Predicting aquatic development and mortality rates of *Aedes aegypti*

Josef Zapletal1*, Himanshu Gupta1, Madhav Erraguntla1, Zach N. Adelman2, Kevin M. Myles2, Mark A. Lawley1

1 Department of Industrial and Systems Engineering, Texas A&M University, College Station, Texas, United States of America, 2 Department of Entomology, Texas A&M University, College Station, Texas, United States of America

* jozinzapletal@tamu.edu

**Abstract**

Mosquito-borne pathogens continue to be a significant burden within human populations, with *Aedes aegypti* continuing to spread dengue, chikungunya, and Zika virus throughout the world. Using data from a previously conducted study, a linear regression model was constructed to predict the aquatic development rates based on the average temperature, temperature fluctuation range, and larval density. Additional experiments were conducted with different parameters of average temperature and larval density to validate the model. Using a paired t-test, the model predictions were compared to experimental data and showed that the prediction models were not significantly different for average pupation rate, adult emergence rate, and juvenile mortality rate. The models developed will be useful for modeling and estimating the upper limit of the number of *Aedes aegypti* in the environment under different temperature, diurnal temperature variations, and larval densities.

**Introduction**

The global burden of mosquito-borne pathogens such as dengue, chikungunya, and Zika virus, has been increasing with changing climate and the expansion of mosquito populations into new areas [1–3]. *Aedes aegypti* are well known to be competent vectors of these pathogens and flourish under temperate climates [4–8]. Feeding primarily on humans and breeding in man-made containers in close proximity, *Aedes aegypti* are among the species of mosquitoes that are contributing to the dispersion of mosquito-borne pathogens in human populations [9,10].

Rainfall and nutrients are necessary factors for the viability of juvenile mosquitoes to hatch and develop, however, several other environmental factors such as exposure to light, average temperature, and diurnal temperature fluctuation ranges have been shown to impact the rates at which juvenile mosquitoes develop [11–24]. Prior studies have shown that larval density within aquatic breeding sites also play a role in the rate at which larvae pupate and pupae emerge as adults [25–30].

In this study, we utilize results from experiments detailed by Zapletal et al. [30] and fit a regression model to predict the rates of *Ae. aegypti* pupation, emergence and mortality under
varying conditions of average temperature, diurnal temperature range, and larval density under conditions of abundant nutrition. Validation using additional experimental data was conducted to assess the validity of the model. Estimating the development rates of *Ae. aegypti* will provide greater insight into the population dynamics of this mosquito species across a variety of environmental conditions.

**Methods**

Experimental data was obtained from previously published results in Zapletal et al. [30]. The laboratory-based experiments performed examined various aspects of aquatic *Ae. aegypti* development including time to first pupation, time of first emergence, maximum rate of pupation, time of maximum rate of pupation, maximum rate of emergence, time of maximum rate of emergence, final average proportion of adult emergence, and average proportion of larval mortality. The experiments conducted observed aquatic development rates under five larval densities (0.2, 1, 2, 4, and 5 larvae/mL of water), two constant temperatures (26.5˚C and 32˚C), and two diurnal temperature ranges (21–32˚C and 26.5–37.5˚C). Diurnal temperatures followed the daily temperature patterns, increasing linearly for nine hours from the low temperature in the morning (07:00) to the high temperature in the late afternoon (16:00) and then decreasing to the low temperature for 15 hours. The larvae were fed with an excess of food so the impact of the lack of nutrition on aquatic development was minimized [30].

Using the data provided, a weighted average of the time to pupation and time to emergence were calculated according to the number of pupae or newly emerged adults recorded at each observation time. The average hourly rate of pupation and emergence was computed by taking the reciprocal of the average time of pupation and emergence, respectively. The hourly juvenile mosquito mortality rate was computed by summing the numbers of dead larvae and pupae and dividing by the total duration of the experiment (in hours). Three different Linear Regression models were developed for average pupation rate, average emergence rate, and juvenile mortality rate.

Each of the linear regression models used 3 basic predictor variables (*X*<sub>1</sub>, *X*<sub>2</sub>, and *X*<sub>3</sub> in Eq 1): (1) average temperature (26.5 or 32˚C), (2) temperature fluctuation (0˚C for constant or 11˚C for diurnal temperature patterns), and (3) larval density (0.2, 1, 2, 4, or 5 larvae/mL of water). Linear regression models for average pupation, emergence, and mortality rates were created using Python 3 in Jupyter. To capture the non-linear effects, polynomial transformation of larval density (up to degree 4; e.g., *X*<sup>4</sup> in Eq 1), and interactions between average temperature, temperature range, and larval density (e.g., *X*<sub>1</sub>*X*<sub>2</sub>, *X*<sub>1</sub>*X*<sub>3</sub>, and *X*<sub>2</sub>*X*<sub>3</sub> in Eq 1) were included in the model. After developing the linear regression models with these linear and non-linear variables, insignificant variables (p-value less than 0.01) were dropped, resulting in the final model presented in Eq 1.

Five of the 20 predictor combinations of average temperature, temperature fluctuation, and larval density were excluded from the model training data and used for validation of the model predictions. These five combinations were excluded randomly, while ensuring that each predictor value was omitted at least once. The excluded combinations are provided in Table 1. Predictions for each excluded combination were calculated using the model and tested against the experimental values obtained by using a paired t-test. Statistical analysis was conducted using Minitab 17.

**Results**

The average rate of pupation was positively correlated with average temperature (*X*<sub>1</sub>), temperature range (*X*<sub>2</sub>), and larval density (*X*<sub>3</sub>). Insignificant variables were eliminated from the
model ($\alpha < 0.01$) and only significant variables were retained in the model. The predictive model structure was identical for average pupation, average emergence, and average mortality rates. The model structure is provided in Eq 1, with the coefficient for each predictor provided for each model in Table 2 and full analysis of each predictor provided in S1–S3 Tables.

**Eq 1. Predictive model structure**

\[
\text{Average Rate} = B_0 + B_1X_1 + B_2X_2 + B_3X_3 + B_4X_3^2 + B_5X_3^3 + B_{12}X_1X_2 + B_{13}X_1X_3 + B_{23}X_2X_3 + B_{123}X_1X_2X_3
\]

The prediction model for average pupation rate was consistent with experimental observations. The model showed a positive correlation between average temperature and average pupation rate, demonstrating that increases in average temperature increased the rate of pupation. Likewise, a positive correlation of temperature range showed that as temperature fluctuation increased an increased average pupation rate was observed. Larval density showed a positive correlation for degree one and two terms, but a negative correlation for degree three and four terms. As observed within the experiments, slower rates of pupation were associated with high larval densities. Two-factor interactions between average temperature and temperature fluctuation range, average temperature and larval density, and the three-factor interaction of all predictors were negatively correlated with average pupation rate. A positive correlation with average pupation rate was found for the interaction of temperature fluctuation range and larval density. Using all predictors and their interactions, the prediction model for average pupation rate produced an adjusted $R^2$ of 0.8236. Paired t-test results (provided in S4 Table) comparing the average predicted and observed pupation rates from excluded combinations provided a p-value of 0.731, showing no significant difference between the average predicted and observed pupation rates.

**Table 1. Excluded predictor combinations used for model validation.**

| Average Temperature (˚C) | Temperature Range (˚C) | Larval Density (larvae/mL) |
|--------------------------|-------------------------|---------------------------|
| 32                       | 0                       | 0.2                       |
| 26.5                     | 0                       | 1                         |
| 26.5                     | 11                      | 2                         |
| 32                       | 0                       | 4                         |
| 32                       | 11                      | 5                         |

**Table 2. Predictor estimates for average pupation, adult emergence, and juvenile mortality rates.**

| Coefficient | Average Pupation Rate | Average Emergence Rate | Average Juvenile Mortality Rate |
|-------------|-----------------------|------------------------|--------------------------------|
| $B_0$       | 1.203x10^{-3}        | -1.961x10^{-4}         | -3.858x10^{-2}                 |
| $B_1$       | 4.567x10^{-4}        | 3.634x10^{-4}          | 1.504x10^{-3}                  |
| $B_2$       | 5.485x10^{-4}        | 3.575x10^{-4}          | 3.565x10^{-3}                  |
| $B_3$       | 1.259x10^{-4}        | 8.520x10^{-2}          | -3.599x10^{-2}                 |
| $B_4$       | 8.573x10^{-3}        | 5.101x10^{-3}          | 3.493x10^{-3}                  |
| $B_5$       | -5.028x10^{-3}       | -3.218x10^{-3}         | 2.438x10^{-3}                  |
| $B_6$       | -3.411x10^{-3}       | -1.832x10^{-3}         | 4.281x10^{-2}                  |
| $B_{12}$    | -2.133x10^{-3}       | -1.420x10^{-3}         | -1.341x10^{-4}                 |
| $B_{13}$    | -1.027x10^{-4}       | -6.841x10^{-3}         | 1.185x10^{-4}                  |
| $B_{23}$    | 1.035x10^{-4}        | 8.593x10^{-7}          | -1.327x10^{-1}                 |
| $B_{123}$   | -5.453x10^{-7}       | -1.423x10^{-7}         | 6.029x10^{-5}                  |

https://doi.org/10.1371/journal.pone.0217199.0001

https://doi.org/10.1371/journal.pone.0217199.t002

https://doi.org/10.1371/journal.pone.0217199.t001
The model for average pupation rate showed comparable results to the prediction model for average adult emergence rate, where the average temperature and temperature fluctuation range showed positive correlations with the average emergence rate. Furthermore, the larval density predictor showed positive correlation for first and second degree terms, but showed a negative correlation for third and fourth degree terms. Hence, as larval density increased, average adult emergence rate decreased. Two-factor interactions between average temperature and temperature fluctuation range, average temperature and larval density, and the three-factor interaction of all predictors were negatively correlated with average adult emergence rate. Only the interaction between temperature fluctuation range and larval density was positively correlated with average adult emergence rate. The prediction model for average adult emergence rate produced an adjusted $R^2$ of 0.8577 and paired t-test results (provided in S5 Table) show no significant difference between predicted and observed adult emergence rates with a p-value of 0.995.

Average juvenile mosquito mortality rates were positively correlated to average temperature and temperature fluctuation range, resulting in increased mortality rates as average temperature and temperature fluctuation rate increased. First degree terms for larval density were negatively correlated with average juvenile mortality rates, however, second, third, and fourth degree terms were positively correlated. This result agrees with what was observed during the experiments, as high levels of mortality were observed as larval density was increased. Two-factor interactions between average temperature and temperature fluctuation range, and temperature fluctuation range and larval density were shown to be negatively correlated with the average juvenile mortality rate. The two-factor interaction between average temperature and larval density, along with the three-factor interaction of all predictors were found to be positively correlated. The prediction model for average juvenile mortality rate produced an adjusted $R^2$ of 0.7412. Results from the paired t-test (provided in S6 Table) show no significant difference between predicted and observed mortality rates with a p-value of 0.535.

**Discussion**

The results show that the models developed are adequate to predict the aquatic development rates of *Aedes aegypti* based on average temperature, temperature fluctuation, and larval density under conditions of plentiful nutrition. Interesting insights from the model include (i) an increase of average temperature results in increased aquatic development rates, (ii) an increase in temperature fluctuation results in increased aquatic development rates, and (iii) an increase in larval density decreases the development rates. The models of pupation, adult emergence, and juvenile mortality rates were validated by testing the models with predictor combination data excluded from the initial model training. Future laboratory experiments will focus on various average temperatures, larval densities, and smaller daily fluctuations of temperature to extend model coverage to micro-fluctuations in diurnal temperature ranges. While the model predictions were consistent with the experimental observations, future experiments will increase model accuracy when applied to predictor combinations outside of those tested experimentally.

Although the prediction models showed no significant difference when compared to the experimental data, there were several limitations to this study. Only two average temperatures and two temperature fluctuation ranges were tested experimentally. While a linear relationship between these predictors is expected, future experiments will be necessary to confirm this pattern. Extension of the predictor models outside of the predictor ranges tested also poses a challenge. It has been demonstrated that mosquito aquatic development slows at both lower thresholds of 16˚C and upper thresholds of 37˚C, with fluctuations in temperature further...
impacting the development rates [31]. Larval density has been shown to slow larval development rates through the presence of juvenile hormone and competition for limited food resources [17,25–29]. As a result, higher larval densities approach a pupation rate asymptote of 0 pupae per hour. Similarly, since the emergence rate depends on the number of larvae that have pupated, the average rate of emergence will follow a similar trend. Average mortality rate will follow an inverse relationship, increasing with larval density and reaching a maximum rate at a threshold density. Further experiments over wider density ranges are required to identify the threshold value and maximum mortality rate.

The experiments presented in Zapletal et al. [30] were conducted under optimal conditions of nutrition for mosquito development with abundant food supply and competition-reducing conditions. Thus the developments rates derived in this work provide an upper bound to the development rates in the field under sub-optimal nutrition availability conditions. Collection of data on micro-climatic conditions showed that the conditions under which mosquitoes develop can vary significantly from the ambient data collected in the environment [32,33]. The preliminary results have shown that temperatures within these micro-climates can be significantly warmer during winter months, allowing mosquitoes to survive even with ambient temperatures falling below 0˚C.

Estimating the impact of different environmental conditions on *Ae. aegypti* will allow for a better understanding of mosquito population dynamics and provide a more accurate prediction of mosquito population. Furthermore, models that can capture these dynamics at a container level, relying on the individual characteristics and micro-climatic conditions at individual breeding sites, will allow for more accurate models of mosquito populations. The models presented in this paper will provide mosquito development rates at different environmental conditions for use in such detailed mosquito population models. Our future effort is focused on developing such a granular mosquito population model that will leverage the development rates derived from the models presented in this paper.

**Conclusion**

With increasing global climate, the number of people impacted by *Ae. aegypti* and the diseases spread by this vector is increasing [34–37]. Prior work has been conducted to understand the impacts of environmental conditions that play a role in the development of these mosquitoes. This study explored the development of regression models to the aquatic development of *Ae. aegypti* by using the results from the experiment conducted by Zapletal et al. [30]. Regression models for rates of pupation, emergence, and mortality were developed using average temperature, temperature fluctuation range, and larval density as predictor variables.

Additional experiments following the same protocol as the prior experiments were conducted to obtain data points for validation. The validation experiments were designed to test an average temperature and larval densities that were not used to build the regression model. The comparison between the predicted and observed values showed that the model was sufficient to predict the average rates of pupation, emergence, and mortality. While the models accounted for high levels of variation in the experiments, further experiments are necessary for the validation of temperature fluctuation range, as well as the upper bounds for larval density.

These regression models will prove to be useful in future modeling efforts to more accurately estimate mosquito populations. While previous mosquito population models have been developed, many of these models have overlooked the importance of larval density in the development rates of juvenile mosquitoes [38–43]. Also, previous work focused on developing population models at specific average temperature and specific diurnal temperature range. The regression models presented in this paper provide the flexibility to perform population
analysis at any setting of average temperature, diurnal temperature range and density (within the bounds of ranges covered in the experiments). Our future research will utilize the prediction models of juvenile *Ae. aegypti* development and environmental data to examine impacts of micro-climatic conditions on mosquito population dynamics and the diseases that they spread throughout human population.

**Supporting information**

S1 Table. Analysis of predictor estimates for average pupation rate. (DOCX)

S2 Table. Analysis of predictor estimates for average adult emergence rate. (DOCX)

S3 Table. Analysis of predictor estimates for average juvenile mortality rate. (DOCX)

S4 Table. Paired t-test and confidence interval for average pupation rate. (DOCX)

S5 Table. Paired t-test and confidence interval for average adult emergence rate. (DOCX)

S6 Table. Paired t-test and confidence interval for average juvenile mortality rate. (DOCX)

**Author Contributions**

**Conceptualization:** Josef Zapletal, Madhav Erraguntla, Mark A. Lawley.

**Data curation:** Josef Zapletal.

**Formal analysis:** Josef Zapletal, Himanshu Gupta.

**Methodology:** Himanshu Gupta, Madhav Erraguntla, Mark A. Lawley.

**Resources:** Zach N. Adelman, Kevin M. Myles.

**Writing – original draft:** Josef Zapletal.

**Writing – review & editing:** Josef Zapletal, Madhav Erraguntla, Zach N. Adelman, Kevin M. Myles, Mark A. Lawley.

**References**

1. O’meara GF, Evans LF Jr, Gettman AD, Cuda JP. Spread of Aedes albopictus and decline of *Ae. aegypti* (Diptera: Culicidae) in Florida. *J Med Entomol.* Oxford University Press Oxford, UK; 1995; 32: 554–562. https://doi.org/10.1093/jmedent/32.4.554 PMID: 7650719

2. Powell JR, Tabachnick WJ. History of domestication and spread of *Aedes aegypti*–A Review. *Mem Inst Oswaldo Cruz.* SciELO Brasil; 2013; 108: 11–17. https://doi.org/10.1590/0074-0276130395 PMID: 24473788

3. Gubler DJ. Epidemic dengue/dengue hemorrhagic fever as a public health, social and economic problem in the 21st century. *Trends Microbiol.* Elsevier; 2002; 10: 100–103. PMID: 11827812

4. Nene V, Wortman JR, Lawson D, Haas B, Kodira C, Tu ZJ, et al. Genome sequence of *Aedes aegypti*, a major arbovirus vector. *Science* (80-). *American Association for the Advancement of Science*; 2007; 316: 1718–1723.

5. Lourenço-de-Oliveira R, Honorório NA, Castro MG, Schatzmayr HG, Miagostovich MP, Alves JCR, et al. Dengue virus type 3 isolation from *Aedes aegypti* in the municipality of Nova Iguaçu, State of Rio de Janeiro. *Memórias do Inst Oswaldo Cruz.* SciELO Brasil; 2002; 97: 799–800.
6. Morrison AC, Zielinski-Gutierrez E, Scott TW, Rosenberg R. Defining challenges and proposing solutions for control of the virus vector Aedes aegypti. PLoS Med. Public Library of Science; 2008; 5: e68. https://doi.org/10.1371/journal.pmed.0050068 PMID: 18351798

7. Scott TW, Nakasaiti A, Day JF, Kitayapong P, Edman JD. A fitness advantage for Aedes aegypti and the viruses it transmits when females feed only on human blood. Am J Trop Med Hyg. ASTMH; 1997; 57: 235–239. PMID: 9288822

8. Gubler DJ, Clark GG. Dengue/dengue hemorrhagic fever: the emergence of a global health problem. Emerg Infect Dis. Centers for Disease Control and Prevention; 1995; 1: 55. https://doi.org/10.3201/eid0102.952004 PMID: 8903160

9. Hemme RR, Thomas CL, Cha dee DD, Severson DW. Influence of urban landscapes on population dynamics in a short-distance migrant mosquito: evidence for the dengue vector Aedes aegypti. PLoS Negl Trop Dis. Public Library of Science; 2010; 4: e634. https://doi.org/10.1371/journal.pntd.0000634 PMID: 20300516

10. Reiter P, Amador MA, Anderson RA, Clark GG. Dispersal of Aedes aegypti in an urban area after blood feeding as demonstrated by rubidium-marked eggs. Am J Trop Med Hyg. ASTMH; 1995; 52: 177–179. PMID: 7872449

11. Bayoh MN, Lindsay SW. Effect of temperature on the development of the aquatic stages of Anopheles gambiae sensu stricto (Diptera: Culicidae). Bull Entomol Res. Cambridge University Press; 2003; 93: 375–381. PMID: 1461976

12. Yang HM, Macoris MLG, Galvani KC, And Richmond MTM, Wanderley DM V. Assessing the effects of temperature on the population of Aedes aegypti, the vector of dengue. Epidemiol Infect. Cambridge University Press; 2009; 137: 1188–1202. https://doi.org/10.1017/S0950268809002040 PMID: 19192322

13. Carrington LB, Seifert SN, Willi ts NH, Lambrechts L, Scott TW. Large diurnal temperature fluctuations negatively influence Aedes aegypti (Diptera: Culicidae) life-history traits. J Med Entomol. Oxford University Press Oxford, UK; 2013; 50: 43–51. https://doi.org/10.1603/me11242 PMID: 23427651

14. Mohammed A, Chadee DD. Effects of different temperature regimens on the development of Aedes aegypti (L.) (Diptera: Culicidae) mosquitoes. Acta Trop. Elsevier; 2011; 119: 38–43. https://doi.org/10.1016/j.actatropica.2011.04.004 PMID: 21549680

15. Carrington LB, Armijos MV, Lambrechts L, Scott TW. Fluctuations at a low mean temperature accelerate dengue virus transmission by Aedes aegypti. PLoS Negl Trop Dis. Public Library of Science; 2013; 7: e2190. https://doi.org/10.1371/journal.pntd.0002190 PMID: 23638208

16. Lambrechts L, Paaij mans KP, Fansiri T, Carrington LB, Kramer LD, Thomas MB, et al. Impact of daily temperature fluctuations on dengue virus transmission by Aedes aegypti. Proc Natl Acad Sci. National Acad Sciences; 2011; 108: 7460–7465. https://doi.org/10.1073/pnas.1101377108 PMID: 21502510

17. Couret J, Dotson E, Benedict MQ. Temperature, larval diet, and density effects on development rate and survival of Aedes aegypti (Diptera: Culicidae). PLoS One. Public Library of Science; 2014; 9: e87468. https://doi.org/10.1371/journal.pone.0087468 PMID: 24498328

18. Bayoh MN, Lindsay SW. Temperature-related duration of aquatic stages of the Afrotropical malaria vector mosquito Anopheles gambiae in the laboratory. Med Vet Entomol. Wiley Online Library; 2004; 18: 174–179. https://doi.org/10.1111/j.0269-283X.2004.00495.x PMID: 15189243

19. Rueda LM, Patel KJ, Axtell RC, Stinner RE. Temperature-dependent development and survival rates of Culex quinquefasciatus and Aedes aegypti (Diptera: Culicidae). J Med Entomol. Oxford University Press Oxford, UK; 1990; 27: 892–898. https://doi.org/10.1093/jmedent/27.5.892 PMID: 2231624

20. Love GJ, Whelchel JG. Photoperiodism and the development of Aedes triseriatus (Diptera: Culicidae). Ecology. JSTOR; 1955; 36: 340–342.

21. Baker FC. The effect of photoperiodism on resting, treehole, mosquito larvae. Can Entomol. Cambridge University Press; 1935; 67: 149–153.

22. Holzapfel CM, Bradshaw WE. Geography of larval dormancy in the tree-hole mosquito, Aedes triseriatus (Say). Can J Zool. NRC Research Press; 1981; 59: 1014–1021.

23. Merritt RW, Dadd RH, Walker ED. Feeding behavior, natural food, and nutritional relationships of larval mosquitoes. Annu Rev Entomol. Annual Reviews 4139 El Camino Way, PO Box 10139, Palo Alto, CA 94303–0139, USA; 1992; 37: 349–374. https://doi.org/10.1146/annurev.en.37.010192.002025 PMID: 1347208

24. Farjana T, Tuno N, Higa Y. Effects of temperature and diet on development and interspecies competition in Aedes aegypti and Aedes albopictus. Med Vet Entomol. Wiley Online Library; 2012; 26: 210–217. https://doi.org/10.1111/j.2013-2915.2011.00971.x PMID: 21781339

25. Hawley WA. The effect of larval density on adult longevity of a mosquito, Aedes sierrensis: epidemiological consequences. J Anim Ecol. JSTOR; 1985; 955–964.
26. Lyimo EO, Takken W, Koella JC. Effect of rearing temperature and larval density on larval survival, age at pupation and adult size of Anopheles gambiae. Entomol Exp Appl. Wiley Online Library; 1992; 63: 265–271.

27. Barrera R. Competition and resistance to starvation in larvae of container-inhabiting Aedes mosquitoes. Ecol Entomol. Wiley Online Library; 1996; 21: 117–127.

28. Mori A, others. Effects of larval density and nutrition on some attributes of immature and adult Aedes albopictus. Trop Med. 1979; 21: 85–103.

29. Wada Y, others. Effect of larval density on the development of Aedes aegypti (L.) and the size of adults. Quaest Entomol. 1965; 1.

30. Zapletal J, Erraguntla M, Adelman ZN, Myles KM, Myles KM, et al. Impacts of diurnal temperature and larval density on aquatic development of Aedes aegypti. PLoS One. Public Library of Science; 2018; 13: e0194025. https://doi.org/10.1371/journal.pone.0194025 PMID: 29513751

31. Carrington LB, Armijos MV, Lambrechts L, Barker CM, Scott TW. Effects of Fluctuating Daily Temperatures at Critical Thermal Extremes on Aedes aegypti Life-History Traits. PLoS One. Public Library of Science; 2013; 8: 1–9. https://doi.org/10.1371/journal.pone.0058824 PMID: 23520534

32. Pincebourde S, Murdock CC, Vickers M, Sears MW. Fine-scale microclimatic variation can shape the responses of organisms to global change in both natural and urban environments. Integr Comp Biol. Oxford University Press; 2016; 56: 45–61. https://doi.org/10.1093/icb/icw016 PMID: 27107292

33. Woods HA, Dillon ME, Pincebourde S. The roles of microclimatic diversity and of behavior in mediating the responses of ectotherms to climate change. J Therm Biol. Elsevier; 2015; 54: 86–97. https://doi.org/10.1016/j.jtherbio.2014.10.002 PMID: 26615730

34. Calvez E, Guillaumot L, Millet L, Marie J, Bossin H, Rama V, et al. Genetic diversity and phylogeny of Aedes aegypti, the main arbovirus vector in the Pacific. PLoS Negl Trop Dis. Public Library of Science; 2016; 10: e0004374. https://doi.org/10.1371/journal.pntd.0004374 PMID: 26799213

35. Richard V, Paoaafaite T, Cao-Lormeau V-M. Vector competence of Aedes aegypti and aedes polynesiensis populations from French Polynesia for chikungunya virus. PLoS Negl Trop Dis. Public Library of Science; 2016; 10: e0004694. https://doi.org/10.1371/journal.pntd.0004694 PMID: 27144888

36. Guillaumot L. Arboviruses and their vectors in the Pacific—status report. Pac Heal Dialog. 2005; 12: 45–52.

37. Fauci AS, Morens DM. Zika virus in the Americas—yet another arbovirus threat. N Engl J Med. Mass Medical Soc; 2016; 374: 601–604.

38. Brady OJ, Johansson MA, Guerra CA, Bhatt S, Golding N, Pigott DM, et al. Modelling adult Aedes aegypti and Aedes albopictus survival at different temperatures in laboratory and field settings. Parasit Vectors. BioMed Central; 2013; 6: 351. https://doi.org/10.1186/1756-3305-6-351 PMID: 24330720

39. De Almeida SJ, Ferreira RPM, Eiras AE, Obermayr RP, Geier M. Multi-agent modeling and simulation of an Aedes aegypti mosquito population. Environ Model Softw. Elsevier; 2010; 25: 1490–1507.

40. Erickson RA, Presley SM, Allen LJS, Long KR, Cox SB. A stage-structured, Aedes albopictus population model. Ecol Model. Elsevier; 2010; 221: 1273–1282.

41. Erickson RA, Presley SM, Allen LJS, Long KR, Cox SB. A dengue model with a dynamic Aedes albopictus vector population. Ecol Model. Elsevier; 2010; 221: 2899–2908.

42. Tran A, L’Ambert G, Lacour G, Benoit-Ritt R, Demarchi M, Cros M, et al. A rainfall- and temperature-driven abundance model for Aedes albopictus populations. Int J Environ Res Public Health. Multidisciplinary Digital Publishing Institute; 2013; 10: 1699–1719. https://doi.org/10.3390/ijerph10051698 PMID: 23624579

43. Jia P, Lu L, Chen X, Chen J, Guo L, Yu X, et al. A climate-driven mechanistic population model of Aedes albopictus with diapause. Parasit Vectors. BioMed Central; 2016; 9: 175. https://doi.org/10.1186/s13071-016-1448-y PMID: 2709065