Overexpression of lncRNA SNGH3 Predicts Unfavorable Prognosis and Clinical Outcomes in Human Cancers: Evidence from a Meta-Analysis

Yaofei Jiang1,2 and Lulu Le1

1The Second Affiliated Hospital of Nanchang University, Nanchang, Jiangxi Province 330000, China
2Department of Radiation and Medical Oncology, Zhongnan Hospital, Wuhan University, Wuhan, Hubei Province 430000, China

Correspondence should be addressed to Lulu Le; ndefy13032@ncu.edu.cn

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1. Introduction

Nowadays, cancer is the most prevalent cause of death and continues to be a serious public health problem with increasing morbidity and mortality [1, 2]. It was reported that about 9.6 million people died of cancers and more than 18.1 million people diagnosed with cancers in 2018 worldwide [3]. Although clinical treatment including surgery, radiotherapy, and chemotherapy, as well as targeted therapy and immunotherapy improved dramatically in the last century, patients’ 5-year survival rate is still low and remains to be enhanced due to malignant progression and deficiencies in early diagnosis and treatment target [4–6]. On account of early diagnosis and treatment that can greatly improve cancer patients’ survival, it is urgent to discover new biomarkers for cancer patients [7].

Long noncoding RNAs (lncRNAs) are mRNA-like transcripts, which consist of more than 200 nucleotides and lack protein-coding ability [8, 9]. Considered genomic “noise” initially, lncRNAs get increasing attention due to their newly found role in various diseases and cellular activity nowadays [10, 11]. Especially in tumor, more and more studies confirm that lncRNAs involve in tumorigenesis and metastasis through various mechanisms, such as sponging miRNAs,
213 of records identified through Pubmed, Web of Science, Cochrane Library, Embase and MEDLINE searching

Additional records identified through other sources (n = 0)

Records after duplicates removed (n = 57)

Records screened (n = 31)

Records excluded (n = 26)

Full-text articles assessed for eligibility (n = 18)

Full-text articles excluded, for no usable data (n = 13)

Studies included in qualitative synthesis (n = 13)

Studies included in quantitative synthesis (meta-analysis) (n = 13)

Figure 1: Flow diagram of the literature retrieval and selection.

Table 1: The main information of enrolled studies in the meta-analysis.

| Study | Year | Country | Tumor types | Sample size | Sample | Detection method | Cutoff value | Follow-up (year) | Outcome measures | Analysis type | NOS |
|-------|------|---------|-------------|-------------|--------|-----------------|--------------|-----------------|-----------------|--------------|-----|
| Chen  | 2019 | China   | Osteosarcoma| 127         | Tissues | qRT-PCR         | ROC curve    | 16 years        | OS CP           | Multivariate  | 9   |
| Fei    | 2018 | China   | Glioma      | 60          | Tissues | qRT-PCR         | Mean         | 5 years         | OS CP           | Univariate    | 8   |
| Hong   | 2018 | China   | Ovarian cancer | 76        | Tissues | qRT-PCR         | Median       | 6 years         | OS CP           | Multivariate  | 9   |
| Ma     | 2020 | China   | Breast cancer | 60         | Tissues | qRT-PCR         | Mean         | NA              | CP              | NA           | 7   |
| Peng   | 2020 | China   | Acute myeloid leukemia | 62    | Bone marrow | qRT-PCR     | Median       | 5 years         | OS CP           | Univariate    | 8   |
| Shi    | 2020 | China   | Non-small cell lung cancer | 32    | Tissues | qRT-PCR         | Median       | 5 years         | OS CP           | Univariate    | 9   |
| Sui    | 2020 | China   | Papillary thyroid carcinoma | 42    | Tissues | qRT-PCR         | Mean         | NA              | CP              | NA           | 8   |
| Tian   | 2019 | China   | Intrathelial cholangiocarcinoma | 52    | Tissues | qRT-PCR         | NA           | 5 years         | OS CP           | Multivariate  | 8   |
| Xuan   | 2019 | China   | Gastric cancer | 60        | Tissues | qRT-PCR         | Median       | 6 years         | OS MFS          | Univariate    | 8   |
| Zahra  | 2019 | Iran    | Breast cancer | 80        | Tissues | qRT-PCR         | Median       | NA              | CP              | NA           | 7   |
| Zhang  | 2016 | China   | Hepatocellular carcinoma | 144   | Tissues | qRT-PCR         | NA           | 5 years         | OS RFS DFS CP  | Multivariate  | 8   |
| Zhang  | 2018 | China   | Hepatocellular carcinoma | 70    | Tissues | qRT-PCR         | NA           | 2 years         | OS CP           | Univariate    | 8   |
| Zheng  | 2019 | China   | Osteosarcoma | 54         | Tissues | qRT-PCR         | Median       | 10 years        | OS              | Univariate    | 9   |
epigenetic regulation, translation regulation, cell differentiation regulation, and therapy resistance [12–14]. The expression and role of different lncRNAs may vary greatly. While some lncRNAs express as tumor suppressor genes, like GAS5 and ANRIL, more lncRNAs functioned as oncogenes, such as NKILA and FAL1 [15–18]. Notably, these lncRNAs are expected to be biomarkers and therapeutic targets for cancers thereby.

Small nucleolar RNA host gene 3 (SNHG3) is a newly identified lncRNA with abnormal expression in various cancers. Previous studies reported that SNHG3 is upregulated in cancers and may act as an oncogene in tumor prognosis, including osteosarcoma [19], glioma [20], ovarian cancers [21], breast cancer [22], and hepatocellular carcinoma [23]. Overexpression of SNHG3 in these was usually associated with poor prognosis and clinical features, like advanced clinical stage, earlier distant metastasis, and tumor size. Furthermore, SNHG3 involved in the pathologic process of tumor, including cell proliferation, migration, EMT, and apoptosis [24]. Collectively, SNHG3 may be a promising biomarker for diagnosis and target for therapy in various cancers. However, the results remain to be confirmed due to contentious outcomes and small sample size in individual studies. Thus, we performed this meta-analysis and review, for the first time, to identify the clinical role of SNHG3 in human cancers and explore its functions.

2. Materials and Methods

The present meta-analysis was constructed and reported according to the PRISMA checklist [25].
### (a) Multivariate

| Study ID | HR (95% CI) | Weight |
|----------|-------------|--------|
| Chen, 2019 | 4.61 (2.10, 10.13) | 11.55 |
| Hong, 2018 | 2.26 (1.05, 4.86) | 12.19 |
| Tian, 2019 | 2.82 (1.24, 6.41) | 10.60 |
| Zhang, 2016 | 3.46 (1.82, 6.58) | 17.28 |
| Subtotal ($I^2 = 0.0\%, P = 0.622$) | 3.20 (2.20, 4.64) | 51.62 |

### (b) Univariate

| Study ID | HR (95% CI) | Weight |
|----------|-------------|--------|
| Fei, 2018 | 1.15 (0.29, 4.55) | 3.77 |
| Peng, 2020 | 1.92 (0.50, 7.37) | 3.95 |
| Shi, 2020 | 2.24 (0.68, 7.38) | 5.03 |
| Xuan, 2019 | 2.11 (0.91, 4.89) | 10.14 |
| Zhang, 2018 | 2.05 (1.13, 3.73) | 19.97 |
| Zheng, 2019 | 1.99 (0.64, 6.21) | 5.52 |
| Subtotal ($I^2 = 0.0\%, P = 0.984$) | 1.97 (1.34, 2.90) | 48.38 |

### Heterogeneity between groups: $P = 0.046$

| Overall ($I^2 = 0.0\%, P = 0.781$) | 2.53 (1.94, 3.31) | 100.00 |

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**Figure 3: Continued.**
2.1. Information Source and Searching Strategy. We searched many databases, including PubMed, Web of Science, Cochrane Library, Embase, and MEDLINE, by retrieved keywords in them from March 1, 2020, to March 5, 2020, to collect all eligible studies for this meta-analysis. We used the following keywords and MeSH terms: ("small nucleolar RNA host gene 3" or "SNHG3") and ("cancer" or "carcinoma" or "neoplasm" or "tumor") and ("prognosis" or "clinical outcome" or "pathological feature" or "survival"). The search syntax is shown in Table S1. Additional records
were identified through searching for references of these retrieval studies.

2.2. Eligibility Criteria. Studies meeting the following criteria were enrolled in this meta-analysis: (a) reported the expression of SNHG3 in tumor tissues; (b) valuable data in regard to association between SNHG3 expression and clinical parameters; (c) providing hazard ratio (HR) with 95% confidence intervals (CI) for survival outcomes directly; (d) sufficient data to calculate the HRs and CIs for survival outcomes; and (e) written in English. Studies meeting one or more of the following criteria were excluded: (a) duplicate publications; (b) based on other diseases but not cancers; (c) animal studies; (d) not available date for survival outcomes and clinical parameters; and (e) letters, case reports, expert opinions, and reviews.

2.3. Data Extraction and Quality Valuation. Two authors (YJ and LL) recorded data and corresponding basic information from included studies and evaluated the quality of them independently, including surname of first author, publication year, country of the study carried out at, cancer type, sample size, specimen, detection method, cutoff value, follow-up time, and analysis method. Any disagreements were resolved by a doctor who is professional in this area. Meanwhile, clinical parameters including age, gender, clinical stage, tumor size, lymph node metastasis, distant metastasis, and histological grade were also extracted. For patients’ survival outcomes, including overall survival (OS), metastasis-free survival (MFS), disease-free survival (DFS), and recurrence-free survival (RFS), we recorded HRs and 95% CIs from studies provided to them directly or using Engauge Digitizer version 4.1 to extract the data from those studies that provided Kaplan-Meier curves only indirectly. The Newcastle-Ottawa Scale (NOS) was used to evaluate the quality of included studies, and studies with NOS score 7-9 were considered of high quality [26].

2.4. Validation by Using Data from TCGA Dataset. Gene Expression Profiling Interactive Analysis (GEPIA) is a newly developed interactive web aiming at analyzing the RNA sequencing expression data from The Cancer Genome Atlas (TCGA) dataset [27]. It was used to explore the SNHG3 expression level in tumor and normal tissues in different kinds of cancers. Kaplan-Meier method and logrank test were applied to calculate the survival analysis to validate the association between SNHG3 expression and OS and DFS.

2.5. Statistical Analysis. STATA software (version 12.0; StataCorp LLC, College Station, Texas) was applied to analyze the HRs and 95% CIs for survival outcomes and ORs and 95% CIs for clinical parameters. Chi squared-based Q test and I² statistics were performed to identify the heterogeneity of the included articles. Fixed-effect model was used for analysis, but if I² > 50% or P value < 0.05, random effect was applied due to significant heterogeneity [28]. Sensitivity analysis was also done by removing one study from the included studies to testify the stability of results for OS. Egger’s funnel regression test and Begg’s funnel plot were performed to evaluate the publication, determined as positive by Pr > |z| ≤ 0.05.

3. Results

3.1. Literature Search. A total of 213 articles were identified after our preliminary search. Subsequently, 57 studies were left for further inspection after removing duplicate articles. Then, we screened their titles, abstracts, or full texts, and 13 studies were finally included in this meta-analysis [19–23, 29–36]. The procedure of study selection is illustrated in Figure 1.

3.2. Characteristics of Included Studies. The main characteristics of the 13 included studies are demonstrated in Table 1. These studies were published between 2016 and 2020, and except one study carried out in Iran, the others were carried out in China. With a total of 919 patients included, the sample size of studies ranged from 32 to 144. These studies reported various cancers, including osteosarcoma (two articles), glioma, ovarian cancer, breast cancer (two articles), acute myeloid leukemia, non-small-cell lung cancer, papillary thyroid carcinoma, gastric cancer, intrahepatic cholangiocarcinoma, and hepatocellular carcinoma (two articles). The level of SNHG3 was quantified with real-time polymerase chain reaction (qRT-PCR). One study measured bone marrow of patients and healthy participants, and the others measured the cancer and matched normal tissues. There were four different cutoff values for dividing patients into high- and low-expression group: five using median, three using mean, one using ROC curve, and three not available. Among these studies, nine studies reported OS, ten studies reported clinical parameters, and one study reported MFS, RFS, and DFS, respectively. With their NOS scores ≥ 7, all included studies showed high quality.

Table 3: Association between overexpression lncRNA SNHG3 and clinical parameters.

| Clinicopathologic parameters | No. of studies | No. of patients | OR  | 95% CL | I² (%) | P      | Model |
|-----------------------------|----------------|----------------|-----|--------|--------|--------|-------|
| Age                         | 9              | 655            | 1.03| 0.74-1.44| 0.0   | 0.545 | Fixed |
| Gender                      | 8              | 589            | 1.06| 0.92-1.21| 0.0   | 0.686 | Fixed |
| Clinical stage              | 8              | 556            | 3.25| 2.23-4.73| 15.9  | 0.305 | Fixed |
| Tumor size                  | 7              | 553            | 1.86| 0.71-4.87| 76.7  | 0.0   | Fixed |
| Lymph node metastasis       | 5              | 262            | 8.96| 4.82-16.68| 19.6  | 0.29  | Fixed |
| Distant metastasis          | 3              | 323            | 2.18| 1.05-4.52| 31.4  | 0.233 | Fixed |
| Histological grade          | 4              | 472            | 2.23| 1.47-3.37| 45.4  | 0.103 | Fixed |

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Overall ($I^2 = 15.9\%, P = 0.305$)

| Study ID  | OR (95% CI) | Weight % |
|-----------|-------------|----------|
| Zahra, 2019 | 3.75 (1.42, 9.91) | 13.39 |
| Ma, 2020    | 5.90 (1.85, 18.78) | 7.64 |
| Shi, 2020   | 9.00 (1.82, 44.59) | 3.18 |
| Sui, 2020   | 15.86 (0.84, 298.86) | 1.27 |
| Tian, 2019  | 5.08 (1.55, 16.64) | 7.71 |
| Zhang, 2016 | 3.47 (1.19, 10.13) | 12.22 |
| Zhang, 2018 | 2.00 (0.95, 4.18) | 32.78 |
| Overall ($I^2 = 15.9\%, P = 0.305$) | 3.25 (2.23, 4.73) | 100.00 |

(a)

Overall ($I^2 = 45.4\%, P = 0.103$)

| Study ID  | OR (95% CI) | Weight % |
|-----------|-------------|----------|
| Fei, 2018 | 7.48 (2.35, 23.81) | 6.92 |
| Hong, 2018 | 1.20 (0.48, 2.99) | 27.36 |
| Ma, 2020    | 3.76 (1.24, 11.38) | 10.61 |
| Tian, 2019  | 1.88 (0.62, 5.65) | 15.00 |
| Zahra, 2019 | 3.28 (0.97, 11.09) | 10.47 |
| Zhang, 2016 | 1.21 (0.50, 2.92) | 29.65 |
| Overall ($I^2 = 45.4\%, P = 0.103$) | 2.23 (1.47, 3.37) | 100.00 |

(b)

Overall ($I^2 = 31.4\%, P = 0.233$)

| Study ID  | OR (95% CI) | Weight % |
|-----------|-------------|----------|
| Chen, 2019 | 1.46 (0.51, 4.16) | 56.69 |
| Tian, 2019 | 5.87 (1.41, 24.40) | 18.07 |
| Zhang, 2016 | 1.17 (0.21, 6.63) | 25.24 |
| Overall ($I^2 = 31.4\%, P = 0.233$) | 2.18 (1.05, 4.52) | 100.00 |

(c)

Figure 4: Continued.
3.3. Association between IncRNA SNHG3 Expression and OS. Ten studies reported the relationship between SNHG3 expression and OS, with a total of 737 cases. Fix-effect model was used to analyze the HR and 95% CI of OS due to absence of apparent heterogeneity among included studies ($I^2 = 0.0$, $P = 0.781$). As shown in Figure 2, the pooled result demonstrated that high expression of SNHG3 was significantly related to poor OS in cancers ($HR = 2.53$, 95% CI: 1.94-3.31).

To further investigate the relationship between SNHG3 expression and OS, we performed subgroup meta-analysis stratified by analysis method (multivariate and univariate analysis), sample size (more or less than 100), follow-up time (more or less than 5 years), and cancer type (gastrointestinal cancer or others). All subgroup analyses showed similar results that high expression of SNHG3 was significantly associated with worse OS in various cancers (Table 2, Figure 3). All results above demonstrated that SNHG3 could be a prognostic factor for cancer patients’ OS.

3.4. Association between IncRNA SNHG3 Expression and MFS, DFS, and RFS. Only one study provided suitable data for MFS, DFS, and RFS, respectively. High expression of SNHG3 was significantly correlated to both unfavorable DFS (HR = 3.89, 95% CI: 1.34-11.3), and RFS (HR = 2.42, 95% CI: 1.14-5.15). But it could not predict worse MFS (HR = 1.39, 95% CI: 0.51-3.84). These results indicated that SNHG3 could prompt the prognosis of patients with cancer.

3.5. Association between IncRNA SNHG3 and Clinical Parameters. Ten studies provided available data for analyses between SNHG3 expression and clinical parameters, including age, gender, clinical stage, tumor size, lymph node metastasis, distant metastasis, and differentiation. ORs and its 95% CIs were adopted for analyses. The results demonstrated that high expression of IncRNA SNHG3 was correlated with later clinical stage (OR = 3.25, 95% CI: 2.23-4.73), higher histological grade (OR = 2.23, 95% CI: 1.47-3.37), distant metastasis (OR = 2.18, 95% CI: 1.05-4.52), and earlier lymph node metastasis (OR = 8.96, 95% CI: 4.82-16.68) (Table 3, Figure 4). Nevertheless, no statistical significance was detected in age (OR = 1.03, 95% CI: 0.74-1.44), gender (OR = 1.06, 95% CI: 0.92-1.21), and tumor size (OR = 1.86, 95% CI: 0.71-4.87) (Table 3).

3.6. Sensitivity Analysis. Then, we executed sensitivity analysis to check the stability of the results of the correlation between SNHG3 expression and OS by removing each study. As shown in Figure 5(a), it was reliable that high expression of SNHG3 was associated with worse OS.

3.7. Publication Bias. In addition, we conducted Egger’s funnel regression test and Begg’s funnel plot to test the publication bias for meta-analysis of the association between SNHG3 expression and OS. There was no significant publication bias based on both Begg’s funnel plot (P = 0.371, Figure 5(b)) and Egger’s funnel regression test (P = 0.22, Figure 5(c)), indicating that our pooled result was credible.

3.8. Validation of the Role of IncRNA SNHG3 in Human Cancers: Based on TCGA Dataset. To further validate the prognostic and clinical value of SNHG3 expression in human cancers, we investigated TCGA dataset. As shown in Figures 6(a) and 6(b), SNHG3 was significantly overexpressed in tumor tissue comparing to normal tissue in various cancers, including bladder urothelial carcinoma, cholangiocarcinoma, colon adenocarcinoma, esophageal carcinoma, liver hepatocellular, lung adenocarcinoma, lung squamous cell carcinoma, prostate adenocarcinoma, rectum adenocarcinoma, stomach adenocarcinoma, and uterine corpus ($P < 0.01$). Moreover, the violin plot demonstrated that SNHG3 expression was also significantly associated with human cancipers’ clinical stage ($P < 0.01$, Figure 6(c)). In addition, the patients were divided into high- and low-SNHG3-expression group according to the median value of its expression. Survival plot was implemented for OS and
Figure 5: (a) Sensitivity analysis of pooled HRs for OS. (b) Begg’s funnel plot of SNHG3 for OS. (c) Egger’s funnel plot of SNHG3 for OS.
Expression − log2 (TPM + 1)

(a)

(b)

(c)

Stage 0 Stage I Stage II Stage III Stage IV Stage X

Figure 6: Continued.
The results have shown that high expression of SNHG3 was significantly associated with worse OS (HR = 1.22, P < 0.01; Figure 6(d)) and DFS (HR = 1.1, P = 0.0028; Figure 6(e)), which ulteriorly confirmed the results of this meta-analysis.

4. Discussion

With the development of second-generation sequencing technology in recent years, studies have shown that IncRNA plays an important role in many biological fields such as tumor development, neuroscience, and ontogeny and is an important regulatory molecule in human genome [37, 38]. Increasing studies demonstrate that IncRNAs are dysregulated in cancers and play an important role in tumor development and progression [39, 40]. For example, IncRNA MALAT1 is upregulated in various cancers. High expression of MALAT1 is associated with poor prognosis of cancer patients, and closely correlated with tumor proliferation, autophagy, and drug resistance [41–43].

Studies have demonstrated that SNHG3 is overexpressed in various cancers and considered to function as a novel oncogene in tumor development. Compared to matched normal tissue, SNHG3 is upregulated in tumor tissues, such as osteosarcoma [19, 36], glioma [20], ovarian cancer [21], breast cancer [22, 34], hepatocellular carcinoma [23, 24, 35], acute myeloid leukemia [29], non-small-cell lung cancer [30], papillary thyroid carcinoma [31], intrahepatic cholangiocarcinoma [32], and gastric cancer [33], and usually brings poor clinical outcomes, including OS, DFS, RFS, stage, grade, tumor size, distant metastasis, and lymph node metastasis. What is more, it was found that the level of SNHG3 is significantly associated with epidermal growth factor receptor 2 (Her-2) and estrogen receptor (ER) status in breast cancer [22, 34]. In acute myeloid leukemia, SNHG3 expression was found to have strong association with platelet count and white blood cell count [29].

In our meta-analysis, we explored the relationship between SNHG3 expression and patients’ prognosis and other clinical parameters. The results demonstrated that overexpressed SNHG3 was significantly associated with poor OS in various cancers (HR = 2.53, 95% CI: 1.94-3.31) and poor DFS (HR = 3.89, 95% CI: 1.34-11.3) and RFS (HR = 2.42, 95% CI: 1.14-5.15) in hepatocellular carcinoma. Additionally, patients with high SNHG3 expression were more prone to have more advanced clinical stage, higher histological grade, earlier distant metastasis, and earlier lymph node metastasis. In brief, SNHG3 could be a promising biomarker for prognosis in pancancers.

Besides, SNHG3 affects tumorigenesis and prognosis of cancers by participating in various biological process, including promoting cell proliferation, tumor migration and invasion, cell cycle progression, inhibiting apoptosis, and drug resistance [23, 24, 44–47]. Furthermore, many mechanism
studies demonstrated that SNHG3 can regulate target genes by miRNA sponge, including miRNA-326 and miRNA-128 in hepatocellular carcinoma [23, 24], miRNA-139-5p in clear cell renal cell carcinoma [44], miRNA-384 in laryngeal carcinoma [45], miRNA-151a-3p and miRNA-196a-5p in osteosarcoma [19, 39], miRNA-758-3p in acute myeloid leukemia [29], and miRNA-182-5p in colorectal cancer [47]. In addition, it could interact with the TGF-β and JAK2/STAT3 pathway in lung cancer [30]. Taken together, SNHG3 involved in many aspects of tumor development and it may serve as a promising therapy target.

There are some limits that exist in this meta-analysis. Firstly, most of the enrolled studies were carried out in China with a relatively small sample, which makes the results more suitable in the Chinese population. Secondly, the cutoff values of SNHG3 expression varied from each study and the actual values could not be obtained, which might bring publication bias. Thirdly, the HR value used for analysis was extracted from the K-M curve in some articles, which inevitably brings some bias. Finally, there may exist small values of SNHG3 expression varied from each study and the actual values could not be obtained, which might bring publication bias. Thirdly, the HR value used for analysis was extracted from the K-M curve in some articles, which inevitably brings some bias. Finally, there may exist small publication bias.

5. Conclusion

This study demonstrated that overexpressed SNHG3 was significantly associated with poor survival, including OS, DFS, and RFS and worse clinical outcomes, including clinical stage, histological grade, distant metastasis, and lymph node metastasis in human cancers. Therefore, SNHG3 can serve as a promising biomarker for predicting patients’ prognosis.

Data Availability

The data supporting the conclusions of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors confirm that there are no conflicts of interest.

Authors’ Contributions

Yaofei Jiang and Lulu Le were responsible for bibliographic search and extraction of data. Yaofei Jiang was responsible for the analysis and interpretation of the data and drafting of the manuscript. Yaofei Jiang and Lulu Le were responsible for the revision of the article.

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Supplementary Materials

Table S1: the search syntax is shown.

References

[1] R. L. Siegel, K. D. Miller, and A. Jemal, "Cancer statistics, 2019," CA: A Cancer Journal for Clinicians, vol. 69, no. 1, pp. 7–34, 2018.
[2] K. D. Miller, L. Nogueira, A. B. Mariotto et al., "Cancer treatment and survivorship statistics, 2019," CA: a Cancer Journal for Clinicians, vol. 69, no. 5, pp. 363–385, 2019.
[3] F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre, and A. Jemal, "Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries," CA: A Cancer Journal for Clinicians, vol. 68, no. 6, pp. 394–424, 2018.
[4] A. A. Wright, N. L. Keating, J. Z. Ayanian et al., "Family perspectives on aggressive cancer care near the end of life," JAMA, vol. 315, no. 3, pp. 284–292, 2016.
[5] S. L. Topalian, J. M. Taube, and D. M. Pardoll, "Neoadjuvant checkpoint blockade for cancer immunotherapy," Science, vol. 367, no. 6477, pp. 674–689, 2019.
[6] A. Huang, L. A. Arrawray, A. Ashworth, and B. Weber, "Synthetic lethality as an engine for cancer drug target discovery," Nature Reviews. Drug Discovery, vol. 19, no. 1, pp. 23–38, 2020.
[7] B. Melichar, "Biomarkers in the treatment of cancer: opportunities and pitfalls," Clinical Chemistry and Laboratory Medicine, vol. 51, no. 7, pp. 1299–1333, 2013.
[8] J. S. Mattick, "The genetic signatures of noncoding RNAs,” PLoS Genetics, vol. 5, no. 4, article e1000459, 2009.
[9] A. M. Schmitt and H. Y. Chang, "Long non-coding RNAs in cancer pathways," Cancer Cell, vol. 29, no. 4, pp. 452–463, 2016.
[10] I. Ulitsky and D. P. Bartel, "lincRNAs: genomics, evolution, and mechanisms," Cell, vol. 154, no. 1, pp. 26–46, 2013.
[11] J. Beermann, M. T. Piccoli, J. Viereck, and T. Thum, "Non-coding RNAs in development and disease: background, mechanisms, and therapeutic approaches," Physiological Reviews, vol. 96, no. 4, pp. 1297–1325, 2016.
[12] C. Jiang, X. Li, H. Zhao, and H. Liu, "Long non-coding RNAs: potential new biomarkers for predicting tumor invasion and metastasis," Molecular Cancer, vol. 15, no. 1, p. 62, 2016.
[13] S. Geisler and J. Coller, "RNA in unexpected places: long non-coding RNA functions in diverse cellular contexts," Nature Reviews. Molecular Cell Biology, vol. 14, no. 11, pp. 699–712, 2013.
[14] F. P. Marchese, I. Raimondi, and M. Huarte, "The multidimensional mechanisms of long noncoding RNA function," Genome Biology, vol. 18, no. 1, p. 206, 2017.
[15] W. Ni, S. Yao, Y. Zhou et al., "Long noncoding RNA GAS5 inhibits progression of colorectal cancer by interacting with and triggering YAP phosphorylation and degradation and is negatively regulated by the m6A reader YTHDF3," Molecular Cancer, vol. 18, no. 1, p. 143, 2019.
[16] Z. Li, X. Yu, and J. Shen, "ANRIL: a pivotal tumor suppressor long non-coding RNA in human cancers," Tumour Biology, vol. 37, no. 5, pp. 5657–5661, 2016.
[17] D. Huang, J. Chen, L. Yang et al., "NKILA lncRNA promotes tumor immune evasion by sensitizing T cells to activation-
induced cell death,” Nature Immunology, vol. 19, no. 10, pp. 1112–1125, 2018.

[18] X. Hu, Y. Feng, D. Zhang et al., “A functional genomic approach identifies FAL1 as an oncogenic long noncoding RNA that associates with BMI1 and represses p21 expression in cancer,” Cancer Cell, vol. 26, no. 3, pp. 344–357, 2014.

[19] J. Chen, Z. Wu, and Y. Zhang, “LncRNA SNHG3 promotes cell growth by sponging miR-196a-5p and indicates the poor survival in osteosarcoma,” International Journal of Immunopathology and Pharmacology, vol. 33, p. 205873841882074, 2019.

[20] F. Fei, Y. He, S. He et al., “LncRNA SNHG3 enhances the malignant progress of glioma through silencing KL2F and p21,” Bioscience Reports, vol. 38, no. 5, 2018.

[21] L. Hong, W. Chen, D. Wu, and Y. Wang, “Upregulation of SNHG3 expression associated with poor prognosis and enhances malignant progression of ovarian cancer,” Cancer Biomarkers, vol. 22, no. 3, pp. 367–374, 2018.

[22] Q. Ma, X. Qi, X. Lin, L. Li, L. Chen, and W. Hu, “LncRNA SNHG3 promotes cell proliferation and invasion through the miR-384/hepatoma-derived growth factor axis in breast cancer,” Human Cell, vol. 33, no. 1, pp. 232–242, 2020.

[23] T. Zhang, C. Cao, D. Wu, and L. Liu, “SNHG3 correlates with malignant status and poor prognosis in hepatocellular carcinoma,” Tumour Biology, vol. 37, no. 2, pp. 2379–2385, 2016.

[24] Q. Zhao, C. Wu, J. Wang et al., “LncRNA SNHG3 promotes hepatocellular tumorigenesis by targeting miR-326,” The Tohoku Journal of Experimental Medicine, vol. 249, no. 1, pp. 43–56, 2019.

[25] “Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015: elaboration and explanation,” BMJ, vol. 354, p. i4086, 2016.

[26] A. Stang, “Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses,” European Journal of Epidemiology, vol. 25, no. 9, pp. 603–605, 2010.

[27] Z. Tang, C. Li, B. Kang, G. Gao, C. Li, and Z. Zhang, “GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses,” Nucleic Acids Research, vol. 45, no. W1, pp. W98–W102, 2017.

[28] Y. F. Jiang, H. Y. Zhang, J. Ke, H. Shen, H. B. Ou, and Y. Liu, “Overexpression of LncRNA GHTET1 predicts an unfavourable survival and clinical parameters of patients in various cancers,” Journal of Cellular and Molecular Medicine, vol. 23, no. 8, pp. 4891–4899, 2019.

[29] L. Peng, Y. Zhang, and H. Xin, “LncRNA SNHG3 facilitates acute myeloid leukemia cell growth via the regulation of miR-758-3p/SRGN axis,” Journal of Cellular Biochemistry, vol. 121, no. 2, pp. 1023–1031, 2019.

[30] J. Shi, J. Li, S. Yang et al., “LncRNA SNHG3 is activated by E2F1 and promotes proliferation and migration of non-small-cell lung cancer cells through activating TGF-β pathway and IL-6/JAK2/STAT3 pathway,” Journal of Cellular Physiology, vol. 235, no. 3, pp. 2891–2900, 2020.

[31] G. Sui, B. Zhang, D. Fei, H. Wang, F. Guo, and Q. Luo, “The LncRNA SNHG3 accelerates papillary thyroid carcinoma progression via the miR-214-3p/PSMD10 axis,” Journal of Cellular Physiology, pp. 1–10, 2020.

[32] D. Tian, X. Wei, H. Zhu, L. Zhu, T. Li, and W. Li, “LncRNA-SNHG3 is an independent prognostic biomarker of intrahepatic cholangiocarcinoma,” International Journal of Clinical and Experimental Pathology, vol. 12, no. 7, pp. 2706–2712, 2019.

[33] Y. Xuan and Y. Wang, “Long non-coding RNA SNHG3 promotes progression of gastric cancer by regulating neighboring MED18 gene methylation,” Cell Death & Disease, vol. 10, no. 10, p. 694, 2019.

[34] Z. Taherian-Esfahani, M. Taheri, S. Dashti, V. Kolhghi-Oskoei, L. Geranpayeh, and S. Ghafouri-Fard, “Assessment of the expression pattern of mTOR-associated lncRNAs and their genomic variants in the patients with breast cancer,” Journal of Cellular Physiology, vol. 234, no. 12, pp. 22044–22056, 2019.

[35] P. F. Zhang, F. Wang, J. Wu et al., “LncRNA SNHG3 induces EMT and sorafenib resistance by modulating the miR-128/CD151 pathway in hepatocellular carcinoma,” Journal of Cellular Physiology, vol. 234, no. 3, pp. 2788–2794, 2019.

[36] S. Zheng, F. Jiang, D. Ge et al., “LncRNA SNHG3/miRNA-151a-3p/RAB22A axis regulates invasion and migration of osteosarcoma,” Biomedicine & Pharmacotherapy, vol. 112, p. 108695, 2019.

[37] S. Müller, S. Raulefs, P. Bruns et al., “Next-generation sequencing reveals novel differentially regulated miRNAs, lncRNAs, miRNAs, sdRNAs and a piRNA in pancreatic cancer,” Molecular Cancer, vol. 14, no. 1, 2015.

[38] S. Serrati, S. De Summa, B. Pilato et al., “Next-generation sequencing: advances and applications in cancer diagnosis,” OncoTargets and Therapy, vol. 9, pp. 7355–7365, 2016.

[39] S. Serghiou, A. Kyriakopoulou, and J. P. A. Ioannidis, “Long noncoding RNAs as novel predictors of survival in human colorectal cancer: a systematic review and meta-analysis,” Molecular Cancer, vol. 15, no. 1, p. 50, 2016.

[40] M. Xiao, Y. Feng, C. Liu, and Z. Zhang, “Prognostic values of long noncoding RNA PVT1 in various carcinomas: an updated systematic review and meta-analysis,” Cell Proliferation, vol. 51, no. 6, article e12519, 2018.

[41] H. YiRen, Y. YingCong, Y. Sunwu et al., “Long noncoding RNA MALAT1 regulates autophagy associated chemoresistance via PI3B-3p sequestration in gastric cancer,” Molecular Cancer, vol. 16, no. 1, p. 174, 2017.

[42] C. Feng, Y. Zhao, Y. Li, T. Zhang, Y. Ma, and Y. Liu, “LncRNA MALAT1 promotes lung cancer proliferation and gefitinib resistance by acting as a miR-200a sponge,” Archives of Bronconeumología, vol. 55, no. 12, pp. 627–633, 2019.

[43] K. Liao, Y. Lin, W. Gao et al., “Blocking lncRNA MALAT1/miR-199a/ZHX1 Axis Inhibits Glioblastoma Proliferation and Progression,” Molecular Therapy - Nucleic Acids, vol. 18, pp. 388–399, 2019.

[44] C. Zhang, Y. Qu, H. Xiao et al., “LncRNA SNHG3 promotes clear cell renal cell carcinoma proliferation and migration by upregulating TOP2A,” Experimental Cell Research, vol. 384, no. 1, p. 111595, 2019.

[45] L. Wang, K. Su, H. Wu, J. Li, and D. Song, “LncRNA SNHG3 regulates laryngeal carcinoma proliferation and migration by modulating the miR-384/WEE1 axis,” Life Sciences, vol. 232, p. 116597, 2019.

[46] L. Liu, J. Ni, and X. He, “Upregulation of the Long Noncoding RNA SNHG3 Promotes Lung Adenocarcinoma Proliferation,” Disease Markers, vol. 2018, 12 pages, 2018.

[47] W. Huang, Y. Tian, S. Dong et al., “The long non-coding RNA SNHG3 functions as a competing endogenous RNA to promote malignant development of colorectal cancer,” Oncology Reports, vol. 38, no. 3, pp. 1402–1410, 2017.