Drug desensitization is the temporary induction of tolerance to a sensitized drug by administering slow increments of the drug, starting from a very small amount to a full therapeutic dose. It can be used as a therapeutic strategy for patients with drug hypersensitivity when no comparable alternatives are available. Desensitization has been recommended for immunoglobulin E (IgE)-mediated immediate hypersensitivity; however, its indications have recently been expanded to include non-IgE-mediated, non-immunological, or delayed T cell-mediated reactions. Currently, the mechanism of desensitization is not fully understood. However, the attenuation of various intracellular signals in target cells is an area of active research, such as high-affinity IgE receptor (FcεRI) internalization, anti-drug IgG4 blocking antibody, altered signaling pathways in mast cells and basophils, and reduced Ca²⁺ influx. Agents commonly requiring desensitization include antineoplastic agents, antibiotics, antituberculous agents, and aspirin/nonsteroidal antiinflammatory drugs. Various desensitization protocols (rapid or slow, multi-bag or one-bag, with different target doses) have been proposed for each drug. An appropriate protocol should be selected with the appropriate concentration, dosage, dosing interval, and route of administration. In addition, the protocol should be adjusted with consideration of the severity of the initial reaction, the characteristics of the drug itself, as well as the frequency, pattern, and degree of breakthrough reactions.

**Keywords:** Drug hypersensitivity; Desensitization, immunologic; Antineoplastic agents; Anti-bacterial agents; Anti-inflammatory agents, non-steroidal
reintroduction of culprit drugs to highly sensitized patients in need of first-line therapies. Desensitization is performed by administration of extremely low doses of the culprit drug, below the threshold for induction of a hypersensitivity reaction, followed by a gradual increase in the dose at fixed time intervals, to allow adaptation to a given amount or concentration; this leads to the development of temporary tolerance to the drug, eventually at its full therapeutic dose [2].

This review presents an overview of the concept of drug desensitization, an update on its mechanism, and how it is applied in clinical practice.

HISTORICAL BACKGROUND

The first instance of drug desensitization was reported in the literature in 1922 by Widal et al. [3] who described nonsteroidal antiinflammatory drug (NSAID)-exacerbated respiratory disease (NERD). Rapid drug desensitization (RDD) was first successfully accomplished in the 1940s during World War II, involving the readministration of penicillin to soldiers who had shown anaphylaxis in response to penicillin [4,5]. At present, desensitization is performed for DHR to various drugs, including antineoplastic agents, antibiotics, antituberculous agents, and aspirin/NSAIDs.

IMMUNOLOGICAL MECHANISM

The desensitization process is known to be antigen-specific, as the level of drug-specific immunoglobulin E (IgE) decreases but the levels of other allergen-specific IgE remain consistent throughout the treatment period [6]. However, the cellular and molecular mechanisms underlying drug desensitization are not yet fully understood. The growing body of clinical knowledge has led to several hypotheses, such as the internalization of high-affinity IgE receptor (FceRI), the generation of anti-drug IgG4 blocking antibodies, alteration of signaling pathways in mast cells and basophils, and reduction of Ca\(^{2+}\) influx [7-11].

Some studies showed that suboptimal doses of drug antigens led to the internalization of antigen/IgE/FceRI complex, thus preventing the activation of mast cells or basophils when the therapeutic dose is administered [12,13]. However, other studies showed that suboptimal doses of drug antigens induce remodeling of the mast cell membrane, which prevents antigen/IgE/FceRI complex internalization and the initiation of hypersensitivity reactions [7,8].

The action of allergen-specific IgG4, which blocks the interaction between allergens and IgE, has been well characterized in allergen immunotherapy. IgG4 may play a role in desensitization, as the level of drug-specific IgG4 increases after desensitization [14,15]. However, IgG4 cannot fully explain the immediate development of immune tolerance and the sustained effect of desensitization [16].

Various proteins in the activation signaling pathways can be altered during desensitization. Syk protein, a tyrosine protein kinase that transduces the activation signal of mast cells and basophils, expression is reduced after desensitization, which inhibits the activation of FceRI by binding of IgE to the sensitized drug [17,18]. In addition, during the desensitization process, FcγRII with immunoreceptor tyrosine-based inhibition motif competitively inhibits FcεRI and dephosphorylates initial signaling molecules, including Syk, through Src homology 2 domain-containing inositol phosphatase 1 (SHIP1), thus blocking the downstream signal transduction to activate mast cells [9,19-21]. Furthermore, the decrease in intracellular Ca\(^{2+}\) ions and their impaired influx by the remodeling of actin filaments also inhibit the degranulation of mast cells [7,8,12,22].

In addition, the role of T cells has been increasingly emphasized. It has been suggested that drug desensitization may lead to attenuation of immunological memory as the degree of DHR and the incidence of breakthrough reactions (BTRs) decrease during the desensitization process [23]. Several cases of successful desensitization for T cell-mediated delayed DHR, such as fixed drug eruptions, have been reported in the literature [24]. The increases of CD4\(^+\)CD25\(^+\) or CD4\(^+\)CD25\(^+\)FOXP3\(^+\) T cells expressing regulatory cytokines such as interleukin 10 (IL-10) and IL-35 after desensitization suggest a role of regulatory T cells [23,25,26].

In summary, desensitization induces temporary tolerance to the culprit drug by attenuating various intracellular signals to prevent the activation of immunological responses in target cells, thus enabling the safe reintroduction of the drug [7].

APPLICATION IN CLINICAL PRACTICE

Overview

The efficacy of drug desensitization is best understood in
IgE-mediated immediate hypersensitivity. In addition, it is also effective for nonallergic immediate reactions, such as infusion-related reactions, which directly trigger mast cells to secrete their granules or cytokines [27].

Two categories of desensitization protocols are currently available: RDD and slow drug desensitization (SDD). RDD is recommended for immediate reactions, both allergic and nonallergic. The most widely used RDD protocol is doubling the dosage every 15 minutes until the therapeutic dose is achieved. SDD is recommended for type IV delayed hypersensitivity reactions with T cell involvement, and can be performed both orally and intravenously. There is as yet no consensus on SDD protocols, including the initial dose, dose increments between steps, and dosing interval. Further clinical experience and research are required to establish the role and efficacy of desensitization for delayed reactions [28,29].

Before performing drug desensitization, the diagnosis should be confirmed by thorough history taking and testing. Desensitization should be considered when no alternative options are available or the culprit drug has far superior efficacy, i.e., the benefits of the culprit drug outweigh the risk. It should be carefully applied in patients with significant risk factors for fatal BTRs, such as those with a history of severe hypersensitivity reactions, like anaphylaxis, Stevens-Johnson syndrome, and toxic epidermal necrolysis, those with cardiovascular or respiratory diseases, and those using beta-blockers. The appropriate protocol should be selected based on the concentration, dosage, dosing interval, and route of administration (oral, intravenous, intramuscular, or subcutaneous). Oral and intravenous protocols are widely used because intramuscular and subcutaneous protocols involve frequent injections. The route of administration can be switched without compromising the efficacy. It is feasible to perform desensitization through intravenous drug administration for meticulous dose adjustment and administer the therapeutic dose in oral form.

H1 blockers, H2 blockers, and glucocorticoids can be used as premedication. Aspirin and montelukast block the end products of the arachidonic acid cascade and decrease the incidence and severity of BTRs. NSAIDs can help to control discomfort, dyspnea, back pain, fever, chills, or myalgia. As mentioned above, the most prevalent causes of DHR include platinum, taxanes, and monoclonal antibodies, which present with different clinical manifestations. DHR to platinum has been reported in 1% to 80% [31-33]. The common causative agents include platinum (e.g., cisplatin, carboplatin, and oxaliplatin), taxanes (e.g., paclitaxel and docetaxel), and monoclonal antibodies (e.g., rituximab, infliximab, trastuzumab, bevacizumab, etc.) [34]. DHR is commonly caused by sensitization with drug-specific IgE, which can be confirmed by skin tests [35-38]. However, non-IgE-mediated reactions, such as infusion-related or solvent-related reactions (e.g., cremophor, polysorbate 80), have also been reported in the literature [32,33,39]. Mast cell-mediated or basophil-mediated reactions present with urticaria, angioedema, laryngeal edema, vomiting, abdominal pain, or anaphylaxis. On the other hand, cytokine-mediated reactions present with generalized rash, abrupt chest discomfort, dyspnea, back pain, fever, chills, or myalgia.

Antineoplastic agents

Antineoplastic agents are repeatedly administered at regular time intervals, which increase the risk of sensitization to these drugs. The incidence of DHR to antineoplastic agents varies according to the type of drug, ranging from 1% to 80% [31-33]. The common causative agents include platinum (e.g., cisplatin, carboplatin, and oxaliplatin), taxanes (e.g., paclitaxel and docetaxel), and monoclonal antibodies (e.g., rituximab, infliximab, trastuzumab, bevacizumab, etc.) [34]. DHR is commonly caused by sensitization with drug-specific IgE, which can be confirmed by skin tests [35-38]. However, non-IgE-mediated reactions, such as infusion-related or solvent-related reactions (e.g., cremophor, polysorbate 80), have also been reported in the literature [32,33,39]. Mast cell-mediated or basophil-mediated reactions present with urticaria, angioedema, laryngeal edema, vomiting, abdominal pain, or anaphylaxis. On the other hand, cytokine-mediated reactions present with generalized rash, abrupt chest discomfort, dyspnea, back pain, fever, chills, or myalgia.
especially when it is resumed after a drug-free period. The correlation between skin test positivity and DHR has been well established. As cross-reactivity can occur between platinum agents, care should be taken when switching agents within platinum-based regimens. As BTRs are common even after previous successful desensitization, shortening of the protocol should be adopted gradually and carefully. In contrast, infusion-related reactions to taxanes tend to occur during the first or second administration. In addition, IgE-mediated reactions can occur in some patients who have specific IgE to the pollen of *Taxus baccata*, a common yew used as a source of taxanes. DHR to taxanes has been reported in 5% to 10% of patients. As taxane-associated BTRs usually occur during the initial attempts of desensitization, the number of steps within desensitization protocols can be safely reduced if the BTR does not occur repeatedly during the early cycles of desensitization. Monoclonal antibodies lead to various types of DHR, including IgE-mediated reactions, cytokine release syndrome, and mixed reactions, which can arise in both the earlier and later courses. The rate of DHR varies according to the degree of humanization and glycosylation pattern, and commonly reported agents include rituximab and cetuximab. Especially for cetuximab, the association with anti-α-1,3-galactose IgE has been observed, which is generated from the consumption of animals that were bitten by a specific type of tick [30].

Various desensitization protocols have been proposed for antineoplastic agents. A protocol from the Adverse Drug Reaction and Desensitization Program in Brigham and Women’s Hospital has been well established worldwide (Table 1) [36]. In this protocol, the culprit drug solution (1:1) is diluted to 1:10, 1:100, and sometimes 1:1,000, and the concentration and rate of administration is escalated stepwise for 15 minutes. This multi-bag protocol is completed in most patients without serious BTR [40-42]. However, multi-bag protocols require excessive time and effort for dilution and changing bags, and increase the risk of exposure of medical staff to antineoplastic agents in the process. In addition, the stability of the drug can be compromised by dilution and administration for a prolonged period.

To overcome these shortcomings, a one-bag protocol was proposed in which a single concentration was used and only the rate of administration was increased (Table 2) [43-46]. Successful completion of desensitization with a one-bag protocol has been reported for all platinum, taxane, and monoclonal antibodies. There were no statistically significant differences regarding the rate of completion or the incidence of BTRs compared to multi-bag methods, while the amount of time required for the preparation was significantly reduced. In addition, the incidence of BTR decreased in the later course of treatment, which was consistent with multi-bag protocols [43-46]. Therefore, it is anticipated that the one-bag protocol can replace the multiple-bag protocol, thus reducing time and effort, maintaining the stability

| Step | Concentration, mg/mL | Rate, mL/hr | Infusion time, min | Dose, mg | Volume, mL |
|------|----------------------|-------------|-------------------|--------|-----------|
| 1    | 0.012                | 5           | 15                | 0.015  | 1.25      |
| 2    | 0.012                | 10          | 15                | 0.030  | 2.5       |
| 3    | 0.012                | 20          | 15                | 0.060  | 5         |
| 4    | 0.012                | 50          | 15                | 0.150  | 12.5      |
| 5    | 0.012                | 100         | 15                | 0.300  | 25        |
| 6    | 0.12                 | 20          | 15                | 0.60   | 5         |
| 7    | 0.12                 | 50          | 15                | 1.50   | 12.5      |
| 8    | 0.12                 | 100         | 15                | 3.00   | 25        |
| 9    | 1.2                  | 20          | 15                | 6      | 5         |
| 10   | 1.2                  | 50          | 15                | 15     | 12.5      |
| 11   | 1.2                  | 100         | 15                | 30     | 25        |
| 12   | 1.2                  | 200         | 135.8             | 543.36 | 452.8     |

Rituximab 600 mg/60 mL was reconstituted with 440 mL normal saline. The concentration of the solution was 1.2 mg/mL. Dose (mg) = rate (mL/hr) × time/60 (hr) × concentration (mg/mL).
of the agents, preventing erroneous dilutions, and reducing the risk of exposure of medical staff to antineoplastic agents.

In addition, the efficacy of chemotherapy is unlikely to be compromised by desensitization. Sloane et al. [42] conducted a large retrospective analysis of patients with recurrent ovarian cancer who underwent RDD for carboplatin; overall survival in the RDD group was non-inferior to that of the regular administration group.

**Antibiotics**

DHR to β-lactams, such as penicillin or cephalosporin, is more common than that to non-β-lactams. Desensitization can be performed for both immediate and delayed hypersensitivity reactions. The protocol should be selected based on patient characteristics, hospital capacity, and physician preferences. It is generally started with 1/1,000 of the therapeutic dose and then increased by 2 to 3-fold every 15 minutes to 5 hours. Oral administration is preferred due to its ease, safety, and effectiveness [47]. Desensitization to penicillin and cephalosporins has been well established. Successful desensitization has also been reported for other β-lactams, such as carbapenem and monobactam, and non-β-lactams, such as vancomycin, clindamycin, metronidazole, macrolides, aminoglycosides, tetracycline, and ciprofloxacin [48-58].

Red man syndrome (RMS) is a distinctive hypersensitivity reaction to vancomycin, ciprofloxacin, teicoplanin, or amphotericin; it is caused by the direct activation of mast cells and secretion of histamine, and is associated with the rate of administration [59]. It can usually be prevented by reducing the rate of administration and pretreating patients with antihistamines. However, in cases of moderate-to-severe RMS, the desensitization protocol can be applied in the absence of appropriate alternative agents. Both RDD and SDD have been studied for RMS; in most cases, RDD can be applied first and, in the case of failure, the protocol can be switched to SDD [60,61]. Successful desensitization to other antimicrobials has also been reported for antifungals, such as amphotericin B, fluconazole, itraconazole, voriconazole, and micafungin, and for antivirals, such as acyclovir, valganciclovir, ribavirin, and nevirapine [62-69].

**Antituberculous agents**

Tuberculosis possesses unique microbial characteristics that require prolonged simultaneous administration of multiple agents. DHR to antituberculous agents is reported in 4% to 5% of patients, which leads to the discontinuation or alteration of medications or prolonged treatment duration [70]. It is common for patients with tuberculosis to be hypersensitive to one or two agents, and delayed reactions, such as maculopapular rash, are more prevalent than immediate reactions, such as urticaria or angioedema [71]. Successful cases of desensitization to antituberculous

### Table 2. An example of a 1-bag, 12-step desensitization protocol for rituximab (600 mg)

| Step | Concentration, mg/mL | Rate, mL/hr | Time, min | Dose, mg | Volume, mL |
|------|----------------------|-------------|-----------|----------|------------|
| 1    | 2.857                | 0.1         | 15        | 0.1      | 0.025      |
| 2    | 2.857                | 0.2         | 15        | 0.1      | 0.05       |
| 3    | 2.857                | 0.4         | 15        | 0.3      | 0.1        |
| 4    | 2.857                | 0.8         | 15        | 0.6      | 0.2        |
| 5    | 2.857                | 1.6         | 15        | 1.1      | 0.4        |
| 6    | 2.857                | 3           | 15        | 2.1      | 0.75       |
| 7    | 2.857                | 6           | 15        | 4.3      | 1.5        |
| 8    | 2.857                | 12.5        | 15        | 8.9      | 3.125      |
| 9    | 2.857                | 25          | 15        | 17.9     | 6.25       |
| 10   | 2.857                | 50          | 15        | 35.7     | 12.5       |
| 11   | 2.857                | 100         | 15        | 71.4     | 25         |
| 12   | 2.857                | 150         | 64.04     | 457.4    | 160.1      |

Rituximab 600 mg/60 mL was reconstituted with 150 mL of normal saline. The concentration of the solution was 2.85714 mg/mL. Dose (mg) = rate (mL/hr) × time/60 (hr) × concentration (mg/mL).
agents, both oral and intravenous, have been reported in the literature since 1990. Through desensitization, 60% to 80% of patients who were hypersensitive to first-line antituberculous agents could continue therapy with those drugs (Table 3) [72]. In addition, second-line agents, such as streptomycin, kanamycin, and cycloserine, have also been continued safely with intramuscular, intravenous, and oral desensitization protocols [73]. In a Korean multicenter study of patients with severe cutaneous adverse reactions to antituberculous agents, the replacement of all drugs did not prevent the recurrence of DHR, possibly due to the development of multi-drug hypersensitivity after drug reaction with eosinophilia and systemic symptoms [74]. However, 84.6% of patients undergoing desensitization succeeded in continuing the original drugs, and their BTRs were no greater than mild.

Both RDD and SDD were studied at intervals of 15 minutes to 1 hour for RDD and a few hours to a day for SDD [75]. RDD has strengths in patient compliance and medication resistance, while SDD has better success rates and safety [29,72,76]. SDD has been recommended for delayed DHR; however, multiple successful RDDs for delayed reactions have also been reported [29].

### Table 3. An example of isoniazid desensitization protocol (target dose: 300 mg)

| Step | Concentration, mg/mL | Volume, mL | Time interval between doses, hr | Administered dose, mg |
|------|----------------------|------------|---------------------------------|-----------------------|
| Day 1|                      |            |                                 |                       |
| 1    | 0.2                  | 0.1        | 0.02                            |                       |
| 2    | 0.2                  | 0.2        | 0.04                            |                       |
| 3    | 0.2                  | 0.5        | 0.1                             |                       |
| 4    | 0.2                  | 1          | 2                               | 0.2                   |
| 5    | 0.2                  | 2          | 2                               | 0.4                   |
| 6    | 0.2                  | 4          | 2                               | 0.8                   |
| 7    | 0.2                  | 8          | 2                               | 1.6                   |
| 8    | 0.2                  | 16         | 2                               | 3.2                   |
| Day 2|                      |            |                                 |                       |
| 9    | 2.0                  | 3          | 6                               | 6                     |
| 10   | 2.0                  | 6          | 2                               | 12                    |
| 11   | 2.0                  | 12         | 2                               | 24                    |
| 12   | 100 mg Tab           | 0.5 Tab    | 2                               | 50                    |
| 13   | 100 mg Tab           | 1 Tab      | 2                               | 100                   |
| 14   | 100 mg Tab           | 2 Tab      | 2                               | 200                   |
| Day 3|                      |            |                                 |                       |
| 15   | 100 mg Tab           | 3 Tab      | 300                             |                       |

### Aspirin and NSAIDs

Hypersensitivity reactions to aspirin/NSAIDs can have varying manifestations, depending on the onset of symptoms and the organs involved [77]. Various protocols for aspirin/NSAIDs, which are adapted from aspirin provocation test protocols, have been suggested [3,78-82]. The mechanism of aspirin/NSAID hypersensitivity is still not fully understood, but non-immunological mechanisms of cyclooxygenase (COX) inhibition are the most plausible. The desensitization process downregulates the production of cysteinyl leukotriene and its receptors, attenuating skin and airway responses [83].

Aspirin/NSAID desensitization is considered for patients with cardiovascular or musculoskeletal diseases who require aspirin or NSAID administration for prolonged periods. It is also considered for patients with aspirin/NERD to improve the overall symptoms and quality of life, and decrease the formation of nasal polyps and sinus infections; however, the effects on asthma outcomes are less consistent [84].

Oral desensitization protocols for aspirin are well established. Notably, the therapeutic dose at the final step differs according to the indication for aspirin/NSAIDs. For cardiovascular or musculoskeletal diseases, a lower dose of aspirin (75 to 100 mg/day) is used. In contrast, for aspirin-exacerbated respiratory disease or chronic sinusitis with or without...
nasal polyps, a higher dose of aspirin (up to 625 mg/day) is administered (Table 4) [84].

For DHR to NSAIDs other than aspirin, it is recommended to switch to drugs that have little COX-1 inhibitory effect (e.g., acetaminophen), drugs that exhibit relatively selective COX-2 inhibition (e.g., nimesulide and meloxicam), or drugs that are highly selective COX-2 inhibitors (e.g., celecoxib and etoricoxib) if well tolerated. The desensitization protocol for NSAIDs can be modified from that for aspirin [77]. The temporary tolerance to aspirin/NSAIDs lasts 48 to 72 hours after desensitization. Therefore, hypersensitivity reactions can recur 2 to 5 days after discontinuation if the therapeutic dose is not continued [85,86].

CONCLUSIONS

Here, we discussed the mechanism of desensitization and its actual implementation for antineoplastic agents, antibiotics, antituberculous agents, and aspirin/NSAIDs. The development of hypersensitivity following repetitive administration of drugs is a major impediment to treatment. Desensitization induces temporary tolerance to the drug by gradually incrementing the dose to reach the therapeutic dose. Drug desensitization should be considered when the benefits of the culprit drug outweigh the risk. However, it should be applied carefully in patients with significant risk factors for fatal BTRs, such as those with a history of severe hypersensitivity reactions. In the desensitization procedure, various BTRs ranging from mild to severe can occur, but most resolve if desensitization is stopped immediately. Further research regarding the mechanism of desensitization and optimal protocols for different medications is warranted.

Conflict of interest
No potential conflict of interest relevant to this article was reported.

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Table 4. An example of aspirin desensitization protocol proposed by Silberman et al. and the EAACI guidelines (target dose: 100 and 325 mg)

| Dose of aspirin, mg | Cardiovascular or musculoskeletal diseases | Aspirin-exacerbated respiratory disease or chronic sinusitis with or without nasal polyps |
|---------------------|-------------------------------------------|------------------------------------------------------------------------------------------------|
| Dosing interval, min| Day 1  | Day 1 | Day 2 |
| 1 mg                | 20–40 mg | 100–160 mg |
| 30                  | 2 mg   |                                  |
| 30                  | 4 mg   |                                  |
| 30                  | 8 mg   | 2     | 40–60 mg | 160–325 mg |
| 30                  | 16 mg  |                                  |
| 30                  | 32 mg  |                                  |
| 30                  | 64 mg  | 2     | 60–100 mg | 325 mg |
| 30                  | 100 mg |                                  |

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