A facile aqueous production of bisphosphonated-polyelectrolyte functionalized magnetite nanoparticles for pH-specific targeting of acidic-bone cells†

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Bone malignancy treatment is being hindered due to the insufficient selectivity of therapeutic nanoparticles towards malignant bone sites. Polyelectrolyte functionalized magnetic nanoparticles having dually specific pH-sensing ability and bisphosphonate moieties, can be an effective solution for selective targeting of bone malignancies. First, polyelectrolyte was prepared via N-carboxy-citraconylation of chitosan (NCCS) followed by successive functionalization with alendronic acid (AL) and fluorescein isothiocyanate (FITC). Then, Fe3O4-NCCS-FITC-AL nanoparticles were synthesized by a facile one-step microwave-assisted aqueous method via in situ surface functionalization. The formation, crystal structure, and surface conjugation of Fe3O4 nanoparticles with polyelectrolytic stabilizer were confirmed by Fourier transform infrared spectroscopy, X-ray diffraction, and thermogravimetric analyses. Synthesized Fe3O4-NCCS-FITC-AL nanoparticles were superparamagnetic, colloidal stable and highly hemocompatible under physiological conditions. Moreover, at pH 5.0, Fe3O4-NCCS-FITC-AL nanoparticles formed a precipitate due to inversion of their surface charge. This pH-dependent charge-inversion drastically changed the interactions with erythrocytes and bones. Selective membranalysis of erythrocytes occurred at pH 5.0. The designed nanoparticles showed enough potential for selective targeting of pathological bone sites in early-stage magnetofluorescent imaging and as a therapeutics carrier to treat malignant bone diseases.

Introduction

Metastatic and metabolic bone malignancies occur in a wide range of ages including children, and the symptoms such as severe bone pain are distressing to various patients. Bone malignancy treatment is being hindered due to the insufficient selectivity of therapeutic nanoparticles towards malignant bone sites. Polyelectrolyte functionalized magnetic nanoparticles having dually specific pH-sensing ability and bisphosphonate moieties, can be an effective solution for selective targeting of bone malignancies. First, polyelectrolyte was prepared via N-carboxy-citraconylation of chitosan (NCCS) followed by successive functionalization with alendronic acid (AL) and fluorescein isothiocyanate (FITC). Then, Fe3O4-NCCS-FITC-AL nanoparticles were synthesized by a facile one-step microwave-assisted aqueous method via in situ surface functionalization. The formation, crystal structure, and surface conjugation of Fe3O4 nanoparticles with polyelectrolytic stabilizer were confirmed by Fourier transform infrared spectroscopy, X-ray diffraction, and thermogravimetric analyses. Synthesized Fe3O4-NCCS-FITC-AL nanoparticles were superparamagnetic, colloidal stable and highly hemocompatible under physiological conditions. Moreover, at pH 5.0, Fe3O4-NCCS-FITC-AL nanoparticles formed a precipitate due to inversion of their surface charge. This pH-dependent charge-inversion drastically changed the interactions with erythrocytes and bones. Selective membranalysis of erythrocytes occurred at pH 5.0. The designed nanoparticles showed enough potential for selective targeting of pathological bone sites in early-stage magnetofluorescent imaging and as a therapeutics carrier to treat malignant bone diseases.

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poly(ethylene glycol) (PEG), poloxamer, polyvinyl alcohol, polyaminoacid, carbohydrates, carboxylate and bisphosphonated polymers have been introduced on the surface of MIONPs. As a result, surface functionalized particles having relatively wider size of 100–200 nm can easily be accumulated and retained in the targeted sites due to their enhanced permeability and retention effect.

Here, bisphosphonate moiety consisting of P–C–P bond is investigated as a bone targeting, pH-sensitive, and dispersing milieu for MIONPs. Because, small organic molecules of bisphosphonates have been found useful to treat various bone malignancies like chondrosarcoma, osteosarcoma, Ewing’s sarcoma, inflammation of bones due to rheumatoid arthritis, periodontal disease, and so on. They are also efficient to inhibit bone resorption due to their ability to bind bone mineral by bis- and tridentate ligation to Ca2+ ions via osteoclastic differentiation and apoptosis. Until now, a number of targeting nano-systems based on bisphosphonates have been developed, and some of them have been tested preclinically for bone metastasis.

But bisphosphonates employed for bone-targeting are non-specific, and as a result, they target both healthy and malignant bone tissues without selectivity resulting in poor performances of therapeutics and adverse side effects. Therefore, development of synthetic methods for MIONPs with targeting ability towards malignant bone cells via facile and biosafe procedures is still a great challenge. pH-responsive systems may introduce selectivity to therapeutics based on bisphosphonate-functionalized MIONPs for metastatic bone diseases as well as primary bone malignancies. Early-stage detection of bone malignancy enables more options for better treatments, resulting in higher survival rates and enhance quality of life.

Ferreira et al. developed bone-targetable pH-sensitive bisphosphonated liposomes and evaluated their cytotoxicity, cardiototoxicity, biodistribution, and used them for treating bone metastasis. Chen et al. reported a bisphosphonated amphiphilic hyperbranched polymer and PEG-based micelles for targeting drugs to malignant bone cells. Stewart et al. reported bisphosphonated acid-sensitive drug-loaded micelles for the treatment of osteosarcoma. Au et al. designed pH-responsive folate conjugated nanoscale MOF bearing bisphosphonate moieties for treating a tumor. The pH responsibility of 1,2-dioleoyl-glycerol-3-phosphatidylethanolamine and cholesteryl hemisuccinate, ester/amide, hydrazine and phosphonate moieties are the key of these pH-sensitive systems.

To the best of our knowledge, there is no report on acid-sensitive charge-inversional bisphosphonated MIONPs for targeting and treatment of malignant bone cells. The present research focuses on a facial aqueous synthetic route to synthesize a novel magnetic nano-biosystem using a polyelectrolytic stabilizer for selective targeting of acidic bone cells. The polyelectrolyte stabilizer was first prepared via N-carboxycitraconylation of chitosan (NC) followed by successive functionalization. Then, in situ synthesis of Fe3O4 and concomitant functionalization was carried out to obtain Fe3O4–NCSS-FITC-AL nanoparticles. The designed nanoparticles would enhance therapeutic efficacy and minimize systemic and off-targeted side effects. In addition, the conjugated polyelectrolytic-NCSS-FITC-AL layer is supposed to improve hemocompatibility, biocompatibility, acid-sensing charge-stability, and selective targeting ability of the MIONPs towards malignant bone sites. We selected AL molecule as a bisphosphonate for bone-targeting component, carboxylate moieties for the acid-sensing, and CS for the biocompatible and reactive scaffold of the polyelectrolytic coating. Fluorescein isothiocyanate (FITC) was employed as a fluorescent probe for facile imaging and detection of materials. The formation, crystal structural, colloidal stability, pH-sensitive charge-inversion ability, hemocompatibility, magnetic property of the nanoparticles were confirmed and evaluated. Erythrocytes membranolytic efficiency, fluorescent properties as well as selectivity of the Fe3O4–NCSS-FITC-AL nanoparticles towards bone mineral (HAp) and a native bone sample under physiological and acidic conditions were also studied.

Results and discussion

Synthesis and characterization of polyelectrolytic NCSS-FITC-AL stabilizer

A simple three-step procedure was used to synthesize polyelectrolytic NCSS-FITC-AL (Scheme 1). In the first step, NCSS was synthesized by partial amidation of the primary amine groups in CS with citraconic anhydride (CAn) for improving dispersity in aqueous media with pH ranging from acidic to weakly basic by the carboxy group. The carboxy group in NCSS is easily transformable to the mono-Na salt via neutralization with aqueous NaOH. Synthesized NCSS is an amorphous fine white powder (Fig. S1a†), whose dispersivity was dramatically improved than CS. It was fully dispersible in neutral deionized distilled water (DDW) due to the addition of hydrophilic carboxylate moieties to CS. The fluorescent group was conjugated in the second step through the conventional nucleophilic addition of the primary amine moieties of NCSS to the isothiocyanate groups of FITC. The resulting product is a yellow-colored fine powder (Fig. S1b†). The NCSS-FITC chain possesses the primary amine, hydroxy, carboxy, and unsaturated ester groups that can be easily conjugated with suitable reagents. Among them, the highly electron-deficient unsaturated ester groups are very prone to nucleophilic addition of amines even under slightly acidic aqueous environments. In the last step, the bisphosphonate moieties were partially introduced via aqueousaza-Michael addition of the primary amine moieties of AL to the alkanyl groups of NCSS-FITC under mildly acidic conditions to form the desired polyelectrolyte, NCSS-FITC-AL. The formed polyelectrolyte was also a light-yellow-colored (Fig. S1c†) very fine powder that was also completely dispersible in DDW.

The successful synthesis of NCSS-FITC-AL was confirmed by energy-dispersive X-ray (EDX) (Fig. S2†), Fourier transform infrared (FTIR) (Fig. S3†), 1H NMR (Fig. S4†), and UV-vis (Fig. S5†) spectroscopic studies. The EDX spectroscopic analysis showed the respective elements present in the NCSS, NCSS-FITC, and NCSS-FITC-AL structures. Especially Na and P originated from the carboxylate and bisphosphonate moieties.
characteristic bands appeared in the FTIR spectra of NCCS, NCCS-FITC, and NCCS-FITC-AL indicated the introduction of the carboxy ($\nu_{C=O}$ at $1406$ cm$^{-1}$), fluorescent ($\nu_{C=\pi}$ at $1450$–1600 cm$^{-1}$), and bisphosphonate ($\nu_{P=O}$ at $1108$ cm$^{-1}$) moieties on CS (Fig. S3†). The $^1$H NMR spectroscopic analysis of CS, NCCS, NCCS-FITC, and polyelectrolytic NCCS-FITC-AL further confirmed the modifications of CS by the signals indicating the introduction of the alkenyl and methylene groups (Fig. S4†). The introduction ratio of the N-citraconyl group was calculated to be $44\%$ to the amine groups from the integral ratio of the peaks assigned to the acetyl and alkenyl protons of NCCS. The introduction ratio of the bisphosphonate moieties was calculated to be $53\%$ with respect to the N-carboxycitraconyl group. The presence of the FITC residues in the NCCS-FITC-AL structure was also confirmed by an absorption band at $495$ nm observed in the UV-vis spectrum (Fig. S5†).

**In situ** aqueous synthesis of Fe$_3$O$_4$ nanoparticles and their concomitant surface functionalization with polyelectrolytic NCCS-FITC-AL

The charge-inversional Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles were prepared by a modified coprecipitation method (Scheme 2). This method involved with **in situ** production of Fe$_3$O$_4$ nanoparticles and concomitant surface functionalization with polyelectrolytic stabilizer, NCCS-FITC-AL, via nucleation, seed generation, and particle growth. When the solutions of the Fe(II) and Fe(III) precursors were added to the dispersion of CS, NCCS-FITC, and NCCS-FITC-AL, a coprecipitation-coating process occurred under microwave irradiation to yield Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles (Scheme 2).

**Scheme 1** Synthesis of NCCS-FITC-AL polyelectrolytic stabilizer from CS.

**Scheme 2** Schematic illustration for **in situ** aqueous synthesis of Fe$_3$O$_4$ nanoparticles and their concomitant surface functionalization with polyelectrolytic NCCS-FITC-AL by MW-assisted one-step coprecipitation-coating method.
NCCS-FITC-AL under N₂, the iron ions were accumulated to the carboxy and bisphosphonate moieties. Rapid nucleation of Fe₃O₄ particles was occurred by the addition of aqueous ammonia solution through hydrolysis and dehydration of the localized iron ions. Then, the dispersion was transferred to a microwave (MW) oven, where MW was irradiated to grow particles at 100 °C for 30 min. In this method, NCCS-FITC-AL served as a scaffold to control the growth of the nanoparticles. NCCS-FITC-AL can also limit the active mass transfer of iron over the overlapping with other bands such as those of the C–O and C–O–C moieties in NCCS-FITC-AL. Hence, the conjugation of functional groups like carboxylate and phosphonate to the Fe₃O₄ surface was confirmed by careful analysis on the comparison of the magnified spectra of Fe₂O₃-NCCS-FITC-AL and NCCS-FITC-AL (Fig. 1B, 700–1500 cm⁻¹). The stretching vibration for P=O in the phosphonate moieties in NCCS-FITC-AL was observed at 1108 cm⁻¹. When the phosphonate groups interacted with iron atoms, the stretching vibration for P=O significantly changed from 1108 to 1133 cm⁻¹. The stretching bands of P–OH were also significantly shifted from 1040 to 1033 cm⁻¹ and 906 to 902 cm⁻¹. In addition, a new peak appeared at 1030 cm⁻¹ assignable the stretching vibration mode of Fe–O–P indicating an effective conjugation of the phosphonate moieties with the surface iron atoms on the Fe₂O₃ nanoparticles. On the other hand, the stretching vibration bands of the carbonyl group in the carboxylate ions were also shifted from 1410 to 1396 cm⁻¹ and 1376 to 1372 cm⁻¹ due to the bidentate chelation to iron atoms on the surface of the Fe₃O₄ nanoparticles. In addition, the peak intensity of the absorption of the carboxylate ion was significantly lowered than that of the phosphonate structure, which demonstrated dominant coordination of the carboxylates to the Fe₂O₃ nanoparticles, and the bisphosphonate moieties mostly remained intact for targeting. The other peaks related to the structure of NCCS-FITC-AL also appeared in shifted positions. For instance, some medium–strong bands were observed at 1645, 1625, and 1617 cm⁻¹, and these bands are assignable to the stretching bands of amide-I and II and C=C overlapped with the bending bands of the –OH, >NH, and –NH₂ moieties. Two distinct sharp bands were visible at 2922 and 2853 cm⁻¹ assignable to the C–H stretching of the CH₂ groups in the amino sugar units and AL. A broad band appeared at 3000–3500 cm⁻¹ is assignable to the stretching vibrations of O–H, >NH, –NH₂, and –C–H bonds.

Characterization of Fe₃O₄-NCCS-FITC-AL nanoparticles

The formation of the charge-inversional Fe₃O₄-NCCS-FITC-AL nanoparticles was confirmed by FTIR and EDX (Fig. S6a†) spectroscopic, and thermogravimetric analyses (Fig. S6c†). A well-washed and fully dried powder sample was used for the FTIR spectroscopic analysis (Fig. 1A). The FTIR spectrum of Fe₃O₄-NCCS-FITC-AL shows two bands at 548 and 573 cm⁻¹ assignable to the stretching vibrations of the Fe–O bond in typical Fe₃O₄. It did not show the Fe–O bands at the higher wavenumber region originating from Fe₂O₃ observable in 600–690 cm⁻¹ (Fig. S6d†). This result demonstrates that Fe₂O₃ nanoparticles were predominantly formed over Fe₃O₄. The oxidation of the iron[II] ions in Fe₃O₄ did not occur by the reaction medium through the polydentate coordination of the carboxylate and bisphosphonate moieties to the Fe atoms on the Fe₃O₄ surface. In addition, NCCS-FITC-AL stabilized the carboxylate and bisphosphonate moieties to the Fe atoms on the Fe₃O₄ surface. In addition, NCCS-FITC-AL stabilized the carboxylate and bisphosphonate moieties to the Fe atoms on the Fe₃O₄ surface. In addition, NCCS-FITC-AL stabilized the carboxylate and bisphosphonate moieties to the Fe atoms on the Fe₃O₄ surface.

Morphology and core-structure of Fe₃O₄-NCCS-FITC-AL nanoparticles

The solid- and liquid-state morphologies and core-structure of the Fe₃O₄-NCCS-FITC-AL nanoparticles were analyzed by scanning electron microscopy (SEM), transmission electron microscopy (TEM), dynamic light scattering (DLS), and X-ray diffraction (XRD) (Fig. 2). The SEM image (Fig. 2A) revealed

![Fig. 1](image-url) Whole (A) and magnified (700–1500 cm⁻¹ region) (B) FTIR spectra of Fe₂O₃-NCCS-FITC-AL nanoparticles (a) and NCCS-FITC-AL (b).
that Fe₃O₄-NCCS-FITC-AL nanoparticles are singly dispersed and almost spherical. The average diameter of Fe₃O₄-NCCS-FITC-AL nanoparticles was calculated to be 27 nm, and the size-distribution histogram is shown in Fig. 2B. The well-dispersed state demonstrates that polyelectrolytic NCCS-FITC AL effectively covering the magnetic Fe₃O₄ core reduced the inter-particle attractions through steric and electrostatic repulsions. The morphology of the core of the Fe₃O₄-NCCS-FITC-AL nanoparticles was analyzed by TEM (Fig. 2D). The TEM image revealed almost similar shapes of the nanoparticles, while the obtained average core size is one-fourth of the size revealed by the SEM image showing the morphology of the particles covered by the polymeric coating in a dried state. Fig. 2C shows the DLS profiles of the Fe₃O₄-NCCS-FITC-AL nanoparticles before and after microwave treatment. The average hydrodynamic diameter (Dₜ) of the nanoparticles was estimated to be 160 nm (black) with a PDI of 0.30. The Dₜ of the Fe₃O₄-NCCS-FITC-AL nanoparticles was increased from 160 to 190 nm (red) after MW treatment and the PDI was also significantly lowered from 0.30 to 0.18. The increment of the size is ascribable to the growth of the particles during the MW-irradiation. The difference between the sizes observed by DLS and SEM is ascribable to the thickness of the swollen layer of NCCS-FITC-AL and the Brownian motion of the particles. XRD analysis was conducted to determine the crystal structure of the core component inside the Fe₃O₄-NCCS-FITC-AL nanoparticles. The XRD profiles of dried, uncoated Fe₃O₄ and Fe₃O₄-NCCS-FITC-AL nanoparticles are illustrated in the 2θ range of 10–70° (Fig. S6b†). The XRD profile of uncoated Fe₃O₄ showed six reflections centered at 30.1°, 35.5°, 43.3°, 53.6°, 57.4°, and 62.8° assignable to the (220), (311), (400), (422), (511), and (440) lattice planes, respectively, of the cubic spinel Fe₃O₄ with the lattice constant a = 8.4040 Å (JCPDS 85-1436). Almost identical patterns were observed for the Fe₃O₄-NCCS-FITC-AL nanoparticles without any additional reflections assignable to any other iron species generated via oxidation during the preparation and storage. The lattice constant for Fe₃O₄-NCCS-FITC-AL nanoparticles was calculated to be 8.3895 Å, which is closer to the lattice constant of bulk magnetite (8.3960 Å), indicating that the in situ prepared Fe₃O₄ nanoparticles were effectively stabilized by polyelectrolytic NCCS-FITC-AL without altering the crystal structure of the Fe₃O₄ core. This result manifests the presence of single-phase-core-Fe₃O₄ inside the Fe₃O₄-NCCS-FITC-AL nanoparticles. Crystallinity and structure of the core component of the Fe₃O₄-NCCS-FITC-AL nanoparticles were further analyzed by high-resolution TEM (HRTEM). Fig. 2E shows a typical HRTEM image showing vividly clear lattice fringes. The interplanar distance is 0.26 nm, which is assignable to the (311) lattice spacing of cubic spinel Fe₃O₄. FTIR, XRD data and TEM analysis indicated the presence of spinel Fe₂O₃ rather than the defective spinel γ-Fe₂O₃ inside the Fe₃O₄-NCCS-FITC-AL nanoparticles.

**Magnetism of Fe₃O₄-NCCS-FITC-AL nanoparticles**

The magnetic response of the dispersed Fe₃O₄-NCCS-FITC-AL nanoparticles in Dulbecco's phosphate-buffered saline (DPBS) was tested using a permanent magnet. The Fe₃O₄-NCCS-FITC-AL nanoparticles are fully dispersible in the absence of external magnetic fields even at the concentration as high as 70 mg mL⁻¹ (Fig. 3A). By contrast, in the presence of a strong...
The nanoparticles were separated completely within a few minutes (Fig. 3B), while bare Fe₃O₄ nanoparticles were instantaneously separated. The excellent dispersity of the Fe₃O₄-NCCS-FITC-AL nanoparticles arises from the polyelectrolytic stabilizer, which effectively stabilizes in situ synthesized Fe₃O₄ while weakens the response towards the external magnetic field. The magnetic property of the charge-inversional Fe₃O₄-NCCS-FITC-AL nanoparticles in the dry-state was also analyzed by vibrating sample magnetometry (VSM) at room temperature (Fig. 3C). The M–H curve of the Fe₃O₄-NCCS-FITC-AL nanocomposite particles showed an open hysteresis loop with negligible remanence and coercivity, indicating the superparamagnetic, single domain, and highly crystalline nature of the charge-inversional Fe₃O₄-NCCS-FITC-AL nanoparticles. The saturation magnetization (Mₛ) value of Fe₃O₄-NCCS-FITC-AL prepared by the MW treatment is 59 emu g⁻¹ in terms of the relative weight composition of Fe₃O₄. This value is higher than previously reported Fe₃O₄ nanocomposites coated with small molecules, carboxy-CS (42 emu g⁻¹) and a cationic polyelectrolyte (46 emu g⁻¹) prepared without using MW treatment, while it is still lower than that of the typical bulk Fe₃O₄ (98 emu g⁻¹). The higher Mₛ value of the composite particles is ascribable to the presence of the iron phosphonate structures on the surface of the particles, which were secured more stabilized spin contributions than that of the magnetically inactive dead surface layer. The Mₛ augmentation has probably originated from the high crystallinity of the Fe₃O₄ core. The obtained result validates that the charge-inversional Fe₃O₄-NCCS-FITC-AL nanoparticles have enough magnetic efficacy for magnetically-induced manipulation for in vivo applications. This MW-assisted synthesis achieves both the excellent magnetic property and the high physiological stability discussed later, which are prerequisites for bio-related applications.

Optical properties of Fe₃O₄-NCCS-FITC-AL nanoparticles

The optical behaviors of the charge-inversional Fe₃O₄-NCCS-FITC-AL nanoparticles were analyzed by UV-vis absorption, fluorescence excitation and emission spectroscopies (Fig. 4). The polyelectrolytic stabilizer, NCCS-FITC-AL, shows a strong absorption band at 495 nm in the UV-vis spectrum. This clear band originates from the FITC moieties of the stabilizer (Fig. S5†). The UV-vis spectrum of Fe₃O₄-NCCS-FITC-AL also showed a shoulder peak around 494 nm for the absorption of Fe₃O₄ core, while the intensity of the absorption by the FITC moieties was lower than that of the polyelectrolytic polymer dispersion due to the lower relative content of the FITC moieties in the composite. The emission spectrum of Fe₃O₄-NCCS-FITC-AL showed an emission peak with the maximum at 516 nm by excitation at 500 nm. Both the emission and excitation peaks accord with the typical fluorescent behavior of the FITC moieties even in the presence of the carboxy and bisphosphonate moieties and the Fe₃O₄ core. The observable fluorescence is sufficient for imaging as described later. This finding agrees well with previous reports. The pH-dependent aggregation-dispersion, charge-inversion, and physiological stability of Fe₃O₄-NCCS-FITC-AL nanoparticles

The aggregation-dispersion and charge-inversional behaviors of the Fe₃O₄-NCCS-FITC-AL nanoparticles were examined at a controlled pH range of 3.5–9.0 (Fig. 5). The Fe₃O₄-NCCS-FITC-AL nanoparticles were initially well-dispersed in DBPS at the examined pH range (Fig. 5A), while selectively aggregated only at pH 5.0 after 2.5 h (Fig. 5B). This pH responsibility was further studied by DLS. The initial Dₕ of Fe₃O₄-NCCS-FITC-AL was almost identical (190–200 nm) regardless of the pH values, and the Dₕ at pH below 4.5 and above 6.5 was stable. By contrast, the Dₕ at the pH range of 5.0–6.0 was gradually increased during the
incubation. The increment of $D_h$ at pH 5.0 was most significant, and the $D_h$ after 2.5 h reached 690 nm. After that, the nanoparticles were fully settled down and the size exceeded the measurable limit of DLS. This pH-dependent variation of $D_h$ can be explained by the $\zeta$-potential (Fig. 5C). At the pH range of 3.5–4.5, the Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles have positive $\zeta$-potentials stabilizing the dispersion by electrostatic repulsions of the ammonium and carboxy groups formed by the protonation of the amine and carboxylate groups. By contrast, at pH 5.0 where the Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles were selectively precipitated and pH 5.5 and 6.0 where the $D_h$ was increased, the $\zeta$-potentials became less positive close to zero charge. The precipitation and aggregation of the nanoparticles occurred by the inter-particle attractions among the zwitterionic surface structure. The $\zeta$-potentials were decreased as the increase of the pH values due to the partial deprotonation of the ammonium structure transforming into the amine groups. The weakly positive $\zeta$-potentials in this precipitating region, not isoelectric and higher than those of our previously reported material\textsuperscript{56} implies that a part of the cationic groups does not contribute to the precipitation behavior. A plausible reason is the lower concentration of the free bisphosphoric and carboxyl groups around the surface contributing to the ionic cross-linkage due to the ligation to the Fe$_3$O$_4$ core as suggested by the FTIR spectroscopic study. Above pH 6.0, the Fe$_3$O$_4$-NCCS-FITC-AL nanocomposite particles showed negative $\zeta$-potential due to the deprotonation of carboxy and phosphonic groups to form carboxylate and phosphonate groups, respectively, resulting in stable dispersion by the ionic repulsions between the nanoparticles. This pH-dependent charge-conversion property of the nanoparticles having targetable bisphosphonate moieties would be beneficial for selective targeting of acidic bone cells by adhering to the negatively charged cell surface and endocytosis. The physiological stability of the charge-inversional Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles dispersed in saline was confirmed by studying their $D_h$ and PDI at physiological conditions (pH 7.4, 37 °C) for 3 weeks (Fig. 5E). Both the $D_h$ (~200 nm) and PDI (below 0.20) of the particles were not significantly changed throughout the observation period because of the electrostatic repulsions among the negatively charged carboxylate and phosphonate moieties of the polyelectrolytes. This result demonstrates that this magnetic and fluorescent nanosystem has capabilities of prolonged circulation and selective deposition in locally acidic environments.

Hemocompatibility of Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles

Hemocompatibility is one of the important prerequisites of biomaterials for intravenous applications. The concentration-dependent hemolytic behaviour of Fe$_3$O$_4$-NCCS-FITC-AL was

Fig. 5  Photo images of pH-dependent aggregation-dispersion (A and B), the pH-dependence of $D_h$ (C), and $\zeta$-potential (D) of Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles in DPBS at 25 °C; and time-course of $D_h$ of Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles in saline (pH = 7.4) at 25 °C (E).
examined using sheep erythrocytes in saline at pH 7.4 and 37 °C for 5 h. Sheep erythrocytes dispersed in a 1% aqueous solution of Triton X 100 and saline were employed as the positive and negative controls, respectively (Fig. 6A). The treatment with Triton X-100 completely hemolyzed the erythrocytes through membrane disruption, and the color was turned into dark red, while the red color of the erythrocytes was retained in the negative control sample. The colors of the erythrocyte suspensions containing Fe₃O₄-NCCS-FITC-AL gradually became slightly darker by trace degrees of hemolysis enhanced as the increase of the concentration of Fe₃O₄-NCCS-FITC-AL (Fig. 6A). At pH 7.4, Fe₃O₄-NCCS-FITC-AL nanoparticles showed only 0.21% hemolysis at 100 μg mL⁻¹, while they showed hemolysis approximately three times higher at 400 μg mL⁻¹ (Fig. 6C). The hemolysis degrees of Fe₃O₄-NCCS-FITC-AL are significantly lower than that of previously reported magnetic nanocomposite particles coated with cationic polyelectrolyte,⁴⁸ carboxy-CS,²⁸ and functionalized carboxy-CS.¹⁸ The morphological change of the sheep erythrocytes in the presence and absence of Fe₃O₄-NCCS-FITC-AL was confirmed by optical microscopy (Fig. 6B). Most of the erythrocytes incubated with Fe₃O₄-NCCS-FITC-AL retained their native morphologies in the same manner with the negative control (Fig. S8†). The improved hemocompatibility of Fe₃O₄-NCCS-FITC-AL plausibly originates from the weakly negative surface charge, which is smaller than natural erythrocytes, resulting in negligible interactions with the negative membrane surface of erythrocytes under these conditions. The hemocompatibility of Fe₃O₄-NCCS-FITC-AL meets the acceptable limit of blood-contacting biomaterials (5%) under the physiological conditions,²⁷,²⁸ indicating the potential of Fe₃O₄-NCCS-FITC-AL for intravenous applications.

**pH-triggered membrane disruption of erythrocytes by Fe₃O₄-NCCS-FITC-AL nanoparticles**

The pH- and concentration-dependent membrane disruption abilities of the charge-inversional Fe₃O₄-NCCS-FITC-AL nanoparticles were tested by hemolysis assay using sheep erythrocytes as a model cell (Fig. 7). The optical images (Fig. 7A) show the relative colour changes of the erythrocytes dispersion with the variations of pHs and concentrations of Fe₃O₄-NCCS-FITC-AL. Erythrocytes dispersed in 1% Triton X100, which fully lysed the erythrocytes was used as positive controle. While erythrocytes dispersed in saline having minimal lysis was regarded as negative controle. The red coloured-dispersion of Fe₃O₄-NCCS-FITC-AL gradually turned into black with decreasing pHs and increasing nanoparticle concentrations. The hemolysis degree of the erythrocytes by the Fe₃O₄-NCCS-FITC-AL nanoparticles was increased with the decrease of pH. This enhancement of hemolysis is ascribable to the decrease in

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Fig. 6 Optical images of sheep erythrocytes dispersed in saline containing Fe₃O₄-NCCS-FITC-AL nanoparticles (A), optical microscopic photograph of sheep erythrocytes dispersed in saline containing 400 μg mL⁻¹ Fe₃O₄-NCCS-FITC-AL nanoparticle (B), and concentration dependence of hemolysis of sheep erythrocytes dispersed in saline with Fe₃O₄-NCCS-FITC-AL (C) (concentration of erythrocyte = 200 μL mL⁻¹, pH = 7.4, incubation time = 5 h, 37 °C).
the negative surface charge of the nanoparticles that lead to enhanced adherence to the erythrocytes membrane surface. The hemolysis degree became significantly higher (18.7%) at the pH range of 5.5 to 5.0, where the negative surface charge of the Fe₃O₄-NCCS-FITC-AL nanocomposite particles changes to positive (Fig. 7B). The positive charge originates from the protonation of the acid-sensing secondary amine, carboxy, and phosphonate groups, enhancing electrostatic interactions with the negatively charged lipid membrane of the erythrocytes under the acidic environments. Another plausible reason for the increase of the hemolysis degree is the hydrophobic interactions between the pendant hydrophobic structures in the polyelectrolyte coating with the lipid membrane of the erythrocytes. At pH 5.0 in the presence of the charge-inversional Fe₃O₄-NCCS-FITC-AL nanoparticles, the erythrocyte membranes were significantly damaged and the erythrocytes were obviously agglutinated (Fig. 8). This pH-sensitive membrane disruption suggests the acidity-responsible membranolytic ability of Fe₃O₄-NCCS-FITC-AL nanoparticles at the pH range from 5.0 to 5.5 identical to the endosomal pH. Similar behaviour was also reported for MIONPs for selective disruption of cancer cells only under the acidic pH.

**pH-dependent bone mineral affinity of Fe₃O₄-NCCS-FITC-AL**

The charge-inversional Fe₃O₄-NCCS-FITC-AL nanoparticles were designed for bone-seeking diagnosis and therapeutic applications. Therefore, we investigated the pH-dependent affinity of Fe₃O₄-NCCS-FITC-AL towards bone minerals. HAp with a monoclinic structure having 2.5 µm of the average length was used as a model bone mineral. HAp has the chemical formula of 3Ca₃(PO₄)₂·Ca(OH)₂, and its surface composes of Ca²⁺, OH⁻, and PO₄³⁻ ions with the protonated phosphate ions depending on pH and coexisting ions. HAp and Fe₃O₄-NCCS-FITC-AL were incubated at 37 °C in saline and the dispersion pH was adjusted to acidic (5.0) and physiological (7.4) conditions. The adhesion ability of the Fe₃O₄-NCCS-FITC-AL nanoparticles to HAp surface was studied by SEM and EDX analyses (Fig. 9). Blank experiments without Fe₃O₄-NCCS-FITC-AL were also performed at both pHs of 5 and 7.4 to observe the effects of the acidic environment (H⁺ rich) and saline (Na⁺ and Cl⁻ ions) on the surface of HAp. Any changes caused by these ions were not observable for both of the control samples (Fig. 9A and C). In addition, adhered particles were not found in the SEM image.
of HAp treated with Fe₃O₄-NCCS-FITC-AL under the physiological conditions (Fig. 9D), which indicates the very poor affinity to HAp. This low affinity is plausibly due to the ionic repulsions between the negative HPO₄²⁻ moieties on the surface of HAp and the negatively charged Fe₃O₄-NCCS-FITC-AL under physiological conditions.⁶⁰,⁶¹ The content of iron on the treated HAp surface was calculated to be below 0.01 wt% from EDS. The SEM image of HAp treated with Fe₃O₄-NCCS-FITC-AL under the acidic pH (5.0) shows many adhered particles indicated by blue arrows (Fig. 9B). The existence of Fe₃O₄-NCCS-FITC-AL was also confirmed by EDS analysis, and the Fe content on the surface of HAp was calculated to be 1.4 wt%. The increased affinity of Fe₃O₄-NCCS-FITC-AL toward HAp can be explained by the strong electrostatic interactions and hydrogen bonding. The surface of HAp under acidic conditions is covered by H₂PO₄⁻ groups, in which the P=O remains facing-up and Ca²⁺ ions are unveiled by the dissolution of anions.⁶⁰-⁶² The Ca²⁺ ions on the HAp surface electrostatically attract the carboxylate and bisphosphonate moieties of the Fe₃O₄-NCCS-FITC-AL nanoparticles. Similar electrostatic interactions were also observed in the previously reported materials bearing bisphosphonate moieties.⁶¹,⁶³-⁶⁵ In addition, the ammonium and P–OH groups present on the surface of the charge-inverted Fe₃O₄-NCCS-FITC-AL nanoparticles also contributed to the attraction by the electrostatic and hydrogen-bonding interactions with the surface groups like H₂PO₄⁻, PO₄³⁻ and P=O moieties of HAp.⁶¹,⁶⁵

This stronger adherence under the acidic conditions, approximately 140% higher affinity than that under the physiological conditions (Fig. S10†), is plausibly ascribed to both the physisorption and chemisorption of the nanoparticles on the HAp surface (Fig. 10). The surface of Fe₃O₄-NCCS-FITC-AL is covered by the anionic bisphosphonate and carboxylate ions and cationic quaternary ammonium ions that electrostatically interact with Ca²⁺ and H₂PO₄⁻ on the surface of HAp, respectively, even though the zeta potential is not so high due the buffering ability of the polyampholytic skeleton. In addition, the dissolution–deposition equilibrium of the phosphonate ions of HAp results in the incorporation of the bisphosphonate groups of Fe₃O₄-NCCS-FITC-AL into the lattice structure of HAp (Fig. 10, right part). These interactions of phosphonates with HAp were reported in previous NMR spectroscopic (³¹P, ¹⁵N, ¹³C, ²H, and ¹H) and chromatographic studies.⁶¹,⁶³,⁶⁶,⁶⁷ This finding revealed that the designed nanosystem has adequate potentials for selective targeting of acidic bone sites.

Affinity of Fe₃O₄-NCCS-FITC-AL nanoparticles towards native bone sample

The affinity and adherence of the charge-inversional Fe₃O₄-NCCS-FITC-AL nanoparticles towards native bone samples were studied by SEM analysis. Bovine leg bone samples were treated with Fe₃O₄-NCCS-FITC-AL at pH 7.4 and 5.0 (Fig. 11). The optical and SEM images of the bone sample are shown in
Both the optical and SEM images of the reference bone showed smooth surface morphology and the absence of any deposited materials except some cracks. The SEM image of the bone sample incubated with Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles at pH 7.4 showed almost identical morphology, and Fe$_3$O$_4$-NCCS-FITC-AL adhered was not observable (Fig. 11C). The poor affinity is due to the electrostatic repulsions between the negatively charged Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles and the bone surface under the physiological conditions. By contrast, at pH 5.0, Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles were adhered on the bone surface, which was almost covered by these nanoparticles (Fig. 11D). This pH dependence, namely high affinity only under acidic conditions, is identical to HAp employed as the model system, while the enhancement in the affinity to the bone under the acidic conditions is more significant than that to HAp. Bone mainly consists of HAp and collagen, whose solubility is higher at pH 5.0 by the positively charged nature.

Fig. 10 Schematic representation of the bindings of Fe$_3$O$_4$-NCCS-FITC-AL nanoparticles to HAp.
the dissolution of charged collagen is a plausible factor for higher affinity. In metastasized bone sites, degradation of collagen is enzymatically promoted, and this acidity-sensitive adsorption is presumably more enhanced. Osteoclasts become activated by cancers and overexpress cathepsin-K degrading collagen fibers networks.\(^5\) This synergy of the pH-triggered charge-conversion and targetable bisphosphonate moieties in the Fe\(_3\)O\(_4\)-NCCS-FITC-AL nanoparticles enabled selective accumulation on HAp unveiled under acidic conditions.

**Conclusions**

In this report, we demonstrated a magnetic nanomaterial, Fe\(_3\)O\(_4\)-NCCS-FITC-AL having pH-controlled hemocompatibility and bone-targeting ability. Fe\(_3\)O\(_4\)-NCCS-FITC-AL was prepared by a simple one-step aqueous process called the *in situ* coprecipitation-coating method without requiring any tedious post modification, toxic organic solvents, capping ligand, stabilizer, and emulsifier. Fe\(_3\)O\(_4\)-NCCS-FITC-AL consists of spherical cores of Fe\(_3\)O\(_4\) with sizes around 10 nm, and the \(D_v\) of 160 nm under physiological conditions suits well for circulation. Fe\(_3\)O\(_4\)-NCCS-FITC-AL is paramagnetic and magnetically guidable. While Fe\(_3\)O\(_4\)-NCCS-FITC-AL is highly dispersible under physiological conditions, it is selectively precipitated at pH 5.0. This pH-dependent change in the dispersibility originates from the charge conversion of the acidic moieties. The hemolytic assay demonstrated excellent hemocompatibility of Fe\(_3\)O\(_4\)-NCCS-FITC-AL at physiological conditions due to the presence negatively charged biocompatible coating. But, Fe\(_3\)O\(_4\)-NCCS-FITC-AL becomes highly hemolytic under acidic conditions owing to the charge reversion ability of the polyelectrolyte. Fluorescent imaging confirmed adherence of Fe\(_3\)O\(_4\)-NCCS-FITC-AL to the membranes of erythrocytes only under acidic conditions. The interactions of Fe\(_3\)O\(_4\)-NCCS-FITC-AL particles with a bone sample and HAp changed with pH-variations in a similar manner with the hemolytic behavior. At pH 7.4, the negatively charged Fe\(_3\)O\(_4\)-NCCS-FITC-AL particles did not interact with HAp and bone sample due to the strong electrostatic repulsions, suggesting the minimal systemic and off-targeted side effects. At acidic pH (5.0), the charge-inverted Fe\(_3\)O\(_4\)-NCCS-FITC-AL was adhered on HAp and the bone sample by the bone-seeking bisphosphonate moieties and the electrostatic interactions, implying the possibility of the selective targeting of malignant bone tissues. Therefore, designed
**FeO$_2$-NCCS-FITC-AL particle** is a promising candidate for early-stage diagnosis and therapy of bone malignancies.

**Experimental section**

**Material and methods**

**Materials.** CS ($M_w = 5.6 \times 10^5$ g mol$^{-1}$), CAn, and AL were purchased from Tokyo Chemical Industry (Tokyo, Japan). FeSO$_4$·7H$_2$O and FITC were purchased from Sigma Aldrich (St. Louis, MO, USA). NH$_4$OH (28%) and FeCl$_3$·6H$_2$O were purchased from Kanto Chemical (Tokyo, Japan). HAp was purchased from Wako Pure Chemical (Osaka, Japan). DPBS was purchased from Gibco Life Technologies (Paisley, UK). Bovine leg bone was purchased from Baticrom Halal Shop (Tokyo, Japan). All other reagents used in this study were lab grade and used without further purification. Water was purified using a Nomura Micro Science (Kanagawa, Japan) MINIPURE TW-300RU water purification system. Deionized distilled water (DDW) was used throughout the study.

**Measurements.** FTIR spectra were documented on a JASCO (Tokyo, Japan) FTIR-460 plus spectrometer using pressed KBr pellets with a scan rate of 4 cm$^{-1}$sec$^{-1}$. £H NMR spectra were recorded on a JEOL (Tokyo, Japan) ECX-400 instrument using tetramethylsilane as an internal standard (400 MHz) at room temperature. Hydrodynamic diameters were measured by DLS on a Malvern Instrument (Malvern, UK) Zetasizer Nano ZS. The temperature. Hydrodynamic diameters were measured by DLS at an accelerating voltage of 30 kV. The powder X-ray diffraction (XRD) patterns were measured on a Rigaku (Tokyo, Japan) Ultima IV RINT D/max-KA diffractometer with Cu Kz radiation ($\lambda = 1.54178$ Å), TEM images were taken using a JEOL (Tokyo, Japan) TEM-2100F field emission scanning electron microscope. Thermogravimetric analysis was carried out on a Seiko (Tokyo, Japan) TG/DSC 6300 (EXSTER6000) at a heating rate of 10 °C min$^{-1}$ under N$_2$. The magnetic properties of the nanoparticles were analyzed on a Riken Denshi (Tokyo, Japan) BHV-30 series vibrating sample magnetometer at ambient conditions. Energy-dispersive X-ray spectra were recorded on a JEOL (Tokyo, Japan) JSM-6510A analytical scanning electron microscope. The optical absorbance was taken using an AS ONE ASV11D UV-visible spectrophotometer. The erythrocytes specimens were observed with an Olympus (Tokyo, Japan) CKX53 microscope. The color pictures of the specimens were captured using a Visualix Pro2 (Kobe, Japan) camera. All images were taken at the same magnification (20×).

**Methods**

**Synthesis of bisphosphonated polyelectrolytic CS derivative:**

$N$-carboxyitraconylation of CS (NCCS). CS was dissolved in 200 mL of aqueous acetate buffer (pH 4.6) by magnetic stirring overnight. The solution was adjusted at pH 5.6 using dilute aqueous NaOH solution. CAn was diluted in methanol (10 mL) and dropwise added to CS solution for 1 h. The reaction was continued for 12 h at room temperature. After the reaction, 10 M NaOH aqueous solution was added to the reaction mixture and stirred for an additional 10 min. The reaction mixture was added dropwise to an excess amount of acetone to precipitate NCCS. The precipitate was separated using a Buchner funnel. The resulting product was purified via dispersing in DDW and reprecipitated in acetone. The purified NCCS was dried in a vacuum desiccator and stored in a refrigerator.

**FITC labeling of NCCS.** NCCS (800 mg) was dissolved in DDW (300 mL). FITC (4.00 mg) was dissolved in 10 mL methanol and the solution was added dropwise to the mixture for 1 h. The reaction was continued in dark conditions for 4 h at room temperature. The reaction mixture was added dropwise to an excess amount of methanol to precipitate the NCCS-FITC. The resulting product was purified by centrifugation at 4000 rpm and washed repeatedly until the decant became completely colorless. The obtained NCCS-FITC was dried in a desiccator in dark and stored in a freezer at $-4$ °C.

**Bisphosphonylation of NCCS-FITC.** In dark conditions, NCCS-FITC was dissolved in DDW (200 mL) by magnetic stirring at room temperature. AL (100 mg) was dissolved in 10 mL DDW and the solution was added dropwise to the dispersion of NCCS-FITC. The reaction was performed at room temperature in dark conditions for 24 h. After the reaction period, the product was precipitated by pouring the mixture into an excess amount of acetone and separated by filtration. The resulting product was purified by dispersing in DDW and reprecipitating in methanol. Bisphosphonated polyelectrolytic NCCS-FITC-AL was obtained by drying in a vacuum desiccator (yield = 85%) and kept at $-15$ °C.

**MW-assisted in situ synthesis and surface functionalization of FeO$_2$ nanoparticles.** Bisphosphonated polyelectrolytic NCCS-FITC-AL was dissolved in deoxygenated DDW (175 mL) by magnetic stirring at room temperature. FeSO$_4$·6H$_2$O (270 mg, 1.00 mmol) and FeCl$_3$·6H$_2$O (139 mg, 0.50 mmol) were dissolved in 15 mL DDW with purging N$_2$. These solutions were mixed in a three-necked round bottom flask (300 mL) under N$_2$. The flask was placed in a preheated thermostat oil bath maintained at 80 °C. Aqueous ammonia solution (5 mL, 28%) was injected at a rate of 1 mL min$^{-1}$. The mixture was magnetically stirred for 10 min. The reaction mixture was then, quickly transferred into a 500 mL conical flask. The flask was placed in a MW oven to be heated at 100 °C for 30 min. The resultant black precipitate was magnetically washed repeatedly with DDW until the supernatant became neutral pH. The solid precipitate was separated at the bottom of the flask and dried in a vacuum desiccator for 24 h. Bare FeO$_2$ was synthesized following the aforementioned procedure without adding the stabilizer, NCCS-FITC-AL.

**Hemocompatibility of FeO$_2$-NCCS-FITC-AL.** Hemolysis activities of the FeO$_2$-NCCS-FITC-AL nanoparticles were assayed by using the following protocol. Preserved sheep whole blood (40 mL) was centrifuged at 2500 rpm for 5 min. The sheep erythrocytes were separated by centrifugation and plasma removal and washed three times with saline (150 mM NaCl aq.). The washed erythrocytes were redispersed in saline (40 mL). The nanoparticles were also dispersed in saline and the dispersion was mixed with erythrocytes suspension. The pH of the mixtures was adjusted to 7.4 using 0.01 M HCl aq. The final
concentrations of the Fe₃O₄-NCCS-FITC-AL nanoparticles were adjusted to 100, 200, and 400 µg mL⁻¹. Triton X-100 (1% aqueous solution) and saline were added to the erythrocytes suspension to prepare the positive (100% lysis) and negative (0% lysis) control samples, respectively. All the samples and controls were incubated for 5 h at 37 °C. The samples were gently swirled once per 20 min for resuspending the erythrocytes. After incubation, the suspension was centrifuged at 10 000 rpm for 5 min to separate the erythrocytes and the supernatants were incubated for 30 min at room temperature to oxidize the released hemoglobin. The optical absorbance of oxyhemoglobin was assessed by absorption at 540 nm. The hemolysis percentage of erythrocytes was calculated using the following equation:

\[
\% \text{ hemolysis} = \left( \frac{\text{Abs}_{\text{sample}} - \text{Abs}_{\text{negative control}}}{\text{Abs}_{\text{positive control}} - \text{Abs}_{\text{negative control}}} \right) \times 100
\]

Membrane morphology study of erythrocytes. The morphology of erythrocytes was studied using a light microscope. The erythrocytes were added to saline containing the charge-conversional Fe₃O₄-NCCS-FITC-AL nanoparticles at 400 µg mL⁻¹, and the pH values were set to 7.4 and 5.0 using 0.01 M HCl aqueous solution. The resulting dispersions were incubated at 37 °C for 5 h, and mildly shaken once every 20 min. After the incubation period, the dispersions were treated with a magnet to remove free nanoparticles. Then, the erythrocytes were cleaned by centrifugal washing and resuspended in saline. A control sample was also made without using nanoparticles. The washed erythrocytes suspension (10 µL) was placed on a glass slide and covered with a coverslip glass. The resulting specimens were observed microscopically.

pH-selective adhesion of Fe₃O₄-NCCS-FITC-AL to HAp and native bone sample. Fe₃O₄-NCCS-FITC-AL nanoparticles (500 µL) were dispersed in saline (10 mL) and the pH values of the dispersions were adjusted to 7.4 and 5.0 using 0.01 M HCl aqueous solution. HAp (20 mg) was added to each dispersion and dispersed by mild shaking. The dispersions were incubated at 37 °C for 3 h and shaken once every 20 min for redispersion. After incubation, the HAp particles were separated by centrifugation at 2000 rpm and washed 7 times with saline. Finally, the washed HAp was analyzed by SEM. For the native bone sample, a small piece of clean bone was added to the dispersion instead of HAp, incubated under the aforementioned conditions, and washed 7 times with saline and water to remove the free and loosely bound nanoparticles. The control bone samples were made without using Fe₃O₄-NCCS-FITC-AL nanoparticles. All the bone samples were also analyzed by SEM and EDS.

Author contributions
M. A. Rahman conceptualized the idea and did formal experiments. He collected and analysed the data, and also explained the results. This research was supervised by B. Ochiai. The manuscript was drafted by M. A. Rahman and revised by B. Ochiai. The final version of the manuscript has approved by both the authors.

Conflicts of interest
The authors declare no competing financial interests that would have been arisen to influence the work reported in this paper.

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