Genetic modification of sorghum for improved nutritional value: state of the problem and current approaches

Abstract

Background: Low digestibility of seed storage proteins (kafirins) and low nutritional grain value are one of the most important directions of genetic improvement of sorghum. In this review, we summarize experiments on improving protein digestibility, and increasing nutritional value of sorghum grain using genetic engineering approaches, primarily, by introduction of genetic constructs that cause the silencing of γ- and/or α-kafirins. Key points of improvement of Agrobacterium-mediated genetic transformation, which was used for introduction of these genetic constructs in sorghum genome, are discussed. Transgenic sorghum harboring the genetic constructs inducing silencing of kafirins are characterized by reduction of kafirin oligomers, an increase of in vitro digestibility of proteins of flour, an increase in the proportion of essential amino acids, lysine, in particular, a changes in the ultrastructure of endosperm protein bodies (in case of α-kafirin silencing), which acquire an irregular shape with invaginations, and modification of endosperm texture, which become floury devoid of the vitreous layer. In some cases transgenic events with vitreous endosperm and high digestibility of kafirins were observed, which, in this connection, may be of interest for practical sorghum breeding. There are evidences that in vitro protein digestibility in transgenic plants with genetic construct for γ-kafirin silencing grown in the field plot was substantially lower than in the plants grown in the pots in greenhouse conditions. These facts indicate that suppression of the synthesis of kafirins occurring at epigenetic level as a result of the expression of the genetic constructs for silencing, is influenced by environment. In this connection, the use of methods of genome editing, in particular, editing the nucleotide sequences of the α- or γ-kafirin genes, opens wider prospects for improving the digestibility of kafirins and increasing the nutritional value of sorghum grain. Current approaches for biofortification of sorghum grain with amino acids and micronutrients are discussed.

Keywords: Agrobacterium-mediated genetic transformation, RNA silencing, kafirins, in vitro protein digestibility, endosperm, Sorghum bicolor (L.) Moench.

Introduction

Global warming is one of the most serious problems that threaten the existence of mankind. The results of observations carried out in different countries clearly demonstrate the steady tendency to increase the average annual temperature on the Earth during the last 150 years. For example, in the Russian Federation, in the next 15 years, in the summer season, the temperature will rise by 1.4-1.5°C in all grain-producing regions. In the next 40 years, depending on the rate of increase of the greenhouse effect, the temperature will rise by 2.0-3.3°C. In this connection, the creation of heat-tolerant and drought-resistant varieties and hybrids, the introduction of drought-resistant crops into agricultural production become extremely urgent problems of genetics and plant breeding.

In this regard, sorghum is of exceptional importance. This relatively high-yield, drought tolerant cereal crop is a reliable source of feed and food grains for regions that are regularly affected by drought, in which sustainable production of traditional cereals such as wheat, maize and barley is challenging. Sorghum is one of the five most important cereal crops cultivated on the Earth. Sorghum grain is the dietary staple for more than 500 million people in 30 countries. Sorghum grain does not contain gluten and can serve as a source of protein for patients suffering from celiac disease (gluten sensitive), who must follow a gluten-free diet. At the same time, sorghum, along with high yield and high drought resistance, is characterized by a number of nutritional constrains, which make it inferior to the other major cereals: lower digestibility of seed storage proteins (kafirins) and starch, low nutritional value of kafirins, which have a low content of essential amino acids, lysine and threonine, in particular.

The reasons of kafirins resistance to proteolytic digestion are multifactorial. Among the factors that cause or may effect this phenomenon there are chemical structure of kafirin molecules, some of which (α- and β-kafirins) are abundant with sulphur-containing amino acids capable to form S–S bonds, resistant to protease digestion; interactions of kafirins with non-kafirin proteins and non-protein components such as polyphenols and polysaccharides; spatial organization of different kafirins in the protein bodies of endosperm cells. It is generally accepted that the peripheral disposition of γ-kafirin in protein bodies reduces digestibility of α-kafirin – the major sorghum seed storage protein located central position in protein bodies and comprising up to 80% of total endosperm kafirins. This hypothesis is supported by studies of protein bodies of the mutant with γ-kafirin silencing, which has been changed from spherical to invaginate; the γ-kafirin located...
at the bottom of invaginations where it should not interfere with the digestion of the α-kafirin. In addition, γ-kafirin is characterized by ability to form oligo- or polymers of high molecular weight, which are resistant to protease digestion. These oligomers comprise α- and γ-kafirins that are linked by disulfide (S–S) bonds. A subsequent study also showed that a sorghum mutant with high digestibility of kafirins has a point mutation in the signal sequence of the α-kafirin gene, which apparently disrupts its deposition in protein bodies.

Improving the kafirins digestibility and increasing the nutritional value of sorghum grains is possible with the help of different genomic and biotechnology tools that target introduction of novel genetic variation leading towards impaired synthesis or modified amino acid composition of kafirins; identification of natural allelic variants of kafirins; increasing the lysine content of sorghum grain by introducing genetic constructs encoding lysine-rich proteins or suppressing synthesis of the enzyme catalyzing lysine (lysine ketoglutarate reductase); the introduction of genetic constructs that cause the silencing of γ- and/or α-kafirin genes; editing nucleotide sequences of γ and/or α-kafirin genes for the purpose of knockout. In this review, we summarized the published reports on improving the digestibility of kafirins and increasing the nutritional value of sorghum grains using genetic engineering approaches and outlined their importance for practical sorghum breeding.

**Sorghum genetic transformation**

The use of biotechnological tools to address the problem of improving the digestibility of kafirins requires reliable methods of introducing genetic constructs into genome of different sorghum lines and reliable methods of plant regeneration from transformed cells. In spite of numerous reports on obtaining transgenic sorghum plants by using microbomabrdiment technique, *Agrobacterium*-mediated genetic transformation, pollen-mediated transformation, genetic transformation of sorghum for a long time was a serious problem (see reviews). However, as a result of intensive research in recent years, significant progress has been made in this area. To date, the methods of introducing transgenes into the genome of sorghum, both by microprojectile transformation, and through *Agrobacterium*-mediated genetic transformation were significantly improved.

Key points of improvement of these methods of genetic transformation have been the use of effective technologies for the induction of embryogenic callus and plant regeneration. For example, the use of M11 culture medium containing high concentrations of amino acids L-asparagine and L-proline combined with elevated phosphate levels significantly stimulates the formation of embryogenic callus in many sorghum lines and prevents the release of specific pigments having toxic effects. This medium increased the frequency of obtaining transgenic sorghum plants in microprojectile transformation and in experiments on *Agrobacterium*-mediated genetic transformation. Increased level of copper ions in the medium for plant regeneration greatly stimulates plant regeneration in sorghum tissue culture, which also contributed to the increase rate of transgenic plants. Recently, using these modifications of nutrient media Do et al. reported on significant increase of frequency of transgenic sorghum plants (up to 14%) in experiments with three strains of *A. tumefaciens*, which contained standard binary vectors.

In addition, in our early studies on the development of methods for obtaining callus cultures with high and stable regeneration capacity from mature sorghum embryos, a high efficiency of a nutrient medium containing a combination of 2,4-D and 6-BAP was shown. Efficiency of the medium with this combination of phytohormones to increase the frequency of transgenic plants in experiments on *Agrobacterium*-mediated genetic transformation has been clearly demonstrated. In this regard, one should mention the report on significant stimulation of callus induction from immature sorghum embryos and plant regeneration by lipoic acid that allowed obtaining high transformation frequency (27.2%) in particle bombardment experiments. Positive effect of lipoic acid might be caused by its anti-oxidative capacity that is important for *in vitro* cultured sorghum embryos.

Alongside with the methods of tissue culture, a significant increase in the frequency of transgenic sorghum plants in experiments on agrobacterial transformation was facilitated by the use of “hypervirulent” strains. These strains contain additional copies of *vir* genes located either on specially designed Ti-plasmids (“superbinary” vectors, such as pSh1) or on additional plasmids (pVIR vectors). It was reported that NTL/Cry59 and AGL1 produced higher transformation frequency than other strains. In our experiments we observed high frequency (9%) of regeneration of transgenic sorghum plants with using AGLO strain. A ternary vector system harboring a disarmed Ti plasmid, an accessory T-DNA-less pVIR plasmid with additional *vir*-genes, and the T-DNA transfer competent binary plasmid increased the frequency of sorghum transformation up to 29% and allowed to obtain transgenic plants in several recalcitrant African sorghum varieties.

Another important factor is the use of effective promoters, which ensure a high level of expression of the marker gene and prevent the regeneration of escapes. The maize ubiquitin 1 promoter (*ubi1*) was shown to provide high level of gene expression in sorghum and is used predominantly with different marker and target genes, although in experiments of Do et al. a higher rate of “escaped” (non-transgenic) plants in comparison with the MAS (mannopine synthase) promoter were observed. It should be noted also that in comparison to gene expression in wheat, the *ubi1* promoter directs very low levels of transgene expression in sorghum. CaMV35S promoter is also used in a number of works, although its efficiency varies at different developmental stages of *T. turgidum* plants being low in young tissues and increased in old tissues.

In addition, a lot of other factors were reported to affect successful genetic transformation of sorghum using *A. tumefaciens*. Among them are addition of poly (vinylpyrrolidone) (PVP) or anti-oxidants (L-cystein or ascorbic acid) into the co-cultivation medium to reduce toxic pigment production by sorghum tissues damaged by agrobacterial cells that is used in numerous reports; cold pretreatment of immature seeds used for isolation of embryos; heat pretreatment of immature embryos; addition of surfactants like Pluronic F-68 or Silwet-701 facilitating transfer of T-DNA in cultivated cells; growing conditions of plants-donors of immature embryos and agrobacterial cells, activation of *vir*-genes, and some others.

Such a variety of factors significantly complicates the production of transgenic plants by *Agrobacterium*-mediated genetic transformation in sorghum tissue culture. A high frequency of regeneration of transgenic plants is observed only with the use of *A. tumefaciens* strains harboring specially designed “superbinary” vectors with additional *vir*-genes. With the use of standard binary vectors, the frequency of transgenic plants reaches 5-8% only in individual experiments, and in most experiments it is much lower. In addition, the introduced transgenes are often subjected to silencing, which makes difficult selection of the transformants and appears to be a major obstacle in the transformation of sorghum.
Recently, it has been demonstrated that the use of vectors containing the genes of the morphogenetic regulators Baby boom (Bbm) and Waschel(Was) significantly increases the frequency of transgenic plants in a number of cereal species, including sorghum.\(^{45,46}\) Moreover, the inclusion in the genetic construct the Cre recombinase gene under the control of a promoter sensitive to desiccation stress in tissue culture and lastP sites flanking the Bbm and Was genes makes it possible to produce transgenic plants bearing only the target gene and not containing the Bbm, Was and Cre. This finding opens wide prospects for the genetic transformation of sorghum in a variety of ways, including improving the nutritional value of this crop.

To overcome the problems associated with agrobacterial transformation under in vitro culture conditions, we proposed a method for producing transgenic sorghum plants in planta by inoculating flowering panicles with a suspension of agrobacterial cells with activated vir-genes.\(^{45}\) In that work we used plants of CMS-line which were pollinated by the pollen of its fertile counterpart and then were inoculated by agrobacterial cells. Using this method, we obtained transgenic plants which inherited transgene (nptII) in T\(_1\) generation. An important point in this method is the need for observing the temperature regime: the plants should be incubated after inoculation under temperature conditions optimal for agrobacterial transformation, i.e. at 23–25°C. Although, later on, we applied this method for fertile sorghum lines and also obtained transgenic plants, which inherited nptII up to T\(_5\) generation, this method requires additional elaboration because of poor reproducibility of the results. Recently, Visarada et al.\(^{46}\) modified procedure for inoculating flowering panicles and obtained transgenic sorghum plants, which inherited transgene (cry1Aa) up to T\(_5\) generation. Yelissety et al.\(^{47}\) suggested another in planta method for genetic transformation of sorghum, by inoculation of the meristematic region of 2-days old germinating seedlings with \(A.\) \textit{tumefaciens} cell suspension. Transformation efficiencies ranged between 34–38% and 26–34% for different vectors and genetic constructs. This method seems to be quite promising; nevertheless it needs additional verification and replication in other laboratories.

**Use of RNA-silencing technology to suppress the synthesis of kafirins**

In recent years, RNA interference technology has been intensively used to suppress the synthesis of seed storage proteins in different species of cereals including wheat,\(^{30-33}\) rice\(^{34-38}\) and maize.\(^{39,40}\) These experiments contributed to obtaining new information on the mechanisms of protein body formation, as well as the role of various classes of prolamins and glutelins in the development of endosperm and the technological properties of flour and dough. For example, in maize, use of genetic constructs containing inverted repeats of α-zein genes (19kDa and 22kDa), allowed to obtain transgenic lines with suppressed synthesis of these proteins.\(^{39,40}\) It has been found that suppression of the synthesis of zeins having a relatively low nutritional value, leads to the accumulation of other proteins with a higher nutritional value. Maize plants with silencing of α-zeins were characterized by a doubled content of essential amino acids - tryptophan and lysine - in their kernels. In these experiments, it was found that silencing the 22kDa α-zein gene resulted in the formation of a floury endosperm. Such a change of the endosperm type was associated with a change in the structure of the protein bodies, namely, with a disturbed incorporation of 19kDa α-zein into the center of the protein body or a change in its interaction with β- and γ-zein.

In the QPM (quality protein maize) lines having vitreous endosperm, various mutants with variations in the structure of protein bodies and endosperm types were obtained by introducing the genetic constructs for RNAi silencing of different zein gene.\(^{41,42}\) It has been demonstrated that an important role in the formation of floury endosperm plays silencing of the γ-zein gene, as a result of which the structure of protein bodies and their connection with starch granules changes.\(^{42}\) In a detailed study it was shown that 27-kDa γ-zein causes initiation of protein bodies.\(^{43}\) Other proteins belonging to the γ-zein family function at the stage of proliferation of protein bodies. Suppression of both 19- and 22-kDa α-zeins significantly suppresses the proliferation of protein bodies, but does not cause their morphological anomalies that arise in the case of reduction of only 22kDa α-zein. Simultaneous suppression of all zein classes leads to a decrease in the number of protein bodies, which in this case have a normal size and morphology.

Intensive studies have been undertaken by several research groups on the induction of RNA-silencing of the kafirin genes in sorghum. The main goal of these experiments was the creation of sorghum lines with improved kafirin digestibility. RNA-silencing was induced by genetic constructs carrying inverted repeats of several kafirin genes (α1, α2, γ1, γ2), separated by the sequence of the intron of the \(ADH1\) gene; these constructs were driven by the maize 19-kDa α-zein promoter.\(^{18,64}\) In another study,\(^{19}\) a genetic construct was used to induce the silencing of the \(γ\)-kafirin, which included the complete sequence of the \(γ\)-kafirin gene under the control of the \(γ\)-kafirin promoter. The sequence of the self-cleaving ribozyme gene of the tobacco mosaic virus was used as a terminator, which expression should destroy \(γ\)-kafirin mRNA. To induce the silencing of α-kafirin, a construct consisting of inverted repeats of α-kafirin separated by the sequence of the intron of the arabidopsis gene encoding the D1 splicesosome protein was used; this construct was driven by the α-zein promoter.\(^{19}\) Later, transgenic sorghum lines with another complex RNAi construct which should suppress synthesis of γ1-, γ2-, and δ-zeins were also obtained.\(^{40}\) In the experiments of each of these groups, lines of transgenic sorghum with suppressed synthesis of γ- and/or α-kafirins and a floury type of endosperm were obtained. Unfortunately, the presence of floury endosperm is a disadvantage of these lines, since the absence of a vitreous layer increases the fragility of the grain and reduces their resistance to damage by the fungal diseases.

In our experiments, we obtained transgenic sorghum plants with genetic construct for RNA-silencing of single 28kDa γ-kafirin gene.\(^{43}\) For this purpose, we used specially designed the binary silencing vector, pNRKAFSIL, which contained a hairpin insert that consisted of an inverted repeat of the 307-bp fragment of the γ-kafirin gene (GeneBank accession number MT368887) and \(ubi1\) intron as the spacer between the arms of the inverted repeat. This construct was driven by the CaMV 35S-promoter. The binary vector pNRKAFSIL was introduced in genome of sorghum line cv. Zheltozernoe 10 (Zh10) by A\(\)b\(\)pro\(b\)acteriu\(m\)-mediated genetic transformation of immature embryos with the \(A.\) \textit{tumefaciens} strain GV 3101. The introduced genetic construct caused reduction of level of the 28-kDa γ-kafirin.\(^{44}\) In addition, significant reduction of the kafirin oligomers of 45- and 66-kDa was observed that was expected assuming the role of the γ-kafirin in kafirin polymerization.\(^{5,7,9,10}\) Inheritance of introduced genetic construct was traced up to T\(_5\) generation by PCR with primers both to \(bar\) gene, which was used as a marker in the genetic construct, and with primers to \(Ubi\)-intron.\(^{45}\)

Remarkably, in our experiments transgenic plants had kernels with both floury endosperm and modified endosperm type (Figure 1). In such kernels, the vitreous layer was significantly reduced and developed as sectors or patches surrounded by floury endosperm.
These kernels resemble the kernels of recombinant sorghum lines obtained by hybridization of highly digestible mutant with floury endosperm (*khihi*) with ordinary sorghum lines with vitreous endosperm. Formation of this endosperm type in our transgenic plants apparently reflects peculiarities of expression of inserted genetic construct during kernel development. Previously, patches of vitreous endosperm were observed in transgenic plants of *T* to30 containing genetic construct for silencing of α- and γ-kafirins, while co-suppression of δ-kafirin and γ-kafirins subclasses synthesis did not change endosperm phenotype in this line. In transgenic plants of P989012 line with genetic construct for silencing of α-, γ- and δ-kafirins kernels had floury phenotype.

Transgenic plants obtained in our experiments also had significantly improved *in vitro* digestibility of endosperm storage proteins. Examples of improved *in vitro* protein digestibility are presented in Figures 2 & 3. Comparison of electrophoretic spectra before and after pepsin digestion of proteins of *T*1 plant #94-2 (almost floury endosperm; Figure 2A, lanes 1, 2) with Zh10 kernels (Figure 2A, lanes 5, 6) revealed that in transgenic plant the amount of undigested α-kafirin monomers and total undigested protein was significantly fewer (in 1.7-1.9 times, according to quantitative analysis of SDS-PAGE images) than in original non-transgenic line. The digestibility value reached 85.4%, whereas in original line this value was about 60% (Table 1). Remarkably, in kernels of transgenic plant #94-3-08 (*T*2 generation) with thick vitreous endosperm (Figure 2A, lanes 3, 4), the differences in kafirin digestion were more pronounced: the amount of undigested monomers was 17.5 times fewer, and the amount of total undigested protein was 4.7 times fewer than in original line. The digestibility value reached 92% (Table 1).

Effect of the RNAi constructs on *in vitro* protein digestibility

The most important effect of kafirins silencing is the increase protein digestibility in both cooked and raw sorghum flour. For example, transgenic plants of *T* to430 carrying genetic construct for silencing of several kafirin sub-classes (α- and γ-kafirin silencing) were characterized by improved *in vitro* protein digestibility of both raw and cooked grain flour: 78 and 61%, respectively; whereas in non-transgenic control these figures were 40-50% and 34-40%, respectively. Genetic construct for silencing only δ- and γ-kafirins also caused improvement of *in vitro* protein digestibility of raw flour but did not affected digestibility of cooked flour. In experiments of Kumar et al., cooked flour obtained from transgenic plants carrying genetic constructs for silencing of γ-kafirin did not differ from non-transgenic control, whereas silencing of α-kafirin improved *in vitro* protein digestibility of cooked flour.
Genetic modification of sorghum for improved nutritional value: state of the problem and current approaches

Table 1  In vitro protein digestibility of sorghum flour from kernels of transgenic plants obtained by genetic transformation with A. tumefaciens GV3101/ pNRKAFSIL.

| Plant                          | Endosperm type                     | Protein digestibility, % |
|-------------------------------|------------------------------------|--------------------------|
| Plants from T1 and T2 families|                                    |                          |
| T1 94-2                       | floury                             | 85.4 c                   |
| T1 94-6                       | floury                             | 85.2 c                   |
| T2 94-2-05                    | modified                           | 74.1 b                   |
| T2 94-2-11                    | floury                             | 90.7 cd                  |
| T2 94-3-08                    | vitreous, irregular                | 92.0 d                   |
| Zheletozernoie 10 (original non-transgenic line) | vitreous                        | 60.4 a                   |
| F                             |                                    | 71.52**                  |
| Plants from T3 families       |                                    |                          |
| T3 94-2-11-2                  | Modified                           | 87.8 b                   |
| T3 94-2-11-3                  | Modified                           | 85.5 b                   |
| T3 94-2-04-2                  | Modified                           | 85.2 b                   |
| T3 94-3-04-1                  | Floury                             | 83.1 b                   |
| T3 94-3-04-1                  | Modified                           | 90.3 c                   |
| T3 94-3-08-2                  | Vitreous, irregular                | 86.2 b                   |
| T3 94-3-08-3                  | Vitreous, irregular                | 88.3 b                   |
| Zheletozernoie 10 (original non-transgenic line) | Vitreous                        | 59.3 a                   |
| F                             |                                    | 68.311**                 |

Each value is a mean from two replications. Data followed by the same letter did not differ significantly (P<0.05) from plant from the same group of families according to Duncan Multiple Range Test. Protein digestibility was calculated as percent ratio of difference between total estimated protein quantity in the control and digested sample to total estimated protein quantity in the control sample.

**Significant at P<0.01.

Plants from T1 generation inherited improved digestibility of kafirins. In these plants, kernels had either floury or modified endosperm (#94-2-04-2; #94-3-04-1) or endosperm with vitreous layer (#94-3-08) (Table 1, Figure 3). Quantitative analysis showed that the level of digestibility of endosperm proteins in these plants was 83-90%, significantly differing from the digestibility of proteins in the original non-transgenic line. One should stress that electrophoretic spectra of digested endosperm proteins of transgenic sorghum plants with the genetic construct for silencing of the γ-kafirin gene are not characteristic of ordinary sorghum cultivars obtained by classical breeding except highly digestible sorghum mutant (hdhl) and its hybrids.69,67 Apparently, a decrease in the level of γ-kafirin increases the digestibility of α-kafirins. This increase may be due to chemical reasons (reduction of polymerization) and/or physical reasons (change in the spatial arrangement of α-kafirins in the protein bodies that increase their availability to pepsin digestion). It should be noted that in vitro protein digestibility of endosperm proteins differ in plants grown in greenhouse in the pots with regular irrigation and in the field plots. When growing under conditions of an experimental field plot kafirin digestibility decreased, in comparison with the previous generations, although it was still significantly higher than the original non-transgenic line.70

Improvement of nutritive value of sorghum grain carrying genetic constructs for RNAi silencing of kafirin genes

An important consequence of the silencing of prolamine genes is the enhancement of the synthesis of other proteins, including those that
have a higher content of essential amino acids, lysine and threonine. Previously, synthesis of such proteins was observed in transgenic plants with silencing of α-zein and α-kafirin. Significant increase of lysine content (up to 3.5g/100g protein, in comparison with 2.1g/100g protein in non-transgenic control) was found in transgenic sorghum plants carrying complex genetic constructs for RNAi silencing of α-, γ-, δ-kafirin genes and lysine-ketoglutarate reductase gene, which control catabolism of free lysine. Maize plants with silencing of a-zeins were characterized by a doubled content in grains of essential amino acids - tryptophan and lysine. In rice, it has been shown that gene silencing of a 13kDa prolamine increases the total lysine content up to 56% as a result of a compensatory rise in the synthesis of lysine-rich glutelins, globulins and chaperones.

In our studies, in the kernels of transgenic plants from the T2 generation with high in vitro protein digestibility, both with modified endosperm and with vitreous endosperm, total amino acid content was significantly reduced for 22.8-40.2%, in comparison with the original non-transgenic line. At the same time, the relative content of two major essential amino acids, lysine and threonine, significantly increased. Lysine proportion increased by 1.6-7.1times: from 1.54% of total amino acid content in the flour of original non-transgenic line to 2.41-2.63% in transgenic plants. Such increase of the relative content of lysine and threonine in transgenic sorghum plants, coupled with a significant reduction of the total level of amino acids, presumably was caused by decrease in the content of α-kafirins poor in lysine and threonine, whereas the synthesis of other proteins remained undisturbed. Accordingly, the relative proportions of lysine and threonine increased. The lower α-kafirins content in transgenic plants relative to the original non-transgenic line is clearly evident in the above electrophoresis photographs (Figure 2 & Figure 3). Perhaps the suppression of the synthesis of γ-kafirin prevents the accumulation of α-kafirins, but does not affect the synthesis of other proteins richer in lysine and threonine.

Such a rebalancing of seed proteome is a common phenomenon for transgenic plants with genetic constructs for RNA-silencing of major seed storage proteins. For maize, it was suggested that developing kernels possess compensatory mechanisms that sense protein content when zein synthesis is interrupted, leading to translation of other mRNAs instead of zein mRNAs. This involves an intracellular signal transduction to attain a predetermined protein level. Remarkably, in knockdown mutant lines of soybean with suppressed synthesis of major storage proteins, the seeds maintained nearly identical levels of total protein compared to the untransformed soybean cultivars. These data suggest that proteome rebalancing in seeds might be a rather common event, providing a constant sink for reduced nitrogen during seed maturation.

It should be noted that the structure of protein bodies, which accumulate prolams in cereals, has changed in transgenic lines with the silencing of α-kafirin. In the endosperm of plants of common sorghum lines, the protein bodies have a regular rounded shape, while in the transgenic lines with α-kafirin silencing the protein bodies acquire an irregular shape with invaginations as it was found in the highly digestible sorghum mutant (Khth). Such protein bodies, γ-kafirin does not form a continuous surface layer. It is believed that such structure of protein bodies increases the contact of the more easily digested α-kafirin with proteases, which contributes to its higher digestibility. At the same time, when the γ-kafirin was silenced, no changes in the shape of protein bodies were observed. However, later study showed that in transgenic lines with γ-kafirin silencing diameter of protein bodies is reduced in comparison with original non-transgenic line. In addition, in one of the studied lines, 42-1, protein bodies were highly irregular in shape, with deep invaginations present at the periphery, while in the line 42-2, the protein bodies had small peripheral indentations that gave the boundary region a cracked appearance.

**Editing of nucleotide sequences of kafirin genes**

The above examples of suppressing the synthesis of kafirins through the introduction of genetic constructs for RNA-silencing is performed by small interfering RNAs (siRNAs) and subsequent mRNA cleavage and translational inhibition, which are important epigenetic mechanisms in plants. However, epigenetic mechanisms in plants are known to be sensitive to growing conditions, to environmental factors (temperature, soil moisture and air, etc.). There are reports that temperature can have a major impact on the extent of gene silencing. Small interfering RNAs (siRNAs) and microRNAs (miRNAs) are key regulators of posttranscriptional gene silencing that includes mRNA cleavage and translational inhibition. Recently it has been shown that mRNA-mediated mRNA cleavage and translational inhibition are dependent on plant growth temperature. Therefore, the effectiveness of suppression of kafirin synthesis by RNA-silencing constructs might be sensitive to environmental conditions. Indeed, in our experiments, in vitro protein digestibility in transgenic plants with genetic construct for γ-kafirin silencing grown in the field plots was substantially lower than in the plants grown in the pots in greenhouse. In this regard, induction of mutations that disrupt functioning of the kafirin genes at the genetic level may be a more effective approach for altering the level of kafirins.

Recently, using CRISPR/Cas9 gene editing approach Li and co-workers induced mutations in a conserved region encoding the endoplasmic reticulum signal peptide of α-kafirins. The target site mutations occurring in each kIC gene include small deletions ranging in length between 1 and 33 bp, as well as a low frequency of small insertions of 1 to 16bp. T1 and T2 seeds showed reduced α-kafirin levels, altered protein body microstructure, and selected T1 events showed significantly increased grain protein digestibility and lysine content. This work opens new perspectives in improvement of nutritional value of sorghum grain using biotechnological approaches.

**Sorghum biofortification with amino acids and micronutrients**

Numerous researches published up to date clearly demonstrate that genetic engineering techniques are quite promising for enrichment of cereal grain with essential amino acids, i.e. lysine, tryptophan, methionine, and micronutrients. By using endosperm-specific promoters, expression of the desired genes may be provided exactly in the kernels, rather than in somatic tissues where the increased content of these amino acids may lead to developmental abnormalities. The gene encoding high lysine analog (HT12 protein) of Hordeum vulgare α-hordothionin protein under the control of the 27kDa maize γ-zein promoter and terminator was introduced in genome of two sorghum lines, P898012 and PHI391, by using Agrobacterium-mediated genetic transformation. The A. tumefaciens strain (LBA4404) contained “super-binary” vector with two unlinked T-DNA cassettes: one cassette contained the lysine-rich HT12 gene and another contained a herbicide-resistant bar gene as a selectable marker, under the control of maize ubiquitin promoter. This co-transformation vector with two
Genetic modification of sorghum for improved nutritional value: state of the problem and current approaches

Sorghum contains very low levels of β-carotene (provitamin A), which is highly important for human health. In order to increase β-carotene content in sorghum grain and ensure its stability during grain storage, the work was carried out on the introduction into the genome of sorghum line (Tx430) the genetic constructs encoding a number of enzymes involved in the carotenoid biosynthesis pathway. These enzymes are: 1-deoxyxylulose 5-phosphate synthase (DXS), the precursor for carotenoid biosynthesis, Zea mays phytene synthase 1 (PSY1) and the Pantoea ananatis carotene desaturase (CRTI), involved in synthesis of phytene and lycopene, respectively, which are the β-carotene precursors (Figure 4). These gene significantly increased β-carotene level in mature seeds of transgenic plants (up to 9.1μg/g vs. 0.5 μg/g in non-transgenic control seeds), although did not prevent its degradation during storage as a consequence of its oxidation. To prevent in planta degradation of β-carotene, the barley HGGT gene encoding homogentisate geranylgeranyl transferase, involved in synthesis of the vitamin E, which has strong antioxidant effect, was introduced into the same genetic construct. It was found that that co-expression of HGGT, stacked with carotenoid biosynthesis genes, enhanced all-trans β-carotene accumulation in the range of 7.3–12.3μg/g and mitigate β-carotene oxidative degradation, resulting in enhanced stability of provitamin A in sorghum seeds. This research is a shining example of the effectiveness of the genetic engineering approach for modifying plant metabolism to meet human needs. It is able to make an impact on millions of people who suffer from vitamin A deficiency.

**Figure 4** Highlights of the enzymes used for sorghum biofortification in the carotenoid biosynthesis pathway.74

Sorghum grain as well as seeds of other cereal store phosphorus in the form of phytic acid, which constitutes about 65–90% of the total phosphorous content in plants. The negatively charged phosphate of phytic acid has strong affinity to chelate divalent ions e.g. calcium, magnesium, and especially with iron and zinc making them insoluble and thus unavailable as nutritional factors. Therefore, it is extremely poor as a dietary source of phosphorous. To enhance bio-availability of micronutrients, an enzyme namely phytase is known to hydrolyze phytic acid. Unfortunately, phytase is not produced in the stomach of monogastric animals and humans. Thus, the presence of phytic acid in cereal foods has become major concern about the deficiency of essential micronutrients.79 This problem may be solved by using transgenic approaches. At present time genetically engineered plants with enhance phytase production in their grains obtained in a number of crops including maize, rice and wheat.80 In sorghum, genetically modified low phytate plants were obtained in the line Tx430. In these plants, the Multidrug Resistance-Associated Protein ATP-binding cassette transporter was silenced that resulted in lower phytate content (3.24–4.91mg/100 g, for 80–86%) compared to their respective null-control plants and increased zinc and iron availability.81

**Conclusion**

Intensive development of methods of genetic transformation allowed creating transgenic sorghum lines carrying specially designed genetic constructs, purposely suppressing the synthesis of endosperm storage proteins - kafirins. Transgenic lines with suppressed kafirin synthesis are characterized by increased digestibility of endosperm proteins and increased content of lysine, and, thus, come close to solving the problem of improving the nutritional value of the grain of this crop. The kernels of transgenic lines with α-kafirin silencing had soft floury endosperm type that prevents their direct commercial use; however these lines may be used as donors of this trait in crosses with sorghum lines adopted to local environments for improvement of their nutritive value. The kernels of transgenic lines with γ-kafirin silencing may have hard vitreous endosperm or modified endosperm having sectors of vitreous endosperm. This fact indicates the possibility of obtaining agronomically valuable sorghum lines with high digestibility of kafirins and hard endosperm, although the lines with γ-kafirin silencing have lower level of in vitro protein digestibility than the lines with α-kafirin silencing. No significant changes in anti-nutritional factors (tannin and phytate content) were detected in transgenic lines with γ-kafirin silencing.84 In addition to the improvement of sorghum protein digestibility, the genetic engineering approaches allowed to increase essential amino acid content, particular lysine, by introducing the genes encoding high-lysine protein, and increase the level of A-provitamin in sorghum grain and bioavailability of micronutrients. However, strict regulations that govern practical usage of these innovations prevent their entering in the market. For until the world regulatory agencies establish guidelines that are science based, that focus on the trait, not the process by which that trait is generated, most valuable technologies will never see the light of day.

**Acknowledgements**

The work has been partially supported by Russian Foundation for Basic Research, project no 16-04-01131.

**Conflict of interest**

Authors declare that there is no conflict of interest.

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