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Structural analogues of existing anti-viral drugs inhibit SARS-CoV-2 RNA dependent RNA polymerase: A computational hierarchical investigation

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PII: S2405-8440(21)00540-5
DOI: https://doi.org/10.1016/j.heliyon.2021.e06435
Reference: HLY 6435

To appear in: HELIYON

Received Date: 12 December 2020
Revised Date: 16 February 2021
Accepted Date: 3 March 2021

Please cite this article as: Hasan, M.K., Kamruzzaman, M., Bin Manjur, O.H., Mahmud, A., Hussain, N., Alam Mondal, M.S., Hosen, M.I., Bello, M., Rahman, A., Structural analogues of existing anti-viral drugs inhibit SARS-CoV-2 RNA dependent RNA polymerase: A computational hierarchical investigation, HELIYON, https://doi.org/10.1016/j.heliyon.2021.e06435.

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Title:
Structural analogues of existing anti-viral drugs inhibit SARS-CoV-2 RNA dependent RNA polymerase: A computational hierarchical investigation

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Abstract:

The Coronavirus Disease 2019 (COVID-19) caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) became a pandemic, resulting in an exponentially increased mortality globally and scientists all over the world are struggling to find suitable solutions to combat it. Multiple repurposed drugs have already been in several clinical trials or recently completed. However, none of them shows any promising effect in combating COVID-19. Therefore, developing an effective drug is an unmet global need. RdRp (RNA dependent RNA polymerase) plays a pivotal role in viral replication. Therefore, it is considered as a prime target of drugs that may treat COVID-19. In this study, we have screened a library of compounds, containing approved RdRp inhibitor drugs that were or in use to treat other viruses (favipiravir, sofosbuvir, ribavirin, lopinavir, tenofovir, ritonavir, galidesivir and remdesivir) and their structural analogues, in order to identify potential inhibitors of SARS-CoV-2 RdRp. Extensive screening, molecular docking and molecular dynamics show that five structural analogues have notable inhibitory effects against RdRp of SARS-CoV-2. Importantly, comparative protein-antagonists interaction revealed that these compounds fit well in the pocket of RdRp. ADMET analysis of these compounds suggests their potency as drug candidates. Our identified compounds may serve as potential therapeutics for COVID-19.

Keywords: COVID-19, RNA dependent RNA polymerase, binding free energy, molecular docking, virtual screening, molecular dynamics simulation
1. Introduction:

SARS-CoV-2 is a novel coronavirus that first emerged in late 2019 at Wuhan of China (Ludwig & Zarbock, 2020). This is the 7th coronavirus species discovered, belonging to the order Nidovirales of the Coronaviridae family. Since its emergence, till 16th February 2021, it has reportedly infected 109,706,925 people and caused 2,419,664 deaths in total (Coronavirus Update (Live): 109,706,925 Cases and 2,419,664 Deaths from COVID-19 Virus Pandemic - Worldometer). Ever since the disease broke out in China, the whole world started taking precautionary measures but failed to stop its rapid spread across the globe. Due to its extreme outbreak, the World Health Organization (WHO) declared a state of emergency and called on a global pandemic on March 11, 2020 (Brinks & Ibert, 2020). Initially, the disease was characterized by fever, coughing, sneezing, shortness of breath, or breathing difficulties (Huang et al., 2020); in severe cases the disease can result in pneumonia and multiple organ failure leading to death. Recent cases complaining of a higher rate of renal impairment indicates its possible role in rendering kidney dysfunction (Z. Li et al., 2020). Similar to an influenza outbreak, COVID-19 causes myocardial injury that may be related to increased viscosity, heightened coagulation cascade, pro-inflammatory effects, or endothelial cell dysfunction caused by SARS-CoV-2 virus (JC et al., 2018; Nguyen et al., 2016). The virus attaches to host cells mostly in the upper respiratory tract via an interaction between spike glycoprotein of the virus and ACE-2 receptor of human (Tai et al., 2020).

The molecular biology of this novel coronavirus has already been vividly studied and several salient features of its genome organization have been elucidated in a number of studies. The RNA genome consists of at least six open reading frames (ORF) (Ahmed et al., 2020). One ORF of special importance is the ORF 1a/b which constitutes two-thirds of the genome and synthesizes two very important polypeptides namely polypeptide 1a and polypeptide 1b which eventually produce 16 nonspecific proteins (NSPs) of the virus. The other 5 ORFs constitute nearly one-third of the genome and produce four structural proteins for the virus, which includes spike glycoprotein (S), envelope glycoprotein (E), membrane protein (M) and nucleoprotein (N) (Woo et al., 2010). Eleven cleavage sites have so far been discovered within NSP5 that generates many important proteins, including RdRp. RdRp plays crucial role in the replication of RNA viruses that makes it an ideal drug target for anti-viral drug development against RNA viruses. For treating many RNA virus infections such as Hepatitis C, Zika and several coronaviruses, many anti-viral drugs have already been developed, which target the viral RdRp enzyme (Elfiky, 2020). Some antiviral drugs, including remdesivir and favipiravir, have been briefly tested against SARS-CoV-2 RdRp, though partially successful further validation is required for the clinical approval to use those for treating SARS-CoV-2 infection (Harrison, 2020; Lung et al., 2020; Zhang1 & Zhou1, 2020).
Over the last ten months, scientists have come up with different ways to alleviate the severity of the disease and reduce its contagiousness (Van Kampen et al., 2020). So far, we have only been able to battle the disease by maintaining social distancing, quarantine and hygiene. Very recently, many countries in the world have already started running the vaccination program (D. D. Li & Li, 2021; Rubin & Longo, 2020). Yet, due to production and budget limitation it will take considerable time to vaccinate the entire population of the world. Importantly, this is uncertain whether the currently developed vaccines will give durable immunity against the virus. In addition, acquiring newer mutations by the virus may render the vaccines ineffective or the virus may make its way to infect vaccinated individuals. Therefore, discovery of effective drugs for treating COVID-19 patients is an urgent need. The enzyme, RdRp plays a pivotal role in viral replication, having no host homologues and, therefore, developing therapeutics that inhibit the enzyme would not only inhibit viral replication but also minimize any potential risks in host cells. A number of FDA-approved RdRp inhibitor drugs, which include remdesivir, favipiravir, sofosbuvir, ribavirin, lopinavir, ritonavir, tenofovir and galidesivir, are effective against a broad range of RNA viruses, including past coronaviruses (G. Li & De Clercq, 2020; Mitjà & Clotet, 2020; Şimşek Yavuz & Ünal, 2020; Sinha & Balayla, 2020). Although remdesivir, an experimental Ebola virus drug, has been approved for emergency use only for hospitalized patients and recent clinical data suggest a moderate clinical improvement by reducing the recovery period (Beigel et al., 2020), however, no impact on mortality; therefore, the potentiality of the drug is not as promising as expected. Thus, identifying novel therapeutics that have the potential to effectively combat the disease is an unmet need. To address this, we, herein, virtually screened FDA approved RdRp inhibitor drugs and their structural analogues (~2400 compounds) based on molecular docking analysis of these compounds in order to identify drug candidates that can potentially inhibit SARS-CoV-2 RdRp. SARS-CoV nsp7 and nsp8 activate and confer the processivity to the nsp12 RNA-synthesizing activity (Subissi et al., 2014). SARS-CoV-2 RNA-dependent RNA polymerase (PDB ID: 6M71) is a complex with those cofactors (nsp7 and nsp8) and nsp12 is a target for nucleotide analog of several antiviral inhibitors and the structure may provide a basis for new antiviral therapeutics (Gao et al., 2020). Based on docking scores, protein-ligand interactions, molecular dynamic simulations and free energy calculation we predicted five RdRp inhibitor drug candidates that may have the potential to combat life-threatening SARS-CoV-2 infection, and therefore, may serve as a cure to treat the disease, addressing the current global need.

2. Methods

2.1. Ligand selection and preparation:

An extensive literature assessment was performed to gather information on antiviral drugs that target and inhibit RdRp enzyme. The ability to block RdRp with various extents has been reported in other RNA
viruses, which includes H1N1 (Lin et al., 2015), HCV2 (Hofmann et al., 2008), HIV3 (Blair & Cox, 2016), SARS (Fani et al., 2020) and MERS (Fani et al., 2020). Eight antiviral compounds: favipiravir (Allen et al., 2020), sofosbuvir (Buonaguro & Buonaguro, 2020), lopinavir (Chakraborty & Das, 2020), tenofovir (Allen et al., 2020), ribavirin (Gish, 2006), ritonavir (Chakraborty & Das, 2020), galidesivir (Allen et al., 2020) and remdesivir (Fani et al., 2020) have been primarily reported to inhibit RdRp of these viruses. To determine their potential in inhibiting SARS CoV-2 RdRp computationally, their SDF (Spatial Data File) files were downloaded from the PubChem database (https://pubchem.ncbi.nlm.nih.gov/). Moreover, structurally similar compounds of the inhibitor drugs were downloaded from Swiss similarity software (Zoete et al., 2016) (http://www.swisssimilarity.ch/) by activating zinc drug-like features. A library of ~2400 compounds has been prepared.

2.2. Macromolecule selection and preparation:

The crystal structure of RdRp of SARS-CoV-2 was collected from the RCSB Protein Data Bank (https://www.rcsb.org/search) (PDB ID: 6M71) (Gao et al., 2020) followed by optimization and processing of the structure by Swiss-PDB viewer software packages (Guex & Peitsch, 1997) (https://spdbv.vital-it.ch/) (version 4.1.0) based on their least energy. Some significant factors, such as side-chain geometry, improper bond order, and missing hydrogen, were observed in the crystal structure of the RdRp protein. PyMol (version 1.1) software package (Adams et al., 2010) (https://pymol.org/2/) was used to remove all the water molecules, hetero atoms, and inhibitor existent in the Crystal structure.

2.3. Virtual screening and ligand preparation

As mentioned above the structural analogues of the inhibitor drugs were obtained from the ZINC database (Sterling & Irwin, 2015) for the best optimal hit against the mentioned target (Figure 1). PyRx (Dallakyan & Olson, 2015) from MGLTools (https://cscr.scripps.edu/mgltools/) was used in the virtual screening executed in the study. For computational drug discovery, PyRx is used as a Virtual Screening software that screens libraries of compounds against potential drug targets according to their target binding affinities.

2.4. Molecular docking studies

The protein-ligand binding interaction of chosen protein-ligand complexes was performed using Autodock Vina (Trott & Olson, 2010). The docking analyses were performed using a semi-flexible docking approach. Briefly, the protein RdRp was kept rigid and ligands were kept flexible. The allowed degrees of freedom for ligand molecules were considered 10. The steps involving the conversion of molecules into pdbqt format, box type, grid box generation, etc. were specified by AutoDock. Since
remdesivir were considered as a potential drug for SARS-CoV-2 based on some initial \textit{in vitro} findings (Shannon et al., 2020) and the site of interaction of the remdesivir to SARS-CoV-2 RdRp has already been determined (Gao et al., 2020). We have chosen the interaction site for determining the inhibitory potential of the selected compounds in molecular docking. Critical 11 amino acid positions of A chain of RdRp (PDB ID: 6m71) is important for remdesivir binding and were selected for creating remdesivir incorporation model. These include K545, R553, R555, V557, D618, S 623, T680, D682, N691, D760 and D761. The grid box was set to size 22.32 Å × 23.91 Å × 32.57 Å (x, y and z) parameters and the center 118.660 Å × 117.665 Å × 131.857 Å (x, y and z) parameters. The docking results were analyzed using Discovery studio visualizer (“BIOVIA Discovery Studio — Discngine,”) and Molecular Operating Environment (Vilar et al., 2008) software, and the docking of ligands to RdRp with the least energy was considered to have a significant binding affinity.

2.5. Drug likeness analysis of the potential drug candidates:

ADMET profiling is a significant pointer in defining the effectiveness and safety of a drug compound before bringing it into commercialization. Due to cost-effectiveness and proper timing in the experimental evaluation of ADMET properties, computational methods have been developed as a practicable alternative in the high-throughput drug discovery process (Shen et al., 2010). The ADMET properties of the 8 top-ranked ligands were calculated by the online ADMET structure-activity database (admetSAR) (Yang et al., 2019) (http://lmmd.ecust.edu.cn/admetsar2/) and pkCSM (Pires et al., 2015) (http://biosig.unimelb.edu.au/pkcsms/) where SMILES (simplified molecular-input line-entry system) strings were utilized throughout the whole process.

2.6. Molecular dynamics simulation with MMGBSA scoring:

Complexes were solvated with TIP3P water model employing a dodecahedron box with a 12 Å packing using tleap available in Amber16 package (Case et al., 2005). The systems were then neutralized with Na+ and Cl− ions until neutrality. The protein, water molecules, and ions were defined by the ff14SB AMBER force field (Duan et al., 2003). Ligands were parameterized by assigning AM1-BCC atomic charges employing the general AMBER force field (GAFF) (Wang et al., 2004). Simulations were run at the NPT ensemble at 310 K via a Langevin thermostat, 1 bar of pressure, and an isotropic Berendsen barostat. Periodic boundary conditions (PBC) were considered with a cutoff of 10 Å for the nonbonded interactions, whereas the electrostatic term was described via the PME method (Darden et al., 1993). A numeric time step of 2 fs and the SHAKE algorithm (Van Gunsteren & Berendsen, 1977) was chosen to constrain bond lengths at individual equilibrium. After constructions and previous MD simulations, a minimization and relaxing protocol was performed comprising of: 1,000 steps of energy minimization by
conjugate gradient method; 1000 ps of pre-relaxation with harmonic restraint for protein and ligand; 1000 ps relaxation removing the ligand restraints; 1000 ps of relaxation with no restrictions to the side chains of the residues 5 Å around the ligand; 1000 ps of NPT relaxation without restrictions for the entire residues around the ligand; 50 ns of NPT MD simulations. The RMSD and clustering analysis was performed using the cpptraj tool in Amber16 to check system equilibration and identification of the most populated conformation during the MD simulations. Images were built using PyMOL (“DeLano, W.L. (2002). The PyMOL Molecular Graphics System. Delano Scientific, San Carlos. - References - Scientific Research Publishing,”).

The Molecular Mechanics Generalized Born Surface Area (MMGBSA) (Belouzard et al., 2012) approach was employed to determine the binding free energy ($\Delta G_{\text{mmgbasa}}$). This analysis was performed over the last 20 ns of MD simulation, selecting a total of 2000 snapshots at time intervals of 10 ps, using implicit solvent models (Feig et al., 2004), and a salt concentration of 0.10 M. The $\Delta G_{\text{mmgbasa}}$ values were determined as previously described (Bello, 2018) and embody average values of triplicate MD simulations.

3. Results:

3.1. Virtual screening, ligand selection and molecular docking studies

In order to identify potential drug candidates targeting SARS-CoV-2 RdRp we adopted a computational approach. Initially, with extensive literature mining, we selected 8 FDA-approved RdRp inhibitor drugs (favipiravir (Allen et al., 2020), sofosbuvir (Buonaguro & Buonaguro, 2020), lopinavir (Chakraborty & Das, 2020), tenofovir (Allen et al., 2020), ribavirin (Gish, 2006), ritonavir (Chakraborty & Das, 2020), galidesivir (Allen et al., 2020) and remdesivir (Fani et al., 2020) that are effective against broad ranges of RNA viruses, including past coronaviruses. We hypothesized that these inhibitor drugs and/or their structural analogues could have the potential to inhibit SARS-CoV-2 RdRp. Following downloading these drugs and their structural analogues (detailed in method section), we have performed an initial virtual screening using PyRx that eventually yielded 8 best compound hits. The structures of the 8 best compound hits are shown in Figure 1. To further validate the initial screen, we later performed molecular docking with AutoDock Vina (Trott & Olson, 2010) and the drug binding scores for the above mentioned approved drugs and their best hit analogues suggest high affinity binding (Table 1 and Table 2), further strengthening our initial findings. It is important to mention that the structural analogues were predicted to have better binding than the existing FDA-approved drugs tested in this study, as evident from the binding scores (Table 1 and Table 2).

3.2. Docking analyses and drug interactions
The chosen 8 compounds predicted (by molecular docking) to bind effectively with RdRp were selected for further analysis (the highest-ranked bound compound in all poses made as a cluster at the ligand-binding sites of these target protein). The analyses were performed to check the efficiency of the ligand and the state of interaction using the Discovery studio visualizer (“BIOVIA Discovery Studio — Discngine,”) and show that all the 8 ligands bind efficiently and interact well with RdRp (Figure 2).

3.3. Stability of simulated systems

The RMSD analysis plots show the mobility of the receptor-ligand complexes and constant values suggest that the system reached equilibrium (Figure 3). The RMSD values of backbone atoms reached an equilibrium state between 20 to 30 ns with RMSD values that oscillated between 2.7 to 3.2 Å. Based on this, further analyses were performed considering the last 30 ns.

3.4. Structural analysis of receptor-ligand complexes through MD simulation

Analysis of the receptor-ligand complexes showed that 3 (ZINC000012863240, ZINC000049581065, and ZINC000072460420) out of 8 ligands lost interactions with the receptor at the beginning of the simulations, therefore, these three compounds were not considered in further analyses. Based on the simulation time in which systems reached equilibrium, a cluster analysis was performed using a cut-off of 2.5 Å to observe the main residues involving in stabilizing the ligand inside the binding site (Figure 2). The ligand orientation at the binding site is shown in Figure 2. ZINC000005605139 compound is coordinated by A524, R525, K591, R594, S652, and D730 through non-polar interactions. Whereas that T526 forms hydrogen bonds with compound through the polar backbone and side-chain atoms, as well as side chains of D422 and D593 (Figure 4A). ZINC000006094731 compound is bound by K515, Y516, A517, S519, K521, and R523 through non-polar interactions, whereas that R525 forms polar interactions through the backbone and side-chain atoms (Figure 4B). ZINC000012161475 compound is coupled through hydrophobic interactions with R523, W587, Y589, R594, N665, M725, D730, D731, A732, V733, V734, and F752, and forms polar interactions with D588 and K591 (Figure 4C). ZINC0000014751834 is bound by S519, P590, M764, and K768 by hydrophobic contacts and polar interactions with backbone atoms of S765 (Figure 4D). ZINC000014882040 compound is coordinated through non-polar contacts with D422, Y425, R523, A524, R594, and K768, and polar interaction with side chains of K591 and N522, and with polar backbone atoms of A524 (Figure 4E). Comparative analysis revealed that most of the interactions are hydrophobic, and R523, A524, R525, and R594 residues were present in almost all the complexes, suggesting an important role of these residues in ligand binding stabilization.

3.5. Binding Free Energy of protein-ligand complexes
To further assess the thermodynamic feasibility of binding of the top-5 ligands to RdRp, MMGBSA method was used to calculate the binding free energy of the protein-ligand complex. Simulation runs were extracted to calculate the MMGBSA free binding energy. All of the five compounds showed negative binding free energy; suggesting successful binding throughout the simulation (Table 3). The molecular mechanic energy showed that Van der Waals energy ($\Delta E_{\text{vdw}} + \Delta G_{\text{pol, sol}}$) was the main force for stabilizing the ligand at the binding site (Table 3). Despite that all the systems exhibited favorable electrostatic contributions ($\Delta E_{\text{ele}}$), the higher nonpolar free energy of desolvation ($\Delta G_{\text{npol,sol}}$) cancelled with the favorable electrostatic contribution. Table 3 also shows that the binding affinities are in the order of ZINC000005605139 > ZINC000012161475 > ZINC000014882040 > ZINC000006094731 > ZINC000014751834. Comparative analysis of the binding free energy ($DG_{\text{mmgbsa}}$) values points out that ZINC000005605139 > ZINC000012161475 exhibit the more favourable $DG_{\text{mmgbsa}}$ value, suggesting that these two perhaps are better candidates to inhibit RdRp than the other three compounds.

3.6. ADMET properties

Drug bioavailability, metabolism and toxicity are important factors that may limit the efficacy of a drug in clinical settings. Therefore, to determine whether the potential drug candidates are effectively bioavailable and have potentially no carcinogenicity to organs in human, the top-5 ligands were assessed based on their ADMET profile (Table 4) and all of the 5 drug candidates were found to have no potential toxicity (residing below toxicity classes of 4 and 5), supporting their safety for human use.

Finally, the top-5 screened ligands ZINC000005605139, ZINC000006094731, ZINC000012161475, ZINC000014751834, and ZINC000014882040 may be considered as potential therapeutic candidates for further exploring their potential in preclinical and clinical settings.

4. Discussion:

The COVID-19 caused by rapidly spreading coronavirus, known as SARS-CoV-2 has been declared as a pandemic outbreak by WHO in March 2020. It has made a devastating impact on the public as well as the social health of more than 200 countries around the globe (https://www.who.int/emergencies/diseases/novel-coronavirus-2019). Therefore, scientific communities from all over the world have come forward to collaborate and make the best effort for the earliest discovery of effective anti-viral drugs for treating COVID-19. The RdRp of SARS-CoV-2 has been sequenced and the X-ray crystallography 3-D structure is available in the PDB databank (Gao et al., 2020.).

Currently, there are no effective therapeutics for treating COVID-19. Therefore, the main focus has been on disease management by prevention of the infection and following other controlling measures.
Recently, some anti-malarial drugs like chloroquine, hydroxychloroquine have given positive results in vitro cell culture systems but further clinical tests are needed for them to be approved as effective anti-viral drugs for treating SARS-CoV-2 infection (Enmozhi et al., 2020). A few other anti-viral drugs, including remdesivir, ribavirin etc., used for treating Hepatitis C and Ebola virus disease, and various vaccine candidates are being studied for effectively tackling the COVID-19 (Shoenfeld, 2020). Of particular interest was remdesivir; however, recently published clinical data showed no significant impact on the reduction in mortality, though the drug does reduce the recovery period (Hillaker et al., 2020).

Inhibition of the replication via inhibiting RdRp has been explored in the development of various antiviral drugs such as favipiravir (Allen et al., 2020), sofosbuvir (Buonaguro & Buonaguro, 2020), lopinavir (Chakraborty & Das, 2020), tenofovir (Allen et al., 2020), ribavirin (Gish, 2006), ritonavir (Chakraborty & Das, 2020), galidesivir (Allen et al., 2020), and remdesivir (Fani et al., 2020). Computational studies have also led to the suggestion of using the RdRp protein as a viable drug target against the current threat (Khan, Khan, et al., 2020). In order to find effective drug, many computational studies have been conducted, which led to in vitro work and showed promising results (Khan et al., 2021; Khan, Ali, et al., 2020) but are not still fully sufficient for mass use. Since there are no effective drugs for treating COVID-19, we adopted a computational approach to identify potential drug candidates. In this study, the inhibitory potential of the candidate drugs targeting SARS-CoV-2 RdRp was predicted via screening from a library of compounds containing existing, approved drugs against RNA viruses and their structural analogues. Careful evaluation of the library by employing various computational screening methods yielded 5 candidate drugs and gave an insight into the potential of these prospective drugs against SARS-CoV-2.

The computational screening approach enables the rapid discovery of promising compounds for developing effective therapeutics against SARS-CoV-2. It has been shown that less binding energy denotes more affinity of a compound for binding to its target (Ortiz et al., 1995). To elucidate the binding affinity to RdRp, the library of compounds was docked against RdRp using the PyRx tool (Dallakyan & Olson, 2015). Since among the approved RdRp drugs the binding site of only remdesivir to SARS-CoV-2 RdRp has already been elucidated as well as the in vitro studies of remdesivir showed some promises we, for our screening, selected 11 critical amino acids in RdRp pocket where the drug interacts. Of note, compounds with a binding affinity of -7.5 Kcal/mol or less are evaluated as potent inhibitors of enzymatic mechanisms (R Yunta, 2016). Based on binding affinities, top ranked 8 compounds were selected for further analysis. None of the approved drugs, including remdesivir, showed significant binding, perhaps these best hits are better candidates than the mother compounds.
The binding affinity determined from the docking study indicates the affinity of an individual ligand and the potency by which the ligand interacts and binds with the target protein at a specific pocket. For the best 8 hit compounds, our docking results were successful as they all were found to have the least binding scores and presence of significant hydrogen bonding at the pocket of SARS-CoV-2 RdRp. (Table 2; Figure 2).

Molecular dynamics simulation was used to validate the docking study. The overreaching goal of our molecular dynamics (MD) simulation study was to investigate the stability and dynamic activity of the receptor protein in the ligand-protein complex system. Protein flexibility plays an essential role and should be considered when designing inhibitors against a target (Salmaso & Moro, 2018). Molecular docking has been performed implementing a single structure while MD simulation presents an in-depth interpretation of the protein-ligand interactions. The stability and dynamic activity of the interaction of RdRp with the selected compounds was studied by molecular dynamics simulation analysis. The MD simulation showed that most of the interactions between RdRp-ligand complexes are hydrophobic, highlighting R523, A524, R525, and R594 amino acid residues that were present in almost all the complexes, indicating a crucial role of these residues in ligand binding stabilization. Analysis of H-bond based interactions suggests that all of the compounds docked with the RdRp with all the available binding interactions and sustain all these interactions throughout the simulation process. Apart from that the receptor-ligand based interactions, all of the docked ligands on an average formed at least two H-bonds with the nearby water molecules, which bestowed further stability of the ligands docked to RdRp.

MMGBSA analysis gave an insight into the binding energy throughout the simulation. The order of binding affinities of the finally selected 5 drug candidates are: ZINC000005605139 > ZINC000012161475 > ZINC000014882040 > ZINC000006094731 > ZINC000014751834. The other three compounds, ZINC000012863240, ZINC000049581065, and ZINC000072460420, were shown to be unstable inside the binding pocket of the SARS-CoV-2 RdRp. The more negative binding free energy (DGmmgbsa value) indicates the higher favorable binding, perhaps suggesting that ZINC000005605139 and ZINC000012161475 are better candidates to inhibit the enzyme compared to the other three compounds (Table 3).

ZINC000005605139 & ZINC000006094731, ZINC000012161475 & ZINC000014882040, ZINC000014751834 are structural analogues of tenofovir, lopinavir and ritonavir, respectively and this study predicted them to be more potent inhibitors of SARS-CoV-2 RdRp compared to their mother drugs. Physicochemical properties of the selected compounds were studied using ADMET (Hodgson, 2001) to determine their drug-likeness properties. ADMET evaluates the molecular properties of a drug that are
crucial for pharmacokinetics of the drug in human body, which includes absorption, distribution, metabolism, and excretion (ADME). Selected drugs showed satisfactory ADMET profile, which may explain by the fact that these are analogues of available drugs for which safety and efficacy have already been proven. Although 4 of the 5 compounds show hepatotoxicity as predicted from ADMET, however, the level of toxicity to the liver, if any, needs to be addressed in preclinical and clinical settings in future.

Our work is limited by the fact that the potential drugs were identified solely by computational approach. Wet lab validation is required which is beyond the scope of our current work, however, is an area that we will try to address in the future.

5. Conclusion:

The present in silico investigation explored the potentiality of the analogues of the available drugs in inhibiting SARS-CoV-2 RdRp. The result of the virtual screening yielded top-5 hit ligands. An ideal drug is presumed to not only have a high affinity to a specific target but also should have the drug likeness properties. Taking into account, for better prediction the binding affinities predicted by MMGBSA, binding modes and further 50 ns MD simulation were also determined, and five ligands (ZINC000005605139, ZINC000006094731, ZINC000012161475, ZINC000014751834, and ZINC000014882040) were found to be the most favorable ones. Considering the safety issue of these drugs, all five compounds showed a good ADMET profile. The identified 5 drug candidates may have therapeutic potential in COVID-19, however, our study offers further preclinical and clinical investigation to prove their safety and efficacy against SARS-CoV-2.

Acknowledgements:

The authors acknowledge Saeed Anwar (University of Alberta) and Sohel Mahmud (Tejgaon College, National University of Bangladesh) for helping with the writing of the manuscript.

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### Table 1. Autodock Vina docking results of the selected antiviral drugs

| Compound   | PMCID       | Score (KJ/mol) |
|------------|-------------|----------------|
| Remdesivir | 121304016   | -6.5           |
| Lopinavir  | 92727       | -7             |
| Ribavirin  | 37542       | -6.2           |
| Galidesivir| 10445549    | -6.8           |
| Sofosbuvir | 45375808    | -7.5           |
| Favipiravir| 492405      | -5.5           |
| Tenofovir  | 464205      | -5.9           |
| Ritonavir  | 392622      | -7.2           |

| Compound                      | IUPAC name                                                                 | Score (KJ/mol) |
|-------------------------------|-----------------------------------------------------------------------------|----------------|
| ZINC000005 605139            | 4-Amino-N'-naphthalen-2-ylsulfonyl-1,2,5-oxadiazole-3-carbohydrazide         | -7.9           |
| ZINC000006 094731            | (7R,8R,9S,10S)-7,8,10-Trihydroxy-7,8,9,10-tetrahydrobenzo[4]pyrene-9-sulfonic acid | -8.1           |
| ZINC000012 161475            | 5-[1-(2-Naphthalen-2-ylacetyl)piperidin-4-yl]-3-(oxolan-2-ylmethyl)-5-propylimidazolidine-2,4-dione | -8.9           |
| ZINC000014 751834            | [3-(4-Hydroxypiperidine-1-carbonyl)-1-(2-phenylethyl)-6,7-dihydro-4H-pyrazolo[4,3-c]pyridin-5-yl]-(1H-indol-5-yl)methanone | -8.9           |
| ZINC000014 882040            | 5-(4-Fluorophenyl)methyl]-5-[1-[2-(3-methylphenyl) acetyl]piperidin-4-yl]-3-(oxolan-3-yl)imidazolidine-2,4-dione | -9.1           |
| ZINC000012 863240            | N-[1-(4a-Hydroxy-1,3,4,5,6,7,8,8a-octahydroisoquinolin-2-yl)-3-(1H-indol-3-yl)-1-oxopropan-2-yl]-4-methylbenzenesulfonamide | -9.1           |
| ZINC000049 581065            | N'-(1,3-Diphenylpyrazole-4-carbonyl)-3-(2-hydroxyethyl)-4-oxophthalazine-1-carboxyrazide | -9.1           |
| ZINC000072 460420            | 7-(3,4-Dihydro-2H-1,5-benzodioxepin-7-yl)-2-(hydroxymethyl)-5-methyl-N-phenyl-1,7-dihydro[1,2,4]triazolo[1,5-a]pyrimidine-6-carboxamide | -8.6           |
Table 2. Autodock Vina docking results of the top-ranking ligands

Table 3. Binding free energy components of protein-ligand systems (in units of kcal/mol).

| System              | $\Delta E_{\text{vdw}}$    | $\Delta E_{\text{ele}}$ | $\Delta G_{\text{ele,sol}}$ | $\Delta G_{\text{ap pol,sol}}$ | $DG_{\text{mm gbsa}}$ |
|---------------------|-----------------------------|--------------------------|------------------------------|---------------------------------|------------------------|
| ZINC000005605139    | -31.90 ± 4.0                | -191.27 ± 13.5           | 186.38 ± 12.6                | -4.92 ± 0.40                   | -41.72 ± 4.0           |
| ZINC000006094731    | -29.08 ± 3.0                | -18.76 ± 6.5             | 30.18 ± 12.0                 | -3.76 ± 0.40                   | -21.43 ± 6.0           |
| ZINC00012161475     | -50.73 ± 4.0                | -32.16 ± 7.5             | 52.51 ± 6.4                  | -5.99 ± 0.50                   | -36.40 ± 5.0           |
| ZINC00014751834     | -29.79 ± 4.0                | -35.13 ± 11.0            | 48.54 ± 8.0                  | -4.09 ± 0.70                   | -20.47 ± 5.0           |
| ZINC00014882040     | -34.84 ± 6.0                | -33.81 ± 10.7            | 48.09 ± 9.7                  | -4.87 ± 0.70                   | -25.43 ± 6.0           |

ADMET results of the top-ranking ligands

| Property             | Model Name | ZINC000005605139 | ZINC000006094731 | ZINC000012161475 | ZINC000014751834 | ZINC000014882040 | Unit                        |
|----------------------|------------|------------------|------------------|------------------|------------------|------------------|-----------------------------|
| Absorption           | Water solubility | -3.533           | -2.891           | -5.178           | -4.127           | -4.263           | Numeric (log mol/L)         |
| Absorption           | Caco2 permeability | 0.492            | 1.237            | 0.846            | 1.307            | 1.159            | Numeric (log Papp in $10^6$ cm/s) |
| Absorption           | Intestinal absorption (human) | 72.698          | 39.7             | 95.097           | 87.731           | 90.718           | Numeric (%) Absorbed         |
| Distribution         | BBB permeability | -1.118           | -1.353           | -0.979           | -0.69            | -0.638           | Numeric (log BB)            |
| Distribution         | CNS permeability | -3.219           | -3.176           | -2.419           | -2.453           | -2.471           | Numeric (log PS)            |
| Metabolism           | CYP1A2     | No               | No               | No               | No               | No               | Categorical                |
| Metabolism | CYP2C19 inhibitor | No | No | Yes | Yes | Yes | Categorical (Yes/No) |
| Metabolism | CYP2C9 inhibition | No | No | Yes | Yes | Yes | Categorical (Yes/No) |
| Metabolism | CYP2D6 inhibition | No | No | No  | No  | No  | Categorical (Yes/No) |
| Metabolism | CYP3A4 inhibition | No | No | Yes | Yes | Yes | Categorical (Yes/No) |
| Toxicity   | AMES toxicity     | No | No | No  | No  | No  | Categorical (Yes/No) |
| Toxicity   | Hepatotoxicity    | Yes| No | Yes | Yes | Yes | Categorical (Yes/No) |
**Figures:**

a) ![Chemical structure](image1)

b) ![Chemical structure](image2)

c) ![Chemical structure](image3)

d) ![Chemical structure](image4)

e) ![Chemical structure](image5)

f) ![Chemical structure](image6)

g) ![Chemical structure](image7)

h) ![Chemical structure](image8)

**Figure 1.** The chemical structures of ligands that were selected based on the initial virtual screen. a) ZINC000005605139, b) ZINC000006094731, c) ZINC000012161475, d) ZINC000014751834 and e) ZINC000014882040, f) ZINC000012863240, g) ZINC000049581065, h) ZINC000072460420.
Figure 2. Molecular docking of probable inhibitory molecules onto RdRp. Panels (a–h) show the 2D structures of the binding interaction of the ligands inside the pocket of SARS Cov-2 RdRp a) ZINC000005605139, b) ZINC000006094731, c) ZINC000012161475, d) ZINC000014751834 and e) ZINC000014882040, f) ZINC000012863240, g) ZINC000049581065, h) ZINC000072460420. The types of interaction are shown at the end of the figure.
Figure 3. RMSD pattern of docked complexes compared to Cov-2 RdRp. a) ZINC000005605139, b) ZINC000006094731, c) ZINC000012161475, d) ZINC000014751834 and e) ZINC000014882040, f) ZINC000012863240, g) ZINC000049581065, h) ZINC000072460420. The RMSD pattern show that all the small molecules docked to the RdRp protein remain relatively stable under the simulated environment.
**Figure 4.** The binding orientation of 5 best ligands inside the receptor binding site of SARS Cov-2 RdRp.

a) ZINC000005605139, b) ZINC000006094731, c) ZINC000012161475, d) ZINC000014751834 and e) ZINC000014882040. The binding orientation indicates stable binding for each of the 5 ligands to the RdRp of Cov-2.