Establishment and Validation of a Novel MRI Radiomics Feature-Based Prognostic Model to Predict Distant Metastasis in Endemic Nasopharyngeal Carcinoma

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Purpose: We aimed to establish a prognostic model based on magnetic resonance imaging (MRI) radiomics features for individual distant metastasis risk prediction in patients with nasopharyngeal carcinoma (NPC).

Methods: Regression analysis was applied to select radiomics features from T1-weighted (T1-w), contrast-enhanced T1-weighted (T1C-w), and T2-weighted (T2-w) MRI scans. All prognostic models were established using a primary cohort of 518 patients with NPC. The prognostic ability of the radiomics, clinical (based on clinical factors), and merged prognostic models (integrating clinical factors with radiomics) were identified using a concordance index (C-index). Models were tested using a validation cohort of 260 NPC patients. Distant metastasis-free survival (DMFS) were calculated by using the Kaplan-Meier method and compared by using the log-rank test.

Results: In the primary cohort, seven radiomics prognostic models showed similar discrimination ability for DMFS to the clinical prognostic model (P=0.070-0.708), while seven merged prognostic models displayed better discrimination ability than the clinical prognostic model or corresponding radiomics prognostic models (all P<0.001). In the validation cohort, the C-indices of seven radiomics prognostic models (0.645-0.722) for DMFS prediction were higher than in the clinical prognostic model (0.552) (P=0.016 or <0.001) or in corresponding merged prognostic models (0.605-0.678) (P=0.297 to 0.857), with T1+T1C prognostic model (based on Radscore combinations of T1 and T1C Radiomics models) showing the highest C-index (0.722). In the decision curve analysis of
Introduction

Nasopharyngeal carcinoma (NPC) is endemic to southern China, southeastern Asia, and northern Africa, where the peak incidence rate is 20–50 cases per 100,000 individuals (1, 2). Advances in NPC diagnosis and treatment using MRI, intensity-modulated radiotherapy, and combined chemotherapy, has significantly improved locoregional control, and distant metastasis is regarded as the main cause of treatment failure (3–5). In a meta-analysis that reviewed 13304 participants, intensity-modulated radiotherapy was associated with better 5-year locoregional control (odds ratio 2.08; 95% confidence interval [CI]=1.82–2.37), while there were no significant differences seen in distant metastasis-free survival (DMFS) compared with conventional radiotherapy (4). Although the current 5-year overall survival rate of NPC patients reach 80–88%, 15–25% of patients develop distant metastasis after treatment, especially in advanced NPC (5, 6), so to identify high-risk patients to guide optimal treatment decisions is vital.

Currently, the TNM staging system is widely used to predict prognosis and establish treatment strategies among patients with NPC. However, patients at the same clinical stage receiving similar treatments often have different outcomes (7). Among NPC patients with distant metastasis after treatment, nearly 80% are stage III or IV (8), when it is difficult to identify high-risk patients. The presence of plasma Epstein-Barr virus (EBV) DNA influences the disease process in NPC patients and is regarded as a potential biomarker with clinical applications (9). Other risk factors have been associated with prognosis in patients with NPC, including age, sex, smoking status, hepatitis B surface antigen, serum lactate dehydrogenase, high-sensitivity C-reactive protein, and microRNA or mRNA; however, results are inconsistent (10–15). Therefore, the prognostic value of these markers needs confirmation.

Radiomics focuses on extracting quantitative features from medical images to find a possible association with tumor phenotypic characteristics (16). Radiomic features were found to be associated with gene and/or protein expression profiles, treatment response, and clinical outcomes in different cancer types (16, 17). Pretreatment MRI radiomics or MRI-based radiomics nomograms were proposed to predict locoregional recurrence, distant metastasis, and early response to induction chemotherapy in endemic NPC (18–29). However, there is no effective radiomics-based model to distinguish different risks levels for distant metastasis among NPC patients.

We conducted a study of multiparametric MRI radiomics features to identify and validate an MRI-based radiomics model discriminating distant metastasis in patients with NPC. Moreover, we compared the accuracy of a radiomics prognostic model (based on selected radiomics features), a clinical prognostic model (based on clinical risk factors), and a prognostic model merging the two, for discriminating distant metastasis in NPC patients.

Methods

Participants

This retrospective study was approved by the Clinical Research Ethics Committee of the Sun Yat-Sen University Cancer Center (SYSUCC); pre-treatment written informed consent was obtained from all patients or their next of kin. The authenticity of the study was validated by uploading the fully raw data onto the Research Data Deposit (RDD) public platform (http://www.researchdata.org.cn), with the approval RDD number as RDDA2021135093. There were 903 patients newly diagnosed with untreated and non-metastatic NPC initially enrolled between January 2010 and November 2012; 125 patients were excluded from analysis (see Supplementary Materials). The remaining 778 NPC patients were randomly assigned in a 2:1 proportion to the primary (n=518) or validation cohort (n=260).

All patients underwent pretreatment evaluation, including clinical examinations of the head and neck region, fiber optic nasopharyngoscopy, neck and nasopharyngeal MRI, chest radiography, abdominal sonography, and whole-body bone scan, and staged according to the eighth American Joint Committee on Cancer TNM staging manual (30). Pretreatment blood samples were collected to determine blood type and the presence or levels of EBV DNA, hepatitis B surface antigen, lactate dehydrogenase, high-sensitivity C-reactive protein, platelets, and leucocytes. A quantitative polymerase chain reaction method was used for detecting plasma EBV DNA (31). Clinical patient characteristics are listed in Table 1.

All patients were treated according to the SYSUCC guidelines for NPC patients, which recommend that stage I patients have to receive radiotherapy alone, stage II radiotherapy alone, or concomitant chemoradiotherapy, and stage III–IVa concomitant chemoradiotherapy with or without induction chemotherapy. All patients received definitive intensity-modulated radiotherapy with 6-MV photons during the entire radiotherapy course.

Abbreviations: CI confidence interval; C-index concordance index; DMFS distant metastasis-free survival; EBV Epstein-Barr virus; NPC nasopharyngeal carcinoma; T1C-w contrast-enhanced; T1-weighted; T1-w T1-weighted; T2-w T2-weighted.
### TABLE 1 | Patient demographic characteristics in the primary and validation cohorts.

| Characteristic                  | Primary cohort (N=518) | Validation cohort (N=260) | P  |
|--------------------------------|------------------------|---------------------------|----|
| Age (years)                    | 44 (38–53)             | 46 (38–52)                | 0.944 |
| Gender                         |                        |                           | 0.389 |
| Male                           | 371 (71.6%)            | 194 (74.6%)               |     |
| Female                         | 147 (28.4%)            | 66 (25.4%)                |     |
| WHO pathologic classification  |                        |                           | 0.253 |
| Type I/II                      | 27 (5.2%)              | 19 (7.3%)                 |     |
| Type III                       | 491 (94.8%)            | 241 (92.7%)               |     |
| T classification               |                        |                           | 0.389 |
| T1                             | 140 (27%)              | 61 (23.5%)                |     |
| T2                             | 60 (11.6%)             | 37 (14.2%)                |     |
| T3                             | 186 (35.9%)            | 103 (39.6%)               |     |
| T4                             | 132 (25.5%)            | 59 (22.7%)                |     |
| N classification               |                        |                           | 0.425 |
| N0                             | 128 (24.7%)            | 53 (20.4%)                |     |
| N1                             | 282 (54.4%)            | 151 (58.1%)               |     |
| N2                             | 69 (13.3%)             | 40 (15.4%)                |     |
| N3                             | 39 (7.5%)              | 16 (6.2%)                 |     |
| AJCC clinical stage (2010)     |                        |                           | 0.181 |
| I                              | 53 (10.2%)             | 19 (7.3%)                 |     |
| II                             | 117 (22.6%)            | 57 (21.9%)                |     |
| III                            | 185 (35.7%)            | 112 (43.1%)               |     |
| IVa                            | 163 (31.5%)            | 72 (27.7%)                |     |
| Treatment regimen              |                        |                           | 0.490 |
| RT alone                       | 71 (13.7%)             | 34 (13.1%)                |     |
| CCRT                           | 201 (38.8%)            | 91 (35%)                  |     |
| IC + CCRT                      | 246 (47.5%)            | 135 (51.9%)               |     |
| EBV DNA (10^3 copies/ml)*      | 232 (44.8%)            | 125 (48.1%)               | 0.262 |
| <1                             | 117 (22.6%)            | 67 (25.8%)                |     |
| <10                            | 129 (24.9%)            | 49 (18.8%)                |     |
| ≥10                            | 40 (7.7%)              | 19 (7.3%)                 |     |
| Blood type                     |                        |                           | 0.145 |
| A                              | 139 (26.8%)            | 56 (21.5%)                |     |
| B                              | 130 (25.1%)            | 67 (25.8%)                |     |
| AB                             | 17 (3.3%)              | 16 (6.2%)                 |     |
| o                              | 232 (44.8%)            | 121 (46.5%)               |     |
| HBsAg                          | 427 (82.4%)            | 210 (80.8%)               | 0.620 |
| Negative                       | 91 (17.6%)             | 50 (19.2%)                |     |
| Positive                       |                         |                           |     |
| LDH (U/L)*                     | 492 (95%)              | 240 (92.3%)               | 0.142 |
| <245                           | 26 (5%)                | 20 (7.7%)                 |     |
| ≥245                           |                         |                           |     |
| hs-CRP (g/ml)*                 | 196 (37.8%)            | 101 (38.8%)               | 0.186 |
| <1                             | 166 (32%)              | 96 (36.9%)                |     |
| ≥3                             | 156 (30.1%)            | 63 (24.2%)                |     |
| Platelet counts (10^3/L)*      |                         |                           | 0.459 |
| <100                           | 4 (0.8%)               | 2 (0.8%)                  |     |
| 100–300                        | 436 (84.2%)            | 227 (87.3%)               |     |
| ≥300                           | 78 (15.1%)             | 31 (11.9%)                |     |
| Leukocyte counts (10^3/L)*     |                         |                           | 0.347 |
| <4                             | 9 (1.7%)               | 9 (3.5%)                  |     |
| ≥4–10                          | 58 (11.3%)             | 36 (13.6%)                |     |
| ≥10                            | 844 (95.5%)            | 591 (88.7%)               |     |
| Follow-up time (months)        |                         |                           | 0.886 |
| Median                         | 84.6 (3.3–104.1)       | 84.4 (6.9–103.9)          |     |

IQR, interquartile range; WHO, World Health Organization; Type I, keratinizing; Type II, non-keratinizing differentiated; Type III, non-keratinizing undifferentiated; T, tumor; N, node; AJCC, American Joint Committee on Cancer; RT, radiotherapy; CCRT, concurrent chemoradiotherapy; IC, induction chemotherapy; EBV DNA, Plasma Epstein-Barr virus DNA; HBsAg, hepatitis B surface antigen; LDH, serum lactate dehydrogenase levels; hs-CRP, high-sensitivity C-reactive protein; min, minimum; max, maximum. *Results before treatment.

Concurrent chemotherapy, induction chemotherapy, and intensity-modulated radiotherapy data are presented in the [Supplementary Material](#).

Complete follow-up data at 3, 5, and 8 years were available for 97.8%, 96.4%, and 87.6% of patients, respectively. Follow-up visits occurred at least once every 3 months during the first 3 years and then once every 6 months after treatment. DMFS was calculated from the date of treatment initiation to that of distant metastasis at any site, death from any cause, or the date of the last follow-up. Distant metastases were assessed using two imaging methods, including chest radiography, abdominal sonography, bone scan, and CT or MRI; elevation of plasma EBV DNA was confirmed by pathological biopsy if necessary.

### MRI Acquisition/Segmentation and Radiomics Feature Extraction

All patients underwent MRI with a 1.5-T system (Signa CV/i; General Electric Healthcare, Chalfont St. Giles, United Kingdom) or 3.0 T system (Siemens Magnetom Tim Trio, Erlangen, Germany), employing the fast spin-echo technique. The scan region imaged ranged from the suprasellar cistern to the inferior margin of the sphenoid wing of the clivus using a head-and-neck combined coil. T1-weighted images (T1-w) in the axial, coronal, and sagittal planes, T2-weighted (T2-w) images in the axial plane, and contrast-enhanced T1-weighted (T1C-w) in the axial, coronal, and sagittal planes were acquired in all patients. Detailed information on MRI scans is shown in [Supplementary Materials](#).

Axial T1-w, T2-w, and T1C-w Digital Imaging and Communications in Medicine (DICOM) images were retrieved from the picture archiving and communication system (PACS) and loaded into AnalyzePro ([https://analyzedirect.com/analyzePRO/](https://analyzedirect.com/analyzePRO/)) for semi-manual segmentation with normalization. One radiologist (Z. Guoyi) with over 10 years of experience in head and neck cancers outlined the tumor contour regions of interest (ROI) on each MRI slice, and each segmentation was validated by a senior radiologist (L. Lizhi) with 20 years of experience in evaluating MRI scans of patients with NPC. Differences were resolved by consensus. Extractions of radiomics features were performed using the open-source PyRadiomics ([http://www.radiomics.io/pyradiomics.html](http://www.radiomics.io/pyradiomics.html)). We extracted 4527 radiomics features for each patient from axial T1-w, T2-w, and T1C-w axial images (1509 features from each unimodal MR image). Data from the radiomics features are shown in the [Supplementary Material](#).

### Statistical Analysis Methods

The interclass correlation coefficient was used to assess the effect of variations in manual segmentation on radiomics feature values from 50 patients randomly extracted from the primary cohort. Features with high reproducibility (median ± standard deviation: interclass correlation coefficient >0.80) were retained for subsequent analysis. After deleting features that had a Pearson correlation coefficient ≥0.75, univariate analysis using the Cox regression model was employed as the first step of feature selection to assess the possible risk factor of DMFS. The least absolute shrinkage and selection operator method was the second feature-selection process for model building.
**Supplementary Figures S1, S2** show the processes of radiomics feature selection and radiomics feature selection using the least absolute shrinkage and selection operator regression model, respectively.

Kaplan-Meier survival curves and log-rank tests were used to evaluate the time-event data. Clinical characteristics between the primary and validation cohorts were compared using the chi-square test for categorical variables and the Mann-Whitney U test for continuous variables. Harrell’s concordance indices (C-indices) were applied to evaluate the discriminating ability of each prognostic model. The establishment flowchart of the clinical prognostic model, radiomics prognostic models, and merged prognostic models is shown in Supplementary Figure S3.

Statistical analyses were performed using R software version 3.2.5. with the following R packages: the caret package for Pearson correlation analysis; the survival package and survminer for Kaplan-Meier survival curves; the glmnet package for least absolute shrinkage and selection operator Cox regression; the ggplot2 package for score plot; the rms package for comparisons between models in terms of C-indices; the pheatmap and gplots packages for heatmaps. All statistical tests were two-tailed with \( P<0.05 \) indicating statistical significance.

**RESULTS**

**Participants**

Table 1 lists the clinical characteristics of patients with NPC. No significant differences were found in clinical characteristics between the primary and validation cohorts (|\( P = 0.142 \) to 0.944). During the follow-up period, 12.9% (67/518) and 13.1% (34/260) of patients developed distant metastases in the primary and validation cohorts, respectively. There was no significant difference between the two cohorts in the distant metastases rate (|\( P = 0.961 \)). The median DMFS time was 84.3 months (range, 3.3–104.1) for the primary cohort, and 84.4 months (range, 6.6–103.9) for the validation cohort.

**Clinical Prognostic Model Building**

Univariate analysis indicated that pre-treatment T stage, N stage, clinical stage, and plasma EBV DNA were associated with DMFS in patients with NPC (Supplementary Table S1). Multivariable analyses further identified that pre-treatment T stage, N stage, and plasma EBV DNA were independent predictors for DMFS (Supplementary Table S2). The clinical prognostic model for DMFS discriminating was based on these three independent factors. The C-index of this model for the primary cohort was 0.736 (95% CI=0.680–0.791), significantly higher than the C-indices for the validation cohort, 0.552 (95% CI=0.457–0.647) (Table 2).

**Radiomics Prognostic Model Building**

Supplementary Table S3 summarizes the features most strongly associated with DMFS based on analysis of the primary cohort.

The T1, T1c, and T2 prognostic models for DMFS prediction were based on selected radiomics features derived from T1-w, T1C-w, and T2-w MRI scans, respectively (Table 3). The T1+T1C, T1+T2, T2+T1C, and T1+T2+T1C prognostic models were based on Radiomics combinations of the T1 and T1C, T1 and T2, T2 and T1C, and T1, T2 and T1C prognostic models, respectively (Supplementary Table S4).

As shown in Table 2, in the primary cohort, seven radiomics prognostic models showed similar DMFS discriminating ability as that of the clinical prognostic model (\( P = 0.070 \) to 0.708). While in the validation cohort, the C-indices for DMFS prediction by the seven models (0.645–0.722) were significantly higher than the C-index calculated using the clinical prognostic model (\( P = 0.016 \) or <0.001). Intriguingly, in the seven radiomics models of the validation cohort, the T1+T1C prognostic model showed the highest C-index (0.722, 95% CI=0.632–0.811) for DMFS discriminating, significantly superior to the C-indices of the T1 (0.676), T2 (0.645), and T1C (0.711) prognostic models (\( P = 0.008, 0.043, \) and 0.048, respectively) and higher than those of the other three radiomics models, although without significant differences (Supplementary Table S4).

**Merged Prognostic Model Building**

To identify whether integrating clinical factors with radiomics prognostic models could increase predictive accuracy, we built seven merged prognostic models (MT1, MT2, MT1C, MT1+T2, MT1+T1C, MT2+T1C, and MT1+T2+T1C) based on combinations of three clinical factors and the T1, T2, T1C, T1+T2, T1+T1C, T2+T1C, and T1+T2+T1C radiomics models, respectively (Supplementary Figure S3). In the primary cohort, merged prognostic models displayed better discriminating ability in DMFS than the corresponding radiomics prognostic models (all \( P < 0.001 \)). While in the validation cohort, the C-indices of seven merged prognostic models (0.605–0.678) for DMFS discriminating were all lower than those of the corresponding radiomics prognostic models; although, there were no significant differences (Table 2), indicating that adding the clinical factors to the radiomics models weakened their discriminating ability.

To confirm this finding, we removed the T stage from the merged prognostic models and constructed seven additional merged prognostic models (rMT1, rMT2, rMT1C, rMT1+T2, rMT1+T1C, rMT2+T1C, and rMT1+T2+T1C) based on the radiomics prognostic model, N stage, and EBV-DNA. The C-indices of these seven models (0.612–0.685) were slightly better than those of the corresponding merged prognostic models in the validation cohort (Table 2).

**Performance of the Clinical and T1+1C Prognostic Model**

Based on the results above, T1+1C radiomics model was considered to provide the best prognostic ability for DMFS (having the highest C-index and Least degree of freedom). We further compared the discriminating ability between the clinical prognostic model and the T1+T1C prognostic model for DMFS from their nomograms, calibration curves, risk score distributions, and decision curve analysis.
TABLE 2 | C-index values of different prognostic models for DMFS prediction in the primary and validation cohorts.

| Prognostic model | Degree of freedom | Primary cohort | Validation cohort |
|------------------|-------------------|---------------|------------------|
|                  |                   | C-index (95% CI) | P†  | P*  | C-index (95% CI) | P†  | P*  |
| Clinical prognostic model | 9                | 0.736 (0.68 0.791) | 0.181 reference | 0.552 (0.457, 0.647) | Reference |
| Radiomics prognostic model | 7               | 0.723 (0.666, 0.780) | 0.181 reference | 0.676 (0.588, 0.764) | <0.001 reference |
| T1                | 5                | 0.715 (0.656, 0.774) | 0.222 reference | 0.645 (0.546, 0.744) | 0.016 reference |
| T1C               | 9                | 0.733 (0.676, 0.791) | 0.708 reference | 0.711 (0.615, 0.807) | <0.001 reference |
| T1+2              | 2                | 0.771 (0.720, 0.823) | 0.175 reference | 0.679 (0.594, 0.763) | <0.001 reference |
| T1+1C             | 2                | 0.757 (0.703, 0.81) | 0.468 reference | 0.722 (0.632, 0.811) | <0.001 reference |
| T2+1C             | 2                | 0.763 (0.712, 0.813) | 0.430 reference | 0.697 (0.599, 0.796) | <0.001 reference |
| T1+2+1C           | 3                | 0.784 (0.737, 0.831) | 0.070 reference | 0.711 (0.622, 0.799) | <0.001 reference |
| Merged prognostic model | 10              | 0.784 (0.731, 0.837) | <0.001 reference | 0.640 (0.558, 0.723) | 0.057 0.511 |
| MT1               | 10               | 0.773 (0.719, 0.826) | <0.001 reference | 0.605 (0.500, 0.710) | 0.055 0.647 |
| MT2               | 10               | 0.791 (0.741, 0.841) | <0.001 reference | 0.661 (0.563, 0.759) | 0.003 0.857 |
| MT1C              | 11               | 0.801 (0.752, 0.850) | <0.001 reference | 0.648 (0.560, 0.735) | 0.004 0.322 |
| MT1+2             | 11               | 0.812 (0.763, 0.860) | <0.001 reference | 0.678 (0.589, 0.767) | 0.001 0.391 |
| MT1+1C            | 11               | 0.806 (0.758, 0.854) | <0.001 reference | 0.653 (0.554, 0.753) | <0.001 0.600 |
| MT2+1C            | 11               | 0.818 (0.771, 0.865) | <0.001 reference | 0.677 (0.587, 0.767) | 0.001 0.297 |
| MT1+2+1C          | 12               | 0.780 (0.726, 0.834) | <0.001 reference | 0.653 (0.570, 0.736) | <0.001 0.843 |
| Remerged prognostic model | 7               | 0.773 (0.719, 0.827) | <0.001 reference | 0.612 (0.508, 0.716) | 0.046 0.964 |
| rMT1              | 7                | 0.799 (0.739, 0.859) | <0.001 reference | 0.671 (0.576, 0.766) | <0.001 0.893 |
| rMT2              | 7                | 0.797 (0.749, 0.848) | <0.001 reference | 0.661 (0.573, 0.748) | <0.001 0.480 |
| rMT1C             | 7                | 0.809 (0.761, 0.857) | <0.001 reference | 0.685 (0.597, 0.774) | <0.001 0.663 |
| rMT1+2            | 8                | 0.804 (0.757, 0.851) | <0.001 reference | 0.665 (0.566, 0.763) | <0.001 0.874 |
| rMT1+1C           | 8                | 0.815 (0.769, 0.862) | <0.001 reference | 0.683 (0.592, 0.775) | <0.001 0.385 |

Note. = C-index = concordance index; DMFS = distant metastasis-free survival; CI = confidence interval. MT1, MT2, MT1C, MT1+T2, MT1+T1C, MT2+T1C and MT1+T2+T1C prognostic models were built, and they integrated clinical risk factors (I stage, N stage, and plasma EBV DNA) with the T1, T2, T1C, T2+1C, T1+T2C, T2+T1C and T1+T2+T1C radiomics prognostic models, respectively. rMT1, rMT2, rMT1C, rMT1+T2, rMT1+T1C, rMT2+T1C and rMT1+T2+T1C radiomics prognostic models were built based on I stage, plasma EBV DNA with the T1, T2, T1C, T2+1C, T1+T2C, T2+T1C and T1+T2+T1C radiomics prognostic models, respectively. rMT1C model.

1P-values were calculated compared with the clinical prognostic model.

*P-values were calculated by comparing with the corresponding radiomics prognostic model. For example, P=0.0511 was the result of the comparison between the MT1 model and the T1 model in the validation cohort.

Nomograms and calibration curves for the clinical and T1+T1C radiomics prognostic models are shown in Figure 1. The T1+T1C radiomics prognostic model predicted post-treatment metastasis risk for patients with a 3-year DMFS probability ≥40% or 5-year DMFS probability ≥30%, while the clinical prognostic model required a 3-year DMFS probability ≥70% or 5-year DMFS probability ≥60% to predicted metastasis risk (Figures 1A1, A2). Calibration curves in the probability of 5-year DMFS by a 1000-iteration bootstrap resampling experiment in the validation cohort also showed the T1+T1C prognostic model had better discriminating ability than the clinical prognostic model. Observations showed better agreement with model predictions when the probability of 5-year DMFS was ≥68.9% in the T1+1C prognostic model, while it had to be ≥83.3% in the clinical prognostic model (Figures 1B2, B4).

We calculated the risk scores in the clinical and T1+T1C prognostic models for each patient and then classified patients into the low- and high-risk groups, with zero as risk score cutoff. The distributions of risk scores and 5-year DMFS in the low- and high-risk groups are shown in Figure 2. As shown, post-treatment metastatic patients were concentrated in the high score area, and the survival curve of the T1+T1C prognostic model showed good prognostic stratification for patients in the low- and high-risk groups in the validation cohort, while such trends were not observed in the validation cohort of the clinical prognostic model. When maximally selected rank statistics were used to generate the optimal risk score cutoff value for the clinical and T1+T1C prognostic models to divide patients into low- and high-risk groups, the results were identical to when zero was used as cutoff value (Supplementary Figures S4, S5).

The decision curve analysis of the validation cohort for the clinical prognostic model, radiomics models T1, T2, T1C, T1+T1C, T1+T2+T1C, MT1+T1C model, and rMT1C model is shown in Figure 3 (other prognostic models are not listed). The T1+T1C prognostic model provided the best performance, while the clinical prognostic model showed no net benefit for patients with a 5-year DMFS probability ≥14.9%.

DISCUSSION

Our radiomics prognostic models, especially the T1+T1C model, had better discriminating ability than either the clinical prognostic model or corresponding merged prognostic models integrating clinical factors with selected radiomics features. All seven radiomics prognostic models could stratify patients into high- and low-risk groups based on significantly different 5-year DMFS rates. To our knowledge, this is the first study to clarify the role of multiple versus single MRI sequence radiomics, of radiomics features versus clinical factors, or the combination of clinical factors and radiomics features to
Since NPC is spatially and temporally heterogeneous (32), plasma EBV DNA does not completely characterize tumor heterogeneity. In this study, the C-index of the clinical prognostic model was 0.552 in the validation cohort, indicating that clinical risk factors were inadequate for distinguishing between different metastasis risk groups and not a reliable method for predicting DMFS in NPC patients.

Radiomics transforms tumor images into detailed quantifications of tumor characteristics to find possible prognostic information for patients (16). Our study showed that radiomics features could be used as biomarkers for DMFS prediction in NPC patients without considering the TNM staging system or other clinical risk factors. There might be two reasons for this result. First, clinical risk factors could not distinguish between different metastasis risks; therefore, adding clinical factors into the radiomics prognostic models could not significantly improve the models’ prediction efficiency. Second, the selected radiomics features might contain prognostic information hidden in clinical factors, and overfitting occurrence might weaken their prediction performance when integrating clinical factors with radiomics prognostic models. Zhang et al. (21) showed that the C-index of a radiomics prognostic model using the TNM staging system or clinical factors (based on age, sex, and hemoglobin) was reduced by 0.009 or 0.013 in the validation cohort, respectively.

### TABLE 3 | Multivariable analysis of radiomic features for the primary cohort.

| Prognostic model | Variable | coefficient | HR (95% CI) | P |
|------------------|----------|-------------|-------------|---|
| T1 radiomics prognostic model | T1_shape_Sphericity | -4.87 | 0.01 (1.78E-04, 0.49) | 0.021 |
| T1_WLHL_GLSZM_LGLZE | -1.47 | 0.03 (0.04, 1.37) | 0.107 |
| T1_WLHL_GLCM_ICM2 | -6.33 | 1.78E-03 (5.51E-05, 5.76E-02) | < 0.001 |
| T1_WLHL_GLCM_ICM2 | 4.93 | 138.93 (2.39, 8.09E3) | 0.017 |
| T1_WLHL_GLCM_ICM2 | -3.41 | 0.03 (6.41E-04, 1.71) | 0.090 |
| T1_log.sigma.3.mnm.3D_NGTDM_Strength | 1.36 | 3.91 (1.19, 12.89) | 0.025 |
| T1_WLHL_NGTDM_Contrast | -14.64 | 4.37E-07 (3.20E-16, 595.36) | 0.172 |
| T2 radiomics prognostic model | T2_WLHL_GLCM_LDHGILE | 8.04E-05 | 1.00008 (1.00006, 1.000115) | 0.034 |
| T2_log.sigma.5.mnm.3D_FOS_Skewness | -0.45 | 0.64 (0.38, 1.06) | 0.084 |
| T2_WLHL_GLSZM_SALGLE | -23.50 | 6.25E-11 (6.92E-18, 5.64E-04) | 0.004 |
| T2_logarithm_NGTDM_Coarseness | 17.62 | 4.48E+07 (1.22E+02, 1.65E+13) | 0.007 |
| T2_WLHL_GLCM_IDMN | 25.21 | 8.86E+10 (1.22E-01, 6.45E+22) | 0.070 |
| T1C radiomics prognostic model | T1C_WLHL_GLCM_Correlation | 6.59 | 7.30E+02 (6.08, 8.77E+04) | 0.007 |
| T1C_WLHL_GLSZM_SAHLGE | 0.03 | 1.03 (1.01, 1.05) | 0.010 |
| T1C_Gradient_GLCM_ICM1 | 9.33 | 1.13E+04 (4.0, 3.23E+08) | 0.075 |
| T1C_Square_GLCM_Correlation | 3.22 | 25.02 (1.95, 2.31E+02) | 0.013 |
| T1C_Gradient_GLSZM_ZE | 0.60 | 1.83 (0.93, 3.58) | 0.079 |
| T1C_square_GGLRM_RE | -1.66 | 0.19 (0.06, 0.61) | 0.005 |
| T1C_log.sigma.3.mnm.3D_GLSZM_ZE | 0.94 | 2.55 (1.07, 6.07) | 0.035 |
| T1C_WLHL_GLSZM_ZE | -1.05 | 0.35 (0.11, 1.06) | 0.063 |
| T1C_Gradient_GLSZM_GLN | 0.04 | 1.04 (0.99, 1.08) | 0.104 |
| T1C_Gradient_GLSZM_ZE | 0.04 | 1.04 (0.99, 1.08) | 0.104 |
| T1C_Gradient_GLSZM_GLN | 0.04 | 1.04 (0.99, 1.08) | 0.104 |
| T1+1C radiomics prognostic model | T1 radscore* | 0.59 | 1.80 (1.25, 2.54) | 0.001 |
| T1C radscore* | 0.75 | 2.12 (1.53, 2.94) | < 0.001 |
| T2 radscore* | 0.42 | 1.52 (1.04, 2.23) | 0.031 |
| T2 radscore* | 0.48 | 1.61 (1.44, 2.28) | 0.007 |
| T1C radscore* | 0.58 | 1.79 (1.26, 2.54) | 0.001 |

**Note:** T1, T1C, and T2 are three common methods used in MRI tumor diagnosis. In single-MRI prognostic models for DMFS, the TIC prognostic model had the highest discriminating ability in our study, followed by the T1 and T2, in both primary and validation cohorts. Differences in discriminating ability may be due to imaging capabilities. T1-w can detect tumor anatomical details; T2-w images are sensitive for detecting effusion or edema; T1C-w images may reflect tumor
FIGURE 1 | (A) Nomograms for 3- and 5-year distant metastasis-free survival (DMFS): (A1) for the clinical prognostic model, (A2) for the T1+T1C prognostic model. The nomogram allows the user to obtain the probability of 3- and 5-year DMFS corresponding to a patient combination of covariates. As an example, locate the patient T stage and draw a line straight upward to the "Points" axis to determine the score associated with that T stage. Repeat the process for each variable and sum of the scores achieved for each covariate; after that, locate this sum on the "Total Points" axis. Draw a line straight down to determine the likelihood of 3- or 5-year DMFS. (B) Calibration curves for predicting 5-year DMFS: (B1) in the primary cohort of the clinical prognostic model, (B2) in the validation cohort of the clinical prognostic model, (B3) in the primary cohort of T1+T1C prognostic model, (B4) in the validation cohort of T1+T1C prognostic model. The Y-axis shows observed survival estimated by the Kaplan–Meier method, and the X-axis shows predicted survival calculated using the prognostic model. The solid lines represent the ideal reference line for which predicted survival corresponds with actual survival.
FIGURE 2 | (A) Risk score distributions: (A1) in the primary cohort of the clinical prognostic model, (A2) in the validation cohort of the clinical prognostic model, (A3) in the primary cohort of the T1+T1C prognostic model, (A4) in the validation cohort of the T1+T1C prognostic model. (B) Kaplan–Meier survival curves of distant metastasis-free survival (DMFS) in patients of the low- and high-risk groups: (B1) in the primary cohort of the clinical prognostic model, (B2) in the validation cohort of the clinical prognostic model, (B3) in the primary cohort of the T1+T1C prognostic model, (B4) in the validation cohort of the T1+T1C prognostic model.
angiogenesis, closely related to tumor invasion and metastasis. For multiple-MRI prognostic models for DMFS, the T1+T1C prognostic model had the best performance in the validation cohort, with a higher C-index than the T1+T2, T2+T1C or T1+T2+T1C prognostic models. Among all prognostic models, the T1+T1C prognostic model also showed the best performance in the decision curve analysis of the validation cohort. Therefore, the radiomics features from T1-w and T1C-w MRI sequences better characterize metastatic biological behaviors than other combinations of the three MRI modalities. Except for one shape feature (sphericity), the other 15 features of the composition of T1+T1C radiomics model are textural features, which reflect tumor heterogeneity. This indicates a radiomic signature of some textural features can be regarded as a potential prognostic biomark to discriminate different metastasis risk groups in NPC patients.

Our study had several limitations. First, the patients in the primary and validation cohorts were enrolled from the same institution, which may reduce the generalizability of our findings. Second, follow-up was incomplete for some patients, which may have led to distortions. Third, although PyRadiomics adheres for the most part to the Image Biomarker Standardization Initiative (IBSI), some small differences have been duly noted for it in terms of quantifying the grey level values, removing features associated with the ROI volume from the very beginning for example. Fourth, other MRI-based radiomics were not included, especially diffusion-weighted imaging and dynamic contrast-enhanced, significant for treatment monitoring and outcome prediction in different cancer types. Thus, the usefulness of diffusion-weighted imaging and dynamic contrast-enhanced MRI-based radiomics in NPC patients should be further explored.

In conclusion, a T1+T1C radiomics prognostic model could be a reliable approach for individual risk discriminating of distant metastasis in nasopharyngeal carcinoma patients. This model provided better prognostic performance than other radiomics prognostic models, clinical risk factors, and combination models, and may facilitate personalized risk stratification and treatment strategies for patients with nasopharyngeal carcinoma. To expand the generalizability of this prognostic model, validation with data from prospective, large sample, multicenter studies is required.

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Medical Ethics Committee of Sun Yat-sen University Cancer center. The patients/participants provided their written informed consent to participate in this study. Written informed consent was obtained from the individual(s) for the publication of any potentially identifiable images or data included in this article.

AUTHOR CONTRIBUTIONS

H-JL, L-ZL, YH, and G-YZ designed the research. H-JL and G-YZ performed the data analysis and drafted the manuscript.
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SUPPLEMENTARY MATERIAL

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