New Approaches in Targeted Therapy for Medulloblastoma in Children

HENRIETTA MAIER, TINA DALIANIS and OURANIA N. KOSTOPOULOU

Department of Oncology-Pathology, Karolinska Institutet, Stockholm, Sweden

Abstract. Medulloblastoma (MB) is the most frequent malignant brain tumor in children. Treatment of MB is based on histopathological and molecular stratification, and includes surgical intervention, often with craniospinal irradiation and adjuvant chemotherapy. Unfortunately, however, this treatment leads to a high morbidity rate, and it does not cure all patients either, with around 30% succumbing to their disease. With improved cancer genomics and better molecular characterization, MB has been classified into four major subgroups, wingless-activated, sonic hedgehog-activated, Group 3, and Group 4 MB, with each comprising additional subtypes (2). Recently disclosed genetic drivers of MB may in the future help improve treatment, and in this way reduce therapy-related toxicity. In this review, we describe the heterogeneity of the MB subgroups, and potential new options for targeted therapy.

Medulloblastoma (MB), the most frequently occurring malignant brain tumor in children, is treated depending on histopathological and molecular stratification, and age, with the current standard of care often including surgery, craniospinal irradiation (CSI) and adjuvant chemotherapy (1). This leads to considerable morbidity among the 70% of patients that are cured, and leaves the remaining 30% in need of better therapeutic options since they are not cured (1-3). With advanced cancer genomics, elucidation of the molecular complexity of MB has resulted in its classification into four major subgroups, wingless-activated (WNT), sonic hedgehog-activated (SHH), Group 3, and Group 4 MB, with each comprising additional subtypes (2). Genomic data have also identified genetic drivers for MB, and, as specific targets, these could potentially help improve present treatment options considerably and thereby reduce therapy-related toxicity (3).

In this review, we describe the heterogeneity of the MB subgroups, and some advances in targeted therapy. We also focus on pathways that drive MB tumorigenesis, and present an overview of some current preclinical and clinical studies targeting the hedgehog (HH), phosphoinositide 3-kinase (PI3K), cyclin-dependent kinase 4 and 6 (CDK4/6) pathways, as well as approaches in immunotherapy. While research continuously sheds more light on the individual genomic complexity of MB, the challenge lies in the comprehensive evaluation and translation of newly obtained data into new therapeutic approaches.

Epidemiology and Symptomatology

Brain tumors are the most frequent solid tumors in children, and among these, MB represents the most common malignant one, accounting for 20% of all pediatric brain tumors (1, 4). The incidence of MB in children is roughly five cases per million individuals, and in adults, around one case per 2 million individuals (4, 5). MB is mainly observed in the first 9 years of life, with a peak incidence between 4 to 7 years of age (6). Although the sex predominance differs between MB subtypes, males are generally 1.5-2 times more frequently affected than females (4).

The World Health Organization has classified MB as a grade IV tumor, usually originating in the cerebellum or the posterior fossa, and shown to spread via the cerebrospinal fluid to the brain and spine (2, 7). In children, MB is mostly located in the midline of the cerebellum, whereas in adults, lateral tumors are more frequent (7). Histologically, the tumor consists of densely packed cells, which are predominantly undifferentiated. If
Molecular Subgroups of Medulloblastoma, Risk Stratification, and Quality of Life

MB is usually divided into four molecular subgroups, named according to the specifically altered cellular pathway or the specific genomic variations they harbor: WNT, SHH, Group 3 and Group 4 MB (Table 1). Within each subgroup, there are further subdivisions (10).

Wingless-activated MB. This subgroup accounts for approximately 10% of all MB cases, and is most common between ages 6-10 years or later, and is relatively equally distributed between males and females; with a 90% 5-year survival rate, it has a very good prognosis compared to other MB subgroups (8, 13). In 80-90% of cases, WNT MB presents somatic activating mutations in exon 3 of CTNNB1, which encodes β-catenin, and these mutations stabilize this protein, and in 85-90% of these same cases, a partial loss of chromosome 6 is observed (1, 8). Stabilized β-catenin accumulates in the nucleus and co-activates transcription factors, which enhance the expression of WNT-responsive genes, which in turn up-regulate cell growth and proliferation (8, 14). In patients, without CTNNB1 mutations, mutations of the tumor suppressor adenomatous polyposis coli (APC) gene are frequent (1, 8, 15). APC normally is able to build a complex with axin 1, axin 2, casein kinase 1 alpha 1 and glycogen synthase kinase 3 beta to drive ubiquitylation and degradation of β-catenin. Loss-of-function mutations of APC, with more active β-catenin, can therefore also constitutively activate WNT signaling. By increasing the understanding of the molecular background of WNT MB, targeted therapies might be helpful in reducing the amounts of chemotherapy agents required. However, targeting the WNT/β-catenin pathway directly would be critical, since it plays an important role during embryonal development, regulating stem cell pluripotency and driving cell growth and proliferation (3, 8). However, since WNT signaling is also required for bone formation, inhibition of WNT can lead to osteoporosis (3).

Sonic hedgehog-activated MB. This accounts for 25-30% of all MB and mainly affects infants (younger than 3 years) and adults, and in the latter, it is the predominant group (60%), while it generally spares children above 3 years old (1). Patients with SHH MB have an overall survival of about 70%, however, outcome strongly depends on additional molecular events, e.g. TP53 status (1, 16). SHH MB is driven by germline or somatic mutations and copy-number variations in genes of the SHH pathway, such as loss-of-function mutations or deletions of protein patched homolog 1 (PTCH1) (43% of all patients) or SUFU negative regulator of hedgehog signaling (SUFU), activating mutations in smoothened (SMO), or amplifications of zinc finger protein GLI1/2 (GLI1/GLI2) or MYCN proto-oncogene (MYCN) (1, 17). These molecular alterations result in a constitutive, ligand-independent activated SHH signaling. Normally, SHH binds PTCH1 leading to a release of SMO, which is responsible for the dissociation of the transcription factor GLI from SUFU (17). Subsequently, unbound GLI is translocated to the nucleus and activates the expression of SHH-activated genes, which drive cellular differentiation, organ formation and post-embryonic tissue regeneration (8, 18). SMO presents a favorable candidate for targeted therapies, but since it has an essential role during development, SMO antagonists can lead to serious growth plate challenges in younger patients (8, 19). Another important characteristic of SHH MB is its TP53 status, since patients harboring TP53 mutations have a 5-year survival rate of around 40%, while those with wild-type TP53 SHH MB have an increased survival rate of approximately 80% (1). Interestingly, TP53 is also commonly mutated in WNT MB but does not affect overall survival (20). Copy-number variations of TP53 or the receptor tyrosine kinase P13K are also commonly observed in SHH MB tumors (1). TP53 has
Table I. Overview of the four subgroups of medulloblastoma. Wingless (WNT)-activated, sonic hedgehog (SHH)-activated, Group 3 and Group 4 medulloblastoma (MB) are compared regarding clinical, molecular and pathological characteristics [adapted from Szalontay and Khakoo (11)].

| MB subgroup | WNT | SHH | Group 3 | Group 4 |
|-------------|-----|-----|---------|---------|
| Clinical data | Age group | Children and adults | Infants and adults | Infants and children | Children and adults |
| Gender distribution | Equal | Good | Male predominance | Male predominance |
| 5-Year survival | Very good | Equal | Poor | Intermediate |
| Metastasis | 97-100% | 67.3-88.5% | 41.9-66.2% | 66.8-82.5% |
| Occurrence | 10% | 30% | 25% | 35% |
| Mutation | CTNNB1, APC | PTCH1, SUFU, SMO, TP53 | OTX2 gain | KDM6A, ZMYM3, KMT2C, KBTBD4 |
| Copy-number alterations | Monosomy of chr6 | MYCN amplification | MYC amplification | Isochromosome 17q |
| | GLI2 amplification | Copy-number variations | Isocromosome 17q | Gain of chromosome |
| | of TP53 or PI3K | of TP53 or PI3K | 17q7 | Deletion of chromosome |
| Tumor location | Cerebellar penduncle/cerebellopontine angle | Cerebellar hemisphere | Midline, ill-defined margins | Midline, no enhancement |
| Origin | Cells in the lower rhombic lip | Cerebellar granule neuron progenitors | Uncertain | Uncertain |
| Histology | Classic | Desmoplastic nodular (MBEN – infant, LC/A – TP53-mutant) | Classic (LC/A – infant) | Classic |

APC: adenomatosis polyposis coli; chr6: chromosome 6; CTNNB1: catenin beta 1; GFI1: growth factor independent 1 transcriptional repressor oncoprotein; GLI2: GLI family zinc finger 2; KBTBD4: kelch repeat and BTB domain containing 4; KDM6A: lysine demethylase 6A; KMT2C: lysine methyltransferase 2C; LC/A: large cell/anaplastic; MBEN: medulloblastoma with extensive nodularity; MYC: MYC proto-oncogene; MYCN: MYCN proto-oncogene; OTX2: orthodenticle homeobox 2; PI3K: phosphoinositide 3-kinase; PTCH1: patched homolog 1; SMO: smoothened; SUFU: SUFU negative regulator of hedgehog signaling; TP53: tumor protein p53; ZMYM3: zinc finger MYM-type containing 3.

a checkpoint role for DNA damage and repair, therefore its dysregulation results in defective apoptosis, whereas the PI3K pathway drives proliferation and cellular growth (21). Based on DNA methylation data, four subclasses (SHHα, SHHβ, SHHγ and SHHδ) represent clinically relevant distinct subgroups (22).

**Group 3 MB.** This subgroup comprises 25% of all MB cases, and mainly affects children 2-5 years of age, with male predominance; it is associated with the worst prognosis (50% overall survival rate) (1). Group 3 MBs are primarily characterized by a high-level of MYC loci amplification but other molecular features, such as presence of isocromosome 17q, loss or gain of chromosome 8q (MYC locus at 8q24), and activation of the growth factor independent 1 transcriptional repressor oncoprotein (GFI1), and orthodenticle homeobox (OTX) amplification are also described (22). Group 3 MB relapses mainly in form of metastases, which are rarely observed in the same location as the primary tumor (23). Due to its dismal prognosis, targeted therapies are of high interest for this subgroup.

**Group 4 MB.** Accounting for 35% of all MB cases, Group 4 MB is more common in males, and while its general prognosis is good, patients with an initially metastatic disease have a much higher risk of relapse (3, 24). No consensus mutation has been identified as most frequent in this MB subgroup but genetic alterations which drive Group 4 MB include mainly the overexpression of the histone-modifying PR/SET domain protein (PRDM6), and mutations in other genes such as lysine demethylase 6A (KDM6A), zinc finger, MYM-type containing 3 (ZMYM3), lysine methyltransferase
2C (KMT2C) and kelch repeat and BTB domain containing 4 (KBTBD4) have also been described (15). In addition, this group is associated with a high burden of chromosomal copy-number variations, with gain of chromosome 7 or 17q, deletion of chromosome 8, 11 or 18p, and isochromosome 17q (11).

Risk stratification. When a patient is considered as having a high risk for poor outcome, factors such as age, extent of resection, metastasis and histological subtype play an important role. Average-risk patients are considered to be older than 3 years of age and have a tumor size <1.5 cm² after surgery. High-risk group patients have >1.5 cm² of tumor after resection and large-cell anaplastic histology (11). However, rather than classifying risk by clinicopathological variables, today risk stratification on a molecular subtype-based approach is recommended (25).

Thereby the low-risk group, with a survival higher than 90%, includes patients younger than 16 years of age with non-metastatic WNT MB, or non-metastatic Group 4 MB with genetic events as loss of chromosome 11 or gain of chromosome 17 (25). The standard-risk group (survival of 75-90%) comprises patients with non-metastatic SHH MB with tumors without a TP53 mutation or MYCN amplification, and those with non-metastatic and non-MYC-amplified Group 3 MB (25). Patients with metastatic non-infant, TP53 wild-type and MYCN-amplified non-metastatic SHH MB and metastatic Group 4 MB are considered as high-risk (survival 50-75%) (25). The very high-risk group has a survival rate of less than 50%, and includes patients with SHH MB harboring TP53 mutations, or Group 3 MB with metastasis (25).

Quality of life. The tumor, as well as its treatment, affects brain development. As a consequence, neurological, neuropsychological and physical disabilities occur in adult survivors of childhood MB (26, 27). Therapeutic interventions such as surgery, alone or together with chemotherapy, and CSI can lead to reduced intelligence, deficits in speed of processing, attention, memory and executive functions. Tumors located in the cerebellum impair motor and behavioral functions (8). Survivors of SHH MB generally have a higher quality of life and intelligence compared to those of other subgroups (28). Of note, 95% of MB-associated deaths are not due to the primary tumor but to recurrent and refractory MB (6).

Standard-of-Care Therapies

Current strategies. Standard treatment for childhood MB consists of surgery, radiation and chemotherapy. The molecular and clinical risk features of each patient should be assessed to provide a maximally curative and minimally harmful treatment (8). After successful surgery, radiation, and adjuvant chemotherapy, patients with WNT MB have an excellent 5-year progression free survival of >95%, while those with SHH MB and Group 4 MB have an intermediate rate of 70-80%, as compared to only 50-60% for those with Group 3 MB (25). Nevertheless, many MB survivors suffer from severe neurocognitive, neuro-endocrine and psychosocial deficits due to standard-of-care therapy (8). There is therefore a strong motivation to identify targeted therapies that are less toxic and more effective, in order to reduce the resulting long-term side-effects.

Surgery. After initial MB diagnosis, gross total resection of the tumor is carried out as a first-line of treatment. The volume of the tumor should be reduced as much as possible, while minimizing the risk of post-operative neurological damage (9). The location of the tumor and its relationship with surrounding structures is highly critical, as neurosurgical procedures carry the risk of developing a cerebellar mutism syndrome (CMS) (11). The incidence of CMS strongly varies within the four subgroups of MB but seems to be independent of tumor volume and location (29). Post-operatively, about 25% of all patients with MB develop difficulties with executive function, linguistic processing, visual spatial abilities and affective modulation, for which risk factors are young age, large tumor volume or midline tumor location (30). Patients with severe CMS are more likely to suffer from long-term neurocognitive effects (29).

Radiation. For children older than the age of 3 years, postoperative radiation is performed as standard of care. Average-risk patients may receive e.g. 23.4 Gy CSI and a higher boost of 55.8 Gy to the tumor bed, usually combined with weekly doses of cytostatic drugs as radiosensitizer 30 days after surgery, while those classified as high-risk patients are treated with e.g. 36 Gy CSI and additional boosts to metastases (31, 32). Notably, however, pediatric MB survivors commonly suffer from a lower intelligence quotient, memory and defective executive functions (31). Proton-beam radiation presents an alternative to photon-based approaches, since it seems to significantly improve acute toxicities and neurocognitive late effects (33). However, alopecia and radiation necrosis are more common with proton radiation (34).

Chemotherapy. When chemotherapy was added to the standard-of-care therapy as an adjunct to surgery and radiation in 1979, it substantially improved progression-free survival and overall survival rates in patients with and without metastatic MB, even with reduction of the CSI dose (35). Cytostatic drugs need to pass the blood–brain barrier, and today, common chemotherapeutic agents are cisplatin, vincristine, cyclophosphamide and lomustine (10). Multi-
agent combinations of these drugs in multiple cycles (mostly 4-9 cycles) have been implemented, where a higher cumulative dose is applied for high-risk patients (8). The event-free survival of patients with MB treated with combinations of cisplatin, lomustine and vincristine or a combination of cisplatin, cyclophosphamide and vincristine were reported to be around 81% or 86%, respectively (35). Since there is a high risk of developing long-term cognitive difficulties from radiation, children under the age of 4 years are in general spared CSI, instead receiving more intense chemotherapy (10).

**Targeted Therapies**

**General introduction.** From a molecular and clinical perspective, MB is a heterogeneous disease classified into four major subgroups, with distinct genetic profiles, and treated accordingly (36). Current standard therapy, mainly composed of maximal surgical resection followed by CSI and chemotherapy, presents severe treatment-related toxicity, and secondary tumors may also occur (36, 37). Reducing these consequences without affecting treatment efficacy is the focus of targeted therapy, aiming to reduce the doses of cytotoxic agents and radiation, as well as optimizing overall survival (38). Tailored therapy for each MB subgroup and subtype is still at its infancy (37). In WNT and SHH MB, some driver mutations and inhibitors interfering in aberrant signaling pathways or targeting somatic mutations have been identified, and are intensively studied in *in vitro* and *in vivo*. Group 4 MB is the least characterized, and Group 3 has the worst prognosis, and here high doses of cytotoxic chemotherapy are still indispensable, so current studies focus on optimizing the doses of cytostatic drugs to balance efficacy and toxicity rather than utilizing targeted therapies (37). Some potential targets for SHH and WNT MB are discussed below, and in addition, some more general targets are presented.

The hedgehog pathway. SHH MB is characterized by a constitutive active HH signaling pathway (37). In intact cells, no HH ligands are present, so PTCH1 inhibits SMO, and thereby inhibits HH signaling. Extracellular HH proteins induce the unbinding of PTCH1 and SMO, resulting in activation of the HH signaling pathway (37). More specifically, SMO translocates into the primary cilium, where it activates the GLI family zinc finger transcription factors (GLI1 and GLI2), which in turn initiate the expression of HH target genes (*GLI, PTCH1*, cyclin D1, snail family transcriptional repressor (*SNAIL*)). Mutations in these target genes may lead to SHH MB (39). Therefore, small molecules targeting upstream factors involved in HH signaling are being investigated as potential treatment.

Vismodegib (GDC-0449) is a Food and Drug Administration (FDA)-approved SMO inhibitor for SHH-dependent cancer but it is not approved for treatment of MB (40). Vismodegib has been shown to be very efficient in tumor suppression in mouse models, and its application induced tumor regression in patients with refractory metastatic MB (19, 40, 41). There are several ongoing clinical trials testing the effect of adjuvant chemotherapy in combination with vismodegib (e.g. NCT01878617). However, mutations in the SMO gene can lead to resistance to SMO inhibitors due to impaired drug binding, and induce disease progression (42). In addition, it was observed that patients with high-risk SHH MB, with an *SUFU* mutation, or *MYCN* and *GLI2* amplifications, did not respond to vismodegib but still developed growth-plate fusions after its application (17, 43). Notably, an alternative clinical trial (NCT01601184) in which patients with SHH MB were treated with vismodegib in combination with the alkylating agent temozolomide was stopped, since no positive effect was reached after the first stage of phase II.

To overcome the limitation of acquired resistance to SMO inhibitors, there is an emerging need to find new therapeutic strategies in which molecules downstream of the HH pathway are targeted. One approach is to epigenetically or directly inhibit the transcription factor GLI, which is considered to be much more efficient than inhibition of SMO, since GLI is a terminal effector of HH signaling and initiates the transcription of HH target genes (44). For instance, the GLI antagonists GANT58 and GANT61 bind to the groove between the zinc finger-2 and -3 of GLI without direct interference with DNA (44). Epigenetically, acetylation of a lysine residue on the C-terminal end of GLI functions as an inhibitory switch and thereby prevents the expression of HH target genes (45). An ongoing study (NCT03904862), in which GLI is blocked by a casein kinase 2 inhibitor (silmitasertib), is currently being evaluated in phase I/II trial for patients with recurrent or CDK4/6 pathway relapsed SHH MB (10).

**PI3K pathway.** Alterations of the signaling pathway of the intracellular lipid kinase PI3K are known to play a crucial role in MB, as well as in other solid tumors, by regulating cellular growth, proliferation and cell survival (46). Activated by growth receptor tyrosine kinases, PI3K recruits and activates protein kinase B, leading to downstream activation of other kinases, e.g. the mammalian target of rapamycin (mTOR), which promotes tumor growth intravasation and invasion, making the inhibition of this pathway favorable for tumor therapy (47). Since PI3K is involved in modulating the survival of MB stem cells following irradiation, agents targeting PI3K and its downstream signaling are considered to be beneficial alone, or in combination with adjuvant chemotherapy (48).

Recently, it was shown that PI3K inhibitors such as alpelisib (BYL719) alone, and in combination with cytostatic
drugs (cisplatin, vincristine or doxorubicin) reduced viability and proliferation in childhood neuroblastoma cell lines, indicating that these combinations might also be used for MB (49). Targeting not only PI3K, but simultaneously the fibroblast growth factor receptor (FGFR) showed an increased sensitivity compared to single-drug treatment, especially of more resistant MB childhood cell lines, and reduced cell viability and proliferation (46). The combinational inhibition of PI3K (BYL719) and mTOR (OSI-027) even enhanced anti-MB effects in vitro and in vivo compared to single-drug use (50). Thereby, the transcription factor GLI involved in HH signaling was identified as a target for simultaneous PI3K and mTOR inhibition (50).

Another therapeutic strategy combined targeting of the PI3K/mTOR pathway and HH signaling. Treatment of MB cells with the HH inhibitor vismodegib and the PI3K inhibitor BEZ235 significantly suppressed cell growth and survival, and in addition, increased cisplatin-mediated cytotoxicity (51). In addition, in vivo experiments with MYC-amplified NOD scid gamma mouse models supported this observation, since combinational treatment with vismodegib and BEZ235, or their respective combination with cisplatin delayed tumor growth and enhanced survival of mice (51). Two ongoing clinical trials are currently investigating the use of the PI3K inhibitor samotolisib (LY3023414) in patients with recurrent MB (NCT03213678, NCT03155620) but both are still in the recruitment phase.

The CDK inhibitor pathway. The cyclin D–CDK–inhibitor of CDK4 (INK4) retinoblastoma (Rb) pathway regulates cell-cycle progression by initiation of the G1–S-phase transition. Active CDK4/6 can phosphorylate the Rb tumor-suppressor protein, so it dissociates from E2F transcription factors and thereby allows for DNA replication and cell-cycle progression (52). Increased CDK4/6 activity can be mediated by overexpression of D-type cyclins, mutations, or amplification of CDK4/6, as shown e.g. in MB, or the loss of cyclin D–CDK4/6 negative regulators, for instance p16INK4A, which can subsequently lead to cancer growth (21, 53). Selective inhibition of CDK4/6 potently arrests the cell cycle and thereby presents a favorable target for cancer treatment. A theoretical concern about these inhibitors is that CDKs also have an essential role in normal cells. However, cells with a dysregulated CDK4/6–INK4–Rb pathway response, should be more sensitive than normal cells, and in this way these endogenous inhibitors may spare CDK2 activity and allow normal cells to proliferate (54).

Three selective CDK4/6 inhibitors, namely palbociclib, ribociclib and abemaciclib, have been approved by the FDA for treatment of estrogen receptor-positive metastatic breast cancer (55). The addition of CDK4/6 inhibitors to established treatments of MB might therefore possibly improve the efficacy and durability of the antitumor response, and may potentially overcome acquired or de-novo treatment resistance. According to a study using patient-derived xenograft mouse models, the CDK4/6–cyclin D–Rb pathway was characterized as a ‘druggable’ pathway for all non-WNT MB (56). Furthermore, experimental data have shown that pretreatment with biologically achievable doses of palbociclib significantly reduced the surviving fraction of tumor cells in response to radiation and increased the sensitizer enhancement ratio (57). Clinical and experimental studies have also demonstrated enhanced cytotoxicity when combining various cytostatic drugs with palbociclib, ribociclib or abemaciclib for several tumor types, e.g. non-small-cell lung carcinoma, breast, ovarian, gastric, pancreatic cancer and others (55, 58–63). Furthermore, addition of CDK4/6 inhibitors enhanced chemotherapy-induced apoptosis of tumor cells in vitro and in vivo contexts in preclinical studies (55). Combining CDK4/6 inhibitors and chemotherapy is therefore an attractive therapeutic strategy, especially since these inhibitors are not associated with cumulative toxicity and can therefore be administered for a long time period (63).

A clinical study from the St. Jude Children’s Research Hospital, USA, has just started and is investigating the combination of the CDK4/6 inhibitor ribociclib with the cytostatic gemcitabine, or in combination with the small molecule trametinib, or with the HH antagonist sonidegib, respectively, to treat recurrent or progressive MB (NCT03434262). Another approach is following the combinational treatment comprising the CDK4/6 inhibitor palbociclib, ribociclib, or abemaciclib, respectively, together with the alkylating agent temozolomide and the cytotoxic alkaloid irinotecan (NCT03709680, and NCT04238819). Two clinical trials investigating the single-drug use of either palbociclib or abemaciclib in patients with recurrent or refractory solid tumors (including MB) are also ongoing (NCT03526250, NCT02644460).

Immunotherapy. Another treatment approach for MB is immunotherapy. In a recent review on immunotherapy and medulloblastoma, the authors concluded that cancer vaccines, oncolytic viral therapy, natural killer cells, and CAR T therapy may all be effective against the innate immunosuppressive properties of MB, and thereby potentially prolong survival (64). Immunotherapy may be beneficial for a variety of cancer types, however, its application in central nervous system tumors is still challenging due to the lack of immunogenic antigens (65). Nevertheless, the variety of dysregulated pathways in MB potentially increases the re-expression of fetal, developmental antigens, which might be targeted since they are not expressed in normal tissue (65, 66). To target such antigens might be an attractive strategy, since it would eliminate cancer cells while sparing important surrounding brain tissue (65, 66).
Another alternative is to use immunotherapy in conjunction with chemotherapy or radiotherapy. Clinical and pre-clinical studies have, for example, shown that simultaneous treatment with vaccination (such as with dendritic cells loaded with brain tumor stem cells or total tumor RNA-loaded autologous lymphocyte transfer) and chemotherapy improved antigen-specific T-cell activity (65, 67). Furthermore, low-dose radiation up-regulated expression of human leukocyte antigen class I and class II in various MB cells by more than 20%, while the high amount of reactive oxygen species generated resulted in increased human epidermal growth factor receptor 2 (HER2) and vascular endothelial growth factor expression, which can both be targeted by antibodies (64, 68). The latter observations imply synergistic effects of low-dose radiation and the application of monoclonal antibodies.

Notably, anti-HER2 chimeric antigen receptor T-cell (CAR T) therapy is a successful approach to treating several types of cancer, and the HER2 receptor is also known to be overexpressed in MB. Furthermore, one study has already shown that CAR T-cells were able to clear MB implanted in the posterior fossa in mice without any significant toxicity (64). However, antigenic escape is a prevalent challenge, therefore CAR T-cells should be designed to target multiple tumor antigens. A clinical trial is currently investigating HER2 CAR T therapy in patients with HER2-positive recurrent central nervous system tumor, including those with MB (NCT03500991).

Another clinical trial currently in phase I and II utilizes a cancer vaccine consisting of RNA-loaded autologous dendritic cells (DCs) which is administered to patients with MB (NCT01326104). In this trial, DCs were matured and activated with specific inflammatory cytokines, and then incubated with amplified tumor messenger RNA, which originated from an individual cDNA library from each patient (69). Finally, the DCs were cultured with T-cells and re-administered together with these T-cells to the respective patient with MB. This procedure is complicated and time-consuming, since tumor cells from each patient have to be collected (64, 69). Unfortunately, several clinical trials testing cancer vaccines in MB have been unsuccessful and were terminated (NCT02332889, NCT01171469, NCT00014573).

Other approaches are also useful, as indicated by a research group showing that MB cells express the poliovirus receptor CD155 on their surface, which allows a poliovirus to infect and inhibit tumor proliferation (70). This serves as an optimal target for oncolytic viral therapy using poliovirus, and has already demonstrated efficacy in vitro (64). CD155 expression was found to be the highest in WNT MB and Group 3 MB, and infection by poliovirus reduced proliferation, resulting in cellular death (70). There are several ongoing clinical studies evaluating cancer vaccines using poliovirus, reovirus, or oncolytic herpes simplex virus (NCT03043391, NCT02444546, NCT03911388).

Finally, programmed death-ligand 1 is a favorable target for checkpoint inhibitors but the complete absence of this ligand in MB has been reported (71). Nevertheless, clinical studies are investigating checkpoint inhibitors against programmed death-ligand 1 in patients with MB (NCT03173950, NCT02359565) (65).

**Summary and Conclusion**

Despite recent advances in cancer genomics, the standard of care for all MB subgroups still generally consists of maximal surgical resection, CSI and adjuvant chemotherapy, leading to numerous severe short- and long-term side-effects. For new therapeutic approaches, major efforts have been made to characterize specific gene mutations or amplifications as possible targets for tailored treatment, with some of them being already investigated in pre-clinical or clinical trials.

Currently, HH pathway inhibitors, already FDA-approved for treatment of other types of tumors, deliver promising clinical trials for the therapy of SHH-driven MB. Other targets such as PI3K or CDK4/6 are highly interesting for personalized therapies, and their use is already being evaluated in several trials for other cancer types. However, one major obstacle is the development of drug resistance to these pathway inhibitors, which could potentially be overcome by combining inhibitors of different pathways. An alternative possibility would be to combine pathway inhibitors, with chemotherapy, since an enhanced sensitivity of MB cancer cells to chemotherapy was revealed when mediated by inhibitor treatment.

In contrast, specific tumor drivers in Group 3 and Group 4 MB have not been well identified, and fewer specific approaches for these subgroups are available. More information on the nature of these tumors and further trials are important in improving patient survival and quality of life.

A different strategy altogether is to treat MB by immunotherapy, where the major challenge lies in the identification of new antigens to engineer therapies with simultaneously more than one target. In this way, immune-suppression can be counteracted, and the patient’s immunity activated by cancer vaccines, viruses, or T-cell therapy.

Finally, continued effort should be made on the molecular characterization of MB, with the aim of personalizing treatment strategy when defining the initial diagnosis, thereby maximally reducing treatment-related toxicity while retaining and improving overall survival. As pre-clinical research is continuously driving the field forward, with large international trials, the biology of individual treatment responses is being more and more understood. Using this comprehensive knowledge, targeted therapies have the potential to improve overall survival as well as the quality of life of MB survivors.
MG, Collins VP, Kieran MW, Cho YJ, Pomeroys SL, Witt O, Brors B, Taylor MD, Schüller U, Korshunov A, Eils R, Wechsler-Reya RJ, Lichter P, Pfister SM and ICGC PedBrain Tumor Project.: Genome sequencing of SHH medulloblastoma predicts genotype-related response to smoothened inhibition. J Clin Oncol 25(3): 393-405, 2014. PMID: 24651015. DOI: 10.1016/j.jco.2014.02.004

18 Choudhry Z, Rikani AA, Choudhry AM, Tariq S, Zakaria F, Ashgar MW, Sarfraz MK, Haider K, Shafiaq AA and Mobassarah NJ: Sonic hedgehog signalling pathway: A complex network. Ann Neurosci 21(1): 28-31, 2014. PMID: 25206052. DOI: 10.5214/ans.0972.7531.210109

19 Petrirena GJ, Masliha-Planchnol J, Sala Q, Pourroy B, Frappaz D, TaboureT E, Graillon T, Gentet JC, Delattre O, Chiotin O and Padovani L: Recurrent extraneural sonic hedgehog medulloblastoma exhibiting sustained response to vismodegib and temozolomide monotherapies and inter-metastatic molecular heterogeneity at progression. Oncotarget 9(11): 10175-10183, 2018. PMID: 29515801. DOI: 10.18632/oncotarget.23699

20 Zhukova N, Ramaswamy V, Remke M, Pfaff E, Shih DJ, Martin DC, Castelo-Branco P, Baskin B, Ray PN, Bouffet E, von Bueren AO, Jones DT, Northcott PA, Kool M, Sturm D, Pugh NJ: Sonic hedgehog signalling pathway: A complex network. Ann Neurosci 21(1): 28-31, 2014. PMID: 25206052. DOI: 10.5214/ans.0972.7531.210109

21 Northcott PA, Shih DJ, Peacock J, Garzia L, Morrissy AS, Zichner T, Stütz AM, Korshunov A, Reimand J, Schüller U, Hill RM, Lindsey JC, Schwalbe EC, Bailey S, Ellison DW, Hawkins C, Malkin D, Clifford SD, Kool M, Karajannis MA, Gardner SL, Zitterbart K, Faria CC, Schüller U, Kren L, Kumabe T, Lonij A, Maier et al: Targeted Therapy of Medulloblastoma (Review)
Pfister SM, Taylor MD and Pomeroy SL: Risk stratification of childhood medulloblastoma in the molecular era: The current consensus. Acta Neuropathol 131(6): 821-831, 2016. PMID: 27040285. DOI: 10.1007/s00401-016-1569-6

26 Moxon-Emre I, Bouffet E, Taylor MD, Laperriere N, Scantlebury N, Law N, Spiegel BJ, Malkin D, Janzen L and Mabbott D: Impact of craniospinal dose, boost volume, and neurologic complications on intellectual outcome in patients with medulloblastoma. J Clin Oncol 32(17): 1760-1768, 2014. PMID: 24516024. DOI: 10.1001/jco.2013.52.3290

27 King AA, Seidel K, Di C, Leisenring WM, Perkins SM, Krull KR, Sklar CA, Green DM, Armstrong GT, Zeltzer LK, Wells E, Stovall M, Ullrich NJ, Oeffinger KC, Robson LL and Packer RJ: Long-term neurologic health and psychosocial function of adult survivors of childhood medulloblastoma/PNET: A report from the Childhood Cancer Survivor Study. Neuro Oncol 19(5): 689-698, 2017. PMID: 28039368. DOI: 10.1093/neuonc/nwx242

28 Bull KS, Kennedy CR, Bailey S, Ellison DW and Clifford SC: Improved health-related quality of life outcomes associated with SHH subgroup medulloblastoma in SIOP-UKCCSG PNET3 trial survivors. Acta Neuropathol 128(1): 141-153, 2014. PMID: 24879579. DOI: 10.1007/s00401-014-1300-4

29 Jabarkheel R, Amayiri N, Yecies D, Huang Y, Toescu S, Nobre L, Mabbott DJ, Sudhakar SV, Malik P, Laughlin S, Swaidan M, Al Hussaini M, Musharbash A, Chacko G, Jones LG, Fisher PG, Hargrave D, Bartels U, Tabori U, Pfister SM, Aquilina K, Taylor MD, Grant GA, Bouffet E, Mankad K, Yeom KW and Kasamwasy V: Molecular correlates of cerebellar mutism syndrome in medulloblastoma. Neuro Oncol 22(2): 290-297, 2020. PMID: 31504816. DOI: 10.1093/neuonc/noz158

30 Schmahmann JD: Neuroanatomy of pediatric postoperative cerebellar cognitive affective syndrome and mutism. Neurology 93(16): 693-694, 2019. PMID: 31527286. DOI: 10.1212/WNL.0000000000083311

31 Merchant TE, Kun LE, Krasin MJ, Wallace D, Chintagumpala MM, Woo SY, Ashley DM, Sexton M, Kellei SJ, Ahern V and Gajjar A: Multi-institution prospective trial of reduced-dose craniospinal irradiation (23.4 Gy) followed by conformal posterior fossa (36 Gy) and primary site irradiation (55.8 Gy) and dose-intensive chemotherapy for average-risk medulloblastoma. Int J Radiat Oncol Biol Phys 70(3): 782-787, 2008. PMID: 17892918. DOI: 10.1016/j.ijrobp.2007.07.2342

32 Hwang EI, Kool M, Burger PC, Capper D, Chavez L, Brabetz M, King AA, Seidel K, Di C, Leisenring WM, Perkins SM, Krull KR, Sklar CA, Green DM, Armstrong GT, Zeltzer LK, Wells E, Stovall M, Ullrich NJ, Oeffinger KC, Robson LL and Packer RJ: Long-term neurologic health and psychosocial function of adult survivors of childhood medulloblastoma/PNET: A report from the Childhood Cancer Survivor Study. Neuro Oncol 19(5): 689-698, 2017. PMID: 28039368. DOI: 10.1093/neuonc/nwx242

33 Thariat J, Hannoun-Levi JM, Sun Myint A, Vuong T and Gérard H: Radiation necrosis in pediatric patients with brain tumors treated with proton radiotherapy. AJNR Am J Neuroradiol 36(8): 1572-1578, 2015. PMID: 26138138. DOI: 10.3174/ajnr.A4333

34 Krailik SF, Ho CY, Finke W, Buchbaum JC, Haskins CP and Shih CS: Radiation necrosis in pediatric patients with brain tumors treated with proton radiotherapy. AJNR Am J Neuroradiol 36(8): 1572-1578, 2015. PMID: 26138138. DOI: 10.3174/ajnr.A4333

35 Packer RJ, Gajjar A, Vezina G, Horke-Adams L, Burger PC, Robertson PL, Bayer L, LaFond D, Donahue BR, Marymont MH, Muraszko K, Langston J and Sposto R: Phase III study of craniospinal radiation therapy followed by adjuvant chemotherapy for newly diagnosed average-risk medulloblastoma. J Clin Oncol 24(25): 4202-4208, 2006. PMID: 16943538. DOI: 10.1200/JCO.2006.06.4980

36 Haltom AR, Toll SA, Cheng D, Maegawa S, Gopalakrishnan V and Khatua S: Medulloblastoma epigenetics and the path to clinical innovation. J Neurooncol 150(1): 35-46, 2020. PMID: 32816225. DOI: 10.1007/s11060-020-03591-9

37 Liu X, Ding C, Tan W and Zhang A: Medulloblastoma: Molecular understanding, treatment evolution, and new developments. Pharmacol Ther 210: 107516, 2020. PMID: 32105673. DOI: 10.1016/j.pharmthera.2020.107516

38 Thompson EM, Ashley D and Landi D: Current medulloblastoma subgroup specific clinical trials. Transl Pediatr 9(2): 157-162, 2020. PMID: 32477916. DOI: 10.21037/tp.2020.03.03

39 Pak E and Segal RA: Hedgehog signal transduction: Key players, oncogenic drivers, and cancer therapy. Dev Cell 38(4): 333-344, 2016. PMID: 27558555. DOI: 10.1016/j.devcel.2016.07.026

40 Rudin CM, Hann CL, Laterra J, Jauch RL, Callahan CA, Fu L, Holcomb T, Stinson J, Gould SE, Coleman B, LoRooso PM, Von Hoff DD, de Sauvage FJ and Low JA: Treatment of medulloblastoma with hedgehog pathway inhibitor GDC-0449. N Engl J Med 361(12): 1173-1178, 2009. PMID: 19726761. DOI: 10.1056/NEJMoa0902903

41 Ajeawung NF, Wang HY, Gould P and Kasnasaran D: Advances in molecular targets for the treatment of medulloblastomas. Clin Invest Med 35(5): E246, 2012. PMID: 23043706. DOI: 10.25011/cim.v35i5.18697

42 Sharpe HJ, Pau G, Dijkstra GF, Basset-Seguin N, Modrusan Z, Janzen L, Mabbott DJ, Sudhakar SV, Malik P, Laughlin S, Swaidan M, Al Hussaini M, Musharbash A, Chacko G, Jones LG, Fisher PG, Hargrave D, Bartels U, Tabori U, Pfister SM, Aquilina K, Taylor MD, Grant GA, Bouffet E, Mankad K, Yeom KW and Kasamwasy V: Molecular correlates of cerebellar mutism syndrome in medulloblastoma. Neuro Oncol 22(2): 290-297, 2020. PMID: 31504816. DOI: 10.1093/neuonc/noz158

43 Robinson GW, Kaste SC, Chemaitilly W, Bowers DC, Laughton S, Smith A, Gottardo NG, Partap S, Bendeel A, Wright KD, Orr BA, Warner WC, Onar-Thomas A and Gajjar A: Irreversible growth plate fusions in children with medulloblastoma treated with a targeted hedgehog pathway inhibitor. Oncotarget 8(41): 69295-69302, 2017. PMID: 29050204. DOI: 10.18632/oncotarget.20619

44 Infante P, Alfonsi R, Botta B, Mori M and Di Marcotullio L: Targeting GLI factors to inhibit the Hedgehog pathway. Trends Pharmacol Sci 36(8): 547-558, 2015. PMID: 26072120. DOI: 10.1016/j.tips.2015.05.006

45 Coni S, Antonucci L, D’Amico D, Di Magna L, Infante P, De Smaele E, Giannini G, Di Marcotullio L, Sprecanti I, Gulino A and Canettieri G: Gli2 acetylation at lysine 757 regulates hedgehog-dependent transcriptional output by preventing its promoter occupancy. PLoS One 8(6): e65718, 2013. PMID: 23762415. DOI: 10.1371/journal.pone.0065718

46 Holzhauer S, Lukoseviciute M, Andonova T, Ursu RG, Dalianis T, Wickström M and Kostopoulou ON: Targeting Fibroblast Growth Factor Receptor (FGFR) and Phosphoinositide 3-kinase (PI3K) signaling pathways in medulloblastoma cell lines.
Anticancer Res 40(1): 53-66, 2020. PMID: 31892552. DOI: 10.21873/anticancer.13925

Yuan TL and Cantley LC: PI3K pathway alterations in cancer: Variations on a theme. Oncogene 27(41): 5497-5510, 2008. PMID: 18794848. DOI: 10.1038/onc.2008.245

Hambardzumyan D, Becher OJ, Rosenblum MK, Pandolfi PP, Manova-Todorova K and Holland EC: PI3K pathway regulates survival of cancer stem cells residing in the perivascular niche following radiation in medulloblastoma in vivo. Genes Dev 22(4): 436-448, 2008. PMID: 18281460. DOI: 10.1101/gad.1627008

Holzhauser S, Lukosevicu M, Papachristofi C, Vasilopoulou C, Herold N, Wickström M, Kostopoulou ON and Dalianis T: Effects of PI3K and FGFR inhibitors alone and in combination, with/without cytostatics in childhood neuroblastoma cell lines. Int J Oncol 58(2): 211-225, 2021. PMID: 33491755. DOI: 10.3892/ijo.2021.5167

Eckerdt F, Clymer J, Bell JB, Beauchamp EM, Blyth GT, Goldman S and Platianis LC: Pharmacological mTOR targeting enhances the antiproliferative effects of selective PI3K inhibitor in medulloblastoma. Sci Rep 9(1): 12822, 2019. PMID: 31492956. DOI: 10.1038/s41598-019-49299-3

Chaturvedi NK, Kling MJ, Coulter DW, McQuire TR, Ray S, Eckerdt F, Clymer J, Bell JB, Beauchamp EM, Blyth GT, Yuan TL and Cantley LC: PI3K pathway alterations in cancer: anti-tumor activities alone/in combination with gemcitabine.

Donson AM, Foreman NK and Vibhakar R: Inhibition of cyclin-dependent kinase 4/6 sensitizes multidrug resistant ovarian cancer cells to paclitaxel by increasing apoptosis. Cell Oncol (Dordr) 40(3): 209-218, 2017. PMID: 28243976. DOI: 10.1007/s13402-017-0316-x

Dowell M, Lowery CD, Shackelford T, Renschler M, Stephens J, Flack R, Blosser W, Gupta S, Stewart J, Webster Y, Dempsey J, VanWye AB, Ebert P, Iversen P, Olsen JB, Gong X, Buchanan S, Houghton P and Stancato L: Abemaciclib is active in preclinical models of ewing sarcoma via multipronged regulation of cell cycle, DNA methylation, and interferon pathway signaling. Clin Cancer Res 24(23): 6028-6039, 2018. PMID: 30131386. DOI: 10.1158/1078-0432.CCR-18-1256

Iyengar M, O’Hayer P, Cole A, Sebastian T, Yang K, Coffman L and Buckanovich RJ: CDK4/6 inhibition as maintenance and combination therapy for high grade serous ovarian cancer. Oncotarget 9(21): 16619-16633, 2018. PMID: 29682173. DOI: 10.18632/oncotarget.24618

Burkhart DL and Sage J: Cellular mechanisms of tumour suppression by the retinoblastoma gene. Nat Rev Cancer 8(9): 671-682, 2008. PMID: 18650841. DOI: 10.1038/nrc2399

Shapiro GI: Cyclin-dependent kinase pathways as targets for cancer treatment. J Clin Oncol 24(11): 1770-1783, 2006. PMID: 16603719. DOI: 10.1200/JCO.2005.03.7689

Roberts PJ, Bisi JE, Strum JC, Combest AJ, Darr DB, Usary JE, Zamboni WC, Wong KK, Perou CM and Sharpless NE: Multiple roles of cyclin-dependent kinase 4/6 inhibitors in cancer therapy. J Natl Cancer Inst 104(6): 476-487, 2012. PMID: 22302033. DOI: 10.1093/jnci/djs002

Roberts PJ, Kumaramassy V, Witkiewicz AK and Knudsen ES: Chemotherapy and CDK4/6 inhibitors: Unexpected bedfellows. Mol Cancer Ther 19(8): 1575-1588, 2020. PMID: 32546660. DOI: 10.1158/1355-7183.MCT-18-1161

Cook Sangar ML, Genovesi LA, Nakamoto MW, Davis MJ, Knobluagh SE, Ji P, Millar A, Wainwright BJ and Olson JM: Inhibition of CDK4/6 by palbociclib significantly extends survival in medulloblastoma patient-derived xenograft mouse models. Clin Cancer Res 23(19): 5802-5813, 2017. PMID: 28637687. DOI: 10.1158/1078-0432.CCR-16-2943

Whiteway SL, Harris PS, Venkataraman S, Ailinova I, Birks DK, Donson AM, Foreman NK and Vibhakar R: Inhibition of cyclin-dependent kinase 6 suppresses cell proliferation and enhances radiation sensitivity in medulloblastoma cells. J Neurooncol 113(2): 113-121, 2013. PMID: 23138228. DOI: 10.1007/s11060-012-1000-7

Gelbert LM, Cai S, Lin X, Sanchez-Martinez C, Del Prado M, Lallena MJ, Torres R, Ajamie RT, Wishart GN, Flack RS, Neubauer BL, Young J, Chan EM, Iversen P, Cronier D, Kreklau E and de Dios A: Preclinical characterization of the CDK4/6 inhibitor LY2835219 in vivo cell cycle-dependent/independent anti-tumor activities alone/in combination with gemcitabine. Invest New Drugs 32(5): 825-837, 2014. PMID: 24919854. DOI: 10.1007/s10637-014-0120-7

Vora SR, Juric D, Kim N, Mino-Kenudson M, Huynh T, Costa C, Lockerman EL, Pollack SF, Liu M, Li X, Lehar J, Wiesmann M, Wartmann M, Chen Y, Cao ZA, Pinzon-Ortiz M, Kim S, Schlegel R, Huang A and Engelman JA: CDK 4/6 inhibitors sensitize PI3KCA mutant breast cancer to PI3K inhibitors. Cancer Cell 26(1): 136-149, 2014. PMID: 25002028. DOI: 10.1016/j.ccr.2014.05.020

Bonelli MA, Di Giacomo G, Fumolaro C, Alfieri R, Quaini F, Falco A, Madeddu D, La Monica S, Cretella D, Ravelli A, Ulivi P, Tebaldi M, Calisti D, Delmonte A, Ampollini L, Carbognani P, Tiseo M, Cavazzoni A and Petronini PG: Combined inhibition of CDK4/6 and PI3K/AKT/mTOR pathways induces a synergistic anti-tumor effect in malignant pleural mesothelioma cells. Neoplasia 19(8): 637-648, 2017. PMID: 28704762. DOI: 10.1016/j.neo.2017.05.003

Gao Y, Shen J, Choy E, Mankin H, Hornick F and Duan Z: Inhibition of CDK4 sensitizes multidrug resistant ovarian cancer cells to paclitaxel by increasing apoptosis. Cell Oncol (Dordr) 40(3): 209-218, 2017. PMID: 28243976. DOI: 10.1007/s13402-017-0316-x
69 Nair SK, Driscoll T, Boczkowski D, Schmittling R, Reynolds R, Johnson LA, Grant G, Fuchs H, Bigner DD, Sampson JH, Gururangan S and Mitchell DA: Ex vivo generation of dendritic cells from cryopreserved, post-induction chemotherapy, mobilized leukapheresis from pediatric patients with medulloblastoma. J Neurooncol 125(1): 65-74, 2015. PMID: 26311248. DOI: 10.1007/s11060-015-1890-2

70 Thompson EM, Brown M, Dobrikova E, Ramaswamy V, Taylor MD, McLendon R, Sanks J, Chandramohan V, Bigner D and Gromeier M: Poliovirus receptor (CD155) expression in pediatric brain tumors mediates oncolysis of medulloblastoma and pleomorphic xanthoastrocytoma. J Neuropathol Exp Neurol 77(8): 696-702, 2018. PMID: 29878245. DOI: 10.1093/jnen/nly045

71 Vermeulen JF, Van Hecke W, Adriaansen EJM, Jansen MK, Bouma RG, Villacorta Hidalgo J, Fisch P, Broekhuizen R, Spliet WGM, Kool M and Bovenschen N: Prognostic relevance of tumor-infiltrating lymphocytes and immune checkpoints in pediatric medulloblastoma. Oncoimmunology 7(3): e1398877, 2017. PMID: 29399402. DOI: 10.1080/2162402X.2017.1398877

Received February 23, 2021
Revised March 3, 2021
Accepted March 4, 2021