PEARLS

Climate change and the emergence of fungal pathogens

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The role of the environment in emerging and reemerging infectious diseases is increasingly recognized [1,2]. Climate change, defined by the United Nations Framework Convention on Climate Change as “a change of climate which is attributed directly or indirectly to human activity that alters the composition of the global atmosphere and which is in addition to natural climate variability observed over comparable time periods” [3] may create environmental pressures that result in new diseases caused by fungi [4]. While viral and bacterial diseases receive most attention as the potential cause of plagues and pandemics, fungi can arguably pose equal or even greater threats: There are no vaccines available yet for fungal pathogens, the arsenal of antifungal agents is extremely limited, and fungi can live saprotrophically, producing large quantities of infectious spores and do not require host-to-host contact to establish infection [5]. Indeed, fungi seem to be uniquely capable of causing complete host extinction [6]. For the vast majority of fungal species, the capacity to grow at elevated temperatures limits their ability to infect and establish in mammals. However, fungi can be trained to evolve thermotolerance, and gradual adaptation to increasing temperature caused by climate change could lead to an increase of organisms that can cause disease [7,8]. In addition, climate change can increase the geographic range of pathogenic species or their vectors, leading to the emergence of diseases in areas where they have not previously been reported [7]. Environmental disruptions due to climate change such as floods, storms, and hurricanes can disperse and aerosolize fungi or implant them via traumatic wounds, resulting in infections by previously very rare or unknown fungal species. Fig 1 summarizes the potential effects of climate change, showing examples of emerging fungi and their consequences, along with the potential for new and currently unknown species to emerge.

Here, we review current literature on the role of climate change in the emergence of fungal diseases. In the context of this review, we consider emerging fungal pathogens to be those that have been reported to have caused disease in plants or animals in the last 30 years, have arisen in novel geographical locations, or have become substantially more problematic, with demonstrated evidence that this emergence is attributed to climate change.

Completely new fungal species hypothesised to have emerged due to climate change

Candida auris

First identified in 2009 from an ear infection in Japan [9], C. auris has since spread globally, with reports from every continent [10]. Little is known about the origins of C. auris; however, its phylogenetic relatives have been isolated from the environment [11], and it is hypothesized to have a nonhuman environmental reservoir with possible dispersal by birds [12]. This yeast
is considered the first “novel” pathogen to have evolved in response to climate change [10,12], although this remains speculative and awaits conclusive evidence; alternative hypotheses for its emergence include expanded farming and aquaculture resulting in increased contact with humans, and environmental contamination with fungicides [11]. C. auris is particularly problematic in healthcare settings where it colonises and spreads to cause nosocomial outbreaks, and it is remarkably resistant to antifungals and disinfectants [13]. Fortunately, C. auris is relatively low in virulence and has only caused infection in people suffering from severe comorbidities [12,13]. Four major genomic clades have been identified for C. auris and these are associated with particular geographic regions: clade I (South Asian), clade II (East Asian), clade III (South African), and clade IV (South American) [10]. Clade II appears to infect the ear only, whereas the other clades have caused invasive infections [14].

Fungal species that have emerged to cause significant outbreaks that appear related to climate change

**Batrachochytrium dendrobatidis**

*Batrachochytrium dendrobatidis* (*Bd*) is an emerging pathogen of amphibians. An environmental chitrid fungus, *Bd* is responsible for the most spectacular loss of amphibian diversity in history [15] and since 1998 has spread to all continents where amphibians are found, decimating populations in Australia and North and South America [16]. *Bd* has also spread to Africa, with the first report in North Africa in 2011 [17]. IPCC data forecasts an increased spread of *Bd* across the northern hemisphere, particularly in the higher-latitude regions that are experiencing warming temperatures [18]. In addition to the increased global range of the pathogen, a shift from cold to hotter temperature has been shown to increase the susceptibility of the amphibian host [19]. Increasing climate change is therefore predicted to cause an ongoing decline in amphibian populations.

**Cryptococcus deuterogattii**

*Cryptococcus deuterogattii*, previously named *Cryptococcus gattii* VGII, was recently elevated to a species within the *Cryptococcus gattii* species complex [20]. *C. gattii* complex species were...
traditionally associated with tropical and subtropical climates [21]; however, in recent years, C. deuterogattii has emerged in the temperate regions of western Canada and the Pacific northwest of North America where it caused hundreds of infections in people and animals [22]. Interestingly, C. deuterogattii has a higher level of thermal tolerance than other C. gattii complex members, indicating that it may have a greater capacity for thermal adaptation [23]. The spread of this fungus has been attributed to anthropogenic dispersal through vehicle wheel wells, footwear, construction and forestry activity (leading to aerial dispersal), and water [24], with climate change suggested as a potential driver [25].

New virulent fungal lineages with adaptations suggested to have evolved in response to climate change

Puccinia striiformis (Rust fungus)

Stripe rust is one of the most devastating global diseases of wheat and is caused by the fungus Puccinia striiformis [26,27]. Wheat yellow (stripe) rust, caused by Puccinia striiformis f. sp. tritici (Pst), has previously shown preference for cold areas but has recently been seen to invade warmer regions [28]. Since 2000, Pst has been reported to have adapted to increased temperatures, with novel strains Pst1, Pst2, and “Warrier,” which are more aggressive and thermotolerant, replacing older strains and spreading to new regions [28]. These strains have now been reported worldwide [30], with major outbreaks occurring in south central United States [29,31] and Australia [32]. Microsatellite genotyping and virulence phenotyping have detected a high degree of variability within the Pst1 and Pst2 lineages, demonstrating an ongoing evolutionary potential of this pathogen [30].

Fusarium head blight

Fusarium head blight (FHB) is a disease of great concern for wheat and for other cereal crops [33]. Caused by members of the Fusarium graminearum species complex (FGSC) [34], infection leads to reduced cereal yield and quality, thereby negatively affecting food security. Outbreaks occur particularly in years with warm and humid weather conditions, resulting in yield losses of up to 75% [35]. Over the past approximately 20 years, some temperate regions have seen a shift from Fusarium culmorum, which is associated with cooler and wetter conditions, to F. graminearum, which favours warmer, humid conditions [36]. Importantly, F. graminearum is more aggressive than F. culmorum and is associated with higher yield loss [37]; hence, the severity of FHB is likely to further increase in future warmer climates. F. graminearum also produces more mycotoxins, and there is evidence that their production increases with temperature and water stress [38], with potential impacts on human and animal health.

Fungal pathogens that have become more problematic due to environmental disruption caused by climate change

Climate change affects rainfall causing floods and drought and can increase tropical cyclone and tornado severity, leading to far-reaching human health impacts. The link between natural disasters and subsequent fungal infections in disaster-affected persons is increasingly recognized, with an excellent review of this area by Benedict and Park [39]. Disaster-associated fungi can be violently displaced and widely disseminated, leading to pulmonary and soft tissue infections by fungal species that may otherwise be uncommon. For example, a severe duststorm in the southern San Joaquin Valley of California in 1977 dispersed Coccidioides immitis from Bakersfield, an area of high endemnicity, to Sacramento County, where it is rare, resulting in more than 100 infections [40]. While this predates the era of significant climate change,
it demonstrates the ramifications of acute environmental disruption, particularly as climate change modelling predicts that the range of *Coccidioides* could expand from 12 to 17 states, with infections increasing by 50% [41]. *Apophysomyces trapeziformis*, a thermotolerant saprotroph that is an extremely rare cause of mucormycosis caused a cluster of cases in patients wounded following the 2011 tornado in Joplin, Missouri [42]. Various other soil-borne fungal pathogens have been speculated to have increased in frequency or range due to climate-induced disruptions, including *Talaromyces marneffei* [43], *Blastomyces*, *Histoplasma* [44], and *Paracoccidioides* [45]. Outside of their normal range, these fungal infections can be challenging to diagnose and they are often refractory to treatment, frequently resulting in poor patient outcomes [40].

**Conclusions and future perspectives**

Unless action is taken to drastically reduce carbon emissions, the global temperature will continue to rise, and the impacts of climate change are predicted to become increasingly severe. Fungi are likely to continue to affect crops and native plants, with expanding ecological range and long-distance dispersal events producing new risks. The threat of novel fungal infections of humans and other mammals is less predictable, and there is a need to understand more about the nexus between ecology and medical mycology, which have traditionally been studied in relative isolation [46]. It is now recognised that many clinically relevant fungal pathogens, including those traditionally considered to be human commensals like *Candida albicans*, can be found in the environment [47], with environmental pressures affecting the evolution of novel traits including virulence and antifungal resistance [48]. Climate change produces a confluence of factors that can act together to drive the emergence of new pathogens (Fig 1). In the clinical setting, it may become increasingly necessary to consider infection by novel or uncommon species, or adapted strains within species, which might otherwise be considered unlikely to be endemic or capable of breaching the mammalian thermal barrier. The era of HIV-AIDS and immunosuppression rewrote the textbooks on medical mycology and what may or may not be considered a potential fungal pathogen; it is likely that climate change will do this again.

**References**

1. Wu X, Lu Y, Zhou S, Chen L, Xu B. Impact of climate change on human infectious diseases: Empirical evidence and human adaptation. Envir Int. 2016; 86:14–23. https://doi.org/10.1016/j.envint.2015.09.007 PMID: 26479830

2. El-Sayed A, Kamel M. Climatic changes and their role in emergence and re-emergence of diseases. Environ Sci Pollut Res Int. 2020; 27:22336–52. https://doi.org/10.1007/s11356-020-08896-w PMID: 32347486

3. Farber DA, Carlarne CP. Climate change law. Ohio St Pub Law Work Paper. 2017;419.

4. Garcia-Solache MA, Casadevall A. Global warming will bring new fungal diseases for mammals. MBio. 2010; 1 (1):e00061–10. https://doi.org/10.1128/mBio.00061-10 PMID: 20689745

5. Casadevall A. Global catastrophic threats from the fungal kingdom: fungal catastrophic threats. Glob Catast Biol Risks. 2019:21–32. https://doi.org/10.1007/82_2019_161 PMID: 31119433

6. Fisher MC, Henk DA, Briggs CJ, Brownstein JS, Madoff LC, McCraw SL, et al. Emerging fungal threats to animal, plant and ecosystem health. Nat. 2012; 484(7393):186–94. https://doi.org/10.1038/nature10947 PMID: 22488624

7. De Crecy E, Jaronski S, Lyons B, Lyons TJ, Keyhani NO. Directed evolution of a filamentous fungus for thermotolerance. BMC Biotechnol. 2009; 9 (1):74. https://doi.org/10.1186/1472-6750-9-74 PMID: 19709419

8. Casadevall A. Climate change brings the specter of new infectious diseases. J Clin Invest. 2020; 130 (2):553–5. https://doi.org/10.1172/JCI135003 PMID: 31904588
9. Satoh K, Makimura K, Hasumi Y, Nishiyama Y, Uchida K, Yamaguchi H. *Candida auris* sp. nov., a novel ascomycetous yeast isolated from the external ear canal of an inpatient in a Japanese hospital. *Microbiol Immunol.* 2009; 53 (1):41–4. https://doi.org/10.1111/j.1348-0421.2008.00083.x PMID: 19161556

10. Rhodes J, Fisher MC. Global epidemiology of emerging *Candida auris*. *Curr Opin Microbiol.* 2019; 52:84–9. https://doi.org/10.1016/j.mib.2019.05.008 PMID: 31279224

11. Jackson BR, Chow N, Forsberg K, Lockhart SR, Welsh R, et al. On the origins of a species: what might explain the rise of *Candida auris*. *J Fungi.* 2019; 5 (3):58. https://doi.org/10.3390/jof5030058 PMID: 31284576

12. Casadevall A, Kontoyiannis DP, Robert V. On the emergence of *Candida auris*: climate change, azoles, swamps, and birds. *mBio.* 2019; 10 (4):e01397–19. https://doi.org/10.1128/mBio.01397-19 PMID: 31337723

13. Chowdhary A, Sharma C, Meis JF. *Candida auris*: a rapidly emerging cause of hospital-acquired multidrug-resistant fungal infections globally. *PLoS Pathog.* 2017; 13 (5):e1006290. https://doi.org/10.1371/journal.ppat.1006290 PMID: 28542486

14. Welsh RM, Sexton DJ, Forsberg K, Vallabhaneni S, Litvintseva A. Insights into the unique nature of the East Asian clade of the emerging pathogenic yeast *Candida auris*. *J Clin Microbiol.* 2019; 57 (4):1–5. https://doi.org/10.1128/JCM.00007-19 PMID: 30760535

15. Skerratt LF, Berger L, Speare R, Cashins S, McDonald KR, Phillott AD, et al. Spread of chytridiomycosis has caused the rapid global decline and extinction of frogs. *EcoHealth.* 2007; 4 (2):125–310.

16. Fisher MC, Garner TW, Walker SF. Global emergence of *Batrachochytrium dendrobatidis* and amphibian chytridiomycosis in space, time, and host. *Ann Rev Microbiol.* 2009; 63:291–310.

17. El Mouden E, Slimani T, Donaire D, Fernández-Beaskoetxea S, Fisher MC, Bosch J. First record of the European chytrid fungus *Batrachochytrium dendrobatidis* in North Africa. *Herpet Rev.* 2011; 42 (1):71–5.

18. Xie GY, Olson DH, Blaustein AR. Projecting the global distribution of the emerging amphibian fungal pathogen, *Batrachochytrium dendrobatidis*, based on IPCC climate futures. *PLoS ONE.* 2016; 11 (8): e0160746. https://doi.org/10.1371/journal.pone.0160746 PMID: 27513565

19. Bradley PW, Brawner MD, Raffel TR, Rohr JR, Olson DH, Blaustein AR. Shifts in temperature influence how *Batrachochytrium dendrobatidis* infects amphibian larvae. *PLoS ONE.* 2019; 14 (9):e0222237. https://doi.org/10.1371/journal.pone.0222237 PMID: 31536533

20. Hagen F, Khayhan K, Theelen B, Kolecka A, Sionov E, et al. Recognition of seven species in the *Cryptococcus gattii/Cryptococcus neoformans* species complex. *Fung Genet Biol.* 2015; 78:76–84. https://doi.org/10.1016/j.fgb.2015.02.009 PMID: 25721988.

21. Herkert PF, Hagen F, Pinheiro RL, Muro MD, Meis JF, Queiroz-Telles F. Ecoepidemiology of *Cryptococcus gattii* in developing countries. *J Fungi.* 2017; 3 (4):62. https://doi.org/10.3390/jof3040062 PMID: 29371578

22. Datta K, Bartlett KH, Baer R, Byrnes E, Galanis E, Heitman J, et al. Spread of *Cryptococcus gattii* into Pacific Northwest region of the United States. *Emerg Infect Dis.* 2009; 15 (8):1185. https://doi.org/10.3201/eid1508.081384 PMID: 19757550

23. Fernandes KE, Dwyer C, Campbell LT, Carter DA. Species in the *Cryptococcus gattii* complex differ in capsule and cell size following growth under capsule-inducing conditions. *mSphere.* 2016; 1 (6):e00350–16. PubMed Central PMCID: PMC28066814. https://doi.org/10.1128/mSphere.00350-16 PMID: 28066814

24. Bartlett KH, Kidd SE, Kronstad JW. The emergence of *Cryptococcus gattii* in British Columbia and the Pacific Northwest. *Curr Infect Dis Rep.* 2008; 10 (1):58–65. https://doi.org/10.1007/s11908-008-0011-1 PMID: 18377181

25. Greer A, Ng V, Fisman D. Climate change and infectious diseases in North America: the road ahead. *CMAJ.* 2008; 178 (6):715–22. https://doi.org/10.1503/cmaj.081325 PMID: 18332386

26. Lyon B, Broders K. Impact of climate change and race evolution on the epidemiology and ecology of stripe rust in central and eastern USA and Canada. *Can J Plan Pathol.* 2017; 39 (4):358–92.

27. Chen W, Wellings C, Chen X, Kang Z, Liu T. Wheat stripe (yellow) rust caused by *Puccinia striiformis f. sp. tritici*. *Mol Plan Pathol.* 2014; 15 (5):433–46. https://doi.org/10.1111/mpp.12116 PMID: 24373199

28. de Vallavieille-Pope C, Bahra B, Leconte M, Zurluhu O, Belaid Y, Maghrebi E, et al. Thermal generalist behaviour of invasive *Puccinia striiformis f. sp. tritici* strains under current and future climate conditions. *Plan Pathol.* 2018; 67 (6):1307–20.

29. Millus E, Seyran E, Mc New R. Aggressiveness of *Puccinia striiformis f. sp. tritici* isolates in the south-central United States. *Plant Dis.* 2006; 90 (7):847–52. https://doi.org/10.1094/PD-90-0847 PMID: 30781019
30. Walter S, Ali S, Kemen E, Nazari K, Bahri BA, Enjalbert J, et al. Molecular markers for tracking the origin and worldwide distribution of invasive strains of *Puccinia striiformis*. Ecol Evol. 2016; 6 (9):2790–804. https://doi.org/10.1002/ece3.2069 PMID: 27066253

31. Milus EA, Kristensen K, Hoymoller MS. Evidence for increased aggressiveness in a recent widespread strain of *Puccinia striiformis* f. *sp. tritici* causing stripe rust of wheat. Phytopathology. 2009; 99 (1):89–94. https://doi.org/10.1094/PHTY-99-1-0089 PMID: 19055439

32. Laradze A, Drum T, Wellings C. Temperature adaptation in Australasian populations of *Puccinia striiformis* f. *sp. tritici*. Plan Pathol. 2014; 63 (3):572–80.

33. Dweba C, Figlan S, Shimelis H, Motaung T, Sydenham S, Mwadzingeni L, et al. *Fusarium* head blight of wheat: Pathogenesis and control strategies. Crop Prot. 2017; 91:114–22.

34. Sarver BA, Ward TJ, Gale LR, Broz K, Kistler HC, Aoki T, et al. Novel *Fusarium* head blight pathogens from Nepal and Louisiana revealed by multilocus genealogical concordance. Fung Genet Biol. 2011; 48 (12):1096–107. https://doi.org/10.1016/j.fgb.2011.09.002 PMID: 22004876

35. Leplat J, Friberg H, Abid M, Steinberg C. Survival of *Fusarium graminearum*, the causal agent of *Fusarium* head blight. A review Agron Sustain Develop. 2013; 33 (1):97–111.

36. Goertz A, Zuehlke S, Spiteller M, Steiner U, Dehne HW, Waalwijk C, et al. *Fusarium* species and mycotoxin profiles on commercial maize hybrids in Germany. Eur J Plant Pathol. 2010; 128 (1):101–11.

37. Jennings P, Coates M, Walsh K, Turner JA, Nicholson P. Determination of deoxyxynivalenol-and nivalenol-producing chemotypes of *Fusarium graminearum* isolated from wheat crops in England and Wales. Plant Pathol. 2004; 53 (5):643–52.

38. Magan N, Medina A, Aldred D. Possible climate-change effects on mycotoxin contamination of food crops pre-and postharvest. Plant Pathol. 2011; 60 (1):150–63.

39. Benedict K, Park BJ. Invasive fungal infections after natural disasters. Emerg Infect Dis. 2014; 20 (3):349. https://doi.org/10.3201/eid2003.131230 PMID: 24565446

40. Schleifer KE, Schleifet RA, Spigler RA, Jibson RW, Harp EL, Marshall GA, et al. A coccidioidomycosis outbreak following the Northridge, Calif. earthquake. JAMA. 1997; 277 (11):904–8. PMID: 9062329

41. Gorris ME, Treseder KK, Zender CS, Randerson JT. Expansion of coccidioidomycosis endemic regions in the United States in response to climate change. GeoHealth. 2019; 3 (10):308–27. https://doi.org/10.1029/2019G H000209 PMID: 32159021

42. Nebiott Fanfair R, Benedict K, Bos J, Bennet SD, Lo Y-C, Adebanjo T, et al. Necrotizing cutaneous mucormycosis after a tornado in Joplin, Missouri, in 2011. New Eng J Med. 2012; 367 (23):2214–25. https://doi.org/10.1056/NEJMo1204781 PMID: 23215557

43. Panackal AA. Global climate change and infectious diseases: invasive mycoses. J Earth Sci Clim Chan. 2011; 2 (2).

44. Azar MM, Loyd JL, Relich RF, Wheat LJ, Hage CA, editors. Current concepts in the epidemiology, diagnosis, and management of histoplasmosis syndromes. Semin Respir Crit Care Med; 2020: Thieme Medical Publishers.

45. Martinez R. New trends in paracoccidioidomycosis epidemiology. J Fungi. 2017; 3(1):1. https://doi.org/10.3390/jf03010001 PMID: 29371520

46. Moro F, Dear medical mycologists, it is time to look outside the box. FEMS Yeast Res. 2020; 20(1):foz080. https://doi.org/10.1093/femsyr/foz080 PMID: 31738413

47. Opulente DA, Langdon QK, BuH KV, Haase MA, Sylvester K, Moriarty RV, et al. Pathogenic budding yeasts isolated outside of clinical settings. FEMS Yeast Res. 2019; 19(3):foz032. https://doi.org/10.1093/femsyr/foz032 PMID: 31076749

48. Verweij PE, Snelders E, Kema GH, Mellado E, Melchers WJ. Azole resistance in *Aspergillus fumigatus*: a side-effect of environmental fungicide use? Lancet Inf Dis. 2009; 9 (12):789–95. https://doi.org/10.1016/S1473-3099(09)70265-8 PMID: 19926038