Intrafamilial phenotypic variation in spinocerebellar ataxia type 23

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Abstract

Background: Spinocerebellar ataxia type 23 (SCA23) is an autosomal dominant cerebellar ataxia caused by pathogenic variants in the prodynorphin gene (PDYN). The frequency of PDYN variants is reportedly very low (~ 0.1%) in several ataxia cohorts screened to date.

Case presentations: We found five cases of SCA23 in two families (mean age at onset: 37.8 ± 5.5 years; mean age at examination: 64.2 ± 12.3 years) with a novel PDYN variant (c.644G > A:p.R215H). We identified marked heterogeneity in the clinical features in Family 1: the proband showed clinical and neuroimaging features suggestive of multiple system atrophy with predominant parkinsonism (MSA-P). Conversely, the proband’s mother with the PDYN p.R215H variant had no subjective symptoms; she had not come to medical attention before our survey, although she showed apparent cerebellar atrophy on brain magnetic resonance imaging (MRI). The other two patients in Family 1 and a patient in Family 2 showed slowly progressive cerebellar ataxia.

Conclusions: We here report two Japanese families with SCA23, one of which showed considerable phenotypic variation in affected members. Our findings support that SCA23 can phenotypically overlap with MSA.

Keywords: Spinocerebellar ataxia type 23 (SCA23), Prodynorphin, Multiple system atrophy (MSA)

Introduction

Spinocerebellar ataxia type 23 (SCA23) is a rare form of autosomal dominant cerebellar ataxias (ADCAs) caused by mutations in the prodynorphin gene (PDYN) [1–5]. This rare disorder was originally identified in a Dutch family: the affected family members presented with late-onset (> 40 years), slowly progressive, pure cerebellar syndromes with hyper-reflexia [6]. Since its discovery, eight SCA23 disease-causing variants in PDYN have been identified, six of which lie exclusively within the PDYN Dyn A and Dyn B encoding regions (amino acid residues 207–236) that form the peptide known as “Big-dynorphin” [1, 2].

To the best of our knowledge, only 20 patients worldwide, including two Japanese patients in the same family, have been confirmed to carry disease-causing PDYN variants to date [1–6]. Here, we report another two Japanese families (family 1 and family 2) with SCA23 carrying a novel PDYN variant, p.R215H. Family 1 included three patients heterozygous and one patient homozygous for the PDYN variant: the proband exhibited clinical and neuroimaging features resembling multiple system atrophy with predominant parkinsonism (MSA-P).

Case report

The clinical details of the patients from Families 1 and 2 are summarized in Table 1.
Family 1: The proband (Fig. 1, II-1) first noted an unsteady gait at 45 years-of-age. At ~ 58 years of age, he developed speech disturbance and right hand tremor. These symptoms gradually worsened, prompting him to visit a hospital at 62 years-of-age. A neurological examination revealed saccadic eye movements, scanning speech, hyperreflexia of all limbs, and parkinsonism such as bradykinesia, rigidity, frozen gait, and a resting tremor affecting both hands (R > L). He also complained of constipation, but did not have orthostatic hypotension or urinary incontinence. He could walk unaided and his ataxia was very mild [Scale for the Assessment and Rating of Ataxia (SARA) score: 5.0/40.0]. Brain magnetic resonance imaging (MRI) revealed mild atrophy of the cerebellar hemisphere and vermis, as well as a thin hyperintense lateral putaminal rim (Fig. 2 a-d). A hot cross bun sign and/or hyperintensities in the middle cerebellar peduncles were not evident. His parkinsonism responded slightly to levodopa (Levodopa/Carbidopa hydrate 300 mg/day) and other anti-parkinsonian drugs (Pramipexole Hydrochloride 1.5 mg/day and Selegiline hydrochloride 0.5 mg/day). Dopamine transporter single-photon emission computed tomography (DAT-SPECT) at 65 years-of-age showed severe bilateral nigrostriatal dopaminergic deficits bilaterally predominantly in the putamen (Fig. 3).

The younger brother of the proband (Fig. 1, II-4) noticed difficulty in speaking smoothly at around 36 years-of-age. A neurological examination at 37 years-of-age revealed scanning speech, dysmetria of the upper limbs, and increased deep tendon reflexes of the legs. A brain computed tomography scan showed cerebellar atrophy and he was subsequently diagnosed with spinocerebellar degeneration (SCD). At 59 years-of-age, he needed a wheelchair because of ataxic gait, and his dysarthria became very severe; his SARA score was 19.0/40.0. Brain MRI scan revealed moderate atrophy of the cerebellar hemisphere and vermis, but brainstem atrophy or

| Table 1: Clinical and neuroimaging features of the patients with SCA23 |
|---------------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Family No.          | 1               | 1               | 1               | 1               | 2               |
| Pedigree number/Sex | II-1 (male)     | II-4 (male)     | II-8 (male)     | I-2 (female)    | II-1 (male)     |
| Age at examination  | 62              | 59              | 52              | 88              | 60              |
| Age at onset (years)| 45              | 36              | 40              | 40              | 30              |
| Disease duration    | 17              | 23              | 12              | 30              |
| Initial symptom     | Unsteadiness    | Dysarthria      | Dysarthria      | Dysarthria      |
| Walking ability     | Unaided         | Wheelchair      | Unaided         | Crutches/cane   |
| SARA                | 5.0/40.0        | 19.0/40.0       | 9.0/40.0        | 14.5/40.0       |
| Dysarthria          | (++)            | (+++)           | (+)             | (−)             |
| EOM disorder        | Saccadic        | (−)             | (−)             | (−)             | Gaze nystagmus  |
| Tremor              | Resting tremor(−)| (−)             | Cerebellar tremor | (−)             |
| Parkinsonism        | (+)             | (−)             | (−)             |
| Deep tendon reflex  | Increased (U/L)| Increased (L)   | Increased (L)   | Normal          |
| Babinski sign       | (−)             | (−)             | (−)             |
| Muscle atrophy & weakness | (−)             | (−)             | (−)             |
| Sensory deficits    | (−)             | (−)             | NA              |
| Urinary symptoms    | (−)             | (−)             | (−)             |
| Atrophy of the cerebrum | (−)             | (−)             | (−)             |
| cerebellum c.       | (+)             | (++)            | (++)            | (++)            |
| Brainstem (pons)    | (−)             | (−)             | (−)             |
| Middle cerebellar peduncle | (−)             | (−)             | (−)             | (++)            |
| Hyperintense lateral putaminal rim | (+)             | (−)             | (−)             | (−)             |
| Hot cross bun sign  | (−)             | (−)             | (−)             | (−)             |
| PDYN (c.644G > A) genotype | Heterozygote | Homozygote | Heterozygote | Heterozygote | Heterozygote |

NA: Information not available. SARA: Scale for the Assessment and Rating of Ataxia. a This patient had no subjective symptoms and had never visited a neurology clinic before we performed a medical survey on the family members of the proband (II-1). b Parkinsonism means bradykinesia, small steps, and frozen gait. c All patients showed atrophy of both cerebellar hemispheres and vermis. (−): none, (+): mild, (++): moderate, (+++): severe, (U): upper extremities, (L): lower extremities
abnormal signals, including a hot cross bun sign or hyperintensities in the middle cerebellar peduncles were not evident.

The proband’s youngest brother (Fig. 1, II-8) developed difficulty in speech and unsteadiness at 40 years-of-age. Brain MRI at 42 years-of-age revealed moderate cerebellar atrophy. A later neurological examination at 52 years-of-age revealed a cerebellar tremor and hyperreflexia of the lower limbs. His gait was ataxic, but he could walk unaided; his SARA score was 9.0/40.0.

Their parents of these affected siblings were first cousins. Their mother (Fig. 1, I-2) was hospitalized for

![Pedigrees of Families 1 and 2 and Sanger sequencing for PDYN](image)

**Fig. 1** Pedigrees of Families 1 and 2 and Sanger sequencing for PDYN. The numbers below the symbols indicate the age (in years, y) at examination and the age at death (d) for the deceased individuals (indicated by the strikethrough). The probands are indicated by the arrows.

![Brain MRI](image)

**Fig. 2** Brain MRI (a–d): Patient II-1 in Family 1 had mild atrophy of the cerebellar hemisphere and vermis. A hyperintense lateral putamen rim (arrows) was seen. (e–h): Patient II-1 in Family 2 had moderate atrophy of the cerebellar hemisphere and vermis. No brainstem atrophy, hot cross bun sign, or atrophy/hyperintensities in the middle cerebellar peduncles was observed in either patient.
crowned dens syndrome at 88 years-of-age, and this was the first time she had undergone any neurological examinations. She also had chronic heart failure and osteoarthritis of the knee joints, but had no subjective complaints of cerebellar ataxia, and was almost independent in her daily life. However, her gait was ataxic and she needed a cane for walking; her SARA score was 14.5/40.0. Brain MRI showed moderate cerebellar atrophy. We assumed that she had cerebellar ataxia like her affected children, but the age of onset was ambiguous. She died of heart failure at 89 years-of-age. The father of the proband (Fig. 1, I-1) died of lung cancer at 70 years-of-age. Although his clinical details were not available, his close family members reported that he did not have dysarthria or a gait disturbance during his lifetime.

Family 2: The proband (Fig. 1, II-1) became aware of slurred speech at 30 years-of-age. At around 45 years-of-age, he noticed a disturbance of skillful movements of both hands that prompted a visit to hospital. Brain MRI scan revealed cerebellar atrophy and he was subsequently diagnosed with SCD that was treated with the thyrotropin-releasing hormone analog, taltirelin. A detailed neurological examination at 60 years-of-age revealed mild dysarthria, horizontal nystagmus, a cerebellar tremor affecting both hands, decreased deep tendon reflex without muscle atrophy or sensory disturbance. He could walk unaided and his SARA score was 7.5/40.0. Brain MRI identified moderate atrophy of the cerebellar hemisphere and vermis, but no atrophy or abnormal signals in the brainstem (Fig. 2e-h).

The proband’s brother (Fig. 1, II-3) exhibited speech disturbance from ~40 years-of-age, but his clinical details were not available. His father (Fig. 1, I-1) died from pneumonia at 84 years-of-age, while his mother (Fig. 1, I-2) had dementia and had been institutionalized in a nursing home. Neither parent was reported to have ataxic symptoms during their lifetime.

**Diagnostic evaluation**

We performed a molecular diagnosis using genomic DNA extracted from a blood sample from each patient. Screening for common repeat expansions for ADCAs (SCA1, SCA2, MJD/SCA3, SCA6, SCA7, SCA8, SCA12, SCA17, and DRPLA) and SCA31 was negative for the proband in each family (II-1 in Family 1, II-1 in Family 2). We then performed targeted re-sequencing for both probands in each family using a TruSight One Sequencing Panel (Illumina) and a MiSeq benchtop sequencer (Illumina). Here we sequenced previously reported causative genes of autosomal dominant and recessive cerebellar ataxias.

We identified a c.644G > A:p.R215H variant in PDYN exon 3 in the proband of each family. Sanger sequencing confirmed that four patients (Family 1: I-2, II-1, and II-8; Family 2: II-1) were heterozygous for this variant, and one patient (Family 1: II-4) was homozygous for this variant (Fig. 1). This variant was registered in the Single Nucleotide Polymorphism database (dbSNP138) as rs201655505, with a minor allele frequency of 0.00003 according to the Exome Aggregation Consortium (ExAC) database [7]. Further analyses using SIFT (http://sift.jcvi.org/), PolyPhen-2 (http://genetics.bwh.harvard.edu/pph2/), and MutationTaster (http://www.mutationtaster.org/) predicted that this missense variant was deleterious. Furthermore, a different missense change at the same amino acid residue, p.R215C, has been demonstrated to be pathogenic [1]. Based on these data, we consider p.R215H as “likely pathogenic” according to the American College of Medical Genetics (ACMG) standards and guidelines [8].
Discussion and conclusions
The clinical phenotype of SCA23 is highly variable, but its cardinal feature is a late onset, slowly progressive cerebellar ataxia [1, 6]. Interestingly, we found marked intrafamilial variation in the clinical and neuroimaging features in the affected members of Family 1. In particular, the proband (II-1) had suspected MSA-P before the diagnosis of SCA23 was made. Because this patient had no urinary dysfunction (incon tinence, urgency, or incomplete bladder emptying) or orthostatic hypotension, he did not fulfill the second consensus criteria of probable or possible MSA [9]. Saigoh et al. reported the first Japanese case who had a hot cross bun sign and hypertensive lateral putaminal rim on brain MRI and suggested that the pontocerebellar tract or striatonigral tract may be damaged in SCA23 [5]. Together with the previous report [5], our cases suggest that the existence of a phenotypic overlap between SCA23 and MSA, both clinically and neuroradiologically.

On the other hand, the parents of the proband in Family 1 did not exhibit signs of cerebellar ataxia until they had reached an advanced age. This finding is in stark contrast to the fact that their affected children developed neurological disorders in the young-adult ages. The low penetrance rate for PDYN variants might have contributed to the marked intrafamilial variation observed in this family. Another possibility is that genetic anticipation occurred in this family, as seen non-repeat expansion mutations in TMEM240 that underlie SCA21 [10, 11].

Family 1 consisted of patients who were heterozygous and homozygous for the variant. The age of onset for the homozygous patient (II-4) was a little younger than that for the heterozygous patients, and the ataxic symptoms were the most severe. As for the pathogenesis of SCA23, both ‘toxic’ gain of function [1, 12, 13] and loss of function [2] mechanisms have been postulated. Although the clinical difference between heterozygous and homozygous patients in Family 1 does not support either a ‘toxic’ gain-of-function or loss-of-function mechanism, it might suggest that the dosage effect of the variant or normal PDYN proteins is directly associated with the clinical severity of SCA23.

In conclusion, we identified two Japanese families with SCA23 likely driven by an underlying, novel PDYN p.R215H variant. The cardinal clinical feature of the patients was a slowly progressive cerebellar ataxia with young-adult onset. Importantly, one patient showed clinical and neuroradiological features similar to MSA-P.

Abbreviations
SCA23: Spinocerebellar ataxia type 23; PDYN: Prodynorphin; ADCA: Autosomal dominant cerebellar ataxia; MSA-P: Multiple system atrophy with predominant parkinsonism; SARA: Scale for the assessment and rating of ataxia; SCD: Spinocerebellar degeneration; ExAC: Exome aggregation consortium; ACMG: American college of medical genetics

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Authors’ contributions
SS, SO, and KY contributed to the conception and design of the study. SS and SO were responsible for collection of the clinical data. KN, TT, and KY conducted genetic testing and analyzed genetic data. All authors read and approved the final manuscript. YK and KY wrote the manuscript and all authors were responsible for its review and critique.

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Availability of data and materials
The data in this study are not publicly available as they could compromise the anonymity of the subjects.

Ethics approval and consent to participate
The Institutional Review Board of Shinshu University School of Medicine approved all methods for this study. After genetic counseling, the patients provided written informed consent for participation in this study.

Consent for publication
Not applicable.

Competing interests
The authors report no competing interests.

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References
1. Bakalkin G, Watanabe H, Jezierska J, Depoorter C, Verschuuren-Bernelmans C, Bazov I, et al. Prodynorphin mutations cause the neurodegenerative disorder spinocerebellar ataxia type 23. Am J Hum Genet. 2010;87:593–603.
2. Jezierska J, Stevanin G, Watanabe H, Fekkes MR, Zagnoli F, Kok J, et al. Identification and characterization of novel PDYN mutations in dominant cerebellar ataxia cases. J Neurol. 2013;260:1807–12.
3. Fogel BL, Lee JY, Lane J, Wahnich A, Chan S, Huang A, et al. Mutations in rare ataxia genes are uncommon causes of sporadic cerebellar ataxia. Mov Disord. 2012;27:442–6.
4. Fawcett K, Mehrabian M, Liu YT, Hamed S, Elahi E, Revesz T, et al. The frequency of spinocerebellar ataxia type 23 in a UK population. J Neurol. 2013;260:856–9.
5. Saigoh K, Mitsui J, Hirano M, Shioyama M, Samukawa M, Ichikawa Y, et al. The first Japanese familial case of spinocerebellar ataxia 23 with a novel mutation in the PDYN gene. Parkinsonism Relat Disord. 2015;21:332–4.
6. Verbeek DS, van de Warrenburg BP, Wesseling P, Pearson PL, Kremer HP, Sinke RJ. Mapping of the SCA23 locus involved in autosomal dominant cerebellar ataxia to chromosome region 20p13–12.3. Brain. 2004;127:2551–7.

7. gnomAD browser | Genome Aggregation Database. https://gnomad.broadinstitute.org/variant/20-1961090-C-T?dataset=gnomad_r2_1. Accessed 18 June 2020.

8. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American college of medical genetics and genomics and the association of molecular pathology. Genet Med. 2015;17:405–24.

9. Gilman S, Wenning GK, Low PA, Brooks DJ, Mathias CJ, Trojanowski JQ, et al. Second consensus statement on the diagnosis of multiple system atrophy. Neurology. 2008;71:670–6.

10. Devos D, Schraen-Maschke S, Vuillaume I, Dujardin K, Nazé P, Willoteaux C, et al. Clinical features and genetic analysis of a new form of spinocerebellar ataxia. Neurology. 2001;56:234–8.

11. Delplanque J, Devos D, Huin V, Genet A, Sand O, Moreau C, et al. TMEM240 mutations cause spinocerebellar ataxia 21 with mental retardation and severe cognitive impairment. Brain. 2014;137:2657–63.

12. Watanabe H, Mizoguchi H, Verbeek DS, Kuzmin A, Nyberg F, Krishtal O, et al. Non-opioid nociceptive activity of human dynorphin mutants that cause neurodegenerative disorder spinocerebellar ataxia type 23. Peptides. 2012;35:306–10.

13. Smeets CJ, Jezierska J, Watanabe H, Duari A, Fokkens MR, Meijer M, et al. Elevated mutant dynorphin a causes Purkinje cell loss and motor dysfunction in spinocerebellar ataxia type 23. Brain. 2015;138:2537–52.

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