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Mask assistance to colorimetric sniffers for detection of Covid-19 disease using exhaled breath metabolites

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ABSTRACT

According to World Health Organization reports, large numbers of people around the globe have been infected or died for Covid-19 due to the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Researchers are still trying to find a rapid and accurate diagnostic method for revealing infected people by low viral load with the overriding goal of effective diagnostic management. Monitoring the body metabolic changes is known as an effective and inexpensive approach for the evaluation of the infected people. Here, an optical sniffer is introduced to detect exhaled breath metabolites of patients with Covid-19 (60 samples), healthy humans (55 samples), and cured people (15 samples), providing a unique color pattern for differentiation between the studied samples. The sniffer device is installed on a thin face mask, and directly exposed to the exhaled breath stream. The interactions occurring between the volatile compounds and sensing components such as porphyrazines, modified organic dyes, porphyrins, inorganic complexes, and gold nanoparticles allowing for the change of the color, thus being tracked as the sensor responses. The assay accuracy for the differentiation between patient, healthy and cured samples is calculated to be in the range of 80%-84%. The changes in the color of the sensor have a linear correlation with the disease severity and viral load evaluated by rRT-PCR method. Interestingly, comorbidities such as kidney, lung, and diabetes diseases as well as being a smoker may be diagnosed by the proposed method. As a powerful detection device, the breath sniffer can replace the conventional rapid test kits for medical applications.

1. Introduction

Based on the World Economic Forum report, the development of breath sensors for rapid detection of diseases is one of the ten emerging technologies that will be highly regarded by researchers and commercial companies in the next two to five years [1]. Exhaled breath is composed of many volatile metabolites, which are classified into amines, ketones, acids, aldehydes, and carbohydrates [2]. The variation in the metabolism concentrations indicates a physiological disorder in the human body, eventually leading to the lung, kidney, and gastrointestinal diseases, airway inflammation, and metabolic disorders (e.g., diabetes, obesity, nonalcoholic fatty liver disease, and hyperlipidemia) [3].

So far, the breath metabolites have been estimated using standard laboratory methods such as gas chromatography (GC) [4], spectroscopic methods including mass spectrometry (MS) [5], proton transfer reaction mass spectrometry (PTR-MS) [6], laser spectroscopy [7], ion mobility spectrometry and/or a combined approach of GC and MS [8]. These methods provide the sufficient information about the type and the amount of metabolites in infected, healthy, and even cured patients. Different metabolic profiles for each sample arise from the appearance, removal, and a concentration change of a certain metabolite [9].

By the advent of SARS-CoV-2 in 2019, and the widespread epidemic...
Covid-19 as a contagious respiratory disease [10], several studies have been performed to evaluate and compare the exhaled breath compositions of Covid-19 patients, healthy people, and individuals infected with other acute respiratory diseases. In this regard, the values of some chemical compounds such as 2, 4-octadiene 1-chloroheptane, nonanal, ethanol, octanol, acetone, butanone, and methanol in the metabolic profiles of the patients infected by Covid-19 have been reported to be different from those by other respiratory disorders [11,12]. The discrimination of healthy samples from the Covid-19 patients were obtained by evaluating the changes in the concentration of some volatile species, consisting of 1-propanol, 3, 6-methylundecane, camphene, beta-cubebene, and iodobenzene [13,14].

Determination of the trace amounts of volatiles, and creation of a unique and reliable response for each exhaled breath sample are the most important features of conventional methods. Nevertheless, collecting virus-infected samples, and transferring them to an isolated ethanal, octanal, acetone, butanone, and methanol in the metabolic consumptions of an excessive volume of materials or reagents, together with collecting virus-infected samples, and transferring them to an isolated healthy controls with an accuracy of 84.0% [30]. The present study proposes an opto-E-nose which is directly exposed to exhaled breath metabolites using a thin face mask without the need of the external force for transferring the sample to the surface of the sensor. The device is made of the low-cost general filter paper, whose configuration comprises a diverse set of sensing elements such as porphyrinazenes, porphyrins, AuNPs, metal ion complexes, and modified organic dyes. This design allows for the measurement of a variety of volatiles, while also increasing the sensor selectivity for recognizing a specific chemical compound, and determining the samples at low concentrations with high sensitivity. Overall, the opto-E-nose is expected to provide a favorable response for differentiating the Covid-19 patients from healthy controls and cured samples, and to have the high ability to simultaneously find the metabolites of other comorbidities. Also, the assay is examined for making a semi-quantitative relationship between the disease severity and the color changes of sensing components.

2. Experimental section

2.1. Instruments and software

Designing and printing the pattern of the proposed sensor was performed by AutoCAD 2016 software and HP LaserJet printer 1200, respectively. A Canon scanner (CanoScan LiDe 220) was used to capture the sensor photos. The resulting images were processed by Image J (1.51 n, National Institutes of Health, USA). Statistical data processing was carried out by MATLAB R2015 and SPSS (Version 22; Chicago, IL, USA) software. The sensing components were spotted on the surface of paper by a microipette (BRAND Transferpette®, Germany). To control the pH of media, a Metrohm 632 pH-meter (Model 780 pH lab) was used.

2.2. Materials and solutions

The organic dyes such as bromophenol red (R1), acridine orange (R2), indigo carmine (R3), toluidine blue (R4), malachite green (R5), phenol red (R6), pararosaniline hydrochloride (R7), thymol blue (R8), methyl red (R9), bromophenol blue (R10), bromopyrogallol red (R11), methyl blue (R12), pyrocatechol violet (Py), gallic acid (GA) and the chemicals like sodium citrate, bovine serum albumin (BSA), polyvinyl pyrrolidone (PVP), 2, 4-dinitrophenylhydrazine, phenylboronic acid (PBA), p-toluenesulfonic acid monohydrate (TsOH), tetrabutylammonium hydroxide (TBAOH), vanadyl sulfate pentahydrate (VOSO_4·5H_2O) were obtained from Sigma Aldrich. The other compounds consist of bromocresol purple (R13), gold (III) chloride trihydrate (HAuCl_4·3H_2O), sodium borohydride (NaBH_4), sodium hydroxide (NaOH), ethanol (EtOH), sulfuric acid (H_2SO_4), nickelf(II) nitrate hexahydrate (Ni(NO_3)_2·6H_2O), copper(II) nitrate trihydrate (Cu(NO_3)_2·3H_2O), iron(II) chloride tetrahydrate (FeCl_2·4H_2O), iron(III) nitrate nonahydrate (Fe(NO_3)_3·9H_2O) and boric acid (H_3BO_3) were provided from Merck chemical company. The pattern of sensor was drawn on the Whatman® Grade NO.2 filter paper. The water soluble tetramethyl quarternized tetracationic porphyrinazenes (tetramethyl tetra-3,4-pyridinoporphyrinato cobalt(II), [Co(3,4-tmpnpa)]^3+, tetramethyl tetra-2,3-pyridinoporphyrinato copper(II), [Cu(2,3-tmpnpa)]^4+, tetramethyl tetra-3,4-pyridinoporphyrinato zinc(II) [Zn(3,4-tmpnpa)]^4+, tetramethyl tetra-3,4-pyridinoporphyrinato iron(II) (Fe(2,3-tmpnpa))] as well as porphyrines ([meso-tetraphenylporphyrin] iron(III) chloride (Fe(III)TPPC), meso-tetrakis(4-chlorophenyl) porphyrin-manganese(III) acetate (Mn(III)(T(4-Cl))PP(OAC)), meso-tetrakis(4-hydroxyphenyl) porphyrin-manganese(III) acetate (Mn(III)(T(4-OH))PP(OAC)), meso-tetra...
tetrathiaporphyrin]-copper(II) (Cu(II)TPP) and [meso-tetraphenylporphyrin]-tin(II) (Sn(II)JTPP)) were synthesized using procedures have been reported previously [31–36]. The sterile thin cloth and three-layer medical masks were prepared from a mask production center in Tehran, Iran.

2.3. Study population

This study was performed in Baqiyatallah Hospital, Tehran, Iran between 2020 and 2021. Same as our previous study [30], the participants were selected from 21 to 80 years. 60 Covid-19 patients whose disease was confirmed by a pulmonologist, and the results of their chest x-ray and rRT-PCR tests were positive, were admitted to this study [30]. These patients did not take any medicine before the admission to the hospital. 55 volunteers of the other medical clinics were chosen as controls who had not been infected by Covid-19 before this study. 15 cured samples were invited to participate in the re-analysis after two months. All experiments were clearly explained to the volunteers. They were asked to sign an informed consent for participating in this study. The medical document of each individual was collected. The demographic information is summarized in Table S1 [30].

2.4. Sensor fabrication

To prepare the paper sensor, the desired pattern was first designed with AutoCAD software, and then printed on a filter paper substrate. The pattern shown in Fig. 1a consists of a square hydrophobic zone (black areas) and 32 circular hydrophilic zones (white areas). The hydrophobic areas were created by the penetration of the printer ink into the texture of the paper, while also blocking the paper holes [37]. To fabricate the sensor array, 0.2 µL of each color indicator (Fig. 1b) was dropped on a hydrophilic substrate (Fig. 1c). The location and contents of each sensing component are shown in Fig. 1b. The developed sensor was pasted onto the center of a plastic strip with size of 2.5 × 5.5 cm (see Scheme 1a-b). The sides of the strip were covered with double-sided adhesive (Scheme 1c). Note that the strip used had good flexibility and mechanical stability.

2.5. Colorimetric detection process

A sterile package containing a sensor array, a thin cloth, and a three-layer medical mask was given to each participant. First, the thin cloth mask was placed on the face (Scheme 1d). The double-sided adhesive was then removed (Scheme 1e). The sensor was stuck to the face mask (Scheme 1f), and covered with the three-layer medical mask to prevent the physical interference and environmental pollutions (Scheme 1g). The participant was asked to normally perform the inhalation and exhalation cycles for a certain period of time without any external force (Scheme 1h). During this time, the participant could continue his daily activities (except sleeping and eating). The exhaled breath metabolites directly interacted with the sensing components, resulting in discoloration of the indicators. These changes were observed by the naked eye (Scheme 1i). The color responses were recorded by a scanner, and analyzed by the image analysis software (Scheme 1j). For each sensing component, the outputs were three numerical values obtained from the difference of the mean values of R, G and B color elements. The difference value was calculated by subtracting the RGB values of the primary sensor from those of the reacted sensor. In total, a vector of 96 data points (32 sensing components × 3 color elements) was obtained for each participant.

2.6. Statistical evaluation of data

To verify the sensor ability to discriminate between the two studied classes of patient-healthy, patient-cured and healthy-cured categories, three matrices with sizes of (115 × 96), (75 × 96) and (70 × 96) were prepared, respectively. Statistical analysis of pattern recognition was performed by principle component analysis-linear discriminate analysis (PCA-LDA) in MATLAB environment. Statistical parameters such as the sensitivity, specificity, accuracy and error rate were calculated for all the three categories. To obtain the total sensor response for each participant, the norm of the data vector known as the Euclidean norm, was calculated as follows:

\[ \text{Euclidean Norm} = \sqrt{\sum_{i=1}^{n} (x_i)^2} \]

where \(x_i\) is the ith numerical value of the data vector. Equation (1) was also used to estimate the total response of a sensing component, involving a data vector with three values related to the RGB color elements.

Two independent sample t-tests were used to compare between the average values of the total sensor responses obtained from the patient and healthy groups. The correlation between the total response of the sensor (or the total response of a sensing component) and other parameters such as age, \(O_2\) saturation, disease severity and viral load value was investigated by the Pearson correlation coefficient. SPSS software was employed to run this statistical analysis.

3. Results and discussion

A sniffer was developed based on a colorimetric sensor array for the detection of exhaled breath volatile metabolites, according to the experimental section. The ability of the sensor to discriminate the patients with Covid-19 from the healthy people was evaluated. The following subsections describe the results of the assay in detail.

Fig. 1. (a) The schematic pattern of sensor designed by AutoCAD, (b) the names of sensing elements and their locations and (c) the image of fabricated sensor array.
3.1. Optimization

The array structure consists of the colorimetric components such as porphyrazines, organic dyes, porphyrins, metal ion complexes, and NPs. The organic dyes can also be combined with additives such as DWES, TBAOH, TsOH and PBA. The amount of each compound and the volume ratio of organic dye: additives in the resultant mixture can affect the sensor response. As well, the time required for the interaction between the metabolites and sensor components must be calculated. These evaluations were performed by optimization experiments in order to find maximum values of the total response of the sensor.

To find the optimal value of each indicator, four different experimental models were designed while Model 1 and Model 4 containing the lowest and highest amounts of colorimetric components, respectively (Fig. S1a). As can be seen in Fig. S1b, the strongest interaction between the metabolites and the sensor components, together with the best sensor response is achieved by using Model 3 for preparing the indicator solutions. The lower values of the indicator are not sufficient to complete the reaction, and the higher values prevent the monitoring of the changes arising from the interactions [38].

In the next step, the organic dyes were mixed with the additives in different volume ratios of (1:1), (2:1), (3:1), (4:1), and (5:1), as seen in Fig. S1c. The results presented in Fig. S1d show that the color changes of the sensing components become more intense with increasing the contribution of the organic dye in the mixture (thus decreasing the additive contribution). In this respect, the highest response is obtained using the volume ratio of (4:1). It is worth noting that the desired reaction does not occur at the lower amount of the additives, and the active sites of indicators are blocked using high amounts of these materials.

Volatile metabolites must be given time to distribute throughout the sensor surface, and interact with its sensing components. The analysis will be stopped once the color changes of all the indicators are fixed, being indicative of the completion of the reaction. Fig. S1e shows that 75 min is required to carry out a desirable experiment.

3.2. Sensor outputs

The discrimination between the patients with Covid-19 disease, the healthy individuals, and other people with non-Covid lung diseases arises from the changes of volatile compounds in the exhaled breath samples [14]. These chemicals are classified in the alkanes, alkenes, aldehydes, ketones, acids, alcohols, and arenes categories [14]. The array components interact with all the volatile metabolites through Lewis acid-base, Bronsted acid-base, electrostatic, H-bonding, charge transfer, π-π, dipole-dipole, and hydrophobic interactions, although color changes may be observed with different intensities depending on the type and amount of the chemical species [20]. Metalloporphyrins
and metallloporphyrinaztes participant in the Lewis acids- bases adduct formation, being attached to analytes by transferring the non-bonding electron pairs [39]. The response of these sensing elements is influenced by the chemical properties of the central metal, the strict hindrance of the macrocyclic aromatic structure, the polarity, chemical hardness, and affinity of the analytes [40]. Unlike porphyrins, the porphyrinaztes contain meso nitrogen atoms, and have a high potential for participation in nucleophilic and H-bonding reactions [41]. In other word, the presence of four electron acceptor pyridine rings in the structure of porphyrinaztes, dramatically increase the Lewis aci"y of their metal centers and conclusively, increasing their tendency for adduct formation by analyte. Organic dyes are proton donors or accep"tors whose color highly depends on the pH of media [42]. To increase reactivity, the organic dyes are mixed with specific additives. Among these substances, DWES tends to do a nucleophilic attack to the carbonyl species [43], whereas PBA binds to diols [44]. The products of both reactions change the concentration of H$_2$O$^-$. Alternatively, TsOH provides the conditions for the interaction between aniline-containing dyes and aldehydes through the Schiff test reaction [45]. Moreover, TBAOH is used to induce facile proton transfer reactions [46]. Metal ion complexes can be employed as sensing elements for metabolite detection. In this case, the analyte replaces the ligand in the complex structure (i.e., the indicator displacement assay) or is simultaneously attacked to the ligand and the metal ion, resulting in the formation of a ternary compound [47]. The stability constant of the metal-ligand complex, and the affinity of the metal ion to the analyte, are the two main factors for this interaction [48]. AuNPs can detect volatile species with a detection limit of 10 ppb [49]. The NP-based arrays provide the fingerprint patterns for chemical materials with different functional groups. The modification of NPs with various capping agents such as GA, BSA, and PVP allows them to interact with a wide range of analytes through electrostatic, H-bonding, and covalent interactions, leading to the color changes of the sensor from red to purple due to the NP aggregation [49].

The proposed sensor array contains 32 chemical compounds related to porphyrinaztes, porphyrins, the organic dyes mixed with DWES, TsOH, TBAOH and PBA, metal ion complexes, and AuNPs. The sensor was exposed to the volunteer exhaled breath metabolites, and the corresponding colorimetric responses were collected after 75 min. The results obtained by the colorimetric and the image analyses of patient, healthy and cured participants are shown in Fig. 2a. All observations are collected in Fig. S2-S4. Also, due to the interaction of the analyte with the sensing component, the partial color changes are clearly represented by the color difference maps (Fig. 2b).

Among all the array components, while S2 ([Zn(3,4-tmtppa)]$^{4+}$), S5 (Bromophenol red + DWES) and S25 (Cu(II)-Py) respond to the volatile metabolites of all the studied samples, no color changes are observed for S4 (Fe(II) (2,3-tmppa)), S7 (Acridine orange + DWES), S16 (Bromophenol blue + TBAOH), S20 (Cu (II) TPP), S21 (Sn (II) TPP), S24 (Fe (II) - Py), S26 (Ni (II)-Py), S28 (Bromopyrogallol red + PBA), S29 (Methyl blue + PBA), and all the NPs (S30–S32). It is indicated that the color of [Co (3,4-tmtppa)]$^{4-}$ (S1) and [Cu (2,3-tmppa)]$^{4-}$ (S3) is turned on in the colorimetric profiles of the patient and healthy samples, respectively. DWES-organic dyes, including bromocresol purple (S6), indigo carmine (S8), malachite green (S10), and phenol red (S11) have a high tendency towards volatile compounds in the patient’s exhaled breath. The markers such as toluidine blue (S9) and pararosaniline (S12) interact with the metabolites of healthy individuals. The dyes combined with TsOH (S13) tend to interact with volatile markers in healthy individuals, whereas TBAOH-integrated indicators (S14 and S15) become discolored in the presence of infected breath samples. Among porphyrins, only Fe(III) TPPCl (S18) responds to the patient metabolites. Both metalloporphyrinaztes with the central metal Mn (S17–S19), and the metal ion complexes of V (IV) and Fe(III) (S22 and S23) are illuminated in the color profile of the healthy individual. Although S27 (Bromocresol purple + PBA) is sensitive to the chemical species of the sample with Covid-19, other PBA-organic dyes are not influenced by the breath compositions of these studied groups.

In order to evaluate the treatment process, patients (15 samples) were asked to participate in this study once again after complete recovery. The test conditions were the same as before. As shown in Fig. 2, the sensor response to the volatile metabolites of the cured samples is different from that obtained by healthy and infected volunteers. The sensing components S1, S8, S11, S15, S18, and S27 are turned off in the color map of the cured samples. In this case, the metabolites responsible for changing the color of these sensors are possibly removed from the exhaled breath profile. However, one can observe the discoloration of S6, S10, and S14, indicating that the body’s metabolic behavior is still affected by the viral infection. Interestingly, some dedicated sensing elements for the healthy samples such as S12, S17, S19, S22 and S23 appear in the color profile of the cured participants, evidencing the progress in the treatment process as well as the accession of the normal situation.

![Fig. 2.](image_url)

(a) The colorimetric response and (b) the colorimetric difference maps of fabricated sensor for patient infected by Covid-19 (P), Healthy control (H) and Cured sample (C). The data was capture after 75 min and at the optimum conditions reported in Fig. S1.
3.3. Discrimination of population

The sensor efficiency in the creation of a difference pattern between the studied samples was evaluated using unsupervised pattern recognition methods such as PCA. For statistical analysis, three different sets, including patient-healthy, patient-cured, and healthy-cured were prepared by forming a data matrix with sizes of (115 × 96), (75 × 96), and (70 × 96), respectively. The score plots obtained by PCA analysis are summarized in Fig. 3, and the classification statistical parameters are collected in Table 1.

Fig. 3a shows the discrimination pattern obtained for the patient-healthy matrix data. The total variance extracted from the first two principal components (PCs) is 93.73%. Accordingly, 47 patients and 46 healthy people with a 95% confidence level are located in the correct sets, whereas 9 patients and 7 healthy samples are misdiagnosed. In total, 6 people (4 patients and 2 healthy samples) are not classified in any group. The parameters obtained from PCA show that the colorimetric method can discriminate the patients from healthy people with a sensitivity of 78.3%, and specificity of 83.6%. The accuracy of the method was calculated to be 80.8%.

The ability of the sensor to differentiate between patient and cured participants is shown in Fig. 3b. The first two PCs comprise 92.9% of the total explained variances. The results reveal that 47 patients and 13 cured samples are correctly diagnosed with a confidence level of 95%. However, 4 infected and 1 cured volunteers fall into the opposite categories. The sensor responses for 9 patients and 1 cured sample are similar to those for the healthy ones. Table 1 illustrates that the proposed method is able to differentiate between members of this set, having the sensitivity and specificity of 78.3% and 86.6%, respectively. In this case, the accuracy was found to be 80%.

For the last set (healthy-cured), the PCA diagrams depicted in Fig. 3c indicate that 91.9% of the total variance is distributed over the first two PCs. This method can classify 46 healthy and 13 cured people with a 95% confidence level. Among the remaining participants, 3 samples are clustered incorrectly, and 8 samples are not placed in the two classes. As seen in Table 1, the sensitivity, specificity and accuracy of this discrimination are equal to 83.6%, 86.6% and 84.3%, respectively.

It should be noted that the adverse sensor responses can be due to the errors in the sampling process, improper transfer of chemical markers from the mask cavities into the sensor texture and low concentration of the markers in the exhaled breath profile due to the mildness of the disease.

3.4. Diagnosis of disease severity

The participants were divided into the following 5 groups based on their disease severity: very mild, mild, moderate, severe, and very severe. The disease severity was determined by a pulmonologist using the vital signs, symptoms, chest imaging, and rRT-PCR results. The total responses of the sensor array and each sensing component were quantified. As illustrated in Fig. 4a, the color of the sensing element (S8) considerably changes by severity of the disease. As depicted in Table S2, an acceptable and significant relationship between these variables is observed due to the high Pearson coefficient (0.946) and low P (<0.001) values.

On the other hand, the relationship between the response of sensory components and viral load value (extracted from the rRT-PCR analysis based on the number of N gene cycle threshold (CT)) was investigated. To this end, the results of PCR were categorized in four sets: low, moderate, high and very high. As is clear in Fig. 4b and Fig. 4c, the response of sensing components (S15 and S27) shows an increasing trend with increasing the viral load value. The Pearson coefficients of 0.922 and 0.931 are obtained for S15 and S27, respectively, according to Table S2. As a result, the PCR analysis information can be qualitatively estimated using the proposed colorimetric sniffer.

3.5. Comorbidities detection

Volunteers may be smokers or have other non-Covid diseases such as cardiovascular disease, chronic kidney disease, asthma, diabetes, chronic obstructive pulmonary disease, chronic liver disease, and hypertension. Metabolites of these diseases can also appear in the breath profile.

Considering the previous studies, volatile compounds such as
acetonitrile and furan derivatives (for smokers) [50], amines and ammonia (for people with chronic kidney disease) [51], acetone, ethanol and methyl nitrate (for diabetics) [52], and isoprene, 4, 7-dimethyl -undecane, 2,6-dimethyl-heptane, acetaldehyde, 2-butyln octanol and methyl isobutyrate (for people with lung disorders) [53] have been reported as effective metabolites for differentiating between the relevant disease and normal cases. For participants of this study, the corresponding comorbidities are listed in Table S1. The developed sensor was exposed to the exhaled breath of people with these diseases. Fig. S5 shows the colorimetric response and colorimetric maps of patients with Covid-19, being smokers or having kidney, diabetes, or lung disorders. As can be seen, the interaction between the exhaled breath of people with diabetes and the sensor components results in the discoloration of S28 (Bromopyrogallol red + PBA). The kidney disease metabolites tend to interact with S16 (Bromophenol blue + TBAOH) and S31 (BSA-AnuNPs). Acidine orange + DWES (S7) responds to volatile compounds in smokers. Finally, the color of Sn (II) TPP (S21) and Fe (II)-Py (S 24) appears in the difference maps of participants with asthma and COPD. The diagnosis ability of the sensor for detection of different diseases is determined to be 85.0% for smokers, 86.6% for kidney disease, 75.0% for diabetes and 75.0% for lung disorders.

Consider that the metabolites of other diseases do not interact with sensing elements that respond to COVID-19 related markers. Also, the specified sensing components turned on in the presence of metabolites of a particular disease. Thus, the proposed sensor provide a unique response for a certain disease.

3.6. Statistical points

The Euclidean norm of the data vector provided by the image analysis was calculated as the total response for each studied sample. The mean of responses for each of patient and healthy groups was equal to 387.88 (± 33.11) and 361.29 (± 22.34), respectively. For all patient and healthy data, the total average was 375.17 (± 31.33). The results of two independent t-tests confirmed that the mean value obtained by the patient samples was higher than that by healthy ones. Thus, the difference obtained (26.59) was statistically significant (P-value < 0.001). It can also be concluded that the Euclidean norm of higher than the total average value (375.17) is indicative of the patient sample, whereas the response of the sensor for a healthy participant has a lower value than the total average.

Furthermore, the effect of age and O2 saturation was investigated on the sensor response. The results of the correlation test between the total response and age of volunteers (Table S2) show that the value of Pearson correlation coefficient is calculated to be 0.024 for patients, and 0.061 for healthy individuals. The corresponding P-values are obtained to be 0.855 and 0.657, giving rise to the lack of a significant relationship between the two variables. From Table S3, no strong and significant correlation is observed between the total response and O2 saturation of all the volunteers. The Pearson correlation coefficient and P-value are 0.117 and 0.395 for the patient group, and 0.178 and 0.192 for the healthy samples. These results explain that the colorimetric response depends on the metabolic changes of the exhaled breath, and is not affected by other variables.

4. Conclusions

For the first time, the diagnosis of a patient with Covid-19 has been performed based on the analysis of exhaled breath metabolites using a colorimetric sniffer. Compared to standard detection methods, the proposed assay was user-friendly because of its simplicity, and low-cost design and fabrication process, as well as being portable and capable of on-site detection. By this method, the volatiles were directly exposed to the sensor without the need of separate and tedious sampling process, providing more desirable sensitivity compared to the PCR method. Unlike the current diagnostic methods, the proposed assay showed high ability to discriminate the cured samples from healthy people, and to detect the comorbidities such as kidney disease, diabetes, and lung disorders, together with their disease severity. This sensor can be easily used in hospitals, medical laboratories, homes, shops, factories and public transportation to identify the disease vectors. The design of the colorimetric breath sensors with the same experimental procedure for detection of other diseases is underway in our research group.

CRediT authorship contribution statement

Mohammad Mahdi Bordbar: Project administration, Methodology, Investigation, Formal analysis, Writing – original draft. Hosein Sama: Conceptualization, Design, Validation, Resources. Ali Hajian: Methodology, Validation, Writing – review & editing. Azarmidokht Sheini: Methodology, Validation. Elham Safaei: Conceptualization, Methodology, Validation. Jasem Aboonajmi: Methodology, Validation. Fabiana Arduini: Conceptualization, Writing – review & editing. Hashem Sharghi: Conceptualization, Validation. Pegah Hashemi: Methodology, Validation. Hosein Khoshsafar: Methodology, Validation. Mostafa Ghanei: Conceptualization, Methodology, Validation, Resources. Hasan Bagheri: Supervision, Conceptualization, Methodology, Writing – review & editing.

Data availability

Data will be made available on request.

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Declaration of competing interest

The authors declare the following competing financial interest(s): Five authors (Mohammad Mahdi Bordbar (M.M.B.), Pegah Hashemi (P.H.), Hosein Khoshsafar (H.Kh.), Mostafa Ghanei (M.Gh) and Hasan Bagheri (H.B)) have filed a provisional patent application on the technology described in this manuscript entitled “A colorimetric sniffer for qualitative and quantitative detection of Covid-19 disease”. The remaining authors declare that they have no competing interests.

Compliance with Ethical Standards

The research ethics committee of Baqiyatallah University of Medical Sciences has approved the project (Approval ID: IR.BMSU.REC.1399.508).

Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.snb.2022.132379.

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