PROBIOTIC CANDIDATE PROTEOLYTIC *Bacillus* sp. COLLECTED FROM MANGROVE OF MARGASARI, LAMPUNG

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PROBIOTIC CANDIDATE PROTEOLYTIC *Bacillus* sp. COLLECTED FROM MANGROVE OF MARGASARI, LAMPUNG. Intensive shrimp culture has encountered many problems, such as declining water quality through disease caused by pathogenic microbes, which affected mortality. This study aimed to determine any potential probiotic from *Bacillus* sp. collected from mangrove in East Lampung, which could be used to improve the cultured shrimps' proteolytic and probiotic activity. This is a descriptive research with sampling and data collection of bacteria from many samples of mangrove. Result shows 128 isolates *Bacillus* from which then it has arrived at five potential probiotic *Bacillus* sp. The study five *Bacillus* sp. has been isolated with potential properties for probiotic (KPP212, IP121, UJ131, UJ132, SB141). Each isolate has characteristics with proteolytic property, growth in a wide range of pH 4–10 and osmotic stress (0–6% NaCl), non-pathogenic, ability for glucose fermentation, non-motile, and has negative catalase activity. The five potential *Bacillus* sp. can be used as probiotics for shrimp farming.

Keywords: Characterisation, *Bacillus* sp., mangrove, probiotic

KANDIDAT PROBIOTIK PADA *Bacillus* sp. PROTEOLITIK DIKOLEKSI DARI HUTAN MANGROVE DI MARGASARI, LAMPUNG. Budidaya udang dengan cara intensif telah menyebabkan banyak masalah seperti penurunan kualitas air yang mengakibatkan munculnya mikroba patogen, sehingga mempengaruhi kematian. Tujuan dari penelitian ini adalah untuk menemukan *Bacillus* sp. yang diisolasi dari beberapa sampel di mangrove dan mengkarakterisasi mikroba terisolasi tersebut untuk penggunaan probiotik. Studi ini merupakan penelitian deskriptif dan koleksi data bakteri dari berbagai jenis sampel mangrove. Dari penelitian ditemukan lima *Bacillus* sp. yang terisolasi dan memiliki sifat potensial untuk probiotik, yaitu KPP212, IP121, UJ131, UJ132, dan SB141, dengan masing-masing isolat memiliki karakteristik bersifat proteolitik, pertumbuhan dalam rentang pH yang luas (4–10) dan taban pada tekanan osmotik (0–6% NaCl), tidak patogen, kemampuan untuk fermentasi glukosa, tidak motil, serta tidak memiliki aktivitas katalase. Kelima bakteri *Bacillus* sp. tersebut dapat digunakan untuk probiotik pada budidaya udang.

Kata kunci: Karakterisasi, *Bacillus* sp., mangrove, probiotik

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I. INTRODUCTION

Mangrove of Margasari Village in the district of Labuhan Maringgai, East Lampung Regency is in 5°51'84" South Latitude–105°64'84" East Longitude covers about 700 hectares which is 6.65% of the total mangrove in Lampung Province. Mangrove has functions ecologically and economically, such as protecting coastal abrasion, brackish water quality control, habitat for many organisms, medicines, and paper pulp. The presence of waste influences mangrove’s ecological activities, decomposition by microorganisms, mineral uptaking by plants, and other biological activities to keep the equilibrium (Kementerian Kehutanan, 2014; Kariada & Andin, 2014). Furthermore, it is necessary to rehabilitate mangrove for further conservation of coastal areas.

Many researchers in various places have also isolated the bacteria potential from mangrove. The study of Deivanai, Bindusara, Prabhakaran, and Bhore (2014) isolated Pantoea ananatis (1MSE1) and Bacillus amyloliquefaciens (3MPE1) bacteria from mangrove, which is interacted positively with rice seedlings, provides significant increase in root and shoot length, fresh weight, and chlorophyll content. The study of Castro et al. (2018) proved that Enterobacter sp. MCR1.48 strain from mangrove endophyte effectively promotes the Acacia polyphylla growth and fitness. The bacteria can be used in the seedling production of the tree. Another researcher, Maulani, Rasi, and Zulkifli (2019), has successfully isolated eighteen endophytic bacteria from mangrove Rhizophora mucronata from Gili Sulat, East Lombok. The 18 isolates of endophytic consist of 15 isolates: Gram-positive bacteria and 3 isolates were Gram-negative bacteria. The endophytic bacteria isolate that had antibiotic activity was B. cereus, P. aeruginosa, S. aureus, and E. coli. On the other hand, the potential fungi from mangrove were also discovered. Hamzah, Lee, Hidayat, Terhem, Hanum, and Mohamed (2018) in Malaysia isolated endophytic fungi from mangrove Rhizophora mucronata. The study found several fungi, i.e. Alternaria, Fusarium, Nigrospora, Pestalotiopsis, Phoma, and Xylaria. After their culture assay for their antagonism activities with the phytopathogenic fungus, Fusarium solani reached 45–66%.

Mangrove plays an important role in the coastal waters, food web and habitat, supporting many different biotas, such as fish, crabs, shrimps, and molluses (Scharler, 2011). Around the Margasari mangrove, there are shrimp farmings. The mangrove has many wastes, which is good nutrition for various bacterial growths. In nature, the bacteria can pass into the digestive tract of animals. Several types of bacteria found in the digestive tract of animals had an important role in improving food and fish’s healthy utilisation (Sarastiti, Suminto, & Sarjito, 2020). Bacillus sp. can be one of the bacteria that can increase the digestibility of fish or shrimp, and it has the potential as a probiotic (Anggriani, Iskandar, & Ankiq, 2012). There was no information related to the isolation of Bacillus sp. from mangrove of Margasari, East Lampung, especially related to those used for probiotics.

The advantage of probiotic technology is that the process is natural and safe. Probiotics had a beneficial effect which includes interacting directly with commensal and pathogenic microorganisms. The probiotics were used for many functions, i.e. (1) to prevent and treat infections, (2) to improve the balance of microorganisms in the small intestine, (3) to produce extracellular enzymes, and (4) to produce beneficial compounds such as vitamins and short-chain fatty acids (Tensiska, 2008).

Isolation and selection of proteolytic bacteria have very good potential to be used as probiotics. The character of the bacteria is needed to increase feed efficiency. The study reported that shrimp and fish feed contained 55.51–67.68% protein (Handayani, 2011).
Other studies have also shown some other microorganisms as probiotic candidates, and they were *Bacillus subtilis*, *Bacillus licheniformis*, *Pseudomonas putida*, *Bacillus bataviensis*, and *Canlobacter* sp. (Rahmawan, Mohamad, Suminto, & Herawati, 2014; Seprianto, Feliatra, & Nugroho, 2017).

Shrimp cultures have been widely growing in Lampung Province, in which intensive shrimp culture deliberates some diseases related to its culture (Taukhid, Supriyadi, & Koesharyani, 2008). To prevent or regulate the disease, therefore, the development of probiotic is necessary. Samosir, Suryanto, and Desrita (2017) established one by developing probiotic from the surrounding ecosystem. Many isolated bacteria with potential sources of probiotic can be collected from mangrove communities, such as from mangrove plant (its root and bark), fishes like milky fish, cuttlefish, mangrove shrimps, molluscs, crabs, and other fishes, as well as from the mud, water and other abiotic factors of the mangrove ecosystem (Pratiwi, Rahayu, & Wahju, 2013; Seprianto, Feliatra, & Nugroho, 2017; Muliani, Nurbaya, Arifudin, & Muharijadi, 2017). Therefore, it is necessary to figure out any potential of the proteolytic *Bacillus* sp. as a local probiotic is collected from the mangrove of Margasari. It could then be applied to the aquaculture digestion, and disease problems, especially in shrimp and fish cultures encountered most in Lampung Province.

**II. MATERIAL AND METHOD**

**A. Sources of Isolated Bacteria**

The sample used to isolate bacteria was collected from mangrove communities in Margasari Village, Labuhan Maringgai, East Lampung Regency. Abiotic and biotic samples were collected from water, mud, fishes, shrimps, crabs, molluscs and cuttlefish found in the mangrove ecosystem, as well as the *Rhizophora* sp. bark and roots.

**B. Isolation of Bacillus sp.**

Isolation of *Bacillus* sp. was done from different sample mass, 10 g of mud, 1 g mangrove root and skin, 1 ml of water, 1 ml intestinal suspension of shrimp, crab, mollusc, and fish (Figure 1). Sample suspension then was made by adding 90 ml physiological salt for mud and 9 ml for water, mangrove root and bark,

![Figure 1. Collected biotic samples](image_url)

Remarks: a. *Penaeus merguiensis* (udang jerbung), b. *Litopenaeus vannamei* (udang kucing) c. *Mugil* sp. (ikan belanak) d. *Mallotus villosus* (ikan kepala batu) e. Epidermis of *Rhizophora* sp. (kulit bakau), f. Root of *Rhizophora* sp. (akar bakau), g. *Scylla serrata* (kepiting bakau) h. *Sepia latimanus* (sotong), i. *Tebunaloa tobi* (ikan pirit), j. *Telescopium telescopium* (siput bakau) k. *Nerita violacea* (keong bulat), and l. *Episesarma* sp. (kepiting pemanjat pohon) and abiotic samples, m. mud, and n. water
shrimp, mollusc, crab and fish. All samples were homogenised by using a vortex mixer at 80°C for 15 minutes. Dilution was made for each sample in series of 10-1 and 10-2. One ml of each was diluted into sample suspension and it was spread into skim milk agar media modification of Sea Water Complete (SWC), followed by incubation for 24 hours at 37°C (Hamtini, 2014). The isolate then was purified by quadrant streak into the SWC agar media.

C. Proteolytic Test

_Bacillus_ sp. isolate was picked using a sterile ose needle and inoculated into SWC media modified with skim milk. The culture then was incubated for 24 hours at 37°C. The observation was made by determining the formed proteolytic index (Hamtini, 2014, Hapsari, Tjahjaningsih, Alamsjah, & Pramono, 2016; Sumardi, Agustrina, Ekowati, & Pasaribu, 2018).

D. Osmotic/Salinity Stress Test

_Bacillus_ sp. isolate was picked using a sterile ose needle and inoculated into modified SWC media with NaCl concentration of 0%, 3%, and 6%. The culture then was incubated for 24 hours at 37°C. The observation was made by determining the number of growing colonies (Subagiyo, Sebastian, Triyanto, & Wilis, 2015).

E. pH-stress Test

_Bacillus_ sp. isolate was picked using a sterile ose needle and inoculated into modified SWC media with pH 4, 7, and 10. The culture then was incubated for 24 hours at 37°C. The observation was made by determining the measurement of growing colonies (Kepel, Widdhi, & Fatimavali, 2020).

F. Pathogenetic Test

_Bacillus_ sp. isolate was picked using a sterile ose needle and inoculated into blood modified SWC media. The culture then was incubated for 24 hours at room temperature (Hamtini, 2014). The observation was made by determining the hemolytic ability of isolate from the change of colours (Figure 2).

G. Characteristic Test

Characterisation of isolate bacteria, presumably _Bacillus_ sp. was done in 2 steps, as follows:

a. Morphological characterisation of colony and cell. Colony characterisation was done by observing the colony formed while cell morphology was made on gram smear (Yulvizar, 2013).

b. Biochemical test

Biochemical test as characterisation of the colony was conducted in different tests, such as catalase, mortality and glucose fermentation.

b.1. Catalase test

Two drops of H_2O_2 was placed in the sterile glass. Then, one ose needle picked of isolate _Bacillus_ sp. was mixed into H_2O_2 in object glass (Yulvizar, 2013).

b.2. Motility test

As much as 1 ose needle isolate _Bacillus_ sp. was placed into SWC agar media SWC. The culture then was incubated for 24 hours at

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**Figure 2.** Hemolysis test in blood agar

Remarks: a. Beta hemolysis, b. Alpha hemolysis, c. Gamma hemolysis (Aryal, 2015)
37°C temperature (Samosir, Suryanto, & Desrita, 2017).

b.3. Glucose fermentation test
As much as 1 ose needle isolate Bacillus sp. was inoculated into liquid SWC agar media and added 1% sugar (glucose, lactose, mannitol, sucrose and mannose). The culture then was incubated for 24 hours at 37°C temperature (Samosir et al., 2017).

III. RESULT AND DISCUSSION

A. Margasari Mangrove
Margasari mangrove is very important for the coastal area of East Lampung. Most of the shrimp or fish cultures/ponds are found in the mangrove belt of East Lampung. It is known that mangrove can support many biotas, including fish, shrimps, molluscs (Figure 1). Margasari mangrove and other mangrove ecosystems provide food for many aquatic biotas and contribute to the biological cycle in coastal waters. With special structures of mangrove plants, such as Rhizophora mucronata, Avicennia marina, Sonneratia alba, and others, the mangrove's ground floor and swamp and water flow within the mangrove connecting provides shelter for many larvae of mangrove biota.

Spawning and nursery become the shelter of many biotas provided by mangrove, and it is affected by microbes activity as a decomposer. These microbes possibly believed to have beneficial uses such as probiotic, antibiotic, and bioactive products and else (Subagiyo, Muhammad, & Wilis, 2017). The recent study also indicated that some bacterial colony could be found in the mud. Another study also found seven isolated bacteria collected from mangrove mud of Wonorejo, Rungkut, Surabaya (Pratiwi, Rahayu, & Wahju, 2013). This diversity found in bacterial colonies was possible since the mangrove area was fully covered by plant debris that can be degraded and used by microorganisms as energy sources (Sinatryani, Moch, Sudarno, & Kustiawan, 2014).

Fourteen bacterial isolation was found from squids which were the highest number of bacterial isolations found among others. Some studies also indicated that four different probiotic isolation was found from carpio fish intestine (Samosir, Suryanto, & Desrita, 2017), while other studies were able to isolate 16 probiotic colonies from shrimp intestine (Febrianti, 2011). Isolation of this probiotic colony mostly was from the intestine/gut of the animal samples. Most of these variety microorganisms play an important role in the digestive system, like produced enzymes (Sarastiti, Suminto, & Sarjito, 2020). In addition, the existing variety of microorganism was also able to compete with the growth of pathogenic bacteria and presumably increased animal immunity. With the normal digestive process, growth was affected, and the animal’s development was the bacterial host (Samosir et al., 2017).

While those in plant parts, such as roots and skins of mangrove plants, 5 and 6 isolated
bacteria were found from each, the roots and skins of the mangrove plants were used since these parts of mangrove plants had to contact with mud and water, which presumably also contained bacteria which can be isolated, and had different characteristics with other bacterial colony found from animal samples. Yet, they had potential characteristic as probiotic candidates.

C. Cell Morphology

The morphology of isolate is correct bacteria, had a similarity, yet only the isolated colony's edge had a different shape (Table 2). In contrast, the cell morphology indicated the same shape and Gram stain, bacillus and positive Gram (Table 2 and Figure 3). Positive Gram stain was indicated by the violet colour of the bacterial cell. The crystal violet was trapped in the thick cellular wall of bacteria with one layer membrane in which bacteria underwent dehydration and shrink after exposure to 96% alcohol (Samosir, Suryanto, & Desrita, 2017).

D. Proteolytic Test

Many different tests were given to the isolated bacteria colony. This caused drastic selection among them. Before the selection was given, 128 isolated bacteria were found, then, with proteolytic selection, 94 isolates were collected. Further selection was made based on the ability to deal with salinity and pH, the Bacillus sp. isolates were reduced to 27 isolates.

### Table 1. Number of *Bacillus* sp. from isolation and selection tests

| Source       | Type of Isolat | Protease Test | Osmotic Stress Test | Pathogen Test |
|--------------|----------------|---------------|---------------------|---------------|
|              | Non Proteolytic | Proteolytic | No growth on pH and salinity stress tests | Growth on pH and salinity stress tests | Pathogen | Non Pathogen |
| Water        | 12             | 1            | 11                  | 7             | 4           | 4           | 0           |
| Mud          | 22             | 4            | 18                  | 16            | 2           | 2           | 0           |
| *Sepia latimanus* | 14           | 9            | 5                   | 4             | 1           | 1           | 0           |
| *Epinesarma* sp.  | 11           | 5            | 6                   | 3             | 3           | 2           | 1           |
| *Scylla serrata*  | 13           | 2            | 11                  | 9             | 2           | 2           | 0           |
| *Tenuacta tibi*   | 8             | 2            | 6                   | 4             | 2           | 1           | 1           |
| *Penaeus merguiensis* | 8            | 2            | 6                   | 3             | 3           | 1           | 2           |
| *Telecopium telecopium* | 7            | 1            | 6                   | 2             | 4           | 3           | 1           |
| *Nerita violacea* | 7             | 0            | 7                   | 5             | 2           | 2           | 0           |
| *Lophopinus vannamii* | 5            | 2            | 3                   | 2             | 1           | 1           | 0           |
| *Rhizopora* sp. root | 5            | 2            | 3                   | 2             | 1           | 1           | 0           |
| *Rhizopora* sp. bark | 6            | 1            | 5                   | 4             | 1           | 1           | 0           |
| *Mallotus villosus* | 7             | 3            | 4                   | 3             | 1           | 1           | 0           |
| *Mugil* sp. | 3             | 0            | 3                   | 3             | 0           | 0           | 0           |
| **Number**   | **128**       | **34**       | **94**              | **67**        | **27**      | **22**      | **5**       |

### Table 2. Morphology of isolat *Bacillus* sp. for the probiotic candidate

| Isolat    | Colony | Cell Morphology | Form | Edges   | Elevation | Colors | Gram | Form |
|-----------|--------|-----------------|------|---------|-----------|--------|------|------|
| KPP212    | Circular | Raised       | Entire | White  | + | Basil |
| IPI121    | Circular | Convex       | Entire | White  | + | Basil |
| UJ131     | Circular | Flat        | Entire | White  | + | Basil |
| UJ132     | Circular | Convex       | Entire | White  | + | Basil |
| SB141     | Irregular | Raised | Lobate  | White  | + | Basil |
The final selection was made for their ability to be pathogenic to ensure that the bacterial colony caused no harm if they were given probiotics. Characteristics of Bacillus sp. could be seen in Table 3.

The protein contained in shrimp feeds is approximately 30-40%. Therefore, proteolytic test was necessarily performed. All the isolated probiotic candidates indicated their ability to degrade casein from media, indicated by clear zone as hydrolysis process surrounding the isolated colony (Figure 3). This ability was also indicated by those colonies of probiotic candidates obtained from other studies (Samosir et al., 2017). The proteolytic ability of Bacillus sp. occurred since the bacteria produced protease (Hamtini, 2014). Protease, as the extracellular enzyme of Bacillus sp. can break the peptide bond of protein into oligopeptida and amino acids (Ilmiah, Nisa, & Budiasih, 2018).

A study on the potential of proteolytic bacteria from mangroves also has been observed by other researchers. Utomo et al. (2019) succeeded in observing the protease enzyme-producing bacteria from the mangrove of Gunung Anyar, Surabaya. Two speices of bacteria was obtained, namely Yersinia enterocolotica and Enterobacter agglomerans. The bacteria have been characterised as the proteolytic enzyme. Other researchers, Castro et al. (2014), isolated endophytic microorganisms from two mangrove species, Rhizophora mangle and Avicennia nitida. They found that mangrove microorganisms demonstrated a diverse range of enzymatic activities. The isolates produced enzymes of amylase, esterase, lipase, protease, and endoglucanase. In this study Bacillus sp. of proteolytic and non-pathogenic was observed as a potential probiotic.

All the collected isolate probiotic candidates showed different proteolytic index. The isolate bacteria with the highest proteolytic index was in code SB141, while the lowest was in code IP121. This different proteolytic index can be seen in each isolate bacteria's clear zone (Figure 4).
Salinity and pH tests were conducted to elucidate selected isolate probiotic candidates’ ability from different salinity and pH stresses of media. Isolate with codes of KPP212, IPI21, UJ131, UJ132, and SB141 survived and grew in salinity of 0%, 3%, and 6% and with pH of 4, 7, and 10.

**E. Bacteria Characterisation**

The five isolated bacteria produced protease and survived in a different range of pH (4–10) and different salinity (0–6%) (Figure 5); meanwhile, their hemolysis activity was in gamma hemolysis or non-pathogenic activity. All the isolated ones also indicated non-motile, and only isolated with code UJ132 had positive catalase activity from biochemical tests. In contrast, in the sugar fermentation test, all isolated ones had positive results in all different types of sugar used for the test.

The ideal of probiotic is survival in many different stress conditions; therefore survival test was conducted. Isolates of KPP212, IPI21, UJ131, UJ132 and SB141 indicated that they
can survive and grew very well in pH and salinity stress (Figure 4).

This ability to survive was similar to those found by another study (Triyanto, Isnansetyo, Prijambada, Widada, & Tarmiawati, 2009) was isolated from the mangrove’s mud. This bacteria colony's survival from different stress of pH and salinity presumably showed that this colony was used to environmental stress, unstable condition, which is very common in estuary ecosystem (Hutabarat, 2000).

Pathogenic or virulent isolates was determined by the degree of clear zone media produced by the isolates. All the isolates probiotic candidates had γ (Gamma) hemolysis characteristic (Table 4). Blood agar as differentiates media was used to determine bacteria’s ability to lyse red blood cells (RBCs) (Hamtini, 2014). The ability of bacteria to lyse RBCs was done by extra-cellular protein produce called haemolysin (Khusnan, Dwi, & Agus, 2018). Pathogenetic in RBCs was defined into three levels, alpha hemolysis, beta hemolysis and gamma hemolysis. Alpha hemolysis occurred when RBCs and hemoglobin were partly lyzed, beta hemolysis occurred when all RBCs and hemoglobin were lyzed, causing the surrounding media to clear. Gamma hemolysis occurred when there was no lysis for both RBCs and hemoglobin, causing no colour change in media (Hamtini, 2014).

In the sugar fermentation test, all of the isolates colony indicated positive fermentation test for sugars such as lactose, mannose, mannitol, glucose and sucrose (Table 4), indicated by the formation of yellow colour media. The change in media colour occurred since fermentation caused acidity of the media in which, by using phenol red as an indicator, it turned to yellow. Acids released in media was produced from the breaking down of sugar by bacteria. In the motility test, all isolates indicated negative results, shown by the bacteria colony’s undispersed growth in their media (Damayanti, Oom, & Effendi, 2018).

A catalase test was done to determine the ability of the isolates colony to produce catalase enzyme. A positive result was shown from the UJ132 isolate, while KPP212, IP121, UJ131, and SB141 isolates indicated negative results (Table 4). A positive result was indicated by an oxygen bulb from mixing of H₂O₂ with isolate bacteria, indicating that catalase enzyme was produced by bacteria and used to break hydrogen peroxide in water and oxygen. Hydrogen peroxide was a compound that interferes with intracellular enzyme activity (Yulvizar, 2013).

Table 4. Bacteria isolated characterisation

| Isolat  | Pr | Stress on pH | Stress on Salinity (%) | Mo | K | Sugar fermentation | Pa | Hemolysis |
|---------|----|--------------|-------------------------|----|---|---------------------|----|-----------|
|         |    | 4 | 7 | 10 | 0 | 3 | 6 | | | Ms | G | S | L | Mt | α | β | γ |
| KPP212  | +  | + | + | + | + | + | + | - | - | + | + | + | + | + | - | - | + |
| IP121   | +  | + | + | + | + | + | + | - | - | + | + | + | + | + | - | - | + |
| UJ131   | +  | + | + | + | + | + | + | - | - | + | + | + | + | + | - | - | + |
| UJ132   | +  | + | + | + | + | + | + | - | - | + | + | + | + | + | - | - | + |
| SB141   | +  | + | + | + | + | + | + | - | - | + | + | + | + | + | - | - | + |

Remarks:

- = no reaction/no growth  α = partial hemolysis
+ = reaction/growth  β = total hemolysis
G = Glucose  γ = no hemolysis
K = Catalase  L = Lactose
Mo = Motility  Ms = Mannose
Mt = Mannitol  Pa = Pathogenic test
Pr = Protease  S = Sukrose
IV. CONCLUSION

This study found about 128 isolated Bacillus sp. from intestinal mangrove biota in Lampung Mangrove Center, such as shrimp, mollusc, fish, and crabs from which 94 isolated Bacillus sp. had proteolytic characters, and 5 of them have very good potentials as a probiotic candidate. The five probiotics were: Bacillus sp. KPP212 collected from climbing crab, Bacillus sp. IP121 collected from fish, Bacillus sp. SB141 collected from the mollusc, Bacillus sp. UJ131 and Bacillus sp. UJ132 collected from shrimp. Diversity of Bacillus sp. and biota are found in various types. Therefore, the mangrove of Margasari of Lampung is very important to conserve.

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