The effect of selenium and polysaccharide of Atractylodes macrocephala Koidz. (PAMK) on endoplasmic reticulum stress and apoptosis in chicken spleen induced by heat stress

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Endoplasmic reticulum (ER) stress and oxidative stress are involved in different types of stress induced injuries. The aim of the present study was to evaluate the effect of selenium (Se), polysaccharide of Atractylodes macrocephala Koidz. (PAMK), and the combination of Se and PAMK on oxidative stress, ER stress and apoptosis in chicken spleen under heat stress (HS) condition. Chickens (1 day old, n = 200) were randomly divided into two groups, the control (Con) group and the HS group. Then these chickens were treated with Se (0.3 mg kg\(^{-1}\)), PAMK (200 mg kg\(^{-1}\)) alone, and the combination of Se (0.3 mg kg\(^{-1}\)) and PAMK (200 mg kg\(^{-1}\)). The inducible NO synthase (iNOS)–NO activities (including NO content and iNOS activity), ER stress-related genes (GRP78, GRP94, ATF4, ATF6 and IRE) and apoptosis-related genes (caspase3 and Bcl-2) were examined by fluorescence quantitative PCR or western blot analysis. Apoptosis levels were also measured using ultrastructural observations. The results indicated that HS induced higher levels of NO content and the activity of iNOS which demonstrated that oxidative stress occurred in the chicken spleen. The ER stress-related genes (GRP78, GRP94, ATF4, ATF6 and IRE) and the apoptosis related gene caspase3 were increased (\(P < 0.05\)), while Bcl-2 was decreased (\(P < 0.05\)) induced by heat stress. These responses were alleviated by the treatment of Se, PAMK alone, and the combination of Se and PAMK (\(P < 0.05\) or not). The results showed that under common conditions, Se and PAMK could improve the immune response by reducing the levels of oxidative stress, ER stress and apoptosis to proper levels; however, under HS condition, Se and PAMK could change the abnormal levels of oxidative, ER damage and apoptosis to ameliorate the injury induced by HS. In addition, there existed a synergistic effect on the modulation of these biomarkers in chicken spleen between Se and PAMK. So both Se and PAMK may play important roles in regulating the immune function in chicken.

Introduction

Various environmental factors, such as extremely ambient temperature, can affect food intake, weight gain, and immunity in the poultry.\(^1\) Heat stress (HS), one of the most serious climate problems of tropical and subtropical countries, and negatively affects the production performance, humoral immunity and antioxidant status of broilers.\(^2\) As one important target of HS, the immune organs can significantly respond to HS. In both humans and animals (for example swine, chicken), the immune response was inhibited by HS and influences the normal performance of the hosts.\(^3\)\(^-\)\(^5\) Oxidative stress may induce apoptosis by intervening in the functional expressions of proapoptotic signal protein genes.\(^6\) Oxidative stress induced by HS has been reported as a primary factor that increases the pathogenesis of several diseases, decreases production results, and number of leucocytes in blood of chickens.\(^7\)\(^-\)\(^8\) Immune regulators alleviate immune system dysfunction and improve the condition of the host by regulating the immune response. Therefore, dietary manipulations are increasingly used as possible approaches to alleviate the harmful effects of high ambient temperature in poultry.\(^9\)

Selenium (Se) is an essential trace element as it plays a crucial role in antioxidant defence mechanisms.\(^9\) Dietary Se can supply antioxidants and is particularly active in maintaining the production performance and breeding in poultry.\(^11\) Se also plays crucial protective effects in the immune organs.\(^12\) Se supplementation has beneficial and detrimental effects on immunity to influenza vaccine in older adults.\(^13\) In addition, Se influences the antioxidation and immune function in heat-stressed broilers.\(^14\) Therefore, Se is usually used as one immune regulator under HS condition in livestock production.

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**Atractylodes macrocephala** Koidz. (AMK) as a traditional Chinese herb medicine has been utilized for at least 2000 years. Polysaccharide of *Atractylodes macrocephala* Koidz. (PAMK) has been shown to have immune regulating functions, and has already widely used in the breeding industry in China. It is reported that PAMK can ameliorate metabolic status and improve immune function in early weaned pigs. However, as one newly identified immune regulator, the possible mechanism by which PAMK influences the immune function is unknown. In our previous studies, we found that the compound of PAMK and Se could alleviate the inflammatory response and regulate the expression of HSPs gene in chicken spleen which induced by heat stress.\(^{16,17}\)

Both Se and PAMK preserve the roles of immune regulation, antioxidation, and or HS protective function. The effect of Se and PAMK on the regulation of immune function and injury under HS condition was less reported. In the current study, the objective was to investigate the effect of Se, PAMK, and the combination of Se and PAMK on NO content, iNOS activity, ER stress and apoptosis in a chicken immune system.

### Materials and methods

#### Chicken diet and sample collection

Chickens were treated humanely, and the experiments received prior ethical approval in accordance with Zhongkai University of Agriculture and Engineering and under the approved protocol number SRM-11. Two hundred chickens were assigned randomly by body weight to one of eight dietary treatments. Each treatment was replicated five times with 5 chickens each. During the first 3 weeks of the experiment, the chambers were not separated and recommended brooding temperatures were applied, so that the temperature was reduced progressively from 33 to 23.9 °C by the end of the third week of age. The control diets were formulated with the Se (1.5 mg kg\(^{-1}\) diet) and PAMK (0 mg kg\(^{-1}\)). The Se supplemental diets were formulated with the Se (3 mg kg\(^{-1}\) diet); the PAMK supplemental diets were formulated with the PAMK (200 mg kg\(^{-1}\)) and the combination group with Se (3 mg kg\(^{-1}\) diet) and PAMK (200 mg kg\(^{-1}\)). Throughout the entire experimental period, the chickens were allowed ad libitum consumption of feed and water, feed intake and body weights were recorded weekly. Clinical symptoms and mortality were also recorded. At the end of the feeding experiment, chickens were killed with sodium pentobarbital. The spleen tissue specimens were quickly removed, minced and stored at –80 °C to determine the index of oxidative stress and isolate the RNA and protein. The remaining tissues were used in the preliminary experiment and served as standby tissues.

#### Temperature and treatments

After 3 weeks, at the recommended brooding temperatures, five cages per dietary treatment were subjected to either a high temperature or the optimum temperature. In one chamber, the ambient temperature was set to 37 ± 1 °C. In the other chamber, the temperature was kept 23.9 °C. The relative humidity was allowed to fluctuate, but not to levels below 55%. For 4 weeks, the chickens were given the same dietary treatment. During the experiment, the chickens were on a lighting schedule of 24 h of light, with an average light intensity of 15 lx.

### Ultrastructural observations

For electron microscopy, spleen tissue specimens were fixed with 2.5% glutaraldehyde in 0.1 M sodium phosphate buffer (pH 7.2) for 3 h at 4 °C, washed in the same buffer for 1 h at 4 °C and postfixed with 1% osmium tetroxide in sodium phosphate buffer for 1 h at 4 °C. The tissues were then dehydrated in a graded series of ethanol starting at 50% for 10 min after two changes in propylene oxide. The tissue specimens were embedded in araldite. Ultrathin sections were stained with Mg-uranyl acetate and lead citrate for transmission electron microscope evaluation.

### Determination of NO content and iNOS activity in spleen

Chicken spleens were homogenized on ice in physiological saline and centrifuged at 700 × g for 15 min at 4 °C before supernatant collection. Here, we detected NO and iNOS as indices of oxidative damage. The NO content and iNOS activity were determined using NO and iNOS activity assay kits (Nanjing Jiancheng Bioengineering Institute, Nanjing, China). The method used in the present study was according to the procedure previously published by Liu with an ELX800 Microplate reader (BioTek Instruments, USA) to detect the OD at 550 and 530 nm, respectively.\(^{18}\)

### Quantitative real-time PCR analysis

The method of quantification used was followed by Yao.\(^{19}\) After quantification, the expression levels of autophagy genes were determined by quantitative reverse transcription PCR using SYBR Premix ExTaq TM (Takara, China) and an ABI PRISM 7500 real-time PCR system (Applied Biosystems). The PCR primers (Table 1) were designed using Oligo Primer Analysis software (version6.0) and synthesized by Invitrogen (Shanghai, China).

| Gene      | Primer sequence                        |
|-----------|----------------------------------------|
| GRP78     | Forward 5’-GAATCCCGCTAAACCAGGAGAGA-3’ |
|           | Reverse 3’-CCTTCACTTCCACACATT-3’       |
| GRP94     | Forward 5’-CAAGACATGGTACGAGAGATG-3’    |
|           | Reverse 3’-TTCCACCTTGGCATCACAGTCA-3’   |
| Bel-2     | Forward 5’-ATCGTCGGCTTTTCTGAGGT-3’     |
|           | Reverse 3’-CAGCCTACTCCGTTGGTCT-3’      |
| Caspase3  | Forward 5’-CATCTTCATCCTGTAATGCAAAG-3’  |
|           | Reverse 3’-CTTCCGGGCTGTGGTGAAGA-3’     |
| ATF4      | Forward 5’-GAATCCGCTAACACAGAGAGA-3’    |
|           | Reverse 3’-CCTGATCACCTCCATGACATC-3’    |
| ATF6      | Forward 5’-CAGGGCTGTACAGGAGAGA-3’      |
|           | Reverse 3’-CTACAGGTGCTCCTCCTACCAT-3’   |
| IRE       | Forward 5’-AAGAATCATCCTCAGGT GG-3’     |
| GADPH     | Reverse 5’-AGCCTTCATACCCCTCTTGTG-3’    |

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Reactions consisted of the following: 10 μl of 2 × SYBR Green I PCR Master Mix (TaKaRa, China), 0.4 μl of 50 × ROX reference Dye II, 0.4 μl of each primer (10 μM), 2 μl of diluted cDNA, and 6.8 μl of PCR-grade water. The PCR program for amplification of detect genes and GAPDH consisted of 95 °C for 30 s followed by 40 cycles at 95 °C for 15 s and 60 °C for 30 s. The results (fold changes) were expressed as \(2^{-\Delta\Delta C_t}\).

**Western blot analysis**

Protein extracts were subjected to SDS-polyacrylamide gel electrophoresis under reducing conditions on 12% gels. Separated proteins were then transferred to nitrocellulose membranes using a tank transfer for 2 h at 200 mA in Trisglycine buffer containing 20% methanol. Membranes were blocked with 5% skim milk for 16–24 h and incubated overnight with diluted primary chicken antibodies against Bcl-2 (1:500, Santa Cruz Biotechnology, USA), caspase3 (1:100, Santa Cruz Biotechnology, USA) and GRP78 (1:1000, polyclonal antibody produced by our lab) followed by a horseradish peroxidase (HRP)-conjugated secondary antibody against goat (Bcl-2) or rabbit (caspase3 and GRP78) IgG (1:1000, Santa Cruz Biotechnology, USA). To verify equal loading of the samples, the membrane was incubated with a monoclonal b-actin antibody (1:1000, Santa Cruz Biotechnology, USA), followed by a HRP-conjugated goat anti-mouse IgG (1:1000) secondary antibody. The signal was detected by X-ray films (Trans Gen Biotech Co., China). The optical density (OD) of each band was determined using the Image VCD gel imaging system, and Bcl-2, caspase3 and GRP78 expression were detected as the OD ratio between Bcl-2, caspase3 and GRP78 and that of β-actin respectively.

**Statistical analysis**

Statistical analysis of all data was performed by using SPSS for Windows (version 13, SPSS Inc., Chicago, IL). When a main effect \((p < 0.05)\) was identified by one-way analysis of variance, mean comparisons were followed. All data were checked for normal distribution and equal variance. Differences between means were assessed by Tukey’s honestly significant difference tests. Data are expressed as mean ± standard deviation. Differences were considered to be significant at \(p < 0.05\).

**Results**

**Ultrastructural changes**

No obvious ultrastructural changes were observed in all the control group (Fig. 1a and b). Under heat stress, the mitochondria from the spleen of HS group were vacuolated and the cristae had degenerated. The apoptotic cells showed typical condensed nuclei with horseshoe-like or crescent-shaped cytoplasmic organelles of an inconspicuous structure accompanied by nucleus shrinkage. The nuclei and organelles of some cells were unclear. The mitochondria were surrounded by fractured ER, and their rough ER were extended (Fig. 1c–e). When treating the HS chicken with Se or PAMK, the mitochondria damage was alleviated (Fig. 1f and g). In addition, the combination of Se and PAMK further enhanced the solo effect of Se and PAMK (Fig. 1h).

**Changes in NO content and iNOS activity**

The effects of Se and PAMK on NO content and iNOS activity in spleen are shown in (Fig. 2). The results indicated that under common condition, there was no different in NO content and iNOS activity via Se treatment or PAMK \((p > 0.05)\). But the combination of Se and PAMK could increase the activities of both NO and iNOS \((p < 0.05)\). Under HS condition, the NO content and iNOS activity were decreased \((p < 0.05)\). When treating the HS chicken with Se or PAMK, the NO content and iNOS activity were decreased \((p < 0.05)\) compared with the HS group. In addition, the combination of Se and PAMK further enhanced the solo effect of Se and PAMK.

![Fig. 1](image1.png) The ultrastructural changes in spleen of chickens in both control and heat stress group.
Effects of Se and PAMK on the expression of ER stress-related genes (GRP78, GRP94, ATF4, ATF6 and IRE) in spleen of chicken which induced by heat stress

The effects of Se and PAMK on the expression of ER stress-related genes in spleen are shown in (Fig. 3). The results indicated that under common condition, there was no different in mRNA level of GRP78, ATF4 and ATF6 gene via Se treatment or PAMK (p < 0.05), and Se treatment or PAMK could decrease the mRNA level of GRP94 and IRE genes. But the combination of Se and PAMK could decrease the mRNA level of all genes (p < 0.05).

Fig. 2  The NO content and iNOS activity. Data are means ± SD, n = 5. Bars without sharing a common letter were significantly different (P < 0.05).

Fig. 3  The mRNA levels of ER stress-related genes (GRP78, GRP94, ATF4, ATF6 and IRE) in spleen of chicken. Data are means ± SD, n = 5. Bars without sharing a common letter were significantly different (P < 0.05).
Under HS condition, the mRNA levels of all the genes were enhanced ($p < 0.05$). When treating the HS chicken with Se or PAMK, the mRNA level of all genes were decreased ($P < 0.05$) compared with the HS group. In addition, the combination of Se and PAMK further enhanced the solo effect of Se and PAMK. 

**Effects of Se and PAMK on the expression of apoptosis-related genes (Bcl-2 and caspase3) in spleen of chicken which induced by heat stress**

The effects of Se and PAMK on the expression of apoptosis-related genes in spleen are shown in (Fig. 4). The results showed that under common condition, Se treatment increased the mRNA level of Bcl-2 ($p < 0.05$), however, PAMK didn't cause any change in the mRNA level ($p < 0.05$). But the combination of Se and PAMK also could increase the mRNA of Bcl-2 gene ($p < 0.05$). The mRNA level of caspase3 in each treated groups were not influenced ($p < 0.05$), except in the compound group. Under HS condition, the mRNA level of Bcl-2 was decreased ($p < 0.05$), but the mRNA level of caspase3 was enhanced ($p < 0.05$). When treating the HS chicken with Se or PAMK, the mRNA level of Bcl-2 was increased ($p < 0.05$), but the levels of caspase3 was decreased ($p < 0.05$) compared with the HS group. In addition, the combination of Se and PAMK further enhanced the solo effect of Se and PAMK.

**Western bolt analysis of GRP78, Bcl-2 and caspase3 levels**

The protein expression levels of GRP78, Bcl-2 and caspase3 were examined by western blots. The results (Fig. 5) showed that under common condition, there was no difference in protein levels of GRP78 and caspase3 genes via Se treatment or PAMK ($p$
these proteins clearly indicated that heat stress could lead to ER stress. In addition, ER lesions were observed by ultrastructural observations in chicken spleen under heat stress condition. In addition, several pathways including IRE1, PERK, and ATF6 were also relate to ER stress. Among them, the PERK pathway enhanced translation of mRNAs, including ATF4.28 When ER stress occurred, the mRNA levels of those gene were increased.28,30

Caspases are key regulators of cell apoptosis, and are involved in ER stressed-induced apoptosis.29 When exposed to glucose-induced apoptosis, caspase3 activity was significantly increased in treated PC12 cells compared to controls.22 Moreover, during the process of apoptosis induced by ER stress, the Bcl-2 mRNA and protein expressions were decreased in lung epithelial cells.31 In the current study, both caspase3 mRNA and protein increased and Bcl-2 mRNA and protein levels decreased in the heat stress group. In addition, apoptosis was observed by ultrastructural observations in chicken spleen. These results showed that ER stress was really related to the occurrence of apoptosis.

Se is an famous antioxidant and plays an important role in immunological functions in humans and many other species.32 Previous studies showed that adequate Se level played an important role in nitric oxide mediated apoptosis induced by cadmium in chicken immune organs.15,34 Additionally, Se influenced the antioxidative and immune functions in HS broilers.33 In the present research, Se could improve the immune functions including oxidative damage, endoplasmic reticulum stress and apoptosis in HS-treated broilers.

Several nutritional approaches such as supplementation of diets with phytochemicals have been used attempted to attenuate the negative effects of heat stress.35 Diets supplemented with tomato pomace could affect the growth, immune, and antioxidant responses of heat stress-exposed broilers.36 Dietary supplementation of either betaine or turmeric rhizome powder alone or in combination could partially ameliorate some of the detrimental effects of HS in broilers.37 Lemon verbena and vitamin C improved some performance data and blood metabolites which somehow suppressed the negative effects of heat stress.38 In our current research, add PAMK also could improve the immune functions, it showed that PAMK had the similar function as the above phytochemicals.

In summary, the present study showed that HS induced the occurrence of oxidative damage, endoplasmic reticulum stress and apoptosis in chicken spleen which affects the immune function.

Both Se and PAMK could serve as the immune regulator under HS condition. And there also existed synergistic effect between Se and PAMK, which may enhance the ability of Se and PAMK to improve immune function.

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