Research Article

Two Functional TP53 Genetic Variants and Predisposition to Keloid Scarring in Caucasians

Andrzej Dmytrzak,1 Agnieszka Boroń,2 Beata Łoniewska,3 Klaudyna Lewandowska,2,4 Iwona Gorący,5 Mariusz Kaczmarczyk,2 and Andrzej Ciechanowicz2

1Aesthetic Med Andrzej Dmytrzak Prywatne Centrum Chirurgii Plastycznej i Rekonstrukcyjnej, ul. Niedziałkowskiego 47, 71-403 Szczecin, Poland
2Department of Clinical and Molecular Biochemistry, Pomeranian Medical University, al. Powstancow Wlkp. 72, 70-111 Szczecin, Poland
3Department of Neonatal Diseases, Pomeranian Medical University, ul. Powstancow Wlkp. 72, 70-111 Szczecin, Poland

Correspondence should be addressed to Andrzej Ciechanowicz; aciech@pum.edu.pl

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Introduction. Keloid is defined as a benign proliferative scar that grows beyond the confines of the original insult to the skin, invading into adjacent normal tissue. The pathogenesis of keloid is complex, and many evidences suggest the influence of genetic factors, among them, the polymorphisms of the TP53 gene encoding tumor protein p53. Objective. To investigate the association of rs1042522 (c.215G>C, p.Arg72Pro) and rs17878362 (16-bp insertion/duplication in intron 3) variants, two most frequently analyzed TP53 functional polymorphisms, and the risk of keloid in Polish patients. Materials and Methods. The rs1042522 and rs17878362 polymorphisms were identified by sequencing genomic DNA extracted from peripheral blood leukocytes of 86 keloid patients and from cordial blood leukocytes of 100 newborn infants consisting control group. Results. The rs1042522 and rs17878362 TP53 genotype distributions both in keloid patients and the control group conformed to the expected Hardy–Weinberg equilibrium. No significant differences in the distribution of rs1042522 and rs17878362 TP53 alleles or genotypes have been found between keloid patients and newborn controls. There is tight, but not complete, linkage disequilibrium between rs1042522 and rs17878362 TP53 polymorphisms (D′ = 0.667, r = 0.448, and p = 0). No significant differences in the distribution of rs1042522 and rs17878362 TP53 haplotypes or diplotypes have been found between keloid patients and newborn controls. Conclusions. Our results suggest the lack of association of rs1042522 and rs17878362 TP53 polymorphisms and their haplotypes or diplotypes with the susceptibility to keloid scarring in Polish patients.

1. Introduction

Keloid scar is defined as a dermal benign fibro-proliferative growth that extends outside the original wound and invades the adjacent dermal tissue due to extensive production of extracellular matrix, especially collagen, which is caused by over expression of cytokines and growth factors. Although many attempts were made to understand the exact pathophysiology and the molecular abnormalities, the pathogenesis of the keloid scar is yet to be determined [1]. The mechanisms of keloid formation include, among others, decreased apoptotic activity, alterations in growth factors, impaired collagen turnover, as well as immunological and genetic contributions [2, 3]. Recently, Glass pointed out either keloid-linked chromosomal loci or candidate genes for keloid, among the latter TP53 [4].

TP53 is a tumor suppressor gene located on chromosome 17p13.1 that encodes protein p53. The tumor protein p53 binds directly to DNA and participates in the regulation of cell cycle checkpoints, DNA repair, and apoptosis and regulates the repair process in response to damaging factors,
including chemicals, radiation, and ultraviolet rays from sunlight [5]. It has been reported that rs1042522 (c.215G>C, p.Arg72Pro) and rs17878362 (16-bp insertion/duplication in intron 3), two most frequently analyzed TP53 polymorphisms, may affect either the function of the p53 protein or its mRNA expression [6–8].

The results of meta-analysis based on 359 keloid cases and 493 healthy controls revealed no association between germlinal p.Arg72Pro mutation and susceptibility to keloid in the Chinese population [9]. To the best of our knowledge in contrast to rs1042522, to date no study for association of intronic rs17878362 TP53 polymorphism with keloid scarring has been carried out.

Therefore, the aim of our study was to analyze the association of rs1042522 and rs17878362 and their haplotypes with the predisposition to keloid in Caucasians.

2. Materials and Methods

The study group consisted of 86 consecutive patients with keloid (aged from 18 to 70 years old), who were treated with surgical excision in the Aesthetic Med, Szczecin, Poland. Characteristics of the studied patients are shown in Supplemental Table 1. The control group consisted of 100 healthy, full-term newborns (52 males and 48 females) randomly chosen from the Newborn DNA Repository at the Department of Clinical and Molecular Biochemistry at the Pomeranian Medical University in Szczecin. All children were breast-fed and free of medication. Twins and infants of mothers with keloid scarring, preeclampsia, hypertension of any cause, diabetes, history of illicit substance use, or antenatal steroid therapy were excluded. Other exclusion criteria were congenital infection, intrauterine growth restriction (i.e., below the 10th percentile birth mass, length, or head circumference), chromosomal aberrations, or congenital malformations. All patients in study and control groups were Poles of European descent. The study was conducted in accordance with the Declaration of Helsinki and was approved by the local bioethics committee at the Pomeranian Medical University in Szczecin. Parental informed consent for cases and parental informed consent for newborn controls were obtained.

Genomic DNA was extracted either from peripheral blood leukocytes (keloid patients) or from cordal blood leukocytes (newborn infants) using a commercially available DNA isolation kit (QIAamp Blood DNA Mini Kit, QIAGEN, Germany). Amplification of the 540-bp TP53 sequence including rs1042522 and rs17878362 was performed by using PCR with 5'-AACCCCAGCCCCCTAGCAGA-GACC-3' as the forward primer and 5'-GGGGATACGGCCAGGATACGGCCAGGATACGG-3' as the reverse primer. Subsequently, PCR amplification products were purified using Exonuclease I and FastAP Thermosensitive Alkaline Phosphatase (ThermoFisher Scientific Inc., Waltham, MA, USA) according to manufacturer procedures. Sequencing of the products used BigDye® Terminator v3.1 Cycle Sequencing Kits (Applied Biosystems, Life Technologies Polska, Warsaw, Poland). Electrophoresis and analysis were performed according to manufacturer procedures using an ABI PRISM 3100-Avant machine (Data Collection Software v2.0, Sequencing Analysis Software v5.4; Applied Biosystems).

Possible divergence of rs1042522 and rs17878362 TP53 genotype frequencies from the Hardy–Weinberg equilibrium was assessed using a χ² test. The association between a pair of analyzed loci (linkage disequilibrium, LD) was tested using the χ² test with the parameter D' and correlation coefficient r. The Hardy–Weinberg equilibrium and LD were analyzed using the “Genetics” package. Frequency differences in frequencies of genotypes, alleles, haplotypes, or diplotypes between groups were tested for statistical significance using the χ² test or Fisher’s exact test, if necessary. Genotype frequencies between groups were then compared by univariate logistic regression in additive, dominant, or recessive mode of inheritance of the risk allele.

The “haplo.score” and “haplo.glm” functions from the “haplo.stats” package were applied to test the effect of haplotypes. A positive Hap.Score value implies that the haplotype occurs more frequently in the keloid patients than control subjects, whereas a negative Hap.Score indicates that the haplotype occurs more frequently in control subjects.

3. Results

The rs1042522 and rs17878362 TP53 genotype distributions both in keloid patients and in the control group conformed to the expected Hardy–Weinberg equilibrium (p = 0.709 and p = 0.864 or p = 0.952 and p = 0.425, respectively). No significant differences in the distribution of rs1042522 and rs17878362 TP53 alleles or genotypes have been found between male newborn controls and female ones (p = 0.773 or p = 0.525 for alleles and p = 0.961 or p = 0.427 for genotypes, respectively). No significant differences in the distribution of rs1042522 and rs17878362 TP53 alleles or genotypes have been found between keloid patients and newborn controls (p = 0.690 or p = 0.496 for alleles and p = 0.184 or p = 0.718 for genotypes, respectively). Univariate logistic regression revealed no significant association between rs1042522 and rs17878362 TP53 polymorphisms and predisposition to keloid in additive mode of inheritance of the risk allele (C allele of rs1042522 or A2 allele of rs17878362, respectively) (Table 1) as well as in dominant (p = 0.107 for rs1042522 or p = 0.440 for rs17878362, respectively) or in recessive (p = 0.206 for rs1042522 or not available for rs17878362, respectively) mode of inheritance of the risk allele.

There is tight, but not complete, linkage disequilibrium between rs1042522 and rs17878362 TP53 polymorphisms (D' = 0.667, r = 0.448, p = 0). No significant association of TP53 haplotypes and the risk of keloid scarring have been found using both global score and haplotype-specific
Table 1: Association analysis of two TP53 gene polymorphisms with keloid in Caucasians.

| Polymorphism (position) | Allele^b (1/2) | Cases 1/2 (%) | Controls 1/2 (%) | P | OR (95% CI)^d |
|-------------------------|----------------|--------------|-----------------|---|---------------|
| rs1042522 (chr. 17: 7579472) | G/C | 137/35 (80/20) | 143/57 (72/28) | 0.690 | 0.63 (0.39–1.04) |
| rs17878362 (chr. 17: 7579690) | A1/A2 | 152/20 (88/12) | 172/28 (86/14) | 0.496 | 0.80 (0.42–1.51) |

^aSNP position was indexed to the NCBI build 37 (GRCh37.p13). ^bAllele 1 and allele 2 were defined as the nonsusceptible allele or the risk allele, respectively. ^cP values for logistic regression in additive mode of inheritance of the risk allele. ^dORs and CIs were calculated using the nonsusceptible allele as a reference.

There are also no significant differences in frequency distribution of TP53 diplootypes between the keloid patients and control group by comparing every diplootype to the reference one (D1) (Table 3).

4. Discussion

Somatic mutations in the TP53 gene encoding tumor protein p53 are one of the most common genetic abnormalities associated with human cancer and have been implicated as causal events in up to 50% of all human malignancies. Germline TP53 mutations also increase the risk of numerous neoplasm types, including breast cancer, leukemia, sarcomas, and central nervous system tumors [5].

The rs1042522 is a G to C transversion at the second position of codon 72 in exon 4 leading to substitution of arginine (CGC) by proline (CCC) in p53 polypeptide chain (c.215G>C, p.Arg72Pro). The transversion is located in the N-terminal proline-rich domain (residues 64–92) of the protein required for the growth suppression and apoptosis mediated by the p53 [10]. It has been reported that the p.Arg72 p53 variant induces apoptosis with faster kinetics and suppresses transformation more efficiently than the p.Pro72 one [6]. In addition, Dumont et al. indicated that the higher efficiency of p.Arg72 variant in triggering cellular apoptosis is due to its greater ability to localize to the mitochondria [7].

In 2005, Zhuo et al., using polymerase chain reaction-reverse dot blot (PCR-RDB) and DNA direct sequencing analyzed Chinese subjects (15 patients with keloid and 15 healthy controls) and revealed that the frequency of both p.Pro72 TP53 allele and p.Pro72 TP53 homozygous genotype in keloid patients was significantly higher than that in the controls [11]. In contrast, Wang et al. also reported in 2005 that p.Arg72 TP53 homozygous genotype in Japanese subjects is associated with the risk for the piercing-induced ear-lobe keloid [12]. Further studies performed exclusively in Chinese patients also yielded conflicting results [13–17].

To the best of our knowledge, our study is a first report about rs1042522 TP53 polymorphism and the predisposition to keloid in subjects of European descent (Poles). The frequency of the minor c.215C (p.Pro72) allele equal 28.5% in our controls was close to values from the 1000 Genomes Project (https://www.ncbi.nlm.nih.gov/variation/tools/1000genomes/) in other European populations, which ranged from 24.2% in CEU (Utah residents with Northern and Western European ancestry) to 30.8% in British individuals in England and Scotland (GBR). The prevalence of p.Pro72 TP53 allele in Poles is also very close to the variant frequencies found in the Czechs and Slovaks, our nearest neighbors of Slavic origin (Czechs 29.1% or 29.4% and Slovaks 25.4%, respectively) [18–20]. We have found no significant differences in the frequency distribution of both rs1042522 TP53 alleles and genotypes between keloid patients and newborn controls. On the other hand, the frequency of minor (p.Pro72) allele in keloid patients was 8% lower as compared with control subjects (20% versus 28%, respectively). Also, Liu [15] and Yan et al. [16] reported of 8% lower frequency of p.Pro72 TP53 allele in Chinese keloid patients as compared with healthy controls. However, it is noteworthy, that the frequency of this allele in Chinese, both in the study (40% or 41%) and control groups (48% or 498%) [15, 16], was significantly higher compared to our data from Caucasians.

The results of meta-analysis based on six studies [11, 13–17] did not confirm the association between rs1042522 TP53 polymorphism and susceptibility to keloids in Chinese patients [9]. However, subsequent subgroup analysis in regard to genotyping method (PCR-reverse dot blot versus PCR-RFLP) revealed contradictory results. The p.Pro72 TP53 allele was associated with the predisposition to keloid scarring only in patients genotyped by using PCR-reverse dot blot [11, 13–15]. In contrast, the association between p.Arg72 TP53 allele and predisposition to keloid has been found in patients analyzed by using PCR-RFLP [16, 17]. And although the authors of meta-analysis argue that different detection methods may be the source of the above heterogeneity, we suppose that the major cause of it is rather population bias evidenced in the 1000 Genomes Project (1KG Project). The p.Pro72 TP53 frequency in CHB (Han Chinese in Beijing), CHS (Southern Han Chinese), and CDX (Chinese Dai in Xishuangbanna) was 45.1%, 40.0%, and 47.3%, respectively. It is also worth emphasizing that the highest p.Pro72 TP53 frequency was observed in populations of Sub-Saharan Africa (63.9% or 68.2% in Nigeria, 70/8% in Gambia, or even 74.7% in Kenya). And despite both these results of 1KG Project and some previous studies [21, 22] strongly supported the hypothesis that the p.Pro72 TP53 frequency is latitude dependent, Shi et al. have found that rs1042522 TP53 polymorphism rather associates with winter temperature [23].

The rs17878362 is a 16-base pair (bp) insertion/duplication in intron 3 of TP53 gene consisting of one copy (A1 allele) or two copies (A2 allele) of the sequence...
Gemignani et al. have found that reduced levels of TP53 mRNA are associated with the A2 allele [8]. It has not escaped of our notice that our study is a first association study of rs17878362 TP53 polymorphism and rs1042522/rs17878362 TP53 haplotypes with the risk of keloid scarring. No significant differences in the frequency distribution of both rs17878362 TP53 alleles and genotypes have been found between keloid patients and controls in our study. The prevalence of A2 TP53 allele in Polish newborns (14%) is also very close to the variant frequencies found in other populations of Slavic descent (15.5% and 17.0% in Czechs, 16.8% in Slovacks, or 12.7% in Russians) [18–20, 24]. We found also that linkage disequilibrium between rs1042522 and rs17878362 TP53 polymorphisms ($D' = 0.667$ and $r^2 = 0.221$) was very close to values in other Slavic populations ($D' = 0.570$ and $r^2 = 0.009$ in Russians or $D' = 0.693$ and $r^2 = 0.220$ in Czechs). No significant differences in the frequency distribution of both rs1042522/rs17878362 TP53 haplotype and diplotype have been found between keloid patients and controls in our study. On the other hand, the frequencies of both “wild-type” haplotype H1 (p.Arg72-A2) and “wild-type” diplotype D1 (H1/H1) were respectively of 7% or 8% higher as compared with control subjects (76% versus 69% for H1 or 56% versus 48% for D1, respectively). Also, Naccarati et al. [18] and Vymetalkova et al. [19] reported very similar H1 frequency in healthy adults in Czech Republic (67.1% or 65.5%, respectively).

We are fully aware that the major limitation of our study is relatively low statistical power due to small sample size. Therefore, we computed the minimal sample size necessary to achieve 80% statistical power for the detection of required ORs. The calculation for rs1042522 TP53 polymorphism was performed using Genetic Power Calculator [25] in different models of inheritance and under the assumptions of 0.09% keloid prevalence [26], minor allele frequency equal 28.5%, case-to-control ratio equal 0.86, and 5% type I error rate ($\alpha$).

5. Conclusion

Our results suggest the lack of association of rs1042522 and rs17878362 TP53 polymorphisms and their haplotypes or diplotypes with the susceptibility to keloid scarring in Polish patients.

Data Availability

The data used to support the findings of our study are included within the article.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

Supplementary Materials

Supplemental Table 1. Clinical characteristics of the keloid patients ($n = 86$). (Supplementary Materials)

References

[1] W. Mari, S. G. Alsabri, N. Tabal, S. Younes, A. Sherif, and R. Simman, "Novel insights on understanding of keloid scar: article review," Journal of the American College of Clinical Wound Specialists, vol. 7, no. 1–3, pp. 1–7, 2016.

[2] D. N. Sayah, C. Soo, W. W. Shaw et al., "Downregulation of apoptosis-related genes in keloid tissues," Journal of Surgical Research, vol. 87, no. 2, pp. 209–216, 1999.
[3] A. S. Halim, A. Emami, I. Salahshourifar, and T. P. Kannan, “Keloid scarring: understanding the genetic basis, advances, and prospects,” *Archives of Plastic Surgery*, vol. 39, no. 3, pp. 184–189, 2012.

[4] D. A. Glass 2nd, “Current understanding of the genetic causes of keloid formation,” *Journal of Investigative Dermatology Symposium Proceedings*, vol. 18, no. 2, pp. S50–S53, 2017.

[5] J. Huszno and E. Grzybowska, “TP53 mutations and SNPs as prognostic and predictive factors in patients with breast cancer,” *Oncology Letters*, vol. 16, no. 1, pp. 34–40, 2018.

[6] M. Thomas, A. Kalita, S. Labrecque, D. Pim, L. Banks, and G. Matlashewski, “Two polymorphic variants of wild-type p53 differ biochemically and biologically,” *Molecular and Cellular Biology*, vol. 19, no. 2, pp. 1092–1100, 1999.

[7] P. Dumont, J. I.-J. Leu, A. C. Della Pietra 3rd, D. L. George, and M. Murphy, “The codon 72 polymorphic variants of p53 have markedly different apoptotic potential,” *Nature Genetics*, vol. 33, no. 3, pp. 357–365, 2003.

[8] F. Gemignani, V. Moreno, S. Landi et al., “A TP53 polymorphism is associated with increased risk of colorectal cancer and with reduced levels of TP53 mRNA,” *Oncogene*, vol. 23, no. 10, pp. 1954–1956, 2004.

[9] Y. Wu, B. Wang, Y. H. Li et al., “Meta-analysis demonstrates association between Arg72Pro polymorphism in the p53 gene and susceptibility to keloids in the Chinese population,” *Genetics and Molecular Research*, vol. 11, no. 2, pp. 1701–1711, 2012.

[10] C. Venot, M. Maratrat, C. Dureuil, E. Conseiller, L. Bracco, and L. Debussche, “The requirement for the p53 proline-rich functional domain for mediation of apoptosis is correlated with specific PIG3 gene transactivation and with transcriptional repression,” *The EMBO Journal*, vol. 17, no. 16, pp. 4668–4679, 1998.

[11] Y. Zhuo, J. H. Gao, S. Q. Luo et al., “p53 gene codon 72 polymorphism and susceptibility to keloid,” *Zhonghua Zheng Xing Wai Ke Za Zhi*, vol. 21, no. 3, pp. 201–203, 2005.

[12] C. M. Wang, H. Hiko, and N. Nakazawa, “Investigation of p53 polymorphism for genetic predisposition of keloid and hypertrophic scar,” *Zhonghua Zheng Xing Wai Ke Za Zhi*, vol. 21, no. 1, pp. 32–35, 2005.

[13] J. Jin, J. H. Gao, and F. Lu, “Clinical experiment of susceptible people to keloid,” *Zhongguo Lin Chuang Jie Pao Xue Za Zhi*, vol. 25, pp. 320–322, 2007.

[14] Y. Zhuo, J. H. Gao, and X. Y. Zeng, “The application of p53 gene detection kit for susceptibility of keloid,” *Zhongguo Mei Rong Yi Xue*, vol. 5, pp. 694–696, 2008.

[15] Y. B. Liu, *The study of impaired apoptosis function of Fas and p53 protein in the fibroblasts derived from keloid*, Ph.D. thesis, Southern Medical University, Guangzhou, China, 2008.

[16] L. Yan, X. Y. Lu, C. M. Wang, and R. Cao, “Association between p53 gene codon 72 polymorphism and keloid in Chinese population,” *Zhonghua Zheng Xing Wai Ke Za Zhi*, vol. 23, pp. 428–430, 2007.

[17] Y. Liu, “Preliminary linkage analysis of keloid susceptibility loci and polymorphisms of correlation genes in Chinese Han population,” Master’s thesis, China Medical University, Shenyang, China, 2007.

[18] A. Naccarati, B. Pardini, V. Polakova et al., “Genotype and haplotype analysis of TP53 gene and the risk of pancreatic cancer: an association study in the Czech Republic,” *Carcinogenesis*, vol. 31, no. 4, pp. 666–670, 2010.

[19] V. Vymetalhokova, P. Soucek, T. Kunicka et al., “Genotype and haplotype Analyses of TP53 gene in breast cancer patients: association with risk and clinical outcomes,” *PLoS One*, vol. 10, no. 7, Article ID e0134463, 2015.

[20] M. Skreiehová, E. Halašová, T. Matáková et al., “Low variability and stable frequency of common haplotypes of the TP53 gene region in colorectal cancer patients in a Slovak population,” *Anticancer Research*, vol. 37, no. 4, pp. 1901–1907, 2017.

[21] G. Beckman, R. Birgander, A. Själander et al., “Is p53 polymorphism maintained by natural selection?,” *Human Heredity*, vol. 44, no. 5, pp. 266–270, 1994.

[22] A. Själander, R. Birgander, A. Kivelä, and G. Beckman, “p53 polymorphisms and haplotypes in different ethnic groups,” *Human Heredity*, vol. 45, no. 3, pp. 144–149, 1995.

[23] H. Shi, S.-J. Tan, H. Zhong et al., “Winter temperature and UV are tightly linked to genetic changes in the p53 tumor suppressor pathway in Eastern Asia,” *The American Journal of Human Genetics*, vol. 84, no. 4, pp. 534–541, 2009.

[24] E. N. Voropaeva, M. I. Voevoda, T. I. Pospelova, and V. N. Maksimov, “Linkage disequilibrium and haplotypes of rs1042522, rs1625895 and rs17878362 gene TP53 markers in patients with diffuse large B-cell lymphoma,” *Molecular Biology*, vol. 48, no. 5, pp. 763–770, 2014.

[25] S. Purcell, S. S. Cherny, and P. C. Sham, “Genetic power calculator: design of linkage and association genetic mapping studies of complex traits,” *Bioinformatics*, vol. 19, no. 1, pp. 149–150, 2003.

[26] E. E. Peacock, J. W. Madden, and W. C. Trier, “Biologic basis for the treatment of keloids and hypertrophic scars,” *Southern Medical Journal*, vol. 63, no. 7, pp. 755–760, 1970.