RESEARCH ARTICLE

Tumor and Plasma Met Levels in Non-Metastatic Prostate Cancer

Deborah R. Kaye1, Peter A. Pinto1, Fabiola Cecchi1, Joseph Reilly1, Alice Semerjian1, Daniel C. Rabe1, Gopal Gupta1, Peter L. Choyke2, Donald P. Bottaro1*

1 Urologic Oncology Branch, Center for Cancer Research, National Cancer Institute, Bethesda, MD, 20892, United States of America
2 Molecular Imaging Program, Center for Cancer Research, National Cancer Institute, Bethesda, MD, 20892, United States of America

Current address: The James Buchanan Brady Urological Institute, The Johns Hopkins University, 600 North Wolfe Street, Baltimore, Maryland, 21287, United States of America
Current address: NantOmics, 9600 Medical Center Drive, Suite 300, Rockville, MD, 20850, United States of America
Current address: George Washington University Medical Faculty Associates, 2150 Pennsylvania Ave NW, Washington, DC, 20037, United States of America
Current address: The University of Chicago, Biological Sciences Division, 924 E. 57th Street, BSLC, Suite 104, Chicago, IL, 60637, United States of America
Current address: Department of Urology, Loyola University Medical Center, 2160 S. First Avenue, Building 54, Room 237A, Maywood, IL, 60153, United States of America

* dbottaro@helix.nih.gov

Abstract

Objective

to measure Met protein content in prostate biopsies guided by fused magnetic resonance and ultrasound imaging, and to measure soluble Met (sMet) protein concentration in plasma samples from patients presenting evidence of prostate cancer.

Patients and Methods

345 patients had plasma samples drawn prior to image-guided biopsy of the prostate. Of these, 32% had benign biopsies. Of the 236 that were positive for prostate adenocarcinoma (PCa), 132 treated by total prostatectomy had Gleason scores of 6 (17%), 7 (55%), 8 (16%), or 9–10 (12%). 23% had evidence of local invasion. Plasma samples were also obtained from 80 healthy volunteers. Tissue Met and plasma sMet were measured by two-site immunoassay; values were compared among clinically defined groups using non-parametric statistical tests to determine significant differences or correlations.

Results

PCa tumor Met correlated significantly with plasma sMet, but median values were similar among benign and malignant groups. Median plasma sMet values were also similar among those groups, although both medians were significantly above normal. Median Met content in primary PCa tumors and sMet concentrations were independent of Gleason score, final pathologic stage and age.
Conclusion

Plasma sMet is not predictive of PCa or its severity in patients with organ-confined or locally invasive disease. Quantitative analysis of Met protein content and activation state in PCa tumor biopsy samples was highly feasible and may have value in follow-up to genomic and/or transcriptomic-based screens that show evidence of oncogenically relevant MET gene features that occur at relatively low frequency in non-metastatic PCAs.

Introduction

Prostate cancer (PCa) is the second most common cancer in men and second most common cause of cancer deaths of men in the United States [1]. Over the last two decades, serum prostate specific antigen (PSA) levels have been used to screen for PCa, to predict disease recurrence after therapy, and to predict metastasis; however, the value of this test for screening and outcome prediction remains controversial [2]. Discordance between PSA level and PCa includes cases where patients with metastatic disease have undetectable PSA levels [3], patients with elevated PSA levels post-treatment who never develop metastatic disease [4], and patients for whom PSA values either do not directly correlate with, or are inversely correlated with, disease progression [4]. Although elevated PSA levels resulting from architectural distortion and low levels due to impaired PSA secretion by tumor cells explain some of these discrepancies [5, 6], concerns about the reliability of PSA for monitoring disease progression provide a strong rationale for identifying alternative markers of disease progression, particularly those that are therapeutically targetable.

The hepatocyte growth factor (HGF) receptor tyrosine kinase, known as Met, is overactivated, overproduced and/or mutated in a wide spectrum of prevalent malignancies [7]. Evidence of HGF/Met signaling in PCa is growing. In prostatectomy specimens, Met protein abundance as determined by immunohistochemical (IHC) staining correlated directly with progressive disease [8] and Met positivity was found in a clear majority of [8, 9], or in all of [10, 11], bone metastases samples examined. Consistent with these observations, Met expression in cultured PCa-derived cell lines was found to be inversely related to androgen receptor expression [8, 12], and phase II clinical trials of cabozantinib, a multikinase inhibitor with potent activity against Met, have reported frequent resolution of bone metastases in PCa patients with advanced disease [13–18]. At least 25 experimental therapeutic agents targeting the HGF/Met pathway have been or are currently being tested in over 200 human clinical trials, and the urgent need for biomarkers of oncogenic HGF signaling is becoming widely recognized [7]. HGF concentration was reported to be significantly elevated in the serum of metastatic PCa patients when compared with patients with localized cancer [19], elevated plasma HGF correlated with decreased survival for patients with hormone refractory metastatic PCa [20], and plasma HGF was found to be an independent predictor of PCa metastasis to lymph nodes and disease recurrence after surgery [21]. Our prior studies of another potential surrogate of HGF signaling, soluble Met ectodomain (sMet), showed that sMet levels in plasma and urine correlated directly with burden of PCa tumor xenografts in mice [22], and that urinary sMet was significantly higher in metastatic PCa patients relative to patients with organ-confined disease or controls not known to have any form of cancer [23].

The present study extends our prior work by adapting the two-site electrochemiluminescent sMet immunoassay for use on detergent extracts prepared from flash frozen primary PCa tumor biopsies, and correlates these measurements with PCa stage, grade, Gleason score and
plasma sMet levels. Since the advent of PSA screening, the pathologic diagnosis of PCa has been based on the use of systematic trans-rectal ultrasound guided biopsies. Advances in multiparametric magnetic resonance imaging (MRI) have allowed high quality visualization of the prostate and can aid in identification of prostate cancer lesions. We have previously reported on the use of a novel MRI and ultrasound (US) fusion biopsy system for the targeted detection of lesions detected on MRI [24, 25]. Taking advantage of this system allowed more efficient sampling of primary PCa tumor tissue to determine absolute Met protein abundance normalized to total sample protein content.

Materials and Methods

Patients and Samples
From 2009 to 2012, primary prostate tumor biopsy specimens and plasma samples were collected from patients who underwent MRI-ultrasound fusion guided prostate biopsy and from patients who proceeded to have radical prostatectomy at the National Cancer Institute (NCI). All patients were enrolled in a NCI Center for Cancer Research (CCR) Institutional Review Board (IRB) approved tissue procurement protocol (NCI-97-C-0147) at the National Institutes of Health (NIH) Clinical Center by informed written consent. Consent documentation records are maintained by the NCI/CCR/IRB and the NIH Clinical Center Office of Protocol Services for a minimum of three years after the completion of the research. Tissue specimens were obtained by MRI-ultrasound fusion guided biopsy using a method reported previously [24, 25]. Serial biopsies from selected sites provided specimens for pathology and protein analysis; the latter were flash frozen on dry ice within 2 minutes of procurement and stored at 80°C prior to analysis. Met and phosphoMet (pMet) protein abundance were measured for biopsy samples selected on the basis of total protein content needed for repeated analysis (14 malignant and 8 benign samples). Tumor tissue samples from 16 patients with pathologically confirmed renal cell carcinoma of various histologies and enrolled in a NCI IRB approved tissue procurement protocol were obtained intraoperatively and processed and stored similarly. Plasma samples were obtained by conventional methods prior to prostatectomy using EDTA as anticoagulant and stored at 80°C prior to analysis. Only patients with a final pathological determination of prostate cancer stage (in those who underwent prostatectomy) and Gleason score were included in the malignant group; all others not known to have any other malignancy were included in the benign group. Plasma samples were also obtained from 80 healthy male volunteers through the NIH Healthy Volunteers program.

Analysis of Tissue Met and Plasma sMet
Met, phosphotyrosyl-Met, and soluble Met ectodomain (sMet) proteins were measured in tissue and plasma samples as described previously [22]. Briefly, streptavidin-coated 96-well plates were blocked, washed with PBS and coated with a biotin-tagged, affinity-purified human Met ectodomain-specific capture antibody (BAF358, R&D Systems). Wells were washed again before adding sample or sMet standard (358-MT, R&D Systems) for 1 h with shaking. After washing with PBS, detection antibody (AF276, R&D Systems) labeled with MSD-Sulfo-Tag (Meso Scale Discovery, Gaithersburg, MD) was added for 1 h with shaking. Wells were washed with PBS before adding read buffer and plates were read in a Meso Scale Discovery SectorImager 2400. Tissue samples were extracted with ice-cold detergent containing buffer with physical disruption with 0.5mm glass beads and shaking in a bead beater (Mini Bead Beater 8, BioSpec Products, Inc., Bartlesville, OK) as described previously [26]. Assays were performed blinded to study endpoint.
Statistical Analyses

Absolute plasma sMet concentration and tissue Met content (corrected to total extracted protein) were compared among groups defined by final pathologic stage (in those who underwent prostatectomy) and Gleason score. Statistical analyses were performed using JMP v10.0.2 (SAS Institute Inc.), Excel 2008 for Mac (Microsoft Inc.) and Prism v6.0f (GraphPad Software Inc.) software packages. Wilcoxon, Mann Whitney and Kruskal-Wallis tests, in conjunction with the Bonferroni multiple test correction where necessary, were used to determine statistical differences between groups. Receiver Operating Curves (ROC) were calculated and Area Under the Curve (AUC) determined using JMP. Cutoff values determined using a 45° line drawn tangential to the ROC curve were used to develop specificity, sensitivity and prediction values.

Reverse-phase protein array (RPPA) and clinical data for a cohort of 352 cases in the Prostate Adenocarcinoma (PRAD) TCGA Provisional data set were obtained from the M.D. Anderson Reverse Phase Protein Array Core. RPPA scores for Met and phosphoMet (pMet) content were analyzed for relationships with T stage, Gleason score and PSA using Prism v6.0f (GraphPad Software Inc.) software. Within the 352 case cohort, Gleason score was available for 352, clinical T stage data for 348, PSA data for 318, clinical N stage data for 309 and clinical M stage data for 327. Of the latter, all but one case were M0, so correlative tests related distant metastases were not performed.

Results and Discussion

Demographic information, medical history, relevant laboratory values and pathology data were obtained for each patient. As shown in Table 1, a total of 345 patients, ages 38–60 yrs, were biopsied; 236 were positive for prostate adenocarcinoma (malignant group), 132 of whom were treated by total prostatectomy. Patients with negative biopsy results (109) with no other known cancer were considered benign. Total prostatectomy specimens from the malignant group ranged in Gleason score from 3+3 = 6 (22/17%) to a group with Gleason score totals of 9 or 10 (16/12%). Local invasion and/or regional lymph node involvement was found in 30 patients (23%) and distant metastases in 7 (5%). Median tissue Met content measured in 8 patients with negative biopsy and 14 with positive biopsy were not significantly different (Mann-Whitney p = 0.4927; Fig 1A). Both median values were significantly lower than the median value obtained for another prevalent cancer of the urinary tract, renal cell carcinoma (RCC; Kruskal-Wallis p < 0.0001; Fig 1A). These findings are consistent with the relative

| Clinical/Pathological Feature                          | Number of Patients (%) |
|--------------------------------------------------------|------------------------|
| Total Number of Patients Biopsied/Treated              | 345                    |
| Median Age (Range)                                     | 60 yrs (38–81)         |
| Negative Targeted Biopsy (benign)                     | 109 (32%)              |
| Positive Biopsy (malignant)                           | 236 (68%)              |
| Total Prostatectomy Specimen                          | 132                    |
| Gleason 6                                              | 22 (17%)               |
| Gleason 7                                              | 73 (55%)               |
| Gleason 8                                              | 21 (16%)               |
| Gleason 9 or 10                                        | 16 (12%)               |
| Organ Confined                                         | 95 (72%)               |
| Local Invasion, Regional Lymph Nodes                   | 30 (23%)               |
| Metastatic                                             | 7 (5%)                 |

doi:10.1371/journal.pone.0157130.001
frequencies of MET gene amplification (determined using GISTIC) and/or mRNA overexpression (2-fold or greater as determined by RNA Seq V2 RSEM) obtained using the cBioPortal \cite{27, 28} to analyze the provisional prostate adenocarcinoma (PRAD) TCGA data set (22/486 cases or 4.5%) vs. clear cell RCC (53/531 cases or 10%). Phosphotyrosyl-Met content was undetectable in these tissue samples, which is likely to reflect minimal receptor activation, since maximal receptor activation stimulated in vitro by the addition of ATP to several of these samples was readily detected (data not shown). Plasma sMet concentration was significantly
correlated with primary tumor Met content in the 14 malignant PCa cases (Spearman 
$r = 0.7143, p = 0.0001$; Fig 1B), consistent with a relationship between Met shedding and malignancy reported previously [22].

As for tissue Met, median plasma sMet values in benign ($n = 109$) and malignant ($n = 236$) patient groups were not significantly different (Mann-Whitney $p = 0.0575$); however, both were significantly higher than the median of a healthy volunteer group (normal; $n = 80$; Spearman $r = 0.2939, p < 0.0001$; Fig 1C). Receiver Operating Characteristic (ROC) analysis of malignant vs. normal groups using a threshold value of 146 ng/ml yielded 79% sensitivity and 94% specificity values (AUC = 0.8309, $p < 0.0001$), but without understanding the basis of elevated sMet levels in benign patients, plasma sMet level appears to have no predictive value for PCa malignancy. A relevant common feature among all patients presenting to the NCI for PCa diagnosis and management is elevated PSA. The median PSA value in the benign cohort was 4.65 ng/ml, significantly above the widely accepted 4.0 ng/ml threshold for prostate biopsy (Wilcoxon Signed Rank Test two-tailed $p < 0.0001$). No correlation was found, however, between plasma sMet concentration and PSA concentration among 274 patients (including benign and malignant cases) whose samples for both tests were drawn at the same time (Spearman $r = 0.05492, p = 0.3652$; Fig 1D). In addition, no significant differences or trends in median sMet concentrations were found on the basis of Gleason score, organ-confined disease, locally invasive disease, or metastasis (Table 2). No correlations were found between plasma sMet vs. age or pathological grade or stage (data not shown).

Reverse-phase protein array (RPPA) analysis results for Met content in a subset (352) of primary tumor samples in the PRAD TCGA data set were negatively correlated with Gleason score ($p = 0.0061$; Table 3 and Fig 2B), clinical T stage ($p = 0.0050$; Table 3 and Fig 2A) and clinical N stage ($p = 0.0102$, Table 3). These findings are consistent with the statistically non-significant trends we observed in sMet values among T/N stage or Gleason subgroups (not shown). RPPA values for pMet content were not significantly correlated with either Gleason score or T/N stage (Table 3 and Fig 2). Met and pMet scores were also not related to one

### Table 2. Group plasma sMet medians, ranges, 95% confidence intervals and comparison to normals (shaded) by Wilcoxon Signed Rank Test.

| Group                  | Plasma sMet (ng/ml) | Median 95% Confidence Interval | Wilcoxon Signed Rank Test |
|------------------------|---------------------|--------------------------------|---------------------------|
|                        | Median | Range    | Actual % Confidence | Lower limit | Upper limit | $W^1$ | $P$ value$^2$ |
| Healthy Volunteer      | 98     | 45–471   | 97                 | 90          | 106         | 5735 | < 0.0001    |
| Benign                 | 300    | 45–3341  | 97                 | 249         | 367         | 7720 | < 0.0001    |
| Malignant              | 239    | 36–3839  | 96                 | 221         | 278         | 26191| < 0.0001   |
| Prostatectomy Specimen | 224    | 36–3034  | 96                 | 199         | 253         | 7720 | < 0.0001    |
| Gleason 6              | 272    | 69–577   | 98                 | 177         | 372         | 249  | < 0.0001    |
| Gleason 7              | 208    | 36–951   | 97                 | 171         | 259         | 2249 | < 0.0001    |
| Gleason 8              | 224    | 42–534   | 97                 | 179         | 323         | 201  | 0.0001      |
| Gleason 9–10           | 257    | 58–425   | 98                 | 107         | 426         | 120  | 0.0008      |
| Organ confined         | 222    | 36–737   | 96                 | 180         | 260         | 4133 | < 0.0001    |
| Local invasion$^2$     | 239    | 44–3034  | 96                 | 199         | 323         | 415  | < 0.0001    |
| Metastatic             | 189    | 58–633   | 98                 | 58          | 633         | 22   | 0.0781      |

$^1$Sum of signed ranks

$^2$Includes regional lymph node involvement

$^3$Two-tailed

doi:10.1371/journal.pone.0157130.t002
Table 3. Spearman correlation analysis of Met content, pMet content, Gleason score and clinical T and N stage for patient tumor samples¹ in the prostate adenocarcinoma (PRAD) TCGA Provisional dataset.

| Parameter | Met² vs Gleason³ | pMet² vs Gleason³ | Met vs T stage⁴ | pMet vs T stage⁴ | Met vs N stage⁵ | pMet vs N stage⁵ |
|-----------|------------------|------------------|-----------------|-----------------|-----------------|-----------------|
| Number of XY Pairs | 352 | 352 | 348 | 348 | 309 | 309 |
| Spearman r | -0.1458 | 0.03617 | -0.1503 | 0.07766 | -0.1460 | 0.08555 |
| 95% confidence interval | -0.2495 to -0.07173 to 0.1432 | -0.2544 to -0.04278 | -0.2565 to 0.1843 | -0.03163 | -0.02962 to 0.1985 |
| P value (two-tailed) | 0.0061 | 0.4988 | 0.0050 | 0.1483 | 0.0102 | 0.1335 |

¹All tissue samples in the PRAD TCGA RPPA Provisional cohort (n = 352) are primary prostate tumor samples
²Content scores determined by RPPA analysis
³N values for each Gleason score subgroup, from 6 to 10, are shown in Fig 2B
⁴N values for each T stage subgroup, from T2a to T4, are shown in Fig 2A; stage data for 4 cases was unavailable
⁵The N0 subgroup contained 246 cases, the N1 subgroup, 62 cases

doi:10.1371/journal.pone.0157130.t003

<Fig 2. A.> Box and whisker plots of Met (upper panel) and pMet content (lower panel; RPPA score, log₂) across clinical T stage subgroups of primary prostate adenocarcinoma biopsies that were obtained as part of the PRAD TCGA Provisional data set. A statistically significant inverse correlation between Met content and higher T stage was found (Table 3). B. Box and whisker plots of Met (upper panel) and pMet content (lower panel; RPPA score, log₂) across clinical Gleason score subgroups of primary prostate adenocarcinoma biopsies that were obtained as part of the PRAD TCGA Provisional data set. A statistically significant inverse correlation between Met content and higher Gleason score was found (Table 3).

doi:10.1371/journal.pone.0157130.g002
another or to PSA (not shown). In these contexts the PRAD TCGA RPPA data for Met and pMet are generally consistent with the findings of our patient study.

Conclusions

Plasma sMet concentration is not predictive of PCa or its severity in patients with organ-confined or locally invasive disease. Analysis of Met protein content and activation state in primary PCa tumor samples obtained by image-guided biopsy as described here was highly feasible and quantitative. The low incidence of MET gene amplification, mutation and/or overexpression in primary prostate tumor samples analyzed by TCGA suggests that Met protein quantitation and activation status may have value in follow-up to genomic and/or transcriptomic-based screens that show evidence of these oncogenically relevant features, to more directly guide therapeutic strategy and for pharmacodynamic evaluation of Met-targeted therapeutics. We believe these findings have value in (1) informing others compelled to do similar work so that resources are used optimally; (2) showing the feasibility and precision of the methods and measurements made in tissue samples obtained with MRI/US-fusion guidance; and (3) improving our understanding of extent and nature of HGF/Met signaling pathway involvement in localized prostate cancer.

Acknowledgments

This work was supported by the Intramural Research Program of the NIH, National Cancer Institute, Center for Cancer Research. We thank Alessio Giubellino for contributions to Met measurements reported here.

Author Contributions

Conceived and designed the experiments: DRK PAP PLC DPB. Performed the experiments: DRK FC JR DCR. Analyzed the data: DRK FC JR AS DCR DPB. Contributed reagents/materials/analysis tools: PAP GG PLC DPB. Wrote the paper: DRK PAP FC AS DCR PLC DPB.

References

1. Jemal A, Siegel R, Ward E, Murray T, Xu J, Thun MJ. Cancer statistics, 2007. CA Cancer J Clin 2007; 57: 43–66. PMID: 17237035
2. Bensalah K, Lotan Y, Karam JA, Shariat SF. New circulating biomarkers for prostate cancer. Prostate Cancer Prostatic Dis 2008; 11:112–20. PMID: 17998918
3. Leibovici D, Spiess PE, Agarwal PK, Tu SM, Pettaway CA, Hitzhusen K, et al. Prostate cancer progression in the presence of undetectable or low serum prostate-specific antigen level. Cancer 2007; 109: 198–204. PMID: 17171704
4. Fall K, Garmo H, Andren O, Bill-Axelson A, Adolfsson J, Adami HO, et al. Prostate-specific antigen levels as a predictor of lethal prostate cancer. J Natl Cancer Inst 2007; 99:526–32. PMID: 17405997
5. Sandblom G, Ladievardi S, Garmo H, Varenhorst E. The impact of prostate-specific antigen level at diagnosis on the relative survival of 28,531 men with localized carcinoma of the prostate. Cancer 2008; 112: 813–9. PMID: 18098207
6. Alhara M, Lebovitz RM, Wheeler TM, Kinner BM, Ohori M, Scardino PT. Prostate specific antigen and gleason grade: an immunohistochemical study of prostate cancer. J Urol 1994; 151: 1558–64. PMID: 7514688
7. Cecchi F, Rabe DC, Bottaro DP. Targeting the HGF/MET signaling pathway in cancer therapy. Expert Opinion on Therapeutic Targets 2012; 16: 553–72. doi: 10.1517/14728222.2012.680957 PMID: 22530990
8. Humphrey PA, Zhu X, Zarnegar R, Swanson PE, Ratliff TL, Vollmer RT, et al. Hepatocyte growth factor and its receptor (c-MET) in prostatic carcinoma. Am J Pathol 1995; 147: 386–96. PMID: 7639332
9. Pisters LL, Troncoso P, Zhou HE, Li W, von Eschenbach AC, Chung LW. c-Met proto-oncogene expression in benign and malignant human prostate tissues. J Urol 1995; 154: 293–8. PMID: 7539865
10. Knudsen BS, Gmyrek GA, Inra J, Scherr DS, Vaughan ED, Nanus DM, et al. High expression of the Met receptor in prostate cancer metastasis to bone. Urology 2002; 60: 1113–7. PMID: 12475693

11. Watanabe M, Fukutome K, Kato H, Murata M, Kawamura J, Shiinaishi T, et al. Progression-linked overexpression of c-Met in prostatic intraepithelial neoplasia and latent as well as clinical prostate cancers. Cancer Lett 1999; 141: 173–8. PMID: 10454259

12. Verras M, Lee J, Xu H, Li TH, Wang Y, Sun Z. The androgen receptor negatively regulates the expression of c-Met: implications for a novel mechanism of prostate cancer progression. Cancer Res 2007; 67: 967–75. PMID: 17283128

13. Smith DC, Smith MR, Sweeney C, Elfiky AA, Logothetis C, Corn PG, et al. Cabozantinib in patients with advanced prostate cancer: results of a phase II randomized discontinuation trial. J Clin Oncol 2013; 31: 412–9. doi: 10.1200/JCO.2012.45.0494 PMID: 23169517

14. Lee RJ, Saylor PJ, Michaelson MD, Rothenberg SM, Smas ME, Miyamoto DT, et al. A dose-ranging study of cabozantinib in men with castration-resistant prostate cancer and bone metastases. Clin Cancer Res 2013; 19: 3088–94. doi: 10.1158/1078-0432.CCR-13-0319 PMID: 23553848

15. Clyne M. Prostate cancer: cabozantinib reduces pain and narcotic use in men with CRPC. Nat Rev Urol 2014; 11: 188.

16. Basch E, Autio KA, Smith MR, Bennett AV, Weitzman AL, Scheffold C, et al. Effects of Cabozantinib on Pain and Narcotic Use in Patients with Castration-resistant Prostate Cancer: Results from a Phase 2 Nonrandomized Expansion Cohort. Eur Urol 2015; 67: 310–8 doi: 10.1016/j.eururo.2014.02.013 PMID: 24631409

17. Gartrell BA, Saad F. Managing bone metastases and reducing skeletal related events in prostate cancer. Nat Rev Clin Oncol 2014; 11: 335–45. doi: 10.1038/nrclonc.2014.70 PMID: 24821212

18. Graham TJ, Box G, Tunariu N, Crespo M, Spinks TJ, Miranda S, et al. Preclinical evaluation of imaging biomarkers for prostate cancer bone metastasis and response to cabozantinib. J Natl Cancer Inst 2014; 106: dju033. doi: 10.1093/jnci/dju033 PMID: 24634505

19. Naughton M, Picus J, Zhu X, Catalona WJ, Vollmer RT, Humphrey PA. Scatter factor-hepatocyte growth factor elevation in the serum of patients with prostate cancer. J Urol 2001; 165: 1325–8. PMID: 11257710

20. Humphrey PA, Halabi S, Picus J, Sanford B, Vogelzang NJ, Small EJ, et al. Prognostic significance of plasma scatter factor/hepatocyte growth factor levels in patients with metastatic hormone-refractory prostate cancer: results from cancer and leukemia group B 150005/9480. Clin Genitourin Cancer 2006; 4: 269–74. PMID: 16729910

21. Gupta A, Karakiewicz PI, Roehrborn CG, Lotan Y, Zlotta AR, Shariat SF. Predictive value of plasma hepatocyte growth factor/scatter factor levels in patients with clinically localized prostate cancer. Clin Cancer Res 2006; 12: 4154–62. PMID: 16857786

22. Athauda G, Giubellino A, Coleman JA, Horak C, Steeg PS, Lee MJ, et al. c-Met ectodomain shedding rate correlates with malignant potential. Clin Cancer Res 2006; 12: 4154–62. PMID: 16857786

23. Russo AL, Jedlicka K, Wernick M, McNally D, Kirk M, Sproull M, et al. Urine analysis and protein networking identify Met as a marker of metastatic prostate cancer. Clin Can Res 2009; 15: 4292–4298.

24. Turkbey B, Xu S, Krukecker J, Locklin J, Pang Y, Bernardo M, et al. Documenting the location of prostate biopsies with image fusion. BJU Int 2011; 107: 53–7. doi: 10.1111/j.1464-410X.2010.09483.x PMID: 20590543

25. Turkbey B, Xu S, Krukecker J, Locklin J, Pang Y, Shah V, et al. Documenting the location of systematic transrectal ultrasound-guided prostate biopsies: correlation with multi-parametric MRI. Cancer Imaging 2011; 11: 31–6. doi: 10.10217/730.2011.0007 PMID: 21450549

26. Cecchi F, Pajalunga D, Fowler CA, Peruzzi B, MacDonald N, Blackman DK, et al. Hepatocyte and vascular endothelial cell growth factor antagonists engineered by targeted disruption of heparan sulfate binding. Cancer Cell 2012; 22: 250–262.

27. Cerami E, Gao J, Dogrusoz U, Gross BE, Sumer SO, Aksoy BA, et al. The cBio cancer genomics portal: an open platform for exploring multidimensional cancer genomics data. Cancer Discov 2012; 2: 401–4. doi: 10.1158/2159-8290.CD-12-0095 PMID: 22588877

28. Gao J, Aksoy BA, Dogrusoz U, Dresdner G, Gross B, Sumer SO, et al. Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. Sci Signal 2013; 6: p11. doi: 10.1126/scisignal.2004088 PMID: 23550210