Anti-CD20 therapies for multiple sclerosis: current status and future perspectives

Monica Margoni1,6 · Paolo Preziosa1,2 · Massimo Filippi1,2,3,4,5 · Maria A. Rocca1,2,5

Received: 7 July 2021 / Revised: 2 August 2021 / Accepted: 3 August 2021 / Published online: 11 August 2021
© Springer-Verlag GmbH Germany, part of Springer Nature 2021

Abstract
Multiple sclerosis (MS) is a chronic inflammatory, demyelinating and neurodegenerative disease affecting the central nervous system (CNS), often characterized by the accumulation of irreversible clinical disability over time. During last years, there has been a dramatic evolution in several key concepts of immune pathophysiology of MS and in the treatment of this disease. The demonstration of the strong efficacy and good safety profile of selective B-cell-depleting therapies (such as anti-CD20 monoclonal antibodies) has significantly expanded the therapeutic scenario for both relapsing and progressive MS patients with the identification of a new therapeutic target. The key role of B cells in triggering MS disease has been also pointed out, determining a shift from the traditional view of MS activity as largely being ‘T-cell mediated’ to the notion that MS-related pathological processes involve bi-directional interactions between several immune cell types, including B cells, both in the periphery and in the CNS. This review provides an updated overview of the involvement of B cells in the immune pathophysiology and pathology of MS. We summarize the rationale regarding the use of anti-CD20 therapies and the results of the main randomized controlled trials and observational studies investigating the efficacy and safety profile of rituximab, ocrelizumab, ofatumumab and ublituximab. Suggestions regarding vaccinations and management of MS patients during COVID-19 pandemic with anti-CD20 therapies are also discussed. Finally, therapies under investigation and future perspectives of anti-CD20 therapies are taken into consideration.

Keywords Multiple sclerosis · Anti-CD20 therapy · Randomized clinical trials · Disease modifying therapy · B cells

Introduction

Multiple sclerosis (MS) is a chronic, inflammatory, demyelinating and neurodegenerative disease affecting the central nervous system (CNS), often leading to the accumulation of irreversible clinical disability.

During recent years, there has been a dramatic evolution in the arsenal of MS treatments. Among the developed therapies, anti-CD20 monoclonal antibodies mediating B-cell depletion, such as rituximab, ocrelizumab and ofatumumab, have received increasing attention. The demonstration of the strong efficacy and good safety profile of these selective B-cell-depleting therapies [1–3] has significantly contributed to expand the therapeutic scenario to treat MS patients.

An increasing amount of data regarding the use of anti-CD20 therapies has emerged. Accordingly, an updated overview is currently needed to summarize the most relevant findings supporting B-cell involvement in MS immune pathophysiology as well as the main results regarding
pharmacology, efficacy and safety of such therapies obtained from the most recent randomized controlled trials (RCTs) and observational studies. Guidelines for the proper timing of vaccinations and recent evidence about the risk of COVID-19 disease in association with anti-CD20 therapies are also discussed. Finally, future perspectives of anti-CD20 therapies and new promising anti-B-cell treatments are described.

Methods

We review the most recent evidence regarding the involvement of B cells in the immune pathophysiology and pathology of MS, the rationale underlying the use of anti-CD20 therapies and the results of the main RCTs and observational studies investigating the efficacy and safety profile of anti-CD20 therapies currently available.

References for this Review were identified through searches of PubMed with the search terms ‘adverse event(s)’, ‘antigen-presenting cell’, ‘atrophy’, ‘B-cell’, ‘blood’, ‘Bruton tyrosine kinase’, ‘CD19’, ‘CD20’, ‘COVID-19’, ‘cerebrospinal fluid’, ‘demyelination’, ‘depleting therapy(ies)’, ‘disability’, ‘disease activity’, ‘disease-modifying’, ‘gadolinium-enhancing lesion(s)’, ‘immunoglobulin’, ‘immunology’, ‘inhibitizumab’, ‘infusion reaction(s)’, ‘lymphocyte’, ‘malignancy(ies)’, ‘monoclonal antibody(ies)’, ‘MRI’, ‘multiple sclerosis’, ‘ocrelizumab’, ‘ofatumumab’, ‘outcomes’, ‘overall drug persistence’, ‘pathology’, ‘phenotype(s)’, ‘prediction’, ‘progression’, ‘primary progressive’, ‘randomized controlled trial’, ‘relapse’, ‘relapsing–remitting’, ‘rituximab’, ‘safety’, ‘SARS-CoV2’, ‘secondary progressive’, ‘T-cell’, ‘T2-hyperintense lesion(s)’, ‘tsolebrutinib’, ‘treatment’, ‘ublituximab’, ‘vaccination’ from 1 January 1979 to 5 July 2021. Articles were also identified through searches of the authors’ own files. Abstract presented at main congresses in the field were also evaluated. Only papers published in English were reviewed. The final reference list was generated on the basis of originality and relevance to the broad scope of this Review, with a focus on articles published during the past 3 years.

The role of B cells in MS immune pathophysiology

Historically, the classical view of MS immune pathophysiology, based on the convergence of studies from MS patients and experimental models, supposed that bouts of MS inflammatory activities are principally mediated by aberrantly activated and/or dysregulated pro-inflammatory CNS-reactive T effector (Teff) CD4 + cells (including T helper 1 [Th1], interleukin-17 [IL-17]-expressing CD4 [Th17], granulocyte–macrophage colony-stimulating factor [GM-CSF] expressing CD4) and CD8 + T cells (including IL-17-expressing CD8 [Tc17], GM-CSF expressing CD8, and mucosal-associated invariant T-cell receptor [TCR]-expressing CD8+ cells). By trafficking into the CNS, these cells are supposed to cause perivascular demyelination, glial cell activation and neuro-axonal injury [4, 5].

During the last few years, there has been a dramatic evolution in several key concepts of MS immune pathophysiology. One update involves a shift from the traditional view of MS disease activity as largely being ‘T-cell mediated’ to the view that MS relapses involve key bi-directional interactions between several immune cell types, including B cells, both in the periphery and in the CNS [4, 6].

This updated conceptual framework of the cellular immunology underlying MS activity has occurred after the demonstration of the strong efficacy of selective B-cell-depleting therapies (such as anti-CD20 monoclonal antibodies), pointing out the key role of B cells in triggering MS disease activity [1–3, 7].

The original impetus for targeting B cells in MS was based on the long-standing recognition of abnormally produced antibodies in the CNS of patients with MS (e.g., increased immunoglobulin [Ig] synthesis rates, cerebrospinal fluid (CSF)-restricted oligoclonal bands, antibodies bound to myelin fragments within phagocytic cells in the CNS parenchyma, Ig and complement detection in demyelinated lesions) [8–11].

Of note, B cells, plasmablasts and plasma cells are increased in the CSF of MS patients and their number is positively associated with intrathecal inflammation and Ig synthesis [12]. Despite this, the antigenic targets of the aberrant immune cell activation in MS remain incompletely defined and the long-term contribution of autoantibodies is largely unknown. Historically, the focus of investigation has been on myelin proteins, such as myelin basic protein (MBP), proteolipid protein (PLP) and myelin oligodendrocyte glycoprotein (MOG) [6, 13]. However, studies of circulating antibodies in MS patients including those directed against myelin antigens (MBP, MOG) [14] and the inward rectifying potassium channel (Kir) 4.1 [15, 16] have not led to the same pathogenic implications of specific CNS-directed antibodies, as those recognized in other conditions such as anti-aquaporin 4 (AQP4) antibodies in neuromyelitis optica spectrum disorders (NMOSD) or anti-MOG associated disease (MOGAD).

Growing evidence suggests that antibody-independent functions of B cells play key roles in mediating disease activity. B cells have been demonstrated to contribute to cascades of cellular immune interaction in the periphery, to act as antigen presenting cells (APCs) to T cells, thus promoting T-cell activation and proliferation, to interact with APCs to influence antigen trafficking, and to be directly involved in
the production of cytokines and chemokines exerting both anti- and pro-inflammatory actions and contributing to oligodendrocyte and neuronal damage (Fig. 1) [4, 6].

B cells are well-known efficient APCs, characterized by the expression of class-II major histocompatibility complex (MHC class II), and specialized in capturing soluble and membrane-tethered antigens, with a higher efficiency in presenting antigens and activating T cells than non-B-cell APCs [6, 17]. Due to the possible relevant role of B cells in processing CNS antigens, B cells could promote an increased activation of Teff, thanks to strong B-cell–T-cell interactions mediated by more than 20 co-stimulatory molecule–receptor pairs, with CD80/86 and their T-cell-activating binding partner CD28 being among the best characterized [6]. In addition to expressing co-stimulatory molecules, B cells can also express co-inhibitory molecules involved in downregulating the responses of Teff, such as the programmed death ligand 1 (PD-L1) and its receptor, programmed death 1 (PD-1) [6].

In MS, B cells are also recognized to have not only an abnormal propensity to produce pro-inflammatory cytokines (interleukin 6 [IL-6], GM-CSF, tumor necrosis factor alpha [TNF-α], and lymphotoxin alpha [LT-α]), but also a deficient capacity to produce regulatory cytokines (such as interleukin 6 [IL-6], GM-CSF, tumor necrosis factor alpha [TNF-α], and lymphotoxin alpha [LT-α]), but also a deficient propensity to produce pro-inflammatory cytokines [4, 6, 18–22]. Due to such an abnormal cytokine response profile, B cells can induce aberrant pro-inflammatory Th1, Th17 and myeloid cell responses, contributing to the cellular immune cascades involved in disease activity [4, 6, 18–22].

### B cells in MS pathology

Pathological studies have consistently shown that B cells significantly contribute to MS pathology in the CNS [23–27]. B-cell infiltrates are significantly higher in MS compared with other inflammatory CNS diseases, especially in patients at early stages of MS and with active lesions.

In early and active focal demyelinating lesions, CD20+ B cells are mainly located focally in the perivascular space of only one or a few larger veins and have pro-inflammatory functions (Fig. 1) [23, 24]. Conversely, a more abundant plasma cell infiltrate can be found in the perivascular space and in the meninges from patients with progressive MS (Fig. 1) [23, 24]. This evidence suggests a gradual differentiation of infiltrating B cells into a stable plasma cell population, showing expression of markers involved in B-cell survival and plasmablast differentiation (CD27 and CD38) [23, 24].

In addition to cascades of the peripheral cellular immune interactions contributing to ‘relapse biology’, there is also an important role for a ‘CNS-compartmentalized’ inflammation that sustains chronic inflammation, demyelination, and neurodegeneration, which can be maintained in the absence of ongoing relapse biology.

This ‘CNS-compartmentalized’ inflammation is characterized by prominent B-cell-rich inflammatory aggregates resembling tertiary lymph follicles that can be found in the meninges of MS patients, mainly within deep cortical sulci, but also in the perivascular spaces (Fig. 1). These inflammatory aggregates in the CNS may provide an environment that fosters B-cell homing, survival and functional activation [28, 29], and, in turn, contribute to degenerative mechanisms [30].

For instance, the extent of meningeal inflammation and the levels of pro-inflammatory cytokines (e.g., interferon gamma [IFN-γ], TNF-α, LT-α, IL-6) in the CSF of MS patients have been associated with the severity of subpial cortical demyelination, promoting also a graded pattern of neuronal loss and microglial activation consistent with a ‘surface-in’ process possibly mediated by one or more toxic substances contained in the CSF [25–27, 31, 32].

### Mechanisms of action of anti-CD20 therapies

CD20 is a transmembrane, non-glycosylated phosphoprotein of 33–37 kDa that is expressed in tetramers associated with lipid rafts on the surface of cell lineage from pre-B cells to naïve and memory B cells (Fig. 2) [33].

Monoclonal antibodies directed against specific targets typically deplete targeted cells through at least four possible different mechanisms: (i) antibody-dependent cellular cytotoxicity (ADCC); (ii) complement-dependent cytotoxicity (CDC); (iii) antibody-dependent cellular phagocytosis (ADCP); and (iv) induction of cell apoptosis. Currently available anti-CD20 monoclonal antibodies induce B-cell depletion mainly through ADCC, CDC and ADCP [33].

The infusion of anti-CD20 monoclonal antibodies promotes a depletion of CD20+ B cells within hours, mainly occurring in the liver [34]. Such a depletion reaches the nadir typically after 8 weeks and can be sustained for several weeks to months according to the posology and the features of the specific anti-CD20 monoclonal antibody.

After anti-CD20 therapies, B-cell repopulation starts in bone marrow and spleen, followed by blood, with a different rate between memory B cells and CD19+ cells [35].

Of note, B-cell counts are usually determined using CD19, which largely overlaps with CD20 during B-cell differentiation, because it is less prone to potential interference in presence of anti-CD20 monoclonal antibodies [35].

B-cell reappearance can be defined when CD19+ B cells reach 1% of lymphocyte counts [36]; however, other criteria are also applied, including 2% of CD19+ B cells [36].
Fig. 1 Summary of the involvement of B cells in the immune pathophysiology and pathology of MS. 

A Roles of B cells in immunity and disimmunity. In MS, B cells are involved in innate immunity, antigen presentation, production of regulatory and pro-inflammatory cytokines, chemokines and autoantibodies.

B, C Overview of the distribution of B cells in the different CNS areas involved in MS pathology. B In an active MS lesion with a central inflamed vein, a demyelinated core, with microglia and macrophages, and a rim of active ongoing demyelination with activated microglia, macrophages with different stages of myelin degradation, and oligodendrocyte injury, the highest density of lymphocytes is seen in the perivascular space of the central vein, with the majority of B cells in the lesion present at this site. C Aggregates of B cells can also be observed in the leptomeninges. This compartmentalized inflammation, characterized by the development of ectopic follicle-like lymphoid aggregates, is mainly driven by B cells, plasma cells, T cells and follicular dendritic cells. Created with biorender.com. See text for further details.

BCR B-cell receptor, Breg regulatory B-cell, Ig immunoglobulin, GM-CSF granulocyte–macrophage colony-stimulating factor, IL-6 interleukin 6, IL-10 interleukin 10, IL-35 interleukin 35, MHC class II major histocompatibility complex class II, NAWM normal-appearing white matter, PD-L1 ligand programmed death ligand 1, TCR T-cell receptor, Tₜeff effector T-cell, TGF-β transforming growth factor beta, TLR Toll-like receptor, TNF-α tumor necrosis factor alpha.
Efficacy of anti-CD20 therapies

Rituximab

Rituximab is a chimeric monoclonal antibody that is commonly prescribed in highly active MS patients, although it is not approved from Food and Drug Administration (FDA) nor European Medicines Agency (EMA) for use in MS (Fig. 2).

Its efficacy has been investigated in several studies as summarized in Table 1.

In particular, in a 48-week phase-II RCT (HERMES), 104 relapsing–remitting (RR) MS patients were randomized to receive either a single administration of i.v. rituximab 1000 mg or placebo on days 1 and 15 [1]. Compared with placebo, patients who received rituximab had a lower ARR at week 24 (p = 0.04), not confirmed at week 48 (p = 0.08), and a reduction of T2-hyperintense lesion volume (LV) from baseline to week 24 (p = 0.008) and 36 (p = 0.004). From week 12, rituximab also reduced Gadolinium (Gd)-enhancing lesions (p ≤ 0.003).

The efficacy of rituximab has been also evaluated in a 96-week phase-II/III RCT of 439 primary progressive (PP) MS patients (OLYMPUS) [37]. Rituximab (two i.v. administrations of 1000 mg 2 weeks apart) was not associated with changes in the proportion of patients developing confirmed disease progression (CDP) (p = 0.14), the primary outcome of the study, nor in brain atrophy rate (p = 0.62). However, it promoted a significant lower increase of T2-hyperintense LV at week 96 (p < 0.001) compared with placebo [37]. In a subgroup analysis, rituximab showed a delayed time to CDP in younger PPMS patients (aged < 51 years) or those with Gd-enhancing lesions at baseline. An additive predictive
| References         | Number and type of patients | Trial design                                           | Clinical findings                                                                                     | MRI findings                                                                                           | Adverse effects                                                                                     |
|--------------------|-----------------------------|-------------------------------------------------------|------------------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------|
| Bar-Or et al. [72] | 26 RRMS                     | Phase I, multicenter open label (72 weeks)            | ↓ mean ARR from b to week 72 (1.27 → 0.18)                                                          | ↓ T2 LV (↓ 119.6 mm³ at week 48, ↓ 272.7 mm³ at week 72)                                              | AEs in all patients (77% mild-moderate, 23% severe)                                               |
|                    |                             |                                                       | ↓ Gd+ lesions from b to week 72 (1.31 → 0)                                                          | ↓ Gd+ lesions from b to week 72 (1.31 → 0)                                                          | No serious AEs                                                                                     |
| Hauser et al. [1]  | 104 RRMS                    | Phase II, randomized, parallel, double-blind, Pbo-controlled study (HERMES-NCT00097188) (48 weeks) | ↓ ARR at week 24 (0.37 with RTX vs 0.84 Pbo, p = 0.04)                                              | ↓ in T2 LV (−163 mm³ with RTX vs +436 mm³ with Pbo at week 24, p = 0.008, −175 mm³ with RTX vs +418 mm³ with Pbo at week 36, p = 0.004) | More IARs with RTX (87.3% vs 40%) after first infusion, opposite after second infusion (20.3% vs 40%), 7.4% severe, remaining mild–moderate |
|                    |                             |                                                       | ↓ in T2 LV (−163 mm³ with RTX vs +436 mm³ with Pbo at week 24, p = 0.008, −175 mm³ with RTX vs +418 mm³ with Pbo at week 36, p = 0.004) | ↓ Gd+ lesions at weeks 12, 16, 20, 24, and 48 (0.5 with RTX vs 5.5 with Pbo; RR = 91%, p < 0.001) | Similar numbers of serious AEs with RTX vs Pbo (13% vs 14.3%)                                      |
|                    |                             |                                                       |                                                     | Similar rate of brain atrophy between groups (−13.1 cm³ with RTX vs −14.0 cm³ with Pbo, p = 0.62) | Similar number of infections (69.6% vs 71.4%)                                                      |
|                    |                             |                                                       |                                                     | ↓ Gd+ lesions (74% at b vs 26% at week 20, p < 0.0001)                                             | No serious AEs (2 withdrawn due to IARs)                                                           |
| Hawker et al. [37] | 439 PPMS                    | Phase II/III, randomized, double-blind, Pbo-controlled study (OLYMPUS-NCT00087529) (96 weeks) | Proportion of patients with 3-month CDP not significantly different at week 96 (RTX = 30.2% vs Pbo = 38.5%, p = 0.14) | Less ↑ of T2 LV at week 96 (median increase: +302.0 mm³ with RTX, +809.5 mm³ with Pbo, p < 0.001) | 16.1% with RTX vs 13.6% Pbo with serious AEs                                                      |
|                    |                             |                                                       | Delayed CDP in patients aged <51 years at b with RTX                                               | Similar rate of brain atrophy between groups (−13.1 cm³ with RTX vs −14.0 cm³ with Pbo, p = 0.62) | 4.5% with RTX vs <1% Pbo serious infections                                                      |
|                    |                             |                                                       |                                                     | Delayed CDP in patients with Gd+ lesions at b with RTX                                             | Mild–moderate IARs with RTX during the first course, decreased to rates comparable to Pbo with successive courses |
|                    |                             |                                                       |                                                     | No effect on T2 and T1 lesion burden                                                              | No serious AEs                                                                                        |
|                    |                             |                                                       |                                                     | ↓ Gd+ lesions (74% at b vs 26% at week 20, p < 0.0001)                                             | Uncomplicated urinary tract infections                                                             |
| Naismith et al. [73]| 32 RRMS                     | Phase II single center (no Pbo-group)—no MRI until week 20 (52 weeks)                             | Stable EDSS during follow-up ↑ MSFC improvement (+0.039 z-score)                                     | No serious AEs (2 withdrawn due to IARs)                                                            | ↓ upper respiratory tract infection                                                                |
|                    |                             |                                                       |                                                     | ↓ Gd+ lesions (74% at b vs 26% at week 20, p < 0.0001)                                             |↓ more AEs with FTY (21%) vs RTX (5%)                                                               |
|                    |                             |                                                       |                                                     | No effect on T2 and T1 lesion burden                                                              | Moderate IARs                                                                                       |
| Alping et al. [74] | 256 RRMS                    | Prospective multicenter (at least 1-year follow-up)                                              | ↓ relapses with RTX vs FTY (1.8% vs 17.6%)                                                          | ↓ Gd+ lesions in RTX vs FTY (1.4% vs 24.2%)                                                         | 3 serious AEs (pyelonephritis, influenza)                                                          |
| De Flon et al. [75]| 75 RRMS                     | Open-label, uncontrolled phase II study (24 weeks)                                               | 5 patients experienced disease activity (2 clinical relapses, 4 MRI activity)                        | ↓ Gd+ lesions (0.028 at b → 0.036 at 6 months)                                                     | ↓ IARs in 7.8%                                                                                        |
| Salzer et al. [52] | 822 MS (557 RRMS, 198 SPMS, 67 PPMS) | Retrospective uncontrolled observational study (mean follow-up 21.8 months) | Stable EDSS in RRMS, ↑ in SPMS/PPMS ↓ ARR (RRMS = 0.044, SPMS = 0.038, PPMS = 0.015)                | ↓ Gd+ lesions (26.2% at b → 4.6% at the last available follow-up)                                 | 89 AEs grade > 2 in 70 patients (infections)                                                       |
| References        | Number and type of patients | Trial design                                                                 | Clinical findings                                                                 | MRI findings                                                                 | Adverse effects                                                                                     |
|------------------|-----------------------------|------------------------------------------------------------------------------|----------------------------------------------------------------------------------|------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------|
| Alcala et al. [76] | 90 MS (31 RRMS, 45 SPMS, 14 PPMS) | Retrospective single-center (follow-up 6 months–5 years)                      | ↓ 88.4% ARR NEDA-3 at 1 year; all MS = 70% RRMS = 74.2%, PMS = 67%                | ↓ Gd+ lesions (2.56 at b → 0.06 at the last available follow-up, \( p < 0.001 \)) | 18.8% IARs 4 SAE (1 agranulocytosis, 3 thrombotic events, 1 death due to pulmonary embolism)     |
| Durozard et al. [77] | 50 RRMS                     | Nationwide retrospective multicenter (median follow-up 1.1 years)             | ↓ ARR (0.8 pre-RTX → 0.18 post-RTX, \( p < 0.001 \))                           | ↓ Gd+ lesions (72% pre-RTX → 8% post-RTX, \( p < 0.001 \))                   | 16 AEs, 10 patients with ≥ 1 AE (mainly infections) 3 SAEs 2 treatment discontinuations due to AE |
| Granqvist et al. [78] | 120 RRMS                    | Retrospective multicenter (follow-up up to 4 years)                          | ↓ ARR with RTX vs injectable DMTs \( p < 0.01 \)                                 | ↓ Gd+ lesions with RTX- (1.7%) vs injectable DMTs (12.6%) and DMF (12.8%)  | No serious AEs with RTX Mild AEs more common for injectable DMTs vs RTX 64 AEs with RTX (71.9% of patients), IARs (25.8%) 2 serious AEs (pyoderma gangrenosum, increase in meningioma size) |
| Yamout et al. [79] | 59 RRMS, 30 PMS              | Retrospective single center (mean follow-up 22.2 months)                      | ↓ ARR (1.07 → 0.11 RRMS and 0.25 → 0.16 PMS) Stable EDSS in both groups NEDA-3 = 74% at 1 year | ↑ of patients free from new MRI lesions (18.6 → 92.6% in RRMS and 43.3 → 82% in PMS) | No serious AEs with RTX Mild AEs more common for injectable DMTs vs RTX 64 AEs with RTX (71.9% of patients), IARs (25.8%) 2 serious AEs (pyoderma gangrenosum, increase in meningioma size) |
| Honce et al. [80] | 55 RRMS                     | Prospective double-blind single center (mean follow-up 1.5 years)            | NEDA-3 = 44.4% with RTX-GA vs 19.2% Pbo-GA                                     | ↑ proportion of patients with new T2 lesions (25.9% RTX-GA vs 61.5% Pbo-GA, \( p = 0.009 \)) | ↑ IARs in RTX 4 serious AEs in RTX, 5 in Pbo                                                    |
| Zecca et al. [81] | 355 MS (188 RRMS, 43 PPMS, 124 SPMS) | Retrospective, uncontrolled, observational study (median treatment 1.9 years) | ↓ ARR vs 1 year before (RRMS = 0.86 → 0.09, \( p < 0.001 \); SPMS = 0.34 → 0.06, \( p < 0.001 \); PPMS = 0.12 → 0.07, \( p = 0.45 \)) | At m12, 15.8% had new T2 and/or Gd+ lesions; 4.1% Gd+ lesions and 13.4% new T2 lesions | 23.7% at least 1 IAR 3.1% serious AE 8 patients withdrew 1 death due to mediastinal neoplasm       |

AE adverse events, ARR annualized relapse activity, CDP confirmed disability progression, DMTs disease-modifying therapies, EDSS expanded Disability Status Scale, FTY fingolimod, GA glatiramer acetate, Gd+ gadolinium-enhancing, IARs infusion-associated reactions, IFN-β1a interferon-β1a, LV lesion volume, MRI magnetic resonance imaging, MSFC multiple sclerosis functional composite, NEDA-3 no evidence of disease activity 3, Pbo placebo, PMS progressive multiple sclerosis, PPMS primary progressive multiple sclerosis, RR relative reduction, RRMS relapsing–remitting multiple sclerosis, RTX rituximab, SPMS secondary progressive multiple sclerosis
effect of age and Gd-enhancing lesions at baseline was found, suggesting that B-cell depletion might be effective in PPMS patients who are younger and have higher inflammatory activity [38].

Although further exploration of efficacy has not been carried out in phase-III RCTs, several other observational studies have confirmed a significant reduction of disease activity with rituximab (Table 1).

Few studies have utilized an intrathecal approach to administer rituximab to target compartmentalized inflammation in progressive MS. In an open-label study in eight progressive MS patients with MRI evidence of meningeal inflammation, a significant and sustained reduction in circulating B cells and a transient drop in CSF B cells were observed, but this did not translate in a change in the number or appearance of leptomeningeal enhancement [39].

Ocrelizumab

Ocrelizumab is a humanized anti-CD20 monoclonal antibody (Fig. 2) approved by the FDA (March 2017) and EMA (January 2018), at a dose of 600 mg i.v. twice yearly, as a therapy for the treatment of highly active relapse-onset MS and PPMS with evidence of disease activity.

A first phase-II RCT explored ocrelizumab efficacy in RRMS patients, assigned to either i.v. low (600 mg; n = 55) or high (2000 mg; n = 55) ocrelizumab administrations divided into two doses on days 1 and 15, i.v. placebo (n = 54) or weekly intramuscular (i.m.) IFNβ-1a (30 µg; n = 54) (Table 2) [40]. RRMS patients treated with ocrelizumab showed a significantly lower ARR (0.13 in the low- and 0.17 in the high-dose group) compared with placebo (0.64) and IFNβ-1a (0.36) group [40]. Change in T2-hyperintense LV did not differ among groups at week 24 (p = 0.2 in the low- and high-dose groups; p = 0.5 in the IFNβ-1a group), whereas the total number of Gd-enhancing lesions was significantly lower in ocrelizumab groups compared with placebo- and IFNβ-1a groups (both p < 0.001).

At week 24, patients initially treated with placebo, 600 mg ocrelizumab and i.m. IFNβ-1a received 600 mg of ocrelizumab, whereas the 2000 mg ocrelizumab group received 1000 mg of ocrelizumab. An interim analysis of the open-label extension (OLE) phase showed a maintained low ARR in ocrelizumab groups (0.06 vs 0.07 in the placebo/ ocrelizumab group and 0.07 in the IFNβ-1a/ocrelixumab group) over 96–144 weeks [41].

Two identical, phase-III RCTs, including 821 (OPERA I) and 835 (OPERA II) RRMS patients [2], further demonstrated ocrelizumab efficacy (Table 2). Patients were randomized (1:1) to i.v. ocrelizumab 600 mg every 24 weeks or subcutaneous (s.c.) IFNβ-1a 44 µg three times per week over 96 weeks. Compared with IFNβ-1a, ocrelizumab showed a higher reduction of ARR (p < 0.001), a lower prevalence of 12-week CDP (p < 0.05), and lower numbers of new/enlarged T2-hyperintense and Gd-enhancing lesions (p < 0.001) [2]. The proportion of RRMS patients with 12-week confirmed disability improvement was also higher with ocrelizumab compared with IFNβ-1a, being significant in the OPERA I (p = 0.01) but not in the OPERA II (p = 0.40) [2]. Percentage of brain volume loss from week 24 to 96 was significantly lower in ocrelizumab- vs IFN-β-1a-group in the OPERA I (p = 0.004), but not in the OPERA II (p = 0.09) [2].

In the 3-year follow-up OLE [42], the cumulative proportion of patients with 24-week CDP was lower in patients who initiated ocrelizumab earlier vs those initially receiving IFNβ-1a (p = 0.014). ARR did not differ between RRMS patients continuing ocrelizumab and those receiving ocrelizumab after switching from IFNβ-1a (p ≥ 0.70); both groups attained an almost complete and sustained suppression of brain MRI lesion activity.

The proportion of RRMS patients with no evidence of disease activity 3 (NEDA-3) (i.e., no relapses, no disability progression, no new/enlarged T2-hyperintense or Gd-enhancing lesions) was also significantly higher in ocrelizumab groups compared with the INFβ-1a groups both in OPERA I and OPERA II (p < 0.001) [43]. In the OLE period, the proportion of patients with NEDA-3 was 65.4% in patients continuously treated with ocrelizumab compared with 55.1% in those switching from IFNβ-1a to ocrelizumab (p < 0.001, relative difference = 19%).

Some preliminary results of the open-label, prospective, single-arm, phase-IIb ENSEMBLE study showed that early RRMS (disease duration ≤ 3 years) treated with ocrelizumab had a high NEDA-3 rate (84.8%) after 1 year of treatment [44]. Similarly, a high 2-year NEDA-3 rate (74.9%) was detected in the primary analysis of the phase-IIb CASTING trial, which evaluated ocrelizumab efficacy in RRMS patients with prior suboptimal response to one or two disease-modifying therapies (DMTs) (Table 2) [45].

Ocrelizumab has been the first DMT showing significant effects in PPMS, as demonstrated in the phase-III RCT (ORATORIO), which evaluated 732 PPMS patients receiving either i.v. ocrelizumab (600 mg) (n = 488) or placebo (n = 244) every 24 weeks for at least 120 weeks [2]. Ocrelizumab reached the primary study endpoint since it was associated with a significant reduction of 12-week CDP (p = 0.03) compared with placebo, further confirmed with the 24-week CDP (p = 0.04) [2]. The prevalence of worsening on timed 25-foot walk test (25-FWT) was also significantly reduced with ocrelizumab (p = 0.04) [2]. Brain T2-hyperintense LV decreased with ocrelizumab and increased with placebo from baseline to week 120 (p < 0.001), and the rate of brain atrophy was significantly lower with ocrelizumab compared with placebo (p = 0.02) [2].

Similarly to rituximab, a post hoc analysis (pooled OPERA studies) suggested that patients who were younger
| References          | Number of patients | Trial design                                                                 | Clinical findings                                                                 | MRI findings                                                                                                           | Adverse effects                                                                                           |
|---------------------|--------------------|-------------------------------------------------------------------------------|----------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------|
| Kappos et al. [40]  | 220 RRMS           | Phase II, randomized, parallel, double-blind, Pbo-controlled study (48 weeks) | ↓ ARR with OCR (low dose = 0.13; high dose = 0.17) vs Pbo (0.64) and INFβ-1a (0.36) | ↓ Gd+ lesions with 600 and 2000 mg OCR (0.6 and 0.2, respectively) vs Pbo (5.5) and INFβ-1a (6.9)                  | ↑ IARs with OCR vs Pbo (44% vs 9%) No ↑ serious AE Similar rate of infections |
| Hauser et al. [2]   | 821 RRMS (OPERA I) | Phase III, randomized, double-blind, active-controlled, parallel group studies (OPERA I) (96 weeks) | ↓ ARR with OCR (46% and 47%, p < 0.001) vs IFN-β1a ↓ proportion of patients with 3-month CDP with OCR (34% and 37%, p < 0.05) | ↓ Gd+ lesions (94% OPERA I; 95% OPERA II, p < 0.001) ↓ new/enlarged T2 lesions (77% OPERA I; 83% OPERA II, p < 0.001) with OCR vs INFβ-1a ↓ percentage of brain volume loss from week 24 to 96 in OPERA I (− 0.57% vs − 0.74%, p = 0.004) but not in OPERA II (− 0.64% vs − 0.75%, p = 0.09) | ↑ IARs 34% OCR vs 10% INFβ-1a or Pbo ↑ infections 59.9% with OCR (vs 54.3% INFβ-1a) in OPERA I, and 60.2% (vs 52.5% with INFβ-1a) in OPERA II → ↑ upper respiratory tract infections with OCR No ↑ serious AE ↑ neoplasm OCR (0.4%) vs INFβ-1a (0.2%) |
| Montalban et al. [2]| 732 PPMS           | Phase III, double-blind, randomized, Pbo-controlled, parallel group study (ORATORIO) (120 weeks) | ↓ proportion of patients with 3-month CDP (32.9% OCR vs 39.3% Pbo, p = 0.03) and 6-month CDP (29.6% vs 35.7%, p = 0.04) | ↓ 34% T2 lesions from b to week 120 (mean change, − 3.4% with OCR vs 7.4% with Pbo, p = 0.001) ↓ loss of brain volume (− 0.90 with OCR vs − 1.09 with Pbo, p = 0.02) | ↑ IARs with OCR (40%) vs Pbo (26%) ↑ infections with OCR (71.4%) vs Pbo (69.9%) → ↑ upper respiratory tract infections with OCR No ↑ SAE ↑ neoplasm with OCR (2.3%) vs Pbo (0.8%) |
| Turner et al. [46]  | 1656 RRMS          | Phase III, randomized, double-blind, active-controlled, parallel group studies (pooled OPERA I and OPERA II) (96 weeks) | ↓ ARR and NEDA-3 re-baselined at week 24 in patients aged < 40 years or with ≥ 1 Gd+ lesion at b with OCR | ↓ Gd+ lesions in patients aged < 40 years or with ≥ 1 Gd+ lesion at b with OCR | AE consistent with past reports and no new safety signals emerged with prolonged treatment |
| Ellwardt et al. [82]| 210 MS (155 RRMS/SPMS, 55 PPMS) | Retrospective, single-center (median follow-up 200 days) | 13% of patients experienced a relapse and 5% experienced a 12-week CDP |                                                                                                                      | 22% AE, 9% IARs Minor infections (8%) and 2 cases of a prolonged herpes labialis I case of toxic drug-induced hepatopathy |
| Hauser et al. [3]   | 702 RRMS           | Open-label extension, phase-III trials (OPERA I and OPERA II) (96 weeks)       | ↓ proportion of patients with 6 months CDP (16.1% with OCR/OCR vs 21.3% with IFN-β-1a/OCR at y5, p = 0.014) NEDA-3=65.4% with OCR/OCR vs 55.1% with IFN-β-1a/OCR (p < 0.001) | Sustained suppression of new brain MRI lesion activity from years 3 to 5. ↓ whole brain volume loss at 5 years vs b in those starting OCR earlier (OCR/OCR = − 1.87% vs IFN-β-1a/OCR = − 2.15%; p < 0.01) | AE consistent with past reports and no new safety signals emerged with prolonged treatment |
| Hartung et al. [44] | 678 RRMS           | Open-label, prospective, single-arm, phase-IIIb study (ENSEMBLE)               | 92.8% of patients free from clinical disease activity NEDA-3=84.8% after 1 year of treatment | 91.3% of patients free from MRI disease activity | AE consistent with past reports |

Table 2 Clinical trials assessing treatment with ocrelizumab in patients with MS
(aged < 40 years) and with baseline disease activity (≥ 1 Gd-enhancing lesions) had a greater treatment benefit with ocrelizumab, relative to IFNβ-1a, than patients who were older and with inactive disease [46].

An interim report from ORATORIO OLE has shown consistent and sustained treatment-associated benefit in multiple measures of CDP and a good safety profile over 6.5 years [47]. Mean percentage changes of brain T2-hyperintense (p < 0.001) and T1-hypointense (p < 0.001) LVs were lower in patients who initiated ocrelizumab early than in those initially receiving placebo from baseline to week 168 [47]. No difference in the rates of whole brain (p = 0.13) and cortical gray matter atrophy (p = 0.38) from baseline to week 144 was found between the two groups.

Some ongoing RCTs and observational studies are investigating additional aspects, including long-term effectiveness and safety of ocrelizumab and of treatment switch from natalizumab or rituximab [48].

**Ofatumumab**

Ofatumumab is the first fully human anti-CD20 monoclonal antibody, with a 20 mg s.c. monthly dosing regimen (Fig. 2), which has been approved by FDA (August 2020) and EMA (March 2021) for the treatment of active relapsing MS forms.

A phase-II RCT evaluated 38 RRMS patients who received two i.v. infusions of ofatumumab 100, 300, or 700 mg or placebo 2 weeks apart (Table 3) [49]. After 24 weeks, ofatumumab promoted an almost complete suppression of brain new/enlarging T2-hyperintense and Gd-enhancing lesions (> 99%).

Another phase-II RCT, the MIRROR study, randomized 232 RRMS patients [50] into placebo, ofatumumab 3, 30 or 60 mg every 12 weeks, or ofatumumab 60 mg every 4 weeks. All patients were treated for 24 weeks and followed up until B-cell repletion. Overall, 26 RRMS patients had a relapse during the first 12 weeks, 11 (42%) of whom during the first 4 weeks. Over 24 weeks, 17 (25%) RRMS patients relapsed in the placebo group compared with three to ten RRMS patients (9–22%) in the ofatumumab groups. Most patients (79%) had unchanged EDSS scores at weeks 12 and 24 [50]. With all ofatumumab regimens, the mean cumulative number of Gd-enhancing lesions was reduced by 65% from baseline to week 12 (p < 0.001), with reductions ≥ 90% for each dose ≥ 30 mg (p < 0.002) [50].

Two recent phase-III RCTs (ASCLEPIOS I and II) in RRMS patients compared the efficacy and safety of ofatumumab (20 mg, s.c. administration every 4 weeks) (465 and 481 RRMS patients) with oral teriflunomide 14 mg daily (462 and 474 patients) [3]. Ofatumumab groups showed a significant lower ARR compared with teriflunomide groups (p < 0.001). In pooled analysis, ofatumumab significantly
Table 3  Clinical trials assessing treatment with ofatumumab in patients with MS

| References            | Number of patients | Trial design                                                                 | Clinical findings                                                                 | MRI findings                                                                 | Adverse effects                                                                 |
|-----------------------|--------------------|------------------------------------------------------------------------------|----------------------------------------------------------------------------------|-------------------------------------------------------------------------------|--------------------------------------------------------------------------------|
| Sorensen et al. [49]  | 38 RRMS            | Phase II, double-blind, randomized, Pbo-controlled study (48 weeks)          | Lower proportion of patients with relapse(s) with OFT vs Pbo (19% vs 25%)         | ↓ new/expanding T2 lesions at week 24 (p < 0.001)                              | Mostly mild-to-moderate severity AEs 2 patients discontinued for grade-2 (pruritic rash, bronchospasm, cough) and grade-3 (pharyngeal edema, erythema, pruritus) AEs |
| Bar-Or et al. [50]    | 232 RRMS           | Phase II, double-blind, randomized, Pbo-controlled study (MIRROR) (48 weeks) | Stable EDSS in 79% of patients at week 12 and 24                                  | ↓ 65% new Gd+ lesions at week 12 (p < 0.001)                                   | ↑ IARs in OFT (52%) vs Pbo (15%) Equivalent infections with OFT and Pbo          |
| Hauser et al. [3]     | 946 (ASCLEPIOS I) and 936 (ASCLEPIOS II) RRMS | Phase III, double-blind, double-dummy, active-controlled studies (ASCLEPIOS I and ASCLEPIOS II) (30 months) | ↓ ARR (ASCLEPIOS I 0.11 OFT vs 0.22 TER, p < 0.001; ASCLEPIOS II 0.10 OFT vs 0.25 TER, p < 0.001) ↓ proportion of patients with 3- and 6-month CDP (10.9% and 8.1% OFT vs 15.0% and 12.0% TER) ↓ Gd+ lesions with OFT vs TER (97.5% ASCLEPIOS I, 93.8% ASCLEPIOS II) ↓ new/enlarging T2 lesions (82% ASCLEPIOS I, 84.5% ASCLEPIOS II) No differences in the annualized rate of brain atrophy OFT vs TER (− 0.28% vs − 0.35% in ASCLEPIOS I; − 0.29% vs − 0.35% ASCLEPIOS II) | Equivalent IARs with OFT (20.2%) vs TER (15%) Equivalent infections with OFT (51.6%) vs TER (52.7%) Equivalent % of neoplasm with OFT (0.5%) vs TER (0.4%) |

*AE* adverse events, *ARR* annualized relapse activity, *CDP* confirmed disability progression, *EDSS* expanded Disability Status Scale, *Gd+* gadolinium-enhancing, *IARs* infusion-associated reactions, *OFT* ofatumumab, *Pbo* placebo, *RRMS* relapsing–remitting multiple sclerosis, *TER* teriflunomide
reduced the risk of 12-week CDP by 34.4% and 32.5% at 3 and 6 months ($p = 0.002$ and $p = 0.012$), whereas percentages of patients with 24-week disability improvement were not significantly different between treatment groups ($p = 0.09$) (Table 3).

Ofatumumab promoted also a more effective suppression of MRI activity compared with teriflunomide, with a significant reduction of new/enlarging T2-hyperintense (ASCLEPIOS I = 82%; ASCLEPIOS II = 84.5%) and Gd-enhancing lesions (ASCLEPIOS I = 97.5%; ASCLEPIOS II = 93.8%). The annualized brain atrophy rate did not differ significantly between the ofatumumab- and teriflunomide groups ($-0.28\%$ vs $-0.35\%$ in ASCLEPIOS I; $-0.29\%$ vs $-0.35\%$ ASCLEPIOS II).

### Safety and tolerability of anti-CD20 therapies

#### Infusion-associated adverse events

Treatment infusion-associated reactions (IARs) are of mild-to-moderate severity. The most common symptoms are fever, headache, rash, nausea, throat irritation, hypotension, and itching. There are no studies comparing the safety profile of different anti-CD20 therapies; evaluating IARs across studies is challenging given different premedication regimens.

In a phase-II RCT, IARs were disclosed in 78.3% of RRMS patients treated with rituximab compared with 40.0% in the placebo group [1].

In the OPERA I and II studies, IARs were reported in 34% of the RRMS patients treated with ocrelizumab compared with 10% of those treated with INFβ-1a or placebo, whereas the prevalence was 40% with ocrelizumab versus 26% with placebo in PPMS from the ORATORIO trial [2]. In the OPERA II and ORATORIO OLE studies [42, 47], IARs incidence was consistent with past reports.

In the MIRROR study, a similar percentage of IARs was reported for ofatumumab (41–66% according to the regimen used vs 15% for placebo) [50]. In ASCLEPIOS I/II, IARs occurred in 20.2% in the ofatumumab group and 15.0% in the teriflunomide group [3].

#### Hypogammaglobulinemia

Although CD20 is not expressed on plasmablasts and plasma cells (Fig. 2), anti-CD20-depleting therapies have been shown to reduce Ig levels. Rituximab has some impact on serum IgM and IgG levels [52], and hypogammaglobulinemia can be found during long-time treatment [53]. In RCTs, ocrelizumab reduced the total serum Ig levels to some extent with greatest impact on IgM [2]. However, there was no association between low IgM levels and serious infections. In the MIRROR study, only two out of 232 (1%) RRMS patients treated with ofatumumab developed decreased IgG levels, leading to study termination [50]. In ASCLEPIOS I and II, no association was observed between a decrease in Ig levels and the incidence of serious/non-serious infections in ofatumumab-treated patients who experienced infections within 1 month prior and until 1 month after a reduction in Ig levels below the lower lymphocyte number [3].

### Infections and COVID-19 interaction

Before starting treatment, the screening for latent infections should be performed. Reactivation of tuberculosis, hepatitis, and human immunodeficiency virus has been reported in patients treated with anti-CD20 therapies [54].

Although the incidence of infections in the placebo (71.4%) and treated (69.7%) groups in the first phase-II RCT of rituximab in RRMS was similar [1], rituximab was associated with an increased incidence of urinary tract infections and sinusitis.

In RCTs with ocrelizumab, the percentage of patients reporting any infection was 59.9% (vs 54.3% in patients treated with INFβ-1a) in OPERA I, 60.2% (vs 52.5% in patients treated with IFNB-1a) in OPERA II and 71.4% in ORATORIO (vs 69.9% in the placebo group). A slightly increased incidence of upper respiratory tract infections was observed in patients treated with ocrelizumab compared with placebo or INF-β-1a [2]. No increased risk of serious infections was reported in any of these studies.

No cases of progressive multifocal leukoencephalopathy (PML) were reported in RCTs with ocrelizumab, whereas ten cases of PML (as of June 2021) have been described in post-marketing surveillance, of which eight were carry-over cases from prior DMTs [55].

According to data from the phase-II/III RCTs, ofatumumab was not associated with an increased risk of infection-related adverse events (AE) compared with placebo and teriflunomide [3, 50].

Recently, the COVID-19 pandemic raised some concerns about immunosuppression in MS, leading to treatment delay or cessation. Some publications have suggested that anti-CD20 therapies in a 6-month schedule (i.e., rituximab and ocrelizumab) may be associated with an increased risk of severe COVID-19 disease and need for hospitalization [56], not confirmed by a recent study [57]. MS patients appear to respond to SARS-CoV2 in a similar way to the general population and high disability or a progressive disease course represent the most relevant risk factors for a severe COVID-19 disease in MS. Moreover, innate immune response, and, probably, anti-viral CD8 T-cell responses play a major role in eliminating the SARS-CoV2 before significant antibody responses have developed, thus B cells do not appear to be an absolute requirement.
for recovery. Duration of exposure might play a role, as suggested by the North American Registry, which disclosed an increased risk of hospitalization in patients treated with rituximab, but not in those treated with ocrelizumab [58].

**Malignancies**

Immunosuppressive drugs could influence immunological tumor surveillance; therefore, long-term data are required to exclude an increased rate of malignancies. In MS patients, rituximab was not associated with an increased neoplastic risk over the long term compared with the general population [59].

In the OPERA I and OPERA II trials, four patients (0.7%) treated with ocrelizumab developed malignancies compared with two patients (0.2%) treated with INF-β-1a [2]. In the ORATORIO trial, 11 patients receiving ocrelizumab (2.3%) developed malignancies (4 breast cancer) compared with two patients (0.8%) in the placebo group [2]. The percentage of patients developing breast cancer in the ocrelizumab-treated group was similar to those expected from epidemiological studies and the incidence decreased during the extension studies [51].

In ASCLEPIOS II/II trials, no unexpected imbalance in the rates of malignancies was observed in ofatumumab-treated patients [3].

**Vaccination**

Patients should complete any required vaccinations at least 6 weeks prior to treatment initiation. Live attenuated or live vaccines are not recommended during treatment and until B-cell recovery, and at least 6 months after the last administration of rituximab [60].

A recent phase-IIIb RCT (VELOCE) [60] provided class-II evidence that peripherally B-cell-depleted ocrelizumab recipients mounted humoral responses to clinically relevant vaccines and the neo-antigen, KLH, although attenuated. For non-live/inactivated vaccines, such as seasonal influenza vaccines, it is recommended to vaccinate patients treated with ocrelizumab since a protective humoral response can be expected, even if attenuated [60]. An expert consensus recently suggested that patients planned to receive ocrelizumab should be vaccinated at least 6 weeks before the first administration, whereas in those already receiving ocrelizumab, vaccinations should be administered at least 3 months after the last infusion [61].

**Therapies currently under investigation and future perspectives**

Few studies have investigated ublituximab, a third-generation glycol-engineered chimeric anti-CD20 antibody promoting B-cell depletion through ADCC (Table 4). In a phase-II study of 48 RRMS patients [62], ublituximab promoted a complete B-cell depletion (> 99%) within 4 weeks. The ARR was 0.07, 93% of the patients were relapse-free at week 48, and no patients demonstrated CDP. T2-hyperintense LV decreased by 8% at week 24 ($p = 0.004$) and 10% at week 48 ($p = 0.016$), whereas the mean number of Gd-enhancing lesions was reduced from 3.8 at baseline to 0 at week 24 ($p = 0.003$) and maintained at week 48 ($p < 0.001$). The most common AEs were IARs that were all grade 1 or 2; no severe AEs were reported. No serious or opportunistic infections and no liver disease occurred. Ublituximab is currently being tested against teriflunomide in two fully enrolled phase-III studies (ULTIMATE I and II) in patients with RRMS (ClinicalTrials.gov identifier: NCT03277261 and NCT03277248).

Given the central role of B cells in MS pathogenesis, new therapeutic strategies directed against B-cell targets are currently under investigations (Table 4). CD19, a member of Ig superfamily, is involved in signal transduction following B-cell-receptor activation and represents one of the main regulator of B-cell activation and humoral immunity [63]. CD19 is more broadly expressed on cells of B lineage since, in contrast to CD20, it is also expressed on pro-B cells, plasmablasts and Ig-producing plasma cells (Fig. 2) [63]. Anti-CD19 therapies produce a more long-lasting B-cell depletion and a marked decline in Ig concentrations [63].

Among the different anti-CD19 monoclonal antibodies, inebilizumab (MEDI-551) is a glycoengineered, afucosylated anti-CD19 antibody having high affinity to FcγRIIA and causing B-cell depletion through ADCC [64]. A 24-week phase-I RCT (ClinicalTrials.gov study identifier: NCT01585766) investigated pharmacological properties and safety of inebilizumab in RRMS patients compared with placebo [65]. Inebilizumab promoted a complete B-cell depletion with all doses (2 i.v. doses, days 1 and 15: 30, 100 or 600 mg; or single s.c. dose on day 1: 60 or 300 mg), with IAR occurring in seven out of 21 (33.3%) RRMS patients and with upper respiratory tract and urinary tract infections, pyrexia and increased blood pressure as the most common AEs. At week 24, neither relapses nor median EDSS score changes occurred in RRMS patients receiving inebilizumab. Moreover, inebilizumab promoted trend in reductions in new/newly enlarging T2-hyperintense (0.4 vs 2.4) and Gd-enhancing lesions (0.1 vs 1.3), with a higher proportion of patients free from new inflammatory activity (75% vs 43%) [65].

Bruton’s tyrosine kinase (BTK) is a cytoplasmic kinase expressed on cells of the hematopoietic lineage, except for T cells, natural killer (NK) cells and plasma cells, and contributes to signal transductions from B-cell receptor (BCR) to the PI3K, MAPK and NF-κB pathways, thus regulating B-cell
| Target | Name and posology | References | Number of patients | Trial design | Efficacy | Safety |
|--------|-------------------|------------|-------------------|-------------|----------|--------|
| CD20   | Ublituximab (150 mg on day 1, 450 or 600 mg on day 15 and 24 weeks) | NCT02738775 [83] | 48 RRMS | Phase II (48 weeks), Pbo-controlled | ARR 0.07 | The most common AEs were IARs that were all grade 1 or 2, no SAE, no serious or opportunistic infections and no liver disease |
| CD19   | Inebilizumab (MEDI-551) (2 i.v. doses, days 1 and 15: 30, 100 or 600 mg; or single s.c. dose on day 1: 60 or 300 mg) | NCT01585766 [65] | 28 RRMS | Phase I (24 weeks) | No relapses and no median EDSS score changes at 24 weeks | IARs occurring in 7 out of 21 (33.3%) RRMS patients and with upper respiratory tract and urinary tract infections, pyrexia and increased blood pressure |
| BTKi   | Evobrutinib (25 mg daily, 75 mg daily or 75 mg twice daily) | NCT02975349 [69] | 228 RRMS, 33 SPMS | Phase II, double-blind, randomized, Pbo or DMF (24 weeks) | 75 mg of evobrutinib once daily: ↓Gd+ lesions vs Pbo at 12 and 24 weeks (1.69 vs 3.85, lesion rate ratio 0.30, p = 0.005) 75 mg of evobrutinib twice daily (1.15, lesion rate ratio 0.44, p = 0.06) No difference in the ARR, relapse-free status or CDP at any dose | Most common AEs: nasopharyngitis and asymptomatic ↑ of aminotransferase levels |
|        | Tolebrutinib (5 mg, 15 mg, 30 mg, 60 mg) | NCT03889639 [70] | 130 RRMS | Phase II, double-blind, Pbo (16 weeks) | ↓new/newly enlarging T2-hyperintense lesions vs Pbo at 12 weeks (2.12 with Pbo, 1.90 with 5 mg, 1.32 with 15 mg, 1.30 with 30 mg, 0.23 with 60 mg) ↓Gd+ lesions vs Pbo at 12 weeks (1.03 with Pbo, 1.39 with 5 mg, 0.77 with 15 mg, 0.76 with 30 mg, 0.13 with 60 mg) | – |

ARR: annualized relapse rate; CDP: clinical deterioration; EDSS: Expanded Disability Status Scale; Pbo: placebo; RRMS: relapsing-remitting multiple sclerosis; SPMS: secondary progressive multiple sclerosis; T2 LV: T2 lesion volume; T2-hyperintense: T2-hyperintense; Gd+: gadolinium; IAR: infusion-related adverse event; SAE: serious adverse event; DMF: dimethylfumarate.
survival, activation, proliferation, and differentiation to plasma cells [66, 67].

Several BTK inhibitors have been proposed for the treatment of hematological malignancies and dysimmune disorders [66, 68]. A recent 24-week phase-II RCT (ClinicalTrials.gov study identifier: NCT02975349) [69] compared oral evobrutinib at different doses (25 mg daily, 75 mg daily or 75 mg twice daily) with placebo or dimethyl fumarate in patients with RRMS or active SPMS. MS patients who received 75 mg of evobrutinib once daily had significantly fewer Gd-enhancing lesions compared with placebo at weeks 12 through 24 (1.69 vs 3.85, lesion rate ratio 0.30, \( p = 0.005 \)), whereas only a trend for patients who received 75 mg of evobrutinib twice daily was found (1.15, lesion rate ratio 0.44, \( p = 0.06 \)) [69]. No significant difference in the ARR, relapse-free status or disability progression at any dose occurred [69]. The most common AEs were nasopharyngitis and asymptomatic increase of aminotransferase levels.

In a 16-week phase-IIb study, 130 patients with RRMS were randomized to receive either placebo or tolebrutinib at different doses (5, 15, 30, and 60 mg) [70]. At week 12, patients treated with tolebrutinib had significantly lower mean number of new/enlarging T2-hyperintense lesions compared with placebo (1.90 with 5 mg, 1.32 with 15 mg, 1.30 with 30 mg, 0.23 with 60 mg vs 2.12 with placebo). Tolebrutinib also reduced the mean number of Gd-enhancing lesions (1.39 with 5 mg, 0.77 with 15 mg, 0.38 with 30 mg, and 0.08 with 60 mg vs 0.89).

Recently, in a subgroup analysis including 61 patients with highly active disease [71] (i.e., one relapse in the year prior to screening and one or more Gd-enhancing lesions on MRI within 6 months prior to screening or nine or more T2-hyperintense lesions at baseline, or two or more relapses in the year prior to screening), at week 12, tolebrutinib reduced the mean number of new/enlarging T2-hyperintense lesions compared with placebo (1.09 with 5 mg, 0.89 with 15 mg, 0.75 with 30 mg, and 0.15 with 60 mg vs 1.44 with placebo). Tolebrutinib also promoted a reduction in the mean number of new Gd-enhancing lesions compared with placebo (0.82 with 5 mg, 0.50 with 15 mg, 0.38 with 30 mg, and 0.08 with 60 mg vs 0.89).

Several other BTK inhibitors are under investigation (see Ref. [66] for a comprehensive review), possibly representing a new therapeutic opportunity for MS patients in the near future although several aspects regarding their mechanism of action, efficacy and safety still need to be fully investigated.
Conclusions

The introduction of anti-CD20 therapies in the MS scenario confirmed the central role of B cells in MS pathogenesis and established a new therapeutic approach for both relapsing and progressive MS patients.

In RRMS, currently approved anti-CD20 monoclonal antibodies (rituximab, ocrelizumab, and ofatumumab) consistently lead to a dramatic reduction of clinical relapses and MRI disease activity together with a significant limitation of disability worsening and brain atrophy progression.

Interestingly, ocrelizumab has been the first effective DMT in delaying disability progression in PPMS. However, efficacy and benefits in PPMS appear partial and more limited compared with those observed in RRMS. These findings suggest that anti-CD20 therapies exert a strong anti-inflammatory activity and that their beneficial neuroprotective effects could be mainly secondary to the prevention of further inflammatory disease activity. The more limited effect in PPMS could be also due to an inefficient depletion of B cells within a CNS-compartmentalized inflammation, possibly due to a limited permeability of anti-CD20 monoclonal antibodies across the blood–brain barrier. This aspect should be better investigated in future studies which should include reliable clinical, laboratory and MRI biomarkers to identify and monitor MS progression.

Data from RCTs, their OLE and from observational studies suggest that anti-CD20 therapies have a favorable short- and medium-term safety profile, with IARs, upper respiratory tract and urinary infections being the most common side effects, although the risk of malignancies should be carefully monitored over long-term.

Several aspects regarding treatment with anti-CD20 therapies deserve further investigations.

Future studies should define how to optimize anti-CD20 therapy administration regimens in terms of dosing, timing and duration of the treatment. Finally, post-marketing and observational studies should be conducted to shed light on beneficial effects and risks over longer term and to better explore the risk of infections and malignancies, especially in older MS patients.

Acknowledgements None.

Author contributions MM: drafting/revising the manuscript, study concept, acquisition and interpretation of the data; PP: drafting/revising the manuscript, study concept, acquisition and interpretation of the data; MF: drafting/revising the manuscript, study concept, acquisition and interpretation of the data; MAR: drafting/revising the manuscript, study concept, acquisition and interpretation of the data.

Funding None.

Conflicts of interest M. Margoni reports grants and personal fees from Sanofi Genzyme, Merck Serono, Novartis and Almirall. She was awarded a MAGNIMS-ECTRIMS fellowship in 2020. P. Preziosa received speaker honoraria from Biogen Idec, Novartis, Merck Serono and ExceMED. He is supported by a senior research fellowship FISM—Fondazione Italiana Sclerosi Multipla—cod. 2019/BS/009 and financed or co-financed with the ‘5 per mille’ public funding. M. Filippi is Editor-in-Chief of the Journal of Neurology and Associate Editor of Radiology, Human Brain Mapping and Neurological Sciences, received compensation for consulting services and/or speaking activities from Almiral, Alexion, Bayer, Biogen, Celgene, Eli Lilly, Genzyme, Merck-Serono, Novartis, Roche, Sanofi, Takeda, and Teva Pharmaceutical Industries, and receives research support from Biogen Idec, Merck-Serono, Novartis, Roche, Teva Pharmaceutical Industries, Italian Ministry of Health, Fondazione Italiana Sclerosi Multipla, and ARSLA (Fondazione Italiana di Ricerca per la SLA). M.A. Rocca received speaker honoraria from Bayer, Biogen, Bristol Myers Squibb, Celgene, Genzyme, Merck Serono, Novartis, Roche, and Teva, and receives research support from the MS Society of Canada and Fondazione Italiana Sclerosi Multipla.

References

1. Hauser SL, Waubant E, Arnold DL, Vollmer T, Antel J, Fox RJ, Bar-Or A, Panzara M, Sarkar N, Agarwal S, Langer-Gould A, Smith CH, HT Group (2008) B-cell depletion with rituximab in relapsing-remitting multiple sclerosis. N Engl J Med 358(7):676–688. https://doi.org/10.1056/NEJMoa0706383
2. Montalban X, Hauser SL, Kappos L, Arnold DL, Bar-Or A, Comi G, de Seze J, Giovannoni G, Hartung HP, Hemmer B, Lublin F, Rammohan KW, Selmañ K, Traboulsi A, Sauter A, Masterman D, Fontoura P, Belachew S, Garren H, Mairon N, Chin P, Wolinsky JS, OC Investigators (2017) Ocrelizumab versus placebo in primary progressive multiple sclerosis. N Engl J Med 376(3):209–220. https://doi.org/10.1056/NEJMoa1606468
3. Hauser SL, Bar-Or A, Cohen JA, Comi G, Correale J, Coyle PK, Cross AH, de Seze J, Leppert D, Montalban X, Selmañ K, Wiendl H, Kerloguen C, Willi R, Li B, Kakarieka A, Tomic D, Goodyear A, Pingili R, Haring DA, Ramanathan K, Merschhemke M, Kappos L, Asclepios I, AIT Investigators (2020) Ofatumumab versus teriflunomide in multiple sclerosis. N Engl J Med 383(6):545–557. https://doi.org/10.1056/NEJMoa1917246
4. Baecker-Allan C, Kaskow BJ, Weiner HL (2018) Multiple sclerosis: mechanisms and immunotherapy. Neuro 97(4):742–768. https://doi.org/10.1016/j.neuron.2018.01.021
5. Dendrou CA, Fugger L, Friese MA (2015) Immunopathology of multiple sclerosis. Nat Rev Immunol 15(9):545–555. https://doi.org/10.1038/nri3871
6. Li R, Patterson KR, Bar-Or A (2018) Reassessing B cell contributions in multiple sclerosis. Nat Immunol 19(7):696–707. https://doi.org/10.1038/s41590-018-0135-x
7. Hauser SL, Bar-Or A, Comi G, Giovannoni G, Hartung HP, Hemmer B, Lublin F, Montalban X, Rammohan KW, Selmañ K, Traboulsi A, Wolinsky JS, Arnold DL, Klingelschmitt G, Masterman D, Fontoura P, Belachew S, Chinn P, Mairon N, Garren H, Kappos L, Opera I, OIC Investigators (2017) Ocrelizumab versus interferon beta-1a in relapsing multiple sclerosis. N Engl J Med 376(3):221–234. https://doi.org/10.1056/NEJMoa1601277
8. Storch MK, Piddlesden S, Halfia M, Ivainnainen M, Morgan P, Lassmann H (1998) Multiple sclerosis: in situ evidence for
antibody- and complement-mediated demyelination. Ann Neurol 43(4):465–471. https://doi.org/10.1002/ana.410430409
9. Genain CP, Cannella B, Hauser SL, Raine CS (1999) Identification of autoantibodies associated with myelin damage in multiple sclerosis. Nat Med 5(2):170–175. https://doi.org/10.1038/5532
10. Lucchinetti C, Bruck W, Parisi J, Scheithauer B, Rodriguez M, Lassmann H (2000) Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination. Ann Neurol 47(6):707–717. https://doi.org/10.1002/1531-8249(200006)47:6<3c:aid-ana3>3.0.co;2-q
11. Obermeier B, Mentele R, Malotka J, Kellermann J, Kumpfel T, Wekerle H, Lottspeich F, Hohlfeld R, Dormann K (2008) Matching of oligoclonal immunoglobulin transcripts and proteomes of cerebrospinal fluid in multiple sclerosis. Nat Med 14(6):688–693. https://doi.org/10.1038/nm1714
12. Cepok S, Rosche B, Grummel V, Vogel F, Zhou D, Sayn J, Sommer N, Hartung HP, Hemmer B (2005) Short-lived plasma blasts are the main B cell effector subset during the course of multiple sclerosis. Brain 128(Pt 7):1667–1676. https://doi.org/10.1093/brain/awh486
13. Bar-Or A (2008) The immunology of multiple sclerosis. Semin Neurol 28(1):29–45. https://doi.org/10.1055/s-2007-1019124
14. Kuhle J, Pohl C, Mehling M, Edan G, Freedman MS, Hartung HP, Polman CH, Miller DH, Montalban X, Barkhof F, Bauer L, Dahms S, Lindberg R, Kappos L, Sandbrink R (2007) Lack of association between antimyelin antibodies and progression to multiple sclerosis. N Engl J Med 356(4):371–378. https://doi.org/10.1038/sj.nm.4002519
15. Li R, Rezk A, Miyazaki Y, Hilgenberg E, Touil H, Shen P, Moore JL, Korn T, Hemmer B (2012) Potassium channel KIR4.1 as an antigenic target in multiple sclerosis. Brain 136(Pt 3):795–806. https://doi.org/10.1093/brain/awr377
16. Srivastava R, Aslam M, Kalluri SR, Schirmer L, Buck D, Tackenberg B, Rothhammer V, Chan A, Gold R, Berthele A, Bennett JL, Korn T, Hemmer B (2012) Potassium channel KIR4.1 as an immune target in multiple sclerosis. N Engl J Med 367(2):115–123. https://doi.org/10.1056/NEJMoa1110740
17. Rodrigues-Pinto D (2005) B cells as antigen presenting cells. Cell Immunol 238(2):67–75. https://doi.org/10.1016/j.cellimm.2006.02.005
18. Bar-Or A, Fazaw L, Fan B, Darlington PJ, Rieger A, Ghorayeb C, Calabresi PA, Waubant E, Hauser SL, Zhang J, Smith CH (2010) Abnormal B-cell cytokine responses a trigger of T-cell-mediated disease in MS? Ann Neurol 67(4):452–461. https://doi.org/10.1002/ana.21939
19. Li R, Rezk A, Miyazaki Y, Hilgenberg E, Touil H, Shen P, Moore CS, Michel L, Althekair F, Rajasekharan S, Gommerman JL, Prat A, Fillatreau S, Bar-Or A, Canadian BCTiMS (2015) Proinflammatory GM-CSF-producing B cells in multiple sclerosis and B cell depletion therapy. Sci Transl Med 7(310):310ra166. https://doi.org/10.1126/scitranslmed.aab4176
20. Duddy M, Niño M, Adatia F, Hebert S, Freedman M, Atkins H, Kim HJ, Bar-Or A (2007) Distinct effector cytokine profiles of memory and naive human B cell subsets and implication in multiple sclerosis. J Immunol 178(10):6092–6099. https://doi.org/10.4049/jimmunol.178.10.6092
21. Barr TA, Shen P, Brown S, Lampropoulou V, Roch T, Lawrie S, Fan B, O’Connor RA, Anderton SM, Bar-Or A, Fillatreau S, Gray D (2012) B cell depletion therapy ameliorates autoimmune disease through ablation of IL-6–producing B cells. J Exp Med 209(5):1001–1010. https://doi.org/10.1084/jem.20111675
22. Li R, Rezk A, Healy LM, Muirhead G, Prat A, Gommerman JL, Bar-Or A, MCBiM Team (2015) Cytokine-defined B cell responses as therapeutic targets in multiple sclerosis. Front Immunol 6:626. https://doi.org/10.3389/fimmu.2015.00626
23. Frischer JM, Bramow S, Dal-Bianco A, Lucchinetti CF, Rauschka H, Schmidbauer M, Laursen H, Sorensen PS, Lassmann H (2009) The relation between inflammation and neurodegeneration in multiple sclerosis brains. Brain 132(5):1175–1189. https://doi.org/10.1093/brain/awp070
24. Machado-Santos J, Saji E, Troscher AR, Paunovic M, Liblau R, Gabriely G, Bien CG, Bauer J, Lassmann H (2018) The compartmentalized inflammatory response in the multiple sclerosis brain is composed of tissue-resident CD8+ T lymphocytes and B cells. Brain 141(7):2066–2082. https://doi.org/10.1093/brain/awy151
25. Magliozzi R, Howell OW, Serafini B, Nicholas R, Paunovic M, Reynolds A, Aloisi F (2007) Meningeal B-cell follicles in secondary progressive multiple sclerosis associate with early onset of disease and severe cortical pathology. Brain 130(4):1089–1104. https://doi.org/10.1093/brain/awm038
26. Magliozzi R, Howell OW, Nicholas R, Cruciani C, Castellaro M, Romualdi C, Rossi S, Pitteri M, Benedetti MD, Gajofatto A, Pizzini FB, Montemezzani S, Rasia C, Capra R, Bertoldo A, Facchiano F, Monaco S, Reynolds A, Calabrese M (2018) Inflammatory intrathecal profiles and cortical damage in multiple sclerosis. Ann Neurol 83(4):739–755. https://doi.org/10.1002/ana.25197
27. Magliozzi R, Serafini B, Rosicarelli B, Chiappetta G, Veroni C, Reynolds A, Aloisi F (2013) B-cell enrichment and Epstein-Barr virus infection in inflammatory cortical lesions in secondary progressive multiple sclerosis. J Neuropathol Exp Neurol 72(1):29–41. https://doi.org/10.1097/NEN.0b013e31827bcf62
28. Meini E, Krumholz M, Hohlfeld R (2006) B lineage cells in the inflammatory central nervous system environment: migration, maintenance, local antibody production, and therapeutic modulation. Ann Neurol 59(6):880–892. https://doi.org/10.1002/ana.20890
29. Touil H, Kobernt A, Lebeurrier N, Rieger A, Saikali P, Lambert C, Fazaw L, Moore CS, Prat A, Gommerman J, Antel JP, Iyoama Y, Nakashima I, Bar-Or A, Canadian BCTiMS (2018) Human central nervous system astrocytes support survival and activation of B cells: implications for MS pathogenesis. J Neuroinflammation 15(1):114. https://doi.org/10.1186/s12974-018-1136-2
30. Mahad DH, Trapp BD, Lassmann H (2015) Pathological mechanisms in progressive multiple sclerosis. Lancet Neurol 14(2):183–193. https://doi.org/10.1016/S1474-4422(14)70256-X
31. Magliozzi R, Howell OW, Reeves C, Roncaroli F, Nicholas R, Serafini B, Aloisi F, Reynolds A (2010) A gradient of neuronal loss and meningeal inflammation in multiple sclerosis. Ann Neurol 68(4):477–493. https://doi.org/10.1002/ana.22230
32. Lucchinetti CF, Popescu BF, Bunyan RF, Moll NM, Roemer SF, Lassmann H, Bruck W, Parisi JE, Scheithauer BW, Giannini C, Weigand SD, Mandrekar J, Ransohoff RM (2011) Inflammatory cortical demyelination in early multiple sclerosis. N Engl J Med 365(23):2188–2197. https://doi.org/10.1056/NEJMoa1100648
33. Meyer S, Evers M, Jansen JHM, Buijs J, Broek B, Reitsma SE, Moerer P, Amini M, Kretschmer A, Ten Broeke T, den Hartog MT, Rijke M, Klein C, Valerius T, Boross P, Leusen JHW (2018) New insights in type I and II CD20 antibody mechanisms-of-action with a panel of novel CD20 antibodies. Br J Haematol 180(6):808–820. https://doi.org/10.1111/bjh.15132
34. Montalvao F, Garcia Z, Celli S, Breart B, Deguine J, Van Rooijen N, Bousso P (2013) The mechanism of anti-CD20-mediated B cell depletion revealed by intravital imaging. J Clin Invest 123(12):5098–5103. https://doi.org/10.1172/JCI70972
35. Cenciioni MT, Mattosco M, Magliozzi R, Bar-Or A, Muraro PA (2021) B cells in multiple sclerosis—from targeted depletion to immune reconstitution therapies. Nat Rev Neurol. https://doi.org/10.1038/s41582-021-00498-5
36. Yang CS, Yang L, Li T, Zhang DQ, Jin WN, Li MS, Su N, Zhang N, Liu Q, Shao ZH, Yu C, Shi FD (2013) Responsiveness to reduced dosage of rituximab in Chinese patients with neuromyelitis optica. Neurology 81(8):710–713. https://doi.org/10.1212/WNL.0b013e3182a1ac7

37. Hawker K, O’Connor P, Freedman MS, Calabresi PA, Antel J, Simon J, Hauser S, Waubant E, Vollmer T, Panitch H, Zhang J, Chin P, Smith CH, OT Group (2009) Rituximab in patients with primary progressive multiple sclerosis: results of a randomized double-blind placebo-controlled multicenter trial. Ann Neurol 66(4):460–471. https://doi.org/10.1002/ana.21867

38. Ingle GT, Sastre-Garriga J, Miller DH, Thompson AJ (2005) Is inflammation important in early PPMS? A longitudinal MRI study. J Neurol Neurosurg Psychiatry 76(9):1255–1258. https://doi.org/10.1136/jnnp.2004.036590

39. Bhargava P, Wicken C, Smith MD, Strowd RE, Cortese I, Reich DS, Calabresi PA, Mowry EM (2019) Trial of intrathecal rituximab in progressive multiple sclerosis patients with evidence of leptomeningeal contrast enhancement. Mult Scler Relat Disord 30:136–140. https://doi.org/10.1016/j.msard.2019.02.013

40. Kappos L, Li D, Calabresi PA, O’Connor P, Bar-Or A, Barkhof F, Yin M, Leppert D, Glanzman R, Tinbergen J, Hauser SL (2011) Ocrelizumab in relapsing-remitting multiple sclerosis: a phase 2, randomised, placebo-controlled, multicentre trial. Lancet 378(9785):1779–1787. https://doi.org/10.1016/S0140-6736(11)61649-8

41. Baker D, Pryce G, James LK, Marta M, Schmierer K (2020) The ocrelizumab phase II extension trial suggests the potential to improve the risk: benefit balance in multiple sclerosis. Mult Scler Relat Disord 44:102279. https://doi.org/10.1016/j.msard.2020.102279

42. Hauser SL, Kappos L, Arnold DL, Bar-Or A, Brochet B, Naimith RT, Traboulsee A, Belachew S, Kobenqen H, Levesque V, Manfrini M, Model F, Hubeaux S, Mehta L, Montalban X (2020) Five years of ocrelizumab in relapsing-remitting multiple sclerosis: the MIRROR Study. Neurology 90(20):e1805–e1814. https://doi.org/10.1212/WNL.000000000005516

43. Ineichen BV, Moridi T, Granberg T, Piehl F (2020) Rituximab treatment for multiple sclerosis. Mult Scler 26(2):137–152. https://doi.org/10.1177/1352458519858604

44. Mok CC (2013) Rituximab for the treatment of rheumatoid arthritis: an update. Drug Des Dev Ther. 8:87–100. https://doi.org/10.2147/DDDT.S41645

45. Patel A, Sul J, Gordon ML, Steinklein J, Sanguinetti S, Pramanik B, Purohit D, Haroutunian V, Williamson A, Koralnik I, Harel A (2021) Progressive multifocal leukoencephalopathy in a patient with progressive multiple sclerosis treated with ocrelizumab monotherapy. JAMA Neurol. https://doi.org/10.1001/jamaneurol.2021.0627

46. Sormani MP, De Rossi N, Carmisciano L, Cordioli C, Moiola L, Radaelli M, Immovilli P, Capobianco M, Trojano M, Zaratin P, Tedeschi G, Comi G, Battaglia MA, Patti F, Salvetti M, Musc-19 Study Group (2021) Disease-modifying therapies. CNS Drugs 35(3):317–330. https://doi.org/10.1007/s40263-021-00804-1

47. Wolinsky JS, Arnold DL, Brochet B, Hartung HP, Montalban X, Naismith RT, Manfrini M, Overell J, Koendgen H, Sauter A, Bennett I, Hubeaux S, Kappos L, Hauser SL (2020) Long-term follow-up from the ORATORIO trial of ocrelizumab for primary progressive multiple sclerosis: a post-hoc analysis from the ongoing open-label extension of the randomised, placebo-controlled, phase 3 trial. Lancet Neurol 19(12):998–1009. https://doi.org/10.1016/S1474-4422(20)30342-2

48. Mancinelli CR, Scarppaza C, Cordioli C, De Rossi N, Rasia S, Turrini MV, Capra R (2021) Switching to ocrelizumab in RRMS patients at risk of PML previously treated with extended interval dosing of natalizumab. Mult Scler 27(5):790–794. https://doi.org/10.1177/1352458520946017

49. Sorensen PS, Lisby S, Grove R, Derosier F, Shackelford S, Havrdova E, Druolvic J, Filippi M (2014) Safety and efficacy of ofatumumab in relapsing-remitting multiple sclerosis: a phase 2 study. Neurology 82(7):573–581. https://doi.org/10.1212/WNL.00000000000125

50. Bar-Or A, Grove RA, Austin DJ, Tolson JM, VanMeter SA, Lewis EW, Derosier FJ, Lopez MC, Kavanagh ST, Miller AE, Sorensen PS (2018) Subcutaneous ofatumumab in patients with relapsing-remitting multiple sclerosis: the MIRROR Study. Neurology 90(20):e1805–e1814. https://doi.org/10.1212/WNL.000000000005516

51. Gelfand JM, Cree BAC, Hauser SL (2017) Ocrelizumab and other CD20(+)-B-cell-depleting therapies in multiple sclerosis. Neurotherapeutics 14(4):835–841. https://doi.org/10.1007/s13240-017-0557-4

52. Salzer J, Svenningsson R, Alping P, Novakova L, Bjorck A, Fink K, Islam-Jakobsson P, Malmeestrom C, Axelsson M, Vagberg M, Sundstrom P, Lycke J, Piehl F, Svenningsson A (2016) Rituximab in multiple sclerosis: a retrospective observational study on safety and efficacy. Neurology 87(20):2074–2081. https://doi.org/10.1212/WNL.0000000000033331

53. Bredouhr J, Barbay E, Pannek J, Tozzini F, Turrini MV, Capra R (2021) Switching to ocrelizumab in RRMS patients at risk of PML previously treated with extended interval dosing of natalizumab. Mult Scler 27(5):790–794. https://doi.org/10.1177/1352458520946017

54. Costello K, Bebo B, Rammohan K, Cutter GR, Cross AH (2021) Improving risk: benefit balance in multiple sclerosis. Mult Scler Relat Disord 66(4):460–471. https://doi.org/10.1016/j.msard.2021.0627

55. Reuler AT, Centonze D, Naylor ML, Nagpal A, Rajbhandari R, Altincatal A, Kim M, Berduse A, Radhakrishnan M, Jung E, Sandrock AW, Smirnakis K, Polpescu C, de Moor C (2021) COVID-19 in patients with multiple sclerosis: associations with disease-modifying therapies. JAMA Neurol. https://doi.org/10.1001/jamaneurol.2021.0627

56. Sormani MP, De Rossi N, Schiavetti I, Carmisciano L, Cordioli C, Moiola L, Radaelli M, Immobellino P, Cappianchio M, Trojano M, Zaratin P, Tedeschi G, Comi G, Battaglia MA, Patti F, Salavetti M, Musc-19 Study Group (2021) Disease-modifying therapies. CNS Drugs 35(3):317–330. https://doi.org/10.1007/s40263-021-00804-1

57. Roder AT, Centonze D, Naylor ML, Nagpal A, Rajbhandari R, Altincatal A, Kim M, Berduse A, Radhakrishnan M, Jung E, Sandrock AW, Smirnakis K, Polpescu C, de Moor C (2021) COVID-19 in patients with multiple sclerosis: associations with disease-modifying therapies. JAMA Neurol. https://doi.org/10.1001/jamaneurol.2021.0627

58. Alping P, Asking J, Burman J, Fink K, Fogdell-Hahn A, Gunnarsen M, Hillert J, Langer-Gould A, Lycke J, Nilsson P, Salzer J, Svenningsson A, Vrethem M, Olsson T, Piehl F, Frisell T (2020)
Cancer risk for fingolimod, natalizumab, and rituximab in multiple sclerosis patients. Ann Neurol 87(5):688–699. https://doi.org/10.1002/ana.25701

60. Bar-Or A, Calkwood JC, Chognut C, Evershed J, Fox EJ, Herman A, Manfrini M, McNamara J, Robertson DS, Stokmaier D, Wendt JK, Winthrop KL, Traboulsee A (2020) Effect of ocrelizumab on vaccine responses in patients with multiple sclerosis: the VELOJO Study. Neurology 95(14):e1999–e2008. https://doi.org/10.1212/WNL.0000000000010380

61. Centonze D, Rocca MA, Gasperini C, Kappos L, Hartung HP, Magiari M, Oreja-Guevara C, Trojano M, Wiendl H, Filippi M (2021) Disease-modifying therapies and SARS-CoV-2 vaccination in multiple sclerosis: an expert consensus. J Neurol. https://doi.org/10.1007/s00415-021-10545-2

62. Fox E, Lovett-Racke AE, Gormley M, Liu Y, Petracca M, Cocozza S, Shubin R, Wray S, Weiss MS, Bosco JA, Power SA, Mok K, Ingele M (2020) A phase 2 multicenter study of ublituximab, a novel glycoengineered anti-CD20 monoclonal antibody, in patients with relapsing forms of multiple sclerosis. Mult Scler. https://doi.org/10.1177/1352458520918375

63. Tedder TF (2009) CD19: a promising B cell target for rheumatoid arthritis. Nat Rev Rheumatol 5(10):572–577. https://doi.org/10.1038/nrrheum.2009.184

64. Chen D, Gallagher S, Monson NL, Herbst R, Wang Y (2016) Inebilizumab, a B cell-depleting anti-CD19 antibody for the treatment of autoimmune neurological diseases: insights from preclinical studies. J Clin Med. https://doi.org/10.3390/jcm5120107

65. Agius MA, Klodowska-Duda G, Maciejowski M, Potemkowski A, Li J, Patra K, Wesley J, Madani S, Barron G, Katz E, Flor A (2019) Safety and tolerability of inebilizumab (MEDI-551), an anti-CD19 monoclonal antibody, in patients with relapsing forms of multiple sclerosis: results from a phase 1 randomised, placebo-controlled, escalating intravenous and subcutaneous dose study. Mult Scler 25(2):235–245. https://doi.org/10.1177/1352458517740647

66. Contentti EC, Correale J (2020) Bruton’s tyrosine kinase inhibitors: a promising emerging treatment option for multiple sclerosis. Expert Opin Emerg Drugs 25(4):377–381. https://doi.org/10.1080/14728214.2020.1822817

67. Torke S, Weber MS (2020) Inhibition of Bruton’s tyrosine kinase as a novel therapeutic approach in multiple sclerosis. Expert Opin Investig Drugs 29(10):1143–1150. https://doi.org/10.1080/13543784.2020.1807934

68. Liang C, Tian D, Ren X, Ding S, Jia M, Xin M, Thareja S (2018) Safety and efficacy of rituximab in relapsing-remitting multiple sclerosis: a 72-week, open-label, phase I trial. Ann Neurol 63(3):395–400. https://doi.org/10.1002/ana.21363

69. Naismith RT, Piccio L, Lyons JA, Lauber J, Tutlam NT, Parks BJ, Trinakos K, Song SK, Cross AH (2010) Rituximab add-on therapy for breakthrough relapsing multiple sclerosis: a 52-week phase II trial. Neurology 74(23):1860–1867. https://doi.org/10.1212/WNL.0b013e3181e24373

70. Alping P, Frisell T, Novakova L, Islam-Jakobsson P, Salzer J, Bjorck A, Axelsson M, Malmestrom C, Fink K, Lycke J, Svenningsson A, Piehl F (2016) Rituximab versus fingolimod after natalizumab in multiple sclerosis patients. Ann Neurol 79(6):950–958. https://doi.org/10.1002/ana.24651

71. Bar-Or A, Calabresi PA, Arnold D, Markowitz C, Shafer S, Kasper LH, Waubant E, Gazda S, Fox RJ, Panzara M, Sarkar N, Agarwal S, Smith CH (2008) Rituximab in relapsing-remitting multiple sclerosis: a 72-week, open-label, phase I trial. Ann Neurol 63(3):395–400. https://doi.org/10.1002/ana.21363

72. Fox E, Lovett-Racke AE, Gormley M, Liu Y, Petracca M, Cocozza S, Shubin R, Wray S, Weiss MS, Bosco JA, Power SA, Mok K, Ingele M (2021) A phase 2 multicenter study of ublituximab, a novel glycoengineered anti-CD20 monoclonal antibody, in patients with relapsing forms of multiple sclerosis. Mult Scler 27(3):420–429. https://doi.org/10.1177/1352458520918375