Risk of Mycoplasma Pneumoniae-Related Hepatitis in MP Pneumonia Pediatric Patients: A Predictive Model Construction and Assessment

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Abstract

Background

A predictive model for risk of *Mycoplasma pneumoniae* (MP)-related hepatitis in MP pneumonia pediatric patients can improve treatment selection and therapeutic effect. However, currently, no predictive model is available.

Methods

374 pneumoniae pediatric patients with/without serologically-confirmed MP infection and 93 health controls were enrolled. Logistic regressions were performed to identify the determinant variables and develop predictive model. Predictive performance and optimal diagnostic threshold were evaluated using area under the receiver operating characteristic curve (AUROC). Stratification analysis by age and MP-IgM titer was used to optimize model's clinical utility. An external validation set, including 84 MP pneumoniae pediatric patients, was used to verify the predictive efficiency. After univariate analysis to screen significant variables, monocyte count (MO), erythrocyte distribution width (RDW) and platelet count (PLT) were identified as independent predictors in multivariate analysis.

Results

We constructed MRP model: \( MO[^{10^9}/L] \times 4 + RDW[\%] - PLT[^{10^9}/L] \times 0.01 \). MRP achieved an AUROC of 0.754 and the sensitivity and specificity at cut-off value 10.44 were 71.72% and 61.00%, respectively in predicting MP-related hepatitis from MP pneumonia. These results were verified by the external validation set, whereas it merely achieved an AUROC of 0.540 in pneumonia without MP infection. The AUROC of MRP was 0.812 and 0.787 in infants and toddlers (0-36 months) and low MP-IgM titer subgroup (1:160-1:320), respectively. It can achieve an AUROC of 0.804 in infants and toddler with low MP-IgM titer subgroup.

Conclusions

MRP is an effective predictive model for risk of MP-related hepatitis in MP pneumonia pediatric patients, especially infants and toddlers with low MP-IgM titer.

Background

*Mycoplasma pneumoniae* (MP) has been considered as the predominant pathogenic bacteria species in community-acquired respiratory tract infection in children\(^1\). Recently, it was reported that extrapulmonary manifestations of MP pneumonia, such as aseptic meningitis, myocarditis, hepatitis and so on, occurred in up to 25% of individuals\(^2\), among which hepatitis is frequently ignored or at least downplayed. Actually, accumulated reports suggest that this MP-related hepatitis is commonly found in children and can manifest as asymptomatic elevation of liver enzymes, depressed multiple coagulation factors or cholestasis\(^3-9\). Therefore, ignoring of MP-related hepatitis will tend to delay the its diagnosis and result in the possibility of serious liver damage during MP pneumonia.
MP pneumonia is recognized as a systemic inflammation characterized as extending beyond the lung and causing other co-morbidities including hepatitis\textsuperscript{10}. Peripheral complete blood cell count (CBC) with differential can simultaneously change with ongoing inflammation process\textsuperscript{11}. In terms of hepatitis, red cell distribution width (RDW), one of the parameters in peripheral CBC with differential, is frequently served as biomarker to predict disease process and mortality in autoimmune hepatitis, hepatitis B virus-related cirrhosis and acute-on-chronic liver failure\textsuperscript{12-15}. Some scores incorporating the peripheral CBC with differential, such as neutrophil-to-lymphocyte ratio (NLR), platelet-to-lymphocyte ratio (PLR), monocyte-to-lymphocyte ratio (MLR) and systemic immune-inflammation index (S\textsubscript{i}) were also used to estimate the severity and outcome of patients with hepatitis B, C and E\textsuperscript{16-19}. Therefore, a novel predictive model derived from parameters of peripheral CBC with differential can also be expected to predict MP-related hepatitis. Additionally, in view of the limited facilities available for monitoring hepatic function in community clinics, this predictive model may assist the community clinicians to screen the high-risk of MP-related hepatitis and improve the treatment. However, currently, there is no available predictive model for clinical practice and the insight of peripheral CBC with differential in predicting is also worth exploring further.

In current study, we firstly investigated whether these common inflammation markers (such as NLR, PLR, MLR and S\textsubscript{i}) can effectively distinguish MP-related hepatitis from MP pneumonia pediatric patients, and then we constructed a new predictive model with capable of predicting MP-related hepatitis. the specificity and efficacy of the predictive model were also verified by validation set and controls. To our knowledge, it is the first report that a novel model, derived from systemic inflammation markers, was constructed and applied to predict the MP-related hepatitis.

**Methods**

**Participants**

The retrospective cohort study enrolled 675 participants. Among these, 562 participants were enrolled from Shandong Provincial Third Hospital, Cheeloo College of Medicine, Shandong University from August 2010 to August 2017. 113 participants were from Affiliated hospital of Yangzhou University from 2013.06 to January 2018. Exclusion criteria were as follows: (1) co-existent severe complication (eg. jaundice, convulsions and dehydration \textit{et al}); (2) overlapping pathogenic infection; (3) receiving antibiotic treatment before admission; (4) having history of hepatotropic and/or non-hepatotropic virus infection (Hepatitis A, B, C and E virus, Epstein-Barr virus and cytomegalovirus), hereditary metabolic liver diseases (Wilson disease and Hereditary hemochromatosis) and drug-associated hepatitis; (5) insufficient medical records. The diagnosis of MP-related hepatitis was established from three aspects: (1) serologically-confirmed MP infection diagnosis: a highly elevated serological MP-IgM titer $\geq 1:320$ (SeroDia\textsuperscript{TM} Mayo MAG\textregistered microparticle agglutination (MAG) assay kit, Fujirebio, Japan). In cases with a lower MP-IgM titer (1:160), a second sample was examined after an interval of 2-3 weeks to confirm a 4-fold titer increase; (2) diagnosis of hepatitis: patients, who had serum alanine aminotransferases (ALT) or aspartate aminotransferase (AST) levels greater than the upper normal limits (ALT: 40 U/L (female) and 50 U/L (male); AST: 35 U/L (female) and 40 U/L (male)) were regarded as having hepatitis; (3) excluding liver biochemistry abnormalities caused by hepatotropic and/or non-hepatotropic virus infection, hereditary metabolic liver disease or drug-associated hepatitis\textsuperscript{20-21}. 

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Study design

As shown in Fig 1, 124 participants were excluded and 551 participants were finally enrolled in the analysis, including 199 MP pneumonia pediatric patients, 175 pneumonia patients without MP infection, 93 healthy controls and 84 patients in an external validation set. Among 199 MP pneumonia patients, 99 patients were diagnosed as MP-related hepatitis and randomly divided into two sets: a training set (n = 55) and the validation set (n = 44), which was used to create and validate the model respectively. Otherwise, patient without presence of elevated serum ALT/ AST level were randomly classified as non-hepatitis set and served as control in both training (n = 50) and validating (n = 50) set at a ratio of 1:1 or so. 175 pneumonia patients without MP infection, including 85 patients diagnosed as hepatitis and 90 patients with non-hepatitis, were used to define the specificity of the model. 93 healthy control participants matching with patients by age and gender, accompanied with an external validation set (including 43 MP-related hepatitis and 41 non-hepatitis) were used to further validate the model’s application scope and predictive efficiency.

The following data were collected within 24 hours at the time of admissions: demographic data, liver biochemistry, immunological marker and peripheral CBC with differential. The liver biochemistry test included ALT, AST and total bilirubin (TBIL); immunological marker was MP-IgM titer; peripheral CBC with differential included WBC and their subtypes: lymphocytes, monocytes (MO), neutrophil, eosinophil, basophil proportion and count, RBC count and related information: hemoglobin, MCV, MCH, MCHC, RDW, PLT count and related information: platelet distribution width (PDW), mean platelet volume (MPV), platelet thrombocytocrit (PCT).

Laboratory evaluation

All tests performance and results verification were conducted in Shandong Provincial Third Hospital, Cheeloo College of Medicine, Shandong University and Affiliated hospital of Yangzhou University. Serological MP-IgM titer and peripheral CBC with differential, were measured from the finger prick blood samples, while liver biochemistry test was measured in samples from the peripheral vein blood samples taken at the same time.

Serum MP-IgM titers were detected by SeroDia™ MAG assay (Fujirebio, Japan). Peripheral CBC with differential was determined using an automated MEK-7222K haematology analyzer (Nihon Kohden, Japan). Liver biochemistry test including assessment ALT, AST levels and TBIL were performed using the Beckman Coulter AU 5800 analyzer (Beckman Coulter Diagnostics, USA). Reference ranges for ALT were 7-40 U/L (female) and 9-50 U/L (male), for AST were 13-35 U/L (female) and 15-40 U/L (male) and for TBIL were 5.1-28 mmol/L.

Statistical analysis

Statistical analysis was performed using SPSS software (version 22.0, SPSS Inc., USA). Data were expressed as Mean ± SD (or median, IQR) for quantitative variables and percentages for qualitative variables. The statistical significance between two groups was determined by t test or rank sum test for continuous variables and Chi-square test or Fisher's exact test for categorical variables. The only endpoints in this study is the diagnosis of MP-related hepatitis. A two-sided p<0.05 was considered statistical significance. Univariate analysis was firstly performed on all variables between patients with and without the study endpoint in training set. All selected variables were analyzed by multivariate analysis to assess the independent risk
factors and were used to construct the predictive model. Predictive efficiency was evaluated using area under the receiver operating characteristic curve (AUROC) and verified in validation set and external validation set. Stratification analysis was used to seek the optimum application scope of the model. The optimal cut-off values was determined using the Youden index based on sensitivity and specificity\textsuperscript{23}.

**Results**

**The participants’ characteristics**

The characteristics of the participants are shown in Table 1 and Table S1. This study includes 238 male and 229 females with the median age being 43 months, ranging from 2 to 158 months. In the training set, the MP-related hepatitis group showed significantly lower PLT count (Mean±SD: 290.13±96.71 vs. 340.44±89.79, \( p = 0.007 \)). Other parameters, such Moncyte count (median, interquartile range [IQR]: 0.6 (0.5, 0.8) vs. 0.4 (0.4, 0.6), \( p = 0.003 \)), RDW (12.0 (11.4, 13.0) vs. 11.5 (11.1, 12.1), \( p = 0.004 \)), PDW (17.34±0.91 vs. 16.80±1.00, \( p = 0.004 \)) and MPV (5.35±1.19 vs. 4.72±0.99, \( p = 0.004 \)) were significantly higher in the MP-related hepatitis group than the non-hepatitis group. No significant difference in MP-IgM titer, age, WBC count, lymphocyte count, neutrophil count, RBC count, hemoglobin and MCV et al. This followed the same trend as in the validation set and indicated that these two sets were well balanced in baseline characteristics.

**Candidate predictors selected and predictive model training**

The purposeful selection procedure began by a univariate analysis of each variable in the training set. A total of 7 variables (MP-IgM titer, MO count, WBC count, RDW, PLT count, PDW and MPV) were selected as determined by statistically significant from the original set of 25 variables in the training set using univariate analysis. Age was also selected by clinical experience. These 8 candidate predictors were further assessed using the multivariate logistic regression analysis. The MO count (odds ratio [OR]: 9.901, 95% confidence interval [CI]: 1.730-56.650, \( p = 0.01 \)), RDW (OR: 1.651, 95%CI: 0.999-2.728, \( p = 0.05 \)) and PLT count (OR: 0.995, 95%CI: 0.990-1.000, \( p = 0.044 \)) were significantly contributing to the prediction accuracy of the model (Table 2). Finally, the predictive model, consisting of 3-predictors with exact coefficient (Regression equation), was as follow: Regression equation = -5.532+2.293×MO \( [\times10^9/L] \)+0.501×RDW[%]-0.005×PLT \( [\times10^9/L] \). The receiver operating characteristic curves (ROC) were plotted in Fig 2A and B with AUROC of 0.749 and 0.711, respectively. We simplified Regression equation into a new applicable equation and named as MRP: MO\( [\times10^9/L] \)×4+RDW[%]-PLT\( [\times10^9/L] \)×0.01. AUROC of MRP in the training and validation set were 0.754 and 0.712, respectively (Fig2C and D). Both Regression equation and MRP were practically of similar accuracy, and MRP can be proposed as a novel predictive model for risk of MP-related hepatitis. In addition, AUROC of MRP in the independent external validation set was 0.728, which also confirms its predictive power (Fig 2E).

**Specificity and clinical utility of MRP**

Due to the derivation of the new predictive model MRP, the specificity needs to be defined carefully to guide MRP clinical application in pediatric pneumonia patients. Pneumonia pediatric patients without MP infection and health controls were introduced to test the specificity of MRP. As shown in Figure 3A, MRP was inefficient for distinguishing the hepatitis from pneumonia patients without MP infection with an AUROC of only 0.540
(95% CI: 0.463-0.616). Although, a slight improvement of MRP predictive efficacy, AUROC of 0.632 (95% CI: 0.557-0.703) was obtained in distinguishing hepatitis from health controls (Fig 3B), MRP predictive efficacy in hepatitis returned to be inefficient (with an AUROC of 0.547, 95%CI: 0.486-0.608) from the combining non-hepatis without MP infection and health controls (Fig 3C). These results indicated that MRP applied only to predict hepatitis in MP pneumonia pediatric patients rather than pneumonia caused by other pathogens.

Based on the ROC curves of MRP, the optimized cut-off value for prediction was set 10.44 to screen MP-related hepatitis (Table 3). Using cut-off value of 10.44 in both training and validation set, 71.72 % of MP pneumonia patients with MRP > 10.44 were verified as MP-related hepatitis (i.e, positive predictive value (PPV) = 64.55%), whereas an MRP ≤ 10.44 has a specificity 61.00 % and a negative predictive value (NPV) of 68.54 %. In the training set, for patients with MRP > 10.44, 40 of 60 (66.67 %) participants were MP-related hepatitis. Simultaneously, for patients with MRP ≤ 10.44, 30 of 45 (66.67 %) participants were non-hepatitis, only 15 (27.27 %) MP-related hepatitis were falsely classified. Meanwhile, the cut-off value for MAP was verified in the validation set. Similarly, in the higher MRP group (> 10.44), 31 of 50 (62.00 %) participants were MP-related hepatitis, while in the lower MRP group (≤ 10.44), 31 of 44 (70.45 %) participants were non-hepatitis, only 13 (29.55 %) MP-related hepatitis were classified incorrectly.

**Optimum applicatin scope of MRP**

To optimize MRP clinical application, stratification analysis by age and MP-IgM titer was performed to improve its diagnosis efficiency in MP-related hepatitis. As detailed in Table S2, stratification analysis by age and MP-IgM titer was performed to improve the predictive efficacy of MRP. Participants were commonly stratified in four subgroups: infants (0-12 months), toddlers (0-36 months), preschoolers (36-60 months) and school-ager (> 60 months). MRP predictive efficacy was strongest in the 0-12 (AUROC = 0.847, 95% CI: 0.727-0.968) and 0-36 (AUROC = 0.812, 95% CI: 0.719-0.904) months old subgroups (Fig 4A, B and C). Meanwhile, the patients were also divided into low (1:160-1:320) and high (1:640-1:1280) MP-IgM titer subgroup. AUROC of MRP in low titer subgroup (0.787, 95% CI: 0.713-0.861) was considerably greater than in high MP-IgM titer subgroup (0.653, 95% CI: 0.551-0.756) (Fig 4D and E). Moreover, MRP can achieve an AUROC of 0.804 (95% CI: 0.701-0.907) in infant and toddler (0-36 months) with low MP-IgM titer (1:160-1:320) subgroup, which indicates the optimum applicatin scope for MRP (Fig 4F).

**Discussion**

MP-related hepatitis, one of the extrapulmonary manifestations of MP pneumonia, has been receiving more attention recently, but for the clinical practice purposes, too little attention is paid for predicting risk of the MP-related hepatitis, especially in MP pneumonia pediatric patients. In this study, these common inflammation markers (including NLR, PLR, MLR and S) were shown as unable to predict MP-related hepatitis from MP pneumonia patients (data not shown). We constructed a novel predictive model, MRP, had comparable accuracy for predicting MP-related hepatitis in MP pneumonia pediatric patients (AUC=0.754), especially in infants and toddlers with low MP-IgM titer (AUC=0.804). The performance of MRP was also validated in an external validation set with similar accuracy (AUC=0.728).
Currently, it is recognized that MP-related hepatitis is closely relative to the systemic inflammatory response induced by MP infection\(^\text{10}\). Therefore, a comprehensive model composed of parameters derived from peripheral CBC with differential has potential to predict MP-related hepatitis. Among the MRP model, RDW has been identified as a biomarker in monitoring disease course and predicting the severity of hepatitis, such as autoimmune hepatitis\(^\text{12}\), hepatitis B-related hepatitis\(^\text{13-14,24-26}\) and nonalcoholic steatohepatitis\(^\text{27}\). RDW levels are usually elevated with progressive liver inflammation. A possible mechanism is that increased inflammatory cytokines in peripheral circulation and liver, such as TNF-\(\alpha\), IL-1\(\beta\) and IL-6, suppress erythrocyte maturation and accelerate the entry of newer, larger reticulocytes into the peripheral circulation, therefore causing the increased RDW\(^\text{28-29}\). Platelet and MO are also included in the MRP model. It was reported that MLR was correlated with risk of death in patients with severe exacerbation of chronic hepatitis B\(^\text{18}\) and can be used in the diagnosis of bacterial infection patients\(^\text{11}\). RDW-to-platelet ratio was a useful indicator for hepatic fibrosis regardless of etiology and can reflect the severity of hepatitis\(^\text{30}\). Therefore, these three parameters are all common biomarkers involved with systemic inflammation and hepatitis. Their combination in the MRP would offer lots of potential advantages in the prediction of MP-related hepatitis, which is caused by MP pneumonia inducing systemic inflammation.

In this study, we enrolled pneumonia patients without MP infection (including hepatitis and non-hepatitis) and healthy control to evaluate the specificity of the MRP model. The low AUC value (only 0.540, 95% CI: 0.463-0.616) suggested that MRP can not apply to screen the hepatitis in these pneumonia patients without MP infection, which in turn ensured that MRP merely can be used to screen hepatitis in MP pneumonia patients. The specificity of MRP application shows some advantages in MP-related hepatitis diagnosis and is more suitable for clinical using. Moreover, we provided an optimal application scope of MRP: the infants and toddlers patients (0-36 months) with low MP-IgM titer (1:160-1:320). Although, we found the largest AUC (0.847, 95% CI: 0.727-0.968) in the infants group (0-12 months), the relatively small-size of positive infants participants (\(n = 17\)) impacted the credibility of the MRP diagnosis efficiency. When we extended to include toddlers patients (age, 0-36 months), the positive participants (\(n = 34\)) size could sufficiently suppose the credibility of AUC (0.812, 95% CI: 0.719-0.904).

Although, some MP-related hepatitis patients (25.00 % in training set and 29.54 % in validation set) were eventually misclassified by the MRP model, this simple model was also an useful method to screen the high-risk of MP-related hepatitis in MP pneumonia pediatric patients, with positive predictive value of 64.55 % and negative predictive value of 68.54 % at 10.44 cut-off point. Actually, as a novel simple noninvasive predictive models, MRP has a fairly high diagnostic efficiency, which can achieve an AUC of 0.804 in the optimum application scope. The misdiagnosed patients can receive liver biochemistry test to exclude eventually and the price can be affordable. However, enhancing the sensitivity of MRP is still needed.

Macrolides are widely used as the first-line antibacterial agents in MP pneumonia treatment\(^\text{31}\). However, macrolide-induced hepatotoxicity, which showed a significantly elevated liver enzyme values or even cholestatic hepatitis, had been reported\(^\text{32-33}\). Pre-existing MP-related hepatitis may aggravate liver injury. Hence, it is necessary to evaluate the high-risk of MP-related hepatitis in MP pneumonia pediatric patients before the antibiotics application. The score of MRP, which can be calculated using only three parameters...
(MO, RDW and PLT) from the peripheral CBC with differential, can be used as “warning-signal” and provide a reference for selecting antibiotic and rational drug use through timely prediction of MP-related hepatitis.

With the facility of finger prick blood sampling, rapid measurement and reliable results, peripheral CBC with differential and MP-IgM titer measurement are commonly available in community clinics. Here the following flow chart can be suggested (see detail in Fig 5): (i) community clinicians diagnose the MP pneumonia by means of lung X-ray examination and serum MP-IgM titer measurement. Meanwhile, the peripheral CBC with differential was also measured; (ii) the score of the MRP was calculated; (iii) if the score of MRP is greater than 10.44, especially in the low MP-IgM titer of infants and toddlers, it is highly recommended for high-level pediatric care (eg. monitoring hepatic function, avoiding/decreasing usage of potentially hepatotoxic antibiotics and so on). Otherwise, they can receive routine usage of oral antibiotics and care at home.

Considering the limited facilities in community clinics and the difficulty in collecting peripheral vein blood samples in pediatric patients, it is convenient for community clinicians to evaluate the risk of MP-related hepatitis and help to select treatment and improve the therapeutic effect.

Our study has several limitations. Although a total of 551 participants from two medical centers were eventually enrolled for development and validation of MRP model, our model also needs to be further validated as yet with large prospective studies from multicenter. Additionally, this model developed from Asian race pediatric patients. Our result may not be representative of other race patients in general. We encourage our model to undergo validation in other races and also in centers outside China.

**Conclusions**

The MRP model was identified as an effective predictive model for risk of MP-related hepatitis in MP pneumonia pediatric patients, especially in infants and toddlers (age, 0-36 months) with low MP-IgM titer (1:160-1:320). What's more, we also provide a flow chart of prediction and management of MP-related hepatitis in MP pneumonia pediatric patients for more convenience in clinical application.

**Abbreviations**

MP, *Mycoplasma pneumoniae*; MO, monocyte; RDW, red cell distribution width; PDW, platelet distribution width; MPV, Mean platelet volume; PCT, Platelet thrombocytocrit; WBC, white blood cell; RBC, red blood cell; IQR, interquartile range; CI, confidence interval; PPV, positive predictive value; NPV, negative predictive value. HAV, hepatitis A virus; HBV, hepatitis B virus; HCV, hepatitis C virus; HEV, hepatitis E virus; EB, Epsten-Barr virus; CMV, cytomegalovirus; ROC, receiver operating characteristic; AUROC, area under the receiver operating characteristic curve; NLR, neutrophil-to-lymphocyte ratio; PLR, platelet-to-lymphocyte ratio; MLR, monocyte-to-lymphocyte ratio; SII, systemic immune-inflammation index; ALT, aminotransferases; AST, aspartate aminotransferase; TBIL, total bilirubin.

**Declarations**

Ethics approval and consent to participate
The study design was approved by the Ethics Committee of Shandong Provincial Third Hospital, Cheeloo College of Medicine, Shandong University and Affiliated hospital of Yangzhou University. Informed consents were obtained from parents/legal guardians for all participants and all methods were carried out in accordance with relevant guidelines and regulations.

**Availability of data and materials**

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

**Competing interests**

The authors declare that they have no competing interests.

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**Authors’ contributions**

Tao Li and Yuna Bi conceptualized and designed this study, drafted the initial manuscript. Tao Li is the recipients of the funding acquisition and supervised this study. Yuna Bi, Yan Ma, Jinhua Zhuo and Lili Zhang collected the demographics, laboratory variables and clinicopathological data from the medical records, carried out the formal analysis and contributed to the investigation. Liyan Yin, Hongling Sheng and Jie Luan assisted in the formal analysis and supervise this study. All authors had reviewed or edited the final manuscript and agree to be accountable for all aspects of the work.

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**Tables**

**Table. 1 Characteristics of the pediatric patients with MP pneumonia (n = 199)**
| Variables (n = 25) | Training Set (n = 105) | Validation Set (n = 94) |
|-------------------|------------------------|------------------------|
|                   | MP-related Hepatitis (n = 55) | Non-Hepatitis (n = 50) | P   | MP-related Hepatitis (n = 44) | Non-Hepatitis (n = 50) | P   |
| Gender            |                        |                        |     |                          |                        |     |
| Male, n (%)       | 28 (51.0)              | 31 (62.0)              | 0.253 | 20 (45.0)                | 16 (32.0)               | 0.181 |
| Female, n (%)     | 27 (49.0)              | 19 (38.0)              |     | 24 (55.0)                | 34 (68.0)               |     |
|                   |                        |                        |     |                          |                        |     |
| Anti-M.P IgM titer# |                      |                        |     |                          |                        |     |
| Low titer, n (%)  | 35 (63.6)              | 26 (52.0)              | 0.227 | 28 (63.6)                | 25 (50.0)               | 0.183 |
| High titer, n (%) | 20 (36.4)              | 24 (48.0)              |     | 16 (36.4)                | 25 (50.0)               |     |
|                   |                        |                        |     |                          |                        |     |
| Age, median (IQR) (Months) |            |                        | 0.717 |                          |                        | 0.883 |
| WBC, median (IQR) (%10⁹/L) |      |                        | 0.105 |                          |                        |     |
| Neutrophil proportion (%) |        |                        | 0.810 |                          |                        |     |
| Lymphocyte proportion (%) |         |                        | 0.695 |                          |                        |     |
| MO proportion (IQR) (%) |          |                        | 0.139 |                          |                        |     |
| Eosinophil proportion (IQR) (%) |       |                        | 0.203 |                          |                        |     |
| Basophil proportion (IQR) (%) |          |                        | 0.075 |                          |                        |     |
| Neutrophil, median (IQR) (%10⁹/L) |      |                        | 0.175 |                          |                        |     |
| Lymphocyte, median | 2.80 (1.70,4.90)       | 2.95 (1.88,3.90)       | 0.790 | 3.19 (2.5,4.41)          | 2.8 (2.1,3.85)          | 0.150 |
|                      | MO, median (IQR) (*10^9/L) | Eosinophil, median (IQR) (*10^9/L) | Basophil, median (IQR) (*10^9/L) | RBC count (*10^{12}/L) | Hemoglobin (g/L) | Hematokrit (%) | MCV (fL) | MCH, median (IQR) (pg) | MCHC, median (IQR) (g/L) | RDW, median (IQR) (%) | PLT (*10^9/L) | PDW (fL) | MPV (fL) | PCT |
|----------------------|-----------------------------|-------------------------------------|----------------------------------|-------------------------|-------------------|----------------|----------|-------------------------|------------------------|-----------------------|----------------|----------|---------|-----|
| (IQR)                | (0.5,0.8)                  | (0.00,0.10)                          | (0.00,0.10)                      | 4.59 ± 0.41             | 127.40 ± 10.68   | 37.88 ± 3.15   | 82.90 ± 5.03 | 27.2 (26.35,28.70)       | 331 (324,344)         | 12.0 (11.4,13.0)       | 290.13 ± 96.71 | 17.34 ± 0.91 | 5.35 ± 1.19 | 0.16 ± 0.09 |
| (10^9/L)             | 0.4 (0.4,0.6)              | 0.04 (0.00,0.10)                     | 0.00 (0.00,0.10)                | 4.52 ± 0.31             | 125.62 ± 9.50   | 37.28 ± 2.63   | 82.55 ± 3.79 | 27.8 (27.05,29.05)       | 335 (331,344)         | 11.5 (11.1,12.1)       | 340.44 ± 89.79 | 16.80 ± 1.00 | 4.72 ± 0.99 | 0.16 ± 0.09 |
|                      | 0.003*                     | 0.791                               | 0.262                            | 0.333                   | 0.371             | 0.295          | 0.695                 | 0.133                   | 0.071                 | 0.002*          | 0.007*          | 0.004*         | 0.004*         |
|                      |                            |                                     |                                  | 4.56 ± 0.42             | 125.05 ± 12.92      | 37.39 ± 2.91   | 82.65 ± 4.43 | 27.5 (26.48,28.28)       | 336 (329.25,341)      | 11.9 (11.4,12.58)       | 297.95 ± 99.43 | 17.23 ± 0.80 | 5.09 ± 0.94 | 0.16 ± 0.09 |
|                      |                            |                                     |                                  | 4.59 ± 0.34             | 128.40 ± 9.18       | 37.90 ± 2.54   | 82.19 ± 6.15 | 27.9 (27.1,28.9)         | 338.5 (336,343)       | 11.4(10.9,11.93)        | 337.48 ± 86.91 | 16.83 ± 0.80 | 4.70 ± 0.82 | 0.15 ± 0.04 |
|                      |                            |                                     |                                  |                         |                   |                |                      |                        |                      |                 |               |               |               |
|                      |                            |                                     |                                  |                         |                   |                |                      |                        |                      |                 |               |               |               |
| NOTE.                | * p < 0.05 for significance; # Low titer was defined as 1:160-1:320 and the high titer as 1:640-1:1280. |

Continuous variables are expressed in the terms of Means ± SD for normal data or median and interquartile range for non-normal data. Comparison tests are performed using *t* test or rank sum test as appropriate. Categorical variables are expressed as n (%) and compared by Chi-square or Fisher exact tests.
Table 2. Univariate and multivariate analysis of variables in Training set of the MP pneumonia pediatric patients

| Variables                              | MP-related Hepatitis vs Non-Hepatitis in Training set |       |       |       |       |       |
|----------------------------------------|--------------------------------------------------------|-------|-------|-------|-------|-------|
|                                        |                                                        | Univariate | Multivariate* |       |       |       |
|                                        |                                                        | P     | B     | S.E.  | Odds ratio | 95% CI    | P     |
| Intercept                              |                                                        | -     | -5.532| 3.313 | -       | -       | 0.095 |
| Age, median (IQR) (Months)             |                                                        | 0.300 | -     | -     | -       | -       | -     |
| Anti-M.P IgM titer                     |                                                        | 0.028*| -     | -     | -       | -       | -     |
| MO, median (IQR) (^10^9/L)             |                                                        | 0.008*| 2.293 | 0.890 | 9.901   | 1.730-56.650 | 0.010 |
| WBC, median (IQR) (^10^9/L)            |                                                        | 0.084§ | -     | -     | -       | -       | -     |
| Neutrophil I, median (IQR) (^10^9/L)   |                                                        | 0.545 | -     | -     | -       | -       | -     |
| Lymphocyte, median (IQR) (^10^9/L)     |                                                        | 0.193 |       |       |         |         |       |
| RDW, median (IQR) (%)                  |                                                        | 0.008*| 0.501 | 0.256 | 1.651   | 0.999-2.728 | 0.050 |
| PLT count (^10^9/L)                    |                                                        | 0.010*| -0.005| 0.003 | 0.995   | 0.990-1.000 | 0.044 |
| PDW (fL)                               |                                                        | 0.006*| -     | -     | -       | -       | -     |
| MPV (fL)                               |                                                        | 0.006*| -     | -     | -       | -       | -     |

**NOTE.** *p < 0.05 for significance; §*p = 0.084 for marginal significance;

#Logist(P)=-5.532+2.293×Mon[^10^9/L]+0.501×RDW[%]-0.005×PLT [×10^9/L].

Table 3. Accuracy of MRP in predicting MP-related Hepatitis and Hepatitis without MP infection in different sets of the pediatric patients
|                                      | MRP cut-off value | Hepatitis | Non-hepatitis | Sensitivity | Specificity | PPV    | NPV    |
|--------------------------------------|------------------|-----------|---------------|-------------|-------------|--------|--------|
| **Total # (N=199)**                  | >10.44           | 71        | 39            | 71.72%      | 61.00%      | 64.55% | 68.54% |
|                                      | ≤10.44           | 28        | 61            |             |             |        |        |
| **Training Set (N=105)**             | >10.44           | 40        | 20            | 72.73%      | 60.00%      | 66.67% | 66.67% |
|                                      | ≤10.44           | 15        | 30            |             |             |        |        |
| **Validation Set (N=94)**            | >10.44           | 31        | 19            | 70.45%      | 62.00%      | 62.00% | 70.45% |
|                                      | ≤10.44           | 13        | 31            |             |             |        |        |
| **Pneumonia pediatric patients without MP infection (N=175)** | >10.44           | 55        | 59            | 64.70%      | 34.44%      | 48.25% | 50.82% |
|                                      | ≤10.44           | 30        | 31            |             |             |        |        |

**NOTE.**#Consisting of the training set and validation set

**Figures**

[Diagram showing the derivation of MRP from various sets and conditions, including pediatric patients with MP pneumonia, pneumonia patients without MP infection, health controls, and MP pneumonia validation set inclusion/exclusion criteria.]
Figure 1

124 participants were excluded and 551 participants were finally enrolled in the analysis, including 199 MP pneumonia pediatric patients, 175 pneumonia patients without MP infection, 93 healthy controls and 84 patients in an external validation set. Among 199 MP pneumonia patients, 99 patients were diagnosed as MP-related hepatitis and randomly divided into two sets: a training set (n = 55) and the validation set (n = 44), which was used to create and validate the model respectively. Otherwise, patient without presence of elevated serum ALT/AST level were randomly classified as non-hepatitis set and served as control in both training (n = 50) and validating (n = 50) set at a ratio of 1:1 or so. 175 pneumonia patients without MP infection, including 85 patients diagnosed as hepatitis and 90 patients with non-hepatitis, were used to define the specificity of the model. 93 healthy control participants matching with patients by age and gender, accompanied with an external validation set (including 43 MP-related hepatitis and 41 non-hepatitis) were used to further validate the model's application scope and predictive efficiency.

Figure 2

The receiver operating characteristic curves (ROC) were plotted in Fig 2A and B with AUROC of 0.749 and 0.711, respectively. We simplified Regression equation into a new applicable equation and named as MRP: M0[^10^9/L]×4+RDW[%]-PLT[^10^9/L]×0.01. AUROC of MRP in the training and validation set were 0.754 and
0.712, respectively (Fig2C and D). Both Regression equation and MRP were practically of similar accuracy, and MRP can be proposed as a novel predictive model for risk of MP-related hepatitis. In addition, AUROC of MRP in the independent external validation set was 0.728, which also confirms its predictive power (Fig 2E).

**Figure 3**

MRP was inefficient for distinguishing the hepatitis from pneumonia patients without MP infection with an AUROC of only 0.540 (95% CI: 0.463-0.616). Although, a slight improvement of MRP predictive efficacy, AUROC of 0.632 (95% CI: 0.557-0.703) was obtained in distinguishing hepatitis from health controls (Fig 3B), MRP predictive efficacy in hepatitis returned to be inefficient (with an AUROC of 0.547, 95% CI: 0.486-0.608) from the combining non-hepatis without MP infection and health controls (Fig 3C). These results indicated that MRP applied only to predict hepatitis in MP pneumonia pediatric patients rather than pneumonia caused by other pathogens.
Figure 4

Participants were commonly stratified in four subgroups: infants (0-12 months), toddlers (0-36 months), preschoolers (36-60 months) and school-ager (> 60 months). MRP predictive efficacy was strongest in the 0-12 (AUROC = 0.847, 95% CI: 0.727-0.968) and 0-36 (AUROC = 0.812, 95% CI: 0.719-0.904) months old subgroups (Fig 4A, B and C). Meanwhile, the patients were also divided into low (1:160-1:320) and high (1:640-1:1280) MP-IgM titer subgroup. AUROC of MRP in low titer subgroup (0.787, 95% CI: 0.713-0.861) was considerably greater than in high MP-IgM titer subgroup (0.653, 95% CI: 0.551-0.756) (Fig 4D and E). Moreover, MRP can achieve an AUROC of 0.804 (95% CI: 0.701-0.907) in infant and toddler (0-36 months) with low MP-IgM titer (1:160-1:320) subgroup, which indicates the optimum applicatin scope for MRP (Fig 4F).
(i) community clinicians diagnose the MP pneumonia by means of lung X-ray examination and serum MP-IgM titer measurement. Meanwhile, the peripheral CBC with differential was also measured; (ii) the score of the MRP was calculated; (iii) if the score of MRP is greater than 10.44, especially in the low MP-IgM titer of infants and toddlers, it is highly recommended for high-level pediatric care (eg. monitoring hepatic function, avoiding/decreasing usage of potentially hepatotoxic antibiotics and so on). Otherwise, they can receive routine usage of oral antibiotics and care at home. Considering the limited facilities in community clinics and the difficulty in collecting peripheral vein blood samples in pediatric patients, it is convenient for community
clinicians to evaluate the risk of MP-related hepatitis and help to select treatment and improve the therapeutic effect.

**Supplementary Files**

This is a list of supplementary files associated with this preprint. Click to download.

- [TableS1.docx](#)
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