Namuncurá Marine Protected Area: an oceanic hot spot of benthic biodiversity at Burdwood Bank, Argentina

Laura Schejter1,2 · Clara Rimondino3 · Ignacio Chiesa4,8 · Juan M. Díaz de Astarloa2,5 · Brenda Doti1 · Rodolfo Elías7 · Mariana Escolar1 · Gabriel Genzano2,7 · Juan López-Gappa4 · Marcos Tatián3 · Diego G. Zelaya8 · Javier Cristobo9 · Carlos D. Perez10 · Ralf T. Cordeiro10 · Claudia S. Bremec2,7

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Abstract The first open-sea (non-coastal) Marine Protected Area (MPA) in Argentina was created in 2013 (and named “Namuncurá”), at Burdwood Bank, an undersea plateau located about 200 km south from Malvinas/Falkland Islands, SW Atlantic Ocean. This contribution represents the most recent and complete checklist of benthic organisms (mainly mega and macrofauna) that summarized 240 taxa collected at three sites in the mentioned area. The benthic richness here documented largely exceeds the values coming from other productive areas in Argentinean waters. Faunistic composition was different among the sites, although several species were shared. Bryozoans presented the highest richness compared to the other groups, followed by mollusks and sponges. These data represent valuable information and a very helpful starting point for incoming research, conservation efforts and management. It is essential to establish the spatial distribution of indicator taxa, like corals, sponges and bryozoans, to better attempt at the protection of benthic biodiversity in this rich Argentinean marine system. This is a key issue considering the notable extension of the MPA compared to other areas in Argentina, the bathymetric variations, particular oceanographic conditions and different fishing circumstances within the area.

Keywords Benthic communities · New Marine Protected Area · Conservation · SW Atlantic Ocean

Introduction

The Burdwood Bank (BB) is an undersea plateau located about 200 km south from Malvinas/Falkland Islands and 150 km east from Isla de los Estados (Tierra del Fuego Province). It comprises nearly 34.000 km² circumscribed by the 200 m isobath, between 54°–55°S and 56°–62°W, with a slight slope extended nearly 370 km east–west. The north–south width of the area varies between 50 and 123 km.
100 km (Fig. 1). The depth of the plateau ranges between 50 and 200 m, where the bottom abruptly breaks into a wall reaching 1100 m to more than 3000 m depth in some regions (Zunino and Ichazo 1979). This area is characterized by the influence of subantarctic waters of 4–5 °C on the bottom and part of the shelf break during the whole year. No thermoclines have been detected in the plateau area, and mean salinity values reached 34 (Zunino and Ichazo 1979; Piola and Gordon 1989; Guerrero et al. 1999). This plateau was first described at the beginning of an Antarctic Exploration Cruise in 1842, commanded by Captain Sir James Ross (Findlay 1867).

Burdwood Bank is an intermediate area between South America and Antarctica and part of the islands and shelf that constitute the Scotia Arc. All these were formerly connected in the supercontinent of Gondwana, but separated since Antarctica’s geographic and thermal isolation through the Cenozoic. Despite geographic distance, depths and oceanographic conditions, the Scotia Arc linked Patagonia and the Antarctic Peninsula, playing a connection role for many marine species (Arntz 2005). Thus, the Patagonia-Scotia Arc-Antarctic Peninsula has become a crucial region of the globe for biological and climatological research (Barnes 2005).

Probably, the first mention (or one of the oldest) regarding the high species richness found at this benthic community at Burdwood Bank was reported by the “Scotia” Expedition, in 1903. One trawl at station no 346 (54°55’S and 57°28’W) reached half a ton (“the largest and richest of the whole Antarctic expedition”), with a preliminary list of 70 species, dominated by sponges and bryozoans (Scottish National Antarctic Expedition 1908). More recently, during the LAMPOS expedition performed by the RV Polarstern (April 2002), a rich benthic community was also found in the east and west slopes of the Burdwood Bank. It was mainly constituted by Porifera and Cnidaria (Arntz and Brey 2003). Currently, BB is formally recognized as a subantarctic area of ecological importance, where biological processes that deserve conservation occurred, when areas of fisheries management and protection were implemented years ago. In 2004, a prohibited area of bottom trawling and fishery of the Patagonian toothfish Dissostichus eleginoides was established at the western portion of the Bank when catches of juveniles exceeded 15% of the total catch (Resolution of the Consejo Federal Pesquero No 3/2004, Argentina). Thereafter, in 2008, another area of permanent prohibition of fisheries was implemented within the central portion of the Bank (currently the “core” of the Marine Protected Area), in order to conserve the marine bottom (Resolution of the Consejo Federal Pesquero No 18/2008, Argentina) (Fig. 1). These regulations were the first steps toward the creation (in 2013) of the first open-sea (non-coastal) Argentine Marine Protected Area (Law 26875, Argentina).

In order to provide the necessary baseline knowledge for the establishment of the Namuncura Marine Protected Area (N MPA), general and historical information on oceanography, biodiversity and fisheries exploitation in the BB was compiled by Falabella et al. (2013). Some reviews (i.e., Zamponi 2008a, b) pointed out the importance of the region based on the presence of vulnerable sessile benthic species like the reported rich coral fauna. Other literature also mentioned the occurrence of several species regarding vulnerable indicators of marine ecosystems (Jones and Lockhart 2011), such as bryozoans, sponges and tunicates. All these organisms have already been recorded at BB (López Gappa 2000; Tatián et al. 2005; Schejter et al. 2012). Criteria of vulnerability of these ecosystems are mainly based on the existence of a necessary high biomass of sessile taxa. The latter are composed by long-living species very sensitive to bottom fishing, high percentages of endemism and very slow recovery times of the community after a perturbation event (FAO 2009; Auster et al. 2011; Portela et al. 2012). Based on these criteria and as a precautionary measure to preserve this ecosystem, a MPA was established.

The N MPA comprises three different sub-areas according to the required protection level. The central one, called “core,” has been defined as the area that contains a representative fraction of the benthic marine biodiversity that requires a strict protection due to the vulnerable characteristics presented by the ecosystem components. No activities are allowed in the core, except for control and monitoring. The core area is surrounded by a “buffer” area, in which activities are allowed after previous authorization (e.g., scientific research, explorations dealing with natural resources and biodiversity, sustainable resources management, restoration and monitoring of global change). Finally, an external “transition” area was defined, in which productive and extractive activities considered in the management plan are allowed. Beyond the transition area, the shelf-break and nearby areas have no protection regulations.

Embedded into the Argentinean “Pampa Azul” Marine National Science Project, the N MPA is a conservation aim that represents a challenge regarding management and planning. It joins the 43 existing Coastal-Marine Protected Areas in Argentina (see Caille et al. 2013). It largely contributes to nearly 4% of marine areas under some kind of protection in the country, supporting the global objective of the Convention on Biological Diversity (CBD) to protect and preserve the biota of the world’s marine and coastal regions. As a consequence, the acquisition of further knowledge of the region in the near future (species composition, abundance, biomass and distribution of benthic species in the bank and the shelf-break area) is expected.
It is well known that bottom fishing may damage or destroy the seafloor habitat and the associated benthic communities (National Research Council 2002). After drastic fishing activities, the ecosystem would experience severe changes in structure and composition (Hinz et al. 2009). Epibenthic and sessile fauna are vulnerable to fishing disturbance, though the level of disturbance is related to the habitat type and may or may not recover after fishing closures or interrupted activities (Queirós et al. 2006; Asch and Collie 2008). Several countries that created MPAs developed action plans and management strategies in order to preserve the biodiversity of vulnerable and sensitive ecosystems, such as coral habitats (Wattage et al. 2011). Protection and conservation issues also apply for the international waters and vulnerable marine ecosystems (Durañón et al. 2012, 2015). Cold-water coral areas remained largely unappreciated and overlooked until 1990s, although their existence was known since the time of Linnaeus (Cairns 2007; Roberts and Cairns 2014). It is also known that more than 65% of all the extant corals (Anthipatharia + Scleractinia + Octocorallia + Stylasteridae) are cold-water species, occurring at depths >200 meters (Cairns 2007). These habitats face considerable threat from ocean acidification (Guinotte et al. 2006), and the future distribution pattern of the benthic communities might be influenced by climate change (Fautin et al. 2009). It is expected that the vulnerable benthic communities recognized near N MPA (BB), as those recently described along the Argentine continental margin (i.e., coldwater coral reefs, coral gardens, sponge beds and rocky environments) (Portela et al. 2012, 2015) and South Georgia Islands (Gaitán et al. 2013a, b) should be protected, studied and monitored from now on.

Fig. 1 Location of the three sub-areas comprised in the Namuncurá MPA, Burdwood Bank (Argentina). Solid circles show the sampling sites during the research cruise onboard the OV "Puerto Deseado," April 2013. Image courtesy of Valeria Falabella.
In this contribution, we present current data on benthic richness of the N MPA based on results from a sampling carried out in April 2013. We provide an updated inventory of benthic taxa collected at the core, buffer and shelf-break areas in order to describe the benthic community in the MPA and to recommend future studies that should be developed for its proper management.

Materials and methods

Four sampling sites were supplementary added to the original campaign planning onboard the Oceanographic Vessel “Puerto Deseado” (CONICET, Argentina) at the return of the Argentinean Antarctic Expedition in April 2013. The objective of the sampling was to perform a preliminary assessment of the Burdwood Bank.

Epibenthic organisms were collected using a bottom otter trawl (14 min, mean trawling time), while infaunal small mollusks were sampled using a small box dredge (45 × 50 × 12 cm). Due to constraints of the sampling methods, only qualitative data have been taken, though visual inspections of the total catch permitted a general assumption of the main taxonomic groups collected in each site.

Due to weather conditions, only three sites (out of the four originally planned) were sampled. These were located at the core, buffer and shelf-break areas of the MPA (Table 1). Epibenthic taxa were sorted, photographed and preserved onboard (frozen, 70 % ethanol and/or formalin–seawater 4 %, depending on the taxa). Taxa were identified at laboratory by different specialists included as co-authors in this paper. Species richness of trawl samples was compared using a similarity index (SI = S/N, where S = taxa in common between samples A and B, and N = cumulative number of taxa in samples A and B), following a similar analysis performed by Lovell and Trego (2003) at Deception Island, Antarctica.

Results

A total of 240 benthic taxa were collected at three different sites at the Namuncurá MPA, Burdwood Bank (Table 2).

One hundred and forty-eight taxa were identified at the core of the MPA (sample volume <90 l). Although not quantitatively assessed, the visual inspection of the general catch revealed that this area was characterized by a relatively high biomass of sponges that corresponded to 16 morphospecies (14 Demospongiae and 2 Calcarea). Few fragments of corals belonging to the stylasterid E. antarctica were also found, with some individuals of the lepadomorph crustacean Ornatoscalpellum gibberum as epibionts. Ten common species of echinoderms were recorded and 12 crustaceans, including 6 decapods comprising the commercial king crabs Lithodes confundens and Paralomis granulosa, 5 peracarids and the above-mentioned lepadomorph. Bryozoans and mollusks presented the highest richness (regarding major groups) in the area: 47 and 25 taxa, respectively. Hydrozoans were represented by 11 taxa, brachiopods by 2 species, ascidians by 8 taxa and pycnogonids by a single taxon, while several polychaetes (13 taxa) were also recorded, although belonging to the infaunal fraction. Ascidians were represented in this station by eight taxa. Most of them were colonial organisms belonging to the family Polyclinidae. Members of this particular Family were rather difficult to identify to the lowest taxonomic level. The reproductive state of the samples collected (asexual phase) was characterized by a regression of the anterior region of the zooids, and the sole presence of phagocytic cells heavily laden with reserves in the postabdomen did not allow us to reach the species or genus level. Four fish species were also caught by the otter trawl.

The sample collected at the station in the buffer area (sample volume <40 l) was mainly composed of brachiopods (living organisms and dead valves) and calcareous polychaete tubes (Serpulidae). A total of 109 taxa were identified. Fifteen echinoderm species (asteroids, sea urchins) were recorded, 48 bryozoans including some big colonies (comprising the lace coral Reteporella magellensis), 8 sponges, 1 hydrocoral (E. antarctica), 5 decapods and 1 amphipod among crustaceans, 8 mollusks, 7 polychaetes, 10 hydrozoans, 1 ascidian and 2 fishes.

A remarkable presence of corals and ophiuroids was detected visually in the shelf break of the bank, at the end of the transition area of the MPA. These coral aggregations at the slope of the shelf break probably caused the breakage of the otter trawl net, hence producing the poorest sample in terms of volume and consequently in taxa richness (sample volume <30 l). A total of 92 taxa were identified,

Table 1 Sampling sites at Burdwood Bank, during the research cruise onboard the OV ARA “Puerto Deseado,” April 2013

| Site         | Latitude (S)    | Longitude (W) | Depth (m) | Surf. temp. (°C) | Salinity |
|--------------|-----------------|---------------|-----------|-----------------|----------|
| Core         | 54°15.78’       | 60°00.43’     | 101       | 7, 52           | 33, 91   |
| Buffer       | 54°15.57’       | 60°34.41’     | 113       | 7, 68           | 33, 9    |
| Shelf break  | 54°15.19’       | 61°40.43’     | 236       | 7, 47           | 33, 9    |
Table 2  Taxa list of the organisms recorded at the three sites (core, buffer and shelf break) of the Namuncurá MPA, Burdwood Bank. Taxa richness per major group and area is indicated

| Taxa                          | Core | Buffer | Shelf break |
|-------------------------------|------|--------|-------------|
| Porifera                      |      |        |             |
| Craniella leptoderma          | 16   | 8      | 13          |
| Calcarea 1                    |      |        |             |
| Calcarea 2                    |      |        |             |
| Callyspongia flabellata       |      |        |             |
| Callyspongia fortis           |      |        |             |
| Coelosphaeridae               |      |        | x           |
| Guitarra sp.                  |      |        |             |
| Halichondrida 1               |      |        | x           |
| Haliclona (Gellius) sp.       |      |        |             |
| Haliclona sp. 3               |      |        | x           |
| Haliclona sp. 4               |      |        | x           |
| Haliclona sp. 1               |      |        |             |
| Haliclona sp. 2               |      |        | x           |
| Haliclona sp. 5               |      |        |             |
| Haplosclerida 1               |      |        |             |
| Haplosclerida 2               |      |        | x           |
| Hexactinellida 1              |      |        | x           |
| Iophon sp.                    |      |        |             |
| Isodictya sp.                 |      |        |             |
| Latrunculia sp.               |      |        |             |
| Lissodendoryx sp.             |      |        |             |
| Microcionidae 1               |      |        | x           |
| Microcionidae 2               |      |        |             |
| Microcionidae 3               |      |        |             |
| Mycale sp. 2                  |      |        |             |
| Mycale sp. 1                  |      |        | x           |
| Myxilla sp. 3                 |      |        | x           |
| Myxilla sp. 1                 |      |        |             |
| Myxilla sp. 2                 |      |        |             |
| Myxillina                     |      |        |             |
| Poecilosclerida               |      |        | x           |
| Sycon sp.                     |      |        |             |
| Tedania cf. charcoti          |      |        |             |
| Tedania massa                 |      |        |             |
| Tedania mucosa                |      |        |             |
| Tedania sp.                   |      |        |             |
| Bryozoa                       | 47   | 48     | 19          |
| Amastigia benemunita          |      |        |             |
| Andreella uncifera            |      |        | x           |
| Arachnopusia monoceros        |      |        |             |
| Beania costata                |      |        | x           |
| Beania inermis                |      |        |             |
| Beania magellanica            |      |        |             |
| Bicirrhis biciliata           |      |        |             |
| Buffonellodes simplex         |      |        | x           |
| Buffonellodes glabra          |      |        | x           |
| Caberea darwinii              |      |        | x           |
| Species                                   | Core | Buffer | Shelf break |
|------------------------------------------|------|--------|-------------|
| Carbasea ovoidea                         | x    | x      |             |
| Catadysis immersum                       | x    | x      |             |
| Cellaria malvinensis                     | x    |        |             |
| Cellarinella dubia                       | x    | x      |             |
| Celleporina bicostata                    | x    |        |             |
| Chaperiopsis galeata                     | x    |        |             |
| Chondriovelum angustilobatum             | x    |        |             |
| Crisia sp.                               | x    |        | x           |
| Disporella sp.                           | x    |        |             |
| Ellisina incrustans                      | x    |        |             |
| Exochella longirostris                   | x    |        |             |
| Fasciculipora ramosa                     | x    |        |             |
| Fenestrulina dupla                       | x    |        |             |
| Fenestrulina horrida                     | x    |        |             |
| Fenestrulina incusa                      | x    |        |             |
| Flustrapora magellanica                  | x    |        |             |
| Foveolaria terrifica                     | x    |        |             |
| Gregarinidra variabilis                  | x    | x      | x           |
| Himantozoum obtusum                      | x    |        |             |
| Hornera sp.                              | x    | x      |             |
| Kenoaplousina fassurata                  | x    |        |             |
| Lacerna hosteensis                       | x    |        |             |
| Menipea flagellifera                     | x    |        |             |
| Menipea patagonica                       | x    |        |             |
| Micropora brevissima                     | x    | x      |             |
| Microporella hyadesi                     | x    | x      |             |
| Neothoa cf. chiloensis                   | x    |        |             |
| Nevianipora milneana                     | x    | x      |             |
| Odontoporella adpressa                   | x    | x      |             |
| Ogivalia elegans                         | x    |        |             |
| Orthoporidoides erectus                  | x    |        |             |
| Osthimosia bicornis                      | x    | x      |             |
| Osthimosia eatonensis                    | x    |        |             |
| Osthimosia magna                         | x    | x      |             |
| Parasmittina dubitata                    | x    | x      |             |
| Plagioecia dichotoma                     | x    | x      |             |
| Plagioecia sp.                           | x    |        |             |
| Platychelely planulata                   | x    |        |             |
| Pseudimonea fissurata                    | x    |        |             |
| Reteporella magellensis                  | x    | x      |             |
| Romanchenia labiosa                      | x    |        |             |
| Smittina julieni                         | x    |        |             |
| Smittina lebruni                         | x    | x      |             |
| Smittina obliata                         | x    | x      |             |
| Smittina smittiana                       | x    | x      |             |
| Smittiodea rhynchota                     | x    |        |             |
| Smittiodea sigillata                     | x    | x      |             |
| Stephanollona longispinata               | x    | x      |             |
|                             | Core | Buffer | Shelf break |
|-----------------------------|------|--------|-------------|
| **Stomatopora eburnea**     | x    | x      | x           |
| **Tricellaria aculeata**    | x    | x      |             |
| **Tubulipora sp.**          | x    | x      | x           |
| **Brachiopoda**             |      |        |             |
| *Lyothyrella uva*           | x    |        |             |
| *Magellania venosa*         | x    | x      | x           |
| *Terebratella dorsata*      | x    |        |             |
| **Cnidaria (Anthozoa)**     |      |        |             |
| *Actiniania 1*              | x    |        |             |
| *Actiniania 2*              | x    |        |             |
| *Actiniania 3*              | x    |        |             |
| *Alcyonium sp.*             | x    |        |             |
| *Flabellium cf. thouarsi*   |      |        |             |
| *Primnoella compressa*      | x    |        |             |
| *Thouarella brucei*         | x    |        |             |
| *Thouarella chilensis*      | x    |        |             |
| *Clavularia sp.*            | x    |        |             |
| **Cnidaria (Hydrozoa)**     |      |        |             |
| *Abietinella operculata*    | x    |        |             |
| *Amphibiseta operculata*    | x    | x      |             |
| *Campanularia sp.*          | x    |        |             |
| *Filellum sp.*              | x    | x      |             |
| *Grammaria magellanica*     | x    | x      |             |
| *Hebella striata*           | x    |        |             |
| *Phialella chilensis*       | x    | x      |             |
| *Sertularella sp. 1*        | x    | x      |             |
| *Sertularella sp. 2*        | x    | x      |             |
| *Sertularella striata*      | x    | x      |             |
| *Errina antarctica*         | x    | x      | x           |
| *Symplectoscyphus sp. 1*    |      |        |             |
| *Symplectoscyphus subdichotomus* | x | | |
| **Mollusca**                |      |        |             |
| *Admete sp.*                | x    |        | x           |
| *Anomacme sp.*              | x    |        |             |
| *Antistreptus sp.*          | x    |        |             |
| *Astarte sp.*               |      |        |             |
| *Ataxocerithium sp.*        | x    |        |             |
| *Australodiris sp.*         | x    |        | x           |
| **Buccinidae**              |      |        |             |
| *Calliostoma sp. 1*         | x    | x      |             |
| *Calliostoma sp. 2*         | x    |        |             |
| *Calliostoma sp.*           | x    |        |             |
| *Carditopsis sp.*           | x    |        |             |
| *Cerithiella sp.1*          | x    |        | x           |
| *Cerithiella sp.2*          | x    |        |             |
| *Cerithiella sp.3*          | x    |        |             |
| *Cylindricalia sp.*         | x    |        |             |
| *Errorina antarctica*       | x    |        |             |
| *Symplectoscyphus subdichotomus* | x | | |
| **Polar Biol**              |      |        |             |
| **123**                     |      |        |             |
| Family                          | Core | Buffer | Shelf break |
|--------------------------------|------|--------|-------------|
| Eulimidae                      |      |        | x           |
| *Eumeta* sp.                   |      |        |             |
| *Eurhomalea* sp.               |      |        |             |
| *Falsimargarita* sp.           |      |        |             |
| *Fissurellidea* sp.            |      |        |             |
| *Fuegotrophon* sp.             |      |        |             |
| *Hiatella* sp.                 |      |        |             |
| *Iothia* sp.                   |      |        |             |
| *Kellia* sp.                   |      |        |             |
| *Lamellaria* sp. 1             |      |        | x           |
| *Limatula* sp.                 |      |        |             |
| *Limopsis* sp.                 |      |        |             |
| *Margarella* sp.               |      |        |             |
| *Neactaeonina* sp.             |      |        |             |
| *Neolepton* sp.                |      |        |             |
| *Onoba* sp.                    |      |        | x           |
| *Pareuthria* sp.               |      |        |             |
| *Solariella* sp.               |      |        |             |
| *Trochita* sp.                 |      |        |             |
| *Trophon* sp. 1                |      |        |             |
| *Trophon* sp. 2                |      |        |             |
| *Turridae*                     |      |        |             |
| *Zygochlamys patagonica*       |      |        |             |
| **Ascidiacea**                 |      |        |             |
| *Aplidium meridianum*          |      |        |             |
| *Aplidium fuegiense*           |      |        |             |
| *Aplidium* sp.                 |      |        |             |
| *Polyclinidae* 1               |      |        |             |
| *Polyclinidae* 2               |      |        |             |
| *Polyclinidae* 3               |      |        |             |
| *Polyclinidae* 4               |      |        |             |
| *Polysyncraton trivolatum*     |      |        |             |
| *Molgula* sp.                  |      |        |             |
| *Styela* sp.                   |      |        |             |
| **Crustacea (Decapoda)**       | 6    | 5      | 3           |
| *Campylonotus vagans*          |      |        |             |
| *Eurypodi*us lateirelle*       |      |        |             |
| *Halicarcinus planatus*        |      |        |             |
| *Lithodes confundens*          |      |        |             |
| *Munida gregaria*              |      |        |             |
| *Pagurus comptus*              |      |        |             |
| *Paralomis granulosa*          |      |        |             |
| **Crustacea (Peracarida)**     | 5    | 1      | 3           |
| *Aega semicarinata*            |      |        |             |
| *Chaetarcturus* sp.            |      |        |             |
| *Cymodopsis* cf. baegeli       |      |        |             |
| *Natatoriana pastorei*         |      |        |             |
| *Acanthoserolis schythei*      |      |        | x           |
Table 2 continued

|                        | Core | Buffer | Shelf break |
|------------------------|------|--------|-------------|
| **Core Buffer Shelf break** |      |        |             |
| *Metepimeria acanthura*  | x    |        |             |
| *Metandania tordi*       | x    |        |             |
| *Leucothoe cf. spinicarpa* | x  |        |             |
| *Jassa alonsoae*         | x    |        |             |
| **Crustacea (Maxillopoda)** | 1   | 0      | 0           |
| *Ornatoscalpellum gibberum* | x  |        |             |
| **Echinodermata**        | 10   | 15     | 14          |
| *Acodontaster sp.*       | x    |        |             |
| *Anasterias sp.*         | x    |        |             |
| *Asteriidae*             | x    |        |             |
| *Astrotoma agassizii*    | x    |        |             |
| *Austrocidaris canaliculata* | x  | x      | x           |
| *Austrocidaris spinulosida* | x  |        |             |
| *Crytasterias turqueti*  | x    |        |             |
| *Diasterias sp.*         | x    |        |             |
| *Diploptaster clarki*    | x    |        |             |
| *Gorgonocephalus chilensis* | x  |        | x           |
| *Henricia sp.*           | x    |        |             |
| *Holothuroidea 1*        | x    |        |             |
| *Holothuroidea 2*        | x    |        |             |
| *Odontaster penicillatus*| x    |        |             |
| *Odontasteridae*         | x    |        |             |
| *Ophiacantha vivipara*   | x    | x      | x           |
| *Ophicactis asperula*    | x    | x      | x           |
| *Ophiolimna antarctica*  | x    |        |             |
| *Ophiura lymani*         | x    |        |             |
| *Perkaster sp.*          | x    |        |             |
| *Porania antarctica*     | x    |        |             |
| *Pseudochinus magellanicus* | x  | x      | x           |
| *Pseudocnus dubiosus*    | x    | x      | x           |
| *Psolus sp.*             | x    |        |             |
| *Pteraster aff. lebruni* | x    |        |             |
| *Schizasteridae*         | x    |        |             |
| *Stereochinus agassizii* | x    |        |             |
| **Pycnogonida**          | 1    | 1      |             |
| *Pantopoda*              | x    |        |             |
| **Polychaeta**           | 13   | 7      | 9           |
| *Idanthyrsus macropalea* | x    |        |             |
| *Nephtyiidae*            | x    |        |             |
| *Nereidiidae*            | x    | x      | x           |
| *Oenoniidae*             | x    |        |             |
| *Orbiniidae*             | x    |        |             |
| *Paraonidae*             | x    |        |             |
| *Eteone sp.*             | x    | x      |             |
| *Polynoidae 1*           | x    |        |             |
| *Polyeunoa laevis*       | x    | x      | x           |
| *Potamilla sp.*          | x    |        |             |
| *Sabellidae 2*           | x    |        |             |
being primnoid corals (Thouarella spp.) the most conspicuous organisms in this site. Two individuals of Flabellum cf. thouarsi, several E. antarctica pieces and 2 Alcyonium sp. colonies were recorded. Ophiura lymani was the most abundant ophiuroid collected, and Gorgonocephalus chilensis was also important in terms of the relative observed biomass in the catch. Other 12 echinoderm taxa, 12 sponges, 9 polychaetes, 1 hydrozoan colony, 1 ascidian, 3 decapods and 3 peracarid crustaceans (isopods), 1 pycnogonid, 3 sea anemones, 1 brachiopod and 2 fish species were recorded.

Similarity index (SI) obtained for core and buffer areas was 0.3066; for buffer and shelf break, it was 0.1994; and for core and shelf break, it was 0.1881.

**Discussion**

This contribution represents the most recent and complete checklist of benthic organisms (mainly mega and macrofauna) that summarized 240 taxa collected at three sites in the Namuncura Marine Protected Area, at Burdwood Bank (Table 2). Only partial information by taxonomic group (e.g., fishes, mollusks, crustaceans) is available from previous studies performed in the current N MPA (see a general compilation at Falabella et al. 2013). The Ocean Biogeographic Information System (OBIS) Web site recorded no information for the three sites here studied and reported only 90 taxa in the transect sampling line that contains the stations that were carried out in the present study (checked at http://www.iobis.org/ in December 2015). Previous studies performed in the slope area of the BB (Arntz and Brey 2003; Schejter et al. 2012) also reported a rich and diverse benthic community. We are well aware that one sample per site is not enough to fully characterize the benthic assemblage, but still, in the present scenario, these data represent a valuable source of information and a very helpful starting point for incoming research, conservation efforts and management. To generate complete inventories at species level of big areas is a very expensive and time-consuming venture. Studies that used taxa higher than species level (e.g., genus and family) as surrogates for predicting species level diversity in marine areas proved to be useful, and taxa assemblages together with the information of endemic, vulnerable and threatened taxa help to evaluate areas that merit conservation efforts (Vanderklift et al. 1998). The National Marine Scientific Policy and Strategy in Argentina includes the scheduling of research cruises in this MPA (among other regions) in the years to come in order to improve data and management of ecologically important areas in this country.

Faunistic composition of the three sampled sites at BB was different, although some species were shared among them (Table 2). The benthic assemblage collected from the buffer area shared more than 50 % of the recorded taxa with the core area (mainly bryozoans, hydrozoans and decapod crustaceans). Only 30 % of the taxa were shared among shelf-break and buffer areas and also 30 % among core and shelf-break regions (mainly bryozoans in both cases). Similarity indexes also reflected these differences among stations, being core and buffer regions more related among them than the shelf-break area. These differences highlighted the importance of a better characterization of

|          | Core | Buffer | Shelf break |
|----------|------|--------|-------------|
| Serpula sp. | x    |        | x           |
| Syllidae 1 | x    |        |             |
| Syllidae 2 | x    |        |             |
| Syllidae 3 | x    |        | x           |
| Syllidae 4 |      |        | x           |
| Syllidae 5 |      |        |             |
| Pista sp.  |      |        |             |
| Terebellidae |     | x      | x           |
| Osteichthyes | 5    | 2      | 2           |
| Patagonothoten ramsayi | x    | x      | x           |
| Cottoperca gobio | x    |        |             |
| Agonopsis chiloensis | x    |        |             |
| Patagonothoten cf. guntheri | x    |        |             |
| Myxine sp. | x    |        |             |
| Total     | 148  | 109    | 92          |
the benthic communities that inhabit the Namuncurá MPA and the nearby shelf break. This huge extension presents not only bathymetric variations and particular oceanographic conditions (Zunino and Ichazo 1979; Piola and Gordon 1989; Guerrero et al. 1999) but also a different disturbance history in the whole area related to fishing restrictions (see Introduction) with the consequent variations on recovery times of the target and nontarget species involved in the fishing operations. Affinities with other benthic communities of the Argentinean shelf are evident. At least at the core region in areas of about 100 m depth, faunal species recorded were frequent taxa at Patagonian scallop benthic community distributed along the Argentinean shelf break front at similar depths (Bremec and Lasta 2002; Ganzano et al. 2009; López Gappa and Landoni 2009; Sánchez et al. 2011; Schejter et al. 2012). On the other hand, it is possible that shelf-break fauna of BB caught at more than 200 m depth is similar to regions sampled at similar depths outside the Argentinean continental shelf in northern latitudes (see Portela et al. 2012, 2015).

Considering the phyla here recorded, bryozoans presented the highest richness compared to the other groups, although among the three areas, the lowest richness was reported for the shelf-break area. This fact could be due to the smaller size of the benthic sample compared to the others. Bryozoa is one of the best studied phyla from the given list; this group has been revised from samples collected from more than 30 years ago to the present by the same taxonomist (López Gappa and Lichtschein 1990; López Gappa 2000). Another important contribution was carried out by Hayward (1980). Also, additional records from outside the MPA coming from the LAMPOS expedition mentioned in the Introduction section can be found in Moyano (2005).

High abundances of recent and fossil corals from Burdwood Bank slope and other nearby areas are known from the literature (e.g., Zamponi 2008a, b; Margolin et al. 2014). In accordance with this, a high diversity of anthozoans (octocorals, scleractinians and sea anemones) was previously found outside the MPA, in the eastern side of the bank (López-González et al. 2003), as well as in the west (Margolin et al. 2014). However, during the present study, anthozoans were only registered in the shelf-break area of the BB, a site where relatively high abundance and richness of these organisms were recorded. Corals, coral gardens and reefs are also present and frequent beyond the shelf break of Argentina (Muñoz et al. 2012; Cairns and Polonio 2013; Portela et al. 2012, 2015), even though they could also be found in lower abundances at the Argentinean shelf, below 200 m depth (Zamponi 2008b).

Sponges were not shared among areas, except for one species (Mycale sp. 1). Although not quantified, this phylum was responsible for the main biomass in the core area, probably the less known area regarding spongofauna. Other previous sponge reports from BB came from sampling sites located in the transition area or beyond the shelf break (López Gappa and Landoni 2005; Schejter et al. 2012).

Peracarid’s species richness collected with the otter trawl underestimated the real values since these organisms, as well as other small invertebrates, require a different sampling method. During the same research cruise, a Rauschert sledge equipped with a 1 mm mesh size took three additional samples in the same sites (material not available for this study). A preliminary revision showed a remarkable high peracarid species richness. However, these results were not included in this study. At least 90 species (including amphipods, isopods, cumaceans and tanaidaceans) were identified and many of them represented new distributional range records (Doti et al. 2014; Chiesa et al. 2015).

The benthic richness here documented largely exceeds the values coming from other productive areas in Argentinean waters where different trawling fisheries are conducted, even considering the limitations of this inventory. In this sense, this area should be considered a hot spot of benthic biodiversity. Other important but less biodiverse areas of the SW Atlantic Ocean are located on the continental shelf between 38°S and 45°S, at the Patagonian shelf-break fishing grounds. In these areas, species richness was not higher than 70 species per site, considering a maximum of 38 megafaunal taxa, 15 bryozoa and about 17 sponges and other epibiotic taxa (see Bremec et al. 2000, 2008; Schejter and Bremec 2007, 2009; Schejter et al. 2011a, b, 2013; Escolar et al. 2009; López Gappa and Landoni 2009). Also, the Argentinean–Uruguayan Common Fishing Zone (35°S–39°S) and the Patagonian shelf (44°S–47°S) (common hake fishery, see Giberto et al. 2014) showed lower values of benthic richness. In addition to the shelf areas mentioned above, 86 taxa of mega and macroinvertebrates were found using a dredge of 10 mm mesh size in a submarine canyon at 43°35′S and 59°33′W, 325 m depth (Bremec and Schejter 2010). The faunal composition of the latter area was similar to that referred to in adjacent external shelf areas, mainly between 90 and 130 m depth (Bremec and Lasta 2002). Moreover, after studying epibiotic relationships in the mentioned canyon, species richness was increased to 127 taxa (Schejter et al. 2014), a value still lower than the richness here reported for BB. Research on benthic biodiversity using samples collected at the Scotia Arc and also referred that BB contributed much to diversity in the study area, such as the case of gorgonians, gastropods and bivalves (Arntz and Brey 2003). Roux et al. (2002) found a richness of a maximum of 27 taxa per site at South Georgia Islands, using demersal trawls. Antarctic studies at Deception Island also revealed a similar richness.
of 35 taxa per sample (Lovell and Trego 2003). Although benthic richness in the different regions of Argentina and Antarctica has been estimated using different sampling methods, we consider that the present results on mega and macrofauna are useful as a general reference regarding the benthic richness of the marine realm.

In the case of the N MPA, an extensive review of scattered benthic information, future sampling cruises, possible ROV images and sediment sampling studies will help to update species inventories. It will also help detect indicator taxa vulnerable to trawling, recognize distribution patterns of the benthic organisms and support management strategies. As epibiotic relationships greatly increase benthic richness in other southern benthic systems, similar studies should also be conducted. It is also essential to establish the spatial distribution of indicator taxa, like corals, sponges and bryozoans, to better attempt to the protection of benthic biodiversity in the rich Argentinean marine system herein studied. Furthermore, this information will provide a stronger idea about the biogeographic importance of the Burdwood Bank, either as center of endemisms and/or as connection for marine benthic species between South America and the Antarctic Peninsula.

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