Bronchoscopic Cryobiopsy for the Diagnosis of Diffuse Parenchymal Lung Disease

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Abstract

Background: Although in some cases clinical and radiographic features may be sufficient to establish a diagnosis of diffuse parenchymal lung disease (DPLD), surgical lung biopsy is frequently required. Recently a new technique for bronchoscopic lung biopsy has been developed using flexible cryo-probes. In this study we describe our clinical experience using bronchoscopic cryobiopsy for diagnosis of diffuse lung disease.

Methods: A retrospective study of subjects who had undergone bronchoscopic cryobiopsy for evaluation of DPLD at an academic tertiary care center from January 1, 2012 through January 15, 2013 was performed. The procedure was performed using a flexible bronchoscope to acquire biopsies of lung parenchyma. H&E stained biopsies were reviewed by an expert lung pathologist.

Results: Twenty-five eligible subjects were identified. With a mean area of 64.2 mm², cryobiopsies were larger than that typically encountered with traditional transbronchial forceps biopsy. In 19 of the 25 subjects, a specific diagnosis was obtained. In one additional subject, biopsies demonstrating normal parenchyma were felt sufficient to exclude diffuse lung disease as a cause of dyspnea. The overall diagnostic yield of bronchoscopic cryobiopsy was 80% (20/25). The most frequent diagnosis was usual interstitial pneumonia (UIP) (n = 7). Three of the 25 subjects ultimately required surgical lung biopsy. There were no significant complications.

Conclusion: In patients with suspected diffuse parenchymal lung disease, bronchoscopic cryobiopsy is a promising and minimally invasive approach to obtain lung tissue with high diagnostic yield.

Introduction

There are over 150 recognized causes of diffuse parenchymal lung disease (DPLD, also known as “interstitial lung disease”, ILD). While in certain circumstances, classic clinical features and radiography are sufficient to establish a diagnosis [1], frequently additional testing including bronchoscopy and/or surgical lung biopsy are required. The ATS/ERS guidelines for diagnosis and treatment of idiopathic pulmonary fibrosis (IPF), the most common and most severe form of idiopathic ILD, recommend surgical lung biopsy if classic radiographic criteria are not found [1]. In two recent clinical trials, 22–54% of IPF patients had undergone surgical lung biopsy to establish the diagnosis [2,3], suggesting that in spite of advances in imaging of DPLDs, a substantial number of patients require biopsy.

Surgical lung biopsy requires endotracheal intubation, general anesthesia, chest tube placement and typically hospitalization for several days. There is also risk of prolonged air leak and/or bronchopleural fistula requiring prolonged hospitalization and/or reoperation, and low but significant risk of mortality [4,5]. Given these potential risks and complications, there is need for a less invasive technique for lung biopsy.

Bronchoscopy is commonly employed in the diagnostic evaluation of DPLD, however for most DPLD’s, transbronchial biopsy (TBBx) specimens have low diagnostic yield. In a large series reviewing 801 DPLD patients who underwent transbronchial biopsies, less than 1/3 established a specific diagnosis, and of those cases nearly all were malignant or infectious in origin [6]. Specifically among patients with idiopathic ILD, the yield appears...
to be low. In one series of patients ultimately found to have IPF, only 3/32 specimens met usual interstitial pneumonia (UIP) criteria [7]. In several other series, 30–34% of transbronchial biopsies had at least 1 feature consistent with UIP/IPF but were overall inconclusive [8,9].

Flexible cryo-probes have been used for bronchoscopic procedures including endobronchial biopsy and tumor ablation with success. Recently, these probes have been employed for peripheral lung biopsy in several small series and have been shown to be safe [10,11,12]. In this study, we sought to examine the diagnostic yield of bronchoscopic cryobiopsy among patients with DPLDs and characterize cryobiopsy specimens in these patients. Some of these results have been presented previously in abstract form at the 2013 American Thoracic Society International Conference, May 19, 2013 [13].

Figure 1. Bronchoscopic cryoprobe and low power images of biopsies demonstrate cryobiopsies are larger than typical forceps biopsies. (A) Bronchoscopic cryoprobe. (B) Gross pathology of a representative cryobiopsy specimen measuring 19 mm in length. (C) Typical transbronchial forceps biopsy. (D) Normal bronchoscopic cryobiopsy sample. All images were taken at low power (40×).

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Materials and Methods

Subjects

This study was approved by the Vanderbilt University Institutional Review Board (IRB # 121556) with a waiver of informed consent as for this study existing data were analyzed in a deidentified manner. Subjects were identified by a retrospective chart review of all patients who had undergone bronchoscopic cryobiopsy at Vanderbilt University Medical Center for the diagnosis of DPLD from January 1, 2012 through October 29, 2012. All subjects had suspected interstitial lung disease based on clinical information, serology testing, and high-resolution computed tomography (HRCT) scans with atypical features requiring lung biopsy. The decision to perform bronchoscopic cryobiopsy was made by the patient’s attending physician as a component of the clinical evaluation of the patient.

Safety outcomes are also reported for a cohort of research subjects that were identified from an ongoing study of asymptomatic family members at-risk for familial interstitial pneumonia (FIP) [14] (Vanderbilt University IRB #87780). Written informed
consent was obtained from subjects prior to enrollment. Eligible subjects were >40 years of age and had a first degree relative with FIP. After enrollment in the study, eligible subjects were administered a modified American Thoracic Society Diffuse Lung Disease 78 (ATS-DLD78) questionnaire. Subjects with a Dyspnea Score <3 were considered asymptomatic as this had previously been shown to be predictive of pulmonary function test abnormalities [15]. All subjects then underwent HRCT with prone positioning as per the study protocol.

Bronchoscopy and Biopsy

Bronchoscopies were performed by an attending interventional pulmonologist with standard monitoring and sedation procedures in the bronchoscopy suite. Seven days prior to the procedure, patients were required to stop all anti-platelet agents: aspirin, clopidogrel and herbal supplements. Patients were administered intravenous conscious sedation by certified registered nurse anesthetist under the direction of an anesthesiologist. A Portex cuffed 8.5 wire-spiral endotracheal tube was positioned in the airway with the bronchoscope (Olympus XT180, Olympus America Inc) with the cuff deflated to permit spontaneous breathing. This allowed rapid re-entry into the airway after the biopsy was done without having to re-navigate through the nose or mouth. Bronchoscopic cryobiopsy was targeted to the areas of abnormality seen on HRCT. Among research subjects with no HRCT abnormalities, biopsies were performed in the posterior and lateral segments of the right lower lobe. The Erbe 1.9 mm cryoprobe (Erbe Elektromedizin GmBH) (Figure 1A) was advanced through the working channel and into the distal parenchyma under fluoroscopic guidance. The probe was withdrawn ~2 cm from the point resistance was met. Then on exhalation a 4-second freeze time was applied and then the probe and bronchoscope were removed en-bloc from the airway, the probe with the frozen biopsy specimen was not pulled into the bronchoscope. The extended probe and frozen specimen was submerged into room temperature saline to speed thawing and removal of specimen. The probe was then reintroduced into the airway to clear bleeding and then wedged into the biopsied subsegment for three minutes. Two biopsies total, each from a different segment were taken for each procedure. The specimens were removed immediately from the saline and placed in formalin.

Clinical Parameters

Clinical data were extracted by retrospective chart review independently by two of the authors; any disagreement was resolved by consensus discussion.
Biopsy Specimens

Biopsy specimens were fixed in 10% formalin and embedded in paraffin. Hematoxylin and eosin (H&E) stained slides were reviewed by the standard protocol of the Vanderbilt clinical pathology laboratory; research samples were processed in an identical manner. Clinical specimens were reviewed under standard clinical care and research specimens were reviewed by an expert lung pathologist who was blinded to subject information.

Diagnostic Yield

Clinical parameters, radiography and biopsy results were reviewed independently by two of the authors who separately recorded a diagnosis for each clinical subject. Biopsies were considered “diagnostic” if after reviewing the clinical information, radiology and pathology both reviewers and the treating clinician reached a uniform diagnosis and no further evaluation (i.e. surgical lung biopsy) was pursued. Disagreement was resolved by consensus discussion between the reviewers and an expert lung pathologist; there was disagreement regarding 2/25 clinical biopsies, both of which were ultimately deemed “non-diagnostic.”

Results

During the study period, 25 subjects underwent cryobiopsy for evaluation of DPLD. An additional 12 asymptomatic subjects at-risk for FIP underwent cryobiopsy as per the study protocol. Subject characteristics are summarized in Table 1. In total, 73 cryobiopsies were performed during the study period. Biopsy characteristics of clinical cryobiopsies are summarized in Table 2. In 7 patients, both upper and lower lobes were sampled; in the remaining 18 subjects, biopsies were taken from multiple segments of the lower lobes. Alveolated lung was present in 24/25 clinical biopsy samples. Typical examples of a forceps biopsy and cryobiopsy are shown in Figure 1. Mean cryobiopsy length was 8.7 mm (range 6–31 mm), and mean biopsy area was 64.2 mm² (range 1.5–136.7 mm²), which is substantially larger than 5–15 mm² reported in several recent studies of transbronchial forceps biopsy [10,12].

Among clinical biopsies a specific clinical-pathologic diagnosis was achieved in 19/25 cases. In one case, normal lung tissue obtained by cryobiopsy was felt sufficient to exclude DPLD as a cause of dyspnea. Three subjects with nondiagnostic cryobiopsies were subsequently referred for surgical lung biopsy, two of which were diagnostic, one demonstrating UIP and another cryptogenic organizing pneumonia (COP). Among the 25 clinical subjects, the most common diagnosis was UIP/IPF (7/25). Other diagnoses are displayed in Figure 2. Representative examples of cryobiopsies from several diagnoses are shown in Figure 3. Among subjects found to have UIP, hallmark features including temporal heterogeneity, microscopic honeycomb cysts and fibroblastic foci from cryobiopsy specimens are demonstrated in Figure 4.

Safety outcomes are summarized in Table 3. During the study period, there were no pneumothoraces or bleeding requiring additional intervention. One day after discharge, one outpatient was admitted to the hospital for overnight observation for hemoptysis that spontaneously resolved. One inpatient was observed one night in the intensive care unit for post-procedural hypoxemia that returned to baseline oxygen requirement within 12 hours. No patients required intubation or use of noninvasive ventilation after the procedure.

Discussion

In this series of twenty-five patients with DPLD who required lung biopsy, in 20 cases (80%) lung tissue obtained from a bronchoscopic cryobiopsy, when combined with clinical and radiographic information, was sufficient to establish a diagnosis. This represents a striking improvement in diagnostic yield.
compared to historical studies evaluating the use of traditional forceps transbronchial biopsies which report diagnostic yield in approximately 30% of cases [7,8,9].

When compared to surgical lung biopsy, performed either via mini-thoracotomy or using video-assisted thoracic surgery (VATS), bronchoscopic cryobiopsy offers several advantages. As we report here, this procedure can be safely performed in the outpatient setting with low risk of need for hospitalization. The risk of general anesthesia, placement of a double-lumen endotracheal tube and single lung ventilation are avoided with bronchoscopic cryobiopsy. Patient discomfort from incisional and/or chest tube site pain is eliminated. We did not observe any pneumothoraces following bronchoscopic cryobiopsy so although we cannot comment on the risk of prolonged air leak or bronchopleural fistula, this suggests it is likely reduced compared to surgical lung biopsy. A previous series reported pneumothorax rates similar to that of transbronchial forceps biopsy [10].

There are several reasons why the diagnostic yield of cryobiopsies may exceed that of forceps biopsies. First, cryobiopsy samples appear to be substantially larger in size than forceps biopsies; in this series the mean longest dimension of cryobiopsy samples was 8.7 mm, 3–5 times larger than those typically obtained using forceps [10,12]. This is accompanied by a substantial increase in biopsy area; the mean biopsy area we report is similar to that observed in a study of lung transplant patients [12].

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Figure 4. Cryobiopsy demonstrates typical features of UIP/IPF. Representative cryobiopsy samples from subjects with UIP demonstrating (A) dense scar with microscopic honeycomb cysts (HC), (B) Temporal heterogeneity, and (C–D) Fibroblastic foci (FF). A and B original magnification 40 ×, C–D 400 ×. doi:10.1371/journal.pone.0078674.g004
patients who underwent bronchoscopic cryobiopsy [12]. This increase in area offers more contiguous alveolated lung for diagnosis of UIP/IF. Similarly, samples more likely contain multiple small airways for evaluation of suspected bronchiolitis. As has previously been reported [10,12], crush artifact caused by the biopsies procedure is minimized as the freezing process does not appear to alter lung architecture.

The primary safety concern of bronchoscopic cryobiopsy is bleeding risk, as the bronchoscope must be withdrawn from the airway to remove the biopsy specimen from the cryoprobe, thus an immediate bronchoscope wedge to tamponade bleeding cannot be achieved. To minimize this risk, all antiplatelet and anticoagulant medications were discontinued at least 7 days before the cryobiopsy procedure. Whenever possible, biopsies were taken from dependent areas of the lung. Once the biopsy was removed from the probe, the bronchoscope was quickly returned to the airway and wedged in the biopsied segment for three minutes to tamponade bleeding and then was slowly withdrawn. We did not observe any bleeding requiring additional intervention or blood transfusion; typical blood loss was 15 milliliters or less.

This study, by nature of its retrospective design, has several limitations. The sample size we report is small and from a single-center where all procedures were performed by an experienced interventional pulmonologist. Therefore, the diagnostic yield and safety outcomes might differ among operators with variable experience. It is possible that referral bias may have enriched the study population for patients/conditions more likely to have “diagnostic” cryobiopsies. Most patients who underwent bronchoscopic cryobiopsy had abnormal pulmonary function tests demonstrating mild-to-moderate restriction with a reduced diffusing capacity for carbon monoxide (DLCO) and “atypical” imaging that did not meet ATS/ERS criteria for definite or probable UIP. While this possibility of a referral bias cannot be completely excluded, all subjects met criteria for referral for surgical lung biopsy at our institution. Two of 25 subjects were found to have malignancy, and 0/25 had sarcoidosis, the two conditions for which traditional transbronchial forceps biopsies is of highest yield, thus we believe the subjects are a representative sample of patients with DPLD referred to a specialty interstitial lung disease clinic at a tertiary care center. In all patients, biopsies were taken from areas of the lung that appeared radiographically abnormal, similar to the approach employed for transbronchial forceps biopsies at this institution wherein two separate affected areas are customarily sampled. The majority of biopsy specimens were taken from lower lobes, and it is possible that this strategy may be subject to sampling error as upper lobes may have been biopsied less frequently than would have been the case with surgical lung biopsy. Blinding of the clinical biopsy samples for pathological review was not possible given the retrospective nature of the study. Finally, no cryobiopsies were performed on subjects with substantial baseline oxygen requirements (>4 Lpm) or patients with respiratory failure requiring mechanical ventilation, thus the safety and yield of bronchoscopic cryobiopsies in this setting is not known.

We believe that this technique has the potential to dramatically change practice in the evaluation of patients with DPLD. For many patients with DPLD and “atypical” imaging findings, bronchoscopic cryobiopsy offers a reasonable likelihood of establishing a diagnosis while avoiding the morbidity of surgical lung biopsy. Further study will be necessary to determine the comparative cost-effectiveness of bronchoscopic cryobiopsy compared to surgical lung biopsy, however we anticipate cryobiopsy may offer substantial cost savings.

**Conclusions**

The evaluation of diffuse parenchymal lung disease is complex, and in many patients, lung biopsy is necessary to establish a diagnosis. Bronchoscopic cryobiopsy can be performed safely (in experienced hands) in patients with DPLD and offers promise of achieving adequate tissue for diagnosis in many patients without the risk and morbidity of surgical lung biopsy. As experience with this technique increases, bronchoscopic cryobiopsy holds promise to become the preferred, first-line approach to lung biopsy in the diagnostic approach to the patient with DPLD.

**Author Contributions**

Conceived and designed the experiments: JAK JMP LHL WEL TSB MPS JEL OBR. Performed the experiments: JAK JMP LHL WEL TSB MPS JEL OBR. Analyzed the data: JAK JMP WRM LS LAG JEJ WEL TSB OBR. Wrote the paper: JAK JMP WRM LHL WEL TSB MPS JEL OBR.

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