Nitric Oxide Synthase Type 1 Methylation Is Associated With White Matter Microstructure in the Corpus Callosum and Greater Panic Disorder Severity Among Panic Disorder Patients

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Objectives: Methylation of the neuronal nitric oxide synthase (NOS1/nNOS) gene has recently been identified as a promising biomarker of psychiatric disorders. NOS1 plays an essential role in neurite outgrowth and may thus affect the microstructure development of white matter (WM) in the corpus callosum (CC), which is known to be altered in panic disorder (PD). We examined the relationship between NOS1 methylation, WM tracts in the CC, and symptoms based on this finding.

Methods: Thirty-two patients with PD and 22 healthy controls (HCs) were recruited after age, gender, and the education level were matched. The cell type used was whole-blood DNA, and DNA methylation of NOS1 was measured at 20 CpG sites in the promoter region. Although 25 patients with PD were assessed with the Panic Disorder Severity Scale (PDSS), diffusion tensor imaging (DTI) scans were only collected from 16 participants with PD.

Results: We observed that the PD group showed lower methylation than did the HCs group and positive correlations between the symptom severity of PD and methylation at CpG4 and CpG9. In addition, CpG9 methylation was significantly correlated with the fractional anisotropy (FA) and mean diffusivity (MD) values of the CC and its major components (the genu and the splenium) in the PD group. Furthermore, path analyses showed that CpG9 methylation offers a mediating effect for the association between the MD values of the genu of the CC and PD symptom severity (95% CI = −1.731 to −0.034).

Conclusions: The results suggest that CpG9 methylation leads to atypical development of the genu of the CC, resulting in higher PD symptom severity, adding support for the methylation of NOS1 as a future prognostic indicator of PD.

Keywords: panic disorder (PD), nitric oxide synthase type I, corpus callosum (CC), DNA methylation, diffusion tensor imaging
INTRODUCTION

Panic disorder (PD) is a frequent anxiety disorder characterized by recurrent accidental panic attacks and unexpected anxiety. PD is considered to be a multifactorial disease resulting from the interactions of multiple genetic and environmental factors. According to previous studies, the 12-month prevalence rate of PD is 0.3%, and the lifetime prevalence rate is estimated at about 0.5% in China (1, 2).

The etiology, neuropathology, and pathophysiology of PD remain elusive. Extensive research has been performed to identify specific biomarkers of PD. Nitric oxide (NO) has been identified as a neuromodulator in the central nervous system (CNS). NO, formed by the enzyme NOS1 and encoded by the NOS1 gene, has the capacity to influence neurodevelopment and growth, the plasticity of synapse, and neurotransmitter release (3–6). NO also plays an important role in many behavioral domains, including aggression (7), anxiety (8), depression, and cognitive functioning (9). Of note is that NO can regulate both the glutamatergic and dopaminergic systems, which are strongly implicated in the biochemical pathology of anxiety (10).

NOS1, located on chromosome 12q24.21 (11), is the central source of NO in the CNS (12, 13). Increasing evidence suggests that NOS1 is implicated in multiple complex neurobiological mechanisms in neuropsychiatric disorders (14–17). For example, Zhou et al. have studied NOS1 and its signal mechanism involved in the pathophysiology of anxiety (18).

Additionally, Kurrikoff et al. demonstrated that females with a short NOS1 ex1f-VNTR allele had higher anxiety scores than did females homozygous for the long alleles when confronted with environmental adversity (15). A recent report by Sarginson et al. with large sample sizes corroborated this finding (19). Thus, exploring DNA methylation may be able to provide a comprehensive view of both the genetic and environmental risk factors for PD. Existing evidence shows that epigenetic modification of NOS1 caused by DNA methylation is related to gene expression and that these NOS1 modifications can affect the generation and bioavailability of NO (20–23). These findings provide preliminary evidence that NOS1 methylation is an indicator of abnormal NO signaling. Also, considering that NO plays an essential role in the neuron and neurite outgrowth in the brain (24–28), this suggests potential influences of NOS1 methylation on brain development through white matter (WM) outgrowth and neurogeny.

In the “fear network model” for PD, the important fear network regions are connected by the corpus callosum (CC) in the bilateral sides (29). The CC is an area dedicated to attention, physical complaints, and related anxiety (30). Structural abnormalities in the CC may predispose patients with PD to panic attacks because of the uneven distributions of the fear network on each side. Thus, the CC likely plays a key role in the pathophysiology of PD (31). Evidence from previous diffusion tensor imaging (DTI) studies not only supports the notion that the CC may play a critical role in PD (32–34) but also revealed an association between altered WM connectivity in PD and dysfunctional clinical symptoms. In particular, anxiety symptoms appear to impact the human WM microstructure, leading us to investigate whether epigenetic modifications (i.e., DNA methylation) contribute to these WM differences (35). DNA methylation provides tools to better understand the impact of PD symptoms on the molecular pathways in PD patients. These markers may provide insights into the potential molecular mediating pathways contributing to the neurodevelopmental effects associated with PD symptoms. Consequently, NOS1 methylation may influence the WM microstructure of CC development in the brain, but this is yet to be investigated.

Therefore, the present study aimed to identify NOS1 methylation and the pathways involved in WM development in PD patients. We examined the levels of NOS1 methylation between patients with PD and healthy controls (HCs) and how the levels of DNA methylation related to the WM microstructure in the CC. We hypothesized that the level of NOS1 methylation would differ between the two groups and are related to the WM microstructure in the CC. This study was expected to contribute to the early intervention and treatment in PD.

MATERIALS AND METHODS

Participants

A total of 54 participants were recruited from the Nanjing Brain Hospital affiliated with Nanjing Medical University between August 2014 and July 2018. Thirty-two adult patients were diagnosed with PD by an experienced psychiatrist conforming to the Diagnostic and Statistical Manual of Mental Disorders (DSM-IV) criteria. In order to reduce the probability of diagnostic errors, all patients were evaluated with the Mini-International Neuropsychiatric Interview (MINI) by two resident physicians. Patients were also assessed with the Hamilton Anxiety Rating Scale (HAMA) (36) and the Panic Disorder Severity Scale (PDSS) (37, 38) on the same day. Magnetic resonance imaging (MRI) and peripheral blood collection were performed at the Nanjing Brain Hospital within 7 days. A deputy chief physician supervised the whole procedure. Twenty-two age- and sex-matched HCs were consecutively recruited from the Nanjing Brain Hospital via word of mouth and public advertisements according to the inclusion and exclusion criteria. HAMA was also used to assess the anxiety levels for HCs.

Inclusion and Exclusion Criteria

In the PD group, the inclusion criteria were as follows: (1) a primary diagnosis of PD by an experienced psychiatrist conforming to the DSM-IV criteria; (2) confirmation of PD diagnosis using MINI; (3) age 18–55 years; and (4) right-handed and masterful at completing all examinations. The exclusion criteria were as follows: (1) any neurological disorders or other psychiatric disorders and major physical or infectious diseases; (2) any comorbid mental illness such as depression, generalized anxiety disorder, bipolar disorder, obsessive–compulsive disorder, schizophrenia, alcohol addiction, social phobia, or eating disorder; (3) history of psychotherapy within 6 months of study enrollment; (4) inability to complete MRI; and (5) pregnancy and lactation.

The inclusion criteria for the HCs were as follows: (1) age 18–55 years; (2) HAMA total score ≤7; and (3) right-handed...
and masterful at completing all examinations. The exclusion criteria were: (1) any neurological disorders or other psychiatric disorders and major physical or infectious diseases; (2) history of mental illness or family history; (3) pregnancy and lactation; (4) history of psychotherapy within 6 months of study enrollment, and (5) inability to complete MRI.

**Analysis of NOS1 DNA Methylation**

The cell type used was whole-blood DNA, and the DNA methylation of NOS1 was measured at 20 CpG sites in the promoter region. Specifically, the genomic regions of interest (ROIs) were analyzed with the geneCpG software and the methylation patterns determined with bisulfite sequencing (see Supplementary Table 1). Firstly, the PCR primer pairs were designed from bisulfate-converted DNA with the Methylation Primer software (see Supplementary Table 2). Genomic DNA was then extracted from frozen samples using a Genomic Tip-500 kit (QIAGEN, Valencia, CA, USA) according to the manufacturer’s protocol, and sulfuric acid was obtained from sulfite using the EZ DNA Methylation™-GOLD Kit (Zymo Research, Irvine, CA, USA). After PCR amplification (HotStarTaq polymerase kit; TAKARA, Tokyo, Japan) and construction of the library, the samples were sequenced using a paired-end sequencing protocol according to the manufacturer’s guidelines (Illumina MiSeq Benchtop Sequencer, San Diego, CA, USA) (39). Normality of the continuous variables was performed with Shapiro–Wilk’s tests. Statistics were performed using t-tests and ANOVA.

**MRI Acquisition and Processing**

The MRI data (3.0T Siemens Medical System scanner, Nanjing Brain Hospital) were obtained using a diffusion tensor echo-planar pulse sequence [matrix = 128 × 128, field of view (FOV) = 240 mm × 240 mm, slice thickness = 3 mm with no interslice gap, repetition time (TR) = 6,600 ms, and echo time (TE) = 93 ms]. For the DTI, the diffusion sensitization gradients were applied in 30 non-collinear directions (b = 1,000 s/mm²), together with non-diffusion-weighted acquisition (b = 0). During the scan (240 volumes), each participant was instructed to remain motionless and to keep the eyes closed. Foam pads and earplugs were used to reduce head movement and noise.

DTI data were processed using the pipeline for analyzing brain diffusion images (PANDA; http://www.nitrc.org/projects/panda) implemented in MATLAB (40). Firstly, all files in Digital Imaging and Communications in Medicine (DICOM) format were converted into Nifti format by MRIcon. Subsequently, using the bet command of FSL (41), the non-diffusion b0 image was used to estimate the brain mask. The purpose of the brain mask was to aid in determining the boundaries of the three dimensions of the brain. Any distortions of the diffusion-weighted images (DWI) caused by eddy current or simple head movement during scanning were corrected by registering the DWI to the b0 image through affine transformation. Then, voxel-wise calculations of the tensor matrix and the calculated diffusion tensor (DT) metrics were estimated, including fractional anisotropy (FA) and mean diffusivity (MD). MD represents the intensity of the diffusion, while FA represents the direction of water diffusion in the brain, both of which indicate the organizational and microstructural integrity of the brain WM tracts. By executing the first command of FSL, non-linear registration was performed on the individual FA and MD images in local space, which were then imported into the FA and MD templates in the Montreal Neurological Institute (MNI) space. We then resampled the diffusion metric (FA and MD) images into the MNI space with a custom spatial resolution (1 × 1 × 1 mm). Finally, the FA and MD images were smoothed with a full width of 6 mm at half of the maximum Gaussian filter.

**Regions of Interest**

To explore the relationship between structural changes and the clinical symptoms in PD patients, we selected the CC and its major components (genu, body, and splenium) as ROIs. The CC mask contained in MRIcon (JHU ICBM-DTI-81 WM labels atlas) (42) included the genu, body, and splenium of the CC. The regional diffusion metrics, FA and MD, were calculated by averaging the values within each region of the CC in DPABI (43). Then, the data were statistically analyzed with SPSS 22.0.

**Statistical Analysis**

Demographic and clinical characteristics were evaluated using chi-square and t-tests. Standard statistical protocol was used for descriptive statistics, followed by the Shapiro–Wilks test (see Supplementary Table 3) to establish differences between the two groups. To investigate region-specific correlations between the PDSS scores and CpG4,9 methylation, we performed Pearson’s correlation analysis using SPSS 22.0. Then, the CpG9 site related to the clinical symptoms was further evaluated using Spearman’s correlation analysis to examine its relationship with the FA and MD values of the ROIs. Simple mediation analyses were performed using PROCESS, and age, gender, and education were included as covariates in the model. PROCESS is based on a regression-based path analysis framework and incorporates mediation and reconciliation into a conditional process model (44). All analyses were based on 5,000 bootstrap samples. When the CI did not include zero, an indirect effect was considered significant.

**RESULTS**

**Sample Characteristics**

From the 40 PD patients and 40 HCs who were invited, 32 PD patients and 22 HCs met the study criteria and completed the assessment. **Table 1** compares the socio-demographic and clinical characteristics of the 32 patients with PD (age ± SD = 33.1 ± 7.4, 16 males) and the 22 controls (age ± SD = 33.3 ± 7.2, 12 males). Patients and controls did not differ with regard to age, sex, or education (all p > 0.05) (**Table 1**). The HAMA total scores were significantly different between patients with PD vs. HCs (**Table 1**). Since seven patients with PD did not fully complete the PDSS or clinical measures, the numbers varied slightly in different categories. The mean PDSS score in patients was 14.8 (n = 25, SD = 5.7), ranging from 8 to 22. DNA methylation data were available for all 54 participants, and neuroimaging data were available for 16 participants with PD.
TABLE 1 | Comparison of the demographic and clinical variables between PD patients and HCs.

|                        | PD patients (n = 32) | HCs (n = 22) | Statistics |
|------------------------|----------------------|--------------|------------|
| Gender                 |                      |              |            |
| Male                   | 16                   | 12           | 0.11       |
| Female                 | 16                   | 10           |            |
| Mean                   | 33.1                 | 33.3         | 0.73       |
| Age (years)            | 50.0                 | 54.5         |            |
| SD                     | 7.4                  | 3.3          |            |
| Education (years)      | 13.9                 | 15.6         | 1.71       |
| HAMA-T                 | 20.3                 | 2.2          | -11.8      |
| PDSS scorea            | 14.8                 | 1.9          | 0.00       |
|                       |                      |              |            |

x² and p-values were obtained with the chi-squared test. T and p-values were obtained with two-sample t-tests. PD, panic disorder; HCs, healthy controls; SD, standard deviation; PDSS, Panic Disorder Severity Scale; HAMA-T, total score of the Hamilton Anxiety Scale. *Twenty-five patients with PD completed the PDSS.

NOS1 Methylation Status

We compared the methylation of each CpG unit in the PD (n = 32) and HC (n = 22) groups using t-tests (Figure 1). We observed that the PD group had significantly lower methylation at CpG4, CpG7, CpG9, CpG10, and CpG15 than did the HC group (p < 0.05). Thus, all subsequent epigenetic analyses were carried out only for these five CpG sites.

Correlation Between Clinical Symptoms and DNA Methylation of NOS1

We then performed Pearson’s correlation analysis in the PD group to confirm the relationship between reduced methylation at five CpG sites and clinical status. A significant positive association was observed between PD symptom severity and the DNA methylation of CpG4 (n = 25, r = 0.412, p = 0.041) and CpG9 (n = 25, r = 0.509, p = 0.009) in the PD group (Figure 2), whereas no significant association was found for CpG7, CpG10, and CpG15 (p > 0.05).

Correlation Between Corpus Callosum Alterations and Clinical Assessments

The FA and MD values of the bilateral CC (including three parts of it) (Figure 3A) were extracted and correlated with NOS1 methylation. As described in the previous section and presented in Figure 2, we observed significant positive correlations between PD severity and NOS1 methylation (at sites CpG4 and CpG9) in the PD group. We then performed Spearman’s correlation analysis between CpG4 and CpG9 methylation and the FA and MD values of the CC in the PD group. As shown in Figure 3A, the results of the correlation analysis showed that the degree of CpG9 methylation was negatively correlated with the MD values in the total CC (n = 16, r = −0.700, p = 0.003), genu (n = 16, r = −0.532, p = 0.036), and the splenium (n = 16, r = −0.737, p = 0.002) (Figure 3B). In contrast, no significant association was found in the body of the CC (p > 0.05). Similarly, we found that the degree of CpG9 methylation was positively correlated with the FA values in the total CC (n = 16, r = 0.535, p = 0.03) and the splenium (n = 16, r = 0.647, p = 0.008) (Figure 3C). The effect was not statistically significant for CpG4.

NOS1 Epigenetic Variation as a Mediator of the Brain and Relationship With Clinical Symptoms

Based on the findings above, we conducted an additional path analysis to examine whether a reduced NOS1 methylation mediated the relationship between the WM microstructure in the CC and PD symptom severity. All phenotypes were examined simultaneously in one model. As shown in Figure 4, the mediation model suggested that NOS1 methylation at CpG9 mediated the connection between the genu of CC and PD symptom severity. The indirect effect of PD on the MD values of the genu of CC through DNA methylation was significant (β = −0.639, 95% CI = −1.731 to −0.034). The effect was not statistically significant for the other regions.

DISCUSSION

The current study extended the association between NOS1 DNA methylation in PD by investigating the relationship between this novel epigenetic risk locus, clinical symptoms, and the microstructure and organization of WM tracts. We observed that the PD group showed lower NOS1 CpG4 and CpG9 methylation than did the HC group, which led us to examine the link between CpG4 and CpG9 methylation and PD symptoms. The results showed that NOS1 methylation at these loci had an apparent positive correlation with current PD symptoms. Finally, path analysis showed that NOS1 methylation at the CpG9 site significantly mediated the relationship between PD and the WM microstructure in the CC. These discoveries extend previous findings by linking NOS1 to alterations in the WM, representing a possible brain endophenotype of oxidative stress in anxiety and broadening its potential clinical service by revealing its value as a promising PD biomarker.

Although the role of NOS1 methylation in the relationship between clinical PD symptoms and the WM microstructure has not been extensively studied, a recent study has shown that the NOS1 ex1f-VNTR genotype is associated with clinical
symptoms and a high MD value in several major WM tracts, including the genu of the CC in patients with psychotic disorders (17). This finding is in line with the notion that NO influences brain development through WM organization. Our study expanded on this by showing that WM alterations in the CC were related to NOS1 methylation in the PD group. Our assumption that the DNA methylation of NOS1 might add to the susceptibility of PD is verified by these results. Additionally, we found that methylation of the NOS1 promoter was significantly reduced in the PD group and was positively associated with current PD severity. This finding is supported by a genome-wide epigenetic study that found the hypomethylation of NOS1 in brain tissue from patients with schizophrenia (23). Additionally, previous studies have found hypomethylation of the glutamate decarboxylase 1 (GAD1) and monoamine oxidase A (MAOA) genes in PD (45, 46), which might very well describe a compensatory mechanism to counteract a genetically driven reduction of NOS1 expression in the prefrontal cortex. Thus, our results of a positive correlation between the methylation of NOS1 and the PDSS scores are in line with these findings because the reduction in methylation can be thought to counteract the genetically driven reduced NOS1 expression in PD. Methylation that occurs in the CpG islands of promoter regions usually suppresses gene expression (47); therefore, a reduced methylation of NOS1 could be associated with increased NOS1 gene expression. NOS1, known as one of the NOS isoforms that produce the signaling molecule NO (48–50), has convincingly been linked to anxiety-like

FIGURE 1 | Frequency of DNA methylation at 20 CpG sites located in the promoter region of the NOS1 gene in patients with panic disorder (PD) and in healthy controls (HCs). *p < 0.05.

FIGURE 2 | Correlations between the clinical symptoms (PDSS scores) and CpG4 and CpG9 DNA methylation in the panic disorder (PD) group. The threshold was set at a significance level of p < 0.05. PDSS, Panic Disorder Severity Scale.
Abnormal WM integrity in the CC in anxiety disorders has been reported in several DTI studies (29, 55, 56). The CC is a non-uniform aggregation of nerve fibers below the cortex next to the longitudinal fissure that works to integrate motor, emotional, and cognitive functions (57). The results of the structural ROI analysis revealed a correlation between a reduced NOS1 methylation and the FA and MD values in the CC in the PD group. As is known, MD represents the average molecular motion, and both the size and integrity of cells can affect it. FA is a measure of axonal integrity and is closely associated with fiber integrity (58). Consistent with previous evidence for a strong genetic influence on the organization and microstructure of WM tracts (59), the negative association between the MD values in the CC and CpG9 methylation, as well as the positive association between the FA values in the CC and the CpG9 methylation in our study, suggested a directional influence of NOS1 on neurite outgrowth. For example, lower FA values or higher MD values can be an indicator of axonal density reduction or axonal mutation. Based on these findings, the hypomethylation of CpG9 may have led to the modulation of NOS1 gene transcription and expression through aberrant outgrowth and organization of the WM microstructure during early brain development. There are

behavior (9, 51, 52) in rodents. In contrast, very few works in humans have found nitrinergic system involvement in PD. Two studies indicated that the levels of NO in the blood refer to diurnal changes in patients with PD (53, 54). Overall, NO is strongly related to the dopamine system and serotonergic neurotransmission, which offers a rationale for the participation of NO in PD.

**FIGURE 3** Scatter plot and line show the association between the degree of CpG9 methylation and the fractional anisotropy (FA) and mean diffusivity (MD) values in the region of interest (ROI) of the brain. (A) ROI: the total corpus callosum (CC). Red represents the genu, orange represents the body, and green represents the splenium of the CC. The statistical threshold for the contrasts was \( p < 0.05 \). (B) Black circles and line represent the MD values of the whole CC, the red circles and line represent the MD values of the genu of the CC, and the green circles and line represent the MD values of the splenium of the CC. (C) Black circles and line represent the FA values of the whole CC and the green circles and line represent the FA values of the splenium of the CC.

**FIGURE 4** The mediating role of CpG9 methylation of NOS1 in the relationship between the mean diffusivity (MD) values of the genu the corpus callosum (CC) and the Panic Disorder Severity Scale (PDSS) scores.
many NOS1 gene products that have not been widely studied, so the precise mechanisms by which NOS1 may influence the CC are unknown. Moreover, no neuroanatomical structure generating NO in a centralized manner has been defined since every NO-producing neuron is independent and synthesizes NO in a non-synchronous way (60). Hence, NO has different effects in the different brain regions and might even be antagonistic on the behavioral level.

Given that DNA methylation may represent a key link between external environmental factors and persistent phenotypic changes, our work provides the first evidence for a mediating role of methylation in NOS signaling in brain development in patients with PD. Although we could not clarify the causality in this cross-sectional study, path analysis showed that NOS1 methylation significantly mediated the association between the MD values of the genu of the CC and clinical symptom severity in PD. There was no similar association for the other components in the model, which suggests that the genu of the CC might be more sensitive to changes in WM microintegrity in PD patients. Aboitiz et al. found that thin fibers are most dense in the genu of the CC (61), indicating a better capacity for interhemispheric transfer. Laitinen et al. reported that psychiatric patients with symptoms of anxiety and tension could immediately benefit from high-frequency electrical stimulation in the genu of the CC (62). These findings highlight the importance of the genu of the CC and its prominent role in the severity of PD symptoms. Furthermore, we found a mediating effect of NOS1 hypomethylation on the relationship between the genu of the CC and the severity of symptoms in PD, while no direct relationship was found between the two. One of the possible reasons for the relevance of the genu of the CC not being observed with the PDSS may be the role negative life events play in PD. Several studies found that early-life stress was associated with increased symptom ratings and meaningful alterations in the CC (63–66). Moreover, DNA methylation has also been regarded as a gene × environment interaction biomarker (46, 67–69). Thus, the DNA methylation mechanism may explain why adversity in early life causes biological changes in psychiatric disorders. Taken together, these findings suggest that the aberrant microstructure of the WM in the genu indirectly affects the severity of PD symptoms, mediated by the lower NOS1 methylation expressed in the CC.

Several limitations should be noted in our study. Firstly, our results are limited given the relatively small sample size in a single center and the cross-sectional nature of our data, so potential selection bias may exist. We did not have the statistical power to draw firm conclusions on the direction of the proposed effects. Secondly, we were unable to assess gene expression and could not evaluate its relationship with the methylation changes. Thirdly, peripheral blood may not necessarily reflect the levels of DNA methylation in the CNS despite previous evidence suggesting that the DNA methylation patterns in peripheral blood cells and several brain areas are highly comparable (70, 71). Finally, confirming the mechanism of epigenetic changes and neural integrity implicated in traumatic stress may need more prospective studies.

In conclusion, these findings extend our understanding of the effects of the increased underlying neuropathological basis on psychiatric disorders by identifying NOS1 methylation as a potentially valuable blood biomarker for WM microstructure in psychiatrically relevant brain regions, which ultimately could be applied to early intervention and treatment. Furthermore, our results provide new insights into a neurobiological pathway from NOS1 epigenetic variation, which may confer increased susceptibility to PD.

DATA AVAILABILITY STATEMENT

The datasets presented in this article are not publicly available due to privacy and ethical restrictions. Requests to access the datasets should be directed to the corresponding author Chun Wang, fm51109@163.com.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by the Ethics Committee of the Nanjing Brain Hospital, affiliate of Nanjing Medical University. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

HX made substantial contributions to initiate, design the study, and wrote the manuscript. HX analyzed and interpreted the data by consulting YZ who was professional with a strength in fMRI, as well as senior fellow apprentice YW and ZM who ever made a lot of contribution on collection of participants. SY, ZH, MP, GL, HD, and HW made substantial contributions in collection of image data. HX revised it critically for important intellectual content with suggestions from all authors. CW led the research training, along with NL, NZ, and YZ provided editing and writing assistance. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/neur.2021.755270/full#supplementary-material

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