Evaluation of dose dependent analgesic response by extracts of *Myristica fragrans* on albino wistar rats: an experimental study

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ABSTRACT

**Background:** The objective of the study was to evaluate analgesic activity of ethanolic extract, methanol and benzene fraction of *Myristica fragrans* on wistar albino rats.

**Methods:** The present study was carried out in the department of pharmacology JNMC AMU and F.H. Medical College, Agra. The analgesic activity was evaluated by employing the Eddy’s hot plate method and tail flick response method. In both the tests, Rats of either sex weighing 150-200 g were used. The total number of animals n=36 were allocated to six groups. Each group consist of six animals each. The response noted in animals that were tested by hot plate method was reaction time for licking/biting of both the paws before and after administration of control & test drugs. However in Tail flick test, the pain threshold response was recorded before and after administration of control & test drugs. The statistical analysis was done by using one-way ANOVA. The data is expressed as Mean±SEM. P<0.05 was considered to be statistically significant.

**Results:** Ethanolic extracts and methanol fraction of *M. fragrans* showed statistically significant (p<0.001) increase in reaction time for licking/biting in hot plate method. On the contrary a significant increase in pain threshold was also recorded in tail flick response test. It is interesting to note that no significant degree of analgesia related to any dose of benzene fraction was observed.

**Conclusions:** The present study reveals the dose dependent significant analgesic activity of the extracts of *M. fragrans* i.e. ethanolic extracts and methanol fraction in both the test. However, the degree of analgesia was recorded significantly higher in groups received higher doses of extracts of *M. fragrans*.

**Keywords:** Analgesic activity, *Myristica fragrans*, Albino wistar rats

INTRODUCTION

Pain is a common subjective phenomenon which brings a patient to physician. It is associated with a number of diseases and is estimated that 80–100% of the population experience back pain at least once in the life time.¹ Non-steroidal anti-inflammatory drugs (NSAIDs) are the main stay of treatment of pain.² It is known fact that the risk of gastrointestinal bleeding and other side effects are associated with acute and chronic use of non-steroidal anti-inflammatory drugs (NSAIDs).³ Keeping in view the gravity of adverse effects of NSAIDs it is necessary to search for new drugs with less adverse effects. In line with this many traditional medicinal plants have been used time to time for the development of new drugs with comparatively less ADRs as compare to NSAIDs.⁴ Recently, many natural medicines derived from medicinal plants, were considered as the effective and safer for the treatment of various diseases including inflammation and pain.⁵ One such medicinal plant namely *Myristica fragrans* is claimed for it is valuable role in reducing pain and inflammation. Hence, it was
found worthwhile to evaluate the extracts of *M. fragrans* for their role in reducing pain in experimental animals. *M. fragrans* (Nutmeg) belongs to the family myristicaceae, is one of the important spices used in indigenous system of medicine in India. Its usefulness is reported in inflammations, cephalgia, helmetsinisiasis, halitosis, dyspepsia, flatulence, nausea, vomiting, diarrhoea, dysentery, colic, asthma, catarrh, neuralgia, lumbago, palpitation, amenorrhoea, menorrhagia, dysmenorrhoea, ulcers, liver and splenic disorders, eye diseases, impotency, skin diseases, freckles, cracks in feet, insomnia, delirium tremens, hyperdysia, cardiac disorders, fever and general debility. Earlier studies showed that ethanolic extract of *M. fragrans* have analgesic activity. The present study is done to validate the earlier study and to screen additionally the effect of its methanol & benzene fraction.

**METHODS**

**Plant materials**

Seeds of *M. fragrans* were obtained from the local market. These were identified and authenticated by Chief Scientist, Raw Material Herbarium and Museum, National Institute of Science Communication and Information Resources (NISCAIR), New Delhi. A sample specimen of plant material was deposited in the NISCAIR bearing voucher number “NISCAIR/RHMD/2013/2345-125-2”.

**Preparation of extract**

*M. fragrans* seeds were shade-dried. Plant material (100g) was powdered by using electrical grinder (REMI auto-mix blender, Vasai, India). The powdered plant material was extracted with 300 ml of absolute alcohol by employing Soxhlet apparatus. The plant extract was filtered, evaporated and dry at 40°C on a water bath. Thus the extract material (semisolid mass) was weighed to calculate its yield in percentage. The final yield of ethanolic extract of *M. fragrans* was 17.94%.

**Preparation of fraction**

One kg powder was made after crushing the dried roots of *M. fragrans*. The air-dried powdered were exhausted with 95% ethanol and the solvent was isolated by steam distillation. Under reduced pressure the extract was concentrated to dark gummy mass. The residue so obtained was fractionated by refluxing in the consecutive order with benzene, ethyl acetate and methanol. The yield of methanol and benzene fraction of *M. fragrans* was 4.17% and 5.31% respectively.

**Animals used**

Wistar albino rats weighing between 150-200 g of either sex were used. The animals were procured from the Central Animal House and were housed in polypropylene cages bedded with husk. They were provided with standard pellet diet (Ashirwad Industries, Chandigarh) and water ad libitum. The animal room was well-ventilated and maintained under standard environmental conditions throughout the experiment (temperature 18-29°C, humidity 30-70%, 12 hour light/dark cycle). Rats were acclimatized to the laboratory condition for 1 week prior to experimental use. The study followed ARRIVE guidelines and was approved by the Institutional Animal Ethics Committee.

**Acute toxicity study**

No toxicity of ethanolic extract of *M. fragrans* seeds was done as ethanolic extract of plant material is claimed to be safe. However, the limit toxicity of methanol and benzene fraction of *M. fragrans* in addition to acute toxicity in accordance to Organization for Economic Cooperation and Development (OECD) Guidelines 425” was done on healthy adult female rats (100-150 g). While following the limit toxicity a dose of 2000 mg/kg was administered to a group of five animals for calculation of LD₅₀ of methanol fraction and they were observed for 14 days. However, no mortality related above limit toxicity dose was noted.

**Drugs**

Pentazocine lactate (Inj. Fortwin, Ranbaxy Lab. Ltd., India).

**Experimental protocol**

**Analgesic activity by Eddy’s hot plate method**

The present study was performed by using the Eddy’s hot plate method (Orchid Scientiﬁc, India). The hot plate is an electrically heated aluminium plate with a temperature ranging between 55°C to 56°C.

Rats of either sex (150-200 g) were used. The response noted was reaction time of licking/biting of both paws. They were similarly screened and those responding in <6 sec were chosen. The paws of rats are very sensitive to heat at temperatures compared to skin.

The selected animals were placed on hot plate to record the response. The reaction time was measured at the interval of 30, 60, 90,120,150,180, 210 and 240 minutes after the administration of control and test drugs. Propylene glycol 0.3 ml/100g p.o. served as control whereas Pentazocine 30 mg/kg i.p. was administered as standard drug. The cut-off time for response reaction was 30 seconds. The plate was wiped clean every time with saline if urination or defecation is found.

The test was done in all 8 groups as shown in Table 1.

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Table 1: Experimental design.

| Groups                  | Medication                     |
|-------------------------|--------------------------------|
| Group I (Normal control)| Propylenglycol 0.3 ml/100 g p.o.|
| Group II (Standard control)| Pentylenglycol 30 mg/kg i.p.  |
| Group III (EEMF200)     | Ethanolic extract of M. fragrans seed 200 mg/kg p.o. |
| Group IV (EEMF400)      | Ethanolic extract of M. fragrans seed 400 mg/kg p.o. |
| Group V (BFMF200)       | Methanol fraction of M. fragrans seed 200 mg/kg p.o. |
| Group VI (MFMF400)      | Ethanol fraction of M. fragrans seed 400 mg/kg p.o. |
| Group VII (BFMF200)     | Benzene fraction of M. fragrans seed 200 mg/kg p.o. |
| Group VIII (BFMF400)    | Benzene fraction of M. fragrans seed 400 mg/kg p.o. |

Analgesic activity by rat tail flick test

The method is based upon the reaction of rats to heat stimulus applied to their tail. It was performed by using the analgesiometer (Orchid Scientifics, India). Rats of either sex (150-200 gm) were placed in restraining holder so that the tail between the hole and tail tip or single point 3-5 cm from the tip of tail are directly kept over heated nichrome wire. The time taken by the rats to withdraw the tail was recorded. Heat intensity was adjusted such that the average withdrawn latency is 3-6 sec and a maximum cut-off time of 15 sec adopted to prevent undue tissue damage. Tail flick latency was tested at 30 min interval for 4 hours. The test was done in 8 groups as shown in Table 1.

Statistical analysis

The statistical analysis was done by using one-way ANOVA. The data is expressed as Mean ± SEM. P<0.05 was considered to be statistically significant.

RESULTS

Effect of ethanolic extract of M. fragrans seed on reaction time in rats on Eddy's hot plate

The rats in the control group responded within cut-off time of 6 seconds in all time periods. The Group II (Standard control) showed significant (p<0.001) increase in reaction time (seconds) as 3.48, 7.05, 8.13, 8.29, 6.96, 5.85 at interval (minutes) of 30, 60, 90, 120,150, 180 respectively. Ethanolic extracts in low (Group III/EEMF200) and high (Group IV/EEMF400 mg/kg) doses showed significant increase in reaction time (seconds) as 4.44 (p<0.05), 4.74 (p<0.001), 5.33 (p<0.001), 5.60 (p<0.001), 4.55 (p<0.01) and 4.82 (p<0.001), 5.17 (p<0.001), 5.99 (p<0.001), 6.35 (p<0.001), 5.14 (p<0.001) at interval (minutes) of 60, 90, 120, 150, 180. It was further noticed that mean response time was higher in the Group IV. The peak effect in both groups was seen at 150 min (Table 2).

Table 2: Effect of ethanolic extract of M. fragrans seed on reaction time in rats on Eddy’s hot plate.

| Groups                  | 0 min | 30 min | 60 min | 90 min | 120 min | 150 min | 180 min | 210 min | 240 min |
|-------------------------|-------|--------|--------|--------|---------|---------|---------|---------|---------|
| Normal control          | 3.89  | 3.76   | 3.89   | 3.78   | 3.95    | 3.90    | 3.89    | 3.88    | 3.85    |
|                          | ±0.1  | ±0.14  | ±0.16  | ±0.15  | ±0.21   | ±0.1   | ±0.1    | ±0.19   | ±0.1    |
| Positive control        | 3.53  | 5.48   | 7.05   | 8.13   | 8.29    | 6.96    | 5.85    | 4.24    | 3.54    |
|                          | ±0.2  | ±0.4***| ±0.2***| ±0.3***| ±0.3*** | ±0.4*** | ±0.46   | ±0.1    |         |
| EEMF 200                | 3.74  | 3.89   | 4.44   | 4.74   | 5.33    | 5.60    | 4.55    | 3.84    | 3.74    |
|                          | ±0.06 | ±0.05  | ±0.05* | ±0.05***| ±0.06***| ±0.06** | ±0.10   | ±0.07   |         |
| EEMF 400                | 3.84  | 4.03   | 4.82   | 5.17   | 5.99    | 6.35    | 5.14    | 4.05    | 3.84    |
|                          | ±0.07 | ±0.08  | ±0.09***| ±0.1***| ±0.1*** | ±0.05***| ±0.04   | ±0.07   |         |

Reaction Time: Mean±SEM (n=6) sec. *p<0.05, **p<0.01, ***p<0.001, as compared to normal control. EEMF- Ethanolic extract of M. fragrans

Table 3: Effect of ethanolic extract of M. fragrans seed on reaction time in rats on tail flick test.

| Groups                  | 0 min | 30 min | 60 min | 90 min | 120 min | 150 min | 180 min | 210 min | 240 min |
|-------------------------|-------|--------|--------|--------|---------|---------|---------|---------|---------|
| Normal control          | 4.12  | 4.11   | 4.12   | 4.13   | 4.13    | 4.12    | 4.13    | 4.12    | 4.10    |
|                          | ±0.01 | ±0.0   | ±0.0   | ±0.0   | ±0.1    | ±0.01   | ±0.01   | ±0.01   | ±0.0    |
| Positive control        | 4.11  | 5.92   | 6.67   | 8.15   | 9.02    | 7.79    | 6.73    | 5.01    | 4.13    |
|                          | ±0.02 | ±0.3***| ±0.1***| ±0.1***| ±0.34***| ±0.20***| ±0.18***| ±0.10***| ±0.02   |
| EEMF 200                | 4.06  | 4.89   | 6.02   | 6.79   | 6.39    | 5.26    | 4.52    | 4.09    | 4.04    |
|                          | ±0.02 | ±0.04  | ±0.2***| ±0.06***| ±0.08***| ±0.11***| ±0.13   | ±0.01   | ±0.02   |
| EEMF 400                | 4.05  | 5.25   | 6.57   | 7.37   | 7.01    | 5.94    | 5.34    | 4.19    | 4.03    |
|                          | ±0.03 | ±0.6***| ±0.01***| ±0.03***| ±0.12***| ±0.10***| ±0.18***| ±0.08   | ±0.04   |

Reaction time: Mean±SEM (n=6) sec. *p<0.05, **p<0.01, ***p<0.001, as compared to normal control. EEMF- Ethanolic extract of M. fragrans
**Effect of ethanolic extract of M. fragrans seed on reaction time in rats on tail flick test**

The rats in the control group responded within cut-off time of 6 seconds in all time periods. The standard group (Group II) showed significant (p<0.001) increase in reaction time (seconds) as 5.92, 6.67, 8.15, 9.02, 7.79, 6.73, 5.01 at interval (minutes) of 30, 60, 90, 120, 150, 180, 210 respectively. Ethanol extract in low (Group III) dose showed significant increase in reaction time (seconds) as 6.02 (p<0.001), 6.79 (p<0.001), 6.39 (p<0.001), 5.26 (p<0.001) at the interval (minutes) of 60, 90, 120 and 150 respectively. Ethanolic extract of high (Group IV/MFMF 200 mg/kg) and high (Group VI/MFMF 400 mg/kg) doses showed significant increase in reaction time (seconds) as 4.66 (p<0.001), 5.23 (p<0.001), 5.53 (p<0.001), 4.59 (p<0.05) and 4.79 (p<0.01), 5.92 (p<0.001), 6.23 (p<0.001), 5.00 (p<0.01) at interval (minutes) of 90,120, 150, 180. It was further noticed that mean response time was higher in the large dose group (Group VI). The peak effect in both groups was seen at 150 mins (Table 4).

**Effect of methanol fraction of M. fragrans seed on tail flick test in rats**

The rats in the control group responded within cut-off time of 6 seconds in all time periods. The Group II (Standard control) showed statistically significant increase in reaction time (seconds) as (p<0.001) 5.86, 6.77, 8.31, 9.13, 7.81, 6.72 and (p<0.01) 4.96 at the interval (minutes) of 30, 60, 90, 120,150, 180 and 210 respectively. Methanol fraction in low (Group V/MFMF 200 mg/kg) dose showed significant increase in reaction time (seconds) as 4.84 (p<0.05), 5.90 (p<0.001), 6.72 (p<0.001), 6.37 (p<0.001), 5.21 (p<0.01) at the interval (minutes) of 30, 60, 90, 120 and 150 respectively. Methanol fraction of high (Group VI/MFMF 400 mg/kg) dose showed significant increase in reaction time (seconds) as 5.25 (p<0.001), 6.49 (p<0.001), 7.25 (p<0.001), 7.07 (p<0.001), 5.85 (p<0.001), 5.23 (p<0.001) at interval (minutes) of 30, 60, 90, 120 and 180 respectively. It was further noticed that mean response time was higher in the large dose group (Group VI). The peak effect in both groups was seen at 90 mins (Table 5).

Table 4: Effect of methanol fraction of M. fragrans on reaction time in rats on Eddy’s hot plate.

| Groups | 0 min | 30 min | 60 min | 90 min | 120 min | 150 min | 180 min | 210 min | 240 min |
|--------|------|-------|-------|-------|--------|--------|--------|--------|--------|
| Normal control | 3.89 | 3.76 | 3.89 | 3.78 | 3.95 | 3.90 | 3.89 | 3.88 | 3.85 |
| Positive control | 4.53 | 4.58 | 7.05 | 8.13 | 8.29 | 6.96 | 5.85 | 4.24 | 3.54 |
| MFMF 200 | 4.36 | 3.84 | 4.31 | 4.66 | 5.23 | 5.53 | 4.59 | 3.79 | 3.72 |
| MFMF 400 | 4.38 | 3.98 | 4.79 | 5.16 | 5.92 | 6.23 | 5.00 | 4.01 | 3.77 |

Reaction Time: Mean±SEM (n= 6) sec. *P<0.05, **P<0.01, ***P<0.001, as compared to normal control. MFMF- Methanol fraction of M. fragrans

Table 5: Effect of methanol fraction of M. fragrans seed on reaction time in rats using tail flick test in rats.

| Groups | 0 min | 30 min | 60 min | 90 min | 120 min | 150 min | 180 min | 210 min | 240 min |
|--------|------|-------|-------|-------|--------|--------|--------|--------|--------|
| Normal control | 4.12 | 4.11 | 4.12 | 4.13 | 4.13 | 4.12 | 4.13 | 4.12 | 4.10 |
| Positive control | 4.11 | 5.92 | 6.67 | 8.15 | 9.02 | 7.79 | 6.73 | 5.01 | 4.13 |
| MFMF 200 | 4.02 | 4.84 | 5.90 | 6.72 | 6.37 | 5.21 | 4.44 | 4.07 | 4.04 |
| MFMF 400 | 4.05 | 5.25 | 6.49 | 7.25 | 7.07 | 5.85 | 5.23 | 4.16 | 4.02 |

Reaction Time: Mean±SEM (n= 6) sec. *P<0.05, **P<0.01, ***P<0.001, as compared to normal control. MFMF- Methanol fraction of M. fragrans
TABLE 6: Effect of Benzene fraction of *M. fragrans* seed on reaction time in rats using Eddy’s hot plate.

| Groups | 0 min | 30 min | 60 min | 90 min | 120 min | 150 min | 180 min | 210 min | 240 min |
|--------|-------|--------|--------|--------|---------|---------|---------|---------|---------|
| Normal control | 3.93 | 3.93 | 3.83 | 3.91 | 3.85 | 3.83 | 3.92 | 3.83 | 3.82 |
| Positive control | 3.84 | 5.95 | 7.22 | 8.39 | 8.77 | 7.46 | 6.41 | 4.67 | 3.86 |
| BFMF 200 | ±0.2 | ±0.2 | ±0.2 | ±0.2 | ±0.2 | ±0.2 | ±0.2 | ±0.2 | ±0.2 |
| BFMF 400 | ±0.0 | ±0.3*** | ±0.4*** | ±0.3*** | ±0.1*** | ±0.2*** | ±0.2*** | ±0.5 | ±0.07 |

Reaction time: Mean±SEM (n=6) sec. *p<0.05, **p<0.01, ***p<0.001, as compared to normal control. BFMF- Benzene fraction of *M. fragrans*.

**Effect of benzene fraction of *M. fragrans* seed on Eddy’s hot plate test**

The rats in the control group responded at all intervals by 6 seconds. The rats in the control group responded within cut-off time of 6 seconds in all time periods. The standard group (Group II) of pentazocine showed significant (p<0.001) increase in reaction time (seconds) as 5.95, 7.22, 8.39, 7.46 and 6.41 at the interval (minutes) of 30, 60, 90, 120, 150 and 180, respectively. Low dose (Group VII/BFMF200 mg/kg) and high dose (Group VIII/BFMF400 mg/kg) of Benzene fraction of *M. fragrans* seed revealed no statistically significant effects (Table 6).

**Effect of benzene fraction of *M. fragrans* seed on tail flick test in rats**

The rats in the control group responded within cut-off time of 6 seconds in all time periods. The Standard group (Group II) showed significant increase in reaction time (Seconds) as (p<0.001) 5.86, 6.77, 8.31, 9.13, 7.81, 6.72 and (p<0.01) 4.96 at the interval (minutes) of 30, 60, 90, 120, 150, 180 and 210 respectively. Low dose (Group VII/BFMF200 mg/kg) and high dose (Group VIII/BFMF400 mg/kg) of benzene fraction of *M. fragrans* seed revealed no statistically significant effects (Table 7).

**DISCUSSION**

The present study reveals the dose dependent significant analgesic activity of the extracts of *M. fragrans* in both the test.

In Eddy’s hot plate and tail flick test, The Standard group of pentazocine showed significant (p<0.001) increase in reaction time. Administration of ethanolic extract of *M. fragrans* in low (200 mg/kg) and high (400 mg/kg) doses showed statistically significant increase in reaction time. Similarly the low dose (200 mg/kg) and high dose (400 mg/kg) of methanol fraction showed significant increase in reaction time. It was also noticed that reaction time was higher in the large dose group in both ethanolic and methanol fraction. Further in ethanolic extract and methanol fraction, the peak effect in both groups was observed at 150 mins. It is interesting to note that low dose (200 mg/kg) and high dose (400 mg/kg) of Benzene fraction of *M. fragrans* seed revealed no statistically significant effects (p>0.05).

Phytochemical examination of *M. fragrans* showed occurrence of actual biological composite like alkaloids, steroids, flavonoids, tannins, phenolics, glycosides in essential oil.9 *M. fragrans* volatile oil is comprised of a mixture of terpenes and alkylbenzene derivatives. Myristicin, safrole and elimicin constitute about 80% of the alkylbenzene derivatives.10 Bioactive compounds including camphene, elemicin, eugenol, isoelemicin, isoeugenol, methoxyeugenol and elimicin were identified as the main constituents of *M. fragrans* seed essential oil.11 Alkaloids are commonly found to have analgesic activities.12 The result obtained in this work demonstrated...
a high analgesic activity at low and high dose of ethanolic extract and methanol fraction of *M. fragrans*.

A study reported that the chemical substance extracted from Myristicaceae plants has reduced the arachidonic acid which is the precursor for prostaglandin synthesis by inhibiting the phospholipase A2. Another study also reported that the antinociceptive activity of plant extracts may be due to inhibition of interleukin-1β and interleukin-8 release by resident peritoneal cells or to suppression of prostaglandins and bradykinin. However, the literature supporting this direct evidence related to antinociceptive action involving either of the mechanism for inhibition of the substances is lacking.

On the contrary a study with regard to the use of scientific methodology employed for evaluation of antinociceptive effect of extracts of *M. fragrans* is in support of reporting that both the responses of hot plate test (paw licking and jumping) integrate at supraspinal structures with the C and Aδ type I and II sensitive fibers participating in this model.

The result of study reveals significant analgesic effect by both hot plate and tail flick tests suggesting enrichment in components (primarily non-lipid and/or aromatic compounds) that might activate a spinally-mediated analgesic pathway. However lack of various constituents in benzene fraction showed no statistically significant analgesic effects.

**CONCLUSION**

We conclude that, the test drug *Myristica fragrans* seed in its ethanolic extract and methanol fraction showed statistically significant increase in reaction time (seconds) on Eddy’s hot plate and tail flick test. The study reveals the dose dependent significant analgesic activity of the extracts of *Myristica fragrans* seed in both the test. Thus the present drug used in traditional medicine might give a solution as an alternative remedy in pain management.

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**Ethical approval:** The study followed ARRIVE guidelines and was approved by the Institutional Animal Ethics Committee (Registration No. 401/RO/C/2001/CPCSEA)

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