Canagliflozin, a sodium–glucose cotransporter 2 inhibitor, normalizes renal susceptibility to type 1 cardiorenal syndrome through reduction of renal oxidative stress in diabetic rats

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INTRODUCTION

Cardiorenal syndromes (CRS) are defined as disorders of the heart and kidney in which acute or chronic dysfunction in one organ might induce acute or chronic dysfunction in another organ1. Of the five types of CRS, type 1 CRS is characterized by acute kidney injury (AKI) as a result of rapid worsening of cardiac function. Indeed, AKI often develops in patients with acute myocardial infarction (MI), and is strongly associated with mortality in patients with MI. Decreased renal blood flow as a result of cardiac dysfunction and elevated renal venous congestion have been suggested to induce type 1 CRS through...
activation of the sympathetic nervous system and the renin–angiotensin–aldosterone (RAA) system. In addition, non-hemodynamic mechanisms, such as increased oxidative stress and chronic inflammation, contribute to type 1 CRS. Diabetes mellitus is an independent risk factor for the development of AKI in patients with MI, and we recently reported that augmented activation of Toll-like receptors (TLRs), germine-encoded innate immune receptors, plays roles in the susceptibility to AKI after MI in diabetic rats. However, how pharmacological treatment of hyperglycemia modifies the diabetes mellitus-induced increase in renal susceptibility to AKI remains unclear.

Sodium–glucose cotransporter 2 (SGLT2) inhibitors are newly developed glucose-lowering agents that inhibit glucose reuptake at the renal proximal tubule where most of the glucose is reabsorbed. Recent clinical trials have shown that treatment with an SGLT2 inhibitor significantly reduced not only cardiovascular events, but also renal events, including decreases in estimated glomerular filtration rate, progression of albuminuria and onset of renal replacement therapy. Amelioration of renal hyperfiltration, high glucose-induced increase in oxidative stress, inflammation and apoptosis are possible mechanisms of protection by SGLT2 inhibitors. Indeed, an SGLT2 inhibitor suppressed oxidative stress and inflammation in the kidneys of animal models. Interestingly, it has been reported that SGLT2 inhibitors increase the circulating level of β-hydroxybutyrate (βOHB), a ketone body, and that βOHB attenuates oxidative stress by upregulating anti-oxidative molecules through inhibition of histone deacetylases (HDACs). In contrast, the initial dip in estimated glomerular filtration rate by SGLT2 inhibitors treatment might increase the risk for AKI, although there is no evidence to support such a possibility. The effects of SGLT2 inhibitors on CRS in diabetes mellitus have not been specifically examined.

In the present study, we examined the effect of canagliflozin, an SGLT2 inhibitor, on the increased susceptibility to AKI after MI in diabetes mellitus. As a model of diabetes mellitus, we used Otsuka Long-Evans Tokushima Fatty rats (OLETF) at ages of 25–30 weeks. The reason for selection of OLETF in the present study is threefold. First, OLETF spontaneously develop diabetes mellitus primarily by hyperphagia as a result of a lack of cholecystokinin-A receptor in the brain, and they show the typical phenotype of type 2 diabetes mellitus (i.e., obesity, hyperinsulinemia and hypertriglyceridemia). Second, OLETF at the ages of 25–30 weeks show an early stage of diabetic nephropathy: lower serum creatinine level with protein urea than that in Long-Evans Tokushima Otsuka rats (LETO), non-diabetes mellitus control rats. Third, we previously found that renal susceptibility to type 1 CRS was higher in OLETF than in LETO. We postulated that a beneficial effect of canagliflozin on diabetes mellitus-induced worsening of CRS, if any, should be detectable in OLETF.

**METHODS**

This study was approved by the Animal Use Committee of Sapporo Medical University (16-060_17-055).

**Animals and experimental protocol**

**Administration of canagliflozin and induction of MI**

**Protocol 1**

Male LETO and OLETF (Sankyo Lab Service, Tokyo, Japan) at the ages of 25–30 weeks were used in all experiments. Rats were subcutaneously pretreated with a vehicle (dimethyl sulfoxide and polyethylene glycol; 1:1 v/v) or canagliflozin (1 mg/kg per day) for 2 weeks using osmotic minipumps (Alzet, Cupertino, CA, USA). After measurement of blood pressure and pulse rate by a tail-cuff method, rats were prepared for induction of MI or a sham operation, as in our previous studies.

In brief, the rats were anesthetized with isoflurane (1–2%), and a marginal branch of the left coronary artery was permanently ligated to induce MI. All rats were allowed ad libitum access to water, but were restricted from food for 12 h after surgery. The rats were divided into six groups: LETO-sham, LETO-MI, OLETF-sham, OLETF-MI, canagliflozin-treated OLETF-sham and canagliflozin-treated OLETF-MI groups.

**Protocol 2**

In this protocol, the effect of fasting on renal susceptibility to CRS in canagliflozin-treated OLETF was examined. OLETF treated with the vehicle or canagliflozin were fasted for 12 h and then the coronary artery of each rat was permanently ligated.

Sampling of blood and kidney tissues was carried out at 12 h after MI, because our previous studies showed that approximately two-thirds of OLETF died during a period of 48 h after MI. After measurements of blood pressure and heart rate under anesthesia, blood samples were taken through a catheter placed in the carotid artery. Then the kidneys were excised, and one kidney was quickly frozen in liquid nitrogen and stored at ~80°C, and the other kidney was fixed with 10% formaldehyde for histology.

Blood levels of glucose, βOHB and angiotensin II were measured using a Glutest-mint (Sanwa Kagaku Kenkyusho, Nagoya, Japan), Precision Xceed (Abbott, Chicago, IL, USA) and Angiotensin II EIA Kit (Sigma-Aldrich, St. Louis, MO, USA), respectively.

**Histological analyses**

Formaldehyde-fixed paraffin sections (3 μm) were stained using primary antibodies against neutrophil gelatinase-associated lipocalin (NGAL; sc-50531, 1:100; Santa Cruz Biotechnology, Santa Cruz, CA, USA) and kidney injury molecule-1 (KIM-1; KIM-1; AF3689, 1:20; R&D Systems, Minneapolis, MN, USA). The NGAL- and KIM-1-positive areas were determined in 10 randomly selected fields from six kidneys in each group. The glomerular area was calculated for analyses of NGAL and KIM-1.

**Cell culture and treatment**

NRK-52E cells, a rat renal proximal tubular cell line, were cultured with Dulbecco’s modified Eagle’s medium containing 10% bovine serum. After serum deprivation for 24 h, cells were pretreated with βOHB (Sigma-Aldrich) for 1 h followed by
24 h-incubation with 3 μmol/L of angiotensin II (Alomone Labs, Jerusalem, Israel). After 24 h, cells were harvested for immunoblotting.

**Messenger ribonucleic acid quantification**

Total ribonucleic acid (RNA) was isolated from frozen tissues by using an RNaseasy Fibrous Tissue Mini Kit (Qiagen, Valencia, CA, USA). First-strand complementary deoxyribonucleic acid (DNA) was synthesized using a SuperScript VILO™ cDNA Synthesis Kit (Life Technologies, Thermo Fisher Scientific, Wallingford, MA, USA). DNA amplification was carried out in ABI PRISM7500 (Life Technologies) by using Taqman Universal PCR Master Mix. Taqman gene expression assays are shown in Table S1. All assays were carried out in duplicate and by the standard curve method using serial complementary DNA dilution. β-Actin served as an internal control.

**Immunoblotting**

Immunoblot analysis was carried out as in our previous studies. Equal amounts of protein were analyzed by immunoblot assays using specific antibodies (Table S1).

**Measurements of lipid peroxidation and xanthine oxidase activity**

Levels of malondialdehyde (MDA) and 4-hydroxynonenal (4HNE), indicators of lipid peroxidation, and xanthine oxidase activity in renal tissues were measured using a Lipid Peroxidation Microplate Assay Kit (FR12; Oxford Biomedical Research, Oxford, MI, USA) and a Xanthine Oxidase Activity Assay Kit (ab102522; Abcam, Cambridge, UK), respectively.

**Statistical analysis**

Data are presented as the mean ± standard error of the mean. One-way ANOVA and Student–Newman–Keuls post-hoc test were used to analyze differences in data among multiple groups. Unpaired Student’s two-tailed t-test was used to determine statistical significance of two data sets. Differences in mortalities in treatment groups were examined by the χ²-test. For all tests, a difference was considered to be statistically significant if P < 0.05. All analyses were carried out with SigmaStat (Systat, San Jose, CA, USA).

**RESULTS**

**Mortality after MI**

In protocol 1, none of the sham-operated rats in the LETO (n = 10), OLETF (n = 10) or canagliflozin-treated OLETF groups (n = 10) died after surgery. In contrast, approximately 30% of the rats died within 12 h after MI: three of 14 rats in the LETO-MI, five of 16 rats in the OLETF-MI and six of 19 rats in the canagliflozin-treated OLETF-MI groups. In protocol 2, five of 15 rats in the OLETF-MI and five of 14 rats in the canagliflozin-treated OLETF-MI groups died within 12 h after MI, suggesting that a fasting condition did not influence the mortality rate in OLETF. Mortality rates did not differ in the study groups with MI.

**Metabolic and hemodynamic profiles**

OLETF had higher bodyweight and higher casual blood glucose levels than those of LETO, and treatment with canagliflozin reduced the blood glucose level without affecting bodyweight in OLETF (Table 1). Blood βOHB levels were similar in LETO, OLETF and canagliflozin-treated OLETF when blood was sampled without fasting. Mean blood pressure, but not heart rate, in a conscious state was higher by ~5 mmHg in OLETF than in LETO, and canagliflozin did not change these parameters. In protocol 2, 12 h of fasting reduced blood glucose levels in OLETF, and there was no statistical difference between glucose levels in OLETF and canagliflozin-treated OLETF. However, blood βOHB level was significantly elevated after 12 h of fasting in canagliflozin-treated OLETF.

At 12 h after the operation (and fasting), blood glucose levels were still higher in OLETF than in LETO, and treatment with canagliflozin significantly reduced the blood glucose level in OLETF (Table 2). The level of βOHB after MI was slightly

| Table 1 | Physiological parameters before induction of myocardial infarction |
|---|---|---|---|---|---|
| | n | Bodyweight (g) | Blood glucose (mg/dL) | β-hydroxybutyrate (mmol/L) | Heart rate (b.p.m.) | Mean blood pressure (mmHg) |
| Protocol 1 (without fasting before surgery) | | | | | | |
| LETO | 21 | 578 ± 15 | 139 ± 4 | 0.42 ± 0.03 | 337 ± 5 | 114 ± 2 |
| OLETF | 21 | 646 ± 10† | 343 ± 28† | 0.35 ± 0.03 | 343 ± 4 | 119 ± 1† |
| OLETF + canagliflozin | 23 | 640 ± 8† | 184 ± 9† | 0.39 ± 0.02 | 350 ± 2 | 119 ± 1† |
| Protocol 2 (fasted before myocardial infarction) | | | | | | |
| OLETF | 10 | 622 ± 10 | 164 ± 17 | 0.54 ± 0.05 | 337 ± 9 | 110 ± 4 |
| OLETF + canagliflozin | 9 | 621 ± 5 | 149 ± 9 | 0.79 ± 0.06§ | 336 ± 12 | 111 ± 4 |

Values are mean ± standard error of the mean. †P < 0.05 versus Long-Evans Tokushima Otsuka rats (LETO). §P < 0.05 versus Otsuka Long-Evans Tokushima Fatty rats (OLETF).
increased in OLETF and markedly increased in canagliflozin-treated OLETF. Although blood urea nitrogen levels were comparable in the sham-operated LETO and OLETF, serum creatinine level was lower in OLETF than in LETO, reflecting glomerular hyperfiltration associated with diabetes in this model. MI did not affect these indices of glomerular function in either LETO or OLETF. Canagliflozin treatment increased blood urea nitrogen level, but not serum creatinine level after MI. Serum angiotensin II level after MI was similar in LETO and OLETF, but not affected by canagliflozin treatment. In protocol 2, although fasting alone did not reduce NGAL levels, the extent of the NGAL-positive areas and the level of NGAL mRNA after MI in OLETF were both significantly decreased in canagliflozin-treated OLETF (Figure 1b-d).

**Immunohistochemical staining of KIM-1** showed few KIM-1 signals in LETO regardless of MI in protocol 1 (Figure 2a). KIM-1-positive cells were detected mostly as proximal tubular epithelial cells in OLETF, and the extent of KIM-1 positive areas was increased in sham-operated OLETF compared with that in LETO, and was further increased after MI. The extent of KIM-1 positive areas in OLETF was not affected by treatment with canagliflozin (Figure 2a,c). In protocol 2, however, canagliflozin with pre-MI fasting decreased the extent of the KIM-1-positive area after MI in OLETF (Figure 2b,c). The expression pattern of KIM-1 mRNA was similar to that of KIM-1 staining (Figure 2d).

**Involvement of renal TLR activation, RAA system and 5’ adenosine monophosphate-activated protein kinase activation** As we recently found involvement of TLR activation in CRS in OLETF, we assessed changes in TLR-mediated signaling after

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**Table 2** Blood analyses and hemodynamic parameters 12 h after surgery

| Protocol 1 (without fasting before surgery) | Protocol 2 (fasted before myocardial infarction) |
|-------------------------------------------|-----------------------------------------------|
| **LETO** | **OLETF** | **OLETF** | **OLETF** |
| **Sham** | **MI** | **+ canagliflozin** | **+ canagliflozin** |
| 10 | 11 | 10 | 13 |
| 129 ± 4 | 137 ± 8 | 196 ± 26 | 176 ± 15 |
| 15.6 ± 0.9 | 190 ± 1.5 | 15.6 ± 0.7 | 15.6 ± 0.7 |
| 0.49 ± 0.02 | 0.48 ± 0.03 | 0.35 ± 0.01 | 0.35 ± 0.02 |
| 7.431 ± 0.013 | 5.05 ± 1.91 | 7.426 ± 0.016 | 7.395 ± 0.008 |
| 350 ± 18 | 384 ± 14 | 280 ± 12 | 264 ± 7 |
| 92 ± 4 | 84 ± 5 | 79 ± 5 | 80 ± 4 |
| 10 | 166 ± 15 | 13 | 123 ± 7 |
| 1.37 ± 0.17 | 31.8 ± 1.7 | 29.9 ± 0.34 | 23.0 ± 0.8 |
| 19.4 ± 0.7 | 86.1 ± 20.2 | 23.0 ± 0.84 | 23.0 ± 0.88 |
| 0.40 ± 0.02 | 0.38 ± 0.02 | 0.37 ± 0.03 | 0.37 ± 0.02 |
| 87.2 ± 37.0 | 48.4 ± 35.0 | 48.4 ± 35.0 | 48.4 ± 35.0 |
| 317 ± 11 | 298 ± 10 | 297 ± 11 | 298 ± 10 |
| 85 ± 4 | 76 ± 3 |

Values are mean ± standard error of the mean. *P* < 0.05 versus Long-Evans Tokushima Otsuka rats (LETO)-Sham. ‡‡P < 0.05 versus LETO-myocardial infarction (MI). ‡P < 0.05 versus Otsuka Long-Evans Tokushima Fatty rats (OLETF)-Sham. †P < 0.05 versus OLETF-MI. ††P < 0.05 versus corresponding sham-operated rats. ††P < 0.05 versus OLETF with fasting. Measurement of angiotensin II was carried out from seven to eight samples.

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**AKI after MI in OLETF**

As in our previous study, periodic acid–Schiff staining in kidney sections did not show detectable abnormalities in tubular cells, such as loss of the brush border, tubular dilation, cast formation and cell lysis, in LETO and OLETF regardless of MI, although sizes of glomeruli were larger in OLETF (data not shown).

In protocol 1, NGAL signals in immunohistochemistry were barely detected in LETO regardless of MI (Figure 1a,c). The extent of NGAL staining of renal tubules in sham-operated OLETF was not significantly different from that in LETO. However, OLETF with MI showed focal NGAL-positive areas in tubular epithelial cells, and the NGAL-positive area was significantly larger than that in sham-operated OLETF (Figure 1a,c). In protocol 2, however, OLETF with canagliflozin treatment with or without fasting did not change the NGAL staining in OLETF. NGAL messenger RNA (mRNA) level in the kidney was higher in sham-operated OLETF than in sham-operated OLETF (Figure 1b-d). The expression level of NGAL was not affected by canagliflozin treatment. In protocol 2, although fasting alone did not reduce NGAL levels, the extent of the NGAL-positive areas and the level of NGAL mRNA after MI in OLETF were both significantly decreased in canagliflozin-treated OLETF (Figure 1b-d).

**Increased KIM-1 and tubular cell lysis** in MI in OLETF was confirmed by immunohistochemical staining of KIM-1 (Figure 1a,c). The extent of KIM-1 positive areas in OLETF was not affected by treatment with canagliflozin (Figure 1a,c). In protocol 2, however, canagliflozin with pre-MI fasting decreased the extent of the KIM-1-positive area after MI in OLETF (Figure 1b,c). The expression pattern of KIM-1 mRNA was similar to that of KIM-1 staining (Figure 1d).
MI by canagliflozin. Expression levels of TLR2, TLR4, interleukin-6, interleukin-1β and chemokine (C-C motif) ligand 3 were higher in OLETF than in LETO, although most of the differences did not reach statistical significance (Figure 3). Treatment with canagliflozin significantly reduced the mRNA level of TLR4, but not those of TLR2 or cytokines and chemokines.

The mRNA levels of these molecules involved in TLR-mediated signaling were comparable in OLETF and canagliflozin-treated OLETF in the fasting protocol. Renal mRNA level of transforming growth factor (TGF)-β was higher in OLETF than LETO after MI, suggesting that renal RAA activity was increased in diabetic rats. However, the
increased renal TGF-β mRNA level in OLETF was not altered by canagliflozin treatment with or without fasting (Figure 3b). In contrast, levels of TGF-β mRNA in the non-infarcted myocardium after MI were similar in LETO and OLETF (data not shown).

The phosphorylation level of 5’ adenosine monophosphate-activated protein kinase (AMPK)α at Thr172 in the kidney at 12 h after MI was unchanged in OLETF compared with that in LETO. The level of phospho-AMPKα was increased in canagliflozin-treated OLETF, although there was no further increase in phospho-AMPKα level by fasting in canagliflozin-treated OLETF (Figure 4b).

Oxidant stress and its regulatory proteins in the kidney

The level of MDA + 4HNE in the kidney after MI was higher in OLETF than in LETO, and this increase in OLETF was
blocked by canagliflozin only when rats were fasted for 12 h before induction of MI (Figure 5a).

Xanthine oxidase activity was significantly increased in OLETF compared with that in LETO after MI. This higher activity in OLETF was not modified by canagliflozin regardless of fasting (Figure 5b). Protein levels of both nicotinamide adenine dinucleotide phosphate oxidase (NOX)2 and NOX4 were significantly increased in OLETF compared with those in LETO. These increases in NOX2 and NOX4 proteins were significantly suppressed by canagliflozin only when rats were fasted for 12 h before MI (Figure 5c,d). The mRNA levels of NOX2 and NOX4 were higher in OLETF than in LETO, and

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**Figure 3** | Expression levels of Toll-like receptors, cytokines and chemokines in the kidney. Expression levels of genes related to (a) Toll-like receptors, and (b) cytokines and chemokines in the kidney; n = 6–11 in each group. *P < 0.05. Cana, canagliflozin; MI, myocardial infarction; Veh, vehicle.
(a) BNP
GAPDH

LETO MI OLETF MI OLETF Cana MI Veh Cana OLETF Fasting MI

(b) P-AMPKα
AMPKα

LETO MI OLETF MI OLETF Cana MI Veh Cana OLETF Fasting MI

(c) Mn-SOD
β-actin

LETO MI OLETF MI OLETF Cana MI Veh Cana OLETF Fasting MI

(d) Catalase
β-actin

LETO MI OLETF MI OLETF Cana MI Veh Cana OLETF Fasting MI

(e) SIRT3
β-actin

LETO MI OLETF MI OLETF Cana MI Veh Cana OLETF Fasting MI
the increased expression levels were also reduced by canagliflozin with fasting (Figure 5e,f).

As shown in Figure 4, protein levels of manganese superoxide dismutase, catalase and SIRT3, a mitochondrial sirtuin, in the kidney after MI were similar in OLETF and LETO, and canagliflozin treatment did not change these levels in OLETF.

**Effects of exogenous βOHb on NOX4 induction**

Angiotensin II (3 μmol/L) increased protein levels of NOX4. Pre-incubation of 1 and 2 mmol/L of βOHb attenuated the increase in NOX4 proteins by 50 and 62%, respectively (Figure 6).

**DISCUSSION**

As in our previous studies, OLETF at the ages of 25–30 weeks showed phenotypes of an early stage of diabetic nephropathy: lower serum creatinine level, indicating glomerular hyperfiltration (Table 2); no histological abnormalities in the kidney, except for larger sizes of glomeruli; and higher level of KIM-1 protein without change in NGAL level (Figures 1,2) compared with those in LETO. After acute MI, protein levels of NGAL and KIM-1 in the kidney were significantly increased by 3.2- and by 1.6-fold, respectively, in OLETF, although blood urea nitrogen and serum creatinine levels were not changed. The impact of MI on the kidney appears modest, and an unchanged level of serum creatinine indicates that renal injury in OLETF after MI is a model of subclinical AKI in patients with CRS. Subclinical AKI is not a trivial injury, as it has a significant impact on clinical outcomes. Inhibition of NOX4 in experimental models of diabetes mellitus afforded marked protection from both structural and functional kidney damage. The contribution of xanthine oxidase to reactive oxygen species-mediated renal injury in diabetes mellitus has also been indicated by the findings that inhibition of xanthine oxidase activity significantly suppressed kidney injury in diabetes mellitus. In the present study, renal expression levels of MDA + 4HNE, NOX2 and NOX4, and xanthine oxidase activity were significantly higher in OLETF than in LETO (Figure 5). As the renal TGF-β mRNA level was higher in OLETF than in LETO (Figure 3b), the increases in NOXs in OLETF might be partly explained by activation of the renal RAA system. Canagliflozin significantly reduced levels of NOX2 and NOX4, but not xanthine oxidase activity (Figure 5), whereas canagliflozin did not significantly change the TGF-β mRNA level. Renal activity of RAA system after an SGLT2 inhibitor was reported to be increased, decreased or unchanged depending on the reports. Of interest, protection against AKI by canagliflozin was detected only when the rats were fasted before induction of MI. Protective effects of canagliflozin on tissue MDA + 4HNE, and on NOX2 and NOX4 proteins were also observed when combined with fasting before MI (Figure 5a,c-f). Hemodynamic parameters, including blood pressure and heart rate, before and 12 h after MI were comparable in canagliflozin-treated OLETF with or without fasting (Tables 1,2). In contrast, the blood βOHb level was twofold higher in canagliflozin-treated OLETF with pre-MI fasting than in canagliflozin-treated OLETF without fasting (0.79 ± 0.06 vs 0.39 ± 0.02 mmol/L; Table 1), whereas blood βOHb levels after MI were increased to 3–4 mmol/L in canagliflozin-treated OLETF regardless of pre-MI fasting (Table 2). As shown in Figure 7, the level of renal NGAL expression was inversely correlated with blood βOHb.
levels before MI, but not with those at 12 h after MI, suggesting a significant association of an increase in βOHB before MI and renal protection from CRS by canagliflozin.

As the correlation between blood βOHB and renal NGAL expression was modest ($r = -0.41$), mechanisms other than increased βOHB are possibly involved in renoprotection by canagliflozin with pre-MI fasting. Activation of AMPK is one of the candidates, but the present results showed that the contribution of AMPK to renoprotection against diabetes mellitus-induced worsening of CRS was limited. It has been suggested that the reduction of oxygen consumption in the proximal tubules and prevention of cellular senescence are the mechanisms of renoprotection by an SGLT2 inhibitor, but whether these mechanisms are involved in amelioration of CRS should be examined in future studies.

As it is actually difficult to maintain the blood βOHB level by administration of exogenous βOHB to a level achieved by SGLT2 inhibitor treatment and fasting in OLETF, we examined the effects of βOHB on NOX4 protein level in vitro. Pre-incubation with βOHB attenuated angiotensin II-induced upregulation of NOX4 protein expression in NRK-52E cells (Figure 6). Recently, it was reported that βOHB functions as an endogenous HDACs inhibitor, and suppresses oxidative stress by upregulation of anti-oxidative proteins, including manganese superoxide dimutase and catalase, in the kidney. Recently, it was reported that exposure of PC12 cells to βOHB attenuated H$_2$O$_2$-induced upregulation of NOX2 and NOX4 proteins. They also showed that knockdown of either HDAC1 or HDAC2 reduced NOX2 and NO4 protein levels. Taken together, the findings suggest that elevation of blood βOHB level before MI is causally related to suppressed expression of NOXs and oxidant stress in the kidney after MI in canagliflozin-treated OLETF with pre-MI fasting. Interestingly, in contrast to suppressed expression of NOXs, upregulation of antioxidative stress proteins was not accompanied by an elevation of blood βOHB levels by canagliflozin and fasting. These results suggest that increased βOHB by canagliflozin treatment protected the kidney from injury downstream of the RAA pathway, but independent from the HDAC pathway. Precise mechanisms of how βOHB attenuate NOXs protein and subsequent renal injury are uncertain and warrant further investigation.

Figure 5 | Effects of canagliflozin and fasting on oxidative stress in the kidney after myocardial infarction. (a) Malondialdehyde (MDA) + 4-hydroxynonenal (4HNE) levels in the kidney. Kidney tissues were sampled 12 h after myocardial infarction (MI) in Long-Evans Tokushima Otsuka rats (LETO), Otsuka Long-Evans Tokushima Fatty rats (OLETF) and canagliflozin (Cana)-treated OLETF. Vehicle (Veh) or canagliflozin-treated OLETF with 12 h fasting before MI were also analyzed; n = 5–6 in each group. (b) Xanthine oxidase activity in the kidney; n = 6 in each group. (c) Representative immunoblots for nicotinamide adenine dinucleotide phosphate oxidase (NOX)2 (left) and summary data of NOX2 protein level normalized by β-actin level (right) in the kidney; n = 6 in each group. (d) Representative immunoblots for NOX4 (left) and summary data of NOX4 protein level normalized by β-actin level (right) in the kidney; n = 6 in each group. (c,d) Samples were run on the same gel but were not contiguous. (e,f) Levels of (e) NOX2 and (f) NOX4 messenger ribonucleic acid (mRNA) in the kidney; n = 6–8 in each group. *P < 0.05. AU, arbitrary unit.

Figure 6 | Effects of exogenous β-hydroxybutyrate (βOHB) on nicotinamide adenine dinucleotide phosphate oxidase (NOX4) protein in vitro. Representative immunoblots (left) and summary data (right) of NOX4 protein level normalized by vinculin are shown. (a) NRK-52E cells were treated with angiotensin II (Ang II) in the presence or absence of βOHB. (b) NRK-52E cells were treated with vehicle or βOHB; n = 6 in each treatment. *P < 0.05. AU, arbitrary unit.
Several reports have shown that treatment with an SGLT2 inhibitor ameliorated diabetic nephropathy by suppression of NOX4 expression and subsequent oxidative stress in diabetes mellitus mice. Thus, the reason why treatment with canagliflozin without fasting failed to suppress NOX4 expression and oxidative stress in the present study is uncertain. However, there are a few possible explanations. First, the duration of SGLT2 inhibitor treatment was shorter in this study than in earlier studies (2 vs 8–12 weeks), which possibly resulted in a lower level of blood ketone bodies. In fact, blood βOHB levels after 12 h fasting were higher after 8 weeks of canagliflozin treatment than that of 2 weeks of treatment (1.08 ± 0.12 vs 0.79 ± 0.06 mmol/L, P < 0.05). Second, as OLETF are hyperphagic due to a lack of cholecystokinin-A receptor in the brain, time periods of fasting might be shorter in OLETF under the condition of ad libitum feeding, leading to less elevation of blood ketone level, compared with those in db/db and Akita mice. In a clinical setting, chronic treatment with an SGLT2 inhibitor increased the blood ketone level throughout the day in patients with diabetes mellitus. Therefore, βOHB-mediated renoprotection would appear in diabetes mellitus patients treated with an SGLT2 inhibitor, but the protection might be attenuated after meal.

There were limitations to the present study. First, AKI was assessed only at 12 h after MI, and the time courses of AKI and impact of AKI on the long-term function of diabetic kidneys were not examined. The high mortality rate (i.e., approximately 60%) of OLETF within 48 h after MI precludes tissue sampling at later time points after MI in OLETF without selection bias in data. Second, treatment with canagliflozin might have affected cardiac function after MI, as SGLT2 inhibitors reportedly reduced heart failure in clinical and experimental studies. However, it is unlikely that canagliflozin had a large impact on the cardiac function 12 h after MI, as myocardial B-type natriuretic peptide levels (Figure 4a) and mortality rates after MI were not different in canagliflozin-treated and -untreated OLETF.

In conclusion, the results of the present study suggest that increased susceptibility to MI-induced AKI in diabetes mellitus rats is mediated by a NOX-induced increase in oxidative stress in the kidney, and is improved by treatment with canagliflozin with an appropriate fasting period.

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**SUPPORTING INFORMATION**
Additional supporting information may be found online in the Supporting Information section at the end of the article.

**Table S1** | Antibodies and Taqman gene expression assays.