Two new species of hydnoid-fungi from India

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Abstract: Two taxa, Hericium yumthangense (Russulales, Agaricomycotina) and Mycoleptodonoides sharmae (Polyporales, Agaricomycotina) are described as new to science from the Shingba Rhododendron sanctuary located in the northern district of Sikkim, India. Macro- and micromorphological characters are described and illustrated for both species, which are compared with allied taxa. ITS rDNA sequences supported H. yumthangense as a rather isolated species within Hericium, the species complexes of which were not resolved due to low interspecific sequence divergence. In the case of M. sharmae, 28S rDNA (D1/D2) data rendered this poorly known genus among well-known taxa of the core-polyporoid clade.

Key words: Abies, Hericiaceae, Himalaya, Meruliciaceae, Rhododendron, Sikkim, taxonomy

INTRODUCTION

Covering an area of 43 sq. km, the Shingba Rhododendron sanctuary is located in the Yumthang valley of the northern district of Sikkim, India, in eastern Himalaya, surrounded by China, Bhutan, and Nepal. It is characterized by temperate silver fir and Rhododendron forest and is a native habitat for Rhododendron nivum. Apart from the trees like Abies densa, Larix griffithiana, Picea spinulosa, Magnolia globosa, M. campbellii, Acer pectinatum, Betula utilis, and more than 40 species of Rhododendron, this natural area is also rich in rare herbaceous flowering plants from various genera, including Primula, Potentilla, Ranunculus, Euphorbia, Roscoea, Heracleum, Rubus, and Aconitum.

During a macrofungal survey of the Shingba Rhododendron sanctuary and adjacent areas by K.D. between August and September 2011, various saprotrophic wood-decaying and mycorrhizal macromycetes were collected. Macro- and micromorphological studies revealed some of them were new taxa including two hydnoid fungi. These are described here as Hericium yumthangense sp. nov. (Hericaceae) and Mycoleptodonoides sharmae sp. nov. (Meruliciaceae). Macro-, micro- and ultrastructural illustrations, together with detailed descriptions and standard reference rDNA (ITS, partial 28S), and mtSSU DNA barcodes are provided.

MATERIALS AND METHODS

Morphological studies

Macromorphological characters were observed and recorded from fresh basidiomata in the field. Colour codes and terms follow the chart in the Flora of British Fungi (Anon.1969; “a” in the descriptions), except that those of spore prints follows Kränzlin (2005; “b” in the descriptions). Field photographs were taken with a Nikon D300s camera.

Micromorphological features were studied from dried samples mounted in a mixture of 5% KOH, 1% phloxin, Congo red and 30% glycerol, and Melzer’s reagent. Drawings were made with a drawing tube (attached to an Olympus CX41 microscope) at 1000×. Basidium length excludes sterigmata-length, and spore-dimensions exclude the dimension of the ornamentations (if any). Measurements are based on 20 examples. Basidiospores are measured in side view and given as KDa-KDc-KDb × KDx-KDz in which KDa is the minimum value for the length, KDb the maximum value for the length, KDb the mean value for the width, KDx the maximum value for the width, KDz the mean value for the width. The quotient of spore indicates the length-width ratio (Q = L/W) and is shown as Qa-Qb-Qc, where Qa refers the minimum quotient value, Qb the maximum quotient value amongst the measured collections, and Qc the mean quotient value.

Scanning electron micrographs of basidiospores were obtained from dry spore prints directly mounted on double-sided adhesive tape, pasted on a metallic specimen-stub, gold coated, and subsequently scanned in high vacuum mode at different magnifications to observe spore-ornamentations. The electron microscopy was carried out with a FEI’s Quanta 200 model scanning electron microscope (SEM) at the S.N. Bose National Centre for Basic Sciences, Kolkata (India).

DNA isolation, PCR, and sequencing

Total genomic DNA was extracted from herbarium specimens using the Jetquick general DNA clean up kit (Genomed), following the given protocols. The PCR for amplification of the
Phylogenetic inference

A search for respective *Hericium* ITS sequences yielded >100 sequences, which were downloaded from the international nucleotide sequence database collaboration (INSDC) database and examined regarding sufficient length, possible alignment ambiguities, and the resulting phylogenies. Since almost no sequence data is yet available for the genus *Mycoleptodonoides* (INSDC contained a single ITS sequence at the time of analysis), and its generic affiliation was unclear, partial 28S (D1/D2) sequences with high similarity to the query were retrieved via a BLAST search, manually filtered in respect to the quality criteria described above, and the dataset was further supplemented with sequences chosen according to Binder et al. (2005). Sequences for both datasets were subsequently aligned, using the EMBL-EBI MAFFT server (http://www.ebi.ac.uk/Tools/msa/mafft/) in standard, default setup. As previously described (Guevara-Guerrero et al. 2011), phylogenies inferred under the maximum-likelihood (ML) criterion (Felsenstein 1981) were conducted with RAxML v. 7.2.7, with search for the best tree under the GTR-MIX approach (Stamatakis et al. 2008). Maximum-likelihood trees for the *Hericium* dataset were rooted with *Laxitextum bicolor*, chosen as outgroup according to Larsson et al. (2004). The phlebioid outgroup (*Phlebia* spp.) for the *Mycoleptodonoides* dataset and clade names were depicted according to Binder et al. (2004). The ITS1-5.8S-ITS2 nuclear ribosomal DNA operon using ITS5/ITS4 was performed under standard conditions (White et al. 1990, Stielow et al. 2010). PCR conditions for amplifying the partial mtSSU and 28S rDNA using ITS5/ITS4 was performed under standard conditions (White et al. 1990, Stielow et al. 2010). PCR conditions for amplifying the partial mtSSU and 28S rDNA using ITS5/ITS4 was performed under standard conditions (White et al. 1990, Stielow et al. 2010). PCR conditions for amplifying the partial mtSSU and 28S rDNA using ITS5/ITS4 was performed under standard conditions (White et al. 1990, Stielow et al. 2010). PCR conditions for amplifying the partial mtSSU and 28S rDNA using ITS5/ITS4 was performed under standard conditions (White et al. 1990, Stielow et al. 2010).
et al. (2005). For depicting the trees, clades comprising at least twenty sequences were collapsed if they were either taxonomically homogeneous or contained only environmental samples. Sequence alignments are included in the online supplementary material.

RESULTS

Phylogenetic inference

The final ITS rDNA alignment of the Hericium dataset contained 100 sequences and 633 positions, for which 276 were considered ambiguously aligned and therefore excluded from the primary alignment. The optimal final ML tree had a log-likelihood of -2687.19 and is shown in Fig. 1. The 28S (D1/D2) rDNA alignment of the Mycoleptodonoides dataset contained 53 sequences and had a total length of 971 positions. The final ML tree is given in Fig. 2; optimal log-likelihood was -4094.56; both alignments are available as supplementary material.

The monophyletic Hericium ingroup was separated into two subtreess. The basal branch comprising the core-subtree with H. erinaceus, H. americanum, H. alpestre, and H. abietis, received strong bootstrap branch support (BS) (86%), while the H. coralloides subtree was not supported by BS. Internal branches for the two “European” and “Asian/ North American” H. coralloides clades were moderately (79%) to strongly supported (91%) respectively, however C. cirrhatus (EU78426) was resolved as a rather isolated taxon close to H. coralloides s.l. in our analysis. Several subclades (like e.g. H. alpestre, H. erinaceus, H. abietis) were not fully resolved and GenBank records inconsistently annotated. The specimen KD-11-146, representing the novel Hericium species, was neither close (with respect to low interspecific sequence variation) to the previously described Indian species (Das et al. 2011), H. bharengense, nor to one of the other subclades, and formed an isolated branch of its own.

For the Mycoleptodonoides dataset, the basal branch separating Antrodia clade 1 from Antrodia clade 2 and the core polyporoid clade received very strong support (100%). Besides the polyphyly of Antrodia among Fomitopsis species in clade 1 (82% BS), the branch inducing the split to clade 2 and the core polyporoid clade was not supported. Deeper internal branches within the

Fig. 2. Phylogenetic tree inferred under the maximum-likelihood (ML) criterion from the LSU (28S) rDNA alignment of the Mycoleptodonoides dataset. Numbers on the branches represent support values from 1000 bootstrap replicates if at least 70%, with branches scaled in terms of the expected number of substitutions per sites. Terminal names are from their original GenBank annotations.
three core clades received moderate to strong BS (70–100%). However, sequence annotations, therefore the original specimen identification, were moderately conflicting based on the publicly available 28S data. The hydnoid genus *Mycoleptodonoides* was nested, with respect to available sequences in the INSDC database, within the core polyphorid clade (Binder et al. 2005) close to the genera *Diplomitoporus* and *Obba*. The basal branch separating the subtrees, comprising *Diplomitoporus*, *Obba*, *Gelatoporia* and *Mycoleptodonoides* from several core polyphorid taxa (i.e. *Polyporus*, *Ganoderma*), received moderate support (78%), yet the isolation to *Antrodia* 2 was not resolved in our analysis (no BS).

Additional DNA barcodes, the ITS, partial LSU, and partial mtSSU sequences (if not used as described above) are deposited at the INSDC database and provide high quality sequence identifiers for the respective taxa. Sequences of the amplified loci were used in respect to availability of sufficient reference data only, i.e. the ITS for *Hericium*, and partial LSU data for positioning *Mycoleptodonoides* among the resupinate taxa, since its generic affiliation was unknown prior to sequencing. The insufficient quantity of mtSSU barcodes, currently available at the INSDC database, does not reach the same level of taxon coverage as when using partial D1/D2 alone. For the *Mycoleptodonoides* specimen it should be noted, that ITS sequence identity to *Mycoleptodonoides aitchisonii* HMJAU4527 (JF430078.1) was 98%, identity (588/602) bp, and gaps (0/602) bp.

**TAXONOMY**

*Hericium yumthangense* K. Das, Stalpers & Stielow sp. nov.
MycoBank MB800641
(Figs 3–5, 9, 10A–B)

**Etymology:** Named after Yumthang, the type locality.

**Diagnosis:** *Basidiomata* 70–100 × 50–80 mm, intricately branched with primary, secondary, tertiayary and quaternary branches arising from a stipe-like rooting base. *Spines* 7–13 mm long, distributed irregularly throughout the branches. *Sporae* 5.0–5.3–6.5 × 4.0–4.6–5.5 μm, subglobose to broadly ellipsoid, ornamentations are of broad ridges and larger isolated warts. *Hyphal system* monomitic, some contextual hyphae encrusted. *Gloeocystidia* to 8 μm wide, capitulate to subcapitate or moniliform.

**Type:** *India:* Sikkim-Yumthang, alt. 3644 m asl, N 27°47’54.8” E 88°42’16.1”, on *Abies densa* in subalpine mixed forest, 30 Aug. 2011, K. Das KD-11-151 (BSHC).

**Notes:** *Hericium yumthangense* is characterized by the stipe-like small rooting base, intricate three tier branching system, 8–13 mm long spines, basidiospores with isolated warts and ridges, and the encrusted contextual hyphae. It can be distinguished from the other four known species of *Hericium* (*H. bharengense*, *H. erinaceus*, *H. coralloides*, and *H. cirratum*) reported from India, by the presence of encrusted hyphae in the context. Moreover, *H. bharengense* has a long distinctly narrow rooting base and spores with comparatively low ornamentation (0.2 μm high) giving a convoluted to brain-like appearance (Das et al. 2011). *Hericium erinaceus* is either unbranched, cushion-like or sparsely branched, bears long spines (to 5 cm long) and has larger basidiospores 4.0–7.0 × 4.0–5.5 μm (Third & Khara 1975). *Hericium coralloides* has shorter spines (3–5 mm long; Third & Khara 1975). While in *H. cirratum*, the basidioma consists of a regular cluster of pileoli united at the base and the spores are smooth (Das & Sharma 2010).

**Description:** *Basidiomata* 70–100 × 50–80 mm, pendent, consisting mostly of five primary branches arising from a distinct rooting stipe-like base, being attached to the living host (*Abies densa*). Primary branches to 13 mm wide, ramified into progressively thinner (to 6 mm wide) fertile secondary branches that bear thinner fertile tertiary branches followed by thinnest fertile quaternary branches ending with clusters of 3–4 spines, white to pale yellow (a: 6F) or yellowish orange. *Spines* 7–13 mm long, moderately to densely distributed irregularly from all surface of secondary, tertiary and quaternary branches, pendent, concolorous with the branches when fresh, darker, yellow (a: 6F) to pale apricot to sienna (a: 11) or cinnamon (a: 10) when dry. *Context* yellowish white, unchanging when bruised or slowly becoming darker, but becoming straw (a: 50) with KOH, pale yellow (a: 4D–5E) with FeSO4. *Odour* pleasant. *Taste* mild or slightly bitter. *Spore print* white (b: 0 Y). *Hyphal system* monomitic; *contextual hyphae* generative, 4.5–11.5 μm wide, thick-walled (wall up to 3.5 μm thick), with frequent branching and conspicuous clamps of variable diam., some with incrustations; *hyphal lamel trama* hyphae 3–14 μm wide, colourless in KOH, comparatively thin-walled (wall to 2 μm thick), branched, with clamps of variable diam; *glyceopterul hyphae* 4.5–8 μm wide, abundant with dense yellowish contents, apex slightly tapering or rounded sometimes moniliform, septa not found. *Basidiospores* 5.0–5.3–6.5 × 4.0–4.6–5.5 μm, subglobose to broadly ellipsoid (Q = 1.09–1.16–1.24), hyaline, amyloid, under light microscope almost smooth to slightly roughened with eccentric apiculus, composed of broad ridges and isolated warts (under SEM), ornamentations up to 0.4 μm high, blunt. *Basidia* 28–35 × 5–6 μm, clavate to subclavate, with basal clamps, 4-spored, sterigmata 4–5 × 1–1.5 μm. *Gloeocystidia* 5–8 μm wide, capitulate to subcapitate or moniliform, emergent to 15 μm, content dense. *Hymenium* and *subhymenium* inamyloid, hyphae of the subhymenium thin-walled, to 5 μm wide.

**Additional specimen examined:** *India:* Sikkim-Yumthang, alt. 3644 m asl, N 27°47’54.8” E 88°42’16.1”, on *Abies densa* in subalpine mixed forest, 30 Aug. 2011, K. Das KD-11-151 (BSHC).
Fig. 3. *Hericium yumthangense* (KD-11-146). A-B. Intricately branched basidiomata. C. Basidiomata showing the stipe-like base and branches. D. Primary branch bearing secondary, tertiary and quaternary branches and spines. E. Secondary branch bearing tertiary and quaternary branches and spines. Distances between two bars = 1 cm.
and quaternary branches. Furthermore, the spores are distinctly larger (5.5–7 μm diam) in *H. americanum* (Ginns 1984). The basidiomata of *H. abietis* are larger (750 × 250 mm), and without any stipe (represented by large solid tubercles) (Harrison 1973).

**Mycoleptodonoides sharmae** K. Das, Stalpers & Stielow sp. nov.
MycoBank MB800642
(Figs 6–8, 9B, 10C–D)

*Etymology*: In recognition of Jai Ram Sharma for his contribution to the Indian mycobiota.

*Diagnosis*: Basidiomata 50–130 mm long, complex, pileate, pileus solitary to concrescent, single pileus 50–90 mm long (diam), fanshaped. Stipe absent or rudimentary. Spines to 6 mm long and 1.5 mm wide, present on the ventral surface of pileus, subulate to flattened. Spores 5.0–6.0–7.5 × 1.5–1.8–2.0 μm, cylindrical to suballantoid or allantoid, smooth, inamyloid. *Gloeocystidia-like hyphal elements* to 5 μm wide, cylindrical, present at the tip of the spine.

*Type*: India: Sikkim: near Yumthang, 3522 m asl, N 27°46'42.4" E 88°42'47.4", on the decaying tree-trunk of an unidentified broad-leaf tree, subalpine mixed forest, 28 Aug. 2011, K. Das, KD-11-122 (BSHC – holotype; CBS herbarium – isotype); GenBank, ITS (JX855031), LSU (JX855030), and mtSSU (JX855032).

*Description*: Basidiomata to 130 mm long, complex, pileate, consisting of imbricate pilei arising from a rudimentary central base. *Pileus* single to concrescent, 50–90 mm diam, fanshaped, convex, depressed towards the place of attachment, glabrous, fissured near the margin; margin regular to lobed, upturned with maturity, dorsal surface somewhat ribbed or veined but smooth towards margin, white when young and fresh, slowly yellowish with darker (a: 6F) margin; turning light yellow (a: 5E) with patches of yellow (a: 6F) and ochraceous (a: 9H) or saffron (a: 49) after drying, ventral surface spiny, light yellow (a: 5E) when young, becoming more distinctly yellow after drying with sienna (a: 11) at margin. Stipe absent or represented by the narrowest part of the pileus giving a rudimentary (never effused) base that attaches to the substrate; attachment never central. Spines to 6 mm long and 1.5 mm wide, decurrent, blunt, subulate to flattened, light yellow (a: 5E). Context fibrous to leathery, chalky white, becoming yellowish after drying, turning cream (a: 4D) with FeSO₄ and KOH. Odour pleasant. Taste indistinctive. Spore print yellowish white (b: 2Y–5Y). Hyphal system monomitic; context of the pileus composed of generative hyphae; hyphae 3.5–15.5 μm wide, frequently branched, septate with clamps, mostly thick walled (to 2.5 μm thick), sometimes inflated, branches frequently form intricate knots. Context of spines composed of generative hyphae; hyphae to 5 μm wide, thin walled towards the tips of spine, slightly thick walled to solid towards the core. *Basidia* 18–50 × 3.5–6 μm, clavate, 4-spored, sterigmata to 5 μm long and 1.5 μm wide. *Basidiospores* 5.0–6.0–7.5 × 1.5–1.8–2.0 μm, cylindrical to suballantoid or allantoid (Q = 2.77–3.24–3.84),

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**Fig. 4. Hericium yumthangense** (KD-11-146). A. Gloeocystidia. B. Basidia. C. Basidiospores. D. Hymenialtramal hyphae. Bars = 10 μm.

**Fig. 5. Hericium yumthangense** (KD-11-146). A. Gloeoplerous hyphae. B. Contextual hyphae. Bars = 10 μm.
Fig. 6. Mycoleptodonoides sharmae (KD-11-122). A. Single to concrescent basidiomata growing on decaying tree-trunk. B. Concrescent basidiomata. C. Dorsal view of single and concrescent basidiomata showing the rudimentary base. D–E. Ventral view of basidiomata showing spines.
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Additional specimens examined: India: Sikkim: near Yumthang 3556 masl, N 27°46′42.4″ E 88°42′47.4″, on the decaying tree-trunk of an unidentified broad-leaf tree, subalpine mixed forest, 28 Aug. 2011, K. Das KD-11-130 (BSHC); ibid., 3556 m asl, N 27°46′46.2″ E 88°42′45.5″, 2 Sept. 2011, K. Das KD-11-187 (BSHC).

Notes: The solitary to concrescent fan-shaped pileus, blunt subulate to flattened spines, cylindrical to allantoid inamyloid spores (longer than 4 μm), monomitic hyphal system, thick-walled to solid generative hyphae in hymenial trama and absence of skeletal hyphae in spines, place the present taxon in the genus *Mycoleptodonoides*. *Mycoleptodonoides sharmae* can be distinguished by the larger spores. They are 5.5–6.5 × 2.0–2.5 μm in *M. aitchisonii*, 4–5 × 1.5–2 μm in *M. vassiljevae*, and 3.5–4.0 × 2.3–3 μm in *M. tropicalis*. In addition, the spores and rudimentary (neither effused nor stipitate) base and habitat, i.e. a decaying tree-trunk in subalpine forest, are diagnostic.

With large cylindrical to suballantoid spores and gloeocystidia-like hyphal elements at the spine tips, *Mycoleptodonoides aitchisonii* (also reported from India) appears morphologically close to *M. sharmae*. However, the presence of comparatively small spores (5.5–6.5 × 2.0–2.5 μm) and extensively effused base in *M. aitchisonii* (Maas Geesteranus 1961) separate it from the species presented here.

**DISCUSSION**

All taxa in *Hericaceae* cause white rot of deciduous and coniferous trees, and the family currently comprises three genera; *Hericium*, *Laxitextum*, and *Dentipelis*. *Hericium*, has *H. coralloides* as the type species. The application of *Hericium s.str.* has been intensively debated due to the loss of the original specimens. Following several nomenclatural actions, including the neotypification of *H. coralloides* (Hallenberg 1983), the genus has been thoroughly investigated with respect to ecology, strain incompatibility, and very recently, species delimitation (Larsson & Larsson 2003, Hallenberg et al. 2012).

The authors of the latter study were investigating phylogenetic species boundaries in *Hericium* (Hallenberg et al. 2012), using the recently ratified universal fungal DNA barcode ITS (Schoch et al. 2012). Similar to our results, the obtained molecular clades were not fully resolved on the basis of ITS sequence data. The same observation...
was made by Das et al. (2011) when describing the new taxon *H. bharengense* from temperate mixed (broad-leaved and coniferous) Himalayan forests in the western district of Sikkim (India). Due to high interspecific ITS sequence divergence within *Hericium*, additional support from morphological and ecological characters is required to ensure a meaningful delimitation between known and new taxa in the genus. The overall poor phylogenetic resolution in our *Hericium* dataset, reflected by the inferred polytomies (Fig. 1), is in agreement with the analysis by Hallenberg et al. (2012). The core clade reflecting *Hericium s.str.*, with *H. americanum/H. erinaceus* and *H. alpestre/H. abietis* was retrieved with poor statistical support (BS) as in the latter work, but with similar distances between species. However, given the very limited resolution of the ITS, protein coding genes, commonly used for the calibration of concatenated gene phylogenies in *Agaricomycotina*, could improve our limited understanding of speciation in *Hericium*. Beside the lack of phylogenetic resolution on the rDNA level, the new species *H. yumthangense*, can be delimitated from existing taxa occurring in India by morphological characters. In particular, the encrusted contextual hyphae, the rooting base and the comparably low spore-ornamentation, approximately 0.2 µm in height, characterize the species. ITS sequence data should not be regarded as absolutely decisive for interpreting this case, and as was the case in Hallenberg et al.’s (2012) study, our findings are well supported morphological characters. Since it was beyond the scope of this study to revise *Hericium s.str.*, other protein coding genes from additional specimens were not sequenced.

In contrast to the intensively studied genus *Hericium*, the number of taxonomic reports on *Mycoleptodonoides* is very limited. Since Nikolajeva (1952) described the type species *M. vassiljevae* from Ussuri, Russia, several recombinations from *Mycoleptodonoides*, such as *M. adusta* and *M. pusilla* into *Mycorrhaphium* (Maas Geesteranus 1962, Imazeki & Hongo 1989), have been made to clarify the generic delimitation. *Mycoleptodonoides* has a monomitic hyphal system, but thick-walled generative hyphae, and spines on

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**Fig. 10.** Habitat. A. Forested area dominated by Abies densa at Yumthang; collection site of *Hericium yumthangense*. B. Trunk of A. densa with basidiomata of *H. yumthangense*. C. Subalpine mixed broad-leaved and coniferous forest; collection site of *Mycoleptodonoides sharmae*. D. Decaying tree-trunk with basidiomata of *M. sharmae*.
the spores larger than 4 μm. In contrast, the morphologically closely related genus *Mychorraphium* is dimitic, with skeletal hyphae, and smaller spores. However, the nature of the skeletal hyphae resembles more closely the situation in *Stereum* than in *Trametes*, and molecular research is necessary to re-evaluate both the more exact taxonomic position and the value of mitism in the species of concern. While the type species, *M. vassiljevae*, is known only from the type locality and northern China, the second accepted species, *M. aitchisonii*, has a wider distribution from subtropical to boreal habitats. Yuan & Dai (2009) reported a third species from tropical southern China, *M. tropicalis*. The species of *Mycoleptodonoides* can be primarily differentiated by their basidiome morphology, which places those which are stipitate or sessile close together.

The relatively well resolved partial 28S phylogeny, but lack of additional sequence data (i.e. protein coding genes), does not allow any certain and well supported positioning of *Mycoleptodonoides* among the "core polyporoid" taxa. However, the high ITS sequence similarity to a Chinese specimen of *M. aitchisonii* (JF430078), verifies our additionally released DNA barcodes (ITS, mtSSU) for *Mycoleptodonoides*. *M. sharmae* can be delimited from all known species by the basidiomata having a rudimentary base, a solitary to concrescent pileus, subulate to flattened spines, and large cylindrical to allantoid basidiospores (5–7.5 × 1.5–2 μm). Moreover, the exceptional subalpine habitat (collected between 3522–3556 m) in Eastern Himalaya, Sikkim-India, is also likely to be an ecological distinction and renders the new taxon unique within this poorly studied genus.

### Revised key to *Mycoleptodonoides* species

1. Pileus stipitate to substipitate ........................................................................................................... 2
   Pileus sessile or with a rudimentary base .......................................................................................... 3

2 (1) Spores 4–5 × 1.5–2 μm, spines to 5 mm long, generative hyphae to 30 μm wide .................. *M. vassiljevae*
   Spores 5.5–6.5 × 2.0–2.5 μm, spines to 7 mm long, generative hyphae to 15 μm wide .................. *M. aitchisonii*

3 (1) Spores cylindrical to suballantoid or allantoid (Q = 2.77–3.84), pileus solitary to concrescent;
   in subalpine forests .......................................................................................................................... *M. sharmae*
   Spores broadly ellipsoid to ellipsoid, never suballantoid to allantoid, (Q = 1.25–1.33),
   pileus never concrescent; in tropical forests .................................................................................. *M. tropicalis*

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