Leaf Wax Hydrogen Isotopes as a Hydroclimate Proxy in the Tropical Pacific

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Abstract

The hydrogen isotope composition ($^2$H/$^1$H ratios) of leaf waxes preserved in sediments is increasingly used to reconstruct past hydroclimate. Here, we extend the global calibration of leaf wax $^2$H/$^1$H ratios to include surface sediments from 23 lakes and swamps on 15 tropical Pacific islands. Leaf wax $^2$H/$^1$H ratios from this new data set are not correlated with regional estimates of mean annual precipitation $^2$H/$^1$H ratios derived from isoscapes or from isotope-enabled general circulation models. Nevertheless, the new data fall within the predicted range of values based on a global calibration compiled from published surface sediments. In our global compilation, we find a strong positive linear correlation between $^2$H/$^1$H ratios of mean annual precipitation and the common leaf waxes n-C29-alkane ($R^2 = 0.73, n = 581$) and n-C28-acid ($R^2 = 0.74, n = 242$). In the tropical Pacific, the largest residuals are no greater than those observed elsewhere, and are likely due to (1) uncertainty in the $^2$H/$^1$H ratios of local precipitation and (2) variability in net fractionation for different plant types. Palynological analyses from the same samples suggest that there is no systematic relationship between any particular type of pollen distribution and deviations from the global calibration line. Overall, our results support the use of leaf wax $^2$H/$^1$H ratios in tropical Pacific lake sediments as proxies for large hydrological changes, especially when paired with $^2$H/$^1$H ratios of source-specific biomarkers. However, the interpretation of such records needs to be informed by careful consideration of local drivers of precipitation isotope variability.

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Leaf Wax Hydrogen Isotopes as a Hydroclimate Proxy in the Tropical Pacific

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Key Points:

- Leaf wax $^2$H/$^1$H ratios are correlated with mean annual precipitation $^2$H/$^1$H ratios globally, but not in the tropical Pacific
- Deviations from the global relationship between precipitation leaf wax $^2$H/$^1$H ratios cannot be predicted from palynological assemblages
- Small range and large uncertainties in estimates of tropical Pacific precipitation $^2$H/$^1$H ratios likely account for poor correlations
Abstract

Hydrogen isotope ratios of sedimentary leaf waxes ($\delta^2$H$_{\text{Wax}}$ values) are increasingly used to reconstruct past hydroclimate. Here, we add $\delta^2$H$_{\text{Wax}}$ values from 19 lakes and four swamps on 15 tropical Pacific islands to an updated global compilation of published data from surface sediments and soils. Globally, there is a strong positive linear correlation between $\delta^2$H values of mean annual precipitation ($\delta^2$H$_{\text{P}}$ values) and the leaf waxes $n$-C$_{20}$-alkane ($R^2 = 0.74$, $n = 665$) and $n$-C$_{28}$-acid ($R^2 = 0.74$, $n = 242$). Tropical Pacific $\delta^2$H$_{\text{Wax}}$ values fall within the predicted range of values based on the global calibration, and the largest residuals from the global regression line are no greater than those observed elsewhere, despite large uncertainties in $\delta^2$H$_{\text{P}}$ values at some Pacific sites. However, tropical Pacific $\delta^2$H$_{\text{Wax}}$ values in isolation are not correlated with estimated $\delta^2$H$_{\text{P}}$ values from isoscapes or from isotope-enabled general circulation models. Palynological analyses from these same Pacific sediment samples suggest no systematic relationship between any particular type of pollen distribution and deviations from the global calibration line. Rather, the poor correlations observed in the tropical Pacific are likely a function of the small range of $\delta^2$H$_{\text{P}}$ values relative to the typical residuals around the global calibration line. Our results suggest that $\delta^2$H$_{\text{Wax}}$ values are currently most suitable for use in detecting large changes in precipitation in the tropical Pacific and elsewhere, but that ample room for improving this threshold exits in both improved understanding of $\delta^2$H variability in plants, as well as in precipitation.

Plain Language Summary

Past precipitation patterns are difficult to reconstruct, limiting our ability to understand Earth’s climate system. Geochemists reconstruct past precipitation by measuring the amount of
heavy hydrogen naturally incorporated into the waxy coating of leaves, which is preserved in mud that accumulates in lakes, soils, and oceans. Heavy hydrogen in leaf waxes is strongly correlated with local precipitation, allowing us to learn about rainfall intensity, temperature, and cloud movement. However, no existing calibration studies include sites from the tropical Pacific, home to the most intense rainfall on the planet and populations that rely on rain for drinking water. We measured heavy hydrogen in leaf waxes from tropical Pacific islands and show that although values are within the global calibration error, no precipitation relationship exists within the region. Plant type distributions do not explain the lack of correlation, which is best attributed to poorly constrained estimates of heavy hydrogen in local rain and the relatively small range of variability within the region. At present, heavy hydrogen from ancient leaf waxes can show large changes in past precipitation, but improved process-level understanding is needed to use this tool to understand smaller changes in the tropical Pacific and elsewhere.

1 Introduction

As Earth warms, precipitation intensity, frequency, and spatial distribution are expected to change over the tropical Pacific (Brown et al., 2011; Tan et al., 2015; Sharmila et al., 2018). These predictions need to be constrained and validated by robust reconstructions of past changes, which are unfortunately limited in this region, in part because of a lack of proxies and archives suitable for producing high resolution, continuous records (Hassall 2017). Existing high-resolution paleohydrologic records have been established from speleothems (Partin et al., 2013; Maupin et al., 2014) and corals (Quinn et al., 1993; Quinn et al., 1998; Hendy et al, 2002; Linsley et al., 2004; Linsley et al., 2006; Calvo et al., 2007; DeLong et al., 2012), but are generally limited to the past 600 years in this region. Lacustrine and swamp sediments can provide longer records with much higher temporal resolution than is possible from slowly
accumulating marine sediments, and are well-established archives of ecological, anthropogenic, and broad climatic changes in the region (Southern, 1986; Hope & Pask, 1998; Stevenson et al., 2001; Prebble & Wilmshurst, 2009; Prebble et al., 2019; Gosling et al., 2020). More recently, such sediments have also been used to reconstruct past hydroclimate change in the western tropical Pacific at higher temporal resolution (Sachs et al., 2009; Smittenburg et al., 2011; Konecky et al., 2016; Richey & Sachs, 2016; Hassall, 2017; Sachs et al., 2018; Sear et al., 2020).

One hydroclimate proxy suitable for tropical lake and swamp sediments in the tropical Pacific is based on the hydrogen isotopic composition of leaf waxes ($\delta^2H_{Wax} = (\overline{2H}/1H)_{Wax}/(\overline{2H}/1H)_{VSMOW} - 1$) (Sachse et al., 2012; Konecky et al., 2016; Hassall, 2017). $\delta^2H_{Wax}$ values are highly correlated with hydrogen isotopes of mean annual precipitation ($\delta^2H_P$) on a global scale and have been applied to reconstruct $\delta^2H_P$ values in diverse locations (Sachse et al., 2012; McFarlin et al., 2019). $\delta^2H_P$ values are related to specific physical processes, and are a chemical signal that can both be transferred to material preserved on geologic timescales, as well as modeled in modern systems with increasing accuracy, making their reconstructions useful for understanding past hydroclimate dynamics (Bowen et al., 2019).

As is typical for organic geochemical proxies, the relationship between $\delta^2H_{Wax}$ and $\delta^2H_P$ has been established through empirical calibrations with surface sediments from lakes and surface soils. These calibration efforts began in Europe (e.g., Sachse et al., 2004; Leider et al., 2013; Nelson et al., 2018) and have been extended to the Americas (e.g., Hou et al., 2008; Polissar & Freeman, 2010; Douglas et al., 2013), East Asia and the Tibetan Plateau (e.g., Jia et al., 2008; Aichner et al., 2010; Bai et al., 2011), and Africa (e.g., Peterse et al., 2009; Garcin et al., 2012; Schwab et al, 2015) (Figure 1). Recent compilations of $\delta^2H_{Wax}$ from surface sediments (McFarlin et al., 2019) and from sediments and soils (Liu & An, 2019) placed these local
calibrations in a global context. However, existing global compilations do not include any tropical Pacific $\delta^2H_{Wax}$ values.

Two considerations make it important to include tropical Pacific data in the global calibration. Firstly, different vegetation types can influence net community $^2H/^1H$ fractionation between precipitation and leaf waxes, which is not constant among plant types or environments (Feakins and Sessions, 2010; Sachse et al., 2012; Kahmen et al., 2013). The unique plant communities on tropical Pacific islands include many endemic species (Gillespie et al., 2013). Additionally, coastal regions or former lagoons on these islands are often covered by mangrove swamps, which consist of trees and shrubs adapted to brackish to hypersaline water. Due to salinity effects, mangrove $\delta^2H_{Wax}$ values may have the opposite response to changes in precipitation intensity as nearby freshwater plants (Ladd & Sachs, 2012; He et al., 2017). However, the impact of mangrove contributions to sedimentary $\delta^2H_{Wax}$ values has not been assessed.

Secondly, there is large uncertainty associated with estimates of $\delta^2H_P$ in the tropical Pacific. Direct measurements of $\delta^2H_P$ from the Global Network of Isotopes in Precipitation (GNIP) are spatially and temporally limited compared to other regions, resulting in large uncertainties for statistical interpolations of $\delta^2H_P$ such as those used for isoscape products and the Online Isotopes in Precipitation Calculator (OIPC; Bowen & Revenaugh, 2003). Estimates of $\delta^2H_P$ from general circulation models (GCMs) in which precipitation isotopes have been incorporated offer another potential calibration target for $\delta^2H_{Wax}$ measurements in the modern tropical Pacific that has not yet been explored.

Here we measured $\delta^2H$ values of seven $n$-alkane and five $n$-alkanoic acid homologues from surface sediments collected from lakes influenced by precipitation from the South Pacific Convergence Zone (SPCZ) and from mangrove swamps influenced by the Intertropical
Convergence Zone (ITCZ). We add new surface sediment $\delta^2$H$_{Wax}$ measurements of two of these compounds ($n$-C$_{29}$-alkane and $n$-C$_{28}$-acid) from 19 lakes and four mangrove swamps on 15 islands distributed throughout the tropical Pacific to an updated global compilation of $\delta^2$H$_{Wax}$ values. We assess whether $\delta^2$H$_{Wax}$ values from tropical Pacific lake and swamp sediments are consistent with the global relationship between $\delta^2$H$_{Wax}$ and modeled $\delta^2$H$_P$ values from a diverse set of algorithms and models. Finally, we use pollen-based vegetation reconstructions to evaluate the influence of plant communities on tropical Pacific $\delta^2$H$_{Wax}$ values.

2 Materials and Methods

2.1 Site description and sample collection

Surface sediments were collected from 19 lakes on 11 islands across the SPCZ region (Figure 1, Table 1), ranging in elevation above mean sea level from 790 m (Lanoto’o, Samoa) to 1 m (Rimatu’u, Oroatera, and Onetahi ponds on Tetiaroa, French Polynesia). Lakes ranged from shallow ephemeral water bodies to an 88 m-deep volcanic crater lake (Lake Lalolalo, Wallis). Most lakes were freshwater systems, except for the brackish (salinity = 17) coastal Lake Dranoniveilomo (Fiji) and Lake Lalolalo (Wallis), which has a freshwater surface lens above saline water (Sichrowsky et al., 2014). Mangrove trees surrounded Lake Dranoniveilomo, while many other sites were located in forested regions, some impacted by human activity, particularly horticulture. Aquatic vegetation covered the surface of some lakes (Table 1). Additional samples were obtained from mangrove swamps located within the ITCZ throughout the Federated States of Micronesia and Guam. All swamps were located at sea level and submerged at high tide. Four or five surface sediment samples were collected from each swamp along a transect from the inland edge to the coast.
Maloney et al. (2019) described the collection of most lake samples from the SPCZ region. New samples include those from Lake Dranoniveilomo, which was cored in 2010 with a Universal Percussion Corer (Aquatic Research, Hope ID, USA) fitted with a 6.6 cm diameter polycarbonate core tube. Vesalea and Nopovois were cored in 2017 with a percussion corer (UWITEC, Mondsee, Austria) equipped with a 6.3 cm diameter polycarbonate tube. Unconsolidated upper sediment from these cores was subsampled at 1 cm intervals in the field and stored frozen in Whirl-Pak plastic bags (Nasco, Fort Atkinson, WI, USA). Analyses of lake surface sediment were restricted to the uppermost 1 or 2 cm of material. A hand trowel was used to collect the upper 1 cm of mangrove sediments in 2012. Samples were stored frozen in Whirl-Pak bags.

Water was collected from the surface of each lake and stored in screw-cap glass vials. Additional water samples were collected from adjacent streams and long-term precipitation integrators such as wells and rain cisterns when available. Samples were stored at 4 °C prior to analysis.

2.2 Leaf wax extraction and purification

Maloney et al. (2019) described lipid extraction, saponification, and column chromatography for all lake surface sediments except Dranoniveilomo, Vesalea, and Nopovois. Samples from Dranoniveilomo were processed following the protocol of Maloney et al. (2019). For all these lake sediments, the acid-containing fraction was eluted with 6 ml 4% acetic acid in diethyl ether from an aminopropyl gel column and the alkane-containing fraction was eluted with 6 ml hexane from a silica gel column. Lipid extraction, saponification, and column chromatography from Vesalea was described by Krentscher et al. (2019), and was identical for
the sample from Nopovois. Lipids from mangrove surface sediments were extracted and divided into compound classes using Si gel column chromatography as in Ladd and Sachs (2017).

For mangrove surface sediments, $n$-alkanes were purified from a Si gel hexane fraction by eluting 8 mL of 100% hexane over 0.5 g of AgNO$_3$-impregnated Si gel (10% by weight). For lake samples, the alkane fraction was urea adducted to isolate unbranched compounds. Fatty acids from lake sediments were methylated with 5% HCl in methanol for 12 hours at 70 °C, and saturated fatty acid methyl esters (FAMEs) were isolated by elution in 8 mL of 4:1 Hex/DCM over 0.5 g of AgNO$_3$-impregnated Si gel (10% by weight). Acid fractions were not analyzed from mangrove surface sediments. Purity and concentrations of $n$-alkanes from mangrove samples were assessed by gas chromatography – flame ionization detection (GC-FID) using the GC program and instrumentation described in Ladd and Sachs (2017). For lake samples, $n$-alkane and $n$-acid homologues were quantified using the same GC program and instrumentation described in Ladd et al. (2018).

2.3 $\delta^{2}$H$_{Wax}$ measurements

Samples were dissolved in hexane at a concentration suitable for hydrogen isotope analyses of $n$-C$_{29}$-alkane or $n$-C$_{28}$-acid when those compounds were sufficiently abundant for analysis by gas chromatography – isotope ratio mass spectrometry (GC-IRMS). $\delta^{2}$H values of other baseline-resolved homologues with peak areas $>$15 Vs are also reported. For mangrove sediment samples, GC-IRMS analyses were conducted with the same GC program and isotopic referencing described in Ladd and Sachs (2017). Lake sediment samples were analyzed with the same GC program and isotopic referencing described in Ladd et al. (2018). Phthalic acid of known isotopic composition (Shimmelmann, Indiana University) was methylated to determine
δ²H values of H added during methylation, which was corrected for using isotopic mass balance (Lee et al., 2017).

2.4 Estimates of δ²H_P values

Estimates of δ²H_P values were extracted from different model products using latitude, longitude, and elevation of each site. Model products included the Online Isotopes in Precipitation Calculator (OIPC) version 3.2 (Bowen & Ravenaugh, 2003; IAEA/WMO, 2015; Bowen, 2020), as well as isotope-enabled climate model contributions to the second Stable Water Isotope Intercomparison Group (SWING2) from the CAM, ECHAM, GISS ModelE, HadAM, isoGSM, LMDZ, and MIROC models (Sturm et al., 2010). OIPC estimated values were obtained manually from the web interface (“OIPC mean annual δ²H”), and also by extraction from the high-resolution spatial gridded data set using a bilinear smooth function to accommodate the proximity of a given location to neighboring pixels and the δ²H values from those pixels (“OIPC extracted mean annual δ²H”). The multi-model mean annual precipitation δ²H value was calculated by averaging predicted values for all climate models that employed spectral nudging (Yoshimura et al., 2008), which includes the ECHAM, GISS (nudged), isoGSM, and LMDZ products.

2.5 Water δ²H and δ¹⁸O analyses

The isotopic composition of most lake and stream water samples were previously analyzed and reported by Maloney et al. (2019). Water samples from the 2012 Micronesian field campaign and from Lake Dranoniveilomo were analyzed by Cavity Ring Down Spectroscopy (CRDS; Li-2130i, Picarro, Santa Clara, CA) using the same conditions and standards as in Maloney et al. (2019). Additional water samples from the 2017 Vanuatu field campaign were
analyzed by Thermal Conversion/Elemental Analysis – Isotope Ratio Mass Spectrometry (TC/EA-IRMS; ThermoFisher Scientific, Bremen, Germany) using the same conditions and standards as in Newberry et al. (2017).

2.6 Pollen counts

Core samples for palynomorph analyses (including pollen and spores) were taken from within the upper portion of the sediment core to determine modern baseline vegetation differences among lakes. Each 1 cm³ sample was processed using standard procedures (10% HCl, hot 10% KOH, and acetolysis) (Moore et al. 1991). Samples were spiked with exotic Lycopodium clavatum L. tablets to allow the palynomorph and charcoal concentrations to be calculated. Counts continued until reaching at least 100 terrestrial palynomorphs. Reference palynomorphs held in the Australasian Pollen and Spore Atlas (apsa.anu.edu.au/) assisted with identification. The vegetation types (primary, secondary, dryland herbs, wetland herbs, etc.) were determined from a regional synthesis of Pacific Island plant ecology (Mueller-Dombois & Fosberg 1998).

3 Results

3.1 δ²H_Wax values in the tropical Pacific

δ²H_Wax values from surface sediments in the tropical Pacific were not correlated with mean annual δ²H_P values as calculated by the OIPC, nor with mean annual precipitation amount as estimated by the Global Precipitation Climatology Project (GPCP) (Adler et al., 2003) (Figure 2). The only lipids with significant correlations with δ²H_P values were dinosterol (data from Maloney et al., 2019), n-C_{16}-acid, and n-C_{18}-acid, and the only significant correlations with the amount of mean annual precipitation were dinosterol, n-C_{18}-acid, n-C_{17}-alkane, and n-C_{33}-alkane.
In almost all cases, correlation coefficients were negative for the relationship between $\delta^2H_{Wax}$ and $\delta^2H_P$ values, and positive for the relationship between $\delta^2H_{Wax}$ values and mean annual precipitation. An exception was $n$-C$_{17}$-alkane, which, similarly to dinosterol, had $\delta^2H$ values that are negatively correlated with mean annual precipitation ($R = -0.95; p = 0.049$) and positively correlated with $\delta^2H_P$ values ($R = 0.95; p = 0.051$) (Figure 2). However, $n$-C$_{17}$-alkane was only abundant enough to measure its $\delta^2H$ values in four samples, making any assessment of these correlations tentative.

3.2 Tropical Pacific $\delta^2H$ values in the global context

Tropical Pacific $\delta^2H$ values of $n$-C$_{29}$-alkanes and $n$-C$_{28}$-acids (the most commonly measured leaf waxes in the literature) were in the range expected based on the global relationship between $\delta^2H_{Wax}$ and $\delta^2H_P$ values (Figure 3). Tropical Pacific $n$-C$_{29}$-alkane $\delta^2H$ values ranged from $-177$ to $-139\%o$, while those of $n$-C$_{28}$-acid ranged from $-175$ to $-119\%o$ (Table 1; Figure 3). Adding these new measurements to an updated global compilation of $\delta^2H_{Wax}$ values from all available surface sediment and soil data sets in non-marine settings (compilations from Liu & An, 2019 and McFarlin et al., 2019, as well as data sets from Nelson, 2013; Bakkelund et al., 2018; Feng et al., 2019; Goldsmith et al., 2019; Li et al., 2019; Wu et al., 2019; Lu et al., 2020; Struck et al., 2020; van der Veen et al., 2020) has minimal impact on the slope, $y$-intercept, or correlation coefficients for the global linear regression (Figure 3).

To compare new measurements from the tropical Pacific to the relationship defined by previously published values, we calculated their residual values from the global linear regression between $\delta^2H_{Wax}$ and $\delta^2H_P$ (excluding new tropical Pacific data). At most tropical Pacific locations, residuals from the global linear regression line were within $\pm 20\%o$, but were greater than this at 5 sites for $n$-C$_{29}$-alkane and 6 sites for $n$-C$_{28}$-acid (Table 1). Only two sites (Lake
Tagamaucia in Fiji and White Lake in Vanuatu) had residuals greater than 20‰ for both compounds.

We also contextualized variability in the tropical Pacific data relative to the global data set by randomly subsampling 17 values from the compiled data 4,000 times and comparing the correlation coefficient between $\delta^2$H$_\text{Wax}$ and $\delta^2$H$_P$ to the range in $\delta^2$H$_P$ values (Figure 4). None of these subsampled data sets had ranges in $\delta^2$H$_P$ values that were as small as the range in the tropical Pacific (smallest range for $n$-C$_{29}$-alkane = 59‰, for $n$-C$_{28}$-acid = 118‰), so we subsampled the compiled data again while restricting the maximum range to 100‰ (1,000 iterations each for the highest, lowest, and middle $\delta^2$H$_P$ values), 50‰ (200 iterations for each possible 50‰ range with maximum $\delta^2$H$_P$ values shifted by 10‰), and 35‰ (100 iterations for each possible 35‰ range with maximum $\delta^2$H$_P$ values shifted by 5‰). Correlation coefficients were typically high (>0.5) when the range of $\delta^2$H$_P$ values was greater than 100‰, and became increasingly scattered below this threshold (Figure 4). The relationship between the correlation coefficient and the range of $\delta^2$H$_P$ values in the tropical Pacific plotted within the range generated by random subsets. However, tropical Pacific correlation coefficients for both compounds were more than one standard deviation below the mean value of random sample sets with a $\delta^2$H$_P$ range between 30 and 35‰ (0.43 ± 0.28 for $n$-C$_{29}$-alkane; 0.22 ± 0.41 for $n$-C$_{28}$-acid) (Figure 4).

In addition to the OIPC, several water-isotope-enabled GCMs also provide estimates of mean annual $\delta^2$H$_P$ values. We extracted $\delta^2$H$_P$ values for all sites with surface sediment or soil $\delta^2$H$_\text{Wax}$ values from each climate model included in the Stable Water Isotope Intercomparison Group, Phase 2 (SWING 2) model comparison. For all models, global $n$-C$_{29}$-alkane and $n$-C$_{28}$-acid $\delta^2$H values were positively correlated with $\delta^2$H$_P$ estimates (Figure 5). For $n$-C$_{29}$-alkane, $\delta^2$H$_\text{Wax}$ values were most highly correlated with $\delta^2$H$_P$ values obtained manually from the OIPC (R = 0.86). The lowest correlation was with $\delta^2$H$_P$ values from HadAM (R = 0.57) (Figure 5). For
n-C_{28}-acid, global $\delta^2H_{\text{Wax}}$ values were most highly correlated with $\delta^2H_P$ values extracted from the CAM model ($R = 0.91$). The lowest correlation was with $\delta^2H_P$ values extracted from the high-resolution spatial gridded OIPC data ($R = 0.82$) (Figure 5). The correlation coefficient obtained manually from the OIPC ($R = 0.86$) was intermediate among the different models (Figure 5).

3.3 Pollen and spore spectra

Palynomorphs from most sites were indicative of human disturbance to the catchment vegetation, as the dominant pollen types are from secondary forest taxa (Figure 6; Table 2). When secondary forest vegetation was not most abundant, fern spores contributed more to the palynomorph sum than any other plant group, except at Lake Hut, where primary forest taxa were most abundant (Figure 6; Table 2). Although wetland plants covered more than 50% of the surface water at three lakes (Onetahi Pond, Lake Tagamucia, and Veselea Pond), wetland herbs and aquatic plants never contributed more than 23% of the observed pollen.

Pollen data are only available for three of the five sites where $n$-C_{29}-alkane residuals from the global $\delta^2H_{\text{Wax}}$ vs. $\delta^2H_P$ relationship were less than $-20\%$ (Figure 6). In two of these (Tagamaucia and White Lake), fern spores were abundant (59% and 39%, respectively) (Figure 6; Table 2). However, at the third site with an $n$-C_{29}-alkane residual less than $-20\%$ (Lake Hut), fern spore concentrations were low and primary forest taxa palynomorphs were most abundant (Figure 6; Table 2). Three sites had $n$-C_{28}-acid residuals from the global $\delta^2H_{\text{Wax}}$ vs. $\delta^2H_P$ relationship less than $-20\%$, of which two have pollen data (Tagamaucia and Dranoniveilomo). Each of these had a high abundance of ferns and wetland plants (80% and 37%, respectively) (Figure 6, Table 2). Three sites had $n$-C_{28}-acid residuals from the global $\delta^2H_{\text{Wax}}$ vs. $\delta^2H_P$ relations that were greater than $20\%$ (Figure 6). One of these, White Lake, had relatively high contributions from ferns. The second, Harai Lake #1, does not have recent pollen data (the most
recent pollen sample is from 11–12 cm), but historically had high contributions from ferns (Table 2). The third, Nopovois, is dominated by secondary forest vegetation (54%) and does not have palynological features that clearly distinguish it from sites where $n$-$C_{28}$-acid $\delta^2$H values adhere more closely to the global relationship (Figure 6; Table 2). Additionally, some sites with high contributions from ferns and wetland plants – Harai Lake #3, Lanoto‘o, and Lake Otas – have $\delta^2$H$_{\text{Wax}}$ values close to the global relationship (Figure 6).

4 Discussion

Although the relationship between $\delta^2$H$_{\text{Wax}}$ and $\delta^2$H$_P$ values lacks any correlation for the tropical Pacific sites in isolation, the values fall within the global scatter around the linear regression of compiled literature values from surface sediments and soils (Figure 3; Table 1). In addition to adding recently published data, our global data set differs from two recent compilations by excluding marine sediments (in contrast to Liu and An, 2019), and including both soils and surface sediments (in contrast to McFarlin et al., 2019). Although leaf waxes in soils and sediments might have different sources and represent different timescales and catchment areas, there is no significant difference in the relationship between $\delta^2$H$_{\text{Wax}}$ and $\delta^2$H$_P$ for either compound (Figure 3c and 3d). The similarity between the soil and sediment compilations suggests that the transit history of leaf waxes from plant to deposition and subsequent preservation varies as much within archive type as it does between them.

The positive linear relationship between $\delta^2$H$_{\text{Wax}}$ and $\delta^2$H$_P$ values in the global compilation remains robust, with R$^2$ values of 0.74 for both $n$-$C_{29}$-alkane (n = 665) and $n$-$C_{28}$-acid (n = 242) (Figure 3). However, considerable scatter around the regression line exists globally and within the tropical Pacific. Large residuals are due to both uncertainty in the y-axis (variable $^2$H/$^1$H fractionation between leaf waxes and water among plant types and environments, discussed in
section 4.1) and in the x-axis (mean annual δ²H_P values and the water source used by plants, discussed in section 4.2).

4.1 Variable hydrogen isotope fractionation during leaf wax synthesis

Although global δ²H_Wax values are well correlated with δ²H_P values of mean annual precipitation (Figures 3, 5), several well-established factors contribute to variability in the net ²H/¹H fractionation between plant waxes and precipitation (α_Wax-P) (Sachse et al., 2012). Variations in α_Wax-P occur among plant functional types (Liu et al., 2006), between leaves and other plant organs (Gamarra & Kahmen, 2015), and with relative humidity (Tipple et al., 2015). Additionally, biosynthetic fractionation between leaf water and waxes can vary seasonally (Newberry et al., 2015), with environmental stresses (Ladd & Sachs, 2015), and with changes in plant metabolism (Cormier et al., 2018). Large differences in α_Wax-P can also exist among plant species growing at the same site (Feakins & Sessions, 2010; Sachse et al., 2012; Eley et al., 2014; He et al., 2020). With the current tropical Pacific data set we can only examine factors that might relate to differences among different types of plants, and not factors that can occur within a single plant, such as metabolic state. We examine three plant groups – mangroves, aquatic plants, and ferns – whose contributions may impact community δ²H_Wax values. By comparing pollen distributions and information about surrounding vegetation at each site with the residuals between δ²H_Wax values and the global calibration line (Figure 6), we demonstrate that changes in vegetation inferred from pollen cannot consistently explain anomalous δ²H_Wax values in tropical Pacific surface sediments.

4.1.1 Mangroves
Mangroves are woody plants that grow in brackish to hypersaline water. They contribute large amounts of organic matter to coastal sediments in the tropics and subtropics (Alongi, 2014). Because mangroves discriminate more against $^2$H as salinity increases (Ladd & Sachs, 2012; He et al., 2017; Ladd & Sachs, 2017), they should have lower $\delta^2$H wax values than nearby freshwater plants. This relationship was recently observed in the Florida Everglades, where mangroves have $\delta^2$H wax values ~50‰ lighter than those from nearby freshwater trees, despite equivalent $\delta^2$H P values (He et al., 2020). Significant contributions of mangrove leaf waxes in coastal areas in the tropical Pacific could result in sedimentary $\delta^2$H wax values that fall below the global calibration line.

Several of the lakes in our calibration set were located in coastal areas, but only one, Dranoniveilomo, had brackish water and mangroves growing directly in its periphery. In this lake $n$-C$_{28}$-acid is significantly depleted relative to the global calibration, but $n$-C$_{29}$-alkane is not (Table 1). Despite the abundant mangroves around Dranoniveilomo, barely any mangrove pollen was found in the sediment, which may reflect different transport mechanisms and catchment areas for leaf waxes and pollen (Table 2). Two coastal lakes in Vanuatu (Otats and Waérowa East) have the highest amounts of mangrove pollen observed in all examined surface sediments (~15%; Figure 6; Table 2). In Lake Otats, $\delta^2$H wax values are close to the values predicted by the global relationship (Table 1). In Lake Waérowa East, they are slightly higher than expected (Table 1), opposite to the expected impact of significant mangrove leaf wax contributions. These data suggest that mangrove leaf waxes are not an important influence on $\delta^2$H wax values in tropical Pacific lake sediments included in this study.

Likewise, Micronesian mangrove swamp surface sediments had $\delta^2$H wax values that were consistent with the global linear regression (Table 1; Figure 3). Additionally, there was little spatial variability in $\delta^2$H wax values throughout each individual mangrove swamp, with 5‰
standard deviations among samples within a single swamp (Table 1). This homogeneity occurred even though samples were collected from sites with surface water salinity ranging from 0-31 at the time of collection. Surface water salinity was dynamic throughout these mangrove swamp surveys, varying temporally and spatially with tides and rain events. Individual mangrove trees with large root networks therefore had access to water with a wide range of salinities and may have opportunistically taken up relatively fresh water that was ultimately used in leaf wax synthesis. Preferential uptake of fresher water by mangroves has been observed previously (Santini et al., 2015; Reef et al., 2015), and could result in all mangroves throughout a swamp using water with similar salinity and isotopic composition, consistent with the surface sediment $\delta^2H_{Wax}$ values observed in transects from Micronesian mangrove swamps.

4.1.2 Aquatic plants

Some of the lakes included in the tropical Pacific survey were partially or completely covered by floating aquatic vegetation (Table 1). Since aquatic plants at diverse sites tend to have lower alkane $\delta^2H_{Wax}$ values than nearby terrestrial plants (Chikaraishi & Naraoka, 2003; Gao et al., 2011; Dion-Kirschner et al., 2020; He et al., 2020), differing relative contributions of leaf waxes from aquatic plants could also reduce sedimentary $\delta^2H_{Wax}$ values. There are a few reasons why aquatic plants may have relatively low $\delta^2H_{Wax}$ values. First, when a lake is mostly covered by water lilies or similar aquatic vegetation, there is a physical barrier to evaporation of lake water, and it may therefore maintain a $\delta^2H$ signal similar to that of precipitation, rather than becoming enriched in $^2H$ due to transpiration, as is the case for leaves exposed to air (Kahmen et al., 2013; Cernusak et al., 2016). Second, it is possible that aquatic plants may exhibit greater biosynthetic fractionation between leaf water and leaf lipid. However, existing investigations of
\( \delta^2 \text{H}_{\text{Wax}} \) values in submerged aquatic plants suggest this is only likely at high salinity, while plants grown in freshwater display similar \( \alpha_{\text{Wax-P}} \) values to other plants (Aichner et al., 2017).

High contributions from aquatic plants could explain why \( \delta^2 \text{H}_{\text{Wax}} \) values at Tagamaucia in Fiji, which is covered in floating sedge islands (Southern et al., 1986), were very \(^2\text{H}\)-depleted relative to the global calibration line (Table 1). However, this relationship was not consistent in all lakes covered by aquatic plants. For example, Lake Veselea in Vanuatu is completely covered by mats of aquatic plants (primarily Persicaria cf. attenuata, Salvinia molesta, and Calystegia soldanella), yet \( \delta^2 \text{H}_{\text{Wax}} \) values from its sediment fell close to the global calibration line (Table 1). Additionally, pollen from wetland herbs and aquatic plants is not consistently associated with large or small residuals from the global calibration line (Figure 6). Aquatic plants may have minimal influence on sedimentary \( \delta^2 \text{H}_{\text{Wax}} \) values because submerged plants are not at risk of desiccation and therefore have little need for the moisture barrier provided by long-chain leaf waxes. They therefore tend to have low concentrations of these compounds (Ficken et al., 2000; Dion-Kirschner et al., 2020; He et al., 2020). Overall, our results suggest that increased presence of aquatic plants does not unequivocally result in decreased \( \delta^2 \text{H}_{\text{Wax}} \) values in tropical Pacific lake sediments.

4.1.3 Ferns

Assessments of \( \delta^2 \text{H}_{\text{Wax}} \) values from ferns are limited, but previous studies show that ferns have similar \( \alpha_{\text{Wax-P}} \) values to many other plant taxa, including lycopsids, gymnosperms, eudicots, and magnoliids (Gao et al., 2014). We therefore had no expectation that sites with large sedimentary contributions of leaf waxes from ferns would diverge from the global \( \delta^3 \text{H}_{\text{Wax}} - \delta^2 \text{H}_\text{P} \) linear regression. However, some of the sites with the largest residuals relative to the global regression line had palynomorph spectra characterized by large contributions of fern spores. This
was particularly true at Lake Tagamaucia in Fiji and White Lake in Vanuatu, and to a lesser extent in Fiji’s Lake Dranoniveilomo (Table 1; Figure 6). However, high accumulation of fern spores did not universally correspond to low $\delta^2H_{\text{Wax}}$ values, for example at Lake Lanoto’o in Samoa and Harai Lakes #1 and #3 in the Solomon Islands (Tables 1 and 3; Figure 6). Of these, $n$-C$_{28}$-acid $\delta^2$H values from the Harai Lakes were much higher than the predicted values from the global regression fit, in direct contrast to the Fijian lakes and White Lake (Table 1; Figure 6).

Additionally, Lake Hut in New Caledonia had the largest $n$-C$_{29}$-alkane residual for any site with pollen data, but did not have much fern pollen (Figure 6). Overall, this suggests that relative contributions of leaf waxes from ferns do not have predictable effects on sedimentary $\delta^2H_{\text{Wax}}$ values. Nevertheless, a shift in $\delta^2H_{\text{Wax}}$ values that coincides with a change in the relative abundance of fern spores in a down-core record may indicate a change in organic matter sources rather than a change in $\delta^2H_P$ values.

4.2 Uncertainty in $\delta^2H_P$ values

Variability in $\alpha_{\text{Wax-P}}$ contributes to uncertainty in the y-axis in the relationship between $\delta^2H_{\text{Wax}}$ and $\delta^2H_P$ (Figure 3). However, unlike most proxy systems, the x-axis calibration target is poorly constrained and likely accounts for a large portion of the linear regression residuals between $\delta^2H_{\text{Wax}}$ and $\delta^2H_P$ values in the tropical Pacific. $\delta^2H_P$ values are not constant throughout the year, and water from different seasons has different residence times in soil, meaning that the $\delta^2$H values of water used by plants is typically not equal to mean annual $\delta^2H_P$ values (Brinkmann et al., 2018). Additionally, $\delta^2H_P$ values from the OIPC represent climatological means, but $\delta^2H_P$ values vary interannually and seasonally. The time period captured by a surface sediment sample (typically a few years to two decades; see Maloney et al., 2019 for sediment accumulation rates at most sites) may differ considerably from the long-term mean. Finally, the robustness of mean
annual estimates provided by the OIPC varies spatially among geographic settings due to the uneven density of $\delta^2$H$_P$ observations (IAEA/WMO, 2015).

Limited $\delta^2$H$_P$ data from some sites in the tropical Pacific mean that $\delta^2$H$_P$ values calculated using OIPC have large uncertainties (Table 1). This is especially problematic for sites in the southeastern portion of the region, since there are no observations of $\delta^2$H$_P$ values from the Solomon Islands, Vanuatu, or New Caledonia in the GNIP database. 95% confidence intervals for $\delta^2$H$_P$ values from sites in Vanuatu are as large as 76‰, considerably larger than the overall range in $\delta^2$H$_P$ values (~30‰) and the range in measured $\delta^2$H$_{Wax}$ values (~40‰ for n-C$_{29}$-alkane and ~55‰ for n-C$_{28}$-acid) in the Pacific (Table 1; Figure 3). For other sites, such as those in Micronesia, OIPC uncertainties may add false confidence to predicted values, as these can be based on a limited number of GNIP observations from several decades ago. For example, the Yap GNIP station has 65 observations collected from 1968 to 1976, and the station on Chuuk (Truk) has 72 observations collected between 1968 and 1977 (IAEA/WMO, 2015).

Climatological means calculated from these data may not be representative of conditions when surface sediment leaf waxes in this study formed. This uncertainty in the independent variable likely contributes to the lack of a regional correlation between $\delta^2$H$_{Wax}$ and $\delta^2$H$_P$ values (Figure 3).

An alternative approach for estimating mean annual $\delta^2$H$_P$ values is to use water-isotope-enabled GCMs (Sturm et al., 2010; Conroy et al., 2013; Steen-Larsen et al., 2017). These estimates of $\delta^2$H$_P$ values have not typically been used in $\delta^2$H$_{Wax}$ calibration studies. Because some of the western Pacific $\delta^2$H$_P$ values derived from OIPC had such large uncertainties, we assessed whether scatter in the global relationship between $\delta^2$H$_{Wax}$ and $\delta^2$H$_P$ could be reduced by using isotope-enabled GCMs to estimate $\delta^2$H$_P$ values (Figure 5).
In general, $\delta^{2}H_{\text{Wax}}$ values from $n$-C$_{28}$-acid are better correlated with $\delta^{2}H_{P}$ values generated by isotope-enabled GCMs than $\delta^{2}H_{\text{Wax}}$ values from $n$-C$_{29}$-alkane. This difference may be due to the distinct spatial distributions of the two calibration data sets, with fatty acid $\delta^{2}H$ values almost exclusively limited to North America (Figure 1). Sites from which $n$-alkane $\delta^{2}H$ values are available are more numerous, more globally distributed, and include many measurements from the Himalayas and Tibetan Plateau (Figure 1). Here, steep elevation gradients may make $\delta^{2}H_{P}$ vary on spatial scales that are smaller than the resolution of most models, and fluvial and aeolian processes may transport waxes between regions with distinct $\delta^{2}H_{P}$ values. Overall, our analysis does not suggest any structural limitations to using the OIPC to estimate $\delta^{2}H_{P}$ values for proxy calibration. However, the limited $\delta^{2}H_{P}$ measurements from the tropical Pacific and resulting large uncertainties in modern estimates from the OIPC or isotope-enabled GCMs remains a considerable challenge for assessing the fidelity of $\delta^{2}H_{\text{Wax}}$ values in this region.

In particular, the orographic effects of mountainous islands may not be adequately captured by either the OIPC or isotope-enabled GCMs, causing particular challenges determining the appropriate calibration target for water isotope proxies. For example, the highest peaks on the island of Espiritu Santo in Vanuatu are nearly 2000 m above sea level. With prevailing winds from the east, the west coast sits in the rain shadow of the mountains and is significantly drier than the eastern part of the island (Terry, 2011). However, the only long-term weather station for the island is located at Pekoa airport in the southeast corner of the island, and there are no local GNIP stations. Therefore, minimal local data is available to inform precipitation isotope models. The OIPC predicts equivalent $\delta^{2}H_{P}$ values for sites on opposite coasts of Espiritu Santo, and predicted $\delta^{2}H_{P}$ values from water-isotope-enabled GCMs on each coast are within a few ‰ of each other. However, the leeward western sites should have lighter $\delta^{2}H_{P}$ values than the eastern sites that receive rain from more marine air masses (Scholl et al., 1996, 2007). This expectation
is supported by the δ²H values of lake and stream waters collected during our field campaigns, which are depleted by 20 – 40 ‰ on the leeward western side of the island compared to the windward east coast (Data Set S2).

4.3 Contrasts with δ²H values of non-leaf wax lipids

In addition to the longer chain n-alkane and n-alkanoic acids that are primarily derived from higher plant waxes, we also measured δ²H values from several compounds of mixed or primarily algal sources. These compounds are typically found in sedimentary records along with leaf waxes, and the different controls on their δ²H values offer the opportunity to more completely resolve sources of down-core variability in δ²H_Wax values. Here we discuss lipids from algal sources, and ubiquitous compounds produced by most organisms.

4.3.1 Algal lipids

Unlike δ²H_Wax values, δ²H values of algal biomarkers are well correlated with tropical Pacific δ²H_P values and mean annual precipitation (Figure 2). This is particularly the case for dinosterol (Maloney et al., 2019), which is primarily produced by dinoflagellates (Volkman, 2003). The δ²H values of n-C₁₇-alkane, which is primarily derived from algae (Cranwell et al., 1987; Meyers, 2003), were also highly correlated with δ²H_P values (consistent with Sachse et al., 2004) and inversely correlated with mean annual precipitation (Figure 2). An inverse correlation between the amount of mean annual precipitation and δ²H_P values (and therefore δ²H values of lipids that track δ²H_P) is expected in a low-latitude maritime regions where the amount effect plays a strong role in determining the isotopic composition of rain (Dansgaard, 1964; Rozanski et al., 1993; Kurita et al., 2009).
One reason why $\delta^{2}H$ values from the algal biomarkers are better correlated with $\delta^{2}H_{P}$ values than $\delta^{2}H_{Wax}$ values might relate to the source water used by each type of organism. Leaf waxes from higher plants growing on land may reflect a temporal bias, as monthly OIPC $\delta^{2}H_{P}$ values can differ by up to 40‰ at the tropical Pacific sites. Higher plants primarily produce leaf waxes soon after setting new leaves, meaning that there may be a seasonal bias in the $\delta^{2}H_{P}$ signal that is transferred to their waxes (Tipple et al., 2013; Freimuth et al., 2017). If algae are productive throughout the year, they may better integrate annual precipitation, therefore resulting in algal lipids that more closely track mean annual $\delta^{2}H_{P}$ values.

Another reason why algal biomarkers may track $\delta^{2}H_{P}$ better than leaf waxes is that they come from a more limited range of potential sources. In addition to the range of plant sources for leaf waxes discussed in section 4.1, many of the mid-chain and relatively long-chained acetogenic compounds, including $n$-$C_{29}$-alkane and $n$-$C_{28}$-acid, can be derived from a mix of terrestrial and aquatic plants (Bush & McInerney, 2013; Andrae et al., 2020; Dion-Kirschner et al., 2020). $n$-$C_{28}$-acid can also be partially derived from microalgal sources (Volkman, 1980; van Bree et al., 2018). Therefore the leaf waxes may represent variable aquatic and terrestrial contributions, while the algal compounds are always aquatically sourced.

Finally, the spatial variability integrated by each type of compound could explain the different trends in their $\delta^{2}H_{Wax}$ values. Algal lipids are produced within the relatively confined space of the lake or pond overlaying the sediments in which they accumulate. Leaf waxes can be derived from plants growing adjacent to their depocenter, but also from further afield in the catchment, and the relative size of the catchment area can differ among water bodies. Additionally, > 20% of leaf waxes accumulating in sediment can come from aerosols, which can be transported long distances and have $\delta^{2}H$ values distinct from local vegetation (Conte et al., 2003; Gao et al., 2014; Nelson et al., 2017; Nelson et al., 2018). Leaf wax aerosols from very
distant sites may have more impact on lake sediments on islands than on continents, since there is a relatively smaller contiguous land area to contribute regional and local waxes. On the other hand, the overall contribution of local leaf waxes may be significantly higher on small islands where all non-local leaf waxes must be carried great distances.

4.3.2 Generic fatty acids

In contrast to the algal specific biomarkers dinosterol and \( n-C_{17} \)-alkane, \( \delta^2H \) values of \( n-C_{16} \) and \( n-C_{18} \) fatty acids were positively correlated with mean annual precipitation and negatively correlated with \( \delta^2H_P \) (Figure 2). These shorter chain fatty acids are synthesized by most organisms, but are frequently attributed to algal sources in aquatic sediments (Huang et al., 2004; Li et al. 2009). Heterotrophic and chemoautotrophic microbes produce short-chain fatty acids that can have \( \delta^2H \) values that differ by several hundred ‰ from those of photoautotrophs grown in similar water (Zhang et al., 2009; Heinzelmann et al., 2015). However, other than in microbial mats (Osburn et al., 2011), sedimentary \( n-C_{16} \) and \( n-C_{18} \) fatty acids typically have fractionation factors consistent with values from photoautotrophs in culture (Li et al, 2009; Zhang et al., 2009; Heinzelmann et al., 2018). In our tropical Pacific data set, fractionation factors between lake water and \( n-C_{16} \) and \( n-C_{18} \)-acids (\( \alpha_{\text{Lipid-Water}} = \frac{\left(^2\text{H} / ^1\text{H}\right)_{\text{Lipid}}}{\left( ^2\text{H} / ^1\text{H}\right)_{\text{Water}}} \)) ranged from 0.773 to 0.920. This large range in \( \alpha_{\text{Lipid-Water}} \) values is consistent with observations of cultures of different types of algae (Zhang and Sachs, 2007; Zhang et al., 2009; Heinzelmann et al., 2015). The \( \delta^2H \) values of the \( n-C_{16} \) and \( n-C_{18} \)-acids in our data set could be influenced by variable contributions from non-photoautotrophs, but could also vary due to differing contributions from different types of algae. In either case, it seems likely that \( \delta^2H \) values of these compounds reflect ecology more than hydroclimate, and their negative correlations with \( \delta^2H_P \) values in our sample set may be a coincidence. Dinosterol and \( n-C_{17} \)-alkane are sourced from a
smaller range of organisms than the near ubiquitous $n$-$C_{16}$ and $n$-$C_{18}$-acids (Cranwell et al., 1987; Meyers, 2003; Volkman, 2003), which could make their $\delta^2$H values more directly related to those of lake water.

4.4 Implications for paleoclimate reconstructions in the tropical Pacific

Although $\delta^2$H$_{Wax}$ values are strongly linearly correlated with $\delta^2$H$_P$ values on a global scale (Figure 3), the large residuals in this relationship indicate that caution should be applied before interpreting relatively small down-core changes in $\delta^2$H$_{Wax}$ values as hydroclimate changes. However, our data do not suggest that there are clear links between vegetation source (as indicated by palynological analyses) and residuals from the global $\delta^2$H$_{Wax}$-$\delta^2$H$_P$ relationship (Figure 6). Rather, one of the largest challenges for interpreting sedimentary $\delta^2$H$_{Wax}$ values in the tropical Pacific are uncertainties associated with modern estimates of $\delta^2$H$_P$ values, given the limited spatial and temporal available of modern observations. Recent isotope modeling work has helped constrain the processes that control $\delta^2$H$_P$ values in this dynamically important region (Conroy et al., 2016; Konecky et al., 2019). Continued effort in this regard is necessary to robustly interpret proxies $\delta^2$H$_P$ values, whether they are derived from $\delta^2$H$_{Wax}$ values or from other archives such as speleothems.

The expected inverse correlation between precipitation amount and OIPC-derived $\delta^2$H$_P$ values in the tropical Pacific, and the correlations between these variables and algal lipid $\delta^2$H values (Maloney et al., 2019), suggest that the uncertainty in $\delta^2$H$_P$ values cannot be solely responsible for the poor correlations associated with $\delta^2$H$_{Wax}$ values (Figure 2). Rather, factors besides $\delta^2$H$_P$ values that influence $\delta^2$H$_{Wax}$ values (variations in seasonality, catchment scales, contributions from organs other than leaves and/or from aquatic sources, or changes in biosynthetic fractionation) result in residuals that are on the order of $\pm25\%$ in the tropical Pacific.
and elsewhere (Figure 3). Any calibration of $\delta^2$H$_{Wax}$ values spanning a relatively small range of $\delta^2$H$_P$ values (like the 32‰ range studied here) is likely to have a poor correlation between the two variables (Figure 4). Correlation coefficients for the tropical Pacific are within the distribution generated by randomly subsampling the global data set while limiting the range in $\delta^2$H$_P$ to 35‰, but fall towards the low end of this range, more than 1 standard deviation below the mean correlation coefficient (Figure 4).

Regional calibrations are expected to have stronger than average correlations by constraining some variables that contribute to scatter in the global relationship between $\delta^2$H$_P$ and $\delta^2$H$_{Wax}$ values. However, the new data from the tropical Pacific exceed a regional scale in many ways, spanning a distance of ~8500 km, larger than even continental-scale studies (e.g., Sachse et al., 2004). The islands included range from low-lying atolls to mountainous volcanoes. The sites differ in seasonality of precipitation, vulnerability to tropical storms, sensitivity to El Niño–Southern Oscillation events, and biodiversity, all of which may impact $\delta^2$H$_{Wax}$ values. The diversity of sites, uncertainty in local estimates of $\delta^2$H$_P$ values, and small $\delta^2$H$_P$ signal relative to the noise in the global calibration, make it unsurprising that $\delta^2$H$_{Wax}$ values are not correlated with $\delta^2$H$_P$ values within the tropical Pacific. However, it is encouraging that tropical Pacific $\delta^2$H$_{Wax}$ values fall within the expected range, and do not have abnormally large residuals from the global calibration line. Together, these results suggest that the processes determining $\delta^2$H$_{Wax}$ values in tropical Pacific lake and swamp sediments are not fundamentally different than elsewhere.

The scatter in the global relationship between $\delta^2$H$_P$ and $\delta^2$H$_{Wax}$ (Figure 3) suggests that down-core changes as large as ~50‰ at any location could be driven by factors other than $\delta^2$H$_P$ values. In practice, at a single site where many variables that contribute to scatter among sites are constant, the threshold for detecting changes in $\delta^2$H$_P$ values may be significantly smaller.

Detailed processed-based studies at the catchment scale (Freimuth et al., 2019, 2020; Dior-
Kirschner et al., 2020) may be more useful for constraining sedimentary $\delta^2 H_{Wax}$ than additional large-scale calibration efforts.

Hydroclimate-driven interpretations of changes in sedimentary $\delta^2 H_{Wax}$ values will be most robust when supported by independent lines of proxy evidence, such as $\delta^2 H$ values of more source specific biomarkers like dinosterol (Smittenberg et al., 2011; Nelson & Sachs, 2016; Richey & Sachs, 2016; Sachs et al., 2018), changes in grain size distributions (Conroy et al., 2008), or changes in the elemental composition of inorganic sediments (Sear et al. 2020; Higley et al., 2018). Continued refinement of a multi-proxy toolbox that includes sedimentary $\delta^2 H_{Wax}$ values offers the best path to confidently reconstructing past hydrologic change.

5 Conclusions

$\delta^2 H_{Wax}$ values from surface sediments from 19 lakes and four swamps on 15 islands throughout the tropical Pacific fall within the overall range of values expected based on a global compilation surface sediment measurements ($R^2 = 0.74$ for both $n$-C$_{29}$-alkane ($n = 665$) and $n$-C$_{28}$-acid ($n = 242$)), and the residuals around the global linear regression between $\delta^2 H_{Wax}$ and $\delta^2 H_P$ are similar in the tropical Pacific and global data sets. Nevertheless, within the tropical Pacific there is no significant correlation between $\delta^2 H_{Wax}$ and $\delta^2 H_P$ values. The lack of correlation regionally is at least partly due to the large uncertainties in $\delta^2 H_P$ values derived from reanalysis data and cannot be ascribed to different vegetation sources within and surrounding the lakes in this study, as deduced from pollen assemblages.

To a first order on a global scale, $\delta^2 H_{Wax}$ values are clearly influenced by $\delta^2 H_P$ values, but the $\delta^2 H_P$ signal spanning the tropical Pacific remains small relative to the noise in the current global calibration. The global $\delta^2 H_{Wax,P}$ calibration remains limited by uncertainties in both the x- and y-axes, and could be improved by better constraints on $\delta^2 H_P$ values. As in other locations,
large changes in $\delta^2$H_{Wax} in sediments from tropical Pacific islands may be caused by variables other than $\delta^2$H$_P$, and could be improved by more catchment-scale, processed-based studies. In particular, interpretations need to consider the possible effects of changing source, growth conditions, and delivery of leaf waxes to sediments. When possible, $\delta^2$H$_{Wax}$ values should be paired with $\delta^2$H values of more source-specific compounds such as dinosterol, which can help distinguish changes in water isotopes from changes in other factors that affect $\alpha_{Wax-P}$ values. As is the case for all paleoclimate proxies, interpretations of $\delta^2$H$_{Wax}$ values are most robust in a multiproxy context.

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Figure captions
Figure 1 Global distribution of leaf wax samples from surface sediments and soils. The left panel shows the locations for $n$-C$_{29}$-alkane (665 sites); the right panel shows the locations for $n$-C$_{28}$-acid (242 sites). Background shading represents annual mean $\delta^2$H$_P$ values from the Online Isotopes in Precipitation Calculator (OIPC) (Bowen & Ravennaugh, 2003; IAEA/WMO, 2015; Bowen, 2020). OIPC does not produce spatial data sets over marine areas, therefore shading is limited to continents.

Figure 2: Correlation coefficients of linear regressions of $\delta^2$H values of all analyzed compounds relative to $\delta^2$H$_P$ values (blue circles) and mean annual precipitation (pink diamonds) in the tropical Pacific. Filled symbols represent significant correlations at the 95% confidence level. Mean annual precipitation is from the Global Precipitation Climatology Project (GPCP) and $\delta^2$H$_P$ values are from the OIPC. Compounds are grouped by source (algal, general, or plant waxes, with increasingly likely terrestrial plant sources associated with longer chain lengths). Dinosterol $\delta^2$H data is from Maloney et al. (2019), all other lipid $\delta^2$H data from this study. Individual measurements are included in Data Set S1.
Figure 3 δ²H values of (a, c) n-C29-alkane and (b, d) n-C28-acid from surface sediments and soils plotted relative to OIPC-derived δ²H_P values, color-coded by region (a, b), and sample type (c, d). In panels a and b, red diamonds are lakes from the SPCZ region and green squares are mangrove swamps in Micronesia (this study), both plotted with error bars. X-axis error bars represent 95% confidence intervals of OIPC values. Y-axis error bars represent 1 standard deviation of measurements from replicate samples from the same lake or swamp and are typically smaller than the marker size. Circles are global values compiled from the literature, color-coded by region. X-axis error bars are not shown for previously published data points, and average 5.2‰ for sites outside the tropical Pacific. Regression statistics in (a) and (b) are shown with and without new Pacific data. Globally compiled data (including tropical Pacific values) do not differ significantly between soils and lacustrine sediments for either (c) n-C29-alkane or (d) n-C28-acid. Shading around linear regressions represents 95% confidence intervals.

Figure 4 Correlation coefficients of random subsamples of 17 values from the global compilation surface sediment and soil of (a) n-C29-alkane and (b) n-C28-acid δ²H values plotted relative to range of δ²H_P values. Subsampled data were taken from the full data set and from restricted δ²H_P ranges as described in the text. Correlation coefficients for the global compilation and each continent are plotted for comparison.

Figure 5 Correlation plots of δ²H values from the global compilation of surface sediments and soils relative to the δ²H_P values from various models (described in section 2.4). Numbers, colors, and the widths of the ellipses correspond to correlation coefficients (R values).

Figure 6 Pollen distributions from surface or near surface sediments in tropical Pacific lake samples plotted relative to residuals from the global δ²H_Wax – δ²H_P calibration line for (a) n-C29-alkane and (b) n-C28-acid. Square and triangle symbols are used to distinguish among multiple sites with the same residual values.

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| Site, Island, Country | Lat. (°N) | Long. (°E) | # of samples | δ²H<sub>P</sub> (‰, VSMOW) | δ²H<sub>n--C<sub>29</sub> alkane</sup> (residual from global relationship) (‰, VSMOW) | δ²H<sub>n--C<sub>28</sub> acid</sup> (residual from global relationship) (‰, VSMOW) |
|-----------------------|-----------|-----------|--------------|-----------------|-------------------------------------------------|-------------------------------------------------|
| **Lakes**             |           |           |              |                 |                                                 |                                                 |
| Rimatu‘u Pond, Tetiaroa, French Polynesia | −17.0249 | 210.4417 | 2            | −25 ± 26        | −177 (−38 ± 12)                                  | −160 ± 17 (−28 ± 19)                              |
| Oroatera Pond, Tetiaroa, French Polynesia | −16.9958 | 210.4591 | 1            | −25 ± 26        | −173 (−34 ± 12)                                  | N.A.                                            |
| Onetahi Pond, Tetiaroa, French Polynesia | −17.0207 | 210.4081 | 1            | −25 ± 26        | −139 (−0 ± 12)                                   | −126 (5 ± 10)                                    |
| Lake Lanoto‘o, Upolu, Samoa | −13.9109 | 188.1726 | 3            | −34 ± 3         | −159 (−12 ± 1)                                   | −148 ± 6 (−9 ± 6)                                 |
| Lac Lalolalo, Wallis, Wallis and Futuna | −13.3017 | 183.7662 | 3            | −23 ± 1         | N.A.                                            | −150 ± 6 (−20 ± 6)                                |
| Lac Lanutavake, Wallis, Wallis and Futuna | −13.3212 | 183.7860 | 2            | −24 ± 2         | N.A.                                            | −140 ± 9 (−9 ± 9)                                 |
| Lake Dranoviteilomo, Vanua Balavu, Fiji | −17.1976 | 181.0441 | 2            | −21 ± 11        | −151 (−15 ± 5)                                   | −173 ± 14 (−44 ± 14)                              |
| Lake Tagamaucia, Teveuni, Fiji | −16.8163 | 180.0601 | 2            | −34 ± 14        | −170 ± 1 (−23 ± 7)                                | −175 ± 1 (−36 ± 5)                                |
| Otas Lake, Efate, Vanuatu | −17.6945 | 168.5850 | 1            | −34 ± 73        | −154 (−6 ± 35)                                   | −136 (2 ± 27)                                    |
| Emaotul Lake, Efate, Vanuatu | −17.7342 | 168.4151 | 3            | −36 ± 76        | −152 ± 9 (−3 ± 36)                                | −130 ± 3 (9 ± 26)                                 |
| White Lake, Thion, Vanuatu | −15.0410 | 167.0892 | 2            | −35 ± 70        | −174 (−25 ± 34)                                   | −119 ± 2 (21 ± 26)                                |
| Waérowa East Lake, Espiritu Santo, Vanuatu | −15.5950 | 167.0788 | 1            | −34 ± 71        | N.A.                                            | −155 (−16 ± 27)                                   |
| Nopovois Pond, Espiritu Santo, Vanuatu | −15.4970 | 166.7357 | 1            | −40 ± 71        | −154 (−1 ± 34)                                   | −122 (21 ± 27)                                    |
| Vesalea Pond, Espiritu Santo, Vanuatu | −15.1589 | 166.6549 | 1            | −40 ± 70        | −157 (−4 ± 34)                                   | N.A.                                            |
| Lake Hut, Grande Terre, New Caledonia | −22.2609 | 166.9526 | 2            | −15 ± 54        | −161 ± 2 (−31 ± 26)                               | −133 ± 1 (−8 ± 20)                                |
| Lake Tava, Tetepare, Solomon Islands | −8.7029  | 157.4503 | 1            | −46 ± 43        | −162 (−4 ± 21)                                   | −156 (−9 ± 16)                                    |
| Lake Rano, Rendova, Solomon Islands | −8.6879  | 157.3243 | 2            | −47 ± 42        | N.A.                                            | −135 ± 5 (13 ± 17)                                |
| Harai Lake #1, Rendova, Solomon Islands | −8.5622  | 157.3556 | 1            | −47 ± 42        | N.A.                                            | −121 (27 ± 16)                                    |
| Harai Lake #3, Rendova, Solomon Islands | −8.5648  | 157.3651 | 2            | −47 ± 42        | N.A.                                            | −134 ± 11 (15 ± 19)                                |
| **Mangrove swamps**   |           |           |              |                 |                                                 |                                                 |
| Sapwalap Swamp, Pohnpei, Fed. States of Micronesia | 6.88  | 158.30 | 5            | −33 ± 2         | −150 ± 5 (−3 ± 5)                                 | N.M.                                            |
| Tol Swamp, Chuuk, Fed. States of Micronesia | 7.35  | 150.60 | 4            | −32 ± 1         | −153 ± 5 (−2 ± 5)                                 | N.M.                                            |
| Sasa Swamp, Guam, United States | 13.45  | 140.73 | 4            | −29 ± 1         | −145 ± 5 (−7 ± 5)                                 | N.M.                                            |
| Galal Swamp, Yap, Fed. States of Micronesia | 9.50  | 138.08 | 5            | −34 ± 1         | −151 ± 6 (−3 ± 6)                                 | N.M.                                            |

1Less precision is provided for latitude and longitude in mangrove swamps because swamp samples were collected along a transect typically spanning > 1 km.

2Mean annual precipitation δ²H values (relative to VSMOW) from OIPC ± 95% confidence intervals.
Mean value of multiple surface sediment measurements from same lake, relative to VSMOW. Uncertainties represent 1 standard deviation. When only one sample was analyzed no uncertainty is reported. Analytical uncertainty for compound specific δ²H measurements is 4‰. “N.A.” = compound was not present or was below detection limit for δ²H measurements. “N.M.” = not measured.

Residuals are offsets from global calibration line of compiled leaf wax δ²H values from the literature. Uncertainties are site specific standard deviations of OIPC δ²H values, and are propagated with standard deviations of leaf wax δ²H values when multiple samples are available from a site.

*Lakes with greater than 50% vegetation cover.
Table 2 Pollen counts from near surface sediments, reported as a percentage of total palynmorphs counted. For each sediment sample, age ranges are presented for the top and bottom depth.

| Site, Island, Country | Depth of pollen sample (cm) | Bacon age at top of interval (year C.E.) | Bacon age at bottom of interval (year C.E.) | Primary forest (%) | Secondary forest (%) | Mangroves (%) | Ferns (%) | Non-vascular plants (%) | Dryland herbs (%) | Wetland herbs (%) | Aquatic plants (%) | Unknown (%) |
|-----------------------|-----------------------------|----------------------------------------|------------------------------------------|-------------------|---------------------|--------------|-----------|-----------------------|------------------|----------------|------------------|-------------|
| Lake Lanoto'o, Upolu, Samoa | 1-2                          | 2013 ± 1                               | 2002 ± 1                                 | 3.0               | 24.6                | 0            | 62.8      | 0                     | 2.7              | 6.9            | 0                | 0           |
| Lac Lalolalo, Wallis, Wallis and Futuna* | 1-3                          | 2001 +8 -14                            | 1991 +14 -19                            | 10.3              | 57.1                | 0            | 16.4      | 0                     | 0.4              | 13.4           | 0.8              | 1.7         |
| Lac Lanutavake, Wallis, Wallis and Futuna | 3-4                          | 1990 +20 -64                           | 1983 +26 -86                            | 13.1              | 69.1                | 0            | 4.3       | 0                     | 6.2              | 4.6            | 0                | 2.7         |
| Lake Dranoniveilomo, Vanua Balavu, Fiji | 2-3                          | 2010 ± 2                               | 2009 ± 3                                 | 21.0              | 33.6                | 0.7          | 17.5      | 1.4                    | 4.2              | 19.6           | 0                | 2.1         |
| Lake Tagamaucia, Teveuni, Fiji# | 2-3                          | 1989 ± 7                               | 1978 ± 10                                | 5.1               | 13.4                | 0            | 58.6      | 0                     | 0.6              | 21.3           | 0                | 1.0         |
| Otas Lake, Efate, Vanuatu | 2-3                          | N.A.                                   | N.A.                                    | 4.7               | 52.1                | 15.6         | 2.7       | 0                     | 0                | 21.8           | 0.8              | 2.3         |
| Emaotul Lake, Efate, Vanuatu | 1-2                          | 2016 ± 3.4                             | 2014 ± 3.4                               | 4.9               | 55.8                | 0            | 12.7      | 0.3                    | 11.4             | 8.4             | 5.2              | 1.3         |
| White Lake, Thion, Vanuatu | 3-4                          | 1997 +23 -14                           | 1991 +30 -19                            | 1.3               | 42.3                | 0            | 39.3      | 0                     | 2.1              | 11.7           | 0                | 3.4         |
| Waérowa East Lake, Espiritu Santo, Vanuatu# | 3-1                          | 2010 ± 3                               | 2009 ± 3                                 | 1.2               | 11.2                | 14.1         | 35.9      | 0                     | 11.2             | 14.7           | 7.7              | 4.1         |
| Nopovois Pond, Espiritu Santo, Vanuatu | 0-1                          | 2017                                   | N.A.                                    | 16.3              | 53.5                | 0            | 15.0      | 0                     | 8.9              | 3.1             | 0.3              | 2.6         |
| Vesalea Pond, Espiritu Santo, Vanuatu# | 0-1                          | 2016 +1 -3                             | 2005 +10 -14                            | 6.5               | 46.2                | 0            | 18.8      | 1.5                    | 11.4             | 6.8             | 4.6              | 4.3         |
| Lac Hut, Grand Terre, New Caledonia | 0-1                          | N.A.                                   | N.A.                                    | 50.1              | 41.1                | 0            | 4.8       | 0                     | 0                | 0.9             | 0                | 3.1         |
| Lake Tavara, Tetepare, Solomon Islands | 8-9                          | 1996 ± 5                               | 1993 ± 6                                 | 6.3               | 23.4                | 6.3          | 54.7      | 0                     | 1.6              | 7.8             | 0                | 0.1         |
| Lake Rano, Rendova, Solomon Islands | 9-10                         | 1969 +17 -16                           | 1960 +21 -20                            | 16.7              | 46.5                | 0            | 29.8      | 0                     | 0                | 4.4             | 0.9              | 1.8         |
| Harai Lake #1, Rendova, Solomon Islands | 11-12                       | 1716 +99 -123                          | 1702 +103 -120                          | 5.4               | 19.4                | 2.2          | 66.7      | 0                     | 0                | 5.4             | 0                | 1.1         |
| Harai Lake #3, Rendova, Solomon Islands | 30-31                       | 1871 ± 85                              | 1866 +86 -84                            | 3.1               | 5.2                 | 0            | 91.8      | 0                     | 0                | 0.0             | 0                | 0           |

*Mean of 2 samples from different sites in these lakes. Age ranges presented represent the full range of possible ages for both sites.

# Lakes with greater than 50% vegetation cover

$ Age ranges are provided from sites with existing age models, the details of which are provided by Maloney et al. (2019), Krentscher et al. (2019), Gosling et al. (2020), and Sear et al. (2020)
Figure 1.
Figure 2.
-1.0  -0.8  -0.6  -0.4  -0.2  0.0  0.2  0.4  0.6  0.8  1.0

-1.0  -0.8  -0.6  -0.4  -0.2  0.0  0.2  0.4  0.6  0.8  1.0

| Compound       | R value δ²H lipid vs δ²H OIPC | R value δ²H lipid vs mm/day (GPCP) |
|----------------|-------------------------------|-----------------------------------|
| n-C₁₆ acid (n = 19) |                               |                                   |
| n-C₁₈ acid (n = 13)  |                               |                                   |
| Dinosterol (n = 21)   |                               |                                   |
| n-C₁₇ alkane (n = 5)   |                               |                                   |
| n-C₂₃ alkane (n = 4)   |                               |                                   |
| n-C₂₅ alkane (n = 6)   |                               |                                   |
| n-C₂₆ acid (n = 12)   |                               |                                   |
| n-C₂₇ alkane (n = 8)   |                               |                                   |
| n-C₂₈ acid (n = 17)   |                               |                                   |
| n-C₂₉ acid (n = 17)   |                               |                                   |
| n-C₃₀ acid (n = 10)   |                               |                                   |
| n-C₃₁ alkane (n = 12)  |                               |                                   |
| n-C₃₃ alkane (n = 12)  |                               |                                   |

Aquatic
Terrestrial
With Tropical Pacific:
\[
m = 0.92 \pm 0.02; b = -117 \pm 2 \\
R^2 = 0.74; p < 0.0001; n = 665
\]
Without Tropical Pacific:
\[
m = 0.93 \pm 0.02; b = -116 \pm 2 \\
R^2 = 0.74; p < 0.0001; n = 648
\]

Soils
\[
m = 0.976 \pm 0.03; b = -115 \pm 2 \\
R^2 = 0.67; p < 0.0001; n = 474
\]

Lakes
\[
m = 0.902 \pm 0.03; b = -114 \pm 2 \\
R^2 = 0.82; p < 0.0001; n = 191
\]

N. America (<25° N)
\[
m = 0.73 \pm 0.03; b = -114 \pm 3 \\
R^2 = 0.74; p < 0.0001; n = 242
\]

Without Tropical Pacific:
\[
m = 0.74 \pm 0.03; b = -113 \pm 3 \\
R^2 = 0.73; p < 0.0001; n = 225
\]

**SPCZ**

| Location          | Intercept (±) | Slope (±) |
|-------------------|---------------|-----------|
| N. America (<25° N) | -114 ± 3     | 0.73 ± 0.03|
| N. America (25 - 60° N) | -113 ± 3   | 0.74 ± 0.03|
| N. America (> 60° N) | -113 ± 3     | 0.74 ± 0.03|

**Soils**

| Location | Intercept (±) | Slope (±) |
|----------|---------------|-----------|
| N. America (<25° N) | -102 ± 5     | 0.860 ± 0.03|
| Switzerland | -102 ± 5   | 0.860 ± 0.03|
| N. America (25 - 60° N) | -102 ± 5    | 0.860 ± 0.03|
| N. America (> 60° N) | -102 ± 5     | 0.860 ± 0.03|

**Lakes**

| Location          | Intercept (±) | Slope (±) |
|-------------------|---------------|-----------|
| N. America (<25° N) | -116 ± 2     | 0.976 ± 0.03|
| Switzerland | -116 ± 2     | 0.976 ± 0.03|
| N. America (25 - 60° N) | -116 ± 2    | 0.976 ± 0.03|
| N. America (> 60° N) | -116 ± 2     | 0.976 ± 0.03|

**Soils**

| Location | Intercept (±) | Slope (±) |
|----------|---------------|-----------|
| N. America (<25° N) | -115 ± 2     | 0.976 ± 0.03|
| Switzerland | -115 ± 2   | 0.976 ± 0.03|
| N. America (25 - 60° N) | -115 ± 2    | 0.976 ± 0.03|
| N. America (> 60° N) | -115 ± 2     | 0.976 ± 0.03|
Figure 4.
Range of $\delta^2H_P$ Values (‰)

Sampling range:

- All data
- 100‰
- 50‰
- 35‰

a) $n$-C$_{29}$-alkane

b) $n$-C$_{28}$-acid
Figure 5.
### Table: Correlation Coefficients

| n-C_{28}-alkane (n = 665) | OIPC | OIPC-extracted | OAM (free) | ECHAM (nudged) | GISS (nudged) | GISS (free) | HadAM (free) | IssGSM (nudged) | LMDZ (nudged) | MIROC (free) | SWING2 nudged mean |
|---------------------------|------|----------------|------------|----------------|---------------|-------------|--------------|----------------|----------------|---------------|------------------|
| n-C_{29}-alkane (n = 665) |      |               |            |                |               |             |              |                |                |               |                  |
| R                         | 0.74 | 0.86          | 0.84       | 0.64           | 0.67          | 0.70         | 0.69         | 0.57           | 0.66           | 0.65          | 0.67             | 0.70             |
| R                         | 0.86 | 0.82          | 0.91       | 0.87           | 0.84          | 0.86         | 0.86         | 0.89           | 0.86           | 0.85          | 0.89             | 0.88             |
Figure 6.
Primary forest  Secondary forest  Mangroves  Non-vascular plants  Dryland herbs  Wetland herbs  Aquatic plants  Ferns

Different shapes indicate different lakes with same residual values

(a) $n$-C$_{29}$-alkane residual

(b) $n$-C$_{28}$-acid residual

Relative abundance (%)

Relative abundance (%)