Substituting Far-Red for Traditionally Defined Photosynthetic Photons Results in Equal Canopy Quantum Yield for CO₂ Fixation and Increased Photon Capture During Long-Term Studies: Implications for Re-Defining PAR

Shuyang Zhen*† and Bruce Bugbee

Crop Physiology Laboratory, Department of Plants Soils and Climate, Utah State University, Logan, UT, United States

Far-red photons regulate shade avoidance responses and can have powerful effects on plant morphology and radiation capture. Recent studies have shown that far-red photons (700 to 750 nm) efficiently drive photosynthesis when added to traditionally defined photosynthetic photons (400–700 nm). But the long-term effects of far-red photons on canopy quantum yield have not yet been determined. We grew lettuce in a four-chamber, steady-state canopy gas-exchange system to separately quantify canopy photon capture, quantum yield for CO₂ fixation, and carbon use efficiency. These measurements facilitate a mechanistic understanding of the effect of far-red photons on the components of plant growth. Day-time photosynthesis and night-time respiration of lettuce canopies were continuously monitored from seedling to harvest in five replicate studies. Plants were grown under a background of either red/blue or white light, each background with or without 15% (50 μmol m⁻² s⁻¹) of far-red photons substituting for photons between 400 and 700 nm. All four treatments contained 31.5% blue photons, and an equal total photon flux from 400 to 750 nm of 350 μmol m⁻² s⁻¹. Both treatments with far-red photons had higher canopy photon capture, increased daily carbon gain (net photosynthesis minus respiration at night), and 29 to 31% more biomass than control treatments. Canopy quantum yield was similar among treatments (0.057 ± 0.002 mol of CO₂ fixed in gross photosynthesis per mole of absorbed photons integrated over 400 to 750 nm). Carbon use efficiency (daily carbon gain/gross photosynthesis) was also similar for mature plants (0.61 ± 0.02). Photosynthesis increased linearly with increasing photon capture and had a common slope among all four treatments, which demonstrates that the faster growth with far-red photon substitution was caused by enhanced photon capture through increased leaf expansion. The equivalent canopy quantum yield among
INTRODUCTION

Plants capture light as fuel for photosynthesis and perceive fluctuations in their radiation environments as signals that trigger changes in plant shape, biochemical composition, developmental stages and resource allocation (Kendrick and Kronenberg, 1994). These adaptive responses are mainly directed by three aspects of light: spectral quality, quantity, and duration. Among the photoreceptors, the red/far-red absorbing phytochromes are of crucial importance in sensing vegetation shade and eliciting shade avoidance responses that often maximize plant growth and fitness in crowded stands (Smith, 1982; Schmitt et al., 1995). Green leaves absorb efficiently in the photosynthetically active region of the spectrum (PAR; 400 to 700 nm) but strongly transmit and reflect longer wavelength far-red photons above 700 nm (Woolley, 1971; Kasperbauer, 1987). This selective attenuation causes the red (R) to far-red (FR) ratio to decrease in forest understories or within dense canopies compared to unfiltered sunlight (Federer and Tanner, 1966; Holmes and Smith, 1977; Casal, 2013). Even before plants are directly shaded, small quantities of additional far-red reflected by neighboring plants are perceived via phytochromes as an early warning signal of competition and induce rapid stem elongation in shade-avoiding seedlings (Ballare et al., 1987). Increased stem (and petiole) extension growth is a most prominent shade avoidance response (Smith and Whitelam, 1997; Franklin, 2008).

Additionally, reduced branching and tillering, lower leaf to stem dry mass ratio, smaller proportion of biomass allocation to the roots, hyponastic (upward bending) leaves with reduced chlorophyll (chl) content, and earlier flowering are among the most frequently observed responses to low R:FR ratios in shade-avoiding species adapted to open habitats (Morgan and Smith, 1979; Kasperbauer, 1987; Halliday et al., 1994; Smith and Whitelam, 1997; Casal, 2012). Those shade-avoidance responses are also induced in agronomic crops under high density monocultures and can cause crops to be more prone to drought and lodging and negatively affect yield, thus are often considered undesirable (Morgan et al., 2002).

Unlike the shade-avoiding species, some plants tolerate shade without showing a strong, or any phytochrome-mediated stem extension growth. Instead, those shade-tolerant species maximize radiation capture through leaf expansion, which is accompanied with thinner leaves and increased fractional biomass allocation to leaves (Gommers et al., 2013). Photosynthetic efficiency is also optimized under shade light environments (i.e., reduced PAR and relatively enriched FR) via adaptive changes in the structure, composition and function of thylakoid membranes, e.g., increased PSII:PSI ratio and lower chl ab ratio (Anderson et al., 1988; Chow et al., 1990a; Valladares and Niinemets, 2008). Both shade-avoiding and -tolerant species perceive shade using the same family of photoreceptors (i.e., phytochromes) and share a number of overlapping responses (e.g., reductions in leaf thickness). The underlying regulatory pathways that lead to the contrasting phenotypic responses between two strategies remain unclear (see Gommers et al., 2013 for an in-depth discussion of potential regulators of shade tolerance responses).

Because crop productivity exhibits a strong linear correlation with radiation intercepted by canopies (Monteith, 1977; Gifford et al., 1984; Beadle and Long, 1985), increased leaf expansion and accelerated canopy closure are desirable traits, especially in annual crops. Recent advances in light emitting diode (LED) technology enable precise control of spectral quality in controlled environment crop production (Massa et al., 2008; Kusuma et al., 2020). Perhaps the most powerful effect is the simulation of shade through FR supplementation, often without reducing light intensity in the PAR region. This change has been reported to promote leaf expansion in various leafy greens, fruiting vegetables, and ornamental species, which increases growth primarily through increased radiation capture (Casal et al., 1987; Goins et al., 2001; Li and Kubota, 2009; Stutte et al., 2009; Park and Runkle, 2017; Kalaitzoglou et al., 2019).

One important difference between simulated shade and natural vegetation shade is that natural shade significantly reduces total photon flux (Casal, 2012). Leaf expansion induced by far-red photons may interact with photon flux as it is partly dependent on supply of photosynthates (Dale, 1988). Heraut-Bron et al. (1999) observed that lowering R:FR ratio induced leaf expansion of white clover when plants were grown under high photosynthetic photon flux density (PPFD) of 320 μmol m$^{-2}$ s$^{-1}$ but not under low PPFD of 110 μmol m$^{-2}$ s$^{-1}$. Leaf expansion induced by far-red supplementation in most simulated shade studies have been conducted under medium to high light conditions. In order to promote beneficial leaf expansion with a high fraction of FR photons, an adequate photon flux in the PAR region is likely needed.

While the photomorphogenic effects of FR photons are largely well characterized, far-red photons (λ > 700 nm) are generally thought to be ineffective for photosynthesis due to their poor photosynthetic efficiency when applied alone (Emerson and Lewis, 1943; McCree, 1972; Inada, 1976). Far-red photons are thus excluded from the current definition of photosynthetic photons. However, pioneering research by Emerson and co-

**Abbreviations**: CUE, carbon use efficiency; DCG, daily carbon gain; LAI, leaf area index, LED, light emitting diode; Canopy $P_{\text{gross}}$ and $P_{\text{net}}$, canopy gross and net photosynthetic rate; PAR, photosynthetically active radiation from 400 to 700 nm; PPFD, photosynthetic photon flux density from 400 to 700 nm; PFD, photon flux density; $R_{\text{dark}}$, dark respiration.
workers found that photosynthetic rate is enhanced under simultaneous illumination of short- and long-wavelength ($\lambda > 685$ nm) photons (Emerson et al., 1957; Emerson and Rabinowitch, 1960). This phenomenon, known as the Emerson enhancement effect, indicates that the effect of different wavelength photons on photosynthesis may not be simply additive. This finding contributed to the subsequent identification of two photosystems with different absorption properties (Hill and Bendall, 1966; Duyzens and Amesz, 1962). More recent work demonstrates that most of the shorter wavelength photons from 400 to 680 nm over-excite photosystem II (PSII), while longer wavelength far-red photons preferentially excite photosystem I (PSI) (Evans, 1987; Hogewoning et al., 2012; Laisk et al., 2014; Zhao et al., 2018). When combining far-red with shorter wavelength photons, the balance of excitation distribution between PSII and PSI is restored, thus leading to increased leaf photochemical efficiency (Zhen and van Iersel, 2017) and photosynthetic rate (Hogewoning et al., 2012; Murakami et al., 2018). Scaling up from leaf to canopy level, Zhen and Bugbee (2020) found that far-red photons (700–750 nm) are equally efficient at driving canopy photosynthesis when added to 400 to 700 nm photons in 14 diverse crop species. These recent findings warrant reconsideration of the photosynthetic value of far-red photons.

Following canopy photon capture and photosynthetic efficiency, the third determinant of daily carbon gain and productivity is the conversion efficiency of carbon fixed in gross photosynthesis into biomass, i.e. carbon use efficiency (Gifford et al., 1984; Bugbee and Monje, 1992). The loss of carbon to respiration at whole-plant/canopy level can be more than 50% of gross photosynthesis (Amthor, 1989). To our knowledge, the effects of far-red photons on canopy quantum yield (moles of CO$_2$ fixed in gross photosynthesis per mole of absorbed photons; a measure of photosynthetic photon use efficiency) and carbon use efficiency (CUE = ratio of C incorporated into biomass to C fixed in gross photosynthesis) during long-term crop cultivation have not been studied. Most previous studies have added far-red photons as a supplement, which complicates interpretation of the results because total photon flux from 400 to 750 nm is not constant.

Our objective was to quantify the effects of far-red substitution for 400–700 nm photons on radiation capture, canopy quantum yield, carbon use efficiency, and biomass allocation of a model crop lettuce. This quantitative approach can provide a mechanistic understanding of the effect of far-red photons on plant growth. Continuous measurement of canopy quantum yield for CO$_2$ fixation can provide additional evidence for changing the definition of photosynthetic active radiation to include far-red photons (700–750 nm).

**MATERIALS AND METHODS**

**Plant Materials**

Lettuce (*Lactuca sativa* cv. Waldmann’s Dark Green) was seeded in 1.7 L containers filled with calcined clay (Greens Grade; Profile, Buffalo Grove, IL) in a glass-covered greenhouse and then moved into a multi-chamber gas exchange system two to four days after emergence. The calcined clay substrate was rinsed with de-ionized water prior to use as described by Adams et al. (2014) and soaked with a hydroponic nutrient solution (Utah hydroponic refill solutions for dicots with FeCl$_3$ and iron-chelating agent EDDHA doubled in concentration; USU Crop Physiology Laboratory, 2018). The pH of the hydroponic solution was adjusted to 5.8 with nitric acid and the electric conductivity of the solution was 1 ± 0.1 mS/cm. This low pH minimized bicarbonate effects on CO$_2$ fluxes from the root-zone (van Iersel and Bugbee, 2000). Seedlings were selected for uniformity and thinned to one plant per container. Five replicate studies were conducted.

**Light Treatments and Growing Conditions**

Sixteen uniform seedlings were moved into a steady-state gas exchange system with four acrylic chambers (100 L/chamber with four plants per chamber; 36 cm x 47 cm x 59 cm; w x l x h), similar to the multi-chamber gas exchange system described by van Iersel and Bugbee (2000). Each chamber consisted of one spectral treatment, with cool white, red, blue, and far-red LEDs (Ray 22; Fluence Bioengineering, Austin, TX, USA) placed on top of the chambers. The four light treatments were: white 350, red/blue (RB) 350, white 300 + far-red (FR) 50, and RB 300 + FR 50 (see Figure 1 for spectral distributions). The numbers following each type of light (e.g. RB 300) indicate the average photon flux density in $\mu$mol m$^{-2}$ s$^{-1}$ [±1 $\mu$mol m$^{-2}$ s$^{-1}$]; measured at 13 locations across the chamber floor area of 0.17 m$^2$ using a spectroradiometer (SS-110; Apogee Instruments, Logan, UT, USA)]. The blue (400–500 nm): green (501–600 nm): red (601–700 nm) ratio was in 31.5:45.2:23.3 in white 350 and 31.5:0.1:68.4 in RB 350. Phytocrome photoequilibrium [PPE, an indicator of the relative amount of active phytochrome in the far-red-absorbing $P_f$ form] under each light spectrum was calculated using the phytochrome photoconversion coefficients by Sager et al. (1988) (Table 1).

Chamber walls were lined with highly reflective Mylar to eliminate side lighting and increase light uniformity. The reflective walls also help to simulate light environment of a canopy because they reflect FR photons similar to neighboring plants. Two of the treatments had FR photons (700–750 nm) substituting for ~15% of the conventionally defined white or RB photosynthetic photons. All four treatments contained 31.5% blue photons and equal total photon flux of 350 $\mu$mol m$^{-2}$ s$^{-1}$ from 400 to 750 nm. Photoperiod was 14/10 h light/dark. Daily light integral was 17.64 mol m$^{-2}$ d$^{-1}$.

Plants were grown under elevated CO$_2$, with the CO$_2$ concentration [CO$_2$] in the pre-chamber air streams enriched to 800 $\mu$mol mol$^{-1}$. CO$_2$ enrichment is a common practice in controlled environment crop production. Controlling CO$_2$ to a constant elevated level also reduces noises in the gas exchange measurements caused by un-steady [CO$_2$] in the ambient air in urban areas. Air flow rate through the chambers was gradually increased from 11 to 37 mmol s$^{-1}$ (15 to 50 standard L min$^{-1}$) from seedling to mature plant stage. Air inside the chambers was mixed with fans (793 L min$^{-1}$; Model A36-B10A-15T2-000, Globe Motors). The [CO$_2$] inside the chambers ranged from
Shoot fresh mass (g) 130\(^b\) 179\(^a\) 129\(^b\) 167\(^a\) (PPE) (0.87) (0.82) (0.84) (0.70) 

Leaf photon absorption 0.927 0.798 0.838 0.686 

Chl content (µmol g\(^\text{dry mass}\)\(^{-1}\)) 40.0 37.5 38.5 34.0 

Leaf area (cm\(^2\)) 309\(^1\) 442\(^3\) 311\(^4\) 422\(^8\) 

% root 16.1\(^a\) 14.7\(^b\) 16.6\(^a\) 15.3\(^a\) 

Leaf area (cm\(^2\)) 309\(^1\) 442\(^3\) 311\(^4\) 422\(^8\) 

Leaf mass per area (g m\(^{-2}\)) 28.2\(^a\) 25.5\(^b\) 27.9\(^b\) 26.3\(^b\) 

Chl content (µmol m\(^{-2}\)) 27.2\(^a\) 21.8\(^b\) 26.7\(^a\) 21.3\(^b\) 

Leaf photon absorption 0.927 0.798 0.838 0.686 

Leaf photon absorption 0.927 0.798 0.838 0.686 

400 to 750 nm. 

Leaf area were expressed on a per chamber basis (i.e. the sum of four plants). Data represent means from five replicate studies with different letters indicate significant differences between means \(P \leq 0.05\); \(n = 5\). Leaf photon absorption of incident photons under each spectral treatment (mol \(\text{absorbed}\) mol\(^{-1}\) \(\text{chamber}\)) was integrated from 400 to 750 nm. 

The pre- and post-chamber air streams were sampled every second using two infra-red gas analyzers (reference and differential IRGA; LI-6252; LI-COR, Lincoln, NE). Each chamber was sampled for 30 s, and then the tubing that connected the chambers to the differential IRGA was purged for 40 s before the next chamber was sampled. Canopy gas exchange rate \([\text{net photosynthetic rate during the light period (P}_{\text{net, light}})]\) and dark respiration rate \([-\text{R}_{\text{dark}}; \text{negative values}) in \mu\text{mol CO}_2 \text{ m}^{-2} \text{ground area s}^{-1}\) was calculated from the mass flow rate and delta \([\text{CO}_2]\) between the pre- and post-chamber air streams and recorded using a datalogger (CR1000; Campbell Scientific, Logan, UT, USA) throughout the entire course of the study. 

To eliminate the mole fraction dilution of \(\text{CO}_2\) analysis by water vapor, the air streams sampled by the IRGAs were first passed through nafion dryers (Perma Pure, Lakewood, NJ, USA) and then columns of magnesium perchlorate to completely remove water vapor. 

To determine the photosynthetic value of FR photons, far-red LEDs in the two FR substitution treatments were turned off for 30 to 40 min during the days prior to harvest. 

\(\text{P}_{\text{net, light}}\) and \(-\text{R}_{\text{dark}}\) was averaged over the 14 h of light period and 10 h of dark period respectively, on a daily basis. The averaged values (in units of mol\(\text{CO}_2\) m\(^{-2}\) ground area h\(^{-1}\)) were used in the subsequent calculations of gas exchange parameters \((\text{Eqs 1–3})\). 

Canopy gross photosynthesis \((\text{P}_{\text{gross}}; \text{g d}^{-1})\) was calculated as: 

\[
\text{P}_{\text{gross}} = (\text{P}_{\text{net, light}} + |\text{R}_{\text{dark}}|) \times 14 \times 0.17 \times 30 \quad (\text{g mol}^{-1})
\]

Where \(|\text{R}_{\text{dark}}|\) was the absolute value of dark respiration. 14 was the light period in hours, and 0.17 was the chamber ground area in m\(^2\). 30 represents grams dry mass per m ole of \(\text{CO}_2\) assimilated, assuming a carbon content of 0.4 g g\(^{-1}\) in plant tissues. This calculation of \(\text{P}_{\text{gross}}\) also assumes that respiration rate was similar in the light and dark. There is ongoing research and discussion on whether respiration rate in the light is similar, higher, or lower than respiration in the dark (Sharp et al., 1984; Atkin et al., 1998; Frantz et al., 2004 and citations therein). We assumed that respiration rate in light and dark was the same, which is the standard assumption in canopy photosynthesis research (van Iersel, 2003). 

Daily carbon gain (DCG; a measure of canopy growth rate in g d\(^{-1}\)) was calculated as: 

\[
\text{DCG} = \text{P}_{\text{gross}} - \text{R}_{\text{dark}}
\]
DCG = \left( P_{\text{net,light}} \times 14\text{ h} - |R_{\text{dark}}| \times 10\text{ h} \right) \times 0.17 \text{ m}^2 \	imes 30 \text{ g mol}^{-1} \quad (2)

Where 14 was the light period in hours, and 10 was the dark period in hours.

Carbon use efficiency (CUE; the ratio of daily net carbon gain to the total amount of carbon fixed in gross photosynthesis) was calculated from Eqs (1) and (2) as:

\[
\text{CUE} = \frac{\text{DCG}}{\text{P}_{\text{gross}}} \quad (3)
\]

**Imaging of Canopy Ground Cover**

Plants were taken out of the gas exchange chambers for 2 to 3 min daily for top down photos of the canopies, taken with a digital camera placed 130 cm above the plant base. Images were analyzed using a Python program, available as open source at https://github.com/jakobottar/green-pixel-analysis. The fraction of canopy ground cover was calculated as the ratio of green pixels (plant tissues) to the total pixel count of an area equal to the chamber ground area as described by Klassen et al. (2004).

**Growth Parameters and Chlorophyll Content**

Plants were destructively harvested 17 to 20 days after transferred into the gas exchange chambers, when the largest canopies were near canopy closure. Shoot fresh weight and total leaf number of all four plants in each spectral treatment were recorded. Leaf chlorophyll content (\(\mu\text{mol m}^{-2}\)) was measured on five representative leaves per plant and averaged over four plants per treatment (MC-100 chlorophyll meter; Apogee Instruments, Logan, UT). Total leaf area per chamber was measured using a leaf area meter (LI-3000; LI-COR, Lincoln, NE, USA). Shoots and roots were oven-dried to a constant mass (at least 48 h at 80°C). Root percent mass was calculated as root dry mass/total shoot and root dry mass). Leaf mass per area (\(\text{g m}^{-2}\)), a measure of leaf thickness, was calculated as leaf dry mass/total leaf area.

**Leaf Photon Absorption and Canopy Photon Capture**

Leaf light absorbance was determined on the day of harvest using a spectroradiometer (PS300; Apogee Instruments) similar to the method of Nelson and Bugbee (2015). The absorbance spectrum was then multiplied by the spectral output of LEDs to obtain the leaf photon absorption under each spectral treatment (mole of photons absorbed by a single layer of leaf per mole of incident photons).

Canopy photon capture (mole of photons absorbed per mole of incident photons) was estimated by multiplying the fraction of canopy ground cover by the leaf photon absorption under each spectral treatment. This gave a good estimate in small canopies with only one layer of leaves. However, the canopy photon capture would be underestimated in larger canopies that had over-lapping layers of leaves. To account for this, the number of leaf layers in each canopy at harvest was calculated as total leaf area at harvest \(\left(\text{m}^2\right)\) divided by the projected canopy ground cover \(\left(\text{m}^2\right)\). The projected canopy ground cover (or projected canopy leaf area) was determined from top down photos \((\text{i.e. percent ground cover multiplied by chamber ground area of 0.17}\text{ m}^2)\). Canopies of lettuce had relatively horizontal leaf orientation and typically did not form over-lapping leaves until one week prior to harvest. The projected canopy leaf area was thus used to estimate total canopy leaf area in young canopies. When leaves started to overlap, total canopy leaf area was interpolated from a regression function (exponential growth) fitted to the canopy leaf area estimated for young canopies and the final total canopy leaf area at harvest. The number of layers of leaves during the week prior to harvest was then calculated as the interpolated total canopy leaf area on each day divided by the projected canopy leaf area.

Using lettuce grown under spectral treatment RB 300 + FR 50 as an example, a single layer of leaves absorbed 0.798 moles of photons (integrated over 700 to 750 nm) per mole of incident photons (Table 1). Photon absorption with more than one layer of leaves was estimated using the Beer–Lambert equation: canopy photon absorption = \(1 - e^{-k \times \text{LAI}}\); where the extinction coefficient \((k = 1.6)\) was derived by setting the photon absorption to 0.798 and LAI to 1. Thus, with two layers of overlapping leaves photon absorption would approximately equal to \(1 - e^{-1.6 \times 2} = 0.959\) mol of photons absorbed per mole of incident photons. With an average of 2.6 layers of leaves at harvest, photon absorption in areas covered by leaves was adjusted to \(1 - e^{-1.6 \times 2.6} = 0.984\) mol of photons absorbed per mole of incident photons.

**Canopy Quantum Yield**

Canopy quantum yield for CO₂ fixation (mole of CO₂ fixed in gross photosynthesis per mole of photons absorbed from 400 to 750 nm) was calculated as:

\[
\text{Canopy quantum yield} = \frac{\text{P}_{\text{gross}}}{\text{canopy absorbed photons}} \quad (4)
\]

Note that \(\text{P}_{\text{gross}}\) here is expressed in units of moles of CO₂ fixed per mole of incident photons for the simplicity of the equation. To convert \(\text{P}_{\text{gross}}\) from g d⁻¹ in Eq. (1) to mol CO₂ mol⁻¹incident photons, the \(\text{P}_{\text{gross}}\) values in g d⁻¹ need to be first divided by 30 g mol⁻¹ (grams of dry mass per mole of carbon assimilated), then divided by 0.17 m² (chamber ground area), and then divided by 17.64 mol incident photons m⁻² d⁻¹ (total daily photon flux density at canopy level).

**Statistical Analysis**

Data were analyzed using regression (linear and sigmoid) in Sigmaplot (Systat Software, San Jose, CA) and ANOVA in Statistical Analysis Systems (SAS Institute, Cary, NC, USA). Mean separation was performed using Fisher’s protected least significant difference (LSD, \(P = 0.05\)). Canopy gas exchange measurements from two representative replicate studies were included in data analysis. Growth parameters and chlorophyll content were obtained from all five replicate studies.
 RESULTS

Biomass, Leaf Area, and Chlorophyll Content
Lettuce grown under the two control treatments without far-red (white 350 and RB 350) had nearly identical dry mass and leaf area at harvest. In contrast, both treatments with 15% FR substitution, either with a white or RB background light, had 29 to 31% increase in total biomass (shoot and root) and a 36–43% increase in leaf area (Table 1). Leaves of plants grown with far-red were thinner with a 4–10% decrease in leaf mass per area, ~20% lower chlorophyll content, and less efficient absorption of the incident photon on single leaf basis (Table 1). Dry mass partitioning to the roots, indicated by % root, did not differ significantly among the four spectral treatments even though it tended to be slightly reduced in the two treatments with FR (Table 1).

Canopy Gas Exchange and Carbon Use Efficiency
Canopy P_{net}, light and R_{dark} under all four spectral treatments increased exponentially from seedling to young plant stage over the first two weeks, followed by slower increases during the third week as plants quickly approached canopy closure (Figure 2; also see Supplementary Figure S1 for additional dataset). Gas exchange rates were identical among all four treatments when uniformly sized seedlings were first moved into the chambers and remained similar between the white 350 and RB 350 treatments throughout the entire course of the study. Plants grown under the two FR substitution treatments, however, gradually showed higher P_{net}, light and R_{dark} (Figure 2) and gained more carbon per day than the control treatments (Figure 3). These measurements were consistent with the differences in plant dry mass under the four treatments. At harvest, dry mass predicted from the cumulative daily carbon gain obtained from gas exchange measurement was 103.9 ± 3.4% of the measured total dry mass in two representative replicate studies.

A decrease in P_{net}, light was detected within 5 min (note that each chamber was sampled every 4 min and 40 s) when the far-red LEDs in the two FR substitution treatments were turned off (Figure 2; insets), indicating that far-red photons directly contribute to canopy photosynthesis.

Carbon use efficiency (CUE) was lower for seedlings and gradually increased as plants matured (Figure 4). CUE for mature plants was similar under all four treatments, with an average of 0.61 ± 0.02 (SD) over the last six days prior to harvest.

Ground Cover and Canopy Photon Capture
Fractional ground cover and canopy photon capture increased over time in a similar manner as gas exchange rates and daily carbon gain (Figure 5; also see Figures 2, 3). The FR substitution treatments consistently had higher fractional ground cover than the control treatments, especially after one week into the study (Figures 5A, B).

The higher fractional ground cover translated into higher canopy photon capture in the FR substitution treatments, although to a smaller extent (Figures 5C, D). This was because 1) leaves of plants grown with far-red were thinner with lower chlorophyll content and less efficient photon absorption especially in the green and far-red regions, and 2) far-red photons in the 15% FR substitution treatments were less efficiently absorbed than the 400 to 700 nm photons in the control treatments (Figure 6 and Table 1). Leaf photon absorption under each of the four light treatments was shown in Table 1.

Canopy Quantum Yield
Canopy quantum yield (moles of CO\textsubscript{2} fixed in gross photosynthesis per mole of absorbed photons integrated over 400 to 750 nm) was similar among all four light treatments, with an average of 0.057 ± 0.002 (SD) (Figure 7). This indicates that the absorbed far-red photons were as efficient as red/blue and white photons.
Correlation Between Canopy Photosynthesis and Photon Capture

Canopy $P_{\text{gross}}$ of all four treatments increased linearly with increasing photon capture and had a common slope; daily carbon gain of all four treatments also showed a common linear correlation with canopy photon capture (Figure 8).

DISCUSSION

Beneficial leaf expansion elicited by far-red has been increasingly explored as means to improve production efficiency of high-value leafy green vegetables and ornamentals in greenhouses and indoor vertical farms (Goins et al., 2001; Li and Kubota, 2009; Stutte et al., 2009; Park and Runkle, 2017; Meng et al., 2019). Most of the previous studies added far-red photons as a supplement to photosynthetic photons within 400–700 nm, and the enhanced plant growth has been, in general, attributed solely to increased radiation capture through leaf expansion.

Far-red supplementation comes with an energy cost and it remains unclear whether it is cost effective to add far-red photons or to increase the traditionally defined photosynthetic photons (400 to 700 nm) by the same amount. Far-red photons contain lower energy and thus can be generated with higher efficacy (moles of photons per unit of electric energy) than red, blue, and green photons using current LED technology (Kusuma et al., 2020). Recent research on the photosynthetic effect of far-red photons has provided compelling evidence that far-red photons (700–750 nm) significantly increase leaf and canopy photosynthesis when added to photons within 400 to 700 nm (Zhen and Bugbee, 2020 and citations therein). These results indicate that far-red photons should be substituted for 400 to 700 nm photons instead of supplemented. Substitution facilitates the separation of the direct effect of far-red photons on canopy photosynthesis (by comparing the canopy quantum yield in treatments with and without far-red substitution) from their indirect effect on plant growth through altering canopy morphology and radiation capture. This provides a mechanistic understanding of the effect of far-red photons on plant growth during long-term crop cultivation.

Leaf Expansion and Phytochrome Photoequilibrium

The increased leaf expansion in the far-red substitution treatments was presumably caused by a decrease in phytochrome photoequilibria (PPE) compared to the control treatments (white or red/blue light) (Table 1). PPE has been widely used to estimate the fraction of phytochromes in the active $P_r$ form relative to the total phytochrome pool (Sager et al., 1988). An inverse linear response between stem elongation rate PPE has been well-established in shade-avoiding species (Morgan and Smith, 1976; Morgan and Smith, 1979) but the relationship between leaf expansion and PPE appears to be species-specific (Smith and Whitelam, 1997; Gommers et al., 2013). The reverse correlation between leaf expansion and PPE in this study is similar to the findings by Park and Runkle (2017). However, the slope of the relationship varied with the background light (Supplementary Figure S2). It is noteworthy that the total canopy leaf area was nearly identical in both treatments with far-red substitution,
large fraction of green photons and a much smaller fraction of red photons compared to the red/blue light (Figure 1). The absorption spectrum of phytochrome ranges from 300 to 800 nm (Sager et al., 1988). Because chlorophylls more efficiently absorb red photons than green photons, the extent of chlorophyll masking effect on the phytochrome response most likely differed under the white and red/blue light. This may be responsible for the different relationships between leaf expansion and estimated PPE under white and red/blue light.

**Leaf Light Absorptance, Canopy Ground Cover, and Radiation Capture**

The increased yield in response to FR supplementation is often attributed to increased photon capture (Goins et al., 2001; Li and Kubota, 2009; Stutte et al., 2009; Park and Runkle, 2017), but canopy photon capture is rarely quantified. Most studies report only leaf area at harvest, which non-linearly correlates with canopy photon capture due to overlapping of leaves. Goins et al. (2001) reported that lettuce grown with 12% added far-red photons consistently had higher canopy ground cover over a two-week growing period than plants that received equal photon flux in the PAR region. As expected, the increased leaf expansion under the far-red substitution treatments led to greater fractional canopy ground cover (Figures 5A, B). Fractional ground cover was found to be highly correlated with canopy photon absorption ($r^2 = 0.99$) of lettuce (Klassen et al., 2004), but the relationship was nonlinear and varied over time. Other factors, including leaf optical properties and the spectral distribution of incident light, can significantly alter spectral effects on canopy photon capture.

Leaf thickness and chlorophyll content affect leaf optical properties (Vogelmann, 1993). Reduced leaf chlorophyll concentration is a nearly universal response to FR irradiation among a wide range of species, regardless of changes in leaf expansion (Morgan and Smith, 1979; Casal et al., 1987; Smith and Whitelam, 1997; Li and Kubota, 2009; Meng et al., 2019). Several causes for this decrease in chlorophyll content have been suggested, including 1) limited photosynthetic supply in shade-avoiding species that had reduced leaf expansion, 2) a dilution effect when leaf expansion is promoted, and 3) a direct effect on chlorophyll biosynthesis due to reduced amount of active Pfr relative to the total pool of phytochromes (see Casal et al., 1987 and references therein). The stimulatory effect of far-red photons on leaf expansion has likely contributed to the 'dilution' of chlorophyll content per unit leaf area. The reduction in leaf chlorophyll content and thickness (indicated by the lower leaf mass per area in Table 1) of lettuce grown with far-red substitution resulted in low leaf light absorptance, especially in the green and far-red regions of the spectrum (Figure 6).

In addition, the far-red photons were less efficiently absorbed by the leaves, which reduced canopy photon capture (Table 1). As a result, the magnitude of increase in canopy photon capture in the FR substitution treatments was smaller than indicated by the fractional ground cover. Therefore, the use of total leaf area or canopy ground cover would likely lead to over-estimation of canopy photon capture.
Predicted Dry Mass From Gas Exchange Measurements

All gas exchange systems measure the carbon assimilation rate in moles of carbon (CO₂) fixed per unit time. To convert \( \text{P}_{\text{gross}} \) and DCG from moles of carbon to grams of biomass, we assumed that the canopy carbon content of lettuce was 0.4 g carbon g\(^{-1}\) dry mass. Carbon fraction in plants has often been assumed to be 43 to 45% (Epstein and Bloom, 2005), but this fraction varies widely with the composition of the biomass (carbohydrates, lignin, proteins, lipids; Amthor, 2010). The carbon fraction thus varies with type of vegetation (leaves, stems and roots) and species (Vertregt and Penning de Vries, 1987; Monje and Bugbee, 1998). It also varies during ontogeny (Ho, 1976; Hadley and Causton, 1984).

Growing conditions also affect carbon fraction. Becker et al. (2015) found that the carbon fraction in leaves of green lettuce decreased from 0.4 to 0.38 g g\(^{-1}\) as nitrogen availability increased. Monje and Bugbee (1998) reported that leaf carbon content of wheat increased from 0.38 under ambient CO₂ to 0.4 g g\(^{-1}\) under elevated CO₂. These changes were small but reproducible. Lettuce is about 85% leaves (Table 1). Roots tend to have a higher carbon content than leaves (0.44 to 0.45 g g\(^{-1}\) in roots of wheat; Monje and Bugbee, 1998). The weighted carbon content of the combined leaves and roots of the plants in these studies in thus estimated to be 0.4 g g\(^{-1}\). With this value, the dry mass at harvest predicted from the cumulative DCG averaged 103.9% of the measured total dry mass. This close match validates the accuracy of the gas exchange measurements.

Canopy Quantum Yield: Short-Term Versus Long-Term Response to Far-Red Photons

Measurements of canopy gas exchange rate coupled with canopy photon absorption enable the calculation of canopy quantum yield, a measure of photosynthetic efficiency expressed as moles of CO₂ fixed in gross photosynthesis per mole of photons...
absorbed. We calculated quantum yield using wavelengths from 400 to 750 nm. To our knowledge, this is the first report of the long-term effect of far-red photons on canopy quantum yield. The lack of difference in canopy quantum yield among the four spectral treatments indicates that the absorbed far-red photons were equally efficient as red/blue or white photons (Figure 7). This is also indicated by the nearly identical relationship between canopy P\text{gross} and photon capture among the treatments (Figure 8A).

On an incident photon flux basis, Zhen and Bugbee (2020) showed that the short-term (hours) response of canopy photosynthesis to far-red photons from 700 to 750 nm was equal to 400 to 700 nm photons when applied simultaneously. Because far-red photons are less efficiently absorbed than 400 to 700 nm photons, the far-red photons had a higher canopy quantum yield on an absorbed photon basis. This difference between short-term and long-term responses indicates that the efficiency of light utilization for photosynthesis may have acclimated to far-red radiation during long-term growth. Possible changes include modifications in leaf thickness, chlorophyll content (Table 1), chl a:b ratio (Gommers et al., 2013; Kalaitzoglou et al., 2019), PSII:PSI ratio, electron transport and photosynthetic capacity, rubisco content and activity (Chow et al., 1990a; Chow et al., 1990b; Anderson et al., 1995), as well as in the biosynthesis of photoprotective pigments carotenoids and anthocyanins (Li and Kubota, 2009; Stutte et al., 2009; Kalaitzoglou et al., 2019).

**Carbon Use Efficiency**

Respiration is often assumed to be a fixed fraction of gross photosynthesis in modeling maximum productivity per unit input of solar energy (Zhu et al., 2008; Zhu et al., 2010; Amthor, 2010). A constant CUE of ~0.5 has been widely used in forest growth models (Waring et al., 1998; DeLucia et al., 2007). Gifford (1994) reported that the ratio of respiration to gross photosynthesis of whole plants (which equals to 1—CUE) was constant around 0.4 (a CUE of ~0.6) in diverse annual and perennial species and was minimally affected by plant age and growth temperature (15 to 30°C). Monje and Bugbee (1998) found that CUE of wheat grown under near-optimal conditions (high light and ample water and nutrients) was constant around 0.6 (except during the first week after emergence and final week when plants senesced) despite changes in carbon partitioning to...
leaves, stems, and seeds. The response was not affected by [CO₂] even though canopy quantum yield and biomass production increased significantly when [CO₂] was elevated to 1200 mol mol⁻¹ (Monje and Bugbee, 1998).

However, reports have conflicted on whether CUE stays constant among species, environmental conditions, and developmental stages (Amthor, 2000; Cannell and Thornley, 2000). A constant CUE means that plants always respire the same fraction of carbohydrates regardless of growth rate. To better understand the link between respiration and growth, respiration is frequently divided into two functional components—growth respiration (that is determined by the synthesis of new biomass (proportional to growth rate) and maintenance respiration (that keeps existing biomass functional) and relative growth rate and growth and maintenance respiration that is affected by the chemical composition of the biomass being synthesized (Penning de Vries et al., 1974; Amthor, 2010). The respiration required to maintain existing biomass per unit time (i.e. maintenance respiration coefficient) is affected by metabolic rate (associated with temperature), but it can also be influenced by the chemical composition, especially tissue nitrogen content (Cannell and Thornley, 2000). van Iersel (2003) showed that CUE can be expressed as function of relative growth rate and growth and maintenance respiration coefficients. Because of the ontogenetic changes in plant tissue composition and relative growth rate, a constant CUE is unlikely unless growth and/or maintenance respiration coefficient change concurrently with relative growth rate. van Iersel (2003) further demonstrated that CUE of lettuce decreased from 0.6 in young plants (24-day old) to 0.2 in old plants (66-day old). This was mainly due to the decrease in relative growth rate and increase in the fraction of maintenance respiration with increasing plant size. The carbon use efficiency of fast-growing and compositionally simple crops like lettuce is expected to be relatively more constant under optimal conditions, where a higher relative growth rate would reduce the importance of maintenance respiration (van Iersel, 2003). Franztz et al. (2004) found that the CUE of rapidly growing lettuce (~15 to 35 day old) under optimal conditions (high light, elevated CO₂, and a constant day/night temperature of 25°C) was stable around 0.62. Decreasing night temperature from 34 to 17°C reduced nighttime respiration rate and allocation to shoot/root, but only slightly altered CUE.

In this study, CUE of young/newly matured lettuce plants (~16–22 days after emergence) was similar under all four treatments, with an average of 0.61 ± 0.02. This is consistent with the values reported for lettuce of similar age by van Iersel (2003) and Franztz et al. (2004). However, CUE of younger seedlings was not determined in the previous studies because of the difficulties in obtaining accurate gas exchange measurements with small plants. With an improved gas-exchange system, we observed that CUE of young lettuce seedlings gradually increased within the first 2 weeks after emergence, similar to the gradual increase in CUE of wheat during the first 8 days after emergence (Monje and Bugbee, 1998). A gradual increase in CUE was also reported in young vinca seedlings as the ratio of respiration to photosynthesis decreased during the first few weeks of plant development (van Iersel and Seymour, 2000). The similarity in CUE of young/newly matured lettuce plants among the spectral treatments indicates that the phenotypical changes caused by FR radiation had little effect on CUE.

CONCLUDING REMARKS

Overall, we found that neither canopy quantum yield nor carbon use efficiency were affected when substituting far-red for traditionally defined photosynthetic photons. This indicates that the absorbed far-red photons (700–750 nm) were equally efficient for photosynthesis when acting synergistically with 400–700 nm photons. Crop yield increased with far-red substitution due to increased leaf expansion and canopy radiation capture mediated by phytochromes during long-term plant cultivation. These data, coupled with previous studies, provide compelling evidence that the current definition of photosynthetically active radiation should be extended to include photons from 700 to 750 nm.

DATA AVAILABILITY STATEMENT

All datasets presented in this study are included in the article/Supplementary Material.

AUTHOR CONTRIBUTIONS

SZ and BB designed the experiment. SZ performed the experiment, analyzed data, and wrote the first draft. SZ and BB discussed the data and revised the manuscript.

FUNDING

This work was supported by the Utah Agricultural Experiment Station, Utah State University; by the NASA-CUBES project award number NNX17AJ31G; and by the USDA-NIFA-SCRI award number 2018-51181-28365 (LAMP Project). Any opinions, findings, and conclusions or recommendations expressed in this material are those of the authors and do not necessarily reflect the views of the National Aeronautics and Space Administration (NASA) or the USDA.

ACKNOWLEDGMENTS

We thank Jakob Johnson for helping with the image analysis of canopy ground cover and Alec Hay for technical support.
SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2020.581156/full#supplementary-material

SUPPLEMENTARY FIGURE 1 | Canopy net photosynthetic rate ($P_{net} \, \text{mmol CO}_2 \text{m}^{-2} \text{s}^{-1}$) of lettuce under red/blue (RB) and white, with or without far-red substitution. The numbers following each type of light (e.g. RB 350) indicate photon flux density in mmol $\text{m}^{-2} \text{s}^{-1}$. Plants were seeded in a greenhouse and moved into a multi-chamber gas exchange system two days after seedling emergence (indicated by Day 0 on the x-axis). Downward pointing arrows indicate when far-red LEDs were turned off.

SUPPLEMENTARY FIGURE 2 | Relationships between estimated phytochrome photoequilibria (PPE) and leaf area of lettuce grown with or without far-red photons under two background light spectra—white and red/blue (RB). The decrease in the estimated PPE under each background light was due to a 15% far-red substitution. All four spectral treatments had equal total photon flux of 350 mmol $\text{m}^{-2} \text{s}^{-1}$ (400 to 750 nm) and 31.5% blue photons.

REFERENCES

Adams, C., Jacobson, A., and Bugbee, B. (2014). Ceramic aggregate sorption and desorption chemistry: implications for use as a component of soilless media. J. Plant Nutr. 37, 1345–1357. doi: 10.1080/01904167.2013.837921

Amthor, J. S. (1989). Respiration and crop productivity (New York, USA: Springer-Verlag).

Amthor, J. S. (2000). The McCree–de Wit–Penning de Vries–Thornley respiration paradigm: 30 years later. Ann. Bot. 86, 1–20. doi: 10.1006/anbo.2000.1175

Amthor, J. S. (2010). From sunlight to photomass: on the potential efficiency of converting solar radiation to phyto-energy. New Phytol. 188, 939–959. doi: 10.1111/j.1469-8137.2010.03505.x

Anderson, J. M., Chow, W. S., and Goodchild, D. J. (1988). Thylakoid membrane organisation in sun/shade acclimation. Funct. Plant Biol. 15, 11–26. doi: 10.1071/PP880011

Anderson, J. M., Chow, W. S., and Park, Y.II (1995). The grand design of photosynthesis: acclimation of the photosynthetic apparatus to environmental cues. Photosynth. Res. 46, 129–139. doi: 10.1007/BF00020423

Atkin, O. K., Evans, R. J., and Siebke, K. (1998). Relationship between the inhibition of leaf respiration by light and enhancement of leaf dark respiration following light treatment. Funct. Plant Biol. 25, 437–443. doi: 10.1071/PP970159

Ballaré, C. L., Sánchez, R. A., Scopek, A. L., Casal, J. J., and Ghera, C. M. (1987). Early detection of neighbour plants by phytochrome perception of spectral changes in reflected sunlight. Plant Cell Environ. 10, 551–557. doi: 10.1111/1365-3040.ep1160091

Beadle, C. L., and Long, S. P. (1985). Photosynthesis – is it limiting to biomass production? Biomass 8, 119–168. doi: 10.1016/0144-4565(85)90022-8

Becker, C., Učević, B., Jukić Spika, M., Kläring, H. P., Krumbein, A., Baldermann, S., et al. (2015). Nitrogen limited red and green leaf lettuce accumulate flavonoid glycosides, caffeic acid derivatives, and sucrose while losing chlorophylls, β-carotene and xanthophylls. PLoS One 10, e0142867. doi: 10.1371/journal.pone.0142867

Bugbee, B., and Monie, O. (1992). The limits of crop productivity. BioScience 42, 494–502. doi: 10.2307/1311879

Bugbee, B. (2004). Nutrient Management in Recirculating Hydroponic Culture. In: Proceedings of the South Pacific Soilless Culture Conference. M. Nichols (ed.). Acta Hortic. 648, 99–112. doi: 10.17660/ActaHortic.2004.648.12

Cannell, M. G. R., and Thornley, J. H. M. (2000). Modelling the components of mineral nutrition of plants: principles and perspectives. 2nd edition (Sunderland, Massachusetts: Sinauer Assoc, Inc.).

Federer, C. A., and Tanner, C. B. (1966). Spectral distribution of light in the forest. Ecology 47, 555–560. doi: 10.2307/1939392

Evans, J. R. (1987). The dependence of quantum yield on wavelength and environmental cues. Func. Plant Biol. 13, 514. doi: 10.1071/PP9880011

Franklin, K. A. (2008). Shade avoidance. New Phytol. 179, 930–944. doi: 10.1111/j.1469-8137.2008.02507.x

Goins, G. D., Ruffe, L. M., Cranston, N. A., Yorio, N. C., Wheeler, R. M., and Sager, J. C. (2001-01-24). Salad crop production under different light wavelengths of red light-emitting diodes (LEDs) (SAE Technical Paper No. 2001-01-2422). doi: 10.4271/2001-01-2422

Gomers, C. M., Visser, E. J., St Onge, K. R., Voesenek, L. A., and Pierik, R. (2013). Shade tolerance: when growing tall is not an option. Trends Plant Sci. 18, 65–71. doi: 10.1016/j.tplants.2012.09.008

Heraut-Bron, V., Robin, C., Varlet-Grancher, C., Afif, D., and Guckert, A. (1999). Light quality (red: far-red ratio): does it affect photosynthetic activity, net CO2 assimilation, and morphology of young white clover leaves? Can. J. Bot. 77, 1425–1431. doi: 10.1139/b99-099

Huang, Y., Luan, X., and Wang, Z. (2019). The role of photosystem II in photosynthesis. Frontiers in Plant Science | www.frontiersin.org September 2020 | Volume 11 | Article 581156
Morgan, D. C., and Smith, H. (1989). A systematic relationship between phytochrome-controlled development and species habitat, for plants grown in simulated natural radiation. *Planta* 145, 253–258. doi: 10.1007/BF00445449

Morgan, P. W., Finlayson, S. A., Childs, K. L., Mullet, J. E., and Rooney, W. L. (2002). Opportunities to improve adaptability and yield in grasses. *Crop Sci.* 42, 1791–1799. doi: 10.2135/cropsci2002.1791

Murakami, K., Matsuda, R., and Fujiwara, K. (2018). A mathematical model of photosynthetic electron transport in response to the light spectrum based on excitation energy distributed to photosystems. *Plant Cell Physiol.* 59, 1643–1651. doi: 10.1093/pcp/pcy085

Nelson, J. A., and Bugbee, B. (2015). Analysis of environmental effects on leaf temperature under sunlight, high pressure sodium and light emitting diodes. *PLoS One* 10, e0138930. doi: 10.1371/journal.pone.0138930

Park, Y., and Runkle, E. S. (2017). Far-red radiation promotes growth of seedlings by increasing leaf expansion and whole-plant net assimilation. *Environ. Exp. Bot.* 136, 41–49. doi: 10.1016/j.envexpbot.2016.12.013

Penning de Vries, F., Brunsting, A. H. M., and van Laar, H. H. (1974). Products, requirements and efficiency of biosynthesis a quantitative approach. *J. Theor. Biol.* 45, 339–377. doi: 10.1016/0022-5193(74)90119-2

Rajapakse, N. C., and Kelly, J. W. (1994). Problems of reporting spectral quality and interpreting phytochrome-mediated responses. *Hortic. Sci.* 29, 1404–1407. doi: 10.21273/HORTSCI.29.12.1404

Sager, J. C., Smith, W. O., Edwards, J. L., and Cryt, K. L. (1988). Photosynthetic efficiency and phytochrome photoequilibria determination using spectral data. *Trans. ASAIE* 31, 1882–1889. doi: 10.13031/2013.39052

Schmitt, J., McCormac, A. C., and Smith, H. (1995). A test of the adaptive plasticity hypothesis using transgenic and mutant plants disabled in phytochrome-mediated elongation responses to neighbors. *Am. Nat.* 146, 937–953. doi: 10.1086/285832

Sharp, R. E., Matthews, M. A., and Boyer, J. S. (1984). Kok effect and the quantum yield of photosynthetic light partially inhibits dark respiration. *Plant Physiol.* 75, 95–101. doi: 10.1104/pp.75.1.95

Siegelman, H. W., and Butler, W. L. (1965). Properties of phytochrome. *Annu. Rev. Plant Physiol.* 16, 383–392. doi: 10.1146/annurev.pp.16.060165.002123

Smith, H., and Whitelam, G. C. (1997). The shade avoidance syndrome: multiple responses mediated by multiple phytochromes. *Plant Cell Environ.* 20, 840–844. doi: 10.1046/j.1365-3040.1997.d01-104.x

Smith, H. (1982). Light quality, photoperception, and plant strategy. *Annu. Plant Physiol.* 33, 481–518. doi: 10.1146/annurev.pp.33.060182.002405

Stutte, G. W., Edney, S., and Serrrit, T. (2009). Photoregulation of bioprotectant content of red leaf lettuce with light-emitting diodes. *Crop Sci.* 49, 78–82. doi: 10.21273/HORTSCI.49.1.78

USU Crop Physiology Laboratory (2018). "Utah Monocot/Dicot Solution". *Nutrients, Paper 2.* Available at: https://digitalcommons.usu.edu/cgi/viewcontent.cgi?article=1001&context=cpl_nutrients

Valladares, F., and Niinemets, Ü. (2008). Shade tolerance, a key plant feature of complex nature and consequences. *Annu. Rev. Ecol. Evol. S.* 39, 237–257. doi: 10.1146/annurev.ecolsys.39.110707.173506

van Iersel, M. W., and Bugbee, B. (2000). A multiple chamber, semicontinuous, crop carbon dioxide exchange system: design, calibration, and data interpretation. *J. Am. Soc. Hortic. Sci.* 125, 86–92. doi: 10.21273/JASHS.125.1.86

van Iserl, M. W., and Seymour, L. (2000). Growth respiration, maintenance respiration, and carbon fixation of vinca: a time series analysis. *J. Am. Soc. Hortic. Sci.* 125, 702–706. doi: 10.21273/JASHS.125.6.702

van Iserl, M. W. (2003). Carbon use efficiency depends on growth respiration, maintenance respiration, and relative growth rate. A case study with lettuce. *Plant Cell Environ.* 26, 1441–1449. doi: 10.1046/j.1365-3040.2003.01067.x

Vertregt, N. F. W. T., and Penning de Vries, F. (1987). A rapid method for determining the efficiency of biosynthesis of plant biomass. *J. Theor. Biol.* 128, 109–119. doi: 10.1016/S0022-5193(87)80034-6

Vogelmann, T. C. (1993). Plant tissue optics. *Annu. Rev. Plant Biol.* 44, 231–251. doi: 10.1146/annurev.pl.44.090193.001311

Waring, R. H., Landsberg, J. I., and Williams, M. (1998). Net primary production of forests: a constant fraction of gross primary production? *Tree Physiol.* 18, 129–134. doi: 10.1093/treephys/18.2.129
Woolley, J. T. (1971). Reflectance and transmittance of light by leaves. *Plant Physiol.* 47, 656–662. doi: 10.1104/pp.47.5.656
Zhen, S., and Bugbee, B. (2020). Far-red photons have equivalent efficiency to traditional photosynthetic photons: Implications for redefining photosynthetically active radiation. *Plant Cell Environ.* 43, 1259–1272. doi: 10.1111/pce.13730
Zhen, S., and van Iersel, M. W. (2017). Far-red light is needed for efficient photochemistry and photosynthesis. *J. Plant Physiol.* 209, 115–122. doi: 10.1016/j.jplph.2016.12.004
Zhen, S., Haidekker, M., and van Iersel, M. W. (2018). Far-red light enhances photochemical efficiency in a wavelength-dependent manner. *Physiol. Plant.* 167, 21–33. doi: 10.1111/ppl.12834
Zhu, X. G., Long, S. P., and Ort, D. R. (2008). What is the maximum efficiency with which photosynthesis can convert solar energy into biomass? *Curr. Opin. Biotech.* 19, 153–159. doi: 10.1016/j.copbio.2008.02.004
Zhu, X. G., Long, S. P., and Ort, D. R. (2010). Improving photosynthetic efficiency for greater yield. *Annu. Rev. Plant Biol.* 61, 235–261. doi: 10.1146/annurev-arplant-042809-112206

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2020 Zhen and Bugbee. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.