CRANIOSYNOSTOSIS

An epistatic explanation

Interactions between two gene variants that rarely cause midline craniosynostosis on their own make the development of the disorder a certainty.

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The skull consists of several bony plates that are joined together at cranial sutures, and progenitor cells that reside in these sutures have a crucial role in supporting the rapid growth of new bone to accommodate the growing infant brain. However, in approximately 1 in 2500 babies the cranial sutures fuse together prematurely (either just before birth or shortly afterwards), resulting in a skull deformity known as craniosynostosis (Wilkie et al., 2010). In this condition, the bones of the skull cannot grow quickly enough to keep up with the growing brain, resulting in increased pressure inside the skull. If craniosynostosis is left untreated, it can result in a loss of vision, hearing problems and chronic headaches.

Although some cases of craniosynostosis have been linked to a number of genetic mutations, 70% of observed cases still have no known genetic cause (Twigg and Wilkie, 2015). A form of craniosynostosis called non-syndromic midline craniosynostosis (metopic and sagittal) craniosynostosis accounts for half of all reported cases (Greenwood et al., 2014; Figure 1), but very few of these cases have been associated with genetic mutations. Now, in eLife, Richard Lifton and colleagues at Yale University School of Medicine and Craniosynostosis and Positional Plagiocephaly Support, Inc. – including Andrew Timberlake as first author – report that two genetic mutations, each of which has little impact on its own, can result in devastating outcomes when they are both present (Timberlake et al., 2016).

Genome-wide association studies have identified several single nucleotide polymorphisms that are associated with non-syndromic midline craniosynostosis (Justice et al., 2012). The most significant of these mutations is located close to the gene for a bone morphogenetic protein (BMP) called BMP2, so this site might act as an enhancer to increase the production of BMP2 (Justice et al., 2012). BMP is known to be important in bone development (Urist, 1965), and previous studies have suggested that altered BMP signaling leads to craniosynostosis (Jabs et al., 1993).

It has been suggested that the sporadic occurrence of non-syndromic craniosynostosis is due to de novo (non-inherited) genetic mutations and/or because these mutations do not consistently produce the same symptoms. To identify genetic mutations associated with midline craniosynostosis, Timberlake et al. sequenced the whole exome (that is, the part of the genome formed by exons) of 191 people who were the first in their family to display the symptoms of non-syndromic midline craniosynostosis; they also sequenced the exomes of both.

Related research article

Timberlake AT, Choi J, Zaidi S, Lu Q, Nelson-Williams C, Brooks ED, Bilguvar K, Tikhonova I, Mane S, Yang JF, Sawh-Martinez R, Persing S, Zellner EG, Loring E, Chuang C, Galm A, Hashim PW, Steinbacher DM, DiLuna ML, Duncan CC, Pelphrey KA, Zhao H, Persing JA, Lifton RP. 2016. Two locus inheritance of non-syndromic midline craniosynostosis via rare SMAD6 and common BMP2 alleles. *eLife* 5: e20125. doi: 10.7554/eLife.20125
parents of 132 of these volunteers. The sequences revealed a number of different de novo mutations in the gene that encodes SMAD6, a molecule that is known to inhibit BMP signaling. Most of the mutations prevent the SMAD6 protein from working.

Timberlake et al. found that SMAD6 mutations produce craniosynostosis in only 9% of cases. However, when an individual has both the SMAD6 mutation and the BMP2 risk allele (which on its own leads to the disorder in just 0.08% of cases), craniosynostosis occurs 100% of the time. Thus, these findings clearly indicate that epistatic interactions – interactions where the effect produced by a given gene depends on the presence of other genes – of rare (SMAD6) and common (BMP2) gene variants are a defining feature of the cause of midline craniosynostosis. As a result, these genetic mutations are likely to account for as many cases of craniosynostosis as the single mutation in a gene called FGFR2 that is currently known as the most frequent cause of craniosynostosis.

Timberlake et al. also show that there is no epistatic interaction between the mutant SMAD6 variant and the other mutation that is known to be associated with midline craniosynostosis (a single nucleotide polymorphism in BBS9; Justice et al., 2012). This finding further supports the hypothesis that an increase in BMP signaling is likely to underlie the symptoms of craniosynostosis.

BMPs can cause bone to form in abnormal places (Urist, 1965), so it is possible that the symptoms of craniosynostosis are caused by increased bone formation “filling in” the cranial suture and depleting the numbers of progenitor cells found there. Bone forms as a result of these osteoprogenitor cells developing into cells called osteoblasts, which secrete the materials that form bone. However, the story may not be so simple, because if increased BMP signaling occurs only in osteoblasts, mice do not develop...
the symptoms of craniosynostosis (Komatsu et al., 2013). Instead, an increased rate of cell death in the osteoprogenitor cell population seems to be the primary cause of craniosynostosis in mutant mice that exhibit excessive BMP signaling. To support this notion, mutant mice treated in the womb with drugs that prevent cell death do not develop skull deformities (Hayano et al., 2015). The obvious next steps are to understand the cellular mechanisms that cause cranial sutures to fuse prematurely, and to confirm whether human patients with mutations in the Smad6 and BMP2 genes experience increased cell death in the sutures.

At present the only option for treating craniosynostosis is repeated reconstructive surgery; however, the work of Timberlake et al. and other groups suggest that an alternative therapeutic approach could be to prevent cell death in sutures. Developing such an approach will require researchers to pool together all the knowledge that has been gained from a wide range of genetics studies in both mice and humans. This is therefore a crucial moment in the search for the best treatment strategy for a large proportion of people with non-syndromic craniosynostosis.

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References
Greenwood J, Flodman P, Osann K, Boyadjiev SA, Kimonis V. 2014. Familial incidence and associated symptoms in a population of individuals with nonsyndromic craniosynostosis. Genetics in Medicine 16:302–310. doi: 10.1038/gim.2013.134
Hayano S, Komatsu Y, Pan H, Mishina Y. 2015. Augmented BMP signaling in the neural crest inhibits nasal cartilage morphogenesis by inducing p53-mediated apoptosis. Development 142:1357–1367. doi: 10.1242/dev.118802
Jabs EW, Müller U, Li X, Ma L, Luo W, Haworth IS, Klisak I, Sparks R, Warman ML, Mulliken JB. 1993. A mutation in the homeodomain of the human MSX2 gene in a family affected with autosomal dominant craniosynostosis. Cell 75:443–450. doi: 10.1016/0092-8674(93)90379-5
Justice CM, Yagnik G, Kim Y, Peter I, Jabs EW, Erazo M, Ye X, Ainehsazan E, Shi L, Cunningham ML, Kimonis V, Roscioli T, Wall SA, Wilkie AO, Stoler J, Ricksmeier JT, Heuzé Y, Sanchez-Lara PA, Buckley MF, Druschel CM, et al. 2012. A genome-wide association study identifies susceptibility loci for nonsyndromic sagittal craniosynostosis near BMP2 and within BBS9. Nature Genetics 44:1360–1364. doi: 10.1038/ng.2463
Komatsu Y, Yu PB, Kamiya N, Pan H, Fukuda T, Scott GJ, Ray MK, Yamamura K, Mishina Y. 2013. Augmentation of Smad-dependent BMP signaling in neural crest cells causes craniosynostosis in mice. Journal of Bone and Mineral Research 28:1422–1433. doi: 10.1002/jbmr.1857
Timberlake AT, Choi J, Zaidi S, Lu Q, Nelson-Williams C, Brooks ED, Bilguvar K, Tikhonova I, Mane S, Yang JF, Sawh-Martinez R, Persing S, Zellner EG, Loring E, Chuang C, Gaim A, Hashim PW, Steinbacher DM, DiLuna ML, Duncan CC, et al. 2016. Two locus inheritance of non-syndromic midline craniosynostosis via rare SMAD6 and common BMP2 alleles. eLife 5:e20125. doi: 10.7554/eLife.20125
Twigg SR, Wilkie AO. 2015. A genetic-pathophysiological framework for craniosynostosis. The American Journal of Human Genetics 97:359–377. doi: 10.1016/j.ajhg.2015.07.006
Urist MR. 1965. Bone: Formtion by autinduction. Science 150:893–899. doi: 10.1126/science.150.3698.893
Wilkie AO, Byren JC, Hurst JA, Jayamohan J, Johnson D, Knight SJ, Lester T, Richards PG, Twigg SR, Wall SA. 2010. Prevalence and complications of single-gene and chromosomal disorders in craniosynostosis. Pediatrics 126:e391–400. doi: 10.1542/peds.2009-3491