Rapidly Progressive Renal Dysfunction in Two Elderly Patients with Renal Enlargement and Medullary Cystic Kidney Disease-like Acute Tubulointerstitial Injury

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Abstract

Medullary cystic kidney disease (MCKD) is a hereditary disease associated with bilateral medullary polycysts and interstitial fibrosis. MCKD is typically associated with slowly progressive renal dysfunction. We herein report two rare elderly cases with enlarged kidneys and rapidly progressive renal dysfunction without myeloperoxidase anti-neutrophil cytoplasmic antibody (MPO-ANCA), PR3-ANCA, or anti-glomerular basement membrane (GBM) antibodies. Renal biopsies revealed extensive tubular dilatation and atrophy with interstitial fibrosis consistent with MCKD. Both patients began hemodialysis therapy a few months later. Our cases suggest a MCKD subgroup among elderly patients with an undefined genetic background, rapidly progressive renal dysfunction, and enlarged kidneys.

Key words: kidney enlargement, rapidly progressive renal failure, medullary cystic kidney disease, elderly patients

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Introduction

Nephronophthisis (NPH) and medullary cystic kidney disease (MCKD) are renal diseases associated with bilateral medullary polycysts and interstitial fibrosis. These two diseases are thought to represent the same disease process according to similar clinical and pathological features, excluding the age of onset and associated genetic mutations. The concept of juvenile nephronophthisis-medullary cystic disease complex (NPH-MCKD) has been proposed (1).

With the progression of tubular damage, tubular functions, such as sodium reabsorption and urine concentration, become impaired. Inflammation and fibrosis occurs in the tubulointerstitial area of the kidneys, ultimately leading to end-stage kidney disease (ESKD). The progression to ESKD is typically gradual, similar to that observed in polycystic kidney disease. NPH is an autosomal recessive (AR) disease that develops in adulthood and slowly progresses to ESKD, characterized by multiple cysts, measuring 1-20 mm, in the corticomedullary boundary.

The clinical features of MCKD are nonspecific and include hypertension, polyuria, and sodium wasting due to an impairment in urinary concentration (2). The pathological features include expanded urinary ducts in the corticomedullary boundary areas, diffuse tubulointerstitial nephritis with tubular atrophy, interstitial fibrosis, and an inflammatory cell infiltrate (3). The kidneys are typically atrophic and decreased in size (4, 5).

We herein report two cases of MCKD in elderly patients with enlarged kidneys and a rapid progression to ESKD. Both patients had no apparent genetic history of kidney disease, and their clinical courses were not typical for traditional hereditary MCKD.
Aminotransferase (AST), 38 IU/L; alanine aminotransferase 142 mg/dL; hemoglobin A1c (HbA1c), 4.8%; aspartate aminotransferase (ALT), 24 IU/L; alkaline phosphatase (ALP), 787 IU/L; gamma glutamyl transpeptidase (γGT), 239 IU/L; IgG, 828 mg/dL; IgM, 700 mg/dL; and IgA, 148 mg/dL. Cryoglobulin was negative and an immunoglobulin (Ig) M (kappa) spike was identified on immunoelectrophoresis. The third complement component (C3) level was 59 mg/dL (normal 50-130 mg/dL); the fourth component (C4) level was 10.5 mg/dL (normal 10-50 mg/dL); hemolytic complement activity via the classical pathway (CH50) was 13 U/mL (normal 25.0-48.0 U/mL); and antinuclear antibody (ANA) was negative. Myeloperoxidase antibodies (MPO), proteinase 3 antibodies (PR3), antineutrophil cytoplasmic antibodies (ANCA), and anti-glomerular basement membrane (anti-GBM) antibodies were not detected. A urinalysis showed 1.17 g protein/g creatinine and 30-49 RBC/high-power field (HPF). The urinary β-2 microglobulin (β-2MG) level was 25,150 ng/mL. Bence Jones protein was negative in the urine. A bone marrow aspirate clot showed hypercellular bone marrow with 3% plasma cells.

At the time of hospitalization, a percutaneous kidney biopsy was not performed because of the patient’s massive ascites. A percutaneous kidney biopsy was performed 1 month later after several paracentesis procedures, allowing the patient to lie in a prone position.

Histological findings revealed tubular dilatation, primarily involving the distal tubules, with extensive fibrosis in the interstitium, consistent with nephropathies-medullary cystic disease complex (NPH-MCKD). There were no remarkable glomerular changes except for global sclerosis consistent with the patient’s age (Fig. 2). Immune deposits of IgG, IgA, IgM, C1q, C3, and fibrinogen were absent, despite the IgM kappa M protein detected in the serum. Electron microscopy revealed irregularities, thinning and disintegration of the tubular basement membrane (TBM). However, it was difficult to explain the patient’s massive ascites and rapidly progressive renal dysfunction according to the histological findings.

Considering her advanced age (84 years of age) and symptomatic relief after paracentesis, no further investigation was done and she was discharged with a serum Cr level of 5.1 mg/dL 2 months after admission. She began hemodialysis therapy 1 month after discharge.

Case 2

A 74-year-old woman was started on antihypertensive therapy 1 year prior to admission when she was noted to have a mild renal impairment with a serum Cr level of 1.1 mg/dL. She was status post-appendectomy at 20 years of age, had a history of ulcerative colitis with rectal cancer at 61 years of age, and a cataract at 68 years of age. She had no family history of renal disease or dialysis. Although her blood pressure was well controlled, her serum Cr level rapidly increased to 1.5 mg/dL at 3 months after the start of antihypertensive therapy and 2.4 mg/dL at 5 months after the start of therapy. She was subsequently referred to our hospital for further evaluation and treatment.

On physical examination, her temperature was 36.5°C, and her blood pressure was 150/79 mmHg. Her conjunctivae were pale and she had lower extremity edema. The following laboratory results were obtained: WBC count, 6,800 cells/mm³, platelet count (plt), 15.8×10⁴ platelets/mm³; white blood cell (WBC) count, 6,800/mm³; hemoglobin (Hb), 8.2 g/dL; hematocrit (Ht), 26.0%; platelet count (plt), 15.8×10⁴ platelets/mm³; total protein (TP), 5.6 g/dL; albumin (Alb), 2.8 g/dL; blood urea nitrogen (BUN), 45 mg/dL; serum Cr, 3.7 mg/dL; sodium (Na), 140 mEq/L; potassium (K), 3.2 mEq/L; chloride (Cl), 104 mEq/L; C-reactive protein (CRP), 0.5 mg/dL; plasma glucose, 142 mg/dL; hemoglobin A1c (HbA1c), 4.8%; aspartate aminotransferase (AST), 38 IU/L; alanine aminotransferase (ALT), 18 IU/L; alkaline phosphatase (ALP), 787 IU/L; gamma glutamyl transpeptidase (γGT), 239 IU/L; IgG, 828 mg/dL; IgM, 700 mg/dL; and IgA, 148 mg/dL. Cryoglobulin was negative and an immunoglobulin (Ig) M (kappa) spike was identified on immunoelectrophoresis. The third complement component (C3) level was 59 mg/dL (normal 50-130 mg/dL); the fourth component (C4) level was 10.5 mg/dL (normal 10-50 mg/dL); hemolytic complement activity via the classical pathway (CH50) was 13 U/mL (normal 25.0-48.0 U/mL); and antinuclear antibody (ANA) was negative. Myeloperoxidase antibodies (MPO), proteinase 3 antibodies (PR3), antineutrophil cytoplasmic antibodies (ANCA), and anti-glomerular basement membrane (anti-GBM) antibodies were not detected. A urinalysis showed 1.17 g protein/g creatinine and 30-49 RBC/high-power field (HPF). The urinary β-2 microglobulin (β-2MG) level was 25,150 ng/mL. Bence Jones protein was negative in the urine. A bone marrow aspirate clot showed hypercellular bone marrow with 3% plasma cells.

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Case 1

An 84-year-old woman had a 3-year history of hypertension, a femoral neck fracture 1 year previously, and a left radial fracture 1 month previously. She had no family history of renal disease or dialysis. She was noted to have renal dysfunction of unknown etiology 1 year prior to admission. She visited a regional clinic for edema and was observed to have renal dysfunction [serum creatinine (Cr), 2.7 mg/dL] and anemia 2 months prior to admission. Since that time, she had developed marked edema and progressive renal dysfunction (serum Cr, 3.7 mg/dL) and was referred to our hospital for further evaluation and treatment. On physical examination, her temperature was 37.1°C, and her blood pressure was 168/63 mmHg. She showed marked abdominal distention due to ascites and also had lower extremity edema without dyspnea. Chest radiography revealed cardiac enlargement due to ascites and also had lower extremity edema. The following laboratory results were obtained: white blood cell (WBC) count, 6,800/mm³; red blood cells (RBC) count, 250×10⁶ RBC/mm³; hemoglobin (Hb), 8.2 g/dL; hematocrit (Ht), 26.0%; platelet count (plt), 15.8×10⁴ platelets/mm³; total protein (TP), 5.6 g/dL; albumin (Alb), 2.8 g/dL; blood urea nitrogen (BUN), 45 mg/dL; serum Cr, 3.7 mg/dL; sodium (Na), 140 mEq/L; potassium (K), 3.2 mEq/L; chloride (Cl), 104 mEq/L; C-reactive protein (CRP), 0.5 mg/dL; plasma glucose, 142 mg/dL; hemoglobin A1c (HbA1c), 4.8%; aspartate aminotransferase (AST), 38 IU/L; alanine aminotransferase (ALT), 18 IU/L; alkaline phosphatase (ALP), 787 IU/L; gamma glutamyl transpeptidase (γGT), 239 IU/L; IgG, 828 mg/dL; IgM, 700 mg/dL; and IgA, 148 mg/dL. Cryoglobulin was negative and an immunoglobulin (Ig) M (kappa) spike was identified on immunoelectrophoresis. The third complement component (C3) level was 59 mg/dL (normal 50-130 mg/dL); the fourth component (C4) level was 10.5 mg/dL (normal 10-50 mg/dL); hemolytic complement activity via the classical pathway (CH50) was 13 U/mL (normal 25.0-48.0 U/mL); and antinuclear antibody (ANA) was negative. Myeloperoxidase antibodies (MPO), proteinase 3 antibodies (PR3), antineutrophil cytoplasmic antibodies (ANCA), and anti-glomerular basement membrane (anti-GBM) antibodies were not detected. A urinalysis showed 1.17 g protein/g creatinine and 30-49 RBC/high-power field (HPF). The urinary β-2 microglobulin (β-2MG) level was 25,150 ng/mL. Bence Jones protein was negative in the urine. A bone marrow aspirate clot showed hypercellular bone marrow with 3% plasma cells.

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Case Reports

Figure 1. Computed tomography of the abdomen in case 1. Computed tomography of the abdomen shows massive ascites and enlarged kidneys with no evidence of cysts.
WBC/mm$^3$; RBC count, 248×$10^6$ RBC/mm$^3$; Hb, 7.3 g/dL; Ht, 21.7%; Plt, 18.2×$10^9$/mm$^3$; TP, 7.1 g/dL; Alb, 4.1 g/dL; BUN, 30 mg/dL; Cr, 3.6 mg/dL; Na, 138 mEq/L; K, 5.3 mEq/L; Cl, 107 mEq/L; CRP, 0.21 mg/dL; plasma glucose, 107 mg/dL; HbA1c, 5.1%; IgG, 1,326 mg/dL; IgM, 76 mg/dL; IgA, 239 mg/dL; C3, 91 mg/dL; C4, 24.7 mg/dL; and CH50, 13 U/mL. Cryoglobulin was negative. ANA was negative, and MPO, PR3, ANCA, and anti-GBM were not detected. A urinalysis showed 1.23 g protein/g creatinine and 6 RBC/HPF. Urinary β-2MG was 27,708 ng/mL.

Abdominal CT showed bilateral kidney enlargement that was not seen on an earlier CT exam 1 year previously. Serial CT images are shown in Fig. 3. Magnetic resonance images (MRI) are shown in Fig. 4. MRI, including coronal magnetic resonance images, did not reveal multiple microcysts in the enlarged kidneys, however, a careful observation revealed multiple cysts along the corticomedullary boundary.

A percutaneous renal biopsy was performed on day 2. Of seven glomeruli, two were globally sclerotic, one was segmentally sclerotic, and two showed glomerular cysts. The other glomeruli were essentially normal. The distal tubules were also dilated, consistent with MCKD, and there was mild interstitial fibrosis and inflammatory cell infiltration (Fig. 5). Electron microscopy revealed irregular layering, spheroidizing, thinning, and partial rupture of the TBM. The patient was treated with corticosteroids [prednisolone (PSL) 30 mg/day] in view of the significantly elevated urinary β-2MG level and interstitial cell infiltration. Immediately after PSL administration, a slight reduction in the serum Cr level was noted and her anemia improved. However, the serum Cr level gradually increased and the Hb level gradually decreased 2 months later, when PSL was tapered. Three months after the start of PSL therapy, the patient developed gastrointestinal bleeding and was found to have a serum Cr level of 7 mg/dL. The gastrointestinal bleed may have been an adverse effect of PSL therapy. However, no apparent bleeding source was detected by gastroscopy, colonoscopy, or enteroscopy. Anemia was corrected with blood transfu-
Figure 4. Magnetic resonance imaging (MRI) of the abdomen: (a) axial section of the abdomen (T1-weighted image) and (b) coronal section of the abdomen (T2-weighted image) in case 2. No apparent microcysts are seen in the enlarged kidneys, including the coronal images, however, a closer observation shows multiple cysts along the corticomedullary boundary in the left kidney in the coronal section (arrows) (b).

Figure 5. Renal biopsy findings in case 2. (a) periodic acid-methenamine-silver (PAM) stain; original magnification 10×, (b) PAS stain original magnification 200×. Light microscopy images show tubular dilatation and atrophy. Cystic tubules show irregularities and thinning of the tubular epithelial cells. Of seven glomeruli, two are globally sclerotic, one is segmentally sclerotic, and two show glomerular cysts. The other glomeruli are intact.

sions, however, the patient’s renal function did not improve and she began hemodialysis therapy 5 months after referral.

**Immunohistochemical findings**

We also performed immunohistochemical staining in both cases for CD10, cytokeratin 7 (CK7), epithelial membrane antigen (EMA), E-cadherin, and cytokeratin (34 beta E12) to identify the level of tubular dilatation (Fig. 6). In case 1, the dilated tubules were negative for CD10, a marker of proximal tubules (6). In contrast, immunostaining results for CK7, EMA, and E-cadherin, markers of the distal tubules and collecting ducts, were positive. Immunostaining results for CK (34 beta E12), a marker of collecting ducts, were also positive (7), indicating that the cysts predominantly arose from collecting ducts in case 1. In case 2, the dilated tubules were positive for CK7 and EMA and weakly positive for E-cadherin and CK (34 beta E12).

**Discussion**

NPH-MCKD is a hereditary disease of the renal tubules, which are important for the reabsorption of sodium and the concentration of urine. Typically, interstitial inflammation and fibrosis occurs, slowly progressing to ESKD, similar to the progression seen in polycystic kidney disease. The genetic mutations associated with NPH-MCKD are shown in Table 1 (8-17).

Recently, a human genetic condition termed “ciliopathy”
has been proposed. Ciliopathy is thought to be caused by a
dysfunction of two functionally and physically associated
organelles, the centromere and the cilium. Autosomal domi-
nant polycystic kidney disease (ADPKD) and NPH-MCKD
are thought to be related to renal cilia dysfunction (ciliopa-
thy), and more than 30 different genetic mutations have
been localized to the cilia and/or centromeres (18, 19).

NPH is inherited in an AR manner. The disease onset oc-
curs in early childhood or adolescence and typically pro-
gresses to ESKD during adolescence. On the other hand,
MCKD is an AD disease. Its onset occurs during adulthood
and slowly progresses to ESKD with the formation of mul-
tiple cysts, measuring 1-20 mm, in the corticomedullary
boundary. The kidneys usually decrease in size and become
atrophic. There is currently no effective treatment to prevent
progression; sodium supplementation and erythropoietin are
used for symptomatic relief.

Two types of MCKD are recognized (MCKD1 and
MCKD2) according to the localizing chromosome and the
onset of ESKD. MCKD1 progresses more slowly than
MCKD2 and has a later onset of ESKD (average age 62
years old; range, 50-70 years). The average age of onset of
ESKD in MCKD2 is 32 years (range, 16-54 years).

Genetic mutations are known to affect mucoprotein
mucin-1 (MUC1) in MCKD1 and uromodulin (UMOD) in
MCKD2 (10). The initial symptoms are polydipsia and poly-
uria due to decreased urine concentration and sodium loss.
The urinalysis is normal early in the disease. Generally,
there is a slow progression to ESKD, similar to the progres-
sion seen in polycystic kidney disease. The typical CT, MRI,
and ultrasound images in MCKD show atrophic kidneys
with multiple cysts in the medulla, particularly at the corti-
comedullary junction (4, 5). The differential diagnosis of
kidney enlargement includes infiltrative diseases (hema-
topoietic malignancy, sarcoidosis, and amyloidosis), autosom-
al dominant polycystic kidney disease, early stage diabetic
nephropathy, acute interstitial nephritis, and pyelonephri-
tis (20). In the present cases, an immunohistological investi-

Figure 6. Immunohistochemical findings in both cases. Immunohistochemical staining for CD10, cyto-
kertatin 7 (CK7), epithelial membrane antigen (EMA), E-cadherin, and cytokeratin (34 beta E12) in
the interstitium of both cases. Original magnification 10×. The tubular basement membrane (TBM)
of dilated tubules in both cases is negative for CD10. In case 1, the TBM of dilated tubules is positive
for CK7, EMA, E-cadherin, and CK (34 beta E12). In case 2, the TBM of dilated tubules is positive
for CK7 and EMA and weakly positive for E-cadherin and CK (34 beta E12).

Table 1. Gene Mutations Associated with NPH-MCKD.

| Reference | Inheritance | Chromosome | Gene | Protein | ESKD (yrs) |
|-----------|-------------|------------|------|---------|------------|
| 8,9       | AR          | 2q13       | NPHP1 | Nephrocystin-1 | 13         |
| 10        | AR          | 9q22-q31   | NPHP2 | Inversin | <1         |
| 11        | AR          | 3q21.1     | NPHP3 | Nephrocystin-3 | 19         |
| 12        | AR          | 1p36.22    | NPHP4 | Nephroretinin | 21         |
| 13        | AR          | 3q21.1     | IQCB1,NPHP5 | Nephrocystin-5 | 13         |
| 14        | AR          | 12q21.33   | CEP290,NPHP5 | Nephrocystin-6 |           |
| 15        | AR          | 16p13.3    | GLIS2 | Zinc finger protein GLIS2 |           |
| 16        | AD          | 1q21       | MUC1 | Mucin-1 | 62         |
| 17        | AD          | 16p12.3    | UMOD | Uromodulin | 32         |

AR: Autosomal Recessive, AD: Autosomal Dominant

Table 1. Gene Mutations Associated with NPH-MCKD.
gation revealed numerous microcysts within the dilated distal tubules and collecting ducts, compatible with MCKD. We postulate that these renal microcysts rapidly developed and replaced the renal parenchyma with tubulointerstitial inflammation, a finding rarely seen in MCKD. The cyst formation and subsequent inflammation played a pivotal role in the pathogenesis of the kidney enlargement and dysfunction. We have found similar case reports from Japan (21-23). These cases, including ours, are summarized in Table 2. All five of these cases showed renal enlargement and MCKD-like tubulointerstitial changes with a rapid progression to ESKD. However, there are no previous reports of patients with massive ascites and rapidly progressive renal failure in MCKD. Taneda et al. also reported a similar case in an elderly patient who was not clinically diagnosed, but had been diagnosed at autopsy with ARPKD according to the renal morphology. The case reported by Taneda et al. also showed renal enlargement and a rapid progression to ESKD in the absence of any apparent genetic mutation. The diagnosis of ARPKD was made according to the presence of liver involvement, including hepatic cysts, jaundice, and elevated liver enzyme levels, findings not typically observed in MCKD (24).

Because these other cases were reported mainly from Japan, it appears that these diseases are not due to regional or racial differences or a new disease type, unlike traditional hereditary MCKD. A greater accumulation of similar cases rapidly progressive renal dysfunction, renal enlargement, and a rapid progression to ESKD in the absence of any apparent genetic mutation. The diagnosis of ARPKD was made according to the presence of liver involvement, including hepatic cysts, jaundice, and elevated liver enzyme levels, findings not typically observed in MCKD (24).

Because these other cases were reported mainly from Japan, it appears that these diseases are not due to regional or racial differences or a new disease type, unlike traditional hereditary MCKD. A greater accumulation of similar cases with genetic and histopathological analyses is important to further our understanding of this disease entity.

We herein reported two unusual cases that presented with rapidly progressive renal dysfunction, renal enlargement, and acute interstitial injury. The pathological findings were consistent with MCKD, however, the patients had no family history of renal disease. Similar cases, lacking the genetic findings of traditional hereditary MCKD, have been reported in Japan. Further pathological and clinical investigations of future cases are required.

The authors state that they have no Conflict of Interest (COI).

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Table 2. Summary of Similar Cases Reported in Japan.

| Reference | Age | Sex | FH | Cr | Hb | CRP | UP | UOB | Uβ2(pg/L) | Outcome |
|-----------|-----|-----|----|----|----|-----|----|-----|----------|---------|
| Case 1    | 84  | F   | (-) | 3.9 | 8.4 | 0.5 | 1+ | 1+  | 25,150   | HD      |
| Case 2    | 74  | F   | (-) | 3.6 | 7.3 | <0.06 | 1+ | -   | 27,708   | HD      |
| 2         | 70  | M   | (-) | 2.3 | 8.3 | 0.37g/Cr | -  | 648 | HD       |
| 22        | 66  | F   | (-) | 5.87| 8.9 | 0.24g/Cr | 1+ | 26,123 | HD       |
| 23        | 63  | F   | (+) | 11.6| 6.2 | 0.06 | 2+ | 1+  | HD       |

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