Primary pulmonary undifferentiated pleomorphic sarcoma (PPUPS)

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ABSTRACT

Undifferentiated pleomorphic sarcoma (UPS) is a high-grade pleomorphic neoplasm with no identifiable line(s) of differentiation using currently available diagnostic techniques. Therefore, it is essentially a diagnosis of exclusion, which requires generous tissue sampling, adequate contextually interpreted immunohistochemistry, and relevant molecular studies. UPS is a common soft tissue sarcoma (historically one of the entities referred to as malignant fibrous histiocytoma (MFH)), which can develop in various organs, but lung involvement is usually due to metastasis. Primary Pulmonary UPS (PPUPS) is exceptionally rare and here we present a 66-year-old man who presented with anemia and weight loss, found to have a 17 cm right lung mass with invasion to the chest wall and diaphragm. Extensive sampling and immunohistochemistry studies failed to reveal any line of differentiation. Upon exclusion of a possible extrapulmonary origin, a diagnosis of PPUPS was rendered. In addition, we reviewed all 84 previously reported cases of PPUPS/PPMFH in the literature since 1979 and summarized the clinical information.

Keywords
Lung; Lung Neoplasms; Sarcoma; Soft Tissue Neoplasms

INTRODUCTION

Undifferentiated pleomorphic sarcomas (UPS), one of the entities previously included in the effete diagnosis of malignant fibrous histiocytoma (MFH), most commonly occurs in deep soft tissue of extremities (thigh) of elderly patients (average 50-70 years). So-called MFH was in the past thought to be of fibrohistiocytic origin; however, further studies (electron microscopy, immunohistochemical techniques, or molecular studies) failed to demonstrate evidence of “fibrohistiocytic” differentiation. Additionally, other unrelated poorly differentiated sarcoma was included in this entity, compounding the non-specific and confusing nature of this diagnostic term. MFH was declassified by World Health Organization (WHO) in 2012, and is no longer listed as a diagnostic entity. Many tumors previously included as MFH have been reclassified as “undifferentiated pleomorphic sarcoma”.\textsuperscript{1}

Primary sarcomas in lung are rare (less than 0.5% of all lung cancers)\textsuperscript{2}, and primary pulmonary undifferentiated pleomorphic sarcomas (PPUPS) are one of the least common primary lung sarcomas, with 84 reported cases in the English literature (Table 1). Weiss SW and Enzinger FM first described soft tissue MFH of 200 cases in 1978.\textsuperscript{3} A year later, Bedrossian et al.\textsuperscript{4} reported the first case of primary pulmonary MFH (PPMFH) in a 51-year-old man.

Yousem and Hochholzer\textsuperscript{5} reported the most extensive series of primary malignant fibrous

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histiocytomas in the lung in 1987 with 22 cases (patient age range of 18 to 80 years). Previous irradiation is a known pathogenic risk factor for soft tissue UPS. Similarly, few reports are available in the literature regarding patients who develop PPUPS years after radiation therapy for another tumor.  

It is unclear whether de-novo PPUPS and radiation associated undifferentiated pleomorphic sarcomas represent the same entity.

Clinical presentations in pulmonary tumors, as with other types of lung cancers, depends more on the tumor location rather than the histological type. Primary pulmonary sarcomas often present as a large peripheral or hilar well-circumscribed mass, and may present as an endobronchial tumor in 10% of cases. The majority of patients present with symptoms of cough, chest pain, hemoptysis, or dyspnea. Radiologic findings can show a solitary mass with or without post-obstructive effects (recurrent pneumonia, bronchiectasis, lobar or segmental atelectasis), and in some cases with extraluminal growth and/or local invasion into adjacent structures.

| Tumor                                | Immunohistochemistry                                                                 | Molecular                                                                                     |
|--------------------------------------|--------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------|
| Sarcomatoid Carcinoma (SC)           | P63+, P40+, PanKeratin+, TTF1+, epithelial markers (MOC31, BerEp4, BG8, B72.3, monoclonal CEA)+ | Gains at chromosomes 8q, 7, 1q, 3q, and 19. KRA5 mutation, EGFR mutation                      |
| Sarcomatoid Mesothelioma             | WT1+, CK5/6+, D2-40+, calretinin+                                                    | Inactivation of CDKN2A at 9p21 on PCR                                                          |
| Angiomatoid fibrous histiocytoma (AFH)| Desmin+/-, CD68+/-, EMA+/-                                                          | EWSR1-CREM1, EWSR1-ATF1, or FUS-ATF1 fusion                                                    |
| Synovial sarcoma (SS)                | TLE1+, Keratin+, EMA+, S100+/-, CD56+, CD99+, Calretinin+/-                         | t(X;18) involving SS18 (SYT) gene                                                            |
| Epithelioid Sarcoma (ES)             | Loss INI, EMA+, Keratin+/-, CD34+-                                                  | SMARCB1 (INI1) gene alterations on (22q11)                                                    |
| Dedifferentiated liposarcoma (DDLPS) | MDM2+, CDK4+, SMA+-/-, Desmin+-/                                                  | Ring and giant marker chromosomes derived from amplification of 12q13–15 (variable amplification of MDM2, SAS, CDK4, HMG2A2) |
| Anaplastic large-cell lymphoma (ALCL)| CD45+, CD30+, ALK-/-                                                               | TCR gene rearrangement, Rearrangement of 2p23 (ALK)                                           |
| Inflammatory myofibroblastic tumor (IMT)| ALK+-/-, SMA+-/-, Desmin+-/-                                                      | Rearrangement of 2p23 (ALK)                                                                   |
| Ewing Sarcoma                        | FLI1+, CD99+                                                                        | t(11;22) and other translocations involving EWSR1 gene                                          |
| Melanoma                             | S100+, SOX10+, MelanA/MART1+, MITF+, Tyrosinase+                                    | BRAF, ARID2, BAP1, GNAQ, HRAS, KIT, NF1, NRAS, and PTEN mutations                              |
| Malignant peripheral nerve sheath tumors (MPNST) | S100+/-, GFAP+-/-, CD34+-/-                                                        | Complex                                                                                       |
| Solitary fibrous tumor (SFT)         | STAT6+, CD34+, BCL2+                                                               | NAB2-STAT6 fusion                                                                              |
| Leiomyosarcoma (LMS)                 | SMA+, Desmin-, Caldesmon+                                                           | Complex                                                                                       |
| Rhabdomyosarcoma (RMS)               | Desmin+, Myogenin+, MyoD1+                                                         | Complex                                                                                       |
| Angiosarcoma                         | Vascular markers (CD31, CD34, FLI1, ERG)+, Keratin +/- in epithelioid angiosarcoma    | MYC (8q24) or FLT4 (VEGFR3) (5q35) amplification, Upregulation of vascular-specific receptor tyrosine kinases (TIE1, KDR, TEK, FLT1) |
| Kaposi Sarcoma                       | HHV8 (LANA)+, vascular markers+, Lymphatic markers (D2-40, LYVE1, Prox1)+           | KSHV/HHV8 with PCR                                                                            |
| Epithelioid hemangioendothelioma (EHE)| Vascular markers+, TFE3+-/-, Keratin +/-                                          | WWTr1-CAMTA1 fusion, YAP1-TFE1 fusion                                                           |
| Alveolar soft part sarcoma (ASPS)    | TFE3+, Desmin+-/-                                                                  | der(17)t(X;17)(p11.2;q25) translocation (ASPS1-TFE1 fusion)                                   |
| Pervascular epithelioid cell tumor (PEComa) | SMA+, Desmin+, HMB45+, MITF+, MART1+                                               | TSC2 mutations, TFE3 gene fusions                                                             |

Table 1. Helpful ancillary tests to differentiate tumors with sarcomatoid features
Since clinical and radiographic features of these tumors are nonspecific, pathological tissue examination is required to differentiate them from the much more common epithelial tumors of the lung. The clinical course of these tumors is generally rapidly progressive and metastasis is common. The majority of patients die within a period of 1 to 72 months. Endobronchial masses showed more favorable prognosis compare to other origins.

Here we report the only case of PPUPS that was diagnosed at UCLA from 2002 to 2019.

CASE REPORT

A 66-year-old man who presented with right sided chest wall pain, weight loss (7 kg), fatigue, and night sweats, without any significant past medical history, was found to have normocytic anemia and a large right inferior hemithorax mass on chest X-ray, which was inseparable from the right hemidiaphragm (Figure 1).

Computed tomography (CT) scan showed a large (15 × 13.6 × 6.2 cm), poorly circumscribed, heterogeneous mass within the right base of the lung involving the pleural surface and mildly protruding into the 8th and 9th intercostal spaces, without eroding of the adjacent bones (Figure 2). Positron emission tomography–computed tomography (PET-CT) scan revealed a large right inferior hemithorax mass with intense fluorodeoxyglucose (FDG) uptake (standardized uptake values (SUVs) of 22.3). The mass abutted the right hemidiaphragm and the chest wall with suggestion of extension into the rib interspaces, most prominent at the right 8th rib interspace. A 6.1 cm right superior paramediastinal mass with intense FDG uptake (SUVMax 16.3) was also seen, suspicious for pleural deposit/metastatic disease (Figure 2).

A CT guided core biopsy was subsequently performed. Histological examination revealed a poorly differentiated malignant neoplasm with epithelioid morphology (Figure 3). Microscopic examination did not reveal any line of differentiation by morphology or upon application of immunohistochemistry; tumoral cells were negative for TTF-1, epithelial (pankeratin, EMA, CAM5.2), mesothelial (calretinin), melanocytic (S100 protein, SOX10, MART1), vascular (CD34), and myogenic (desmin, caldesmon) markers. Therefore, the favored diagnosis was poorly differentiated malignant epithelioid neoplasm.

Figure 1. Chest X-Ray (A) anteroposterior view; (B) lateral view showing a large right lower lung mass.

Figure 2. Imaging study of the thorax. CT scan (A - axial plane, C - coronal plane) shows a 15 × 6 cm poorly circumscribed right lower lung mass protruding into the 8th and 9th intercostal spaces. (B - axial plane and D - coronal plane) PET scan shows a 15 cm right inferior lung mass and a 6 cm right superior paramediastinal mass with high FDG uptake.

Figure 3. Photomicrograph of the CT-Guided biopsy showed a poorly differentiated malignant epithelioid neoplasm (H&E, 10X).
Subsequently, the patient underwent surgical wedge resection of right lower and middle lobes, along with the adjacent right chest wall. Gross examination showed a well-circumscribed light tan and firm mass with areas of necrosis and hemorrhage, which abutted the bone without any gross or microscopic evidence of invasion. The mass measured 17.5 cm × 6.5 cm × 1.5 cm (Figure 4).

Representative sections showed a vaguely nodular mass consists of spindle to epithelioid neoplastic cells arranged in sheets and fascicles some in a storiform pattern. Large, bizarre pleomorphic cells with round to oval nuclei, prominent nucleoli, and moderate amounts of cytoplasm were observed (Figure 5A and 5B). Areas of necrosis (Figure 5C) and high mitotic activity, including atypical mitosis (Figure 5D), were identified.

All surgical margins were free of malignancy and no lymph node metastasis was identified. As with the biopsy, the neoplastic cells did not show any immunoreactivity to epithelial, mesothelial, or glandular markers; negative staining for pankeratin, CAM5.2, Keratin 5/6, p63, calretinin, WT1, D2-40, TTF1, S100, SOX10, MART1, HMB45, desmin, caldesmon, EMA, CD34, STAT6, C-Kit, DOG1, myogenin, MyoD1, CD21, CD23, CD35, chromogranin, TLE1, BCL2, and CD99 (Figure 6). Since no evidence of extrapulmonary origin

Figure 4. Gross examination of the lung mass resection.

Figure 5. Photomicrographs of the tumor showing in A – Vaguely nodular high-grade spindle to epithelioid neoplasm with adjacent rib bone (H&E, 3X); B – Neoplastic cells with adjacent normal lung parenchyma (H&E, 14X); C – Undifferentiated pleomorphic sarcoma with areas of necrosis (H&E, 10X); D – Pleomorphic cells with high mitotic activity, atypical mitoses, chronic inflammatory cell infiltrate and focal necrosis (H&E, 12.5X).
was identified and the tumor did not demonstrate any line of differentiation in our extensive work up, the diagnosis of primary pulmonary undifferentiated pleomorphic sarcoma (PPUPS) was rendered.

Final Pathologic Diagnosis: (i) Undifferentiated pleomorphic sarcoma, high grade, 17.5 cm; (ii) Surgical margins are uninvolved.

The patient was further treated with radiotherapy and multiple cycles of adjuvant chemotherapy (7 days cycle per month with Ifosfamide 1500mg/m2 CIV on days 1-7). Patient tolerated the treatment without any evidence of local recurrence or metastasis up to this date (4 months follow up).

DISCUSSION

Primary lung sarcomas are rare, and represents less than 0.5% of lung malignancies. Cameron and Miller and Allen reported primary lung sarcomas in 0.15% and 0.3% of lung neoplasms in studies of 6000 and 10134 patients with primary lung malignancies respectively. Among the primary pulmonary sarcomas, undifferentiated pleomorphic sarcoma (UPS) is one of the rarest one with less than 84 reported cases. UPS is defined as a high-grade pleomorphic neoplasm that shows no discernible microscopic evidence of any specific form of differentiation (e.g., lipoblasts, bone formation, epithelial structures) using currently available diagnostic techniques. Fletcher CD in a retrospective study on re-analysis of 159 tumors showed that just 26% of previously diagnosed MFH cases were “true” UPSs, and more than half of cases showed an identifiable line of differentiation. The main core of diagnosis of UPS is to exclude other malignant tumors that display similar morphological findings such as malignant melanomas, sarcomatoid carcinomas, anaplastic lymphomas, sarcomatoid mesotheliomas, or other sarcomas (e.g., dedifferentiated liposarcoma and pleomorphic rhabdomyosarcoma, etc.). Therefore, it is a diagnosis of exclusion, which requires careful

Figure 6. Photomicrographs of the tumor. Immunohistochemistry studies shows no line of differentiation (Magnification x10).
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examination of tissue and generous sampling of the specimen to establish a correct diagnosis. This diagnosis should not be made definitively on a biopsy specimen, because the entire mass is not present in the biopsy materials. Some authors believe that the category of UPS serves primarily as a “wastebasket” for a heterogeneous group of unclassifiable neoplasms with pleomorphic morphology.

UPS characteristically shows positive staining for histiocytic markers (CD68, α1-antichymotrypsin, vimentin), a reason why such tumors were identified as “malignant fibrous histiocytoma” (MFH) in the past. Staining for TTF-1, S-100 protein, desmin, actin, myoglobin, caldesmon, D2-40, and calretinin is negative. In some cases, keratin staining may be positive, which makes it difficult to differentiate sarcomatoid carcinomas (SCs) from UPS. Stronger cytokeratin immunoreactivity along with more differentiated carcinomatous elements, as well as immunoreactivity to other epithelial markers (such as P63) can be helpful in the diagnosis of SCs. A wider than usual panel of immunohistochemical studies is necessary to rule out other neoplasms that can resemble UPS including other types of sarcomas, sarcomatoid carcinoma, melanoma, or mesothelioma. (Table 1). Special staining, electron microscopy, or microscopy with ultraviolet surface excitations have not shown strong utility in the diagnosis of UPSs.

Non-specific small droplets of neutral fat and PAS-positive, diastase-resistant droplets may be seen especially in the giant cells in this tumor, which probably reflect a degenerative change. Electron microscopic features are helpful for evaluation of better-differentiated tumors; for example, high aspect ratio surface microvilli, perinuclear tonofilaments, and hyaluronic acid crystals are seen in more differentiated mesotheliomas. However, when the tumor is poorly differentiated, the ultrastructural findings are rarely helpful. The genetic profile of primary pulmonary mesenchymal tumors shows complex and nonspecific cytogenetic aberrations, similar to their soft tissue counterparts. Molecular studies can be helpful to rule out other tumors with similar histological findings (Table 1). Molecular mechanisms responsible for primary pulmonary UPS formation and progression are unknown.

It is also important to know that UPS is a relatively common soft tissue tumor and most cases of UPS found in the lung represent metastasis from an extrapulmonary origin. Therefore, a careful clinical evaluation to exclude a possible extrapulmonary site of origin is necessary before diagnosing primary pulmonary UPS (PPUPS).

We provide a brief review of literature on PPUPS (previously described as PPMFH) in Table 2. 85 cases have been reported in the English literature since 1979.

Table 2. Review of literature of primary pulmonary undifferentiated pleomorphic sarcoma/primary pulmonary fibrous histiocytoma PPUPS/PPMFH.

| # | Year | Reference | Age | Sex | Location | Size (cm) | LN | Tx | Survival (mos) | F/U |
|---|------|-----------|-----|-----|----------|----------|----|----|---------------|-----|
| 1 | 1979 | Bedrossian et al. | 51 | M | LLL/RML | 3 | N | L | 14 | DOD |
| 2 | 1979 | Kern et al. | 53 | M | RLL | 8 | N | L | 12 | DOD |
| 3 | 1980 | Chowdhury et al. | 52 | F | RLL | 5 | U | C | 4 | DOD |
| 4 | 1981 | Paulsen et al. | 53 | F | LLL | 4 | N | L | 36 | DOD |
| 5 | 1982 | Mills et al. | 60 | F | RLL | 10 | N | L | 18 | AWD |
| 6 | 1982 | Srumpai et al. | 41 | M | RLL | 9 | U | L | 18 | DOD |
| 7 | 1983 | Misra et al. | 45 | M | RLL | 16 | P | X | 10 | DOD |
| 8 | 1984 | Larsen et al. | 75 | M | RUL | 2.5 | N | R | 10 | NED |
| 9 | 1984 | Lee et al. | 62 | M | LLL | 6 | N | L | 12 | NED |
| 10 | 1984 | Lee et al. | 54 | M | LUL | 7 | N | C | 7 | DOD |
| 11 | 1984 | Lee et al. | 69 | M | RUL | 8 | N | Pn,X | 8 | NED |

AWD = alive with disease; C = chemotherapy; DNED = dead with no evidence of disease; DOD = death of disease; DWED = death of non-related cause with evidence of disease; E = endoscopic resection; F/U = follow up; F = female; L = lobectomy; LLL = left lower lobe; LN = lymph node; LUL = left upper lobe; M = male; N = negative; NED = no evidence of disease; Nt = no treatment; Pn = pneumonectomy; P = positive; R = resection; REB = right endobronchial; RLL = right lower lobe; RML = right middle lobe; RUL = right upper lobe; Tx = treatment; U = unavailable data; X = radiotherapy.
| #  | Year | Reference                          | Age | Sex | Location | Size (cm) | LN | Tx | Survival (mos) | F/U |
|----|------|------------------------------------|-----|-----|----------|----------|----|----|----------------|-----|
| 12 | 1984 | Lee et al.23                        | 62  | F   | LLL      | 5        | N  | L,X | 120            | NED |
| 13 | 1984 | Lee et al.23                        | 67  | M   | LUL      | 4        | N  | L   | 60             | NED |
| 14 | 1984 | Lessel and Erbstösser24            | 35  | F   | RLL      | 25       | U  | Nt  | 12             | DOD |
| 15 | 1984 | Silverman and Coalson25            | 56  | M   | LUL      | 8        | -  | C   | 3              | AWD |
| 16 | 1985 | Tanino et al.26                    | 75  | F   | LLL      | 5        | +  | Nt  | 5              | DOD |
| 17 | 1986 | Venn et al.27                      | 32  | F   | RML/RUL  | U        | U  | L   | 18             | NED |
| 18 | 1986 | Venn et al.27                      | 62  | M   | RUL      | 8        | U  | L   | 60             | NED |
| 19 | 1986 | Venn et al.27                      | 61  | F   | LUL      | U        | U  | Pn  | 15             | DWED |
| 20 | 1986 | Venn et al.27                      | 62  | M   | LUL      | U        | U  | X   | 2              | AWD |
| 21 | 1987 | Hsiu et al.28                      | 71  | F   | RUL      | 5        | N  | L   | 10             | NED |
| 22 | 1987 | Juettner et al.29                  | 68  | M   | LLL      | 20       | N  | Nt  | 12             | DOD |
| 23 | 1987 | Juettner et al.29                  | 58  | M   | RLL      | 5.5      | P  | L   | 12             | DOD |
| 24 | 1987 | Ismailer et al.30                  | 12  | F   | RUL      | 9        | N  | L   | 12             | AWD |
| 25 | 1987 | Yousem and Hochholzer5             | 54  | F   | RLL      | 1.7      | N  | L   | 108            | NED |
| 26 | 1987 | Yousem and Hochholzer5             | 33  | M   | RUL      | 3.8      | N  | L   | 84             | NED |
| 27 | 1987 | Yousem and Hochholzer5             | 59  | M   | RLL      | 5.9      | N  | L   | 65             | NED |
| 28 | 1987 | Yousem and Hochholzer5             | 73  | F   | LUL      | 8.5      | P  | Pn  | 36             | NED |
| 29 | 1987 | Yousem and Hochholzer5             | 64  | M   | RUL      | 5        | N  | L,X | 16             | NED |
| 30 | 1987 | Yousem and Hochholzer5             | 42  | F   | LLL      | 3        | N  | L   | 122            | NED |
| 31 | 1987 | Yousem and Hochholzer5             | 57  | F   | RUL      | 4        | N  | Pn  | 1              | DNED |
| 32 | 1987 | Yousem and Hochholzer5             | 80  | M   | LUL      | 3        | N  | L   | 1              | DNED |
| 33 | 1987 | Yousem and Hochholzer5             | 74  | M   | LUL      | U        | N  | Nt  | 2              | DOD |
| 34 | 1987 | Yousem and Hochholzer5             | 18  | M   | RLL      | 10       | N  | L   | 1              | DOD |
| 35 | 1987 | Yousem and Hochholzer5             | 46  | F   | RUL      | 6        | P  | L,X | 8              | DOD |
| 36 | 1987 | Yousem and Hochholzer5             | 52  | F   | RLL      | U        | N  | C   | 9              | DOD |
| 37 | 1987 | Yousem and Hochholzer5             | 52  | F   | LUL      | 4        | P  | L,C,X| 72             | DOD |
| 38 | 1987 | Yousem and Hochholzer5             | 74  | F   | RUL      | 14       | P  | L   | 24             | DOD |
| 39 | 1987 | Yousem and Hochholzer5             | 69  | F   | RUL      | 8        | N  | X   | 36             | DOD |
| 40 | 1987 | Yousem and Hochholzer5             | 40  | F   | LLL      | 4        | N  | L,X | 24             | DOD |

AWD = alive with disease; C = chemotherapy; DNED = dead with no evidence of disease; DOD = death of disease; DWED = death of non-related cause with evidence of disease; E = endoscopic resection; F/U = follow up; F = female; L = lobectomy; LLL = left lower lobe; LN = lymph node; LUL = left upper lobe; M = male; N = negative; NED = no evidence of disease; Nt = no treatment; Pn = pneumonectomy; P = positive; R = resection; REB = right endobronchial; RLL = right lower lobe; RML = right middle lobe; RUL = right upper lobe; Tx = treatment; U = unavailable data; X = radiotherapy.
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### Table 2. Continued...

| #   | Year | Reference                                      | Age | Sex | Location | Size (cm) | LN | Tx | Survival (mos) | F/U |
|-----|------|-----------------------------------------------|-----|-----|----------|-----------|-----|----|----------------|-----|
| 41  | 1987 | Yousem and Hochholzer² | 74  | M   | RML     | U          | N   | X  | 8              | DOD |
| 42  | 1987 | Yousem and Hochholzer² | 19  | M   | LUL     | U          | N   | L,C,X         | 14  | DOD |
| 43  | 1987 | Yousem and Hochholzer² | 63  | M   | LLL     | 7          | N   | Nt | 14             | DOD |
| 44  | 1987 | Yousem and Hochholzer² | 36  | M   | RLL     | 3          | N   | R  | 12             | DOD |
| 45  | 1987 | Yousem and Hochholzer² | 32  | M   | LLL     | 11         | P   | Pn,C,X        | 3   | DOD |
| 46  | 1988 | Casey and Peddle³ | 21  | M   | RUL     | 3          | N   | L  | 96             | NED |
| 47  | 1988 | Casey and Peddle³ | 46  | M   | LLL     | 10         | N   | L  | 8              | NED |
| 48  | 1988 | McDonnell et al.³² | 73  | F   | LLL     | 6.5        | N   | L  | 3              | DOD |
| 49  | 1988 | Palmer et al.³³ | 62  | F   | RLL     | U          | N   | L  | 14             | DOD |
| 50  | 1989 | White et al.³⁴ | 55  | M   | RUL     | U          | U   | Nt | 4              | DOD |
| 51  | 1990 | In et al.³⁵ | 43  | F   | RLL     | U          | U   | C,X | U              | U   |
| 52  | 1990 | Marchán and Pérez³⁶ | 10  | F   | LLL     | 5          | N   | L  | U              | U   |
| 53  | 1993 | Higashiyama et al.³⁷ | 49  | F   | RLL     | 6          | P   | Pn | U              | NED |
| 54  | 1995 | Kamath et al.³⁸ | 56  | M   | RLL     | 10         | U   | Nt | 3              | DOD |
| 55  | 1996 | Gómez-Román and Val-Bernal³⁹ | 61  | M   | RUL     | 3          | U   | R  | 9              | NED |
| 56  | 1996 | Halyard et al.⁴⁰ | 51  | F   | LLL     | 10         | N   | L,X | 60             | NED |
| 57  | 1996 | Halyard et al.⁴⁰ | 77  | M   | RML     | 2.2        | N   | L  | 36             | NED |
| 58  | 1996 | Halyard et al.⁴⁰ | 40  | M   | LLL     | 11         | P   | R  | 6              | DOD |
| 59  | 1996 | Halyard et al.⁴⁰ | 57  | F   | LUL     | 7.5        | U   | L  | 1              | DOD |
| 60  | 1996 | Shah et al.⁴¹ | 9   | M   | LUL     | 6          | U   | L,C,X        | 36  | NED |
| 61  | 1997 | Nistal et al.⁴² | 12  | F   | LUL     | 7          | U   | C,X | 5              | AWD |
| 62  | 1997 | Barbas et al.⁴³ | 37  | M   | RML/RLL | 10         | N   | Pn | 6              | DNED |
| 63  | 2000 | Fujita et al.⁴⁴ | 65  | F   | LLL     | 12         | U   | Nt | 6              | DOD |
| 64  | 2000 | Herrmann et al.⁴⁵ | 57  | M   | RUL     | 13         | U   | L  | 12             | NED |
| 65  | 2001 | Nonaka et al.⁶ | 59  | M   | U       | 4.5        | U   | U  | U              | U   |
| 66  | 2002 | Alhadab et al.⁴⁶ | 56  | M   | LUL/LLL | U          | U   | Nt | 4              | DOD |
| 67  | 2002 | Etienne-Mastroianni et al.² | 47  | M   | U       | U          | U   | L,X | 3              | NED |
| 68  | 2003 | Wang et al.⁴⁷ | 86  | M   | LLL     | 15         | U   | Nt | 2              | DOD |
| 69  | 2007 | Maeda et al.⁴⁸ | 62  | M   | LUL     | 4.5        | P   | L  | 24             | DNED |
| 70  | 2007 | Rzyman et al.⁴⁹ | 58  | M   | LUL     | 4          | N   | Pn | 121            | NED |
| 71  | 2007 | Rzyman et al.⁴⁹ | 61  | M   | RUL     | 7.5        | N   | L  | 7              | DNED |
| 72  | 2007 | Rzyman et al.⁴⁹ | 75  | M   | RUL     | 8          | P   | Pn | 4              | DOD |
| 73  | 2007 | Rzyman et al.⁴⁹ | 61  | F   | LUL     | 3          | N   | L  | 2              | DOD |
| 74  | 2007 | Rzyman et al.⁴⁹ | 54  | M   | RUL     | 9          | P   | L  | 3              | DOD |
| 75  | 2008 | Noh et al.⁵⁰ | 58  | F   | RUL     | 5          | N   | L,X | 5              | NED |
| 76  | 2010 | Maitani et al.⁵¹ | 18  | F   | LUL     | 2.2        | U   | L  | 36             | NED |

AWD = alive with disease; C = chemotherapy; DNED = dead with no evidence of disease; DOD = death of disease; DWED = death of non-related cause with evidence of disease; E = endoscopic resection; F/U = follow up; F = female; L = lobectomy; LLL = left lower lobe; LN = lymph node; LUL = left upper lobe; M = male; N = negative; NED = no evidence of disease; Nt = no treatment; Pn = pneumonectomy; P = positive; R = resection; REB = right endobronchial; RLL = right lower lobe; RML = right middle lobe; RUL = right upper lobe; Tx = treatment; U = unavailable data; X = radiotherapy.
including our case. 51 out of 85 case were male and 35 were female (M:F ratio of 1.45). The patient age ranged from 9 to 85 years with a mean age of 54. The tumor size ranges from 1.7 to 25 cm, with average size of 7.3 cm, and mean survival of 23.73 months. Even though PPUPS has a high local recurrences and mortality rate, long term survival has been reported in some cases, even more than 10 years.\textsuperscript{5,23,49,52}

**CONCLUSION**

Primary pulmonary undifferentiated pleomorphic sarcoma (PPUPS) is an exceptionally rare tumor and is essentially a diagnosis of exclusion. The approach should be to first establish the absence of any particular line of differentiation, which requires proper sampling, histological analysis, and immunohistochemistry studies. Other ancillary tests such as molecular studies and electron microscopy could also possibly be helpful. The next step is to exclude any possible extrapulmonary origin by clinical examination or by other means such as PET-CT scan. PPUPS is a highly malignant sarcoma with a poor prognosis, with surgery being the primary treatment in most cases. Postoperative chemotherapy has also been reported to be beneficial in some cases. We report here the only case of PPUPS, which was diagnosed in the past 17 years at UCLA, after an extensive work up. The patient was treated with surgical resection and post-operative chemotherapy. With 4 months follow up, there is no evidence of local recurrence or distant metastasis.

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