Animal models of intellectual disability: towards a translational approach

Carla A. Scorza,1 Esper A. Cavalheiro1II

1Disciplina de Neurologia Experimental. Universidade Federal de São Paulo/Escola Paulista de Medicina. (UNIFESP/EPM). São Paulo, Brazil. II APAE – São Paulo, Brazil.

Intellectual disability is a prevalent form of cognitive impairment, affecting 2–3% of the population in the industrialized world.1 ID is characterized by significant limitations both in intellectual functioning and in adaptive behavior as expressed in conceptual, social and practical adaptive skills. Moreover, ID is one of the few disorders for which the etiology and pathogenesis are still poorly understood. ID is accepted if the intelligence quotient (IQ) is lower than 70 and is considered borderline from 70 to 85.3,4 However, a multidimensional framework for understanding ID depicts how human functioning and the manifestation of ID involve the dynamic, reciprocal engagement among intellectual ability, adaptive behavior, health, participation, context and individualized supports.2 ID is a condition of great concern for public health and society. The frequency of ID-related cognitive dysfunction is alarming, considering that pharmacological intervention is currently non-existent.5 In this context, ID is one of the more important topics in medical science. It has a complex etiology, with an intricate interplay of genetic and environmental factors, but the causal mechanisms are not understood. ID is one of the few clinically important disorders for which the etiopathogenesis is still poorly understood. In this sense, in order to improve human health, scientific discoveries must be translated into practical applications. Such discoveries typically begin at “the bench” with basic research and then progress to the clinical level, or the patients “bedside”. Scientists are increasingly aware that this bench-to-bedside approach to translational research is really a two-way street.6 Scientists provide clinicians with new tools for use with patients and for assessment of their impact, and clinical researchers make novel observations about the nature and progression of a condition, which often stimulates basic investigation.7 This is the first area of translational research called “bench-to-bedside and back”. The second area concerns research aimed at enhancing the adoption of best practices in the community. As an example, improvements in medical intervention for people with Down syndrome (DS) have led to a substantial increase in their longevity, with the estimated life expectancy in developed countries increasing from an average of 12 years in the 1940s to an average of 57.8 years for women and 61.1 years for men.7,8 In this sense, animal models have been invaluable in ID research, since studies in humans have a very limited capacity to explain basic mechanisms of disease. Animal models facilitate the developmental analysis of the pathogenesis of abnormalities, particularly during crucial stages of organogenesis. Even if appropriate tissues were available in humans, it would be impossible to carry out any investigations other than static biochemical or morphological examinations. Consequently, a great deal of the knowledge that has improved our understanding of several pathologies has derived from appropriate animal
models. This is certainly the case of pathologies associated with ID. Arguably the single most essential element in animal-based research, identifying and selecting the most appropriate animal model, is also the most challenging. Different species have species-specific behavioral repertoires shaped by their evolutionary history. The evolutionary process, by definition, actually refers to the vast differences that exist between species, and even between cells within a given species. Thus, modeling of human-like symptoms in animals should be based primarily on an expectation of functional similarity of the displayed behavioral strategy, rather than on one of behavior equivalency. The crucial point is not whether a mouse, for example, would show a given cognitive impairment, but, rather, how a cognitive impairment would manifest itself in this mouse. The challenge that basic science research needs to meet is to make use of a comparative approach to benefit the most from what each model can tell us. Anyhow, the use of animal models of ID are critical for determining the basic mechanisms, the neurobiological substrates and the neural basis of cognitive function as well as for testing the efficacy of potential therapeutic drugs and the neurotoxicity of environmental contaminants and drugs of abuse, among others.

The causes of ID are extremely heterogeneous and although a cause for ID has been diagnosed in about half of cases, it has been estimated that half of all cases are due to environmental factors and half to genetic factors. Environmental factors include prenatal exposure of the fetus to toxic substances (e.g., alcohol, drugs), environmental contaminants, radiation, infection, malnutrition, illness of the mother (e.g., exposure to toxoplasmosis, cytomegalovirus and rubella), etc. In addition, multiple problems, during or after birth, may also culminate in ID. This review shall describe several animal models of potential ID risk factors. We have organized this review as shown in Table 1.

### GENETIC MODELS

About one-third of all genetic disorders show some neurological involvement, and many of these represent neurodegenerative diseases of infancy. Genetic or genetically influenced conditions rank among the leading causes of organically based mental retardation. Genetically modified mice are currently the most commonly used approach to investigate the role of a specific genetic alteration and to model pathologies leading to intelligence disability. The mouse is the most widely used laboratory species to provide insights linking specific genes to biological functions. Its wide use is primarily because, among mammals, the mouse is most amenable to genetic manipulation. Furthermore, our extensive knowledge of the genome, physiology and behavior of the mouse makes it possible to interpret the effects of gene manipulations.

### Down syndrome

DS caused by trisomy of human chromosome 21 (HSA21), shows an incidence of about 1 in 733 live births. DS is characterized by abnormalities in learning, memory and language; some degree of intelligence disability is essentially universal. Furthermore, individuals with trisomy 21 may show a wide range of pathological features, such as heart disease, sterility, higher incidence of leukemia, immune system perturbations and premature aging. Moreover, they also display muscular hypotonia, deposition of Alzheimer-like plaques and neurofibrillary tangles in the brain in the third decade of life. Lejeune et al. showed that an extra copy of human chromosome 21 (now HSA21) was found in DS. The discovery provided the first evidence for a genetic substrate of intelligence disability. Although DS pathogenesis may be complex, all changes must arise from excess genetic material on HSA21, which encodes 300 known and predicted genes. Mouse models of DS have been developed using syntenies between HSA21 and MMU16, MMU10 and MMU17. Available mouse models carry extra fragments of MMU16 or of HSA21 that cover all of HSA21 (chimeric HSA21) or MMU16 (Ts16); some carry large parts of MMU16 (Ts65Dn, Ts1Cje, Ms1Cje), whereas others have reduced contiguous fragments covering the D21S17-ETS2 region or single transfected genes. Among these, Ts65Dn and Ts1Cje mouse models have been extensively used to study DS neurobiological phenotypes. The Ts65Dn mouse, with an extra copy of about 104 mouse genes orthologous to those on HSA21, shows a number of developmental and functional parallels with DS. These include changes in behavior, alterations in the structure of dendritic spines in the cortex and hippocampus, and failed hippocampal long-term potentiation (LTP).

### Table 1 - Animal models.

| Genetic models               | Down syndrome                  |
|----------------------------|--------------------------------|
| Fragile X syndrome         |                                |
| Rett syndrome              |                                |
| Inborn errors of metabolism|                                |
| Neurotoxic insults          |                                |
| Selective neurotoxin-induced model |                |
| Neurotoxic treatment of the developing fetus |             |
| Environmental neurotoxic insults |                          |
| Maternal exposure to infections |                            |
| Congenital toxoplasmosis    |                                |
| Congenital cytomegalovirus  |                                |
| Congenital rubella          |                                |
| Maternal exposure to teratogens |                            |
| Fetal alcohol syndrome      |                                |
| Malnutrition                |                                |
| Brain lesions at early development |                |
| Hypoxic-ischemic insult     |                                |
| Pediatric head trauma       |                                |
| Status epilepticus          |                                |
| Age-related changes         |                                |
| Down syndrome Ts65Dn mouse  |                                |
| Transgenic mouse models of amyloid deposition |         |
| Cholinergic receptor knockout mice |              |
the mechanisms by which the anatomical, chemical and neurophysiological brain abnormalities underlying ID arise from deregulation of gene expression? Major questions are still unanswered. Genetically modified mouse models have been helping to investigate the contributions of specific gene alterations and gene–environment interactions to the phenotype of several forms of ID.

Fragile X syndrome
X-linked disorders may explain why males show a 25–35% higher incidence of mental disabilities than females. It is imperative to gain knowledge of the mechanisms specific to the X-chromosome (e.g., X-inactivation, gene regulation) that may be relevant to understanding some forms of X-linked mental retardation. Fragile X syndrome (FXS) is the leading cause of heritable intelligence disability, affecting about 1 in 1400 males and 1 in 2500 females. FXS is caused by a trinucleotide expansion in the fragile X mental retardation 1 gene (fmr1) that prevents the expression of the encoded protein, called fragile X mental retardation protein (FMRP). The genetic defect is an unstable region of DNA on the X chromosome that becomes highly expanded when transmitted through the maternal lineage. FXS is a common form of mental retardation associated with attention deficit, autistic behavior, hyperactivity and epileptic seizures. The phenotype of FXS is reproduced in fmr1 knockout (KO) mice that, among others, have regionspecific altered expression of some gamma-aminobutyric acid (GABA) receptor subunits.

Rett syndrome
Rett syndrome (RTT) is an X-linked autism spectrum disorder found almost solely in females. Clinical characteristics include abnormal motor gait, stereotypic hand wringing movements and autistic-like behavior. Affected girls also exhibit speech abnormalities and severe intellectual disability in most cases. One peculiar aspect of this disorder is that individuals appear normal at birth, then, between 6 and 18 months, they begin to lose some already acquired skills, such as communication, language and motor coordination. RTT is caused by mutations in MECP2, encoding methyl CpG-binding protein 2. Since the discovery of MECP2 mutations as the genetic cause of RTT in 1999, the understanding of MeCP2 function has evolved. In past years, researchers have produced mice with genetically altered MeCP2 that displayed some features of Rett in order to characterize the biological, pathological and behavioral features of these mice and have compared them with the human condition. Combined epigenomic approaches of MeCP2 binding, methylation and gene expression have demonstrated that MeCP2 binds preferentially to intergenic and intronically and sparsely methylated promoters of active genes. While autism is strongly heritable, most cases of autism are expected to be due to a combination of genetic, environmental and epigenetic factors. As in the case of RTT, genetic disorders on the autism spectrum affecting epigenetic pathways include Angelman, Prader–Willi and 15q duplication syndromes. Thus, armed with intense research, improved understanding and therapies for RTT, and perhaps a subset of autism cases, are certain to follow.

Inborn errors of metabolism
Metabolic disorders result from the absence or abnormality of an enzyme or its cofactor, leading to either accumulation or deficiency of a specific metabolite. Optimal outcome for children with inborn errors of metabolism (IEM) depends upon prompt recognition, evaluation and management of these disorders. Delay in diagnosis may result in acute metabolic decompensation, progressive neurologic injury, or even death. Animal models of metabolic diseases especially focus on the pathophysiology mechanisms due to imbalances in amino acids, mucopolysaccharides, purines, lipids and carbohydrates, or the dysfunction of cellular organelles (e.g., mitochondria, peroxisomes, lysosomes, or Golgi) that contribute to intelligence and developmental disabilities as well as to the development of therapeutic strategies (i.e., genetic or pharmacological) that aid in the diagnosis and clinical management of these IEM disorders. Specific examples of metabolic disorders associated with intelligence disability include phenylketonuria, Lesch–Nyhan, galactosemia and adrenoleukodystrophy. In the 70s in Brazil, Benjamin Schmidt and colleagues started a project called “A national plan for study and detection of IEM disorders that could lead to mental deficiency”. In the sequence, Brazil established the first Newborn Screening Programme for IEM in Latin America for the detection of phenylketonuria and other IEM capable of causing intelligence disability. However, today, the few data available lead to the conclusion that neither newborn screening nor phenylketonuria treatment centers cover all the Brazilian cases. In addition, most phenylketonuria patients have to leave their home states to seek better treatment. Moreover, the required specific foods are not readily available and are expensive. New options are being researched, nevertheless, there is much to be done, mainly on food research and production.

ENVIRONMENTAL CONDITIONS
There are a host of environmental conditions that affect biological risk for intelligence and developmental disabilities, among them are (a) neurotoxic insults, (b) maternal exposure to infections (e.g, toxoplasmosis, cytomegalovirus and rubella), (c) maternal exposure to teratogens (e.g., alcohol), (d) malnutrition.

Neurotoxic insults
Selective neurotoxin-induced models. Many individuals with intellectual disability exhibit chronic aberrant behavior that includes hyperactive, stereotyped, aggressive and self-injurious behaviors (SIBs). SIB is a severe symptom in Lesch–Nyhan syndrome (LNS), a genetic disorder associated with deficiency of the enzyme hypoxanthine-guanine phosphoribosyltransferase (HPRT). HPRT-deficient mice provided the first demonstration that genetically engineered mice could be produced as models for a specific human disease. It has been shown that these mice presented reduced dopamine along with microstructural anatomical abnormalities, demonstrating that HPRT deficiency leads to a loss of basal ganglia dopamine through a metabolic process. Unfortunately, the main limitation of these mice is that they do not exhibit some of the more complex aspects of the LNS phenotype, such as the consequences of uric acid overproduction or the neurobehavioral syndrome. As a result, they are of limited value for directly addressing these aspects of the disorder. In this sense, another animal model has been developed to address the functional significance of dopamine
loss due to neurobehavioral syndrome. Because the prominent neurobiological deficit in LNS is a loss of dopaminergic neurons, reduction of dopamine 6-hydroxydopamine in rats during their development is proposed as a model of dopamine deficiency in LNS as well as in individuals with ID and SIB, who may have this neurotransmitter deficiency. Accompanying the profound loss of dopamine was an increase in striatal serotonin content. When the neonatally lesioned rats were challenged as adults with systemically administered L-DOPA or with muscimol administration into the substantia nigra reticulata (SNR), SMB was observed, a response not observed in unlesioned rats. Since D3-dopamine antagonist blocked the SMB response to L-DOPA, it was proposed that D3-dopamine receptors were critical to this behavioral response.

**Neurotoxic treatment of the developing fetus.** The administration of the alkylating neurotoxin methylazoxymethanol acetate (MAM, 25 mg/kg sc) to pregnant rats on day 14 or 15 of gestation induces, in the offspring, a marked microencephaly. MAM treatment severely reduces intrinsic neuronal populations including GABAergic and glutamatergic neurons and produces a shrunken cortex relatively hyperinnervated by noradrenergic and cholinergic neurons. MAM-induced brain and behavioral abnormalities provide an animal model of congenital microencephaly associated with intelligence disability. The MAM offspring showed no increase in mortality, but weighed less than controls. In addition, MAM-treated rats display a characteristic hyperactivity. This hyperactive state was accompanied by notable impairments in the acquisition of spatial instrumental learning tasks. At 2 months of age, the MAM offspring showed pronounced deficit in learning a water maze; this deficit was not abated when animals were tested again at 6 months of age, implicating long-term learning impairments.

**Environmental neurotoxic insults.** Neurotoxicity is a potentially major contributor to the etiology of many types of intelligence and developmental disabilities. For instance, models of developmental risk in neurotoxicology have guided environmental regulation to reduce the likelihood of neurotoxic effects. On the other hand, models of developmental risk for ID aim for prevention and early intervention. Over the past three decades, researchers have found that remarkably low-level exposure to environmental toxins (e.g., heavy metals, polychlorinated biphenyls (PCBs), pesticides and herbicides, organic solvents, environmental tobacco smoke, radiation and endotoxins) has been linked to intellectual impairment, learning disabilities, behavioral problems, spontaneous abortions, or preterm births. Lead is probably the most-studied environmental contaminant with respect to the effects of developmental exposure on cognition in children or animal models. PCBs were banned in the 1970s but their extreme stability allowed them to bioaccumulate over time, such that they remain one of the most prevalent environmental contaminants. In this case, animal research has helped to reveal the differential effects of neurodevelopmental exposure to PCBs in producing significant disruption of tasks that require executive function. As a testament to the usefulness of animal models, recent follow-up studies of several human PCB cohorts have employed domain-specific tests such as the Wisconsin Card Sorting Task and continuous performance tasks that assess specific aspects of executive control. These recent assessments have yielded results that are strikingly consistent with the findings in animal models, and they highlight the utility of animal models for predicting specific cognitive domains that are likely to be impacted in exposed human populations.

**Maternal exposure to infections.** Different animal models of congenital toxoplasmosis have previously been developed in mice, rats, sheep and guinea pigs. The intrauterine protozoan *Toxoplasma gondii* is a widespread opportunistic parasite of humans and animals, and transmission from animals to humans occurs mainly through oocysts excreted in the feces of infected cats, and meat products from farm animals contaminated with viable tissue cysts. Pregnant women who are infected with *T. gondii* usually remain asymptomatic, although they can still transmit the infection to their fetuses, with severe consequences. Primary infection during pregnancy may result in neonatal death or in severe congenital defects such as hydrocephalus, mental retardation and retinochoroiditis, which may occur at birth or during development. An effective vaccine should protect against both acute and chronic infection. The development of suitable laboratory models is essential for evaluation of the efficacy of the different recombinant subunit vaccine candidates against congenital toxoplasmosis.

**Congenital and perinatal infections with cytomegalovirus.** These are responsible for considerable short- and long-term morbidity in infants. Cytomegalovirus (CMV) is the most common congenital viral infection in the developed world, occurring in 0.5–2% of all births and is a common cause of neurodevelopmental injury, including mental retardation and sensorineural hearing loss (SNHL). Its clinical manifestations range from asymptomatic forms (90% of cases) to severe fetal damage and, in rare cases, death due to abortion. Furthermore, 10–15% of children who are asymptomatic at birth may develop late sequelae after a period of months or even years. Animal models are imperative for the understanding of the host-virus relationship, which can be exploited to delineate the mechanisms of pathogenesis to develop therapeutic strategies to prevent CMV-associated diseases; however, CMVs are highly species specific. Consequently, each CMV species is prevalent environmental contaminant with respect to the effects of developmental exposure on cognition in children or animal models. In this case, animal research has helped to reveal the differential effects of neurodevelopmental exposure to PCBs in producing significant disruption of tasks that require executive function. As a testament to the usefulness of animal models, recent follow-up studies of several human PCB cohorts have employed domain-specific tests such as the Wisconsin Card Sorting Task and continuous performance tasks that assess specific aspects of executive control. These recent assessments have yielded results that are strikingly consistent with the findings in animal models, and they highlight the utility of animal models for predicting specific cognitive domains that are likely to be impacted in exposed human populations.

**Congenital rubella syndrome.** Most of the environmental conditions that affect the biological risk for intelligence and developmental disabilities are preventable. In several cases, we, as a society, have been remarkably successful in preventing their effects. Congenital rubella syndrome (CRS) is a case in point. In 1963–65 there was an epidemic of more than 25,000 cases of CRS in the United States. However, since the development of the rubella vaccine in 1969, the incidence has dropped markedly. In 1996 only two cases of CRS were reported in the United States. Although Brazil strives to achieve high immunization coverage through the routine vaccination program and follow-up campaigns, rubella incidence increased in the country between 2006 and 2007. For this reason, in 2008, Brazil launched a major and successful...
The main defects associated with CRS are deafness, eye defects such as cataracts, cardiovascular defects and central nervous system damage, leading to intelligence disability. It is common to try to study human pathologies by establishing animal models and certainly that was the case for CRS. However, several discrepancies have been found in such studies. Low incidences of cataracts have been reported in the offspring of infected pregnant rats, rabbits and monkeys. In contrast, other studies with pregnant animals (e.g., rats, mice, rabbits, ferrets and monkeys) have not shown these defects. Moreover, abnormalities of the heart, cerebral blood vessels, and inner ear have been reported in some studies, but incidences are very low and inconsistent. Taken together, animal models of CRS have not contributed much to the understanding of their pathogenesis. On the other hand, the use of animals as models for microbiological infections has been a fundamental part of infectious disease research. Thus, the use of animal models for a myriad of bacterial and viral diseases has led to the production of vaccines for diseases such as rubella.

Maternal exposure to teratogens

Fetal alcohol syndrome. Fetal alcohol syndrome (FAS) and the subsets of individuals with attenuated phenotype subsumed under the umbrella term of fetal alcohol spectrum disorder (FASD) provide clinicians with a challenge. Individuals with FASD have a higher incidence of impairments in social adaptive and executive function, and a higher degree of psychopathology than the general population. The diagnosis of FAS is based on the occurrence of the following main symptoms: growth deficiency, dysmorphic characteristics, central nervous system dysfunctions, intelligence disability, delays in sensorimotor development, attention deficit, memory disorders, hyperactivity and sleep disturbances. Animal models of FASD have been used to demonstrate the specificity of the teratogenic effects of alcohol and some of the underlying changes in the central nervous system and to explore ways to ameliorate the effects of alcohol. The effects of acute alcohol exposure on specific days during the embryonic period have been studied fairly extensively in rodents. Exposure on gestation days 7 and 8 in mice has shown to result in craniofacial defects similar to those seen in FAS (e.g., micrognathia, low-set ears, short philtrum, cleft palate, cleft lip), as well as brain anomalies (e.g., microcephaly, exencephaly, deficiencies in cerebral hemispheres, striatum, olfactory bulbs, limbic structures, corpus callosum, lateral ventricles). In addition, ocular defects (anophthalmia, microphthalmia, corneal and lens anomalies) were associated with acute alcohol exposure on gestation day 7. Research has indicated that environmental enrichment and voluntary exercise have been shown to ameliorate some of the effects of alcohol during development, but the roles of enhanced social interactions in the case of enrichment, social housing and voluntary exercise need to be more fully delineated. In addition, findings of alterations in maternal care of the alcohol-exposed offspring, epigenetic effects and their relationship to social behavior in animal models of FASD are a fruitful area of research.

Malnutrition

Malnutrition is a worldwide health problem; it exists in many forms, affects the developing and mature nervous system, and has acute and chronic health implications. Malnutrition in children has particularly severe consequences for growth, development, health and well-being, both on a short- and on a long-term basis, including a high risk for ID. Malnutrition was associated with 54% of deaths in children in developing countries. In developed countries, childhood malnutrition occurs mostly secondary to chronic diseases, and it may be aggravated by frequent hospital stays and diagnostic examinations. Most concepts basic to nutritional investigations have been derived from animal studies. One distinct characteristic of the research on protein energy malnutrition and child development is the selective focus on learning and intelligence. Historically, work in animals on the functional consequences of energy and protein deprivation has moved in two directions: one approach aims the understanding of the developmental changes in the anatomy and biochemistry of the brain in experimental animals, while the other one describes the compromise of behavioral competence.

BRAIN LESIONS AT EARLY DEVELOPMENT

Hypoxic-ischemic insult

Despite major advances in monitoring technology and knowledge of fetal and neonatal pathologies, hypoxic-ischemic encephalopathy (HIE) remains a serious condition that causes significant mortality and long-term morbidity. HIE represents a common cause of chronic handicapping conditions such as cerebral palsy, intelligence disability, learning disability and epilepsy. In an established model of perinatal HIE brain damage, neonatal animals undergo carotid artery ligation and hypoxia. Animal models have long been established in the study of neonatal HIE and have been found suitable for acute and chronic studies. Multiple therapeutic trials are currently being conducted in animal models to explore various methods of interventional therapy against HIE-induced brain damage, with recent breakthroughs in the demonstration of a neuroprotective effect of some anti-apoptosis agents and other methods, such as hypothermia. Premature births result in an increased likelihood of intelligence disability. Premature infants are at high risk of intracranial hemorrhage, asphyxia and the damaging effects of subsequent HIE. Existing models of pediatric hypoxic-ischemic brain damage studied the effects of glutamate in the postnatal day 7 rat, which is considered analogous to the newborn human. In this sense, the newborn rat is considered analogous to the late gestational human. In order to model preterm hypoxic ischemic brain damage events, it has been proposed that treatment of newborn rats with muscimol mimics the initiation of a cell death cascade induced by hypoxia or injury in premature infants and is analogous to the accepted method of glutamate administration to the week-old rat pup to model the newborn human. Following this line of evidence, investigators proposed that the excitatory drive through the GABA receptor in early development may be an important contributor to some forms of brain damage in premature infants. Furthermore, they also investigated the effect of postnatal day 0 and 1 muscimol treatment by performing a pre-weaning version of the water maze and an open field maze on postnatal day 21, two tasks that are sensitive to hippocampal function. Taken together, it has been proposed that rats exposed to excessive GABA receptor activation over the first 2 days of postnatal life serve as a model for pre-term infant brain damage.
Pediatric head trauma

Trauma to the developing brain still represents a poorly explored field. Perinatal head trauma can lead to intellectual disability among other severe consequences. Head trauma is the leading cause of death and disability in the pediatric population. In efforts to model pediatric head trauma, Bitigau et al. have developed a model for head trauma in infant rats in an attempt to study mechanisms of neurodegeneration in the developing brain. In order to morphologically characterize two distinct types of brain damage they have demonstrated that, in the developing rat brain, apoptosis and not excitotoxicity determined neuropathologic outcome following head trauma. In this sense, they suggested that radical scavengers and tumor necrosis factor inhibitors may be useful in the treatment of pediatric head trauma. In this line of evidence, trauma triggers both excitotoxic and apoptotic neurodegeneration in the developing rat brain. Excitotoxic neurodegeneration develops and subsides rapidly (within hours), whereas apoptotic cell death occurs in a delayed fashion over several days following the initial traumatic insult.

Status epilepticus

Status epilepticus (SE) is a medical and neurologic emergency. In Brazil, the annual occurrence of SE is about 90,000 cases. Thus, increased awareness of presentation, etiologies and treatment of SE is essential in the practice of critical care medicine. Elegant animal studies in the 1970s and 1980s revealed significant damage to the brain after 30 min of seizure activity, even with control of blood pressure, respiration and body temperature. Damage to brain tissues resulting from recurrent convulsive seizures induced by chemicals such as pilocarpine or kainic acid treatment may also constitute models of intellectual disability. These animal models are useful for the study of SE-induced epileptogenesis and neurological sequelae, especially during early brain development. Our laboratory group showed several permanent abnormalities in rats subjected to multiple SE induced by pilocarpine (ip) during early development (7–9 days old). In adulthood, these animals presented frequent episodes of continuous complex spiking activity and high-voltage ictal discharges and evidence of severe cognitive deficits. SE may result in important plastic changes in critical periods of brain maturation leading to long-lasting epileptogenesis, as manifested by electrographic epileptiform discharges, behavioral deficits and in vitro hyperexcitability of hippocampal networks.

AGE-RELATED CHANGES

Down syndrome Ts65Dn mouse

As described above, DS results from trisomy of all or part of human chromosome 21, which generally accounts for triplication of at least 100 genes. Among these is the gene encoding amyloid protein precursor (APP), as well as genes that upregulate APP expression. Beta-amyloid is overproduced in DS individuals throughout life. The partial trisomy 16 (Ts65Dn) mouse is considered the gold standard of DS mouse models. Ts65Dn mice have an extra copy of the distal aspect of mouse chromosome 16, a segment homologous to human chromosome 21 that contains much of the genetic material responsible for the DS phenotype, including three copies of APP. Ts65Dn mice show developmental delay during the postnatal period as well as abnormal behaviors in both young and adult animals that may be analogous to mental retardation. Additionally, by 6 months of age, Ts65Dn mice begin progressive, age-related decline in choline acetyltransferase levels and cognitive function, features common to adult DS and Alzheimer patients. Moreover, although the Ts65Dn brain is normal on gross examination, there is age-related degeneration of septohippocampal cholinergic neurons and astrocytic hypertrophy, markers of Alzheimer disease pathology that are also present in elderly DS individuals. These findings suggest that Ts65Dn mice may be useful to study certain developmental and degenerative abnormalities in the DS brain.

Cognitive impairment in transgenic mouse models of amyloid deposition.

The identification of the Aβ peptide as a major component of amyloid deposited in brain vessels and subsequently of parenchymal plaques in the brains of Alzheimer’s subjects led to a focus on this molecule as a key element in the pathophysiology of Alzheimer disease. Subsequent work found that some mutations causing the disease occurred in the amyloid precursor protein (APP) that is processed, in some circumstances, into the Aβ peptide. As for other disorders, development of animal models to understand amyloid pathology became an important research goal. Hsiao et al. were the first to demonstrate that APP transgenic mice had both amyloid deposits and memory deficit. They demonstrated deficits in a reference memory version of the open-pool water maze, with the deficits first appearing at an age when the amyloid plaques started to appear (10–11 months). APP transgenic mice are a very good model of amyloid deposition. The patterns of deposition, regional distribution and even the anatomical localization of the short and long variants mimic the human disease. The APP mouse phenotype also consistently includes progressive memory impairment. This phenotype appears to be due to Aβ accumulation and not overexpression of APP, as the BACE1-null background, which overproduces APP but not Aβ, rescues the memory phenotype. In addition, a number of manipulations, most notably immunotherapies, have been found to regulate the memory phenotype. However, the mechanisms mediating memory deficiencies are not clear.

Cholinergic receptor knockout mice

The essential involvement of the cholinergic system in both preclinical and clinical aspects of cognition processes has been extensively proposed. The literature shows that currently approved drugs for the treatment of Alzheimer disease are cholinesterase inhibitors, which exert their efficacy apparently through stimulation of both muscarinic acetylcholine receptors (mAChRs) and nicotinic acetylcholine receptors (nAChRs). Transgenic and KO mouse models are particularly useful for studies of complex neurobiological problems such as mAChR KO mice, nAChR KO mice as well as acetylcholinesterase (AChE) KO mice. To illustrate, in the AChE−/− mice, the M(1), M(2) and M(4) mAChRs showed a striking 50–80% decreased expression in brain regions associated with memory. In addition, a large body of evidence has demonstrated that nicotine has a clinical utility for cognitive enhancement. However, for most diseases in the central nervous system, the coexistence of malfunctions in multiple subtypes of the...
same receptor or in multiple neurotransmitters typically contribute to a complex phenotype such as cognition.

Therefore, cholinergic-based strategies will likely remain as one approach to understanding and to rational drug development for the treatment of Alzheimer disease and other forms of dementia and cognitive impairment.

**CONCLUSION**

ID is a lifelong condition and it is a daunting societal problem. ID results from many different genetic and environmental insults. Although tremendous progress has been made, pharmacological intervention is currently still non-existent and therapeutic strategies remain limited. However, similarities across the spectrum of ID-related disorders argue that common mechanisms underlie the manifestation of learning and memory deficits in intellectual disabled individuals. Interestingly, many of the histological features noted in the brains of subjects with DS parallel phenotypes that have been found in the brains of individuals with other classes of ID, such as genetic disorders, including inborn errors of metabolism and non-genetic insults. Nevertheless, ID is a clinically important disorder for which the etiology and pathogenesis are still poorly understood. The past decades have seen extraordinary progress in understanding, preventing and treating the behavioral and cognitive deficits associated with ID. Doubtless, animal models have been invaluable in ID investigation, and continued progress will require programmatic research. Progress in understanding and treating ID will require translational research efforts. Translation research means the transformation of knowledge through successive fields of research from basic scientific discovery to public health impact.

**REFERENCES**

1. Branci I, Bichler Z, Berger-Sweeney J, Ricceri L. Animal models of mental retardation: from gene to cognitive function. Neuroscience and Behavioral Rev. 2003;27:141–53, doi: 10.1016/S0306-4522(03)00016-2.

2. A/AIDD. American Association on Intellectual and Developmental Disabilities. Intellectual Disability: definition, classification, and systems of supports. The A/AIDD Ad Hoc Committee on Terminology and Classification. 11th ed. Washington, DC: A/AIDD; 2010.

3. Luckassen R, Poulier DL, Pollock JE, Reiss S, Schalock RL, Snell ME, et al. Mental Retardation: Definition, Classification, and Systems of Supports. 9th ed. Washington, DC: American Association on Mental Retardation; 1992.

4. Shogren KA, Turnbull HR. Public policy and outcomes for persons with intellectual disability: extending and expanding the public policy framework of A/AIDD’s 11th Edition of Intellectual Disability: Definition, Classification, and Systems of Supports. Intellect Dev Disabil. 2010;48:375–86, doi: 10.3337/jir.95:6256–61, doi: 10.1073/pnas.95.11.6256.

5. Fernandez F, Garner CC. Over-inhibition: a model for developmental disability. Trends Neurosci. 2007;30:497–503, doi: 10.1016/j.tins.2007.07.005.

6. NIH Common Fund Translational Research Overview (2006). Available from: http://nihroadmap.nih.gov

7. Bittles AH, Bower C, Hussain R, Glasson EJ. The four ages of Down syndrome. Eur J Public Health. 2007;17:221–5, doi: 10.1093/eurpub/ckl013.

8. Lott IT, DiGangi Jr. Cognitive deficits and associated neurological complications in individuals with Down’s syndrome. Lancet. 2010;376:23–33, doi: 10.1016/S0140-6736(10)60829-2.

9. Lejeune J, Gauthier M, Turpin R. Study of somatic chromosomes from mongoloid children. C R Hebd Seances Acad Sci. 1959;248:1721–2.

10. Kurtzberg J, Alston SE, Archana S, et al. Impaired short- and long-term memory in Ts65Dn mice, a model for Down syndrome. Neurosci Lett. 1995;195:143–6, doi: 10.1016/0304-3940(95)02152-6.

11. Lejeune J, Gauthier M, Turpin R. Study of somatic chromosomes from mongoloid children. C R Hebd Seances Acad Sci. 1959;248:1721–2.

12. Bucan M, Abel T. The mouse: genetics meets behaviour. Nat Rev Genet. 2003;4:585–94, doi: 10.1038/nrg1169.

13. Fernandez F, Garner CC. Over-inhibition: a model for developmental disability. Trends Neurosci. 2007;30:497–503, doi: 10.1016/j.tins.2007.07.005.

14. Shogren KA, Turnbull HR. Public policy and outcomes for persons with intellectual disability: extending and expanding the public policy framework of A/AIDD’s 11th Edition of Intellectual Disability: Definition, Classification, and Systems of Supports. Intellect Dev Disabil. 2010;48:375–86, doi: 10.3337/jir.95:6256–61, doi: 10.1073/pnas.95.11.6256.

15. Bucan M, Abel T. The mouse: genetics meets behaviour. Nat Rev Genet. 2003;4:585–94, doi: 10.1038/nrg1169.

16. Giralt S, Capra CM, Furlow DA, et al. A more human model of congenital muscular dystrophy. Cell. 1994;78:23–33.

17. Curtia G, Longo D, Biagini G, Jones RSG, Avoli M. Downregulation of tonic GABAergic inhibition in a mouse model of fragile X syndrome. Cereb Cortex. 2009;19:1515–20, doi: 10.1093/cercor/bhn159.

18. Dunn HG, MacLeod PM. Rett syndrome: review of biological abnormalities. Can J Neurol Sci. 2001;28:16–29.
38. Amir RE, Van den Veyer BI, Wan M, Tran CQ, Francke U, Zoghbi HY. Rett syndrome is caused by mutations in X-linked MECP2, encoding methyl-CpG-binding protein 2. Nat. Genet. 1999;23:185–8, doi: 10.1038/15610.

39. Guy J, Hendrich B, Holmes M, Martin JE, Bird A. A mouse MeCP2-null mutation causes neurological symptoms that mimic Rett syndrome. Nat Genet. 2001;27:322–6, doi: 10.1038/85906.

40. Chen RZ, Akbarian S, Tudor M, Jaenisch R. Deficiency of methyl-CpG binding protein-2 in CNS neurons results in a Rett-like phenotype in mice. Nat Genet. 2002;31:243–54, doi: 10.1038/ng762-02067-8.

41. Shahbazian M, Young J, Yuva-Paylor L, Spencer C, Antalffy B, Noebels J, et al. Mice with truncated MeCP2 recapitulate many Rett syndrome features and display hyperexcitability of histone H3. Neuron. 2002;35:243–54, doi: 10.1016/s0896-6273(02)00768-7.

42. LaSalle JM, Yasui DH. Evolving role of MeCP2 in Rett syndrome and autism. Epigenomics. 2009;1:119–30, doi: 10.2217/epi.09.19.

43. Campagnol M. An approach to the diagnosis of inherited metabolic disease. Arch Dis Child Educ Pract Ed. 2010;95:44, doi: 10.1136/adc.2008.151183.

44. Garcia-Cazorla A, Wolf NI, Serrano M, Moog U, Pérez-Dueñas B, Póp P, et al. Mental retardation and inborn errors of metabolism. J Inherit Metab Dis. 2009;32:597–608.

45. de Carvalho TM, dos Santos HP, dos Santos IC, Vargas PR, Pedrosa J. Newborn screening: a national public health programme in Brazil. J Inherit Metab Dis. 2007;30:615, doi: 10.1007/s10545-007-0650-7.

46. Boulton A, Baker G, Martin-Iverson M, editors. Animal Models of Intellectual Disability. CliftonNJ:Humana Press;1991, pp. 273–314.

47. Monteiro LTB, Carvalho J, Ferreira EM, Rice DC. Animal Models of Cognitive Impairment Produced by Developmental Lead Exposure. In: Levin ED, Buccafusco JJ, editors. Animal Models of Cognitive Impairment. CRC Press Taylor & Francis Group, LLC. Boca Raton, FL: 2006.

48. Jacobson J, Jacobsen SW. Prenatal exposure to polychlorinated biphenyls and attention at school age. J Pediatrics. 2003;143:780–8, doi: 10.1067/s0022-3476(03)00577-8.

49. Stewart P, Fitzgerald S, Reinhum J, Gump B, Lonky E, Darvill T, et al. Increased PCB exposure, the corpus callosum, and response inhibition. Environmental Health Perspect. 2003;111:1670–7, doi: 10.1289/ehp.6173.

50. Buxton D, Finlayson J. Experimental infection of pregnant sheep with Toxoplasma gondii: pathological and immunological observations on the placenta and foetus. J Comp Pathol. 1986;96:319–33, doi: 10.1016/S0022-3476(03)00572-6.

51. Dubey JP, Shen SK. Rat model of congenital toxoplasmosis. Infect. Immun. 1991;59:3301–3.

52. Breese GR, Criswell HE, Duncan DE, Mueller RA. A dopamine uptake inhibitor GBR-12909 induction of aberrant behaviors in animal models of dopamine dysfunction. J Int Neurosci. 2002;20:323–33, doi: 10.1016/S0736-5748(02)00584-0.

53. Breese GR, Knapp DJ, Criswell HE, Müller RA. A dopamine deficiency model of Lesch-Nyhan disease—the neonatal-6-OHDA-lesioned rat Brain Res Bull. 1990;25:477–84.

54. Fisher MH, Welker C, Waisman H. Generalised growth retardation in rats induced by maternal methylazoxymethanol acetate. Teratol. 1972;6:223–32, doi: 10.1002/tera.1420050212.

55. Johnston MV, Coyle JT. Histological and neurochemical effects of fetal treatment with methylylzoxomethanol on rat neocortex in adulthood Brain Res. 1979;170:135–55.

56. Verhees CV, Fernandez K, Dams MA, Haddad RK. Pervasive hyperactivity and long-term learning impairments in rats with induced microencephaly from prenatal exposure to methylylzoxomethanol. Developmental Brain Res. 1984;13:1–10, doi: 10.1016/0165-3806(84)90134-2.

57. Archer T, Hard E, Hansen S. Animal models of mental retardation. In: Boulton A, Baker G, Martin-Iverson M, editors. Animal Models of Psychiatry II. Neuromethods. CliftonNJ:Humana Press;1991, pp. 273–314.

58. Schroeder SR. Mental retardation and developmental disabilities influenced by environmental neurotoxic insults. Environ Health Perspect. 2000;108 Suppl 2:350S–3S. doi: 10.1289/ehp.00229.x.

59. Cook CS, Novotny AZ, Sulik KK. Fetal alcohol syndrome: eye malformations in a mouse model. Arch Ophthalmol. 1987;105:1576–81.

60. Buscher WS, Walsh DA, McEwen SE, Lipson AH. Some teratogenic properties of ethanol and acetaldehyde in implications for the study of fetal alcohol syndrome research. Ment Retard Dev Dis Res Rev. 1998;4:13–23, doi: 10.1002/1532-4428(1998)4:1<13::AID-MRDR5>3.0.CO;2-0.

61. Eskenazi B, Harley K, Bradman A, Weltzien E, Jewell NP, Barr DB, et al. Mental Retardation and inborn errors of metabolism. J Inherit Metab Dis. 2007;30:615, doi: 10.1007/s10545-007-0669-9.

62. Lanphear BP. Congressional Briefing in the American Association on Mental Retardation’s Environmental Health Initiative, Learning Disabilities Association of America’s Healthy Children Project Collaborative on Health and the Environment’s Learning and Developmental Disabilities Initiative May 10, 2005. Available from: http://www.aamr.org...
89. Roobey T, Raja TN, Moustojiannis AN. Animal models for the study of perinatal hypoxic-ischemic encephalopathy: a critical analysis. Early Hum Dev. 1997;47:115–46, doi: 10.1016/S0378-3798(96)01773-2.

90. Wagner BP, Nedelcu J, Martin E. Delayed posts ischemic hypothermia improves long-term behavioral outcome after cerebral hypoxia-ischemia in neonatal rats. Pediatr Res. 2002;51:354–60, doi: 10.1203/00006450-200203000-00015.

91. Lee JH, Lee YK, Ishikawa M, Koga K, Fukunaga M, Miyakoda G, et al. Cilostazol reduces brain lesion induced by focal cerebral ischemia in rats—an MRI study. Brain Res. 2003;994:91–8, doi: 10.1016/j.brainres.2003.09.021.

92. Wang Y, Cheung PT, Shen GX, Wu EX, Cao G, Bart I, et al. Hypoxic-ischemic brain injury in the neonatal rat model: relationship between lesion size at early MR imaging and irreversible infarction. AJNR Am J Neuroradiol. 2006;27:51–4.

93. Holmes GL, Khazipov R, Ben-Ari Y. New concepts in neonatal seizures. Brain Res. 2002;925:27–35, doi: 10.1016/S0006-8993(02)02033-9.

94. Simon NP. Long-term neurodevelopmental outcome of asphyxiated newborns. Clin Perinatol. 1999;26:767–78.

95. Yager RGY. Pathophysiology of perinatal brain damage. Brain Res Rev. 1999;30:107–34, doi: 10.1016/S0165-0173(99)00009-0.

96. Rice JE, Vannucci RC, Brierly JB. The influence of immaturity on hypoxic–ischemic brain damage in the rat. Ann Neurol. 1981;9:131–41, doi: 10.1002/ana.410090206.

97. Nunez JL, Alt JJ, McCarthy MM. A new model for prenatal brain damage. I. GABAA receptor activation induces cell death in developing rat hippocampus. Exp Neurol. 2003;181:258–69, doi: 10.1016/S0014-4886(03)00053-0.

98. Nunez JL, Alt JJ, McCarthy MM. A novel model for prenatal brain damage II. Long-term deficits in hippocampal cell number and hippocampal dependent behavior following GABA receptors activation. Exp Neurol. 2003;181:270–80, doi: 10.1016/S0014-4886(03)00051-2.

99. Bittigau P, Sifrininger M, Felderhoff-Mueser U, Hansen HH, Ikonomidou C. Neuropathological and biochemical features of traumatic injury in the developing brain. Neurotox Res. 2003;5:475–490, doi: 10.1007/s10068-003-03318.

100. Bittigau P, Pohl D, Sifrininger M, Shimizu H, Ikeda M, Ishimaru M, et al. Modeling pediatric head trauma: mechanisms of degeneration and potential strategies for neuroprotection. Restor Neurol Neurosci. 1998;13:11–23.

101. Garzon E. Estado de Mal epileptico. J Epilepsy Clin Neurophysiol. 2008;14Suppl:27–11.

102. Meldrum BS, Horton RW. Physiology of status epilepticus in primates. Arch Neurol. 1973;28:1–9.

103. Nevander G, Ingvar M, Auer R, Siesjo BK. Status epilepticus in well-oxygenated rats causes neuronal necrosis. Ann Neurol. 1985;18:281–90, doi: 10.1002/ana.410180303.

104. Santos NF, Marques RH, Correia L, Sinigaglia-Coimbra R, Calderazzo L, Sanabria ER, et al. Multiple pilocarpine-induced status epilepticus in developing rats: a long-term behavioral and electrophysiological study. Epilepsia. 2000;41Suppl 6:557–63, doi: 10.1111/j.1528-1157.2000.tb0358.x.

105. Milgram NW, Isen DA, Mandel D, Palantzas H, Pepkowski MJ. Deficits in spontaneous behavior and cognitive functions following systemic administration of kainic acid. Neurotoxicology. 1988;9:611–24.

106. Wolvetsang EW, Bradfield OM, Tynms M, Zunavsek S, Hatzistavrou V, Kola I, et al. The chromosome 21 transcription factor ETS2 transactivates the beta-APP promoter: implications for Down syndrome. Biochim Biophys Acta. 2003;1628:105–10.

107. Netzer WJ, Powell C, Nong Y, Blundell J, Wong L, Dufi K, et al. Lowering beta-amyloid levels rescues learning and memory in a Down syndrome mouse model. PLoS One. 2010;5:e10943, doi: 10.1371/journal.pone.0010943.

108. Akesson EC, Lamber JP, Narayanswami S, Gardiner K, Bechtel LJ, Davison MT. T65Dn-localization of the translocation breakpoint and trisomic gene content in a mouse model for Down syndrome. Cytogenet Cell Genet. 2001;93:270–6, doi: 10.1159/000056997.

109. Granholm AC, Sanders LA, Cnric LS. Loss of cholinergic phenotype in basal forebrain coincides with cognitive decline in a mouse model of Down’s syndrome. Exp Neurol. 2000;161:647–63, doi: 10.1006/exnr.1999.7289.

110. Holtzman DM, Santucci D, Kilbridge J, Chua-Couzens J, Fontana DJ, Daniels SE, et al. Developmental abnormalities and age-related neurodegeneration in a mouse model of Down syndrome. Proc Natl Acad Sci USA. 1996;93:13333–8, doi: 10.1073/pnas.93.23.13333.

111. Glenner GG, Wong CW. Alzheimer’s disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein. Biochem Biophys Res Commn. 1984;120:3885, doi: 10.1016/S0006-291X(84)80190-4.

112. Coote A, Chartier-Harlin MC, Mullan M, Brown J, Crawford F, Fidani L, et al. Segregation of a missense mutation in the amyloid precursor protein gene with familial Alzheimer’s disease. Nature. 1991;349:704–6, doi: 10.1038/349704a0.

113. Hsiao K, Chapman P, Nilsen S, Eckman C, Harigaya Y, Younkin S, et al. Correlative memory deficits, A beta elevation, and amyloid plaques in transgenic mice. Science. 1996;274:99, doi: 10.1126/science.274.5284.99.

114. Morris RG, Garrud P, Rawlins JN, O’Keefe J. Place navigation impaired in rats with hippocampal lesions. Nature. 1982;297:681, doi: 10.1038/349704a0.

115. Ohno M, Sametksy EA, Younkin LH, Oakley H, Younkin SG, Citron M, et al. BACE1 deficiency rescues memory deficits and cholinergic dysfunction in a mouse model of Alzheimer’s disease. Neuron. 2003;30:821–7, doi: 10.1016/S0896-6273(03)00810-9.

116. Terry AV Jr, Buccafusco JJ. The cholinergic hypothesis of age and Alzheimer’s disease-related cognitive deficits: recent challenges and their implications for novel drug development. J Pharmacol Exp Ther. 2003;306:821–7, doi: 10.1124/jpet.102.041616.

117. Zhang L. Cholinergic Receptors Knockout Mice.Chapter 11. In: Levin ED, Buccafusco JJ, editors. Animal Models of Cognitive Impairment. Boca Raton: CRC Press; 2006.

118. Volpicealli-Daley LA, Duyksen EG, Lockridge O, Levey AI. Altered hippocampal muscarinic receptors in acetylcholinesterase-deficient mice. Ann Neurol. 2003;53:788–96, doi: 10.1002/ana.10589.