Research Article

Downregulation of miR-3934 in Peripheral Blood Mononuclear Cells of Asthmatic Patients and Its Potential Diagnostic Value

Wenyu Wang, Jing Wang, Hong Chen, Xiaofei Zhang, and Kaiyu Han

1Department of Respiratory and Critical Medicine, The Second Affiliated Hospital of Harbin Medical University, China
2Department of Respiratory Medicine, Harbin Chest Hospital, China

Correspondence should be addressed to Kaiyu Han; hankaiyu2002@163.com

Received 22 September 2020; Revised 7 December 2020; Accepted 30 December 2020; Published 9 January 2021

Academic Editor: Shtaywy S. Abdalla

Copyright © 2021 Wenyu Wang et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. The present study focused on the potential clinical significance of miR-3934 in the occurrence and development of asthma. Methods. 80 asthma and 80 healthy controls were recruited in this study. The peripheral blood mononuclear cells (PBMCs) and serum samples of the asthma patients as well as the healthy controls were isolated, and the expression levels of miR-3934 in PBMCs were examined by RT-qPCR methods. Furthermore, the relationship between the level of miR-3934 in PBMCs and the disease severity has been analyzed, and the potential diagnostic value of miR-3934 was evaluated by the receiver operating characteristics (ROC) curve. Finally, the expression level of IL-6, IL-8, and IL-33 have been detected using the ELISA kits, and Pearson’s correlation analysis was performed to investigate the relationship between the level of miR-3934 in PBMCs and the serum expression of those inflammatory cytokines in asthma patients. Results. miR-3934 was dramatically decreased in PBMCs of the asthma patients, and miR-3934 was markedly reduced in PBMCs of patients with severe asthma vs. mild asthma. Furthermore, ROC analysis showed that levels of miR-3934 in PBMCs can distinguish asthma patient, especially the severe asthma patients from the controls. Finally, the levels of miR-3934 in PBMCs were negatively correlated with the serum levels of IL-6, IL-8, and IL-33 in asthma patients, respectively. Conclusions. miR-3934 was downregulated in PBMCs of asthmatic patients and may function as a potential diagnosis biomarker.

1. Introduction

Asthma is a common chronic airway inflammatory disease. In recent years, its incidence has been on the rise all over the world [1–3]. Severe asthma has seriously affected human health and increased socioeconomic burden [4, 5]. The receptor for advanced glycation end products (RAGE) is a multiligand receptor that belongs to the immunoglobulin superfamily. Accumulating evidence has indicated that RAGE is considered a key mediator of asthma pathogenesis [6]. RAGE is highly expressed in the lung tissues of patients with various respiratory diseases including asthma [7–9]. It is speculated that inhibiting RAGE expression may play a role in reducing airway inflammation in asthma.

MicroRNA (miRNA) is a group of small noncoding RNAs, 18-25 nucleotides in length. They act as gene inhibitors by negatively regulating the expression of their target genes. Previous studies have shown that miRNA may play an important role in the pathogenesis of different diseases, such as cancer, cardiovascular diseases, and autoimmune diseases [10–13]. It has also been found that some miRNAs are abnormally expressed in asthma [14–18]; however, few studies have focused on the expression of miRNAs in PBMC of asthma patients and their possible underlying mechanisms.

First, with bioinformatics analysis (TargetScan), we noticed that miR-3934 is complementary to the 3’ untranslated region (UTR) in RAGE. The miR-3934 was verified by luciferase assay to bind with the sequence “ACACCUGA” in RAGE. Moreover, it was observed that the overexpression of miR-3934 can suppress the expression level of RAGE. Therefore, we predicted that miR-3934 may be a potential negative regulatory sequence for RAGE. Considering that the RAGE expression was significantly high in PBMCs from asthma patients, this study is aimed at investigating the potential clinical value of detecting the expression level of miR-3934 in the PBMC of patients with asthma.
2. Material and Methods

2.1. Patients. The study recruited 80 patients with asthma who were diagnosed in the Second Affiliated Hospital of Harbin Medical University between January 2018 and September 2019. At the same time, 80 healthy volunteers without a history of lung disease, allergies, or respiratory infections were used as a control group. This study does not include candidates with autoimmune diseases, heart disease, AIDS, and other serious or systemic diseases. The severity of asthma is defined according to the Global Asthma Management and Prevention Strategy (updated in 2018) as follows: (1) mild: asthma that is well controlled with as-needed reliever medication alone or with low-intensity controller treatment such as low dose ICS; (2) moderate: asthma that is well controlled with low dose ICS/LABA or with high/moderate dose ICS; (3) severe: asthma that requires high-dose ICS/LABA to prevent it from becoming “uncontrolled” or asthma that remains “uncontrolled” despite this treatment [19]. The ethics documents were obtained from the Ethics Committee of the Second Affiliated Hospital of Harbin Medical University. The study obtained the informed consent of all candidates. In addition, the clinical data was collected for analysis, including demographic information and peripheral blood cell. The blood samples were collected while diagnosing asthma. After centrifugation, the serum samples were stored in a refrigerator at -80°C for further analysis.

2.2. Quantitative Real-Time RT-PCR. Because asthma is an immune disease, our group focused on the roles of immune cells in asthma. Therefore, we focused on the expression of miRNA in PBMCs. The level of miR-3934 in PBMCs was detected by qRT-PCR. Briefly, PBMCs were separated from blood samples as previously described [20]. The total RNAs were extracted from cells using TRIzol reagent (Invitrogen) and then reverse-transcribed into cDNAs using PrimeScript RT Master Mix (Takara, Dalian, China). Furthermore, the Hairpin-it™ miRNAs qPCR Quantitation Kit (Takara, Dalian, China) was used for RT-PCR reactions. ABI 7900 Biosystems (Applied Biosystems Life Technologies, Foster City, USA) was used for amplification by the following program: 95°C for 30 secs, 40 cycles of 95°C for 5 secs, and 60°C for 30 secs. U6 was employed for normalization, and primers were designed by Genescript (Nanjing, China). The level of miR-3934 in each sample was quantified by the 2-ΔΔCt method.

2.3. Enzyme-Linked Immunosorbent Assay. The expression level of IL-6, IL-8, and IL-33 in serum samples of asthmatic patients, as well as healthy subjects, were detected by the
enzyme-linked immunosorbent assay (ELISA) using commercially available kits (Nanjing Jiancheng Bio-Engineering Institute Co., Ltd., Nanjing, China). Procedures strictly followed the manufacturer’s instructions.

2.4. Statistical Analysis. Continuous variables were presented as the median (interquartile range), and categorical variables were described as number and percentage (%). Differences between two normal distribution groups were compared by Student’s t-test using the GraphPad Prism v7.0 software (GraphPad Software, CA, USA). Receiver operating characteristics (ROC) curve analysis was applied to predict the potential diagnostic value of miR-3934 for asthma. Pearson’s correlation analysis was used for correlation studies. Significance was considered when the p value was less than 0.05.

3. Results

3.1. Downregulation of miR-3934 in PBMCs of Asthmatic Patients Compared with Healthy Controls. The expression levels of miR-3934 in PBMCs of asthmatic patients and that of healthy volunteers were compared by RT-PCR analysis. There were no significant differences between the asthma group and the control group in terms of age, gender, smoke history, allergy history, and white blood cell count (Table 1, p > 0.05). It has been observed that miR-3934 expression was dramatically decreased in PBMCs of asthmatic patients in comparison with that of healthy controls (Figure 1, p < 0.001). Furthermore, patients were divided into a mild (n = 52) and severe group (n = 28) based on the severity of the disease as previously described [19]. The association between miR-3934 levels in PBMCs of patients and severity

![Figure 2: Potential diagnostic value of miR-3934 in peripheral blood mononuclear cells (PBMCs) for asthma. (a) Receiver operating characteristic (ROC) analysis for miR-3934 to distinguish between asthmatic patients and healthy controls. (b) ROC analysis for miR-3934 to distinguish between mild asthmatic patients and healthy controls. (c) ROC analysis for miR-3934 to distinguish between severe asthmatic patients and healthy controls.](image-url)
of the disease has also been analyzed. We found that compared with patients with mild asthma, miR-3934 levels markedly decreased in PBMCs of patients with severe asthma (Figure 1(b), \( p < 0.01 \)).

3.2. Potential Diagnostic Value for Detecting miR-3934 Levels in PBMCs of Asthmatic Patients. The potential diagnostic values of miR-3934 for asthma were evaluated by receiver-operating characteristic curves (ROC). As Figure 2(a) shows, the area under curve (AUC) value for miR-3934 levels in PBMCs to distinguish asthmatic patients from healthy subjects was 0.8041 (Figure 2(a), 95% confidence interval (CI), 0.7366–0.8715). Moreover, when asthmatic patients were divided into mild and severe groups, the AUC value of miR-3934 was 0.7666, to distinguish between mild asthmatic patients from healthy subjects (Figure 2(b), 95% CI, 0.6880–0.8452), and 0.8737 to distinguish between severe asthmatic patients and healthy subjects (Figure 2(c), 95% CI, 0.8074–0.9399). The above results indicated that miR-3934 may serve as a sensitive biomarker for the early diagnosis of asthma, particularly for those with severe asthma.

3.3. Proinflammatory Cytokines Were Increased in Serum of Asthmatic Patients. Proinflammatory factors in all subjects, including IL-6, IL-8, and IL-33, were evaluated by ELISA. The results showed that the levels of IL-6, IL-8, and IL-33 were markedly increased in serum of patients with asthma in comparison with that of healthy subjects (Figure 3, \( p < 0.01 \)).

Figure 3: Interleukin-6 (IL-6), IL-8, and IL-33 were overexpressed in the serum of asthmatic patients. **\( p < 0.01 \).

Figure 4: Correlation between miR-3934 expression and levels of interleukin-6 (IL-6), IL-8, and IL-33 in asthmatic patients.
3.4. Correlation between miR-3934 Expression in PBMCs and Serum Levels of IL-6, IL-8, and IL-33 in Asthmatic Patients. We performed Pearson’s correlation analysis to analyze the relationship between the miR-3934 expression in PBMCs and serum levels of some proinflammatory cytokines in asthmatic patients (Figure 4). We found that the miR-3934 expression levels in PBMCs negatively correlated with serum expression of IL-6 ($r = -0.3726$, $p = 0.0007$), IL-8 ($r = -0.3656$, $p = 0.0009$), and IL-33 ($r = -0.2323$, $p = 0.0381$) in asthmatic patients, respectively.

4. Discussion

In the current work, we investigated the potential clinical significance of miR-3934 in asthma. The main finding is that miR-3934 is significantly reduced in PBMC of asthmatic patients, and the level of miR-3934 in PBMC can distinguish asthmatic patients from healthy controls. And the level of miR-3934 is negatively correlated with the severity of the disease. Finally, the level of miR-3934 in PBMCs is negatively correlated with the expression of certain inflammatory cytokines in the serum of asthmatic patients.

The roles of miRNAs in asthma have been discussed in previous work, e.g., miR-221 and miR-485-3p were upregulated in asthma and may serve as potential biomarkers [21]; miR-192-5p has been reported to attenuate remodeling of the airway as well as autophagy in asthma [14]. miR-3934 is a recently identified miRNA, and its role in asthma has not been discussed. The results of the bioinformative analysis showed that there are many miRNAs that target 3′UTR of RAGE. Based on the unpublished results of our group, miR-3934 is the most significantly differently expressed miRNA between patients and healthy controls; more importantly, the direct targeting relationship between miR-3934 and RAGE was confirmed by due luciferase report assay. Therefore, miR-3934 was selected as a target in the present study.

Although it is more convenient to examine the expression of miRNAs in the serum or plasma samples, there are many studies focused on miRNA expression as biomarkers in PBMCs in different diseases, especially for immune diseases. Since asthma is an immune disease, in the current study, miRNA levels in PBMCs of 80 patients and healthy controls were compared. The results showed that miR-3934 was downregulated in asthma. This suggested that miR-3934 might be involved in asthma pathogenesis. Moreover, ROC analysis confirmed that miR-3934 levels in PBMCs can distinguish between disease intensity of asthmatic patients, especially between severe asthmatic patients and healthy controls. Overall, our study revealed that miR-3934 levels in PBMCs may function as potential diagnostic markers for asthma. Screening the expression of miR-3934 in PBMC may help identify asthma patients who are prone to severe diseases. Preventive targeted therapy for these patients is expected to reduce the frequency of acute attacks and reduce the economic burden.

It has been widely accepted that asthma is an inflammatory disease, and inflammation has been considered an important event during the occurrence and progress of asthma [22–25]. Several cytokines were found to be abnormally expressed in asthma serum and are associated with disease development [22, 25]. IL-6, IL-8, and IL-33 are known as proinflammatory cytokines. In accordance with previous reports, we found that they markedly increased in asthmatic patients in comparison with healthy controls [19, 26–28]. Moreover, a negative correlation was observed between miR-3934 expressions in PBMCs and inflammatory cytokine levels. Interestingly, none of these proinflammatory cytokines has been predicted as the target gene of miR-3934. Therefore, we speculate that some important inflammatory regulators may be direct targets of miR-3934, and downregulation of miR-3934 in PBMC may be related to the expression of these molecules and consequently activate inflammatory signaling and lead to increased expression of proinflammatory cytokines. In other words, the upregulation of miR-3934 in PBMC may play a protective role in the occurrence of asthma.

In this research, we focused on the clinical implication of miR-3934 in asthma, and its hypothesized mechanism, mainly through serological analyses. Furthermore, the targets of miR-3934 in asthma should be identified by performing cell and animal studies.

In conclusion, miR-3934 was downregulated in PBMCs of asthmatic patients and may function as a potential diagnostic marker. Although the current work needs to be further confirmed by larger sample size, the results of this study have proposed potential clinical implications of miR-3934 in early diagnosis and management of asthma.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors report no conflict of interest.

Authors’ Contributions

Han KY was responsible for the study conception and design and critically reviewed the manuscript. Chen H was responsible for the data collection. Zhang XF was responsible for the data analysis and interpretation. Han KY, Wang Jing, and Wang WY were responsible for the manuscript drafting. Han KY was responsible for the statistics. All of the authors have read and approved the final version of the manuscript.

Acknowledgments

This study was supported by the funds from the National Natural Science Foundation of China (NO. 81700021) and was supported by funds from the Wu Jieping Medical Foundation (NO.320.6750.18493).
References

[1] M. G. Matera, P. Rogliani, L. Calzetta, and M. Cazzola, “TSLP Inhibitors for asthma: current status and future prospects,” Drugs, vol. 80, pp. 449–458, 2020.

[2] T. Irewall, L. Soderstrom, A. Lindberg, and N. Stenfors, “High incidence rate of asthma among elite endurance athletes: a prospective 4-year survey,” Journal of Asthma, pp. 1–7, 2020.

[3] W. Barcik, B. RCT, M. Sokolowska, and B. B. Finlay, “Inhibitors for asthma: current status and future prospects,” Drugs, vol. 80, no. 7, pp. 2243–2253, 2020.

[4] T. Irewall, L. Soderstrom, A. Lindberg, and N. Stenfors, “The role of lung and gut microbiota in the pathology of asthma,” Immunity, vol. 52, no. 2, pp. 241–255, 2020.

[5] K. Han, M. Suzuki, M. Yamaguchi et al., “Parental quality of life and self-efficacy in pediatric asthma,” Journal of Asthma, pp. 1–8, 2020.

[6] T. Haselkorn, S. J. Szefler, B. E. Chipp et al., “Disease burden and long-term risk of persistent very poorly controlled asthma: TENOR II,” The Journal of Allergy and Clinical Immunology: In Practice, vol. 8, no. 7, pp. 2243–2253, 2020.

[7] S. J. M. Hoornhorst, A. T. Lo Tam Loi, S. D. Pouwels et al., “Advanced glycation endproducts and their receptor in different body compartments in COPD,” Journal of Allergy and Clinical Immunology, vol. 144, no. 3, pp. 796–808, 2019.

[8] M. Choi, H. Jeong, S. Kim, M. Kim, M. Lee, and R. TJN, “Targeted delivery of Chlβ4Chlβ4 siRNA to alveolar macrophages using ternary complexes composed of HMG and oligoarginine micelles,” Nanoscale, vol. 12, no. 2, pp. 933–943, 2020.

[9] S. Zhang, Y. Shi, Y. Zhang et al., “miRNA-5119 regulates immune checkpoints in dendritic cells to enhance breast cancer immunotherapy,” Cancer Immunology, Immunotherapy, vol. 69, pp. 951–967, 2020.

[10] W. Peng, S. Yu, A. M. Handler et al., “miRNA-1-3p is an early embryonic male sex-determining factor in the Oriental fruit fly Bactrocera dorsalis,” Nature Communications, vol. 11, no. 1, pp. 932, 2020.

[11] L. Lou, M. Tian, J. Chang, F. Li, and G. Zhang, “miRNA-192-5p attenuates airway remodeling and autophagy in asthma by targeting MMP-16 and ATG7,” Biomedicine & Pharmacotherapy, vol. 122, article 109692, 2020.

[12] S. Bartel, S. La Grutta, G. Cilluffo et al., “Human airway epithelial extracellular vesicle miRNA signature is altered upon asthma development,” Allergy, vol. 75, no. 2, pp. 346–356, 2020.

[13] R. Niu, X. Xiao, B. Liu, Y. Li, Y. Zhong, and L. Ma, “Retraction Note: Inhibition of airway inflammation in a cockroach allergen model of asthma by agonists of miRNA-33b,” Scientific Reports, vol. 9, no. 1, article 16790, 2019.

[14] Y. Chen, Z.-D. Mao, Y.-J. Shi et al., “Comprehensive analysis of miRNA–mRNA–lncRNA networks in severe asthma,” Epigenomics, vol. 11, no. 2, pp. 115–131, 2019.

[15] D. Lacedonia, G. P. Palladino, M. P. Foschino-Barbaro, G. Scioscia, and G. E. Carpanagno, “Expression profiling of miRNA-145 and miRNA-338 in serum and sputum of patients with COPD, asthma, and asthma–COPD overlap syndrome phenotype,” International Journal of Chronic Obstructive Pulmonary Disease, vol. 12, pp. 1811–1817, 2017.

[16] X. G. Jiang, X. D. Yang, Z. Lv, and P. H. Zhuang, “Elevated serum levels of TNF-α, IL-8, and ECP can be involved in the development and progression of bronchial asthma,” Journal of Asthma, vol. 55, no. 2, pp. 111–118, 2018.

[17] K. Han, M. Suzuki, M. Yamaguchi et al., “The in vitro effects of advanced glycation end products on basophil functions,” International Archives of Allergy and Immunology, vol. 155, no. s1, pp. 64–70, 2011.

[18] F. Liu, H. B. Qin, B. Xu, H. Zhou, and D. Y. Zhao, “Profiling of miRNAs in pediatric asthma: upregulation of miRNA-221 and miRNA-485-3p,” Molecular Reports, vol. 6, no. 5, pp. 1178–1182, 2012.

[19] Y. Zhang, C. Zhao, X. Zhao et al., “Interleukin-25 in sputum as a marker of airway inflammation in children asthma,” Science China Life Sciences, vol. 62, no. 12, pp. 1605–1608, 2019.

[20] T. Hassegawa, T. Okazawa, H. Uga, H. Kurata, and A. Mori, “Serum CXCL9 as a potential marker of Type 1 inflammation in the context of eosinophilic asthma,” Allergy, vol. 74, no. 12, pp. 2515–2518, 2019.

[21] G. S. Whitehead, S. Hussain, R. Fannin et al., “TLRS activation exacerbates airway inflammation in asthma,” Lung, vol. 198, no. 2, pp. 289–298, 2020.

[22] X. G. Liu, J. Li, L. J. Zheng, B. Han, and F. Huang, “Interleukin-36 receptor antagonist alleviates airway inflammation in asthma via inhibiting the activation of interleukin-36 pathway,” International Immunopharmacology, vol. 81, article 106200, 2020.

[23] D. Dimitrova, V. Youroukova, E. Ivanova-Todorova, K. Tumangelova-Yuzeir, and T. Velikova, “Serum levels of IL-5, IL-6, IL-8, IL-13 and IL-17A in pre-defined groups of adult patients with moderate and severe bronchial asthma,” Respiratory Medicine, vol. 154, pp. 144–154, 2019.

[24] K. Gorska, P. Nejman-Gryz, M. Paplinska-Goryca, P. Korczynski, M. Prochorec-Sobieszek, and R. Krenke, “Comparative study of IL-33 and IL-6 levels in different respiratory samples in mild-to-moderate asthma and COPD,” COPD: Journal of Chronic Obstructive Pulmonary Disease, vol. 15, no. 1, pp. 36–45, 2018.

[25] N. Turan, M. J. Edwards, S. Bates et al., “IL-6 pathway upregulation in subgroup of severe asthma is associated with neutrophilia and poor lung function,” Clinical & Experimental Allergy, vol. 48, no. 4, pp. 475–478, 2018.