Minireview

The Role of Hippo Pathway in Cancer Stem Cell Biology

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The biological significance and deregulation of the Hippo pathway during organ growth and tumorigenesis have received a surge of interest in the past decade. The Hippo pathway core kinases, MST1/2 and LATS1/2, are tumor suppressors that inhibit the oncogenic nuclear function of YAP/TAZ and TEAD. In addition to earlier studies that highlight the role of Hippo pathway in organ size control, cell proliferation, and tumor development, recent evidence demonstrates its critical role in cancer stem cell biology, including EMT, drug resistance, and self-renewal. Here we provide a brief overview of the regulatory mechanisms of the Hippo pathway, its role in cancer stem cell biology, and promising therapeutic interventions.

Keywords: cancer stem cell, Hippo pathway, metastasis, YAP/TAZ

INTRODUCTION

The Hippo pathway was originally identified by genetic screens to identify tumor suppressors involved in Drosophila melanogaster tissue growth control. Deletion of the Hippo pathway components in Drosophila resulted in dramatic tissue overgrowth phenotype that was later recapitulated in transgenic mouse models (Dong et al., 2007; Harvey et al., 2003; Huang et al., 2005; Wu et al., 2003; Xu et al., 1995). Recent advances in identification of the mammalian Hippo pathway components and functional implications highlight the role of Hippo pathway in organ development, tumorigenesis, tissue regeneration and stem cell self-renewal (Park and Guan 2013; Yu et al., 2015; Zanconato et al., 2016).

The core Hippo pathway components include a cytoplasmic kinase module and a nuclear transcriptional module. The kinase module is composed of mammalian STE20-like protein kinase 1 (MST1) and MST2, which phosphorylate and activate downstream kinases, large tumor suppressor 1 (LATS1) and LATS2. Recent studies indicate the MAP4K family as part of the kinase module parallel to MST1/2 by directly phosphorylating LATS (Meng et al., 2015). The major function of the Hippo kinase cascade is to inhibit the oncogenic transcriptional module composed of yes-associated protein (YAP), transcriptional co-activator with PDZ-binding motif (TAZ), and TEA domain family members (TEAD). YAP/TAZ function as transcriptional co-activators, which translocate between the cytoplasm and the nucleus, and induce target gene expression involved in cell proliferation and anti-apoptosis via interaction with the TEAD family of transcription factors. Genetic evidence in mice shows that YAP and TAZ (two homologs of Drosophila Yorkie) are functionally redundant during development and regeneration (Nishioka et al., 2009; Xin et al., 2013). When the cytoplasmic Hippo kinase module is ‘on’, MST1/2 activates LATS1/2, which in turn phosphorylates and inactivates YAP/TAZ. Phosphorylated YAP/TAZ are either retained in the cytoplasm via 14-3-3 interaction or subjected to proteasomal or autophagy-induced degradation, and consequently, TEAD-mediated gene transcription is suppressed. By contrast,
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Fig. 1. Regulation of YAP and TAZ by the Hippo pathway. The core inhibitory kinase module of the Hippo pathway is composed of MST1/2, MAP4K, and LATS1/2. The transcriptional module is composed of YAP/TAZ and TEAD. (A) When the Hippo pathway is 'off', YAP/TAZ are dephosphorylated, accumulated, and they translocate into the nucleus to bind the transcription factors, TEAD1–TEAD4, which enable target gene transcription involved in cell proliferation. (B) When the Hippo pathway is turned 'on', LATS1/2 directly phosphorylate YAP/TAZ, which inhibit nuclear import of YAP/TAZ via 14-3-3-mediated cytoplasmic retention, and ubiquitination-mediated proteasomal and autolysosomal degradation. TEAD transcriptional activity is suppressed by VGLL4. LATS indicates large tumor suppressor; MST, mammalian STE20-like protein kinase; YAP, Yes-associated protein; TAZ, transcriptional co-activator with PDZ-binding motif; TEAD, TEA domain family members; VGLL4, transcription cofactor vestigial-like protein 4.

when the Hippo kinases are inactive, dephosphorylated YAP/TAZ maneuver into the nucleus and induce TEAD target gene expression (Meng et al., 2016). In this view, it is well accepted that the core Hippo kinases are tumor suppressors and members of the transcriptional module are oncogenes (Fig. 1). Moreover, although the function of TEAD is largely regulated by YAP/TAZ (Zhao et al., 2008a), recent studies uncover the Hippo-YAP-independent regulatory mechanisms of TEAD that include post-translational modifications and changes in subcellular localization (Chan et al., 2016; Lin et al., 2017; Noland et al., 2016).

UPSTREAM SIGNALS OF THE HIPPO PATHWAY

Hippo pathway and soluble factors

G protein-coupled receptor (GPCR) Ligands

The G protein-coupled receptors (GPCRs) and its ligands, such as mitogenic hormones and growth factors, were discovered as the first soluble signals that regulate the Hippo pathway. YAP/TAZ are activated through Gα12/13-coupled GPCR ligands, including lysophosphatidic acid (LPA), sphingosine-1-phosphate (S1P), Wnt3a, Wnt5a/b, thrombin, thromboxane A2, as well as Kaposi sarcoma-associated herpesvirus (Feng et al., 2016; Liu et al., 2015; Miller et al., 2012; Mo et al., 2012; Park et al., 2015; Yu et al., 2012). Gα12/13 inhibits LATS1/2 and activates YAP/TAZ via Rho GTPase-dependent actin polymerization. Gα11-coupled GPCR ligands, such as endothelin-1 and estrogen also activate YAP/TAZ (Wang et al., 2017; Zhou et al., 2015). Moreover, constitutively activated YAP/TAZ are oncogenic drivers and therapeutic targets in patients with uveal melanoma harboring hyperactive Gα11 mutations (Feng et al., 2014; Yu et al., 2014). Different isoforms in the protein kinase C (PKC) family, key downstream effectors of Gα11, can both activate or inhibit YAP/TAZ (Gong et al., 2015).

Apart from Gα12/13 and Gα11, the Gαs-coupled GPCR ligands such as glucagon and epinephrine, as well as the downstream effectors, cAMP and protein kinase A (PKA), inhibit YAP/TAZ activity (Kim et al., 2013; Yu et al., 2012; 2013). The omega-3 polyunsaturated fatty acids, docosahexaenoic acid (DHA), and eicosapentaenoic acid (EPA) inhibit YAP/TAZ via GPR40/120-Gαs-PKA (Zhang et al., 2016). On the other hand, Gαs is an oncogene frequently mutated in human cancers (O’Hayre et al., 2013). The status and function of Hippo and YAP/TAZ activity are yet unknown in these hyperactive Gαs-driven cancers.

Wnt signaling

Among various morphogens important in development, the Wnt proteins are particularly noteworthy. Recent studies have revealed a complicated cross talk between the Wnt and Hippo-YAP/TAZ pathways. Both canonical Wnt/β-catenin and β-catenin-independent alternative Wnt signaling activate YAP/TAZ via distinct mechanisms. Upon Wnt3a stimulation, canonical Wnt signaling activates both YAP/TAZ and β-catenin similarly through dissociation from the destruction complex (Azzolin et al., 2012; 2014). In this context,
YAP/TAZ activity is indispensable for Wnt-induced biological responses. However, Wnt can activate YAP/TAZ via β-catenin-independent mechanisms. APC, a critical component of the destruction complex, regulates Hippo-YAP activity independent of β-catenin (Cai et al., 2015). Alternative Wnt signaling triggered by both Wnt3a and Wnt5a/b ligands activates YAP/TAZ via the Fzd/Ror-Grb2/Rho GTPases-LATS1/2 pathway which in turn inhibits β-catenin-induced transcription (Park et al., 2015). Mechanistically, YAP/TAZ have been shown to inhibit Wnt/β-catenin signaling by inhibition of DVL nuclear translocation or induction of secreted Wnt inhibitors such as DKK1 and Wnt5a/b (Barry et al., 2013; Park et al., 2015; Seo et al., 2013; Varelas et al., 2010a). Inhibition of Wnt/β-catenin signaling by YAP/TAZ is important during intestinal stem cell regeneration (Barry et al., 2013; Gregoireff et al., 2015), mesenchymal stem cell differentiation (Park et al., 2015; Seo et al., 2013), and induction of naïve pluripotent stem cells (Qin et al., 2016). Similarly, YAP/TAZ-dependent liver growth and HCC formation were suppressed by Wnt/β-catenin signaling (Kim et al., 2017). The context-dependent regulation between YAP/TAZ and Wnt/β-catenin signaling requires further investigation.

**PHYSICAL CUES**

**Cell density and contact inhibition**

Stringent regulation of cell growth upon cell-cell contact is critical for proper organ growth and function since loss of cell contact inhibition of proliferation (CIP) leads to tissue overgrowth, hyperplasia, and tumorigenesis. The Hippo pathway has been shown to mediate cell-cell contact-induced growth inhibitory signals (Ota and Sasaki 2008; Zhao et al., 2008b). During cell-cell contact, components of adherens junction and tight junctions, such as E-cadherin, α-catenin, and Crumbs, activate the Hippo pathway and inhibit YAP/TAZ-induced cell proliferation (Kim et al., 2011; Schlegelmilch et al., 2011; Varelas et al., 2010). In normal cells, YAP/TAZ activation by loss of cell-cell contact during cell migration and proliferation is important for wound healing. In addition, YAP-TEAD transcriptional regulation by contact inhibition is important for proper embryonic development (Nishiohka et al., 2009). However, hyperactive YAP/TAZ during epithelial-to-mesenchymal transition (EMT) results in tumorigenesis due to loss of contact inhibition (Zhao et al., 2007).

**Mechanotransduction**

Mechanical cues including extracellular matrix (ECM) stiffness, cell attachment or detachment, cell geometry, and cytoskeletal tension are potent regulators of YAP/TAZ (Aragona et al., 2013; Dupont et al., 2011; Zhao et al., 2012). Physical attachment of cells to stiff matrix are widely spread out and cells harbor active nuclear YAP/TAZ, whereas, YAP/TAZ are sequestered in the cytoplasm when cells are grown on soft ECM and poorly spread. Similarly, cell attachment to ECM activates YAP/TAZ through Rho GTPases or FAK-Src-P38 pathway, whereas cell detachment inhibits YAP/TAZ in a LATS-dependent manner, leading to anoikis (Kim and Gumbiner 2015; Zhao et al., 2012). Integrins are critical mediators that sense ECM stiffness and relay signals for cell survival and proliferation. Integrins and the focal adhesion complex have been shown to regulate YAP/TAZ (Elbediwy et al., 2016; Serrano et al., 2013; Wang et al., 2016c). Mechanical strain has been shown to induce E-cadherin-dependent, β-catenin-independent YAP activation, which is required for cell cycle entry (Benham-Pyle et al., 2015).

YAP/TAZ in vascular endothelial cells are activated by disturbed blood flow-induced shear stress, whereas laminar flow-induced unidirectional shear stress inhibits YAP/TAZ (Wang et al., 2016b; 2016c). YAP is also required for shear stress-induced migration and invasion of cancer cells (Lee et al., 2017). These implicate YAP/TAZ as mediators of mechanotransduction that are involved in a wide range of pathological conditions.

**STRESS SIGNALS**

**Nutrient stress**

Extracellular nutrients such as glucose and amino acids regulate cell metabolism and proliferation. Cancer cells rely on glycolysis, the Warburg effect, as their main energy source to generate ATP and this regulates YAP/TAZ activity. Glucose starvation-induced energy stress activates LATS1/2 and AMP-activated protein kinase (AMPK), which in turn phosphorylates and inhibits YAP/TAZ activity (Mo et al., 2015; Wang et al., 2015). AMPK can also directly phosphorylate AMOTL1, which regulates LATS and YAP simultaneously (DeRan et al., 2014). Interestingly, recent findings indicate a critical role of YAP in glucose metabolism in cancer cells. YAP promotes glucose transporter 3-induced glucose uptake, which can further activate YAP by promoting YAP-TEAD interaction via direct binding of phosphofructokinase (PFK-1) to TEAD (Enzo et al., 2015; Wang et al., 2015).

Amino acids can regulate YAP/TAZ through the TSC-mTORC1 pathway. Loss-of-function TSC mutations activate YAP by mTORC1-mediated inhibition of autophagy (Liang et al., 2014). In addition, YAP-induced miR-29 expression suppresses PTEN translation, which in turn activates the PI3K-mTOR pathway (Tumaneng et al., 2012). Also, under nutrient-limiting conditions, YAP/TAZ activates mTORC1 via inducing the TEAD target gene, LAT1 amino acid transporter (Hansen et al., 2015). Inhibition of cholesterol synthesis can also suppress YAP/TAZ activity by inducing defect in isoprenylation and membrane localization of Rho GTPases (Sorrentino et al., 2014).

**Cellular and environmental stresses**

Cells and tissues are constantly challenged by various cellular and environmental stresses, and the Hippo pathway has been suggested to play critical roles against stress signals. Cytokinesis failure and extra centrosomes activate the Hippo pathway, which induces YAP/TAZ inhibition and p53 stabilization, thereby preventing tumorigenesis by G1 phase cell cycle arrest (Ganem et al., 2014). In addition, mitotic arrest by anti-tubulin drugs regulates YAP via cyclin-dependent kinase 1 (CDK1)-induced phosphorylation (Yang et al., 2013; Zhao et al., 2014). ER stress dynamically regulates the Hippo pathway in cancer stem cells.
pathway. During the adaptive stage of UPR, PERK-eIF2α-ATF4 axis activates YAP to prevent cell death, however, activation of Hippo signaling by prolonged ER stress inhibits YAP and promotes apoptosis (Wu et al., 2015).

In addition to intracellular stresses, Hippo-YAP/TAZ pathway responds to various external environmental stresses. Oxidative stress activates the Hippo pathway, which in turn, antagonizes YAP-FOXO1 complex that leads to cell death in the mouse heart (Shao et al., 2014a). By contrast, hypoxia inhibits the Hippo pathway by LATS2 degradation through SIAH2-induced ubiquitination, which in turn activates YAP/TAZ (Ma et al., 2015). Hyperosmotic stress elicits TAZ-NFAT5 interaction by ABL kinase-induced tyrosine phosphorylation of TAZ (Jang et al., 2012). Osmotic pressure and gravity also regulate YAP (Hong et al., 2017; Porazinski et al., 2015). In addition to YAP/TAZ, a recent study highlights the effect of cellular and environmental stresses on TEAD localization. High cell density and osmotic stress promote TEAD cytoplasmic translocation via p38 MAPK-independent and -dependent mechanisms, respectively (Lin et al., 2017).

**HIPPO PATHWAY IN TUMORIGENESIS**

**Cancer stem cell**
A specialized subset of tumor cells that had undergone epithelial-to-mesenchymal transition (EMT) has been proposed to harbor unique properties, such as self-renewal and tumor-initiating potential. These cancer stem cells (CSCs) are responsible for drug resistance, metastasis, and recurrence, which are the major causes of cancer mortality (Shibue and Weinberg 2017). Therefore, exploring CSC-specific signaling mechanisms and characteristics is clinically important for better-targeted anticancer treatment.

The Hippo-YAP pathway regulates cell fate and differentiation of progenitor cells during normal organ development and in the context of cancer. YAP can also dedifferentiate and expand undifferentiated stem/progenitor cells such as mature hepatocytes into progenitor cells (Yimlamai et al., 2014). Similarly, YAP/TAZ activation leads to induction of CSC properties in a wide range of human cancers (Fig. 2). In breast cancer cells, YAP has been shown to occupy mammary stem cell signature gene promoters to induce breast CSCs (Kim et al., 2015). In addition, gene expression data from breast cancer tissues with high CSC content overlap with YAP/TAZ-induced gene expression, and TAZ expression is enriched in breast CSCs with CD44high/CD24low phenotype, which is required to sustain their self-renewal and tumor-initiating capacities (Cordenonsi et al., 2011). YAP is also a major inducer of CSC properties by direct upregulation of SOX9 (Song et al., 2014). Similarly, YAP activation by SOX2 is important in maintaining CSCs in osteosarcoma and glioblastoma (Basu-Roy et al., 2015). Glucocorticoid hormone-induced YAP activation expands chemoresistant breast CSCs (Sorrentino et al., 2017). Together, ample amount of evidence indicates that YAP/TAZ play critical roles in CSCs maintenance and cancer progression. However, CSC-specific regulatory mechanisms of YAP/TAZ and Hippo pathway remain unclear.

**EMT**
The epithelial-to-mesenchymal transition (EMT) is a normal developmental program that has been hijacked by cancer cells to trigger abnormal cell migration and invasion. Therefore, EMT has emerged as a critical regulator of CSC phenotype and prerequisite for metastasis (Overholtzer et al., 2006; Shibue and Weinberg 2017). Importantly, induction of EMT in epithelial cells results in CSC characteristics, such as increased mesenchymal traits, increased stem-cell marker expression, enhanced ability to form mammosphere, and more efficient tumor initiation in vivo.

Numerous studies indicate that YAP/TAZ activation drives cell transformation by inducing EMT. YAP/TAZ are well known to promote EMT and metastasis via activation of TEAD transcription factors (Lamar et al., 2012; Lei et al., 2008; Overholtzer et al., 2006; Zhang et al., 2009a; Zhao et al., 2008a). Loss of Hippo kinases or induction of YAP/TAZ activates TEAD-mediated target gene transcription, which results in increased expression of mesenchymal markers, such as vimentin and N-cadherin, and inhibition of epithelial markers, including E-cadherin, simultaneously. Nuclear accumulation of YAP/TAZ is required for epithelial cells to undergo TGFβ-induced EMT, and YAP-TEAD interaction promotes tumor growth and metastasis of breast cancer and melanoma cells (Lamar et al., 2012; Varelas et al., 2010). In addition, YAP interacts with ZEB1, an EMT-inducing transcriptional repressor, and together, ZEB1/YAP target genes promote CSC traits and predict poor survival in breast cancer (Lehmann et al., 2016). Similarly, EMT-inducing transcription factors, Snail/Slug, bind YAP/TAZ-TEAD to control mesenchymal stem cell self-renewal (Tang et al., 2016). Interestingly, YAP expression was critical for the progression of various KRAS-driven cancers, and YAP/KRAS converged on FOS to promote EMT that contributes to KRAS oncogenic addiction (Shao et al., 2014b).
Drug resistance
Chemotherapy and targeted therapy are currently the major treatments for patients with cancer. Unfortunately, these conventional therapies often fail to eradicate carcinoma cells that enter the CSC state, thereby permitting CSC-mediated clinical relapse.

Tumor cells with high YAP/TAZ activity display resistance to chemotherapeutics, which is in part due to the CSC characteristics acquired by YAP/TAZ activation. YAP activation transforms prostate epithelial cells and confers androgen-insensitive state and castration resistance in vivo (Zhang et al., 2015). Recently, it was shown that inhibition of LATS2 by miR-302/367 induces YAP activation and confers CSC state in prostate cancer cells (Guo et al., 2017). YAP/TAZ activation was shown to induce metastatic activity and drug resistance to chemotherapeutic drugs such as paclitaxel and doxorubicin in breast CSCs (Bartucci et al., 2015; Cordenonsi et al., 2011). Numerous studies suggest that Hippo pathway inhibition or YAP/TAZ activation sustains cancer cell survival against DNA-damaging agents such as UV, radiation, cisplatin, taxol, and fluorouracil (5-FU) in a wide number of tumor types (Cheng et al., 2016; Ciamporcero et al., 2016; Fernandez et al., 2012; Mao et al., 2014; Zhao et al., 2014).

YAP/TAZ also promote resistance to targeted therapies. YAP promotes resistance to drugs targeting RAF and MEK in tumor cells harboring BRAF, KRAS, and NRAS mutations, which is a major clinical challenge (Lin et al., 2015). YAP activation by actin remodeling is one of the mechanisms by which YAP induces resistance to BRAF inhibitors in melanoma cells (Kim et al., 2016).

Importantly, well-established YAP/TAZ-TEAD target genes of secreted ligands mediate drug resistance. Connective tissue growth factor (CTGF) confers resistance to chemotherapeutic drugs including doxorubicin and paclitaxel in breast cancer, osteosarcoma, and glioblastoma multiforme (GBM) (Tsai et al., 2014; Wang et al., 2009; Yin et al., 2010). In addition, the prototype alternative Wnt ligand WNT5A, as well as its receptors Ror1/2, has been shown to induce resistance to targeted BRAF inhibitors in melanoma cells (Anastas et al., 2014; O’Connell et al., 2013; Park et al., 2015). Moreover, in PIK3CA mutant breast tumors, WNT5A is one of the most upregulated genes, which correlates with activation of YAP/TAZ-WNT5A axis by PIK3CA H1047R mutation (Cizkova et al., 2010; Park et al., 2015). The ligand for epidermal growth factor receptor (EGFR), amphiregulin (AREG), is also a transcriptional target of YAP, which is known to cause resistance to chemotherapy and receptor tyrosine inhibitors such as gefitinib in non-small-cell lung cancers (Busser et al., 2010; Tung et al., 2017; Zhang et al., 2009b).

These indicate that targeting YAP/TAZ-induced secreted ligands could improve the efficacy in patients with cancer via non-cell-autonomous mechanisms.

PHARMACOLOGICAL INTERVENTIONS
Cancer remains to be one of the leading causes of mortality worldwide. CSCs that have undergone EMT, are capable of metastasizing, and confer drug resistance. As discussed above, the Hippo-YAP/TAZ pathway has emerged as an oncogenic driver that confers CSC traits in cancer cells. An ideal target for small-molecule therapeutics is protein kinases, which function as oncogenes. Unfortunately, since the Hippo core kinases, MST1/2 and LATS1/2, are tumor suppressors, researchers have been searching for therapeutic options beyond the Hippo kinases (Fig. 3). However, recent progress suggests that inhibition of Hippo kinases may also be beneficial in cancer treatment.
Verteporfin
Verteporfin, a FDA-approved photosensitizer to treat macular degeneration, is currently the most widely used YAP inhibitor in research laboratories (Liu-Chittenden et al., 2012). Verteporfin inhibits YAP-TEAD-induced target gene transcription via disrupting the binding between YAP and TEAD, or upregulating 14-3-3 expression (Liu-Chittenden et al., 2012; Wang et al., 2016a). Verteporfin blocks YAP-induced liver tumorigenesis and uveal melanoma in vivo, thereby demonstrating the therapeutic significance of disrupting YAP/TAZ-TEAD interactions in cancer cells (Feng et al., 2014; Liu-Chittenden et al., 2012; Yu et al., 2014). Verteporfin exerts its anti-tumor effect via directly binding to YAP. In addition to TEADs, other transcription factors, such as SMAD, p73, and AP1 serves as transcriptional partners of YAP/TAZ (Strano et al., 2001; Varelas et al., 2008; Zanconato et al., 2015), however, the effect of verteporfin on their transcriptional activity is unknown. Also, YAP-independent cytotoxic effect of verteporfin has been reported, which suggests caution when interpreting the effect of verteporfin on YAP activity (Dasari et al., 2017).

VGLL4-mimicking peptide
VGLL4, which is the only member of VGLL family that carries two TDU domains, was initially reported to bind TEAD1 in cardiac myocytes and suppress TEAD1-dependent \( \alpha_1 \)-adrenergic activation (Chen et al., 2004). Recently, VGLL4 was identified as an endogenous inhibitor of YAP-TEAD interaction by competing with YAP, thereby preventing TEAD-induced cell growth and tumorigenesis in D. melanogaster as well as mouse models (Koontz et al., 2013; Zhang et al., 2014). Unlike other negative regulators of YAP/TAZ, such as Hippo kinases, angiomotin, 14-3-3, and \( \alpha \)-catenin that are located in the cytoplasm, VGLL4 is one of the few endogenous YAP inhibitors that resides in the nucleus. Based on the structure of VGLL4-TEAD4 complex, a peptide that mimics VGLL4 inhibitory function against YAP has recently been developed (Jiao et al., 2014). Treatment with VGLL4-mimicking peptide suppresses human primary gastric cancer cell growth, providing a rationale for peptide-based therapeutic strategy to treat YAP-driven human cancers.

Statins
Statins were identified as potent YAP inhibitors from FDA-approved drug library screening to search for drugs that induce cytoplasmic translocation of YAP/TAZ in breast cancer cells (Sorrentino et al., 2014). Statins are drugs that are used to lower cholesterol levels in patients with hypercholesterolemia by inhibiting 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase. Inhibiting the conversion of HMG-CoA to mevalonate reduces geranylgeranyl pyrophosphate synthesis required for proper Rho GTPase activity. Consequently, inactivation of Rho GTPases by treatment with statins inhibits YAP/TAZ (Sorrentino et al., 2014; Wang et al., 2014). Statin treatment of breast cancer cells decreased their CSC properties, such as self-renewal, and inhibited in vitro and in vivo tumor initiation. In addition to clinical studies showing the anti-inflammatory effects of statins, the inhibitory mechanisms of YAP may provide a rationale to reposi-

LAT5 kinase inhibitors
Hippo core kinases are well established as tumor suppressors since loss of NF2, Sav1, MST1/2, and Mob1 in mouse models as well as in humans leads to cancer development (Yu et al., 2015). Since small-molecule kinase activators are hardly available, Hippo kinases remain to be poorly druggable targets. However, a recent study identified an unexpected role of LAT51/2 kinase in suppressing anti-tumor immunity (Morishii et al., 2016). LAT51/2 suppressed Toll-like receptors-MYD88/TRIF pathway-mediated type I interferon response, thereby enhancing anti-tumor immune response. Therefore, inhibition of upstream Hippo kinases as a monotherapy or in combination with immune checkpoint inhibitors can be an attractive strategy to suppress tumor progression. In addition to anticancer effects, drugs targeting the Hippo pathway may be valuable for wound healing and tissue regeneration.

FUTURE PERSPECTIVE

The Hippo pathway plays an important role in tumorigenesis by promoting CSC characteristics, such as EMT, chemoresistance, metastatic potential, and self-renewal. Despite its relevance in tumorigenesis, oncogenic driver mutations in the core components of the Hippo pathway are relatively rare. Therefore, in addition to targeted therapy, pharmacological modulation of signal transduction pathways that crosstalk with the Hippo pathway or inhibition of YAP/TAZ target genes by combination therapy may be promising approaches to target YAP/TAZ activity in CSCs. More importantly, limitations of current CSC markers hinder precise isolation of CSCs from bulk heterogeneous cancer cell populations. Therefore, identifying the CSC-specific oncogenic signaling network of YAP/TAZ will provide attractive avenues to eradicate CSC and treat cancer progression.

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