Denosumab improves clinical manifestations of hypophosphatemic osteomalacia by adefovir-induced Fanconi syndrome: a case report

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Abstract

Background: Adefovir dipivoxil is a nucleotide analogue that is approved for treatment of chronic hepatitis B. Adefovir dipivoxil is associated with proximal tubular dysfunction, resulting in Fanconi syndrome, which can cause secondary hypophosphatemic osteomalacia. We describe a case of a patient with hypophosphatemic osteomalacia secondary to Fanconi syndrome induced by adefovir dipivoxil concomitantly with osteoporosis in whom clinical symptoms were improved by adding denosumab (a human monoclonal antibody targeting the receptor activator of nuclear factor-κB ligand) to preceding administration of vitamin D3.

Case presentation: A 60-year-old Japanese man had been receiving low-dose adefovir dipivoxil (10 mg/day) to treat chronic hepatitis B for approximately 5 years. He presented to an orthopedic surgeon with severe pain of the right hip and no trauma history, and fracture of the neck of the right femur was identified. In addition, 99mTc-hydroxymethylene diphosphate scintigraphy revealed significantly abnormal uptake in the bilateral ribs, hips, and knees, and he was therefore referred to our university hospital for evaluation of multiple pathological fractures. We diagnosed hypophosphatemic osteomalacia due to Fanconi syndrome induced by adefovir dipivoxil therapy. Although we reduced the patient's adefovir dipivoxil dose and added calcitriol (active vitamin D3), he did not respond and continued to complain of bone pain. Several bone resorption markers and bone-specific alkaline phosphatase were also persistently elevated. Therefore, we added denosumab to vitamin D3 supplementation for treatment of excessive bone resorption. Two months after initiation of denosumab, his hip and knee pain was relieved, along with a decrease in serum alkaline phosphatase and some bone resorption markers.

Conclusions: Although denosumab is not generally an appropriate treatment for acquired Fanconi syndrome, it may be useful for patients who have hypophosphatemic osteomalacia due to adefovir dipivoxil-induced Fanconi syndrome associated with excessive bone resorption. However, clinicians should keep in mind that if denosumab is administered to patients with hypophosphatemic osteomalacia accompanied by excessive bone resorption, adequate vitamin D and/or phosphate supplementation should be done before administration of denosumab.

Keywords: Adefovir dipivoxil, Fanconi syndrome, Osteomalacia, Osteoporosis, Chronic hepatitis B, Denosumab

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Background
Adefovir dipivoxil (ADV) is an acyclic nucleotide analogue of adenosine monophosphate that is widely used to treat chronic hepatitis B. There is considerable evidence that long-term administration of ADV, even at a low dose, causes renal tubular dysfunction due to nephrotoxicity [1–3]. Fanconi syndrome can be induced by generalized proximal tubular dysfunction due to ADV therapy, resulting in hypophosphatemic osteomalacia with pathological fractures [1–3].

Denosumab is a human monoclonal antibody targeting the receptor activator of nuclear factor-κB ligand (RANKL) that is employed for the treatment of osteoporosis [4]. In general, denosumab would not be considered appropriate for patients with hypophosphatemic osteomalacia due to Fanconi syndrome, because denosumab rapidly inhibits osteoclast-dependent bone and calcium resorption and subsequently induces hypocalcemia. However, we encountered a 60-year-old man with hypophosphatemic osteomalacia due to ADV-induced Fanconi syndrome in whom clinical manifestations improved after denosumab was added to vitamin D₃ supplementation, as reported here.

Case presentation
A 60-year-old Japanese man was referred to our hospital for evaluation of severe bone pain and pathological fracture of the neck of the right femur. He had been receiving treatment for chronic hepatitis B with lamivudine (100 mg/day) and ADV (10 mg/day) since December 2006. In June 2013, he noticed low-back pain and then developed severe pain in the right hip. One month later, he also developed pain of the great toe during walking and was referred to an orthopedic surgeon at our hospital. Fracture of the neck of the right femur was found, despite no history of trauma (Fig. 1). In addition, ⁹⁹ᵐTc-hydroxymethylene diphosphate scintigraphy revealed significantly abnormal uptake in the bilateral ribs, hips, and knees (Fig. 2). In August 2013, he was referred to our outpatient clinic for evaluation of multiple pathological fractures.

On examination, his body mass index was 18.0 kg/m², temperature was 36.7 °C, blood pressure was 151/86 mmHg, and pulse rate was 67 beats/min (regular). He had generalized bone pain and gait disturbance. His past medical history was appendicitis in 1967 and stomach polyps in 2011. In his family medical history, there was pancreatic cancer, but there was no liver disease. His regular medications were adefovir and ursodeoxycholic acid. He had smoked three packs of cigarettes per day for 30 years, but he had quit since 51 years old. He drinks 350 ml/day of beer. Laboratory tests showed marked elevation of alkaline phosphatase (ALP) (1223 U/L), as well as hypophosphatemia (1.9 mg/dl) and mild hypocalcemia (8.5 mg/dl). His serum creatinine was slightly elevated, whereas serum 1α,25(OH)₂ vitamin D₃ was relatively low at 26.4 pg/ml (reference range, 20.0–60.0 pg/ml) (Table 1).

Urinalysis showed glycosuria (2+) and proteinuria (1+). Urinary β₂-microglobulin was markedly elevated.
### Table 1 Laboratory data on admission

| Chemistry                  | Reference range | Reference range |
|----------------------------|-----------------|-----------------|
| AST                        | 12 U/L          | 13–30           |
| ALT                        | 7 U/L           | 10–42           |
| T-Bil                      | 0.5 mg/dl       | 0.4–1.5         |
| LDH                        | 141 U/L         | 124–222         |
| ALP                        | 1223 U/L        | 106–322         |
| ALP1                       | 3 %             |                 |
| ALP2 + ALP3                | 91 %            |                 |
| ALP5                       | 6 %             |                 |
| LAP                        | 49 U/L          | 30–70           |
| gGTP                       | 12 U/L          | 13–64           |
| ChE                        | 267 U/L         | 215–464         |
| TP                         | 6.6 g/dl        | 6.6–8.1         |
| Alb                        | 4.1 g/dl        | 4.1–5.1         |
| BUN                        | 14 mg/dl        | 8.0–20.0        |
| Cre                        | 1.44 mg/dl      | 0.65–1.07       |
| eGFR                       | 40.2 ml/min/1.73m² |            |
| UA                         | 1.5 mg/dl       | 3.7–7.0         |
| Na                         | 143 mEq/L       | 138–145         |
| K                          | 4 mEq/L         | 3.6–4.8         |
| Cl                         | 113 mEq/L       | 101–108         |
| Ca                         | 8.5 mg/dl       | 8.8–10.1        |
| IP                         | 1.9 mg/dl       | 2.7–4.6         |
| Glucose                    | 93 mg/dl        | 55–110          |
| HbA1c                      | 4.8 %           | 4.3–5.8         |
| CRP                        | 0.1 mg/dl       | ≤0.14           |
| Serum β₂-microglobulin     | 3.5 mg/dl       |                 |
| Hepatitis marker           |                 |                 |
| Hbs-Ag                     | 250 IU/ml       | <0.05           |
| Hbs-Ag                     | 0.4 S/CO        | <1.0            |
| Hbe-Ab                     | 99.6 %Inh       | <50.0           |
| Hematology                 |                 |                 |
| WBC                        | 5200 /μl        | 3300–8600       |
| RBC                        | 447 10³/μl      | 4.35–5.55       |
| Hb                         | 16.5 g/dl       | 13.7–16.8       |
| Hct                        | 47.6 %          | 40.7–50.1       |
| Plt                        | 16.3 10³/μl     | 15.8–34.8       |
| Bone metabolic parameters  |                 |                 |
| TRACP-5b                   | 781 mU/dl       | 170–590         |
| Bone ALP                   | 112 μg/L        | 3.7–20.9        |
| Deoxypyridinoline          | 6.7 nmol/mMol Cre | 2.1–5.4   |
| U-NTx                      | 216.1 mmol BCE/mMol Cr | 13.0–66.2 |
| calcitonin                 | 16 pg/ml        | <952            |
| intact-PTH                 | 43.6 pg/ml      | 8.7–79.6        |
| PTHrP                      | <1.1 pmol/L     | <1.1            |
| 1,25(OH)₂D₃, Vit.D3        | 264 pg/ml       | 20.0–60.0       |
| FGF23                      | <5 pg/ml        |                 |

### Table 1 Laboratory data on admission (Continued)

| Immunology                  | Reference range | Reference range |
|----------------------------|-----------------|-----------------|
| Antinuclear antibody        | 20 times        | <20             |
| C3                         | 74 mg/dl        | 65–135          |
| C4                         | 13 mg/dl        | 13–53           |
| CH50                       | 41 μl/ml        | 30–50           |
| IgG                        | 1031 mg/dl      | 680–1620        |
| IgA                        | 301 mg/dl       | 84–438          |
| IgM                        | 87 mg/dl        | 57–288          |
| IgE                        | 3 IU/ml         | <295            |
| Serum immunoelectrophoresis| M protein (+)    |                 |
| Urinary immunoelectrophoresis| BJ protein (+)  |                 |
| Blood gas analysis          |                 |                 |
| pH                         | 7.328           | 7.350–7.450     |
| PCO₂                       | 40.3 mmHg       | 35.0–45.0       |
| PO₂                        | 110 mmHg        | 85.0–105.0      |
| HCO₃⁻                      | 20.5 mmol/L     | 23.0–28.0       |
| BE                         | –4.7 mmol/L     | –4.6            |
| AG                         | 4.8 mmol/L      | 8.0–12.0        |
| Urinalysis                 |                 |                 |
| pH                         | 6.5             |                 |
| U-glucose                  | 100 mg/dl       |                 |
| U-blood                    | –               |                 |
| BJ-protein                 | +               |                 |
| Urinary NAG                | 78 IU/L         | 0.3–1.15        |
| Urinary β₂-microglobulin   | 138,885 μg/g Cr |                 |
| Na                         | 73 mEq/L        |                 |
| K                          | 24 mEq/L        |                 |
| Cl                         | 91 mEq/L        |                 |
| Ca                         | 29.4 mg/dl      |                 |
| IP                         | 43 mg/dl        |                 |
| UA                         | 30 mg/dl        |                 |
| Cre                        | 48 mg/dl        |                 |
| %TRP                       | 41.59 %         |                 |
| FEUA                       | 46.3 %          |                 |

**Abbreviations:** AG Anion gap, Alb Albumin, ALP Alkaline phosphatase, ALT alanine aminotransferase, AST aspartate aminotransferase, BCE Bone collagen equivalents, BE Base excess, BJ protein Bence-Jones protein, BUN Blood urea nitrogen, CH50 Total hemolytic complement, CHE Cholinesterase, Cre Creatinine, CRP C-reactive protein, eGFR Estimated glomerular filtration rate, FEUA Fractional excretion of uric acid, FGF23 Fibroblast growth factor 23, gGTP γ-Glutamyl transpeptidase, Hb Hemoglobin, HbA1c Hemoglobin A1c, Hb-Ab Hepatitis B e antigen antibody, Hbs-Ag Hepatitis B surface antigen, HCO₃⁻ Bicarbonate, Hct Hematocrit, Ig Immunoglobulin, IP inorganic phosphorus, LAP Leukocyte alkaline phosphatase, LDH Lactate dehydrogenase, NAG N-acetyl-β-D-glucosaminidase, NTx Cross-linked N-telopeptide of type I collagen, PCO₂ Partial pressure of carbon dioxide, P/O Partial pressure of oxygen, PTH Parathyroid hormone, PTHrP Parathyroid hormone-related protein, RBC Red blood cells, T-Bil Total bilirubin, TP Total protein, TRACP-5b Tartrate-resistant acid phosphatase 5b, %TRP Percentage tubular reabsorption of phosphate, UA Urinalysis, WBC White blood cells
at 138,885 μg/g creatinine (Cr), and tubular reabsorption of phosphate was significantly decreased to 41.59% (reference range for percentage tubular reabsorption of phosphate, 80–94%) (Table 1). On the basis of these results, we diagnosed hypophosphatemic osteomalacia secondary to Fanconi syndrome caused by ADV therapy.

Dual-energy X-ray absorptiometry showed an extremely low bone mineral density with a mean lumbar T-score of −3.6 SD. Several bone resorption markers were highly elevated (urinary cross-linked N-telopeptide of type I collagen, 216.1 nmol bone collagen equivalents/mmol; urinary deoxypyridinoline, 6.7 nmol/mmol Cr; serum tartrate-resistant acid phosphatase 5b, 781 mU/dl) (Table 1). Taken together, these findings suggested that the patient had excessive bone resorption combined with hypophosphatemic osteomalacia.

To treat his condition, we first reduced the dose of ADV from 10 mg daily to 10 mg every other day and administered calcitriol (1.0 μg/day) because he had both hypophosphatemia and mild hypocalcemia. In October 2013, he underwent prosthetic replacement of the head of the right femur. However, his generalized bone pain was not relieved by these measures, and several bone resorption markers remained very high, as did serum ALP despite treatment for osteomalacia. In June 2016, we added denosumab (60 mg subcutaneously), a human monoclonal antibody that inhibits RANKL, to ongoing vitamin D therapy in an attempt to suppress persistently high bone resorption. Two months after initiation of denosumab, his hip and knee pain were relieved, along with a decrease in serum ALP and several bone resorption markers (Figs. 3 and 4a–c). Urinary β2-microglobulin decreased gradually after addition of denosumab to vitamin D₃. After 9 months of denosumab treatment, the patient’s mean lumbar T-score increased from −2.0 SD to −1.4 SD (Fig. 4d). We administered denosumab 60 mg every 6 months, and currently he continues to receive denosumab.

**Discussion and conclusions**

We present a case of a 60-year-old man who had hypophosphatemic osteomalacia secondary to acquired Fanconi syndrome caused by low-dose ADV therapy (10 mg/day). Osteomalacia is a metabolic bone disease characterized by a defective mineralization of the osteoid matrix synthesized by osteoblasts, leading to an accumulation of nonmineralized bone. Osteomalacia is usually associated with vitamin D deficiency or hypophosphatemia. Fanconi syndrome results from generalized dysfunction of the proximal renal tubules, which results in impaired reabsorption of amino acids, glucose, uric acid, bicarbonate, and phosphate, with increased urinary excretion of these solutes [5]. Thus, hypophosphatemic osteomalacia can occur in patients with Fanconi syndrome.

ADV is a nucleotide analogue of adenosine monophosphate that is widely used for the treatment of chronic hepatitis B. The mechanisms by which ADV induces nephrotoxicity remain to be determined, but ADV may impair tubular transport, increase apoptosis, or cause mitochondrial injury in the renal tubular epithelium [6]. It has been reported that ADV-related nephrotoxicity is dose-dependent, with a low dose of ADV (10 mg/day) generally being safe and well tolerated [7]. However, there is evidence that even low-dose ADV can induce nephrotoxicity, including Fanconi syndrome, especially in Asian patients [2, 3, 8, 9]. Recently, there has been an increase of reports regarding Fanconi syndrome associated with long-term ADV therapy, even at low doses, in patients from Japan and other Asian countries [8].
In addition, it has been suggested that the incidence of hypophosphatemic osteomalacia due to Fanconi syndrome associated with low-dose ADV therapy may be higher than previously thought [9].

In patients with ADV-induced Fanconi syndrome and/or hypophosphatemic osteomalacia, withdrawal or dose reduction of ADV should be performed immediately [5], and oral phosphate, calcium, or vitamin D₃ should be added as necessary. In our patient, the dose of ADV was immediately reduced from 10 mg daily to 10 mg every other day. We also initiated treatment with calcitriol (1.0 μg/day) because our patient had hypophosphatemia and slightly low serum calcium and vitamin D₃ levels. Generalized renal tubular injury caused by ADV inhibits 1α-hydroxylase activity with subsequent reduction of the 1,25-dihydroxyvitamin D₃ level, leading to a decrease in intestinal calcium and phosphate absorption that can contribute to development of osteomalacia [3]. However, generalized bone pain was not relieved in our patient, and several bone resorption markers remained very high, despite ADV dose reduction and vitamin D₃ supplementation. The serum level of bone-specific ALP also remained high.

Interestingly, our patient had persistent elevation of bone resorption despite receiving treatment for osteomalacia. Previous studies have shown that bone resorption is occasionally increased in patients with hypophosphatemic osteomalacia because osteoclasts are unable to resorb nonmineralized osteoid [10]. Accordingly, our patient may have a mixed form of osteoporosis and osteomalacia (i.e., osteoporomalacia) [11]. Therefore, we added denosumab (anti-RANKL monoclonal antibody) to vitamin D₃ supplementation in order to suppress bone resorption and treat his generalized bone pain. Two months after starting denosumab therapy, the patient’s hip and knee pain showed improvement, together with a decrease in serum bone-specific ALP and bone resorption markers.

Denosumab is a human immunoglobulin G2 monoclonal antibody that inhibits bone resorption by targeting RANKL, which is involved in osteoclast differentiation [4]. Several studies have demonstrated that denosumab is effective for reducing the risk of fracture in women with postmenopausal osteoporosis [12]. Unlike bisphosphonates (another class of potent antiresorptive agents), denosumab does not cause more adverse events in patients with impaired kidney function, because renal insufficiency does not affect its pharmacokinetics or pharmacodynamics [13]. Thus, we chose denosumab for our patient because he had Fanconi syndrome with generalized proximal tubular dysfunction caused by ADV therapy.

In general, antiresorptive agents such as bisphosphonates and denosumab may not be appropriate for treating hypophosphatemic osteomalacia or other forms of osteomalacia, regardless of the degree of renal insufficiency and vitamin D level. Severe and prolonged hypocalcemia was reported after a single injection of denosumab (60 mg) in a patient with osteomalacia due to Fanconi syndrome [14], because calcium homeostasis is dependent on high bone turnover in osteomalacia. Monitoring of the serum calcium level also is mandatory to prevent severe hypocalcemia when denosumab is initiated in all forms of osteomalacia. Very recently, there was another case report of hypophosphatemia osteomalacia secondary to Fanconi syndrome in which bone pain was worsened by administration of denosumab [15]. Unlike in these reports, we observed that 2
months after initiation of denosumab, hip and knee pain of our patient was relieved along with a decrease in serum ALP and some bone resorption markers. We speculate that denosumab had worked well in our patient because of an adequate administration of vitamin D₃ prior to denosumab. We speculate that denosumab may be an option for patients who have hypophosphatemic osteomalacia due to ADV-induced Fanconi syndrome with concurrent enhancement of bone resorption and/or osteoporosis. However, clinicians should keep in mind that if denosumab is administered to patients with hypophosphatemic osteomalacia accompanied by persistent excessive bone resorption despite treatment for osteomalacia, adequate vitamin D and/or phosphate supplementation should be done before administration of denosumab.

Abbreviations
ADV: Adefovir dipivoxil; AG: Anion gap; ALb: Albumin; ALP: Alkaline phosphatase; ALT: alanine aminotransferase; AST: aspartate aminotransferase; BCE: Bone collagen equivalents; BE: Base excess; BJ protein: Bence-Jones protein; BUN: Blood urea nitrogen; CH50: Total hemolytic complement; Che: Cholinesterase; Crea: Creatinine; CRP: C-reactive protein; eGFR: Estimated glomerular filtration rate; FEUA: Fractional excretion of uric acid; FGF23: Fibroblast growth factor 23; gGT: γ-Glutamyl transpeptidase; Hb: Hemoglobin; Hba1c: Hemoglobin A1c; Hb-Ab: Hepatitis B e antigen antibody; Hbs-Ag: Hepatitis B surface antigen; HCO₃⁻: Bicarbonate; Hct: Hematocrit; Ig: Immunoglobulin; LAP: Leukocyte alkaline phosphatase; LDH: Lactate dehydrogenase; NTx: Cross-linked N-telopeptide of type I collagen; Pco₂: Partial pressure of carbon dioxide; PCT: Platelets; PO₂: Partial pressure of oxygen; PTH: Parathyroid hormone; PTHrP: Parathyroid hormone-related protein; RANKL: Receptor activator of nuclear factor-κB ligand; RBC: Red blood cells; T-Bil: Total bilirubin; TP: Total protein; TRACP-5b: Tartrate-resistant acid phosphatase 5b; %TRP: Percentage tubular reabsorption of phosphate; UA: Urinalysis; WBC: White blood cells

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Availability of data and materials
The datasets used and analyzed during this study are available from the corresponding author on reasonable request.

Authors’ contributions
TK, TI, and YA analyzed data and wrote the manuscript. TJ and MS carried out the clinical treatment and follow-up of the patient. MK and SS collected the data. TK and IU advised on and reviewed this report. All authors read and approved the final manuscript.

Ethics approval and consent to participate
It was not required to submit the case to the institutional ethics committee.

Consent for publication
Written informed consent was obtained from the patient for publication of this case report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

Competing interests
The authors declare that they have no competing interests.

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