Spectrum of antibacterial activity and mode of action of a novel tris-stilbene bacteriostatic compound

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The spectrum of activity and mode of action of a novel antibacterial agent, 135C, was investigated using a range of microbiological and genomic approaches. Compound 135C was active against Gram-positive bacteria with MICs for *Staphylococcus aureus* ranging from 0.12–0.5 μg/ml. It was largely inactive against Gram-negative bacteria. The compound showed bacteriostatic activity in time-kill studies and did not elicit bacterial cell leakage or cell lysis. Checkerboard assays showed no synergy or antagonism when 135C was combined with a range of other antibacterials. Multi-step serial passage of four *S. aureus* isolates with increasing concentrations of 135C showed that resistance developed rapidly and was stable after drug-free passages. Minor differences in the fitness of 135C-resistant strains and parent wildtypes were evident by growth curves, but 135C-resistant strains did not show cross-resistance to other antibacterial agents. Genomic comparison of resistant and wildtype parent strains showed changes in genes encoding cell wall teichoic acids. 135C shows promising activity against Gram-positive bacteria but is currently limited by the rapid resistance development. Further studies are required to investigate the effects on cell wall teichoic acids and to determine whether the issue of resistance development can be overcome.

Antimicrobial resistance (AMR) is one of the most problematic public health issues that society faces today. As an example, the prevalence of methicillin-resistant *Staphylococcus aureus* (MRSA) increased from 5% in 1980 to 50% of documented *S. aureus* infections in 20001. The increase in AMR is placing significant pressure on essential health care systems around the world2 as infections caused by resistant bacteria are associated with higher levels of morbidity and mortality and increased health care costs3. Furthermore, in 2016 more than 2800 people died from infectious diseases in Australia4, highlighting the need for effective antibiotic therapy as a critical component of reducing both morbidity and mortality associated with infectious diseases.

Naturally-occurring stilbenes are known for their diverse bioactivities5 including anti-cancer (phoyunbenes6, halophilol A7 and combretastatins8), anti-diabetic (rumexoid9) and antimalarial activity (stilbenes isolated from *Artocarpus integer*10). Many stilbene derivatives have also shown activity against bacteria11 including a compound isolated from *Combretum woodii* leaves12.

A previous study of a promising antibacterial stilbene compound found in silico led to triacid derivative 135C (Fig. 1). In this previous medicinal chemistry study, 135C was found to exhibit antibacterial activity13 against several Gram-positive bacteria in vitro, including methicillin-resistant *Staphylococcus aureus* (MRSA), at concentrations comparable to those required for existing antibiotics14. Our reference compound 135C contains a tri-carboxylic acid functionality and a tri-styrene motif, and its structural class is not seen in the current antimicrobial literature. Other natural stilbenes with antibacterial activity, such as resveratrol15, have been studied in detail but none contain this tri-styrene motif. Thus, given that this compound falls outside any known existing antibacterial classes, the benefits from developing it as a potential antibacterial agent are considerable, as there may be a reduced likelihood of observing antibacterial cross-resistance16 and it is possible that the compound has a novel mode of antibacterial action.
The aim of the current research was to continue investigations into the antibacterial activity of compound 135C. In particular, the spectrum of activity against a wide range of organisms was determined and the mode of action was investigated. The spectrum of antibacterial activity and synergy with conventional antibacterial agents was determined using broth microdilution techniques. Detection of the leakage of intracellular material that absorbs at 260 nm, lysis of cells and time-kill experiments were also performed to investigate potential modes of action. Lastly, the development of multi-step resistance to 135C was investigated by serial passage using four S. aureus strains. A genomic comparison of the resultant 135C-resistant mutant strains and wildtype parent strains was also performed.

Materials and Methods

Bacterial strains. Reference and clinical strains (n = 93) were obtained from the School of Biomedical Sciences at The University of Western Australia (UWA) and PathWest Laboratory Medicine WA. The Gram positive isolates (n = 71) comprised staphylococci (n = 40), streptococci (n = 20), micrococci (n = 10) and Clostridium difficile (n = 1). The Gram negative isolates (n = 22) included Moraxella spp. (n = 10), Escherichia coli (n = 3), Pseudomonas aeruginosa (n = 3), Prevotella bivia (n = 1) and Bacteroides spp. (n = 5).

Compounds. Compound 135C was synthesised as described previously and purity was confirmed by $^1$H and $^{13}$C NMR spectroscopy. Compound 135C was a yellow solid, and was stored under argon and protected from light at −20°C. Stock solutions of 135C in dimethyl sulfoxide (DMSO) were prepared fresh on each testing occasion and were diluted as required. The maximum concentration of DMSO present in bioactivity assays was 10%.

Susceptibility tests. Minimum inhibitory concentrations (MICs) and minimum bactericidal concentrations (MBCs) were determined using the broth microdilution methods recommended by the Clinical and Laboratory Standards Institute. Briefly, compound 135C was serially diluted two-fold in 100 µL volumes of sterile distilled water in a 96-well microtitre tray (Falcon, Becton Dickinson, USA). Inocula were prepared by culturing organisms overnight on blood agar, then suspending several colonies in 0.85% saline. Each suspension was inoculated with a 100 µL volume of inocula and microtitre trays were incubated at 37°C for 24 h, or 48 h for anaerobes, under the appropriate conditions. For a subset of Gram-negative bacteria, MICs were also determined.

Determination of antibacterial synergy. MICs of 135C in combination with a range of antibiotics with varying mechanisms of action were determined against S. aureus NCTC 6571 to investigate whether synergy or antagonism occurred. The antibiotics tested were ciprofloxacin, rifampicin, vancomycin, erythromycin, gentamicin and oxacillin. The checkerboard broth microdilution method was used as described previously. Briefly, each antibiotic was serially diluted two-fold in 50 µL volumes in a 96-well microtitre tray (Falcon, Becton...
Dickinson, USA), after which 50 μL of 135C at varying concentrations (ranging from 0.06–4 μg/ml) was added to each dilution series. The remainder of the assay was performed as described above. For each well of the microtitre plate that corresponded to a MIC, the fractional inhibitory concentration index (FIC) was determined using the equation: FIC = (MIC of drug A in combination/MIC of drug A alone) + (MIC of drug B in combination/MIC of drug B alone). The FICmin and FICmax were determined as the lowest and the highest FIC, respectively. The interaction was deemed synergistic if the FIC was < 0.5, additive if the FIC was between 0.5 and 1.0, indifferent if the FIC was between 1.0 and 2.0 and antagonistic if the FIC was > 2.0. This assay was performed in triplicate per antibacterial agent/135C combination.

**Cellular leakage and lysis.** Bacteriolysis and cell leakage assays were performed as previously described. Inocula were prepared from an overnight culture of S. aureus NCTC 6571 in Mueller-Hinton broth (MHB) by diluting to 0.5 McFarland in fresh MHB. Bacterial suspensions were then left untreated (untreated control) or were treated with 2, 32 (1 × MBC) and 320 μg/ml (10 × MBC) 135C. Treatments and the control were incubated at 35°C with shaking at 80 rpm. Samples were removed immediately after inoculation (time 0), and further samples were taken at 15, 30, 60, 120 and 240 min. Cell lysis was determined by measuring the optical density (OD) of each sample at 600 nm. The leakage of intracellular contents was determined by filtering samples (0.45 μm) to obtain cell-free filtrates and determining the OD260 at time zero and 240 min only. The OD of all samples was determined against the appropriate blank at the appropriate wavelength. Each assay was performed in triplicate and mean values were determined.

**Time-kill kinetics.** Bacterial killing assays were performed as previously described. Inocula were prepared as described for the cell leakage and lysis assays and were exposed to 135C at 0 (untreated control), 32 and 320 μg/ml of 135C in MHB. Treatments and controls were incubated at 35°C with shaking at 80 rpm and samples were removed at 0, 15, 30, 60, 120 and 240 min. Samples were immediately serially diluted ten-fold in 0.01 M phosphate buffered saline (PBS) at pH 7.0 and 10 μL aliquots were spot-inoculated in duplicate onto MHA. Agar plates were incubated at 37°C overnight before determining the viable count. The assay was conducted on two separate occasions.

**Serial passage with compound 135C to evaluate resistance development.** Four S. aureus strains (NCTC 10442, ATCC 29213 and clinical isolates RPH 15913 and 37845) were investigated for the development of multi-step resistance to 135C by serially passaging in increasing concentrations of the compound, using methods as previously described. Briefly, MICs were determined using the CLSI broth microdilution method as described above. The optical density of all wells was then determined (at 600nm) and serial passage was performed by removing the contents of the well with the highest concentration of 135C that had an OD600 value of approximately 0.1, and adding this culture to a freshly prepared dilution series in a 96-well microtitre tray. This was repeated at 24 h intervals for a total of 10 passages, for each of the four organisms. The compound concentration across each new passage was based on the MIC from the previous passage. At the end of the serial passage experiment, each organism was cultured from the highest turbid well onto blood agar (BA) and all were then passaged daily on drug-free media (BA) for 10 days. MICs of 135C were determined again after the third and tenth drug-free passages. After three drug-free passages, MICs were also determined for ciprofloxacin, erythromycin, gentamicin, oxacillin, rifampin, vancomycin, kanamycin and chloramphenicol to evaluate whether antibacterial cross-resistance was evident. The entire experiment was performed four times.

**Relative fitness of S. aureus isolates serially passaged with 135C.** The relative fitness of strains was evaluated as described previously. Briefly, serially-passaged S. aureus isolates and parent strains were cultured overnight on brain heart infusion agar (BHIA) plates, then 2–3 well-isolated colonies were inoculated into 15 ml of brain heart infusion broth (BHB) and incubated at 37°C with shaking at 80 rpm. After overnight growth, all cultures were diluted to an OD600 of 0.01 and 150 μl of this was added to 15 ml of BHB to give an inoculum concentration of approximately 10⁶ CFU/ml. Cultures were incubated at 37°C with shaking at 125 rpm for 24 h. The OD600 of cultures was determined every hour for 1–7 h and then at 24 h. The entire assay was performed four times.

**Genomic DNA preparation and whole genome sequencing.** Following subculture of S. aureus isolates on blood agar for 24 h, genomic DNA was extracted using a Gentra Puregene Yeast/Bact. Kit [Qiagen, Hilden, Germany] and multiplexed paired-end (PE) libraries were generated using standard Nextera XT protocols [Illumina, CA, USA]. Whole genome sequencing (WGS) of eight S. aureus strains (4 generated mutants, 4 wildtype) was performed using a MiSeq benchtop sequencer [Illumina], generating 300 bp PE reads. Sequence data for this study has been deposited in the European Nucleotide Archive (ENA) under study PRJEB21492 (sample accessions ERS1797685-ERS1797692). To examine genetic differences between wildtype parent strains and mutant progenitors, high-resolution single nucleotide variant (SNV) analysis was performed, as previously described. The highly annotated genome of S. aureus strain MRSA-252 [Genbank accession BX571856, multilocus sequence type (ST) 36] was used as a reference chromosome for read mapping (to a median depth of 45X across the 2.9 Mbp chromosome, range 26.9–52.6X) and SNV calling.

**Results**

**Susceptibility tests.** Susceptibility testing of a range of aerobic bacterial species showed that compound 135C was active against Gram-positive bacteria and also active against the Gram negative species Moraxella catarrhalis (Table 1). The most susceptible organism was S. aureus, with MICs of 135C ranging from 0.12–0.5 μg/ml. However, the MBCs for all species tested were ≥ 32 μg/ml (Table 1). Given this large difference between the MICs and MBCs, the activity of 135C was deemed to be bacteriostatic. When MICs against S. aureus NCTC 6571 were determined under anaerobic conditions, the MIC decreased from 0.5 to 0.004 μg/ml (seven doubling-dilutions). Susceptibility tests against a range of anaerobic bacteria showed an MIC of 16 μg/ml for Clostridium difficile, and a range of 32–256 μg/ml for Prevotella bivia and five Bacteroides spp. strains.
time zero, indicating that growth was inhibited but that cell death did not occur under these conditions.

Table 1. Minimum inhibitory and bactericidal concentrations of 135C. \(^1\)CNS, coagulase-negative = Staphylococcus hominis, S. warneri, S. saprophyticus, S. capitis and S. haemolyticus.

| Organism                  | MIC (µg/ml) | n  | Range | MIC\(_{50}\) | MIC\(_{90}\) | MBC (µg/ml) |
|---------------------------|-------------|----|-------|--------------|--------------|-------------|
| Micrococcus spp.          | 10          | 8–32 | 16    | >32          | >32          | >32         |
| Moraxella catarrhalis     | 10          | 1–32 | 8     | 32           | >32          | >32         |
| S. aureus (methicillin resistant) | 10 | 0.12–0.5 | 0.25 | 0.5 | 32–256 | |
| S. aureus (methicillin susceptible) | 10 | 0.25–0.5 | 0.25 | 0.5 | 32–128 | |
| Staphylococcus epidermidis | 10          | 1–32 | 2     | 4            | >32          | >32         |
| Staphylococcus spp. (CNS) | 10          | 0.5–32 | 4     | >32          | >32          | >32         |
| Streptococcus pneumoniae  | 10          | 1–16 | 4     | 16           | >32          | >32         |
| Streptococcus pyogenes    | 10          | 8–32 | 8     | 16           | >32          | >32         |

Table 2. Minimum inhibitory concentrations of 135C for Gram-negative bacteria in the presence and absence of PMBN. n.d. = no data. PMBN concentration 5 µg/ml\(^{-1}\).

| Organism                  | MIC (µg/ml) | | | | | |
|---------------------------|-------------|---|---|---|---|
| E. coli ATCC 43889        | >512        | 128 | 64 | 2 |
| E. coli NCTC 10538        | >512        | 32 | 256 | 4 |
| E. coli ATCC 25922        | >512        | 128 | 128 | 2 |
| P. aeruginosa NCTC 6749   | 512         | 256 | >512 | n.d. |
| P. aeruginosa ATCC 27853  | 512         | n.d. | 256 | 1 |
| P. aeruginosa ATCC 25668  | 512         | 512 | 512 | 8 |

**MICs in the presence of PMBN.** The activity of 135C against E. coli and P. aeruginosa was enhanced in the presence of PMBN (Table 2). The effect was more pronounced for E. coli than for P. aeruginosa, with the prior exhibiting at least a three-doubling-dilution increase in susceptibility to 135C in the presence of PMBN.

**Synergy experiments.** Antibacterial synergy data were generated using oxacillin, gentamicin, erythromycin, vancomycin, rifampicin and ciprofloxacin (Fig. 2). The \(\Sigma FIC\) was calculated to determine whether each combination of antibacterial agent/135C was considered synergistic, additive, indifferent or antagonistic (Table 3). Although there were several \(\Sigma FIC\) values that were less than 0.5, the results overall showed largely additive and indifferent activity for all six antibacterial agents, indicating negligible synergistic activity with 135C.

**Leakage, lysis and time-kill experiments.** Treatment of S. aureus NCTC 6571 with the highest concentration compound 135C (320 µg/ml, 10 × MBC) did not lead to any decrease in OD\(_{600}\), indicating that cell lysis did not occur. Similarly, experiments investigating whether exposure to 135C resulted in the leakage of intracellular contents did not show an appreciable change in OD\(_{590}\) after 2 h, indicating that the leakage of intracellular materials (nucleic acids) did not occur and that the cell membrane likely remained intact. Time-kill experiments with S. aureus NCTC 6571 did not show cell death at any concentration of 135C, with bacterial growth occurring in the presence of 2 µg/ml and 32 µg/ml similar to the drug-free control. At 320 µg/ml (10 × MBC), cell viable counts remained similar to time zero, indicating that growth was inhibited but that cell death did not occur under these conditions.

**Multi-step serial passage of S. aureus isolates with compound 135C.** Four S. aureus isolates were serially passaged with increasing concentrations of compound 135C to investigate the development of resistance. After the first passage, MIC values for all four isolates had increased by at least 3-fold (Fig. 3) After ten passages with increasing concentrations of compound 135C, MICs for all strains had increased from initial values of 0.12–0.25 µg/ml to 32–64 µg/ml. To investigate whether resistance was stable, isolates were subsequently passed 10 times without 135C, and MICs were again determined. For all four strains, the MICs after 10 drug-free passages were 32 µg/ml, which is similar to the final values (32–64 µg/ml) obtained after the 10 serial passages. This indicates that changes in susceptibility were stable, and are possibly due to a defined genetic mutation. The susceptibility of serially passaged S. aureus isolates was determined for a range of antibacterial agents selected to cover five major mode of action targets for antibiotics within bacterial cells. Cross-resistance to an antibacterial agent could indicate a shared, or at least similar, mode of antibacterial action. However, parent and serially passaged resistant strains did not differ in susceptibility (Table 4). Therefore, this part of the study did not offer any new information on the mode of action of compound 135C. Growth experiments conducted with both wildtype and serially-passaged 135C-resistant strains (Fig. 4) showed minor differences in growth patterns, with the greatest difference seen for S. aureus ATCC 29213.

**Genomic comparison of generated mutants and wildtypes.** High-quality SNVs were detected in all four S. aureus mutant strains, relative to their wildtype counterparts (Table 5). Three of the four 135C-resistant
strains showed nonsynonymous nucleotide changes (point mutations) in genes associated with teichoic acid bio-
synthesis (tagH, tagA, and tagG). These missense mutations resulted in codon changes and non-synonymous amino acid changes in the respective tag proteins.

Figure 2. Isobolograms showing interactions between 135C and antibacterial agents. The dotted gradient line represents the theoretical line of additivity.

| Antibiotic   | FIC_{min} | FIC_{max} | Interaction |
|--------------|-----------|-----------|-------------|
| Ciprofloxacin| 0.5       | 1.125     | Additive    |
| Erythromycin | 0.376     | 1.125     | Additive    |
| Gentamicin   | 0.375     | 1.125     | Additive    |
| Oxacillin    | 0.531     | 1.015     | Additive    |
| Rifampicin   | 0.375     | 1.125     | Additive    |
| Vancomycin   | 1         | 2.25      | Indifferent |

Table 3. Minimum and maximum Fractional Inhibitory Concentration (FIC) indices of antibacterial agent/135C combinations.
Discussion

The current study showed that the novel compound 135C was active largely against Gram-positive bacteria and confirmed the lack of activity observed previously against Gram-negative bacterial species. Compound 135C showed highest activity against S. aureus, and was slightly less active against other Gram positive organisms, for

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Table 4. Evaluation of S. aureus isolates passaged with 135C for cross-resistance to antibacterial agents. CHL, chloramphenicol; CIP, ciprofloxacin; ERY, erythromycin; GEN, gentamicin; KAN, kanamycin; OXA, oxacillin; RIF, rifampin; VAN, vancomycin. *Clinical isolate.
Table 5. Analysis of heterozygous sites in wildtype and mutant strains of *S. aureus*. WT, wildtype. MUT, mutant. MRSA-252 Genbank accession BX571856. *Clinical isolates. p. denotes amino acid change. †Base pair change on forward (plus) or reverse (minus) DNA strand.

reasons that remain to be determined. Poor activity against most Gram-negative bacteria may be due to a number of mechanisms including outer membrane impermeability and active efflux systems. It has been shown previously that the outer membrane permeabiliser PMBN interacts with the lipopolysaccharide of the outer membrane of Gram-negative bacteria, and PMBN has been used to sensitise different bacteria to many hydrophobic antibacterial agents such as novobiocin, erythromycin and clindamycin. PMBN alone has negligible activity and does not cause the leakage of periplasmic proteins from bacteria but allows the entry of hydrophobic antibacterial compounds into the cell by facilitating hydrophobic diffusion through the outer membrane. The activity of 135C was enhanced in the presence of PMBN, suggesting that the outer membrane has a role in limiting or preventing the entry of this compound into the cell. Further evidence of outer membrane involvement is shown by the susceptibility of the unusual Gram negative *M. catarrhalis* to 135C, which is likely due to the lack of long O-antigen polysaccharide chains in the lipopolysaccharide structure of its outer membrane, which are normally present in Gram-negative bacteria. The absence of these lipooligosaccharide chains results in the increased permeability of the bacterial envelope, which may contribute to the susceptibility of this organism to 135C.

When *S. aureus* species switch from aerobic to anaerobic growth, changes in gene expression occur, for example the presence of glycolytic enzymes, and comparatively low amounts of tricarboxylic acid cycle enzymes. The MIC of 135C against *S. aureus* NCTC 6571 decreased from 0.5 μg/ml to 0.004 μg/ml under anaerobic conditions. This suggests that under such conditions, the compound may be oxidised to a less active derivative, or that the compound may be metabolised differently under anaerobic conditions.

The mode of action of 135C against *S. aureus* was investigated using several different approaches. The first approach was to investigate effects against the cell wall and cell membrane, as these are two major targets for current antibacterial agents and experiments to quantify one or both are often performed to elucidate the mode of action of novel antibacterial agent. Unfortunately, the results from the cell leakage and lysis assays indicated that neither of these sites is likely to be major targets for 135C.

Subsequently, studies investigating antibacterial synergy were conducted. Synergy between different antibacterial compounds can be determined using the checkerboard method where multiple concentrations of two different compounds are tested in combination. There are many examples of antibacterial combinations that have been shown to be synergistic, and synergistic interactions have the potential to shed light on the mode of action of an antibacterial agent. Examples include compounds that are active against the cell wall enhancing the uptake of aminoglycosides, as well as combinations of agents active on the cell wall. Compound 135C was tested in combination with antibacterial agents from different classes, and in this study only compound combinations that may contribute to the susceptibility of this organism to 135C were investigated.

Investigation of the development of resistance to 135C by *S. aureus* isolates showed rapid and stable resistance, which limits the potential usefulness of the compound. Often, the acquisition of bacterial resistance is accompanied by a fitness cost to bacteria, typically evident as a reduction in growth rates in vivo and in vitro, decreased invasiveness or a lower cell density. Data from the current study indicated only minor differences in the fitness of parent and serially passaged strains, suggesting that fitness was not substantially altered.

Genomic comparison of the four 135C-resistant mutants to their wildtype parent strains provided interesting insight into a potential relationship between 135C and wall teichoic acids (WTAs) (Table 5). Of the four 135C-resistant strains, three contain mutations within the teichoic acid glycerol (tag) genes, *tagH*, *tagA*, and *tagG*. These three genes are involved in protein products associated with teichoic acid biosynthesis (*tagA*), and their ATP-binding cassette (ABC) transporter subunits (*tagH* and *tagG*). The changes identified in the 135C-resistant mutants suggest that WTAs may be the target site of 135C, or that the compound may disrupt the regular function of WTAs. This finding is also helps to explain the lack of activity against Gram negative bacteria, as teichoic acids are absent in Gram negative bacteria.

Of note was that the changes in all three *tag* genes were nonsynonymous substitutions. These missense mutations (single nucleotide alterations) result in codon changes that lead to different amino acids. These changes occurred in only three of the four strains, with these three strains all being methicillin-resistant while the fourth strain is methicillin susceptible. The amino acid changes in *S. aureus* 3784546C and NCTC 10442 both involved an arginine substitution. In both cases, the substitution led to amino acids with shorter (less flexible), more hydrophobic and non-polar side chains (glycine and isoleucine). For *S. aureus* RPH 15913, the mutation resulted in an amino acid change from asparagine to lysine. The lysine side chain contains a longer (more flexible) and
less hydrophobic moiety than asparagine, which is in contrast to what was observed in the previous two strains. However, for all three strains the mutation resulted in a less polar amino acid. The polarity of the amino acid changes may be a significant factor if WTAs are the binding sites of 135C. Structurally, 135C is a symmetrical molecule with three polar carboxylic acid moieties. It is therefore plausible that a change in the polarity of an amino acid in WTAs may have an effect on the ability of the acid moieties within this compound to hydrogen bond to a potential binding site. Therefore, from a structural perspective, the changes in the amino acid side chains may be significant if the tag genes are indeed the binding sites of 135C.

It was interesting to note that S. aureus ATCC 29213 (methicillin-susceptible) was the only strain without a change in a tag gene. A plausible explanation may be that this strains’ WTA genetic profile is different to that of the methicillin-resistant strains. The mode of action of the β-lactam antibiotic methicillin involves the inhibition of penicillin-binding proteins (PBPs), which are essential in the synthesis of the peptidoglycan layer that makes up the cell wall. Since WTAs are embedded in the peptidoglycan layer, the resistance mechanism of MRSA may have also affected these sites. It is also important to note that WTAs are classified as virulence factors and, as such, 135C may have a downstream effect on the ability of Gram-positive bacteria to proliferate in host organisms.

To summarise, our study revealed 135C to be a bacteriostatic agent with activity against a range of Gram-positive pathogens. The compound was largely inactive against Gram-negative bacteria, which may be due, in part, to the impermeability of the Gram-negative outer membrane and absence of teichoic acids within the cell wall. When combined with conventional antibiotics 135C did not show significant synergistic nor antagonistic activity. The absence of leakage of bacterial cell contents and absence of cell lysis indicate that 135C is unlikely to have a target within these sites. Resistance studies showed that S. aureus rapidly developed stable resistance to 135C, after as little as a single passage. Comparison of the whole genome sequence of the 135C-resistant mutants to their wildtype counterparts revealed a change in teichoic acid-associated tag genes. Changes in these genes indicate a possible interaction between 135C and WTAs as part of its mode of action. Additional studies with bacterial strains that have specific WTA mutations may help to clarify the role of WTA in susceptibility to 135C. This may also offer insight into why S. aureus in particular, is highly susceptible compared to other Gram positive organisms. In conclusion, lack of activity against Gram negative pathogens, absence of bactericidal activity and the rapid development of resistance all significantly limit the potential usefulness of the compound. It remains possible that additional chemical modification of the compound may lead to further novel compounds with improved efficacy, broader spectrum of activity and reduced resistance capacity.

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Acknowledgements
NYM and DRK were supported by Australia Research Training Program Scholarships.

Author Contributions
N.Y.M. and K.A.H. wrote the main manuscript text and prepared Figures 1–4 and Tables 1–4. D.R.K. prepared Table 5. N.Y.M., D.R.K., K.A.H., S.G.S., A.J.M. and T.V.R. all reviewed the manuscript.

Additional Information
Competing Interests: The authors declare no competing interests.

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