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Association between Exposure to Environmental Tobacco Smoke and Biomarkers of Oxidative Stress Among Patients Hospitalised with Acute Myocardial Infarction

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Abstract

Objective: To determine whether exposure to environmental tobacco smoke was associated with oxidative stress among patients hospitalised for acute myocardial infarction.

Design: An existing cohort study of 1,261 patients hospitalised for acute myocardial infarction.

Setting: Nine acute hospitals in Scotland.

Participants: Sixty never smokers who had been exposed to environmental tobacco smoke (admission serum cotinine ≥3.0 ng/mL) were compared with 60 never smokers who had not (admission serum cotinine ≤0.1 ng/mL).

Intervention: None.

Main outcome measures: Three biomarkers of oxidative stress (protein carbonyl, malondialdehyde (MDA) and oxidised low-density lipoprotein (ox-LDL)) were measured on admission blood samples and adjusted for potential confounders.

Results: After adjusting for baseline differences in age, sex and socioeconomic status, exposure to environmental tobacco smoke was associated with serum concentrations of both protein carbonyl (beta coefficient 7.96, 95% CI 0.76, 15.17, p = 0.031) and MDA (beta coefficient 10.57, 95% CI 4.32, 16.81, p = 0.001) but not ox-LDL (beta coefficient 2.14, 95% CI –2.84, 13.21, p = 0.703).

Conclusions: Exposure to environmental tobacco smoke was associated with increased oxidative stress. Further studies are required to explore the role of oxidative stress in the association between environmental tobacco smoke and myocardial infarction.

Introduction

Exposure to environmental tobacco smoke (ETS) is associated with an increased risk of developing cardiovascular disease. A meta-analysis, conducted in 1999, reported a pooled relative risk of 1.25 (95% CI 1.17, 1.32), and demonstrated a dose-response relationship [1]. Even brief exposure to ETS can produce changes in platelet activation and endothelium-dependent vasodilation [2] of a comparable magnitude to those observed in active smokers [3]. Side-stream smoke contains higher concentrations of toxic gases and small, respirable particles than mainstream smoke [4]. Therefore, ETS exposure carries up to 90% of the risk of cardiovascular events associated with active smoking [5].

ETS exposure is also associated with cardiovascular disease progression. In the apolipoprotein-E deficient murine model, exposure to ETS for 21 days increased atherosclerotic lesions by 76% [5], and in a general population cohort, those exposed to ETS had 20% higher progression of carotid intima media thickness over three years [5]. Animal experiments suggest that ETS exposure results in larger infarctions and an increased risk of ventricular hypertrophy [6,7], with evidence of a dose response in the former [7]. ETS is associated with prognosis following acute coronary events. In a cohort of 2,172 patients admitted for either
myocardial infarction or unstable angina, those exposed to ETS had higher concentrations of both troponin I and creatine kinase and were at higher risk of adverse events (death or rehospitalisation) within 30 days (adjusted RR 1.61, 95% CI 1.14–2.0) [8]. There was a dose-response in relation to the number of years the index cases had lived with their smoking partner [9]. We previously used data on never smokers, from an existing cohort of patients hospitalized for acute myocardial infarction, to investigate the association between ETS exposure and early adverse events [9]. ETS exposure was associated with increased risk of all-cause deaths, cardiovascular deaths, and fatal/non-fatal acute myocardial infarctions within 30 days of admission, with clear evidence of dose relationships for all three [9]. It is plausible that oxidative stress may be involved in the mechanism by which ETS exposure increases risk. Therefore, the aim of the current study was to use the same cohort to investigate whether there was an association between ETS exposure and oxidative stress.

**Methodology**

**Main study**

Written, informed consent was obtained from participants. Approval for the consent process and overall study were obtained from the West of Scotland Multi-Centre Research Ethics Committee. Written, informed consent was obtained to undertake the interviews and perform assays on the residual serum from clinical blood samples taken on admission to hospital. The multi-centre research ethics committee also approved inclusion of patients who died before consent could be obtained; including use of their blood samples and extraction of information from their case-notes.

We previously recruited a cohort of patients admitted with acute myocardial infarction to nine Scottish, acute hospitals between 2005 and 2007 [10]. Acute myocardial infarction was defined as a detectable cardiac troponin I or T concentration in a patient admitted as an emergency for chest pain. Over the study period, troponin assays were performed routinely on all patients admitted to Scottish hospitals with chest pain. Therefore, this case definition could be applied consistently across all hospitals and on all patients, irrespective of date, time or route of admission. We excluded patients with other causes of raised troponin (types 2–5 myocardial infarction): renal failure, thromboembolic disease, myocarditis and coronary revascularisation.

In order to ensure complete case ascertainment, the hospital laboratories produced daily lists of patients with detectable serum troponin concentrations. Dedicated research nurses identified all eligible patients on the lists and completed structured interviews to obtain information on age, sex and postcode of residence, as well as self-reported smoking status and information on ETS exposure. In Scotland, there are 6,505 datazones, based on postcode of residence, with a mean population of 800. The Scottish Index of Multiple Deprivation (SIMD) for each datazone is derived from (http://www.scotland.gov.uk/Topics/Statistics/SIMD). The SIMD has been used to derive quintiles of socioeconomic status for the Scottish population; ranging from 1 (most deprived) to 5 (least deprived). We used postcode of residence to categorize our study participants according to these quintiles.

All samples were centrifuged and stored locally at −20°C before being transported on dry ice. One aliquot from each patient was sent to the ABS Laboratory (London, UK) where cotinine assays were performed using gas chromatography with a specific nitrogen/phosphorous detector GC-NPD. Cotinine and the internal standard 5-methyl cotinine were extracted using dichloroethane from 100 µL of serum after alkalisation using sodium hydroxide. The lower limit of detection was 0.1 ng/mL.

**Sub-study**

In the current study, we compared the 60 patients who classified themselves as never smokers, had a serum cotinine concentration of ≤0.1 mg/mL and for whom a second aliquot of serum was available, with a randomly selected 60 patients from the 369 patients who classified themselves as never smokers, had a serum cotinine concentration of ≥3.0 mg/mL and ≥12 mg/mL and for whom a second aliquot was available. Three biomarkers of oxidative stress (protein carbonyls, malondialdehyde (MDA) and oxidised LDL (ox-LDL)) were assayed by the Fre Radic Research laboratory (University of the Highlands & Islands, Inverness, UK) who were blinded to participants’ cotinine concentration. MDA and ox-LDL assays were performed on all 120 samples. There was insufficient sample available to perform protein carbonyl on 14 of the 120 samples.

**Malondialdehyde (MDA).** MDA was measured in defrosted samples using a commercially available kit (Cell Biolabs Incorporated, cat 10005020). The samples (100 µL) and standards were treated with butylated hydroxytoluene prior to incubation with SDS lysis solution (5 min, room temperature) and addition of TBA reagent (15 min, 95°C). After cooling and centrifugation (15 min, 5000 g), supernatant was aspirated and treated with an equivalent volume of butanol prior to further centrifugation (5 min, 10,000 g). The sample was then transferred to a 96-well microplate and absorbance read at λ = 532 nm; quantification was achieved against a standard curve (0–125 µM).

**Oxidised low density lipoprotein (Ox-LDL).** Oxidised LDL was assayed using a competitive enzyme linked immuno-sorbant assay (Mercodia, Cat 10-1158-01). Oxidised LDL in the sample competes with a fixed amount of oxidised LDL bound to the microtiter well for the binding of biotin-labelled specific antibodies (4E6). After washing to remove unreactive sample components, the biotin labelled antibody bond to the well is detected by HRP-conjugated streptavidin, detected by reaction with 3,3′,5,5′-tetramethylbenzidine. The reaction is stopped by adding acid to give a colorimetric endpoint that is read spectrophotometrically at λ = 450 nm. The results were calculated by expressing calibrators and standards as a percentage inhibition of the mean absorbance of the maximum binding at zero inhibition (B0). Cubic spline regression analysis was used to construct the standard curve.

**Protein carbonyls (PCs).** PCs were assayed in defrosted samples (100 µL) using a commercially available kit (Cayman assay kit; cat 10003020). Briefly, the assay measures the reaction of PCs with 2,4-dinitrophenylhydrazine (DNPH) reaction to form a Schiff base and the corresponding hydrazine. The amount of protein–hydroxylamine produced is quantified spectrophotometrically (λ = 375 nm).

**Statistical analyses**

Mann Whitney U tests were used to compare the concentrations of oxidative stress markers between never smokers with high and low exposure to ETS. Separate linear regression models were used to determine whether ETS exposure was associated with each of the oxidative stress biomarkers. Multivariate analyses were used to determine whether any associations remained after adjusting for the potential confounding effects of age, sex and socioeconomic deprivation. All tests were two-sided and statistical significance was defined as p<0.05.
Results

Never smokers who had high levels of ETS exposure were significantly more likely to live in socioeconomically deprived areas (Table 1). They were older and more likely to be female but neither reached statistical significance (Table 1). Exposed individuals had significantly higher concentrations of plasma MDA (Table 1 & Figure 1). Their higher concentrations of plasma protein carbonyl were of borderline statistical significance and there was no significant difference in plasma ox-LDL concentrations (Table 1).

After adjusting for the potential confounding effects of age, sex and socioeconomic status, exposure to environmental tobacco smoke was associated with serum concentrations of both protein carbonyl (beta coefficient 7.96, 95% CI 0.76, 15.17, p = 0.031) and MDA (beta coefficient 10.57, 95% CI 4.32, 16.81, p = 0.001) but not ox-LDL (beta coefficient 2.14, 95% CI −8.94, 13.21, p = 0.703) (Table 2).

Discussion

Among patients admitted to hospital for acute myocardial infarction, exposure to ETS was associated with increased concentrations of two plasma markers of oxidative stress (protein carbonyls and MDA). Future studies are required to determine whether oxidative stress plays a causal role in the association between ETS and myocardial infarction.

Several observational studies have demonstrated an association between exposure to ETS and oxidative stress among children.

Table 1. Summary characteristics of patients hospitalised for myocardial infarction categorised by exposure to environmental tobacco smoke.

|                  | Cotinine ≤0.1 | Cotinine ≥3.0 | P value |
|------------------|---------------|---------------|---------|
|                  | N = 60        | N = 60        |         |
|                  | N (%)         | N (%)         |         |
| Sex              |               |               |         |
| Male             | 43 (72)       | 39 (65)       | 0.434   |
| Female           | 17 (28)       | 21 (35)       |         |
| SIMD5            |               |               |         |
| 1 (deprived)     | 15 (25)       | 42 (70)       | <0.001  |
| 2                | 10 (17)       | 10 (17)       |         |
| 3                | 8 (13)        | 2 (3)         |         |
| 4                | 11 (18)       | 3 (5)         |         |
| 5 (affluent)     | 16 (27)       | 3 (5)         |         |
|                  | Median (IQR)  | Median (IQR)  |         |
| Age (years)      | 68 (57, 78)   | 72 (62, 75)   | 0.149   |
| Protein carbonyl | 19.49 (14.22, 25.16) | 22.99 (17.05, 32.49) | 0.052 |
| MDA (units)      | 27.65 (21.26, 35.86) | 40.59 (32.83, 51.88) | <0.001 |
| Ox LDL (units)   | 62.19 (48.15, 79.96) | 61.61 (45.76, 89.44) | 0.586  |

MWU, chi square for trend. N number; SIMD5 Scottish Index of Multiple Deprivation quintile; MDA; LDL; IQR interquartile range. doi:10.1371/journal.pone.0081209.t001
Table 2. Univariate and multivariate linear regression analyses of the association between exposure to environmental tobacco smoke and three biomarkers of oxidative stress.

| Biomarker       | Univariate | Multivariate* |
|-----------------|------------|---------------|
|                 | Beta coeff | 95% CI P value | Beta coeff | 95% CI P value |
| Protein carbonyl | 6.78       | 0.52, 13.04   | 0.034      | 7.96          | 0.76, 15.17 | 0.031 |
| MDA             | 12.82      | 7.30, 18.34   | -0.001     | 10.57         | 4.32, -16.81 | 0.001 |
| Ox LDL          | 4.02       | -5.72, 13.76  | 0.415      | 2.14          | -8.94, 13.21 | 0.703 |

*adjusted for age, sex, SIMD5.

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Kosecik et al. demonstrated a significantly lower total anti-
oxidative response among 83 children aged 9–13 years who were
exposed to ETS at home than among 61 children of similar age
who were not exposed [11]. Aycicek et al. studied 84 infants under
28 weeks of age [12]. Those infants who were exposed to ETS at
home had significantly lower concentrations of plasma vitamin C,
thiol, TAC 1 and TAC 2 concentrations and significantly higher
concentrations of plasma total peroxide, OSI1 and OSI2. Zalata et
al. compared 23 exposed children under 8 years of age with 23
who were not exposed [13]. The former had significantly higher
concentrations of MDA and significantly lower concentrations of
 glutathione peroxidase and tocopherol fractions, with evidence of
dose response relationships. Aycicek and Ipek examined fetal cord
blood and reported significantly higher lipid hydroperoxide, total
oxidant status and oxidative stress indices among women who
were exposed to ETS [14]. Shermatov et al. studied the peripheral
lymphocytes of 27 children who had been exposed to ETS and 27
who had not [15]. The former had significantly higher “total
oxidant status”, oxidative stress indices and DNA damage assessed
by alkaline comet assay. In a study by Kim et al., there was a
significant, independent association between urinary MDA and
PM2.5 among elderly subjects but not children, suggesting that the
effect of ETS exposure on oxidative stress may increase with age
[16]. In 1998, Howard et al. studied non-smokers who were not
exposed to ETS at home [5]. Those who were exposed to ETS in
the workplace had higher concentrations of catalase and
 glutathione peroxidase and higher oxidative DNA mutagen 8-
hydroxy-2'-deoxyguanosine.

Two intervention studies have examined the effect of ETS
exposure on oxidative stress. Kato et al. exposed 30 healthy
Japanese men, aged 25–39 years, to ETS for 30 minutes [17].
Exposure had no significant effect on the 15 smokers. In the 15
non-smokers, percentage flow-mediated vasodilation decreased
and plasma 8-isoprostanate increased significantly to concentrations
comparable to those of the active smokers. Kostikas et al.
undertook a randomised controlled cross-over trial in which they
exposed 18 never smokers to bar/restaurant levels of ETS for one
hour [18]. Following exposure, H2O2 in the exhaled breath
condensate increased and remained significantly higher four hours
after exposure. In a study of 29,579 Chinese non-smokers, Clark et
al. demonstrated an interaction with diet, whereby ETS exposure
was associated with coronary heart disease mortality only in the
sub-group with a diet low in fibre and anti-oxidants [19].

In a rat model, Valenti et al. demonstrated that the effect of
ETS exposure on cardiovascular regulation was mediated via its
influence on catalase activity [20]. Al-Arifa et al. demonstrated that
exposing rats to ETS resulted in increased mRNA expression of
oxidative stress genes as well as significant increases in a number
of markers of oxidative stress (heme oxygenase 1, catalase,
cyclooxygenase and glutathione S-transferase) [21]. Raji et al.
showed that exposure of rat aortic rings to cigarette smoke
impaired nitric oxide-mediated endothelial function via increased
generation of superoxide anion [22]. In a subsequent in vitro
study, using bovine, rat and human tissues, they demonstrated that
the thiol-reactive stable compounds in cigarette smoke activated
NADPH oxidase and increased endothelial superoxide anion
production, thereby reducing NO bioactivity and resulting in
endothelial dysfunction [23]. Based on rat models, Gentner and
Weber postulated that increased lung neutrophils or pulmonary
CYP1A1 may be responsible for the increase in oxidative stress
following exposure to ETS and may, in turn, be related to the
observed failure of blood pressure to fall during periods of sleep
and a possible increase in arterial stiffness [24].

To the best of our knowledge, this is the first study of the
association between exposure to ETS and oxidative stress among
patients suffering acute myocardial infarction. By its very nature,
oxidative stress is a difficult parameter to measure effectively in
vivo. Oxidative damage to biomolecules is an accepted surrogate
measure of oxygen-centred free radical generation and a range of
plasma markers of lipid and protein peroxidation have been
identified [25]. However, there is considerable debate as to the
benefits of one measure over another and, in studies where more
than one marker has been measured, there are not necessarily
cosistent effects across different markers of oxidative stress in
response to the same stimulus, suggesting that different markers
might reflect disturbances in different processes that contribute to
oxidative stress. With this in mind, our approach was to measure a
range of markers to provide as broad a picture as possible, whilst at
the same time potentially contributing additional information as to
the mechanisms involved in the process. Three markers were
selected: MDA and ox-LDL as markers of lipid peroxidation, and
protein carbonyls as a marker of protein oxidation. MDA is an
end-product of lipid peroxidation that has been associated with a
range of risk factors for cardiovascular disease, including cigarette
smoke [26]. Ox-LDL is a key mediator of atherosclerosis and
plasma concentrations have been found to be elevated in a range of
cardiovascular diseases [26,27]. Plasma ox-LDL and MDA
concentrations may also be associated with plaque instability
[27,29]; with studies suggesting higher concentrations among
patients presenting with acute MI compared to those with stable
angina [27,28] and, among patients with coronary artery disease,
MDA concentration are higher in patients with thin-cap atheroma
and complex plaques [29]. Protein carbonyls, on the other hand,
represent oxidation in a different compartment in the plasma that
have been shown to be associated with smoking in some studies
[30] and with age, ethnicity and some socio-economic factors
amongst smokers, but not with urinary cotinine in another [31].
Given the relevance of these three markers in our patient group, it
was considered sensible to measure them all where sufficient
sample was available.
The lack of association of cigarette smoke exposure with ox-LDL is perhaps surprising, given that previous studies have shown an association of this marker with both smoking and with acute MI, but this marker is perhaps the most susceptible to dietary influences, such as recent meals containing cholesterol or oxidised cholesterol, which could not be controlled for in this study. Nevertheless, the finding that other markers of oxidative stress in the lipid (MDA) and aqueous protein (carbonyls) plasma compartments both show an association with ETS exposure is supportive of the notion that oxidative stress caused by smoke inhalation is implicated in acute MI in vulnerable individuals. Furthermore, the discrepancy in the findings between different markers of oxidative stress highlights the need for multiple markers to be measured, given that each will be the product of different processes with different formation and clearance rates. In our analyses we were able to adjust for the potential confounding effects of age, sex and area deprivation. However, we did not have access to information on lifestyle risk factors such as physical activity, diet and adiposity. Therefore, residual confounding is possible. Our findings were based on relatively small numbers of patients and merit corroboraton in larger studies.

References

1. He J, Vupputuri S, Allen K, Peress MR, Huguet J, et al. (1999) Passive smoking and the risk of coronary heart disease: a meta-analysis of epidemiologic studies. N Engl J Med. 340: 920–926.
2. Celermajer DS, Adams MR, Clarkson P, Robinson J, McCreadie R, et al. (1996) Passive smoking and impaired endothelium-dependent arterial dilation in healthy young adults. N Engl J Med 334(3): 150–154.
3. Ambrose JA, Barna RS (2003) The pathophysiology of cigarette smoking and cardiovascular disease: An update. JACC 43: 1731–1737.
4. Raupach T, Schäfer K, Konstantinides S, Andreas S (2006) Secondhand smoke as an acute threat for the cardiovascular system: a change in paradigm. Eur Heart J 27(4): 301–309.
5. Howard DJ, Ota RB, Briggs LA, Hampton M, Prissos CA (1998) Environmental tobacco smoke in the workplace induces oxidative stress in employees, including increased production of 8-hydroxy-2′-deoxyguanosine. Cancer Epidemiology, Biomarkers and Prevention 7: 141–146.
6. Torok J, Gvozdjakova A, Kucharska J, Balazovjech I, Kysela S, et al. (2000) Passive smoking impairs endothelium-dependent relaxation of isolated rabbit arteries. Physiol Res 49: 135–141.
7. Zhu BQ, Sun YP, Sudhir K, Selek S, Glantz SA, et al. (1994) Exposure to environmental tobacco smoke increases myocardial infarct size in rats. Circulation 90: 1282–1290.
8. Panagiotakos DB, Pitsavos C, Stefanadis C (2007) Chronic exposure to second hand smoke and 30 day prognosis of patients hospitalized with acute coronary syndromes: the Greek study of acute coronary syndromes. Heart 93: 309–212.
9. Pell JP, Hsu S, Colebe S, Newby DE, Pell AC, et al. (2009) Second-hand smoke exposure and survival following acute coronary syndrome: Prospective cohort study of 1,261 consecutive admissions among never smokers. Heart 95 (17): 1415–1418.
10. Pell JP, Hsu S, Colebe S, Newby DE, Pell AC, et al. (2008) Smoke-free legislation and hospitalizations for acute coronary syndrome. New Engl J Med 359(3): 482–491.
11. Kosecik M, Erel O, Sevine E, Selek S (2005) Increased oxidative stress in children exposed to passive smoking. Int J Cardiol 100(1): 61–64.
12. Ayicak A, Erel O, Kocyigit A (2005) Increased oxidative stress in infants exposed to passive smoking. Eur J Pediatr 164(12): 775–778.
13. Zalata A, Yalas S, El-Bakary A, ElSheikha HM (2007) Increased DNA damage in children caused by passive smoking as assessed by comet assay and oxidative stress. Mutat Res 629(2): 140–147.
14. Ayicak A, Ipek A (2008) Maternal active or passive smoking causes oxidative stress in cord blood. Eur J Pediatr 167(1): 81–85.
15. Shermatos K, Zeyrek D, Yildirim F, Kilic M, Cebi N, et al. (2012) DNA damage in children exposed to secondhand cigarette smoke and its association with oxidative stress. Indian Pediatr 49(12): 958–962.
16. Kim K, Park EY, Lee KH, Park JD, Kim YD, et al. (2009) Differential oxidative stress response in young children and the elderly following exposure to PM2.5. Environ Health Prev Med 14(1): 60–66.
17. Kato-T, Inoue T, Morosuka T, Yoshimoto N, Node K (2006) Short-term passive smoking causes endothelial dysfunction via oxidative stress in nonsmokers. Can J Physiol Pharmacol 84(5): 523–529.
18. Kostikas K, Minas M, Nikolau E, Papaioannou AI, Laakis P, et al. (2013) Secondhand smoke exposure induces acutely airway acidification and oxidative stress. Respir Med 107(2): 172–179.
19. Clark ML, Butler LM, Koh WP, Wang R, Yuan JM (2013) Dietary fiber intake modifies the association between secondhand smoke exposure and coronary heart disease mortality among Chinese non-smokers in Singapore. Nutrition 29(11–12): 1304–1309.
20. Valenti VE, de Abreu LC, Sato MA, Ferreira C, Adami F, et al. (2012) Sidestream cigarette smoke effects on cardiovascular responses in conscious rats: involvement of oxidative stress in the fourth cerebral ventricle. BMC Cardiovasc Disord 12: 22.
21. Al-Arifi MN, Maayah ZH, Alishamrani AA, Khorashy HM (2012) Impact of cigarette smoke exposure on the expression of cardiac hypertrophic genes, cytochrome P450 enzymes, and oxidative stress markers in rats. J Toxicol Sci 37(2): 1083–1090.
22. Raji L, DeMaster EG, James EA (2001) Cigarette smoke-induced endothelium dysfunction: role of superoxide anion. J Hypertens 19: 891–897.
23. James EA, DeMaster EG, Tian RN, Raji L (2004) Stable compounds of cigarette smoke induce endothelial superoxide anion production via NADPH oxidase activation. Arterioscler Thromb Vasc Biol 24: 1031–1036.
24. Gentner NJ, Webey LP (2012) Secondhand tobacco smoke: arterial stiffness, and altered circadian blood pressure patterns are associated with lung inflammation and oxidative stress in rats. Am J Physiol Heart Circ Physiol 302(3): H818–H825.
25. Lee R, Margaritis M, Channon KM, Anoniades C (2012) Evaluating oxidative stress in human cardiovascular disease: methodological aspects and considerations. Curr Med Chem 19: 2504–2520.
26. Sanderson KJ, van Rij AM, Wade CR, Sutherland WH (1995) Lipid peroxidation of circulating low density lipoproteins with age, smoking and in peripheral vascular disease. Atherosclerosis 118: 45–51.
27. Ebara S, Ueda M, Naruko T, Haze K, Itoh A, et al. (2001) Elevated levels of oxidized low density lipoprotein show a positive relationship with the severity of acute coronary syndromes. Circulation 103: 1955–1960.
28. Holvoet P, Vanhaecke J, Janssens S, Van de Werf F, et al. (1998) Oxidized LDL and malondialdehyde-modified LDL in patients with acute coronary syndromes and stable coronary artery disease. Circulation 98: 1487–1494.
29. Tajika K, Okamatsu K, Takano M, Inami S, Yamamoto M, et al. (2012) Malondialdehyde-modified low density lipoprotein is a useful marker to identify patients with vulnerable plaque. Circulation 76: 2211–2217.
30. Kapaki E, Liappas I, Lyras L, Paraskevas GP, Mamali I, et al. (2007) Oxidative damage to plasma proteins in patients with chronic alcoholic dependence: the effect of smoking. In Vivo 21: 525–528.
31. Chih-Ching Y, Barr RG, Powell CA, Mesia-Vela S, Wang Y, et al. (2008) No effect of cigarette smoking dose on oxidized plasma proteins. Environ Res 106: 219–225.

Conclusion

Oxidative stress increases the risk of developing cardiovascular disease and disease progression and it is now well established that active smoking predisposes to oxidative stress. Our study adds to increasing evidence that ETS exposure may also be associated with oxidative stress. This mechanism may play a role in the association between ETS exposure and myocardial infarction.

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Author Contributions

Conceived and designed the experiments: SJH JPP. Performed the experiments: ILM. Analyzed the data: DEN. Contributed reagents/materials/analysis tools: ILM. Wrote the paper: JPP. All authors interpreted the results, revised the manuscript and approved the final version.

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