Expression of the serotonin 1b receptor in experimental pulmonary hypertension

B. Rondelet*, R. Van Beneden**, F. Kerbau†, S. Motte*, P. Fesler*, K. McEntee*, S. Brimioulle*, J-M. Ketelslegers#, R. Naeije*

Expression of the serotonin 1b receptor in experimental pulmonary hypertension. B. Rondelet, R. Van Beneden, F. Kerbau, S. Motte, P. Fesler, K. McEntee, S. Brimioulle, J-M. Ketelslegers, R. Naeije. ©ERS Journals Ltd 2003.

ABSTRACT: The pathogenesis of pulmonary arterial hypertension (PAH) remains uncertain. Both the serotonin and endothelin (ET) systems are believed to be involved. Recent studies pointed to the importance of the serotonin 2B receptor as a limiting step.

The current authors investigated the lung tissue expression of serotonin receptors and of the serotonin transporter (5-HTT) by real-time-quantitative polymerase chain reaction in chronic overcirculation-induced PAH in growing piglets, with and without treatment with the dual ET receptor blocker bosentan. Pulmonary haemodynamic changes were described by pulmonary arterial resistance.

Three months after the surgical anastomosis of the left subclavian artery to the pulmonary arterial trunk, there was a shift of the impedance spectra to higher ratios of pressure and flow moduli, with increases in both 0 Hz impedance and characteristic impedance, and these changes were completely prevented by bosentan therapy. There was an increase in the expression of the serotonin 1B receptor. There was no change in the expression of the 5-HTT, and of the serotonin 2B, 1D, and 4 receptors. The overexpression of the serotonin 1B receptor was partially prevented by bosentan therapy.

The present authors conclude that this early pulmonary arterial hypertension model is characterised by an endothelin receptor-dependent increased expression of the serotonin 1B receptor.

Eur Respir J 2003; 22: 408–412.

The pathogenesis of pulmonary arterial hypertension (PAH), defined as primary pulmonary hypertension (PPH) with identifiable associated conditions [1], remains incompletely understood. Biological abnormalities have been identified in all the compartments of the pulmonary arterial wall [2]. They concern the bone morphogenetic protein-transforming growth factor β (TGF-β) signalling pathway, the serotonin system, smooth muscle cell potassium channels, extracellular matrix remodelling with increased tenasin and matrix proteinases, monoclonal endothelial cell proliferations in plexiform lesions, inflammatory reactions, activated coagulation, and an endothelium-derived vasodilator (mainly nitric oxide (NO) and prostacyclin)-vasoconstrictor (mainly thromboxane A2 and endothelin (ET)) imbalance [2]. However, it remains unknown which of these perturbations initiates the disease. There has been interest in the serotonin system, because of an epidemic of PAH related to the intake of serotoninergic appetite suppressant drugs [3] and the report of increased serotonin levels in PAH patients [4]. Serotonin causes pulmonary vasoconstriction, which is believed to be mediated mainly through serotonin receptors 5-HT1B/1D and 5-HT1A, and remodelling, which appear to require the serotonin transporter (5-HTT) [2, 5]. A series of studies point to the 5-HT1B, rather than the 5-HT2A receptor, as the mediator of serotonin-induced pulmonary vasoconstriction [5], while an overexpression of the 5-HTT has been reported in lung tissue from PPH patients [6].

Much of the difficulties in unravelling the pathobiology of PAH result from the impossibility of studying early stages of the disease. The current authors recently reported on the Blalock-Taussig procedure in growing piglets as a model of congenital left-to-right shunting-associated early PAH, characterised by significant pulmonary hypertension, medially hypertrophy, and overexpression of the ET, NO, and vascular endothelium growth factor systems, that could apparently be completely prevented by the dual ET-A and ET-B receptor blocker bosentan [7].

While the publication of this work was in progress, a report drew attention to the possible determinant role of the 5-HT2B receptor [8]. The present authors therefore re-investigated lung tissue from the experiments to investigate the expression of all possible components of the serotonin system. Since the haemodynamic data had been stored as high fidelity pressure and flow signals, the pulmonary vascular changes were also reanalysed with impedance spectra calculations, which could possibly be more sensitive to pulmonary vascular remodelling than pulmonary vascular resistance (PVR) determinations [9].

*Laboratory of Physiology, Free University of Brussels. #Unit of Diabetes and Nutrition, Catholic University of Louvain. ^Dept of Intensive Care of the Erasme University Hospital, Brussels, Belgium. *Unité de Réanimation de Chirurgie Cardiaque, Timone Hospital, Marseille, France.

Correspondence: R. Naeije
Laboratory of Physiology
Free University Brussels
Erasmus Campus CP 604
Lennik road 808
B-1070 Brussels
Belgium
Fax: 32 25554124
E-mail: rnaeije@ulb.ac.be

Keywords: Endothelin experimental pulmonary arterial hypertension pulmonary vascular impedance serotonin systemic-to-pulmonary shunting

Accepted after revision: May 22 2003
Material and methods

Thirty piglets were included in this study, which had been approved by the Committee on the Care and Use of Animals in Research of the Brussels Free University School of Medicine. Ten of the animals had a sham operation. In the other 20 animals, the left subclavian artery was anastomosed to the pulmonary arterial trunk (the so-called "Blalock-Taussig" operation). Two animals died in the postoperative course. The other 28 animals were investigated 3 months later. The placebo-treated shunted animals presented with an increase in PVR and small arterioles med. hypertrophy. These changes were completely prevented by bosentan therapy. The shunted animals had increased expressions of the pulmonary ET, the inducible NO, and the VGEF systems. These results have been reported elsewhere [7].

The instantaneous pulmonary pressures and flow signals were digitised with a sampling rate of 200 Hz, stored and analysed on a personal computer. For each data collection, intervals of five end-expiratory heartbeats were analysed. Pulmonary vascular impedance (PVZ) was calculated from the Fourier series expressions for pressure and flow signals as previously reported [10]. In this analysis, the impedance at 0 Hz \( Z_0 \) corresponds to total PVR, that is the ratio between mean pulmonary artery pressure \( (P_{pa}) \) and cardiac output \( (Q) \). Characteristic impedance \( Z_c \) was calculated as the average of impedance moduli taken between 2 and 15 Hz. The first harmonic modulus \( (Z_1) \) and the first harmonic phase angle were derived from PVZ spectra. Instantaneous pressure waves were separated into their forward and backward components [11]. The reflected wave was characterised by its amplitude and time intervals between the electrocardiographic R wave and the foot, the upward zero crossing, the peak and the downward zero crossing of the waves.

The haemodynamic measurements were obtained after ensuring steady-state conditions (stable heart rate and vascular pressures, which were continuously monitored) for 60 min after clamping of the shunt.

Whole pulmonary tissue messenger ribonucleic acid levels were measured by Sybr Green® (Applied Biosystems, Foster City, CA, USA) real-time-quantitative polymerase chain reaction (RTQ-PCR). Total ribonucleic acid (RNA) was extracted from frozen samples (400 mg) using TRIzol\textsuperscript{®} reaction (RTQ-PCR). Total ribonucleic acid (RNA) was extracted from frozen samples (400 mg) using TRIzol\textsuperscript{®} (Gibco Life Technologies, Paisley, UK) as outlined by the manufacturer. RNA pellets were dissolved in 400 \( \mu \)L ribonuclease (RNase) free \( \text{H}_2\text{O} \). RNA concentration was measured by ultraviolet spectrophotometry \( (\lambda=260 \text{ nm}) \). RNA concentrations were adjusted to 0.25 \( \mu \)g\text{\muL}^{-1} \) using RNase free \( \text{H}_2\text{O} \) and stored at \(-80^\circ\text{C} \). Reverse transcription (RT) was performed in a total volume of 20 \( \mu \)L, containing 1 \( \mu \)g of total RNA, 7.5 \( \mu \)M of random hexamers, Buffer 1, 220 \( \mu \)M of each deoxyribonucleotide (dNTP), 9 mM dithiotreitol, 20 U RNase inhibitor (Applied Biosystems), and 50 U Superscript enzyme (Gibco BRL, Life Technologies, Merelbeke, Belgium). First strand complimentary deoxyribonucleic acid (cDNA) synthesis was performed in the GeneAmp PCR system 2400 (Perkin Elmer, Foster City, CA, USA) as follows: 22°C for 10 min, 42°C for 1 h and 99°C for 5 min. After RT, final volume was adjusted to 40 \( \mu \)L using RNase free \( \text{H}_2\text{O} \). Samples were stored at \(-20^\circ\text{C} \). Using already described sequences for 5-HT1B (GenBank\textsuperscript{®} Y11867), 5-HT1D (GenBank\textsuperscript{®} Y11868), 5-HT2B (GenBank\textsuperscript{®} Z48174), 5-HT4 (GenBank\textsuperscript{®} Z48175) and 5-HTT (GenBank\textsuperscript{®} XM171519), specific pig primers were designed on Primer Express (Applied Biosystems). Using those porcine specific sequences, primers were produced on an automated synthesiser (Applied Biosystems) according to the manufacturer protocols and purified by high performance liquid chromatography. The primer sequences for Sybr Green® RTQ-PCR are summarised in table 1. Sybr Green® RTQ-PCR analysis was carried out using the ABI Prism 5700 sequence detector (Applied Biosystems). Amplification mixture (25 \( \mu \)L) containing 5 ng of cDNA, 2.5 \( \mu \)L Sybr Green® buffer, 250 \( \mu \)M dNTP, 3 mM MgCl\text{\textsubscript{2}}, 400 \( \eta \)M of each primer and 0.625 \( \text{U} \) of Amp Taq Gold Polymerase (Applied Biosystems) were processed as follows: 95°C for 10 min, followed by 40 cycles of 15 s at 95°C and 1 min at 60°C. Each cDNA was carried out twice. Each plate included tw o wells without template (no template control) for every mix to control contamination. For every mix a positive control was also included in duplicate.

For RTQ-PCR, statistical analysis was carried out using the 9 cycle threshold \( (Ct) \) value \( (Ct\text{gene of interest}-Ct\text{corresponding report gene}) \). Relative gene expression was obtained by \( \Delta \Delta Ct \) methods \( (\Delta Ct\text{sample-}\Delta Ct\text{calibrator}) \) using the sham-operated pig group as a calibrator, for comparison of every unknown sample gene expression levels. The conversion between \( \Delta \Delta Ct \) and relative gene expression levels was: fold induction=\( 2^{\Delta \Delta Ct} \) [12].

Values are reported as mean±SEM. Effects of shunt and drugs were analysed by a repeated measures analysis of variance (ANOVA). When the F-ratio of the ANOVA reached a \( p<0.05 \) critical value, post hoc comparisons were done using Scheffe’s tests [13]. A value of \( p<0.05 \) was considered statistically significant.

Results

The piglets with a systemic-to-pulmonary shunting during 3 months, had, after clamping of the shunt, a persistent pulmonary hypertension with a mean \( P_{pa} \) at 33±1 mm-Hg versus 21±1 mm-Hg in the controls. This increase in \( P_{pa} \) was completely prevented by bosentan therapy [7].

Shunt-induced pulmonary hypertension was associated with an upwards shift of the PVZ spectrum, with increases in \( Z_0, Z_1 \), and \( Z_c \), a nonsignificant displacement of the first minimum of the ratio of pressure and flow moduli to higher

| Gene | Primer sequences |
|------|------------------|
| **HPRT** | \( 5’-TCAGGCAGTATATAATCCAAAGATG-GT\text{\textunderscore}3’ \) |
| Sense | \( 5’-AGTCTGCGTTATATCCAAACTTCC\text{\textunderscore}3’ \) |
| Antisense | \( 5’-TCCTGCTCTTTCCGTGATGGA\text{\textunderscore}3’ \) |
| **5-HT1B** | \( 5’-GACCCGAGGCCCAACTGATAA\text{\textunderscore}3’ \) |
| Sense | \( 5’-GACCCGAGGCCCAACTGATAA\text{\textunderscore}3’ \) |
| Antisense | \( 5’-GTCCTAGGAGTTGGACCCA\text{\textunderscore}3’ \) |
| **5-HT1D** | \( 5’-ATAGTAAACGCCCGGACACCG\text{\textunderscore}3’ \) |
| Sense | \( 5’-ATAGTAAACGCCCGGACACCG\text{\textunderscore}3’ \) |
| Antisense | \( 5’-AGACAATGGCAATCTGGACCA\text{\textunderscore}3’ \) |
| **5-HT2B** | \( 5’-GGGTGATGTTGGACCTT\text{\textunderscore}3’ \) |
| Sense | \( 5’-GGGTGATGTTGGACCTT\text{\textunderscore}3’ \) |
| Antisense | \( 5’-TGTTGGCATTTTCCAGTGATG\text{\textunderscore}3’ \) |
| **5-HT4** | \( 5’-CCGTGCTGTACCTTCTTCTCT\text{\textunderscore}3’ \) |
| Sense | \( 5’-CCGTGCTGTACCTTCTTCTCT\text{\textunderscore}3’ \) |
| Antisense | \( 5’-TQCCGATGTTTACCCAGGTC\text{\textunderscore}3’ \) |
| **5-HTT** | \( 5’-TGGAACACTGGCAACCTGCAC\text{\textunderscore}3’ \) |
| Sense | \( 5’-TGGAACACTGGCAACCTGCAC\text{\textunderscore}3’ \) |
| Antisense | \( 5’-CCAGGTTGTTTTCCTCGG\text{\textunderscore}3’ \) |

HPRT: hypoxanthine-guanine phosphoribosyl transferase; ET-1: endothelin 1; 5-HT1B, 5-HT1D, 5-HT2B, 5-HT4: serotonin receptors; 5-HTT: serotonin transporter.

Table 1. – Primers used for real-time-quantitative polymerase chain reaction in porcine pulmonary tissue
frequencies, and no change in phase angle (fig. 1, table 2). There was an earlier return of the backward wave, which was of unchanged amplitude and duration (table 2). All these changes in PVZ spectra and wave reflection were completely prevented by bosentan therapy (fig. 1, table 2).

Shunt-induced pulmonary hypertension was associated with increased expression of the 5-HT1B receptor, from 1.00±0.39 (mean±SEM) to 2.35±0.42 units of relative gene expression (p<0.01) while the expressions of the 5-HT2B, 5-HT1D and 5-HT4 receptors remained unchanged (fig. 2). The expression of the 5-HT7 also remained unchanged. Bosentan therapy was associated with a diminished increase of the expression of the 5-HT1B receptor to 1.76±0.45 units of relative gene expression (p<0.1 versus placebo-treated pulmonary hypertensive animals, and p>0.05 versus sham-operated controls).

**Discussion**

The present results show that chronic overcirculation-induced pulmonary hypertension in growing piglets as a model of early congenital left-to-right shunting-induced PAH, is associated with an increased expression of the 5-HT1B receptor, with no significant change in the expressions of other components of the serotonin system previously implicated in the pathogenesis of PAH, such as the 5-HT2B receptor or the 5-HTT. Therapy with the dual ET-1 receptor blocker bosentan partially prevented the increase in 5-HT1B receptor expression.

Studies on the mechanisms of PAH have been limited until now by a lack of satisfactory experimental models. Most studies have been devoted to pulmonary hypertension induced by chronic hypoxic exposure, or monocrotaline injection in rats. Hypoxic pulmonary hypertension in rodents is associated with an only moderate increase in Ppa [14], and clinical hypoxic pulmonary hypertension differs from PAH in clinical presentation, histology and response to treatment [1]. Injection of monocrotaline induces severe pulmonary

| Table 2.—Pulsatile pulmonary haemodynamics in piglets 3 months after a sham operation, or systemic to pulmonary artery shunting, with intake of either a placebo or bosentan |
|-------------------------------|-------------------|-------------------|
|                             | Sham              | Shunt             |
| **Piglets n**                | 9                 | 10                | 9                 |
| **Z0** dyn.s.cm⁻⁵.m⁻²         | 524±28            | 895±35*           | 470±25*           |
| **Z1** dyn.s.cm⁻⁵.m⁻²         | 60±5              | 125±11*           | 62±5*             |
| **Zc** dyn.s.cm⁻⁵.m⁻²         | 68±6              | 112±7*            | 67±4*             |
| **fmin** Hz                  | -0.65±0.11        | -0.50±0.09        | -0.45±0.07        |
| **Amp** mm.Hg                | 3.5±0.3           | 4.3±0.5           | 3.1±0.5           |
| **tfoot** ms                 | 182±16            | 113±9*            | 128±19            |
| **tpos** ms                  | 233±19            | 168±7*            | 185±26            |
| **tpeak** ms                 | 326±18            | 253±15*           | 264±21            |
| **tneg** ms                  | 434±17            | 398±15            | 370±28            |
| **Cycle** ms                 | 506±22            | 520±30            | 428±35            |

Fig. 1.—Composite pulmonary vascular impedance spectra in piglets 3 months after a sham operation (●), or systemic to pulmonary artery shunting, with intake of either a placebo (▲) or bosentan (□). a) The shunt was associated with increases of 0 Hz impedance (Z0; *: p<0.05, shunt versus baseline; #: p<0.05 shunt versus bosentan), first harmonic impedance and characteristic impedance (Zc; : p<0.05 shunt versus baseline; : p<0.05 versus bosentan), all of which were completely prevented by bosentan therapy. b) Phase angle was not different in the three groups.

Fig. 2.—Bar graphs showing relative lung tissue messenger ribonucleic acid content for serotonin receptor (5HT1B, -1D, -2B, -4, and serotonin transporter (5HT-T) in piglets 3 months after a sham operation (●), or systemic to pulmonary artery shunting, with intake of either a placebo (▲) or bosentan (□). Shunt-induced pulmonary hypertension was associated with an increased expression of the 5-HT1B receptor, and this was partially prevented by bosentan therapy. **: p<0.01, placebo-treated pulmonary hypertensive piglets versus sham-operated controls, *: p<0.05 placebo versus bosentan-treated pulmonary hypertensive piglets.
hypertension [15], but is preceded by permeability lung oedema [16], which is not a feature of PAH. Typical PAH is a classically described complication of congenital heart disease with left-to-right shunts [1, 17].

Previous attempts to reproduce PAH associated with systemic left-to-right shunting often led to disappointingly moderate increases in \( P_{pa} \), due to poorly reactive animal species, such as dogs, or insufficient duration, pressure, or volume flow of surgically implanted shunting [18–21]. In the present study, a Blalock-Taussig operation was performed in as young as possible growing pigs, allowing for shunt flow to increase progressively with the growth of the animals, exposing the pulmonary circulation to as high as possible volume flow and pressure. This approach produced marked pulmonary arteriolar medial hypertrophy and severe pulmonary hypertension, with mean \( P_{pa} \) between 30–40 mmHg at normalised pulmonary blood flow, approaching values found in symptomatic PAH patients [7]. The observation of medial hypertrophy as the major morphological change in overcirculation-induced PAH is compatible with earlier pathological studies, which showed this aspect to be the most early change, characteristic of less severe and postoperatively reversible case [22]. Predominant localisation of pulmonary vascular changes at the smallest pulmonary arterioles explains a previously reported absence of significant changes in partitioning of PVR as determined by single occlusion, and shift of \( P_{pa}Q \) plots in parallel to higher pressures [7]. However, whether the present experimental model really represents congenital left-to-right cardiac shunt-associated PAH remains unproved in the absence of intimal and adventitial remodelling and plexiform lesions described in patients with this type of pulmonary hypertension [16, 21]. Longer periods of observation will be necessary to validate the overcirculation-induced PAH piglet model.

Standard pulmonary haemodynamic investigations rely on mean \( P_{pa} \) and mean Q determinations, with derived PVR calculations, and neglect the information contained in pulmonary pressure and flow waves [9]. This information can be apprehended using spectral analysis and derived PVZ computations [9]. Accordingly, the current authors wondered whether ET receptor blocker therapy would have completely prevented PVZ changes in the shunted piglets in which mean \( P_{pa}Q \) relationships were returned to normal [7]. In the present study, the shunted piglets presented with a shift of PVZ spectra to higher ratios of pressure and flow moduli and an increase in \( Z_C \), which is in keeping with previous observations [19]. There was also an earlier return of reflected waves. All these pulsatile haemodynamic changes are compatible with a proximal increase in pulmonary arterial elastance [23]. However, the morphometry showing predominantly peripheral medial hypertrophy, culminating at 50–100 \( \mu \)m diameter arterioles [7], would suggest that increased elastance could be entirely accounted for by increased pulmonary artery distending pressures, rather than to vessel wall remodelling.

Overcirculation-induced PAH was associated with increased circulating ET-1, increased pulmonary tissue gene expression for ET-1 and ET-B receptor, and pulmonary endothelial ET-1 protein, indicating activation of the pulmonary endothelial ET system [7]. The dual ET receptor blocker bosentan prevented this early PAH, as also reported in hypoxic- or monocrotaline-induced pulmonary hypertension [14, 15], suggesting that the ET system might be a nonspecific initiating mechanism of pulmonary arterial remodelling.

Several observations point to serotonin as a potential mediator of PAH. Serotoninergic appetite suppressants are associated with an increased risk of developing PPH [3], and plasma serotonin is increased in PAH [4]. Serotonin binds to at least 17 different subtypes [5]. Vasoconstricting effects of serotonin appear to be mediated mainly by the 5-HT2A receptors in the systemic arteries, and by the 5-HT2B or the 5-HT1D receptors in the pulmonary arteries [6]. It has been shown that the circulating levels of serotonin measured in PAH patients contract isolated human arterioles, and that this effect is mediated by the 5-HT1B receptor [24]. On the other hand, serotonin exerts potent mitogenic effects on pulmonary artery smooth muscle cells that appear to require the expression of a 5-HTT [2, 5]. This 5-HTT has been shown to be overexpressed in lungs from transplanted PAH patients [6].

Most recently LAUNAY et al. [8] reasoned that the 5-HT2B could also be involved. Indeed, the active metabolite of dexfenfluramine, which is the serotoninergic appetite suppressant drug most clearly related to an increased risk of developing PAH [3], is a selective 5-HT2B receptor. The authors observed that chronic hypoxia-induced pulmonary hypertension in mice was associated with vascular proliferation, elastase activity and TGF-\( \beta \) levels, that were potentiated by dexfenfluramine treatment. In contrast, hypoxic mice with genetically or pharmacologically inactive 5-HT2B receptors did not develop pulmonary hypertension and associated elastase and TGF-\( \beta \) changes [8]. This observation would explain why dexfenfluramine is associated with PAH while it’s inhibiting effects on the 5-HTT, shared by serotoninergic antidepressant drugs, should actually protect against pulmonary arterial remodelling.

The current authors could not find primers to investigate the expression of the 5-HT2A receptor. Therefore, the importance of the 5-HT2A receptor in this PAH model is not known. Another important limitation to the present study is that no antibodies were available to measure tissue serotonin and transporter protein levels, which would have allowed more to be known about the functional state of all the components of the serotonin receptors. In addition, the question of the nature of possible interactions between ET and serotonin receptors remains unresolved.

However, the present results are compatible with a participation of the serotonin system, essentially by means of an increased expression of the serotonin receptor 5-HT1B rather than that of the serotonin receptor 5-HT2B or of the serotonin transporter, in the most early stages of congenital left-to-right shunt-induced pulmonary arterial hypertension.

Acknowledgements. Supported by grant number 3.4567.00 from the Fonds de la Recherche Scientifique Médicale, and by a grant from the Foundation of Cardiac Surgery, Belgium. B. Rondelet was a fellow of the Erasmus Foundation, Brussels, Belgium. S. Motte is "Aspirant" from the Fonds National de la Recherche Scientifique, Belgium.

References

1. Fishman AP. Clinical classification of pulmonary hypertension. Clin Chest Med 2001; 22: 385–391.
2. Eddahibi S, Morrell N, d’Ortho MP, Naeije R, Adnot S. Pathobiology of pulmonary arterial hypertension. Eur Respir J 2002; 20: 1559–1572.
3. Abenhaim L, Moride Y, Brenot F, et al. Appetite-suppressant drugs and the risk of primary pulmonary hypertension. International Primary Pulmonary Hypertension Study Group. Eur J Med 1996; 335: 509–516.
4. Herve P, Launay JM, Scrobocchi ML, et al. Increased plasma serotonin in primary pulmonary hypertension. Am J Med 1995; 99: 249–254.
5. MacLean M, Herve P, Eddahibi S, Adnot S. 5-hydroxytryptamine and the pulmonary circulation: receptors, transporters and relevance to pulmonary arterial hypertension. Br J Pharmacol 2000; 131: 161–168.
6. Eddahibi S, Humbert M, Fadel E, et al. Serotonin transporter overexpression is responsible for pulmonary artery smooth muscle hyperplasia in primary pulmonary hypertension. *J Clin Invest* 2001; 108: 1141–1150.

7. Rondelet B, Kerbaul F, Motte S, et al. Bosentan for the prevention of overcirculation-induced experimental pulmonary arterial hypertension. *Circulation* 2003; 107: 1329–1335.

8. Launay JM, Herve P, Peoc’h K, et al. Function of the serotonin 5-hydroxytryptamine 2B receptor in pulmonary hypertension. *Nat Med* 2002; 8: 1129–1135.

9. Grant BJB, Lieber BB. Clinical significance of pulmonary arterial input impedance. *Eur Respir J* 1997; 10: 1933–1934.

10. Pagnamenta A, Bouckaert Y, Wauthy P, Brimioulle S, Naeije R. Continuous versus pulsatile pulmonary hemodynamics in canine oleic acid lung injury. *Am J Respir Crit Care Med* 2000; 162: 936–940.

11. Westerhof NP, Sipkema G, van den Bos C, Elzinga G. Forward and backward waves in the arterial system. *Cardiovasc Res* 1972; 6: 648–656.

12. Winer J, Jung CK, Shackel I, Williams PM. Development and validation of real-time quantitative reverse transcriptase-polymerase chain reaction for monitoring gene expression in cardiac myocytes in vitro. *Anal Biochem* 1999; 270: 41–49.

13. Winer BJ. Statistical principles in experimental design. 2nd Edn. New York, McGraw-Hill, 1971; pp. 514–603.

14. Eddahibi S, Raffestin B, Clozel M, et al. Protection from pulmonary hypertension with an orally active endothelin receptor antagonist in hypoxic rats. *Am J Physiol* 1995; 268: H828–H835.

15. Okada M, Yamashita C, Okada M, Okada K. Role of endothelin-1 in beagles with monocrotaline-induced pulmonary hypertension. *Circulation* 1995; 92: 114–119.

16. Plestina R, Stoner HB. Pulmonary edema in rats given monocrotaline pyrrole. *J Pathol* 1972; 106: 235–249.

17. Hoffman JI, Rudolph AM, Heymann MA. Pulmonary vascular disease with congenital heart lesions: pathologic features and causes. *Circulation* 1981; 64: 873–877.

18. Rendas A, Lennox S, Reid L. Aorta-pulmonary shunts in growing pigs. Functional and structural assessment of the changes in the pulmonary circulation. *J Thorac Cardiovasc Surg* 1979; 77: 109–118.

19. De Canniere D, Stefanidis C, Brimioulle S, Naeije R. Effects of a chronic aortopulmonary shunt on pulmonary hemodynamics in piglets. *J Appl Physiol* 1994; 77: 1591–1596.

20. Hopkins RA, Hammon JW, McHale PA, Smith PK, Anderson RW. Pulmonary vascular impedance analysis of adaptation to chronically elevated blood flow in the awake dog. *Circ Res* 1979; 45: 267–274.

21. Parviz M, Bousamra M, Chammas JH, et al. Effects of chronic pulmonary recirculation on pulmonary vasomotor tone. *Ann Thorac Surg* 1999; 67: 522–527.

22. Heath D, Edwards JE. The pathology of pulmonary vascular disease: a description of six grades of structural changes in the pulmonary arteries, with special reference to congenital cardiac septal defects. *Circulation* 1958; 18: 533–547.

23. Nichols WW, O’Rourke MF. McDonald’s Blood Flow in Arteries. 4th Edn. London, Edward Arnold, 1998.

24. Morecroft I, Healey RP, Prentice HM, Kirk A, MacLean MR. 5-HT receptors mediating contraction in human small pulmonary arteries: importance of the 5-HT1B receptor. *Br J Pharmacol* 1999; 128: 730–734.