Multimodal characterization of Yucatan minipig behavior and physiology through maturation

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Brain injuries induced by external forces are particularly challenging to model experimentally. In recent decades, the domestic pig has been gaining popularity as a highly relevant animal model to address the pathophysiological mechanisms and the biomechanics associated with head injuries. Understanding cognitive, motor, and sensory aspects of pig behavior throughout development is crucial for evaluating cognitive and motor deficits after injury. We have developed a comprehensive battery of tests to characterize the behavior and physiological function of the Yucatan minipig throughout maturation. Behavioral testing included assessments of learning and memory, executive functions, circadian rhythms, gait analysis, and level of motor activity. We applied traditional behavioral apparatus and analysis methods, as well as state-of-the-art sensor technologies to report on motion and activity, and artificial intelligent approaches to analyze behavior. We studied pigs from 16 weeks old through sexual maturity at 35 weeks old. The results show multidimensional characterization of minipig behavior, and how it develops and changes with age. This animal model may capitulate the biomechanical consideration and phenotype of head injuries in the developing brain and can drive forward the field of understanding pathophysiological mechanisms and developing new therapies to accelerate recovery in children who have suffered head trauma.

Brain disorders, diseases and injuries caused by external forces, such as sports collisions, car accidents, falls, violent attacks and blasts, remain challenging to model experimentally1–4. The nature and the extent of the injury depends upon a complex array of anatomical, physiological, and biomechanical parameters, including the size of the brain, thickness of skull, the ratio of grey to white brain matter, age at the time of injury, as well as the energy, angle, and acceleration of the impact itself1–4. A single event or series of events may result in changes in cognition, as well as sensorimotor and physiological impairments that are often difficult to measure in experimental models4–6.

An emerging animal model for brain injury research is the domestic pig (sus scrofa domesticus). There are distinct advantages to using a large animal model for in vivo brain injury research7–10. Compared to rodent models, which are the most commonly used animal models for central nervous system injury studies, pigs are much more similar to humans in terms of brain size11, neuroanatomy11, brain organization and morphology12,13, neurodevelopment5,14,15, and neuroinflammatory mechanisms16–18. Due to the strong similarities between pigs and humans, pigs are an ideal species for modeling pediatric brain injuries. All mammalian species experience a transient period of rapid brain growth termed the “brain growth spurt”, and the brain is recognized as being particularly vulnerable to the effects of developmental damage during this time15,19. While rodents are postnatal brain developers, and sheep and non-human primates are prenatal brain developers, humans and pigs, being perinatal brain developers, both experience a brain growth spurt near the time of birth14.

Indeed, the pig has long been considered a highly valuable animal model for biomedical research20–23. Pigs have frequently been used for research in toxicology24,25, diabetes26,27, imaging28,29, and cardiovascular disease30,31. For more than a decade, pigs have also been gaining popularity as a desirable large animal model for

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neuroscience, cognitive and behavioral research32–38. Agricultural bred pigs are most commonly used in research due to their wide availability and relative low price. However, these pigs can be particularly challenging to work with as they grow quickly and body weight at maturity can easily reach 300 kg32.

Recently, laboratory purpose-bred miniature pigs (minipigs) are being used more frequently in research studies. Breeds such as the Yucatan and Hanford pigs reach an adult body weight of 70–90 kg, and micropig breeds such as the Gottingen and Sinclair pigs reach an adult weight of 35–55 kg32. The compact size and the docile temperament of these minipigs make them an attractive model for brain research.

It is conceivable that porcine cognitive and motor performance is dynamic and changes with age, thus it remains necessary to comprehensively characterize minipig behavior over time. We have conducted a battery of behavioral and physiological tests in Yucatan minipigs using established behavioral testing methods, as well as state-of-the-art wearable sensors and artificial intelligence (AI) methods to analyze behavior. We tested their neuropsychological parameters including executive functions, processing speed, anxiety and learning and memory. We also tested motor and physiological characteristics including gait analysis, circadian rhythms, and daily activity levels. We describe their cognitive, sensorimotor, and physiological function from adolescence (4 months old) through maturity (7 months old) 39. This multimodal understanding of healthy Yucatan minipig behavior and how their behavior is dynamic throughout development is essential for identifying behavioral and physiological changes induced by brain injury and disease and to test the efficacy of potential therapeutic strategies.

Results

Neuropsychological screening for executive function, anxiety, willingness to explore a new environment and locomotion were performed using the open field test40. The open field test has been used in many animal species, including rodents41–43, cows44, sheep45, as well as pigs46–48. In this study, pigs were placed in a 1.83 m × 1.83 m open chamber and their activity was recorded using overhead cameras for 10 min, once per week. We used deep-learning artificial intelligence (AI) software, DeepLabCut49,50, to track pig locomotor activity in the open field arena. Figure 1 shows the distance travelled by pigs (n = 4) in the open field from 18 weeks of age to 36 weeks of age. The results show that on average the pigs walked 217 ± 20 m, but between the age of 27–29 weeks old there was a marked increase in walking (396 ± 19 m). RStudio analysis of coordinate data extracted from DeepLabCut allowed us to calculate the spatial distribution of the pigs’ movement and we found that pigs spend much of their time near the door/entrance to the arena. Figure 1 shows the distance travelled by pigs (n = 4) in the open field from 18 weeks of age to 36 weeks of age. The results show that on average the pigs walked 217 ± 20 m, but between the age of 27–29 weeks old there was a marked increase in walking (396 ± 19 m). RStudio analysis of coordinate data extracted from DeepLabCut allowed us to calculate the spatial distribution of the pigs’ movement and we found that pigs spend much of their time near the door/entrance to the arena. Figure 2 characterizes attempts by the pigs to escape the arena defined as any behavior where the pigs pushed on the walls or door of the chamber. Figure 2a illustrates the number of individual events where pigs would exhibit this behavior, whereas 2b shows the cumulative seconds pigs spent engaged in escape behaviors. Figure 3 shows heatmap representations of pig location within the chamber. Figure 3a is a composite heatmap of all 4 pigs during the open field test across all weeks overlaid with an overhead still image of a pig during the test period. Our results confirm our observations that pigs spend much of the test period in the area immediately surrounding the door (within 30 cm). Further analysis shows that pigs’ spatial distribution differs with age. Figure 3b is a heatmap representation of one pig at 18 weeks of age, whereas 3c shows the heatmap for the same pig at 36 weeks of age. When pigs are younger, they are less inclined to explore the arena. These data are indicative that young pigs may be more anxious than adults.

Healthy pigs exhibit an affinity for the region of the arena closest to the door. This is consistent with findings showing that pigs spend much of their time in the arena attempting to escape.

Measurements of learning and memory, anxiety and depression were performed using the novel object recognition test51,52. The novel object task takes place in two parts: the habituation phase and the test phase. Briefly, pigs (n = 4) were brought to the behavior chamber and presented with two identical objects during the habituation phase. Pigs were individually allowed to explore these objects for 10 min. Each pig was returned to the housing room and after a 15-min inter-trial interval, the pig was brought back to the behavioral chamber for

Figure 1. Locomotor activity in the open field. The total average distance minipigs traveled in the open field arena. Data show mean and SEM (n = 4). A Pearson correlation analysis between age and distance traveled yields an r of 0.149 and a p value of 0.567.
the test phase. The pig was presented with one of the objects from the habituation phase (familiar object), and one object they had never seen before (novel object). The quantity and duration of contacts with both the familiar and novel objects were recorded. The test took place once per week from 19 to 36 weeks. Figure 4 shows results from the test phase of the novel object recognition task. Figure 4a illustrates the cumulative duration of contact with the familiar and novel objects. Throughout development, pigs display a significant preference for exploring an unknown, new object. Statistical analysis shows that an unpaired t-test between contacts with the novel object and familiar object yields a p value less than 0.0001. These results indicate that the difference between contacts with the objects is not the result of chance. 4b shows the percent of time the pigs interacted with the novel and
familiar objects out of the total time pigs interacted with objects. These results further confirm that healthy pigs prefer interacting with the novel object.

We have developed a novel test for assessing executive function, processing speed and spatial learning and memory which we term the baited ball pit. This test was inspired by the cognitive hole-board test and is designed to utilize the natural rooting behavior of the pig. In this test, the pig is required to find six apple slices that are hidden in the ball pit. The slices are always hidden at the same location and the time it takes the pig to retrieve each of the six slices is determined. We hypothesize that this test can be used as a measure of situational memory and contextual conditioning. Figure 5 demonstrates that throughout development the pig became increasingly faster at identifying the hidden objects. On the first week that the test was administered at age 19 weeks, the pigs completed it within 26.5 ± 0.23 s, but increased their speed at finding the object by 100% when they reached 36 weeks old (7.8 ± 0.09 s; n = 4).

The pigs’ natural circadian rhythms were assessed in 5-month-old pigs. We combined results from activity tracker and night-vision video recording to determine the pigs wake and sleep cycles. Figure 6 shows that pigs have consistent sleep cycles, waking around 7 am and sleeping around 11 pm, sleeping for an average of 8.7 h (± 0.2 h). We also acquired high resolution kinematics and gait analysis. Three-dimensional (3D) kinematic data has the ability to capture unique parameters that are unable to be identified through video alone. Additionally, location and movements of anatomical points can be quantified with high levels of accuracy through a robust calibration and use of a multi-camera system. Thus, for this study, motion capture was used to obtain initial 3D
movement profiles of a pig during a walking trial. As proof of concept, initial kinematic data were calculated for a single pig. The camera space permitted capture of one full gait cycle. One gait cycle is defined as the following touch point in sequence: left hind limb to left front limb followed by right hind limb touch and right front limb.

Three rigid marker pods were placed on the spine of the pig. Each pod consisted of three reflective markers with one located on the head (covering the base of the skull to the second cervical vertebra), a second on the shoulders (directly over the third thoracic vertebrae), and a third on the rear (directly over the sixth lumbar vertebrae). Additionally, four single markers were placed on the front and hind limbs distal to the condyle of the humerus and distal to the lateral condyle of the femur, respectively.

Initial kinematic analyses of the pig gait trials were performed. This initial analysis included the average linear velocity, along with angle data. The angle data were rotations computed about three axes that were created at each pod. For example, an axis perpendicular to the base of the pod was used to define left/right rotation of the head. An anterior axis along the base of the pod was developed and rotation about that axis was left/right lateral tilt and finally a medial/lateral axis was used to define the up/down—or “nodding” motion. These same three axes were computed on the other two pods as well. Figure 8 shows the marker system used on the pigs (left) and the local coordinate systems created at each pod along with the leg markers (right).

Data were analyzed of each pod with respect to the fixed reference system (i.e. global coordinate system). The average linear velocity of this pig was 1.93 m/s. Data from this pig showed lateral tilt ranges around 12°. Additionally, the head flexion–extension or “nodding” had a range of approximately 17°. Left to right rotation of the head ranged for this trial showed a 30° movement to one side as the pig moved its head in an attempt to turn. Similar motions were obtained for each of the marker pods located on the shoulders and buttocks regions.

Discussion

The minipig possess many physiological and anatomical attributes that are similar to humans, making it an attractive animal model for biomedical research and an emerging translational model in neurology and neuroscience. The ability to produce in the minipig brain an injury of similar nature to humans, and the opportunity to acquire neuropsychological and motor performance measurements, make this animal model invaluable in the field of neurology and rehabilitation. It is of particular interest to understand how different features of the behavior change throughout age. In adults, recovery after brain injury is considered when performance returns to pre-morbid levels, but in children and adolescents this is more complex. During the time that it may require for children who suffered brain injury to return to pre-morbid levels, other children may have reached new

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Figure 6. Activity tracking/Fitbit step counting. 12-h (8:00 am–8:00 pm) graph showing steps recorded by an activity tracker (Fitbit) on pigs averaged over 5 days, broken into 15-min bins. Pigs were most active in the middle of the day, between noon and 4 pm.

Figure 7. Sleep and circadian rhythms. Sleep–wake graph showing that pigs tend to fall asleep between 10 pm and 12 am and tend to wake around 7 am. Pigs slept for an average of 8.7 h per night.
milestones and the critical period of acquiring or improving these abilities may have passed, leading to long term impairments. Therefore, characterizing the dynamic of cognitive, sensorimotor, and physiological functions in the minipig model is critical for translational research.

We have used a battery of neuropsychological tests to evaluate processing speed, learning and memory, and anxiety in the minipigs. These tests are particularly relevant for assessing the severity of brain injury in animal models and are highly translational in human research. Similar to humans, the minipigs show increased learning capabilities, long-term memory, and increased self-confidence with age.

The physiological measures are also relevant for neurology and rehabilitation research. Brain injury is often accompanied by reduced quality of sleep and sleep disturbances. The results demonstrate that a combination of activity tracker and video recording can provide valuable data regarding these aspects of behavior.

We have used AI to analyze 400 min of video recording, which would have taken as much time, or more, to be manually analyzed. Machine learning and AI methods are becoming increasingly useful in behavioral research and provide new opportunity to analyze large sets of data with high temporal and spatial resolution and in less time than it would have taken by a manual user.

This work also demonstrates the initial data sets possible with a 3D kinematic gait analysis. In the future we will take rotations of each pod location (head, shoulders, rear) and compare the motions between each whereas the above data are the movement of the pods with respect to a global system. Future work will use this approach to quantify and compare changes in movement pre and post trauma within and across pigs. We will gather data on several healthy pigs to obtain a profile of healthy motions. Additionally, pigs with injuries or diseases could be tested to identify how gait parameters are affected by a specific disease or injury.

While there exists an ample amount of literature describing pig anatomy and physiology, studies investigating the complexity of pig psychology and behavior are less abundant. Much of this scarcity can likely be attributed to the numerous challenges associated with the use of large animals in research. Due to handling concerns, agriculture pigs are typically used only for acute studies, with the age cap at 5–6 weeks of age. This tends to prevent long-term behavioral studies from being done.

This study extends the toolbox of behavioral and physiological tests in the Yucatan minipig. These comprehensive measurements of different aspects of behavior can be useful to measure performance after major and mild injuries and may be sensitive to subtle changes in performance that are often difficult to diagnose clinically.

Methods

Animals. All experimental protocols have been approved by the Michigan State University Institutional Animal Care and Use Committee. All experiments were conducted in compliance with guidelines set by the American Veterinary Medical Association.

Four Yucatan minipigs, 2 female 2 male, were used in this study (Premier BioSource, CA). Behavioral testing began when pigs were 4-months of age. Pigs were pair housed in an enriched environment, fed nutritionally complete feed twice per day, with unrestricted access to water on a 12-h (7:00–19:00) light cycle.

Behavioral chamber. Behavioral experiments, with the exception of gait analysis, took place in an open chamber measuring 1.83 m × 1.83 m. The walls of the chamber were made of commercially available PVC board and measured 1 m in height. A bullet camera (Omron Sentech) was suspended overhead to record locomotion. The concrete floor of the chamber was hosed down between subjects.

Behavioral experiments. Open Field: Pigs were individually led to the behavioral chamber. Each pig was allowed to freely explore the chamber for 10 min and locomotor activity was recorded by the camera suspended overhead. This test was performed once per week.

Motion tracking was conducted using DeepLabCut software. Software was trained to recognize the different body parts of the pig in various poses. This training set was used to analyze all open field videos, and coordinate data for various body parts was extracted. Coordinate data was imported to RStudio where custom-written code was used to calculate the distance traveled by body parts of interest (back of head reported here). RStudio was also used to generate heat maps allowing us to better understand where the pig spends most of their time in the chamber.
Novel Object: Pigs were individually led to the behavioral chamber. Pigs were first exposed to two identical toys and all activity was recorded by the camera suspended overhead. This initial habituation phase lasted 10 min. Each pig was then returned to the housing room for a 15-min inter-trial interval. Following this interval, the pig was again led to the behavioral chamber and presented with one familiar object from the habituation phase and one new/novel object that had not been seen before. The test phase lasted 10 min, after which the pig was returned to the housing room. Times at which pigs were engaged with either the novel or familiar object were recorded. Cumulative time (in seconds) spent with each object is reported here.

Ball Pit: Prior to testing, handlers placed a plastic wading pool (91.44 × 91.44 × 17.53 cm) in the behavioral chamber and filled it with colorful plastic balls (5.59 cm diameter). Six slices of apple were buried in a pentagonal shape around the perimeter of the pool with one apple slice in the center. Pigs were then individually led to the behavioral chamber. Cameras, both overhead and handheld, recorded as the pigs searched for and retrieved apple slices. The duration of this test was less than 5 min.

Analysis was conducted by manually watching the videos and tracking the timestamp at which each apple slice was retrieved. The latency between successful retrievals was calculated, and a mean latency for each trial was determined.

Gait Analysis: Pigs were trained to tolerate wearing commercially available miniature swine harnesses and to walk on a lead with the handler. Pigs were outfitted with coban wrap around each leg, on the head, and around the body both posterior to the front limbs and anterior to the back limbs. Reflective markers were attached to the coban along the spine of the pig in the direction of movement (x-axis); one on the head covering the base of the skull and first cervical vertebrae, one on the shoulders over the third thoracic vertebrae, and one on the rear over the sixth lumbar vertebrae. Markers were also placed on each of the legs. The test setup consisted of a 6-meter-long runway with 6 motion capture cameras (Qualisys, Sweden) with 3 cameras positioned on each side.

The Qualisys motion capture system exports .mat files with three-dimensional positions of each reflective marker and rotational as well as positional data of marker pods in a global coordinate system. Files were imported to MATLAB and positional data were used to create local coordinate systems on each marker pod with origins at their respective geometric centers. Ranges of head, trunk, and rear tilt/rotation were determined by the vectors from the local pods with respect to the global coordinate system. Position data of the markers on the legs were used to identify a gait cycle.

Activity Tracking: Fitness tracking devices (Fitbit®) were attached to miniature swine harnesses. Pigs wore harnesses and fitness trackers for several hours per week. Step data was extracted from the Fitbit® app.

Circadian Rhythms: Wide angle cameras (Amazon Cloud Cam©, WYZE Cam©) were used to record pigs in the housing room 24/7. Times that the pigs wake in the morning and fall asleep in the evening were recorded.

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References

1. Taylor, C. A., Bell, J. M., Breiding, M. J. & Xu, L. Traumatic brain injury-related emergency department visits, hospitalizations, and deaths—United States, 2007 and 2013. MMWR Surveill. Summ. 66(9), 1–16. https://doi.org/10.15585/mmwr.ss6609a1 (2017).
2. Kochanek, P. M., Wallisch, J. S., Bayır, H. & Clark, R. S. Pre-clinical models in pediatric traumatic brain injury—challenges and lessons learned. Childs Nerv. Syst. 33(10), 1693–1701 (2017).
3. Oser, N. D. & Dixon, C. E. The controlled cortical impact model: applications, considerations for researchers, and future directions. Front. Neurol. 7, 134. https://doi.org/10.3389/fneur.2016.00134 (2016).
4. Schreiten, D. J. & Shapiro, A. M. A quantitative review of the effects of traumatic brain injury on cognitive functioning. Int. Rev. Psychiatry 15(4), 341–349 (2003).
5. Conrad, M. S., Dilger, R. N. & Johnson, R. W. Brain Growth of the Domestic Pig (Sus scrofa) from 2 to 24 weeks of age: a longitudinal MRI study. Dev. Neurosci. 34(4), 291–298. https://doi.org/10.1159/000339311 (2012).
6. Dai, J. X., Ma, Y. B., Le, N. Y., Cao, J. & Wang, Y. Large animal models of traumatic brain injury. Neurosci. Lett. 607, 1–6. https://doi.org/10.1016/j.neulet.2015.11.029 (2016).
7. Dai, J. X., Ma, Y. B., Le, N. Y., Cao, J. & Wang, Y. Large animal models of traumatic brain injury. Neurosci. Lett. 607, 1–6. https://doi.org/10.1016/j.neulet.2015.11.029 (2016).
8. Duhaime, A. C. Large animal models of traumatic injury to the immature brain. Dev. Neurosci. 28(4–5), 380–387. https://doi.org/10.1159/000094164 (2006).
9. Margulies, S. S. et al. Establishing a clinically relevant large animal model platform for TBI therapy development: using cyclosporin A as a case study. Brain Pathol. 25(3), 289–303. https://doi.org/10.1111/bpa.12247 (2015).
10. Sorby-Adams, A. J., Vink, R. & Turner, R. J. Large animal models of stroke and traumatic brain injury as translational tools. Am. J. Physiol. Regul. Integr. Comp. Physiol. 315(2), R165–R190. https://doi.org/10.1152/ajpregu.00163.2017 (2018).
11. Gieling, E. T., Schuurman, T., Nordquist, R. E. & van der Staay, F. J. The pig as a model animal for studying cognition and neurobehavioral disorders. Curr. Top. Behav. Neurosci. 7, 359–383. https://doi.org/10.1007/7854_2010_112 (2011).
12. Brezulcz, J., Swafford, B. & Thompson, W. Study of motor cortex of domestic pigs. Am. J. Vet. Res. 27(120), 1369–2000 (1966).
13. Craner, S. L. & Ray, R. H. Somatosensory cortex of the neonatal pig. I. Topographic organization of the primary somatosensory cortex (SI). The J. Comp. Neurol. 306(1), 24–38. https://doi.org/10.1002/cne.903060103 (1991).
14. Dobbing, J. & Sands, J. Comparative aspects of the brain growth spurt. Early Hum. Dev. 3(1), 79–83 (1979).
15. Davison, A. N. Biochemical Correlates of Brain Structure and Function (Academic Press, 1977).
16. Fairbairn, L., Kapetanovic, R., Sester, D. P. & Hume, D. A. The mononuclear phagocyte system of the pig as a model for understanding human innate immunity and disease. J. Leukoc. Biol. 80(6), 855–871 (2011).
17. Dawson, H. D., Chen, C., Gaynor, B., Shao, J. & Urban, J. F. The porcine translational research database: a manually curated, genomics and proteomics-based research resource. BMC Genomics 18(1), 643. https://doi.org/10.1186/s12864-017-4009-7 (2017).
18. Wolford, K. L. et al. Rapid neuroinflammatory response localized to injured neurons after diffuse traumatic brain injury in swine. Exp. Neurol. 299, 85–94 (2017).
19. Finnie, J. W. Comparative approach to understanding traumatic injury in the immature, postnatal brain of domestic animals. Aust. Vet. J. 90(8), 301–307. https://doi.org/10.1111/1751-0813.2012.00955.x (2012).
20. Landy, J. J., Crowdon, J. H. & Sandberg, R. L. Use of large, germfree animals in medical research. JAMA 178(11), 1084–1087 (1961).
21. Bustad, L. K. & McClellan, R. O. Swine in biomedical research. Science 152(3728), 1526–1530 (1966).
22. Douglas, W. R. Of pigs and men and research. Space Life Sci. 3(3), 226–234 (1972).
23. Tumbleson, M. E. editor. Swine in Biomedical Research. Conference on Swine in Biomedical Research. University of Missouri–Columbia (USA), Plenum Press (1985, 1986).
24. Swindle, M. M., Makin, A., Herron, A. J., Clwb, F. J. & Frazier, K. S. Swine as models in biomedical research and toxicology testing. Vet. Pathol. 49(2), 344–356. https://doi.org/10.1177/0300983811402886 (2012).
25. Schneider, N. R., Bradley, S. L. & Andersen, M. E. Toxicology of cyclotrimethylenetrinitramine distribution and metabolism in the rat and the miniature swine. Toxicol. Appl. Pharmacol. 39(3), 531–541 (1977).
26. Phillips, R., Panepinto, L., Spangler, R. & Westmoreland, N. Yucatan miniature swine as a model for the study of human diabetes mellitus. Diabetes 31(Supplement 1), 30–36 (1982).
27. Wolf, E., Braun-Reichhart, C., Streckel, E. & Renner, S. Genetically engineered pig models for diabetes research. Transgenic Res. 23(1–2), 27–38 (2014).
28. Boettcher, A. N. et al. Human ovarian cancer tumor formation in severe combined immunodeficient (SCID) pigs. Front. Oncol. 9, 9 (2019).
29. Spuentrup, E. et al. Molecular magnetic resonance imaging of atrial clots in a swine model. Circulation 112(3), 396–399 (2005).
30. Hughes, H. Swine in cardiovascular research. Lab. Anim. Sci. 36(4), 348–350 (1986).
31. Shimokawa, H. et al. Coronary artery spasm induced in athersclerosis miniature swine. Science 221(4610), 560–562 (1983).
32. Lind, N. M. et al. The use of pigs in neuroscience: modeling brain disorders. Neurosci. Biobehav. Rev. 31(5), 728–751 (2007).
33. Gieling, E. T., Nordquist, R. E. & van der Staay, F. J. Assessing learning and memory in pigs. Anim. Cogn. 14(2), 151–173. https://doi.org/10.1007/s10071-010-0364-3 (2011).
34. Kornum, B. R., Thygesen, K. S., Nielsen, T. R., Knudsen, G. M. & Lind, N. M. The effect of the inter-phase delay interval in the spontaneous object recognition test for pigs. Behav. Brain Res. 181(2), 210–217. https://doi.org/10.1016/j.bbr.2007.04.007 (2007).
35. Glud, A. N. et al. Direct MRI-guided stereotaxic viral mediated gene transfer of alpha-synuclein in the Gottingen minipig CNS. Acta Neurobiol. Exp. (Wars). 71(4), 508–518 (2011).
36. Fang, M. et al. Myelination of the pig's brain: a correlated MRI and histological study. Neuroimage 14(3), 102–108. https://doi.org/10.101159/000086292 (2005).
37. Gould, T. D., Dao, D. T. & Kovacsics, C. E. The Open Field Test. Mood and Anxiety Related Phenotypes in Mice 1–20 (2009).
38. Hall, C. S. Emotional behavior in the rat. I. Defecation and urination as measures of individual differences in emotionality. J. Comp. Psychol. 18(3), 385 (1934).
39. Lu, H. et al. Transcranial magnetic stimulation facilitates neurorehabilitation after pediatric traumatic brain injury. Sci. Rep. 5, 14769. https://doi.org/10.1038/srep14769 (2015).
40. Cywiak, C. et al. Non-invasive neuromodulation using tEMS and the electromagnetic-perceptive gene (EPG) facilitates plasticity after nerve injury. Brain Stimul. 13(6), 1774–1783. https://doi.org/10.1016/j.brs.2020.10.006 (2020).
41. Rousay, A. & Le Neindre, P. Social influences on the reactivity of heifers: implications for learning abilities in operant conditioning. Appl. Anim. Behav. Sci. 25(1–2), 149–165 (1990).
42. Winfield, C., Syme, G. & Pearson, A. Effect of familiarity with each other and breed on the social behaviour of sheep in an open field. Appl. Anim. Ethol. 71(1), 67–75 (1981).
43. Donald, R. D., Healy, S. D., Lawrence, A. B. & Rutherford, K. M. Emotionality in growing pigs: is the open field a valid test?. Physiol. Behav. 104(5), 906–913 (2011).
44. Shea-Moore, M. The effect of genotype on behavior in segregated early-weaned pigs tested in an open field. J. Anim. Sci. 76(1), 100 (1998).
45. Belharz, R. & Cox, D. Social dominance in swine. Anim. Behav. 15(1), 117–122 (1967).
46. Mathis, A. et al. DeepLabCut: markerless pose estimation of user-defined body parts with deep learning. Nat. Neurosci. 21(9), 1281–1289 (2018).
47. Hunt, R. D. et al. Swimming direction of the glass catfish is responsive to magnetic stimulation. PLoS ONE 16(3), e0248141. https://doi.org/10.1371/journal.pone.0248141 (2021).
48. Haig, A., Chen, J.-Y. & O'Driscol, K. Variations in the behavior of Pigs during an open field and novel object test. Front. Vet. Sci. https://doi.org/10.3389/fvets.2020.00607 (2020).
49. Antonides, A., Schoonderwoerd, A. C., Nordquist, R. E. & van der Staay, F. J. Very low birth weight piglets show improved cognitive performance in the spatial cognitive holeboard task. Anim. Cogn. 18(3), 906–913 (2015).
50. Bhatti, Z. Gait analysis and biomechanics of quadruped motion for procedural animation and robotic simulation. Bahria Univ. J. Inf. Commun. Technol. (BUJICT) 10(2) (2017).
51. Duhaime, A. C. et al. Head injury in young children: mechanisms, injury types, and ophthalmologic findings in 100 hospitalized patients younger than 2 years of age. Pediatrics 90(2 Pt 1), 179–185 (1992).
52. Taylor, S. R., Smith, C., Harris, B. T., Costine, B. A. & Duhaime, A. C. Maturation-dependent response of neurogenesis after traumatic brain injury in children. J. Neurosurg. Pediatr. 12(6), 545–554. https://doi.org/10.3171/2013.8.PEDS13154 (2013).
53. Mangeot, S., Armstrong, K., Colvin, A. N., Yeates, K. O. & Taylor, H. G. Long-term executive function deficits in children with traumatic brain injuries: assessment using the behavior rating inventory of executive function (BRIEF). Child Neuropsychol. 18(4), 271–284. https://doi.org/10.1076/chin.8.4.271.13503 (2002).
54. Taylor, H. G. et al. A prospective study of short- and long-term outcomes after traumatic brain injury in children: behavior and achievement. Neuropsychology 16(1), 15–27 (2002) (Epub 2002/02/21 PubMed PMID: 11853533).
55. Li, N. et al. Evidence for impaired plasticity after traumatic brain injury in the developing brain. J. Neurotrauma. 31(4), 395–403. https://doi.org/10.1089/neu.2013.3059 (2014).
56. Shin, S. S. et al. Transcranial magnetic stimulation and environmental enrichment enhances cortical excitability and functional outcomes after traumatic brain injury. Brain Stimul. https://doi.org/10.1016/j.brs.2018.07.050 (2018).
57. Baker, E. W. et al. Controlled cortical impact severity results in graded cellular, tissue, and functional responses in a piglet traumatic brain injury model. J. Neurotrauma. https://doi.org/10.1089/neu.2017.5551 (2018).
58. Friess, S. H. et al. Neurocritical care monitoring correlates with neurologic outcome in a swine model of pediatric traumatic brain injury. Neurosurgery 69(5), 1139–1147. https://doi.org/10.1227/NEU.0000003182288a1 (2011).
59. Karlsson, M. et al. Neuroprotective effects of cyclosporine in a porcine pre-clinical trial of focal traumatic brain injury. J. Neurotrauma. https://doi.org/10.1089/neu.2018.5706 (2018).
65. Kilbaugh, T. J. et al. Cyclosporin A preserves mitochondrial function after traumatic brain injury in the immature rat and piglet. *J. Neurotrauma.* 28(5), 763–774. https://doi.org/10.1089/neu.2010.1635 (2011).
66. Duhaime, A. C. et al. Magnetic resonance imaging studies of age-dependent responses to scaled focal brain injury in the piglet. *J. Neurosurg.* 99(3), 542–548. https://doi.org/10.3171/jns.2003.99.3.0542 (2003).
67. Barshikar, S. & Bell, K. R. Sleep disturbance after TBI. *Curr. Neurrol. Neurosci. Rep.* 17(11), 1–7 (2017).
68. Luther, M., Poppeart Cordts, K. M. & Williams, C. N. Sleep disturbances after pediatric traumatic brain injury: a systematic review of prevalence, risk factors, and association with recovery. *Sleep* 43(10), zsaa083 (2020).
69. Tham, S. W. et al. The longitudinal course, risk factors, and impact of sleep disturbances in children with traumatic brain injury. *J. Neurotrauma* 29(1), 154–161 (2012).
70. Mathias, J. & Alvaro, P. Prevalence of sleep disturbances, disorders, and problems following traumatic brain injury: a meta-analysis. *Sleep Med.* 13(7), 898–905 (2012).
71. Rutherford, K. M., Donald, R. D., Lawrence, A. B. & Wemelsfelder, F. Qualitative behavioural assessment of emotionality in pigs. *Appl. Anim. Behav. Sci.* 139(3–4), 218–224 (2012).
72. Düpjan, S., Ramp, C., Kanitz, E., Tuchscherer, A. & Puppe, B. A design for studies on cognitive bias in the domestic pig. *J. Vet. Behav.* 8(6), 485–489 (2013).
73. Schlinger, H. D. Behavior analysis and behavioral neuroscience. *Front. Hum. Neurosci.* 9, 210 (2015).

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**Author contributions**
A.N. and G.P. designed all experiments and wrote the manuscript. A.N., R.H., and J.F. performed all experiments. N.A. and T.B. performed gait analysis experiments. A.N., R.H., K.M., A.C., A.V. and G.P. were involved in the discussions and preparation of the manuscript. All authors reviewed and approve of the manuscript.

**Competing interests**
The authors declare no competing interests.

**Additional information**
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