Visceral Adiposity Index (VAI) Is Predictive of an Altered Adipokine Profile in Patients with Type 2 Diabetes

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Abstract

Aims: Although there is still no clear definition of “adipose tissue dysfunction” or ATD, the identification of a clinical marker of altered fat distribution and function may provide the needed tools for early identification of a condition of cardiometabolic risk. Our aim was to evaluate the correlations among various anthropometric indices [BMI, Waist Circumference (WC), Hip Circumference (HC), Waist/Hip ratio (WHR), Body Adiposity Index (BAI) and Visceral adiposity Index (VAI)] and several adipokines [Visfatin, Resistin, Leptin, Soluble leptin receptors (sOB-R), Adiponectin, Gherlin, Adipsin, PAI-1, vascular endothelial growth factor (VEGF), Hepatocyte growth factor (HGF) TNF-α, hs-CRP, IL-6, IL-18] in patients with type 2 diabetes (DM2).

Materials and Methods: Ninety-one DM2 patients (age: 65.25 ± 6.38 years; 42 men and 49 women) in stable treatment for the last six months with metformin in monotherapy (1.5–2 g/day) were cross-sectionally studied. Clinical, anthropometric, and metabolic parameters were evaluated. Serum adipokine levels were assayed with Luminex based kits.

Results: At the Pearson’s correlation, among all the indices investigated, VAI showed a significant correlation with almost all adipokines analyzed [Visfatin, Resistin and hsCRP (all p < 0.001); Adiponectin, sOb-R, IL-6, IL-18, HGF (all p < 0.010); Gherlin and VEGF (both p < 0.05)]. Through a two-step cluster analysis, 55 patients were identified with the most altered adipokine profile (patients with ATD). At a ROC analysis, VAI showed the highest C-statistic [0.767 (95% CI 0.66–0.84)] of all the indices.

Conclusions: Our study suggests that the VAI, among the most common indexes of adiposity assessment, shows the best correlation with the best known adipokines and cardiometabolic risk serum markers. Although to date we are still far from clearly identifying an ATD, the VAI would be an easy tool for clearly mirroring a condition of cardiometabolic risk, in the absence of an overt metabolic syndrome.

Introduction

Metabolically complicated obesity and its associated insulin resistance are increasingly prevalent and contribute to increasing the prevalence of type 2 diabetes and cardiovascular disease [1–3], two major causes of morbidity and mortality in our population. Excessive triglyceride storage in adipocytes, due to weight increase, is known to induce changes in adipose tissue (AT), such as adipocyte insulin resistance (resulting in higher lipolytic activity), decreased adiponectin production, increased TNF-α, IL-6 production and increase in several other adipokines. These changes are characteristic of “adipose tissue dysfunction” (ATD) or “adiposopathy” [4] which is often associated with diabetes mellitus, hypertension and diabetic dyslipidemia [low high-density lipoprotein cholesterol (HDL-C) and raised triglycerides]. However, this condition is not necessarily associated with obesity; it can occur with a mild increase in body weight, even within the nonobese range [5,6]; conversely, the threshold may not be reached even in the presence of obesity [7]. Therefore, the existence of simple tools for clinical evaluation of this condition would allow better identification of people at risk for systemic metabolic complications and lay the foundation for developing more effective strategies to prevent such complications as ectopic fat deposition and systemic insulin resistance.

In 2010 we modelled the Visceral adiposity index (VAI), a gender-specific mathematical index based on simple anthropometric [BMI and waist circumference (WC)] and metabolic [Triglycerides (TG) and HDL Cholesterol (HDL)] parameters, as a presumed surrogate marker of adipose tissue function and distribution, independently linked to insulin sensitivity and cardiometabolic risk in the general population [8]. In the last three years, the VAI has been reported on in 30 publications, in which its capacity to express cardiometabolic risk and possible ATD was identified.
The most important results were obtained in populations at metabolic risk not always having an overt metabolic syndrome, such as the general population [9–12], post-menopausal women [13] and women with Polycystic Ovary Syndrome [14]. Recently, in a population of young women with PCOS, the VAI proved to be able to replace visceral Computed Tomography scanning as a marker for visceral adiposity and to predict insulin resistance [15]. These findings have led to the proposal of a precise VAI cut-off to differentiate the “metabolically healthy polycystic ovary syndrome” from the “metabolically unhealthy polycystic ovary syndrome” [16].

Also, in patients with non-alcoholic fatty liver disease, the VAI is linked to significant fibrosis [17,18], while it is not more powerful than WC in discriminating steatosis from steatohepatitis [19]; in patients with HCV, the VAI showed an independent association with both steatosis and necroinflammatory activity and has a

### Table 1. Clinical characteristics and pattern of adipocytokine secretion in the patients with type 2 diabetes.

|                      | Women No 49 | Men No 42 | Total No 91 |
|----------------------|-------------|-----------|-------------|
| **Age (years)**      | 62.37±6.67  | 62.12±6.10| 62.25±6.38  |
| **Duration of disease (years)** | 9.53±6.50  | 9.95±5.40 | 9.73±5.99   |
| **BMI (Kg/m²)**      | 30.35±4.94*| 28.36±4.09| 29.43±4.65  |
| **Waist Circumference (cm)** | 103.02±12.43| 101.93±10.10| 102.52±11.36|
| **Hip Circumference (cm)** | 110.30±11.66*| 103.66±7.40| 107.24±10.42|
| **Waist/Hip ratio**   | 0.93±0.10*  | 0.98±0.04 | 0.95±0.08   |
| **Body Adiposity Index** | 37.09±6.27**| 30.31±4.17| 33.96±6.35  |
| **Visceral Adiposity Index** | 2.64±1.56* | 1.87±0.94 | 2.29±1.36   |
| **HbA1c (%)**        | 6.94±0.68   | 6.88±0.57 | 6.92±0.63   |
| **HbA1c (mmol/mol)** | 52±7.40     | 52±6.20  | 52±6.90     |
| **HOMA-IR**          | 2.98±3.02   | 3.26±3.44 | 3.11±3.21   |
| **Fasting Insulin (pmol/l)** | 61.75±63.33 | 68.94±72.31 | 65.07±67.33 |
| **Fasting C-peptide (nmol/l)** | 0.14±0.16  | 0.16±0.15 | 0.15±0.15   |
| **Urinary Albumin-to-Creatinine Ratio (UACR) (mg/g)** | 16.13±47.53 | 17.05±26.31 | 16.55±38.99 |
| **Fasting lipids**   |             |           |             |
| **Total Cholesterol (mmol/l)** | 4.67±0.78  | 4.53±0.97 | 4.60±0.87   |
| **HDL Cholesterol (mmol/l)** | 1.31±0.31  | 1.20±0.32 | 1.25±0.32   |
| **LDL Cholesterol (mmol/l)** | 2.75±0.82  | 2.79±0.91 | 2.77±0.86   |
| **Triglycerides (mmol/l)** | 1.45±0.61  | 1.36±0.51 | 1.41±0.57   |
| **NEFA (mmol/l)**    | 0.54±0.27*  | 0.39±0.24 | 0.47±0.26   |
| **Fasting adipocytokines** |             |           |             |
| **Leptin (ng/dl)**   | 13.74±12.02**| 3.59±4.79 | 9.06±10.65  |
| **Soluble leptin receptor (sOb-R) (ng/ml)** | 19.89±6.18* | 23.10±6.40 | 21.37±6.46  |
| **Leptin/sOb-R ratio** | 0.89±1.09** | 0.20±0.34 | 0.57±0.90   |
| **Adiponectin (µg/ml)** | 10.95±12.10 | 13.24±14.37 | 12.01±13.17 |
| **Visfatin (ng/ml)** | 0.64±0.56   | 0.61±0.50 | 0.62±0.53   |
| **Resistin (ng/ml)**  | 3.78±4.36   | 4.42±5.28 | 4.07±4.79   |
| **Ghrelin (pmol/l)** | 31.49±9.96  | 30.83±7.66 | 31.19±8.93  |
| **Adipsin (µg/ml)**  | 0.63±0.35   | 0.70±0.30 | 0.67±0.33   |
| **PAI-1 (ng/ml)**    | 83.51±78.96 | 83.78±86.79 | 83.63±82.20 |
| **TNF-α (pg/ml)**    | 1.43±1.62   | 1.49±1.64 | 1.46±1.62   |
| **IL-6 (pg/ml)**     | 2.24±2.42   | 2.21±2.18 | 2.22±2.30   |
| **IL-18 (pg/ml)**    | 41.75±27.32 | 39.13±24.47 | 40.54±25.93 |
| **VEGF (pg/ml)**     | 106.26±90.91 | 117.81±104.69 | 111.59±97.13 |
| **HGF (pg/ml)**      | 351.82±185.87 | 385.22±209.67 | 367.23±196.80 |
| **Ins-CRP (mg/l)**   | 0.50±0.67   | 0.97±4.44 | 0.72±3.04   |

Women vs. Men:

* p<0.05,
**p<0.001; Student’s t Test after testing for equality of variance (Levene test) and after natural log transformation of several variables (ln).

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direct correlation with viral load [20]; in patients with acromegaly, the VAI appears to be associated with disease activity, adiponectin levels and insulin sensitivity and secretion, and is influenced independently of GH levels [21].

However, in all these studies the ability of the VAI to express the adipose tissue function was only a hypothesis, few studies existing that have evaluated the correlation between it and levels of adipocytokine [9,17,21].

The aim of this study is to evaluate the relationship between the VAI and other common anthropometric measures of fat distribution [BMI, WC, Hip Circumference (HC), Waist/Hip ratio (WHR), Body Adiposity Index (BAI)], on the one hand, and a complete panel of adipocytokines [Resistin, Leptin, Soluble Leptin Receptor (sOB-R), Leptin/sOB-R ratio, Adiponectin, Visfatin, Ghrelin, Adipsin, PAI, Vascular-endothelial growth factor (VEGF), TNF-α, hs-CRP, IL-6, IL-18, Hepatocyte growth factor (HGF)], on the other, in patients with type 2 diabetes mellitus (DM2).

Materials and Methods

Subjects
This study was approved by the Institutional Review Board at the Faculty of Medicine of the University of Palermo. At the time of observation all patients regularly signed an informed consent for the scientific use of their data.

Ninety-one consecutive Caucasian patients with DM2 (mean age 62.25±6.38 years; range: 51–75), followed up in our dedicated

Table 2. Bivariate correlations between anthropometric measures and adipocytokine, lipid and glycemic parameters.

|                  | BMI       | WC        | HC         | WHR       | BAI       | VAI       |
|------------------|-----------|-----------|------------|-----------|-----------|-----------|
| Adipocytokines   |           |           |            |           |           |           |
| Ln Leptin        | 0.454***  | 0.400***  | 0.535***   | −0.045    | 0.542***  | 0.239*    |
| sOb-R            | −0.436*** | −0.412*** | −0.300**   | −0.204    | −0.239*   | −0.278**  |
| Ln Leptin/sOb-R  | 0.491***  | 0.435***  | 0.526***   | 0.008     | 0.521***  | 0.270**   |
| Ln Adiponecit    | −0.002    | 0.053     | 0.109      | −0.060    | 0.006     | −0.330**  |
| Visfatin         | 0.116     | 0.115     | 0.093      | 0.036     | 0.087     | 0.375***  |
| Ln Resistin      | 0.045     | 0.217*    | 0.103      | 0.156     | −0.028    | 0.379***  |
| Ghrelin          | −0.228*   | −0.296**  | −0.108     | −0.263*   | −0.045    | −0.225*   |
| Adipsin          | −0.008    | 0.115     | 0.078      | 0.055     | 0.004     | 0.025     |
| PAI-1            | 0.013     | 0.107     | 0.008      | 0.153     | −0.004    | 0.086     |
| Ln TNF-α         | −0.007    | 0.003     | −0.206     | 0.213*    | −0.180    | 0.170     |
| IL-6             | 0.145     | 0.187     | 0.015      | 0.233*    | 0.083     | 0.300**   |
| Ln IL-18         | −0.052    | −0.095    | 0.001      | −0.134    | 0.076     | 0.340**   |
| VEGF             | 0.072     | 0.097     | −0.097     | 0.250*    | −0.197    | 0.211*    |
| HGF              | 0.076     | 0.200     | 0.065      | 0.177     | 0.039     | 0.298**   |
| Ln hs-CRP        | 0.155     | 0.233*    | 0.171      | 0.120     | 0.158     | 0.419***  |
| LIPID PATTERN    |           |           |            |           |           |           |
| Total Cholesterol| −0.082    | −0.165    | −0.078     | −0.122    | 0.046     | 0.316**   |
| HDL Cholesterol  | −0.087    | −0.065    | 0.113      | −0.194    | 0.145     | −        |
| Triglycerides    | 0.107     | 0.051     | 0.069      | 0.003     | 0.115     | −        |
| LDL Cholesterol  | −0.075    | −0.152    | −0.144     | −0.041    | −0.062    | 0.286**   |
| NEFA             | 0.060     | −0.011    | 0.042      | −0.051    | 0.025     | 0.327**   |
| GLYCEMIC PATTERN |           |           |            |           |           |           |
| HOMA-IR          | 0.373***  | 0.465***  | 0.421***   | 0.122     | 0.236*    | 0.324**   |
| Fasting C-Peptide| 0.228*    | 0.250*    | 0.325**    | −0.028    | 0.129     | 0.228*    |
| Fasting Insulin  | 0.336**   | 0.449***  | 0.426***   | 0.097     | 0.220*    | 0.329**   |
| Fasting glucose  | 0.184     | 0.262*    | 0.098      | 0.216*    | 0.075     | 0.031     |
| 2 h after breakfast glucose | −0.067 | 0.026 | −0.105 | 0.162 | −0.076 | 0.166 |
| 2 h after lunch glucose | 0.024 | 0.089 | 0.006 | 0.089 | −0.037 | 0.069 |
| 2 h after dinner glucose | 0.070 | 0.157 | 0.094 | 0.078 | 0.085 | 0.128 |
| HbA1c            | 0.127     | 0.268*    | 0.132      | 0.202     | 0.125     | 0.250*    |

BMI: Body Mass Index; WC: Waist Circumference; HC: Hip Circumference; WHR: Waist/Hip ratio; BAI: Body Adiposity Index; VAI: Visceral adiposity Index; sOb-R: Soluble leptin receptor; PAI-1: Plasminogen activator inhibitor-1; TNF-α: Tumor necrosis factor alpha; IL-6: Interleukin 6; IL-18: Interleukin 18; VEGF: Vascular endothelial growth factor; HGF: Hepatocyte growth factor; hs-CRP: high-sensitivity C reactive protein.

Bivariate Pearson correlation: the value indicates $r$ coefficient.

*p < 0.050;**p < 0.010; ***p < 0.001;
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Outpatients Clinic (from January 1st 2011 to December 31st 2012), were cross-sectionally studied. Inclusion criteria were a 2-year or longer history of DM2 at study entry, in stable treatment for the last six months with metformin (1.5–2 g/day). Only patients were included who had HbA1c ≥8 (64 mmol/mol), in order to avoid the effects of severe glucotoxicity on the adipocytokines. The following were excluded: patients with type 1 diabetes, patients with previous treatment with other anti-diabetic drugs in the past six months, patients with morbid obesity, pendulous abdomen, severe hypertriglyceridemia and/or or use of...
fibrates [22]. Also excluded were patients with macro- or microvascular complications (except 7 patients with simple preproliferative retinopathy). Patients were not excluded that had essential hypertension (58 patients). In the patients included a complete medical history, a complete physical examination, body weight, height, waist and hip circumference were recorded. Waist circumference (WC) was measured midway between the inferior margin of the last rib and the crest of the ilium in a horizontal plane by the measurer sitting by the subject and fitting the tape snugly, but not compressing soft tissues. Hip circumference (HP) was measured around the pelvis at the point of maximal protrusion of the buttocks. Fasting blood samples and a morning spot urine sample were collected for biochemical analyses. All participants were instructed to perform home glucose monitoring (one full day glucose profile per week: fasting, 2 h after breakfast, 2 h after lunch and 2 h after dinner).

Anthropometric Indices

BMI was calculated as body weight (in kilograms) divided by the square of height (in meters). WHR and was calculated by dividing the waist circumference by the hip circumference. Body adiposity Index (BAI) was calculated using the following formula [23]:

\[
BAI = ((\frac{HC}{\text{height}}) \times 1.5) - 1.8
\]

HC is expressed in centimeters and height in meters.

The VAI was calculated as described [8,22] using the following sex-specific equation, where TG is triglyceride levels expressed in mmol/l and HDL is HDL-cholesterol levels expressed in mmol/l:

\[
\text{Females: } VAI = \left( \frac{WC}{36.58 + (1.89 \times BMI)} \right) \times \left( \frac{TG}{0.81} \right) \times \left( \frac{1.52}{HDL} \right)
\]

\[
\text{Males: } VAI = \left( \frac{WC}{39.68 + (1.88 \times BMI)} \right) \times \left( \frac{TG}{1.03} \right) \times \left( \frac{1.31}{HDL} \right)
\]

**Table 3. Differences in anthropometric and glycometabolic characteristics between cluster 1 and cluster 2.**

|                      | Cluster 1 | Cluster 2 |
|----------------------|-----------|-----------|
|                      | No 36     | No 55     |
| Age (years)          | Mean ± SD | Mean ± SD | p        |
|                      | 62.83 ± 5.48 | 61.87 ± 6.92 | 0.486    |
| Duration of disease (years) | 10.56 ± 6.45 | 9.18 ± 5.66 | 0.301    |
| BMI (Kg/m²)          | 28.33 ± 4.08 | 30.15 ± 4.90 | 0.060    |
| Waist Circumference (cm) | 100.36 ± 12.07 | 103.93 ± 10.76 | 0.155    |
| Hip Circumference (cm) | 107.61 ± 11.19 | 107 ± 9.98 | 0.791    |
| Waist/Hip ratio      | 0.93 ± 0.08 | 0.97 ± 0.08 | 0.036    |
| Body Adiposity Index | 33.27 ± 6.53 | 34.41 ± 6.26 | 0.412    |
| Visceral Adiposity Index | 1.57 ± 0.75 | 2.76 ± 1.47 | <0.001   |
| HbA1c (%)            | 6.83 ± 0.59 | 6.97 ± 0.66 | 0.285    |
| HbA1c (mmol/mol)     | 51 ± 6.4 | 53 ± 7.2 | 0.285    |
| HOMA-IR              | 2.46 ± 2.59 | 3.53 ± 3.51 | 0.099    |
| Fasting Insulin (pmol/l) | 54.40 ± 60.47 | 72.06 ± 71.14 | 0.208    |
| Fasting C-peptide (nmol/l) | 0.13 ± 0.10 | 0.16 ± 0.18 | 0.229    |
| Fasting glucose (mmol/l) | 7.36 ± 1.27 | 7.69 ± 1.69 | 0.292    |
| 2 h after breakfast glucose (mmol/l) | 6.79 ± 1.07 | 6.86 ± 1.74 | 0.806    |
| 2 h after lunch glucose (mmol/l) | 7.51 ± 1.41 | 7.59 ± 1.96 | 0.821    |
| 2 h after dinner glucose (mmol/l) | 7.31 ± 1.74 | 7.98 ± 1.98 | 0.093    |
| Urinary Albumin-to-Creatinine Ratio (UACR) (mg/g) | 19.83 ± 55.35 | 14.41 ± 23.15 | 0.581    |

Differences in anthropometric and glycometabolic characteristics between cluster 1 and cluster 2. Student’s t Test after testing for equality of variance (Levene test). doi:10.1371/journal.pone.0091969.t003

**Assay**

The BioPlex Pro Human Diabetes 10-plex assay (BioRad, Milan, Italy) was used to quantitate the following diabetes-related analytes: C-peptide (nmol/l), Ghrelin (pg/ml), Leptin (ng/ml), PAI-1 (ng/ml), Resistin (ng/ml) and visfatin (pg/ml). The BioPlex Pro Human Diabetes Adipsin and Adiponectin duplex assay (BioRad, Milan, Italy) was used for Adipsin (ng/ml) and Adiponectin (µg/ml) assays. A customized BioPlex 3-plex assay (BioRad, Milan, Italy) was used for the quantitation of serum IL-6 (pg/ml), TNF-α (pg/ml) and VEGF (pg/ml). A customized BioPlex duplex assay (BioRad, Milan, Italy) was used for the quantitation of IL-18 (pg/ml) and HGF (pg/ml). All these assays were run on a BioPlex 200 workstation (BioRad, Milan, Italy). Human sOb-R (ng/ml) was assayed using an ELISA sandwich enzyme immunoassay (Human leptin receptor ELISA, BioVendor, Heidelberg, Germany). Serum non-esterified fatty acids (NEFA) (mmol/l) were assayed after overnight fasting using an enzymatic colorimetric method (Randox NEFA assay FA115, Randox Laboratories, County Antrim, UK). CRP (mg/l) was determined by a high
sensitivity immunonephelometric method (CardioPhase hs CRP, SIEMENS) on the BNTN II System (SIEMENS). Blood glucose levels (mg/dl) were measured using an electrochemical system (Glucocard, Menarini Diagnostics, Italy). Total cholesterol, HDL cholesterol, triglycerides and urinary albumin and creatinine were measured in our laboratory using standard assays. HbA1c was determined by High Pressure Liquid Chromatography (HPLC) with ion-exchange resin (HA8121, Hi-AutoA1c, Menarini, Italy). LDL cholesterol levels were calculated with Friedewald’s formula. The conversion factors for the International System (SI) were the following: glucose (mg/dl vs. mmol/l; X 0.0555), insulin (mUI/l vs. pmol/l; X 6.945), total cholesterol (mg/dl vs. mmol/l; X 0.0259), visfatin (pg/ml vs. ng/ml; :1000), Ghrelin (pg/ml vs. pmol/l; X 0.296), Adipsin (ng/ml vs. mg/ml; :1000).

Statistical Methods

The Statistical Packages for Social Sciences SPSS version 17 and MedCalc version 11.3 were used for data analysis. Baseline characteristics were presented as mean ± Standard Deviation (SD) for continuous variables; rates and proportions were calculated for categorical data. Normality of distribution for quantitative variables was assessed by the Shapiro-Wilk test. Some variables (Leptin, Leptin/sOb-R ratio, Adiponectin, Resistin, TNF-α, IL-18 and hs-CRP) did not show normal distribution, and the natural logarithmic transformed (Ln) values of each adipocytokine for statistical analysis were used. Differences between the two groups were detected by the unpaired Student’s t test for continuous variables (after testing for equality of variance: Levene test) and by the χ²-test and Fisher’s exact test (when appropriate) for categorical variables. Correlations among anthropometric indices and adipocytokines were analysed using the Pearson’s test.

In order to identify patients with Hypothetical ATD (HATD), we performed a two-step cluster analysis using log-likelihood distance measures. This technique can detect latent relationships within a complex data set between patients with multiple distinct characteristics. Cluster analysis was applied using an a priori number of fixed clusters (cluster 1 and cluster 2) and the following continuous variables were included in the analysis: Ln Leptin/sOb-R ratio, Ln Adiponectin, Ln Resistin, Ln NEFA and Ln TNF-α. The selection of the variables included in the analysis was made on the basis of the Silhouette Coefficient obtained by including various combinations of adipocytokines. The 5 variables included produced a Silhouette Coefficient >0.5, indicative of a good partitioning of data. The two clusters identified were designated as “normal adipose tissue function” (cluster 1) and HATD (cluster 2). For comparison between cluster 1 and cluster 2, the group sizes gave 74.84% power to detect a moderate effect size (Cohen’s d = 0.5) using T-test, with alpha at 0.05. Post-hoc power analysis was performed using G*Power Version 3.1.6 software. The receiver-operating characteristic (ROC) curve analysis was conducted using the MedCalc software v. 9.3.8.0 for Windows [which uses calculation of the area under the curve (C-statistic) and 95% confidence intervals by the technique of Hanley and McNeil], to evaluate the diagnostic performance of the various anthropometric indices (BMI, WC, HC, WHR, BAI, VAI) for predicting HATD. The differences between C-statistics were calculated by the method of Hanley and McNeil.

A p value of <0.05 was considered statistically significant.

Results

The 91 patients studied consisted in 49 women and 42 men; the clinical characteristics, the glycometabolic state and the adipocytokine pattern are summarized in Table 1. Regarding the
anthropometric indices, the women showed a higher BMI (30.35±4.94 vs. 28.36±4.09 kg/m², p = 0.039), HC (110.30±11.66 vs. 103.66±7.40 cm, p = 0.001), BAI (37.09±6.27 vs. 30.31±4.17, p<0.001), VAI (2.64±1.56 vs. 1.87±0.94, p = 0.005) and a lower WHR (0.93±0.10 vs. 0.98±0.04, p = 0.012). Regarding the adipocytokine profile, only significantly higher levels of leptin (13.74±6.02 vs. 3.59±4.79 ng/dl, p<0.001) and significantly lower levels of sOb-R (19.89±6.18 vs. 23.10±6.40 ng/ml, p = 0.018) were found in women as compared to men. No other gender difference was found.

Comparison of Various Anthropometric Indices with the Adipocytokines, Lipid and Glycemic Parameters

**BMI.** BMI showed a significant positive correlation with Ln Leptin, Ln Leptin/sOb-R ratio, HOMA IR, fasting C-peptide and fasting Insulin; by contrast, a significant inverse correlation was observed with sOb-R and Ghrelin (Table 2).

**WC.** WC showed a significant positive correlation with Ln Leptin, Ln Leptin/sOb-R ratio, Ln Resistin, Ln hs-CRP, HOMA IR, fasting C-peptide and fasting Insulin, fasting Glucose and HbA1c; a significant inverse correlation was observed with sOb-R and Ghrelin (Table 2).

**HC.** HC, excluding Ghrelin, showed similar significant correlations to BMI (Table 2).

**WHR.** WHR significantly correlated with Ghrelin, Ln TNF-α, IL-6, VEGF and fasting glucose.

**BAI.** BAI showed significant correlations only with Ln Leptin, sOb-R, Ln Leptin/sOb-R ratio, HOMA IR and fasting Insulin.

**VAI.** VAI is the anthropometric index that showed the largest number of correlations with adipocytokines. Significant positive correlations were found with Ln Leptin, Ln Leptin/sOb-R ratio, Visfatin, Ln Resistin, IL-6, Ln IL-18, VEGF, HGF, Ln hs-CRP, Total Cholesterol, LDL Cholesterol, NEFA, HOMA IR, fasting C-peptide and fasting Insulin. Significant inverse correlations were observed with sOb-R, Ghrelin and Ln Adiponectin.

Two-Step Cluster Analysis

In order to identify a group of patients with the most altered adipocytokine profile, classifiable as patients with HATD, a two-step cluster analysis was performed (based on the selection of Ln Leptin/sOb-R ratio, Ln Adiponectin, Ln Resistin, Ln NEFA and Ln TNF-α). Two clusters were identified (Cluster 1 = 36 patients; Cluster 2 = 55 patients) of which cluster 2 showed the worst adipocytokine profile (Figure 1). Regarding the specific parameters of diabetes (duration of disease, HbA1c, HOMA IR, glycemic profile), there were no significant differences between the two clusters (Table 3). Significant differences were observed for lipids and adipocytokines. Indeed, cluster 2 showed significantly higher levels of triglycerides (1.58±0.61 vs. 1.14±0.38 mmol/l, p<0.001), NEFA (0.54±0.28 vs. 0.36±0.20 mmol/l, p = 0.001) and lower levels of HDL cholesterol (1.43±0.37 mmol/l, p<0.001); no difference was found for total cholesterol and LDL cholesterol. Furthermore, cluster 2 identified patients with higher levels of Ln Leptin (1.78±1.22 vs. 1.15±1.18 ng/dl, p = 0.017), Ln Leptin/sOb-R ratio (1.17±0.91 vs. 0.82±0.60 ng/ml, p = 0.031), Visfatin (0.74±0.61 vs. 0.45±0.33 ng/ml, p = 0.014), Ln TNF-α (0.32±0.89 vs. 0.81±0.34 pg/ml, p<0.001), Ln hs-CRP (411.45±193.95 vs. 299.67±183.32 pg/ml, p = 0.007) and lower levels of sOb-R (23.16±5.16 ng/ml, p = 0.022) and Ln Adiponectin (1.17±1.22 vs. 2.83±0.81 μg/ml, p<0.001). No significant differences were found for Adipsin, Ghrelin, PAI-1, IL-6 and IL-18.
ROC Curve Analysis

ROC curve analysis was performed for BMI, WC, HC, WHR, BAI and VAI to evaluate their diagnostic performance for predicting an HATD (Table 4).

C-statistics were then compared to identify which parameter was a better indicator for HATD. The C-statistics were 0.588 (95% CI 0.48-0.69) for BMI, 0.573 (95% CI 0.46-0.67) for WC, 0.507 (95% CI 0.40-0.61) for HC, and 0.574 (95% CI 0.46-0.67) for WHR, 0.537 (95% CI 0.42-0.64) for BAI and 0.767 (95% CI 0.66-0.84) for VAI. VAI showed a significantly higher C-statistic compared to BMI (difference: 0.178; 95% CI: 0.03-0.32; SE: 0.07; p = 0.0014), WC (difference: 0.194; 95% CI: 0.04-0.34; SE: 0.07; p = 0.009), HC (difference: 0.260; 95% CI: 0.11-0.40; SE: 0.07; p < 0.001), WHR (difference: 0.193; 95% CI: 0.04-0.33; SE: 0.07; p = 0.009) and BAI (difference: 0.230; 95% CI: 0.08-0.37; SE: 0.07; p = 0.001) (figure 2). No difference in C-statistic was found among the other anthropometric indices.

Conclusions

We demonstrate in this study that the VAI is a good indicator of an altered adipokine profile and its associated cardiometabolic risk.

The emerging paradigm of ATD supports the view that adipose tissue dysregulation (altered fat distribution and function) might play a crucial role in the pathogenesis of insulin resistance and atherosclerosis. From this viewpoint, atherosclerotic cardiovascular disease and DM2 seem to arise from a “common soil”, characterized by altered adipokine production, increase in the lipolytic activity of adipose tissue and low-grade inflammation [24–26]. In addition, recently the hypothesis that subcutaneous AT has a limited capacity to increase its mass safely has been gaining increasing recognition [27,28]. This capacity is individually determined by a wide range of environmental and genetic factors. The absence of adaptive hyperplasia of the subcutaneous adipose tissue could be associated with visceral obesity and excess ectopic fat.

We studied the ability of various common anthropometric indices to correlate with a wide panel of adipokines. Though the literature data show a continued involvement of new adipokines, we chose to study some of those that have more scientific evidence about their ability to express a condition of ATD [29,30].

In our DM2 patients there were no gender differences except as regards the leptinemic system: leptin levels are higher in women than in men and the level of its soluble receptor is inversely proportional; this known datum could be an expression of a state of relative leptin resistance in women with diabetes [31]. Also supporting this hypothesis is the observation of higher levels of NEFA in women, which may reflect the reduction of anti-lipotoxic activity carried out by leptin [32].

Among all the anthropometric indices studied the VAI is the one that showed the greatest number of correlations with the adipokines and the lipids (including NEFA). The VAI was able to express both the altered endocrine function of adipose tissue and the state of relative leptin resistance and low-grade inflammation, which are all alterations present in a state of ATD. Particularly strong is the correlation with visfatin (adipocytokine prevalently secreted by visceral adipose tissue, which according to some authors may have adipogenic effects and is a potential mediator of obesity related insulin resistance in rodents, but still today a subject of controversy in human obesity and human epidemiologic studies) [33,34], resistin (a potential mediator of obesity related insulin resistance in rodents, but still today a subject of controversy in human obesity and type 2 diabetes or in other populations, mainly in the absence of an overt metabolic syndrome) [10].

In conclusion, although still lacking prospective studies that can attribute to the VAI a prognostic role on cardiovascular risk, given the simplicity of WC and BMI measurement and Triglycerides and HDL Cholesterol assessment, we suggest that the VAI would be an easy tool for the evaluation of the cardiometabolic risk linked to glucose toxicity (for these aspects the most important parameter is always HbA1c), but it might mirror the residual cardiovascular risk despite the normalization of glycemic, lipid and hemodynamic parameters [41]. Finally, in our study we also identified a precise cut-off of the VAI able to identify a HATD. This cut-off of 2 seems to be comparable to the cut-off point identified, in the same age range, in a previous study (in which in the general population an ATD dysfunction is hypothesized in subjects with metabolic syndrome) [10].

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Author Contributions

Conceived and designed the experiments: MCA. Performed the experiments: MCA VT. Analyzed the data: MCA. Contributed reagents/materials/analysis tools: GP GM SM. Wrote the paper: MCA CG.
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