Peels of Selected Fruits with Antioxidant and Anthelmintic potential

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Abstract

Helminth infestation occurs through one or more intestinal parasitic larvae like Ascaris, whipworm, hookworm, tapeworms etc. This is a serious problem encountered in most of the developing countries, predominantly seen in tropical and subtropical zones. Morbidity and mortality rates are increasing year after year. Various extracts of the peels of Punica granatum, Musa sapientum and Malus pumila were tested for their anti oxidant and anthelmintic potential. In hydrogen peroxide method, at a conc. of 100 µg/ml, Chloroform, methanol and aqueous extracts of apple peels produced 82.28%, 82.91% and 86.6% ; pomegranate peel extracts showed 77.51%, 85.97% and 75.98 % and banana peel extracts produced 85.16%, 83.18%, and 85.43% free radical scavenging activity respectively. All the extracts at a dose of 100 µg/ml showed significant ferric reducing power when compared to the standard ascorbic acid. In the anthelmintic activity determination, methanol extract (40mg/ml) of banana peels showed shortest time of paralysis and death when compared to the standard albendazole. Therefore the peels of selected fruits have potential use in helminth infestations and in free radical mediated diseases.

Keywords
Free radical, Anthelmintic activity, Ferric reducing power, peel extracts.

Introduction

Helminthiasis means infection of the human body with parasitic worms such as round worms, pinworms etc. The worms usually involve the intestinal tract but sometimes they may invade other organs. It is a serious problem not only in human beings but also livestock especially in tropics (Dhar et al, 1982). The ailment is particularly predominant in developing countries in association with poor Organization performs and insufficient regulating measures.

Anaemia is a major consequence of infection with GI nematodes, they have a detrimental effect on the host's nutritional status, also cause a loss of appetite and affect the host's physical, cognitive and social development. Influx of these infections can cause morbidity, and occasionally death, by bargaining nutritional status, affecting cognitive processes, persuading tissue responses, such as granuloma, and provoking intestinal obstruction or rectal prolapse in livestock.
In a hyperinfective form the worms are concentrated in the intestinal and respiratory tracts. They also spread throughout many organs, including the central nervous system. There is increased risk of meningitis and other secondary bacterial infections making the condition more worse. Control of helminthiasis is based on drug treatment, biological control, hygiene and health education as reported by WHO. Anthelmintic herbs possess properties that destroy helminths and protozoal parasitic worms or prevent their growth or replication and can be used therapeutically in the treatment of helminthiasis. Some of the medicinal plants possessing anthelmintic activity include Hunteria umbellata, Combretum mucronatum, Raw garlic (Allium sativum), Elaeophor biadrupifera, Hillaria latifolia, citrus aromaticum, Citrus medica, Citrus aurantifolia, Diospyros mespiliformis, Cucurbita maxima, Cucurbit amoschata, Cucurbita aromatic, Hagenia abyssinica etc.

Reactive oxygen species (ROS) have been found to play an important role in the initiation and/or progression of various diseases such as atherosclerosis, inflammatory diseases, cancer and cardiovascular disease. Thus, recent studies have investigated the potential of plant products as antioxidants against various diseases induced by free radicals (Pasupathi et al, 2009 and Farber, 1994). Peels of pomegranate (Punica granatum), the inedible portion contains as much as three times the total amount of polyphenols, including condensed tannins and catechins, gallocatechins and prodelphinidins. It is used in heart problems, treatment of cough and cold, used in dental hygiene, anti-aging and skin cancer (Ricci and Giamperi, 2006; Kotamballi, 2002). Peels of banana (Musa sapientum), contain protein, crude lipid, carbohydrate and crude fibre. It is used in treatment of heavy metal contamination from river water, also used as a means of filtration for heavy metals and radionuclides occurring in water produced by the nuclear and fertilizer industries (Chang-Peng Yang et al., 2001; Pingyi Zhang, Roy et al., 2005).

Apple (Malus pumila) peels are rich in Alpha-Linolenic-Acid, Asparagine, D-Catetin, Isoquercetin, Hyperoside, Ferulic-Acid, Farnesene, Neoxatin, Phosphatidyl-Choline, Reynoutrin, Sinapic-Acid, Caffeic-Acid, Chlorogenic-Acid, P-Hydroxy-Benzoic-Acid, P-Coumaric-Acid, Avicularin, Lutein, Quercitin, Rutin, Ursolic-Acid, Protocatechuic-Acid, and Silver (Edgar Pastene et al., 2009; Kelly Wolfe et al., 2003).

**Materials and Methods**

Chloroform (Merck Specialities Pvt. Ltd.), Methanol (Loba chemicals), Hydrochloric acid (Hi-Pure chemicals), Sulphuric acid (Hi-pure chemicals), Ammonia ((Loba chemicals), Sodium hydroxide (SD fine chemicals), Lead acetate (Hi-Pure chemicals), Sodium nitroprusside (Lobachemicals), Acetic anhydride (Loba chemicals), Tannic acid, Molish reagent (Loba chemicals), Picric acid, Potassium hydroxide (Loba chemicals), Libbermann-Burchard’s reagent, Potassium dichromate (Hi-Pure chemicals), Copper sulphate, Hydrogen peroxide, Saline solution, Ascorbic acid, DMSO (Loba chemicals), Potassium ferricyanide, Gum acacia, Sodium chloride (Loba chemicals), Albendazole (local drug store).

**Extraction**

The fruits of Punica granatum, Malus pumila, Musa sapientum were procured from nearby fruit market and authenticated in the Department of Botany, P.B. Siddhartha College of Arts and Science, Vijayawada. The peels were manually separated and
subjected to drying under shade and a specimen was preserved for future reference. The dried peels were made into powder in a mechanical grinder. The powdered plant material is subjected to successive extraction using different solvents like chloroform, methanol, and water. Extraction with chloroform and methanol was done by maceration. Aqueous extract is obtained using soxhlet apparatus. The following procedure was followed:

The powder is macerated with chloroform for 6 days. Then the contents are filtered by vacuum filtration. The marc is retreated with same solvent for another 3 days. The contents are filtered, combined and concentrated.

The marc is dried and extracted with methanol in the same manner. After methanol extraction, the solid material was sufficiently dried and extracted with water in a soxhlet apparatus for eight hours period. The extract was concentrated. All the extracts were stored in a desiccator.

**Preliminary Phytochemical Screening**

The extracts obtained were weighed and yield was calculated Table1. Preliminary phytochemical screening was carried out following standard procedures (Khandelwal, 2004; Kokate, 2006). The extracts were abbreviated as follows:

Chloroform extract: *Punica granatum* (Pomegranate) (PCE), *Malus pumila* (Apple) (ACE), *Musa sapientum* (Banana) (BCE)

Methanol extract: *Punica granatum* (Pomegranate) (PME), *Malus pumila* (Apple) (AME), *Musa sapientum* (Banana) (BME)

Aqueous extract: *Punica granatum* (Pomegranate) (PAE), *Malus pumila* (Apple) (AAE), *Musa sapientum* (Banana) (BAE)

**In vitro Evaluation of Anthelmintic Activity**

Indian adult earthworms (*Pheritima posthuma*) were used to study anthelmintic activity due to anatomical and physiological resemblance to intestinal round worm parasites (Vidyarthi, 1967; Thompson, 1995). Earthworms were collected from the water logged areas and then washed with normal saline to remove soil and fecal matter. The earthworms of 5-8 cm length and 0.2-0.3 cm in width were used for all experimental protocols. They were authenticated in the department of Zoology, Siddhartha Arts and Science College, Vijayawada.

**Experimental Procedure**

Anthelmintic activity was evaluated by the method of Ajaieyoba et al (2001). Fourteen groups of approximately equal size Indian earth worms consisting of six earthworms in each group were used for the study. Groups were treated according to the protocol.

**Preparation of Test Samples**

Samples for evaluation of anthelmintic activity were prepared by dissolving 2.5gm of crude extract in 25 ml of 1% gum acacia solution prepared in normal saline. 20 and 40 mg/ml conc. were used in the study.

**Preparation of Standard**

A standard drug Albendazole is procured from the nearby medical store. A solution of 50mg/ml conc. is prepared in 1% gum acacia prepared in normal saline solution.

**Procedure**

The samples were taken in petriplates and adult healthy earth worms(n=6) were introduced into them. Observations were
made for the time taken to paralyze and time for death of individual worms. Paralysis was said to occur when the worms do not revive even when introduced into normal saline. Death was concluded when worms lost their motility, followed by fading away of their body colour.

**Evaluation of Anti-Oxidant Activity**

(a) **Hydrogen Peroxide Method** (Kumaran, 2007; Srinivasan, 2007).

A solution of hydrogen peroxide (20 mM) was prepared in phosphate buffer saline (pH 7.4), different concentrations of plant extract and standard ascorbic acid solution viz. 10, 20, 40, 60, 80 and 100 μg/ml in methanol (1 ml) where added to hydrogen peroxide solution (2 ml). Absorbance of hydrogen peroxide was determined at 230 nm after 10 min against a blank. Solution containing phosphate buffer without hydrogen peroxide is used as a blank. For each concentration, a separate blank sample was used for background subtraction. The antioxidant activity of the extract was expressed as IC50. All the tests were performed in triplicate and the graph was plotted with the average of three observations.

The percentage inhibition activity was calculated from \( \frac{(A_0 - A_1)}{A_0} \times 100 \), where \( A_0 \) is the absorbance of the control and \( A_1 \) is the absorbance of extract/standard.

**Ferric Reducing Power Determination**

Reducing power of the extracts was determined by Yen and Chen method (Yen GC and Chen, 1995). Different concentrations of fruit peel extracts and standard (STD) ascorbic acid solution viz. 20, 40, 60, 80 and 100 μg/ml in 1 ml of methanol were mixed with phosphate buffer (2.5 ml, 0.2 M pH 6.6) and Potassium ferricyanide [K3Fe(CN)6] (2.5 ml, 1%). The mixture was incubated at 50°C for 20 min. A portion (2.5 ml) of trichloroacetic acid (10%) was added to the mixture, which was then centrifuged at 3,000 g (rpm) for 10 min at room temperature. The upper layer of solution (2.5 ml) was mixed with distilled water (2.5 ml) and ferric chloride(FeCl3) (0.5 ml, 0.1%). A blue-green colour was obtained in the reaction, whose O.D was measured at 700 nm. An increase in the absorbance of the reaction mixture indicated increased reducing power. All the tests were performed in triplicate and the graph was plotted with the average of three observations.

**Results and Discussion**

Yield was calculated using formula:

\[
\% \text{ Yield} = \frac{\text{Weight of the extract}}{\text{Weight of the plant material}} \times 100
\]

The percentage yield of fruit peel extracts is given in Table 1. The maximum percentage of yield was reported with methanol extract, followed by aqueous extract and chloroform extract.

**Preliminary Phytochemical Screening**

The chloroform extracts of all the three plants contain alkaloids, fats and fixed oils, steroids and triterpenoids, and carbohydrates. Methanol extract of pomegranate peels contains carbohydrates, glycosides, steroids and triterpenoids. Methanol extract of apple and banana peels contains carbohydrates, fats and fixed oils, glycosides, steroids and triterpenoids. Aqueous extract of pomegranate was found to contain alkaloids, glycosides, tannins and carbohydrates. Apple aqueous extract contains carbohydrates and tannins. Banana
aqueous extract contains glycosides and carbohydrates

Basic identification of the chemical constituents of a plant drug is possible with phytochemical screening. The therapeutic activity of any plant drug resides in its phytochemicals. A knowledge of the chemical constituents helps in disclosing the pharmacological activities.

**Evaluation of Anthelmintic Activity**

The anthelmintic activity of selected fruit peel extracts were evaluated using Indian earthworms against the standard albendazole. The results of Anthelmintic activity are shown in Table 3. Results are expressed as an average of six observations. Among the extracts, methanol extract(40mg/ml) of banana peels showed shortest time of paralysis and death. At the test dose of 40mg/ml, the time required to paralyze the worms was 17.5± 0.09, min. and death was recorded at 22.5 ± 0.19 min. whereas the time required to paralyze and kill the worms with standard (Albendazole) was 45.0± 0.45and 55.0±0.20 min. respectively. The activity of methanol extract was comparable to that of standard drug Albendazole with respect to time taken to paralyze and to kill the worms. All the extracts showed dose related response. Significant activity was observed with chloroform, methanol and aqueous extracts of *Punica granatum, Musa sapientum, Malus pumila*. The potency of the extracts was inversely proportional to the time taken for paralysis /death of the worms. The control group animals were alive up to 24hrs.

Increasing order of potency of the extracts is: BME >AAE >AME> PAE > BAE > PME

Chemotherapeutic agents act as anthelmintics by disruption of neuromuscular physiology, blockade of energy metabolism, disrupting reproductive system which is highly efficient in these parasites(Martin,1997; Chatterjee,1967; Thornton,1977). The tested extracts may act as anthelmintics by anyone or a combination of these mechanisms. As the extracts caused paralysis as well as death, they can be confirmed to possess potent anthelmintic activity. Some synthetic phenolic anthelmintics interefers with energy of helminth parasites by uncoupling oxidative phosphorylation. Albendazole by increasing chloride ion conductance of worm muscle membrane produces hyperpolarization and reduced excitability that leads to muscle relaxation and flaccid paralysis that results in expulsion of the worms by peristalsis. The anthelmintic activity of the extract may be attributed to the presence of active components such as glycosides, triterpenoids, alkaloids, and tannins that may act by one or the other mechanisms as explained above. The anthelmintic properties of these phytoconstituents are well documented (Vigar, 1984; Bate Smith, 1962; Harekrishna, 2010; Bhakta, 2013; Ajay Sharma, 2011).

**Evaluation of Anti-oxidant Activity**

Free radical scavenging activity of the selected extracts was carried out by H2O2 method. The results are displayed in Table 4, fig 2,3&4. The scavenging activity of the standard ascorbic acid and various peel extracts after incubation for 10 minutes was observed to increase with an increase in their concentration.
Table 1 Percentage Yield

| S.no | Solvent     | Name of the extract | %yield |
|------|-------------|---------------------|--------|
| 1    | Chloroform  | PCE                 | 3      |
| 2    |             | ACE                 | 2      |
| 3    |             | BCE                 | 2.5    |
| 4    | Methanol    | PME                 | 5      |
| 5    |             | AME                 | 3      |
| 6    |             | BME                 | 4.5    |
| 7    | Aqueous     | PAE                 | 4      |
| 8    |             | AAE                 | 3      |
| 9    |             | BAE                 | 3.5    |

Table 2 Preliminary Phytochemical Screening

| SL.NO | TEST                        | PCE | PME | PAE | ACE | AME | AAE | BCE | BME | BAE |
|-------|------------------------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| 1     | Test for alkaloids           | +   | +   | +   | +   | -   | +   | +   | -   |     |
| 2     | Test for carbohydrates       | +   | +   | +   | +   | +   | +   | +   | +   | +   |
| 3     | Test for fats and fixed oils | +   | -   | -   | +   | -   | +   | +   | -   |     |
| 4     | Test for glycosides          | -   | +   | -   | -   | +   | -   | -   | -   | +   |
| 5     | Test for specific glycosides | -   | -   | +   | -   | -   | -   | -   | -   |     |
| 6     | Test for tannins             | -   | -   | +   | -   | -   | +   | -   | -   |     |
| 7     | Test for steroids and        | +   | +   | -   | +   | +   | -   | +   | +   | -   |
|       | triterpinoids                |     |     |     |     |     |     |     |     |     |
| 8     | Test for flavanoids          | -   | -   | -   | -   | -   | -   | -   | -   | -   |

+ indicates presence, - indicates absence

Fig. 1 Radical scavenging activity of Pomegranate peels

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**Fig. 1** Radical scavenging activity of Pomegranate peels

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**Table.1** Percentage Yield

| S.no | Solvent     | Name of the extract | %yield |
|------|-------------|---------------------|--------|
| 1    | Chloroform  | PCE                 | 3      |
| 2    |             | ACE                 | 2      |
| 3    |             | BCE                 | 2.5    |
| 4    | Methanol    | PME                 | 5      |
| 5    |             | AME                 | 3      |
| 6    |             | BME                 | 4.5    |
| 7    | Aqueous     | PAE                 | 4      |
| 8    |             | AAE                 | 3      |
| 9    |             | BAE                 | 3.5    |

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**Table.2** Preliminary Phytochemical Screening

| SL.NO | TEST                        | PCE | PME | PAE | ACE | AME | AAE | BCE | BME | BAE |
|-------|------------------------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| 1     | Test for alkaloids           | +   | +   | +   | +   | +   | -   | +   | +   | -   |
| 2     | Test for carbohydrates       | +   | +   | +   | +   | +   | +   | +   | +   | +   |
| 3     | Test for fats and fixed oils | +   | -   | -   | +   | -   | +   | +   | -   |     |
| 4     | Test for glycosides          | -   | +   | -   | +   | -   | -   | -   | -   | +   |
| 5     | Test for specific glycosides | -   | -   | +   | -   | -   | -   | -   | -   | -   |
| 6     | Test for tannins             | -   | -   | +   | -   | -   | +   | -   | -   |     |
| 7     | Test for steroids and        | +   | +   | -   | +   | +   | -   | +   | +   | -   |
|       | triterpinoids                |     |     |     |     |     |     |     |     |     |
| 8     | Test for flavanoids          | -   | -   | -   | -   | -   | -   | -   | -   | -   |

+ indicates presence, - indicates absence
**Fig. 2** Radical Scavenging Activity of Apple Pomegranate Peels

![Graph showing Apple anti-oxidant activity with different doses of various samples.]

**Fig. 3** Radical Scavenging activity of Banana Peels

![Graph showing Banana anti-oxidant activity with different doses of various samples.]

**Table 3** Anthelmintic Activity

| SL.NO | SAMPLE     | DOSE (mg/ml) | RESPONSE          | Paralysis Time (Min) | Death Time (Min) |
|-------|------------|--------------|-------------------|----------------------|------------------|
| 1     | CONTROL    |              |                   | NO RESPONSE         | NO RESPONSE      |
| 2     | PME        | 20           | 42.5 ± 0.11       | 50.5 ± 0.3           |
| 3     | PME        | 40           | 32.5 ± 0.14       | 44.5 ± 0.08          |
| 4     | PAE        | 20           | 49 ± 0.16         | 54.5 ± 0.11          |
| 5     | PAE        | 40           | 22.5 ± 0.06       | 29.0 ± 0.15          |
| 6     | AME        | 20           | 62.5 ± 0.18       | 74 ± 0.14            |
| 7     | AME        | 40           | 21.5 ± 0.16       | 32.5 ± 0.07          |
| 8     | AAE        | 20           | 90 ± 0.09         | 100 ± 0.08           |
| 9     | AAE        | 40           | 20 ± 0.12         | 35 ± 0.18            |
| 10    | BME        | 20           | 55 ± 0.06         | 71 ± 0.16            |
| 11    | BME        | 40           | 17.5 ± 0.09       | 22.5 ± 0.19          |
| 12    | BAE        | 20           | 40 ± 0.17         | 52.5 ± 0.05          |
| 13    | BAE        | 40           | 30 ± 0.01         | 40 ± 0.14            |
| 14    | ALBENDAZOLE (STD) | 50   | 45 ± 0.45         | 55 ± 0.20            |
Table 4: H2O2 Free Radical Scavenging Activity of Fruit Peel Extracts

| Conc (µg/ml) | PCE | PME | PAE | ACE | AME | AAE | BCE | BME | BAE | STD |
|--------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| 10           | 60.97 | 36.96 | 33.86 | 50.08 | 52.24 | 41.54 | 62.94 | 53.05 | 49.01 | 63.12 |
| 20           | 62.94 | 40.1 | 39.74 | 61.6 | 65.55 | 58.18 | 66.63 | 57.91 | 63.3 | 68.25 |
| 40           | 67.8 | 68.7 | 50.26 | 66.72 | 68.52 | 72.57 | 75.26 | 72.75 | 71.49 | 74.19 |
| 60           | 72.84 | 77.51 | 57.28 | 74.37 | 73.29 | 73.74 | 79.31 | 74.91 | 75.71 | 84.26 |
| 80           | 74.73 | 81.83 | 67.8 | 76.97 | 77.24 | 79.49 | 82.55 | 77.96 | 84.26 | 89.47 |
| 100          | 77.51 | 85.97 | 75.98 | 82.28 | 82.91 | 86.6 | 85.16 | 83.18 | 85.43 | 90.01 |

Table 5: Reducing Power of Various Peel Extracts

| Conc (µg/ml) | PCE | PME | PAE | ACE | AME | AAE | BCE | BME | BAE | STD |
|--------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| 10           | 0.412 | 0.31 | 0.2 | 0.219 | 0.218 | 0.35 | 0.3 | 0.353 | 0.165 | 0.198 |
| 20           | 0.43 | 0.335 | 0.225 | 0.238 | 0.25 | 0.364 | 0.308 | 0.36 | 0.225 | 0.242 |
| 40           | 0.439 | 0.365 | 0.265 | 0.26 | 0.275 | 0.375 | 0.358 | 0.374 | 0.27 | 0.287 |
| 60           | 0.505 | 0.39 | 0.283 | 0.305 | 0.283 | 0.38 | 0.37 | 0.385 | 0.358 | 0.297 |
| 80           | 0.511 | 0.445 | 0.36 | 0.385 | 0.415 | 0.41 | 0.445 | 0.398 | 0.38 | 0.31 |
| 100          | 0.518 | 0.512 | 0.435 | 0.425 | 0.42 | 0.425 | 0.465 | 0.435 | 0.425 | 0.362 |

Fig. 5: Pomegranate Peels Reducing Activity

Fig. 6: Apple Peels Reducing Activity
All the peel extracts exhibited significant hydrogen peroxide scavenging activity when compared to the standard ascorbic acid at similar concentrations. Among various extracts of *Malus pumila*, aqueous extract showed potent free radical scavenging activity which was comparable to that of the standard, ascorbic acid. *Punica granatum*, methanol extract and *Musa sapientum* aqueous extracts also showed significant activity. At 100 µg/ml conc. Chloroform, methanol and aqueous extracts of apple peels produced 82.28%, 82.91% and 86.6% ; pomegranate peel extracts showed 77.51%, 85.97% and 75.98% and banana peel extracts produced 85.16%, 83.18%, and 85.43% free radical scavenging activity respectively. The % scavenging activity as exhibited by standard in this conc. was 90.01. Alkaloids, tannins, steroids and triterpinoids are the chief principles identified in the tested extracts. All these extracts possessed antioxidant activities. Tannins have a strong free radical scavenging activity and anti-inflammatory activity. They regulate cellular activities of inflammation related cells, macrophages, lymphocytes, Neutrophils. They also modulate metabolizing enzymes like, Phospholipase A2, Cyclooxygenase (COX), Lipoxygenase (LOX) and Nitric oxide synthase (NOS) (Weisburger, 1999; Nabavi, 2008).

Hydrogen peroxide itself is not very reactive, it can generate the highly reactive hydroxyl radical (OH) through the Fenton reaction. Thus, the scavenging of hydrogen peroxide is an important antioxidant defense mechanism (Rice-Evans et al., 1995; Nabavi et al., 2009; Ying Chen et al., 2014; Rajesh Mandade et al., 2011; Sanchez, 2006).

\[
\text{Fe}^{2+} + \text{H}_2\text{O}_2 + \text{Fe}^{3+} + \text{OH} + \text{OH}^- 
\]

The decomposition of hydrogen peroxide to water involves the transfer of electrons as in the following reaction:

\[
\text{H}_2\text{O}_2 + 2\text{H}^{+} + 2\text{e}^- \rightarrow 2\text{H}_2\text{O}
\]

Electron donating ability of the tested peels may be responsible for their free radical scavenging activity. Increasing order of the free radical scavenging activity of various extracts of the fruit peels is as follows:

AAE > PME > BAE > BCE > BME > AME > ACE > PCE > PAE.

The results of Ferric reducing ability of the tested extracts are shown in table 5, fig 5, 6 & 7. Ferric reducing power determines the electron-donating capacity of an antioxidant (Kumar et al., ). The reduction of the ferric ion (Fe$^{3+}$) to ferrous ion (Fe$^{2+}$) is measured by the intensity of the resultant blue-green solution which has a maximum...
absorption at 700nm, and an increase in absorbance is indicative of higher reducing power of the antioxidant. The reducing power of the extracts was found to increase progressively over the concentration range studied. Extract at a dose of 100 µg/ml showed significant ferric reducing power when compared to the standard. Chloroform extracts of all the peels exhibited potent action when compared to methanol and water extracts.

In conclusion, based on the literature survey peels of selected fruits were evaluated for anthelmintic activity and anti-oxidant activity as they were rich in important constituents. Peels, which are considered to be a waste product were experimentally proved to possess promising anthelmintic activity and antioxidant activity. There is a lot of scope to work in this field and reveal the hidden secrets of the fruit peels. Therefore, the fruit peels of pomegranate, banana and apple can be used in the treatment of helminth infestations and in free radical mediated diseases.

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