mTORC1 controls long-term memory retrieval

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Understanding how stored information emerges is a main question in the neurobiology of memory that is now increasingly gaining attention. However, molecular events underlying this memory stage, including involvement of protein synthesis, are not well defined. Mammalian target of rapamycin complex 1 (mTORC1), a central regulator of protein synthesis, has been implicated in synaptic plasticity and is required for memory formation. Using inhibitory avoidance (IA), we evaluated the role of mTORC1 in memory retrieval. Infusion of a selective mTORC1 inhibitor, rapamycin, into the dorsal hippocampus 15 or 40 min but not 3 h before testing at 24 h reversibly disrupted memory expression even in animals that had already expressed IA memory. Emetine, a general protein synthesis inhibitor, provoked a similar impairment. mTORC1 inhibition did not interfere with short-term memory retrieval. When infused before test at 7 or 14 but not at 28 days after training, rapamycin impaired memory expression. mTORC1 blockade in retrosplenial cortex, another structure required for IA memory, also impaired memory retention. In addition, pretest intrahippocampal rapamycin infusion impaired object location memory retrieval. Our results support the idea that ongoing protein synthesis mediated by activation of mTORC1 pathway is necessary for long but not for short term memory.

The ability to recall past events is a major determinant of survival strategies in all species. Memory retrieval is the act of making stored information available for immediate use and is considered to be a complex and probably multistage process by which previously acquired information is reactivated from a latent state to a state that permits its expression. Not always but in most cases it is also accompanied by a behavioral output in the animal that constitutes the experimental assessment of a given memory. Memories reactivation at the time of retrieval seems to be a rapid process since animals may retrieve as soon as they receive, usually without notice, the CS or other cues including US remindful stimuli, another related memory recall or an emotional context associated.

A body of evidence suggest that long-term memories (LTM) rely on functional and structural changes at synapses, as Ramón y Cajal had proposed more than a century ago1–3. Neurons and their synapses that are reactivated when the animals are demanded to retrieve are believed to be those that have been changed through the molecular processes that underlie memory formation4,4. But in contrast to this initial stage, the information about the molecular mechanisms of memory retrieval is surprisingly scarce and fragmentary, despite its necessity for utilizing learned information. Some neurotransmitters and their receptors including metabotropic and AMPA glutamate receptors are required for memory retrieval in the hippocampus and related structures, while β-adrenoreceptors and cholinergic muscarinic receptors have neuromodulatory action5–11. In addition, several protein kinases including ERK1/2, PI3K and PKA are necessary for retrieval of fear-motivated memories in the hippocampus9,10,12. Despite the fact that protein synthesis has been extensively reported as a key factor in memory consolidation and reconsolidation, its role on memory retrieval remains controversial. Experiments using anisomycin as a general protein synthesis inhibitor (PSI) yielded negative or conflicting results13–15. However, Nader and coworkers11 recently found that pretest intramygdala infusion of anisomycin or rapamycin (rapa), a specific mTORC1 inhibitor, impaired auditory fear memory retrieval. These findings suggest that ongoing protein synthesis induced by mTORC1 pathway is required in the amygdala to enable memory retrieval.

mTORC1 is a serine/threonine kinase complex that modulates cell proliferation and growth, metabolism, autophagy, and mRNA translation initiation16. In the brain it has a prime function in regulating synaptic plasticity and memory formation, via the control of protein synthesis, including dendritic translation of synaptic proteins17–20.

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Therefore, to further investigate the role of mTORC1 in memory retrieval we determined whether mTORC1 is required for retrieval of an inhibitory avoidance (IA) task, a fear-motivated and hippocampus-dependent learning experience. We found that inhibition of hippocampal mTORC1 using infusions of rapa before test (TS) sessions at 1, 7 or 14 days after training provoked a marked and reversible impairment of recent and remote IA LTM expression, but not when mTORC1 inhibition was performed before testing for short-term or for 28 days long-term memory. Inhibition of mTORC1 before LTM testing in the retrosplenial cortex, another brain region involved in IA LTM retrieval21, also resulted in transient impairment in memory expression. LTM expression of object location (OL) was also affected by pretest inhibition of mTORC1.

Results
Pretest infusion of rapamycin or emetine impaired IA LTM retrieval. We chose to use IA training because it is acquired in a single and brief training session, which makes it ideal for investigating the neural and molecular events associated with memory retrieval without interference from previous expression of the learned behavior which occurs in multi-trial tasks22.

Confirming and extending recent findings in auditory fear memory retrieval using intraamygdala infusions of PSIs11, we found that intrahippocampal infusions (Fig. 1a) of two different types of PSI, emetine (eme) or rapa, 15 min before an IA TS session 24 h after the first TS without any infusion. *p < 0.01, Veh vs. Rapa, 1°TS Veh vs. Rapa, and Rapa vs. reTS groups; **p < 0.001, Rapa vs. 1°TS Rapa groups, one-way ANOVA followed by Newman-Keuls Test, n = 9-10. Data are expressed as mean ± SEM of TR or TS session step-down latency.
eme, which is a general PSI. The deficit on memory retrieval caused by rapa was transient, because a subsequent TS session performed 5 h after the first one showed normal IA retention performance (Fig. 1b, dark gray bar). Moreover, this reversible blockade of memory retrieval by intrahippocampal rapa was also seen in trained rats that expressed IA memory at a 24 h TS and the inhibitor of mTORC1 was given 15 min before a second TS carried out 48 h after training (Fig. 1d, p < 0.05, rapa vs. veh groups, one-way ANOVA followed by Newman-Keuls Comparison Test, n = 9–10). When infused outside the intended area in the dorsal hippocampus rapa did not affect retrieval. For instance, it did not impair retention performance when infused in the dorsal thalamic region (n = 3) or the hippocampal hilus region (n = 2) (data not shown). Taking together, these findings indicate that ongoing protein synthesis is required in the dorsal hippocampus to retrieve IA memory.

The effect of intrahippocampal rapa on IA LTM expression was also seen when the compound was infused 40 min before subjecting the animals to a TS session carried out 24 h after training (Fig. 2, p < 0.05, rapa 40 min compared to veh group, Newman-Keuls Comparison Test after one-way ANOVA, n = 9 per group). However, administration of rapa into the dorsal hippocampus 3 h before the TS session provoked no impairment on memory retrieval (Fig. 2, p > 0.05, rapa3h vs. veh animals, Newman-Keuls Comparison Test after one-way ANOVA, n = 9 per group).

**Rapamycin affected LTM, but not STM recall.** We next determined whether the effect of rapa on memory expression depends on the age of LTM to be retrieved. As shown in Fig. 3b,c the inhibition of mTORC1 15 min before TS sessions at 7 or 14 days after training impaired IA LTM retention performance (Fig. 3b,c, p < 0.05, rapa 7d vs. veh groups and rapa 14d vs. veh groups, Newman-Keuls Comparison Test after one-way ANOVA, n = 11–12 and n = 6–7 respectively). In both cases, a retest carried out 5 h after TS restored retrieval (Fig. 3b,c, dark gray bars). At 28 days after IA training, some authors had reported that memory expression becomes muscimol and CNQX-insensitive after pretest intrahippocampal infusions23,24, suggesting that IA memory might become hippocampus-independent. Consistent with these previous works, the intrahippocampal infusion of rapa 15 min before 28 days TS provoked no alterations in memory retrieval (Fig. 3d, p > 0.05, rapa28d vs. veh groups, Student’s t Test, n = 9 per group). Moreover, mTORC1 is not required for retrieval of short-term memory (STM), because the infusion of rapa into the dorsal hippocampus 15 min before a TS session performed 2 h after training did not alter IA memory retention (Fig. 3a, p > 0.05, Student’s t Test, n = 8–12).

Memories have been proposed to rely on endogenous and exogenous substances state-dependent process25,26. The aim of the next experiment was to determine whether the impairing effect of pretest rapa on memory retrieval could be attributable to a mismatch between the pharmacological state of the animal in which information was acquired and the one in which it was recalled rather than as a consequence of retrieval molecular process disruption27,28. Four groups of rats received two infusions of vehicle or rapa, one delivered immediately after training and the other 15 min before testing (veh-veh, veh-rapa, rapa-veh and rapa-rapa). A one-way ANOVA showed a significant effect of treatment (p < 0.01, F = 6.54). The main finding of this experiment is that rapa impaired memory when given after training or before testing as well as when delivered at both time points (Fig. 4, veh-veh differs from all the others groups, Newman-Keuls Comparison Test after one-way ANOVA), indicating that alterations in the internal state of the rat is not responsible for the effect of pretest mTORC1 inhibition on memory retrieval.

Several non-specific factors affecting sensory-motor and emotional processes can influence retention performance of an IA task. For this reason, we tested whether bilateral rapa infusion into the dorsal hippocampus affects locomotor or exploratory activity. As shown in Fig. 5, rapa had no discernible effects on these parameters
in an open field (p > 0.05, Student’s t Test for both rearings and crossings of veh vs. rapa groups, n = 12 per group). Taking together with the experiment showing that pretest rapa did not affect STM retrieval (Fig. 3a), these results indicate that the impairing effect of rapa on LTM retention is not due to non-specific actions on stepping-down latency.

**Pretest rapamycin infusion on other brain regions also impaired memory retrieval.** Several brain regions are simultaneously involved in memory retrieval, inasmuch as the memories on which retrieval is based certainly are not confined to any single brain region29,30. Taking this into account, we next determined whether mTORC1 in the retrosplenial cortex (RSC), a region involved in consolidation and expression of IA training21, is required for memory retrieval. As shown in Fig. 6b, the pretest infusion of rapa into the anterior part of RSC (Fig. 6a) transiently blocked IA memory retrieval in a reversible way (Fig. 6b, p < 0.05, Newman-Keuls Comparison Test after one-way ANOVA). Pretest rapamycin infusion impaired OL memory retrieval. OL memory is the ability to discriminate the familiar location of previously encountered objects31,32. We chose this OL memory in order to assess if the disruptive effect of rapa on IA memory can be replicated in a non aversive hippocampus-dependent task. Figure 7

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**Figure 3.** Rapamycin effect depends on the age of the memory retrieved. (a) Rapamycin (Rapa) was administered 15 min before a test (TS) session carried out 2 h after training (TR). p > 0.05, Vehicle (Veh) vs. Rapa short term memory groups, unpaired Student’s t Test, n = 8–12. Data are expressed as mean ± SEM of TR or TS session step-down latency. In the next experiments, Rapa was administered 15 min before a TS session carried out (b) 7 (*p < 0.05, Veh vs. Rapa 7d and Veh vs. reTS groups, one-way ANOVA followed by Newman-Keuls Comparison Test, n = 11-12), (c) 14 (*p < 0.05, Veh vs. Rapa 14d groups, one-way ANOVA followed by Newman-Keuls Comparison Test, n = 6-7) or, (d) 28 days after TR (*p > 0.05, Vehs. Rapa 28 d groups, unpaired Student’s t test n = 9 per group). Data are expressed as mean ± SEM of TR or TS session step-down latency.
shows that the discrimination index significantly decreased with intrahippocampal administration of rapa 30 min before TS session of the OL task (p < 0.05, veh vs. rapa group, Student’s t test, n = 7-8).

**Discussion**

The ability to recall past event is of great importance in determining our uniqueness as individuals. In this regard, retrieval is critical for memory: without its retrieval it is not possible to ensure we have a given memory. The main finding of the present study is that a general PSI, eme, and a selective inhibitor of mTORC1, rapa, infused into the dorsal hippocampus before IA testing greatly impaired memory retrieval. Given that rapa has less nonspecific effects than a general PSI like eme, we chose to use the specific inhibitor of mTORC1 to further characterize the
involvement of ongoing protein synthesis on memory retrieval processing. The amnesic effect of rapa is limited to LTM, and does not depend on a drug-induced change in the internal state of the animal. Interestingly, STM retrieval does not require mTORC1 activity. This result cannot be explained by alterations in locomotor or exploratory activity, since rapa had no discernible effect on locomotion and exploration in an open field. Also, intrahippocampal rapa administration impaired memory expression of a non-aversive OL memory. Together, these findings endorse the idea that regulation of translation by mTORC1 at the moment of retrieval is required for the expression of LTM.

Remarkably, at each time point tested, the effect of rapamycin infusion on memory retrieval was transient since a 5 h re-test showed full memory recovery. This finding supports the idea that the infusion of rapa into the dorsal hippocampus produced no functional or anatomical damage and, is consistent with the fact that rapa administration 3 h prior testing had no effect on memory recall, suggesting that rapa has a limited action after its in vivo infusion.

Our present findings extend those of Lopez et al., who demonstrated that anisomycin and rapa delivered into the amygdala before testing impaired the retrieval of an auditory fear-conditioning. Because the inhibition
of mTORC1 in the RSC before testing also impaired IA memory retrieval (Fig. 6) and that rapa also affect OL memory retrieval (Fig. 7), we are tempted to suggest that the requirement of mTORC1 in memory retrieval is a general feature of memory recall in mammals.

In contrast to the amnesic effect of rapa when administered 15 min pretest at 1, 2, 7 or 14 days, mTORC1 in IA memory retrieval was not required when animals were tested 28 days after training, probably because IA memory trace is no longer in the dorsal hippocampus 4 weeks after IA training. Interestingly, this time point coincides with the period of time at which the hippocampus becomes totally insensitive to the impairing effect of pretest infusions of muscimol33 or CNQX 23, an AMPA/Kainate glutamate receptor antagonist. Our present findings using rapa are also consistent with experiments showing that disruption of hippocampal function mostly hindered recent LTM without affecting remote LTM29,34,35 although other authors had reversibly abolished remote memory recall by a brief optogenetic inhibition of CA1 36.

STM recall is also not affected by pretest infusions of rapa into the dorsal hippocampus, suggesting that mTORC1 activity is not necessary for the retrieval of IA memory at short times after training. This finding parallels those showing that STM does not depend on new protein synthesis 3,37. It is interesting to note here that several behavioral experiments in rodents indicate the requirement of mTORC1 in the formation, consolidation and reconsolidation of LTM, but not STM 38,22,30,39.

Also, we have shown that the rapa effect can be extended to an OL memory task, since intrahippocampal infusion of rapa before TS session hindered OL memory retrieval as shown in Fig. 7. Considering our present findings, the effect of mTORC1 blockade on memory retrieval appears to be valence-independent and the role of mTORC1 pathway on this memory phase can be generalized to several forms of associative learning tasks.

What are the upstream regulators of the activity of mTORC1? Extracellular activators of the mTORC1 pathway with relevance to memory retrieval include brain-derived neurotrophic factor (BDNF) and glutamate 17,40,41. Dendritic mTORC1 is activated by glutamate action on NMDA receptors to control long term potentiation (LTP) 42 and metabotropic glutamate receptors activation results in enhancing ERK and AKT-mTOR pathways controlling protein synthesis and long-term depression 43,44. Intracellular activators of mTORC1 such as ERK 1/2 and AKT have also important modulatory effects on retrieval of IA memory 45.

A prevailing view tells us that retrieval regroups the transformation of patterns of brain activity produced during initial learning. In other words, reactivation of distributed ensembles of neurons in different brain regions that were active during initial learning is necessary for the subsequent retrieval of the memory. If this is the case, how are the patterns of activity in the hippocampus generated and controlled during retrieval? Many years ago, it was suggested that all or part of the mechanisms occurring during encoding have to be reestablished at the moment of recall in order to successfully retrieve memory 27,28. However, our findings using posttraining and/or pretest administration of rapa (Fig. 4) did not support this idea. Later on, molecular pharmacological data showed that the biochemical changes underlying IA consolidation are in part similar to those of retrieval. Indeed, some types of glutamate receptors and the activity of PKA, PKC and MAPKs are necessary for both consolidation and retrieval, and the modulation by D1 dopamine receptors, β-adrenoceptors, SHT 1α receptors and cholinergic muscarinic receptors is similar in both cases 7,8,45,46. But the nature and the timing of the role played by each signaling pathway in consolidation is different from that of retrieval. In addition, other mechanisms crucial for encoding or consolidation of IA LTM like NMDA receptors and CaMKII are in general terms not necessary for memory retrieval.

Besides changes in the modulation of translation, brief inhibition of mTORC1 by rapa affects biological processes such as ion homeostasis, regulation of membrane potential, regulation of secretion and synaptic vesicle trafficking 47. These findings suggest that acute inhibition of mTORC1 may greatly impact synaptic function and as a consequence impairs memory retrieval.

In conclusion, we found that the activity of mTORC1 close in time to memory recall is required for the normal expression of aversive and non-aversive LTM in the rat. Together with the seminal work of Nader group in amygdala 48, our findings open new lines of research regarding the molecular mechanisms underlying memory retrieval.

**Materials and Methods**

**Subjects.** Male adult Wistar rats weighing 200–250 g on arrival (Facultad de Ciencias Exactas y Naturales, University of Buenos Aires) were housed in groups of five per cage and kept with water and food ad libitum under a 12 light/dark cycle (lights on at 7 A.M.) at a constant temperature of 23 °C. Experiments took place during the light phase of the cycle. Animals were handled for 3 min for 3 consecutive days before the experiment to avoid emotional stress.

All the experimental protocols used for this study followed the guidelines of the National Institutes of Health Guide for Care and Use of Laboratory Animals and were approved by the Animal Care and Use Committees of the University of Buenos Aires (CICUAL).

**Surgery.** Rats were bilaterally implanted under deep ketamine/xylazine anesthesia (100 and 5 mg/kg, respectively) with a 22-gauge guide cannulae aimed to the dorsal CA1 region of the hippocampus (AP - 3.9 mm, LL ± 3.0 mm, DV - 3.0 mm from Bregma) or to the retrosplenial cortex (AP - 3.9 mm, LL ± 0.5 mm, DV - 1.8 mm from Bregma). Coordinates were based on Paxinos and Watson Atlas 49. Cannulae were fixed to skull with dental acrylic. At the end of the surgery, animals were injected with a single dose of meloxicam (0.2 mg/kg) as analgesic and gentamicin (2.5 mg/kg) as antibiotic. Behavioral procedures took place 5–7 days after surgery.

**Inhibitory Avoidance Task.** After recovery from surgery, IA was performed as described previously 22. During training, rats were placed on a 3 cm high, 9 cm wide platform at left of a 47 × 25 × 30 cm opaque acrylic box whose floor was a grid made of a series of parallel 1 mm-caliber steel bars spaced 1 cm apart. As they stepped
down onto the grid they received a 3 s, 0.7 mA scrambled foot shock and the latency to step down with all four paws was measured. Rats were tested for retention 2 h, 24 h, 2, 7, 14 or 28 days after training, depending on the experiment. In the TS the procedures were similar except that the foot-shock was omitted and the time spent on the platform was evaluated for a maximum of 300 s. Typical IA 24 h step-down latency was 150 ± 30 s. In the case where the reversibility of the rapi effect was examined, animals were retested 5 h after the first TS.

Object Location Task. The OL task consisted of 3 days of 20 min habituation sessions in the absence of the objects, a 10 min TR and a 3 min TS. Before habituation, all rats were handled 2 min daily for 3 days. The experimental apparatus was a 60 cm wide × 40 cm long × 50 cm high acrylic box with a transparent frontal wall and hatched back wall, while laterals walls were white with different visual clues. On the TR day, the objects (two identical glass bottles) were located in the arena in two adjacent corners and animals were allowed to explore for 10 min. During TS, performed 24 h after training, one of the objects was switched to a new position and exploration time was recorded again. A rat was scored as exploring an object when it orientates its head towards the object with the nose within 1 cm of the object with behaviors including sniffing, touching and gnawing. Sitting on the object, looking up while resting against the object or any time where the rat simply propped the forepaws onto the object with the nose pointing away from it were not counted as exploration. Rats were excluded from the analysis if they explore one of the objects more than 65% of total objects–exploration time during TR. In both TR and TS, rats with total objects–exploration time lower than 10 s were excluded. The exploration time was measured using a hand stopwatch and results are expressed as a discrimination index: $\text{[Exploration time of new location (Tn) - Exploration time of familiar location (TF)]/(Tn + TF)}$.

From rat to rat, the familiar or novel OL of the object in TS was counterbalanced. The box and the objects were thoroughly cleaned between trials.

Open Field Test. To evaluate locomotor activity animals were exposed to an open field. The OF arena consisted in a square box of 50 × 50 × 39 cm, with black walls and floor, which was divided into nine quadrants by white stripes. Animals were left to explore for 5 min and the exploratory activity was measured as the number of crossings between squares and the number of rearings registered minute by minute.

Drug Infusion. For intracerebral infusions, 30-gauge needles connected to Hamilton syringes were used. For rapa, the volume infused was 0.5 µl/side and the infusion rate was 0.5 µl/min. For eme, the volume infused was 1 µl/side and the infusion rate was 1 µl/min. Infusions were delivered through a needle extending 1 mm beyond the tip of the guide cannula. Injectors were left in place for an additional min following infusion before they were removed carefully to minimize backflow. During the procedure the animals were slightly restrained with the hands, without provoking any evident stress. Drugs and doses were as follows: rapa (specific mTORC1 inhibitor) was removed carefully to minimize backflow. During the procedure the animals were slightly restrained with the

Cannula placement. Cannula placement was verified after the end of the behavioral procedures by infusions of 0.5 µl of a solution of 4% methylene blue in saline. Animals were killed by decapitation 15 min later and histological localization of cannula placement was performed, taking the extension of the dye as an indicator of the presumable diffusion of rapa. Infusions spread with a radius between 0.5 and 1.0 mm3 as described before21,22,50 was similar to that found using 3H-muscimol or fluorescent muscimol51,52 and is in agreement with published data from our group26, in which rhodamine labeled alpha-bungarotoxin infusions match those observed in the schema of Fig. 1a. Infusions were found to be correct (i.e., within 1.5 mm3 of the intended site) in 94% of the animals. Only the behavioral data from animals with the cannula located in the intended site were included in the final analysis.

Data analyses. Statistical analysis and graphs of behavioral data were performed with unpaired Student’s t test when comparing two groups and one-way ANOVA Test followed by Newman-Keuls Comparison Test when comparing 3 or more groups using Graph Pad Prism 5® software (Graphpad, La Jolla, CA, USA). In all the cases, α level was set at 0.05. IA data in the bar graphs were expressed as mean ± SEM of training or test session step-down latency. In addition, training effect was always found significant; thus, for the sake of visual simplicity symbols showing significant differences between step-down latencies of training and testing were omitted.

References
1. Kandel, E. R. The molecular biology of memory storage: a dialogue between genes and synapses. Science 294(5544), 1030–1038 (2001).
2. Kandel, E. R., Dudai, Y. & Mayford, M. R. The molecular and systems biology of memory. Cell 157(1), 163–186 (2014).
3. McGaugh, J. L. Memory—a century of consolidation. Science 287(5451), 248–251 (2000).
4. Redondo, R. L. et al. Bidirectional switch of the valence associated with a hippocampal contextual memory engram. Nature 513(7518), 426–430 (2014).
5. Nader, K. Memory traces unbound. Trends Neurosci 26(2), 65–72 (2003).
6. Riedel, G. et al. Reversible neural inactivation reveals hippocampal participation in several memory processes. Nat Neurosci 2(10), 898–905 (1999).
7. Szapiro, G. et al. Participation of hippocampal metabotropic glutamate receptors, protein kinase A and mitogen-activated protein kinases in memory retrieval. Neuroscience 99(1), 1–5 (2000).
8. Szapiro, G. et al. Facilitation and inhibition of retrieval in two aversive tasks in rats by intrahippocampal infusion of agonists of specific glutamate metabotropic receptor subtypes. Psychopharmacology (Berl) 156(4), 397–401 (2001).
9. Szapiro, G. et al. Molecular mechanisms of memory retrieval. Neurochem Res 27(11), 1491–1498 (2002).
10. Thomas, S. A. Neuromodulatory signaling in hippocampus-dependent memory retrieval. Hippocampus 25(4), 415–431 (2015).
11. Lopez, J., Gamache, K., Schneider, R. & Nader, K. Memory retrieval requires ongoing protein synthesis and NMDA receptor activity-mediated AMPA receptor trafficking. J Neurosci 35(6), 2465–2473 (2015).
12. Shen, X. et al. PI3 kinase signaling is required for retrieval and extinction of contextual memory. Nat Neurosci 8(7), 925–931 (2005).
13. Berman, D. E. & Dudai, Y. Memory extinction, learning anew, and learning the new: dissociations in the molecular machinery of learning in cortex. Science 291(5512), 2417–2419 (2001).
14. Dubue, J. D., McKinney, T. L., Treit, D. & Dickson, C. T. Intrahippocampal Anisomycin Impairs Spatial Performance on the Morris Water Maze. J Neurosci 35(31), 11118–11124 (2015).
15. Vianna, M. R., Szapiro, G., McGaugh, J. L., Medina, J. H. & Izquierdo, I. Retrieval of memory for fear-motivated training initiates extinction requiring protein synthesis in the rat hippocampus. Proc Natl Acad Sci USA 98(21), 12251–12254 (2001).
16. Hay, N. & Sonenberg, N. Upstream and downstream of mTOR. Genes Dev 18(16), 1926–1945 (2004).
17. Takei, N. et al. Brain-derived neurotrophic factor induces mammalian target of rapamycin-dependent local activation of translation machinery and protein synthesis in neuronal dendrites. J Neurosci 24(44), 9760–9769 (2004).
18. Schratt, G. M., Nigh, E. A., Shen, W. G., Hu, L. & Greenberg, M. E. BDNF regulates the translation of a select group of mRNAs by a mammalian target of rapamycin-phosphorylidyinositol 3-kinase-dependent pathway during neuronal development. J Neurosci 24(33), 7366–7377 (2004).
19. Hoeffer, C. A. & Klann, E. mTOR signaling: at the crossroads of plasticity, memory and disease. Trends Neurosci 33(2), 67–75 (2010).
20. Liu-Yesucevitz, L. et al. Local RNA translation at the synapse and in disease. J Neurosci 31(45), 16086–16093 (2011).
21. Katche, C. et al. On the role of retrosplenial cortex in long-lasting memory storage. Hippocampus 23(4), 295–302 (2013).
22. Bekinschtein, P. et al. mTOR signaling in the hippocampus is necessary for memory formation. Neuronbrot Learn Mem 87(2), 303–307 (2007).
23. Bianchini, M. et al. Memory expression is blocked by the infusion of CNQX into the hippocampus and/or the amygdala up to 20 days after training. Behav Neural Biol 59(2), 83–86 (1993).
24. Gonzalez, M. C. et al. Medial prefrontal cortex dopamine controls the persistent storage of aversive memories. Front Behav Neurosci 8, 498 (2014).
25. Overton, D. A. State-Dependent or “Dissociated” Learning Produced with Pentobarbital. J Comp Physiol Psychol 57, 3–12 (1964).
26. Izquierdo, I. & Dias, R. D. Memory as a state dependent phenomenon: role of ACTH and epinephrine. Behav Neural Biol 38(1), 144–149 (1983).
27. Spear, N. E. Retrieval of memory in animals. Psychological Review 80(3), 163–194 (1973).
28. Izquierdo, I. & Ferreira, M. B. Diazepam prevents post-training drug effects related to state dependency, but not post-training memory facilitation by epinephrine. Behav Neural Biol 51(1), 73–79 (1989).
29. Frankland, P. W. & Bontemp, B. The organization of recent and remote memories. Nat Rev Neurosci 6(2), 119–130 (2005).
30. Izquierdo, I. et al. The connection between the hippocampal and the striatal memory systems of the brain: a review of recent findings. Neurotox Res 10(2), 113–121 (2006).
31. Dere, E., Huston, J. P. & De Souza Silva, M. A. Integrated memory for objects, places, and temporal order: evidence for episodic-like memory in mice. Neuronbrot Learn Mem 84(3), 214–221 (2005).
32. Ennaceur, A. & Delacour, J. A new one-trial test for neurobiological studies of memory in rats. 1: Behavioral data. Behav Brain Res 31(1), 47–59 (1988).
33. Gonzalez, M. C., Villar, M. E., Izquierdo, I., Medina, J. H. & Bayley, P. J. The neuroscience of remote memory. Curr Opin Neurobiol 18(2), 185–196 (2007).
34. Squire, L. R. & Bayley, P. J. The neuroscience of remote memory. Curr Opin Neurobiol 17(2), 185–196 (2007).
35. Sutherland, R. J. et al. Retrograde amnesia after hippocampal damage: recent vs. remote memories in two tasks. Hippocampus 11(1), 27–42 (2001).
36. Goshen, I. et al. Dynamics of retrieval strategies for remote memories. Cell 145(3), 678–689 (2011).
37. Kandel, E. R. The molecular biology of memory: cAMP, PKA, CRE, CREB-1, CREB-2, and CREB. Mol Brain 5, 14 (2012).
38. Parsons, R. G., Gafford, G. M. & Helmstetter, F. J. Translational control via the mammalian target of rapamycin pathway is critical for the formation and stability of long-term fear memory in amygdala neurons. J Neurosci 26(50), 12977–12983 (2006).
39. Roesler, R. Molecular mechanisms controlling protein synthesis in memory reconsolidation. Neuronbrot Learn Mem 142(1 Pt A), 30–40 (2017).
40. Lenz, G. & Arruver, J. Glutamatergic regulation of the p7056 kinase in primary mouse neurons. J Biol Chem 280(46), 38121–38124 (2005).
41. Slipczuk, L. et al. BDNF activates mTOR to regulate Glur1 expression required for memory formation. PLoS One 4(6), e6007 (2009).
42. Gong, R., Park, C. S., Abbassi, N. R. & Tang, S. J. Roles of glutamate receptors and the mammalian target of rapamycin (mTOR) signaling pathway in activity-dependent dendritic protein synthesis in hippocampal neurons. J Biol Chem 281(27), 18802–18815 (2006).
43. Banko, J. L. et al. The translation repressor 4E-BP2 is critical for eIF4F complex formation, synaptic plasticity, and memory in the hippocampus. J Neurosci 25(42), 9581–9590 (2005).
44. Hou, L. & Klann, E. Activation of the phosphoinositide 3-kinase-Akt-mammalian target of rapamycin signaling pathway is required for metabolotropic glutamate receptor-dependent long-term depression. J Neurosci 24(28), 6532–6561 (2004).
45. Barros, D. M. et al. Molecular signalling pathways in the cerebral cortex are required for retrieval of one-trial avoidance learning in rats. Behav Brain Res 114(1-2), 183–192 (2000).
46. Izquierdo, I. & MaGruA, J. L. Behavioural pharmacology and its contribution to the molecular basis of memory consolidation. Behav Pharmacol 11(7-8), 517–534 (2000).
47. Null, F. et al. Analysis of Proteins That Rapidly Change Upon Mechanistic/Mammalian Target of Rapamycin Complex I. Protein 7 (PARK7) as a Novel Protein Aberrantly Expressed in Tuberous Sclerosis Complex (TSC). Mol Cell Proteomics 15(2), 426–444 (2016).
48. Paxinos, G. & Watson, C. The Rat Brain in Stereotaxic Coordinates (Elsevier Ltd), Vol. 1, pp. 33–37 (1997).
49. Alberini, C. M. The role of protein synthesis during the labile phases of memory: revisiting the skepticism. Neurobiol Learn Mem 89(3), 234–246 (2008).
50. Bernabeu, R., Cammarota, M., Izquierdo, I. & Medina, J. H. Involvement of hippocampal AMPA glutamate receptor changes and the CAMPProtein kinase A/CREB-P signalling pathway in memory consolidation of an avoidance task in rats. Braz J Med Biol Res 30(8), 961–965 (1997).
51. Allen, T. A. et al. Imaging the spread of reversible brain inactivations using fluorescent muscimol. J Neurosci Methods 171(1), 30–38 (2008).
52. Martin, J. H. & Ghez, C. Pharmacological inactivation in the analysis of the central control of movement. J Neurosci Methods 86(2), 145–159 (1999).
53. Tomaiuolo, M., Gonzalez, C., Medina, J. H. & Piriz, J. Lateral Habenula determines long-term storage of aversive memories. Front Behav Neurosci 8, 170 (2014).

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**Author Contributions**
C.K., J.H.M. designed research; M.P., A.B.L. and C.K. performed research; M.P., C.K., J.H.M. and A.B.L. analyzed data; and J.H.M., M.P. and C.K. wrote the paper. The authors declare no conflict of interest.

**Additional Information**

**Competing Interests:** The authors declare no competing interests.

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