Immanent conditions determine imminent collapses: nutrient regimes define the resilience of macroalgal communities

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Predicting where state-changing thresholds lie can be inherently complex in ecosystems characterized by nonlinear dynamics. Unpacking the mechanisms underlying these transitions can help considerably reduce this unpredictability. We used empirical observations, field and laboratory experiments, and mathematical models to examine how differences in nutrient regimes mediate the capacity of macrophyte communities to sustain sea urchin grazing. In relatively nutrient-rich conditions, macrophyte systems were more resilient to grazing, shifting to barrens beyond 1 800 g m⁻² (urchin biomass), more than twice the threshold of nutrient-poor conditions. The mechanisms driving these differences are linked to how nutrients mediate urchin foraging and algal growth: controlled experiments showed that low-nutrient regimes trigger compensatory feeding and reduce plant growth, mechanisms supported by our consumer–resource model. These mechanisms act together to halve macrophyte community resilience. Our study demonstrates that by mediating the underlying drivers, inherent conditions can strongly influence the buffer capacity of nonlinear systems.

1. Introduction

Identifying where critical thresholds occur in systems characterized by nonlinear dynamics is fundamental to objectively quantifying their resilience [1–3]. Ecosystems as diverse as freshwater lakes, grasslands, coral reefs, and macroalgal communities show hysteretic behaviour [4]; after collapse, these systems may not recover their initial state, even when stress conditions abate. These altered states are typically maintained by increases in the abundance of key species that reinforce stabilizing feedback [5]. There is growing recognition of external stressors as triggers of state shifts, and an increasing interest in understanding their underlying mechanisms [6–9]. Several critical state-changing agents have been identified, including overfishing, pollution and abnormal nutrient loading, population outbreaks of grazers, and infrequent disturbances like storms, fires, temperature anomalies, and other stochastic events [2,10]. Among the best described of these shifts occurs when herbivorous sea urchins, released from predation, quickly overtake near-shore macrophyte communities (i.e. kelp forests),
Figure 1. A macroalgal community grows up to a carrying capacity $N$ following the solid blue line in the absence of herbivores. A population of herbivores consumes this macroalgal community at a rate represented by the solid green line (intermediate pressure, May 1977). Under this consumption curve, two stable states exist: a barren state (red point) and a well-structured macroalgal state (blue point). One unstable state exists (orange point) in which situations on the left precipitate barren formation (consumption $\succ$ growth) and situations on the right enhance the macroalgal community stability (consumption $\prec$ growth). The distance between the unstable point and red points represents the barren precipitation state and the distance between the unstable point and the carrying capacity $N$ represents the macroalgal state resilience.

Reducing uncertainty in complex systems requires a better handle on how context-specific underlying conditions modify ecosystem processes. Attempts to anticipate thresholds have focused on examining characteristics of boundary conditions as signals of impending change [1,2,18,19]. These may manifest as subtle changes in the variance and skew of key system variables, self-organized patchiness, or a slowing down in ecosystem dynamics [20–23]. These changes in system behaviour serve as powerful early-warning indicators presaging major state shifts—either catastrophically [19] or more continuously [24]. Typically, signals have been derived from ecosystem models or by hind-casting of systems that have already experienced shifts. Finding meaningful predictive metrics that work in real-world situations is still elusive. These indicators are essentially phenomenological and dependent on long-term monitoring [19]. Their advantage is that they provide transcendent insights on system behaviour applicable across systems. However, they are not geared to illuminating specific mechanisms underlying transitional responses for a particular ecosystem. Identifying these causal mechanisms may require acknowledging that several interdependent biotic and abiotic processes drive the functioning of the system. Understanding these mechanisms would allow for a clearer evaluation of ecosystem resilience, providing managers with unambiguous directions in prioritizing ameliorative measures.

Mediterranean rocky macroalgal communities are useful models to explore how regional conditions may mediate ecosystem transitions: they show nonlinear responses, are relatively simple, and occur in conditions that differ considerably in their inherent nutrient regimes [11,25,26]. Overfished, these systems often shift to urchin-dominated barrens when their populations cross critical thresholds [11,26,27] (herbivory rates surpass plant growth (Figure 1)). We examined if inherent nutrient regimes mediate where thresholds occur in response to urchin biomass [25]. Additionally, we examined potential mechanistic pathways by which nutrients modify these thresholds. We hypothesize that nutrient regimes can determine the relationship between growth rate and consumption by (i) influencing herbivore consumption rates based on food quality and/or (ii) modifying macroalgal growth based on nutrient availability. Given these mechanisms, macroalgal communities in low-nutrient regimes will experience sudden shifts to barrens at lower urchin biomasses than in high-nutrient regimes. We used complementary approaches to test these mechanisms including empirical observations and controlled...
laboratory and field experiments. We developed a simple consumer–resource model incorporating these mechanisms to explore how nutrients mediate ecosystem state changes.

2. Material and methods

(a) Study system and principal objectives

Shallow Mediterranean rocky shores are dominated by macroalgae, constituting a very structured and diverse community [28,29]. Although growing in oligotrophic conditions, nutrient regimes vary considerably between regions: western continental shores typically receive high-nutrient inputs (i.e. riverine), while the eastern Mediterranean and islands are comparatively nutrient poor [30]. Together with the fish, Sarpa salpa, Paracentrotus lividus is among the most important herbivores in Western Mediterranean macroalgal communities, precipitating regime shifts when outbreaks occur [26,31]. Predator overfishing releases urchin populations from predation pressure, and macroalgal systems give way to barrens under sustained herbivory [26,32]. Once created, several reinforcing feedbacks maintain this new state. These include enhanced post-settlement survival of sea urchins, reduced potential for algal recruitment, reductions in the recruitment of predatory fish, and the facilitation of other herbivores [33–35]. We (i) used observational studies to establish the relationship between nutrient levels and critical transitions in macrophyte communities, (ii) identified the mechanisms underlying these transitions to understand how contrasting nutrient regimes influence resilience, and (iii) built a predictive model to explain how these transitions vary with inherent conditions. First, we examined how algal cover changed along a gradient of nutrient conditions. For the second objective, we assessed two potential mechanisms underlying nutrient-mediated thresholds. For the third objective, we used mean percentage cover per urchin biomass. As the change point detection requires discrete cover values applied to detect abrupt changes in many other datasets [1].

Our observations indicated a threshold in the response of algal cover to urchin biomass. We examined this with change point detection methods using the package strucchange [1,38] in R [39] for each region. The algorithm assesses whether different parts of datasets require different fits to a linear regression. To assess the significance of every potential change point, an F-test (Chow test) was computed. This test is typically used in time-series for systems exposed to disturbances, but can be applied to detect abrupt changes in many other datasets [1].

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(ii) Mechanisms underlying the thresholds

We explored mechanisms underlying nutrient-mediated thresholds using a combination of controlled laboratory and field experiments.

(i) Feeding response. In the laboratory, we tested if macroalgal nutrient content affected per capita urchin herbivory rates. We used the Mediterranean seaweed Cystoseira mediterranea as a model forage species because it is clearly preferred by P. lividus [31]. Sea urchins and macroalgae were collected in the same area (41.7 N 2.8 E). Half the macroalgae was fertilized (F) in aquaria (approx. 10 l) with running seawater for 3 days using 6 g of fertiliser (12N : 8P : 16 K) while the other non-fertilized (NF) were kept in natural seawater aquaria. To assess food quality, we measured leaf nitrogen (%N) in fronds from each region, NT = 5 per treatment. Fronds were powdered and analysed for nitrogen concentration. Fertilization successfully increased the nutrient content by approximately 9% (electronic supplementary material, figure S2; p-value < 0.01). All collected urchins were starved for 3 days in a holding aquarium (approx. 1 000 l). They were then transferred to six independent aquaria for testing. Each aquarium was divided into six compartments. Five of these accommodated a single urchin, and one was left without urchins (n = 30). We fed each the urchins with 4 g (weet weight) of the non-fertilized algae while the rest were fed with 4 g of the fertilized algae (n = 15 urchins per treatment). Compartments left without urchins, served as procedural controls to account for non-feeding-related algal losses. After 6 days we weighed the remaining algae and consumption was estimated by subtracting the final from the initial weight and dividing by total feeding.
time (6 days). No significant change in fertilized or unfertilized algal wet weight was detected in procedural controls. We used a linear mixed effects model to test for differences in consumption rates between fertilized and unfertilized treatments. The full model included the dependent variable ‘consumption rate’, the fixed factor ‘treatment’ (two levels: fertilized and unfertilized), and the random effect ‘aquarium’ (six levels—the six aquaria) to account for the variance among animals kept within the same aquarium. Each effect was dropped sequentially, and we selected the best model using the Akaike Information Criterion [40].

(ii) Nutrient-mediated algal growth. To measure how nutrients influenced algal growth in field conditions, we used two bare rock areas (completely overgrazed areas, 0% algal cover), one in each nutrient region. There, three herbivore exclusion cages of 50 × 50 cm (2 cm mesh size) were established. We measured changes in total algal cover (of both erect and turf algae) inside each plot after one month in both high- and low-nutrient conditions. We used change in algal cover as an indirect, comparative index of growth [41,42]. We measured algal cover using the same methods employed in our field surveys (see above). We used one-way ANOVAs to test for differences in growth using R [39]. Prior to the analysis, assumptions of normality and homogeneity of variances were checked both visually and statistically (i.e. Shapiro test and Bartlett test, respectively).

(iii) Feeding response and nutrient-mediated growth (field experiment). To further validate these mechanisms, we conducted a manipulative experiment to independently estimate the effects of nutrient-mediated growth and feeding response in real-world conditions. The experiment was conducted in an oligotrophic location (41.0° N 9.0° E) at approximately 8 m depth in a landscape dominated by isolated boulders during the maximum vegetative growth period for most algal species in the study area (May to September). Twelve isolated boulders (approx. 2 m² in area) were randomly selected and used as independent plots (more than 5 m apart) in a total area of about 1 000 m². All urchins were removed and algae scraped off with a metal brush to ensure similar initial conditions. Treatments were randomly assigned to plots: three fertilized (nutrient-enriched, without urchins: N⁺U₀), three fertilized and had urchins added at a density of 10 individuals m⁻² (nutrient-enriched, with urchins: N⁺U₁), three unfertilized and had sea urchins added at a density of 10 individuals m⁻² (not enriched, with urchins N₀U₁) and finally, three left unfertilized and without urchins (not enriched, without urchins: N₀U₀). Enriched conditions were achieved by fixing 20 × 5 × 5 cm nylon mesh bags (1 mm mesh size), containing 150 g fertilizing pellets (15N : 9P : 9 K) to a tile. The effectiveness of fertilization was assessed from water samples collected at randomly selected plots. Samples were taken approximately 1 cm from the nutrient source, filtered (0.45 μm filter size) and analysed using a continuous-flow AA3 Auto-Analyzer to determine the seawater nitrate concentration (mg l⁻¹). Fertilization was always successful: N concentrations in fertilized plots were about 98% higher compared with unfertilized ones (p < 0.01). The number of urchins in Uᵢ treatments was period-ically controlled [33]. Algal cover on the boulders was estimated after six weeks using four random photographic quadrats (400 cm² in size) of the surface of each plot. Nutrient-mediated growth in the absence of herbivory was calculated as the percentage difference in cover between fertilized and unfertilized treatments:

\[
\text{nutrient-mediated growth} = 100 \times \frac{N_i U_0 - N_0 U_0}{N_i U_0}.
\]  

(Urchin addition treatments integrated the effects of both nutrient-mediated growth and feeding response:)

\[
\text{nutrient-mediated growth + feeding response} = 100 \times \frac{N_i U_1 - N_0 U_1}{N_i U_1}.
\] 

This allowed us to estimate the effect of compensatory feeding alone, by subtracting the effects of nutrient-mediated growth (equation (2.1)) from the total unconsumed algae (equation (2.2))

\[
\text{feeding response} = \text{equation (2.2)} - \text{equation (2.1)}.
\]

(iii) Modelling nutrient-mediated thresholds

We developed a simple consumer–resource model incorporating the mechanisms of nutrient-mediated growth and compensatory feeding to predict differences in tipping points. In the model, algal cover was introduced as a logistic growth curve while the effects of urchin herbivory were modelled as a Holling Type II equation. As a consequence, the interplay between algal demographics and urchin herbivory determined a dynamic equilibrium for algal cover through time. In this model, nutrients influenced both algal growth rates and urchin herbivory—the second by mediating handling time within the Holling Type II Equation (see electronic supplementary material, appendix A).

3. Results

(a) Determining thresholds under different nutrient regimes

There was clear evidence that the macroalgal systems we sampled showed alternate states in both high- and low-nutrient regions (figure 2). Sudden changes in the community state (per cent cover) occurred when the stressor (urchin biomass) crossed critical values ( tipping point). In high-nutrient conditions, results show that this tipping point was breached at urchin biomasses more than twice (figure 2a) that of low-nutrient conditions (figure 2b). Threshold analyses confirmed the existence of regime shifts in both nutrient regimes; thresholds were found at significantly different levels of the stressor (urchin biomass; figure 2c,d) in different nutrient conditions. Macroalgal communities in low-nutrient conditions shifted abruptly to urchin barren when stressor values crossed 736 g m⁻² sea urchin biomass (approx. 20 sea urchins m⁻² of 5 cm TD, 380–1 250 g m⁻², 95% confidence intervals; figure 2d,f). By contrast, in high-nutrient conditions canopy-forming algae were still present at biomasses around 1 832 g m⁻² (approx. 40 sea urchins m⁻², 1 484–2 494 g m⁻², 95% confidence intervals; figure 2c,e); beyond this level however, these systems also collapse to urchin-dominated barrens.

(b) Mechanisms underlying thresholds

Urchins showed significant foraging plasticity, adapting to lower nutrient conditions by increasing feeding rates (compensatory feeding). In the laboratory, urchin grazing rates were 25% higher when offered non-fertilized C. mediterranea compared with fertilized algae (figure 3b; p < 0.01; electronic supplementary material, figure S4). In field experiments, urchins in unfertilized plots ate considerably more than in fertilized plots. After accounting for nutrient-mediated growth, compensatory feeding amounted to 35.5% of total consumption in the high-nutrient regime, caged sites recovered 100% of their original algal biomass—growing significantly more in nutrient-rich regimes as well as in fertilized plots. After a month of herbivore exclusion, barren sites in both regions were recolonized by a very similar assemblage of mixed turfs and erect algae. However, while in the high-nutrient regime, caged sites recovered 100% of their original algal biomass.
Figure 2. Bubble plots of the percentage of algal cover for different sea urchin biomass for the two regions sampled: (a) Catalan coast (n = 237) and (b) Sardinia Island (n = 185). Bubble size is proportional to the number of times a specific combination of urchin biomass and macroalgal cover was recorded. Dashed red lines indicate the position of thresholds. Photos are representative of typical barren areas in each region. Change point analysis results for high- (c) and low- (d) nutrient regions. Black dots show the mean algal cover at different levels of the stressor (urchin biomass in grams per square metre of wet weight). Red horizontal lines represent the mean algal cover before and after the threshold. Grey areas represent the confidence intervals (95%) around change points detected. The F statistic (Chow test) obtained for the forward threshold is presented for both high- (e) and low-nutrient regions (f). F statistic peaks around the area where the change point has been detected.

Figure 3. Conceptual diagram describing the two mechanisms evaluated; the compensatory feeding and nutrient-mediated growth of the macroalgal community and sea urchin population described in figure 1a. Under low-nutrient regimes sea urchins compensate feeding (i.e. increased consumption rates) (b) mean consumption rate on fertilized C. mediterranea $F = 0.53 + 0.04$ g day$^{-1}$ (n = 15) and on non-fertilized C. mediterranea $NF = 0.71 + 0.03$ g day$^{-1}$ (n = 15) in laboratory experiments after 6 days. At the same time under low-nutrient regimes, the macroalgal community presents retarded growth rates, (c) mean percentage cover of erect and turf algae in the high-nutrient region $HN = 100\% + 0\%$ and low nutrient region $LN = 29.3\% + 7.42\%$ after one month of herbivore exclusion.
algal cover, in the low-nutrient regime, only approximately 30% of the substrate was colonized (figure 3c; \( p < 0.01 \); electronic supplementary material, figure S4). Fertilization experiments confirmed these results; fertilized plots (N0U0) showed a 35% increase in algal cover compared to controls (NoU0) (electronic supplementary material, S3).

(c) Modelling nutrient-mediated thresholds

The consumer-resource model identified clear bistability in this system (figure 4), with macroalgae rapidly shifting to barrens beyond a threshold. This threshold varied with nutrient conditions, governed largely by differences in growth rates and handling time. Additionally, as forage encounter rates are typically high compared with algal growth rates, it requires urchin populations to reduce very close to zero before the barren is likely to shift back to algal forests (figure 5, electronic supplementary material; appendix A). This was true for both nutrient regimes. This matches closely with the empirical data and, when parametrized with ecologically meaningful values, predicts thresholds very close to those observed in the field.

4. Discussion

While nonlinear dynamics of temperate macrophyte communities have long been recognized, identifying where nonlinearities lie has resisted prediction. Our results indicate that nutrient regimes can strongly determine the buffer capacity of macroalgal communities, mediating the amount of herbivory the system can withstand. Results from field, laboratory, and modelling approaches, all converged to give us a clear understanding of how nutrients mediate the grazing resilience of these near-shore benthic communities. Specifically, oligotrophic regions are much less resilient to grazing compared with nutrient-rich regions (figure 5). We documented a clear shift to barrens at around half the sea urchin biomass in oligotrophic systems (relative to high-nutrient areas), indicating that they are intrinsically less able to cope with grazing. Barren states in both nutrient regimes were remarkably stable. Our model highlights the existence of hysteresis indicating that it would require an almost-complete disappearance of sea urchins before these systems recover their macroalgal state. This is consistent with other studies [43,44]. We identified the mechanisms underlying differences in these shifts, showing that both compensatory grazing and reduced algal growth act together in oligotrophic systems, endowing them with roughly half the buffer capacity of more nutrient-rich systems (figures 3 and 5).

Understanding and quantifying how context-specific conditions influence threshold dynamics will take us one step closer towards reducing the inevitable surprise of nonlinear ecological systems.

Temperate macrophyte communities appear particularly prone to catastrophic shifts [11,26], with sea urchin overgrazing (linked to population outbreaks) being the primary trigger of these events. While it is uncertain from our work what drives differences in urchin populations, studies have highlighted the importance of both supply-side processes governing recruitment and settlement [45] as well as subsequent top-down control by fish predators [26,46–48] in driving urchin population dynamics. Temperate macroalgal systems show catastrophic state-shifting behaviour independently of local and regional conditions. We documented sudden shifts to urchin barrens in both nutrient regimes, linked clearly to increases in urchin abundance. The difference was in where thresholds lay, with relatively nutrient-rich systems more than twice as resilient to grazing compared with nutrient-poor systems. Given their susceptibility to discontinuities, determining these boundary conditions is all the more important to manage temperate macrophyte communities in functionally healthy states. This is critical given that our model shows strong hysteretic behaviour, an indicator of the resilience of the altered community states [11,25]. While low-nutrient conditions make macrophyte systems much more prone to barrens, recovery from this state appears to be independent of nutrient regimes (figure 4, electronic supplementary material; appendix A). The model suggests that as long as algal encounter rates are much higher than algal regrowth, recovery from barrens will always be very protracted. While the backward process needs to be interpreted with caution, it indicates that erect algae are likely to recover only when urchin abundances reach close to zero, regardless of nutrient conditions. Once barrens are created, urchin populations generally do not collapse although individual growth rates may decrease as they switch to feeding less nutritious encrusting algae [25,44]. In macroalgal-barren systems, urchins continue scraping the substrate, maintaining areas free of algae. In addition, other barren-associated biota may also play significant roles in enhancing the stability of barrens [49]. Such reinforcing feedbacks make recovery difficult, emphasizing the need to prevent collapses from occurring. Being able to accurately predict transitions is essential for preventive action to be effective.

Our findings contribute to understanding the role environmental factors play in determining the resilience of macroalgal-dominated reefs. Wernberg et al. recently described the collapse of kelp forests following a severe heatwave [9]. Additionally, Verge's empirically demonstrated how warming-mediated increases in fish herbivory trigger system collapses [50]. These recent examples highlight the urgent need to uncover the mechanisms underlying the worldwide decline of temperate macroalgal reefs [51]. Our work shows that inherent conditions can be critical drivers of buffer capacity by influencing both plant growth and herbivore feeding responses. Hence, very
oligotrophic systems may be particularly vulnerable to herbivore outbreaks. Cardona et al. [52] described how, after a pulse in productivity in a low-nutrient region, the abundance of urchins increased, resulting in dramatic reductions in macroalgal communities. In the eastern Mediterranean, with significantly lower nutrient conditions than the western basin [30], the spread of the herbivorous fish *Siganus* spp. through the Suez Canal has led to extreme depletions of canopy-forming algae and, where well-developed macrophyte communities were once dominant, now bare rock prevails [29, 53]. Acknowledging this differential vulnerability may require designing context-specific strategies for managing these systems based on measurable differences in their inherent buffer capacity.

Our work also explores the potential mechanisms that can explain the differential resilience of these ecosystems. Under relatively nutrient-poor conditions, macroalgae showed clearly reduced rates of growth and urchins offset the low quality of plants by increasing their feeding rates to meet nutritional requirements [54]. Both compensatory feeding and nutrient-mediated growth work together (figure 4) making macroalgal systems in low-nutrient regimes shift to barrens at much lower herbivore levels compared with communities in high-nutrient conditions. The underlying nutrient regime determines the degree to which macroalgal growth can support urchin consumption before collapsing. As we demonstrate, low-nutrient regimes increased rates of consumption by herbivores (compensatory feeding, figure 3b; electronic supplementary material, figure S5) while simultaneously reducing the growth capacity of macroalgae (figure 3c; electronic supplementary material, figure S5). This maximized grazing: primary production ratio caused a faster shift to a new macroalgal-free urchin barren (figure 3). These overshoots are much more likely to occur in the characteristic nutrient-poor conditions of islands making them much less resilient to herbivory compared with other nutrient-rich systems. Under extremes of high-nutrient conditions the situation may change. In these scenarios (relatively rare in the Mediterranean), macroalgal community composition may change to fast-growing species, making the system vulnerable to collapse through a completely different set of mechanisms including shading and overgrowth [55]. Based on our current results however, we suspect that oligotrophic systems may follow inherently different trajectories than non-oligotrophic systems, and need to be addressed separately.

Several studies have focused on determining signals of impending collapse in nonlinear systems [2, 19, 23]. They identify potentially useful proxies (critical slowing down, increasing variance and skewness, etc.) that may herald approaching thresholds. Identifying these signals depends on reliable long-term monitoring, and adequate demonstration that signals correlate with hysteretic change. There have been few real-world examples where these leading indicators have been able to predict imminent collapse in time for ameliorative action. Essentially phenomenological, these approaches assume that the underlying mechanisms will vary from context to context. Our work indicates that, where it is possible to unpack these underlying drivers, it can help substantially in identifying where and why tipping points occur. Additionally, clarifying the mechanisms that govern these dynamics allows determining the role inherent conditions play in mediating critical thresholds. Taken together, it provides a way forward to make regime-specific predictions of buffer capacity of systems at local to regional scales. These are admittedly much more difficult to establish in more complex systems where multiple mechanisms act in several synergistic and antagonistic ways. Our contention is that, even in these more complex ecosystems, inherent conditions may predispose the system to very different dynamics, implying very different ecosystem responses. It is critical to shift attention to a more mechanistic understanding of the ecological processes that govern nonlinear systems. Determining how feedbacks interact with context-specific conditions will help considerably improve the predictive power of resilience models, reducing the surprise

![Figure 5. Buffer capacity of macroalgal community and barrens in each nutrient regime. The bubble graph shows macroalgal percentage cover related to sea urchin biomass in gram per metre square (wet weight) in the two regions. The dashed red line represents the threshold after a tipping point F2 is reached. F1 represents the tipping point from which the recovery of the macroalgal state is possible. Both F1 and F2 are set in the position determined by the consumer-resource mathematical model (see electronic supplementary material, appendix A). The schematic drawing below shows the resilience of each stable state in both nutrient regimes. The valleys represent the alternative stable states and the depth of the valley represents the resilience of that particular state.](http://rspb.royalsocietypublishing.org/)
inherent in identifying thresholds and improving our ability to manage systems characterized by nonlinear behaviours.

Data accessibility. Data from surveys are stored in the open access Temperate Reef Base (http://temperatereefbase.imas.utas.edu.au/static/landing.html) and Data deposited in Dryad: http://dx.doi.org/10.5061/dryad.7k3k00 [56].

Authors’ contributions. All authors conceived the idea and decided the experimental design. J.B., A.P., S.O., G.C., and T.A. collected data on macroalgal cover and sea urchin biomass. L.P., J.B., A.P., S.O., and G.C. designed and conducted field experiments. D.A. developed the mathematical model. J.B., J.F.P., and T.A. performed laboratory and statistical analyses. J.B., R.A., T.A., and J.R. designed figures and schemes. J.B., R.A., and T.A. wrote the manuscript with contributions from all authors.

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