The Thermogenic Responses to Overfeeding and Cold are Differentially Regulated

Courtney M. Peterson1, Virgile Lecoultre1,2, Elizabeth A. Frost1, Jonathan Simmons1, Leanne M. Redman1, and Eric Ravussin1

Objective: Brown adipose tissue (BAT) is a highly metabolic tissue that generates heat and is negatively associated with obesity. BAT has been proposed to mediate both cold-induced thermogenesis (CIT) and diet-induced thermogenesis (DIT). Therefore, it was investigated whether there is a relationship between CIT and DIT in humans.

Methods: Nine healthy men (23 ± 3 years old, 23.0 ± 1.8 kg m⁻²) completed 20 min of cold exposure (4°C) 5 days per week for 4 weeks. Before and after the intervention, CIT (the increase in resting metabolic rate at 16°C relative to 22°C) was measured by a ventilated hood indirect calorimeter, whereas DIT was measured as the 24-h thermic response to 1 day of 50% overfeeding (TEF150%) in a respiratory chamber.

Results: After the cold intervention, CIT more than doubled from 5.2% ± 14.2% at baseline to 12.0% ± 11.1% (P = 0.05), in parallel with increased sympathetic nervous system activity. However, 24-h energy expenditure (2,166 ± 206 vs. 2,118 ± 188 kcal day⁻¹; P = 0.15) and TEF150% (7.4% ± 2.7% vs. 7.7% ± 1.6%; P = 0.78) were unchanged. Moreover, there was no association between CIT and TEF150% at baseline or post-intervention, nor in their changes (P ≥ 0.47).

Conclusions: Cold acclimation resulted in increased CIT but not TEF150%. Therefore, it is likely that CIT and DIT are mediated by distinct regulatory mechanisms.

Introduction

Negative energy balance is required to reduce body weight and is achieved by reducing energy intake, increasing energy expenditure (EE), or both. The recent identification of active brown adipose tissue (BAT) in adults (1-5) has revived interest in targeting BAT to increase the facultative components of EE and potentially promote weight loss (6). In response to cold exposure, BAT generates heat to help maintain core body temperature, a phenomenon termed cold-induced thermogenesis (CIT) (6). The heat induced by BAT is generated via mitochondrial uncoupling through mitochondrial uncoupling protein 1 (UCP1) and is enabled by BAT’s multiple lipid droplets, abundant mitochondria, high vascularity, and rich innervation by the sympathetic nervous system (SNS). These unique characteristics serve to fuel an intense metabolic activity, as demonstrated by a twofold increase in blood flow (7-9), a several-fold increase in glucose and free fatty acid uptake (3,8,10-12), and an increase in oxidative metabolism (10,13) upon cold stimulation.

Since the recent finding that active BAT is present in a majority of adults (2,4,8,10,14-17), several studies have reported that BAT activity is inversely associated with BMI (1,2,4,5,14-16), suggesting a role for BAT in the regulation of energy balance. Acute cold exposure typically increases EE by 5-17% (2,9,11,12,15,17-27), in parallel with increases in BAT activity (12,14,28). However, it has been reported that CIT is significantly lower in overweight and BAT-negative individuals (2,9,11,15,17,21,25,26,28). Interestingly, longer-term cold exposure upregulates BAT activity (and possibly mass) (1,4,13,14,19,28,29) and increases CIT (14,24,28,30), resulting in modest body weight reduction (19,28) and slightly improved glucose metabolism (11,19,27,29). Thus, habitual exposure to cold may provide a simple way to enhance EE and to improve metabolic health through BAT-mediated CIT.

In addition to CIT, BAT has also been hypothesized to regulate diet-induced thermogenesis (DIT). Rothwell and Stock initially observed that overfeeding resulted in less weight gained than anticipated but that the effect was inhibited by propranolol (31), suggesting that SNS stimulation of BAT may be involved; these changes occurred in parallel with an increase in BAT mass. Later studies showed that even at thermoneutrality, UCP1 knockout mice are

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1 John S. McIlhenny Skeletal Muscle Physiology Laboratory, Pennington Biomedical Research Center, Baton Rouge, Louisiana, USA.
2 Faculty of Sciences, Department of Medicine, University of Fribourg, Switzerland.

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heavier than their wild-type counterparts (32) and that BAT transplantation reduces body weight (33). Since then, several studies in rodents (20,31,34) and humans (18,19,34) have reported associations between postprandial EE and BAT or CIT; however, many other studies refute this association (20,27,35-38).

Since the overall relationships among BAT activity, CIT, and DIT are unclear, we designed a study to determine whether the magnitude of changes in EE in response to cold (CIT) and overfeeding in humans are correlated. To assess the potential for DIT (which requires long-term overfeeding studies), we measured the thermic response during 1 day of 50% overfeeding (TEF\textsubscript{150\%}) in a respiratory chamber. TEF\textsubscript{150\%} represents the increase in postprandial EE due to the consumption of 150% of the daily energy requirement divided by the ingested calories. This measure can be viewed as halfway between the thermic effect of food (TEF; excess energy expended in response to a single meal) and DIT (adaptive dietary thermogenesis in response to long-term overfeeding). We hypothesized that cold acclimation would increase both CIT and TEF\textsubscript{150\%} and that their changes would be correlated, thus suggesting that CIT and DIT are regulated by similar mechanisms.

Methods

Participants

Eleven participants enrolled in this clinical trial (NCT01898949) at Pennington Biomedical Research Center (Baton Rouge, LA). Participants were recruited via electronic advertising targeting healthy, lean men (BMI between 18.5-25 kg m\textsuperscript{-2}) between the ages of 18 and 35. Potential participants were excluded for smoking, chronic alcohol consumption (>3 drinks/day), current use of medication, recent changes in body weight (>2 kg in the prior 6 months), impaired fasting glucose (>100 mg dl\textsuperscript{-1}), regular intense exercise (>3 times/week), or chronic disease. The study was approved by the Pennington Biomedical Institutional Review Board and was conducted in accord with the Declaration of Helsinki. All participants provided written informed consent prior to participation.

Study design

To increase the capacity for non-shivering thermogenesis and likely BAT activity, participants spent 20 min per day, five consecutive days per week, for 4 weeks in a cold room (4°C). This protocol was piloted to see whether intense cold exposure for short periods of time could be used as an effective cold exposure intervention, thus reducing participant time burden. In the cold room, participants wore light clothing (T-shirt, shorts and light shoes, e.g., flip flops) and a wrist watch to monitor heart rate variability (HRV) to track SNS activity. Participants stood for the duration of the cold exposure, and compliance to the protocol was closely monitored. Before and after the cold exposure intervention, EE in response to acute cold exposure (CIT) and overfeeding (TEF\textsubscript{150\%}) were measured according to a 2-day testing protocol (Figure 1). Body composition was measured by DXA (Hologics, Bedford, MA), and EE was measured by indirect calorimetry using a ventilated hood system, with concomitant HRV assessment, in both thermoneutral (22°C) and cold (16°C) conditions. Core temperature was monitored throughout the EE measurements using an ingestible telemetric temperature capsule (CorTemp, HQInc, Palmetto, FL). The following day, vital signs (blood pressure, pulse, temperature) were measured, and a fasting blood sample was collected to measure glucose (Beckman Coulter DXC600) and insulin (Siemen Immulite 2000). Participants were then admitted to the respiratory chamber for 24 h and fed an overfeeding diet (50% above energy requirements) at thermoneutrality (22°C).

Cold-induced thermogenesis

Resting metabolic rate (RMR) was measured at thermoneutrality and in response to acute cold by indirect calorimetry using a ventilated hood system (Max II Metabolic Cart; AEI Technologies, Naperville, IL). Participants refrained from performing vigorous physical activity and from consuming caffeine-containing drinks for 24 h prior to testing. In the morning after an overnight fast, participants rested supine at thermoneutrality (22°C) for 30 min, and then RMR was measured for 30 min. Participants were then rolled on a wheelchair into a cold room (16°C). After 90 min of acute cold exposure, the
30-min RMR measurement was repeated. CIT was calculated as the percent increase in RMR following acute cold exposure relative to RMR in the thermoneutral condition. In addition, HRV was measured during the 8 min prior to each RMR measurement using a heart rate chest strap and monitor (Polar S810, Polar Electro Oy, Kempele, Finland).

Thermic effect of food for 1 day of 50% overfeeding (TEF150%)
EE was measured over 24 h in a respiratory chamber (39) at thermoneutrality (22°C) while participants consumed 150% of their weight maintenance energy requirements. Energy requirements were estimated using the Dietary Reference Intakes equation (40) for resting EE (REE), adjusted with an activity factor of 1.3. At 0730 h, participants swallowed a core temperature capsule, and fasting blood was drawn for measurement of glucose and insulin. Participants were dressed in medical scrubs and then entered the chamber and were instructed to limit motion and lay supine on the bed for the first hour of measurement to assess REE. Thereafter, participants were allowed to move freely in the chamber, but exercise was not permitted. Breakfast, lunch, an afternoon snack, and dinner were served at 0900 h, 1330 h, 1600 h, and 1900 h, respectively, and provided an overall macronutrient composition of 15% protein, 50% carbohydrate, and 35% fat. Participants were required to eat their meals within 30 min. Lights were turned off at 2230 h, and sleeping metabolic rate (SMR) was calculated between 0200 h and 0500 h for all minutes with radar activity <1%. Participants were awoken at 0630 h the following morning, and they exited the chamber at 0715 h. Participants collected all their urine during their chamber stay for assessment of urinary nitrogen and subsequent determination of protein oxidation. Physical activity within the chamber was quantified as percent of time in motion as measured by radar, and EE attributed to this spontaneous physical activity (SPA) was calculated as total daily EE minus REE and TEF150%. TEF150% was determined by subtracting REE from the y-intercept of the regression of EE versus % activity using data averaged in 15-min intervals, as previously described (39).

Heart rate variability
Autonomic function was assessed by spectral analysis of HRV using a heart-rate belt and a wrist recorder (Polar S810, Polar, USA). Breathing frequency was controlled at a rate of 15 breaths per minute using a metronome. Spectral analysis of heart rate data was performed using the Kubios HRV software (Kubios HRV, v. 2.1; Department of Applied Physics, University of Eastern Finland, Kuopio, Finland). For frequency domain data, a discrete Fourier transformation algorithm was used, and the spectral power was quantified by integrating the areas under the curve for the following frequency bands: very low frequency (VLO; 0.007 to 0.035 Hz), low frequency (LO; 0.035 to 0.15 Hz), high frequency (HI; 0.15 to 0.5 Hz), and total power (0.007 to 0.5 Hz). The spectral power in the VLO and LO bands were taken to be indices of SNS activity, while the power in the HI band was taken as an index of parasympathetic activity.

Statistical analysis
Because this was a pilot study with a novel cold exposure intervention, no power analysis was performed. Data are expressed as mean ± SD. The effect of the cold exposure intervention was assessed using a two-tailed Student’s paired t test, except where otherwise noted. To determine both the separate effects of and the interaction between the cold intervention and ambient temperature condition, 2 × 2 repeated measures ANOVAs were also performed on HRV, CIT, and EE data. The correlation between TEF150% and CIT was considered to be the primary endpoint. Correlations were performed using parametric least squares linear regression. The Type I error rate was set at α = 0.05.

Results
Participant characteristics
Nine out of 11 healthy, male participants recruited from the Greater Baton Rouge area completed the study; two participants were dropped from the study for non-compliance with the study intervention. The nine completers (8 Caucasian, 1 Hispanic) had a mean age of 23 ± 3 years and a mean BMI of 23.0 ± 2.2 kg m⁻² (Table 1).

Effect of the cold intervention on cardiometabolic health
After the 4-week cold intervention, participants lost an average of 0.5 ± 0.5 kg (from 71.5 ± 6.5 to 71.0 ± 6.6 kg; P = 0.03), which is consistent with weight loss reported in other studies (19,28). Repeated cold exposure decreased systolic blood pressure from 114 ± 6 to 110 ± 6 mm Hg (P = 0.05), while diastolic blood pressure declined from 70 ± 6 to 67 ± 3 mm Hg (P = 0.04). Fasting glucose and insulin (Table 1) were unaffected by the cold intervention (P > 0.15), with no change in insulin resistance as estimated by HOMA-IR (P = 0.62). Baseline heart rate was 64 ± 5 beats per minute (bpm), and the cold intervention tended to modestly decrease it by 3 ± 5 bpm (P = 0.09). Core temperature at thermoneutrality was unchanged (P = 0.88).

| Table 1: Participant characteristics and cardiometabolic indices |
|----------------|-------|-------|
| Age (years)    |       |       |
| Weight (kg)    | 71.5 ± 6.5 | -0.5 ± 0.5 | 0.03** |
| BMI (kg m⁻²)   | 23.0 ± 1.8 | -0.2 ± 0.2 | 0.03** |
| % Body fat     | 22.2 ± 5.7 | -    | -    |
| Systolic blood pressure (mm Hg) | 114 ± 6 | -4 ± 4 | 0.05** |
| Diastolic blood pressure (mm Hg) | 70 ± 6 | -3 ± 3 | 0.04** |
| Heart rate (bpm) | 64 ± 5 | -3 ± 5 | 0.09* |
| Core temperature (°C) | 36.7 ± 0.1 | 0.0 ± 0.1 | 0.88 |
| Fasting glucose (mg dl⁻¹) | 90 ± 4 | 2 ± 4 | 0.15 |
| Insulin (mU l⁻¹) | 4.7 ± 1.7 | 0.3 ± 2.2 | 0.83 |
| HOMA-IR         | 0.84 ± 0.42 | 0.09 ± 0.50 | 0.62 |
| Total cholesterol (mg dl⁻¹) | 167 ± 26 | -    | -    |
| LDL (mg dl⁻¹)   | 95 ± 17 | -    | -    |
| HDL (mg dl⁻¹)   | 53 ± 11 | -    | -    |
| Triglycerides (mg dl⁻¹) | 92 ± 38 | -    | -    |

Values are mean ± SD. *P < 0.10. **P < 0.05, according to a two-tailed paired t test.
Effect of the cold intervention on CIT
At baseline under thermoneutral conditions, RMR was 1702 ± 124 kcal/24 h and remained unchanged following the 4-week cold intervention (1636 ± 184 kcal/24 h; \( P = 0.14 \)). Under acute mild cold conditions (16°C), RMR increased from 1782 ± 198 kcal/24 h during baseline testing to 1824 ± 214 kcal/24 h during post-intervention testing, and there was no significant temperature by time interaction (\( P = 0.36 \)). CIT was 5.2% ± 14.2% at baseline and increased to 12.0% ± 10.6% after the 20 sessions of cold exposure (\( P = 0.05 \)), indicating that cold acclimation did indeed occur (Figure 2). Shivering was not observed by research staff or reported by the participants at any time during the RMR testing in the mild cold conditions (nor during the respiratory chamber testing), indicating that the increase in EE was due to non-shivering thermogenesis. Lastly, there was no statistically significant change in the respiratory quotient (RQ) in response to either acute cold exposure or the cold intervention (\( P \geq 0.10 \); data not shown).

Effect of the cold intervention on EE in response to overfeeding
Participants were overfed by 50.2% ± 4.6% at baseline versus 53.1% ± 3.4% post-cold acclimation (\( P = 0.24 \)). Twenty-four-hour EE in response to overfeeding in thermoneutral conditions (Figure 3) was similar at baseline (2166 ± 206 kcal day\(^{-1}\)) and following the 4-week cold intervention (2118 ± 188 kcal day\(^{-1}\); \( P = 0.15 \)). The 50% overfeeding resulted in a robust value of 7.4% ± 2.7% for TEF\(_{150\%}\) before the cold intervention, and this value was unchanged at 7.7% ± 1.6% after the intervention period (\( P = 0.78 \)). The other subcomponents of EE—SMR and EE from SPA—were unchanged following the cold intervention (Figure 3; \( P \geq 0.15 \)). We also observed no differences in fat, protein, or carbohydrate oxidation (\( P \geq 0.15 \); data not shown). Similarly, there was no difference in 24-h core temperature during overfeeding (36.90 ± 0.35°C vs. 36.90 ± 0.29°C; \( P = 1.00 \)) before and after the cold intervention.

Effect of the cold intervention on SNS activity
Both acute cold exposure and the cold intervention increased HRV as measured by the standard deviation of RR intervals, relative to baseline at thermoneutrality (\( P = 0.02 \); Table 2). Similarly, SNS activity as indicated by VLO and LO spectral power was significantly increased in response to both acute cold exposure and after cold acclimation, relative to baseline testing thermoneutrality. At baseline, acute cold exposure stimulated VLO (\( P = 0.04 \)) and total power (\( P = 0.05 \)) by 2.3- and 1.7-fold, respectively. Interestingly, the cold intervention increased VLO power by threefold (\( P = 0.03 \)) and increased total power by twofold (\( P = 0.02 \)) under thermoneutral conditions. Under mild cold conditions (16°C), however, the cold intervention did not significantly affect spectral power (\( P \geq 0.24 \)); however, the large inter-individual responses to the cold intervention may obscure any modest changes.

Correlations between CIT and TEF\(_{150\%}\)
Despite cold acclimation-induced increases in CIT and SNS activity, TEF\(_{150\%}\) did not increase (\( P = 0.78 \)). Contrary to our hypothesis, no correlations were observed between CIT and TEF\(_{150\%}\) at baseline or following the cold intervention, nor did the changes in CIT and TEF\(_{150\%}\) values correlate with each other (\( P \geq 0.47 \); Figure 4). However, baseline TEF\(_{150\%}\) did negatively correlate with the change in TEF\(_{150\%}\) (\( r = -0.83 \); \( P = 0.006 \)), which likely reflects reversion to the mean.

Discussion
Since the recent discovery of functional BAT in adults (1-5) and its negative correlation with BMI (1,2,4,5,14-16), there has been a flurry of research to determine whether targeting BAT activity can be used to treat obesity. While much focus is on enhancing CIT or SNS activity, BAT may mediate other facultative components of EE, such as DIH, as first proposed in 1979 by Rothwell and Stock (31).

In this study, we investigated whether the thermic response to 1 day of 50% overfeeding and CIT are mediated by similar mechanisms in humans. To maximize CIT and BAT activity, we exposed nine healthy men to a 4-week cold acclimation protocol; cold acclimation is known to enhance BAT activity (13,14,19,28,29), as well as CIT (14,28,30). Following the 4-week cold intervention, CIT more than doubled from 5.2% ± 14.2% to 12.0% ± 11.1%, a finding
TABLE 2 HRV in thermoneutral (22°C) and cold conditions (16°C)

|                          | Baseline          | Post             | P value |
|--------------------------|-------------------|------------------|---------|
| **Thermoneutral conditions (22°C)** |                   |                  |         |
| Mean RR interval (ms)    | 1,045 ± 185       | 1,106 ± 147      | 0.20    |
| SD of RR intervals (ms)  | 96 ± 16           | 122 ± 38         | 0.02**  |
| VLO power (ms²)          | 3,658 ± 2,818     | 10,887 ± 8,438   | 0.03**  |
| L0 power (ms²)           | 2,660 ± 2,217     | 3,547 ± 1,710    | 0.08*   |
| HI power (ms²)           | 1,843 ± 1,269     | 1,878 ± 1,156    | 0.94    |
| Total power (ms²)        | 8,160 ± 2,544     | 16,313 ± 9,263   | 0.02**  |
| **Cold conditions (16°C)** |                   |                  |         |
| Mean RR interval (ms)    | 1,042 ± 198       | 1,037 ± 159      | 0.91    |
| SD of RR intervals (ms)  | 119 ± 36          | 138 ± 44         | 0.28    |
| VLO power (ms²)          | 8,225 ± 5,837     | 13,506 ± 11,298  | 0.24    |
| L0 power (ms²)           | 3,121 ± 2,758     | 3,450 ± 2,505    | 0.58    |
| HI power (ms²)           | 3,021 ± 2,989     | 2,005 ± 1,356    | 0.19    |
| Total power (ms²)        | 14,366 ± 9,186    | 18,961 ± 12,849  | 0.38    |

Four weeks of cold acclimation increased very low frequency (VLO), low frequency (L0), and total power, confirming that the cold acclimation upregulated SNS activity. However, the increases only reached statistical significance under thermoneutral conditions. Values are mean ± SD.

*P < 0.10.  **P < 0.05, according to a two-tailed paired t test.

consistent with other studies (14,28,30). For instance, van der Lans and colleagues demonstrated a 7% absolute increase in CIT following exposure to mild cold (16°C) for 6 h day⁻¹ for 10 days (14), while Yoneshiro and colleagues reported comparable results after 2 h of daily exposure to mild cold (19°C) for 6 weeks (28). Our results suggest that enhancing the metabolic response to cold may also be achieved by exposing participants to colder temperatures (4°C) but for shorter durations (e.g., 20 min day⁻¹ over 4 weeks), thereby significantly reducing participant time burden. Interestingly, in response to our cold intervention, blood pressure slightly but significantly improved following the cold intervention, but HOMA-IR did not. In addition, our HRV results are consistent with those of other studies [e.g., (27)] and demonstrate that the 4-week cold intervention increased SNS activity. Both BAT activity and the facultative part of DIT are known to be mediated by SNS activity. Therefore, if the same SNS mechanism mediates both CIT and DIT, we expected that the cold acclimation (which enhances SNS activity even in thermoneutral conditions) would also enhance TEF₁₅₀%.

However, despite the increases in CIT and SNS activity following 4 weeks of cold acclimation, the thermic response to 50% overfeeding was unchanged (7.4% ± 2.7% before versus 7.7% ± 1.6% after; *P = 0.78). In addition, core body temperature, 24-h EE, and its subcomponents were not altered by the intervention (*P ≥ 0.15). Furthermore, CIT and TEF₁₅₀% were not associated at baseline or post-intervention, nor did the changes in CIT and TEF₁₅₀% correlate with each other (*P ≥ 0.47). In sum, this suggests that DIT and CIT are mediated through different pathways.

Prior rodent and human studies have provided conflicting results on whether BAT and/or CIT are related to DIT (18-20,31,32,34-38).

For instance, Rothwell and Stock (31) originally reported enhanced oxygen consumption in response to overfeeding in rats, in parallel with increases in BAT mass, but a later study showed that BAT activity accounted for only 2-3% of DIT in the same rodent model (37). Similarly, the finding that *UCP1* knockout mice had reduced DIT (32) was later refuted (36,38). The knockout animals were hyperphagic, and DIT was erroneously defined as the ability of animals to respond to adrenergic stimulation (36).

Six prior studies in humans have attempted to measure postprandial responses to a meal in the context of BAT, CIT, or cold exposure. One study measured BAT activity by PET/CT in response to 100% overfeeding and found that none of the participants had increased BAT activity with overfeeding, despite 75% of participants having increased BAT activity in response to cold (35). Two studies with EE measurements in respiratory chambers have reported that TEF was associated with CIT and/or BAT activity (19,34); however, in both studies, TEF was not measured directly, but rather either 24-h EE or postprandial EE (both include EE attributed to physical activity) was used instead. In one of the studies (19), impossibly high values of TEF in excess of 30% were reported. Notably, a third study measuring postprandial EE using the same method came to the opposite conclusion and reported data suggesting that TEF is somewhat blunted upon mild cold exposure (27). Overall, these mixed results underscore that measuring postprandial EE without accounting for activity levels should not be used to estimate TEF.

Only two human studies have directly measured TEF in the context of cold exposure or BAT activity, although neither incorporated both overfeeding and measurement of CIT. One study by Vosselman and colleagues (20) observed no direct relation between either CIT or postprandial BAT activity and TEF in response to a single meal. The second study, by van Marken Lichtenbelt et al. (18), found no difference in TEF in response to *ad libitum* food intake measured at 16°C in comparison to thermoneutrality (22°C). We therefore performed the first *bona fide* investigation of the relationship between CIT and TEF in response to a structured 24-h overfeeding paradigm, and we report that TEF values before and after the cold intervention were not associated with CIT. Our study does have some limitations.

Figure 4 Relationship between 50% overfeeding (TEF₁₅₀%) and cold-induced thermogenesis (CIT). There was no correlation between baseline or post-intervention values of CIT and TEF₁₅₀% (shown above), or in the changes in TEF₁₅₀% and CIT values (*P ≥ 0.47).
such as the small number of participants, not measuring BAT activity or muscle shivering activity, and measuring TEF by respiratory chamber method, which is less reproducible than by ventilated hood system. Nonetheless, our results and evidence from other studies indicate that CIT and DIT are probably mediated by different regulatory mechanisms.

The hypothesis that CIT and DIT are mediated by different mechanisms is supported by suggestive but preliminary evidence from mechanistic investigations. First, insulin infusion increases BAT glucose uptake but not tissue perfusion (unlike with cold exposure), suggesting that BAT thermogenesis is not triggered by feeding (8). Second, unlike in rodents, Muzik et al. (9) showed that most of the increase in BAT EE during cold exposure in humans is mediated by changes in blood flow, and thus BAT can account for only up to 15-25 kcal day⁻¹.

In conclusion, our observations indicate that as little as 20 min of daily exposure to cold (4°C) enhances CIT, in parallel with increased SNS activity. Combined with evidence from previous reports, our results suggest that the mechanisms underlying CIT are probably different from those mediating the thermogenic response to acute overfeeding.

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