Introduction

Hypericum perforatum L. (St. John’s wort) is a member of the Hypericum genus, which includes some of the most widely used herbal substances (8,19). This plant possesses antimicrobial properties and is used to treat depression; however, it has been implicated in drug interactions (4,12,16). Herbal remedies can alter a patient’s hematological status and cause abnormal blood cell count results (1), but the impact of St. John’s wort on bone marrow is unknown. In this case report, we describe a patient who developed fatal bone marrow necrosis associated with use of St. John’s wort.

Case Report

A 22-year-old male was admitted to our hospital with fever, oral ulceration and fatigue. He had been well until 7 days earlier, when he developed a sore throat and a fever of 39.2 °C. That evening, he visited the Ear, Nose and Throat Department of another hospital. The only abnormalities detected on physical examination were temperature 40.2 °C and pharyngeal hyperemia. A chest radiograph was unremarkable. Laboratory investigations revealed normal serum levels of blood urea nitrogen, creatinine, glucose, and normal urine analysis. No hematologic testing was done. A throat swab was taken, and the culture grew normal flora. Although the patient was diagnosed with a viral syndrome he was started empirical amoxicillin therapy. However, when his fever persisted he came to our hospital.

On admission to our center, a complete history is taken before a physical exam. He had no history of travel to rural/agricultural areas, nor any exposure to animals. He did not use alcohol, tobacco or illicit drugs. There was a history of depression, and the patient had been taking fluoxetine (Prozac®, Lilly) 90 mg/day for 3 months. For the 3 weeks prior to admission, he had also been taking roughly 1000 mg/day of flowering herbs (Hypericum perforatum L., known as St. John’swort) for treatment of depression. A complete blood count on the first day of hospitalization showed agranulocytosis and normocytic anemia. His condition worsened, and he developed progressive dysphagia. A bone marrow biopsy on day 3 revealed bone marrow necrosis. After the diagnosis was established (day 3 of hospitalization), treatment with granulocyte colony-stimulating factor 48 U/day, intravenous immunoglobulin 400 mg/kg, and amphotericin B 100 mg/day was initiated. The patient did not respond, and died within one week of the diagnosis. This case suggests that Hypericum species may lead to severe hematologic toxicity, with conditions involving bone marrow necrosis.
Serum biochemistry analysis on admission revealed total bilirubin 4.9 mg/dL, direct bilirubin 3.4 mg/dL, serum albumin 3.1 g/dL, blood urea nitrogen 22 mg/dL, creatinine 1.0 mg/dL, lactate dehydrogenase 202 IU/L, alanine aminotransferase 404 IU/L, and aspartate aminotransferase 97 IU/L. By the third day of hospitalization, the levels of these two liver enzymes had dropped to 68 IU/L and 30 IU/L, respectively. Cytokine levels (IL-1β, IL-2R, IL-6, IL-8, IL-10, TNF-alpha) in plasma were analyzed by using chemiluminescent method and immulite hormon autoanalyser (Diagnostic Product Corporation, IL, USA). All cytokine levels in plasma except IL-1β were found to be higher than normal range (Tab. 1). Serologic testing was also done on the first day of hospitalization. The results for hepatitis B and C viruses, human immunodeficiency viruses (HIV) -1 and -2, parvovirus B19, Epstein-Barr virus (EBV) immunoglobulin (Ig) M, herpes simplex viruses 1 and 2, cytomegalovirus IgG and IgM, rubella, Brucella spp., typhoid fever, syphilis, and Toxoplasma IgM were all negative.

On the second day of hospitalization, chest radiography and abdominal computed tomography showed nothing remarkable. Computed tomography of the paranasal sinuses revealed sinusitis on the right side. Electrocardiography showed sinus tachycardia (rate 120 beats/min). Abdominal ultrasound was normal. Multiple blood, bone marrow and urine cultures were also done on day 2, and none of these showed growth. A culture of material from the oropharynx grew mixed flora, and the majority of the isolates were Klebsiella spp. and Enterobacter spp. The hemoglobin level decreased from 10 to 6 g/dL within 3 days, and schistocytes were seen in peripheral blood smear on day 3, which reflects microangiopathic hemolytic anemia.

Based on the abnormal findings on the complete blood cell count and the peripheral smear, the bone marrow of the posterior iliac crest was aspirated and biopsied on the third day of hospitalization. The aspirate showed marrow necrosis, with only a few mononuclear cells and histiocytes preserved. Detailed histopathologic examination of the bone marrow revealed marked decreased numbers of hematopoietic cell series, which are megakaryocytic, erythroid and myeloid series, and cellularity was 15%. Focal bone marrow necrosis was established with necrotic cells and fat necrosis. There were no phagocytic activity and necrotic cell debris around the necrotic focus. With these findings the

#### Tab. 1: Hematologic and immunologic data for the case.

|                       | Day of Admission | Day Three | Normal Range |
|-----------------------|------------------|-----------|--------------|
| Hematocrite           | 31               | 18        | 40–53        |
| MCV (fl)              | 81               | 80        | 80–95        |
| Hb (g/dL)             | 10               | 6         | 12–16        |
| RBC (M/mm³)           | 3.6              | 1.9       | 4.5–5.9      |
| WBC (M/mm³)           | 0.64             | 0.23      | 4.50–11.0    |
| PLT (K/mm³)           | 464.000          | 83.000    | 130.000–400.000 |
| Differential count (%)|                  |           |              |
| Neutrophils           | 1.92             | 1.95      | 40–72        |
| Lymphocytes           | 75.8             | 76.2      | 20–47        |
| Monocytes             | 20.7             | 22.2      | 3–8          |
| Eosinophils           | 0.17             | 1.26      | 0–8          |
| Fibrinogen (g/L)      | 4.30             | 4.30      |              |
| Partial thromboplastin time (s) | 56.2 | 30.7 | 26–36 |
| Prothrombin time      | 19.60            | 15.2      | 11–14        |
| INR                   | 1.9              | 1.40      | 0.85–1.15    |
| D-dimer (µg/ml)       | 0.72             |           | 0–10         |
| ESR (mm/h)            | 47               | 36        | 0–20         |
| Antinuclear antibodies| negative         |           |              |
| c-ANCA                | negative         |           |              |
| IL-1β (pg/ml)         | 5.00             |           | 0–5          |
| IL-2R (U/ml)          | 7310             |           | 223–710      |
| TNF-alpha (pg/ml)     | 15.8             |           | 0–8.10       |
| IL-6 (pg/ml)          | 266.00           |           | 0–9.7        |
| IL-8 (pg/ml)          | 3467             |           | 0–62         |
| IL-10 (pg/ml)         | 18.70            |           | 0–9.10       |

INR: International normalization ratio; IL: Interleukin; TNF: Tumor necrosis factor; CRP: C-reactive protein, ANCA: Antineutrophil cytoplasmic antibodies.
The patient received meropenem (Meronem®, AstraZeneca) 6 g/day and amikacin (Mikasin®, Abfar) 1 g/day before and after the diagnosis was established. Repeat blood testing on the second and third days of treatment showed that the neutropenia was becoming more severe. The patient developed progressive dysphagia. Endoscopic examination of the esophagus showed candidal esophagitis. To manage the bone marrow necrosis and associated clinical signs, we initiated treatment with granulocyte colony-stimulating factor (filgrastim, Neupogen®, Roche) 48 U/day, a 400 mg/kg bolus of intravenous I g (Endobulin S/D®, Baxter) to suppress cytokines, and amphotericin B (Fungizone®, Bristol-Myers Squibb) 100 mg/day. However, there was no response to treatment, and the patient died of cardiac arrest on the eighth day of hospitalization.

Fig. 1: Focal bone marrow necrosis (arrow)(H&E x100).

Discussion

The most prominent finding in this case was bone marrow necrosis. It has been reported that beside agranulocytosis, anemia and thrombocytopenia can be observed (90% and 80% respectively) in patients with bone marrow necrosis (11). In our case with agranulocytosis, thrombocytopenia developed following microangiopathic hemolytic anemia, which often occurs in the case of bone marrow necrosis (7,10). Necrosis of normal hematopoietic elements with gelatinous transformation has been described in patients with hematopoietological disease, adenocarcinoma, typhoid fever, systemic lupus erythematosus and tuberculosis (7,9,14,15). Other causes of marrow necrosis include typhoid fever, systemic lupus erythematosus and tuberculosis. The patient had no history of exposure to any other chemical or potential toxins.

There are two explanations for this patient’s dramatic illness. First, the condition may have been initiated by an infection that resulted in histiocytic proliferation and macrophage reactivation with excessive cytokine release. Epstein-Barr virus, cytomegalovirus, parvovirus B19, HIV-1 and -2, and herpes virus are the most common causes of histiocytic proliferation and inhibition of normal hematopoietic cells in adults (5). Epstein-Barr virus infection has a number of different oral manifestations. Affected patients frequently develop cervical lymphadenopathy and may exhibit palatal petechiae, but erosions and ulcers are not typical. Cytomegalovirus usually does not cause oral ulceration (3,5). In our case, the absence of any risk factors for HIV infection made this diagnosis unlikely. Further, all the serology results for viral disease were negative.

A second possible explanation is bone marrow necrosis, and elevated liver enzymes might have been unexplained toxic effects of the St. John’s wort the patient was taking for depression. Use of herbal preparations is increasing globally in parallel with alternative medicine. St. John’s wort is one of several herbal products used to treat depression (4,8). Previous studies have shown that Hypericum spp. have a number of biological activities. Hypericin, a derivative and the active compound of St. John’s wort, is known to have antioxidant and antimicrobial activity (2,6,13,17). Two reports suggest that St. John’s wort has anticancer activity, and that it may act by inducing apoptosis and inhibition of cell growth (19). Other research indicates that this herb may also affect drug metabolism by inducing the cytochrome P 450 enzyme (12). St. John’s wort has in vitro immunomodulatory effects on lymphocytes, and may alter cytokine release (10,18). The presence of high plasma cytokine levels in our case might reflect such immunological effects that can lead to bone marrow necrosis.

Herbal remedies are used throughout the world, and this report highlights a rare association of herb use and bone marrow necrosis. This case underlines the need for better assessment of the safety of these products.

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