Novel nose poke-based temporal discrimination tasks with concurrent in vivo calcium imaging in freely moving mice

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Abstract

The hippocampus has been known to process temporal information as part of memory formation. While time cells have been observed in the hippocampus and medial entorhinal cortex, a number of the behavioral tasks used present potential confounds that may cause some contamination of time cell observations due to animal movement. Here, we report the development of a novel nose poke-based temporal discrimination task designed to be used with in vivo calcium imaging for the analysis of hippocampal time cells in freely moving mice. First, we developed a ten second held nose poke paradigm for use in mice to deliver a purer time metric for the analysis of time cell activity in hippocampus CA1. Second, we developed a temporal discrimination task that involves the association of held nose poke durations of differing lengths with differential spatial cues presented in arms on a linear I-maze. Four of five mice achieved successful temporal discrimination within three weeks. Calcium imaging has been successfully performed in each of these tasks, with time cell activity being detected in the 10s nose poke task, and calcium waves being observed in discrete components of the temporal discrimination task. The newly developed behavior tasks in mice serve as novel tools to accelerate the study of time cell activity and examine the integration of time and space in episodic memory formation.

Keywords: Time, Timing, Temporal discrimination, Space, Memory, Hippocampus, Time cell, Entorhinal cortex, In vivo calcium imaging

Main text

While the hippocampus (HPC) is traditionally known for processing spatial information [1], it also processes temporal information which is important for the formation of episodic memories [2–5]. Neurons that repeatedly fire at a specific moment during the delay period of behavior tasks have been found in the hippocampus and medial entorhinal cortex (EC), and are referred to as time cells [3–7]. Time cell activity has been observed during the delay periods of treadmill running tasks, trace eye blink conditioning, spatial working memory tasks, timing tasks in virtual reality and of delayed matching to sample tasks [3, 6–11]. The activity of these time cells and their sequences during delay or interval periods is considered to link events in time, and remember specific timing in memory formation [3–5]. While time cells can be identified during the delay periods of timing-related behavior tasks even with animal body movement, it would be more desirable to analyze time cell activity in an immobile condition [8–10, 12].

In this study, we aim to deliver a purer temporal metric for the analysis of time cells with minimal contamination from body movement, speed, or head direction, which will be integrated with time by the HPC-EC circuit, while also depending on an attention-requiring task, rather than a passive wait period. To achieve this, we have designed a task in which mice are trained to actively hold a nose poke for 10 s in order to open a door leading to a reward zone. This task is designed to be paired with in vivo calcium imaging from hippocampal CA1 pyramidal cells during the held nose poke period in order to identify time cell activity in larger ensembles of neurons simultaneously. All animal lines were obtained
Fig. 1 (See legend on next page.)
from Jackson labs and bred in house. Five 26–27 week-old C57BL6J male mice (Jax 000664) and three 22–29 week-old Somatostatin (SST)-Cre transgenic male mice on a C57BL6 background (Jax 028864) were injected with 500 nl of adenovirus 5 (AAV5)-calmodulin-dependent protein kinase II alpha (CaMKIIα) with 500 nl of adeno-associated virus 5 (AAV5) expressing DAPI: blue. Lens placement was just above the hippocampus. The implant consisted of a GRIN lens (1 mm diameter) contained within a glass bottomed cannula. Stacked GCaMP6f image acquired through the miniature fluorescent microscope over 10 min of imaging from CA1 at 20 Hz. Specifically noted cells correspond to traces in d, d) Relative fluorescence changes (ΔF/F %) for five hippocampal CA1 cells identified in c. e) Nose poke behavior apparatus. Mice are trained to hold a 10 s nose poke at a nose poke port in a gated chamber (width: 8 cm, length 14 cm, height 3 cm) at the start of a linear track (Width: 8 cm; length: 30 cm, height: 3 cm). Nose poke activity was detected by an infrared photointerrupter module positioned adjacent to the nose poke port exterior opening. After holding the nose poke for the required amount of time, the door opens (orange), allowing access to the reward zone at the end of the track. f) Long-duration nose poke learning curve. Each blue line indicates the performance of individual mice. Numbers at the end of each line indicate the number of mice represented by that terminus as several of the mice had overlapping performance levels at various points in the training procedure. g) Upper panel: Three examples of hippocampal time cells imaged during successful nose poke intervals. Traces show the relative fluorescence changes (ΔF/F %) across multiple 10 s held nose poke trials for each cell. The color code plot at the bottom represents the averaged trace for each cell. h) Nose poke-based temporal discrimination behavior apparatus. Mice were trained to associate a 2.5 s held nose poke with a left or right turn (randomized per mouse), and a 10 s poke with the opposite direction. The apparatus consists of a central nose poke port-equipped starting box (width: 8 cm, length 10 cm, height 3 cm) with two doors (orange) and two arms (width: 8 cm, length 50 cm, height 3 cm) on the left and the right. i) Learning curves for the nose poke-based temporal discrimination task in mice. Each line indicates the performance of 1 mouse; Three C57BL6J mice (B6–11, B6–12, B6–13), and Two SST-Cre transgenic mice (SST-Cre-6, SST-Cre-9). Dashed line indicates 70% threshold of statistical significance (Binomial test, P < 0.05). * indicates 70% or higher success rate on 2 consecutive days. j) Calcium imaging during the temporal discrimination task. Relative fluorescence change (ΔF/F %) for a total of 20 (out of 98 imaged) cells have been presented for successful trials associated with a 2.5 s nose poke (left panel, cells A-J) and a 10 s nose poke (right panel, cells K-T). The trace activity data has been temporally aligned with movement tracking data showing the distance in cm of the animal's head from the center point (nose poke port) of the l-maze. Vertical dashed lines delineate discrete periods of the trials.
Data Processing Software v1.2 and ImageJ, as previously examined [13–15] (Fig. 1d,g). We obtained individual calcium activity from 302 cells across 3 mice (90, 50, and 162 = 302 cells in total) during the nose poke task, indicating that we successfully developed the nose poke task with in vivo calcium imaging in freely moving mice. Then, we analyzed individual calcium activity during successful nose poke periods to identify cell activity. We observed 22% of hippocampal CA1 pyramidal cells that were repeatedly activated at a specific moment during successful nose poke periods in at least one fifth of the total trials (total of 90 cells were imaged from mouse "SST-Cre-9", Fig. 1g). Some neurons had phasic responses within the first 2 s (6/20), and others activated later with responses at the middle (seconds 2–5, 5/20; seconds 5–8, 2/20) or end (seconds 8–10, 7/20) of that period (Fig. 1g, upper). We also characterized time cells as an ensemble of neurons imaged simultaneously in a single mouse, and observed that the mean peak of calcium activity for each time cell occurred at sequential moments and that the ensemble of time cells bridges the entire nose poke period (Fig. 1g, lower).

We then further developed a nose poke-based temporal discrimination task in mice. Mice which were previously trained for the 10 s nose poke task were used in this task. Five mice were habituated to the new apparatus for two days. The apparatus consists of a nose poke port located between two linear arms which are gated with mechanical doors. Each of the arms had unique spatial features on the floor, with the left arm having vertical black and white stripes 3 cm thick, and the right having black polka dots 1.5 cm in diameter on a white background. The walls of the room had additional unique visual cues comprised of shapes and patterns made from vertical or horizontal strips of tape (Fig. 1h, Additional file 1: Figure S1). Mice were trained to associate a 2.5 s tone with one arm, and a 10 s tone with the other for 7 days. After training, we started the temporal discrimination task in which mice were randomly given a short or long poke threshold (Fig. 1i). In the temporal discrimination task, upon reaching the threshold of the randomly assigned 2.5 or 10 s nose poke, both doors open and the mouse must choose which arm is associated with the poke duration it performed to open the doors. Each mouse was given 20 trials per day, with four training repetitions given prior to each testing period. The statistical threshold for considering successful acquisition of temporal discrimination was set at a 70% correct response rate for 20 trials (Binomial test, \( P < 0.05 \)) on two consecutive days. Four of five mice were observed to successfully discriminate within 22 days (Fig. 1i). Furthermore, we successfully obtained individual calcium activity during the nose poke, running and reward periods in the temporal discrimination task (Fig. 1j).

In this study, we developed novel nose poke-based temporal discrimination task with in vivo calcium imaging in freely moving mice. While time cells have been observed using head-fixed animals [8, 9], we believe that our task in freely moving mice can be useful to study the neural mechanisms of temporal discrimination, and furthermore will provide added variety to the extant behavioral approaches for examining the integration of time and space with the application of transgenic strains and viral manipulations.

**Supplementary information**

Supplementary information accompanies this paper at https://doi.org/10.1186/s13041-019-0515-7.

**Additional file 1: Figure S1A.** Basic nose poke apparatus consisting of a gated start chamber with a nose poke port within a linear track (left panel). The Arduino unit serving as the processing unit and main output of nose poke timing and experimental logs is shown in the upper right. The center panels show close up views of the nose poke port, and the photointerrupter module used as the nose poke sensor. The surface above the nose poke port prevents the mouse from climbing over and interacting with the photointerrupter from the opposite side. The door and servo mechanism are shown in the right panel. The door has extended surfaces past the sides of the maze to prevent the mouse from climbing around it. **B** Layout of the I-maze used in temporal discrimination testing. Each of the arms has a unique spatial cue associated with it. The nose poke port (inset) and door mechanisms are similar to those in the basic nose poke task. Highlighted here are other common features with the nose poke task. A speaker is used to deliver simultaneous auditory cues during active pokes. A TTL output delivers time stamp data to the nVoke system temporal allowing linkage between nose pokes and calcium spike events. A rheostat is used for program selection, and programs are triggered and terminated using the manual triggers.

**Abbreviations**

AAV: Adeno-associated virus; CaMKII: Calcium/calmodulin-dependent protein kinase II alpha; EC: Entorhinal cortex; GRIN lens: Gradient-Index lens; HPC: Hippocampus; SST: Somatostatin.

**Acknowledgements**

We thank all members of the Kitamura laboratory for their support. We thank Drs. Hiroshi Ito and Emilio Kroppf for their discussion in developing the ideas that led to this project. We also thank Dr. Jason Paris and Dr. Shari Birnbaum for their helpful conversations on behavioral approaches, and Dr. Patrick Sternkowski for his advice on calcium imaging analysis.

**Authors’ contributions**

WDM and TK conceived the study. WDM, JY, SKO, and TK designed experiments. WDM conducted experiments. WDM and HO analyzed data. WDM, HO, JY, SKO, and TK interpreted data. WDM and TK wrote the manuscript. All authors read and approved the final manuscript.

**Funding**

This work was supported by grants from Endowed Scholar Program (TK), Human Frontier Science Program (TK), Brain Research Foundation (TK), Faculty Science and Technology Acquisition and Retention Program (TK), Mary and John Osterhaus for their donation and to the Brain & Behavior Research Foundation (TK), and The Whitehall Foundation (TK).

**Availability of data and materials**

Data and scripts for analysis and behavioral apparatus operation will be made available by the corresponding author upon reasonable request.

**Ethics approval**

All animal experiments were approved by the UT Southwestern Medical Center IACUC (Protocol# 2017–102301).

**Consent for publication**

NA.
Competing interests
The authors declare that they have no competing interests.

Received: 5 September 2019 Accepted: 21 October 2019
Published online: 06 November 2019

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