Estimating the survival of unobservable life stages for a declining frog with a complex life history

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Abstract. Demographic models enhance understanding of drivers of population growth and inform conservation efforts to prevent population declines and extinction. For species with complex life histories, however, parameterizing demographic models is challenging because some life stages can be difﬁcult to study directly. Integrated population models (IPMs) empower researchers to estimate vital rates for organisms that have cryptic or widely dispersing early life stages by integrating multiple demographic data sources. For a stream-inhabiting frog (Rana boylii) that is declining through much of its range in Oregon and California, USA, we collected egg-mass counts and capture–mark–recapture data on adults from two populations in California to ﬁt IPMs that estimate adult abundance and the survival rate of both marked and unobserved life stages. Estimates of adult abundance based on long-term monitoring of egg-mass counts showed that study populations ﬂuctuated greatly inter-annually but were stable at longer timescales (i.e., decades). Adult female survival during 5–6 yr of capture–mark–recapture study periods was nearly equal in each population. Survival rate of R. boylii eggs to the subadult stage is low on average (0.002) but highly variable among years depending on post-oviposition stream ﬂow. Population viability analysis showed that survival of adult and subadult life stages has the greatest proportional effect on population growth; the survival of egg and tadpole life stages, however, is more malleable by management interventions. For example, simulations showed head-starting of tadpoles, salvaging stranded egg masses, and limiting aseasonal pulsed ﬂows could dramatically reduce the threat of extirpation. This study demonstrates the value of integrating multiple demographic data sources to construct models of population dynamics in species with complex life histories.

Key words: altered ﬂow regime; Bayesian modeling; environmental stochasticity; integrated population model; population viability analysis; Rana boylii.
INTRODUCTION

Species extinction is occurring at an unprecedented rate (Barnosky et al. 2011, IPBES 2019). Actions to protect whole ecosystems can be effective to slow biodiversity loss, especially when a master variable, such as the flow regime of a river, has profound influences on survival of many taxa (Poff et al. 1997, 2010, Palmer and Ruhí 2019). Still, extinction can be idiosyncratic (Colchero et al. 2019, Howard et al. 2019). Demographic knowledge at a fine-grained level of detail is essential (Akçakaya and Sjögren-Gulve 2000), because species may face disparate stressors across their ranges, and life stages within a species can display incongruent risks from similar stressors (Otero et al. 2011, Sturrock et al. 2020). Structured population models, which condense demography into a few key age- or stage-specific vital rates, have proven useful for identifying the environmental factors, biological traits, and anthropogenic stressors driving growth or decline (Brook et al. 2000, Caswell 2001). Here, we develop an integrated population model (IPM) to quantify the efficacy of measures, including the management of water flow in rivers with dams, designed to minimize extinction risk of an imperiled frog. We also address the difficulties of vital rate estimation for hard-to-study life stages.

Because of complex life histories and cryptic life stages, vital rates can be challenging to quantify for a wide range of taxa that are emblematic of the current biodiversity crisis, from lepidoptera to amphibians (Dirzo et al. 2014, Scheele et al. 2019). The life history of a free-living larval stage followed by a dispersing post-metamorphic stage can complicate parameterization of models (Williams et al. 2002, Oberhauser et al. 2017, Earl 2019). In particular, survival of recently metamorphosed individuals can be difficult to estimate in the wild when recapture rates for marked individuals are low (Kupferberg et al. 2009). This has pushed researchers to use in situ enclosures that might not fully reflect natural conditions (Altwegg and Reyer 2003). When vital rates are highly uncertain and models are parameterized using independent data sources, demographic models, such as matrix models, can produce unrealistic estimates of population growth rates (Schaub and Abadi 2011). Despite the challenges posed by high uncertainty in estimates of vital rates for early life stages, many authors have used stage-based population models to study the demography of amphibians (Earl 2019). Integrated population models can partially resolve the aforementioned challenges by explicitly integrating different data types into a joint likelihood and can produce demographic estimates with greater precision and accuracy than independent models (Abadi et al. 2010). Integrated population models can enable the estimation of survival and potentially can uncover environmental covariates of survival for unobserved life stages (Abadi et al. 2017).

Integrated population models hold promise for improving conservation of species with breeding migrations and complex life cycles, such as the Foothill Yellow-Legged Frog (*Rana boylii*). This stream and river dweller, endemic to Oregon and California (USA), has a life cycle synchronized with the region’s winter flood/summer drought flow regime and punctuated by spring migration of adults from resident habitats to lek sites where they mate and lay eggs. Some populations inhabit tributary streams too cool and shaded to support timely larval development; wider sun-lit channels where tadpoles graze on abundant periphyton are necessary for metamorphosis prior to the next flood season (Catenazzi and Kupferberg 2013). Thus, the quantity of clutches of eggs laid in mainstem channels reflects adult female abundance in adjacent tributaries. Several factors have driven the extirpation of *R. boylii* from almost two thirds of its historical range (Lind 2005). Flow regulation by dams creates stressors for early life stages (Kupferberg et al. 2012, Catenazzi and Kupferberg 2013), while conditions for post-metamorphic life stages in un-dammed tributaries remain unimpaired. Other stressors include invasive species that thrive under artificial low flow or drought conditions (Kupferberg 1997), chytridiomycosis (Adams et al. 2017b), habitat destruction (Davidson et al. 2002), and synergies among all these factors (Adams et al. 2017a). Artificial fluctuation in stream flow released from dams is a particularly potent cause of early life stage mortality because it can scour or strand a substantial portion of an annual cohort of egg masses and larvae (Lind et al. 1996, Kupferberg et al. 2012). Cold water from the hypolimnion of upstream...
reservoirs released in the summer shifts periphyton communities toward taxa inedible by tadpoles (Furey et al. 2014), reinforcing the effects of low temperatures on reduced tadpole growth and higher mortality (Catenazzi and Kupferberg 2013, Wheeler et al. 2015). Despite such mechanistic knowledge of the causes of this species’ decline, critical information gaps about survival of _R. boylii_ remain, in part due to dispersal of young of the year away from the mainstem lek sites into complex fluvial networks. Although monitoring studies have tracked trends in adult _R. boylii_ abundance using egg-mass counts (Kupferberg et al. 2012), and manipulative experiments have provided insight into mortality of pre-metamorphic life stages (Kupferberg et al. 2012), capture-mark-recapture (CMR) data suitable for estimating post-metamorphic survival are limited, and survival of pre-reproductive life stages remains largely unknown. Quantifying these vital rates in systems without anthropogenic impairment is a necessary step to project the viability of populations when stressors are added and to assess the need to protect _R. boylii_ under the federal Endangered Species Act (United States Fish and Wildlife Service 2015).

In this study, we use an IPM to model the demography of robust populations of _R. boylii_ inhabiting free-flowing streams at two northern California sites where the aforementioned anthropogenic stressors are largely absent (Hurdygurdy Creek in the Smith River watershed and Fox Creek and its confluence with the South Fork Eel River). Starting with that reference model, we then conduct scenario analysis to project the viability of populations in rivers with dams or in un-dammed rivers where increasing flow variability associated with climate change and disease presents future threats. Our IPM integrates long-term monitoring of egg-mass counts (7 yr at one site and 27 yr at the other), CMR data from adult frogs, fecundity, and survival of eggs to hatching to estimate recruitment to the subadult stage and survival of adult and subadult frogs, and to quantify trends in adult female abundance for these two populations. Importantly, the integration of multiple data sources, and the linkage between population trends and demographic rates, produces precise and realistic estimates of survival, including survival from egg to subadult, which is not directly observed. Using these demographic vital rates to parameterize a stochastic population model allowed for an examination of how different conservation and water management scenarios could affect population viability. By simulating the effects of increasing survival to adulthood (as would occur in a zoo rearing and reintroduction effort to bolster small populations) or decreasing recruitment from the egg stage (as when ill-timed releases from dams occur), we quantify anthropogenic influences on the relative probability of persistence vs. extirpation for _R. boylii_ populations.

**METHODS**

**Data sources**

We compiled data on the survival of each life stage of _R. boylii_ and the fecundity of adult females from published literature and unpublished data (Appendix S1: Table S1). The number of eggs in individual egg masses (fecundity) and embryo survival rates (proportion of eggs that hatched) were counted from the South Fork Eel River (hereinafter “SF Eel River”) in 1992 (n = 3) and 2007 (n = 20; Kupferberg et al. 2009).

We collected CMR data from adult and subadult _R. boylii_ at Fox Creek, a tributary of the SF Eel River, from 2014 to 2019 (Fig. 1). We sampled for one or two primary periods each summer, between June and early September. Each primary period consisted of 2–6 secondary periods of sampling. We sampled frogs using visual encounter surveys both during the day (between 09:00 and 18:00 hours) and at night (between 20:00 and 01:00 hours). We marked frogs with passive integrated transponder (PIT) tags and determined sex based on the presence or absence of nuptial pads.

We marked and counted egg masses along an approximately 5.2-km reach of the SF Eel River each spring/summer from 1993 to 2019 using the census protocol described by Kupferberg et al. (2012). We performed two to six egg-mass surveys per year (average = 3.8 surveys) between the beginning of April and mid-June; egg masses were marked to prevent double counting. For the Fox Creek IPM, we included egg masses counted at the lek sites immediately upstream and downstream of the confluence of Fox Creek (where CMR surveys took place) and the SF Eel River (reach length = 507 m). We documented egg-
mass mortality from stranding and scouring on
the SF Eel River from 1992 to 2013 and 2017 to
estimate the relationship between egg-mass sur-

vival and post-oviposition stream flow (follow-
ing Kupferberg et al. 2012).

At Hurdygurdy Creek, we collected CMR data
from adult R. boylii from 2002 to 2008, primarily
from April to June, with additional surveys in
August and September 2002 and 2004, and
August to November in 2003. Survey effort was
higher from 2002 to 2004 (26–41 survey dates
over 5–8 months/yr) than from 2005 to 2008 (3–12
survey dates over 2–3 months). We counted egg
masses along a 1.56-km reach of Hurdygurdy
Creek during 12–25 surveys (average = 17.7)
from April to June each year. Egg-mass mortality
was not tracked at Hurdygurdy Creek.

**Integrated population model**

The IPM links the survival rate of each life
stage and the fecundity of adult females to create
a demographic model for R. boylii (Fig. 2). The
survival rate of adult frogs is informed by both
the CMR data and the annual egg-mass counts
(Fig. 2). Egg-mass counts are also used to esti-
mate the number of adult females present in the
population each year. The inclusion of egg-mass
counts as an estimate of adult female abundance
constrains all vital rate parameters to values that
match the temporal trend in the abundance of
adult frogs in the breeding population. Likewise,
the estimate of adult survival from CMR data
constrains estimates of trends in adult female
abundance.

The IPM is a female-based population model
with three stages: egg (age 0), subadult (age 2),
and adult (ages 3+). The number of eggs pro-
duced in year \( t \) is the product of the number of
adult females, \( N_{ad}[t] \), and the average number
of eggs per egg mass, \( f \), divided by 2 (assuming
a 1:1 sex ratio of eggs; Eq. 1). The survival rate
from the egg stage to subadult stage (\( \phi_{em} \)) is the
product of the survival rate of egg masses in the
face of stranding and scouring (\( \phi_{em} \)), the
survival rate of embryos hatching to become tadpoles ($\phi_h$), and the survival rate from the tadpole stage until the 2-yr-old subadult stage ($\phi_{t.s}$; Eq. 2). Individuals take two years to grow from the egg stage to the subadult stage (Eq. 3) and reach the adult stage (sexual maturity) at 3 yr of age. Empirical data are available for each survival parameter except $\phi_{t.s}$, which is estimated by leveraging the relationships between the other parameters in the IPM (Fig. 2). The number of female recruits that survive to two years old is the product of the recruit survival rate ($\phi_{rec}$) and the expected number of eggs produced two years earlier, $N_{egg}[t-2]$. The survival rate of subadult frogs from 2 yr old to 3 yr old is assumed to be equal to the annual survival rate of adult frogs ($\phi_a$). The number of adult frogs at time $t$, $N_{ad}[t]$, is the sum of adults present at time $t-1$ that survive to time $t$ and the number of subadults present at time $t-1$ that survive to time $t$ (Eq. 4).

$$N_{egg}[t] = N_{ad}[t] \times \frac{f}{2}$$  \hspace{1cm} (1)

$$\phi_{rec} = \phi_{em} \times \phi_h \times \phi_{t.s}$$  \hspace{1cm} (2)

$$N_{sub}[t] = N_{egg}[t-2] \times \phi_{rec}$$  \hspace{1cm} (3)

$$N_{ad}[t] = N_{ad}[t-1] \times \phi_a[t-1] + N_{sub}[t-1] \times \phi_a[t-1].$$  \hspace{1cm} (4)

We analyzed the CMR data for adult and subadult $R. boylii$ using a Bayesian implementation of a Cormack-Jolly-Seber (CJS) model (Cormack 1964, Jolly 1965, Seber 1965, Kéry and Schaub 2012). The CJS model conditions on first capture and estimates the recapture rate ($p$) of marked individuals and their survival rate ($\phi_a$) between primary sampling periods. We used a binomial
indicator of captured (1) or not (0) for each year of sampling; if we sampled frogs during multiple primary periods within a year, we collapsed data into a single binomial response. Because we cannot separate true survival from emigration, \( \phi_a \) is an estimate of apparent survival of adult and subadult frogs. We tested for an effect of frog sex on \( p \) and \( \phi_a \) in each CMR dataset by fitting sex-dependent models and calculating the difference between females and males for each parameter.

We used a state-space model to analyze trends in egg-mass counts (Kéry and Schaub 2012). Female \( R. \) boylii, like many ranid frogs, lay one clutch per breeding season, so the number of egg masses is effectively a count of the number of adult females present and breeding in a given year (Zweifel 1955). The egg-mass count results from an observation process that samples the true underlying number of breeding adult females. The observation process is modeled using a Poisson distribution, and both over- and under-counting are possible, although under-counting is unlikely given our method of marking egg masses. The IPM does not explicitly include immigration or emigration, and we assume that all adult females breed each year. These assumptions can be violated for amphibian populations (Earl 2019) but were necessary because we had no data to estimate these parameters. We calculated the long-term log stochastic growth rate (log \( \lambda \)) for each population using the estimated abundance of adult females each year (Caswell 2001). To assess how annual population growth rates were related to time-varying vital rates, we calculated the correlation between \( \lambda(t) \) and adult survival that year (\( \phi_{a,t} \)) and egg-mass survival three years prior (\( \phi_{em,t-3} \)), following the methodology of Kéry and Schaub (2012).

We summarized the fit of each sub-model with a Bayesian P value, where \( P \) equals the proportion of iterations in which the fit is better for replicate data generated by the model than the observed data. In general, Bayesian P values near 0.5 indicate good fit of the model to data, and values far from 0.5 indicate the model fits the replicate data better (near 1) or much worse (near 0) than the observed data (see Appendix S1; Kéry and Schaub 2012).

We used uninformative Beta(1,1) priors for mean \( \phi_a \) and mean \( p \). We allowed adult survival and recapture probability to vary among years using a logit-normal random effect (Table 1). Survival from tadpole to subadult stage (\( \phi_{h,s} \)) was constant because we lacked data to estimate annual variation in this parameter. We used an uninformative Beta(1,1) prior for \( \phi_{h,s} \). The prior for hatching survival (\( \phi_{h} \), the proportion of eggs that hatch into tadpoles) in the Hurdygurdy Creek IPM was a Beta(3.8, 0.75) distribution based on empirical data from the SF Eel River study reach (Table 1).

We modeled egg-mass survival at the SF Eel River as a function of the natural log of the ratio of maximum to minimum post-oviposition stream flow (hereinafter “flow magnitude index”; Kuperberg et al. 2012) using a binomial regression with the observed number of egg masses that survived stranding and scouring as the response. For Hurdygurdy Creek, egg-mass survival was modeled using an informative Beta(6,1) prior based on observed egg-mass survival rates at the SF Eel River study reach (Table 1).

We ran IPMs in JAGS version 4.3.0 (Plummer 2003) accessed through R version 3.6.3 (R Core Team 2020) using the runjags package (Denwood 2016). We ran IPMs on five independent chains of 2,000,000 iterations each after a burn-in of 100,000 iterations, with chains thinned by a factor of 100 to produce a final posterior of 100,000 samples. We inspected trace plots and calculated the Gelman-Rubin diagnostic, \( R \) (Brooks and Gelman 1998), to assess mixing of chains and convergence. All parameters had \( R < 1.01 \) and exhibited no signs of failure to converge. For parameter estimates, we report the median of the posterior distribution followed by the 95% highest posterior density interval (HPDI) in parentheses. R and JAGS codes to replicate the IPM are available on ScienceBase (Rose et al. 2020).

Population viability analysis

We used vital rate estimates from the IPM to parameterize a stochastic population model for a population viability analysis (PVA). The goal of the PVA was to compare the sensitivity of population viability to changes in the survival rate of early life stages (eggs and tadpoles) and survival of adult and subadult frogs, not to estimate actual probabilities of focal populations going extinct. We projected the future trend of populations under different scenarios over a time period of 20 yr, similar to the study period for the SF Eel...
River population. Shorter time horizons are preferable for PVA projections because extinction probabilities over longer time periods (e.g., 50 or 100 yr) are sensitive to initial population growth rates and environmental conditions over longer time horizons are highly uncertain (Fieberg and Ellner 2000). We incorporated both temporal variance in vital rates (i.e., process variance, Morris and Doak 2002) and parametric uncertainty (i.e., sampling variance or variance in estimates of survival rates from the IPM) to fully account for uncertainty in projections of population viability under different management scenarios (McGowan et al. 2011). We ran all PVAs for 10,000 simulations. For each simulation, we drew survival and fecundity parameters from the posterior distribution of the Fox Creek IPM to reflect uncertainty in vital rates. We then used the value drawn for that simulation for every year of the projection interval. We drew the adult survival value for each year \( \phi_a(t) \) from a normal distribution with a mean equal to the estimate from the posterior draw and a standard deviation based on a random draw from the posterior for the standard deviation of the temporal random effect on adult survival \( \sigma_{\phi_a} \).

We evaluated scenarios representing unregulated and regulated (i.e., dammed) streams. For the unregulated and regulated stream scenarios, the flow magnitude index was drawn randomly each year from a gamma distribution based on empirical stream discharge (m³/s) data from the SF Eel River (1990–2019) and Alameda Creek below Calaveras Reservoir (1996–2019), respectively (Appendix S1: Fig. S1; Kupferberg et al. 2012). We based egg-mass survival each year on the estimated relationship between \( \phi_{em} \) and flow magnitude (Fig. 3); heavy flows after oviposition (e.g., from late spring rains) can reduce survival of egg masses by scouring them from their attachment sites (Lind et al. 1996, Kupferberg et al. 2012).

### Table 1. Vital rate parameters, prior distributions, and posterior summaries for the integrated population models from Fox Creek and Hurdygurdy Creek. An entry of “...” indicates a derived parameter for which no prior distribution was specified.

| Parameter by population | Symbol | Prior | Median | SD | Lower | Upper |
|-------------------------|--------|-------|--------|----|-------|-------|
| Fox                     |        |       |        |    |       |       |
| Mean adult survival     | \( \phi_a \) | Beta(1, 1) | 0.462 | 0.047 | 0.373 | 0.558 |
| Tadpole to subadult survival | \( \phi_{ts} \) | Beta(1, 1) | 0.002 | 0.001 | 0.002 | 0.003 |
| Hatching survival       | \( \phi_h \) | Beta(1, 1) | 0.815 | 0.002 | 0.811 | 0.820 |
| Egg-mass survival       | \( \phi_{em} \) | ... | 0.889 | 0.148 | 0.515 | 0.980 |
| Survival to recruitment | \( \phi_{rec} \) | ... | 0.002 | 0.000 | 0.001 | 0.003 |
| Mean recapture probability | \( p \) | Beta(1, 1) | 0.474 | 0.130 | 0.233 | 0.754 |
| Mean fecundity per egg mass | \( f \) | Normal(1500, 316) | 1527 | 115 | 1301 | 1753 |
| Temporal variation in \( \phi_a \) | \( \sigma_{\phi_a} \) | Half-Cauchy(0, 1) | 0.334 | 0.153 | 0.021 | 0.616 |
| Temporal variation in \( p \) | \( \sigma_{p} \) | Half-Cauchy(0, 1) | 1.011 | 0.630 | 0.275 | 2.332 |
| Logit-scale intercept of \( \phi_{em} \) | \( \alpha_{em} \) | Normal(2.3, 0.55) | 3.668 | 0.225 | 3.228 | 4.109 |
| Slope of flow effect on \( \phi_{em} \) | \( \beta_{flow} \) | Normal(0, 100) | -0.980 | 0.090 | -1.158 | -0.808 |
| Hurdygurdy               |        |       |        |    |       |       |
| Mean adult female survival | \( \phi_{af} \) | Beta(1, 1) | 0.483 | 0.091 | 0.319 | 0.674 |
| Mean adult male survival | \( \phi_{am} \) | Beta(1, 1) | 0.286 | 0.056 | 0.180 | 0.402 |
| Tadpole to subadult survival | \( \phi_{ts} \) | Beta(1, 1) | 0.003 | 0.002 | 0.001 | 0.007 |
| Hatching survival       | \( \phi_h \) | Beta(3.8, 0.75) | 0.846 | 0.188 | 0.407 | 1.000 |
| Egg-mass survival       | \( \phi_{em} \) | Beta(6, 1) | 0.871 | 0.141 | 0.551 | 1.000 |
| Survival to recruitment | \( \phi_{rec} \) | ... | 0.002 | 0.001 | 0.001 | 0.003 |
| Mean female recapture probability | \( p_f \) | Beta(1, 1) | 0.162 | 0.071 | 0.053 | 0.313 |
| Mean male recapture probability | \( p_m \) | Beta(1, 1) | 0.749 | 0.105 | 0.536 | 0.931 |
| Mean fecundity per egg mass | \( f \) | Normal(1526, 350) | 1637 | 314 | 1040 | 2258 |
| Temporal variation in \( \phi_{af} \) | \( \sigma_{\phi_af} \) | Half-Cauchy(0, 1) | 0.402 | 0.290 | 0.000 | 0.974 |
| Temporal variation in \( p_{af} \) | \( \sigma_{p_{af}} \) | Half-Cauchy(0, 1) | 0.523 | 0.420 | 0.000 | 1.359 |

Notes: Parameters for temporal variation in survival \( (\sigma_{\phi_a}) \) and recapture probability \( (\sigma_{p}) \) are presented on the logit scale. Lower and upper limits from the posterior distribution represent the highest posterior density interval. SD is standard deviation.
the summer reduce tadpole survival by reducing algal resources, slowing growth, and washing tadpoles downstream (Kupferberg et al. 2009, 2011, Catenazzi and Kupferberg 2013). For the regulated stream scenario, the number of summer pulsed flows from upstream dams was drawn each year from a Poisson (rate = 1) distribution. The survival rate of tadpoles during each pulsed flow was 0.61 (Kupferberg et al. 2009). No summer pulsed flows occur in the
unregulated scenario. Adult and subadult survival rates are equal for the regulated and unregulated stream scenarios.

For both the regulated and unregulated scenarios, we simulated the effect of altering (1) survival from egg to subadult stage ($\phi_{rec}$) and (2) survival of subadult and adult life stages ($\phi_{a}$) by a set percentage (from +50% to −50% in 10% increments) on the probability of quasi-extirpation. We chose these simulated reductions in survival to reflect empirical estimates of the effect of known stressors for each life stage. Recent evidence indicates that declines in some populations of *R. boylii* are related to drought-associated outbreaks of chytridiomycosis caused by *Batrachochoytium dendrobatidis* (hereinafter Bd; Adams et al. 2017a). During outbreak events, mortality rates of post-metamorphic frogs in the closely related *Rana muscosa* and *Rana sierrae* can be very high (Rachowicz et al. 2006, Briggs et al. 2010). In anuran populations in the western United States where Bd is endemic, the effects of chytridiomycosis on post-metamorphic survival ranged from a 6% reduction to as high as 55% (Russell et al. cosis on post-metamorphic survival ranged from anuran populations in the western United States high (Rachowicz et al. 2006, Briggs et al. 2010). In

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For each of the above scenarios, we ran simulations with the initial abundance of adult females (\(N_{0,ad}\)) set to 25, 50, 100, and 250 individuals/km based on the range of egg-mass densities (no./km) observed at SF Eel River, Hurdygurdy Creek, and other rivers in central and northern California (Catenazzi and Kupferberg 2017). We set a quasi-extirpation threshold of 10 adult females per river km; if the abundance of adult females dropped below this threshold, we considered the simulated population functionally extinct. We calculated the probability of quasi-extirpation as the proportion of simulations that dropped below the quasi-extirpation threshold during the 20-yr projection interval. We ran the PVA using custom R code that is available on ScienceBase (Rose et al. 2020).

**Results**

We made 571 captures of 340 frogs at Fox Creek from 2014 to 2019, including 145 females, 182 males, and 13 individuals of unknown sex. Of these 340 frogs, 270 frogs were captured in only one year, 53 were captured in two years, 15 were captured in three years, and two were captured in four years. The CJS (\(P = 0.36\)) and egg-mass count sub-models (\(P = 0.56\)) of the Fox Creek IPM showed no evidence of a lack of fit to the observed data. There was no evidence for differences in survival (\(\phi_{l} - \phi_{d} = 0.014, 95\% \text{ HPDI} = -0.14 \text{ to } 0.17\)) and recapture rates (\(p_{r} - p_{m} = -0.001, 95\% \text{ HPDI} = -0.24 \text{ to } 0.23\)) between female and male *R. boylii* at Fox Creek. Therefore, we focused our inference on an IPM where we pooled captures of both sexes to estimate adult survival (Table 1). The estimated mean annual adult survival rate at Fox Creek was 0.46 (95% HPDI = 0.37–0.56). When transformed back to the probability scale, the standard deviation of annual variation in adult survival was 0.08 (0.01–0.14) or approximately 17% of the average annual survival rate of adults at Fox Creek.

The median estimate of survival from the egg to subadult stage (\(\phi_{rec}\)) for a year with average post-oviposition flow was 0.0016 (95% HPDI = 0.0007–0.0026; Appendix S1: Fig. S2). There was a negative relationship between egg-mass survival and the post-oviposition flow magnitude index (\(\phi_{flow} = -0.98, -1.16 \text{ to } -0.81\); egg-mass
survival was lower in years with a greater difference between maximum and minimum post-oviposition flows (Fig. 3). Given the relationship between egg-mass survival and the magnitude of post-oviposition flows, the proportion of eggs that survive to reach the subadult stage ($\phi_{rec}$) can vary over time. For the lowest post-oviposition flow magnitude measured at the SF Eel River, survival from egg to the subadult stage would be 0.0018 (0.0011–0.0027), whereas for the highest measured post-oviposition flow magnitude, survival to subadult would only be 0.0006 (0.0004–0.0010). With an average of 1526 eggs per egg mass, assuming a 1:1 sex ratio and no immigration or emigration of recruits, only 1.3 (0.8–1.8) females per egg mass are expected to reach two years of age on average; recruitment of two-year-old frogs could vary from 0.5 (0.3–0.8) females per egg mass in a bad flow year to 1.4 (0.9–2.0) females in a good flow year.

The abundance of adult female *R. boylii* at Fox Creek fluctuated from 1993 to 2019 (Fig. 3 A) and closely mirrored fluctuations of egg-mass counts in the larger SF Eel River study reach (Appendix S1: Fig. S3). The average population growth rate, log $\lambda_0$, indicated a stable population at Fox Creek over this time period (median = 0.000, 95% HPDI = −0.012 to 0.011). Despite the apparent stability in the long term, the abundance of adult females fluctuated substantially from year to year. The annual population growth rate, $\lambda[t]$, at Fox Creek was positively correlated with $\phi_a[t]$ ($r = 0.36$; 95% HPDI = −0.05 to 0.68) and with egg-mass survival three years earlier, $\phi_{em}[t−3]$ ($r = 0.32$; 0.14–0.49). Although the median correlation between $\lambda[t]$ and $\phi_{em}[t−3]$ was lower, this relationship was less uncertain than the relationship between $\lambda[t]$ and $\phi_a[t]$ (Fig. 4). The effect of low $\phi_{em}$ on $\lambda[t]$ is apparent for two observed declines in $N_{ad}$. High post-oviposition flow magnitude resulted in low $\phi_{em}$ in 2005, which was followed by a decline in $N_{ad}$ from 2008 to 2009 with estimated $\lambda = 0.74$ (0.51–1.02). Similarly, low $\phi_{em}$ in 2009 was followed by a decline in $N_{ad}$ from 2012 to 2013, with estimated $\lambda = 0.72$ (0.51–0.97; Fig. 3). Stream discharge at the SF Eel River study reach during the post-oviposition period appeared more variable from 1990 to 2019 than during the mid-20th century, with years of both higher and lower post-oviposition flow magnitude recorded from 1946 to 1970 (Appendix S1: Fig. S4).

![Fig. 4. Correlation between annual population growth rate, $\lambda_0$, and time-varying demographic rates of *Rana boylii* at Fox Creek, California, USA. (A) Adult survival, $\phi_a$, and (B) survival of egg masses to stranding or scouring, $\phi_{em}$. Black circles are posterior medians, and gray lines are 95% highest posterior density intervals (HPDIs) for each parameter. $r$ is the median correlation coefficient between $\lambda_t$ and the survival parameters, followed by the 95% HPDI in parentheses. $P(r > 0)$ is the posterior probability that the correlation coefficient is greater than zero.](https://www.esajournals.org/doi/10.1890/160306)
We made 1052 captures of 319 frogs at Hurdygurdy Creek from 2002 to 2008, including 121 females and 198 males. Of these 319 frogs, 258 were captured in only one year, 47 in two years, 12 in three years, and 2 in four years. The CJS ($P = 0.19$) and egg-mass count sub-models ($P = 0.57$) of the Hurdygurdy Creek IPM showed no evidence of a lack of fit to the observed data. Sex differences were apparent in the survival and recapture rates of adult frogs from Hurdygurdy Creek; females had higher survival and lower recapture rates than males (Table 1). The estimated survival rate of adult females at Hurdygurdy Creek (0.48, 0.32–0.67) was similar to that of those at Fox Creek. The standard deviation of annual variation in female adult survival was 0.10 (<0.01–0.23) or approximately 21% of the average annual survival rate of adults at Hurdygurdy Creek. The estimated survival rate from egg to subadult at Hurdygurdy Creek was also similar to that at Fox Creek (0.0019, 0.0008–0.0035), although the median estimate for $\phi_{rec}$ was slightly higher at Hurdygurdy Creek (Appendix S1: Fig. S2). The abundance of adult female $R. boylii$ at Hurdygurdy Creek was stable from 2002 to 2008 (Fig. 3B; median log $\lambda_a = 0.019$, −0.034 to 0.072).

Of the scenarios evaluated in the PVA, the biggest influence on quasi-extirpation probability was whether the simulation represented a regulated or unregulated stream. The probability of quasi-extirpation over a 20-yr period was much higher for simulated populations in regulated streams, which experienced pulsed summer flows that reduced tadpole survival, compared to unregulated streams without pulsed flows (Fig. 5). Within a given flow scenario, reducing $\phi_a$ produced a rapid increase in the probability of quasi-extirpation. In comparison, reducing $\phi_{rec}$ had a weaker effect on quasi-extirpation, particularly for the unregulated scenario. Increasing the survival rate of adults and subadults resulted in much lower probabilities of quasi-extirpation compared to equivalent increases in the survival rate from egg to subadult.

**Discussion**

In contrast to the pernicious loss of amphibian biodiversity globally (Scheele et al. 2019), the population growth rate for both focal populations of $R. boylii$ was stable when assessing the study period as a whole, despite inter-annual fluctuations in abundance and recruitment, as well as multi-year periods of apparent decline. Measuring stability of populations requires a long time series relative to a species’ generation time (Connell and Sousa 1983); indeed, had we sampled the Fox Creek population for only a five- or 10-yr period we might have concluded it was declining (e.g., from 2011 to 2015) or growing (2002–2011). The dynamic equilibrium of the populations is notable in light of the survival rate of adult female $R. boylii$ (0.46–0.48), which is lower than estimates of adult survival for three declining $Rana$ species in California: 0.73 in $R. sieracae$ (Fellers et al. 2013), 0.80 in $R. muscosa$, and 0.58 in $Rana draytonii$ (Russell et al. 2019), but similar to reintroduced $Rana pretiosa$ in Oregon (0.40; Duarte et al. 2017). A potential explanation for the apparent stability might be that episodes of recruitment into the adult population in some years compensate for periods of low recruitment and low adult survival. This interpretation is supported by: (1) the estimated threefold variation in survival from egg to subadult, driven by fluctuations in the magnitude of stream flow following oviposition; and (2) the positive correlation between annual population growth rates for and the estimated survival rate of egg masses laid three years earlier. An alternative explanation for the observed stability is that immigration of adult females from outside of our sampling area offset losses due to death and emigration from the sampling area. Because the model estimated apparent survival, true survival could be higher if females emigrated from our study area and were unavailable for capture. It is unlikely that immigration and emigration were the main drivers of the observed dynamics at Fox Creek, because we observed similar fluctuations in egg-mass counts for a much longer reach of the SF Eel River over the same time period. The congruence in vital rate estimates from the two models suggests similar demographic processes govern population growth for these two populations inhabiting unregulated streams in northwestern California. Population densities of $R. boylii$ in northwestern California are robust relative to those in Oregon (Olson and Davis 2009), the Sierra Nevada, and Coast Ranges of central and southern California (California
Department of Fish and Wildlife 2019). Therefore, demographic patterns observed in this study might not be found in other parts of the species’ range where hydrologic regimes differ (e.g., snowmelt driven in the Sierra or xeric conditions in the south).

The importance of recruitment to the population dynamics and persistence of R. boylii heightens the need for precise estimates of the survival rate of early life stages. We did not have data that directly informed the survival rate from the egg stage to the subadult stage. However, based

Fig. 5. Quasi-extirpation probability over 20 yr as a function of the percentage change in survival rate of different life stages of Rana boylii. Each scenario was run for 10,000 simulations with a quasi-extinction threshold of 10 adult females per river km and an initial population of (A) 25, (B) 50, (C) 100, or (D) 250 adult females per river km. Red lines represent the regulated stream scenario, and black lines represent the unregulated stream scenario. \( \phi_{\text{rec}} \) = change in survival from egg to subadult. \( \phi_a \) = change in survival of all adults and subadults.
upon the fecundity of *R. boylii*, adult survival rates (estimated from CMR data), and trends in the adult female population (from egg-mass counts), we were able to obtain estimates of the proportion of eggs that survive to two years of age. Because we had concurrent egg-mass counts as an index of adult female abundance and CMR data to estimate adult survival, the IPM could constrain vital rate estimates to realistic values that matched the observed relative stability in abundance averaged over the time series for each population. Leveraging multiple data sources that provide information about the same vital rate yields more precise and realistic estimates and is one of the great advantages of IPMs (Schaub and Abadi 2011), although estimates of additional parameters can be sensitive to model assumptions (Riecke et al. 2019).

The average survival of early life stages is not the ultimate driver of the population dynamics of *R. boylii*; however, mortality of eggs and tadpoles varies greatly from year to year due to hydrologic conditions (Kupferberg et al. 2012) and is likely to increase in variability as climate changes (Granatham et al. 2018). In some years, survival from egg to subadult is higher than our average estimate, which could result in an episode of high recruitment, whereas in other years, survival to subadult is lower, potentially resulting in little recruitment. Such episodic recruitment events are common among stream-breeding fish, where environmental variability can be strongly linked to larval survival and productivity (Sakaris and Irwin 2010, Lobón-Cerviá 2014). Therefore, predicting the viability of any population of *R. boylii* is fraught, because its fate will be inextricably linked to the joint effect of temperature and precipitation on hydrological conditions in the future. Even these robust populations may not be immune to the effects of climate change. Our retrospective analysis of post-oviposition flow fluctuation in the SF Eel River revealed greater variability from 1990 to 2019 than during the mid-20th century, and California streamflow predictions for the mid-21st century (2040–2069) forecast that trend will continue. Projections informed by global climate models indicate that May (typically when most *R. boylii* breeding occurs) will experience the largest decline in monthly flow relative to a baseline (1975–1999) in coastal northern California, and change in May flows will be the greatest in the xeric central and southern regions (Granatham et al. 2018), where *R. boylii* is largely extirpated (Adams et al. 2017b).

Population models for organisms as disparate as butterflies and lizards show that modeling the response of each life stage is important for evaluating the outcome of future environmental scenarios (Radchuk et al. 2013, Levy et al. 2015). Creating detailed population models can be challenging for species with cryptic life stages, however. Rather than forecasting the trajectories of our study populations, the best use of PVA is to evaluate the relative risk of extirpation under different hypothetical management scenarios (Lindenmayer and Possingham 1996, Akçakaya and Sjögren-Gulve 2000). Our approach of combining multiple data sources in an IPM to estimate a vital rate that was not directly observable and using vital rate estimates to evaluate relative extinction risk under different management scenarios could be applied to many species with difficult-to-study life stages. Two key considerations for applying our methods are (1) the life cycle model of the focal species must reflect the data available to inform each vital rate parameter (i.e., one cannot estimate multiple vital rates that are each lacking empirical data) and (2) simulated scenarios should reflect natural or human-induced perturbations of the estimated vital rates to explore which interventions and vital rates influence population viability the most.

For *R. boylii*, PVA scenarios could include efforts to mimic ecologically relevant aspects of a river’s natural flow regime, to augment survival of eggs and tadpoles via in situ rearing, or other types of conservation translocation. Each of these management methods is increasingly employed to address biodiversity loss and reverse population extirpation (Poff et al. 1997, Harding et al. 2016, Bubac et al. 2019). Population viability analysis results demonstrated that reduced survival rates of early life stages (eggs, tadpoles), designed to simulate the effect of ill-timed releases of water in regulated rivers, could have a large effect on the probability of extirpation. This result reinforces the finding from a regional study of population trends in *R. boylii*, which concluded that altered flow regimes lead to declines in abundance and extirpation downstream of large dams (Kupferberg et al. 2012), as well as
the global finding that reversing the degradation of riverine biota depends on imitating natural flow regimes (Palmer and Ruhi 2019).

In our simulation of disease effects, decreasing the survival of subadult and adult frogs by 10–20% led to a much-increased probability of extirpation in modeled populations, even those in unregulated streams. A recent die-off of post-metamorphic *R. boylii* at a stream in central California, likely caused by chytridiomycosis, raises concerns that conditions favorable to Bd outbreaks, such as drought that crowds animals together or the presence of Bd-tolerant bullfrogs, could cause population-level declines (Adams et al. 2017a). Bd epizootics in other *Rana* species can lead to high mortality in post-metamorphic life stages, resulting in rapid extirpation (Briggs et al. 2010). In populations of western *Rana* species with enzootic Bd, adult female survival was reduced by 6–12%, but for western toads (*Anaxyrus boreas*), survival was reduced by up to 55% (Russell et al. 2019). Based on our PVA, even small reductions in adult survival could be enough to drive population declines and increase the risk of extirpation. We detected Bd on frogs from Fox Creek in 2018 and 2019 (USGS, unpublished data), but the potential effect of the pathogen on the survival of post-metamorphic frogs in this population is not known.

Although altering the survival of adult and subadult frogs had a greater influence on population viability than an equivalent change to the survival from egg to subadult, pre-metamorphic life stages are likely more amenable to management intervention. Head-starting of early life stages is a popular strategy to rescue amphibian species on the verge of extinction (Griffiths and Pavajeau 2008). In addition to the aforementioned effects of flow on survival of egg masses, it has been shown that *R. boylii* tadpoles can survive to metamorphosis at a higher rate when reared in an in situ enclosure and fed high-quality food (Catanzazi and Kupferberg 2013). The difference in egg-mass survival between benign and highly volatile post-oviposition flow regimes can be >50%, illustrating the benefits of avoiding high post-oviposition flows in regulated streams. In contrast, it is unlikely that management actions could increase adult survival even 1.5–1.75 times greater than its average value of 0.46–0.48. Compared to the survival of early life stages, the natural variability of adult survival was low, even though the 2014–2019 CMR period at Fox Creek included both the most severe multi-year drought of the last 1200 yr (Griffin and Anchukaitis 2014) and extreme winter flooding. This stability in adult survival could mean population growth is driven more by the variation in early life stage survival (Gaillard et al. 2000). It is valuable to consider not only the proportional effect of management actions, but also the range of vital rates that are possible in natural populations (Mills et al. 1999).

There are gaps in the data we have available to estimate demographic rates for *R. boylii*, and future studies could improve on this work by using novel methods to collect empirical data that could fill these gaps. For example, in the PVA we assumed that the survival rate of post-metamorphic life stages was the same in regulated and unregulated streams. It would be valuable to collect concurrent egg-mass counts, adult CMR data, and fecundity data from a population of *R. boylii* inhabiting a regulated stream to see whether the demography of post-metamorphic life stages differs from unregulated streams. Also, we assumed that all females breed each year and therefore that the number of egg masses could represent the abundance of adult females in the population. Female ranid frogs might not always breed annually; for example, female *R. pretiosa* are known to skip breeding in some years (Turner 1960). If adult female *R. boylii* skip years of breeding, then our estimated vital rates could be biased, because only changes in survival rates were able to account for increased numbers of egg masses in certain years in our model (e.g., 2006, 2011, 2017). It would be valuable to link the CMR data for adult females to the egg-mass count data by tracking the identities of which females return to the oviposition sites to breed each year. Because we conducted our CMR sampling at Fox Creek in the non-breeding, summer habitat, our captures of adult females are not always representative. In contrast to the survival of adult females, the number of egg masses deposited in certain years may represent the abundance of adults that breed that year.
environmental factors that drive females’ reproductive decisions.

**SUMMARY AND CONCLUSIONS**

Collecting concurrent egg-mass count and CMR data and analyzing these data in an IPM enabled us to obtain precise estimates of adult survival for *R. boylii* and to estimate the low survival rate of unobserved early life stages. Researchers studying species with complex life histories can employ IPMs to gain inference into the vital rates of life stages that are challenging to study directly. A relatively small increase in effort expended, to collect data that complement an existing study, can have great benefits for modeling population dynamics. Still, our understanding of the population dynamics of *R. boylii* would benefit from more data on the survival of unobserved life stages, such as recently metamorphosed frogs, along with potential drivers of temporal variation in adult survival. Given the increased probability of extirpation of *R. boylii* in regulated streams due to low survival of pre-metamorphic life stages, threats that could increase the mortality of adult frogs, such as drought-induced outbreaks of chytridiomycosis, could greatly threaten extant populations. Active conservation efforts, such as habitat enhancement, flow management, and head-starting of early life stages, may be needed for the continued persistence of populations.

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**LITERATURE CITED**

Abadi, F., C. Barbraud, and O. Gimenez. 2017. Integrated population modeling reveals the impact of climate on the survival of juvenile emperor penguins. Global Change Biology 23:1353–1359.

Abadi, F., O. Gimenez, R. Arlettaz, and M. Schaub. 2010. An assessment of integrated population models: bias, accuracy, and violation of the assumption of independence. Ecology 91:7–14.

Adams, A. J., S. J. Kuperberg, M. Q. Wilber, A. P. Pessier, M. Greifrud, S. Bobzien, V. T. Vredenburg, and C. J. Briggs. 2017a. Extreme drought, host density, sex, and bullfrogs influence fungal pathogen infection in a declining lotic amphibian. Ecosphere 8:e01740.

Adams, A. J., A. P. Pessier, and C. J. Briggs. 2017b. Rapid extirpation of a North American frog coincides with an increase in fungal pathogen prevalence: historical analysis and implications for reintroduction. Ecology and Evolution 7:10216–10232.

Akçakaya, H. R., and P. Sjögren-Gulve. 2000. Population viability analyses in conservation planning: an overview. Ecological Bulletins 48:9–21.

Altwegg, R., and H. Reyer. 2003. Patterns of natural selection on size at metamorphosis in water frogs. Evolution 57:872–882.

Barnosky, A. D., et al. 2011. Has the Earth’s sixth mass extinction already arrived? Nature 471:51–57.

Briggs, C. J., R. A. Knapp, and V. T. Vredenburg. 2010. Enzootic and epizootic dynamics of the chytrid fungal pathogen of amphibians. Proceedings of the National Academy of Sciences USA 107:9695–9700.

Brook, B. W., J. J. O’Grady, A. P. Chapman, M. A. Burgman, H. R. Akçakaya, and R. Frankham. 2000. Predictive accuracy of population viability analysis in conservation biology. Nature 404:385–387.

Brooks, S. P., and A. Gelman. 1998. General methods for monitoring convergence of iterative simulations. Journal of Computational and Graphical Statistics 7:434–455.

Bubac, C. M., A. C. Johnson, J. A. Fox, and C. I. Cullingham. 2019. Conservation translocations and post-release monitoring: identifying trends in failures, biases, and challenges from around the world. Biological Conservation 238:108239.

California Department of Fish and Wildlife. 2019. A status review of the Foothill Yellow-legged Frog (*Rana boylii*) in California. California Department of Fish and Wildlife, Sacramento, CA.

Caswell, H. 2001. Matrix population models. Second edition. Sinauer, Sunderland, Massachusetts, USA.

Catenazzi, A., and S. J. Kuperberg. 2013. The importance of thermal conditions to recruitment success in stream-breeding frog populations distributed...
across a productivity gradient. Biological Conservation 168:40–48.

Catenazzi, A., and S. J. Kupferberg. 2017. Variation in thermal niche of a declining river-breeding frog: from counter-gradient responses to population distribution patterns. Freshwater Biology 62:1255–1265.

Colchero, F., et al. 2019. The diversity of population responses to environmental change. Ecology Letters 22:342–353.

Connell, J. H., and W. P. Sousa. 1983. On the evidence needed to judge ecological stability or persistence. American Naturalist 121:789–824.

Cook, D. G., S. White, P. White, and E. White. 2012. Rana boylii (Foothill Yellow-legged Frog). Upland movement. Herpetological Review 43:325–326.

Cormack, R. M. 1964. Estimates of survival from the sighting of marked animals. Biometrika 51:429–438.

Davidson, C., H. B. Shaffer, and M. R. Jennings. 2002. Spatial tests of the pesticide drift, habitat destruction, UV-B, and climate-change hypotheses for California amphibian declines. Conservation Biology 16:1588–1601.

Denwood, M. J. 2016. runjags: an R package providing interface utilities, model templates, parallel computing methods and additional distributions for MCMC models in JAGS. Journal of Statistical Software 71:1–25.

Dirzo, R., H. S. Young, M. Galetti, G. Ceballos, N. J. B. Isaac, and B. Collen. 2014. Defaunation in the Anthropocene. Science 345:401–406.

Duarte, A., C. A. Pearl, M. J. Adams, and J. T. Peterson. 2017. A new parameterization for integrated population models to document amphibian reintroductions. Ecological Applications 27:1761–1775.

Earl, J. E. 2019. Evaluating the assumptions of population projection models used for conservation. Biological Conservation 237:145–154.

Fellers, G. M., P. M. Kleeman, D. A. W. Miller, B. J. Halstead, and W. A. Link. 2013. Population size, survival, growth, and movements of Rana sierrae. Herpetologica 69:147–162.

Fieberg, J., and S. P. Ellner. 2000. When is it meaningful to estimate an extinction probability? Ecology 81:2040–2047.

Furey, P. C., S. J. Kupferberg, and A. J. Lind. 2014. The perils of unpalatable periphyton: Didymosphenia and other mucilaginous stalked diatoms as food for tadpoles. Diatom Research 29:267–280.

Gaillard, J.-M., M. Festa-Bianchet, N. G. Yoccoz, A. Loison, and C. Toigo. 2000. Temporal variation in fitness components and population dynamics of large herbivores. Annual Review of Ecology and Systematics 31:367–393.

Grantham, T. E. W., D. M. Carlisle, G. J. McCabe, and J. K. Howard. 2018. Sensitivity of streamflow to climate change in California. Climatic Change 149:427–441.

Griffin, D., and K. J. Anchukaitis. 2014. How unusual is the 2012–2014 California drought? Geophysical Research Letters 41:9017–9023.

Griffiths, R. A., and L. Pavajeau. 2008. Captive breeding, reintroduction, and the conservation of amphibians. Conservation Biology 22:852–861.

Harding, G., R. A. Griffiths, and L. Pavajeau. 2016. Developments in amphibian captive breeding and reintroduction programs. Conservation Biology 30:340–349.

Hayes, M. P., C. A. Wheeler, A. J. Lind, G. A. Green, and D. C. Macfarlane. 2016. Foothill Yellow-legged Frog conservation assessment in California. General Technical Report PSW-GTR-248. U.S. Department of Agriculture, Albany, CA.

Howard, C., C. H. Flather, and P. A. Stephens. 2019. What drives at-risk species richness? Environmental factors are more influential than anthropogenic factors or biological traits. Conservation Letters 12:e12624.

IPBES. 2019. Summary for policymakers of the global assessment report on biodiversity and ecosystem services of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services. Page 56 in S. Díaz et al. editors. Report of the Plenary of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services on the Work of Its Seventh Session. IPBES, Bonn, Germany.

Jolly, G. M. 1965. Explicit estimates from capture-recapture data with both death and immigration-stochastic model. Biometrika 52:225–247.

Kéry, M., and M. Schaub. 2012. Bayesian population analysis using WinBUGS: a hierarchical perspective. Academic Press, Waltham, Massachusetts, USA.

Kupferberg, S. J. 1997. Bullfrog (Rana catesbeiana) invasion of a California river: the role of larval competition. Ecology 78:1736–1751.

Kupferberg, S. J., A. J. Lind, and W. J. Palen. 2009. Pulsed flow effects on the Foothill Yellow-Legged Frog (Rana boylii): population modeling. California Energy Commission, Sacramento, CA.

Kupferberg, S. J., A. J. Lind, V. Thill, and S. M. Yarnell. 2011. Water velocity tolerance in tadpoles of the Foothill Yellow-legged Frog (Rana boylii): swimming performance, growth, and survival. Copeia 2011:141–152.

Kupferberg, S. J., W. J. Palen, A. J. Lind, S. Bobzien, A. Catenazzi, J. Drennan, and M. E. Power. 2012. Effects of flow regimes altered by dams on survival, population declines, and range-wide losses of California river-breeding frogs. Conservation Biology 26:513–524.
Levy, O., L. B. Buckley, T. H. Keitt, C. D. Smith, K. O. Boateng, D. S. Kumar, and M. J. Angilletta. 2015. Resolving the life cycle alters expected impacts of climate change. Proceedings of the Royal Society B: Biological Sciences 282:20150837.

Lind, A. J. 2005. Reintroduction of a declining amphibian: determining an ecologically feasible approach for the Foothill Yellow-legged Frog (Rana boylii) through analysis of decline factors, genetic structure, and habitat associations. Dissertation. University of California, Davis, Davis, California, USA.

Lind, A. J., H. H. Welsh, and R. A. Wilson. 1996. The effects of a dam on breeding habitat and egg survival of the Foothill Yellow-legged Frog (Rana boylii) in northwestern California. Herpetological Review 27:62–67.

Lindenmayer, D. B., and H. P. Possingham. 1996. Ranking conservation and timber management options for Leadbeater’s Possum in southeastern Australia using population viability analysis. Conservation Biology 10:235–251.

Lobón-Cerviá, J. 2014. Recruitment and survival rate variability in fish populations: Density-dependent regulation or further evidence of environmental determinants? Canadian Journal of Fisheries and Aquatic Sciences 71:290–300.

McGowan, C. P., M. C. Runge, and M. A. Larson. 2011. Incorporating parametric uncertainty into population viability analysis models. Biological Conservation 144:1400–1408.

Mills, L. S., D. F. Doak, and M. J. Wisdom. 1999. Reliability of conservation actions based on elasticity analysis of matrix models. Conservation Biology 13:815–829.

Morris, W. F., and D. F. Doak. 2002. Quantitative conservation biology. Theory and practice of population viability analysis. Sinauer, Sunderland, Massachusetts, USA.

Moyle, P. B. 1973. Effects of introduced Bullfrogs, Rana catesbeiana, on the native frogs of the San Joaquin Valley, California. Copeia 1973:18–22.

Oberhauser, K., R. Wiederholt, J. E. Diffendorfer, D. Semmens, L. Ries, W. E. Thogmartin, L. Lopez-hoffman, and B. Semmens. 2017. A trans-national monarch butterfly population model and implications for regional conservation priorities. Ecological Entomology 42:51–60.

Olson, D. H., and R. J. Davis. 2009. Conservation assessment for the Foothill Yellow-legged Frog (Rana boylii) in Oregon, version 2.0. U.S.D.A. Forest Service Region 6 and U.S.D.I. Bureau of Land Management Interagency Special Status Species Program.

Otero, J., A. J. Jensen, J. H. L’Abée-Lund, N. C. Stenseth, G. O. Storvik, and L. A. Vøllestad. 2011. Quantifying the ocean, freshwater and human effects on year-to-year variability of one-sea-winter Atlantic salmon angled in multiple Norwegian Rivers. PLOS ONE 6:e24005.

Palmer, M., and A. Ruhli. 2019. Linkages between flow regime, biota, and ecosystem processes: implications for river restoration. Science 365:eaaw2087.

Plummer, M. 2003. JAGS: a program for analysis of Bayesian graphical models using Gibbs sampling. Pages 1–8 in K. Hornik, F. Leisch & A. Zeileis editor, Proceedings of the 3rd International Workshop on Distributed Statistical Computing (DSC 2003). Technische Universität Wien, Vienna, Austria.

Poff, N. L., et al. 2010. The ecological limits of hydrologic alteration (ELOHA): a new framework for developing regional environmental flow standards. Freshwater Biology 55:147–170.

Poff, N. L. R., J. D. Allan, M. B. Bain, J. R. Karr, K. L. Prestegaard, B. D. Richter, R. E. Sparks, and J. C. Stromberg. 1997. The natural flow regime. BioScience 47:769–784.

R Core Team. 2020. R: a language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria.

Rachowicz, L. J., R. A. Knapp, J. A. T. Morgan, M. J. Stice, V. T. Vredenburg, J. M. Parker, and C. J. Briggs. 2006. Emerging infectious disease as a proximate cause of amphibian mass mortality. Ecology 87:1671–1683.

Radchuk, V., C. Turture, and N. Schtickzelle. 2013. Each life stage matters: the importance of assessing the response to climate change over the complete life cycle in butterflies. Journal of Animal Ecology 82:275–285.

Riecke, T. V., P. J. Williams, T. L. Behnke, D. Gibson, A. G. Leach, B. S. Sedinger, P. A. Street, and J. S. Sedinger. 2019. Integrated population models: model assumptions and inference. Methods in Ecology and Evolution 10:1072–1082.

Rose, J. P., S. J. Kupferberg, C. A. Wheeler, P. M. Klemman, and B. J. Halstead. 2020. Code and Data to fit an Integrated Population Model for the Foothill Yellow-legged Frog, Rana boylii in northern California. U.S. Geological Survey. https://doi.org/10.5066/P9N019EK

Russell, R. E., et al. 2019. Effect of amphibian chytrid fungus (Batrachochytrium dendrobatidis) on apparent survival of frogs and toads in the western USA. Biological Conservation 236:296–304.

Sakaris, P. C., and E. R. Irwin. 2010. Tuning stochastic matrix models with hydrologic data to predict the population dynamics of a riverine fish. Ecological Applications 20:483–496.

Schaub, M., and F. Abadi. 2011. Integrated population models: a novel analysis framework for deeper insights into population dynamics. Journal of Ornithology 152:S227–S237.
Scheele, B. C., et al. 2019. Amphibian fungal panzootic causes catastrophic and ongoing loss of biodiversity. Science 363:1459–1463.

Seber, G. A. F. 1965. A note on the multiple-recapture census. Biometrika 52:249–259.

Sturrock, A. M., S. M. Carlson, J. D. Wikert, T. Heyne, S. Nusslè, J. E. Merz, H. J. W. Sturrock, and R. C. Johnson. 2020. Unnatural selection of salmon life histories in a modified riverscape. Global Change Biology 26:1235–1247.

Turner, F. B. 1960. Population structure and dynamics of the western spotted frog, *Rana p. pretiosa* Baird & Girard, in Yellowstone Park, Wyoming. Ecological Monographs 30:251–278.

Twitty, V., D. Grant, and O. Anderson. 1967. Amphibian orientation: an unexpected observation. Science 155:352–353.

United States Fish and Wildlife Service. 2015. Endangered and threatened wildlife and plant; 90-day findings on 31 petitions. Federal Register 80:37568–37579.

Wheeler, C. A., J. B. Bettaso, D. T. Ashton, and H. H. Welsh Jr. 2015. Effects of water temperature on breeding phenology, growth, and metamorphosis of Foothill Yellow-legged Frogs (*Rana boylii*): a case study of the regulated mainstem and unregulated tributaries of California’s Trinity River. River Research and Applications 31:1276–1286.

Williams, B. K., J. D. Nichols, and M. J. Conroy. 2002. Analysis and management of animal populations. Academic Press, San Diego, California, USA.

Zweifel, R. G. 1955. Ecology, distribution, and systematics of frogs of the *Rana boylii* group. University of California Publications in Zoology 54:207–292.

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