Epstein-Barr Virus (EBV) infection related thrombocytosis: A Case Report

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Introduction

EBV infection can occur in any part of the world and infection is common in early childhood and the second peak can occur during late adolescence, most of the EBV infections in infants and young children are asymptomatic. Whereas around 75% of infections in adolescents may present with infectious mononucleosis, common symptoms of this infection are sore throat, malaise, headache, abdominal pain and common signs are lymphadenopathy, fever, pharyngitis, hepatomegaly, splenomegaly, rash. Some adults (less than 0.5%) have been reported to have life-threatening hematological complications like subcapsular splenic hemorrhage or splenic rupture. Other hematological complications: Coombs test positive hemolytic anemia, severe thrombocytopenia, neutropenia, and aplastic anemia. Many cases of ITP turning to aplastic anemia have been found in the literature. IgM and IgG antibodies directed against the Epstein-Barr virus capsid antigen (VCA) are usually present at the onset of clinical illness because of the long viral incubation period. IgM levels wane approximately three months later; thus, they are a good marker of acute infection. IgG VCA antibodies persist for life and are a marker of EBV infection.

Case Report:

A seventeen year-old male presented with complaints of easy fatigability of 4 months duration, abdominal pain of 1 months duration and fever for 5 days. He had decreased energy associated with fatigue, decreased appetite. His sister also noticed pallor. The easy fatigability was progressive and was associated with palpitation on mild exertions because of which he had to take rest. His abdominal pain was mainly over epigastrium and left upper quadrant of abdomen and was dull aching in type associated with nauseating feeling and loss of appetite and occasional vomitings. The fever was intermittent not associated with chills and sweating, with maximum documented temperature being 102 degree F. The patient was febrile and pale and ill-looking young male with normal other vital signs. Per abdominal findings were suggestive of palpable nontender spleen around 2 cm below left subcostal margin along mid clavicular line, otherwise no other significant systemic findings were noted.

Initial laboratory results showed WBC: 2800 per cmm (neutrophils 20%, lymphocytes 79%, monocytes 0%, eosinophils 1%), Hb: 6.4 g/dL, Hct: 17%, platelets: 59000/μL, RBC: 1.91 million/μL, MCV: 91, Serum ferritin 382(13-400 ng/ml), Serum iron 12.4 mcg/dl (65-175 μg/dL), TIBC 279 mcg/dl (228-428 mcg/dl), iron saturation 44% (21-42%), Serum folate 3.9 ng/ml (5.2-20) and B12- 765 pg/ml (200-1100 pg/ml) and Reticulocyte of 0.8%.

Peripheral smear showed macrocytic, microcytic and hypochromic occasional fragmented RBCs, decreased WBC with relative lymhocytosis, no blast, and reduced platelets without atypical cells and Chest X-ray was normal.

Abstract

EBV infection can occur worldwide. We managed a 17 year old male who developed transient pancytopenia followed by thrombocytosis concurrent with EBV infection. To the best of our knowledge we have not encountered any cases of thrombocytosis concurrent with EBV infection before. Our case was diagnosed as a case of EBV infection with the help of serology and managed conservatively. In our case, though it is uncommon to find thrombocytosis due to EBV infection, it is possible that following pancytopenia concurrent with the EBV infection, the patient might have developed secondary or reactive thrombocytosis due to elevated endogenous levels of thrombopoietin, interleukin-6, catecholamines.

Keywords: Epstein-Barr Virus (EBV), infection, thrombocytosis.

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The fragmented RBCs in smear could be due to hemolytic anemia caused by EBV infection itself. The bone marrow biopsy and aspiration showed hypercellular marrow with erythroid hyperplasia with mixed erythroid maturation with increased megakaryopoiesis, this could be possible because the patient presented to us probably during recovery phase from acute infection. USG abdomen showed borderline splenomegaly, and anti-HCV, HBsAG and HIV, anti-HAV, HEV were nonreactive, Direct Coombs’ Test- negative, scrub typhus, brucella and leptospiro-antibodies negative, mantoux test negative, serum total bilirubin-1.5mg/dl with direct component-0.7mg/dl, SGPT- 36U/l, SGOT-38U/l, ALP-43U/l, serum LDH-1536U/l, total protein-6.4g/dl, albumin-4.5g/dl, calcium-8.6mg/dl, phosphorus-4.3mg/dl, stool for occult blood-negative, K39-negative, RA factor-negative, ANA-negative, malarial antigen negative and haptoglobin-0.5mg/dl (decreased). The high LDH, decreased haptoglobin could be parts of hemolysis concurrent with EBV infection.

The EBV(VCA) IgG=4.76 (positive if >1.1 index value) and IgM=1 (equivocal if 0.9-1.1, positive if more than 1.1, negative if less than 0.9 index value).

The patient was given blood transfusion for anemia following supportive treatment, after about 7 days of admission he developed thrombocytosis, with maximum platelets count reaching up to 10,5900/mm³ which fell on its own to normal range within a period of 1 week, during which he was carefully observed for possible complications. He didn’t develop any complications of thrombocytosis. After 2 weeks of in-hospital medical management his Hb-13.8 g/dl, total count-7700/mm³ with neutrophils-67%, lymphocytes-31%, and eosinophils-less than 0.9 index value.

Discussion:
Aplastic anemia after primary EBV infection or reactivation of EBV infection is enumerated very well in the literature, but thrombocytosis following EBV infection has not been documented. Our case is development of transient pancytopenia which later on was followed by thrombocytosis apparently following EBV infection. We think this case could be one of the rarest cases of EBV infection causing thrombocytosis.

The most common cause of thrombocytosis is a reactive process. The platelet count as such can not differentiate primary from reactive thrombocytosis. In one study with series of 732 patients with platelet counts 500,000 per cubic millimeter or higher, around 88% had secondary thrombocytosis, the most common causes were major surgery, infection, cancer and chronic inflammation.

Likewise, in another series of 280 hospitalized patients with platelet count of 1 million per cubic millimeter or higher, around 82% cases had reactive thrombocytosis.

Reactive thrombocytosis can be caused by increased endogenous levels of thrombopoietin, IL-6 and catecholamines. These mediators are produced because of inflammation, infection, and neoplastic conditions or situations causing stress. The levels of thrombopoietin are high in reactive thrombocytosis. Elevated plasma levels of IL-6 plays an important role in acute phase response of inflammatory and neoplastic processes. IL-6 increases the expression of thrombopoietin messenger RNA in the liver.

Thus, IL-6 may be a key mediator of the increased synthesis of thrombopoietin and the consequent reactive thrombocytosis.

Conclusion
In our case, it is possible that following pancytopenia concurrent with the EBV infection, the patient might have developed thrombocytosis but it’s difficult to say whether it’s clonal (primary) or reactive type but the scenario may favour reactive type which in turn may be speculated to be caused by elevated endogenous levels of thrombopoietin, interleukin-6, other cytokines, or catecholamines which are not measured in this case. This could well be a tug of war between the myelosuppressive effects of EBV itself and the increasing levels of endogenous mediators during recovery phase of the infection.

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