Recovery of thicket in a revegetated limestone mine

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The opencast extraction of limestone at the PPC Cement (Pty) Ltd Loerie quarry in the Eastern Cape, South Africa, has led to disturbance of the immediate environment. Large tracts of thicket have been removed during mining and exhausted quarry areas had to be revegetated. Revegetation commenced 16 years ago and as a result of progressive revegetation efforts since then, five seral stages were available for study. This presented an opportunity to describe the successional progression that occurs where thicket topsoil was used for revegetation. The quarry lies near the Gamtoos fault and both thicket and mountain fynbos is found in proximity to the mine. Before mining, the area was covered by thicket and quarry floors were revegetated by covering landscaped areas with thicket topsoil. The vegetation that developed in the quarry had a very low floristic relationship to fynbos. After 16 years, the vegetation also had a low similarity to thicket, even though the soils used for revegetation contained thicket propagules. The oldest seral stage (16 years) was a Rhus incisa–Panicum deustum thicket in which only 46% of the species were mature thicket species. In order to develop thicket more rapidly, active intervention will be required. This could take the form of planting of saplings of key thicket species. A trial planting of such saplings showed high survival success of most species. Long-term (50 year) monitoring will be required to determine whether this approach was successful.

Introduction

Plant succession is a directional cumulative change in the species that occupy a given area through time (Barbour et al. 1987). Typically successional processes (replacement of some species by others) lead to a climax condition that then remains relatively stable over a substantial period of time. If disturbance causes a regression to an earlier successional stage (sere) and the successional process then follows the same path it did before, a disturbed system should eventually achieve complete recovery (Cairns 1989). These ‘Clementsian’ ideas do not hold for restoration ecology since in revegetated areas, the species representing the earlier successional stages may no longer be present for re-invasion, and other species not present during the earlier developmental period, may be the post-disturbance colonisers.

Application of successional theory to derelict land has not been easy, as substrates do not always correspond to natural ones (Harris et al. 1996). It is difficult to make predictions on the succession of vegetation in highly disturbed environments. For instance, limestone quarries are not alike with respect to the environmental conditions that will direct successional pathways in plant growth. Many attempts at rehabilitating opencast mineral extraction stands have failed because practitioners rigidly followed the methods of past successes (Harris et al. 1996). This is especially true when European methods have been used in arid and semi-arid lands in southern Africa.

The opencast extraction of limestone at a quarry near Loerie, about 50km west of Port Elizabeth, has caused severe damage to the natural thicket and underlying soils. Entire hillsides have been cut away in order to access underlying blocks of limestone. The overlying material, or overburden was removed by ripping or blasting. After removal of the limestone, mined areas and hillsides were shaped and terraced with this overburden and then top dressed with a thin (100–200mm) layer of soil taken from thicket areas earmarked for new excavations. This revegetation effort commenced in the early 1980s. After consultation with the Department of Botany at the University of the Witwatersrand it was decided to leave these areas to revegetate through natural succession processes (Shaw and Black 1985). There was thus no irrigation or addition of fertiliser and seed to initiate revegetation in these areas. This method has been used in a number of areas within the quarry over the past 16 years.

If environmental factors such as aspect, slope and macroclimate are similar for all stands, comparison between stands of different age is possible. Further influences on the development of rehabilitated vegetation such as the origin and quality of the topsoil, the distance from an available seed source (MacMahon 1990) and the boundary shape of the rehabilitated site that affects the colonisation pattern (Hardt and Forman 1989) need to be considered.
All the revegetated stands at the Loerie quarry showed close similarity in these environmental factors and could thus be used as a chronosequence to study the process of succession in the natural return of indigenous vegetation on the stands. It was hoped that by using topsoil from previously vegetated areas that the natural vegetation would return through succession. Quarry management was under the impression that the natural vegetation of the area was beginning to recover at these stands and this study was initiated to determine if this was so.

It is generally believed that thicket will not recover after massive disturbance. While intact thicket has high stability and resilience (Cowling 1984, Everard 1987) evidence available to date (e.g. Stuart-Hill 1991, Moolman and Cowling 1994) indicates that it is likely that areas cleared of thicket will follow the non-equilibrium model of community dynamics (DeAngelis 1987) rather than develop through to a stable climax state comparable to the intact thicket found prior to disturbance. A number of ecologists have shown that there is little or no regeneration of thicket through the establishment of seedlings (Aucamp and Tainton 1984, Stuart-Hill and Danckwerds 1988, Stuart-Hill 1991, Moolman and Cowling 1994). It is believed that the climate has become drier over the past 6000 years or so and the lack of a moist and shaded environment inhibits seedling establishment (Stuart-Hill 1991). La Cock (1991) and Thrash (1998) suggest that regeneration through seedlings can only take place under the canopy of certain nurse plant species that provide a suitable microclimate for their establishment. La Cock (1991) believed that the distribution of seedlings at the quarry followed no discernible pattern of seedings that is responsible for the failure of the shrub component of thicket to re-colonise poorly vegetated areas rather than the lack of germination and consequent establishment of seedlings.

The use of a chronosequence to study succession has been used extensively in the regeneration of dune thicket after mining by Richards Bay Minerals (Cairns 1989, Moll 1992, Van Aarde and Smit 1997). These studies have been invaluable in the information base that they have provided to the mining company. The advantage of having different aged stands to compare is that predictions can be made as to the development and management of future revegetation trials (Bell 1987, Bradshaw 1990). Key species and successional processes can be identified and manipulated at an early stage thereby speeding up the process of revegetation and reducing costs for the mining company. Cairns (1989) feels that the opportunity offered by rehabilitated areas for the study of ecological processes and functioning cannot be overemphasised.

There is a paucity of succession studies in thicket, probably because of the common belief that the system will not regenerate. Few, if any studies have investigated the regeneration of thicket over a long time period. The primary aim of this study was to assess whether thicket is returning to the revegetated areas through succession in the Loerie quarry. The success of the revegetation programme is assessed and predictions are made as to the climax vegetation expected for the quarry.

Materials and Methods

Study site

Loerie has a temperate climate. Daily summer temperatures for Patensie, a small town 30km from Loerie, range from 16–30°C (means calculated from daily temperatures supplied by the South African Weather Bureau for 1993–1999). Winter temperatures range from 8–21°C. The average annual rainfall is 730mm per year. The area experiences a bimodal rainfall pattern with peaks in the autumn months (March–May), receiving an average of 63mm per month, and in the spring months (September–November) an average of 73mm per month. There are very few incidences of hail on record and virtually no frost.

The natural thicket surrounding the Loerie quarry showed similarities to the Pterocelastrus–Euclea community of Kaffrarian Thicket as well as to the Euclae undulata–Brachylaena ilicifolia community of Kaffrarian Succulent Thicket described by Cowling (1984). The vegetation similarly can be related to Xeric Kaffrarian Thicket or Valley Thicket (Lubke 1996) and Mesic Succulent Thicket as described by Everard (1987). Species such as Aloe africana, Carissa haematocarpa, Euclea undulata var. undulata, Grewia occidentalis, Scutia myrtina and Sideroxylon inerme subsp. inerme found in thicket of the study area, are also listed as dominant in the communities described by these two authors (Cowling 1984, Everard 1987). This vegetation forms a closed low forest with a high proportion of trees forming the canopy between 4m and 6m in height (Everard 1987). The vegetation has a low succulent component and is dominated by a woody canopy with species of strong Afromontane affinity (Cowling 1984). The thicket of the study area is more open within the canopy than Mesic Succulent Thicket as there are fewer spinescent shrubs, woody creepers and succulent species (Everard 1987).

Elements from both Mesic Succulent Thicket (of the lower Gamtoos Estuary) and Xeric Kaffrarian Thicket (of the upper Gamtoos Valley) (Everard 1987) occur at Loerie quarry. This accounts for the high beta diversity in intact thicket at the quarry. Aspect and geology separates thicket types at Loerie. Mesic Succulent Thicket occurs on the upper and drier north facing slopes of valleys and Valley Thicket in pockets in the deeper valleys and on moist, south facing slopes. This gradient has important implications with regards to the succession of the revegetated stands. All revegetation stands are north facing in aspect and have a shallow soil profile that leads to a drier environment similar to that where Mesic Succulent Thicket occurs. It would thus be expected that Mesic Succulent Thicket would revegetate at these stands as opposed to Valley Thicket that requires a deep soil profile, a more shaded southerly aspect and a relatively moist environment.

Vegetation assessment

Quadrats were sampled in revegetated areas of one year, two years, six years, 10 years and 16 years old, intact thick- et and fynbos. Plant species were recorded with a measure of their canopy cover. Plots were 100m² for revegetated
areas (Carey and Brooks 1985, Camp and Weisser 1991, Moll 1992), 10m² for fynbos (Cowling 1984) and 40m² for thicket (a belt transect of 20m x 2m wide). Belt transects were selected as the appropriate sampling method in thicket as they provide a less subjective assessment of plant cover than that of a large quadrat (Cowling 1984, Everard 1987, Louw 1993). Access and manoeuvrability within the thicket is also extremely difficult and thus belt transects are an easier way of effectively assessing the composition of the vegetation.

Comparison to fynbos

Ordination (DCA) of the samples for the revegetated stands shows that thicket and fynbos separate with a group between them that contained the revegetated stands (Figure 1). The fynbos samples plotted separately from the other two groups on the right hand side of the diagram indicating a high degree of dissimilarity from thicket and revegetated samples. The ordination points of revegetated samples lie closer to those of thicket than those of fynbos.

The species ordination of the same analysis showed complete separation of fynbos species from the rest (data not shown). Members of the Proteaceae (Leucadendron salignum, Protea exima), Restionaceae (Hypodiscus striatus) and Ericaceae (Erica chamissonis var. chamissonis, Erica sessiliflora var. sessiliflora) were dominant in the fynbos community.

Classification (TWINSpan) of species (Figure 3) also showed high dissimilarity between fynbos and the other communities (an eigenvalue of 0.8).

Comparison to thicket

Thicket and revegetated stand species showed some similarity in that they split at the second division level at an eigenvalue of around 0.3 (Figure 3). However, separation between

![Figure 1: Ordination (DCA) of samples of revegetated stands (circles), thicket (squares) and fynbos (asterisks). Cumulative % variance of species data = 18.6%; sum of unconstrained eigenvalues = 7.178](image-url)

Sideroxylon inerme subsp. inerme and Tarchonanthus camphoratus were planted in groups of three. Survival of these saplings was monitored.

Results
| Species name                        | Code   | 1-year | 2-year | 6-year | 10-year | 16-year | Fynbos | Thicket |
|------------------------------------|--------|--------|--------|--------|---------|---------|--------|---------|
| Abutilon sonneratianum (Cav.) Sweet | Abutsonn | 1      | 2      | 6      | 10      |         |        |         |
| *Acacia cyclops A. Cunn ex G. Don* | Acaccycl | 1      | 2      | 6      | 10      | 16      |        |         |
| Acacia karroo Hayne               | Acackrr | 2      | 6      | 10     | 16      | T       |        |         |
| *Agrostis montevidensis* Spreng. ex Nees | Agrormont | 6      |        |        |         |         |        |         |
| Aizoone globoides L. f.           | Aizogolin | 1      | 6      | 10     |         |         |        |         |
| Aizoone rigidum L. f. var. rigidum | Aizorigi | 1      | 2      |        |         |         |        |         |
| Aloe africana Mill.               | Aloeafri |        |        |        |         |         | F      | T       |
| Aloe ferox Mill.                  | Aloefero |        |        |        |         |         | T      |         |
| Aloe pluridens Haw.               | Aloeplur |        |        |        |         |         | T      |         |
| Anthospermum aethiopicum L.       | Anthaeth |        |        |        |         |         | F      |         |
| *Argemone mexicana L.*            | Argemexi | 1      |        |        |         |         |        |         |
| Asclepias physocarpa (E. Mey.) Schltr | Asclphys | 1      | 2      | 6      |         |         |        |         |
| Astephanus marginatus Decne.      | Astemarg |        |        |        |         |         |        |         |
| Athanasia dentata (L.) L.         | Alhadent |        |        |        |         |         |        | F       |
| Atriplex semibaccata R. Br. var. appendiculata Aell. | Atrisemi | 1      | 2      | 6      | 10     |         |        |         |
| Azima tetracantha Lam.            | Azimtetr | 2      |        |        |         |         | T      |         |
| Berkheya rigida (Thunb.) H. Bol. & Wolley-Dod ex Adamson & Salter | Berkrgi | 6      | 10     | 16     |         |         |        |         |
| *Bidens pilosa L.*                | Bidepilo | 1      | 2      |        |         |         |        |         |
| Brachylaena ilicifolia (Lam.) Phill. & Schweick | Braclic |        |        |        |         |         | T      |         |
| *Bromus diandrus Roth*            | Bromdian |        |        |        | 16      |         |        |         |
| Bulbostylis hispidula (Vahl) R. Haines | Bulbhis | 10     |        |        |         |         |        | T       |
| Calpurnia aurea (Ait.) Benth. subsp. aurea | Calpaure | 10      | 16     |         |         |         |        |         |
| Capparis sepia L. var. citrifolia (Lam.) Tölken | Cappsespi |        |        |        |         |         | T      |         |
| Carissa haematocarpa (Eckl.) A. DC. | Carinhaem | 16     |        |         |         |         |        |         |
| Caroprous edulius (L.) L. Bol.    | Carpedul | 2      |        | 10     |         |         |        |         |
| Cassine aethiopica Thunb.         | Cassaeth |        |        |        |         |         | T      |         |
| Cassine peragua L.                | Casspera |        |        |        |         |         | T      |         |
| Cassine tetragona (L.) Loes.      | Cassstetr |        |        |        |         |         | T      |         |
| Celtis africana Burm. f.          | Celtafri |        |        |        |         |         | T      |         |
| Chamaeaesce inaequilatera (Sind.) Sojak | Chaminae | 1      | 2      | 6      |         |         |        |         |
| Cheilanthes viridis (Forssk.) Swartz var. viridis | Cheiviri |        |        |        |         |         | T      |         |
| Chironia baccifera L.             | Chirbacc |        |        |        |         |         | T      |         |
| Chrysanthemoides monilifera (L.) T. Norl. | Chrymone | 2      | 6      | 10     | 16     |         |        |         |
| Chrysosoma ciliata L.             | Chrycili | 2      | 6      | 10     |         |         |        |         |
| *Ciclospermum leptophyllum* (Pers.) Eichler | Ciclept | 1      | 2      | 6      |         |         |        |         |
| *Cirsium vulgar* (Savi) Ten.       | Cirsvulug | 1      | 2      | 6      | 10     |         |        |         |
| Cliftonia ilicifolia L. var. ilicifolia | Cliflic |        |        |        |         |         | F      |         |
| Clutia daphnoides Lam.            | Clutdaph | 6      | 10     | 16     | 16     |         |        |         |
| *Coryza albida Spreng.*           | Coryalbi | 2      | 6      |         |         |         |        |         |
| Conyza acabrida DC.               | Conyascab | 2      | 6      | 10     |         |         |        |         |
| Crassula ovata (Mill.) Druce      | Crassovat |        |        |        |         |         | T      |         |
| Crassula tetragona L. subsp. tetragona | Crasstetr |        |        |        |         |         | T      |         |
| Cussonia spicata Thunb.           | Cussspic |        |        |        |         |         | T      |         |
| Cynanchum ellipticum (Harv.) R.A. Dyer | Cynaeli | 2      |        | 16     |         |         | T      |         |
| Cynodon dactylon (L.) Pers.       | Cynodact | 1      | 2      | 6      | 10     |         |        |         |
| *Cynura stramonium L.*            | Datustra |        |        |        |         |         | T      |         |
| Dietes indoides (L.) Sweet ex Klatt | Dietrid |        |        |        |         |         | T      |         |
| Digitaria eriantha Steud.         | Digieria |        |        |        |         |         | F      |         |
| Diospyros dichophylla (Gand.) De Winter | Diosdich | 2      | 10     | 16     | 16     |         |        |         |
| Dipogon lignosus (L.) Verdc.      | Dipolign | 2      | 10     |         |         |         | T      |         |
| Doyyalis rotundifolia (Thunb.) Thunb. & Harv. | Dovyrotu |        |        |        |         |         | T      |         |
| Drostanthemum hispidum (L.) Schwant. var. platypetalum (Haw.) Schwant. | Droshisp | 2      | 6      | 10     |         |         |        |         |
| Eylopappus rhinocerots (L.f.) Less. | Elyrthinh | 6      | 10     | 16     |         |         |        |         |
| Eragrostis curvula (Schrad.) Nees | Eragcurv | 2      | 6      | 10     | 16     |         |        |         |
| Eragrostis obtusa Munro ex Fical. & Hiern | Eragobtu | 2      |        |        |         |         |        |         |
| Erica chamissonis Klotzch ex Benth. var. chamissonis | Ericahech |        |        |        |         |         | F      |         |
| Erica copiosa Wendl. var. copiosa | Ericcopi |        |        |        |         |         | F      |         |
| Erica sessiliflora L. f. var. sessiliflora | Ericaess |        |        |        |         |         | F      |         |
### Table 1 cont.

| Species name | Code | 1-year | 2-year | 6-year | 10-year | 16-year | Fynbos | Thicket |
|--------------|------|--------|--------|--------|---------|---------|--------|---------|
| Erica sp. 1 | Ericasp1 |        |        |        |         | F       |        |         |
| Erica sp. 2 | Ericasp2 |        |        |        |         | F       |        |         |
| Eriocephalus africanus L. | Eriofri | 6 | 10 | 16 | F | T |
| Euclea natalensis A. DC. subsp. capensis F. White | Euclnata | | | | | | T |
| Euclea undulata Thunb. var. undulata | Euclundu | | | | | | T |
| Euphorbia fimbriata Scop. | Euphfimb | | | | | | T |
| Euphorbia kraussiana Bernh. var. kraussiana | Euphkrau | 6 | 10 | | | | |
| Euphorbia triangularis Desf. | Euphtria | | | | | | T |
| Exomis microphylla (Thunb.) Aell. var. axyroides (Fenzl) Aell. | Exommicr | 2 | 6 | 10 | 16 | | |
| Felicia fasicularis | Felifasi | 2 | 10 | | | | |
| Gladiolus sp. | Gladisp | | | | | | F |
| *Gnaphalium pensylvanicum* Willd. | Gnappens | | 6 | | | | |
| Grewia occidentalis L. | Grewocci | 2 | 10 | | | | T |
| Grewia robusta Burch. | Grewrobu | | | | | | T |
| Gymnosporia buxifolia (L.) Szyszyl. | Maythete | 2 | 6 | 10 | | | |
| Helichrysum anomalum Lees. | Helianom | | | | | | F |
| Helichrysum cymosum (L.) D. Don subsp. cymosum | Helicymo | 2 | 6 | 10 | | | F |
| Helichrysum felinum Lees. | Helifeli | | | | | | F |
| Helichrysum nudifolium (L.) Lees. | Helinudi | | | | | | F |
| Helichrysum petiolare Hilliard & Burtt | Helipeti | | | | | | F |
| Helichrysum rosom (Berg.) Lees. var. rosom | Helirosu | 6 | 10 | 16 | | | T |
| Hermania althaoides Link | Hermalth | 2 | 6 | 10 | 16 | | T |
| Hermania flammaea Jacq. | Hermflam | 6 | 10 | 16 | | | F |
| Hermania mucronulata Turcz. | Hermmucr | 6 | 10 | | | | |
| Hippobromus pauciflorus (L. f.) Radlk. | Hipppauc | | | | | | T |
| Hypodiscus striatus (Kunth.) Mast. | Hypostri | 6 | | | | | |
| *Inula graveolens* Desf. | Inulgrav | | 2 | | | | |
| *Lantana camara* L. | Lantcama | 6 | | | | | |
| *Lantana rugosa* Thunb. | Lantrugo | 2 | 6 | 10 | 16 | | T |
| Ledebouria ensifolia (Eckl.) S. Venter | Ledeensi | | | | | | |
| Lepidium africanum (Burm. f.) DC. subsp. africanum | Lepiafri | 2 | | 10 | | | F |
| Leucadendron salignum Berg. | Leucalsi | | | | | | T |
| Lycium feroxissimum Miers | Lycifero | | | | | | T |
| Maerua cafra (DC.) | Maercafr | | | | | | T |
| Medicago polymorpha L. | Medipoly | 1 | 6 | | | | |
| Melhania didyma Eckl. & Zeyh. | Melhidy | 1 | 6 | 10 | 16 | | |
| Melica racemosa Thunb. | Melirace | 6 | | | | | |
| Merxmuelleria disticha (Nees) Conert | Merxdist | 6 | | 16 | | | |
| Mesembryanthemum aitonis Jacq. | Meseaito | 1 | 2 | | | | |
| Metalasia muricata (L.) D. Don | Metamuri | 6 | | | | | F |
| Mohria cafforum (L.) Desv. var. cafforum | Mohrcaff | | | | | | F |
| Myrphysium asparagusoides (L.) Wild. | Myrsapar | | | | | | T |
| *Nicotiana glauca* R.C. Grah. | Nicoglau | 1 | 2 | 6 | | | |
| Oedera genistifolia (L) Anderb. & Bremer | Oedegeni | 6 | | 16 | | | T |
| Olea europaea L. subsp. africana (Mill.) P.S. Green | Oleaeuro | 6 | 16 | | | | T |
| *Opuntia ficus-indica* (L.) Mill. | Opunficu | | | | | | T |
| Panicum deustum Thunb. | Panideus | 1 | 2 | 6 | 10 | 16 | | T |
| Panicum maximum Jacq. | Panimaxi | | | | | | T |
| Pappea capensis Eckl. & Zeyh. | Pappcape | | | | | | T |
| Passerina vulgaris Thoyad. | Passvulg | | | | | | F |
| Pavenia praemorsa (L. f.) Cav. | Pavpave | 10 | 16 | | | | T |
| Pelargonium peltatum (L.) L’ Hérit | Pelapel | | | | | | T |
| Pelargonium reniforme Curtis. | Pellarote | 6 | | | | | F |
| Pelargonium urosepalus (L.) L’ Hérit | Pelauros | | | | | | F |
| *Pennisetum clandestinum* Chiouv. | Pennclan | 1 | 2 | | | | |
| *Physalis angulata* L. | Physangu | 2 | 16 | | | | |
| *Picris echioideae* L. | Picrechi | 1 | 6 | | | | |
| Pittosporum viridiflorum Sims | Pittviri | | | | | | T |
| *Plantago lanceolata* L. | Pilanlac | 6 | | | | | T |
| Plumbago auriculata Lam. | Plumauri | | | | | | T |
| Polygala myrtifolia L. | Polymyr | 6 | 10 | 16 | | | T |
### Table 1 cont.

| Species name | Code     | 1-year | 2-year | 6-year | 10-year | 16-year | Fynbos | Thicket |
|--------------|----------|--------|--------|--------|---------|---------|--------|---------|
| *Prota*parasagus africanus (Lam.) Oberm. | Protafi | 2      | 10     |        |         |         |        |         |
| *Prota*parasagus crassicladus (Jessop) Oberm. | Protcras | 10     |        |        |         |         | T      |         |
| *Prota*parasagus densiflorus (Kunth.) Oberm. | Protdens |        |        |        |         |         | T      |         |
| *Prota*parasagus denudatus (Kunth.) Oberm. | Protdenu |        |        |        |         |         | T      |         |
| *Prota*parasagus racemosus (Willd.) Oberm. | Prottrace | 10     | 16     |        |         |         | T      |         |
| *Prota*parasagus setaceus (Kunth.) Oberm. | Protseta | 10     |        |        |         |         | T      |         |
| *Prota*parasagus suaveolens (Burch.) Oberm. | Protsuav | 10     |        |        |         |         | T      |         |
| *Prota*parasagus subulatus (Thunb.) Oberm. | Protsubu |        |        |        |         |         | T      |         |
| *Protea* exima (Salisb. ex Knight) Fourc. | Proteixim |        |        |        |         |         | F      |         |
| *Pseudognaphalium* luteo-album (L.) Hilliard & Burtt. | Pseulute | 2      |        |        |         |         |        |         |
| *Pt*aeoxylon obliquum (Thunb.) Radlk. | Ptaeobil |        |        |        |         |         | T      |         |
| *Pterocelastrus* tricuspidatus (Lam.) Sond. | Ptertric |        |        |        |         |         | T      |         |
| *Pteronia* incana (Burm.) DC. | Pterinca |        | 16     |        | F       |         | T      |         |
| *Putterfickia* pyracantha (L.) Szszyz. | Putfpyra |        |        |        |         |         | T      |         |
| *Rh*ioacarpus capensis (Harv.) A. DC. | Rhioicape |        |        |        |         |         | T      |         |
| *Rhoicissus* digitata (L. f.) Gilg & Brandt | Rhoidigi |        |        |        |         |         | T      |         |
| *Rhoicissus* tridentata (L. f.) Wild & Drum. | Rhoidrig |        |        |        |         |         | T      |         |
| subsp. tridentata | Rhotird |        |        |        |         |         | T      |         |
| *Rhus* crenata Thunb. | Rhuscren | 2      |        | F      |         |         |        |         |
| *Rhus* glauca Thunb. | Rhusglau |        |        | F      |         |         |        |         |
| *Rhus* incisa L. f. var. effusa (Presl.) R. Fernandes | Rhusinci | 2      | 10     | 16     |         |         | T      |         |
| *Rhus* laevigata L. | Rhuslave | 2      | 6      | 10     |         |         | T      |         |
| *Rhus* pterota Presl. | Rhuspter | 6      | 10     |         |         |         |        |         |
| *Rhus* undulata Jacq. | Rhusundu |        |        |        |         |         | T      |         |
| *Ricinus* communis L. | Ricicomm | 2      |        |        |         |         |        |         |
| *Salsola* kali L. | Salskali | 1      |        |        |         |         |        |         |
| *Salv*ia triangularis Thunb. | Salvtria | 6      | 10     | 16     |         |         | T      |         |
| *Sarc*ostemma viminal (L.) R. Br. | Sarvimi |        |        |        |         |         | T      |         |
| *Schkuhrijia* pinnata (Lam.) Cabr. | Schkpinn | 2      | 6      |        |         |         | T      |         |
| *Schotia* afra (L.) Thunb. var. afra | Schoafr |        |        |        |         |         | T      |         |
| *Schotia* latifolia Jacq. | Scholati | 2      |        |        |         |         | T      |         |
| *Sc*oloplia mundii (Eckl. & Zeyh.) Warb. | Scolmund |        |        |        |         |         | T      |         |
| *Sc*utia myrtina (Burm. f.) Kurz | Scutmryt | 16     |        |        |         |         | T      |         |
| *Selag*o corymbosa L. | Selacory | 6      | 10     | 16     |         |         | F      |         |
| *Sen*ecio ilicifolius L. | Senelic | 1      | 2      | 6      |         |         | T      |         |
| *Senecio* inaequidens DC. | Seneinae | 6      |        |        |         |         |        |         |
| *Senecio* linifolius L. | Senelini |        |        |        |         |         | F      |         |
| *Sid*a ternata L. f. | Sidatem | 2      | 6      | 10     |         |         | T      |         |
| *Sideroxylon inerme* L. subsp. inerme | Sideine |        |        |        |         |         | T      |         |
| *Solan*um hermannii Dun. | Solaherm | 1      | 2      | 6      | 10     |         |        |         |
| *Solan*um nigrum L. | Solanigr | 1      |        |        |         |         |        |         |
| *Sonchus* oleraceus L. | Soncole | 1      |        |        |         |         |        |         |
| *Sporobolus* africanus (Poir.) Robyns & Tournay | Sporafri | 1      | 2      | 6      | 10     | 16     | F      |         |
| *Sporobolus* fimbiatus (Trin.) Nees | Sporfmb | 2      | 6      | 10     |         |         |        |         |
| *Sutherlandia* frutescens (L.) R. Br. | Suthfrut | 1      |        |        | 10     |         |        |         |
| *Tag*etes minuta L. | Tagemnu | 1      | 6      |        |         |         |        |         |
| *Tar*axacum officinale Weber | Taraoiffi | 2      | 10     |        |         |         |        |         |
| *Tarchonathus* camphoratus L. | Tarcamp | 10     | 16     |        |         |         | T      |         |
| *Tephrosia* capensis (Jacq.) Pers. | Tephcape | 1      | 2      | 6      | 10     | 16     | F      |         |
| *Themeda* triandra Forsk. | Themtria |        |        |        |         |         | T      |         |
| *Viscum* rotundifolium L. f. | Viscrotu |        |        |        |         |         | T      |         |
| *Watsonia* longifolia J.W. Mathews & L. Bol. | Watslong |        |        |        |         |         | F      |         |
| *Withania* somnifera (L.) Dun. | Withsomn | 1      | 2      | 6      | 10     | 16     |        |         |
| *Xanthium* spinosum L. | Xantspin | 1      |        |        |         |         |        |         |
| *Zanthoxylum* capense (Thunb.) Harv. | Zantcape |        |        |        |         |         | T      |         |
the thicket and the revegetated stand samples occurred at a high eigenvalue of over 0.8 (Figure 2). *Scutia myrtina* serves as the indicator species for the thicket community.

There is a clear separation between the thicket and revegetated stands in ordination space and the revegetated stands generally plot in groups according to their age. The x-axis represents a succession gradient for the revegetated stands that tends towards that of thicket (arrow in Figure 4). There is a progression of samples from the one year stands (labelled A in Figure 4) through the stands of two and six years (labelled B and C in Figure 4) to the older seral stands of ten and sixteen years (labelled D and E respectively in Figure 4). The wide dispersion of thicket samples suggests high beta diversity.

The species ordination (DCA) of thicket and revegetated stands presents the successional replacement of species (Figure 5). Pioneer species lie to the left of an x-ordination score of 1 (dashed line, Figure 5). Thicket species plot on the right of the x-ordination score of 5 (solid line, Figure 5). The seral species are scattered between these succession-
al extremes indicating gradual replacement of pioneer species with later successional ones, and in the older stands, some climax thicket species are found: e.g. *Grewia occidentalis*; *Olea europaea* subsp. *africana*; *Protasparagus racemosus* and *Schotia latifolia*.

Revegetated stands split at the third classification level where samples of one-year old stands separate from the rest at an eigenvalue of 0.5. *Tagetes minuta* was the indicator for the pioneer successional stage. Classification at the fourth level of division separated two- and six-year stands from older stands. The six- and ten-year stages could not be separated and were indicated by *Cynodon dactylon* while the oldest stage was a *Rhus incisa* var. *effusa*--*Panicum deustum* community.

Total vegetation cover of the revegetated stands increased with age (Figure 6). The vegetation cover of the 16-year stand was similar to that of thicket. The fynbos community had a lower total percentage cover than that of the thicket and the stands revegetated for 10 years and 16 years. The herbaceous layer had a steady increase in cover from one year to 10 years, after which there was a decline. The total cover of both the tree and shrub strata increased with time up to 16 years corresponding with the reduction in the herbaceous understory. The thicket community had the highest cover of tree species (150%, Figure 6). By contrast, the fynbos community was composed primarily of a herbaceous layer as well as large contribution (150%) by the shrub component.

**Figure 5:** Ordination (DCA) of species of revegetated stands (left) and thicket (right) vegetation. The arrow represents the successional progression. Species codes are listed in Table 1

**Figure 6:** The total cover (as %) of different strata of the revegetated stands (1–16 years), thicket (T) and fynbos (F)
Species richness in the 1-year stand was almost half that found in the 2-year, 6-year and 10-year old stands (Figure 7). Species richness was lower in the 16-year stand. Thicket had the highest species richness (81 species) and fynbos the lowest.

The species similarity of the 16-year old revegetated stands showed the highest similarity to thicket (Table 2) with 46% of the species found at the 16-year site being thicket species. Similarity increased with age of the revegetated stands. The 16-year site showed a very low similarity to fynbos (15%).

Soil moisture content and organic matter content increased significantly with the age of the revegetated stands (Figure 8; moisture: F = 7.6; df = 6; P < 0.001; organic matter: H' = 24.4; df = 6; P < 0.001). Early seral stages (1-year, 2-year, 6-year) showed no significant difference for either average soil moisture or organic matter content (P > 0.05). Soil samples taken from the fynbos site had a significantly lower (P < 0.05) soil moisture and organic matter content than thicket, 10-year or 16-year stands. Thicket and older revegetated soils had almost twice the organic matter content of fynbos soils.

Intervention by planting saplings of thicket species appears to be an option as survival rates of over 70% were recorded three years after planting for *Acacia karroo*, *Azima tetracantha*, *Rhus undulata*, *Schotia afra* and *Sideroxylon inerme* (Table 3). With the exception of *Pterocelastrus tricuspidatus*, some saplings of all the other species survived (lowest of 25%, Table 3).

### Discussion

#### Revegetation

The method of revegetation implemented at Loerie quarry in the 1980s was to let the stands revegetate through what
Barbour et al. (1987) refer to as secondary succession. It was known that the soil used for the revegetation stands was obtained from areas previously covered by thicket and it was presumed that this vegetation type would regenerate through secondary succession as a result. However, the regeneration of thicket through a soil-stored seed bank is considered unlikely based on the findings of Aucamp and Tainton (1984), Stuart-Hill and Danckwerts (1988), Stuart-Hill (1991), Moolman and Cowling (1994).

**Secondary succession**

Using both the ordination and classification of samples and species it was possible to define successional species replacement within the revegetation samples (Figures 1–3). The early successional seral stage consisted of a dominance of weedy annual species (*Picris echidodes*, *Salsola kali*, *Tagetes minuta* and *Xanthium spinosum*). This was replaced by weedy perennial species (*Berkheya rigida*, *Cirsium vulgare*, *Nicotiana glauca* and *Ricinus communis*). The third seral stage consisted of grass and ground cover species such as *Atriplex semibaccata* var. *appendiculata*, *Cynodon dactylon*, *Mesembryanthemum altonis*, * Panicum maximum* and *Sporobolus africanus*. The sub-climax stages (older than 10 years) contained tree and shrub species dominated by *Acacia karroo*, *Clitria daphnoides*, *Polygala myrtifolia*, *Rhus incisa* and *Rhus laevigata*. This successional pathway follows that of Connell and Slayter’s (1977) facilitation and tolerance modes of succession: facilitation where early pioneer plants alter the soil and site characteristics so as to make it suitable for other species to establish and tolerance is where the slower growing competitors eventually exclude the early dominant ruderal species.

The cover abundance of the tree and shrub component of revegetated stands increased with age of the stand and corresponds to a decrease in the herbaceous layer (Figure 6). The reduction in the herbaceous layer could be a result of canopy shading in that they are unable to compete for the sub-climax seral species light. This is supported by the decrease in species richness that occurred in the 10-year and 16-year stands (Figure 7) where trees and shrubs became dominant.

The low species richness of the 16-year old thicket (half that of intact thicket) indicates that it does not resemble that of thicket in terms of diversity (Figure 7), however, species composition was 46% similar to thicket.

The reduction in the understorey forb component and the consequent increase in the woody component (Figure 6) suggest that succession at the Loerie quarry is still midway through the facilitation and tolerance modes (Connell and Slayter 1977). An important observation is that the succulent and geophytic component was absent from the 16-year site. Cowling (1984) and Everard (1987) emphasise the importance of the succulent and geophytic components in thicket. Many of the rare and endemic species found in thicket are of these life forms (Dyer 1937, Cowling 1984, Everard 1987). The absence of these components in the 16-year stand further indicates that the revegetation stands have not yet developed into thicket. It is also unlikely that lower-storey succulents and geophytes will return to the system in time.

Resources in the succulent and geophytic life forms are usually allocated towards water storage and the build up of reserves and their reproductive strategy favours asexual rather than sexual reproduction (Cowling 1984). The low production of seed implies that a long time span is needed for the re-colonisation of species through seed. These strategies enable the species to survive adverse climatic conditions such as erratic rainfall (Cowling 1984) but hinder their return to successional areas.

The percentage distribution of the life forms in fynbos in comparison with the 16-year old revegetated stand further excludes fynbos as the vegetation type returning in these stands. There is a very large geophyte component in the fynbos vegetation in comparison with the vegetation from all the other stands analysed (Figure 7). The geophytic life form protects the underground vegetative parts from fire and thus allows the species to flourish after fire events. It is well established that fire is a driving force in the functioning of fynbos vegetation (Cowling 1984, Low and Rebelo 1996) and the absence of this important geophytic component in the 16-year revegetated stand rules out the possibility that the vegetation is developing towards fynbos.

The tree species dominant in the 16-year old revegetated stand were dominated by *Acacia karroo*, *Rhus incisa* and *Polygala myrtifolia*. Moll (1992) had similar findings in a study on the rehabilitation of Dune Thicket at Richards Bay. The dominance of *Acacia karroo* in 10-year old revegetated Dune Thicket was believed to have caused a reduction in the herb and grass layer of these stands. Although species richness was reduced after 10 years it was found that after 50 years the *A. karroo* trees eventually died and were replaced by Dune Thicket tree and shrub species leaving only a few large individuals. Moll (1992) further reported on the importance of this species as a nurse species allowing shade tolerant thicket species to germinate under the canopy. It is believed that the build-up of organic matter and the retention of moisture under these thickets provides favourable environmental conditions for the establishment and growth of secondary thicket species that will eventually form the climax community in the rehabilitation of Dune Thicket (Moll 1992, Van Aarde and Smit 1997). It is possible that *Acacia karroo* might fulfil a similar function at Loerie with regards to the re-establishment of key thicket species. The tall *A. karroo* trees serve as perches for birds that have been recognised as the primary dispersers of seed in thicket (La Cock 1991). Observation suggests that these trees serve a function in this regard, with seedlings of *Clitria daphnoides*, *Polygala myrtifolia*, *Rhus incisa* and *Scutia myrtina* often found at the base of the *A. karroo* trees. These species are all dominant in *Euclea–Pterocelastrus* community of Kaffrarian Thicket described by Cowling (1984). It is possible that the microclimate under the canopy of the *A. karroo* trees is favourable for seed germination and the spinescent nature of the trees may also offer protection to the seedlings from herbivores.

It is interesting to note that this nurse tree association was also found under a few large *Acacia cyclops* trees found on the older revegetated stands. Seedlings of the same species found under *A. karroo* trees were also found under *A. cyclops*. Many large *A. cyclops* individuals had fallen over
due to the canopy of the trees becoming top-heavy. Seedlings of *Cluitia daphnoides*, *Polygala myrtifolia*, *Rhus incisa* and *Scuita myrtina* were found growing among the recently uprooted roots as well as underneath the branches of the fallen tree. Seeds of thicket species also germinated in the soil around the base of the fallen trees and this could be due to higher nutrient levels in the soil that would be associated with this leguminous alien species. Litter nitrogen inputs are greater for the alien acacias and this leads to an increase in the surface soil and litter layer nitrogen content (Witkowski 1991). It was also interesting to note that very few *A. cyclops* seedlings were found in association with the fallen adult trees indicating that seed from this species is unable to germinate and compete with the vegetation that has re-established.

Soil moisture and organic matter content increased with the age of the revegetated stands to a value similar to that of thicket (Figure 8). This can be attributed to the increase in vegetation cover. As the vegetation re-establishes itself at the revegetated stands, the increase in plant cover provides a favourable microclimate for the retention of moisture in the upper layers of the soil. Soil moisture content in the fynbos areas was low in comparison to the revegetated stands and thicket. The lower soil moisture content in the fynbos area could be attributed to the nature of the soil substrate. The soils of South Eastern Mountain Fynbos at Loerie are of the Oak-leaf soil form (Macvicar et al. 1977) and have a low clay content and are made up of sandy and gravel material that has a lower water holding capacity than the deeper Hutton soil. A lower abundance of thicket is found (Cowell 1984). A higher vegetation cover means that organic matter will be returned to the soil through leaf litter and the senescence of annual and perennial plants (Moll 1992).

**Predicted climax state**

Louw (1993) hypothesised that degraded thicket in the Loerie region would more likely regenerate towards Mountain fynbos rather than to the original thicket. This hypothesis can be rejected on the basis of the data presented. Analysis of the species composition, physiognomy and richness suggests that the vegetation is still in a state of flux and has not reached a climax state.

Although revegetation of the various stands has not returned them to the original vegetation state, vegetation has established and the soil environment has changed over a relatively short period of time. It is clear that the use of topsoil from areas previously covered with thicket provides the physical, chemical and biological attributes necessary for the rapid revegetation of mine areas. It is evident that the soil contains a large seed bank that contributes greatly to the speedy recovery of vegetation of these areas.

The older revegetated stands showed little invasion by exotic species with a few large *Acacia cyclops* individuals being found at the 10-year and 16-year old stands. Many of these individuals have already senesced and may be providing a habitat for the establishment of shrub species. It is evident from the small number of *A. cyclops* seedlings that the ground cover at these stands is preventing the germination of seed and the establishment of thickets. It is important that the revegetated areas be protected from massive disturbance and fire as this will stimulate the germination of *A. cyclops* seed and the reduction in the ground cover will give the species a chance to establish.

It is widely recognised by many ecologists (Cowling 1984, Everard 1987, Lubke *et al.* 1986, La Cock 1991) that thicket does not recover after massive disturbance. This idea, however, is based on limited research with little data from long term monitoring of disturbed areas. It is imperative that areas such as Loerie be monitored over a long period to test this theory. It is also important that further research into other biological mechanisms such as soil dispersal, plant-animal interactions, soil nutrients and soil mycorrhizae be implemented in order to provide a greater understanding of the process of succession at Loerie quarry and of thicket in general.

Using ordination distance as a measure of successional progression, it can be predicted that the vegetation will require at least 50 years ($r^2 = 0.874$, $n = 5$, $P < 0.05$) to develop to a state that can be considered to be near-natural thicket (calculated from a power fit of time since rehabilitation against the mean distance of the samples from a 10% fringe around the ordination spread of natural thicket).

Knowledge gained from studying the processes of revegetation and succession at the Loerie quarry may be used to speed up revegetation programmes in the future. By planting some of the indicator and nurse tree species identified in this study it may be possible to revegetate land in a much shorter period of time, with reduced labour and financial input.

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