REVIEW PAPER

Diversity in sphingolipid metabolism across land plants

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Abstract

Sphingolipids are essential metabolites found in all plant species. They are required for plasma membrane integrity, tolerance of and responses to biotic and abiotic stresses, and intracellular signalling. There is extensive diversity in the sphingolipid content of different plant species, and in the identities and roles of enzymes required for their processing. In this review, we survey results obtained from investigations of the classical genetic model Arabidopsis thaliana, from assorted dicots with less extensive genetic toolkits, from the model monocot Oryza sativa, and finally from the model bryophyte Physcomitrium patens. For each species or group, we first broadly summarize what is known about sphingolipid content. We then discuss the most insightful and puzzling features of modifications to the hydrophobic ceramides, and to the polar headgroups of complex sphingolipids. Altogether, these data can serve as a framework for our knowledge of sphingolipid metabolism across the plant kingdom. This chemical and metabolic heterogeneity underpins equally diverse functions. With greater availability of different tools for analytical measurements and genetic manipulation, our field is entering an exciting phase of expanding our knowledge of the biological functions of this persistently cryptic class of lipids.

Keywords: Arabidopsis thaliana, ceramide, desaturase, glycosyl inositol phosphorylceramide (GIPC), metabolic diversity, Oryza sativa, Physcomitrium patens, sphingolipid.

Introduction

Sphingolipids are essential metabolites found in all eukaryotic cells. They have superficial structural similarity to glycerolipids, the more common components of biological membranes, but have distinct chemistry, metabolism, and functions. Their unusual nature was the reason for their ‘sphingo-’ denomination; in 1884, the biochemist and physician Johann L.W. Thudichum used this term to describe these components of brain tissue, which he found confounding, in reference to the riddle of the sphinx in ancient Greek mythology (Thudichum, 1884). The scope of our understanding of sphingolipids is broader today, but arguably no less puzzling.
Across kingdoms, sphingolipids are essential for plasma membrane integrity, signalling, cell polarity and polar secretion, programmed cell death, and cellular responses to both biotic and abiotic factors in the environment. Their functions can also be highly specialized in different lineages. They have been extensively studied in animal systems due to their multiple, essential functions in the nervous system, and diseases associated with defects in sphingolipid metabolism (reviewed in Hannun and Obeid, 2018). Sphingolipid functions are equally essential for plant life, yet our understanding here is comparatively vague. Nevertheless, recent improvements in analytical methods for detecting and quantifying sphingolipids (Markham et al., 2006; Markham and Jaworski, 2007; Buré et al., 2011, 2014, 2016; Cacas et al., 2012, 2013, 2016; Tarazona et al., 2015; Ishikawa et al., 2016), as well as genome editing technologies available in different plant species (reviewed in Hahne et al., 2019), will greatly facilitate research. A clearer picture of both the biochemical and functional diversity of plant sphingolipids is currently emerging.

Plant sphingolipids can be classified in four broad groups: long-chain bases (LCBs), ceramides, glucosyl ceramides (GlcCers), and glycosyl inositol phosphorylceramides (GIPCs) (Zäuner et al., 2010) (Fig. 1). LCBs are the simplest form, and also the defining moiety of all sphingolipids (Warnecke and Heinz, 2003). They are synthesized by condensation of palmitoyl-CoA with serine to generate 3-ketosphinganine, catalysed by a heterodimeric serine palmitoyl transferase (SPT), consisting of LCB1 and LCB2a/b subunits (Tamura et al., 2001; Chen et al.,

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Fig. 1. Representative structures of a free long chain base (LCB), ceramide (Cer), glucosylceramide (GlcCer), and glycosyl inositol phosphorylceramide (GIPC). Enzymes responsible for modification or addition of moieties are indicated in bold. DES/SΔ4D, degenerative spermatocyte/sphingolipid Δ4 desaturase/dihydroceramide desaturase; SLD/SΔ8D, sphingolipid Δ8 desaturase; LOH/CerS, longevity-assurance gene one homologue/ceramide synthase; FAH, fatty acyl hydroxylase; ADS/SFD, acyl-CoenzymeA desaturase/sphingolipid fatty acyl desaturase; GCS, glucosylceramide synthase; IPCS, inositol phosphorylceramide synthase; IPUT, inositol phosphorylceramide glucuronosyltransferase; GMT/GINT, GIPC mannosyl transferase/glucosamine inositolphosphorylceramide transferase; SBH/S4H, sphingoid base hydroxylase/sphinganine-4-hydroxylase.
globosides and gangliosides, are also common (Kolter, 2011). Cerebrosides, and more extensively decorated glycosphingolipids, simple glycosphingolipids with a single sugar are referred to as both plants and fungi (Warnecke and Heinz, 2003). In animals, kingdoms. Glycosphingolipids with a single sugar headgroup, sent across eukaryotes, but take on different forms in different

There is also remarkable structural diversity in sphingolipids within the plant lineage alone. This can be expected from a theoretical standpoint, as extensive modifications to the LCB, fatty acid, and headgroup moieties are possible. Empirical data now show that within particular species and even particular tissues, a wide variety of different ceramides are produced and accumulate, both in free form and in complex sphingolipids. The default LCB structure is dihydroxylated, saturated sphinganine (or 1,3-dihydroxyphosphatidylcholine) with the hydroxylations resulting from the original condensation of palmitoyl-CoA and serine (Fig. 2). Trihydroxylated LCB (i.e., t18:0, 4-hydroxyphosphatidylcholine), produced by SPHINGOID BASE HYDROXYLASE (SBH)/SPHINGANINE-4-HYDROXYLASE (S4H) is commonly referred to as sphingosine. t18:0 is common to and abundant in plants; however, its distribution is not strictly limited to the plant lineage. Desaturation at the Δ4 position of the LCB is also possible (sphingosine, or Δ4-sphingenine), but a mutually exclusive alternative to hydroxylation at this site. This Δ4 desaturation, catalysed by DEGENERATIVE SPERMATOCYTE/SPHINGOLIPID Δ4 DESATURASE/DIHYDROCERAMIDE DESATURASE (DES/Δ4D), is particularly interesting as it is one of few trans unsaturations produced in nature (Ternes et al., 2002). Finally, Δ8 unsaturation is also commonly observed in plant LCBs, and can occur in combination with either 4-hydroxylation or Δ4 desaturation. Catalysed by SPHINGOLID Δ8 DESATURASE (SLD/Δ8D), this desaturation is observed in several phyla, including plants, fungi, diatoms, and invertebrates; however, it is especially interesting in plants, where it is unique in that it can produce both cis–(Z) and trans–(E) unsaturated products (Sperling et al., 1998; Beckmann et al., 2002; Chen et al., 2012). SLDs in other systems produce only trans products (Sato et al., 2019). Modifications of the fatty acid moiety of the ceramide also contribute to sphingolipid structural diversity. These include hydroxylation at the α-position catalysed by FATTY ACYL HYDROXYLASE (FAH) (König et al., 2012; Nagano et al., 2012), desaturation at either the n–8
modification in chain length between 16 and 26 carbon atoms, which is dependent on specificity of the condensing enzyme of the fatty acid elongase complex. Ceramide synthases (CerS/LAG ONE HOMOLOGUEs) link free LCBs to acyl-CoAs to form ceramides, and have specificity for both substrate groups; therefore, they also contribute to ceramide composition in a major way (Markham et al., 2011; Ternes et al., 2011a).

Another source of sphingolipid diversity is the headgroup structure of GIPCs. These vary in the identities and number of glycosylations, with some work indicating that up to 16 sugars can be added to the core GlcA-IPC structure (Cacas et al., 2012). GIPCs are subclassified based on the number of additional sugars added to the GlcA-IPC structure, with A-series bearing one additional sugar, B-series two, etc. The challenges inherent in isolating, detecting, and accurately identifying these structures push the limits of modern analytical methods, although substantial progress has been made in recent years. Profiling of diverse species by Cacas et al. (2013) revealed patterns in the number of sugar headgroups added to different lineages. It is equally fascinating that multiple recent studies have uncovered specific functions for specific GIPC modifications in plant physiology and biotic interactions (Grison et al., 2015; Lenarčič et al., 2017; Jiang et al., 2019; Moore et al., 2021).

Molecular genetic studies in selected model organisms have unveiled diversity in sphingolipid modifications among plant lineages, and different physiological and metabolic consequences of sphingolipid modifications. Importantly, novel features uncovered in specific model systems are seldom specific to the lineages in which we first observe or report on them. This highlights the importance of using multiple, diverse model systems in foundational plant research, and cautions against extrapolating generalizations about sphingolipid forms and their metabolic and functional significance. Describing the diversity of plant sphingolipids and determining the physiological relevance of these features is a work in progress.

In this review, we will present our current understanding of sphingolipid metabolism based on classical and emerging model systems: the molecular genetic model Arabidopsis thaliana, other dicot models that have been extensively chemically profiled including Solanum lycopersicum and Nicotiana tabacum, the monocot model Oryza sativa, and finally the model bryophyte Physcomitrium patens. Broad features of the sphingolipid profiles of these models are summarized in Fig. 3. We will discuss selected features of plant sphingolipid synthesis that vary between lineages, with an emphasis on LCB desaturations, fatty acid desaturations, and GIPC headgroups. These findings lay a framework for further exploration of sphingolipid functions in plants.

**Fig. 2.** Structures of LCB moieties of plant sphingolipids.

or n-9 position catalysed by ACYL-COA DESATURASE (ADS) (Smith et al., 2013) or SPHINGOLIPID FATTY ACYL DESATURASE (SFD) (Resemann et al., 2021), and

**Foundational work in Arabidopsis thaliana**

**Profiling A. thaliana sphingolipid content**

The variation in physical properties of plant sphingolipids makes it difficult to simultaneously extract and analyse all
classes: free LCBs, ceramides, GlcCers, and GIPCs. Additionally, there is inherent difficulty in working with the amphipathic and high molecular weight GIPCs. Profiling of sphingolipids is therefore often carried out on isolated and targeted fractions, for example after TLC separation, or after microsome enrichment in the case of GIPCs. The first global, quantitative analysis of A. thaliana sphingolipid content was published by Markham et al. (2006). This work systematically compared methods for sphingolipid extraction and thereby provided a basis for future work using rosette leaves from the popular molecular genetic model. Analysis of all sphingolipids was carried out and integrated to allow quantitative comparisons between classes. This work has since been expanded by the analysis of different organs and developmental stages (Luttgeharn et al., 2015; Kehelpannala et al., 2021), and by numerous mutant studies. While the primary focus in Markham et al. (2006) was A. thaliana, complementary profiling of Solanum lycopersicum (tomato) and Glycine max (soybean) provided an impression of the chemical diversity present within the dicot lineage (discussed in the following section).

Four methods were compared for optimized sphingolipid extraction; subsequently, neutral and anionic lipids were purified by solid-phase separation, fractionated by preparative HPLC, and finally sphingolipids were analysed by electrospray ionization MS (ESI-MS) (Markham et al., 2006). The LCB moiety profile of neutral sphingolipids (effectively, GlcCers) and anionic sphingolipids (GIPCs) was measured. Neutral sphingolipids include both ceramides and GlcCers, but because GlcCers make up the vast majority of the neutral sphingolipid pool, the neutral lipid profile is primarily representative of these, not of free ceramides. The neutral sphingolipid pool was largely made up of t18:1Δ8E/Z, with a 40% greater amount of Δ8Z than E, as well as d18:1Δ4E/Z. Anionic sphingolipids mainly contained t18:1Δ4E/Z, with a 10-fold greater amount of Δ4E than Z. These differences in the number of hydroxylations, and stereochemistry of the Δ8 double bond, suggest that there are distinct ceramide pools that are shunted into either complex sphingolipid pathway. Because both cis and trans products are produced by a single type of desaturase in plants, SΔ8D, the identity of the desaturase enzyme itself does not determine the downstream flux into GlcCer or GIPC pools.

**LCB desaturations in A. thaliana**

Although Δ8 desaturation is a predominant modification in both complex sphingolipid types, blockage of Δ8 desaturation by mutation of the two SLD/SΔ8D loci in A. thaliana resulted in depletion of GlcCers, and accumulation of GIPCs, with both containing t18:0 in lieu of t18:1Δ8. Remarkably, this substantial modification of sphingolipid metabolism resulted in only subtle effects on plant growth under optimal conditions. Mild developmental phenotypes were observed in cold temperatures, suggesting a function for Δ8 desaturation in cold stress tolerance (Chen et al., 2012). Δ4-Desaturated LCBs account for only a small proportion of the total LCBs in A. thaliana, and they are only detected in floral tissues. Analysis of loss-of-function des/SΔ4D mutants revealed no obvious phenotypes (Michaelson et al., 2009).

Compared with complex sphingolipids, the free ceramide pool of leaves has unique LCB content, made up of roughly equimolar amounts of t18:1Δ4E and t18:0 (Markham et al., 2006). This may suggest that desaturation of the LCB moiety of ceramides could be a determinant or a limiting factor for complex sphingolipid synthesis. Alternatively, this could be interpreted as evidence that ceramide pool composition is only partially determined by flux into downstream products.

**Fatty acyl content of sphingolipids in A. thaliana**

Profiles of the fatty acid moieties of free ceramides, GlcCers, and GIPCs are also unique, reflecting differences in the activity of ceramide synthases, fatty acid desaturases, and fatty acid hydroxylases. Broadly, in A. thaliana, there is an enrichment of C22–26 very-long-chain fatty acids (VLCFAs) in GIPCs, and in GlcCers there is a split between C16 long-chain fatty acids and C24 VLCFAs (Markham et al., 2011). This trend is strongly influenced by the specificity of the ceramide synthases LOH1/2/3 [named after longevity assurance gene (LAG) One Homologue] for both their LCB and fatty acyl-CoA substrates, with LOH1 and LOH3 showing preference for trihydroxy-LCB and VLCFAs, and LOH2 for dihydroxy-LCB and LCFA substrates (Markham et al., 2011; Ternes et al., 2011).

The fatty acid moieties of most free ceramides, GlcCers, and GIPCs are hydroxylated at the α-position in A. thaliana; the enzymes responsible for this modification, FATTY ACID...
HYDROXYLASE (FAH) 1/2, have been characterized by mutant analysis (Mitchell and Martin, 1995; König et al., 2012; Nagano et al., 2012). Double mutants had reduced GlcCer content and accumulated LCBs and free ceramides, suggesting that ceramide hydroxylation influences complex sphingolipid assembly. GIPCs were not measured in this analysis. The fh1 single mutant was hypersensitive to oxidative stress, and the double mutant had a mild growth defect and enhanced resistance to a biotrophic pathogen, together suggesting a function for sphingolipids harbouring α-hydroxylated VLCFAs in stress responses (König et al., 2012; Nagano et al., 2012).

The VLCFA moieties of A. thaliana sphingolipids are also often monounsaturated, in free ceramide, GlcCer, and GIPC pools. The primary desaturase required for this modification is ACYL-COA DESATURASE-LIKE2 (ADS2) (Smith et al., 2013), based on the characterization of mutant and overexpression lines. The desaturase is presumed to use acyl-CoA substrates, as the strongest effects of its genetic manipulation were observed in the acyl-CoA pool, and influenced both glycerolipid and sphingolipid content. The ADS gene family consists of nine members in A. thaliana, most of which have been only partially characterized by heterologous expression in yeast.

**Regulation of free LCB and ceramide content in A. thaliana**

While the vast majority of sphingolipids synthesized in A. thaliana are destined to become complex sphingolipids via the addition of sugar headgroups, a small fraction remains as free LCBs and ceramides. These simpler forms are metabolic fates with their own physiological functions. LCBs and ceramides are potent signalling molecules, with activities dependent on their phosphorylation status (reviewed in Berkey et al., 2012; Luttgeharm et al., 2016). LCBs and LCB phosphates (LCB-Ps) are broadly considered to have antagonistic activity in cell death signalling, with LCBs triggering cell death, and LCB-Ps mitigating or down-regulating this process (Berkey et al., 2012). However, this antagonism was recently called into question by Glenz and co-workers, who demonstrated that competitive uptake of the phosphorylated and non-phosphorylated forms could account for some of the observed antagonism of LCB-Ps against LCB-induced PCD (Glénz et al., 2019). This important physiological response certainly requires further investigation, particularly the identification of additional genetic components associated with PCD signalling. The kinases responsible for phosphorylating LCBs, SPHINGOSINE KINASE 1 and SPHINGOSINE KINASE 2, have primarily been studied in A. thaliana (Coursol et al., 2005; Worrall et al., 2008; Alden et al., 2011; Guo et al., 2011). LCB-Ps can be depleted either by dephosphorylation by LONG-CHAIN BASE PHOSPHATE PHOSPHATASEs (LCB-PP1 and LCB-PP2) (Worrall et al., 2008) or by cleavage of the LCB backbone to produce hexadecanal and phosphoethanolamine by LONG-CHAIN BASE PHOSPHATE LYASE (DPL) (Tsengay et al., 2007).

Phosphorylation of ceramide is catalysed by a ceramide kinase, ACCELERATED CELL DEATH5 (ACD5) (Greenberg et al., 2000; Li et al., 2003; Wang et al., 2008). A phosphatase catalysing the dephosphorylation of ceramide phosphates has not been reported in plants.

There are several families of ceramidases that can hydrolyse ceramides to their LCB and fatty acid components; these are classified as acid, neutral, or alkaline ceramidases based on the optimal pH for their activity. To date, only neutral and alkaline ceramidases have been identified and characterized in plants. Their activities are tightly associated with both pathogen-triggered PCD-like signalling as well as developmentally controlled PCD, autophagy, and cellular turgor pressure (Chen et al., 2015; Li et al., 2015; Wu et al., 2015; Zheng et al., 2018; Zienkiewicz et al., 2020). Interestingly, some ceramidases also have the capacity to synthesize ceramides by reverse activity (Okino et al., 2003).

**Complex sphingolipids in A. thaliana**

Identification of GCS in A. thaliana was enabled by homology to GCS genes in fungi and animals. However, characterization of a gcs mutant and delineation of the physiological role of GlcCers was impeded by its severe, seedling-lethal phenotype (Msanne et al., 2015). In contrast to the phenotype of mutant seedlings, mutant calli grew normally, suggesting that gcs mutants are impaired in cellular differentiation. Despite this deleterious effect of full loss of function on plant viability, GlcCer content reduced to as low as 2% of wild-type levels. A phosphatase catalysing the dephosphorylation of ceramide phosphates has not been reported in plants.

The initial characterization of GIPC structures was established from corn and tobacco in the mid 1900s (Carter et al., 1958; Kaul and Lester, 1975); more recent work with ESI-MS/MS and MALDI-MS has revealed a wide diversity of GIPC structures within and among different plant species (Cacas et al., 2013; Buré et al., 2014). Product and precursor ion scans of the major anionic sphingolipids of A. thaliana by Markham et al. (2006) suggested a headgroup structure of hexose-hexuronic acid–inositol phosphoceramide—an A-series GIPC. Precise identification of the headgroups was not possible with the ESI-MS approach used here. Subsequent work combining
ESI-MS/MS with MALDI-MS/MS resolved the headgroup profiles of GIPC structures in detail, for both *A. thaliana* leaves and cell cultures (Buré et al., 2011, 2016).

IPCs are common to plants, fungi, and protists (Gronnier et al., 2016). The genes responsible for transfer of the inositol phosphate headgroup to ceramide was identified previously in *Saccharomyces cerevisiae* (AUR1) (Nagiec et al., 1997) and in several protozoans (INOSITOL PHOSPHATE CERAMIDE SYNTHASE, IPCS) (Denny et al., 2006); however, the first plant IPCS was identified serendipitously via a forward genetic screen for enhanced disease responses (Wang et al., 2008). Three paralogues in *A. thaliana* have since been characterized (Mina et al., 2010). Total loss of function is impossible to assess, as triple knockouts are lethal, and single knockouts do not show any obvious defects. Curiously, several additional proteins with sequence similarity to IPCSs, named PHLOEM LOADING MODULATORS (PLMs), have also been identified in *A. thaliana*, which are required for plasmodesmal function (Yan et al., 2019). The biochemical function of PLMs remains unknown, and the specific physiological functions and biochemical specificities of IPCSs have yet to be distinguished.

In contrast to IPCS genes, the gene responsible for transferring the hexuronic acid moiety, determined to be glucuronic acid (GlcA) (Kaul and Lester, 1975), *IPUT*, was identified by reverse genetics (Rennie et al., 2014). Its product is encoded by a single gene in *A. thaliana*, and is thought to be plant specific (Cacas et al., 2012). A null *iput* mutant could not be obtained, as the mutant lesion was found to be non-transmissible through pollen due to a defect in ovule targeting (Tartaglio et al., 2017). Transient complementation of the mutation with a pollen-specific promoter enabled characterization of mutant sporophytic tissue, which had developmental defects (Tartaglio et al., 2017). Recently, the partial loss-of-function *moca1* allele of *IPUT*1 was identified in a forward genetic screen for ionic stress sensitivity, implicating GIPCs in salt sensing and initiation of cellular responses to ionic stress (Jiang et al., 2019).

The identity of the hexose residues added onto GlcA of the GIPC was explored by Mortimer et al. (2013), during the isolation and characterization of a Golgi nucleotide-sugar transporter (GONST) that yielded many surprising and interesting results. Similar to other sphingolipid-deficient mutants, *gonst1* mutants have a dwarf phenotype and elevated salicylic acid levels, resulting in spontaneous lesion formation. Recent work on a close homologue of *GONST1*, *GONST2*, revealed similar activities for the two genes *in planta*, but different functions determined by their expression profiles. This work also highlighted the effect of disruptions to GIPC synthesis on plant susceptibility to biotrophic pathogens, here *Golovinomyces orontii* (Jing et al., 2021). Biochemical characterization of *gonst1* revealed that the loss of this transporter, which has the capacity to transport GDP-mannose, -galactose, and -fucose sugars, resulted in an ~75% reduction in Hex-GIPC, and an exponential increase in GlcA-IPC lacking further glycosylation. Interestingly, monosaccharide composition analysis of isolated Hex-GIPC headgroups identified arabinose, galactose, glucose, and mannose, but only mannose content was reduced in the *gonst1* mutant. In line with this, characterization of GIPC MANNOSYL TRANSFERASE (GMT) mutants, defective in the enzyme required for the transfer of the hexose onto GIPC, revealed the strongest reduction in mannose content in the mutants. Again, other sugar moieties were detected in GIPCs, with glucose particularly abundant and variable in content (Fang et al., 2016). In both of these studies, it was concluded that mannose-GlcA-IPC is the major Hex-GlcA-IPC (and GIPC, overall) species present in *A. thaliana*. It remains unclear how depletion of the supply of mannose for addition to GIPCs in both of these mutants results in such a strong decrease in total Hex-GIPCs, when other sugars were also detected as abundant hexose moieties in the headgroup.

**Expanded insight from other dicots**

**Profiles of non-models and insight into functions of LCB desaturation**

Profiling of the sphingolipid content of *S. lycopersicum* and *G. max* by Markham et al. (2006) revealed substantial variation in sphingolipid content within the dicot lineage. Within the neutral sphingolipids, an entirely different LCB moiety predominated in *S. lycopersicum*, d18:2Δ4,E,8E,Z. In *G. max*, both the LCB most abundant in *A. thaliana*, t18:1Δ4,E,Z, and that most abundant in *S. lycopersicum*, d18:2Δ4,E,8E,Z, were observed. The significance of these differences in LCB moieties remains unclear. In contrast to the variation in ceramide structure of GlcCers, the predominant ceramide backbone of GIPCs in all three species studied by Markham et al. (2006) was t18:1/h24:0; from this, it can be suggested that trihydroxylated LCBs have conserved use in GIPC assembly.

The absence of d18:1Δ4 (sphingosine) as a major LCB moiety in any of these model plants stands in stark contrast to animals, where sphingosine is an important and abundant LCB. In animals, its derivative sphingosine-1-phosphate (S-1-P) is known to be a potent signalling molecule (Hannun and Obeid, 2018). Low levels of S-1-P have been detected in plants, and these increase upon drought stress (Ng et al., 2001). Application of S-1-P to *Commelina communis* (Asiatic dayflower) and *A. thaliana* has been reported to affect stomatal guard cell turgor in response to drought stress (Ng et al., 2001; Courson et al., 2003). Whether it is a physiologically relevant signalling molecule remains uncertain since, as mentioned, *A. thaliana Δ4* desaturase (*des/sΔ4d*) mutants unable to produce sphingosine and S-1-P did not have any obvious growth defects (Michaelson et al., 2009).

The distribution of LCB moieties that predominate in different plant lineages was explored in detail by Islam et al. (2012). Here, the desaturation of di-hydroxy LCBs was surveyed across a broad sampling of species. Unsaturation of Δ4 was the primary monounsaturation in sampled non-seed plants and angiosperms,
while Δ8 unsaturation was detected as the predominant mono-unsaturated LCB only in some select angiosperm species, including A. thaliana, and was predominant in gymnosperms. The authors concluded that Δ4 monounsaturation is ancestral. Very recently, Zhang (2021) proposed that the availability of dihydroxy LCB substrate is a major factor determining whether the Δ4–unsaturated LCB moiety accumulates, and demonstrated in A. thaliana that modifying the amount of ceramides bearing dihydroxy LCB moieties resulted in changes in the activity of a heterologously expressed Δ4 desaturase. However, due to the high activity level of the endogenous Δ8 desaturase, the Δ4 product accumulating in these transgenic lines was d18:2. Regardless of the mechanistic reason for Δ4- or Δ8-unsaturated LCB accumulating, the physiological function of Δ4-unsaturated LCB remains of interest. Islam et al. (2012) suggested that investigation of a model plant accumulating Δ4-monounsaturated LCBs, in addition to Δ8-accumulating A. thaliana, would be useful to improve our understanding of plant sphingolipid metabolism in general. To date, the only other species in which Δ4 desaturase mutants have been characterized are O. sativa and P. patens (Sato et al., 2019; Gömann et al., 2021a), discussed in the following sections.

Use of inhibitors of sphingolipid metabolism and insight into reaction order

Profiling sphingolipid content in diverse model and non-model species can be especially insightful when paired with the application of inhibitors of sphingolipid assembly. Inhibitors are valuable tools in these systems that are less tractable by genetic manipulation, which nevertheless offer specific advantages with respect to their development and physiology. Inhibitors often cause growth and developmental defects that mimic mutants defective in sphingolipid assembly. Many chemicals have been identified that affect sphingolipid metabolism, including several mycotoxins that inhibit ceramide synthesis (Chen et al., 2020), 1-phenyl-2-decanoylamino-3-morpholino-1-propanol (PDMP), which inhibits GlcCer synthesis (Msamne et al., 2015), and myriocin, which inhibits LCB synthesis (Miyake et al., 1995; Chen et al., 2021), among others. The ceramide synthase inhibitors fumonisin B₁ (FB₁) from Fusarium, and Alternaria alternata lycopersici (AAL) toxin, have been particularly well used in plant research. Treatment of duckweed (Lemma paucisostata), S. lycopersicum plants, S. lycopersicum leaf discs, or N. tabacum callus with either FB₁ or AAL toxin consistently resulted in the accumulation of d18:0 and t18:0 LCBs (Abbas et al., 1994). Other sphingoid bases could be detected in small amounts after these treatments, but their deduced structures were inconsistent with known LCBs. Direct treatment of L. paucisostata cultures with d18:0, t18:0, and d18:1Δ4 LCBs produced similar growth defects to the mycotoxins, supporting the notion that FB₁ and AAL truly act as ceramide synthase inhibitors, and that their toxic effects are mediated by excessive accumulation of LCB substrate upon ceramide synthase inhibition (Tanaka et al., 1993).

The effect of these mycotoxins on sphingolipid content may also help to reveal the order of reactions during ceramide assembly; accumulation of only d18:0 and t18:0 upon FB₁ or AAL treatment suggests that hydroxylation occurs on LCB substrates, and that desaturation, at either the Δ4 or Δ8 position, uses ceramide substrates. This model is now also supported by mutant studies in A. thaliana. In the hydroxylase mutant sbl1 sbl2, there is exponential accumulation of di-hydroxylated free LCBs and all sphingolipids incorporating di-hydroxylated LCB moieties, as well as a total depletion of tri-hydroxylated LCBs and sphingolipids containing tri-hydroxylated LCB moieties (Chen et al., 2008). In contrast, in the sld1 sld2 mutant lacking the Δ8 desaturase (SLD/Δ8D), the ceramides, GlcCers, and GIPCs with t18:0 LCB moieties accumulate exponentially, at the expense of t18:1, while the free saturated LCBs show only a modest increase (Chen et al., 2012). Cumulatively, these data present a logical and consistent model of ceramide assembly and modification.

Diversity in headgroup composition

Beyond the variations in the ceramide content of different plant species, vast chemical diversity is observed in the headgroup content of the GIPCs. Structural diversity in GIPC headgroups has arguably been best described in N. tabacum (Buré et al., 2011; Cacas et al., 2016) and derived Bright Yellow 2 (BY-2) culture lines, owing to the utility and establishment of BY-2 in other fields of research, and the simplicity of this cell line as a source of uniform, non-differentiated plasma membrane. In contrast to the Hex-GlcA-IPC content of A. thaliana, the main species of GIPCs in N. tabacum were determined to be hexamine-GlcA-IPC and N-acetylated hexamine-GlcA-IPC (Buré et al., 2011). These studies of cell cultures have consistently revealed an increase in the extent of glycosylation when compared with developed organs containing multiple cell and tissue types; enrichment of detergent-insoluble membranes from BY-2 cells revealed that GIPCs could constitute up to 60% of the lipids in this fraction. The roles of GIPCs in the functionality of these membrane domains is poorly understood, but an exciting and growing field (Gronnier et al., 2016; Mamode Cassim et al., 2019, 2021).

Clues from a model monocot: Oryza sativa

Profiling O. sativa sphingolipid content and insight into LCB desaturations

Ishikawa et al. (2016) replicated the methodology from Markham et al. (2006) for analysis of the O. sativa sphingolipidome. An additional step in their processing was the enrichment of sphingolipid classes with commercial solid-phase extraction (SPE) cartridges. Similar to G. max, O. sativa accumulates neutral sphingolipids with both d18:2Δ4E,Δ8E/Z and t18:1Δ8E/Z. The cumulative data suggest that there is substantial flexibility in the identity of the ceramide backbone used for GlcCer synthesis, and that this does not show any obvious lineage-specific
correlations. Although d18:1 is a minor component of GlcCers and free ceramides, the predominant desaturation in both classes was at the Δ4 position, conforming to the trends reported by Islam et al. (2012). The GIPC content of O. sativa was distinct from that of the dicots described above in that its predominant backbone structure was t18:0, not t18:1. The t18:0 backbone made up >80% of the GIPC class. O. sativa GlcCer Δ8 unsaturation was primarily Et18:0 backbone made up >80% of the GIPC class. Its predominant backbone structure was t18:0, not t18:1. The unsaturated fatty acids (Adem et al., 2016), and GlcCers of lupin bean and mung bean lack unsaturated fatty acids (Adem et al., 2021). Comparison of GlcCer profiles (Lynch and Dunn, 2004) from Spinacia oleracea (spinach) leaf (Ohnishi et al., 1983), Zea mays (maize) leaf (Ohnishi et al., 1988), Secale cereale (rye) leaf (Cahoon and Lynch, 1991), Triticum aestivum grain (Fujino and Ohnishi, 1983), and G. max (Ohnishi and Fujino, 1982) revealed accumulation of unsaturated fatty acids (hydroxy-24:1) only in S. cereale, which the review authors associated with this species’ cold hardiness. This correlation warrants further exploration through metabolic profiling: in more species, under cold stress conditions, and in the GIPCs of the species previously only profiled for GlcCers. Importantly, in their characterization of A. thaliana ADS2, Smith et al. (2013) did note that the ADS gene family appeared to be lacking in monocots, with no homologues found in the genomes of O. sativa, Brachypodium distachyon, or Z. mays; thorough analysis of the phylogeny of the ADS gene family will therefore also be essential to understanding the role of sphingolipid fatty acyl unsaturation in plant biology.

**Fatty acyl unsaturation in sphingolipids of O. sativa**

A particularly striking feature of the O. sativa sphingolipid profile in comparison with A. thaliana is the complete absence of unsaturated fatty acid moieties in all sphingolipid classes: free ceramides, GlcCers, and GIPCs. Fatty acyl unsaturation is a common and often prevalent feature of some profiled plant species, where it has been suggested (Fukuchi-Mizutani et al., 1998; Smith et al., 2013) to have a physiological role in cold stress adaptation. Oryza sativa is not unique in lacking ceramide fatty acyl unsaturation; unsaturated fatty acyl moieties are also absent from both GlcCers and GIPCs of tobacco (Cacas et al., 2016), and GlcCers of lupin bean and mung bean lack unsaturated fatty acids (Adem et al., 2021). Comparison of GlcCer profiles (Lynch and Dunn, 2004) from Spinacia oleracea (spinach) leaf (Ohnishi et al., 1983), Zea mays (maize) leaf (Ohnishi et al., 1988), Secale cereale (rye) leaf (Cahoon and Lynch, 1991), Triticum aestivum grain (Fujino and Ohnishi, 1983), and G. max (Ohnishi and Fujino, 1982) revealed accumulation of unsaturated fatty acids (hydroxy-24:1) only in S. cereale, which the review authors associated with this species’ cold hardiness. This correlation warrants further exploration through metabolic profiling: in more species, under cold stress conditions, and in the GIPCs of the species previously only profiled for GlcCers. Importantly, in their characterization of A. thaliana ADS2, Smith et al. (2013) did note that the ADS gene family appeared to be lacking in monocots, with no homologues found in the genomes of O. sativa, Brachypodium distachyon, or Z. mays; thorough analysis of the phylogeny of the ADS gene family will therefore also be essential to understanding the role of sphingolipid fatty acyl unsaturation in plant biology.

**GIPC headgroup assembly in O. sativa**

Analysis of the GIPC content of O. sativa (Ishikawa et al., 2016) revealed an abundance of GIPCs with two additional sugar residues added on to the glucuronic acid—B-series GIPCs—compared with the predominance of A-series GIPCs described in A. thaliana (Mortimer et al., 2013) and Nicotiana benthamiana (Cacas et al., 2016). The accumulation of B-series GIPCs is a general trend observed in monocots, whereas dicots typically accumulate more A-series (Cacas et al., 2013). This is now recognized to have physiological significance in the context of plant–pathogen interactions. Necrosis- and ethylene-inducing peptide-1-like (NLP) proteins, which are produced by many bacterial, fungal, and oomycete pathogens and are necessary for infection, were found to directly bind GIPC headgroups. The distinction between A- and B-series has been demonstrated to be a determining factor for NLP binding, and therefore for pathogenesis (Lenartčík et al., 2017).

Another interesting feature of the GIPC profile of O. sativa (Ishikawa et al., 2016) is the abundance of hexosamine (HexN)/N-acetylated hexosamine (HexNAc)—containing headgroups, which prompted the investigation of the glycosyltransferases responsible for adding these moieties
(Ishikawa et al., 2018). As described in the previous section, HexN/HexNAc as moieties of GIPCs have also been detected in dicots, for example N. tabacum (Buré et al., 2011), and also specifically in seeds and flowers of A. thaliana (Tellier et al., 2014; Luttgegharm et al., 2015); the preference for Hex versus HexN/HexNAc is therefore not specific to the dicot or monocot lineages. However, the addition of these two headgroup moieties has been thoroughly compared and investigated in O. sativa, and so we will address this topic here. In contrast to the diversity of possible Hex substrates for Hex-GIPC synthesis, glucosamine (GlcN)/N-acetylated glucosamine (GlcNAc) is the only hexosamine detected in plants, and it is therefore assumed to be the moieties present on GIPCs (Ishikawa et al., 2018). UDP-GlcNAc is abundant in plants, while UDP-GlcN has not been detected (Ito et al., 2014; therefore, Ishikawa et al. (2018) predicted that GlcNAc is transferred onto GlcA-IPC, and that GlcN-GlcA-IPC is a product of deacetylation on the assembled GIPC.

The glycosyltransferases responsible for transfer of Hex, GMT, and HexNAc, GINT, belong to the same carbohydrate-active enzyme (CAZY) glycosyltransferase family, GT64. However, there is a great deal of variety within this family. The enzymes responsible for these transfers in A. thaliana are only 29% identical; AtGINT is approximately double the length of AtGMT, and contains an additional exostosin domain. The phenotypes of A. thaliana gnt (Fang et al., 2016) and O. sativa gnt (Ishikawa et al., 2018) mutants were similar, with stunted growth and development, and cell death lesions. However, there are multiple lines of evidence that the distinct presence or profile of either product Hex-GlcA-IPC or HexNAc-GlcA-IPC is physiologically relevant. Complementation of A. thaliana gnt mutants with either A. thaliana or O. sativa GINT recovered total GIPC levels, with GlcNAc replacing Hex, but only partially rescuing the developmental phenotype of these mutants. Further evidence of the significance of Hex/HexNAc content of GIPCs was recently found in the roots and nodules of the model legume Medicago truncatula by Moore et al. (2021). MrGINT1, the gene required for transferring HexNAc headgroups to GIPCs, is expressed in tissues synthesizing perimicrobial membranes that directly interact with both bacterial (nodulating rhizobia) and fungal (arbuscular mycorrhiza) root symbionts. MrGINT1, and presumably its HexNAc-GIPC products, was found to be required for maintaining these important symbioses.

And a model moss: Physcomitrium patens

Profilng P. patens sphingolipid content and analysis of mutant phenotypes

The sphingolipid profile of the model bryophyte Physcomitrium patens was recently published, lending greater diversity to the list of examined plant species (Resemann et al., 2021). The methods used were based on Markham et al. (2006) and Tarazona et al. (2015). Major ceramide species are t18:0 LCB linked to hydroxy-22:0, -24:0, and -24:1 fatty acids. The GlcCer content of P. patens consisted almost entirely of 4:8:2 LCB with hydroxy-20:0 fatty acid. This is a unique feature of the P. patens sphingolipidome in comparison with other investigated plants, from the perspective that the LCB and fatty acid moiety of GlcCer is so homogeneous. The GIPCs used t18:0 exclusively as an LCB moiety, amide-linked to hydroxylated 20:0, 22:0, 24:0, and 24:1 fatty acids. Both A- and B-series GIPCs were detected, with both Hex and HexNAc headgroups. Mutant analysis by Steinberger et al. (2021) reported a greater variety of GIPC headgroups, including one C-series GIPC with both Hex and HexNAc headgroups containing a mixture of pentose and hexose sugars in P. patens were also reported by Cacas et al. (2013). The discrepancy among these reports of the GIPC content of P. patens highlights the difficulty inherent in targeted profiling of an essentially unknown target; although tremendous progress has been made in this field, establishment of robust and reproducible extraction and measurement is very much a work in progress.

Standard methods for cultivating P. patens gametophytic tissues in vitro, with the possibility of manipulating development by nutritional supplementation, or complementing metabolic deficiencies, have proven to be valuable tools for the analysis of sphingolipid-deficient mutants that have severe developmental defects. Two groups recently and independently generated and characterized P. patens mutants deficient in the LCB hydroxylase SPHINGANINE C-4 HYDROXYLASE (S4H)/SPHINGOID BASE HYDROXYLASE (SBH) (Gömann et al., 2021; Steinberger et al., 2021). The analogous sbh1 sbh2 loss-of-function mutant of A. thaliana was unable to develop beyond early vegetative growth, necessitating the use of RNAi to characterize the effects of a deficiency in tri-hydroxylated LCBS. In contrast, the P. patens mutants were viable and could be cultivated in parallel with wild-type plants. The simple developmental patterning of P. patens also facilitated the analysis of mutant phenotypes affecting growth, cell division, and differentiation. As severe, non-viable, and/or pleiotropic phenotypes, as well as complex developmental patterning, have presented major obstacles to the analysis and characterization of many sphingolipid-deficient vascular plant mutants, the simplicity of P. patens makes it an appealing model for further work on sphingolipids, both as structural components of cell membranes and as essential signalling molecules.

GlcCer assembly in P. patens

The homogeneous GlcCer content of P. patens facilitated further investigation of the order and importance of reactions required for GlcCer synthesis in this species, via characterization of loss-of-function glucosyl ceramide synthase (gcs) and sphingolipid delta 4 desaturase (des/sΔ4d) mutants (Gömann et al., 2021a). Multiple alleles of both single mutants were nearly completely devoid of GlcCers, and Δ4d mutants additionally lacked any LCB unsaturation, at either Δ4 or Δ8. This result suggests that
in *P. patens*, Δ4 unsaturation precedes Δ8 desaturation, and is a prerequisite for both Δ8 desaturation and addition of the hexose headgroup. A similar result was obtained and similar conclusions were drawn from study of *Pichia pastoris* (*Ternes et al., 2011b*). In contrast, in *A. thaliana*, which largely accumulates Δ8-monounsaturated LCBs in lieu of Δ4 as moieties of free ceramides, GlcCers, and GIPCs, Δ8 unsaturation is not required for Δ4 desaturase activity (*Chen et al., 2012*). Perhaps a better comparison for *PpsΔ4d* is with *Odes* mutants, as O. *sativa* also accumulates Δ4-unsaturated LCB moieties. In *Odes*, Δ8-unsaturated LCBs accumulate to compensate for the loss of Δ4, Δ8-di-unsaturated LCBs, indicating that Δ4 unsaturation is not a universal prerequisite for Δ8 unsaturation in species where Δ4 unsaturation is prevalent. However, this is only true in total LCB analysis. The loss of Δ4 unsaturation sharply reduces the production of GlcCers, which primarily contain d18:2 LCBs (*Sato et al., 2019*). Surprisingly, a similar observation was reported in the analysis of the *A. thaliana sΔ4d* mutant. In this mutant, in a species where Δ4 appears to have less physiological consequence, there was a substantial reduction in the total amount of GlcCer, which exceeded the specific reduction in d18:2 (*Michaelson et al., 2009*). Altogether, these results present a mixed model of the importance of Δ4 versus Δ8 LCB desaturation relative to each other, but suggest that Δ4 desaturation is broadly preferred for GlcCer assembly. It remains unclear whether these observations are due to substrate specificity of the downstream enzymes, metabolite channeling into different pools with different metabolic fates, or interaction and/or regulation between the enzymes.

Strikingly, although both *Ppgcs* and *PpsΔ4d* mutants were nearly completely devoid of GlcCers, with *sΔ4d* accumulating only trace amounts of GlcCer, the *gcs* mutants presented severe developmental defects while *sd4d* mutants were effectively indistinguishable from the wild type. The authors speculated that this contrast could be due to either accumulation of ceramides in the *gcs* mutant, or the trace quantities of GlcCer in *sd4d* being sufficient to maintain plant health. Generation and characterization of a double mutant will be essential to answer this question (*Gömann, 2020*). Results from *A. thaliana gcs*, which did not accumulate ceramide precursors but nevertheless presented a strong developmental phenotype, would hint that the latter hypothesis is more likely.

**Sphingolipid fatty acyl unsaturation in *P. patens***

Unsaturated fatty acids were abundant moieties of ceramides and GIPCs, and were detected in trace amounts in non-d18:2/h20:0 GlcCer. The detection of unsaturated fatty acids in *P. patens* could be argued to fit with the requirement for this modification for cold hardiness. This notion was further explored via characterization of the *P. patens* SPHINGOLIDIP FATTY ACYL DESATURASE (SFD; *Resemann et al., 2021*). Remarkably, this desaturase is from an entirely distinct desaturase family from the characterized *A. thaliana* ADS2 catalysing the analogous reaction. Sphingolipid fatty acid desaturation, in both of these clearly distinct model systems, catalysed by clearly distinct enzymes, may be similarly required for adaptation to cold stress. This points to convergent evolution of mechanisms of cold stress tolerance, between bryophyte and vascular plant lineages which diverged >500 million years ago. Further investigation of both the ADS and SFD families is warranted, as little is known about both and as yet based on very few representative members. Additionally, the distribution of these enzymes, and whether these or another as yet unidentified group of desaturase enzymes is present in monocots will be essential for our understanding of the function of sphingolipid fatty acyl desaturation in plants.

**Outlook**

Our understanding of plant sphingolipid metabolism has expanded in recent years, as a product of substantial, focused work by many research groups. While chemical profiling of sphingolipids in various plants has been done for some time, more recently the increased accessibility of non-model systems for genetic manipulation has provided a convenient, complementary toolkit. Pairing exploration via these two approaches will provide greater insight into both sphingolipid metabolism and function in plants. Further, tools that are flexible in the choice of a host will enable targeted investigations where unexpected or particularly interesting sphingolipid profiles are identified, for example in BY-2 cells. Equally importantly, these tools can be used in systems where growth and developmental phenotypes can most easily be studied, as discussed in the context of *P. patens*. Exciting topics that our field may be well positioned to investigate include the distribution and roles of the ADS- and SFD-like genes among plant lineages, the threshold level of GlcCers required for normal growth and development, the inferred functions of PLM proteins in sphingolipid metabolism, and the contributions of more extensively decorated GIPCs to plant biology.

**Conflict of interest**

The authors declare no conflict of interest.

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References

Abbas HK, Tanaka T, Duke SO, Porter JK, Wray EM, Hodges L, Sessions AE, Wang E, Merrill AH Jr, Riley RT. 1994. Fumonisin- and AAL-toxin-induced disruption of sphingolipid metabolism with accumulation of free sphingoid bases. Plant Physiology 106, 1085–1093.

Adem AA, Belete A, Soboleva A, Frolov A, Tessema EN, Gebre-Mariam T, Neubert RHH. 2021. Structural characterization of plant glucosylceramides and the corresponding ceramides by UHPLC-LTQ-Orbitrap mass spectrometry. Journal of Pharmaceutical and Biomedical Analysis 192, 113677.

Alden KP, Dhondt-Cordelier S, McDonald KL, Reape TJ, Ng CK, McCabe PF, Leaver CJ. 2011. Sphingolipid long chain base phosphates can regulate apoptotic-like programmed cell death in plants. Biochemical and Biophysical Research Communications 410, 574–580.

Beckmann C, Rattke J, Oldham NJ, Sperling P, Heinz E, Boland W. 1994. Characterization of a Δ4-sphingolipid desaturase from higher plants: a stereochromic and mechanistic study on the origin of E,Z isomers. Angewandte Chemie International Edition 41, 2298–2300.

Berkey R, Bendigeri D, Xiao S. 2012. Sphingolipids and plant defense/disease: the “death” connection and beyond. Frontiers in Plant Science 3, 68.

Buré C, Cacas J-L, Badoc A, Mongrand S, Schmitter J-M. 2016. Branched glycosylated inositolidophospho sphingolipids in plants revealed by MS3 analysis. Journal of Mass Spectrometry 51, 305–308.

Buré C, Cacas JL, Mongrand S, Schmitter JM. 2014. Characte rization of glycosyl inositol phosphorylceramides from plants and fungi by mass spectrometry. Analytical and Bioanalytical Chemistry 406, 995–1010.

Buré C, Cacas JL, Wang F, Gaudin K, Domergue F, Mongrand S, Schmitter JM. 2011. Fast screening of highly glycosylated plant sphingolipids by tandem mass spectrometry. Rapid Communications in Mass Spectrometry 25, 3131–3145.

Cacas JL, Buré C, Furt F, Maaloup JP, Badoc A, Cluzet S, Schmitter JM, Antajen E, Mongrand S. 2013. Biochemical survey of the polar head of plant glycosylinositolphosphoceramides unravels broad diversity. Phytochemistry 96, 191–200.

Cacas JL, Buré C, Grosjean K, et al. 2016. Revisiting plant plasma membrane lipids in tobacco: a focus on sphingolipids. Plant Physiology 170, 367–384.

Cacas JL, Melser S, Domergue F, Joublès J, Bourdenx B, Schmitter JM, Mongrand S. 2012. Rapid nanoscale quantitative analysis of plant sphingolipid long-chain bases by GC-MS. Analytical and Bioanalytical Chemistry 403, 2745–2755.

Cahoon EB, Lynch DV. 1991. Analysis of glucocerebroside of rye (Secale cereale L. cv Puma) leaf and plasma membrane. Plant Physiology 95, 58–68.

Carter HE, Gigk RH, Law JH, Nakayama T, Weber E. 1958. Biochemistry of the sphingolipides: XI. Structure of phytoglycolipide. Journal of Biological Chemistry 233, 1309–1314.

Chao DY, Gable K, Chen M, et al. 2011. Sphingolipids in the root play an important role in regulating the leaf ionome in Arabidopsis thaliana. The Plant Cell 23, 1061–1081.

Chen J, Li Z, Cheng Y, Gao C, Guo L, Wang T, Xu J. 2020. Sphinganine-analog myctoxcins (SAMe): chemical structures, bioactivities, and genetic controls. Journal of Fungi 6, 312.

Chen LY, Shi DQ, Zhang WJ, Tang ZS, Liu J, Yang WC. 2015. The Arabidopsis alkaline ceramidase TGD1 is a key turgor pressure regulator in plant cells. Nature Communications 6, 6030.

Chen M, Han G, Dietrich CR, Dunn TM, Cahoon EB. 2006. The essential nature of sphingolipids in plants as revealed by the functional identification and characterization of the Arabidopsis LOC1 subunit of serine palmitoyltransferase. The Plant Cell 18, 3576–3593.

Chen M, Markham JE, Cahoon EB. 2012. Sphingolipid Δ8 unsaturation is important for glucosyloceramide biosynthesis and low-temperature performance in Arabidopsis. The Plant Journal 69, 769–781.

Chen M, Markham JE, Dietrich CR, Jaworski JG, Cahoon EB. 2008. Sphingolipid long-chain base hydroxylation is important for growth and regulation of sphingolipid content and composition in Arabidopsis. The Plant Cell 20, 1862–1878.

Chen Q, Xu F, Wang L, Luo X, Wang Q, Meng Q, Huang L, Ma C, Li G, Luo M. 2021. Sphingolipid profile during cotton fiber growth revealed that a phytoceramide containing hydroxylated and saturated VLCFA is important for fiber cell elongation. Biomolecules 11, 1352.

Chueasiri C, Chunthong K, Pitnjam K, Chakhonkaen S, Sangarwut N, Sangsawang K, Suksarunprom M, Michaelson LV, Napier JA, Muangprom A. 2014. Rice ORMDL controls sphingolipid homeostasis affecting fertility resulting from abnormal pollen development. PLoS One 9, e106386.

Coursel S, Fan LM, Le Stunff H, Spiegel S, Gilroy S, Assmann SM. 2003. Sphingolipid signaling in Arabidopsis guard cells involves heterotrimic G proteins. Nature 423, 651–654.

Coursel S, Le Stunff H, Lynch DV, Gilroy S, Assmann SM, Spiegel S. 2005. Arabidopsis sphingosine kinase and the effects of phytosphingosine-1-phosphate on stomatal aperture. Plant Physiology 137, 724–737.

Denny PW, Shams-Eldin H, Price HP, Smith DF, Schwarz RT. 2006. The protozoan inositol phosphoceramide synthase: a novel drug target that defines a new class of sphingolipid synthase. Journal of Biological Chemistry 281, 28200–28209.

Fang L, Ishikawa T, Rennie EA, et al. 2016. Loss of inositol phosphoceramide sphingolipid mannosylation induces plant immune responses and reduces cellulose content in Arabidopsis. The Plant Cell 28, 2991–3004.

Fujino Y, Ohnishi M. 1983. Sphingolipids in wheat grain. Journal of Cereal Science 1, 159–168.

Fukuchi-Mizutani M, Tasaka Y, Tanaka Y, Ashikari T, Kusumia M, Murata N. 1996. Characterization of Δ9 acyl- lipid desaturase homologues from Arabidopsis thaliana. Plant & Cell Physiology 37, 247–253.

Glenz R, Schmalhaus D, Krischke M, Mueller MJ, Waller F. 2019. Elevated levels of phosphorylated sphingolipid bases do not antagonize sphingobase- or fumonisin B1-induced plant cell death. Plant & Cell Physiology 60, 1109–1119.

Gömann J. 2020. Sphingolipid biosynthesis in Phycomitrella patens. PhD dissertation, University of Göttingen.

Gömann J, Herrfurth C, Zienkiewicz K, Haslam TM, Feussner I. 2021a. Sphingolipid Δ4-desaturation is an important metabolic step for glycosyloceramide formation in Phycomitrella patens. Journal of Experimental Botany 72, 5569–5583.

Gömann J, Herrfurth C, Zienkiewicz K, Ischebeck T, Haslam TM, Feussner I. 2021b. Sphingolipid Δ4-desaturation influences plant growth and callose deposition in Physcomitrium patens. New Phytologist 231, 297–314.

Greenberg JT, Silverman FP, Liang H. 2000. Uncoupling salicyl acid-dependent cell death and disease-related responses from disease resistance in the Arabidopsis mutant acc5. Genetics 156, 341–350.

Grison MS, Brocard L, Fouillen L, et al. 2015. Specific membrane lipid composition is important for plasmodesmata function in Arabidopsis. The Plant Cell 27, 1228–1250.

Gronnier J, Germain V, Gouguet P, Cacas JL, Mongrand S. 2016. GIPC: glycosyl inositol phospho ceramides, the major sphingolipids on earth. Plant Signaling & Behavior 11, e1152438.

Guo L, Mishra G, Taylor K, Wang X. 2011. Phosphatidic acid binds and stimulates Arabidopsis sphingosine kinases. Journal of Biological Chemistry 286, 13336–13345.

Hahn O, Tomlinson L, Nogué F. 2019. Precision genetic engineering tools for next-generation plant breeding. Plant Cell Reports 38, 435–436.

Hannan YA, Obeid LM. 2018. Sphingolipids and their metabolism in physiology and disease. Nature Reviews. Molecular Cell Biology 19, 175–191.
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Hunter JE, Brandsma J, Dymond MK, Koster G, Moore CM, Postle AD, Mills RA, Attard GS. 2018. Lipidomics of Thalassiosira pseudonana under phosphorus stress reveal underlying phospholipid substitution dynamics and novel diglycosylercyclamide substitutes. Applied and Environmental Microbiology 84, e02032-02017.

Imai H, Ohnishi M, Hotsubo K, Kojima M, Ito S. 1997. Sphingoid base composition of cerebrosides from plant leaves. Bioscience, Biotechnology, and Biochemistry 61, 351–353.

Ishikawa T, Fang L, Rennie EA, et al. 2018. GLUCOSAMINE INOSITOLPHOSPHORYLCERAMIDE TRANSFERASE1 (GINT1) is a GlicNAc-containting glycosylercyclamide glycosyltransferase. Plant Physiology 177, 938–952.

Ishikawa T, Ito Y, Kawai-Yamada M. 2016. Molecular characterization and targeted quantitative profiling of the sphingolidpome in rice. The Plant Journal 88, 681–693.

Islam MN, Jacquetom MP, Coursol S, Ng CK. 2012. Sphingoosine in plants—more riddles from the Sphinks? New Phytologist 193, 51–57.

Ito J, Herter T, Baidoo EE, et al. 2014. Analysis of plant nucleotide sugars by hydrophilic interaction liquid chromatography and tandem mass spectrometry. Analytical Biochemistry 448, 14–22.

Ito S, Ohnishi M, Fujino Y. 1985. Investigation of sphingolipids in pea. Journal of Biochemistry 97, 539–540.

Jiang Z, Zhou X, Tao M, et al. 2019. Plant cell-surface GIPC sphingolipids sense salt to trigger Ca2+ influx. Nature 567, 341–346.

Jing B, Ishikawa T, Soltis N, et al. 2021. The Arabidopsis thaliana nucleotide sugar transporter GONST1 is a functional homolog of GONST1. Plant Direct 5, e00309.

Kaul K, Lester RL. 1975. Characterization of inositol-containing phosphosphingolipids from tobacco leaves: isolation and identification of two novel, major lipids: N-acetylgalcosamidoglucurononido inositol phosphorylceramide and glucosamidoglucurononido inositol phosphorylceramide. Plant Physiology 55, 120–123.

Kehelpannala C, Rupasinghe T, Pasha A, Esteban E, Hennessy T, Bradley D, Ebert B, Provant NJ, Roessner U. 2021. An Arabidopsis lipid map reveals differences between tissues and dynamic changes throughout development. The Plant Journal 107, 287–302.

Kimberlin AN, Majumder S, Han G, Chen M, Cahoob RE, Stone JM, Dunn TM, Cahoob EB. 2013. Arabidopsis 56-amino acid serine palmitoyltransferase-interacting proteins stimulate sphingolipid synthesis, are essential, and affect mycotoxin sensitivity. The Plant Cell 25, 4627–4639.

Kolter T. 2011. A view on sphingolipids and disease. Chemistry and Physics of Lipids 164, 590–596.

König S, Feusssner K, Schwartz M, Kaeber A, Iven T, Landeseinfeld M, Ternes P, Karlovsky P, Lipka V, Feusssner I. 2012. Arabidopsis mutants of sphingolipid fatty acid α-hydroxylases accumulate ceramides and sialylates. New Phytologist 196, 1086–1097.

Lenarčič T, Albert I, Böhm H, et al. 2017. Eudicot plant-specific sphingolipids determine host selectivity of microbial NLP cryoplates. Science 358, 1431–1434.

Li J, Bi FC, Yin J, Wu JX, Rong C, Wu JL, Yao N. 2015. An Arabidopsis neutral ceramide mutant noer1 accumulates hydroxyceramides and is sensitive to oxidative stress. Frontiers in Plant Science 6, 460.

Li J, Yin J, Rong C, Li KE, Wu JX, Huang LQ, Zeng HY, Sahu SK, Yao N. 2016. Orosomucoid proteins interact with the small subunit of serine palmitoyltransferase and contribute to sphingolipid homeostasis and stress responses in Arabidopsis. The Plant Cell 28, 3038–3051.

Liang H, Yao N, Song JT, Luo S, Lu H, Greenberg JT. 2003. Ceramides modulate programmed cell death in plants. Genes & Development 17, 2636–2641.

Luttgrehm KD, Cahoob EB, Markham JE. 2016. Substrate specificity, kinetic properties and inhibition by furonisin B1 of ceramide synthase isoforms from Arabidopsis. The Biochemical Journal 473, 593–603.

Luttgrehm KD, Kimberlin AN, Cahoob RE, Cerny RL, Napier JA, Markham JE, Cahoob EB. 2015. Sphingolipid metabolism is strikingly different between pollen and leaf in Arabidopsis as revealed by compositional and gene expression profiling. Phytochemistry 115, 121–129.

Lynch DV, Dunn TM. 2004. An introduction to plant sphingolipids and a review of recent advances in understanding their metabolism and function. New Phytologist 161, 677–702.

Mamode Cassim A, Gouguet P, Gronnier J, Laurent N, Germain V, Grison M, Boutté Y, Gerbeau-Pissot P, Simon-Plass F, Mongrand S. 2019. Plant lipids: key players of plasma membrane organization and function. Progress in Lipid Research 73, 1–27.

Mamode Cassim A, Navon Y, Gao Y, et al. 2021. Biophysical analysis of the plant-specific GIPC sphingolipid reveals multiple modes of membrane regulation. Journal of Biological Chemistry 296, 100602.

Markham JE, Jaworski JG. 2007. Rapid measurement of sphingolipids from Arabidopsis thaliana by reversed-phase high-performance liquid chromatography coupled to electrospray ionization tandem mass spectrometry. Rapid Communications in Mass Spectrometry 21, 1304–1314.

Markham JE, Li J, Cahoob EB, Jaworski JG. 2006. Separation and identification of major plant sphingolipid classes from Arabidopsis. Journal of Biological Chemistry 281, 22684–22694.

Markham JE, Molino D, Gissot L, Bellec Y, Hématy K, Marion J, Belcam R, Palaquji JC, Satiat-Jeneuinae B, Faure JD. 2011. Sphingolipids containing very-long-chain fatty acids define a secreory pathway for specific polar plasma membrane protein targeting in Arabidopsis. The Plant Cell 23, 2362–2378.

Michaelson LV, Zäuner S, Markham JE, et al. 2009. Functional characterization of a higher plant sphingolipid α4-deasaturase: defining the role of sphingosine and sphingosine-1-phosphate in Arabidopsis. Plant Physiology 149, 487–498.

Miller R, Wu G, Deshpande RR, et al. 2010. Changes in transcript abundance in Chlamydomonas reinhardtii following nitrogen deprivation predict diversion of metabolism. Plant Physiology 154, 1737–1752.

Mina JG, Okada Y, Wansadhipathi-Kannagara NK, Pratt S, Shams-Eldin H, Schwarz RT, Steel PG, Fawcett T, Denny PW. 2010. Functional analyses of differentially expressed isoforms of the Arabidopsis inositol phosphoceramidase synthase. Plant Molecular Biology 73, 399–407.

Mitchell AG, Martin CE. 1995. A novel cytochrome b5-like domain is linked to the carboxyl terminus of the Saccharomyces cerevisiae Δ9 fatty acid desaturase. Journal of Biological Chemistry 270, 29766–29772.

Miyake Y, Kozutsumi Y, Nakamura S, Fujita T, Kawasaki T. 1995. Serine palmitoyltransferase is the primary target of a sphingosine-like immunosuppressant, ISP-1/myriocin. Biochemical and Biophysical Research Communications 211, 396–403.

Moore WM, Chan C, Ishikawa T, Rennie EA, Wipf HM, Benitez V, Kawai-Yamada M, Mortimer JC, Scheller HV. 2021. Reprogramming sphingolipid glycosylation is required for endosymbiont persistence in Medicago truncatula. Current Biology 31, 2374–2385.

Mortimer JC, Yu X, Albrecht S, et al. 2013. Abnormal glycosphingolipid mannosylation triggers salicylic acid-mediated responses in Arabidopsis. The Plant Cell 25, 1881–1894.

Msanne J, Chen M, Luttgrehm KD, Bradley AM, Mays ES, Paper JM, Boyle DL, Cahoob RE, Schrick K, Cahoob EB. 2015. Glucosylercerramides are critical for cell-type differentiation and organogenesis, but not for cell viability in Arabidopsis. The Plant Journal 84, 188–201.

Nagano M, Takahara K, Fujimoto M, Tutsunomi U, Uchiyama H, Kawai-Yamada M. 2012. Arabidopsis sphingolipid fatty acid 2-hydroxylases (AtFAH1 and AtFAH2) are functionally differentiated in fatty acid 2-hydroxylation and stress responses. The Plant Cell 149, 1138–1148.

Nagiec MM, Nagiec EE, Baltisberger JA, Wells GB, Lester RL, Dickson RC. 1997. Sphingolipid synthesis as a target for antifungal drugs: complementation of the inositol phospholipidcereamide synthase defect in a mutant strain of Saccharomyces cerevisiae by the AUR1 gene. Journal of Biological Chemistry 272, 9809–9817.

Ng CK, Carr K, McAlinsh MR, Powell B, Hetherington AM. 2001. Drought-induced guard cell signal transduction involves sphingosine-1-phosphate. Nature 410, 596–599.

Ohnishi M, Fujino Y. 1982. Sphingolipids in immature and mature soybeans. Lipids 17, 803–810.
Ohnishi M, Ito S, Fujino Y. 1983. Characterization of sphingolipids in spinach leaves. Biochimica et Biophysica Acta 752, 416–422.

Ohnishi M, Imai H, Kojima M, Yoshida S, Murata N, Fujino Y, Ito S. 1988. Separation of cerebrosides species in plants by HPLC and their phase transition temperature. Proceedings of the ISF–JOCS World Congress 11, 930–936.

Okino N, He X, Gatt S, Sandhoff K, Ito M, Schuchman EH. 2003. The reverse activity of human acid ceramidase. Journal of Biological Chemistry 278, 29948–29953.

Posse de Chaves E, Sipione S. 2010. Sphingolipids and gangliosides of the nervous system in membrane function and dysfunction. FEBS Letters 584, 1748–1759.

Rennie EA, Ebert B, Miles GP, et al. 2014. Identification of a sphingolipid α-gluconuronosyltransferase that is essential for pollen function in Arabidopsis. The Plant Cell 26, 3314–3325.

Resemann HC, Herrfurth C, Feussner K, et al. 2021. Convergence of sphingolipid desaturation across over 500 million years of plant evolution. Nature Plants 7, 219–232.

Santos FC, Marquês JT, Bento-Oliveira A, de Almeida RFM. 2020. Sphingolipid-enriched domains in fungi. FEBS Letters 594, 3698–3718.

Sato M, Nagano M, Jin S, Miyagi A, Yamaguchi M, Kawai-Yamada M, Ishikawa T. 2019. Plant-unique cis/trans isomerism of long-chain base unsaturation is selectively required for aluminum tolerance resulting from glucosylceramide-dependent plasma membrane fluidity. Plants 9, 19.

Smith MA, Dauk M, Ramadan H, Yang H, Seamons LE, Haslam RP, Beaudoin F, Ramirez-Erosa I, Forseille L. 2013. Involvement of Arabidopsis ACYL-COENZYME A DESATURASE-LIKE2 (At2g31360) in the biosynthesis of the very-long-chain monounsaturated fatty acid components of membrane lipids. Plant Physiology 161, 81–96.

Sperling P, Zähringer U, Heinz E. 1998. A sphingolipid desaturase from higher plants—identification of a new cytochrome b₆ fusion protein. Journal of Biological Chemistry 273, 28590–28596.

Steinberger AR, Merino WO, Cahoon RE, Cahoon EB, Lynch DV. 2021. Disruption of long-chain base hydroxylation alters growth and impacts sphingolipid synthesis in Physcomitrella patens. Plant Direct 5, e6396.

Tamura K, Mitsuhashi N, Hara-Nishimura I, Imai H. 2001. Characterization of an Arabidopsis cDNA encoding a subunit of serine palmitoyltransferase, the initial enzyme in sphingolipid biosynthesis. Plant & Cell Physiology 42, 1274–1281.

Tanaka T, Abbas HK, Duke SO. 1993. Structure-dependent phytotoxicity of fumonisins and related compounds in a duckweed bioassay. Phytochemistry 33, 779–785.

Tarazona P, Feussner K, Feussner I. 2015. An enhanced plant lipidomics method based on multiplexed liquid chromatography-mass spectrometry reveals additional insights into cold- and drought-induced membrane remodeling. The Plant Journal 84, 621–633.

Tartaglio V, Rennie EA, Cahoon R, Wang G, Baidoo E, Mortimer JC, Cahoon EB, Scheller HV. 2017. Glycosylation of inositol phosphocholine sphingolipids is required for normal growth and reproduction in Arabidopsis. The Plant Journal 89, 278–290.

Tellier F, Maia-Grondard A, Schmitz-Afonso I, Faure JD. 2014. Comparative plant sphingolipidomic reveals specific lipids in seeds and oil. Phytochemistry 103, 50–58.

Ternes P, Feussner K, Werner S, Lerche J, Iven T, Heilmann I, Riezman H, Feussner I. 2011a. Disruption of the ceramide synthase LOH1 causes spontaneous cell death in Arabidopsis thaliana. New Phytologist 192, 841–854.

Ternes P, Franke S, Zähringer U, Sperling P, Heinz E. 2002. Identification and characterization of a sphingolipid Δ4-desaturase family. Journal of Biological Chemistry 277, 25512–25518.

Ternes P, Wobbe T, Schwarz M, et al. 2011b. Two pathways of sphingolipid biosynthesis are separated in the yeast Pichia pastoris. Journal of Biological Chemistry 286, 11401–11414.

Thudichum J. 1884. A treatise on the chemical constitution of the brain: based throughout upon original researches. Glasgow Medical Journal 22, 363–364.

Tsegaye Y, Richardson CG, Bravo JE, Mulcahy BJ, Lynch DV, Markham JE, Jaworski JG, Chen M, Cahoon EB, Dunn TM. 2007. Arabidopsis mutants lacking long chain base phosphate lyase are fumonisin-sensitive and accumulate trihydroxy-18:1 long chain base phosphate. Journal of Biological Chemistry 282, 28195–28206.

Wang W, Yang X, Tangchalarubana S, et al. 2008. An inositolphosphorylceramide synthase is involved in regulation of plant programmed cell death associated with defense in Arabidopsis. The Plant Cell 20, 3163–3179.

Warnecke D, Heinz E. 2003. Recently discovered functions of glucosylceramides in plants and fungi. Cellular and Molecular Life Sciences 60, 919–941.

Worrall D, Liang YK, Alvarez S, Holroyd GH, Spiegel S, Panagopoulos M, Gray JE, Hetherington AM. 2008. Involvement of sphingosine kinase in plant cell signalling. The Plant Journal 56, 64–72.

Wu JX, Li J, Liu Z, Yin J, Chang ZY, Rong C, Wu JL, Bi FC, Yao N. 2015. The Arabidopsis ceramide acyltransferase ACET1 functions in disease resistance and salt tolerance. The Plant Journal 81, 787–790.

Yan D, Yadav SR, Paterlini A, et al. 2019. Sphingolipid biosynthesis modulates plasmodesma ultrastructure and phloem unloading. Nature Plants 5, 604–615.

Yang B, Li M, Phillips A, Li L, Ali U, Li Q, Lu S, Hong Y, Wang X, Guo L. 2021. Nonspecific phospholipase C4 hydrolyzes phosphosphingolipids and sustains plant root growth during phosphate deficiency. The Plant Cell 33, 766–780.

Zäuner S, Ternes P, Warnecke D. 2010. Biosynthesis of sphingolipids in plants (and some of their functions). Advances in Experimental Medicine and Biology 688, 249–263.

Zhang D. 2021. Characterization of sphingolipid biosynthesis and modification in plants. PhD thesis, University of Nebraska.

Zheng P, Wu JX, Sahu SK, Zeng HY, Huang LQ, Liu Z, Xiao S, Yao N. 2018. Loss of alkaline ceramidase inhibits autophagy in Arabidopsis and plays an important role during environmental stress response. Plant, Cell & Environment 41, 837–849.

Zienkiewicz A, Gömann J, Herrfurth C, Liu YT, Meldau M, Ishikawa T. 2019. Plant-unique cis/trans isomerism of long-chain base hydroxylation alters growth and immunity in Arabidopsis. The Plant Journal 81, 767–780.