Background

Atopic Dermatitis (AD) is a chronic relapsing inflammatory disease characterized by inflammatory cell infiltration in skin. It is widely admitted that AD is a systemic disease with both atopic and nonatopic co-morbidities [1, 2]. However, AD may occur at any age, AD starts during the first two years of life [3]. AD is characterized by erythema, edema, lichenification, excoriations, oozing and crusting. AD mainly occurs on ankles, wrists, face, hands and feet, neck and upper chest [4]. Pruritus is a pivotal feature of AD and bring out comorbidities such as psychological distress, sleep loss and increased healthcare utilization. It may be accompanied with symptoms of bronchial asthma, nasal mucosa inflammation and conjunctivitis [5]. Severe cases of AD is correlated with poorer overall health as it causes many disorders such as impaired sleep and increased healthcare utilization [6].

Patients with AD are characterized by skin barrier dysfunction leading to penetration of allergens into the skin, associated with an increased cutaneous inflammation [7]. Many genetics factors might spoil diverse aspects of barrier function. In addition, environmental exposures may affect the barrier function such as soaps, detergents, exogenous proteases and repetitive scratching [8].

Crocin is the major component of Gardenia jasminoides Family Rubiaceae. Crocin was proved to treat AD via inhibition of histamine [9] and blocking of ERK-MAPK/NFκB/STAT1 [10]. Therefore, the goal of this study was to determine the effect of crocin on E-cadherin/AKT signaling pathways in atopic dermatitis.

Materials and Methods

Animals

The local ethical committee approved the animal protocol. Four-week-old BALB/c mice were housed under pathogen-free con-
ditions with normal 12-h light-dark cycle. Mice were distributed into four groups with ten mice each:

**Control group.** Hair of the back of mice was shaved and rubbed with mixture of acetone: olive oil (3:1) was as well as the back of both ears of mice.

**Crocin treated control group.** Mice were given 20 mg/kg crocin (Sigma Aldrich Chemicals Co., St Louise, MO, USA) subcutaneously three times per week for three weeks.

**DNCB-induced AD group.** After 24 hours of removal of the dorsal hair, 100 µl of 1% 2,4-dinitrochlorobenzene (DNCB) solution in acetone: olive oil (3:1) was utilized on back skin and back of both ears for sensitization. After five days, 150 µl of 0.2% DNCB was applied on the skin of the back and behind both ears three times a week for three weeks [11, 12].

**Crocin treated group.** After induction of AD, mice were treated with 20 mg/kg of crocin subcutaneously three times per week for another three weeks.

**Animal sacrifice and collection of samples**

Animals were sacrificed by decapitation. Skin samples were removed and stored at -80°C.

**Morphologic analysis of skin tissue**

Five-micrometer thickness sections were cut from paraffin blocks, stained with toluidine blue and inspected in a blind manner using Nikon Digital Camera (Japan).

**ELISA determination**

Commercially available ELISA kits was used for determination of skin levels of E-cadherin, AKT, syndecan-1 (Thermo Fisher Scientific Inc., Waltham, MA, USA) and MMP9 (R&D Systems, Inc. Minneapolis, MN, USA) following the manufacture protocol.

**Quantitative real-time polymerase chain reaction (RT-PCR)**

RNeasy Mini kit (Qiagen, USA) was used for separation of total RNA. Next, Maxima® SYBR Green/Fluorescein qPCR Master Mix (Fermentas, USA) was used for estimation of total amount of RNA followed by reverse-transcription of 1µ RNA into cDNA by QuantiTect® Reverse Transcription Kit (Qiagen, USA). AKT, MMP9 and syndecan-1 mRNA levels were determined by Maxima® SYBR Green/Fluorescein qPCR Master Mix by Rotor-Gene Q (Qiagen, USA). Meanwhile, GAPDH was implemented as an internal reference. The specific PCR primers (Table 1) were prepared by the help of Primer Express 3.0 (Applied Biosystems, USA).

**Statistical analysis**

The mean ± standard error was used. Sample normality were checked by Kolmogorov-Smirnov test was used. Comparison between groups was evaluated by ANOVA followed by post hoc Bonferroni correction tests. SPSS version 20 was used for statistical computations. Statistical significance was predefined as P ≤ 0.05.

**Results and Discussion**

Atopic dermatitis resulted in 61% reduction in the skin level of E-cadherin when compared with the control group. Treatment of AD mice with crocin resulted in about 2-fold elevation in E-cadherin as compared with AD mice but still significantly less than the control group. Crocin did not affect the E-cadherin of control group (Figure 1). E-cadherin is a transmembrane protein and one of the tight junction [13]. The lesion of AD in humans is associated with down-regulation of E-cadherin [14]. In the inflamed skin lesion of AD patients, off note, E-cadherin expression enhances the induction of T regulatory cells in mice [15]. Crocin was reported to reduce the ability of cells invasion, migration and adhesion by increasing the expression E-cadherin in melanoma metastatic model in mice [16] and prostate cancer cell lines [17]. Up to our knowledge, this is the first time to examine the effect of crocin on E-cadherin in atopic dermatitis model in mice.

Histological observations of skin section of AD mice stained with toluidine blue declared the presence of acanthosis of epidermis (broad arrows) and infiltration with mast cells (thin arrows). All these effects were attenuated by crocin (Figure 2).

Skin of AD mice showed increased gene expression of the cell migration marker, AKT, MMP9 and syndecan-1 by 2.97-, 2.39- and 3.07-fold increase respectively in comparison with control mice (Figure 3 and 4). Moreover, AD resulted in elevated skin protein levels of AKT, MMP9 and syndecan-1 by 3.34-, 2.18- and 3.40-fold increase in comparison with control mice. Treatment of mice with Crocin decreased both gene and protein expression AKT, MMP9 and syndecan-1 in AD mice without affecting the

### Table 1. The primer sets used:

| Primer  | Accession number | Sequence (sense, antisense) |
|---------|------------------|-----------------------------|
| AKT     | NM_009652.3      | 5’-ACAGCATGTTGGCAGAAGAA-3’, 5’-GTGCTGGAGGACAACGACT-3’ |
| MMP9    | NM_013588        | 5’-AACATCTGGCCATCCACACC-3’, 5’-GCAGAAGTTCTTTGGCCTGC-3’ |
| Syndecan-1 | Z22532    | 5’-CAACCTGGCCTCCATGAGAC-3’, 5’-CAACCTGGCCAACAGAC-3’ |
| GAPDH   | M32599          | 5’-ACACAGTCCATGCCATC -3’, 5’-CACCCCTGTTGCGTAGGC-3’ |

[Source: Alyoussef A. Crocin Ameliorated Atopic Dermatitis Induced in Mice by Inhibiting E-Cadherin/AKT Pathway. Int J Clin Dermatol Res. 2020;8(1):248-252.]
Figure 1. Effect of atopic dermatitis (AD) alone and in combination with 20 mg/kg crocin on skin levels of E-cadherin. *: significant difference compared with the control groups at p<0.05. #: significant difference compared with atopic dermatitis group at p<0.05.

Figure 2. Effect of atopic dermatitis (AD) alone and in combination with 20 mg/kg crocin on mice skin sections from control (a), treated control (b), atopic dermatitis (c) and treated atopic dermatitis (d) mice, stained with toluidine blue. Sections showing acanthosis of the epidermis of the skin (broad arrows) and infiltration of mast cells (thin arrows).

Figure 3. Effect of atopic dermatitis (AD) alone and in combination with 20 mg/kg crocin on gene expression (a) and protein level of AKT (b) in skin of mice. *: significant difference compared with the control groups at p<0.05. #: significant difference compared with atopic dermatitis group at p<0.05.
Figure 4. Effect of atopic dermatitis (AD) alone and in combination with 20 mg/kg crocin on gene expression of matrix metalloproteinase 9 (MMP9, a) and syndecan-1 (c) as well as the protein level of MMP9 (b) and syndecan-1 (d) in skin of mice. *: significant difference compared with the control groups at p<0.05. #: significant difference compared with atopic dermatitis group at p<0.05.

Figure 5. Mechanism of action of crocin in atopic dermatitis.
control mice, however, their levels still significantly higher than control mice. The serine/threonine kinase AKT is a downstream effector that is activated by several stimuli, such as growth factor stimulation. AKT is a central component of the activated signaling pathway present in the endothelial cells that helps in epidermal terminal differentiation. Activation of AKT1 results in hyperkeratosis [18]. The phosphorylation of AKT induces the transcription of many pro-inflammatory cytokines such as TNF-α and interleukins [20]. In addition, activation of AKT up-regulates MMP9 enhancing cell invasion and migration [21]. In turn, MMP9 helps the proteolytic activation of syndecan-1 and releasing it in the surrounding media helping cell migration [22, 23]. AKT was previously reported to be involved in crocin activity on endothelium [24], against α-galactose aging model in the hippocampus of Wistar rats [25], angiogenesis in male rats [26] and on diabetic retinopathy in microglial Cells [27]. However, no previous study illustrated the effect of AKT/MMP9/syndecan-1 on atopic dermatitis.

Conclusion

We illustrated that crocin could ameliorate DNCB-induced AD in mice. Crocin protection proceed via activation of E-cadherin leading to the inhibition of cell migration pathway (Figure 5).

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