Short communication

Anticancer effects of radiation therapy combined with Polo-Like Kinase 4 (PLK4) inhibitor CFI-400945 in triple negative breast cancer

Armen Parsyan a, b, c, d, *, Jennifer Cruickshank e, Kelsey Hodgson e, Drew Wakeham e, Sierra Pellizzari d, Vasudeva Bhat e, d, David W. Cescon e, f

a Department of Surgery, St Joseph’s Health Care and London Health Sciences Centre, Western University, London, Ontario, N6A 4V2, Canada
b Department of Oncology, Western University, London, Ontario, N6A 5W9, Canada
c London Regional Cancer Program, London Health Sciences Centre, Western University, London, Ontario, N6A 5W9, Canada
d Department of Anatomy and Cell Biology, London Regional Cancer Program, Western University, London, Ontario, N6A 5C1, Canada
e Princess Margaret Cancer Centre, University Health Network, University of Toronto, Toronto, Ontario, M5G 2C1, Canada
f Department of Medical Oncology and Hematology, Princess Margaret Cancer Centre, University Health Network, University of Toronto, Toronto, Ontario, M5G 2C1, Canada

Article info

Article history:
Received 26 December 2020
Received in revised form 7 March 2021
Accepted 29 March 2021
Available online 1 April 2021

Keywords:
Triple-negative breast cancer
Radiotherapy
Polo-like kinase 4
CFI-400945
Radiosensitization
Patient-derived organoids

Abstract

Development of novel multimodality radiotherapy treatments in metastatic breast cancer, especially in the most aggressive triple negative (TNBC) subtype, is of significant clinical interest. Here we show that a novel inhibitor of Polo-Like Kinase 4 (PLK4), CFI-400945, in combination with radiation, exhibits a synergistic anti-cancer effect in TNBC cell lines and patient-derived organoids in vitro and leads to a significant increase in survival to tumor endpoint in xenograft models in vivo, compared to control or single-agent treatment. Further preclinical and proof-of-concept clinical studies are warranted to characterize molecular mechanisms of action of this combination and its potential applications in clinical practice.

© 2021 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Introduction

Local disease control at primary or metastatic sites in patients with metastatic or inoperable breast cancer is commonly addressed with radiotherapy. However, outcomes remain poor, and progression in irradiated areas is not uncommon [1–4]. Development of novel combined modality radiotherapy treatments in metastatic or inoperable disease, especially in the most aggressive triple negative breast cancer (TNBC) subtype, is therefore of significant clinical interest. With the development of more selective chemical inhibitors, there has been a renewed interest in agents targeting regulators of genomic stability and cell cycle which can further exacerbate numerical chromosomal instability (CIN) and lead to cellular lethality [5,6]. Ionizing radiation induces genotoxic stress and aneuploidy and may thus synergize with agents targeting cell cycle checkpoints [7–9]. CIN itself mediates susceptibility to radiation [10,11], and therefore therapies that induce CIN may increase radiosensitivity. Polo-like kinase 4 (PLK4) is a key regulator of the cell cycle and centriole duplication that is aberrantly expressed in breast cancer and is associated with CIN [12,13]. PLK4 was identified as a promising anticancer therapeutic target in TNBC, and a first-in-class inhibitor, CFI-400945, has been recently characterized [14,15]. CFI-400945 exacerbates CIN, has been shown to have antitumor activity in preclinical models, including TNBC, and is being evaluated in clinical trials in patients with metastatic breast cancer, and other cancer types [14,16–20]. We therefore assessed whether radiation and CFI-400945 exhibit combined anticancer effects in breast cancer cell lines and patient-derived organoids (PDO).

Methods

Colony formation assays were performed for breast cancer cell
In vitro radiation was performed using an X-RAD 320 irradiator (Accela), where cells were treated with various doses of radiation. The media was replaced and supplemented with varying concentrations of CFI-400945 or vehicle control. Synergy of dose-response matrix data was assessed by Bliss synergy score using SynergyFinder Software [23]. All patient samples (Supplementary Table 1) were obtained after participants’ consent and used in accordance with a research ethics board-approved protocol (REB#159481). Organoids were generated and propagated as previously described [24]. Organoid cancer cells were plated (2000 cells/well) in Basement Membrane Extract, Type II (Bio-Techne) in 48-well plates. Animal experiments were performed under an institutionally approved protocol (#1113). MDA-MB-231 cells (2 × 10^6) were injected into the mammary fat pad of NOD/SCID mice. Tumor endpoint survival curve (left) using Kaplan-Meier analysis. P-value was calculated using log-rank test. No significant effects on animal general health and weight were observed, and no detectable metastases (liver and lungs) were seen on the autopsy in all study arms. Significantly smaller proportion of mice developed tumor ulceration in the combination treatment arm compared to other treatment arms.

**Fig. 1.** Effects of CFI-400945 and Radiation in Triple Negative Breast Cancer In Vitro and In Vivo Models. A. Combined CFI-400945 and radiation resulted in dose-dependent reductions in colony formation in MDA-MB-231. The number of colonies in each treatment arm were normalized to the respective control (no-radiation, no-drug). B. Synergy of combined CFI-400945 and radiation was observed across several dose levels. Bliss synergy scores, calculated with SynergyFinder, are displayed in the heatmap, where intensity of red indicates higher degree of synergy. C. Organoid formation assay using various concentrations of CFI-400945 alone (top panel) or in combination with 3 Gy radiation (bottom panel) in BPTO19, generated from a chest wall metastasis of a TNBC patient. Bright-field microscopy (at 4× magnification) images were taken 28 days following treatment. The number of organoids was counted by 2 independent observers in at least 3 random fields per each well. The counts were normalized to respective unirradiated controls in each group. Bar represents 100 μm. D. Effect of CFI-400945 and radiation in BPTO19. Average number of organoids was normalized to that of control (no-radiation, no-drug). Averages and standard deviations (SD, bars) of replicate experiments are presented. E. In Vivo effects of CFI-400945 and Radiation on MDA-MB-231 Xenografts in NOD/SCID mice. Tumor endpoint survival curve (left) using Kaplan-Meier analysis. P-value was calculated using log-rank test. No significant effects on animal general health and weight were observed, and no detectable metastases (liver and lungs) were seen on the autopsy in all study arms. Significantly smaller proportion of mice developed tumor ulceration in the combination treatment arm compared to other treatment arms.

### Abbreviations

- **TNBC**: triple negative breast cancer
- **BME**: basement membrane extract
- **CIN**: chromosomal instability
- **CDK4/6**: cyclin-dependent kinase 4/6
- **PLK4**: Polo-Like Kinase 4
- **PDO**: patient-derived organoids
- **PDXO**: patient-derived organoids established from a xenograft
- **NOD/SCID**: Nonobese diabetic/severe combined immunodeficiency

**Fig. 1.** Effects of CFI-400945 and Radiation in Triple Negative Breast Cancer In Vitro and In Vivo Models. A. Combined CFI-400945 and radiation resulted in dose-dependent reductions in colony formation in MDA-MB-231. The number of colonies in each treatment arm were normalized to the respective control (no-radiation, no-drug). B. Synergy of combined CFI-400945 and radiation was observed across several dose levels. Bliss synergy scores, calculated with SynergyFinder, are displayed in the heatmap, where intensity of red indicates higher degree of synergy. C. Organoid formation assay using various concentrations of CFI-400945 alone (top panel) or in combination with 3 Gy radiation (bottom panel) in BPTO19, generated from a chest wall metastasis of a TNBC patient. Bright-field microscopy (at 4× magnification) images were taken 28 days following treatment. The number of organoids was counted by 2 independent observers in at least 3 random fields per each well. The counts were normalized to respective unirradiated controls in each group. Bar represents 100 μm. D. Effect of CFI-400945 and radiation in BPTO19. Average number of organoids was normalized to that of control (no-radiation, no-drug). Averages and standard deviations (SD, bars) of replicate experiments are presented. E. In Vivo effects of CFI-400945 and Radiation on MDA-MB-231 Xenografts in NOD/SCID mice. Tumor endpoint survival curve (left) using Kaplan-Meier analysis. P-value was calculated using log-rank test. No significant effects on animal general health and weight were observed, and no detectable metastases (liver and lungs) were seen on the autopsy in all study arms. Significantly smaller proportion of mice developed tumor ulceration in the combination treatment arm compared to other treatment arms.
Statistically significant prolongation of survival times. P-values were calculated using a log-rank test (Prism 8, GraphPad Software).

**Results**

In the MDA-MB-231 TNBC cell line (Fig. 1A), synergy of combined treatment was observed at 10 and 20 nM of CFI-400945 (Fig. 1A) and was greatest at 10 nM of CFI-400945 and 2 Gy of radiation (Fig. 1B). Similar synergistic relationships were observed in two other TNBC lines, MDA-MB-468 and MDA-MB-436 (Supplementary Fig 1). In the TNBC PDO model, BPTO19, 1 nM of CFI-400945 had no significant effect on organoid formation, while at 50 nM no organoid formation was observed (Fig. 1C and D). At 5 nM concentration, CFI-400945 alone caused 2.1 times decrease in colony formation. Irradiation with 3 Gy reduced the number of organoids by 1.6 times, compared to no-irradiation control. When combined with CFI-400945 at 1 or 5 nM, irradiation resulted in an average 2.8- and 5.6-fold decrease in organoid formation respectively, compared to no treatment control. Similar effects were observed in 2 other organoid models, BXT081 and BXT064 (Supplementary Fig 2). We then assessed the combination of orally delivered CFI-400945 with a single fraction of targeted radiation to MDA-MB-231 xenografts in NOD/SCID mice (Fig. 1E). In the control arm, median survival to tumor humane endpoint was 32 days, which was increased to 43, 57 and 75 days for radiation-only, CFI-400945-only and combination treatment respectively (Fig. 1E). Statistically significant ($p < 0.0005$) improvements in survival comparing combination versus single treatments or control groups were observed. Similarly, analysis of the median time to tumor volume tripling showed that combination treatment resulted in a significantly prolonged tumor volume tripling time, compared to control, drug-only or radiation-only arms (Supplementary Fig 3). No significant effects on animal general health and weight were observed.

**Discussion**

Augmenting the effects of radiation with chemotherapeutic agents or targeted therapy is an attractive strategy to improve survival and quality of life of cancer patients, through achieving synergistic antitumor activity and minimization of the overlapping toxic effects [25,26]. This strategy has not been incorporated into the standard clinical management of breast cancer, though chemotherapy is sometimes used, and targeted combinations are being explored in clinical trials [27]. Our results demonstrate combination synergy and radiosensitizing effects of CFI-400945 across multiple preclinical TNBC models. We show that the combined delivery of these treatments in vivo results in significant prolongation of survival and delay in growth of tumors. CFI-400945 has been shown to be safe and well-tolerated in humans and is being tested in Phase 2 clinical trials in patients with advanced/metastatic breast cancer [19,20]. Emerging data suggest synergistic effects of combining radiotherapy with other novel therapeutic compounds, particularly those that affect cell cycle control and exacerbate CIN. In patients with breast cancer brain metastases receiving CDK4/6 inhibitors, delivery of radiotherapy has been reported to confer prolonged survival outcomes [28]. Ongoing clinical trials are evaluating CDK4/6 inhibitors in combination with radiotherapy in patients with bone metastases and oligometastatic breast cancer [29,30]. Similar to our results, an inhibitor of the spindle assembly checkpoint control protein TTK has been shown in preclinical studies to confer radiosensitization effects and substantially improved tumor control in TNBC [31]. Our findings support development of further preclinical studies of this combination and its molecular mechanisms, as well as early clinical trials to evaluate safety/toxicity in patients with metastatic breast cancer.

**Declaration of competing interest**

D.W.C serves as a consultant for Agendia, Dynamo Therapeutics, AstraZeneca, Exact Sciences, GSK, Merck, Novartis, Pfizer, Puma and Roche; receives research support (to institution) from GSK. Merck and Pfizer and holds intellectual property as co-investigator on a patent related to biomarkers for TTK inhibitors. The other authors declare no potential conflicts of interests.

CFI-400945 was developed by the Therapeutics group at the authors’ institution (University Health Network, Toronto, ON), and has been licensed to Treadwell Therapeutics.

**Acknowledgments**

This research was conducted with the support of the Terry Fox Research Institute (New Frontiers Research Program PPG-1064, to D.W.C) and Stand Up To Cancer Canada – Canadian Cancer Society Breast Cancer Dream Team Research Funding, with supplemental support of the Ontario Institute for Cancer Research through funding provided by the Government of Ontario (Funding Award Number: SU2C-AACR-DT-18-15). Stand Up To Cancer Canada is a program of the Entertainment Industry Foundation Canada. Research funding is administered by the American Association for Cancer Research International — Canada, the Scientific Partner of SU2C Canada. This research was also supported by the Young Investigator Startup Grant, Department of Surgery, Western University and the London Regional Cancer Program Catalyst Grant for Translational Cancer Research, Western University (London, ON) to A.P.

A. P. was supported by the Strategic Training in Transdisciplinary Radiation Science for the 21st (STARS21, Terry Fox Foundation, Princess Margaret Radiation Medicine Program and Cancer Centre), Princess Margaret Cancer Centre Scholarship (Toronto, ON); Department of Surgery, University of Toronto (ON); the Clinician Scientist Award, Department of Surgery, Western University and the Academic Medical Organization of Southwestern Ontario (AMOSO) Opportunities Fund (London, ON).

V. B. was supported by Western Postdoctoral Fellowship (Western University, London, ON) and the Breast Cancer Society of Canada Postdoctoral Fellowship. S.P was supported by the Translational Breast Cancer Research Unit Trainee Scholarship (Source: Breast Cancer Society of Canada), London, ON.

We thank Drs Kelsie L. Thu and Isabel Soria-Bretones for input on experimental methodologies. PDO and PDO models were developed in collaboration with the Princess Margaret Living Biobank (PMLB, Toronto, ON).

**Appendix A. Supplementary data**

Supplementary data to this article can be found online at https://doi.org/10.1016/j.breast.2021.03.011.
References

[1] Rades D, Fischer D, Veninga T, Stalpers LJ, Schild SE. Prognostic factors for survival and intracerebral control after irradiation for brain metastases from gynecological cancer. Gynecol Oncol 2009;114:506–8.

[2] Milano MT, Katz AW, Zhang H, Okunieff P. Oligometastases treated with stereotactic body radiotherapy: long-term follow-up of a prospective study. Int J Radiat Oncol Biol Phys 2012;83:878–86.

[3] Oral C, Guler OC, Yildirim BA. Treatment outcomes of breast cancer liver metastasis treated with stereotactic body radiotherapy. Breast Cancer 2018;42:150–8.

[4] Dellas E. Does radiotherapy have curative potential in metastatic patients? The concept of local therapy in oligometastatic breast cancer. Breast Care (Basel) 2011;6:363–8.

[5] Dominguez-Brauer C, Thu KL, Mason JM, Blaser H, Bray MR, Mak TW. Targeting mitosis in cancer: emerging strategies. Mol Cell 2015;60:524–36.

[6] Thu KL, Soria-Bretones I, Mak TW, Cescon DW. Targeting the cell cycle in breast cancer: towards the next phase. Cell Cycle 2018;17:1871–85.

[7] Citrin DE, Mitchell JB. Mechanisms of normal tissue injury from irradiation. Semin Radiat Oncol 2017;27:316–24.

[8] Cho YH, Kim SY, Woo HD, Kim YJ, Ha SW, Chung HW. Delayed numerical chromosome aberrations in human fibroblasts by low dose of radiation. Int J Environ Res Publ Health 2015;12:15162–72.

[9] Niu N, Qin Y, Fridley BL, Hou J, Kalari KR, Zhu M, et al. Radiation pharmacogenomics: a genome-wide association approach to identify radiation response biomarkers using human lymphoblastoid cell lines. Genome Res 2010;20:1482–92.

[10] Balkhoun SF, Kabeche L, Wood MD, Lucarius CD, Qu D, Laughney AM, et al. Numerical chromosomal instability mediates susceptibility to radiation treatment. Nat Commun 2015;6:5990.

[11] Yao D, Xu X, Yang L, Zhu J, Wan J, Shen L, et al. Patient-derived organoids predict chemoradiation responses of locally advanced rectal cancer. Cell Stem Cell 2020;26:17–26 e6.

[12] Ianevski A, He L, Aittokallio T, Tang J. SynergyFinder: a web application for analyzing drug combination dose-response matrix data. Bioinformatics 2017;33:2413–5.

[13] Sachs N, de Ligt J, Kopper O, Gogola E, Bounova G, Weeber F, et al. A living Biobank of breast cancer organoids captures disease heterogeneity. Cell 2018;172:373–386 e10.

[14] Rosenzweig KE, Gomez JE. Concurrent chemotherapy and radiation therapy for inoperable locally advanced non-small-cell lung cancer. J Clin Oncol 2017;35:6–10.

[15] Sharma RA, Plummer R, Stock JK, Greenhalgh TA, Ataman O, Kelly S, et al. Clinical development of new drug-radiotherapy combinations. Nat Rev Clin Oncol 2016;13:627–42.

[16] Loap P, Loirat D, Berger F, Ricci F, Vincent-Salomon A, Ezzili C, et al. Combination of olaparib and radiation therapy for triple negative breast cancer: preliminary results of the RADIOPARP phase 1 trial. Int J Radiat Oncol Biol Phys 2021;109:436–40.

[17] Veitch ZW, Cescon DW, Denny T, Yonemoto LM, Fletcher G, Broks R, et al. Safety and tolerability of CFI-400945, a first-in-class, selective PLK4 inhibitor in advanced solid tumours: a phase 1 dose-escalation trial. Br J Canc 2019;121:318–24.

[18] Cescon D, Pezo R. CFI-400945 in patients with advanced/metastatic breast cancer. In: ClinicalTrials.gov, editor. Accessed February 23, 2021.

[19] Franken NA, Rodermond HM, Spa J, Haveman J, van Bree C. Clonogenic assay of cells in vitro. Nat Protoc 2006;1:2315–9.

[20] Yao Y, Xu X, Yang L, Zhu J, Wan J, Shen L, et al. Patient-derived organoids predict chemoradiation responses of locally advanced rectal cancer. Cell Stem Cell 2020;26:17–26 e6.

[21] Ianevski A, He L, Aittokallio T, Tang J. SynergyFinder: a web application for analyzing drug combination dose-response matrix data. Bioinformatics 2017;33:2413–5.

[22] Sachis N, de Ligt J, Kopper O, Gogola E, Bounova G, Weeber F, et al. A living Biobank of breast cancer organoids captures disease heterogeneity. Cell 2018;172:373–386 e10.

[23] Rosenzweig KE, Gomez JE. Concurrent chemotherapy and radiation therapy for inoperable locally advanced non-small-cell lung cancer. J Clin Oncol 2017;35:6–10.

[24] Sharma RA, Plummer R, Stock JK, Greenhalgh TA, Ataman O, Kelly S, et al. Clinical development of new drug-radiotherapy combinations. Nat Rev Clin Oncol 2016;13:627–42.

[25] Loap P, Loirat D, Berger F, Ricci F, Vincent-Salomon A, Ezzili C, et al. Combination of olaparib and radiation therapy for triple negative breast cancer: preliminary results of the RADIOPARP phase 1 trial. Int J Radiat Oncol Biol Phys 2021;109:436–40.

[26] Figura NB, Potluri TK, Mohammad H, Oliver DE, Arrington JA, Robinson TJ, et al. CDK 4/6 inhibitors and stereotactic radiotherapy in the management of hormone receptor positive breast cancer brain metastases. J Neuro Oncol 2019;144:581–9.

[27] Jeong J. Local treatment in ER-positive/HER2-negative oligo-metastatic breast cancer (CLEAR). ClinicalTrials.gov; November 2018;23.

[28] Figura NB, Potluri TK, Mohammad H, Oliver DE, Arrington JA, Robinson TJ, et al. CDK 4/6 inhibitors and stereotactic radiotherapy in the management of hormone receptor positive breast cancer brain metastases. J Neuro Oncol 2019;144:581–9.

[29] Jeong J. Local treatment in ER-positive/HER2-negative oligo-metastatic breast cancer (CLEAR). ClinicalTrials.gov; November 2018;23.

[30] Torres M. Radiation Therapy, Palbociclib, and Hormone Therapy in Treating Breast Cancer Patients with Bone Metastasis (ASPIRE). NCT03691493. October 1, 2018. ClinicalTrials.gov.

[31] Chandler BC, Moubadler L, Ritter CL, Liu M, Cameron M, Wilder-Romans K, et al. TTFields inhibition radiosensitizes basal-like breast cancer through impaired homologous recombination. J Clin Invest 2020;130:958–73.