N-terminal heterogeneity of parenchymal and vascular amyloid-β deposits in Alzheimer’s disease

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Abstract

Aims: The deposition of amyloid-β (Aβ) peptides in the form of extracellular plaques in the brain represents one of the classical hallmarks of Alzheimer’s disease (AD). In addition to ‘full-length’ Aβ starting with aspartic acid (Asp-1), considerable amounts of various shorter, N-terminally truncated Aβ peptides have been identified by mass spectrometry in autopsy samples from individuals with AD.

Methods: Selectivity of several antibodies detecting full-length, total or N-terminally truncated Aβ species has been characterized with capillary isoelectric focusing assays using a set of synthetic Aβ peptides comprising different N-termini. We further assessed the N-terminal
heterogeneity of extracellular and vascular Aβ peptide deposits in the human brain by performing immunohistochemical analyses using sporadic AD cases with antibodies targeting different N-terminal residues, including the biosimilar antibodies Bapineuzumab and Crenezumab.

Results: While antibodies selectively recognizing Aβ1-x showed a much weaker staining of extracellular plaques and tended to accentuate cerebrovascular amyloid deposits, antibodies detecting Aβ starting with phenylalanine at position 4 of the Aβ sequence showed abundant amyloid plaque immunoreactivity in the brain parenchyma. The biosimilar antibody Bapineuzumab recognized Aβ starting at Asp-1 and demonstrated abundant immunoreactivity in AD brains.

Discussion: In contrast to other studied Aβ1-x-specific antibodies, Bapineuzumab displayed stronger immunoreactivity on fixed tissue samples than with sodium dodecyl sulfate-denatured samples on Western blots. This suggests conformational preferences of this antibody. The diverse composition of plaques and vascular deposits stresses the importance of understanding the roles of various Aβ variants during disease development and progression in order to generate appropriate target-developed therapies.

Keywords
Alzheimer disease; amyloid; N-terminal truncation; capillary isoelectric focusing immunoassay; antibody; Abeta

Introduction
The deposition of amyloid-β (Aβ) peptides in the form of extracellular plaques in the brain parenchyma is one of the characteristic neuropathological features of Alzheimer’s disease (AD). These Aβ peptides are derived from the larger amyloid precursor protein (APP) and generated by sequential proteolytic cleavage events carried out by the so-called β- and γ-secretases [1]. The Aβ cascade hypothesis postulates a central role of Aβ in the disease process [2] and is supported by genetic findings in familial early onset AD (EOAD) cases. In these patients, autosomal dominant mutations in the genes encoding APP or presenilins (PSEN) have been identified which inevitably lead to EOAD [3,4]. On the other hand, a lack of a robust correlation between extracellular amyloid pathology and the cognitive status of AD patients has questioned the validity of this concept [5,6]. The recent failures of several clinical trials employing therapeutic antibodies directed against Aβ peptides fuelled additional scepticism [7–13]. Aβ immunoprecipitation combined with matrix-assisted laser desorption ionization time-of-flight (TOF) mass spectrometry in brain tissue samples from AD subjects revealed the presence of a variety of Aβ peptides with differences in length due to N- and C-terminal truncations [14–20]. A shortened N-terminus and a longer C-terminus appear to facilitate peptide aggregation into beta-sheet fibrils, and thereby increase their neurotoxicity as compared to full-length Aβ peptides. In addition to N- or C-terminal truncations, post-translational protein modifications such as the formation of N-terminal pyroglutamate residues (AβPE3-x, AβPE11-x) [21,22] or Aβ peptide phosphorylation [23,24] have been reported. Furthermore, N-truncated Aβ variants may act as initiators of amyloid deposition [25]. Analysis of amyloid core preparations revealed the presence of Aβ peptides beginning with every residue between aspartic acid (Asp)-1 and glutamate (Glu)-11, but
with major signals for peptides starting with phenylalanine (Phe)-4, serine (Ser)-8 and Glu-11 [26]. Aβ4-42 has been detected by surface-enhanced laser desorption/ionization TOF (SELDI-TOF) mass spectrometry with strong signals in post mortem brain samples from aged controls, patients with vascular dementia and AD patients [27]. Among the different N-truncated Aβ forms, Aβ4-42 was found to be the major N-truncated Aβ species in parenchymal Aβ plaques [28]. In contrast, cerebrovascular amyloid was reported by Miller et al. to contain mainly Aβ peptides starting with residues 1 or 2 [26]. However, more recently also Aβ4-40 or Aβ4-42 peptides have been described in cases with congophilic amyloid angiopathy (CAA) or AD + CAA [16]. The conformational/aggregation state of Aβ peptides is an additional aspect to take into consideration when discussing the validity of Aβ as a therapeutic target. Besides the classical ‘Amyloid Hypothesis’ that sees amyloid deposition as a critical pathogenic event in AD [2], the role of soluble Aβ oligomers as key actors in synaptic dysfunction, neuron loss and memory deficits has grown in favour. Today, it is generally accepted that the apparently pathologically inert amyloid fibrils, which are found in plaques, originate from a nearly irreversible aggregation of monomeric Aβ peptides through toxic protofibrillar intermediates [29]. The function of amyloid plaques in this scenario is still a matter of controversial debates, whether they act as a reservoir of toxic amyloid oligomers or possess buffering properties [30]. While several preclinical immunotherapy studies employing anti-Aβ antibodies have demonstrated beneficial effects with regard to reduced extracellular plaque pathology [31,32], others also reported that passive immunization might cause severe vascular alterations such as increased frequency of haemorrhages [33,34]. Considering the controversial outcome of the immunotherapies in AD so far, a better understanding of the spatial localization or distribution of specific Aβ variants (e.g. N-terminal truncated or C-terminal elongated), and ultimately their role in the aetiology of the disease, is of high importance. Moreover, given that different Aβ conformation and/or aggregation states might have different roles in the pathology, the affinity of antibodies used in immunotherapy to bind to the different Aβ states is an additional crucial characteristic to take into consideration. In this study, a variety of antibodies detecting N-terminal Aβ variants were characterized in detail and used to assess the N-terminal heterogeneity of extracellular and vascular Aβ peptide deposits in human brain samples.

Materials and methods

Antibody characterization by capillary isoelectric focusing immunoassay

In order to evaluate the selectivity of a set of different antibodies detecting either full-length or N-terminal truncated variants of Aβ ending at position 40 as well as 42, a capillary isoelectric focusing immunoassay (CIEF-immunoassay) was employed as published previously [35,36]. In brief, synthetic Aβ peptides with different N-termini were separated on a Peggy Sue device (Protein Simple, San Jose, CA, USA) by isoelectric focusing in microcapillaries. Next, the detection was carried out using mouse monoclonal antibodies 80C2 (#218231, Synaptic Systems, Goettingen, Germany, 20 μg/ml) and 82E1 (IBL International, Hamburg, Germany, 20 μg/ml), both detecting full-length Aβ starting with Asp-1, antibody 18H6 (1.9 μg/ml) [20] detecting truncated Aβ starting with Phe-4 and the generic Aβ antibody 4G8 (BioLegend, San Diego, CA, USA, 2 μg/ml). The rabbit
polyclonal generic antibody 24311 [37] and the humanized murine monoclonal biosimilar antibodies of Bapineuzumab [38] (14 μg/ml) and Crenezumab [38] (17 μg/ml), which have been used in clinical trials, were also used in the assay. The synthetic Aβ peptides Aβ1–40/42, Aβ2–40/42, Aβ3–40/42, pyroglutamate AβpE3–40/42, Aβ4–40/42, Aβ5–40 and Aβ11–40 were purchased from AnaSpec (Fremont, CA, USA). The synthesis of Aβ3–40/42 (corresponding to APP669–711/713) has been described previously [39] (generous gift of Dr H.-J. Knölker, Dresden). Peptide stock solutions (1 mg/ml) were prepared in DMSO or, in the case of AβpE3–40, in 0.1% NH₃(aq) (~0.1% NH₃(w/v)) and stored at ~80°C. Working aliquots were thawed only once. Following further dilution in 20 mM bicine pH 7.6, 0.6% 3-[(3-cholamidopropyl) dimethylammonio]-1-propanesulphonate (CHAPS), peptides were mixed in a 1:4 ratio with G2 Premix containing a pH5–pH8 nested gradient, as well as fluorescent pH standards and a DMSO inhibitor mix (all reagents were obtained from ProteinSimple).

The automated CIEF-immunoassay was carried out on the Peggy Sue platform (ProteinSimple) as described previously for the NanoPro platform (ProteinSimple, San Jose, CA, USA) [35,36]. Each microcapillary contained a final peptide concentration of 100 ng/ml for peptides ending at position 40 and 50 ng/ml for peptides ending at position 42, corresponding to ~50 and 25 pg peptide per capillary, respectively, based on an internal capillary volume of roughly 0.5 μl. Chemiluminescence detection was carried out using peroxidase-labelled anti mouse secondary antibodies (with 4G8, 80C2 and 82E1) or biotinylated anti-rabbit, anti-mouse or antihuman IgG plus peroxidase-conjugated streptavidin (in case of 24311, 18H6 and the humanized biosimilar antibodies Bapineuzumab and Crenezumab).

**Western blot analysis under denaturing conditions**

The sensitivity of the antibody 82E1 and the biosimilar Bapineuzumab antibody was further analysed by urea-bicine/bis-tris/Tris/sulphate sodium dodecyl sulphate (SDS)-PAGE followed by western immunoblotting. Stock solutions of synthetic Aβ1–40 were prepared in sample buffer (0.36 M bis-tris, 0.16 M bicine, 15% wt/vol sucrose, 1% wt/vol SDS and 0.0075% wt/vol bromophenol blue) and stored at ~80°C. Different amounts of Aβ1–40 peptides (25–100 pg and 25–100 ng respectively) were loaded and separated by urea-bicine/bis-tris/Tris/sulphate SDS-PAGE and analysed by immunoblotting as previously described [40]. The blotting membranes were blocked in 2% enhanced chemiluminescence (ECL) advance blocking agent (GE Healthcare Life Sciences, Little Chalfont, UK) in phosphate-buffered saline (PBS) with 0.075% Tween 20 (PBS-T) for 1h at room temperature and probed with the primary antibody 82E1 or the biosimilar Bapineuzumab antibody (0.5 μg/μl in PBS-T) at 4°C overnight. After three washing steps with PBS-T, biotinylated goat anti-mouse IgG (1:3000 in PBS-T; Life Technologies, Carlsbad, CA, USA) for 82E1 and biotinylated goat anti-human IgG (DAKO, Glostrup, Denmark, 1:3000) for Bapineuzumab, followed by streptavidin-HRP, were employed as secondary reagents. Chemiluminescent signals obtained with ECL Prime Western Blotting Detection Reagent (GE Healthcare Life Sciences) were recorded.

**Human brain samples**

Paraffin-embedded human brain samples from sporadic AD (n = 20; mean age 84.45 ± 6.45, average post mortem interval (PMI) = 4.37 ± 0.56 h; 14 females, 6 males), nondemented...
controls (NDC; n = 11; mean age 84.18 ± 7.17, average PMI = 8:11 ± 5:54 h; 7 females, 4 males) and Down syndrome (DS; n = 2; mean age 61.0 ± 4.24, average PMI = 5:12 ± 1:21 h; 1 female, 1 male) were obtained from the Netherlands Brain Bank (Table 1). Samples exclusively from the medial frontal gyrus region of the brain were used in the analysis. This study was approved by the ethical committee of the University Medical Center Göttingen.

**Immunohistochemical staining and semi-quantitative analyses**

Immunohistochemistry was performed on 4 μm paraffin sections as previously described [41]. In brief, following deparaffinization in xylene and rehydration in a descending series of ethanol (100%, 95%, 70%), endogenous peroxidases were blocked by incubation in 0.3% H2O2 in PBS for 30 min. Sections were boiled in 0.01 M citrate buffer and incubated in 88% formic acid for antigen retrieval. Prior to the incubation with the primary antibodies, nonspecific-binding sites were blocked by treatment with 4% skim milk in PBS containing 10% foetal calf serum for 1 h at room temperature. The following antibodies were used for the detection of different Aβ variants: 80C2 (mouse mAb, Synaptic Systems, 2 μg/ml) and 82E1 (mouse mAb, IBL International, Hamburg, Germany, 1 μg/ml) for the detection of Aβ1–x; D3E10 (rabbit mAb, Cell Signaling Technology, Danvers, MA, USA, 1:1000) against Aβ42; 24311 (rabbit pAb [37], 1:500) against the N-terminus of Aβ; 18H6 (mouse mAb, 1.9 μg/ml [20]) and 029–2 (guinea-pig pAb [36], 2.36 μg/ml) for the detection of Aβ starting with Phe at position 4; and pAb77 (rabbit pAb [40], 2.32 μg/ml) against Aβ2–x. Lastly, the humanized biosimilar Bapineuzumab ([38,42], 0.7 μg/ml) and Crenezumab ([38,42]; 0.85 μg/ml) antibodies were used in this study. Biotinylated secondary anti-guinea pig (Dianova, Hamburg, Germany), anti-mouse, anti-rabbit and anti-human antibodies (all DAKO) were used in a 1:200 dilution. Staining was visualized using the ABC method, with a Vectastain kit (Vector Laboratories, Burlingame, CA, USA) and diaminobenzidine as chromogen. Counterstaining was carried out with haematoxylin. In order to perform a semi-quantitative analysis of the staining, the sections were evaluated in a blinded fashion using a BX51 microscope (Olympus, Hamburg, Germany) equipped with a Moticam Pro 282 camera (Motic, Wetzlar, Germany). The following semi-quantitative scoring criteria were used to assess parenchymal and vascular staining: 0 = no staining, 0.5 = barely detectable staining, 1 = weak staining, 2 = moderate staining, 3 = extensive staining. Heat maps were plotted using GraphPad Prism 8.3.0 (GraphPad, San Diego, CA, USA).

**Results**

**Determination of antibody selectivity using CIEF-immunoassays**

In the present report, a comparative analysis of the immunohistochemical staining profile of antibodies detecting either full-length or N-terminally truncated Aβ species was carried out. First, we studied the selectivity of the different antibodies using the CIEF immunoassay employing a variety of synthetic N-terminal truncated Aβ peptide variants. Antibody 4G8, directed against a central epitope (17–24) which is present in all peptides utilized in the current analysis, was used a control antibody. As expected, 4G8 detected the entire set of Aβ peptides (Aβ3–40, Aβ1–40, Aβ2–40, Aβ3–40, AβpE3–40, Aβ4–40, Aβ5–40, Aβ11–40) (Figure S1). Next, antibodies presumed to detect selectively either full-length Aβ species starting with Asp-1 or N-terminally truncated species, such as Aβ4–x starting with a phenylalanine...

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April 28, 2021 (GraphPad, San Diego, CA, USA).
residue at position 4, were analysed. The mouse monoclonal antibodies 82E1 and 80C2 detected the N-terminus of A\(\beta\) and produced a clear signal for A\(\beta\)\(_{1-40}\) without appreciable cross-reactivity with either the N-terminally elongated peptide variant A\(\beta\)\(_{-3-40}\) or the N-terminally truncated species such as A\(\beta\)\(_{2-40}\), A\(\beta\)\(_{3-40}\) or A\(\beta\)\(_{4-40}\) under the tested conditions. Thus, it appears that these two particular antibodies require a free N-terminal aspartic acid residue in position 1 of the A\(\beta\) sequence (Asp-1) (Figure 1). In contrast, the mouse monoclonal antibody 18H6 showed a clear signal only with synthetic A\(\beta\) peptides starting with the Phe residue (A\(\beta\)\(_{4-x}\)), without any noticeably cross-reactivity with longer or shorter A\(\beta\) peptides tested, thereby confirming its high selectivity for A\(\beta\)\(_{4-x}\) variants (Figure 1). The polyclonal antibody 24311 showed a profile comparable to 4G8, except for the fact that it did not detect elongated A\(\beta\)\(_{-3-40}\) peptides and only weakly reacted with full-length A\(\beta\)\(_{1-40}\) (Figure 1). Finally, the N-terminal selectivity of the recombinant biosimilar human antibodies Bapineuzumab and Crenezumab was determined. While Bapineuzumab only detected the full-length A\(\beta\)\(_{1-40}\) peptides without showing signals for either elongated (A\(\beta\)\(_{-3-40}\)) or any of the N-terminally truncated A\(\beta\) variants (Figure 1), Crenezumab detected the entire range of A\(\beta\) peptides employed (Figure 1), resembling the peptide pattern detected by the pan-specific control antibody 4G8 (Figures S1 and S2). As A\(\beta\) species ending at 42 could have a very distinct conformation that might eventually alter immunoreactivity towards epitopes even located at the N-terminus, we additionally tested the detection of selected A\(\beta\)\(_{42}\) variants with antibodies 4G8, 80C2, 18H6 and the biosimilar antibodies Bapineuzumab and Crenezumab. In qualitative terms, the tested antibodies showed the same detection profile as with the A\(\beta\)\(_{40}\) variants (Figures S2 and S3).

**Comparative A\(\beta\) staining profiles in human sporadic AD, NDC and DS cases**

Most of the sporadic AD cases, as well as the two DS samples, demonstrated high abundance of both extracellular amyloid plaques and vascular deposits detected by the pan-A\(\beta\) antibody 24311 (Figures 2A and 3A), as well as the A\(\beta\)\(_{42}\)-selective antibody D3E10 (Figures 2B and 5). In contrast, antibodies 82E1 (Figure 2C) and 80C2 (Figures 2D and 4B), selectively recognizing Asp-1, showed a much weaker staining of extracellular plaques, but intensely stained cerebrovascular amyloid deposits (Figure 3B,C). In general, 82E1 detected a more widespread pattern of extracellular deposits, while the staining profile of both antibodies largely overlapped with regard to parenchymal or meningeal vascular amyloid accumulation. The polyclonal antibody Ab77, directed against A\(\beta\)\(_{2-x}\) starting with an alanine residue, detected only few plaques in most of the AD cases (Figures 2E and 5). In contrast, A\(\beta\)\(_{2-x}\)-positive plaques were found in substantial amounts in both DS cases, as well as in meningeal and parenchymal vessels in the majority of AD cases (Figure 5).

Both antibodies capable of recognizing the free Phe residue at position 4 (029–2 and 18H6) showed abundant amyloid plaque immunoreactivity in the brain parenchyma of individuals suffering from sporadic AD (Figure 2F) and detected vascular deposits in approximately half of the AD cases (Figure 5). Finally, the two biosimilar recombinant antibodies Crenezumab and Bapineuzumab were analysed. While Crenezumab demonstrated considerable immunoreactivity and showed a weak to moderate staining pattern in the majority of sporadic AD patients analysed (Figures 2G and 5), Bapineuzumab detected extracellular plaques in all AD and DS cases with mainly moderate to high intensity (Figures 2H and 5).
Both antibodies showed a largely concordant staining profile with regard to parenchymal and meningeal vascular deposits in AD cases with intense immunoreactivity in the DS cases (Figure 5; Figure S4).

In the control cases considered here, the overall immunoreactivity was predominantly negative, with four out of 11 cases displaying a weak to moderate extracellular amyloid plaques staining with the pan-A\(\beta\) antibody 24311 and A\(\beta\)\(_{42}\) antibody D3E10. The control cases were largely negative with the antibodies recognizing N-terminal truncated A\(\beta\) variants and parenchymal and meningeal vascular deposits were absent with all the analysed antibodies (Figure 5; Figures S5 and S6).

A comparative analysis of 24311 (pan-Ab), 80C2 (A\(\beta\)\(_{1–x}\)), 18H6 (A\(\beta\)\(_{4–x}\)) and Bapineuzumab underscored the above-mentioned differences in immunoreactivity with regard to full-length A\(\beta\) peptides. While 24311, 18H6 and Bapineuzumab detected the identical extracellular A\(\beta\) deposit in parallel sections with comparable sensitivity; the signal generated by 80C2 was strongly reduced (Figure 4, asterisk). Interestingly, all four antibodies showed a comparable staining intensity of parenchymal vessels (Figure 4, arrowhead), corroborating the quantification of parenchymal CAA (Figure 5).

**Evidence of a conformational preference of the biosimilar antibody Bapineuzumab**

In the immunohistochemical analysis, the biosimilar antibody Bapineuzumab detected substantially more extracellular amyloid deposits than the other A\(\beta\)\(_{1–x}\)-specific antibodies 80C2 and 82E1. Thus, it appears that under the experimental conditions employed (i.e. formalin fixation and pretreatment with formic acid for sensitization), Bapineuzumab is more sensitive than the other A\(\beta\)\(_{1–x}\)-specific antibodies tested. To assess whether in addition to the amino acid sequence (primary structure), the conformation of the A\(\beta\) peptides may impact the strength of the recognition by the different antibodies, we compared Bapineuzumab and 82E1 by urea-bicine/bis-tris/Tris/sulphate SDS-PAGE followed by Western blotting. Different amounts of A\(\beta\)\(_{1–40}\) peptides (25–100 pg and 25–100 ng respectively) were separated on SDS gels containing 8 M urea, blotted onto PVDF membranes and probed with either Bapineuzumab or 82E1 antibodies. While no appreciable signals were observed with amounts up to 100 pg A\(\beta\)\(_{1–40}\) with Bapineuzumab (Figure 6A), even minute amounts as low as 25 pg of A\(\beta\)\(_{1–40}\) were clearly detected with 82E1 (Figure 6B). The comparatively much higher detection sensitivity of 82E1 in the urea Western blot assays suggests a strong impact of the A\(\beta\) peptide conformation on its binding by Bapineuzumab. While in immunohistochemistry (i.e. without preanalytical A\(\beta\) denaturation with SDS and Urea), Bapineuzumab displayed very high detection sensitivity, it appeared to be substantially less sensitive on Urea- SDS-PAGE-Western blots.

**Discussion**

The presence of A\(\beta\)\(_{40}\) and A\(\beta\)\(_{42}\) peptides has been extensively evaluated in human brain. Due to the commercial availability of A\(\beta\) end-specific antibodies, a multitude of neuropathological and biochemical studies dealing with the role of these peptides during ageing and the course of AD have been published in recent years. In addition to full-length A\(\beta\) starting with Asp-1 (e.g. A\(\beta\)\(_{1–40}\), A\(\beta\)\(_{1–42}\)), a variety of N- and C-terminally truncated A\(\beta\)
peptides can be detected in human brain samples [14,15,43–45]. Mass spectrometry allows for the unequivocal identification of Aβ variants present in brain amyloid but provides only limited information in terms of spatial localization or distribution of specific Aβ species. On the other hand, the use of Aβ antibodies in applications such as immunohistochemistry is largely dependent on the careful characterization of antibody selectivity and sensitivity. We therefore set out to determine the specificity of a variety of commonly used Aβ antibodies using CIEFs, by employing a wide range of N-terminally elongated (Aβ−3–40), full-length (Aβ1–40), as well as N-terminally truncated (Aβ2–40, Aβ3–40, Aβ4E3–40, Aβ4–40, Aβ5–40, Aβ11–40) Aβ peptide variants. The widely used 4G8 antibody, recognizing a central epitope within the Aβ sequence, detected all Aβ species employed as anticipated and served as a control antibody. We further confirmed the high selectivity of 80C2 [46] and 18H6 [20], for Aβ1–x and Aβ4–x respectively.

The immunohistochemical analysis employing this entire set of antibodies against full-length, N-truncated, Aβ C-terminus (Aβ42) or total Aβ revealed the expected heterogeneity. While both 80C2 and 82E1 (detecting Asp-1) on the one hand, or 029–2 [36] and 18H6 (detecting Phe-4) on the other showed an exactly concordant profile in terms of selectivity in the CIEF assays, the staining patterns with regard to extracellular amyloid plaque as well as vascular deposition revealed some distinctions. 82E1 showed a more widespread pattern of extracellular deposits compared to 80C2, disclosing a pattern associable to a classical neuritic plaque staining. While 029–2 and 18H6 intensely stained amyloid plaque cores, the latter appeared to present a more abundant pattern in the cases employed in this study. This might be attributed to differences in antibody detection sensitivities, as well as an additional impact of Aβ peptide conformation beyond sequence specificity. As expected, sporadic AD cases showed a more intense staining pattern than the employed NDC cases in general. The substantial immunoreactivity of NDC case 2 with several antibodies can be attributed to the presence of medium densities of amyloid deposits (amyloid stage B, see Table 1).

The PMI is a variable parameter in neuropathological analyses that might influence protein and nucleic acid integrity. In this study, the average PMI of the AD samples (<5 h) and control samples (<9 h) were relatively short, which might have only a limited impact on protein denaturation. It has been shown by others that a PMI of over 50 h did not change the immune staining profiles of several proteins [47], and no correlation was found between PMI and the relative abundance of most of the N-truncated variants analysed here [18].

Despite several setbacks [7–13], passive immunization with anti-amyloid monoclonal antibodies is still debated as a potential disease-modifying therapy, and clinical trials are ongoing [48]. We analysed the biosimilar Bapineuzumab and Crenezumab antibodies, which both had been evaluated in clinical trials and did not meet their primary endpoints [8,49,50]. While Crenezumab, similar to the pan-specific anti Aβ monoclonal 4G8, detected the whole set of Aβ variants tested, Bapineuzumab selectively detected Aβ1–40 starting with Asp and did not detect N-terminally elongated or truncated Aβ species. Crenezumab has been described as a fully humanized immunoglobulin isotype G4 (IgG4) monoclonal antibody that binds to Aβ oligomers with high affinity, while also maintaining the ability to bind to other forms of Aβ [51]. Atomic structures show that Crenezumab and the clinical antibody
Solanezumab bind Aβ peptides in a virtually identical conformation of consecutive residues in the central epitope (~Aβ13–24) [52,53], containing the Aβ aggregation nucleation site KLVFFA. This is consistent with our results from the CIEF assay, where Crenezumab behaved in the same way as the widely used 4G8 antibody (epitope Aβ17–21/23) [54,55] and the findings of Watt and colleagues in ex vivo SELDI-TOF experiments in AD-affected tissue [36].

The observed selectivity of Bapineuzumab for the free N-terminal Asp residue confirms and extends earlier studies. Surface plasmon resonance binding response curves resulted in essentially identical $k_d$ values of Bapineuzumab for different peptides starting with Asp-1, such as Aβ1–28, Aβ1–40 and Aβ1–42. No binding was detected between the antibody and Aβ species with an altered N-terminus [38]. Analysis of the crystal structure of the humanized antigen-binding fragment in complex with Aβ1–28 revealed that the free N-terminus was critical for Aβ binding. This structure analysis suggests that Bapineuzumab recognizes the N-terminal end of Aβ in a helical conformation that is stabilized by five putative intramolecular hydrogen bonds [53]. The experiments performed in this study support these assumptions, showing in addition that neither elongated (Aβ1–40) or peptides starting with an N-terminal alanine residue (Aβ2–40) are recognized. With regard to the immunohistochemical staining of parenchymal plaques, Bapineuzumab seems to stain a higher proportion of extracellular deposits than 80C2 or 82E1, which are also directed against the free N-terminal Asp residue present in full-length Aβ peptides. Under the strongly denaturing conditions employed in the Urea-SDS PAGE/Western blot analysis, the opposite was observed with 82E1 detecting Aβ1–40 peptides at ~1000-fold lower concentrations than Bapineuzumab. We conclude that, the helical conformation of the Aβ-N-terminus recognized by Bapineuzumab [53] is probably essentially destroyed by heating the samples in the presence of SDS prior to electrophoresis, but not by formalin fixation and formic acid treatment used in immunohistochemistry. Thus, Bapineuzumab, but not 82E1, appears to be particularly sensitive to specific conformational properties of Aβ.

The conformational preference of antibodies may play a critical role both at a clinical level, when employed in, for example immunotherapy, as well as when they are utilized as research tools in preclinical settings. This adds to the complexity determined by the heterogeneous composition of plaques and the differences in spatial localization or distribution of specific Aβ species and impedes approaches to unravel disease mechanisms and find potentially suitable targets in AD.

**Supplementary Material**

Refer to Web version on PubMed Central for supplementary material.

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Figure 1.
Assessment of antibody selectivity by capillary isoelectric focusing immunoassay. Synthetic amyloid-β (Aβ) peptides with different N-termini were separated by isoelectric focusing in microcapillaries, immobilized by a photochemical reaction and probed with the indicated primary antibodies. Chemiluminescence detection was achieved with peroxidase-labelled anti mouse antibodies (80C2, 82E1). In case of 24311, 18H6 and the humanized biosimilar antibodies Bapineuzumab and Crenezumab, biotinylated anti-rabbit, anti-mouse or anti-human IgG plus peroxidase-conjugated streptavidin were employed. For each one of the indicated primary antibodies, a mixture of Aβ1–40, Aβ2–40 and Aβ5–40 (electropherogram A), a mixture of AβpE3–40, Aβ4–40 and Aβ11–40 (B), Aβ3–40 (C) and N-terminally elongated Aβ3–40 (APP669–711) (D) were assessed. To facilitate the evaluation of the selectivity for specific N-terminal Aβ variants, the electropherograms were scaled for each primary antibody according to the maximum signal that was observed. Note that the signals obtained with the humanized biosimilar antibodies Bapineuzumab and Crenezumab were substantially higher than with the mouse monoclonals. Presumably, in these two cases the combination of a biotinylated anti human antibody and peroxidase-conjugated streptavidin provided highly efficient signal amplification.
Figure 2.
Immunohistochemical staining of extracellular amyloid-β (Aβ) peptides in a sporadic Alzheimer’s disease (AD) case (AD case 20). Parallel sections of the frontal cortex of a sporadic AD case were stained with the pan-Aβ antibody 24311 (A), the Aβ42-specific antibody D3E10 (B), the Aβ1–x-specific antibodies 82E1 (C) and 80C2 (D), the Aβ2–x-specific antibody Aβ77 (E), the Aβ3–x-specific antibody 18H6 (F), as well as the biosimilar human antibodies Crenezumab (G) and Bapineuzumab (H). Scale bar: A–H = 100 μm.
Figure 3. Aβ-immunoreactivity of parenchymal vessels in a sporadic Alzheimer’s disease (AD) case (AD case 5). Parallel sections of the frontal cortex of a sporadic AD case were stained with the pan-Aβ antibody 24311 (A), the Aβ1–42-specific antibodies 82E1 (B) and 80C2 (C), as well as the biosimilar human antibody Bapineuzumab (D). Scale bar: A–D = 100 μm.
Figure 4.
High magnification images of 24311 (A), 80C2 (B), 18H6 (C) and Bapineuzumab (D) antibodies in adjacent sections (Alzheimer's disease case 18). While immunoreactivity of all antibodies is comparable in parenchymal vessels (arrowhead), 80C2 shows considerably less staining in parenchymal extracellular deposits compared to the other antibodies (*). Scale bar: A–D = 33 µm.
Figure 5.
Semiquantitative analyses of the immunostaining. Heat maps illustrating the results of the immunostaining of parenchymal plaques (A), parenchymal (B) and meningeal CAA (C). AD, sporadic Alzheimer’s disease; NDC, nondemented control; DS, Downs’ syndrome; CAA, cerebral amyloid angiopathy.
Figure 6.
Differential sensitivity of N-terminal-specific antibodies Bapineuzumab and 82E1 under denaturing conditions. Under denaturing conditions in the SDS-PAGE, Bapineuzumab (A) shows a roughly 1000-fold lower sensitivity compared to 82E1 (B), which is capable of detecting Aβ1–40 peptides in amounts as low as 25 pg.
| Case     | Age (years) | Gender | ApoE | Braak stage | Amyloid | PMI (h) | CAA   |
|----------|-------------|--------|------|-------------|---------|---------|-------|
| Sporadic AD cases | | | | | | | |
| AD 1     | 86          | f      | 3/3  | IV          | B       | 04:10   |        |
| AD 2     | 90          | f      | 4/4  | VI          | C       | 04:00   | Mild  |
| AD 3     | 85          | f      | 3/3  | VI          | C       | 03:30   | Mild  |
| AD 4     | 88          | f      | 3/3  | IV          | C       | 06:15   | Distant |
| AD 5     | 87          | f      | 4/3  | VI          | C       | 05:00   | Distant |
| AD 6     | 92          | m      | 3/3  | IV          | C       | 05:30   | Distant |
| AD 7     | 91          | m      | 3/3  | IV          | C       | 03:45   | Distant |
| AD 8     | 75          | f      | 4/4  | IV          | C       | 06:15   | Distant |
| AD 9     | 93          | f      | 3/3  | IV          | C       | 06:30   | Distant |
| AD 10    | 74          | m      | 4/3  | V           | C       | 06:00   | Distant |
| AD 11    | 78          | f      | 3/3  | V           | C       | 03:25   | Distant |
| AD 12    | 82          | f      | 4/4  | V           | C       | 04:05   | Distant |
| AD 13    | 77          | m      | 3/3  | V           | C       | 04:50   | Distant |
| AD 14    | 89          | f      | 4/3  | V           | C       | 03:05   | Distant |
| AD 15    | 81          | m      | 3/3  | V           | C       | 04:00   | Distant |
| AD 16    | 79          | m      | 3/3  | V           | C       | 03:30   | Distant |
| AD 17    | 79          | m      | 3/3  | V           | C       | 06:15   | Distant |
| AD 18    | 70          | f      | 3/3  | V           | C       | 04:15   | Distant |
| AD 19    | 70          | f      | 4/3  | V           | C       | 04:50   | Distant |
| AD 20    | 83          | s      | 3/3  | V           | C       | 06:30   | Distant |
| AD 21    | 91          | m      | 3/3  | V           | C       | 06:00   | Distant |
| AD 22    | 72          | f      | 3/3  | V           | C       | 03:05   | Distant |
| AD 23    | 89          | f      | 4/3  | V           | C       | 04:05   | Distant |
| AD 24    | 78          | f      | 3/3  | V           | C       | 03:05   | Distant |
| AD 25    | 70          | m      | 3/3  | V           | C       | 03:52   | Distant |
| AD 26    | 84          | m      | 3/3  | V           | C       | 06:00   | Distant |
| AD 27    | 70          | m      | 3/3  | V           | C       | 07:30   | Distant |
| AD 28    | 84          | m      | 3/3  | V           | C       | 09:00   | Distant |

| NDC 1    | 82          | f      | 3/3  | I           | A       | 11:00   | Distant |
| NDC 2    | 82          | f      | 3/3  | I           | A       | 04:45   | Distant |
| NDC 3    | 78          | f      | 3/3  | I           | A       | 06:00   | Distant |
| NDC 4    | 78          | f      | 3/3  | I           | A       | 04:05   | Distant |
| NDC 5    | 73          | m      | 3/3  | I           | A       | 03:52   | Distant |
| NDC 6    | 70          | m      | 3/3  | I           | A       | 06:00   | Distant |
| NDC 7    | 84          | m      | 3/3  | I           | A       | 09:00   | Distant |

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| Case  | Age (years) | Gender | ApoE | Braak stage | Amyloid | PMI (h) | CAA  |
|-------|-------------|--------|------|-------------|---------|---------|------|
| NDC 8 | 87          | f      | n.d. | II          | 0       | 05:30   | No   |
| NDC 9 | 93          | f      | 3/3  | II          | 0       | 05:50   | No   |
| NDC 10| 87          | m      | 3/3  | III         | A       | 06:10   | No   |
| NDC 11| 90          | f      | 3/3  | III         | A       | 06:05   | No   |
| DS 1  | 64          | f      | 3/3  | V           | C       | 04:15   | Distinct |
| DS 2  | 58          | m      | 4/3  | VI          | n.d.    | 06:10   | Distinct |

ApoE, apolipoprotein E; AD, Alzheimer’s disease; NDC, nondemented control subject; DS, Down syndrome; PMI, post mortem interval; n.d., not determined; CAA, cerebral amyloid angiopathy.