Sorption and desorption of Cr(VI) ions from water by biochars in different environmental conditions

Aleksandra Tytłak · Patryk Oleszczuk · Ryszard Dobrowolski

Abstract In the present research, the potential of two biochars produced by the thermal decomposition of wheat straw (BCS) and wicker (BCW) for Cr(VI) ions removing from wastewater was investigated. The pH and the presence of chlorides and nitrates were also investigated. The Freundlich and Langmuir models were applied for the characterization of adsorption isotherms. The Langmuir model has better fitting of adsorption isotherms than the Freundlich model. The sorption process can be described by the pseudo second-order equation. The optimal adsorption capacities were obtained at pH 2 and were 24.6 and 23.6 mg/g for BCS and BCW, respectively. X-ray photoelectron spectroscopy (XPS) studies confirmed that Cr(III) ions were the most abundant chromium species on the biochars’ surface. The results indicated that the sorption mechanism of Cr(VI) on biochar involves anionic and cationic adsorption combined with Cr(VI) species reduction.

Keywords Sorption · Chromium · Biochar · Environmental conditions · XPS · SEM-EDS

Introduction

Chromium occurs in the environment in natural forms as a ferric chromite (FeCr₂O₄), crocoite (PbCrO₄), and chrome ochre (Cr₂O₃) (Mohan and Pittman 2006). Moreover, it is delivered into the environment from anthropogenic sources which are caused by the wide exploitation of chromium in the industry. This element and its compounds have applications in electroplating, metal finishing, petroleum refining, magnetic tapes, pigments, leather tanning, wood protection, chemical manufacturing, brass, electrical and electronic equipment, nuclear power plants, and catalysis (Kuo and Bembenek 2008; Gottipati and Mishra 2010). Chromium contaminants released by these industries are input into the soil, surface water, drinking water, and groundwater, and afterwards pass through the cell membrane of the living organism and to the food chain to the human body.

The most common species of chromium present in the environment are hexavalent chromium—Cr(VI) and trivalent chromium—Cr(III). Different states of valence affect their contrary properties such as toxicity, bioavailability, and mobility (Kotaś and Stasicka 2000). Cr(VI) is toxic, mutagenic, carcinogenic, teratogenic on biological systems, soluble, and mobile; moreover, it is thermodynamically metastable in the soil and exists in anionic and neutral species as a chromates CrO₄²⁻ and Cr₂O₇²⁻ and bichromates HCrO₄⁻, H₂CrO₄, and HCr₂O₇²⁻ (Choppala et al. 2012). Cr₂O₇²⁻ forms predominantly in a low pH and high chromium concentration conditions, whereas CrO₄²⁻ are the most abundant forms at pH higher than 6.5 (Mohan and Pittman 2006). Cr(III) is nontoxic and ranked as a micronutrient because of its being an essential component of the human diet and having influence on fat, glucose (GFT, glucose tolerance factor), and cholesterol metabolism of mammals (Kimbrough et al. 1999). Cr(III) forms a stable inorganic ion as [Cr(H₂O)₅]³⁺ and hexacoordinate complexes (Dhal et al. 2013).

Considering the different properties of chromium species and their impact on the living organisms, especially with regards to their identification, the determination and effective reduction of Cr(VI) to Cr(III) have become recently the main...
subject of many studies (Kimbrough et al. 1999; Mohan and Pittman 2006; Dobrowolski and Otto 2010).

The concentration of the Cr(VI) input to the environment from industrial applications must be reduced to below 0.05 mg/L before discharging into the surface water, as is established by the US EPA (Baral and Engelken 2002). To reduce Cr(VI) concentration in water, different techniques usually are used, e.g., chemical precipitation, ion exchange, membrane separation, ultrafiltration, sedimentation, and adsorption. These techniques are not free from disadvantages, e.g., high cost, low efficiency, low selectivity, high-energy requirements, and secondary toxic waste generation. Adsorption of contaminants onto activated carbon (AC) because of its high efficiency and simplicity of design is still one of the most frequently used methods (Mohan and Pittman 2006). The highly developed porosity, extended surface area, microporous structure, high adsorption capacity, and high degree of surface reactivity make the AC a good sorbent for wastewater treatment. However, there is a significant limitation to the application of these materials: commercial ACs are expensive. Because of their high price, the necessity in the preparation of low-cost carbon materials has become an important issue for many researchers (Deveci and Kar 2013).

An interesting replacement of AC can be the biochar (BC). BC is a special type of charcoal produced by the thermal decomposition of biomass under relatively low temperature (<700 °C) and limited oxygen conditions (Oleszczuk et al. 2013). Due to the specific surface area and high sorption capacity in regard to heavy metals ions, the biochar could be used as an effective adsorbent of these contaminants (Beesley et al. 2011; Kolodyńska et al. 2012). The presence of many functional groups on the biochar surface, e.g., carboxylic, alcohol, and hydroxyl, makes an opportunity to form complexes between these groups and heavy metal ions (Tang et al. 2013). Despite of the fact that BCs have become the subject of many studies recently, there is still not enough comprehensive information about their potential applications in the removal of Cr(VI) from water. Because of the growing chromium pollution, there is a necessity to establish a method which can use satisfactory biochar properties for the selective chromium removal.

The aim of this study was the evaluation of Cr(VI) adsorption onto two different biochars as a tool of reduction and immobilization of its toxic forms from aqueous media. The basic parameters having an influence on adsorption capacity were determined. The adsorption capacities were studied based on the initial runs of the adsorption isotherms. Taking into account further practical applications of biochars in water treatment, the influence of chlorides and nitrates on adsorption ability was also investigated.

Materials and methods

Biochar characterization

Biochars were produced by the thermal decomposition of biomass at a temperature range from 350 °C (start of combustion) to 650 °C (temperature of maximal combustion) in limited oxygen conditions (1–2 %). BCs were produced from a wicker (BCW) and a wheat straw (BCS) and provided by Fluid SA (Sędziszów, Poland) and Mostostal Sp. z. o. o. (Wroclaw, Poland), respectively.

The properties of biochars were studied by the standard methods, and the results are presented in Table 1. The pH values were obtained potentiometrically in 1 mol/L potassium chloride after 24 h in the liquid/soil ratio of 10. The cation exchange capacity (CEC) and available forms of phosphorous, potassium, and magnesium were determined according to procedures for soil analysis recommended by van Reeuwijk (1992). The total organic carbon content (TOC) was measured using TOC-VCSH (Shimadzu) with solid sample module (SSM-5000). The total nitrogen (Nt) was determined by the Kjeldahl’s procedure without the application of Dewarda’s alloy (Cu-Al-Zn alloy reducer of nitrates and nitrates).

The SEM studies were performed on Tesla BS-301 microscope Quanta 3D FEG operating at 15.0 keV. The FTIR/PAS spectrum of the biochars was recorded by means of the Bio-Rad Excalibur 3000 MX spectrometer equipped with photoacoustic detector MTEC 300 with infrared detection of resulting CO2, H2, and NO2, using TOC-VCSH (Shimadzu) with solid sample module (SSM-5000). The total nitrogen (Nt) was determined by the Kjeldahl’s procedure without the application of Dewarda’s alloy (Cu-Al-Zn alloy reducer of nitrates and nitrates). For analyzing the structure of used BCs, low-temperature (77.4 K) nitrogen adsorption-desorption isotherms were obtained using Micromeritics ASAP 2405N adsorption analyzer. On the basis of the standard BET method, the specific surface areas $S_{BET}$ were calculated. The CHN elemental analyzer (Carlo-Erba NA-1500) via high-temperature catalyzed combustion, followed by infrared detection of resulting CO2, H2, and NO2, was used for determination of carbon, hydrogen, and nitrogen content in BCs.

Batch sorption experiment

The standard stock solution of Cr(VI) (520 mg/L) was prepared by proper dissolution of $K_2Cr_2O_7$ powder (POCH, Gliwice, Poland) in redistilled water. The calibration curve was established using the standard solutions of Cr(III) prepared in 2 mol/L HNO3 by dilution from 1,000 mg/L stock solution (Merck, Darmstadt, Germany). Moreover, Suprapur nitric acid (65 %) (POCH, Gliwice, Poland), Suprapur hydrochloric acid (36 %) (POCH, Gliwice, Poland), sodium...
chloride (POCH, Gliwice, Poland), and potassium nitrate
(POCH, Gliwice, Poland) were used. Sodium hydroxide and
hydrochloric acid solutions were used for pH adjustment.

The effect of time on the adsorption of Cr (VI) ions onto
BCS and BCW vs. time intervals up to 28 h at the initial
concentration of Cr(VI) at the level of 100 mg/L and the solid
materials’ mass around 0.2±0.03 g at pH of 2 were investi-
gated. The adsorption capacities of Cr(VI) onto the biochars
were determined by studying the initial runs of adsorption
isotherms. The equilibrium isotherms were obtained at initial
pH of 2, Cr(VI) concentrations from 1 to 600 mg/L range,
equilibrium time of 24 h, and temperature of 25 °C. The
effect of pH on Cr(VI) adsorption onto the tested biochars was
carried out using series of solutions, keeping the
basic spectrometer parameters, were 2.9 and 13.5 L/min,
acetylene and air flow, 357.9 nm and slit width of 0.2 nm were selected for the
measurement. Moreover, the values of lamp current of 7 mA. The wavelength of
357.9 nm and slit width of 0.2 nm were selected for the
measurement. Moreover, the values of acetylene and air flow,
the basic spectrometer parameters, were 2.9 and 13.5 L/min,
respectively. The limit of Cr detection was at the level of
0.14 mg/L.

The optimization of Cr(VI) ions adsorption onto BCs was
conducted at 25 °C. Particular measuring points were acquired
for the adsorption system as follows: 50 mL of chromium(VI)
solution and 0.2 g of biochar. The equilibrium adsorption
value in the solid material a (mg/g) was calculated from the
following equation:
\[ a = \frac{(c_1-c) \cdot V}{m} \]  \hspace{1cm} (1)
where \( c_1 \) is the initial concentration of Cr(VI) (mg/L), \( c \) is the
equilibrium concentration of Cr(VI) (mg/L), \( V \) is the volume of the solution (L), and \( m \) is the mass of the solid material (g).

Data analysis

Pseudo first-order equation and pseudo second-order equation
were used for recognizing the reaction order of obtained
Cr(VI) sorption kinetics onto the examined biochars. Kinetic
data were fitted to the following equations:
\[ \ln (a_{eq}-a_t) = \ln a_{eq} - k_1t \] \hspace{1cm} (2)
\[ \frac{t}{a_t} = \frac{1}{k_2 a_{eq}^2} + \frac{1}{a_{eq}} \cdot \frac{t}{a_{eq}} \] \hspace{1cm} (3)
where \( a_t \) is the amount adsorbed (mg/g) at time \( t \), \( a_{eq} \) is the
amount adsorbed (mg/g) at equilibrium time, and \( k_1 \) (1/min) and \( k_2 \) (g/mg min) are the rate constants of the pseudo first-
order equation and pseudo second-order equation, respectively.

Two nonlinear isotherm models (Freundlich and Langmuir)
were tested to fit the experimental data: the Langmuir model
\[ a = a_m \cdot \frac{(Kc_{eq})^n}{1 + (Kc_{eq})^n} \] \hspace{1cm} (4)
and Freundlich model, in which the linear form can be written
as follows:
\[ \ln a = \ln(a_mK) + n \ln c \] \hspace{1cm} (5)
where \( a \) is the Cr(VI) ions’ adsorbed amount (mg/g) onto BCs
at the equilibrium of Cr(VI) chromium concentration \( c \) (mg/L),

Table 1: Physicochemical properties of BCS and BCW biochars

| Biochar | pH | CEC | Available forms of | Elemental composition | Ash | SBET | PV |
|---------|----|-----|-------------------|----------------------|-----|------|----|
|         |    |     |  P2O5 K2O Mg       | C H N O               |     |      |    |
| BCS     | 9.9| 530 | 540 2,824 163      | 54 1.8 0.9 2.3       | 41  | 26.3 | 0.026 |
| BCW     | 9.1| 143 | 122 772 32         | 70 3.2 0.8 17.9      | 8.6 | 11.4 | 0.0061 |

pH in KCl, CEC the cation exchange capacity (mmol/kg), P2O5, K2O, Mg available forms of phosphorous, potassium, and magnesium (mg/100 g), CHNO the contribution (%) of carbon, hydrogen, nitrogen, and oxygen, Ash ash content (%), H/C ratio of hydrogen to carbon, (O+N)/C polarity index, O/C ratio of oxygen to carbon,SBET specific surface area (m2/g), PV pore volume (cm3/g)
$a_m$ is the maximum adsorbed amount needed to form a monolayer on adsorbent surface (sorption capacity), $K$ is the Langmuir constant, and $n$ ($0 \leq n \leq 1$) characterizes the quasi-Gaussian energetic heterogeneity of the adsorption system.

**Results and discussion**

**Biochar properties**

The CEC values for BCS and BCW are distinctly different and equal to 530 and 143 mmol/kg, respectively (Table 1). For BCS, $K^+$ ions dominated in the sorptive complex, next to $\text{Ca}^{2+}$, $\text{Mg}^{2+}$, and $\text{Na}^+$. BCW ions demonstrate a different order. The $\text{Ca}^{2+}$ ions were predominant, followed by $\text{K}^+$, $\text{Na}^+$, and $\text{Mg}^{2+}$. Considering the availability of the selected forms of elements, the highest concentration was determined for potassium, which is lower for phosphorous and magnesium, without regard to the biochars’ type. BCW has higher $\text{H}/\text{C}$, $(\text{O} + \text{N})/\text{C}$, and $\text{O}/\text{C}$ ratios than BCS. The lowest $\text{H}/\text{C}$ ratio for BCS was associated with the aromatic structure, with BCS having the higher carbonization level than for BCW. Moreover, comparing the $\text{O}/\text{C}$ ratio values, it can be claimed that BCW material presents more hydrophilic structures; in addition, the highest $(\text{O} + \text{N})/\text{C}$ ratio, named the polarity index, indicates the higher polarity for this material. Furthermore, the BCs studied have different values of the specific surface area, 26.3 and 11.4 m$^2$/g for BCS and BCW, respectively. This parameter is correlated with the pore area value, which is also significantly higher in the case of BCS.

The IR spectra of examined biochars (Fig. 1) are complicated because of the presence of multiple bands in a wide range of wavenumbers, which is also in correlation with the amount of functional groups on the surface of the biochar. At about 3,650 and 3,202 cm$^{-1}$ in BCS, the spectra have centered band characteristics for stretching vibration of hydrogen-bonded hydroxyl groups of alcohols and carboxylic acids, respectively. In the same spectrum, the band being in correlation with hydroxyl groups of water at about 3,433 cm$^{-1}$ can be seen. At about 3,354 and 3,358 cm$^{-1}$ in the both spectra, the bands attributed to the primary amine groups’ presence on the biochars’ surface can be seen. The vibration at 3,068 cm$^{-1}$ in BCW spectrum is associated with vinyl and acrylic C-H groups. The band at about 3,034 cm$^{-1}$ is attributed to the $\text{=C-H}$ stretching of aromatic compounds. The aliphatic C-H bands at the region 2,867–2,932 cm$^{-1}$ can be found in the BCW spectrum, but there are no adsorption bands of those types in BCS spectrum. The bands at 1,751 and 1,685 cm$^{-1}$ in the BCS spectrum and the similar band at 1,701 cm$^{-1}$ in the second biochar spectra are the adsorption bands of C=O. The vibration at 1,595 cm$^{-1}$ is associated with C=O in ketones, carboxylates, and quinones or with the C=C stretching in the aromatic components [30]. The bands in the range 1,400–1,050 cm$^{-1}$ could be correlated with C-O and O-H presence in carboxylic acids (band at 1,356 cm$^{-1}$) and phenols or C-O-C in ethers. The bands in the region of 1,200–1,000 cm$^{-1}$ are attributed to the C-H deformation in the aromatic structures. The bands centered at 1,055, 957, and 783 cm$^{-1}$ can be responsible for Si-O vibrations of inorganic materials in BCS [31]. The bands at 874 and 873 cm$^{-1}$ in BCS and BCW spectra, respectively, can be associated with C-H bending in aromatic compounds.

**Sorption kinetics**

Cr(VI) demonstrated relatively fast sorption kinetics on tested biochars (Fig. 2). The equilibrium adsorption of Cr(VI) onto BCW was reached quicker than onto BCS. The equilibrium
time was achieved after 21.5 h for BCS and 18 h for BCW. It is thought that the adsorption of Cr(VI) onto BCs proceeds with surface reduction of chromium(VI) to chromium(III) in the form of aqueous complex [Cr(H₂O)₅]³⁺ and adsorption of this complex on the biochar surface (Dobrowolski and Otto 2010). The experimental data suggest that 18 h (BCW) and 21.5 h (BCS) will be sufficient to reach the equilibrium state of Cr by tested biochars under other adsorption conditions determined as an optimum in this research. The differences between times of reaching the equilibrium for BCs may be caused by the presence of the different functional groups, acidic and basic, on the surface of the studied BCs. There are more varied functional groups on the BCS surface than BCW (Fig. 1) which are equal with the higher quantity of adsorption centers which could bond the chromium ions. Moreover, it can be the effect of different values of \( S_{\text{BET}} \). BCS has higher specific surface area than BCW, 26.3 and 11.4 m²/g, respectively, and the equilibrium state is reached longer in the case of this biochar. In comparison to the previous studies, the obtained data are in agreement with most studies concerning the Cr(VI) ions adsorption (Aliabadi et al. 2012; Zhang et al. 2013). The \( a_{\text{eq}} \) values for the studied BCs were 14.36 mg/g (BCS) and 10.28 mg/g (BCW). Moreover, the sorption capacities calculated from pseudo second-order equation were consistent with the experimental value and were 14.78 mg/g (BCS) and 10.31 mg/g (BCW).

Sorption isotherms

The maximum sorption capacities obtained for both biochars were similar and equal to 24.6 mg/g for BCS and 23.6 mg/g for BCW. The linear dependencies obtained are plotted for the each isotherm model and are shown in the Fig. 4. In Table 3, the corresponding constants and correlation coefficients are reported. For both examined BCs, Langmuir model gave a slightly better fit and provided the best correlation to the isotherms data. The correlation coefficients were 0.990 and 0.978 for BCS and BCW, respectively. However, the studied Cr(VI) adsorption isotherms are in good agreement.
with the linear form of the Freundlich equation for higher concentration of chromium(VI) (Fig. 4). The decreasing of \( n \) value to zero causes the increasing adsorption heterogeneity. According to the data presented in Table 3, the \( n \) value obtained for BCS material is smaller than for BCW material, which is associated with greater energetic heterogeneity of BCS and the smaller value for BCW biochar.

The greater adsorption capacity for BCS was partially correlated with higher CEC value. This effect can be also explained by the pore volume of investigated biochars. Cr(VI) ions are adsorbed on the BCS’ surface in form of \([\text{Cr(H}_2\text{O)}_6]^{3+}\). The pore volume has significant influence on adsorption process of these cations because of their size. On the basis of Table 1, BCS has higher PV, which can be one of the few explanations of the higher adsorption value of this material. The transport of chromium ions through BCW pores was probably more difficult.

The O/C index should be also considered as a possible reason of higher adsorption capacity of BCS. BCS is more hydrophobic than BCW, which can make this property responsible for stronger electrostatic interaction between \([\text{Cr(H}_2\text{O)}_6]^{3+}\) and the BCS surface.

**Table 2** Parameters of pseudo first-order and pseudo second-order kinetic models for Cr(VI) adsorption onto BCS and BCW biochars

| Biochar | Kinetic model         | Pseudo first order | Pseudo second order |
|---------|-----------------------|--------------------|---------------------|
|         | \( k_1 \) \( R^2 \) \( a_{eq} \) | \( k_2 \) \( R^2 \) \( a_{eq} \) |
| BCS     | 0.0052 0.749 –        | 0.00059 0.983 14.78 |
| BCW     | 0.0051 0.778 –        | 0.00039 0.992 10.31 |

\( a \), the amount adsorbed (mg/g) at time \( t \), \( a_{eq} \), the amount adsorbed (mg/g) at equilibrium time, \( k_1 \) (1/min) and \( k_2 \) (g/mg min) the rate constants of the pseudo first-order equation and pseudo second-order equation, respectively, \( R \) regression coefficient

**Effect of pH**

It has been proved that pH is one of the most important parameters having an influence on the adsorption capacity of adsorbent for heavy metal ions removal from the aqueous solutions (Kołodyńska et al. 2012; Aliabadi et al. 2012; Deveci and Kar 2013). In Fig. 5, the adsorption abilities of chromium in an equilibrium pH are shown depending on the biochar tested. The variations in chromium(VI) sorption ability in the examined pH range may be partly related to pH dependency of the chromium species present in the aqueous solution and onto the BCS’ surface. At pH 2, Cr(III) ions exist in aqueous solution as \([\text{Cr(H}_2\text{O)}_6]^{3+}\) which has associated water molecules, whereas Cr(VI) is unstable. These molecules can be exchanged with the hydroxyl ions which depend on the pH values. Changes in acidic groups present on the surface of the biochar cause the variations of pH. The maximum adsorption of Cr(VI) ions onto BCS was achieved at pH 2. For both of BCS in the pH range from 1 to 7, the same trend was observed, and the differences between biochars were only noticeable above pH 7. It is caused by the uncompleted reduction of Cr(VI) and mixed Cr(III) and Cr(VI) adsorption processes. At pH lower than 2, in acidic conditions, the reduction of Cr(VI) to Cr(III), followed by the adsorption by nonspecific sorption or surface chelation, is the main mechanism of Cr(VI) adsorption. At pH 2.0–3.2, Arslan and

![Fig. 3](image1.png)

*Fig. 3* The kinetic fitting plots 1 pseudo first-order equation and 2 pseudo second-order for the BCS and BCW

![Fig. 4](image2.png)

*Fig. 4* Langmuir (1) and Freundlich (2) isotherms for chromium(VI) adsorption onto BCS and BCW
Pehlivan (2007) noticed that carboxylic acid centers could be appreciably deprotonated and exist in the form of COO$^-$ which can bind to Cr(III) ions. Furthermore, Gardea-Torresdey et al. (2000) reported that Cr(III) can be bound to the carboxyl groups containing oxygen. In higher pH values, acidic surface groups enhanced the adsorption of Cr(III) onto the biochar. These kinds of groups were identified by FTIR (Fig. 1) and X-ray photoelectron spectroscopy (XPS) (Fig. 8) (which will be discussed later) techniques onto the BCs’ surface.

Effect of NO$_3^-$ and Cl$^-$

The influence of oxidants and reducing agents on Cr(VI) adsorption ability on the biochar is important. Nitrates have quite different impacts on Cr(VI) adsorption onto BCs than chlorides (Fig. 6). In the case of nitrates, the decrease of Cr(VI) adsorption to 74 % (BCS) and 77 % (BCW) was observed at the concentration of these ions below the level of 0.001 mol/L. The concentration above 0.001 mol/L of KNO$_3$ does not cause the decreasing adsorption, which acquires the constant value for both BCs. For the highest ions’ concentration, chlorides cause the decrease of the adsorption value to 49 and 57 % for BCS and BCW, respectively. These results are compared with the adsorption obtained for the Cr(VI) solution without interferences’ presence and with optimal pH value. These drastic effects can be caused by the competitive interactions between Cr(VI) and nitrates or chlorides towards biochar adsorption centers. The more drastic impact of chlorides on Cr(VI) ions’ adsorption can be explained by the fact that for examined BCs, the adsorption was nonspecific or through the diffuse layer. A competitive interaction between CrO$_4^{2-}$ and Cl$^-$ takes place. The differences between the interferences impact on adsorption can be related to higher tendency chlorides for binding to the aliphatic group presented in the structure of BC.

Desorption study

Batch desorption experiments were conducted in order to investigate whether chromium adsorption on investigated biochars is reversible or not. The desorption studies of Cr(VI) ions in relation to HCl and HNO$_3$ concentrations were performed. The desorption kinetics of chromium forms was similar for both materials and does not exceed 20 min (Fig. 7). The maximum desorption of Cr(VI) ions from the

| Biochar | Langmuir | Freundlich |
|---------|----------|------------|
|         | $K_L$    | $a_m$ | $R^2$ | $K$ | $1/n$ | $R^2$ |
| BCS     | 0.017    | 28.1  | 0.990 | 0.06 | 1.45  | 0.888 |
| BCW     | 0.012    | 28.1  | 0.978 | 0.08 | 1.47  | 0.952 |

$a_m$ the maximum adsorbed amount (sorption capacity) (mg/g), $K_L$ the Langmuir constant—the quasi-Gaussian energetic heterogeneity of the adsorption system, $R$ regression coefficient, $K$, $n$ empirical constants indicative of sorption capacity and sorption intensity.
biochars’ surface cannot be achieved even if concentrated HCl or HNO₃ was applied. Additionally, for nitric acid as a desorptive agent, the lowest desorption is observed for BCW (51 %) and the greatest for BCS (79 %). A different effect is observed for hydrochloric acid, wherein the lowest desorption is for BCS (39 %) and greatest for BCW (47 %). This study confirmed that even the application of concentrated hydrochloric or nitric acid does not cause the total desorption of Cr(VI) ions from the biochar surface. Moreover, it should be noticed that chromium(VI) adsorption process onto BCs is irreversible, which can be caused by the surface precipitation of Cr(OH)₃. The differences between desorption obtained in the case of both desorptive agents can be related to the fact that the adsorption of Cr(III) species proceeds by different functional groups, acidic and basic, on the BCS and BCW surfaces. Furthermore, the different structures of studied biochars (different S_BET) (Table 1) can be responsible for uncompleted desorption.

**XPS study of chromium-loaded BCs**

The information about the distribution of element species loaded onto the biochars’ surface and the confirmation of proposed Cr(VI) ions adsorption mechanism were obtained using XPS technique. In Fig. 8, the XPS spectrum of the Cr₂p region is shown. In both element species, Cr(III) and Cr(VI) were detected on the BCs’ surface. The fitted peaks for the Cr₂p region and their possible assignments implicate that the main species as a result of the chromium adsorption onto BCs are Cr(III) and Cr(VI), which represent about 76 and 24 %, respectively. It can be caused by uncompleted reduction of Cr(VI) to Cr(III) on the biochars’ surface. Significant bands at the binding energies of 577 – 580 were used for the identification of the chromium state of valence (Cr₂p₃/2 region). The bands at binding energy of 577.9 and 577.7 for BCS and BCW, respectively, indicate the presence of Cr(III) species (CrCl₃ and Cr(OH)₃). Furthermore, in the same Cr₂p₃/2 region, the bands of 579.7 and 579.5 confirmed the Cr(VI)
species’ loading on the BCs’ surface (CrO₃), in respect to BCS and BCW.

On the basis of the XPS data, it is confirmed that Cr(III) is the most abundant species on the BCs’ surface, and the main mechanism of chromium(VI) adsorption onto these materials is the surface reduction of Cr(VI) to Cr(III). This effect determines the selectivity of the applied method. After the uncompleted reduction, both chromium species are adsorbed on the BCs’ surface by oxygen-containing functional groups. On the basis of Saha and Orvig (2010) classification, the reaction of Cr(VI) ions with biomaterials can proceed by four mechanisms: (1) anionic adsorption, where Cr(VI) ions are adsorbed through electrostatic interaction, without Cr(VI) reduction by porous material; (2) adsorption coupled with reduction, where Cr(VI) is adsorbed and completely reduced by porous material to Cr(III) form; (3) anionic and cationic adsorption, where Cr(VI) is reduced by porous material to Cr(III) which is finally adsorbed onto BCs; and (4) reduction and anionic adsorption, where Cr(VI) is...
adsorbed and the remainder is reduced to Cr(III). Accordingly, the Cr(VI) adsorption onto BCS and BCW can proceed by mixed 1 and 3 mechanisms, because both Cr(VI) and Cr(III) species are present on the biochars’ surface.

SEM-EDX study

To evaluate the surface physical morphology and the porous structure of examined biochars, SEM analysis was performed. Figure 9 presents the SEM images of chromium-loaded BCs and confirmed their porous structure. Furthermore, the Cr elemental mapping shows the presence of chromium onto biochars in the form of an irregular island on material edge. That irregular contribution of chromium loaded onto biochar surface confirms their heterogeneous structure.

Conclusions

The tested biochars were suitable for the removal of Cr(VI) ions from wastewater. The studied BCS were characterized by similar chromium sorption capacities. The Cr(VI) adsorption process was highly pH-dependent and the pH value of 2 was optimal for Cr(VI) removal from the solution. These acidic conditions are favorable for geological sample analysis because precipitation of transition metal ions can be excluded. The obtained experimental adsorption results were successfully fitted to the Langmuir isotherm model. The partial irreversibility process and X-ray photoelectron spectroscopy data confirmed that the main Cr(VI) adsorption mechanism onto BCs is the surface reduction of Cr(VI) to Cr(III). It is worth to pointing out that the tested biochars could be used as an effective, low-cost adsorbent for Cr(VI) removal from aqueous solutions under the determined optimal experimental conditions. The disadvantage of the long-range application of these materials is the inability of repeated usage caused by uncompleted desorption of Cr species from biochars’ surface. The physicochemical improvement processes could be applied for enhancing the adsorptive properties of the studied BCS.

Acknowledgments

The project was funded by the National Science Centre grant on the basis of decision number DEC-2012/07/E/ST10/00572.

Open Access

This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

References

Agrafioti E, Kalderis D, Diamadopoulos E (2014) Arsenic and chromium removal from water using biochars derived from rice husk, organic solid wastes and sewage sludge. J Environ Manag 133:309–314. doi:10.1016/j.jenvman.2013.12.007

Aliabadi M, Khazaiei F, Fakhraee H, Mousavian MTH (2012) Hexavalent chromium removal from aqueous solutions by using low-cost biological wastes: equilibrium and kinetic studies. Int J Environ Sci Technol 9:319–326. doi:10.1007/s13762-012-0045-7

Arslan G, Pehlivan E (2007) Batch removal of chromium(VI) from aqueous solution by Turkish brown coals. Bioresour Technol 98: 2836–2845. doi:10.1016/j.biortech.2006.09.041

Baral A, Engelenk RD (2002) Chromium-based regulations and greening in metal finishing industries in the USA. Environ Sci Pol 5:121–133. doi:10.1016/S1462-9011(02)00029-X

Beesley L, Moreno-Jiménez E, Gomez-Eyles JL et al (2011) A review of biochars’ potential role in the remediation, revegetation and restoration of contaminated soils. Environ Pollut 159:3269–3282. doi:10.1016/j.envpol.2011.07.023

Choppala GK, Bolan NS, Megharaj M et al (2012) The influence of biochar and black carbon on reduction and bioavailability of chrome in soils. J Environ Qual 41:1175–1184. doi:10.2134/jeq2011.0145

Deveci H, Kar Y (2013) Adsorption of hexavalent chromium from aqueous solutions by biochars obtained during biomass pyrolysis. J Ind Eng Chem 19:190–196. doi:10.1016/j.jiec.2012.08.001

Dhal B, Thatoi HN, Das NN, Pandey BD (2013) Chemical and microbial remediation of hexavalent chromium from contaminated soil and mining/metallurgical solid waste: a review. J Hazard Mater 250–251:272–291. doi:10.1016/j.jhazmat.2013.01.048

Dobrowski R, Otto M (2010) Study of chromium(VI) adsorption onto modified activated carbons with respect to analytical application. Adsorption 16:279–286. doi:10.1007/s10450-010-9240-3

Gardea-Torresdey JL, Tiemann KJ, Armendariz V et al (2000) Characterization of Cr(VI) binding and reduction to Cr(III) by the agricultural byproducts of Avena monida (Oat) biomass. J Hazard Mater 80:175–188. doi:10.1016/S0304-3894(00)00301-0

Gottipati R, Mishra S (2010) Process optimization of adsorption of Cr(VI) on activated carbons prepared from plant precursors by a two-level full factorial design. Chem Eng J 160:99–107. doi:10.1016/j.cej.2010.03.015

Kimbrough DE, Cohen Y, Winer AM et al (1999) A critical assessment of chromium in the environment. Crit Rev Environ Sci Technol 29:1–46. doi:10.1080/10643389991259164

Kolodyńska D, Wnętrzak R, Leahy J et al (2012) Kinetic and adsorptive characterization of biochar in metal ions removal. Chem Eng J 197: 295–305. doi:10.1016/j.cej.2012.05.025

Kotar J, Stasiak A, Stasiak Z (2000) Chromium occurrence in the environment and methods of its speciation. Environ Pollut Barking Essex 107:263–283

Kuo S, Bembenek R (2008) Sorption and desorption of chromate by wood shavings impregnated with iron or aluminum oxide. Bioresour Technol 99:5617–5625. doi:10.1016/j.biortech.2007.10.062

Mohan D, Pittman CU (2006) Activated carbons and low cost adsorbents for remediation of tri- and hexavalent chromium from water. J Hazard Mater 137:759–762. doi:10.1016/j.jhazmat.2006.06.060

Olęszczuk P, Jośko I, Kuśmiercz M (2013) Biochar properties regarding to contaminants content and ecotoxicological assessment. J Hazard Mater 260:375–382. doi:10.1016/j.jhazmat.2013.05.044

Saha B, Orvig C (2010) Biosorbent for hexavalent chromium elimination from industrial and municipal effluents. Coord Chem Rev 254: 2959–2972. doi:10.1016/j.ccr.2010.06.005

Tang J, Zhu W, Kooekana R, Katayama A (2013) Characteristics of biochar and its application in remediation of contaminated soil. J Biosci Bioeng 116:653–659. doi:10.1016/j.jbiosc.2013.05.035

Van Reeuwijk LP (1992) Procedures for soil analysis. International Soil Reference and Information Centre. Wageningen, The Netherlands

Zhang W, Mao S, Chen H et al (2013) Pb(II) and Cr(VI) sorption by biochars pyrolyzed from the municipal wastewater sludge under different heating conditions. Bioresour Technol 147:545–552. doi:10.1016/j.biortech.2013.08.082