Overexpression of retinoblastoma-binding protein 4 contributes to the radiosensitivity of AGS gastric cancer cells via phosphoinositide3-kinase/protein kinase B pathway suppression

XIAOXI JIN1, RUI JIANG2, YONGSHENG XIANG2, ZHEN FAN2, ZHIWEI WU2, BO YANG2, LUJUN YANG2, SHANSHAN WEI2 and YAN YANG1

1Laboratory Department; 2Department of Hematology, Jingmen No. 1 People's Hospital, Jingmen, Hubei 448000, P.R. China

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Abstract. In the present study, the effects and underlying mechanism of RbAp48 on the radiosensitivity of AGS gastric cancer cells was investigated. Cell proliferation was determined with an MTT assay. Flow cytometry was performed to evaluate the cell cycle and apoptosis. Reverse transcription-quantitative polymerase chain reaction and western blot analysis were performed to detect mRNA and protein expression, respectively, including RbAp48, phosphoinositide 3-kinase (PI3K) and protein kinase B (Akt). The results revealed that radiation enhanced the expression level of RbAp48 in AGS cells, and that RbAp48 combined with radiation reduced AGS cell proliferation. In addition, RbAp48 combined with radiation resulted in G1 phase arrest and induced apoptosis via regulation of the PI3K/Akt pathway. In conclusion, it was demonstrated that overexpression of RbAp48 enhanced the radiosensitivity of AGS gastric cancer cells via suppression of PI3K/Akt pathway activity, suggesting that RbAp48 may hold potential as a gene therapeutic strategy in the future, aiding in the treatment of gastric cancer.

Introduction

Gastric cancer is one of the most common malignant tumors worldwide. A number of factors contribute to the development of gastric cancer, of which the pathogenesis is highly complex (1,2). *Helicobacter pylori* infection is listed as a class 1 carcinogenic factor for gastric cancer by the World Health Organization, and high salt and high nitrate diets may also be risk factors for gastric cancer development (3). Genetic factors, environmental factors and bacterial infections ultimately affect the occurrence and progression of gastric cancer (4,5). It has been reported that although gastric cancer treatment and prognosis has greatly improved in China, the incidence of gastric cancer remains high (6). As there is a lack of knowledge of specific symptoms, the diagnosis of gastric cancer at an early stage is difficult. Gastrectomy is a widely used strategy in gastric cancer therapy. However, the prognosis of patients with gastric cancer at advanced stages is unsatisfactory (7). Therefore, a better understanding of the occurrence and progression of gastric cancer is of scientific significance.

The primary target molecule of radiotherapy is DNA. The mechanism of cell DNA damage repair is initiated by radiation exposure, which activates cell cycle arrest, thereby promoting repair of injury (8). If DNA fails to repair, it may result in cell death, necrosis or senescence (8). DNA strand breaks (DSBs) induced by radiation exposure are closely associated with cell death. DSB repair is associated with radiosensitivity (9). The effectiveness of therapy of gastric cancer primarily depends on the sensitivity of the tumor to radiotherapy (10-12). Radiation resistance has become key to further deterioration of tumors, thus the study of radiosensitization has become more prevalent.

Gene therapy is being increasingly recognized in tumor therapy. Tumor radiosensitivity is associated with its internal molecular biological mechanism. It has been demonstrated that the abnormal expression of a number of oncogenes and tumor suppressor genes may affect tumor cell apoptosis, radiosensitivity and patient prognosis (13). The potential combination of tumor gene therapy and radiotherapy has therefore been suggested, to ultimately reduce normal tissue damage and enhance the effects of radiotherapy (14). Numerous tumor gene therapies have been investigated in vitro experiments and have exhibited beneficial effects, such as cellular tumor antigen p53 (P53), which has successful results in clinical trials, achieving desirable treatment outcomes (15-17). Retinoblastoma-binding protein 4 (RbAp48) is a member of the WD-40 protein family and was originally identified as a retinoblastoma protein (Rb) binding protein (18). E2F transcription factor (E2F) 1 and RbAp48 interaction is mediated by Rb and histone deacetylase (HDAC) and results in the inhibition of E2F regulatory gene transcription, which are important cell cycle regulatory proteins (19).
The underlying mechanisms of gastric cancer radiosensitivity remain unclear. The present study aimed to investigate the effect and underlying mechanisms of RbAp48 on gastric cancer cell radiosensitivity.

Materials and methods

Cell culture. The human gastric cancer cell line (AGS) was purchased from Shanghai Gefan Biotechnology Co., Ltd. (Shanghai, China). The cells were maintained in RPMI-1640 medium (Fisher Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 10% fetal bovine serum (Gibco; Thermo Fisher Scientific, Inc.) in a 37˚C incubator with 5% CO₂.

Cell transfection and grouping. pcDNA3.1, pcDNA3.1-RbAp48, RbAp48 siRNA and non-specific scrambled siRNA vectors were obtained from Invitrogen (Thermo Fisher Scientific, Inc.). The vectors were transfected at a final concentration of 100 nmol/l transfection (20). AGS cells were transfected with pcDNA3.1 (mock), pcDNA3.1-RbAp48 (RbAp48), RbAp48 siRNA (si-RbAp48; 5′-CAGGGGATACA CGGGAGTAGT-3′) and non-specific scrambled siRNA (NC; 5′-ACGUGACACGUUCGGAGAATTT-3′) vectors using EndoFectin™ Max transfection reagent (GeneCopoeia, Inc., Rockville, MD, USA) at 37˚C for 48 h. Following transfection, cells were lysed for western blot analysis and RT-qPCR to verify transfection efficiency.

There were five AGS cell treatment groups: Control (treated with PBS), mock (treated with pcDNA3.1), control+RAD (treated with 6 Gy radiation), mock+RAD (treated with pcDNA3.1 and radiation) and the RbAp48+RAD group (treated with pcDNA3.1-RbAp48 and radiation), in the early stage of the experiment. There were seven treatment groups in the advanced stage of the experiment, including: pcDNA3.1 (Mock), pcDNA3.1 and 6 Gy radiation (RAD), si-RbAp48, si-RbAp48+RAD, pcDNA3.1-RbAp48 (RbAp48), pcDNA3.1-RbAp48 and radiation (RbAp48+RAD), as well as pcDNA3.1-RbAp48, 6 Gy radiation and 50 ng/ml insulin-like growth factor-1 (IGF-1) (RbAp48+RAD+IGF-1). IGF-1 was used as an agonist of the phosphoinositide 3-kinase (PI3K)/protein kinase B (Akt) pathway, and was used to treat cells at 37˚C for 48 h.

Cell radiation. Cells were digested with 0.25% trypsin and counted with a hemocytometer prior to radiation exposure. Cells were subsequently transferred into new culture bottles, each containing 10⁶ cells. Cells were maintained in the incubator overnight at 37˚C with 5% CO₂. Following this, cells were treated with 0, 2, 4, and 6 Gy 6 MV-X ray at room temperature for 3 h using a PRIMUS™ linear accelerator (Siemens AG, Munich, Germany). This was a preliminary experiment to determine the appropriate dose of radiation for the subsequent experiments.

Cell proliferation analysis. Cell proliferation was determined by an MTT assay. Cells (6x10⁵/ml) in the logarithmic phase were sowed into the wells of 96-well plates and incubated for 12 h at 37˚C with 5% CO₂. Along with a control group, cells were treated with PBS, 6 Gy doses of radiation, and 100 nmol/l pcDNA3.1-RbAp48 and pcDNA3.1 vectors. The cells were subsequently maintained at 37˚C with 5% CO₂ for 12, 24 and 48 h. A volume of 50 µl MTT solution was added into each well, and cells were transferred to the incubator for 4 h. Subsequently, 100 µl dimethyl sulfoxide was added into each well. The absorbance was read at wavelengths of 570 and 630 nm using a microplate reader (cat. no. SMR16.1; Uscn Life Sciences, Inc., Wuhan, China). Cell proliferation was determined in terms of the percentage of cell survival.

Flow cytometry (FCM). FCM was used for the analysis of the cell apoptosis and cell cycle. For cell apoptosis analysis, cells were harvested following transfection for 48 h and fixed in 70% ethanol at room temperature overnight. Cells prepared for assessment were first washed with PBS and subsequently resuspended in Annexin V-fluorescein isothiocyanate and propidium iodide (PI; Shanghai Yeasen Biotechnology Co., Ltd., Shanghai, China) at 37˚C for 30 min. A flow cytometer (FACS Calibur; BD Biosciences, Franklin Lakes, NJ, USA) was used to assess cell apoptosis.

Cell cycle analysis. Following digestion with 0.25% trypsin, ~5x10⁶ cells were plated into 6-well plates and incubated for 24 h at 37˚C with 5% CO₂. Following treatment with radiation and mock or RbAp48 vector, the medium was removed and cells were washed with PBS three times. Cells were digested with 0.25% trypsin, placed in 15 ml centrifuge tubes and centrifuged for 5 min at 1,000 x g at 4˚C and supernatant was subsequently discarded. Cells were resuspended following washing with PBS, and subsequently centrifuged for 5 min at 1,000 x g at 4˚C, and the supernatant was discarded. Pre-cooled 70% ethanol (4˚C; 1 ml) was added into cells and the cells were gently blown with a pipette. Following this, cells were stored in a refrigerator at 4˚C overnight. Cells were centrifuged for 5 min at 1,000 x g at 4˚C, ethanol was discarded and cells were washed with PBS three times. PBS (500 µl) containing PI (50 µg/ml), RNase A (100 µg/ml) and Triton X-100 (0.2%) was added to the cells, which were incubated in the dark for 30 min at 4˚C. The results were detected by FCM and cell cycle analysis was performed with FlowJo 10 software (FlowJo LLC, Ashland, OR, USA).

Reverse transcription-quantitative polymerase chain reaction (RT-qPCR) analysis. TRIzol® reagent (Tiangen Biotech Co., Ltd., Beijing, China) was used to extract total RNA from cells. According to the manufacturer’s protocols, 2 µg of RNA was used for cDNA synthesis using the First Strand cDNA Synthesis kit (Sigma-Aldrich; Merck KGaA, Darmstadt, Germany). RT-qPCR was performed using the SYBR Green Premix reagent (TakaraBio, Inc., Otsu, Japan) in an ABI 7500 Thermocycler (Thermo Fisher Scientific, Inc.). The PCR thermocycling conditions were as follows: Initial denaturation for 10 min at 95˚C, 40 cycles of 95˚C for 5 sec and 65˚C for 31 sec, followed by 95˚C for 15 sec, 60˚C for 1 min, 95˚C for 15 sec and a final extension at 72˚C for 10 min and finally held at 4˚C. β-actin was used as the internal control for normalization. The primers used for RT-qPCR were listed in Table 1.

Western blotting. Treated cells were lysed in radiimmunoprecipitation lysis and extraction buffer (Thermo Fisher Scientific, Inc.) on ice. Cells were broken into pieces with an
ultrasonic cell disruptor. Supernatant was collected following centrifugation at 1,000 x g at 4˚C for 10 min. According to the manufacturer's protocols, protein concentration was determined using a bicinchoninic acid assay reagent (Bio-Rad Laboratories, Inc. Hercules, CA, USA). An equal quantity of proteins (50 µg lane) was separated using 10% SDS-PAGE. Proteins obtained were transferred to nitrocellulose membranes for 1.5 h. Membranes were blocked in 5% low-fat dried milk at room temperature for 2 h. The following primary antibodies were incubated with membranes at 4˚C overnight: Anti-RbAp48 (1:1,000; cat. no. ab1765; Abcam, Cambridge, MA, USA), anti –cyclin B1 (1:1,000; cat. no. ab72; Abcam), anti-cell division control protein 2 homolog (Cdc2; 1:2,000; cat. no. ab12568; Abcam), anti-phosphorylated (p)-Cdc2 (1:1,000; cat. no. ab258965; Abcam), anti-p-M-phase inducer phosphatase 3 (Cdc25c; 1:500; cat. no. ab62191), anti-Cdc25c (1:1,000, cat. no. ab32444; Abcam), anti-cleaved caspase-3 (1:1,000; cat. no. ab2302; Abcam), anti-cleaved caspase-7 (1:500; cat. no. ab32042; Abcam), anti-cleaved caspase-9 (1:1,000; cat. no. ab1324; Abcam), anti-cleaved poly (ADP-ribose) polymerase (PARP; 1:5,000; cat. no. ab532064; Abcam) anti-p-Pi3K (1:1,000; cat. no. ab182651; Abcam), anti-Pi3K (1:1,000; cat. no. ab151549; Abcam) anti-p-Akt (1:5,000; cat. no. ab81283; Abcam), anti-Akt (1:5,000; cat. no. ab182729; Abcam) and anti-β-actin (1:1,000; cat. no. ab8227; Abcam) at 4˚C. Rabbit anti-mouse immunoglobulin G (IgG; cat. no. 58802; 1:7,000; CST Biological Reagents Co. Ltd., Shanghai, China), goat anti-mouse IgG (cat. no. ab7064; 1:8,000; Abcam) and mouse anti-goat IgG (cat. no. BA1074; 1:7,000; Invitrogen; Thermo Fisher Scientific, Inc.) horseradish peroxidase (HRP)-conjugated secondary antibodies were incubated with the membranes at room temperature for 2 h. Bands were visualized using enhanced chemiluminescent reagent (EMD Millipore, Billerica, MA, USA). Images were captured using the Fujifilm LAS-3000 Imager imaging system (Fuji Photo Film Co., Ltd., Tokyo, Japan) and the software used was the LAS-3000 Image Reader (Fuji Photo Film Co., Ltd.).

Statistical analysis. Results are presented as the mean ± standard error of mean using SPSS 20.0 (IBM Corp., Armonk, NY, USA). Data were analyzed by one-way analysis of variance, followed by Tukey's post-hoc test. Each experiment was repeated three times. P<0.05 was considered to indicate a statistically significant difference.

Results

Radiation enhances the expression level of RbAp48 in AGS cells. RT-qPCR and western blot analysis was performed to evaluate the expression levels of RbAp48 in AGS cells under various experimental conditions. AGS cells were treated with increasing intensities of radiation (2, 4 and 6 Gy). The results
revealed that the mRNA (Fig. 1A) and protein (Fig. 1B) expression of RbAp48 was significantly higher in cells exposed to radiation compared with the control group, in a dose-dependent manner (P<0.05). Additionally, following AGS cell transfection with pcDNA3.1-RbAp48 plasmids, the mRNA (Fig. 1C) and protein (Fig. 1D) expression levels of RbAp48 were significantly higher than control and mock groups (P<0.05). Furthermore, the mRNA (Fig. 1E) and protein (Fig. 1F) levels of RbAp48 in the si-RbAp48 transfected group were significantly lower, compared with the control and NC groups (P<0.05). These data demonstrated that radiation increased RbAp48 expression and that the transfection efficiency of RbAp48 was high in AGS cells.

**RbAp48 overexpression combined with radiation exposure causes G2 cell cycle arrest.** In order to identify the mechanism of AGS cell growth inhibition with combination of RbAp48 and radiation exposure, cell cycle analysis was performed. As presented in Fig. 2B and C, the percentage of cells in the G2 phase was markedly higher in the RbAp48+RAD group (48.98%), compared with the other groups (control, 10.47%; Mock, 9.29%; control+RAD, 22.85%; mock+RAD, 23.36%). This phenomenon indicated that overexpression of RbAp48 in combination with radiation exposure arrested cells in the G2 phase.

Furthermore, the expression levels of cell cycle associated factors were evaluated in each group by RT-qPCR and western blot analysis. It was demonstrated that radiation significantly reduced the mRNA expression of cyclin B1, and enhanced the expression of P53 and cyclin dependent kinase inhibitor (P21; Fig. 3A; P<0.05). When cells were exposed to a combination of radiation and RbAp48 overexpression, the downregulation of cyclin B1 expression and upregulation of P53 and P21 expression was more significant (P<0.01). As presented in Fig. 3B, western blot analysis demonstrated that compared with the other groups, RbAp48 overexpression combined with radiation significantly decreased the protein expression of cyclin B1 (P<0.001). Furthermore, in the RbAp48+RAD group, the protein expression of p-Cdc2 (Tyr15) in AGS cells was significantly increased (Fig. 3C), and p-Cdc25c expression...
was significantly reduced (Fig. 3D; P<0.01). Based on the aforementioned results, it was concluded that RbAp48 overexpression combined with radiation exposure resulted in G2 cell cycle arrest, through regulation of associated cell cycle factor expression levels.

**RbAp48 combined with radiation induces AGS cell apoptosis.** In order to determine the effect of RbAp48 and radiation on cell apoptosis, FCM analysis was performed in the present study. It was demonstrated that the percentage of apoptotic cells was significantly increased in the control+RAD and mock+RAD groups compared with the control (P<0.01), indicating that radiation induced AGS cell apoptosis. Additionally, the percentage of apoptosis in the RbAp48+RAD group was significantly higher than the other groups (Fig. 4). These data suggested that RbAp48 in combination with radiation further induced AGS cell apoptosis.

The expression levels of associated apoptosis proteins were presented in Fig. 5. With regards to the RT-qPCR data (Fig. 5A), the expression levels of caspase-3, caspase-7, caspase-9, PARP and Bax were significantly higher (P<0.01), while the expression level of Bcl-2 was significantly lower (P<0.05) in the RAD groups compared with the control. In addition, expression levels of caspase-3, caspase-7, caspase-9, PARP and Bax were further increased and Bcl-2 expression was further decreased in the RbAp48+RAD group, compared with the control+RAD and mock+RAD groups. Furthermore, the western blotting data of caspase-3, cleaved caspase-7,
cleaved caspase-9, and cleaved PARP expression displayed a similar trend (Fig. 5B). Therefore, it was demonstrated that RbAp48 combined with radiation induced AGS cell apoptosis through regulation of associated apoptosis marker expression.

**RbAp48 combined with radiation inhibits the PI3K/Akt pathway.** Western blotting was also performed to explore the functional mechanism of RbAp48 in combination with radiation in AGS cell growth inhibition. The expression levels of p-PI3K, PI3K, p-Akt, and Akt were detected. In the RAD groups, the protein expression levels of p-PI3K were significantly lower than the control and mock groups. However, the expression levels of total PI3K were not significantly different among the five groups (Fig. 6A). Additionally, compared with the control and mock groups, protein expression levels of p-Akt were markedly reduced by radiation, while the expression levels of total Akt were not significantly different among the five groups (Fig. 6B). Furthermore, overexpression of RbAp48 resulted in further reductions in p-PI3K and p-Akt expression, compared with the radiation only groups (P<0.01). Therefore, it was confirmed that RbAp48 combined with radiation affected the PI3K/Akt pathway activity, which may have been involved in AGS cell growth inhibition and induction of AGS cell apoptosis.

**si-RbAp48 represses cell apoptosis and RbAp48 in combination with radiation promotes cell apoptosis via PI3K/Akt pathway inhibition.** FCM analysis was performed in order to...
analyze the effect of si-RbAp48 on cell apoptosis, and to determine if the combination of RbAp48 and radiation induced apoptosis via PI3K/Akt pathway analysis. The results revealed that compared with the mock group, the apoptotic cell number in the si-RbAp48 group was reduced, and was significantly increased in the RbAp48 overexpression group. In addition, cells treated with a combination of RbAp48 and radiation had the highest proportion of apoptotic cells. Furthermore, compared with the RbAp48+RAD group, the number of apoptotic cells in the RbAp48+RAD+IGF-I group was significantly lower (P<0.05; Fig. 7). Therefore, it was demonstrated that si-RbAp48 repressed cell apoptosis, and RbAp48 in combination with radiation promoted cell apoptosis via PI3K/Akt pathway inhibition.

**Discussion**

RbAp48, as a binding protein of tumor suppressor protein Rb, is involved in various biological functions, including chromatin assembly, histone modification and nucleosome...
remodeling (21,22). Studies have demonstrated that the Rb/RbAp48-associated histone acetyl transferase complex participates in the inhibition of E2F regulatory gene transcription (23-27). It has been reported that RbAp48 promotes radiosensitivity in breast, melanoma and cervical cancer cells (28,29). However, the effects of RbAp48 in gastric cancer remain unclear.

AGS gastric cancer cells were used in the present study. Radiation treatment was administered and the mRNA and protein expression of RbAp48 had significantly increased in the AGS cells, suggesting that RbAp48 is a radiation-inducible protein in gastric cancer. This finding provided the basis for further investigation. Furthermore, compared with the cells treated with radiation only, cell proliferation was significantly inhibited in cells overexpressing RbAp48 in addition to radiation exposure. The mechanism underlying this phenomenon may be associated with the inhibition of oncogene expression and increased cancer suppressor gene expression (30).

In the present study, gastric cancer cell proliferation was significantly inhibited following treatment with RbAp48 in combination with radiation. However, the mechanism underlying the inhibition of AGS cell proliferation by this combination may be complex. The present preliminary study suggested that RbAp48 arrested the AGS cells at the G2 phase and induced apoptosis. These processes may serve a critical role in RbAp48-induced radiosensitivity.

Radiation-induced cell responses include cell cycle arrest, apoptosis and DNA repair, as a complex result of multiple gene involvement (31). In response to radiation-induced DNA impairment, DNA repair, G2 phase arrest and apoptosis predominantly occur (32,33). In the present study, it was demonstrated that the G2 phase arrest was increased by radiation, and this effect was further increased by RbAp48 overexpression. Typically, the G1 and G2 phases are more sensitive to radiation, whereas the S phase is the least sensitive. Thus, the G2 phase is considered a key point in cancer radiotherapy (31). Previous research has reported that RbAp48 may enhance the proportion of HS-578T cells in the G2 phase (28), which was in accordance with the results of the present study.

Radiation leads to apoptosis in cancer cells, indicating that apoptosis is a key mechanism in tumor cell radiotherapy. The detection and signaling system of DNA impairment controls cell cycle checkpoints and results in subsequent cell death. The present study revealed that overexpression of RbAp48 significantly increased the apoptotic rate in combination with radiotherapy. Additionally, RbAp48 silencing markedly attenuated apoptosis in radiotherapy-treated AGS cells. These results demonstrated that RbAp48 is a radiotherapy-induced protein in gastric cancer cells, which promoted the radiosensitivity of gastric cancer cells through increased cell apoptosis.

Cell cycle arrest, DNA repair and apoptosis initially occur in response to DNA impairment. Cyclin B1, Cdc2 and Cdc25c
are critical to the radiographic response and mediate the cell cycle when DNA is impaired (34-38). The P53 gene is a tumor suppressor gene (39), and the P21 gene is an important member of the cyclin dependent kinase inhibitor family (40). P53 and P21 coordinate the association between cell cycle, DNA replication and repair, and thus closely associate tumor inhibition with cell cycle control (39-42). The present results demonstrated that overexpression of RbAp48 in combination with radiation significantly reduced the expression of cyclin B1 and p-Cdc25c, and enhanced the expression of p-Cdc2, P53 and P21.

Caspases are key elements in the apoptotic pathway and participate in apoptosis signal transduction and execution. Research has indicated that caspase-3, -7 and -9 are involved in radiation-induced cell apoptosis (43,44). In the present study, overexpression of RbAp48 in combination with radiotherapy markedly enhanced AGS cell apoptosis. Furthermore, RT-qPCR and western blotting data indicated that overexpression of RbAp48 in combination with radiation significantly reduced the expression levels of p-PI3K and p-Akt. In addition, the percentage of apoptotic cells in the RbAp48+RAD+IGF-1 group was significantly lower than that of the RbAp48+RAD group. Therefore, it was confirmed that overexpression of RbAp48 in combination with radiation increased apoptosis via inhibition of PI3K/Akt pathway activity.

Radiotherapy is one of the major treatment options for malignant tumors. Though tumor cells cannot be completely removed by surgery, surgery in combination with radiotherapy increases the possibility of patient survival (52,53). However, certain tumor cells (glioma and pancreatic tumor) are not sensitive to radiation, which consequently results in treatment failure (54,55). Therefore, improving radiosensitivity is key to improving the clinical outcomes of radiotherapy. Current clinical radiotherapy sensitization drugs, including cisplatin, 5-fluorouracil and gemcitabin, are effective; however, these drugs may additionally result in damaging effects in normal tissues (56,57). Therefore, developing specific radiation-sensitive agents for tumors should be the direction of future radiotherapy research.

In conclusion, overexpression of RbAp48 in combination with radiation arrested AGS cells in G2 phase and induced apoptosis via regulation of the PI3K/Akt pathway (50,51). Therefore, it was hypothesized that AGS cell apoptosis, induced by RbAp48 overexpression in combination with radiation, was associated with PI3K/Akt pathway regulation. In the present study, the results of the RT-qPCR and western blot analysis demonstrated that overexpression of RbAp48 in combination with radiation significantly downregulated the expression levels of p-PI3K and p-Akt. In addition, the percentage of apoptotic cells in the RbAp48+RAD+IGF-1 group was significantly lower than that of the RbAp48+RAD group. Therefore, it was confirmed that overexpression of RbAp48 in combination with radiation increased apoptosis via inhibition of PI3K/Akt pathway activity.
RbAp48 may be a potential genetic therapeutic target in gastric cancer. However, the specific effects of RbAp48 overexpression in healthy tissues remain unknown and require further investigation.

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Availability of data and materials
All data generated and analyzed during this study are included in this published article.

Authors' contributions
XJ, RJ, YX, BY and YY made substantial contributions to the conception and design of the experimental scheme. ZF, ZW, LY and SW analyzed and interpreted data. XJ was a principal contributor in writing the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate
Not applicable.

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Not applicable.

Competing interests
The authors declared that they had no competing interests.

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