Regulation of capillary hemodynamics by $K_{\text{ATP}}$ channels in resting skeletal muscle

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Abstract

ATP-sensitive K$^+$ channels ($K_{\text{ATP}}$) have been implicated in the regulation of resting vascular smooth muscle membrane potential and tone. However, whether $K_{\text{ATP}}$ channels modulate skeletal muscle microvascular hemodynamics at the capillary level (the primary site for blood-myocyte O$_2$ exchange) remains unknown. We tested the hypothesis that $K_{\text{ATP}}$ channel inhibition would reduce the proportion of capillaries supporting continuous red blood cell (RBC) flow and impair RBC hemodynamics and distribution in perfused capillaries within resting skeletal muscle. RBC flux ($f_{\text{RBC}}$), velocity ($V_{\text{RBC}}$), and capillary tube hematocrit (Hct$_{\text{cap}}$) were assessed via intravital microscopy of the rat spinotrapezius muscle ($n=6$) under control (CON) and glibenclamide (GLI; $K_{\text{ATP}}$ channel antagonist; 10 µM) superfusion conditions. There were no differences in mean arterial pressure (CON: 120 ± 5, GLI: 124 ± 5 mmHg; $p > 0.05$) or heart rate (CON: 322 ± 32, GLI: 337 ± 33 beats/min; $p > 0.05$) between conditions. The %RBC-flowing capillaries were not altered between conditions (CON: 87 ± 2, GLI: 85 ± 1%; $p > 0.05$). In RBC-perfused capillaries, GLI reduced $f_{\text{RBC}}$ (CON: 20.1 ± 1.8, GLI: 14.6 ± 1.3 cells/s; $p < 0.05$) and $V_{\text{RBC}}$ (CON: 240 ± 17, GLI: 182 ± 17 µm/s; $p < 0.05$) but not Hct$_{\text{cap}}$ (CON: 0.26 ± 0.01, GLI: 0.26 ± 0.01; $p > 0.05$). The absence of GLI effects on the %RBC-flowing capillaries and Hct$_{\text{cap}}$ indicates preserved muscle O$_2$ diffusing capacity (DO$_2$m). In contrast, GLI lowered both $f_{\text{RBC}}$ and $V_{\text{RBC}}$ thus impairing perfusive microvascular O$_2$ transport (Qm) and lengthening RBC capillary transit times, respectively. Given the interdependence between diffusive and perfusive O$_2$ conductances (i.e., %O$_2$ extraction×DO$_2$m/Qm), such GLI alterations are expected to elevate muscle %O$_2$ extraction to sustain a given metabolic rate. These results support that $K_{\text{ATP}}$ channels regulate capillary hemodynamics and, therefore, microvascular gas exchange in resting skeletal muscle.

KEYWORDS

ATP-sensitive K$^+$ channel, blood flow, intravital microscopy, microcirculation, red blood cell
INTRODUCTION

The skeletal muscle capillary vascular bed provides the largest surface area for gas and substrate exchange within the body (Hirai et al., 2019; Poole et al., 2011; Poole, 2019). Regulation of skeletal muscle capillary hemodynamics is mediated primarily at the arteriolar level (Joyner & Casey, 2015; Kindig & Poole, 2001; Laughlin et al., 2012; Segal, 2005). Several complex and often interacting mechanisms are responsible for alterations in arteriolar resistance and thus vascular conductance. Among these processes, K⁺ channels constitute the dominant ion conductance of the vascular smooth muscle cell determining membrane potential, contractile activity, and thus vascular tone (Foster & Coetzee, 2016; Jackson, 2017; Tyckoki et al., 2017).

ATP-sensitive K⁺ (K<sub>ATP</sub>) channels constitute one of the, at least, five distinct classes of K⁺ channels regulating vascular smooth muscle function (Foster & Coetzee, 2016; Jackson, 2017; Tyckoki et al., 2017). Inhibition (closing) of K<sub>ATP</sub> channels decreases K⁺ efflux leading to depolarization of the smooth muscle cell. Voltage-gated Ca<sup>2+</sup> channels then transduce membrane depolarization into increased Ca<sup>2+</sup> influx promoting vascular muscle contraction (i.e., vasoconstriction). In vitro and in vivo studies have thus used pharmacological K<sub>ATP</sub> channel inhibition to evaluate its potential role in setting vascular tone and, consequently, tissue perfusion. Sulfonylureas such as glibenclamide (GLI), which are employed clinically in the treatment of non-insulin-dependent diabetes (Montvida et al., 2018), represent the most frequently used class of K<sub>ATP</sub> channel inhibitors. To date, however, there is a lack of agreement regarding the role of K<sub>ATP</sub> channels in the regulation of resting vascular tone and blood flow within skeletal muscle (Foster & Coetzee, 2016; Jackson, 2017; Tyckoki et al., 2017). K<sub>ATP</sub> channel inhibition with GLI has produced conflicting effects on resting skeletal muscle arteriolar diameter (Hammer et al., 2001; Hodnett et al., 2008; Jackson, 1993; Lu et al., 2013; Murrant & Sarelius, 2002; Saito et al., 1996; Xiang & Hester, 2009) and bulk blood flow (Bank et al., 2000; Colburn, Holdsworth, et al., 2020; Duncker et al., 2001; Farouque & Meredith, 2003a,b; Holdsworth et al., 2015; Vanelli & Hussain, 1994). Importantly, the functional role of K<sub>ATP</sub> channels within the skeletal muscle capillary network (i.e., the primary site for blood-myocyte O₂ exchange) remains to be determined. Resolution of the mechanisms modulating capillary red blood cell (RBC) hemodynamics has direct relevance to transcapillary O₂ flux and cellular energetic status.

The purpose of this study was to evaluate the regulation of capillary RBC hemodynamics by K<sub>ATP</sub> channels in resting skeletal muscle. Using intravital microscopy techniques combined with the rat spinotrapezius preparation, we tested the hypothesis that acute local K<sub>ATP</sub> channel inhibition with GLI would impair key determinants of capillary diffusive and perfusive O₂ conductances. Specifically, GLI was anticipated to reduce the proportion of capillaries supporting continuous RBC flow and impair microvascular hemodynamics and distribution (i.e., RBC flux, velocity, and capillary tube hematocrit) in perfused capillaries within resting skeletal muscle. Confirmation of these hypotheses would support a role for K<sub>ATP</sub> channels in the regulation of skeletal muscle capillary hemodynamics (and thus microvascular gas exchange) under resting conditions.

METHODS

Intravital microscopy was used to evaluate the in vivo spinotrapezius muscle microcirculation in healthy young male Sprague-Dawley rats (n = 6; ~3–4 months old; 386 ± 26 g). Rats were maintained in accredited facilities (Association for the Assessment and Accreditation of Laboratory and Animal Care) under a 12:12 h light–dark cycle with food and water provided ad libitum. All experimental procedures and protocols were approved by the Institutional Animal Care and Use Committee of Kansas State University and followed the guidelines established by the National Institutes of Health.

Surgical procedures

2.1.1 | Anesthesia and catheter placement procedures

All rats were anesthetized initially with a 5% isoflurane-O₂ mixture and maintained subsequently on 2–3% isoflurane-O₂ (Butler Animal Health Supply). Anesthetized rats were kept on a heating pad to maintain core temperature at ~37–38°C as measured via a rectal probe. The right carotid artery was cannulated (PE-10 connected to PE-50; BD IntraMedic Polyethylene Tubing) for continuous measurements of mean arterial pressure and heart rate (MAP and HR, respectively; PowerLab; ADInstruments). The caudal artery was cannulated (PE-10 connected to PE-50; BD IntraMedic Polyethylene Tubing) for continuous measurements of mean arterial pressure and heart rate (MAP and HR, respectively; PowerLab; ADInstruments). The caudal artery was cannulated (PE-10 connected to PE-50; BD IntraMedic Polyethylene Tubing) for continuous measurements of mean arterial pressure and heart rate (MAP and HR, respectively; PowerLab; ADInstruments). The caudal artery was cannulated (PE-10 connected to PE-50; BD IntraMedic Polyethylene Tubing) for continuous measurements of mean arterial pressure and heart rate (MAP and HR, respectively; PowerLab; ADInstruments).
allows for clear visualization of capillary structure and hemodynamics via light transmission microscopy (Gray, 1973; Poole et al., 1997); b) it can be exteriorized without neural or substantial vascular disruption (Bailey et al., 2000; Gray, 1973; Poole et al., 1997); c) a physiological sarcomere length can be maintained throughout the experimental protocol (thereby preventing muscle overstretching and adverse microcirculatory effects) (Poole et al., 1997; Welsh & Segal, 1996); and d) its mixed fiber-type composition and oxidative capacity are similar to those of the human quadriceps (Delp & Duan, 1996; Leek et al., 2001), thus representing a useful analog of human locomotor muscle. Following catheter placement procedures, the left spinotrapezius was exposed and exteriorized as described previously (Bailey et al., 2000; Gray, 1973; Poole et al., 1997) with minimal fascial removal to limit tissue damage and microcirculatory disturbances (Mazzoni et al., 1990). Briefly, the caudal end of the muscle was isolated from its origin and sutured at equidistant points to a horseshoe-shaped manifold. The rat was then placed on a water circulation-heated (38°C) Lucite platform and the manifold secured with the ventral aspect of the muscle reflected upwards for microscopic observation. The preparation was frequently superfused with Krebs–Henseleit bicarbonate-buffered solution (2.0 mM CaCl2, 2.4 mM MgSO4, 4.7 mM KCl, 22 mM NaHCO3, 131 mM NaCl; pH 7.4; equilibrated with 5% CO2 and 95% N2 at ~38°C) and exposed surrounding tissue covered with Saran wrap (Dow Brands). As noted earlier, the spinotrapezius was maintained at physiological sarcomere length (2.6 ± 0.1 µm) to prevent stretch-induced reductions in capillary blood flow (Poole et al., 1997; Welsh & Segal, 1996).

2.2 | Experimental protocol

2.2.1 | Intravital video microscopy

Microcirculatory fields located midway between arteriolar and venular ends within the mid-caudal region of the spinotrapezius that provided optimal clarity were selected randomly for the study. Microcirculatory images were obtained via a bright-field microscope (Nikon Eclipse E600-FN) equipped with a non-contact illuminated lens (x40, numerical aperture 0.8) and viewed in real time on a high-resolution color monitor (Sony Trinitron PVM-1954Q) under a final magnification of x1,184 (as confirmed by initial calibration of the system with a stage micrometer; MA285; Meiji Techno). Images were recorded by a video camera (JVC KY-F55B) and stored on a computer for subsequent offline analyses.

2.2.2 | Experimental design

Once the spinotrapezius muscle was positioned on the platform, a quiescent period of at least 15 min was allowed before any data were acquired. Intravital microscopy recordings were then made under two separate superfusion conditions: control (Krebs–Henseleit; CON) and KATP channel inhibition (glibenclamide; GLI). GLI was the last treatment due to its long-lasting effects (Thomas et al., 1997) and the possibility of incomplete washout with Krebs–Henseleit. All superfusion solutions were maintained at approximately 38°C. The spinotrapezius was superfused with each solution for 3 min (average flow rate of 1 ml/min) followed by a 30 min equilibration period as employed by previous microcirculatory studies (Hodnett et al., 2008; Jackson, 1993; Lu et al., 2013; Murrant & Sarelius, 2002; Saito et al., 1996). Microcirculatory fields were chosen randomly from each rat (based on clear visualization of sarcomeres, fibers, and capillaries) and each recorded for ~1–1.5 min. At the end of the experimental protocol, rats were killed with intra-arterial pentobarbital sodium overdose (100 mg/kg) followed by pneumothorax.

2.2.3 | Acute local KATP channel inhibition

The pharmacological sulphonylurea derivative GLI (494 g/mol; 5-chloro-N-[4-(cyclohexylureidosulfonyl)phenethyl]-2-methoxybenzamide; Sigma-Aldrich; St. Louis, MO) was used to inhibit KATP channels via superfusion (topical application) of the spinotrapezius muscle. GLI stock solutions were made fresh daily with the solvent dimethyl sulfoxide (DMSO) and diluted with control (Krebs–Henseleit) superfusate. The final working superfusate had a GLI concentration of 10 µM and contained <0.01% DMSO. This GLI concentration was based on previous microcirculatory research using similar drug delivery methods with the rat spinotrapezius and hamster cheek pouch and cremaster muscles (Cohen & Sarelius, 2002; Hammer et al., 2001; Hodnett et al., 2008; Jackson, 1993; Lu et al., 2013; Murrant & Sarelius, 2002; Saito et al., 1996; Xiang & Hester, 2009). Previous studies have shown that ≤0.05% DMSO solutions have no significant effects on resting skeletal muscle arteriolar diameter or reactivity (Cohen et al., 2000; Jackson, 1993) or KATP channel currents in the absence or presence of ATP (Mele et al., 2014; Tricarico et al., 2006). Recent reports from our laboratory have shown no differences in arterial blood PO2, PCO2, O2 saturation, hematocrit, pH, [lactate] or [glucose] following superfusion of the rat spinotrapezius muscle with KATP channel agonists (pinacidil; 5 mg/kg) and antagonists (GLI; 5 mg/kg) (Holdsworth et al., 2017).
### 2.3 Analysis of muscle capillary hemodynamics

Within each preparation, only those microcirculatory fields with the best overall clarity were selected for further examination via frame-by-frame techniques (30 frames/s; Dartfish) as described previously (Kindig et al., 1999, 2002; Poole et al., 1997). Sarcomere length was determined from sets of 10 consecutive in-register sarcomeres (i.e., distance between 11 consecutive A-bands) measured in parallel to the muscle fiber longitudinal axis. This procedure was repeated 3 times where sarcomeres were visible to obtain a mean sarcomere length for each field. Comparisons between control and GLI conditions were made within the same microcirculatory field for each rat and, whenever possible, between the same capillaries (which occurred in ~47% of cases; i.e., 14 of 30 vessels). The percentage of RBC-perfused vessels was established as (no. of capillaries supporting RBC flow/total no. of visible capillaries per field) × 100. Vessels demonstrating impeded or stopped flow (i.e., stationary or no visible RBC flow) for ≥10 s were regarded as non-flowing capillaries. Five capillaries supporting continuous RBC flow were selected randomly for hemodynamics analysis from each microcirculatory field based on clarity. For all capillaries in which hemodynamics was evaluated and where the capillary endothelium was clearly visible on both sides of the lumen, capillary luminal diameter (d_{cap}) was measured at two random sites per capillary (2–3 measurements/site). RBC velocity (V_{RBC}) was determined by following the RBC path length over several frames and for the maximum capillary length over which the cells remained in crisp focus. RBC flux (f_{RBC}) was measured by counting the number of cells in a capillary passing an arbitrary point over several frames. These measurements were performed three times per capillary. For each capillary in which hemodynamics data were measured, capillary tube hematocrit (Hct_{cap}) was calculated as follows:

\[
\text{Hct}_{\text{cap}} = \frac{\text{vol}_{\text{RBC}} \times f_{\text{RBC}}}{\pi \times \left(\frac{d_{\text{cap}}}{2}\right)^2 \times V_{\text{RBC}}} \tag{1}
\]

where vol_{RBC} is RBC volume and assumed to be 61 µm^3 (Altman & Dittmer, 1974) and capillaries were approximated as circular in cross section (Desjardins & Duling, 1990; Kindig et al., 1998).

### 2.4 Statistical analyses

Statistical analyses were performed using a commercially available software package (SigmaPlot 11.2; Systat Software). Data distribution was assessed via the Shapiro–Wilk test for normality. Central hemodynamics (MAP and HR) and spinotrapezius muscle capillary structure (d_{cap}) and hemodynamics (%RBC-flowing capillaries, f_{RBC}, V_{RBC}, and Hct_{cap}) data were compared between control and GLI conditions using paired Student’s t-tests. Coefficients of variation were calculated as (SD/mean) × 100 and compared between control and GLI conditions using the methods described by Forkman (2009). Pearson’s product-moment correlations and linear regression analyses were used to examine relationships between f_{RBC} and V_{RBC}. Linear regression slope comparisons were performed as described by Zar (1984). Significance was accepted at p < 0.05. Data are reported as mean ± SE.

### 3 RESULTS

#### 3.1 Central hemodynamics

As expected based on the topical drug delivery method employed herein (i.e., local superfusion of the spinotrapezius muscle), no differences in MAP (CON: 120 ± 5, GLI: 124 ± 5 mmHg; p > 0.05) or HR (CON: 322 ± 32, GLI: 337 ± 33 beats/min; p > 0.05) were observed between conditions.

#### 3.2 Spinotrapezius muscle capillary structure and hemodynamics

The percentage of capillaries supporting continuous RBC flow was not altered between conditions (Figure 1; p > 0.05). Capillaries subjected to hemodynamics analysis supported RBC flow prior to, and continued to do so, following K_{ATP} channel inhibition...
inhibition (i.e., during CON and GLI, respectively). In these RBC-flowing capillaries, GLI reduced both \( f_{\text{RBC}} \) and \( V_{\text{RBC}} \) (Figures 2 and 3; \( p < 0.05 \) for both) but not Hct\(_{\text{cap}}\) (CON: 0.26 ± 0.01, GLI: 0.26 ± 0.01; \( p > 0.05 \)). GLI-induced alterations in muscle capillary hemodynamics occurred in the absence of changes in \( d_{\text{cap}} \) between conditions (CON: 4.9 ± 0.1, GLI: 4.9 ± 0.1 µm; \( p > 0.05 \)). Frequency histograms of capillary \( f_{\text{RBC}} \) and \( V_{\text{RBC}} \) are shown in Figure 3. Microvascular blood flow heterogeneity, as evaluated by the individual capillary coefficient of variation, was not different between conditions with respect to \( f_{\text{RBC}} \) (CON: 61.8, GLI: 57.4%; \( p > 0.05 \)), \( V_{\text{RBC}} \) (CON: 49.4, GLI: 47.8%; \( p > 0.05 \)) or Hct\(_{\text{cap}}\) (CON: 23.6, GLI: 22.3%; \( p > 0.05 \)). Similar results were found when comparing heterogeneity between microvascular fields in terms of \( f_{\text{RBC}} \) (CON: 21.4, GLI: 21.6%; \( p > 0.05 \)), \( V_{\text{RBC}} \) (CON: 17.4, GLI: 22.9%; \( p > 0.05 \)), and Hct\(_{\text{cap}}\) (CON: 13.5, GLI: 11.1%; \( p > 0.05 \)).

Consistent with the lack of changes in Hct\(_{\text{cap}}\) with GLI (and the relationship described in Eq. 1 above), no significant differences were observed in the slope of the \( f_{\text{RBC}}/V_{\text{RBC}} \) relationship between conditions (individual capillaries; CON: 8.75, GLI: 9.44; individual muscles; CON: 7.46, GLI: 12.25; \( p > 0.05 \) for both). Therefore, Figure 4 presents linear correlations and regression analyses for the combined CON and GLI data. Capillary \( f_{\text{RBC}} \) and \( V_{\text{RBC}} \) were significantly correlated.

**FIGURE 2** Capillary red blood cell flux (top panel) and velocity (bottom panel) during control (CON; \( n = 6 \)) and K\(_{\text{ATP}}\) channel inhibition (glibenclamide, GLI; \( n = 6 \)) conditions. Dashed lines represent individual muscle data. *\( p < 0.05 \) vs. CON

**FIGURE 3** Relative frequency histograms of capillary red blood cell flux (top panel) and velocity (bottom panel) during control (CON; \( n = 6 \)) and K\(_{\text{ATP}}\) channel inhibition (glibenclamide, GLI; \( n = 6 \)) conditions. Arrows show mean values. *\( p < 0.05 \) vs. CON
This investigation examined, for the first time, the regulation of capillary hemodynamics by K<sub>ATP</sub> channels in resting skeletal muscle. Superfusion of the rat spinotrapezius with the sulfonylurea GLI was used to locally inhibit K<sub>ATP</sub> channels in vivo. The principal novel findings are as follows: a) inconsistent with our hypothesis, GLI did not reduce the proportion of capillaries supporting continuous RBC flow or Hct<sub>cap</sub>; and b) consistent with our hypothesis, GLI lowered microvascular blood flow (i.e., both f<sub>RBC</sub> and V<sub>RBC</sub>) in perfused capillaries. These data suggest that K<sub>ATP</sub> channels, via modulation of arteriolar vascular conductance, regulate microvascular hemodynamics at the capillary level (the primary site for blood-myocyte O<sub>2</sub> diffusion) (Hirai et al., 2019; Poole et al., 2011; Poole, 2019) in resting skeletal muscle.

4 | DISCUSSION

This investigation examined, for the first time, the regulation of capillary hemodynamics by K<sub>ATP</sub> channels in resting skeletal muscle. Superfusion of the rat spinotrapezius with the sulfonylurea GLI was used to locally inhibit K<sub>ATP</sub> channels in vivo. The principal novel findings are as follows: a) inconsistent with our hypothesis, GLI did not reduce the proportion of capillaries supporting continuous RBC flow or Hct<sub>cap</sub>; and b) consistent with our hypothesis, GLI lowered microvascular blood flow (i.e., both f<sub>RBC</sub> and V<sub>RBC</sub>) in perfused capillaries. These data suggest that K<sub>ATP</sub> channels, via modulation of arteriolar vascular conductance, regulate microvascular hemodynamics at the capillary level (the primary site for blood-myocyte O<sub>2</sub> diffusion) (Hirai et al., 2019; Poole et al., 2011; Poole, 2019) in resting skeletal muscle.

4.1 | Intravital microscopy preparation

Surgical exteriorization of the rat spinotrapezius muscle was performed as described previously (Bailey et al., 2000; Gray, 1973; Poole et al., 1997) with minimal fascial disturbance to curtail tissue damage and related microcirculatory consequences. Accordingly, previous studies from our laboratory indicate that the surgical exteriorization requisite for transmission intravital microscopy as performed herein does not impair the microvascular integrity or responsiveness of the spinotrapezius muscle (Bailey et al., 2000).

In the present investigation, spinotrapezius sarcomere length was set at physiological values (2.6 ± 0.1 µm) to prevent stretch-induced capillary luminal diameter reductions (Poole et al., 1997) and/or sympathetically mediated decreases in arteriolar blood flow (Welsh & Segal, 1996; cf. Kindig & Poole, 2001). Capillary structure (d<sub>cap</sub>) and hemodynamics (%RBC-flowing vessels, f<sub>RBC</sub>, V<sub>RBC</sub>, and Hct<sub>cap</sub>) data obtained herein during the control condition are consistent with those published previously in the resting rat spinotrapezius and hamster cheek pouch and cremaster muscles (Copp et al., 2009; Kano et al., 2005; Kindig et al., 1998, 1999, 2002; Kindig & Poole, 2001; Poole et al., 1997; Richardson et al., 2003; Russell et al., 2003; Sarelius & Duling, 1982). Our data are also consistent with the well-documented lower than systemic Hct<sub>cap</sub> values found in resting skeletal muscle (Poole et al., 2011; Poole, 2019). Based on both theoretical and empirical studies, potential mechanisms for the systemic versus capillary tube hematocrit difference include a) the presence of the endothelial surface layer (i.e., the often termed “glycocalyx”); b) the Fahraeus effect; and c) plasma skimming at arteriolar bifurcations (Desjardins & Duling, 1990; Ellsworth et al., 2009; Pries et al., 1986; Secomb et al., 1998).

As reviewed recently (Poole et al., 2011; Poole, 2019), a compelling body of evidence demonstrates that the vast majority of skeletal muscle capillaries support continuous RBC perfusion at rest. Our data are consistent with this notion showing that ~85% of capillaries supported continuous f<sub>RBC</sub> during both CON and GLI conditions (Figure 1).

4.2 | Local K<sub>ATP</sub> channel inhibition with GLI superfusion

As expected, GLI-induced alterations in microvascular hemodynamics (f<sub>RBC</sub> and V<sub>RBC</sub>; Figure 2) occurred in the absence of changes in d<sub>cap</sub> between conditions. This suggests...
increased vascular resistance at sites upstream of the capillary bed consistent with the well-established arteriolar control of skeletal muscle blood flow (Joyner & Casey, 2015; Kindig & Poole, 2001; Laughlin et al., 2012; Segal, 2005). Our data are thus in agreement with some, but not all, reports of decreased arteriolar diameter (Hammer et al., 2001; Hodnett et al., 2008; Jackson, 1993; Lu et al., 2013; Murrant & Sarelius, 2002; Saito et al., 1996; Xiang & Hester, 2009) and bulk blood flow (Bank et al., 2000; Colburn, Weber, et al., 2020; Duncker et al., 2001; Faroqule & Meredith, 2003a,b; Holdsworth et al., 2015; Vannelli & Hussain, 1994) following KATP channel inhibition in the resting skeletal muscle. Potential reasons for this discrepancy include species differences, experimental models, GLI doses, KATP channel density distribution, compensatory vasodilation by redundant pathways, muscle fiber type, and arteriolar size and branch order. Although providing an invaluable framework for understanding the control of vascular tone and tissue perfusion, previous measurements of arteriolar diameter and bulk blood flow responses with GLI presented no information concerning the distribution of microvascular flow within the capillary network. Moreover, it was not known whether KATP channel inhibition would impact uniformly RBC-perfused capillaries or whether the proportion of RBC-perfused capillaries could be reduced. Evaluation of capillary hemodynamics within those vessels supporting continuous RBC flow is critical to identifying how KATP channels may modulate blood-myocyte O2 and substrate exchange. This is especially true when considering that the capillary network provides the largest surface area for gas and substrate exchange within the skeletal muscle microcirculation (Hirai et al., 2019; Poole et al., 2011; Poole, 2019).

The potential for KATP channels to modulate skeletal muscle O2 exchange must be considered in light of the interdependence between diffusive and perfusive conductances setting fractional O2 extraction (Roca et al., 1992):

$$\%\text{O}_2\text{ extraction} = 1 - e^{-\frac{\text{DO}_2 m}{\beta \text{Q}_m}}$$  \hspace{1cm} (2)

where DO2m is muscle O2 diffusing capacity, $\beta$ is the slope of the O2 dissociation curve in the physiologically relevant range, and Qm is muscle blood flow. Theoretical models of skeletal muscle O2 exchange indicate that DO2m is determined by structural (capillary-to-fiber ratio, capillary length, and $d_{cap}$) and functional ($Hct_{cap}$ at constant arterial O2 saturation) elements (Federspiel & Popel, 1986; Groebe & Thews, 1990). Capillary-to-fiber ratio, capillary length, and $\beta$ are not expected to be affected by acute GLI treatment as employed herein. Moreover, we observed no differences in $d_{cap}$ or $Hct_{cap}$ between CON and GLI. It thus seems that KATP channels do not modulate significantly DO2m in resting skeletal muscle. On the other hand, the current GLI-induced reductions in $f_{RBC}$ (which largely dictates Qm within the microcirculation) (Berg & Sarelius, 1996) are anticipated to elevate the $\text{DO}_2 m/\text{Q}_m$ ratio within RBC-perfused capillaries and, therefore, necessitate higher muscle fractional O2 extraction for a given metabolic rate according to Eq. 2 above. Importantly, the functional consequence of lowered $f_{RBC}$ with GLI (Figure 2, top panel) is impaired O2 delivery per capillary (QO2cap). Disregarding the small amount of O2 dissolved in plasma (and, therefore, its relevance to capillary gas exchange), QO2cap can be estimated as the product of $f_{RBC}$, the hemoglobin content per RBC (17 $\times$ 10−12 g per RBC) (Altman & Dittmer, 1974) and the O2 carrying capacity of hemoglobin (at 89% saturation, assuming arterial PO2 = 80 mmHg in anesthetized rats) as described previously (Kindig et al., 1998):

$$\text{QO}_2\text{cap} = f_{RBC} \times (\text{g Hb} \times \text{RBC}^{-1}) \times (1.34 \times \text{ml O}_2 \times \text{g Hb}^{-1} \times 0.89)$$  \hspace{1cm} (3)

Using the equation above, KATP inhibition with GLI superfusion of the resting spinotrapezius herein reduced mean QO2cap by approximately 27% from 4.08 to 2.96 $\times$ 10−10 ml O2/s. Interestingly, this estimation is strikingly similar to the ~28% reduction in total hindlimb skeletal muscle blood flow of conscious resting rats following GLI administration (Colburn, Holdsworth, et al., 2020).

GLI-induced alterations in $V_{RBC}$ (Figure 2, bottom panel) underscore the potential of KATP Channels to further modulate capillary gas exchange via changes in RBC transit time. Given the ~25% decrease in $V_{RBC}$ with GLI observed herein and the mean capillary length within the rat spinotrapezius (i.e., 430 μm based on previous morphometric analyses by Gray, 1984), total capillary RBC residence time is calculated to increase from ~1.8 s during control to ~2.4 s following KATP channel inhibition. While it is acknowledged that these calculations may underestimate true residence time due to the actual RBC path length being somewhat longer than the anatomical path length (Sarelius, 1986), the amount of error introduced should not preferentially bias either superfusion condition. Longer RBC transit times provide the mechanistic basis for the elevated fractional O2 extraction expected with GLI as discussed above.

The proportional reductions in $f_{RBC}$ and $V_{RBC}$ with GLI were such that Hctcap remained unchanged between conditions (vide supra its mathematical description; Eq. 1). It is interesting that the relationship between $f_{RBC}$ and $V_{RBC}$ in the resting skeletal muscle may also be preserved with aging and some disease states. Accordingly, previous studies from our laboratory have shown that the microvascular dysregulation characteristic of aging (Copp et al., 2009; Russell et al., 2003), type I diabetes (Kindig et al., 1998), and chronic heart failure (Kindig et al., 1999; Richardson et al., 2003) does not appear to affect resting Hctcap. As noted above, Hctcap is a primary determinant of DO2m and, therefore, diffusive O2 conductance within the microcirculation (Federspiel & Popel, 1986; Groebe & Thews, 1990). This derives from the low O2 diffusivity in plasma and the particulate nature of blood,
which render only the capillary surface area in close proximity to the RBC functional for diffusion at any given time. The current Hct\text{cap} data thus indicate preserved capillary surface area available for diffusive O₂ exchange within continuously RBC-perfused vessels following Kₐ₅₆ channel inhibition.

An important question for future studies is whether the impairments in resting capillary hemodynamics seen here after Kₐ₅₆ inhibition are also present during muscle contractions and, if so, their potential implications for exercise tolerance. In the resting spinotrapezius, GLI lowered f\text{RBC} and V\text{RBC} (i.e., reduced Qm; Figure 2) whilst not affecting Hct\text{cap} or the %RBC-flowing capillaries (i.e., unaltered DO₂m as noted above), thereby predisposing to higher O₂ extraction (i.e., higher DO₂m/Qm ratio; Eq. 2). While there has been no prior evaluation of Kₐ₅₆ channel inhibition on contracting muscle capillary blood flow per se, studies on arteriolar diameter and bulk blood flow have produced contrasting evidence regarding the role of Kₐ₅₆ channels in functional hyperemia (Tykocki et al., 2017). Nonetheless, recent reports from our laboratory indicate that GLI lowers microvascular O₂ partial pressures (PO₂) as well as estimated DO₂m in the contracting spinotrapezius and mixed gastrocnemius muscles (Colburn, Weber, et al., 2020; Holdsworth et al., 2016). Under these circumstances, intramyocyte PO₂ is expected to fall to generate the driving pressure (ΔPO₂) and possibly elevate intramyocyte O₂ diffusing capacity by and by further deoxygenating myoglobin as necessary to maintain a given metabolic rate as described by Fick's law of diffusion:

\[ \dot{V}O₂ = DO₂m \times \Delta PO₂ \]  

where VO₂ corresponds to the rate of O₂ flux, DO₂m is the diffusing capacity as defined above and ΔPO₂ the O₂ partial pressure difference between the microvascular and intramyocyte spaces. Reduction in intramyocyte PO₂ can be problematic because it promotes depletion of finite muscle phosphocreatine and glycogen stores, intracellular accumulation of metabolites (including ADP, Pi, and H⁺) and acid-base perturbations associated with fatigue (Hogan et al., 1992; Wilson et al., 1977). Therefore, based on the above observations, potential impairments in contracting muscle capillary hemodynamics with GLI could result in pernicious consequences for oxidative metabolism and exercise tolerance (Colburn, Weber, et al., 2020; Lu et al., 2013).

Despite marked reductions in RBC hemodynamics (i.e., decreased f\text{RBC} and V\text{RBC}; Figure 2), GLI did not change microvascular blood flow heterogeneity (as evaluated by the coefficient of variation of f\text{RBC}, V\text{RBC}, and Hct\text{cap}) in the resting spinotrapezius. Due to the nature of our experimental protocol (i.e., assessment of the same microcirculatory fields, and whenever possible, capillaries between conditions), these data indicate that any spatial vascular/metabolic mismatch present at rest was not exacerbated with GLI. Whether the same holds true for the contracting skeletal muscle (amid dynamic temporal and spatial changes in O₂ delivery-utilization) remains to be determined. Importantly, the consequences of microvascular blood flow heterogeneity assume greater significance during exercise when high O₂ fluxes (which can rise >100-fold compared to rest) are required to support oxidative metabolism.

4.3 | Clinical implications

Sulfonylureas are the most popular second-line treatment prescribed for type 2 diabetes mellitus (Montvida et al., 2018), promoting insulin release via inhibition of pancreatic Kₐ₅₆ channels. However, systemic inhibition of Kₐ₅₆ channels with these oral medications may also compromise key components of capillary gas exchange within skeletal muscle as revealed herein (i.e., lowered f\text{RBC} and V\text{RBC}; Figure 2). The latter microvascular impairments compound with those described previously in the diabetic skeletal muscle (Kindig et al., 1998; Padilla et al., 2006) thus questioning the therapeutic value of sulfonylurea medications for diabetic patients. Systemic sulfonylurea administration has also been reported to reduce contracting muscle bulk blood flow (Holdsworth et al., 2015; Keller et al., 2004; Thomas et al., 1997) and submaximal and maximal exercise tolerance (Colburn, Weber, et al., 2020; Lu et al., 2013). Although no decrements in in-capillary DO₂m with GLI were found herein at rest, potential reductions in the proportion of capillaries supporting continuous RBC flow with sulfonylurea medications during contractions would be expected to impair muscle glucose uptake (via reductions in exchange surface area). As such, exercise intolerance may be exacerbated in diabetics by systemic sulfonylurea medications impairing Kₐ₅₆ channel regulation of skeletal muscle capillary hemodynamics.

4.4 | Experimental considerations

Superfusion of the spinotrapezius muscle with GLI was employed herein to locally inhibit Kₐ₅₆ channels in vivo. This topical drug delivery method was chosen to prevent the potential confound of alterations in central hemodynamics (e.g., MAP, HR, systemic vascular resistance), myocardial hemodynamics and function (e.g., coronary vascular conductance and O₂ delivery, left ventricular relaxation rate), sympathetic nerve activity, visceral organ blood flow, blood insulin and glucose concentration, among others with systemic drug administration as reported previously (Colburn, Holdsworth, et al., 2020; Colburn, Weber, et al., 2020; Duncker et al., 2001; Farouque & Meredith, 2003a; Holdsworth et al., 2015, 2016; Rocha et al., 2020; Thomas et al., 1997). Nonetheless, it is not possible to pinpoint the
site of GLI action (e.g., across the vascular network and/or among distinct cell types) with the current experimental protocol. Although our GLI concentration and superfusion protocol were based on previous microcirculatory studies (Cohen & Sarelius, 2002; Hammer et al., 2001; Hodnett et al., 2008; Jackson, 1993; Lu et al., 2013; Murrant & Sarelius, 2002; Saito et al., 1996; Xiang & Hester, 2009) as described earlier, this approach could lead to non-specific vasodilation (Jiang et al., 2007; Tykocki et al., 2017) and potentially underestimate the contribution of K\textsubscript{ATP} channels to microvascular control. It is noteworthy that, despite the latter considerations and the modest sample size (n = 6), significant GLI-induced reductions in f\textsubscript{RBC} and V\textsubscript{RBC} were observed in the resting spinotrapezius muscle (Figures 2 and 3). Post hoc power calculations indicate that a total of 48 animals would be needed to detect significant changes in the %RBC-flowing capillaries between conditions.

Previous evidence indicates that muscle capillaries increase in diameter by ~0.5–1.0 µm from their arteriolar to venular ends (Smaje et al., 1970). Although d\textsubscript{cap} measurements herein were not normalized for the distance from arteriolar or venular ends, our d\textsubscript{cap} assessment at random sites along the capillary length is not anticipated to introduce systematic errors. Moreover, capillary structural and hemodynamics analyses of the same microcirculatory fields within a given animal (and, in 47% of the cases, within the same capillary between CON and GLI conditions) likely minimized the potential for biases (including spatial vascular/metabolic heterogeneity, and heterogeneous f\textsubscript{RBC}, V\textsubscript{RBC} and Hct\textsubscript{cap} distributions) to be expressed in the present results. The lack of changes in d\textsubscript{cap} in the face of reduced f\textsubscript{RBC} and V\textsubscript{RBC} with GLI (Figure 2) is consistent with upstream arteriolar regulation of skeletal muscle blood flow as mentioned above (Joyner & Casey, 2015; Kindig & Poole, 2001; Laughlin et al., 2012; Segal, 2005).

4.5 Summary and conclusions

Local inhibition of K\textsubscript{ATP} channels with GLI did not reduce the proportion of capillaries supporting continuous RBC flow or Hct\textsubscript{cap} in the resting rat spinotrapezius muscle. These data are indicative of preserved muscle O\textsubscript{2} diffusing capacity (DO\textsubscript{2}m) with GLI. In contrast, GLI lowered both f\textsubscript{RBC} and V\textsubscript{RBC} thus impairing perfusive microvascular O\textsubscript{2} transport (Qm) and lengthening RBC capillary transit times, respectively. Considering the interdependence between diffusive and perfusive O\textsubscript{2} conductances (i.e., %O\textsubscript{2} extraction determined largely by the local DO\textsubscript{2}m/Qm; vide Eq. 2 above) (Roca et al., 1992), such GLI-induced alterations in capillary hemodynamics are expected to elevate muscle fractional O\textsubscript{2} extraction to sustain a given metabolic rate. Taken together, the current results suggest that K\textsubscript{ATP} channels regulate capillary hemodynamics (and, therefore, microvascular gas exchange) in resting skeletal muscle.

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DISCLOSURE

No conflicts of interest, financial or otherwise, are declared by the authors.

AUTHOR CONTRIBUTIONS

DMH, TIM, and DCP conceived and designed research; DMH, AT, JCC, and TDC performed experiments; DMH analyzed data; DMH, TIM, and DCP interpreted results of experiments; DMH prepared figures and drafted manuscript; DMH, AT, JCC, TDC, TIM, and DCP approved final version of the manuscript.

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