Contact Heat Evoked Potentials in China: Normal Values and Reproducibility

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Background: Contact heat evoked potentials (CHEPs) is used to diagnose small fiber neuropathy (SFN). We established the normal values of CHEPs parameters in Chinese adults, optimized the test technique, and determined its reproducibility.

Methods: We recruited 151 healthy adults (80 men; mean age, 37 ± 14 years). CHEPs was performed on the right forearm to determine the optimal number of stimuli, and then conducted at different sites to establish normal values, determine the effects of demographic characteristics and baseline temperature, and assess the short- (30 min) and long-term (1 year) reproducibility. N2 latency/height varied with age and sex, while P2 latency/height and N2–P2 amplitude varied with age. The optimal number of stimuli was three.

Results: N2 latency/height (t = 5.45, P < 0.001) and P2 latency/height (χ² = −4.06, P < 0.001) decreased and N2–P2 amplitude (t = −5.01, P < 0.001) and visual analog scale score (χ² = −5.84, P < 0.001) increased with increased baseline temperature (35 vs. 32°C). CHEPs parameters did not differ with time (baseline vs. 30 min vs. 1 year).

Conclusion: We established normal CHEPs values in Chinese adults. We found that CHEPs parameters changed with baseline temperature and that the short- and long-term test reproducibility were satisfactory.

Keywords: contact heat evoked potentials, normal values, reproducibility, small fiber neuropathy, intraepidermal nerve fiber density

INTRODUCTION

Small fiber neuropathy (SFN) is a peripheral neuropathy that affects small-caliber thinly myelinated Aδ and unmyelinated C fibers (McCarthy et al., 1995; Said, 2003; Hoitsma et al., 2004). No consensus yet exists on the standard diagnostic criteria for SFN. Some studies report that intraepidermal nerve fiber density (IENFD) assessment is the most reliable diagnostic test for SFN (Hoitsma et al., 2004; Lauria et al., 2010a). Other studies use a combination of sensory and autonomic symptoms (not otherwise explainable) as well as intact large-fiber function on examination (normal vibration sense and nerve conduction studies) with abnormal IENFD and/or

Abbreviations: CHEPs, contact heat evoked potentials; SFN, small fiber neuropathy; IENFD, intraepidermal nerve fiber density; NCS, nerve conduction studies; EMG, electromyography.
abnormal temperature threshold testing; if any two of these criteria are met, then SFN can be diagnosed (Lauria et al., 2010b; Tesfaye et al., 2010). However, IENFD measurement requires skin biopsy and immunohistochemical staining, and the sensitivity of IENFD for SFN depends largely on the chosen cut-off values (Nebuchennykh et al., 2009). Furthermore, although this test has been widely applied in Western countries, it is not commonly available in China. Temperature threshold testing requires subject cooperation, although standardized protocols have been published, the technique varies among laboratories. Some experts’ reviews highlighted the role of neurophysiology in the diagnostic work-up of SFN (Terkelsen et al., 2017). Therefore, novel, simple, and standardized diagnostic tools of neurophysiology for SFN are required in China.

Contact heat evoked potential (CHEP) is a neuroelectrophysiologic technique in which heat stimuli consisting of rapid changes in temperature (70°C/s) are applied to the skin to evoke cerebral electroencephalographic responses conveyed by Aδ fibers (Lagerburg et al., 2015; Granovsky et al., 2016). The recorded cortical responses—namely N2 latency, P2 latency, and N2-P2 amplitude—can be used to appraise the function of small peripheral nerve fibers (Chen et al., 2006; Chao et al., 2007; Kramer et al., 2012a, 2013). The CHEP test is objective and simple, and it is a safer approach than laser-evoked potentials for evaluating small peripheral nerve fibers (Le Quesne et al., 1990; Magerl et al., 1999; Le Pera et al., 2002; Truini et al., 2005; Chao et al., 2008). In SFN, CHEPs are sensitive and correlate with IENFD (Chen et al., 2001; Le Pera et al., 2002; Atherton et al., 2007; Chao et al., 2010; Casanova-Molla et al., 2011). Furthermore, CHEPs are not only used in the diagnosis of SFN, but also in spinal disorders (Kramer et al., 2012a).

However, to diagnose SFN using CHEPs, the establishment of normal CHEP values is vital. A CHEP study of 35 normal controls was performed in Taiwan (Chen et al., 2006), and another study established normal CHEP values in a Dutch population of 97 subjects (Lagerburg et al., 2015). Furthermore, a multicenter study determined normal CHEP values in 226 subjects from Brazil, Israel, Japan, Spain, and the United States (Granovsky et al., 2016), and a Swiss study identified CHEP normative data in cervical dermatomes in 101 healthy subjects (Jutzeler et al., 2016). However, different studies have revealed conflicting relationships of CHEP parameters with variables such as gender, age, and height (Chen et al., 2006; Lagerburg et al., 2015; Granovsky et al., 2016). Moreover, no large-scale studies of normal CHEP values have been conducted in China, where the diagnosis of SFN is restricted.

Therefore, it is necessary to establish normal CHEP values in the Chinese population. Furthermore, different studies have used differing numbers of CHEP stimuli (Atherton et al., 2007; Chao et al., 2007, 2008, 2010; Casanova-Molla et al., 2011; Lagerburg et al., 2015; Granovsky et al., 2016). In this study, we aimed to determine the optimal number of stimuli to avoid habituation and simplify the CHEP test. In addition, it is meaningful to study the reproducibility of the CHEP test and baseline temperature on the results obtained.

In this study, we aimed to optimize the CHEP testing technique (to determine the optimal number of stimuli), determine the short-term and long-term reproducibility of the CHEP test, and establish the normal values of CHEP parameters in a large sample of healthy Chinese adults.

**MATERIALS AND METHODS**

**Participants**

Healthy participants were recruited from hospital volunteers and through advertisements between November 20, 2014 and December 31, 2016. All participants underwent neurological examinations, laboratory examinations, and nerve conduction studies (NCS) at outpatient departments in a comfortable, temperature-controlled room. Neurological examinations and NCS were performed by two experienced neurologists. Laboratory examinations were performed by two clinical laboratory physicians of Chinese PLA General Hospital. To be eligible for inclusion in the study, participants were required to meet the following criteria: (1) age ≥ 18 years; (2) no sensory symptoms or signs on neurological examination; (3) normal results on laboratory tests; (4) normal results on NCS; (5) absence of diseases that may cause polyneuropathy (e.g., diabetes mellitus, metabolic syndrome, and systemic illnesses like sarcoidosis or malignancy); (6) no alcohol abuse (arbitrarily defined as drinking at least 4 international units per day) (Saunders et al., 1993) or smoking history; (7) no history of hereditary diseases (e.g., hereditary motor and sensory neuropathy, hereditary sensory and autonomic disease, and Fabry disease); (8) no history of medication with neurotoxic drugs; and (9) no skin ulceration or infection at CHEP-stimulation sites.

This study was approved by the Ethics Committee of Chinese PLA General Hospital and was conducted in accordance with the Declaration of Helsinki. Written informed consent was obtained from each participant enrolled in this study.

**Laboratory Examinations**

The following laboratory tests were conducted: complete blood count, erythrocyte sedimentation rate, C-reactive protein, renal and hepatic function, lipid profile, fasting glucose, glycosylated hemoglobin, oral glucose tolerance test, folate and vitamin B12, thyroid function, antinuclear antibody, anti-extractable nuclear antigens antibody, serum protein electrophoresis, and tumor markers. The diagnoses of impaired fasting glucose, impaired glucose tolerance, and diabetes were based on the diagnostic criteria published by the World Health Organization (2006).

**Electrophysiological Examination**

All participants underwent NCS. Skin temperature was maintained at 32°C or above during the examinations. NCS were performed on the median, ulnar, tibial, peroneal, and sural nerves using the Keypoint electromyography (EMG) system (Medtronic, Inc., Minneapolis, United States). The results were
measured according to the normal reference values used by the EMG Laboratory of Chinese PLA General Hospital.

**Experimental Setup**

CHEP stimulation was conducted at the following sites:

1. Forearm (FA), at approximately the upper border of the distal third of the volar aspect of the right forearm (Dermatome T1),
2. Leg (LE), above the lateral malleolus at the upper border of the distal third of the right leg (Dermatome L5),
3. Cervical spine (C7), at the spinous process of C7 (Dermatome C3) and
4. Thoracic spine (T12), at the spinous process of T12 (Dermatome T12).

Five sessions of CHEP stimulation were performed (Figure 1). Session 1 served as a pilot study in which CHEP was performed on the right FA with 3, 5, and 10 stimuli to determine the optimal number of stimuli; the baseline temperature was set to 32°C. The order of 3, 5, and 10 stimuli series was performed randomly. The participants of the pilot study were not included in the main study. In session 2, CHEP was conducted on the right FA, right LE, C7, and T12, using the number of stimuli determined in session 1 and a baseline temperature of 32°C. The order of stimulation sites was randomized across participants. In session 3, CHEP stimulation was performed on the right FA with a baseline temperature of 32°C 1 year later to assess its long-term repeatability (session 5).

The visual analog scale (VAS) was used to evaluate pain intensity during the CHEP sessions. The VAS comprises a 10-cm-long horizontal line anchored by verbal descriptors of no pain (score of 0) and worst pain imaginable (score of 10). The participants were asked to place a line perpendicular to the VAS at the point that represented the average pain during CHEP stimulation.

**Contact Heat Evoked Potentials**

CHEP stimulation was performed in a temperature-controlled room maintained between 20 and 24°C. A CHEP stimulator (PATHWAY, Sensory Analyzer System; Medoc Ltd., Ramat Yishai, Israel) was used to apply heat pulses at different body sites. The thermode had a diameter of 27 mm and an area of 572.5 mm². It was used to deliver contact heat stimuli by increasing the baseline temperature from 32°C to a peak temperature of 51°C at a rate of 70°C/s.

Evoked potentials were recorded and analyzed using the Keypoint EMG system, which has a sensitivity of 20 µV/div and a bandpass filter of 0.1–50 Hz. The electrodes were placed on Cz and Fz according to the International 10–20 system, and referenced to linked ears. Before stimulation, all participants received 2 stimuli per body site for familiarization. The participants were instructed to keep their eyes open in a fixed, neutral position to avoid blink artifacts during the test. The participants did not know when the CHEP stimuli would be given. To avoid habituation, the thermode was moved slightly after each stimulus, which restricted in a certain area (similar distance to the recording electrode). The interstimulus interval was set between 10 and 18 s. The researchers kept the
thermode in contact with the skin surface by using pressure. Two experienced researchers evaluated the recorded cortical responses and recordings with blink, muscle, or movement artifacts or other stimulation or recording interferences were eliminated. Impedance was kept below 5 kohm during the test. N2 latency, P2 latency, and N2–P2 amplitude at the four body sites were independently determined by two researchers. Disagreements between the two researchers were resolved through discussion. N2 latency/height and "P2 latency/height" (latency divided by height) (Lagerburg et al., 2015).

**Statistical Analysis**

Statistical analysis was performed using SPSS version 19.0 software (SPSS Inc., Chicago, IL, United States). A non-parametric test (Kruskal Wallis Test) was performed to compare CHEPs parameters (N2 latency, P2 latency, and N2–P2 amplitude) of different number of stimuli (3, 5, or 10). Logistic regression analysis was conducted to determine the relationship of CHEP parameters with age and gender, as well as their interaction. Normal values were established using the normal distribution method. The paired t-test and non-parametric test were performed to determine the effects of different baseline temperatures (32 vs. 35°C). Test-retest reproducibility was performed to assess the short- and long-term reproducibility of CHEP (Lagerburg et al., 2015). The data were expressed as mean ± standard deviation or median (interquartile range). A two-tailed P-value < 0.05 was considered statistically significant.

**RESULTS**

**Demographic Data**

A total of 151 participants (80 males and 71 females) were enrolled into this study and the mean age of the participants was 37 ± 14 years (range, 21–68 years) (Table 1). All of the volunteers were of Han Chinese ethnicity. The entire examination took approximately 90 min. The CHEP procedure was well tolerated by all of the subjects. In four participants, no N2–P2 amplitudes were elicited in the case of the right LE at a baseline temperature of 32°C, but were elicited when the baseline temperature was raised to 35°C.

The participants were stratified into five age groups: age 20–29 years (25 males, 21 females), 30–39 years (21 males, 16 females), 40–49 years (12 males, 12 females), 50–59 years (10 males, 12 females), and 60–69 years (12 males, 10 females). As

| Table 1 | Demographics, clinical and laboratory examinations of the participants. |
|---------|-------------------------------------------------|
| **Variable**                              | **151**                                                                 |
| Number of participants                     | 151                                                                   |
| Examination age (mean ± SD, year)          | 37 ± 14 (range, 21–68)                                           |
| Sex (male/female, n)                       | 80/71                                                                 |
| Height (cm)                                | 168.2 ± 9.4 (range, 156–185)                                       |
| Neurological examination                   | No sensory symptoms or signs                                       |
| Laboratory examination                     | -                                                                    |
| NCS                                         | -                                                                    |
| Median motor and sensory nerve             | -                                                                    |
| Ulnar motor and sensory nerve              | -                                                                    |
| Peroneal motor and sensory nerve           | -                                                                    |
| Sural motor and sensory nerve              | -                                                                    |

- negative.

| Table 2 | Results of contact heat evoked potential stimulation of the right forearm with three, five, and ten stimuli. |
|---------|---------------------------------------------------------------------------------------------------------------|
| Parameters | 3 stimuli | 5 stimuli | 10 stimuli | N | χ² | P     |
| NL/H     | 2.11 (0.22) | 2.10 (0.21) | 2.10 (0.21) | 69 | 7.81 | 0.056 |
| PL/H     | 2.79 (0.33) | 2.82 (0.35) | 2.81 (0.29) | 69 | 0.137 | 0.934 |
| N2–P2  | 46.00 (24.70) | 37.7 (26.90) | 29.7 (24.00) | 69 | 54.64 | <0.001 |

NL/H, N2 latency/height (ms/cm); PL/H, P2 latency/height (ms/cm); N2–P2 amplitude (µV); IQR, interquartile range; Values are expressed as Median (IQR).

| Table 3 | Influence of age, sex, and the interaction of sex*age on contact heat evoked potential parameters. |
|---------|-------------------------------------------------------------------------------------------------|
| Parameters | Site | Mean ± SD | N | Sex | Age | Sex*Age |
| NL/H     | FA     | 2.1 ± 0.27 | 151 | 0.176 | 0.114 | 0.017 |
| PL/H     | C7     | 1.87 ± 0.37 | 151 | 0.295 | 0.024 | 0.104 |
| NL/H     | T12    | 2.13 ± 0.22 | 151 | 0.752 | 0.002 | 0.785 |
| NL/H     | LE*    | 2.75 ± 0.41 | 147 | 0.374 | 0.033 | 0.203 |
| PL/H     | FA     | 2.72 ± 0.36 | 151 | 0.468 | 0.221 | 0.155 |
| PL/H     | C7     | 2.56 ± 0.38 | 151 | 0.37 | 0.582 | 0.166 |
| PL/H     | T12    | 2.8 ± 0.31 | 151 | 0.598 | 0.015 | 0.792 |
| PL/H     | LE*    | 3.39 ± 0.41 | 147 | 0.787 | 0.71 | 0.577 |
| N2–P2 amplitude | FA | 42.56 ± 18.41 | 151 | 0.626 | 0.117 | 0.441 |
| N2–P2 amplitude | C7 | 40.1 ± 17.25 | 151 | 0.903 | 0.17 | 0.975 |
| N2–P2 amplitude | T12 | 38.02 ± 18.06 | 151 | 0.328 | 0.057 | 0.153 |
| N2–P2 amplitude | LE* | 34.94 ± 21.69 | 147 | 0.064 | <0.001 | 0.053 |

NL/H, N2 latency/height (ms/cm); PL/H, P2 latency/height (ms/cm); FA, forearm; LE, leg. *In 4 patients, recordings could not be made at the leg site at a baseline temperature of 32°C.
N₂ and P₂ latency are related to height, the data were presented as “N₂ latency/height” and “P₂ latency/height” (Lagerburg et al., 2015). The average height of the participants was 168.2 ± 9.4 cm (range, 156–185 cm).

**Optimal Number of Stimuli**

A pilot study of 69 participants (39 males and 30 females) was performed involving CHEP stimulation of the right FA with a varying number of stimuli (3, 5, or 10). The mean age of the participants was 35 ± 12 years (range, 21–65 years). The median values of the CHEP parameters according to the number of stimuli is shown in Table 2. N₂ latency/height ($\chi^2 = 7.81$, $P = 0.056$) and P₂ latency/height ($\chi^2 = 0.137$, $P = 0.934$) did not significantly differ with the number of stimuli. However, N₂–P₂ amplitude ($\chi^2 = 54.64$, $P < 0.001$) significantly decreased with an increase in the number of stimuli (Figure 2).

**Effect of Baseline Temperature**

The comparison of CHEP parameters at the FA and VAS scores at different baseline temperatures is shown in Table 5. N₂ latency/height ($t = 5.45$, $P < 0.001$) and P₂ latency/height ($\chi^2 = -4.06$, $P < 0.001$) significantly decreased and N₂–P₂ amplitude ($t = -5.01$, $P < 0.001$) and VAS score ($\chi^2 = -5.84$, $P < 0.001$) significantly increased with an increase in baseline temperature (Figures 4–6).

**Reproducibility of Contact Heat Evoked Potential Parameters**

A comparison of CHEP parameters at the FA during different sessions is shown in Table 6. We found no significant differences in CHEP parameters at different time intervals (baseline vs. 30 min later vs. 1 year later). The test-retest reliability of the latency/height was good.

**DISCUSSION**

This research established normal values of CHEP parameters in a large sample of healthy Chinese adults. We found that N₂ latency/height was affected by both age and sex, while P₂ latency/height and N₂–P₂ amplitude were affected by age. Three was the optimal number of stimuli, and a higher baseline temperature (35°C) could be used to elicit responses if these
Normal values of contact heat evoked potential parameters in the Chinese population.

TABLE 4

| Parameters | 32°C | 35°C | N | t(2) | P  |
|------------|------|------|---|-----|----|
| NL/H       | 2.09 ± 0.19 | 1.95 ± 0.19 | 147 | 5.45 | <0.001 |
| PL/H       | 2.71 (0.43) | 2.49 (0.34) | 147 | −4.06 (χ²) | <0.001 |
| N₂–P₂      | 40.29 ± 18.84 | 53.01 ± 19.96 | 147 | −5.01 | <0.001 |
| VAS        | 5 (3) | 7 (3) | 147 | −5.84 (χ²) | <0.001 |

TABLE 5 | Contact heat evoked potential parameters for the right forearm and VAS scores at different baseline temperatures.

Parameters | 32°C | 35°C | N | t(2) | P |
|------------|------|------|---|-----|---|
| NL/H       | 2.09 ± 0.19 | 1.95 ± 0.19 | 147 | 5.45 | <0.001 |
| PL/H       | 2.71 (0.43) | 2.49 (0.34) | 147 | −4.06 (χ²) | <0.001 |
| N₂–P₂      | 40.29 ± 18.84 | 53.01 ± 19.96 | 147 | −5.01 | <0.001 |
| VAS        | 5 (3) | 7 (3) | 147 | −5.84 (χ²) | <0.001 |

NL/H, N₂ latency/height (ms/cm); PL/H, P₂ latency/height (ms/cm); N₂–P₂, N₂–P₂ amplitude (µV); VAS, visual analog scale; IQR, interquartile range. Values are expressed as mean ± SD or Median (IQR).

In this study, sex affected the value of N₂ latency/height at the FA, while age affected the values of N₂ latency/height at the FA, P₂ latency/height at T12, and N₂–P₂ amplitude at the LE. These findings are not completely consistent with those of previous studies (Chen et al., 2006; Lagerburg et al., 2015; Granovsky et al., 2016). A study of 35 Taiwanese subjects indicated that the values of CHEP parameters were not correlated with age, sex,
and body height (Chen et al., 2006); however, the sample size of this study was relatively small, and its conclusions may not be definitive. A Dutch study of 97 healthy controls revealed that N2 latency/height and P2 latency/height were significantly correlated with age, and N2–P2 amplitude was significantly correlated with age and sex (Lagerburg et al., 2015). The authors of this study proposed that the correlation of sex with N2 and P2 latency may be attributable to height differences between men and women. A recent multicenter study of 226 healthy subjects from Brazil, Israel, Japan, Spain, and America found that women had larger amplitudes and shorter latencies than men (Granovsky et al., 2016). However, body height was not an influencing factor in their study, and therefore, it is unclear whether the above differences in CHEP parameters were caused by sex-specific differences in body height. Moreover, in the above study, age had a significant influence only on CHEP parameters at the L1 and C7 sites, and sex had a significant influence only on P2 latency in the leg (Granovsky et al., 2016).

Thus, different studies have revealed conflicting relationships of CHEP parameters with age, sex, and height (Truini et al., 2005; Chen et al., 2006; Chao et al., 2007; Lagerburg et al., 2015; Granovsky et al., 2016) possibly due to differences in sample size, ethnicity, and parameter processing method. However, we conducted CHEP tests in 151 healthy Chinese subjects and established normal CHEP values for the Chinese population, which is vitally important in the diagnosis of SFN in China.

We found that N2–P2 amplitude varied greatly among subjects, and that N2 and P2 latency were more reliable parameters to evaluate small nerve fiber function. Different studies have used differing numbers of CHEP stimuli, from as few as 5 to as many as 20 stimuli (Atherton et al., 2007; Chao et al., 2007, 2008, 2010; Casanova-Molla et al., 2011; Haefeli et al., 2014; Lagerburg et al., 2015; Granovsky et al., 2016). In this study, we conducted a pilot study to determine the optimal number of stimuli (3, 5, or 10) at the FA, and found that N2 latency/height and P2 latency/height did not significantly differ with the number of stimuli. However, N2–P2 amplitude significantly decreased with an increase in the number of stimuli. Furthermore, 3 stimuli provided the same efficacy as 10 stimuli. Fewer numbers of stimuli were time-consuming, reduced the misery of participants during CHEP test and avoided across site habituation for CHEP test was applied on multiple sites. Therefore, we applied 3 stimuli at each body site in subsequent sessions to establish the normal values, which simplified the CHEP test and made it more efficient. In addition, blink and startle artifacts were carefully monitored and eliminated to ensure the efficacy of the test. The possible pitfalls of reduced stimuli include 3 stimuli may reduce the accuracy of the CHEP test if blink and startle artifacts were not carefully monitored, and larger sample test should be done to further confirm the efficacy of 3 stimuli.

In our study, N2 latency/height and P2 latency/height were significantly shorter at a baseline temperature of 35°C than at 32°C, and N2–P2 amplitude and VAS scores were significantly higher at the higher temperature value, which is consistent with the findings of other studies (Kramer et al., 2012a, 2013; Lagerburg et al., 2015). In four subjects, no recordings were obtained at the LE site at a baseline temperature of 32°C, but were obtained when the temperature was increased to 35°C. Similar observations have been reported in previous studies (Itskovich et al., 2000; Chen et al., 2006; Warbrick et al., 2009; Kramer et al., 2012a; Lagerburg et al., 2015).

Possible explanations for these findings include (1) the higher baseline temperature reduces the time required to reach the peak temperature of 51°C, leading to the activation of a greater number of nerve fibers, and (2) at the higher temperature, the ultra-late C-fiber response does not influence the late Aδ response (Truini et al., 2007; Lagerburg et al., 2015).

We also confirmed the short-term and long-term repeatability of CHEP testing by showing that CHEP parameters did not change significantly with time (baseline vs. 30 min later vs. 1 year later). Most studies have focused on the short-term reproducibility of CHEP stimulation, and found excellent reproducibility (Kramer et al., 2012b; Lagerburg et al., 2015), which is consistent with our findings. Ruscheweyh et al. (2013) examined the long-term (6 months) reproducibility of CHEP, and found that both amplitude and latency changed over time (Ruscheweyh et al., 2013). Possible reasons for this include seasonal differences in skin conductivity and psychological effects (Ruscheweyh et al., 2013). In our study, we included only healthy subjects and excluded seasonal effects by using a time interval of 1 year, and were able to confirm the short- and long-term reproducibility of CHEP.

The correlation of CHEP parameters with age, sex, and body height differs among different studies, and thus, studies with larger sample sizes are required to further investigate this issue.
CONCLUSION
We have established the normal values of CHEP parameters in a relatively large Chinese cohort, which facilitated the diagnosis of SFN in China. Only 3 stimuli were required to obtain reliable results on CHEP testing, and this has possibly simplified the examination technique. We found that CHEP parameters changed significantly with baseline temperature, but the short-term and long-term reproducibility of CHEP stimulation was excellent, which is useful in the diagnosis and follow-up of SFN.

DATA AVAILABILITY STATEMENT
The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

ETHICS STATEMENT
The studies involving human participants were reviewed and approved by Ethics Committee of Chinese PLA General Hospital. The patients/participants provided their written informed consent to participate in this study.

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AUTHOR CONTRIBUTIONS
BS and XH designed the study. BS and ZC conducted the study. BS, HW, and ZC collected and analyzed the data. HW, FC, and FY contributed samples collection and intellectual input. BS drafted and wrote the manuscript. XH revised the manuscript critically for intellectual content. All authors gave intellectual input to the study and approved the final version of the manuscript.

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