Generalist hydrocarbon-degrading bacterial communities in the oil-polluted water column of the North Sea

Panagiota-Myrsini Chronopoulou,1,16 Gbemisola O. Sanni,1,15 Daniel I. Silas-Olu,1 Jan Roelof van der Meer,2 Kenneth N. Timmis,3 Corina P. D. Brussaard4‡ and Terry J. McGenity1* 1School of Biological Sciences, University of Essex, Wivenhoe Park, Colchester CO4 3SQ, UK. 2Department of Fundamental Microbiology, University of Lausanne, Lausanne, Switzerland. 3Institute of Microbiology, Technical University Braunschweig, Braunschweig, Germany. 4Department of Biological Oceanography, Royal Netherlands Institute for Sea Research (NIOZ), Den Burg, The Netherlands.

Summary

The aim of this work was to determine the effect of light crude oil on bacterial communities during an experimental oil spill in the North Sea and in mesocosms (simulating a heavy, enclosed oil spill), and to isolate and characterize hydrocarbon-degrading bacteria from the water column. No oil-induced changes in bacterial community (3 m below the sea surface) were observed 32 h after the experimental spill at sea. In contrast, there was a decrease in the dominant SAR11 phylotype and an increase in Pseudoalteromonas spp. in the oiled mesocosms (investigated by 16S rRNA gene analysis using denaturing gradient gel electrophoresis), as a consequence of the longer incubation, closer proximity of the samples to oil, and the lack of replenishment with seawater. A total of 216 strains were isolated from hydrocarbon enrichment cultures, predominantly belonging to the genus Pseudoalteromonas; most strains grew on PAHs, branched and straight-chain alkanes, as well as many other carbon sources. No obligate hydrocarbonoclastic bacteria were isolated or detected, highlighting the potential importance of cosmopolitan marine generalists like Pseudoalteromonas spp. in degrading hydrocarbons in the water column beneath an oil slick, and revealing the susceptibility to oil pollution of SAR11, the most abundant bacterial clade in the surface ocean.

Introduction

The global use of crude oil has plagued the marine environment with numerous major oil-pollution incidents, resulting in devastating environmental damage to marine habitats with serious socio-economic implications. A recent example is the largest offshore spill in the history of the USA, which occurred when the Deepwater Horizon rig exploded, releasing several hundred million litres of oil into the Gulf of Mexico (Crone and Tolstoy, 2010). Crude-oil components are toxic and stressful to marine organisms, including microorganisms (Sikkema et al., 1995), and if they reach the shore they can persist for decades (Atlas and Bragg, 2009). However, many microbes have developed pathways for hydrocarbon metabolism, some to the extent that they thrive only in the presence of crude-oil components (Dyksterhouse et al., 1995; Yakimov et al., 1998; 2004a; Teramoto et al., 2011; McGenity, 2014). Following oil contamination in marine systems, these obligate hydrocarbonoclastic bacteria (such as Alcanivorax, Cycloclastis, Thalassolituus, and Oleibacter species) typically bloom and become dominant members of the prevailing microbial communities (Kasai et al., 2002; Cappello et al., 2007; McKew et al., 2007b; Teramoto et al., 2009; Vila et al., 2010).

These specialized bacteria are often found in close contact with oil at the oil-water / oil-sediment interface (Schneiker et al., 2006; Gertler et al., 2009; Cappello and Yakimov, 2010). In experimental oil-contaminated tidal mudflat mesocosms, Coulon and colleagues (2012) showed that Alcanivorax sp. constituted almost 50% of the community in the tidal biofilms floating above oiled mudflat cores. Thalassolituus sp. dominated the microbial communities present in the oily phase of water samples obtained from production wells in Canada (Kryachko...
et al., 2012). And, in beach-simulating mesocosms, *Alcanivorax* and *Cycloclasticus* dominated the surfaces of seawater-immersed oil-coated gravel after the addition of nutrients (Kasai et al., 2002). Microbes like *Alcanivorax borkumensis* are exquisitely adapted to biofilm formation, oil solubilization and degradation, and oil-induced stress tolerance (Schneiker et al., 2006; Sabirova et al., 2008).

However, oil floating on the surface of marine waters is broken down into small droplets, dispersed and integrated into the water column to different degrees depending on many factors including wave action (ITOPF, 2002; Gros et al., 2014). Known obligate hydrocarbonoclastic bacteria are not universally encountered in the water column below an oil slick, for example in the Gulf of Mexico (Redmond and Valentine, 2011). Therefore, we hypothesize that below the oil-water interface, specialized hydrocarbonoclastic bacteria are replaced by generalist marine bacteria, perhaps better adapted to grow on low levels or more soluble components of crude oil. Besides, there is paucity of information on the effect of crude oil spills on marine bacteria that play vital roles in global biogeochemical cycles. Therefore, we sought to investigate the impact of an oil spill on pelagic marine bacteria, both in situ and in mesocosms, that were not in direct contact with the oil slick; and to test the hypothesis that some common marine bacteria would be inhibited by the oil. Consequently, we have carried out comparative studies of bacterial community changes in the water column during a small experimental spill of light crude oil in the North Sea and in oil-enriched 1-m³ seawater mesocosms. Furthermore, we have isolated, identified and characterized generalist hydrocarbon-degrading microorganisms from both systems.

**Results**

**Bacterial community changes during the experimental oil spill and in the oil-enriched on-board mesocosms**

Temporal and treatment effects on the bacterial communities in the experimental oil spill at sea and the oil-enriched enclosed mesocosms on-board the ship were investigated by denaturing gradient gel electrophoresis (DGGE) analysis of the bacterial 16S rRNA gene. There was no obvious difference between water-column bacterial communities inside and outside of the experimental light crude oil spill at sea over 32 h (Fig. 1). In the on-board mesocosms, where samples could be taken closer to the oil slick; and to test the hypothesis that some common marine bacteria would be inhibited by the oil. Consequently, we have carried out comparative studies of bacterial community changes in the water column during a small experimental spill of light crude oil in the North Sea and in oil-enriched 1-m³ seawater mesocosms. Furthermore, we have isolated, identified and characterized generalist hydrocarbon-degrading microorganisms from both systems.

Fig. 1. DGGE profiles of amplified bacterial 16S rRNA genes from 1.5 m (out) and 3 m (in) below the sea surface during the experimental oil spill (outside and inside the spill respectively) at different times. Refer to the Experimental Procedures for an explanation of the experimental-spill and sampling strategy.

Bacterial identification by sequencing DGGE bands was done for the on-board mesocosms, focusing on those bands that were most influenced by oil addition (Fig. 2A, Table 1). The main bacteria detected belong to the alphaproteobacterial SAR11 group and the gammaproteobacterial genus *Pseudoalteromonas*. Bands similar to SAR11 types (97–100% 16S rRNA gene identity) were present in the non-oiled mesocosm (B1), in the less oiled mesocosm (BII) and at early time points of the most oiled mesocosm (BIII). However, when oil was present at high concentration or lower concentration but for longer, many bands (D, M1, M2, N) representing SAR11 bacteria, were absent. Nevertheless, some SAR11-related bands
persisted in oiled mesocosms, namely A, K1, K2. In contrast, the bands from *Pseudoalteromonas* spp. increased in intensity in oiled mesocosms, for example band J1, appearing at day 6 in the less-oiled mesocosm, and band J2 present at day 4 and 6 in the most oiled mesocosm (Fig. 2A). Band L, which was present at day 6 in the less-oiled mesocosm, was also similar (98% 16S rRNA gene sequence identity) to *Pseudoalteromonas* sp. Although the UV-treated and oiled mesocosm (BIV) is a very artificial system, having selectively killed certain microbes, the dominant bands over time were most similar to *Pseudoalteromonas haloplanktis*. For example,
bands I1 (99%) and I2 (99%), present at all time points were both similar to this species. Band E1 was also similar (99%) to Pseudoalteromonas sp. In this mesocosm there were also some dense bands, such as band B (97%) and bands F1 (99%) and F2 (100%), similar to Alteromonas spp.

**Isolation and characterization of oil-degrading strains**

Although DGGE analysis suggests that Pseudoalteromonas spp. are probably the main microbes responsible for the degradation of oil in the on-board mesocosms, it is known that many species of this genus do not degrade hydrocarbons. Consequently, Pseudoalteromonas-related bacteria were cultivated to test the role they play in hydrocarbon degradation.

Samples from the experimental spills (Spill-in), accidental slicks (Acc-in) and their non-contaminated controls (Spill-out, Acc-out) and oil-enrichment mesocosms were inoculated into the hydrocarbon-enrichment media, and after subculturing, 216 strains were isolated (Table 2); 47 from the experimental spills [40 strains from inside the spill (Spill-in) and 7 from outside the spill (Spill-out)], 50 from the accidental slicks [25 each from inside the slicks (Acc-in) and outside (Acc-out)], 119 from the on-board mesocosms.

Most strains were isolated from the tetradecane (77 strains) and crude oil (76 strains) enrichment cultures, while 19 strains were isolated from enrichment cultures with 1-methylnaphthalene as the sole carbon source. Sequencing of 16S rRNA genes from all 1-methylnaphthalene-grown isolates and selected isolates from other hydrocarbon-enrichments revealed a predominance of generalist hydrocarbon-degrading members.

### Table 1. Sequencing results of the 16S rRNA gene DGGE bands from the on-board mesocosms.

| Band | Oil Band int. | Mesocosms - Time | Closest Relative | Group | Accession No | Id (%) |
|------|---------------|-----------------|-----------------|-------|--------------|--------|
| A    | —             | BII-2 day       | SAR11 bacterium  | Alpha | JO859613     | 99     |
| B    | ↑             | BIV-6 day       | Alteromonas macleodii | Alpha | JN710109     | 100    |
| C    | ↓             | BIV-2 day       | Pseudoalteromonas sp. M71_D76 | Alpha | HM799000     | 99     |
| E1   | ↑             | BII-2 day       | Pseudoalteromonas sp. MSI02 | Gamma | CP003973     | 97     |
| E2   | ↑             | BIV-4 day       | Pseudoalteromonas sp. 28ox | Gamma | JQ411283     | 99     |
| F1   | ↑             | BIV-4 day       | Alteromonas sp. B348 | Gamma | FN95800     | 99     |
| F2   | ↑             | BIV-6 day       | Alteromonas tagae BCR 17571 | Gamma | NR_043977    | 100    |
| G    | ↓             | BII-2 day       | SAR11 bacterium  | Alpha | JN710109     | 100    |
| H    | ↓             | BII-2 day       | Uncultured marine bacterium | Beta | AF419359     | 99     |
| I1   | ↑             | BIV-4 day       | Pseudoalteromonas sp. BB68 | Gamma | FR693362     | 99     |
| I2   | ↑             | BII-6 day       | Pseudoalteromonas sp. BB68 | Gamma | FR693362     | 99     |
| J1   | ↑             | BII-6 day       | Alteromonas marina ECSMB2 | Gamma | JM66469     | 100    |
| J2   | ↑             | BII-4 day       | Pseudoalteromonas sp. BSI20652 | Gamma | BAD71000269 | 98     |
| K1   | ↓             | BII-2 day       | SAR11 bacterium  | Alpha | JO859613     | 99     |
| K2   | —             | BII-2 day       | SAR11 bacterium  | Alpha | JO859613     | 99     |
| L    | ↑             | BII-6 day       | Pseudoalteromonas marina ECSMB2 | Gamma | JM66469     | 99     |
| M1   | ↓             | BII-2 day       | SAR11 bacterium  | Alpha | JO859613     | 99     |
| M2   | ↓             | BII-2 day       | SAR11 bacterium  | Alpha | JO859613     | 99     |
| N    | ↓             | BII-2 day       | SAR11 bacterium  | Alpha | JO859613     | 99     |

Sequences are based on approximately 130 bp of the 16S rRNA gene. A BLAST search was performed against the NCBI database.

- **a.** Arrows indicate increase (↑) or decrease (↓) over time in oiled mesocosms; Dash symbol (—) indicates no change in band intensity.
- **b.** BI: no oil (control); BII: 2 L of Arabian light crude oil; BIII: 5.5 L of Arabian light crude oil; BIV: 2 L of Arabian light crude oil and UV light-treated (killed control).
- **c.** Alpha = Alphaproteobacteria; Beta = Betaproteobacteria; Gamma = Gammaproteobacteria.
- **d.** Percentage partial 16S rRNA gene sequence identity.

### Table 2. Summary of strains isolated from hydrocarbon-enrichments of North Sea samples.

| Environment | Carbon Source | Media | Number of Isolates |
|-------------|---------------|-------|--------------------|
|             | TOTAL         | T     | F     | P     | M     | Solid | Liquid |
| ON-BOARD   | BI            | 25    | 6     | 8     | 9     | 2     | 12     | 13     |
| MESOCOSMS  | BII           | 26    | 12    | 9     | 3     | 2     | 9      | 17     |
| EXPERIMENTAL | Spill-in     | 40    | 13    | 17    | 10    | 0     | 19     | 21     |
| ACCIDENTAL | Spill-out     | 7     | 3     | 1     | 3     | 0     | 0      | 0      |
| SLICKS     | Acc-in        | 25    | 9     | 10    | 5     | 1     | 14     | 11     |
|            | Acc-out       | 25    | 8     | 9     | 6     | 2     | 8      | 17     |
| TOTAL      | 216           | 77    | 76    | 44    | 19    | 102   | 114    |

- **a.** BI: no oil (control); BII: 2 L of Arabian light crude oil; BIII: 5.5 L of Arabian light crude oil; BIV: 2 L of Arabian light crude oil and UV light-treated (killed control).
- **b.** Spill-in: inside experimental oil spill; Spill-out: outside the experimental spill.
- **c.** Acc-in: inside accidental slicks; Acc-out: outside accidental slicks.
- **d.** T: tetradecane; F: weathered Forties crude oil; P: pristane; M: 1-methylnaphthalene.

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belonging to Gammaproteobacteria but not obligate hydrocarbonoclastic bacteria (Fig. 3). With the exception of strain NS168 related to Glaciecola sp. (100% 16S rRNA gene sequence identity), all the isolates from samples obtained from both within and outside the spill and accidental slicks belonged to the genus Pseudoalteromonas (99.4–100% identity). Strain NS163 (99% identity to Roseovarius strain AP6), isolated from a 1-methylnaphthalene-enrichment, was the only strain belonging to Alphaproteobacteria.

A greater variety of microbes was isolated from the UV-treated control mesocosm (BIV) on-board the ship; in addition to Pseudoalteromonas, a member of the genus Pseudomonas (90% identity to Pseudomonas...
stutzeri 3C68, 93.2% identity to Pseudomonas sp. W1-1) and several isolates belonging to the genus Halomonas (99.4–99.8% identity) were isolated from 1-methylnaphthalene enrichments from mesocosm BIV.

Utilization of different growth substrates

A growth experiment was set up in order to investigate the range of carbon sources that these isolates could utilize for growth, mainly focusing on hydrocarbons. Based on the phylogenetic tree (Fig. 3), eight phylogenetically distinct Pseudoalteromonas strains (two from each hydrocarbon enrichment), six phylogenetically distinct strains of Halomonas, and the Glaciecola and Roseovarius strains, were selected.

The results of these assays are summarized in Fig. 4. All the isolates were able to grow with Forties crude oil, pyruvate and the amino acids alanine and arginine as sole carbon sources. With the exception of strain Glaciecola NS168 all strains grew on glucose. Acetate and tetradeacane supported growth of all Pseudoalteromonas strains, while only three strains of Halomonas (NS159, NS161, NS162) grew with tetradeacane. However, no isolates grew on benzoate, cyclohexane, benzene, toluene, pyrene or biphenyl; while methanol supported growth of four strains of Halomonas (NS159, NS161, NS162 and NS165), Glaciecola (NS168) and Roseovarius (NS163). Some isolates grew on a wide range of hydrocarbons whereas others seemed to specialize in particular classes of hydrocarbons. For example, Pseudoalteromonas strain NS50 grew on linear alkanes (decane, tetradeacane and eicosane), branched alkanes (pristane and squalane) and PAHs (phenanthrene, anthracene and fluorene). Pseudoalteromonas strain NS151, Halomonas strains NS159, NS161, NS162 and Pseudoalteromonas NS164, isolated from 1-methylnaphthalene enrichments, did not grow on any other aromatic hydrocarbon but grew on \(n\)-alkanes and branched alkanes. Generally, linear alkanes supported the growth of most of the isolates.

Crude-oil biodegradation by selected isolates

The components of crude oil consumed by isolates NS163 (99% 16S rRNA gene sequence identity to Roseovarius sp. AP6), NS168 (100% identity to Glaciecola sp. MOLA 37), NS159 (99.6% identity to Halomonas sp. MAN K33) and isolate NS164 (99% identity to Pseudoalteromonas elyakovii) were identified by growing them on 1% v/v weathered Forties crude oil for 12 days (except strain NS164 that was incubated for 28 days) and measuring the hydrocarbon components remaining. There was a significant reduction of Total Petroleum Hydrocarbon (TPH) (Tukey’s HSD: \(P < 0.05\)) in all cultures, confirming that all the isolates were oil degraders, with isolates NS159 (Halomonas sp.) and NS168 (Glaciecola sp.) exhibiting similar degradative potentials (data not shown).

These four isolates exhibited a preference for alkanes to PAHs. Total alkanes significantly reduced by 36.9, 42.1, 44.7% for strains NS163, NS168 and NS159 respectively.

Fig. 4. Phenogram of sequenced isolates based on the pattern of growth substrate utilization. All negative for Biphenyl, Pyrene, Toluene, Benzene, Cyclohexane and Sodium Benzoate. All positive for Pyruvate, L-Alanine, L-Arginine, Forties crude oil. Square brackets contain substrates utilized by corresponding isolate. (Glu = Glucose; Al = L-Alanine; Ac = K-acetate; Me = Methanol; Sq = Squalane; T = Tetradeacane; Pr = Pristane; De = Decane; He = Hexacosane; Phn = Phenanthrene; Ei = Elcosane; Fl = Fluorene; An = Anthracene). * = One out of two cultures grew on the substrate.

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after 12 days of incubation, and by 54% for strain NS164 after 28 days (Tukey’s HSD: $P < 0.05$). In contrast, total PAH values were unchanged.

The extent of biodegradation of individual alkane components (C$_{11}$–C$_{33}$) in Forties crude oil by the four isolates was assessed (Fig. 5). Strain NS159 significantly degraded all alkane components although those in the range of C$_{19}$–C$_{28}$ (including the branched-chain alkanes) were least degraded. Strain NS163 and NS168 did not significantly degrade C$_{11}$, C$_{12}$ and C$_{20}$ $n$-alkanes (Tukey’s HSD: $P > 0.05$), but all other alkanes were degraded. *Pseudoalteromonas* strain NS164 had a different degradation profile, significantly degrading most of the alkane components (Tukey’s HSD: $P < 0.05$) except pristane and C$_{29}$ and C$_{31}$ alkanes.

**Discussion**

Open-sea experimental spill versus the on-board mesocosms

We took this rare opportunity to investigate the effects of a light crude-oil spill on the bacterial community in the marine water column under natural conditions. When sampling from inside the experimental spill, it was essential to minimize disturbance of the oil so that other measurements would not be compromised and to avoid breakup of the slick, and so sampling was performed 3 m below the surface. The open-sea oil slick had dissipated by 32 h, and, for the reasons described by Brussaard and colleagues (2010), production of a larger, contained slick was not feasible. Therefore, in parallel, on-board mesocosms were established to simulate a more severe oil spill in a restricted area, e.g. in a harbour or bay. DGGE analysis revealed that bacterial communities in the on-board mesocosms changed rapidly upon oil addition, with a decline in most SAR11 phylotypes and a concomitant increase in *Pseudoalteromonas* spp. (see later discussion). In contrast, the bacterial community composition in the water column inside and outside the experimental oil spill did not change over the course of the 32-hour experiment. Three factors are likely to have contributed to this difference between open-sea and enclosed-mesocosm experiments. 1) The timescale of the on-board mesocosm was much longer (7 days), and
indeed most differences became apparent after 2 days, and so insufficient time had elapsed to allow a large population of hydrocarbon degraders to develop in the open-sea experiment. 2) The availability of oil was lower in the open-sea experiment, due to: horizontal spreading of the Arabian light crude oil, lower oil loading per unit area, and greater depth of sampled water, and thus distance from the oil source. 3) In contrast to the simulation of an enclosed spill, the open sea would have received a flow of seawater from uncontaminated areas, containing a new supply of nutrients and microbes. These findings emphasize that complementary approaches, including experimental oil spills at sea, are required to investigate the microbial responses to crude oil; and that scaling-up data from mesocosms to the open sea requires careful consideration.

Negative impact of crude oil on the SAR11 clade

Dominant bands in the uncontaminated mesocosm (BI) and in the less-oiled mesocosm (BII) or at early stages in the most oiled one (BIII) were similar (97–100%) to SAR11, a clade of Alphaproteobacteria and the most abundant phylotype in the surface ocean (Morris et al., 2002). SAR11 varies widely in spatial and temporal abundance, but generally represents the dominant bacterioplankton taxon in the North Sea, constituting up to 37% (Giebel et al., 2011) or 47% (Sperling et al., 2012) of total bacterial 16S rRNA genes.

Candidatus Pelagibacter ubique strain HTCC1062 is the first cultured member of the clade and has one of the smallest genomes of autonomously replicating bacteria (Giovannoni et al., 2005). Evidence from this strain and in-situ studies have shown that SAR11 bacteria are predominantly chemoorganoheterotrophs, utilizing glucose, amino acids, trimethylamine N-oxide and dimethylsulfoniopropionate (Malmstrom et al., 2004; Laghdass et al., 2012; Lidbury et al., 2014), but when starved of organic carbon, growth is enhanced by light-energy capture via proteorhodopsin (Steindler et al., 2011). Importantly, our study has provided experimental evidence that a prolonged dose of light crude oil has a negative impact on this bacterial clade that has such a pivotal role in marine ecology and biogeochemical processes: most SAR11-derived bands decreased in intensity with time in the oiled mesocosms compared with the non-oiled mesocosm, and were absent after day 4 in the mesocosm receiving most oil (BIII). The detrimental effects of oil on SAR11 phyatypes were also observed in diesel-polluted mesocosms containing surface water from a coastal area of southwest Mallorca (Lanfranconi et al., 2010). Nogales and colleagues (2007) also noted the absence of SAR11 in polluted harbour waters. Moreover, Redmond and Valentine (2011) found SAR11 to be among the dominant bacteria in surface water collected from three sites of the Deepwater Horizon spill that had a thin oil sheen, but they were undetected or represented by <1% of 16S rRNA sequences in two sites with a thicker oil coating. Also in the Gulf of Mexico, but this time in the oil plume in deeper water, most of the abundant taxa showed an increase in gene expression; in contrast SAR11 transcript abundance was lower inside than outside the plume, but not significantly so (Rivers et al., 2013). Little attention is given to the impact of crude oil on marine bacteria, yet we showed that there are clear dose- and time-dependent effects on SAR11. It will be important to determine what effect spills of different magnitude may have on such microbes, as well as the resulting impact on cycling of elements (e.g. N, P and Fe) that are required for efficient hydrocarbon degradation. The mechanisms by which crude oil inhibits SAR11 cells remain to be elucidated, but may include: direct interaction of hydrocarbons with cells, e.g. damaging membranes; reduced light availability; potentially affecting sensory processes or energy generation; and/or indirect processes, such as enhanced competition with hydrocarbon-degrading microbes for nutrients.

The importance of nutritionally diverse bacteria, such as Pseudoalteromonas spp., in hydrocarbon degradation in the marine water column

Pseudoalteromonas spp. are ubiquitous in marine environments, representing on average 2.6% of the surface bacterioplankton along a latitudinal gradient (Wietz et al., 2010), but less than 1% in one location in the southern North Sea (Eilers et al., 2000). Pseudoalteromonas-related bands became dominant in oiled mesocosms (BII, BIV), and Pseudoalteromonas spp. were the most commonly isolated bacteria, including from the open-sea spill. This, coupled with the demonstrated ability of these isolates to degrade a wide range of hydrocarbons, implies that they were the main microbes responsible for hydrocarbon degradation in the water column. No obligate hydrocarbonoclastic bacteria (OHCBs) were detected or isolated from the experimental spill or the mesocosms. This was not due to the choice of media, because in another study of mudflat sediments using the same procedures, OHCBs belonging to the genera Alcanivorax, Cycloclasticus and Thalassolituus were isolated in abundance (Sanni et al., unpublished). OHCBs such as Alcanivorax borkumensis and Marinobacter hydrocarbonoclasticus form biofilms that aid their attachment to oil-water interfaces where they carry out degradation (Schneiker et al., 2006; Duran, 2010), and so they may well have been present on the unsampled oil-water surface. However, an intriguing possibility is that the anti-biofilm activity of Pseudoalteromonas (Dheilly et al., 2012).
2010; Klein et al., 2011) may have inhibited oil-attached OHCBs.

The closest known relatives of the oil-degrading Pseudoalteromonas strains isolated in this study (e.g. P. haloplanktis, P. elyakovii, P. distincta, P. atlantica) are not known to degrade hydrocarbons. However, representatives of this genus have been detected in a variety of oil-contaminated environments and oiled microcosms, and shown to degrade hydrocarbons. Notably, in one of the two surface water samples from the Gulf of Mexico that had a thick layer of oil, and that was mentioned previously because of the absence of SAR11 (Redmond and Valentine, 2011), Pseudoalteromonas spp. were isolated from plume water samples during the Deepwater Horizon spill (Gutierrez et al., 2013). Pseudoalteromonas spp. increased in abundance in deeper waters of the Gulf-of-Mexico plume, after hydrocarbon exposure, especially after alkanes had been depleted and aromatics remained (Dubinsky et al., 2013). Also, Pseudoalteromonas clones were detected in seawater microcosms amended with crude oil and incubated at 4°C and 20°C (Yakimov et al., 2004b; Coulon et al., 2007), and in PAH enrichment-microcosms of seawater amended with nutrients over a period of 9 weeks (McKew et al., 2007a). Pseudoalteromonas spp. have been isolated from phenantherene, chrysenene and naphthalene-enrichment cultures of marine sediments (Melcher et al., 2002; Hedlund and Staley, 2006; Cui et al., 2008). The biodegradation potential of Pseudoalteromonas spp. for both aliphatic and aromatic compounds of crude oil has been confirmed in a consortium derived from arctic ice and seawater (Deppe et al., 2005) and in a consortium from the Korean Western coast (Cho and Oh, 2012), whereas a strain of Pseudoalteromonas has been found to preferentially metabolize short-chain alkanes in the range of C_{12}-C_{16} in arctic sediments (Lin et al., 2009). All of our Pseudoalteromonas strains grew on a range of carbon and energy sources, and varied in the breadth of utilization of hydrocarbons, with all but three strains able to grow on straight-chain and branched alkanes as well as PAHs, but none grew on toluene. In contrast, OHCBs, like Alcanivorax and Cyloclasticus spp., tend to be more specialized, degrading alkanes and PAHs, respectively (see Cappello and Yakimov, 2010; Staley, 2010).

Interactions between microbes are as important in oil degradation as they are in other biogeochemical processes (McGenity et al., 2012), and the ability of Pseudoalteromonas spp. to remove potential competitors has been mentioned previously. Pseudoalteromonas spp. may also be beneficial to co-existing microbes by producing EPS that has the potential to serve as a biosurfactant (Nichols et al., 2005; Saravanan and Jayachandran, 2008; Cho and Oh, 2012). It will be important to elucidate the relative contribution of generalists like Pseudoalteromonas and specialists like Alcanivorax, and to understand to what extent they interact. Pseudoalteromonas spp. will be at an advantage owing to their higher abundance in unpolluted seawater, allowing them to become established more rapidly after an oil spill; however, the burden of carrying genes that are not required for hydrocarbon biodegradation is likely to make them less competitive over time.

Other hydrocarbon-degrading isolates

Whereas Pseudoalteromonas was both shown to be abundant in oiled mesocosms and readily cultivable on hydrocarbon-enriched marine media, hydrocarbon-degrading strains from other genera were also isolated but not shown to have increased in abundance in the oiled mesocosms. Six strains were related to species of Halomonas (more than 99% percent identity), all of which derived from UV-treated and oiled mesocosm BIV and had been enriched with 1-methylnaphthalene as sole carbon source. Like Pseudoalteromonas strains, they grew on a wide range of hydrocarbons, including branched and straight-chain alkanes and PAHs. GC-MS analysis of one strain demonstrated metabolism of almost all the alkane components of crude oil but not the aromatic components within 12 days, implying a preference for alkanes when both alkane and PAHs are present as crude oil components. Halomonas spp. have been reported to degrade aromatic (Melcher et al., 2002; Garcia et al., 2004; Cui et al., 2008) and aliphatic hydrocarbons (Pepi et al., 2005; Mnif et al., 2009) in marine and hypersaline environments.

One strain had 99% 16S rRNA sequence identity with two members of the Roseobacter clade: Roseovarius sp. AP6 which was isolated by Alonso-Gutierrez and colleagues (2009) from shoreline samples contaminated with oil from the Prestige spill, and Roseovarius sp. DG944 found in association with an alga, Gymnodinium catenatum, which causes paralytic shellfish poisoning in marine waters (Green et al., 2004). Roseobacter spp. were found in abundance in both n-alkane- (McKew et al., 2007a) and oil- (Brakstad and Lødeng, 2005) enriched seawater and also in oil-contaminated seawater (Prabagaran et al., 2007). Similarly, species of Roseovarius have been implicated in the degradation of PAHs (Harwati et al., 2007; Wang et al., 2008; Vila et al., 2010), n-alkanes and pristane (Harwati et al., 2007).

Species of Glaciecola are usually found in polar seas and their coastal environments, Arctic and Antarctic sea-ice in association with marine invertebrates (Romanenko et al., 2003; Yong et al., 2007) and clones have been detected in oil-contaminated samples from such
environments (Prabagaran et al., 2007; Brakstad et al., 2008; Reberg et al., 2011). Yakimov and colleagues (2004b) also isolated a strain of *Glaciecola* from tetradecane-enrichments of Antarctic seawater. However, these studies did not present evidence to support the direct participation of these species in hydrocarbon degradation, whereas our *Glaciecola* strain NS168 grew on most of the *n*-alkanes tested and on pristane.

It is noteworthy that the four isolates (*Roseovarius* sp. strain NS163, *Glaciecola* sp. strain NS168, *Halomonas* sp. strain NS159 and *Pseudoalteromonas* sp. strain NS164) did not degrade any of the PAH components of crude oil during the period of incubation (as evidenced by GC-MS analysis), even though they were all isolated on 1-methylnaphthalene-enrichments and the first three strains grew on phenanthrene in the growth substrate assay. This implies that the strains have a preference for alkanes when both alkane and PAHs are present as crude oil components but could metabolize some PAHs in the absence of alkanes.

**Selection of Alteromonas sp. by UV-light treatment**

Although the primary purpose of the UV-treated mesocosm (BIV) was to allow analysis of *in-situ* hydrocarbon biodegradation, it was a source of several strains of hydrocarbon-degrading microbes. Hydrocarbons were a selective force in this mesocosm, but also a dense band indicating the presence of *Alteromonas*, not detected in other oiled mesocosms, could be a consequence of the resistance of representatives of this genus to ultraviolet light (Agogué et al., 2005) and their ability to grow rapidly on the organic matter released from cells killed by UV light in the same way that they are often associated with phytoplankton blooms (Tada et al., 2012) and exudates (Nelson and Carlson, 2012).

**Environmental implications**

Several factors have been proposed to explain the patchy horizontal distribution of important marine taxa, such as SAR11, in marine environments (Sperling et al., 2012); however, our study shows that consideration should be given to oil-pollution incidents. Also, a ten-fold enrichment of *Pseudoalteromonas* spp., and absence of SAR11, in the sea-surface microlayer compared with underlying North Sea water (Franklin et al., 2005), may in part be explained by the accumulation of hydrocarbons in the sea-surface microlayer (Wurl and Obbard, 2004) and the demonstrated sensitivity of SAR11 to, and predilection of *Pseudoalteromonas* spp. for, hydrocarbons.

It has been proposed that ecological damage caused by oil spills might be reduced by the addition of microbes like *Alcanivorax* spp. which, although naturally present in contaminated sites, require considerable time to reach population densities needed for maximum biodegradation rates. The work presented here shows that such bioaugmentation strategies may require multiple species with different specialities, including oil-attached specialists like *Alcanivorax* and planktonic generalists such as *Pseudoalteromonas*.

**Experimental procedures**

**Sample site and sample collection**

The experimental oil spill. Details of the permitted, controlled, experimental spill and on-board mesocosms are provided by Brussaard and colleagues (2010). As part of an EC project FACEiT (Fast-Advanced Cellular and Ecosystem Information Technologies), an experimental oil spill was organized by the Royal Netherlands Institute for Sea Research (NIOZ), Texel, in collaboration with the Netherlands Ministry of Transport, Public Works and Water Management, RWS Noordzee (RWS-NZ); the spill was carried out on 8 May 2008 in Netherlands Exclusive Economic Zone waters. To aid the visualization and tracking of the oil slick, 5 kg of the fluorescent dye rhodamine WT (25 L 20% w/v) was mixed with 1 m$^3$ of seawater and discharged together with 5000 L of Arabian Light crude oil into Dutch waters. In addition to the dye, two drifting buoys were used to locate the position of the oil slick especially when the rhodamine had diluted beyond the limit of detection and it thus became difficult to follow the slick. Time zero (0 h) corresponds with the end-point of oil and dye addition, which took approximately 30 min. In order to prevent RV Pelagia from disturbing the experimental spill, a rubber boat was dispatched to collect water samples from inside the spill (Spill-in) using a specially designed sampler that gently pumped seawater from 3 m depth into a metallic jerry can. The tube used was attached to a metallic inlet that was passed closed through the surface oil layer, preventing oil droplets from contaminating the sample. Samples were taken at 0, 3, 9, 12, 24 and 32 h after addition of oil. Within 30 min of these time points, corresponding control samples were collected from waters at least 500 m from the spill with no apparent oil on the surface, and ensuring no cross-contamination with the added oil by accounting for wind, tide and current. These samples, called ‘Spill-out’, were taken at a depth of 1.5 m using Niskin bottles (10 L each) mounted on a Rosette sampler equipped with Seabird conductivity-temperature-depth (CTD) sensors and a natural chlorophyll autofluorescence sensor.

Accidental slicks. Samples were collected from oil slicks encountered in shipping lanes within a section of the Dutch EEZ rich of oil platforms (southeast of Dogger bank) (Acc-in) at 1.5 m water depth. Corresponding control samples (Acc-out) were collected in clean waters outside the slicks at the same depth.
Oil-enriched enclosed mesocosms. In addition to the experimental spill, four 1 m³ mesocosms (called Cube vessels) were set up on board the RV Pelagia on the 6th May, 2008. The Cube vessels were filled with seawater (1000 L) from the Dogger Bank, a biologically productive location containing waters with no obvious oil contamination and high biomass. Cube 1 (B1) served as the non-oiled control, Cube 2 (B2) contained 2 L of Arabian Light Crude oil, Cube 3 (B3) contained 5.5 L of crude oil, and Cube 4 (B4) contained 2 L of oil, but was treated with UV light to sterilize the mesocosm ('killed control'). Each Cube vessel had three taps: at the top, middle and bottom. As the level of water in each mesocosm reduced, samples were taken from the tap closest to the surface of the water (approximately 15–20 cm from the surface).

The mesocosms were sampled on the 7th May, 2008 before the addition of oil at 08:15 (t = 0), then at 2 h, 6 h, 12 h after oil addition. Further samples were obtained daily at 13:00 for the next 7 days.

Initial processing of samples

Aliquots of seawater sampled from all environments were immediately inoculated into hydrocarbon-enrichment media while the rest were stored on board the RV Pelagia and transported at 4°C back to the University of Essex lab. Microbial cells were also immediately collected on 0.2 μm pore size filters by filtering 1.2 L of seawater samples with Nalgene bottle-top filter units. Seawater samples (pristine, experimental spill and accidental spills) were filtered with Sartorius gridded cellulose nitrate filters (47 mm diameter, 0.2 μm pore size) while samples from the Cube vessels were filtered with Durapore PVDF filters (47 mm diameter, 0.22 μm pore size). The filters were initially stored at 4°C for 12 h before centrifugation in a Sorvall Biofuge Stratos centrifuge at 12,000 g for 15 min at 4°C to pellet cells. Total hydrocarbon was solvent extracted from the liquid contents of the serum bottles as described by Coulon and colleagues (2007), and then frozen at – 80°C.

Hydrocarbon enrichment cultures

Hydrocarbon enrichment cultures in liquid media were established by inoculating water samples (200 μL) into tubes containing 10 ml of ONR7a medium (Dyksterhouse et al., 1995) using 1% v/v filter-sterilized tetradecone, weathered Forties crude oil, pristane or 1-methylnaphthalene as sole carbon sources. Tubes containing ONR7a medium with hydrocarbons had been prepared, capped and crimp-sealed with PTFE-lined silicon septa prior to inoculation. Subculturing was done by carrying out 10-fold serial dilutions of the cultures and spreading 50 μL from the 10⁻², 10⁻⁵ and 10⁻⁸ dilutions on ONR7a agar plates containing sterile GF/C filter papers soaked with 125 μL of the hydrocarbons on the lids of the Petri dishes.

All hydrocarbon enrichment cultures using solid media were prepared with washed agar. These enrichments were prepared by spreading 100 μL of seawater samples (that had been preserved at 4°C for a maximum of three weeks) onto ONR7a agar with hydrocarbon-soaked GF/C filter papers placed on the lids of the Petri dishes. Each of the four hydrocarbons (125 μL) was used as sole carbon source. All cultures were incubated at 12°C, representing the in-situ temperature at the time of sampling.

Nucleic acid extraction, polymerase chain reaction, denaturing gradient gel electrophoresis, and sequencing

DNA was extracted from cultures by boiling the bacterial cells in diethyl pyrocarbonate-treated water (DEPC water), followed by PCR amplification of bacterial 16S rRNA genes using primers described by Lane (1991). The 16S rRNA amplicons of bacterial isolates were sent to GATC Biotech (http://www.gatc-biotech.com/en/index.html) for sequencing.

DNA from filtered water was extracted following bacterial cell lysis with phenol : chloroform : isoamylalcohol (25:24:1 v/v) as described by McKew and colleagues (2007a). PCR of bacterial 16S rRNA gene from this DNA was performed using primers described by Muyzer and colleagues (1993), followed by denaturing gradient gel electrophoresis (DGGE) employing a denaturing gradient from 40 to 60% (McKew et al., 2007a). Bands were excised and sequenced on an ABI Prism 3100 Genetic Analyser.

Full details of these methods and associated analyses are given in the supporting information.

Accession numbers

The DNA sequences reported in this study were deposited in the EMBL database under the accession numbers HE961976 to HE962017.

Growth substrate assays

The ability of selected isolates to utilize different carbon sources was examined by growing the isolates (100 μL) in duplicate tubes containing sterile ONR7a liquid medium (10 ml) and various carbon sources (0.1% v/v). The growth substrates included Forties crude oil, n-alkanes (decane, tetradecane, eicosane and hexacosane), branched alkanes (pristane and squalane), cyclic alkane (cyclohexane), aromatic hydrocarbons (benzene, toluene, phenanthrene, anthracene, pyrene, fluorene and biphenyl), amino acids (L-alanine and L-arginine), D-glucose, Na-benzoate, Na-pyruvate, K-acetate and methanol. Details are provided in the supporting information.

Biodegradation of crude oil by selected isolates

Detailed biodegradation studies were carried out on four selected isolates. Washed, suspended cells (200 μL) were inoculated in triplicate into serum bottles (125 ml) containing sterilized ONR7a medium (20 ml) with weathered Forties crude oil (1% v/v) and incubated on a rotary shaker (Innova 2300; New Brunswick Scientific) in the dark at 100 rpm for 12 days (isolate NS164 was incubated for 28 days of 12 h light-dark cycles). After incubation, the cultures were vigorously vortexed for 10 s to dislodge bacterial cells from the crude oil and centrifuged in a Sorvall Biofuge Stratos centrifuge at 8,500 g for 15 min at 4°C to pellet cells. Total hydrocarbon was solvent extracted from the liquid contents of the serum bottles as described by Coulon and colleagues (2007), and hydrocarbons were quantified using a Thermo Trace GC gas chromatograph attached to a Thermo Trace DSQ® mass spectrometer. Details are given in the supporting information.

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Statistical analysis

Similarity between DGGE profiles was calculated based on a manually produced binary matrix of the presence/absence of DGGE bands. A proximity matrix based on the Pearson correlation coefficient was used to construct a Multidimensional Scaling (MDS) plot in XLSTAT Version 2008.1.02. A phenogram based on the growth substrate utilization pattern of sequenced isolates was generated with PAST (Paleontological statistics) 2.17 software package (Hammer et al., 2001) using the Ward's method and Euclidean distance to demonstrate the dissimilarities in substrate utilization by the isolates. Significant hydrocarbon degradation was determined by analysis of variance coupled with Tukey’s HSD and Dunnett’s two sided tests. These tests were performed with XLSTAT 2011 (Addinsoft™).

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Conflict of interest

None declared.

References

Agogué, H., Joux, F., Obernosterer, I., and Lebaron, P. (2005) Resistance of marine bacterioneuston to solar radiation. Appl Environ Microbiol 71: 5282–5289.
Alonso-Gutierrez, J., Figueras, A., Albaiges, J., Jimenez, N., Vinas, M., Solanas, A.M., and Novoa, B. (2009) Bacterial communities from shoreline environments (Costa da Morte, northwestern Spain) affected by the Prestige oil spill. Appl Environ Microbiol 75: 3407–3418.
Atlas, R., and Bragg, J. (2009) Bioremediation of marine oil spills: when and when not – the Exxon Valdez experience. Microb Biotechnol 2: 213–221.
Brakstad, O.G., and Lødeng, A.G. (2005) Microbial diversity during biodegradation of crude oil in seawater from the North Sea. Microb Ecol 49: 94–103.
Brakstad, O.G., Nonstad, I., Faksness, L.G., and Brandvik, P.J. (2008) Responses of microbial communities in Arctic sea ice after contamination by crude petroleum oil. Microb Ecol 55: 540–552.
Brussaard, C.P.D., Peperzak, L., Witte, Y., and Huisman, J. (2010) An experimental oil spill at sea. In Handbook of Hydrocarbon and Lipid Microbiology. Timmis, K.N., McGenity, T.J., van der Meer, J.R., and de Lorenzo, V. (eds). Berlin Heidelberg: Springer, pp. 1723–1728.
Cappello, S., Caruso, G., Zampino, D., Monticelli, L.S., Maimone, G., Denaro, R., et al. (2007) Microbial community dynamics during assays of harbour oil spill bioremediation: a microscale simulation study. J Appl Microbiol 102: 184–194.
Cho, S.H., and Oh, K.H. (2012) Removal of Crude Oil by Microbial consortium isolated from oil-spilled area in the Korean western coast. Bull Environ Contam Toxicol 89: 680–685.
Coulon, F., McKeown, B.A., Osborn, M.A., McGenity, T.J., and Timmis, K.N. (2007) Effects of temperature and biostimulation on oil-degrading microbial communities in temperate estuarine waters. Environ Microbiol 9: 177–186.
Coulon, F., Chronopoulou, P., Fahy, A., Paissé, S., Gofi-Urriza, M., Peperzak, L., et al. (2012) Central role of dynamic tidal biofilms dominated by aerobic hydrocarbonoclastic bacteria and diatoms in the biodegradation of hydrocarbons in coastal mudflats. Appl Environ Microbiol 78: 3638–3648.
Crone, T.J., and Tolstoy, M. (2010) Magnitude of the 2010 Gulf of Mexico oil leak. Science 330: 634.
Cui, Z., Lai, Q., Dong, C., and Shao, Z. (2008) Biodiversity of polycyclic aromatic hydrocarbon-degrading bacteria from deep sea sediments of the Middle Atlantic Ridge. Environ Microbiol 10: 2138–2149.
Deppe, U., Richnow, H.H., Michaels, W., and Antranikian, G. (2005) Degradation of crude oil by an arctic microbial consortium. Extremophiles 9: 461–470.
Dheilly, A., Soum-Soutera, E., Klein, G.L., Bazire, A., Compere, C., Haras, D., and Dufour, A. (2010) Antibiofilm activity of the marine bacterium Pseudoalteromonas sp. strain 3J6. Appl Environ Microbiol 76: 3452–3461.
Dubinsky, E.A., Conrad, M.E., Chakraborty, R., Bill, M., Borglin, S.E., Hollibaugh, J.T., et al. (2013) Succession of hydrocarbon-degrading bacteria in the after-math of the Deepwater Horizon oil spill in the Gulf of Mexico. Environ Sci Technol 47: 10860–10867.
Duran, R. (2010) Marinobacter. In Handbook of Hydrocarbon and Lipid Microbiology. Timmis, K.N., McGenity, T.J., van der Meer, J.R., and de Lorenzo, V. (eds). Berlin Heidelberg: Springer, pp. 1725–1735.
Dyksterhouse, S.E., Gray, J.P., Herwig, R.P., Lara, J.C., and Staley, J.T. (1995) Cycloclasticus pugetii gen. nov., sp. nov., an aromatic hydrocarbon-degrading bacterium from marine sediments. Int J Syst Bacteriol 45: 116–123.
Eilers, H., Pernthaler, J., Glöckner, F.O., and Amann, R. (2000) Culturability and in situ abundance of pelagic bacteria from the North Sea. Appl Environ Microbiol 66: 3044–3051.
Franklin, M.P., McDonald, I.R., Bourne, D.G., Owens, N.J.P., Upstill-Goddard, R.C., and Murrell, J.C. (2005) Bacterial diversity in the bacterioneuston (sea surface microlayer): the bacterioneuston through the looking glass. Environ Microbiol 7: 723–736.
Garcia, M.T., Mellado, E., Ostos, J.C., and Ventosa, A. (2004) Halomonas organivorans sp. nov., a moderate halophile able to degrade aromatic compounds. Int J Syst Evol Microbiol 54: 1723–1728.

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K.N., McGenity, T.J., van der Meer, J.R., and de Lorenzo, V. (eds). Berlin Heidelberg: Springer, pp. 1738–1748.
Cappello, S., Caruso, G., Zampino, D., Monticelli, L.S., Maimone, G., Denaro, R., et al. (2007) Microbial community dynamics during assays of harbour oil spill bioremediation: a microscale simulation study. J Appl Microbiol 102: 184–194.
Cho, S.H., and Oh, K.H. (2012) Removal of Crude Oil by Microbial consortium isolated from oil-spilled area in the Korean western coast. Bull Environ Contam Toxicol 89: 680–685.
Coulon, F., McKeown, B.A., Osborn, M.A., McGenity, T.J., and Timmis, K.N. (2007) Effects of temperature and biostimulation on oil-degrading microbial communities in temperate estuarine waters. Environ Microbiol 9: 177–186.
Gertler, C., Gerdzs, G., Timmis, K.N., and Golyshin, P.N. (2009) Microbial consortia in mesocosm bioremediation trial using oil sorbents, slow-release fertilizer and bioaugmentation. *FEMS Microbiol Ecol* 69: 288–300.

Giebel, H.-A., Kalhoefer, D., Lemke, A., Thole, S., Gahl-Janssen, R., Simon, M., and Brinkhoff, T. (2011) Distribution of *Roseobacter* RCA and SAR11 lineages in the North Sea and characteristics of an abundant RCA isolate. *ISME J* 5: 8–19.

Giovannoni, S.J., Bibbs, L., Cho, J.C., Stapels, M.D., Desiderio, R., Vergin, K.L., et al. (2005) Proteorhodopsin in the ubiquitous marine bacterium SAR11. *Nature* 438: 82–85.

Green, D.H., Llewellyn, L.E., Negri, A.P., Blackburn, S.I., and Bolch, C.J. (2004) Phylogenetic and functional diversity of the cultivable bacterial community associated with the paralytic shellfish poisoning dinoflagellate *Gymnodinium catenatum*. *FEMS Microbiol Ecol* 47: 345–357.

Gros, J., Deedar, N., Würz, B., Wick, L.Y., Brussaard, C.P.D., Huisman, J., et al. (2014) First day of an oil spill on the open sea: early mass transfers of hydrocarbons to air and water. *Environ Sci Technol* 48: 9400–9411.

Gutierrez, T., Singleton, D.R., Berry, D., Yang, T., Aitken, L., Hedlund, B., and Teske, A. (2013) Hydrocarbon-degrading bacteria enriched by the Deepwater Horizon oil spill identified by cultivation and DNA-SIP. *ISME J* 7: 2091–2104.

Hammer, Ø., Harper, D.A.T., and Ryan, P.D. (2001) PAST: paleontological statistics software package for education and data analysis. *Palaeontol Electronica* 4: 1–9.

Harwati, T.U., Kasai, Y., Kodama, Y., Susilaningsih, D., and Watanabe, K. (2007) Characterization of diverse hydrocarbon-degrading bacteria isolated from Indonesian seawater. *Microbes Environ* 22: 412–415.

Hedlund, B., and Staley, J. (2006) Isolation and characterization of *Pseudoalteromonas* strains with divergent polycyclic aromatic hydrocarbon catabolic properties. *Environ Microbiol* 8: 178–182.

IOPF (2002) Fate of marine oil spills. The International Tanker Owners Pollution Fertilization Limited 2: 1–8.

Kasai, Y., Kishira, H., and Harayama, S. (2002) Bacteria belonging to the genus *Cyclocaslicus* play a primary role in the degradation of aromatic hydrocarbons released in a marine environment. *Appl Environ Microbiol* 68: 5625–5633.

Klein, G.L., Soum-Soutera, E., Guede, Z., Bazire, A., Compere, C., and Dufour, A. (2011) The anti-biofilm activity secreted by a marine *Pseudoalteromonas* strain. *Biofouling* 27: 931–940.

Kryachko, Y., Dong, X., Sensen, C.W., and Voordouw, G. (2012) Compositions of microbial communities associated with oil and water in a mesothermic oil field. *Antonie Van Leeuwenhoek* 101: 493–506.

Laghdass, M., Catala, P., Caparros, J., Oriol, L., Lebaron, P., and Obermisterer, I. (2012) High contribution of SAR11 to microbial activity in the North West Mediterranean Sea. *Microb Ecol* 63: 324–333.

Lane, D.J. (1991) 16S/23S rRNA sequencing. In *Nucleic Acid Techniques in Bacterial Systematics*. Stackebrandt, E., and Goodfellow, M. (eds). Chichester: John Wiley & Sons, pp. 115–175.

Lanfranconi, M.P., Bosch, R., and Nogales, B. (2010) Short-term changes in the composition of active marine bacterial assemblages in response to diesel oil pollution. *Microb Biotechnol* 3: 607–621.

Lidbury, I., Murrell, J.C., and Chen, Y. (2014) Trimethylamine N-oxide metabolism by abundant marine heterotrophic bacteria. *Proc Natl Acad Sci USA* 111: 2710–2715.

Lin, X., Yang, B., Shen, J., and Du, N. (2009) Biodegradation of Crude Oil by an Arctic Psychrotrophic Bacterium *Pseudoalteromonas* sp. P29. *Curr Microbiol* 59: 341–345.

McGenity, T.J. (2014) Hydrocarbon biodegradation in intertidal wetland sediments. *Curr Opin Biotechnol* 27: 46–54.

McGenity, T.J., Folwell, B.D., McKew, B.A., and Sanni, G.O. (2012) Marine crude-oil biodegradation: a central role for interspecies interactions. *Aquat Biosyst* 8: 10.

McKew, B.A., Coulon, F., Osborn, A.M., Timmis, K.N., and McGenity, T.J. (2007a) Determining the identity and roles of oil-metabolizing marine bacteria from the Thames estuary, UK. *Environ Microbiol* 9: 165–176.

McKew, B.A., Coulon, F., Yakimov, M.M., Denaro, R., Genovese, M., Smith, C.J., et al. (2007b) Efficacy of intervention strategies for bioremediation of crude oil in marine systems and effects on indigenous hydrocarbonoclastic Bacteria. *Environ Microbiol* 9: 1562–1571.

Malmstrom, R.R., Kiene, R.P., Cottrell, M.T., and Kirchman, D.L. (2004) Contribution of SAR11 bacteria to dissolved dimethylsulfiniopropionate and amino acid uptake in the North Atlantic ocean. *Appl Environ Microbiol* 70: 4129–4135.

Melcher, R.J., Apitz, S.E., and Hemmingsen, B.B. (2002) Impact of irradiation and polycyclic aromatic hydrocarbon spiking on microbial populations in marine sediment for future aging and biodegradability studies. *Appl Environ Microbiol* 68: 2858–2868.

Menf, S., Chamkha, M., and Sayadi, S. (2009) Isolation and characterization of *Halomonas* sp. strain C2SS100, a hydrocarbon-degrading bacterium under hypersaline conditions. *J Appl Microbiol* 107: 785–794.

Morris, R.M., Rappé, M.S., Connon, S.A., Vergin, K.L., Siebold, W.A., Carlson, C.A., and Giovannoni, S.J. (2002) SAR11 clade dominates ocean surface bacterioplankton communities. *Nature* 420: 806–810.

Muyzer, G., De Waal, E.C., and Uitterlinden, A.G. (1993) Profiling of complex microbial populations by denaturing gradient gel electrophoresis analysis of polymerase chain reaction-amplified genes coding for 16S rRNA. *Appl Environ Microbiol* 59: 695–700.

Nelson, C.E., and Carlson, C.A. (2012) Tracking differential incorporation of dissolved organic carbon types among diverse lineages of Sargasso Sea bacterioplankton. *Environ Microbiol* 14: 1500–1516.

Nichols, C.M., Lardilère, S.G., Bowman, J.P., Nichols, P.D., A.E. Gibson, J., and Guézenec, J. (2005) Chemical characterization of exopolysaccharides from Antarctic marine bacteria. *Microb Ecol* 49: 578–589.

Nogales, B., Aguiló-Ferretjans, M.M., Martín-Cardona, C., Lalucat, J., and Bosch, R. (2007) Bacterial diversity, composition and dynamics in and around recreational coastal areas. *Environ Microbiol* 9: 1913–1929.
Pepi, M., Cesaro, A., Liut, G., and Baldi, F. (2005) An antarctic psychrotrophic bacterium Halomonas sp. ANT-3b, growing on n-hexadecane, produces a new emulsifying glycolipid. *FEMS Microbiol Ecol* 53: 157–166.

Prabagaran, S.R., Manorama, R., Delille, D., and Shivaji, S. (2007) Predominance of Roseobacter, Sulfitobacter, Glaciecola and Psychrobacter in seawater collected off Ushuaia, Argentina, Sub-Antarctica. *FEMS Microbiol Ecol* 59: 342–355.

Redmond, M.C., and Valentine, D.L. (2011) Natural gas and temperature structured a microbial community response to the Deepwater Horizon oil spill. *Proc Natl Acad Sci USA* 109: 20292–20297.

Rivers, A.R., Sharma, S., Tringe, S.G., Martin, J., Joye, S.B., Redmond, M.C., and Valentine, D.L. (2011) Natural gas and temperature structured a microbial community response to the Deepwater Horizon oil spill. *Proc Natl Acad Sci USA* 109: 20292–20297.

Romanenko, L.A., Zhukova, N.V., Rohde, M., Lysenko, A.M., Røberg, S., Østerhus, J.I., and Landfald, B. (2011) Dynamics of bacterial community exposed to hydrocarbons and oleophilic fertilizer in high-Arctic intertidal beach. *Polar Biol* 34: 1455–1465.

Sabirova, J.S., Chernikova, T.N., Timmis, K.N., and Golyshin, P.N. (2008) Niche-specificity factors of a marine oil-degrading bacterium *Alcanivorax borkumensis* SK2. *FEMS Microbiol Lett* 285: 89–96.

Saravanan, P., and Jayachandran, S. (2008) Preliminary characterization of exopolysaccharides produced by a marine biofilm-forming bacterium *Pseudoalteromonas rutenica* (SBT 033). *Lett Appl Microbiol* 46: 1–6.

Schneiker, S., Martins Dos Santos, V.A., Bartels, D., Bekel, T., Brecht, M., Buhrmester, J., et al. (2006) Genome sequence of the ubiquitous hydrocarbon-degrading marine bacterium *Alcanivorax borkumensis*. *Nat Biotechnol* 24: 997–1004.

Sikkema, J., De Bont, J.A., and Poolman, B. (1995) Mechanisms of membrane toxicity of hydrocarbons. *Microbiol Rev* 59: 201–222.

Sperling, M., Giebel, H.A., Rink, B., Grayek, S., Stanev, J., Stanev, E., and Simon, M. (2012) Differential effect of hydrographic and biogeochemical properties on SAR11 and *Roseobacter* RCA populations in the southern North Sea. *Aquat Microb Ecol* 67: 25–34.

Staley, J.T. (2010) *Cyclolasticus*: A genus of marine polycyclic aromatic hydrocarbon degrading bacteria. In *Handbook of Hydrocarbon and Lipid Microbiology*. Timmis, K.N., McGinley, T.J., van der Meer, J.R., and de Lorenzo, V. (eds). Berlin Heidelberg: Springer, pp. 1782–1786.

Steindler, L., Schwabach, M.S., Smith, D.P., Chan, F., and Giovannoni, S.J. (2011) Energy starved *Candidatus Pelagibacter* ubique substitutes light-mediated ATP production for endogenous carbon respiration. *PLoS ONE* 6: e19725.

Tada, Y., Taniguchi, A., Sato-Takabe, Y., and Hamasaki, K. (2012) Growth and succession patterns of major phylogenetic groups of marine bacteria during a mesocosm diatom bloom. *J Oceanogr* 68: 509–519.

Teramoto, M., Suzuki, M., Okazaki, F., Hatamati, A., and Harayama, S. (2009) *Oceano bacter*-related bacteria are important for the degradation of petroleum aliphatic hydrocarbons in the tropical marine environment. *Microbiology* 155: 3362–3370.

Teramoto, M., Ohuchi, M., Hatamati, A., Darmayati, Y., Widyastuti, Y., Harayama, S., and Fukunaga, Y. (2011) *Oleibacter marinus* gen. nov., sp. nov., a bacterium that degrades petroleum aliphatic hydrocarbons in a tropical marine Environment. *Int J Syst Evol Microbiol* 61: 375–380.

Vila, J., Maria Nieto, J., Mertens, J., Springael, D., and Grifoll, M. (2010) Microbial community structure of a heavy fuel oil-degrading marine consortium: linking microbial dynamics with polycyclic aromatic hydrocarbon utilization. *FEMS Microbiol Ecol* 73: 349–362.

Wang, B., Lai, Q., Cui, Z., Tan, T., and Shao, Z. (2008) A pyrene-degrading consortium from deep-sea sediment of the West Pacific and its key member *Cyclolasticus* sp. P1. *Environ Microbiol* 10: 1948–1963.

Wietz, M., Gram, L., Jørgensen, B., and Schramm, A. (2010) Latitudinal patterns in the abundance of major marine bacterioplankton groups. *Aquat Microb Ecol* 61: 179–189.

Wurl, O., and Obbard, J.P. (2004) A review of pollutants in the sea-surface microlayer (SML): a unique habitat for marine organisms. *Mar Pollut Bull* 48: 1016–1030.

Yakimov, M.M., Golyshin, P.N., Lang, S., Moore, E.R., Abraham, W.R., Lunsdorf, H., and Timmis, K.N. (1998) *Alcanivorax borkumensis* gen. nov., sp. nov., a new, hydrocarbon-degrading and surfactant-producing marine bacterium. *Int J Syst Bacteriol* 48: 339–348.

Yakimov, M.M., Giuliani, L., Denaro, R., Crisafi, E., Chemikova, T.N., Abraham, W.R., *et al.* (2004a) *Thalassolituus oleivorans* gen. nov., sp. nov., a novel marine bacterium that obligately utilizes hydrocarbons. *Int J Syst Evol Microbiol* 54: 141–148.

Yakimov, M.M., Gentile, G., Bruni, V., Cappello, S., D’Auria, G., Golyshin, P.N., and Giuliani, L. (2004b) Crude oil-induced structural shift of coastal bacterial communities of rod bay (Terra Nova Bay, Ross Sea, Antarctica) and characterization of cultured cold-adapted hydrocarbonoclastic bacteria. *FEMS Microbiol Ecol* 49: 419–432.

Yong, J.J., Park, S.J., Kim, H.J., and Rhee, S.K. (2007) *Glaciecola agarilytica* sp. nov., an agar-digesting marine bacterium from the East Sea, Korea. *Int J Syst Evol Microbiol* 57: 951–953.

### Supporting information

Additional Supporting Information may be found in the online version of this article at the publisher’s web-site:

*Fig. S1.* DGGE profiles of bacterial communities in the non-oiled (BI) and the most oiled (BIII) on-board mesocosms over time, based on 16S rRNA gene.

**Appendix S1.** Supporting experimental procedures.