Borneo coral reefs subject to high sediment loads show evidence of resilience to various environmental stressors

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For reefs in SE Asia the synergistic effects of rapid land-development, insufficient environmental policies and a lack of enforcement has led to poor water quality and compromised coral health from increased sediment and pollution. Those inshore turbid coral reefs, subject to significant sediment inputs, may also inherit some resilience to the effects of thermal stress and coral bleaching. We studied the inshore turbid reefs near Miri, in northwest Borneo through a comprehensive assessment of coral cover and health in addition to quantifying sediment-related parameters. Although Miri Reefs had comparatively low coral species diversity, dominated by massive and encrusting forms of Diploastrea, Porites, Montipora, Favites, Dipsastrea and Pachyseris, they were characterised by a healthy cover ranging from 22-39%. We found a strong inshore to offshore gradient in hard coral cover, diversity and community composition as a direct result of spatial differences in sediment at distances <10 km. As well as distance to shore, we included other environmental variables like reef depth and sediment trap accumulation and particle size that explained 62.5% of variation in benthic composition among sites. Miri’s reefs showed little evidence of coral disease and relatively low prevalence of compromised health signs including bleaching (6.7%), bioerosion (6.6%), pigmentation response (2.2%), scars (1.1%) and excessive mucus production (0.5%). Tagged colonies of Diploastrea and Pachyseris suffering partial bleaching in 2016 had fully (90-100%) recovered the following year. There were, however, seasonal differences in bioerosion rates, which increased five-fold after the 2017 wet season. Differences in measures of coral physiology, like that of symbiont density and chlorophyll a for Montipora, Pachyseris and Acropora, were not detected among sites. We conclude that Miri’s reefs may be in a
temporally stable state given minimal recently dead coral and a limited decline in coral cover over the last two decades. This study provides further evidence that turbid coral reefs exposed to seasonally elevated sediment loads can exhibit relatively high coral cover and be resilient to disease and elevated sea surface temperatures.
Borneo coral reefs subject to high sediment loads show evidence of resilience to various environmental stressors.

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ABSTRACT

For reefs in SE Asia the synergistic effects of rapid land-development, insufficient environmental policies and a lack of enforcement has led to poor water quality and compromised coral health from increased sediment and pollution. Those inshore turbid coral reefs, subject to significant sediment inputs, may also inherit some resilience to the effects of thermal stress and coral bleaching. We studied the inshore turbid reefs near Miri, in northwest Borneo through a comprehensive assessment of coral cover and health in addition to quantifying sediment-related parameters. Although Miri Reefs had comparatively low coral species diversity, dominated by massive and encrusting forms of Diploastrea, Porites, Montipora, Favites, Dipsastrea and Pachyseris, they were characterised by a healthy cover ranging from 22-39%. We found a strong inshore to offshore gradient in hard coral cover, diversity and community composition as a direct result of spatial differences in sediment at distances <10 km. As well as distance to shore, we included other environmental variables like reef depth and sediment trap accumulation and particle size that explained 62.5% of variation in benthic composition among sites. Miri’s reefs showed little evidence of coral disease and relatively low prevalence of compromised health signs including bleaching (6.7%), bioerosion (6.6%), pigmentation response (2.2%), scars (1.1%) and excessive mucus production (0.5%). Tagged colonies of Diploastrea and Pachyseris suffering partial bleaching in 2016 had fully (90-100%) recovered the following year. There were, however, seasonal differences in bioerosion rates, which increased five-fold after the 2017 wet season. Differences in measures of coral physiology, like that of symbiont density and chlorophyll a for Montipora, Pachyseris and Acropora, were not detected among sites. We conclude that Miri’s reefs may be in a temporally stable state given minimal recently dead coral and a limited decline in coral cover over the last two decades. This study provides further
INTRODUCTION
Turbid reefs are commonly regarded as marginal reefs living near their environmental limits (Kleypas et al., 1999; Guinotte et al., 2003; Perry and Larcombe, 2003; Palmer et al., 2010; Goodkin et al., 2011). As such, these reefs are traditionally perceived to be in a reduced health status (Kleypas, 1996; Kleypas et al., 1999) and more sensitive to rising sea surface temperatures (SST; Nugues and Roberts, 2003; Crabbe and Smith, 2005; Fabricius, 2005; Woolridge, 2008). Yet, there is growing evidence that turbid reefs may actually be more resilient to future climate change effects (Goodkin et al., 2011; Morgan et al., 2017) and serve as refugia for surviving corals (Cacciapaglia and van Woesik, 2015; 2016; Morgan et al., 2016). This has been demonstrated on turbid reefs that experience significant sediment and nutrient inputs, yet are still characterized by high coral cover, low bleaching, and rapid recovery rates from bleaching and cyclonic events (Larcombe et al., 2001; Browne et al., 2010; Richards et al., 2015; Morgan et al., 2016). Studying the level of resilience and survival of turbid reefs in different environmental settings will provide clearer insights into the future of reefs subject to climate change (Guinotte et al., 2003; Hennige et al., 2010; Richards et al., 2015).

Despite elevated resilience to naturally turbid conditions, many inshore turbid reefs face threats from local pressures, largely related to declining water quality and increased sediment input. In South East (SE) Asia, 95% of reefs are threatened from local sources and, therefore, are regarded as the most endangered reefs globally (Burke et al., 2011). From the 1980’s to early 2000’s hard coral cover on reefs in SE Asia declined from 45% to 22% at an average rate of 2% loss per year (Bruno and Selig 2007). Most SE Asian reefs are located in close proximity to countries with rapidly emerging economies and fast population growth (Wilkinson, 2006; Burke et al., 2011; Heery et al., 2018). They are further characterised by poorly developed environmental policies, inadequate regulation, lack of enforcement, a shortage of institutional and technical capacity, insufficient community support and involvement, and conflicts and tensions between stakeholders (Fidelman et al., 2012). The synergistic effects of these factors has led to poor water quality on many inshore reefs via pollution and sediment input derived by rapid land development, and over-fishing activities (McManus, 1997; Wilkinson, 2006). As a consequence,
sedimentation rates are high with SE Asian coastal systems experiencing the highest siltation loads globally (Kamp-Nielsen et al., 2002; Syvitski et al., 2005).

Nearshore coral reefs along the north central section of Sarawak, on the island of Borneo, are highly diverse with an estimated 518 fish species (Shabdin, 2014) and 203 hard coral species from 66 genera (Elcee Instrumentation, 2002). Sarawak is a deforestation hotspot with only 3% of its forest cover intact (Bryan et al., 2013). Ongoing deforestation and poor land use practices are a growing threat for these biological diverse reefs that also support local fisheries and an expanding dive tourism industry (Elcee Instrumentation, 2002). As such, in 2007 a marine park (the Miri-Sibuti Coral Reef National Park; MSCRNP) that covered 11,020 km² was established to promote and protect 30 coral reefs adjacent to Miri, the second largest town in Sarawak. In 2001, a broad assessment of coral reef health within the park indicated that live coral cover was approximately 35-50% and recently dead coral cover was 0.5% (Elcee Instrumentation, 2002).

Subsequent Reef Check surveys in 2010 and 2014 concluded these same reefs were experiencing multiple stressors, but were in ‘fair’ condition (~40% hard coral cover; Reef Check, 2010;2014). However, despite these surveys there is limited quantitative data on coral health and biodiversity (Shabdin, 2014), and more importantly no comprehensive assessment of environmental drivers of reef health. For example, the Baram River (10 km north of the reef complex), is known to discharge $2.4 \times 10^{10}$ kg yr$^{-1}$ of sediments into the coastal zone (Nagarajan et al., 2015), such that sediment and nutrient influx are considered to be the greatest threat to these poorly studied reefs (Pilcher and Cabanban, 2000; Ferner, 2013; Shabdin, 2014). Without thoroughly quantifying sediment impacts on corals, no conclusions can be made on coral tolerance levels, the drivers of community composition and future resilience to both local and global pressures. Given the Baram River delta is in a destructive phase due to rising sea level (Lambiase et al., 2002), together with the increased frequency and intensity of rainfall events and plans for future modification of both the river and adjacent land development (Nagarajan et al., 2015), it is likely that threats from sediments will only increase.

The reefs within the MSCRNP provide a valuable opportunity to address several knowledge gaps on turbid coral reef health and their potential resilience to local and global threats. The last comprehensive assessment of coral cover on Miri’s reefs was in 2001 (Elcee Instrumentation,
2002), with no assessments of coral taxa health and disease for any Sarawak reef recorded to date. In particular, coral disease studies are rarely undertaken on SE Asian reefs largely due to a lack of resources and expertise (Green and Bruckner, 2000; Raymundo et al., 2005; Heintz et al., 2015). The lack of quantitative data on the health and stability (as defined by resistance, resilience and maintenance of key functional groups) of these reefs coupled with ongoing unsustainable land use practices in Sarawak, raises concerns over their long-term viability. This is of particular concern as Sarawak reefs have been estimated to provide 6 million AUD per year in tourism and 13.5 million AUD from fisheries (Elsee Instumentation, 2002). We argue there is an urgent need for a comprehensive assessment of coral cover and health measured alongside key environmental and sediment-related parameters. The key objectives of this study therefore are to: 1) quantify benthic cover and health, 2) compare the prevalence of impaired health in the dominant coral species, and 3) identify key parameters related to sediment delivery that are associated with benthic cover and health along an inshore to offshore gradient. These data will improve our understanding of turbid coral reefs composition and potential resilience to both local and global stressors, and promote current management strategies that aim to protect inshore turbid reefs from future changes to land use.

MATERIALS & METHODS

Study sites

The study was conducted on three low profile submerged patch reefs (Eve’s Garden, Anemone Garden and Siwa Reef) in the MSCRNP (Fig. 1). These sites were of a comparable depth (5-15 m) and size (<0.11 km²), and do not have the typical windward-leeward zonation patterns. As such, benthic composition over the reefs are comparatively homogenous. Eve’s Garden (EG) is a shallow inshore reef close to shore (7.3 km) with a coral community dominated by plate and massive corals such as Pachyseris sp. and Porites sp. (Ferner, 2013). Anemone Garden (AG) is further offshore (11.7 km) and consists of a considerable density of anemone colonies, with plating forms of Acropora sp. and exceptionally large massive Porites sp. and Diploastrea sp. colonies (1-5 m length). Siwa Reef (SW) situated further to the south is the most biologically diverse of the studied reefs consisting of encrusting and massive coral forms (Ferner, 2013). These reefs lie on an inshore to offshore transect from the Baram (sediment influx 2.4 x 10¹⁰...
kg.year\(^{-1}\); Nagarajan et al., 2015) and Miri River mouths, located to the north of EG (10 km from Miri river and 30 km from Baram river).

Physical (temperature, light, turbidity and sediment trap accumulation) and biological (benthic cover, coral health) data were collected at the end of the dry season (15\(^{th}\) September to 20\(^{th}\) October 2016) and during the wet season (11\(^{th}\) May to 3\(^{rd}\) June 2017). At each of the three reefs, six replicate line transects (20 m), separated by 20 m intervals to ensure independence were run across the reef surface (EG = 8-12 m; AG =10-14 m; SW= 8-14 m). These reefs are not characterized by typical windward and leeward reef edges, but are low profile patch reefs where the majority of the reef sits in one relatively flat plane, sloping gently on all sides to the sea floor. As such, all transects were laid out along the same axis across the flat section of each reef.

Physical data collection
Seasonal changes in light (measured with Photosynthetic Irradiance Recording System by Odyssey, New Zealand) and temperature (measured with HOBO Pro V2 loggers, Australia) were recorded every 10 minutes from September 2016 for 9 months (temperature at EG and AG) and 12 months (light at EG). To capture changes in suspended sediment loads over a tidal cycle, turbidity loggers were deployed (in a horizontal position) for two weeks at the end of the 2016 dry season (September; EG and SW) and end of the 2017 wet season (May; EG; AQUA logger 210/310TY, Aquatech, UK). Data on cloud cover, rainfall and wind speed over the period from October 2016 to October 2017 was retrieved from the online database World Wide Weather (2017).

To assess small-scale spatial variation in sediment trap accumulation, four sediment traps per three transects (8 traps in total per reef) were deployed at each reef in September 2016 to collect sediments during the NE monsoon. Each trap consisted of 3 cylindrical PVC plastic containers (diameter of 7.6 cm by 15 cm high) attached to a metal rod positioned 30 cm above the substrate (Storlazzi et al., 2011). The traps remained in-situ until May 2017. To determine if trapped sediments were from local resuspension or transported on to the reef, 500 g of benthic sediment at the base of each trap was sampled. The content of each trap was emptied into a labelled
plastic bag and stored at -20°C until further analysis at the Curtin University Sarawak Laboratory facilities (Laboratory SK2 204), Malaysia.

Sediment samples were analysed for weight and particle size characteristics. Frozen samples were thawed and allowed to settle overnight. Water remaining on the surface was filtered (0.45 nm filter paper) to capture the fine suspended sediments. The sediments (washed, settled and filtered) were oven-dried at 60 °C for 2-3 weeks and weighed to the nearest 0.001 g.

Sedimentation accumulation rate (g cm\(^{-2}\) day\(^{-1}\)) was calculated as the weight of sediment trapped (g) divided by the number of days the trap was deployed and the surface area of the trap (cm\(^2\)).

For the grain size analysis, the settled dry sediments were manually homogenized and weighed before sieving. The sediments were homogenized using a pestle and mortar given the sediments were mostly sand and loosely aggregated. Sediments were separated into 5 class fractions (>1 mm, 500 to <1000 μm, 250 to <500 μm, 125 to <250 μm and 63 to 125 μm) by placing the sieve stack on a mechanical shaker for 20 minutes. Each of the 5 sediment fractions were weighed to the nearest 0.001 g.

**Biological data collection**

**In water data collection**

The benthic cover and coral diversity (to genus level) in September 2016 were assessed using the photographic transect method (Bégin et al., 2013). Photographs were taken using a Canon Powershot G7 mark II digital camera at a fixed height of 0.75 m above the transect line every 1 m along the transect (n = 21). Photographs (0.5 m\(^2\)) were analysed in Coral Point Count (CPCe) with a uniform grid of 25 points to calculate benthic cover for each of 8 categories (hard coral, soft coral, recently dead coral, turf algae, macroalgae, crustose coralline algae, sponge, abiotic substrate) (CPCe; Kohler and Gill, 2006). Recently dead coral (<1 yr) was considered to include dead coral that had visible corallites and limited algae growth. The hard coral category was further subdivided into 38 genera common to the Indo-Pacific region according to Kelley (2009).

To assess seasonal fluctuations in coral reef health, signs of compromised health (disease, bleaching, bioerosion, pigmentation response, excessive mucus production, scars) were recorded in September 2016 and May 2017. The belt transect methodology was used to cover a wider area...
along the transect line via a zig-zag pattern 1 m either side of the transect line (40 m² for each 20 m transect). Coral colonies within each belt transect were identified to genus level and classified as either healthy or affected by an impaired health sign (Beeden et al., 2008). Signs of bioerosion included the presence of organisms such as Christmas tree worms, boring bivalves and sponges, and bleaching was further subdivided into whole, partial, focal and non-focal bleaching (as defined in Beeden et al., 2008). To determine if bleached corals recovered or died, 14 coral colonies from EG and SW in both sampling seasons that showed signs of bleaching were tagged and photographed (4 *Diploastrea* sp., 6 *Pachyseris* sp., 4 *Porites* sp.). The percentage of bleached tissue was assessed from scaled photographs using CPCe software (1=normal, 2=pale, 3=0-20%, 4=20-50%, 5=50-80% and 6=80+% bleached). While this is a low sample size, the data is included to provide further insight into the recovery potential of corals on these reefs.

**Symbiont density and chlorophyll a analysis**

In May 2017, fragments of three coral genera (*Montipora* sp., *Pachyseris* sp. and *Acropora* sp.) were collected from EG, AG and SW for chlorophyll *a* and symbiont density analysis. Higher chlorophyll *a* and symbiont densities are typically recorded on turbid reefs (Browne et al., 2015) as this increases the coral’s ability to photosynthesis under low light levels as they acclimate to suspended sediments (Hennige et al., 2010). Fragments (5-10 cm for branching corals and ~10 x 10 cm for foliose corals) were collected using cutters and placed in plastic bags. Samples were placed on ice during transportation back to the laboratory where they were stored at -80 °C until further analysis. Symbiont density and chlorophyll *a* content were quantified following the removal of coral tissue from the skeleton. The protocol for extracting tissue was adapted from Ben-Haim et al. (2003) (Supplementary material).

**Statistical analysis**

Univariate statistical analysis was conducted in R Studio Desktop version 1.1.383. Prior to analysis, normal distribution and homogeneity of variances were checked using the Shapiro Wilk test and the Levene’s test, respectively. To assess if there were significant differences in benthic cover (hard coral, soft coral, algae) and diversity among sites a one way analysis of variance (ANOVA, n= 6, α= 0.05) was used followed by a Tukey HSD post-hoc test (Bonferroni method), if necessary. Significant differences in the prevalence of compromised health signs
(bleaching, bioerosion, mucus production, pigmentation and scars) among sites and between seasons were identified for both total hard coral cover and for the most abundant coral genera (Porites, Pachyseris, Montipora, Diploastrea, Acropora) using a Full Factorial ANOVA (FF ANOVA, n= 6, α= 0.05) and a Tukey HSD post-hoc test. If required, a log10 transformation was carried out for datasets to meet homogeneity of variance. However, as the bleaching recovery was assessed using a scale, these data were tested using a Wilcoxon test to determine if there had been a significant recovery in tagged coral colonies between years. To determine differences in physiology (chlorophyll a content and zooxanthellae density) between the three coral genera sampled (Acropora n=17, Pachyseris n=13, Montipora n=15) and across sites, a non-parametric Kruskal Wallis test was performed. Furthermore, to evaluate cell health differences between the three genera and among reefs, the percentage of cells from each grade were compared using the Kruskal-Wallis test. Differences in sediment trap accumulation rates was tested among reefs (Kruskal – Wallis). In addition particle size characteristics (median, fine/course fraction) among reefs, and between the trapped sediments and the benthic sediments were also tested (FF ANOVA, n=18).

Permutational multivariate analysis was conducted in PRIMER-7 version 7.0.13. A Distance-based Linear Model (DISTLM) was used to determine how much of the variation in community assemblage (hard coral cover=HCC, soft coral cover=SCC, algae, recently dead coral cover=DCC, H’ index, number of coral genera) among transects and reefs was driven by depth, distance from the two nearby river mouths, distance from shore and differences in sediment trap accumulation rates and particle size characteristics. Depth was included in the analysis as depth is known to influence sediment dynamics (Wolanski et al., 2005) as well as declines in light associated with suspended sediments (Falkowski et al., 1990). A distance-based resemblance matrix was created for the biological data set using Bray- Curtis similarity values following a square-root transformation and for the environmental data using Euclidean distances and normalised values. A DISTLM, using the BEST fit model with the Akaike’s Information Criterion (AIC) and 9,999 permutations was performed using the resemblance matrices. The multivariate scale relationship between the predictor (environmental) and response variables (biological) was presented on a plot with a distance-based redundancy analysis (dbRDA; Legendre and Anderson, 1999). To investigate whether environmental factors contributed to
differences in health status among sites again a DISTLM model was used followed by dbRDA plotting as above. Predictor variables included substrate structure (hard coral cover (HCC), diversity) and physical conditions (depth, sediment trap accumulation rate, particle size characteristics, distance from both river mouths and distance from shore). Hard coral cover (HCC) and diversity were used since higher HCC can contribute to a greater probability of impaired coral health (Bruno and Selig, 2007). In contrast, reefs that are more diverse can lower susceptibility as it reduces the quick spread of a disease (Raymundo et al., 2005; Aeby et al., 2011). As sediment data were obtained at the end of the wet season (May 2017), these were used to explain the 2017 health data. For the 2016 coral health data, which had no associated sediment data, only sampling year, HCC and coral diversity together with distance from shore and rivers were used as explanatory variables.

RESULTS

Physical parameters

The dry season was characterized by less variable, warmer SST’s (mean monthly range = 30.0 to 30.7 °C; sup Fig. 2), greater in-water light penetration (mean monthly range at EG = 156 to 320 μmol photons m\(^{-2}\) s\(^{-1}\)) and reduced rainfall (mean monthly rainfall range = 78 to 166 mm) and cloud cover (Fig. 2). In contrast, the wet season was cooler (mean monthly range = 28 to 30.1 °C) with higher rainfall (mean monthly range 126 to 234 mm) and reduced light levels on the reef (mean monthly range at EG = 19 to 150 μmol photons m\(^{-2}\) s\(^{-1}\)). Wind speeds were also slightly elevated during the wet season months (Fig. 2d). Mean sediment trap accumulation rates following the wet season ranged from 13 to 28 mg cm\(^{-2}\) day\(^{-1}\), with a rate almost three times higher at EG compared to AG and SW (H\(^2\) = 10.3, p<0.005; Fig. 3). Site differences in potential sediment load were also observed during the dry season with higher and more variable turbidity recorded at the nearshore EG reef (mean monthly range = <1 to 24 FTU) than at SW (mean monthly range = 1-7 FTU) located 10 km further south from the large Baram river mouth (sup Fig. 3).

All three reefs were dominated by sand (>98%), with the median particle size of benthic sediments significantly increasing (F\(^2\)=13.6, p<0.005) with distance from the mouths of the Baram and Miri rivers (Fig. 4). Benthic sediments at SW comprised 58% of very coarse sand,
nearly three times that of EG (20%) (F\(^3\)=24.9, p<0.001; PH: SW>EG,AG; sup Fig. 4) and a significantly smaller proportion of medium/fine sands (F\(^2\)=17.2, p=<0.001; PH: SW>AG>EG).

In contrast there was little difference in the median particle size from the sediment traps among sites (F\(_2\)=2.25, p=0.133), although particle sizes of the benthic sediment were significantly greater compared to the trapped sediments (F\(_1\)=60.93, p<0.001).

**Benthic cover**

Hard coral cover increased with distance from the major sediment source (Baram River) and varied significantly among sites (F\(_2\)=5.3, p=0.01; PH: SW>EG). SW had the highest HCC (39.3%) and EG almost half the HCC (21.9%; Fig. 5). Soft corals also varied significantly but declined with increasing distance from the major sediment source (H\(_2\)= 8.6, p=0.01; MWPH: EG>AG, SW) with EG having nearly 15-fold higher cover than SW. Turf algae dominated the algal community and contributed to 52-57% of all reefs’ benthos. However, there was no significant difference in turf algal cover among reefs (F\(_2\)=0.103, p>0.05). Recently dead coral cover was consistently low among sites (4.25%) as was crustose coralline algae, which was typically covered in turf algae.

In total 28 genera were recorded (Table 1). Coral diversity was considerably different among sites (F\(_2\)=4.6, p=0.03; PH: SW>EG) with SW having the highest richness and 25 genera (H’=1.93), and EG and AG 16 and 14 genera, respectively (H’ ~1.4). The surveyed sites were composed of similar communities, with the most dominant genera including *Diploastrea* sp., *Porites* sp., *Montipora* sp., *Favites* sp., *Dipsastrea* sp. and *Pachyseris* sp. (Table 1). All other species comprised a small fraction of the community (<2% cover). Most notable differences in the composition were with the high cover of *Diploastrea* sp. at AG and EG, *Galaxea* sp. at EG, and *Acropora* and *Montipora* sp. at SW.

**Coral reef health**

Of the compromised health signs recorded at each reef, the five most commonly observed were mucus production (0.5 ± 0.3%), pigmentation response (2.2 ± 0.7%), bioerosion (6.6 ± 2%), bleaching (6.7 ± 0.9%) and scars (1.1 ± 0.4%; Fig. 6). No diseases per se were observed except at EG where one colony of massive *Porites* sp. had ulcerative white spots. Despite a decline in
the prevalence of compromised health along an inshore to offshore gradient following the dry season in 2016, total prevalence (sum of the five commonly observed signs) was not statistically significant among sites and seasons (p>0.05; Table 2). The prevalence of mucus production by corals at Eves Garden (5%), however, was nearly five times that of other reefs (F₂=3.6; p<0.05; EG<AG, SW), and SW recorded the lowest levels of pigmentation prevalence (Table 2; F₂=5.3; p<0.05; AG>SW). In contrast, bioerosion was comparatively similar among sites within each season, but increased five-fold from 2.7 ± 0.6% to 10 ±1.3% following the 2017 wet season (Table 1; F=20.2; p<0.001; 2017>2016; Fig 7). During both seasons, overall bleaching prevalence was ≤10% with partially bleached the most common form and whole bleaching the least observed (sup Fig. 5). Bleaching prevalence declined from 8.1 ± 1.4% following the dry season to 5.4 ± 1.1 % after the wet season. Although this decline was not statistically significant (F=3.3; p=0.08), the recovery of bleached corals that had been tagged the year before was significant (p=0.002). The average bleaching scale dropped from 3.9 ± 0.4 to 1.6 ± 0.2 (Fig. 8) with all Diploastrea sp. and Pachyseris sp. colonies recovered by 90-100% in 2017.

Patterns of compromised health differed among five representative coral genera (Acropora sp., Montipora sp., Pachyseris sp., Diploastrea sp. and Porites sp.). Acropora sp. displayed the least signs of stress in both seasons (<3.5%). Porites sp. were the most compromised (2016 = 50.8 ± 6%; 2017 = 72 ± 5%; Fig. 9) and the only coral genera with a significant increase in stress symptoms (p=0.004), because of a 40% increase in bioerosion after the wet season (F₁=10.17; p<0.001; Table 3). Montipora sp. and Diploastrea sp. also suffered from an increase in bioerosion between sampling seasons, although this was not statistically significant (p>0.05; Table 3). Despite a slight increase in the number of bleached Porites sp. corals, bleaching occurrence for the other four corals declined, most notably for Pachyseris sp. (55% to 3%; F₁=9.03; p=0.008). Furthermore, the most abundant genus Porites sp. was the only coral to show elevated signs of pigmentation (>10%) although this health sign was less prevalent at SW, the most offshore site (F²=5.3; p=0.01; Table 3).

For the three coral genera, Montipora sp., Pachyseris sp. and Acropora sp., there was no significant difference in symbiont density (H = 4.0397, df=2, p>0.05) and chlorophyll a among sites (H = 2.3769, p>0.05) although SW scored the highest of both measures (3.2*10⁶ ±5.5
Symbiont density differed among the three coral genera (chi-square = 23.1, df=2, p<0.001; MWPH: AC>MT,PH) with Acropora sp. scoring four and five times higher symbiont densities (sup Fig. 6). Over 50% of the symbions observed were healthy (stage 1; sup Fig. 7a) with slightly more healthy cells observed at SW (H=1.7, p>0.05) and marginally more degraded cells (stage 5) observed at AG (H=3.4, p>0.05). Among genera, Acropora had a greater number of healthy cells (69 ±3.9%) than both Montipora (49.4±5) and Pachyseris (52.6 ± 4.8; H= 14.4, p<0.001; sup Fig. 7b).

**Environmental associations with benthic cover and health**

Environmental variables (depth, sediment trap accumulation rate, distances from shore/river mouth, concentration of silt/fine/coarse particles, median particle size) explained 62.5% of the variation in benthic composition among reefs. Key drivers (p<0.05) were distance from river mouth (30.3%) and shore (1%), median particle size (16.4%), and sediment trap accumulation rate (2.3%; Table 4). Variability among sites was higher than within, with sediment trap accumulation rate and particle size a key driver of benthos at EG and AG, and distance of river and shore more closely associated with SW (Fig. 11).

To determine key drivers of coral health, two DistLM models were run. The first model included health data from both sampling seasons, with six explanatory variables (season, HCC, diversity, distance from river mouth and shore, and depth). The second model included health data and sediment related variables following the wet season and sediment trap contents (sediment trap accumulation rate, concentration of silt/fine/course sediments, median particle size). For the first model, year, HCC and diversity significantly explained <31% of the variation in coral health among transects and sites (Table 5). Sites within a sampling season were separated along a HCC and diversity gradient (Fig. 12), with transects at SW typically characterised by higher HCC and diversity but lower prevalence of scars, pigmentation and bleaching (sup Fig. 8). Furthermore, repeat transects were separated between seasons, with those completed in 2017 recording higher bioerosion, but lower bleaching and pigmentation (sup Fig. 7), supporting our previous results. Of the sediment drivers, the BEST model included both silt and the course sediments, which explained 18% of the variations in coral health in 2017. Higher sediment trap accumulation rates, although not statistically significant (p=0.06; Table 5),
explained 7% of the variation in health, and were most often associated with higher prevalence of pigmentation, bioerosion and bleaching (sup Fig. 9).

**DISCUSSION**

The three reef sites in the MSCRNP are characterised by healthy coral cover yet low coral diversity. Average live coral cover among the three reefs was 30%, ranging from 22% at EG to 39% at Siwa Reef. This is lower than reefs to the north in Sabah, with reports of live coral cover from 23 to 75% (Pilcher and Cabanban, 2000; Chou and Tun, 2002; Lee, 2007; Praveena et al., 2012; Waheed et al., 2016), but greater than the average coral cover for the wider Pacific region, estimated at 22% in 2003 (Bruno and Selig, 2007). Previous assessment of coral cover in the early 2000s on the Miri reefs range from 28% (Pilcher and Cabanban, 2000) to 22-58% (Elcee Instumentation, 2002). Although the higher coral cover reported by the latter study is most likely an artefact of the methodology used (ex-situ Acoustic Ground Discrimination System), which can result in the misidentification and, therefore, quantification of live coral cover. Regardless, our data suggest that coral cover at Miri’s reefs has been relatively stable over the last two decades. Miri’s coral cover is comparable to both turbid and clear water reefs (Roy and Smith, 1971; Loya, 1976; Larcombe et al., 2001; Wesseling et al., 2001; Palmer et al., 2010; Goodkin et al., 2011), yet diversity was comparatively low (14 to 25 genera per reef) for the Coral Triangle region. Turak and Devantier (2010) reported 391 coral species (~70 genera) on reefs near Brunei (~80 km from Miri), and Teh and Cabanban (2007) reported 120 species within 71 hard coral genera for Banggi Island in Sabah. A comprehensive biodiversity assessment of all 30 reefs with the MSCRNP in 2001 reported 66 genera (203 coral species; Elcee Instumentation, 2002). We only observed a third of the number of coral genera (n=28), which is expected given we surveyed only 10% (n=3) of the reefs surveyed in 2001. However, this report also found that coral diversity was highly variable among reefs, with an average of nine coral genera per transect. It is likely that MSCRNP reefs found further to the south and in deeper (15-35 m) offshore waters but outside the scope of this study (characterized by different environmental conditions) include several coral species not observed at our shallow nearshore sites, which are influenced by terrestrial sedimentation from both natural and anthropogenic processes.
Low diversity at the surveyed sites is likely the result of poor water quality in the nearshore shallow coastal zone. The inshore reefs of Miri are found in a narrow depth range between 7 to 15 m, hence there is a complete lack of reef structure at 1-5 m. These very shallow depths, however, are often characterised by a distinct set of coral species (Morgan et al., 2016; DeVantier and Turak, 2017). Given that lack of reef structure at these shallow depths in Miri, this in part may explain lower coral diversity than on reefs to the north in Brunei and Sabah that have reached sea level. But these inshore reefs are also characterised by high levels of terrigenous sediments, which can also reduce coral diversity (Rogers, 1990; Fabricius, 2005).

High sediment input from rivers are typically correlated with high nutrient loads that can lead to increase in reef algal biomass (De'ath et al., 2012). Algal cover on all three reefs was high (>50%) compared to reefs in northern Borneo (0 to 29%; Waheed et al., 2015), and will most likely be competing with corals for reef space. Some coral taxa will be less resilient to both sediments and algal competition resulting in lower coral diversity (Fabricius et al., 2005; De'ath and Fabricius, 2010). In Indonesia, Edinger et al. (1998), recorded lowest coral diversity on reefs with algae cover reaching 46%. Reduced diversity was also attributed to land pollution as well as destructive and over-fishing practices that destroy the reef structure and reduce fish biomass thereby removing the top-down control on algal growth by herbivore browsers (Hughes, 1994; Rogers and Miller, 2006; De'ath and Fabricius, 2010). In Miri, overfishing and poor land management practices have been a long-term concern for the regional government (Elcee Instumentation, 2002) but there are limited funds to actively protect the reefs (Teh and Teh, 2014) and collect data on these impacts.

Low coral diversity does not necessarily suggest a degraded reef condition. Typically, low diversity in nature results in lower resilience (Raymundo et al., 2005) and community stability (Bellwood et al., 2004). Yet there is growing evidence to suggest that a few but tolerant species can maintain reef resilience to local and global impacts, and implies that the diversity-resilience links need further investigation (Bellwood et al., 2004; Fabricius et al., 2005; Nystrom et al., 2008). A recent study on relatively undisturbed and well protected reefs in the Philippines that looked to identify site specific benchmarks for coral diversity, measured high coral cover (>30%) at the majority of sites, but low generic diversity (10 to 25 coral genera per 75 m by 25 m area; Licuanan et al., 2017). This highlights that high diversity is not necessarily a key benchmark for
a healthy reef system. As well as assessing diversity on a reef, it is important to determine if and how coral community structure has changed over time. Significant shifts in coral composition can affect the reef’s ecological function such as framework building, habitat complexity and food source diversity (Aronson et al., 2004; Pratchett, 2005; Graham et al., 2006). At six reef sites on the Great Barrier Reef coral communities shifted over 12 years towards a high abundance of *Porites* spp. and soft corals; a community assemblage that is less likely to re-establish to the pre-disturbance coral assemblage (Johns et al., 2014). Inshore reefs in Miri are similarly dominated by massive corals including *Porites* sp. and *Diploastrea* sp., with some (e.g. EG) also characterized by high soft coral cover (>10%). While we have no long-term data sets to evaluate change in both diversity and composition, Miri’s reefs may have experienced a community shift due to reduction in water quality. Yet our tendency as coral reef ecologists to focus on coral cover, composition and diversity, has resulted in a misconception as to what constitutes an overall healthy reef (Vroom, 2011). Some reefs may naturally be dominated by non-coral organisms, such as calcifying algae that are equally important for reef accretion and stability, but possibly less resilient to climate change. Thus our perception of the reefs current state and its future trajectory are likely inaccurate and need adjusting to go beyond diversity assessments.

The MSCRNP reef community can best be described as representative of turbid reefs in the Indo-Pacific. The dominant coral species include several genera (*Acropora*, *Montipora*, *Porites*, *Pachyseris*, *Favites* and *Galaxea* spp.) that have been observed on nearshore reefs in Singapore (Chou, 1988; Dikou and van Woesik, 2006), the Great Barrier Reef (GBR) (Ayling and Ayling, 1991; Larcombe et al., 2001; Browne et al., 2010; Morgan et al., 2016), Thailand (Tudhope and Scoffin, 1994), Hong Kong (Goodkin et al., 2011) and Sabah (Pilcher and Cabanban, 2000). These corals are considered to be more resilient to sediment influx either through: 1) enhanced photo-acclamatory abilities required during periods of low light (e.g. *Stylophora*; Dubinsky et al., 1984; Browne et al., 2014), 2) active sediment removal processes by the coral polyp (e.g. *Goniastrea*; Rogers, 1990; Erftemeijer et al., 2012), 3) enhanced mucus production to remove settled sediments (e.g. *Porites*; Bessell-Browne et al., 2017) or, 4) morphological advantages that result in greater degree of vertical growth thereby reducing tissue mortality from sediment burial (e.g. *Acropora* and *Montipora*; Erftemeijer et al., 2012). There were also distinct differences in the community assemblages particularly between Siwa Reef and Eve’s Garden. Siwa Reef had a
mixed assemblage of branching, foliose and massive corals, whereas Eve’s Garden was
dominated by massive corals, such as *Porites* sp. and *Diploastrea* sp. These coral community
differences suggest there are significant differences in environmental drivers (including
sediments) over a comparatively small spatial scale (10 km’s).

The inshore to offshore gradient in hard coral cover and diversity, and differences in coral
composition is heavily influenced by the spatial differences in sediment related parameters. Over
62% of the variation in benthic cover at our three reef sites is explained by differences in depth,
sediment trap accumulation rates and distance from sediment sources as well as sediment particle
size characteristics. Consequently, we saw a significant increase in both coral cover and diversity
with increasing distance from the river mouths. Similar observations have been reported from
Indonesia and Puerto Rico, where hard coral cover nearly halved towards shore (Loya, 1976;
Edinger et al., 2000), and in Hong Kong, where inshore coral cover was 20% lower than offshore
(Goodkin et al., 2011). Reduced coral cover occurs because of low larval recruitment as a
consequence of limited hard substrate following sediment settling (Birrell et al., 2005; Fabricius,
2005; Dikou and van Woesik, 2006), colony mortality caused by anoxic conditions that occur
under sediment layers (Rogers, 1983; Riegl and Branch, 1995; Wesseling et al., 2001), or lower
coral growth as a result of limited light or more energy used for sediment clearing (Browne,
2012). The sediment particle size and source (marine versus terrestrial) are considered equally
important to sediment volume in assessing the impacts of sediments on coral health (Weber et
al., 2006). A recent study demonstrated that as the percentage of terrestrial sediments increases,
there are greater declines in coral cover (Bégin et al., 2016). The significantly lower hard coral
cover and diversity at EG than at SW could be driven in part by a higher volume of terrestrial
sediments from the Baram and Miri rivers. Although we did not assess sediment origin,
sediment trap accumulation rates at EG were over double that at AG and SW, which may be due
to the reefs closer proximity to the two river mouths. However, it could also be the result of
increased sediment resuspension in shallow water or a combination of these factors. Sediment
traps do not provide a comprehensive assessment of sediment dynamics on reefs, and given that
our traps were out for 7 months, we recognize that our monthly sediment trap accumulation rates
can only be compared among our study sites and not to other studies. Regardless, it is likely that
river flow and sediments are influencing reef health, but these reefs appear to be in a temporally
stable state given low recently dead coral cover (4.35%) and the limited decline in coral cover over the last two decades.

The prevalence of impaired health signs was dominated by bioerosion and pigmentation response with no signs of coral disease (with one exception). These health indicators are typically related to high sediment and nutrient influx. High levels of bioerosion in particular has been linked to land based pollution whereby lower light, from high turbidity, reduces CaCO$_3$ density (Risk and Sammarco, 1991; Lough and Barnes, 1992) weakening the coral skeleton and increasing susceptibility to bioeroders (e.g. molluscs, worms etc.; Prouty et al., 2017). Furthermore, even modest increases in nutrient levels can lead to an increase in the abundance of bioeroding organisms shifting a reef community from one of net production to net erosion (Hallock and Schlager, 1986; Hallock, 1988; Prouty et al., 2017). Bioerosion levels were significantly greater following the wet season when the impact of sediments on the Miri reefs were elevated as indicated by declines in light and higher suspended sediment loads. Conversely, pigmentation rates were higher following the dry season. Pigmentation is an indicator of immune function in response to a stressor (Willis et al., 2004; Palmer et al., 2009). These stressors have been related to settling sediments (Pollock et al., 2014) or lesions from abrasion or scars (Willis et al., 2004), or for the case of Miri reefs elevated SST’s recorded in the region in 2016 leading to the moderate bleaching event as observed by the diving operators and fisherman. Spatially, pigmentation rates were significantly lower at Siwa, which may suggest that corals at the least sediment impacted site were also less stressed than at AG and EG. Sediments can also promote diseases in corals (Voss and Richardson, 2006; Haapkyla et al., 2011; Pollock et al., 2014) with Black Band Disease and White Plague widely observed in the Indo-Pacific (Harvell et al., 2007; Beeden et al., 2008), although generally low (~8% of current global records) in SE Asian reefs compared to the Caribbean (Green and Bruckner, 2000). Suggested explanations for this include poor reporting of coral diseases and relatively high coral diversity that might aid in diminishing a quick spread of a disease (Raymundo et al., 2005). At Miri, the more likely explanation of low to no coral diseases are more resilient individual corals and coral species, and potentially limited connectivity with nearby coral populations, although this remains speculative until further work is carried out.
Hard coral cover and diversity also explained a significant portion of the variation in coral health. Miri reefs with a higher frequency of impaired health at sites recorded less coral cover and diversity. In a recent study by Miller et al., (2015) reefs in Sabah had four common coral diseases at varying frequencies (<0.1 to 0.6 per affected colonies in an m²) and signs of tissue necrosis and pigmentation responses. They found a positive correlation between disease frequency and coral cover, which suggested that host density was a key driver of disease prevalence and compromised health. This relationship is due to reduced distances between colonies, and greater shading and competition by fast growing species as coral cover increases (Bruno and Selig, 2007). In Miri, we see the reverse trend suggesting that factors other than host density are driving coral health, potentially changing sediment loads and finer sediment particles not present in Sabah. However other variables often associated with sediment such as nutrient levels and pollution such as heavy metal loads are also worth investigating.

Variable species composition among sites would also partly explain the spatial variation in coral health. Different coral taxa have different susceptibilities to bioerosion, bleaching, disease and compromised health (Raymundo et al., 2005; Couch et al., 2014; Heintz et al., 2015). In Miri signs of pigmentation and bioerosion were most prominent on massive Porites sp. colonies. Porites sp., although typically considered a harder coral taxa (Raymundo et al., 2005) tolerant of turbid waters, often have the most lesions, highest tissue loss and pigmentation response (Tribollet et al., 2011; Pollock et al., 2014; Heintz et al., 2015) as well as being a target for disease (Raymundo et al., 2005). The level of bleaching observed in Porites at Miri was comparable to other abundant coral genera, but recovery potential after 9 months was lower, possibly due to other stress symptoms. Bleaching was the most common sign of impaired health among coral taxa, most commonly observed in Pachyseris, Porites, Montipora, Dipsastrea and Acropora spp. (in declining order). A comprehensive study by Marshall and Baird (2000) of 40 coral taxa on the GBR found the same coral taxa were highly (>50% bleached or dead) or severely (>15% dead) susceptible to thermal stress. In contrast, the other five most abundant corals at Miri (Diploastrea, Favites, Galaxea, Echinopora, Merulina spp.) are considered to be less sensitive to rising SST’s (Marshall and Baird, 2000; Guest et al., 2016). However, bleaching susceptibility does vary considerably according to the thermal history of a region. For example, Acropora sp. is susceptible to bleaching on some reefs (Marshall and Baird, 2000; Pratchett et
al., 2013; Hoogenboom et al., 2017), but was less susceptible on other reefs (e.g. Singapore following the 2010 bleaching event: Guest et al., 2012). Only ~5% of Acropora sp. colonies in Miri showed signs of thermal stress, which suggests moderate thermal tolerance to high SST’s. High levels of algal density in coral tissue are also linked to higher thermal stress resistance (Glynn, 1993; Stimson et al., 2002) due to the symbionts providing a greater concentration of mycosporine-like amino acids that protect corals from UV radiation (Xu et al., 2017). Symbiont densities measured at Miri were high (mean = 2.4*10^6 cells per cm^2) but comparable to corals on other turbid reefs like those from Singapore (e.g. 0.5 to 3*10^6 cells per cm^2 (Browne et al., 2015). However, it was Acropora sp. that had significantly higher symbiont density than the more frequently bleached Montipora sp. and Pachyseris sp. Our results suggest that resilience to stress for these corals is a complex, but synergistic relationship between the level and frequency of environmental stressors, community composition and a coral’s adaptability to increased SST.

In 2016, a severe coral bleaching event occurred throughout the Indo-Pacific region. The impacts of this event were thoroughly assessed on the GBR, where over 90% of reefs bleached resulting in the loss of 29% of shallow water coral cover (Great Barrier Reef Marine Park Authority, 2016). In January to March 2016, SST along the northern shore of Borneo were in the highest 10% of global records since 1990 (Great Barrier Reef Marine Park Authority, 2016). SST reported by NOAA for Brunei peaked in May to June 2016 at 31°C (the bleaching threshold temperature; Fig. 12). During this time there was 1 to 2.5 Degree Heating Weeks (DHW) and mid-level bleaching warnings. SST remained at ~30°C until January 2017 (National Oceanic & Atmospheric Administration, 2018), which agree with our in-water assessment of SST during September 2016 to early 2017 (sup Fig. 2). This suggests that while corals at Miri were subject to elevated SST’s for 5 or more months, our surveys revealed comparatively low bleaching rates (~10% of colonies bleached), and high recovery rates (as suggested by the tagged corals; >90%). This suggests that these nearshore turbid water reefs are resilient to high SST’s supporting the growing body of evidence that turbid reefs bleach less severely and frequently than their clear-water counterparts (Marshall and Baird, 2000; Heintz et al., 2015; Morgan et al., 2017). Low bleaching and high recovery rates of Miri reefs is possibly due to nearshore coral assemblages more frequent exposure to higher temperatures than their offshore deeper conspecifics, resulting in the development of adaptive mechanisms (Marshall and Baird, 2000; Guinotte et al., 2003;
Guest et al., 2016; Morgan et al., 2017). It may also be due to lower UV light penetration that
602 can exacerbate temperature stress (Courtial et al., 2017), or potentially from higher heterotrophy,
603 which increases the supply of essential metals to the symbionts thus sustaining them through
604 elevated temperatures (Ferrier-Pagès et al., 2018). This study further suggests that while turbid
605 reefs are potentially more resilient to elevated SST, the mechanism/s responsible for this
606 resilience remain unclear.

CONCLUSIONS
609 In conclusion, the MSCRNP reefs are characterized by relatively high coral cover, low
610 prevalence of impaired health and are composed of a few but tolerant coral taxa. Low recently
611 dead coral cover and potentially limited decline in coral cover over the last two decades indicates
612 these reefs are stable despite elevated sediment inputs and regular exposure to thermal stress
613 events. There are, however, potential risks from proposed coastal and in-land developments
614 given we found that sediment related parameters have resulted in an on- to offshore gradient in
615 coral cover, diversity and health. Furthermore, high bioerosion and algae cover indirectly
616 suggests high nutrient influx, most likely from the Baram River. The high prevalence of
617 bioerosion observed in *Porites* sp. colonies is a concern given that this coral is a key reef
618 framework builder, and any notable declines in *Porites* sp. health will reduce coral reef
619 complexity and habitat availability for other invertebrate and fish species. Currently, there is no
620 baseline data on spatial and temporal changes in river outputs and sediment plume dynamics
621 within the MSCRNP, which is crucial in evaluating future threats to these reefs. Local
622 management agencies will need to address this knowledge gap if they plan to develop strategies
623 that address the potential impacts of changing land use on MSCRNP. The reefs current health
624 state and elevated stress tolerance does, however give hope that these reefs could be resilient to
625 future climate change but only if local water quality does not deteriorate.

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Figure 1

Map of south China Sea with enlarged map of study area, showing locations of the three reefs, Miri city and the closest rivers.

(Image credit Hedwig Krawczyk modified from Natural Earth - Free vector and raster map data).
Figure 2

Average monthly data for A. light, B. wind speeds, C. cloud cover, and D. rain fall.

Light data was collected at EG as part of this study whereas wind, cloud and cover data was taken from the worldwideweatheronline.com website (error bars = SE).
Figure 3

Average sedimentation rates at the three surveyed sites (error bars = SE).
Figure 4

Particle size data from the sediment traps and the benthos at EG, AG and SW.

![Graph showing particle size distribution in different locations.](image)
Figure 5

Average percentage benthic cover at EG, AG and SW. Sites are organised from inshore to offshore (error bars = SE).
Figure 6

Signs of impaired health

A. Excessive mucus and slothing, B. Feeding scars from parrotfish, C. Christmas tree worms and bivalves, D. pigmentation response in *Porites* sp. E. Non-focal bleaching, and F. Partial bleaching. Black arrow points to associated impaired health sign.
Figure 7

Average prevalence of the dominant signs of impaired health across all three surveyed sites (EG, AG, SW) following the 2016 dry season and 2017 wet season (error bars = SE).
Figure 8

Average bleaching scale for the three coral genus across the three survey sites (EG, AG, SW) that were tagged in September 2016 following the warm dry season and cooler wet season (error bars = SE).

(1=normal, 2=pale, 3=0-20% bleached, 3=21-50% bleached, 4=51-80% bleached, 5=81-100% bleached)
Figure 9

Prevalence of the most common impaired health signs following the 2016 dry season and the 2017 wet season for the five most common observed coral genus across all three sites surveyed (EG, AG< SW).
Figure 10

Average symbiont density (A) and chlorophyll a pigment density (B) across the three coral species assessed (*Acropora*, *Monitpora* and *Pachyseris*) at EG, AG and SW (error bars = SE).
Figure 11

Distance-based redundancy analysis (dbRDA) plot with an AIC criterion selection illustrating the significant environmental factors (p<0.05) that influence community composition at EG, AG and SW.

The length and direction of the vectors represent the strength of the correlation (circle denotes a correlation of 1) and direction (+/-) of the relationship with transects (points plotted) at each site.
Figure 12

Distance-based redundancy analysis (dbRDA) plot with an AIC criterion selection illustrating the that influence coral health at at EG, AG and SW.

Significant explanatory variables (p<0.05; HCC = hard coral cover, diversity = coral diversity, year = Sept 2016 and May 2017). The length and direction of the vectors represent the strength and direction (+/-) of the relationship with transects (points plotted) at each site. (Image credit: Amitay Moody).
Table 1 (on next page)

Average (%) coral cover of the 28 genera observed at the three surveyed reefs illustrating the 10 most dominant coral genus.
| Genus                | Eve’s Garden | Anemone’s Garden | Siwa reef |
|---------------------|-------------|------------------|-----------|
| Acropora (branching)| 0.07 ± 0.07 | 2.60 ± 0.40      |           |
| Diploastrea (massive)| 14.80 ± 1.60| 10.60 ± 3.70    | 0.40 ± 0.10 |
| Echinopora (encrusting)| 0.50 ± 0.14  | 1.90 ± 1.60     |           |
| Dipsastrea          | 0.90 ± 0.30 | 3.44 ± 0.40      | 3.60 ± 2.00 |
| Favites (massive)   | 1.70 ± 0.80 | 2.40 ± 0.86      | 5.10 ± 1.60 |
| Galaxea             | 3.00 ± 1    | 0.62 ± 0.20      | 0.90 ± 0.30 |
| Merulina            | 1.60 ± 1.5  | 0.10 ± 0.03      | 1.33 ± 0.80 |
| Montipora (plate)   | 1.30 ± 100  | 2.09 ± 1.10      | 8.60 ± 3.00 |
| Pachyseris (plate)  | 2.10 ± 1.10 | 0.50 ± 0.30      | 2.00 ± 1.30 |
| Porites (massive/plate) | 5.70 ± 2.80 | 7.30 ± 1.50      | 7.30 ± 2.30 |
| Astreopora          |             |                  | 0.90 ± 0.60 |
| Caulastrea          | 0.07 ± 0.19 | 0.04 ± 0.04      |           |
| Ctenaxis (solitary) | 0.07 ± 0.07 | 0.62 ± 0.15      | 0.14 ± 0.09 |
| Echinophyllia       | 0.30 ± 0.30 |                  | 0.06 ± 0.06 |
| Fungia              |             |                  | 0.10 ± 0.01 |
| Goniatra            |             | 0.10 ± 0.03      | 0.04 ± 0.04 |
| Goniopora           | 0.03 ± 0.03 |                  |           |
| Heliofungia         | 0.10 ± 0.10 |                  |           |
| Leptoria            | 0.03 ± 0.03 |                  | 0.08 ± 0.08 |
| Leptoseris          | 0.17 ± 0.17 |                  | 1.60 ± 1.50 |
| Montastrea          |             | 0.04 ± 0.04      |           |
| Oxypora             | 0.03 ± 0.03 |                  | 0.17 ± 0.17 |
| Pectinia            |             | 0.08 ± 0.08      |           |
| Physogryra          |             | 0.17 ± 0.17      |           |
| Platgyra (massive)  | 0.90 ± 0.80 | 1.79 ± 1.60      | 0.60 ± 0.40 |
| Psammocora          | 0.10 ± 0.10 |                  |           |
| Symbiophyllia       | 0.40 ± 0.20 |                  | 0.69 ± 0.30 |
| Turbinaria          |             | 0.68 ± 0.68      |           |
Statistical results from two-way ANOVA of the total impaired health and each impaired health indicator with site (EG = Eves Garden, AG = Anenomes Garden, SW = Siwa) and season (2016, 2017), and the interaction.

Table 2 (on next page)
| Health sign            | Factor       | df | F value | p value | Post hoc       |
|------------------------|--------------|----|---------|---------|----------------|
| Total impaired health  | Site         | 2  | 0.25    | 0.780   |                |
|                        | Season       | 1  | 1.11    | 0.300   |                |
|                        | Site*Season  | 2  | 0.15    | 0.860   |                |
| Bleaching              | Site         | 2  | 0.19    | 0.830   |                |
|                        | Season       | 1  | 3.30    | 0.080   |                |
|                        | Site*Season  | 2  | 0.69    | 0.510   |                |
| Mucus                  | Site         | 2  | 3.60    | **0.040** | EG<AG,SW      |
|                        | Season       | 1  | 0.15    | 0.700   |                |
|                        | Site*Season  | 2  | 7.20    | **0.003** |                |
| Bioerosion             | Site         | 2  | 0.87    | 0.430   |                |
|                        | Season       | 1  | 20.20   | **<0.001** | 2017>2016    |
|                        | Site*Season  | 2  | 3.80    | 0.040   |                |
| Pimentation            | Site         | 2  | 5.30    | **0.010** | AG>SW         |
|                        | Season       | 1  | 1.00    | 0.320   |                |
|                        | Site*Season  | 2  | 0.82    | 0.440   |                |
| Scars                  | Site         | 2  | 0.10    | 0.910   |                |
|                        | Season       | 1  | 0.33    | 0.570   |                |
|                        | Site*Season  | 2  | 2.59    | 0.090   |                |
Statistical results from two-way ANOVA of the total impaired health and each impaired health indicator for the 5 most dominant coral genera with site and season and the interaction.

If impaired health result is missing then it was not observed for that coral genus. Sites; EG = Eves Garden, AG = Anenomes Garden, SW = Siwa: Seasons; 2016, 2017.
| Species  | Health sign | Factor      | df  | F value | p value | Post hoc     |
|----------|-------------|-------------|-----|---------|---------|--------------|
| Porites  | Total       | Site        | 2   | 1.71    | 0.202   |              |
|          |             | Year        | 1   | 10.17   | **0.004** | 2017>2016   |
|          |             | Site*year   | 2   | 4.00    | **0.031** |              |
| Bleaching| Site        | 2           | 0.36| 0.701   |         |              |
|          | Year        | 1           | 0.08| 0.774   |         |              |
|          | Site*year   | 2           | 1.81| 0.185   |         |              |
|          | Mucus       | Site        | 2   | 6.72    | **0.034** | EG>SW        |
|          |             | Year        | 1   | 2.64    | 0.104   |              |
|          |             | Site*year   | 2   | 1.61    | 0.219   |              |
|          | Bioerosion  | Site        | 2   | 8.79    | **0.001** | 2016>2017   |
|          |             | Year        | 1   | 21.79   | <0.001  |              |
|          |             | Site*year   | 2   | 6.29    | **0.006** |              |
|          | Pimentation | Site        | 2   | 2.64    | 0.126   |              |
|          |             | Year        | 1   | 2.49    | 0.104   |              |
|          |             | Site*year   | 2   | 2.09    | 0.145   |              |
|          | Scars       | Site        | 2   | 0.46    | 0.637   |              |
|          |             | Year        | 1   | 0.38    | 0.543   |              |
|          |             | Site*year   | 2   | 2.25    | 0.126   |              |
|          | Pachyseris  | Site        | 2   | 0.30    | 0.744   |              |
|          |             | Year        | 1   | 9.02    | **0.008** | 2016>2017   |
|          |             | Site*year   | 2   | 0.14    | 0.869   |              |
|          | Bleaching   | Site        | 2   | 0.37    | 0.699   |              |
|          |             | Year        | 1   | 9.69    | **0.006** | 2016>2017   |
|          |             | Site*year   | 2   | 0.11    | 0.897   |              |
|          | Bioerosion  | Site        | 2   | 0.49    | 0.622   |              |
|          |             | Year        | 1   | 1.42    | 0.249   |              |
|          |             | Site*year   | 2   | 0.39    | 0.685   |              |
| Montipora| Total       | Site        | 2   | 0.77    | 0.476   |              |
|          |             | Year        | 1   | 1.65    | 0.211   |              |
|          |             | Site*year   | 2   | 1.45    | 0.254   |              |
|          | Bleaching   | Site        | 2   | 2.06    | 0.149   |              |
|          |             | Year        | 1   | 0.29    | 0.594   |              |
|          |             | Site*year   | 2   | 0.73    | 0.494   |              |
|          | Bioerosion  | Site        | 2   | 0.83    | 0.449   |              |
|          |             | Year        | 1   | 0.83    | 0.371   |              |
|          |             | Site*year   | 2   | 0.68    | 0.519   |              |
| Diploastrea| Total    | Site        | 2   | 0.66    | 0.527   |              |
|          |             | Year        | 1   | 0.10    | 0.752   |              |
|          |             | Site*year   | 2   | 2.54    | 0.104   |              |
|          | Bleaching   | Site        | 2   | 0.63    | 0.541   |              |
|          |             | Year        | 1   | 1.69    | 0.209   |              |
|          | Mucus       | Site        | 2   | 0.58    | 0.570   |              |
|                |       |      |      |
|----------------|-------|------|------|
| **Bioerosion** |       |      |      |
| Year           | 1     | 2.75 | 0.113|
| Site*year      | 2     | 0.71 | 0.502|
| Site           | 2     | 1.64 | 0.220|
| Year           | 1     | 0.86 | 0.364|
| Site*year      | 2     | 0.99 | 0.391|
| **Acropora**   |       |      |      |
| Total          |       |      |      |
| Site           | 2     | 1.92 | 0.171|
| Year           | 1     | 0.22 | 0.644|
| Site*year      | 2     | 0.14 | 0.872|
| Bleaching      |       |      |      |
| Site           | 2     | 1.27 | 0.300|
| Year           | 1     | 1.02 | 0.323|
| Site*year      | 2     | 0.64 | 0.538|
**Table 4** (on next page)

PERMANOVA results highlighting the significant drivers that explain variation in benthic community assemblage among reefs in 2017
| Explanatory variable      | p value | Pseudo-F | $R^2$ |
|---------------------------|---------|----------|-------|
| Depth                     | 0.094   | 2.3      | 0.010 |
| Dist. Baram River         | **0.002** | 7.0      | 0.303 |
| Dist. Shore               | **0.007** | 5.1      | 0.008 |
| Sedimentation rate        | **0.025** | 3.9      | 0.023 |
| Course sediments          | 0.069   | 2.7      | 0.001 |
| Fine sediments            | 0.070   | 2.7      | 0.100 |
| Silt                      | 0.153   | 1.9      | 0.015 |
| Median particle size      | **0.010** | 5.0      | 0.164 |
Table 5 (on next page)

PERMANOVA results highlighting the significant drivers in coral health.

The top panel are the results of a DistLM that includes substrate structure and physical conditions among reefs and across both sampling seasons, and the bottom panel are the results of a DistLM that includes data from the sediment traps among reefs in 2017 only.
### Explanatory variable

| Parameter                  | p value | F    | R²  |
|----------------------------|---------|------|-----|
| Year                       | 0.003   | 5.0  | 0.128 |
| HCC                        | 0.042   | 2.8  | 0.052 |
| Diversity                  | 0.003   | 5.1  | 0.129 |
| Dist. Baram River          | 0.304   | 1.3  | 0.019 |
| Dist. Shore                | 0.521   | 0.8  | 0.020 |
| Depth                      | 0.467   | 0.9  | 0.017 |

### Sediment variable

| Parameter                  | p value | F    | R²  |
|----------------------------|---------|------|-----|
| Sedimentation rate         | 0.059   | 2.4  | 0.070 |
| Course sediments           | 0.031   | 2.9  | 0.152 |
| Fine sediments             | 0.031   | 2.9  | 0.030 |
| Silt                       | 0.067   | 2.3  | 0.110 |
| Median particle size       | 0.083   | 2.2  | 0.024 |