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Inflammatory and hematologic markers as predictors of severe outcomes in COVID-19 infection: A systematic review and meta-analysis

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ABSTRACT

Background: Laboratory testing is commonly performed in patients with COVID-19. Each of the laboratory parameters has potential value for risk stratification and prediction of COVID-19 outcomes. This systematic review and meta-analysis aimed to evaluate the difference between these parameters in severe and nonsevere disease and to provide the optimal cutoff value for predicting severe disease.

Method: We performed a systematic literature search through electronic databases. The variables of interest were serum procalcitonin, albumin, C-reactive protein (CRP), D-dimer, and lactate dehydrogenase (LDH) levels in each group of severity outcomes from COVID-19.

Results: There were a total of 4848 patients from 23 studies. Our meta-analysis suggest that patients with severe COVID-19 infections have higher procalcitonin, (mean difference 0.07; 95% CI 0.05–0.10; p < 0.00001), CRP (mean difference 36.88; 95% CI 29.10–44.65; p < 0.00001), D-Dimer (mean difference 0.43; 95% CI 0.31–0.56; p < 0.00001), and LDH (mean difference 102.79; 95% CI 79.10–126.49; p < 0.00001) but lower levels of albumin (mean difference −4.58; 95% CI −5.76 to −3.39; p < 0.00001) than those with nonsevere COVID-19 infections. The cutoff values for the parameters were 0.065 ng/mL for procalcitonin, 38.85 g/L for albumin, 33.55 mg/L for CRP, 0.635 μL for D-dimer, and 263.5 U/L for LDH, each with high sensitivity and specificity.

Conclusion: This meta-analysis suggests elevated procalcitonin, CRP, D-dimer, and LDH and decreased albumin can be used for predicting severe outcomes in COVID-19.

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1. Introduction

In December 2019, new emerging cases of atypical pneumonia were first reported in Wuhan, Hubei Province, China. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is the pathogen of this atypical pneumonia outbreak, called coronavirus disease 2019 (COVID-19), which targets the lower respiratory tract and other organs expressing the ACE2 receptor. Respiratory and airborne droplets are the main routes of transmission of this disease. The World Health Organization (WHO) declared this disease a public health emergency [1]. As of July 13th, 2020, a total of 13,082,304 cases of COVID-19 were recorded worldwide, with a total number of deaths reaching 572,551 [2]. Based on its features, this disease can be divided into ordinary, mild, severe, and critically ill types [3]. The initial symptoms and signs in COVID-19 patients are usually very mild, and the infection can even be asymptomatic. However, the disease can deteriorate over a short period of time (between 7 and 10 days) into acute respiratory distress syndrome (ARDS) and other multiorgan complications due to rapid viral replication and cytokine storms [4,5]. This abrupt onset of disease progression has contributed to an increase in the mortality rate of the disease. Severe comorbidities have also been demonstrated to be associated with the development of severe COVID-19, such as hypertension, diabetes mellitus, dyslipidemia, thyroid disease, cardiovascular disease, dementia, and pulmonary disease [6-9]. Therefore, prompt identification and containment, which are achievable through strict surveillance and early diagnosis, are very important.

One of the tests that physicians perform most often in the setting of COVID-19 is laboratory testing. During the detection, treatment, and follow-up of COVID-19, physicians frequently check various laboratory parameters to see the dynamic changes in each. These laboratory
parameters have been suggested for risk stratifications in COVID-19, as timely detection of disease progression is crucial for appropriate management and intervention. Currently, combinations of several laboratory tests have been used in some settings to show the hyperinflammatory state and prognostication. These combinations include the neutrophil to lymphocyte ratio (NLR) and the lymphocyte to C-reactive protein ratio (LCR) [10]. Based on the pathophysiology of severe COVID-19, which involves a hyperinflammatory state, coagulation cascade, and multiorgan dysfunction [11], several biomarkers that represent each of those conditions, such as CRP, procalcitonin, D-dimer, LDH, and albumin, might be helpful to predict the outcome of COVID-19. Unfortunately, there are still missing puzzle pieces regarding the most significant laboratory markers and the cutoff values that can differentiate severe and nonsevere outcomes. This systematic review and meta-analysis aimed to analyze the differences in several biomarkers, including serum procalcitonin, CRP, D-dimer, LDH, and albumin, in severe and nonsevere disease, as well as the cutoff value for each biomarker to predict severe outcomes in COVID-19 infection.

2. Material and methods

2.1. Eligibility criteria

We conducted a systematic review and meta-analysis study. Studies were included in this review if they met the following inclusion criteria: representation for clinical questions (P: positive/confirmed cases of COVID-19; I: a group of patients with severe COVID-19; C: a group of patients with nonscere COVID-19; O: information on laboratory parameters such as procalcitonin, albumin, C-reactive protein (CRP), D-dimer, and lactate dehydrogenase (LDH) levels), the type of study was a randomized control trial, cohort, clinical trial, case cohort, and crossover design, and if the full-text article was available. The following types of articles were excluded: articles other than original research (e.g., review articles or commentaries); case reports; articles not in the English language; articles on research in pediatric populations (17 years of age or younger); and articles on research in pregnant women.

2.2. Search strategy and study selection

We performed a systematic literature search from PubMed, PubMed Central, and Google Scholar with the search terms: ‘Characteristics’ OR ‘Laboratory parameters’ AND ‘COVID-19’ OR ‘Coronavirus disease 2019’. Duplicate results were removed. The remaining articles were independently screened for relevance by their abstracts with two authors. The full texts of residual articles were assessed according to the inclusion and exclusion criteria. The search started on July 6, 2020 and was finalized on July 13, 2020. The study was carried out according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines.

2.3. Data extraction and quality assessment

Data extraction was performed independently by two authors, and we used standardized forms that included author, year, study design, number of participants, age, sex, serum procalcitonin, albumin, CRP, D-dimer, LDH, and severe COVID-19.

The variables of interest in our meta-analysis were serum procalcitonin, serum albumin, serum CRP, serum D-dimer, and serum LDH concentrations in mean ± SD or median (interquartile range) from each group of severe COVID-19 and nonscere COVID-19 infections. Severe COVID-19 infection was defined as patients who had any of the following features at the time of or after admission: (1) respiratory distress (≥30 breaths per min); (2) oxygen saturation at rest ≤ 93%; (3) ratio of the partial pressure of arterial oxygen (PaO2) to a fractional concentration of oxygen inspired air (FiO2) ≤ 300 mmHg; or (4) critical complications (respiratory failure, septic shock, and/or multiple organ dysfunction/failure) or admission to the ICU [12].

Two investigators independently evaluated the quality of the included cohort and case-control studies using the Newcastle–Ottawa Scale (NOS) [13]. The selection, comparability, and exposure of each study were broadly assessed, and studies were assigned a score from zero to nine. Studies with scores ≥ 7 were considered of good quality. They also independently evaluated the quality of the included case-series studies using the Joanna Briggs Institute Critical Appraisal Checklist for Case Series [14].

2.4. Statistical analysis

The software programs Review Manager 5.4 (Cochrane Collaboration) and Comprehensive Meta-Analysis version 3 were used for meta-analysis. Continuous variables were calculated using the inverse-variance formula with random effects models regardless of heterogeneity. The effect estimate was reported as the mean difference (MD) and its standard deviation (SD) along with its 95% confidence interval (CI). Heterogeneity was assessed by using the I2 statistic, and values of <25%, 26–50%, and >50% were considered low, moderate, and high degrees of heterogeneity, respectively. The p-value was two-tailed, and the statistical significance was set at ≤ 0.05. When data were reported as medians and interquartile ranges, we converted them to means and standard deviations for meta-analysis pooling using the formula by Wan et al. [15] The mean values of parameters that were found to be significant in the meta-analysis were then used to generate receiver operating characteristic (ROC) curves using SPSS ver. 24 to determine the area under the curve (AUC). The optimal cutoff for parameters with a significant p-value (two-tailed) was determined using Youden’s index [16], and the corresponding sensitivity and specificity for the cutoff were also calculated. Subgroup analysis comparing prospective and retrospective studies was performed for each component of variable of interest. A funnel plot, Begg’s rank correlation method [17], and Egger’s weighted regression method [18] were adopted to statistically assess publication bias (p < 0.05 was considered statistically significant).

3. Results

3.1. Study selection and characteristics

An initial search yielded 34,005 records, and 23,460 records remained after the removal of duplicates. A total of 23,408 records were excluded after screening the titles/abstracts. After evaluating 52 full texts for eligibility, 23 full-text articles were excluded because they did not have a control/comparison group, and 5 were excluded because they had no outcome of interest. Twenty-four studies were included in the qualitative synthesis and meta-analysis (Fig. 1) [19-41]. There were a total of 4848 patients from 23 studies. Of a total of 23 included studies, 19 were retrospective cohorts, 3 were prospective cohorts and 1 was a case-series study. All studies involved adult patients over 18 years old and used RT-PCR from respiratory tract samples as a confirmatory test for COVID-19 infections. The clinical characteristics of the included studies are summarized in Table 1.

3.2. Quality of study assessment

Studies with various study designs, including cohort and case series, were included in this review and were assessed accordingly with the appropriate scale or tool. The Newcastle Ottawa Scale (NOS) was used to assess the cohort and case-control studies (Table 2), while the Joanna Briggs Institute Critical Appraisal checklist was used for case series studies (Table 3). All included studies were rated ‘good’ based on the criteria used in the Newcastle Ottawa Scale (NOS) and the Joanna Briggs Institute Critical Appraisal checklist. In conclusion, all studies were deemed fit to be included in the meta-analysis.
3.3. Outcomes

3.3.1. Albumin levels

Fifteen studies (n = 4744) reported the levels of serum albumin in each group of outcomes. Decreased albumin was associated with severe disease based on our meta-analysis, with high heterogeneity (mean difference (MD) = −4.58; 95% CI = −5.76 to −3.39; p < 0.00001; I² = 87%; random-effects modeling) (Fig. 2A). The ROC curve for albumin parameters is shown in Fig. 3A, demonstrating that serum albumin provides good discrimination (AUC = 0.827, p = 0.002) between severe

| Study                  | Number participants | Type of study           | Laboratory parameter                  | Severe disease (n, %) | Age (years, mean ± SD) | Non-Severe disease (n, %) | Age (years, mean ± SD) |
|------------------------|---------------------|-------------------------|--------------------------------------|-----------------------|------------------------|--------------------------|------------------------|
| Almazeedi et al. [19]  | 1096                | Retrospective cohort    | Procalcitonin, Albumin, CRP, D-Dimer | 42 (3.8%)             | 54.8 ± 11              | 1054 (96.2%)            | 71.1 ± 15               |
| Alshukry et al. [20]   | 193                 | Retrospective cohort    | Procalcitonin, Albumin, CRP, D-Dimer | 22 (11.4%)            | 52.3 ± 13.5            | 171 (88.6%)             | 64.6 ± 15.7            |
| Cheng et al. [21]      | 456                 | Retrospective cohort    | Procalcitonin, Albumin, CRP, D-Dimer | 251 (55%)             | 59.8 ± 17.4            | 205 (45%)               | 80.1 ± 17.4            |
| Dreher et al. [22]     | 50                  | Prospective cohort      | Procalcitonin, CRP, D-Dimer, LDH      | 24 (48%)              | 63.3 ± 8.8             | 32 (62%)                | 73.1 ± 12.5            |
| Duan et al. [23]       | 348                 | Retrospective cohort    | Procalcitonin, CRP, D-Dimer          | 20 (5.7%)             | 58 ± 15                | 328 (94.3%)             | 64 ± 15                |
| Feng et al. [24]       | 406                 | Retrospective cohort    | Procalcitonin, Albumin, CRP, D-Dimer | 54 (13.3%)            | 57.6 ± 14              | 352 (86.7%)             | 66.7 ± 15.7            |
| Gao et al. [25]        | 43                  | Retrospective cohort    | Procalcitonin, CRP, D-Dimer          | 15 (34.8%)            | 45.2 ± 7.6             | 28 (65.2%)              | 42.9 ± 14              |
| Gong et al. [26]       | 189                 | Retrospective cohort    | Procalcitonin, CRP, Albumin, D-Dimer | 28 (14.8%)            | 63.3 ± 12.9            | 161 (86.2%)             | 68.6 ± 21.4            |
| Huang et al. [27]      | 41                  | Prospective cohort      | Procalcitonin, Albumin, D-Dimer, LDH  | 13 (31.7%)            | 50.3 ± 14.8            | 28 (68.3%)              | 49.1 ± 12.2            |
| Jiang et al. [28]      | 60                  | Retrospective cohort    | D-Dimer                              | 8 (13.3%)             | 56.3 ± 27.4            | 52 (86.7%)              | 60.3 ± 42.2            |
| Khamsi et al. [29]     | 63                  | Retrospective cohort    | CRP, D-Dimer, LDH                     | 24 (38%)              | 50 ± 17                | 39 (62%)                | 57 ± 16                |
| Lv et al. [30]         | 270                 | Retrospective cohort    | Procalcitonin, CRP, D-Dimer          | 155 (57.4%)           | 58.6 ± 47.4            | 115 (42.6%)             | 54.3 ± 41.4            |
| Shang et al. [31]      | 443                 | Retrospective cohort    | Procalcitonin, Albumin, CRP, D-Dimer | 139 (31.3%)           | 63.6 ± 14              | 304 (68.7%)             | 57.3 (14.8)            |
| Shi et al. [32]        | 134                 | Retrospective cohort    | Procalcitonin, Albumin, D-Dimer, CRP | 46 (34.3%)            | 56 ± 14.8              | 88 (65.7%)              | 40 ± 15.5              |
| Sun et al. [33]        | 18                  | Prospective cohort      | Albumin, CRP, D-Dimer, LDH            | 10 (55.3%)            | 59 ± 38.5              | 8 (44.5%)               | 24.6 ± 33.3            |
| Wan et al. [34]        | 135                 | Retrospective case series | Procalcitonin, Albumin, CRP, D-Dimer | 40 (29.6%)            | 60.3 ± 15.5            | 95 (70.4%)              | 42 ± 11.8              |
| Wang et al. [35]       | 45                  | Retrospective cohort    | Albumin, D-Dimer, LDH                 | 10 (22.2%)            | 44.3 ± 25.1            | 35 (77.8%)              | 38.6 ± 34              |
| Wang et al. [36]       | 138                 | Retrospective cohort    | Procalcitonin, CRP, D-Dimer          | 36 (26%)              | 67 ± 15.5              | 102 (74%)               | 50 ± 18.5              |
| Wei et al. [37]        | 167                 | Retrospective cohort    | Procalcitonin, Albumin, CRP, D-Dimer | 30 (17.9%)            | 49 ± 12.6              | 137 (82.1%)             | 40.8 ± 15.4            |
| Yang et al. [38]       | 200                 | Retrospective cohort    | Procalcitonin, Albumin, CRP, D-Dimer | 29 (14.5%)            | 71 ± 13.4              | 171 (85.5%)             | 52 ± 16.2              |
| Yi et al. [39]         | 100                 | Retrospective cohort    | Procalcitonin, CRP, D-Dimer          | 49 (49%)              | 60.6 ± 14              | 51 (51%)                | 48 ± 16.2              |
| Zhang et al. [40]      | 140                 | Retrospective cohort    | Procalcitonin, CRP, D-Dimer          | 56 (40.3%)            | 58.6 ± 45.9            | 82 (59.5%)              | 51.8 ± 38.5            |
| Zhang et al. [41]      | 113                 | Retrospective cohort    | Procalcitonin, Albumin, CRP, LDH      | 61 (53.9%)            | 53.6 ± 13.3            | 52 (46.1%)              | 34.2 ± 19.6            |
COVID-19 and nonsevere COVID-19 infections, with an optimal cutoff of 38.85 g/L, yielding a sensitivity of 66.7% and a specificity of 93.3% (Table 4).

3.3.2. C-reactive protein (CRP) levels

Nineteen studies \((n = 4558)\) reported the levels of serum CRP in each group of outcomes. Elevated CRP was associated with severe disease based on our meta-analysis, with high heterogeneity \((\text{mean difference (MD) 36.88; 95% CI 29.10–44.65; } p < 0.00001; \mu^2 = 84\%; \text{random-effects modeling})\) \((\text{Fig. 2B})\). The ROC curve for CRP parameters is shown in \(\text{Fig. 3B}\), demonstrating that serum CRP provides good discrimination \((\text{AUC = 0.891, } p < 0.0001; \mu^2 = 89\%; \text{random-effects modeling})\) \((\text{Fig. 2B})\). The ROC curve for D-dimer parameters is shown in \(\text{Fig. 3C}\), demonstrating that serum D-dimer provides good discrimination \((\text{AUC = 0.836, } p < 0.001)\) between severe COVID-19 and nonsevere COVID-19 infections with an optimal cutoff of 0.635 μg/L, yielding a sensitivity of 75% and a specificity of 90% (Table 4).

3.3.3. D-dimer levels

Twenty-one studies \((n = 4426)\) reported the levels of serum D-dimer in each group of outcomes. Elevated D-dimer was associated with severe disease based on our meta-analysis, with high heterogeneity \((\text{mean difference (MD) 0.43; 95% CI 0.31–0.56; } p < 0.00001; \mu^2 = 83\%; \text{random-effects modeling})\) \((\text{Fig. 2C})\). The ROC curve for D-dimer parameters is shown in \(\text{Fig. 3C}\), demonstrating that serum D-dimer provides good discrimination \((\text{AUC = 0.836, } p < 0.001)\) between severe COVID-19 and nonsevere COVID-19 infections with an optimal cutoff of 0.635 μg/L, yielding a sensitivity of 75% and a specificity of 90% (Table 4).

### Table 2

Newcastle-Ottawa quality assessment of observational trials.

| First author, year | Study design | Selection | Comparability | Outcome | Total score | Result |
|--------------------|--------------|-----------|---------------|---------|-------------|--------|
| Almazeedi et al. [19] | Cohort       | ****      | **            | ***     | 9           | Good   |
| Alshukry et al. [20] | Cohort       | ***       | **            | ***     | 8           | Good   |
| Cheng et al. [21]   | Cohort       | ***       | **            | ***     | 8           | Good   |
| Dreher et al. [22]  | Cohort       | **        | **            | ***     | 7           | Good   |
| Duan et al. [23]    | Cohort       | ****      | **            | ***     | 9           | Good   |
| Feng et al. [24]    | Cohort       | ***       | **            | ***     | 8           | Good   |
| Gao et al. [25]     | Cohort       | **        | **            | ***     | 7           | Good   |
| Gong et al. [26]    | Cohort       | ***       | **            | ***     | 8           | Good   |
| Huang et al. [27]   | Cohort       | ***       | **            | ***     | 8           | Good   |
| Jiang et al. [28]   | Cohort       | ***       | **            | ***     | 8           | Good   |
| Xhamsis et al. [29] | Cohort       | ***       | **            | ***     | 8           | Good   |
| Lv et al. [30]      | Cohort       | ***       | **            | ***     | 8           | Good   |
| Shang et al. [31]   | Cohort       | ***       | **            | ***     | 8           | Good   |
| Shi et al. [32]     | Cohort       | ***       | **            | ***     | 8           | Good   |
| Sun et al. [33]     | Cohort       | ***       | **            | ***     | 7           | Good   |
| Wang et al. [35]    | Cohort       | ***       | **            | ***     | 7           | Good   |
| Wang et al. [36]    | Cohort       | ***       | **            | ***     | 8           | Good   |
| Wei et al. [37]     | Cohort       | ***       | **            | ***     | 7           | Good   |
| Yang et al. [38]    | Cohort       | ***       | **            | ***     | 8           | Good   |
| Yi et al. [39]      | Cohort       | ***       | **            | ***     | 7           | Good   |
| Zhang et al. [40]   | Cohort       | ***       | **            | ***     | 9           | Good   |
| Zhang et al. [41]   | Cohort       | ***       | **            | ***     | 8           | Good   |

Each (*) means one score given to that criteria, so *** means the score of the study under that criteria is 3.
Fig. 2. Forest-plot analysis for serum albumin (A), CRP (B), D-Dimer (C), LDH (D), and procalcitonin (E) in severe and non-severe COVID-19.
Fig. 3. ROC-curve analysis for serum albumin (A), CRP (B), D-Dimer (C), LDH (D), and procalcitonin (E) for predicting severe COVID-19 infection.
−3.63), p < 0.00001, I² = 0%, random-effect modeling; MD = 50.63; (95% CI 11.00–90.26), p = 0.01; I² = 0%; random-effect modeling; MD = 2.45; 95% CI 1.48–3.42; p < 0.00001; I² = 0%; random-effect modeling; MD = 126.43; 95% CI 52.15–200.71; p = 0.0009; I² = 50%; random-effect modeling.) On the other hand, subgroup analysis for retrospective studies (mean difference (MD) 0.07; 95% CI 0.04–0.09; p < 0.00001; I² = 92%; random-effect modeling) showed a lower but more significant mean difference (MD) for procalcitonin levels between severe and nonsevere COVID-19 compared with prospective studies (mean difference (MD) 0.75 (95% CI −0.76–2.25), p = 0.33, I² = 83%, random-effect modeling).

### 3.2.7. Publication bias

The funnel-plot analysis showed a relatively symmetrical inverted funnel plot for the albumin (Fig. 4A), CRP (Fig. 4B), D-dimer (Fig. 4C), and procalcitonin (Fig. 4D), an parameters but showed an asymmetrical shape for the LDH (Fig. 4D) parameter, indicating possible publication bias. Meanwhile, rank-correlation Begg’s test and regression-based Egger’s test were not statistically significant for albumin and CRP parameters, showing no indication of publication bias, but were statistically significant for procalcitonin, D-dimer, and LDH parameters, showing a possible indication of publication bias (Table 4).

### 4. Discussion

This systematic review and meta-analysis included 24 articles with quite large samples of COVID-19 patients. Our meta-analysis suggests that patients with severe COVID-19 infections have higher procalcitonin, CRP, D-dimer, and LDH levels but lower levels of albumin than those with nonsevere COVID-19 infections.

Albumin is a natural colloid that is abundant in plasma. It is exclusively synthesized in the liver and serves several purposes, such as maintaining intravascular oncocytic pressure, acting as a carrier of several different endogenous and exogenous compounds, and maintaining the acid-base balance, and is often used as a marker of nutritional status and particular disease severity (e.g., liver cirrhosis) [42]. Several conditions can decrease the levels of serum albumin, such as decreased albumin production (e.g., advanced stage of hepatic cirrhosis and increased cataabolism due to systemic illness), nutritional deficiencies (e.g., kwashiorkor), and increased loss of albumin (e.g., renal loss, gut loss, and extravascular loss). Systemic inflammatory, such as what happens in sepsis, will increase systemic vascular permeability and capillary leakage, causing albumin to shift toward the extravascular space. In such patients, the synthesis of albumin is also disturbed, making hypoalbuminemia more profound [43]. In the case of COVID-19, the low levels of albumin in severe disease may be caused by malfunctioning system organs (vascular permeability, renal, and gastrointestinal), contributing to a greater extent of albumin excretion. Moreover, albumin has been found to have the ability to downregulate ACE2 receptors [44], which is crucial for modulating COVID-19 infection; therefore, low levels of albumin will possibly result in upregulation of ACE2 receptors and an increase in COVID-19 infectivity.

C-reactive protein (CRP) is a sensitive biomarker for inflammation. As an acute-phase inflammatory mediator, CRP is synthesized and released by the liver to the bloodstream, contributing to the host's resistance against invading pathogens [45]. The elevation of CRP may also be caused by bacterial coinfection that occurs in severe COVID-19. Moreover, a robust inflammatory response that occurs in severe disease may cause the levels of CRP to increase significantly.

D-dimer is one marker associated with thrombotic events and may also be elevated in infections such as influenza, SARS, and CAP [46,47]. In COVID-19 patients, elevation of the D-dimer level is common and is associated with disease severity and mortality. D-dimers are fragments produced when plasmin cleaves fibrin to break down clots. Therefore, every process that increases fibrin production or breakdown will elevate plasma D-dimer levels. It is assumed that in severe COVID-19 (critically ill) patients, proinflammatory cytokines and the coagulation cascade, including D-dimer, are activated [48]. Studies also suggest that under inflammatory conditions, dysregulation of the coagulation cascade in SARS-CoV-2 infection results in diffuse alveolar damage with cellular fibromyxoid exudates, pneumocyte desquamation, and formation of a hyaline membrane, pulmonary edema with hyaline membrane formation, and interstitial mononuclear inflammatory infiltrates dominated by lymphocytes [49,50]. This event shifts the alveolar hemostatic balance toward prothrombotic activity. Proinflammatory cytokines also contribute to endothelial injury, which could activate coagulation and inhibit fibrinolysis in patients with severe COVID-19 [51]. This elevation of D-dimer suggests that there is a hypercoagulable state contributing to the severity of the disease and increased mortality.

Lactate dehydrogenase (LDH) is a cytoplasmic glycolytic enzyme that catalyzes the reversible conversion of L-lactate and pyruvate with concomitant interconversion of NADH and NAD+. It is present in the cytoplasm of all human tissues, with higher concentrations in the liver, heart, and skeletal muscle [52]. It has been reported that elevated LDH levels are one of the most common findings in patients infected with MERS-CoV [53] and H5N1 [54] and are also one of the biomarkers most strongly associated with ARDS mortality [55]. Multiple organ injury and decreased oxygenation with upregulation of the glycolytic pathway can result in abnormal values because the acidic extracellular pH resulting from infection and tissue injury will activate metalloproteases and enhance macrophage-mediated angiogenesis [56]. Increased LDH in severe COVID-19 suggested possible subclinical tissue damage. LDH itself is released when cellular necrosis happens; therefore, in severe infections in which a large number of cells typically undergo necrosis, high serum LDH levels can be observed. In particular, severe COVID-19 infections that mainly affect the lungs will release a high amount of LDH isozyme commonly found in lung tissue (LHD isoenzyme 3) [57,58].

A peptide precursor of the hormone calcitonin, PCT, has been widely investigated as a promising biomarker for the initial investigation of a bacterial infection [59]. An elevated serum PCT is often found in patients with sepsis and septic shock [60]. While it is still controversial whether PCT can accurately distinguish bacterial or viral pneumonia [61], PCT-guided therapy in acute respiratory infections may reduce antibiotic exposure [62]. Bacterial infections trigger extrathoroidal synthesis of PCT, which is actively maintained by elevated values of IL-6, IL-1β, and TNF-α, while viral infections hinder PCT production due to interferon-γ [63]. In this meta-analysis we found that an elevated serum PCT was associated with severe COVID-19.
Our systematic review and meta-analysis have several strengths. First, we included all published studies, therefore, the risk of publication bias was minimized. Second, the pooled effect estimates of our meta-analysis were very precise. Third, we also provided a cutoff value for each parameter with high sensitivity and specificity based on ROC curve analysis to help physicians predict the severe outcome of COVID-19.

Nonetheless, this review has its limitations. Given the observational design of the included studies and the retrospective data collection, the possibility that the observed association between each laboratory parameter and the severity of COVID-19 was affected by bias or confounding factors should still be considered. The asymmetrical inverted funnel plot for serum LDH and the significant p-values from Begg’s and Egger’s tests for serum procalcitonin, D-dimer, and LDH imply the presence of publication bias. Another limitation is that the total sample size of our meta-analysis was not very high due to the limited number of studies that provided data regarding laboratory parameters that matched our inclusion criteria. We also combined the results from prospective and retrospective cohort studies. Finally, there is an unclear association between the time at which biomarkers were tested and the determination of severe or nonsevere COVID-19; however, we believe that early laboratory testing since the patients were first admitted to the emergency unit can still be used to monitor and anticipate the progression of nonsevere to severe COVID-19; therefore, timely and suitable management can be performed.

Further research can explore another potential laboratory parameter to predict the severity or mortality of COVID-19. A cutoff value for new potential parameters should also be included to help physicians predict severe COVID-19.

5. Conclusion

This meta-analysis suggests serum procalcitonin, CRP, D-dimer, and LDH are elevated in patients with severe COVID-19 infection, while serum albumin levels are lower in severe illness compared with nonsevere COVID-19 infections. Further study is needed concerning specific threshold levels of these laboratory markers. Finally, COVID-19 patients with high levels of serum procalcitonin, CRP, D-dimer, and LDH and low levels of serum albumin should be monitored closely to minimize the risk of progression to severe disease.

Authorship

All authors whose names appear on the submission:

1) made substantial contributions to the conception or design of the work; or the acquisition, analysis, or interpretation of data; or the creation of new software used in the work;
2) drafted the work or revised it critically for important intellectual content;
3) approved the version to be published; and
4) agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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Declaration of Competing Interest

The authors declare no conflict of interest regarding this article.

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None.
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