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To cite this version:
Laura N. Azandi, Tariq Stévart, Bonaventure Sonké, Murielle Simo-Droissart, Tania d’Haijère, et al.. Taxonomic description and pollination ecology of Cyrtorchis okuensis (Orchidaceae, Angraecinae), a new species endemic to the Cameroon Volcanic Line. Plant Ecology and Evolution, 2021, 154, pp.483-496. 10.5091/plecevo.2021.1823. hal-03463042

HAL Id: hal-03463042
https://hal.inrae.fr/hal-03463042
Submitted on 2 Dec 2021

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Taxonomic description and pollination ecology of *Cyrtorchis okuensis* (Orchidaceae, Angraecinae), a new species endemic to the Cameroon Volcanic Line

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**Background and aims** – A recent taxonomic and molecular study of the genus *Cyrtorchis* revealed three new species. Here, we describe one of these new species, endemic to the Cameroon Volcanic Line (CVL), and compare it to its closest relatives. Its conservation status is provided along with detailed information on its ecology and reproductive biology.

**Material and methods** – A detailed examination of 21 herbarium specimens was performed using standard practices of herbarium taxonomy. The conservation status of the new species was assessed using the IUCN Red List Categories and Criteria. Furthermore, we investigated the reproductive biology of the new species in an easily accessible subpopulation in Cameroon and tested the hypothesis of hawkmoth pollination in the genus *Cyrtorchis*.

**Key results** – The newly discovered species, here named *C. okuensis*, is restricted to the montane forest (1600 to 2500 m elevation) of the CVL and is currently assessed as Near Threatened (NT). Among angraecoid orchids in which hawkmoth pollination is recorded (~16 species), *C. okuensis* has the shortest nectar spur (19–40 mm). In situ observations revealed hawkmoths with short proboscis of the genus *Hippotion* (*H. celerio*, *H. eson*, and *H. osiris*) as the main pollinators. The rate of pollination and fruiting under natural conditions was low and among the 448 surveyed flowers only 38.8% (174/448 flowers) had their pollinia removed, 19% produced fruits (87/448 flowers), and 16% (70/448 flowers) set mature fruits.

**Conclusion** – *Cyrtorchis okuensis* is clearly differentiated from *C. submontana* by its longer and wider floral bracts, longer anther caps, and wider triangular dorsal sepal.

**Keywords** – Angraecoid orchids; epiphyte; hawkmoth; IUCN Red List Categories and Criteria; pollination; systematics.

INTRODUCTION

A recent study combining morphological and molecular data on the African angraecoid genus *Cyrtorchis* tested the monophyly of its two sections and re-assessed the circumscription of each described species (Azandi et al. 2021). This study revealed three novelties, and among these, one is endemic to the Cameroon Volcanic Line (CVL). Some of the specimens ascribable to this taxon have been wrongly identified as *C. guillaumetii* (Pérez-Vera) R.Rice, *C. brownii* (Rolfe) Schltr., or *C. submontana* Stévart, Droissart
& Azandi because of their similarities in leaf shape and size and in inflorescence and spur lengths.

African angraecoid orchids include a high proportion of species characterised by white, long-spurred, nectariferous, and night-fragnant flowers, features which are typical of hawkmoth pollination or sphingophily (Martins & Johnson 2007; Micheneau et al. 2009). These pollination syndromes or combination of floral traits are known to have independently evolved to attract hawkmoths (Ashworth et al. 2015). In tropical Africa, several studies on this pollination syndrome have been performed in its eastern part (see Martins & Johnson 2007, 2013), its southern part (see Luyt & Johnson 2001; Johnson & Raguso 2016; Johnson et al. 2017), and in Madagascar (see Nilsson et al. 1985, 1987, 1992; Nilsson 1992, 1998; Wasserthal 1997); yet such studies are missing for Central Africa. Most of these previous studies on angraecoid orchids pollination have demonstrated that the long nectar spurs of orchids and the proboscis of pollinators are involved in evolutionary processes resulting in the remarkable angraecoid floral specialization. The reproductive success of these orchids is known to be pollinator-mediated and generally results in low fruit set due to low visiting frequency of hawkmoths (Micheneau et al. 2009; Amorim et al. 2014). In-depth knowledge about the pollination ecology of threatened species would enable the identification and description of the factors limiting their reproduction and allow proposing adequate conservation measures for both plants and their pollinators (Senapathi et al. 2015).

During our field trips in Cameroon, we discovered a large and accessible subpopulation of the new species in Oku National Park (North-West Region). That subpopulation provided us with the opportunity to test Cribb’s (1989) suggestion that the considerable resemblance of the flowers in the genus Cyrtorchis derives from a common hawkmoth pollination syndrome (i.e. sphingophily), a hypothesis that remains unverified to date.

The present contribution intends to describe the new Cyrtorchis species revealed by the recent study combining morphological and phylogenetic data on Cyrtorchis spp. and to describe, for the first time in Central Africa, aspects of reproductive biology of an angraecoid orchid genus.

**MATERIAL AND METHODS**

**Taxonomy**

**Material and morphological examination** – Taxonomic work is based on a detailed examination of 21 specimens of the novelty, which have been collected in different locations in Cameroon, Bioko (Equatorial Guinea), and Nigeria, 15 of which were collected since 1997 by our team (see Stévart et al. 2020). Specimens include 10 dried and 11 spirit-preserved samples and are deposited in the following herbaria: BRLU, FHI, K, MA, P, WAG, and YA (acronyms according to Thiers continuously updated). For stereomicroscopic observations, flowers from dried materials were initially boiled in water. Morphological investigations were performed using a Zeiss stereo microscope Stemi SV11, both for spirit-preserved and herbarium materials. Morphology of seeds was observed under a light microscope Olympus BX51; quantitative parameters (length, width, and shape of viable seeds and embryos, and number of testa cells) were measured in μm with the image acquisition and processing software ArchiMed microvision v.6.1.3. Measurements, colours, and other details given in the description below are based on living material, spirit-preserved, herbarium specimens, and data derived from field notes.

**Description, geographical distribution, and conservation status assessment** – The terminology used to describe structures and shapes follows the Systematics Association Committee for Descriptive Biological Terminology (1962a, 1962b) and the Kew Plant Glossary (Beentje 2010). The geographical distribution was determined from data given on the herbarium sheets. The distribution map was prepared using the software ArcGIS v.10.5.1 (ESRI 2017). The risk of extinction of the novelty was assessed by applying the International Union for Conservation of Nature (IUCN) Red List Categories and Criteria (IUCN 2012, 2019) and using GeoCAT (Bachman et al. 2011) to calculate the area of occupancy (AOO) and extent of occurrence (EOO).

**Ecology and reproductive biology**

**Study site** – Flowering phenology and floral visitors of the novelty were investigated in the Mount Oku National Park in May 2017. Mount Oku, the second highest mountain on the West African mainland, with an elevation of about 3,011 m a.s.l. (Asanga 2002), is part of the Bamenda Highlands, in the North-West Region of Cameroon (6°12′N, 10°32′E). Mount Oku is located within the Kilum-Ijim forest massif, which is the largest remnant of West African upper montane forests (Maisels et al. 2000). The vegetation includes a mosaic of montane forest, montane grassland, and sub-alpine habitats (Cheek et al. 2000; Asanga 2002).

Our ecological survey was conducted in 2017 in an area of wooded savannah, at the upper montane forest-grassland edge, located on the north-western part of Mount Oku crater lake, which rises to about 2,400 m elevation (fig. 1A). This site was chosen because of the abundance of mature individuals of the novelty within reach, to explore their ecology and reproductive biology. We planned to return to complete our data with more and improved camera traps. However, since mid-2017, troubles in this part of Cameroon prevented all access to the study area. No improvement of the situation seems conceivable in the short- and medium-term.

**Ecology, phenology, and pollination reproductive success** – We studied the ecology of the new Cyrtorchis species in about one hectare of natural vegetation. Phytosociological surveys of 2 m length each (see Stévart 2003) were made on branches of labelled host trees of 33 mature Cyrtorchis individuals bearing one to four inflorescences (one individual per survey) (supplementary file 1). Herbarium vouchers were collected for each of the host trees along with the epiphytes growing with the new species for subsequent identification. To study its reproductive biology, monitoring of flowering from anthesis (opening of the first flower on an individual) was conducted in situ from 5 to 29 May 2017 (25 days). We daily recorded (a) the proportion of individuals with open
flowers, (b) the number of open flowers and their opening date, and (c) the proportion of flowers visited by insects and fruit set (pollinia removal from the gynostemia for male fitness and pollinia load on stigmas or fruit set for female fitness).

**Pollinator observation and syndromes** – To identify the pollinators of this new species, we used the interval-programming function of a waterproof digital camera (Camera Bushnell Natureview HD MAX), using the settings to take pictures at 1-minute intervals, and also by motion detection in the field of vision on five individuals located at least 10 m apart from each other (see details in supplementary file 1). Furthermore, during two nights (on 6–7 May 2017 from 6 pm to midnight, and on 8–9 May 2017 from 7 pm to 2 am), light traps with mercury vapor lamps (Sylvania HQL bulb, 250 W), installed near the surveyed population, were used to investigate and collect potential pollinators (i.e., hawkmoths and settling moths). Captured moth specimens were examined to detect pollinia on their body. A subsequent analysis relating the pictures recorded with the cameras and the daily survey of each flower was used to determine whether a specific species is a passive visitor or an effective pollinator of the novelty. To further confirm the potential pollinator identity, the relationship between the proboscis and spur length was assessed for hawkmoths seen on the camera footage foraging the flowers. The proboscis length of individual hawkmoths was measured to the nearest millimetre (from the base near the head to the tip) with a ruler (Martins & Johnson 2007; Peter & Venter 2017). The spur length of fully developed flowers was measured from BRLU herbarium specimens (n = 9). The captured hawkmoths specimens were sent to Rodolphe Rougerie, a sphingid specialist at the Muséum national d’histoire naturelle in Paris for identification.

**Breeding system** – To test the ability of the new species to reproduce by autonomous self-pollination, we bagged three individuals (with a total of five inflorescences and with 20 closed flower buds) with fine nets prior to flower anthesis and we monitored their development in natural condition until the end of flowering. Hand pollinations were set up ex situ in the Elak village, to investigate the new species’ breeding system, by performing 114 hand-pollinations including 58 cross-pollinations (20 individuals; 58 flowers) and 56 self-pollinations (19 individuals; 56 flowers) on 39 cultivated specimens. The Shapiro-Wilk Normality test (Royston 1982; Dytham 2011) was performed prior to statistical difference

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**Figure 1** – *Cyrtorchis okuensis* in its natural habitat, the shrubby savannah around the volcanic lake Oku. **A.** Panoramic view of *C. okuensis* habitat in the survey area. **B.** Individuals of *C. okuensis* growing on a 2 meter-long branch of *Gnidia glauca*. **C.** A flowering plant of *C. okuensis* growing with mosses *Leptodontium viticulosoides*, and the ferns *Lepisorus excavatus* and *Asplenium theciferum*. Photographs by Vincent Droissart (A) and Laura Azandi (B, C).
testing to evaluate if fruit set followed a normal distribution pattern. To statistically evaluate the difference between the fruit set according to the pollination treatment (self- or cross-pollination) a Wilcoxon rank-sum test was performed, which is recommended for the comparison of two groups of non-parametric datasets (fruit set for self- and cross-pollination treatments), as observed here (e.g. McDonald 2014). These analyses were performed using R v.3.4.4 (R Core Team 2020).

**RESULTS**

**Taxonomy**

*Cyrtorchis okuensis* Droissart, Azandi & M.Simo, sp. nov. (fig. 2) – *Cyrtorchis aff. guillaumetii* “Oku” (Azandi et al. 2021). – Type: CAMEROON • Région du Nord-ouest, arrondissement d’Oku. Savane boisée sur le flanc est du lac Oku; 6°12′18.667″N, 10°27′1.306″E; 28 May 2017; Azandi L. 26; holotype: BRLU; isotype: YA.

**Diagnosis** – *Cyrtorchis okuensis* is similar to *C. submontana* but differs in having wider floral bracts (7–12.5 × 8–13.7 mm vs 4–6.5 × 5–7 mm), longer anther caps (2.5–3 vs 1–2 mm), and wider triangular dorsal sepal (4–5 mm vs 2–3 mm). These species also differ by their habitat: *C. submontana* is found in submontane and evergreen moist forest (between 550 and 1,200 m), whereas *C. okuensis* is restricted to the upper montane vegetation (between 1,600 and 2,500 m) along the CVL.

**Description** – Epiphytic herb, erect, monopodial, with short to long branched stems, 3–18.5 cm long bearing 3–13 alternate leaves with a 16–21.5 cm leaf span. Roots 15.5–20.8 cm long and 0.3–0.4 cm diameter, pale green and covered by mosses. Leaves closely elliptic, distichous, subimbricate, fleshy, olive green, margins entire, 4.6–11.3 × 1.2–3 cm, unequally bilobed at the apex, with rounded lobes. Inflorescences up to 4, axillary or below the leaves, 4.4–9 cm long, 3–12-flowered, flowers 3.3–4.6 cm long with floral bracts 7–12.5 × 8–13.7 mm and usually acuminate at the apex. Flowers white, fading to orange, 3–4.7 cm long and 1–1.8 cm in diameter. Dorsal sepal triangular, 8–13 × 4–5.5 mm. Lateral sepals lanceolate, acute at the apex, 8–15.5 × 3–5.2 mm. Petals similar to lateral sepal but shorter and usually acuminate at the apex, 8–13 × 3–4 mm. Lip triangular with acuminate fleshy apex, 8.5–12 × 5–6 mm; spur incurved sometime sigmoid, 1.9–4 cm long. Anther cap whitish, turning brown with senescence, obovate in the upper part and with narrowed lower part, slightly toothed in the margins and covering the base of the rostellum 2.5–3 × 1.5–2 mm. Pollinia 2, oblate from side rarely ovoid. Column stout and short, 1–2 mm long. Viscidium saddle-shaped, with a uniformly hyaline texture, bifid at the basal apex, 2–4 mm. Stipites spathulate or slightly clavate with acute apices pointing towards the middle side, insertion point at the rear third of the length of the viscidium, 1.5–2.5 mm long. Pedicel with ovary 10.5–19.5 mm long; ovary triquetrous. Rostellum trident, 2.5–4 mm long, midlobe straight, acute, lateral lobes linear, about three times as long as the midlobe, pendulous and parallel to each other with rounded apices. Fruits narrowly ellipsoid capsules, triquetrous with three smooth sides, narrowly ellipsoid, turning yellowish-green at maturity, 12.9–24.5 × 4.5–10.8 mm. Seeds fusiform, 149–194 × 70–93 μm, rarely ellipsoid with ovoid to ellipsoid embryo, 117–141 × 56–76 μm, with two testa cells in an individual seed coat along the longitudinal axis of seed.

**Position** – *Cyrtorchis okuensis* belongs to section Homococolleticon Summerh. based on the structure of its viscidium, uniformly hyaline in texture, and its thin apices of the stipites.

**Distribution** – Endemic to the Cameroon Volcanic Line: Nigeria, Cameroon, and Bioko (Equatorial Guinea) (fig. 3).

**Habitat and ecology** – Restricted to montane vegetation (moist forest, forest-grassland edge, and wooded savannah from 1,600 to 2,500 m), *Cyrtorchis okuensis* grows on mossy branches of *Gnidia glauca* (Fresen.) Gilg (Thymelaeaceae), *Albizia gummifera* (J.F.Gmel.) C.A.Sm. (Fabaceae), and *Prunus africana* (Hook.F.) Kalkman (Rosaceae). On these phorophytes, *C. okuensis* is usually found growing with mosses, lichens, ferns, and other orchid species (see details below in “Ecology and reproductive biology”).

**Phenology** – The flowering period occurs between April and June with a flowering peak in May. Fruit maturation lasts about 230 days on average (from June to January).

**Etymology** – The epithet *okuensis* refers to Oku, a subdivision in the North-West Region of Cameroon where the large, surveyed subpopulation was discovered around Lake Oku. The term Oku also refers to the people who live in this area and their native language.

**Red list conservation status** – The species is given a Red List status of Near Threatened [NT]. The species is known from 21 specimens in Nigeria, Cameroon, and Bioko (Equatorial Guinea), which represent 21 occurrences. The extent of occurrence (EOO) of *Cyrtorchis okuensis* is estimated to be 38,319.1 km², far exceeding the upper limit of the Vulnerable category (20,000 km²) under the subcriterion B1, while its area of occupancy (AOO) is estimated to be 68 km², which falls within the limits of the Endangered category under the subcriterion B2.

In Nigeria, *C. okuensis* has been collected in the Boshi Extension Forest Reserve. In Bioko, the species is known from the Pico Basilé National Park and the Reserva Científica de la Caldeira de San Carlos (at Lago Moka). In Cameroon, the species is recorded in three protected areas: the Mount Oku and the Mount Manengouba National Parks (where some occurrences are below the lower altitudinal limit of effective protection) and the Bafut Ngemba Forest Reserve. In unprotected sites, the species habitat is mainly subject to small-scale shifting agriculture, the logging activities on *Prunus africana*, and the fuelwood collection for small-scale subsistence.

Based on the most serious plausible threat, which is shifting agriculture, the 21 occurrences represent 16 locations (sensu IUCN 2019), which is more than 10, the upper limit of the Vulnerable category under the condition ‘a’ of subcriterion B2. *Cyrtorchis okuensis* is not severely fragmented and there are no extreme fluctuations...
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**Figure 2 – Cyrtorchis okuensis (Azandi 26).** A. Habit and inflorescence. B. Flower, frontal. C. Flower, side view. D. Fruit (cultivated plant Y 6377 BA, fruit ID 499). E. Labellum with spur frontal. F. Dorsal sepal. G. Lateral sepal. H. Petal. I. Anther cap, side view. J. Anther cap, frontal. K. Viscidium with stipites frontal (*Droissart et al. (Ombrière de Yaoundé) 5109*). L. Column, frontal. M. Column, side view. N. Ovary section. O. Pollinia. Drawing by Tania D’haijère.
in its parameters (EOO, AOO, number of locations or subpopulations, and number of mature individuals). The projected ongoing loss of its habitat leads us to predict a continuing decline in the number of subpopulations in four Regions in Cameroon (in Bangem near the Banyang-Mbo Wildlife Sanctuary in the South-West Region, in Bambuto in the West Region, in Mount Manengouba near Nkongsamba in the Littoral Region, in Nkambe and the Bafut Ngemba Forest Reserve in North-West Region), in mature individuals, in its AOO and its EOO. Since two (a and c) of the three conditions under subcriterion B2 are not met, *C. okuensis* cannot be regarded as a threatened species. Considering that shifting agriculture will lead to the loss of four locations in the near future, reducing the number of locations to 12, *Cyrtorchis okuensis* is assigned a IUCN Red List status of Near Threatened (NT).

**Additional specimens examined** – NIGERIA • Boshi Extension Forest Reserve; 6°18′38.16″N, 9°12′46.8″E; 1600 m; 23 May 1971; *van Meer P.P.C. 1765*; FHI web.

CAMEROON • Province du Nord-Ouest, Lac à proximité du village d’Oku; 6°11′48.77″N, 10°27′38.74″E; 2300 m; 3 Sep. 2011; *Droissart V. et al. 894*; BRLU • Belo, Mont Oku, autour du lac; 6°11′8.2″N, 10°25′19.88″E; 2300 m; 21 Apr. 2014; *Droissart V. et Simo M. 1693*; BRLU, YA • ibid.; 5 May 2014; *Droissart V. et al. (Ombrière de Yaoundé) 5046*; BRLU • ibid.; 16 May 2014; *Droissart V. et al. (Ombrière de Yaoundé) 5074*; P, YA • Belo, Mont Oku, savane boisée autour du lac; 6°12′14.4″N, 10°26′E; 2313 m; 28 Apr. 2017; *Droissart V. et al. (Ombrière de Yaoundé) 7163*; BRLU • Forêt du mont Oku; 6°13′51.6″N, 10°31′33.6″E; 15 May 2017; *Droissart V. et al. (Ombrière de Yaoundé) 7174*; BRLU • Entre Kumbo et Ndop; 6°11′1.32″N, 10°39′32.94″E; 9 Jun. 2011; *Simo M. et al. (Ombrière de Yaoundé) 2848*; BRLU • Près du point trigonométrique de Mbem (40 km ESE de Nkambe); 6°26′59.75″N, 11°2′40.92″E; 1800 m; 11 Jul. 1904; *Letouzey R. 8860*; P, YA • Bamenda district, along the footpath from the rest house to number 6 plot in Bafut Ngemba Reserve; 5°55′0″N, 10°13′E; 19 May 1959; *Daramola B.O. FHI 41182*; FHI, K • Région du Sud-Ouest, crête NW Muetan Aku, 9 km SW Bangem; 5°5′4.04″N, 9°45′59.93″E; 2 Jun. 1982; *Villiers J.F. 1397*; BRLU, P n.v. • Province de l'Ouest, Bayangam, village d'Afinita; 5°17′55.7″N, 10°25′50.52″E; 1650 m; 2 Jun. 2014; *Droissart V. et al. (Ombrière de Yaoundé) 5117*; BRLU • Bamputo; 5°44′N, 10°4′E; 2500 m; 21 Nov 1972; *Jacques-Félix H. 5441*; K n.v., P • ibid.; 2300 m; May 1940; *Jacques-Félix H. 5483*; P • Baham; 5°17′12.47″N, 10°22′32.98″E; 1920 m; 28 Apr. 2017; *Droissart V. et al. (Ombrière de Yaoundé) 7164*; BRLU • ibid., 15 Jun. 2017; *Droissart V. et al. (Ombrière de Yaoundé) 7203*; YA • Dschang district, Massif Manengouba, village de Bouroakou to crater lakes; 5°0′0″N, 9°49′48″E; 2133 m; 18 Nov. 1968; *Sanford W.W. 5553*; K • Province du littoral, Melong, Mont Manengouba;

**Figure 3** – Distribution of *Cyrtorchis okuensis* in Central Africa, along the Cameroon Volcanic Line. Map created with ArcGIS v.10.5.1 (ESRI 2017). © Esri and its licensors, all rights reserved.
Identification key to *Cyrtorchis okuensis* and related species (amended from Azandi et al. 2016)

1. Leaves thin and flexible, equally bilobed at the apex ........... *Cyrtorchis ringens* (Rchb.f.) Summerh.

1’. Leaves fleshy and rigid, unequally bilobed at the apex ........................................ 2

2. Floral bract imbricate, longer than the pedicel and ovary ........... *Cyrtorchis brownii* (Rolfe) Schltr.

2’. Floral bract subimbricate or well-space, shorter than the pedicel and ovary ........................................ 3

3. Inflorescence loose at the base and densely flowered near the apex ........................................... *Cyrtorchis guilaumetii* (Pérez-Vera) R.Rice

3’. Inflorescence not loose at the base with flowers distributed all along ........................................ 4

4. Anther cap 2.5–3 mm long, dorsal sepal triangular, 4–5 mm wide ........................................... ........................... *Cyrtorchis okuensis* Droissart, Azandi & M.Simo

4’. Anther cap 1–2 mm long, dorsal sepal narrowly triangular, 2–3 mm wide ........................................... ........................... *Cyrtorchis submontana* Stéwart, Droissart & Azandi

Ecology and reproductive biology

Ecology, phenology and pollination reproductive success – In the shrubby savannah surveyed around Lake Oku, the main host trees were *Gnidia glauca* with 32 host trees and *Prunus africana* with only one host tree. *Gnidia glauca*, a small bushy tree growing in a large, monodominant subpopulation in the study area, was always associated with a high number of individuals of *C. okuensis*, bearing up to 13 individuals (fig. 1B). Specimens of *C. okuensis* were found growing on branches of 4 to 30 cm in diameter, under the canopy of both host trees at 1 to 4 m above ground level.

On the 33 phytosociological branch surveys, the species was found growing in association with mosses such as *Daltonia minuta* Thér. (Daltoniaceae) and *Leptodontium viticulosoides* (P.Beauv.) Wijk & Margad. (Pottiaceae), with the lichen *Usnea barbata* (L.) F.H.Wigg. (Parmeliaceae), with the ferns *Asplenium aethiopicum* (Burm.f.) Becherer, *A. theciferum* (Kunth) Mert., *Lepisorus excavatus* (Bory ex Willd.) Ching, and *Pleopeltis macrocarpa* (Bory ex Willd.) Kauf., with *Kalanchoe arietina* (Andrees) Haw. (Crassulaceae), and with the orchids *Klycanthae hueae* (Schltr.) Farminhão, Stéwart & Droissart and *Polystachya alpina* Lindl.

The time of the opening of the first and the last flower on a given individual varied from two to 10 days for the 16 individuals bearing one to two inflorescences (with up to 19 flowers in total) and from 5 to 28 days for the 14 individuals bearing 3 to 4 inflorescences (with up to 31 flowers in total). This timing of flowering was not estimated for the three individuals for which the anthesis had already passed at the beginning of the survey. The daily monitoring of each of the 33 labelled individual conducted during 25 days enabled us to measure the frequency of flowering and visiting events. Flowering peaks (i.e. the maximum number of flowers opened the same day) in the *C. okuensis* subpopulation surveyed were registered on the 16th, 17th, and 21st day (fig. 4A). More than 50% (17 individuals) of all the 33 surveyed individuals were already flowering on the 13th day and by the end of the survey all the 33 individuals had at least one open flower. Concerning the visitation rate, flower visits started on the 5th day till the end of the monitoring session with a maximum observed on the 11th and 19th day (see fig. 4B). Eight hawkmoth visits recorded with cameras on the 9th, 12th, 16th, 17th, and 23rd day matched daily observations of the number of visited flowers (fig. 4C).

In natural conditions, the rate of pollinarium removal averaged 38.8% (174 flowers out of 448 flowers) for the 33 individuals. Successful pollen deposit on the stigma, indirectly measured by fruit set rate, was 19.4% (87 fruits/448 flowers), and 15.6% reached maturity (70 matured fruits/448 flowers) after an average of 230 days (table 1).

Pollinator observation and syndromes – A total of 72,476 pictures (more than 1207 hours of observation) were recorded with four camera traps, showing visits of at least seven different insects, one spider, and two bird species (the northern double-collared sunbird (*Cinnyris reichenowi* Sharpe, 1891) and the oriole finch (*Lirmurgus olivaceus* Fraser, 1843)). During daytime, flowers were mostly visited by birds and flies, while hawkmoths, settling moths, and grasshoppers were the main visitors at night. Apart from birds, which ate the perianth parts and with possible nectar robbery, and hawkmoths, which only foraged on flowers for nectar, the other insects were passive visitors.

The opening of the flowers occurred only at night. A slight floral scent was detected during daytime when getting near to the flowers. At dusk, from about 6 pm, a jasmine-like scent began to emanate from the flowers and was detectable a few meters away. Scent release began from about 7:30 pm and remained strong throughout the night. Hawkmoth
Table 1 – Breeding system and reproductive success of *Cyrtorchis okuensis* under natural and controlled pollination experiments. Values in brackets represent the percentage of sample size (total number of flowers used or fruits produced) per treatment.

| Treatments                          | Number of individuals | Number of flowers | Number of flowers with pollinia removed (%) | Number of fruits set (%) | Number of mature fruits (%) |
|-------------------------------------|-----------------------|-------------------|-------------------------------------------|--------------------------|----------------------------|
| **In situ**                         |                       |                   |                                           |                          |                            |
| Pollinators excluded (bagged flowers) | 3                     | 20                | –                                         | –                        | –                          |
| Natural pollinations                | 33                    | 448               | 174 (38.8)                                | 87 (19.4)                | 70 (15.6)                  |
| **Ex situ (controlled pollinations)** |                       |                   |                                           |                          |                            |
| Manual self-pollinations            | 19                    | 56                | –                                         | 36 (64.2)                | 21 (37.5)                  |
| Manual cross-pollinations           | 20                    | 58                | –                                         | 51 (88)                  | 27 (46.5)                  |

**Figure 4** – Reproductive phenology of *Cyrtorchis okuensis* per day in the studied population (total of 33 individuals) from 5 to 29 May 2017. **A.** Daily number of open flowers **B.** Daily number of flowers visited. **C.** Daily number of hawkmoth visits recorded with camera traps.
visits mostly occurred at dusk between 6 pm and 1 am with a peak of visits being between 10 pm and 1 am (table 2). The combined camera survey and light trap design enabled us to capture nine individuals of potential effective pollinators of *C. okuensis* (fig. 5A), representing three species of the genus *Hippotion* Hübner, 1819, namely *H. celerio* (Linnaeus, 1758), *H. eson* (Cramer, 1779), and *H. osiris* (Dalman, 1823) (fig. 5B–D). Unfortunately, none of the recorded nor captured hawkmoth specimens were caught with the pollinia attached on any part of their bodies. Nevertheless, based on pollinia removal and/or deposit observed during daily monitoring, as well as fruit set after their visits, we infer that one or more of these hawkmoth species is the effective pollinator of *C. okuensis* in the Mount Oku area.

Morphologically, the mean spur length of *C. okuensis* was 28.2 mm (sd = 0.64, n = 9). This value matches the length of the proboscis of all nine captured *Hippotion* specimens (*H. celerio* (5 individuals), *H. eson* (3), and *H. osiris* (1)), which ranged between 20 and 42 mm, with a mean of 31 mm (sd = 0.86, n = 9). Based on these spur and tongue lengths, the majority of these species of hawkmoth are likely to forage on the flowers in this subpopulation. There was no significant difference in the spur and proboscis length (*p* = 0.89).

**Breeding system assessment** – None of the flowers tested for autonomous self-pollination (bagged inflorescence) produced fruits, suggesting that *C. okuensis* requires a pollinator to set fruit (table 1). The species is however self-compatible as the self-pollination treatments resulted in 64.2% fruit set. Nevertheless, cross-pollination shows a higher success rate with a fruit set of 88%. These two controlled pollination treatments did however not differ significantly for fruit set success (*W* = 138, *p* > 0.05).

**DISCUSSION**

*Cyrtorchis okuensis* is the angraceoid orchid with the shortest spur that is pollinated by hawkmoths

Our formal description of *Cyrtorchis okuensis* constitutes an additional step towards a global revision of the genus *Cyrtorchis*. The current study not only confirms pollination by hawkmoths in the genus *Cyrtorchis*, as predicted by
Cribb (1989), but also brings down the lower limit of spur lengths associated with hawkmoth pollination within the angraecoid group (~800 species; see Simo-Droissart et al. 2018; Farminhão et al. 2021). *Cyrtorchis okuensis* exhibits clear floral adaptations to hawkmoth pollination, such as the white night-scented flowers with a “relatively” long nectar-rewarding spur (Martins & Johnson 2007, 2013; Vogel 2012; Johnson & Raguso 2016; Johnson et al. 2017). These white long-spurred orchids with night-scented flowers are common in Africa (Dressler 1981), with data supporting that about 50% of the African orchids are pollinated by hawkmoths, based on the high frequency of this floral syndrome (Martins & Johnson 2007, 2013; Johnson & Raguso 2016; Johnson et al. 2017). However, the spur length of *C. okuensis* appears to be relatively short (1.9–4 cm vs 4.4–24 cm long) compared to most of the other known African continental angraecoids (~16 species) in which hawkmoth pollination syndromes are documented. Luyt & Johnson (2001) previously reported this pollination syndrome for *Mystacidium venosum* Harv. ex Rolfe, whose spur length ranges from 2.5 to 5.5 cm and Martins & Johnson (2007) later described it for *Aerangis confusa* J.Stewart with an average spur length of 4.5 cm. As observed for other short-spurred angraecoid species (i.e. *M. venosum* and *A. confusa*), there is a good match between the spur length in *C. okuensis* and the proboscis length of the pollinators. This might suggest that a strong selective pressure of the proboscis on spur length might have played a role.

Figure 5 – Pollinators of *Cyrtorchis okuensis* observed around the Lake Oku (North-West Cameroon). A. Pictures recorded with a camera trap showing a *Hippotion* sp. foraging on *C. okuensis* flowers. B–D. Sphinx moths captured during light trap sessions. B. *Hippotion celerio*. C. *H. eson*. D. *H. osiris*. Photographs by Laura Azandi.
Three different species of *Hipppotion* (*H. celerio, H. eson, and H. osiris*) were identified as effective pollinators of *C. okuensis*. These results are in line with previous studies, as *H. eson* was also observed feeding on flowers of *Mystacidium venosum* (Luyt & Johnson 2001) and *H. celerio* was reported to be a pollinator of *Aerangis confusa* in Kenya (Martins & Johnson 2007). Although it was difficult to identify pollinators at the species level, the confirmed presence of the three captured *Hipppotion* spp. and their quite similar proboscis length advocate in support of them being effective pollinators of *C. okuensis*. This was previously found in a study of Alexandersson & Johnson (2002) on *Gladiolus longicollis* Baker, in which moths with matching proboscis lengths were effective agents of pollen removal and deposition. Additionally, more recent studies suggested that hawkmoths are highly polyphagous and readily feed on flowers that have tubes much shorter than their proboscis lengths, while plants adapted to hawkmoths are more specialized, particularly when they are long tubed (Martins & Johnson 2013; Amorim et al. 2014). Given the slight difference in size between the average length of the pollinators’ proboscis and the spur of *C. okuensis*, and the fact that pollinia in the genus *Cyrtorchis* are generally placed along the proboscis (Laura Azandi, pers. obs. on *Cyrtorchis letouzeyi* and *C. chailuana*), we can assume that there is a selection pattern for short spurs as described by Ellis & Johnson (2010) in their sex-specific selection model. This model states that if selection is made through the male, this would favour an evolution towards shorter spurs than proboscis because pollinia export is potentially highest in short-spurred species as pollinia are placed far from the base of the proboscis and could reach both the stigmas of long and short-spurred flowers. Our autonomous self-pollination experiment provides strong evidence that *C. okuensis* fruit production depends on external pollinators. Under natural conditions, hawkmoth visitation events on *C. okuensis* flowers seem relatively rare (see fig. 4 and table 2) and resulted in low reproductive success (pollen removal and fruit set) as only 19.4% of the 448 flowers we surveyed in May 2017 produced fruits. Amorim et al. (2014) and Micheneau et al. (2009) also reported low visitation and fruit set rates of orchid flowers when pollination is accomplished by hawkmoths.

**Importance and conservation of the Cameroon Volcanic Line**

The CVL includes a chain of isolated volcanic or plutonic mountain peaks that covers ~40,877 km², stretching from Annobon Island in the Gulf of Guinea to the Mandara Mountains in the Far North Region in Cameroon. It includes a broader continental part and a smaller oceanic portion in the form of four major islands (Ayonghe et al. 1999; Frodin 2001; Sainje et al. 2017). The CVL is known for its rich and diverse flora with high levels of endemism combined with many threats (Sossef et al. 2017; Droissart et al. 2018), which led to its classification of a biodiversity hotspot (Cheek et al. 2000, 2004; Myers et al. 2000; Barthlott et al. 2005). Plant diversity comprises about 4000 species (Droissart et al. 2018). The orchid family represents one of the most diverse plant groups of the CVL (Cable & Cheek 1998; Cheek et al. 2000, 2004; Onana & Cheek 2011; Harvey et al. 2004) including 33 strictly endemic taxa, distributed in the submontane (17 taxa), montane (12 taxa), and both submontane and montane strata (4 taxa) (Droissart 2009). The new orchid described here further stresses the need for additional fieldwork in these mountains, specifically for sites that have so far been little or not explored. This seems particularly the case for the forests of the Cross River National Park, the Boshii Extension Forest Reserve, and the Gashaka-Gumti National Park in Nigeria, and for the Rumpi Hills area in Cameroon. Additional material from African herbaria (e.g. Forest Herbarium Ibadan (FHI)) should be carefully examined to identify putative unidentified or misidentified orchid material.

With over 200 plant species considered as threatened (Cheek et al. 2004; Onana & Cheek 2011) and with more than 80 endemic species (Cheek et al. 2004; Frankie 2004; Sainje et al. 2005, 2010; Sainje 2012, 2016), the CVL is endangered by increasing threats due to human activities. Land use change and deforestation have resulted in degraded forest areas, fragmented populations, and the CVL is therefore impoverishment on all three levels of biodiversity (i.e. genetic, species, and ecosystem diversity). It has been estimated that over 96% of the original forest cover of the Bamenda highlands above 1,500 m elevation has been lost (Cheek et al. 2000). Hence, a regional synthesis, including recent species diversity checklists coupled with Important Plant Area assessments (Darbyshire et al. 2017), is urgently needed to better manage and conserve the remaining biodiversity of the CVL.

**Threats to the host trees of Cyrtorchis okuensis**

Our results identified *Gnidia glauca* and *Prunus africana* as main host trees of *C. okuensis*. This disparity in the number of host trees identified in this region could be closely related to the fact that our study was conducted at a grassland edge of a natural and monospecific *G. glauca* forest (Momo et al. 2017), but also to the strong threat to *P. africana* over the past several years by unsustainable harvesting in many countries where it occurs. Indeed, this species is used by pharmaceutical companies to manufacture a drug used in treating benign prostatic hyperplasia and prostate gland hypertrophy (Stewart 2003). Cunningham (2005) reported that over the last 40 years, *P. africana* bark harvest for its medicinal properties has shifted from subsistence use to large-scale commercial use for international trade. Cameroon comprises large parts of the current species distribution, and in the Kilum-Ijim area, *P. africana* is traditionally used to treat malaria and stomach ache (Nsorn & Dick 1992). Currently listed as Vulnerable on the IUCN Red List (IUCN 2020), the species is also included in Appendix II of the Convention on International Trade in Endangered Species of Flora and Fauna (CITES) (Betti 2011). Therefore, its overexploitation could have led to a drastic reduction of its populations.

Although *G. glauca* seems less threatened, it is not exempt of risks from future overexploitation as it is also widely used in traditional pharmacopoeia, in nanomedicine, in domestic service in the form of rope or yarn made from bark fibres,
and as an insecticide or piscicide (Ghosh et al. 2012; Avana-Tientcheu et al. 2018). A recent study (Avana-Tientcheu et al. 2018) conducted in the Oku area revealed that G. glauca was more vulnerable in forests than in savannah, because of the proximity of forest stands to neighbouring villages makes them more easily accessible than savannah stands. The IUCN conservation status of G. glauca has not yet been assessed and would help to propose effective conservation measures. As with P. africana (Avana 2006), domestication should be also considered for G. glauca to be able to meet potential high future demands (Avana-Tientcheu et al. 2018).

SUPPLEMENTARY FILE

Supplementary file 1 – Detailed information about the inflorescences and flowers per surveyed individual and about the camera survey sessions. The IDs followed by a * are those surveyed with cameras.

https://doi.org/10.5091/plecevo.2021.1823.2579

ACKNOWLEDGEMENTS

We express our gratitude to the Institut de Recherche pour le Développement (IRD) for financial support provided for field activities in Oku. We are also grateful to the American Orchid Society (AOS) and the Académie de Recherche et d’Enseignement Supérieur (ARES) for funding the PhD activities of the first author in Cameroon and her stay at the herbarium of the Université Libre de Bruxelles, and to the Fonds de la Recherche Scientifique (FNRS-FRIA) for the grant to Tania D’hajière. We also express our gratitude to the National Geographic Society (Grant C303-15, Vincent Droissart as PI) who supported ex situ conservation activities (orchid living collection and seedbank) in Cameroon. We thank all the conservation staff of the Mount Oku National Park and the traditional authorities for allowing us to work in the reserve and access to their facilities. We are grateful to M. Samuel Bilack for his support during the field trip and Roger Kamgang who initiated us to insect light trapping. We gratefully acknowledge the curators of various herbaria (BR, BRLU, K, P, WAG, and YA) for making their collections available for direct examination and loan. We express our sincere gratitude to the curator of the Lepidoptera at the Muséum national d’histoire naturelle in Paris, Dr Rodolphe Rougerie, for his helpful comments on moths’ collection and management, and for the identification of hawkmoths collected in the field. We also acknowledge Dr Vincent Deblauwe for reporting and providing pictures of herbarium sheets of Cyrtorchis okuensis from the Forest Herbarium of Ibadan in Nigeria. We are grateful to Nicolas Texier for his help in providing the distribution map and to João N.M. Farminhão for his constructive comments on the description of the species and for reviewing an earlier version of the manuscript. We finally wish to express our gratitude to Dr Elmar Robbrecht, Dr Thierry Pailler, and an anonymous reviewer who provided a thoughtful and helpful review of the manuscript.

REFERENCES

Alexandersson R. & Johnson S.D. 2002. Pollinator-mediated selection on flower-tube length in a hawkmoth-pollinated Gladiolus (Iridaceae). Proceedings of the Royal Society of London Series B: Biological Sciences 269: 631–636. https://doi.org/10.1098 rspb.2001.1928

Amorim F.W., Wyatt G.E. & Sazima M. 2014. Low abundance of long-tongued pollinators leads to pollen limitation in four specialized hawkmoth-pollinated plants in the Atlantic Rain Forest, Brazil. Naturwissenschaften 101(11): 893–905. https://doi.org/10.1007/s00114-014-1230-y

Asanga C. 2002. Case study of exemplary forest management in Central Africa: community forest management at the Kilum-Ijim mountain forest region, Cameroon. Food and Agricultural Organization of the United Nations, Forestry Department.

Ashworth L., Aguilar R., Martén-Rodríguez S., et al. 2015. Pollination syndromes: a global pattern of convergent evolution driven by the most effective pollinator. In: Pontarotti P. (ed.) Evolutionary biology: diversification from genotype to phenotype: 203–224. Springer, Cham. https://doi.org/10.1007/978-3-319-19932-0_11

Avana-M.L. 2006. Domestication de Prunus africana (Hook.f.) Kalkam (Rosaceae): étude de la germination et du bouturage. PhD thesis, Université de Yaoundé, Cameroon.

Ayonghe S.N., Mafany G.T., Ntasin E. & Samalang P. 1999. Seismically activated swarm of landslides, tension cracks, and a rockfall after heavy rainfall in Bafaka, Cameroon. Natural Hazards 19(1): 13–27. https://doi.org/10.1023/A:1008041205256

Azandi L., Stevart T., Sonké B., Simo-Droissart M., Avana M.L. & Droissart V. 2016. Synoptic revision of the genus Cyrtorchis Schltr. (Angraecinae, Orchidaceae) in Central Africa, with the description of a new species restricted to submontane vegetation. Phytotaxa 267(3): 165–186. https://doi.org/10.11646/phytotaxa.267.3.1

Azandi L., Droissart V., Sonké B., et al. 2021. A near comprehensive phylogenetic framework gives new insights towards a natural classification of the African genus Cyrtorchis (Angraecinae, Orchidaceae). Taxon: 1–27. https://doi.org/10.1002/tax.12511

Bachman S., Moat J., Hill A. W., de la Torre J., & Scott B. 2011. Supporting Red List threat assessments with GeoCAT: geospatial conservation assessment tool. In: Smith V. & Penel L. (eds) e-Infrastructures for data publishing in biodiversity science. ZooKeys 150: 117–126. https://doi.org/10.3897/zookeys.150.2109

Bartholw J., Putke J., Ratnapoor D., Kier G. & Kreft H. 2005. Global centers of vascular plant diversity. Nova Acta Leopoldina 92: 61–83.

Beenetje H. 2010. The Kew plant glossary, an illustrated dictionary of plant terms. Royal Botanic Gardens, Kew, Richmond.

Betti J.L. 2011. Stock of Prunus africana stems on the mount Cameroon forest. African Journal Plant Science 5(13): 781–789. https://doi.org/10.5897/AJPS.9000229
Cable S. & Cheek M. 1998. The plants of Mount Cameroon: a conservation checklist. Royal Botanic Gardens, Kew, Richmond.

Cheek M., Onana J.-M. & Pollard B.J. 2000. The plants of Mount Oku and the Ijm Ridge, a conservation checklist. Royal Botanic Gardens, Kew, Richmond.

Cheek M., Pollard B.J., Darbyshire I., Onana J.-M. & Wild C. 2004. The plants of Kupe, Mwanenguba and the Bakossi Mountains, Cameroon: a conservation checklist. Royal Botanic Gardens, Kew, Richmond.

Cribb P.J. 1989. Orchidaceae 3. In: Polhill R.M. (ed.) Flora of Tropical East Africa: 413–651. Balkema, Rotterdam.

Cunningham A.B. 2005. CITES significant trade review of Prunus africana. In: Review of Significant Trade in specimens of Appendix-II species. Sixteenth meeting of the Plants Committee, Lima (Peru), 3–8 July 2006. PC16 Doc.10.2: 3–55. Available from https://citeseerx.ist.psu.edu/viewdoc/summary?doi=10.1.1.133.558 [accessed 3 Nov. 2021].

Darbyshire I., Anderson S., Asatryan A., et al. 2017. Important plant areas: revised selection criteria for a global approach to plant conservation. Biodiversity and Conservation 26: 1767–1800. https://doi.org/10.1007/s10531-017-1336-6

Dressler R.L. 1981. The orchids: natural history and classification. Harvard University Press, Cambridge.

Droissart V. 2009. Etude taxonomique et biogéographique des plantes endémiques d’Afrique centrale atlantique: le cas des Orchidaceae. PhD thesis, Université Libre de Bruxelles, Belgium.

Droissart V., Dauby G., Hardy O.J., et al. 2018. Beyond trees: biogeographical regionalization of tropical Africa. Journal of Biogeography 45(5): 1153–1167. https://doi.org/10.1111/jbi.13190

Dytham C. 2011. Choosing and using statistics. A biologist’s guide, Third edition. Wiley-Blackwell Science Ldt, USA.

Ellis A.G. & Johnson S.D. 2010. Gender differences in the effects of floral spur length manipulation on fitness in a hermaphrodite orchid. International Journal of Plant Science 171 (9): 1010–1019. https://doi.org/10.1086/656351

ESRI 2017. ArcGIS for desktop. Version 10.5.1. ESRI, Redlands, CA.

Farminhão J.N.M., Verlynde S., Kaymak E., et al. 2021. Rapid radiation of anagroides (Orchidaceae, Angraeciinae) in tropical Africa characterised by multiple karyotypic shifts under major environmental instability. Molecular Phylogenetics and Evolution 159(4): 107105. https://doi.org/10.1016/j.ympev.2021.107105.

Franke T. 2004. Afrothrixismia sangei (Burmanniaceae), a new myco-heterotrophic plant from Cameroon. Systematics and Geography of Plants 74: 27–33. https://www.jstor.org/stable/3668554

Frodin D.G. 2001. Guide to standard floras of the world: an annotated, geographically arranged systematic bibliography of the principal floras, enumerations, checklists and chorological atlases of different areas. Cambridge University Press, Cambridge.

Ghos S., Patil S., Ahire M., et al. 2012. Gnidia glauca flower extract mediated synthesis of gold nanoparticles and evaluation of its chemocatalytic potential. Journal of Nanobiotechnology 10(1): 17. https://doi.org/10.1186/1477-3155-10-17

Harvey Y., Pollard B.J., Darbyshire I., Onana J.-M. & Cheek M. 2004. The plants of Bali Ngemba Forest Reserve, Cameroon: a conservation checklist. Royal Botanic Gardens, Kew, Richmond.

IUCN 2012. IUCN Red List Categories and Criteria. Version 3.1. Second edition. Available from https://www.iucnredlist.org/resources/categories-and-criteria [accessed 1 Mar. 2020].

IUCN 2019. Guidelines for Using the IUCN Red List Categories and Criteria. Version 14. Available from https://www.iucnredlist.org/resources/redlistguidelines [accessed 1 Mar. 2020].

IUCN 2020. The IUCN Red List of Threatened Species. Version 2020-1. Available from https://www.iucnredlist.org/species/33631/9799059 [accessed 4 Jun. 2020].

Johnson L.A. & Raguso R.A. 2016. The long-tongued hawkmoth pollinator niche for native and invasive plants in Africa. Annals of Botany 117: 25–36. https://doi.org/10.1093/aob/mcv137

Johnson S.D., More M., Amorim F.W., et al. 2017. The long and the short of it: a global analysis of hawkmoth pollination niches and interaction networks. Functional Ecology 31: 101–115. https://doi.org/10.1111/1365-2435.12753

Luyt R. & Johnson S.D. 2001. Hawkmoth pollination of the African epiphytic orchid Mystacidium venosum, with special reference to flower and pollen longevity. Plant Systematic and Evolution 228: 49–62. https://doi.org/10.1007/s0060601700036

Maisels F.G., Cheek M. & Wild C. 2000. Rare plants on Mount Oku summit, Cameroon. Oryx 34(2): 136–140. https://doi.org/10.1016/j.oryx.2000.01017.x

Martins D.J. & Johnson S.D. 2007. Hawkmoth pollination of aerangoid orchids in Kenya, with special reference to nectar sugar concentration gradients in the floral spurs. American Journal of Botany 94(4): 650–659. https://doi.org/10.3732/ajb.94.4.650

Martins D.J. & Johnson S.D. 2013. Interactions between hawkmoths and flowering plants in east africa: polyphagy and evolutionary specialization in an ecological context. Biological Journal of the Linnean Society 110: 199–213. https://doi.org/10.1111/bij.12107

McDonald J.H. 2014. Handbook of biological statistics. Third edition. Sparky House Publishing, Baltimore, Maryland.

Micheneau C., Johnson S.D. & Fay M.F. 2009. Orchid pollination: from Darwin to the present day. Botanical Journal of the Linnean Society 161(1): 1–19. https://doi.org/10.1111/j.1095-8339.2009.00995.x

Momo S.M.C., Kamga Y.B., Nguefos V.F., Tiokeng B., Avana-Tientchou M.L. & Kemeuzeu V.A. 2017. Diversité floristique et variation altitudinale de la structure des formations a Gnidia glauca (Fresen) Gilg. dans les forêts communautaires de Kilum-Limé (Nord-Ouest Cameroun). European Scientific Journal 13(6): 17–37. https://doi.org/10.19044/esj.2017.v13n6p17

Nyland M., Mitteimer R.A., Mitteimer C.G., da Fonseca G.A.B. & Kent J. 2000. Biodiversity hotspots for conservation priorities. Nature 403: 853–858. https://doi.org/10.1038/35002501

Nilsson L.A., Jonsson L., Rason L. & Randrianjohany E. 1985. Monophyly and pollination mechanisms in Angraecum arachnites Schltr. (Orchidaceae) in a guild of long-tongued hawk-moths (Sphingidae) in Madagascar. Biological Journal of the Linnean Society 26(1): 1–19. https://doi.org/10.1111/j.1095-8312.1985.tb01549.x

Nilsson L.A., Jonsson L., Ralison L. & Randrianjohany E. 1987. Angraecoid orchids and hawkmoths in central Madagascar:
specialized pollination systems and generalist foragers. 
Biotropica 19(4): 310–318. 
https://www.jstor.org/stable/2388628

Nilsson L.A. 1992. Orchid pollination biology. Trends in Ecology &
Evolution 7(8): 255–259. 
https://doi.org/10.1016/0169-5347(92)90170-G

Nilsson L.A. 1998. Deep flowers for long tongues. Trends in
Ecology & Evolution 13: 259–260. 
https://doi.org/10.1016/S0169-5347(98)01509-2

Nilsson L.A., Rabakonandrianina E. & Pettersson B. 1992. Exact
tracking of pollen transfer and mating in plants. Nature 360: 
666–668. 
https://doi.org/10.1038/360666a0

Nsom C.L. & Dick J. 1992. An ethno-botanical tree survey of the
Kom area. Unpublished document. Ijim Mountain Forest
Project.

Onana J.M. & Cheek M. 2011. The Red data book of the flowering
plants of Cameroon. Royal Botanic Gardens, Kew, Richmond.

Peter C.I. & Venter N. 2017. Generalist, settling moth pollination
in the endemic South African twig epiphyte, Mystacidium
pusillum in the endemic South African twig epiphyte, Mystacidium
pusillum Harv. (Orchidaceae). Flora 232: 16–21. 
https://doi.org/10.1016/j.flora.2016.11.014

R Core Team 2020. R: a language and environment for statistical
computing. R Foundation for Statistical Computing, Vienna. 
Available from https://www.R-project.org [accessed 10 Aug.
2021].

Royston J.P. 1982. An extension of Shapiro and Wilk’s W test for
normality to large samples. Applied Statistics 31: 115–124. 
https://doi.org/10.2307/2347973

Sainge M.N., Franke T. & Agerer R. 2005. Afrothismia korupensis
(Burmanniaceae, tribe Thismiaceae) from Korup National Park,
Cameroon. Wildenovia 35: 287–291. 
https://doi.org/10.3372/wi.35.35209

Sainge M.N., Franke T., Merckx V. & Onana J.-M. 2010. Distribution of myco-heterotrophic (saprophytic) plants of
Cameroon. In: van der Burgt X., van der Maesen J. & Onana
J.-M. (eds) Systematics and conservation of African plants:
Distribution of myco-heterotrophic (saprophytic) plants of
Cameroon. Royal Botanic Gardens, Kew, Richmond.

Sainge M.N. 2012. Systematics and Ecology of Thismiaceae of
Cameroon. MS thesis, University of Buea, Cameroon.

Sainge M.N. 2016. Patterns of distribution and endemism of plants in the Cameroon Mountains: a case study of Protected Areas in
Cameroon: Rumpi Hills Forest Reserve (RHFMR and the Kimbi
Fungom National Park (KFNP). Technical Report, Tropical
Plant Exploration Group (TroPEG), Buea, Cameroon.

Sainge M.N., Onana J.-M., Nchu F., Kenfack D. & Peterson A.T.
2017. Botanical sampling gaps across the Cameroon Mountains. 
Biodiversity Informatics 12: 76–83. 
https://doi.org/10.17161/bi.v12i0.6707

Senapathi D., Biessmeijer J.C., Breeze T.D., Kleijn D., Potts S.G. &
Carvalheiro L.G. 2015. Pollinator conservation - The difference
between managing for pollination services and preserving
pollinator diversity. Current Opinion in Insect Science 12: 93–
101. https://doi.org/10.1016/j.cois.2015.11.002

Simo-Droissart M., Plunkett G.M., Droissart V., et al. 2018. New
phylogenetic insights toward developing a natural generic
classification of African angrecoid orchids (Vandeae, 
Orchidaceae). Molecular Phylogenetics and Evolution 126: 
241–249. https://doi.org/10.1016/j.ympev.2018.04.021

Sosef M.S.M., Dauby G., Blach-Overgaar A., et al. 2017. Exploring
the floristic diversity of tropical Africa. BMC Biology 15(1): 15. 
https://doi.org/10.1186/s12915-017-0356-8

Stévat T. 2003. Etude taxonomique, écologique et
phytographique des Orchidaceae en Afrique centrale
atlantique. PhD thesis,Université Libre de Bruxelles, Belgium.

Stévat T., Akouangou E., Andriamahefarivo L., et al. 2020. The
Missouri Botanical Garden. In: Hermans J., Hermans C.,
Linsky J. & Li C.W. (eds) World orchid collections: 26–43,
Taiwan Orchid Growers Association, Tainan, Taiwan.

Stewart K.M. 2003. The African cherry (Prunus africana): can
lessons be learned from an over-exploited medicinal tree? 
Journal of Ethnopharmacology 89(1): 3–13. 
https://doi.org/10.1016/j.jep.2003.08.002

Systematics Association Committee for Descriptive Biological 
Terminology 1962a. II. Terminology of simple symmetrical
plane shapes (chart 1a). Taxon 11: 145–148. 
https://doi.org/10.2307/1216718

Systematics Association Committee for Descriptive Biological 
Terminology 1962b. Ila. Terminology of simple symmetrical
plane shapes (chart 1), addendum. Taxon 11: 245–245. 
https://doi.org/10.2307/1217034

Thiers B. continuously updated. Index Herbariorum: a global
directory of public herbaria and associated staff, New York
Botanical Garden’s Virtual Herbarium. Available from: 
http://sweetgum.nybg.org/ih/ [accessed 5 Feb. 2014].

Vogel S. 2012. Floral biological syndromes as elements of diversity 
within tribes in the flora of South Africa. Shaker, Aachen.

Wasserthal L.T. 1997. The pollinators of the Malagasy star orchids
Angraecum sesquipedale, A. sororium and A. compactum
and the evolution of extremely long spurs by pollinator shift. 
Botanica Acta 110 (5): 343–359. 
https://doi.org/10.1186/s12915-017-0356-8

Communicating editor: Elmar Robbrecht.

Submission date: 14 Oct. 2020
Acceptance date: 12 Aug. 2021
Publication date: 23 Nov. 2021