Feasibility of sputum testing for detection of COVID-19

Arvind Sharma*, Tej Pratap Singh

Department of Community Medicine, N.S.C.B. Medical College, Jabalpur, Madhya Pradesh, India

Received: 22 January 2021
Revised: 25 March 2021
Accepted: 30 March 2021

*Correspondence:
Dr. Arvind Sharma,
E-mail: drarvindsharmajbp@yahoo.co.in

Copyright: © the author(s), publisher and licensee Medip Academy. This is an open-access article distributed under the terms of the Creative Commons Attribution Non-Commercial License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

ABSTRACT

For the prevention of COVID-19 spread, early and accurate detection is important. Specimens are collected through respiratory mucosal surfaces with nasopharyngeal or oropharyngeal swabs, from infected patients are highly complex. Sputum testing could preferably be a more convenient technique for the detection of COVID-19 as being noninvasive method, which could easily be collected by having a patient cough deeply to produce and expel phlegm which could be in compliance to patient in comparison to Nasopharyngeal swab (NPS). Consequently, broader testing than the current methods of nasal or throat swabs will significantly increase the number of people screening, leading to more effective control of the spread of COVID-19. Nonetheless, a comparison of the saliva-based assay with current swab test is needed to understand what and how we can benefit from this newly developed assay. Therefore, in this review article, we aimed to summarize the feasibility of sputum testing in relation to Covid-19. Any implementation of clinical sampling for diagnosis should take into considerations of the sensitivity of assays, risks to healthcare professionals, and global shortage of equipment.

Keywords: COVID-19, SARS-CoV-2, Sputum sample, Sputum testing

INTRODUCTION

In late December 2019, a cluster of zoonotic pneumonia cases in Wuhan, China, was reported and galloping pace over the world. Since then, the 2019 novel coronavirus was categorized as a public health outbreak of international concern by the World Health Organization on 30 January 2020, and therefore, this was declared as global pandemic on 11 March 2020, and the disease was named COVID-19.

Currently, viral loads are routinely measured to monitor severe viral respiratory tract infections for clinical progression, response to treatment, cure and relapse. To diagnose COVID-19, Nucleic acid testing, most commonly Reverse Transcriptase Polymerase Chain Reaction (RT-PCR), plays a significant role. Specimens are collected through respiratory mucosal surfaces with nasopharyngeal or oropharyngeal swabs, from infected patients, as recommended by the WHO for the detection of Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2), which eventually require adequate human resources like trained medical staff with their expertise, guards to avoid overcrowding, and appropriate logistics for timely and effective diagnosis. The same guideline also recommended collection of lower respiratory specimens: sputum (if produced) and/or endotracheal aspirate or bronchoalveolar lavage in patients with more severe respiratory disease.

Consideration to use sputum as a diagnostic fluid

COVID-19 pandemic brought diagnostic and technical challenges to testing laboratories, for its ability to provide accurate and rapid test results, as lack of attention on various parameters like, sample collection (i.e., poor-quality collection), specimen collection devices (including swab material and transport media), specimen transport and storage, testing outside the diagnostic window, and non-compliance to the patient as the process...
of NPS sampling is painful as well as unaccepted by many in the community. These parameters would highly raise the possibility of false-negative results, which seems to be contagious for the community and also found delayed to mitigate the transmission. So as to overcome these challenges, sputum testing could preferably be more convenient technique for the detection of COVID-19 as being non-invasive method, which could easily be collected by having a patient cough deeply to produce and expel phlegm which could be compliance to patient in comparison to Nasopharyngeal swab (NPS). Secondly, it also provides with less expertise staff, fewer logistics, easy to collect on the patient’s home and the site, no aerosol generation, and no transport media required for storage. In NTEP, all peripheral health workers are previously sensitized to the procedure of sample collection and they have already trained regarding the process therefore health care workers, as well as community, are already accustomed with the process of sputum collection. Home collection of samples could be advised for sputum collection of suspects along with integrated TB-COVID laboratory services under NTEP program, which would decrease the burden of medical colleges and district hospitals laboratories.

In a systematic review and meta-analysis conducted, concluded that the detection rate of SARS-CoV-2 for oropharyngeal swabs were 43%, for NP swabs 54%; and for sputum, it was maximum i.e., 71%. The SARS-CoV-2 was most reliably detected in sputum samples, which contained the highest viral load, followed by nasal swabs. It revealed that options of sputum test, are more accurate in diagnosing SARS-CoV-2 compared to the NPS, despite of the NPS being the most commonly used test for diagnosis. Various other studies also reported that in COVID-19 diagnosed individuals, the SARS-CoV-2 detection rate ranged about 32% to 65% for oropharyngeal swabs, 25% to 70% of collected nasopharyngeal swabs, and 48% to 90% for sputum.

Hence, it was found that the rate of SARS-CoV-2 detection was significantly higher in sputum than either oropharyngeal swabs or NP swabs. The higher sensitivity for the detection of COVID-19 in sputum is also supported by previously reported studies on the detection of other non-COVID-19 respiratory viruses.

**DISCUSSION**

As mentioned previously, an accurate identification of respiratory viruses are critically affected by the source of clinical specimens. While several studies on up to 15 common respiratory viruses suggest that the use of nasopharyngeal swabs provides a higher sensitivity than that of nasopharyngeal washings or oropharyngeal swabs, this is not necessarily the case for SARS-CoV-2, as the infectivity and the predilection for transmission may differ significantly between viruses. In addition, even if a given type of clinical specimen offers a relatively higher accuracy in diagnosis, it remains an open question whether the technique-demanding test is the most needed during a pandemic with the global shortage of medical supplies as of today. Study done by Chen et al reveals that some patients who develop a productive cough at the later course of the disease may still be positive with SARS-CoV-2 RNA in the sputum despite the NPS became negative. Similarly, Wolfel et al. have demonstrated the presence of infectious viral particles in the lower respiratory tract, the detection of SARS-CoV-2 RNA in sputum, particularly those with low to moderate CT values, likely indicate the possibility of persistent infection. A testing algorithm for sputum may need to be developed with the support of various other branches of medical specialists. Therefore, testing of sputum may be necessary for a certain patient population indicated as below: (i) hospitalized individuals...
with endotracheal intubation before the NPS samples could be taken for COVID-19 evaluation (endotracheal aspiration may be indicated in this situation); (ii) patients by the Centre for Disease Control. However, self-collected sputum (with proper instructions) in a defined patient population would provide several advantages over NPS, such as the discomfort associated with NPS sampling.

Table 1: Summary description of studies and reports included, with sample size and key findings.

| Authors            | Sample size and respondents                                                                 | Key findings                                                                                                                                 |
|--------------------|---------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------|
| Mohammadi et al.   | 11 studies that met inclusion criteria, with SARS-CoV-2 testing results from a total of 3442 respiratory tract specimens. | Compared to NPS sampling, sputum testing resulted in significantly higher rates of SARS-CoV-2 RNA detection while OPS testing had lower rates of viral RNA detection. |
| Falsey Ann et al.  | A total of 532 subjects admitted for 556 respiratory illnesses were enrolled. A total of 189 virus strains were identified by RT-PCR | 23% were positive by NTS (Nose throat swab) alone, 33% were positive only with sputum samples.                                               |
| Jeong et al.       | 154 adults who were admitted or presented to the clinics of Gil Medical Center with acute respiratory symptoms were recruited from 1 November 2012 to 31 March 2013. 154 specimens of nasopharyngeal swabs and sputum were taken. | The positive rate was 53% (81/154) for nasopharyngeal swabs and 68% (105/154) for sputum (p<0.001).                                                  |
| Branche et al.     | During the four winters from 2008 to 2012, 965 respiratory illnesses were evaluated. A viral infection was identified in 295 of 965 patients (31%), of which 7 subjects had two different viruses for a total of 302 viruses identified using the in-house uniplex RT-PCR assays. | Of the 302 viral detections, 124 (41%) were positive in both the Nose Throat Swab and sputum samples, 105 (35%) were positive by the sputum sample alone, and 73 (24%) were positive by the NTS alone. |
| Chenyao Lin et al. | Paired specimens of throat swabs and sputum were obtained from 52 COVID-19 suspected patients. All patients received RT-PCR assays in both throat swabs and sputum specimens at the same time. | The positive rates of SARS-CoV-2 from sputum specimens was 76.9% and for throat swabs 44.2%. The findings showed that the positive rate from sputum specimens was significantly higher than that from throat swabs (p=0.001) |

CONCLUSION

The current procedures, however, are technically complex, often inconvenient for patients, and require personal protective equipment. Sputum testing, on the other hand, is feasible and requires less expertise staff also has readily accepted in the community. The diagnostic testing is crucial for controlling the COVID-19 pandemic. Any implementation of clinical sampling for diagnosis should take into considerations of the sensitivity of assays, risks to healthcare professionals, and the global shortage of equipment.

Funding: No funding sources
Conflict of interest: None declared
Ethical approval: Not required

REFERENCES

1. World Health Organization. Coronavirus disease (COVID-19) situation reports, 2020. Available at: https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200121-sitrep-1-2019-ncov.pdf?sfvrsn=20a99c10_4. Accessed on 15 Dec 2020.
2. World Health Organization. Coronavirus disease (COVID-19) situation reports, 2020. Available at: https://www.who.int/docs/default-source/coronaviruse/situation-reports/20200311-sitrep-51-covid-19.pdf?sfvrsn=1ba62e57_10. Accessed on 15 Dec 2020.
3. WHO Laboratory testing for 2019 novel coronavirus (2019-nCoV) in suspected human cases. Interim guidance, Jan 17, 2020. Available at://C:/Users/HP/Downloads/WHO-COVID-19-
Effect of Throat Washings on Detection of SARS-CoV-2 in Adults with Respiratory Illness: A Systematic Review and Meta-Analysis. EBioMedicine. 2020;59:102903.

5. Yoon JG, Yoon J, Song JY, Yoon SY, Lim CS, Seong H, et al. Clinical Significance of a High SARS-CoV-2 Viral Load in the Saliva. J Korean Med Sci. 2020;35(20):195.

6. Yu F, Yan L, Wang N, Yang S, Wang L, Tang Y, et al. Quantitative Detection and Viral Load Analysis of SARS-CoV-2 in Infected Patients. Clin Infect Dis. 2020;71(15):793-98.

7. Kim ES, Chin BS, Kang CK, Kim NJ, Kang YM, Choi JP, et al. Korea National Committee for Clinical Management of COVID-19: Clinical Course and Outcomes of Patients with Severe Acute Respiratory Syndrome Coronavirus 2 Infection: a Preliminary Report of the First 28 Patients from the Korean Cohort Study on COVID-19. J Korean Med Sci. 2020;35(13):142.

8. Wang W, Xu Y, Gao R, Lu R, Han K, Wu G, et al. Detection of SARS-CoV-2 in Different Types of Clinical Specimens. JAMA. 2020;323(18):1843-4.

9. Wu J, Liu J, Li S, Peng Z, Xiao Z, Wang X, et al. Detection and analysis of nucleic acid in various biological samples of COVID-19 patients. Travel Med Infect Dis. 2020;37:101673.

10. Pan Y, Zhang D, Yang P, Poon LLM, Wang Q. Viral load of SARS-CoV-2 in clinical samples. Lancet Infect Dis. 2020;20(4):411-2.

11. Zhang W, Du RH, Li B, Zheng XS, Yang XL, Hu B, et al. Molecular and serological investigation of 2019-nCoV infected patients: implications of multiple shedding routes. Emerg Microbes Infect. 2020;9(1):386-9.

12. Guo WL, Jiang Q, Ye F, Li SQ, Hong C, Chen LY, et al. Effect of Throat Washings on Detection of 2019 Novel Coronavirus. Clin Infect Dis. 2020;71(8):1980-1.

13. Han H, Luo Q, Mo F, Long L, Zheng W. SARS-CoV-2 RNA more readily detected in induced sputum than in throat swabs of convalescent COVID-19 patients. Lancet Infect Dis. 2020;20(6):655-656.

14. Falsoye AR, Formica MA, Walsh EE. Yield of sputum for viral detection by reverse transcriptase PCR in adults hospitalized with respiratory illness. J Clin Microbiol. 2012;50(1):21-4.

15. Jeong JH, Kim KH, Jeong SH, Park JW, Lee SM, Seo YH. Comparison of sputum and nasopharyngeal swabs for detection of respiratory viruses. J Med Virol. 2014;86(12):2122-7.

16. Branche AR, Walsh EE, Formica MA, Falsoye AR. Detection of respiratory viruses in sputum from adults by use of automated multiplex PCR. J Clin Microbiol. 2014;52(10):3590-6.

17. Lieberman D, Lieberman D, Shimoni A, Keren-Naas A, Steinberg R, Shemer-Avni Y. Identification of respiratory viruses in adults: nasopharyngeal versus oropharyngeal sampling. J Clin Microbiol. 2009;47(11):3439-43.

18. Li L, Chen QY, Li YY, Wang YF, Yang ZF, Zhong NS. Comparison among nasopharyngeal swab, nasal wash, and oropharyngeal swab for respiratory virus detection in adults with acute pharyngitis. BMC Infect Dis. 2013;13:281.

19. Mandel ID. The diagnostic uses of saliva. J Oral Pathol Med. 1990;19(3):119-25.

20. Chen L, Zhao J, Peng J, Li X, Deng X, Geng Z, et al. Detection of SARS-CoV-2 in saliva and characterization of oral symptoms in COVID-19 patients. Cell Prolif. 2020;53(12):12923.

21. Hung KF, Sun YC, Chen BH, Lo JF, Cheng CM, Chen CY, et al. New COVID-19 saliva-based test: How good is it compared with the current nasopharyngeal or throat swab test? J Chin Med Assoc. 2020;83(10):891-4.

22. Chen C, Gao G, Xu Y, Pu L, Wang Q, Wang L, et al. SARS-CoV-2-Positive Sputum and Feces After Conversion of Pharyngeal Samples in Patients With COVID-19. Ann Intern Med. 2020;172(12):832-4.

23. Wölfel R, Corman VM, Guggemos W, Seilmaier M, Zange S, Müller MA, et al. Virological assessment of hospitalized patients with COVID-19. Nature. 2020;581(7809):465-9.

24. Thwe PM, Ren P. Analysis of sputum/tracheal aspirate and nasopharyngeal samples for SARS-CoV-2 detection by laboratory-developed test and Panther Fusion system. Diagn Microbiol Infect Dis. 2021;99(1):115228.

25. Lin C, Xiang J, Yan M, Li H, Huang S, Shen C. Comparison of throat swabs and sputum specimens for viral nucleic acid detection in 52 cases of novel coronavirus (SARS-CoV-2)-infected pneumonia (COVID-19). Clin Chem Lab Med. 2020;58(7):1089-4.

Cite this article as: Sharma A, Singh TP. Feasibility of sputum testing for detection of COVID-19. Int J Res Med Sci 2021;9:1494-7.