Mitochondrial mechanisms and therapeutics in ischaemia reperfusion injury

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Abstract
Acute kidney injury (AKI) remains a major problem in critically unwell children and young adults. Ischaemia reperfusion (IR) injury is a major contributor to the development of AKI in a significant proportion of cases and mitochondria are increasingly recognised as being central to this process through generation of a burst of reactive oxygen species early in reperfusion. Mitochondria have additionally been shown to have key roles in downstream processes including activation of the immune response, immunomodulation, and apoptosis and necrosis. The recognition of the central role of mitochondria in IR injury and an increased understanding of the pathophysiology that undermines these processes has resulted in identification of novel therapeutic targets and potential biomarkers. This review summarises a variety of therapeutic approaches that are currently under exploration and may have potential in ameliorating AKI in children in the future.

Keywords Acute kidney injury · Children · Ischaemia reperfusion injury · Mitochondria · Reactive oxygen species · Succinate

Acute kidney injury in paediatric nephrology
Acute kidney injury (AKI) remains a major problem in critically unwell children and young adults and is recognised as a major risk factor for the development of chronic kidney disease [1, 2]. The reported incidence of AKI varies between studies; a consequence in part of different case mix of institutions and variable definitions of AKI. In a multinational study, 26.9% of patients admitted to paediatric intensive care units were observed to have AKI. Importantly, in these children, AKI was an independent risk factor for morbidity and mortality, highlighting the urgent need for the development of effective therapies to prevent and treat AKI [3, 4]. Ischaemia reperfusion (IR) injury is a major contributor to the development of AKI in a significant proportion of children. Mitochondria are increasingly recognised to have a fundamental role in the pathogenesis of IR injury, and with the development of novel mitochondria-targeted therapies, there is increasing interest in the application of such therapies to the management of AKI [5].

Ischaemia reperfusion injury and reactive oxygen species
Reperfusion injury is the paradoxical, pathological exacerbation of tissue injury that occurs on re-oxygenation of an organ that has previously been subjected to a period of ischaemia [6]. While early research into this process focused on the heart [7–10], it is now increasingly recognised that the underlying pathophysiological process is common to a wide range of disorders, including AKI, stroke, intestinal ischaemia, multi-organ failure, hypovolaemic shock and organ dysfunction after transplantation [6].

It is well recognised that the dominant injurious effector upon reperfusion is an early burst of reactive oxygen species (ROS). Mitochondria are increasingly recognised as the key source of these ROS, through the generation of a burst of superoxide upon reperfusion, and these findings have been corroborated across a range of tissue types, including renal tissue [11, 12]. Whilst there are a number of alternative sources of superoxide, including the xanthine oxidase pathway and NADPH oxidases that are thought to be important in
renal IR injury [13], activation of these pathways seems to occur after, and be secondary to, the initial mitochondrial burst of superoxide formation [11, 14].

Mitochondrial generation of reactive oxygen species

Reactive oxygen species generation by mitochondria has long been known to occur during both physiological and pathological conditions. However, until recently, superoxide production during reperfusion was presumed to be the result of generalised dysregulation of the electron transport chain, with electrons leaking at multiple non-specific sites when oxygen was re-introduced to a system in biochemical disarray following a period of ischaemia [11].

Contrary to this view, Chouchani et al. recently identified a specific metabolic pathway in which superoxide was generated through reverse electron transport at complex I of the electron transport chain. Moreover, this process was shown to be driven by the pool of the citric acid cycle metabolite, succinate, that accumulates during ischaemia (Fig. 1) [15].

This burst of mitochondrial superoxide leads to the activation of a plethora of pathways that cause tissue injury. Direct cellular or mitochondrial damage through lipid peroxidation or protein carbonylation results in disruption of adenosine triphosphate (ATP) generation, dysregulation of calcium levels and induction of the mitochondrial permeability transition pore (MPTP) with subsequent activation of necrosis and apoptosis. Tissue damage can also occur indirectly through activation of the innate and adaptive immune response by damage-associated molecular patterns (DAMPs). These DAMPs can be cellular (e.g. high mobility group box 1 (HMGB1), hyaluronan [16]) or directly mitochondrial in origin. Mitochondrial ROS, generated by IR injury, have been shown to act directly as a DAMP. Furthermore, opening of the MPTP in response to mitochondrial dysfunction releases other mitochondrial DAMPs [17, 18] such as mtDNA, cytochrome c, succinate and N-formyl peptides [19, 20]. These processes provide a putative mechanism for the impact of mitochondrial dysfunction on longer-term renal function after an episode of AKI. The identification of a unifying, specific pathway also provides a potential explanation for the vast array of interventions and therapies that have previously been shown to ameliorate IR injury [11].

Mitochondrial therapeutic strategies in IR injury

The recent progress in our understanding of the pathophysiological mitochondrial mechanisms that underpin IR injury has led to an array of potential applications of mitochondria as both targets for therapeutic strategies and as biomarkers of disease severity. The therapeutic strategies can be grouped into the following areas and are discussed in more detail below: limiting oxidative stress and mitochondrial ROS generation, reducing tubular cell death through necrosis and apoptosis, moderating mitochondrial dynamics and mitochondrial immunomodulation.

Mitochondria oxidative stress

The fundamental role of mitochondrial oxidative stress in a wide range of pathologies including IR injury has been extensively reported in the literature and has provided a strong rationale for the use of antioxidants as a therapeutic intervention. Unfortunately, the clinical translation of non-specific antioxidants has been almost universally disappointing. This paradox can be interpreted in one of two ways. Either reactive oxygen species do not have a role in the pathophysiology of these diseases or the antioxidants are not adequately delivered to the appropriate region of the cell to prevent oxidative damage. This second argument is supported by the increasing recognition of the important physiological roles of ROS within the cell and the recognition of the integral role of the burst of mitochondrial ROS in mediating IR injury. Targeting antioxidants to mitochondria, therefore, provides an approach that could both explain this paradox and provide a novel therapeutic strategy in IR injury [5, 21].

Bioactive molecules and drugs, including antioxidants, have been targeted to mitochondria in vivo using both lipophilic cations and mitochondrial targeted peptides (reviewed in detail elsewhere [21]). Both these approaches lead to a rapid and significant accumulation of the targeted compound within the mitochondria. This approach increases potency whilst enabling a lower dose to be administered, minimising off-target effects and toxicity. Triphenylphosphonium (TPP) is a lipophilic cation that is rapidly taken up into mitochondria and concentrated several hundred-fold due to the large mitochondrial membrane potential in vivo. Covalent linkage of bioactive molecules or drugs to TPP has been used for a wide range of compounds [21–24]. The most extensively investigated of these is MitoQ. The bioactive molecule of MitoQ is ubiquinone. This is rapidly reduced in mitochondria to the chain-breaking antioxidant ubiquinol, which directly scavenges mitochondrial ROS, inhibiting downstream lipid peroxidation and mitochondrial damage. MitoQ has been shown to protect against oxidative injury in a variety of animal models and has been used in phase II human trials [25, 26]. In models of renal IR injury, MitoQ has recently been demonstrated to reduce markers of oxidative injury, renal function and tissue injury following IR injury [27–29].

Another approach is through the use of peptide delivery systems including the Szeto-Schiller (SS) peptides and the mitochondrial penetrating peptides. The precise mechanism of action of these molecules is not understood, but they have
been proposed to protect mitochondria by interacting with cardiolipin [30]. SS peptides have also been shown to ameliorate renal IR injury in rodents and have been studied in other models of IR pathologies [31]. The lead compound in this group, SS-31, has been investigated in larger animal models and is currently the subject of a human clinical trial investigating its efficacy in ameliorating IR injury post-angioplasty for renal artery stenosis [32].

The recognition of a specific metabolic pathway that drives mitochondrial ROS production during IR injury opens up the possibility of a novel therapeutic strategy that acts upstream of ROS generation [15], namely competitive inhibition of succinate dehydrogenase, which has been shown to ameliorate IR injury in a variety of in vivo models [15, 33]. The metabolic signature of ischaemic succinate accumulation has been demonstrated in a wide range of tissues including human myocardial [34] and renal tissue [35]. Contrary to these findings, some authors have questioned the translation of these findings in small animals to human tissues [36]. An inter-species difference in mechanisms of mitochondrial ROS generation is unlikely, given the very early evolutionary origin of mitochondria, and it is essential to rule out differences in experimental methodology as a source of conflicting data. Therefore, preventing succinate accumulation by inhibiting succinate dehydrogenase activity remains a potentially important, but as yet unexplored, area of therapy in renal IR injury. Furthermore, the demonstrated therapeutic potential in other models of IR injury make it an appealing mechanism that warrants future investigation (Fig. 2).

**Tubular death through apoptosis and necrosis**

Mitochondria are recognised to be integral to the processes of necrosis and apoptosis which underlie tubular injury and cell death following IR injury [12]. In mammalian cells, apoptosis is initiated through two major but interconnected pathways: death receptors and mitochondria. The mitochondrial pathway is characterised by an increase in the permeability of the outer...
mitochondrial membrane (MOMP) with the release of pro-apoptotic factors such as cytochrome c. The B-cell lymphoma 2 (Bcl-2) family proteins are important regulators of MOMP, both in a positive and negative capacity [37]. The release of cytochrome c, and other proteins, from the intermembrane space triggers the formation of the apoptosome which consists of cytochrome c, apaf-1 and caspase-9. This then activates downstream caspase-activation pathways resulting in apoptosis. However, despite the potential promise of therapies designed to inhibit apoptosis in AKI, it has yet to be realised in routine clinical practice [38]. A promising approach that is currently in phase II human clinical trials to treat AKI is the use of a small interfering RNA (siRNA) that temporarily inhibits expression of the stress response gene p53 [39, 40]. Other approaches that target the same pathway include disrupting steps in the apoptotic pathway using small molecules, caspase inhibitors and recombinant proteins [12].

Mitochondrial biogenesis, mitophagy and dynamics

Mitochondria are highly dynamic organelles existing not as solitary, isolated entities but as a complex, interconnected network that undergoes continuous biogenesis, fusion, fission and the selective removal by autophagy, termed mitophagy. These processes are all essential for normal mitochondrial and cellular function [41, 42] and have been shown to be implicated in AKI.

Altered mitochondrial dynamics contributes to changes in mitochondrial energetics, cellular injury and repair following AKI [42]. A variety of mammalian proteins have been identified as regulators of mitochondrial fission and fusion including the pro-fusion proteins, mitofusin 1 and 2, and OPA1, and the pro-fission protein dynamin-related protein1 (DRP1) [5, 43, 44]. The activation of DRP1 results in the translocation of DRP1 to the outer mitochondrial membrane promoting mitochondrial fission and exacerbating AKI. Pharmacological inhibition of DRP1 in a mouse model of AKI reduces mitochondrial fission and ameliorates AKI in vivo [45]. Sirtuin 3 (SIRT3) has also been shown to have a functional role in mitochondrial dynamics, preserving mitochondrial integrity by preventing DRP1 translocation. SIRT3 upregulation was shown to be protective in vitro in human proximal tubular epithelial cells damaged by cisplatin. Furthermore, in a murine model of AKI, upregulation of SIRT3 resulted in a decrease in mitochondrial fission whilst SIRT3 deficiency in the Sirt3−/− mice exacerbated cisplatin-induced AKI [46].

In the murine kidney, mitophagy has been shown to be highly active [47, 48], with an integral role in moderating...
tissue injury in the kidney. Ablation of key genes that regulate autophagy, such as autophagy-related protein 7 (ATG7) and ATG5, has been shown to exacerbate AKI in vivo [48, 49]. There is also evidence that there is crosstalk between the cell death machinery and mitophagy. Deletion of the pro-apoptotic protein BAK has been shown to be renoprotective in models of IR injury. This renoprotective effect was associated with a decrease in the release of cytochrome c and mitochondrial fragmentation [37].

Following AKI, the resolution of kidney injury and return of function is primarily through restoration of cellular function rather than regeneration and cell proliferation. Therefore, moderating mitochondrial biogenesis may provide another therapeutic avenue in AKI [42, 50]. Peroxisome proliferator-activated receptor-γ coactivator-1α (PGC-1α) has been identified as a key regulator of mitochondrial biogenesis and PGC-1α knock-out mice have been shown to be more susceptible to kidney injury [51]. More recently, sirtuins have also been shown to have regulatory roles in mitochondria biogenesis, and the SIRT1 activator SRT1720 has been shown to augment mitochondrial recovery and tubular function in the rat in vivo following IR injury [52]. There is also emerging evidence that the β2-antagonist formoterol has effects as an activator of mitochondrial biogenesis and can enhance recovery of mitochondria and kidney function following IR injury [5, 53].

**Mitochondrial immunomodulation**

Mitochondria are increasingly being recognised as having critical roles in activating and moderating the immune system through a range of pathways [17, 54, 55]. In adult kidney transplantation, there is evidence to suggest that prolonged cold ischaemia impacts on long-term graft survival. Despite significant improvements in short-term outcomes, the

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**Fig. 3** The role of mtDNA in activating the innate immune response. Ischaemic damage-induced loss of membrane potential and mitochondrial swelling can lead to opening of a non-selective permeability transition pore (MPTP) in the mitochondrial inner membrane that releases mitochondrial molecules, like cytochrome c, ATP, ROS, N-formyl peptides and mtDNA, into the cytosol. Released mitochondrial components can act as mitochondrial damage associated molecular patterns (mtDAMPs). When released, they function as signals for injury in cells and activate the innate immune response. MtDNA, due to its similarity to bacterial DNA, can activate the TLR9 dependent immune response and in turn stimulate activation of the transcription factor NF-κB and therefore expression of the cytokine IL-6, which is released from the cell and functions as a stimulus for immune cells. In addition, mtDNA is involved in the activation of the NLRP3 inflammasome that senses cytosolic DNA and stimulates the caspase-1-dependent release of IL-1β and IL-18. Furthermore, the ER-linked STING pathway can be activated inducing the expression of interferon type 1 and its inflammatory signalling pathways.
prevalence of chronic allograft dysfunction remains largely unchanged [56]. The emerging evidence that mitochondria are not only integral to IR injury but also have fundamental roles in the immune response suggests there may be new avenues to improve long-term graft outcomes. Mitochondria, for example, are thought to be involved in regulation of conversion of M1 inflammatory macrophages to M2 anti-inflammatory cells. Both metformin and rotenone have been shown to facilitate this switch, possibly mediated through actions on complex I by reducing reverse electron transport and ROS [17, 57, 58]. Furthermore, the inhibition of ROS generation by inhibiting SDH has also been shown to limit pro-inflammatory responses and boost anti-inflammatory responses [58, 59].

Recently, it has also been suggested that maladaptive repair following AKI may be responsible for the progression of renal disease and development of chronic kidney disease in affected individuals [60]. Interestingly, the cellular changes underlying maladaptive repair, most notably cellular senescence in tubular epithelial cells and adoption of a pro-fibrotic phenotype, mimic those of kidney ageing [60, 61].

Mitochondrial DNA release as a biomarker of kidney injury

The damage to mitochondria associated with IR injury results in the opening of the MPTP with the consequent release of a number of mitochondrial components into the cytosol. These function as DAMPs, activating an innate immune response and driving the systemic inflammatory response associated with IR injury. mtDNA is an example of such a DAMP (Fig. 3). A number of studies have examined the role of circulating mtDNA in the blood of patients. The levels of mtDNA have been shown to increase in the circulation following trauma [18] and more recently have been shown to correlate with mortality in intensive care unit patients [62]. It has also been investigated in the clinical context of severe lung injury and AKI, and shows promise as a potential plasma biomarker [27, 62–67]. Increased plasma mtDNA levels can be found in murine models of AKI and have been shown to be linked to an elevated innate immune response via activation of the TLR9 pathway [68]. In addition to its detection in the circulation, mtDNA can also be detected in the urine and levels have been shown to be significantly increased in the urine of patients after AKI [69–72]. The recognition that the kidney has a role in the clearance of DAMPs from plasma and the efficacy of dialysis in decreasing the amount of circulating mtDNA in patients may have future implications for our understanding of the pathogenesis of IR injury in AKI and the role of dialysis in managing IR injury [73–75].

Summary

AKI remains a major problem in children and IR injury is either the primary aetiology or is implicated in a significant proportion of cases. Mitochondrial dysfunction or damage are increasingly recognised as fundamental to IR injury, generating the burst of ROS that initiates downstream tissue injury. They also have key roles in a variety of downstream processes, including the direct activation of the innate immune response, immunomodulation, and apoptosis and necrosis. Plasma and urinary mtDNA may have roles as biomarkers of IR injury in the future and there are a number of therapeutic strategies that are being explored to ameliorate mitochondrial dysfunction. We look forward with interest to the translation of these promising strategies to the management of AKI in children in the future.

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Compliance with ethical standards

Conflicts of interest M.P.M. holds patents in the area of mitochondrial therapies and has a financial interest in a company Antipodean Inc. that is commercialising mitochondrial therapies. J.L.M., A.V.G., T.E.B. and K.S.P. have no conflicts of interest.

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