Minireview

Vascular disrupting agents in clinical development

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Growth of human tumours depends on the supply of oxygen and nutrients via the surrounding vasculature. Therefore tumour vasculature is an attractive target for anticancer therapy. Apart from angiogenesis inhibitors that compromise the formation of new blood vessels, a second class of specific anticancer drugs has been developed. These so-called vascular disrupting agents (VDAs) target the established tumour vasculature and cause an acute and pronounced shutdown of blood vessels resulting in an almost complete stop of blood flow, ultimately leading to selective tumour necrosis. As a number of VDAs are now being tested in clinical studies, we will discuss their mechanism of action and the results obtained in preclinical studies. Also data from clinical studies will be reviewed and some considerations with regard to the future development are given.

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Tumour-related angiogenesis is essential for tumour growth and metastases formation. It is a complex process in which vascular endothelial growth factor (VEGF) produced by tumour cells plays a predominant role (Ferrara et al., 2003). Binding of VEGF to the transmembrane endothelial VEGF tyrosine kinase receptors (VEGFR) type 1 or 2 initiates a cascade of intracellular signaling pathways resulting in endothelial cell proliferation and the formation of new blood vessels. Apart from VEGF, basic fibroblast growth factor, platelet-derived growth factor, interleukin-8 and insulin-like growth factor are proangiogenic factors. Natural antiangiogenic factors produced by tumour and host cells are tumour necrosis factor alpha (TNF-α), serotonin (5-HT), nitric oxide (NO), thrombospondin, angiostatin and endostatin.

Inhibiting angiogenesis has become a challenge in the development of a totally new class of anticancer drugs as was already acknowledged in 1971 by Folkman (1971). Angiogenesis inhibitors can be divided into two groups, monoclonal antibodies (Moabs) and small molecule tyrosine kinase inhibitors (TKIs). Bevacizumab is a humanised Moab targeting VEGF, which has shown clinical activity in combination with cytotoxic chemotherapy in metastatic colorectal cancer, non-small cell lung cancer (NSCLC) and breast cancer (Hurwitz et al., 2004; Ramaswamy et al., 2006; Sandler et al., 2006). As single agent bevacizumab has demonstrated activity in metastatic renal cell carcinoma (Yang et al., 2003).

Apart from Moabs, a large number of small-molecule VEGFR TKIs have been explored in clinical studies. Results in randomised studies in renal cell carcinoma with the broadspectrum TKIs sunitinib and sorafenib have resulted in their regulatory approval for this disease (Escudier et al. 2007; Motzer et al., 2007). Most other TKIs so far have either only been tested in smaller phase I and II studies, or have failed to show meaningful effects in larger randomised phase III studies (Koehne et al., 2006). Theoretically, it is conceivable that angiogenesis inhibitors will exert optimal activity in a situation of minimal residual disease with high-angiogenic potency such as could be the case in the adjuvant setting.

Although inhibiting angiogenesis thus seems to be successful in various conditions, an urgent need for more optimal treatment options in metastatic disease exists.

For example, what to do with the already established tumour-related vasculature? Vascular targeting strategies can be divided into two different approaches: as mentioned above an antiangiogenic approach, but apart from that a so-called vascular disrupting approach has emerged (Siemann et al., 2005). Vascular disrupting agents (VDAs) target endothelial cells and pericytes of the already established tumour vasculature. Although this approach looks very interesting from a theoretical point of view, one of the critical issues one could raise is that of tumour specificity: do VDAs selectively target tumour-related endothelium or is there a more general vascular targeting effect with a risk of subsequent ischaemic complications? In the following sections we will discuss in more detail the hypothesised mechanisms of action of VDAs and will review results of preclinical and clinical studies performed so far. Finally, we will give some thoughts on were to go with VDAs in future studies.

VDAs AND THEIR TARGET

Indirect killing of tumours by compromising their vascularisation is a potentially attractive anticancer treatment approach. On the one hand drug resistance is not likely to appear because the targeted endothelial cell has much greater genetic stability than neoplastic cells (Kerbel and Folkman, 2002). Also drug delivery is likely to be without compromise, as the endothelium of the tumour
vascularity is easily accessible. Lastly, and theoretically, vascular shutdown is likely to result in a massive ‘downstream’ tumour cell killing.

Selective vascular shutdown suggests a structural difference in endothelium of tumour vessels compared to that of normal vessels. Indeed, tumour vasculature is, among others, marked by a high rate of endothelial cell proliferation, the absence of pericytes, abnormalities in the basement membrane and often an increased vascular permeability. Structurally, disorganised, tortuous, thin-walled vessels are seen that lack smooth muscle and pericyte coats and innervation (Figure 1) (Kakolyris et al., 2000; Konerding et al., 2001). Blood flow frequently is sluggish and at times might be stationary or even in a reversed direction (Tozer et al., 1990). Vessel diameters are irregular and lengths between branches are long, resulting in a high resistance to blood flow. A small decrease in perfusion pressure, which has little effect in normal tissue, therefore can be catastrophic to tumours.

Finally, endothelial cells are highly dependent on tubulin cytoskeleton for their motility, invasion, attachment, alignment and proliferation (Denekamp, 1982).

Most VDAs induce changes in endothelial cell shape by disruption of the cytoskeleton and cell-to-cell junctions. This results in increased permeability to proteins and an increased interstitial fluid pressure, which might be sufficient to reduce vessel diameter. Plasma leakage also leads to increased blood viscosity resulting in decreased blood flow and roulaux formation. Another factor contributing to the vascular shutdown is the activation of platelets through contact with basement membrane components, which are exposed. All together this cascade of events results in vascular shutdown more selectively in tumour endothelium than normal endothelium. As stated previously, it is suggested that the inhibition of blood flow and the subsequent compromised supply of oxygen and nutrients will induce necrosis of many tumour cells downstream.

Compared to the antiangiogenic approach of both TKI and Moab, the vascular disrupting approach therefore seems to be cytotoxic rather than cytostatic. However, in preclinical models it has been observed that following exposure to a VDA, only the centre of a tumour becomes necrotic, with a viable rim (Figure 2) remaining in the periphery. This rim of viable tumour cells presumably survives because it derives nutritional support (most likely via diffusion) from adjacent normal blood vessels that are typically less responsive to VDAs.

To demonstrate biological activity of VDAs in preclinical and clinical studies, noninvasive techniques such as dynamic contrast-enhanced magnetic resonance imaging (DCE–MRI) and positron emission tomography scans (PET) have been explored. This type of imaging can demonstrate changes in tumour perfusion and tumour viability (Beauregard et al., 1998; Galbraith et al., 2003). Although data to date look very promising, one must realise that both techniques are not yet validated to predict antitumour activity or real patient benefit in the clinical situation.

VDAs IN CLINICAL DEVELOPMENT

Vascular disrupting agents have been divided into two types, small molecule and ligand directed VDAs. We will focus on small molecule VDAs because they are in a more advanced stage of clinical development. Small molecule VDAs can be divided into two groups; the tubulin-binding agents and the flavonoids. Their mechanism of action is somewhat different as will be discussed. Tubulin-binding agents are combretastatin, AVE8062, ZD6126, ABT-571, MN-029 and the dolatastatin derivative TAZT-1027. Of the flavonoids only 5,6-dimethylxanthenone-4-acetic acid (DMXAA) will be discussed.

I. TUBULIN-BINDING AGENTS

These agents work by acting at the colchicines-binding site of the β-subunit of endothelial tubulin, resulting in depolymerisation of microtubules and disorganisation of actin and tubulin. Disruption of the endothelial cytoskeleton results in conformational changes leading to loss of blood flow. In addition to this, a recent study showed that the typical microtubule-destabilising agent combretastatin A4 phosphate (CA4P) also selectively disrupts the VE-cadherin/β-catenin complex interfering with cell–cell contact (Vincent et al., 2005). Loss of this cell–cell contact increases vascular permeability leading to increased interstitial pressure and additional loss of blood flow. In addition to these effects, the already mentioned loss of cell–cell contact results in the exposure of the already often abnormal basement membrane, which in turn can result in the induction of the coagulation cascade with subsequent thrombus formation. Tumour-related endothelial cells are much more sensitive to the activity of tubulin-binding agents than normal endothelial cells (Chaplin and Dougherty, 1999).

Combretastatin A4 phosphate

Combretastatin A4 phosphate is a water-soluble prodrg of combretastatin A4 (CA4). Following administration, CA4P is rapidly cleaved to CA4 and binds tubulin at or close to the colchincines-binding site (McGown and Fox, 1989). One of the first in vivo studies showed rapid, extensive and irreversible vascular shutdown and haemorrhaghic necrosis following a single dose of CA4P. A pronounced and sustained reduction in functional...
vascular volume was observed following drug administration at a dose much lower than the maximum-tolerated dose (MTD) (Dark et al., 1997). Histologically as well as DCE–MRI studies in preclinical models show that the antivascular effects of CA4 are restricted to the core of the tumour, leaving viable tumour cells at the periphery (Beauvergaret al., 1998). Combretastatin A4 shows different activity in normal and tumour endothelium in preclinical models; Tozer et al. (1999) showed a 100-fold decrease in blood flow in p22 carcinosarcomas with a much smaller reduction in blood flow in the spleen, skeletal muscle and brain. No significant reduction in blood flow was seen in heart, kidney and intestine.

Three phase-I trials of CA4P in humans have been published (Table 1). In the first study by Rustin et al. (2003a) CA4P was given weekly for 3 weeks followed by a week gap. Thirty-four patients with advanced solid tumours received 167 infusions. Up to 40 mg m⁻², the only drug-related toxicity was tumour pain in 35%. Tumour pain was not considered a dose-limiting toxicity (DLT) because it could be controlled by analgesics. Tumour viability and tumour blood flow were assessed by PET and DCE–MRI (Galbraith et al., 2003). Dose-limiting toxicity were fatal ischaemia in previously irradiated bowel, vasovagal syncope, motor neuropathy and reversible ataxia. Other side-effects were hypertension (35%), hypotension (30%), tachycardia (53%), bradycardia (24%), nausea (21%), fatigue (23%), visual disturbance (9%) and dyspnoea (6%). The drug was generally well tolerated and no myelosuppression, alopecia and mucositis were seen. One partial response was seen (metastatic adenocortical carcinoma). The recommended phase-II dose of 52–68 mg m⁻² was based upon clinical tolerability and the assessment of biological activity by means of PET and DCE–MRI analysis.

In a second phase-I study, Stevenson et al. (2003) used a daily infusion for 5 days every 3 weeks. Thirty-seven patients received 133 cycles. Dose-limiting toxicities were tumour pain, reversible sensorimotor neuropathy, syncope and dyspnoea. No cardiotoxicity or electrocardiographic changes were seen. One patient with metastatic sarcoma had a partial response, and 14 patients showed stable disease. The recommended phase-II dose was 52 mg m⁻².

Dowlati et al. (2002) used a once every 3 weeks schedule. Twenty-five patients received 107 cycles. Dose-limiting toxicities were cardiac ischaemia and dyspnoea in two patients with pre-existing cardiovascular disease. A significant decline in gradient peak tumour blood flow by DCE–MRI was observed in six patients treated at 60 mg m⁻². One complete response was observed in a patient with anaplastic thyroid cancer, whereas two patients experienced freedom from disease progression lasting more than 12 months. Dosages up to 60 mg m⁻² as a 10-min infusion defined the upper boundary of the MTD. Cooney et al. (2004) determined the cardiovascular safety profile of CA4P in the same patient cohort. They observed asymptomatic QTc prolongation as DLT. Apart from this, two patients had an acute coronary syndrome within 24 h after the infusion of CA4P.

All mentioned studies used a different dosing schedule (weekly, 3-weekly, daily for 5 days every 3 weeks) and showed that CA4P was safe, well tolerated and lacking haematologic toxicity. In all studies MTDs of 50–60 mg m⁻² were set with consistent indications of antivascular effects observed by either DCE–MRI or PET. Currently CA4P is further explored as single agent in phase-II studies in patients with advanced anaplastic thyroid cancer. Apart from single-agent approaches, CA4P has been studied in combination with carboplatin. Combretastatin A4 was given 3-weekly (27–36 mg m⁻²) 60 min after carboplatin (AUC 4–5). Dose-limiting toxicity was trombocytopenia (Bilenker et al., 2005).

In another ongoing study, induction chemotherapy using doxorubicin and cisplatin is followed by CA4P and radiation therapy in patients with newly diagnosed advanced anaplastic thyroid cancer.

Finally, CA4P is currently being explored in combination with carboplatin and paclitaxel in patients with advanced solid tumours.

**Table 1** Currently published phase-I studies

| Drug     | Company       | Treatment schedule | Dose range       | DLT*              | References                  |
|----------|---------------|--------------------|------------------|-------------------|-----------------------------|
| CA4P     | Oxigen        | d 1, 8, 15 q4w     | 5–114 mg m⁻² iv. | Bowel ischaemia,  | Rustin et al                  |
|          |               | d 1–5 q3w         | 6–75 mg m⁻² iv.  | tumour pain, vagal | Stevenson et al               |
|          |               | d 1 q3w           | 18–90 mg m⁻² iv. | syncope, cardiac   | Dowlati and Cooney et al      |
|          | Sanofi Aventis| d 1, 8, 15 q4     | 4.5–40 mg m⁻² iv. | ischaemia, dyspnoea| Tolcher et al                  |
| AVE8062  |               |                   |                  | Hypoxia, transient  |                            |
| ZD6126   | Astra Zeneca  | weekly             | 5–28 mg m⁻² iv.  | myocardial ischaemia,  | Beerepoort et al               |
|          |               | d 1 q3w           | 5–40 mg m⁻² iv.  | ischaemia         | LoRusso et al                 |
|          |               | d 1 q3w           | 5–112 mg m⁻² iv. | Hypotension, transient  | Gadgeel et al                |
|          |               |                   |                  | neuromotor symptoms |                            |
| ABT-751  | Abbott        | q.d. 7 days q3w   | 200–300 mg po    | Illeus, constipation,  | Hande et al                   |
|          |               | b.i.d. 7 days q3w | 125–175 mg po    | abdominal pain, neuropathy,  |                            |
| MN-029   | Medicinova    | 3-weekly           | 4–180 mg m⁻² iv. | Fatigue           | Ricart et al                  |
| TZT-1027 | Daichi        | d 1, 8 q3w        | 1.35–2.7 mg m⁻² | Reversible cardiac  | de Jonge et al                |
|          | Pharmaceuticals| d 1, 8 q3w + carb | 1.6–2.0 mg m⁻²  | ischaemia         | Greytakte et al               |
|          |               | o AUC 4–5          | 1.35–3.0 mg m⁻²  | Neuropathy        | Schoeffl et al                |
| DMXAA    | Antisoma      | Weekly             | 6–4900 mg m⁻² iv.| Reversible urinary  | Rustin et al                  |
|          |               | d 1 q3w           | 6–4900 mg m⁻² iv.| incontinence, visual | Jameson et al                 |

DLT = dose-limiting toxicity; iv. = intravenously. Ref. = References.
As in vivo studies have shown synergistic activity of AVE8062 with oxaliplatin and docetaxel (Demers et al, 2006), clinical studies exploring these combinations are currently also ongoing.

**ZD6126**

In preclinical models this agent demonstrated significant anti-tumour activity. Stasis of blood flow was seen at doses 1/8–1/16 of the MTD and occurred specifically in tumour tissue (Davis et al, 2002). Two phase-I studies, in which ZD6126 was given thrice weekly, have been presented (Gadgeel et al, 2002; LoRusso et al, 2002) (Table 1). One patient showed asymptomatic, reversible cardiac ischaemia with subsequent demonstration of coronary artery disease. Maximum-tolerated dose was set at 112 mg m⁻², whereas biological activity indicated by a sustained decrease in tumour blood flow measured by DCE-MRI occurred at doses above 80 mg m⁻². A third phase-I study has recently been published (Beerepoot et al, 2006). Here, ZD6126 was given weekly to 32 patients. Dose-limiting toxicity consisted of myocardial infarction and was observed at a dose of 10 mg m⁻² in one patient. This patient was found to have a history of ischaemic heart disease. Two patients treated at 28 mg m⁻² experienced DLT, one each with pulmonary embolus (disease-related) and asymptomatic decrease in left ventricular ejection fraction. Maximum-tolerated dose was set at 20 mg m⁻². In all three studies ZD6126 was well tolerated and only showed mild side effects such as anaemia, nausea, vomiting and constipation. So far no objective tumour responses have been observed. Currently, ZD6126 is being explored in metastatic renal-cell carcinoma.

**ABT-751**

ABT-751 is a sulphonamide molecule that can be given orally, and has shown significant antitumour activity in a variety of tumour models (Ozawa et al, 2001). In a phase-I study, 39 patients with solid tumours were given ABT-751 once or twice daily for 7 days every 3 weeks (Table 1) (Hande et al, 2006). Dose-limiting toxicities were ileus and neuropathy at 300 mg daily. In the twice daily schedule, grade 3 ileus, constipation, abdominal pain and fatigue were observed. One minor response and four patients with stable disease lasting for 6 months were observed. The MTD and recommended phase-II doses for ABT-751 were 250 mg daily and 150 mg twice daily for 7 days every 3 weeks. Phase I/II studies are currently ongoing, evaluating the safety and efficacy of ABT-751 in combination with pemetrexed or docetaxel in patients with NSCLC.

**MN-029**

One preclinical study was published using a rodent KHT sarcoma model. After intraperitoneal injection of 100 mg kg⁻¹ a significant reduction in the functional vessel number was seen. Treatment with MN-029 resulted in dose-dependent tumour cell killing. Effects were enhanced by combining the agent with radiation and cisplatin chemotherapy (Shi and Siemann, 2003). Only one ongoing phase-I study has been reported so far (Table 1). In this study, 28 patients with various solid tumours received 110 cycles. Dose-limiting toxicity consisted of reversible cardiac ischaemia at 180 mg m⁻². Seven patients had stable disease after 3 cycles. Dynamic contrast-enhanced magnetic resonance imaging analysis indicated significant dose-dependent reductions in tumour blood flow. Accrual at 225 mg m⁻² continues (Ricart et al, 2006).

**TZT-1027**

TZT-1027 is a synthetic derivative of dolastatin-10 with cytotoxic and antivascular activity. Three different treatment schedules have been explored in phase-I trials (Table 1). Schoffski et al (2004) performed a phase-I study in which 21 patients received TZT-1027 infusions at 3-weekly intervals. Dose-limiting toxicities were neutropenia, fatigue and short lasting peripheral neuropathy. Anorexia, alopecia and constipation were also seen. The recommended phase-II dose was set at 2.7 mg m⁻². A second phase-I study, exploring day 1 and 8 every 3 weeks administration in 17 patients showed comparable DLTs as well as pain in the infusion arm lasting 1–2 days at a dose of 2.7 mg m⁻² (DeJonge et al, 2005). Other side effects included nausea, fatigue, vomiting and diarrhoea. One patient with metastatic liposarcoma had an ongoing partial response for more than 54 weeks. The recommended dose for phase-II studies of TZT-1027 in this study was set at 2.4 mg m⁻². A third phase-I study explored the combination of TZT-1027 with carboplatin in 14 patients (Greystone et al, 2006). Dose-limiting toxicity consisted of neutropenia and grade 3 ileus. Other toxicities were comparable to those mentioned above. No pharmacokinetic interaction between carboplatin and TZT-1027 was observed. One patient with metastatic adenocarcinoma of the pancreas showed a partial response lasting 181 days. The recommended phase-II doses of TZT-1027 in combination with carboplatin AUC 5 was set at 1.6 mg m⁻².

**II. FLAVONOIDS**

**DMXAA**

5,6-Dimethylxanthone-4-acetic acid is an active analogue of flavone acetic acid causing DNA damage to endothelial cells that induces apoptosis in preclinical models (Corbett et al, 1986). In response to vascular damage 5-HT is released by platelets that further enhances the vascular effects (Baguley et al, 1997). Although the exact mechanism of action of DMXAA is unknown, its activity involves pathways leading to upregulation of the nuclear transcription factor NfκB, which leads to production of TNF-α and other cytokines (Ching et al, 2002). Tumour blood flow decreases and 5-HT levels increase. In addition, NO is produced in response to DMXAA, improving blood flow and vascular permeability, increasing the effects of TNF-α and 5-HT (Thomsen et al, 1991). How these forces oppose each other is unknown. Two phase-I trials have been published so far (Table 1). Rustin et al (2003b) treated 46 patients with weekly infusions and documented rapidly reversible DLTs like urinary incontinence, visual disturbance (blurring, colour disturbance and photophobia) and anxiety. No tumour pain was seen. Maximum-tolerated dose was set at 3700 mg m⁻². At dose levels of 650 mg m⁻² and above a dose-dependent increase of 5-HT concentrations in plasma was seen. There was one unconfirmed partial response at 1300 mg m⁻². In a second study, in which 63 patients received 3-weekly infusions, comparable DLTs were observed with additional confusion, slurred speech, tremor and possible left ventricular failure (Jameson et al, 2003). Asymptomatic transient QTc-prolongation was seen in 13 patients at high doses. One partial response was seen in a patient with cervical carcinoma. Maximum-tolerated dose was set at 3700 mg m⁻². Two randomised phase-II studies combining DMXAA with conventional chemotherapeutics have recently been published. Gabra, (2006) randomised 55 patients with recurrent ovarian cancer to receive paclitaxel (175 mg m⁻²), carboplatin (AUC 6) and DMXAA (1200 mg m⁻²). Preliminary data revealed no additional toxicity owing to the addition of DMXAA. Efficacy assessments are pending. In 78 patients with NSCLC, McKeeage (2006) also found no additional toxicity when carboplatin and paclitaxel were combined with DMXAA. Initial response data suggest additional benefit from triple treatment compared to conventional therapy Currently, the efficacy and safety of DMXAA in combination with docetaxel is assessed in a phase-II study in patients with hormone refractory metastatic prostate cancer.
FUTURE DEVELOPMENTS

Vascular disrupting agents are a new class of antivascular anticancer agents that are currently undergoing clinical studies. At this moment, mainly (ongoing) phase-I studies have been presented, although some compounds have already entered phase-II testing either as single agent or in combination with chemotherapeutics. Therefore, the real (added) value in terms of patient benefit cannot be fully assessed yet.

What distinguishes VDAs from other vascular targeting agents, how can we optimally assess their biological and clinical activity and how should these agents be taken forward?

When assessing the toxicity pattern observed so far in the various clinical studies described, it is obvious that with regard to mechanism of action tumour specificity is most likely to be of critical importance. Vascular disrupting agents disrupt the established abnormal tumour vasculature by targeting the immature dysmorphic endothelial cells. As mentioned earlier tumour endothelium is more vulnerable to the activity of VDAs, and therefore in the end selective tumour vascular shutdown is likely to occur. However, based upon the pattern of side effects observed in clinical studies, normal vascular endothelium seems to be affected by VDAs as well. Cardiac ischaemia and cardiac arrhythmias as well as reversible neurologic complications (the latter not with all agents) seem to underscore this issue and probably will remain dose limiting in future studies. Of critical importance therefore will be the assessment of biological activity at doses that can be administered safely. This will probably mean that in the design of early clinical studies the concept of looking for MTD will have to be replaced by the concept of looking for optimal biological dose, hereby assuming that the therapeutic window of these agents will permit us to do so. At this moment the probable optimal way to assess biologic or antivascular activity of VDAs is by repeated dynamic scanning, and therefore the role of DCE–MRI and PET analysis must be validated further. Vascular shutdown and decreased tumour blood flow as an indication of biological activity have meanwhile been demonstrated by DCE–MRI and/or PET analysis, whereas the occurrence of tumour pain following administration of several compounds also can be considered as an indication of biological and perhaps clinical activity. Whether biological activity will result in tumour size reduction and meaningful patient benefit has to be evaluated further in phase II and randomised phase-III studies. The assessment of cardiac and neurological complications that can occur in these studies necessitates optimal communication between oncologists and other specialists, and this will also be important when patients willing to be enrolled in studies are being screened (Heeckeren van, 2006). This is, as we think, a nice example of the tremendous practical consequences the introduction of a new class of anticancer drugs could and even should have.

When looking at the observed biological effects induced by VDAs, the induction of central tumour necrosis whereas leaving a viable rim at the periphery seems to be a consistent finding. Probably, this means that there is a rationale to combine VDAs with other treatment strategies. Many theoretical combinations can be thought of, and combining VDAs with conventional cytotoxic treatment is already being pursued to a rather large extent. Apart from this, the addition of an angiogenesis inhibiting agent following VDA administration conceivably could induce ‘synergistic’ antiangiogenic activity leading to a complete growth inhibition and a subsequent state of dormancy of the ‘centrally killed’ tumour mass. This observation has meanwhile indeed been made in preclinical models (Siemann and Shi, 2004). In addition, the combination of low-dose chemotherapy following the administration of a VDA could also be considered. Here one could think of the concept of metronomic chemotherapy, whereby the administration of a cytotoxic drug could also be considered.

Exploring a combination of VDAs and epidermal growth factor receptor (EGFR) inhibitors theoretically could also be an interesting approach; here one could speculate that tumour cells in the viable rim will become apoptotic and die when being deprived of their growth-stimulating factors such as EGF.

Effectiveness of combination therapies often depends on the sequence of administration. First increasing the vascular permeability (within minutes) allowing accumulation of a cytotoxic agent in the tumour and then inducing a shutdown of blood flow (within hours) could probably kill large amounts of tumour cells. However, as certain chemotherapeutics have their own vascular side effects (arterial thrombosis following administration of cisplatin), the alternative sequence of first administering the VDA followed by the administration of a cytotoxic drug could also be considered. Irrespective of the sequence administered, these combination therapies should be carefully chosen and closely monitored.

From a theoretical perspective, radiation therapy probably must precede VDA administration because optimal blood flow and oxygen radical formation might be necessary for an optimal synergistic effect. Preclinical models have meanwhile shown such a synergistic effect. Finally, and considered as a very specific example, in tumours of the extremities, one could think of isolated limb perfusion with VDAs after clamping; Biologically active concentrations of VDAs could probably be achieved with only limited systemic exposure, thus leading to optimal and selective tumour cell killing while preserving the heart, central nervous system and other potential organs at risk. Monitoring antivascular effects with either angiography, DCE–MRI or PET would be challenging.

In conclusion, VDAs are a new and promising class of targeted anticancer agents. Although their safety currently is the major focus of research, results that will show anticancer activity are likely to follow soon.

Their unique mechanism of action merits thorough and extensive exploration, both as single agent as well as in combination with other treatment modalities. If these studies are performed adequately, with a close observation of toxicity, it is to be expected that in the years to come a clear picture of their role in anticancer treatment can be established.

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