Research Article

Survival of Viral Biowarfare Agents in Disinfected Waters

Mary Margaret Wade, 1 Amanda E. Chambers, 1 Joseph M. Insalaco, 2 and Alan W. Zulich 1

1 Edgewood Chemical Biological Center, U.S. Army, RDECOM, Edgewood Area, Aberdeen Proving Ground, Aberdeen, MD 21010, USA
2 Science Applications International Corporation, P.O. Box 68, Gunpowder Branch, Aberdeen, MD 21010, USA

Correspondence should be addressed to Mary Margaret Wade, mary.m.wade@us.army.mil

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1. Introduction

Since the tragic events of 2001, concern among many government agencies has risen with regard to protecting the nation’s critical water infrastructure. And although chlorination is commonly used in the U.S. for disinfecting drinking water [1, 2], little data currently exists with regard to the persistence of biowarfare agents in chlorinated water, particularly viral biowarfare agents. Studies have been performed examining chlorine inactivation of bacterial biothreat agents [3–5], but few studies have examined free chlorine inactivation of viral biothreat agents, such as those that cause viral hemorrhagic fevers, viral encephalitis, or smallpox. One study examining the survival of the vaccine strain of Venezuelan equine encephalitis (VEE) virus in liquids reported that VEE strain TC-83 each separately to a final concentration of approximately 1 × 106 PFU/mL, and survival was assessed by plaque assay. Both viruses were inactivated by 1 mg/L free available chlorine (FAC) and 2 mg/L total bromine within one hour. In conclusion, these results demonstrate that both chlorine and bromine are effective disinfectants against vaccinia virus and VEE strain TC-83 at the concentrations tested.

1.1. Formulated Tap Water Preparation.

Formulated tap water was prepared using ASTM Type I deionized water and restored to 4°C with a shelf life of 6 months (see Table 1 for list of stock solutions). Once stock solutions were prepared, approximately 500 mL of ASTM Type I deionized...
Table 1: Preparation of formulated tap water.

| Chemical        | Concentration of Stock (mg/liter) | Amount of Stock added (mL) | Final concentration (mg/liter) |
|-----------------|-----------------------------------|---------------------------|-------------------------------|
| NaHCO₃          | 10,000                            | 10.0                      | 100                           |
| MgSO₄·7H₂O      | 1,000                             | 13.4                      | 13.4                          |
| K₂HPO₄          | 1,000                             | 0.7                       | 0.700                         |
| KH₂PO₄          | 1,000                             | 0.3                       | 0.300                         |
| (NH₄)₂SO₄       | 100                               | 0.1                       | 0.0100                        |
| NaCl            | 100                               | 0.1                       | 0.0100                        |
| FeSO₄·7H₂O      | 10.0                              | 0.1                       | 0.001                         |
| NaNO₃           | 1,000                             | 1.0                       | 1.00                          |
| CaSO₄           | 1,000                             | 27.0                      | 27.0                          |
| Humic acid      | 1,000                             | 1.0                       | 1.00                          |
| Fulvic acid     | 1,000                             | 1.0                       | 1.00                          |

(a) IHSS Suwannee River Humic Acid Standard, Cat. No. 1S101H.
(b) IHSS Suwannee River Fulvic Acid Standard, Cat. No. 1S101F.

2.5. Cell Lines and Virus Source. Vero cells (CCL-81), BSC-40 cells (CRL-2761), and BHK-21 cells (CCL-10) were obtained from ATCC (Manassas, VA). Vaccinia virus strain WR and Venezuelan Equine Encephalitis virus strain TC-83 (VEE TC-83) were also obtained from ATCC. Vaccinia virus served as a biosafety level 2 surrogate for Variola major (smallpox) and the vaccine strain of VEE (TC-83) also served as a surrogate for virulent VEE. All cells were grown at 37°C with 5% CO₂ in Dulbecco’s Modified Eagle’s Medium (DMEM) containing 10% heat-inactivated fetal bovine serum (FBS). Cells were passed on a twice weekly basis, and media were replaced daily.

2.6. Vaccinia Propagation. Vaccinia virus was propagated in BHK-21 monolayer cultures at 37°C. Cells were infected with virus for 1 hour in DMEM at 37°C. Following the absorption period, media were removed and replaced with fresh DMEM containing 10% FBS. Infected cells were harvested 48 hours postinfection and centrifuged at 650 × g at 4°C for 10 minutes. Pellets were resuspended in cell culture medium, freeze-thawed for three cycles, sonicated for four minutes on ice, and centrifuged at 650 × g at 4°C for 10 minutes. The supernatant served as the source for virus. The resulting virus served as the source of vaccinia for all experiments described below.

2.7. VEE Propagation. VEE strain TC-83 virus was propagated in Vero monolayer cultures at 37°C. Cells were infected with virus for one hour in DMEM at 37°C. Following the absorption period, media were removed and replaced with DMEM containing 10% FBS at 37°C. Infected cells were harvested and centrifuged at 650 × g at 4°C for 10 minutes. The supernatant was saved and stored at –80°C and served as the source of VEE strain TC-83 for all experiments described below.

2.8. Plaque Assays. BSC-40 cells (host of vaccinia virus) or Vero cells (host of VEE TC-83) were plated at a density of 3.0 × 10⁵ cells per well in twelve-well plates. Cells were allowed to reach confluence overnight at 37°C in DMEM containing 10% FBS. Prior to dilution, the vaccinia stock was sonicated on ice for 30 seconds. The virus was serially diluted from 1:10² to 1:10⁹ in DMEM. The medium was removed from the cells, and the diluted virus was added. Virus absorption was for one hour at 37°C with occasional shaking. Following absorption, medium was removed and replaced with minimal essential medium (MEM) containing 5% FBS and 1% SeaPlaque Agarose. Forty-eight hours after
infection, the cells were fixed in 7% formaldehyde for one hour. The agarose layer was removed, and cells were fixed for an additional hour in 7% formaldehyde. After removing the formaldehyde, plaques were visualized by staining with 0.01% crystal violet for 30 minutes. The plaques were counted, and the results were reported in plaque forming units per milliliter (pfu/mL).

2.9. Inoculation, Incubation, and Sampling. Survival of vaccinia virus strain WR and VEE TC-83 in chlorinated and brominated water (preparation of each described above) was determined by inoculating each water matrix with virus to a final concentration of approximately \(1 \times 10^6\) PFU/mL. At various time intervals, 1 mL of the spiked water sample was removed and sodium thiosulfate was added to a final concentration of 0.005% to quench any remaining disinfectant (Sigma Chemical Company, St. Louis, MO). The samples were serial diluted, and plaque assay was performed as described above. All water matrices tested were filter sterilized prior to inoculation. All samples were incubated at room temperature (21°C) after inoculation, and each virus was tested separately in each water. Military relevant time points were selected per guidance from the funding agency.

3. Results

Survival of vaccinia virus strain WR and VEE strain TC-83 in both chlorinated and brominated water over time is presented in Figures 1 and 2, respectively. All data is presented as the number of PFU/mL recovered over time by plaque assay. As shown in Figures 1 and 2, both viruses persisted in positive control samples, which consisted of formulated tap water without disinfectant, with no decrease in viability for the time points tested. However, in the presence of 1 mg/L FAC and 2 mg/L total bromine, neither virus was infectious at the earliest time point tested of one hour or at any subsequent time points tested.

4. Discussion

Although disinfection of water supplies is common practice in the U.S., limited data is currently available with regard to the length of time viral biothreat agents can survive in those waters. Both chlorination and bromination of water are practiced by branches of the military and therefore both were included at military relevant concentrations in the present study in order to ascertain their effectiveness as disinfectants against selected viruses. Survival of vaccinia virus strain WR and VEE strain TC-83 in formulated tap water with bromine or chlorine was monitored over time at room temperature using plaque assay to assess infectivity of the virus. Both disinfectants proved to be effective sanitizers against the viral biowarfare agents tested in a minimal amount of time making this study one of the first to report the survival of viral biothreat agents in chlorinated, and, moreover, brominated water.

The U.S. Army and other government entities, in an effort to protect national and military water supplies, are placing great emphasis on developing rapid detection and identification technologies for biological agents in water. However, based on data provided in this study, monitoring chlorine or bromine levels in water supplies to ensure
that adequate levels of disinfectant are present could prove sufficient for maintaining safe water supplies. Additional testing with a greater number of agents will be required before this decision can be made, and these tests are already underway to evaluate whether additional agents are easily killed by chlorination and bromination.

Acknowledgment

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References

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