Abstract. 3D printing has emerged as an advanced manufacturing technology in the field of pharmaceutical sciences. Despite much focus on enteral applications, there has been a lack of research focused on potential benefits of 3D printing for parenteral applications such as wound dressings, biomedical devices, and regenerative medicines. 3D printing technologies, including fused deposition modeling, vat polymerization, and powder bed printing, allow for rapid prototyping of personalized medications, capable of producing dosage forms with flexible dimensions based on patient anatomy as well as dosage form properties such as porosity. Considerations such as printing properties and material selection play a key role in determining overall printability of the constructs. These parameters also impact drug release kinetics, and mechanical properties of final printed constructs, which play a role in modulating immune response upon insertion in the body. Despite challenges in sterilization of printed constructs, additional post-printing processing procedures, and lack of regulatory guidance, 3D printing will continue to evolve to meet the needs of developing effective, personalized medicines for parenteral applications.

KEY WORDS: 3D printing; parenteral; personalized medicines; printability.

INTRODUCTION

3D printing has revolutionized the way researchers approach developing treatments for patients in recent years. 3D printing is an additive manufacturing technology in which objects are constructed in a layer-by-layer fashion. Layer adhesion can be achieved via heat fusion, ultraviolet light (UV), and through chemical bonding depending upon the type of 3D printing technology used. Common techniques include fused deposition modeling (FDM), vat polymerization (VP), and powder bed printing. Although its history can be traced back to the 1980s, 3D printing was not well studied for pharmaceutical applications until the mid-2000s. In 2015, the US Food and Drug Administration (FDA) approved the first 3D printed drug product Spritam®, a fast disintegrating orodispersible tablet containing levetiracetam for epilepsy treatment [1]. Spritam® is produced using ZipDose® technology, which is a proprietary powder bed-based 3D printing technology capable of producing highly porous tablets. The FDA approval of this product was pivotal as it demonstrated the commercial success of 3D printed drug products. There have been numerous research articles and reviews highlighting applications of 3D printing technology in oral dosage forms. However, there is a lack of literature and research centered on 3D printing for parenteral applications.

3D printing allows for quick and flexible design and production of patient-personalized parenteral medicines, with precise control over size and shape, porosity, and mechanical properties of printed constructs [2]. For example, 3D printing technology enables researchers to produce scaffolds with tunable drug release kinetics by modulating pore size/architecture as well as shape of printed constructs [3]. Through optimization of materials and printing parameters, 3D printed constructs can be fabricated exhibiting porous architecture with mechanical properties that more closely mimic native tissue, resulting in more biocompatible constructs favoring cell adhesion and proliferation, suitable for regenerative applications [4]. In addition, 3D printing technology has evolved to allow for multi-material printing, which allows scientists to harness the benefits of each material in a single dosage form. This enhanced design flexibility has paved the way for the development of complex constructs, such as fabricating prints with a core/shell structure to enhance either...
stent patency or improve vascularization for bone regeneration [5,6].

The present review highlights recent parenteral applications aided by 3D printing, current challenges, and future perspectives of this emerging manufacturing technology.

KEY ASPECTS OF FABRICATING PARENTERAL DOSAGE FORMS VIA 3D PRINTING

Types of 3D Printing Technology

Extrusion-Based 3D Printing

FDM is a popular type of extrusion-based 3D printing technology used to produce parenteral dosage forms. As shown in Fig. 1a, FDM works by feeding a filament, typically consisting of a thermoplastic material (or blend of materials) into a high temperature nozzle to melt the material before being extruded onto the lower temperature build plate, where it is then cooled and solidified [7]. It allows for the production of 3D printed constructs in a quick and efficient manner, ideal for rapid prototyping [8]. Aside from material properties, final product quality is governed by FDM process parameters, including extrusion temperature, layer thickness, and nozzle diameter of the printing head [9]. Common filament materials used include biocompatible thermoplastic polymers such as polyvinyl alcohol (PVA), poly(lactic acid) (PLA), and polyvinylpyrrolidone [10]. One disadvantage of FDM technology is that typically, printing material needs to be inserted into a nozzle in the form of a solid filament, which does not exist for many pharmaceutical materials [10]. Thus, companion techniques, such as hot melt extrusion (HME), may be used to transform pharmaceutical grade materials, including active pharmaceutical ingredients (API), into FDM-suitable filaments [8]. Filaments can also be impregnated with an API solution during the production of filaments, usually via HME [11]. However, thermolabile therapeutics are not suitable for extrusion via FDM, due to potential degradation concerns [12]. It is worth mentioning that extrusion-based bioprinting at room or body temperature using bioinks has shown attractive clinical potential in achieving personalized treatment. For example, Long et al. developed a personalized 3D printed wound dressing composed of chitosan and pectin using an extrusion-based bioprinter, with the ability to control dimensional properties such as thickness (e.g., layer height of 0.25 mm) and pore size, while allowing for a facile lidocaine incorporation for immediate pain relief [13].

Vat Polymerization

In the case of VP technology, a build plate moves along the z-axis inside of a vat containing liquid resin, consisted of photopolymerizable monomer(s) and photoinitiator(s) (Fig. 1b). Once exposed to a specific wavelength of light (dependent on resin/printing material), polymerization of the monomer resin occurs [14]. This process continues, layer-by-layer, as unreacted functional groups in the previous layer are polymerized under light exposure, causing adherence to the current layer, resulting in layer formation [15]. API’s and excipients can be blended with a resin, effectively becoming trapped in the polymer matrix upon photopolymerization [16]. The two main VP techniques are “stereolithography” (SLA), which uses a UV laser beam, and “digital light processing” (DLP), which uses UV light from a projector, to cure the resin. Compared to SLA, DLP is more efficient as it can cure an entire layer at one time, reducing overall printing time. However, objects printed via SLA have a better spatial resolution (down to 25 μm) than DLP (35–100 μm), mainly due to the small optical spot size of SLA lasers [17]. VP printed objects typically undergo a post-printing curing process to ensure complete polymerization and further improve mechanical integrity of the prints, while reducing potential toxicity associated with the presence of residual monomers and oligomers [18,19]. SLA parameters (such as laser power and scan speed) and general VP parameters (e.g., resin characteristics and curing duration) contribute to the overall resolution of final prints [20]. VP printing can circumvent issues associated with other 3D printing technologies, such as avoiding thermal degradation of thermolabile therapeutics [21]. More importantly, VP techniques can achieve the highest printing resolution among all 3D printing technologies and hence process great translational potential in personalized implants. VP printing has been successfully used for parenteral applications, such as producing hydrogels for nerve and tissue regeneration [22–25]. However, the clinical applications of VP printing are still limited due to the lack of photopolymers with suitable mechanical properties and biocompatibility. Another major drawback to VP printing is that most commercial VP printers do not allow for multi-resin printing, thus limiting material selection and print design. To address this challenge, Konasch et al. developed a hybrid additive manufacturing technique combining both SLA and inkjet printing technologies to produce poly(ethylene glycol) diacrylate (PEGDA)-based matrices with multiple drug depots [26]. Essentially, a modified SLA printer was used to build the layers of the matrix system, consisting of PEGDA and lithium phenyl-2,4,6-trimethylbenzoyl-phosphinate (LAP) as the photoinitiator. Two inkjet print-heads were used to deposit two materials layer-by-layer inside of the PEGDA-based matrix and modulated drug release was achieved by changing spatial positioning of such depots. Other researchers have attempted to produce multilayered constructs by manually pausing prints, swapping resin tanks, and continuing the print, which is inefficient and negates the autonomy inherent to the 3D printing process [27].

Powder Bed Printing

Powder bed printing encompasses inkjet printing and selective laser sintering (SLS) technologies. Typically, powder bed printed constructs exhibit a minimum feature size down to approximately 50 μm [28]. Inkjet printing, illustrated in Fig. 1c, is a technique in which droplets of a binder solution are dispensed through printing heads, driven either by piezoelectric or thermal processes, onto a thin layer of bulk material, positioned in the powder bed [29]. Inkjet printing parameters including nozzle diameter and binder rheological properties have been shown to play a key role in modulating binder droplet size, which in turn impacts printing precision [30]. Huang et al. produced levofloxacin implants with predefined microstructures via inkjet printing. It was observed that more
complex drug release (e.g., bimodal and pulsatile) can be achieved via this method in comparison to the traditional compression method, as 3D printing allows for the flexibility to incorporate multiple types of structures such as reservoir and matrix architectures in one dosage form [31]. A disadvantage associated with inkjet printing is caused by the binder hitting the powder bed and displacing powder, leading to sub-surface depletion zones. SLS is very similar to inkjet printing, instead using a laser to bind particles/powder together to form layers, as opposed to depositing a binding solution (Fig. 1d) [32]. Xia et al. developed SLS printed nano-hydroxyapatite/poly-ε-caprolactone (PCL) scaffolds with a highly porous architecture (150 μm in layer thickness and 70–79% porosity) and sustained rhBMP-2 release for improved bone defect repair [33]. One main drawback associated with SLS is potential degradation of payload when exposed to high energy lasers used in the printing process, which has limited its pharmaceutical applications [14].

**Design and Personalization**

Additive manufacturing techniques enable the design and production of patient-centric dosage forms with precise control over dimension and microstructure, a feat not achieved through traditional manufacturing techniques, like compression. These factors ultimately play a key role in modulating drug release kinetics [34]. Computer-aided design (CAD), preparation, and evaluation have shown great application prospects in the field of 3D printing. Firstly, 3D scanning combined with digital modeling can greatly improve the accuracy of models and printlets, promoting personalized clinical use. Dosage forms (e.g., implants, wound dressing) can be efficiently designed in a CAD software to precisely match patient anatomy, typically through the use of medical imaging data such as computer tomography (CT) or magnetic resonance imaging (MRI) scans (as illustrated in Fig. 2). Then, a software such as Slicer can be used to open these files, typically in a digital imaging and communications in medicine (DICOM) format. Next, segmentation is used to partition the image into different sections of interest (i.e., tissue, organs), and the image is saved as an STL file (3D readable format) and only the region of interest that has been sectioned off will be saved. A slicing software such as Chitubox can then be used to open up STL files and slice them to determine the number of layers in the final 3D printed object, before finally being printed. For example, customized molars with carious cavities were obtained via 3D scanning and FDM printing, and personalized dental fillers with high mechanical strength and “on-demand” drug release characteristics were fabricated [35]. In another study, CT images were used to create a CAD model of orbital floor implants, and a bio-compatible polycarbonate ISO (PC-ISO) material was used to print implants for the treatment of orbital fractures [36].

Computer-aided methods have also been used to visualize and optimize the printing process. For example, finite element method (FEM) was used to elucidate the mechanism of FDM process by simulating the stress-strain behavior of filament during extrusion (Fig. 3) [37]. Computational fluid dynamics (CFD) was used to understand the melt flow field in the printing head during extrusion and provide useful information for further formulation and process optimization [38]. CFD has also been used to predict the flow velocity within different nozzle geometries during bioprinting and to establish a viability-stress-time-viscosity mathematical relationship [39]. Furthermore, computer-aided methods have been used for in vitro and in vivo evaluation of printlets. FEM was used to mimic the biomechanical properties of implants.
by simulating the Von Mises stress and strain distribution, while CFD was used to visualize the two-liquid mixing process and predict the perfusion process [40].

Printability

In general, the term “printability” relates to the deformation resistance of material(s) during and after printing, which is influenced by factors including mechanical properties, thermal properties, and/or gelation mechanism of the material(s) [41]. Adequate mechanical properties are important to enable successful deposition of multiple layers during the printing process [42]. With respect to bioprinting, in which biological materials (e.g., cells) are blended with traditional scaffolding materials, cell survival rate post-printing is also a key parameter that contributes to the overall printability of the printing ink [43].

Printability of extrusion-based systems can be impacted by a variety of factors, including material properties (e.g., swelling, mechanical, and rheological) as well as printing parameters such as nozzle size, air pressure, and printing speed [44]. Yang et al. investigated the printability of a gelatin-based thermosensitive extrudable paste and found that addition of 25% microcrystalline cellulose resulted in filament with enhanced mechanical properties, and thus improved deformation resistance [38]. It has been previously reported that a shear thinning material with a viscosity ranging from 400 to 3000 mm² s⁻¹ exhibits a rapid gelling time, allowing for successful deposition and layering and hence, high fidelity prints [45,46]. Printing fidelity of VP processes is influenced by parameters such as photopolymer concentration, addition of plasticizers, and printing parameters (e.g., layer height and exposure time) [40]. Light attenuating additives (i.e., tartrazine, coccine) are commonly used in VP resins to absorb excess light, allowing for controllable photopolymerization, resulting in formation of layers with desired thickness [47]. In addition, post-printing curing procedures involving exposure to UV light and elevated temperature may lead to shape deformations, thus reducing overall printability [48]. Powder bed printability can be attributed to parameters including powder particle size and binder over-spreading. In general, binder over-spreading can lead to prints with reduced dimensional accuracy [49]. Specifically, inkjet printability is largely dependent upon properties of ink, nozzle, and actuator. Ink properties such as viscosity and surface tension impact the resolution and uniformity of printed constructs. Nozzle properties (e.g., nozzle diameter and nozzle-substrate distance) impact Reynolds, Weber, and Ohnesorge numbers which are dimensionless numbers used to describe printability and droplet formation from inkjet printers [50]. In addition, inkjet printers rely on actuators, which can include piezoelectric actuation, electromagnetic forces, thermal actuation, and pneumatic pumps to eject ink droplets from the nozzle by overcoming

Fig. 2. Workflow diagram depicting process of fabricating a therapeutic loaded (indicated by green triangles) and personalized 3D printed wound dressing, including a 3D design of construct via computer aided design software, such as Tinkercad; b Export design as stereolithography/standard triangle language (STL) file into slicing software, such as Chitubox, and slice into layers; c Upload the STL file into 3D printer and execute printing; and d Apply wound dressing to affected area to release payload as desired.
ink surface tension [51]. Actuator type and its parameters can impact droplet size and overall print quality of constructs. Similarly, powder composition and properties such as particle size and polymer molecular weight (MW) impact printability and drug release behavior of SLS printed constructs [52]. Finer powder results in structures exhibiting enhanced green strength, smoother surface, quicker drug release, and reduced porosity, as well as an overall improvement in mechanical properties [53–55]. Laser properties (e.g., laser energy density) can also have an impact on SLS printability by altering powder bed temperature [32]. Lastly, high laser scan speeds have resulted in constructs exhibiting increased porosity, leading to rapid drug release and reduced mechanical properties due to shortened contact time between laser and powder [56,57].

Printing Materials

Materials used in 3D printed parenteral constructs need to be biocompatible to minimize immune response in the body, in addition to demonstrating suitable mechanical properties to ensure sufficient printability. Considerations such as ability to promote cell adhesion and proliferation should also be taken into account for parenteral applications including bone and tissue scaffolds [58]. Some materials commonly used in 3D printed parenteral constructs are listed in Tables I and II.

Synthetic Materials

Synthetic polymers such as polyesters, PVA, and polyurethane (PU) typically have more reproducible polymer characteristics and desirable mechanical properties (e.g., tensile strength and elastic modulus) compared to natural materials, which makes them more suitable for 3D printing applications [61]. Polyesters. Biodegradable polyester-based synthetic polymers, such as PLA, poly(lactic-co-glycolic acid) (PLGA), and PCL, are relatively hydrophobic and inherently biologically inert. Owing to their excellent biocompatibility and tunable mechanical properties, PLA and PLGA, a class of aliphatic polyesters, are suitable for 3D printing applications [62,63]. Tappa et al. developed FDM printed PLA-based osseous fixation devices, including surgical screws, pins, and bone plates [64]. The 3D printed PLA devices exhibited compressive strengths between 20 and 500 MPa, demonstrating feasibility in orthopedic applications. In another study, Wang et al. developed a 3D printed bilayer membrane, consisting of a PLGA nanofiber outer layer (layer height: 0.05 μm) and alginate hydrogel inner layer (layer height: 100 μm), designed to mimic the epidermal and dermal layers of the skin for use as a wound dressing with demonstrated accelerated wound healing ability in vivo [65]. Combining polyester materials with other polymers such as PU has been used to further enhance mechanical properties of polyesters [66]. However, the hydrophobicity of polyesters results in inadequate cell adhesion and poor osteogenesis, as well as potential bacterial adhesion and biofilm formation [67]. Thus, 3D printed polyester-based constructs have been functionalized with biomolecules such as collagen, minocycline, and hydroxyapatite to improve cell adhesion and promote bone regeneration [68,69]. In addition, chemical structure modifications have been used to improve cell binding and hydrophilicity of polyesters [70].

Photopolymers. Biocompatible photopolymers such as PEGDA, PEG dimethacrylate (PEGDMA), and gelatin methacrylate (GelMA) are commonly used in VP technology (Table II). PEGDMA hydrogels exhibit similar compressive modulus to musculoskeletal tissue, making them a suitable
choice for bone regeneration applications. A bioprinting setup, consisting of a Hewlett-Packard (HP) Deskjet thermal inkjet printer modified with an overhead UV lamp, was used to produce 3D printed PEGDMA bone constructs with Irgacure I-2959 as the photoinitiator [71]. In another study, Zhou et al. developed a GelMA-based bioink suitable for DLP printing containing LAP photoinitiator and a hyaluronic acid (HA) derivative to create functional living skin for skin regeneration applications [59]. Another photopolymer, polylactide (PLA) (L-form) (PLA), poly(lactic-co-glycolic acid) (PLGA), poly(ethylene glycol) (PEG) diacrylate (PEGDA), gelatin methacrylate (GelMA), and hyaluronic acid (HA) are generally non-biodegradable but biocompatible and have been extensively used in various medical applications including vascular grafts, catheters, heart valves, and wound dressings. Recently, researchers have synthesized biodegradable PU to expand their biomedical applications [76,77]. Interestingly, PU are comprised of alternating hard and soft segments, the former owing to long-chain diols, which leads to enhanced elasticity, and the latter responsible for overall material strength due to the presence of crystalline regions [78]. Jung et al. developed 3D printed thermoplastic PU-based tracheal prostheses with higher tensile strength and enhanced flexibility compared to native trachea tissue. The microporous architecture of the 3D printed prostheses promoted biological interactions by allowing for cellular infiltration and facilitating ingrowth of connective tissue [79].

### Table I. Representative Materials Used in 3D Printing of Parenteral Dosage Forms

| Material                      | Tg (°C) | Tm (°C) | Tensile strength (MPa) | Elongation at break (%) | Degradation duration (months) | Ref       |
|-------------------------------|---------|---------|------------------------|-------------------------|------------------------------|-----------|
| Synthetic                     |         |         |                        |                         |                              |           |
| PLA (L-form)                  | 60–70   | 173–178 | 60–70                  | 2–6                     | >24                          | [119,153–157] |
| PLA (DL-form)                 | 45–60   | NA      | 40                     | 1–6                     | 12–16                        |           |
| PLGA (82/18)                  | 50      | 135–145 | 60–70                  | 2–6                     | 12–18                        |           |
| PDLGA (50/50)                 | 40–50   | NA      | 40–50                  | 1–4                     | 1–2                          |           |
| PCL                           | (–65)–(–54) | 55–63 | 23                    | >4000                   | 24–36                        |           |
| PLA/PCL (70/30)               | 20      | 100–125 | 18–22                  | >100                    | 12–24                        |           |
| PU                            | –73     | 64      | 1.8                    | 65                      | >6                           | [158,159] |
| PVA                           | 85      | 180–228 | 3.2–4.6                | 52–500                  | 2–3                          | [160–162] |
| Natural                       |         |         |                        |                         |                              |           |
| CS                            | 140–150 | NA      | 19–24                  | 13–20                   | >3                           | [163–165] |
| Alginate                      | 81      | NA      | 0.3–0.9                | 99–193                  | 10                           | [166–168] |
| HA                            | (–48)–(–80) | NA | 0.06–0.1              | >600                    | 10                           | [169–171] |
| SF (B. mori)                  | 175     | 256     | 1.5–15.9               | 10–50                   | 10                           | [172–175] |
| Collagen                      | 35      | 28–36   | 0.1–0.12               | 380                     | ≈1                           | [176–178] |
| Gelatin                       | 18–28   | 29–37   | 0.196–0.35            | 5–10                    | >1                           | [179–181] |

*PLA,* poly(lactic acid); *PLGA,* poly(lactic-co-glycolic acid); *PDLGA,* poly(DL-lactide-co-glycolide); *PCL,* polycaprolactone; *PU,* polyurethane; *PVA,* poly(vinyl alcohol); *CS,* chitosan; *HA,* hyaluronic acid; *SF,* silk fibroin

### Table II. Representative Photopolymers Used in 3D Printing of Parenteral Constructs

| Material | UV wavelength (nm) | Photoinitiator | Photoabsorber | Mechanical properties | Ref       |
|----------|--------------------|----------------|---------------|-----------------------|-----------|
| PEGDA    | 405                | LAP            | Orange G dye  | NA                    | [26,182]  |
| PEGDMA   | 315–400            | Irgacure I-2959 NA | Compressive/storage modulus: 37–500 Pa/14–70 kPa | [69,183,184] |
| GelMA    | 365                | LAP            | NA            | Young’s modulus: 31 kPa | [59]      |
| PPF      | 365                | Irgacure I-2959 NA | Compression/tensile stiffness: 394/463 N/mm | [60]      |

*PEGDA,* poly(ethylene glycol) diacrylate; *PEGDMA,* poly(ethylene glycol) methacrylate; *GelMA,* gelatin methacrylate; *PPF,* polypropylene fumarate
Natural Polymers

Natural polymers (e.g., chitosan, collagen, and gelatin) possess better biocompatibility and biological activity, including supporting cell attachment and differentiation, compared to synthetic polymers, making them an ideal choice for use in bioprinting applications [80]. However, natural polymers typically exhibit weaker mechanical properties than synthetic polymers, which may lead to a reduction in printability and hence unsuccessful prints. To overcome this hurdle, strategies such as forming composites with “stronger” materials (e.g., tricalcium phosphate, graphene), increasing crystallinity, and optimizing cross-linking conditions have enabled successful printing of constructs with enhanced mechanical properties [81].

Polysaccharide-Based Materials. Chitosan (CS), a cationic linear polysaccharide, has been shown to accelerate wound repair by promoting tissue growth and differentiation [82]. CS has also been shown to accelerate the formation of osteoblasts, leading to enhanced bone regeneration, and promote connective tissue regeneration [83]. Physicochemical properties (such as solubility, crystallinity, and degradation) of CS can be modulated by altering CS MW and degree of deacetylation [84]. CS has been used in 3D printing of both soft tissue (e.g., wound dressing) and hard tissue applications such as bone regeneration [85]. However, CS alone has relatively weak mechanical properties and poor printability. Thus, blending other materials (such as gelatin) with CS has been shown to improve mechanical properties, resulting in a higher fidelity print [86]. In a recent study, Intini et al. developed a FDM printed CS-raffinose scaffold for diabetes-related wound healing [87]. Addition of raffinose has been shown to enhance the mechanical properties, wettability, and hydrophilicity of CS films, thus promoting tissue regeneration in a rat model [88].

Alginate is a polysaccharide isolated from the cell walls of brown algae. Alginate can be cross-linked with calcium ions (Ca$^{2+}$) in a facile manner to produce constructs that effectively mimic an extracellular matrix (ECM) structure [89]. The use of alginate in wound dressings has resulted in accelerated wound healing, due to its ability to maintain a moist environment and minimize bacterial infection [65]. Thus, alginate remains an excellent material choice for parenteral applications including wound dressings and tissue regeneration. Composite materials containing alginate and other polymers (e.g., gelatin) have been used to improve mechanical properties of alginate [90]. Li et al. developed graphene oxide (GO)-coated 3D printed alginate/gelatin scaffolds with enhanced mechanical strength as well as improved osteogenic differentiation and cell adhesion for bone regeneration applications [91].

HA, a linear biodegradable polysaccharide, has a ubiquitous presence and serves numerous roles in the human body, such as maintaining the ECM structure by interacting with proteoglycans and link proteins, in addition to acting as a signal molecule by interacting with various cell surface receptors, thereby mediating cellular functions [92]. Therefore, HA is an ideal material for use in parenteral applications such as wound healing and tissue/cartilage engineering. However, similar to other natural polymers, HA exhibits unfavorable mechanical properties, leading to low shape fidelity and poor printability for 3D printing applications. To overcome this limitation, Ouyang et al. developed extrusion-based 3D printed HA scaffolds via dual cross-linking (i.e., supramolecular and UV cross-linking) for use in cartilage and tissue engineering applications [42].

Protein-Based Materials. Over the last decade, silk derived from Bombyx mori (B. mori) silkworms has gained much attention across a host of biomedical applications including drug delivery and tissue regeneration, due to its impressive mechanical properties, biocompatibility, and processability [93]. The two main components of B. mori silk are as follows: (1) sercin (SS), a glue-like outer protein coating that is commonly removed via a degumming process to enhance overall mechanical properties; and (2) fibroin (SF), an insoluble inner core protein that provides mechanical stability [94]. Silk has been shown to exhibit enhanced tensile modulus and strength, compared to traditional well-studied polymers (e.g., collagen and PLA). Silk (without SS) exhibits an over 300-fold increase in modulus and approximately a 100-fold increase in ultimate tensile strength when compared to collagen. It also exhibits a 5-fold increase in modulus and a nearly 14-fold increase in ultimate tensile strength when compared to PLA [93]. In general, SF is an attractive bioink, as it can maintain cell viability and structural integrity of a 3D scaffold. Recently, a stem cell laden SF/gelatin hydrogel was bioprinted using an in-house designed multi-head deposition-based 3D printer [95]. Impressively, the printed SF/gelatin hydrogel maintained cell viability for more than 30 days.

Collagen is the most abundant protein ubiquitous in the human body, exhibiting a rod shaped quaternary structure formed via the entanglement of three left handed helices [96]. Collagen has been used to mimic the ECM structure in vitro, and has been shown to promote cell adhesion, proliferation, and migration of various types of cells, including bone marrow mesenchymal cells for tissue engineering applications [97]. Nocera et al. developed a porous collagen (type I)-based scaffold using an in-house extrusion-based 3D printer equipped with syringes and 21G needles [98]. The 3D printed scaffolds can support cell attachment and proliferation of fibroblast cells without cytotoxicity.

Gelatin, a type of linear peptide (MW: 15 to 250 kDa), is produced via heat and enzymatic denaturation of collagen [99]. Cross-linking of gelatin can be accomplished via chemical and enzymatic reactions, in addition to physical cross-linking which can be accomplished by heating gelatin solution to around 40–50°C, before cooling below 30°C at which point a semi-solid gel is formed [99]. Negrini et al. developed chemically cross-linked, gelatin-based scaffolds for adipose tissue engineering applications using an extrusion-based 3D printer [100]. Results showed that the scaffolds remained stable for 21 days and exhibited similar mechanical properties as native adipose tissue and supported adipogenic differentiation.

3D PRINTING IN PARENTERAL APPLICATIONS

3D printing technology has been successfully utilized to fabricate parenteral constructs such as implants, stents, and wound dressings (Table III).
### Table III. Examples of 3D Printing Technology for Parenteral Applications

| Materials | 3D printer | Design | Applications | Ref |
|-----------|------------|--------|--------------|-----|
| Implants  |            |        |              |     |
| PLLA      | SLA        | Anatomically relevant spherical or cylindrical shape | Sustained release of multiple chemotherapeutics for 12 weeks for osteosarcoma therapy. | [185] |
| PLA, PVA, PCL | FDM      | Rod-shaped implants containing different sized “windows” | Sustained payload release from implants modulated via the “windows.” | [101] |
| PCL, PLGA | Extrusion-based | Patches with different shaped pores | Sustained 5-fluoracil release over 4 weeks for pancreatic cancer therapy. | [186] |
| Calcium phosphate | Inkjet printer | | Co-delivery of multiple antibiotics for the treatment of bone osteomyelitis. | [102] |
| PLA, collagen, hydroxyapatite | FDM | | Combination of the macroporous architecture and antibiotic release allowing vascularization while against bone infection. | [103] |
| PLA, PCL | FDM | | Shape-dependent progesterone release for contraceptive purposes. | [104] |
| Biomedical devices |            |        |              |     |
| PCL, PLA | FDM | | | |
| PCL, sulfated CS (26SCS) | Extrusion-based | Biodegradable polymer-coated stents | Stents exhibited excellent anti-coagulant activity and biocompatibility for cardiovascular disease management. | [188] |
| PLA, polydopamine, PEI, heparin | Extrusion-based | Biodegradable polymer-coated stents | PVA-based biliary stents with resisted biofilm formation and enhanced stent patency for biliary obstruction. | [75] |
| PVA, collagen, PCL, cholangiocyte | FDM | Stem cell-coated biliary stent | Sustained delivery of multiple therapeutics with similar mechanical properties as conventional coronary stents (elastic modulus 400 MPa) | [189] |
| PCL, graphene | FDM | Multi-drug eluting stent | Flexible, self-expanding stents with reduced stent migration for cardiovascular disease management. | [190] |
| PLA, TPU | FDM | PLA/TPU stent with spiral patterns with controllable spiral angle, thickness, and pitch | | |
| Wound dressing | | | | |
| Pectin, CS | Extrusion-based | Hydrogel scaffold | Wound dressings exhibited good bioadhesion strength (86.5–126.9 g), while maintaining a moist environment for skin wound healing. | [13] |
| PCL, FPLA, PEGDA, PEG | FDM/SLA | Personalized anti-acne patches/masks | Personalized acne treatment with salicylic acid based on patient scans. | [191] |
| Chitosan, genipin, PEG | Extrusion-based | Films | Mucoadhesive and swellable films for payload release to promote skin wound healing. | [192] |
| CS, raffinose | FDM | Wound dressing with controllable microarchitecture | CS scaffolds promoted tissue regeneration in a diabetes-related skin wound rat model. | [87] |
| SS, GelMA | Extrusion-based | Transparent hydrogel scaffold with controllable pore sizes | Wound dressing designed for real-time monitoring of wound healing process. | [193] |
| PU, HA | FDM | Scaffolds designed to release two biomolecules | Sustained release of multiple therapeutics to accelerate wound healing process for cartilage defect. | [194] |
| PLGA, alginate | Extrusion-based | Bilayer membrane designed to mimic the skin dermis and epidermis | Porous bilayer wound dressing to enhanced wound repair or be used as a skin substitute. | [65] |

PLL, poly(L-lactide); PEI, polyethyleneimine; TPU, thermoplastic polyurethane; FPLA, Flex EcoPLA™; SS, silk sericin; PEG, polyethylene glycol; FDM, fused deposition modeling; SLA, stereolithography
Long-Acting Implants/Inserts

Long-acting implants/inserts have been widely utilized for various clinical applications, including contraception, cancer treatment, and localized delivery of anesthetics and antibiotics [105–108]. 3D printed implants can be designed to achieve tunable, sustained drug release through precise control over implant shape, size, and microstructure. In a recent study, Stewart et al. produced 3D printed rod-shaped PVA/PLA implants via FDM, with designed “windows” to modulate drug release [101]. Implants with smaller “windows” and a decreased number of total “windows” resulted in slower payload release. Impressively, these implants, dip-coated with a PCL polymer mixture, can retard payload release for up to 300 days. In order to provide personalized vaginal rings to avoid pelvic inflammatory disease and uterine perforations [109], Fu et al. developed 3D printed PLA/PCL composite vaginal rings with customized shapes (i.e., “O,” “Y,” and “M”) to accurately mimic the structure of female anatomy [104]. All printed vaginal rings exhibited shape-dependent progesterone release for 7 days. Similarly, Tappa et al. developed 3D printed vaginal inserts containing either estrogen or progesterone [110]. The inserts were fabricated to mimic clinically relevant surgical meshes, intrauterine devices, and pessaries, and demonstrated excellent biocompatibility and sustained payload release (Fig. 4).

3D printing technology also allows for seamless integration of multiple API’s in a single dosage form for combination therapy. Qiao et al. developed 3D printed PLGA scaffolds for the combination therapy of doxorubicin and cisplatin against breast cancer [111]. The 3D printed scaffolds (pore sizes > 200 μm) were produced using a customized E-jet printer and were capable of delivering both drugs in a controlled release manner for up to 30 days, demonstrating synergistic antitumor effect of the combination therapy. Won et al. developed a 3D printed core (alginate and dexamethasone)/shell (PCL and bevacizumab) structured rod using a multi-head bioprinter for the treatment of retinal vascular diseases [112]. The rods exhibited sustained bevacizumab release over 60 days and dexamethasone release over 7 days, leading to suppressed angiogenesis over a 4-week period in a rat model.

Regenerative Applications

3D printing technology has shown promising in regenerative applications, particularly for treating bone defect, as it can accurately and quickly produce customizable scaffolds with defined microstructures and precise control over factors (such as shape, porosity, and mechanical properties), all of which impact magnitude of osteogenesis and angiogenesis [5,113–116]. For example, Zhang et al. developed a multifunctional bioceramic scaffold capable of promoting vascularized bone regeneration to treat large segmental bone defects [5]. Hollow-pipe-packed silicate bioceramic (BRT-H) scaffolds with a core/shell structure were produced via a modified extrusion-based 3D printer. The synergistic effect of the hollow channel structures produced via 3D printing and ionic components (e.g., silicon, magnesium, and calcium) of the alginate-based scaffold led to enhanced tissue growth and vascularization [117]. Similarly, Martin et al. engineered a multifunctional 3D printed PLA scaffold via FDM for bone regeneration [103]. The printed PLA scaffold exhibited a lattice-shaped structure with a controllable pore size of 1000 μm and a porosity around 55%. Multifunctionalization via a combination of collagen, minocycline, and hydroxyapatite, aided by scaffold porosity, resulted in improved antibacterial/antibiofilm properties while promoting osteogenesis. Calcium phosphate scaffolds (CPS) containing antibiotics (i.e., rifampin and vancomycin) have been developed via inkjet 3D printing for osteomyelitis therapy [102]. 3D printed porous CPS implants allowed for a 6-fold increase in vancomycin release compared to manually molded poly(methyl methacrylate) spacers, resulting in a reduction in mean bacterial load.

Implantable Biomedical Devices

Stents/Drug-Eluting Stents

Stents have been widely used to widen the affected blood vessels and restore blood flow for treating cardiovascular diseases, a leading cause of death around the world. Factors including stent strut thickness and structure (i.e., shape, geometry) have been shown to have a substantial impact on overall mechanical properties (e.g., radial force, radial recoil, and flexibility) of stents and hence stent effectiveness [118]. 3D printing technology allows for precise control over stent shape and dimensions using either a single material or combinations of multiple materials, to achieve desired mechanical and physical properties depending on application site, which is crucial to stent effectiveness [119]. Moreover, conventional methods (e.g., laser cutting) to produce metallic and polymeric stents can negatively impact overall stent microstructure, leading to microcracks [120]. 3D printing can minimize damage to stent microstructure by avoiding the use of high temperatures inherent to conventional laser cutting manufacturing. Recently, 3D printing technology has been implemented to produce biodegradable polymer-based stents with structure flexibility, ideal for ease of insertion, while maintaining a rigid structure to support the blood vessel. Guerra et al. developed biodegradable stents consisting of either PLA filament, PCL filament, or a combination of both via FDM printing [6]. PLA and PCL exhibit vastly different mechanical properties and degradation profiles, which when used alone, are insufficient for use in stent applications. However, when used together, composite stents can achieve more desirable mechanical properties. Composite stents composed of a PLA core and PCL shell exhibited a Young’s modulus around 1400 MPa and about 3% degradation over the span of 6 weeks, suitable for stent applications.

Drug-eluting stents (DES) can not only physically provide structure to keep the blood vessel open, but also release multiple therapeutics designed to treat post-surgical side effects such as inflammation. Therapeutics can be blended with polymers to create a drug-loaded filament for 3D printing, or coated on the surface of printed stents [121]. Kim et al. developed a 3D printed PCL DES, using a deposition-based 3D printer, to treat recurrent obstructive salivary gland disease, commonly caused by the buildup of salivary stones [121]. The stent shape was derived from CT images to mimic salivary ducts after removal of salivary...
stones. The DES was designed and printed within 7 min, and showed sustained amoxicillin release (up to 28 days) to treat *S. aureus* and a relatively faster cefotaxime release (up to 3 days) to treat *E. coli*, resulting in enhanced antimicrobial activity via the combination therapy [122].

**Wound Dressings**

Additive manufacturing has shown great promise in wound healing applications. Wound healing is a complex process, involving hemostasis, inflammation, proliferation, and remodeling [123]. Ideally, tissue-engineered constructs should facilitate the biological function (e.g., suitable angiogenesis and re-epithelialization) and closely resemble the structural organization of the native tissue, which is a feat that can be accomplished through 3D printing technology [124]. 3D printing allows for precise control over the spatial distribution of biological components, biomaterials, and therapeutics, to enhance cell migration and proliferation, and accelerating the overall wound healing process while reducing inflammation and scarring [125,126]. The 3D printed wound dressing demonstrated excellent flexibility and similar adhesive strength when compared to marketed wound dressings. Similarly, Hung et al. developed a PU scaffold using a self-modified, low temperature FDM printer. Either sustained co-delivery or sequential delivery of combination therapeutics can be achieved by modulating scaffold designs and printing parameters [127]. 3D printing can also produce patient-specific wound dressings that match anatomically complex architecture. For example, Muwaffak et al. developed a patient-specific 3D printed antimicrobial wound dressing made of PCL incorporated with metal ions (e.g., copper, zinc, or silver) [128]. A 3D scanner was used to obtain images of a patient’s ear and nose, which were then uploaded and edited in a 3D printing software. Results showed that 3D printed personalized dressings can match the anatomical complexity of a patient while providing sustained release of metal ions for 72 h, resulting in effective inhibition against *S. aureus*.

**Traditional Medical Devices**

Additive manufacturing has also been utilized to produce prototype traditional medical devices such as autoinjectors and ophthalmic devices. EpiPen®, one of the most well-known examples of autoinjectors, used to treat acute anaphylaxis by delivering epinephrine via IM route [129]. Typically, EpiPen® is only capable of administering a single dose of epinephrine, which may be inadequate for patients requiring an additional dose to alleviate their symptoms [130,131]. Thus, researchers have developed 3D printed prototypes designed to meet the need of administering multiple doses of epinephrine, manufactured using 3D printers such as the Stratasys OBJET CONNEX 500, the Stratasys Dimension, and the MakerBot Replicator [132]. Researchers have also developed more sustainable and eco-friendly alternatives to traditional disposable autoinjectors by designing 3D printed reloadable autoinjectors, containing replaceable epinephrine-loaded cartridges [133]. The use of 3D printing in the field of ophthalmology has also been on the rise, with the emergence of more biocompatible materials which can reduce the risk of rejection and irritation [134]. For example, 3D printed corneas have been produced from biocompatible materials such as alginate, collagen, and human stem cells, and have been designed to match patient specific corneal geometrical and thickness specifications [135]. Researchers are continuing to make strides in other ophthalmic applications, such as producing 3D printed intraocular...
lenses (IOL), which require careful consideration of other parameters, such as refractive index (RI) of 3D printed layers, and 3D printed retinas, the success of which depends on the ability to successfully print multiple retinal cell types [134].

**FUTURE PERSPECTIVES AND CHALLENGES**

3D printing has tremendous potential in personalized medicines via parenteral routes. It has been successfully utilized to print cells in specific and predetermined spatial arrangements, which closely mimic the cellular organization of native tissue for tissue regeneration applications. Researchers have now turned their attention to using this additive manufacturing technology to print entire organs to solve organ donor shortage and immune rejection issues [136]. In the future, this technology may become more popular in hospital and emergency room settings, as 3D printing allows for rapid fabrication of clinically relevant and on-demand constructs. Another intriguing avenue that researchers have begun exploring is the combination of 3D printed constructs and biomedical electronics. 3D printed implants/inserts offer an array of advantages over implants fabricated via traditional molding/extrusion methods, such as the ability to achieve patient-specific characteristics, load multiple therapeutics in one dosage form while avoiding incompatibilities between drugs, in addition to maintaining precise control over microstructure and mechanical properties, and drug release kinetics [137]. On the other hand, biomedical electronics have been used to achieve externally controlled drug delivery, including hormone-releasing microchips and miniaturized neural drug delivery systems [138,139]. Recently, Kong et al. developed an FDM printed, Bluetooth-enabled gastric resident electronic device capable of on-demand release of antimicrobial and hormonal agents [140]. In this application, 3D printing allows for the fabrication of an insert with precise dimensions and the seamless integration of multiple materials, including PLA and PU to amplify the adhesion strength between the materials, to achieve gastric retention over 36 days.

Despite the tremendous potential that 3D printing technology offers, there are some challenges that need to be addressed before this technology becomes mainstream in manufacturing of parenteral constructs. General 3D printing considerations, such as material selection, printing parameters, post-printing treatment, and material toxicity concerns, need to be addressed prior to achieving a successful print [141]. The development of new materials or printing inks is the key to the success in propelling 3D printing-based parenteral applications. Scientists often need to modify commercially available materials to satisfy certain printing requirements, such as improving mechanical properties for adequate printability [38]. In addition to impacting printability, mechanical properties of 3D printed constructs play a key role in modulating cell-scaffold interactions, especially in terms of cell adhesion ability and stent patency [142]. Products manufactured via certain 3D printing techniques, such as VP, require an additional post-printing curing step to enhance mechanical integrity of the product, adding to the complexity of the overall 3D printing process [18]. This step may also have a negative impact on loaded therapeutics, which can lead to compromised biocompatibility. Another important factor to consider is the sterilization of parenteral prints. Traditional sterilization techniques include exposure to gamma-irradiation, ethylene oxide gas, UV irradiation, ethanol washing, and autoclaving [143]. A recent study investigated the efficacy of sterilization techniques, including plasma irradiation and autoclave steam sterilization (121°C and 134°C), for surgical guides and implants [144]. It was concluded that both plasma irradiation and autoclave steam sterilization are suitable sterilization methods, and that high temperature steam sterilization caused significant deformation of 3D printed implants. While these sterilization methods are promising, they are not suitable for all 3D printed constructs, as high temperature sterilization methods can potentially compromise the integrity and efficacy of prints containing temperature sensitive polymers and/or thermobiodegradable therapeutics. In addition, while personalized constructs can be produced, the time it takes to 3D print individual prints far exceeds the time it takes to commercially manufacture them as a result of the layer-by-layer addition process inherent to 3D printing. Furthermore, there is a need to develop multi-material 3D printers to enable flexible construct design and combination therapy. Last but not the least, toxicological effects of materials used in the 3D printing process remain a paramount concern for researchers. Additives such as photoinitiators and cross-linking agents are typically required in the 3D printing process, to achieve successful prints. However, unreacted resin components found in VP type 3D printing techniques have been shown to present cytotoxicity concerns, and are a key reason many 3D printing procedures involve wash steps to remove unreacted material [145]. Two photoinitiators commonly used in the 3D printing process of parenteral application including LAP and Irgacure I-2959 have demonstrated increased cytotoxicity levels at elevated concentrations [145,146]. Alternative photoinitiators, such as riboflavin, have been shown to exhibit UV cross-linking ability while remaining non-cytotoxic, despite exhibiting lower print resolution due to longer reaction times [147]. Other cross-linking agents used in the development of parenteral delivery systems, such as glutaraldehyde, which has been used in combination with chitosan to produce hydrogels, has been shown to demonstrate mutagenic and neurotoxic properties [148]. Thus, materials used in the 3D printing process of parenteral applications must be carefully selected to ensure that final printed products remain biocompatible and non-toxic.

Despite the tremendous potential 3D printing has to offer for parenteral applications, regulatory guidance on characterization and assessment methods as well as process validation methodology remains scarce. While dozens of 3D printed medical devices have received FDA approval such as dental crowns and bone plates, only one pharmaceutical drug product, Spritam® medication, has been approved by the FDA [149,150]. Clinical trials are underway for other 3D printed medical devices, such as 3D printed patient-specific intramedullary guide and 3D printed denture framework. Most recently, the FDA held a public workshop in 2014 and issued a guidance in 2017 covering technical considerations for additive manufactured medical devices, such as information regarding design and manufacturing considerations for 3D printed medical devices [151]. This guidance recommends
material controls, describing specifications for raw materials including particle size, viscosity, and filament dimensions, should be well controlled. The guidance also recommends to understand and document the impact of post-processing steps involved in residue removal and sterilization, including heat or chemical treatments, on final product performance and properties. Furthermore, process validation, including assessments on device dimensions, feature geometry, and material properties, must be performed on final prints to ensure quality is maintained for parts produced in a single build cycle and between multiple build cycles. Lastly, final product mechanical properties such as modulus, yield strength, and creep should be investigated once all post-processing, cleaning, and sterilization steps have been performed. While this guidance provides insightful information, numerous regulatory concerns remain unaddressed, including regulation of 3D printed on-demand personalized products at hospitals and pharmacies and the regulation of printer ink and 3D printer manufacturing [152]. There is a growing interest to use 3D printing to produce parenteral dosage forms, and with this increased appeal, more specific and defined regulations will need to be established.

CONCLUSION

Although still in its infancy, 3D printing has already demonstrated tremendous potential for producing parenteral constructs. The importance of producing on-demand, personalized medications tailored to patient anatomy and disease conditions cannot be overstated. In addition, the ability to progress from design to prototype in a matter of hours allows scientists and physicians to quickly and efficiently test out various designs and therapeutic regimens until a desirable treatment is obtained. Additive manufacturing techniques also allow for the flexibility to combine multiple therapeutics in a single dosage form in a controllable and organized fashion. Biomedical devices and implantable scaffolds can be printed with controllable dimensions and microstructures, leading to tunable degradation and drug release characteristics, in addition to playing a key role in modulating cell proliferation and migration abilities. Thus, despite not being an optimal solution for large-scale manufacturing, the use of 3D printing for parenteral applications will continue to rise, to meet the growing demand for patient-centric medications.

DECLARATIONS

Conflict of Interest The authors declare that they have no conflict of interest.

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