Characterization of Breast Tumors Using Diffusion Kurtosis Imaging (DKI)

Dongmei Wu1,2,3, Guanwu Li1,3, Junxiang Zhang4, Shixing Chang3*, Jiani Hu2, Yongming Dai5

1 Shanghai Key Laboratory of Magnetic Resonance, East China Normal University, Shanghai, China, 2 Department of Radiology, Wayne State University, Detroit, Michigan, United States of America, 3 Department of Radiology, Yue Yang Hospital of Integrated Traditional Chinese and Western Medicine, Shanghai University of Traditional Chinese Medicine, Shanghai, China, 4 Department of Radiology, The First Affiliated Hospital of Bengbu Medical College, Bengbu, Anhui Province, China, 5 Philips Healthcare, Shanghai, China

Abstract

Aim: The aim of this study was to investigate and evaluate the role of magnetic resonance (MR) diffusion kurtosis imaging (DKI) in characterizing breast lesions.

Materials and Methods: One hundred and twenty-four lesions in 103 patients (mean age: 57 ± 14 years) were evaluated by MR DKI performed with 7 b-values of 0, 250, 500, 750, 1,000, 1,500, 2,000 s/mm² and dynamic contrast-enhanced (DCE) MR imaging. Breast lesions were histologically characterized and DKI related parameters—mean diffusivity (MD) and mean kurtosis (MK)—were measured. The MD and MK in normal fibroglandular breast tissue, benign and malignant lesions were compared by One-way analysis of variance (ANOVA) with Tukey’s multiple comparison test. Receiver operating characteristic (ROC) analysis was performed to assess the sensitivity and specificity of MD and MK in the diagnosis of breast lesions.

Results: The benign lesions (n = 42) and malignant lesions (n = 82) had mean diameters of 11.4 ± 3.4 mm and 35.8 ± 20.1 mm, respectively. The MK for malignant lesions (0.88 ± 0.17) was significantly higher than that for benign lesions (0.47 ± 0.14) (P < 0.001), and, in contrast, MD for benign lesions (1.97 ± 0.35 (10⁻³ mm²/s)) was higher than that for malignant lesions (1.20 ± 0.31 (10⁻³ mm²/s)) (P < 0.001). At a cutoff MD/MK 1.58 (10⁻³ mm²/s)/0.69, sensitivity and specificity of MD/MK for the diagnosis of malignant were 79.3%/84.2% and 92.9%/92.9%, respectively. The area under the curve (AUC) is 0.86/0.92 for MD/MK.

Conclusions: DKI could provide valuable information on the diffusion properties related to tumor microenvironment and increase diagnostic confidence of breast tumors.

Introduction

Breast MRI has been increasingly used in diagnosing patients with suspicious breast lesions owing to its non-invasive nature and excellent soft-tissue contrast as compared with other diagnostic imaging modalities [1–4]. In particular, diffusion MRI is a non-invasive MR imaging technique that allows in vivo characterization and quantification of the molecular water diffusion in tissues [5–7]. Specifically, measurement of functional parameters that reveal the water diffusion in microscopic environment can serve to characterize the pathological conditions of breast lesions [8]. For instance, mean diffusivity (MD) is a measure of the water diffusivity in diffusion-weighted imaging (DWI). The measurement of such water diffusion rate has been shown to help distinguish between normal fibroglandular breast tissue and benign or malignant lesions [9–17]. However, benign lesions and malignant cancers cannot always be discriminated accurately from each other because of the confounding overlap in their diffusion rate values. As such, advanced diffusion MR imaging techniques which may provide better characterization of breast tumor tissues are highly desirable to assist the classification and differential diagnosis of breast lesions.

Tumor invasiveness (grade and aggressiveness) is highly related to the pathophysiologic features of tumor tissues such as...
cytological patterns or organization, cellular structure and density [10], which are indirectly reflected in molecular diffusion properties of water molecules. Conventional diffusion MRI techniques always assume a Gaussian diffusion (i.e., free and unrestricted diffusion) of water protons. In fact, water diffusion in the complex biological tissues has a non-Gaussian distribution of water displacement profile in the presence of diverse barriers and compartmentalization that restrict the free displacement of water molecules, instead of a simplified Gaussian probability density function [19–23]. Indeed, non-mono-exponential diffusion-weighted (DW) signal decay has been observed with high b-values, likely due to restricted water diffusion associated with the underlying microstructures [23–26]. Accordingly, measurement of diffusional non-Gaussianity (i.e., diffusion kurtosis), a measure of diffusional heterogeneity, by means of diffusion kurtosis imaging (DKI) may allow improved characterization of water diffusion properties in the tumor microenvironment.

Recently, DKI parameters have been proved to be able to indicate microstructural changes within cerebral glioma tissue that affect the way that molecule diffuse and to allow better differentiation among cerebral glioma grades than those of conventional DWI [27,28]. Moreover, the applications of DKI in characterization of the non-gaussian diffusion behavior have been successfully extended from cerebral gliomas to hepatic fibrosis [29], hepatic carcinoma [30] and prostate cancer [31]. In this study, we hypothesize that DKI could provide additional information about the water diffusion in the breast tumor microenvironment as compared with the conventional DWI. The aim of this study was to investigate and evaluate the role of DKI in characterizing breast lesions by examining the relationship between DKI parameters and tumor types in human patients with breast tumors.

Materials and Methods

Ethics Statement

All research procedures were approved by our institutional review boards (Yue Yang Hospital of Integrated Traditional Chinese and Western Medicine and The First Affiliated Hospital of Bengbu Medical College). Written informed consent for each study was obtained independently for all patients.

Study Population and Pathological Examination

In total, 103 female patients (mean age = 57±14 years; age range = 24 to 87 years) with suspicious breast lesions (N = 124) were included in this study from September 2012 to June 2014. Following breast MRI, needle or excision biopsies were performed on patients to obtain lesion samples. All biopsy specimens were fixed in 10% formalin and embedded in paraffin. The specimens were then cut and stained with hematoxylin and eosin (H&E), followed by histological examination. The lesion classification based on pathological analysis of biopsy specimens was considered as standard reference. Each breast lesion was assigned as either benign or malignant. There were 42 benign lesions in 30 patients, including 17 fibroadenomas, 9 fibrocytic changes, 16 cysts; and 82 malignant lesions in 73 patients, including 27 infiltrating ductal carcinomas, 21 ductal carcinomas in situ, 23 infiltrating lobular carcinomas, 1 lobular carcinoma in situ.

Breast MR Imaging

Bilateral breast MR imaging was performed on two 3.0 Tesla MR systems (MAGNETOM Verio; Siemens Medical Solutions, Erlangen, Germany; Achieva TX, Philips Healthcare, Best, The Netherlands) with a maximum gradient strength of 45(40) mT/m and a maximum slew rate of 200 T/m/s. 46 patients (mean age = 59±14 years; age range = 31 to 87 years) were scanned in a prone position on Siemens scanner with a dedicated phased-array 4-channel bilateral breast coil; while 57 patients (mean age = 55±13 years; age range = 24 to 79 years) were scanned in prone position on Philips scanner with a dedicated phased-array 7-channel bilateral breast coil. T2-weighted images were first acquired with interleaved multi-slice turbo spin-echo (TSE) sequence with repetition time (TR) = 3000 ms, echo time (TE) = 61 ms, Inversion Time (TI) = 230 ms, turbo factor = 16, field of view (FOV) = 320×320 mm², acquisition matrix size = 320×320, slice thickness = 4.5 mm, number of slices = 30, parallel imaging acceleration factor = 2 using generalized autocalibrating partially parallel acquisition (GRAPPA, Siemens) or Sensitivity Encoding (SENSE, Philips), number of averages (NA) = 2, and acquisition time = 2 minutes. For DKI, interleaved multi-slice DWI was performed using a single-shot spin-echo planar imaging (EPI) sequence with 7 b-values of 0, 250, 500, 750, 1000, 1500 and 2000 s/mm² in x, y and z directions with TR = 4000 ms, TE = 100 ms, receiver bandwidth = 1184 Hz/Pix, FOV = 166×380 mm², acquisition matrix = 84×192, slice thickness = 7.0 mm, number of slices = 12, parallel imaging acceleration factor = 2 using GRAPPA or SENSE, fat suppression achieved by spectral adiabatic inversion recovery (SPAIR), “distortion correction” = Yes, NA = 1, and acquisition time = 2.7 minutes. Dynamic contrast-enhanced T1-weighted MRI was then performed using fat-saturated three-dimensional (3D) fast low-angle shot (FLASH) sequence with TR = 4.73 ms, TE = 1.71 ms, flip angle (FA) = 10°, FOV = 320×320 mm², acquisition matrix = 384×384, slice thickness = 1.2 mm, number of slices = 128, parallel imaging acceleration factor = 2 using GRAPPA or SENSE, NA = 1, and acquisition time = 50 sec for each time point, before (pre-contrast) and five times after injection (post-contrast) of gadolinium chelate (Magnevist; Bayer Healthcare, Berlin, Germany) at a dose of 0.1 mmol/kg at a rate of 2 mL/s followed by a flush of saline solution.

Image and Data Analysis

MR images were analyzed in MATLAB (MathWorks, Natick, MA). Apparent diffusion coefficient (ADC) and apparent kurtosis coefficient (AKC) along each diffusion gradient direction were derived per voxel simultaneously by least-square fitting the DW signals non-linearly to:

\[
S(b) = S_0 \exp \left(-b \cdot ADC + \frac{1}{6} b^2 \cdot AKC \right)
\]

where \(S(b)\) is the DW signal at a particular b-value and \(S_0\) is the signal without diffusion weighting. Note that we excluded the b0 (i.e., b-value =0 s/mm²) images from the numerical fitting to minimize blood perfusion effects on the DW signal decay. Background voxels with signal intensities below a threshold value in the b0 images were excluded from the fitting to increase efficiency. MD and MK maps were then calculated as the average of the parametric ADC and AKC maps along all applied diffusion gradient directions, respectively [23]. Contrast-enhancement maps were calculated as the subtraction images of the post-contrast and pre-contrast dynamic contrast-enhanced T1-weighted images. Contrast-enhancement curve (percentage signal intensity enhancement versus image frame) was also evaluated. Region of interest (ROI) analysis was performed manually by delineating breast lesions on the DW b1000 (b-value =1000 s/mm²) images with reference to the contrast-enhanced T1-weighted images. Care was taken to avoid region’s cyst and necrosis that were hyperintense on
both T2W images and MD maps, as well as regions of adipose tissue that were hypointense on b0 image. ROI with similar size was also defined in normal fibroglandular breast tissue in the contralateral breast. Lesion and normal fibroglandular breast tissue ROIs were then transferred and used for MD and MK measurements on the MD and MK maps. Lesion sizes were defined as the longest dimension on DCE-MRI. Results were expressed as mean ± standard deviation (SD). Prior to the analysis of DKI parameters for lesions, the study performed on two different scanners necessitates a pilot analysis to evaluate the vendor-specific differences of DKI. An independent samples t-test was used to compare DKI parameters for the ROIs of normal fibroglandular breast tissue, which were divided into two groups according to the scanners. Then, One-way analysis of variance (ANOVA) with Tukey’s multiple comparison test was performed to compare the MD and MK values among normal fibroglandular breast tissue, benign and malignant tissues. A P-value of less than 0.05 was considered statistically significant. In addition, receiver operating characteristic (ROC) analysis was carried out to determine suitable MD and MK cutoff values for discrimination between benign and malignant lesions and to assess the diagnostic performance of DKI.

### Results

57 ROIs of normal fibroglandular breast tissue (31.5 ± 23.7 mm, range = 7–86 mm) obtained on Siemens scanner had mean values of 2.22 ± 0.41 (10^{-3} \text{ mm}^2/\text{s}) and 0.55 ± 0.15 for MD and MK respectively, while 67 ROIs of normal fibroglandular breast tissue (24.2 ± 10.2 mm, range = 11–70 mm) obtained on Philips scanner had mean values of 2.14 ± 0.36 (10^{-3} \text{ mm}^2/\text{s}) and 0.53 ± 0.14 for MD and MK respectively. No significant differences were found for MD (P = 0.23) and MK (P = 0.54) from these two groups.

Table 1 summarizes the breast tumor types in the patient population. The mean sizes of benign and malignant lesions were 11.4 ± 3.4 mm (range = 7–18 mm) and 35.8 ± 20.1 mm (range = 11–67 mm), respectively. Figure 1 shows the DW signal decay of breast lesion in two patients with benign and malignant lesions separately, along with the fitted curve using mono-exponential model and DKI model (Eq. 1).

Figure 2 shows the T2-weighted TSE images, b0 images, MD maps, MK maps, as well as the contrast enhancement maps, Maximum Intensity Projection (MIP) of enhancement maps, and contrast enhancement curves of a patient with benign breast tumor – fibroadenomas. Similarly, Figure 3 shows the T2-weighted TSE images, b0 images, MD maps, MK maps, as well as the contrast enhancement maps, MIP of enhancement maps, and contrast enhancement curves of a patient with malignant carcinoma – infiltrating ductal carcinomas.

Figure 4 compares the MD and MK values in contralateral fibroglandular tissues, benign and malignant lesions. The MD values are 2.18 ± 0.49 (10^{-3} \text{ mm}^2/\text{s}), 1.97 ± 0.35 (10^{-3} \text{ mm}^2/\text{s}) and 1.20 ± 0.31 (10^{-3} \text{ mm}^2/\text{s}), while MK values are 0.54 ± 0.18, 0.47 ± 0.14, 0.88 ± 0.17 for contralateral fibroglandular tissues.

---

**Table 1. Tumor types confirmed by pathological examination.**

| Benign          | Malignant          |
|-----------------|--------------------|
| FAs (n = 17, 11.8 ± 3.1 mm) | IDC (n = 27, 37.3 ± 22.8 mm) |
| FC (n = 9, 9.6 ± 2.3 mm)   | DCIS (n = 21, 29.0 ± 9.0 mm)  |
| Gysts (n = 16, 12.1 ± 3.9 mm) | ILC (n = 23, 37.8 ± 21.1 mm)  |
|                 | LCIS (n = 11, 39.8 ± 25.7 mm) |

Note. – FC = fibrocystic changes; FAs = fibroadenomas; DCIS = ductal carcinomas in situ; IDC = infiltrating ductal carcinomas; LCIS = lobular carcinoma in situ; ILC = infiltrating lobular carcinoma.

doi:10.1371/journal.pone.0113240.t001

---

**Figure 1.** DW signal decay of breast lesion at 7 b-values of 0, 250, 500, 750, 1000, 1500 and 2000 s/mm² was fitted using mono-exponential model and DKI model: a) a 59 year-in-old woman with fibroadenomas and b) a 43 year-in-old woman with ductal carcinomas in situ.

doi:10.1371/journal.pone.0113240.g001
benign and malignant lesions, respectively. The results reveals that both MD and MK values in fibroglandular tissue, benign and malignant lesions are significantly different ($P<0.001$).

Figure 5 shows the ROC curves for the evaluation of the ability of MD and MK values to differentiate between benign and malignant lesions. Results of ROC analysis are presented in Table 2. The area under curve (AUC) for MD was 0.86 with 95% confidence interval being 0.79–0.92. Using $1.58 \times 10^{-3} \text{mm}^2/\text{s}$ as MD cutoff value between benign and malignant lesions, the sensitivity was 79.3% and specificity was 92.9%; 20.7% were misclassified as benign, while 7.1% were misclassified as malignant. For MK, the AUC was 0.92 and the 95% confidence interval was 0.83–0.96. Using 0.69 as MK cutoff value between benign and malignant lesions, the sensitivity was 84.2% and specificity was 92.9%; 15.8% were misclassified as benign, while 7.1% were misclassified as malignant.

**Discussion**

Accurate diagnosis and classification of breast lesions have always been challenging using conventional mammography and ultrasound, especially in dense fibroglandular breasts [32]. Improved characterization of breast tumor type and degree of malignancy of breast cancer could greatly assist treatment planning and hence help improve patient’s outcome. Diffusion MRI is a non-invasive functional imaging technique for providing...
valuable information on the water diffusion properties in tumor microenvironment. In particular, DKI can potentially augment conventional diffusion techniques for better breast lesion characterization. Recent studies have shown that DKI offers a method to evaluate the non-gaussian diffusion behavior in complex biological tissues in various brain diseases, including ischemic stroke, Parkinson diseases, traumatic brain injuries, and brain gliomas [27,33–36]. The present study aimed to examine the capability of DKI in differentiating benign from malignant breast lesions.

Our study results demonstrate the potential utility of DKI for the characterization of breast lesions. The results of ROC analysis suggest that the use of MK lead to a higher sensitivity and specificity and lower percentage for misclassification than those of MD in determining benign and malignant breast lesions, indicating a better diagnostic performance, figure 5. Our study results also show that significantly higher MK values are found in malignant lesions, indicative of higher non-Gaussian diffusion, i.e., higher cellular complexity, than in benign lesions, as theoretically expected.

The behavior of DW signal with higher b-values, whether in benign or malignant lesions, is better characterized by DKI model rather than mono-exponential model, figure 1. It has been suggested that molecular motion of water becomes more restricted because of decreasing extracellular space as tumor cells proliferate and hence cellularity increases. This may reflect microstructural differences between benign and malignant tissues. Moreover,

Figure 3. A 51 year-in-old woman with infiltrating ductal carcinomas, indicated by the white arrow: a) T2-weighted TSE image; b) DW image at b = 0; c) MD map; d) MK map; e) contrast enhancement map; f) MIP of enhance map; g) contrast enhancement curves and h) histological specimen.

doi:10.1371/journal.pone.0113240.g003
moderate correlation between water diffusivity and tumor cellularity has been illustrated previously. Changes in DKI parameters may reflect physiologic and morphologic alterations associated with breast tumor tissues. MK may be related to the degree of microstructural complexity. DKI may add valuable indications of microstructural changes to conventional diffusion techniques for the characterization of breast tumors, however, the mechanisms underlying the differences in MK alteration between benign tumors and malignant cancers have yet to be determined.

Although DCE-MRI using gadolinium chelates offers high sensitivity in detecting breast lesions and hence has been widely used for locating multiple small breast tumors, it suffers from low specificity in characterizing tumor tissues [37]. Indeed, certain benign breast lesions enhance in a similar fashion to malignant cancers [38,39], which is confirmed in our study that the contrast enhancement curves for both benign and malignant lesions are in the same plateau pattern, figure 2 and 3. As a result, contrast enhancement is thought to be attributed to the proliferating activity of the hyperplastic or neoplastic parenchymal cells, in addition to the vessel density associated with tumor angiogenesis [40,41]. Therefore, it is sometimes impossible to distinguish enhancing benign tumors from malignant carcinomas unambiguously based on the contrast enhancement patterns and morphologic features alone. Nonetheless, diffusion MRI could complement DCE-MRI in these circumstances [42–46]. In addition, the results of this study suggest that the potential toxicity of gadolinium chelates limits the accessibility of DCE-MRI for patient with compromised kidney functions. In contrast, DKI does not require contrast agent administration and hence is more suitable for multiple longitudinal follow-up studies for monitoring tumor growth and response to therapy.

It is worthwhile to note that pronounced microvascular perfusion associated with tumor angiogenesis may lead to bi-exponential DW signal decay in breast tumor tissue [47,48]. Indeed, MR signal attenuation in a voxel of highly perfused tissue is associated with the combined effects of molecular water diffusion and blood perfusion in the presence of magnetic field gradient [49,50]. The contribution of micro-perfusion to the DW signal loss arises from intra-voxel spin dephasing due to pseudorandom motion of moving blood protons in the microvasculature, also known as intra-voxel incoherent motion [49]. Nevertheless, the blood microcirculation only dominates the pseudo-diffusion signal attenuation prominently at low b-values [8]. As such, we performed diffusion kurtosis analysis by fitting the DW signals with b-values starting from a non-zero b-value (250 s/mm²) to minimize the effects of pseudorandom vascular perfusion [51–53] in the current study.

Breasts consist of an exceptionally high content of fatty tissue. It has been shown that adipose tissue in breasts yields significantly lower MD value than normal fibroglandular tissue [54]. As a result, effective fat suppression is essential in breast DKI to minimize any partial volume effects from intravoxel fat signals [55]. It has also been suggested that the choice of fat suppression

Figure 4. Box plot distribution: a) MD values for fibroglandular tissue, benign and malignant lesions; b) MK values for fibroglandular tissue, benign and malignant lesions. Outliers are also represented. Top and bottom of each box represent 25% and 75% percentiles of the MD and MK values, respectively. Horizontal line inside each box represents median value.

doi:10.1371/journal.pone.0113240.g004

Figure 5. Receiver operating characteristic curve for MD and MK values used as predictors of malignancy in 124 breast lesions in 103 patients. Straight diagonal line spanning the middle of the graph indicates an AUC of 0.5.

doi:10.1371/journal.pone.0113240.g005
they exhibit low MD value.

By exceptionally low signal intensity on non-DW b0 images, even in breast adipose tissue, our study shows that breast adipose tissue can be easily identified. Note that breast adipose tissue homogeneity could suppress fat signal in breast effectively as shown in our study. Note that breast adipose tissue can be easily identified by exceptionally low signal intensity on non-DW b0 images, even they exhibit low MD value.

In our study, DKI was performed based on a single-shot spinecho EPI sequence with 7 b-values in three orthogonal directions, and the MK was computed by averaging AKC over all directions. Indeed, the use of average AKC for the approximation of MK took an assumption of isotropic diffusion for simplicity, which might introduce the rotation variations and hence deteriorate the validity of MK in the cases of anisotropic diffusion. For improvement, a tensor could be used to describe the dependency of the measured diffusional kurtosis on the direction of the diffusion sensitizing gradients [23]. Nonetheless, the use of kurtosis tensor has been challenged for its lengthy acquisition time and complex postprocessing [58], involving at least 15 independent components of the tensor [23]. As such, rapid protocol and robust postprocessing approaches targeted for diffusion kurtosis tensor imaging remain necessary for the future DKI investigation.

Several potential limitations exist in the current study. First, we did not evaluate the influences of menopausal status and menstrual cycle on DKI parameters in breast tumor tissues. Slight variation in normal breast MD owing to hormonal fluctuations and hence water content throughout menstrual cycle has been reported previously [59]. Second, the findings in the current study were preliminary, a further multi-center study with large cohort is necessary. Further clinical evaluation on a larger patient population is warranted. Third, the non-linear squared fitting method used for DKI parameters computation in this study is susceptible to the noise for the low signal-to-noise data at high b-values. Therefore, advanced method such as multi-step weighted linear least squared (WLLS) approach [60], which could provide high performance in terms of accuracy/precision, is necessary. Fourth, histopathological evaluation is subject to both observer variation and variability based on the spatial focus of observation. Fifth, malignant lesions with central necrosis often show high ADC values in the area of necrosis and the rim of the lesion may be too thin for correct ROI placement. Nonfocal mass lesions as often seen in DCIS may not be categorized correctly with DWI even with a small ROI due to diffuse tumor spread and partial volume effects.

In conclusion, the results of this study suggest that DKI could provide valuable information on the diffusion properties related to tumor microenvironment by the simultaneous quantification of both MD and MK. DKI could improve tissue characterization of breast lesions and increase diagnostic confidence of breast tumors. Further studies with larger sample size are warranted to explore the full potential of DKI for non-invasive imaging of human breast lesions in clinical setting. Once fully validated, breast DKI may also serve as a non-contrast breast screening imaging technique, avoiding unnecessary biopsies.

**Author Contributions**

Conceived and designed the experiments: DMW GWL. Performed the experiments: GWL JXZ. Analyzed the data: DMW GWL YMD JNH. Contributed reagents/materials/analysis tools: DMW GWL SJC YMD JNH. Wrote the paper: DMW GWL YMD.

---

**Table 2. Receiver operating characteristic (ROC) analysis for the accuracy of MD and MK in the diagnosis of breast lesions.**

| Parameters | AUC | Cutoff Value | Sensitivity | Specificity | P value |
|------------|-----|--------------|-------------|-------------|---------|
| MD         | 0.86| 1.58 (10^−3 mm²/s) | 79.3%       | 92.9%       | <0.001  |
| MK         | 0.92| 0.69         | 84.2%       | 92.9%       | <0.001  |

Note: AUC = area under the curve; MD = Mean Diffusivity; MK = Mean Kurtosis.

doi:10.1371/journal.pone.0113240.t002

---

**References**

1. Rankin SC (2000) MRI of the breast. Br J Radiol 73 (872): 806–18.
2. Kneeshaw PJ, Turnbull LW, Drew PJ (2005) Current applications and future direction of MR mammography. Br J Cancer 81 (1): 4–10.
3. Sinha S, Sinha U (2009) Recent advances in breast MRI and MRS. NMR Biomed 22 (1): 3–16.
4. Petralia G, Bonollo L, Prisco F, Demeo P, Bellomi M (2011) Breast MR with special focus on DW-MRI and DCE-MRI. Cancer Imaging 11: 76–80.
5. Wesbege GE, Moseley ME, Elman RL (1984) Translational molecular self-diffusion in magnetic resonance imaging. II. Measurement of the self-diffusion coefficient. Invest Radiol 19 (6): 491–8.
6. Le Bihan D, Breton E, Lallemand D, Grenier P, Cabanas E, et al. (1986) MR imaging of intravoxel incoherent motions: application to diffusion and perfusion in neurologic disorders. Radiology 161 (2): 401–7.
7. Basser PJ (1995) Inferring microstructural features and the physiological state of tissues from diffusion-weighted images. NMR Biomed 8 (7-8): 333–44.
8. Sinha S, Sinha U (2002) Functional magnetic resonance of human breast tumors: diffusion and perfusion imaging. Ann N Y Acad Sci 980: 95–115.
9. Guo Y, Cai YQ, Cai ZL, Gao YG, An NY, et al. (2002) Differentiation of clinically benign and malignant breast lesions using diffusion-weighted imaging. J Magn Reson Imaging 16 (2): 172–8.
10. Sinha S, Lucas-Quezada FA, Sinha U, DeBruhl N, Bassett LW (2002) In vivo diffusion-weighted MRI of the breast: potential for lesion characterization. J Magn Reson Imaging 15 (6): 695–704.
11. Woodhams R, Matsunaga K, Iwabuchi K, Kau S, Hata H, et al. (2005) Diffusion-weighted imaging of malignant breast tumors: the usefulness of apparent diffusion coefficient (ADC) value and ADC map for the detection of malignant breast tumors and evaluation of cancer extension. J Comput Assist Tomogr 29 (5): 649–49.
12. Rubesova E, Griell AS, De Maertelaer V, Metens T, Chao SL, et al. (2006) Quantitative diffusion imaging in breast cancer: a clinical prospective study. J Magn Reson Imaging 24 (2): 319–24.
13. Marini C, Iacconi C, Giannelli M, Cilotti A, Moretti M, et al. (2007) Quantitative diffusion-weighted MR imaging in the differential diagnosis of breast lesions. Eur Radiol 17 (10): 2646–55.
14. Lo GG, Ai V, Chan JK, Li KW, Cheung PS, et al. (2009) Diffusion-weighted magnetic resonance imaging of breast lesions: first experiences at 3 T. J Comput Assist Tomogr 33 (1): 63–9.
15. Costantini M, Berti P, Rinaldi P, Buù E, Giardina G, et al. (2010) Diffusion-weighted imaging in breast cancer: relationship between apparent diffusion coefficient and tumour aggressiveness. Clin Radiol 65 (12): 1005–12.
16. Eh Khoras RH, Jacobs MA, Mezban SD, Huang P, Kamel IR, et al. (2010) Diffusion-weighted imaging improves the diagnostic accuracy of conventional 3-T breast MR imaging. Radiology 256 (1): 64–73.
17. Partridge SC, Demartini WB, Kurland BF, Eby PR, White SW, et al. (2010) Differential diagnosis of mammographically and clinically occult breast lesions on diffusion-weighted MRI. J Magn Reson Imaging 31 (3): 562–70.
10. Ducatman BS, Emery ST, Wang HH (1995) Correlation of histologic grade of breast carcinoma with cytologic features on fine-needle aspiration of the breast. Mod Pathol 8 (5): 539–43.

19. Le Bihan D (1995) Molecular diffusion, tissue microdynamics and microstructure. NMR Biomed 8 (7–8): 375–86.

20. Alexander DC, Barker CJ, Arridge SR (2002) Detection and modeling of non-Gaussian apparent diffusion coefficient profiles in human brain data. Magn Reson Med 48 (2): 331–40.

21. Liu C, Ramm R, Acar R, Moseley ME (2004) Characterizing non-Gaussian diffusion by using generalized diffusion tensors. Magn Reson Med 51 (5): 924–37.

22. Assaf Y, Freidlin RZ, Rohde GK, Basser PJ (2004) New modeling and experimental framework to characterize hindered and restricted water diffusion in brain white matter. Magn Reson Med 52 (5): 965–78.

23. Jensen JH, Helpern JA, Ramani A, Lu H, Katzynski K (2005) Diffusional kurtosis imaging: the quantification of non-gaussian water diffusion by means of magnetic resonance imaging. Magn Reson Med 53 (6): 1452–60.

24. Jensen JH, Helpern JA (2010) MRI quantification of non-Gaussian water diffusion by kurtosis analysis. NMR Biomed 23 (7): 690–710.

25. Wu GX, Churger MM (2010) MR diffusion kurtosis imaging for neural tissue characterization. NMR Biomed 23 (7): 836–46.

26. De Santis S, Gabrielli A, Palombo M, Maraviglia B, Capuani S (2011) Non-Gaussian diffusion imaging: a brief practical review. Magn Reson Imaging 29 (10): 1410–6.

30. Rosenkranz AB, Sigmund EE, Winnick A, Niver BE, Spieler B, et al. (2012) Diffusion kurtosis indices: preliminary experience in live human explants. Magn Reson Imaging 30 (10): 1534–40.

33. Jansen JF, Stambuk HE, Kouchner JA, Shukla-Dave A (2010) Non-gaussian analysis of diffusion-weighted MR imaging in head and neck squamous cell carcinoma: A feasibility study. AJNR Am J Neuroradiol 31 (4): 741–8.

34. Bones N, Bihl J, Hebi F, Hedges J, Smith R, et al. (2019) The variability of fibroadenoma in contrast-enhanced dynamic MRI mammography. AJR Am J Roentgenol 165 (5): 1331–4.

35. Hochman MG, Ogel SG, Powell CM, Schnall MD, Reynolds CA, et al. (1997) Fibroadenomas: MRI imaging appearances with radiologic-histopathologic correlation. Radiology 204 (1): 123–9.

40. Bone B, Wiiberg MK, Parrado C, Falkmer U, Aspelin P, et al. (1998) Mechanism of contrast enhancement in breast lesions at MR imaging. Acta Radiol 39 (5): 494–500.

41. Stomper PC, Winston JS, Herman S, Klapperstein DL, Arredondo MA, et al. (1997) Angiogenesis and dynamic MR imaging gadolinium enhancement of malignant and benign breast lesions. Cancer Res Treat 45 (1): 39–46.

42. Kinoshita T, Yashiro N, Hara N, Funato H, Fukushima E, et al. (2002) Diffusion-weighted half-Fourier single-shot turbo spin echo imaging in breast tumors: differentiation of invasive ductal carcinoma from fibroadenoma. J Comput Assist Tomogr 26 (6): 1042–6.

43. Yabuuchi H, Matsuo Y, Okafuji T, Kamitani T, Soeda H, et al. (2008) Enhanced mass on contrast-enhanced breast MR imaging: lesion characterization using combination of dynamic contrast-enhanced and diffusion-weighted MR images. J Magn Reson Imaging 28 (5): 1157–65.

44. Partridge SC, DeMartini WB, Kurland BF, Eby PR, White SW, et al. (2009) Quantitative diffusion-weighted imaging as an adjunct to conventional breast MRI for improved positive predictive value. AJR Am J Roentgenol 193 (6): 1716–22.

45. Yabuuchi H, Matsuo Y, Kamitani T, Setoguchi T, Okafuji T, et al. (2010) Non-linear like enhancement on contrast-enhanced breast MR imaging: lesion characterization using combination of dynamic contrast-enhanced and diffusion-weighted MR images. Eur J Radiol 75 (1): e126–32.

46. Partridge SC, Rahbar H, Murthy R, Chai X, Kurland BF, et al. (2011) Improved diagnostic accuracy of breast MRI through combined apparent diffusion coefficients and dynamic contrast-enhanced kinetics. Magn Reson Med 65 (6): 1579–87.

47. Param Y, Bendel P, Margalit R, Dogani H (2004) Water diffusion in the different microenvironments of breast cancer. NMR Biomed 17 (4): 170–80.

48. Sigmund EE, Cho GY, Kim S, Finn M, Mccoidal M, et al. (2011) Intravoxel incoherent motion imaging of tumor microenvironment in locally advanced breast cancer. Magn Reson Med 65 (5): 1347–57.

49. Le Bihan D, Betton E, Lallemant D, Aubin ML, Vignaud J, et al. (1988) Separation of diffusion and perfusion in intravoxel incoherent motion MR imaging. Radiology 168 (2): 497–505.

50. Yamada I, Aung W, Himeiro Y, Nakagawa T, Shibuya H (1999) Diffusion coefficients in abdominal organs and hepatic lesions: evaluation with intravoxel incoherent motion echo-planar MR imaging. Radiology 210 (3): 617–23.

51. Bogner W, Gruber S, Pinker K, Grauber G, Stadlbauer A, et al. (2009) Diffusion-weighted MR for differentiation of breast lesions at 3.0 T: how does selection of diffusion protocols affect diagnosis? Radiology 253 (2): 341–51.

52. Pereira FP, Martins G, Figueiredo E, Domingues MN, Domingues RG, et al. (2009) Assessment of breast lesions with diffusion-weighted MRI: comparing the use of different b values. AJR Am J Roentgenol 193 (4): 1030–5.

53. Peters NH, Vincken KL, van den Bosch MA, Luijten PR, Mali WP, et al. (2010) Quantitative diffusion weighted imaging for differentiation of benign and malignant breast lesions: the influence of the choice of b-values. J Magn Reson Imaging 31 (5): 1160–5.

54. Englander SA, Utag AM, Brem R, Glionc JD, van Zijl PC (1997) Diffusion-weighted human imaging. NMR Biomed 10 (7): 348–52.

55. Partridge SC, Singer L, Sun R, Wilmes LG, Kliota CS, et al. (2011) Diffusion-weighted MRI: influence of intravoxel fat signal and breast density on breast tumor conspicuity and apparent diffusion coefficient measurements. Magn Reson Imaging 29 (9): 1215–21.

56. Winkel E, Geppert C, Schulz-Wenditl R, Uder M, Kifer B, et al. (2007) Diffusion weighted imaging in breast MRI: comparison of two different pulse sequences. Acad Radiol 14 (9): 1077–83.

57. Baron P, Dorriss MD, Kappert P, Oudkerk M, Sijens PE (2010) Diffusion-weighted imaging of normal fibroglandular breast tissue: influence of microperfusion and fat suppression technique on the apparent diffusion coefficient. NMR Biomed 23 (4): 389–403.

58. Hansen B, Land TE, Sangill R, Jepsen SN (2013) Experimentally and computationally fast method for estimation of a mean kurtosis. Magn Reson Med 69 (6): 1754–60.

59. Partridge SC, McKinnon GC, Henry RG, Hylton NM, et al. (2000) Menstrual cycle variation of apparent diffusion coefficients measured in the normal breast using MRI. J Magn Reson Imaging 14 (4): 433–8.

60. Verraat J, Sijbers J, Sarnaet S, Leemans A, Jeurissen B (2013) Weighted linear least squares estimation of diffusion MRI parameters: strengths, limitations, and pitfalls. Neuroimage 1 (81): 335–46.