Growth performance and emergence of invasive alien Rumex confertus in different soil types

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*Rumex confertus* is known to be one of the most serious invasive weed species infesting meadows, pastures and ruderal lands throughout the Central Europe. *Rumex confertus* was grown in pot experiments using 8 soil types at 5 concentrations of nitrogen (N) and phosphorus (P). Based on harvest data, the variables, seedlings emergence, root: shoot (R: S) ratio, N and P concentration, chlorophyll content, Relative Colimitation Index (RCI) and dry matter allocation to plant components, were determined. N and P addition stimulated the growth of plants in different soils, as reflected by a significant increase in seedling growth parameters such as total plant biomass and shoot biomass. Across all soil types, emergence of seedlings was negatively affected by very high N, but positively affected by increased P availability. This study indicates that Dystric Arenosol, Entic Podzol, Brunic Arenosol and Calcaric Leptosol are unfavorable for *R. confertus* growth, excluding *R. confertus* completely. Moreover, evidence suggests that plant growth is limited by both N and P, therefore *R. confertus* could be controlled by reducing available N and P content in the soil.

Invasion of exotic plant species is a global phenomenon that poses a critical problem for the conservation and management of many ecosystems1-2. Plant invasion is influenced by various environmental factors including both abiotic and biotic factors. Soil type is one of the essential abiotic factors which might affect plants growth. It has long been recognized that that invasive plant species have a wide environmental tolerance that allows them to grow and development on different types of soil, enabling them to easily invade new habitats3.

In many terrestrial ecosystems, nitrogen (N) and phosphorus (P) are usually considered the two most common and primary nutrient for plant growth at the biochemical level and are therefore known to play a crucial role in plant growth, especially in invasive plants4-8. N is the nutrient element that plays a crucial role in photosynthesis, plant production, protein synthesis and all enzymatic activity, whereas P is involved in photosynthesis, the energy transfer within cells, the transport of carbohydrates and, together with N, it is an important structural element in DNA and RNA9-12.

Empirical data shown that both N and P, are more apt to be deficient in natural soils than the others13. As N and P availability increases due to agricultural runoff, ecosystems can become more vulnerable to invasion as some invasive plants respond more positively than natives to N- and P-enrichment14. For example, Thorpe et al.15 found that phosphorus concentration in the invasive *Centaurea maculosa* was more than twice that of native species and phosphorus uptake by this invasive species was six times greater than that by the native, *Lupinus argenteus*. Which of the main nutrients limits the growth, development and eventually settling of new habitats depends on the chemical composition of the soil and the availability of nutrients and on the balance between their inputs and outputs16.

Several studies have revealed a strong interaction between these two mineral nutrients in plant growth17-19. For example, positive interaction between N and P which leads to increase in P absorption have reported for *Eucalyptus grandis* plants20. In many terrestrial ecosystems soil pH, soil texture and soil aeration are thought to be the key factors affecting nutrients interaction21-23.

*Rumex confertus*, native to Eastern Europe and Asia, is known to be one of the most serious invasive weed species in Central Europe, but its preferred soil types are not documented. It is also necessary to know the individual (N or P) and combined (N + P) effects on seedling emergence, or subsequent growth of *R. confertus*. In particular I addressed the following questions: (1) how emergence of seedlings as well as the growth of shoots and roots of
Physicochemical properties of the soil (organic matter content, pH and macronutrient content [N, P, K])

Table 1.

| Soil types          | C (%) | N (%) | C:N | K (mg 100 g) | P (mg 100 g) | pH-H2O | pH-KCL | Soil particle size (% mm) |
|---------------------|-------|-------|-----|--------------|--------------|--------|--------|--------------------------|
| Dystric Arenosol    | 0.89  | 0.06  | 14.8| 3.4          | 2.3          | 3.5    | 3      | >2.0                     |
| Entic Podzol        | 1.25  | 0.08  | 15.6| 3.6          | 2.5          | 4.8    | 4.2    | 0.0–0.05                 |
| Brunic Arenosol     | 1.9   | 0.18  | 10.6| 4.8          | 2.8          | 4.9    | 3.4    | 0.05–0.002               |
| Leptosol            | 2.78  | 0.21  | 13.2| 4.6          | 3.5          | 7.7    | 7.3    | <0.002                   |
| Fibric Histosol     | 4.16  | 0.38  | 10.9| 8.5          | 3.9          | 5.0    | 4.6    |                          |
| Fluvic Cambisol     | 5.15  | 0.56  | 9.2 | 13.3         | 8.3          | 6.9    | 6.6    |                          |
| Dystric Gleysol     | 2.82  | 0.29  | 9.7 | 6.4          | 3.7          | 5.8    | 5.3    |                          |
| Eutric Cambisol     | 3.67  | 0.46  | 8.0 | 14.7         | 6.8          | 6.9    | 6.0    |                          |

Comparison in physicochemical properties among different types of cultivation soil.

As shown in Table 1, physicochemical properties were different in different soil types. The contents of C, N, K, P were highest in Fluvic Cambisol, they were 6, 9, 4 and 3 times higher, respectively, than those in Dystric Arenosol, which was the poorest soil. Generally all soils were poor in extractable P. Taking pH-KCL value into account, the studied soils ranged from acidic (Dystric Arenosol – pH 5.5) to alkaline (Leptosol – pH 7.3). Soil C levels varied from 0.89% (Dystric Arenosol) to 5.15% (Fluvic Cambisol), and total soil N levels varied from 0.06% (Dystric Arenosol) to 0.56% (Fluvic Cambisol).

The influences of four types of soil on the growth – visual observations.

In this experiment I selected four representative soil types with quite different textures and physicochemical properties, i.e. Dystric Arenosol (loose texture, the low pH level), Entic Podzol (loose texture, the low pH level), Brunic Arenosol (moderate texture, the low pH level) and Leptosol (dense texture and rich in calcium carbonate). The four soil types used in this experiment had different physicochemical properties. The results of this experiment indicated that during 10 weeks seedlings died shortly after they started growing or else showed poor growth in four soils. The data were not treated statistically as only few plants survived in each soil culture. The seedlings were not measured for aboveground biomass and belowground biomass because of the poor growth conditions and lack of new leaves. Four seedlings which survived on Brunic Arenosol remained at the seedling stage throughout the experiment. During the course of experiment the plants assumed a pale greenish hue. The seedlings grown on Dystric Arenosol and Entic Podzol both contained less nutrients than the other soil types and had lower pH.

Effects of soil type and nutrients on total dry weight, shoot and root biomass and root to shoot biomass (R: S) ratio.

Significant (P < 0.05) differences were observed in the growth of *R. confertus* plants on the soil used in terms of plant biomass (Table 2). In general, the control non-fertilized plants grew better in the Fluvic Cambisol than in the other three soils i.e., Eutric Cambisols, Fibric Histosol and Dystric Gleysols. The total biomass of plants grown in Fluvic Cambisol was about 19–46% greater than of those grown in Fibric Histosol or Dystric Gleysols. Not only the total biomass but also the shoot biomass varied depending on the soil types. It was the largest on Fluvic Cambisol and the smallest on Dystric Gleysol. The R: S tended to be lower in Fluvic Cambisol than in the other three soils.

The growth of *R. confertus* was stimulated by both N and P fertilization. The plants treated with N1 or P1 fertilizer had significantly (Tukey’s test, P < 0.05) greater biomass production (plant dry weight and shoot dry weight) than the control (i.e., 0 g N and P) and the effect was most pronounced in N2 or P2 (i.e., 140 g N and 37.5 g P). In addition, across all soil types, P1 and P2 (i.e., 25 g and 37.5 g) had greater effect than N1 and N2 (i.e., 70 g N and 140 g P). There was also a significant (Tukey’s test, P < 0.05) interaction effect of N and P for total biomass, shoot and root biomass and found that values for these parameters were highest in seedlings treated with N2 + P2 (i.e.,
140 g N and 37.5 g P). Across all the soil types, the total dry biomass was largest at N2 + P2 combination on Fluvic Cambisol. In contrast to the shoots, growth of the roots appeared to be decreased with increasing amounts of inorganic N and P in the soil.

The R: S ratio was significantly (Tukey’s test, P < 0.05) greater levels of chlorophyll than those treated with N1 + P1, N1 + P2, N2 + P1, and N2 + P2 exhibit the lowest R: S ratio. There was also a N-P interaction effect: R: S ratio tended to be lower when N was added in combination (Tukey’s test, P < 0.05).

### Leaf N and P concentrations

The leaf concentrations of N and P differed significantly (P < 0.01) between plants grown on four soil types and tended to increase with increasing level of soil applied N and P (Table 3). The control plants grown on Fluvic Cambisol without N or P fertilization showed the highest N or P leaf concentrations, significantly exceeding other soils (P < 0.001), whereas the lowest N or P leaf concentrations were observed in Dystric Gleysol without N or P. The ranking was: Fluvic Cambisol > Eutric Cambisol > Fibric Histosol > Dystric Gleysol. Fertilization altered leaf N and P concentrations. Plants fertilized with N or P exhibited higher N concentrations than plants without N or P. Across all soil types, N concentrations increased with addition of N alone (Tukey’s test, P < 0.001), P alone (Tukey’s test, P < 0.05), and N and P together (Tukey’s test, P < 0.05). The total foliar P concentration followed a similar trend as the total N with soil type and fertilization (Table 3). Regardless of the soil type, the highest leaf concentrations of N or P were always noted in the plants fertilized with the highest N or P supply. In all analyzed soils, N: P ratios were below 15.

### Leaf chlorophyll content

Different types of the cultivation soil had significant effects (Tukey’s test, P < 0.05) on chlorophyll content in R. confertus (Fig. 1) which was the lowest in the plants grown in Dystric Gleysol. Compared to this in the plants grown in Fluvic Cambisol this parameter was significantly increased by 23% (P < 0.01). The supply of both N and P had significant (Tukey’s test, P < 0.05) positive effects on the chlorophyll content. The plants treated with N1 had significantly (Tukey’s test, P < 0.05) greater levels of chlorophyll than those of the control (0 g N) while in those treated with N2 this parameter was highest. Similarly, the plants treated with P1 or P2 had significantly (Tukey’s test, P < 0.05) greater levels of chlorophyll than those treated with

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**Table 2.** Mean values and their standard deviations for total dry weight (DW), shoot and root biomass, root to shoot (leaf + stem DW) (R: S) ratio of R. confertus plants grown with and without fertilization in four different natural soil types. Values in a column with different letters are significantly different from each other according to Tukey’s HSD post hoc test with Bonferroni correction at P < 0.01.

| Soil types | Nutrients added | Total dry weight (g DW plant⁻¹) | Shoot biomass (g DW plant⁻¹) | Root biomass (g DW plant⁻¹) | Root: shoot (R: S) ratio |
|------------|----------------|---------------------------------|------------------------------|-----------------------------|--------------------------|
|            |                | 140 ± 5.0                    | 13.7 ± 0.5                 | 12.5 ± 0.5                 | 0.97                     |
|            |                | 130 ± 5.0                    | 13.4 ± 0.5                 | 12.3 ± 0.5                 | 0.97                     |
|            |                | 120 ± 5.0                    | 13.1 ± 0.5                 | 12.0 ± 0.5                 | 0.97                     |
|            |                | 110 ± 5.0                    | 12.8 ± 0.5                 | 11.8 ± 0.5                 | 0.97                     |
|            |                | 100 ± 5.0                    | 12.5 ± 0.5                 | 11.5 ± 0.5                 | 0.97                     |
|            |                | 90 ± 5.0                     | 12.2 ± 0.5                 | 11.3 ± 0.5                 | 0.97                     |
|            |                | 80 ± 5.0                     | 11.9 ± 0.5                 | 10.7 ± 0.5                 | 0.97                     |
|            |                | 70 ± 5.0                     | 11.6 ± 0.5                 | 10.4 ± 0.5                 | 0.97                     |
|            |                | 60 ± 5.0                     | 11.3 ± 0.5                 | 10.1 ± 0.5                 | 0.97                     |
|            |                | 50 ± 5.0                     | 11.0 ± 0.5                 | 9.8 ± 0.5                  | 0.97                     |
|            |                | 40 ± 5.0                     | 10.7 ± 0.5                 | 9.5 ± 0.5                  | 0.97                     |
|            |                | 30 ± 5.0                     | 10.4 ± 0.5                 | 9.2 ± 0.5                  | 0.97                     |

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The leaf concentrations of N and P differed significantly (P < 0.01) between plants grown on four soil types and tended to increase with increasing level of soil applied N and P (Table 3). The control plants grown on Fluvic Cambisol without N or P fertilization showed the highest N or P leaf concentrations, significantly exceeding other soils (P < 0.001), whereas the lowest N or P leaf concentrations were observed in Dystric Gleysol without N or P. The ranking was: Fluvic Cambisol > Eutric Cambisol > Fibric Histosol > Dystric Gleysol. Fertilization altered leaf N and P concentrations. Plants fertilized with N or P exhibited higher N concentrations than plants without N or P. Across all soil types, N concentrations increased with addition of N alone (Tukey’s test, P < 0.001), P alone (Tukey’s test, P < 0.05), and N and P together (Tukey’s test, P < 0.05). The total foliar P concentration followed a similar trend as the total N with soil type and fertilization affecting leaf P concentration (Table 3). Regardless of the soil type, the highest leaf concentrations of N or P were always noted in the plants fertilized with the highest N or P supply. In all analyzed soils, N: P ratios were below 15.

### Leaf chlorophyll content

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types of soil. The values are means ± standard deviations. The values in a column with different letters are significantly different from each other according to Tukey's HSD post hoc test with Bonferroni correction at 

| Soil Types          | Nutrients added | N (mg g⁻¹ DW) | P (mg g⁻¹ DW) | N: P ratio (g g⁻¹) |
|---------------------|-----------------|---------------|---------------|-------------------|
|                     | Control         | 18.5 ± 1.2    | 1.47 ± 0.6    | 12.6              |
|                     | N1              | 21.45 ± 0.25  | 1.68 ± 0.15   | 12.8              |
|                     | N2              | 25.87 ± 0.27  | 2.08 ± 0.25   | 12.4              |
|                     | P1              | 27.08 ± 0.28  | 2.15 ± 0.17   | 12.6              |
|                     | P2              | 28.98 ± 0.24  | 2.32 ± 0.31   | 12.9              |
|                     | N1 + P1         | 32.35 ± 0.41  | 2.45 ± 0.26   | 13.3              |
|                     | N2 + P2         | 36.22 ± 0.36  | 2.56 ± 0.46   | 14.1              |
| Fluvic Cambisol     | Control         | 17.6 ± 0.32   | 1.42 ± 0.14   | 12.4              |
|                     | N1              | 20.36 ± 0.25  | 1.66 ± 0.11   | 12.3              |
|                     | N2              | 24.12 ± 0.31  | 1.88 ± 0.34   | 12.8              |
|                     | P1              | 26.06 ± 0.37  | 1.99 ± 0.36   | 13.1              |
|                     | P2              | 28.34 ± 0.28  | 2.06 ± 0.42   | 13.7              |
|                     | N1 + P1         | 30.37 ± 0.24  | 2.15 ± 0.31   | 14.1              |
|                     | N2 + P2         | 32.89 ± 0.36  | 2.218 ± 0.43  | 14.4              |
| Eutric Cambisol     | Control         | 15.38 ± 0.35  | 1.32 ± 0.19   | 11.7              |
|                     | N1              | 18.67 ± 0.41  | 1.53 ± 0.25   | 12.2              |
|                     | N2              | 21.25 ± 0.39  | 1.65 ± 0.32   | 12.9              |
|                     | P1              | 23.41 ± 0.32  | 1.85 ± 0.37   | 12.7              |
|                     | P2              | 25.13 ± 0.35  | 1.97 ± 0.24   | 12.8              |
|                     | N1 + P1         | 27.34 ± 0.41  | 2.11 ± 0.34   | 13.0              |
|                     | N2 + P2         | 30.90 ± 0.35  | 2.24 ± 0.23   | 13.8              |
| Fibric Histosol     | Control         | 12.67 ± 0.41  | 1.29 ± 0.25   | 9.8                |
|                     | N1              | 15.25 ± 0.39  | 1.42 ± 0.22   | 10.7              |
|                     | N2              | 16.41 ± 0.32  | 1.49 ± 0.27   | 11.0              |
|                     | P1              | 21.32 ± 0.26  | 1.58 ± 0.29   | 13.5              |
|                     | P2              | 24.13 ± 0.25  | 1.67 ± 0.24   | 14.4              |
|                     | N1 + P1         | 26.04 ± 0.41  | 1.81 ± 0.24   | 14.3              |
|                     | N2 + P2         | 28.90 ± 0.35  | 1.97 ± 0.23   | 14.7              |
| Dystric Gleysol     | Control         | 18.67 ± 0.41  | 1.53 ± 0.25   | 12.2              |
|                     | N1              | 21.25 ± 0.39  | 1.65 ± 0.32   | 12.9              |
|                     | N2              | 23.41 ± 0.32  | 1.85 ± 0.37   | 12.7              |
|                     | P1              | 25.13 ± 0.35  | 1.97 ± 0.24   | 12.8              |
|                     | P2              | 27.34 ± 0.41  | 2.11 ± 0.34   | 13.0              |
|                     | N1 + P1         | 30.90 ± 0.35  | 2.24 ± 0.23   | 13.8              |
|                     | N2 + P2         | 32.89 ± 0.36  | 2.218 ± 0.43  | 14.4              |

Table 3. The leaf concentrations of nitrogen (N) and phosphorus (P) (mg g⁻¹ DW), N: P ratio (expressed as mass ratio gN/gP) of Rumex confertus plants fertilized with N and P (individually or in combination) in four types of soil. The values are means ± standard deviations. The values in a column with different letters are significantly different from each other according to Tukey’s HSD post hoc test with Bonferroni correction at P < 0.01. N, P, N + P = factorial additions of nitrogen and phosphorus. DW represents dry weight.

| Source of variation | Soil type (S) | Nitrogen (N) | Phosphorus (P) | S × N | S × P | N × P | S × N × P |
|---------------------|---------------|--------------|----------------|-------|-------|-------|-----------|
| Total dry biomass   | 34.78***      | 19.85***     | 18.85***       | 3.23* | 8.05* | 12.89**| 18.43*    |
| Shoot dry biomass   | 25.54***      | 17.87**      | 14.78**        | 5.6*  | 8.9*  | 9.8*  | 6.7*      |
| Root dry biomass    | 17.56**       | 18.21**      | 12.21**        | 9.8*  | 12.4* | 7.51* | 6.0*      |
| R: S ratio          | 5.32*         | 6.56*        | 10.4*          | 6.56* | 8.23* | 8.34* | 5.46*     |
| Total chlorophyll content | 16.31**     | 18.96**     | 18.96**        | 11.78*| 18.6* | 7.65* | 11.05*    |
| Foliar N-concentration | 15.54**   | 16.56*      | 10.32*         | 13.67*| 13.22*| 16.12*| 16.8*     |
| Foliar P-concentration | 14.65*    | 17.34*      | 16.21*         | 14.42*| 15.51*| 10.4* | 14.8*     |

Table 4. Results of three-way ANOVA (numbers represent F-values) for the effects of the soil type (S), nitrogen (N) and phosphorus (P) on the total dry biomass, shoot dry biomass, root dry biomass, total chlorophyll content and foliar N and P concentrations of Rumex confertus seedlings grown with N and P (individually or in combination) in four different natural soils. There were four soil types (Fluvic Cambisol, Eutric Cambisol, Fibric Histosol, Dystric Gleysol), four nitrogen treatments (N1, N2, N1 + P1, N2 + P2) and four phosphorus treatments (P1, P2, N1P1, N2P2). Each treatment had four replicates. S: soil types, N: nitrogen, P: phosphorus. *P < 0.01, **P < 0.01, ***P < 0.01. S: soil types, N: nitrogen, P: phosphorus. Data log transformer.

0 g N (control) regardless of the soil type. A significant interaction (P < 0.05) effect of N and P on the levels of chlorophyll was also observed, the total chlorophyll was highest in the plants treated with N2 + P2 (Fig. 1).

The three-way ANOVA results revealed that all main factors (soil type ‘S’, nitrogen ‘N’, phosphorus ‘P’) and their interactions had significant effects on morphological and physiological variables (Table 4). The parameters, such as total biomass, root and shoot dry biomass were significantly impacted by all the main effects and interactions; while R: S ratio weakly responded both to single nutrient application and their interactions (Table 6). In
addition, the effect of the interaction between nutrient treatments and soil type was found for total chlorophyll content, leaf N and P concentration.

Relative Colimitation Index (RCIN) (i.e. an index of the relative limitation of biomass production by N and P). In all soils, the mean RCIN index for N was between 0.5 and 0.7, while RCIP for P was between 0.7 and 0.9 (Table 5).

Rate of emergence. The three-way ANOVA indicated that the soil type, nitrogen, phosphorus and their interaction significantly affected final seedling emergence percentage (Table 6). In Fluvic Cambisol, higher emergence values were recorded than in Eutric Cambisol (Fig. 2), but in Eutric Cambisol seedlings were green and substantially more vital than in Fluvic Cambisol. The mean emergence over all levels of nutrient availability was 49, 45, 40 and 38% for Fluvic Cambisol, Eutric Cambisol, Fibric Histosol and Dystric Gleysol, respectively. In all soil types, the emergence was lowest in both treatments with N2 applications (N2 and N2 + P2) and these treatments were significantly different from the P1 and P2 treatments. The highest field emergence, was recorded in P1 followed by P2 treatment.

Figure 1. Mean (±SD) leaf chlorophyll content (mg cm⁻²) in *Rumex confertus* plants in relation to the soil types and fertilization treatments. Different letters denote significant differences between mean values (ANOVA, Tukey’s HSD post hoc test with Bonferroni correction at *P* < 0.01). Data log transformer.
Jasione montana dient. Low soil pH-values and low water contents limit plant nutrient uptake. In addition, high H ion activity in these conditions and in appropriate amounts become toxic to plants. It is possible that Al or Mn ions may exclude P in the plants. However, there are no consistent views about the critical pH value for growth of plants. Several reports indicate maximal Al toxicity at pH around 4.5 but other reports – at pH around 5.5. In acidic soils (soil solution pH < 6.0), iron and aluminum amounts are high, which causes either fixation or removal of P from the soil solution. Thus, the species composition of plant communities is determined by competition for nutrients rather than light. Plant species typical of sandy soils have evolved numerous adaptations to the xerothermic microclimatic conditions for example morphological attributes, seed dormancy rhythms and capacity of clonal spread which are not present in R. confertus. Although no measurements were taken, visual assessment revealed that the plants grown on sandy soils poor in N showed symptoms typical of this element deficiency; they were small with pale, chlorotic foliage and showed visible darkening of the foliage and pronounced purple discoloration. Typical signs of P deficiency were found, including increased R: S ratio and reduced biomass. Under natural conditions the majority of these plants undoubtedly would die within a short time in such surroundings, and even if a few managed to survive for a longer period probably competition from species of psammophilous grasslands Spergulo morisonii-Corynephoretum canescens (Tx. 1928) Libb. 1933 e.g., Achillea millefolium agg., Corynephorus canescens, Helichrysum arenarium, Hieracium pilosella, Hypochaeris radicata and Jasione montana would ultimately lead to their death.

Soil variables limiting plant growth and development differ according to the position of the soil in the acidity gradient. Low soil pH-values and low water contents limit plant nutrient uptake. In addition, high H ion activity in these soils elevates concentrations of exchangeable aluminium (Al) or manganese (Mn) ions, which under specific conditions and in appropriate amounts become toxic to plants. It is possible that Al or Mn ions may exclude P from Dystric Arenosol, Entic Podzol and Brunic Arenosol. De Graaf et al. observed that Arnica montana and Cirsium dissectum showed Al toxicity, indicated by poor root development, yellowish leaves and reduced contents of Mg and P in the plants. However, there are no consistent views about the critical pH value for growth of plants. Several reports indicate maximal Al toxicity at pH around 4.5 but other reports – at pH around 5.5. In acidic soils (soil solution pH < 6.0), iron and aluminum amounts are high, which causes either fixation or removal of P from the soil solution. Thus, the availability of inorganic P to plants at soils with pH levels < 5.0. Very little is known about germination and seedling establishment on dry acidic soils. It seems that the low water holding capacity of the soil is the main factor limiting germination and seedling survival. Furthermore, mosses such as Polytrichum piliferum, which may amount to 70% of total vegetation cover, seem to inhibit germination of herbs and grasses.

The poor development and growth of R. confertus on a calcareous Leptosol in the experiment 1 is probably results from the high pH (pH 8–8.5) and exchangeable calcium (Ca). Lime chlorosis in plants grown on calcareous Leptosol was probably attributed to Fe deficiency. In addition, at higher pH levels, calcium may react with phosphorus, creating unavailable phosphate compounds for plants, which may cause poor growth of R. confertus on Leptosol. This is consistent with the pattern found in R. obtusifolius. High demand for certain trace elements (Fe, Mn, Zn, Cu), which are particularly insoluble at high pH, could also be a possible explanation for the absence of R. confertus from calcareous soils.

### Table 5. Relative Colimitation Index (RCI) in relation to soil types and fertilization treatments.

| Soil types                  | RCI_{S} | RCI_{N} | RCI_{P} | RCI_{S\times N} | RCI_{S\times P} | RCI_{N\times P} | RCI_{S\times N\times P} |
|-----------------------------|---------|---------|---------|-----------------|-----------------|-----------------|------------------|
| Fluvic Cambisol             | 0.6     | 0.6     | 0.8     | 0.8             |                 |                 |                  |
| Eutric Cambisol             | 0.5     | 0.5     | 0.9     | 0.8             |                 |                 |                  |
| Fibric Histosol             | 0.6     | 0.6     | 0.7     | 0.7             |                 |                 |                  |
| Gleysol                     | 0.5     | 0.7     | 0.8     | 0.8             |                 |                 |                  |

Table 5. Relative Colimitation Index (RCI) in relation to soil types and fertilization treatments. In all soils, the mean RCI_{S} index for N was between 0.5 and 0.7, while RCI_{P} for P was between 0.7 and 0.9.

### Table 6. Results of three-way ANOVA of characteristics of final seedling emergence of R. confertus in relation to soil types and fertilizer treatments.

| Independent variables | Final emergence percentage |
|-----------------------|---------------------------|
| Soil type (S)         | 46.37***                  |
| Nitrogen (N)          | 23.57**                   |
| Phosphorus (P)        | 21.24**                   |
| S × N                 | 6.45*                     |
| S × P                 | 8.46**                    |
| N × P                 | 18.41*                    |
| S × N × P             | 14.49*                    |

Table 6. Results of three-way ANOVA of characteristics of final seedling emergence of R. confertus in relation to soil types and fertilizer treatments. There were four soil types (Fluvic Cambisol, Eutric Cambisol, Fibric Histosol, Gleysol), four nitrogen treatments (N1, N2, N1P1, N2P2) and four phosphorus treatments (P1, P2, N1 + P1, N2 + P2). Each treatment had four replicates. Figures represent F-values. *P < 0.01, **P < 0.01, ***P < 0.001. Data log transformer.

### Discussion

The development and morphology of a particular plant species can be modified by the soil type in which the plant grows. This may explain the significant differences in the growth parameters and nutrient requirements between R. confertus plants growing on the tested soil types. For example, the results from experiments 1 indicated that the poor development of R. confertus in the Dystric Arenosol, Entic Podzol, Brunic Arenosol and Leptosol was probably a reflection of the low levels of available nitrogen and phosphorus in these soils. Low nutrients availability along with low soil water content are the limiting factors for plant growth on sandy soils i.e., Dystric Arenosol, Entic Podzol and Brunic Arenosol are, thus the species composition of plant communities is determined by competition for nutrients rather than light. Plant species typical of sandy soils have evolved numerous adaptations to the xerothermic microclimatic conditions for example morphological attributes, seed dormancy rhythms and capacity of clonal spread which are not present in R. confertus. Although no measurements were taken, visual assessment revealed that the plants grown on sandy soils poor in N showed symptoms typical of this element deficiency; they were small with pale, chlorotic foliage and showed visible darkening of the foliage and pronounced purple discoloration. Typical signs of P deficiency were found, including increased R: S ratio and reduced biomass. Under natural conditions the majority of these plants undoubtedly would die within a short time in such surroundings, and even if a few managed to survive for a longer period probably competition from species of psammophilous grasslands Spergulo morisonii-Corynephoretum canescens (Tx. 1928) Libb. 1933 e.g., Achillea millefolium agg., Corynephorus canescens, Helichrysum arenarium, Hieracium pilosella, Hypochoeris radicata and Jasione montana would ultimately lead to their death.

Soil variables limiting plant growth and development differ according to the position of the soil in the acidity gradient. Low soil pH-values and low water contents limit plant nutrient uptake. In addition, high H ion activity in these soils elevates concentrations of exchangeable aluminium (Al) or manganese (Mn) ions, which under specific conditions and in appropriate amounts become toxic to plants. It is possible that Al or Mn ions may exclude P from Dystric Arenosol, Entic Podzol and Brunic Arenosol. De Graaf et al. observed that Arnica montana and Cirsium dissectum showed Al toxicity, indicated by poor root development, yellowish leaves and reduced contents of Mg and P in the plants. However, there are no consistent views about the critical pH value for growth of plants. Several reports indicate maximal Al toxicity at pH around 4.5 but other reports – at pH around 5.5. In acidic soils (soil solution pH < 6.0), iron and aluminum amounts are high, which causes either fixation or removal of P from the soil solution. This limits the availability of inorganic P to plants at soils with pH levels < 5.0. Very little is known about germination and seedling establishment on dry acidic soils. It seems that the low water holding capacity of the soil is the main factor limiting germination and seedling survival. Furthermore, mosses such as Polytrichum piliferum, which may amount to 70% of total vegetation cover, seem to inhibit germination of herbs and grasses.

The poor development and growth of R. confertus on a calcareous Leptosol in the experiment 1 is probably results from the high pH (pH 8–8.5) and exchangeable calcium (Ca). Lime chlorosis in plants grown on calcareous Leptosol was probably attributed to Fe deficiency. In addition, at higher pH levels, calcium may react with phosphorus, creating unavailable phosphate compounds for plants, which may cause poor growth of R. confertus on Leptosol. This is consistent with the pattern found in R. obtusifolius. High demand for certain trace elements (Fe, Mn, Zn, Cu), which are particularly insoluble at high pH, could also be a possible explanation for the absence of R. confertus from calcareous soils.
Different types of cultivation soil in the experiment 2 had different impacts on traits of *R. confertus* (Table 4). For example, both the total plant dry weight and shoot biomass were highest in Fluvic Cambisol, higher by 46% and 60% respectively, as compared to the plants in Dystric Gleysol in which their performances were poorest. The differences in these parameters between two types of soil were statistically significant (*P* < 0.05). The relatively poor growth recorded in Dystric Gleysols could be due to poor root growth, resulting in subsequent reduction in the roots absorptive surface area for both water and nutrients.

The R: S ratio may often provide some estimation of nutrient conditions of soils. The mechanisms which regulate nutrient allocation to shoots and roots remain unclear. The nutrient supply hypothesis proposed that plants could increase the relative amount of carbon allocated to the roots (resulting in increased R: S dry weight ratios) under deficiency of N or P but not of K or Mg or when other environmental factors such as Al toxicity.

**Figure 2.** Effects of different fertilizer treatments on field emergence of *Rumex confertus* seedlings grown in four soils. Values in a column with different letters are significantly different from each other according to Tukey’s HSD test with Bonferroni correction at *P* < 0.01). Data log transformer.
limit root growth. Consistently with my first hypothesis, it was found that the shoot biomass increased, and that of roots decreased, with increasing N and P availability. Consequently, R: S ratios were lowered by high nutrient availability in all soil types. The R: S ratio for *R. confertus* also decreased as applied P increased consistent with observations on other species, for example for *Trifolium hirtum* and *T. subterraneum* [15], and *Agave lechuguilla* [16]. Similarly, the R: S ratio decreases as N availability is increased for *Atriplex littoralis* [17] and *Dactyliis glomerata* [18]. However, in some species, such as the perennial leguminous shrub *Lespedeza davurica* and the perennial herbaceous grass *Bothriochloa ischaemum*, P generally stimulates root growth [19]. In other cases, however, not only shoot growth but also root growth of invaders was enhanced by increasing N supply [20].

Numerous papers reported positive interaction between N and P which led to increase in soil P absorption and higher yields [18,20]. As predicted (hypothesis 2) P fertilization improved leaf nutrient status more than N fertilization, which can explain greater effect of the former on plant growth. In this study across all soil types, N concentrations increased with addition of N alone, P alone, and N and P together. However, N content was higher with P fertilization than with N fertilization.

Foliar N concentrations of *R. confertus* in this experiment were comparable to those measured by Vondráčková et al. [21] in *R. obtusifolius* and *R. acetosa*. Several recent studies showed opposite results, i.e. N concentrations in leaves did not increase with soil N concentration [22].

Nutrient limitation may be estimated by indirect measurements of some parameters, the most common of which include (1) nutrient availability in soil [23]; (2) plant investments in acquiring particular nutrients [24]; (3) patterns of limitation assessed by analyzing colimitation indices for N and P nutrients [25], and (4) ratios of nitrogen to phosphorus (N: P) in plant foliage [26–28]. In this study Relative Colimitation Index (RCIN) and the leaf N: P ratio were used as indicators to determine limitation types. It is widely reported that plant growth is N-limited at low N: P values [29–32]. As predicted (hypothesis 3) the shoot biomass of *R. confertus* increased with the N and P addition rate, indicating co-limitation by N and P. Growth co-limitation by N and P was also indicated by the increase in N and P concentrations with increasing N and P supplies. Tomassen et al. [33] showed that the N: P ratios could not predict the responses of individual plant species to fertilization because the N: P ratio tool developed by Koerselman & Meuleman [34] and by Verhoeven et al. [35] is based on the results obtained for the plant community as a whole (i.e. bulk vegetation) rather than for individual species. However, Tessier and Rayna [36] postulated that N: P ratios may be used to indicate nutrient limitation in individual species. To quantify the relationships between N and P, a Relative Colimitation Index (RCIN) was calculated [37]. The results of this study showed that the mean relative colimitation index for P was between 0.5 and 0.7, and for N between 0.7 and 0.9, suggesting that growth was co-limited by N and P.

If N and P are among the macronutrients considered most limiting to plants growing in grassland soils, plants growing on soils of higher fertility should be less responsive to N and P addition than those growing on soils poor in these macronutrients [38]. My research results support to this hypothesis. In this study, addition of these two macronutrients individually and/or together enhanced biomass production more in poor soils than in soils with high fertility.

Leaf chlorophyll content indicates their photosynthetic capacity because low content of chlorophyll limits the process of photosynthesis and, consequently, leads to a decrease in biomass production in the plants [39]. Chlorophyll synthesis depends upon mineral nutrition [40–42]. In vascular plants, high chlorophyll content is typically correlated with increased N uptake [43–44]. For example, it might be due to the availability of N, which plays role in chlorophyll formation. As predicted (hypothesis 4) total chlorophyll contents significantly increased following fertilization with N and P in different soil type. The effects of nutrient addition on chlorophyll content were in agreement with the previous findings [45,46].

In this study, seedlings emergence was negatively affected by very high N availability but positively affected by increased P availability in the soil. It was caused by a negative impact of high NH₄⁺ and/or NO₃⁻ on seed germination [47]. However, growth tends to be higher with a high N supply. This is consistent with the pattern found in *R. obtusifolius* and *R. crispus* [48–49].

The results suggest that the poor emergence and subsequent growth of seedlings in Dystric Gleysol was caused by the negative impact of soil compaction. Effect of soil compaction on poor emergence and *R. confertus* growth observed in this study can be due to decreased root growth. Soil compaction affects soil physical characteristics in grasslands [50]. The most obvious physical effect is lack of oxygen, an increase in soil bulk density and the pore spaces reduction [51–52].

In this study, plants grown in pots filled with soil from the original ecological environment in which they usually grow. However, this study was not designed to reveal the effects of litter traits on germination and seedling emergence. A litter layer sometimes forms a mechanical barrier that prevents seedling emergence and changes environmental factors such as light, soil moisture, and temperature. The effect of litter on seedling emergence and consequently on seedling recruitment in grassland ecosystems could play negative or positive role depending on micro-environmental conditions, thickness of the litter layer and species [53–54]. Therefore, the results of the present study concerning seedlings emergence and growth in pot experiments may therefore have a more general validity despite the use of natural soil.

**Conclusion**

To my knowledge, this study evaluated for the first time the interactive effects of soil types and plant nutrition on the growth and seedling emergence of *R. confertus* plants. Based upon these results it is evident that the development and growth of the *R. confertus* were influenced by nitrogen and phosphorus fertilization. Furthermore, P fertilization increased growth more than N fertilization in all four soils used. The concentration and ratio of these fertilizers were important in determining growth and development. Both N and P fertilization affected growth
The results reveal the reasons for the distribution of *R. confertus*. The ability to make use of available nutrients in order to increase biomass and to produce diaspore is one of the reasons for the settlement success of *R. confertus* in grasslands and meadows in the Warta river valley. The nutrient supply to each site can either be related to the factors associated with the river Warta itself (such as its water quality) or with site-specific factors, such as agricultural pollution (inflows of groundwater and surface water). Under these conditions the concentration of nitrogen and phosphorus is sufficiently high to allow *R. confertus* to colonize such sites and become established. Further research is needed to evaluate the effect of grassland management and nutrient availability on plant performance in temperate grasslands and meadows.

**Materials and Methods**

**Study species.** *Rumex confertus* Willd (Russian or Asiatic dock), probably originating in the Eastern Europe and Asia, where it thrives on meadow-steppes and glades in forest-steppe has been recently observed to successfully colonise Central Europe. The reason for its sudden increase in population growth is not known, although an extended period of germination, rapid growth and high production of small seeds, regeneration of plants form fragments of underground organs, quick and large biomass production and lack of native herbivores were suggested to facilitate its success and spread in Europe. In Poland, where it is an exotic plant, *R. confertus* is considered an aggressive colonizer of meadows, wet ditches, riparian-scrub, roadides, railway tracks and embankments. Under nutrient-rich soil conditions, *R. confertus* is a very strong competitor. *R. confertus* occurs over a wide range of soil types in Poland, whereas it typically grows on moist soils in its native habitat (personal observations). *Rumex confertus* is a perennial herb (Polygonaceae) with stems of 60–120 cm height. It flowers in July and produces one or usually more floreoleances that can reach up to 0.3 m or more. The seeds of *R. confertus* undergo a seasonal dormancy cycle with deep dormancy in winter and early spring and a low level of dormancy in early autumn.

**Experimental design and growth conditions.** Mature *R. confertus* seeds were collected (September 2017) from twenty plants selected randomly, taking care not to favor tall or small, growing in the valley of the Warta River (60 km west of Lodz; N 51°96', E18°79'; central Poland); the seeds were mixed to obtain one representative seed sample. The collection site was a high productive wet meadow on Eutric Cambisol soil. Subsequently, the seeds were stored at room temperature, in paper bags in the dark for two weeks. Deformed and damaged seeds were discarded, and healthy seeds were exposed to cold stratification (±5°C in darkness) until use. Prior to sowing the fertilizers were added and thoroughly mixed into the soil. Up to this time, fertilizers were not used routinely. Three experiments were conducted during 2018 in a greenhouse conditions under natural illuminations with a 12–15-h photoperiod (no extra illumination). The temperature by day was 20–30°C, night 15–20°C and relative humidity of 30–50%. The objectives, growth conditions, soil treatments, and statistical analyses for each experiment are described below. Germination of the seeds tested under laboratory conditions in a 12-h day/night light at 22°C directly before establishment of the experiment was 60–70%. In both experiment 1 and 2, I used 5 L polythene pots 25 cm in diameter and 22 cm deep (491 cm²) filled (leaving 1 cm brim at the top) with air dried soil (approximately, 7 kg of soil). Subsequently, ten seeds of *R. confertus* were sown (1–2 cm deep) in each pot and after 3 weeks of growth, the seedlings were thinned to one plant in each pot. The seedlings were irrigated with tap water daily. Every week, the pots were reshuffled to avoid light/shade or any other greenhouse effects. Nutrient treatments were initiated 2 days after planting. There were four replicate plants for all treatments in all experiments. Germination succeeded well (70–80%) with all soils and was completed within 14 d. The pots were covered with aluminum foil to prevent evaporation. At the end of an experiment the plants were harvested and washed in deionised water for a short time.

**Soil sampling.** The soils for the experiments were collected from eight selected locations, situated in central Poland, which represent eight different habitats (Table 7). All soils were collected from fallows under multispecies plant community that was devoid of alien plants. Soil samples were taken from the top 20 cm of each soil and then was used as the growth substrate. At each site, 4 m² (2 × 2 m) plot with homogeneous soil was designated. In each plot five soil cores with a length of 20 cm and a diameter of 3 cm were collected with a soil borier (one core at each corner of the square and one core at the centre of the square). These five cores were mixed up into a single bulk sample for each plot.

**Soil substrate analysis.** The soil samples were air-dried at 25°C for 3 days, after removal of roots, they were ground and passed through a 2-mm sieve to remove rocks, then thoroughly mixed. Next the soils were analyzed by the Laboratory of Chemical-Agricultural Station in Lodz. Soil composites were analyzed for texture, pH, carbon (C), available P, potassium (K), total N and carbon/nitrogen (C:N ratio). The soil particle size distributions of each sample was determined using the laser diffraction method: clay (<0.002 mm), silt (0.002–0.05 mm), sand (0.05–2.0 mm) and soil skeleton particles (>2.0 mm). Organic matter contents of the soil, were estimated as loss on ignition (550°C, 3 h). Soil pH was determined by mixing a slurry of ten g of air-dried soil with 5 mL deionized water (pHkg), and also after adding 5 drops of 1 M KCl solution to the slurry (pHkg). The Tiurin method was applied to assess the total carbon content. Total N was extracted using micro-Kjeldahl method based on the wet oxidation of organic matter using 30 M sulfuric acid (H₂SO₄), and then, the concentration was determined using the indophenol blue method. The contents of phosphorus and potassium were determined based on the Egner-Riehm method.

Total N and P content in leaf tissue was determined by acid digestion (340°C) of ground (to fine powder using a ball mill) after dried (60°C for 72 h) plant material (subsamples of 100 mg) with a mixture of salicylic acid and
sulphuric acid using a selenium mixture’s catalyst (a modification of the Kjeldahl method). Concentrations in the digests were determined colorimetrically using the molybdenum blue method for phosphorus and the indophenol blue method for nitrogen.

**Experiment 1.** To gain some indication of the tolerance of plants to the soil type they were grown from seeds in soils which do not support this species under natural conditions. Establishment of *R. confertus* form seeds was studied in a greenhouse experiments using four contrasted soil types: Dystric Arenosol, Entic Podzol, Brunic Arenosol and Leptosol. The duration was 10 weeks: sowing date 8 May, harvest date 18 July, 2018. Subsequently, ten seeds of *R. confertus* were sown (1–2 cm deep) in each pot and after 3 weeks of growth, the seedlings were thinned to one plant in each pot. A total number of 12 pots were randomly divided into for groups for soil type.

**Experiment 2.** This was a two-factor experiment involving soil type and nutrient treatments. A fertiliser experiment was carried out to determine whether nitrogen or phosphorus was limiting plant growth at four soil types which support this species under natural conditions. Based on the characteristics of the natural distribution of *R. confertus*, I selected four representative soil types (i.e., Calcaric Leptosol, Fibric Histosol, Fluvic Cambisol and Eutric Cambisol) from four different sites with quite different textures and physiochemical properties. The sites fulfilled the following conditions: (i) having well established and still expanding populations of *R. confertus*; (ii) having sufficiently homogenous soil; (iii) the survey should include a range of habitats from nutrient poor grasslands to highly productive meadows, and (iv) little or no active grazing or herbicide use. At each site where this invader species was found, one plot was marked out (from which soil samples were taken) nearby the areas that had no evidence of invasion.

From the beginning of May 2018 until the end of September 2018 the plants were grown under glasshouse conditions. The seeds were sown as in experiment 1. At sowing time (10 May) the dry, granular fertilizers were added and thoroughly mixed into the soil. Seven nutrient treatments i.e., single-nutrient (N and P) and dual-nutrient (N + P, herein NP) were used in this research, differing in the amount of N or P: N1, N2, P1, P2, N1 + P1, N2 + P2, and control without any fertilized input (Table 8). Nitrogen was applied as ammonium nitrate with lime (NH4NO3 + CaCO3, containing 27.5% N, 10% Ca) — N treatment and P was applied as super phosphate (Ca(H2PO4)2 + CaSO4, 8.5% P, 20% Ca, 10% S) — P treatment. There were also combined N and P control (only demineralised water) treatments as indicated in Table 1. Fertilizer rates were calculated on a surface area basis. The design was a 4 × 7 factorial (soil type × N–P treatments) for a total of 28 replicated four times and arranged in a randomized complete block, i.e. 112 pots per experiment with each pot being considered an experimental unit.

The plants were harvested after a growth period of five months and their biomass divided into roots and shoots (i.e., stems with leaves). The roots were first washed thoroughly with tap water to remove soil adhered to below-ground organs. The total plant biomass (g plant⁻¹) was the sum of root, stem and leaf biomass. Dry weight of the plant parts was measured after drying the material for at least 72 h at 60°C. Dry biomass was obtained by weighing individual floral parts in a microbalance. From these primary data the root to shoot (R: S) ratio was calculated as the root dry biomass divided by shoot biomass. To collect and prepare leaf samples the first mature leaf (blade plus petiole) was taken from each plant per treatment combination in each soil type. After drying, the leaves were ground with a miniature coffee mill to pass through a 0.7-mm mesh screen. The leaf samples were then analyzed for N and P concentrations (% dry mass), as in the soil samples.

Relative Colimitation Index (RCIi) was used to quantify the relationships between N and P. The equation was as follows:

$$\text{RCI}_i = \frac{(B_N - B_C)/(B_{N+P} - B_C)}{(B_{N+P} - B_C)/(B_{N+P} - B_C)}$$  (1)

where, Bc is the biomass of unfertilized plants (control), BN is the biomass of plants fertilized with N, and BN+P is the biomass of plants fertilized with N and P. RCIi value between 0 and 0.5 indicates that the plants are more classically co-limited by N and P than they are primarily limited by N. If RCIi is between 0.5 and 1, plants are primarily limited by N. By analogy to N, an index was also calculated for P.
Leaf chlorophyll content was determined by measuring the ratio between the chlorophyll fluorescence at 735 nm and 700–710 nm \(^{76}\) by a hand-held instrument (CCM300, Opti-Sciences, Inc., Hudson, USA), using standard manufacturer-recommended protocols. The working principal and design of CCM-300 is based on the work of Gitelson \textit{et al.} \^{76} using the equation (Chl) = 634 × F735/F700 + 391. The chlorophyll content (mg/m\textsuperscript{2}) of the same plant in each treatment was measured in recently fully expanded leaf (from the tip, center, and base of each leaf) per each plant. Mean values of the three sections were used for data analysis.

**Statistical analysis of the data.** Analyses of variance (ANOVA) were performed for total plant biomass, Chl content, leaf tissue mineral element concentrations as well as R: S and N: P values. Tukey’s post-hoc test was used to identify differences between treatments. Differences were considered significant at a probability of \(P < 0.05\). The effects of soil types and different N and P concentrations on morphological and physiological variables were analyzed by three-way factorial ANOVA. Normality was verified with the Shapiro-Wilk test. Prior to statistical analysis, the data were log-transformed to make the variance less dependent on the means and to fit a normal distribution. Statistical analyses were performed using the Statistica 13.0 package \^{29}.

**Data availability**

All data generated or analyzed during this study are included in this published article.

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### Table 8. Fertilizer treatments and the amount of nutrients application. *Number in bracket is equivalent to the total amount in kg ha\(^{-1}\) of the applied nutrients.

| Treatment | Nutrients added (mg pot\(^{-1}\) year\(^{-1}\)) |
|-----------|------------------------------------------|
|           | N | P     |
| Control   | — | —     |
| N1        | 70 (150) | —     |
| N2        | 140 (300) | —     |
| P1        | — | 25 (50) |
| P2        | — | 37.5 (75) |
| N1 + P1   | 70 (150) | 25 (50) |
| N2 + P2   | 140 (300) | 37.5 (75) |
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Author contributions
J. Kołodziejek conceived and designed the study, performed the experiments, collected the data, prepared the analysis and figures, and wrote the manuscript.

Competing interests
The author declares no competing interests.

Additional information
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