Multiple-dose versus single-dose gonadotropin-releasing hormone agonist after first in vitro fertilization failure associated with luteal phase deficiency: A randomized controlled trial

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Abstract

Objective: To evaluate the efficacy and safety of multiple- versus single-dose gonadotropin-releasing hormone agonist (GnRH-a) addition to luteal phase support (LPS), in patients with a first in vitro fertilization (IVF) failure associated with luteal phase deficiency (LPD).

Methods: Eighty patients with a first IVF failure associated with LPD were randomly assigned into single-dose and multiple-dose GnRH-a groups. In the second IVF attempt, patients in the single-dose group were given standard LPS plus a single dose of GnRH-a 6 days after oocyte retrieval. Patients in the multiple-dose group received standard LPS plus 14 daily injections of GnRH-a. Children conceived were followed up for 2 years.

Results: Pregnancy (67.5% vs. 42.5%), clinical pregnancy (50.0% vs. 22.5%), and live birth rates (42.5% vs. 20.0%) were significantly higher in the multiple-dose versus single-dose GnRH-a group. Patients in the multiple-dose GnRH-a group had significantly higher progesterone levels 14 days after oocyte recovery (35.9 vs. 21.4 ng/mL). No significant difference existed in the status at birth or developmental and behavior assessments of 2-year-old children conceived in both groups.

Conclusions: Daily addition of GnRH-a to standard LPS can achieve better pregnancy outcomes with a sustained safety profile in patients with a first IVF failure associated with LPD.

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Gonadotropin-releasing hormone agonist, luteal phase deficiency, in vitro fertilization failure, long-term safety, luteal phase support, pregnancy outcomes

Introduction
Luteal phase deficiency (LPD) is mainly caused by impaired secretory function of the corpus luteum, resulting in low estradiol and progesterone levels and shortening of the luteal phase. In the context of in vitro fertilization (IVF) attempts, LPD is common in follicular stimulation using any kind of ovarian stimulation protocol. Luteal phase support (LPS) is considered essential to correct LPD in infertility treatment. The first LPS modalities were the administration of human chorionic gonadotropin (hCG) and progesterone. However, LPS with hCG increases the risk of ovarian hyperstimulation syndrome compared with progesterone. Progesterone for LPS can be administered vaginally, orally, or intramuscularly, but the optimal route of administration has not been established. Some other modalities are under investigation; for example, estrogen, steroids, ascorbic acid, and acupuncture.

Recent studies have proposed the use of a gonadotropin-releasing hormone agonist (GnRH-a) as LPS. The first report of GnRH-a in LPS by Tesarik et al. retrospectively demonstrated that GnRH-a administration at the time of implantation could significantly improve implantation and birth rates and enhance embryo developmental potential. Several studies have since exhibited a positive effect of GnRH-a as LPS.

LPD is well accepted as one of the first factors contributing to infertility. Moreover, if LPD cannot be diagnosed correctly, it contributes to IVF failure. The diagnostic criteria of LPD remain a matter of debate. Some studies define LPD as a single progesterone level below 10 ng/mL (31.8 nmol/L) in the mid-luteal phase. Recent studies have revealed that a cutoff serum progesterone level of 15 ng/mL (47.7 nmol/L) on the day of pregnancy test exhibited 100% sensitivity for detecting LPD. Zafardoust et al. reported that single-dose GnRH-a in the luteal phase could improve implantation and clinical pregnancy rates in patients with previous IVF-embryo transfer (ET) failure. Mendoza-Tesarik et al. found that GnRH-a treatment for 2 weeks could improve pregnancy and birth rates in patients with a first IVF failure associated with LPD. However, there have been relatively few studies on the effect and long-term safety of GnRH-a as LPS in patients with previous IVF failure. In addition, optimal use of GnRH-a remains undetermined. The present study aimed to explore the efficacy and safety of single-dose and multiple-dose GnRH-a as LPS in patients with first IVF failure associated with LPD.

Materials and methods
Patients
This was a prospective randomized controlled study in a 1:1 allocation ratio conducted in the assisted reproduction technology unit of a tertiary university.
hospital in Beijing. Patients who underwent a second IVF-ET treatment between January 2013 and January 2016 were considered for participation in our study. Patients were divided into two groups using the random block allocation method. The inclusion criteria were as follows: (1) between 22 and 40 years old; (2) failure to achieve clinical pregnancy in their first IVF attempt with LPD after ET; (3) treatment cycles with controlled ovarian hyperstimulation (COH) and fresh ETs. LPD during the first attempt was defined as a serum progesterone level <15 ng/mL on day 14 after oocyte retrieval in patients receiving standard LPS treatment. Women with an abnormal uterine cavity or with frozen ET cycles were excluded.

The study was approved by the Ethics Committee of Beijing Chaoyang Hospital. All procedures that involved human participants were in accordance with the ethical standards of the Institutional and National Research Committee, with the 1964 Declaration of Helsinki and its later amendments or comparable ethical standards. The Consolidated Standards of Reporting Trials (CONSORT) recommendations were followed in this study. Informed consent was obtained from all participants enrolled for the study.

**COH protocol**

Ovarian stimulation was carried out using recombinant human follicle stimulating hormone (rFSH; Gonal-F; Merck Serono, Geneva, Switzerland) and human menopausal gonadotropin (hMG, Livzon, Zhuhai, China). Serial transvaginal ultrasound was performed to monitor follicular growth. The doses of rFSH and hMG were adjusted according to follicular development and serum estradiol and luteinizing hormone (LH) levels. When at least three follicles reached a mean diameter of 18 mm, ovulation was induced with a single dose of 10,000 IU of recombinant human chorionic gonadotropin (hCG). After 36 hours, transvaginal ultrasound-guided oocyte retrieval was performed. Fertilization was achieved with intracytoplasmic sperm injection (ICSI) in all couples, and ET was performed on day 3 following ICSI. One or two embryos were implanted depending on maternal age. All women were treated with vaginal 8% progesterone gel, 90 mg/d (Crinone, Merck Serono, Rockland, MA, USA) plus dydrogesterone tablets (Duphaston, Abbott Laboratories, Chicago, IL, USA), 20 mg/d, starting on the day of oocyte retrieval and continuing until the pregnancy test performed 14 days after ET.

Data on participants’ characteristics, COH, and embryology were collected. These included maternal age; type, duration, and cause of infertility; ovarian reserve assessment [cycle day 3 anti-Müllerian hormone (AMH) and FSH levels]; duration and total dose of gonadotropin (Gn) treatment; endometrial thickness on the day of hCG administration; number of retrieved oocytes; and number of embryos transferred.

All participants were randomized into one of two groups by a computer-generated program. Women allocated to the single-dose GnRH-a group received a single dose of 0.1 mg decapetyl 6 days after oocyte retrieval, whereas women allocated to the multiple-dose GnRH-a group received an additional daily injection of 0.1 mg decapetyl for 14 days starting from the day of oocyte retrieval. Investigators and participants were not blinded to treatment allocation.

**Primary and secondary effect outcome measures**

Primary effect outcomes of the study were pregnancy outcomes, including pregnancy rate, clinical pregnancy rate, and live birth
rate. Pregnancy was assessed by measuring serum β-hCG levels 14 days after embryo transfer, and clinical pregnancy was confirmed by the presence of an intrauterine gestational sac on ultrasonography 5 weeks after embryo transfer. Clinical pregnancy rate was calculated as the number of clinical pregnancies divided by the number of ET procedures. Live birth rates were defined as the percentage of assisted reproductive technology (ART) cycles started that resulted in a live birth. The outcome “birth defect” was defined as a structural defect of the body that affects quality and viability of life and that requires medical intervention. The secondary effect outcomes were serum progesterone concentrations, measured on day 14 after ET.

Safety outcome measures
Safety outcome was based on measures of motor, cognitive, language, and behavioral development of children conceived by the study cohort. When IVF-ET children were 24 months old, developmental-behavior assessments were performed. The developmental-behavior assessments consisted of four domains: motor, cognitive, language, and behavioral development. The Bayley Scales of Infant Development (3rd ed., Bayley-III), a multidisciplinary battery, was used to assess motor, cognitive, and language development. The motor scale of Bayley-III assesses gross and fine motor skills; the cognitive scale measures non-verbal activities including sensorimotor development, object exploration, and concept formation; and the language scale estimates receptive and communicative skills. The Bayley-III scores were converted to a standardized mean value of 50 and a standard deviation (SD) of 10, with higher scores reflecting better performance. Behavior assessment was assessed using the Child Behavior Checklist (CBCL). The CBCL consists of 100 items describing sleep problems, withdrawal, somatic problems, depression, and aggressive and destructive behavior. Raw scores were normalized into T-scores with a mean value of 50 and an SD of 10.

Statistical analysis
A pilot study at our institution (8 patients per group) demonstrated a 25% difference in pregnancy rates between two groups. Power analysis indicated that a sample size of at least 37 patients per group would provide a power of 80% at an α level of 0.05 (two-tailed test). Thus, we aimed to enroll 40 patients in each group. Continuous data were expressed as mean ± SD or median (interquartile range) and were compared with the Student’s t test, Mann-Whitney U test, or Wilcoxon rank sum test. Categorical data were expressed as number (percentage) and compared with Fisher’s exact test or the Chi-square test. We performed subgroup analyses in both groups by comparing singleton and multiple pregnancies. Statistical significance was considered at a two-tailed P < 0.05. Statistical analyses were performed using SPSS version 22.0 (IBM Corp., Armonk, NY, USA).

Results
Participant characteristics at baseline
Eighty patients were enrolled in this study, in two groups: single-dose GnRH-a (n = 40) and multiple-dose GnRH-a (n = 40). The study flowchart is depicted in Figure 1. The baseline characteristics of the patients in the two groups, including maternal age, BMI, menstrual cycle, infertility cause, and duration of infertility, were similar, as shown in Table 1.
Fresh cycle characteristics

The fresh cycle characteristics of the patients are depicted in Table 2. We found no differences in AMH level, FSH level, Gn dose, Gn duration, average endometrial thickness, average number of retrieved oocytes, or average number of transferred embryos between the two groups.

IVF-ET pregnancy outcomes

Compared with the single-dose GnRH-a group, patients in the multiple-dose GnRH-a group demonstrated higher rates of pregnancy ($P = 0.025$), clinical pregnancy ($P = 0.011$), and live birth ($P = 0.030$), as shown in Table 3. There were no significant differences between patients in the single-dose GnRH-a and multiple-dose GnRH-a groups regarding delivery outcomes: preterm birth [1/8 (12.5%) vs. 1/17 (5.9%)] and low birth weight [1/8 (12.5%) vs. 1/17 (5.9%)]. All pregnancies resulted in the birth of healthy babies.

Serum progesterone levels

There was no difference in serum progesterone level on the day of ET between the two groups. However, on day 14 after ET, serum progesterone levels of patients in the multiple-dose GnRH-a group were significantly higher than those of the single-dose GnRH-a group ($P < 0.001$; Table 4). In the subgroup analysis (Table 5), both pregnant and nonpregnant patients of the multiple-dose GnRH-a group had higher serum progesterone levels ($P = 0.002$ and $P = 0.003$, respectively) than those of the single-dose GnRH-a group.
Table 1. Baseline characteristics of the study cohort.

| Variable                  | Single-dose GnRH-a group (n = 40) | Multiple-dose GnRH-a group (n = 40) | P-value |
|---------------------------|-----------------------------------|-------------------------------------|---------|
| Maternal age, years       | 28.6 ± 4.6                        | 27.2 ± 4.9                          | 0.192   |
| BMI, kg/m²                | 22.6 ± 6.3                        | 22.9 ± 6.6                          | 0.836   |
| Duration of infertility, years | 3.8 ± 0.9                           | 3.5 ± 0.7                          | 0.100   |
| Menstrual cycle, days     | 28.3 ± 3.8                        | 29.6 ± 4.2                          | 0.151   |
| Type of infertility, n (%) |                                   |                                     | 0.356   |
| Primary infertility       | 17 (42.5)                         | 23 (57.5)                           |         |
| Secondary infertility     | 13 (32.5)                         | 27 (67.5)                           |         |
| Infertility cause         |                                   |                                     | 0.757   |
| Tubal abnormality         | 12 (30.0)                         | 16 (42.1)                           |         |
| Male factor               | 11 (27.5)                         | 8 (21.1)                            |         |
| Cervical factor           | 7 (17.5)                          | 6 (15.8)                            |         |
| Ovulation failure         | 5 (12.5)                          | 4 (10.5)                            |         |
| Endometriosis             | 2 (5.0)                           | 4 (10.5)                            |         |
| Unexplained               | 3 (7.5)                           | 2 (5.0)                             |         |

GnRH-a, gonadotropin-releasing hormone agonist; BMI, body mass index.

Table 2. Fresh cycle characteristics of the study cohort.

| Variable                  | Single-dose GnRH-a group (n = 40) | Multiple-dose GnRH-a group (n = 40) | P-value |
|---------------------------|-----------------------------------|-------------------------------------|---------|
| AMH, ng/mL                | 4.3 ± 1.3                         | 4.1 ± 1.2                           | 0.477   |
| FSH, IU/L                 | 6.5 ± 2.1                         | 6.2 ± 1.7                           | 0.485   |
| Gn duration, days         | 10.5 ± 2.2                        | 10.2 ± 1.9                          | 0.516   |
| Gn dose, U                | 1998.7 ± 516.2                    | 1945.1 ± 505.9                      | 0.640   |
| Endometrial thickness, mm | 11.9 ± 1.9                        | 11.3 ± 1.7                          | 0.141   |
| Retrieved oocytes         | 10.2 ± 4.2                        | 9.5 ± 3.9                           | 0.442   |
| Transferred embryos       | 1.8 ± 0.2                         | 1.9 ± 0.3                           | 0.083   |

GnRH-a, gonadotropin-releasing hormone agonist; AMH, anti-Müllerian hormone; FSH, follicle-stimulating hormone; Gn, gonadotropin.

Table 3. Pregnancy outcomes following in vitro fertilization-embryo transfer.

| Variable                  | Single-dose GnRH-a group (%)     | Multiple-dose GnRH-a group (%)    | P-value |
|---------------------------|----------------------------------|----------------------------------|---------|
| Pregnancy rate, n (%)     | 17 (42.5)                        | 27 (67.5)                        | 0.025   |
| Clinical pregnancy rate, n (%) | 9 (22.5)                       | 20 (50.0)                        | 0.011   |
| Live birth rate, n (%)    | 8 (20)                           | 17 (42.5)                        | 0.030   |
| Singleton pregnancy, n (%) | 7 (17.5)                        | 14 (35.0)                        |         |
| Twin pregnancy, n (%)     | 1 (2.5)                          | 3 (7.5)                          |         |

GnRH-a, gonadotropin-releasing hormone agonist.
Table 4. Luteal phase characteristics of the study cohort.

| Variable                          | Single-dose GnRH-a group (n = 40) | Multiple-dose GnRH-a group (n = 40) | P-value |
|----------------------------------|-----------------------------------|-----------------------------------|---------|
| Serum progesterone (ng/mL)       |                                   |                                   |         |
| Day of ET                        | 12.8 ± 3.8                        | 12.6 ± 3.7                        | 0.812   |
| Day 14 after ET                  | 21.4 ± 10.9                       | 35.9 ± 6.2                        | <0.001  |

GnRH-a, gonadotropin-releasing hormone agonist; ET, embryo transfer.

Table 5. Subgroup analysis of luteal phase characteristics.

| Variable                          | Single-dose GnRH-a group | Multiple-dose GnRH-a group | P-value |
|-----------------------------------|--------------------------|----------------------------|---------|
| Pregnant patients, n              | 17                       | 27                         |         |
| Serum progesterone on day 14 after ET (ng/mL) | 31.5 ± 10.6           | 42.9 ± 11.5               | 0.002   |
| Nonpregnant patients, n           | 23                       | 13                         |         |
| Serum progesterone on day 14 after ET (ng/mL) | 13.9 ± 6.3             | 21.3 ± 7.3                | 0.003   |

GnRH-a, gonadotropin-releasing hormone agonist; ET, embryo transfer.

Table 6. Long-term follow-up of children conceived by the study cohort.

| Variable                          | Single-dose GnRH-a group | Multiple-dose GnRH-a group | P-value |
|-----------------------------------|--------------------------|----------------------------|---------|
| Number of live births, n          | 9                        | 20                         |         |
| Bayley scores                     |                          |                            |         |
| Motor skills                      | 105.6 ± 16.3             | 101.5 ± 17.9               | 0.563   |
| Cognition and language skills     | 100.3 ± 17.5             | 102.7 ± 20.5               | 0.763   |
| CBCL scores                       | 45.8 ± 9.8               | 47.5 ± 9.7                 | 0.667   |

GnRH-a, gonadotropin releasing hormone agonist; CBCL, Child Behavior Checklist.

**Long-term follow-up of children conceived by the study cohort**

Table 6 summarizes the results of Bayley-III motor, cognitive, and language assessments and CBCL behavior assessment performed when the children of IVF-ET patients were 24 months old. We observed no significant differences in motor, cognitive, language, or behavioral outcomes between the two groups.

**Discussion**

LPD has been identified in all ARTs. To overcome this issue, additional LPS has been routinely used during ART-stimulated cycles to improve pregnancy outcomes.\(^\text{16}\) GnRH-a is considered a novel LPS. In 1993, Balasch et al.\(^\text{17}\) reported the first study of inadvertent administration of GnRH-a in the mid-luteal phase, which did not compromise pregnancy outcomes but
rather enhanced implantation rates. A series of studies have since demonstrated that addition of GnRH-a during the luteal phase can significantly increase rates of clinical pregnancy, ongoing pregnancy, and live births.\(^8,18–21\)

Although the exact mechanism of GnRH-a in the mid-luteal phase remains unclear, the major beneficial effect of GnRH-a administration in this phase is postulated to work at three levels: the corpus luteum, the endometrium, and the embryo. GnRH-a can support corpus luteum maintenance by activating the secretion of LH by pituitary gonadotropic cells.\(^{22}\) GnRH-a can directly bind GnRH receptors on the endometrium, simulate the production of angiogenic growth factors, and decrease natural killer cytotoxicity, making the endometrium more favorable for embryo implantation.\(^{23}\) Additionally, some researchers believe that GnRH-a can have a direct beneficial effect on early embryo development, as indicated by increased hCG, progesterone, and estradiol levels.\(^{6,23}\)

Many studies have confirmed that a single administration of GnRH-a can enhance the clinical outcomes of IVF-ET treatment.\(^{6,8,9,14}\) Administration of multiple boluses of GnRH-a in LPS protocols has become more common. Pirard et al.\(^{24}\) published the results of a prospective, randomized controlled clinical trial, in which daily administration of GnRH-a, as the only LPS for 10 to 16 days, achieved rates of pregnancy, implantation, and clinical pregnancy comparable to the standard LPS with intravaginal progesterone. Bar-Hava et al.\(^{25}\) showed that daily continuous administration of GnRH-a for 2 weeks, as the only LPS, supported and enabled fresh embryo implantation with satisfactory clinical pregnancy rates. Recently, Mendoza-Tesarik et al.\(^{15}\) found that daily supplementation of GnRH-a during luteal support for 2 weeks in patients with a first IVF failure resulted in higher pregnancy and clinical pregnancy rates in their second IVF attempts. However, no study has compared single-dose and multiple-dose GnRH-a as LPS for IVF-ET. To the best of our knowledge, ours is the first study to make this comparison. Our data showed that daily repeated GnRH-a administration had pronounced beneficial effects on pregnancy, clinical pregnancy, and live birth outcomes. This improvement over the single-dose regimen probably reflects the short-action duration of GnRH-a, which has a half-life of 3 hours in plasma after a subcutaneous injection of 1.0 mg.\(^{26}\)

Although GnRH-a, as LPS, appears to exert beneficial effects on the achievement of clinical pregnancy, concerns remain regarding the safety of GnRH-a. In 1998, Cahill\(^{27}\) reported 346 unexpected spontaneous pregnancies in patients exposed to GnRH-a for 10 to 20 days in early pregnancy. Among these pregnancies, congenital abnormality and pregnancy loss rates were 2.5% and 15%, respectively, approximately the same as the rates in the population at large. However, Sahin et al.\(^{28}\) reported that GnRH-a triggered IVF cycles and had higher ectopic pregnancy rates relative to hCG-triggered cycles, which was probably caused by the decreased receptivity of the endometrium due to insufficient luteal support. Zhou et al.\(^{29}\) showed that addition of a single dose of GnRH-a to progesterone did not increase the risk of complications during pregnancy, at delivery, or postpartum. A recent study published by Bar-Hava et al.\(^{25}\) revealed that daily administration of GnRH-a for 2 weeks did not have any long-term adverse effects. Our study used the Bayley-III and CBCL scores to assess the developmental behaviors at 2 years old of children conceived by the study cohort. The Bayley-III and CBCL scores of the study cohort were comparable to those of ART-conceived children reported by Balayla.
et al.,30 and Zhan et al.,31 which indicates the safety of GnRH-a as LPS.

In this prospective randomized controlled study, we investigated and compared the efficacy and safety of single and multiple doses of GnRH-a with progesterone as LPS on pregnancy, delivery, and postpartum outcomes. We selected patients who had a first IVF failure associated with LPD. Our results indicated that continuous supplementation of GnRH-a could support the luteal phase, as indicated by higher serum progesterone levels in the mid-luteal phase in the multiple-dose GnRH-a group, which resulted in higher rates of pregnancy, clinical pregnancy, and live birth compared with the single-dose GnRH-a group. Additionally, during the 2-year follow-up, children conceived by the study cohort had developmental behaviors similar to those reported in ART-conceived children.

Our study had several limitations. First, the study cohort consisted of a relatively small sample from a single institution. Additional studies with multi-institutional cohorts are warranted to validate the study findings. Second, although we speculate that GnRH-a alone is sufficient for LPS, patients in both GnRH-a groups of the present study were also administered transvaginal progesterone, primarily for ethical reasons. We intend to solve this issue in a future study.

In summary, in the present study, we showed that daily addition of GnRH-a to luteal support achieved better pregnancy outcomes with a sustained safety profile in patients who had a first IVF failure associated with LPD.

Declaration of conflicting interest

The authors declare that there is no conflict of interest.

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References

1. Fritz MA. The modern infertility evaluation. Clin Obstet Gynecol 2012; 55: 692–705. DOI: 10.1097/GRF.0b013e31825cab55.
2. Daya S and Gunby J. Luteal phase support in assisted reproduction cycles. Cochrane Database Syst Rev 2004; 3: CD004830. DOI: 10.1002/14651858.CD004830.
3. Pirard C, Donnez J and Loumaye E. GnRH agonist as luteal phase support in assisted reproduction technique cycles: results of a pilot study. Hum Reprod 2006; 21: 1894–1900. DOI: 10.1093/humrep/del072.
4. Ciampaglia W and Cognigni GE. Clinical use of progesterone in infertility and assisted reproduction. Acta Obstet Gynecol Scand 2015; 94: 17–27. DOI: 10.1111/aogs.12770.
5. Fatemi HM. Simplifying luteal phase support in stimulated assisted reproduction cycles. Fertil Steril 2018; 110: 1035–1036. DOI: 10.1016/j.fertnstert.2018.08.019.
6. Tesarik J, Hazout A and Mendoza C. Enhancement of embryo developmental potential by a single administration of GnRH agonist at the time of implantation. Hum Reprod 2004; 19: 1176–1180. DOI: 10.1093/humrep/deh235.
7. Van Der Linden M, Buckingham K, Farquhar C, et al. Luteal phase support for assisted reproduction cycles. Cochrane Database Syst Rev 2011; 10: CD009154. DOI: 10.1002/14651858.CD009154.pub2.
8. Oliveira JB, Baruffi R, Petersen CG, et al. Administration of single-dose GnRH agonist in the luteal phase in ICSI cycles: a meta-analysis. Reprod Biol Endocrinol 2010; 8: 107. DOI: 10.1186/1477-7827-8-107.
9. Boyle PC, De Grooth T, Andralojc KM, et al. Healthy singleton pregnancies from restorative reproductive medicine (RRM) after
failed IVF. Front Med (Lausanne) 2018; 5: 210. DOI: 10.3389/fmed.2018.00210.

10. Tesarik J, Mendoza N and Mendoza-Tesarik R. The neglected luteal phase after natural conception: rescue by early progesterone supplementation. Curr Opin Gynecol Obstet 2019; 2: 216–220. DOI: 10.18314/cogo.v2i2.1722.

11. Jordan J, Craig K, Clifton DK, et al. Luteal phase defect: the sensitivity and specificity of diagnostic methods in common clinical use. Fertil Steril 1994; 62: 54–62. DOI: 10.1016/s0015-0282(16)56815-0.

12. Van Zonneveld P, Te Velde ER and Koppeschaar HP. Low luteal phase serum progesterone levels in regularly cycling women are predictive of subtle ovulation disorders. Gynecol Endocrinol 1994; 8: 169–174. DOI: 10.3109/09513599409072451.

13. Aslih N, Ellenbogen A, Shavit T, et al. Can we alter pregnancy outcome by adjusting progesterone treatment at mid-luteal phase: a randomized controlled trial. Gynecol Endocrinol 2017; 33: 602–606. DOI: 10.1080/09513590.2017.1298742.

14. Zafardoust S, Jeddi-Tehrani M, Akhondi MM, et al. Effect of administration of single dose GnRH agonist in luteal phase on outcome of ICSI-ET cycles in women with previous history of IVF/ICSI failure: a randomized controlled trial. J Reprod Infertil 2015; 16: 96–101.

15. Mendoza-Tesarik R, Mendoza N, López CC, et al. GnRH agonist treatment of luteal phase deficiency in HCG-triggered IVF cycles: a matched case-control study. Reprod Biomed Online 2019; 39: 225–230. DOI: 10.1016/j.rbmo.2019.03.215.

16. De Ziegler D, Ayoubi JM, Frydman R, et al. Luteal phase support in assisted reproductive technologies: from here to there. Fertil Steril 2018; 109: 57–58. DOI: 10.1016/j.fertnstert.2017.10.031.

17. Balasch J, Martinez F, Jove I, et al. Inadvertent gonadotrophin-releasing hormone agonist (GnRHa) administration in the luteal phase may improve fecundity in in-vitro fertilization patients. Hum Reprod 1993; 8: 1148–1151.

18. Papanikolaou EG, Platteau P, Albano C, et al. Achievement of pregnancy three times in the same patient during luteal GnRH agonist administration. Reprod Biomed Online 2005; 10: 347–349.

19. Platteau P, Gabbe M, Talbot M, et al. Two consecutive pregnancies during inadvertent gonadotropin-releasing hormone agonist desensitization. Fertil Steril 2000; 73: 1244–1246.

20. Tan HH, Yeong CT and Loh KE. Perinatal outcome of pregnancies after inadvertent exposure to gonadotrophin-releasing hormone analogue. Aust N Z J Obstet Gynaecol 2006; 46: 336–340. DOI: 10.1111/j.1479-828X.2006.00602.x.

21. Van Der Linden M, Buckingham K, Farquhar C, et al. Luteal phase support for assisted reproduction cycles. Cochrane Database Syst Rev 2015; 7: CD009154. DOI: 10.1002/14651858.CD009154.pub3.

22. Pirard C, Donnez J and Loumaye E. GnRH agonist as novel luteal support: results of a randomized, parallel group, feasibility study using intranasal administration of buserelin. Hum Reprod 2005; 20: 1798–1804. DOI: 10.1093/humrep/deh830.

23. Tesarik J, Hazout A, Mendoza-Tesarik R, et al. Beneficial effect of luteal-phase GnRH agonist administration on embryo implantation after ICSI in both GnRH agonist- and antagonist-treated ovarian stimulation cycles. Hum Reprod 2006; 21: 2572–2579. DOI: 10.1093/humrep/dei173.

24. Pirard C, Loumaye E, Laurent P, et al. Contribution to more patient-friendly ART treatment: efficacy of continuous low-dose GnRH agonist as the only luteal support-results of a prospective, randomized, comparative study. Int J Endocrinol 2015; 2015: 727569. DOI: 10.1155/2015/727569.

25. Bar-Hava I, Mizrahi Y, Karfunkel-Doron D, et al. Intranasal gonadotropin-releasing hormone agonist (GnRHa) for luteal-phase support following GnRHa triggering, a novel approach to avoid ovarian hyperstimulation syndrome in high responders. Fertil Steril 2016; 106: 330–333. DOI: 10.1016/j.fertnstert.2016.04.004.

26. Lorusso F, Depalo R and Selvaggi L. Relationship between gonadotropin releasing hormone agonist dosage and in vitro
fertilization outcome. *Gynecol Endocrinol* 2004; 18: 69–73.

27. Cahill D. The risks of GnRH agonist administration in early pregnancy. Ovulation Induction Update ‘98. Parthenon, London, 1998, pp.97–106.

28. Sahin S, Ozay A, Ergin E, et al. The risk of ectopic pregnancy following GnRH agonist triggering compared with hCG triggering in GnRH antagonist IVF cycles. *Arch Gynecol Obstet* 2015; 291: 185–191. DOI: 10.1007/s00404-014-3399-x.

29. Zhou W, Zhuang Y, Pan Y, et al. Effects and safety of GnRH-a as a luteal support in women undertaking assisted reproductive technology procedures: follow-up results for pregnancy, delivery, and neonates. *Arch Gynecol Obstet* 2017; 295: 1269–1275. DOI: 10.1007/s00404-017-4353-5.

30. Balayla J, Sheehy O, Fraser WD, et al. Neurodevelopmental outcomes after assisted reproductive technologies. *Obstet Gynecol* 2017; 129: 265–272. DOI: 10.1097/AOG.0000000000001837.

31. Zhan QT, Pan PP, Xu XR, et al. An overview of studies on psychological well-being in children born following assisted reproductive technologies. *J Zhejiang Univ Sci B* 2013; 14: 947–960. DOI: 10.1631/jzus.B1300101.