We tested the association between PON1 L55M, Q192R and I102V polymorphic variants and PC risk in Polish men. DNA from 110 consecutive, newly diagnosed patients hospitalized because of PC and DNA from 110 men – volunteers, healthy at the time of the study. PCR-RFLP method. In our study the average age at PC diagnosis of 55MM genotype carriers from families fulfilling Hereditary Prostate Criteria (HPC) was statistically significantly lower (by 7 years) than the average age at the disease diagnosis of 55MM carriers from families without HPC (54.6 ±6.7 vs. 61.9 ±5.4, respectively, p = 0.03). The probability of 5-year survival for the 55MM carriers was 81.3%, compared to 95.7% for non-carriers (p = 0.08, tendency). This is the first study in Polish men evaluating the impact of PON1 genetic polymorphisms on prostate cancer development and its clinical course. PON1 55MM variant may be probably associated with younger age at PC onset in men from families with HPC and with a shorter survival. However, more extensive studies on a larger number of PC patients, possibly from various populations, are necessary to confirm, and extend our findings.

Key words: prostate cancer; PON1 L55M, Q192R and I102V polymorphisms, predisposition, clinical characteristics.

Introduction

A cancer has become a major public health problem and global burden because of the high morbid-
ity and mortality rate. According to the American Cancer Society, in 2017, prostate cancer (PC) was the com-
monest diagnosed malignancy and the third lead-
ing cause of cancer mortality in men in the US [1].
There were 161 360 new diagnoses of prostate cancer and 26 730 deaths from the disease [1, 2]. Genetic
basis of prostate cancer is very complex. Thus, iden-
tification of PC high predisposition genes, as well as prevention or early diagnosis of the disease are ex-
tremely urgent. Some studies show that BRCA1, BRCA2, CHEK2, NBS1 or HOXB13 gene mutations which play an important role in cell cycle regulation, DNA repair and embryonic development of various organs are associated with higher risk of PC but none of these is a high risk gene for prostate cancer development. Literature analysis indicates that redox
signaling plays an important role in malignant tumor cell transformation through reactive oxygen species (ROS) mediated function. Dysregulation of this redox balance is implicated in the development of various diseases including cancer [3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15].

An antioxidant paraoxonase 1 (PON1) is an important endogenous free radical scavenger which plays a role in preventing the effects of systemic oxidative stress which is defined as a redox imbalance between prooxidant and antioxidant systems with either an overproduction of oxidants or a deficiency of antioxidant agents [16]. Under oxidative stress, excess ROS can cause damage to many cellular and extracellular constituents, including DNA, proteins and lipids [17]. The physiological role of PON1 protein is not exactly known, however increasing evidence indicates that it can assure protection against oxidative damage by hydrolysing lipid hydroperoxides and by protecting the low density lipoproteins from oxidative modifications [18].

The human PON1 protein is coded by PON1 gene located on the long arm of chromosome 7 (7q21.3). Its most common polymorphisms localize in the coding region: a leucine (L) to methionine (M) substitution at position 55 (L55M, p.Leu55Met, c.163T>A) and a glutamine (Q) to arginine (R) substitution at position 192 (Q192R, p.Gln192Arg, c.575A>G). The L55M [rs854560] and Q192R [rs662] polymorphisms affect paraoxonase 1 (PON1) serum concentration and activity of PON1 gene [19, 20, 21]. It was suggested that PON1 55M allele could correlate with higher PON1 activity than PON1 55L allele. In another study, it was observed that the differences in the PON1 serum levels linked to the L55M polymorphism were due to the reduced stability of the PON1 55M protein [22]. The hydrolyzing activity of the protein encoded by the 192RR, second the most common genotype of PON1 gene, is 8-fold higher than that encoded by the 192QQ. This polymorphism also affects the serum protein concentration. Homozygous 192RR individuals have higher enzyme concentrations than the homozygous 192QQ individuals [15, 23, 24]. Additionally, Marchesani et al. detected new polymorphic variant in exon 4, an A→G substitution at nucleotide position 304 resulting in an isoleucine → valine substitution at codon 102 of the PON1 gene. The authors indicated that PON1 1102V is associated with decreased serum paraoxonase stability [25].

Variations due to the presence of single nucleotide polymorphisms in antioxidant enzyme genes may contribute to the interindividual differences of transcript levels and enzyme activities, which might play a significant role in the prostate cancer development [14, 22]. Therefore, the aim of the present study was to investigate the relation between PON1 L55M, Q192R and 1102V polymorphisms and prostate cancer risk in Polish men. Additionally, we measured the impact of these variants and their constructed haplotypes on clinical course of the disease, including overall survival time.

**Material and methods**

The material of the investigations were archival DNA samples stored in the Department of Clinical Genetics CM UMK in Bydgoszcz from 2005 to 2007. DNA was isolated from EDTA anticoagulated peripheral blood of 110 consecutive, newly diagnosed prostate cancer (PC) patients, from all over Poland, regardless of age at PC onset, family history and histological type of cancer. They were hospitalized because of PC at Department of Urology of the J. Bziel University Hospital in Bydgoszcz. The age at PC diagnosis ranged from 45 to 75 years (the mean age 59.9 ± 5.9). Family history was analyzed either by the construction of a family pedigree or the completion of a standardized questionnaire by patients. All cases of first- and second-degree relatives diagnosed with prostate cancer and their age at disease diagnosis were recorded. The estimation of patient families as those with a history suggesting hereditary risk of prostate cancer (HPC) was performed on the basis of criteria defined by Carter et al. [26] and Cybulski et al. [27]. Among 110 prostate cancer patients, 25 (22.7%) originated from families suspected of Hereditary Prostate Cancer (Fig. 1).

Information about the PSA level before the surgical operation was available for 97 patients, about grade of prostate cancer for 101 patients, and about tumor stage for 99 patients. PC cases were divided into two groups based on the incidence of PC in the family history: sporadic (HPC = 0) and hereditary prostate cancer (HPC ≥ 1). The Spearman correlation coefficient was used to assess the relationships between PON1 polymorphisms and clinicopathological parameters. The significance level was set at p < 0.05.
stage for 102 patients. Data on survival were available for 103 PC patients.

In 61 of 110 (55.45%) families, an aggregation of cancers of the breast, stomach, colon, ovary, lung, larynx, bladder and kidney, as well as melanoma was present, in addition to prostate cancer.

Control group consisted of DNA samples isolated from peripheral blood of 110 men, volunteers, healthy at the time of the investigations, i.e. (without prostate cancer on the basis of PSA concentration and/or digital rectal examination – DRE). The medical examinations were performed in them as a part of PC prophylaxis. Control molecular test was offered to all men with correct result of DRE and with a normal age-specific value of PSA. The age of men from control group ranged from 46 to 74 years (the mean age was 59.9 ±6.6) and matched to the PC group.

The study protocol was approved by the Ethics Committee of the Collegium Medicum Nicolaus Copernicus University in Bydgoszcz, Poland. Every PC patient and a men from control group gave their written informed consent for the use of their DNA sample for genetic testing.

Polymorphisms analysis

Genomic DNA was isolated from peripheral blood leukocytes by the standard salting-out method. The PCR-RFLP was used for determining the L55M and Q192R polymorphisms, and Sanger sequencing method for determining the I102V. The primers for amplification of coding regions including L55M, Q192R and I102V were designed using Primer3 software (http://frodo.wi.mit.edu/).

The PCR amplification was done for the fragments of PON1 gene with following pairs of primers: Forward (F): 5' – GAAGAGTGATGTATAGCCCCA G – 3' together with Reverse (R): 5' – TTTAATCCAGAGCTAATGAAAGCC – 3' for determination of the L55M polymorphism and F: 5' – TATTGTGTGCTGTGGGACCTGA G – 3' together with R: 5' – CACGCTAAACCGAAAATACATCTC – 3' for determination of the Q192R polymorphism. The conditions and components of PCR reaction are available on request. The restriction enzyme N1aIII (Biolabs) was used for determination of the L55M. Digestion with N1aIII generated the following fragments: for 55M allele fragments of 127 bp and 42 bp, for 55L allele a single fragment of 169 bp. The Alw1 (Biolabs) restriction enzyme was used to identify the possible Q192R genotypes. The Alw1 digestion generated the following fragments: for 192R allele fragments of 63 bp and 36 bp and for 192Q allele – a single fragment of 99 bp. The digested PCR products of the two PON1 polymorphisms were separated on 3% agarose gel and visualized using Midori Green DNA staining.

The I102V polymorphism was genotyped using Sanger sequencing. The primers for amplification of PON1 coding region were: 5’ – CTTCGATGGGGCAAGGATA – 3’ (F) and 5’ – CCTCACCAGCTCCAACTCTC – 3’ (R). The amplified DNA amplion was sequenced using BigDye terminator v3.1 Cycle Sequencing Kit (Life Technologies), according to manufacturer protocol. Sequencing products were analyzed on the ABI prism 3130 Genetic Analyzer (Applied Biosystems, Life Technologies). All sequenc- es were compared with the PON1 RefSeq sequence (NG_008779) for I102V polymorphism detection. Sequencing analysis of the coding region of PON1 was performed in all men from study and control groups.

Statistical analysis

The polymorphisms frequencies in prostate cancer patients and controls were compared. To evaluate the effects of three Single Nucleotide Polymorphisms (SNPs) on prostate cancer risk, haplotypes were constructed. Haplotypes frequencies were estimated using statistical software. Odds ratios (ORs) with 95% confidence intervals (CIs) were used to assess the strength of the associations. ORs were generated from two-by-two tables and statistical significance was assessed with the Fisher’s exact test or the chi-square test when appropriate. Mean age at PC diagnosis was compared between polymorphism carriers and non-carriers using Student’s t-test. For the survival analysis, the patients were followed from the date of biopsy (confirmation of prostate cancer) until death, if applicable or the end of observations. In living patients five year survival was analyzed. The vital status and the date of death were requested from the Polish Ministry of Health. The Kaplan-Meier curves for the probability of survival of PON1 polymorphic variant carriers and non-carriers have been estimated.

Results

The comparison of the PON1 L55M and Q192R genotypes and haplotypes frequencies between prostate cancer patients and healthy controls were shown in Table I and Table II. The comparison of the both groups of men did not show any statistically significant differences or trends in genotypes and haplotypes distribution. However, the 55MM homozygous genotype was detected in 14.5% of PC patients vs 17.3% of controls (p = 0.58, NS) and the 192RR homozygous genotype was detected in 5.5% of PC patients vs. 4.6% of controls (p = 0.77, NS). The PON1 55LM+192QQ haplotype was the most frequently found in both prostate cancer and control groups (26.6% and 26.9%, respectively). The PON1 55MM+192RR haplotype was detected in no man. A PON1 102II wild type homozygous genotype was
detected in all PC patients and in all men from control group, therefore, in further analyzes it was omitted.

We observed almost 2-fold higher 55MM genotype frequency in PC patients from families with HPC compared to its frequency in patients from families without HPC (20.0% vs. 12.9%). The difference was not statistically significant (p = 0.38). However, the average age at PC diagnosis of 55MM genotype carriers from families fulfilling HPC criteria was statistically significantly lower (by 7 years) than the average age at the disease diagnosis of 55MM carriers from families without HPC (54.6 ± 6.7 vs. 61.9 ± 5.4, respectively, p = 0.03).

The age at PC diagnosis of the 192RR carrier (66 years) from a family with sporadic prostate cancer was statistically significantly older than the average age at PC diagnosis of 192RR carriers (56.3 ± 1.7) from families with one PC case and other cancers aggregation. The difference was statistically significant (p = 0.01), however, it was calculated on the basis of the age at PC diagnosis of only one man with 192 RR.

Data on survival were available for 103 investigated PC men. There were 3 deaths recorded among sixteen 55MM homozygous genotype carriers and six deaths among 87 non-carriers. The probability of 5-year survival for the 55MM carriers was 81.3%, compared to 95.7% for non-carriers (p = 0.08, tenancy). Figure 2 shows the Kaplan-Meier curves for these PON1 genotype carriers.

The probability of 5-year survival for the 192RR carriers was 100%, compared to 93.2% for carriers of other genotypes (p = 0.44, NS). The Kaplan-Meier curves for PON1 Q192R prostate cancer patients are presented on Fig. 3.

In the present study, both L55M and Q192R polymorphisms genotype frequencies were not related to
the patients age. We did not observed a statistically significant difference between the average age at PC onset of 55MM genotype carriers compared to the average age at PC onset of 55LM and 55LL genotypes carriers (59.6 ± 6.6 vs. 60.0 ± 5.8, respectively, p = 0.84). Similar results were obtain for the analysis of the average age at PC onset of 192RR genotype carriers compared to 192QR and 192QQ genotypes carriers (59.7 ± 5.5 vs. 59.9 ± 5.6, respectively, p = 0.92). The 192RR genotype was detected in 4 (3.7%) PC men and in one (0.9%) man from the control group between 51 and 60 years of age (OR = 3.78, p = 0.24, NS).

In 18.8% of PC men, 55MM carriers, prostate cancer cells were characterized by a low degree of differentiation (SG ≥ 8), compared to 10.3% of men with 55LL genotype and 11.3% of men with 55LM genotype. Moreover, the cancers showed a high level of progression at the time of diagnosis (T3/4) in 31.3% of men with 55MM (in 10% of men with 55LL and in 15.1% of men with 55LM). 15.4% of men with 55MM had preoperative PSA > 20 ng/ml compared to 7.9% of men with 55LL and 5.7% of men with 55LM. However, all above differences were no statistically significant. No statistically significant differences in the presence of PSA concentration > 20 ng/ml, high grade Gleason Score (GS ≥ 8) or advanced TNM stage (T3/4), depending on Q192R carrier status, were also observed.

Discussion

This is the first study in Polish men evaluating the impact of PON1 genetic polymorphisms on prostate cancer development and its clinical course. The oxidative stress has a strong impact on cancer formation and development. However, the human body has several protective systems with an ability to remove harmful free radicals. In the present study we did not find any statistically significant relationship of the L55M and Q192R genotypes and their constructed haplotypes with PC, however, some observations are worth noting. In contrast to our results, Antognelli et al. found a relationship between PON1 55LM-MM and 192QR polymorphisms and PC risk in the Italian population [28]. Fang et al. performed a meta-analysis based on 25 studies and found association between 55LM heterozygous genotype compared to 55LL wild type genotype with increased prostate cancer risk [29]. Zhang et al. analyzed 8 112 cases and 10 037 controls from 32 published case-control studies to assess the connection between PON1 192R allele with many types of cancers including prostate cancer. They showed that 192QR compared to 192QQ and 192RR+QR compared to 192QQ were associated with an increased PC risk (OR = 1.782, p = 0.000 and OR = 1.281, p = 0.056, respectively) [30]. In another meta-analysis, based on 21 studies, Chen et al. indicated that PON1 L55M was a risk factor for prostate cancer in the heterozygote model (55LM vs. 55LL: OR=1.304, p=0.067) [31].

In the present study, no relationship was observed for I102V with prostate cancer. The 102II homozygous genotype was detected in all PC patients and 192QR or 192QQ carriers compared to patients with 192RR genotype, the green line the survival for 55LM or 55LL carriers and the red line the survival for 55MM carriers and the green line the survival for 192QR or 192QQ carriers.
Additionally, these authors indicated that L55M or Q192R were not associated with PC [25]. The size of prostate cancer increased risk associated with the I102V may be different in Polish and Finnish men. Comparing to Polish ethnically healthy homogenous population, Finns have a relatively small founder population and many immigrants from other countries, compared to Polish population.

In the present study, the 55M frequency was higher in men from HPC families than in men from families without HPC (20% vs. 12.9%, NS). The average age at PC diagnosis was statistically significantly younger (by 7 years) in 55MM carriers from HPC families than those from families without HPC. Thus, the carrying of 55M may be associated with hereditary prostate cancer and a younger age at PC onset, however, in order to make unambiguous conclusions, more studies are needed, on larger groups of patients. The 192 RR frequency was similar in men from HPC families and those from families without HPC (4% vs. 6%). This indicates that there is no relationship between PON1 192 RR genotype and hereditary form of the disease.

The Kaplan-Meier curves analysis showed that overall survival time was shorter for 55MM carriers compared to other genotype carriers with PC. About 14.4% less patients with 55MM than 55LM and 55LL survived 5 years from the time of PC diagnosis.

It seems also that carrying of 192RR is a good prognostic factor in relation to the overall survival time of PC men. All 192 RR carriers survived at least 5 years from the moment of PC diagnosis (and even over 7.5 years).

The 192RR genotype probably may be associated with the age lower than 60 years at PC onset. It is worth noting that 192RR was detected in 6.9% of PC men compared to 1.9% of healthy men < 60 years of age, however, this difference was not statistically significant (p = 0.24).

We suggest that the presence of 55M homozygous genotype may be probably associated with an aggressive form of prostate cancer. The similar observations were made by Stevens et al. in the US population. They observed the 2.18-fold higher risk of aggressive PC (T3/4; GS ≥ 8) in 192QR/55LM heterozygous ha haplotype carriers, compared to the population risk (OR = 2.18; p ≤ 0.05). Authors suggested that men with PON1 L55M and Q192R genotypes that reduce paraoxonase 1 – mediated ROS detoxification capacity may have increased risk of aggressive prostate cancer [8].

Concluding, we suggest that PON1 55MM variant may be probably associated with younger age at PC onset in men from families with HPC and with a shorter survival. However, more extensive studies on a larger number of PC patients, possibly from various populations, are necessary to confirm, and extend our findings. Performing similar investigations on a larger group of PC patients, also in different populations, will be promising. PON1 polymorphisms usefulness as a biomarkers for screening, diagnosis and measurement of treatment efficacy with respect to prognosis and survival in prostate cancer patients should be determined.

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The authors declare no conflict of interest.

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