Poly(ADP-RIBOSE) polymerase-1 (Parp-1) antagonizes topoisomerase I-dependent recombination stimulation by P53

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ABSTRACT

PARP-1 interacts with and poly(ADP-ribosyl)ates p53 and topoisomerase I, which both participate in DNA recombination. Previously, we showed that PARP-1 downregulates homology-directed double-strand break (DSB) repair. We also discovered that, despite the well-established role of p53 as a global suppressor of error-prone recombination, p53 enhances homologous recombination (HR) at the RARα breakpoint cluster region (bcr) comprising topoisomerase I recognition sites. Using an SV40-based assay and isogenic cell lines differing in the p53 and PARP-1 status we demonstrate that PARP-1 counteracts HR enhancement by p53, although DNA replication was largely unaffected. When the same DNA element was integrated in an episomal recombination plasmid, both p53 and PARP-1 exerted anti-recombinogenic rather than stimulatory activities. Strikingly, with DNA substrates integrated into cellular chromosomes, enhancement of HR by p53 and antagonistic PARP-1 action was seen, very similar to the HR of viral minichromosomes. siRNA-mediated knockdown revealed the essential role of topoisomerase I in this regulatory mechanism. However, after I-SceI-meganeuclease-mediated cleavage of the chromosomally integrated substrate, no topoisomerase I-dependent effects by p53 and PARP-1 were observed. Our data further indicate that PARP-1, probably through topoisomerase I interactions rather than poly(ADP-ribosyl)ation, prevents p53 from stimulating spontaneous HR on chromosomes via topoisomerase I activity.

INTRODUCTION

PARP-1 plays fundamental roles in the recruitment and modulation of enzymatic and regulatory factors involved in transcription, DNA replication, repair and recombination [reviewed in (1–3)]. Importantly, PARP-1 catalyses poly(ADP-ribosyl)ation of many of these proteins including itself and dissociates from DNA after auto-modification (4). PARP-1 is enzymatically activated by binding to single-strand breaks (SSBs) and participates in base excision repair (5,6). Given that PARP-1 also recognizes double-strand breaks (DSBs), interacts with Ku70/80, the catalytic subunit of DNA-dependent protein kinase (DNA-PKcs), and the RecQ helicase WRN, and regulates the biochemical activities of DNA-PKcs and WRN (7–10), several groups investigated potential activities of PARP-1 in DSB repair. Moreover, two recent studies showed that dysfunction of homology-directed DSB repair sensitizes cells to PARP inhibition suggesting that PARP enzymatic activity is required to avoid the accumulation of lesions which are repaired by homologous recombination (HR) (11,12).

DSBs are caused spontaneously during physiological DNA processing in replication, immunoglobulin gene diversification and meiosis and can arise from exogenous DNA-damaging agents, including ionizing radiation or cancer chemotherapeutic agents. The two major pathways of DSB repair are nonhomologous end joining (NHEJ) and HR (13,14). In the NHEJ pathway, Ku70 and Ku80 bind the DSB, followed by recruitment and activation of DNA-PKcs, which mediates synopsis and recruits XRCC4 and DNA Ligase IV. In mammalian cells HR requires a protein complex comprising Mre11, Rad50 and Nbs1 for DSB recognition and end resection to yield 3′-ssDNA tails. Subsequent strand exchange between the processed ssDNA and an intact homologous duplex are catalysed by Rad51. This reaction is facilitated by the DNA end-protecting protein Rad52, the DNA-dependent ATPase and SNF2/SWI2 family member Rad54, as
well as by the Rad51 paralogs Rad51B, Rad51C, Rad51D, Xrcc2 and Xrcc3. The breast cancer related gene product BRCA2 is thought to assist Rad51 filament assembly on ssDNA coated by replication protein A (RPA) (15). BLM and WRN, mutated in Bloom’s and Werner’s syndrome, respectively, unwind DNA, and WRN additionally exhibits exonuclease activity. These enzymes may play a role in resolving aberrantly paired structures, particularly during error-prone Rad51-dependent recombination at stalled replication forks (16).

Waldman and Waldman (17) observed lower frequencies of illegitimate recombination after treatment with an inhibitor of poly(ADP-ribosylation). Consistently, Rudat et al. (18) found that overexpression of the mere DNA-binding domain of PARP-1 (PARP-DBD), which causes trans-dominant inhibition of poly(ADP-ribosylation), inhibits rejoining of ionizing radiation-induced DSBs, suggesting that PARP-1 counteracts NHEJ when stably bound to DNA strand interruptions. On the other hand, in a recent biochemical study, Audebert et al. (19) discovered that PARP-1 provides synapsis activity on a novel route for DSB rejoining that complements the DNA-PK-dependent pathway. In agreement with the latter idea, poly(ADP-ribosyl) colocalizes with the break sensor Mre11, when nascent opened ends are generated during the coding end resolution stage of V(D)J recombination, but only in DNA-PK-α-negative cells (20). High levels of sister chromatid exchange (SCE) in cells from PARP-1-/- mice indicated a role for PARP-1 in protecting against excess HR (21,22). Recently, Shibata et al. (23) also provided in vivo evidence for a role of PARP-1 in suppressing deletion and insertion mutations which accompany chromosomal rearrangements in response to alkyl treatment. Consistently, Schultz et al. (24) noticed a hyperrecombination phenotype in cells from PARP-1-/- mice as well as after treatment of wild-type cells with an inhibitor of poly(ADP-ribosylation), as indicated by increased nuclear Rad51 foci formation. However, no significant rate change was seen in homology-directed repair of a single DSB after chemical PARP-1 inhibition, although similar treatments caused stimulation of intrachromosomal (25) and extrachromosomal (26) HR as well as homology-directed gene targeting (27). In our previous work, we used a rapid, fluorescence-based recombination assay in order to better discriminate between early DSB repair and subsequent apoptotic processes, which indirectly may enhance DSB repair (28). The results demonstrated a reduction of homology-directed DSB repair frequencies by exogenously expressed PARP-1. On the other hand, using a similar assay, Yang et al. (29) did not find evidence for interference of PARP-1 with DSB repair, although enhancement of Rad51 foci formation and SCE in cells from PARP-1-/- mice was confirmed. Additionally, delayed S-phase progression of PARP-1-/- cells was detected after replication arrest, suggesting a role in the recombinative reactivation of stalled replication forks.

The p53 tumor suppressor protein represents another PARP-1 target and has been established as an antagonist of error-prone HR events (30–35). Accumulating evidence has provided support for interference of PARP-1 with DSB repair, although similar treatments caused stimulation of intrachromosomal (25) and extrachromosomal (26) HR. As a consequence, PARP-1 inhibits rejoining of PARP-1 target and has been established as an antagonist of DSB repair (28). The results demonstrated a reduction of homology-directed DSB repair frequencies by exogenously expressed PARP-1. On the other hand, using a similar assay, Yang et al. (29) did not find evidence for interference of PARP-1 with DSB repair, although enhancement of Rad51 foci formation and SCE in cells from PARP-1-/- mice was confirmed. Additionally, delayed S-phase progression of PARP-1-/- cells was detected after replication arrest, suggesting a role in the recombinative reactivation of stalled replication forks.

RPA, BRCA1, BRCA2, BLM and/or WRN, and the DNA intermediates of homologous exchange. Similarly, p53 was also found to inhibit error-prone NHEJ (36–38). Surprisingly, when applying a specifically designed SV40-based recombination test, we discovered that p53 upregulates recombination within a fragment of the RARα breakpoint cluster region (bcr), which comprises two perfect topoisomerase I recognition sequences and is responsive to the topoisomerase I inhibitor camptothecin (39). A functional link between p53 and topoisomerase I had already been suggested by the fact that p53 forms stable complexes with topoisomerase I and enhances topoisomerase I-mediated relaxation of supercoiled DNA (40,41). Mutant analyses showed that members of the Rad52 epistasis group are involved in the repair of topoisomerase I-DNA complexes, thus indicating a critical role of the homology-directed pathway (42,43).

Based on the findings that PARP-1 interacts with and poly(ADP-ribosyl)ates p53 and topoisomerase I, which both have been implicated in HR (30–32,39,44–49), we examined the effect of PARP-1 on p53- and topoisomerase I-dependent recombination. For this purpose we adapted and applied our cellular assay systems for analyses of recombination within extrachromosomal plasmid DNA substrates, SV40 minichromosomes and cellular chromosomes (35,39,50). To distinguish possible influences of direct physical interactions from those due to enzymatic activities of PARP-1, we additionally analysed the C-terminally truncated PARP-1 mutant PARP-DBD. We demonstrate that p53 stimulates recombination through topoisomerase I, whereas PARP-1 abrogates this effect, and we define the conditions required for these processes.

MATERIALS AND METHODS

Plasmid constructs and topoisomerase I knockdown

The plasmids for extrachromosomal HR measurements were constructed by Sal I insertion of the 0.3 kb RARα bcr fragment (39) in both orientations in place of the hygromycin resistance cassette in the pHr-EGFP/3′EGFP plasmid (36). Thus, two plasmids, pHr-EGFP/3′EGFP-Rarαdfwd and pHr-EGFP/3′EGFP-Rarαrdrev were constructed, with the Rarα bcr fragment being localized between two disrupted EFGP genes.

Plasmids pPARP31, pPARP6 (51,52) and pCMV-p53 (BD Biosciences Clontech, Heidelberg) direct overexpression of full-length PARP-1, the DNA-binding domain of PARP-1, and p53, respectively, have been described previously. DNA-modifying enzymes were purchased from New England Biolabs, Frankfurt/Main.

To generate SV40 genomes carrying a 398 bp fragment from the 3′-untranslated region (3′-UTR) of the PARP gene without topoisomerase sequences (53), we transferred the HindIII fragment after blunt end formation from the PARP 3′-UTR into the Smal site within the Cla linker of shortened SV40 genome derivatives (39). The resulting vector set with a pUC-SV40-Cla, a pUC-SV40-tsVP1(196Y)-Cla and a pUC-SV40-tsVP1(290T)-Cla derivative carried the foreign sequence in the same orientation. Foreign sequences and insertion sites were verified by use of the ABI PRISM Big Dye Ready Reaction Cycle Sequencing kit and an ABI 377 sequencer.
To generate the plasmid pSUPER-Topol, which directs the synthesis of siRNAs targeting topoisomerase I mRNA 
(54) the following oligonucleotides were synthesized (Thermoxygen, Ulm): AR-TopoI10-fwd, 5'-GATCCCCAGGAGAGGACTGCTTCCCTGTGCTTTTGGGAA-3'; AR-TopoI10-rev, 5'-AGCTTTCTCAAAAAAGCAGAGAATGCTACGGTCGCCGAGAGGTCTGCTTTGAAA-3'; AR-TopoI9-rev, 5'-AGCTTTCTCAAAAAACAGATGTGGAAGTCTCTTCCAGCTTCATCAACTGTTTTGAAAA-3'; AR-TopoI9-fwd, 5'-GATCCCCAGTGTGGAAGTCTCTTCCAGCTTCATCAACTGTTTTGAAAA-3'; AR-TopoI11-rev, 5'-AGCTTTCTCAAAAAACAGATGTGGAAGTCTCTTCCAGCTTCATCAACTGTTTTGAAAA-3'; AR-TopoI11-fwd, 5'-GATCCCCAGTGTGGAAGTCTCTTCCAGCTTCATCAACTGTTTTGAAAA-3'. Whereas pSUPER plasmid containing AR-TopoI11-fwd/AR-TopoI11-rev showed no knockdown activity, both plasmids with AR-TopoI9-fwd/AR-TopoI9-rev and AR-TopoI10-fwd/AR-TopoI10-rev were positive. pSUPER-Topol with AR-TopoI10-fwd/AR-TopoI10-rev was the most effective and was, therefore, employed in further experiments. To verify RNA interference under recombination assay conditions (see below), we electropropated KMOV(HR-EGFP/3'EGFP) cells (36) with 15 μg pSUPER-TopoI (AR-TopoI10-fwd/AR-TopoI10-rev) DNA plus empty vector, p53 expression plasmid and/or PARP-1 expression plasmid. Subsequently, cells were returned to growth conditions for 48–72 h and topoisomerase I protein expression was analysed by western blotting. In these different cell types, topoisomerase I was downregulated to 40–50% of the protein expression levels in control cells correspondingly transfected with pSUPER plus empty vector, p53 expression plasmid, and/or PARP-1 expression plasmid (mean values from two to four western blots as calculated from topoisomerase I-specific band intensities after normalizing with actin or tubulin-specific band intensities).

Mammalian cell culture, cell cycle profiles and viral DNA synthesis

For SV40-based recombination assays LMV, LMV-PARP, LMV-p53her and LMV-p53her/PARP cell lines, described in Süss et al. (28), were used. Briefly, LMV is derived from the parental line LLC-MK2 from rhesus monkey (Macaca mulatta) kidney, stably expressing the estradiol-inducible transcriptional transactivation factor Gal4ERVP. LMV-PARP cells additionally express PARP activity as a measure of chromosomal recombination. To measure recombination frequencies, the ratios between the values from double infections with tsVP1-SV40 mutants and from control infections with the same infectious units of wtVP1-SV40 were determined for each single assay condition. This procedure served to normalize recombination frequencies, in order to exclude rate deviations caused by growth regulatory effects with the individual cell line used, differences in cell lethality, and alterations in virus propagation related to the specific DNA element inserted and/or alterations of transcriptional and translational activities. During recombination assays, the two tsVP1-SV40 mutants and the control wtVP1-SV40 virus carried the same foreign sequence of interest.

To quantify EGFP reconstitution as a measure of chromosomal recombination frequencies KMOV(HR-EGFP/3'EGFP) cells were electropropated at 200 V and 1050 mF with a total amount of 40 μg plasmid DNA. For p53, PARP-1 and PARP-DBD expression 5 μg of plasmid DNA were included each and replaced by the corresponding empty vectors in controls. For experiments involving pSUPER-Topol we used 15 μg of the plasmid mediating RNA interference. For experiments with targeted cleavage of the chromosomally integrated HR-EGFP/3'EGFP recombination substrate, 5 μg of pCMV-I-ScsI were also included. After electropropation cells were cultivated for 72 h at 37°C. Subsequently, cells were analysed by flow cytometry by use of a FACS® Calibur FACScan (Becton & Dickinson, Heidelberg) using the 488 nm laser line for excitation in combination with the filters used for green (FL1) and orange (FL2) fluorescence detection.
One hundred thousand living cells were analysed each to distinguish between EGFP-positive and -negative cells by the diagonal gating method in the F1/Fl2 dot plot (36,56). To determine the individual expression levels of EGFP in the presence or absence of p53, PARP-1, PARP-DBD and/or topoisomerase I, each experiment included a cotransfection with a control plasmid, which carried wild-type EGFP at the acceptor position. The fraction of EGFP-positive cells was individually determined and was used to normalize each single recombination frequency, to exclude rate deviations related to growth regulatory effects, differences in growth, cell lethality, transcriptional and translational
activities (36). The statistical significance of differences was determined using Student’s t test for unpaired samples.

To assess recombination with extrachromosomal substrate DNA, KMV cells were transfected with Lipofectamine™ 2000 Reagent (Invitrogen, Karlsruhe) and 4 μg of total plasmid DNA. The recombination plasmids pH-EGFP/3'EGFP-RRR-fwd and pH-EGFP/3'EGFP-RRR-rev were applied at 0.5 μg each. For p53, PARP-1 and PARP-DBD expression, 0.5 μg of plasmid DNA were included each and substituted by the corresponding empty vectors in controls. After cultivation for 48 h at 37°C expression of the reactivated EGFP was measured via FACS® cytometry as described above.

**Western blot analysis**

Immunoblot analysis was done essentially as described in Akyüz et al. (36). Briefly, total cell extracts were prepared from KMV or KMV(HR-EGFP/3'EGFP) cells 48 h after transfection under recombination assay conditions. Proteins were separated on 8% polyacrylamide–SDS gels and transferred to Immobilon-P Transfer Membranes (Millipore, Schwabach). The presence of topoisomerase I was revealed by the human antiserum Sci-70 (1:5,000; Topogen, Port Orange) followed by incubation with peroxidase-conjugated secondary antibody (Biomol Rockland, Vienna, Austria). For western blot analyses of PARP-1 and PARP-DBD we used goat polyclonal IgG N-20 (1:10,000; Santa Cruz, Heidelberg), for human p53 we applied the monoclonal antibody DO1 (1:10,000; Calbiochem, Bad Soden) and anti-mouse IgG peroxidase-conjugated secondary antibodies, respectively (Biomol Rockland, Vienna; Dianova/Jackson Immuno, Hamburg). To verify equal loading, tubulin or actin were detected by goat serum I-19 (1:1,000; Santa Cruz, Heidelberg) and the mAb DM1A (1:1,000; abcam, Cambridge, MA), respectively. Western blot signals were visualized by enhanced chemiluminescence (SuperSignal West Dura; Pierce/Perbio Science, Bonn).

**RESULTS**

**Effect of PARP-1 on the p53-dependent regulation of recombination between SV40 minichromosomes**

Although p53 has been established as a surveillance factor that downregulates HR in a DNA sequence-independent manner, we recently observed a stimulatory effect of p53 on recombination within a 0.3 kb DNA fragment from the RARα bcr (33–35, 39). There has been controversy about the functional links of PARP-1 to p53 (31,57–63) and about the possible role of PARP-1 in homology-directed DNA repair (24,28,29). We therefore applied our previously described SV40-based assay (39,50) in order to examine recombination under conditions where p53 exerts a positive or a negative regulatory effect. The SV40-based assay relies on the genetic exchange between SV40 genomes with different VP1-mutations causing temperature sensitivity (Figure 1a). Virus particles were produced at the permissive temperature (32°C) and DNA recombination was tested after double infection of target cells at the non-permissive temperature (39°C). Genetic reconstitution of wild-type VP1 (wtVP1) virus genomes was monitored by plaque assays at 39°C using supernatants from the doubly infected cultures. Plaque formation after wtVP1-SV40 infection was scored in parallel in order to normalize PFUs from coinfection experiments. For the analysis of RARα bcr-dependent recombination we generated virus particles from shortened SV40 genome variants [SV40-tsVP1(290T)-Cla, SV40-tsVP1(196Y)-Cla, SV40-wtVP1-Cla] with the 0.3 kb RARα bcr sequence cloned into a synthetic ClaI restriction site adjacent to the mutations causing temperature sensitivity (Figure 1a).

To address whether PARP-1 plays a role during recombination between SV40 minichromosomes, we made use of a representative set of primate cell lines conditionally expressing exogenous PARP-1 in a wild-type p53-positive or -negative background, respectively (28). The parental line LMV represents an LLC-MK2 derivative expressing the estradiol-inducible transcription factor Gal4ERVP, which directs expression of PARP-1 from pGC-PARP-1 plasmid. LMV subclones with stably integrated pGC-PARP-1 expression plasmid (LMV-PARP), with pSV53her plasmid for constitutive expression of the estradiol-responsive fusion protein containing wild-type p53 and the estradiol-binding domain of the human estrogen receptor (LMV-p53her), and with both plasmids (LMV-p53her/PARP) have been characterized previously with respect to protein expression levels, transcriptional, cell cycle regulatory and pro-apoptotic responses (28). Consistent with our earlier findings (39), expression of wild-type p53 in LMV-p53her as compared with LMV cells caused a 3.8-fold stimulation (P = 0.034) of recombination between SV40 genomes carrying the RARα bcr fragment (Figure 1b). Similar recombination frequencies were measured for LMV and LMV-PARP cells, i.e. no major influence of PARP-1 on RARα bcr-dependent recombination was...
detected. Most interestingly, however, we did not see recombination enhancement in LMV-p53her/PARP cells suggesting that PARP-1 counteracts p53-mediated stimulation of recombination between SV40 minichromosomes carrying the RARα bcr subregion. In sharp contrast to the results obtained with RARα bcr SV40 variants, recombination between SV40 genomes without foreign sequence was repressed by wild-type p53 in LMV-p53her cells (3.5-fold, \( P = 0.008 \)) (Figure 1c). LMV-PARP cells showed a 1.8-fold reduced recombination frequency. A downregulatory effect of PARP-1 was not detected in LMV-p53her/PARP cells, possibly due to the formation of DSBs by apoptotic processes which overcome anti-recombinogenic activities of p53 and PARP-1 (28). Possibly, this effect is seen only at specific response sequences or at low basal recombination frequencies, i.e. below \( 5 \times 10^{-4} \). Figure 1d shows additional control data involving a genomic sequence, which is similarly sized as the RARα bcr fragment (0.4 kb) and displays recombination-promoting features (recombination frequency for LMV: \( 67 \times 10^{-4} \)), but lacks strong topoisomerase I recognition sites. The results depicted in this figure confirm that wild-type p53 in LMV-p53her represses rather than stimulates recombination, when topoisomerase I sites are missing in the foreign DNA element (3.1-fold, \( P = 0.025 \)). PARP-1 expression in LMV-PARP cells caused a 3.5-fold downregulation. Expression of both p53 and PARP-1 in LMV-p53her/PARP cells repressed recombination 3.7 fold.

**Effect of PARP-1 and p53 on viral DNA synthesis**

Since replication arrest had been shown previously to stimulate recombination between SV40 minichromosomes (64,65), the impact of p53 and PARP-1 on RARα bcr-dependent DNA synthesis was examined. We measured \( [\text{3H}] \) thymidine incorporation into viral DNA at various times after infection (Figure 2). As reported previously (39), de novo DNA synthesis was slightly reduced by wild-type p53 (LMV-p53her), both for SV40 genomes without foreign sequence (control) and for RARα bcr comprising derivatives. Compared with LMV, expression of PARP-1 in LMV-PARP and LMV-p53her/PARP cells did not cause any major changes in the \( [\text{3H}] \) thymidine incorporation pattern compared with the control SV40 and the RARα bcr-variant. However, we cannot fully exclude a minor influence of replication on the recombination activities determined. Nevertheless, effects on recombination frequencies by PARP-1 expression in a wild-type p53-positive or -negative background failed to show a close correlation with changes in DNA replication rate.
p53 and PARP-1 downregulate recombination in extrachromosomal plasmid substrates

To further delineate the influence of p53 and PARP-1 in HR at the RARα bcr subregion, we investigated possible SV40-specific effects. For this purpose, we constructed plasmids pHREGFP/3'EGFP-RARαfwd and pHREGFP/3'EGFP-RARαrev, which allow assessment of HR between mutated EGFP genes (HR-EGFP and 3'EGFP) in the vicinity...
of the RARα bcr fragment (Figure 3a). This enabled us to use the fluorescence-based assay (36) which had successfully been used to study p53 and PARP-1 in DSB repair (28). To quantify recombination frequencies in this transient transfection assay, EGFP recombination is monitored, i.e. the ratios between green fluorescent cells and the total number of cells in the population are determined by flow cytometry. In order to relate cellular recombination events to the fraction of successfully transfected cells, we performed cotransfections with wild-type EGFP control plasmid parallel to each recombination experiment. We also wanted to exclude possible indirect effects related to DNA structure and, therefore, inserted the RARα bcr fragment between HR-EGFP and 3'EGFP in both orientations (Figure 3a).

To study homology-directed repair as a function of p53 activity, we lipofected KMV cells with the recombinant plasmids pHr-EGFP/3'EGFP-RARαfwd or pHr-EGFP/3'EGFP-RARαrev and pBS control plasmid or pCMV-p53. pCMV-p53 directs expression of wild-type p53 in KMV cells, which are derived from the parental line K562 with homozgyously deleted p53 (36). Surprisingly, KMV cells ectopically expressing p53 (Figure 3c) showed a reduction in recombination frequency by 54% rather than an increase (Figure 3b). Having shown that PARP-1 counteracts p53-dependent recombination increases at the RARα bcr in the SV40-based assay, we investigated the role of PARP-1 with the episomal substrates pHr-EGFP/3'EGFP-RARαfwd and pHr-EGFP/3'EGFP-RARαrev. To separate the enzymatic PARP-1 function from its DNA-binding activity we also tested the effect of overexpression of the mere DNA-binding domain of PARP-1 (PARP-DBD), which is devoid of enzymatic activity and causes transcriptional dominant inhibition of poly(ADP-ribosylation) (51,52). For this purpose, KMV cells were not only transfected with one of the recombination plasmids together with or without pCMV-p53 but also with plasmids pPARP31 or pPARP6 which direct expression of PARP-1 and PARP-DBD in KMV cells, respectively (Figure 3c) (28). For PARP-1 and PARP-DBD expression in the absence of p53, we observed a reduction by 21% (P = 0.000) and 16% (P = 0.030), respectively, with pHr-EGFP/3'EGFP-RARαfwd as a recombinant substrate, which was similar to the reduction of 27% (P = 0.000) and 26% (P = 0.000) with pHr-EGFP/3'EGFP-RARαrev (Figure 3b). In the presence of p53, expression of PARP-1 or PARP-DBD did not induce any additional, significant changes in recombination frequency at the given protein/DNA substrate ratios (28). Equivalent results were obtained with a similarly sized 0.4 kb fragment from the late region of the SV40 genome containing one topoisomerase I recognition sequence and a 1.3 kb hygromycin resistance cassette with multiple topoisomerase I recognition sequences (see below), thereby excluding a major regulatory influence from the specific DNA sequence (data not shown). Taken together, both p53 and PARP-1 as well as the PARP-DBD diminished HR activities with plasmid substrates independently of DNA sequence content, length and orientation between the substrates for homologous exchange.

p53 stimulates HR on cellular chromosomes via topoisomerase I and PARP-1 counteracts this stimulatory effect

So far, stimulation of recombination by p53 has only been observed with SV-40 minichromosomes (39). To clarify whether chromosomal features are critical for this effect, we applied the well-characterized, p53-deficient KMV cell line with a stably integrated HR-EGFP/3'EGFP recombination substrate (36). Within HR-EGFP/3'EGFP, mutated EGFP genes are flanking a spacer region consisting of a hygromycin resistance cassette with altogether eight perfect topoisomerase I recognition sequences (Figure 3a). KMV(HR-EGFP/3'EGFP) cells were electroporated with pBS or pCMV-p53 and concomitantly with pSUPER control plasmid or pSUPER-TopoI, which directs the synthesis of siRNAs targeting topoisomerase I mRNA, thereby causing a decrease in topoisomerase I protein level (Figure 3c). Topoisomerase I was silenced previously in HCT116 cells, when using a vector stably expressing siRNA hairpins called pREP4/Topo1 (66). p53 expression resulted in a 2.9-fold increase of spontaneous HR (P = 0.011; Figure 3b). Downregulation of endogenous topoisomerase I in cells without p53 led to a 51% reduction of the recombination frequency (not significant; P = 0.087). Strikingly, knocking down topoisomerase I expression fully suppressed the recombination increase caused by p53 expression (5.1-fold, P = 0.002). Next, KMV(HR-EGFP/3'EGFP) cells were additionally transfected with the PARP-1 and PARP-DBD recombination plasmids pPARP31 and pPARP6, respectively. In the absence of p53, neither PARP-1 nor PARP-DBD altered recombination frequencies significantly. Reminiscent of the influence of PARP-1 on HR between RARα bcr fragment carrying SV40 genomes, we did not detect any significant p53-mediated recombination.

Figure 3. Influence of p53, PARP-1 and PARP-DBD expression on recombination with plasmid episomes. (a) Design of the recombination plasmids comprising the RARα bcr sequence. The EGFP-based recombination plasmid HR-EGFP/3'EGFP (36) was modified such that the RARα bcr fragment (39) was positioned as spacer between the following DNA elements serving as substrates for homology-directed repair: HR-EGFP, an internally mutated EGFP gene under the control of the CMV promoter, following a puromycin resistance cassette, and the N-terminally truncated 3'EGFP gene. Within the spacer region the RARα bcr fragment was inserted in the forward and the reverse orientations to construct plasmids pHr-EGFP/3'EGFP-RARαfwd and pHr-EGFP/3'EGFP-RARαrev, respectively. (b) Recombination after p53, PARP-1 or PARP-DBD expression. KMV cells (36) were subjected to lipofection with the plasmids pBS or pCMV-p53 together with pBS, pPARP31 or pPARP6, for expression of wild-type p53 (wtp53), full-length PARP-1 and PARP-DBD, respectively. For determination of recombination frequencies either plasmid pHr-EGFP/3'EGFP-RARαfwd (RARα bcr forward) or pHr-EGFP/3'EGFP-RARαrev was cotransfected (RARα bcr reverse). After lipofection cells were cultivated for 48 h. Reconstitution of EGFP was monitored by FACS® analysis and recombination frequencies determined as the fraction of green fluorescent cells within the population of non-fluorescent cells. Individual transfection efficiencies served to normalize each single recombination frequency. Relative recombination frequencies were calculated by taking frequencies in control cells with pHr-EGFP/3'EGFP-RARαfwd or pHr-EGFP/3'EGFP-RARαrev as 100% (absolute values: 4.3 × 10−2 for forward, 4.2 × 10−2 for reverse). Mean values and SEM from six to nine recombination measurements are shown. (c) Protein expression. p53-negative KMV cells were lipofected with pHr-EGFP/3'EGFP-RARαrev together with pBS (control) and pCMV-p53 or with pBS (control), pPARP6, and pPARP31 and total cell extracts prepared after 48 h of continued cultivation. Immunoblotting with the antibody DO1 and goat polyclonal N-20 serum was performed to visualize expression of wild-type p53 (wtp53), and PARP-1 and PARP-DBD, respectively. The asterisk marks a band that is detected by N-20 serum independently of the inclusion of expression plasmid.
enhancement on the chromosomally integrated HR-EGFP/3'EGFP substrate after cotransfection with pPARP31 or pPARP6. Similar observations were also made in cotransfection experiments with pSUPER-Topol. Importantly, FACS® analysis of PI stained cells demonstrated that neither topoisomerase I knockdown nor p53, PARP-1 or PARP-DBD expression had a major influence on cell cycle distribution or apoptosis induction under the conditions of the assay (Figure 4d). Altogether these data indicated that diminished topoisomerase I expression as well as increased PARP-1 and PARP-DBD levels counteract spontaneous recombination on cellular chromosomes in the presence of p53.
Role of p53, PARP-1 and topoisomerase I in DSB repair after targeted cleavage of chromosomal substrate DNA

Having established a p53-topoisomerase I pathway in the regulation of spontaneous recombination events, we were interested in the functional links between p53 and topoisomerase I with respect to homology-directed DNA repair if DSBs were artificially introduced by meganuclease I-Sce I. For this purpose we performed recombination measurements with KMV(HR-EGFP/3’EGFP) cells as shown in Figure 4 and additionally co-electroporated the meganuclease I-Sce I expression plasmid pCMV-I-SceI. HR-EGFP/3’EGFP was designed such that an I-Sce I recognition sequence is positioned within HR-EGFP allowing targeted cleavage of this stably integrated recombination marker gene (Figure 5a) (36). I-Sce I expression caused a 20-fold overall increase of recombination frequencies (Figures 4b and 5b). p53 expression resulted in a 88% decrease of recombination (Figure 5b). In cells without p53, PARP-1 or PARP-DBD expression caused reduction of frequencies by 32 and 21%, respectively. No additional PARP-1-dependent changes were observed in cells electroporated with pCMV-p53. A similar picture emerged in pSUPER-TopoI cotransfection experiments. From these results we conclude that topoisomerase I has no major influence on homology-directed repair once a DSB has been introduced into the target, both in the presence and absence of wild-type p53 or PARP-1.

DISCUSSION

We and others (24,28,29) showed previously that PARP-1 has a negative regulatory role in HR, but it remained unclear whether PARP-1 is involved in homology-directed repair of DSBs, reactivation of stalled replication forks and/or alternative HR pathways. In this work, we showed that PARP-1 expression reduces homology-directed repair of a targeted DSB on chromosomes as well as HR on different extrachromosomal plasmid substrates, i.e. in a manner independent of DNA replication and of specific DNA sequences. PARP-DBD was similarly effective, strongly indicating that the underlying mechanism is binding of the DNA substrate via the ends or secondary structures (67). Supporting this concept, Rudat et al. (18) found that overexpression of PARP-DBD inhibits rejoining of DSBs. p53 and topoisomerase I both represent PARP-1-binding partners and poly(ADP-ribosyl)ation substrates and play a role in recombination (30–32,35,39,44,45,49). Therefore, we investigated more closely possible links between PARP-1, p53 and topoisomerase I in spontaneous and DSB-triggered HR. Functional interactions between PARP-1 and wild-type p53 may well have been overlooked in previous studies using mutant p53 hamster cells, embryonic stem cells, in which p53 is inactivated due to its cytoplasmic localization or embryonic fibroblasts with p53 gene mutations frequently occurring during early passages (24,29).

Our results from RNA interference experiments firstly demonstrate that p53 can stimulate HR in a manner strictly depending on topoisomerase I, which is consistent with our previous report revealing a similar degree of recombination enhancement by either p53 expression or camptothecin treatment (39). This stimulatory effect was detectable only if the DNA recombination substrate, which carried topoisomerase I recognition sequences in the neighborhood of the exchange site, was localized on viral or cellular chromosomes. This finding could mean that the cooperation between p53 and topoisomerase I depends on chromatin structure. In this regard, it is interesting to note that Rubbi and Milner (68) discovered a role of p53 in regulating chromatin remodelling which may give access to the sites of topoisomerase I action. Topoisomerase I preferentially interacts with supercoiled DNA, which alternatively may be the basis for recruiting the enzyme to DNA in the context of chromatin (69). Recombination enhancement by p53 was no longer seen, when DSBs were introduced within the DNA substrate. Instead, a net decrease of HR was monitored after wild-type p53 expression. Similarly, in the presence of naked plasmid DNA, the role of wild-type p53 in controlling the fidelity of HR dominated over its topoisomerase I-dependent HR upregulation (35,70). These observations strongly suggested that p53 and topoisomerase I cooperate in a pathway increasing the recombinogenicity of the DNA substrate which becomes irrelevant upon availability of highly recombinogenic DNA.

The recombination stimulatory role of p53 might simply be achieved through stimulation of the topoisomerase I relaxation activity to relieve torsional strain (40,41). Another possible explanation has come from biochemical observations made by Grose and co-workers (46–48). These authors demonstrated that a topoisomerase I–DNA complex can be

Figure 4. Recombination on cellular chromosomes after expression of p53, PARP-1 and PARP-DBD, and concomitant knockdown of endogenous topoisomerase I. (a) Structure of the DNA substrate for homology-directed repair in the context of cellular chromosomes. Substrate HR-EGFP/3’EGFP was chromosomally integrated in the p53-negative parental KMV line to yield KMV(HR-EGFP/3’EGFP) cells (36). HR-EGFP/3’EGFP comprises a puromycin resistance cassette, the HR-EGFP cassette, and the 3’EGFP gene (Figure 3). The spacer region contains a hygromycin resistance cassette (SV40 promoter, closed square; hygromycin resistance gene, open square) with eight perfect topoisomerase I recognition sequences. (b) Recombination as a function of p53, PARP-1 and topoisomerase I. KMV(HR-EGFP/3’EGFP) cells were co-electroporated with pBS or pCMV-p53 (wtp53) together with pBS, pPARP31 (PARP-1) or pPARP6 (PARP-DBD) and concomitantly with pSUPER or pSUPER-Topol (topoisomerase I knockdown) as indicated. Coelectroporations were followed by further cultivation for 72 h. Mean recombination frequencies and SEM were determined from 10 recombination measurements each. For the determination of relative recombination frequencies, frequencies in control cells with pBS were taken as 100% (3 × 10^-5). EGFP was typically expressed in 39–57% of the cells electroporated with the 12 different plasmid mixtures. (c) Downregulation of topoisomerase I expression. KMV(HR-EGFP/3’EGFP) cells were electroporated with 40 μg of pBS (control) or 15 μg of pSUPER-Topol mediating RNA interference plus 25 μg of pBS control plasmid and cultivated for 48 h under conditions of the recombination assay. Endogenous topoisomerase I levels were visualized via immunodetection with the human antiserum Sc1-70. Topoisomerase I from total cellular homogenates appeared as the full-length protein of 100 kDa and as a truncated version. Equal loading was verified by rehydration of the western blot with anti-actin antibodies. (d) Cellular features after topoisomerase I knockdown. Cell cycle distribution and apoptosis were analysed flow cytometrically for propidium iodide stained KMV(HR-EGFP/3’EGFP) cells 72 h after transfection and cultivation under the conditions of the recombination assay. The percentage of cells (cellular fraction) with sub-G1 DNA content (first bar), in G1 (second bar), S (third bar) and G2 phase (fourth bar) are illustrated for cells co-electroporated with pBS or pCMV-p53 (wtp53) together with pBS, pPARP31 (PARP-1) or pPARP6 (PARP-DBD) and concomitantly with pSUPER or pSUPER-Topol (topoisomerase I knockdown) each. Mean values and SD from two measurements each were calculated.
recognized by one additional topoisomerase I molecule to form the so-called topoisomerase I double cleavage complex. Release of one cleavage complex together with the oligonucleotide encompassing the DNA lesion creates an ssDNA gap that provides an entry site for a foreign, complementary DNA strand. p53 has a significant stimulatory effect on this double cleavage reaction and on the resulting recombination-mediated repair events.

When p53-mediated HR stimulation was seen on SV40 or cellular chromosomes, coexpression of PARP-1 antagonized this topoisomerase I-dependent effect (summarized in Table 1). SSBs, including those produced by topoisomerase I–DNA complex stabilization, are known to become converted into DSBs at the replication fork (71). Thus, anti-recombinogenic effects of PARP-1 could theoretically stem from its activities in the repair of SSBs (72). However, it has been well documented that p53 inhibits rather than stimulates HR in response to DSB formation at the replication fork, which is why SSB repair activities are unlikely to play a role during the p53- and topoisomerase I-dependent HR enhancement observed in this study (64,73). PARP-1 expression did not have a major influence on the DNA replication pattern or

Table 1. Comparative analysis of recombination data for p53 and PARP-1

| Recombination substrates at topoisomerase I recognition sequences | Protein expression |
|---------------------------------------------------------------|-------------------|
| Protein expression                                             | P53   | PARP-1 | p53 and PARP-1 |
| RARα bcr-SV40 minichromosomes                                   | ↑a    | ⇔b     | ⇔         |
| Naked plasmid DNA                                              | ⇩c    | ⇩      | ⇩         |
| Cellular chromosomes                                           | ↑↓    | ⇩      | ⇩         |
| Cellular chromosomes, I-Sce I cleavage                        | ⇩     | ⇯      | ⇯         |

*aSignificant increase of recombination frequency.  
*bNo significant change of recombination frequency.  
*cSignificant decrease of recombination frequency.
on S-phase progression under the conditions of the SV40-
and EGFP-based recombination measurements, respectively,
which altogether argues against a critical role of replicational
stress in the antagonistic effect of PARP-1 described here.
PARP-1 has also been shown to play a fundamental role
in chromatin condensation and decondensation, particularly
through trans-poly(ADP-ribosylation) of histones and high-
mobility group proteins (1,3). Additionally, binding of
PARP-1 to DNA is enhanced specifically by histones (74).
Therefore, PARP-1 could act on the topoisomerase I pathway
via chromatin remodelling activities. However, under condi-
tions of p53-mediated recombination enhancement, but
without exogenously expressed p53, PARP-1 did not exert
significant recombination regulatory activities. This phe-
notomen suggests that PARP-1 does not have a direct
regulatory influence on topoisomerase I-dependent recom-
bination events in general. Rather PARP-1 appears to revert
the effect of p53 on topoisomerase I specifically. These results
may be surprising because PARP-1 downregulates DSB repair
independently of p53 [(28), this study]. In conclusion, a bio-
chemical activity of PARP-1 that is different from its catalytic
activity must be responsible for the modulation of p53/
topoisomerase I activities leading to increased recombination.

Major clues to the mechanism underlying the antagonistic
role of PARP-1 in p53/topoisomerase I-promoted recombi-
nation may come from the results obtained with PARP-DBD.
PARP-DDB represents a dominant negative PARP-1 mutant
with respect to poly(ADP-ribose)ylation (51,52). Ferro and
Olivera (75) reported that covalent poly(ADP-ribose)ylation
inhibits topoisomerase I. On the other hand, Malanga and
Althaus (76) saw that poly(ADP-ribose) formation enhances
topoisomerase I-mediated break rescaling. Consistent with
the latter report, it was shown that recruitment of XRCC1
and DNA ligase III by PARP-1 as well as rejoining of DSBs via
the novel PARP-1/XRCC1/DNA ligase III-dependent route
relies on poly(ADP-ribose) synthesis (5,6,19,77). Given that
poly(ADP-ribosylation) is suppressed in PARP-DDB but not
PARP-1 expressing cells, these mechanisms cannot explain
the anti-recombinogenic effect described in this study, because
we did not obtain significantly different results with PARP-1
and PARP-DDB in EGFP-based measurements of spontan-
eous HR on cellular chromosomes. However, mapping studies
of the PARP-1 domain interacting with topoisomerase I
showed that PARP-DDB is sufficient for topoisomerase I
binding (45). Interestingly, p53 contacts the core of
topoisomerase I (amino acids 140–484), i.e. a region overlap-
ing with the PARP-1 interactions sites from amino acid 209–
400 and 626–765 (45,78,79). Therefore, it is conceivable that
PARP-1 competes with p53 for binding to topoisomerase I
or even destabilizes p53/topoisomerase I–DNA complexes
(Figure 6).

Covalent topoisomerase I–DNA complexes are normally
transient but can be stabilized by the chemotherapeutic
agent camptothecin (80) and by various DNA lesions (81).
p53 increases dissociation of topoisomerase I covalently
bound to DNA, which represents the limiting step during
topoisomerase I-mediated repair, particularly, when it is
trapped on damaged DNA (48). According to a recent report
(82), p53 may also trigger proapoptotic degradation of topoi-
somerase I covalently bound to DNA and thereby accelerate
complete removal. In contrast, PARP-1 (via the PARP-DDB)
appears to increase the binding affinity of topoisomerase I
to substrate DNA and to stimulate the formation of coval-
ent topoisomerase I–DNA complexes (45,79). Therefore,
although both factors cause a net increase in topoisomerase I
activity (45,46,49), opposing effects of p53 and PARP-1
on topoisomerase I–DNA complex dynamics could be
the molecular basis for the direct antagonizing activity in
recombination.

Wild-type p53 was demonstrated to recruit topoisomerase I
to the genomic DNA in response to DNA damage (41,83,84).
Using live cell imaging Yung et al. (49) demonstrated that
PARP-1 and topoisomerase I colocalize throughout the cell
cycle, but in response to DNA damage PARP-1-topoisomerase
I interactions are disrupted due to PARP-1 auto-modification.
Therefore, topoisomerase I may be connected to different
cellular processes by specific complex partners that contact
topoisomerase I in a mutually exclusive manner. According
to this model, p53 may represent the molecular link to recom-
bibinative repair of specific DNA lesions such as complex
DSBs which are generated by ionizing irradiation and repara-
ted slowly. PARP-1 may block this activity in undamaged,
cycling cells and rather link topoisomerase I to transcription
and DNA replication. Any shift of the delicate balance
between the different topoisomerase I functions will have
major implications for cellular genome stability, because
trapped topoisomerase I–DNA complexes initiate illegitimate
recombination events with topoisomerase I molecules bound
elsewhere in the genome (41,84,85).

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REFERENCES

1. Huber,A., Bai,P., Menissier de Murcia,J. and de Murcia,G. (2004)
PARP-1, PARP-2 and ATM in the DNA damage response: functional
synergy in mouse development. DNA Repair, 3, 1103–1108.
2. Bürkle,A. (2005) Poly(ADP-ribose): The most elaborate metabolite of
NAD+. FEBS J., 272, 4576–4589.
3. Faraoone-Mennella,M.R. (2005) Chromatin architecture and functions:
the role(s) of poly(ADP-RIBOSE) polymerase and poly
(ADPribosylation of nuclear proteins. Biochim. Cell Biol., 83, 396–404.
4. Ferro,A.M. and Olivera,B.M. (1982) Poly(ADP-riboseylation) in vitro.
Reaction parameters and enzyme mechanism. J. Biol. Chem., 257,
7808–7813.
5. Masson,M., Niedergang,C., Schreiber,V., Muller,S., Menissier-de
Murcia,J. and de Murcia,G. (1998) XRCC1 is specifically associated with
poly(ADP-ribose) polymerase and negatively regulates its activity
following DNA damage. Mol. Cell. Biol., 18, 3563–3571.
6. Schreiber,V., Ame,J.C., Dolle,P., Schultz,L., Rinaldi,B., Fraulob,V.,
Menissier de Murcia,J. and de Murcia,G. (2002) Poly(ADP-ribose)
polymerase-2 (PARP-2) is required for efficient base excision
DNA repair in association with PARP-1 and XRCC1. J. Biol. Chem.,
277, 23028–23036.
26. Semionov, A., Cournoyer, D. and Chow, T.Y. (1999) Inhibition of the poly(ADP-ribose) polymerase by PARP-1 and p53 independently function in regulating double-strand break repair in primate cells. Nucleic Acids Res., 23, 669–680.

28. Süss, S., Scholz, C.J., Bürkle, A. and Wiesmüller, L. (2004) Poly(ADP-ribose) polymerase (PARP-1) and p53 independently function in regulating double-strand break repair in primate cells. Nucleic Acids Res., 23, 669–680.

29. Yang, Y.G., Cortes, U., Patnaik, S., Jasin, M. and Weng, Z.Q. (2004) Ablation of PARP-1 does not interfere with the repair of DNA double-strand breaks, but compromises the reactivation of stalled replication forks. Oncogene, 23, 3872–3882.

30. Wiesierska-Gadek, J., Bugajska-Schretter, A. and Cerni, C. (1996) ADP-ribosylation of p53 tumor suppressor protein: mutant but not wild-type p53 is modified. J. Cell. Biochem., 62, 90–101.

31. Vaziri, H., West, M.D., Allopp, R.C., Davison, T.S., Wu, Y.S., Aronow, H.H., C. and Poiesz, B.G. and Benchimol, S. (1997) ATM-dependent telomere loss in aging human diploid fibroblasts and DNA damage lead to the post-translational activation of p53 protein involving poly(ADP-ribose) polymerase. EMBO J., 16, 6018–6033.

32. Kumari, S.R., Mendoza-Alvarez, H. and Alvarez-Gonzalez, R. (1998) Functional interactions of p53 with poly(ADP-ribose) polymerase (PARP) during apoptosis following DNA damage: a novel poly(ADP-ribose)ylation of p53 by exogenous PARP and noncovalent binding of p53 to the Ms 85000 proteolytic fragment. Cancer Res., 58, 5075–5078.

33. Bertrand, P., Saintigny, Y. and Lopez, B.S. (2004) p53’s double life: transactivation-independent repression of homologous recombination. Trends Genet., 20, 235–240.

34. Sengupta, S. and Harris, C.C. (2005) p53: traffic cop at the crossroads of DNA repair and recombination. Nature Rev. Mol. Cell Biol., 6, 44–55.

35. Gatz, S.A. and Wiesmüller, L. (2006) p53 in recombination and repair. Cell Death Differ. in press.

36. Akyüz, N., Boehden, G.S., Süss, S., Rimek, A., Preuss, U., Scheidtmann, K.-H. and Wiesmüller, L. (2002) DNA substrate dependence of the p53-mediated regulation of double-strand break repair. Mol. Cell. Biol., 22, 6306–6317.

37. Lin, Y., Waldman, B.C. and Waldman, A.S. (2003) Suppression of high-fidelity double-strand break repair in mammalian chromosomes by pifithrin-alpha, a chemical inhibitor of p53. DNA Repair, 2, 1–11.

38. Dahm-Daphi, J., Hubbe, P., Horvath, P., El-Awady, R.A., Bouffard, K.E., Powell, S.N. and Willers, H. (2005) Nonhomologous end-joining of site-specific but not of radiation-induced DNA double-strand breaks is reduced in the presence of wildtype p53. Oncogene, 24, 1663–1672.

39. Boehden, G.S., Röste, A., Marschalek, R., Stocking, C. and Wiesmüller, L. (2004) Recombination at chromosomal sequences involved in leukemogenic rearrangements is differentially regulated by p53. Carcinogenesis, 25, 1305–1313.

40. Albor, A., Kaku, S. and Kulesz-Martín, M. (1998) Wildtype and mutant forms of p53 activate human topoisomerase I: a possible mechanism for gain of function in mutants. Cancer Res., 5, 2091–2094.

41. Gobert, C., Składanowski, A. and Larsen, A.K. (1999) The interaction between p53 and DNA topoisomerase I is regulated differently in cells with wildtype and mutant p53. Proc. Natl Acad. Sci. USA, 96, 10355–10360.

42. Vance, J.R. and Wilson, T.E. (2001) Repair of DNA strand breaks by the overlapping functions of lesion-specific and non-lesion-specific DNA 3’ to 5’ exo activities. Mol. Cell. Biol., 21, 7191–7198.

43. Ponten, B., Redon, C., Rao, Y.A., Seiler, J.A., Sordet, O., Takemura, A., Antony, S., Meng, L., Liao, Z., Kohlhagen, G. et al. (2003) Repair of and checkpoint response to topoisomerase I-mediated DNA damage. Mutat. Res., 532, 173–203.

44. Ferro, A.M., Higgins, N.P. and Olivera, B.M. (1983) Poly(ADP-ribosylation) of a DNA topoisomerase. J. Biol. Chem., 258, 6000–6003.

45. Bauer, P.L., Chen, J.H., Kenedi, E., Kenessy, L., Buki, K.G., Kirsten, E., Hakama, A., Hwang, J.I. and Kun, E. (2001) Molecular interactions between poly(ADP-ribose) polymerase (PARP-1) and topoisomerase I (Topo I): identification of topology of binding. FEBS Lett., 506, 239–242.

46. Soe, K., Hartmann, H., Scholl, B., Stevsner, T. and Grosse, F. (2002) The tumor suppressor protein p53 stimulates the formation of the human topoisomerase I double cleavage complex in vitro. Oncogene, 21, 6614–6623.

47. Stephan, H., Grosse, F. and Soe, K. (2002) Human topoisomerase I cleavage complexes are repaired by a p53-stimulated recombination-like reaction in vitro. Nucleic Acids Res., 30, 5087–5093.
48. Soc,K. and Grosse,F. (2003) p53 stimulates human topoisomerase I activity by modulating its DNA binding. *Nucleic Acids Res.*, 31, 6585–6592.

49. Yung,T.M., Sato,S. and Satoh,M.S. (2004) Poly(ADP-ribose)lation as a DNA damage-induced post-translational modification regulating poly(ADP-ribose) polymerase-1 topoisomerase I interaction. *J. Biol. Chem.*, 279, 39686–39696.

50. Wiesmüller,L., Cammenga,J. and Deppert,W.W. (1996) In vivo assay of p53 function in homologous recombination between simian virus 40 chromosomes. *J. Virol.*, 70, 737–744.

51. Küpper,J.H., de Murcia,G. and Bürkle,A. (1990) Inhibition of poly(ADP-ribosylation) by overexpressing the poly(ADP-ribose) polymerase DNA-binding domain in mammalian cells. *J. Biol. Chem.*, 265, 18721–18724.

52. Küpper,J.H., Mülller,M., Jacobson,M.K., Tatsumi-Miyajima,J., Coyle,D.L., Jacobson,E.L. and Bürkle,A. (1995) Trans-dominant inhibition of poly(ADP-ribose)lation sensitizes cells against gamma-irradiation and N-methyl-N′-nitro-N-nitrosoguanidine but does not limit DNA replication of a poliovirus replicon. *Mol. Cell. Biol.*, 15, 3154–3163.

53. Will,K., Warnecke,G., Wiesmüller,L. and Deppert,W. (1998) Specific interaction of mutant p53 with regions of MARs with a high potential for base-unpairing. *Proc. Natl. Acad. Sci. USA*, 95, 13681–13686.

54. Brummelkamp,T.R., Bernard,R. and Agami,R. (2002) A system for stable expression of short interfering RNAs in mammalian cells. *Science*, 296, 550–553.

55. Zink,D., Mayr,C., Janz,C. and Wiesmüller,L. (2002) Association of p53 with the DNA replication fork. *J. Biol. Chem.*, 277, 27622–27629.

56. Ku¨pper,J.H., Mu¨ller,M., Jacobson,M.K., Tatsumi-Miyajima,J., Saintigny,Y. and Lopez,B.S. (2002) PARP-1 modifies the function of p53 in PARP-1 and topoisomerase I interaction. *Oncogene*, 21, 488–492.

57. Soye,K. and Grosse,F. (2003) p53 stimulates human topoisomerase I DNA-binding domain in mammalian cells. *J. Biol. Chem.*, 268, 2593–2600.

58. Park,S.Y. and Cheng,Y.C. (2005) Poly(ADP-ribose) polymerase-1 could be recruited by replication-mediated G1 arrest response following ionizing radiation. *J. Biol. Chem.*, 280, 11008–11016.

59. Wiener,R., Gagne,J.P., Vaziri,H., Poirier,G.G. and Benchimol,S. (2003) Poly(ADP-ribose) polymerase-1 is a positive regulator of the p53-mediated G1 arrest response following ionizing radiation. *J. Biol. Chem.*, 278, 18914–18921.

60. Junz,C. and Wiesmüller,L. (2002) Wild-type p53 inhibits replication-associated homologous recombination. *Oncogene*, 21, 5229–5233.

61. Boeddehn,G.S., Baumann,C., Sielhorst,S. and Wiesmüller,L. (2005) Wild-type p53 stimulates homologous recombination upon sequence-specific binding to the ribosomal gene cluster repeat. *Oncogene*, 24, 4183–4192.

62. Sordet,O., Liao,Z., Liu,H., Antony,S., Stevens,E.V., Kohlhagen,G., Fu,H. and Pommier,Y. (2004) Topoisomerase I-DNA complexes contribute to arsenic trioxide-induced apoptosis. *J. Biol. Chem.*, 279, 33968–33975.

63. Potaman,V.N., Shlyakhtenko,L.S., Oussatcheva,E.A., Lyubchenko,Y.L. and Soldatenkov,V. (2005) Specific binding of poly(ADP-ribose) polymerase-1 to cruciform hairpins. *J. Mol. Biol.*, 348, 609–615.

64. Robbi,C.P. and Milner,J. (2003) p53 is a chromatin accessibility factor for nucleotide excision repair of DNA damage. *EMBO J.*, 22, 975–986.

65. Madden,K.R., Stewart,L. and Chambon,P.J. (1995) Preferential binding of human topoisomerase I to superhelical DNA. *EMBO J.*, 14, 5399–5409.

66. Dudenhofer,C., Rohaly,G., Will,K., Deppert,W. and Wiesmüller,L. (1998) Specific mismatch recognition in heteroduplex intermediates by p53 suggests a role in fidelity control of homologous recombination. *Mol. Cell. Biol.*, 18, 5332–5342.

67. Strumberg,D., Fison,A.A., Smith,M., Hickey,R., Malkas,L. and Pommier,Y. (2000) Conversion of topoisomerase I cleavage complexes on the leading strand of ribosomal DNA into 5-phosphorylated DNA double-strand breaks by replication runoff. *Mol. Cell. Biol.*, 20, 3977–3987.

68. Dantzer,F., de La Rubia,G., Menissier-de Murcia,J., Hostomsky,Z., de Murcia,G. and Schreiber,Y. (2000) Base excision repair is impaired in mammalian cells lacking Poly(ADP-ribose) polymerase-1. *Biochemistry*, 39, 7559–7569.

69. Sauvigny,Y. and Lopez,B.S. (2002) Homologous recombination induced by replication inhibition, is stimulated by expression of mutant p53. *Oncogene*, 21, 488–492.

70. Sastry,S.S. and Kun,E. (1988) Molecular interactions between DNA, poly(ADP-ribose) polymerase, and histones. *J. Biol. Chem.*, 263, 1505–1512.

71. Ferro,A.M. and Olivera,B.M. (1984) Poly(ADP-ribose)ylation of DNA topoisomerase I from calf thymus. *J. Biol. Chem.*, 259, 547–554.

72. Malanga,M. and Alhau,F.R. (2004) Poly(ADP-ribose) reactivates stalled topoisomerase I and induces strandbreak resealing. *J. Biol. Chem.*, 279, 5244–5248.

73. Leppard,J.B., Dong,Z., Mackey,Z.B. and Tomkinson,A.E. (2003) Physical and functional interaction between DNA ligase IIIalpha and poly(ADP-ribose) polymerase 1 in DNA single-strand break repair. *Mol. Cell. Biol.*, 23, 5919–5927.

74. El-Hizawi,S., Lagowski,J.T. and Albor,A. (2002) Induction of gene amplification as a gain-of-function phenotype of mutant p53 proteins. *Cancer Res.*, 62, 3264–3270.

75. Ferro,A.M. and Cheng,Y.C. (2005) Poly(ADP-ribose) polymerase-1 could facilitate the religation of topoisomerase I-linked DNA inhibited by camptothecin. *Cancer Res.*, 65, 3894–3902.

76. Svejstrup,J.Q., Christiansen,K., Gromova,I.I., Andersen,A.R. and Westergard,O. (1991) New technique for uncoupling the cleavage and religation reactions of eukaryotic topoisomerase I. The mode of action of camptothecin at a specific recognition site. *J. Mol. Biol.*, 222, 669–678.

77. Pourquier,P., Ueng,L.M., Fertala,J., Wang,D., Park,H.J., Essigmann,J.M., Bjornsti,M.A. and Pommier,Y. (1999) Induction of reversible complexes between eukaryotic DNA topoisomerase I and DNA-containing oxidative base damages, 7,8-dihydro-8-oxoguanine and 5-hydroxycytosine. *J. Biol. Chem.*, 274, 8516–8523.

78. Tomicic,M.T., Christmann,M. and Kaina,B. (2005) Topotecan-triggered degradation of topoisomerase I is p53-dependent and impacts cell survival. *Cancer Res.*, 65, 8920–8926.

79. Mao,Y., Okada,S., Chang,L.S. and Muller,M.T. (2000) p53 dependence on the leading strand of ribosomal DNA into 5-phosphorylated DNA double-strand breaks by replication runoff. *Mol. Cell. Biol.*, 20, 3977–3987.

80. Svejstrup,J.Q., Christiansen,K., Gromova,I.I., Andersen,A.R. and Westergard,O. (1991) New technique for uncoupling the cleavage and religation reactions of eukaryotic topoisomerase I. The mode of action of camptothecin at a specific recognition site. *J. Mol. Biol.*, 222, 669–678.

81. Pourquier,P., Ueng,L.M., Fertala,J., Wang,D., Park,H.J., Essigmann,J.M., Bjornsti,M.A. and Pommier,Y. (1999) Induction of reversible complexes between eukaryotic DNA topoisomerase I and DNA-containing oxidative base damages, 7,8-dihydro-8-oxoguanine and 5-hydroxycytosine. *J. Biol. Chem.*, 274, 8516–8523.

82. Tomicic,M.T., Christmann,M. and Kaina,B. (2005) Topotecan-triggered degradation of topoisomerase I is p53-dependent and impacts cell survival. *Cancer Res.*, 65, 8920–8926.

83. Macaya,O., Chagla,L.S. and Muller,M.T. (2000) p53 dependence of topoisomerase I recruitment to a specific recognition site. *J. Mol. Biol.*, 222, 669–678.

84. Radford,I.R. (2002) Model for the initiation of ionizing action of camptothecin at a specific recognition site. *J. Mol. Biol.*, 222, 669–678.