Clinical and Molecular Perspectives of Monogenic Hypertension

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Abstract: Advances in molecular research techniques have enabled a new frontier in discerning the mechanisms responsible for monogenic diseases. In this review, we discuss the current research on the molecular pathways governing blood pressure disorders with a Mendelian inheritance pattern, each presenting with a unique pathophysiology. Glucocorticoid Remediable Aldosteronism (GRA) and Apparent Mineralocorticoid Excess (AME) are caused by mutations in regulatory enzymes that induce increased production of mineralocorticoids or inhibit degradation of glucocorticoids, respectively. Geller syndrome is due to a point mutation in the hormone responsive element of the promoter for the mineralocorticoid receptor, rendering the receptor susceptible to activation by progesterone, leading to hypertension during pregnancy. Pseudohypoaldosteronism type II (PHA-II), also known as Gordon’s syndrome or familial hyperkalemic hypertension, is a more variable disorder typically characterized by hypertension, high plasma potassium and metabolic acidosis. Mutations in a variety of intracellular enzymes that lead to enhanced sodium reabsorption have been identified. In contrast, hypertension in Liddle’s syndrome, which results from mutations in the Epithelial sodium Channel (ENaC), is associated with low plasma potassium and metabolic alkalosis. In Liddle’s syndrome, truncation of one the ENaC protein subunits removes a binding site necessary protein for ubiquitination and degradation, thereby promoting accumulation along the apical membrane and enhanced sodium reabsorption. The myriad effects due to mutation in phosphodiesterase 3A (PDE3A) lead to severe hypertension underlying sodium-independent autosomal dominant hypertension with brachydactyly. How mutations in PDE3A result in the phenotypic features of this disorder are discussed. Understanding the pathologies of these monogenic hypertensive disorders may provide insight into the causes of the more prevalent essential hypertension and new avenues to unravel the complexities of blood pressure regulation.

Keywords: Hypertension, monogenic, aldosteronism, mineralocorticoid, liddle’s syndrome, ENaC, WNK, PDE3A.

1. INTRODUCTION

The purpose of this paper is to describe human hypertension disorders that are due to alterations in single genes. These include Glucocorticoid Remediable Aldosteronism (GRA), Liddle’s syndrome, Apparent Mineralocorticoid Excess (AME), Pseudohypoaldosteronism type II (PHA-II), Geller syndrome, and Autosomal Dominant Hypertension with Brachydactyly (ADHB). The research done on these rare genetic disorders has shed much light on the pathways and mechanisms involved in blood pressure regulation. These disorders are often categorized as low renin forms of hypertension. Patients with these disorders present with unique pathological states offer new insight into the causes of primary hypertension. Genes responsible for hereditary forms of pheochromocytoma and familial paraganglioma have also been identified but are beyond the scope of this review.

1.1. Background

Hypertension is a chronic medical condition characterized by elevated arterial blood pressure. Hypertension has been considered as a major risk factor of coronary heart disease [1], stroke, aneurysms, kidney failure and arterial disease, and is classified as either primary or secondary. Primary hypertension, also called essential or idiopathic hypertension, is a form of elevated blood pressure with no identifiable cause. Studies have shown that out of all cases of diagnosed hypertension, 95% of cases are diagnosed as this type [2]. The underlying mechanisms are currently believed to be a combination of genetic and environmental factors. Secondary hypertension results from an identifiable underlying cause, often involving kidney or endocrine diseases and tumors. It can also occur as a side effect of certain medications. Only 5% of hypertensive cases are diagnosed as secondary. Of these, a subgroup can be further identified to be due to mutations in a single gene product or protein. According to the Center of Disease Control and Prevention, about 70 million American adults are hypertensive. Annual costs
of health care and medication to treat people with hypertension are over $46 billion. In 2013, more than 360,000 deaths in the United States reported hypertension as a primary or secondary cause [3, 4].

2. GLUCOCORTICOID REMEDIABLE ALDOSTERONISM

GRA is an autosomal dominant disorder that results in hypertension, high serum aldosterone, low renin activity and abnormal adrenal steroid production. It is also referred to as aldosterone synthase hyperactivity, glucocorticoid suppressible hyperaldosteronism or familial hyperaldosteronism type I.

2.1. Background

GRA was first discovered and studied in the 1960’s, and since then has been studied by multiple investigators across the scientific community. Sutherland et al. (1966) and Salti et al. (1969) first described a father and son who showed symptoms of hypertension with increased aldosterone secretion but low renin activity [5, 6]. Ganguly et al. (1981) diagnosed a 7-year-old boy with GRA, which led to the discovery that his mother and grandmother also had an identical phenotype. A urinary analysis failed to identify an absolute "aldosterone stimulating factor." This suggested that GRA was a distinct disorder from idiopathic aldosteronism [7]. Another study reported a family diagnosed with GRA when a saline infusion failed to lower aldosterone levels and administration of a loop diuretic and low Na + diet did not increase renin activity. This study also showed that the decline in plasma aldosterone secretion when a patient is upright, seen in aldosterone producing adenomas, is also observed in GRA. Gordon et al. (1995) reported the phenotypic heterogeneity of GRA in over 21 members from a kindred with more than 1,000 decedents of an English convict, who was sent to Australia in 1837 for highway robbery. Gordon was the first to refer to GRA as familial hyperaldosteronism type I [8]. Shortly thereafter, Gates et al. (1996) reported two large pedigrees with GRA that were confirmed my genetic analysis. At the time, GRA patients were clinically indistinguishable from patients with essential hypertension. Gates suggested that GRA is an underdiagnosed condition [9]. Stowasser et al. (1999) studied ten normotensive subjects with GRA that took no medication. Results showed normal plasma and normal upright aldosterone levels. However, in subjects with GRA and hypertension, the results demonstrated a failure of aldosterone to increase at least 50% (normal) in subjects sitting in an upright position, as well as a failure to raise aldosterone after an angiotensin II infusion. Studies also showed that aldosterone levels correlated significantly with cortisol levels, but not with plasma renin activity. Aldosterone levels were suppressed after treatment with dexamethasone, suggesting that Adrenocorticotropic Hormone (ACTH) was regulating aldosterone production [10].

In 2000, Stowasser et al. studied GRA patients with varying degrees of hypertension. They showed that males were affected more severely than females and that the degree of hybrid gene-induced aldosterone overproduction contributed to the severity of hypertension [11]. Multatero et al. (2002) discovered the presence of a chimeric gene in the affected GRA members of through four of five-generations in a pedigree from Sardinia [12].

2.2. Physiology

The adrenal glands are endocrine glands that produce a variety of hormones including: glucocorticoids (cortisol), mineralocorticoids (aldosterone), androgens (dehydroepiandrosterone [DHEA]), epinephrine and norepinephrine. Glucocorticoids, mineralocorticoids, and androgens are steroid hormones and share the same biosynthetic pathways. Each is unique in their evolution, regulation, and effects. They are all derived from cholesterol and synthesized in differentiated layers within the cortex of the adrenal gland (Fig. 1). The zona glomerulosa is the outermost layer of the cortex. This is the main site of mineralocorticoid (aldosterone) synthesis [13].

Aldosterone is involved in the regulation of electrolytes and blood pressure. The final step is the formation of aldosterone from corticosterone by the enzyme aldosterone synthase. Aldosterone synthase is a steroid hydroxylase cytochrome P450 enzyme and is exclusively expressed in mitochondria of the zona glomerulosa [14]. Aldosterone synthase is encoded on chromosome 8 by the CYP11B2 gene. It converts corticosterone the final product aldosterone [13].

Studies show that calcium (Ca 2+) acts as a transcription factor for aldosterone synthase though interaction at the 5' flanking region of the CYP11B2 gene [15]. Aldosterone is stimulated by multiple factors: a decrease in blood pressure via the stretch receptors in the heart, acidosis, an increase in circulating angiotensin II (ANG II), or elevated plasma K + concentration. Increases in plasma K + concentrations are the most potent stimulator, and act by depolarizing the cell and opening voltage-gated Ca 2+ channels, which stimulate aldosterone synthase expression and activity. Studies have shown that sustained aldosterone production requires a higher intracellular Ca 2+ concentration [16, 17].

In normal conditions, Adrenocorticotropic Hormone (ACTH) has only a minor role in regulating aldosterone and acts indirectly by stimulating the expression of its precursor molecules.

Aldosterone exerts its effects mainly in the distal tubule and collecting ducts of the nephron. By binding to the nuclear mineralocorticoid receptor of the principal cells, it upregulates and/or activates the basolateral Na+/K- pumps (Na/K-ATPase), leading to greater K + secretion. Aldosterone also upregulates the expression of epithelial sodium channels (ENaC) on the apical membrane of the principal cells of the renal collecting ducts. This leads to an increase in sodium (Na+) reabsorption and allows chloride (Cl -) to be reabsorbed to maintain electrochemical balance. It also stimulates hydrogen (H +) secretion in exchange for K + to regulate pH. Lastly, the urinary loss of K + results in lower plasma K + concentration, thereby indirectly stimulating reabsorption of NaCl by the thiazide-sensitive sodium-chloride cotransporter (NCC) [13].

The zona fasciculate is the middle layer of the cortex. This is the main site of glucocorticoid (cortisol) synthesis. It
is the largest layer, accounting for up to 80% of the volume of the entire adrenal cortex. Cortisol is also a steroid hormone and is involved in regulating aspects of multiple systems that pertain to metabolism, electrolyte balance, stress, and immune responses. It is regulated by the hypothalamus and pituitary glands. Within the paraventricular nucleus of the hypothalamus, Corticotrophin-Releasing Hormone (CRH) is produced. When released into the median eminence, CRH travels through the hypothalamo-hypophyseal portal system to the anterior pituitary. CRH stimulates corticotropes to secrete ACTH, a peptide hormone that stimulates the secretion of glucocorticoids. The ACTH receptor is expressed primarily on adrenocortical cells in the zona glomerulosa. It has rapid effects that stimulate delivery of cholesterol to the mitochondria, for the first step of steroidogenesis via the P450 side chain cleavage enzyme (P450scc). It also stimulates lipoprotein influx into the cortical cells. Chronically, it stimulates the transcription of the genes coding for steroidogenic enzymes (P450scc) and steroid 11β-hydroxylase. It also stimulates the transcription of genes encoding subunits of the mitochondrial oxidative phosphorylation system, which is thought to be necessary for the enhanced energy needs due to ACTH stimulation [18].

Cortisol is synthesized from cholesterol and is the main hormone secreted from the adrenal cortex in humans and many other species. Cortisol is metabolized from its precursor 11-deoxycortisol via the steroid enzyme 11β-hydroxylase in a final step of the pathway. The 11β-hydroxylase, also referred to as cytochrome P450 11B1 and is encoded by the CYP11B1 gene. White et al. (1987) stated that the CYP11B1 gene is present on chromosome 8 and later localized to chromosome 8q21 [19, 20]. 11β-Hydroxylase expression is found on the inner mitochondrial membrane of the zona glomerulosa and the zona fasciculata. It is not only responsible for the metabolism of cortisol, but also for the conversion of deoxycorticosterone to corticosterone, the precursor to aldosterone. Cortisol is also metabolized by the 11β-hydroxysteroid dehydrogenase system (11βHSD) which requires two enzymes: 11β-HSD1, which converts inactive cortisone to active cortisol and requires NADPH as a cofactor; 11β-HSD2, which converts cortisol to cortisone and requires an NAD⁺ cofactor [21]. Cortisol’s main effects on the kidneys include increasing glomerular filtration rate, renal plasma flow, Na⁺ reabsorption, K⁺ secretion and water diuresis.

The zona reticularis is the inner most layer of the cortex and is superficial to the medulla. This layer is responsible for the production of precursor weak androgens such as Dehydroepiandrosterone (DHEA) and androstenedione, from cholesterol.
2.3. Pathophysiology

GRA is a cause of primary hyperaldosteronism. The genes encoding 11β-hydroxylase and aldosterone synthase, CYP11B1 and CYP11B2, respectively, are both on chromosome 8 within close proximity to each other. The two genes encoding each enzyme share 95% sequence homology and have the same intron-exon structures (Fig. 2). During DNA replication in patients with GRA, an unequal crossing over during replication results in a chimeric gene where CYP11B2 comes under the regulation of the promoter regulated by ACTH. Thus, transcription of aldosterone synthase is enhanced by ACTH and leads to increased formation of aldosterone. (A higher resolution / colour version of this figure is available in the electronic copy of the article).

2.4. Clinical Findings, Diagnosis, and Treatment

The most common side effects of GRA include severe hypertension with low plasma renin activity (PRA) and mild hypernatremia. The onset of the disease is often seen early in life. Most GRA patients have normal or only mildly decreased plasma levels of K⁺, despite the apparent hyperaldosteronism effects. The reason why is not well understood. In a retrospective study on 27 GRA pedigrees, early hemorrhagic stroke (EHS) was an associated feature in nearly half of the pedigrees. There were no symptoms of EHS in the GRA negative family members in each pedigree.

GRA is often misdiagnosed as essential hypertension. Early signs of GRA include early onset of hypertension, family history, and refractory hypertension when given standard treatment. Testing for the plasma aldosterone to PRA ratio may help screening test to identify high aldosterone with suppressed PRA [24]. This ratio and a dexamethasone suppression test, used to measure urinary hybrid steroids (18-oxocortisol and 18-hydroxycortisol), can both be used as diagnostic aids to differentiate elevated aldosterone that is abnormally under the influence of ACTH. Direct genetic analysis is the best way to diagnose GRA and treatment can be achieved with multiple approaches. Low dose glucocorticoids are effective as a treatment due to negative feedback on ACTH, thereby removing its stimulation of aldosterone. Mineralocorticoid Receptor (MR) antagonists such as spironolactone or eplerenone are effective alternatives due to the competitive inhibition of the MR therefore suppressing the effect of aldosterone. ENaC antagonists like amiloride have also been used effectively to inhibit ENaC, thereby...
diminishing Na⁺ reabsorption and K⁺ secretion. Antihypertensive drugs affecting the renin-angiotensin system, such as ACE-inhibitors or β-blockers, are ineffective in treating GRA because PRA is already suppressed.

3. LIDDLE’S SYNDROME

Liddle’s syndrome is an autosomal dominant genetic disorder characterized by severe high blood pressure associated with low plasma renin and aldosterone. Very low plasma K⁺ concentrations and metabolic alkalosis are also present.

3.1. Background

Liddle’s syndrome was first described by Liddle et al. (1963) as hypertension associated with hypokalemia that was not caused by hyperaldosteronism but by problems in the distal tubule. Family studies of patients with this disease displayed an autosomal dominant inheritance. Several investigators confirmed the original findings by Liddle and colleagues [25-28]. They discovered that either amiloride or triamterene - both K⁺ sparing diuretics that inhibit ENaC - along with a low Na⁺ diet provide an effective treatment of both the hypertension and hypokalemia. Importantly, they showed that spironolactone, a competitive inhibitor of the Mineralocorticoid Receptor (MR), was not an effective treatment for Liddle’s syndrome.

Botero-Velez et al. (1994) studied a brother and sister who presented with hypertension and hypokalemia with low aldosterone levels. High ratios of Na⁺: K⁺ in sweat and saliva,
and a lack of effect by spironolactone to treat the hypertension excluded mineralocorticoid excess as the cause. They postulated that constitutive activation of any component of ENaC or the MR within the collecting tubule could explain the syndrome [29]. Thirty-five years later, the first in vivo results showing an increase in ENaC activity in a patient with Liddle’s Syndrome by measuring transnasal potential difference were reported. Nasal potential difference measurements are now used as a simple clinical test [30]. Shimkets et al. (1994) discovered a mutation in the beta subunit of ENaC involving a premature stop codon that causes a truncated C terminus in the cytoplasm. This same premature stop or frameshift mutation in the same C terminus domain was identified in four additional family members [31]. Hansson et al. (1995) also showed that Liddle’s Syndrome can also result in truncation of the C terminus of the gamma subunit of ENaC [32]. Truncation of the C terminus increases expression of ENaC on the apical membrane, thereby enhancing Na+ reabsorption by the collecting duct. They discovered a conserved motif in the C terminus of all three subunits of ENaC that, when mutated, produce the effects observed with Liddle’s Syndrome. Results showed that the truncation increased the surface expression of ENaC on the apical membrane [33].

3.2. Physiology

Under normal conditions, hypertension and low potassium would both suppress aldosterone production and secretion (see above).

Within the kidney aldosterone acts on the nuclear MR localized in the collecting duct, specifically in principal cells. Aldosterone upregulates and activates the Na+/K+-ATPase on the basolateral membrane. This creates a gradient for the reabsorption of Na+ from the tubular fluid and secretion of K+ into the lumen of the collecting duct. Uptregulation of ENaC expression on the apical surface of the distal nephron is induced by aldosterone [13, 34]. ENaC plays a vital role in Na+ and K+ homeostasis in coordination with the basolateral Na+/K+-ATPase. The function of ENaC is critical and contributes crucially to the maintenance of total body Na+ content and blood pressure. ENaC structure consists of three different subunits (alpha, beta and gamma) and is a heterotrimeric protein [34]. Each subunit contains two transmembrane helices and an extracellular loop. The extracellular loops contain multiple, highly conserved cysteine residues, proposed to function as receptors. Both the amino- and carboxy-termini are intracellular. The C-terminus contains proline-rich motifs (PPxY). Studies have suggested that ENaC contains a central ion channel along its central symmetry axis between the three subunits. Opening of the ion pathway is thought to involve a coordinated rotation of the second transmembrane spanning domain of each subunit [35]. The complex regulation of ENaC synthesis and trafficking are reviewed thoroughly by Palmer et al. (2012) [36].

Retrieval of ENaC from the apical membrane is also carefully regulated. ENaC is marked for retrieval and degradation by ubiquitination via the E3 ubiquitin ligase, Nedd4-2. The main function of ubiquitin protease system is to degrade marked proteins in the cell by proteolysis. The gene for NEDD4 is highly conserved in all eukaryotes. It is widely expressed in the body and is involved in the regulation of multiple metabolic processes [37]. It binds to ENaC at the C-terminus of the beta and/or gamma subunits to mark it for degradation [38, 39].

3.3. Pathophysiology

Liddle’s syndrome is caused by heterozygous mutations in the SCNN1B and/or the SCNN1G genes located on chromosome 16, which result in a truncated C-terminus on either the beta or gamma subunits of ENaC. The mutant ENaC protein no longer has a binding site for Nedd 4-2 and therefore, cannot mark ENaC for proteolytic degradation.

The result of the inhibited breakdown of ENaC leads to increased ENaC expression on the apical membrane which causes increased Na+ reabsorption. This correlates with the severe hypertension seen in patients with Liddle’s syndrome [31]. The increase in Na+ entry into the principal cell activates the Na+/K+-ATPase, resulting in increased flux of K+ into the cell at the basolateral side thereby facilitating K+ secretion into the lumen (Fig. 3). The result is low urinary Na+ and high urinary K+ excretion culminating in hypertension and hypokalemia. The influx of Na+ also leads to a more negative lumen potential that is permissive for H+ secretion by nearby type A intercalated cells and results in metabolic alkalosis.

3.4. Clinical Findings, Diagnosis, and Treatment

Patients with Liddle’s Syndrome have salt-sensitive hypertension that develops early in childhood, hypokalemia, metabolic alkalosis, low PRA and low aldosterone. Patients are recommended to eat a low Na+ diet and are treated with a K+ sparing diuretic such as amiloride or triamterene. Both amiloride and triamterene are direct inhibitors of ENaC and therefore block Na+ reabsorption via the channel. By inhibiting Na+ influx, the hypokalemia is also alleviated though patients may require potassium supplementation as well. The increased arterial pressure suppresses aldosterone via negative feedback, and due to this, competitive MR inhibitors of aldosterone such as spironolactone are not an effective treatment.

4. APPARENT MINERALOCORTICOID EXCESS

AME is an autosomal recessive disease that results in hypertension and hypokalemia with low levels of renin and aldosterone.

4.1. Background

Initial publications of documented clinical cases that presented with symptoms of AME began in the 1970’s, with patients displaying effects of mineralocorticoid excess but with low to undetectable levels of aldosterone [40, 41]. The hypertensive symptoms in these cases responded well to diuretics such as triamterene and spironolactone. Studies showed an increase in the half-life of cortisol, implying a deficiency in the enzyme 11β-Hydroxysteroid Dehydrogenase type 2 (11βHSD2) that converts cortisol to its less active form, cortisone [42, 43]. It was believed that AME results in a lack of specificity by the Mineralocorticoid Receptor (MR) [44]. Several studies have shown that the MR has equal af-
finity to both cortisol and aldosterone in vitro [44, 45]. In studies done in vivo, the MR has substantially higher affinity for aldosterone. Finally, in 1983, it was proposed that cortisol was the mineralocorticoid that induced hypertension. Patients later diagnosed with AME were treated with the glucocorticoid hormone, hydrocortisone, but unexpectedly experienced aggravated effects of hypertension and hypokalemia [43].

Studies were conducted to identify 11βHSD as the cause of AME. The search first led to the discovery of 11βHSD1 in rat liver homogenates. 11βSD1 is also an oxoreductase but favors the conversion of cortisone to cortisol. No mutations were found on the gene encoding 11βHSD1 in patients with AME. Additional studies done on the distal nephron in rabbits showed a separate NAD-dependent isoform of 11βHSD. In 1994, cDNA-encoding 11βHSD2 was isolated and cloned in kidney cells of sheep, humans and rabbits. One year later, the first identification of a mutation on the 11βHSD2 gene was discovered within an Iranian mutations have since been discovered in the 11βHSD2 gene that abolishes enzyme activity, resulting in AME [46, 47].

AME is very rare. In the past 25 years less than 100 patients worldwide have been diagnosed with the disease. Such rare autosomal recessive mutations are usually explained by endogamy, consanguinity, or the founder effect. A study done in 1995 showed that seven of the eight families with AME appeared to conform to one of the three explanations [48].

4.2. Physiology

In the kidneys, active circulating cortisol is degraded to its inactive form cortisone via 11βHSD2 [49]. Cortisol is a steroid hormone produced in the zona fasciculata of the adrenal gland. Its release is regulated by stress and low blood glucose levels. Its functions include stimulating gluconeogenesis and anti-inflammatory pathways and regulating electrolyte balance. Within the kidneys, cortisol activates its glucocorticoid receptor which stimulates water diuresis, glomerular filtration rate, and renal plasma flow [50]. Cortisol is oxidized by 11βHSD2, which is from a family of oxidoreductases that act on donor hydroxyl groups and converts them to keto-groups, forming cortisone. This reaction is dependent on the ratio of NADP+/NADPH. 11βHSD2 is transcribed from the HSD11B2 gene, located on chromosome 16q22. The gene is approximately 6kb in length containing 5 exons. The two isoforms of 11βHSD only contain 20% sequence homology, suggesting they belong to different gene families. 11βHSD2 is colocalized with the MR in the distal tubules and collecting ducts of the nephron and is specifically localized in the endoplasmic reticulum of principal cells as well as in epithelial tissues and in the brainstem. In contrast to cortisol, cortisone binds very poorly to MR. Thus, by facilitating the conversion of cortisol to cortisone, 11βHSD2 activity in the principal cell prevents activation of MR by cortisol such that the MR in the distal nephron responds almost exclusively to aldosterone.

4.3. Pathophysiology

AME is due to a deficiency in the enzyme 11βHSD2, resulting in an increase in serum cortisol (Fig. 3). Circulating cortisol levels are normally 100-to 1000-fold higher than those of aldosterone, particularly in the morning. Failure of cortisol conversion to cortisone results in inappropriate activation of the MR by cortisol [51]. When the MR is activated by mineralocorticoids and even glucocorticoids, clinical signs of hyperaldosteronism ensue. The MR is expressed in many tissues: kidney, CNS, heart and sweat glands. In the principal cells of the kidney, MR activation specifically increases Na⁺ reabsorption through ENaC, leading to an increase in extracellular volume. K⁺ secretion is concurrently increased within these cells via the Renal Outer Medullary K⁺ channel (ROMK) for the combined effect hypertension and hypokalemia. The resultant change in intratubular electrical potential also facilitates H⁺ secretion by adjacent α-intercalated cells, leading to enhanced bicarbonate reabsorption and metabolic alkalosis [52-54]. AME-like clinical presentation can also be induced by ingesting large amounts of licorice, which contains glycyrrhizic acid that is hydrolyzed to glycyrrhetinic acid. In turn, glycyrrhetinic acid is metabolized into 3-beta-D-(monoglucuronyl)-18-beta-glycyrrhetinic acid which is capable of inhibiting 11βHSD2. The increased levels of cortisol in the collecting duct leading to MR activation and an “acquired” form of AME. Notably, glycyrrhizic acid is absent from licorice produced in the United States, but may still be found in licorice flavored chewing tobacco or imported licorice [55].

4.4. Clinical Findings, Diagnosis, and Treatment

AME presents early on in life with clinical features of low birth weight and severe hypertension. It is associated with metabolic alkalosis and severe hypokalemia. Diagnosis can be made by measuring the ratio of urinary cortisol metabolites, tetrahydrocortisol and allotetrahydrocortisol to the urinary concentration of tetrahydrocortisone. A normal ratio is 1:1. In a patient with AME, ratios ranged from 6.7 to 33 [56]. The optimal diagnostic test is the measurement of urine samples following 11-1 tritiated cortisol injection. In patients with AME, the infused cortisol is converted to detectable cortisone only 0-6% of what is observed in healthy individuals. However, this technique is not widely used due to the rarity of tritiated cortisol. Patients with AME are favorably treated with spironolactone to lower Na⁺ reabsorption and K⁺ secretion.

5. GELLER SYNDROME

Geller Syndrome is a form of hypertension caused by a heterozygous mutation of the MR that leads to altered nuclear receptor ligand selectivity and activation. In Geller syndrome, steroid hormones such as progesterone have increased affinity for the MR, leading to enhanced activation of mineralocorticoid signaling cascades that increase Na⁺ reabsorption and K⁺ secretion. Patients with this mutation often present at a young age with hypertension, diminished PRA, and low serum aldosterone. During pregnancy when progesterone levels are dramatically elevated, this mutation induces severe hypertension.

5.1. Background

Pregnancy-related hypertension results in complications in 6-10% of pregnancies and leads to increases in maternal
and perinatal mortality [57]. Hypertension occurring during gestation is characterized by one of four conditions: pre-existing hypertension, gestational hypertension and pre-eclampsia, pre-existing hypertension plus superimposed gestational hypertension with proteinuria and unclassified hypertension. In many of these cases, delivery of the infant resolves symptoms, suggesting pregnancy-specific factors as a pathologic source. Geller et al. (2000) tested the hypothesis that gain-of-function mutations in the MR could induce hypertension. Seventy-five patients with early onset of hypertension were screened, of which a 15-year-old boy with severe idiopathic hypertension was found to be heterozygous for a missense mutation in the MR hormone binding domain. Eleven of 23 relatives of this child were evaluated and were found to have been diagnosed with severe hypertension before the age of 20. Three individuals died due to heart failure before the age of 50. The early onset of hypertension in this pedigree co-segregated with the MR mutation, strongly suggesting a Mendelian inheritance pattern.

5.2. Physiology

As described in prior sections, the binding of aldosterone to the MR in the principal cell of the kidney tubules leads to translocation to the nucleus where it binds to the hormone response element of the promoter region of genes regulated by aldosterone. The result is increased expression of ENaC and Na/K-ATPase. The Glucocorticoid Receptor (GR) and MR possess close structural similarity such that cortisol has high affinity for both GR and MR, indicating common ancestry [58]. However, aldosterone demonstrates clear MR specificity. To study the specificity of MR to aldosterone, Rogerson et al. created a series of chimeras from both receptors. Using this strategy, they identified amino acids 804-874 of the MR ligand-binding domain that were essential for aldosterone binding. Further analysis of these chimeras from this group and others found that the amino acid region 820-844 on the surface of the molecule determines ligand-binding specificity for each receptor subtype. Site-directed mutagenesis found 12 of the 16 amino acids that differ between GR and MR in the 820-844 region were responsible for directing specificity to aldosterone [59].

5.3. Pathophysiology

Wild type MR (MRWT) requires a steroid 21-hydroxyl group to permit binding and activation by steroid hormones such as aldosterone - a group that is not found on the progesterone molecule. Ligand binding induces various conformational changes involving the bending of several receptor helices to form a surface for coactivator binding [60]. A point mutation causing substitution of a leucine for a serine at codon 810 (S810L; MR L810), located on chromosome 4q31 alters the conformation of the hormone-binding domain. Subsequent studies indicate that the substituted lysine in MR L810 permits bending of the side-chain that forms van der Waals interactions when bound by smaller steroids such as progesterone. These interactions significantly increase binding affinity when compared to that of aldosterone. Progesterone levels typically increase extensively during pregnancy, suggesting that the MR L810 mutation could result in increases in ENaC and Na/K-ATPase expression and activity, thereby augmenting Na+ reabsorption and provoking gestational hypertension (Fig. 3). Indeed, in one such pregnant patient blood pressure rose to levels of 210/120 mmHg with low serum potassium levels low and nearly undetectable aldosterone levels.

In men and non-pregnant women, the severe hypertension observed with the MR L810 mutation is not likely explained by elevated progesterone levels. Cortisol levels are significantly more abundant than aldosterone in men and bind the MR with similar affinity [61, 62]. However, in healthy individuals, MR activation is prevented by metabolism of cortisol to corticosterone. Interestingly, in vitro analysis of MR L810 activity in the presence of cortisol by-products clearly demonstrated the ability of corticosterone and 11-dehydrocorticosterone to bind and activate the MR with high affinity. These results suggest cortisol by-products as the pathologic source in men and non-pregnant women with the MR L810 mutation [63].

5.4. Clinical Findings, Diagnosis and Treatment

Patients with Geller syndrome need to consume a low salt diet. Pregnancy should be closely monitored. Due to the altered binding parameters of the ligand binding domain, MR L810 has increased affinity for the traditional MR antagonist, spironolactone. This may result in paradoxical activation of the MR, inducing an agonist reaction and exacerbating the hypertension and electrolyte disturbances.

6. PSEUDOHYPOALDOSTERONISM TYPE II

PHA is a heterogeneous group of genetic disorders involved in electrolyte balance. Although, there are multiple types and subsets of PHA, this review focuses on PHA-II, also known as Gordon’s syndrome, since it is associated with hypertension. PHA-II is an autosomal dominant disorder characterized by hypertension, hyperkalemia and metabolic acidosis. It is associated with both low renin and low aldosterone levels [64-67]. In contrast, PHA-I results in salt wasting and hypotension, which is beyond the focus of this review. For more information on PHA-I, the reader is referred to excellent recent reviews [68, 69].

6.1. Background

In 1970, Gordon et al. reported a case of hyperkalemia and hypertension with low PRA and aldosterone that responded to dietary salt restriction [70]. Brautbar et al. (1978) treated a similar patient who had four family members with the same symptoms. The inability to increase K+ excretion by exogenous mineralocorticoid treatment pointed to a defect in K+ regulation in the distal nephron. Treatment with an ion exchange resin reduced the hyperkalemia and ameliorated the hyperchloremic acidosis [71]. Several other reports followed [72-75]. Schambelan et al. (1981) first referred to Gordon’s syndrome as PHA-II; a syndrome of chronic mineralocorticoid-resistant hyperkalemia and hypertension [76]. Pasman et al. (1989) studied a 14-year-old boy with Gordon’s syndrome that presented with secondary hyperkalemic periodic paralysis. Treatment with hydrochlorothiazide normalized plasma K+ levels, aldosterone and PRA and cured the periodic paralysis [77]. Take et al. (1991) studied a
Japanese family with three affected members and were the first to suggest increased NaCl reabsorption in the thiazide-sensitive segment of the nephron and autosomal dominant transmission [78], later shown to be a linkage disorder with chromosome 1 and 17 [65].

Wilson et al. (2001) identified two genes that associated with PHA-II. These genes encoded isoforms of the with-no-lysine (WNK) family of serine-threonine kinases, WNK1 and WNK4. Identification of these proteins propelled research and precipitated advances in the understanding of their actions within the kidney. Further research lead to the discovery of two additional proteins, CUL3 and KLHL3, that are involved in the proteasomal degradation of WNK proteins. Mutations in these have also been associated with PHA-II [66, 67].

6.2. Physiology

The WNK1 gene is located on chromosome 12 and produces four alternatively spliced isoforms: L-WNK1 and KS-WNK1 are most relevant to Gordon’s syndrome. L-WNK1 is the full-length isoform and is expressed throughout the body. Kidney-specific WNK1 (KS-WNK1) is expressed exclusively in the kidneys [79, 80]. Both proteins help maintain electrolyte homeostasis by regulating the thiazide transporter, NaCl-cotransporter (NCC) and/or the renal outer medullary potassium channel (ROMK). NCC is expressed apically in the distal tubules and is responsible for the reabsorption of ~5-10% of filtered NaCl. It is inhibited by thiazides. ROMK is an ATP-dependent K⁺ channel expressed in the thick ascending limb and in the principal cells of the collecting ducts. It is involved in K⁺ recycling and K⁺ secretion, respectively [81, 82].

The regulation of Na⁺ reabsorption and K⁺ secretion by the WNK kinases is complex and the mechanisms continue to be updated. L-WNK1 activates SPS-1-related proline/alanine-rich kinase (SPAK) via phosphorylation. In turn, SPAK phosphorylates NCC thereby increasing its trafficking and membrane expression [83]. By stimulating NCC activity, WNK1 promotes increased NaCl reabsorption. Moreover, L-WNK1 directly inhibits ROMK activity in the collecting duct. Inhibition of ROMK promotes decreased K⁺ secretion [84, 85]. KS-WNK1, in turn, can inhibit L-WNK1 and L-WNK1 can interact with WNK4 (via a conserved HQ domain), to promote further “fine tuning” of the intracellular regulatory mechanisms controlling Na⁺ and K⁺ handling by this nephron segment [86].

The gene encoding WNK4 is located on chromosome 17. The function of WNK4 has been more difficult to elucidate due to apparent paradoxical effects. WNK4 stimulates NCC and inhibits surface expression of ROMK via a dynamin-dependent mechanism that does not require ubiquitination [85, 87]. WNK4 has also been implicated in increasing paracellular Cl⁻ reabsorption by phosphorylation of claudins [88-91]. WNK4 also regulates ENaC, but its major effects appear to be on NCC as a knockout of WNK4 results in a Gitelman-like phenotype [92]. Notably, intracellular Cl⁻ concentration modulates WNK4 function via a chloride-sensing motif [93]. WNK4 maintains basal phosphorylation of NCC via SPAK. Under conditions of low intracellular Cl⁻, NCC is stimulated whereas high intracellular Cl⁻ inhibits. Moreover, membrane potential at the apical surface of the distal convoluted tubule cells (related to extracellular K⁺) also plays a role. Mutations in WNK1 or WNK4 have been identified in a relative small number of individuals with PHA-II. This observation suggested the presence of mutations in other intracellular proteins. The ubiquitin proteasome system is a protein degradation pathway that is critical to processes such as cell cycle progression, signal transduction, protein expression and quality control [87]. Proteolytic specificity occurs via a protein complex comprised of an E3-ubiquitin ligase, a Cullin protein and an adaptor protein. The Cullins provide a scaffold for E3 ligase subunits, which are enzymes that confer substrate specificity and are required for ubiquitination of targeted proteins. In turn, the Cullin proteins interact with a family of adaptor proteins. More specifically, Cullin3 (CUL3) binds to a domain on Kelch-like protein 3 (KLHL3), a subclass of complex/tramtrack/bric-a-brac proteins. Within the nephron, CUL3 binds with KLHL3. Kelch domains within KLHL3 bind to WNK proteins and target them for degradation [94-98].

6.3. Pathophysiology

Direct mutations in the WNK1 and WNK4 genes are associated with PHA-II, but account for only a small number of reported cases (Fig. 4). Mutant WNK1 genes lead to overexpression of L-WNK1. The resultant activation of SPAK leads to enhanced phosphorylation of NCC, increased NaCl reabsorption and hypertension. Overexpression of WNK1 can inhibit WNK4 activity, further promoting additional NCC phosphorylation and NaCl reabsorption. Concomitant inhibition of ROMK results in decreased K⁺ secretion and hyperkalemia.

Mutations in WNK4 genes tend to cluster within a non-catalytic domain that is otherwise highly conserved and is critical for binding to KLHL3. Disruption of the binding to KLHL3, ubiquitination and subsequent proteolysis are impaired leading to increased levels of WNK4 and downstream phosphorylation of NCC via SPAK. Together with simultaneous inhibition of ROMK, these mutations in WNK4 also lead to the PHA-II phenotype [99-101].

Mutations in CUL3 or KLHL3 also result in PHA-II. The pathologic changes in the CUL3 gene to date have resulted in variants that impair splicing of exon 9. There are 16 currently known mutations to CUL3 which ultimately result in the aberrant splicing of exon 9 and the subsequent in-frame fusion of exons 8 and 10. The product is a protein lacking 57 amino acids that is unable to develop ubiquitin-ligase complexes or bind substrates. The resultant in-frame deletion of this motif in CUL3 disrupts proteolytic degradation of the WNKs. Likewise, pathologic dominantly inherited changes in the KLHL3 gene tend to be missense mutations that produce an abnormal C-terminal Kelch propeller domain that alters substrate binding. In contrast, recessively inherited KLHL3 mutations are more widely distributed along the gene and include frameshift or truncation of the protein. All in all, these variants disrupt binding to CUL3, WNK4 or WNK1 such that WNK4 levels are increased due to attenuated proteolysis. The mechanisms have been discussed in greater detail in the recent review by Murthy et al. [98].
6.4. Clinical Findings, Diagnosis and Treatment

Importantly, the clinical phenotype of patients with PHA-II, or Gordon’s syndrome, is the same for mutations in any of the four proteins: WNK1, WNK4, CUL3 or KLHL3. Mutations in CUL3, however, are associated with a more severe presentation with higher blood pressures, more profound hyperkalemia and acidosis and an earlier onset [66]. Dramatic improvement in PHA-II is observed with inhibition of NCC by thiazide diuretics. A Na+ and K+ restricted diet is also advisable [75, 76, 98].

7. AUTOSOMAL DOMINANT HYPERTENSION WITH BRACHYDACTYLITY

Autosomal dominant hypertension with brachydactyly (ADHB) is a rare autosomal dominant disorder characterized by brachydactyly type E, short stature and severe age-dependent hypertension. When left untreated, death due to stroke often occurs before the age of 50.

7.1. Background

Bilginturan et al. (1973) first described a Turkish family with brachydactyly involving the metacarpal and phalangeal bones that co-segregated 100% with hypertension. An extensive pedigree of the family revealed an autosomal dominant inheritance pattern [102]. More than twenty years later, investigation of six members from the same family, five affected and one unaffected individuals, revealed that the affected patients were not salt sensitive and had normal renin, aldosterone and catecholamnergic responses to volume depletion and volume expansion. Investigators were able to pinpoint the disorder to chromosome 12p12.2-p11.2 by linkage analysis. Further studies of the original pedigree as well as several other reports from families unrelated to the original pedigree showed mutations in the same gene [102-104]. Toka et al. (1998) also reported that the brachydactyly was type E; due to the association with short stature and severe hypertension [105].

Magnetic resonance imaging (MRI) with angiography identified unilateral or bilateral posterior inferior cerebellar artery or vertebral artery loops contacting the ventrolateral medulla in the posterior fossa in all affected patients. Unaffected individuals did not display these loops. They proposed that the loops could be responsible for hypertension [105].

Linkage analysis of the hypertension with brachydactyly and the neurovascular anomaly with chromosome 12 markers revealed a LOD score of 9.2; meaning the odds that the two traits being linked are better than 1,000,000,000:1. Gong et al. (2003) performed a genome-wide parametric linkage analysis and discovered a locus for essential hypertension on chromosome 12 (HYT4), that overlaps with the locus associated with ADHB. They proposed two candidate genes as the underlying cause of the hypertension: phosphodiesterase 3A (PDE3A) and the sulfonlylurea...
(PDE3A) and the sulfonylurea receptor 2 (SUR2) which is a subunit of an ATP-sensitive K⁺ channel [109].

7.2. Physiology

When the PDE3A gene is normally expressed, it encodes phosphodiesterase 3A, a member of the cGMP-inhibited cyclic nucleotide phosphodiesterase (cGI-PDE) family. The cGI-PDEs are capable of hydrolyzing both cGMP and cAMP and are expressed prominently in platelets, oocytes, heart and vascular smooth muscle cells (VSMC). These enzymes play a vital role in many cellular processes by regulating the duration and amplitude of intracellular cyclic nucleotide signals [110]. Low levels of cAMP can promote cell proliferation. PDEs also can mediate platelet aggregation and regulate vascular smooth muscle contraction, arteriogenesis and arterial remodeling [111].

PDEs are also involved during early stages of osteogenesis [112]. Studies show PDE3A expression in developing limbs and chondrogenesis in mice [113]. Parathyroid related protein (PTHrP) is a member of the parathyroid hormone family that acts as an autocrine, endocrine, intracrine and paracrine hormone. PTHrP can inhibit VSMC proliferation and regulate endochondral bone development and growth by maintaining the endochondral growth plate at a constant width. Endochondral ossification is one of the two essential processes in fetal development of the mammalian skeletal system, vital in the rudimentary formation, growth and healing of long bones. PDEs share an identical N-terminus as PTH and can stimulate most of the effects of PTH, including enhanced bone resorption and distal tubule Ca⁺⁺ reabsorption, while inhibiting proximal tubular phosphate transport [114, 115].

7.3. Pathophysiology

Bahring et al. (2008) studied all affected members of the four Turkish, Canadian and American families and showed that all had mutations on chromosome 12. They predicted the single exon is a stem-loop structure essential in noncoding RNA processing [116].

Maass et al. (2015) performed whole-exome sequencing on the Turkish family and identified a heterozygous missense mutation in the PDE3A gene. Studies on six different families revealed six independently clustered heterozygous missense mutations in exon 4. The altered amino acids at residues: 445, 447 and 449, belong to a highly conserved domain. Adjacent to the mutations are two residues, Ser428 and Ser438, which can be activated by either protein kinase A or protein kinase C. This gain of function results in increased cAMP hydrolysis and lowered cAMP levels. Functional analysis revealed the mutations increase protein kinase A-mediated PDE3A phosphorylation. This results in a gain of function in PDE3A activity. The increase in cAMP hydrolysis causes reduced levels of phosphorylated vasodilator-stimulated phosphoprotein, [110, 117, 118]. In vitro studies show an increased expression of smooth muscle actin alpha, calponin and transgelin in mesenchymal stem-cell-derived

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Fig. (5). Diagnostic approach to secondary hypertension with emphasis on monogenic hypertensive disorders. Diagnostic tests are summarized in Table 1. *Note that familial forms of pheochromocytoma have also been associated with genetic changes. All are autosomal dominant and are listed here with their respective genes: Von-Hippel-Lindau (VHL), multiple endocrine neoplasia type 2 (RET-proto-oncogene), neurofibromatosis type 1 (NF1) and familial paraganglioma (SDH and its subunit genes). (A higher resolution / colour version of this figure is available in the electronic copy of the article).
VSMCs from affected individuals. Thus, enhanced activity of PDE3A promotes lower cAMP levels in VSMC which, in turn, increases neointimal proliferation and remodeling of the arteries and neurovascular structures. Hypertension ensues from the attendant increases in vasoconstriction and peripheral vascular resistance.

Although platelet function was not altered, dysregulation of parathyroid-related protein (PTHrP) which regulates epithelial-mesenchymal interactions and endochondral bone development was observed. They hypothesized that these mutations cause the hypertension by contributing to an increase in peripheral vascular resistance along with the characteristic skeletal changes [112, 119].

7.4. Clinical Signs, Diagnosis and Treatment

Patient with ADHB typically present with short stature, severe salt-independent hypertension and altered baroreflex regulation. They will also suffer from type E brachydactyly, which is characterized by thickening and shortening of metacarpals and phalanges. The disease can be hard to diagnose in prepubescent children. Neurovascular contact at the rostroventrolateral medulla is also associated with ADHB and likely contributes to the barodysregulation [108, 120]. Hypertension occurs in childhood and progresses in a time-dependent manner. Stroke is typically associated with untreated ADHB, with early mortality. Diagnosis is typically made during childhood and early treatment can reduce the chances of stroke [121]. Studies have shown significant reduction of hypertension in combination or single drug therapy with beta-blockers, alpha-blockers, Ca^2+ channel blockers, or an ACE inhibitor. Measures to increase cyclic GMP to offset the cAMP deficiency are being studied but not yet approved [122].

### CONCLUSION

Hypertension remains a major health issue as the leading cause of cardiovascular morbidity and mortality in the U.S. and the world. In the overwhelming number of cases, the mechanism(s) underlying pathological sustained elevations in arterial pressure remain unknown. Table 1 summarizes the six categories of monogenic hypertension discussed in this review and their clinical and diagnostic parameters. In addition, Fig. 5 shows a simplified diagnostic approach for identifying individuals who may potentially possess one of these genetic forms of hypertension. Ultimately, genetic confirmation is the only definitive test.

Although these disorders may encompass only a small proportion of individuals suffering from hypertension, they provide invaluable insights into the genetics and complex regulatory systems involved in blood pressure regulation. They have also shed light on some of the cellular functions that underscore hypertension that may be multifactorial. These unique pathologic states have provided a framework for the discovery of additional mechanisms of hypertension and innovative targets for therapeutic development.

### LIST OF ABBREVIATIONS

- **ACE**: Angiotensin Converting Enzyme
- **ACTH**: Adrenocorticotropic Hormone
- **ADHB**: Autosomal Dominant Hypertension with Brachydactyly
- **AME**: Apparent Mineralocorticoid Excess
- **ANG II**: Angiotensin II
- **CUL3**: Cullin 3
CONSENT FOR PUBLICATION

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CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

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REFERENCES

[1] Lewington S, Clarke R, Qizilbash N, Peto R, Collins R. Prospective Studies C. Prospective Studies Collaboration. Age-specific relevance of usual blood pressure to vascular mortality: A meta-analysis of individual data for one million adults in 61 prospective studies. Lancet 2002; 360(9349): 1903-13. http://dx.doi.org/10.1016/S0140-6736(02)11911-8 PMID: 12493255

[2] Carretero OA, Oparil S. Essential hypertension. Part I: definition and etiology. Circulation 2000; 101(3): 329-35. http://dx.doi.org/10.1161/01.CIR.101.3.329 PMID: 10645931

[3] Nwankwo T, Yoon SS, Burt V, Gu Q. Hypertension among adults in the United States: National Health and Nutrition Examination Survey, 2011-2012. NCHS Data Brief 2013; 133(133): 1-8. PMID: 24171916

[4] Mozaffarian D, Benjamin EJ, Go AS, et al. American Heart Association Statistics Committee and Stroke Statistics Subcommittee. Heart disease and stroke statistics—2015 update: A report from the American Heart Association. Circulation 2015; 131(4): e29-e322. http://dx.doi.org/10.1161/CIR.0000000000000152 PMID: 25520374

[5] Sutherland DJ, Ruse JL, Laidlaw JC. Hypertension, increased aldosterone secretion and low plasma renin activity relieved by dexamethasone. Can Med Assoc J 1966; 95(22): 1109-19. PMID: 4288576

[6] Salti IS, Stiefel M, Ruse JL, Laidlaw JC. Non-tumorous “primary” aldosteronism. J. Type relieved by glucocorticoid (glucocorticoid-remediable aldosteronism). Can Med Assoc J 1969; 101(1): 1-10. PMID: 5793351

[7] Ganguly A, Grim CE, Weinberger MH. Anomalous postural aldosterone response in glucocorticoid-suppressible hyperaldosteronism. N Engl J Med 1981; 305(17): 991-3. http://dx.doi.org/10.1056/NEJM198110223051706 PMID: 6268979

[8] Gordon RD. Heterogeneous hypertension. Nat Genet 1995; 11(1): 6-9. http://dx.doi.org/10.1038/ng0995-6 PMID: 7550315

[9] Gates LJ, MacConnachie AA, Lifton RP, Haines NE, Benjamin N. Variation of phenotype in patients with glucocorticoid remediable aldosteronism. J Med Genet 1996; 33(1): 25-8. http://dx.doi.org/10.1136/jmg.33.1.25 PMID: 8825044

[10] Stowasser M, Huggard PR, Rossetti TR, Bachmann AW, Gordon RD. Biochemical evidence of aldosterone overproduction and abnormal regulation in normotensive individuals with familial hyper-aldosteronism type I. J Clin Endocrinol Metab 1999; 84(11): 4031-6. http://dx.doi.org/10.1210/jcem.84.11.6159 PMID: 10566645

[11] Stowasser M, Bachmann AW, Huggard PR, Rossetti TR, Gordon RD. Severity of hypertension in familial hyperaldosteronism type I: Relationship to gender and degree of biochemical disturbance. J Clin Endocrinol Metab 2000; 85(6): 2160-6. http://dx.doi.org/10.1210/jc.85.6.2160 PMID: 10852446

[12] Mulatero P, di Cella SM, Williams TA, et al. Glucocorticoid remediable aldosteronism: Low morbidity and mortality in a four-generation italian pedigree. J Clin Endocrinol Metab 2002; 87(7): 3187-91. http://dx.doi.org/10.1210/jc.87.7.8647 PMID: 12107222

[13] Nussey S, Whitehead S. Endocrinology: An integrated approach. Oxford 2001. PMID: 20821847

[14] Cumrow KM, Tusie-Luna MT, Pascoe L, et al. The product of the CYP11B2 gene is required for aldosterone biosynthesis in the human adrenal cortex. Mol Endocrinol 1991; 5(10): 1513-22. http://dx.doi.org/10.1210/mend-5-10-1513 PMID: 1775135

[15] Bassett MH, White PC, Rainey WE. The regulation of aldosterone synthase expression. Mol Cell Endocrinol 2004; 217(1-2): 67-74. http://dx.doi.org/10.1016/j.mce.2003.10.011 PMID: 15134803

[16] Williams GH, Dluhy RG. Aldosterone biosynthesis. Interrelationship of regulatory factors. Am J Med 1972; 53(5): 595-605. http://dx.doi.org/10.1016/0002-9344(72)90156-8 PMID: 3432886

[17] Brown RD, Strott CA, Liddle GW. Site of stimulation of aldosterone biosynthesis by angiotensin and potassium. J Clin Invest 1972; 51(6): 1413-8. http://dx.doi.org/10.1172/JCI106937 PMID: 4336939

[18] Hanukoglu I, Feuchtwanger R, Hanukoglu A. Mechanism of corticoterpin and cAMP induction of mitochondrial cytochrome P450 system enzymes in adrenal cortex cells. J Biol Chem 1990; 265(33): 20602-8. PMID: 2173715

[19] Chua SC, Szabo P, Vetek A, Grzeschik KH, John M, White PC. Cloning of cDNA encoding steroid 11 beta-hydroxylase (P450sc11). Proc Natl Acad Sci USA 1987; 84(20): 7193-7. http://dx.doi.org/10.1073/pnas.84.20.7193 PMID: 3499608

[20] White PC. Genetics of steroid 21-hydroxylase deficiency. Recent Prog Horm Res 1987; 43: 305-36.
the metabolic syndrome: A hypothesis. J Clin Endocrinol Metab 2009; 94(8): 2692-701.

http://dx.doi.org/10.1210/jc.2009-0370 PMID: 19470627

[52] Bailey MA, Unwin RJ, Shirley DG. In vivo inhibition of renal 1/beta-hydroxysteroid dehydrogenase in the rat stimulates collecting duct sodium reabsorption. Clin Sci (Lond) 2001; 101(2): 195-8. http://dx.doi.org/10.1042/cs1010195 PMID: 11473496

[53] Gaeggerer HP, Gonzalez-Rodriguez E, Jaeger NF, et al. Mineralocorticoid versus glucocorticoid receptor occupancy mediating aldosterone-stimulated sodium transport in a novel renal cell line. J Am Soc Nephrol 2005; 16(4): 878-91. http://dx.doi.org/10.1618/asn.2004121110 PMID: 15743993

[54] Choi KB. Hypertensive hypothalamic disorders. Electrolyte Blood Press 2007; 5(1): 34-41. http://dx.doi.org/10.5049/EBP.2007.5.1.34 PMID: 24459498

[55] Faresne RV, Biglieri EG, Shackleton CH, Irony I, Gomez-Fuentes R. Liricorice-induced hypermineralocorticoidism. N Engl J Med 1991; 325(17): 1223-7. http://dx.doi.org/10.1056/NEJM1991102423251706 PMID: 1922210

[56] Palermo M, Quinkler M, Stewart PM. Apparent mineralocorticoid excess syndrome: An overview. Aq Bras Endocrinol Metabol 2004; 48(5): 687-96. http://dx.doi.org/10.1590/S0004-27302004000500015 PMID: 15761540

[57] WHO Recommendations for Prevention and Treatment of Pre-Eclampsia and Eclampsia. Geneva: WHO Guidelines Approved by the Guidelines Review Committee 2011.

[58] Baker ME. Adrenal and sex steroid receptor evolution: environmental implications. J Mol Endocrinol 2001; 26(2): 119-25. http://dx.doi.org/10.1681/ASN.2004121110 PMID: 15743993

[59] Roy C. Familial pseudohypoaldosteronism (apropos of 5 cases). Afr Fr Pediatr 1977; 34(1): 37-54. PMID: 30001368

[60] Lampal M, Rappaport R, Dechaux M, Morin C. Familial dominant pseudohypoaldosteronism. Lancet 1978; (18054): 51. http://dx.doi.org/10.1016/S0021-9738(78)90404-X PMID: 75436

[61] Lee MR, Morgan DB. Familial hyperkalaemia responsive to benzothiadiazine diuretic. Lancet 1980; 1(8173): 879. http://dx.doi.org/10.1016/S0140-6736(80)91378-1 PMID: 6103235

[62] Licht JH, Amundson D, Hseuh WA, Lombardo FY. Familial hyperkalemic acidosis. J Clin Endocrinol Metab 1985; 54(214): 161-76. PMID: 3885297

[63] Schambelan M, Sebastian A, Rector FC Jr. Mineralocorticoid-resistant renal hyperkalaemia without salt wasting (type II pseudohypoaldosteronism): role of increased renal chloride reabsorption. Kidney Int 1981; 19(5): 716-27. http://dx.doi.org/10.1038/ki.1981.72 PMID: 7026872

[64] Pasman JW, Gabreëls FJ, Semmekrot B, Renier WO, Monnens LA. Hyperkalaemic periodic paralys in Gordon’s syndrome: A possible defect in atrial natriuretic peptide function. Ann Neurol 1989; 26(3): 392-5. http://dx.doi.org/10.1002/ana.410260314 PMID: 2529811

[65] Take C, Ikeda K, Kurasawa T, Kurokawa K. Increased chloride reabsorption as an inherited renal tubular defect in familial type II pseudohypoaldosteronism. N Engl J Med 1991; 324(7): 472-6. http://dx.doi.org/10.1056/NEJM199102143240707 PMID: 1988833

[66] Xu B, English JM, Wilsbacher JL, Stippec S, Goldsmith EJ, Cobb MH. WKIN, a novel mammalian serine/threonine protein kinase lacking the catalytic lysine in subdomain II. J Biol Chem 2000; 275(22): 16795-801. http://dx.doi.org/10.1074/jbc.275.22.16795 PMID: 10828064

[67] Wang Z, Yang CL, Ellison DH. Comparison of WKIN and WKIN kinase and inhibiting activities. Biochem Biophys Res Commun 2004; 317(3): 939-44. http://dx.doi.org/10.1016/j.bbrc.2004.03.132 PMID: 15081430

[68] Faure S, Delaloy C, Leprivey V, et al. WKIN kinases, distal tubular ion handling and hypertension. Nephrol Dial Transplant 2003; 18(12): 2463-7. http://dx.doi.org/10.1093/ndt/gfg526 PMID: 14605263

[69] Hebert SC, Gamba M. Molecular cloning and characterization of the renal diuretic-sensitive electronegative sodium-potassium)-chloride cotransporters. Clin Investig 1994; 72(9): 692-4. http://dx.doi.org/10.1007/BF00212991 PMID: 7849450

[70] Moriguchi T, Urushiyama S, Hisamoto N, et al. WKIN regulates phosphorylation of cation-chloride-coupled cotransporters via the STE20-related kinases, SPAK and ORS1. J Biol Chem 2005; 280(52): 42858-93. http://dx.doi.org/10.1074/jbc.M510042200 PMID: 16263722

[71] Yang CL, Zhu X, Wang Z, Subramanya AR, Ellison DH. Mechanisms of WKIN1 and WKIN4 interaction in the regulation of thiazide-sensitive NaCl cotransport. J Clin Invest 2005; 115(5): 1379-87.
http://dx.doi.org/10.1172/JCI22452 PMID: 15841204

[85] Lazark A, Liu Z, Huang CL. Antagonistic regulation of ROMK by long and kidney-specific WNK1 isomorphs. Proc Natl Acad Sci USA 2006; 103(5): 1615-20.

[86] Hadchouel J, Ellison DH, Gamba R. Regulation of renal electrolyte transport by WNK and SPAK-OSR1 kinases. Annu Rev Physiol 2016; 78: 367-99.

[87] Segref A, Hoppe T. Think locally: control of ubiquitin-dependent protein degradation in neurons. EMBO Rep 2009; 10(1): 44-50.

[88] Yamauchi K, Rai T, Kobayashi K, et al. WNK4 phosphorylates ser(206) of claudin-7 and promotes paracellular Cl(−) permeability. FEBS Lett 2007; 581(20): 3887-91.

[89] Takahashi D, Mori T, Nemura N, et al. WNK4 is the major WNK positively regulating NCC in the mouse kidney. Biosci Rep 2014; 34(3): e01007.

[90] Terker AS, Zhang C, Ersaperme KJ, Gamba G, Yang CL, Ellison DH. Unique chloride-sensing properties of WNK4 permit the distal nephron to modulate potassium homeostasis. Kidney Int 2016; 89(1): 126-34.

[91] Funakawa M, He YJ, Borchers C, Xiong Y. Targeting of protein ubiquitination by BTB-Cullin 3-Roc1 ubiquitin ligases. Nat Cell Biol 2003; 5(11): 1001-7.

[92] Lu X, Wei Y, Rebol J, et al. BTB proteins are substrate-specific adapters in an SCF-like modular ubiquitin ligase containing CUL-3. Nature 2003; 425(6955): 316-21.

[93] Zimmerman ES, Schulman BA, Zheng N. Structural assembly of cullin-RING ubiquitin ligase complexes. Curr Opin Struct Biol 2010; 20(6): 714-21.

[94] Murthy M, Kurz T, O’Shaughnessy KM. WNK signalling pathways in blood pressure regulation. Cell Mol Life Sci 2017; 74(7): 1261-80.

[95] Wilson FH, Kahle KT, Wilson DH, Latorre A, et al. A novel WNK is regulated by the sodium-chloride cotransporter and potassium channel. Hypertension 2005; 46(2): 295-300.

[96] Gamba R. Role of WNK kinases in regulating tubular salt and potassium transport and in the development of hypertension. Am J Physiol Renal Physiol 2005; 288(2): F245-52.
Zhao H, Guan Q, Smith CJ, Quilley J. Increased phosphodiesterase 3A/4B expression after angioplasty and the effect on VASP phosphorylation. Eur J Pharmacol 2008; 590(1-3): 29-35. http://dx.doi.org/10.1016/j.ejphar.2008.05.016 PMID: 18571642

Maass PG, Aydin A, Luft FC, et al. PDE3A mutations cause autosomal dominant hypertension with brachydactyly. Nat Genet 2015; 47(6): 647-53. http://dx.doi.org/10.1038/ng.3302 PMID: 25961942

Schuster H, Toka O, Toka HR, et al. A cross-over medication trial for patients with autosomal-dominant hypertension with brachydactyly. Kidney Int 1998; 53(1): 167-72.

Toka O, Maass PG, Aydin A, et al. Childhood hypertension in autosomal-dominant hypertension with brachydactyly. Hypertension 2010; 56(5): 988-94. http://dx.doi.org/10.1161/HYPERTENSIONAHA.110.156620 PMID: 20837885

Toka O, Tank J, Schächterle C, et al. Clinical effects of phosphodiesterase 3A mutations in inherited hypertension with brachydactyly. Hypertension 2015; 66(4): 800-8. http://dx.doi.org/10.1161/HYPERTENSIONAHA.115.06000 PMID: 26283042