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Itm2a, a Target Gene of GATA-3, Plays a Minimal Role in Regulating the Development and Function of T Cells

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Abstract

The integral membrane protein 2a (Itm2a) is one of the BRICHOS domain-containing proteins and is structurally related to Itm2b and Itm2c. It is expressed preferentially in the T lineage among hematopoietic cells and is induced by MHC-mediated positive selection. However, its transcriptional regulation and function are poorly understood. Here we showed Itm2a to be a target gene of GATA-3, a T cell-specific transcription factor. Deficiency of Itm2a had little impact on the development and function of polygonal T cells but resulted in a partial defect in the development of thymocytes bearing a MHC class I-restricted TCR, OT-I. In addition, Itm2a-deficient mice displayed an attenuated T helper cell-dependent immune response in vivo. We further demonstrated that Itm2b but not Itm2c was also expressed in T cells, and was induced upon activation, albeit following a kinetic different from that of Itm2a. Thus, functional redundancy between Itm2a and Itm2b may explain the minimal phenotype of Itm2a deficiency.

Introduction

GATA-3 is the third member of the GATA family of transcription factors [1]. Among hematopoietic cells, GATA-3 is expressed nearly exclusively in the T lineage, and is critical for the development of CD4+ T helper (Th) cells [2], the differentiation of Th helper type 2 (Th2) cells [3,4], the function of regulatory T (Treg) cells [5,6], and the activation/homeostasis of CD8+ cytolytic T (Tc) cells [7,8]. However, it is still not fully understood how GATA-3 executes its function. Several GATA-3 targets genes have been identified. For example, GATA-3 directly regulates the expression of Th-POK (encoded by Zbtb7b) [9], a transcription factor that is also essential for the development of CD4+ T cells [10,11]. However, reconstitution of Th-POK was unable to rescue the development of GATA-3-deficient CD4+ T cells, suggesting the presence of additional GATA-3 target genes in mediating its function.

Itm2a is a type II transmembrane protein and contains 263 amino acid residues [12]. It is highly conserved between human and mouse. Itm2a belongs to the Type II Integral Membrane protein (Itm) family, which also includes Itm2b and Itm2c [13]. Members of this protein family contain a BRICHOS domain, which possesses chaperone activity [14], a transmembrane domain at the N-terminus, and a furin cleavage site at the C-terminus. Among the Itm family members, Itm2b is the most extensively studied protein. The ectodomain of Itm2b can undergo at least three steps of protease-mediated cleavage [15,16]. The first cleavage occurs at the C-terminus and is mediated by furin. The remaining Itm2b protein can be further cleaved by ADAM10 protease, releasing the C-terminal half, including the BRICHOS domain, into extracellular compartments. The remaining N-terminal half of Itm2b can then undergo regulated intramembrane proteolysis mediated by signal peptide peptidase (SPP) 2a and 2b. This intramembrane proteolytic event yields an intracellular domain (ICD) and a secreted low molecular weight peptide. Two types of mutation of Itm2b have been linked to familial British Dementia (FBD) and familial Danish Dementia (FDD) [17,18]. Both mutations result in the addition of 10 amino acid residues to the furin-cleaved C-terminal peptide, which accumulates in CNS forming the pathogenic plaques found in these patients. However, how the addition of 10 amino acids leads to the development of dementia is still unclear. Given the high degree of similarity in protein structure among itm2 proteins, it is very likely that such protease-mediated cleavages also occur on Itm2a. Evidence supporting this notion however is still lacking.

Like GATA-3, Itm2a is also preferentially expressed in cells of the T lineage among hematopoietic cells according to the Immgen database [http://www.immgen.org]. Thus far, there are only a handful of published studies providing clues regarding the role of Itm2a in T cells. Itm2a was identified as a gene that was induced by MHC-mediated positive selection of double positive (DP) thymocytes [19]. Peripheral naive Th and Tc cells expressed a low level of Itm2a, which was transiently induced when T cells were stimulated with concanavalin A. Three recent transcription profile studies independently found that the expression of Itm2a was significantly lower in Treg cells than in effector Th cells [20-22].
Marson et al further identified Itm2a as one of the most down-regulated genes in Treg cells and suggested that its expression was suppressed by Foxp3, the master transcription factor of Treg cells [21]. In addition, Itm2a was one of approximately 15 genes including CD3, Lck, and TCRβ, preferentially expressed in pediatric T-ALL [23]. Lately, Itm2a was linked to Graves’ disease in a staged genome-wide association study [24], further strengthening its role in the immune system.

In this report, we show that the expression of Itm2a was reduced in GATA-3-deficient thymocytes and that Itm2a is transactivated by GATA-3. Although polyclonal thymocytes of Itm2a-deficient mice develop normally, Itm2a-deficient mice display a partial defect in the development of thymocytes bearing the MHC class I-restricted OT-I TCR but not the MHC class II-restricted OT-II TCR. Surprisingly, deficiency of Itm2a has little impact on the differentiation, activation, and function of T cells in vitro. However, Itm2a KO mice are not as efficient as wild type mice in mounting humoral immune responses against a model Th cell-dependent antigen. We further show that Itm2b was also expressed in T cells and very likely compensated for the loss of Itm2a.

### Materials and Methods

#### Mice

The targeting construct of Itm2a was created through a recombineering-based method. The detailed strategy will be provided upon request. The targeting construct was then introduced into 129X1b hybrid ES cells by inGenious Targeting Laboratory (Stony Brook, NY) to create floxed Itm2a mice. The Itm2a gene in the floxed allele was then deleted with ELIIa-cre (The Jackson Laboratories) to create Itm2a-deficient mice, which were backcrossed to C57BL/6 mice for five generations before use. OT-I and OT-II mice (The Jackson Laboratories) to create Itm2a-deficient mice. Male or female mice aged 4–12 weeks were used. In all experiments, littermates were used as controls.

#### Ethics statement

All animals were housed under specific pathogen-free conditions, and experiments were performed in accordance with the institutional guidelines for animal care under a protocol (#07-017) approved by the Institutional Animal Care and Use Committee (IACUC) of Dana-Farber Cancer Institute.

#### Activation of T cells and in vitro differentiation of Th cells

Total cells were collected from spleens and activated with plate bound anti-CD3 (ranging from 0.5 μg/ml to 1 μg/ml), OT-II peptide (OVA 323-339), or OT-I peptide (SIINFEKL, OVA 257-264) at different concentrations. After 3 days, cells were collected and stained with surface markers. The supernatant was collected and used for cytokine production by ELISA. For in vitro differentiation of Th cells, CD4+ T cells were purified from spleens and lymph nodes by magnetic cell sorting (Miltenyi Biotec). The cells were stimulated with 1 μg/ml plate-bound anti-CD3 and 2 μg/ml soluble anti-CD28 under Th1 (3 ng/ml IL-12 plus 10 μg/ml anti-IL-4) or Th2 (10 ng/ml IL-4 plus 10 μg/ml anti-IFN-γ) conditions. For Th17 differentiation, total CD4+ cells were cultured in the presence of WT irradiated splenocytes (1:3 ratio) and 3 ng/ml TGF-β1 and 20 ng/ml IL-6. Recombinant human IL-2 (100 U/ml) was added after 24 h, and the cells were expanded in complete medium containing IL-2. On day 7, the cells were restimulated with 50 ng/ml PMA and 1 mM ionomycin. The cytokine production was examined by intracellular cytokine staining.
performed using the Brilliant SYBR Green QPCR kit according to the manufacturer’s protocol (Stratagene) on a MX-3000P apparatus (Stratagene) using the following cycling conditions: denaturation at 95°C for 30 s, annealing at 56°C for 60 s, and extension at 72°C for 30 s. Primer sets were designed using the Primer3 web utility. Levels of mRNA were adjusted for differences in actb (encoding beta-actin) expression. The following primers are used: Itm2a 5’-CGCAGCTGTCGAGCTCAATAT-3’ and 5’-CATCTCCAGATGAGCCCATCC-3’; Itm2b 5’-GAGGTTGACGTTCACACTCGG-3’ and 5’-CTCTGTCCAAACCG-GAACCAC-3’; Itm2c 5’-GTCGCCATCAAGGCTGATAAA-3’ and 5’-AGGCCCATGAGCATTACCC-3’; actb 5’-GGCTGTATTCCCCTCCATCG-3’ and 5’-CCAGTTGGTAACATGCCATGT-3’.

ELISA

Sandwich ELISA was performed using the following monoclonal antibody pairs (BD Biosciences): anti-IL-2 (JES6-1A12)/

Figure 2. Deficiency of Itm2a has little impact on the differentiation of immune cells. A. Generation of Itm2a-deficient mice. Schematic diagrams of Itm2a gene (top), the targeting construct (middle), and deleted allele (bottom) are shown. Exons are represented with gray boxes and numbered. Regions for homologous recombination are marked with dashed lines. Neo and TK stand for neomycin cassette and thymidine kinase. B. T cells were purified from control (WT) and two Itm2aKO mice and stimulated with anti-CD3 for 72 hours. Cell extract from the stimulated cells was probed with anti-Itm2a and anti-Hsp90. C. Thymocytes of WT and Itm2aKO mice were enumerated, and the means and standard deviations are shown in the left panel. D & E. The thymocytes were also stained with anti-CD4/anti-CD8 (D) and anti-α/β TCR, and TCR<sup>high</sup> cells were further stained with antibodies against various V<sub>β</sub> chains indicated in E. F. The thymocytes were stained with anti-CD4/anti-Foxp3 and anti-TCR/NK1. G. Splenocytes and lymph node cells of WT and Itm2aKO mice were stained with anti-CD4 and anti-CD8. H. The splenocytes were also stained with anti-TCR/anti-B220. B220<sup>+</sup> cells were then further separated by anti-IgM/anti-IgD staining. The data shown in D–H are representative FACS plots of at least three independent experiments.

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biotin-anti–IL-2 (JES6-5H4), and anti–IFN-γ (R4-6A2)/biotin-anti–IFN-γ (XMG1.2).

Plasmid, transfection, and luciferase assay

An approximately 2 kb genomic fragment containing +28 to -1964 in relation to the transcriptional start site of the murine Itm2a gene was amplified and cloned into the BglII site of the PGL2-Basic (Promega) to create Itm2a-Luc. The IL-13-Luc and pcDNA3-GATA3 were previous reported [25]. In all luciferase assays, murine M12 B cells were transfected with 10 μg of luciferase reporter, 5 μg of pcDNA3 vector, and 5 μg pTK-Renilla with BIORAD GENE PULSER II set at 280 V and 0.975 F. Luciferase activity was determined in duplicate with Dual-Luciferase Reporter Assay System (Promega, Madison, WI). The firefly luciferase activity obtained from each sample was normalized against the renilla luciferase activity from the same sample.

CTL killing

CTL killing assay was adapted from a published protocol [26]. Briefly, total splenocytes from WT and KO mice were cultured with irradiated splenocytes from Balb/c (H-2d) mice for 7 days and viable cells were collected as effector cells. EL4 (H-2b) or P815 (H-2d) target cells were label with CellTrack Far Red DDAO-SE (Molecular Probes Eugene, OR) at 37°C for 10 min. Effector and target cells (1 × 10⁶) were mix at different ratio and incubated at 37°C, 5% CO2 in a humidified incubator for 4 hours. Cells were collected, fixed and stained with anti-activated caspase-3 antibody (BD Bioscience CA).

In vivo immunization and Th recall response

Mice were immunized with 20 μg trinitrophenyl-conjugated keyhole limpet hemocyanin KLH (TNP-KLH) (Biosearch Technologies, Novato, CA) emulsified in complete Freund adjuvant (CFA) through a subcutaneous injection. Immunized mice were sacrificed two weeks later. For recall response, 1 × 10⁶ lymph node cells were stimulated with 20 μg/ml TNP-KLH for 3 days,
Generation of Itm2a-deficient mice

The expression of Itm2a is induced in activated T cells [19]. To investigate the function of Itm2a in T cells, we transduced activated Th cells with a retrovirus expressing Itm2a. However, we found that overexpression of Itm2a had little impact on the activation, survival, and cytokine production of Th cells (data not shown). Therefore we set out to generate Itm2a-deficient (Itm2aKO) mice. We inserted a neomycin cassette and two loxp sites into the Itm2a allele (Figure 2A). We then deleted the Itm2a gene in germline with the EII-a Cre transgene. Itm2aKO mice were born according to Mendelian ratios and had normal development and growth up to 6 months of age (data not shown). T cells obtained from Itm2aKO mice contained no detectable Itm2a protein (Figure 2B), confirming the complete deletion of the Itm2a gene. This lack of developmental defect is consistent with the results of a recent publication [27], in which an independently generated Itm2a-deficient mouse strain was studied.

T cell development is normal in Itm2aKO mice

Although Itm2a transgenic mice have a subtle aberration in the expression of CD8 [19], we found that deficiency of Itm2a had no apparent impact on the development of thymocytes. The total number of thymocytes and the percentage of CD4 single positive (SP), CD8SP and CD4–CD8– thymocytes were very comparable between WT and Itm2aKO mice (Figure 2C and 2D). In addition, the mean fluorescence intensity of CD4, CD8, HSA, TCR, and Qa2 was normal in Itm2aKO thymocytes (data not shown). We further investigated the TCR repertoire with antibodies against various Vβ chains and found no apparent difference between WT and Itm2aKO mice (Figure 2E). The numbers of Foxp3+ Treg cells and natural killer T (NKT) cells were also normal (Figure 2F). Similarly, there was no detectable difference in the number and CD4/CD8 ratio of T cells in peripheral lymphoid organs (Figure 2G). Furthermore, the development and expression of various differentiation markers of other immune cells including B cells and myeloid cells were unaltered in the absence of Itm2a (Figure 2H and data not shown).

Normal activation of Itm2aKO T cells

We then examined if loss of Itm2a would affect T cell activation. We stimulated splenocytes with escalating doses of anti-CD3 for 3 days. The induction of activation markers including CD25 and CD69, and cell recovery were comparable between WT and Itm2aKO CD4+ T cells and found no apparent defect (Figure 3A and data not shown). There was also no difference in the production of IFN-γ (Figure 3B). In addition, Itm2aKO Th cells differentiated normally into Th1, Th2, and Th17 cells in vitro (Figure 3C). We also examined CTL activity against MHC mismatched targets of Itm2aKO CD8+ T cells and found no apparent defect (Figure 3D).
Figure 5. Itm2a regulates the thymic development but not the activation of OT-I cells. A. Thymocytes of WT/OT-I and KO/OT-I mice were stained with anti-V\(\beta\)5, anti-\(\alpha\)/\(\beta\) TCR, anti-CD4, and anti-CD8, and, representative TCR/V\(\beta\)5 FACS plots are shown in the left column. The CD4/CD8 FACS plots of TCR\(^+\)V\(\beta\)5\(^+\) cells are shown in the right column. The DP, post-selected CD4\(^+\)CD8\(^{dull}\) (PS), and CD8SP populations of TCR\(^+\)V\(\beta\)5\(^+\) cells are gated. B. The numbers of DP, PS, and CD8SP V\(\beta\)5\(^+\) cells were enumerated from three pairs of mice. The means and standard deviations are shown. Statistical analysis was performed with Student's t tests. C. The histograms of CD69 and CD5 of indicated V\(\beta\)5\(^+\) thymocytes of WT/OT-I and KO/OT-I mice are overlaid and shown. D. Splenocytes and lymph nodes cells of WT/OT-I and KO/OT-I mice were stained with anti-V\(\beta\)5 and anti-CD8. Representative CD8/V\(\beta\)5 FACS plots and the absolute numbers of CD8\(^+\)V\(\beta\)5\(^+\) cells are shown. E. Splenocytes of WT/OT-I and KO/OT-I mice were stimulated with ovalbumin peptides at the indicated concentrations for 3 days. The percentages of live cells were determined based on the FSC/SSC gate and are
Taken all together, Itm2a was dispensable for the development and function of polyclonal T cells.

**Itm2a regulates the development of OT-I but not OT-II thymocytes**

To address whether Itm2a deficiency might affect the development of monoclonal thymocytes, we first crossed Itm2aKO mice with OT-II TCR transgenic mice, which bear an MHC class II-restricted TCR recognizing a peptide derived from ovalbumin. Introduction of the OT-II TCR comparably increased the percentage and number of CD4SP thymocytes in both WT and Itm2aKO mice (Figure 4A, upper row). The number of mature OT-II CD4+ thymocytes was normal in the absence of Itm2a (Figure 4A, lower row). We further stimulated WT/OT-II and Itm2aKO/OT-II splenocytes with the cognate ovalbumin peptide and found no difference in the expansion and activation of OT-II CD4+T cells (Figure 4B). Thus, deficiency of Itm2a very likely does not affect MHC class II-mediated selection and antigen-specific activation of CD4+ T cells.

We then introduced the OT-I TCR transgene, a MHC class I-restricted TCR, into Itm2aKO mice. In OT-I mice, nearly all TCR+ cells are also positive for Vb5 (Figure 5A). Interestingly, we found that deficiency of Itm2a led to an approximately 50% reduction in the number of mature Vb5+CD8+SP OT-I thymocytes (Figure 5A and 5B). One possible explanation for the reduction in the number of mature OT-I thymocytes in the absence of Itm2a is that Itm2a is required for the optimal positive selection. Upon positive selection by MHC, DP thymocytes up-regulate CD69 and CD5, and become CD4+CD8dull before transiting into SP cells. However, the induction/expression of CD69 and CD5 in DP and post-selected CD4+CD8dull Vb5+ thymocytes was very comparable between WT/OT-I and Itm2aKO/OT-I mice (Figure 5C). In addition, the absolute

![Figure 6. Impaired immune response to Th cell-dependent antigens in the absence of Itm2a.](image-url)
The induction of activation markers (Figure 5E). The cognate peptide resulted in comparable cell expansion and stimulation of WT and Itm2aKO OT-I splenocytes with the number of OT-I T cells in peripheral lymphoid organs (Figure 5D); the Itm2aKO/OT-I CD8SP thymocytes appeared to be normal based on the down-regulation of HSA and the induction of Qa2. Apoptosis of CD8SP cells because there was no apparent difference in the number was observed (Figure 5B). This data strongly suggest that deficiency of Itm2a has little impact on MHC-mediated functional compensation from Itm2b, which is also highly expressed in T cells. Thus, the combined function of Itm proteins in the adaptive immunity is rather modest probably due to functional compensation from Itm2b and/or Itm2c. A potential functional redundancy between Itm2a and Itm2b may compensate for the loss of Itm2a. We therefore compared the expression of these three genes in thymocytes and activated Th cells. We found that the transcript level of Itm2b was almost 10 times higher than that of Itm2a in thymus and activated Th cells. In contrast, the transcript of Itm2c was almost undetectable. In agreement with the published data, the protein level of Itm2a gradually increased in response to in vitro stimulation and peaked at day 2 after stimulation (Figure 7B), but quickly faded away after day 4. The level of Itm2b protein also increased in activated T cells but peaked at day 3-4. Unlike Itm2a, the level of Itm2b was maintained up to seven days after stimulation. This result suggests that Itm2b very likely compensates for the loss of Itm2a. Itm2b-deficient mice have been generated, but their immune phenotype has never been reported. It will of great interest to examine the development and function of T cells in mice deficient in Itm2b or both Itm2a and Itm2b.

In summary, Itm2a is a target gene of GATA-3. However, the phenotype of Itm2a deficiency was rather modest. This modest phenotype raises the possibility that Itm2b and/or Itm2c may compensate for the loss of Itm2a. We therefore examined the expression of these three genes in thymocytes and activated Th cells. We found that the transcript level of Itm2b was almost 10 times higher than that of Itm2a in thymus and activated Th cells. We found that the transcript level of Itm2b was almost 10 times higher than that of Itm2a in thymus and activated Th cells. However, its role in the adaptive immunity is rather modest probably due to functional compensation from Itm2b, which is also highly expressed in T cells. Thus, the combined function of Itm proteins remains to be determined.

Possible functional redundancy among the members of the Itm2 family

Our data indicate that Itm2a is a target gene of GATA-3. However, the phenotype of Itm2a deficiency was rather modest. This modest phenotype raises the possibility that Itm2b and/or Itm2c may compensate for the loss of Itm2a. We therefore compared the expression of these three genes in thymocytes and activated Th cells. We found that the transcript level of Itm2b was almost 10 times higher than that of Itm2a in thymus and activated Th cells. In contrast, the transcript of Itm2c was almost undetectable. In agreement with the published data, the protein level of Itm2a gradually increased in response to in vitro stimulation and peaked at day 2 after stimulation (Figure 7B), but quickly faded away after day 4. The level of Itm2b protein also increased in activated T cells but peaked at day 3-4. Unlike Itm2a, the level of Itm2b was maintained up to seven days after stimulation. This result suggests that Itm2b very likely compensates for the loss of Itm2a. Itm2b-deficient mice have been generated, but their immune phenotype has never been reported. It will of great interest to examine the development and function of T cells in mice deficient in Itm2b or both Itm2a and Itm2b.

In summary, Itm2a is a target gene of GATA-3 in thymocytes. It is required for the optimal development of OT-I thymocytes and humoral responses to Th cell-dependent antigens. However, its role in the adaptive immunity is rather modest probably due to functional compensation from Itm2b, which is also highly expressed in T cells. Thus, the combined function of Itm proteins remains to be determined.

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Author Contributions

Conceived and designed the experiments: TST SYP ICH. Performed the experiments: TST SYP. Analyzed the data: TST SYP ICH. Contributed reagents/materials/analysis tools: TST SYP ICH. Wrote the paper: TST ICH. PLOS ONE | www.plosone.org 8 May 2014 | Volume 9 | Issue 5 | e96535

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