c-erbB-2 expression in benign and malignant breast disease

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Summary
An antibody, 21N, raised against a synthetic peptide from the predicted sequence of the c-erbB-2 protein has been used immunocytochemically in a retrospective study of formalin fixed paraffin embedded breast biopsies. Fourteen out of 103 infiltrating ductal carcinomas exhibited positive membrane staining. Fifty-four of these tumours had lymph node involvement of which nine contained stained cells. These were all cases where the primary tumour was positive. In this series there was no correlation between c-erbB-2 overexpression and lymph node status. In five of the positive cases studied there was an associated in situ component which was also positively stained. Ten out of 24 pure intraduct carcinomas showed membrane staining, but none of the 149 benign conditions studied, which included 22 radial scars and 13 cases of atypical ductal proliferation, demonstrated the pattern of staining associated with overexpression. It is concluded that the c-erbB-2 protein is overexpressed in a minority (~14%) of infiltrating ductal carcinomas and only in cells that are cytologically malignant. Overexpression of c-erbB-2 is considered in relation to pathogenesis.

The c-erbB-2 gene is a normal cellular gene which is present on chromosome 17 (Fukushige et al., 1986) and encodes a protein which has close sequence homology with the epidermal growth factor receptor (Coussens et al., 1985; Bargmann et al., 1986a). This gene was originally identified in ethyl nitrosourea induced rat neuroblastoma cell lines as an activated oncogene called neu (Padhy et al., 1982; Bargmann et al., 1986b). Neu was shown to encode a 185,000 molecular weight protein, which from its structure (Coussens et al., 1985; Bargmann et al., 1986a,b) and localisation to the membrane (Drebin et al., 1984) suggests that it may be a receptor for an as yet unidentified growth factor. In rat cell lines the oncoenic-neu gene differs from the cellular-neu gene by a single base change (glutamic acid to valine) in the putative transmembrane region (Bargmann et al., 1986b).

Clinical interest in the gene in breast cancer was initiated by a report from Slamon’s group which indicated that amplification of this gene may be related to poor prognosis, with the claim that amplification alone had greater prognostic value than hormonal-receptor status in lymph node positive patients (Slamon et al., 1987). Work on cell lines had previously shown that in human breast carcinoma the c-erbB-2 gene could be overexpressed by a number of mechanisms, one of which was amplification (Kraus et al., 1987). More recently of perhaps more biological significance was the observation by Di Fiore et al. (1987) that c-erbB-2 needed to be overexpressed in NIH/3T3 cells (5–10 times) to be transforming. The fact that the cellular gene could be transforming without the point mutation supported the view that overexpression of this gene in breast carcinomas could have significance for the pathogenesis of this disease. Subsequent to the Slamon paper other groups, including our own, have confirmed the high incidence of c-erbB-2 amplification in breast cancer (van de Vijver et al., 1987; Venter et al., 1987; Varley et al., 1987; Zhou et al., 1987) and two of these have also indicated a correlation between amplification and poor prognosis (Varley et al., 1987) and between gene amplification and lymph node metastases (Zhou et al., 1987).

On the basis of the c-erbB-2 protein sequence, Gullick’s group produced two polyclonal antibodies to c-terminal peptides (Gullick et al., 1987). Using one of the antibodies it was demonstrated that amplification, which was present in 12 out of 36 tumours, correlated with overexpression of the c-erbB-2 protein, measured immunocytochemically and by Western blotting (Venter et al., 1987). In a subsequent publication using the same series of tumours we have shown that strong membrane staining using one of the antibodies, 21N, correlated with gene amplification (Gusterson et al., 1987). Measurement of EGF receptor and oestrogen receptor status in the same series showed no significant correlation with c-erbB-2 overexpression (Gusterson et al., 1987). This preliminary study, which was carried out on formalin fixed paraffin embedded material, indicated the utility of antibody 21N for retrospective analyses of tumours and benign breast diseases. The present paper describes an extensive immunocytochemical study of infiltrating ductal carcinomas, lymph node metastases, intraduct carcinomas and a range of benign breast disease, including 22 radial scars and 13 cases of atypical ductal proliferation.

Materials and methods

Tissues used
This study was carried out on 103 cases of infiltrating ductal carcinoma, selected into two groups on the basis of the lymph node status of the patients at the time of primary breast surgery and division of the patients into those that had relapsed within one year and those that were disease free after five years.

In addition 24 cases of intraduct carcinoma were examined and 150 examples of benign breast disease taken from material obtained at the Guildford Breast Screening Programme. Access to this material gave us the opportunity of examining 22 stellate scars and 13 cases of atypical ductal proliferation. All of the material was formalin fixed and paraffin embedded.

Antibody and immunocytochemistry
Antibody 21N, raised to a synthetic peptide at the c-terminal end of the c-erbB-2 protein, has been shown to precipitate a 190,000 mol. wt glycoprotein from human cells (Gullick et al., 1987). In a previous study it had been demonstrated that using antibody 21N membrane staining correlated with c-erbB-2 gene amplification (Gusterson et al., 1987). It has been shown by Dr N. Hynes that membrane staining with this antibody also correlates with increased c-erbB-2 protein expression (personal communication). On the basis of these

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data all of our analyses have been carried out using the antibody titrated against a tumour sample with a 12-fold gene copy number and strong membrane staining. In this study we are therefore aiming to identify the stages in tumour development at which overexpression occurs, the clinical significance of overexpression, and whether overexpression correlates with any identifiable morphological change.

Immunocytochemical staining was carried out using the indirect immunoperoxidase technique. Dewaxed sections were washed in PBS, and endogenous peroxidase blocked with 0.1% phenylhydroxyine hydrochloride in PBS for 5 min at room temperature. After washing in PBS, sections were exposed to the first antibody diluted to 3.3 μg ml⁻¹ in PBS for 1.5 h at room temperature. This concentration was based on a previous titration of the antibody against a tumour with a known 12-fold amplification of the c-erbB-2 gene (Gusterson et al., 1987).

After washing in PBS, sections were incubated with 100 μl sheep anti-rabbit peroxidase conjugate (Dako, High Wycombe, Bucks), at a dilution of 1 in 100, for 1 h. The colour was developed using diaminobenzidine (DAB) [10 mg DAB in 100 ml Tris buffer (pH 7.2) 0.1 M, 100 ml M.O, 66 μl H₂O₂] for 5 min. Sections were dehydrated, cleared and mounted in histomount (National Diagnostics). Control sections included omission of the first antibody and prior absorption of the antibody with peptide (1 mg ml⁻¹) at room temperature for 2 min.

Results

Morphological analyses

Of the 103 infiltrating carcinomas in this study there were 93 ductal carcinomas, 9 lobular carcinomas and 1 medullary. Strong membrane staining was seen in 14 of the ductal carcinomas together with a weak cytoplasmic blush (Figure 1a,b). In two of the lobular carcinomas there was cytoplasmic staining which was both diffuse and granular in type. The single medullary carcinoma was negative. Fifty-seven cases of infiltrating carcinoma had lymph node metastases. The primary tumours were stained in parallel with representative sections of the lymph nodes. Nine primary tumours of the original 14 positive cases had lymph node involvement

Figure 1 (a) shows strong membrane staining of an infiltrating ductal carcinoma. The apical membranes of the cells adjacent to the lumena are negative. (b) is a parallel section in which the antibody has been prior absorbed with the peptide – this extinguishes all staining (x 250).
and an identical pattern of staining was seen in the metastatic deposits. There was no evidence of positive staining in any of the metastases where the primary was negative (Table I). This led us to conclude that overexpression of this protein was not related to the metastatic process. Of the 14 positive cases, five had an in situ component and in all instances the staining pattern was identical with the infiltrating component, suggesting that the protein was overexpressed in a small percentage of tumours as a relatively early event in tumorigenesis. In all of the cases where there was strong membrane positivity the tumours were relatively homogeneously positive.

On the basis of the findings in these five intraduct carcinomas it was decided to extend the study to benign lesions and intraduct carcinoma biopsies from a breast screening centre where large numbers of small tumours and atypical lesions were available. The aim was to identify the earliest morphological changes associated with membrane positivity.

Ten of the 24 intraduct carcinomas from the screening centre were positively stained on the membrane. Where a single row of tumour cells lined the ducts the staining was concentrated on the lateral and basal membranes with absence of staining on the luminal surface (Figure 2). Where there was multilayering tumour cells were positive on their adjacent membranes. There was a clear delineation between the positive tumour cells and the adjacent normal cells. In no instances were cells strongly positive on the membranes that were not cytologically malignant. In some micropapillary intraduct carcinomas there was a loss of staining on the cells towards the lumen. This was associated with central necrosis and was interpreted as due to cellular degeneration.

The results of the benign lesions examined is shown in Table II. Only one case of apocrine metaplasia showed membrane staining and this was weak and on three cells in one cyst wall. The staining was interpreted as due to an artefact produced by fixation. The moderate cytoplasmic staining, which was a consistent finding in apocrine metaplasia in all of the material, when concentrated in a submembrane location as a result of fixation would give the impression of membrane staining. In one case of atypical epitheliosis there was marked cytoplasmic staining of cytologically abnormal cells which were present in a lobule, the significance of which is difficult to assess. There was no evidence of membrane positivity in any of the other cases examined including 22 stellate scars.

Clinical correlation

Data were available on 102 of the carcinoma cases for analysis using Chi-squared test with Yates correction. There was no correlation between the 21N staining and nodal status ($x^2=0.2$) or disease free interval ($x^2=0.4$) (Table III). When the data were analysed comparing 21N positive and 21N negative patients in relation to disease free survival and overall survival (Figures 3 and 4) overexpression of c-erbB-2 had no prognostic significance ($x^2=1.45$ and 3.37 respectively).

Discussion

This communication describes the distribution of the c-erbB-2 protein in the human breast. From these data we have concluded that overexpression of the c-erbB-2 protein can be visualized immunocytochemically as membrane staining in approximately 14% of breast carcinomas. In this small series there is no significant correlation of staining with tumour progression or lymph node involvement. There was also no relationship to either tumour grade or stage. Although not statistically significant, both the overall survival data and the disease free survival show a trend indicating that 21N positivity may be a poor prognostic factor. A much larger series with a longer follow-up is required to confirm this observation. The morphological studies would suggest that overexpression is an early event in tumorigenesis but in the classical two-stage carcinogenesis model would be considered as post initiation.

In this study overexpression of the c-erbB-2 protein has an apparently higher incidence (40%) in pure intraduct carcino-

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**Table I** 21N staining in primary tumours and lymph node metastases.

| Lymph node | + ve | - ve |
|------------|------|------|
| Primary tumour | 9    | 0    |
| - ve        | 0    | 45   |

**Figure 2** Sections show an intraduct carcinoma. There is strong staining of the pleomorphic tumour cells. Both the normal ductal cells and the surrounding stromal cells are negative. Note the absence of staining of the luminal membrane of the tumour cells (×250).

**Table II** 21N staining in intraduct carcinoma and benign breast disease.

|                | Positive | Weak cytoplasmic | Negative | Total |
|----------------|----------|------------------|----------|-------|
| Ductal carcinoma in situ | 10 (M)   | 8                | 6        | 24    |
| Epitheliosis     | 0        | 2                | 4        | 6     |
| Atypical epitheliosis | 1 (C)  | 3                | 9        | 13    |
| Radial scars     | 1 (C)    | 1                | 20       | 22    |
| Papilloma        | 0        | 0                | 6        | 6     |
| Adenosis         | 0        | 2                | 4        | 6     |
| Blunt duct adenosis | 0    | 0                | 14       | 14    |
| Sclerosing adenosis | 0    | 0                | 4        | 4     |
| Apocrine metaplasia | 1 (C-M)| 24               | 7        | 32    |
| Cysts (including duct ectasia and involution) | 0 | 0 | 22 | 22 |
| Fibroadenomatoid hyperplasia     | 0        | 1                | 2        | 3     |
| Fibroadenoma     | 0        | 5                | 16       | 21    |

M = Membrane; C = Cytoplasmic.
was increased (Gusterson, unpublished). There was not, however, an increase in the number of tumours with membrane staining. Under such conditions all of the cells in the section were stained including inflammatory cells, fibroblasts and smooth muscle in vessel walls. The strong membrane staining observed in some of the tumours, which correlates with overexpression, is also seen in cultured breast carcinoma cells where BT474 cells which have a 125-fold increase in mRNA for c-erbB-2 compared with MCF7 cells (Kraus et al., 1987) show strong membrane staining while MCF7 cells are only weakly cytoplasmically positive (Gusterson, unpublished).

A number of facts indicate that the c-erbB-2 protein may have an important role in the pathogenesis of human breast cancer. Firstly the structure of the protein and its membrane localisation suggest that it may be a growth factor receptor (Coussens et al., 1985; Bargmann et al., 1986a). Secondly the c-erbB-2 gene is consistently overexpressed and/or amplified in a significant proportion of infiltrating breast carcinomas in a number of series. The recent demonstration by Di Fiore et al. (1987) that the transforming ability of this gene is related to the level of expression of the protein further supports the view that the demonstration of an over expressed protein in a proportion of breast carcinomas may be related to a specific association with the mechanism of transformation rather than an incidental observation. Further support for a growth regulatory role of c-erbB-2 is the observation that a monoclonal antibody reactive with the extracellular domain exerted an anti-tumour effect on neu-transformed NIH3T3 cells (Drebin et al., 1986). In order to investigate the function of this protein further it will be necessary to produce antibodies that recognise the human external domain. Such antibodies may act as agonists or produce blocking of the putative binding site. They may also provide reagents for radio-localisation studies in a small but easily definable group of breast carcinomas. The absence of membrane staining in normal tissues indicates that this latter approach may be worth pursuing.

Although it is suggested that the c-erbB-2 protein may be a receptor for an unknown growth factor it is worth considering other possibilities. The sevenless gene in Drosophila encodes a putative transmembrane protein that has a similar structure and sequence homology to hormone receptors including the epidermal growth factor receptor (EGFR) (Hafen et al., 1987). The sevenless protein is required for the formation of the R7 photoreceptor in each ommatidium of the compound eye. It has been suggested that the protein controls this differentiation step by providing positional information, either by cell–cell interaction or through locally diffusible factors (Tomlinson et al., 1987). By analogy it could be speculated that the EGFR, which is known to be expressed on both proliferating and terminally differentiated cells (Gusterson et al., 1984) and the c-erbB-2 protein with its overexpression on adjacent cell membranes may also be involved in cell–cell interactions. A study of c-erbB-2 expression in the human embryo and in particular in the developing breast may provide further clues to the function of this molecule.

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