Introduction

Maize heat shock protein of 101 kDa (HSP101) is essential for thermotolerance induction in this plant. The mRNA encoding this protein harbors an IRES element in the 5’ UTR that mediates cap-independent translation initiation. In the current work it is demonstrated that hsp101 IRES comprises the entire 5’UTR sequence (150 nts), since deletion of 17 nucleotides from the 5’ end decreased translation efficiency by 87% compared to the control sequence. RNA structure analysis of maize hsp101 IRES revealed the presence of three stem-loops toward its 5’ end, whereas the remainder sequence contains a great proportion of unpaired nucleotides. Furthermore, HSP90 protein was identified by mass spectrometry as the protein preferentially associated with the maize hsp101 IRES. In addition, it has been found that eIFiso4G rather than eIF4G initiation factor mediates translation of the maize hsp101 mRNA.
and structural analysis of maize hsp101 IRES. These studies identified critical stem-loops needed for IRES activity. In addition, mass spectrometry identification of factors bound to hsp101 IRES element revealed HSP90 as a factor specifically associated to the IRES competent complex under cap-independent conditions. Finally, in vitro translation of hsp101 mRNA showed selectivity for eIFiso4G rather than eIF4G translation initiation factor.

Results

Functional analysis of the maize hsp101 IRES

To identify the minimal region of the maize hsp101 IRES driving internal initiation of translation, 5’ end deletions of the IRES sequence inserted in pBIC-5 construct were generated. The pBIC-5 construct, containing an active maize hsp101 IRES [2], harbors 60 nts belonging to the genomic region upstream of the transcription start site (+1) (Figure 1a). Thus, to verify whether these extra 60 nts alters the hsp101 IRES activity, a construct named 101+1 (starting at the +1 transcription start site) was generated. In addition, 5’ end deletions of 77 and 110 nts were introduced in constructs 101Δ17 and 101Δ50, respectively (Figure 1a). These sequences were inserted in the pBIC bicistronic vector in which translation of the first reporter gene (CAT) is cap-dependent while translation of the second reporter gene (LUC) is cap-independent (Figure 1b). pBIC-5, 101+1, 101Δ17 and 101Δ50 RNAs synthesized in vitro were translated using the rabbit reticulocyte lysate (RRL) translation system in a cap-dependent manner. In parallel, translation of these RNAs was analyzed in lysates expressing the Lb protease of FMDV, as previously done with the pBIC-5 IRES sequence [2]. This protease cleaves eIF4G [26], rendering the cell-free translation system cap-independent. In the presence of Lb protease, LUC protein was synthesized from RNAs pBIC-5 and, particularly, 101+1 (insert in Figure 1c). In contrast, very low levels of luciferase were detected in lanes loaded with reaction products driven by 101Δ17 RNA. The fold-increase of LUC protein synthesis was plotted to compare translation initiation efficiency among the different IRES constructs. To this end, the intensity of LUC polypeptide was normalized to the value obtained for pBIC-5 RNA (the construct with proven IRES activity) set to 100% (Figure 1c). Translation efficiency of luciferase from mRNA 101+1 was threefold higher than the control RNA pBIC-5 in the presence of Lb. In contrast, translation efficiency of mRNAs 101Δ17 and 101Δ50 decreased severely (83% and 90%, respectively) in comparison to the control pBIC-5 (Figure 1c). These results demonstrated that the complete 5’UTR sequence of hsp101 RNA was required for IRES-dependent translation initiation under conditions that compromise cap-dependent initiation, here illustrated by Lb protease coexpression.

Given that the sequence in 101+1 RNA was identified as the authentic maize hsp101 IRES functional region, we analyzed the secondary structure of this RNA in order to identify potential RNA motifs mediating internal initiation, likely shared with other IRES elements.

Secondary structure of maize IRES hsp101

To characterize the secondary structure in solution of maize hsp101 functional IRES, 101+1 RNA was treated with N-methylisatoic anhydride (NMIA), which reacts with the single-stranded 2’-hydroxyl groups of nucleotides to form a stable 2’-O-aduct [27]. This modification was identified by primer extension reaction as reverse transcriptase (RT) stop sites [28] (Figure 2a). The intensity of RT stops were quantified by densitometry and then normalized to the intensity of the full-length product obtained in each RT reaction set to 100%. Four ranges of reactivity were chosen, colored black, blue, yellow and red (0, >20, >50 and < 50, respectively) in Figure 2b. Secondary structure of the functional maize hsp101 IRES was obtained by using the values for NMIA reactivity in the mFOLD software; nucleotides are presented in a color scale reflecting their reactivity to NMIA (Figure 2c). The RNA structure revealed a large proportion of nucleotides that are reactive to NMIA; thereby unpaired, with the exception of 3 stem-loops located at the 5’ end of the RNA. The first hairpin comprises nucleotides 9–21, thus including most of the 17 nucleotides whose deletion leads to a loss of IRES activity. Together, these results demonstrate the importance of this hairpin for internal translation initiation. Furthermore, consistent with the defective phenotype of the deletion of the first hairpin, a construct lacking 50 nucleotides at the 5’ end eliminating the 3 hairpins also yielded fully defective IRES elements.

Isolation of RRL proteins bound to IRES under cap-independent conditions

Once the secondary structure of the functional region of maize hsp101 IRES was elucidated, we set up to identify candidate proteins potentially interacting with this element. For this, poly(A)-tailed RNAs were generated to capture the proteins that bind to maize hsp101 IRES taking advantage of the poly(A) affinity to oligo dT cellulose. In addition, untreated and Lb protease-treated lysates were used to favor cap-dependent or cap-independent conditions, respectively. No differences were observed in the pattern of proteins purified with the RNAs tested using untreated lysates (Figure 3a). In contrast, two bands of ~50 kDa and ~75 kDa were specifically associated to 101+1 sense RNA using the Lb-treated RRL (red arrows in Figure 3b). Notably, these bands were absent in samples pulled down with the antisense 101+1as RNA. These results indicate that the 75 and 50 kDa factors specifically interact with the authentic hsp101+1 RNA. Lack of binding of these factors to the RNAs derived from pGEM-T 101+1, 101Δ17 and 101Δ50 could be explained at least in part by differences in the RNA structure induced by 30 nt polylinker sequence located at their 3’ end that preclude their interaction. Mass spectrometry (LS/MS/MS) identification of these bands revealed that the ~75 kDa band was the molecular chaperone HSP90 (Figure 3c), as demonstrated by the presence of 4 unique peptides (yellow marks in Figure 3c). The second protein could not be unambiguously identified. We conclude that HSP90 protein forms part of the maize hsp101 IRES translatable complex assembled under cap-independent conditions.

Functional relevance of HSP90, eIF4G and eIFiso4G in maize hsp101 translation

To confirm that the HSP90 protein was involved in maize hsp101 IRES-driven translation, we conducted cap-independent in vitro translation assays. Additionally, two potential inhibitory reagents were added to the translation system. One of them, radicicol is an HSP90 specific ligand [29]. The second one was a specific antibody HSP90 (anti-HSP90 Ab). Under cap-independent conditions both, Radicicol and anti-HSP90 Ab inhibited translation of the 101+1 mRNA was (Figure 4). These data revealed the importance of HSP90 for maize hsp101 mRNA translation. However, addition of commercial purified human HSP90 protein to the system did not affect translation efficiency of 101+1 (Figure 4), suggesting that the level of HSP90 protein present in the lysate is sufficient for IRES activity.

Plants have two isoforms of eIF4G, being eIFiso4G preferentially used for translation initiation of mRNAs with 5’UTR stable
secondary structure, with hypermethylated cap structures or actively involved in IRES-dependent translation [30,31]. Thus, since many viral and cellular IRES elements depend on eIF4G factor, we investigated the role of eIF4G and eIFiso4G factors for maize hsp101 translation. For this purpose, in vitro translation assays were performed using the wheat germ system (WGE) depleted from either eIF4F or eIFiso4F (eIF4E-eIF4G and eIFiso4E-eIFiso4G, respectively) using m7GTP-sepharose. Then, addition of wheat recombinant eIF4G or eIFiso4G to the translation system was conducted to determine whether any of these factors affected hsp101 IRES-directed translation. No significant difference in hsp101 translation efficiency was observed when eIF4G was added to the system compared to the depleted lysate. However, translational efficiency increased by 40% when eIFiso4G was added to the translation system compared to the control. Furthermore, hsp101 translation efficiency increased by 60% when both factors were added to the system (Figure 5). These results suggest that there is selectivity on the use of factors used to initiate translation, and that maize hsp101 IRES has preference for eIFiso4G over eIF4G to form the active translational complex.

Discussion

Eukaryotic mRNA translation initiation depends on the cap structure located at the 5’end [3]. However, under conditions that inhibit cap-dependent translation, activation of IRES-dependent translation facilitates cell survival. IRES elements, usually located at the 5’ UTR of some mRNAs of eukaryotic cells and their viruses, do not present extensive similarities in their primary sequence or RNA structure. Therefore, definition of the boundaries of IRES elements as well as their structure has to be experimentally delimited case by case. This can be accomplished by functional analysis of the candidate sequences, using cap-independent translation conditions [11]. Earlier studies have shown that some IRES elements are active in heterologous systems.

![Maize Hsp101 IRES Element](image_url)
In agreement with this, the maize hsp101 mRNA contains an IRES element which is active under eIF4G cleavage induced by the Lb protease activity in RRL [2]. In the current study, and using the same system, it is shown that deletions or insertions at the 5′ end of the authentic hsp101 5′UTR affect the translation efficiency. Thus, the entire 5′UTR sequence of the heat shock proteins message (about 150 nts long) is required for efficient internal initiation of translation. Specifically, we have found that three stem-loops within the first 50 nucleotides are essential for IRES-dependent translation initiation (Figure 1). Similar results were reported for the hsp70 IRES element of Drosophila melanogaster and Homo sapiens. In both cases, the complete sequence is required for cap-independent translation, since short deletions of the 5′UTR reduced IRES activity by 75% to 95% [35,36].

The rich diversity of IRES structures in conjunction with their response to different conditions suggests the existence of multiple mechanisms driving IRES function [13,17]. Therefore, it is very important to elucidate their RNA structure. Here we have used nucleotide modification by NMIA to analyze the secondary structure of the hsp101 IRES, a methodology that reports local flexibility of RNA. The maize hsp101 IRES secondary structure is the first experimental IRES structure reported for plant heat shock mRNAs (Figure 2). Structural domains identified in the maize hsp101 IRES include three stem-loops SL1 (9–21 nts), SL2 (27–34 nts) and SL3 (43–61 nts). This is followed by two large loops L1 (62–84 nts) and L2 (107–121 nts) flanked by 2 short stems, S1 and S2 (87–103 nts). Finally, a single-stranded region (126 to 150 nts) is located upstream of the AUG start codon (Figure 2). This RNA structure differs from other IRES elements [37–39]. In support of the biological relevance of this region, it is worth mentioning that the nucleotide sequence of this mRNA region is highly conserved among different Zea mays sequences deposited in databases, despite being outside of the coding region.

The relevance of RNA structure for IRES function is supported by the fact that deletions of the stem-loop SL1 in construct 101Δ17 abrogated translation. Likewise, the construct lacking the three stem-loops at the 5′end also failed to drive protein synthesis. This

**Figure 2. SHAPE structural analysis of maize hsp101 IRES.** a) Primer extension analysis of 101+1 RNA. The first four lanes (UACG) show hsp101+1 sequence, obtained with the 5′ end labeled primer. Lanes 5 and 6 show the primer extension products obtained with 101+1 RNA untreated (−) or treated (+) with NMIA, respectively. b) SHAPE reactivity of 101+1 RNA. Reactivity was normalized relative to the full-length product (set to 100%). Blue bars represent values <20; yellow bars represent values <50 and red bars represent values >50. c) Maize hsp101 IRES secondary structure. Colors of the nucleotides reflect SHAPE reactivity values.

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result suggests that a defective structure lacking SL1 is unable to interact with a trans-acting element. Conversely, the large single-stranded region presumably provides the binding site of proteins necessary to form the active translational complex under heat stress conditions, including HSP90 and the product of hsp101 message. Along this idea, recent data have shown that mRNAs associated with stress responses tend to have more single-stranded nucleotides at the 5'UTR, longer maximal loop length and higher free energy [40], as found in the RNA structure of the hsp101 IRES reported in the current study. On the other hand, it has been also noted that comparison of RNA structures of mRNA activated under stress response were poorly predicted in silico, emphasizing the need to determine the RNA structure biochemically.

Our results showed that the length and structure of the IRES is important for its function. However, IRES-binding proteins are also essential. Our experimental approach identified two proteins (50 kDa and 75 kDa) that were coprecipitated with the IRES element. The 75 kDa protein was identified as the molecular chaperone HSP90, a protein belonging to a family of ancient conserved factors found in all organisms including plants and animals [41], which has been reported to form stress granules and P-bodies under stress conditions [42]. This protein shows a large degree of amino acid conservation between animals and plants [43]. The active role of HSP90 to form the translational complex and initiate cap-independent translation was demonstrated by absence of translation of hsp101+1 in the presence of radicicol or the HSP90 antibody (Figure 4). HSP90 has also been reported to participate in the formation of the translational complex of the hepatitis C virus (HCV) IRES [44]; in this case, HSP90 interacts with the eIF3c subunit, preventing its ubiquitination and thus, impairing its degradation by the proteasome pathway.

Previous evidences has shown that eIF4G initiation factor is required for cap-independent translation driven by various IRES elements in the presence of the Lb protease (cap-independent).

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Figure 3. Identification of proteins associated to maize hsp101 IRES. Purification of IRES-protein complexes using RRL translation system (a), or Lb-treated RRL translation system (b) with RNAs indicated at the top. Equal amounts of samples containing the proteins that coprecipitated with the indicated RNAs were loaded on SDS-PAGE and detected by silver staining. Red arrows depict two bands specifically copurifying with 101+1 sense RNA using the Lb-treated RRL. c) Sequence of the 75 kDa protein that binds to maize hsp101 IRES. The 75 kDa band (indicated by an arrow in lane 2 of panel b) was sequenced and identified as the molecular chaperone HSP90. The results yielded 4 unique peptides (shaded in yellow), 4 unique spectra, 5 total spectra, 54/565 amino acids (10% coverage).

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Figure 4. Effect of HSP90 protein on hsp101+1 translation initiation. Equal amounts of in vitro synthesized RNAs 101+1 were used to program translation using RRL in the presence of Radicicol (lane 3), anti-HSP90 (lane 4), HSP90 protein (lane 5), or DMSO (lane 6). Proteins were resolved by SDS–PAGE and visualized by autoradiography of the dry gel. Synthesis of luciferase driven by the indicated IRES elements in the presence of the Lb protease (cap-independent).

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Maize Hsp101 IRES Element

Figure 5. Hsp101+1 translation in a wheat system (WGE) depleted from elf4E, elfiso4E, elf4G and elfiso4G. a) Maize hsp101 translation in the complete WGE system. Lane 1: background signal obtained without added RNA (negative control); lane 2: hsp101+1-driven translation adding recombinant elf4G factor (4); hsp101+1 translation adding recombinant elfiso4G factor (5); hsp101+1 translation adding both elf4G and elfiso4G (6). b) Histogram of cap-independent translation efficiency of RNA 101+1 when elf4G and elfiso4G are added to the system. Values represent the mean (±SD) of three independent assays.

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elements [13]. In the case of maize hsp101 IRES activity, elfiso4G was more active than elf4G. This result might be explained by the fact that the factor generally used for plant mRNA translation is elfiso4G [30,45] suggesting that this characteristic also prevails for IRES from plant mRNAs. This is in accordance with the fact that hsp101 IRES is specific to maize and thus, conserves a preference for factors from the same origin. Some cellular and viral IRES elements also require elf4G factor [46–48] for translation initiation. However, there are no sequence similarities with the maize hsp101 IRES sequence, suggesting that the interaction can be based on a structural RNA domain, possibly helped by the interaction with other interacting factors.

Our results raised the possibility that SL1 could provide the binding-site for proteins such as HSP90 or elfiso4G, a possibility that needs to be tested in the future. Interestingly, the results obtained in the current work suggest that transfer of the IRES hsp101 of maize to other plant mRNAs could confer translational advantage under heat stress and thus, alleviate problems of thermo-tolerance in plants.

In summary, this work reports the first experimental secondary structure of the IRES of a cellular mRNA of plants [hsp101], the participation of HSP90 in the maize hsp101 complex and finally, the preference for elfiso4G for translation initiation under cap-independent conditions.

Material and Methods

Constructs

The IRES sequence amplification reactions were performed as previously reported [2] using the PXE101 plasmid with the hsp101 cDNA as template including the 5’UTR region (Figure 1a). Recognition sequence for the restriction enzyme SacI was added to the primers used in the amplification reactions. Construct hsp101+1 was generated with the pair of primers forward hsp101+1 5’-CGAGCTCGACAACTTTCAACCAGA-3’/reverse RV101* 5’-AGCAAGCCATGAAATCCGAGACACACTCTC-GACG-3’. For constructs hsp101Δ17 and hsp101Δ50, the pair of primers forward hsp101-A 5’-CGAGCTCGAAACACTAGCC- GAAGCACC-3’/reverse RV101* and forward hsp 101-B 5’-CGAGCTCGACACTTGGTGGATATCATCTC-3’/reverse RV101* respectively, were used. PCR products were inserted into the pGEM-T EASY cloning vector to generate pGEM-101+1, pGEM-101Δ17 and pGEM-101Δ50, respectively. The fragments were then excised with SacI to replace the IRES sequence in pBIC-5 generating pBIC-hsp101+1, pBIC-hsp101Δ17 and pBIC-hsp101Δ50 constructs. RNAs derived from pGEM-T constructs were synthesized using T7 RNA polymerase with PstI linearized plasmids. Bicistronic RNAs pBIC-5, 101+1, 101Δ17 and 101Δ50 were synthesized in vitro using HpaI linearized plasmids and T7 RNA polymerase [2].

In vitro translation

Cap-dependent and cap-independent translation was examined either in rabbit reticulocyte lysate (RRL) or Wheat Germ Extract (WGE). For cap-dependent translation analysis, a mix of RRL (17.5 μL), RNasin (1 μL), amino acids mix without methionine (1 μL), the bicistronic RNA (100 ng) and [35S] methionine (1 μL, 10 μCi) was prepared in a final volume of 35 μL with DEPC-treated water. The mix was incubated 60 min at 30°C; at the end of the reaction, 1 μL of each sample was obtained and the synthesized proteins were resolved by SDS-PAGE. Gels were dried and exposed to a phosphorscreen to be analyzed by the QUANTITY ONE software (BIORAD). Values represent the mean (±SD) of three independent assays.

To generate the cap-independent RRL translation system, 50 ng of mRNA encoding the Lb protease of FMDV, a protein that cleaves elf4G preventing cap-dependent translation [14], was added to the RRL mix and incubated at 30°C for 15 minutes. Subsequently, 100 ng of each bicistronic mRNA were added to the mix and incubated for another 45 minutes prior to analyze the translation products as indicated above. When indicated, the HSP90 inhibitors, Radicicol (SIGMA) and HSP90 antibody (Santa Cruz), or the commercially purified HSP90 protein (100 ng) (BIOMART) were added to the hsp101 IRES-driven translation in separate reactions.

Similarly, WGE was used either complete (cap-dependent) or elf4E and elfiso4E-depleted (cap-independent). For cap-independent translation, elf4E and elfiso4E were depleted from WGE using m’GTP sepharose [49]. To this end, 150 μL of m’GTP sepharose were centrifuged at 1500 rpm for 1 min to eliminate the supernatant and the resin was washed twice with 2
volumes of m 7 GTP buffer (100 mM KCl, 20 mM HEPES, pH 7.6, 0.2 mM EDTA, 10% glycerol, 0.05 mM PMSF, 0.5 mM DTT). Then, 50 µl of WGE were added to the resin and the mix was incubated for 30 minutes at 4°C with gentle shaking. Finally, the sample was centrifuged at 1500 rpm for 1 min at 4°C, the supernatant was recovered and used as the WGE-depleted lysate. The translation reaction mix contained 12.5 µl of WGE depleted, 1 µl of amino acids mix without methionine, 100 ng of hsp101+1 RNA, 1 µl [35S] methionine (10 µCi), 100 ng of eIF4G or eIFiso4G and DEPC-treated water to reach a volume of 25 µl. The mix was incubated for 60 min at 25°C. Newly synthesized proteins were resolved by SDS-PAGE loading 1 µl of each sample. Gels were dried and exposed to a phosphorimager screen.

**Determination of the maize hsp101 IRES secondary structure by Selective 2′-Hydroxyl Acylation Primer Extension (SHAPE)**

In order to determine the IRES secondary structure, hsp101+1 RNA was synthesized with T7 RNA polymerase using pBIC-hsp101+1 plasmid linearized with BbsI [50]. SHAPE reactivity assays were carried out as described [27] with some modifications [26]. Briefly, hsp101+1 RNA (2 pmol) were resuspended in 12 µl 0.5X TE buffer, heated for 2 min at 95°C and then cooled for 2 min at 4°C. 6 µl of 3.5-RNA folding mix (333 mM HEPES, pH 8.0, 20 mM MgCl₂, 333 mM NaCl) were added and incubated for 20 min at 37°C. This mix was divided into two tubes (9 µl in each tube); 1.5 µl NMI (65 mM) [27] was added to one of the tubes (+) and 1.5 µl DMSO to the other (−). Either reaction was incubated for 45 min at 37°C. At the end of the incubation period, 90 µl H₂O, 4 µl 5 M NaCl, 1 µl glycerin (20 mg/ml), 2 µl 100 mM EDTA, pH 8 and 350 µl absolute ethanol were added to each reaction tube. Samples were precipitated and the pellet RNA was resuspended in 20 µl of 0.5X TE. Ten microliters of each RNA sample, NMI (+) and (−), were transferred to a 200 µl tube; 0.5 µl of the [35S]-labeled oligonucleotide (GGGCTTCCTTTAATGGTTTTGCG) using polynucleotid kinase (PRMEGA) were added and incubated first, for 5 min at 65°C, then for 5 min at 35°C and finally at 4°C for 1 min. Three microliters of reverse transcriptase (RT) Superscript III buffer, 1U RNasin, 0.25 µl 100 mM DTT, 1.5 µl 10 mM dNTPs and 0.25 µl H₂O were added to each sample and further incubated for 1 min at 52°C. Then, 0.25 µl Superscript III RT (INVITROGENE) was added and the mix and incubated for 30 min at 52°C. At the end of the reaction, 0.4 µl of 4 M NaOH were added to each sample. Samples were precipitated by adding 1 volume of AcNH₄, 1 µl glycerin and 75 µl absolute ethanol. Pellets were resuspended in 10 µl H₂O plus 4 µl formamide buffer (90% formamide). Primer extension products were resolved on denaturing 6% polyacrylamide 7 M urea gels, in parallel to a sequence ladder prepared with the same 5′ end-labeled primer. Gels were dried and exposed to a radiography film for quantification. SHAPE reactivity data was analyzed as described [50].

**Purification of IRES-associated proteins**

In order to identify the proteins interacting with hsp101 IRES (either sense hsp101+1 or antisense orientation inserted in the bicistronic constructs), 101+1, 101Δ17 and 101Δ50 RNAVs (derived from pGEM-T EASY), were synthesized adding a poly-A tail using poly-A polymerase (AMBION) as described by the manufacturer. From each RNA sample, 2.5 µg were transferred to 100 µl of binding buffer (10 mM Tris, pH 7.5, 100 mM KCl, 2 mM MgCl₂) containing 30 µl oligo dT sepharose. Samples were incubated for 30 min at 4°C with gentle shaking and then washed twice with binding buffer to remove unbound RNA. Then, 100 µg of RRL and 10 µg total RNA (nonspecific competitor) were added to a final volume of 100 µl binding buffer and incubated for 60 min at 4°C with gentle shaking. Samples were washed twice with binding buffer followed by washing buffer (10 mM Tris, pH 7.5, 100 mM KCl, 0.5 mM MgCl₂) to remove unbound proteins to the IRES-oligo dT sepharose complex. IRES-protein complexes were resolved by SDS-PAGE and visualized by silver staining in triplicate experiments. Thin slices containing differential IRES-bound proteins were excised and identified by mass spectrometry (LC/MS/MS) at Quebec Genomics Center.

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**Author Contributions**

Conceived and designed the experiments: ESdJ ASJG. Performed the experiments: ASJG NF. Analyzed the data: ASJG NF EMS ESdJ. Contributed to the writing of the manuscript: ESdJ EMS.

**References**

1. Nieto-Seto´le J, Martínez LM, Pance G, Cassab GI, Alagon A, et al. (2002) Maize HSP101 plays important roles in both induced and basal thermotolerance and primary root growth. Plant Cell 14: 1621–1635.
2. Dinkova TD, Zepeda H, Martínez-Salas E, Martínez LM, Nieto-Seto´le J, et al. (2005) Cap-independent translation of maize Hsp101. Plant J 41: 722–731.
3. Soneberg N, Himemachi AG (2009) Regulation of translation initiation in eukaryotes: mechanisms and biological targets. Cell 136: 731–745.
4. Komar AA, Hatzoglou M (2011) Cellular IRES-mediated translation: the war of ITAFs in pathophysiological states. Cell Cycle 10: 229–240.
5. Spriggs KA, Bushell M, Willis AE (2010) Translational regulation of gene expression during conditions of cell stress. Mol Cell 40: 228–237.
6. Coblhoid LC, Spriggs KA, Haines SJ, Doblyn HC, Hayes C, et al. (2008) Identification of internal ribosome entry site (IRES)-trans-acting factors for the Myx family of IRESs. Mol Cell Biol 28: 40–49.
7. Stoneley M, Willis AE (2004) Cellular internal ribosome entry segment structures, trans-acting factors and regulation of gene expression. Oncogene 23: 3200–3208.
8. Yanguess E, Castro-Sanz AB, Fernandez-Bautista N, Oliveros JC, Castaño-113 (2013) Analysis of genome-wide changes in the translationome of Arabidopsis seedlings subjected to heat stress. PloS One 8: e71425.
9. Baird SD, Turcotte M, Korneluk RG, Holcik M (2006) Searching for IRES RNA: 12: 1755–1785.
10. Martinez-Salas E (2008) The impact of RNA structure on picornavirus IRES activity. Trends Microbiol 16: 230–237.
19. Jaag HM, Kavchuk I, Rohde W, Fischer R, Emans N, et al. (2003) An unusual internal ribosomal entry site of inverted symmetry directs expression of a potato leafroll potyvirus replication-associated protein. Proc Nat Acad Sci USA 100: 8939–8944.

20. Koh DC, Wong SM, Liu DX (2003) Synergism of the 3′-untranslated region and an internal ribosome entry site differentially enhances the translation of a plant virus coat protein. J Biol Chem 278: 20565–20573.

21. Zrenko V, Galleh DR (2005) Cap-independent translation of tobacco etch virus is conferred by an RNA pseudoknot in the 5′-leader. J Biol Chem 280: 26813–26824.

22. Dorokhov YL, Ivanov PA, Komarova TV, Skulachev MV, Atabekov JG (2006) An internal ribosome entry site located upstream of the crucifer-infecting tobravirus coat protein (CP) gene can be used for CP synthesis in vivo. J Gen Virol 87: 2693–2697.

23. Karretnikov A, Lehto K (2007) The RNA2 5′-leader of Blackcurrant reversion virus mediates efficient in vivo translation through an internal ribosomal entry site mechanism. J Gen Virol 88: 286–297.

24. Fernandez-Miragall O, Hernandez C (2011) An Internal Ribosome Entry Site Directs Translation of the 3′-Gene from Polarogenium Flower Break Virus. Genes 2: 6: e22617.

25. Mardanova ES, Zanchuk LA, Skulachev MV, Ravin NV (2008) The 5′ untranslated region of the maize alcohol dehydrogenase gene contains an internal ribosome entry site. Gene 420: 11–16.

26. Kirchweyer R, Ziegler E, Llampheir BJ, Waters D, Labig HD, et al. (1994) Foot-and-mouth disease virus leader protease: purification of the L1s form and determination of its cleavage site on eIF-4 gamma. J Virol 68: 5677–5684.

27. Wilkinson KA, Merino EJ, Weeks KM (2006) Selective 2′-hydroxyl acylation analyzed by primer extension (SHAPE): quantitative RNA structure analysis at single nucleotide resolution. Nat Protoc 1: 1610–1616.

28. Fernandez N, Garcia-Saccarstan A, Ramajo J, Briones C, Martinez-Salas E (2011) Structural analysis provides insights into the modular organization of picornavirus IRES. Virology 409: 251–261.

29. Uehara Y (2003) Natural product origins of Hsp90 inhibitors. Current Cancer Drug Targets 3: 325–330.

30. Mayberry LK, Allen ML, Nitka KR, Campbell L, Murphy PA, et al. (2011) Directs Translation of the 3′-Gene from Pelargonium Flower Break Virus Genomic RNA: Implications for Infection. PLoS One 6: e22617.

31. Patrick RM, Browning KS (2012) The eIF4F and eIFiso4F Complexes of Plants: An Evolutionary Perspective. Comp Funct Gen 2012: 267814.

32. Woodlalay KE, Lazaridou K, Belsham GP, Carter MJ, Roberts LO (2001) The 3′ untranslated region of Rhopalosiphum padi virus contains an internal ribosome entry site which functions efficiently in mammalian, plant, and insect translation systems. J Virol 75: 10244–10249.

33. Urvin P, Yi L, Martin H, Atkinson H, Gillmarrt PM (2000) Functional characterization of the EMCV IRES in plants. Plant J 24: 583–589.

34. Wong SM, Koh DC, Liu D (2008) Identification of plant virus IRES. Meth Mol Biol 451: 123–133.

35. Rubtsova MP, Szovat DV, Dmitriev SE, Ivanov DS, Prassolov VS, et al. (2003) Distinctive properties of the 5′-untranslated region of human hsp70 mRNA. J Biol Chem 278: 22353–22356.

36. Hernandez G, Vazquez-Fanzola P, Sierra JM, Rivera-Pumar R (2004) Internal ribosome entry site drives cap-independent translation of reaper and heat shock protein 70 mRNAs in Drosophila embryos. RNA 10: 1783–1797.

37. Le Quesne JP, Stoneley M, Fraser GA, Willis AE (2001) Derivation of a structural model for the e-myc IRES. J Mol Biol 310: 111–126.

38. Dorot L, Lavelle G, Cloie P, Martinez-Salas E (2013) Using RNA inverse folding to identify IRES-like structural subdomains. RNA Biology 10: 1842–1852.

39. Xia X, Holcik M (2009) Strong eukaryotic IRESs have weak secondary structure. PLoS One 4: e1136.

40. Ding Y, Tang Y, Kwok CK, Zhang Y, Beilruga PC, et al. (2014) In vivo genome-wide profiling of RNA secondary structure reveals novel regulatory features. Nature 565: 696–700.

41. Chen B, Zhong D, Monteiro A (2006) Comparative genomics and evolution of the HSP90 family of genes across all kingdoms of organisms. BMC Genomics 7: 156.

42. Gupta RS (1995) Phylogenetic analysis of the 90 kDa heat shock family of protein sequences and an examination of the relationship among animals, plants, and fungi species. Mol Biol Evol 12: 1063–1073.

43. Matsumoto K, Minami M, Shimosaki F, Suzuki Y, Ake K, et al. (2011) Hsp90 is involved in the formation of P-bodies and stress granules. Biochem Biophys Res Commun 407: 720–724.

44. Ujino S, Nishiharu H, Sugiyama R, Suzuki H, Hisaki T, et al. (2012) The interaction between human initiation factor eIF3 subunit c and heat-shock protein 90: a necessary factor for translation mediated by the hepatitis C virus internal ribosome entry site. Virus Res 163: 390–395.

45. Lellis AD, Allen ML, Aertker AW, Tran JK, Heilis DI, et al. (2010) Deletion of the eIFiso4G subunit of the Arabidopsis eIFiso4F translation initiation complex impairs health and viability. Plant Mol Biol 74: 249–263.

46. Kopolauva VG, Potova TV, Heleni CU, Shatuyk IN (1996) Translation eukaryotic initiation factor 4G recognizes a specific structural element within the internal ribosome entry site of encephalomyocarditis virus RNA. J Biol Chem 271: 18599–18604.

47. Lopez de Quinto S, Martinez-Salas E (2000) Interaction of the eIF4G initiation factor with the aphylovirus IRES is essential for internal translation initiation in vivo. RNA 6: 1390–1392.

48. Spriggs KA, Cobbold LG, Jopling CL, Cooper RE, Wilson LA, et al. (2009) Canonical initiation factor requirements of the Myc family of internal ribosome entry segments. Mol Cell Biol 29: 1565–1574.

49. Galleh DR, Browning KS (2001) eIF4G functionally differs from eIFiso4G in vivo. RNA 6: 1380–1392.

50. Fernandez N, Fernandez-Miragall O, Ramajo J, Garcia-Saccarstan A, Bellora N, et al. (2011) Structural basis for the biological relevance of the invariant apical stem in IRES-mediated translation. Nucleic Acids Res 39: 8572–8583.