FELINE UVEITIS
An ‘intraocular lymphadenopathy’

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Classifying uveitis in clinically useful ways

The uvea contains familiar tissues and cell types (eg, lymphocytes, smooth muscle, blood vessels), is inflamed by familiar antigens (infectious agents, neoplasia, autoantigens) and reacts with the five cardinal signs of inflammation seen elsewhere (heat, pain, swelling, redness and loss of function). There are numerous systems for categorizing uveitis. Those emphasized here permit directed and prioritized diagnostic testing, guide selection of appropriate therapy while test results are pending, and inform predictions regarding visual outcome and, in some cases, prognosis for life.

What parts of the uvea are involved?

Uveitis is inflammation of the middle vascular (uveal) tunic of the eye, which is located between the outer fibrous (corneoscleral) tunic and the inner neurosensory (retinal) tunic (Fig 1). The uvea is composed of just three structures: the choroid posteriorly and the iris and ciliary body anteriorly. These anatomic divisions explain terms frequently used to categorize uveitis. Anterior uveitis (sometimes termed iridocyclitis) is inflammation of the iris and ciliary body. Posterior uveitis is inflammation of the choroid; however, because the retina and choroid are so intimately associated anatomically and functionally, retinal inflammation inevitably occurs along with choroiditis and so posterior

Practical relevance

Feline uveitis can be a subtle, insidious, painful, vision-threatening disease with causes that can sometimes be fatal. It is essential that clinicians remain alert to its various clinical presentations, thoroughly diagnose cases once detected, and treat the primary cause whenever possible.

Clinical challenges

In the majority of patients, a cause is not found and aggressive immunomodulating therapy of what may become a chronic or recurrent immune-mediated disease must be instigated. As with immune-mediated diseases elsewhere, this involves local or systemic immunosuppression with slow tapering and frequent monitoring. The aim of this review is to aid diagnosis and therapy of uveitis by likening it to inflammation elsewhere (because it is more similar than it is different) while highlighting differences (because these are helpful).

Global importance

Feline uveitis is similar in its presentation throughout the world. Although the list of infectious causes may vary in composition or order of likelihood, idiopathic, immune-mediated and neoplastic causes of feline uveitis are universal.

Patient group

Patients of either gender and all ages and breeds are affected by uveitis.

Evidence base

Despite the fact that feline uveitis is a serious and common disorder, the peer-reviewed literature regarding this disease is somewhat limited. Approximately half the publications are review articles, case reports or case series. The majority of prospective and retrospective research describes epidemiologic surveys of antibodies, antigens and organism DNA in serum and aqueous humor.
Uveitis has few pathognomonic signs and these are notably more subtle in cats than they are in dogs. Therefore, uveitis in cats often goes undetected by owners and untreated by veterinarians until potentially blinding sequelae such as glaucoma, cataracts and retinal detachment or degeneration occur.

uveitis is usually called chorioretinitis. Other anatomic classifications include panuveitis (inflammation of the choroid, iris and ciliary body), and pars planitis or intermediate uveitis (inflammation of the posterior ciliary body, or pars plana, and anterior choroid).

While these anatomic divisions are useful, inciting antigens and effector cells and cytokines rarely totally respect them and pure anterior or posterior uveitis should never be assumed. For example, in a patient with hyphema a dilated fundic examination should always be conducted for the presence of chorioretinal hemorrhage. Likewise, a patient with chorioretinitis should always undergo intraocular pressure (IOP) measurement (tonometry) to assess any involvement of the anterior uvea. That said, some diagnostic, therapeutic and prognostic decisions can be made following determination of whether the patient has predominantly anterior and/or posterior uveal involvement.

- **Diagnostic implications**: Although many causative agents of uveitis can lead to panuveitis (eg, feline infectious peritonitis [FIP] virus), many tend to produce signs that predominate in either the anterior uvea (such as feline immunodeficiency virus [FIV]) or the posterior uvea (such as the systemic mycoses). Therefore, clinically localizing the uveitis to one or both anatomic sites can assist with ranking differential diagnoses as well as choosing and prioritizing diagnostic tests (Table 1).

- **Treatment implications**: Posterior segment disease must be treated via the systemic route while anterior segment inflammation may be treated with topical application of drugs that penetrate the cornea. Hence knowing the extent of uveal involvement is critical before choosing a route of therapy. Additionally, humans report that inflammation of the posterior uvea is not as painful as anterior uveitis. If this is also true in cats, then analgesia may be not be necessary in patients showing no ciliary body involvement.

- **Prognostic implications**: Although anterior segment inflammation can be blinding, chorioretinitis is more likely to be associated with vision-threatening outcomes. This reinforces the importance of posterior segment examination before and throughout the course of treatment.

What is the likely cause of the uveitis?
The causes of uveitis are diverse and sometimes elusive. Even with very complete diagnostic testing, a cause is not discovered in as many as 70% of cats with uveitis. Therefore, although feline uveitis can be subdivided into infectious, inflammatory, traumatic, neoplastic causes, etc, the most clinically relevant classifications are both two-category systems.

First, uveitis can be categorized (following detailed diagnostic investigation) as idiopathic or as having an identified cause. While this categorization may at first sight seem too broad to be of any clinical value, it actually permits some critical decisions to be made about therapy. For example, if a cause is identified, then therapy may be targeted at prompt and specific reduction or removal of the antigen (organism, neoplasm, etc) that is driving the immune response with minimal non-specific immunosuppression. An example would be a cat diagnosed with systemic cryptococcosis. Specific antifungal therapy with minimal and only local (ocular) immunomodulation is required and, if successful, should lead to resolution of the uveitis, perhaps with some scarring. By contrast, if after thorough diagnostic testing a cause cannot be found, then the antigen cannot be medically reduced or resolved and this patient will require aggressive non-specific immune damping with the expectation that recurrence might be likely. These two scenarios exemplify the fact that elimination of known causes of uveitis that could progress or even become fatal with immunosuppression is critical, and that categorizing patients into those with a known cause and those with idiopathic uveitis is a very important approach to treating these cats.

The second broad etiologic system involves categorizing uveitis as due to endogenous or exogenous causes. Examples of exogenous causes include trauma, lens luxation and ulcerative keratitis; the last produces a so-called 'reflex uveitis' via stimulation of the trigeminal nerve. Exogenous causes are usually more obvious than endogenous causes and tend to be associated with other ocular signs, which can be identified during a thorough ophthalmic examination. By contrast, endogenous uveitis includes those cases induced by immune-mediated, infectious or neoplastic disease, as well as idiopathic disease. In cats, endogenous causes tend to be more common, produce more subtle
clinical signs, require more diagnostic verification, and be more resistant to treatment than exogenous causes.

**TABLE 1** Typical clinical course, predominant location and suggestive signs of common causes of uveitis

| Cause                        | Clinical course | Typical location | Suggestive signs* |
|------------------------------|-----------------|------------------|-------------------|
| Trauma                       | Acute           | Anterior uveitis | Hyphema; AC fibrin; miosis; aqueous flare; hypotony |
| Reflex uveitis (due to ulcerative keratitis) | Acute           | Anterior uveitis | Miosis; aqueous flare; hypopyon (if ulcer is infected); hypotony |
| FIP                          | Subacute        | Panuveitis       | KP; aqueous flare; AC fibrin; hypopyon; retinal vascular engorgement and increased tortuosity; perivascular chorioretinal granulomas; retinal detachment |
| Lymphoma                     | Subacute        | Anterior uveitis | Hypopyon; hyphema; AC fibrin; aqueous flare; iridal thickening; iridal nodules; ruberosis iridis; iris bombe; secondary glaucoma |
| Systemic mycoses             | Subacute        | Panuveitis       | Hypopyon; hyphema; AC fibrin; aqueous flare; iridal thickening; ruberosis iridis; iridal nodules; ruberosis iridis; secondary glaucoma; chorioretinal granulomas; retinal detachment |
| Lens-induced uveitis         | Phacolytic (chronic) | Anterior uveitis | Aqueous flare; iridal thinning/atrophy; ruberosis iridis; posterior synechiae; mature/hypermature cataract; secondary glaucoma |
| Idiopathic                   | Chronic or recurrent | Anterior intermediate uveitis | Iridal thinning/atrophy; iridal nodules; ruberosis iridis; aqueous flare; KP; snow banking; vitreous debris/infiltirates; posterior synechiae; cortical cataract; secondary glaucoma |
| Primary uveal neoplasia      | Chronic         | Anterior uveits or chorioretinitis depending on site of tumor | Anteriorly located | Hyphopyon; hyphema; AC fibrin; aqueous flare; anterior iridal displacement; ruberosis iridis; vitreous debris/infiltirates; secondary glaucoma |
| FIV                          | Chronic         | Intermediate uveits | Vitreous debris/infiltirates; snow banking; iridal thinning/atrophy; ruberosis iridis; aqueous flare; posterior synechiae; cortical cataract; secondary glaucoma |

**How long has the patient been affected?** Classification of uveitis as acute or chronic is very important since exogenous uveitis tends to be acute and self-limiting; cases associated with systemic diseases tend to be subacute; and idiopathic cases tend to become chronic or recurrent (Table 1). Historical data gained through careful interview of the client and from repeat examinations will assist in determining chronicity. Additionally, there are a number of clinical signs (discussed later) seen only with chronicity.

**Is the uveitis bilateral or unilateral?** Bilateral uveitis tends to be associated with systemic causes and unilateral uveitis is more consistent with idiopathic disease or exogenous causes. However, this distinction cannot always be relied on and signs of uveitis may be very asymmetric in their magnitude or stage. Therefore, it is critical that both eyes are examined to avoid misdirected diagnostic testing, inadequate therapy, or an inaccurate prognosis with potentially blinding or even fatal consequences.

**Clinical signs of uveitis: a functional approach**

Uveitis has few pathognomonic signs and these are notably more subtle in cats than they are in dogs. Therefore, uveitis in cats often goes undetected by owners and untreated by veterinarians until potentially blinding sequelae such as glaucoma, cataracts and retinal detachment or degeneration occur. For these reasons, clinicians must maintain a high index of suspicion regarding uveitis in all cats with ocular disease and even those with non-specific signs such as lethargy, ‘hiding’, anorexia or fever.

An appreciation of the diversity and importance of normal anterior uveal functions facilitates recognition of the classic clinical signs and devastating consequences of uveitis (Table 2).
The anterior uvea is responsible for:
- aqueous humor formation by the ciliary body epithelium and therefore nutrition of the avascular cornea and lens;
- formation of the blood–aqueous barrier (BAB) by the ciliary body epithelium and vascular endothelium;
- determination of retinal illumination due to pupil size;
- accommodation due to alteration of lens curvature by the ciliary body musculature; and
- regulation of aqueous humor outflow through the iridocorneal angle (conventional outflow) and uveal parenchyma (non-conventional or uveoscleral outflow).

Given this diverse list, it is not surprising that minor disruptions in uveal function during uveitis can have far-reaching and vision-threatening effects within the globe.

The final function of the uveal tract listed in Table 2 (antigen recognition and processing) is critical in the pathogenesis of chronic or recurrent uveitis. Because there are no intraocular lymphatic vessels and therefore no ‘draining’ lymph node for intraocular tissues, the uvea plays a major role in immune surveillance and response within the eye. Its role can be likened to that of a lymph node or an extension of the reticuloendothelial system. In acute uveitis, this function is vital for normal identification, processing and clearance of inciting antigens. However, ongoing unbridled immunologic activity within the uvea, sometimes in response to an autoantigen or unrecognized foreign antigen, is responsible for chronic, low-grade, immune-mediated uveitis and serious sequelae. The extracocular analogy would be a reactive lymph node. Thus uveitis can be thought of as ‘intraocular lymphadenopathy’, since the list of differential considerations is almost identical for lymphadenopathy and uveitis, especially in chronic uveitis. This also introduces the therapeutically critical concept that, in idiopathic uveitis, recurrence is likely as immunomodulation is tapered.

### Manifestations of heat, pain, swelling and redness in eyes with uveitis

As mentioned, uveitis manifests as one or a combination of the five cardinal signs of inflammation: heat, pain, swelling, redness and loss of function. Heat is not readily palpated in the eye, as it would be in other soft tissue swellings such as cellulitis. Therefore, particular attention must be paid to evidence of the other four changes.

Intraocular pain is sometimes difficult to interpret in animals, especially cats. As in dogs, uveal pain in cats can manifest as blepharospasm or epiphora; however, cats seem far more likely to show subtle and less localizing signs such as lethargy, anorexia, or reduced interaction with their own or environment. This seems more akin to humans who complain of symptoms such as photophobia and headache or brow-ache when they have uveitis.

Swelling is an important sign of iridocyclitis. However, since the ciliary body cannot be observed clinically, iridal swelling must be relied on solely for this sign. Observation of iridal swelling requires that the eye is examined using a source of magnification (such as the Optivisor) in association with a bright and focal light source (such as the Finoff transilluminator) directed very obliquely across the globe (Fig 2). The normal iris is highly textured, with crypts and ridges across its entire anterior face (Fig 3a). In cats with uveitis, these become obscured by iridal swelling and the anterior iris face appears ‘muddy’ or flattened (Fig 3b). Cellular infiltration may also be evident as a homogeneous change in color of the iris, particularly in cats with lighter irides, or as nodular swellings.

Redness is one of the most overt signs of inflammation elsewhere in the body, but scleral injection is very subtle in many cats with uveitis.
It is made even less obvious by the fact that so little of the sclera is visible when the eyelids are in their normal open position. Therefore, feline eyes must be examined particularly carefully and great importance attributed to even subtle congestion of blood vessels present over the sclera. The temptation to diagnose any redness of this region as conjunctivitis must be avoided. Unlike dogs, uveitis in cats tends to produce more subtle vascular engorgement than conjunctivitis does. In addition, uveitis does not tend to produce notable chemosis (conjunctival edema), whereas conjunctivitis frequently does. Finally, it is important to distinguish deep episcleral vessels (which become injected in cases of uveitis, other intraocular diseases and deep corneal disease) from superficial conjunctival vessels (which indicate conjunctivitis or superficial corneal disease; Fig 4). Compared with superficial conjunctival vessels, deep episcleral vessels tend to be straighter, less mobile, of larger diameter, and to branch less. They also appear to stop 1–2 mm before they reach the corneoscleral limbus whereas conjunctival vessels proceed to and sometimes loop back at the limbus.

A minority of cats (in particular brachycephalic individuals) may normally have a single large conjunctival or episcleral blood vessel. This should not confuse the diagnosis since cats with uveitis and episcleral injection will reliably have other evidence of uveitis including engorgement of multiple vessels rather than a single vessel. Congestion of the iridal vessels themselves is typically disguised from the examiner by melanocytes and iridophores. When it is noted, redness of the iris usually indicates neovascularization and not congestion (see later discussion of chronic changes).

Loss of function produces the major clinical signs of uveitis
The final hallmark of inflammation – loss of function – produces most of the pathognomonic or highly characteristic signs of uveitis (Table 2). It is critical to be alert to these signs.

Breakdown of the blood–aqueous barrier
Breakdown of the BAB is a pathognomonic sign of uveitis. Therefore the anterior chamber should be comprehensively examined for the presence of free-floating cells or proteins that would normally remain within the intravascular compartment. This is no different to inflammation elsewhere with exudation of intravascular contents into the interstitial space. In the eye, the aqueous humor is equivalent to the interstitial space such that the extent and nature of exudation can be observed. Hypopyon (white blood cells; Fig 5), hyphema (red blood cells; Figs 5 and 6) and fibrin (Fig 7) are usually relatively obvious, whereas specialized examination techniques are necessary to detect aqueous flare (albumin and other small proteins suspended in the aqueous humor) and keratic precipitates (KPs) (white blood cells and inflammatory proteins clumped against the corneal endothelial surface; Fig 3b).

Aqueous flare is best detected using magnification and a very focal, intense light source in a totally darkened room. A slit lamp is ideal; however, the beam produced by the smallest circular aperture on a direct ophthalmoscope held as closely as possible to the cornea and viewed transversely with magnification will also provide excellent results (Fig 8). In the normal eye, a focal reflection is seen where the light strikes the cornea. The beam is then invisible as it traverses the almost protein- and cell-free aqueous humor in the anterior chamber and is visible again as a focal reflection on the anterior lens capsule, and then as a diffuse beam through the body of the normal lens. If uveitis has allowed leakage of serum proteins into the aqueous humor, this will scatter (and make visible) the light beam as it passes through the anterior chamber. A beam of light that is visible traversing the anterior chamber and joining the reflections from the cornea and anterior lens capsule therefore indicates aqueous flare.

Keratic precipitates may be observed during routine examination of the cornea (Fig 3b) but, when they are very small or few in number, their visualization may be enhanced using retroillumination.
illuminate each eye. The fundic reflection is used to assess the patient’s nose from at least arm’s length so as to equally examine the examiner’s eye and directed over the bridge of the transilluminator or direct ophthalmoscope held close to the best detected using retroillumination (see box) with the miosis. While the ciliary body cannot be observed during spasm as miosis) and ciliary body (evident as spasm of smooth muscle within the iris (evident as pain). Other signs of uveal dysfunction (Table 2) are Miosis and hypotony. Similarly, the absence of one or more of the types of exudate does not allow a cause to be ruled out.

Miosis and hypotony
Other signs of uveal dysfunction (Table 2) are attributable to prostaglandin-mediated spasm of smooth muscle within the iris (evident as miosis) and ciliary body (evident as pain). While the ciliary body cannot be observed during the ophthalmic examination, miosis can. Subtle miosis is best detected using retroillumination (see box) with the transilluminator or direct ophthalmoscope held close to the examiner’s eye and directed over the bridge of the patient’s nose from at least arm’s length so as to equally illuminate each eye. The fundic reflection is used to assess the ciliary body and subtle uveitis, IOP may be within the normal range (approximately 10–20 mmHg) but notably (> 20%) lower than the normal eye and so can still be classified as hypotony. If uveitis is complicated by impaired aqueous outflow, the IOP will be higher than the opposite eye and sometimes above normal. Coincident secondary glaucoma is a major concern in such cases.

In my experience, tonometry is one of the most sensitive tests for monitoring uveitis during treatment (see later).

Corneal edema
The cornea owes its transparency to a number of factors, among which a state of relative dehydration that is maintained largely by the corneal endothelium is critically important. With breakdown of the BAB and alterations in aqueous outflow, the composition of the aqueous humor is altered. Increased concentrations of inflammatory mediators and consumption of nutrients usually present are associated with dysfunction of the corneal endothelium and subsequent corneal edema. This appears as a bluish discoloration and loss of transparency of the cornea that can be very subtle in cats compared with dogs (Fig 4). It is best seen by viewing the cornea from directly in front of the cat while directing a focal light transversely across the cornea.

Clinical signs associated with chronic uveitis and its sequelae
As with inflammation at other sites of the body, chronic uveitis induces a different set of changes from those seen with acute uveitis. Like elsewhere, these are associated with scarring (fusion of one tissue to another and cicatrization), chronic dysfunction and neovascularization (Table 2). The presence of vascular components within the anterior chamber due to breakdown of the BAB can cause clogging (in the subacute phase) and ultimately scarring (in the chronic stage) of the aqueous outflow tract. This in turn causes raised IOP and ultimately glaucoma. Adhesions may also occur between adjacent tissues, most notably the iris and lens.
(posterior synechiae) or iris and corneal endothelium (anterior synechiae). Posterior synechiae are evident as melanotic debris adherent to the anterior lens capsule or changes in pupil shape (dyscoria), position (corectopia) or size (miosis or mydriasis). If posterior synechiae become extensive, accumulation of aqueous humor in the posterior chamber will cause a forward bowing of the iris (iris bombe') and secondary glaucoma. Peripheral anterior synechiae are less visible during ophthalmic examination but further obstruct the aqueous outflow and manifest as glaucoma. If severe enough, scarring within the globe may undergo cicatrization with subsequent contracture of the globe (phthisis) or tractional retinal detachment.

Altered aqueous humor composition and circulation also cause a relative malnutrition of the lens and inner cornea, with resultant cataracts and keratitis (corneal edema, vascularization, fibrosis, etc), respectively, once this becomes chronic. Lens luxation may occur due to enzymatic lysis or phagocytosis of the lens zonules, secondary to cataract development, or as a sequel to buphthalmos due to secondary glaucoma. Neovascularization of the face of the iris (rubeosis iridis) is a pathognomonic sign of subacute or chronic active uveitis.

What is a reasonable diagnostic approach to a cat with uveitis?

Having a high degree of clinical suspicion and performing a targeted clinical examination with appropriate ocular diagnostic testing (retroillumination, assessment of aqueous flare and IOP, and a dilated fundic examination – see box on the right) will ensure that uveitis is diagnosed when present. However, detecting uveitis is the beginning of the diagnostic process – not the end. Confirming or eliminating all suspected etiological diagnoses is the essential next step.

By conducting a thorough general physical and ophthalmic examination as well as gathering a focused history, the initial goal is to categorize the uveitis as present in a well or systemically ill patient; unilateral or bilateral; exogenous or endogenous; acute or chronic; and as involving the anterior uvea, choroid, or both. This will help to prioritize the diagnostic and ultimately the therapeutic approaches. I like to assemble evidence that strongly supports or does not strongly support the need for further diagnostic testing (Fig 10). The majority of cases of feline endogenous uveitis are infectious, neoplastic or immune-mediated in origin. Application of the standard ‘DAMNIT-V’ list will identify some less common causes of uveitis.

Since immune-mediated uveitis is a diagnosis of exclusion the emphasis in the following discussion is the diagnostic approach to patients with suspected infectious or neoplastic uveitis.

**Diagnostic tests for uveitis**

The following tests are essential to detect evidence of uveitis and should be conducted in the order shown:

- **Retroillumination** – to look for:
  - Anisocoria (especially due to miosis)
  - Discoridia
  - Corectopia
  - Iridal atrophy
  - Opacities in the clear ocular media (KPs, hypopyon, hyphema, vitreous debris, posterior synechiae, secondary cataract)

- **Oblique illumination of eye** – to look for:
  - Corneal edema
  - Iridal swelling/nodules
  - Iridal thinning/atrophy
  - Rubeosis iridis
  - Posterior synechiae
  - Iris bombe'
  - Hypopyon
  - Hyphema
  - Fibrin in the anterior chamber

- **IOP measurement (tonometry)** – to look for:
  - Decreased or low-normal pressure, suggesting uncomplicated uveitis
  - Elevated or high-normal pressure, suggesting uveitis complicated by glaucoma

- **Pupil dilation** – to look for:
  - Resistance to dilation
  - Snow banking

- **Assessment of aqueous flare** – to look for:
  - Serum proteins in the anterior chamber due to breakdown of the blood–aqueous barrier

- **Fundic examination** – to look for:
  - Signs of posterior uveitis (retinal detachment/degeneration, choriretinal granulomas, hemorrhage, edema)
  - Vitreous debris/infiltrates
  - Snow banking

- **Fluorescein staining** – to look for:
  - Corneal ulceration, which suggests exogenous (axonal) uveitis. Corneal ulcers also preclude use of topical corticosteroids

**Diagnostic approach for infectious uveitis**

An etiologic diagnosis in a case of infectious uveitis is usually reached in one or more of four ways.

Detection of infectious agent within ocular tissues or fluids Detection of the infectious agent directly within the eye is best suited to larger organisms. For example, migrating or...
aberrant parasitic larvae can be detected without magnification within the anterior chamber or vitreous.\textsuperscript{3,4} Fungal organisms, protozoa or bacteria\textsuperscript{5} may be detected histologically within biopsy tissue or cytologically following paracentesis of the anterior chamber, subretinal space or vitreous body. Culture of these aspirates may also be attempted; however, retrospective studies reveal that this is a low-yield approach.\textsuperscript{1,6} More recently, polymerase chain reaction (PCR) has been used to detect organism DNA in ocular aspirates or tissue samples. However, due to the extreme sensitivity of this technique certain organisms may be found intraocularly or through surface or blood contamination at the time of sampling in healthy animals, thus lowering the diagnostic sensitivity of this approach. Additionally, since breakdown of the BAB is a hallmark of uveitis, any organism present within the bloodstream can leak into the inflamed eye whether or not it is the cause of the inflammation. Therefore, intraocular detection of such organisms does not prove causation of the uveitis. For these various reasons, false negative and false positive detection of intraocular organisms or their DNA may occur in cats with uveitis.

Serologic evidence of the agent
The value of serology for diagnosing the cause of uveitis varies greatly with the organism. Circulating antibodies suggest exposure to that organism but do not necessarily suggest that exposure was recent, let alone current. For example, immunoglobulin (Ig) G titers to \textit{Toxoplasma gondii} may remain elevated for many years in healthy animals.\textsuperscript{7} IgM titers have been assessed as a means of detecting more recent infection. However, like IgG, detection of this antibody class simply reflects exposure to a given organism and not necessarily causation of the current clinical signs; furthermore, IgM titers may not be elevated in animals undergoing recurrent rather than primary disease.\textsuperscript{8} An additional complication is that antibody tests can be cross-reactive with related but sometimes non-pathogenic organisms. An example of this is the failure of serologic assays to discriminate between antibodies directed at the FIP virus and those targeting the feline enteric coronaviruses.

Finally, antibody tests typically do not differentiate natural exposure to the pathogenic organism from vaccinal exposure to a modified, non-pathogenic variant. A long-recognized example of this phenomenon is feline herpesvirus 1 (FHV-1), while the recent release of an FIV vaccine means that this organism must now be added to that list. Together, these shortfalls lead to serious concerns regarding the etiologic diagnosis of feline uveitis based on serologic testing alone. In one study, equivalent numbers of cats with and without uveitis had circulating antibodies to FHV-1,\textsuperscript{9} while in another study circulating antibodies specific to \textit{Bartonella} species were more common and of greater titer in cats without uveitis than in cats with uveitis.\textsuperscript{10}

Regardless, there are some organisms and situations in which antibody testing is useful. For example, because FIV is not usually cleared by the immune system, detection of circulating antibodies to FIV in an unvaccinated cat is considered confirmation of infection with that virus (although does not confirm that any uveitis present is caused by the virus). Conversely, negative titers can sometimes be used to exclude that organism as a cause, especially in cats with chronic uveitis. Detection of circulating antibodies, as is done for feline leukemia virus (FeLV) or \textit{Cryptococcus} species, is also extremely useful in confirming current infection with those organisms. For most other organisms, though, the major use of serology seems to be large, controlled epidemiologic studies.

Evidence of agent-specific intraocular antibody production
The questionable value of antibody detection in the serum has led to interest in the diagnostic value of intraocular antibody production by lymphocytes within the uvea. However, breakdown of the BAB permits serum antibodies to leak into the eye and these must be differentiated from locally produced antibodies. This is done by calculating the ratio of the antibody of interest to total antibody within the serum and within an intraocular fluid (typically aqueous humor). This is called the Goldmann-Witmer coefficient and is expressed as a C value (see box, page 175). If this ratio is greater intraocularly than in the serum (ie, C value > 1), intraocular antibody production is assumed.

However, the diagnostic utility of the Goldmann-Witmer coefficient is reduced by knowledge that non-specific immune stimula-
tion can cause specific intraocular antibody production. In one study, naive cats and cats previously exposed to *T. gondii* were exposed subcutaneously to inactivated *T. gondii* antigens or to Freud’s adjuvant. Previously infected cats produced intraocular antibody (C values > 1) even when stimulated with adjuvant alone. Of even more interest, *Toxoplasma*-naive cats had C values > 1 when stimulated with inactivated *Toxoplasma* antigens. These findings have two important implications. First, they confirm that the uvea can behave like a reactive lymph node in response to a distant and sometimes unrelated antigen. Secondly, they suggest that higher C value cutoffs might have more clinical utility. While this requires further investigation, there is some emerging support for this hypothesis.

Detection of suggestive changes outside the eye

In many cats with uveitis the organism may be detected at extraocular sites. This is particularly true for systemic mycoses where cytologic assessment of tissue aspirates, nasal discharge or draining skin tracts may yield evidence of a pathogen known also to cause uveitis. Although not pathognomonic, changes noted on evaluation of a serum biochemistry panel and complete blood count (CBC) can permit prioritizing of differential diagnoses or direct further testing for those organisms that cause systemic changes in conjunction with uveitis.

Diagnostic approach for neoplastic uveitis

Reaching an etiologic diagnosis in a patient with neoplastic uveitis tends to be somewhat easier than for infectious uveitis and relies on cytologic or histologic demonstration of tumor within the eye or at a distant site with signs consistent with metastasis. As such, the diagnostic approach replicates the investigation of neoplasia at other sites.

Intraocular neoplasia tends to manifest in one of three ways – breakdown of the BAB (with hyphema, hypopyon, intraocular fibrin or aqueous flare), ocular dysfunction as a result of a mass effect (dyscoria, anisocoria, corectopia, glaucoma, etc.), or as a visible mass within the eye. Uveal masses can often be missed because the majority of the uvea (choroid, ciliary body and posterior iris) is not visible during the customary external ocular examination, and because the ciliary body and posterior iris cannot be examined even after full pupil dilation. Thus, ocular neoplasia, whether it is a discrete mass hidden from view (such as a ciliary body adenoma) or diffuse uveal invasion of a metastatic tumor (such as lymphoma), should always be considered in patients with endogenous uveitis.

As for infectious agents, the diagnostic search typically begins outside the eye with a thorough general physical examination, CBC, serum biochemistry profile, urinalysis, and imaging of the abdominal and thoracic cavities. Due to the association with lymphoma, serologic assessment of FeLV and FIV status is wise. Any masses elsewhere, as well as lymph nodes, should always be aspirated. Since cytologic abnormalities are not always associated with lymphadenomegaly, aspiration of palpably normal lymph nodes should be considered, as there is no ‘draining’ lymph node for the eye, any lymph node can be aspirated. If a diagnosis cannot be reached with extraocular testing, then cytologic assessment of an aspirate of aqueous or vitreous humor, subretinal fluid (if the retina is detached) or an intraocular mass can be undertaken by those trained and experienced in these techniques.
What are the most common causes of endogenous anterior uveitis in cats?

The known causes of endogenous anterior uveitis in cats are expanding but still too few to explain the majority of cases. To my knowledge, the most recent reasonably large retrospective analyses of feline uveitis were published in 1991.1,12 Following very complete clinical examination and clinicopathological, serological and histopathological testing at that time, a cause was discovered in only 30% of 53 cats with uveitis in one of these studies. The most commonly identified agent in these cats was FeLV and the most commonly identified cause of uveitis was lymphosarcoma or melanoma.1 In the second study, 139 cats (158 globes) with such severe disease that their eyes were enucleated or the cats died or were euthanized were retrospectively assessed. A cause was found in only 67% of cases; the most common diagnoses were FIP, FeLV-associated lymphosarcoma and trauma.12

Infectious agents

Table 3 lists the microbiological causes of feline uveitis. Only the more commonly implicated and better understood agents are discussed here and, as with all diagnostic enquiries, clinicians should consider the previous and current geographic locale in which the patient has resided when ranking the likelihood of each agent.

Viral

Perhaps one of the best understood viral causes of uveitis is FIP. Signalment (younger patients often from a multiset environment),13 clinicopathologic findings (hyperglobulinemia, lymphocytopenia, and seropositivity to feline coronavirus)13,14 as well as a highly cellular panuveitis (KP, hypopyon, choroidal granulomas) often permit FIP to be elevated as a prime differential consideration in affected animals (Table 1). The majority of cases are seen with the non-effusive or ‘dry’ form of the disease and typically ocular signs coexist with other systemic signs such as fever, lethargy, anorexia or neurologic signs15,16. The major differential consideration is lymphoma and therefore this is one of the circumstances where aqueocentesis is worth considering. Although pathognomonic cytologic changes are not seen with FIP, patients with lymphoma may have neoplastic cells within the anterior chamber. Additionally, I have found the positive (89%) and negative (99%) predictive values of the triad of hyperglobulinemia, lymphocytopenia and seropositivity to feline coronavirus reported by Sparkes et al14 to be particularly useful in diagnosing cats with uveitis believed to be due to FIP.

Aqueous humor samples from affected cats can be tested for the presence of feline coronavirus DNA. However, this does not provide definitive evidence of FIP as being the cause of uveitis because the PCR assay cannot differentiate FIP from enteric coronaviruses, because enteric coronaviruses in white blood cells can be detected with this assay, and because white blood cells are present intraocularly in uveitis resulting from many different causes.17–19

Cats presented with anterior uveitis are sometimes infected with FIV or FeLV; however, the role of these retroviruses is not always directly causal. In fact, FeLV is not a proven cause of anterior uveitis in cats20 and ocular disease is a very uncommon finding in cats seropositive for this virus.21,22 By contrast, FIV is suggested to be a cause of pars planitis,21,22 and one study found 84% of cats with advanced FIV infection to have lymphocytic/plasmacytic iridocyclitis when examined histologically.21 This is sometimes clinically evident as inflammatory cell accumulation in the anterior vitreous immediately behind the iris.23 However, this lesion, which is termed ‘snow banking’ (Fig 11), is not pathognomonic. In fact, it is an incidental finding in some patients with no apparent discomfort or other obvious signs of active uveitis and may represent slower removal of inflammatory cells from the vitreous than from the anterior chamber.

Notwithstanding, both retroviruses should be on the list of differential considerations for feline uveitis because they are important causes of lymphoma, which is the most common neoplastic cause of uveitis in cats. In particular, FIV-infected cats develop B-cell lymphoma,25 which can occur within the eye without lymph node or bone marrow involvement.26 FIV and FeLV can also permit superinfection with uveitogenic organisms via immunosuppression.

Bacterial

Theoretically, any organism responsible for bacteremia could also cause uveitis, but, in practice, this is extremely rare; perhaps due to the integrity of the normal blood–ocular barrier. However, there is increasing evidence that Bartonella species...

**TABLE 3** Infectious causes of feline uveitis

| Viral         | Bacterial                     | Fungal/algal | Parasitic   | Protozoal  |
|---------------|-------------------------------|--------------|-------------|------------|
| FIP15,16      | Bartonella species10,26,29    | Cryptococcus neoformans53† | Cuterebra4† | Leishmania species5 |
| FeLV47*       | Mycobacterium species46,48    | Histoplasma capsulatum16† | Toxoplasma gondii52 |
| FIV47         | Encephalitozoon species64†    | Blastomyces dermatitidis23† |            |            |
| FHV9          | Borrelia burgdorferi11†       | Candida albicans51†         |            |            |

*Via immunosuppression or oncogenesis, †Chorioretinitis predominates, ‡Seroprevalence data only (no clinical evidence)
are associated with and may even cause uveitis in some cats. An initial case report described a cat with uveitis without any other detectable cause. There was notable intraocular production of Bartonella species-specific antibodies (C value = 4.4) but intraocular detection of Bartonella DNA or culture was not attempted. The cat’s uveitis was unresponsive to a 4-week course of topical and systemic corticosteroids, but responded promptly and completely when doxycycline was added to the treatment regimen.

Subsequently, a number of epidemiologic studies have revealed that circulating antibodies to Bartonella species are common in experimentally infected cats and cats with uveitis, and that intraocular antibody production and organism DNA are more common in cats with uveitis and in experimentally infected cats than in cats without uveitis. However, neither the presence nor titer magnitude of Bartonella species-specific antibodies in serum are diagnostically reliable.

**Fungal/algal**

The fungi capable of causing systemic mycosis in cats frequently cause uveitis, albeit the organisms encountered vary somewhat with geographic region. Histoplasma capsulatum and Cryptococcus species tend to be the most commonly reported. Less common causes include Blastomyces dermatitidis and Coccidioides immitis. Other fungi and algae are sometimes seen, particularly in immunocompromised patients. Although all can cause anterior uveitis, fungal elements seem to have a predilection for the posterior segment and so chorioretinitis predominates.

Regardless of ocular location, fungal uveitis tends to be extremely granulomatous, manifesting as ‘greasy’ KP’s, a highly cellular and fibrinous hypopyon, and granulomas within the retina and choroid, often with exudative retinal detachments. Most affected cats are systemically ill and fungal elements may be detected at extraocular sites; especially skin tracts, lungs, lymph nodes and bone. In patients with a blind and painful globe the diagnosis may reasonably be achieved by therapeutic enucleation with histopathology since the fungal elements are typically numerous and can be seen on light microscopy. Antigen detection is particularly useful for diagnosing cryptococcosis.

**Protozoal**

Protozoal species may also cause feline uveitis. Of these, T gondii has been the most extensively studied using techniques such as fulfillment of Koch’s postulates, intraocular detection of organisms, detection of circulating T gondii-specific antibodies or antigens, response to therapy, and detection within aqueous humor of T gondii-specific intraocular antibody production, antigens, DNA and immune complexes. Taken together, data from these studies make it clear that T gondii is associated with uveitis in some cats; especially chorioretinitis in systemically ill cats during primary exposure to the organism. However, the role of T gondii in the more insidious anterior uveitis of older, otherwise healthy cats and the correct means of diagnosing T gondii as the cause of disease in those cats are not well established. While reactivation of tissue cysts is one proposed mechanism, there is accumulating evidence that the presence of organisms within the eye may not be necessary. Rather, immunopathology due to an autoantigen, molecular mimicry or loss of immunotolerance may be involved. Unfortunately, no test(s) can currently be recommended to definitively diagnose toxoplasmic uveitis in systemically well cats because many normal cats are seropositive for the organism, intraocular antibody production can be induced even in T gondii-naive patients, and T gondii DNA can be detected in aqueous humor samples from cats without uveitis. However, lack of evidence of intraocular organisms along with evidence of immunopathology does endorse the use of topical corticosteroids in such patients.

Increasing evidence that Leishmania species can cause uveitis in dogs, as well as case reports of Leishmania amastigotes within the iris and ciliary body of cats with granulomatous panuveitis, suggest that this organism causes uveitis in cats. Intraocular examination via light microscopy, immunohistochemical assessment and PCR analysis appear to be useful for diagnosis.

**Neoplasia**

While the most common primary intraocular neoplasm is melanoma, this typically causes little or no uveitis. By sharp contrast, the most common metastatic ocular neoplasm – lymphoma – tends to be associated with marked breakdown of the BAB with hypopyon formation, fibrin exudation into the anterior chamber, and hyphema. Curiously, the degree of pain associated with this disease often appears less than might be expected from the severity of other signs of intraocular inflammation. The exception to this is when secondary glaucoma occurs, which can be quite frequent due to the highly cellular and fibrinous nature of the anterior chamber exudate.

Lymphoma can cause unilateral uveitis, sometimes without extraocular involvement. If the tumor cannot be diagnosed from an extraocular site, detection of neoplastic lymphocytes in an aqueocentesis sample may be helpful. Diagnostic testing aimed at discovering evidence of the tumor elsewhere should be performed first, however, both for staging the disease and because it is less invasive.

Despite the severity of intraocular inflamma-
Immunomodulatory agents
Corticosteroids are highly potent, available in topical or systemic forms, relatively inexpensive, generally well tolerated by cats, and can be administered at anti-inflammatory or immunosuppressive dosages. For these reasons, they are commonly used for feline uveitis. Their systemic use should be reserved until a definitive cause responsive to corticosteroids has been found or, failing this, causes known to be worsened by glucocorticoids have been adequately eliminated. In particular, systemic mycoses must be ruled out as potential causes. Likewise, patients in which lymphoma is possible and which would benefit from multidrug chemotherapy should not be treated with systemic corticosteroids alone.

By contrast, topical corticosteroids may be used safely even when an infectious or neoplastic cause might prevent systemic administration of the same drugs. This is because systemic effects are insignificant with short-term topical application. It is possible that topically administered corticosteroids may alter cytolitic findings and so, if safe, their use should perhaps be delayed until ocular centesis has been performed. Topical corticosteroids should never be used in the face of corneal ulceration because they can be associated with rapid worsening of the ulcer due to superinfection, collagenolysis, local immunosuppression and delayed wound healing. Prednisolone acetate (1% or 0.125%) and dexamethasone (0.1%) will penetrate intact corneal epithelium and reach the anterior uveal tract. Hydrocortisone (as found in many combined antibiotic–corticosteroid ophthalmic preparations) does not penetrate intracocularly and should not be used. The frequency of application should be tailored to the severity of the uveitis, starting as frequently as q 2 h and tapering as a clinical response is noted.

When safe, corticosteroids should be administered systemically for posterior uveitis and when more significant immunomodulation is necessary, or when corneal ulceration prohibits their topical use. Typical doses of prednisolone range from 1 mg/kg q 12 h when notable inflammation is present to 0.5 mg/kg once daily when a more moderate anti-inflammatory effect is desired. As with topical corticosteroids, dose and dose frequency of systemically administered glucocorticoids should be carefully reduced based entirely on clinical evidence of waning disease. In cats with acute/subacute uveitis, this can be fairly rapid. Cats with chronic idiopathic (immune-mediated) uveitis require slow tapering (eg, a halving of dose or dose frequency every 2–3 weeks), with the expectation that inflammation may return below a critical dose. In these patients, returning to the previously effective dose will be necessary. Some cats will suffer herpetic recrudescence when receiving corticosteroids, regardless of route.

Compared with corticosteroids, non-steroidal anti-inflammatory drugs (NSAIDs) are not immunoosuppressive, and in some countries are more expensive, sold in smaller volumes and may not be available in ointment form. These limitations make them a generally less useful group of drugs than corticosteroids for cats with uveitis. However, they may be preferred in patients with diabetes or other endocrinopathies in which corticosteroid use may not be wise. They can also be administered systemically in place of corticosteroids when systemic infectious disease is suspected or proven, or until lymphosarcoma is eliminated as a differential consideration. As such they may make an excellent choice for initial control of inflammation while likely causes are being ruled in or out. The above general comments regarding dose frequency and route of administration of corticosteroids apply equally to NSAIDs. Also, this class of drugs is known to reactivate human herpesviruses and could do likewise in cats with FHV-1.

Iridocycloplegic agents
Parasympatholytic drugs such as atropine have multiple favorable actions in eyes with uveitis and form a critical component of treatment. These drugs paralyze the parasympathetically innervated iris sphincter and ciliary body muscles, causing mydriasis and cycloplegia, respectively.

Pupil dilution has numerous important effects. It reduces leakage of vascular elements into the aqueous humor by causing radial blood vessels within the iris stroma to ‘concertina’ (providing a physiological tamponade); iris surface area is decreased (from which inflammatory mediators and vascular components originate); uveal vascular endothelial permeability is reduced; and the risks and consequences of posterior synecchiation are diminished. However, ‘bunching’ of the iris in the periphery does increase the risk of anterior synecchia and potentially obstruction of the iridocorneal angle.

Cycloplegia reduces ocular pain but also increases resistance to aqueous outflow. Therefore, pupil dilution and cycloplegia are desirable in all cases of uveitis except those where secondary glaucoma is present or likely. In these patients, the effect of mydriasis on IOP can be tested by a single application of the short acting drug tropicamide followed by tonometry when the pupil is fully dilated. If IOP is increased by tropicamide, atropine should not be administered. If atropine therapy is initiated, IOP should be rechecked regularly and the drug discontinued if IOP increases above normal. An ophthalmic ointment formulation rather than a solution should be used because atropine is bitter and passage down the nasolacrimal duct can cause violent salivation and frothing, which is harmless but disturbing to the cat and its owner. Atropine should be applied to effect. Since cycloplegia cannot be observed, the pupil is used for monitoring dose. Depending on the severity of uveitis, once to twice daily application may be needed for the first day or two to open the pupil. Subsequently, once to twice weekly application will often keep the pupil mydriatic. Posterior synecchia will not be resolved with atropine and will prohibit use of pupil size as an indicator of drug efficacy. However, atropine should still be administered to patients with synecchia since the analgesia resulting from cycloplegia should not be affected.

Autoimmunity
The only common type of uveitis in cats in which autoimmunity is confirmed is lens-
induced uveitis. This occurs in two forms: phacolytic uveitis, where there is slow leakage of lens proteins across an intact lens capsule, as occurs with advanced cataracts; and phacolastic uveitis, which is a more severe form associated with lens capsule rupture and resultant rapid leakage of large amounts of lens proteins. In both forms (phacolastic particularly so), lens proteins are released in amounts that exceed immunotolerance, leading to a true autoimmune response. Phacolytic uveitis occurs far less frequently in cats than in dogs due to the relatively low incidence of feline cataracts; especially diabetic cataracts with which this syndrome is most commonly seen. By contrast, phacolastic uveitis represents a significant concern in cats not only due to its severity but because a percentage of cats with traumatic lens injuries develop post-traumatic intraocular sarcoma, which can be fatal, usually through extraorbital invasion but sometimes due to metastasis. For this reason, eyes with traumatic lens capsule rupture should be aggressively treated and very carefully monitored for the life of the cat. Traumatized eyes that are blind should be enucleated prior to development of this tumor.

Aside from lens-induced uveitis, many other uveitides are proposed to be immune-mediated although specific autoantigens have not been identified. Various immunological mechanisms are possible including loss or failure of development of immunotolerance to autoantigens usually sequestered within the eye behind the blood–ocular barrier or exposure of autoantigens during breakdown of the blood–ocular barrier due to an unrelated inflammatory insult. These autoantigens remain the target of ongoing immunologic activity after the primary insult has been resolved. Molecular mimicry, whereby autoantigens sufficiently like foreign peptides of infectious agents initiate or perpetuate intraocular immune responses, may also be important in immune-mediated uveitis. This has been shown for other species such as the horse, in which the cornea and lens contain proteins very similar to those found in lepto-prirosis. Finally, it is possible that normal host proteins within the eye become altered such that they become antigenic.

These hypotheses help explain the common scenario whereby a thorough antigen search is unrewarding and the patient improves with empirical anti-inflammatory therapy, only to deteriorate as this is tapered below a critical level.

What are the major treatment decisions?

Treatment of feline anterior uveitis must be tailored to the individual case based on proven or suspected cause, severity, anatomical location, chronicity, and the presence of systemic or other intraocular disease. Regardless, some general therapeutic guidelines are possible. Optimal treatment relies on identification and removal or reduction of the causative antigen; however, this is rarely possible. Additionally, all patients with uveitis need their intraocular inflammation controlled rapidly and completely, since it is painful and produces vision-threatening sequelae. Thus, immunomodulating drugs form the mainstay of therapy for uveitis. The major decisions are therefore which immunomodulating drugs should be given, via what route and at what dose (see box, page 178).

Potential sequelae and monitoring of treatment

Prompt specific treatment of uveitis with tapering of therapy based on reduction of clinical signs may result in some sequelae but these are usually mild and should be static. If they are not, this suggests chronic or recurrent uveitis and further investigations and treatment are necessary. Classic sequelae include corneal fibrosis, cataract or posterior synechiae. None of these changes should result in pain or, unless severe, vision disturbance. By contrast, more severe, unrecognized, persistent or recurrent uveitis frequently results in a blind and sometimes painful globe.
George, a 16-month-old male castrated domestic shorthair cat, presented with ‘cloudiness’ of both eyes which his owners had observed over the past 24 h.

Pertinent history  George was found as a feral kitten. After a short period in a humane shelter, he was adopted by his current owners at about 5 months of age. He now lives indoors with one other indoor cat. His owners state that there is a rat problem in the home but that neither George nor the other cat hunts. Other than a short episode of upper respiratory disease many months ago, which was diagnosed as FHV-1 infection, George has been in good health since they obtained him. Over the past 3–4 weeks George’s owners have noticed a reduction in his energy levels: he has been playing less with his favorite toy, and more recently has just been sitting quietly on the couch. Over the past week, George has appeared ‘wobbly’, more lethargic, and his appetite has decreased. Yesterday, they observed the cloudiness in his eyes. He has no history of vomiting, diarrhea, coughing or sneezing, and his thirst and urination patterns appear normal.

Ophthalmic examination On full ophthalmic examination both eyes were open and appeared comfortable, and George behaved as if he was at least partially sighted. There was no overt ocular discharge from either eye. On retroillumination, the fundic reflection from the left eye (OS) was generally reduced or ‘hazy’, especially in the ventral half; no fundic reflection could be elicited from the right eye (OD). No overt anisocoria was noted. Sequential examination of both eyes revealed normal eyelids and third eyelids. The anterior chamber of both eyes (OU) contained a fibrin clot and 1+ (out of 4+) aqueous flare. The anterior chamber OU was notably shallow due to a markedly swollen iris, potentially with iris bombé. There was marked rubecosis iridis OU. The left pupil was displaced medially (corectopia). The direct pupillary light reflex was present OS and a consensual reflex was present from OD to OS. It was not possible to assess movements OD due to the anterior chamber fibrin. Neither pupil dilated after two topical applications of tropicamide. This (along with the anterior chamber fibrin) limited examination of the lens, vitreous and fundus, but no further abnormalities were noted OU. Fluorescein stain was not retained by either cornea and was visible within 2 mins at both nares.

The most common sequelae include cataracts (20–36% of eyes), lens luxation (11–18%), glaucoma (16–46%) and enucleation (29%). Many cats experience more than one of these and thus frequent and careful monitoring of a patient with uveitis is essential. This should be performed as for patients with immune-mediated disease elsewhere, with gradual tapering of medications and re-examination at doubling intervals assuming there is improvement; more often if there is not. Re-examination and tapering of medications should be continued until there is complete resolution of every clinical sign of active uveitis.

I believe that tonometry is the most sensitive test with which to monitor uveitis during treatment because subtle hypotony (sometimes only relative to the contralateral eye) can continue long after other more overt signs have normalized. Ongoing treatment of these patients may prevent or delay the development of sight-threatening ocular complications.
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The fact that George has bilateral chronic, endogenous, fibrinous uveitis and is systemically unwell strongly supports stage 1 testing (see box, page 175). Stage 2 testing could be initiated simultaneously or delayed for prioritization based on stage 1 test results if preferred. George’s owners elected to perform all stage 1 and some stage 2 tests.

Measurement of IOP
IOP should always be measured in cats with uveitis. George’s IOP was 20 mmHg OD and 14 mmHg OS. Given that marked anterior uveitis should be associated with low IOP; these pressures are consistent with ocular hypertension/glucoma OD and suggest it may be beginning OS.

General physical examination
George was apparently bright and responsive. He was in good body condition and weighed 6.1 kg. His temperature was 39.2°C and his pulse and respiratory rates were 138 beats per minute and 24 breaths per minute, respectively. No abnormalities were detected on thoracic auscultation; however, the kidneys appeared enlarged and there was a mobile caudal abdominal mass. Lymph nodes were normal on palpation.

CBC, biochemistry profile and urinalysis
Hematology revealed lymphopenia and a mature neutrophilia, which was interpreted as a stress leukogram. There were no abnormalities on the serum biochemistry panel and urinalysis.

Retroviral testing
FeLV, FIV and coronavirus serology were all negative.

Further diagnostics
Abdominal ultrasound revealed bilaterally enlarged kidneys with considerable hypoechoic tissue in the subcapsular space and mild pelvic dilation. A large intramural mass was also noted in the distal jejunum. Several mesenteric lymph nodes were mildly enlarged, hypoechoic and round. There was scant ascites. The intestinal mass and kidneys were aspirated for cytologic assessment.

5 Which of the following best ranks (in decreasing order) the likely differential diagnoses for George’s uveitis?
(a) Toxoplasmosis, lymphoma, bartonellosis.
(b) Lymphoma, FIP, metastatic neoplasia.
(c) Idiopathic/immune-mediated, melanoma, FIV.
(d) FeLV, idiopathic/immune-mediated, trauma.

Diagnosis
Toxoplasmosis was the most likely diagnosis for George’s chronic fibrinous uveitis. Endogenous fibrinous uveitis is a systemically occurring disease and is usually unilateral. Primary ocular toxoplasmosis is thought to be a systemic, lymphocytic inflammatory response to infection. This systemic response may be due to: 1. disseminated toxoplasmosis; 2. pseudoneoplastic reaction (ankylosing spondylitis-like uveitis); 3. immune-mediated (primary or secondary). Feline uveitis is a consideration when to review the history of the cat in order to rule out other causes of the disease.

Case outcome
George’s uveitis was treated with oral antibiotics, glucocorticoids and immunosuppressives, and the uveitis resolved over the next 6 weeks.George’s owners elected to have George neutered and have planned future monitoring for toxoplasmosis reactivation.

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