HEPATOPROTECTIVE EFFECTS OF CORILAGIN FOLLOWING HEMORRHAGIC SHOCK ARE THROUGH AKT-DEPENDENT PATHWAY

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ABSTRACT—Corilagin, a component of Phyllanthus urinaria extract, possesses antioxidant, thrombolytic, antatherogenic, and hepatoprotective properties, but the mechanism underlying these effects remains unclear. Previous studies showed that the Akt (protein kinase B) signaling pathway exerts anti-inflammatory and organ protective effects. The aim of this study was to investigate the mechanism of action of corilagin and determine whether these effects are mediated through the Akt-dependent pathway in a trauma-hemorrhagic shock-induced liver injury rodent model. Hemorrhagic shock was induced in male Sprague-Dawley rats; mean blood pressure was maintained at 35 mm Hg to 40 mm Hg for 90 min, followed by fluid resuscitation. During resuscitation, three doses of corilagin alone (1 mg/kg, 5 mg/kg, or 10 mg/kg, intravenously) were administered. Furthermore, a single dose of corilagin (5 mg/kg) with and without Wortmannin (1 mg/kg, PI3K inhibitor), Wortmannin alone, or vehicle was administered. Twenty-four hours after resuscitation, plasma alanine aminotransferase and hepatic aspartate aminotransferase concentration and hepatic parameters were measured. One-way ANOVA was used for statistical analysis. Hepatic myeloperoxidase activity and the concentrations of plasma alanine aminotransferase and aspartate aminotransferase, interleukin-6 (IL-6), tumor necrosis factor-α (TNF-α), intercellular adhesion molecule-1, and cytokine-induced neutrophil chemoattractant-1 (CINC-1) and CINC-3 increased following hemorrhagic shock. These parameters were significantly attenuated in corilagin-treated rats following hemorrhagic shock. Hepatic phospho-Akt expression was also higher in corilagin-treated rats than in vehicle-treated rats. The elevation of phospho-Akt was abolished by combined treatment with Wortmannin and corilagin. Our results suggest that corilagin exerts its protective effects on hemorrhagic shock-induced liver injury, at least, via the Akt-dependent pathway.

KEYWORDS—Akt, corilagin, hepatic injury, trauma-hemorrhage

INTRODUCTION

Traumatic injury with hemorrhagic shock is an important clinical issue (1). Severe hemorrhagic shock results in circulating blood supply insufficiency, leading to tissue damage and vital organ dysfunction (2). The liver is a major solid intra-abdominal organ that is supplied with rich blood, and can cause severe hepatic injury following hemorrhagic shock (3). Previous studies showed that hemorrhagic shock results in high production of proinflammatory mediators and cytokines, such as interleukin-6 (IL-6), tumor necrosis factor-α (TNF-α), cytokine-induced neutrophil chemoattractant-1 (CINC-1), and CINC-3 (4).

Following hemorrhagic shock, neutrophils are activated by pro-inflammatory chemokines/cytokines (4). The activated neutrophils are recruited to adhesion molecules and induce severe organ injury (4). Intercellular adhesion molecule-1 (ICAM-1) is upregulated after trauma-hemorrhage, which may enhance neutrophil adhesion to the vascular endothelium (5). IL-6 and TNF-α also play important roles in neutrophil infiltration and hepatic injury (4).

The phosphoinositide-3-kinase (PI3K)/Akt (protein kinase B) signaling pathway regulates cell survival in response to various injuries (6, 7). Previous studies showed that activation of the PI3K/Akt pathway decreases the overproduction of proinflammatory mediators and adhesion molecules to protect organs from injury (5).

Phyllanthus urinaria is a popular herb belonging to the Phyllanthaceae family and has been used in traditional anti-diabetic, antiviral, and gastrointestinal treatments (8). In addition, P. urinaria extracts also possess antioxidant and anti-inflammatory activities (9). Previous studies reported that P. urinaria extracts can inhibit the phagocytic activity of neutrophils, particularly by suppressing reactive oxygen species production (10) and reducing TNF-α expression in radiation-induced brain inflammation (11). In addition, P. urinaria has shown antiviral potential against A16, enterovirus 71, and coxsackievirus A16 infections (12). Corilagin is an important component of P. urinaria extract. Previous reports indicated that corilagin has many medical benefits, including antihypertensive, anticancer, antihyperalgesic, thrombolytic, and hepatoprotective properties (13–16).

Although various protective effects of corilagin have been reported, it has not been examined if this herb has any salutary effects following hemorrhage. Furthermore, the precise mechanism by which this agent produces organ protection effects.

The authors report no conflicts of interest.

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remains unknown. We hypothesized that the protective effects of corilagin after hemorrhagic shock are exerted through the Akt-dependent pathway. Experimental animals were treated with corilagin alone and in combination with the PI3K inhibitor Wortmannin following hemorrhagic shock. The effects of these treatments were examined and the plasma concentrations of alanine aminotransferase (ALT) and aspartate aminotransferase (AST), hepatic tissue myeloperoxidase (MPO) activity, pro-inflammatory mediator levels, and Akt expression were determined following hemorrhagic shock.

MATERIALS AND METHODS

Animals

Male Sprague–Dawley rats were used in this study. Rats were obtained from the National Science Council Experimental Animal Center of Taiwan. All experimental procedures and protocols were approved by the Institutional Animal Care and Use Committee of Chang Gung Memorial Hospital. The animal experiments followed the guidelines of the Animal Welfare Act and The Guide for Care and Use of Laboratory Animals from the National Institutes of Health.

Experimental groups

Thirty-six male Sprague–Dawley rats (275–325 g) were randomly assigned to six groups (n = 6/group) for evaluation of the dose–response effect of corilagin on hepatic injury following hemorrhagic shock. Groups receiving corilagin (0 mg/kg, 1 mg/kg, 5 mg/kg, or 10 mg/kg) and sham groups were also included in the initial studies. An additional 36 male Sprague–Dawley rats were also randomly divided into six groups (n = 6/group). Rats in the hemorrhagic shock groups were divided into four groups and treated with corilagin (5 mg/kg), corilagin and Wortmannin (1 mg/kg, intravenously), Wortmannin alone, or vehicle; sham groups were also included.

Rat hemorrhagic shock and resuscitation model

All animals were placed in individual air-conditioned cages (humidity: 70–75%) with controlled temperature (24–25°C) and lighting (12:12 light–dark cycle: lights on from 06:00 to 18:00) in an animal house. One week before the experiments, all experimental animals were sent to the animal house and allowed to adapt to the environment. Basal feed and water was provided. Before the experiment, male Sprague–Dawley rats were fasted overnight but allowed free access to water. Hemorrhagic shock and resuscitation was performed as described previously (4).

Briefly, a 5-cm midline abdominal laparotomy was performed on anesthetized rats by isoflurane inhalation to induce tissue trauma, and then the abdominal wound layers were sutured in layers. Under anesthesia, polyethylene catheters (PE-50; BD Biosciences, Franklin Lakes, NJ) were placed in the right femoral vein and bilateral femoral arteries, fixed, and the incision sites were closed. Before the experiment, male Sprague–Dawley rats were fasted overnight but allowed free access to water. Hemorrhagic shock and resuscitation was performed as described previously (4).

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Assessment of hepatic injury

Blood samples were collected in a heparin tube and the plasma was separated by centrifugation at 24 h following trauma-hemorrhage or sham surgery. Hepatic injury was determined by measuring plasma ALT and AST levels using a colorimetric analyzer (Dri-Chem 3000, Fuji Photo Film Co, Tokyo, Japan).

Determination of MPO activity

The MPO activity assay in homogenates of rat liver tissues was measured as described previously (4). Frozen liver tissue samples were thawed and suspended in pH 6.0 phosphate buffer containing 0.5% hexadeccyltrimethylammonium bromide (Sigma, St. Louis, MO). All samples were sonicated on ice, centrifuged at 12,000 × g for 15 min at 4°C, and then aliquots were transferred into phosphate buffer (pH 6.0) containing 0.167 mg/mL O-dianisidine hydrochloride and 0.0005% hydrogen peroxide (Sigma). The change in absorbance was measured at a wavelength of 460 nm with a spectrophotometer for 5 min. MPO activity was calculated using a standard curve generated using human MPO (Sigma), and values were normalized according to the protein concentration.

Measurement of CINC-1, CINC-3, ICAM-1, IL-6, and TNF-α levels

Liver tissues were homogenized in potassium phosphate buffer (1:100 weight/volume; pH 7.4) containing protease inhibitors (Complete Protease Inhibitor Cocktail; Boehringer, Mannheim, Germany). The homogenates were centrifuged at 2,000 × g (4°C) for 20 min and the supernatants were analyzed for the presence of IL-6, TNF-α, ICAM-1, CINC-1, and CINC-3 using ELISA kits (R&D Systems, Minneapolis, Minn) according to the manufacturer’s instructions, and as described previously (4). Aliquots of the supernatant were used to determine protein concentration using the Bio-Rad DC Protein Assay (Bio-Rad, Hercules, Calif.).

Western blot assay

Rat liver tissues were homogenized in buffer as described previously (4). Protein aliquots were used to determine protein concentration using the Bio-Rad DC Protein Assay (Bio-Rad). Samples were mixed with ≥ 4x sample buffer, electrophoresed on sodium dodecyl sulfate-polyacrylamide gels, and electro-phoretically transferred onto nitrocellulose membranes. The membranes were incubated with antibodies for total Akt protein, phosphorylase-Akt (Ser473) (Cell Signaling Technology, Beverly, Mass), or GAPDH (Abcam, Cambridge, UK) overnight at 4°C. The membranes were then incubated with horseradish peroxidase-conjugated goat antimouse antibody or goat antirabbit antibody for 1.5 h at room temperature. After the final wash, the membranes were probed by enhanced chemiluminescence (Amersham, Amersham, UK) and exposed to film for autoradiography.

Statistical analysis

The InStat 3.0 biostatistics program (Graph Pad Software Inc, San Diego, Calif) was used for statistical analysis. The results are presented as the mean ± standard error of the mean (SEM). The data were analyzed using one-way analysis of variance and the Tukey test, and differences were considered significant when P < 0.05.

RESULTS

Dose–response effects of corilagin on plasma AST and ALT levels

Hemorrhagic shock caused a significant increase in plasma AST and ALT levels at 24 h after resuscitation (Fig. 1, A and B). Treatment with corilagin at 1 mg/kg, 5 mg/kg, or 10 mg/kg was used to evaluate the beneficial effects of corilagin on the attenuation of liver injury following hemorrhagic shock. As shown in Figure 1, 1 mg/kg, 5 mg/kg, or 10 mg/kg corilagin administration had beneficial effects. However, the effects of corilagin were equivalent when administered at a dose of 5 mg/kg or 10 mg/kg.

Alteration in plasma AST and ALT levels

At 24 h after sham operation, there was no significant difference in plasma AST and ALT levels between vehicle- and corilagin-treated groups (Fig. 2, A and B). Plasma AST and ALT levels were significantly increased in trauma-
hemorrhaged rats. Treatment with corilagin (5 mg/kg) attenuated the hemorrhagic shock-induced increase in plasma AST and ALT levels. We next investigated whether the protective effects of corilagin in attenuating hepatic injury following hemorrhagic shock were mediated via Akt-mediated activity. In the group of corilagin-treated trauma-hemorrhaged rats cotreated with the PI3K inhibitor Wortmannin, the corilagin-induced reduction in plasma AST and ALT levels was abolished (Fig. 2).

**Alteration in hepatic MPO activity**

As shown in Figure 3, hepatic MPO activity was observed in sham-operated or trauma-hemorrhaged animals with and without corilagin treatment. In sham-operated rats, corilagin did not alter hepatic MPO activity. Hemorrhagic shock significantly increased hepatic MPO activity in vehicle-treated animals. Furthermore, treatment with corilagin attenuated the increase in hepatic MPO activity. Co-administration of the PI3K inhibitor Wortmannin with corilagin prevented corilagin-mediated attenuation of hepatic MPO activity after hemorrhagic shock.

**Alteration of hepatic CINC-1, CINC-3, and ICAM-1 levels**

As shown in Figures 4A, B and 5, no significant change in hepatic CINC-1, CINC-3, and ICAM-1 levels between the vehicle- and corilagin-treated sham groups were observed. Hemorrhagic shock significantly elevated hepatic CINC-1, CINC-3, and ICAM-1 levels. The increase in hepatic CINC-1, CINC-3, and ICAM-1 levels was reduced by corilagin treatment. The corilagin-mediated reduction in CINC-1, CINC-3, and ICAM-1 levels was abolished by co-administration of the PI3K inhibitor Wortmannin.
There was no significant difference in hepatic IL-6 and TNF-α expression between the sham groups (Fig. 6). Following hemorrhagic shock, hepatic IL-6 and TNF-α concentrations were significantly higher in vehicle-treated rats than in sham animals (Fig. 6, A and B). Treatment with corilagin (5 mg/kg) attenuated the hemorrhagic shock-induced increase in IL-6 and TNF-α expression. Co-administration of corilagin with the PI3K inhibitor Wortmannin prevented the corilagin-induced decrease in IL-6 and TNF-α concentrations.

Activity of hepatic Akt protein

As shown in Figure 7, hepatic Akt protein expression between the sham and trauma-hemorrhaged rats was not significantly different. Phosphorylation of Akt is typically used to determine Akt activation. The Akt phosphorylation status was significantly decreased after hemorrhagic shock, and treatment of corilagin restored Akt phosphorylation to the observed levels in sham animals. The increase in phosphorylated-Akt induced by corilagin was abolished when the PI3K inhibitor Wortmannin was co-administered with corilagin.

DISCUSSION

In the present study, we investigated the protective effects of corilagin on rodent hepatic injury following hemorrhagic shock and evaluated the role of the Akt-dependent pathways in corilagin-mediated hepatoprotection. Hepatic injury-related parameters, including plasma AST and ALT concentrations, hepatic MPO activity, and CINC-1, CINC-3, ICAM-1, IL-6, and TNF-α levels were significantly elevated at 24 h after
ICAM-1 and CINC-3 are potent chemoattractants for neutrophils and have been correlated with tissue MPO activity in injured tissue (17). Our results showed that hemorrhagic shock induced a marked increase in hepatic MPO activity and CINC-1 and CINC-3 levels, which were attenuated by corilagin treatment. In addition, neutrophil migration requires the interaction with multiple adhesion molecules. ICAM-1 is an important adhesion molecule in neutrophils for adhesion onto the vascular endothelium after organ injury (5). In this study, hemorrhagic shock resulted in a significant increase in hepatic ICAM-1 levels, which was accompanied by hepatic MPO activity elevation. Furthermore, MPO activity and ICAM-1 levels were alleviated in corilagin-treated hemorrhagic shock rats.

IL-6 and TNF-α are important pro-inflammatory mediators and their expression reflects the severity of tissue injury and influence on organ function (4). A previous study showed that liver injury leads to marked increases in hepatic IL-6 and TNF-α (4). Furthermore, adhesion molecule expression and cytokine release are related to IL-6 and TNF-α stimuli (4). In this study, we found that the hepatic IL-6 and TNF-α expression in rats was elevated following hemorrhagic shock. Treatment of the animals with corilagin significantly decreased hepatic IL-6 and TNF-α concentrations. These findings suggest that corilagin reduces inflammatory cytokine and adhesion molecules expression to regulate hepatic inflammation.

The PI3K/Akt pathway is an important intracellular signaling pathway involved in cell survival and tissue function maintenance (7). Previous studies have reported that activation of the PI3K/Akt pathway protects organs against hypoxia or ischemia injury (6). Additionally, PI3K influences neutrophil function and infiltration in the injured liver (18). Previous reports have also indicated that up-regulation of the PI3K/Akt signaling pathway decreased neutrophil accumulation and cytokine/adhesion molecule overproduction after hemorrhagic shock (19). Our previous studies also suggested that the PI3K/Akt pathway plays a critical role in organ protection following hemorrhagic shock (5).

*Phyllanthus urinaria,* a traditional herbal medicine, has been reported to possess different biomedical characteristics including antipathogenic, anticancer, and antihepatotoxicity effects (12, 14, 16). Studies showed that *P. urinaria* extracts have numerous biological activities including anti-oxidative, anti-inflammatory, antiapoptotic, and hepatoprotective effects (16). Corilagin, an important *P. urinaria* extract, has been reported to exhibit antiviral activity against hepatitis B virus infection (20) and antitumor activity against Hep3B hepatoma in mice (21). Corilagin was also shown to alleviate post-parasiticide liver fibrosis in a *Schistosomiasis*-infected animal model (22). In addition, corilagin protected against galactosamine/lipopolysaccharide-induced liver injuries through anti-oxidative stress and antilipid peroxidation effects, as well as by inhibiting apoptosis (23). In view of the above reported effects, we evaluated whether the PI3K/Akt pathway is involved in mediating the hepatoprotective effects following corilagin treatment. Our results demonstrated that the corilagin-induced attenuation of hepatic enzymes and pro-inflammatory mediators were blocked by the PI3K inhibitor Wortmannin. The elevation in Akt phosphorylation by corilagin following trauma-hemorrhagic shock was abolished by co-administration of Wortmannin. The current results indicate that the hepatic protective effects of corilagin are in part mediated by the Akt-dependent pathway.

It should be noted that the present study examined the mechanism of the salutary effects of corilagin following hemorrhagic shock at a single time, i.e., 24 h after treatment. Thus, it remains unclear if the same or different mechanisms are
involved in the early or late protective effects by this agent. Furthermore, although we examined the mechanism of corilagin following hemorrhagic shock, we did not examine if this agent had any protective effects on survival in this study not did we examine if it had protective effects on hepatic architecture, function of other organs such as the heart, intestine and brain, or functional aspects of the liver following hemorrhagic shock. However, since the liver enzymes did decreasing with corilagin it would suggest that there were some protective effects on hepatic architecture. Nonetheless, we did not measure any functional aspects such as synthesis of acute phase synthesis of the liver under those conditions and thus it remains unclear if this agent also had salutary effects of the functional aspects of the liver. Our primary focus in this study was on examining the potential mechanism of hepatoprotection by corilagin and we hope to examine the other aspects listed above in our future studies. Wortmannin can have effects on other signaling pathways such as mammalian target of rapamycin, p38 mitogen-activated protein kinases, polo-kinase, and insulin receptor activation, depending on the concentration and cell type (24, 25). These actions might contribute to the hepatoprotective effects of corilagin following trauma-hemorrhagic shock. In addition, corilagin may function by different mechanisms/pathways. Corilagin is a weak carbonic anhydrase inhibitor (26). It remains unknown if the effect is related to p-Akt. Additional studies are needed to precisely elucidate the mechanism by which corilagin attenuates hepatic injury following trauma-hemorrhagic shock.

In summary, we determined the hepatoprotective mechanism of corilagin from *P. urinaria* extract in a trauma-hemorrhagic shock rodent model. Corilagin treatment effectively ameliorated neutrophil accumulation, production of pro-inflammatory mediators, and elevation of enzymes released following hepatic injury. When Akt activation was blocked, the hepatoprotective effects of corilagin were abolished. Our results suggest that the Akt-dependent pathway plays a critical role in corilagin-mediated hepatoprotection following trauma-hemorrhagic shock. These findings indicate that corilagin from *P. urinaria* extract is a potential therapeutic adjunct that can be used for organ protection following low flow conditions.

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