Nationwide distribution of varicella-zoster virus clades in China

Songtao Xu1†, Mukai Chen2†, Huanying Zheng3†, Haiyan Wang4†, Meng Chen5, Jianhui Zhou6, Wang Shuang6, Pengbo Yu7, Chaofeng Ma8, Jian He9, Daxing Feng10, Zhu Zhen1, Zhang Yan1, Mao Naiying1, Aili Cui1, Qiuhua Wu1, Mengyuan Qi11, Chongshan Li12*, Xiaoguang Xu11* and Wenbo Xu1*

Abstract

Background: In 2010, a universal nomenclature for varicella-zoster virus (VZV) clades was established, which is very useful in the monitoring of viral evolution, recombination, spread and genetic diversity. Currently, information about VZV clades has been disclosed worldwide, however, there are limited data regarding the characterization of circulating VZV clades in China, even where varicella remains widely epidemic.

Methods: From 2008 to 2012, clinical samples with varicella or zoster were collected in General Hospital in eight provinces and analyzed by PCR, restriction endonuclease digestion and sequencing. The viral clades were determined by analysis of five single nucleotide polymorphisms (SNPs) within the 447-bp fragment of open reading frame (ORF) 22, and the restriction fragment length polymorphisms (RFLPs) of ORF 38 (PstI), ORF 54 (BglII) and ORF 62 (SmaI) were evaluated to understand genetic diversity of VZV and determinate varicella vaccine adverse event (VVAE).

Results: Seventy-seven varicella and 11 zoster samples were identified as being positive for VZV. The five SNPs profile showed that the majority of VZV strains belonged to clade 2, but clade 5 and clade 4 strains were also found in Guangdong. The RFLPs analysis of the DNA fragments of ORF 38, 54 and 62 showed that 85 of these samples were characterized as PstI+ BglII- SmaI-, and the remaining three VZV strains from varicella patients were characterized as PstI- BglII+ SmaI+ which is the genetic profile of VVAEs.

Conclusions: The study suggested that the predominant clade 2 VZVs had been continually circulating since at least the 1950s in China. Nearly all VZV strains except VVAEs possessed the genetic profile of PstI + BglII + SmaI-. However, the other clades were also found to be co-circulating with clade 2, especially in the border regions. These results highlighted the need for the constant and broad use of virologic surveillance to provide an important genetic baseline for varicella control and vaccination programs in China.

Keywords: Varicella-zoster virus, Clade, Varicella vaccine adverse event, Genetic profile, China
Background

Varicella-zoster virus (VZV) is highly contagious and causes two diseases. The primary infection results in varicella (chickenpox), which usually occurs early in life. Subsequently, the virus establishes a lifelong latent infection in the sensory nerve ganglia, which reactivates to cause zoster (shingles) under conditions of declining immunity that are most commonly associated age [1].

Genotyping of VZV contributes to reveal viral diversity, recombination, evolution patterns and transmission pathways. To achieve these goals, several groups have published different schemes by using molecular techniques such as the sequencing, restriction fragment length polymorphisms (RFLPs) and single nucleotide polymorphisms (SNPs) since the 1990s, however, the different nomenclature methods will not facilitate interchange of information [2–7]. In 2010, Breuer et al. summarized the previous nomenclature methods and proposed a novel nomenclature for VZV: clade 1–5 and two putative clades (VI and VII). The new universal nomenclature will be useful for the interchange and comparison of genotyping data worldwide [6, 8, 9].

The universal childhood varicella vaccination program was initiated in the United States (US) in 1995 [10, 11]. Currently, although varicella vaccines are licensed and available throughout the world, the varicella vaccination has been recommended as a universal childhood immunization in only a small number of countries including Canada, Germany, Australia, Korea and Japan, where high coverage rates have been attained, and vaccination has resulted in a significant decline in varicella-related incidence, morbidity and mortality [12, 13]. Due to the absence of varicella vaccination programs in most countries, varicella remains a worldwide epidemic and was estimated that the global varicella disease burden includes 4.2 million severe cases and 4200 deaths annually [14]. In China, the varicella vaccine is not included in the national immunization program, which results in poor varicella control and epidemic conditions, but it's available for purchase on private basis for vaccination of children. However, application of the live-attenuated varicella vaccine may cause varicella vaccine adverse events (VVAEs). Surveillance of the VVAE plays a key role in evaluating side effects of the vaccine and helps physicians in appropriate decision making.

This study describes the clade distribution and genetic profile of circulating VZV strains throughout China, and distinguishes vaccine strains from wild-type strains. These data provide important genetic background that will facilitate efforts for controlling varicella and zoster in China in the future.

Methods

Ethics statement

This study was approved by the Ethics Committee of National Institute for Viral Disease Control and Prevention, China Center for Disease Control and Prevention. Individual written informed consent was obtained from the parents or guardians of all participants.

Clinical samples

From 2008 to 2012, the suspected varicella or zoster samples were collected from outpatients who were clinically diagnosed with varicella or zoster in General Hospitals. These patients stayed residentially in eight provinces (Beijing, Jilin, Guangdong, Henan, Shaanxi, Gansu, Sichuan, and Shandong) located in eastern, southern, western, northern and central of mainland China. These provinces cross subtropical (Guangdong) and temperate climates (the other provinces) and should be sufficiently representative (Fig. 1).

Viral DNA extraction and polymerase chain reaction (PCR)

Viral DNA was extracted from 200 μl vesicle fluid or throat swab samples using the QIAamp DNA Mini Kit (Qiagen, Valencia, CA). To characterize viral clades and distinguish vaccine strains from wide-type strains, the PCR amplification of ORF 22, 38, 54 and 62 were performed in accordance with previously described protocols [15–18]. One Chinese vaccine strain and one wide type strain were used for the positive controls, the DNase-free water was used for the negative control. The 447-bp region of ORF 22 (Strain Dumas, Genbank No. X04370) encompassing five SNPs (37902, 38019, 38055, 38081, and 38177) was sufficient to characterize between viral clades [2, 6]. However, discrimination between clades 1 and 3 required the addition of a sixth SNP (39394) to differentiate between the two clades. The genetic markers of VZV that are most commonly considered in vaccine adverse events and epidemiological studies include the RFLPs of ORFs 38 (PstI), 54 (BglII) and 22 (SmaI) [16, 19]. The PCR assays were performed using the PCR amplification kit (Takara, Dalian, China) according to the manufacturer’s instruction. The template amplifications were performed using a GeneAmp 9700 thermocycler (Applied Biosystems, Grand Island, NY, USA). The PCR products were purified using a QIAquick Gel Extraction Kit (Qiagen) and evaluated by electrophoretic separation on 2% agarose gels.

Sequencing and restriction enzyme reactions

The 447-bp region of ORF 22 were sequenced using an ABI PRISM 3100 genetic analyzer (Applied Biosystems, Hitachi, Japan) according to the manufacturer’s instruction, and the sequence data were assembled and edited using Sequencher software version 4.0.5 (GeneCodes,
Ann Arbor, MI). The five SNPs were analyzed for VZV clades identification using BioEdit version 7.0 (Tom Hall, North Carolina State University, Raleigh, NC). Restriction endonuclease digestion of the PCR products (ORF38, ORF54 and ORF62) was performed with 8 μl of the PCR products, 15 U endonuclease (PstI, BglII and SmaI) (Takara, Dalian, China), and 2 μl of the accompanying 10× endonuclease buffer. The final reaction volume was adjusted to 20 μl with DNase- and RNase-free water. The analysis of RFLPs was performed according to the protocols previously described in the literature [2, 16].

**Results**

**Case confirmation**

From 2008 to 2012, a total of 126 clinical samples were collected in eight provinces, including 115 throat swabs from 115 varicella patients and 11 vesicular fluids from eleven zoster patients. All patients were born and spent their childhoods in China. No information was available about the VZV exposure history including varicella vaccination, except for five patients with mild varicella within 3 weeks after varicella vaccination. The ages of 115 patients with varicella ranged between 3 and 25 years (mean age 7.5 years); the ages of 11 patients with zoster ranged between 40 and 68 years (mean age 50 years).

A total 88 of 126 samples were tested VZV positive using the PCR screening of ORF 22 (447-bp), including 77 throat swabs from varicella patients and all 11 vesicular fluids from zoster patients. The throat swabs from three of the five varicella patients with mild varicella after varicella vaccination were also positive.

**Viral clade determination**

The 447-bp fragment (ORF 22) of 88 VZV samples was successfully sequenced, the five SNPs profile (37902: G, 38019: G, 38055: C, 38081: C and 38177: A) disclosed that most of endemic VZV strains were attributed to their childhoods in China. No information was available about the VZV exposure history including varicella vaccination, except for five patients with mild varicella within 3 weeks after varicella vaccination. The ages of 115 patients with varicella ranged between 3 and 25 years (mean age 7.5 years); the ages of 11 patients with zoster ranged between 40 and 68 years (mean age 50 years).

A total 88 of 126 samples were tested VZV positive using the PCR screening of ORF 22 (447-bp), including 77 throat swabs from varicella patients and all 11 vesicular fluids from zoster patients. The throat swabs from three of the five varicella patients with mild varicella after varicella vaccination were also positive.

**Viral clade determination**

The 447-bp fragment (ORF 22) of 88 VZV samples was successfully sequenced, the five SNPs profile (37902: G, 38019: G, 38055: C, 38081: C and 38177: A) disclosed that most of endemic VZV strains were attributed to
clade 2; however, one clade 5 (37902: A, 38019: G, 38055: T, 38081: C and 38177: G) strain from a 7-year-old child and one clade 4 (37902: A, 38019: A, 38055: C, 38081: C and 38177: A) strain from a 64-year-old elder with zoster were found in Guangdong in 2009. The analysis of the sixth SNP (in the position 39394) was not performed due to the lack of clade 1 or 3 strains (Table 1).

Distinguishing vaccine strains from wild-type of VZV strains

For these positive VZV strains, three PCR amplifications of ORF 38 (350-bp), 54 (222-bp) and 62 (268-bp) were performed to distinguish vaccine strains from wild-type strains. The three target bands were obtained for each of the samples and digested with PstI, BglI and SmaI enzymes, respectively. The 350-bp amplification products were cleaved to the 250 bp and 100 bp fragments by PstI in most of the samples, except for three samples that were collected from the Henan and Shaanxi provinces. The 222-bp amplification products were cleaved to 137 bp and 85 bp by BglI in all of the samples. The two genetic markers that were characterized in the most VZV strains possessed the PstI + BglI + pattern, with the exception of 3 samples (PstI-BglI+). For the 268-bp fragment, most VZV samples were cleaved to 153-bp, 79-bp, and 36 bp (SmaI-) except the 3 strains that were cleaved to 112-bp, 79-bp, 41-bp and 36 bp (SmaI+). The results of the analysis of the three genetic makers indicated that most of the VZVs belonged to wild-type strains, whereas the three VZV strains from patients within 3 weeks after varicella vaccination possessed the genetic profile (PstI-BglI + SmaI+) of the Oka varicella vaccine (Table 1).

**Table 1** Genetic profile of eighty-eight VZV strains in eight provinces

| Province       | Collection time | No. of V or Z | Clade | Residue at ORF 22 position | ORF38 (PstI) | ORF54 (BglI) | ORF62 (SmaI) |
|----------------|-----------------|---------------|-------|----------------------------|-------------|-------------|-------------|
| Guangdong 2008–2010 | V(21), Z(10)    | 2  G G C C A | +     | +                         | -           | -           |             |
| Guangdong 2009    | V(1)            | 4  A A C C A | +     | +                         | -           | -           |             |
| Guangdong 2009    | Z(1)            | 5  A G T C G | +     | +                         | -           | -           |             |
| Sichuan 2009      | V(9)            | 2  G G C C A | +     | +                         | -           | -           |             |
| Beijing 2009      | V(18)           | 2  G G C C A | +     | +                         | -           | -           |             |
| Henan 2010        | V(1)            | 2  G G C C A | +     | +                         | -           | -           |             |
| Henan 2010        | V(2)            | 2  G G C C A | -     | +                         | +           | +           |             |
| Shaanxi 2012      | V(1)            | 2  G G C C A | -     | +                         | +           | +           |             |
| Shandong 2012     | V(2)            | 2  G G C C A | +     | +                         | -           | -           |             |
| Gansu 2012        | V(7)            | 2  G G C C A | +     | +                         | -           | -           |             |
| Jilin 2011–2012   | V(15)           | 2  G G C C A | +     | +                         | -           | -           |             |

1. V varicella; Z herpes zoster
2. All genome positions are based on the published sequence for the VZV reference strain: Dumas (Clade 1; Accession # XO4370)
3. Numbers in parenthesis indicate the number of varicella and/or zoster strains
4. *in right corner indicates the VVAEs

**Discussion**

Information about the molecular epidemiology of VZV has been proven very useful for characterizing endemic strains, tracing transmission pathways, and distinguishing vaccine strains from wild-type strains. In 2010, Breuer proposed a novel nomenclature for VZV: clade 1–5 and two putative clades (VI and VII) that were identified by Roman numerals until confirmed to be a clade. The new universal nomenclature will be useful for the interchange and comparison of genotyping data worldwide [6]. According to the universal nomenclature, the worldwide distribution of VZV clades has been established. Clade 1 and clade 3 represent the dominate clades of the circulating VZV in Europe and the Americas, where the clade 4 and 5 strains are frequently identified due to the immigration of people with African origin. The clade VI strains were also found in France (10 %) and Italy (11 %), and one might speculate that clade VI strains are more common than clade 3 strains in Southern Europe [20]. To date, only one clade VII strain was exclusively isolated in the US in 2002 [21]; in Asia, VZV clade 2 has been the dominant clade according to previous publications from Korea, Japan, and China; however, clade 2 strains were not found circulating in India, Nepal and Bangladesh, where clade 4 and 5 strains were dominant [2, 22–24].

Several previous publications (mostly in Chinese) have disclosed information about the genotypes and genetic profiles of VZV, but all of them focused on one province or prefecture and were limited [2, 23, 25–32]. Overall, 420 VZV strains collected from 115 varicella patients and 305 zoster patients were characterized, which were detected during the period from 2000 to 2015 in 13 provinces (Table 2). In this study, 88 VZV strains collected from
Table 2 Clade summary of the VZV strains isolated in mainland China according to previous publications

| Province        | Collection time | No. of V or Z | Clade       | References |
|-----------------|-----------------|---------------|-------------|------------|
| Guangdong       | before 2000     | V (3)         | 4           | [2]        |
| Beijing         | before 2000     | V (3)         | 1 or 3      | [2]        |
| Shanghai        | 2007            | V (8)         | 2           | [25, 26]   |
| Hubei           | 2008            | V (16), Z (80)| 2           | [32]       |
| Anhui           | 2007–2008       | V (2), Z (17)| 2           | [23]       |
| Liaoning        | 2009            | V (3)         | 2           | [33]       |
| Inner Mongolia  | 2010            | V (3)         | 2           | [28]       |
| Zhejiang        | 2009–2011       | V (25)        | 2 (23), 5   | [37]       |
| Fujian          | 2010–2011       | V (10), Z (34)| 2           | [41]       |
| Tibet           | 2011            | V (6), Z (4) | 5 (3), 4 (4), 2 (3) | [27] |
| Xinjiang        | 2012–2013       | V (4), Z (116)| 2 (79), 5 | [30]       |
| Shanxi          | 2013            | V (12)        | 2           | [31]       |
| Hunan           | 2013–2015       | V (20), Z (54)| 2           | [29]       |

1. V varicella; Z herpes zoster
2. Numbers in parenthesis indicate the number of varicella and/or zoster strains

eight provinces where no VZV investigation was reported were characterized (Table 1). With the incorporation of previously published data, this study contributes towards the knowledge of the molecular epidemiology profile of prevalent VZVs circulating in mainland China.

In northern China (Jilin, Liaoning, Beijing, Shanxi and Inner Mongolian provinces), all clinical samples from varicella patients since 2009 belonged to clade 2 [28, 31, 33]. However, three clade 1 or 3 VZV strains circulating before 2000 were identified in Beijing by Loparev [2]. We speculate that the three clade 1 or 3 strains, which were either imported or random cases, were not the predominate clades in light of the current studies. In western China, 120 VZV strains including four varicella and 116 zoster samples were described in Xinjiang. The results showed that four varicella strains belonged to clade 2 and 116 zoster strains were distributed into 3 clades: 75 in clade 2, 40 in clade 5, and one in clade 1 or 3. Geography might cause multiple clade strains to co-circulate because it borders Gansu province, where clade 2 strains predominate. Additionally, Southern Xinjiang borders Pakistan and India, where clade 5 strains have been circulating [34, 35]. Northern Xinjiang borders Russia, where the clade 1 and 3 predominate [36]. The genetic profiles of the RFLPs were characterized as 120 PstI+, 119 BglII+, one BglII- and 120 SmaI-, which indicated that the genetic profile of most strains was PstI + BglII+, except for one PstI + BglII- strain which is the genetic profile of strains from Europe and North America [30]. In Tibet, ten positive VZV strains were identified and belonged to three clades, three in clade 2, three in clade 1 or 3, and four in clade 5, which is probably due to the special geographical location, as it borders Pakistan, India and Nepal, where clades 1 or 3 and clade 5 predominate [2]. However, information about VZV clades is currently unavailable for Yunnan which is also a very special location because it borders Myanmar, Vietnam and Laos where the characterization of the VZV clade has never been reported. The strengthening of the virologic surveillance is critical in Yunnan in the near future. In southern China (Guangdong, Hunan and Hubei provinces), a total of 206 positive VZV strains, those isolated from 141 varicella and 65 zoster cases, were identified. All of them belonged to clade 2, except for one, which belonged to clade 5, and one that belonged to clade 4 and was found in Guangdong. The clade 5 strain was isolated from a 7-year-old varicella patient. The clade 4 strain was isolated from a 63-year-old zoster patient. Loparev et al. also published that three clade 4 strains circulating before the year 2000 were found in Guangdong [2]. These results suggest that the minority clade 4 and 5 strains might have been circulating in Guangdong for a long time. This situation might result from frequent international communication, especially among the hundreds of thousands of Africans residing in Guangdong. However, this should be further validated by studies with large sample numbers in the future. In eastern China (Shandong, Anhui, Shanghai, Zhejiang, and Fujian provinces), the clade characterization had been already highlighted, and all VZV strains from varicella or zoster patients belonged to clade 2 except two imported clade 5 strains based on the epidemiological survey in Zhejiang (Tables 1 and 2) (Fig. 1) [37, 38].

Primary infection with VZV usually occurs during childhood, at approximately 10 years of age [39]. Previously published data have shown that the strains of virus causing zoster in a patient are identical to the primary varicella strain that infected that patient [5]. The current study shows that although other clades of VZV strains were found to co-circulate in the border regions such as Xinjiang, Tibet and Guangdong, the majority of VZV strains from zoster patients belonged to clade 2. The oldest patient was 68 years old, which indicated that the clade 2 VZV strains had been continually circulating in China since at least the 1950s.

Since the varicella vaccine is a live attenuated vaccine, varicella-like rash after vaccination is rare but can occur and is defined as a varicella vaccine adverse event (VVAE), an increasingly common side effect of varicella vaccination. The data of the Vaccine Adverse Event Reporting System of the U.S. showed the mild varicella-like rash would occur in 3 to 5 % of varicella vaccine recipients [5]. The RFLP of a variable region is a useful
tool to better understand VZV genetic characterization and determination of varicella or zoster vaccine adverse event. Previously RFLPs analysis showed that the majority of wild-type strains from Europe and North America (temperate climate) were PstI+ BglII−. Wild-type strains in Africa, Asia or the Caribbean (tropical and subtropical climates) were PstI+ BglII+, whereas the Oka vaccine strain (vOka) was PstI-BglII+, and the Japanese Oka-like wild-type strains were PstI-BglII+/PstI + BglII+, which indicates that the RFLP analysis of the PstI and BglII failed to distinguish wild-type strains from Oka vaccine strains [8, 16, 18]. In 2000, RFLP analysis of the SmaI in ORF 62 was developed, which had been used to distinguish vaccine strains from the wild-type strains. This marker has been shown to be stable and can be confidently used to identify vaccine strains [16]. However, with the identification of more unique SNPs within the vaccine virus, it may be preferable to sequence the SNPs in order to identify vaccine strains [9]. In this study, three VVAEs from varicella patients in Shaanxi and Henan were identified according to the genetic profile of RFLPs: PstI-BglII+ SmaI+ [16, 40], which is limited report about the VVAEs in China until now. Although the varicella vaccine is not included in the national immunization program in China, with the improvement of prevention awareness of varicella, more children will be vaccinated, which could result in more VVAEs. Therefore, it is very important to enhance the ability to determine VVAEs that will enable physicians to make informed decisions and to relieve the burden of disease in patients.

Conclusions

In conclusion, this study confirms that VZV clade 2 represented the dominant clade nationwide and has been continuously circulating in China since at least the 1950s. There were other clade strains co-circulating with clade 2 in the border regions. The genetic profile of the 3 RFLPs within ORF 38, 54 and 62 was usually PstI + BglII + SmaI−, except for three VVAEs (PstI-BglII + SmaI+). This report identified the clade distribution of the circulating VZV strains throughout China and determined the VVAEs. These results emphasize the identification of VVAEs and the importance of a more systematic collection of VZV strains, especially in the border regions, to propose more effective control strategies and to establish an accurate genetic baseline for VZVs in China in the near future.

Abbreviations

ORF: Open reading frame; RFLP: Restriction fragment length polymorphism; SNP: Single nucleotide polymorphism; WAE: Varicella vaccine adverse event; VZV: Varicella-zoster virus

Acknowledgements

We are thankful to the staff of the provincial CDCs who collected the clinical samples and provided the epidemiological data. We also thank Dr. D. Scott Schmid, Dr. Li Jin, Dr. Chris Maple, Dr. Judith Breuer and Dr. Mona Marin for their assistance with the preparation of this manuscript.

Funding

This study was supported by grants from the National Natural Science Foundation of China (project no.813302477), the Key Technologies R&D Program of National Ministry of Science (2013ZX10004-022, 2013ZX10004-101, 2012YQ03026104).

Availability of data and materials

Raw data is available upon request from the first author (Dr. Songtao Xu, National Institute for Viral Disease Control and Prevention, China Center for Disease Control and Prevention, Beijing City 102206, China, Email: xstt8868@sina.com).

Authors’ contributions

SX designed and performed the experiments, drafted and reviewed manuscript; MC, HZ, CM, JZ, WS, HW, PY, CM, JH and DF collected clinical samples; SX, MC, HZ, ZZ, ZY, MN, AC, LJ, CL, QW, MQ, XX and WX participated in study design and coordination and helped to review the manuscript; CL, XX and WX conceived of the study. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable.

Ethics approval and consent to participate

Written informed consent was obtained from the patient or his/her legal guardian. This study was approved by the Institutional Ethics Committee, National Institute for Viral Disease Control and Prevention, China Center for Disease Control and Prevention, China (Reference code: IVDC 2013-008).

Author details

1National Institute for Viral Disease Control and Prevention, China Center for Disease Control and Prevention, Beijing City 102206, China. 2The First Affiliated Hospital of Sun Yat-sen University, Guangzhou City 510080, Guangdong Province, China. 3Guangdong Center for Disease Control and Prevention, Guangzhou City 510030, Guangdong Province, China. 4Shandong Center for Disease Control and Prevention, Jinan City 250014, Shandong Province, China. 5Beijing Center for Disease Control and Prevention, Beijing City 100030, China. 6Jilin province Center for Disease Control and Prevention, Changchun City 130021, Jilin Province, China. 7Shaanxi Center for Disease Control and Prevention, Xi’an City 710012, Shaanxi Province, China. 8Xi'an city Center for Disease Control and Prevention, Xian City 710031, Shaanxi Province, China. 9Sichuan Center for Disease Control and Prevention, Chengdu City 610014, Sichuan Province, China. 10Henan Center for Disease Control and Prevention, Zhengzhou City 450006, Henan Province, China. 11Department of Neurosurgery, Second Affiliated Hospital of Dalian Medical University, Dalian 116027, Liaoning Province, China. 12Shanghai Center for Disease Control and Prevention, Shanghai City 200336, China.

Received: 26 November 2015 Accepted: 22 September 2016
Published online: 07 October 2016

References

1. Overturf GD. American Academy of Pediatrics. Committee on Infectious Diseases. Technical report: prevention of pneumococcal infections, including the use of pneumococcal conjugate and polysaccharide vaccines and antibiotic prophylaxis. Pediatrics. 2000;106(2 Pt 1):367–76.
2. Lopez VN, Gonzalez A, Deleon-Carnes M, Tipple G, Fickenscher H, Torfason EG, Schmid DS. Global identification of three major genotypes of varicella-zoster virus: longitudinal clustering and strategies for genotyping. J Virol. 2004;78(15):8349–58.
3. Muir WB, Nichols R, Breuer J. Phylogenetic analysis of varicella-zoster virus: evidence of intercontinental spread of genotypes and recombination. J Virol. 2002;76(4):1971–9.

Go back...
11. Goldman GS, King PG. Review of the United States universal varicella vaccination program: summary of VZV Nomenclature Meeting 2008, Barts and the London School of Medicine and Dentistry, 24–25 July 2008. J Gen Virol. 2010;91(Pt 4):821–8.

12. Lu L, Suo L, Li J, Zhai L, Zheng Q, Pang X, Bialek SR, Wang C. A varicella zoster virus phylogenetic analysis of a Beijing outbreak in a school with high one-dose vaccination coverage. J Virol. 2012;86(1):594–601.

13. Lu L, Wang C, Suo L, Li J, Liu W, Pang X, Seward JF. Genotype clinical isolates in Australia and New Zealand. J Virol. 2007;81(23):12850–12858.

14. Loparev VN, Rubtcova EN, Bostik V, Tzaneva V, Sauerbrei A, Robo A, Breuer J, Grose C, Norbert P, Tippels GA. A proposal for a common nomenclature for viral clades that form the species varicella-zoster virus. J Clin Microbiol. 2009;47(1):38–45.

15. Loparev VN, Argaw T, Krause PR, Takayama M, Schmid DS. Improved drug-resistant Mycobacterium tuberculosis genotype clinical isolates in Australia and New Zealand. J Virol. 2007;81(23):12850–12858.

16. Loparev VN, Argaw T, Krause PR, Takayama M, Schmid DS. Improved drug-resistant Mycobacterium tuberculosis genotype clinical isolates in Australia and New Zealand. J Virol. 2007;81(23):12850–12858.

17. Lu L, Suo L, Li J, Zhai L, Zheng Q, Pang X, Bialek SR, Wang C. A varicella outbreak in a school with high one-dose vaccination coverage. Beijing, China. Vaccine. 2012;30(34):5094–8.

18. Lu L, Wang C, Suo L, Li J, Liu W, Pang X, Seward JF. Varicella disease in Beijing in the era of voluntary vaccination, 2007 to 2010. Pediatr Infect Dis J. 2013;32(8):e314–9.

19. Lu L, Wang C, Suo L, Li J, Liu W, Pang X, Seward JF. Varicella disease in Beijing in the era of voluntary vaccination, 2007 to 2010. Pediatr Infect Dis J. 2013;32(8):e314–9.

20. WHO. Varicella and Herpes Zoster Vaccination Position Paper. 2014.

21. Loparev VN, Rubtcova E, Rubtsova EN, Bostik V, Tzaneva V, Sauerbrei A, Robo A, Breuer J, Grose C, Norbert P, Tippels GA. A proposal for a common nomenclature for viral clades that form the species varicella-zoster virus. J Clin Microbiol. 2009;47(1):38–45.

22. Loparev VN, Argaw T, Krause PR, Takayama M, Schmid DS. Improved drug-resistant Mycobacterium tuberculosis genotype clinical isolates in Australia and New Zealand. J Virol. 2007;81(23):12850–12858.

23. Liu J, Wang M, Gan L, Yang S, Chen J. Genotyping of clinical varicella-zoster virus isolates collected in China. J Clin Microbiol. 2009;47(5):1418–23.

24. Schmidt-Chanasit J, Sauerbrei A. Evolution and world-wide distribution of varicella-zoster virus clades. Infect Genet Evol. 2011;11(1):1–10.

25. Yang Jixing JL-w, et al. Research of genotype of varicella-zoster virus in Shanghai. Chin J Health Lab Technol. 2008, 18:223–225.

26. Li Chongshan LL, et al. Gene characteristics of varicella-zoster virus. Disease Surveillance. 2009;24(3):168–71.

27. Liu Lan DB-z, et al. Research on the genotypes of clinical varicella-zoster virus isolateds in Tibet. Chin J Microbiol Immunol 2012, 32(1):934–938.

28. Lu Ai-tao GW-d, et al. Genotype analysis of the pathogen of an outbreak of varicella. Prac Prev Med. 2012, 19:1302–1304.

29. Peng Yan-qian ZZJ-w, et al. The main genotype of Varicella-zoster virus circulating in Hunan. Chin Foreign Medl Res. 2015; 13(25):142–144.

30. Qu Yuan-yuan Yh, et al. Genotypic analysis of clinical varicella-zoster virus isolations in Xinjiang. Clin J Inpr Skin Dis. 2015; 51:526–530.

31. Shi Wei LJ, et al. Genotype analysis of varicella-zoster virus in Yinzge of Talyuan in 2013. J Prac Med. Techniques. 2015;22(4):352–354.

32. Zhou Xiao-yong ZZJ-l, et al. Genotype analysis of varicella-zoster virus in Wuhan. Chin J Dermatol. 2009;42(10):688–690.

33. Wang yan My, et al. Genotype analysis of varicella-zoster virus in Liaoning province. Chin J Public Health. 2012; 28(1):76–78.

34. Kaushik KS, Lahit K, Chamber SK, Gupta RM, Kumar S, Kapila K, Karade S. Molecular characterization of clinical varicella-zoster strains from India and differentiation from the oka vaccine strain. Jpn J Infect Dis. 2008;61(1):65–7.

35. Iroue H, Motani-Saito H, Sakurada K, Ikegaya H, Yajima D, Hayakawa M, Sato Y, Otuska K, Kobayashi K, Nagasawa S, et al. Determination of the geographical origin of unidentified cadavers based on geographical differences in genotype of varicella-zoster virus. J Med Virol. 2010;82(5):903–8.

36. Springfeld C, Sauerbrei A, Filusich A, Konstandin M, Hartschuh W, Sauer P, Encke J, Stremmel W, Schnitzler P. Fatal varicella in an immunocompromised adult associated with a European genotype E2 variant of varicella zoster virus. J Clin Virol. 2009;44(1):70–3.

37. Hu Feng-jiao JS-lea: Chickenpox cases monitoring and epidemic strains gene polymorphism analysis in Ningbo, Zhejiang. Inter J Epidemiol Infect Dis. 2013;4:254–257.

38. Grose C. Pargaea and the Out-of-Africa Model of Varicella-Zoster Virus Evolution and Phylogeography. J Virol. 2012;86(18):9558–65.

39. Gunnar Jr JW, Whitley RJ. Clinical practice. Herpes zoster. N Engl J Med. 2002;347(5):340–6.

40. Hondo R, Yogo Y, Yoshida M, Fujima A, Itoh S. Distribution of varicella-zoster virus strains carrying a PstI-site-less mutation in Japan and DNA change responsible for the mutation. Jpn J Exp Med. 1989;59(6):233–7.

41. Chen Ting-ting WH, et al: Genotyping of varicella-zoster virus clinical strains isolated in Fujian, China. Chin J Microbiol Immunol. 2012;32(7):614-617.