Effects of retinoic acid isomers on apoptosis and enzymatic antioxidant system in human breast cancer cells

Tae-Kyong Hong and Yang Cha Lee-Kim
Department of Food and Nutrition, Yonsei University, 134 Shinchon-dong, Seodaemun-gu, Seoul 120-749, Korea

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Abstract
Retinoic acids (RAs) modulate growth, differentiation, and apoptosis in normal, pre-malignant & malignant cells. In the present study, the effects of RA isomers (all-trans RA, 13-cis RA, and 9-cis RA) on the cell signal transduction of human breast cancer cells have been studied. The relationship between RAs and an enzymatic antioxidant system was also determined. Estrogen-receptor (ER) positive MCF-7 and ER-negative MDA-MB-231 human breast cancer cells were treated with different doses of each RA isomers, all-trans RA, 13-cis RA, or 9-cis RA. Treatment of RA isomers inhibited cell viability and induced apoptosis of MCF-7 cells as a result of increased caspase activity in cytoplasm and cytochrome C release from mitochondria. All-trans RA was the most effective RA isomer in both cell growth inhibition and induction of apoptosis in MCF-7 cells. However, no significant effect of RA isomers was observed on the cell growth or apoptosis in ER-negative MDA-MB-231 cells. In addition, activities of antioxidant enzymes such as catalase and glutathione peroxidase were decreased effectively after treatment of RA in MCF-7 cells, whereas SOD activity was rarely affected. Thus, the present data suggest that all-trans RA is the most potential inducer of apoptosis and modulator of antioxidant enzymes among RA isomers in MCF-7 human breast cancer cells.

Key Words: Breast cancer cells, RA Isomers, apoptosis, antioxidant enzymes

Introduction
Among the many investigations of breast cancer, there has been much interest recently in retinoic acid (RA) metabolites used in i) cellular growth process, ii) activating gene transcription by specific nuclear receptors, iii) inhibiting cancer cell growth by several different pathways (Freemantle et al., 2003; Marill et al., 2003; Napoli, 1996; Ralhan & Kaur, 2003; Smith et al., 1992). Previous studies also suggest an intimate relationship between cancer and serum RA level; possibly in prevention of cancer cell proliferation or development into second malignancy in breast and liver cancer (Grønemeyer & Miturski, 2001; Manor et al., 2003; Prakash et al., 2001). Treatment with RA in other types of cancer, ischemia or pathophysiology of acne has been shown to be effective as well (Brodowicz et al., 1999; Freemantle et al., 2003; Nelson et al., 2006; Sato et al., 2008). Many studies have been conducted on molecular and cellular mechanisms of RA in the past ten years. In particular, the roles of RA in control and gene expression of cancer cell’s initiation and apoptosis, in multiple signal pathways inside cancer cells, and in the correlation between cancer cells and RA receptors have been investigated (Chambon, 1996; Hayashi et al., 2001; Marill et al., 2003). However, recent clinical experiments that used RA show no effect in reducing first stage or second stage lung cancer for cigarette smokers. In addition, due to the potential toxicity of RA, detailed research is needed on the effect of physiological dose of RA through a close examination of its mechanisms at the cellular level (Freemantle et al., 2003; Ralhan & Kaur, 2003; Windhorst & Nigra, 1982).

When free radicals and reactive oxygen species (ROS) accumulate in the process of aerobic cell metabolism, it consequently results in diseases and aging by inducing damage to tissues and cells. ROS are also known to be involved in cancer growth, cancer cell regeneration and differentiation, and cell signaling systems (Drisko et al., 2003). The enzymatic defense mechanisms with superoxide dismutase (SOD), catalase, and glutathione peroxidase participate in converting superoxide anion to H2O2, and then to H2O. Although turnover volume for catalase is very high, its affinity to H2O2 is relatively low. Therefore, when H2O2 accumulates in cells in even low concentration, it can cause oxidative damage to DNA, which then induces cancer or cell death (Nordberg & Arner, 2001). According to studies of relationship between RA and antioxidation, RA stabilizes protein levels in brain cells and the activity of SOD at the mRNA level, thereby reduces oxidative stress and apoptosis (Ahlemeyer et al., 2001). On the other hand, according to other researches, among RA isomers, all-trans RA suppresses cell growth of malignant myeloblastic leukemia and induces its apoptosis, and
is known to control the activity levels of antioxidant enzymes in rat sertoli cells. Many different aspects of cellular level mechanisms of RA are currently being discovered. However, inconsistency of the results requires more in-depth research (Conte da Frota et al., 2006; Xu et al., 2002).

In this report, we have investigated to measure the influence of RA isomers (all-trans RA, 13-cis RA, and 9-cis RA) on apoptosis and on enzymatic antioxidant system in breast cancer cell lines using the activity levels of caspases and antioxidant enzymes.

**Materials and Methods**

**Chemicals and reagents**

All-trans RA, 13-cis RA, and 9-cis RA were purchased from Sigma Chemical Co. (St. Louis, MO). Growth media RPMI-1640 and other growth supplements, heat-inactivated fetal bovine serum (FBS), penicillin streptomycin, Hank’s Balanced Salt Solutions (HBSS) were obtained from Gibco-BRL (Eggenstein, Germany). A spectrophotometric assay kit for SOD activity and a colormetric kit for apoptosis were purchased from R&D Systems (Wiesbaden, Germany). Caspase-8&-9 fluorescent assay kits were purchased from Peptron Co. (Daejeon, Korea).

**Cell culture and treatment of cells**

Human breast ER-positive cells MCF-7 and ER-negative cells MDA-MB-231 obtained from the American Type Culture Collection (Rockville, MD) were grown in Roswell Park Memorial Institute (RPMI) medium-1640 containing HEPES, sodium bicarbonate, penicillin streptomycin, and 10% fetal bovine serum at 37°C in a humidified atmosphere of 5% CO2 in air. Around 80% confluent cells were passaged by standard trypsinization procedures. Cells were seeded at a density of 3x10⁴ cells/100 dish. The RA isomer solutions dissolved in ethanol (final ethanol concentration was 0.1%) were diluted in culture medium to final concentrations of 10, 100, and 1,000 nM. Cells were treated for up to 4 days. All solutions containing RA isomers were handled under yellow light. The range of RA isomers were changed every 48 hrs and cells were harvested and counted with a haemocytometer. Each sample was counted in duplicate, and each experiment was done in triplicate (Prakash et al., 2001).

**MTT assay**

Following incubation with various reagents, the rate of cell viability was assessed by colormetric measurement of 3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyl tetrazolium bromide (MTT) reduction (Rishi et al., 2003). Values from each treatment were calculated as a percentage relative to the untreated control (defined as 100% survival).

**Apoptosis detection assay**

MCF-7 and MDA-MB-231 cells placed in dishes filled with 3.7% buffered formaldehyde solution for 7 min. at room temperature (RT) were washed twice with 1X PBS. The cells were post-fixed in 100% methanol for 20 min and washed twice with 1X PBS. A portion of 50 μl of cytonin solution was added to each dish and the cells were incubated for 15 min at RT. Fixed cells were washed with 200 μl of DNase-free water, to which 50 μl of 2.5% H2O2 solution was added and the cells were incubated for 5 min at RT. After washing the cells again, 150 μl of 1X TdT labeling buffer was added, and the cells were incubated for 5 min. The cells were washed, and 50 μl of labeling reaction mixture was added followed by incubation at 37°C for 1 hr. The cells were placed in 150 μl of 1X TdT stop buffer for 5 min and washed twice with 1X PBS for 2 min. The cells were placed in 50 μl of streptavidin-HRP solution and incubated at RT for 10 min. Cells were washed four times with 200 μl of PBS and 0.1% Tween 20. The cells were then placed in 100 μl of TACS-Sapphire and incubated at RT for 30 min in the dark. The reaction was stopped by adding 100 μl of 2N HCl. The absorbance of apoptotic cells, which could be detected by labeling of terminal deoxynucleotidyl transferase (TdT) and TACS-Sapphire, was measured at 450 nm (Kuypers et al., 1996; Pigault et al., 1994)

**Caspase activity assay**

To determine caspase-3 activity, western blot analysis (Nelson et al., 2006) was used. Fluorometric assay was used to measure activities of caspase-8 &-9 (Kuida, 2000; Stennicke & Salvesen, 2000; Villa et al., 1997).

**Determination of cytochrome C releasere**

The release of cytochrome C into the cytosol of MCF-7 cells treated with 1 μM of RA isomers for 4 days was measured by a western blot (Nagase et al., 2001) For the detection of cytochrome C, the lysate from the last centrifugation and mitochondrial fractions were subjected to 12% SDS-PAGE. Proteins in the gel were transferred onto prewetted polyvinylidene difluoride (PVDF)- nitrocellulose filters (Bio-Rad, Cambridge, MA). Membranes were blocked with 5% non-fat, dry milk in TBST at RT for 1hr to reduce nonspecific antibody binding. Membranes were washed with PBS three times and incubated with cytochrome C specific antibody. Specific bands of interest were detected by Enhanced Chemiluminescence (ECL; Amersham, Buckinghamshire, U.K.).
Antioxidant enzyme activity assay

Activities of SOD, catalase and glutathione peroxidase were measured by spectrophotometry (Arruda et al., 1996; Bai et al., 1999; Sutherland & Learmonth, 1997).

Statistical analysis

For statistical analysis, each experiment was carried out in triplicate sets and the SAS program was used. Data were expressed as mean ± S.D. The differences among mean values were assessed by one-way analysis of variance (ANOVA) coupled with Duncan’s multiple range test or student’s t test. A p-value <0.05 was considered to be statistically significant.

Results

Effect of RA isomer concentrations on breast cancer cell growth

In the ER-positive MCF-7 cell line, the cancer cell growth was blocked by the high concentration of RA. In particular, all-trans RA showed greater inhibition of cancer cell growth than other isomers did, showing the lowest cell survival rate of 61% at 1 μM concentration. This result was followed by 13-cis RA, which showed a 67% cell survival and 9-cis RA resulted in 75% survival, which showed the least influence in breast cancer cell growth. On the other hand, in the ER-negative MDA-MB-231 cell line, all-trans RA did show a better suppression of cancer cell growth (79% cell survival) than the other isomers similar to shown in ER-positive MCF-7 cell line. The effect on cell growth inhibition in the ER-negative MDA-MB-231 cells was not as significant as in ER-positive MCF-7 cell line (Fig. 1).

Changes in apoptosis by RA isomers

In both ER-positive MCF-7 and ER-negative MDA-MB-231 cell lines, as the concentration of RA isomers (all-trans RA, 13-cis RA, and 9-cis RA) became higher, the number of cell deaths increased (Fig. 2). All-trans RA in the MCF-7 cell line at 1 μM concentration showed the greatest suppression of cancer cell growth up to 41%, followed by 13-cis RA up to 37%, and 9-cis RA up to 28%. In the MDA-MB-231 cell line, the induction of apoptosis was considerably lower, with all-trans RA showing 18% cell death, 13-cis RA showing 16%, and 9-cis RA showing 11% (Fig. 2). These results indicate that RA isomers are closely related to sensitivity and resistance of breast cancer cells. According to several research results, the ER positive breast cancer cells are known to be related to sensitivity of breast cancer cells due to RA, and exert influence on cell growth inhibition, while ER-negative breast cancer cells are reported to have more effect on cells’ resistance than on sensitivity (Freemantle et al., 2003; Gallagher, 2002).

Changes in caspase-3 activity and inhibition by RA isomers

The effect of RA isomers on cell death of the ER-positive MCF-7 cell line was the maximum at 1 μM of RA isomers. Examining with a western blot, the changes in the activity of
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Caspase-3 activity was measured by Western blot using caspase cleavage specific antibody.

Fig. 3. Effect of retinoic acid (RA) isomers on caspase-3 activity in MCF-7 cells.
MCF-7 cells were seeded at 3x10^4 cells and incubated with 1 μM RA isomers. Caspase-3 activity was measured by a Western blot using caspase cleavage specific antibody.

(a) Control All-trans RA 13-cis RA 9-cis RA
(b) Control All-trans RA Caspase inhibitor

Fig. 4. Effect of all-trans retinoic acid (RA) on caspase inhibition of MCF-7 cells.
MCF-7 cells were seeded at 3x10^4 cells and incubated with 1 μM all-trans RA. Caspase inhibitor was used by Ac-DEVD-CHO. A: Caspase-3 activity by caspase inhibitor in ER-positive MCF-7 cells. B: Effect of caspase inhibitor on viability of MCF-7 cells. Results are expressed as relative % of control value. Values are means ± S.D. of three independent experiments.

Changes in cytochrome C release in mitochondria by RA isomers

Cytochrome C release in mitochondria was measured using a western blot. The results showed that all-trans RA exerted the most influence on the cytochrome C release, similar to the results for caspase-3 activity. However, 9-cis RA and 13-cis RA released cytochrome C in mitochondria, more so with 9-cis RA than with 13-cis RA. This indicates that in programmed cell death, RA isomers act differently in mechanisms of apoptosis (Fig. 5).

Changes in caspase-8 and caspase-9 activities by RA isomers

For inducing apoptosis, pro-caspase-8 of the death receptors is activated first followed by other caspases activations resulted
Discussion

Many studies have uncovered therapeutic abilities of RA in breast cancer by either blocking cancer regeneration or inducing cancer apoptosis (Freemantle et al., 2003; Marill et al., 2003). Although scientists have focused on molecular and cellular mechanisms of RA for the last decade, research on RA’s multiple signal pathways is still scarce. The current study focused on the effect of RA isomers (all-trans RA, 13-cis RA, and 9-cis RA) on the cell growth and cell death in breast cancer cell lines, ER-positive MCF-7 cells and ER-negative MDA-MB-231 cells, as well as on the effect of antioxidant systems in the cell lines. In ER-positive MCF-7 cell line, as the concentration of RA isomer treatment increased, the growth of cancer cells was inhibited as compared to that of the control. All-trans RA specifically displayed the most significant suppression of cancer cell growth over the other isomers. While all-trans RA showed stronger suppression of cancer cell growth than the other isomers in ER-negative MDA-MB-231 cell line similar to the effect on the ER-positive MCF-7 cell line, it produced a higher cell survival rate than that of 9-cis RA in ER-positive MCF-7 cell line. It has been reported that breast cancer is closely related to the influence of hormones and cancer cell growth shows significant differences depending on the protein binding affinity of hormone receptors (Hara et al., 2000; Marill et al., 2003; Napoli, 1996; Ott & Lachance, 1979; Paik et al., 2003; Soprano et al., 2004). ER-positive breast cancer cells are known to respond more sensitively to RA isomers in their inhibition of cancer cell growth in contrast to ER-negative breast cancer cells, which are known to influence cancer cell resistance (Gallagher, 2002; Nagpal & Chandraratna, 2000; Xu et al., 1999).

The current study indicated that the inhibition of breast cancer cell growth was associated with apoptosis as shown in the MCF-7 cell line where all-trans RA induced the highest level of apoptosis, followed by 13-cis RA and 9-cis RA. It seemed that there was no significant cell death effect in the MDA-MB-231 cell line. In accordance with the previous study (Prakash et al., 2001), all-trans RA induced greater apoptosis than the other RA isomers in the current study. In normal cells, all-trans RA arrested the G1 phase of the cell cycle without apoptosis induction, whereas in breast cancer cell line, MCF-7, RA not only arrested G0/G1 phases but also induced apoptosis at low concentrations (Dietze et al., 2002; Dimberg & Oberg, 2003; Rishi et al., 2002). Therefore, it is plausible that the effects mentioned above seem to be associated with the caspase activities and cytochrome C as shown in the current study. RA isomers may affect the regulation of Bcl-2 family protein and the release of cytochrome C in mitochondria (thus called a key regulator of caspase cascade). Cytochrome C combines with Apaf-1 and pro-caspase-9 to form apoptosome, which activates caspase-3 resulted in the activation of the apoptosis pathway. It is interesting to note that the current study indicates that RA isomer treatments increase the activities of caspase-9 and -3. Furthermore, apoptosis is activated when pro-caspase-8 of the death receptor is firstly activated and successively activates other caspases,
thereby activating a caspase cascade. When apoptosis is induced, the typical morphological change is known to occur due to the division of key cellular protein that results from the caspase cascade (Hengartner, 2000). In this study, the morphological change was also occurred by treatment of RA isomers (Data not shown) and the activity of caspase-8 was also increased in breast cancer cells. Caspases involved in apoptosis are known to be largely classified as initiator caspase-2, -8, -9, -10 and effector caspase-3, -6, -7 (Hengartner, 2000; Melino et al., 1997). Overall, the results obtained from the current study by treating RA isomers (all-trans RA, 13-cis RA, and 9-cis RA) in estrogen receptor (ER) positive MCF-7 cell line verify that caspase-8, -9, and -3 are the main effector of the cell death program (Raloff, 2000; Ueda (1999). Inhibition of proliferation and induction of apoptosis in soft tissue sarcoma cells by interferon-alpha and retinoids. Bj Cancer 80:1350-1358.

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