Safety issues and new rapid detection methods in traditional Chinese medicinal materials

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Abstract The safety of traditional Chinese medicine (TCM) is a major strategic issue that involves human health. With the continuous improvement in disease prevention and treatment, the export of TCM and its related products has increased dramatically in China. However, the frequent safety issues of Chinese medicine have become the ‘bottleneck’ impeding the modernization of TCM. It was proved that mycotoxins seriously affect TCM safety; the pesticide residues of TCM are a key problem in TCM international trade; adulterants have also been detected, which is related to market circulation. These three factors have greatly affected TCM safety. In this study, fast, highly effective, economically-feasible and accurate detection methods concerning TCM safety issues were reviewed, especially on the authenticity, mycotoxins and pesticide residues of medicinal materials.

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Abbreviations: AA, aristolochic acid; Afs, aflatoxins; DON, deoxynivalenol; GICA, gold immunochromatographic assay; LOD, limit of detection; OTA, ochratoxin A; PAs, pyrrolizidine alkaloids; SNP, single nucleotide polymorphism; SSCP, single-strand conformation polymorphism; ZEN, zearalenone

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1. Safety issues of traditional Chinese medicine (TCM)

1.1. Safety issues of adulterants and toxic TCM in the market

Various TCM materials are confusing because of historical and geographical reasons and this confusion brings enormous danger to the TCM safety. Potent toxic substances, including aconite, aristolochic acid, anticholinergic, podophyllin, grayanotoxin, pyrrolizidine alkaloids, matrine, gelsemine, teucvin and strychnine, were easily misidentified, erroneously substituted with other herbs or intentionally adulterated for greater benefit. Traditional identification methods recognize materials by the morphological characteristics of TCM; such methods mainly depend on the expertise of the person who identifies. Once misidentified, the TCM can cause serious toxicity problems. The problem is, however, that we are facing the lack of the experts in TCM identification. At present, the most commonly used detection platforms are based on analytical laboratory instruments. These approaches fail to meet the purpose of rapid on-site analysis in the quarantine clearance of quarantine-related departments.

1.2. Mycotoxin-related safety problems of TCM materials

Medicinal plants are the main raw materials in TCM production. These plants may be infected by fungi and mycotoxins during their growth in fields or in the process of harvest and storage, thereby increasing the odds of significant health problems induced by TCM (e.g., teratogenesis, immunotoxicity or even cancer). Currently, 500 different mycotoxins have been recognized, among which the most common and of particular interest are aflatoxins (AFs), ochratoxins, fumonisins and deoxynivalenol (DON). Medicinal plants, such as platycladi seed and raw malt, are often infected by AFs. The positive rate of AFs present in Nelumbo nucifera (Gaertn.) is up to 70%, in which the content of 30% AF B1 samples and the total content of 25% AF samples both exceeds the standard limits of 5 μg/kg and 10 μg/kg, respectively. Moldy licorice samples are infected by both AFs and ochratoxin A (OTA), and their infection levels are relatively high. In OTA investigations of 57 medicinal material samples distributed in six regions in China, the results showed that the positive rate of molded samples by storage is 74%, and that of un-molded samples is only 8%. The OTA content of partial samples exceeded the standard limit set by the European Union, implying serious undetected toxicity for clinical drug use. Furthermore, Semen Cocis listed in the ‘Medicine Food Homology’ could be easily infected by zearalenone (ZEN) besides DON, and Baohe pills made from the powder of Semen Cocis are also easily infected by DON. In recent years, foreign scholars reported that 5% OTA in licorice root was transferred into boiling tea, and 1% OTA was transferred into impregnated tea. The above results demonstrate the urgent need for the monitoring of mycotoxin residues during TCM production.

1.3. Pesticide-related safety problems of TCM materials

More than 12,000 pesticides exist throughout the world. The pesticides mainly found in TCM materials include organochlorine, organophosphorus, pyrethroid and carbamate pesticides. Although organochlorine pesticides have been banned for many years, their residues may still exist in TCM because of their stable nature. Moreover, these residues are uneasy to decompose and can be stored in water, soil or biological organisms for a long time. Long-term use of TCM may lead to exposure to pesticide residues beyond safety limits, resulting in bioaccumulation and poisoning. Some cause-and-effect relationships of pesticides (e.g., arsine and organochlorine) have been clearly established. Epidemiological investigations showed that the risk of cancer is increasing in rural areas, including leukemia, malignant brain tumor, testicular cancer, multiple myeloma and lymphoma. Washing methods can be used to remove residual water-soluble pesticides from the plants successfully. However, to remove most fat-soluble pesticides, which possess high biological attachment coefficient, strong penetrability, and can easily enter plant, washing is much ineffective.

In view of the above, the establishment of accurate, rapid and simple methods for safety monitoring of TCM materials is urgently needed.

2. New rapid detection methods of the safety monitoring of TCM materials

2.1. Rapid detection of authenticity

2.1.1. DNA barcode

DNA barcode was proposed for the first time by Canadian zoologist Paul Hebert in 2003. It is a new molecular diagnostic technology that identifies species using a recognized standard short sequence in the genome. Gregory believed that global DNA barcode innovation research would become a “big science” program after the human genome project. Miller explained and popularized DNA barcoding in “the Renaissance of DNA barcode and taxonomy”. DNA barcode has become a global research highlight and direction for biological taxonomy in both academic journals and lay media. DNA barcoding technology has superseded the limitations imposed by traditional morphological identification methods. By establishing a TCM identification database, the digitalized DNA barcode moves TCM identification methods from morphological identification to molecular identification.

Based on standardized DNA barcodes and universal primers, the DNA barcode method is universal. By comparing sequences among species, identification can be easily processed without the taxonomic knowledge of a specialist. DNA barcoding is not restricted by morphological characteristics and physiological conditions. Researchers can accurately determine the information of a species by analyzing DNA sequences. Chen et al. first developed ITS2 as a DNA barcoder of medicinal plants, established a plant barcode identification system that was mainly based on ITS2, and used psbA-trnH as a complementary barcode. This system has high identification efficiency in Rosaceae, Asteraeaceae, and many other families or genera. Chen et al. also developed an animal barcode identification system, which was mainly based on CO1, and used ITS2 as a complementary barcode. Chen et al. completed the construction of a standardized DNA barcode database of TCM materials and identification website (http://www.tcmbarcode.cn/en). With this platform, rapid identification of original plants, pills, powder, tissues, or cells can be realized. The TCM barcode database will become permanent data that can be improved by adding new research sequences from taxonomists. Based on the database of the TCM barcode, Chen et al. designed the DNA barcode identification software for many companies to meet their requirements of rapid detection. Chen’s research proposed a new perspective for potential universal barcode sequence identification of all land plants, stimulating wide
discussion. The proposed barcode ITS2 has become a hotspot in international plant barcode research.

DNA barcoding can achieve rapid, accurate and automated identification of species without material specificity. In basic laboratories, researchers use DNA barcoding to detect and identify a large number of samples of herbal materials. The whole process of rapid identification can be completed within 4 h, which meets the requirement of entry-exit inspection and quarantine where identification demands a rapid, high-throughput, sensitive, accurate test for TCM materials. DNA barcoding has good application potential in toxic TCM identification. Our group have developed a special molecular method which could identify all the TCM materials containing aristolochic acid (AA), and we have also designed special primer to identify all the TCM materials containing retronecine-type and otonecine-type pyrrolizidine alkaloids (PAs).

On the other hand, authentication of TCM and their adulterants were widely carried out by using species-specific PCR and microarray. With the dramatic reduction in the cost of high-throughput sequencing, full-length sequencing of the chloroplast gene sequences can be used to find one suitable sequence as a DNA barcode or the entire chloroplast genome can be used as the ultra-barcode. Another alternative is the application of the single nucleotide polymorphism (SNP) method.

2.1.2. SNP identification

Based on DNA barcoding analysis, Chen et al. detected two stable SNPs for Panax ginseng and Panax quinquefolius authentication, whereas Liao et al. obtained two SNPs for identifying Panax notoginseng. With the development of DNA barcoding, increasing amounts of SNPs have been discovered. A series of detection methods for SNPs has been explored, such as single-strand conformation polymorphism (SSCP) and invader assay with dual-color fluorescence polarization detection. However, all these methods have some shortcomings, such as complex methodology, time-consuming steps, and expensive instruments, all of which discourage wide implementation. Therefore, microarray-in-a-tube, gold nanoparticles (GNPs), and nucleic acid test strips have high potential for fast detection of SNPs.

2.1.2.1. Microarray-in-a-tube

Microarray-in-a-tube is a novel DNA microarray technique in which specific nucleic acid probes are immobilized on the inner surface of a converted Eppendorf tube cap. Different from conventional glass microarray, the probes are arranged on a plastic substrate by agarose film. An inner vessel to store the hybridization solution is placed in the sealed tube. After amplification, the tube can be inverted and hybridization can be performed without contamination. Liu et al. successfully detected single-base mutations of HIV-1 resistance through microarray-in-a-tube. Considering the quenching ratio, single base mismatch discrimination ratio, and time-cost, Wang et al. proved that this method has an advantage over the traditional chip. Moreover, Liu et al. detected four respiratory tract viruses using microarray-in-a-tube accompanied with reverse transcription-PCR. The sensitivity of the system for virus detection can reach 10^2 copies/μL. Liu et al. prepared kits for several viruses. Microarray-in-a-tube can be used in the detection of similar clinical respiratory viruses, such as distinguishing the SARS virus from other viruses. The major advantages of the method are multivirus detection and elimination of contamination.

Currently, microarray-in-a-tube is a fast and feasible technology in the detection of SNPs. Based on the theory, SNPs in TCM can be detected through microarray-in-a-tube. Investigations on SNP detection kits for P. ginseng, P. quinquefolius, Ophiocordyceps sinensis, and other expensive medicinal materials are currently underway.

2.1.2.2. GNP technology

GNP is an SNP detection method based on color reaction. It is based on a method in which single- or double-stranded DNA has different electrostatic interactions among the GNPs. GNP is based on the characteristic that DNA bases are strictly complementary paired and hybridized to form double-stranded DNA; it uses the color change or aggregation condition as the signal to determine whether the detected sequence of a target gene has mutations or not. Given that GNPs have unique physical and chemical properties, this methodology can improve the accuracy and stability of biological detection.

In recent years, the use of GNPs for gene mutation detection and analysis of SNPs has shown rapid development in the research field. Many scholars have contributed to the methods of gene mutation and SNP detection by GNPs. For example, Rothberg designed a new method according to the dynamics of double-stranded DNA melting, and their results showed that the method can detect a minimum of 100 fmol of target DNA within 5 min. Their probes did not need thiol modification and the PCR products did not require purification, so the detection steps were further simplified and the cost was reduced. Bao's studies have shown that GNP probes combined with gene chip have many advantages, such as simple operation, short operating time, specificity and high sensitivity. GNP technology is convenient in gene mutation detection and SNP analysis.

GNPs are easily prepared and stored. Their advantages include high detection sensitivity and simple observation. With this technology, the herb-specific detection probes, complementary target sequence, and oligonucleotide DNA with single-base mutation sequences can be designed according to SNP sites. At room temperature, detection probes are hybridized between complementary sequences and single-base mutation sequences in the sequence buffer. With the addition of NaCl solution, the GNP solution produces distinctly different color changes in two hybridization solutions. Thus, the authenticity of medicines can be identified in a fast, effective, and stable manner. GNPs do not require special markers, such as fluorescent dye or expensive equipment, so this technology can achieve low-cost, high-throughput, high sensitivity and high automation detection. This method can be used for on-site testing of TCM materials.

2.1.2.3. A nucleic acid amplification test strip method

Isothermal nucleic acid amplification is a methodology that extends the length of target DNA sequences or increases their copy numbers at a specific temperature. Compared with PCR, this technique can conduct amplification in an isothermal period, thereby eliminating the requirements for instruments. Moreover, the temperature control system can be operated by a heating module, water bath, or other simple instruments. Hangzhou USTAR Bio-tech Limited developed a fast detection technique for SNPs. This technique contains one-step PCR and a nucleic acid test strip, which is a detection method for specific extension products. In this technique, the regions containing SNPs are amplified unspecifically at first. The SNP sites are then specifically amplified by allele-specific PCR. Finally, the specific amplification products are detected by a nucleic acid test strip.

Wang et al. designed a suite of loop-mediated isothermal amplification primers for the sequences of the exogenous gene Cry1ab/ac in Bt-transgenic crops. Given the method's high reliability, specificity and stability, it can be used in the rapid
on-site detection of Bt-transgenic crops. Qin et al.\textsuperscript{61} established cross-priming amplification for the rapid detection of \textit{Vibrio cholerae}, and the method was proved to have high specificity and stability. Wang et al.\textsuperscript{62} developed a new method for the rapid detection of mtDNA G1178A mutation based on the SNP test strip; the result of the SNP test strip was identical to that of DNA sequencing. Zhang et al.\textsuperscript{63} used this method to detect \textit{Mycobacterium tuberculosis} in sputum, and found that this method is fast (within 2 h), sensitive, and easy to operate (it does not rely on expensive equipment). Zhang et al.\textsuperscript{64} developed a nucleic acid test strip method to detect \textit{Bursaphelenchus xylophilus}, and proved that the method can be applied in the rapid identification of \textit{B. xylophilus} in entry-exit inspection and quarantine. The test strip technique has also been reported in other fields of biology\textsuperscript{65–68}.

The author's research group has established a sophisticated database of the DNA barcode of TCM plants. More and more SNP(s) have been discovered for identification. Using specific-primer isothermal PCR combined with nucleic acid test strip technique, we can design specific rapid detection kits for some TCM materials, allowing the direct detection of amplification results without electrophoresis, PCR, and other expensive equipment. Therefore, fast on-site detection can be conducted.

### 2.1.3. Information platform for geographic traceability of TCM

Chinese herbal medicine traceability technology permits circulation information on herbal medicines to be recorded and traced\textsuperscript{69}. This technology ensures the safe use of herbal medicines. Barcode technology is presently the main technology for traceability. A barcode is a set of graphics arranged by certain encoding rules for storing information, and it can be divided into 1D and 2D barcodes. Each code system has its own specific character set and validation functions\textsuperscript{70}. Compared with the 1D barcode, the 2D barcode is extensively used in many fields because it can store large data and encode numbers, letters, and characters. The 2D barcode systems have become popular for media, traceable security system, business cards, social networking, marketing, and electronic payments in China\textsuperscript{71,72}. Several studies have applied the 2D barcode in food traceability\textsuperscript{70,73–75}. 2D barcode not only can be used for the origin traceability of food and herbal medicine, but also can be used in tracking entire production progress\textsuperscript{76}. Yan et al.\textsuperscript{77} applied the 2D barcode for GAP production progress of herbal medicine, and developed 2D barcode-based GAP production patterns. Jin et al.\textsuperscript{78} provided a new method of medicinal slice warehousing by 2D barcode-based medicinal slice logistics management. Liu et al.\textsuperscript{79} successfully converted DNA barcoding information into 2D barcodes. We have developed an automated process of DNA barcode sequences by converting them into colorful 1D barcodes and 2D barcodes. Users can conveniently use mobile terminals, such as mobile phones, to obtain DNA barcode sequences by scanning barcode images, and submit the sequences to the world's largest DNA barcode database of TCM (http://www.tcmbarcode.cn/en/) for analysis. Barcode traceability technology is often combined with databases and networks. The barcode acts as a carrier for information transfer and network as a bridge in information flow, and the database acts as a warehouse for traceable information storage. Information of each phase of circulation is stored in a database through the network and converted into a 2D barcode, whereas traceability information can be obtained by scanning a 2D barcode image and searching the database. This combination is convenient for recording and management of information, and can achieve rapid transmission and information retrieval. Considering the popularity of smartphones, 2D barcode traceability technology no longer needs to rely on a specific barcode reading machines or software. The advantages and disadvantages of these fast identification methods were listed in (Table 1).

### 2.2. Rapid detection technology of mycotoxins

To ensure the quality, safety and efficacy of different products, various analytical techniques have been applied for the detection of mycotoxins in foodstuffs, medicinal plants and their derivative products. Conventional analytical methods of mycotoxin detection involve chromatographic analyses, such as TLC, HPLC, GC, and more recently, techniques such as LC/MS and GC/MS. Most of these methods have high sensitivity, and they were developed for quantitative and qualitative analyses of mycotoxins. To monitor and control the contamination of mycotoxins, rapid detection technology has become the research focus because of its relative simplicity, convenience, accuracy, and efficiency.

#### 2.2.1. Gold immunochromatographic assay (GICA)

GICA is a solid-phase marker immunoassay technique that combines colloidal gold labeling technology and immunoassay, with chromatography analysis technology. The technique not only has the characteristics of good stability and low-cost but also has intuitive and reliable results, suitable for semi-quantitative and quantitative rapid detection of mycotoxins. Nowadays, gold nanoparticles have been extensively employed as immobilizing different biological receptors, e.g., enzyme, DNA, antigen/antibody and other biomolecules. Wang et al.\textsuperscript{80} found that colloidal gold immunochromatographic dual strip can rapidly and accurately detect samples containing zearalenone and fumonisin B1. The limit of detection (LOD) for fumonisin B1 is 1.0 ng/mL. Multiple testing based on immune colloidal gold test strip has become a new trend for the detection of mycotoxins by immunological methods\textsuperscript{81}.

GICA is an ideal selection in biotechnological systems because of its inherent advantages (e.g., easy preparation and good biocompatibility). However, owing to the limitation of antigenic epitope on the small biomolecules and a narrow linear range, innovative and powerful techniques are being developed for the amplification of detectable signal. The present limitations of GICA include the small size of the biomolecule as epitope and narrow linear range of assays.

#### 2.2.2. Biochip technology

Biochip is a new technology developed through life science and microelectronics in recent years. It has incomparable advantages compared with conventional methods including high-throughput, multi-parameter synchronization analysis; fully automatic, rapid analysis; high accuracy and sensitive analysis. In antibody chip technology, various mycotoxins and other hazardous chemicals can be monitored simultaneously, thereby greatly reducing the time of sample extraction and detection and improving efficiency. Wang et al.\textsuperscript{82} designed an immunochip to simultaneously quantify the contents of AFB1, AFM1, DON, OTA, T-2 toxin, and ZON and the present LODs majorly showed relatively lower.

Although biochip technology has a lot of advantages in advanced optical biosensors for sensing analytes, this technique is expensive in analytical cost, and requires complex labeling process and professional technicians with specialized equipment.
2.2.3. Biosensor technology

Biosensor technology, a prominent technique for preliminary screening of the toxicity of samples, can be defined as having a sensing element for the selective detection of a target and method to transduce the interaction as a measurable signal. The biosensor’s high sensitivity, selectivity, low cost, simplicity, miniaturization, portability and integration in automated devices make it a reliable and usable alternative for monitoring mycotoxins.

Numerous types of biosensors have been developed. Sapsford et al. detected the content of AFB$_1$ employed by Array Biosensor, and reported a low LOD of 0.3 ng/mL. Wang et al. presented a novel suspension array technology for quantifying AFB$_1$, DON, T-2 toxin and ZEN simultaneously and quantitatively in corn and peanut, and the levels of LOD were better than those obtained using HPLC. Yuan et al. designed a surface plasmon resonance biosensor to detect OTA directly, and reported a dramatically improved LOD of 0.042 ng/mL.

In conclusion, these biosensors can achieve the sensitivity and selectivity required for the very strict regulatory limits from legislation. Nowadays, its stability, reproducibility and life-time coupled with the differences in tested samples, less species and instability of antibody, many studies on the biosensors are still at the experimental stage.

2.2.4. Aptamer identification technology

Aptamers, single-stranded oligonucleotides of DNA or RNA sequences, can specifically bind to target molecules in a complex matrix. Aptamer technologies have been widely applied in the analysis of mycotoxins. Yang et al. presented the colorimetric detection of OTA using OTA’s aptamer and unmodified GNP. In addition, Wang et al. developed a new type of structure-switching aptasensor to simultaneously detect OTA and FB$_1$.

Aptamers have the ability under certain physicochemical conditions to fold into defined three-dimensional conformations, which facilitate specific interaction with target molecules having high affinity constants. Based on novel, molecular recognition, aptamer identification technology has great potential for application in the rapid detection of mycotoxins because of high accuracy, precision and specificity. The combination of aptamer recognition techniques and novel nano-materials has been used in various optical and electrochemical analytical methods for mycotoxin analysis.

Consequently, aptamers have emerged owing to inherent advantages compared with antibodies, such as non-requirement of immunization from animals, more chemical and thermal stability, and less variability. Particularly, aptamers are not susceptible to denaturation in the presence of solvents commonly used in the extraction of mycotoxins.

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Table 1 Comparison of advantages and disadvantages of different detection methods.

| Identification method | Advantage | Disadvantage | Scope of application in TCM |
|-----------------------|-----------|--------------|-----------------------------|
| DNA barcoding         | Sequence-based | Require advanced equipment | All medicinal plants and animals |
|                       | Very high accuracy |                  |                              |
|                       | Universal | Need comparison in database |                              |
| Microarray in a tube  | SNP detection | Need specific primer | Some closely related species |
|                       | On-site detection | Not universal |                              |
|                       | Not require advanced equipment |                  |                              |
|                       | After method have been established, single test cost lower than CNY 2.00 |                  |                              |
|                       | Database-free |                  |                              |
| Nano-Au               | High accuracy | Need specific primer | Some closely related species |
|                       | SNP detection | Not universal |                              |
|                       | On-site detection |                  |                              |
|                       | Not require advanced equipment |                  |                              |
|                       | Low cost |                  |                              |
|                       | Database-free |                  |                              |
|                       | Can be developed into a species specific kit |                  |                              |
| Nucleic acid amplification test strip | High accuracy | Need specific primer | Some closely related species |
|                       | SNP detection | Not universal |                              |
|                       | On-site detection |                  |                              |
|                       | Not require advanced equipment |                  |                              |
|                       | Low cost |                  |                              |
|                       | Database-free |                  |                              |
|                       | Can be developed into a species specific kit |                  |                              |
| Traceability system   | High accuracy | Need database | All medicinal plants and animals |
|                       | High universality |                  |                              |
|                       | Very rapid |                  |                              |
|                       | Not require advanced equipment |                  |                              |
|                       | Low cost |                  |                              |

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2.3. Rapid detection technology of pesticide residues

Classical instrumental analytical techniques for the determination of pesticides in a large number of samples have been developed. These methods involve GC and LC coupled with various detectors, GC-MS, GC-MS/MS, or LC-MS. In recent years, GC-MS/MS has been intensively used for the determination of pesticides. Although chromatography-based methods are sensitive and reliable, they require sophisticated equipment, skilled analysts, and time-consuming sample preparation steps. Moreover, organic solvents used in the detection process may lead to environmental pollution.

3. Conclusions and prospects

The quality control of medicinal materials has always been a weak link in the Chinese medicine industry, and it has affected the sustainable development of the TCM industry. Low-cost, on-site fast detection of TCM with the capability of easy operation and large-scale implementation is an urgent must, which depends on not only the availability of portable equipment, but also simplifying the techniques and promoting new detection principles. The future development of the rapid testing of Chinese medicine have three goals: (1) exploring mobile species identification systems and developing kits for SNP detection based on GNPs, microarray-in-a-tube, and nucleic acid amplification test strip; (2) establishing a fast detection system for mycotoxins and pesticide residues of Chinese medicine; (3) establishing a TCM traceability system in China, using some specific kits unlimited from the environment and infrastructure, which can satisfy the requirements for fast detection in remote and undeveloped areas.

3.1. Setting up mobile species identification system (2D DNA barcoding)

Given the rapid development in DNA barcoding, all species are expected to have a unique DNA barcode in the future. Hence, the technique of DNA barcoding has enormous potential applications. The application of handheld rapid species identification equipment will bring great convenience in entry-exit inspection and quarantine work. Compared with traditional DNA sequences (about 200–600 bp) the 2D code has larger information capacity and adjustable size, which is suitable for direct labeling of samples. The 2D code contains more DNA information. Moreover, it will be easier to operate and be more convenient to collect and conduct remote authentication for 2D code using a communication terminal camera with scanner function and wireless communication function. A species identification system within a mobile application will make authentication more convenient and rapid. The DNA barcode for species identification is a global method that can improve the identification of TCM to form a unified international standard. It can greatly improve the identification level of Chinese medicinal materials by establishing a Chinese herbal medicine DNA barcode database. Moreover, this global method can speed up the process of modernization of TCM, and provide a scientific basis for the control of raw TCM materials. This method is of great significance to the internationalization of the Chinese herbal medicine identification standard.

3.2. Setting up a fast detection system of mycotoxin and pesticide residue in the production of TCM

The design of a rapid detection platform is the primary problem in the quality control of TCM. To set up a fast detection system of mycotoxins and pesticide residues in the production of TCM, the management of TCM supervision departments and other relevant units should select different methods for analyzing various mycotoxins according to their detection technology and detection means to control the quality and safety of TCM. Future research should focus on the identification and quantification of AFs (AFG2, AFG1, AFB2, AFB1), OTA, DON, ZEN, fumonisins (FB1, FB2) and citrinin. According to their detection technology and detection means, the selection of different analytical methods of pesticide residues for the control of the quality and safety of TCM should meet the social demand for rapid identification of medicinal species. Analytical methods must meet the drug test, customs, security functions, and companies’ demands for rapid identification of medicinal species, which will ensure the safety and efficacy of TCM.
3.3. Setting up the traceability of TCM

Blind transplanting of medicinal materials has led to the decline in medicinal quality and counterfeit medicines in production or sales, which may increase the risk of accidents in TCM102. Establishing herb medicine traceability can not only protect the authenticity of a drug, but also manage the information of seed, planting, processing, and marketing. A traceable system has great application potential in the Chinese herb medicine industry. In consideration of cost and rapidity, barcode traceability can be considered a main technology tool. Once safety problems occur, potential problems can be traced and further characterized by molecular biology or fingerprinting.

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References

1. Ng KY, Cheng CL, Xu HX. Safety issues of Chinese medicine: a review of intoxication cases in Hong Kong. Chin Herb Med 2009;1:29–39.
2. Dickens P, Tai YT, But PPH, Tomlinson B, Ng HK, Yan KW. Fatal accidental aconite poisoning following ingestion of Chinese herbal medicine: a report of two cases. Forensic Sci Int 1994;67:55–8.
3. Chan TY. Incidence of herb-induced aconite poisoning in Hong Kong: impact of publicity measures to promote awareness among the herbalists and the public. Drug Saf 2002;25:823–8.
4. Chan TY. Aconite poisoning: a global perspective. Vet Hum Toxicol 1994;36:326–8.
5. Deng JF, Lin TJ, Kao WF, Chen SS. The difficulty in handling poisonings associated with Chinese traditional medicine: a poison control center experience for 1991–1993. Vet Hum Toxicol 1997;39:106–14.
6. Poon WT, Lai CK, Ching CK, Tse KY, So YC, Chan YC, et al. Aconite poisoning in camouflag. Hong Kong Med J 2006;12:456–9.
7. Lo SHK, Mo KL, Wong KS, Poon SP, Chan CK, Lai CK, et al. Aristolochic acid nephropathy complicating a patient with focal segmental glomerulosclerosis. Nephrol Dial Transplant 2004;19:1913–5.
8. Kumana CR, Ng M, Lin HJ, Ko W, Wu PC, Todd D. Hepatic veno-occlusive disease due to toxic alkaloid in herbal tea. Lancet 1983;322:1360–1.
9. Yang MH. Research progress in fungi and mycotoxin infection of medicinal plants and their products. Guizhou Agric Sci 2008;36:59–63.
10. Yang MH, Chen JM, Zhang XH. Immunoaffinity column clean-up and liquid chromatography with post-column derivatization for analysis of aflatoxins in traditional Chinese medicine. Chromatographia 2005;62:499–504.
11. Liu SY, Qiu F, Yang MH. Determination of aflatoxins in nelumbinis semen by immunoaffinity column clean-up and HPLC-FLD with online post-column photochemical derivatization and LC-MS/MS confirmation. China J Chin Mater Med 2012;37:305–9.
12. Wei RW, Yang XL, Qiu F, Yang MH, Qin JP. Simultaneous determination of aflatoxin B1, B2, G1, G2 and ochratoxin A in Glycyrrhiza uralensis analyzed by HPLC-MS/MS. Food Control 2013;32:216–21.
13. Yang L, Wang L, Pan J, Xiang L, Yang M, Logrieco AF. Determination of ochratoxin A in traditional Chinese medicinal plants by HPLC-FLD. Food Addit Contam Part A Chem Anal Control Expo Risk Assess 2010;27:989–97.
14. Yue YT, Zhang XF, Pan JY, Zhen Y, Wu J, Yang MH. Determination of deoxynivalenol in medicinal herbs and related products by GC-ECD and confirmation by GC-MS. Chromatographia 2010;71:533–8.
15. Kong WJ, Li JY, Qiu F, Wei JH, Xiao XH, Zheng Y, et al. Development of a sensitive and reliable high performance liquid chromatography method with fluorescence detection for high-throughput analysis of multi-class mycotoxins in Coix seed. Anal Chim Acta 2013;799:68–76.
16. Pietri A, Rastelli S, Bertuzzi T. Ochratoxin A and aflatoxins in liquorice products. Toxins 2010;2:758–70.
17. Herrera M, Herrera A, Ariño A. Estimation of dietary intake of ochratoxin A from liquorice confectionery. Food Chem Toxicol 2009;47:2002–6.
18. Ariño A, Herrera M, Estopañan G, Juan T. High levels of ochratoxin A in licorice and derived products. Int J Food Microbiol 2007;114:366–9.
19. Rong WG, Guo H, Yang H. Current research status in China on pesticide contamination of plant material used in making Chinese herbal medicines. Agrochemicals 2006;45:302–8.
20. Miao Q, Kong WJ, Wei JH, Yang SH, Yang MH. Analysis and effective control of pesticides residues in traditional Chinese medicine. Chin J Pestic Sci 2012;14:63–70.
21. Hebert PD, Cywinska A, Ball SL, deWaard JR. Biological identifications through DNA barcodes. Proc Biol Sci 2003;270:313–21.
22. Schindel DE, Miller SE. DNA barcoding a useful tool for taxonomists. Nature 2005;435:17.
23. Miller SE. DNA barcoding and the renaissance of taxonomy. Proc Natl Acad Sci USA 2007;104:4775–6.
24. Chen SL, Pang XL, Song JY, Shi LC, Yao H, Han JP, et al. A renaissance in herbal medicine identification: from morphology to DNA. Biotechnol Adv 2014;32:1237–44.
25. Chen SL, Yao H, Han JP, Liu C, Song JY, Shi LC, et al. Validation of the ITS2 region as a novel DNA barcode for identifying medicinal plant species. PLoS One 2010;5:e8613.
26. Pang XH, Song JY, Zhu YJ, Xie CX, Chen SL. Using DNA barcoding to identify species within Euphorbiaceae. Planta Med 2010;76:1784–6.
27. Han JP, Shi LC, Li MH, Yao H, Song JY, Xu HX, et al. Relationship between DNA Barcoding and chemical classification of Salvia L. medicinal herbs. Planta Med 2009;75:416.
28. Gao T, Yao H, Song JY, Liu C, Zhu YJ, Ma XY, et al. Identification of medicinal plants in the family Fabaceae using a potential DNA barcode ITS2. J Ethnopharmacol 2010;130:116–21.
29. Luo K, Chen SL, Chen KL, Song JY, Yao H, Ma XY, et al. Assessment of candidate plant DNA barcodes using the Rutaceae family. Sci China Life Sci 2010;53:701–8.
30. Pang XH, Song JY, Zhu YJ, Xu HX, Huang LF, Chen SL. Applying plant DNA barcodes for Rosaceae species identification. Cladistics 2011;27:165–70.
31. Han JP, Shi LC, Chen XC, Lin YL. Comparison of four DNA barcodes in identifying certain medicinal plants of Lamiaceae. J Syst Evol 2012;50:227–34.
32. Gao T, Yao H, Song JY, Zhu YJ, Liu C, Chen SL. Evaluating the feasibility of using candidate DNA barcodes in discriminating species of the large Asteraceae family. BMC Evol Biol 2010;10:324.
33. Yao H, Song JY, Liu C, Luo K, Han JP, Li Y, et al. Use of ITS2 region as the universal DNA barcode for plants and animals. PLoS One 2010;5:e13102.
34. Han JP, Zhu YJ, Chen XC, Liao BS, Yao H, Song JY, et al. The short ITS2 sequence serves as an efficient taxonomic sequence tag in
comparison with the full-length ITS. Biomed Res Int 2013;2013:741476.
36. Wolf M, Chen SL, Song JY, Ankenbrand M, Müller T. Compensatory base changes in ITS2 secondary structures correlate with the biological species concept despite intragenomic variability in ITS2 sequences—a proof of concept. PLoS One 2013;8:e66726.
37. Carles M, Cheung MKL, Moganti S, Dong TSM, Tsim KW, Ip NY, et al. A DNA microarray for the authentication of toxic traditional Chinese medicinal plants. Planta Med 2005;71:580–4.
38. Tsai PY, Woo HS, Wong MS, Chen SL, Fong WF, Xiao PG, et al. Genotyping and species identification of Fritillaria by DNA chips. Acta Pharm Sin 2003;38:185–90.
39. Che J, Tang L, Liu Y, He W, Chen F. Molecular identity of Crocus sativus and its misused substitutes by ITS sequence. China J Chin Mater Med 2007;32:668–71.
40. Li XW, Yang Y, Henry RJ, Rossetto M, Wang YT, Chen SL. Plant DNA barcoding: from gene to genome. Nucleic Acids Res 2005;33:7122–31.
41. Li QS, Li Y, Song JY, Pang XH, Han JP, Chen SL. A fast SNP identification and analysis of intraspecific variation in the medicinal Panax species based on DNA barcoding. Gene 2013;530:39–43.
42. Liao BS, Han JP, Chen XC, Chen SL, inventors; Chinese Academy of Medical Sciences Institute of Medicinal Plant Development, assignee. Method for rapid identification of Ophiocordyceps Sinensis. China Patent CN102888456A. 2013 Jan 23.
43. Lu ZH, Liu QJ, Wang H, inventors; Southeast University, assignee. Flushing-free PCR amplification tube capable of directly detecting gene. China patent CN1448500. 2003 Oct 15.
44. Liu QJ. Development of three novel integrated DNA microarrays and detection instruments [dissertation]. Nanjing: Southeast University; 2006.
45. Wang H, Li J, Liu HP, Liu QJ, Mei Q, Wang YJ, et al. Label-free hybridization detection of a single nucleotide mismatch by immobilization of molecular beacons on an agarose film. Nucleic Acids Res 2002;30:e61.
46. Liu QJ, Bai YF, Ge QY, Zhou SX, Wen T, Lu ZH. Microarray-in-a-tube for detection of multiple viruses. Clin Chem 2007;53:188–94.
47. Zhang JF, Sun LP, Li H, Wang XY, Zhang QQ. Gold nanoparticles application in gene mutation detection and SNP analysis. Chin J Biochem Mol Biol 2008;24:489–95.
48. Bao YP, Huber M, Wei TF, Marla SS, Storhoff JJ, Müller UR. SNP identification in amplified human genomic DNA with gold nanoparticle probes. Nucleic Acids Res 2005;33:e15.
49. Charrier A, Candoni N, Liachenko N, Thibaudau F, 2D aggregation and selective desorption of nanoparticle probes: a new method to probe DNA mismatches and damages. Biosens Bioelectron 2007;22:1881–6.
50. Li JS, Chu X, Liu YL, Jiang JH, He ZM, Zhang ZW, et al. A colorimetric method for point mutation detection using high-fidelity DNA ligase. Nucleic Acids Res 2005;33:e168.
51. Pang LL, Li JS, Jiang JH, Shen GL, Yu RQ. DNA point mutation detection based on DNA ligase reaction and nano-Au amplification: a piezoelectric approach. Anal Biochem 2006;358:99–103.
52. Zhao WA, Chuanman W, Brook MA, Li YF. Simple and rapid colorimetric biosensors based on DNA aptamer and noncrosslinking gold nanoparticle aggregation. ChemBioChem 2007;8:727–31.
53. Elghanian R, Storhoff JJ, Mucic RC, Letsinger RL, Mirkin CA. Selective colorimetric detection of polynucleotides based on the distance-dependent optical properties of gold nanoparticles. Science 1997;277:1078–81.
54. Mao X, Xu H, Zeng QX, Zeng LW, Liu GD. Molecular beacon-functionalized gold nanoparticles as probes in dry-reagent strip biosensor for DNA analysis. Chem Commun 2009;21:3065–7.
89. Wu SJ, Duan N, Ma XY, Xia Y, Wang HX, Wang ZP, et al. Application of ePS platform in logistics management of small package of TCM decoction pieces storehouse. *China Pharm* 2013;24:271–2.

90. Liu C, Shi LC, Xu X, Li H, Xing H, Liang D, et al. DNA barcode goes two-dimensions: DNA QR code web server. *PLoS One* 2012;7:e35146.

91. Miao Q, Kong WJ, Yang SH, Yang MH. Comparison of sample preparation methods combined with gas chromatography with electron-capture detection for the analysis of multipesticide residues in lotus seeds. *J Sep Sci* 2013;36:2010–9.

92. Liu QZ, Kong WJ, Qiu F, Wei JH, Yang SH, Zheng YG, et al. One-step extraction for gas chromatography with flame photometric detection of 18 organophosphorus pesticides in Chinese medicine health wines. *J Chromatogr B* 2012;885–886:90–6.

93. Hu YC, Wan L, Zhang JM, Yang F, Cao JL. Rapid determination of pesticide residues in Chinese materia medica using QuEChERS sample preparation followed by gas chromatography–mass spectrometry. *Acta Pharm Sin B* 2012;2:286–93.

94. Lee KG, Jo EK. Multiresidue pesticide analysis in Korean ginseng by gas chromatography-triple quadrupole tandem mass spectrometry. *Food Chem* 2012;134:2497–503.

95. Chen LN, Song FR, Liu ZQ, Zheng Z, Xing JP, Liu SY. Multi-residue method for fast determination of pesticide residues in plants used in traditional Chinese medicine by ultra-high-performance liquid chromatography coupled to tandem mass spectrometry. *J Chromatogr A* 2012;125:132–40.

96. Pareja L, Fernández-Alba AR, Cesio V, Heinzen H. Analytical methods for pesticide residues in rice. *TrAC, Trends Anal Chem* 2011;30:270–91.

97. Wilkowska A, Biziuk M. Determination of pesticide residues in food matrices using the QuEChERS methodology. *Food Chem* 2011;125:803–12.

98. Zhang Q, Sun Q, Hu BS, Shen Q, Yang G, Liang X, et al. Development of a sensitive ELISA for the analysis of the organophosphorus insecticide fenithion in fruit samples. *Food Chem* 2008;106:1278–84.

99. Marty JL, Garcia D, Rouillon R. Biosensors: potential in pesticide detection. *TrAC, Trends Anal Chem* 1995;14:329–33.

100. Ferentinos KP, Yialouris CP, Blouchos P, Moschopoulou G, Kintziou S. Pesticide residue screening using a novel artificial neural network combined with a bioelectric cellular biosensor. *BioMed Res Int* 2013;2013:815319.

101. Qie ZW, Ning BA, Liu M, Bai JL, Peng Y, Song N, et al. Fast detection of atrazine in corn using thermometric biosensors. *Analyst* 2013;138:5151–6.

102. Hu SL. *Genuine traditional Chinese medicine*. Harbin: Heilongjiang Science and Technology Press; 1989.