Possible role of intestinal stem cells in the pathophysiology of irritable bowel syndrome

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Abstract

The pathophysiology of irritable bowel syndrome (IBS) is not completely understood. However, several factors are known to play a role in pathophysiology of IBS such as genetics, diet, gut microbiota, gut endocrine cells, stress and low-grade inflammation. Understanding the pathophysiology of IBS may open the way for new treatment approaches. Low density of intestinal stem cells and low differentiation toward enteroendocrine cells has been reported recently in patients with IBS. These abnormalities are believed to be the cause of the low density of enteroendocrine cells seen in patients with IBS. Enteroendocrine cells regulate gastrointestinal motility, secretion, absorption and visceral sensitivity. Gastrointestinal dysmotility, abnormal absorption/secretion and visceral hypersensitivity are all seen in patients with IBS and haven been attributed to the low density the intestinal enteroendocrine cells in these patients. The present review conducted a literature search in Medline (PubMed) covering the last ten years until November 2019, where articles in English were included. Articles about the intestinal stem cells and their possible role in the pathophysiology of IBS are discussed in the present review. The present review discusses the assumption that intestinal stem cells play a central role in the pathophysiology of IBS and that the other factors known to contribute to the pathophysiology of IBS such as genetics, diet gut microbiota, stress, and low-grade inflammation exert their effects through affecting the intestinal stem cells. It reports further the data that support this assumption on genetics, diet, gut microbiota, stress with depletion of glutamine, and inflammation.

Key words: Diet; Gut enteroendocrine cells; Gut microbiota; Low grade inflammation; Stress

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Core tip: The pathophysiology of irritable bowel syndrome (IBS) is not completely understood. Understanding the pathophysiology of IBS may enable us to find an effective treatment for this disorder. The density of intestinal stem cells is low in patients with IBS. Moreover, the differentiation of stem cells into enteroendocrine cells is abnormal. It seems that these abnormalities in intestinal stem cells is the cause of the low density of enteroendocrine cells seen in patients with IBS. It is believed that the low density of enteroendocrine cells is behind the gastrointestinal dysmotility, abnormal secretion/absorption and hypersensitivity observed in patients with IBS. This review presents the observations that suggest that the factors known to contribute to the pathophysiology of IBS may exert their effects through affecting the intestinal stem cells.

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INTRODUCTION

Irritable bowel syndrome (IBS) is a wide spread condition affecting 12.1% of the world population[1,2]. The prevalence of IBS differs considerably between different parts of the world with the lowest prevalence in Asia and the highest in South America[1]. The cardinal symptom of IBS is intermittent abdominal pain accompanied by altered bowel habits and abdominal bloating/distention[3]. There is no biochemical, radiological or clinical test/examination for diagnosing IBS and the IBS diagnosis is based on symptoms assessment[4]. IBS reduces significantly the patients’ quality of life in the same degree as major chronic diseases such heart failure, renal failure, diabetes, and inflammatory bowel disease[5-7]. It has been reported that 12%-14% of primary care patient visit, and 28% of referrals to gastroenterologists are IBS patients[5-7] and consequently IBS patients are more common in the healthcare than patients with diabetes, hypertension or asthma[8,9]. There is no effective treatment for IBS and the available treatment in clinic is directed to symptom relief[4].

Several factors are known to play pivot role in pathophysiology of IBS. These factors are genetics, diet, gut microbiota, gut endocrine cells, stress and low-grade inflammation[2,10]. Abnormalities in the intestinal stem cells has been reported recently[11-13]. The present review aimed at discussing the possibility that the factors known to contribute to the pathophysiology of IBS may exert their effects through affecting the intestinal stem cells. The present review conducted a literature search in MEDLINE (PubMed) covering the last ten years until November 2019, where articles in English were included. Articles about the intestinal stem cells and their possible role in the pathophysiology of IBS are discussed.

FCACORS INVOLVED IN THE PATHOPHYSIOLOGY OF IBS

Genetics

Studies of family history and family cluster as well as twin studies provided strong evidences that IBS is hereditary[14-21]. However, the possible mutant gene(s) responsible for IBS is/are not found yet[4].

Diet

Patients with IBS avoid certain food items as they believe they worsen/trigger their symptoms[22-26]. However, there is no difference in intake of calories, or the meal patterns between IBS patients and community controls[23,27,28]. The effect of diet on IBS symptoms cannot be explained by food allergy/intolerance[29]. However, it is generally accepted that poorly absorbed carbohydrates and fibers play an important role in development IBS symptoms[29,30]. The intake of low fermentable oligo-, di-, monosaccharides and polyols-diet and National Institute for Health and Care Excellence-modified diet improve both symptoms and quality of life in IBS patients[22,29,31,32]. However, a recent review and meta-analysis showed that there is very low quality evidence showing that low fermentable oligo-, di-,...
monosaccharides and polyols diet reliefs IBS symptoms\(^3\).

Based on a case report published in 1978, non-celiac gluten sensitivity was coined\(^{34-36}\). In this case, a patient without celiac disease, suffered from abdominal pain and diarrhea who experienced symptoms improved when she used gluten-free diet. Several studies showed that withdrawal of wheat products in patients with non-celiac IBS-like symptoms improve these symptoms\(^{37-42}\). However, a double-blind placebo-controlled study showed that it is fructan in the wheat rather than gluten that trigger IBS symptoms\(^{43}\). In a recently published meta-analysis concluded that there is insufficient evidence that gluten-free diet improves IBS symptoms\(^3\).

Gut microbiota

The gastrointestinal microbiota comprises 12 different bacteria phyla, but most of the gut bacteria belongs to the Proteobacteria, Firmicutes, Actinobacteria and Bacteroidetes\(^{44,45}\). The anaerobic Firmicutes and Bacteroidetes phyla dominate the bacterial population in the intestinal of healthy adults, with a few members from of the Proteobacteria and Actinobacteria phyla\(^{46,47}\). A low microbial diversity in the gut (dysbiosis) has been reported to be associated with several diseases\(^{47,48}\).

In healthy subjects, the intestinal microbiota composition is affected by the individual genetic composition and environmental factors one is exposed for\(^{44,48}\). The intestinal microbiota in IBS patients differs from that of healthy subjects\(^{48-51}\), and have a lower diversity (dysbiosis)\(^{48-51}\). It is believed that this difference in the intestinal microbiota plays a pivot role in the pathophysiology of IBS\(^{49}\).

Gastrointestinal endocrine cells

The gastrointestinal endocrine cells are scattered in-between the epithelial cells facing the gut lumen (Figure 1)\(^{52-54}\). These cells are localized to the stomach, small- and large intestine\(^{53}\). Among the different segments of the gastrointestinal tract the density of the endocrine cells is highest in the duodenum (Figure 2)\(^{55}\). These cells secret over 10 different hormones that interact and integrate with the enteric, autonomic and central nervous system to regulate: Gastrointestinal motility, secretion of enzymes and bile acid, absorption of nutrients, visceral sensation, gastrointestinal cell proliferation, local immune defense and appetite\(^3,22,52,55-69\). These cells have sensory microvilli that project into the gastrointestinal lumen that sense gastrointestinal lumen contents and respond by releasing their hormones into the lamina propria\(^{70-82}\). These hormones can act locally on the nearby structures (paracrine mode of action) or reach the blood stream and act on far structure (endocrine mode of action)\(^{70-82}\).

Several abnormalities in different endocrine cell types of the stomach, small- and large intestine have been described in IBS patients (Figure 3)\(^{53,83-97}\). Generally, IBS patients have a lower gut endocrine cell density than healthy subjects\(^{52}\).

Stress

Stress is defined as an acute threat, real or perceived, to the homeostasis of an organism\(^{10}\). Stress is a known factor that trigger/worsen the IBS symptoms\(^9,98\). The exact mechanisms by which stress affects IBS are not exactly known. However, the negative effect of stress on IBS symptoms is believed to be caused by an interaction between the gut and the central nervous system (gut-brain axis)\(^{10}\).

Low grade inflammation

Intestinal low-grade inflammation is believed to be a factor that contribute to the pathophysiology of IBS\(^8\). Low grade inflammation in the intestinal mucosa occurs only in a subset of IBS, i.e., post-infectious IBS, but not in sporadic (non-specific) IBS\(^{86,91-102}\).

THE ROLE OF INTESTINAL STEM CELLS IN THE PATHOPHYSIOLOGY OF IBS

Intestinal stem cells

Each intestinal crypt contains four to six pluripotent (stem) cells\(^{103}\). Stem cell perform 2 activities, namely self-renewal by dividing into identical stem cell (clonogeny) to maintain a constant number of stem cells and differentiation progeny\(^{103}\). In the differentiation progeny, the stem cells differentiate into all cell types of the villus epithelium through 2 cell lineages: The secretory lineage giving raise to goblet cells, endocrine cells and Paneth cells, and the absorptive lineage giving raise to absorptive enteroendocrine cells. This differentiation takes place through a series of precursors (progenitors) (Figure 4)\(^{68,69,104-122}\).
The relation between the abnormalities in intestinal stem cells and enteroendocrine cells

As mentioned previously, the densities of the gastrointestinal endocrine cells are lower than that of healthy subjects\cite{53,83-97}. The cell density of Musashi 1, and neurogenin 3 immunoreactive cells in the small and large intestine of patients with IBS are lower than that of healthy subjects (Figures 5 and 6)\cite{11-13}. Musashi 1 is marker for intestinal stem cells and their early progeny, and neurogenin 3 is expressed in early intestinal endocrine cell progenitors originated from stem cells\cite{103,113-118}. The low densities of enteroendocrine cells in patients with IBS could be explained by the abnormalities in intestinal stem cells\cite{119}. Thus, low densities of Msi-1 and NEUROG3 small and large intestine in IBS patients indicate that the intestinal stem cells in these patients exhibit reduced clonogenic activity and low differentiation progeny toward endocrine cells\cite{119,120}.

HYPOTHESIS

Based on the data presented above, one may hypothesized that IBS patients may have a gene mutation controlling the number of the stem cells and/or NEUROG3 gene mutation. Furthermore, environmental factors such as diet, inflammation, stress and gut microbiota may affect the stem cells and their progeny (Figure 7). This hypothesis gets support from the following facts: (1) Low density of intestinal endocrine cells has been described in patients with congenital malabsorptive diarrhoea, which is an autosomal recessive disorder\cite{121}. The low density of intestinal endocrine cells in this disorder is caused by loss-of-function mutations in NEUROG3 gene\cite{121}. Similarly, low density of intestinal endocrine cells has been observed in small-intestine allograft rejection, and in NEUROG3-knockout mice\cite{117,120}. The low density of intestinal endocrine cells in these conditions was associated with a reduction in number of intestinal neurogenin 3 cells\cite{117,120}; (2) Changing from the common Norwegian diet to a National Institute for Health and Care Excellence-modified diet, which improved symptoms and quality of life in IBS patients is associated with changes in the densities of gastrointestinal cells\cite{91,122-127}; (3) Modulation of the intestinal microbiota by fecal microbiota transplantation improved both symptoms and the quality of life in patients with IBS\cite{128}. This improvement was accompanied by a change in in the densities of enteroendocrine cells\cite{128}; (4) Glutamine is the main energy source for intestinal enterocytes and plays a major role in intestinal homeostasis and other physiological functions\cite{129-133}. Stress, infection or inflammation cause a depletion of...
In a randomized placebo-controlled study, dietary glutamine supplements improved symptoms in patients with post-infectious IBS\(^ {134} \). Glutamine have a trophic effect on the intestinal stem cells and promotes stem cell differentiation\(^ {129,135,136} \). One may speculate that stress results in the depletion of glutamine, which causes disturbance in the differentiation of the intestinal cells. This in turn would cause low density in enteroendocrine cells and the development of IBS symptoms; and (5) In animal models of human ulcerative colitis and Crohn’s disease, the changes in enteroendocrine cells have been found to be strongly correlated with changes in the intestinal stem cells and their differentiation progeny toward intestinal endocrine cells\(^ {137,138} \).

**CLINICAL IMPLICATIONS**

Assuming that IBS is caused by abnormalities in stem cells, which in turn caused by genetic and environmental factors, intestinal stem cell transplantation might be an effective tool in the treatment of IBS.

**CONCLUSION**

The intestinal stem cells appear to play a central role in the pathophysiology of IBS. Factors that are known to be involved in the pathophysiology of IBS exert their effects probably through affecting the intestinal stem cells.
Figure 3 Chromogranin immunoreactive cells in the duodenum of a healthy subject and of a patient with irritable bowel syndrome. A: Chromogranin immunoreactive cells in the duodenum of a healthy subject; B: Chromogranin immunoreactive cells in the duodenum of a patient with irritable bowel syndrome (IBS); C: Chromogranin A cells in the colon of a healthy control; D: Chromogranin A cells in the colon of a patient with IBS. Chromogranin A is a common marker for enteroendocrine cells. The density of Chromogranin A in the duodenum and colon of patients with IBS is lower than that of healthy subjects.

Figure 4 The intestinal stem cell divides into 2 identical cells (clonogenic). One of these cells remains inactive, while the other stem cell differentiates into all cell types of the villus epithelium through 2 cell lineages. The secretory lineage giving rise to goblet cells, entodocrine cells and Paneth cells, and the absorptive lineage giving rise to absorptive enterocytes. This differentiation occurs through a series of progenitors. The observations that the density of stem cells and progenitors for enteroendocrine cells, led to the assumption of gene mutations affecting the stem cell and neurogenin 3 gene.
Figure 5 Musashi 1 immunoreactive cells in duodenum. A: A healthy subject; B: A patient with irritable bowel syndrome (IBS). Musashi 1 is a marker for intestinal stem cells and their early progeny. The density of Musashi 1 cells in healthy subjects is higher than that of the IBS patients. Furthermore, Musashi 1 cells in healthy subjects appear to have more proliferation activity than that of IBS patients.

Figure 6 Neurogenin 3 immunoreactive cells in the duodenum. A: A healthy control; B: A patient with irritable bowel syndrome. Neurogenin 3 is expressed in the nuclei (brown) of early intestinal endocrine cell progenitors. The healthy subjects have higher density of neurogenin 3 than irritable bowel syndrome patients.

Figure 7 Schematic drawing to illustrate the hypothesis that intestinal stem cells play a central role in the pathophysiology of irritable bowel syndrome. The abnormalities in the intestinal stem cells can be caused by gene mutation or by environmental factors such as diet, intestinal microbiota, stress and low-grade inflammation.
El-Salhy M. Stem cells in IBS

REFERENCES

1. Canavan C, West J, Card T. The epidemiology of irritable bowel syndrome. Clin Epidemiol 2014; 6: 71-80 [PMID: 24523597 DOI: 10.2147/CLEP.S40245]

2. El-Salhy M. Recent developments in the pathophysiology of irritable bowel syndrome. World J Gastroenterol 2015; 21: 7621-7636 [PMID: 26167062 DOI: 10.3748/wjg.v21.i25.7621]

3. El-Salhy M. Irritable bowel syndrome: diagnosis and pathogenesis. World J Gastroenterol 2012; 18: 5151-5163 [PMID: 23066308 DOI: 10.3748/wjg.v18.i37.5151]

4. El-Salhy M, Gilja OH, Hatlebakk JG. Overlapping of irritable bowel syndrome with erosive esophagitis and the performance of Rome criteria in diagnosing IBS in a clinical setting. Mol Med Rep 2019; 20: 787-794 [PMID: 31180516 DOI: 10.3892/mmr.2019.10284]

5. Talley NJ, Gabriel SE, Harmsen WS, Zinsmeister AR, Evans RW. Medical costs in community subjects with irritable bowel syndrome. Gastroenterology 1995; 109: 1376-1371 [PMID: 7498636 DOI: 10.1016/0016-5085(95)90738-6]

6. Jones R, Lydeard S. Irritable bowel syndrome in the general population. BMJ 1992; 304: 87-90 [PMID: 1737146 DOI: 10.1136/bmj.304.6819.87]

7. Hungin AP, Whorwell PJ, Tack J, Mearin F. The prevalence, patterns and impact of irritable bowel syndrome: an international survey of 40,000 subjects. Aliment Pharmacol Ther 2003; 17: 643-650 [PMID: 12641512 DOI: 10.1046/j.1465-236X.2003.01456.x]

8. Schuster MM. Defining and diagnosing irritable bowel syndrome. Am J Manag Care 2001; 7: S246-S251 [PMID: 11474699]

9. Mitchell CM, Drossman DA. Survey of the AGA membership relating to patients with functional gastrointestinal disorders. Gastroenterology 1987; 92: 1282-1284 [PMID: 3557021 DOI: 10.1016/s0016-5085(87)91099-7]

10. Kouturek PC, Brzozowski T, Kouturek SJ. Stress and the gut: pathophysiology, clinical consequences, diagnostic approach and treatment options. J Physiol Pharmacol 2011; 62: 591-599 [PMID: 23314561]

11. El-Salhy M, Gilja OH. Abnormalities in ileal stem, neurogenin 3, and enteroendocrine cells in patients with irritable bowel syndrome. BMJ Gastroenterol 2017; 17: 90 [PMID: 28764761 DOI: 10.1186/s12876-017-0643-4]

12. El-Salhy M, Hatlebakk JG, Hausken T. Reduction in duodenal endocrine cells in irritable bowel syndrome is associated with stem cell abnormalities. World J Gastroenterol 2015; 21: 9577-9587 [PMID: 26327765 DOI: 10.3748/wjg.v21.i32.9577]

13. El-Salhy M, Patcharatrakul T, Hatlebakk JG, Hausken T, Gilja OH, Gonlachanvit S. Enteroendocrine, Musashi 1 and neurogenin 3 cells in the large intestine of Thai and Norwegian patients with irritable bowel syndrome. Scand J Gastroenterol 2017; 52: 1331-1339 [PMID: 28853308 DOI: 10.1080/00365521.2017.1371793]

14. Locke GR, Zinsmeister AR, Talley NJ, Fett SL, Melton LJ. Familial association in adults with functional gastrointestinal disorders. Mayo Clin Proc 2000; 75: 907-912 [PMID: 10994826 DOI: 10.4065/75.9.907]

15. Kalantar JS, Locke GR, Zinsmeister AR, Beiglely CM, Talley NJ. Familial aggregation of irritable bowel syndrome: a prospective study. Gut 2003; 52: 1703-1707 [PMID: 14633946 DOI: 10.1136/gut.52.12.1703]

16. Kanazawa M, Endo Y, Whitehead WE, Kano M, Hongo M, Fukudo S. Patients and nonconsultors with irritable bowel syndrome reporting a parental history of bowel problems have more impaired psychological distress. Dig Dis Sci 2004; 49: 1046-1053 [PMID: 15309909 DOI: 10.1023/b:ddas.0000034570.52305.10]

17. Morris-Yates A, Talley NJ, Boyce PM, Nandurkar S, Andrews G. Evidence of a genetic contribution to functional bowel disease. Am J Gastroenterol 1998; 93: 1311-1317 [PMID: 9707057 DOI: 10.1111.j.1572-0241.1998.440_x.x]

18. Levy RI, Jones KR, Whitehead WE, Feld SI, Talley NJ, Corey LA. Irritable bowel syndrome in twins: heredity and social learning both contribute to etiology. Gastroenterology 2001; 121: 799-804 [PMID: 11606493 DOI: 10.1053/gast.2001.279095]

19. Lemoa A, Zaman M, Jones M, Talley NJ. Influence of genetics on irritable bowel syndrome, gastrooesophageal reflux and dyspepsia: a twin study. Aliment Pharmacol Ther 2007; 25: 1343-1350 [PMID: 17509102 DOI: 10.1111/j.1365-2036.2007.03526.x]

20. Wojcieszyn MK, North KE, Pedersen NL, Sullivan PF. Irritable bowel syndrome: a co-twin control analysis. Am J Gastroenterol 2007; 102: 2220-2229 [PMID: 17897337 DOI: 10.1111.j.1572-0241.2007.01479.x]

21. Bengtson MB, Roming T, Vatn MH, Harris JR. Irritable bowel syndrome in twins: genes and environment. Gut 2006; 55: 1754-1759 [PMID: 17008366 DOI: 10.1136/gut.2006.097287]

22. El-Salhy M, Ostgaard H, Gundersen D, Hatlebakk JG, Hausken T. The role of diet in the pathogenesis and management of irritable bowel syndrome (Review). Int J Mol Med 2012; 29: 723-731 [PMID: 22367773 DOI: 10.3892/ijmm.2012.926]

23. Ostgaard H, Hausken T, Gundersen D, El-Salhy M. Diet and effects of diet management on quality of life and symptoms in patients with irritable bowel syndrome. Mol Med Rep 2012; 5: 1382-1390 [PMID: 22446969 DOI: 10.3892/mmr.2012.813]

24. El-Salhy M, Gilja OH, Gundersen D, Hatlebakk JG, Hausken T. Interaction between ingested nutrients and gut endocrine cells in patients with irritable bowel syndrome (review). Int J Mol Med 2014; 34: 363-371 [PMID: 24953595 DOI: 10.3892/ijmm.2014.1811]

25. El-Salhy M, Mazzawi T, Hausken T, Hatlebakk JG. Interaction between diet and gastrointestinal endocrine cells. Biomed Rep 2016; 4: 651-656 [PMID: 27284402 DOI: 10.3892/br.2016.649]

26. El-Salhy M, Lilbo E, Reinemo A, Salmeøid L, Hausken T. Effects of a health program comprising reassurance, diet management, probiotic administration and regular exercise on symptoms and quality of life in patients with irritable bowel syndrome. Gastroenterology Insights 2010; 21: [DOI: 10.4041/gi.pen10.06]

27. Jarrett M, Heitkemper MM, Bond EF, Georges J. Comparison of diet composition in women with and without functional bowel disorder. Gastroenterol Nurs 1994; 16: 253-258 [PMID: 8057160 DOI: 10.1097/00001610-199406000-00004]

28. Saito YA, Locke GR, Weaver AL, Zinsmeister AR, Talley NJ. Diet and functional gastrointestinal disorders: a population-based case-control study. Am J Gastroenterol 2005; 100: 2743-2748 [PMID: 16393229 DOI: 10.1111/j.1572-0241.2005.00288.x]

29. El-Salhy M, Gundersen D. Diet in irritable bowel syndrome. Nutr J 2015; 14: 36 [PMID: 25880820 DOI: 10.1186/s12937-015-0022-3]
El-Salhy M, Hatlebakk JG, Gilsø OH, Hausken T. The relation between celiac disease, non-celiac gluten sensitivity and irritable bowel syndrome. *Nutr J* 2015; 14: 92 [PMID: 26345589 DOI: 10.1186/s12977-015-0080-6]

El-Salhy M, Gundersen D, Hatlebakk JG, Hausken T, Watson RR. Diet and irritable bowel syndrome, with a focus on appetite-regulating hormones. In: Watson RR. Nutrition in the prevention and treatment of abdominal obesity. San Diego: Elsevier. Watson RR. 2014; 5-16

El-Salhy M. Diet in the pathophysiology and management of irritable bowel syndrome. *Cleve Clin J Med* 2016; 83: 663-664 [PMID: 27618354 DOI: 10.3949/ccjm.18-016019]

Dionne J, Ford AC, Yuan Y, Chey WD, Lacey BE, Saito YA, Quigley EMN, Moayyedi P. A Systematic Review and Meta-Analysis Evaluating the Efficacy of a Gluten-Free Diet and a Low FODMAPs Diet in Treating Symptoms of Irritable Bowel Syndrome. *Am J Gastroenterol* 2018; 113: 1290-1300 [PMID: 30046155 DOI: 10.1038/s41395-018-0195-4]

Ellis A, Linaker BD. Non-celiac gluten sensitivity? *Lancet* 1978; 1: 1358-1359 [PMID: 78118 DOI: 10.1016/s0140-6736(78)92427-3]

Boettcher E, Crowe SE. Dietary proteins and functional gastrointestinal disorders. *Am J Gastroenterol* 2013; 108: 728-736 [PMID: 23557359 DOI: 10.1038/ajg.2013.97]

Saporen A, Bai JC, Ciacci C, Dolinsk J, Green PH, Hadjivassiliou M, Kaukinen K, Rostami K, Sanders DS, Schumann M, Ulbrich R, Villalta D, Volta U, Catassi C, Fasano A. Spectrum of gluten-related disorders: consensus on new nomenclature and classification. *BMC Med* 2012; 10: 13 [PMID: 22313950 DOI: 10.1186/1741-7015-10-13]

Campanella J, Biagi F, Bianchi PI, Zaneelati G, Marchese A, Corazza GR. Clinical response to gluten withdrawal is not an indicator of coeliac disease. *Scand J Gastroenterol* 2008; 43: 1511-1514 [PMID: 18699173 DOI: 10.1080/00365520802200036]

Vazquez-Roque MI, Camilleri M, Smyrk T, Murray JA, Marietta E, O’Neill JL, Carlson P, Lamsam J, Janzow D, Eckert D, Burton D, Zinsmeister AR. A controlled trial of gluten-free diet in patients with irritable bowel syndrome-diarrhea: effects on bowel frequency and intestinal function. *Gastroenterology* 2013; 144: 903-911 e3 [PMID: 23577155 DOI: 10.1053/j.gastro.2013.01.040]

Kaukinen K, Turjanmaa K, Mäki M, Partanen J, Väänänen R, Reunala T, Collin P. Intolerance to cereals is not specific for coeliac disease. *Scand J Gastroenterol* 2000; 35: 942-946 [PMID: 11063153 DOI: 10.1080/0365552070022995]

Carrocio A, Mansueto P, Iacono G, Sorensi M, D’Alcamo A, Cavataio F, Brusa I, Florena AM, Ambrosiano G, Seidita A, Pirrone G, Rimi GB. Non-celiac wheat sensitivity diagnosed by double-blind placebo-controlled challenge: exploring a new clinical entity. *Am J Gastroenterol* 2012; 107: 1898-1906 [PMID: 22825566 DOI: 10.1038/ajg.2012.236]

Biesiekierski JR, Newnham ED, Irving PM, Barrett JS, Haines M, Doeecke JD, Shepherd SJ, Muir JG, Gibson PR. Gluten causes gastrointestinal symptoms in subjects without celiac disease: a double-blind randomized placebo-controlled trial. *Am J Gastroenterol* 2011; 106: 508-514 [PMID: 21224837 DOI: 10.1038/ajg.2010.487]

Shahbakhshani B, Sadeghi A, Malekzadeh R, Khatavi F, Etemadi M, Kalantari E, Rostami-Nejad M, Rostami K. Non-Celiac Gluten Sensitivity Has Narrowed the Spectrum of Irritable Bowel Syndrome: A Double-Blind Randomized Placebo-Controlled Trial. *Nutrients* 2015; 7: 4542-4554 [PMID: 26569620 DOI: 10.3390/nu7064542]

Skodje GI, Sarna VK, Minelle IH, Rolfsen KL, Muir JG, Gibson PR, Veieriod MB, Henriksen C, Lundin KEA. Fructan, Rather Than Gluten, Induces Symptoms in Patients With Self-Reported Non-Celiac Gluten Sensitivity. *Gastroenterology* 2018; 154: 529-539 [PMID: 29102613 DOI: 10.1053/j.gastro.2017.10.040]

Thursby E, Juge N. Introduction to the human gut microbiota. *Biochem J* 2017; 474: 1823-1836 [PMID: 28552250 DOI: 10.4252/bj20160510]

Hugon P, Dufour JC, Polier C, Fouinier PE, Sallah K, Raoult D. A comprehensive repertoire of prokaryotic species identified in human beings. *Lancet Infect Dis* 2017; 17: 728-736 [PMID: 28512250 DOI: 10.1016/S1473-3099(16)00293-5]

Ley RE, Hamady M, Lozupone C, Turnbaugh PJ, Ramey RR, Bircher JS, Schlegel ML, Tucker TA, Schrenzel MD, Knight R, Gordon JI. Evolution of mammals and their gut microbes. *Science* 2008; 320: 1647-1651 [PMID: 18497261 DOI: 10.1126/science.1157225]

Krisis M, Hazleton KZ, Nubsacher NM, Martin CG, Lozupone CA. Low diversity gut microbiota dysbiosis: drivers, functional implications and recovery. *Curr Opin Microbiol* 2018; 44: 34-40 [PMID: 29026705 DOI: 10.1016/j.mib.2018.07.003]

Wilson BC, Vatanen T, Cutiflet WS, O’Sullivan JM. The Super-Donor Phenomenon in Fecal Microbiota Transplantation. *Front Cell Infect Microbiol* 2019; 9: 2 [PMID: 30719428 DOI: 10.3389/fcimb.2019.00002]

El-Salhy M, Massaquoi T. Fecal microbiota transplantation for managing irritable bowel syndrome. *Expert Rev Gastroenterol Hepatol* 2018; 12: 439-445 [PMID: 29403330 DOI: 10.1080/17474124.2018.1447330]

CASEN C, Vede HC, Selkjell M, Hegge FT, Karlsson MK, Cimnjesiewska E, Dzankovic S, Frøyland C, Nestestog R, Engstrand L, Munkholm P, Nielsen OH, Rogler G, Simrën M, Öhman L, Vatn MH, Rudi K. Deviations in human gut microbiota: a novel diagnostic test for determining dysbiosis in patients with IBS or IBD. *Aliment Pharmacol Ther* 2015; 42: 71-83 [PMID: 25973666 DOI: 10.1111/apt.13236]

Enck P, Mazarak N. Dysbiosis in Functional Bowel Disorders. *Front Microbiol* 2017; 8: 296-306 [PMID: 29609452 DOI: 10.3389/fcmib.2019.00487]

El-Salhy M, Sein I, Chopin L, Gundersen D, Hatlebakk JG, Hausken T. Irritable bowel syndrome: the role of gut neuroendocrine peptides. *Front Biosci (Elite Ed)* 2012; 4: 2783-2800 [PMID: 22652678 DOI: 10.2741/cfs03]

El-Salhy M, Gundersen D, Hatlebakk JG, Hausken T. Irritable bowel syndrome: diagnosis, pathogenesis and treatment options. New York: Nova Science Publishers, 2012: 35-78

El-Salhy M, Hatlebakk JG, Hausken T. Diet in Irritable Bowel Syndrome (IBS): Interaction with Gut Microbiota and Gut Hormones. *Nutrients* 2019; 11: pii: E1824 [PMID: 31349793 DOI: 10.3390/nu11081824]

Maue GM, Coates MD, Moses PL. Review article: intestinal serotonin signalling in irritable bowel syndrome. *Aliment Pharmacol Ther* 2006; 23: 1067-1076 [PMID: 16611266 DOI: 10.1111/j.1365-2036.2006.02858.x]

Wade PR, Chen J, Jaffe B, Kassem IS, Blakely RD, Gershon MD. Localization and function of a 5-HT transporter in crypt epithelia of the gastrointestinal tract. *J Neurosci* 1996; 16: 2352-2364 [PMID: 8601815 DOI: 10.1523/JNEUROSCI.16-07-03252.1996]
El-Salhy M. Stem cells in IBS  

57 Gershon MD, Tack J. The serotonin signaling system: from basic understanding to drug development for functional GI disorders. Gastroenterology 2007; 132: 397-414 [PMID: 17241888 DOI: 10.1053/j.gastro.2006.11.002]

58 Gershon MD. 5-Hydroxytryptamine (serotonin) in the gastrointestinal tract. Curr Opin Endocrinol Diabetes Obes 2013; 20: 14-21 [PMID: 23222533 DOI: 10.1097/MED.0b013e32835b703]

59 Gershon MD. Serotonin is a sword and a shield of the bowel: serotonin plays offense and defense. Trans Am Clin Climatol Assoc 2012; 123: 268-280 [PMID: 2303993]

60 El-Salhy M, Mazzawi T, Gunderson D, Hatlebakk JG, Hausken T. The role of peptide YY in gastrointestinal diseases and disorders (review). Int J Mol Med 2013; 31: 275-282 [PMID: 23292145 DOI: 10.3892/ijmm.2012.1222]

61 Dubrasquet M, Bataille D, Gespach C. Oxyntomodulin (glucagon-37 or bioactive enteroglucagon): a potent inhibitor of pentagastrin-stimulated acid secretion in rats. Biosci Rep 1982; 2: 391-395 [PMID: 6125221 DOI: 10.1042/bj0119301]

62 Schjøldegaard BT, Baldissera FG, Mortensen PE, Holst JJ, Christiansen J. Oxyntomodulin: a potential hormone from the distal gut. Pharmacokinetcis and effects on gastric acid and insulin secretion in man. Eur J Clin Invest 1988; 18: 499-503 [PMID: 3147901 DOI: 10.1111/j.1365-2362.1988.tb01046.x]

63 Schjøldegaard J, Mortensen PE, Myhre J, Christiansen J. Oxyntomodulin from distal gut. Role in regulation of gastric and pancreatic functions. Dig Dis Sci 1989; 34: 1411-1419 [PMID: 2670478 DOI: 10.1007/bf01538076]

64 Dakin CL, Small CJ, Batterham RL, Neary NM, Cohen MA, Patterson M, Ghatai MA, Bloom SR. Peripheral oxyntomodulin reduces food intake and body weight gain in rats. Endocrinology 2004; 145: 2687-2695 [PMID: 15091546 DOI: 10.1210/en.2003-1338]

65 Wynne K, Park AJ, Small CJ, Patterson M, Ellis SM, Murphy KG, Wren AM, Frost GS, Meenan K, Ghatai MA, Bloom SR. Subcutaneous oxyntomodulin reduces body weight in obese subjects: a double-blind, randomized, controlled trial. Diabetes 2005; 54: 2390-2395 [PMID: 16046306 DOI: 10.2337/diabetes.54.8.2390]

66 Camilleri M. Peripheral mechanisms in irritable bowel syndrome. N Engl J Med 2012; 367: 1626-1635 [PMID: 23049224 DOI: 10.1056/NEJMr1107084]

67 Jianu MCS, Fossmark R, Syversen U, Hauso O, Waldum HL. A meal test improves the specificity of chromogranin A as a marker of neuroendocrine neoplasia. Tumour Biol 2010; 31: 373-380 [PMID: 20480408 DOI: 10.1007/s13277-010-0045-5]

68 Gunawanenene AR, Corfe BM, Staton CA. Classification and functions of enterodocrine cells of the lower gastrointestinal tract. Int J Exp Pathol 2011; 92: 219-231 [PMID: 21518048 DOI: 10.1111/j.1365-2613.2011.00767.x]

69 May CL, Kaestner KH. Gut endocrine cell development. Mol Cell Endocrinol 2010; 323: 70-75 [PMID: 20259333 DOI: 10.1016/j.mce.2009.12.009]

70 Sandström O, El-Salhy M. Ageing and endocrine cells of human duodenum. Mech Ageing Dev 1999; 108: 39-48 [PMID: 10566038 DOI: 10.1016/s0047-6374(98)00154-7]

71 El-Salhy M. Ghrelin in gastrointestinal diseases and disorders: a possible role in the pathophysiology and clinical implications (review). Int J Mol Med 2009; 24: 727-732 [PMID: 19856511 DOI: 10.3892/ijmm.2009.00285]

72 Tolhurst G, Reimann F, Gribble FM. Intestinal sensing of nutrients. Handb Exp Pharmacol 2012; 309-335 [PMID: 22429821 DOI: 10.1007/978-3-642-24716-3_14]

73 Lee J, Cummings BP, Martin E, Sharp JW, Stanhope KL, Havel PJ, Raybould HE. Glucose sensing by gut endocrine cells and activation of the vagal afferent pathway is impaired in a rodent model of type 2 diabetes mellitus. Am J Physiol Regul Integr Comp Physiol 2012; 302: R657-R666 [PMID: 22106540 DOI: 10.1152/ajpregu.00345.2011]

74 Parker HE, Reimann F, Gribble FM. Molecular mechanisms underlying nutrient-stimulated incretin secretion. Expert Rev Mol Med 2010; 12: e1 [PMID: 20047700 DOI: 10.1017/S146239940900132X]

75 Raybould HE. Nutrient sensing in the gastrointestinal tract: possible role for nutrient transporters. J Physiol Biochem 2008; 64: 349-356 [PMID: 19391461 DOI: 10.1007/bf03174091]

76 Saa Gabriel A, Nakamura E, Uneyama H, Torii K. Taste, visceral information and exocrine reflexes with glutamate through umami receptors. J Med Invest 2009; 56 Suppl: 209-217 [PMID: 20224183 DOI: 10.2152/jmi.56.200]

77 Rudholm T, Wallin B, Theodorsson E, Näslund E, Helfström PM. Release of regulatory gut peptides somatostatin, neurotensin and vasoactive intestinal peptide by acid and hyperosmolar solutions in the intestine in conscious rats. Reg Pept 2009; 152: 8-12 [PMID: 20680430 DOI: 10.1016/j.regpep.2009.03.002]

78 Sterrini C, Anselmi L, Rozengurt E. Enteroendocrine cells: a site of ‘taste’ in gastrointestinal chemosensing. Curr Opin Endocrinol Diabetes Obes 2008; 15: 73-78 [PMID: 18180566 DOI: 10.1097/MED.0b013e3282f5b373]

79 Sterrini C. Taste receptors in the gastrointestinal tract. IV. Functional implications of bitter taste receptors in gastrointestinal chemosensing. Am J Physiol Gastrointest Liver Physiol 2007; 292: G457-G461 [PMID: 17095755 DOI: 10.1152/ajpgi.00411.2006]

80 Buchan AM. Nutrient Tasting and Signaling Mechanisms in the Gut III. Endocrine cell recognition of luminal nutrients. Am J Physiol 1999; 277: G1103-G1107 [PMID: 10600808 DOI: 10.1152/ajpgi.1999.277.6.G1103]

81 Montero-Hadadje M, Elias S, Chevalier L, Benard M, Tanguy Y, Turquier V, Galas L, Yon L, Malagon MM, Drouich A, Gasman S, Anouar Y. Chromogranin A promotes peptide hormone sorting to mobile granules in constitutively and regulated secreting cells: role of conserved N- and C-terminal peptides. J Biol Chem 2009; 284: 12420-12431 [PMID: 19173339 DOI: 10.1074/jbc.M805072200]

82 Shoshansharzadeh P, Zhang D, Chich JF, Gasnier C, Schneider F, Hafkel Y, Aunis D, Metz-Boutigue MH. The antimicrobial peptides derived from chromogranin/secretogranin family, new actors of innate immunity. Regul Pept 2010; 165: 102-110 [PMID: 19932135 DOI: 10.1016/j.regpep.2009.11.014]

83 Wendelbo I, Mazzawi T, El-Salhy M. Increased serotonin transporter immunoreactivity intensity in the ileum of patients with irritable bowel disease. Mol Med Rep 2014; 9: 180-184 [PMID: 24213511 DOI: 10.3892/mmr.2013.1784]

84 El-Salhy M, Wendelbo I, Gunderson D. Reduced chromogranin A cell density in the ileum of patients with irritable bowel syndrome. Mol Med Rep 2013; 7: 1241-1244 [PMID: 23426642 DOI: 10.3892/mmr.2013.1325]

85 El-Salhy M, Vanli K, Ozdar V, Hausken T. Abnormal small-intestinal endocrine cells in patients with...
Johnson T, Wiedemann LM, Barrett TA, Hood L, Wu H, Li L. PTEN-deficient intestinal stem cells initiate
expressed in crypt base columnar cells of mouse small intestine.

Okano H, Chiba T. Candidate markers for stem and early progenitor cells, Musashi-1 and Hes1, are
Identification of a putative intestinal stem cell and early lineage marker; musashi-1.

Lee CS, Kaestner KH. Clinical endocrinology and metabolism. Development of gut endocrine cells.

Korinek V, Begthel H, Peters PJ, Clevers H. Identification of stem cells in small intestine and colon by marker gene
Tracking down the stem cells of the intestine: strategies to identify adult stem cells.

Barker N, Clevers H. Tracking down the stem cells of the intestine: strategies to identify adult stem cells.

Barbara G, De Giorgio R, Stanghellini V, Corinaldesi R. Activated mast cells in proximity to
neurons of the rectum in patients with irritable bowel syndrome.

El-Salhy M, Gundersen D, Hatlebakk JG, Hausken T. Chromogranin A cell density as a diagnostic marker for
lymphomatous colitis. Dig Dis Sci 2012; 57: 3154-3159 [PMID: 22699394 DOI: 10.1007/s10620-012-2249-6]

Mazzawi T, Hausken T, Gundersen D, El-Salhy M. Effects of dietary guidance on the symptoms, quality of life and habitual dietary intake of patients with irritable bowel syndrome. Mol Med Rep 2013; 8: 845-852 [PMID: 23820783 DOI: 10.3892/mmr.2013.1356]

El-Salhy M, Gilja OH, Hatlebakk JG, Hausken T. Stomach antral endocrine cells in patients with irritable bowel syndrome. Int J Mol Med 2014; 34: 967-974 [PMID: 25110039 DOI: 10.3892/ijmm.2014.1887]

Sjölund K, Ekman R, Wierup N. Covariation of plasma ghrelin and motilin in irritable bowel syndrome. Peptides 2010; 31: 1109-1112 [PMID: 20338210 DOI: 10.1016/j.peptides.2010.03.021]

Wang SH, Dong L, Luo JY, Gong J, Li L, Xu XL, Han SP. Decreased expression of serotonin in the jejunal and increased numbers of mast cells in the terminal ileum in patients with irritable bowel syndrome. World J Gastroenterol 2007; 13: 6641-6647 [PMID: 18023907 DOI: 10.3748/wjg.v13.45.6401]

Park JH, Rhee PL, Kim G, Lee JH, Kim YH, Kim JI, Rhee JC, Song SY. Enterocoid cell count correlates with visceral hypersensitivity in patients with diarrhoea-predominant irritable bowel syndrome. Neurogastroenterol Motil 2008; 20: 539-546 [PMID: 18771769 DOI: 10.1111/j.1365-2982.2006.00771.x]

Coates MD, Mahoney CR, Linden DR, Sampson JE, Chen J, Blaszky J, Crowell MD, Sharkey KA, Gershon MD, Mawe GM, Moses PL. Molecular defects in mucosal serotonin content and decreased serotonin reuptake transporter in ulcerative colitis and irritable bowel syndrome. Gastroenterology 2004; 126: 1657-1664 [PMID: 15181159 DOI: 10.1053/j.gastro.2004.03.013]

El-Salhy M, Wendelbo I, Gundersen D. Serotonin and serotonin transporter in the rectum of patients with irritable bowel disease. Mol Med Rep 2013; 8: 451-455 [PMID: 23778763 DOI: 10.3892/mmr.2013.1525]

Soares RL. Irritable bowel syndrome: a clinical review. World J Gastroenterol 2014; 20: 12144-12160 [PMID: 25322249 DOI: 10.3748/wjg.v20.s1.12144]

Winston AP, Biddle WL, Ibhaia PS, Miner PB. Terminal ileal mucosal mast cells in irritable bowel syndrome. Dig Dis Sci 1993; 38: 1590-1595 [PMID: 8359068 DOI: 10.1007/BF01303164]

O'Sullivan M, Clayton N, Breslin NP, Harman I, Bonnar C, McLaren A, O'Morain CA. Increased mast cells in the irritable bowel syndrome. Neurogastroenterol Motil 2000; 12: 449-457 [PMID: 11012945 DOI: 10.1046/j.1365-2982.2000.00221.x]

Barbara G, De Giorgio R, Stanghellini V, Cremon C, Corinaldesi R. A role for inflammation in irritable bowel syndrome? Gut 2002; 51 Suppl 1: i41-i44 [PMID: 12077063 DOI: 10.1136/gut.51.suppl_1.i41]

Barbara G, Stanghellini V, De Giorgio R, Cremon C, Cottrell GS, Santini D, Pasquinielli G, Morselli-Labate AM, Grady EF, Breslin NP, McLaren A, O'Morain CA. Increased mast cells in the proximal colon correlate with abdominal pain in irritable bowel syndrome. Gastroenterology 2004; 126: 693-702 [PMID: 14988823 DOI: 10.1053/j.gastro.2003.11.055]

Montgomery RK, Breault DT. Small intestinal stem cell markers. J Anat 2008; 213: 52-58 [PMID: 18638070 DOI: 10.1111/j.1469-7580.2008.00925.x]

Barker N, Clevers H. Tracking down the stem cells of the intestine: strategies to identify adult stem cells. Gastroenterology 2007; 133: 1755-1760 [PMID: 18005454 DOI: 10.1053/j.gastro.2007.10.029]

Barker N, van de Wetering M, Clevers H. The intestinal stem cell genes. Dev Genes 2008; 22: 1856-1864 [PMID: 18628392 DOI: 10.1110/sgd.164008]

Barker N, van Es JH, Kuipers J, Kujala P, van den Born M, Cozijnsen M, Haegeman A, Korving J, Begthel H, Peters PJ, Clevers H. Identification of stem cells in small intestine and colon by marker gene Lgr5. Nature 2007; 449: 1003-1007 [PMID: 17934449 DOI: 10.1038/nature06196]

Korinek V, Barker N, Moerer P, van Donselaar E, Huls G, Peters PJ, Clevers H. Depletion of epithelial stem-cell compartments in the small intestine of mice lacking Tcf-4. Nat Genet 1998; 19: 379-383 [PMID: 9697701 DOI: 10.1038/1270]

Cheng H, Leblond CP. Origin, differentiation and renewal of the four main epithelial cell types in the mouse small intestine. V. Unitarian Theory of the origin of the four epithelial cell types. Am J Anat 1974; 141: 537-561 [PMID: 4440635 DOI: 10.1002/aja.10101410407]

Le Douarin NM, Teillet MA. The migration of neural crest cells to the wall of the digestive tract in avian embryo. J Embryol Exp Morphol 1973; 36: 31-48 [PMID: 4729950]

Rawson BB, Andrew A. Origin and differentiation of gut endocrine cells. Histol Histopathol 1993; 8: 567-580 [PMID: 8358116]

Hoffman J, Kuhnert F, Davis CR, Kuo CJ. Wnts as essential growth factors for the adult small intestine and colon. Cell Cycle 2004; 3: 554-557 [PMID: 15044853 DOI: 10.4161/cc.3.5.858]

Lee CS, Kaestner KH. Clinical endocrinology and metabolism. Development of gut endocrine cells. Best Pract Res Clin Endocrinol Metab 2004; 18: 453-462 [PMID: 15533769 DOI: 10.1016/j.beem.2004.08.008]

Potten CS, Booth C, Tudor GL, Booth D, Brady G, Harley P, Ashton G, Clarke R, Sakakibara S, Okano H. Identification of a putative intestinal stem cell and early lineage marker; musashi-1. Differentiation 2003; 71: 28-41 [PMID: 12556601 DOI: 10.1046/j.1432-0436.2003.00754.x]

Kayahara T, Sawada M, Takishii S, Fukushi H, Seno H, Fukuzawa H, Suzuki K, Hiai H, Kageyama R, Okano H, Chiba T. Candidate markers for stem and early progenitor cells, Musashi-1 and Hes1, are expressed in crypt base columnar cells of mouse small intestine. FEBS Lett 2003; 535: 131-135 [PMID: 12568091 DOI: 10.1016/S0014-5793(02)03896-6]

Ne HC, Yin T, Grindley JC, Tian Q, Sato T, Tao WA, Dirisana R, Porter-Westphal KS, Hembree M, Johnson T, Wiedemann LM, Barrett TA, Hood L, Wu H, Li L. PTEN-deficient intestinal stem cells initiate
intestinal polyposis. Nat Genet 2007; 39: 189-198 [PMID: 17237784 DOI: 10.1038/ng1928]

Fishbein TM, Novitskyy G, Loug DH, Matsumoto C, Kaufman SS, Shetty K, Zaslavoff M. Rejection reversibly alters enteroendocrine cell renewal in the transplanted small intestine. Am J Transplant 2009; 9: 1620-1628 [PMID: 19219821 DOI: 10.1111/j.1660-6433.2009.02681.x]

Schonhoff SE, Giel-Moloney M, Leiter AB. Minireview: Development and differentiation of gut endocrine cells. Endocrinology 2004; 145: 2639-2644 [PMID: 15044535 DOI: 10.1210/en.2004-0051]

Schonhoff SE, Giel-Moloney M, Leiter AB. Neurogenin 3-expressing progenitor cells in the gastrointestinal tract differentiate into both endocrine and non-endocrine cell types. Dev Biol 2004; 278: 443-454 [PMID: 15187225 DOI: 10.1016/j.ydbio.2004.03.013]

El-Salhy M, Hausken T, Gilja OH, Hatlebakk JG. The possible role of gastrointestinal endocrine cells in the pathophysiology of irritable bowel syndrome. Expert Rev Gastroenterol Hepatol 2017; 11: 139-148 [PMID: 27927062 DOI: 10.1080/17474124.2017.1269601]

Jenny M, Uhl C, Roche C, Duluc I, Guillermin V, Guillenot F, Jensen J, Keding M, Gradwohl G. Neurogenin3 is differentially required for endocrine cell fate specification in the intestinal and gastric epithelium. EMBO J 2002; 21: 6338-6347 [PMID: 12456641 DOI: 10.1093/emboj/cdf649]

Wang J, Cortina G, Wu SV, Tran R, Cho JL, Tsai M, Bailey TJ, Janmrich M, Ament ME, Teem WR, Hill ID, Vargas JH, Gershman G, Farmer DG, Reyun L, Martin MG. Mutant neurogenin-3 in congenital malabsorptive diarrhea. N Engl J Med 2006; 355: 270-280 [PMID: 16857267 DOI: 10.1056/NEJM-Mou054288]

Mazzawi T, El-Salhy M. Changes in small intestinal chromogranin A-immunoreactive cell densities in patients with irritable bowel syndrome after receiving dietary guidance. Int J Mol Med 2016; 37: 1247-1255 [PMID: 26987104 DOI: 10.3892/ijmm.2016.2523]

Mazzawi T, El-Salhy M. Dietary guidance and ileal enteroendocrine cells in patients with irritable bowel syndrome. Exp Ther Med 2016; 12: 1398-1404 [PMID: 27588061 DOI: 10.3892/etm.2016.3491]

Mazzawi T, Gundersen D, Hausken T, El-Salhy M. Increased gastric chromogranin A cell density following changes to diets of patients with irritable bowel syndrome. Mol Med Rep 2014; 10: 2322-2326 [PMID: 25744555 DOI: 10.3892/mmr.2014.2498]

Mazzawi T, Gundersen D, Hausken T, El-Salhy M. Increased chromogranin A cell density in the large intestine of patients with irritable bowel syndrome after receiving dietary guidance. 2015; 823897 [PMID: 25918524 DOI: 10.1155/2015/823897]

Mazzawi T, Hausken T, Gundersen D, El-Salhy M. Effect of dietary management on the gastric endocrine cells in patients with irritable bowel syndrome. Eur J Clin Nutr 2015; 69: 519-524 [PMID: 25097063 DOI: 10.1038/ejcn.2014.151]

Mazzawi T, Hausken T, Gundersen D, El-Salhy M. Dietary guidance normalizes large intestinal endocrine cell densities in patients with irritable bowel syndrome. Eur J Clin Nutr 2016; 70: 175-181 [PMID: 26603880 DOI: 10.1038/ejcn.2015.191]

Chen Y, Arslan G, El-Salhy M, Gilja OH, Hatlebakk JG, Hausken T. Effect of fecal microbiota transplantation on the symptoms and duodenal enteroendocrine cells in patients with irritable bowel syndrome. United Eur Gastro 2016; 4: 677-677 [DOI: 10.26226/moreressor.57c53841d462b80296c9c83f]

Chen S, Xia Y, Zhu G, Yan J, Tan C, Deng B, Deng J, Yin Y, Ren W. Glutamine supplementation improves intestinal cell proliferation and stem cell differentiation in weaning mice. Food Nutr Res 2018; 62 [PMID: 30083086 DOI: 10.2291/ijn.v62.1439]

Chen Y, Tsai YH, Tseng BJ, Tseng SH. Influence of Growth Hormone and Glutamine on Intestinal Stem Cells: A Narrative Review. Nutrients 2019; 11 [PMID: 31426533 DOI: 10.3390/nu11081941]

Chen Y, Tseng SH, Yao CL, Li C, Tsai YH. Distinct Effects of Growth Hormone and Glutamine on Activation of Intestinal Stem Cells. JPN/J Parenteral Enteral Nutr 2018; 42: 642-651 [PMID: 28510488 DOI: 10.1177/0148607117709435]

Kim MH, Kim H. The Roles of Glutamine in the Intestine and Its Implication in Intestinal Diseases. Int J Mol Sci 2017; 18 [PMID: 28498331 DOI: 10.3390/ijms18051051]

Wang B, Wu G, Zhou Z, Dai Z, Sun Y, Ji Y, Li W, Wang W, Liu C, Han F, Wu Z. Glutamine and intestinal barrier function. Amino Acids 2015; 47: 2143-2154 [PMID: 24965526 DOI: 10.1007/s00726-014-1773-4]

Zhou Q, Verne ML, Fields IZ, LeFante JJ, Basra S, Salameh H, Verne GN. Randomised placebo-controlled trial of dietary glutamine supplements for postinfectious irritable bowel syndrome. Gut 2019; 68: 996-1002 [PMID: 30108163 DOI: 10.1136/gutjnl-2017-315136]

Curoha A, Gruetter R, Do KQ, Duarte JMN. Social isolation stress and chronic glutathione deficiency have a common effect on the glutamine-glutamate ratio and myo-inositol concentration in the mouse frontal cortex. J Neurochem 2017; 142: 767-775 [PMID: 28646450 DOI: 10.1111/jnc.14116]

Harnett NG, Wood KH, Ference EW, Reid MA, Lahti AC, Knight AJ, Knight DC. Glutamate/glutamine concentrations in the dorsal anterior cingulate vary with Post-Traumatic Stress Disorder symptoms. J Psychiatric Res 2017; 91: 169-176 [PMID: 28478230 DOI: 10.1016/j.jpsychires.2017.04.010]

El-Salhy M, Umezawa K, Hatlebakk JG, Gilja OH. Abnormal differentiation of stem cells into enteroendocrine cells in rats with DSS-induced colitis. Mol Med Rep 2017; 15: 2106-2112 [PMID: 28259897 DOI: 10.3892/mmr.2017.6266]

El-Salhy M, Mazzawi T, Umezawa K, Gilja OH. Enteroendocrine cells, stem cells and differentiation progenitors in rats with TNBS-induced colitis. Int J Mol Med 2016; 38: 1743-1751 [PMID: 27779708 DOI: 10.3892/ijmm.2016.2787]
