A review of traditional pharmacological uses, phytochemistry, and pharmacological activities of *Tribulus terrestris*

Wenyi Zhu, Yijie Du, Hong Meng, Yinmao Dong and Li Li*

**Abstract**

*Tribulus terrestris* L. (TT) is an annual plant of the family Zygophyllaceae that has been used for generations to energize, vitalize, and improve sexual function and physical performance in men. The fruits and roots of TT have been used as a folk medicine for thousands of years in China, India, Sudan, and Pakistan. Numerous bioactive phytochemicals, such as saponins and flavonoids, have been isolated and identified from TT that are responsible alone or in combination for various pharmacological activities. This review provides a comprehensive overview of the traditional applications, phytochemistry, pharmacology and overuse of TT and provides evidence for better medicinal usage of TT.

**Keywords:** Tribulus terrestris, Traditional uses, Phytochemical, Pharmacological activities

**Introduction**

TT is an annual plant of the family Zygophyllaceae, which is commonly known as *Tribulus*, Hard thorns, and goat head in China. It is mainly planted in the Mediterranean and in sub-tropical regions such as India, China, South America, Mexico, Spain, Bulgaria, and Pakistan. It is a small, prostrate, 10–60 cm high, hirsute or silky hairy shrub. The leaves are opposite, often unequal, paripinnate, pinnate from 5 to 8 pairs and elliptical or an oblong lanceolate. The fruits from the five mericarps are ax-shaped, 3–6 mm long, and arranged radially and have a diameter of 7–12 mm and a hard texture. The root is slender, fibrous, cylindrical and frequently branched, bears a number of small rootlets and is light brown in colour [1]. The fruits and roots of TT, as a folk medicine, have been used for thousands of years in China. Over the last several years, it has been certified for its pharmaceutical activities for improving sexual function and cardiac protection and providing anti-urolithic, antidiabetic, anti-inflammatory, anti-tumour and antioxidants effects. In the current review, we present and analyse the ethnobotanical use and the phytochemical and pharmacological activities of TT. These up-to-date research observations will be helpful in understanding the characteristics and superiorities of this traditional Chinese medicine and will be applicable in developing new products and herbal medicines in the future.

**Traditional pharmacological uses**

TT is native to south-eastern and Mediterranean Europe, temperate and tropical Asia and Africa, and northern Australia. The use of TT from ancient times occurred in the traditional medicine of major cultures in these geographical areas, such as traditional Chinese medicine, traditional Indian medicine (Ayurveda), and the traditional medicine of south-eastern Europe, and this has defined its ethnopharmacological relevance as a medicinal plant [2]. As a traditional Chinese Medicine, it was listed as a top grade medicine in the earliest extant Chinese pharmaceutical monograph “Shen Nong Ben Cao Jing” [3]. In Chinese Pharmacopoeia [4], the fruits of TT have been used for tonifying the kidneys and as a diuretic and cough expectorant that improves eyesight and for the treatment of skin pruritus, headache and vertigo, and mammary duct blockage. In India, the fruits have been used in the treatment of infertility, impotence, erectile dysfunction and low libido in Ayurveda. In addition, the roots and fruits are considered to
have cardiotoxic properties [5]. In Sudan, TT has been used as demulcent and in nephritis and the treatment of inflammatory disorders [5]. In addition, it has been used for diuretic and uricosuric effects in Pakistan [6]. Modern investigation showed that the chemical constituents steroidal saponins and flavonoids with the prominent anti-inflammatory and antiaging activities of TT were the main contributors to the traditional pharmacological activities.

**Phytochemical investigations**

Many different compounds with a variety of biological properties and chemical structures have been identified from TT, including steroidal saponins, flavonoids, glycosides, phytosterols, tannins, terpenoids, amide derivatives, amino acids, and proteins. Among the different types of constituents, steroidal saponins and flavonoids are considered to be the most important metabolites with various bioactivities.

**Steroidal saponins**

Spirostanol and furostanol saponins are considered the most characteristic chemicals in TT. To date, 108 kinds of steroidal saponins have been isolated from TT (1–108). Among them, there are 58 kinds of spirostane saponins (1–58) and 50 kinds of furostane saponins (59–108). The steroidal saponins, such as protodioscin and protogracilin, are thought to confer TT unique biological activities. Skeletal types of steroidal aglycones in TT are shown in Figs. 1, 2. Steroidal saponins(aglycones) in TT are shown in Table 1.

**Flavonoids**

The flavonoids of TT are mainly derivatives of quercetin, kaempferol and isorhamnetin. Quercetin (109), isoquercitrin (110), rutin (111), quercetin-3-O-gent (112), quercetin-3-O-gentr (113), quercetin-3-O-rha-gent (114), quercetin-3-O-gent-7-O-glu (115) are flavonoids with quercetin as the basic parent structure [34–36]. Isorhamnetin (116), isorhamnetin-3-O-glu (117), isorhamnetin-3-O-gent (118), isorhamnetin-3-O-rutinoside (119), isorhamnetin-3-O-gentr (120), isorhamnetin-3,7-di-O-glu (121), isorhamnetin-3-O-p-coumarylglu (122), isorhamnetin-3-O-gent-7-O-glu (123), isorhamnetin-3-O-gentr-7-O-glu (124) are flavonoids with isorhamnetin as the basic parent structure [30, 32, 37]. Kaempferol (125), kaempferol-3-O-glu (126), kaempferol-3-O-gent (127), kaempferol-3-O-rutinoside (128), kaempferol-3-O-gent-7-O-glu (129), tribuloside (130) are flavonoids with kaempferol as the basic parent structure [35, 36, 38, 39]. Structures of flavonoids in TT are shown in Fig. 3.

**Alkaloids**

Tribulusamide C (131), tribulusterine (132), tribulusin A (133), harmine (134), harman (135), harmhol (136), tribulusimide C (137), terrestriamide (138), N-trans-coumaroilytraine (139), N-trans-cafeoilytraine (140), terrestribisamide (141) are the main alkaloids isolated from the stems, leaves, and fruits of TT [40–45]. The nuclear mainly belong to β-carboline alkaloids and amide alkaloids. Structures of the alkaloids in TT are shown in Fig. 4.

**Others**

Other components of TT include organic acids, amino acids and other substances. Organic acids isolated from TT are benzoic acid [46], vanillic acid, 2-methyl benzoic acid, ferulic acid [42], succinic acid, palmitic acid monoglyceride, succinic acid, docosanoic acid [47], Tribulus acid [48] and others. The main amino acids are alanine and threonine [49]. In addition, TT also contains 4-ketopinoresinol [50], uracil nucleic acid [46], coumarin [47], emodin, and physcion [51].

**Pharmacological activities**

TT has long been used in traditional Chinese and Indian systems of medicine for the treatment of various ailments, especially for improving sexual function, the prevention and treatment of cardiovascular diseases, and diabetes. It also has hepatoprotective, antioxidant, anti-inflammatory, antibacterial, antiaging, and antitumour activities.

**Improving sexual function**

The active extracts and constituents of TT could improve sexual function through activating aphrodisiacs and improving fertility in men. It could also activate sexual desire in postmenopausal women. It is widely believed and insistently advertised that TT possesses aphrodisiac and pro-sexual activities due to its ability to increase testosterone or testosterone precursor levels and this view is outdated [2].

**Aphrodisiac activation**

Erectile dysfunction (ED) is a sexual disorder characterized by the inability to achieve or maintain a sufficiently rigid erection [52]. Analysis of phytochemical and pharmacological studies in humans and animals revealed an important role for *T. terrestris* in treating erectile dysfunction and sexual desire problems. Rats were fed a standard diet treated with Mucuna pruriens, *T. terrestris*, and Ashwagandha (300 mg kg⁻¹) for 8 weeks. The results indicated that the extract of TT was comparatively more potent than the two others. These herbs are potent
enhancers of sexual function and behaviour by increasing the testosterone levels and regulating the NF-κB and Nrf2/HO–1 pathways in male rats [53].

The hormonal effects of TT were evaluated in primates, rabbits and rats to identify its usefulness in the management of ED [54]. Blood samples were analysed for testosterone (T), dihydrotestosterone (DHT) and dehydroepiandrosterone sulphate (DHEAS) levels using a radioimmunoassay. TT increased some of the sex hormones, which is possibly due to the presence of protodioscin in the extract. The results indicated that TT may be useful in mild to moderate cases of ED.

The aphrodisiac properties of the furostenol glycoside fraction of \textit{T. terrestris} extract (TT-FG) were previously studied [55]. Adult Wister rats were castrated and divided into five groups of six animals each and treated with TT-FG (5, 10, and 25 mg kg$^{-1}$, p.o.) once daily through subcutaneous injections for 14 days. After the acute (1 day) and subacute (7 and 14 days) treatments with the TT-FG, there was an increase in mounting frequency (MF), intromission frequency (IF), and ejaculation latency (EL) and a decrease in mounting latency (ML), intromission latency (IL), and post-ejaculation interval (PEI) and serum testosterone levels in the blood.

There was a randomized, double-blind, placebo-controlled, clinical trial as a piece of evidence for aphrodisiac activation function of TT. 180 males aged between 18 and 65 years with mild or moderate ED and with or without Hypoactive sexual desire disorder (HSDD) were randomized in a 1:1 ratio to the two treatments groups (TT or placebo). The TT group received 2 tablets (500 mg) Tribestan orally three times daily after meals for 12 weeks. Each tablet contains the active substance TT herba extractum siccum 250 mg (content of furostanol saponins not less than 112.5 mg). And the placebo group were treated by a identical appearance, colour and taste one. The results showed that there was significant differences of IIEF (International Index of Erectile Function) score between the two groups (p < 0.0001) after 3 months, but no differences in the incidence of adverse effects [56]. It can therefore be assumed that TT can improve sexual function.

**Improvement in fertility**

In the literature, it has been concluded that the ethanol extract of \textit{T. terrestris} (EETT) influences spermatogenesis, as shown by the evident changes in the tubular compartment of the testes, such as increases in the total tube length, tubular volume and height of the seminiferous epithelium. The hexanic and aqueous soluble fraction in the methanol fractions promoted changes in the intertubular compartment because they increased the nuclear volume, cytoplasmic volume and individual volume of Leydig cells in male Wistar rats [57].

Another animal study describes the protective role of TT against AlCl$_3$-induced adverse effects on male reproductive organs and fertility. High dosages of TT (100 mg kg$^{-1}$ day$^{-1}$) in AlCl$_3$-treated mice restored the
body weight, sex organ relative weights, sperm count, motility, viability, epididymal sialic acid, seminal vesicular fructose, serum testosterone, antioxidant enzymes [superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase-1 (GPx)], mating ability and fertility [58].

TT was reported to cause reproductive system enhancement and possess antioxidant activity, which may assist in the choice of drugs for longer durations that can be prescribed safely without affecting the fertility potential in males. A high dose of the fruit extract of TT (200 mg kg$^{-1}$ day$^{-1}$) restored metronidazole
| No. | Chemicals                                  | Aglycones | $R_1$ | $R_2$ | $R_3$ | $R_4$ | $R_5$ | $R_6$ | 25-C | Refs. |
|-----|-------------------------------------------|-----------|-------|-------|-------|-------|-------|-------|------|-------|
| 1   | Tigogenin                                 |           | OH    | H     | H     | H     | H     | R     |      | [7]   |
| 2   | Neotigogenin                              |           | OH    | H     | H     | H     | H     | R     |      | [8]   |
| 3   | Tigogenin-3-O-$\beta$-glc(1 → 4)-$\beta$-o-gal (A) |           | Ra    | H     | H     | H     | H     | R     |      | [9]   |
| 4   | Terrestrosin F                            |           | Rb    | H     | H     | H     | H     | R     |      | [10]  |
| 5   | Desgalactotigogenin                       |           | Rc    | H     | H     | H     | H     | R     |      | [9, 11] |
| 6   | Gitoiin (B)                               |           | Rd    | H     | H     | H     | H     | R     |      | [11]  |
| 7   | Tigogenin-3-O-$\beta$-o-xyxyl(1 → 2)-[$\beta$-o-xyxyl(1 → 3)]-[$\beta$-o-glc(1 → 4)-[$\alpha$-rha(1 → 2)]-[$\beta$-o-gal (C) |           | Re    | H     | H     | H     | H     | R     |      | [9, 11, 12] |
| 8   | Tribulosin                                |           | Re    | H     | H     | H     | H     | S     |      | [13]  |
| 9   | Terrestrosin A                            |           | Rf    | H     | H     | H     | H     | R,S   |      | [11]  |
| 10  | Terrestrosin B                            |           | Rg    | H     | H     | H     | H     | R,S   |      | [11]  |
| 11  | Gitoiin                                   |           | OH    | OH    | H     | H     | H     | H     | R     | [7, 14] |
| 12  | Neogitoiin                                |           | OH    | OH    | H     | H     | H     | H     | S     | [15]  |
| 13  | Gitoiin-3-O-$\beta$-o-gluc(1 → 4)-$\beta$-o-gal |           | Ra    | OH    | H     | H     | H     | R     |      | [16]  |
| 14  | F-gitoiin                                 |           | Rd    | OH    | H     | H     | H     | R     |      | [11]  |
| 15  | Desglccolanatigogen                       |           | Rd    | OH    | H     | H     | H     | H     |      | [11]  |
| 16  | 2SR,5a-spiro-2a,3$\beta$-dihydroxy-3-$\beta$-o-0-gl(1 → 4)-$\beta$-o-gal |           | Ra    | OH    | H     | H     | H     | R,S   |      | [16]  |
| 17  | Terrestrosin E                            |           | Rf    | OH    | H     | H     | H     | R,S   |      | [11, 16] |
| 18  | Hecogenin                                 |           | OH    | $\approx$ | O | H     | H     | H     | R     | [7, 17] |
| 19  | Neohecogenin sapogenin                    |           | OH    | $\approx$ | O | H     | H     | H     | S     | [17]  |
| 20  | Agovoside A                               |           | Rh    | $\approx$ | O | H     | H     | H     | R     | [12]  |
| 21  | Hecogenin-3-O-$\beta$-o-gluc(1 → 4)-$\beta$-o-gal (D) |           | Ra    | H     | $\approx$ | O | H     | H     | R     | [9, 18] |
| 22  | Hecogenin-3-O-$\beta$-o-gluc(1 → 2)-$\beta$-o-gluc(1 → 4)-$\beta$-o-gal (E) |           | Ri    | H     | $\approx$ | O | H     | H     | R     | [9]   |
| 23  | Hecogenin-3-O-$\beta$-o-xyxyl(1 → 2)-$\beta$-o-gluc(1 → 4)-$\beta$-o-gal (F) |           | Rj    | H     | $\approx$ | O | H     | H     | R     | [9]   |
| 24  | Hecogenin-3-O-$\beta$-o-gluc(1 → 2)-[$\beta$-o-xyxyl(1 → 3)]-[$\beta$-o-gluc(1 → 4)-[$\alpha$-rha(1 → 2)]-[$\beta$-o-gal (G) |           | Rd    | H     | $\approx$ | O | H     | H     | R     | [9, 12] |
| 25  | Terrestrosin D                            |           | Rk    | H     | $\approx$ | O | H     | H     | H     | R     | [11, 12] |
| 26  | Hecogenin-3-O-$\beta$-o-xyxyl(1 → 2)-[$\beta$-o-xyxyl(1 → 3)]-[$\beta$-o-gluc(1 → 4)-[$\alpha$-rha(1 → 2)]-[$\beta$-o-gal (H) |           | Re    | H     | $\approx$ | O | H     | H     | R     | [12]  |
| 27  | Neohecogenin-3-O-$\beta$-o-gluc |           | Ri    | H     | $\approx$ | O | H     | H     | S     | [8]   |
| 28  | Terreside B                               |           | Ra    | H     | $\approx$ | O | H     | H     | H     | S     | [19]  |
| 29  | Terreside A                               |           | Ri    | H     | $\approx$ | O | H     | H     | H     | R     | [19]  |
| 30  | Tettetressosin C                          |           | Rf    | H     | $\approx$ | O | H     | H     | R,S   | [11, 13] |
| 31  | 2SR,5a-spiro-12-one-3-O-$\beta$-o-xyxyl(1 → 2)-[$\beta$-o-xyxyl(1 → 3)]-[$\beta$-o-gluc(1 → 4)-[$\alpha$-rha(1 → 2)]-[$\beta$-o-gal |           | Re    | H     | $\approx$ | O | H     | H     | R,S   | [20]  |
| 32  | Hecogenone                                |           | $\approx$ | O | H     | $\approx$ | O | H     | H     | R     | [7]   |
| 33  | 2SR-5a-spiro-3,6,12-triketo               |           | $\approx$ | O | H     | $\approx$ | O | H     | H     | R     | [7]   |
| 34  | Sarasasaponin                            |           | OH    | H     | H     | H     | H     | S     |      | [21]  |
| 35  | Isotetrestrosin B8                        |           | Rg    | H     | H     | H     | H     | S     |      | [21]  |
| No. | Chemicals | Aglycones | R<sub>1</sub> | R<sub>2</sub> | R<sub>3</sub> | R<sub>4</sub> | R<sub>5</sub> | R<sub>6</sub> | 2S-C | Refs. |
|-----|-----------|-----------|--------------|--------------|--------------|--------------|--------------|--------------|-------|-------|
| 36  | Chlorogenin | I         | OH           | H            | H            | OH           | H            | H            | R     | [22]  |
| 37  | Terrestrinin U | I       | H            | Rm           | H            | H            | Rl           | H            | S     | [23]  |
| 38  | Terrestrinin I | I       | Ra           | H            | =O           | H            | RI           | H            | S     | [24]  |
| 39  | 23S,25S-spiro-24-one-3β,23-diol-3-O-[α-L-rha-(1→2)]-O-[β-D-glc-(1→4)]-β-D-gal | I       | Rn           | H            | H            | H            | =O           | OH           | S     | [25]  |
| 40  | 24S,25S-spiro-3β,24-diol-3-O-[α-L-rha-(1→2)]-O-[β-D-glc-(1→4)]-β-D-gal | I       | Rn           | H            | H            | H            | OH           | H            | S     | [25]  |
| 41  | 23S,24R,25R-5α-spiro-3β,23,24-triol-3-O-[α-L-rha-(1→2)]-O-[β-D-glc-(1→4)]-β-D-gal | I       | Rn           | H            | H            | H            | OH           | OH           | R     | [26]  |
| 42  | 23S,24R,25S-5α-spiro-3β,23,24-triol-3-O-[α-L-rha-(1→2)]-O-[β-D-glc-(1→4)]-β-D-gal | I       | Rn           | H            | H            | H            | OH           | OH           | S     | [26]  |
| 43  | Diosgenin | II        | OH           | H            | R            |             |               |               | R     | [14]  |
| 44  | Yamogenin | II        | OH           | H            | S            |             |               |               |       | [10]  |
| 45  | Trillin | II        | RI           | H            | R            |             |               |               | R     | [27]  |
| 46  | Trillarin | II        | Ro           | H            | R            |             |               |               |       | [27]  |
| 47  | Dioscin | II        | Rp           | H            | R            |             |               |               | R     | [28]  |
| 48  | Gracillin | II        | Rq           | H            | R            |             |               |               |       | [27]  |
| 49  | Diosgenin-3-O-α-L-rha-(1→3)]β-D-glc | II       | Rh           | H            | R            |             |               |               | R     | [27]  |
| 50  | Tribestin | II        | Rs           | H            | R            |             |               |               | R     | [28]  |
| 51  | Ruscogenin | II       | OH           | OH           | R            |             |               |               |       | [14]  |
| 52  | Saponin C | II        | OH           | Rt           | R            |             |               |               | R     | [29]  |
| 53  | 25R-spiro-3,5-diene | III     | H            |             | R            |             |               |               |       | [14]  |
| 54  | 25R-spiro-3,5-dien-12one | III     | =O           |             | R            |             |               |               |       | [17]  |
| 55  | 25R-spiro-4-ene-3,12-dione | IV      | =O           | H            | H            | R            |             |               |       | [7, 17] |
| 56  | 25R-spiro-4-ene-3,6,12-trione | IV      | =O           | H            | =O           | H            | R            |             |       | [7]   |
| 57  | 25R-spiro-24-hydroxy-3,12-dione | IV      | =O           | H            | H            | OH           | R            |             |       | [17]  |
| 58  | 25R-spiro-2αβδ-hydroxy-4-ene-12one | IV      | OH           | OH           | H            | H            | R            |             |       | [17]  |
| 59  | 25R-sa-furo-22-methoxy-3β,26-dihydroxy-Oββ-D-glc-3-Oβα-D-xyl-(1→2)]-Oββ-D-glc-(1→4)-[α-L-rha-(1→2)]β-D-gal (J) | V       | Re           | H            | H            | OH            | Rl           | R,S        |       | [9, 12, 21] |
| 60  | Neoprotodioscin | V       | Rp           | H            | H            | OH           | Rl           | R          |       | [29]  |
| 61  | Neoprototribestin | V       | Rs           | H            | H            | OH           | Rl           | R          |       | [24]  |
| 62  | Terrestrinin B | V       | Re           | H            | H            | OH           | Rl           | R          |       | [12]  |
| 63  | Tetterestosin H | V       | Re           | H            | H            | OH           | Rl           | R,S        |       | [16]  |
| 64  | Terrestroneoside A | V       | Re           | H            | H            | OCH<sub>3</sub> | Rl           | Uncertain  |       | [30]  |
| 65  | Terrestrosin F | V       | Ra           | OH           | H            | OH           | Rl           | R          |       | [16]  |
| 66  | Terrestrosin G | V       | Ra           | OH           | H            | OH           | Rl           | R,S        |       | [16]  |
| 67  | Terrestrosin H | V       | Ra           | OH           | H            | OH           | Rl           | R          |       | [16]  |
| 68  | 25S,5α-furo-12-one-3β,22α,26-triroyoxy-Oββ-D-glc-(1→4)]β-D-gal | V       | Ru           | H            | =O           | OH           | Rl           | R          |       | [31]  |
| 69  | 25S,5α-furo-12-one-3β,22α,26-triroyoxy-Oββ-D-glc-(1→4)]β-D-gal | V       | Ru           | H            | =O           | OH           | Rl           | S          |       | [20]  |
| 70  | 25S,5α-furo-12-one-3β,22α,26-triroyoxy-Oββ-D-glc-(1→4)]β-D-gal | V       | Rg           | H            | =O           | OH           | Rl           | S          |       | [20]  |
| No. | Chemicals                        | Aglycones | R₁ | R₂ | R₃ | R₄ | R₅ | R₆ | 2S-C | Refs. |
|-----|---------------------------------|-----------|----|----|----|----|----|----|------|-------|
| 71  | Terrestrosin I                  |           | V  |    |    |    |    |    |      | [16]  |
| 72  | 5α-furo-12-one-3β,22,26-trihydroxy-O-β-D-gluc-3-β-D-gluc-1→3)-[β-D-gal-(1→2)]-β-D-gluc(1→4)-β-D-gluc | V          | Rv | H  | =O | OH | RI |    |      | Uncertain |
| 73  | Terrestrinin R                   |           | V  |    |    |    |    |    | =O  | RI    | [23]  |
| 74  | Terrestrinin S                   |           | V  |    |    |    |    |    | =O  | RI    | [23]  |
| 75  | Terrestrinin F                   |           | V  |    |    |    |    |    | =O  | RI    | [24]  |
| 76  | 26-O-β-D-gluc-25R:5α-furo-2α,3β,22α,26-tetraol-3-O-β-D-gluc(1→2)-O-β-D-gluc(1→4)-β-D-gluc | V          | Ri | OH | H  | CH₃| RI |    |      | [25]  |
| 77  | Pseudoprotodioscin(K)            |           | VI |    |    |    |    |    | Rp  | OH    | R     | [21]  |
| 78  | Methyl pseudo-diosgenin          |           | VI |    |    |    |    |    | Rp  | OCH₃  | R     | [32]  |
| 79  | Prototribestin                   |           | VI |    |    |    |    |    | Rs  | OH    | R     | [32]  |
| 80  | Methyprototribestin              |           | VI |    |    |    |    |    | Rs  | OCH₃  | R     | [32]  |
| 81  | Protogracillin                   |           | VI |    |    |    |    |    | Rq  | OH    | H     | [8]   |
| 82  | Terrestrosin J                   |           | VI |    |    |    |    |    | Rf  | OH    | RI    | [16]  |
| 83  | Tribol                           |           | VI |    |    |    |    |    | Rp  | H     | OH    | [28]  |
| 84  | Terrestrinin K                   |           | VI |    |    |    |    |    | Rm  | OH    | H     | [23]  |
| 85  | Terrestrosin K                   |           | VII|    |    |    |    |    | Rf  | H     | =O H  | CH₃   | [16]  |
| 86  | 25R:5α-furo-12-one-20R:26-di-O-β-D-gluc-3-O-β-D-gluc(1→4)-β-D-gluc | VII        | Ra | H  | =O | H  | CH₃| RI | RS  | [31]  |
| 87  | 25R:5α-furo-12-one-20R:26-di-O-β-D-gluc-3-β-D-gluc(1→2)-O-β-D-gluc(1→4)-β-D-gluc | VII        | Rf | H  | =O | H  | CH₃| RI | RS  | [16]  |
| 88  | 5α-furo-12-one-2022-ene-3β,26-dihydroxy-O-β-D-gluc-3-O-β-D-gluc(1→3)-[β-D-gal-(1→2)]-β-D-gluc(1→4)-β-D-gluc | VII        | Rv | H  | =O | H  | CH₃| RI | Uncertain | [18] |
| 89  | Tribulosaponin A(L)              |           | VII|    |    |    |    |    | Rp  | H     | H₂    | CH₃   | S     | [21]  |
| 90  | Tribulosaponin B(M)              |           | VII|    |    |    |    |    | Rg  | H     | H₂    | CH₃   | R     | [21]  |
| 91  | 26-O-β-D-gluc-25R:5α-furo-20(22)-ene-2α,3β,26-triol-3-O-β-D-gluc(1→2)-O-β-D-gluc(1→4)-β-D-gluc | VII        | Rn | H  | =O | OH | OCH₃| RI | R   | [25]  |
| 92  | 26-O-β-D-gluc-25R:5α-furo-12-one-3β,22α,26-triol-3-O-β-D-gluc(1→2)-O-β-D-gluc(1→4)-β-D-gluc | VII        | Ri | H  | =O | OH | OR  | RI | R   | [33]  |
| 93  | 26-O-β-D-gluc-25S:5α-furo-22-methoxy-2α,3β,26-triol-3-O-β-D-gluc(1→2)-O-β-D-gluc(1→4)-β-D-gluc | VII        | Ri | OH | H  | OH | OCH₃| RI | S   | [33]  |
| 94  | Terrestrinin A                   |           | VIII|    |    |    |    |    | R   | S     | [12]  |
| 95  | Terrestrinin Q                   |           | VIII|    |    |    |    |    | R   | R     | [23]  |
| 96  | Terrestrinin C                   |           | IX  |    |    |    |    |    | OH  | OH    | =O H  | H₂    | RI    | R  | [24]  |
| 97  | Terrestrinin D                   |           | IX  |    |    |    |    |    | =O  | H     | =O H  | H₂    | RI    | R  | [4]   |
| 98  | Terrestrinin E                   |           | IX  |    |    |    |    |    | =O  | H     | =O  | =O H  | RI    | R  | [24]  |
| 99  | Terrestrinin G                   |           | IX  |    |    |    |    |    | =O  | H     | OH   | H₂    | RI    | R  | [24]  |
| 100 | Terrestrinin H                   |           | IX  |    |    |    |    |    | =O  | H     | =O H  | H₂    | Rv    | R  | [24]  |
| 101 | Terrestrinin J                   |           | X   |    |    |    |    |    | Rm  | OH    | RI    | R     | [23]  |
| 102 | Terrestrinin L                   |           | XI  |    |    |    |    |    | Rm  | OH    | RI    | R     | [23]  |
| 103 | Terrestrinin M                   |           | XI  |    |    |    |    |    | Rx  | OH    | RI    | R     | [23]  |
| 104 | Terrestrinin N                   |           | XI  |    |    |    |    |    | Ry  | OH    | RI    | R     | [23]  |
| 105 | Terrestrinin O                   |           | XII |    |    |    |    |    | Ry  | H     | RI    | R     | [23]  |
| 106 | 26-O-β-D-gluc-25R:5α-furo-20(22)-ene-2α,3β,26-triol-3-O-β-D-gluc(1→2)-O-β-D-gluc(1→4)-β-D-gluc | XII       | Ri | OH | RI | R | R | [25]  |
| No. | Chemicals                  | Aglycones | R<sub>1</sub> | R<sub>2</sub> | R<sub>3</sub> | R<sub>4</sub> | R<sub>5</sub> | 25-C | Refs. |
|-----|----------------------------|-----------|--------------|--------------|--------------|--------------|--------------|------|-------|
| 107 | Terrestrinin P             | XIII      | Rm           | Rl           | R             |               |               | R    | [23]  |
| 108 | Terrestrinin T             | XIV       | Rm           | Rl           | R             |               |               | R    | [23]  |

Ra: Oβ-D−Gal-[1→4]-β-D−Glc; Rb: O−Glc/Rha = 2:1; Rc: Oβ-D−Gal-[1→2]-β-D−Xyl; Rf: Oβ-D−Glc-[1→4]-β-D−Glc; Rg: Oβ-D−Gal-[1→2]-α-L−Rha; Rh: Oβ-D−Glc-[1→4]-β-D−Glc; Rj: Oβ-D−Gal-[1→4]-β-D−Glc/[1→3]-β-D−Xyl; Rk: Oβ-D−Gal-[1→4]-β-D−Glc/[1→3]-β-D−Xyl; Rl: Oβ-D−Glc/[1→4]-β-D−Glc; Rm: Oβ-D−Gal-[1→2]-α-L−Rha; Rn: Oβ-D−Gal-[1→4]-β-D−Glc; Ro: Oβ-D−Glc/[1→2]-β-D−Glc; Rp: Oβ-D−Glc/[1→2]-α-L−Rha; Rs: Oβ-D−Glc/[1→4]-β-D−Glc; Rt: Oβ-D−Glc/[1→2]-α-L−Rha; Ru: Oβ-D−Glc/[1→3]-α-L−Rha; Rv: Oβ-D−Gal-[1→4]-β-D−Glc/[1→3]-α-L−Xyl; Rw: Oβ-D−Glc/[1→2]-β-D−Glc; Rx: Oβ-D−Gal-[1→4]-β-D−Glc; Ry: Oβ-D−Gal-[1→4]-β-D−Glc; Rz: Oβ-D−Glc/[1→4]-β-D−Glc; R[1→3]-β-D−Xyl; R[1→2]-β-D−Xyl;
(MTZ)-induced spermatogenic inhibition and reduced the epididymal sperm count. The restoring potential of TT against MTZ-induced alterations in the spermatogenesis appears to be due to the presence of antioxidative flavonoids rather than steroidal saponins [59].

The in vitro addition of TT extract to human sperm could affect male fertility capacity. The incubation of human semen with 40 and 50 μg mL$^{-1}$ of TT extract significantly enhanced the total sperm motility, number of progressive motile spermatozoa, and curvilinear velocity over 60–120 min of holding time. Overall, the sperm viability significantly improved [60].

**Libido-enhancing activity**

HSDD is defined in Diagnostic and Statistical Manual of Fourth Edition as persistent or recurrent deficiency (or absence) of sexual fantasies/thoughts, and/or desire for or receptivity to sexual activity, which causes personal distress [61]. TT was considered to be a safe alternative for the treatment of HSDD in postmenopausal women because it was effective in reducing symptoms with few side effects through a randomized, double-blinded, placebo-controlled trial (A total of 45 healthy sexually active postmenopausal women who reported a diminished libido were selected to participate in the study and were randomly assigned to receive 750 mg day$^{-1}$ of TT or a placebo for 120 days). Its probable mechanism of action involves an increase in the serum levels of free and bioavailable testosterone [62].

Other clinical research established that regarding the treatment in the domains of desire and sexual interest of 74 postmenopausal women with sexual dysfunction, the TT treatment (250 mg, orally three times a day for 90 days) was considered to be effective in treating sexual problems among menopausal women [63].

**Antiurolithic activity**

The fruits of TT have long been used in traditional systems of medicine for the treatment of various urinary diseases including urolithiasis. Calcium oxalate is a major type of crystal found in kidney stones. Calcium oxalate is classified into two types: calcium oxalate monohydrate stones (COM) and calcium oxalate dihydrate (COD). Many medicinal plants have been used for centuries for the treatment of urinary stones in spite of the lack of rationale behind their use. The aqueous extract of TT fruits and its fractions were studied to evaluate its antiurolithic potential using different models. The inhibitory potency of the plant was tested on the nucleation and growth of the most commonly occurring kidney stones and COM. The results showed that the bioactive $n$-butanol fraction, due to higher contents of quercetin,
diosgenin and tannic acid, has a protective capacity rather than a curative property against urolithiasis [64].

A protein (60 kDa) purified from TT showed the highest similarity with carotenoid cleavage dioxygenase 7 (CCD7) of Arabidopsis thaliana after matching peptide mass fingerprints with the MASCOT search engine. CCD7 belong to a family of dioxygenases, which possess five characteristic conserved histidines spread throughout their primary protein sequence. Histidine is said to induce the conversion of oxalate to formic acid and carbon dioxide (CO₂). The purified protein decreased cell injury induced by oxalate in a concentration dependent manner and showed the ability to inhibit calcium oxalate (CaOx) crystallization in vitro [65].

Human clinical data indicated that TT extract may be useful in the treatment of urolithiasis. After oral administration of the extract, the levels of mean citrate, oxalate, proteins and glycosaminoglycan in patients’ 24 h urine samples decreased significantly. Urine volume and phosphate level in the serum were not altered significantly.
in the urolithic patients [66]. It was concluded that TT extract was useful in the treatment of urolithiasis.

**Antidiabetic activity**

Diabetes mellitus is a metabolic disorder with chronic hyperglycaemia, which results from a defect in insulin secretion, insulin action, or both [67]. The gross saponins of *T. terrestris* (GSTT) showed inhibitory activity against α-glucosidase. In addition, it showed the inhibition activities of a postprandial increase in blood glucose and improvement in insulin dependent diabetes symptoms [68]. Animal experiments indicated that GSTT significantly reduced the postprandial blood glucose levels by intragastric administration of sucrose in normal rats and type 2 diabetic rats but did not affect the postprandial blood glucose levels of the rats with intragastric administration of glucose [69]. Clinical trials proved that the water extract of *T. terrestris* (WETT) has an antidiabetic activity. The fasting blood glucose, 2-h postprandial glucose, glycated haemoglobin and lipid profile of diabetic women treated with TT extract (1000 mg day⁻¹) for three months were lowered compared to those of the placebo group [70].

**Prevention and treatment of cardiovascular diseases**

Presently, the clinical treatments are thrombolysis and nerve protection. Thrombolysis has a significant effect. However, it is limited by a narrow therapeutic time window. Therefore, the development of neuroprotective agents is of great significance. Studies have shown that GSTT has a neuroprotective effect on cerebral ischaemia injury, and these saponins have been commercially available as active compounds in traditional Chinese medicine formulations, such as “Xin-nao-shutong”, which has been used for the treatment of cardiovascular disease [71]. Meanwhile, TT plays an important role in the treatment of cardiovascular disease with anti-myocardial ischaemia and myocardial ischaemia–reperfusion injury. GSTT has a protective effect on myocardial ischaemia–reperfusion injury. GSTT reduced the levels of lactate dehydrogenase (LDH), methane dicarboxylic aldehyde (MDA), tumor necrosis factor (TNF)-α and interleukin (IL)-6, increased SOD and the rate of apoptosis, and improved the structure of cardiomyocytes in rats [72]. Moreover, GSTT could improve coronary flow and heart function and increase adenosine triphosphate (ATP) activity in myocardial ischaemia–reperfusion injury [73]. The methanol extract of *T. terrestris* (METT) fruits, which mainly contains ferulic acid, phloridzin and diosgenin, had an effect on mitochondrial dysfunction in a cell-based (H9c2) myocardial ischaemia model. The extract guards the mitochondria via its antioxidant potential [74]. The cellular and molecular mechanisms of the prevention activity against artherosclerosis occurs when GSTT significantly suppresses the increase in cell proliferation induced by angiotensin II, significantly suppresses the increase in the intracellular production of hydrogen peroxide (H₂O₂) induced by angiotensin II, significantly inhibits the increase in intracellular free Ca²⁺ induced by H₂O₂, significantly inhibits the increase in phospho-ERK1/2 induced by angiotensin II, and significantly inhibits the increase in the mRNA expressions of c-fos, c-jun and pck-a induced by angiotensin II [75].

TT significantly suppressed the proliferation of ox-LDL-induced human umbilical vein endothelial cells (HUVECs) and the apoptosis rate. It also prolonged the HUVEC survival time and postponed the cells’ decaying stage (from the 69 h to over 100 h). TT normalized the increased mRNA expressions of PI3Kα and Socs3. It also decreased the mRNA expressions of Akt1, AMPKα1, JAK2, LepR and STAT3 induced by ox-LDL. The most notable changes were for JAK2, LepR, PI3Kα, Socs3 and STAT3. It is thought that the JAK2/STAT3 and/or PI3K/AKT pathway might be a very important pathway that is involved in the mechanism of TT as a vascular protective agent [76].

In addition, TT functions as a protector of the myocardium. It supported cardioprotective properties against myocardial ischaemia, protected myocardial cells and reduced the apoptosis rate induced by oxidative stress damage [77]. Inhibition of cardiac muscle cell apoptosis occurs when GSTT reduces cell apoptosis through regulating protein expression of Bcl-2 and Bax [78].

**Protective activity in neuronal cells**

TT has a protective effect for neuron injury mainly via its anti-inflammatory and antioxidant effects. GSTT has a neuroprotective effect on cerebral ischaemia–reperfusion injury in rats by suppressing NF-κB, TNF-α and IL-1β. It plays a neuroprotective role in rat cerebral ischaemia reperfusion injury by inhibiting the inflammatory response and PPARγ protein expression [79]. GSTT decreased the damage to PC12 cells induced by H₂O₂ in the increased mRNA expressions of PI3Kα and Socs3. It also decreased the mRNA expressions of Akt1, AMPKα1, JAK2, LepR and STAT3 induced by ox-LDL. The most notable changes were for JAK2, LepR, PI3Kα, Socs3 and STAT3. It is thought that the JAK2/STAT3 and/or PI3K/AKT pathway might be a very important pathway that is involved in the mechanism of TT as a vascular protective agent [76].

After cerebral haemorrhaging, brain tissue generates many free radicals that causes lipid peroxidation. GSTT significantly increased the SOD content and decreased the MDA and NO levels in plasma and brain tissue to attenuate neuron injury [81].

The apoptosis of retinal ganglion cells (RGCs) is an important cause of glaucoma. TT can block the optic nerve injury pathway and enhance the survival of the optic nerve to protect the optic nerve [82, 83]. It was reported that TT could reduce the degeneration of RGCs
and the retinal nerve fibre layer in hyper-intraocular pressure rabbits by intravenous administration with TT sterilization powder [84].

**Improvement of athletic ability activity**

Athletic fatigue is generally measured by the levels of testosterone and corticosterone, and the testosterone and corticosterone (T/C) ratio. Herbs and herbal combinations have been used to improve athletic ability through several ways that mimic epinephrine effects, mimic testosterone effects, and increase the productions of corticotropin and cortisol. TT contains gitonin, protodioscin, and tribulosaponins A and B, which are believed to mimic testosterone-like effects in humans because of the similarities of their chemical structures [85, 86]. The main effect is an increase of testosterone anabolic and androgenic action via the activation of endogenous testosterone production [87].

The administration of GSTT (120 mg kg\(^{-1}\)) can prolong the time to exhaustion and increase body mass, relative mass, and protein levels of gastrocnemius in overtrained rats. The level of testosterone can directly affect the motor ability of the body and its restoration. Corticosterone can accelerate the decomposition of proteins in the body [88]. Treatment of rats with GSTT during overtraining dramatically increased the serum level of testosterone and led to a significant decrease in the serum level of corticosterone. The T/C ratio with GSTT was much higher than that with the blank control.

In addition, the cognate receptor of testosterone is AR. IGF-1 is closely related to muscle mass, conservation of the musculoskeletal system, the metabolic rate, and muscle strength. GSTT resulted in a significant increase in AR in gastrocnemius and significantly suppressed the overtraining-induced increase in IGF-1R in the liver. It was concluded that GSTT significantly improves exercise performance due to changes in the androgen–AR axis and IGF-1R signalling [89].

**Antitumour activity**

GSTT is likely to affect the processes of apoptosis and metastasis of cancer cells. The overexpression of CXCR4 has been associated with the formation of metastases and poor prognosis of patients with breast and other types of cancer. CCR7 is reportedly correlated with lymphatic metastasis and poor prognosis in breast cancers. The product of the BCL2 gene is a mitochondrial membrane protein that blocks apoptosis. After implying a cell-specificity for GSTT, CXCR4 expression was reduced in both cell lines, and CCR7 and BCL2 levels decreased only in tumourigenic MCF-7 cells [90].

The anticancer mechanism of terrestrosin D was detected by observing in vitro Caspase-3 activity and vascular endothelial growth factor secretion and the in vivo anticancer effect of the PC-3 xenograft mouse model. It was concluded that terrestrosin D inhibited tumour growth through the inhibition of tumour angiogenesis. In addition, GSTT has a preventive efficacy against UVB-induced carcinogenesis. The photo protective effect of GSTT is tightly correlated with the enhancement of NER gene expression and the blocking of UVB-mediated NF-κB activation [91].

**Antibacterial activity**

The antibacterial activity of TT had been widely studied. A total of 50% of *H. pylori* strains were sensitive to a concentration of 1000 mg mL\(^{-1}\) of total extract of TT by the in vitro cup plate method [92]. GSTT inhibited the *Candida albicans* ACS1, ACS2, ERG1, ERG2, ERG6, ERG7, ERG11, ERG25, ERG26 and ERG27 genes, which are directly involved in the ergosterol synthesis pathway. An anti-fungi agent, GSTT may function through direct binding to sterol on the cell membrane and may inhibit ERG gene expression in *C. Albicans* [93]. TT was extracted with different solvents (methanol, petroleum ether, chloroform, and ethanol). The results showed that methanol extract has the highest inhibition zone for *Bacillus cereus*, *Escherichia coli* and *Staphylococcus aureus*. For *Staphylococcus aureus* and *Pseudomonas aeruginosa*, WETT also had a certain inhibitory effect [94]. The ethanol extract of TT exhibited good antibacterial activity against *Streptococcus mutans*, *Streptococcus sanguis*, *Actinomyces viscosus*, *Enterococcus faecalis*, *Staphylococcus aureus*, and *Escherichia coli*. Complexes of *T. terrestris*, *Capsella bursa-pastoris*, and *Glycyrrhiza glabra* had synergistic effects compared with those of any of the herbs alone [95].

**Antioxidant activity**

TT exhibited effective antioxidant activity in a concentration-dependent manner by 2,2-dii-(4-tert-octylphenol)-1-picyrylhydrazyl (DPPH), \(H_2O_2\), and superoxide scavenging activity, as well as the FRAP (Ferric reducing antioxidant power) assay [96]. The experiment proved that the antioxidant effect of GSTT is excellent and that it could improve SOD activity and MDA content for chronically high intraocular pressure in rabbits [97]. Compared with the ethanolic extraction, the butanol extract (1 mg ml\(^{-1}\)) was rich in saponin and had notable quenching of nitric oxide (90.30%), hydroxyl radicals (90.02%), and hydrogen peroxide radicals (89%) [98]. Diosgenin from the callus of *T. terrestris* was found to have great antioxidant activity [99].

**Anti-inflammmatory activity**

The EETT and *N*-trans-\(\rho\)-caffeoyl tyramine isolated from TT had marked anti-inflammatory activities [100]. EETT and *N*-trans-\(\rho\)-caffeoyl tyramine inhibited the productions of nitric oxide (NO), TNF-\(\alpha\), IL-6 and IL-10 in

lipopolysaccharide (LPS) stimulated RAW264.7 cells in a dose dependent manner. In addition, N-trans-\(\rho\)-caffeoyl tyramine markedly suppressed the expression of cyclooxygenase (COX)-2 and the production of prostaglandin E2 (PGE2) through decreasing p-\(\eta\)-JNK expression.

METT (200 and 400 mg kg\(^{-1}\)) showed a dose-dependent inhibition of rat paw volume in a carrageenan-induced rat paw edema model. The TT extract and diclofenac sodium (a COX-inhibitor) were injected 30 min prior to carrageenan. The results showed that both drugs can reduce the paw volume 1–4 h after injection of carrageenan by inhibiting the releases of histamine, serotonin and kinins in the early phase. Furthermore, the anti-inflammatory effect of 400 mg kg\(^{-1}\) of TT extract is equivalent to that of 20 mg kg\(^{-1}\) of Diclofenac sodium [101].

**Hepatoprotective activity**

GSTT can ameliorate injured liver cells and have a protective effect on acute hepatic injury in mice induced by tripterygium glycosides. GSTT can significantly increase the levels of SOD and GPx, decrease the level of MDA in serum, suppress Caspase-3 expression and improve the ultrastructure of liver tissue in a mouse model. Caspase-3 is a class of hydrolytic protease, and its activation plays an important role in hepatocyte apoptosis. GSTT can interrupt the cascade in the process of apoptosis by reducing the expression of Caspase-3. The mechanism of its hepatoprotective activity may be related to the antioxidant activity, the influence on metabolism regulation and the repression of apoptosis of liver cells, which effectively reduces the level of Caspase-3 in liver tissue [102].

**Anthelmintic and larvicidal activity**

METT resulted in wormicidal activity by inhibiting spontaneous motility (paralysis) and causing death with lower doses. The effects were comparable with that of Albendazole [103]. In addition, TT exhibited high larvicidal activity. *Anopheles stephensi*, *Aedes aegypti* and *Culex quinque-fasciatus* have been identified as the primary vectors of malaria, dengue fever and lymphatic filariasis, respectively, in this part of the desert. The larvicidal potential of TT was evaluated by calculating the mortality percent of *A. stephensi*, *A. aegypti* and *C. quinque-fasciatus*. The results showed that the fruits were a more potent form regarding larvicidal activity than the leaves [104].

**Anticarious activity**

TT was certified to have an anticarious effect. *Streptococcus mutans* is an important oral pathogen that causes dental caries. The anticarious effect of TT was evaluated for inhibiting *S. mutans* bacteria. *In vitro* studies showed that the extract exhibited antibacterial activity for inhibiting *S. mutans* growth in a dose dependent manner. Meanwhile, TT extract suppressed the adherence of *S. mutans* to saliva-coated hydroxyapatite (S-HA), which simulated teeth, and inhibited the formation of water-insoluble glucans [105].

**Antiaging and memory improvement activity**

GSTT can effectively increase SOD activity, decrease MDA and hydroxyproline (Hyp) in the whole blood of \(\pi\)-galactose-induced senile mice. Compared with the ageing model group, the GSTT group showed a thicker dermis and more compactly arranged fibre content. The skin morphology of the GSTT group was close to that of the normal group [106].

Ageing is accompanied by a decline in memory, but GSTT can improve memory impairment. A study showed that GSTT significantly improved obtained memory disorder, consolidated memory disorder and recovered memory disorder [107]. The effect of the water extract of TT fruits on learning and memory ability in rodents was evaluated by recording the time of reaching the reward chamber (TRC) in the Hebb William Maze and the transfer latency (TL) in the T-zema. The results showed that the water extract of TT fruits significantly reduced the time of arrival at the maze in a dose-dependent manner [108].

**Absorption enhancer**

TT promotes absorption. The biopharmaceutics classification system (BCS) is a scientific classification method based on solubility in vitro and permeability of drugs in the intestine. Metformin hydrochloride (HCl) is a BCS class III drug with a high solubility and poor absorption characteristics. Therefore, it is necessary to increase the intestinal permeability of drugs to improve their bioavailability. The experiment indicated that TT can enhance the absorption of Metformin HCl in a goat intestine [109]. The absorption enhancement effect of TT was concluded by the presence of saponin.

**Toxicity**

An animal study investigated the acute toxicity of METT (2 g kg\(^{-1}\), given orally to 5 mice for 14 days). The methanol extracts mainly consisted of flavonoids, anthraquinones, phenols/tannins, and steroids/triterpenes. As a result, there were no toxic symptoms or mortality observed in any animals and no obvious differences between the treated and control animals regarding behavioural changes and toxicological signs (general behaviour, motor activities, aggressiveness, reaction to noise, reaction to pinch, state of the tail and state of excrement) [110].

The genotoxic potential of TT extracts, as assessed by a Comet assay in a rat kidney cell line and by an Ames
assay in *Salmonella typhimurium* strains, was evaluated [111]. The METT had relatively higher genotoxic activities (2400 mg mL\(^{-1}\) METT, tDNA%: 11.43) and cytotoxic activities (IC\(_{50}\) = 160 mg mL\(^{-1}\)) than WETT and the chloroform extracts of *T. terrestris* (CETT) but did not damage the deoxyribonucleic acid (DNA), whereas the 300 mg mL\(^{-1}\) WETT might induce frame shift mutations when metabolically activated. WETT showed oestrogenic activity at concentrations higher than 27 mg mL\(^{-1}\) (2.6-fold), and none of the extracts had androgenic activity.

**Conclusions**

The traditional pharmacological activities of TT focused on improving sexual function and cardiotoxic properties. Modern investigation showed that steroidal saponins and flavonoids with the prominent antiaging and anti-inflammatory activities were the main contributors to the traditional pharmacological activities. While the clinical trials with TT are scarce, and randomized placebo controlled clinical trials should be done in future. In addition, we should give more attention to the traditional curative effect of skin pruritus. TT maybe have more utilization as cosmetic plant materials on skin. A critical assessment of the results presented in this review may provide scientific evidence for the reasonable utilization of TT and promote further investigation for the development of new herbal medicine and health products.

**Abbreviations**

ATP: adenosine triphosphate; CaOx: calcium oxalate; CAT: catalase; CCD\(_2\): carotenoid cleavage dioxygenase 7; CETF: chloroform extracts of *T. terrestris*; CO\(_2\): carbon dioxide; COD: calcium oxalate dihydrate; CDM: calcium oxalate monohydrate stones; COX: cycloxygense; DHEAS: dehydroepiandrosterone; DHT: dihydrotestosterone; DNA: deoxyribonucleic acid; DPPH: 2,2-di-(4-tert-octylphenol)-1-picyrilhydrazyl; ED: erectile dysfunction; EFTT: ethanol extract of *T. terrestris*; EL: ejaculation latency; FRAP: ferric reducing antioxidant potential; GC/MS: gas chromatography/mass spectrometry; GPx-1: glutathione peroxidase 1; GSTT: gross saponins of extract of *T. terrestris*; HCo: hydrochloride; H\(_2\)O\(_2\): hydrogen peroxide; HSDD: hypoxia reactive sexual desire disorder; HUVECs: human umbilical vein endothelial cells; Hyp: hydroxyproline; IIEF: International Index of Erectile Function; IF: intromission frequency; IL-1: interleukin; LDH: lactate dehydrogenase; LTD: low density lipoprotein; LPS: lipopolysaccharide; MDA: methane dicarboxylic aldehyde; METT: methanolic extract of *T. terrestris*; MF: mounting frequency; ML: mounting latency; MTZ: metronidazole; NO: nitric oxide; PEI: post-ejaculation interval; PGE\(_2\): prostaglandin E2; RGCs: retinal ganglion cells; S-HA: saliva-coated hydroxyapatite; SOD: superoxide dismutase; T: testosterone; T/C: testosterone and corticosterone; TL: transfer latency; TNF: tumor necrosis factor; TRC: reward chamber; TT: *Tribulus terrestris* L.; TT-FG: the furostenol glycoside fraction of *T. terrestris*; WETT: water extract of *T. terrestris*.

**Authors’ contributions**

WZ, YD, and LL were involved in preparing the manuscript. HM and YD participated in discussions of views represented in the paper. All authors read and approved the final manuscript.

**Acknowledgements**

The authors gratefully acknowledge the financial support from the National Natural Science Foundation of China (NO. 31501402).

**Competing interests**

The authors declare that they have no competing interests.

**Availability of data and materials**

All data are fully available without restriction.

**Consent for publication**

All authors agree to publish this article.

**Funding**

This paper is financially supported by the National Natural Science Foundation of China (NO. 31501402).

**Publisher’s Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Received:** 21 March 2017  **Accepted:** 3 July 2017  **Published online:** 11 July 2017

**References**

1. Chhatre S, Nesari T, Somani G, Kanchan D, Sathaye S (2014) Phytopharmaceutical overview of *Tribulus terrestris*. Pharmacogn Rev 8(15):45–51
2. Neychev V, Mitev V (2016) Pro-sexual and androgen enhancing effects of *Tribulus terrestris* L.: fact or fiction. J Ethnopharmacol 179:345–355
3. Shang ZJ (2008) Annotation of Shen Nong Ben CaoJ ing. Academy
4. Chinese Pharmacopoeia Commission (2015) Chinese pharmacoepoia (volume I). China Medical Science Press, Beijing, p 352
5. Mohammed MS, Khalid HS, Osman WJA, Mudattahir AK (2014) A review on phytochemical profile and biological activities of three anti-inflamatory plants used in sudanese folklic medicine. Am J Pharm Tech Res 4(4):1–14
6. Akram M, Asif HM, Akhtar N, Shah PA, Uzair M, Shaheen G et al (2011) *Tribulus terrestris* Linn.: a review article. J Med Plants Res 5(16):3601–3605
7. Xu YX, Chen HS, Liu WY, Gu ZB, Liang HQ (1998) Two sapogenins from *Tribulus terrestris*. Phytochemistry 49:199–201
8. Mahato SB, Sahu NP, Gangui AN, Kazumoto M, Toshio K (1981) Steroidal glycosides of *Tribulus terrestris* Linn. J Chem Soc Perkin I 9:2405–2410
9. Xu YX, Chen HS, Liang HQ, Gu ZB, Liu WY, Leung WN et al (2000) Three new saponins from *Tribulus terrestris*. Planta Med 66:545–550
10. Tomova M, Panova D, Wulfson NS (1974) Steroid saponins and saponins IV. Saponins from *Tribulus terrestris*. Planta Med 25:231–237
11. Wang Y, Othann K, Kasiar R, Yasamaki K (1996) Steroidal saponins from fruits of *Tribulus terrestris*. Phytochemistry 42:1417–1422
12. Huang JW, Tan CH, Jiang SH, Zhu DY (2003) Terrestrinins A and B, two new steroidal sapogenins from *Tribulus terrestris*. J Assian Nat Prod Res 4(4):1–14
13. Deepak M, Dipankar G, Prashanth D, Ahsa MK, Armit A, Venkataraman BV (2002) Tribulosin and β-sitosterol-dglucoside.the anthelmintic principles of *Tribulus terrestris*. Phytomedicine 9:753–756
14. De Kock WT, Enslin PR (1958) Chemical investigation of photosensitization diseases of domestic animals I. Isolation and characterization of steroidal sapogenins from *Tribulus terrestris*. Afr Chem Inst 11:33–36
15. Sharma HC, Narula JL (1977) Chemical investigation of flowers of *Tribulus terrestris*. Chem Era 13:15–17
16. Wang Y, Othahn K, Kasiar R, Yasamaki K (1997) Steroidal sapogenins from fruits of *Tribulus terrestris*. Phytochemistry 45:811–881
17. Huang JW, Jiang SH, Tan CH, Zhu DY (2002) Structural elucidation of three new steroidal sapogenins, Chin J Org Chem 22:917–921
18. Wu G, Jiang S, Jiang F, Zhu D, Wu H, Jiang S (1996) Steroidal glycosides from *Tribulus terrestris*. Phytochemistry 42:1677–1681
19. Xu YJ, Xie SX, Zhao HF, Han D, Xu TH, Xu DM (2001) Studies on the chemical constituents from *Tribulus terrestris*. Acta Pharm Sin 36(10):750–753
20. Cai LF, Wu YJ, Zhang JG, Pei FK, Xu YJ, Xie SX et al (2001) Steroidal sapogenins from *Tribulus terrestris*. Planta Med 67:196–198
21. Bedir E, Khart A, Walker LA (2002) Biologically active steroidal glycosides from *Tribulus terrestris*. Pharmazeu 57:491–493
22. Gheorghiu A, Ionescu-Matiu E (1968) Presence of chlorogenin next to diogenin and gitogenin in *Tribulus terrestris*. Ann Pharm 26:745–798
23. Wang ZF, Wang BB, Zhao Y, Wan FX, Sun Y, Guo RJ et al (2016) Furostanol and spirostanol saponins from Tribulus terrestris. Molecules 21:429
24. Kang LP, Wu RL, Yu HS, Pang X, Liu J, Liu LF et al (2014) Steroidal saponins from Tribulus terrestris. Phytochemistry 107:182–189
25. Lan S, Gang C, Feng SG, Wei W, Li ZF, Chen H et al (2009) Steroidal saponins from Tribulus terrestris. Steroids 74:399–403
26. Su L, Feng SG, Qiao L, Zhou YZ, Yang RP, Pei YH (2010) Two new steroidal saponins from Tribulus terrestris. Chin Chem Lett 21:33
27. Matschenko HE, Gulemetova R, Kintya PK, Shashkov AS (1990) A sulfated glycoside from the preparation “Tribestan”. Khim Prir Soedin 6:49–652
28. Conrad J, Dinchev D, Klaiber I, Mika S, Kostova I, Kraus W (2004) A novel furostanol saponin from Tribulus terrestris of Bulgarian origin. Fitoterapia 75:171–122
29. Wilkins AL, Miles CO, De Kock WT, Erasmus GL, Basson AT, Kellerman (2002) A new steroidal saponin from Tribulus terrestris. J Asian Nat Prod Res 4:29–35
30. Sun WJ, Gao J, Tu GZ, Guo Z, Zhang Y (2002) A new steroidal saponin from Tribulus terrestris. Acad Pharm Sin 34:759–761
31. Cai LF, Jing FY, Zhang JG, Pei FK, Xu YJ, Liu SY et al (1999) Steroidal saponins from Tribulus terrestris. Z Naturforsch 57c:33–38
32. Liu T, Lu X, Wu B, Chen G, Hua HM, Pei YH (2010) Two new steroidal saponins from Tribulus terrestris. J. Asian Nat Prod Res 12:30–35
33. Kostova I, Dinchev D, Rentsch GH, Dimitrov V, Livanova A (2002) Two new sulfated furostanol saponins from Tribulus terrestris. Z Naturforsch 57c:33–38
34. Qu NN, Yang SS (2007) Extraction and determination of chemical constituents of flavonoids in Tribulus terrestris. J Liaoning Univ Tradit Chin Med 9(3):182–183
35. Saleh NAM, Ahmed AA, Abdalla MF (1982) Flavonoid glycosides of Tribulus pentandrus and T. ptychostemon. Phytochemistry 21(8):195–2000
36. Marzieh M, Yekta S (2008) Flavonoid Glycosides from Tribulus terrestris L. Sin. J Asian Nat Prod Lett 16:243–247
37. Yang FK, Zhang ML, Quan XL, Xue ML, Cai HX, Yang J (2014) Determination and comparison HPLC of total flavonoids in different parts of Tribulus terrestris. J Ayurveda Clinical Res 4(4):61–64
38. Su L, Feng SG, Qiao L, Zhou YZ, Yang RP, Pei YH (2009) Two new steroidal saponins from Tribulus terrestris. J. Asian Nat Prod Res 11(1):38–43
39. Bhutani SP, Chhibber SS, Seshadri TR (1969) Flavonoids of the fruits and leaves of Tribulus terrestris: constitution of tribuloside. Phytochemistry 8(1):299–303
40. Zhang XP, Wei N, Huang Q, Tan YF, Jin DJ (2012) A new furoyl amide derivative from the fruits of Tribulus terrestris. Nat Prod Res 26(20):1922–1925
41. Wu TS, Shi LS, Kuo SC (1999) Alkaloids and other constituents from Tribulus terrestris. Phytochemistry 50(8):1411–1415
42. Lv AL (2007) Chemical constituents of Tribulus terrestris L. Shenyang Pharmaceutical University, Shenyang
43. Wang Y (1989) Pharmacological action and chemical components of Tribulus terrestris L. (A Review). J Beijing Univ Trad Chin Med 6:30–32
44. Lv AL, Zhang N, Sun MG, Huang YF, Sun Y, Ma HY et al (2008) One new cinamnic imide derivative from the fruits of Tribulus terrestris. Nat Prod Res 22(11):1007–1010
45. Ren YJ, Chen HS, Yang YZ, Zhou H (1994) Isolation and identification of a new cinamnic acid from Tribus terrestris. Acta Pharm Sin 29(303):204–206
46. Wang RV, Chen SY, Yu CY (2009) Chemical constituents of Tribulus terrestris L. J Univ Chem Technol (Nat Sci Ed) 36:79–82
47. Li RH (2006) Studies on bioactive compounds and quality evaluation of Tribulus terrestris L. J Liaoning Univ Tradit Chin Med
48. Chen HS, Chen QJ, Xuan WD (2004) A new organic acid from Tribulus terrestris. J. Liaoning Univ Tradit Chin Med 2(1):126–130
49. Wang XD, Shao JX (1994) Determination of amino acids in Tribulus terrestris L. Amino Acid Biotic Resour 16(2):29–30
50. Lv AL, Zhang Y, Ma HY, Wang D, Dang Q, Pei YH (2007) Chemical constituents of Tribulus terrestris L. Chin J Med Chem 3:170–172
51. Liu J, Chen HS, Xu YX, Zhang WD, Liu WY (2003) Studies on chemical constituents of Tribulus terrestris L. J Acad Second Military Med Univ 24(2):221–222
52. Custers D, Van PN, Courbeille P, Aperis S, Deconinck E (2017) Chromatographic fingerprinting as a strategy to identify regulated plants in illegal herbal supplements. Talanta 164:490–502
53. Sahin K, Othan C, Akdemir F, Tuzcu M, Gencoglu H, Sahin N et al (2016) Comparative evaluation of the sexual functions and NF-κB and NF2 pathways of some aphrodisiac herbal extracts in male rats. BMC Complement Altern Med 16:1–11
54. Kalamegam G, Adaikan PG (2008) The hormonal effects of Tribulus terrestris and its role in the management of male erectile dysfunction—an evaluation using primates, rabbit and rat. Phytomedicine 15:44–54
55. Tyagi RM, Aswair UM, Mohan V, Bodhanakar SL, Zambare GN, Thakuresh PA (2008) Study of furostrol glycoside fraction of Tribulus terrestris on male sexual function in rats. Pharm Biol 46(3):191–198
56. Kamenov Z, Fileva S, Kalinov K (2017) Evaluation of the efficacy and safety of Tribulus terrestris, in male sexual dysfunction—a prospective, randomized, double blinded, placebo-controlled clinical trial. Maturitas 81(1):26
57. Oliveira NNP, Felix MAR, Pereira TCS, Rocha LGP, Miranda JR, Zangemonino MK et al (2015) Sperm quality and testicular histomorphometry of wistar rats supplemented with extract and fractions of fruit of Tribulus terrestris L. Brazarch Biol Techn 58(6):891–897
58. Kumari M, Singh P (2015) Protective role of Tribulus terrestris on alu-minium chloride-induced reproductive toxicity in the male laboratory mouse. Int. J Pharm Pharraceutical Sci 7:295–298
59. Kumar P, Singh P (2015) Tribulus terrestris ameliorates metronidazole-induced spermatozoogenic inhibition and testicular oxidative stress in the laboratory mouse. Indian J Pharmacol 47(3):304–310
60. Khaleghi S, Bakhiani M, Assadmabnnia A, Esmaeeli F (2016) Tribulus terrestris extract improves human sperm parameters in vitro. J Evid Based Complement Altern Med 22(3):1–6
61. Bitzer J, Giraldd A, Plass J (2013) Sexual desire and hypoactive sexual desire disorder in women: introduction and overview. standard operating procedure. J Sexual Med 10(1):36–49
62. De Souza KZD, Vale FBC, Geber S (2016) Efficacy of Tribulus terrestris for the treatment of hypoactive sexual desire disorder in postmenopausal women: a randomized, double-blinded, placebo-controlled trial. Meno-parise 23(11):1252–1256
63. Postgo S, Lima SMRR, Yamada SS, Reis BFD, Silva GMDD, Aoki T (2016) Assessment of the effects of Tribulus terrestris on sexual function of menopausal women. Rev Bras Ginecol Obstet 38:140–146
64. Sharma H, Khan W, Ahmad S (2017) In vitro and ex vivo approach for anti-ultrolicithal potential of bioactive fractions of gokhru with simulta-neous HPLC analysis of six major metabolites and their exploration in rat plasma. Pharm Biol 55(1):701–711
65. Aggarwal A, Tandong S, Singla SK, Tandong (2012) A novel antilithic protein from Tribulus terrestris having cytoprotective potency. Protein Peptide Lett 19(8):812–819
66. Arasararathnam V, Balakumar S, Senthuran A, Rajendraprasad R (2010) A study of Tribulus terrestris extract on risk factors for urinary stone in normal subjects and urological patients. J Nat Sci Found Sri Lanka 39(3):187–191
67. Van FA, Lucassen PL, Akkermans RP, Van de Lisdonk EH, Rutten GE, Van PA (2008) Study of furostenol glycoside fraction of Tribulus terrestris from Bulgarian origin. Fitoterapia 79(4):221–222
68. Ren YJ, Chen HS, Yang YZ, Zhou H (1994) Isolation and identification of a new cinamnic acid from Tribus terrestris. Acta Pharm Sin 29(303):204–206
74. Reshma PL, Sainu NS, Mathew AK, Raghu KG (2016) Mitochondrial dys-
function in H9c2 cells during ischemia and amelioration with Tribulus terrestris. Life Sci 152:220–230
75. Li M, Guan Y, Liu J, Zhai F, Zhang X, Guan L (2013) Cellular and molecular mechanisms in vascular smooth muscle cells by which total saponin extracted from Tribulus terrestris protects against atherosclerosis. Cell Physiol Biochem 32(5):1299–1308
76. Jiang YH, Yang CH, Li W, Wu S, Meng XQ, Li DN (2016) Aqueous extracts of Tribulus terrestris protects against oxidized low-density lipoprotein-induced endothelial dysfunction. Chin J Integr Tradit West Med 22(3):193–200
77. Reshma PL, Lekshmi VS, Sankar V, Raghu KG (2015) Tribulus terrestris (Linn.) Attenuates Cellular Alterations Induced by Ischemia in H9c2 Cells Via Antioxidant Potential. Phytother Res 29(6):933–943
78. Guo Y, Yin HJ, Shi DZ (2006) Effect of xinnao shutong capsule on cardiac muscle cell apoptosis and protein expressions of bcl-2 and bax in hyperlipidemia rats after myocardial infarction. Chin J Integr Tradit West Med 26(6):541–544
79. Fg Zhai, Li HZ, Zhou FB, Lin F, Guan LX (2015) Effects of saponins of Tribulus terrestris on PPARγ and NF-kB signaling pathways expression in rat brain following cerebral ischemic injury. Med Recapitulate 21(4):4539–4540
80. Jiang EP, Su XJ, Li H, Yang SJ (2008) Protective effects and mechanisms of gross saponins of Tribulus terrestris on apoptosis of PC12 cells induced by H2O2. Chin Tradit Herbal Drugs 39(9):1368–1371
81. Li LB, Li J, Li H (2006) Protective effects of gross saponins of Tribulus terrestris on experimental intracerebral hemorrhage in rats. J Harbin Med Univ 40(2):299–102
82. Wang ZJ (2011) Progress on protective mechanisms of single Chinese crude drug and its effective ingredients on optic nerve of glaucoma. Herald Med 26(10):1191–1193
83. Sheng YM, Meng XL (2007) Research advance on the effect of several Chinese traditional medicine on optic nerve protection. Herald Med 26(10):1191–1193
84. Liao MY, Huang LN, Zeng P (2009) Effect of Tribulus terrestris L. on the retinal ganglion cells. Int Eye Sci 9(2):282–283
85. Bucci LR (2000) Selected herbal and human exercise performance. Am J Clin Nutr 72(2 Suppl):6245–6365
86. Di PM (1995) Anabolic steroids substitutes from plants and herbs? Drugs Sports 3:10–12
87. Saudan C, Baumer N, Emery C (2008) Short term impact of Tribulus terrestris intake on doping control analysis of endogenous steroids. Forensic Sci Int 178(1):7–10
88. Jing QQ (1998) The effect of overload-training on the pritary-gonalad axis of rats. Zhejiang Sport Sci 20(1):43–46
89. Yin L, Wang Q, Wang X, Song LN (2016) Effects of Tribulus terrestris saponins on exercise performance in overtraining rats and the underlying mechanisms. Can J Physiol Pharmacol 94(1):1193–1201
90. Gorenova TE, Bozhanov SS, Lozanov VS, Mitov VI, Karieva RP, Georgieva EI (2015) Changes in gene expression of CCR1, CCR2, and CBL, after treatment of breast cancer cells with saponin extract from Tribulus terrestris. Neoplasma 62(1):27–33
91. Sisto M, Lisi S, D’Amore M, De Lucro R, Carati D, Castellana D et al (2012) Saponins from Tribulus terrestris L. protect human keratinocytes from UVB-induced damage. J Photochem Photobiol B: Biol 117:193–201
92. Vala MH, Goudatzhi M, Moghadha SN, Nejad MK, Jahangiri S, Golamí M (2013) In vitro assessment of Tribulus terrestris aqueous extract and Benzoxacin fraction against Helicobacter pylori isolates from biopsy samples of Iranian patients. Novel biomed 1(3):84–87
93. Zhang JD, Zheng XU, Cao YB, Jun GU, Jiang YY (2011) Study on regulation of ERG genes expression in Candida albicans by a new anti-fungi agent TTS-12. Chin Pharm J 46(16):1229–1234
94. Kiran B, Lalitha V, Raveesha KA (2011) In vitro evaluation of aqueous and solvent extract of Tribulus terrestris L. leaf against human bacteria. Int J Pharm Tech Res 3(3):1897–1903
95. Soleimampour S, Sedighinia FS, Safipour AA, Zarif R, Ghazvini K (2015) Antibacterial activity of Tribulus terrestris and its synergistic effect with Capsella bursa-pastoris and Glyceria stratiotes against oral pathogens: an in vitro study. Avicenna J Phytomed 5(3):210–217
96. Bhuvad S, Nitheshwark K (2016) Assessment of free radical scavenging activity of ten madhurakanthda drugs through UV spectroscopic and chromatographic technique. J Pharm Pharmacol 8(3):92–96
97. Li N, Huang LN, Zeng P (2013) Influence of gross saponins from Tribulus terrestris L. on SOD activity and MDA content for chronic high intracranial pressure in rabbit. Int Eye Sci 13(5):854–856
98. Hemalatha S, Hari R (2013) Comparative antioxidative activities of crude ethanolic and saponin rich butanol extracts of Tribulus terrestris fruits. Int J Pharm Bio Sci 4(4):784–793
99. Yogendra KG, Vimala Y, Yogendra KG (2014) Antioxidant activity and RP-HPLC analysis of diosgenin from the callus of Tribulus terrestris Linn. Int J Res Ayurveda Pharm 5(3):343–346
100. Ko HI, Ahn EK, Oh J.S (2015) N-trans-p-cafeoyl tyramine isolated from Tribulus terrestris exerts anti-inflammatory effects in lipopolysaccharide-stimulated RAW 264.7 cells. Int J Med Microbiol 364(6):1042–1048
101. Babuoro B, Rayalakshimi G, Venkatesham A, Kiran G, Shyam Sunder A, Gangla Rao B (2009) Anti-inflammatory and antimicrobial activities of methanolic extract of Tribulus terrestris Linn plant. Int J Chem Sci 16(2):209–218
102. Hu DH (2009) Effect of gross saponins from Tribulus terrestris on hepatic apoptosis in mice’s acute hepatic injury induced by tripterygium glycosides. Hebei University Chinese Medicine, Heibe
103. Parimala DR (2011) In-vitro anthelmintic activity of methanolic extract of aerial parts of Tribulus terrestris Linn. J Global Pharma Technol 3(6):1–3
104. Bansal SK, Singh KV, Sharma S (2014) Larvicidal potential of wild mustard (Cleome viscosa) and gokhru (Tribulus terrestris) against mosquito vectors in the semi-arid region of Western Rajasthan. J Environ Biol 35(2):327–332
105. Oh HK, Park SJ, Moon HD, Jun SH, Choi NY, You YO (2011) Tribulus terrestris inhibits caries-inducing properties of Streptococcus mutans. J Med Plants Res 5(25):6061–6066
106. Zhu JW (2011) Effect of gross saponins from Tribulus terrestris on skin morphology of g-catalase-induced aging mice. Chin J Gerontol 31(3):4628–4630
107. Zhang J, Zhang DS, Yan C.L (2007) Effects of GSTT on dysmenia in mice. Pharmocol Clin Chin Mater Clin Med 23(5):47–49
108. Prabhu N (2014) Effect of Tribulus terrestris on learning and memory in wistar rats. Phcog J 4(6):68–71
109. Ayyanna C, Chandra Mohan Rao G, Sasikala M, Somasekhar P, Arun Kumar N, Pradeep Kumar MVS (2012) Absorption enhancement studies of ten madhuraskandha drugs through UV spectroscopic and chromatographic technique. J Pharm Pharmacol 46(1):4118–4125
110. EI-Shaibany A, Al-Habori M, Al-Tahami B (2015) Anti-hyperglycaemic activity of Tribulus terrestris L. against oral pathogens: an in vitro study. Avicenna J Phytomed 5(3):210–217
111. Abudayyak M, Jannuzzi AT, Özhan G, Alpertunga B (2015) Investigation on the toxic potential of Tribulus terrestris in vitro. Pharm Biol 53(4):469–476