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RESEARCH ARTICLE | Neurogastroenterology and Motility

Histamine causes influx via T-type voltage-gated calcium channels in an enterochromaffin tumor cell line: potential therapeutic target in adverse food reactions

Beatrix Pfanzagl,1 Victor F. Zevallos,3 Detlef Schuppan,3 Roswitha Pfragner,4 and Erika Jensen-Jarolim1,2

1Institute of Pathophysiology and Allergy Research, Center for Pathophysiology, Infectiology, and Immunology, Medical University of Vienna, Vienna, Austria; 2The Interuniversity Messerli Research Institute, University of Veterinary Medicine Vienna, Medical University of Vienna, Vienna, Austria; 3Division of Molecular and Translational Medicine, Department of Medicine I, Johannes Gutenberg University, Mainz, Germany; and 4Otto Loewi Research Center for Vascular Biology, Immunology, and Inflammation, Medical University of Graz, Graz, Austria

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INTRODUCTION

At present, the triggers of adverse food reactions in individual patients are not entirely predictable. Innate mechanisms and neuronal factors may amplify the reactions, especially with preexisting inflammatory conditions. While most research so far has been dedicated to mast cells, eosinophils, and macrophages, we propose here that enterochromaffin (EC) cells might participate in and amplify adverse food symptoms or impact the threshold levels for clinical symptoms.

EC cells are dispersed throughout the intestinal mucosa and release serotonin (5-hydroxytryptamine) to the serosal side upon stimulation by nutrients or other chemical mediators, acidic pH, or mechanical distortion (6, 25). Their apical microvilli project in the gut lumen. They are found in close contact to nerve endings of submucosal neurons of the enteric reflex pathways, which are largely cholinergic (26, 36). Cholinergic agonists like acetylcholine (ACh) induce a rise in intracellular calcium ([Ca2+]i) that triggers exocytosis of serotonin-containing vesicles to the basolateral side (46). ACh (26) and serotonin (21, 29, 62) enhance gut secretion and motility, and high levels of serotonin can cause diarrhea (60). A role of serotonin in the pathology of inflammatory bowel disease, diarrhea-predominant irritable bowel syndrome (IBS), and celiac disease is also discussed (19, 20, 30).

The P-STS cell line (45, 48), isolated from a poorly differentiated neuroendocrine tumor of the terminal ileum, grows with a stable genotype (48) and was established as a reliable EC cell line by showing stable expression of the neuroendocrine vesicle components and tryptophanhydroxylase-1, the rate-limiting enzyme for synthesis of serotonin expressed specifically in enteroendocrine cells (44). The neuroendocrine identity of P-STS cells was confirmed by Hofving et al. (27), as was the neuroendocrine origin of to wheat amylase-trypsine inhibitors, suggesting that enterochromaffin cells are not directly involved in nongluten wheat sensitivity.

amylase trypsine inhibitors; calcium channels; enterochromaffin; histamine; otilonium bromide

NEW & NOTEWORTHY We show that histamine synergistically enhances the intracellular calcium response to the physiological agonist acetylcholine in human ileal enterochromaffin tumor cells. This synergistic activation and cell activation by histamine alone largely depend on T-type voltage-gated calcium channels and are inhibited by the antispasmodic otilonium bromide. The cells showed no response...
part of the TLR4-MD2-CD14 complex. (LPS), these effects of ATI require the presence of CD14 as cytokines. As established for bacterial lipopolysaccharideysis that are taken up with wheat and related cereals (67).

The [Ca²⁺], response to ACh plus histamine is more than additive. There was also a weak response to histamine 1 receptor (H₁R) that had not been noticed previously. We investigated the role of external calcium and of L-, N-, P-, R-, and T-type voltage-gated calcium channels (VGCC) in the responses of P-STS cells to ACh and histamine. Voltage-dependent calcium currents in primary ileal neurendocrine tumor cells cultivated for a few days have been attributed mainly to L-type VGCC (22).

Intracellular calcium can also be mediated by transient receptor potential (TRP) channels. Some of them can be activated by food constituents like cinnamonaldehyde [transient receptor potential channel ankyrin 1 (TRPA1)], capsaicin [transient receptor potential channel vanilloid 1 (TRPV1)], or piperine, trans-cinnamaldehyde (99%). A-23187 Ca²⁺ ionophore, nifedipine, GSK-1016790A, MPA, LPS from Escherichia coli 055:B5, bovine serum albumin (>98%), ACh chloride, histamine dihydrochloride, monosodium L-glutamic acid, and sodium butyrate were from Sigma Aldrich (St. Louis, MO).

**Isolation of wheat ATI.** Wheat ATI was isolated as described elsewhere (67). Briefly, wheat kernels were ground to flour, sieved (250 μm), defatted with methanol/diethyl ether, dried, and extracted with 50 mM ammonium bicarbonate/0.3 M sodium chloride (pH 7.8) at 4°C. The extract was fractionated with ammonium sulfate. The fraction precipitating between 1.8 and 4 M ammonium sulfate was redissolved, extensively dialyzed against 10 mM ammonium bicarbonate/0.3 M sodium chloride (pH 7.8), clarified by centrifugation, filtered (0.22 μm), and freeze-dried, resulting in a preparation with an ATI content of >60% of total protein known to contain mainly the isoforms CM3, 0.19, and 0.28, as determined by mass spectrometry (67). Biological activity and endotoxin content below 0.1 ng/ml were assayed as described elsewhere (67). Before use, the ATI preparation was resuspended in water at a concentration of 30 mg dry wt/ml.

**Immunofluorescence microscopy and [Ca²⁺], imaging.** For immunofluorescence microscopy, cells were grown on cover slips and analyzed as described previously (44). Briefly, cells were fixed with 4% paraformaldehyde in phosphate-buffered saline (PBS) and permeabilized for 5 min with 0.2% vol/vol Triton X-100 in PBS. Blocking and incubation with antibodies took place in 5% fetal calf serum in PBS at room temperature. Nuclei were stained with Hoechst dye 33342. Where indicated, antibodies were preincubated for 2 h at room temperature with or without a fivefold (weight/weight) amount of blocking peptide corresponding to their epitope before addition to the fixed and permeabilized cells.

**METHODS**

**Cell culture.** P-STS midgut neurendocrine tumor cells [semi-adherent, originally isolated from a World Health Organization III neurendocrine tumor of the terminal ileum (48)] were grown in a 1:1 mixture of medium-199 and Ham’s F-12 nutrient mixture supplemented with 10% heat-inactivated fetal calf serum, 100 U/ml penicillin G, and 100 μg/ml streptomycin at 37°C in a humidified atmosphere containing 5% CO₂. HEK 293 human embryonic kidney cells and THP-1 monocytes were grown in DMEM and RPMI 1640 medium, respectively, containing the same supplements. THP-1 cells were differentiated for 2 days with 200 nM phorbol 12-myristate,13-acetate (PMA) before immunofluorescence staining or treatment with LPS. All cell lines were tested for mycoplasma contamination by staining with Hoechst dye 33342.

**Antibodies and reagents.** Mouse anti-human TLR4 antibody (clone 76B357.1) was from Novus Biologicals (Littleton, CO); rabbit anti-human phospho-NF-κB p65(Ser⁵²⁶) (clone 93H1) was from Cell Signaling Technology (Cambridge, UK); mouse anti-human RelA/NF-κB p65 (clone 532301) and recombinant human tumor necrosis factor-α (TNF-α) were from R&D Systems (Minneapolis, MN); rabbit anti-human TRPV4 (directed against an epitope in the middle region) and the corresponding blocking peptide were from Aviva Systems Biology (San Diego, CA); Alexa Fluor 647-labeled mouse anti-human CD14 was from Biolegend (San Diego, CA); goat Alexa Fluor 488-labeled secondary fluorescent antibodies and fluo 4-AM were from Life Technologies (Carlsbad, CA); mouse anti-human H₁R (G-11) and the corresponding blocking peptide as well as anti-mouse IgG-horseradish peroxidase were from Santa Cruz Biotechnology (Dallas, TX); α-agatoxin IVA, 4-methylhistamine dihydrochloride, mepramine maleate, mibefradil dihydrochloride, and ranitidine hydrochloride were from Tocris (Bristol UK); α-conotoxin GVIA was from Smartox Biotechnology (Saint-Egrève, France); SNX-482 was from Peptanova (Sandhausen, Germany); oltionium bromide was from Seleckchem (Munich, Germany); l-epinephrine, capsaicin, piperine, trans-cinnamaldehyde (99%), A-23187 Ca²⁺ ionophore, nifedipine, GSK-1016790A, MPA, LPS from Escherichia coli 055:B5, bovine serum albumin (>98%), ACh chloride, histamine dihydrochloride, monosodium L-glutamic acid, and sodium butyrate were from Sigma Aldrich (St. Louis, MO).

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**[Ca²⁺], imaging with fluo 4-AM, quantification with Image J, and statistical analysis of [Ca²⁺].** responses from ranked data with the unpaired two-tailed t-test assuming unequal variances were done as described previously (44). Briefly, the cells were grown in 24-well plates to very low density and stained 45 min at room temperature with fluo 4-AM in serum-free medium. The medium was exchanged for 200 μl of pure medium, and, after 25 min of incubation at room temperature, photographs were taken before and after careful addition of potential agonists in 200 μl pure medium. The [Ca²⁺], response of fluo 4-AM-stained P-STS cells to addition of the indicated substances was determined as fluorescence enhancement 10 s after the start of
substance addition in relation to baseline fluorescence, if not indicated otherwise. The number of experiments conducted (n) is given in the legends for Figs 1–10.

For evaluation of the effect of external calcium, cells were washed two times with HEPES-buffered saline, pH 7.4, without Ca\textsuperscript{2+} after incubation with fluo 4-AM and then incubated for 25 min in the same buffer with or without 1 mM Ca\textsuperscript{2+} before agonist addition.

RNA isolation, reverse transcription, and polymerase chain reaction. RNA from 50% confluent P-STS cells was isolated, purified, and quantified with TRIzol (Thermo Fisher Scientific, Waltham, MA) following the manufacturer’s instructions. In a total volume of 20 μL, cDNA was synthesized from 0.7 μg RNA using the iScript cDNA Synthesis Kit (Bio-Rad) as described by the manufacturer and kept at –20°C until use. Polymerase chain reaction (PCR) amplifications were performed with 0.1 μL cDNA. *Thermus aquaticus* Taq DNA polymerase and deoxyribonucleotides were from Sigma Aldrich. The amplification process was started with incubation at 95°C for 5 min. After 35–42 cycles (see below) elongation was completed for 7 min at 72°C. The following primer pairs (ordered from Thermo Fisher Scientific) and PCR conditions were used successfully: human H4R forward 5'-GAG AAC ACC AAG TTG GTC ACC A-3' and reverse 5'-TGA ATC ATG ATG TAG ATG TGC C-3' [product size 632 and 812 bp (32)] with 2.5 mM MgCl\textsubscript{2} and 42 cycles (30 s at 95°C, 1 min at 60°C, 1 min at 72°C); human H2R PrimerBank (53) primer pair ID 149158708c2 (forward 5'-CTG AGC ACT ATC TGG CCT TTC GTC-3' and reverse 5'-AGG ATG TTT ATA GGC ATG ACG A-3', product size 158 bp) with 1.5 mM MgCl\textsubscript{2} and 35 cycles (30 s at 95°C, 1 min at 54°C, 1 min at 72°C); human free fatty acid receptor 3 (FFAR3) PrimerBank primer pair ID 227430361c2 (forward 5'-GTA CAC GCT TCC AGC ACT ATC TGC TTG GTC-3' and reverse 5'-GAT CAC GCT TCC AGC ACT ATC TGC TTG GTC-3', product size 233 bp) with 1.5 mM MgCl\textsubscript{2} and eight cycles with annealing at 60°C (30 s at 95°C, 1 min at 60°C, 1 min at 72°C) plus 32 cycles at 54°C (30 s at 95°C, 1 min at 54°C, 1 min at 72°C); human TLR4 PrimerBank primer pair ID 373432602c1 (forward 5'-AGA CCT GGA GAG CAT GCA CAC AAA CTAAACCAGCCA-3', expected product size 147 bp, conditions as for human H2R); PCR products and Thermo Scientific GeneRuler DNA Ladder Mix were separated on 1% agarose and stained with ethidium bromide.

![Fig. 1. The intracellular calcium concentration ([Ca\textsuperscript{2+}]\textsubscript{i}) response to fluo 4-AM-stained P-STS cells to histamine and other substances. For pre-incubation (PI), the indicated substances were added to the cells 10 min before agonist addition. HA, histamine. *p < 0.05 and **p < 0.01. A: synergistic enhancement of cell activation by simultaneous addition of 1 μM ACh and 10 μM histamine. The sum of the fluorescence enhancement by addition of ACh or histamine alone calculated separately for each experiment [ΔF (ACh + HA)] was compared with fluorescence enhancement after simultaneous addition (4th column, n = 18). B: histamine (10 μM) also enhances the [Ca\textsuperscript{2+}]\textsubscript{i} response to ACh when added 10 min earlier (n = 9). C: the Ca\textsuperscript{2+} ionophore A-23187 (100 nM) did not enhance the [Ca\textsuperscript{2+}]\textsubscript{i} response to 10 μM histamine or 1 μM ACh (n = 12 and 6, respectively). D: histamine (10 μM), but not the histamine 4 receptor (H4R) agonist 4-methylhistamine (4-mHA, 10 μM), causes cell activation when added alone (n = 8, left). Activation by histamine is inhibited by preincubation with the histamine 1 receptor (H1R) inhibitor mepyramine (1 μM, n = 6, right). E: preincubation with the histamine 2 receptor (H2R) antagonist ranitidine (10 μM) had no significant effect on the [Ca\textsuperscript{2+}]\textsubscript{i} response to histamine (n = 13). Because cells reacted slowly in this experiment, significance was calculated from measurements taken 20 s after substance addition.

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Statistical analysis of [Ca\(^{2+}\)]\(_i\) imaging experiments. Because it was obvious that increases in fluorescence calculated from the [Ca\(^{2+}\)]\(_i\), imaging experiments were not normally distributed, statistical significance of the difference of mean values was calculated from ranked data with the unpaired two-tailed t-test assuming unequal variances (for a detailed discussion, see Ref. 44). To increase statistical reliability and strength, treatments were always conducted in parallel, resulting in equal sample numbers in each treatment group.

RESULTS

The [Ca\(^{2+}\)]\(_i\) response to ACh is synergistically enhanced by histamine while histamine alone evokes a small [Ca\(^{2+}\)]\(_i\) rise via H\(_1\)R. P-STS cells reacted with a strong increase in [Ca\(^{2+}\)]\(_i\), to histamine or the H\(_4\)R agonist 4-mHA when added together with a small amount of ACh (44), while histamine or 4-mHA alone did not evoke a significant response in these previous experiments. Here we demonstrate that ACh and histamine in fact acted synergistically in this cell activation when added simultaneously (Fig. 1A) and were equally effective when histamine was added 10 min before ACh (Fig. 1B). In our previous work (44) we suggested that the observed synergism of ACh and histamine might simply be the result of an elevation of [Ca\(^{2+}\)]\(_i\), when either compound caused an increased reaction to the second compound. A thorough investigation now showed that the reactions to histamine or ACh alone were not significantly influenced by increasing [Ca\(^{2+}\)]\(_i\), with the calcium ionophore A-23187 (Fig. 1C).

Further experiments showed that 10 μM histamine alone elicited a small rise in [Ca\(^{2+}\)]\(_i\), not observed with the H\(_1\)R agonist 4-mHA alone (Fig. 1D, left). This reaction, unnoticed in our previous investigations (44), was completely inhibited by the H\(_1\)R antagonist mepyramine (Fig. 1D, right). In our previous work (44) we showed that histamine 3 receptor (H\(_3\)R) activation had an inhibitory effect on the [Ca\(^{2+}\)]\(_i\) response of P-STS cells to epinephrine, histamine, and ACh. Epinephrine, histamine, or ACh was added to the cells at the indicated concentrations, and the increase in fluo 4-AM fluorescence was determined (n = 7). *P < 0.05 and **P < 0.01.

P-STS cells to ACh, ACh plus histamine, or histamine alone. We did not detect any effect of the histamine 2 receptor (H\(_2\)R) antagonist ranitidine on the [Ca\(^{2+}\)]\(_i\) response of P-STS cells to ACh or ACh plus histamine. Here we tested the effect of ranitidine on cell activation by histamine alone, but again without any significant effect (Fig. 1E). In addition, the [Ca\(^{2+}\)]\(_i\) response of P-STS cells to ACh and epinephrine was compared (Fig. 2). The cells reacted only weakly to a challenge with 100 μM epinephrine while showing a strong response to 1 μM ACh.

T-type VGCCs play a prominent role in the [Ca\(^{2+}\)]\(_i\) response to ACh and histamine. Experiments conducted in medium prepared without Ca\(^{2+}\) indicated a contribution of external Ca\(^{2+}\) to the [Ca\(^{2+}\)]\(_i\) response to ACh or histamine (Fig. 3). To define the pathway of Ca\(^{2+}\) influx, several inhibitors of VGCCs were tested for their ability to reduce cell activation by ACh (Fig. 4A) and ACh plus histamine (Fig. 4B). Inhibitors of L-, N-, P-, and R-type VGCCs (nifedipine [5 μM, IC\(_{50}\) ~0.3 μM for L-type channels (50)], ω-conotoxin GVIA, ω-agatoxin IVA, and SNX-482, respectively) had no significant effect, whereas the T-type channel inhibitor mibebradil (5 μM, IC\(_{50}\) ~0.3 μM for T-type channels (54)) effectively reduced the increase in [Ca\(^{2+}\)]\(_i\) in response to ACh and ACh plus histamine. Likewise, mibebradil inhibited the [Ca\(^{2+}\)]\(_i\) increase...
induced by histamine alone (Fig. 4C), suggesting an important role for T-type channels with all tested agonists without involvement of L-type channels. The participation of P-, N-, and R-type channels in the response to the employed agonists cannot be completely excluded, since the inhibitor concentrations used were rather low to avoid unspecific inhibition. Similar inhibitor concentrations were, however, inhibitory in other systems (4, 42, 51).

An even better inhibitor than mibefradil was the antispasmodic otilonium bromide, an inhibitor of T- and L-type channels (54), but also of muscarinic ACh receptors (35). It already strongly inhibited the [Ca\(^{2+}\)]_{i} response to ACh plus histamine at a concentration of 1 \(\mu\)M. In accordance with its ability to block T-type channels at similar concentrations as mibefradil (54), otilonium bromide (5 \(\mu\)M) also inhibited the response to histamine alone (Fig. 4D).

[Ca\(^{2+}\)]_{i}, response to agonists of TRPA1, TRPV1 and TRPV4. A role in visceral nociception has been observed for TRPV1, TRPA1, and TRPV4 (5). Upon stimulation, these channels mediate influx of Ca\(^{2+}\) and other cations. We tested the effect of the TRPV1 agonist capsaicin from hot peppers, the TRPA1 agonist cinnamaldehyde from the spice cinnamon, and the TRPV4 agonist GSK-1016790A on P-STS cells in the absence (Fig. 5A) and presence (Fig. 5B) of ACh, all rendering no significant effects on [Ca\(^{2+}\)]_{i}. Piperine, an agonist of TRPV1 and TRPA1 and a constituent in pepper, was also without effect (Fig. 5A). Visceral hypersensitivity induced by an agonist of TRPV4 was previously found to be potentiated by...
histamine in vivo and in isolated sensory neurons (14). Likewise, a prominent role of TRPV4 has been seen in cell activation by histamine in human keratinocytes (15), as has a facilitation of GSK-1016790A-induced signaling by TRPV1 in HEK 293 cells expressing TRPV1 and TRPV4 (32). This led us to investigate whether preincubation with different combinations of histamine, capsaicin, and GSK-1016790A had an effect on cell activation by small amounts of ACh (Fig. 5C), but no significant enhancement was detected. Notably, in this experiment the histamine concentration was only 2 μM, a concentration that does not always show a clear enhancing effect on the ACh-induced [Ca2+]i response (also see Ref. 44). This histamine concentration was chosen to allow for further enhancements of [Ca2+]i by coadded substances.

[Ca2+]i, response to selected nutrient components and degradation products. More important dietary constituents like glutamate, butyrate (a fiber degradation product produced in the intestine), and ATI were tested for their effect on [Ca2+]i in PSTS cells. None of these compounds caused cell activation when added alone (Fig. 6, A and C). The increase in fluorescence upon addition of ATI apparently was because of autofluorescence of ATI, as shown by comparison with ATI added to Hoechst dye 33342-stained fixed cells instead of live fluorescent 4-AM-stained cells using the same experimental procedure (Fig. 6C), and by the fact that, after the initial increase in fluorescence resulting from ATI addition (ATI autofluorescence), there was no further change in fluorescence (Fig. 6E). Because ATI have been shown to activate TRL4, the reaction of P-STS cells to the TRL4 ligand LPS was tested for comparison (Fig. 6C). Even in the presence of 1% heat-inactivated fetal calf serum, which effectively supplemented components of the TRL4 signaling pathway in intestinal epithelial cells (13), [Ca2+]i showed no response to LPS (Fig. 6D). Glutamate and butyrate were also added together with ACh (Fig. 6A). Although glutamate was without significant effect, butyrate strongly enhanced the response to ACh. In this series of experiments (Fig. 6A), the reaction to 1 μM ACh was very weak, which prompted additional experiments. The enhancing effect of butyrate on the response to ACh remained significant (Fig. 6B).

Effect of long incubation with histamine, ATI, LPS, or TNF-α on [Ca2+]i responses to agonists. Because histamine, ATI, or TNF-α are all substances with a suspected role in delayed food-intolerance reactions, IBS, or inflammatory bowel disease (IBD), we further tested whether longer incubation with these substances would have an effect on the susceptibility of P-STS cells to histamine. Preincubation with histamine for 18–20 h before removal of the medium and start of the labeling procedure for [Ca2+]i imaging appeared to desensitize P-STS cells to histamine, ACh, and ACh plus histamine (Fig. 7A). Preincubation with ATI or LPS had no enhancing effect on the response to histamine or ACh plus histamine (Fig. 7, B and C). After preincubation with TNF-α, implicated in tissue inflammation in IBD, there was still no response to ATI (Fig. 7E) or LPS (Fig. 7D) but an enhanced response to histamine alone (Fig. 7D). The response to ACh plus histamine was too variable to allow any conclusion to be drawn (Fig. 7D).

P-STS cells express H1R, H4R, FFAR2, FFAR3, TRPV4, and TRL4 mRNA. Our [Ca2+]i imaging experiments indicated that P-STS cells expressed functional H1R and H4R and suggested expression of the short-chain fatty acid receptors FFAR2 or FFAR3 because the cells reacted to butyrate in the presence of

Fig. 5. Effect of transient receptor potential (TRP) channel agonists on the intracellular calcium concentration ([Ca2+]i) response of P-STS cells to ACh. ACh plus histamine, or histamine alone. For preincubation (PI) the indicated substances were added to the cells 10 min before agonist addition. HA, histamine. A: [Ca2+]i response is detectable with different TRP agonists (n = 6–8, final DMSO concentration 0.02% because of preparation of some TRP agonist stocks in DMSO). B: [Ca2+]i response is detectable with different TRP agonists added together with 1 μM ACh (n = 7, final DMSO concentration 0.02%). C: combining the transient receptor potential channel vanilloid 1 (TRPV1) agonist capsaicin and the transient receptor potential channel vanilloid 4 (TRPV4) agonist GSK-1016790A with each other or with histamine during preincubation had no discernible effect on the [Ca2+]i response to ACh. Capsaicin, 50 nM; GSK-1016790A, 100 nM; HA, 2 μM; ACh, 1 μM; n = 8.
ACh. This was confirmed by detection of the corresponding mRNAs for all of those genes by PCR after reverse transcription (Fig. 8). We also investigated the presence of TRPV4 and TLR4 mRNAs. Although the cells did not respond to the TRPV4 agonist GSK-1016790A or the TLR4 ligand LPS, the mRNAs corresponding to these receptors were expressed (Fig. 8, lanes 3 and 14). Sequences for several of the primer pairs used were from the PrimerBank project and were designed to give good results with 40 cycles at 60°C with 1.5 mM MgCl2 and an annealing time of 30 s per cycle. This was confirmed for the TLR4 primer pair ID373432602c1 and the FFAR2 primer pair ID227430361c2 (Fig. 8, lanes 3 and 5). With the primer bank primers ID209862766c1 and ID373432602c1 for FFAR3 and TRPV4, respectively, no PCR product was detectable using these conditions (Fig. 8, lanes 7 and 8). A PCR product of FFAR3 with the right size was obtained with the primer pair ID209862766c2, albeit with different cycling conditions (Fig. 8, lane 10).

P-STS cells express H1R, TRPV4, TLR4, and CD14 protein. Protein expression of H1R, TRPV4, TLR4, and CD14, another component of the LPS receptor (Fig. 9D), was verified by immunofluorescence microscopy using blocking peptides or other cell lines as positive or negative controls (Fig. 9).

NF-κB activation after treatment with LPS is undetectable in P-STS cells. Because TLR4 and CD14 protein was detected in P-STS cells but without any [Ca2+]i response to LPS, we investigated whether LPS treatment would initiate NF-κB signaling in these cells. In addition to nuclear translocation of the p65/RelA subunit of NF-κB in the nucleus after release
from its complex with cytoplasmic IκB following LPS-induced IκB phosphorylation, LPS is known to induce phosphorylation of Ser536 of p65 and migration of the phosphorylated protein in the nucleus as an early event of NF-κB activation (39, 66).

Immunofluorescence microscopy showed that there was no change in phospho-p65 staining in P-STS cells even after 270 min (Fig. 10B, top) while, in PMA-differentiated THP-1 cells, p65/RelA phosphorylated at Ser536 migrated from the cytoplasm in the nucleus within 70 min after addition of LPS (Fig. 10A, bottom). Similarly, p65 distribution remained unchanged in P-STS cells with LPS (Fig. 10B, bottom) while, in control experiments in THP-1 cells, the p65 protein was found primarily in the nucleus 21 h after LPS addition (Fig. 10A, bottom). Treatment of P-STS cells with H2O2 for 30 min as a positive control induced migration of p65/RelA in the nucleus (Fig. 10B, bottom).

DISCUSSION
Our work establishes that the previously observed (44) enhancement of the reaction of P-STS cells to ACh by histamine is not only additive but also synergistic. A more thorough investigation of the effect of Ca2+ ionophore added to either ACh or histamine alone showed that this synergistic effect

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**Fig. 7.** Effect of prolonged incubation of P-STS cells with histamine, amylase-trypsin inhibitors (ATI), lipopolysaccharide (LPS), or tumor necrosis factor-α (TNF-α) on the reactivity to agonists. For preincubation (PI) the indicated substances were added to the cells for 18–20 h, and the medium was exchanged before staining with fluo 4-AM. HA, histamine. If not indicated otherwise, ACh (1 μM), ACh (1 μM) plus histamine (2 μM), histamine (10 μM), LPS (10 μg/ml), or ATI were added to the cells, and the intracellular calcium concentration ([Ca2+]i) response was measured. A: preincubation with histamine (n = 6). B: preincubation with ATI (n = 5). C: preincubation with LPS (because of high reactivity of the cells only 0.5 μM ACh was added in these experiments, n = 8). D: preincubation with 10 ng/ml TNF-α (n = 6, except for addition of histamine with n = 15). E: time course of fluorescence increase after addition of ATI to cells preincubated with 10 ng/ml TNF-α. As deduced from Fig. 5C, the initial increase in fluorescence after addition of ATI is the result of ATI autofluorescence, but there is no further increase in fluorescence with ATI, indicating a lack of [Ca2+]i response.
The susceptibility of P-STS cells to activation by histamine cannot be explained by a mere rise in [Ca$^{2+}$]. The synergistic effect of ACh and histamine is also seen when the cells are preincubated with histamine before addition of ACh. This suggests that pulsing of ACh secreted by nerve endings and reaching EC cells located in intestinal crypts could evoke a similar response when histamine concentrations are elevated following mast cell activation. Degranulated mucosal mast cells and elevated histamine levels were detected in patients with IBD and in a subset of IBS patients with increased numbers of intestinal mast cells (reviewed in Refs. 7 and 33). Multiple observations indicate a role of mast cells in gut sensorimotor dysfunction and related diarrhea in IBD and IBS. The susceptibility of P-STS cells to activation by histamine suggests that these symptoms might be partly attributed to increased serotonin release by EC cells in response to elevated histamine secretion by mast cells.

In comparison with the strong response evoked by histamine in the presence of ACh, presumably via H$\text{H}_1$R signaling (15), the response to histamine alone appeared weak and may play a minor role in serotonin secretion. Unfortunately, the low serotonin content of P-STS cells does not allow easy detection of secretion (44). Inhibition by the H$\text{H}_1$R antagonist mepyramine, a first-generation antihistamine, showed that the [Ca$^{2+}$]i response evoked by histamine alone is attributable to H$\text{H}_1$R signaling. The H$\text{H}_1$R agonist 4-mHA alone did not cause any significant [Ca$^{2+}$]i response. Regarding the other histamine receptors, our previous work has demonstrated that the H$\text{H}_2$R agonist methemiprop had an inhibitory effect on P-STS activation by ACh, ACh plus histamine, or histamine alone (44). As shown here, the H$\text{H}_3$R inhibitor ranitidine did not have any significant effect on the [Ca$^{2+}$]i response evoked by histamine. Epinephrine, an agonist sometimes used to stimulate secretion in neuroendocrine cells, evoked only a very weak [Ca$^{2+}$]i response in P-STS cells compared with ACh. This is in accordance with the important physiological roles of ACh and serotonin as excitatory stimuli of gut motility (43) and underlines the suitability of the P-STS cell line as a model for intestinal serotonin-secreting cells.

Activation of H$\text{H}_1$R by histamine is known to mobilize intracellular Ca$^{2+}$ stores and to cause influx of extracellular Ca$^{2+}$ via voltage-dependent and -independent ion channels in excitable cells (reviewed in Ref. 38). This influx of external calcium is mainly responsible for secretion in response to histamine. The [Ca$^{2+}$]i response to histamine, as well as to ACh alone, was also augmented by external Ca$^{2+}$ in P-STS cells.

VGCC are thought to elevate Ca$^{2+}$ locally, and some of them have been shown to interact directly with the secretory machinery (see below). Additional Ca$^{2+}$ entry pathways like TRP channels may trigger or modulate exocytosis (10, 41). We investigated the participation of VGCC in the [Ca$^{2+}$]i response evoked by histamine. The [Ca$^{2+}$]i response to histamine, as well as to ACh alone, was also augmented by external Ca$^{2+}$ in P-STS cells. The [Ca$^{2+}$]i response for ACh and ACh plus histamine using selective inhibitors for L-, N-, P-, R-, and T-type VGCC. Only the T-type channel inhibitor mibefradil showed a significant inhibitory effect. Mibefradil inhibited the [Ca$^{2+}$]i response for histamine alone and for ACh and ACh plus histamine. In contrast to the other VGCC channels mentioned, T-type VGCC are low-voltage-activated channels that ensure secretion at membrane potentials near the resting conditions important for basal hormone release or sustained release during mild stimulations (reviewed in Refs. 12 and 63). There is ample evidence for their participation in neurotransmitter release in neuroendocrine cells, although they lack the consensus synaptic protein interaction (synprint) site allowing N- and P/Q-type VGCC to bind proteins of the secretion machinery. An increased density of T-type VGCC is seen as a response to stressors in many cells and tissues, presumably lowering the action potential firing thresholds.

**Fig. 8.** P-STS cells express histamine 1 receptor (H$\text{H}_1$R), histamine 4 receptor (H$\text{H}_4$R), free fatty acid receptor 2 (FFAR2), free fatty acid receptor 2 (FFAR3), and transient receptor potential channel vanilloid 4 (TRPV4) mRNA. Detection of mRNAs by reverse transcription-polymerase chain reaction (PCR). Lanes 7 and 8, no PCR fragment detectable with the primer bank primers ID20986276c1 and ID33342602c1 for FFAR3 and TRPV4. Lanes 12 and 13, no PCR fragment of TRPV4 detectable with 1.5 and 2.0 mM MgCl$_2$ instead of 2.5 mM MgCl$_2$ as used in lane 14. TLR4, Toll-like receptor 4.
threshold (12). Thus, the importance of T-type VGCC for histamine-induced and ACh-induced cell activation in P-STS cells may be a typical feature of human EC cells. T-type channel blockers have been suggested as promising candidates for novel analgesics potentially useful for treating visceral pain and discomfort (3). Otilonium bromide, a spasmolytic with significant therapeutic effect in IBS and minimal systemic absorption from the intestine (3, 11, 16, 59), is known to inhibit T- and L-type VGCC and muscarinic ACh receptors (27, 31). Accordingly, otilonium bromide effectively inhibited the synergistic enhancement of the \([\text{Ca}^{2+}]_i\) response to ACh by histamine and the response to histamine alone, confirming the results with mibefradil. This suggests that otilonium bromide might be useful not only in IBS but also in histamine intolerance and food allergy by obviating excessive dietary restrictions that might have detrimental effects.

A role in visceral nociception and hypersensitivity, an important clinical problem for the patients, has been suggested for TRPA1, TRPV1, and TRPV4 (5, 37). Accordingly, agonists of these cation channels, including the food constituents capsaicin, cinnamaldehyde, and piperine, were tested for their effect on the \([\text{Ca}^{2+}]_i\) response of P-STS cells in the absence or presence of ACh, histamine, or both agonists. These experiments gave no indication of the presence of functional TRPA1, TRPV1, or TRPV4. Because there is evidence suggesting that TRPV4 expression is increased in the intestine of IBS patients and that in these patients elevated levels of histamine and serotonin are implicated in the development of visceral hypersensitivity (reviewed in Refs. 23 and 64), we looked for expression of TRPV4 in P-STS cells. We detected both TRPV4 mRNA and protein. Further research will be necessary to clarify why the cells do not react to the TRPV4 agonist...
GSK-1016790A. Possible explanations include low TRPV4 localization at the cell surface or expression of nonfunctional TRPV4. In this respect, it is interesting that, in primary human endothelial cells, most TRPV4 channels remained silent even during maximal stimulation with GSK-1016790A (56).

There was a weak [Ca\(^{2+}\)]\(i\) response by butyrate when added together with ACh. Induction of serotonin secretion by butyrate would be consistent with the necessity for increased gut motility when microbial activity is high to pursue digestion and prevent bacterial overgrowth in the small intestine (52). Short-chain fatty acid concentrations of 70–140 mM have been observed in the proximal colon (65), and about one-tenth of this concentration was found in human ileum (17). Short-chain fatty acids are agonists of the free fatty acid receptors FFAR2/G protein-coupled receptor 43 and FFAR3/G protein-coupled receptor 41 and induced a [Ca\(^{2+}\)]\(i\) response when transfected in Chinese hamster ovary cells (34). They are involved in hormone secretion by intestinal neuroendocrine cells (8). Butyrate and FFAR2 have been implicated in serotonin secretion by neuroendocrine cells (2, 47). We show that P-STS cells express mRNA of both FFAR2 and FFAR3.

Upon addition of glutamate, glutamate plus ACh, or ATI, there was no [Ca\(^{2+}\)]\(i\) response in P-STS cells. Because the IL-8 response to ATI in human monocyte-derived dendritic cells depends heavily on TLR4 and CD14 (28), we tested the response of P-STS cells to LPS, another ligand of the TLR4-MD-2-CD14 complex. Despite the presence of TLR4 and CD14 protein confirmed by immunofluorescence microscopy, the cells showed neither a [Ca\(^{2+}\)]\(i\) response to LPS nor discernible migration of NF-κB p65/RelA or Ser\(^{336}\)-phosphorylated p65/RelA to the nucleus. Whereas preincubation with TNF-α simulating inflammatory conditions considerably increased the [Ca\(^{2+}\)]\(i\) response to histamine alone, long-term incubation with ATI or LPS had no such effect. No protein-altering insertions/deletions or rare single nucleotide polymorphisms have been found in the P-STS TLR4 sequence (27). It is unknown, however, whether P-STS cells express MD-2, which was critical for LPS responses in intestinal epithelial cell lines and a CHO cell line (1, 49). In accordance with our results, in a previous study, LPS evoked only an insignificant release of serotonin in human EC cells isolated from normal mucosa, whereas in EC cells from Crohn’s colitis mucosa, there was an increased serotonin secretion in response to LPS (31). Control of signaling to commensal microorganisms is extremely important in the intestinal epithelium. Low expression of Toll-like receptors and accessory proteins may be a means of avoiding hyperresponsiveness (61) (reviewed in Refs. 40 and 58). Additional control mechanisms of TLR signaling include location of TLRs in intracellular compartments or at the basolateral membrane and negative regulation of signaling by inhibitory factors. An investigation into the complexity of this regulatory network in P-STS cells would exceed the scope of this work.

Recent evidence indicates that LPS can directly activate the cation channels TRPA1 and TRPV4 and induce an increase in [Ca\(^{2+}\)]\(i\). (9). Interaction of LPS with TRPV4 has been shown to sensitize TRPV1 to capsaicin (37). These properties of LPS might be the physiological reason for the lack of functional TRPA1, TRPV1, and TRPV4 in P-STS cells, since constant exposition to LPS of cells exposed to the gut lumen otherwise might activate these channels.

In conclusion, in this work, we show work that, in addition to H\(_2\)R and H\(_4\)R, P-STS ileal EC cells express the H\(_3\)R and are also sensitive to histamine in the absence of ACh. Histamine and ACh together cause a synergistic increase in [Ca\(^{2+}\)]\(i\) that is inhibited by the spasmolytic oltionium bromide, a drug ameliorating symptoms in IBS. Oltionium bromide also inhibits cell activation by histamine alone. These results suggest that it might be therapeutically useful in histamine intolerance and food allergy. Our data indicate that the T-type VGCC participates in the increase in [Ca\(^{2+}\)]\(i\) evoked by histamine, which is a requirement for serotonin secretion. Pretreatment with TNF-α considerably increased the [Ca\(^{2+}\)]\(i\) response to histamine alone, suggesting that patients with IBD or IBS with a tendency toward inflammation might have increased susceptibility to histamine and activation of mast cells. The cells showed no response to wheat amylase-trypsin inhibitors, suggesting that EC cells are not directly involved in nongluten wheat sensitivity.

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No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS
B.P. and E.J.-J. conceived and designed research; B.P. and V.F.Z. performed experiments; B.P. analyzed data; B.P., V.F.Z., D.S., and E.J.-J. interpreted results of experiments; B.P. prepared figures; B.P. drafted manuscript; B.P., V.F.Z., D.S., R.P., and E.J.-J. approved final version of manuscript; V.F.Z., D.S., R.P., and E.J.-J. edited and revised manuscript.

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