Asymmetric Synthesis of Oxygenated Monoterpenoids of Importance for Bark Beetle Ecology

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ABSTRACT: Herein we report the asymmetric syntheses of a number of oxygenated terpenoids that are of importance in the chemical ecology of bark beetles. These are pinocamphones, isopinocamphones, pinocarvones, and 4-thujanols (= sabinene hydrates). The camphones were synthesized from isopinocamphone, the pinocarvones from β-pinene, and the thujanols from sabinene. The NMR spectroscopic data, specific rotations, and elution orders of their stereoisomers on a chiral GC-phase (β-cyclodextrin) are also reported. This enables facile synthesis of pure compounds for biological activity studies and identification of stereoisomers in mixed natural samples.

The colonization of trees by bark beetles is generally influenced by an intricate release of chemical signals in strict chronological order. The signals recruit conspecifics to a suitable host tree, and later in the colonization, other compounds are produced to convey to conspecifics that this tree is becoming overexploited.1 Thereby competition for food and larvae is avoided. These attractant chemical signals can originate from metabolized monoterpenoids, like for the noxious larger spruce bark beetle, Ips typographus, where cis-verbolen is converted to verbolenone, but the pheromones can also be synthesized de novo.2 In recent work with semiochemicals for tree-killing bark beetles we have encountered a number of oxygenated monoterpenoids that are physiologically active (antiattractive, i.e., reduce the effect of aggregation pheromone) in these beetles (Figure 1).3,4

We recently published that the production of oxygenated monoterpenoids is related to tree stress and that it might be a signal for a suitable or nonsuitable host for bark beetles.3 Investigations by gas chromatographic electroantennographic detection (GC-EAD) of monoterpenones by us4 and Kalinova et al.5 revealed that both isopinocamphones and pinocamphones elicit antennal responses in I. typographus. There are relatively few syntheses of pinocamphone and isopinocamphone reported. In one report in Chinese, Wang et al. reacted α-pinene with borane to obtain diisopinocamphethylborane, which was oxidized to isopinocamphone by sodium perborate to afford isopinocamphone.6 The isopinocamphone was finally oxidized by H2O2 with vanadium phosphorus oxide as catalyst to yield isopinocamphone. Pitinova-Stekrova and co-workers utilized different titanosilicate catalysts to convert α-pinene to obtain campholenic aldehyde.7 Some of these catalysts produced pinocamphone as side-product. In another report, thermolysis of α-pinene epoxide in supercritical anhydrous isopropanol afforded up to ~25% pinocamphone, but in an inseparable mixture of oxygenated monoterpenoids.8 These syntheses do not yield pure stereoisomers or are tedious and have low yields. For short and convenient synthesis without heating, we developed a simple method using pure isopinocamphone stereoisomers that are commercially available.

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Pinocarvone is reported as a pheromone for the southern pine beetle (Dendroctonus frontalis) and has been found in the hindguts of male white pine cone beetles, Conophthorus coniperda. In the GC-EAD analysis of I. typographus, pinocarvone gave strong antennal responses, indicating biological activity. Pinocarvone has been isolated from Eucalyptus oil and has been produced by oxidation of \( \beta \)-pinene with SeO\(_2\). In this reaction, myrtenal is formed by a rearrangement, and this byproduct was either overlooked or lost during spinning band distillation.

In previous reports, we reported that one of the two trans-4-thujanol stereoisomers showed strong GC-EAD activity for bark beetles and that this (+)-trans-thujanol is a field-active semiochemical for the bark beetle, I. typographus. Blazytė-Cereskienė et al. reported that young spruce trees release more 4-thujanol than older trees and that 4-thujanol plays an important role in both host defense and tree choice by bark beetles. Thus, 4-thujanol is seemingly an indicator of healthy strong trees which should be avoided and could be of interest in forest protection. Several publications on the synthesis of 4-thujanol have been published, including the biotransformation of \( \alpha \)-pinene to \( cis \)-4-thujanol using the microorganism Fusarium saloni, Baeckström’s synthesis of \( trans \)-4-thujanol from \( cis \)-3-thujol, Galpin’s synthesis of the \( trans \) isomers from methyl vinyl ketone, Cheng’s synthesis of \( cis \)-4-thujanol, and Fanta’s synthesis of \( trans \)-thujanol. However, all these synthetic procedures involve many steps and/or expensive starting materials, and there are no effective synthetic routes for all possible stereoisomers. In order to develop a short synthesis of all stereoisomers of \( trans \)-4-thujanol for the investigation of GC-EAD activity, herein these were synthesized from commercial \( \alpha \)-pinene. The absolute configuration of each stereoisomer was unambiguously assigned by the deduction from the original \( \alpha \)-pinene in combination with NMR spectroscopy and chiral-phase GC-MS analysis.

These compounds are obviously important in bark beetle ecology, and some of them act as indicators of tree health; they are also of interest for managing bark beetle populations. It is well known that the stereochemistry of pheromones and other semiochemicals is often extremely important. Thus, there is a need to develop analytical procedures to be able to use enantioselective gas chromatography to differentiate between the stereoisomers of these semiochemicals in a biological sample, as well as their facile synthesis. We herein report the syntheses, specific rotations, and elution orders on a chiral GC phase (\( \beta \)-cyclodextrin) of pinocamphones, isopinocamphones, pinocarvones, and 4-thujanols (sabine hydrate).

**RESULTS AND DISCUSSION**

**Synthesis of Isopinocamphones and Pinocamphones.** Scheme 1 summarizes the syntheses of the four stereoisomers of pinocamphone. The pure enantiomers of isopinocamphone (1 and 2) were separately oxidized with pyridinium dichromate (PDC) to obtain both enantiomers of isopinocamphone in >98% optical purity. The oxidation was improved by adding silica gel to the reaction mixture, which prevents the formation of lumps and gel and in turn leads to higher yields and easier filtration at workup.

To produce pinocamphones, NaOEt was used to epimerize C-2 of isopinocamphone. The thermodynamic equilibrium seems to be 4:1 in favor of pinocamphone, and thus, 20% of isopinocamphone had to be removed by chromatography to obtain pure pinocamphone (Scheme 1).

### Table 1. Enantioselective GC-FID and Specific Rotations of Isopinocamphones, Isopinocamphones, and Pinocamphones

| s. no | compound | \( \tau_R \) | specific rotation | class |
|-------|----------|--------------|-------------------|-------|
| 1     | (1S,2S,3S,5R)-(+)-isopinocamphone | 11.69 | +13.4 (c 1.0, DCM) | isopinocamphones |
| 2     | (1R,2R,3S,5S)-(-)-isopinocamphone | 11.06 | +12.2 (c 1.0, EtOH) | pinocamphones |
| 3     | (1S,2S,5R)-(-)-pinocamphone | 11.37 | +20.2 (c 1.0, EtOH) | (PC) |
| 4     | (1R,2S,5S)-(+)-pinocamphone | 11.53 | +22.7 (c 1.0, EtOH) | (PC) |

**GC Elution Order of Isopinocamphones and Pinocamphones.** In the analysis by GC equipped with HP-5MS or DB-5MS columns, the pinocamphones eluted before the isopinocamphones. In the GC analysis using a chiral-phase column (Cyclosil B), \((-)\)-pinocamphone (5) eluted before \((+)-\)pinocamphone (6) and \((+)-\)isopinocamphone (4) before \((-)\)isopinocamphone (3) (Table 1 and Figure 2).

**Synthesis of Pinocarvone Stereoisomers.** There are only a few syntheses of pinocarvone published, and usually

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**Figure 2.** Mix of pinocamphones and isopinocamphones separated by enantioselective GC. The temperature program was isothermal 110 °C on a Cyclosil B column.
pinocarvone has been synthesized by oxidation of α-pinene or β-pinene (Scheme 2). One example is the Crich synthesis

**Scheme 2. Synthesis of Pinocarvones**

of pinocarvone from β-pinene (7) using perfluorooctyl selenic acid, where focus was on the preparation of the catalyst, and the yield was ca. 40%.27 However, a serious drawback with 3). The diol was cleaved using periodate to yield sabina ketone

1.0, EtOAc); (−)-pinocarvone (7) using SeO₂/DCM (Scheme 2). The mixture of pinocarvone and myrtetal was subsequently oxidized with H₂O₂/NaH₂PO₄ and NaClO₂ to remove the myrtenal in the form of myrtenic acid by silica gel chromatography (see GC-chromatogram, Figure 3). On the enantioselective GC-column

![Figure 3](https://dx.doi.org/10.1021/acs.jnatprod.0c00669) 3. Gas chromatograms before and after oxidation of myrtetal by NaClO₂/H₂O₂.

Figure 3. Gas chromatograms before and after oxidation of myrtenal by NaClO₂/H₂O₂.

phase (−)-pinocarvone (9) elutes before (+)-pinocarvone (8) (Figure 4). The ¹H NMR spectroscopic data of pinocarvone have been published,²⁸ but here we also provide ¹³C NMR spectroscopic data.

**Specific Rotation of Pinocarvone Stereoisomers.** The sign of specific rotation changed when pinocarvone to pinocarvone, i.e., (−)-β-pinene yielded (+)-pinocarvone (8) and (+)-β-pinene yielded (−)-pinocarvone (9). The specific rotations were as follows: (+)-pinocarvone (8) [α]₂⁰D +30.8 (c 1.0, EtOAc); (−)-pinocarvone (9) [α]₂⁰D −29.6 (c 2.0, EtOAc).

**Synthesis of 4-Thujanol (Sabinene Hydrate) Stereoisomers.** (−)-Sabinene (10) (86% ee) was subjected to a mild permanganate oxidation yielding sabinenediol (11) (Scheme 3). The diol was cleaved using periodate to yield sabina ketone (12). Chirality of the sabina ketone was confirmed by use of specific rotation and a GC column (β-cyclodextrin phase).⁵⁹ The ketone was reacted with MeLi. The methyl group attacked stereoselectively from the sterically less hindered side of the carbonyl, resulting in a 10:1 excess of cis-forms (13 plus enantiomer 16) over the corresponding trans-forms of thujanol (14 plus enantiomer 15) (i.e., cis-4-thujanol is the major diasteromer formed). As the ratio of stereoisomers in the sabinene (10) was (−)-93: (+)-7, it was easy to differentiate the (+)-(1R,4S)-4-thujanol (15), (−)-(1S,4R)-4-thujanol (14), (+)-(1R,4R)-4-thujanol (16), and (−)-(1S,4S)-4-thujanol (13), in a ratio of 1:9:4:86, by use of the enantioselective GC column.

The NMR spectrum and specific rotation proved that the isomer purchased from Sigma-Aldrich was the (+)-trans-isomer, and the major product could be assigned as (−)-cis-4-thujanol (13), based on the retention of ring configuration in the synthesis sequence, as well as regioselective considerations and reported NMR spectroscopic data.⁵⁰,⁵¹

**GC Elution Order of 4-Thujanol Stereoisomers.** On the HP-3MS GC column, the trans diastereomers eluted first. On the β-cyclodextrin column the first peak of four synthetic isomers coeluted with the commercial (+)-trans-4-thujanol stereoisomer (15) purchased from Sigma-Aldrich, and the last peak coeluted with the isolated (−)-cis-thujanol (13). The elution order of all isomers was (+)-trans, (−)-trans, (+)-cis, (−)-cis (Figure 5). The elution order is in accordance with those reported by Larkov et al.⁵² and Marriott et al.²³

**Specific Rotation of Sabina Ketone and 4-Thujanol Stereoisomers.** It should be noted that (−)-sabinene (10) yields (+)-sabina ketone (12), which is subsequently transformed to thujanols with (−)-cis-thujanol (13) as the major isomer (Scheme 3). The commercial sabinene (apparently from a natural source) has a specific rotation of −73 (c 1.0, EtOH) and −81 (c 1.0, DCM). Moreover, the chemical purity of the commercial sabinene (10) was only 75%, with 25% β-pinene as an impurity and with an ee of 86%. Sabina ketone (12) was obtained in 86% ee and with specific rotations of [α]₂⁰D +24 (c 1.0, EtOH) and [α]₂⁰D +33.5 (c 1.0, EtOAc) after removal of byproducts (pinene ketones) by chromatography. The (−)-cis-thujanol isomer (13) produced in the last step had, after chromatography, an optical purity of 91% ee and a specific rotation of −40 (c 0.5, DCM). The commercial (+)-trans-4-thujanol (15) (Sigma-Aldrich) had a specific rotation of +29.8 (c 0.5, DCM).

**EXPERIMENTAL SECTION**

**General Experimental Procedures.** Optical rotations were recorded in EtOH, EtOAc, and DCM on a 2019 model Rudolph automatic polarimeter (APIII) manufactured by Rudolph Research Analytical (Hackettstown, NJ, USA). NMR spectra were recorded in CDCl₃ on Bruker 400 and Varian 500 MHz spectrometers. The GC-MS instrument was an Agilent 6890 GC and 5973 mass detector and an Hewlett-Packard with a FID detector (Palo Alto, CA, USA). Helium was used as carrier gas. Two types of columns, a nonpolar column (HP-5MS, film thickness = 0.25 μm; Agilent Technologies 19091S-433) and a chiral-phase capillary column (Chirobiotic-B, 30 m × 0.25 μm, i.d. 0.25 mm, J&W Scientific, via Scantech Nordic AB, Jonsered, Sweden), were used. Mass spectra were obtained by electron impact ionization (70 eV). The general gas chromatography temperature
The program for both columns was as follows: initial temperature 50 °C (hold for 2 min), raised to 200 °C with 10 °C/min (hold for 15 min) (splitless). When a different temperature program was used for resolution of enantiomers (on the Cyclosil-B column), the temperature program is described in the figure legends of the GC chromatograms.

The synthesized compounds were purified on silica gel column chromatography using 230−400 mesh ultra pure silica.

### Synthesis of the Four Stereoisomers of Pinocamphone (Scheme 1)

#### Scheme 3. Synthesis of 4-Thujanol Stereoisomers

Figure 5. Chromatograms of chiral-phase GC of 4-thujanol isomers in the mixture and chromatograms of isolated (+)-trans- and (−)-cis-4-thujanol isomers. The temperature program: Initial oven temp 40 °C (hold for 3 min) and increased to 150 °C at 3 °C/min and finally increased to 250 °C at 15 °C/min. It was kept for 10 min at the final temperature of 250 °C.

**Figure 4. GC elution order of pinocarvone stereoisomers. Separated on GC equipped with a Cyclosil B column. In the chiral analysis (−)-pinocarvone eluted earlier than (+)-pinocarvone.**

(A) (−)-Pinocarvone (9), (B) (+)-pinocarvone (8), and (C) mix of pinocarvone enantiomers. The temperature program: Initial oven temperature was 40 °C (held for 5 min) increased to 150 °C at 3 °C/min and finally increased to 220 °C at 10 °C/min (held for 5 min at the final temperature).
(+)-Isopinocamphone (4). In analogy with the procedure used for the (+)-antipode, (1R,2R,3S,5S)-2,6,6-trimethylbicyclo[3.1.1]heptan-3-one [(1R,2R,3S,5S)-isopinocamphone] (2) (12.21 g, 79.3 mmol) was oxidized with PDC (60 g, 160 mmol) to yield 75% (90.3 ± 59.4 mmol) (1R,2R,5S)-2,6,6-trimethylbicyclo[3.1.1]heptan-3-one ((+)-isopinocamphone) (4) after column chromatography. \([\alpha]_D^{25} = +11.2 (c \ 1.0, 

EtOH), 98\% ee. The NMR data were identical to the data for the (-)-isomer. GC-MS m/z. See other enantiomer.

(-)-Pinocamphone (5). (1S,2R,3S)-2,6,6-Trimethylbicyclo[3.1.1]-heptan-3-one. To a solution of (-)-isopinocamphone (3) (3.04 g, 20.0 mmol) dissolved in EtOH (10 mL) was added NaOEt in EtOH (21% w/w, 11 mL, 34 mmol), and the mixture stirred for 24 h at RT. Water (50 mL) and EtO (50 mL) were added after 24 h, when the 4:1 equilibrium ratio between (-)-pinocamphone (5) and (-)-iso pinocamphone (4) had been established. The aqueous phase was washed with EtOAc (2 × 50 mL), and the combined ether phases were washed with 15 mL of water to remove EtOH. After drying over MgSO₄, filtration, and evaporation, 15 mL of toluene was added before subsequent evaporation. The removal of water/EtOH by azetropic distillation with toluene was repeated twice. The clear amber-colored residue was subjected to MPLC, yielding 2.9 g (19.1 mmol) of (-)-pinocamphone (5) (in ppm) 5.93 (1H, s), 5.98 (1H, s), 2.74 (1H, t, J = 5.9 Hz), 2.69−2.64 (1H, m), 2.63 (1H, d, J = 2.6 Hz), 2.50 (1H, dd, J = 19.3 and 2.7 Hz), 2.18 (1H, dd, J = 5.8 and 3.0 Hz), 1.33 (3H, s, H-8), 1.27 (1H, d, J = 10.5 Hz), 0.78 (3H, s, H-9). \[^{13}C\] NMR (CDCl₃, 125 MHz) δ (in ppm) 214.9, 39.5, 33.7, 33.2, 32.6, 23.6, 19.5, 19.3, 19.1; GC-MS m/z 108 (100%), 81, 53, 107, 79, 41, 77, 150 (M⁺), 69, 91, 122 (intensity of decreasing order). NMR spectrum data are in accordance with reported data [14].

Synthesis of 4-Thujanol Isomers (Scheme 3). Synthesis of Sabinedanol (11). To a solution of (-)-sabinedanol (10) 86% ee (1 g, 7.4 mmol) in THF (3 mL) was added KMnO (2.3 g, 14.6 mmol) in water (4 mL) over a period of 2 h. The mixture was stirred for another hour before the precipitate was filtered off. The filtrate was extracted with EtOAc (2 × 50 mL), and the combined organic layers were washed with brine and dried over Na₂SO₄. The solution was concentrated by rotary evaporation to yield 875 mg (5.1 mmol) of crude sabinediol (11) (70% yield). \[^{1}H\] NMR (CDCl₃, 500 MHz) δ (in ppm) 3.55 (2H, ap t, J = 11.6 Hz), 2.45 (1H, br s, −OH), 2.26 (1H, d, J = 17.8 Hz), 1.95−1.89 (1H, m), 0.78 (3H, H, H-9). \[^{13}C\] NMR (CDCl₃, 125 MHz) δ (in ppm) 202.2, 149.1, 117.6, 48.3, 42.6, 40.41, 38.6, 32.5, 26.1, 22, 16; GC-MS m/z 90.6 m/z 81 (100%), 108, 53, 107, 135, 79, 41, 77, 150 (M⁺), 69, 91, 122 (intensity of decreasing order).

Synthesis of α-4-Thujanol Isomers (Scheme 3). Synthesis of α-Sabinedanol (11a). To a solution of α-sabinedanol (10) 86% ee (1 g, 7.4 mmol) in THF (3 mL) was added KMnO (2.3 g, 14.6 mmol) in water (4 mL) over a period of 2 h. The mixture was stirred for another hour before the precipitate was filtered off. The filtrate was extracted with EtOAc (2 × 50 mL), and the combined organic layers were washed with brine and dried over Na₂SO₄. The solution was concentrated by rotary evaporation to yield 875 mg (5.1 mmol) of crude sabinediol (11) (70% yield). \[^{1}H\] NMR (CDCl₃, 500 MHz) δ (in ppm) 3.55 (2H, ap t, J = 11.6 Hz), 2.45 (1H, br s, −OH), 2.26 (1H, d, J = 17.8 Hz), 1.95−1.89 (1H, m), 0.78 (3H, H, H-9). \[^{13}C\] NMR (CDCl₃, 125 MHz) δ (in ppm) 202.2, 149.1, 117.6, 48.3, 42.6, 40.41, 38.6, 32.5, 26.1, 22, 16; GC-MS m/z 90.6 m/z 81 (100%), 108, 53, 107, 135, 79, 41, 77, 150 (M⁺), 69, 91, 122 (intensity of decreasing order).

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0.62 (1H, dd, J = 5.2 and 3.6 Hz), 0.28 (1H, dd, J = 8.0 and 5.2 Hz);
13C NMR (CDCl₃, 100 MHz) δ (in ppm) 79.2, 36.0, 33.5, 33.1, 32.5,
27.9, 25.4, 19.5, 19.4, 11.1; GC-MS m/z 71 (100%), 111, 93, 81, 43,
139, 121, 79, 69, 55, 136, 154 (M⁺) (decreasing order of intensity).
(+)-trans-(1S,4S)-4-Thujaanol (15) (from Sigma-Aldrich, Schnell-
dorf, Germany).

Notes

Authors

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GC chromatograms, 13C NMR and 1H NMR spectra (PDF)

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Notes
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