Human *Toxoplasma gondii* infection in Nigeria: a systematic review and meta-analysis of data published between 1960 and 2019

Solomon Ngutor Karshima and Magdalene Nguvan Karshima

**Abstract**

**Background:** Over 70% of the world’s population is infected by *Toxoplasma gondii*, a pathogen capable of causing cerebral toxoplasmosis in HIV patients and neonatal complications like miscarriage, chorioretinitis, hydrocephalus, cerebral calcification and foetal death in the third trimester of pregnancy. In spite of this, the burden of this zoonotic pathogen is poorly understood in Nigeria. The aim of the present study therefore, is to determine the burden of *T. gondii* among normal individuals, HIV patients and pregnant women as well as the distribution of the infection across Nigeria.

**Methods:** Using the PRISMA guidelines, we conducted a systematic review and meta-analysis of data retrieved from six electronic databases (AJOL, Google Scholar, PubMed, Scopus, Science Direct and Web of Science). Pooled prevalence (PP) and heterogeneity were determined by the random-effects model and the Cochran’s Q-test respectively. The quality of each study and publication bias were assessed by the 9 point Joanna Briggs Institute Critical Appraisal Instrument and the Egger’s regression asymmetry test respectively, while the robustness of a pooled estimate was tested by the single study omission analysis.

**Results:** Exactly 5834 of the 16,230 individuals examined for *T. gondii* infection by 50 studies across 17 Nigerian States were positive for the infection. Overall PP was 32.92% (95% CI: 27.89, 38.37), with a range of 14.41% (95% CI: 5.32, 33.54) to 86.82% (95% CI: 66.13, 95.69) across sub-groups. Pooled prevalence was significantly higher (*p* < 0.001) among pregnant women (40.25%; 95% CI: 33.19, 47.73) and HIV patients (31.68, 95% CI: 20.53, 45.41) than normal individuals (23.32, 95% CI: 17.25, 30.75). *T. gondii* prevalence declined by over 58% during the 59 years reviewed.

**Conclusion:** *Toxoplasma gondii* infection is moderately prevalent in Nigeria. Highest prevalence estimates were observed among pregnant women and in the south-south region. For effective control of the disease in Nigeria, a holistic approach involving on-farm, environmental, public health and animal components are suggested.

**Keywords:** Geographical distribution, HIV patients, Normal individuals, Pregnant women, Prevalence, *Toxoplasma gondii*
Background

Toxoplasma gondii is an obligate intracellular protozoan zoonotic pathogen of almost all warm-blooded animals including humans and birds [1]. The infection is worldwide in distribution and over 70% of the world’s population is infected [2, 3]. Domestic cats are the definitive hosts, and represent the main source of infection through oocysts passed in their faeces. The pathogen is currently a global problem which is present in every country of the world [4].

Human infection may result via several routes including contact with infected cats, the consumption of animal tissues infected by cysts of T. gondii, the ingestion of food or water contaminated with oocysts excreted in the faeces of cats, blood transfusion and intrauterine [5–7]. Transmission is influenced by factors such as environmental conditions, host immune status, cultural behaviour, individual’s hygienic practices, type of food and cooking methods [8, 9].

In immunocompetent individuals, Toxoplasma infection may be asymptomatic or self-limiting. However, immunocompromised conditions like HIV infection may alter the clinical course of T. gondii infection [10, 11]. HIV infection may cause reactivation of the asymptomatic Toxoplasma infection resulting in neurological signs like headache, disorientation, drowsiness, hemiparesis, reflex changes and convulsion [12–14]. About 25–50% of HIV immunocompromised individuals may show the signs of cerebral toxoplasmosis [15] and Toxoplasma infection is now a known cause of morbidity and mortality in people living with HIV and AIDS (PLWHA) [12].

Toxoplasma induced neonatal complications especially during third trimester of pregnancy may include miscarriage, chorioretinitis, hydrocephalus, cerebral calcification and foetal death [16, 17]. The risk of transmission of Toxoplasma infection from mother to child intrauterine is increased during the third trimester of pregnancy [18, 19]. Continental prevalence of T. gondii across the world ranged between 4.3–75.0% in Africa [20–23], 14.0–96.3% in Asia [24–26], 6.8–51.8% in Europe [27–29], 10.6–13.0% in North America [30–32] and 26.3–80.0% in South America [33–35].

Despite the association of toxoplasmosis with immunocompromised conditions and the increasing number of PLWHA and pregnant women in Nigeria, the nationwide prevalence and burden of T. gondii infection are poorly understood. In this study, we reported the burden of T. gondii infections among normal individuals, HIV patients and pregnant women in Nigeria. It is envisaged that the present finding will enable stakeholders and policy makers in the health sector to re-strategize on the control of toxoplasmosis, thus reducing the burden of the disease especially in Nigerian pregnant women.

Methods

Study protocol and literature search procedure

We conducted a systematic review and meta-analysis using the guidelines provided by Moher [36] for Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA). Inclusion of data for quantitative synthesis was based on the PRISMA checklist (Additional file 1), and the infection of humans with Toxoplasma gondii was the outcome of interest. The review protocol was registered on PROSPERO International prospective register of systematic reviews with registration number CRD42019135416 and available from: http://www.crd.york.ac.uk/PROSPERO/display_record.php?ID=CRD42019135416.

Six electronic databases: African Journals OnLine (AJOL), Google Scholar, PubMed, Scopus, Science Direct and Web of Science were systematically searched between 1st June and 30th April 2020 for literature published on T. gondii infection in humans in Nigeria between 1960 and 2019. Additional studies were obtained through searching of list of references of retrieved studies and by contacting editors of Nigerian biomedical journals. Full articles with only visible online abstracts were requested from authors and editors of the publishing journals through phone calls or e-mails. The MeSH search string employed in PubMed was “toxoplasmosis” OR “Toxoplasma” OR “Toxoplasma gondii” OR “Toxoplasma infections” AND “Prevalence” OR “Seroepidemiology” AND “Humans” OR “Healthy individuals” OR “HIV patients” OR “Pregnant women” AND “North-central” OR “North-eastern” OR “North-western” OR “South-eastern” OR “South-south” OR “South-western” AND “Nigeria”. Search on AJOL was carried out on journal by journal bases within biomedical journals indexed in the database. Due to the large volume of data in Google Scholar, the study customised article search in this database based on year of publication for ease of sorting articles.

Requirements for inclusion

Studies were subjected to two stages of screening for either inclusion or exclusion. In the first stage, studies were screened by scanning through titles for exclusion of duplicates. Second stage screening involved detailed review of abstract and full text for removal of irrelevant studies and identification of relevant information. A study was considered for inclusion if it had the following characteristics: (i) it was carried out in Nigeria, (ii) it was published in English, (iii) it was carried out and published between January 1960 and December 2019, (iv) it was a cross sectional or prevalence study, (v) it stated the study location, (vi) it clearly stated the number of sample size and positive cases, (vii) it reported T. gondii in humans, and (viii) it stated the target population.
Studies that did not meet these inclusion criteria and all unpublished articles were excluded.

Quality assessment
The quality of each article analysed was assessed independently using the 9 point Joanna Briggs Institute (JBI) critical appraisal instrument for studies reporting prevalence data [37]. The JBI checklist posed nine questions viz.: (1) Was the sample frame appropriate to address the target population? (2) Were study participants recruited in an appropriate way? (3) Was the sample size adequate? (4) Were the study subjects and setting described in detail? (5) Was data analysis conducted with sufficient coverage of the identified sample? (6) Were valid methods used for the identification of the condition? (7) Was the condition measured in a standard, reliable way for all participants? (8) Was there appropriate statistical analysis? (9) Was the response rate adequate, and if not, was the low response rate managed appropriately (Additional file 2)? Answers to the aforementioned questions for individual studies were respectively assigned scores of 0 or 1 for no or yes answers, while U or NA were used when a study does not clearly answer the question or when the question was not applicable to the study. For a study to be included in the quantitative synthesis, it was required to have a minimum quality assessment score of 6 (66.7%); that is answering yes to at least 6 of the 9 questions on the checklist.

Extraction of data
To ensure data validation and increase the likelihood of detecting errors, literature search, screening of articles, selection of articles for eligibility and data extraction were performed by both authors (SNK and MNK) independently. However, in cases of discrepancies, both authors crosschecked data simultaneously and discussed issues until consensus was reached. Data pulled out from each published study were name of author, the year the study was carried out, the year it was published, sample size, number of positive cases, study location, study design, method of diagnosis, diagnostic target and characteristics of study population. Where specified, the gender and ages of study population were also extracted and individuals within the age brackets ≤17 years of age were categorised as children while those ≥18 years of age were categorised as adults. Where the study year for any article was not stated, the year preceding its publication year was considered as the year it was conducted.

In our PICOS, we answered questions such as: (1) What is the burden of Toxoplasma gondii infection in normal individuals, HIV patients and pregnant women from Nigeria? (2) Is the burden greater among immunocompromised than normal individuals? (3) What is the distribution pattern of the infection across Nigeria? For the purpose of the present study, an individual was said to be infected with T. gondii only if the individual tested positive for the parasite by microscopic, serological or molecular techniques, normal individuals refers to individuals without any history of pregnancy or any immunocompromised conditions like HIV/AIDS and neoplasia, pregnancy refers to a state where a woman carried an embryo or foetus for a period of ±9 months and HIV patients refer to people living with HIV/AIDS. More so, for the purpose of our analysis, diagnostic methods like polymerase chain reaction (PCR) and immunochromatography which were utilized by only one study, latex agglutination test (LAT) which was utilized by only two studies and studies with unidentified methods of diagnosis were grouped as others.

Data collation and analyses
Preliminary analyses including summations, subtractions, divisions, multiplications and estimation of percentages were conducted using Microsoft Excel. Statistical and meta-analyses were carried out using Graph-Pad Prism version 4.0 and Comprehensive Meta-Analysis version 3.0 respectively. Prevalence of individual studies was determined by expressing the proportion of positive cases of T. gondii infection and sample size as percentages.

Pooling, sub-group and heterogeneity analyses
Pooled prevalence and their 95% Confidence Interval (CI) were estimated by the random-effects model [38]. Sub-group analyses were performed based on ages (Adult, children), gender (female, male), characteristics of study population (normal individuals, HIV patients and pregnant women) and geographic regions (North-central, north-east, north-west, south-east, south-south and south-west). Others were diagnostic methods (Dye test, ELISA, HAT, LAT, PCR, and RSAT), study period (1960–1975, 1976–1990, 1991–2005 and 2006–2019) as well as sample size (≤ 150, 151–300, 301–450 and > 450).

Heterogeneity among studies was evaluated using the Cochrane’s Q-test while the percentage variation among studies due to heterogeneity was quantified using the formula $I^2 = 100 \times \left( \frac{Q-df}{Q} \right)$; where Q is Cochrane’s heterogeneity statistic and df is the degree of freedom which is determined by subtracting one from the number of studies analysed. I-square values of 0, 25, 50 and 75% were considered no, low, moderate and substantive heterogeneities respectively [39, 40].

Publication bias, sensitivity and meta-regression analyses
Publication bias (across-study bias) was examined by funnel plots while the statistical significance was assessed by the Egger’s regression asymmetry test [41]. The unbiased estimates were calculated using the Duval
and Tweedie non-parametric ‘fill and trim’ linear random method [42]. The robustness of a pooled estimate was tested by the single study omission analysis, and a study was considered to have no influence on the pooled prevalence if the pooled estimate without it (i.e. number of studies = 49) was within the 95% confidence limits of the overall pooled prevalence when number of studies equals 50 [43]. Meta-regression analysis was performed for different sub-groups including year of conduct of study, diagnostic methods, geographic regions as well as age, gender and characteristics of the study population to determine the possible sources of heterogeneity.

Results

Literature search and eligible studies

The procedure for selection of eligible studies is presented in Fig. 1. A total of 79 studies resulted from the search of six electronic databases. Twenty five of the studies were duplicates and were removed after screening of titles. Fifty four of the studies were subjected to detailed review of abstract and full text. Four studies were thereafter removed for insufficiency of data on sample sizes and number of cases \((n = 3)\) and lack of information on study location \((n = 1)\). Fifty studies were subjected to the quantitative synthesis. None of the studies assessed for quality by the JBI critical appraisal instrument was excluded for lack of merit. Quality scores ranged between 6 and 8 (66.67–88.89%) of a total of 9 scores (Table 1 and Additional file 3).

Characteristics of eligible studies

Table 1 shows the characteristics of the eligible studies. Fifty studies examined 16,230 individuals for *Toxoplasma gondii* infection among Nigerians and reported prevalence rates ranging between 2.00 and 88.24%. Nine, 4, 10, 8 and 19 of the studies were reported in the north-central, north-eastern, north-western, south-south and the south-western regions respectively. Three studies utilized rapid slide agglutination test (RSAT) for diagnosis, 4 each were diagnosed using dye and haem-agglutination tests, 28 of the studies utilized enzyme linked immunosorbent assay (ELISA) while 11 studies utilized other methods (immunochromatographic test 1, LAT 2, PCR 1 and unidentified tests 7). One study each targeted *Toxoplasma*-immunoglobulin M (IgM) and deoxyribonucleic acid (DNA), 20 studies targeted *Toxoplasma*-immunoglobulin G (IgG), 17 targeted both IgG and IgM, while 11 studies failed to state their diagnostic targets. Two studies were reported between 1960 and 1975, 6 between 1976 and 1990, 5 between 1991 and 2005 and 37 between 2006 and 2019. Twelve, 18, 13 and 7 of the studies had sample sizes of \(\leq 150\), 151–300, 301–450 and > 450 respectively.

Spatial distribution of eligible studies

The 50 studies were reported across 5 of the 6 regions of Nigeria as presented in Fig. 2. The nine (18.00%) studies reported across the north-central region were distributed as follow: one each in Kwara and Niger States, 2 each in Abuja and Benue State, and 3 in Plateau State. Four (8.00%) studies were reported in Borno State, North-east Nigeria while the 10 (20.00%) studies reported across the north-west region were from Kaduna and Sokoto (3 studies each) and 4 from Kano. One study each was reported from Akwa Ibom and Delta States and 3 each from Edo and Rivers States totally 8 (16.00%) studies from the south-south while the 19 (38.00%) studies distributed across the south-west; 1, 2, 6 and 10 were from Ogun, Osun, Oyo and Lagos States respectively. No study was reported in the south-east region. 

---

![Flow diagram for the selection of eligible studies](image)
| Study year | Study location | Region | Method of Diagnosis | Diagnostic targets | Characteristics of study population | Sample size | Cases | Prev. (%) 95% CI | QAS Study Ref |
|------------|----------------|--------|---------------------|--------------------|-------------------------------------|-------------|-------|-----------------|---------------|
| 2015 Osun   | South-west    | ELISA  | IgG and IgM         | Pregnant women     | 391 186 4757 42.53–52.65 6 | [44]        |
| 1999–04 Kwara | North-central | ELISA  | Not stated          | HIV-positive individuals | 60 25 41.67 29.07–55.12 7 | [45]        |
| 1989 Kaduna | North-west    | HAT    | Not stated          | Pregnant women     | 834 329 3945 36.11–42.86 8 | [46]        |
| 2009 Lagos  | South-west    | –      | IgG                 | HIV-positive individuals | 460 230 5000 45.34–54.66 7 | [47]        |
| 2009 Lagos  | South-west    | ELISA  | IgG                 | Pregnant women     | 179 73 4078 33.51–48.36 7 | [48]        |
| 2011 Sokoto | North-west    | RSAT   | IgG                 | HIV-positive individuals | 84 22 2619 17.20–36.93 6 | [49]        |
| 2011 Sokoto | North-west    | LAT    | IgG                 | Abattoir workers   | 75 38 5067 38.86–62.42 6 | [50]        |
| 2012 Sokoto | North-west    | RSAT   | IgG                 | Pregnant women     | 173 48 2775 21.22–35.05 6 | [51]        |
| 2018 Oyo    | South-west    | ELISA  | IgG and IgM         | Blood donors       | 248 106 4274 36.50–49.16 7 | [52]        |
| 2011 Benue  | North-central | ELISA  | IgG                 | HIV-positive individuals | 360 39 1083 7.82–14.51 7 | [53]        |
| 1985 Delta  | South-south   | IgG    | Not stated          | Pregnant women and blood donors | 1650 972 5891 56.49–61.30 7 | [54]        |
| 2013 Oyo    | South-west    | PCR    | DNA                 | Pregnant women     | 179 49 2737 20.99–34.53 6 | [55]        |
| 2008 Kaduna | North-west    | ELISA  | IgG and IgM         | Pregnant women     | 374 112 2995 25.35–34.87 8 | [56]        |
| 2010 Lagos  | South-west    | ELISA  | IgG and IgM         | Pregnant women     | 276 111 4022 34.38–46.26 6 | [57]        |
| 2016/17 Akwa Ibom | South-south | ELISA  | IgG                 | Abattoir workers   | 339 189 5575 50.29–61.12 7 | [58]        |
| 1993 Oyo    | South-west    | –      | IgG                 | Lymphoid neoplasia patients | 162 44 2716 20.48–34.70 6 | [59]        |
| 2018 Oggun  | South-west    | RSAT   | IgG and IgM         | Young adults with history of ocular infections | 150 3 200 0.41–5.73 8 | [60]        |
| 2011 Borno  | North-east    | ELISA  | IgG                 | HIV positive individuals | 190 42 2211 16.42–28.68 7 | [61]        |
| 2013/14 Lagos | South-west   | LAT    | Not stated          | Primary school children | 382 91 2383 19.64–28.42 7 | [62]        |
| 2016 Kano   | North-west    | ELISA  | IgM                 | Obstetric patients  | 320 24 750 4.86–10.95 6 | [63]        |
| 1960 Kano   | North-west    | Dye Test| Not stated          | Chorioretinitis patients | 17 15 8824 63.56–98.54 7 | [64]        |
| 2011/12 Edo | South-south   | ELISA  | IgG and IgM         | Psychotic individuals | 280 90 3214 26.71–37.96 7 | [65]        |
| 2008 Borno  | North-east    | ELISA  | IgG                 | Normal individuals  | 180 43 2389 17.9–30.8 7 | [66]        |
| 2011 Abuja  | North-central | ELISA  | IgG and IgM         | HIV-positive individuals | 341 30 880 6.0–12.3 6 | [67]        |
| 2014 Borno  | North-east    | ELISA  | IgG and IgM         | Pregnant women     | 360 176 4889 43.6–54.2 8 | [68]        |
| 2013 Rivers | South-south   | ELISA  | IgG and IgM         | Pregnant women     | 288 189 6563 59.8–71.1 8 | [69]        |
| 2018 Edo    | South-south   | ICT    | IgG and IgM         | HIV-positive and healthy individuals | 1500 113 753 6.3–9.0 6 | [70]        |
| 2008 Kaduna | North-west    | ELISA  | IgG and IgM         | HIV-positive and healthy individuals | 219 78 3562 29.3–42.4 8 | [71]        |
| 1980 Kano   | North-west    | HAT    | Not stated          | Pregnant women and male patients | 20 7 3500 15.4–59.2 7 | [72]        |

Table 1: List and characteristics of eligible studies.
| Study year | Study location | Region       | Method of Diagnosis | Diagnostic target        | Characteristics of study population                         | Sample size | Cases | Prev. (%) | 95% CI       | QAS | Study Ref |
|------------|----------------|--------------|---------------------|--------------------------|------------------------------------------------------------|-------------|-------|------------|-------------|-----|-----------|
| 1991       | Oyo            | South-west   | Dye Test            | IgG                      | Pregnant women                                            | 352         | 273   | 77.66      | 72.83–81.81 | 8   | [76]      |
| 2008       | Lagos          | South-west   | –                   | Not stated               | HIV-positive individuals                                   | 400         | 14    | 3.50       | 1.93–5.80   | 6   | [77]      |
| 2016       | Rivers         | South-south  | ELISA               | IgG                      | Healthy individuals                                       | 800         | 329   | 41.13      | 37.69–44.62 | 7   | [78]      |
| 2009       | Lagos          | South-west   | –                   | IgG                      | HIV-positive individuals                                   | 83          | 71    | 85.54      | 76.11–92.30 | 7   | [79]      |
| 1984       | Plateau        | North-central| HAT                 | Not stated               | Pregnant women and other hospital patients                 | 210         | 48    | 22.86      | 17.36–29.14 | 6   | [80]      |
| 2010       | Lagos          | South-west   | ELISA               | IgG                      | HIV-positive individuals                                   | 380         | 206   | 54.21      | 49.05–59.30 | 7   | [81]      |
| 2013       | Borno          | North-east   | ELISA               | IgG                      | Pregnant women                                            | 90          | 20    | 22.22      | 14.13–32.21 | 6   | [82]      |
| 2015       | Lagos          | South-west   | HAT                 | IgG and IgM              | HIV-positive and normal individuals                        | 65          | 2     | 30.80      | 0.37–10.68  | 6   | [83]      |
| 1985       | Niger          | North-central| –                   | Not stated               | Hospital patients                                          | 176         | 139   | 78.98      | 72.21–84.75 | 6   | [84]      |
| 2016       | Osun           | South-west   | ELISA               | IgG and IgM              | Pre-school aged children                                  | 272         | 19    | 6.99       | 4.26–10.69  | 7   | [85]      |
| 1980       | Oyo            | South-west   | ELISA               | Not stated               | Children                                                  | 66          | 11    | 16.67      | 8.63–27.87  | 6   | [86]      |
| 2004       | Plateau        | North-central| –                   | IgG                      | HIV-positive individuals                                   | 363         | 115   | 31.68      | 26.92–36.74 | 7   | [87]      |
| 2006       | Plateau        | North-central| ELISA               | IgG                      | Normal individuals                                         | 144         | 30    | 20.83      | 14.52–28.39 | 6   | [88]      |
| 2011       | Abuja          | North-central| ELISA               | IgG                      | Pregnant, HIV-positive and normal individuals             | 216         | 68    | 31.48      | 25.35–38.13 | 8   | [89]      |
| 1969       | Oyo            | South-west   | Dye Test            | Not stated               | Normal individuals                                         | 6           | 5     | 83.33      | 35.88–99.58 | 6   | [90]      |
| 2016       | Rivers         | South-south  | ELISA               | IgG and IgM              | Pregnant women                                            | 213         | 58    | 27.23      | 21.37–33.73 | 7   | [91]      |
| 2014       | Kano           | North-west   | ELISA               | IgG and IgM              | HIV-positive pregnant women                                | 273         | 93    | 34.07      | 28.46–40.02 | 8   | [92]      |
| 2015       | Edo            | South-south  | ELISA               | IgG and IgM              | HIV-positive individuals                                   | 342         | 208   | 60.82      | 55.42–66.03 | 8   | [93]      |

CI: Confidence interval; ELISA: Enzyme linked immunosorbent assay; HAT: Haemagglutination test; ICT: Immunochromatographic test; IgG: Immunoglobulin G; IgM: Immunoglobulin M; LAT: Latex agglutination test; PCR: Polymerase chain reaction; QAS: Quality assessment score; RSAT: Rapid slide agglutination test; Prev: Prevalence
Pooled prevalence and heterogeneity analysis

Pooled prevalence and heterogeneities are presented in Tables 2 and 3 as well as Figs. 3 and 4. Fifty studies reported 5834 positive cases of *Toxoplasma gondii* infection among 16,230 individuals examined from 17 Nigerian States. Overall PP was 32.92% (95% CI: 27.89, 38.37; Q-\(p\) < 0.001) with ranges of 28.58 (95% CI: 16.28, 45.17; Q-\(p\) < 0.001) to 41.00% (95% CI: 25.61, 58.37; Q-\(p\) < 0.001) across geographic regions, 14.41% (95% CI: 5.32, 33.54; Q-\(p\) < 0.001) to 74.29% (95% CI: 57.49, 86.06; Q-\(p\) < 0.001) across methods of diagnosis, 28.75 (95% CI: 23.44, 34.71; Q-\(p\) < 0.001) to 86.82% (95% CI: 66.13, 95.69; Q-\(p\): 0.760) across study periods and 30.22 (95% CI: 19.87–43.07; Q-\(p\) < 0.001) to 36.64% (95% CI: 24.15, 51.23; Q-\(p\) < 0.001) across sample size.

Pooled prevalence of *T. gondii* infection among adults and children were 36.93% (95% CI: 29.04, 45.59; Q-\(p\) < 0.001) and 18.52% (95% CI: 6.02, 44.63; Q-\(p\) < 0.001) respectively. Gender-based PP were 30.61% (95% CI: 22.95, 39.51; Q-\(p\) < 0.001) and 31.85% (95% CI: 21.79, 43.94; Q-\(p\) < 0.001) for females and males respectively. Pooled prevalence reported in both HIV-patients 31.68% (95% CI: 20.53, 45.41; Q-\(p\) < 0.001) and pregnant women 40.25% (95% CI: 33.19, 47.73; Q-\(p\) < 0.001) were significantly higher (\(p\) < 0.001) than that among normal individuals 23.32% (95% CI: 17.25, 30.75; Q-\(p\): 0.001). Overall heterogeneity was 97.64% with a range of 85.76 to 99.17% (Tables 2 and 3).

Publication bias, sensitivity and meta-regression analyses

The funnel plots (Fig. 5) and their respective bias coefficients for studies published in Nigeria as a whole (b: -3.48; 95% CI: -7.53, 0.58; \(p\): 0.091), northern (b: -3.10; 95% CI: -7.90, 1.71; \(p\): 0.194) and southern regions (b: -5.03; 95% CI: -11.05, 0.99; \(p\): 0.098) suggest insignificant publication bias. No outlying study capable of causing publication bias was identified and removed by the Duval and Tweedie’s trim and fill method (Additional file 4). As in Additional file 5, the sensitivity tests showed that all single-study omission estimates were within the 95% CI of the overall PP.

Meta-regression analysis (Tables 2 and 3) suggests that geographic regions (Q: 1.89; \(df\): 4; \(p\): 0.756), sample size (Q: 0.53; \(df\): 3; \(p\): 0.913), age (Q: 0.57; \(df\): 1; \(p\): 0.451) and gender of participants (Q: 0.04; \(df\): 1; \(p\): 0.835) were unlikely to be the sources of heterogeneity in the present
### Table 2: Pooled prevalence of *T. gondii* infection in humans in Nigeria based on sub-groups

| Variables               | No. of Studies | Sample size | Cases | Prev. (%) | Pooled Estimates (95% CI) | Heterogeneity | Meta-regression | P-value | OR (95% CI) |
|-------------------------|----------------|-------------|-------|-----------|---------------------------|---------------|-----------------|---------|-------------|
|                         |                |             |       |           |                           |               |                 |         |             |
| Region                  |                |             |       |           |                           |               |                 |         |             |
| South-west              | 19             | 5133        | 1880  | 31.70     | 23.86, 40.74              | 618.15        | 97.09           | < 0.001 | 0.756       |
| South-south             | 8              | 5412        | 2148  | 41.00     | 25.61, 58.37              | 825.41        | 99.15           | < 0.001 | –0.15       |
| North-west              | 10             | 2389        | 766   | 32.52     | 25.03, 41.03              | 116.68        | 92.29           | < 0.001 | 0.10        |
| North-east              | 4              | 820         | 281   | 28.58     | 16.28, 45.17              | 58.84         | 94.90           | < 0.001 | 0.41        |
| North-central           | 9              | 2476        | 759   | 29.58     | 18.54, 43.66              | 313.91        | 97.45           | < 0.001 | Reference    |
| MOD                     |                |             |       |           |                           |               |                 |         |             |
| RSAT                    | 3              | 407         | 73    | 14.41     | 5.32, 33.54               | 23.60         | 91.53           | < 0.001 | < 0.001     |
| Others                  | 11             | 4386        | 1169  | 34.79     | 21.32, 51.24              | 731.36        | 98.63           | < 0.001 | –1.76       |
| HAT                     | 4              | 1129        | 386   | 23.71     | 12.85, 39.58              | 35.54         | 91.56           | < 0.001 | –2.34       |
| ELISA                   | 28             | 8283        | 2941  | 31.62     | 26.39, 37.37              | 740.87        | 96.36           | < 0.001 | –1.90       |
| Dye Test                | 4              | 2025        | 1265  | 74.29     | 57.49, 86.06              | 46.33         | 93.53           | < 0.001 | Reference    |
| Study period            |                |             |       |           |                           |               |                 |         |             |
| 2006–2019               | 37             | 11,708      | 3586  | 28.75     | 23.44, 34.71              | 1420.79       | 97.47           | < 0.001 | 0.003       |
| 1991–2005               | 5              | 1543        | 722   | 44.60     | 27.46, 63.13              | 172.73        | 97.68           | < 0.001 | –2.07       |
| 1976–1990               | 6              | 2956        | 1506  | 41.93     | 27.81, 57.51              | 218.12        | 97.71           | < 0.001 | –2.19       |
| 1960–1975               | 2              | 23          | 20    | 86.82     | 66.13, 95.69              | 0.09          | 0.00            | 0.760  | Reference   |
| Sample size             |                |             |       |           |                           |               |                 |         |             |
| > 450                   | 7              | 6690        | 2524  | 36.64     | 24.15, 51.23              | 721.84        | 99.17           | < 0.001 | 0.913       |
| 301–450                 | 13             | 4704        | 1663  | 30.22     | 19.87, 43.07              | 790.96        | 98.48           | < 0.001 | –0.09       |
| 151–300                 | 18             | 3976        | 1398  | 33.68     | 27.00, 41.08              | 354.78        | 95.21           | < 0.001 | 0.06        |
| ≤ 150                   | 12             | 860         | 249   | 32.59     | 18.93, 50.04              | 160.68        | 93.15           | < 0.001 | Reference    |
| Overall                 | 50             | 16,230      | 5834  | 32.92     | 27.89, 38.37              | 2071.70       | 97.64           | < 0.001 |             |

CI: Confidence interval; ELISA: Enzyme-linked immunosorbent assay; HAT: Haemagglutination test; LAT: Latex agglutination test; PCR: Polymerase chain reaction; RSAT: Rapid slide agglutination test; P*: Inverse variance index; MOD: Method of diagnosis; OR: Odds Ratio; Prev: Prevalence; Q-p: Cochran’s p-value

### Table 3: Pooled prevalence of *Toxoplasma gondii* infections in relation to age, gender and characteristics of study population

| Variables               | No. of Studies | Sample size | Cases | Prev. (%) | Pooled Estimates (95% CI) | Heterogeneity | Meta-regression | P-value | OR (95% CI) |
|-------------------------|----------------|-------------|-------|-----------|---------------------------|---------------|-----------------|---------|-------------|
|                         |                |             |       |           |                           |               |                 |         |             |
| Age*                    |                |             |       |           |                           |               |                 |         |             |
| Children                | 7              | 1365        | 551   | 18.52     | 6.02, 44.63               | 342.16        | 98.25           | < 0.001 | 0.451       |
| Adults                  | 7              | 2790        | 1224  | 36.93     | 29.04, 45.59              | 106.08        | 94.34           | < 0.001 | 0.37        |
| Total                   | 14             | 4155        | 1775  | 29.55     | 21.19, 39.55              | 448.80        | 97.10           | < 0.001 | Reference   |
| Gender                  |                |             |       |           |                           |               |                 |         |             |
| Male                    | 15             | 2391        | 1052  | 31.85     | 21.79, 43.94              | 353.07        | 96.04           | < 0.001 | 0.835       |
| Female                  | 22             | 4263        | 1667  | 30.61     | 22.95, 39.51              | 620.71        | 96.62           | < 0.001 | Reference   |
| Total                   | 37             | 6654        | 2719  | 31.23     | 25.15, 38.02              | 981.86        | 96.33           | < 0.001 |             |
| Target Population       |                |             |       |           |                           |               |                 |         |             |
| Pregnant women          | 20             | 6560        | 2990  | 40.25     | 33.19, 47.73              | 600.05        | 96.83           | < 0.001 | 0.034       |
| Healthy individuals     | 11             | 1372        | 313   | 23.32     | 17.25, 30.75              | 70.24         | 85.76           | < 0.001 | 0.35        |
| HIV patients            | 16             | 4061        | 1163  | 31.68     | 20.53, 45.41              | 767.23        | 98.05           | < 0.001 | Reference   |
| Total                   | 47             | 11,993      | 4466  | 32.94     | 27.61, 38.74              | 1614.29       | 97.15           | < 0.001 |             |

P*: Inverse variance index; CI: Confidence interval; HIV: Human immunodeficiency virus; MOD: Method of diagnosis; OR: Odds Ratio; Prev: Prevalence; Q-p: Cochran’s p-value
analysis. However, the year of conduct of the studies (Q: 13.72; \(df\): 3; \(p\): 0.003), methods of diagnosis employed by the studies (Q: 20.65; \(df\): 4; \(p\) < 0.001) and characteristics of study population (Q: 6.00; \(df\): 2; \(p\): 0.034) might be the possible causes of the heterogeneity in our analysis.

**Discussion**

Adequate understanding of the nationwide burden of toxoplasmosis is essential for effective prevention and control of the disease and its associated complications in immunocompromised individuals. Here, data from individual surveillance studies were harmonised across Nigeria with the sole aim of providing epidemiological information that may serve as a guide for disease monitoring and control in Nigeria.

We observed an overall PP of 32.92% which is consistent with a global report of 34.2% for middle income countries [94]. Our finding is however, higher than the 27.9% reported from Mexico [95] but lower than the 74.7% prevalence reported from Ethiopia [96]. *T. gondii* infection also showed geographical variations across Nigeria with the highest prevalence observed in the south-south region. Corroborating our finding, geographical variations within countries have been documented elsewhere. For instance, in sub-Saharan Africa, studies from Ethiopia reported prevalence of 68.4, 81.4 and 88.2% in the north-western, central and southern regions respectively [19, 22, 97]. More so, in Asia, studies from China documented prevalence of 8.4, 10.3, 12.3 and 21.6% in Anhui, Shanghai, Jilin and Yunnan provinces respectively [98–101]. Differences in weather conditions, eating habits, levels of environmental contamination with *T. gondii* oocysts, personal hygiene and human-cat contacts [96, 102] may be possible explanations for these variations across regions and countries.

The majority of the studies included in the analysis were diagnosed using ELISA. This may be due to the rapidity and accuracy of the method, convenience, ease of use, high sensitivity and specificity, cost effectiveness and the global acceptability of the test [103–105]. Studies diagnosed using the dye test recorded the highest disease prevalence probably due to its high specificity and sensitivity [106]. The serological tests conducted by the individual studies analysed targeted IgG (test for convalescent infection), IgM (test for recent or active infections) and their combination. Interestingly, 85% of the studies conducted on pregnant women detected IgM to identify activeness of infection which is a significant decision making stage to ameliorate or prevent congenital consequences based on stage of pregnancy.

The study showed over 58% decline in the PP of *T. gondii* in Nigeria during the 59 years under review (1960–2019). The decline in recent years could have resulted from controlled movement of cats, improved...
**Fig. 5** Funnel plots of standard error by logit event rate for pooled prevalence of *T. gondii* in **a** Nigeria, **b** northern Nigeria and **c** southern Nigeria.
living standards, life styles and feeding habits that influence the transmission of this pathogen from cats and food animals to man. Other factors that may be responsible for the variations in *T. gondii* prevalence across study periods may include skills of the researchers involved as well as the sensitivity and specificity of the diagnostic methods employed by the individual studies. The majority of the studies were published within the twenty-first century in agreement with global report on *T. gondii* [94]. Increased interest on the pathogen arising from its association with severe complications in immunocompromised conditions like HIV infection, neoplasia and pregnancy may be a possible explanation.

The prevalence of *T. gondii* infection was higher in adults than children concurring reports from Mozambique [107] and Thailand [108]. This may be explained by the habits of eating undercooked roasted meat outside the home and possible intimacy of adults with pet cats. Corroborating a report from China [101], the present study observed a significantly higher prevalence in males than females. Contrary to our finding, Al-Qurashi et al. [7], Domingos et al. [107] and Tegegne et al. [109] reported higher prevalence in females in Saudi Arabia, Mozambique and Ethiopia, respectively.

Prevalence of *T. gondii* showed substantive heterogeneity among eligible studies reported across Nigeria. Meta-regression analysis suggests that this high heterogeneity might be from sources including years in which the studies were conducted, methods of diagnosis employed by the individual studies, characteristics of the study population and not from study locations, sample sizes, as well as age and gender of participants. The sensitivity tests showed that all single-study omission estimates were within the 95% CI of the overall prevalence, suggesting that the pooled prevalence of *T. gondii* was not substantially influenced by any single study. No significant publication bias was observed at alpha level of 0.05 with either Egger’s test or Duval and Tweedie’s method. The findings of the publication bias, sensitivity test and meta-regression substantiate the validity and reliability of the present analysis.

The PP of 40.25% observed among Nigerian pregnant women is higher than the range of 6.1–25.0% reported in countries like China [102, 110], Thailand [108] and Mexico [111]. The high prevalence among pregnant women is of major concern particularly in the ability of *T. gondii* to undergo intrauterine transmission and induce neonatal complications such as miscarriage, chorioretinitis, hydrocephalus, cerebral calcification and foetal death during pregnancy [16, 17]. *T. gondii* PP in HIV-patients in Nigeria was 31.68% concurring with reports of 36.3 and 38.0% from Thailand [112] and South Africa [113] respectively. The present finding is however; grossly lower than reports of 87.45% [17] and 90.0% [114] from Ethiopia. Another major concern with this finding is the risk of HIV changing the asymptomatic course of *T. gondii* in these individuals to a severe cerebral toxoplasmosis which is manifested by headache, disorientation, drowsiness, hemiparesis, reflex changes and convulsion [12–14].

The public health implications of the present finding in a developing country like Nigeria with a collapse primary health care system which is supposed to take care of the health of the rural majority are increased morbidity and associated complications in pregnant women and PLWHA. Two approaches are pertinent to the control of *T. gondii* in Nigeria. First, reviewing disease control programmes in Nigeria to include staging of pregnancy and IgG avidity test particularly in *Toxoplasma*-seropositive pregnant women for possible treatment to prevent congenital consequences and the restriction of cat movement. Second, stakeholders in the area of veterinary public health can also ensure on-farm good agricultural practices as well as standardized veterinary meat inspection to curtail the zoonotic transmission of this pathogen from companion and food animals to man.

The present study has several limitations despite its contribution to knowledge. We could not include some potentially relevant studies which would have added to the understanding of *T. gondii* infection in Nigeria due to insufficiency of data. A whole region (south-east) was not represented in the analysis because no study was published from the region. Studies analysed were published from only 17 of the 36 States in Nigeria including the Federal Capital Territory and were concentrated in the south-west (*n* = 19). Over 95% of the studies included in the analysis relied on serological methods of diagnosis which are unreliable in immunocompromised people [42]. This suggests a possible underestimation of the PP reported by the present study in immunocompromised people in Nigeria. The substantive heterogeneity observed among studies indicates variations across studies which could be due to several factors including year of conduct of study, methods of diagnosis and characteristics of study population. This suggests that the present finding may not represent an absolute *T. gondii* situation in Nigeria, but may provide a guide for disease control policies and the direction for future studies.

**Conclusion**

*T. gondii* is moderately prevalent in Nigeria especially in the south-south region and among pregnant women and HIV-patients. The study showed over 58% decline in the PP of *T. gondii* during the 59 years reviewed (1960–2019). Age was a determining factor in the prevalence of *T. gondii* infection in Nigeria. To effectively control the disease, a holistic approach involving on-farm, environmental, public health and animal components are suggested.
Supplementary information

Supplementary information accompanies this paper at https://doi.org/10.1186/s12889-020-00915-7.

Additional file 1. Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) checklist.
Additional file 2. JBI critical appraisal instrument for studies reporting prevalence data.
Additional file 3. Quality assessment scores for eligible studies.
Additional file 4. Results for Egger regression analysis.
Additional file 5. Results for single study omission analysis.

Abbreviations

AIDS: Acquired immunodeficiency virus; AJOL: African Journals OnLine; CI: Confidence interval; df: degree of freedom; ELISA: Enzyme linked immunosorbent assay; HAT: Haemagglutination test; HIV: Human immunodeficiency virus; ICT: Immunochromatographic test; IP: Inverse variance index; LAT: Latex agglutination test; MOD: Method of diagnosis; PCR: Polymerase chain reaction; PLWHA: People living with HIV/AIDS; PP: Pooled prevalence; Preval: Prevalence; PRISMA: Preferred Reporting System for Systematic Reviews and Meta-Analyses; Q: Cochran's heterogeneity statistic; Qp: Cochran's p-value; RSAT: Rapid slide agglutination test

Acknowledgements

We are grateful to the Editor-In-Chief of Nigerian Journal of Parasitology for providing some of the articles included in the analysis, and to all authors whose articles are included in this study. We also appreciate Mrs. Juliana Tije for assisting in literature search.

Authors’ contributions

SNK: Conceived and designed the study, SNK and MNG: conducted literature search, identified articles, screened articles and extracted data. SNK: Conceived and designed the study, SNK and MNG: conducted literature search. Both authors read and approved the final manuscript.

Funding

Not applicable.

Availability of data and materials

The data supporting the conclusion of this article are all included in the article and Additional files 1-5.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

Author details

1Department of Veterinary Public Health and Preventive Medicine, University of Jos, PMB 2084, Jos, Nigeria. 2Department of Parasitology and Entomology, Modibbo Adama University of Technology, Yola, Adamawa State, Nigeria.

Received: 31 January 2020 Accepted: 29 May 2020
Published online: 06 June 2020

References

1. Dubey JP. Toxoplasmosis of animals and humans. 2nd ed. Boca Raton: CRC Press; 2010.
2. Paul RT, Pierpaolo M. The global burden of congenital toxoplasmosis: a systematic review. Bull Wld Hlth Organ. 2013;91:501–8.
3. Doudou Y, Renaud P, Coralie L, Jacqueline F, Hypolite S, Hypolite M. Toxoplasmosis among pregnant women: high seroprevalence and risk factors in Kinshasa, Democratic Republic of Congo. Asian Pac J Trop Biomed. 2014;4:69–74. https://doi.org/10.1016/S2221-1691(14)60211-2.
4. Pappas G, Roussos N, Falagas ME. Toxoplasmosis snapshots: global status of Toxoplasma gondii seroprevalence and implications for pregnancy and congenital toxoplasmosis. Int J Parasitol. 2009;39:1385–94. https://doi.org/10.1016/j.ijpara.2009.04.003.
5. Al-Hamedi T, Behbehani K. Epidemiology of toxoplasmosis in Kuwait. II. Toxoplasma gondii in the desert rodent Aeniones crassus. Tran R Soc Trop Med Hyg. 1980;74:745–6.
6. Abbas SA, Basalamah A, Serebour F, Alfonso M. The prevalence of Toxoplasma gondii antibodies in Saudi women and the outcome of congenital infection among new-borns in Saudi Arabia. Saudi Med J. 1986;7:346–54.
7. Al-Qurashi AR, Ghandour AM, Obied OE, Al-Mulhim A, Makki SM. Sero-epidemiological study of Toxoplasma gondii infection in the human population in the eastern region. Saudi Med J. 2001;22(1):13–8.
8. Dabritz HA, Conrad PA. Cats and Toxoplasma: implications for public health. Zoon Pub Hlth. 2010;57:34–52.
9. Caballero-Ortega H, Uribe-Salas FJ, Conde-Glez CJ, Cedillo-Pelaez C, Vargas-Villavicencio JA, Luna-Pastén H, et al. Sero-prevalence and national distribution of human toxoplasmosis in Mexico: analysis of the 2000 and 2006 national health surveys. Tran R Soc Trop Med Hyg. 2012;106(1):653–9. https://doi.org/10.1016/j.trstmh.2012.08.004.
10. Welker Y, Molina JM, Poirot C, Ferchal F, Decazes JM, Lagrange P, et al. Interaction between human immunodeficiency virus and Toxoplasma gondii replication in doubly infected monocyte cells. Infect Immun. 1993;56(14):1596–8.
11. Bala S, Englund G, Kovacs J, Wahl L, Martin M, Sher A, et al. Toxoplasma gondii soluble products induce cytokine secretion by macrophages and potentiate in vitro replication of a monocytic strain of HIV. J Eukaryot Microbiol. 1994;41:75.
12. Barratt JL, Harkness J, Marriott D, Ellis JT, Stark D. Importance of non-enteric protozoan infections in immunocompromised people. Clin Microbiol Rev. 2010;23:795–836.
13. Robert-Gangneux F, Darde ML. Epidemiology of and diagnostic strategies for toxoplasmosis. Clin Microbiol Rev. 2012;25(2):264–96. https://doi.org/10.1128/CMR.00513-11.
14. Machala L, Kodym P, Maly M, Gelenky M, Beran O, Jilich D. Toxoplasmosis in immunocompromised patients. Epidemiol Mikrobiol Imunol. 2015;64:59–65.
15. Alonso R, Heiman-Patterson T, Mancall EL. Cerebral toxoplasmosis in acquired immune deficiency syndrome. Arch Neurol. 1984;41:321–3.
16. Torgerson PR, Macpherson CN. The socioeconomic burden of parasitic zoonoses: global trends. Vet Parasitol. 2011;182(1):79
17. Walle F, Kebede N, Tsegaye A, Kassa T. Sero-prevalence and risk factors for Toxoplasmosis in HIV infected and non-infected individuals in Bahir Dar, Northwest Ethiopia. Parasit Vectors. 2013;6(15):8 http://www.parasitesandvectors.com/content/6/1/15.
18. Besieres MH, Benebi A, Cassaing S, Fillaux J, Cambus JP, Berry A, et al. Diagnosis of congenital toxoplasmosis: prenatnal and neonatal evaluation of methods used in Toulouse University Hospital and incidence of congenital toxoplasmosis. Mem Inst Oswaldo Cruz. 2009;104(2):389–92.
19. Gebremedhin EZ, Abebe AH, Tesemma TS, Tullu KD, Medhin G, Vitale M, et al. Sero-epidemiology of Toxoplasma gondii infection in women of child-bearing age in central Ethiopia. BMC Infect Dis. 2013;13(101):95. https://doi. org/10.1186/1471-2334-13-101.
20. Zamir LN, Savva B, Wheeler RB, Hira SK, Luo NP, Kaleebu P, et al. Toxoplasmosis serology in Zambian and Ugandan patients infected with the human immunodeficiency virus. Tran R Soc Trop Med Hyg. 1991;85(2):227–9. https://doi.org/10.1016/0035-9203(91)90034-V.
21. Khalil KM, Ahmed AA, Elrayah LE. Sero-prevalence of Toxoplasma gondii infection in humans in Khartoum state, Sudan. Intern J Trop Med. 2012;7:143–50.
22. Yohanes T, Debalke S, Zemene E. Latent Toxoplasmosis gondii infection in women of childbearing age in central Ethiopia. BMC Infect Dis. 2013;13(101):96. https://doi.org/10.1186/1471-2334-13-101.
23. Miremba MM, Mushil MF, Kivambe C. High sero-prevalence of Toxoplasma gondii IgG antibodies among HIV/AIDS patients with immunological failure attending a tertiary hospital in North-Western Tanzania. Tanz J Hlth Res. 2016;18:1–4.
24. Hagiwara E, Ito A, Shirai A, Kawada K, Okubo T, Amano T, et al. Seroprevalence of anti-Toxoplasmosis IgG Antibody in Japanese patients with HIV infection. Kansenshogakujutsu Zassi. 2001;75:703–4.
71. Ogoina D, Onyemelukwe GC, Musa BO, Obiako R. Sero-prevalence of IgM and IgG antibodies to Toxoplasma infection in healthy and HIV-positive adults from northern Nigeria. J Infect Dev Ctries. 2013;7(5):398–403. https://doi.org/10.3855/jidc.0000.

72. Okoh AEJ, Asgborlahoe DR, Momoh M. Toxoplasmosis in Nigeria- a serological survey. Trop Anim Health Prod. 1981:13:37–43.

73. Okwuzi JO, Odunukwe NN, Ezechi OC, Gbajabiamila TA, Musa AZ, Ezeobi PM, et al. Toxoplasma gondii infection in HIV/AIDS: prevalence and risk factors. Afr J Clin Exp Microbiol. 2014;15:297–102. https://doi.org/10.4314/ajcem.v15i2.7.

74. Okwuzi JO, Osubanjo AO, Ezechi OC, Idowu ET, Odunukwe NN, Okoye RN, et al. Toxoplasma gondii infection among HAART-naive HIV-positive patients and HIV-negatives in Lagos state, Nigeria. Intern J Microbiol Immunol Res. 2013:1:8–1.

75. OlusI T, Gross U, Ajayi J. High incidence of toxoplasmosis during pregnancy in Nigeria. Scand J Infect Dis. 1996;28(6):545–6.

76. Onadeko MO, Joyson DH, Payne RA. The prevalence of Toxoplasma infection among pregnant women in Ibadan, Nigeria. Trop Med Hyg. 1992(5):94–7.

77. Onakoya AO, Odemyi MG, Arilaba OT, Akinsola FB. Ocular findings in acquired immunodeficiency syndrome patients in Lagos, Nigeria. Nig Quart J Hosp Med. 2012;22(1):152–7.

78. Onosakpome EO, Wokem GN, Abah AP. Comparison of ELISA and Rapid Immunomagnetic tests in diagnosis of toxoplasmosis in Port Harcourt, Nigeria. Intern J Trop Dis. 2020;20(1):54–9.

79. Oshinaie EO, Osubanjo NJ. Ojini FL, Danesi MA. A preliminary study of the frequency of focal neurological deficits in HIV/AIDS patients sero-positive for Toxoplasma gondii IgG in Lagos, Nigeria. Nig Quart J Hosp Med. 2010;20(3):104–7.

80. Osyemi TIQ, Synge EMM, Asgborlahe DR, Agbaave R. The prevalence of Toxoplasma gondii antibodies in man in plateau state and meat animals in Nigeria. Trans R Soc Trop Med Hyg. 1985;79(1):31–35.

81. Ouakoualou, Wu, Akanmwc SA, Ofamoh NJ, Onyiaohor IV, Akinde RA, et al. Sero-prevalence of Toxoplasma gondii IgG antibody in HIV-infected patients at the Lagos University teaching hospital. HIV/AIDS Res Palliative Care. 2013;1:101–5. https://doi.org/10.2147/HIV.315532.

82. Ouyoloye SX, Igila-Atisibe M, Ajayi B, Lawan MA. Serological screening for anti-natal toxoplasmosis in Madugun municipal council, Borno state, Nigeria. Afr J Exp Microbiol. 2014;15(2):91–6. https://doi.org/10.4314/ajcem.v15i2.6.

83. Sanoyao AO, Oyibo WA, Itetemew NC, Badaru OS. Sero-prevalence study of parasitic infections among HIV positive and negative patients in Lagos, Nigeria. Afr J Clin Exp Microbiol. 2016;17(2):160–5.

84. Sal W, Rosegger H. Schneeweiss H, Withalm H, Schuhmann G. Serological investigations in Nigeria for anthropozoonoses in human sera: brucellosis, echinococcosis, toxoplasmosis, chlamydial diseases, leptospirosis, rickettsiosis (Coxella burnetii and Rickettsia conorii). J Hyg Epidemiol Microbiol Immun. 1987;31(4):493–5.

85. Sowemimo OA, Wuc T, Leel W, Akanmwc SA, Ofamoh NJ, Ojini FL, Danesi MA. Determination of IgG, IgM and IgA antibodies prevalence and associated risk factors among HIV patients. Asian J Clin Exp Med. 2008;39(2):625.

86. Sowemimo OA, Wuc T, Leel W, Akanmwc SA, Ofamoh NJ, Ojini FL, Danesi MA. A preliminary study of the frequency of focal neurological deficits in HIV/AIDS patients sero-positive for Toxoplasma gondii IgG in Lagos, Nigeria. Nig Quart J Hosp Med. 2010;20(3):104–7.

87. Uneke CJ, Duhlinska DD, Njoku MO, Ngwu BAF. Sero-prevalence of acquired immunodeficiency syndrome patients in Lagos, Nigeria. Afr J Clin Exp Microbiol. 2014;15(2):91–5.

88. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

89. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

90. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

91. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

92. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

93. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

94. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

95. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

96. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

97. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

98. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

99. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

100. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.

101. Wang L, He L, Meng D, Chen Z, Wen H, Fang G, et al. Sero-prevalence and risk factors of Toxoplasma gondii infection in healthy and HIV-positive children in Xuhui District of Shanghai. Chin J Parasitol Parasit Dis. 2016;17(2):88.
case-control study in eastern China. Biomed Res Int. 2015;2015:6. https://doi.org/10.1155/2015/170278.

113. Shimelis T, Tebeje M, Tadesse E, Tegbaru B, Terefe A. Sero-prevalence of latent Toxoplasma gondii infection among HIV-infected and HIV-uninfected people in Addis Ababa, Ethiopia: A comparative cross-sectional study. BMC Res Notes. 2009;2(213):5. https://doi.org/10.1186/1756-0500-2-213.

114. Chemoh W, Sawangjaroen N, Siripaitoon P, Andiappan H, Hortiwakul T, Sermwittayawong N, et al. Toxoplasma gondii – prevalence and risk factors in HIV-infected patients from Songklanagarind hospital, Southern Thailand. Fron Microbiol. 2015;6(1304). https://doi.org/10.3389/fmicb.2015.01304.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.