Optogenetic dissection of basolateral amygdala contributions to intertemporal choice in young and aged rats

Caesar M Hernandez¹, Caitlin A Orsini², Chase C Labiste¹, Alexa-Rae Wheeler¹, Tyler W Ten Eyck¹, Matthew M Bruner¹, Todd J Sahagian³, Scott W Harden³, Charles J Frazier³, Barry Setlow², Jennifer L Bizon¹*

¹Department of Neuroscience, University of Florida, Gainesville, United States; ²Department of Psychiatry, University of Florida, Gainesville, United States; ³Department of Pharmacodynamics, University of Florida, Gainesville, United States

Abstract Across species, aging is associated with an increased ability to choose delayed over immediate gratification. These experiments used young and aged rats to test the role of the basolateral amygdala (BLA) in intertemporal decision making. An optogenetic approach was used to inactivate the BLA in young and aged rats at discrete time points during choices between levers that yielded a small, immediate vs. a large, delayed food reward. BLA inactivation just prior to decisions attenuated impulsive choice in both young and aged rats. In contrast, inactivation during receipt of the small, immediate reward increased impulsive choice in young rats but had no effect in aged rats. BLA inactivation during the delay or intertrial interval had no effect at either age. These data demonstrate that the BLA plays multiple, temporally distinct roles during intertemporal choice, and show that the contribution of BLA to choice behavior changes across the lifespan.

Introduction Intertemporal choice refers to decisions between rewards that differ with respect to both their magnitude and how far in the future they will arrive. Biases in intertemporal choice, whether manifesting as extreme impulsivity or patience, strongly associate with psychiatric disease. For example, enhanced preference for smaller, immediate rewards (greater impulsive choice) is a hallmark of attention deficit hyperactivity disorder and substance use disorders (Bickel et al., 2014; Hamilton et al., 2015; Patros et al., 2016), whereas pronounced preference for delayed gratification is characteristic of the eating disorder anorexia nervosa (Steinglass et al., 2012; Kaye et al., 2013; Decker et al., 2015). Independent of psychopathology, intertemporal choice both associates with life outcomes and changes across the lifespan (Denburg et al., 2007; Boyle et al., 2012; Beas et al., 2015; Hess et al., 2015). Contrary to economic models predicting that older individuals should account for reduced time on the horizon in making intertemporal choices, healthy older adults actually exhibit a marked increase in preference for delayed outcomes (Green et al., 1996; Green et al., 1999; Jimura et al., 2011; Löckenhoff et al., 2011; Mata et al., 2011; Samanez-Larkin et al., 2011; Eppinger et al., 2012). Although this pattern of choice behavior is sometimes characterized as ‘wisdom’, increased preference for delayed over immediate rewards may also be maladaptive. For example, biases toward delayed gratification in older adults could contribute to inappropriately conservative financial strategies that forgo expenditures necessary to maintain quality of life.

The neural circuits underlying age-associated changes in intertemporal choice remain poorly understood. Relevant to elucidating this circuity is the fact that intertemporal choice is a...
multicomponent process that involves a series of temporally distinct cognitive operations (Rangel et al., 2008; Fobbs and Mizumori, 2017). Specifically, most decisions begin with representations of past choices, as well as some idea of the outcomes associated with each choice option. These representations are weighted by one’s motivation to obtain the choice outcomes at the time of the decision. A second phase of decision making occurs after a choice is made and involves evaluating the outcome to determine the degree to which it matches its predicted value. Feedback from this evaluative process can be used to adjust representations of the options to guide future choices.

Both deliberation before a choice and outcome evaluation after a choice are supported by a network of brain structures that mediate reward processing, prospection, planning, prediction error, and value computations (Peters and Büchel, 2011; Orsini et al., 2015a; Bailey et al., 2016; Fobbs and Mizumori, 2017). The basolateral amygdala (BLA), which forms associations between cues or actions and their outcomes (Johansen et al., 2012; Wassum and Izquierdo, 2015), plays a central role in decision making and has been specifically implicated in both deliberative and evaluative processing (Schoenbaum et al., 1998; Schoenbaum et al., 1999; Ghods-Sharifi et al., 2009; Peters and Büchel, 2011; Zuo et al., 2012; Grabenhorst et al., 2012; Zangemeister et al., 2016; Orsini et al., 2017). The BLA also undergoes structural and functional alterations with advanced age, and BLA neural activity during intertemporal decision making is attenuated in aged rats (Lolova and Davidoff, 1991; Rubinow and Juraska, 2009; Rubinow et al., 2009; Roesch et al., 2012; Burke et al., 2014; Prager et al., 2016; Samson et al., 2017). It remains unclear, however, how age-associated changes in BLA recruitment actually influence intertemporal choice.

Hernandez et al. eLife 2019;8:e46174. DOI: https://doi.org/10.7554/eLife.46174
Optogenetic tools have been employed previously to define temporally-specific roles of BLA during deliberation and outcome evaluation in young rats performing a decision-making task involving risk of punishment (Orsini et al., 2017). Specifically, BLA inactivation at discrete timepoints in the decision process shifted choice behavior in opposite directions, highlighting multiple roles for BLA information processing in risky decision making. The present study used a similar optogenetic approach to define the roles of BLA neural activity in intertemporal choice (Figure 1) and to further determine if the roles of BLA change across the lifespan.

Results

Electrophysiological confirmation of light-induced inhibition of BLA neurons expressing eNpHR3.0.

Virally-transduced neurons were identified by mCherry expression and targeted for whole-cell patch clamp recordings using a combination of epifluorescence and differential interference contrast microscopy. Virally-transduced BLA neurons examined in slices from young and aged animals did not differ with respect to input resistance (Young: 122.6 ± 20.7 MΩ, n = 26 cells; Aged: 120.2 ± 11.6 MΩ, n = 28 cells; t_{52}=0.101, p=0.920), whole cell capacitance (Young: 139.7 ± 7.54 pF, n = 26 cells; Aged: 138.8 ± 6.91 pF, n = 28 cells; t_{52}=0.081, p=0.936), or current required to maintain the membrane potential at −70 mV (Young: −104.34 ± 15.9 pA; n = 26 cells; Aged: −116.0 ± 13.3 pA, n = 28 cells; t_{52}=0.566, p=0.574). Young and aged neurons filled with biocytin and visualized with 2-photon mediated epifluorescence microscopy were multipolar and had substantial dendritic branching, consistent with the morphology of BLA principal neurons (Figure 2A,E).

Light pulses (1 s duration) produced similar outward currents in young and aged virally-transduced BLA neurons, as observed in voltage clamp (young: 78.8 ± 10.6 pA, n = 26 cells, aged: 89.1 ± 8.4 pA, n = 28 cells, t_{52}=0.764, p=0.448, Figure 2B,F). Activation of eNpHR3.0 in this manner was consistently sufficient to silence both young and aged neurons when firing under a moderate (~50–200 pA) load (Figure 2C,G). Additional experiments were conducted to confirm that age differences did not emerge with longer light pulses (4 s) that matched or exceeded durations used in the in vivo experiments. A 4 s light-induced activation of eNpHR3.0 produced similar outward currents in young and aged cells (young: 55.2 ± 11.9 pA, n = 16 cells; aged: 55.9 ± 6.8 pA, n = 16 cells, t_{30}=0.050, p=0.961, Figure 2D,H). Finally, because long term activation of eNpHR3.0 plausibly could alter chloride gradients across the cell membrane, evidence for rebound excitation after 4 s light pulses was evaluated in both young and aged neurons. Overall, voltage clamp experiments measured in the first 500 msec after the light pulse revealed similar mean currents in young and aged neurons (young: 0.6 ± 1.6 pA, n = 16 cells, aged: 4.1 ± 2.1 pA, n = 16 cells, t_{30}=1.351, p=0.187). Similarly, only 1 of 61 neurons examined in current clamp (across both ages and light durations) that were silent before exposure to light fired any action potentials within 1 s of cessation of the light pulse. Overall, these data indicate that light-induced activation of eNpHR3.0 produces robust inhibition of virally-transduced BLA principal neurons, and that these effects do not significantly vary with age. The data further demonstrate that rebound excitation after eNpHR3.0 activation in BLA is minimal and unlikely to be functionally impactful.

Fiber placement and AAV transduction

Expression of mCherry was used to confirm viral transduction in the BLA of rats used in behavioral studies that were injected with either AAV5-CamKIIα-eNpHR3.0-mCherry (AAV-eNpHR3.0, black circles in Figure 3) or AAV-CamKIIα-mCherry alone (AAV-control, white circles in Figure 3). Cannula placements were centered in the BLA, and the brain volumes virally transduced by AAV-eNpHR3.0 and AAV-control (calculated from the atlas of Paxinos and Watson, 2005) were comparable in young and aged rats.

Effect of age on intertemporal choice performance

Previous work shows that aged rats display attenuated discounting of delayed rewards (Simon et al., 2010; Hernandez et al., 2017). Therefore, prior to inactivation sessions, delays were adjusted on an individual basis to ensure that all rats’ choice performance was within the same parametric space (Figure 4A). This approach allowed a comparable range of effects from BLA.
inactivation to be observed in both young and aged rats, without concern for ceiling or floor effects. Figure 4B shows the actual delays used in the second and third blocks to achieve roughly 66% and 33% choice of the large reward, respectively, plotted as a function of age. A two-factor ANOVA (age × delay block) comparing the actual delays indicated the expected main effect of block ($F(2,26) = 18.685$, $p < 0.001$, $\eta^2_p = 0.606$, $1-\beta = 0.930$), as well as a main effect of age ($F(1,13) = 6.402$, $p = 0.025$, $\eta^2_p = 0.330$, $1-\beta = 0.648$) and an age × delay block interaction ($F(2,26) = 6.913$, $p = 0.004$, $\eta^2_p = 0.347$, $1-\beta = 0.891$). Post hoc analyses comparing the actual delays of young and aged rats in blocks 2 and 3 indicated that aged rats required longer delays than young to achieve comparable preference for large vs. small rewards (Block 2: $t(13) = -2.234$, $p = 0.044$, Cohen’s $d = 1.114$, $1-\beta = 0.480$; Block 3: $t(13) = -2.660$, $p = 0.020$, Cohen’s $d = 1.328$, $1-\beta = 0.625$). Consistent with this analysis, aged rats in comparison to young rats had a greater indifference point (the delay at which rats showed equivalent preference for large and small rewards; $t(13) = 2.168$, $p = 0.049$, Cohen’s $d = 1.080$, $1-\beta = 0.457$; Figure 4C).

Effects on choice behavior of BLA inactivation during deliberation

Inactivation of the BLA during the deliberation epoch ($n = 8$ young and $n = 7$ aged) significantly increased choice of the large reward to the same extent in young and aged rats, particularly at long delays (Figure 5A). A three-factor ANOVA (laser condition × age × delay block) indicated a main effect of laser condition ($F(1,13) = 103.507$, $p < 0.001$, $\eta^2_p = 0.888$, $1-\beta = 1.000$) but no main effect of age ($F(1,13) = 0.089$, $p = 0.770$) nor an age × laser condition interaction ($F(1,13) = 1.838$, $p = 0.198$). A reliable main effect of delay block was observed ($F(2,26) = 112.005$, $p < 0.001$, $\eta^2_p = 0.896$, $1-\beta = 1.000$), as was an interaction between laser condition and delay block ($F(2,26) = 38.369$, $p < 0.001$, $\eta^2_p = 0.747$, $1-\beta = 1.000$). Follow-up analyses, conducted to further explore the laser condition × delay block interaction, compared the effects of inactivation at each block. This analysis indicated that BLA inactivation significantly increased choice of the large reward in blocks 2 ($t(14) = -6.494$, $p < 0.001$, Cohen’s $d = 1.724$, $1-\beta = 0.995$) and 3 ($t(14) = -9.434$, $p < 0.001$, Cohen’s $d = 2.228$, $1-\beta = 1.000$), but not in block 1 in which rats of both ages strongly preferred the large reward, even under control conditions ($t(14) = -0.323$, $p = 0.751$).
Effects on choice behavior of BLA inactivation during the small reward

In direct contrast to the effects of BLA inactivation during deliberation, BLA inactivation during the small reward epoch (n = 6 young and n = 6 aged) significantly decreased choice of the large reward only in young rats (Figure 5C). A three-factor ANOVA (laser condition × age × delay block) indicated main effects of laser condition (F(1,10) = 5.131, p = 0.047, η² = 0.339, 1-β = 0.534) and delay block (F(2,20) = 248.854, p < 0.001, η² = 0.961, 1-β = 1.000), but no interaction between laser condition and delay block (F(2,20) = 1.317, p = 0.290). Notably, although there was no main effect of age (F(1,10) = 0.941, p = 0.355), the effects of BLA inactivation during small reward delivery did reliably interact with age (laser condition × age: F(1,10) = 7.127, p = 0.024, η² = 0.416, 1-β = 0.673). To better define the nature of this interaction, follow-up analyses using two-factor ANOVAs (laser condition × delay block) were performed on choice behavior separately in young and aged rats. BLA inactivation significantly decreased choice of the large reward in young rats (main effect of laser condition: F(1,5) = 18.226, p = 0.008, η² = 0.785, 1-β = 0.922, main effect of delay block: F(2,10) = 173.588, p < 0.001, η² = 0.972, 1-β = 1.000; laser condition × delay block: F(2,10) = 3.829, p = 0.058) but not in aged rats (main effect of laser condition: F(1,5) = 0.061, p = 0.814; main effect of delay block: F(2,10) = 93.015, p < 0.001, η² = 0.949, 1-β = 1.000; laser condition × delay block: F(2,10) = 0.185, p = 0.834).

Because different delays to large reward delivery were required to achieve comparable levels of choice preference in young and aged rats, it is possible that the absence of BLA inactivation effects...
in aged rats was due to the different delay durations employed rather than age differences per se. To address this possibility, an additional analysis was conducted in which aged rats were sub-grouped into those with delays comparable to young ('aged delay-matched') and those with delays that exceeded young ('aged delay-unmatched'). A multi-factor ANOVA was conducted with age subgroup (two levels: aged delay-matched and aged delay-unmatched) as the between-subjects factor and laser condition (two levels: laser on or off) and block (three levels: blocks 1, 2, and 3) as within-subjects factors. Critically, this analysis revealed no choice difference between aged delay-matched and aged delay-unmatched subgroups when the BLA was inactivated during the small reward delivery (main effect of sub-group: $F_{(1,4)}=0.180, p=0.694$; main effect of laser condition: $F_{(1,4)}=0.050, p=0.834$; main effect of block: $F_{(2,8)}=80.518, p<0.001, \eta^2=0.953, 1-\beta=1.000$; laser condition $\times$ sub-group: $F_{(1,4)}=0.082, p=0.789$; laser condition $\times$ block: $F_{(2,8)}=0.167, p=0.849$; sub-group $\times$ block: $F_{(2,8)}=0.328, p=0.729$; laser condition $\times$ sub group $\times$ block: $F_{(2,8)}=0.515, p=0.616$). These results indicate that it is unlikely that the different delay durations contributed to the age difference in the role of the BLA during small reward delivery.

**Altered choice strategy resulting from BLA inactivation during the deliberation and small reward epochs**

The data above show that BLA inactivation during the deliberation and small reward epochs altered choice behavior in different directions (i.e., BLA inactivation during deliberation increased choice of the large reward in both young and aged rats, whereas BLA inactivation during small reward delivery decreased choice of the large, delayed reward in young rats but had no effect in aged rats). A trial-by-trial analysis was conducted on these data to determine the effects of BLA inactivation on two distinct behavioral strategies that could mediate these shifts in choice preference. Specifically,
during the deliberation epoch, this analysis determined the degree to which BLA inactivation influenced rats to ‘shift’ to the large reward option following a choice of the small reward on the previous trial, versus ‘stay’ with the large reward option following choice of the large reward on the previous trial. In the small reward epoch, the analysis assessed the degree to which BLA inactivation influenced rats to ‘shift’ to the small reward following choice of the large reward on the previous trial, versus ‘stay’ with the small reward following choice of the small reward on the previous trial.

As shown in Figure 5B, the percentage of trials during deliberation epoch inactivation on which a large reward choice was followed by a second large reward choice (large-stay) did not differ as a function of laser condition or age (main effect of laser condition: $F_{(1,13)}=2.563$, $p=0.605$; main effect of age: $F_{(1,13)}=0.282$, $p=0.605$; laser condition $\times$ age: $F_{(1,13)}=0.153$, $p=0.702$). In contrast, a similar analysis conducted on the percentage of trials on which a choice of the small reward was followed by choice of the large reward (small-shift) revealed a main effect of laser condition but no effect of age (main effect of laser condition: $F_{(1,13)}=40.051$, $p<0.001$, $\eta^2_p=0.755$, 1-$\beta=1.000$; main effect of age: $F_{(1,13)}=0.425$, $p=0.526$; laser condition $\times$ age interaction: $F_{(1,13)}=0.003$, $p=0.954$). Planned paired-samples t-tests showed that a significant increase in shifting after a choice of the small reward was evident in both young ($t_{(7)}=4.095$, $p=0.005$, Cohen’s $d=1.802$, 1-$\beta=0.917$) and aged ($t_{(6)}=5.342$, $p=0.002$, Cohen’s $d=2.442$, 1-$\beta=0.987$) rats. This finding indicates that the effects on choice behavior of BLA inactivation during deliberation result from rats shifting choices toward the large reward following a choice of the small reward.

Applying a parallel analysis to sessions in which inactivation took place during the small reward epoch yielded a different pattern of results (Figure 5D). BLA inactivation during the small reward epoch significantly increased the percentage of trials on which a small reward choice was followed by a second small reward choice (small-stay; main effect of laser condition: $F_{(1,10)}=6.026$, $p=0.034$, $\eta^2_p=0.376$, 1-$\beta=0.601$; main effect of age: $F_{(1,10)}=2.421$, $p=0.151$; laser condition $\times$ age interaction: $F_{(1,10)}=3.519$, $p=0.090$). Planned paired-samples t-tests showed that young rats were more likely to repeat the small reward choice on subsequent trials (small-stay; $t_{(5)}=3.593$, $p=0.016$, Cohen’s $d=1.694$, 1-$\beta=0.754$) but this pattern was not observed in aged rats ($t_{(5)}=0.363$, $p=0.732$). In contrast, neither BLA inactivation nor age influenced the percentage of trials on which a choice of the large reward was followed by a choice of the small reward (large-stay; main effect of laser condition: $F_{(1,10)}=1.120$, $p=0.315$; main effect of age: $F_{(1,10)}=0.105$, $p=0.753$; laser condition $\times$ age: $F_{(1,10)}=0.086$, $p=0.775$).

### Effects of BLA inactivation during the deliberation and small reward epochs on other task performance measures

Other task measures were compared between BLA inactivation and baseline conditions in both deliberation and small reward epochs using a mixed-factor ANOVA, with age as the between-subjects factor and laser condition as the within-subjects factor. As shown in Table 1, the number of trials completed in a session did not differ as a function of laser condition or age in either the deliberation or small reward outcome epochs ($F_s<3.431$, $p>0.094$). Similarly, as shown in Table 2, latency to press either the small or large reward lever did not differ as a function of laser condition or age in either epoch ($F_s<4.149$, $p>0.069$). See Tables 1 and 2 for full statistical results of these analyses.

### Effects of BLA inactivation during epochs associated with the large reward

Choosing the large reward lever resulted in a variable delay period that was followed by large (three food pellets) reward delivery. The effects of BLA inactivation during the delay and large reward delivery epochs were initially tested in separate sessions ($n=6$ young and $n=6$ aged). Subsequently, the effects of BLA inactivation across both the delay and large reward epochs were tested in a subset of these rats ($n=3$ young and $n=3$ aged).

### Effects of BLA inactivation during the delay epoch

The effects of BLA inactivation during the delay epoch were tested in delay blocks 2 and 3 using a three-factor ANOVA (laser condition $\times$ age $\times$ delay block). As expected, there was a main effect of delay block ($F_{(2,20)}=146.811$, $p<0.001$, $\eta^2_p=0.936$, 1-$\beta=1.000$) such that both young and aged rats
decreased their choice of the large reward as the delay prior to the large reward increased (Figure 6A). Compared to baseline, however, no reliable differences in choice behavior resulted from BLA inactivation during the delay epoch ($F_{1,10} = 0.005, p=0.947$), nor was there an interaction between laser condition and delay block ($F_{2,20} = 0.002, p=0.998$). Similarly, there were neither main effects nor interactions associated with age (main effect of age: $F_{1,10} < 0.001, p=0.996$; age × delay).

### Table 1. Effects of BLA inactivation on number of trials completed per session.

| Epoch         | Age     | Laser condition | Mean   | SEM   | Statistical comparisons |
|---------------|---------|-----------------|--------|-------|-------------------------|
| Deliberation  | Young   | Off (Baseline)  | 52.688 | 0.81  | Laser condition: $F_{1,13} = 0.180, p=0.678$ |
|               |         | On (Inactivation) | 51.625 | 0.94  |
|               | Aged    | Off (Baseline)  | 51.714 | 0.87  |
|               |         | On (Inactivation) | 53.429 | 1.003 |
| Outcome       | Young   | Off (Baseline)  | 52.667 | 0.394 |
| (small reward)|         | On (Inactivation) | 53.667 | 0.635 |
|               | Aged    | Off (Baseline)  | 52.639 | 0.394 |
|               |         | On (Inactivation) | 53.167 | 0.635 |

Raw data for this table are provided in Table 1—source data 1. DOI: https://doi.org/10.7554/eLife.46174.011

The following source data is available for Table 1:
Source data 1. Hernandez et al. Source data 1. DOI: https://doi.org/10.7554/eLife.46174.012

### Table 2. Effects of BLA inactivation on lever response latencies.

| Epoch         | Age     | Lever | Laser condition | Mean (sec) | Std. error | Statistical analysis |
|---------------|---------|-------|-----------------|------------|-----------|----------------------|
| Deliberation  | Young   | Large | Off (Baseline)  | 1.385      | 0.174     | Laser condition:
|               |         |       | On (Inactivation) | 1.442     | 0.204     |
|               | Aged    | Large | Off (Baseline)  | 1.083      | 0.187     |
|               |         |       | On (Inactivation) | 1.543     | 0.326     |
|               |       | Small | Off (Baseline)  | 0.956      | 0.186     |
|               |         |       | On (Inactivation) | 1.108     | 0.218     |
|               | Aged    | Small | Off (Baseline)  | 1.043      | 0.187     |
|               |         |       | On (Inactivation) | 1.120     | 0.326     |
| Outcome       | Young   | Large | Off (Baseline)  | 1.322      | 0.136     |
| (small reward)|         |       | On (Inactivation) | 1.217     | 0.141     |
|               | Aged    | Large | Off (Baseline)  | 1.107      | 0.149     |
|               |         |       | On (Inactivation) | 0.938     | 0.054     |
|               |       | Small | Off (Baseline)  | 0.912      | 0.136     |
|               |         |       | On (Inactivation) | 0.870     | 0.141     |
|               | Aged    | Small | Off (Baseline)  | 0.962      | 0.149     |
|               |         |       | On (Inactivation) | 0.804     | 0.054     |

Raw data for this table are provided in Table 2—source data 1. DOI: https://doi.org/10.7554/eLife.46174.013

The following source data is available for Table 2:
Source data 1. Hernandez et al. Source data 1. DOI: https://doi.org/10.7554/eLife.46174.014
Effects of BLA inactivation during both delay and large reward epochs

One potential explanation for the null effects of BLA inactivation during the delay and large reward epochs is that, given the role of the BLA in integration of rewards and costs, inactivation may only be effective when conducted during both of these epochs. To evaluate this possibility, rats were tested while the BLA was inactivated during both the delay and large reward epochs; however, continuous inactivation across both epochs yielded no effects on choice performance. As shown in Figure 6C, a three-factor ANOVA (laser condition × age × delay block) revealed the expected main effect of delay block (F(2,20) = 193.743, p < 0.001, η² = 0.924, 1 - β = 1.000) but no main effects or interactions involving laser condition or age (main effect of laser condition: F(1,4) = 0.757, p = 0.433; main effect of age: F(1,10) = 0.125, p = 0.731; laser condition × age × delay block: F(2,20) = 0.437, p = 0.652; laser condition × age: F(1,10) = 0.697, p = 0.423; laser condition × age × delay block: F(2,20) = 0.664, p = 0.526).

Effects of BLA inactivation during both delay and large reward epochs

Unlike the effects of BLA inactivation during the small reward epoch, inactivation of BLA during the large reward epoch did not alter choice performance in either young or aged rats compared to baseline (Figure 6B). As expected, there was a main effect of delay block (F(2,20) = 120.846, p < 0.001, η² = 0.924, 1 - β = 1.000) such that both young and aged rats decreased their choice of the large reward as the delay to large reward delivery increased. Compared to baseline, however, no reliable differences in choice behavior resulted from BLA inactivation during the large reward epoch (F(1,10) = 0.125, p = 0.731), nor was there an interaction between laser condition and delay block (F(2,20) = 0.133, p = 0.876). Similarly, there were neither main effects (F(1,10) = 0.249, p = 0.629) nor interactions associated with age (age × delay block: F(2,20) = 0.437, p = 0.652; laser condition × age: F(1,10) = 0.697, p = 0.423; laser condition × age × delay block: F(2,20) = 0.664, p = 0.526).

Effects of BLA inactivation during the large reward epoch

Unlike the effects of BLA inactivation during the small reward epoch, inactivation of BLA during the large reward epoch did not alter choice performance in either young or aged rats compared to baseline (Figure 6B). As expected, there was a main effect of delay block (F(2,20) = 120.846, p < 0.001, η² = 0.924, 1 - β = 1.000) such that both young and aged rats decreased their choice of the large reward as the delay to large reward delivery increased. Compared to baseline, however, no reliable differences in choice behavior resulted from BLA inactivation during the large reward epoch (F(1,10) = 0.125, p = 0.731), nor was there an interaction between laser condition and delay block (F(2,20) = 0.133, p = 0.876). Similarly, there were neither main effects (F(1,10) = 0.249, p = 0.629) nor interactions associated with age (age × delay block: F(2,20) = 0.437, p = 0.652; laser condition × age: F(1,10) = 0.697, p = 0.423; laser condition × age × delay block: F(2,20) = 0.664, p = 0.526).
Figure 5. Effect of BLA inactivation during the deliberation and small reward epochs. (A) Inactivation of the BLA during the deliberation epoch (prior to a choice) resulted in a significant increase in preference for the large, delayed reward in both young (n = 8) and aged (n = 7) rats. (B) Effects of BLA inactivation during the deliberation epoch on trial-by-trial choice strategies. This analysis revealed that the increased choice of the large, delayed reward caused by BLA inactivation during deliberation (panel A) was due to an increase in the percentage of trials on which rats shifted to the large, delayed reward following a choice of the small, immediate reward. (C) Inactivation of the BLA during the small reward epoch resulted in a significant decrease in preference for the large, delayed reward in young (n = 6), but not aged (n = 6), rats. (D) Effects of BLA inactivation during the small reward epoch on trial-by-trial choice strategies. This analysis revealed that the decreased choice of the large, delayed reward in young rats caused by BLA inactivation during the small, reward epoch (panel C) was due to an increase in the percentage of trials on which rats 'stayed' on the small, immediate reward following a choice of this reward on the previous trial. In contrast, BLA inactivation during the same epoch in aged rats had no effect on trial-by-trial choice strategies. In all panels, error bars represent standard error of the mean (SEM). *p<0.05, **p<0.01, ***p<0.001, main effect of inactivation; ***p<0.001, inactivation × delay block interaction. Raw data for these graphs are provided in Figure 5—source data 1.

DOI: https://doi.org/10.7554/eLife.46174.009

The following source data is available for figure 5:

Source data 1. Hernandez et al. Figure 5—source data 1.
DOI: https://doi.org/10.7554/eLife.46174.010

effect of age: F(1,4)=0.306, p=0.610; laser condition × delay block: F(2,8)=0.979, p=0.417; laser condition × age: F(2,8)=0.053, p=0.949; laser condition × age × delay block: F(2,8)=0.159, p=0.856.

Effects of BLA inactivation during the intertrial interval
To confirm the temporal specificity of the BLA inactivation effects, rats (n = 6 young, n = 6 aged) were tested while the BLA was inactivated during the intertrial interval (ITI). Although the expected main effect of delay block was observed (F(2,20)=116.459, p<0.001, ηp²=0.921, 1-β=1.000), BLA inactivation during the ITI did not alter choice performance compared to baseline in young or aged rats (main effect of laser condition: F(1,10)=0.082, p=0.780; main effect of age: F(1,10)=0.042, p=0.842;
Figure 6. Effect of BLA inactivation during outcomes associated with choice of the large reward. (A) Inactivation of the BLA during the delay epoch resulted in no change in choice performance in either young (n = 6) or aged (n = 6) rats. (B) Inactivation of the BLA during the large reward epoch resulted in no change in choice performance.
Effects of light delivery into BLA in rats with control virus (AAV5-CamKIIα-mCherry)

To control for non-specific effects of light delivery and viral transduction (e.g., changes in tissue temperature and off-target transduction effects), the effects of light delivery in rats virally transduced with a control virus that did not contain the eNpHR3.0 gene were tested during behavioral epochs in which BLA inactivation influenced choice behavior (i.e., deliberation: n = 4 young and n = 4 aged rats; and small reward: n = 4 young rats).

Effects of light delivery during the deliberation epoch

Light delivery during the deliberation epoch in rats virally transduced with a control virus had no effects on choice performance (Figure 8A). A three factor ANOVA (laser condition × age × delay block) indicated the expected main effect of delay block ($F_{(2,12)}=100.272, p<0.001, \eta^2_p=0.944, 1-\beta =1.000$) but no main effects or interactions involving laser condition or age (main effect of laser condition: $F_{(1,6)}=0.128, p=0.733$; main effect of age: $F_{(1,6)}=0.055, p=0.823$; laser condition × age: $F_{(1,6)}=0.028, p=0.874$; laser condition × delay block: $F_{(2,12)}=0.121, p=0.887$; age × delay block: $F_{(2,12)}=0.105, p=0.902$; laser condition × age × delay block: $F_{(2,12)}=0.434, p=0.658$).

Effects of light delivery during the small reward epoch

Light delivery during the small reward epoch in young rats transduced with control virus also failed to influence choice performance (Figure 8B). A two factor ANOVA (laser condition × delay block) indicated the expected main effect of delay block ($F_{(2,6)}=46.712, p<0.001, \eta^2_p=0.940, 1-\beta =1.000$) but no main effect of laser condition ($F_{(1,3)}=0.359, p=0.592$) or laser condition × delay block interaction ($F_{(2,6)}=0.173, p=0.845$).

Discussion

Previous work employing permanent lesions or pharmacological approaches to manipulate BLA activity (Winstanley et al., 2004; Churchwell et al., 2009) has clearly demonstrated a critical role for this structure in intertemporal decision making. Specifically, BLA inactivation using such techniques results in more impulsive choices that favor immediate over delayed rewards. Lesion and pharmacological inactivation approaches, however, do not differentiate between temporally-discrete stages of decision making (e.g., deliberation, delay, outcome evaluation), which have been shown in other contexts to involve distinct neural circuitry (Peters and Büchel, 2011; Fobbs and Mizumori, 2017). The current study employed optogenetic tools in order to parse temporally-discrete contributions of BLA to intertemporal choice in both young and aged rats.

Role of BLA in outcome evaluation in young rats

A large literature supports a role for BLA in assigning and updating the value of stimuli and events (Hatfield et al., 1996; Málková et al., 1997; Baxter et al., 2000; Baxter and Murray, 2002; Shiflett and Balleine, 2010; Izquierdo et al., 2013; Parkes and Balleine, 2013; Wassum and Izquierdo, 2015). With respect to decision making, the evaluative process mediated by BLA after an
outcome has been received appears to involve acquisition and/or integration of information about the negative properties of that outcome. For example, previous work from our laboratory using a risky decision-making task demonstrated that BLA inactivation during receipt of a large, punished reward increased subsequent choice of this option over a small but safe reward (Orsini et al., 2017). Given this previous finding, it was somewhat surprising that in the current study, BLA inactivation during the large reward following the delay and/or the delay interval itself had no effect on choice behavior. These data demonstrate that the aversive properties of delays that bias choice behavior toward immediate options are not critically mediated by BLA. Importantly, however, BLA inactivation during the small reward epoch did reliably bias young rats toward choices of the small, immediate reward. This bias, which mimics that produced by BLA lesions (Winstanley et al., 2004; Churchwell et al., 2009), indicates that BLA is specifically important for evaluating and integrating the aversive properties that make the small reward less attractive than the large. Indeed, the trial-by-trial analysis showed that BLA inactivation rendered rats more likely to ‘stay’ with choices of the small reward after selection of that option on the previous trial, as though the negative feedback about that small reward ‘not being good enough’ had been attenuated. While these data are consistent with the idea that the BLA processes information about the aversive properties of outcomes in order to bias future behavior toward more favorable options (Ghods-Sharifi et al., 2009; Orsini et al., 2017), they also suggest that final integration of the values of both reward magnitude and delay occurs outside of the BLA. Given that working memory appears to contribute to the ability to delay gratification during intertemporal choice (Shamosh et al., 2008; Bobova et al., 2009; Shimp et al., 2015; Hernandez et al., 2017), it is likely that brain regions such as the hippocampus and prefrontal cortex mediate at least some components of information processing during the delay period. As such, one might predict that temporally-selective inactivation of these structures while waiting for a large reward (i.e., the delay interval) would influence future choice of that delayed option (Churchwell et al., 2009; Mariano et al., 2009; Abela and Chudasama, 2013; Sonntag et al., 2014; Yates et al., 2014). It should also be noted that BLA might be more critically involved in integrating information about delays and reward magnitude under other intertemporal choice conditions, such as in the presence of a cue that predicts large reward delivery during the
Role of BLA during deliberation in young rats

In contrast to results obtained during outcome evaluation, BLA inactivation during deliberation in young rats increased choice of the large, delayed reward. This shift toward less impulsive choice is opposite of that produced by inactivation methods such as lesions, which inhibit the BLA throughout the entire decision process (Winstanley et al., 2004; Churchwell et al., 2009). The prior study from our lab investigating risky decision making also showed that BLA inactivation during the deliberation epoch produced effects on choice behavior that were opposite those produced by neurotoxic lesions (Orsini et al., 2015b; Orsini et al., 2017). Specifically, during choices between small, safe and large, risky rewards, BLA lesions increase risky choice whereas selective optogenetic BLA inactivation during deliberation attenuates risky choice. Together, these data indicate a critical role for BLA during deliberation, which is normally overshadowed by its role in outcome evaluation. Importantly, although the risky decision-making and intertemporal choice tasks are analogous in design, performance on the two tasks is orthogonal and involves dissociable neural mechanisms (Floresco et al., 2008; Churchwell et al., 2009; Simon et al., 2011; Mitchell et al., 2014; Orsini et al., 2015a; Orsini et al., 2018; Bailey et al., 2016). Thus, the similar pattern of results observed with BLA manipulations suggests a common role for this structure in multiple forms of decision making. Specifically, BLA activity during deliberation under normal conditions may be important for ascribing incentive salience to the choice options, signaling their motivational value, or how much they are ‘wanted’, in the moment of the decision. Consequently, choice may be biased toward the more immediate reward in the intertemporal choice task, and toward the larger, albeit riskier, reward in the risky choice task. When this signal is removed (e.g., during BLA inactivation), rats are more likely to wait in the intertemporal choice task and less likely to risk punishment in the risky choice task to obtain the large reward. Indeed, the trial-by-trial analysis in the current study shows that BLA inactivation during deliberation renders rats more likely to shift their choices to the delayed reward following a choice of the immediate reward, as though the incentive properties...
driving the immediate choice have been attenuated. This interpretation agrees with evidence from other behavioral contexts. For example, an intact BLA is necessary for the potentiating influence of reward-predictive cues on instrumental responding for reward (Everitt et al., 2003), as well as for maintaining effortful choices of preferred options (Hart and Izquierdo, 2017).

### Age differences in intertemporal choice

Across species, aging is accompanied by an increased ability to delay gratification (Green et al., 1994; Green et al., 1999; Simon et al., 2010; Jimura et al., 2011; Löckenhoff et al., 2011; Samañez-Larkin et al., 2011; Eppinger et al., 2012; Hernandez et al., 2017). Previous work from our labs showed that relative to young rats, aged rats display greater preference for large, delayed over small, immediate rewards in a ‘fixed delays, block design’ intertemporal choice task. This difference is not readily attributable to age-related deficits in cognitive flexibility, working memory, or food motivation, nor is it attributable to impairments in reward or temporal discrimination (Simon et al., 2010; Hernandez et al., 2017). The present study replicated these prior findings using a task variant in which the fixed delays/block design employed in our previous work was maintained, but the delays to large reward delivery were adjusted on an individual basis to obtain equivalent levels of choice preference in young and aged rats. Under these conditions, aged rats required longer delays to achieve levels of choice preference comparable to young, consistent with the idea that delays are less effective at discounting reward value in aged compared to young rats.

### Age differences in the role of BLA in intertemporal choice

Inactivation of BLA during the deliberation epoch decreased impulsive choice in both young and aged rats. In contrast, BLA inactivation during the small reward epoch increased impulsive choice in young rats but had no effect in aged rats. Importantly, this lack of effect in aged rats is unlikely to be attributable to age-related impairments in viral transduction or optogenetic efficacy. Histological reconstruction showed comparable BLA viral transduction in young and aged rats (Figure 3). Moreover, in vitro electrophysiological experiments showed that halorhodopsin-transduced BLA neurons in both age groups were silenced to a similar degree in response to light pulses (Figure 2). Most importantly, inactivation during the deliberation epoch in the same aged rats used to test the effects of inactivation during the small reward epoch produced effects on behavior that were as robust as those in young rats (Figure 5A). These latter data provide direct, in vivo verification that the absence of behavioral effects following BLA inactivation during the small reward epoch in aged rats cannot be ascribed to attenuated halorhodopsin efficacy. It is possible that aged rats’ bias toward the large, delayed reward could have resulted in insufficient parametric space in which to observe optogenetically-induced shifts in choice behavior. The intertemporal choice task was explicitly designed to address this possibility by adjusting the delays to equate baseline choice preference in young and aged rats (Figure 4A). Additional analyses were conducted to ensure that the age difference in the delay to large reward delivery that resulted from this design did not itself influence the role of BLA in intertemporal choice. Specifically, aged rats were divided into subgroups based on whether their delays to large reward delivery matched or exceeded the range of young. The effects of BLA inactivation were identical between these aged subgroups, and in particular, BLA inactivation during the small reward epoch had no effect in either subgroup. These data indicate that the different delay durations experienced by young and aged rats do not account for the age differences in the role of BLA in intertemporal choice.

The distinct effects of BLA inactivation in young and aged rats could suggest that aged subjects fundamentally make decisions differently than young, relying less heavily on evaluation of choice outcomes (Löckenhoff et al., 2011; Mather et al., 2012; Samañez-Larkin and Knutson, 2015; Pachur et al., 2017). For example, whereas young adults readily incorporate new information to guide decisions, older adults tend to rely more heavily on previously-learned, ‘crystallized’ knowledge for decision making (Horn, 1982; Mather, 2006; Mata et al., 2011). If the cognitive structure of the decision process differs, the absence of BLA inactivation effects during outcome evaluation in aged rats may not necessarily reflect BLA dysfunction. For example, the increased reliance of older adults on crystallized knowledge for decision making has been attributed in part to vulnerability in neural structures such as hippocampus, which compromises encoding of new information and prospective memory (Del Missier, 2015; Shadlen and Shohamy, 2016; Hu et al., 2017; Levin et al.,...
Moreover, there is substantial evidence in aged humans and rodents for recruitment of brain circuits that are distinct from those engaged by young subjects during complex cognitive operations, even when performance is equated (Antonenko and Flöel, 2014; Lighthall et al., 2014; Tomás Pereira et al., 2015; Wang et al., 2015). The cause-effect relationships in the influences of aging on neural versus cognitive ‘restructuring’ are difficult to disentangle. It is certainly possible, however, that the fact that aged rats don’t use the BLA for outcome evaluation during intertemporal decision making reflects neural dysfunction and/or compensation associated with other brain regions.

While the current findings do not necessarily reflect age-associated neurobiological impairments within the BLA, prior evidence does suggest that this region is susceptible to age-associated changes (Rubinow and Juraska, 2009; Rubinow et al., 2009; Roesch et al., 2012; Prager et al., 2016). Indeed, although BLA neuron number remains relatively stable with age, baseline firing rate of BLA neurons in vivo is reduced in aged rats (Almaguer et al., 2002; Roesch et al., 2012), and a recent electrophysiological recording study reported enhanced β-power in BLA of aged rats during reward evaluation in a probabilistic decision-making task (Samson et al., 2017). Projections from the BLA to the nucleus accumbens (NAc) may be particularly relevant for reward outcome evaluation. Pharmacological disconnection of the BLA and NAc impairs discrimination between a devalued vs. a non-devalued food reward (Shiflett and Balleine, 2010), and optogenetic inactivation of BLA terminals in NAc during outcome evaluation increases preference for the ‘risky’ option in a probabilistic decision-making task (Bercovici et al., 2018). These results support the idea that BLA projections to NAc are responsible for mediating negative feedback regarding choice outcomes. Notably, Eppinger et al. (2013) showed blunted activity in ventral striatum during reward prediction errors in older adults performing a learning task. Together with the current study, these findings suggest that the BLA-NAc circuit is disengaged during decision making in older adults.

Unlike BLA inactivation during outcome evaluation, inactivation during deliberation in both young and aged rats mimicked the attenuated impulsive choice observed in aging (Figure 4C; Simon et al., 2010; Hernandez et al., 2017). This effect is only observed using temporally-discrete optogenetic inhibition during deliberation, and not during outcome evaluation or with experimental methods such as lesions or pharmacological inactivation that inhibit the BLA across all stages of the decision process (Winstanley et al., 2004; Churchwell et al., 2009). Such data suggest that in young rats, activity in BLA circuits involved in outcome evaluation may be the primary driver of choice behavior. The failure to engage such outcome evaluation circuits in aging, however, may ‘unmask’ the contributions of BLA during deliberation. According to this hypothesis, structural or functional changes in BLA that occur with aging would thus exert their influence on intertemporal choice through a putative ‘deliberation circuit’, perhaps involving BLA projections to prefrontal cortex (PFC; Burgos-Robles et al., 2017). The BLA contains co-distributed neurons that send distinct efferent projections to PFC and NAc (Pérez-Jaranay and Vives, 1991; Ambroggi et al., 2008; Shiflett and Balleine, 2010; Dilgen et al., 2013; McGarry and Carter, 2017). These distinct populations of BLA efferents may subserve unique roles in intertemporal choice, and further, may be differentially susceptible to aging. In other words, attenuated impulsive choice in aged rats might reflect a failure to engage a BLA-NAc ‘outcome evaluation’ circuit in combination with a hypoactive BLA-PFC ‘deliberation’ circuit that mimics the effect of BLA inactivation during deliberation. Future experiments applying circuit-based optogenetic approaches to the study of decision making in aging should be helpful for further elucidating the neurobiological substrates of age-associated alterations in intertemporal choice.

Conclusions
The current experiments demonstrate several unique roles for BLA activity in intertemporal choice. First, these data demonstrate a novel role for BLA in promoting impulsive choices when deciding whether to delay gratification. Moreover, these experiments defined a second role for BLA in attenuating impulsive choices, by providing negative feedback about the inadequacy of small vs large reward. Notably, this latter role in negative feedback does not extend to the aversives properties of the delay, indicating that the integrated valuation of reward and costs occurs outside of the BLA. Finally, the current experiments demonstrate that these temporally distinct roles of BLA in decision making change in aging. Specifically, aged rats do not appear to use BLA in any form of outcome evaluation. Moreover, the effects of age on intertemporal choice are mimicked by inactivation of
BLA during deliberation. These findings suggest complex effects of aging within and/or outside BLA, which may uniquely impact distinct BLA efferent circuits. This study is among the first to apply optogenetic techniques to the study of cognitive aging. The findings offer unique insights into how BLA mediates intertemporal choice, and show that optogenetic approaches can be used to complement and extend our understanding of how changes in neural activity guide behavioral alterations in aged subjects.

Materials and methods

Subjects
Young (6 months old, n = 24) and aged (24 months old, n = 19 male Fischer 344 x Brown Norway F1 hybrid (FBN) rats were obtained from the National Institute on Aging colony (Charles River Laboratories) and individually housed in the Association for Assessment and Accreditation of Laboratory Animal Care International-accredited vivarium facility in the McKnight Brain Institute building at the University of Florida in accordance with the rules and regulations of the University of Florida Institutional Animal Care and Use Committee and National Institutes of Health guidelines. The facility was maintained at a consistent temperature of 25°C with a 12 hr light/dark cycle (lights on at 0600) and free access to food and water except as otherwise noted. Rats were acclimated in this facility and handled for at least one week prior to initiation of any procedures. A subset of rats completed only some of the behavioral epochs due to lost headcaps and premature death, and some rats were excluded entirely for misplaced injections. Only the final numbers of rats included in each analysis are provided below.

Surgical procedures
Surgical procedures were performed as in our previous work (Orsini et al., 2017). Rats were anesthetized with isofluorane gas (1–5% in O₂) and secured in a stereotaxic frame (David Kopf). An incision along the midline over the skull was made and the skin was retracted. Bilateral burr holes were drilled above the BLA and five additional burr holes were drilled to fit stainless steel anchoring screws. Bilateral guide cannulae (22-gauge, Plastics One) were implanted to target the BLA (antero-posterior (AP): −3.25 mm from bregma, mediolateral (ML): ±4.95 mm from bregma, dorsoventral (DV): −7.3 mm from the skull surface) and secured to the skull using dental cement. A total of 0.6 μL of a 3.5 × 10¹² vg/ml titer solution (University of North Carolina Vector Core) containing AAV5 packaged with either halorhodopsin (CamKIIα-eNpHR3.0-mCherry, n = 8 young and n = 7 aged rats) or mCherry alone (CamKIIα-mCherry, n = 4 young and n = 4 aged rats) was delivered through the implanted cannulae at a rate of 0.6 μL per min. Stainless steel obturators were placed into the cannulae to minimize the risk of infection. Immediately after surgery, rats received subcutaneous injections of buprenorphine (1 mg/kg) and meloxicam (2 mg/kg). Buprenorphine was also administered 24 hr post-operation, and meloxicam 48–72 hr post-operation. A topical ointment was applied as needed to facilitate wound healing. Sutures were removed after 10–14 days and rats recovered for at least 2 weeks before food restriction and behavioral testing began.

In vitro electrophysiology
For in vitro electrophysiological verification of functional halorhodopsin (eNpHR3.0), young (n = 4) and aged (n = 3) rats underwent surgery as described above except that neither guide cannulae nor skull screws were implanted. Following a 3–4 week survival time, rats were deeply anesthetized via i.p. injection of ketamine (75–100 mg/kg) and xylazine (5–10 mg/kg). The brain was rapidly cooled via transcardial perfusion with cold oxygenated sucrose-laden artificial cerebrospinal fluid (ACSF) containing (in mM): 205 sucrose, 10 dextrose, 1 MgSO₄, 2 KCl, 1.25 NaH₂PO₄, 1 CaCl₂, and 25 NaHCO₃. Rats were then decapitated, brains extracted and coronal slices (300 μm) prepared using a Leica VT 1000 s vibratome. Slices were incubated for 30 min at 37°C in oxygenated low-calcium ACSF containing (in mM): 124 NaCl, 10 dextrose, 3 MgSO₄, 2.5 KCl, 1.23 NaH₂PO₄, 1 CaCl₂, and 25 NaHCO₃, after which they were transferred to room temperature for a minimum of 30 min prior to experimentation. During recording experiments, slices were bathed in ACSF containing (in mM): 125 NaCl, 11 dextrose, 1.5 MgSO₄, 3 KCl, 1.2 NaH₂PO₄, 2.4 CaCl₂, and 25 NaHCO₃, maintained at 28–30°C. The pipette (internal) solution contained (in mM): 125 K-gluconate, 10 phosphocreatine, 1
MgCl₂, 10 HEPES, 0.1 EGTA, 2 Na₂ATP, 0.25 Na₃GTP, and 5 biocytin, adjusted to pH 7.25 and 295 mOsm with KOH. BLA neurons were visualized using a combination of IR-DIC and epifluorescence microscopy using an Olympus BX51WI microscope and a TTL-controlled light source (X-Cite 110 LED light source, XF102-2 filter set, Omega Optical, excitation 540–580 nm, emission 615–695 nm, also used for in vitro activation of eNpHR3.0 for 1 or 4 s). Patch pipettes were prepared with a Flaming/Brown type pipette puller (Sutter Instrument, P-97) from 1.5 mm/0.8 mm borosilicate glass capillaries (Sutter Instruments) and pulled to an open tip resistance of 4–7 MΩ using internal solution and ACSF noted above. Electrophysiological recordings were performed using a Multiclamp 700B amplifier and Digidata 1440A digitizer (Axon Instruments/Molecular Devices). Electrophysiological data were collected at 20 kHz and low-pass filtered at 2 kHz.

At the conclusion of experiments, a subset of slices was transferred to 10% formalin (4°C, 24 hr) to allow for post hoc histological analysis. Slices were washed in PBS, permeabilized in PBS containing 0.1% Triton-X, and incubated in streptavidin conjugate with fluorophore (1:1000, 594 nm, ThermoFisher S32356). Slices were then mounted onto slides and coverslipped using VECTASHIELD. Morphological reconstruction was achieved by creating an all-in-focus maximum intensity projection of a Z-series acquired with a two-photon laser scanning epifluorescence microscope (810 nm excitation).

Behavioral testing procedures
Apparatus
Testing was conducted in four identical standard rat behavioral test chambers (Coulbourn Instruments) with metal front and back walls, transparent Plexiglas side walls, and a floor composed of steel rods (0.4 cm in diameter) spaced 1.1 cm apart. Each test chamber was housed in a sound-attenuating cubicle and was equipped with a custom food pellet delivery trough fitted with a photobeam to detect head entries (TAMIC Instruments) located 2 cm above the floor and extending 3 cm into the chamber in the center of the front wall. A nosepoke hole equipped with a 1.12 W lamp for illumination was located directly above the food trough. Food rewards consisted of 45 mg grain-based food pellets (PJAI; Test Diet, Richmond, IN, USA). Two retractable levers were positioned to the left and right of the food trough (11 cm above the floor). A 1.12 W house light was mounted near the top of the rear wall of the sound-attenuating cubicle. For optical activation of eNpHR3.0, laser light (561 nm, 8–10 mW output at the fiber tip, Shanghai Laser and Optics Century) was delivered through a patch cord (200 μm core, Thor Labs) to a rotary joint (1 x 2, 200 μm core, Doric Lenses) mounted above the operant chamber. At the rotary joint, the light was split into two outputs. Tethers (200 μm core, 0.22 NA, Thor Labs) connected these outputs to the bilateral optic fibers (200 μm core, 0.22 NA, 8.3 mm in length; Precision Fiber Products) implanted in the BLA (Orsini et al., 2017). A computer running Graphic State 4.0 software (Coulbourn Instruments) was used to control the behavioral apparatus and laser delivery, and to collect data.

Behavioral shaping and initial training
The intertemporal choice task was based on a design by Evenden and Ryan (1996) and was used previously to demonstrate age-related alterations in decision making in both Fischer 344 (Simon et al., 2010) and FBN (Hernandez et al., 2017) rats (Figure 1). Prior to commencement of behavioral testing, both young and aged rats were food-restricted to 85% of their free-feeding weights over the course of 5 days and maintained at these weights for the duration of the experiments. Rats were initially shaped to lever press to initiate delivery of a food pellet into the food trough and were then trained to nosepoke to initiate lever extension. Each nosepoke initiated extension of either the left or right lever (randomized across pairs of trials), a press on which yielded a single food pellet. After two consecutive days of reaching criterion performance (45 presses on each lever), rats began testing on the intertemporal choice task.

Intertemporal choice task
Each 60 min session consisted of 3 blocks of 20 trials each. The trials were 60 s in duration and began with a 10 s illumination of both the nosepoke port and house light. A nosepoke into the port during this time extinguished the nosepoke light and triggered lever extension. Any trials on which rats failed to nosepoke during this 10 s window were scored as omissions. Each 20-trial block began
with two forced choice trials, in which either the right or left lever was extended, in order to remind rats of the delay contingencies in effect for that block. These forced choice trials were followed by 18 free choice trials, in which both levers were extended. For all trials, one lever (either left or right, counterbalanced across age groups) was always associated with immediate delivery of one food pellet (the small reward), and the other lever was associated with three food pellets (the large reward) delivered after a variable delay. Lever assignment (small or large reward) remained consistent throughout testing. Within a session, the duration of the delay preceding large reward delivery increased across the three blocks of trials. The actual delay durations were adjusted individually for each rat, such that the percent choice of the large reward corresponded to roughly 100% in block 1, 66% in block 2, and 33% in block 3. On all trials, rats were given 10 s to press a lever, after which the levers were retracted, and food was delivered into the food through. If rats failed to press a lever within 10 s, the levers were retracted, lights were extinguished, and the trial was scored as an omission. An inter-trial interval (ITI) followed either food delivery or an omitted trial, after which the next trial began.

Rats were initially trained for 15 sessions on the intertemporal choice task. They were then lightly anesthetized and optic fibers (Precision Fiber Products) were inserted into the guide cannulae such that they extended 1 mm past the end of the guide cannulae and were cemented in place. After recovery, rats resumed training but were now tethered to the rotary joint.

Evaluation of optogenetic inactivation during specific task epochs
The effects of temporally-discrete optogenetic inhibition of BLA were tested in both young and aged rats using a within-subjects design. Data from sessions occurring just prior to inactivation sessions (in which rats did not receive light delivery) were used as the baseline against which to compare the effects of BLA inactivation. Task epochs in which the BLA was inactivated included: deliberation (light delivery began 500 ms prior to illumination of the nosepoke light and continued until the rat pressed the lever, for a maximum of 10 s); small reward delivery (light delivery began when food was dispensed and remained on for 4 s); large reward delivery (light delivery began when food was dispensed and remained on for 4 s); delay (light delivery began upon pressing the large reward lever and remained on throughout delay intervals ranging between 2–24 s); large reward delivery + delay (light delivery began upon pressing the large reward lever and remained on until 4 s after the large reward was dispensed), intertrial interval (ITI; light delivery began 14 s after reward was dispensed and continued for 4 s). Finally, the sequence of BLA inactivation sessions during each task epoch was counterbalanced across rats.

Vector expression and cannula placement histology
After completion of behavioral testing, rats were administered a lethal dose of Euthasol (sodium pentobarbital and phenytoin solution; Virbac, Fort Worth, TX, USA) and perfused transcardially with a 4°C solution of 0.1M phosphate buffered saline (PBS), followed by 4% (w/v) paraformaldehyde in 0.1M PBS. Brains were removed and post-fixed for 24 hr then transferred to a 20% (w/v) sucrose solution in 0.1M PBS for 72 hr (all chemicals purchased from Fisher Scientific, Hampton, NH, USA). Brains were sectioned at 35 μm using a cryostat maintained at −20°C. Sections were then incubated with rabbit anti-mCherry antibody (ab167453, Abcam, Cambridge, MA, USA) diluted in blocking solution consisting of 3% normal goat serum, 0.3% Triton-X-100 in 0.1M PBS for 1 hr at room temperature. Sections were then incubated with donkey anti-rabbit conjugated to AlexaFluor-488 (1:300) for 2 hr at room temperature. Following primary incubation, sections were rinsed in 0.1M TBS and incubated in blocking solution containing the secondary antibody (donkey anti-rabbit conjugated to AlexaFluor-488, 1:300) for 2 hr at room temperature. After rinsing in 0.1M TBS, sections were mounted on electrostatic glass slides and coverslipped using Prolong Gold containing DAPI (Thermo Fisher Scientific, Waltham, MA, USA). Slides were sealed with clear nail polish and sections were visualized at 20X using an Axio Imager 2 microscopy system (Carl Zeiss Microscopy, LLC, Thornwood, NY, USA) to assess mCherry expression in BLA neurons. Cannula placements and mCherry expression were mapped onto plates adapted from the rat brain atlas of Paxinos and Watson (2005).
**Experimental design and statistical analysis**

**Evaluation of age differences in halorhodopsin effects on BLA neuronal activity**

Data analysis was performed using OriginLab and custom electrophysiology analysis software written by CJF. Electrophysiological measures were compared between young and aged cells (obtained from \( n = 2-4 \) rats per age) using an independent samples t-test.

**Evaluation of age differences in intertemporal choice under baseline conditions**

Raw data files were extracted using a Graphic State 4.0 analysis template that was custom-designed to extract the number of lever presses on each lever (large or small rewards) during forced and free choice trials in each trial block. First, age differences in intertemporal choice performance were tested by analyzing the actual delays used to achieve the target 100%, 66% and 33% choice of the large reward in blocks 1, 2 and 3, respectively. Actual delays were compared using a mixed-factor ANOVA, with age (two levels: young and aged) as the between-subjects factor and block (three levels: blocks 1–3) as the within-subjects factor. Second, the choice indifference point, or the delay at which a rat showed equivalent choice of the small and large reward, was calculated and compared between young and aged rats. Choice indifference points were calculated by fitting a trend line to each rat’s percent choice of the large reward at each delay block. The slope-intercept formula, \( y = mx + b \) (where ‘\( y \)’ is percent choice of the large reward, and ‘\( x \)’ is delay), was then used to solve for the number of seconds (\( x \)) at which \( y = 50\% \) choice of the large reward (the delay at which the rat was equally likely to choose the large or small reward). Choice indifference points were compared between young and aged rats using an independent samples t-test. Alpha was set to 0.05 for all statistical analyses. For statistical results that reached the threshold for significance, \( \eta_p^2 \) (partial eta squared) and Cohen’s \( d \) were used to report the effect size for mixed-factor ANOVAs and t-tests, respectively, and \( 1-\beta \) was used to report the observed power.

**Evaluation of BLA inactivation on intertemporal choice**

Power analyses based on data from an initial cohort of rats (\( n = 3 \)) were used to determine sample sizes necessary to evaluate the effects of BLA inactivation on choice behavior. These analyses indicated the presence of large effect sizes (greater than 1.0), and that \( n = 6 \) rats should be sufficient to detect effects of BLA inactivation, with a power to detect significant differences of 0.95. The effects of light delivery were tested separately for each task epoch (deliberation, small reward delivery, large reward delivery, delay, delay + large reward delivery, and ITI). For each epoch, comparisons were made using a mixed factor ANOVA (laser condition \( \times \) age \( \times \) delay block), with age as the between-subjects factor (two levels: young and aged), and laser condition (two levels: laser on or off) and delay block (three levels: delay blocks 1–3) as within-subjects factors. To better understand significant main effects or interactions, post hoc analyses were conducted in each age group separately using a repeated-measures ANOVA (laser condition \( \times \) block). Note that for those epochs in which effects of BLA inactivation during the delay were tested, data analyses were confined to blocks 2 and 3, as there was no delay in block 1.

**Evaluation of choice strategy resulting from BLA inactivation**

Additional analyses were conducted to better understand the shifts in choice performance following BLA inactivation during the deliberation and small reward epochs. Graphic State 4.0 templates were created to assess trial-by-trial choices during baseline and BLA inactivation sessions for the deliberation and small reward epochs. Trials were categorized based on choices made on the previous trial. For the deliberation epoch, trials were categorized as ‘small-shift-to-large’ or ‘large-stay-on-large’. For the small reward delivery epoch, trials were categorized as ‘large-shift-to-small’ or ‘small-stay-on-small’. The number of trials in each category was divided by the total number of trials in that session and expressed as a percentage. For each task epoch, percentages of trials in each category were compared using a mixed-factor ANOVA with age as the between-subjects factor (two levels: young and aged) and laser condition as the within-subjects factor (two levels: laser on or off).
Effects on other task performance measures resulting from BLA inactivation
Other task measures were compared between BLA inactivation and baseline conditions in task
epochs in which BLA inactivation produced significant changes in choice behavior. Specifically, on
free choice trials, response latency (the time between lever extension and a lever press) was com-
pared. Previous work shows that response latencies can differ for large and small reward levers
(Hernandez et al., 2017) and hence analyses were conducted separately for each lever using data
from delay block 2, during which rats made roughly equivalent numbers of choices on each reward
lever. Response latency and total number of trials completed were compared using a mixed factor
ANOVA (laser condition × age).

Acknowledgements
We thank Vicky S Kelly, Shannon C Wall, and Bonnie I McLaurin for assistance with surgeries and
behavioral testing, Wanhui Sheng for assistance with two-photon microscopy, and Jeff Thinschmidt
for assistance with patch clamp recording. Supported by R01AG029421 (JLB), RF1AG060778 (JLB,
BS, CJF), the McKnight Brain Research Foundation (JLB), a McKnight Predoctoral Fellowship and the
Pat Tillman Foundation (CMH), and a Thomas H. Maren Fellowship and K99DA041493 (CAO).

Additional information

| Funder                        | Grant reference number | Author                      |
|-------------------------------|------------------------|-----------------------------|
| McKnight Brain Research       | R01AG029421            | Jennifer L Bizon            |
| Foundation                    |                        |                             |
| National Institutes of Health | R01AG029421            | Jennifer L Bizon            |
| McKnight Foundation           |                        | Caesar M Hernandez          |
| Pat Tillman Foundation        |                        | Caesar M Hernandez          |
| Thomas H. Maren Foundation    | K99DA041493            | Caitlin A Orsini            |
| National Institutes of Health | RF1AG060778            | Charles J Frazier, Barry Setlow, Jennifer L Bizon |

The funders had no role in study design, data collection and interpretation, or the
decision to submit the work for publication.

Author contributions
Caesar M Hernandez, Conceptualization, Data curation, Formal analysis, Investigation, Methodology,
Writing—original draft, Writing—review and editing; Caitlin A Orsini, Conceptualization, Formal
analysis, Investigation, Methodology, Writing—review and editing; Chase C Labiste, Alexa-Rae
Wheeler, Tyler W Ten Eyck, Matthew M Bruner, Todd J Sahagian, Scott W Harden, Investigation;
Charles J Frazier, Conceptualization, Software, Formal analysis, Supervision, Methodology, Project
administration, Writing—review and editing; Barry Setlow, Jennifer L Bizon, Conceptualization,
Resources, Supervision, Funding acquisition, Methodology, Writing—original draft, Project adminis-
tration, Writing—review and editing

Author ORCIDs
Caesar M Hernandez https://orcid.org/0000-0001-9690-5119
Scott W Harden http://orcid.org/0000-0002-0757-1979
Barry Setlow https://orcid.org/0000-0001-9133-9445
Jennifer L Bizon https://orcid.org/0000-0002-9517-5844
Ethics
Animal experimentation: This research was conducted in accordance with the rules and regulations of the University of Florida Institutional Animal Care and Use Committee (protocol number 201604961) and National Institutes of Health guidelines.

Decision letter and Author response
Decision letter https://doi.org/10.7554/eLife.46174.023
Author response https://doi.org/10.7554/eLife.46174.024

Additional files
Supplementary files
- Transparent reporting form
  DOI: https://doi.org/10.7554/eLife.46174.021

Data availability
All data generated or analyzed during this study are included in the manuscript and supporting files.

References
Abela AR, Chudasama Y. 2013. Dissociable contributions of the ventral hippocampus and orbitofrontal cortex to decision-making with a delayed or uncertain outcome. European Journal of Neuroscience 37:640–647. DOI: https://doi.org/10.1111/jen.12071, PMID: 23190048
Almaguer W, Estupiñan B, Uwe Frey J, Bergado JA. 2002. Aging impairs amygdala-hippocampus interactions involved in hippocampal LTP. Neurobiology of Aging 23:319–324. DOI: https://doi.org/10.1016/S0197-4580(01)00278-0, PMID: 11804717
Ambroggi F, Ishikawa A, Fields HL, Nicola SM. 2008. Basolateral amygdala neurons facilitate reward-seeking behavior by exciting nucleus accumbens neurons. Neuron 59:648–661. DOI: https://doi.org/10.1016/j.neuron.2008.07.004, PMID: 18760700
Antonenko D, Füel A. 2014. Healthy aging by staying selectively connected: a mini-review. Gerontology 60:3–9. DOI: https://doi.org/10.1159/000354376, PMID: 24080587
Bailey MR, Simpson EH, Balsam PD. 2016. Neural substrates underlying effort, time, and risk-based decision making in motivated behavior. Neurobiology of Learning and Memory 133:233–256. DOI: https://doi.org/10.1016/j.nlm.2016.07.015, PMID: 27427327
Baxter MG, Parker A, Lindner CC, Izquierdo AD, Murray EA. 2000. Control of response selection by reinforcer value requires interaction of amygdala and orbital prefrontal cortex. The Journal of Neuroscience 20:4311–4319. DOI: https://doi.org/10.1523/JNEUROSCI.20-11-04311.2000, PMID: 10818166
Baxter MG, Murray EA. 2002. The amygdala and reward. Nature Reviews Neuroscience 3:563–573. DOI: https://doi.org/10.1038/nnrn875, PMID: 12094212
Beas BS, Setlow B, Gregory R, Samanez-Larkin BJL. 2015. Modeling Cost-Benefit Decision Making in Aged Rodents. In: In: Aging and Decision Making: Empirical and Applied Perspectives. Elsevier. p. 16–40.
Bercovici DA, Princz-Lebel O, Tse MT, Moorman DE, Floresco SB. 2018. Optogenetic dissection of temporal dynamics of Amygdala-Striatal interplay during risk/Reward decision making. Eneuro 5:ENEURO.0422-18.2018. DOI: https://doi.org/10.1523/ENEURO.0422-18.2018, PMID: 30627636
Bickel WK, Koffarnus MN, Moody L, Wilson AG. 2014. The behavioral- and neuro-economic process of temporal discounting: a candidate behavioral marker of addiction. Neuropharmacology 76:518–527. DOI: https://doi.org/10.1016/j.neuropharm.2013.06.013, PMID: 23806805
Bobeva L, Finn PR, Rickert ME, Lucas J. 2009. Disinhibitory psychopathology and delay discounting in alcohol dependence: personality and cognitive correlates. Experimental and Clinical Psychopharmacology 17:51–61. DOI: https://doi.org/10.1037/a0014503, PMID: 19186934
Boyle PA, Yu L, Wilson RS, Gamble K, Buchman AS, Bennett DA. 2012. Poor decision making is a consequence of cognitive decline among older persons without Alzheimer’s disease or mild cognitive impairment. PLOS ONE 7:e43647. DOI: https://doi.org/10.1371/journal.pone.0043647, PMID: 22916287
Burgos-Robles A, Kimchi EY, Izadmehr EM, Porzenheim MJ, Ramos-Guasp WA, Nieh EH, Felix-Ortiz AC, Namburi P, Leplla CA, Presbrey KN, Anandalingam KK, Pagan-Rivera PA, Ananta M, Beyeler A, Tye KM. 2017. Amygdala inputs to prefrontal cortex guide behavior amid conflicting cues of reward and punishment. Nature Neuroscience 20:824–835. DOI: https://doi.org/10.1038/nn.4553, PMID: 28436980
Burke SN, Thome A, Plange K, Engle JR, Trouard TP, Gothard KM, Barnes CA. 2014. Orbitofrontal cortex volume in area 11/13 predicts reward devaluation, but not reversal learning performance, in young and aged monkeys. Journal of Neuroscience 34:9905–9916. DOI: https://doi.org/10.1523/JNEUROSCI.3918-13.2014, PMID: 25057193
Hernandez et al. eLife 2019;8:e46174. DOI: https://doi.org/10.7554/eLife.46174
Hu X, Uhle F, Fließbach K, Wagner M, Han Y, Weber B, Jessen F. 2017. Reduced future-oriented decision making in individuals with subjective cognitive decline: a functional MRI study. Alzheimer’s & Dementia: Diagnosis, Assessment & Disease Monitoring 6:222–231. DOI: https://doi.org/10.1016/j.dadm.2017.02.005

Izquierdo A, Darling C, Manos N, Pozos H, Kim C, Ostrander S, Cazares V, Stepp H, Rudebeck PH. 2013. Basolateral amygdala lesions facilitate reward choices after negative feedback in rats. Journal of Neuroscience 33:4105–4109. DOI: https://doi.org/10.1523/JNEUROSCI.4942-12.2013, PMID: 23447618

Jimura K, Myerson J, Hilgard J, Keighley J, Braver TS, Green L. 2011. Domain independence and stability in young and older adults’ discounting of delayed rewards. Behavioural Processes 87:253–259. DOI: https://doi.org/10.1016/j.beproc.2011.04.006, PMID: 21550384

Johansen JP, Wolff SB, Løthi A, LeDoux JE. 2012. Controlling the elements: an optogenetic approach to understanding the neural circuits of fear. Biological Psychiatry 71:1053–1060. DOI: https://doi.org/10.1016/j.biopsych.2011.10.023, PMID: 22169096

Kaye WH, Wierenga CE, Bailar UF, Simmons AN, Bischoff-Grethe A. 2013. Nothing tastes as good as skinny feels: the neurobiology of anorexia nervosa. Trends in Neurosciences 36:110–120. DOI: https://doi.org/10.1016/j.tins.2013.01.003, PMID: 23333342

Levin N, Kritman M, Maroun M, Akirav I. 2017. Differential roles of the infralimbic and prelimbic areas of the prefrontal cortex in reconsolidation of a traumatic memory. European Neuropsychopharmacology 27:900–912. DOI: https://doi.org/10.1016/j.euroneuro.2017.06.007, PMID: 28647452

Lighthall NR, Huettel SA, Cabeza R. 2014. Functional compensation in the ventromedial prefrontal cortex improves memory-dependent decisions in older adults. Journal of Neuroscience 34:15648–15657. DOI: https://doi.org/10.1523/JNEUROSCI.2888-14.2014, PMID: 25411493

Löckenhoff CE, O’Donoghue T, Dunning D. 2011. Age differences in temporal discounting: the role of dispositional affect and anticipated emotions. Psychology and Aging 26:274–284. DOI: https://doi.org/10.1037/a0023280, PMID: 21534688

Lolova I, Davidoff M. 1991. Changes in GABA-immunoreactivity and GABA-transaminase activity in rat amygdaloid complex in aging. Journal For Hirnforschung 32:231–238. PMID: 1791303

Málková L, Gaffan D, Murray EA. 1997. Excitotoxic lesions of the amygdala fail to produce impairment in visual learning for auditory secondary reinforcement but interfere with reinf ercer devaluation effects in rhesus monkeys. The Journal of Neuroscience 17:6011–6020. DOI: https://doi.org/10.1523/JNEUROSCI.17-15-06011.1997, PMID: 9221797

Mariano TY, Bannerman DM, McHugh SB, Preston TJ, Rudebeck PH, Rudebeck SR, Rawlins JN, Walton ME, Rushworth MF, Baxter MG, Campbell TG. 2009. Impulsive choice in hippocampal but not orbitofrontal cortex-lesioned rats on a nonspatial decision-making maze task. European Journal of Neuroscience 30:472–484. DOI: https://doi.org/10.1111/j.1460-9568.2009.06837.x, PMID: 19651777

Mata R, Josef AK, Samanez-Larkin GR, Hertwig R. 2011. Age differences in risky choice: a meta-analysis. Annals of the New York Academy of Sciences 1235:18–29. DOI: https://doi.org/10.1111/j.1749-6632.2011.06200.x, PMID: 22023565

Matther M. 2006. A Review of Decision-Making Processes. In: Weighing the Risks and Benefits of Aging. US: National Academies Press.

Matther M, Mazar N, Gorlick MA, Lighthall NR, Burgeno J, Schoeke A, Ariely D. 2012. Risk preferences and aging: the "certainty effect" in older adults’ decision making. Psychology and Aging 27:801–816. DOI: https://doi.org/10.1037/a0030174, PMID: 23066800

McGarry LM, Carter AG. 2017. Prefrontal cortex drives distinct projection neurons in the basolateral amygdala. Cell Reports 21:1426–1433. DOI: https://doi.org/10.1016/j.celrep.2017.10.046, PMID: 29117549

Mitchell MR, Weiss VG, Beas BS, Morgan D, Bizon JL, Setlow B. 2014. Adolescent risk taking, cocaine self-administration, and striatal dopamine signaling. Neuropsychopharmacology 39:955–962. DOI: https://doi.org/10.1038/npp.2013.295, PMID: 24145852

Orsini CA, Moorman DE, Young JW, Setlow B, Floresco SB. 2015a. Neural mechanisms regulating different forms of risk-related decision-making: insights from animal models. Neuroscience & Biobehavioral Reviews 58:147–167. DOI: https://doi.org/10.1016/j.neubiorev.2015.04.009, PMID: 26072028

Orsini CA, Trotta RT, Bizon JL, Setlow B. 2015b. Dissociable roles for the basolateral amygdala and orbitofrontal cortex in decision-making under risk of punishment. Journal of Neuroscience 35:1368–1379. DOI: https://doi.org/10.1523/JNEUROSCI.3586-14.2015, PMID: 25632115

Orsini CA, Hernandez CM, Singhal S, Kelly KB, Frazier CJ, Bizon JL, Setlow B. 2017. Optogenetic inhibition reveals distinct roles for basolateral amygdala activity at discrete time points during risky decision making. The Journal of Neuroscience 37:11537–11548. DOI: https://doi.org/10.1523/JNEUROSCI.2344-17.2017, PMID: 2 9079687

Orsini CA, Heshmati SC, Garman TS, Wall SC, Bizon JL, Setlow B. 2018. Contributions of medial prefrontal cortex to decision making involving risk of punishment. Neuropharmacology 139:205–216. DOI: https://doi.org/10.1016/j.neuropharm.2018.07.018, PMID: 30009836

Pachur T, Mata R, Hertwig R. 2017. Who dares, who errs? disentangling cognitive and motivational roots of age differences in decisions under risk. Psychological Science 28:504–518. DOI: https://doi.org/10.1177/0956797616687729, PMID: 28406375

Parkes SL, Balleine BW. 2013. Incentive memory: evidence the basolateral amygdala encodes and the insular cortex retrieves outcome values to guide choice between goal-directed actions. Journal of Neuroscience 33:8753–8763. DOI: https://doi.org/10.1523/JNEUROSCI.5071-12.2013, PMID: 23678118
Patros CH, Alderson RM, Kasper Lj, Tarle SJ, Lea SE, Hudec KL. 2016. Choice-impulsivity in children and adolescents with attention-deficit/hyperactivity disorder (ADHD): A meta-analytic review. Clinical Psychology Review 43:162–174. DOI: https://doi.org/10.1016/j.cpr.2015.11.001, PMID: 26602954

Paxinos G, Watson C. 2005. The Rat Brain in Stereotaxic Coordinates. New York: Elsevier Academic.

Pérez-Jaranay JM, Vives F. 1991. Electrophysiological study of the response of medial prefrontal cortex neurons to stimulation of the basolateral nucleus of the amygdala in the rat. Brain Research 564:97–101. DOI: https://doi.org/10.1016/0006-8993(91)91357-7, PMID: 1777825

Peters J, Büchel C. 2011. The neural mechanisms of inter-temporal decision-making: understanding variability. Trends in Cognitive Sciences 15:227–239. DOI: https://doi.org/10.1016/j.tics.2011.03.002, PMID: 21497544

Prager EM, Bergstrom HC, Wynn GH, Braga MF. 2016. The basolateral amygdala γ-aminobutyric acidergic system in health and disease. Journal of Neuroscience Research 94:548–567. DOI: https://doi.org/10.1002/jnrr.23690, PMID: 26586374

Rangel A, Camerer C, Montague PR. 2008. A framework for studying the neurobiology of value-based decision making. Nature Reviews Neuroscience 9:545–556. DOI: https://doi.org/10.1038/nrn2357, PMID: 18545266

Roesch MR, Esber GR, Bryden DW, Cerri DH, Haney ZR, Schoenbaum G. 2012. Normal aging alters learning and attention-related teaching signals in Basolateral amygdala. Journal of Neuroscience 32:13137–13144. DOI: https://doi.org/10.1523/JNEUROSCI.2393-12.2012, PMID: 22993430

Rubinow MJ, Drogos LL, Juraska JM. 2009. Age-related dendritic hypertrophy and sexual dimorphism in rat basolateral amygdala. Neurobiology of Aging 30:137–146. DOI: https://doi.org/10.1016/j.neurobiologyaging.2008.06.004, PMID: 17570563

Rubinow MJ, Juraska JM. 2009. Neuron and glia numbers in the basolateral nucleus of the amygdala from preweaning through old age in male and female rats: a stereological study. The Journal of Comparative Neurology 512:717–725. DOI: https://doi.org/10.1002/cne.21924, PMID: 19065620

Samanez-Larkin GR, Mata R, Radu PT, Ballard IC, Carstensen LL, McClure SM. 2011. Age differences in striatal delay sensitivity during intertemporal choice in healthy adults. Frontiers in Neuroscience 5:126. DOI: https://doi.org/10.3389/fnins.2011.00126, PMID: 22110424

Samanez-Larkin GR, Knutson B. 2015. Decision making in the ageing brain: changes in affective and motivational circuits. Nature Reviews Neuroscience 16:278–289. DOI: https://doi.org/10.1038/nrn3917, PMID: 25873038

Samson RD, Lester AW, Duarte L, Venkatesh A, Barnes CA. 2017. Emergence of β-Band oscillations in the aged rat amygdala during discrimination learning and decision making tasks. Eneuro 4:Eneuro.0245-17.2017. DOI: https://doi.org/10.1523/ENEURO.0245-17.2017, PMID: 29034315

Schoenbaum G, Chiba AA, Gallagher M. 1998. Orbitofrontal cortex and basolateral amygdala encode expected outcomes during learning. Nature Neuroscience 1:155–159. DOI: https://doi.org/10.1038/40760, PMID: 10195132

Schoenbaum G, Chiba AA, Gallagher M. 1999. Neural encoding in orbitofrontal cortex and basolateral amygdala during olfactory discrimination learning. The Journal of Neuroscience 19:1876–1884. DOI: https://doi.org/10.1523/JNEUROSCI.19-05-01876.1999, PMID: 10024371

Shadlen MN, Shohamy D. 2016. Decision making and sequential sampling from memory. Neuron 90:927–939. DOI: https://doi.org/10.1016/j.neuron.2016.04.036, PMID: 27253447

Shamosh NA, Deyoung CG, Green AE, Reis DL, Johnson MR, Conway AR, Engle RW, Braver TS, Gray JR. 2008. Individual differences in delay discounting: relation to intelligence, working memory, and anterior prefrontal cortex. Psychological Science 19:904–911. DOI: https://doi.org/10.1111/j.1467-9280.2008.02175.x, PMID: 18947356

Shifflett MW, Balleine BW. 2010. At the limbic-motor interface: disconnection of basolateral amygdala from nucleus accumbens core and shell reveals dissociable components of incentive motivation. European Journal of Neuroscience 32:1735–1743. DOI: https://doi.org/10.1111/j.1465-9291.2010.07439.x, PMID: 21044174

Shimp KG, Mitchell MR, Beas BS, Bizon JL, Setlow B. 2015. Affective and cognitive mechanisms of risky decision making. Neurobiology of Learning and Memory 117:60–70. DOI: https://doi.org/10.1016/j.nlm.2014.03.002, PMID: 26642448

Simon NW, LaSarge CL, Montgomery KS, Williams MT, Mendez IA, Setlow B, Bizon JL. 2010. Good things come to those who wait: attenuated discounting of delayed rewards in aged Fischer 344 rats. Neurobiology of Aging 31:853–862. DOI: https://doi.org/10.1016/j.neurobiaging.2008.06.004, PMID: 18657883

Simon NW, Montgomery KS, Beas BS, Mitchell MR, LaSarge CL, Mendez IA, Bañuelos C, Vokes CM, Taylor AB, Haberman RP, Bizon JL, Setlow B. 2011. Dopaminergic modulation of risky decision-making. Journal of Neuroscience 31:17460–17470. DOI: https://doi.org/10.1523/JNEUROSCI.3772-11.2011, PMID: 22131407

Sonntag KC, Brenhouse HC, Freund N, Thompson BS, Puhl M, Andersen SL. 2014. Viral over-expression of D1 dopamine receptors in the prefrontal cortex increase high-risk behaviors in adults: comparison with adolescents. Psychopharmacology 231:1615–1626. DOI: https://doi.org/10.1007/s00213-013-3399-8, PMID: 24408208

Steinglass JE, Figner B, Berkowitz S, Simpson HB, Weber EU, Walsh BT. 2012. Increased capacity to delay reward in anorexia nervosa. Journal of the International Neuropsychological Society 18:773–780. DOI: https://doi.org/10.1017/S1355617712000446, PMID: 22591835

Tomás Pereira I, Gallagher M, Rapp PR. 2015. Head west or left, east or right: interactions between memory systems in neurocognitive aging. Neurobiology of Aging 36:3067–3078. DOI: https://doi.org/10.1016/j.neurobiaging.2015.07.024, PMID: 26281759
Wang WC, Dew IT, Cabeza R. 2015. Age-related differences in medial temporal lobe involvement during conceptual fluency. Brain Research 1612:48–58. DOI: https://doi.org/10.1016/j.brainres.2014.09.061, PMID: 25305568

Wassum KM, Izquierdo A. 2015. The basolateral amygdala in reward learning and addiction. Neuroscience & Biobehavioral Reviews 57:271–283. DOI: https://doi.org/10.1016/j.neubiorev.2015.08.017, PMID: 26341938

Winstanley CA, Theobald DE, Cardinal RN, Robbins TW. 2004. Contrasting roles of basolateral amygdala and orbitofrontal cortex in impulsive choice. Journal of Neuroscience 24:4718–4722. DOI: https://doi.org/10.1523/JNEUROSCI.5606-03.2004, PMID: 15152031

Yates JR, Perry JL, Meyer AC, Gipson CD, Chamigo R, Bardo MT. 2014. Role of medial prefrontal and orbitofrontal monoamine transporters and receptors in performance in an adjusting delay discounting procedure. Brain Research 1574:26–36. DOI: https://doi.org/10.1016/j.brainres.2014.06.004, PMID: 24928616

Zangemeister L, Grabenhorst F, Schultz W. 2016. Neural basis for economic saving strategies in human Amygdala-Prefrontal reward circuits. Current Biology 26:3004–3013. DOI: https://doi.org/10.1016/j.cub.2016.09.016, PMID: 27773572

Zeeb FD, Floresco SB, Winstanley CA. 2010. Contributions of the orbitofrontal cortex to impulsive choice: interactions with basal levels of impulsivity, dopamine signalling, and reward-related cues. Psychopharmacology 211:87–98. DOI: https://doi.org/10.1007/s00213-010-1871-2, PMID: 20428999

Zuo Y, Wang X, Cui C, Luo F, Yu P, Wang X. 2012. Cocaine-induced impulsive choices are accompanied by impaired delay-dependent anticipatory activity in Basolateral amygdala. Journal of Cognitive Neuroscience 24:196–211. DOI: https://doi.org/10.1162/jocn_a_00131, PMID: 21916564