CSF albumin and immunoglobulin analyses in childhood neurologic disorders

ABSTRACT

Objective: To evaluate the utility of qualitative and quantitative analyses of CSF immunoglobulins as part of the diagnostic workup of CNS inflammatory conditions.

Methods: One hundred eighty-nine children who underwent CSF investigation for their neurologic condition had CSF and serum testing to (1) qualitatively identify oligoclonal band (OCB) patterns and (2) quantitatively measure the immunoglobulin (Ig) G index and albumin quotient (QAlb). Case notes were retrospectively reviewed and patients were grouped according to whether their primary diagnosis was due to an inflammatory (n = 104) or noninflammatory (n = 85) etiology.

Results: CSF-restricted OCBs were found in 20/104 (19%) of the inflammatory group compared with 4/85 (5%) of the noninflammatory group (p = 0.0036). Mirrored OCBs were found in 13/104 (12.5%) of the inflammatory group compared with 5/85 (6%) of the noninflammatory group (p = 0.14). IgG index and QAlb were significantly higher in patients with an inflammatory etiology. However, a raised IgG index (>0.85) and QAlb (>0.049) were seen in both groups, with QAlb abnormalities seen more frequently in the inflammatory group (p = 0.0028).

Conclusions: Both methods were informative in identifying inflammatory mechanisms. Abnormalities were more commonly, but not exclusively, seen in primary inflammatory conditions. The qualitative and quantitative evaluation collectively revealed additional positive results than when done in isolation. Neurology Neuroimmunol Neuroinflammation 2014;1:e10; doi: 10.1212/NXI.0000000000000010

GLOSSARY

ICD-10 = International Classification of Diseases, tenth revision; Ig = immunoglobulin; MS = multiple sclerosis; OCB = oligoclonal band; QAlb = albumin quotient.

CSF analysis of immunoglobulins (Igs; often IgG) includes a qualitative analysis of concurrent sera and CSF to identify the 5 characteristic oligoclonal band (OCB) patterns.1 Type 1 is a normal pattern where no bands are identified. A type 2 pattern indicates intrathecal synthesis, where bands are seen only in the CSF. When the pattern of bands seen is identical in both sera and CSF, a “mirrored” type 4 pattern is recorded, demonstrating that the IgG has passively diffused into the CNS. Sometimes the pattern identified has identical shared bands but additional CSF-specific bands, a type 3 pattern. On rare occasions, a type 5 pattern is seen, in which a monoclonal IgG band is identified in serum and CSF (detailed description provided in reference 1). In addition, the CSF and sera can be quantitatively analyzed by measuring the albumin quotient (QAlb = AlbCSF/AlbSERUM) and IgG index (IgG Index = IgGCSF/IgGSERUM)/(AlbCSF/AlbSERUM) to evaluate blood-brain barrier dysfunction.1

The quantitative analysis of sera and CSF has little added value to the qualitative analysis of bands in the diagnosis of multiple sclerosis (MS),1 although it is less clear whether this is the case across the range of neurologic disorders. Two studies from more than 2 decades ago have studied qualitative and quantitative CSF analysis in a range of neurologic conditions. The first study highlighted the additional value of testing serum and CSF together and identified identical bands in the serum in 50% (56/112) of the patients, suggesting a systemic immune response.2 The second study, which

From Children’s Neuroscience, Evelina London Children’s Hospital at Guy’s & St Thomas’ NHS Trust, King’s Health Partners AHSC, Lambeth Palace Road, London.

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The first pediatric study, was very informative but included only 33 children (out of the 161 studied) with inflammatory conditions. A contemporary Australian study reported the diagnostic value of qualitative CSF IgG analysis in a range of childhood-onset neurologic diseases. Therefore, the aim of this study was to evaluate the utility of the qualitative and quantitative evaluation of the CSF when investigating children with CNS inflammatory conditions.

### Table 1

**ICD-10 inflammatory diseases of the central and peripheral nervous system and the qualitative (oligoclonal band pattern) and quantitative (IgG index and QAlb) analysis**

| ICD-10          | Type 1, no OCB | Type 2, intrathecal | Type 3, mirrored and intrathecal | Type 4, mirrored | Raised IgG Index (>0.85) | Raised QAlb (>4.9 × 10⁻¹) |
|-----------------|----------------|---------------------|----------------------------------|-----------------|------------------------|--------------------------|
| All inflammatory (n = 104) | 72 (69) | 10 (10) | 10 (10) | 13 (13) | 21 (20) | 27 (26) |
| Monophasic ADS (n = 29)* | G36; G37.3; G37.8 | 17 (59) | 2 (7) | 4 (14) | 7 (24) | 5 (17) | 8 (28) |
| Non-MS relapsing demyelination (n = 5) | G37.9 | 3 (60) | 2 (40) | 0 | 0 | 1 (20) | 2 (40) |
| MS (n = 3)* | G35 | 0 | 3 (100) | 0 | 0 | 2 (67) | 0 |
| CNS infections (n = 11)* | G01; G02; G04; G05 | 8 (72) | 2 (18) | 1 (9) | 0 | 4 (36) | 3 (27) |
| Antibody-mediated encephalitis (n = 5) | G05.8 | 5 (100) | 0 | 0 | 0 | 0 | 0 |
| Encephalopathies (immune) (n = 15) | G05.8 | 12 (80) | 0 | 2 (13) | 1 (7) | 2 (13) | 4 (27) |
| CNS vasculitis (n = 3) | G08; I67.6 | 1 (33) | 1 (33) | 1 (33) | 0 | 0 | 1 (33) |
| Peripheral demyelination (n = 14) | G61 | 10 (71) | 0 | 0 | 4 (29) | 4 (29) | 8 (58) |
| Cranial neuropathies (n = 8)* | G50–54 | 6 (75) | 0 | 2 (25) | 0 | 2 (25) | 1 (13) |
| Postinfectious ataxia (n = 5) | G09 | 5 (100) | 0 | 0 | 0 | 0 | 0 |
| Movement disorder (immune) (n = 4) | G09 | 2 (67) | 0 | 0 | 1 (33) | 1 (33) | 0 |
| Uveitis (n = 3) | H20 | 3 (100) | 0 | 0 | 0 | 0 | 0 |
| All noninflammatory (n = 85) | 76 (89) | 3 (4) | 1 (1) | 5 (6) | 14 (16) | 7 (8) |
| Seizure disorder (n = 22) | G40 | 19 (87) | 1 (5) | 0 | 2 (9) | 2 (9) | 2 (9) |
| Neuroregression (n = 13) | G31 | 11 (85) | 0 | 0 | 2 (15) | 4 (31) | 2 (15) |
| Neurometabolic disorder (n = 11) | E70–90 | 10 (91) | 1 (9) | 0 | 0 | 4 (36) | 1 (9) |
| Headaches (n = 8; 6 IIH, 2 migraine) | G43; G93.2; R51 | 8 (100) | 0 | 0 | 0 | 0 | 0 |
| Developmental delay (n = 6) | F70–79 | 5 (83) | 1 (17) | 0 | 0 | 0 | 0 |
| Movement disorder (n = 6) | G20–26 | 6 (100) | 0 | 0 | 0 | 0 | 0 |
| Other encephalopathies (Wernicke, hypertensive, drug toxicity, PRES) (n = 5) | E51.2; I67.4; G92 | 3 (60) | 0 | 1 (20) | 1 (20) | 1 (20) | 1 (20) |
| Malignancy (n = 4) | C00–C75 | 4 (100) | 0 | 0 | 0 | 1 (25) | 0 |
| Non-CNS infections (n = 4) | B34; P36.1 | 4 (100) | 0 | 0 | 0 | 1 (25) | 0 |
| Functional (n = 3) | C69–C72 | 3 (100) | 0 | 0 | 0 | 0 | 0 |
| Vascular (n = 2) | I63 | 2 (100) | 0 | 0 | 0 | 1 (50) | 0 |
| Peripheral neuropathy (n = 1) | G62 | 1 (100) | 0 | 0 | 0 | 0 | 0 |

Abbreviations: ADS – acquired demyelinating syndromes; HSV – herpes simplex virus; ICD-10 – International Classification of Diseases, tenth revision; Ig – immunoglobulin; IIH – idiopathic intracranial hypertension; MS – multiple sclerosis; OCB – oligoclonal band; PRES – posterior reversible encephalopathy syndrome; QAlb – albumin quotient; VZV – varicella-zoster virus.

Data are n (%), except for the ICD-10 column.

* Demyelinating phenotypes were classified based on International Pediatric MS Study Group criteria into monophasic ADS (15 acute disseminated encephalomyelitis, 2 optic neuritis, 7 transverse myelitis, and 4 other clinically isolated syndrome) and relapsing phenotypes. One child with non-MS relapsing demyelination was diagnosed with neuromyelitis optica.

* Twelve children were diagnosed with MS at our center during the study period. Four had CSF analysis done at the referring institution, 2 had a firm clinical diagnosis (and had not had CSF studies), and 3 were between the ages of 16 and 18 years and hence were not included.

* All 11 patients with a diagnosis of CNS infection had the relevant serum and CSF investigations to confirm the diagnosis; of these, 3 patients had bacterial meningitis/myelitis, 4 had CSF PCR-proven viral encephalitis (3 HSV, 1 VZV), 2 had clinical syndrome associated with rising viral titers (1 H1N1, 1 influenza A), and 2 had presumed viral encephalitis.

* Four patients with 7th nerve palsy, 2 with 6th, 1 with 3rd, and 1 with multiple cranial neuropathies.
METHODS Between 2007 and 2012, a total of 189 consecutive children (ages 3 months to 16 years, median age 8 years) who underwent CSF investigation for their suspected inflammatory neurologic condition at a tertiary pediatric neurology center had CSF and serum testing to (1) qualitatively identify OCB patterns type 1–5 by isoelectric focusing on agarose gels, followed by immunoblotting as previously described; and (2) quantitatively measure the IgG index and QAlb, as previously reported. If multiple samples were tested (n = 11), results from the first sample were reported and used in analysis. CSF IgG analysis was not used in designating the classification of the patients’ diagnosis. In our institution, the investigations protocol for a child with a suspected inflammatory disorder includes both qualitative and quantitative CSF Ig analysis.

Patient case notes were retrospectively reviewed (Y.H., R.S., V.F.) and patients were classified (Y.H., M.A., M.L.) using the ICD-10 nervous system (V.F.) and patients were classified (Y.H., M.A., M.L.) using the Pediatric MS Study Group criteria into monophasic acquired demyelinating syndromes (acute disseminated encephalomyelitis, optic neuritis, transverse myelitis, or other clinically isolated syndrome) and relapsing phenotypes. Patients with autoimmune encephalopathies were subdivided into those with a known neuronal autoantibody and those with probable clinical diagnosis, as previously described. All patients with a diagnosis of CNS infection had the relevant serum and CSF investigations to confirm the diagnosis. Descriptive statistics were used to summarize the key components of patient data. Fisher exact (2-tailed) and Mann-Whitney tests were used for comparison of clinical details and investigatory findings in the different groups.

Standard protocol approvals, registrations, and patient consents. Institutional approval was obtained to perform this evaluation as a clinic audit (Ref 1782).

RESULTS The qualitative and quantitative CSF and sera IgG abnormalities and QAlb are reported in table 1, stratified by presence of a primary inflammatory pathology. Patients with an inflammatory condition were significantly more likely to have abnormal results (51/104 [49%] compared to 26/85 [31%]; p = 0.012, Fisher exact test). Abnormal qualitative results were identified in 33/104 (32%) and 9/85 (11%), respectively (p = 0.0007, Fisher exact test). Intrathecal CSF OCBs (types 2 + 3) were found in 20/104 (19%) (type 2, n = 10; type 3, n = 10) of the inflammatory group compared with 4/85 (5%) (type 2, n = 3; type 3, n = 1) of the noninflammatory group (p = 0.0036, Fisher exact test). Mirrored OCBs (type 4) were found in 13/104 (12.5%) of the inflammatory group compared with 5/85 (6%) of the noninflammatory group (p = 0.14, Fisher exact test). The quantitative analyses are reported in table 2 and illustrated in the figure. The absolute measures of IgG index and QAlb are significantly higher in patients with an inflammatory etiology (table 2, p = 0.048 and p = 0.0098, respectively; Mann-Whitney test). Raised IgG index (>0.85) was seen in both groups (table 2), whereas QAlb (>0.049) abnormalities were seen more frequently in the inflammatory group (p = 0.0028, Fisher exact test). The qualitative and quantitative evaluation collectively revealed additional positive results than when done in isolation (figure 1, A and B). Overall, the likelihood ratio of a positive test (type 2–4 pattern vs type 1 pattern) to help differentiate inflammatory from noninflammatory causes was 3.1 (95% confidence interval 1.6–6.0).

DISCUSSION In this study of 189 children, both quantitative and qualitative analyses were informative in identifying inflammatory mechanisms. Intrathecal OCBs were present in 32% of patients with inflammatory CNS disorders. OCBs were also identified in the noninflammatory group, albeit at a lower frequency (11%). Four patients had type 2 and 3 OCBs in the noninflammatory group (table 1). All these patients did not have features atypical to their diagnosis. These data are similar to those reported by Sinclair et al., who identified intrathecal OCBs in 11/58 (19%) of patients with inflammatory conditions and in none of 142 patients with noninflammatory conditions. In contrast, we additionally identified 11 patients (predominantly in the inflammatory group) with a combination mirrored and intrathecal OCB pattern (type 3).

To date, our study is the first contemporary cohort to evaluate the utility of the quantitative analysis in a range

| Comparison of quantitative results between the inflammatory and noninflammatory groups additionally subdivided into patients with normal and abnormal oligoclonal band pattern |
|-----------------|-----------------|-----------------|
| Age, y          | Inflammatory (n = 105) | Noninflammatory (n = 85) | p Value |
| IgG index (n = 77, 54) | 0.71 (0.11–7.16) | 0.62 (0.13–4.94) | 0.0484 |
| IgG index >0.85  | 21/77 (27)       | 14/54 (26)       | NS     |
| QAlb >4.9 × 10−5 (n = 83, 63) | 0.0033 (0.00035–0.046) | 0.0026 (0.00032–0.03) | 0.0098 |
| Isolated OCB only (type 2 + 3) (normal IgG index, QAlb) | 4/20 (20) | 2/9 (22) | NS     |
| Isolated OCB only (type 4) (normal IgG index, QAlb) | 4/13 (31) | 1/5 (20) | NS     |

Abbreviations: Ig = immunoglobulin; NS = nonsignificant; OCB = oligoclonal band; QAlb = albumin quotient. Data are median (range) or n/N (%).
of childhood neurologic disorders. Raised IgG index in isolation was not found in significantly higher incidence in the inflammatory group and did not have specificity for a specific diagnosis or disease group, whereas raised QAlb was seen more frequently in patients with inflammatory conditions. Nevertheless, IgG index and QAlb were raised in isolation (with normal OCB pattern) in 16/35 (46%) and 24/34 (71%) patients, respectively. This additional information, although identified in both groups, is clinically relevant. Because systemically derived albumin gets into the CSF through passive diffusion and cannot be produced in the CNS, the QAlb is a useful method to assess the blood-CSF barrier and, when raised, indicates barrier impairment. In contrast, raised Ig levels in the CSF may be secondary to either blood-brain barrier disruption or intrathecal synthesis; therefore, the IgG index is a more sensitive measure of focal IgG production as it adjusts for IgG leakage due to barrier dysfunction and serum albumin and IgG concentration. Quantitative analysis may also be useful when there is significant damage or impairment to the blood-brain barrier and the OCB patterns may be masked by the background of other proteins.

It is important to acknowledge that our cohort has inherent biases. As this was a pragmatic clinical study, the higher representation of some of the immune-mediated conditions may simply reflect the lengths physicians go to investigate them, in contrast to conditions where the clinical diagnosis is based on good diagnostic criteria, such as MS, or a disease biomarker, such as the autoimmune encephalitides. Sometimes patients with incomplete investigations performed prior to referral would not have a repeat CSF analysis and were subsequently excluded from the study. Thus, our study may not reliably evaluate for specific disorders, such as MS, that are more susceptible to these influencing factors. Nevertheless, both quantitative and qualitative analyses of CSF are useful in identifying the group of inflammatory disorders.

The early neurologic features of inflammatory disorders can be indistinguishable from a range of other “noninflammatory” disorders; CSF analysis would have been performed in patients who are subsequently diagnosed with a range of these conditions, as in reflected in our cohort. However, this has provided important information on the possible inflammatory mechanisms in these disorders. In our study, both qualitative and quantitative abnormalities were seen in patients with noninflammatory disorders, in keeping with previous reports of immune and inflammatory mechanisms being implicated in a broad range of neurodegenerative disorders and epilepsies. Hence, when evaluating the diagnostic accuracy of such an analysis that identifies an inflammatory mechanism, care needs to be taken when CNS disorders are classified only according to their primary pathology.

Taken together, our data support the use of OCB analysis in pediatric inflammatory neurologic conditions, as has been previously reported in other pediatric and adult cohorts. However, both the IgG index and QAlb can be additionally informative and should be performed as a standard evaluation as they may also have utility in monitoring inflammatory mechanisms and therapy response. Further studies should evaluate the additional utility of these biomarkers, either individually or in combination with other markers of inflammation such as CSF reactivity, imaging, and CSF neopterins, in evaluating inflammatory conditions and/or mechanisms.

**AUTHOR CONTRIBUTIONS**

Yael Hacohen: drafting/revising the manuscript, study concept or design, analysis or interpretation of data, acquisition of data. Rahul Singh: acquisition of data, drafting/revising the manuscript. Vhari Forsyth: acquisition
of data, drafting/revising the manuscript. Michael Absoud: drafting/revising the manuscript, study concept or design, statistical analysis, analysis or interpretation of data. Ming Lim: drafting/revising the manuscript, study concept or design, study supervision.

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