CHICKEN EGG WHITE — CHARACTERISTICS OF ITS PROPERTIES AND THE PROSPECTS FOR FUNCTIONAL FOODS DEVELOPMENT

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Abstract

The overview presents the literature data and the results of our own research on prospects of using the chicken eggs as the basis of functional foods. The composition of chicken eggs and their components, characteristics of egg white proteins properties are presented thereto. The biologically active compounds included into egg composition are analyzed. The data on the biological value of egg white are given. The characteristic of egg white foaming ability is presented. It has been shown that the ability of proteins to form stable intermolecular structures, especially with partially denaturated proteins, allows them forming viscoelastic superficial films that ensure foam stability. The high foaming ability of chicken egg protein macromolecules is directly related to their interphase properties, i.e. the ability to form interphase layers at the "liquid — gas" interface. The foaming properties of the various egg proteins are not equal, and therefore they contribute to foaming properties at various extents. The model of egg white proteins gelation is considered and the factors influencing the gelation process are described. It has been shown that very important changes in proteins properties are caused by denaturation. The proteins lose their ability to hydrate; the protective aqueous shell around the globules disappears, the proteins stick together, grow larger and lose solubility. This process is called coagulation. The influence of denaturation and aggregation on variations of protein properties is described below. Data on protein fortification with functional ingredients (calcium, iodine, plant polyphenols) and creation of functional egg and meat foods are presented here.

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Introduction

The priority task of state policy in our country in sphere of nutrition is development of health-improving foods production in order to preserve and enhance the health of population, to prevent diseases caused by insufficient and unbalanced nutrition. The practical implementation of this task assumes, in particular, the health of population, to prevent diseases caused by insufficient and unbalanced nutrition. The practical implementation of this task assumes, in particular, development, production and supply of wide range of health-improving foods to the domestic consumer market, including functional foods, in accordance to appropriate regulatory and methodological provisions harmonized with international requirements.

The concept of healthy nutrition assumes an increase of efficiency of nutrition factor to the preservation and improvement of public health within the state. This factor provides necessity for development of biotechnologies, targeted to foods modifying by including ingredients of high biological and nutritional value. This leads to creation of an expanded range of functional foods (FF).

The promising source of food raw materials with high biological FF value are chicken eggs. Chicken eggs contain all food and biologically active substances necessary for healthy development [1]. Chicken egg is a natural functional food; it is a unique source of numerous substances of high biological and nutritional value due to significant content of protein balanced in amino acid profile. It contains a complete lipid complex, wide range of macro- and microelements, and vitamins. The average chemical composition is constant, fluctuations in the content of macro- and microelements, and vitamins. The average chemical composition is constant, fluctuations in the content of eggs, laid by one species of poultry, depend on the diet of layers (which is especially peculiar for lipid fraction) (Table 1) [2].

Table 1. Chemical composition of a chicken egg

| Components | Mass fraction, % |
|------------|-----------------|
| Moisture   | 75.33–76.07     |
| Protein    | 11.34–12.31     |
| Fat        | 10.2–10.3       |
| Carbohydrates | 1.0   |
| Inorganic compounds | 0.8 |
| Melange    | 88.65–87.06     |
| White      | 49.91–50.29     |
| Yolk       | 1.61–1.63       |
| Shell      | 3.30–3.41       |

Eggs contain a lot of biologically active compounds, including those which possess antimicrobial, immunomodulatory, antioxidant, anticarcinogenic, hypotensive and other properties [3–5].
Information on this extensive issue can be partially obtained from a number of scientific reviews publications [3–8], nevertheless it is necessary to briefly characterize the chicken egg proteins, since they constitute the most important component in the composition of the functional egg foods being developed now.

Egg composition
Egg white contains simple proteins (ovalbumen, ovoconalbumen, ovoglobulin) and complex proteins — glycoproteins or mucoproteins (ovomucoid and ovomucin) (Table 2).

The major egg white proteins are ovalbumen (54%), ovotransferrin (12%), ovomucoid (11%), lysozyme (3.5%) and ovomucin (3.5%). Minor proteins include avidin (0.05%), cystatin (0.05%), ovomacroglobulin (0.5%), ovo-flavoprotein (0.8%), ovogloboprotein (1.0%), and ovo-inhibitor (1.5%) [3,5,6].

Biological value of the egg white
In order to give quantitative characteristic to the quality of food protein, the parameter “biological value” (BV) in nutritional science is used. BV is the degree of food nitrogen retention in a growing body (which degree depends on the amino acid composition and other structural features of protein), and efficiency of food nitrogen utilization to maintain nitrogen balance in humans. The amino acid composition, content and ratio of essential amino acids determine the BV of a protein — the parameter that reflects the degree of protein utilization. The degree of digestion, absorption and utilization of protein is significantly influenced by food manufacture technology and methods of food processing at the food processing plants. Therefore, when evaluating BV, its true digestibility is taken into account. The parameter “The protein digestibility-corrected amino acid score” (or PDCAAS for short) [9], which is equal to the amino acid rate multiplied by true digestibility, has been introduced. Its highest reference value is 1.0. Comparative data on true digestibility and PDCAAS for chicken egg white and proteins obtained from various food sources are presented below in the Table 3.

Table 3. Corrected amino acid index of proteins digestibility

| Food          | Protein content % | True digestibility | Amino acid score | PDCAAS |
|---------------|-------------------|--------------------|-----------------|--------|
| Casein        | 94.7              | 99                 | 1.19            | 1.00   |
| Chicken egg white | 87               | 100                | 1.19            | 1.00   |
| Beef protein  | 95.2              | 98                 | 0.94            | 0.92   |
| Soy protein isolate | 92.2     | 98                 | 0.94            | 0.92   |

Gel-forming properties of chicken egg
Currently the share of eggs and their separate components in formulations of a wide range of foods is increasing steadily, which is explained by their properties: emulsifying ability and thermal gelation (coagulation) [7].

Foaming capacity of egg white proteins
Poultry egg white is an aqueous protein solution with 84–89% of water. It also contains minor amounts of carbohydrates, vitamins and minerals. Proteins, which make up more than 90% of the egg white dry matter, are heat-sensitive and prone to surface denaturation, which explains their unique technological properties. As it’s known, foams are the dispersed systems consisting of many gas bubbles separated by thin (only single colloidal particle thick) films of liquid. Adding of surfactants, which reduce surface tension, contribute to foaming. However, stable foaming requires the presence of high-molecular compounds in the system, for example, proteins which form two-sided strong adsorption layers on the surface of thin films of liquid. Egg white proteins are amphiphilic and feature relatively high surface hydrophobic properties; therefore, they diffuse towards the air-water phase interfaces, where they are adsorbed effectively. This flexibility of molecules allows proteins to rearrange their conformational structure at the interface, which leads to significant decrease in the superficial tension at these interfaces. The ability of proteins to form stable intermolecular structures, especially partially denatured proteins, allows them forming viscoelastic superficial films that ensure foam stability. The high foaming capacity of chicken egg protein macromolecules is directly

| Protein          | % in protein | pH   | Molecule weight (kDa) | Internal viscosity, 100 cm²/g | Tdenaturation (°C) | –SH | S–S | Notes                                  |
|------------------|-------------|------|-----------------------|-------------------------------|--------------------|-----|-----|----------------------------------------|
| Ovalbumen        | 54          | 4.5–4.6 | 45000              | 0.043                         | 75–84              | 4   | 1   | Phosphoglyco-protein; 4 SH-groups     |
| Ovotransferrin   | 12–13       | 6.1–6.6 | 76000–80000          | 0.084                         | 61–65 (76.5; Al³⁺) | —   | 15  | Glycoprotein, binds to complexes with iron and other metals |
| Ovomucoid        | 11          | 3.9–4.3 | 28000              | 0.055                         | 77                 | —   | 9   | Glycoprotein, trypsin inhibitor       |
| Ovomucin         | 3.5         | 4.5–5.0 | 110000             | 2.10                          | —                  | —   | —   | Glycoprotein, fibrous, viscous       |
| Lysozyme         | 3.4–3.5     | 10.7  | 14300–14600         | 0.027                         | 69–77              | —   | 4   | Spherical protein; 4 — S5 — links; features lytic activity |
| Ovo-inhibitor    | 1.5         | 5.1–5.2 | 44000–49000         | —                             | —                  | —   | —   | Inhibits trypsin and chymotrypsin    |
| Ovoglyco-protein | 1.0         | 3.9   | 24000–24400         | —                             | —                  | —   | —   | Glycoprotein                          |
| Ovoflavoprotein  | 0.8         | 4.0–4.1 | 32000–35000        | —                             | —                  | —   | 2   | Binds riboflavin                      |
| Ovomacro-globulin| 0.5         | 4.5–4.7 | 760000–900000       | 0.065                         | —                  | —   | —   | Glycoprotein                          |
| Avidin           | 0.5         | 9.5–10 | 55000–68300         | —                             | —                  | —   | 1   | Binds biotin                          |
related to their interfacial properties, i.e., the ability to form interfacial layers at the liquid-gas interface. The foaming properties of various proteins of chicken egg white are not equal, and therefore they contribute to the formation of its foaming properties in various extents (Table 4).

Table 4. Interfacial characteristics of the major proteins of egg white [8]

| Protein       | Superficial tension (mN/m) | Foaming coefficient (cm/g per minute) |
|---------------|----------------------------|--------------------------------------|
| Ovalbumen     | 51.8                       | 0.59                                 |
| Ovotransferrin| 42.4                       | 0.34                                 |
| Lysozyme      | 42                         | 0.12                                 |
| Ovomucoid     | 39                         | 0                                    |
| Ovomucin      | No data                    | 0                                    |
| Mixture of these proteins in the same proportion as in egg white | 46.7 | 3.08 |

Egg white is considered to be a reference model for foaming properties, which other animal and plant proteins are compared to [10–15]. Despite numerous comparative studies of the properties of some individual egg white proteins and attempts to rank them according to these properties [10,16,17], it is almost impossible because of complexity of egg white proteins composition and potentially possible synergistic interactions of individual proteins. All those complexities make it almost impossible to identify the special role of each individual protein in the foaming process [18]. It is very difficult to predict the foaming properties of any mixture of proteins, since individual proteins may compete among themselves for the surface, or they may substitute each other at the interface [19]. However, the foaming capacities of individual proteins isolated from egg white are always less than the foaming capacity of egg white as a whole. Egg white foams are the main component of such foods as meringues, nougat and light biscuits. Another peculiar feature of egg white-based foams is the dependence of their characteristics on the quality of the original egg white [20]. Since the foaming properties of dense egg white and liquid egg white differ from each other [21], all factors affecting the ratio of these fractions in the egg white (i.e., shelf life of the egg, age and breed of laying hen) also affect the foaming capacity of the egg white [22–26]. Another extremely important parameter is the accuracy of separation of egg white and yolk, since even very small amounts of yolk (0.022%) in the egg white significantly reduce its foaming capacity [27]. Heat treatment also reduces the foaming capacity of the egg white. Industrial pasteurization can reduce by 10% both the foaming capacity of the egg white and the foam stability as well [20,28, 29]. The dripping of liquid phase from the foam can be increased threefold after its pasteurization. Heating the protein during drying also worsens the foaming properties of the egg white, and the foaming may be decreased by 10 to 60% due to pH of the egg white during drying, and may decrease the foam stability by 20%.

**Gelation properties of egg white**

The process of egg proteins gelation is described in details by the model of thermal gelation of globular structure proteins [5]. In this model gelation is considered as a two-stage process, sequentially including denaturation and aggregation. The scheme is as follows: native protein → denatured protein (long chains) → aggregated protein (associated mesh).

Denaturation results in very important changes in proteins properties. First of all, they lose their ability to hydrate, the aqueous protective shell around the globules disappears, they join together, grow larger and lose their solubility. This process is called coagulation.

When eggs are heated, the proteins that make up their composition, changes first. Due to the thermal denaturation of protein substances at 50–55°C, local opacities are formed in the transparent mass, and gradually spread out. At 65°C the entire mass of protein get thicker, and at 75°C it turns into a solid opaque mass of a very delicate consistency. At 80°C the gel is obtained which is able to keep its shape, and along with further heating (above 85°C) it becomes more and more dense. The degree of compaction of the protein gel depends on heating duration. Chicken egg white has 2 main endothermic points at 60–65°C and 80–85°C, which corresponds to the temperatures of ovotransferrin and ovalbumen denaturation [30,31].

Differences in opacity of samples with different salt content are associated with differences in protein clots sizes and rate of protein aggregation. Salt NaCl causes changes in conformation of egg proteins proportionally to increase in the ionic strength of the solution. Increasing the solubility of protein molecules increases their susceptibility to thermal denaturation. The high content of NaCl leads to coagulation of protein, thus playing a decisive role in the process of its aggregation.

At the first stage of gelation the “folded” macromolecules of globular proteins “unfold”, thus forming the denatured proteins, and release their hydrophobic “internal” structure. At the second stage of gelation (aggregation), macromolecules of denatured proteins interact with each other to form even higher molecular weight aggregates, which in its turn also interact with each other, which ultimately leads to the formation of three-dimensional gel structure [32]. Comparison of the denaturation rate with the aggregation rate helps determine the characteristics of the obtained gel. The wide range of factors affect protein aggregation, which is important because changing the aggregation rate in relation to the denaturation rate affects the characteristics of the obtained gel. Some of these factors are listed below.

Electrostatic charge (pH) can alter the charge distribution among the side chains of amino acids and, accordingly, either decrease or increase protein-protein interactions. The main factor in heat-induced aggregation of ovalbumen (pH 4.5–4.6) is the degree of electrostatic repulsion between the protein molecules of exposed denaturation. The aggregation rate of a heated 1.8% ovalbumen solution
is higher at pH=5.5 and 8.5 than at pH=7.0. At a pH of 5.5 the opacity builds up more rapidly.

The concentration of protein in solution also affects aggregation rate. The high concentration of protein causes the molecules to come closer together to form aggregates at lower temperatures.

The pH value required for achieving the same opacity that indicates protein aggregation, increases as the protein concentration increases, and the pH shifts to alkaline direction from the isoelectric point from 6 to 11, and decreases when the pH shifts to acidic direction from 4.5 to 3.0. There is an assumption [5] that the denaturation rate causes a low rate of aggregation, because the forces of attraction between denatured protein chains are low. The resulting gel will have a fine mesh of protein chains, will be more transparent, and will be characterized by less syneresis than the gel obtained at the high aggregation rate. A larger mesh of protein chains will result to the opaque gel with large internodes capable of retaining a solvent that is easily pressed out from the matrix. Conditions that favor denaturation, such as high or low pH, reduce aggregation of globular proteins. In this case, due to the high charge of the molecules, denaturation based on protein-solvent interactions prevails rather than aggregation caused by protein-protein interactions. A gel mesh with a certain degree of structural regularity can be obtained if the aggregation rate prevails rather than aggregation caused by interactions prevails rather than aggregation caused by denatured proteins. In general, gelation is associated with an imbalance between mutually attractive (van der Waals) and mutually repulsive (electrostatic, steric) interactions that determine the spatial structure of the protein in solution. The protein structure can be modified in various ways. Examples of the modification are given in the monograph by V. N. Izmailova and P. A. Rebinder “Structure formation in protein systems” and by other authors [34]. Both during acidification and alkalinization the native globular proteins can irreversibly “rearrange” into proteins of the fibrillar type. The formation of thixotropic ovalbumen gels due to addition of acetic acid may result from formation of hydrogen bonds between acetic acid and protein. Ovalbumen gets denatured in presence of urea and alkali, when this protein forms the gel. Alkaline ovalbumen gels melt the faster, the higher is pH of the gel. Ovalbumen gels can also be prepared at high pressures which cause denaturation changes in protein macromolecules.

The results of heat treatment research are of particular interest, as the heat treatment is widely used in the food industry for formation of gels from egg white and yolk. In aqueous solutions of ovalbumen at room temperature the induction period of structure formation got shorter with increasing protein concentration. At the same time, the strength of the gel structure increased due to an increase in the number of intermolecular contacts per volume unit, accompanied by conformational changes in protein macromolecules. The process of denaturation of ovalbumen, exposed to acid or alkali, was accelerated by heating. While discussing the nature of the interactions that determine the strength of the resulting ovalbumen gels, the authors come to the conclusion that it is determined by hydrogen bonds and hydrophobic interactions. In this case electrostatic interactions are excluded due to the formation of the gel structure in strongly acidic and strongly alkaline media. Thus, the structure of ovalbumen gels is similar to thixotropic coagulation structural meshes, where the particles of the dispersed phase or macromolecules aggregates are bound together by van der Waals forces which occur between hydrocarbon hydrophobic amino acid residues of ovalbumen macromolecules during protein denaturation.

The type and force of interaction between denatured protein molecules depend on their structure and, in particular, on the “extent of unfolding” of protein molecules at the end of the denaturation stage. These interactions also depend on the general physical and chemical conditions in the system, which can both restrict and contribute to gelation, i. e. respectively increase or decrease the level of aggregation and, conversely, decrease or increase the degree of denaturation, after which these interactions begin [35]. These mechanisms were intensively researched on the samples of thermally induced gelation of egg white and ovalbumen, while the pecu-
Liar attention was paid to the influence of the ionic strength of a solution on the structure and characteristics of the resulting gels [32, 36–39]. When heated at high ionic strength, ions shield the charges of protein molecules, which process promotes hydrophobic interactions [40]. In these conditions aggregates of partially denatured proteins are formed at random. These aggregates create cloudy opaque gels with low rates of rigidity [hardness], low elasticity and poor water-holding capacity (WHC). On the other hand, at low ionic strength the higher electrostatic repulsive forces slow down the aggregation process [32] and promote protein denaturation. Finally, the further process of aggregation includes the interaction of hydrophobic sections of molecules with the formation of linear polymer aggregates. In media with the higher ionic strength these aggregates can interact to form gels with good processing properties.

The thermal gelation capacity has been used to develop technologies and new types of functional egg foods, as well as meat foods with a high share of eggs included in their composition.

Another important parameter to monitor the quality of protein gelation is the pH value. Close to their isoelectric point (pH ~ 5) the proteins are more prone to the formation of randomly arranged aggregates, similar to those obtained at high ionic force. On the contrary the gelation properties of protein are expressed in the best way within the alkaline pH range [41,42]. On the other hand at low pH (2.0) the gelation temperature decreases and the rheological properties of the gel worsen, which is explained by decrease in protein solubility [32]. The composition of the albumen fraction in chicken egg whites also affects their aggregation. The denaturation temperature of conalbumen, globulin, ovalbumen and lysozyme is 57.3°C; 72.0°C; 71.5°C and 81.5°C, respectively, ovomucin and ovomucoid do not coagulate at these temperatures. The lysozyme-based gel is the strongest gel, the globulin-based gel is less strong. Conalbumen gels feature the greatest drainage rate. In binary mixtures of the albumen fraction of chicken egg whites, the mixtures get aggregated close to the denaturation temperature of the least thermostable protein. The lysozyme-globulin gel is the strongest, while the ovomucoid-ovalbumen gel features the least strength [43].

**Egg white coagulation**

Accidental aggregation of already denatured protein molecules, when polymer-polymer interactions dominate over polymer-solvent interactions, leads to protein coagulation.

Coagulation is the process of particles adhesion that leads to formation of large aggregates. As a result of coagulation the system loses its sedimentation stability. There are two stages of coagulation. The first of these is the latent coagulation. At this stage the particles grow larger, but still keep their sedimentation stability. The formation of disulfide bonds and their impact on hydrophobic amino acid residues are involved in the first stage of coagulation. Proteins with a higher percentage of hydrophobic amino acids are classified as the proteins of coagulating type, while proteins with a lower percentage of hydrophobic amino acids are the proteins of gelation type. Further heating causes the egg albumen to polymerize and form a mesh. Many globular proteins with different sulphhydril groups can form thermally induced gels.

The second stage is the evident coagulation. At this stage the particles lose their sedimentation stability. If the density of the particles is higher than the density of the dispersion medium, those particles fall out. Coagulation can be caused by various factors: heat, high pressure, presence of salts, alkalis, acids, alcohols and denaturing agents, e.g. urea. In cooking conditions, the egg white coagulates at a temperature of 57–60°C, while the yolk or whole egg coagulates at slightly higher temperature (65–70°C). Adding of certain organic acids or cooking salt can raise the upper threshold of the protein thermal coagulation, which mechanism is used for eggs pasteurization [30,31,43,44].

**Changes in the functional parameters of egg white in relation to the heating temperature**

During coagulation and aggregation of proteins the disulfide bonds are redistributed, the content of Sh-groups changes, which leads to pH change. Thus, pH is a parameter that characterizes the changes in protein denaturation process — coagulation and aggregation.

The research conducted by Stefanova I. L. et al. [45] shows (Figure 1) that while heating the egg white with citric acid and cooking salt, the pH of the protein changes as follows:
- it decreases till the temperature of 65°C;
- then it gradually grows up to the original value till the temperature of (75–80) °C;
- at the temperature of 82°C pH increases sharply and grows up from 7.2–7.4 units to 8.6 units at the temperature of 92°C.

These data prove that protein coagulation occurs at a temperature of (82±2) °C. At the temperatures above 82°C the egg white clot keeps thickening at temperatures up to 88°C.

When the egg white is heated to the temperature of 80°C, the coagulated protein forms a suspension that slightly compacts, but does not separate from the whey while centrifuging at 3,500 rpm. With a further increase in temperature a clot is formed, the amount of which increase along with the increase of temperature up to 86–88°C. Then the mass of the clot does not change; only further compaction of the clot occurs.

**Composition and yield of coagulated protein depending on the level of egg white heating**

The level of heating directly affects the yield and protein composition (Figure 2). The separation of the clot begins at 82°C. At the temperatures up to 84°C, the yield of coagulated egg white increases by more than 15%. At 88°C the yield is the highest. A further increase of temperature leads to a slight decrease in yield of protein.

The temperature of denaturation transitions depends on adding of acids, alkalis and salts to protein solutions [45].
It is shown in the research [45] that while raising the amount of added citric acid from 0.10 to 0.13% simultaneously with heating up to a temperature of 86 °C, the yield of protein increases from 61 to 66% and decreases with a further buildup of acid concentration (Figure 3). The dependence of protein yield on amount of the introduced acid when the mixture is heated to a temperature of 88 °C has the same pattern, but the yield is higher. The highest yield is observed at 86 °C and 88 °C, and it accounts for 66.2% and 68.5%, respectively. Thus, the optimal citric acid concentration is 0.13%.

When the temperature of the final heating raises up to 88 °C, the yield of coagulated protein increases while the amount of drained whey decreases. The composition of the coagulated protein changes (Table 5). It was found when coagulation temperature increases and, accordingly, the yield, the content of protein and dry matter in the coagulated egg white (clot) increases, while 92.65% of the protein remains in the coagulated protein, and only 7.35% of the protein remains in the whey. The dry matter in the coagulated egg white accounts for 85.3% of dry matter in the original egg white.

Table 5. Content of protein and dry matter in coagulated egg white

| No. of test | Temperature, °C | Yield, % | Mass fraction of protein, % | Mass fraction of dry matter, % |
|-------------|----------------|----------|-----------------------------|-------------------------------|
| 1           | 82             | 46.4     | 14.8 ± 0.5                  | 17.89 ± 0.35                 |
| 2           | 84             | 61.6     | 13.8 ± 0.3                  | 16.73 ± 0.33                 |
| 3           | 86             | 65.0     | 14.1 ± 0.5                  | 17.25 ± 0.28                 |
| 4           | 88             | 70.0     | 14.1 ± 0.4                  | 17.30 ± 0.28                 |
| 5           | 90             | 69.0     | 14.2 ± 0.4                  | 17.41 ± 0.30                 |

Note: mass fraction of protein in the original egg white — 10.6 ± 0.3%, mass fraction of dry matter — 12.86 ± 0.26%
Coagulation of egg white can significantly affect its allergenic potency. Egg allergy is based on the allergy for the egg white. Sensitization to the protein components of chicken eggs is often accompanied with an allergy for the yolk and eggs of some other poultry species. Among young children the allergy for egg white is the second most common food allergy after allergy for cow’s milk protein [44]. The overall prevalence of allergy for chicken egg white among the children in European countries is about 2.5% [46]. Reducing the allergenic potency of food proteins is an important problem for food technology, which faces the challenge of manufacturing of specialized hypoallergenic foods. Although food proteins are denatured in result of heat treatment, no decrease in their allergenic potency in the general case can be guaranteed, since the allergenic sectors of the protein in some cases are short fragments of the polypeptide sequence which are resistant to denaturation. Nevertheless, the literature contains data on decrease in the allergic potencies and properties of food proteins during intense heating and cooking [47].

It may be possibly explained that the denatured food antigens are more actively attacked by digestive proteases in comparison with intact protein and, therefore, the absorption of their antigenic structures is reduced. Heat treatment destroys the conformational epitopes of egg whites, which cause the immune system of an allergic predisposed person to form IgE antibodies. A brief description of the potential allergenic properties of a range of chicken egg whites is presented in the research [48]. The authors note the complexity of the antigenic composition of the raw chicken egg white, and also state that 13 egg proteins have allergenic properties and ovalbumen (OVA) and ovomucoid (OVM) are allergic to the greatest extent. As noted, OVA is phosphoprotein with MW of 44 kDa. Protein is relatively poorly attacked by proteases and can be absorbed in the digestive tract in an undigested form. OVA is one of the most important food allergens. The protein is thermolabile (it easily denatures when heated to form an insoluble gel), but its allergenic potency decreases slightly. The antigenic determinants of OVA, which are recognized by antibodies of the IgE class, appear to be consequent [49]. OVM, due to its high resistance to proteases of the human gastrointestinal tract, quite easily penetrates the intestinal barrier and causes allergic sensitization. The antigenic structures of OVM are thermolabile, and the allergenic potency of this protein decreases sharply when heated. This is apparently

### Table 6. Mineral composition of coagulated protein

| Product               | Ca (µg/kg) | Mg (µg/kg) | K (µg/kg) | Na (µg/kg) | Fe (µg/kg) | P (µg/kg) |
|-----------------------|------------|------------|-----------|------------|------------|-----------|
| Coagulated clot       | 26.4 ± 1.8 | 56.9 ± 9.1 | 1047.3 ± 123 | 54413 ± 598 | 4.7 ± 0.3 | 16.37 ± 2.3 |

### Table 7. The content of protein and fat in the egg, depending on the mode of heat treatment

| Egg (50 g)     | Protein (g) | Calories | Fat (g) |
|----------------|-------------|----------|---------|
| Raw egg        | 6.3         | 72       | 4.7     |
| Hardboiled egg | 6.3         | 78       | 5.3     |
| Scrambled eggs | 5.0         | 75       | 5.5     |
| Poached egg    | 6.2         | 69       | 4.7     |
| Omelet         | 5.3         | 77       | 5.8     |
| Coagulated egg white | 7.5     | 30       | —       |
| Coagulated melange | 7.4   | 87       | 6.4     |

Figure 3. Dependence of coagulated protein yield on amount of added acid at different levels of heating

![Figure 3](image_url)
facilitated by the discovery that the allergenic epitopes of OVM are conformational, in contrast to OVA which are not. The promising use of thermally induced coagulation of egg white and/or chicken egg melange to reduce the allergenic potency of the resulting food is evidenced by the data that quite often people who are allergic to chicken egg whites are able to tolerate them when egg whites are heat-treated [50].

It was shown in the research [51] that thermal coagulation of egg white acidified with citric acid provides a 15-fold decrease in the original antigenicity in comparison with native egg white (Figure 4). The content of intact ovalbumen, the antigenicity of which is taken as 100%, is 2.2% and 33% respectively in the coagulated and original lyophilized egg white. The obtained results prove a decrease in the potential allergenic properties of coagulated egg white and are an important additional argument for the prospects of using the egg whites in the composition of mass market foods and in specialized foods also [52,53].

A wide range of coagulated egg foods has been developed on the basis of coagulated egg whites [33].

The coagulation process provides for fortification of chicken eggs with minor compounds. Stefanova I. L. et al. in their research [54] proved that fortification of protein in the coagulation process with iodine and calcium provides 25–30% of the daily demand for calcium and iodine.

The content of calcium in the coagulated protein, depending on the level of its heating during egg white coagulation (i.e. adding of a mineral fortifier in amount of 1% of the egg white mass during fortification of egg white with calcium and iodine) decreases together with rising of coagulation temperature from 84 °C to 90 °C, and calcium content amounts to 551.98; 518.95; 470.86 and 439.00 mg/100 g protein, respectively (Figure 5). When the temperature rises from 84 °C to 90 °C, calcium is lost at level 25.2, 28.4, 35.0, 39.4%. Basically, calcium losses occur due to its excretion with whey.

The content of calcium in the fortified melange (Figure 5) at the introduction of calcium at dose of 725 mg/100 g of melange is 425.4; 392.4; 399.4 and 396.2 mg/100 g within the range of temperatures of 86 °C; 88 °C; 90 °C; 92 °C, respectively, and it practically does not change within the temperature range 88–92 °C. Calcium losses are 44.9–45.9%. Melange binds 54.1–55.1% of the added calcium.

Egg white binds more calcium: 60.6–65.0% versus 54.1–55.1% bound by melange, despite more considerable whey drainage (23.2% for egg white at 88 °C versus 9.2% for melange at 90 °C).

The content of iodine in the coagulated protein, depending on the temperature of final coagulation (86 °C; 88 °C; 90 °C; 92 °C), showed that when fortifying the protein with seaweed powder containing 456 μg of iodine per 100 g of egg white, the mass fraction of iodine is respectively 298; 253; 311 and 281 μg/100 g of coagulated egg white, iodine losses are 34.6; 44.5; 31.8 and 38.4%, respectively. The loss of iodine during egg whites fortification during the coagulation process is lower than when iodine is added at the moment of the food formulation. When the melange is fortified with seaweed powder in amount of 0.2%, the mass fraction of iodine in the coagulated melange is 0.203; 0.243; 0.258; 0.273 μg/100 g respectively, depending on the heating temperature (86 °C; 88 °C; 90 °C; 92 °C). The share of iodine bound with melange is lower in comparison with egg white fortification with iodine, which is apparently related to the lower protein content in melange. (Figure 6).

The high frequency of metabolic syndromes, type 2 diabetes mellitus and concomitant clinical complications determine the relevance of development and creation of a wide range of new functional foods for their use in the diet of people with disorders of carbohydrate and / or fat metabolism. The results of clinical and experimental studies, so far accumulated by world nutritional science, prove the hypolipidemic and hypcholesterolaemic effects of a wide range of polyphenolic compounds.

The authors of the research [55] implemented a significant comprehensive work for development of functional specialized foods based on the egg whites fortified with cranberry polyphenolic compounds. The sorption of anthocyanins by the chicken eggs whites in the process of hydrolysis occurs to the greatest extent when the egg white is
heated to 82 ± 1°C. In this case the yield of the functional food ingredient (FFI) is 70.6–74.0% (Table 8). The anthocyanin profile is represented mainly by cyanidin-3-galactoside, peonidin-3-galactoside (Table 9).

It is possible to substitute poultry meat with coagulated egg products (egg white, yolk and melange) in amount of 15–25 in the production of semi-finished foods%, which led to creation of whole range of foods with high biological value and low fat content [56–58].

Table 8. Anthocyanin content in FFI depending on the coagulation temperature

| Temperature of coagulation, °C | 100% juice | 80 | 82 | 84 | 86 |
|-------------------------------|------------|----|----|----|----|
| 100% juice                    | 17.56      | 3.14 | 3.79 | 2.97 | 1.96 |

* Content in the solid food

Table 9. Anthocyanin profile in FFI depending on the coagulation temperature

| Anthocyanin                  | Content, % of anthocyanin amount |
|------------------------------|----------------------------------|
| Cyanidin-3-galactoside       | 28.2 | 21.1 | 21.8 | 21.8 | 19.1 |
| Cyanidin-3-glucoside         | 3.0  | 1.9  | 1.9  | 1.9  | 3.1  |
| Cyanidin-3-arabinoside       | 19.0 | 18.7 | 19.6 | 19.1 | 17.1 |
| Peonidin-3-galactoside       | 31.4 | 35.2 | 34.1 | 34.6 | 36.3 |
| Peonidin-3-glucoside         | 5.5  | 6.8  | 6.5  | 6.7  | 7.4  |
| Peonidin-3-arabinoside       | 12.9 | 16.3 | 16.0 | 15.9 | 17.0 |
| Malvidin-3-arabinoside       | traces | traces | traces | traces | traces |

Conclusions

Eggs are not only the unique mono food necessary in human nutrition. Eggs contain egg whites of a high biological value and biologically significant macro- and micronutrients. Egg whites are also a multifunctional raw material component for development and creation of new modern foods for various aims and tasks. The study of the functional and technological properties of a whole egg and its components, the influence of various parameters of technological processes on the physical and chemical properties of an egg, and ability to preserve the biologically significant key characteristics allows creating a wide range of healthy foods.

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