The effectiveness of antibiotic cement-coated nails in post-traumatic femoral and tibial osteomyelitis – comparative analysis of custom-made versus commercially available nails

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Abstract. Background: The first objective of this retrospective study was to assess infection control rates in patients with chronic post-traumatic osteomyelitis (CPTO) of the femur or tibia treated with antibiotic cement-coated nails. The second objective was to compare the efficacy of custom-made nails versus commercially available antibiotic-coated nails in terms of infection control and need for reoperation. Methods: We reviewed a consecutive series of CPTO patients treated with antibiotic-coated nails who had a minimum follow-up of 24 months. We recorded the characteristics of the initial injury, the type of nail used, cement–nail debonding, infecting microorganisms, operating time, infection control, need for reoperation, and failure rate. We performed a comparative analysis between nails manufactured in the operating room (i.e., custom-made) and those commercially available. Results: Thirty patients were included. The affected bones were the femur (n = 15) and the tibia (n = 15). Twenty-one of the 30 initial injuries were open fractures. Staphylococcus aureus was the most frequently isolated microorganism (50 %). Sixteen patients were treated with custom-made nails and 14 with commercially available antibiotic-coated nails. At the time of extraction, four out of five custom-made antibiotic-coated nails experienced cement–bone debonding. Commercial nails were associated with shorter operating times (p < 0.0001). The overall infection control rate was 96.66 %. Eight (26.66 %) patients needed reoperation. There was one failure (3.33 %) in the group treated with custom-made antibiotic-coated nails. We did not find significant differences between nail types in terms of reoperation, infection control, and failure rate. Conclusions: The use of antibiotic cement-coated nails proved useful in CPTO treatment. Commercially available nails had significantly shorter operating times and did not present cement–bone debonding during removal. Our results seem to indicate that both nail types are similar in terms of infection control and reoperation rates.

1 Introduction

Post-traumatic osteomyelitis (PTO) is a challenging condition both for surgeons and patients (Hake et al., 2015). It can occur after the surgical treatment of closed or open fractures, though it is more frequent in the latter. Its reported prevalence ranges from 0.7 % to 33 % (Patzakis and Zalavras, 2005; Patzakis et al., 1986; Barger et al., 2017; Chadayammuri et al., 2017; Beals and Bryant, 2005).

At present, patient management practices include aggressive surgical debridement, bone stabilization, adequate soft tissue coverage, and the administration of specific antimicrobials depending on the infecting microorganism (Patzakis and Zalavras, 2005; Beals and Bryant, 2005; Tetsworth and Cierny, 1999; Cierny et al., 2003). Another important aspect to take into consideration is dead space management generated by the resection of devitalized tissue, bone sequestrum, and/or the removal of previous osteosynthesis (Hake et al.,...
host immune response and systemic antibiotic therapy tend to be weak (Cierny et al., 2003; Koury et al., 2017; Gosselin et al., 2004; Wasko and Borens, 2013). The use of antibiotic-loaded polymethylmethacrylate (PMMA) beads described by Klemm (1979) has proven effective for the local administration of antimicrobials; however, when the medullary cavity is affected by the infection – due to dissemination or colonization of an intramedullary nail – placing and removing the beads from this cavity is usually difficult (Klemm et al., 1998; Barger et al., 2017). In these cases, an antibiotic-coated intramedullary metal core can be used (Barger et al., 2017; Koury et al., 2017; Paley and Herzenberg, 2002; Bhadra and Roberts, 2009). Since they were first described, these devices have demonstrated efficacy in the treatment of long bone osteomyelitis with infection control rates of up to 80%–100% (Klemm et al., 1998; Bhadra and Roberts, 2009; Conway et al., 2014; Wasko and Kaminski, 2015; Makhdom et al., 2020).

Kanakaris et al. (2014) reported a 96% success rate in a series of 24 PTO patients at 21 months of follow-up with the use of rods as a metal core.

Among other advantages, these intramedullary devices offer high local antibiotic concentration and may even provide mechanical support – depending on the type of metal core used in the manufacturing process (Barger et al., 2017; Kim et al., 2014).

While many studies in the literature have assessed the efficacy of antibiotic cement-coated nails (Wasko and Borens, 2013; Conway et al., 2014; Wasko and Kaminski, 2015; Kanakaris et al., 2014; Kim et al., 2014), few have made a comparative assessment of the results obtained with custom-made nails made in the operating room versus commercially available nails.

The main objective of this retrospective study was to assess the effectiveness of antibiotic cement-coated nails to control infections in a series of patients with chronic post-traumatic osteomyelitis (CPTO) of the femur or tibia. Our second objective was to compare infection control and re-operation and failure rates of custom-made versus commercially available antibiotic-coated nails.

## 2 Materials and methods

We retrospectively reviewed our department’s database to identify consecutive patients treated with an antibiotic cement-coated nail for post-traumatic osteomyelitis between January 2010 and December 2017. The diagnosis of osteomyelitis was suspected when some of the following clinical, laboratory or imaging parameters were found: swelling, redness, pain, fever (>38°C), or active fistula or wound drainage with bone exposure or osteolysis; white blood cell count, erythrocyte sedimentation rate, and C-reactive protein; or cloacae, sequestra, involucrum, or periostitis on radiographs and/or MRI. Diagnosis was confirmed according to the Center for Disease Control and Prevention (CDC) guidelines, when at least one of the following criteria was present: growth of a microorganism in bone or soft tissue culture of intraoperative samples, histopathological analysis compatible with osteomyelitis of intro-operative bone samples, and at least two of the clinical criteria mentioned above (Centers for Disease Control and Prevention, 2016). Injuries of more than 3 months of evolution from onset or initial surgery were defined as chronic (Hotchen et al., 2017).

The data obtained for this initial sample were cross-checked against medical records and radiographic images kept at our institution to select the final cohort. This study was conducted with the approval of our institution’s ethics committee.

We included patients aged ≥18 years old with a CPTO diagnosis of the femur or tibia treated with antibiotic cement-coated intramedullary nailing and a minimum follow-up of 24 months after nailing.

Exclusion criteria involved patients whose histopathological diagnosis of osteomyelitis (based on intraoperative samples) could not be confirmed on medical records, and those with non-union at the time of antibiotic nailing or segmental bone defects (SBDs) secondary to debridement or injury were excluded (Cierny–Mader type IV osteomyelitis) (Cierny et al., 2003).

### 2.1 Treatment protocol

Each case was analyzed separately. The surgery started with the removal of the previous osteosynthesis material. According to the preoperative plan, we made a bony window where the bone was most affected (e.g. sequestrum) and performed an aggressive surgical debridement, resecting sequestra, cloacae, and devitalized tissue until healthy bleeding (paprika sign) was reached (Tetsworth and Cierny, 1999; Patzakis and Zalavras, 2005; Simpson et al., 2001; Blanchette et al., 2018). The medullary cavity was progressively reamed and washed with 6L of saline solution (Tetsworth and Cierny, 1999; Beals and Bryant, 2005). We took multiple (at least five) bone samples – including reamed material – for the histopathological analysis, culture, and antibiotic sensitivity test (Morgenstern et al., 2018; Chadayammuri et al., 2017; Hake et al., 2015). Afterwards, we implanted an antibiotic-coated nail 1.5–2mm smaller in diameter than the last reamer used. When the last reamer was ≤13mm, the metal core selected was an Ender nail (5mm). When the last reamer was larger, an interleaving nail was implanted.

After surgery, patients were started on empirical systemic antibiotics (vancomycin 1g/12h; ceftazidime 2g/8h) until they were switched to specific ones according to culture results of intra-operative samples. According to the procedures of our institution, intraoperative samples were processed within 6h. This process started with the homogeniza-
tion of the tissues in brain–heart infusion (BHI) broth, and then they were inoculated on aerobic and anaerobic sheep blood agar, chocolate agar, and thioglycolate broth. Aerobic and anaerobic plates were incubated aerobically at 37°C in 5%–7% CO2 for 7 d and anaerobically at 37°C for 14 d. Samples in thioglycolate were incubated for 14 d and subsequently cultured on blood agar. Identification of germs that grew on the plates and antibiograms was performed with standard techniques.

Clinical and radiological controls were performed on weeks 3 and 6, on months 3, 6 and 12, and annually thereafter. Our institution’s infectious disease department was in charge of antibiotic treatment and monitoring.

2.2 Nail coating technique
In order to design the metal core, plastic chest tubes of different diameters were used as molds based on preoperative planning and the diameter of the medullary canal. In all cases, the tube inner diameter was at least 2–3 mm larger than the diameter of the nail to be used. The tube was filled with antibiotic cement, and the nail was inserted. In the case of interlocking nails, a Steinmann pin was passed through the nail locking holes before adding the cement. The tube was allowed to set, sectioned lengthwise, and removed (Fig. 1). Physicians used Simplex cement (Stryker; Mahwah, NJ, USA) to manufacture the nails, adding vancomycin 2 g and gentamicin 0.5 g/40 g of cement.

Commercial nails (Subiton, Laboratorios SL, Buenos Aires, Argentina) had a stainless-steel core, interlocked, with 1 mm of gentamicin-impregnated cement (according to the manufacturer’s specifications). According to medical records, surgeons used five nails with 665.0 mg of gentamicin, four with 698.1 mg, two with 732.6 mg, two with 642.4 mg, and one with 777.1 mg.

2.3 Definitions
Treatment was defined as successful when the infection was eradicated and the patient showed no clinical, laboratory, or radiographic signs or symptoms of infection for at least 24 months.

Additional procedure was defined as any reoperation performed after placement of the antibiotic-coated nail. Infection recurrence was defined as an initially controlled infection that relapsed and required new surgical treatment and nail exchange with or without systemic antimicrobial therapy. (Hake et al., 2015; Tice et al., 2003). Treatment failure was defined as the absence of infection control at the end of follow-up.

2.4 Variables analyzed
We analyzed patients’ demographics, comorbidities, and relevant data about initial injury. Patients were classified using the Cierny–Mader classification based on comorbidities and osteomyelitis characteristics (Cierny et al., 2003).

We recorded all intraoperative complications related to the removal of the previous implant and the bone window, as well as the antibiotic used in the cement of custom-made nails manufactured in the operating room (Ender or interlocking nail) and in commercial nails, the soft tissue coverage procedures, operating time, and the results of intraoperative cultures.

Infection-related laboratory markers were also documented: erythrocyte sedimentation (ESR), normal value (nv) < 16 mm/h; C-reactive protein (CRP), nv: < 0.3 mg/dL, and white blood cell count (WBC), nv: < 9000/mm3, at the time of treatment and on the last control in patients’ clinical records. The development, if any, of antibiotic-related allergies, nephrotoxicity, and ototoxicity was also recorded.

Infection control, reoperation, and failure rates at the end of follow-up were also registered.

2.5 Statistical analysis
Statistical analysis was performed using GraphPad PRISM-8.2 software, Inc., San Diego, USA. Quantitative data were expressed as median values and interquartile range (IQR), and non-continuous data were expressed as frequency and percentage. Fisher’s nonparametric or Mann–Whitney tests were used to establish significant associations between variables. A value of p < 0.05 was considered statistically significant.

3 Results
Out of the 55 patients identified in our database, 20 were excluded for presenting non-union or segmental bone defects, two for being under 18 years of age, two for not complying with minimum follow-up requirements, and one for lack of histopathological confirmation of chronic osteomyelitis in medical records.

3.1 Study population and patient characteristics
The final series was made up of 30 patients, 27 males and 3 females, with a median age of 33 (IQR 24–43) years. As regards pre-operative comorbidities, 4 patients were smokers, 2 were diabetic, and 1 presented chronic renal disease. None had a history of heart disease, anemia, poor vascularity (chronic lymphedema, venous stasis, major vessel compromise or arteritis), rheumatoid arthritis, immune deficiency, steroids or immunosuppressant medication, malnutrition, drug or alcohol abuse, or body mass index > 30. Twenty-eight patients were American Society of Anesthesiologists (ASA) I–II, and two were ASA III. Table 1 presents a description of patients.

The femur was the affected bone in 15 cases (9/15–60.00% open fractures) and the tibia in 15 cases (11 open fractures). We recorded all intraoperative complications related to the removal of the previous implant and the bone window, as well as the antibiotic used in the cement of custom-made nails manufactured in the operating room (Ender or interlocking nail) and in commercial nails, the soft tissue coverage procedures, operating time, and the results of intraoperative cultures.

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fractures, 73.3%). All patients had previously undergone osteosynthesis, and the most frequent method used was intramedullary nailing (25; 83.33%) – in four cases with antibiotic-coated nails. The median time between initial injury and antibiotic nail treatment was 10 (IQR 6.5–15) months. According to the Cierny–Mader classification, 18 (60.00%) bone infections were classified as type 3A, 8 (26.66%) as type 1A, 3 (10.00%) as type 1B, and 1 (3.33%) as 3B.

Complications related to the removal of previous implants occurred in 75.00% (three out of four) of the cases in which a custom-made antibiotic-coated nail was removed. In these cases, there was cement debonding inside the medullary canal, and a second bony window had to be opened for removal (Fig. 2). There were no complications related to the removal of non-coated intramedullary nails. We recorded no fractures associated with the bony window performed during debridement. Of the 30 patients, 13 (43.33%) required soft-tissue coverage procedures. Nine of these procedures were performed on the leg (eight local muscle flaps and one cross-leg flap). Three thigh coverage defects were treated with negative pressure wound therapy and skin grafts.

### Microbiological results

The mean number of samples sent for culture and histopathological analysis was five (range four–eight). Microbiological analysis of intraoperative samples revealed 28 (93.3%) positive results, while all histopathological samples were positive. *Staphylococcus aureus* was the most frequently isolated germ (14/28; seven methicillin-resistant and seven methicillin-sensitive). Of the 14 staphylococcal infections, nine were controlled, and five required reoperations. One out of two *Pseudomonas aeruginosa* infections and two out of six polymicrobial infections required reoperation. Failure occurred in a *Proteus mirabilis* infection. We found no significant differences in reoperations when infections were generated by an *S. aureus* vs. another microorganism \((p = 0.99)\) or between mono- vs. polymicrobial infections \((p = 0.64)\).

### Laboratory outcomes

Humoral parameters showed a significant decrease in 29 cases at the end of the study. ESR decreased from 44.32 ± 18.81 mm/h pre-operatively to 12.96 ± 6.13 mm/h \((p = < 0.0001)\) at the end of the study. CRP went from 9.15 ± 9.96 to 0.3 ± 0.29 mg/dL \((p = < 0.0001)\), and WBC went from 13 518 ± 3155 to 7 400 ± 2068 \((p = < 0.0001)\). There were no cases of antibiotic allergy, nephrotoxicity, or ototoxicity.

### Infection control rate

The overall median follow-up was 69 (range: 45–96) months. At the end of follow-up, 29 (96.6%) patients were clinically well and showed no clinical, radiological, or laboratory ev-
idence of infection. The infection control rate without the need for additional procedures was 70.00% (n = 21).

3.5 Overall reoperation and failure rates

Eight (26.66% – patients: 23 to 30) patients needed additional surgical debridement and replacement of the antibiotic-coated nail. In two of them, soft-tissue-covering procedures were also performed (p: 23; 27). The median time to reoperation was 2.5 (range; 1–12) months. It should be noted that during nail removal, two of the custom-made antibiotic-coated nails showed nail–cement debonding. The overall rate for this complication was 62.50% (five out of eight). In all cases the type of nail used for reoperation was the same as the one implanted in the first surgery. None of the patients who underwent these procedures required another surgery. When we comparatively analyzed the characteristics of reoperated and non-reoperated patients, we found no significant differences in terms of age (p = 0.18), sex (p = 0.99), affected bone (p = 0.68), open versus closed fractures (p = 0.52), number of previous surgeries (p = 0.18), time from the initial trauma to antibiotic-coated nail placement (p = 0.88), presence of soft tissue coverage defects (p = 0.42), use of commercial versus custom-made nails (p = 0.99), or microorganism isolated in intraoperative samples (SA vs. other (p = 0.99); mono- vs. polymicrobial p = 0.64).

There was one failure (3.33%) in a patient with a Gustilo 3B gunshot proximal tibial fracture referred to our center with an active fistula. The patient responded well to the initial treatment with an antibiotic-coated nail, but 11 months later his infection recurred. When the treatment was repeated, the patient is currently under treatment.

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**Table 1. Summary description of the preoperative characteristics, treatment and evolution of the patients in the series.**

| Patients | Gender | Bone injury Gustilo | Prev. surgery (n) | Cierny– Mader | Treatment: previous implant/ATB nail type | Soft tissue coverage | Microbiology | Outcome | Follow-up (months) |
|----------|--------|---------------------|------------------|--------------|------------------------------------------|---------------------|--------------|---------|-------------------|
| 1        | M45    | T 3B                | 4                | 3A           | IMN/ILN                                  | LMF                 | P. mirabilis | Failure | 38                |
| 2        | M56    | T 2                 | 2                | 3A           | IMN/EN                                   | –                   | MSSA         | Control | 45                |
| 3        | M36    | F                   | 2                | 3A           | IMN/EN                                   | –                   | C-N          | Control | 92                |
| 4        | M35    | T                   | 4                | 1A           | IMN*/A/EN                                | LMF                 | MSSA         | Control | 120               |
| 5        | M22    | F 3B                | 2                | 3A           | EF/EN                                    | VAC SG              | MRSA         | Control | 121               |
| 6        | M41    | T                   | 2                | 3A           | IMN/EN                                   | –                   | MSSA         | Control | 110               |
| 7        | M70    | F 3B                | 2                | 3B           | IMN*/A/ILN                               | –                   | CoNS         | Control | 50                |
| 8        | M27    | F 2                 | 2                | 1A           | IMN/ILN                                  | –                   | MRSA         | Control | 32                |
| 9        | M24    | T 2                 | 2                | 1A           | IMN/EN                                   | –                   | E. faecalis  | Control | 47                |
| 10       | M18    | F                   | 3                | 3A           | IMN/EN                                   | –                   | MRSA         | Control | 42                |
| 11       | M20    | F                   | 2                | 3A           | IMN/ILN                                  | –                   | MSSA         | Control | 40                |
| 12       | M69    | F 3B                | 9                | 1B           | Plate/EN                                 | –                   | P²           | Control | 67                |
| 13       | M47    | F 2                 | 2                | 1B           | IMN/IN                                   | VAC SG              | MSSA         | Control | 68                |
| 14       | F18    | T 3C                | 3                | 3A           | EF/IN                                    | LMF                 | P. aeruginosa | Control | 100               |
| 15       | M29    | T 3B                | 10               | 1A           | IMN/IN                                   | LMF                 | P³           | Control | 100               |
| 16       | F32    | F 2                 | 2                | 3A           | IMN/IN                                   | –                   | E. faecalis  | Control | 62                |
| 17       | M33    | T 3B                | 10               | 3A           | IMN/A/IMN                                | LMF                 | P. acnes     | Control | 89                |
| 18       | M39    | T 3B                | 3                | 3A           | IMN/A/IMN                                | LMF                 | Control     | 83                |
| 19       | M53    | F                   | 1                | 1A           | IMN/IN                                   | –                   | MSSA         | Control | 52                |
| 20       | M72    | T                   | 1                | 1B           | IMN/IN                                   | –                   | C-N          | Control | 67                |
| 21       | M31    | F                   | 2                | 3A           | IMN/IN                                   | –                   | CoNS         | Control | 46                |
| 22       | M41    | F 3B                | 4                | 3A           | IMN*/A/IN                                | LMF                 | MRSA         | Reop.   | 56                |
| 23       | M45    | T 3A                | 3                | 3A           | IMN/ILN                                  | –                   | MSSA         | Reop.   | 56                |
| 24       | M29    | F 3B                | 3                | 1A           | IMN/ILN                                  | –                   | MRSA         | Reop.   | 56                |
| 25       | M30    | F 2                 | 6                | 1A           | IMN/EN                                   | VAC SG              | MRSA         | Reop.   | 130               |
| 26       | M24    | T 3B                | 7                | 3A           | IMN/EN                                   | LMF                 | Poly¹        | Reop.   | 79                |
| 27       | M36    | T 3B                | 8                | 1A           | EF/IN                                    | C-LEG               | P. aeruginosa | Reop.   | 120               |
| 28       | M35    | F                   | 7                | 3A           | IMN/IN                                   | –                   | MRSA         | Reop.   | 55                |
| 29       | M31    | T 1                 | 1                | 3A           | Plate/IN                                 | –                   | MRSA         | Reop.   | 45                |
| 30       | F22    | F 2                 | 4                | 3A           | IMN/IN                                   | LMF                 | P⁰          | Reop.   | 42                |

Gender: M: male; F: female. Injury: F: femur; T: tibia. Previous surgery: n: number. IMN: intramedullary nail; IMN A: antibiotic coated nail; EF: external fixation; EN: Ender nail; ILN: interlocking nail; IN: industrial nail; LMF: local muscle flap; VAC: vacuum-assisted closure; SG: skin graft; C-LEG: cross-leg; C-N: culture-negative. MRSA: methicillin-resistant Staphylococcus aureus; MSSA: methicillin-sensitive Staphylococcus aureus; P. aeruginosa: Pseudomonas aeruginosa; CoNS: Coagulase negative Staphylococcus; E. Faecalis: Enterococcus faecalis; P. mirabilis: Proteus mirabilis; P. acnes: Propionibacterium acnes; P¹: MSSA + Enterococcus cloacae; P²: MRSA + Proteus mirabilis + Klebsiella blee; P³: P. aeruginosa + CoNS + MSSA; P⁴: CoNS + Proteus mirabilis; P⁵: P. aeruginosa + Enterococcus cloacae; P⁶: MRSA + Klebsiella blee: Reop.: reoperation.
Figure 2. Osteomyelitis of the tibia treated with a custom-made ATB-cemented nail at another center. (a–c) AP and lateral radiographs showing intracanal cement debonding and clinical image with active fistula in the leg. (d) Higher zoom image of the lateral radiograph, to highlight the cement remnants inside the tibia. (e, f) Intraoperative images, showing the locations where the bone windows were made (at the level of the fistula and distally for the removal of the intracanal cement) and the cement remnants removed. (g, h) AP and lateral radiographs at 30 months after reaming and placement of an ATB-cemented nail with infection control. (i) Good evolution of the treatment of the soft tissue coverage defect.

Figure 3. (a, b) AP and lateral radiographs of a patient with a history of open femur fracture treated with an endomedullary nail, 15 years of evolution. The patient had an active fistula at the level of the middle third of the external aspect of the thigh. Note the intracanal pathological images (white arrows). (c, d) AP and lateral radiographs 2 months postoperatively, after treatment with the commercial nail. (e, f) Control radiographs 48 months postoperatively with infection control.

3.6 Commercial nails vs. custom nails

Sixteen custom-made nails were used. In 10 of these cases, the metal core was an Ender nail, while the other 6 were treated with interlocking nails. The remaining 14 received commercial antibiotic-coated nails (Fig. 3).

In this series, the comparative analysis between custom-made and commercial nails showed no significant differences in preoperative variables (Table 2).

We found no significant differences between nail types in terms of infection control \( (p = 0.99) \), reoperation rates \( (p = 0.99) \), or failure rates \( (p = 0.99) \).

4 Discussion

The main study finding is that infection control in CPTO was achieved in 96.66 % of the patients using antibiotic cement-coated nails.

The use of antibiotic-loaded cement or PMMA has proven effective to administer local antimicrobials, fill voids, and achieve > 90 % healing rates in infected joint replacements (Hanseen, 2005). Antibiotic-coated nails present an alternative to extend this concept and represent a successful therapeutic option widely supported by the literature (Beals and Bryant, 2005; Qiang et al., 2007; Thonse and Conway, 2007).

Several manufacturing techniques have been described for these devices, like using pins, rods, and flexible or locking nails (Patzakis and Zalavras, 2005; Gosselin et al., 2004; Pailey and Herzenberg, 2002). These techniques offer the advantage of delivering high doses of antibiotics locally, with few or no systemic adverse effects (Zalavras et al., 2004). The peak of antibiotic elution is reached 24 h after placement and remains stable until day 10. This elution is maintained at high doses until week 10 and may persist in lower concentrations for up to 36 weeks (Nelson et al., 1994). In addition, interlocking nailing also provides mechanical stability, if needed (Thonse and Conway, 2007).

Conway et al. (2014) published a series of 43 patients with infected long-bone non-unions (Cierny–Mader type IV) treated with custom-made antibiotic-coated nails. They reported 100 % consolidation and infection control with a re-
### Table 2. Comparative analysis between patients treated with custom-made and commercial nails.

|                                | Overall (n 30) | Custom-made (n 16) | Commercial nails (n 14) | p < 0.05 |
|--------------------------------|----------------|--------------------|------------------------|----------|
| **Gender (male) n (%)**        |                |                    |                        |          |
|                               | 27 (90)        | 16 (100)           | 11 (78.57)             | 0.09     |
| **Age median (IQR)**           | 33 (24–43)     | 32.5 (24–45)       | 34 (30.5–42.5)         | 0.7      |
| **Risk factors n (%)**         |                |                    |                        |          |
| Diabetes                       |                |                    |                        |          |
|                               | 2 (6.66)       | 1 (6.25)           | 1 (7.14)               | 0.9      |
| Smoking                        |                |                    |                        |          |
|                               | 4 (13.33)      | 3 (18.75)          | 1 (7.14)               | 0.6      |
| CRD                             |                |                    |                        |          |
|                               | 1              | 1                  | –                      | 0.9      |
| ASA I–II                       |                |                    |                        |          |
|                               | 28 (93.33)     | 15 (93.75)         | 13 (92.85)             | 0.9      |
| ASA III–IV                     |                |                    |                        |          |
|                               | 2 (6.66)       | 1 (6.25)           | 1 (7.14)               |          |
| **Bone – open fracture n**     |                |                    |                        |          |
| Femur                          | 15–9           | 9–6                | 6–3                    | 0.3      |
| Tibia                          | 15–11          | 7–5                | 9–6                    | 0.3      |
| **Number of previous surgery median (range)** | 3 (1–10) | 2.5 (2–9) | 3 (1–10) | 0.7 |
| **Previous osteosynthesis type n (%)** | 30     | 16               | 14                     | –        |
| IMN                            | 25 (83.33)     | 14–2* (87.5)       | 11–2* (78.57)          |          |
| EF                             | 3 (10)         | 1 (6.25)           | 2 (14.28)              | 0.7      |
| Plate                          | 2 (6.66)       | 1 (6.25)           | 1 (7.14)               | 0.6      |
| **Months from injury to ATB nail median (IQR)** | 10 (6.5–15) | 9.5 (6.5–12) | 12 (6–20.7) | 0.18 |
| **Cierny–Mader n (%)**         |                |                    |                        |          |
| 1A                             | 8 (26.66)      | 5 (31.25)          | 3 (21.43)              | 0.4      |
| 3A                             | 18 (60.00)     | 9 (56.25)          | 9 (64.28)              |          |
| 1B                             | 3 (10.00)      | 1 (6.25)           | 2 (14.28)              | 0.6      |
| 3B                             | 1 (3.33)       | 1 (6.25)           | –                      | 0.9      |
| **ATB nail (metal core) n (%)** |                |                    |                        |          |
| Ender                          | 10 (33.33)     | 10 (62.50)         | –                      |          |
| Locking nail                   | 20 (66.66)     | 6 (37.50)          | 14 (100)               |          |
| **ATB in PMMA n (%)**          |                |                    |                        |          |
| Vancomycin + gentamicin        | 16 (53.33)     | 16 (100)           |                        |          |
| Gentamicin                     | 14 (46.66)     | 14 (100)           |                        |          |
| **Soft tissue coverage**       | 13 (43.33)     | 6 (37.50)          | 7 (50.00)              | 0.7      |
| **Surgical time median (IQR)** | 123            | 130.5              | 100.5                  | < 0.001  |
|                                | (110.3–131.8)  | (122.8–138)        | (94.2–121)             |          |
| **Intraoperative culture n (%)** |               |                    |                        |          |
| MRSA                           | 7 (23.33)      | 5 (31.25)          | 2 (14.28)              | 0.4      |
| MSSA                            | 7 (23.33)      | 5 (31.25)          | 2 (14.28)              | 0.4      |
| Polymicrobial                   | 6 (20.00)      | 2 (12.50)          | 4 (28.57)              | 0.3      |
| Culture-negative               | 2 (6.66)       | 1 (6.25)           | 1 (7.15)               | 0.9      |
| Others                          | 8 (26.66)      | 3 (18.75)          | 5 (37.71)              |          |
| **Infection control n (%)**    | 29 (96.60)     | 15 (93.7)          | 14 (100)               | 0.9      |
| **Reoperation n (%)**          | 8 (26.66)      | 4 (25.00)          | 4 (28.57)              | 0.9      |
| **Failure n (%)**              | 1 (3.33)       | 1 (6.25)           | –                      | 0.9      |
| **Follow-up months, median (IQR)** | 65       | 60.5              | 64.5                   | 0.8      |
|                                | (45–96)        | (42.7–105.5)       | (44.2–91.7)            |          |

CRD: chronic renal disease. *: ATB nail. IMN: intramedullary nail. EF: external fixation. Gent: gentamicin. Van: vancomycin. MRSA: methicillin-resistant Staphylococcus aureus. MSSA: methicillin-sensitive Staphylococcus aureus. P1: MSSA + Enterococcus cloacae. P2: MRSA + Proteus mirabilis + Klebsiella blee. P3: P. aeruginosa + CoNS + MSSA. P4: CoNS + Proteus mirabilis. P5: P. aeruginosa + Enterococcus cloacae. P6: MRSA + Klebsiella blee. The p value corresponds to the comparative analysis between custom-made and industrial nails.
Infections are associated with a higher risk of recurrence than S. aureus. On the other hand, Jorge et al. (2018) reported that polymicrobial PTO is associated with worse outcomes. In our series, 33.33% (2 out of 6) polymicrobial infections needed reoperation.

The population analyzed in this series and treated either with custom-made or commercial antibiotic-coated nails obtained similar results, and we observed no significant differences in their preoperative characteristics. In our analysis, devices manufactured in the OR were associated with longer operating times and some difficulties at the time of nail removal due to nail–cement debonding. While other authors have reported a 20% incidence of this complication (Wasko and Borens, 2013; Thonse and Conway, 2007; Shyam et al., 2009), it was more than 3 times higher in our series, as fragment extraction added to the complexity of the procedure and the need for an extra bone window. On the other hand, commercially available nails were easier to remove and did not show nail–cement debonding, possibly due to the greater strength of the cement used in their manufacturing process. Finally, we found no significant differences between the two types of nails in terms of infection control and need for reoperation.

The limitations of this study are those inherent to retrospective studies with a small sample. This might account for the lack of statistical significance of some of the variables analyzed producing a type 2 error. This number was limited as we excluded Cierny–Mader type IV osteomyelitis cases to obtain a more homogeneous population – especially with regard to procedures related to SBD reconstruction. Another limitation is the lack of a control group or functional analysis. On the other hand, the strengths of this study include the detailed characterization of patients with a complex and difficult to treat condition such as CPTO using antibiotic-coated nails. The literature does not include many comparative studies on the results of patients treated with commercial nails versus those treated with custom-made nails. Finally, our study presents an acceptable follow-up (minimum of 24 months) considering most recurrences occur during the first postoperative year.

5 Conclusion

Along with the classic treatment pillars for chronic osteomyelitis, the use of antibiotic cement-coated nails proved useful in the challenging treatment of post-traumatic chronic osteomyelitis. Our comparative analysis of custom-made and commercial antibiotic-coated nails did not show significant differences in terms of infection control and reoperation rates for patients without non-union or segmental bone defects. However, the use of custom-made nails made in the operating room was associated with longer operating times and nail–cement debonding at the time of extraction. In order to determine the external validity of this study, it is necessary to conduct properly designed studies in a larger population.
ethical statement. This retrospective chart review study, involving human participants, was in accordance with the ethical standards of the institutional and national research committee and with the 1964 Helsinki Declaration and its later amendments. The Bioethics Committee of British Hospital of Buenos Aires approved this study (protocol number 853). Personal information was removed from any published material, in agreement with the guidelines of the European General Data Protection Regulation, so as to protect the privacy of patients.

Data availability. All data generated and analyzed during this study are included in this published article and are available from the corresponding author on reasonable request.

Author contributions. All authors contributed to the study conception and design. Material preparation, data collection, and analysis were performed by GG, LPA, JAR, HdS, and CAP. The first draft of the manuscript was written by GG, and all authors commented on previous versions of the manuscript. All authors read and approved the final paper.

Competing interests. The contact author has declared that neither they nor their co-authors have any competing interests.

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References

Barger, J., Fragomen, A. T., and Rozbruch, S. R.: Antibiotic-coated interlocking intramedullary nail for the treatment of long-bone osteomyelitis, J. Bone Joint Surg. Rev., 5, 1–10, https://doi.org/10.2106/JBJS.RVV.16.00095, 2017.

Beals, R. K. and Bryant, R. E.: The treatment of chronic open osteomyelitis of the tibia in adults, Clin Orthop Relat Res., 433, 212–217, https://doi.org/10.1097/01.blo.0000150462.41498.f, 2005.

Bhadra, A. K. and Roberts, C. S.: Indications for antibiotic cement nails, J. Orthop. Trauma, 23, S26–30, https://doi.org/10.1097/BOT.0b013e31819f27a, 2009.

Blanchette, K. A. and Wenke, J. C.: Current therapies in treatment and prevention of fracture wound biofilms: why a multifaceted approach is essential for resolving persistent infections, J. Bone Joint Infect., 3, 50–67, https://doi.org/10.7150/jbji.23423, 2018.

Centers for Disease Control and Prevention: CDC/NHSN Surveillance Definitions for Specific Types of Infections, available at: https://www.cdc.gov/nhsn/pdfs/pscmanual/17pscnosinfdef_current.pdf, last access: 9 December 2016.

Cierny 3rd, G., Mader, J. T., and Penninck, J. J.: A clinical staging system for adult osteomyelitis, Clin. Orthop. Relat. Res., 414, 7–14, https://doi.org/10.1097/01.blo.000088564.81746.62, 2003.

Chadayaammuri, V., Herbert, B., Hao, J., Mavrogenis, A., Quispe, J. C., Kim, J. W., Young, H., Hake, M., and Mauffrey, C.: Diagnostic accuracy of various modalities relative to open bone biopsy for detection of long bone posttraumatic osteomyelitis, Eur. J. Orthop. Surg. Traumatol., 27, 871–875, https://doi.org/10.1007/s00590-017-1976-y, 2017.

Conway, J., Mansour, J., Kotze, K., Specht, S., and Shabbati, L.: Antibiotic cement-coated rods: an effective treatment for infected long bones and prosthetic joint non-unions, Bone Joint J., 96-B, 1349–1354, https://doi.org/10.1302/0301-620X.96B10.33799, 2014.

Finelli, C. A., Dos Reis, F. B., Fernandes, H. A., Dell’Aquila, A., Carvalho, R., Miki, N., Francioci, Z., Abdalla, R., and Salles, M. J. C.: Intramedullary reaming modality for management of postoperative long bone infection: a prospective randomized controlled trial in 44 patients, Patient Saf. Surg., 13, 39, https://doi.org/10.1186/s13037-019-0215-3, 2019.

Garcia Del Pozo, E., Collazos, J., Carton, J. A., Camporro, D., and Asensi, V.: Factors predictive of relapse in adult bacterial osteomyelitis of long bones, BMC Infectious Diseases, 18, 635, https://doi.org/10.1186/s12879-018-3550-6, 2018.

Gosselin, R. A., Roberts, I., and Gillespie, W. J.: Antibiotics for preventing infection in open limb fractures, Cochrane Database Syst. Rev., 1, CD003764, https://doi.org/10.1002/14651858.CD003764.pub2, 2004.

Hake, M. E., Oh, J. K., Kim, J. W., Ziran, B., Smith, W., Hak, D., and Muffrey, C.: Difficulties and challenges to diagnoses and treat post-traumatic long bone osteomyelitis, Eur. J. Orthop. Surg. Traumatol., 2015, 1–3, https://doi.org/10.1007/s00590-014-1576-z, 2015.

Hanseen, A. D.: Local Antibiotic delivery vehicles in the treatment of musculoskeletal infection, Clin. Orthop. Relat. Res., 437, 91–96, https://doi.org/10.1097/01.blo.0000175713.30506.77, 2005.

Hotchen, A. J., McNally, M. A., and Sendi, P.: The Classification of Long Bone Osteomyelitis: A Systemic Review of the Literature, J. Bone Joint Infect., 2, 167–174, https://doi.org/10.7150/jbji.21050, 2017.

Jorge, L. S., Fucuta, P. S., L., M. G., Nakazone, M. A., de, J. A., Chueire, A. G., and Costa, M. J.: Outcomes and Risk Factors for Polymicrobial Posttraumatic Osteomyelitis, J. Bone Joint Infect., 3, 20–26, https://doi.org/10.7150/jbji.22566, 2017.

Jorge-Mora, A., Amhaz-Escanlar, S., Fernandez-Pose, S., Garcia-Iglesias, A., Mandia-Mancebo, F., Franco-Trepat, E., Guillan-Fresco, M., and Pino-Miguez, J.: Commercially available antibiotic-laden PMMA-covered locking nails for the treatment of fracture-related infections - A retrospective case analysis of 10 cases, J. Bone Joint Infect., 4, 155–162, https://doi.org/10.7150/jbji.34072, 2019.

Kanakaris, N., Gudipati, S., Tosounis, T., Harwood, P., Britten, S., and Giannoudis, P. V.: The treatment of intramedullary osteomyelitis of the femur and tibia using the Reamer-Irrigator-Aspirator system and antibiotic cement rods, Bone Joint J., 96-B, 783–788, https://doi.org/10.1302/0301-620X.96B6.32244, 2014.

Kim, J. W., Cuellar, D. O., Hao, J., Seligson, D., and Mauffrey, C.: Custom-made antibiotic cement nails: A comparative
G. Garabano et al.: Effectiveness of antibiotic cement-coated nails

study of different fabrication techniques, Injury, 45, 1179–1184, https://doi.org/10.1016/j.injury.2014.03.006, 2014.
Klemm, K.: Gentamicin-PMMA-beads in treating bone and soft tissue infections, Zentralbl Chir., 104, 934–942, 1979.
Klemm, K., Henry, S. L., and Seligson, D.: The treatment of infection after interlocking nailing, Technicol Orthop., 3, 54–61, 1998.
Koury, K. L., Hwang, J. S., and Sirkin, M.: The antibiotic nail in the treatment of long bone infection: Technique and results, Orthop Clin North Am., 48, 155–165, https://doi.org/10.1016/jocl.2016.12.006, 2017.
Makhdom, A. M., Buksbaum, J., Rozbruch, S. R., Cunha, R. D., and Fragomen, A. T.: Antibiotic Cement-Coated interlocking Intramedullary Nails in the Treatment of Septic Complex Lower Extremity Reconstruction; A Retrospective Analysis with Two year Minimum Follow up, J. Bone Joint Infect., 5, 176–183, https://doi.org/10.7150/jbji.46570, 2020.
Morgenstern, M., Athanasou, N. A., Ferguson, J. Y., Metsemakers, W. J., Atkins, B. L., and McNally, M. A.: The value of quantitative histology in the diagnosis of fracture-related infection, Bone Joint J., 100-B, 966–972, https://doi.org/10.1302/0301-620X.100B7.BJJ-2018-0052.R1, 2018.
Nelson, C. L., Hickmon, S. G., and Harrison, B. H.: Elution characteristics of gentamicin-PMMA beads after implantation in humans, Orthopedics, 17, 415–416, 1994.
Paley, D. and Herzenberg, J. E.: Intramedullary infections treated with antibiotic cement rods: preliminary results in nine cases, J. Orthop. Trauma, 16, 723–729, https://doi.org/10.1097/00005131-200211000-00007, 2002.
Patzkis, M. J. and Zalavras, C. G.: Chronic posttraumatic osteomyelitis and infected non-union of the tibia: current management concepts, J. Am. Acad. Orthop. Surg., 13, 417–427, https://doi.org/10.5435/00124635-200510000-00006, 2005.
Patzkis, M. J., Wilkins, J., and Wiss, D. A.: Infection following intramedullary nailing of long bones. Diagnosis and management, Clin. Orthop. Relat. Res., 212, 182–191, 1986.
Qiang, Z., Jun, P. Z., Jie, X. J., Hang, L., Bing, L. J., and Cai, L. F.: Use of antibiotic cement rod to treat intramedullary infection after nailing: preliminary study in 19 patients, Arch. Orthop. Trauma Surg., 127, 945–951, https://doi.org/10.1007/s00402-007-0315-x, 2007.
Shyam, A. K., Sancheti, P. K., Patel, S. K., Rocha, S., Pradhan, C., and Patil, A.: Use of antibiotic cemented impregnated intramedullary nail in treatment of infected non-union of long bones, Indian J. Orthop., 43, 396–402, https://doi.org/10.4103/0019-5413.55468, 2009.
Simpson, A. H. R. W., Deakin, M., and Latham, J. M.: Chronic osteomyelitis. The effect of the extent of surgical resection on infection-free survival, J. Bone Joint Surg. Br., 83, 403–407, https://doi.org/10.1302/0301-620x.83b3.10727, 2001.
Tetsworth, K. and Cierny 3rd, G.: Osteomyelitis debridement techniques, Clin. Orthop. Relat. Res., 360, 87–96, https://doi.org/10.1097/00003086-199903000-00011, 1999.
Thonse, R. and Conway, J.: Antibiotic Cemented-Coated Interlocking Nail for the treatment of infected non-unions and segmental bone defects, J. Orthop. Trauma, 21, 258–268, https://doi.org/10.1097/BOT.0b013e31803ea9e6, 2007.
Tice, A. D., Hoaglund, P. A., and Shoultz, D. A.: Risk factors and treatment outcomes in osteomyelitis, J. Antimicrob. Chemother., 51, 261–1268, https://doi.org/10.1093/jac/dkg186, 2003.
Wasko, M. K. and Borens, O.: Antibiotic cement nail for the treatment of posttraumatic intramedullary infections of the tibia: mild-term results in 10 cases, Injury, 44, 1057–1060, https://doi.org/10.1016/j.injury.2013.05.001, 2013.
Wasko, M. K. and Kaminski, R.: Custom-made antibiotic cement nails in orthopaedic trauma: review of outcomes, new approaches and perspectives, BioMed Research Int., 2015, 387186, https://doi.org/10.1155/2015/387186, 2015.
Zalavras, C. G., Patzkis, M. J., and Holtom, P.: Local Antibiotic therapy in the treatment of open fractures and osteomyelitis, Clin. Orthop. Relat. Res., 427, 86–93, https://doi.org/10.1097/01.blo.0000143571.18892.8d, 2004.