Richardson, T. G., Richmond, R. C., North, T. L., Hemani, G., Davey Smith, G., Sharp, G. C., & Relton, C. L. (2019). An integrative approach to detect epigenetic mechanisms that putatively mediate the influence of lifestyle exposures on disease susceptibility. *International Journal of Epidemiology*, 48(3), 887-898. https://doi.org/10.1093/ije/dyz119

Publisher's PDF, also known as Version of record

License (if available):
CC BY

Link to published version (if available):
10.1093/ije/dyz119

Link to publication record in Explore Bristol Research
PDF-document

This is the final published version of the article (version of record). It first appeared online via Oxford University Press at https://academic.oup.com/ije/article/48/3/887/5525259. Please refer to any applicable terms of use of the publisher.

University of Bristol - Explore Bristol Research

General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available:
http://www.bristol.ac.uk/pure/about/ebr-terms
An integrative approach to detect epigenetic mechanisms that putatively mediate the influence of lifestyle exposures on disease susceptibility

Tom G Richardson, Rebecca C Richmond, Teri-Louise North, Gibran Hemani, George Davey Smith, Gemma C Sharp and Caroline L Relton

MRC Integrative Epidemiology Unit (IEU), Population Health Sciences, Bristol Medical School, University of Bristol, Oakfield House, Oakfield Grove, Bristol BS8 2BN, UK

*Corresponding author. MRC Integrative Epidemiology Unit, Population Health Sciences, Bristol Medical School, University of Bristol, Oakfield House, Oakfield Grove, Bristol BS8 2BN, UK. E-mail: Tom.G.Richardson@bristol.ac.uk

Abstract

Background: There is mounting evidence that our environment and lifestyle has an impact on epigenetic regulatory mechanisms, such as DNA methylation. It has been suggested that these molecular processes may mediate the effect of risk factors on disease susceptibility, although evidence in this regard has been challenging to uncover. Using genetic variants as surrogate variables, we have used two-sample Mendelian randomization (2SMR) to investigate the potential implications of putative changes to DNA methylation levels on disease susceptibility.

Methods: To illustrate our approach, we identified 412 CpG sites where DNA methylation was associated with prenatal smoking. We then applied 2SMR to investigate potential downstream effects of these putative changes on 643 complex traits using findings from large-scale genome-wide association studies. To strengthen evidence of mediatory mechanisms, we used multiple-trait colocalization to assess whether DNA methylation, nearby gene expression and complex trait variation were all influenced by the same causal genetic variant.

Results: We identified 22 associations that survived multiple testing ($P < 1.89 \times 10^{-7}$). In-depth follow-up analyses of particular note suggested that the associations between DNA methylation at the ASPSCR1 and REST/POL2RB gene regions, both linked with reduced lung function, may be mediated by changes in gene expression. We validated associations between DNA methylation and traits using independent samples from different stages across the life course.

Conclusion: Our approach should prove valuable in prioritizing CpG sites that may mediate the effect of causal risk factors on disease. In-depth evaluations of findings are
necessary to robustly disentangle causality from alternative explanations such as horizontal pleiotropy.

**Key words:** DNA methylation, Mendelian randomization, mediation, smoking, ALSPAC, ARIES

**Introduction**

Epigenetic factors have long been postulated as molecular mechanisms that are responsible for mediating the effect of causal risk factors on disease outcomes.\(^1\)\(^-\)\(^4\) However, evidence implicating specific mechanisms across the genome in this regard is scarce. Recent epigenome-wide association studies (EWAS) have identified hundreds of associations between risk factors and variation in DNA methylation levels\(^5\)\(^-\)\(^8\), an epigenetic regulation process known to play a key role in cell development and disease susceptibility.\(^9\)\(^,\)\(^10\) Interpreting the findings of EWAS is a challenging endeavour, particularly due to reverse causation and unmeasured confounding factors that may influence results.\(^11\)\(^-\)\(^13\)

An approach that can be used to address this challenge is Mendelian randomization (MR), a method by which genetic variants can be used as a proxy for modifiable exposures to infer causality among correlated traits.\(^14\)\(^-\)\(^16\) Research undertaken using the Accessible Resource for Integrated Epigenomics Studies (ARIES) resource\(^17\) has identified thousands of CpG sites where variation in DNA methylation levels is associated with genetic variants, known as methylation quantitative trait loci (mQTL).\(^18\) Undertaking MR analyses using mQTL as instrumental variables can therefore be used to investigate potential downstream effects of changes in DNA methylation levels on human health and disease risk. Furthermore, under the assumptions of two-sample MR (2SMR), it is possible to harness findings from large-scale, well-powered genome-wide association studies (GWAS) to obtain effect estimates for mQTL on hundreds of complex traits across the human phenome.

We have undertaken a study that builds upon the methodology established by two-step epigenetic MR\(^19\) to assess the potential downstream effects of DNA methylation variation associated with smoking\(^5\) (Figure 1). We focus on this exposure as a key risk factor to illustrate this approach as previous studies have provided strong evidence that it acts as a determinant of DNA methylation levels.\(^20\)\(^-\)\(^22\) Moreover, large EWAS of prenatal smoking have identified hundreds of CpG sites across the genome associated with DNA methylation levels, which is attractive when investigating consequences of epigenetic changes as reverse causation is extremely unlikely (i.e. prenatal smoke exposure influences DNA methylation and not vice versa). We have investigated consequences of these putative molecular changes across the human phenome using findings from 643 large-scale GWAS. Next we applied multiple-trait colocalization to discern whether variation in DNA methylation levels may influence traits via changes in the transcription of nearby genes to associated CpG sites. Finally, we have undertaken sensitivity analyses to explore associations in detail, as well as validation analyses using an independent sample of young individuals to investigate potential in utero effects.

**Methods**

**Study population**

The Avon Longitudinal Study of Parents and Children (ALSPAC) is a population-based cohort study investigating genetic and environmental factors that affect the health...
and development of children. The study methods are described in detail elsewhere (http://www.bristol.ac.uk/alspac). Briefly, 14,541 pregnant women residents in the former region of Avon, UK, with an expected delivery date between 1 April 1991 and 31 December 1992, were eligible to take part in ALSPAC. Detailed information and biosamples have been collected on these women and their offspring at regular intervals, and are available through a searchable data dictionary (http://www.bris.ac.uk/alspac/researchers/our-data/). Written informed consent was obtained for all study participants. Ethical approval for the study was obtained from the ALSPAC Ethics and Law Committee and the Local Research Ethics Committees.

DNA methylation used in the study was derived using blood samples obtained from 1018 ALSPAC mother-offspring pairs (mothers at two timepoints and their offspring at three timepoints) as part of the Accessible Resource for Integrated Epigenomic Studies (ARIES) project. The Illumina HumanMethylation450 (450 K) BeadChip array was used to measure DNA methylation at over 480,000 sites across the epigenome. Genotype data were available for all ALSPAC individuals enrolled in the ARIES project, which had previously undergone quality control, cleaning and imputation at the cohort level. Details on methylation assays, genotyping and imputation can be found in the Supplementary Material, available as Supplementary data at IJE online.

Identifying eligible CpG sites using results from EWAS and the mQTL database

Prior to undertaking analysis, we identified CpG sites across the epigenome that current findings and accessible datasets allowed us to investigate. As an example, we obtained a list of 2303 CpG sites that were associated with prenatal smoking based on findings from Joubert et al., using a false discovery rate (FDR)<0.01 in their analysis adjusted for cell counts. Next, we evaluated which of these CpGs we were able to instrument using an mQTL. In this study we used the mQTL database (http://www.mqtldb.org) for this purpose, which reports SNP-methylation effect estimates obtained from the ARIES project (http://www.ariesepigenomics.org.uk/). However, there will likely be alternative resources to this with larger sample sizes in the forthcoming years. Moreover, future research may eventually be able to access mQTL effects derived from different tissue types than whole blood as undertaken in this study.

As we wanted to use mQTLs as instrumental variables for changes in DNA methylation influenced by prenatal smoke exposure, we have only used results from the birth timepoint in ARIES (i.e. based on DNA methylation data derived from cord blood). Conditionally independent mQTLs were identified in ARIES using the COJO-slct routine in GCTA and applying a threshold of $P < 1.0 \times 10^{-7}$. This threshold was merely applied as a heuristic to identify instruments for our analysis though and future
applications may wish to apply alternative criteria. Furthermore, we only identified cis-mQTLs as suitable instrumental variables for our analysis, i.e. genetic variants associated with proximal DNA methylation levels (<1 MB distance away). This is because trans-mQTLs are more likely to be prone to horizontal pleiotropy, i.e. where the genetic variant influences DNA methylation and the associated complex trait via two independent biological pathways. As CpG sites in ARIES are typically only independently associated with a complex trait via two independent biological pathways. As such, future resources that uncover a larger number of conditionally independent mQTL to instrument CpG sites may contemplate their inclusion in study designs.

Complex trait data from large-scale GWAS and the UK Biobank study

We next obtained observed effects for each mQTL identified in the previous stage of our analysis on a range of complex traits using large-scale studies available from the MR-Base platform (http://www.mrbase.org).27 The following inclusion criteria were used to select complex traits to be analysed:

- Study samples must be larger than 1000
- Effects reported genome-wide for over 100 000 genetic variants
- Either European or mixed populations
- Reported beta, standard error (SE) and effect alleles for variants

We also downloaded GWAS summary statistics from the Neale lab28 for complex traits not represented in MR-Base. When effect estimates for complex traits were not available for an mQTL, we identified a proxy instrument in high linkage disequilibrium (\(r^2 > 0.8\)) using genotyped data from Europe individuals as part of phase 3 (version 5) of the 1000 genomes project.29 Although we have evaluated associations on a wide array of traits in this study, future applications may wish to investigate hypothesis-driven relationships depending on the environmental exposure of interest.

Statistical analysis

Two-sample Mendelian randomization

We used 2SMR to evaluate whether variation in DNA methylation at CpG sites associated with our exposure (i.e. prenatal smoke exposure in this study) were also associated with complex trait variation (Figure 1). When DNA methylation levels at CpG sites only had a single cis-acting instrument (i.e. independent cis-mQTL), MR analyses were undertaken using the Wald ratio test.30 For single genetic variant \(j\), this can be calculated by taking the ratio of the gene–outcome association (denoted by \(\hat{Y}_j\)) and the gene–exposure association (denoted by \(\hat{Y}_j\)) estimates:

Where two or more genetic instruments were available for a CpG site, we used the inverse variance weighted (IVW) method to obtain MR effect estimates:

\[
\hat{\beta}_{IVW} = \frac{\sum j^n_2 \sigma_{Yj}^2 \hat{\beta}_j}{\sum j^n_2 \sigma_{Yj}^2}
\]

where \(\sigma_{Yj}\) is the SE of the gene–outcome estimate for variant \(j\). Although the majority of CpG sites in our study could only be instrumented using a single genetic variant, we expect the number of independent mQTL for a given CpG site to increase with larger sample size mQTL studies. As such, future applications of our approach may benefit from using the IVW method to harness multiple independent cis-acting variants as instrumental variables. Furthermore, this will allow MR sensitivity analyses to be undertaken, such as assessing heterogeneity and conducting leave-one-out analyses.

Multiple-trait colocalization

When investigating how changes in DNA methylation may influence downstream traits in an MR framework, it is important to evaluate whether association signals may be a product of linkage disequilibrium between two separate causal variants (i.e. one influencing changes in DNA methylation and another influencing trait variation).31,34 Furthermore, if changes in DNA methylation at a CpG site truly influence complex trait variation, then they are likely to be doing so via changes in the expression of a nearby gene. We therefore applied multiple-trait colocalization35 to investigate associations that survived multiple testing comparisons in the 2SMR analysis (i.e. 0.05/number of CpG-trait combinations analysed). This allowed us to assess whether the genetic variant responsible for changes in DNA methylation is also responsible for both variation in nearby gene expression and the associated complex trait.

Multiple-trait colocalization was applied using mQTL data from the Focus on Mother’s (FOM) timepoint in ARIES (mean age: 47.5), GWAS summary data for the associated complex trait and expression quantitative trait loci (eQTL) data derived from whole blood that was obtained from the GTEx consortium v7 (\(n = 369\)).36 mQTL data from the FOM timepoint was used to evaluate
whether effects observed in the previous analysis using cord blood persisted until later life. We ran this analysis multiple times to systematically investigate colocalization with the expression of all genes within 100 kb of the CpG site of interest. Analyses were only undertaken if there were at least 50 variants (minor allele frequency \( \geq 5\% \)) in common between all three datasets. As recommended by the authors of this approach, a posterior probability of association (PPA) of 80% or higher was considered evidence of colocalization. Additionally, multiple testing was not accounted for in these analyses as the authors of the multiple-trait colocalization paper demonstrated that after 500 000 simulations there was no indication of inflated false-positive rates using this method (even with a PPA threshold of 30%). If a CpG site was associated with multiple correlated traits, only the trait with the highest PPA threshold of 30%). If a CpG site was associated with multiple correlated traits, the trait with the highest PPA in the colocalization analysis was taken forward.

**Orienting directions of effect between traits**

For associations that provided evidence of colocalization in the previous analysis, we evaluated the direction of effect between 2SMR estimates and observed effects from the Joubert study.\(^5\) This allowed us to orient direction of effects between prenatal smoke exposure, DNA methylation and the complex traits. Such evaluations are important to validate that directions of effect between environmental exposures and outcomes are consistent with evidence from the literature.

We also undertook an analysis in ALSPAC to investigate the association between maternal smoking and any traits with evidence of association from our MR and multiple-trait colocalization analysis. This was to evaluate the proportion of the effect mediated via changes in DNA methylation levels along the causal pathway between our exposure and associated outcome. Analyses in this study were adjusted for maternal education, age and sex.

To investigate evidence of reverse causation, we evaluated the association between complex traits (i.e. modelled as our exposure) and DNA methylation (i.e. our outcome) in a 2SMR framework. As a sensitivity analysis, we also used the MR directionality test to assess the direction of causality between molecular and complex traits.\(^37\) This test was applied three times for each association, to evaluate the direction between each pair of traits (i.e. DNA methylation, gene expression and complex trait). This allowed us to undertake an exploratory analysis regarding the chain of direction between all three traits concerned.

**Comparison of findings using two independent samples**

We also repeated 2SMR analyses for the associations that provided evidence of colocalization using two independent samples. Along with validating findings in separate populations, this stage of our approach can be valuable in terms of exploring the temporality of associations. For instance, as we selected prenatal smoking as our exposure in this study, we were interested in assessing the effect estimates for findings at an early stage in the life course.

For this we undertook an initial analysis within the ALSPAC cohort, using mQTL effects for ARIES participants found at https://github.com/eptgr/prenatal-CpGs.Powercalculator using base R graphics. Code used in this project can be found at https://github.com/eptgr/prenatal-CpGs. Power calculations based on the effect sizes identified in this study were undertaken using the MR power calculator.\(^40\)

**Results**

Assessing the impact of DNA methylation variation at birth on the human phenotype

There were 412 unique CpG sites associated with prenatal smoke exposure (FDR<0.01)\(^5\) that met our inclusion
criteria based on data from the ARIES project\(^7\) (Figure 1a, Supplementary Table 1, available as Supplementary data at IJE online). We then assessed the potential effect of changes in DNA methylation on 643 complex traits at these CpG sites using 2SMR (Figure 1b, Supplementary Table 2, available as Supplementary data at IJE online).

Overall there were 22 CpG-trait associations across 12 distinct CpG sites that survived multiple testing corrections \(P < 1.89 \times 10^{-7}\) (Table 1 and Supplementary Table 3, available as Supplementary data at IJE online). Among these associations were measures of anthrometry (height and waist-hip-ratio), lung function [forced vital capacity and forced expiratory volume in 1 second (FEV1)], cardiovascular traits (low-density lipoprotein cholesterol and systolic blood pressure) and bone mineral density. A flowchart illustrating the analysis pipeline used in this study to identify and evaluate these effects can be found in Figure 2.

### Table 1. Top findings from Mendelian randomization analysis for prenatal smoke exposure-associated CpG sites and complex traits

| CpG site   | Gene       | Complex trait       | MR Beta (SE) | MR P          | PPA\(_{abc}\) |
|------------|------------|---------------------|--------------|---------------|--------------|
| cg26930078 | C17orf53   | BMD                 | 0.093 (0.010) | 1.50 \times 10^{-20} | 1.84 \times 10^{-05} |
| cg02812767 | LOXL1      | FVC                 | -0.051 (0.006) | 3.78 \times 10^{-20} | 0.220 |
| cg08685733 | C17orf53   | BMD                 | -0.108 (0.013) | 5.78 \times 10^{-18} | 0.176 |
| cg25313468 | REST       | Height              | -0.064 (0.008) | 5.37 \times 10^{-17} | 9.61 \times 10^{-06} |
| cg06105699 | ASPSCR1    | FEV1                | 0.053 (0.007) | 7.29 \times 10^{-15} | 1 |
| cg23184042 | DLX6AS     | BMD                 | -0.038 (0.005) | 1.26 \times 10^{-14} | 1.31 \times 10^{-04} |
| cg14150774 | QSOX2      | FVC                 | 0.038 (0.005) | 1.99E -12 | 5.31 \times 10^{-05} |
| cg18883198 | TMEM57     | LDL                 | -0.099 (0.016) | 2.72 \times 10^{-10} | 5.22 \times 10^{-06} |
| cg01401641 | TSHZ3      | SBP                 | 0.040 (0.007) | 4.25 \times 10^{-09} | 0.158 |
| cg01307174 | ARPP-21    | Worrying            | 0.025 (0.004) | 2.65E -08 | 0.003 |
| cg25313468 | REST       | FEV1                | -0.030 (0.006) | 4.86 \times 10^{-8} | 0.955 |
| cg18089426 | DLK1       | Age at menarche     | 0.099 (0.019) | 9.00 \times 10^{-08} | 1.09 \times 10^{-04} |
| cg06070002 | PRDX1      | Waist-to-hip ratio  | 0.056 (0.011) | 1.08 \times 10^{-07} | 9.62 \times 10^{-05} |

MR Beta (SE) units are in standard deviations (SD) meaning that 1 SD increase in DNA methylation relates to X SD change in the trait. CpG site, ID based on Illumina mappings; Gene, as reported by Joubert et al.\(^7\) or nearest gene if not specified; MR, Mendelian randomization; SE, standard error; \(P\), P-value; PPA\(_{abc}\), highest posterior probability of association of colocalization between all 3 traits i.e. DNA methylation (a), gene expression (b) and complex trait (c) (PPAs surviving the 0.8 threshold used in this study are in bold). BMD, bone mineral density; FVC, forced vital capacity; FEV1, forced expiratory volume in 1 second; LDL, low-density lipoproteins; SBP, systolic blood pressure; Worrying, a Yes/No questionnaire based outcome in response to the question ‘Do you worry too long after embarrassment?’

Eliciting putative epigenetic mechanisms in disease using multiple-trait colocalization

If DNA methylation levels at a CpG site truly have an impact on human health and disease, then we would expect the transcription of nearby genes to also be involved. Therefore, to strengthen evidence of a causal association, we integrated data on gene expression from the GTEx consortium\(^36\) for all genes within a 100 kb distance of the CpG sites associated with complex traits. As DNA methylation data in this study was only available using data from whole blood, we only used gene expression data also derived from this tissue type from GTEx. Applying multiple-trait colocalization provided evidence at two CpG sites that variation in gene expression, DNA methylation and complex traits were driven by the same underlying genetic variant (Supplementary Table 4, available as Supplementary data at IJE online). These were cg06105699 and cg25313468 which are located at the promoter regions of the ASPSCR1 and REST genes respectively. DNA methylation at cg06105699 colocalized with measures of lung function and the expression of ASPSCR1 (highest \(PPA = 1\)), whereas cg25313468 colocalized with measures of lung function and the expression of POL2RB (highest \(PPA = 0.955\)). Figure 3 illustrates the overlapping distributions of effects for genetic variants at the ASPSCR1 region on FEV1, ASPSCR1 expression and DNA methylation at cg06105699. Supplementary Figure 1, available as Supplementary data at IJE online, provides a similar plot illustrating the effect at the REST/POL2RB locus. There was also evidence of colocalization at one other CpG site for DNA methylation and complex traits but not gene expression (cg08685733 near C17orf53, associated with bone mineral density) (Supplementary Table 5, available as Supplementary data at IJE online).

Orienting the direction of effect between prenatal smoke exposure, molecular traits and outcomes

Using observed estimates reported by the EWAS of prenatal smoking\(^5\) we were able to infer that prenatal smoke exposure was putatively associated with reduced lung function via methylation at the ASPSCR1 and REST/POL2RB loci. Prenatal smoke exposure was negatively associated with DNA methylation at ASPSCR1 [Beta: \(-0.008\) (SE: 0.002)]
and our MR analysis suggests that reduced DNA methylation at this CpG site correlates with reduced lung function [MR Beta: 0.053 (MR SE: 0.007)]. At the REST/POLR2B locus, prenatal smoke exposure was positively associated with DNA methylation levels [Beta: 0.012 (SE: 0.003)] and our MR analysis suggests that increased methylation at this CpG site also may influence reduced lung function [MR Beta: 0.030 (MR SE: 0.006)].

We next undertook MR analyses to evaluate reverse causation (i.e. whether genetic liability to variation in lung function has an effect on DNA methylation at these CpG sites), although there was no strong evidence of this (lowest $P = 0.026$ across 20 tests) (Supplementary Table 6, available as Supplementary data at IJE online). A sensitivity analysis using the MR directionality test complemented these findings, as it suggested the direction of effect between molecular and complex traits at each locus was that DNA methylation influences gene expression, which subsequently influences lung function (Supplementary Table 7, available as Supplementary data at IJE online). Although we may hypothesize that this chain of events is the most parsimonious explanation for associations based on the results of our study, in-depth function studies are necessary to robustly demonstrated that this is the true underlying causal pathway.

Our analysis with regard to maternal smoking and FEV1 in ALSPAC identified weak evidence of association (Beta = −0.02, SE = 0.01, $P = 0.07$, $n = 5417$). However,
the direction of effect in this analysis was consistent with oriented effects between prenatal smoke exposure, CpGs and FEV1 (i.e. maternal smoking is associated with reduced lung function). We were however unable to investigate the proportion of effect potentially mediated between our exposure and outcome via changes in DNA methylation in this analysis. Future studies interested in applying our approach in larger samples sizes may benefit from investigating the proportion potentially mediated along putative causal pathways using well-powered effect estimates.

Validation of results using multiple timepoints over the life course

We investigated the observed associations at the two CpG sites of interest during early stages in the life course using data from the ALSPAC cohort. There was weak evidence that DNA methylation levels at either CpG site was associated with FEV1 at age 8 (ASPSCR1—MR Beta (SE) = −0.017 (0.010), P = 0.086, n_outcome = 5552; REST/POL2RB—MR Beta (SE) = 0.014 (0.012), P = 0.226, n_outcome = 4644). However, evidence was stronger at age 15.5 despite having a smaller sample size (ASPSCR1—MR Beta (SE) = −0.073 (0.029), P = 0.013, n_outcome = 3852; REST/POL2RB—MR Beta (SE) = 0.078 (0.032), P = 0.016, n_outcome = 3240). Full results from this analysis can be found in Supplementary Table 8, available as Supplementary data at IJE online.

Discussion

In this study we have proposed a pipeline of analytical techniques to investigate the potential functional implications of associations between lifestyle risk factors and changes in DNA methylation levels, as opposed to an in utero effect due to maternal smoke exposure. Our split sample MR analysis using data from the UK Biobank supported this, as the observed associations were not confined to the subset of individuals who reported exposure to maternal smoking around birth (Supplementary Table 9, available as Supplementary data at IJE online).

We also evaluated these effects using data obtained during adulthood that provided strong evidence of association between DNA methylation at these loci and their relevant traits (ASPSCR1—MR Beta (SE) = −0.025 (0.005), P = 9.23 × 10^{-08}, n_outcome = 307 638; REST/POL2RB—MR Beta (SE) = 0.034 (0.005), P = 2.90 × 10^{-13}, n_outcome = 307 638). This was undertaken in an independent sample of individuals to the analysis in adolescence, as we used mQTL data from the BIOS QTL browser and the UK Biobank study. These findings therefore help validate the robustness of these associations, as well as suggesting that own smoke exposure may be more likely to be responsible for putative effects rather than maternal smoke exposure. Moreover, the direction of MR effects at both loci were consistent between the population of adults and adolescents (Supplementary Table 8, available as Supplementary data at IJE online).
measures of DNA methylation. We have illustrated this approach by evaluating CpG sites previously linked with maternal smoke exposure, where the strongest evidence of association suggested that putative smoke exposure-associated epigenetic changes may potentially influence reduced lung function at the ASPSCR1 and REST/POL2RB loci via changes in gene expression. The other association signals detected by this study require further validation to robustly link DNA methylation variation with later life outcomes, although upcoming datasets across various tissue types will facilitate such endeavours. Furthermore, this proposed pipeline should prove valuable for future studies that harness these forthcoming datasets and findings of large-scale EWAS. Doing so will help uncover putative epigenetic mechanisms that may play a mediatory role between risk factors and disease.

DNA methylation variation at either the ASPSCR1 or REST/POL2RB locus has not previously been associated with measures of lung function by published EWAS. A possible explanation for this could be the reasonably modest sample sizes analysed in these studies (n = 100 and n = 172, respectively), which may be attributed to the current costs of DNA methylation sequencing assays. This highlights the strength of our pipeline in terms of leveraging summary statistics from a large-scale consortium in a 2SMR framework, thus circumventing the requirement of having DNA methylation and trait measurements in the same population. Furthermore, this allows our pipeline to evaluate associations with a wide array of traits, which again would be challenging in a one-sample setting. We note however that as larger sample sizes become available for EWAS it would be valuable to compare observational effect estimates with findings from MR analyses. This is particularly important given that observational estimates may be biased and disguise a true underlying relationship.

There was an inverse relationship between prenatal smoke exposure and lung function potentially mediated by epigenetic variation at ASPSCR1 and REST/POL2RB, which is in line with evidence from the literature concerning exposure to tobacco smoke early in the life course and later-life lung function. We also observed consistent directions of effect and associations between DNA methylation and lung function at these loci in two separate populations. Interestingly, the association observed using individuals from the ALSPAC cohort was stronger when using measures of lung function from the age 15 timepoint compared with the age 8 one, despite having a smaller sample size (n = 3852 and 3240 vs n = 5552 and 4644 respectively). This implies that these associations are unlikely to be attributed to an in utero effect due to maternal smoke exposure. Furthermore, it may be that the most parsimonious explanation for these results is that both genetics and risk factors such as prenatal smoke exposure collectively influence traits via changes in DNA methylation. That being said, this approach should still prove useful for future studies interested in identifying in utero effects, although as demonstrated by this study we advise using a cohort of young individuals to evaluate findings. Identifying specific epigenetic mechanisms that can explain in utero effects is a challenging but potentially extremely rewarding area of research.

The association signal we detected in this study at cg25313468 corresponds to a CpG site that resides near the promoter region of the REST gene. However, the strongest evidence of co-localization was detected with the expression of the neighbouring POL2RB gene, along with DNA methylation at this CpG site and FEV1. The functional gene responsible for this putative association is therefore unclear, as is whether there may be in fact multiple genes at this locus whose transcription may be influenced by potential changes in DNA methylation due to prenatal smoke exposure. The REST gene encodes the RE1-silencing transcription factor that has been reported to be a transcriptional repressor. Although this has been principally linked to neuronal genes in non-neuronal tissues, there is also previous evidence suggesting that REST functions as a master regulator of gene repression in response to hypoxia (deprivation of adequate oxygen reaching tissues). There is also previous evidence that maternal RE1-silencing transcription factor regulates the expression of 158 genes, which may have implications in both development and long-term phenotypes. In contrast, evidence implicating the POL2RB gene at this locus is less clear, although epigenetic regulation of POL2RA has been previously linked with childhood wheezing due to maternal stress.

At the other CpG site with evidence of association in this study, concerning DNA methylation at cg06105699, there was evidence of colocalization with two nearby genes (ASPSCR1 and LRR45). ASPSCR1 is a candidate gene for alveolar soft part sarcoma (ASPS), a rare type of soft tissue sarcoma that typically occurs in young patients and often spreads to the lungs, brain and bone. There is evidence to suggest that this is due to oncogenic rearrangement with the transcription factor gene TFE3. The function of LRRCA5 is less clear, although it is known to play a role in centrosome cohesion. Further analyses are therefore required to separate co-expression between these proximal genes to better understand the potential effect at this locus.

Our proposed split sample analysis using UK Biobank data should prove useful for future studies wishing to evaluate putative associations for certain exposures, such as prenatal smoking that was investigated in this study. Furthermore, exploring associations between an exposure
and outcome across multiple timepoints can enhance understanding of temporal effects. In this study we undertook these types of analyses to demonstrate that associations are more likely to be attributed to own smoking (or possibly environmental smoke exposure) rather than an in utero effect. However, we appreciate that the feasibility of undertaking these analyses in future studies may depend on the accessibility of appropriate datasets in large sample sizes. As such, design choices should be carefully considered before applying our pipeline to investigate a research question. Longitudinal cohorts such as ALSPAC can be extremely useful in terms of exploring the temporality of effects, whereas richly phenotyped resources such as UK Biobank are powerful for independent validation. We also undertook some power calculations based on the effect sizes observed in our own study regarding lung function traits (which typically ranged from 0.03 to 0.05 standard deviation increase in outcome per standard deviation change in exposure). These results suggest that harnessing summary statistics from biobank-scale datasets considerably helps in achieving adequate statistical power for the types of analyses proposed in our study (Supplementary Figure 2, available as Supplementary data at IJE online).

A limitation of this study is the lack of tissue-specific data at birth, which is why we have used DNA methylation data derived from whole blood. Incorporating DNA methylation and gene expression data from different tissue types should therefore improve this analysis pipeline’s potential to help elucidate epigenetic mechanisms in disease, as both are known to be tissue-specific. This is the phenomenon whereby a gene’s function may be restricted to certain tissue types that may be relevant for a given disease. This is important as DNA methylation data derived from cord blood may only be acting as a proxy for a more relevant tissue type. For example, in this study this could be lung tissue given the associations we detected with measures of lung function. In-depth tissue-specific evaluations of the association signals detected in whole blood would therefore be worthwhile once such datasets concerning DNA methylation become available. Added complexities (such as multiple testing) when analysing various tissue types will need to be carefully considered in these study designs. However, as sample sizes for publicly available molecular datasets increase, so too will the necessary power required to detect associations similar to the two CpG sites highlighted in our study. This should facilitate mechanistic insight into the causal pathway between risk factors and later life outcomes, as well as disentangling co-expression between neighbouring genes, which is another current limitation in the field when evaluating signals. However, in the meantime there is increasing evidence that mQTL derived from blood act as a surprisingly adequate proxy for other tissue types.

The approach demonstrated by this study should prove valuable in prioritizing genes to investigate the biological mechanisms that may explain associations between modifiable risk factors and disease susceptibility. The rewards for this challenging endeavour are promising, as it will allow us to use epigenetic factors as an indicator for early disease prognosis and subsequently improve health care and treatment.

Supplementary data
Supplementary data are available at IJE online.

Funding
This work was supported by the Integrative Epidemiology Unit which receives funding from the UK Medical Research Council and the University of Bristol (MC_UU_00011/1 and MC_UU_00011/5). This work was also supported by CRUK (grant number C18281/A19169) and the ESRC (grant number ES/N000498/1). T.G.R. is a UKRI Innovation Research Fellow (MR/S003886/1). G.H. is supported by the Wellcome Trust [208806/Z/17/Z].

Acknowledgements
We are extremely grateful to all the families who took part in this study, the midwives for their help in recruiting them and the whole ALSPAC team, which includes interviewers, computer and laboratory technicians, clerical workers, research scientists, volunteers, managers, receptionists and nurses. The UK Medical Research Council and Wellcome (Grant ref: 102215/2/13/2) and the University of Bristol provide core support for ALSPAC. GWAS data were generated by Sample Logistics and Genotyping Facilities at the Wellcome Trust Sanger Institute and LabCorp (Laboratory Corporation of America) using support from 23andMe. Methylation data in the ALSPAC cohort were generated as part of the UK BBSRC funded (BB/I025751/1 and BB/I025263/1) Accessible Resource for Integrated Epigenomic Studies (ARIES). UK Biobank data were analysed as part of projects 8786 and 15825. This publication is the work of the authors and T.G.R. will serve as guarantor for the contents of this paper.

Conflict of interest: None declared.

References
1. Zhang Y, Wilson R, Heiss J et al. DNA methylation signatures in peripheral blood strongly predict all-cause mortality. Nat Commun 2017;8:14617.
2. Feinberg AP, Koldobskiy MA, Gondor A. Epigenetic modulators, modifiers and mediators in cancer aetiology and progression. Nat Rev Genet 2016;17:284–99.
3. Richmond RC, Al-Amin A, Smith GD, Relton CL. Approaches for drawing causal inferences from epidemiological birth cohorts: a review. Early Hum Dev 2014;90:769–80.
4. Hande DE, Castro R, Loscalzo J. Epigenetic modifications: basic mechanisms and role in cardiovascular disease. Circulation 2011;123:2145–56.
5. Joubert BR, Felix JF, Yousef P et al. DNA methylation in newborns and maternal smoking in pregnancy: genome-wide consortium meta-analysis. Am J Hum Genet 2016;98:680–96.

6. Joubert BR, den Dekker HT, Felix JF et al. Maternal plasma folate impacts differential DNA methylation in an epigenome-wide meta-analysis of newborns. Nat Commun 2016;7. doi: 10.1038/ncomms10577.

7. Wahl S, Drong A, Lehe B et al. Epigenome-wide association study of body mass index, and the adverse outcomes of adiposity. Nature 2017;541:81–86.

8. Sharp GC, Salas LA, Monnereau C et al. Maternal BMI at the start of pregnancy and offspring epigenome-wide DNA methylation: findings from the pregnancy and childhood epigenetics (PACE) consortium. Hum Mol Genet 2017;26:4067–85.

9. Kulik M, Heath S, Bibikova M et al. Epigenomic analysis detects widespread gene-body DNA hypomethylation in chronic lymphocytic leukemia. Nat Genet 2012;44:1236–42.

10. Grundberg E, Meduri E, Sandling JK et al. Global analysis of DNA methylation variation in adipose tissue from twins reveals links to disease-associated variants in distal regulatory elements. Am J Hum Genet 2013;93:876–90.

11. Karlsson Linner R, Marioni RE, Rietveld CA et al. Epigenome-wide analysis detects widespread gene-body DNA hypomethylation in chronic lymphocytic leukemia. Nat Genet 2012;44:1236–42.

12. Richmond RC, Relton CL, Davey Smith G. What evidence is required to suggest that DNA methylation mediates the association between prenatal famine exposure and adulthood disease? Sci Adv 2018;4: eaao4364.

13. Richmond RC, Relton CL, Davey Smith G. What evidence is required to suggest that DNA methylation mediates the association between prenatal famine exposure and adulthood disease? Sci Adv 2018;4: eaao4364.

14. Davey Smith G, Ebrahim S. ‘Mendelian randomization’: can genetic epidemiology contribute to understanding environmental determinants of disease? Int J Epidemiol 2003;32:1–22.

15. Davey Smith G, Hemani G. Mendelian randomization: genetic anchors for causal inference in epidemiological studies. Hum Mol Genet 2014;23:R89–98.

16. Relton CL, Davey Smith G. Mendelian randomization: applications and limitations in epigenetic studies. Epigenomics 2015;7:1239–43.

17. Relton CL, Gaunt T, McAredle W et al. Data resource profile: accessible resource for integrated epigenomic studies (ARIES). Int J Epidemiol 2015;44:1181–90.

18. Gaunt TR, Shihab HA, Hemani G et al. Systematic identification of genetic influences on methylation across the human life course. Genome Biol 2016;17:61. doi: 10.1186/s13059-016-0926-z.

19. Relton CL, Davey Smith G. Two-step epigenetic Mendelian randomization: a strategy for establishing the causal role of epigenetic processes in pathways to disease. Int J Epidemiol 2012;41:161–76.

20. Lee KW, Pausova Z. Cigarette smoking and DNA methylation. Front Genet 2013;4:132.

21. Jochanes R, Just AC, Marioni RE et al. Epigenetic signatures of cigarette smoking. Circ Cardiovasc Genet 2016;9:436–47.

22. Richmond RC, Simpkin AJ, Woodward G et al. Prenatal exposure to maternal smoking and offspring DNA methylation across the life course: findings from the Avon Longitudinal Study of Parents and Children (ALSPAC). Hum Mol Genet 2015;24:2201–17.

23. Boyd A, Golding J, Macleod J et al. Cohort profile: the ‘children of the 90s’—the index offspring of the Avon Longitudinal Study of Parents and Children. Int J Epidemiol 2013;42:111–27.

24. Fraser A, Macdonald-Wallis C, Tilling K et al. Cohort profile: the avon longitudinal study of parents and children: ALSPAC mothers cohort. Int J Epidemiol 2013;42:97–110.

25. Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait analysis. Am J Hum Genet 2011;88:76–82.

26. Yang J, Ferreira T, Morris AP et al. Conditional and joint multiple-SNP analysis of GWAS summary statistics identifies additional variants influencing complex traits. Nat Genet 2012;44:369–75, S1–3.

27. Hemani G, Zheng J, Elsworth B et al. The MR-Base platform supports systematic causal inference across the human phenome. Elife 2018;7. doi: 10.7554/elife.34408.

28. Neale lab. Rapid GWAS of Thousands of Phenotypes for 337,000 Samples in the UK Biobank. 2017. http://www.neale.lab.is/blog/2017/7/19/rapid-gwas-of-thousands-of-phenotypes-for-337000-samples-in-the-uk-biobank (19 July 2017, date last accessed).

29. 1000 Genomes Project, Abecasis GR, Altshuler D et al. A map of human genome variation from population-scale sequencing. Nature 2010;467:1061–73.

30. Wald A. The fitting of straight lines if both variables are subject to error. Ann Math Statist 1940;11:284–300.

31. Bowden J, Davey Smith G, Haycock PC, Burgess S. Consistent estimation in mendelian randomization with some invalid instruments using a weighted median estimator. Genet Epidemiol 2016;40:304–14.

32. Burgess S, Butterworth A, Thompson SG. Mendelian randomization analysis with multiple genetic variants using summarized data. Genet Epidemiol 2013;37:638–65.

33. Richardson TG, Zheng J, Davey Smith G et al. Mendelian randomization analysis identifies CpG sites as putative mediators for genetic influences on cardiovascular disease risk. Am J Hum Genet 2017;101:590–602.

34. Richardson TG, Haycock PC, Zheng J et al. Systematic Mendelian randomization framework elucidates hundreds of CpG sites which may mediate the influence of genetic variants on disease. Hum Mol Genet 2018;27:3293–304.

35. Giambartolomei C, Zhenli Liu J, Zhang W et al. A Bayesian framework for multiple trait colocalization from summary association statistics. Bioinformatics 2018;34:2538–45.

36. Carithers LJ, Moore HM. The genotype-tissue expression (GTEx) project. Biopreserv Biobank 2015;13:307–08.

37. Hemani G, Tilling K, Davey Smith G. Orienting the causal relationship between imprecisely measured traits using GWAS summary data. PLoS Genet 2017;13:e1007081.

38. Bonder MJ, Luijk R, Zhernakova DV et al. Disease variants alter transcription factor levels and methylation of their binding sites. Nat Genet 2017;49:131–38.

39. Sudlow C, Gallacher J, Allen N et al. UK biobank: an open access resource for identifying the causes of a wide range of complex diseases of middle and old age. PLoS Med 2015;12:e1001779.

40. Brion MJ, Shakhbazov K, Visscher PM. Calculating statistical power in Mendelian randomization studies. Int J Epidemiol 2013;42:1497–501.
41. Lee MK, Hong Y, Kim SY, Kim WJ, London SJ. Epigenome-wide association study of chronic obstructive pulmonary disease and lung function in Koreans. *Epigenomics* 2017;9:971–84.

42. Bell JT, Tsai PC, Yang TP et al. Epigenome-wide scans identify differentially methylated regions for age and age-related phenotypes in a healthy ageing population. *PLoS Genet* 2012;8:e1002629.

43. Lodrup Carlsen KC, Jaakkola JJ, Nafstad P, Carlsen KH. In utero exposure to cigarette smoking influences lung function at birth. *Eur Respir J* 1997;10:1774–79.

44. Rehan VK, Asotra K, Torday JS. The effects of smoking on the developing lung: insights from a biologic model for lung development, homeostasis, and repair. *Lung* 2009;187:281–89.

45. Horsthemke B. A critical view on transgenerational epigenetic inheritance in humans. *Nat Commun* 2018;9:2973.

46. Chong JA, Tapia-Ramirez J, Kim S et al. REST: a mammalian silencer protein that restricts sodium channel gene expression to neurons. *Cell* 1995;80:949–57.

47. Schoenherr CJ, Anderson DJ. The neuron-restrictive silencer factor (NRSF): a coordinate repressor of multiple neuron-specific genes. *Science* 1995;267:1360–63.

48. Cavadas MA, Mesnieres M, Crifo B et al. REST is a hypoxia-responsive transcriptional repressor. *Sci Rep* 2016;6. doi: 10.1038/srep31355.

49. Moravec CE, Samuel J, Weng W, Wood IC, Sirotkin HL. Maternal Rest/Nrsf regulates zebrafish behavior through snap25a/b. *J Neurosci* 2016;36:9407–19.

50. Trump S, Bieg M, Gu Z et al. Prenatal maternal stress and wheeze in children: novel insights into epigenetic regulation. *Sci Rep* 2016;6. doi: 10.1038/srep28616.

51. Folpe AL, Deyrup AT. Alveolar soft-part sarcoma: a review and update. *J Clin Pathol* 2006;59:1127–32.

52. Hodge JC, Pearce KE, Wang X, Wiktor AE, Oliveira AM, Greipp PT. Molecular cytogenetic analysis for TFE3 rearrangement in Xp11.2 renal cell carcinoma and alveolar soft part sarcoma: validation and clinical experience with 75 cases. *Mod Pathol* 2014;27:113–27.

53. He R, Huang N, Bao Y, Zhou H, Teng J, Chen J. LRRC45 is a centrosome linker component required for centrosome cohesion. *Cell Rep* 2013;4:1100–07.

54. Sonawane AR, Platig J, Fagny M et al. Understanding tissue-specific gene regulation. *Cell Rep* 2017;21:1077–88.

55. eGTEx Project. Enhancing GTEx by bridging the gaps between genotype, gene expression, and disease. *Nat Genet* 2017;49:1664–70.

56. Qi T, Wu Y, Zeng J et al. Identifying gene targets for brain-related traits using transcriptomic and methylomic data from blood. *Nat Commun* 2018;9:2282.