Anti-hypertensive vasodilatory action of *Gynura procumbens* mediated by kaempferol 3-O-rutinoside [version 1; peer review: 2 approved with reservations]

Syahirah Shahlehi, Aziemah Azizi, Asrin Tengah, Siti Nornadhirah Amdani, Mark I. R. Petalcorin

PAPRSB Institute of Health Sciences, Universiti Brunei Darussalam, Jalan Tungku Link, Gadong, BE1410, Brunei

**Abstract**

**Introduction:** *Gynura procumbens* (GP), otherwise known as longevity spinach or “Sambung Nyawa” in Malay, is an evergreen herb found in Africa and Southeast Asian countries (including Brunei) used traditionally to treat various diseases such as fever, diabetes and hypertension. We examined GP’s vasodilatory action to determine its possible role via the cholinergic-mediated pathway.

**Methods:** GP leaves were prepared by filtration and evaporation to obtain the aqueous (AEGP) and methanol (MEGP) extracts followed by screening for phytochemical constituents. The total phenol, total flavonoid and flavonol contents were determined using the corresponding Folin–Ciocalteau, and aluminium colorimetric methods and the presence of kaempferol 3-O-rutinoside in the extracts was detected using HPLC analysis. Organ bath studies were conducted to determine the vasodilatory activity using intact and denuded isolated rat aortic rings by exposure to either increasing concentration of extracts (0.25, 0.5, 1.0, and 2.0 mg/mL) or 10 µg/mL kaempferol 3-O-rutinoside in the presence or absence of acetylcholine (ACh) after pre-contraction by noradrenaline (NA).

**Results:** MEGP contained more phytochemical constituents and higher content of total flavonoid and total flavonol but less phenolic content than AEGP. Furthermore, MEGP yielded a 20% elevated amount of kaempferol 3-O-rutinoside than AEGP. Both extracts significantly amplified ACh-endothelium dependent vasodilation and mediated relaxation at 1 mg/mL in endothelium-intact and endothelium-denuded aortic rings with MEGP as a more effective vasodilator than AEGP. Overall, these results imply the involvement of extracts in potentiating cholinergic pathway, which might be mediated by kaempferol, as shown by its vasorelaxation effects in endothelium-intact and –denuded aorta.

**Conclusions:** The present findings demonstrate that the vasodilatory activities of the two *Gynura procumbens* extracts, AEGP and MEGP, in...
thoracic aorta rings isolated from rats are potentially mediated via a cholinergic pathway through the action of a flavonoid particularly kaempferol 3-O-rutinoside.

**Keywords**
Gynura procumbens, Hypertension, Vasodilation, Cholinergic pathway, Kaempferol, Acetylcholine, Muscarinic receptors
Introduction

Hypertension persists as high arterial pressure with systolic and diastolic blood pressure (BP) exceeding 140 and 90 mmHg, respectively\(^1\) contributing to cardiovascular diseases (CVDs), which are the leading cause of death globally. An estimated 17.9 million people die annually from CVDs, representing 31\% of all global deaths\(^2\). More than 90\% of hypertension is termed as essential or primary hypertension with unknown causes but influenced by environmental, genetic and behavioural factors\(^3\), including high salt intake, smoking, age, gender, obesity, diabetes, sedentary lifestyles, chronic alcohol consumption and family history of hypertension\(^4,5\). The remaining 10\% of the cases fall under secondary hypertension caused by potentially reversible biochemical or mechanical pathology associated with renal or adrenal diseases\(^6,7\).

A continuous effort has been made to search for a safer and more effective method to control hypertension worldwide. Herbal medicines have been used since the beginning of human civilisation to control hypertension and other cardiovascular disorders including congestive heart failure, atherosclerosis, cerebral and venous insufficiency, arrhythmia and ischemic heart diseases\(^8\). Traditional herbs have become significant sources for drug research in pharmaceutical settings as alternatives to conventional synthetic drugs such as thiazide and captopril for hypertension\(^9\). *Gynura procumbens* (longevity spinach or “Sambung Nyawa”) is an evergreen herb that is used traditionally in Africa and Southeast Asia including Brunei, Indonesia, Thailand and Malaysia to treat various diseases such as hypertension, rashes, kidney disease, fever and diabetes\(^10\)-\(^13\). Belonging to the Asteraceae family, this plant is small in size (around 1-3 m in height) with oval-shaped green leaves, fleshy stems and purple tint\(^14\), and can thrive in partial, shaded or full sun as long as the soil is moist. The leaves of this plant are traditionally consumed as salad, juice or tea for health benefits.

Various studies have demonstrated the antihypertensive activity of GP both in-vitro and in-vivo\(^10\)-\(^13\),\(^15\)-\(^16\), acting to lower the blood pressure by stimulating vasodilation besides heart stabilisation and diuretic effects. However, its mechanism of inducing vasodilation is not well understood. Activation of the parasympathetic division of autonomic nervous system induces relaxation of the blood vessel\(^17\) via the cholinergic pathway by releasing acetylcholine (ACh) from the nerve terminal in blood vessel walls, which in turn stimulates nitric oxide (NO) production that triggers dilation of vascular smooth muscle. The identity of GP’s active ingredient for vasodilation is likely to be the flavonoids, which may be responsible for its antihypertensive effect\(^10\)-\(^13\),\(^18\),\(^19\). Kaempferol is a flavonoid that is present in the sub-fraction of the ethanolic extract of GP\(^20\)-\(^21\), and kaempferol 3-O-rutinoside is one of the various types of kaempferol that has been identified in GP\(^4\). In this present study, we investigated GP’s vasodilatory action and, then, determined its possible activation of the cholinergic pathway mediated by kaempferol 3-O-rutinoside.

Methods

Chemicals and Reagents

All chemicals used for methanol extraction, phytochemical content determination including total phenolic, flavonoid, flavonol and high liquid performance liquid chromatography (HPLC) were obtained from Merck, Germany except for Dragentorff’s reagent, mercuric chloride, potassium iodide, iodine, quercetin and kaempferol 3-O-rutinoside, which were purchased from Sigma Aldrich, USA. All reagents for organ bath studies including those to prepare Kreb’s Henseleit solution (Kreb’s solution), sodium chloride (NaCl), potassium chloride (KCl), potassium dihydrogen phosphate (KH\(_2\)PO\(_4\)), glucose, sodium hydrogen carbonate (NaHCO\(_3\)) and calcium chloride (CaCl\(_2\)) were purchased either from Sigma Aldrich or Merck.

Preparation of solution

Kreb’s solution was prepared by dissolving compounds in distilled water and adding calcium chloride last with continuous stirring to eliminate the formation of a salt precipitate. For the organ bath study, drugs used were dissolved following the manufacturer’s instructions and GP extracts were dissolved in Kreb’s solution. For HPLC experiment, kaempferol 3-O-rutinoside was dissolved in methanol.

Plant collection and verification

GP was purchased from Rimba Horticulture Centre, Brunei Darussalam. Leaf samples were then verified by Dr Johan Willem Frederick Slik (Associate Professor in Phylodiversity, Faculty of Science, Universiti Brunei Darussalam). The herbal specimen (dried leaves) was deposited into Universiti Brunei Darussalam Herbarium (UBDH) with a sample number 17H8182.

Drying of *G. procumbens* leaves and powdered preparation

Fresh leaves of GP were cleaned, weighed, and oven-dried at 60°C for three days. The dried leaves were then crushed using a blender (Waring 800s, USA) to produce fine powder and filter-sieved in dry form using a mechanical sieve (0.3 mm mesh hole size).

Preparation of aqueous (AEGP) and methanol extracts of *G. procumbens* (MEGP)

Aqueous extract of *Gynura procumbens* (AEGP) was prepared as described previously\(^22\). Briefly, the powdered leaves were macerated twice in distilled water (1:10 ratio of solute to solvent) and kept at 60°C water bath for three days. After filtration, the filtrate was centrifuged at 3000 rpm for 20 min, and the supernatant was recovered, and frozen at -80°C overnight. The sample was then lyophilised by freeze-drying for three days to yield a brown powder of AEGP\(^23\) that was stored at 4°C until further use.

MEGP was prepared as described previously\(^24\) with slight modifications, using ultrasonic extraction (UE) with 95\% methanol in 300 W ultrasonic bath. Briefly, the powdered leaves were
placed in a beaker and added with methanol at a 1:20 (solute: solvent) ratio before subjected with ultrasound at 25°C for 30 min. After extraction, the sample was filtered using filter paper, and the powder residues were added again with methanol of 1:20 (solute: solvent) ratio in another cycle of ultrasound. The filtrate obtained was evaporated with a rotary evaporator (Yamato RE301, Japan) at 63°C, and left to dry at 60°C in a drying cabinet for five days yielding a dark greenish-brown sticky solid MEGP that was stored at 4°C until further use. The percentage of yield was calculated using the following equation: \[ \% \text{ Yield} = \frac{\text{Mass of plant crude extract}}{\text{Mass of powder plant in thimble}} \times 100\%. \]

**Phytochemical screening**

The extracts were tested using the standard procedures for phytochemical constituents, including alkaloids, glycosides, flavonoids, saponins, steroids, triterpenoids and tannins. For stock extract solution preparation, AEGP and MEGP (1 g each) were completely dissolved in 100 mL of water and methanol, respectively. The stock solutions obtained were used for phytochemical screening whenever necessary as some of the methods required the solid form of the extracts. Qualitative results were expressed as (+) for the presence and (-) for the absence of phytochemicals. Each test was done by visual observation of colour change or formation of a precipitate after the addition of specific reagents. Hence, the experiments were only carried out once.

**Test for alkaloids**

Around 15 mg of each extract was separately stirred in 6 mL of 1% HCl at room temperature water bath for 5 min and then filtered using Whatman filter paper (grade 42, 55 mm). The filtrates were divided into two equal parts and used for Dragendorff’s and Mayer’s tests. The orange-red precipitate formation indicated the presence of alkaloids using Dragendorff’s test upon addition of 1 mL of Dragendorff’s reagent (potassium bismuth iodide solution) to the first extract filtrate. Using Mayer’s test, the presence of alkaloids was indicated by the cream-coloured precipitate or turbid solution formation upon addition of 1 mL of Mayer’s reagent (potassium mercuric iodide solution) to the second filtrate.

**Test for flavonoids**

The presence of flavonoids was detected by the formation of an intense yellow solution that turned into colourless upon addition of a few drops of 1 M HCl to a mixture of 1 mL of each extract and 0.2 mL of 1 M NaOH.

**Test for glycosides**

Using Borntrager’s test, the presence of anthraquinone glycosides was detected by the formation of rose-pink to red colour ammoniacal layer upon addition of diluted (10%) ammonia to the lower layer of chloroform that was recovered from boiling and filtering a mixture of 1 mL of each extract, 1 mL of 5% sulphuric acid (H₂SO₄) and 2 mL chloroform.

**Test for saponins**

The presence of saponins was detected by the formation of a topmost foamy layer upon thoroughly mixing by hand a dilution of 1 mL of each extract in 20 mL of distilled water for 15 min.

**Test for steroids**

The presence of steroids was detected by the colour change of the upper layer into red and sulphuric acid (H₂SO₄) layer, and into fluorescent yellow upon addition of 10 mL of chloroform and an equal volume of concentrated H₂SO₄ into 1 mg of each extract.

**Test for tannins**

The presence of tannins was detected by the development of precipitate upon addition of 1% potassium dichromate solution to an equal volume of each extract.

**Test for triterpenoids**

The formation of reddish-violet colour detected the presence of triterpenoids upon addition of 2 mL of chloroform, 1 mL of acetic anhydride and 1 mL of concentrated H₂SO₄ to 5 mg of each dry extract.

**Determination of total phenolic, total flavonoids and total flavonol contents**

**Total phenolic content.** The total phenolic content of each plant extract was determined using Folin–Ciocalteau assay. Briefly, 20 μL of extracts, 100 μL of 2 N Folin–Ciocalteu phenol reagent and 1.58 mL of distilled water were mixed thoroughly. After 8 min, 300 μL of Na₂CO₃ was added into the mixture and allowed to react at room temperature for two hours before measuring the absorbance at 765 nm using a spectrophotometer. A standard calibration curve of gallic acid (0-4 mg/mL) was prepared, and results were expressed as mg of gallic acid equivalent (GAE) per g of extract. Experiments were replicated four times.

**Total flavonoid content.** Total flavonoid content of each extract was determined using aluminium colourimetric method using quercetin (0-200 μg/mL) as a standard. Briefly, 0.5 mL of extract and 0.5 mL of the standard were placed in separate test tubes with 0.1 mL of 10% aluminium chloride, 0.1 mL of 1 M potassium acetate, 1.5 mL of 80% methanol and 2.8 mL of distilled water, and mixed thoroughly. A blank was prepared in the same manner where 0.5 mL of distilled water was used instead of the sample or standard, and the aluminium chloride replaced by distilled water. All tubes were incubated at room temperature for 30 min, and the absorbance was measured at 415 nm using the spectrophotometer. Experiments were carried out four times. Results were expressed as mg of quercetin equivalent (QE) per gram of extract.

**Total flavonol content.** Total flavonol content of each extract was analysed using aluminium chloride colourimetric method as described previously. Briefly, quercetin was used to make
a standard calibration curve ranging from 0 to 100 μg/mL. The test tube containing 1 mL of extract or 1 mL of standard solution was added with 1 mL of 2% aluminum chloride and 3 mL of 5% sodium acetate and then mixed well. The mixture was then centrifuged at 3000 rpm for 20 min to get a clear solution. The absorbance of the standard and sample was measured at 440 nm using the spectrophotometer. Experiments were replicated four times. Results were expressed as mg of quercetin equivalent (QE) per gram of extract.

High-performance liquid chromatography (HPLC) analysis of kaempferol-O-rutinoside

HPLC analysis was performed as previously described using Agilent 1200 series HPLC system (Agilent LabX, USA) equipped with an autosampler and coupled to a variable UV wavelength detector. Liquid chromatography was performed on a reversed-phase C18 column (internal diameter 4.6 mm, height 250 mm, and particle size 5 mm). The mobile phase consisted of two solvents [A] 5% CH₃COOH, acetic acid and [B] CH₃CN, acetonitrile) using the following elution program: 0–10 min 70% A isocratic, 10–20 min gradient to 40% A and then 20–30 min 40% A isocratic again with a flow rate of 1.0 mL/min at 30°C to detect peaks at 367 nm. Standard calibration was done using kaempferol 3-O-rutinoside at a concentration ranging from 0 μg/mL to 100 μg/mL. After the calibration curve was generated, both extracts with or without spiking with kaempferol 3-O-rutinoside (100 μg/mL) were run using the same elution program. Experiments were carried out in triplicates.

Animals

A total of 25 adult male Sprague Dawley (SD) rats at age of 16–20 weeks, weighing from 250 to 300 g, were obtained from Universiti Brunei Darussalam animal house. The animals were housed in standard environmental conditions (24°C, 60–70% humidity) on a 12 hour-light-dark cycle feeding. The animals were given standard rat pellets and tap water ad libitum.

Statement on ethical issues

The study was conducted following the ethical guidelines of the Laboratory Animal Care (NIH publication #85-23, revised in 1985) and approved by the University Research Ethics Committee (UREC) of PAPRSB Institute of Health Sciences, Universiti Brunei Darussalam.

Vessel preparation and setting-up of the organ bath

After sacrificing the rats by cervical dislocation, their descending thoracic aorta was removed and placed in a petri dish on ice containing Kreb’s buffer (118.2 mM of NaCl, 4.7 mM of KCl, 2.5 mM of CaCl₂, 25 mM of NaHCO₃, 1.2 mM of MgSO₄, 1.2 mM of KH₂PO₄, and 11.7 mM of glucose). The rat aorta was dissected, cleared of fats including connective tissues, and cut into rings of 3 mm long. The denudated endothelium was prepared by inserting fine forceps into the lumen of the aorta with gentle rotation. The aortic rings were mounted using two triangular-shaped platinum wire holders in the vessel lumen and suspended horizontally in an organ bath with 10 mL of Kreb’s buffer, thermoregulated at 37°C, and bubbled with carbogen to a resting tension of 1.0 g (Figure 1A). One of the holders was fixed to the bottom of the chambers, and the other was connected to a force-displacement transducer (Grass® Force Displacement Transducer FT03) for measuring isometric tension. The rings were equilibrated at 1.0 g resting tension for 20 min, with bathing solution replacement every 10 min, and tension readjustment to 1.0 g if necessary. The aortic rings were tested for viability using 40 mM KCl before conducting experiments (Figure 1B). Isometric contraction and relaxation were recorded using a Force Displacement Transducer. The transducer signals were displayed, stored and analysed on a computer using AD Instruments model 4/25T with Power Lab Software version 7. The responses (in grams) were recorded using LabChart Reader 7.

Experimental protocols

The following experiments were conducted using both endothelium-intact and endothelium-denuded aorta, except for the protocol on the addition of noradrenaline (NA) with the extracts or kaempferol, whose effects were previously determined and ruled out as NA-independent regardless of the tissue preparation.

Effects of AEGP and MEGP on aortic rings pre-contracted with NA

The aortic rings were pre-contracted by adding 50 nM of NA to induce transient and stable contraction for 60 min to generate the first response curve. Then, the aortic rings were washed with Kreb’s buffer three times, allowing the baseline to stabilise for 10 min before pre-contracted again with 50 nM of NA. After the tonic response or contraction became stable for 20 min, increasing concentrations of AEGP or MEGP (0.25, 0.5, 1 and 2 mg/mL) were added cumulatively at 10 min interval to obtain the second response curve.

Effect of AEGP and MEGP on aortic rings pre-contracted with NA

The aortic rings were pre-contracted with 50 nM of NA for 10 min, followed by addition of 100 μM of ACh for 50 min to obtain the first response curve. Then, the aortic rings were washed with Kreb’s buffer three times, allowing the baseline signal to stabilise for 10 min. The aortic rings were pre-contracted again with 50 nM of NA for 10 min followed by addition of 100 μM of ACh for another 10 min before adding AEGP or MEGP cumulatively in increasing concentrations (0.25, 0.5, 1 and 2 mg/mL) every 10 min to obtain the second response curve.

Effect of kaempferol 3-O-rutinoside on NA-induced contraction

The aortic rings were pre-contracted with 50 nM of NA for 30 min to obtain the first response curve. Then, the aortic rings were washed with Kreb’s buffer three times, allowing the basal tension signal to stabilise for 10 min. The rings were pre-contracted again with 50 nM of NA for 10 min, followed by addition of 10 μg/mL kaempferol 3-O-rutinoside for 20 min to generate the second response curve. This experiment was to rule out the role of kaempferol in enhancing the ACh-mediated vasodilation.

Effect of kaempferol 3-O-rutinoside on ACh-mediated vasodilation

The aortic rings were pre-contracted with 50 nM of NA for 10 min, followed by addition of 100 μM of ACh for 20 min to obtain the first response curve. Then, the aortic rings
were washed with Kreb’s buffer three times, allowing the basal tension signal to stabilise for 10 min. The rings were pre-contracted again with 50 nM of NA followed by addition of 100 μM of ACh before adding 10 μg/mL kaempferol at 10 min interval to generate the second response curve.

**Figure 1. Isolated organ bath system.** (A) Schematic set-up of the organ bath. The aortic rings were mounted using two-triangle-shaped platinum wire holders, suspended horizontally in an organ bath containing 10 mL of the above solution, thermoregulated at 37°C, and bubbled with carbogen with a resting tension of 1 g. Isometric contraction and relaxation were recorded using Force Displacement Transducer. Transducer signals were displayed and stored on a computer. (B) A summarised protocol of one of the organ bath experiments. Before each experiment, the aortic rings were stimulated with 40 mM KCl to produce a contraction, which represents a reference response, ensuring the viability of the rings just before the start of the experiment. Then the buffer solution was replaced for three times, and baseline was stabilised before adding 50 nM of NA for 10 min to induce stable contractions, and 100 μM of ACh was added for 50 min afterwards to produce the first curve. Then, the aortic rings were washed with Kreb’s buffer three times, allowed to stabilise, then added again with 100 μM ACh for 50 min in pre-contracted rat-aorta before adding extracts cumulatively to obtain the second curve.

**Statistical analysis**
Calculations and statistical analysis were performed using GraphPad Prism (Version 6.0) software. Contractile responses were expressed in grams of tension (g). The vasorelaxation due to ACh, the addition of extracts and kaempferol 3-O-rutinoside,
was expressed as a percentage relaxation in response to contraction caused by NA. All results were expressed as mean ± standard error of the mean (SEM) per experimental group with n representing the number of aortae used in each experiment. Data were analysed using two-way analysis of variance (ANOVA), which was followed by Dunnett’s test for significant differences between the control and the experimental treatments, except for the data of kaempferol 3-O-rutinoside with the addition of Ach, which was analysed using unpaired Student’s t-test. The line graph for determining the potent vasodilator between the two extracts was constructed using linear regression. P-values of less than 0.05 (P<0.05) were considered statistically significant.

Results
MEGP produced higher percentage yield with more flavonoids and flavonols than AEGP
The percentage yield extracted from powdered leaves of GP using aqueous extraction (AEGP) or methanol extraction (MEGP) is shown in Table 1. After drying the extracted fluid, MEGP gave a higher percentage yield of 8.7%, producing a dark greenish-brown sticky solid material, while AEGP yielded 5.5%, generating a light brown fluffy material. The phytochemical analysis is presented in Table 2, which shows that flavonoids and triterpenoids were detected in both AEGP and MEGP while saponins, steroids and tannins were only seen in MEGP. On the other hand, alkaloids were only detected in AEGP but not MEGP. Glycosides were absent in all extracts. Table 3 presents the total phenolic, flavonoid and flavonol contents in both types of GP extracts stated as mg GAE/g extract for total phenolic content and mg QE/g extract for total flavonoid and flavonol contents. AEGP has higher total phenolic content of 28.23±4.68 mg GAE/g extract, which is twice as that of MEGP (13.82 ± 2.42 mg GAE/g extract). In contrast, MEGP has four times more total flavonoid (10.37 ± 0.07 mg QE/g extract) and two times more total flavonol (4.72 ± 0.04 mg QE/g extract) content than AEGP (2.52 ± 0.21 and 2.35 ± 0.03 mg QE/g extract, respectively). The total phenolic content of GP extracts was based on the standard gallic acid curve obtained at y=0.4332x+0.0339 (r²=0.993), while the total flavonoid and flavonol contents were calculated from a standard quercetin curve using the linear regression equations y=0.0029x+0.0458 (r²=0.995) and y=0.0045x+0.0339 (r²=0.996), respectively.

Kaempferol 3-O-rutinoside was detected in both extracts at a higher percentage in MEGP than AEGP
The results of HPLC screening using a reverse-phase C-18 column with a gradient system of acetic acid-acetonitrile combination as the mobile phase for both extracts revealed that when using the spiking technique with the standard reference compound of 100 μg/mL kaempferol 3-O-rutinoside, the height of peaks for both extracts was explicitly increased with the retention time observed at 1.4 min after injection. The amount of kaempferol 3-O-rutinoside calculated based on the difference of the height of peaks at 1.4 min between the standard reference and the extracts were 57.86 μg/mL for AEGP and MEGP, respectively. Figure 2 shows the actual HPLC chromatograms for AEGP and MEGP, with and without spiking of 100 μg/mL kaempferol 3-O-rutinoside as the standard reference.

Both extracts mediated relaxation in aortic rings pre-contracted with NA in organ bath studies
Both AEGP and MEGP extracts were cumulatively added at 10 min interval in increasing concentrations (0.25, 0.5, 1 and 2 mg/mL). Addition of these extracts started at 20 min after the aortic rings were pre-contracted with 50 nM of NA. At the lowest concentration, both AEGP and MEGP produced a

Table 1. Percentage yield of aqueous extract of Gynura procumbens (AEGP) and methanoic extract of Gynura procumbens (MEGP).

| Extract type | Description of the solid extract obtained | Yield (%) |
|--------------|------------------------------------------|-----------|
| AEGP         | A light brown fluffy material             | 5.5       |
| MEGP         | A dark greenish-brown sticky solid material | 8.7       |

Table 2. Phytochemical screening of aqueous extract of Gynura procumbens (AEGP) and methanoic extract of Gynura procumbens (MEGP).

| Constituents | Specific test   | Results | Aqueous | Methanol |
|--------------|----------------|---------|---------|----------|
| Alkaloids    | Dragendorff's test | -       | -       |
|              | Mayer's test     | +       | -       |
| Flavonoids   | Sodium hydroxide test | +       | +       |
| Glycosides   | Borntrager's test | -       | -       |
| Saponins     | Frothing test    | -       | +       |
| Steroids     | Salkowski test   | -       | +       |
| Tannins      | Potassium dichromate test | -       | +++     |
| Triterpenoids|                | +++     | ++      |

*: weak presence, **: moderate presence, +++: strong presence, and -: Absent.

Table 3. Total phenolic, flavonoid and flavonol contents of aqueous extract of Gynura procumbens (AEGP) and methanoic extract of Gynura procumbens (MEGP). Values are expressed as mean of mg of GAE or QE per gram of extract ±S.E.M; n=4.

|                | Total phenolic content | Total flavonoid content | Total flavonol content |
|----------------|-----------------------|-------------------------|------------------------|
| AEGP           | 28.23±4.68            | 2.52±0.21               | 2.35±0.03              |
| MEGP           | 13.82±2.42            | 10.37±0.07              | 4.72±0.04              |

GAE: Gallic acid equivalent, QE: Quercetin equivalent, G. procumbens: Gynura procumbens, SEM: Standard error of the Mean.
minimum relaxation of 12 ± 4.46% and 18 ± 3.22%, respectively (Figure 3). At the highest concentration, both AEGP and MEGP produced a maximum relaxation of 95 ± 4.53% and 112 ± 6.07%, respectively (Figure 3). In the pattern of relaxation induced by both extracts, the percentage relaxation of NA-induced contraction in rat aorta was significantly higher when exposed to increasing concentration of GP extracts, especially from 0.5 mg/mL to 2 mg/mL for both extracts, as compared with that of the control NA alone (Figure 3).

Both extracts enhanced ACh-mediated response in intact and denuded endothelium rat aorta

In endothelium-intact rat aorta, both AEGP and MEGP extracts were cumulatively added at 10 min interval in increasing concentrations (0.25, 0.5, 1 and 2 mg/mL) at 10 min interval, starting at 20 min after adding 100 μM of ACh. Addition of these extracts started at 30 min following addition of 100 μM of ACh. At the lowest concentration, both AEGP and MEGP produced the minimum relaxation of 61 ± 13.80% and 72 ± 18.40%, respectively (Figure 4A). At the highest concentration, both AEGP and MEGP produced the maximum relaxation of 87 ± 4.30% and 112 ± 12.3%, respectively (Figure 4A). Relaxation induced by both extracts was significantly different from ACh control alone, especially at 0.5 mg/mL for AEGP and 1 mg/mL for both AEGP and MEGP (Figure 4A).

In endothelium-denuded rat aorta, both AEGP and MEGP extracts were separately added in increasing and cumulative concentrations (0.25, 0.5, 1 and 2 mg/mL) at 10-min intervals. In both cases, the addition of these extracts started at 30 min following addition of 100 μM of ACh. At the lowest concentration, both AEGP and MEGP produced the minimum relaxation of 46 ± 15.70% and 42 ± 13.10%, respectively (Figure 4B). At the highest concentration, both AEGP and MEGP produced the maximum relaxation of 79 ± 5.60% and 114 ± 8.60%, respectively (Figure 4B). Overall, there was a pattern of relaxation induced by both extracts, where AEGP significantly enhanced relaxation at 1 mg/mL, while MEGP enhanced relaxation at a wider range of 0.5 mg/mL to 2 mg/mL (Figure 4B).

**MEGP was more potent vasodilator than AEGP**

Exposure of endothelium-intact rat aorta to either extract when added cumulatively produced a similar vasodilatory effect as a reduction in the percentage of NA peak response.
Exposure to increasing concentration of AEGP up to 1 mg/mL reduced the contraction of rat aorta from 39 ± 13.9% to 0 ± 17.1%, and MEGP from 40 ± 18.4% to 5 ± 14.0%. ACh alone without the addition of extracts still showed a 23 ± 4.90% contraction. However, at 2 mg/mL, both extracts had a differential effect, in which MEGP caused further reduction from 5 ± 14.0% to 0 ± 12.3% contraction, whereas AEGP produced a rise of contraction from 0 ± 17.1% to 18 ± 10.2%.

Exposure of endothelium-denuded rat aorta to either extract also produced a similar vasodilatory effect, in which the percentage contraction of NA peak response at concentrations reaching up to 1 mg/mL was reduced from 65 ± 9.35% to 0 ± 6.60% for AEGP and 72 ± 13.1% to 21 ± 9.88% for MEGP, respectively. In comparison, ACh alone without the addition of any extract was only able to reduce the contraction from 80 ± 9.31% to 50 ± 7.58%. However, at 2 mg/mL both extracts had a different effect in which MEGP caused a further reduction of contraction from 21 ± 9.88% to 0 ± 8.64%, whereas AEGP produced a rise of contraction from 0 ± 6.60% to 8 ± 5.61%. Overall, Figure 5 demonstrates the vasodilatory action of both GP extracts, expressed as the normalised data for the
Effect of both AEGP and MEGP observed as a reduction in the contraction of endothelium-intact and denuded aortic rings that were pre-contracted with 50 nM of NA.

Kaempferol 3-O-rutinoside reduced NA-mediated contraction

To elucidate the role of kaempferol-3-O-rutinoside in inducing relaxation, we first determined the vasodilatory effect of kaempferol without the application of ACh in the pre-contracted aorta rings. Kaempferol 3-O-rutinoside was added for 20 min, which started at 10 min after pre-contraction with 50 nM of NA. Addition of 10 μg/mL of kaempferol 3-O-rutinoside caused rapid relaxation (Figure 7), which was significantly increased from 9.8 ± 1.56% to 84.9 ± 18.7% at 20 min and from 17.5 ± 5.24% to 94.6 ± 5.59% at 30 min, respectively, as illustrated in Figure 6.

Kaempferol 3-O-rutinoside enhanced the ACh-mediated response in endothelium-intact and -denuded rat aorta

The pre-contraction of aortic rings with intact and denuded endothelium were supplemented with 10 μg/mL of kaempferol 3-O-rutinoside at 20 min after addition of 100 μM of ACh to determine the involvement of kaempferol 3-O-rutinoside in potentiating the cholinergic pathway. Addition of kaempferol 3-O-rutinoside significantly enhanced the vascular relaxation evoked by ACh in rat aorta with and without the endothelium. Kaempferol 3-O-rutinoside elicited a maximum relaxation of 76.0 ± 4.66% and 70.7 ± 0.73% with or without endothelium, respectively. In comparison, the control produced relaxation of 41.7 ± 6.47% and 19.9 ± 6.69%, with and without endothelium, respectively (Figure 7).

Raw data for each experiment performed in this study are available, see Underlying data.

Discussion

Various studies have been carried out to identify the antihypertensive ability of GP extracts\cite{8-10,13-16}. The presence of bioactive compounds in GP is believed to contribute to its beneficial properties such as flavonoids, phenolic acids, alkaloids, terpenoids, glycosides and sterols\cite{12}. In this present study, the antihypertensive property of both crude extracts of AEGP and MEGP was investigated in vitro to determine whether they were able to potentiate the cholinergic mediated vasodilation. Limited studies showed the possible contribution of the cholinergic pathway in enhancing the relaxation effect of GP extracts. Successful extractions with water or methanol from the dried and powdered leaves of GP were achieved, producing the fluffy brown powder and the sticky dark greenish-brown substance as yields for AEGP and MEGP, respectively. The characteristics of AEGP in our study were similar to those reported by Lee et al.\cite{23} and Kamaruzaman and Mat Noor\cite{31}. There was neither an established guideline nor any known protocols for methanol-based extraction for GP plant leaves. However, the characteristics of the dry methanol extracts, yielding a dark brown sticky solid was demonstrated by Korwar et al.\cite{32}, similar to our findings. The percentage yields for both extracts were different, which might be due to the polarity of the solvents used\cite{33}. Given an equal initial amount of fresh weight for the plant parts, the aqueous solvent yielded less extract than those extracted with the methanol solvent, suggesting that the plant GP possesses more methanol-soluble components\cite{33}. Our finding is in concordance with a study done by Njume et al.\cite{34} and Ngo et al.\cite{35}, where absolute methanol or a mixture of 50% (v/v) water with other solvents such as alcohol or acetone was recommended as the solvent of choice to produce high levels of extractable ingredients.

Our results highlighted the observation that both extracts, which were obtained using different solvents, contained...
Figure 6. The percentage of relaxation induced by kaempferol 3-O-rutinoside in the pre-contracted rat aorta in comparison with noradrenaline (NA) alone. Data presented as mean ± SEM. The n values for NA alone and NA with kaempferol 3-O-rutinoside are 5 and 3, respectively. ****P<0.05 for the addition of kaempferol 3-O-rutinoside at 20 and 30 min vs control, NA alone.

Figure 7. The percentage relaxation induced by acetylcholine (Ach) in the absence or presence of 10 μg/mL of kaempferol 3-O-rutinoside in endothelium-intact rat aortic rings pre-contracted with noradrenaline. (A) Endothelium-intact rat aorta (B) Endothelium-denuded rat aorta. Data presented as mean ± SEM. The n values for Ach alone and kaempferol 3-O-rutinoside are 6 and 4, respectively in endothelium-intact rat aorta. In contrast, the n values for Ach alone and kaempferol 3-O-rutinoside are 5 and 3, respectively in endothelium-denuded rat aorta. **P<0.05 for the addition of kaempferol 3-O-rutinoside at 30 min vs control, Ach alone in both endothelium-intact and –denuded rat aorta.

Phenolic compounds account for the majority of pharmacological actions. For several decades, phenolic compounds are regarded as having a powerful free radical scavenging activity. As many aspects of CVDs are associated with high oxidative stress, the consumption of polyphenol-rich food that combats oxidative stress can help lower the risk of chronic diseases, including cardiovascular disorders. In this present study, the total phenolic content of AEGP is higher than MEGP. However, the latter was shown to contain more total flavonoids and total flavonol content. The results obtained for the flavonoids are in agreement with the previous report on GP by Afandi et al., showing that MEGP contains higher total flavonoid content than AEGP, but the amount of phenolic content was found lowest in AEGP among all the other extract preparations. This variation could be due to differences in the process of extractions and the solvents used. Various factors could contribute to the efficiency of extraction, such as the type and concentration of solvents, pH, time, temperature and solid-liquid ratios.

Besides, the different types and amount of phenolic and non-phenolic compounds extracted are dependent on the extraction solvent used. The results showing that AEGP contained a lower percentage of total flavonoid and total flavonol compounds.
The presence of flavonoids in both extracts suggests they have a considerable role in various pharmacological effects, especially the hypotensive action, which might explain the BP-lowering observation and claims in the traditional practice of consuming the GP leaves to treat hypertension. Experimental evidence has shown that the flavonoids in GP extracts are responsible for an antihypertensive effect. Kaempferol, which is one of the significant flavonoids dominantly present in the sub-fraction of the ethanolic extract of GP, was also detected in both extracts in our study. Kaempferol 3-O-rutinoside was detected in a higher amount from MEGP than AEGP. After quantifying this active ingredient, we decided to test its vasodilatory activity in the organ bath system using rat aorta to elucidate its hypotensive contribution. We first added the extracts after NA-induced contraction of aortic rings to confirm the vasodilatory action of both extracts. Our results highlighted the vascular relaxation effect indicating the antihypertensive property of this plant and confirming reports from previous studies that GP extracts might reduce blood pressure by vasodilatation. Next, we investigated whether there is a possible contribution of the cholinergic-mediated pathway in the vasodilatory effect of the extracts. Our present finding demonstrated that the vasorelaxation activity of the extracts further enhanced the ACh-mediated vasodilation in the endothelium-intact aorta, primarily upon exposure to 0.5 mg/mL AEGP and 1 mg/mL of both AEGP and MEGP. These results support the findings of Kaur et al., which showed that ACh administration to GP-treated rats in vivo significantly reduced the mean arterial pressure (MAP) and diastolic pressure (DP) of spontaneously hypertensive rat (SHR), and that of Kim et al., which detected an increased of NO levels in the serum of Gynura procumbens water extract (GPWE)-treated SHR. Our data further revealed that the extracts significantly exerted their relaxation effect in the endothelium-denuded aorta, suggesting that the vasodilatory action of AEGP and MEGP extracts is also mediated directly through the vascular smooth muscles. The vascular smooth muscle cells are located within the media of the vascular bed, directly modulating the vascular tone primarily via the changes in sarcoplasmic free Ca²⁺ concentration. Various signalling mechanisms could trigger the response of the smooth muscles through an ACh endothelium-independent pathway as the adrenergic pathway. In the adrenergic pathway, α₁ and α₂ adrenoceptors mediate contraction of the smooth muscles by coupling to Gq and activating phospholipase C (PLC), thereby increasing the concentration of Inositol 1,4,5-triphosphate (IP₃), which in turn, causes the rise in intracellular Ca²⁺ concentration. A previous study showed that GP extracts could mediate relaxation via α₁ receptors in vivo. PE (an α adrenoceptor agonist) and methoxamine (a selective α₁ adrenoceptor agonist) induced a significant decrease in presor response of GPWE-treated rats. From our findings, we speculate that the GP extracts might also mediate relaxation via adrenergic pathway by activating α and β adrenoceptors. A more favored explanation for this action, however, is based on the evidence that we obtained so far, which pointed towards the cholinergic pathway. It is known that ACh fails to induce relaxation when the endothelium is denuded and removed, suggesting that vasorelaxation of smooth muscle is inhibited when the formation of NO is impaired due to activation of M₁ and M₃ receptors, hence causing contraction instead. Thus, we focused more on investigating the cholinergic endothelium-independent pathway via stimulation of M₁ and M₃ muscarinic receptors that inhibit the contraction. Overall, our follow-up study elucidated and confirmed that the underlying mechanism of action for both extracts was mediated via the cholinergic pathway, with a significant contribution from kaempferol.

Investigation on the vasodilators between these two extracts was carried out in both endothelium-intact and endothelium-denuded rat aorta. Results have shown that upon exposure at the same concentration of 2 mg/mL, MEGP produced a more potent vasodilatory effect than that of AEGP. Although at a lower concentration, both extracts responded in a dose-dependent manner, AEGP seemed to reverse the effect into slight contraction upon reaching the concentration of 2 mg/mL, unlike MEGP, which continued to cause further relaxation. Drug effects are caused by the interaction of the drug molecules and the receptor. At low concentration of the drug, many binding sites are still available; however, further increasing the drug concentration causes more drug-receptor complex formation and a substantial increase in drug effect until all of the receptor binding sites are occupied. Further increasing the drug concentration causes receptors to become saturated, leading to no or little effect of the drug. Thus, the slight reverse effect of contraction by AEGP at 2 mg/mL indicates the possible saturation of all the receptors binding sites for the relaxation effect of the blood vessels, limiting any further vasodilation. Thus, in using AEGP, a possibly saturated concentration exceeding 2 mg/mL would not cause any relaxation effect but a contraction of the blood vessels instead.

Flavonoids rich food, primarily via consumption of botanicals, is often associated with a reduction of cardiovascular diseases. Kaempferol 3-O-rutinoside was detected and quantified as one of the active ingredients in GP. Thus, we decided to test its hypotensive property in an organ bath system using rat aorta. We first carried out the experiments to confirm the relaxant effect of kaempferol 3-O-rutinoside in rat aortic rings, resulting in vasodilatory activity after NA-induced contraction. This result is in agreement with those of previous studies that demonstrated the vasodilatory effects of kaempferol in different vascular beds. Similar findings have also been reported on the vasorelaxant properties of other flavonoids such as quercetin, myricetin, rutin and apigenin. Kaempferol, however, is considered as the most effective among all flavonoids in potentiating vasorelaxation, thus might have more significant potential in preventing and treating CVDs. We assessed the involvement of kaempferol 3-O-rutinoside in potentiating ACh-dependent endothelium-mediated vasodilation, and ACh-independent mediated contraction. Our results showed that kaempferol elicited the relaxation response produced by ACh in both endothelium-intact and endothelium-denuded rat aorta. Our observation suggests that kaempferol enhanced the relaxation in rat aortic rings possibly via endothelium-dependent and endothelium-independent cholinergic pathway, following previous reports, which is possibly mediated by kaempferol 3-O-rutinoside, as illustrated in Figure 8.
Figure 8. Effect of extracts in endothelium and smooth muscle cells of rat aorta possibly via activation of the endothelium-dependent cholinergic pathway or directly via smooth muscle. The figure is a simplified representation showing the main component associated with acetylcholine mediated endothelium vasodilation in the isolated rat aorta (IRA) model and showing the involvement of both extracts in inducing relaxation in vascular endothelium or vascular smooth muscle. In the presence of endothelium, both extracts (AEGP/MEGP) mediate vasodilation possibly by acting on the muscarinic ACh receptor $M_3$ by producing NO. When the endothelium is disrupted, the extracts mediate relaxation on vascular smooth muscle possibly via endothelium-independent cholinergic pathway by acting on the muscarinic ACh receptor $M_2$ and the muscarinic ACh receptor $M_3$ or via adrenergic pathway by acting on α and β adrenoceptors. The vasorelaxant effect produced by the extracts is mediated by kaempferol, particularly, kaempferol 3-O-rutinoside. ACh; Acetylcholine $M_2$: Muscarinic acetylcholine receptor $M_2$; $M_3$: Muscarinic acetylcholine receptor $M_3$; α: alpha adrenoceptors; β: beta-adrenoceptors; NO: Nitric oxide; GC: Guanylyl cyclase; cGMP: cyclic guanosine monophosphate; AEGP: Aqueous extract of G. procumbens; MEGP: Methanol extract of G. procumbens.

Summary and conclusion
The result of this study shows that crude extracts of AEGP and MEGP possessed different characteristics and physical appearance, as well as the percentage yield. MEGP was a dark greenish-brown sticky solid whereas AEGP was a light brown fluffy material. The percentage yield of MEGP was higher than AEGP.
The result of this study also reveals the presence of essential phytoconstituents in both AEGP and MEGP, and MEGP was found to contain more active constituents than AEGP. Besides, MEGP also contained higher total flavonoid and flavone contents than AEGP, although the amount of phenolic content was lower in MEGP. HPLC study has demonstrated the presence of kaempferol 3-O-rutinoside, its vasorelaxation effects in endothelium-intact and endothelium-denuded aorta. This present study also demonstrates the vasodilatory effect of the two extracts, in thoracic aorta tissues isolated from rats, possibly via the contribution of the cholinergic mediated pathway. It is also concluded that the main active vasodilatory component in both extracts causing such hypotensive effect is the flavonoid particularly kaempferol 3-O-rutinoside. However, further study is necessary to elucidate its mechanism of action.

**Data availability**

**Underlying data**

Open Science Framework: Anti-hypertensive Vasodilatory Action of Gynura procumbens. [https://doi.org/10.17605/OSF.IO/W6XJV](https://doi.org/10.17605/OSF.IO/W6XJV)

This project contains “Anti-hypertensive Vasodilatory Action of Gynura procumbens.xlsx”, which includes raw data for all experiments performed in this study.

Data are available under the terms of the Creative Commons Attribution 4.0 International licence (CC-BY 4.0).

**References**

1. Chobanian AV, Bakris GL, Black HR, et al.: Seventh report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure. Hypertension. 2003; 42(6): 1206–1252. PubMed Abstract | Publisher Full Text

2. World Health Organization: Cardiovascular diseases (CVDs). 2019. Reference Source

3. Oparil S, Zaman MA, Calhoun DA: Pathogenesis of Hypertension. Annuals of Internal Medicine. 2003; 139: 761–776. PubMed Abstract | Publisher Full Text

4. Bolivar JJ: Essential Hypertension: An Approach to Its Etiology and Neurogenic Pathophysiology. Jnt J Hypertens. 2013; 2013: 547809. PubMed Abstract | Publisher Full Text | Free Full Text

5. Beggs S, Cosgrove M, Hatfield NT, et al.: Introductory Clinical Pharmacology. 7th ed edn (Lippincott Williams & Wilkins, 2003). Reference Source

6. Tabassum N, Ahmad F: Role of natural herbs in the treatment of hypertension. Pharmacogn Rev. 2011; 5(9): 30–40. PubMed Abstract | Publisher Full Text | Free Full Text

7. Freel EM, Connell JMC: Mechanisms of hypertension: the expanding role of aldosterone. J Am Soc Nephrol. Mechanisms 2005; 16(8): 1993–2001. PubMed Abstract | Publisher Full Text | Free Full Text

8. Kaur N, Kumar R, Yarn MF, et al.: Antihypertensive effect of gynura procumbens water extract in spontaneously hypertensive rats. International Journal of Applied Research in Natural Products. 2013; 6(3): 20–27. Reference Source

9. Ng HK, Poh TF, Lam SK, et al.: Potassium channel openers and prostacyclin play a crucial role in mediating the vasorelaxant activity of Gynura procumbens. BMC Complement Altern Med. 2013; 13: 188–188. PubMed Abstract | Publisher Full Text | Free Full Text

10. Kaur N, Ihsan Akwadhi A, Ali RB, et al.: Cardio-Vascular Activity of Gynura Procumbens Merr. Leaf Extracts. Int J Pharm Sci Res. 2012; 3: 1401–1405. Publisher Full Text

11. Jarikasem S, Siirantikorn S, Charuwichitratana S, et al.: Antithrombic Effects of Gynura procumbens. 2013; 2013: PubMed Abstract | Publisher Full Text | Free Full Text

12. Mou KM, Dash PR: Comprehensive Review on Gynura Procumbens Leaves. International Journal Of Pharmacognosy. 2016; 3(4): 167–174. Publisher Full Text

13. Kim MJ, Lee HJ, Wiroyiwigado S, et al.: Antithrombic Effects of Gynura procumbens Extract in Spontaneously Hypertensive Rats. J Med Food. 2006; 9(4): 587–590. PubMed Abstract | Publisher Full Text

14. Abrikia OSS, Yam MF, Asmawi MZ, et al.: Effects of extracts and fractions of gynura procumbens on rat atrial contraction. J Acupunct Meridian Stud. 2013; 6(4): 199–207. PubMed Abstract | Publisher Full Text

15. Hoo SZ, Lee CN, Mok SL, et al.: Gynura procumbens Merr. decreases blood pressure in rats by vasodilatation via inhibition of calcium channels. Clinics (Sao Paulo). 2011; 66(1): 143–150. PubMed Abstract | Publisher Full Text | Free Full Text

16. Poh TF, Ng HK, Hoo SZ, et al.: Gynura procumbens causes vasodilation by inhibiting angiotensin II and enhancing bradykinin actions. J Cardiovasc Pharmacol. 2013; 61(5): 378–384. PubMed Abstract | Publisher Full Text

17. Azimah Aziz Al, Petekcin MR: Developing a phosphodiesterase-5 inhibitor assay to detect counterfeit drugs containing phosphodiesterase-5 inhibitors using spectrophotometry [version 1; peer review: 1 approved, 1 approved with reservations]. F1000Research. 2019; 8: 1692. Publisher Full Text

18. Wang H, Zhou JW, Fu DH, et al.: Gynura procumbens ethanolic extract suppresses osteosarcoma cell proliferation and metastasis in vitro. Oncol Lett. 2013; 4(1): 113–117. PubMed Abstract | Publisher Full Text | Free Full Text

19. Tan HL, Chan KG, Puspaparaj P, et al.: Gynura procumbens: An Overview of the Biological Activities. Front Pharmacol. 2016; 7: 52. PubMed Abstract | Publisher Full Text | Free Full Text

20. Algarini K, Menger KY, Atangwoh II, et al.: Hypoglycemic and anti-hyperglycemic study of Gynura procumbens leaf extracts. Asian Pac J Trop Biomed. 2013; 3(5): 358–366. PubMed Abstract | Publisher Full Text | Free Full Text

21. Kaewseenan N, Sutthikum V, Siriamornpun S: Potential of Gynura procumbens leaves as source of flavonoid-enriched fractions with enhanced antioxidant capacity. J Funct Foods. 2015; 12: 120–128. Publisher Full Text

22. Afandi A, Zulkifili MH, Sadikun A, et al.: Antioxidant Properties of Gynura Procumbens Extracts and Their Inhibitory Effects on Two Major Human Recombinant Cytochrome P450s using a High Throughput Luminescence Assay. Asian J Pharm Clin Res. 2014; 7: 5–10. Reference Source

23. Lee HW, Hakim P, Rabu A, et al.: Antidiabetic effect of Gynura procumbens leaves extracts involve modulation of hepatic carbohydrate metabolism in streptozotocin-induced diabetic rats. J Med Plant Res. 2012; 6(5): 796–812. Publisher Full Text

24. Trushvea S, Trunkova D, Bankova V: Different extraction methods of biologically active components from propolis: a preliminary study. Chem Cent J. 2007; 1: 13–13. PubMed Abstract | Publisher Full Text | Free Full Text

25. Ahmad A, Alkarkhi AFM, Hena S, et al.: Extraction, Separation and Identification of Chemical Ingredients of Elephantopus Scaber L. Using Factorial Design of Experiment. International Journal of Chemistry. 2009; 1: 36–49. Publisher Full Text
26. Iqbal E, Salim KA, Lim LBL: Phytochemical screening, total phenolics and antioxidant activities of bark and leaf extracts of Goniothalamus velutinus (Airy Shaw) from Bruneli Darussalam. J King Saud Univ Sci. 2015; 27: 224-232. Publisher Full Text

27. Hossain MA, Al-Raqmi KAS, Al-Mijizy ZH, et al.: Study of total phenol, flavonoids contents and phytochemical screening of various leaves crude extracts of locally grown T. hymus vulgaris. Asian Pac J Trop Biomed. 2013; 3(9): 705-710. PubMed Abstract | Publisher Full Text | Free Full Text

28. Srinivas P, Samatha T, Valya G, et al.: Phytochemical Screening and Antimicrobial Activity of Leaf Extract of Wrightia tomentosa. International journal of biological sciences. 2013; 2: 23–27. Reference Source

29. Kulevanova S, Stefova M, Panovska TK, et al.: HPLC Identification and determination of myricetin, quercetin, kaempferol and total flavonoids in herbal drugs. Macedonian pharmaceutical bulletin. 2002; 48: 25–30. Publisher Full Text

30. Shahlehi S, Petalcorin MR: Anti-hypertensive Vasodilatory Action of Gynura procumbens. Open Science Framework. 2020. http://www.doi.org/10.17605/OSF.IO/W6XJF

31. Kamara-Kanu KA, Mat Noor M: Gynura procumbens Leaf Improves Blood Glucose Level, Restores Fertility and Libido of Diabetic-Induced Male Rats. Sains Malays. 2017; 46: 1471-1477. Publisher Full Text

32. Konwar PG, Beknai AK, Patil BS, et al.: A study on phytochemical investigation of Drynaria Quercifolia Linn Rhizome. Int J Pharm Sci Res. 2010; 1: 148-158. Publisher Full Text

33. Enne AC, et al.: Antiplasmodial and Antioxidant Evaluation of the Methanol and Aqueous Extracts of Sacrocephalus latifolius. Scied Journal of Analytical Biochemistry. 2018; 1: 1-9.

34. Njume C, Jide AA, Ndip RN: Aqueous and Organic Solvent-Extracts of Selected South African Medicinal Plants Possess Antimicrobial Activity against Drug-Resistant Strains of Helicobacter pylori: Inhibitory and Bactericidal Potential. Int J Mol Sci. 2011; 12(9): 5652-5655. PubMed Abstract | Publisher Full Text | Free Full Text

35. Ngo TV, Scarlett C, Bowyer MC, et al.: Impact of Different Extraction Solvents on Bioactive Compounds and Antioxidant Capacity from the Root of Selicia chinensis L. Journal of Food Quality. 2017; 1: 1-8. Publisher Full Text

36. Ndip RN, Tarkang AEM, Mbullah SM, et al.: In vitro anti-Helicobacter pylori activity of extracts of selected medicinal plants from North West Cameroon. J Ethnopharmacol. 2007; 114(3): 452-457. PubMed Abstract | Publisher Full Text

37. Fraga CG, Galilea M, Verstraeten SV, et al.: Basic biochemical mechanisms behind the health benefits of polyphenols. Mol Aspects Med. 2010; 31(6): 435-445. PubMed Abstract | Publisher Full Text

38. Aroyl S, Baniya MK, Danekhu K, et al.: Total Phenolic Content, Flavonoid Content and Antioxidant Potential of Wild Vegetables from Western Nepal. Plants (Basel). 2019; 8(4): 96. PubMed Abstract | Publisher Full Text | Free Full Text

39. Vazquez D, Rodriguez-Mateos A, Corona G, et al.: Polypolins and Human Health: Prevention of Disease and Mechanisms of Action. Nutrients. 2010; 2(11): 1106-1131. PubMed Abstract | Publisher Full Text | Free Full Text

40. Cheng YC, Sheen JM, Hu WL, et al.: Polypolins and Oxidative Stress in Atherosclerosis-Related Ischemic Heart Disease and Stroke. Oxid Med Cell Longev. 2017; 2017: 8526438. PubMed Abstract | Publisher Full Text | Free Full Text

41. Goscik K, Duthie GG, Stewart D, et al.: Bioactive polyphenols and cardiovascular disease: chemical antagonists, pharmacological agents or xenobiotics that drive an adaptive response? Br J Pharmacol. 2017; 174(1): 1209-1225. PubMed Abstract | Publisher Full Text | Free Full Text

42. Aybaster O, Isik E, Sahin S, et al.: Optimization of ultrasonic-assisted extraction of antioxidant compounds from blackberry leaves using response surface methodology. Industrial Crops and Products. 2013; 44: 558-565. Publisher Full Text

43. Tan HL, Chan KG, Puspaprajah P, et al.: Gynura procumbens: An Overview of the Biological Activities. Front Pharmacol. 2016; 7: 52. PubMed Abstract | Publisher Full Text | Free Full Text

44. Walsh MP: Regulation of vascular smooth muscle tone. Can J Physiol Pharmacol. 1994; 72(8): 919-936. PubMed Abstract | Publisher Full Text | Free Full Text

45. Sumi Y, Woehler T, Chen Y, et al.: Adrenergic receptor activation involves ATP release and feedback through purinergic receptors. Am J Physiol Cell Physiol. 2010; 299(5): C1118–1126. PubMed Abstract | Publisher Full Text | Free Full Text

46. Srinivas P, Samatha T, Valya G, et al.: Cholinergic submandibular effects and muscarinic receptor expression in blood vessels of the rat. Arch Oral Biol. 2008; 53(7): 605-616. PubMed Abstract | Publisher Full Text | Free Full Text

47. Srivastava D, Tiwari P, Tiwari B, et al.: Expression of muscarinic receptor subtypes in salivary glands of rats, sheep and man. Arch Oral Biol. 2008; 53(1): 66-74. PubMed Abstract | Publisher Full Text

48. Salavanki S, Gunasekaran B, Shaharuddin NA, et al.: Antiatherosclerotic Effects of Plant Flavonoids. Biomed Res Int. 2014; 2014: 486528. PubMed Abstract | Publisher Full Text | Free Full Text

49. Hacker M, Messer W, Bachmann KA: Pharmacology: Principles and Practice. (Academic Press) 2009. Reference Source

50. Wang C, Mehdendale S, Calway T, et al.: Botanical Flavonoids on Coronary Heart Disease. Am J Chin Med. 2012; 39(4): 661-671. PubMed Abstract | Publisher Full Text | Free Full Text

51. Al-maasarani S, Gunasekaran B, Shaharuddin NA, et al.: Antiinflammatory effects of Gynura procumbens. J Ethnopharmacol. 2008; 114(1): 148–158. PubMed Abstract | Publisher Full Text | Free Full Text

52. Chandradevan M, Simoh S, Mediani A, et al.: UHPLC-ELISA-Orbitrap-MS Analysis of Biochemically Active Extracts from Gynura procumbens (Lour.) Merr. and Cleome gynandra L. Leaves. Evid Based Complement Alternat Med. 2020; 2020: 3238561. PubMed Abstract | Publisher Full Text | Free Full Text

53. Perez-Vicinalo F, Ibarra M, Cogolludo AL, et al.: Endothelium-Independent Vasodilator Effects of the Flavonoid Quercetin and Its Methylated Metabolites in Rat Conductance and Resistance Arteries. J Pharmacol Exp Ther. 2002; 302(1): 66–72. PubMed Abstract | Publisher Full Text

54. Xu YC, Leung SW, Leung GPH, et al.: Kaempferol enhances endothelium-dependent relaxation in the porcine coronary artery through activation of large-conductance Ca(2+)-activated K(+) channels. Br J Pharmacol. 2015; 172(2): 3003–3014. PubMed Abstract | Publisher Full Text | Free Full Text

55. Mahobiya A, Singh TU, Rungsung S, et al.: Kaempferol-induced vasorelaxation via endothelium-independent pathways in rat isolated pulmonary artery. Pharmacol Rep. 2018; 70(5): 863–874. PubMed Abstract | Publisher Full Text | Free Full Text

56. Osz OY, Ertug PU, Karabulut E, et al.: Dose-dependent differential mechanism of quercetin-induced vasodilatations in isolated perfused rat mesenteric vascular bed. International Journal of Pharmacology. 2016; 12(4): 379–386. Publisher Full Text
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Yun Shin Sew

Malaysian Agricultural Research & Development Institute (MARDI), Seri Kembangan, Malaysia

This article by Shahlehi et al. reported interesting findings about the vasodilatory effects of two different plant extracts of Gynura procumbens. The authors have shown the one extracted using methanol solvent, MEGP contains higher amount of flavonoids particularly kaempferol 3-O-rutinoside than the water solvent, AEGP. Following that, the authors showed MEGP works better for in vitro vasodilatory performance than AEGP. The authors also claim a probable role of kaempferol 3-O-rutinoside in both extracts as the main causal factor via experimental observations. There are several comments on this article as the followings:

1. The authors should state the volume of individual AEGP and MEGP used for HPLC analysis in the material and method section.

2. Table 1 should be removed as those are simple information which could be just included in the text.

3. There are discrepancies in the usage of asterisk mark for statistical significance level (referring to Figure 3, 4 and 7) which might confuse the readers. Single asterisk often indicate P<0.05 and double asterisk for P<0.01, need to standardize the usage of asterisk(s) in all figures.

4. It was a good experimental design in such a way that the relaxant effect of kaempferol 3-O-rutinoside in rat aortic rings was investigated. However the amount of kaempferol 3-O-rutinoside tested was pretty low (10ug/mL). Could this be tested with higher amount of kaempferol 3-O-rutinoside since as high as 57.86 ug/mL and 78.35 ug/mL of this compound was found in AEGP and MEGP extract, respectively? Given that higher concentration of extract showed adverse effect of vasodilatory action in Figure 5, will any adverse effect happen when higher kaempferol 3-O-rutinoside is being used (mimicking the concentration of the compound found in each extract)?

5. Just an additional suggestion for the future works of this study. Though the methanol extract of Gynura procumbens performs better than the one extracted using water (as shown
in the present study), methanol extracts of herbs are often considered to be more toxic than water extracts. Therefore, it is necessary to investigate the safety level of this methanol extract. A follow-up in vivo study using rat model should be conducted in future.

Is the work clearly and accurately presented and does it cite the current literature?
Yes

Is the study design appropriate and is the work technically sound?
Yes

Are sufficient details of methods and analysis provided to allow replication by others?
Yes

If applicable, is the statistical analysis and its interpretation appropriate?
Partly

Are all the source data underlying the results available to ensure full reproducibility?
Yes

Are the conclusions drawn adequately supported by the results?
Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Molecular and safety analysis of food and herb products

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

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address are listed below.

Background:
- Authors are asked to comment on reference made to Abrika et al 2013, where they looked at the effect of GP flavonoid fraction on rat arterial contraction as I am contending lack of novelty based on this.

Results:
- Tables 1 and 3 are unnecessary. Information could easily be stated in text without the need of a table.
- Figure 5 is the critical figure in this manuscript. However, error bars are not visible. Is there a reason why there is only n=3 for each treatment arm in figure A and n>5 in figure B? Authors are also asked to keep Y-axis scale consistent between both figures.

Is the work clearly and accurately presented and does it cite the current literature?
Yes

Is the study design appropriate and is the work technically sound?
Yes

Are sufficient details of methods and analysis provided to allow replication by others?
Yes

If applicable, is the statistical analysis and its interpretation appropriate?
Yes

Are all the source data underlying the results available to ensure full reproducibility?
Yes

Are the conclusions drawn adequately supported by the results?
Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Cell biology and drug delivery

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.
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