Variants in PRKCE and KLC1, Potential Negative Regulators of Type I Psoriasis

Jianxiao Xing
Taiyuan central hospital of shanxi medical university

Ying Wang
Taiyuan central hospital of shanxi medical university

Xincheng Zhao
Taiyuan central hospital of shanxi medical university

Junqin Li
Taiyuan central hospital of shanxi medical university

Ruixia Hou
Taiyuan central hospital of shanxi medical university

Xuping Niu
Taiyuan central university of shanxi medical university

Guohua Yin
Taiyuan central hospital of shanxi medical university

Xinhua Li
Taiyuan central hospital of shanxi medical university

kaiming zhang (✉ zhangkaiming@sina.com)
Taiyuan central hospital of shanxi medical university

DOI: https://doi.org/10.21203/rs.3.rs-147917/v1

License: ©  This work is licensed under a Creative Commons Attribution 4.0 International License. Read Full License
Abstract

Background: Psoriasis is a multifactorial disease with a complex genetic predisposition. The pathophysiology of psoriasis is associated with genetic variants, especially in negative regulatory genes. To better characterize gene variants in psoriasis and identify the relationship between clinical characteristics and variant genes in its pathogenesis. DNA was extracted from 282 type psoriasis patients and purified, and 13 variable genes were amplified and sequenced using the Sanger method.

Results: Among the 13 investigated genes, the variants frequencies of protein kinase C epsilon (PRKCE) (c.240T>C, 35.9% vs 47.7%, P< .05) and kinesin light chain 1 (KLC1) (c.216A>G, 2.9% vs 98.1%, P< .01) were significantly lower in patients than in normal Asian individuals. Additionally, we found considerable differences in the relationship between variants in genes CADM2, JPH2, SPTLC3 and clinical characteristics stratified by medical history and family history. Moreover, MEGF6 gene variants (39.52% vs. 22.50%, χ²=3.83, p < .05) showed a stronger association with the group as mild (PASI≤10) than with the group as heavy.

Conclusions: Our results provide a comprehensive correlation analysis of negative regulatory genes that are regulated in psoriasis. This integrated analysis offers novel insight into the pathogenic mechanisms involved in psoriasis.

Background

Psoriasis is a common, genetic, chronic inflammatory skin disease that causes red, flaky patches on the skin [1]. The worldwide prevalence of psoriasis is approximately 3.2%, or more than 7.4 million people, as estimated by Rachakonda et al [2]. Psoriasis has a high recurrence rate and is considered a complex disease due to environmental and genetic factors [3].

The genetic component is partly explained by its association with certain susceptibility loci [4]. At present, more than 70 susceptibility genes are associated with psoriasis [5–6]. The strongest susceptibility locus lies within the major histocompatibility complex (MHC) [7]. Although most studies have focused on psoriasis susceptibility genes, others have shown that certain genetic variants play a negative regulatory role in the pathogenesis of psoriasis. There was also article found the association of TRAF3IP2-rs33980500 variant with the susceptibility to psoriasis [8].

Because monozygotic twins carry the theoretically same genetic information, they are more conducive to the study of susceptibility genes than sporadic populations. Recently, we obtained the whole genome sequences of 9 pairs of monozygotic twins with psoriasis discordance. In addition to the C3 de novo mutation[9], we reported 13 loci that were significantly different between normal individuals and patients with psoriasis in this paper. Interestingly, the 12 of 13 loci found in the normal individuals whose twin's medical history are longer than 10 years, indicating the 12 loci play a negative regulatory role in the pathogenesis of psoriasis, especially in long medical history. According to the onset age, vulgaris psoriasis were classified as early onset, vulgaris psoriasis were classified as type (late onset, onset age ≥ 40)[10]. Many scholars suggested a genetic influence upon its age at onset, especially in the type psoriasis[11–12]. To further investigate the contribution of these variants, we additionally examined these gene variants in 282 type psoriatic patients and analyzed the relation between variants and clinical manifestation based on the monozygotic twins investigation.

Results

13 variant loci were filtered in the monozygotic twins

In this study, blood sample from 9 pairs of monozygotic twins with psoriasis discordance was analyzed by whole genome sequencing. We obtained 13 variants loci, such as kinesin light chain 1 (KLC1, c.216A > G), protein kinase C epsilon (PRKCEc.240T > C), Multiple epidermal growth factor-like domains 6 (MEGF6, c.96 A > G), Cell adhesion molecule 2 (CADM2, c.263 A > G), Multiple epidermal growth factor-like domains 11 (MEGF11, c.80 T > G), Neural cell adhesion molecule 2 (NCAM2, c.236 G > T), Dipeptidyl peptidase like 6 (DPP6, c.174 G > T), Myosin heavy chain 14 (MYH14, c.461 T > C), Serine palmitoyltransferase long chain base subunit 3 (SPTLC3, c.481 T > C), FA complementation group C (FANCC, c.134 C > T), DENN domain containing 5B (DENND5B, c.96 A > G) Junctophilin 2 (JPH2, c.195 T > G) and RNA polymerase II associated protein 3 (RPAP3, c.146 C > G) (Table 1). Interestingly, all of the 13 above mentioned variant loci were found only in the normal homozygote, but none of them in psoriatic homozygote. Variant exist in normal populations, indicating their potential prevention to psoriasis.
Table 1
Thirteen mutant loci in 9 pairs of monozygotic twins

| Chromosome | SNP  | variants loc | Gene      |
|------------|------|--------------|-----------|
| Chr1       | rs2821008 | c.96 A > G   | MEGF6     |
| Chr3       | rs10511083| c.263 A > G  | CADM2     |
| Chr15      | rs72742862| c.80 T > G   | MEGF11    |
| Chr20      | rs761206  | c.195 T > G  | JPH2      |
| Chr21      | rs2826733 | c.236 G > T  | NCAM2     |
| Chr2       | rs3738896 | c.240 T > C  | PRKCE     |
| Chr7       | rs35660473| c.174 G > T  | DPP6      |
| Chr9       | rs2121200 | c.134 C > T  | FANCC     |
| Chr12      | rs11168200| c.146 C > G  | RPAP3     |
| Chr19      | rs788336  | c.461 T > C  | MYH14     |
| Chr20      | rs6041870 | c.481 T > C  | SPTLC3    |
| Chr12      | rs1259410 | c.288 T > C  | DENND5B   |
| Chr14      | rs861536  | c.216 A > G  | KLC1      |

The variation frequency of PRKCE and KLC1 were lower than normal individuals

Among 614 loci in 282 psoriasis patients, 240 loci in protein kinase C epsilon (PRKCE) were mutated (c.240T > C), and the variation frequency in patients with psoriasis (36.3%) was significantly lower than that in normal Asian individuals (47.7%, P < 0.05). Moreover, 216 loci in kinesin light chain 1 (KLC1) were mutated (c.216A > G), and the variation frequency in patients with psoriasis (1.2%) was significantly lower than that in normal Asian individuals (98.1%, P < 0.01). In addition, the mean variation frequencies of the genes MEGF6 (20.2% vs 25.9%), CADM2 (71.4% vs 86.1%), MEGF11(59.2% vs 61.8%), NCAM2(32.4% vs 33.6%), DPP6 (81.2% vs 83.9%), MYH14 (17.7% vs 22.0%), SPTLC3(64.5% vs 66.1%), FANCC (4.4% vs 4.60%) and DENND5B (0.53% vs 0.6%) were lower in patients with psoriasis than in normal individuals, though all of P value was higher than 0.05. However, the variation frequencies of JPH2 (86.1% vs 85.12%) and RPAP3 (28.0% vs 26.9%) were higher in patients with psoriasis than in normal individuals (Fig. 1).

Occurrence of psoriasis and its relationship with the sociodemographic characteristics of participants

Medical history, the PASI score and family history were used to clarify the different clinical subtypes. Therefore, the relationship between medical history, the PASI score, family history and variant genes was investigated. CADM2(χ² = 9.29, P < 0.05), JPH2(χ² = 8.47, P < 0.05), SPTLC3(χ² = 20.65, P < 0.01) was significantly different between patients with different medical histories(Fig. 2). Analysis of the PASI score revealed a significant association with variation in MEGF6 (χ² = 6.06, P < 0.05). Next, we further analyzed the relationship between family history and variant genes and found that CADM2(χ² = 6.08, P < 0.05), JPH2(χ² = 10.98, P < 0.01), SPTLC3(χ² = 8.51, P < 0.01) was significantly associated with family history, suggesting that the variation in CADM2,JPH2, SPTLC3 may be the risk factor for psoriasis in individuals who have a family member with psoriasis.

Univariate and multivariate analysis of family history on disease characteristics

For research purposes, we rearranged the population data and divided individuals with psoriasis into two groups, as shown in Table 2. Research has shown that with the extension of time, new mutations will emerge and old ones will be repaired. Therefore, to eliminate the impact of time on mutation gene, we divided into two groups based on medical history and then compare different PSAI and family history of patients. In the under 20 years medical history group and over 20 year medical history group, we analyzed the relationship between gender, age, PASI, family history and gene mutation. For the comparison of family history of two group, we found that CADM2(χ² = 5.89,94.87% vs 76.56%, p<0.05) was a risk factor for having family history with type II psoriasis. Similarly
when compared to patients with under 20 years medical history group and over 20 year medical history group, there were differences in \( MEGF6 (\chi^2 = 3.86, 39.52\% \text{ vs } 22.5\%, \ p<0.05) \) according to PASI of type II psoriasis.

### Table 2

| Medical history<20 | Medical history ≥ 20 |
|------------------|------------------|
| \( \chi^2 \) | \( \chi^2 \) |
| PASI | Family history | Family history | PASI | Family history | Family history |
| \( \leq 10 \) | (YES) | (NO) | \( >10 \) | (YES) | (NO) |
| MEGF6 | 39.52% | 22.50% | 3.83 | 41.03% | 31.25% | 1.02 | 40.24% | 22.22% | 3.58 | 36.84% | 36.92% | 0.00 |
| CADM2 | 83.87% | 72.50% | 2.55 | 94.87% | 76.56% | 5.89 | 73.17% | 75.00% | 0.04 | 71.05% | 73.85% | 0.09 |
| MEGF11 | 87.10% | 85.00% | 0.11 | 76.92% | 89.06% | 2.72 | 79.27% | 80.56% | 0.03 | 71.05% | 84.62% | 2.72 |
| JPH2 | 99.19% | 95.00% | 2.96 | 100.00% | 95.31% | 1.88 | 95.12% | 91.67% | 0.54 | 97.37% | 92.31% | 1.12 |
| NCAM2 | 57.26% | 52.50% | 0.28 | 56.41% | 56.25% | 0.00 | 52.44% | 61.11% | 0.76 | 63.16% | 50.77% | 1.49 |
| PRKCE | 49.19% | 42.50% | 0.54 | 33.33% | 51.56% | 3.26 | 57.32% | 61.11% | 0.15 | 63.16% | 55.38% | 0.60 |
| DPP6 | 94.35% | 92.50% | 0.18 | 92.31% | 98.44% | 2.44 | 91.46% | 97.22% | 1.31 | 89.47% | 93.85% | 0.64 |
| FANCC | 8.06% | 10.00% | 0.15 | 10.26% | 6.25% | 0.54 | 9.76% | 5.56% | 0.57 | 7.89% | 9.23% | 0.05 |
| RRAP3 | 44.35% | 57.50% | 2.10 | 51.28% | 50.00% | 0.02 | 48.78% | 41.67% | 0.51 | 39.47% | 49.23% | 0.92 |
| MYH14 | 31.45% | 42.50% | 1.64 | 33.33% | 28.13% | 0.31 | 31.71% | 22.22% | 1.10 | 31.58% | 27.69% | 0.18 |
| SPTLC3 | 81.45% | 87.50% | 0.78 | 87.18% | 81.25% | 0.62 | 91.46% | 83.33% | 1.69 | 94.74% | 84.62% | 2.39 |
| DENND5B | 0.00% | 0.00% | 0.00 | 0.00% | 0.00% | 0.00 | 2.44% | 2.78% | 0.01 | 2.63% | 1.54% | 0.15 |
| KLC1 | 3.23% | 7.50% | 1.35 | 2.56% | 4.69% | 0.29 | 0.00% | 0.00% | 0.00 | 0.00% | 0.00% | 0.00 |

### Discussion

Based on screening studies and genome-wide association studies (GWASs), a large number of susceptibility genes that are involved in the pathogenesis of complex diseases, such as cancer, psoriasis, and other major human diseases, have been identified [17–18]. Concerning psoriasis, more than 70 susceptible genes have been identified in the last 30 years [19–20]. In our study, we use next-generation sequencing technique to assess the status of 13 genes, and we finally found that \( PRKCE \) and \( KLC1 \) had mutated in psoriasis patients. The results of the study indicated that genetic variation has a very close correlation with clinical features.

A pathogenic role for \( PRKCE \) has been found in several types of cancers, as it is able to promote proliferation and inhibit apoptosis [21]. Constitutively active \( PRKCE \) has been found in small cell lung cancer [22] and epithelial cells of the colon [23] associated with tumor formation [24]. The \( PRKCE \) acts as a regulator of PLD activity, and this inhibition is mediated by its regulatory domain [25]. Moreover, the novel partial loss-of-function defect in \( PRKCE \) impairs AKT activation via compromised mTORC2 complex function [26]. According to studies of patients with psoriasis, the PI3K/AKT pathway is an important signaling pathway that regulates the hyperproliferation of keratinocytes [27]. It has also been reported that a variation in the phosphorylation site of \( KLC1 \), Ser517/Ser520, affects AMP-activated protein kinase (AMPK) to suppress low glucose concentrations and block granule movement [28]. In this study, we demonstrated that the variation rates of \( PRKCE \) and \( KLC1 \) were significantly lower in psoriasis patients than in normal Asian patients, consistent with the results reported by Salo-Mullen EE [29], indicating that the described \( PRKCE \) and \( KLC1 \) variants may be a negative regulatory mechanism for the onset of psoriasis.

Medical history and the PASI score are commonly used in clinical trials on psoriasis, and this scoring algorithm greatly expands options for quantifying treatment outcomes in cost-effectiveness analyses of psoriasis therapies [30]. To further evaluate the relationship between variant genes and clinical characteristics, we grouped patients with psoriasis according to sex, age, medical history, PASI score and family history. The variation rate was distinctly different according to medical history, PASI score and family.
history. We found that the CADM2, JPH2 and SPTLC3 variation have correlation with more than 20 years of medical history in individuals with type 1 psoriasis. Moreover, moderate-to-severe (PASI score ≤ 10) variants were more commonly observed in individuals with a MEGF6 variation. Indeed, several studies have shown the significant impact of the onset of psoriasis not only in patients with a medical history and a high PASI score but also in their family and close relatives [31–32]. The literature supports some differences between familial and sporadic cases of psoriasis [33]. Differences in the strength of the association between the variant gene and family history have been reported for the MHC, including a strong association of HLA-C*06 and HLA-B*27 with psoriasis [34]. Similarly, in our study, we found that the variation frequency of CADM2, JPH2 and SPTLC3 was higher in the familial group than in the sporadic group. Our findings are important for the promotion of large panels in patient populations with psoriasis and for clinical genetic testing for patient management. However, the specific regulatory mechanisms involved need to be explored in the skin cells of psoriasis patients.

**Conclusion**

We demonstrated that the variation rates of KLC1 and PRKCE were significantly lower in psoriasis than normal, indicating these two genes could serve as potential novel negative regulatory genes for type psoriasis. We showed considerable differences in the relationship between clinical characteristics and variant genes.

**Methods**

**Samples**

MZ Twins Samples from 8 pairs of Chinese MZ twins who were discordant in psoriasis and samples from the twins’ parents were collected with consent at Taiyuan Central Hospital.

282 patients diagnosed with vulgaris psoriasis were recruited from the outpatient center of the Taiyuan Central Hospital. All patients included in this study were of Han Chinese descent. Individuals with psoriasis were diagnosed by at least two dermatologists based on clinical and histopathological manifestations, and their clinical information was collected through a comprehensive clinical check-up by professional investigators. Self-reported information from a standard questionnaire was used to collect demographic and other characteristics (severity, medical history, and family history) from the patients and to exclude any other systemic, infectious, autoimmune, atopic, or malignant disease and to determine whether they received systemic treatment in the six months prior to collection. None of the patients had hypertension, gout, asthma or multiple coffee spots.

**Characteristics of the subjects**

The clinical features of MZ Twins Samples (4 pairs of women and 4 pairs of men, range from 11 to 51 years with the mean age 30.0 years) were shown in Table S1.

The clinical characteristics of the study subjects are shown in Table S2. In this study, we investigated 282 Chinese Han individuals with type psoriasis. Their age of onset is less than 40 and pertain to early onset, which all belong to the type 1 patients with psoriasis.

Regarding of medical history, PASI and family history, 282 patients were grouped, respectively. The medical history group was divided into two ranks: less than 20 years (58.2%) and more than 20 years (41.88%). Patients were selected based on disease severity, which was assessed using PASI score[13]. The number of patients in the mild group (PASI score ≤ 10) was 209(74.1%), and the number of patients in the moderate-to-severe group (PASI score > 10) was 73(25.9%). Patients were also grouped according to family history: positive (27.3%) and negative (49.3%).

**Whole Genome Sequencing And Variant Identification**

Each qualified sample of genomic DNA from whole blood of 9 twins was sequenced on the Illumina HiSeq platform using paired-end reads according to the manufacturer's instructions. The following types of reads were removed: (1) containing a sequencing adapter; (2) low quality base > 50%; and (3) 'N' base > 10%. All clean data from each sample were mapped to the human reference genome HG19 by Burrows Wheeler Aligner software (BWA, v0.75) [14]. Duplicate reads were marked by Picard tools. We used 3 different
methods to call SNPs and InDels and get overlap of them to ensure the accuracy of variant calling: SAMTOOLS mpileup [15], recommended best practices for variant analysis with the Genome Analysis Toolkit (GATK) [16] and Freebayes. We identified the DNM discordance between co-twins by checking the concordance genotype between them. We genotyped our DNMs with Sanger sequencing to confirm our DNM detection accuracy.

**DNA Extraction**

Genomic DNA was extracted from the peripheral whole blood of the 282 patients using a Blood Genomic DNA Midi Kit (Cwbio Biotech, Beijing, China) according to the manufacturer’s instructions. All DNA samples were dissolved in water and stored at -20°C until use.

**Sanger Sequencing**

Sequencing primers (Table S3) were designed for the 13 single-nucleotide polymorphisms (SNPs) using Primer Premier 5.0. Genomic DNA was amplified using the Bio-Rad PCR System. Thermal cycling was performed as follows: 5 min at 96°C for 10 cycles (20 seconds at 96°C; 30 seconds for touchdown at 52–62°C; and 60 seconds at 72°C), followed by 35 cycles (20 seconds at 96°C; 30 seconds at 52°C; and 60 seconds at 72°C), and ending with 5 min at 72°C.

**Statistical analysis**

Information on the variants frequency in 8624 normal Asian individuals was obtained from the Pubvar database(https://www.pubvar.com/). Amplicons were bidirectionally sequenced using an ABI 3730 system. We performed sequence analysis by using Variants Surveyor software. Variants included hybrid variants and homozygous variants. The variation frequency was calculated with the following equation: (hybrid + homozygous × 2)/282/2. Differences in patient demographics (e.g., stage, sex, age, severity, medical history, and family history) were evaluated with SPSS version 18.0. The chi-square ($\chi^2$) test was used to test the relationship between psoriasis and the investigated factors. Statistical significance was set at $P < 0.05$.

**Abbreviations**

PASI
Psoriasis Area Severity Index
GWASs
genome-wide association studies
AMPK
AMP-activated protein kinase
MZ
monozygotic
SNPs
single-nucleotide polymorphisms

**Declarations**

**Ethics approval and consent to participate**

All participants provided written informed consent. The study protocol was approved by the ethics committee of Taiyuan Central Hospital.

**Consent to publish**

All participants provided written informed consent.

**Availability of data and materials**
All data generated or analysed during this study are included in this published article.

**Competing interests**

The authors declare that they have no competing interests.

**Funding**

The authors are grateful for funding from the National Natural Science Foundation of China (nos. 81773336, 81602768, and 81803146).

**Authors’ Contributions**

XJ, WY: Put forward new thoughts the research and draft paper. ZX: Responsible for DNA extraction. LJ, HR, NX: Analyze the data. YG, LX, ZK: Responsible for the final version. All authors read and approved the final manuscript.

**Acknowledgements**

We would like to thank AJE (https://www.aje.cn/) for English language editing.

**Author information**

Shanxi Key Laboratory of Stem Cells for Immunological Dermatosis, Institute of Dermatology, Taiyuan Central Hospital of Shanxi Medical University, No. 5 Dong San Dao Xiang, Jiefang Road, Taiyuan 030009, China

Jianxiao Xing, Ying Wang, Xincheng Zhao, Junqin Li, Ruixia Hou, Xuping Niu, Guohua Yin, Xinhua Li, Kaiming Zhang

**Corresponding Author**

Kaiming Zhang, Tel.: +86-0351-5656080, E-mail: zhangkaiming@sina.com

**References**

1. Lin Y, Liu L, Sheng Y, Shen C, Zheng X, Zhou F, Yang S, Yin X, Zhang X. A catalog of potential putative functional variants in psoriasis genome-wide association regions. PLoS One. 2018;13(5):e0196635.
2. Rachakonda TD, Dhillon JS, Florek AG, Armstrong AW. Effect of tonsillectomy on psoriasis: a systematic review. J Am Acad Dermatol. 2015;72(2):261–75.
3. Tang L, Cheng Y, Zhu C, Yang C, Liu L, Zhang Y, Wen L, Zhang X, Zhou F, Yang S. Integrative methylome and transcriptome analysis to dissect key biological pathways for psoriasis in Chinese Han population. J Dermatol Sci. 2018;91(3):285–91.
4. Ammar M, Bouchlaka-Souissi C, Soumaya K, Bouhaha R, Ines Z, Bouazizi F, Doss N, Dhaoui R, Ben Osman A, Ben Ammar-El Gaaied A et al. Failure to find evidence for deletion of LCE3C and LCE3B genes at PSORS4 contributing to psoriasis susceptibility in Tunisian families. Pathol Biol (Paris), 2014; 62(1), 34–37.
5. Wang Y, Li Y, Hao M, Liu X, Zhang M, Wang J, Xiong M, Shugart YY, Jin L. Robust Reference Powered Association Test of Genome-Wide Association Studies. Front Genet. 2019;10:319.
6. Rahman P, Elder JT. Genetics of psoriasis and psoriatic arthritis: a report from the GRAPPA 2010 annual meeting. J Rheumatol, 2012;39(2): 431–433.
7. Fan X, Wang H, Sun L, Zheng X, Yin X, Zuo X, Peng Q, Standish KA, Cheng H, Zhang Y, et al. Fine mapping and subphenotyping implicates ADRA1B gene variants in psoriasis susceptibility in a Chinese population. Epigenomics. 2019;11(4):455–67.
8. Dębiak T, Soczewa E, Boer M, Różewicka-Czabańska M, Wiśniewska J, Serrano-Fernandez P, Mirecka A, Paszkowska-Szczur K, Lubinski J, Krysztoforska L, et al. Common variants of ZNF750, RPTOR and TRAF3IP2 genes and psoriasis risk. Arch Dermatol Res. 2014;306(3):231–8.
9. Li J, Lin H, Hou R, Shen J, Li X, Xing J, He F, Wu X, Zhao X, Sun L, et al. Multi-omics study in monozygotic twins confirm the contribution of de novo mutation to psoriasis. J Autoimmun. 2020;106:102349.
10. Henseler T, Christophers E. Psoriasis of early and late onset: characterization of two types of psoriasis vulgaris. Am Acad Dermatol. 1985;13:450–6.
11. Sun LD, Cheng H, Wang ZX, Zhang AP, Wang PG, Xu JH, Zhu QX, Zhou HS, Ellinghaus E, Zhang FR, et al. Association analyses identify six new psoriasis susceptibility loci in the Chinese population. Nat Genet. 2010;42(11):1005–9.

12. Hüffmeier U, Lascorz J, Becker T, Schürmeier-Horst F, Magener A, Ekici AB, Endele S, Thiel CT, Thorna-Usynski S, Mössner R, et al. Characterisation of psoriasis susceptibility locus 6 (PSORS6) in patients with early onset psoriasis and evidence for interaction with PSORS1. J Med Genet. 2009;46(11):736–44.

13. Klein RJ, Zeiss C, Chew EY, Tsai JY, Sackler RS, Haynes C, Henning AK, SanGiovanni JP, Mane SM, Mayne ST, et al. Complement factor H polymorphism in age-related macular degeneration. Science. 2005;308(5720):385–9.

14. Gharbi H, Ben Hassine I, Soltani I, Safra I, Ouerhani S, Bel Haj Othmen H, Teber M, Farah A, Amouri H, Toumi NH, et al. Association of Genetic Variants in ARID5B, IKZF1 and CEBPE with Risk of Childhood de novo B-Lineage Acute Lymphoblastic Leukemia in India. Asian Pac J Cancer Prev, 2016:17;3989–3995.

15. Zhao Y, Forst CV, Sayegh CE, Wang IM, Yang X, Zhang B. Molecular and genetic inflammation networks in major human diseases. Mol Biosyst. 2016;12(8):2318–41.

16. Liu QP, Wu LS, Li FF, Liu S, Su J, Kuang YH, Chen C, Xie XY, Jiang MH, Zhao S, et al. The association between GJB2 gene polymorphism and psoriasis: a verification study. Arch Dermatol Res. 2012;304(9):769–72.

17. Choon SE, Lai NM, Mohammad NA, Nanu NM, Tey KE, Chew SF. Clinical profile, morbidity, and outcome of adult-onset generalized pustular psoriasis: analysis of 102 cases seen in a tertiary hospital in Johor, Malaysia. Int J Dermatol. 2014;53:676–84.

18. Chen H, Toh TK, Szeverenyi I, Ong RT, Theng CT, McLean WH, Seielstad M, Lane EB. Association of skin barrier genes within the PSORS4 locus is enriched in Singaporean Chinese with early-onset psoriasis. J Invest Dermatol. 2009;129:606–14.

19. Stuart PE, Nair RP, Ellinghaus E, Ding J, Tejasvi T, Gudjonsson JE, Li Y, Weidinger S, Eberlein B, Gieger C, et al. Genome-wide association analysis identifies three psoriasis susceptibility loci. Nat Genet. 2010;42:1000–4.

20. McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernytsky A, Garimella K, Altshuler D, Gabriel S, Daly M, et al. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. Genome Res. 2010;20:1297–303.

21. Jain K, Basu A. The Multifunctional Protein Kinase C-epsilon in Cancer Development and Progression. Cancers (Basel). 2014;6(2):860–78.

22. Baxter G, Oto E, Daniel-Issakani S, Strulovic B. Constitutive presence of a catalytic fragment of protein kinase C epsilon in a small cell lung carcinoma cell line. J Biol Chem. 1992;267(3):1910–7.

23. Perlett GP, Folini M, Lin HC, Mischak H, Piccinini F, Tashjian AH Jr. Overexpression of protein kinase C epsilon is oncogenic in rat colonic epithelial cells. Oncogene. 1996;12(4):847–54.

24. Gorin MA, Pan Q. Protein kinase C epsilon: an oncogene and emerging tumor biomarker. Mol Cancer. 2009;8:9.

25. Pan Y, Yuan C, Cheng C, Zhang Y, Ma Y, Zheng D, Zheng S, Li Y, Jin Y, Sun Y, et al. Frequency and clinical significance of NF1 mutation in lung adenocarcinomas from East Asian patients. Int J Cancer. 2019;144(2):290–6.

26. Alcantara D, Elmslie F, Tetreault M, Bareke E, Hartley T, Care4Rare Consortium, Majewski J, Boycott K, Innes AM, Dyment DA, et al. O'Driscoll SHORT syndrome due to a novel de novo mutation in PRKCE (Protein Kinase Cvarepsilon) impairing TORC2-dependent AKT activation. Hum Mol Genet. 2017;26(19):3713–21.

27. Zhang M, Zhang X. The role of PI3K/AKT/FOXO signaling in psoriasis. Arch Dermatol Res. 2019;311(2):83–91.

28. McDonald A, Fogarty S, Leclerc I, Hill EV, Hardie DG, Rutter GA. Rutter Cell-wide analysis of secretory granule dynamics in three dimensions in living pancreatic beta-cells: evidence against a role for AMPK-dependent phosphorylation of KLC1 at Ser517/Ser520 in glucose-stimulated insulin granule movement. Biochem Soc Trans. 2010;38(1):205–8.

29. Solmaz D, Bakirci S, Kimyon G, Gural EK, Dogru A, Bayindir O, Dalkicilc E, Ozisler C, Can M, Akar S, et al. The impact of having family history of psoriasis or psoriatic arthritis on psoriatic disease. Arthritis Care Res (Hoboken). 2020;72:36–83.

30. Matza LS, Brazier JE, Stewart KD, Pinto L, Bender RH, Kircik L, Jordan J, Kim KJ, Mutebi A, Viswanathan HN, et al. Developing a preference-based utility scoring algorithm for the Psoriasis Area Severity Index (PASI). J Med Econ. 2019;22(9):936–44.

31. López-Esteban JL, Sánchez-Carazo JL, Sulleiro S. Effect of a family history of psoriasis and age on comorbidities and quality of life in patients with moderate to severe psoriasis: Results from the ARIZONA study. J Dermatol. 2016;43(4):395–401.
32. Wason JM, Dudbridge F. A general framework for two-stage analysis of genome-wide association studies and its application to case-control studies. Am J Hum Genet. 2012;90:760–73.

33. Chandran V, Schentag CT, Brockbank JE, Pellett FJ, Shanmugarajah S, Toloza SM, Rahman P, Gladman DD. Familial aggregation of psoriatic arthritis. Ann Rheum Dis. 2009;68(5):664–7.

34. Raposo I, Carvalho C, Bettencourt A, Da Silva BM, Leite L, Selores M, Torres T. Psoriasis pharmacogenetics: HLA-Cw*0602 as a marker of therapeutic response to ustekinumab. Eur J Dermatol. 2017;27(5):528–30.

Figures

**Figure 1**

Differences in variation frequencies between psoriasis patients and normal individuals
Figure 2

Cartesian inspection analysis in patients based on gender, age, the PASI score, medical history of psoriasis and a family history of psoriasis. A. The percentage of variant gene for the sex group. B. The percentage of variant gene for the age group. C. The percentage of variant gene for the PASI group. D. The percentage of variant gene for medical history group. E. The percentage of variant gene for family history group.