Distribution and phylogeography of the genus *Mattirolomyces* with a focus on the Asian *M. terfezioides* haplotypes

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ABSTRACT

*Mattirolomyces* is an edible commercial sequestrate genus that is globally distributed. From the five described taxa of this genus, *Mattirolomyces terfezioides* is the most common species in Asia. Our recent attempts to locate *M. terfezioides* outside its current distribution area in China documented its first records in areas of poplar trees with the lowest known temperature and precipitation averages ever recorded for this species. This peculiar ecology was not reflected on the species-morphological features nor on its phylogenetic position in the genus. The first attempt to apply the phylogenetic network approach to *Mattirolomyces* revealed its geographic origin in the Asian-Pacific areas prior to frequent long-distance migration events. Based on data from recent study areas, we found that the collections from Inner Mongolia and the Shanxi province were similar to European collections. Asian haplotypes were less distant from the outgroup comparing to collections from Europe, supporting the hypothesis that *M. terfezioides* was originated from this Chinese area and was subsequently transported to Europe. Exploring *M. terfezioides* ecology and its mycorrhiza potential to grow in association with poplars would be of great importance for planning cultivation projects of this valuable desert truffle species in Central and Eastern China, a currently underexploited economic sector that deserves further ecological and *M. terfezioides* mycorrhizal synthesis investigations.

INTRODUCTION

The genus *Mattirolomyces* (Tuberaceae, Pezizales) was called the “Mattirolo fungus” by Fisher in 1938 after Mattirolo which was the first to describe the type species of this genus. This last species Mattirolo named in 1887 as *Choiromyces terfezioides* (Fischer, 1938), however, Moreno, Alvarado & Manjón (2012) confirmed later its separate generic position from the genus Choiromyces. The type specimen was firstly collected from clay agricultural soils in a non-typical ecological location in Piemonte, Northern Italy, and was considered...
at that time as a potential symbiotic partner of *Prunus avium* (*Mattirolo, 1887*). The *Mattirolomyces* taxon belongs to the ascomycetous desert truffles. All known species of this genus form sequestrate to hypogeous sporocarps (*Fischer, 1938*). Currently, five species in the genus *Mattirolomyces* have been shown to have a wide geographical distribution, commonly collected from areas with low and variable average rainfall and high summer temperatures (*Kagan-Zur et al., 2014*). The type species of the genus *Mattirolomyces terfezioides* (Mattir.) E. Fisch was recorded in Europe and Asia (*Fischer, 1938*); *Mattirolomyces spinosus* (Harkn.) Kovács, Trappe & Alsheikh was collected in North America and Pakistan (*Kovács et al., 2011*); *Mattirolomyces mulpu* Kovács, Trappe & Claridge was reported from Australia (*Trappe, Kovács & Claridge, 2010*); *Mattirolomyces austroafricanus* (Marasas & Trappe) Kovács was found from South Africa (*Trappe, Kovács & Claridge, 2010*); and *Mattirolomyces mexicanus* Kovács, Trappe & Claridge was described in Mexico (*Kovács et al., 2011*). The distribution pattern of these five species suggests a wide geographical (presumably global) range of the genus (*Kagan-Zur et al., 2014*) with most records coming from Eastern and Southeastern Europe (*Glejdura & Kunca, 2012; Kagan-Zur et al., 2014; Assyov & Slavova, 2016*).

The desert regions of the Southern Hemisphere have low and variable average rainfall and high summer temperatures. *Mattirolomyces* spp. and other desert truffles species have a long history of regular hunt and harvest for human consumption since prehistoric times (*Trappe, Kovács & Claridge, 2010*). However, *M. terfezioides* is rarely recognized as a valuable commercial species in Europe and Asia (*Boa, 2004*). *Mattirolomyces* spp. sporocarps are traditionally collected, sold, and consumed under the name *Terfezia terfezioides* (Mattir.) Trappe (*Trappe, 1971*), a synonym of *Mattirolomyces terfezioides*. Despite their great values as mycorrhizal species and culinary delicacy, this species is not well-known in the Northern Hemisphere (*Kovács, Jakucs & Bagi, 2007*). Most available collections of *M. terfezioides* are from Hungary (Europe) and Northern China, namely Beijing, Hebei Province, and Shanxi Province. Most of the Chinese collections date back several decades, with the most recent dating to 1986 (*Wang, Liu & Sun, 2017*). The economic and culinary value of the Chinese *M. terfezioides* collections have not yet been evaluated. Furthermore, this species has been considered long-lost in China by many mycologists.

We were motivated by a recently discovered *M. terfezioides* collection from the desert areas of Inner Mongolia, China to revive the study of the Chinese genus, characterize the current ecological span of the species, and prepare a morphology-based description of the Chinese collection. Due to a low number of records and available nucleotide sequences, no phylogeographic insight into the genus is currently available. Therefore, we aimed to position the Chinese collections of *M. terfezioides* in a phylogenetic network of the whole genus, focusing on the relationship between the Chinese collections and the collections from other areas worldwide, in order to ultimately hypothesize the genus’s geographic origin.
MATERIALS AND METHODS

Study site and sampling

The most temperate continental part of China, Inner Mongolia, has a cold semi-arid (BSk) to cold desert (BWk) climate (Peel, Finlayson & McMahon, 2007). Although the occurrence of Mattirolomyces was not previously recorded in this area, the genus was found in most of its neighboring provinces, which indicated its potential for fruiting in Inner Mongolia as well. Sporocarps were primarily sought for in ecosystems that were suitable for Mattirolomyces (Kagan-Zur & Akyuz, 2014). When selecting sampling microlocations, we targeted the known ectomycorrhizal partner sites with pines and black locusts (Kagan-Zur & Akyuz, 2014), as well as young plantations of Populus alba L. ssp. pyramidalis (Bunge) W.Wettst. at various locations in the Baotou area, Inner Mongolia, China, between September and October 2018 and 2019.

The main climatic characteristics of the broader area where M. terfezioides was repeatedly collected were: an elevation of about 1,070 m in the surveyed area, average temperature of around 8.5 °C, lowest temperature of minus 27.6 °C, and highest temperature of 40.4 °C. The average annual rainfall in this area over the last 18 years is 301.6 mm (the minimum rainfall was 175.9 mm in 2005, and the maximum rainfall was 465.2 mm in 2003). The average annual rainfall in 2018 and 2019 was 364.8 and 327.6 mm, respectively, according to data from Inner Mongolia Meteorology. The soils are sandy, hyphal aggregates connected the roots with above ground parts of the plants, and the sporocarps of this fungus developed from the hyphal aggregates.

Sporocarps were collected by raking the soils following procedure in Castellano, Trappe & Luoma (2004). All sporocarps were photographed in situ with a Canon EOS 60D camera (Canon, Tokyo Japan), then dried in a forced-air dryer and kept in the Herbarium and Fungarium of Baotou Teachers’s College under accession number Fan0273.

Determining soil physical and chemical properties

The soil samples were taken from the immediate vicinity of the fruiting sporocarps to a depth of 10 cm. Soil pH was measured in 1M KCl (1:5 w/v). Organic C and total N were analyzed using the CHNS-analyzer system (Elementar Analysen Systeme GmbH, Hanau, Germany) with the burning method at 450 °C and 1,250 °C, respectively (Liu et al., 2012). We determined total organic matter, available phosphorus content, and available potassium as well as the total content of water-soluble salts following the standardized operation procedures (Pansu & Gautheyrou, 2007).

Morphological observation

The macro-morphological characterization of ascomata was performed using a stereomicroscope (Motic K400; Motic Asia, Kowloon, Hong Kong) following the Mattirolomyces morphological characters description of Kovács et al. (2011). The micro-morphological features were determined on 30 spores and asci using a light microscope (Motic BA410E with a Moticam2506 camera; Motic Asia, Kowloon, Hong Kong). Melzer’s reagent and Cotton blue chemical reactions were also used in order to improve the morphological identification. The spore morphology and ornamentation were
examined using scanning electron microscopy (SEM). The observations were performed on a desiccated spore suspension coated with platinum-palladium using a vacuum metallizing machine (Hitachi E-1010; Hitachi, Tokyo, Japan). Electron microscope images were obtained with a Hitachi S-530 (Hitachi, Tokyo, Japan) scanning electron microscope.

**DNA extraction, PCR amplification, and sequencing**

DNA extraction, PCR amplification of the complete rDNA ITS region using primers ITS1f/ITS4 (White et al., 1990) and Taq PCR Master Mix (Biobasic, Markham, ON, Canada) as well as sequencing were carried out according to Wang, Liu & Sun (2017). PCR products were purified and sequenced at the Chengdu Institute of Biology, Chinese Academy of Sciences, Chengdu, Sichuan, China. The rDNA ITS sequence obtained in the present study was deposited in GenBank under the accession number listed in Table 1.

**Phylogenetic analyses**

Available and compete nuclear rDNA ITS sequences from the genus *Mattirolomyces* were retrieved from GenBank (Benson et al., 2013) and UNITE databases (Kõljalg et al., 2013) on December 12, 2019. We conducted a nucleotide search using the basic local alignment search tool (BLAST) with our representative sequence to found additional sequences that were potentially misnamed in the searched databases. A local *Mattirolomyces* spp. nuclear rDNA ITS sequence database was built based on available and reliable sequences, and we selected environmental parameters from the corresponding original papers of the sequences or directly from online databases (Table 1), with meteorological data from the latest FLUXNET synthesis dataset, the FLUXNET2015 database (http://fluxnet.fluxdata.org/data/fluxnet2015-dataset/data-processing/).

DNA sequences were assembled in BioEdit v5.0.9 (https://bioedit.software.informer.com). MEGA7.0 software (Kumar, Stecher & Tamura, 2016) was then used for multiple sequence alignment and phylogenetic analysis. The internal MEGA7 plug-ins were used for sequence alignment (ClustalW), testing for the best nucleotide substitution model (Model Test), maximum likelihood phylogenetic analysis (ML phylogenetic analysis), and construction of the phylogenetic tree. Kimura’s 2-parametric model was selected as the best model for a distance calculation of a given dataset. In order to evaluate the stability of the ML evolutionary tree topology, 1,000 bootstrap repetitions were run. Using the Bayesian method, we calculated Bayesian inference with MrBayes v. 3.1.2 (Ronquist & Huelsenbeck, 2003) and an HKY+G model. Four Markov chains were run for two runs from random starting trees for 1 million generations, until the split deviation frequency value <0.01. Every 100th generation was sampled. The Bayesian inference tree was visualized in FigTree 1.4.2. Branches that received bootstrap from ML ≥60% and Bayesian posterior probabilities (BPP) ≥0.95 were considered significantly supported.

For the phylogenetic network analysis, the same nucleotide dataset was realigned in MAFFT v. 7.304b (Katoh & Standley, 2013) and analyzed with a Median joining approach in Network5 (Bandelt, Forster & Röhl, 1999). The phylogenetic network constructed in Network5 was modified and annotated in the GNU General Public License program Inkscape 0.91 (https://inkscape.org/release/inkscape-0.91/).
| Species              | GenBank accession numbers | Geographic origin                                      | Climate (Köppen-Geiger climate classification) | Potential(∗) symbiotic partners | Sequence reference                  |
|----------------------|---------------------------|-------------------------------------------------------|-----------------------------------------------|--------------------------------|-------------------------------------|
| Mattirolomyces terfezioides | KT963177                | China, Hebei Province, Wanxian                          | Dwa                                           | Robinia pseudoacacia           | Wang, Liu & Sun (2017)              |
| Mattirolomyces terfezioides | KT963175                | China, Beijing City                                    | Dwa                                           | Robinia pseudoacacia           | Wang, Liu & Sun (2017)              |
| Mattirolomyces terfezioides | AJ305170                | Italy, Ravenna                                         | Cfa                                           | n/a                            | Kovács et al. (2001)               |
| Mattirolomyces terfezioides | AJ305169                | Hungary, Great Hungarian Plain, Kunfehértó             | Cfb                                           | n/a                            | Kovács et al. (2001)               |
| Mattirolomyces terfezioides | AJ272442                | Hungary, Great Hungarian Plain, Órbottyán               | Dfa/Dfb                                       | n/a                            | Kovács et al. (2001)               |
| Mattirolomyces terfezioides | AJ305045                | Hungary, Great Hungarian Plain, Mogyoród                | Dfa/Dfb                                       | n/a                            | Kovács et al. (2001)               |
| Mattirolomyces terfezioides | AJ272443                | Hungary, Great Hungarian Plain, Gyál                    | Dfa/Dfb                                       | n/a                            | Kovács et al. (2001)               |
| Mattirolomyces terfezioides | AJ306556                | Hungary, Great Hungarian Plain, Kunfehértó             | Cfb                                           | n/a                            | Kovács et al. (2001)               |
| Mattirolomyces terfezioides | AJ272444                | Hungary, Great Hungarian Plain, Órbottyán               | Dfa/Dfb                                       | n/a                            | Kovács et al. (2001)               |
| Mattirolomyces terfezioides | AF276681                | Hungary, Surány                                        | Dfa                                           | n/a                            | Díez, Manjon & Martin (2002)       |
| Mattirolomyces terfezioides | AJ272445                | Hungary, Süllysáp                                      | Dfa/Dfb                                       | n/a                            | Kovács et al. (2001)               |
| Mattirolomyces terfezioides | GQ231754                | France, Provence-Alpes-Côte d’Azur, Le Thor            | Csa                                           | n/a                            | Trappe, Kovács & Claridge (2010)   |
| Mattirolomyces terfezioides | AJ306555                | Hungary, Great Hungarian Plain, Kunfehértó             | Cfb                                           | n/a                            | Kovács et al. (2001)               |
| Mattirolomyces terfezioides | AF276680                | Hungary, Csomád                                        | Dfa                                           | n/a                            | Kovács et al. (2001)               |
| Mattirolomyces terfezioides | KT025693                | South Korea, Buk-myeon, Taegunang                       | Dwa                                           | Robinia pseudoacacia           | Ka et al. (2015)                   |
| Mattirolomyces terfezioides | KT963176                | China, Shanxi Province, Taiyuan                         | BSκ                                           | Robinia pseudoacacia           | Wang, Liu & Sun (2017)              |
| Mattirolomyces terfezioides | JP908728                | Italy                                                 | n/a                                           | Robinia pseudoacacia           | Osmundson et al. (2013)            |
| Mattirolomyces terfezioides | AJ875015                | Hungary                                               | Dfa/Dfb                                       | Robinia pseudoacacia           | Bratek et al. (1996)               |
| Mattirolomyces terfezioides | KT963178                | China, Shanxi Province, Taiyuan                         | BSκ                                           | Robinia pseudoacacia           | Wang, Liu & Sun (2017)              |
| Mattirolomyces terfezioides | AJ875016                | Hungary                                               | Dfa/Dfb                                       | Robinia pseudoacacia           | Bratek et al. (1996)               |
| Mattirolomyces terfezioides | MN619773                | China, Inner Mongolia, Baotou                          | BSκ                                           | Populus alba L. ssp. pyramidalis | this study                         |
| Mattirolomyces spinosus  | HQ660384                | Pakistan, Punjab, Sheikhupura                          | BSh                                           | n/a                            | Kovács et al. (2011)               |

(Continued)
RESULTS

Taxonomy

Over 2 years of hunting for hypogeous fungi in *Mattirolomyces*-like habitats, we collected two independent collections made up of a total of 32 sporocarps, all from *Populus alba* ssp. *pyramidalis* plantations.

The morphological description of the collections from Inner Mongolia affiliate them to *Mattirolomyces terfezioides* (Mattir.) E. Fisch., as described by Fischer (1938). Ascomata (fresh specimens, Fig. 1A) were hypogeous or subepigeous, 4–5 cm in diam., subglobose to irregular massy, white, surface smooth to scabrous, lobed and furrowed; gleba solid, firm with minute pockets, white with narrow white veins (Fig. 1C). Taste and odor were strongly sweet when fresh. Dark brown nombril (0.5–1.0 cm in diam.) was found in some specimens as hyphal aggregates, attached with the base of the sporocarps (Fig. 1B). Paraphyses were absent.

Microscopic features: peridium thin, 120–280 mm thickness, not differentiated from the gleba, composed of inflated hyphae and irregular, hyaline cells; gleba composed of interwoven septate hyphae 7.5–11(20) μm broad, with some free hyphal ends; asci randomly arranged in gleba, 8- or 10-spored, hyaline, globose to ellipsoid, pockety, saccate, cylindrical or clavate, (55) 65–95 (117) × (26) 35–45 (60) μm, sessile or occasionally sub-stipitate with a short stalk, disintegrating with age, thin walled, readily separable from gleba hyphae, nonamyloid (Figs. 1D and 1E); ascospores hyaline to pale yellow, globose, (12) 14–19 (22) μm in diam. excluding the ornamentation (Figs. 1D and 1E); ornamentation of blunt spines connected in an irregular alveolate reticulum, 1–4 μm high, mostly have a de Bary bubble and are uniguttulate, walls 1.5–2 μm thick (Fig. 1F).
In terms of ecology, all collections were found in the vicinity of *Populus alba* L. ssp. *pyramidalis* (Bunge) Wettst. Soils were sandy to finely sandy with a history of extensive management practices. Soils have relatively high water-soluble salt content (1.29 g kg\(^{-1}\)), neutral pH (7.34), containing 1.49 g kg\(^{-1}\) of total nitrogen, 46.4 mg kg\(^{-1}\) of available phosphorus, and 29.82 g kg\(^{-1}\) of organic matter. The average annual precipitation for the collections from the sampled region of Inner Mongolia (area of the Bao Tou City) were more similar to European collections, Mediterranean collections from several countries, and Continental collections from Hungary, than to other Asian collections (Table 1, Fig. 2). The Inner Mongolian collections were from sites with larger winter/summer temperature differences and lower winter rainfall averages when compared to the other collections included in this study (Fig. 2).
Phylogenetic analysis

Bayesian and ML phylogenetic analyses resulted in a strongly supported, topologically identical phylogenetic tree with a well-supported major clade containing the studied specimen from Inner Mongolia which clustered together with other *M. terfezioides* from China, Hungary, Italy, and South Africa (Fig. 3).

A phylogenetic network analysis (Fig. 4) separated all the five recognized taxa in the genus *Mattirolomyces* with an unexpected higher diversity displayed in *M. terfezioides*. The phylogenetic distance of the outgroup (*Elderia arenivaga*) from the genus *Mattirolomyces* indicated a poor yet the most optimal selection regarding the available taxa and their sequences. At the base of the *Mattirolomyces* cluster, three lineages were disclosed. The first lineage led to three clusters: one directed to South Africa with *M. austroafricanus*, the second with a more basal position of *M. spinosus* from south Asia (the collection was from Pakistan), and a phylogenetically close collection from the United States, with a distinct sub-cluster of closely related *M. mexicanus* from Mexico and distantly-related (based on the comparison of the number of mutated sites) *M. mulpu* from Australia. For all four mentioned recent taxa, the number of available nucleotide sequences was low. The third lineage formed a cluster of *M. terfezioides* that showed higher intraspecific diversity and a recognizable geographic pattern with more basal haplotypes from China, followed by collections from S. Korea. At this point of evolution, there was a jump of haplotypes from Asia (China) to Europe (Hungary, Italy, France), and no supported intra-Europe geographic pattern was recognized.

**DISCUSSION**

*Mattirolomyces terfezioides* is a commonly collected edible hypogeous fungus best known for its traditional use in the desert regions of the Southern Hemisphere (*Trappe, Kovics & Claridge, 2010*). Although it has been used for culinary purposes since ancient Persian Empire (*ibid.*), its recent distribution outside its optimal ecological zone has not been explored. There are two well-recognized areas of distribution: the Mediterranean and
Pannonian basin in Europe (Kagan-Zur et al., 2014; Kovács et al., 2001) and Beijing, along with neighboring regions in China. The taxonomic characteristics of the Inner Mongolian collections fit well in the concept of the Mattirolyces terfezioides morphological species (Fischer, 1938; Mattirolo, 1887) and also the phylogenetic species (Díez, Manjon & Martin, 2002).

We present the first example of a global phylogenetic network study of the genus Mattirolyces and its corresponding geographic distribution pattern in M. terfezioides. Phylogenetic networks are known to give a better insight into species ecology and distribution (Fig. 4), an approach frequently used for its ability to visualize evolutionary relationships between nucleotide sequences and depict microevolution events such as the
Figure 4 A rooted phylogenetic network of *Mattirolomyces* based on the sequence dataset of the complete ITS region. Black dots represent recent taxa, gray dots represent ancestral stages/nodes. The *M. terfezioides* rDNA ITS sequence obtained in the present study is indicated by a black triangle. Values on mutation vectors represent the number of mutations between two nodes. Names of existing taxa and their geographic origin (country of collections) are given. Phylogenetic network was constructed and tested with a median joining approach.
geographical distribution of populations (Huson & Bryant, 2006). The geographic
distribution of *Mattirolomyces* indicates that the origin of the genus was the current
Asia-Pacific areas prior to frequent long-distance migration events, one of which brought
*M. terfezioides* to eastern and Northern China. Based on the ecology of recently studied
areas, the collections from Inner Mongolia and the Shanxi province were both similar to all
European collections and also shared the same node in the phylogenetic networks. This
supported the idea that climatic conditions were an important evolutionary drive in this
species, and that the European *M. terfezioides* species were probably originated from the
China area in Asia. *M. terfezioides* haplotypes from Europe appeared to form more
terminal leaves on the network, indicating their more recent arrival to this area, which is
additionally supported by the unresolved haplotype distribution between two main
suitable habitats: Mediterranean areas and the Pannonian basin (Kovács et al., 2001).
The observed diversification and lack of any further geographic or ecological
microevolutionary structure in the European haplotypes in the more terminal leaves of the
network additionally support the theory of *M. terfezioides* recent arrival to this area and/or
lack of evolutionary pressure.

Our collections are originated from the continental steppe areas of Inner Mongolia in
China, an area characterized by cold semi-arid climates with hot dry summers and cold
winters with little snowfall, classified as BSk according to the Köppen-Geiger climate
classification, and where climate-related indicators point towards a severe spatial
desertification risk (Spinoni et al., 2015). Presently, the area still experiences little rainfall
with a low average yearly precipitation and lower average yearly temperatures and winter
extremes (>5 degrees lower) compared to any other known *Mattirolomyces terfezioides*
areas. Our findings indicated that this species survives in dryer and colder conditions, at
least outside its fruiting period, than previously reported (Gógán Csorbainé et al., 2008),
and is becoming further endangered due to projected future climate changes (He et al.,
2019). The same collections also showed ecological discrepancies from other currently
known species ecologies. *Mattirolomyces terfezioides* is usually found under *Robinia
pseudoacacia* L., but rarely under artificially planted *Diospyros kaki* Thunb., *Prunus avium*
(L.) L. or diverse families of Leguminosae, Ebenaceae, and Rosaceae in Southern and
Central Europe and in Northern China (Fischer, 1938; Bratek et al., 1996; Wang, Liu &
Sun, 2017). *R. pseudoacacia* is native to the Southern Appalachian and Ozark Mountains of
the United States (Huntley, 1990), and was introduced to Europe, Asia, Australia, South
America, and Africa mainly as an ornamental plant, or was cultivated to revegetate
disturbed sites or for agricultural and commercial uses in recent centuries (Keresztesi,
1988). Its mycorrhizal association with *Mattirolomyces terfezioides* is most likely secondary
since phylogeographically basal haplotypes of *M. terfezioides* originated from Asia and not
from North America. All our *M. terfezioides* were collected close to poplars, including pure
poplar plantations. As far as we know, this is the first study that shows that *M. terfezioides*
is potentially associated with *Populus alba*, a tree species in the family of *Salicaceae* with a
wide distribution in Europe and central Asia (Palancean et al., 2018). *Populus* spp. are
among ectomycorrhizal hosts for *M. terfezioides*, despite *Populus* spp. form a dual
mycorrhiza with the ratio between ectomycorrhiza and arbuscular mycorrhiza depending
on specific soil conditions (Neville et al., 2002). Since the European poplar species are most closely related to the Asian species (Cervera et al., 2005), they could be potential mycorrhizal partner for the European *M. terfezioides* species.

In addition, climate change, recent industry, urbanization, and other land use conversion factors are threatening the survival of *M. terfezioides* in the wild, and maybe the reason for its long-lost status in China. *Populus* spp. have only recently become the dominant species in Northern China and are mainly used for the restoration of degraded arid and semi-arid landscapes, combating desertification, and drought resilience strategies (FAO, 2016). All *Populus*-planted areas are sites where *M. terfezioides* has a potential to grow. These areas may also serve to protect and preserve this rare desert fungus, as long as suitable agricultural practices for its cultivation are developed and supported, especially in rural, arid, and semiarid areas. However, *M. terfezioides* in China, especially among the Mongol people, remains underexploited and would require further ecological and *M. terfezioides* mycorrhizal synthesis investigations in order to fully develop agricultural practices for its sustainable cultivation. *M. terfezioides* could become an excellent model not only to develop local economy in rural areas, but also to highlight the importance of non-timber forest-related products in otherwise industrial forest tree plantations.

**CONCLUSION**

The first record of *M. terfezioides* and its distribution in Inner Mongolia outside its current distribution area in China with the lowest known temperature and precipitation averages for this species. Our first attempt at phylogenetic network analysis in the genus *Mattirolomyces* revealed its geographic origin was in Asia-Pacific areas prior to frequent long-distance migration events. *M. terfezioides* seems to be originated from Inner Mongolia and the Shanxi province of China in Asia and was subsequently transported to Europe. Exploring *M. terfezioides* ecology and its potential to grow with poplars also increase its potential for cultivation and consumption in Central and Eastern China. This is a completely underexploited possibility among Mongols in China, and it deserves further ecological and mycorrhizal investigations on *M. terfezioides* in arid areas of China which should be carried out in the near future.

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**ADDITIONAL INFORMATION AND DECLARATIONS**

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Competing Interests
The authors declare that they have no competing interests.

Author Contributions
• Jie Wei conceived and designed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
• Tine Grebenc analyzed the data, prepared figures and/or tables, and approved the final draft.
• Xuan Zhang performed the experiments, prepared figures and/or tables, and approved the final draft.
• SiMin Xiang performed the experiments, authored or reviewed drafts of the article, and approved the final draft.
• Yongjun Fan conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.

DNA Deposition
The following information was supplied regarding the deposition of DNA sequences:
The sequence is available at GenBank: MN619773.1.

Data Availability
The following information was supplied regarding data availability:
The raw measurements are available in the Figs. S1–S7 and Table 1.

Supplemental Information
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