INTRODUCTION

Retinoic Acid (ATRA) has been shown to regulate several biological processes, in particular during embryonic development in mammals. Concerning reproduction, ATRA was shown to be crucial for the differentiation of gonocytes and spermatogonia and to induce the onset of meiosis in female primordial germ cell (PGCs)/oogonia and in spermatogonia from puberty onward. STIMULATED BY RETINOIC ACID GENE 8 (STRA8) protein has been proposed as the molecular effecter of such promoting ATRA action. In line with this concept, mouse Stra8 gene, originally identified in Embryonal Stem (ES) and Embryonal Carcinoma (EC) cells after...
In the first helical region of the STRA8-HLH domain, there is a basic Nuclear Localization Sequence that might mediate DNA binding to EBox sequence. However, STRA8 lacks the first glutamate and last arginine residues of the ERXR motif for EBox recognition.

In the present paper, we aimed to characterize the action of STRA8 as a transcriptional regulator and to investigate whether its HLH domain, by mediating the interaction with others HLH protein/s including the germ cell specific bHLH factor SOHLH1 could modulate their transcriptional function.

2 | MATERIALS AND METHODS

2.1 | Cell culture

HEK293T cells and P19 Embryonal Carcinoma (EC) cells (ATCC) were grown in Dulbecco’s modified Eagle’s medium (DMEM· High glucose) with 1% penicillin and streptomycin, 2 mmol/L L-Glutamine, 0.1 mmol/L non-essential amino acid (Sigma-Aldrich), and 10% foetal bovine serum (FBS) (Lonza) under standard culture conditions.

2.2 | Male and female germ cells isolation

Postnatal male germ cells were obtained from 10 days post-partum (dpp) CD1 albino mice as reported. When indicated, separation of KIT-positive from KIT-negative male germ cells was performed by magnetic-activated cell sorting (mini-MACS) with CD117 conjugated microbeads (Miltenyi Biotech) as previously described. Female germ cells were isolated from 14.5 and 16.5 days post-coitum (dpc) ovaries following digestion for 10 minutes in Trypsin/EDTA solution (Lonza) and mechanical disaggregation in a monodispersed cell suspension. The cells were cultured for 30 minutes at 37°C and 5% CO₂ in air in DMEM and samples of cells remaining in suspension, roughly consisting of 70%-80% of oocytes, were collected for RNA preparation.

2.3 | DNA constructs

Plasmids expressing the fusion protein GFP-STRA8 and Myc-STRA8 were constructed as reported. pcDNA3-STRA8 and GST-STRA8 was obtained by subcloning the coding sequence of mouse Stra8 by EcoRI/XhoI digestion of pcDNA3-Myc-Stra8 within pcDNA3.1 vector (Promega) and the C-terminus of pGEX-4T (GE Healthcare) by using restriction enzymes EcoRI and Sall. The deletion mutants GST-HLH-only-STRA8, Myc-HLH-only-STRA8 (aa 1-84), GST-ΔHLH-STRA8 and Myc-ΔHLH-STRA8 (aa 99-393) were obtained amplifying corresponding fragments by using pcDNA3-Myc-Stra8 as template and primers (1-4) as indicated in Table 1. For c-Kit promoter
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TABLE 1 Sequences of the primers used in this study

|   | Primer Facebook | Sequence       |
|---|-----------------|----------------|
| 1 | HLH-Str8 fr      | AGGAATTCTAGGCCACCCCTGGAG |
| 2 | HLH-Str8 rev     | AGGTGAGCTTACAGCTTCTTCTCC |
| 3 | ΔHLH-Str8 fr     | AGGAATCCCCAAAGCTTACAGAGA |
| 4 | ΔHLH-Str8 rev    | AGGTGAGCTTACAGCTCTGGAAAG |
| 5 | c-KIT-LUC-fr     | AGGATCGACCTTCTGGAGATCGCTTTT |
| 6 | c-KIT-LUC-rev    | AGGTCACTGTCACGATCCTGCAAG |
| 7 | Myc-Sohlh1 fr    | AGGAATCATGGGGTGCCCTGGT |
| 8 | Myc-Sohlh1 rev   | AGGTTCAGTCAGGGGGAAATGC |
| 14| E47 for          | ACAGATGAGGTGCTGTCCCTG |
| 15| E47 rev          | TCACAGGTCGCCAGCTGGAATT |
| 16| Stra8 fr         | GTCCCTGGTGTCCACCAAG |
| 17| Stra8 rev        | CACCCGAGGCTCAAGCTTC |
| 18| Gapdh for        | AACTTTGGCATGTGGGAAG |
| 19| Gapdh rev        | CCCTGTTCTACTCCCCAATG |
| 20| Kit fr           | GAGAAGTGACTGTCCCTG |
| 21| Kit rev          | TCATTCTGTATGCTTCTG |
| 22| L34 fr           | GGTGTCAGGCGACCTAGGAT |
| 23| L34 rev          | GTGCTTTCCACCTTCTTG |

construct, (pGL3-CMV-Kit-LUC), a 2149 bp fragment (~2525 to ~376) of mouse Kit promoter (the sequence was derived from EPD Eukaryotic promoter database at https://epd.epfl.ch/mouse/mouse_database.php?db=mouse), was amplified by P19EC cells genomic DNA using primers listed in Table 1 and ligated to the XhoI and NcoI sites of pGL3-CMV-LUC vector (gently provided by Prof. C. Sette). The coding sequence of mouse Sohlh1 (GeneBank NM_001001714), were amplified by PCR from total RNA obtained from P10 male germ cells using primers listed in Table 1 and cloned in BamH1/XhoI sites of pcDNA3 vector. The sequences of the vectors were verified by DNA sequencing (BMR Genomics, Italy). pcDNA3-Flag-E47 was generously provided by Prof. Y. Yokota (University of Fukui, Japan), pGEX-E47 was obtained by Prof. Pura Munoz (Pompeu Fabra University, Barcelona), while V. Saccone (Centro Europeo Ricerca sul cervello, CERC, Rome, Italy) provided pHA-MYOD and pGEX-MYOD plasmids.

2.4 | Cell transfection

HEK293T cells were transiently transfected with the different plasmids by using the jetPei™ Polyplus transfection reagent (Polyplus-Transfection SA, SIC, Italy), according to the manufacturer’s protocol. 2.5 x 10⁶ P19EC cells were electroporated with 3 µg total DNA by using Cell Line Nucleofector kit and AMAXA nucleofector device II (C-20 program) (Amaxa).

2.5 | Immunofluorescence analysis

For immunohistochemistry, serial 6 µm thick sections were obtained from testes of 10 dpp mice, fixed in buffered formalin and paraffin embedded. Slides were dewaxed, rehydrated and microwaved in 10 mmol/L sodium citrate buffer, pH 6 for 20 minutes. After blocking with 10% goat serum (GS), sections were incubated with rabbit polyclonal anti-STR8 (1:400 Ab49405, Abcam) or rabbit polyclonal anti-E47 antibodies (1:200 N-649 sc763, Santa Cruz Biotechnology) diluted in PBS/0.1% BSA/0.1% Triton overnight. After washing with PBS/0.5% Tween, 1:400 goat anti-rabbit (Alexa Fluor 568, Thermo Fisher Scientific Inc) were used as secondary antibodies for 1 hour incubation at RT. Hoechst in PBS was added for 5 minutes and the samples were observed under a TCS SP5 laser-scanning confocal microscope (LEICA microsystem, Switzerland). For HA-MYOD and FLAG-E47 immunolocalization in HEK293T, cells attached to poly-L-lysine-coated coverslips were transfected as indicated and 24 hours after transfection cells were fixed with 4% (v/v) paraformaldehyde for 10 minutes. The cells were permeabilized with 0.1% Triton X-100 in PBS for 5 minutes at RT and after blocking with 10% GS for 45 minutes, the anti-HA antibody (1:500, MMS-101P Covance) and the anti-FLAG antibody (1:500, F3165 Sigma-Aldrich) were applied for 1 hour at RT. After washing with PBS, cells were incubated with goat anti-mouse antibodies (Alexa Fluor, Molecular Probes) for 45 minutes and with Hoechst for 5 minutes. The samples were observed under a Leica CTR600 microscope with a 40X objective.

2.6 | Immunoprecipitation

Transfected HEK293T cells, P10 dpp-isolated male germ cells or P19 EC cells cultured with ATRA (1 µmol/L) for 24 hours were lysed (20 minutes at 4°C) in buffer containing 50 mmol/L Tris-HCl (pH 7.6), 150 mmol/L NaCl, 2 mmol/L EDTA, 0.5% NP-40, 3% glycerol, 10 µg/mL phenylmethylsulfonyl fluoride, and a protease inhibitor mix (Sigma-Aldrich). When indicated, nuclear extracts were
prepared from P10 dpp-isolated male germ cells by homogenizing cells in hypotonic buffer (10 mmol/L Tris/HCl pH 7.4, 10 mmol/L NaCl, 2.5 mmol/L MgCl₂, 1 mmol/L DTT, protease inhibitor cocktail (PIC) (Sigma-Aldrich)). After incubation on ice for 7 minutes, samples were centrifuged at 700 g for 7 minutes. Pelleted nuclei were resuspended in hypotonic buffer supplemented with 90 mmol/L NaCl and 0.5% Triton, sonicated and centrifuged (5000 g for 15’) on 30% sucrose cushion. 1 mg of proteins were pre-cleared with Dynabeads protein G (Life Technologies) for 30 minutes then the extracts were incubated with Dynabeads protein G coupled with control IgG or specific primary antibodies for 2 hours at 4°C under rotation. After washing in the same buffer used for the lysis, precipitates were analysed by Western blotting.

2.7 | Purification of GST fusion proteins

The BL21 strain of *Escherichia coli* was transformed with plasmid pGEX-STRÅ8 (full length and corresponding deletions), pGEX-MYOD or with pGEX-4T1 for expression of GST and grown at 37°C in LB medium until A₆₀₀ was 0.7, at which time isopropyl-β-thiogalactopyranoside (IPTG) was added for 3 hours at a final concentration of 0.5 mmol/L. Bacteria were collected by centrifugation and the pellet was resuspended in GST-extraction buffer (GEB: 20 mmol/L Tris·HCl pH 7.4, 1 mol/L NaCl and 0.2 mmol/L EDTA) with a PIC (Sigma-Aldrich). The suspensions were sonicated and debris removed by centrifugation. Fusion proteins were affinity purified by adsorption to glutathione-agarose beads (Sigma-Aldrich) for 2 hours at 4°C and eluted in the Elution Buffer (EB: 20 mmol/L Glutathione, 100 mmol/L Hepes pH 7.6, 1 mmol/L DTT and 0.1 mmol/L EDTA). Protein concentrations were determined by SDS-polyacrylamide gel electrophoresis using purified BSA as standard and Coomassie gel staining.

2.8 | Pull-down assay

Transfected HEK293T cells and male germ cells were lysed in a pull-down buffer (PB: 25 mmol/L Tris·HCl pH 7.5, 150 mmol/L NaCl, 0.5% Triton-X100, 1 mmol/L EDTA, 5% glycerol and PIC) and protein concentration was determined using a BCA protein assay kit (Pierce). When indicated, GST or GST fusion proteins were cross-linked to the glutathione-agarose beads with Dimethyl pimelimidate (DMP, Sigma-Aldrich). Stock concentration: 13 mg/mL). Briefly, GST protein bound to the GST-agarose were washes 2 times with 200 mmol/L Triethanolamine pH 8.9 and incubated for 1 hour with the cross-linking solution (50 mmol/L DMP in 200 mmol/L Triethanolamine pH 8.9). Cross-link reaction was blocked with 100 mmol/L ethanolamine pH 8.9, and the beads were washed in PBS and used for pull-down experiments. Cell extracts (500 μg-1 mg) were pre-cleared on glutathione-agarose beads for 1 hour at 4°C, then incubated under constant shaking with GST or GST fusion proteins, that were cross-linked or not to the beads, for 2 hours at 4°C. After three washes with the PB buffer, adsorbed proteins were eluted in SDS sample buffer and analysed by Western blotting.

2.9 | Western blot analysis

Cell extracts or immunoprecipitated proteins were diluted in PBS sample buffer and boiled for 5 minutes. Protein was separated on either 10% or 8% SDS-PAGE gels and transferred to PVDF Transfer Membrane Hybond™ (Amersham Bioscience). Membranes were saturated with 5% non-fat dry milk in PBS containing 0.1% Tween20 (PBST) for 1 hour at RT. The antibodies (anti -c-MYC sc764, anti-GFP sc9996, anti-GST sc9996, anti-E47 sc763, anti-GAPDH sc-32233 were from Santa Cruz Biotechnology; anti-Flag F3165 was from Millipore; anti STRÅ8 Ab49405, anti-SOHHL1 Ab41520 and anti-HISTONE H3 Ab1791 were from Abcam; anti c-KIT gently provided by Prof. S. Dolci) were diluted in PBST buffer and added to the PVDF membrane for 1 hour at RT or overnight at 4°C followed by incubation with the appropriate horseradish peroxidase-conjugated secondary antibodies (Amersham Bioscience) for 45 minutes at RT. The STRÅ8 and SOHLH1 immunoprecipitated protein were detected after incubation with the peroxidase-conjugated anti-rabbit IgG light chain specific (Jackson Immuno Research). All proteins were detected with ECL plus detection reagents (Amersham Bioscience) and visualized by chemiluminescence.

2.10 | Luciferase reporter assay

4 × 10⁴ HEK293T cells or P19 EC cells were seeded in 24 wells and transfected with 200 ng of the LUC reporters. The p4RE-LUC (Dr V. Saccone, (Centro Europeo Ricerca sul cervello, CERC, Rome, Italy), consists of 4 copies of right EBox from Muscle Creatine Kinase (MCK) enhancer. AP1, RARE and SXXCRE-DNA-binding sequences were gently provided by Prof. C. Sette, Catholic University of the Sacred Heart, Rome, Italy; Dr D. Baretto, Instituto di Biomedicina de Valencia, Valencia, Spain and Dr PJS Stork, Vollum Institute, Portland, respectively. pGL3-CMV-Kit-LUC was described before. Each well also received 10 ng of a pRL-TK Vector (Promega) to normalize for transfection efficiency. At 48 hours after transfection, cells were washed three times with PBS and scraped in 100 μL of reporter lysis buffer (Promega). Luciferase activity in 20 μL of the cell extracts was quantified using the Dual-Luciferase Reporter assay system (Promega). Each extract was assayed three times with a Hitox luminometer (RadTech, Italy). The Firefly luciferase activity was divided by Renilla luciferase activity and the transcriptional effect expressed as relative activity compared to the control groups.

2.11 | Total RNA isolation, RT-PCR and qRT-PCR

Total RNA was extracted from the cells using Trizol reagent (Invitrogen) following manufacturer instructions. 1 μg of RNA was retrotranscribed using M-MLV reverse transcriptase (Promega).
25 ng of cDNA was used as template for PCR (GoTaq, Promega) and reactions were analysed on agarose gels. For quantitative RT-PCR (qPCR) experiments, SYBR green PCR master mix (Kapa Biosystem) was used following manufacturer’s instructions. L34 gene expression was used for normalization in qPCR experiments. All primers used in these experiments were indicated in Table 1.

### 2.12 Statistical analyses

All experiments were replicated at least three times. Data were expressed as mean ± Standard Deviation (SD). Student’s t-test and one-way ANOVA analysis were performed using Graphpad Prism software.
3 | RESULTS

3.1 | STRA8 interacts with itself in vitro and in vivo through the HLH domain

Since STRA8 possesses a well conserved HLH domain (aa 17-84) that could mediate protein-protein interaction, we tested the ability of the protein to interact with itself. GST and GST-STRA8 fusion proteins were used in pull-down assays with lysates obtained from HEK293T transfected with pcDNA3-Myc-STRA8 or from 10 dpp male germ cells. As shown in Figure 1, both recombinant (1A) and endogenous (1B) STRA8 were able to specifically bind to GST-STRA8 and not to GST alone, thus indicating that the protein efficiently interacts in the in vitro assay. We further investigated whether STRA8 was able to form complexes in intact cells by a co-immunoprecipitation (CO-IP) assay. Efficient co-precipitation of transfected Myc-STRA8 with GFP-STRA8 (Figure 1C) and endogenous STRA8 (Figure 1D) was observed.

To verify if HLH domain was involved in the interaction, we generated two deletion mutants of STRA8, one consisting of the first 84 amino acids (Myc-HLH-only-STRA8, GST-HLH-only-STRA8) and another in which the HLH domain was deleted (aa 99-393, Myc-ΔHLH-STRAT8, GST-ΔHLH-STRA8). We repeated GST pull-down and co-immunoprecipitation assays with these mutants. Lysates obtained from HEK293T cells transfected with pcDNA3-Myc-STRA8 were used in pull-down assays with the fusion proteins GST-HLH-only-STRA8 or GST-ΔHLH-STRA8 and as shown in Figure 1E, the interaction in vitro was evident only with the HLH-only-containing mutant. HEK293T cells were transfected with pcDNA3-Myc-ΔHLH-STRA8 and after 24 hours of culture lysates were immunoprecipitated with an anti-MYC antibody. As shown in Figure 1F, deletion of the HLH domain of STRA8 resulted in a complete loss of the interaction with the full length GFP-STRA8, thus indicating that this STRA8's domain is involved in the interaction.

3.2 | STRA8 interacts in vitro and in vivo with class I bHLH E47 through its HLH domain

Tissue specific bHLHs transcription factors are known to form homo and heterodimers with class I bHLH factors through their HLH domain and this is important for efficient DNA binding. With the aim to identify bHLH transcription factors with which STRA8 interacts in germ cells, we concentrated on E proteins, a class of widely expressed bHLH partners. In particular, we investigate Tcf3/E47 gene, a splicing product of the Tcf2a gene that is known to be involved in multiple differentiation processes. Single cell RNA-sequencing analysis from mouse testis in the perinatal period or from sorted KIT-negative and KIT-positive cells, indicated that E47 is expressed in both spermatogonia populations. Moreover, this gene is also expressed in the preleptotene cells. To verify the expression of E47 in male germ cells, we performed RT-PCR with total RNA obtained from immunomagnetic-purified undifferentiated (KIT-negative) and differentiating (KIT-positive) P10 dpp germ cells. As shown in Figure 2A, we confirmed that E47 was expressed in both cell populations. As a control for the enrichment of the cell separation, Stra8 expression was evaluated, confirming its presence prevalently in the KIT-positive germ cell population (Figure 2A). Immunolocalization of STRA8 in the testis from P10 mice showed an uneven distribution of the protein in different tubules as expected with spermatogonia and preleptotene positive cells (Figure 2B). E47, on the other hand, is expressed in the nuclei of the cells in each tubule thus indicating that STRA8-positive cells also express the bHLH factors at this age. RT-PCR performed in pre-meiotic (12.5 dpc) and meiotic (13.5-14.5 dpc) female germ cells obtained by mouse embryos, showed that E47 transcript was detected in all female germ cell samples analysed (Figure 2C).

By using the co-immunoprecipitation assay, we next investigated whether STRA8 and E47 interacted. A Myc-tagged Stra8 expression vector was transfected into HEK293T cells together with pcDNA3-E47-Flag using a Flag-empty vector as a control and total extracts were subjected to immunoprecipitation with an anti-FLAG antibody. The results in Figure 3A showed that STRA8 was co-immunoprecipitated with E47 and the specificity of the immunoprecipitation was confirmed with an isotype-matched nonspecific mouse IgG. Co-immunoprecipitation performed using total lysates from P10-isolated male germ cells, that include both STRA8 and E47 expressing cells (Figure 2B) confirmed that the anti-E47 antibody co-immunoprecipitated STRA8 also in these cells (Figure 3B). The same result was obtained when anti-STRA8 antibody was used to immunoprecipitate E47 from nuclear extracts of the germ cells (Figure 3C).

To analyse which domain of STRA8 was important for the interaction with E47, we performed a GST pull-down assay. Cross-linked GST-STRA8 and GST-MYOD fusion proteins were used for interaction with Flag-E47 that was transiently transfected in HEK293T cells. GST protein was used as a negative control. As shown in Figure 3D, GST-STRA8 as GST-MYOD specifically interacted in vitro with E47 whereas the control GST protein exhibited no interaction. When this assay was repeated with the deletion mutants GST-HLH-only-STRA8 and GST-ΔHLH-STRA8 fusion proteins, only the mutant with the HLH domain formed a specific complex with E47 (Figure 3E), thus indicating that this region mediates also the interaction of STRA8 with the bHLH factor.

3.3 | STRA8 inhibits bHLH-dependent transcriptional activity through its HLH domain in transfected cells

Although STRA8 is described as a HLH-transcriptional regulator, in the literature it is not clear if the HLH domain is involved in its transcriptional activity, also because when the HLH region was deleted or mutated, STRA8 did not localize to the nucleus. Moreover, STRA8 did not bind the regulated gene promoters to the canonical EBox motif recognized by the tissue specific bHLH transcription factors. Performing a sequence alignment of STRA8 and different bHLH factors, including those expressed in germ cell (MEIOSIN, SOHLH1, SOHLH2) with Clustal Omega program
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(https://www.ebi.ac.uk), we verified that its basic N-terminal region did not actually included the determinants for the EBox recognition20,21 (Figure S1). Therefore, we hypothesize that STRA8, binding to bHLH factors might negatively modulate their action as the basic region-lacking ID protein do.21 To test this, we investigated if STRA8 was able to repress the activity of an EBox reporter stimulated by two bHLH proteins such as MYOD and E47. Transient transfection experiments were performed in HEK293T using the 4RE-LUC reporter and vectors expressing STRA8, E47 and MYOD in different combinations as indicated. As shown in Figure 4A, both MYOD and E47, alone or in combination, were able to significantly stimulate the luciferase transcription, while STRA8 failed to induce the transactivation above the basal level. Conversely, when STRA8 was co-expressed with E47 and MYOD, a significant dose-dependent inhibitory effect on luciferase transactivation induced by E47 and MYOD homodimers and E47/MYOD heterodimers was observed (Figure 4A,B). The reporter used in these assays consists of four copies of Ebox from Muscle Creatin Kinase (Mck) enhancer and it has been shown that the heterodimer MYOD/E47 is more efficient in the trans-activation.30 For this reason, we hypothesized that STRA8 inhibited the MYOD-induced stimulation of reporter activity by interfering with the complex formed by MYOD and endogenous E47. Immunolocalization of E47 and MYOD in transfected HEK293T cells indicated that the two TFs were localized in the nucleus both in the presence and in absence of STRA8-expressing vector (Figure S2). To determine whether the inhibitory activity of STRA8 was related to the HLH domain, the Myc-HLH-only-STRA8 deletion mutant that localizes in the nucleus15 was used in the transcription assay. As shown in Figure 4C, HLH-only-STRA8 reduced the stimulation of luciferase activity by E47-MYOD heterodimers in a dose-dependent manner.

To investigate whether the inhibitory effect of STRA8 was specific on EBox motif, we evaluated the activity of over-expressed Myc-Stra8 in reporter transactivation experiments in which luciferase was under the control of different DNA-binding motifs. As shown in Figure 4D, STRA8 did not affect the basal level of each luciferase reporter and did not interfere with the stimulatory activity of CREB through the AP1 or CRE elements and of RARx through the RARE responsive elements.

3.4 STRA8 interacts with the germ cell specific bHLH factor SOHLH1 and represses its induced c-KIT expression in vitro

SOHLH1 and SOHLH2 (SPERMATOGENESIS- AND OOGENESIS-SPECIFIC BHLH TRANSCRIPTION FACTOR 1 AND 2) are two germ cell specific bHLH transcription factors that have been recently reported to be essential for spermatogonia29-33 and oocyte34-36 differentiation. SOHLH1 and SOHLH2 are able to form homo- and heterodimers.32,33 To examine whether STRA8 was able to interact with SOHLH1, we first evaluated the expression of the proteins in the P10 male germ cells suspension including a heterogeneous population of differentiating spermatogonia and preleptotene/early leptotene spermatocytes. As shown in Figure S3, in isolated germ cells, 85% of STRA8-positive cells were present that expressed different amount of the proteins and 57% of the cells expressing SOHLH1. We then performed GST pull-down assays incubating GST-STRA8 fusion protein with total extracts obtained from HEK293T cells transfected with pcDNA3-Myc-SOHLH1 or from P10 dpp male germ cells. As shown in Figure 5, GST-STRA8 strongly interacted in vitro with over-expressed-SOHLH1

FIGURE 2 E47 expression in postnatal male germ cells and embryonic female germ cells. A, RT-PCR analysis of Stra8, E47 and Gapdh (as a control) expression in germ cells isolated from P10 dpp testis. Undifferentiated KIT- and differentiating KIT+ cells were purified through immunomagnetic sorting as indicated in Materials and Methods. B, Immunodetection of STRA8 and E47 in histological adjacent sections from 10 ddp mouse testis. C, RT-PCR analysis of Stra8, E47 and Gapdh (as a control) expression in female germ cells obtained from ovaries at the indicated embryonal ages.
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(Figure 5A) and with endogenous protein (Figure 5B), whereas the control GST protein exhibited no interaction with either proteins (Figure 5A, B). We then performed co-immunoprecipitation experiments by using total cell extracts obtained from HEK293T cells co-transfected with STRA8- and SOHLH1-expressing vectors. As showed in Figure 5C, recombinant STRA8 was able to interact with over-expressed SOHLH1 also in intact cells. The interaction between STRA8 and SOHLH1 was evident also when the co-immunoprecipitation experiments were repeated immunoprecipitating SOHLH1 from male germ cells nuclear extracts incubated with STRA8 antibody (Figure 5D). It has been demonstrated that SOHLH1 can positively regulate c-Kit expression by binding its promoter at EBox containing sequence. 29,32 To analyse the effect of STRA8 on such stimulatory action by SOHLH1, we first cloned the c-Kit-regulatory promoter region (~2525 to ~376 bp) including the two EBoxes bound by SOHLH1 in KIT+-spermatogonia 29 in pGL3-CMV-LUC vector. P19EC cells were then transfected with Myc-SOHLH1, with or without Myc-tagged STRA8 expression vector, and luciferase activity was measured 48 h after transfection. As shown in Figure 5E c-Kit promoter activity was induced by SOHLH1 construct, while recombinant STRA8 had no effect. Moreover, the co-transfection of STRA8 with SOHLH1 vector reduced the transcriptional activation in a dose-dependent manner. The inhibitory activity of STRA8 was related to the HLH domain because when the Myc-HLH-only-STRA8 deletion mutant was used in the transcription assay, it was able to reduce the stimulation of luciferase activity by SOHLH1 in a dose-dependent manner (Figure 5F).

To verify if STRA8 was also able to negatively modulate the expression of c-Kit, we over-expressed SOHLH1 in P19EC cells (that expresses low level of c-Kit) with or without Myc-STRA8. After 24 hours from transfection, we observed that SOHLH1 induced a significant increase of the c-Kit transcript and when STRA8 was co-transfected with the bHLH factor, it was able to almost completely abolish the c-Kit stimulation by SOHLH1 (Figure 5G). This result was also observed when c-KIT protein expression was analysed by Western blot analysis (Figure 5H).
DISCUSSION

In the mouse embryonal ovary and preleptotene spermatocytes, a complex transcriptional programme of meiosis beginning regulated by STRA8 has been recently revealed.\textsuperscript{13,14,37} The most part of involved genes appear to be engaged in cell cycle regulation, double strand break repair and chromosome synapsis and most of them are expressed before the beginning of the meiotic prophase 1. Curiously, a significant percentage of the STRA8-activated genes is not meiosis/germ cells specific. Accordingly, STRA8 might function...
FIGURE 5 STRA8 interacts with SOHLH1 and down-regulates SOHLH1-induced c-KIT expression in vitro. A-B, Western blot analysis with anti-Myc (A) and anti-SOHLH1 (B) antibodies of the pull-down assays performed with total lysates (TL) from HEK293T expressing pcDNA3-Myc-SOHLH1 (A) or from P10 dpp-isolated male germ cells and fusion proteins GST-TRA8 and GST (as negative control). The amount of purified GST fusion proteins used in the assay was determined by Coomassie staining (A,B lower panels). C, Western blot analysis with anti-TRA8, anti-Myc and anti-GAPDH antibodies of the immunoprecipitation performed with control mouse IgG or anti-Myc antibody of total lysates (TL) of HEK293T expressing pcDNA3-Myc-SOHLH1 and/or pcMV-TRA8. D, The co-immunoprecipitation assay was repeated with nuclear extracts obtained from P10 dpp-isolated male germ cells. SOHLH1 immunoprecipitated by anti TRA8 antibody was revealed in Western blot with anti SOHLH1 antibody (upper panel) and as a control of immunoprecipitation, the membranes was incubated with STRA8 and HISTONE H3 antibodies (lower panel). E, The bar graph represents the relative luciferase activity in HEK293T cells transfected with c-Kit promoter reporter in combination or not with pcDNA3-Myc-SOHLH1 and/or different amount of pcDNA3-Myc-TRA8. A schematic representation of the Eboxes in the c-kit promoter is showed. Data represent mean ± SD of three replicates. *P < 0.05; **P < 0.001; ns, not significant related to control (ordinary one-way ANOVA, Tukey’s multiple comparisons test). F, Different concentrations (300 ng) of Myc-HLH-only Stra8 mutant were analysed in the same Ebox-mediated luciferase assay (mean ± SD, ***P < 0.001; ****P < 0.0001). G-H, qPCR (G) and Western blot (H) analyses of c-KIT expression in P19EC cells transfected with Myc-TRACTA8 and/or Myc-SOHLH1 vectors or control plasmid (mock). G. Fold change of c-KIT expression relative to Gapdh expression was calculated by the ΔΔCq method. Data represent mean ± SD of three replicates. **P < 0.001; ns, not significant (ordinary one-way ANOVA, Tukey’s multiple comparisons test). H, Western blot analysis of c-KIT expression in P19EC cells transfected as above. Over-expression of recombinant proteins was evaluated with anti-Myc antibodies. GAPDH was evaluated as loading control.
expressed in differentiating KIT-positive spermatogonia. Besides, it resulted expressed also in preleptotene enriched germ cell population and up-regulated by ATRA treatment as STRA8 itself. In male spermatogonia, SOHLH1 and SOHLH2 form homo- and heterodimers and bind chromatin at EBox sequences upstream of their own genes and other genes which are essential for spermatogonia differentiation, as for example, c-KIT. The latter, is fundamental in the regulation of the mitotic activity that characterizes the differentiating population of spermatogonia (A1/A2/A3/A4/intermediate/B spermatogonia) and its expression is dependent by ATRA signalling as SOHLH1 and STRA8. It has been hypothesized that the competence to enter meiosis is related to the number of mitotic divisions controlled by the KL/KIT complex. In fact, KIT expression is down-regulated at the time of meiotic entry in spermatocyte as well as in embryonic female PGCs. In line with the notion that the differentiating spermatogonia are therefore heterogeneous cell populations, we previously observed that in prepuberal testes there are cells SOHLH1+/c-KIT+/STRA8+ (differentiating spermatogonia), but also cells SOHLH1+/c-KIT-/STRA8; this latter population could represent early differentiating spermatogonia or the competent-late pre-meiotic spermatogonia. Since c-KIT is important for spermatogonia differentiation and in fact is expressed at high levels in such population, we hypothesize that STRA8 takes part to its negative regulation at the beginning of meiosis interfering with SOHLH1 in late differentiating spermatogonia and preleptotene spermatocytes.

In conclusion, it is possible to postulate that STRA8 is crucial to regulate the spermatogonia cell cycle and differentiation and at the same time for triggering the meiotic programme in different ways. First, with a positive feedback mediated by its direct binding to its own promoter (a consensus STRA8-binding motif CNCCTCAG is located −782 bp from TSS) and to the promoter of the positively regulated genes. Moreover, STRA8 is able to modulate negatively genes regulated by bHLH transcription factors by interfering with their DNA recognition capability. The positive and negative modulatory role of a transcriptional regulator is common also to other transcription factors that can activate or repress target genes in different way, depending on a number of factors such as their concentration, the binding/recruitment of additional proteins to the transcriptional complex or following post-translation modification and not least the epigenetic status of the target cells. The importance of better understanding the c-KIT and STRA8 regulation is not only related to spermatogenesis, but extend also to testicular germ cell tumours. Alteration as mutations, copy number and expression of c-KIT and c-KIT-PI3K pathway are significantly associated to seminoma. STRA8 and the premature germ cell differentiation are associated to susceptibility to testicular teratomas. Therefore, other than for fertility preservation, it is important to understand how STRA8 works to design future therapeutic strategies.

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CONFLICT OF INTEREST
The authors confirm that are not conflicts of interest.

AUTHOR CONTRIBUTIONS
Maria Giovanna Desimio: Conceptualization (equal); Data curation (equal); Formal analysis (equal); Investigation (equal); Eleonora Cesari: Conceptualization (equal); Data curation (equal); Investigation (equal); Massimo De Felici: Conceptualization (equal); Funding acquisition (equal); Supervision (equal); Writing-original draft (equal).
Donatella Farini: Conceptualization (equal); Data curation (equal); Formal analysis (equal); Supervision (equal); Writing-original draft (equal); Writing-review & editing (equal).

DATA AVAILABILITY STATEMENT
All data generated during the study are available from the corresponding author on request.

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SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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