In Vivo Recognition of Vascular Structures by Near-Infrared Transillumination †

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Abstract: Transillumination is a very well-known non-invasive optical technique that relies on the use of non-ionizing radiation to obtain information about the internal morphology of biological tissues. In a previous work, we implemented a laser-based illuminator operating at a wavelength of 850 nm, combined with a CMOS digital camera and narrow-band optical detection that showed great potential for in vivo imaging. A great advantage is the use of low-cost semiconductor lasers, driven by a very low current (about 11 mA, spatially distributed as a 6-by-6 matrix covering a 25 cm² area). Thanks to the strong absorption of hemoglobin at this wavelength, we have collected raw data of vascular structures that have been further processed to achieve images with much better quality. In particular, here we show that a higher contrast can be attained by the expansion of gray level histograms to exploit the full range, 0–255. This elaboration can be, for instance, exploited for the recognition of vascular structures with better resolution. Examples are reported relative to hand dorsal vein patterns and live chick embryos’ blood vessels. Analyses can be successfully performed without applying any thermal or mechanical stress to the human tissue under test and without damaging or puncturing any parts of the eggshell.

Keywords: transillumination; in vivo imaging; chick embryo; hand dorsal vein pattern; VCSEL

1. Introduction

Optical transillumination is a non-invasive method for imaging that allows the investigation of the internal structure of thin portions of biological tissues [1–4]. It relies on the use of non-ionizing radiation, resulting, thus, in a totally safe diagnostic tool. The transillumination analysis consists of illuminating the sample with a light source and collecting the radiation that is transmitted through the tissue under test. The propagation of the photons is conditioned by absorption, scattering, and reflection effects taking place inside the tissue: hence, the acquired images carry important information about the morphology and the health condition of the sample. In biological tissues, absorption effects are mainly due to water; macromolecules, such as proteins and lipids; and pigments, such as hemoglobin [5,6]. In particular, in the wavelength range from 600 to 1200 nm (the so-called “diagnostic and therapeutic window”) the absorption of water is much lower than that of oxygenated and deoxygenated hemoglobin. The choice of light sources emitting in this spectral region produces results that are particularly interesting when studying highly perfused tissues. As it concerns scattering, only a small amount of light is redirected but this phenomenon still prevents the formation of high-definition images comparable with the results from more complex diagnostic techniques that make use of ionizing radiations. Nevertheless, transillumination can be exploited as
a first approach to perform preliminary analysis, in place of more complex and invasive tests [7,8]. Hence, it is applied for the investigation of hydroceles, hydrocephalus, caries, malignant lumps, and blood vessel patterns. Commercial medical devices are available: they are based on light-emitting diodes (LEDs) emitting visible light and they can be used only in a darkened environment [9,10]. Transillumination is also exploited for photoplethysmography (PPG), a non-invasive optical method that allows reconstruction of a signal related to the change of blood volumes inside the blood vessels of the tissue under test. The PPG signal obtained with pulse oximetry looks similar to the arterial pressure wave, but its waveform appears distorted. This happens because the PPG analysis is carried out by applying the sensor typically to a fingertip or to an earlobe that is subjected to mechanical pressure. This stress activates alpha adrenergic receptors which affect the arteries and veins vasoconstriction (narrowing the blood vessels).

To solve this issue and in view of the technological advancement done in the past years in the fabrication of light sources and detectors operating in the wavelength range 800–1000 nm, in previous work we proposed and demonstrated a portable vertical cavity surface emitting laser (VCSEL)-based instrumental setup for morpho-functional imaging of in vivo biological tissues [11,12]. We employed the optoelectronic system to acquire pictures and videos of human hands and chick embryos inside eggshells and to extract vital sign information. In this manuscript, we show how the raw images can be further processed (by expansion and equalization of the gray level histograms) in order to obtain better quality images with higher contrast, for a more precise identification of blood vessel patterns.

2. Materials and Methods

The optoelectronic transillumination system for in vivo imaging featured an illuminator composed of 36 VCSELs arranged in a 6-by-6 matrix covering an area of 25 cm². Their pumping current was driven by a custom-designed circuit. A digital CMOS camera was employed for image acquisition (Figure 1).

![Figure 1. Portable optoelectronic configuration for transillumination-based imaging. Abbreviations: VCSEL—vertical cavity semiconductor emitting laser; NIR—near-infrared.](image-url)
In particular, the employed VCSELs (OPV 332, OPTEK Technology, TTElectronics, Woking, UK) were characterized by a nominal peak emission wavelength of 850 nm, particularly interesting for our purpose as it falls within the diagnostic and therapeutic window. Each VCSEL had a small divergence angle of 4°, was driven by a direct current (DC) of 11 mA, and emitted an optical power of about 6 mW. The acquisition system was positioned at a distance of 50 cm from the biological tissue under examination and obtained images that can be processed and subsequently archived. It consisted of a CMOS camera (GS3-U3-41C6NIR-C, CMOS sensor 1″, 2048 × 2048 pixels, Point Grey Research Inc., Richmond, BC, Canada), a long-wavelength-pass optical filter with 780 nm cut-on wavelength (MidOpt LP780, Midwest Optical Systems, Inc., Palatine, IL, USA), and a 10-nm-bandpass optical filter centered at 850 nm (FBH850-10, Thorlabs Inc., Newton, NJ, USA). The use of the filter was fundamental for the collection of the NIR photons scattered from the biological tissue and the rejection of ambient light, thus allowing the system to perform the measurements in standard daylight conditions. The camera was USB 3.0 interfaced to a laptop using dedicated software (FlyCapture2, Point Grey Research Inc.). The acquired images were then processed with MATLAB.

3. Results and Discussion

To demonstrate the performance of the system, raw images were acquired first by transilluminating the upper limbs of human volunteers at rest. During the procedure, the subjects were seated in a comfortable position and the biological tissues were not subjected to any kind of thermal stress or mechanical constriction. Before the test, the volunteers provided their agreement to take part in the study and to publish their images. Grayscale pictures were acquired in standard ambient light conditions using exposure times of the order of hundreds of ms and allowed the visualization of vascular structure that appears darker because light is strongly absorbed by the hemoglobin present in the blood flowing in the vessels.

In particular, Figure 2 shows the data related to the transilluminated hand of a male dark-skinned volunteer. Figure 2a reports the raw image: morphological details are visible but with lower contrast with respect to pictures acquired on subjects with white skin [11, 12], because of the high pigmentation of the epidermis. Hence, with the aim of obtaining higher contrast, the image was further processed using MATLAB software. The original picture was cropped (Figure 2b) to eliminate un-significant borders and the histogram relative to the density distribution function of the gray levels was calculated (Figure 2c). Figure 2c shows that the gray levels of the image pixels are concentrated only in a limited range of the histogram because of the low contrast. The cropped picture was processed by contrast adjustment, a procedure that remaps image intensity values to cover the entire range of gray level 0–255 maintaining the shape of the distribution of the original image, as visible from the histogram of Figure 2e. The modified image (Figure 2d) has a higher definition and tiny details of the venous tree are now recognizable.

Since the setup is portable, it was employed also to carry out in field monitoring of the growth of chicken embryos inside the eggshell. Figure 3a–c reports the original picture of the fecundated egg at day 20 of incubation, a cropped area of the image, and its histogram, respectively.
Figure 2. Transilluminated hand of a dark-skinned male subject. (a) Original image with selected region for cropping (red rectangle). (b) Cropped region. (c) Histogram of the cropped area showing distribution of the gray levels. (d) Processed higher quality image after contrast adjustment. (e) Histogram of the processed picture, showing the exploitation of the full range 0–255.

The pixel distribution shows that only a limited range of gray levels is present in the selected area. For this reason, the blood vessels inside the eggshell are barely recognizable and the image looks very dark. Figure 3d, with its histogram (Figure 3e), reports the processed image by contrast enhancement. In this case, a further elaboration step was computed, i.e., histogram equalization (Figure 3g). While the expansion is limited to stretching the histogram without changing its shape, the purpose of the equalization is to change it in such a way to obtain a distribution of constant density. In the final picture (Figure 3f), the blood vessels are finally recognizable without uncertainty. The white small dots are due to the local egg-shell porosity.
Figure 3. Transilluminated chicken embryo inside the eggshell at day 20 of incubation. (a) Original image with selected cropped area (red rectangle). (b) Cropped image. (c) Histogram of the selected area. (d) Processed picture after the step of contrast adjustment and (e) its histogram. (f) Final image after equalization (g) and its histogram.

4. Conclusions

We employed our house-built near-infrared transillumination setup to acquire functional images of in vivo biological tissues and we processed the acquired data to obtain better quality and higher contrast pictures. First, the image processing sequence was tested on a male dark-skinned volunteer. In subjects with high pigmentation of the skin, it is more difficult to clearly distinguish the dorsal vein tree because of partial absorption of the light from the epidermis. By processing the original image, it was possible to obtain a better contrasted picture and to visualize, also, tiny details of the vessel pattern. Moreover, the setup, which is portable, was used also to perform in field monitoring of fecundated chicken eggs. Pictures of the eggs containing embryos were collected and processed by contrast enhancement and further histogram equalization. Elaborated images were of better quality and allowed for more precise recognition of the blood vessels. Future works could focus on a more sophisticated image processing to use this transillumination system for biometric recognition and validation [13].

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References

1. Andersson-Engels, S.; Jarlman, O.; Berg, R.; Svanberg, S. Time-resolved transillumination for medical diagnostics. Opt. Lett. 1990, 15, 1179.
2. Berg, R.; Jarlman, O.; Svanberg, S. Medical transillumination imaging using short-pulse diode lasers. Appl. Opt. 1993, 32, 574.
3. Srinivasan, R.; Singh, M. Laser backscattering and transillumination imaging of human tissues and their equivalent phantoms. IEEE Trans. Biomed. Eng. 2003, 50, 724–730.
4. Durduran, T.; Choe, R.; Baker, W.B.; Yodh, A.G. Diffuse optics for tissue monitoring and tomography. Rep. Prog. Phys. 2010, 73, 076701.
5. Jacques, S.L. Spectral imaging and analysis to yield tissue optical properties. J. Innov. Opt. Health Sci. 2009, 2, 123–129.
6. Jacques, S.L. Optical properties of biological tissues: A review. Phys. Med. Biol. 2013, 58, R37–R61.
7. Gonzalez, J.; Roman, M.; Hall, M.; Godavarty, A. Gen-2 Hand-Held Optical Imager towards Cancer Imaging: Reflectance and Transillumination Phantom Studies. Sensors 2012, 12, 1885–1897.
8. Jung, Y.J.; Roman, M.; Carrasquilla, J.; Erickson, S.J.; Godavarty, A. Non-contact deep tissue imaging using a hand-held near infrared optical scanner. J. Med. Diagnostic Methods 2015, 4, 169.
9. Breastlight™. Available online: https://www.breastlight.com/ (accessed on 8 October 2019).
10. Diaphanoscopy—Stihrle Electronic. Available online: https://www.stihler electronic.de/products/diaphanoscopy.html (accessed on 8 October 2019).
11. Merlo, S.; Bello, V.; Bodo, E.; Pizzurro, S. A VCSEL-Based NIR Transillumination System for Morpho-Functional Imaging. Sensors 2019, 19, 851.
12. Merlo, S.; Bello, V.; Bodo, E.; Catalano, R.; Pizzurro, S.; Rossi Borghesano, M. NIR transillumination system for in vivo functional imaging. In Proceedings of the SPIE Optics + Optoelectronics 2019, Prague, Czech Republic, 1–4 April 2019; Lieberman, R.A., Baldini, F., Homola, J., Eds.; Volume 11028, p. 38.
13. Crisan, S. A Novel Perspective on Hand Vein Patterns for Biometric Recognition: Problems, Challenges, and Implementations; Springer: Cham, Switzerland, 2017; pp. 21–49.