Multifaced regulator: RNA binding proteins and their roles in hematopoiesis

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Abstract
Despite the conventional definition of RNA binding proteins (RBPs) as controlling the metabolism of their bound RNAs, more and more RBPs are found to function through distinct ways in complex biological processes. With the recent discovery of transcriptional regulation activity of some RBPs, a hypothesis that RBPs could be multifaceted regulators orchestrating gene expression has emerged. Hematopoiesis is a stepwise process that needs to be fine-tuned to keep the subtle balance between hematopoietic stem cell (HSC) stemness maintenance and downstream lineage commitment. Although the classic RBPs account for the posttranscriptional regulation in hematopoiesis, the importance and multiple regulatory capacities of RBPs have not been well-characterized. In this review, we summarize the recent findings of large-scale screening of novel RBPs and their novel transcriptional regulation potentials. In hematopoietic system, this kind of multifaceted regulators account for nearly a half of functional RBPs. Therefore, further studies on identifying this new kind of multifaceted RBPs and clarifying their regulatory mechanisms would help us better understand the precise and complex regulatory networks of gene expression in hematopoiesis.

Keywords: Hematopoiesis, RNA binding protein

1. RBPS ARE DIVERSIFIED REGULATORS INVOLVED IN DIFFERENT REGULATION PATHWAYS

So far, there are approximately 20,000 protein-coding genes in human genome, of which over 1500 have been annotated as RNA binding proteins (RBPs) that regulate the biogenesis, fate, and function of RNAs. Conventional RBPs take part in the formation of ribonucleoprotein complex and govern RNA processing to maintain fundamental gene expression. RBPs recognize and bind to specific sequences or structures in RNA molecules via a set of defined RNA binding domains (RBDs), although studies on decoding ribonucleoprotein structures revealed the existence of protein–RNA interactions that do not require classical RBDs, and this unconventional type of protein–RNA interaction is more common than anticipated.

The canonical regulation mode of an RBP is to take “good care” of its target RNAs on different aspects; however, emerging evidences have indicated that the RNA-modulating pathway is not universally applicable, as some RBPs can be recruited by RNA, while the others can work independently, for instance, PKR (interferon-induced, double-stranded RNA-activated protein kinase) and RIGI (retinoic acid-inducible gene I protein) can be activated by abnormal RNA molecules derived from virus replication and TRIM25 can activate the production of interferon upon the stimulation of viral RNA molecules. Furthermore, several RBPs have been reported to act on chromatin, thus regulating gene transcription. For example, Lin28A binds to genomic regions near the transcription start sites and activates gene transcription by interfering with DNA methylcytosine oxidase (Tet1). Another study in Arabidopsis thidopsis revealed that AGO1, a classical RBP involved in RNA interference, could also bind to specific chromatin regions with SWI/SNF complex to activate gene expression. Moreover, it was reported that hnrNPU (heterogeneous nuclear ribonucleoprotein U) bound to the active chromatin regions and facilitated the maintenance of the 3D structure of chromatin together with CTCF, RAD21 and other chromatin structural proteins. Currently, the traditional conception of RBPs is being overturned gradually, and increasing evidences prompt us to redefine the role of RBP in multiple biological processes not only as a key regulator of RNA metabolism.

2. RBPS PLAY CRITICAL ROLE IN HEMATOPOIESIS AT THE POSTTRANSCRIPTIOAL LEVEL

Hematopoiesis is a stepwise process that requires multilayered regulation to keep a subtle balance between hematopoietic stem cell (HSC) stemness maintenance and downstream lineage commitment.
Any perturbation in the balance would lead to severe pathological phenotypes. On the other hand, efforts to discover novel key regulators or pathways that regulate the hematopoietic homeostasis may offer great opportunities to manipulate the composition of hematopoietic cell pool, thus shedding lights on clinical management of malignant hematological diseases.

Although numerous studies focused on transcriptional regulation have proved the crucial role in hematopoiesis, transiently activating the pluripotency of HSC and its capacity to differentiate into all blood cell subtypes by cell fate decision make it an extraordinary complex system that must be controlled precisely at multiple levels. Besides the classical “checkpoint” function on the way from DNA to RNA, the regulatory events occurring on the RNA molecules have won their reputation as a detrimental process in hematopoiesis in recent years. This posttranscriptional regulation mechanism forms an additional level for the rapid and precise orchestration of protein expression during hematopoiesis. Importantly, it is well-recognized that RBPs are the key players involved in this regulation network.

So far, several RBPs have been identified to regulate hematopoiesis via regulating RNA splicing, RNA modification, RNA translation, or RNA decay. The RNA splicing is the major component of RBP-regulated pathways. For example, the RBP Rbm15 regulates the splicing of c-Mpl receptor and affects the thrombopoietin signaling pathway, thus contributing to the quiescence and proliferation of HSC. Another splicing factor Srsf2 has a role in HSC production and hematopoietic reconstitution in mouse model. Previously, we identified QKI5 could activate the processing of primary miR-124-1 and decreased QKI5 during erythropoiesis led to concomitant reduction of mature miR-124, which facilitated erythrocyte maturation. Furthermore, we also confirmed that RBP KSRP promoted the processing of primary miR-129, thus inducing granulocyte differentiation at the level of monocyte–macrophage differentiation. The most well-characterized modification on RNA is N6-methyladenosine (m6A), which is dominated by the “writer” protein METTL3 and METTL14. METTL3 depletion induces loss of m6A modification, which then triggers the expression of hematopoietic differentiation-associated genes; therefore, Mettl3-deficient mice displayed T cell activation deficiency. RBP Musashi-2 (MSI2), a repressor of mRNA translation, has a key role in HSC self-renewal and lineage determination by modulating TGF-β signaling pathway. RBP ZFP36 has been reported to control erythroid differentiation by regulating RNA decay. RBP Musashi-2 (MSI2), a repressor of mRNA translation, has a key role in HSC self-renewal and lineage determination by modulating TGF-β signaling pathway. RBP ZFP36 has been reported to control erythroid differentiation by regulating RNA decay. However, the current knowledge of RBP’s function in hematopoietic system is still very limited and mainly focus on the RNA-binding-associated activities; thus, further investigation of multifunctional potency of the hematologic RBPs is highly demanded.

3. INSIGHTS INTO THE MULTILAYERED REGULATORY POTENCY OF NOVEL RBPS IN HEMATOPOIETIC SYSTEM

Currently benefited from the development of large-scale screening approaches for novel RBPs, we can have a glimpse
of the newly identified multipotent RBPs in hematopoiesis. According to two most commonly used databases, RBPdb and ATTRACTION, there are only 441 classical RBPs annotated (Fig. 1A). However, Castello et al identified 837 proteins that could bind to RNA directly using a poly(A)-baiting protein capture screening system. Among these recognized RBPs, 602 were found to interact with RNA for the first time (Fig. 1B). Another outstanding work carried out by Trendel et al described a method named “XRNAX,” which extracted protein-cross-linked RNAs based on organic phase separation. “XRNAX” could purify broad-spectrum RNA and also provided a new protocol for RNA binding protein screening, which picked up 1238 RBPs, among them 995 were newly identified (Fig. 1B). These two studies revealed in total 1137 novel RBPs that accounted for up to 70% of all RNA–protein interaction events as compared to classical ones that only hit less than 30%.

As for hematopoietic system, we realized that the importance of RBP in hematopoiesis should be recharacterized because of the ever-expanding list of RBPs. Therefore, we obtained 954 hematopoiesis-associated genes from QuickGO database. When we investigated the gene sets, only 24 classical RBPs were involved in hematopoietic system. However, 66 RBPs from those newly discovered RBP set were found to be associated with different pathways in hematopoiesis (Fig. 1C). The increasing number of RBPs demonstrated its crucial role of RNA-mediated functions in hematopoietic system.

Indeed, the field of RBPs is far beyond our current understanding, because not only a good deal of potential RBPs are needed to be uncovered, but also the inadequate knowledge of the multilayered regulation capacity of RBPs. A research profiling the human protein–DNA interactome revealed 4191 DNA binding proteins (DBPs). It is notable that when we looked into this large set of RBPs, we observed 405 newly identified RBPs and 219 classical RBPs (Fig. 1D). These discoveries prompted us to reconsider the definition between DBP and RBP, and the multiple regulatory potentials of RBP on both posttranscriptional and transcriptional levels. For the hematopoietic system, we found 37 RBPs that were predicted to have the ability to bind both RNA and DNA. They accounted for about 40% in all hematopoietic RBPs, suggesting a rather common phenomenon for RBPs to have multifunction in one system. Moreover, from the Encode project, we found 92 RBPs with their defined ChIP-seq data, further proving that RBPs indeed have multiple regulatory capacities, while 7 of them were functionally proved in hematopoietic system.

Taken together, in complex systems such as hematopoiesis, regulators tend to function precisely at multiple levels. Case studies and large-scale screening have demonstrated that RBPs could regulate gene expression at different levels. Development of new techniques will definitely provide new insights of RBPs and their novel regulatory pathways.

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REFERENCES

[1] Gerstberger S, Hawker M, Tuschl T. A census of human RNA-binding proteins. Nat Rev Genet 2014;15(12):829–845.
[2] Dreyfuss G, Kim VN, Kataoka N. Messenger-RNA-binding proteins and the messages they carry. Nat Rev Mol Cell Biol 2002;3(3):195–203.
[3] Lunde BM, Moore C, Varami G. RNA-binding proteins: modular design for efficient function. Nat Rev Mol Cell Biol 2007;8(6):479–490.
[4] Ramakrishnan V. The ribosome emerges from a black box. Cell 2014;159(5):979–984.
[5] Steitz TA. A structural understanding of the dynamic ribosome machine. Nat Rev Mol Cell Biol 2008;9(3):242–253.
[6] Behrmann E, Loerke J, Budkevich TV, et al. Structural snapshots of actively translating human ribosomes. Cell 2015;161(4):845–887.
[7] Matera AG, Wang Z. A day in the life of the spliceosome. Nat Rev Mol Cell Biol 2014;15(2):108–121.
[8] Papasaiikos P, Valcarcel J. The spliceosome: the ultimate RNA chaperone and sculptor. Trends Biochem Sci 2016;41(1):33–45.
[9] Plaschka C, Lin PC, Nagai K. Structure of a pre-catalytic spliceosome. Nature 2017;546(7660):617–621.
[10] Dabo S, Meurs EF. dsRNA-dependent protein kinase PKR and its role in stress, signaling and HCV infection. Viruses 2012;4(11):2598–2635.
[11] Habjan M, Pichlmair A. Cytosplasmic sensing of viral nucleic acids. Curr Opin Virol 2015;11:31–37.
[12] Rehwinkel J. RNA sensing: the more RIG-I the merrier? EMBO Rep 2013;14(9):751–752.
[13] Guich MU, Shao YC, Joo CH, et al. TRIM25 RING-finger E3 ubiquitin ligase is essential for RIG-I-mediated antiviral activity. Nature 2007;446(7138):916–920.
[14] Zeng Y, Yao B, Shin J, et al. Lin28A binds activators and promotes Tert to regulate gene expression. Mol Cell 2016;61(1):153–160.
[15] Liu C, Xin Y, Xu L, et al. Arabidopsis ARGONAUTE 1 binds chromatin to promote gene transcription in response to hormones and stresses. Dev Cell 2018;44(3):348–361. e347.
[16] Fan H, Lv P, Hua X, et al. The nuclear matrix protein HNRNP maintains 3D genome architecture globally in mouse hepatocytes. Genome Res 2018;28(2):192–202.
[17] Nerlov C, Querfurth E, Kaulsa H, Graf T. GATA-1 interacts with the myeloid PU.1 transcription factor and represses PU.1-dependent transcription. Blood 2000;95(8):2534–2535.
[18] Wilson NK, Foster SD, Wang X, et al. Combinatorial transcriptional control in blood/stem/progenitor cells: genome-wide analysis of ten major transcriptional regulators. Cell Stem Cell 2010;7(4):532–544.
[19] Grech G, von Lindern M. The role of translation initiation regulation in haematopoiesis. Comp Funct Genomics 2012;2012:677640.
[20] Xiao N, Laha S, Das S, Morlock K, Jesneck JL, Raffel GD. Ott1 regulates primary miR-124-1 processing via a distal RNA motif during hematopoiesis. EMBO Rep 2014;15(11):1479–1485.
[21] Komeno Y, Huang YJ, Qiu J, et al. SRSF2 is essential for hematopoiesis, and its myelodysplastic syndrome-related mutations dysregulate alternative pre-mRNA splicing. Mol Cell Biol 2015;35(17):3071–3082.
[22] Kim E, Ilagan JO, Liang Y, et al. SRSF2 mutations contribute to myelodysplasia by mutant-specific effects on exon recognition. Cancer Cell 2017;27(5):617–630.
[23] Wang F, Song W, Zhao H, et al. The RNA-binding protein QKIS regulates primary miR-124-1 processing via a distal RNA motif during erythropoiesis. Cell Res 2017;27(3):416–439.
[24] Zhao H, Wang X, Yi P, et al. KSRP specifies monocytic and granulocytic differentiation through regulating miR-129 biogenesis and RUNX1 expression. Nat Commun 2017;8(1):1428.
[25] Meyer KD, Jaffrey SR. The dynamic epitranscriptome: N6-methyladenosine and gene expression control. Nat Rev Mol Cell Biol 2014;15(5):313–326.
[26] Barbieri I, Tzelepis K, Pandolfi L, et al. Promoter-bound METTL3 maintains myeloid leukemia by m6A-dependent translation control. Nature 2017;552(7683):126–131.
[27] Yu LP, Piccirillo BF, Cheng Y, et al. The N6-methyladenosine (m6A)-forming enzyme METTL3 controls myeloid differentiation of normal hematopoietic and leukemia cells. Nat Med 2017;23(11):1369–1376.
[28] Li HB, Tong J, Zhu S, et al. m(6)A mRNA methylation controls T cell homeostasis by targeting the IL-7/STAT5/SOCS pathways. *Nature* 2017;548(7667):338–342.

[29] Park SM, Deering RP, Lu Y, et al. Musashi-2 controls cell fate, lineage bias, and TGF-beta signaling in HSCs. *J Exp Med* 2014;211(1):71–87.

[30] Stumpo DJ, Broxmeyer HE, Ward T, et al. Targeted disruption of Zip36L2, encoding a CCHC tandem zinc finger RNA-binding protein, results in defective hematopoiesis. *Blood* 2009;114(12):2401–2410.

[31] Zhang L, Prak L, Rayon-Estrada V, et al. ZFP36L2 is required for self-renewal of early burst-forming unit erythroid progenitors. *Nature* 2013;499(7456):92–96.

[32] Castello A, Fischer B, Eichelbaum K, et al. Insights into RNA biology from an atlas of mammalian mRNA-binding proteins. *Cell* 2012;149(6):1393–1406.

[33] Trendel J, Schwarzl T, Horos R, et al. The human RNA-binding proteome and its dynamics during translational arrest. *Cell* 2019;176(1–2):391–403. e319.

[34] Ashburner M, Ball CA, Blake JA, et al. Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat Genetics* 2000;25(1):25–29.

[35] The Gene Ontology Consortium. The Gene Ontology Resource: 20 years and still GOing strong. *Nucleic Acids Res* 2019;47((D1):D330–D338.

[36] Hu S, Xie Z, Onishi A, et al. Profiling the human protein–DNA interactome reveals ERK2 as a transcriptional repressor of interferon signaling. *Cell* 2009;139(3):610–622.

[37] Davis CA, Hitz BC, Sloan CA, et al. The Encyclopedia of DNA elements (ENCODE): data portal update. *Nucleic Acids Res* 2018;46((D1):D794–D801.