Association of vitamin D receptor gene polymorphisms with metabolic syndrome: a case–control design of population-based cross-sectional study in North China

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Abstract

Background: Metabolic syndrome (MS) increases a risk of developing cardiovascular disease (CVD) and type 2 diabetes mellitus (T2DM). The Vitamin D Receptor gene (VDR) may be important for developing MS. The aim of this study is to investigate the correlation between the VDR gene polymorphisms and MS in North China.

Methods: A case–control study included 391 participants with MS according to the MS diagnostic criteria of International Diabetes Federation 2005 (IDF2005) and 400 controls was conducted on the basis of a cross sectional study which was performed from 2008 to 2012 in Ningxia Hui Autonomous Region, China. Anthropometric data, blood pressure and blood samples were collected in the field investigation. Blood biochemistry analyses were carried out in the laboratory. Two single-nucleotide polymorphisms (SNPs) in the VDR gene, BsmI (rs1544410 A > G) and FokI (rs 2228570 C > T), were genotyped.

Results: The difference in the occurrence of genotypes in BsmI between individuals with MS and the control group was significant. Compared with genotype Bb/bb and allele b, genotype BB and allele B showed higher frequencies in MS cases than controls, which suggested they were risk factors. In addition, the genotype BB carriers with MS presented a higher waist circumference, while genotype FF for the FokI polymorphism was correlated with lower BMI in subjects with MS.

Conclusion: Our study suggests that the VDR gene polymorphisms appear to be associated with MS in the Northern Chinese population. Allele B and BB genotype for BsmI are risk factors for MS. The BsmI polymorphism seems to influence waist circumference, while the FokI polymorphism influence BMI in subjects with MS.

Keywords: Vitamin D Receptor gene, Polymorphisms, Metabolic syndrome

Background

Metabolic syndrome (MS) includes a variety of metabolic components in abnormal pathological state, which contains abnormal plasma glucose, dyslipidemia, high blood pressure, and high cholesterol [1]. Patients with MS have a greater risk of developing cardiovascular disease (CVD) and type 2 diabetes mellitus (T2DM) [2-4].

For the past few years, the prevalence of MS has been on the rise in adults globally, increasing from about 20% to 40% [5-8] in multiple groups of people, and is becoming a serious global health burden. Although insulin resistance [9] and central obesity [10] are considered to be its primary etiological factors, genetics undoubtedly plays an important role as well. In particular, genetic influence may be different among various ethnic groups [11].

The Vitamin D receptor (VDR) is a member of the steroid hormone receptor family that acts as a transcriptional activator of many genes. The DNA polymorphisms that
have been often reported for the VDR gene are (described in both restriction sites and dbSNP info): BsmI (rs1544410 A > G), FokI (rs 2228570 C > T), TaqI (rs731236 T > C), and Apal (rs 7975232 C > T). In past years, many studies have found that these polymorphisms are related to bone mineral density (BMD) [12], calcium metabolism [13], tuberculosis, hepatitis [14] and cancer [15,16]. Additionally, some of these polymorphisms have been recently identified to be associated with type 1 diabetes [17], T2DM [18], and insulin secretion [19].

Although the molecular mechanisms of VDR in components of MS remain unclear, a relationship between 25(OH)D3 and the disorders of MS has been previously demonstrated [20-23]. Studies have also discussed VDR gene polymorphisms for associations with the components of MS, which suggest that two major VDR gene polymorphisms (BsmI and FokI) seemed to influence BMI, insulin resistance, and serum HDL cholesterol [24,25]. However, little is known about the role of VDR gene polymorphisms in MS for Chinese population.

Therefore, we suppose the two single-nucleotide polymorphisms (SNPs), BsmI (rs1544410 A > G) and FokI (rs 2228570 C > T), of the VDR gene intricately affect the development of MS. This study was designed to determine the contribution of VDR polymorphisms to MS in a population from North China.

Materials and methods

Participants

A cross-sectional study was conducted from 2008 to 2012 in Guyuan County, Qingtongxia County and Pingluo County, of Ningxia Hui Autonomous Region, China. Stratified cluster sampling was applied to select two villages (one Hui ethnic village and one Han ethnic village) in each county, totaling six villages. During the study period, 4614 subjects aged 24–75 were interviewed with a questionnaire, followed by anthropometric measurements (height, weight, waist circumference), and blood pressure determination. Pregnant or breastfeeding women, patients with severe mental disease and other serious illness were excluded from the study. The subjects enrolled did not report to have chronic viral infection, cold or acute respiratory infection, or any type of surgery in the week preceding the study. Meanwhile, 2615 participants had blood drawn via venipuncture for laboratory measurements, and finally 1169 blood samples were selected for genotyping by mechanical sampling (k = 1). Based on diagnostic criteria (International Diabetes Federation IDF-2005), 391 cases with MS and 400 controls were selected from the same population to conduct the case–control study. The study protocol had been approved by the Medical Ethics Review Committee of Ningxia Medical University. All participants signed the consents on enrollment after they received written and verbal information about the study.

All participants were interviewed by trained persons with a standard closed-ended questionnaire. The contents of the questionnaire included general demographic characteristics, blood pressure, blood lipid, blood sugar, smoking, physical activity status, drinking and related diseases (including hypertension, coronary heart disease, stroke and diabetes medical history and family history, individual treatment, etc.).

Anthropometric measurements and blood pressure

Standing height was measured using a portable ruler. Body weight was also measured using a weight scale (Omron, China). All measurements were performed by well-trained investigators. The reading of measurements was accurate within 0.1 cm or 0.1 kg, and the average of the two collected measures was recorded for further analysis. Body mass index (BMI) was calculated as kg/m². Obesity was defined as BMI ≥ 30. Arterial blood pressure was measured three times in sitting position using an electronic sphygmomanometer (Omron-HEM 7301-IT, China). All participants were at rest for at least 30 minutes before the measurement was collected. Patients with average blood pressures ≥140/90 mmHg or taking antihypertensive medication in 2 weeks were classified as being hypertensive.

Blood biochemistry analyses

Blood samples were drawn from the antecubital vein at the hours of 6 to 8 AM, after at least 10 hours of fasting and avoidance of alcohol. Two sets of fasting blood samples were collected separately from each subject in sodium fluoride potassium oxalate tubes (for glucose) and lithium heparin vacuum tubes (for lipids). The latter collections were then centrifuged and kept at −80°C until analysis. Fasting glucose was immediately determined by One Touch Ultra 2 (LifeScan, USA). Serum levels of HDL- cholesterol, LDL- cholesterol, and triglycerides were measured by enzymatic assay (CHOD-PAP, Roche Diagnostics GmbH). All analyses were carried out through an automatic biochemical analyzer (COBASE 501, Roche Diagnostics GmbH). Subjects were further divided into two groups, cases with MS and controls without MS, according to the IDF definition. For a subject to be defined as having MS, they must have: Central obesity (waist circumference ≥ 90 cm in males, ≥ 80 cm in females) and BMI > 30 kg/m² along with any two of the following four factors: triglycerides ≥ 150 mg/dL (1.7 mmol/L); HDL-cholesterol < 40 mg/dL (1.03 mmol/L) in males, < 50 mg/dL (1.29 mmol/L) in females; blood pressure, systolic BP ≥ 130 or diastolic BP ≥ 85 mmHg; fasting glucose ≥ 5.6 mmol/L (100 mg/dL) [1].
Genomic DNA was extracted from the whole blood with the SE Blood DNA Kit (OMEGA, USA). After extraction, 30 samples of the extracted material were randomly selected for validation by agarose gel electrophoresis. SNP genotyping of BsmI and FokI in the VDR gene was performed using a technology known as SNPsCan™[26-28] (Genesky Biotechnologies, China). SNPsCan™ is a patent technology and developed on double ligation and multiplex fluorescence PCR with high genotyping accuracy (>99.9%) and call rate (>98%). Specifically, highly multiple ligation products in different length were generated on account of high specificity of the ligase reaction. Meanwhile, the ligation probes and templates were further lengthened through double ligation. With blue fluorescent dye modified universal primers, ligation products were amplified by Thermal Cycler (ABI, USA) and then detected through fluorescent probes. With blue fluorescent dye modified universal primers, ligation products were amplified by Thermal Cycler (ABI, USA) and then detected through fluorescent probes.

Statistical analysis
Statistical analysis was conducted using SPSS statistical software (version 14.0 SPSS Corp, College Station, TX) and descriptive data were presented by mean and standard deviation. The Kolmogorov-Smirnov test was used to analyze normality of the distribution of each variable. Comparisons between any two groups were performed using independent t-tests. Chi-square analysis was applied to examine the variation of sex, genotype and gene frequency in different groups. Odds ratios (ORs) and their 95% confidence interval (CI) were computed for the risk alleles from logistic regression analysis. Statistical significance was set at P < 0.05.

Results
General characteristics and components of MS
Characteristics of the subjects with and without MS are shown in Table 1. In our study, the mean age of the 391 participants with MS was 53 (±11) years, and the mean age of 400 controls was 53 (±11) years. Compared with the controls, BMI, waist circumference, blood pressure, fasting glucose, and triglycerides were significantly higher in individuals with MS, while HDL cholesterol was lower. Both VDR BsmI and FokI genotypic distributions in controls were in Hardy-Weinberg equilibrium (P = 0.762 and 0.376, respectively) (Table 2).

The distribution of VDR BsmI and FokI polymorphisms
As shown in Table 3, both genotypic and allelic frequencies of VDR BsmI and FokI polymorphisms were presented for participants with or without MS. Genotypes of BsmI and FokI are normally expressed as dominant homozygous genotype “BB, FF”, heterozygous genotype “Bb, Ff”, and recessive homozygous genotype “bb, ff”. There were differences in genotypes and gene frequencies of VDR BsmI between the MS cases and controls, with P-values of 0.011 and 0.03 respectively. Frequency of genotype BB and Bb was significantly different between individuals with MS and without MS, after comparing all of the three genotypes with each other (χ² = 7.022, P = 0.008). However, there was no difference in FokI polymorphism.

VDR polymorphisms and MS risk
The difference in the occurrence of the genotypes in BsmI between individuals with MS and the control group was significant (P trend = 0.02). The frequencies of genotype BB and combined Bb/bb in individuals with MS were 89% and 11% respectively, in the control group were 82% and 18% respectively, which suggested that the genotype BB was a risk factor (P = 0.006; OR = 1.771; 95% CI, 1.179-2.661). Compared with genotype BB, genotype Bb was likely to be a protective factor (P = 0.009; OR = 0.575; 95% CI, 0.381-0.869). The incidence of B and b alleles for the BsmI polymorphism in the two group were statistically significant (allele B vs. b; P = 0.005), which indicated allele b
could be a protective factor. However, it seemed no significant evidence to be risk factors or protective factors to FokI polymorphism in individuals with MS (Table 4).

Moreover, the genotypes of BsmI and FokI were combined and generated as 9 new genotypes like BBFF to analyze the cumulative effect of the polymorphisms, as shown in Table 4. But there were no significant differences in the risk of MS ($P_{\text{trend}} = 0.187$).

### Discussion

The results showed that VDR polymorphisms may influence the emergence of MS. The VDR BsmI (rs1544410 A > G) polymorphism appeared to associate with MS. Specifically, allele B and genotype of BB in BsmI were associated with MS, while allele b and genotype Bb seemed to play a protective role. The role of genotype bb was unclear, because of its limited frequency. When discussed VDR gene polymorphisms with the components of MS, we found the genotype BB carriers with MS presented a higher waist circumference. In addition, genotype FF for the VDR FokI (rs2228570 C > T) polymorphism was correlated with lower BMI in MS. Our findings were different with previous researches which demonstrated the VDR FokI polymorphism was related with triglycerides and HDL-cholesterol levels [25].

A few studies have demonstrated that VDR gene polymorphisms are related to obesity, diabetes, insulin sensitivity and insulin secretion [17,19,29]. VDR BsmI and FokI polymorphisms have been previously reported to be associated with anthropometric and biochemical parameters describing MS. Trzmiel et al. (2008) found that VDR BsmI polymorphism seemed to influence BMI, while the FokI VDR polymorphism appeared to affect insulin sensitivity and serum HDL cholesterol in men [24]. However, Lwow et al. (2008) indicated that VDR BsmI polymorphism did not seem to predispose postmenopausal women to obesity and insulin resistance, but the genotype BB was connected with dyslipidemia [30]. Frey et al. (2003) did not find evidence for the association of VDR polymorphisms with glycemia either [31]. Results of these study seemed to conflict with the pleiotropic effect of the VDR gene
in individuals with MS, and this result was not found in other populations [32]. Moreover, interactions in circulating glucose, triglyceride, cholesterol and insulin levels were not well explained [33,34]. For this reason, whether these differences come from ethnic variations or interactions in components of MS still needs further exploration, and we should not focus on the disorders in MS separately.

The VDR gene is not a main influencing factor to the variability of circulating levels of vitamin D, according to genome-wide association (GWA) studies [17,35,36]. However, it is clear that the VDR gene plays a primary role in the pleiotropic actions of 1,25(OH)2D3 [37] and in insulin secretion [19,38]. Serum 25-hydroxyvitamin D levels is inversely related to percentage body fat content in healthy women [20] and incident hypertension [21]. Furthermore, McKeown et al. (2011) found that the predicted 25-hydroxyvitamin D score might be an important determinant for change in fasting plasma glucose concentration in the Framingham offspring study [22]. Unfortunately, our study did not include serum 25-hydroxyvitamin D, but we could assume that serum 25-hydroxyvitamin D is the focus leading metabolic disorders if the interactions we discussed above were taken into consideration.

**Conclusion**

This study revealed a significant association of VDR BsmI polymorphism with MS in the Northern Chinese population. Allele B and genotype BB for BsmI are risk factors for MS. The BsmI polymorphism seemed to influence waist circumference, while the FokI polymorphism influence BMI in subjects with MS. Thus, we propose that the BsmI and FokI polymorphisms of VDR are potential prognostic variables which may predict the risk of developing MS. Even so, further examination should be carried out on large population. Also, genome wide association studies are still needed to evaluate the direct effect of these polymorphisms on MS.

**Abbreviations**

MS: Metabolic syndrome; VDR: Vitamin D Receptor gene; CVD: Cardiovascular disease; T2DM: Type 2 diabetes mellitus; IDF2005: International Diabetes Federation 2005; SNPs: Single-nucleotide polymorphisms; BMD: Bone mineral density; BMI: Body mass index; WC: Waist circumference; BPS: Systolic blood pressure; BPD: Diastolic blood pressure; FBG: Fasting blood-glucose; TG: Triglycerides; HWE: Hardy–Weinberg equilibrium; CI: Confidence interval; GWA: Genome-wide association.

**Competing interests**

The authors declare that they have no competing interests.

**Authors’ contributions**

Conception of research idea and Secure funding: YZ, YHZ; Study Design: YZ, YHZ, SL; Implementation (training; Data collection): SL, YZ, JH, YNJ, HLF, XYC, XMF, JJ, HXX, XYL; Study coordination and management: YZ, YHZ; Data management: YZ, ZYH; Data analysis and drafting paper: SL, YZ. All authors read and approved the final manuscript.

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Table 5 FokI and BsmI polymorphisms and MS components

|                | BsmI (rs1544410 A > G) | P       | FokI (rs 2228570 C > T) | P       |
|----------------|------------------------|---------|------------------------|---------|
|                | BB                     | Bb + bb | FF                     | Ff + ff |
| **MS**         |                        |         |                        |         |
| BMI            | 26.1 ± 3.1             | 25.7 ± 2.2 | 25.1 ± 2.5*            | 26.3 ± 3.1 | 26.4 ± 2.9 | 0.005|
| WC(cm)         | 90 ± 6.6               | 87.6 ± 6.3 | 88.7 ± 5.5            | 89.7 ± 7 | 90 ± 6.6 | 0.284|
| BPS(mmHg)      | 136.4 ± 19.8           | 141.9 ± 22.7 | 135.9 ± 20            | 137.5 ± 20.6 | 136.6 ± 20.4 | 0.829|
| BPD(mmHg)      | 84.1 ± 11.4            | 85 ± 12.5 | 84 ± 11.8            | 84.8 ± 11.3 | 83.2 ± 11.8 | 0.438|
| FBG(mmol/L)    | 6.1 ± 1.2              | 6.4 ± 2.2 | 6.2 ± 1.6            | 6.1 ± 1 | 6.2 ± 1.5 | 0.496|
| TG(mmol/L)     | 2.1 ± 1.2              | 2 ± 1.2  | 2.1 ± 0.9            | 2.1 ± 1.3 | 2 ± 1.2 | 0.821|
| HDL-C(mmol/L)  | 1.3 ± 0.4              | 1.4 ± 0.3 | 1.2 ± 0.3            | 1.3 ± 0.3 | 1.3 ± 0.5 | 0.077|
| **Control**    |                        |         |                        |         |
| BMI            | 22.7 ± 2.8             | 22 ± 2.4 | 22.5 ± 2.9            | 22.5 ± 2.6 | 22.5 ± 2.8 | 0.995|
| WC(cm)         | 79.7 ± 7.6             | 76.3 ± 6.1 | 78.7 ± 8            | 79.2 ± 6.5 | 79.2 ± 8.7 | 0.849|
| BPS(mmHg)      | 124.9 ± 18.3           | 124.9 ± 18.3 | 124.9 ± 18.6       | 126.8 ± 19.3* | 121.5 ± 15.6 | 0.045|
| BPD(mmHg)      | 77.2 ± 11.1            | 77.1 ± 10.8 | 78.2 ± 10.5        | 78.1 ± 11.8 | 74.9 ± 9.4* | 0.031|
| FBG(mmol/L)    | 5.6 ± 0.9              | 5.6 ± 0.5 | 5.5 ± 0.8            | 5.7 ± 1 | 5.5 ± 0.5 | 0.118|
| TG(mmol/L)     | 1.3 ± 1.2              | 1.2 ± 0.6 | 1.3 ± 0.6            | 1.3 ± 1.5 | 1.2 ± 0.6 | 0.789|
| HDL-C(mmol/L)  | 1.5 ± 0.3              | 1.5 ± 0.3 | 1.4 ± 0.3            | 1.5 ± 0.3 | 1.5 ± 0.3 | 0.596|

*significant for the genotype Ff/ff; **significant for the genotype ff.
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