Accounting for Time: Circadian Rhythms in the Time of COVID-19

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Abstract The COVID-19 pandemic has necessitated novel approaches and collaborative efforts across multiple disciplines. It is known that various aspects of our physiology and response to pathogens are under tight clock control. However, the assimilation of circadian biology into our clinical and research practices is still evolving. Using a focused review of the literature and original analyses of the UK Biobank, we discuss how circadian biology may inform our diagnostic and therapeutic strategies in this pandemic.

Keywords vaccines, translational medicine, chronotherapy, COVID-19, circadian, immunity

The coronavirus disease 2019 (COVID-19) epidemic, which as caused by severe acute respiratory coronavirus-2 (SARS-CoV2), has prompted a scramble for safe and effective treatments. Our focus remains on vaccine development and repurposing approved drugs. However, reviewing our current understanding of chronobiology as applicable to other viral pathogens may help us realize opportunities to harness the power of circadian biology for SARS-CoV-2.

Circadian rhythms are evolutionarily conserved pathways that serve as a means for living organisms to adapt to the environment. It is an anticipatory system, preparing us to deal with challenges to our resilience, including infections. Molecular clockworks are highly conserved, tightly regulated, and exhibit a high degree of redundancy (Takahashi, 2017). Directed by a master clock in the suprachiasmatic nucleus, peripheral tissue clocks are among the most prominent biological networks that integrate cross-tissue functions, such as are intrinsic to cardiovascular homeostasis, metabolism, and the immune response (Yang et al., 2013).

The clock may be relevant to the pathophysiology and treatment of COVID-19 in the following ways:

1. The time of infection relative to the circadian day: The molecular clock determines the response to infection via a wide range of pathogens in many species because of circadian control of the immune system (Curtis et al., 2014; Man et al., 2016; Zhuang et al., 2019; Stone et al., 2012; Ehlers et al., 2018). For example, we have demonstrated that mice infected with influenza virus at dusk have a 3-fold higher mortality than those infected at dawn. This did not result from attenuation of the viral burden but rather from modulation of the immune response to infection. This circadian variation in lethality was abolished when the clock was broken in immune cells and...
pulmonary epithelial cells by deletion of the nonredundant core clock gene, Bmal1 (Sengupta et al., 2019). Consistent with this observation, we discovered an interesting association between circadian rhythms and the need for hospitalization for influenza. Approximately 100,000 participants in the UK Biobank have activity measurement for 1 week using a wrist-worn accelerometer device (Doherty et al., 2017). Activity was classified using a machine-learning algorithm (Willetts et al., 2018) and summarized to participant-level measures of sleep, activity, and rhythmic patterns. We found that having a robust circadian amplitude in activity levels was associated with less likelihood of having been treated for influenza (false-discovery rate–corrected q value $<2 \times 10^{-26}$) and other lower respiratory infections, as well as other viral and bacterial infections (q value $<10^{-9}$; Fig. 1). Might the time of infection contribute to the diversity of clinical response to SARS-CoV-2? The entry of SAR-CoV2 virus depends on the cellular receptor angiotensin converting enzyme 2 (ACE2; Hoffmann et al., 2020), a common target for antihypertensive therapy. ACE2 itself is a negative regulator of the renin-angiotensin-aldosterone system (RAAS), which is known to oscillate under circadian control (Ohashi et al. (Ohashi et al., 2017; Nonaka et al., 2001). There are no published reports of a circadian pattern of expression of ACE2 protein or mRNA. However, given its close connection to the oscillatory RAAS system, it is still plausible that ACE2 expression is affected by circadian rhythms, thus directly influencing cellular access by SARS-CoV-2.

As we reopen institutions and design schedules and workspaces, we could test whether time of day influences the susceptibility to or severity of COVID-19 infection. Historically, shift workers have been known to have increased susceptibility to many diseases (Smith and Eastman, 2012) and demonstrate differences in their immunological profile modified by their chronotype and the time from last night shift (Loef et al., 2019). However, in the context of the pandemic, the central question is whether the risk of infection among shift workers is influenced by the time of their shift. This is applicable to all shift workers, including health care workers, but it has particular relevance to the clustered outbreaks in workplaces where social distancing has been a challenge, such as the meat-processing industry (Waltenberg et al., 2020). In many ways, the COVID-19 pandemic presents us with a natural experiment. Thus, an intriguing possibility would be to leverage the many well-designed contact-tracing initiatives to address the time-of-day–specific susceptibility of various populations.

2. Circadian variation in the immune system across the day: Control of the immune system by the molecular clock includes regulation of inflammatory cytokines (Gibbs et al., 2012; Early et al., 2018; Keller et al., 2009), neutrophil maturation (Adrover et al., 2019), myeloid and lymphoid cell trafficking (Druzd et al., 2017), and migration as well as development and differentiation of immune cells, such as Th17 cells (Yu et al., 2013). Similarly, various aspects of human physiology have been shown to oscillate across the circadian day (Skarke et al.,

Figure 1. The top 10 infection phenotypes associated with relative circadian amplitude among participants of the UK Biobank, by $-\log_{10}$ of the Benjamini-Hochberg q-values. Cases were extracted by lifetime occurrence of ICD-10 codes and associations computed by a linear model controlling for age, sex, socioeconomic status, body mass index, smoking status, and self-reported overall health. Relative amplitude was computed as a ratio (M10-L5/M10+L5) where M10 and L5 are the participant's average activity levels of the ten hours of highest activity and five hours of the lowest activity, respectively (Witting et al., 1990).
Thus, although there is evidence that the various components of the immune system oscillate in a circadian manner, this is not usually factored into sample collection for immunophenotyping (Mathew et al., 2020). Sample collection is typically driven by the availability of patient and staff and/or the relation to drugs already administered, and it is agonistic to timing. Given the magnitude of oscillations in immune cell number and function in model systems and humans, time stamping such samples to account for this variable can limit the source of biological noise, refine data interpretation, and reduce the required sample size for studies of COVID-19.

3. Circadian biology and time of vaccination: There is evidence from animal studies that both the innate immune system (Silver et al., 2012) and the adaptive system (Suzuki et al., 2016), such as the CD8 cell-intrinsic clock (Nobis et al., 2019), influence the immune response to vaccination in model systems. In a cluster randomized design study in humans, morning vaccination with the inactivated influenza vaccine resulted in higher antibody responses than afternoon vaccination in adults older than 65 years (Long et al., 2016). However, this may be confounded by factors such as vaccination history (Cobey and Hensley, 2017). In the case of a novel pathogen such as SARS-CoV-2, there is the opportunity to perform a controlled comparison of morning versus evening vaccination to determine if time conditions the immune response. Another related factor that has been shown to affect vaccine response, at least for influenza immunization, is sleep deprivation (Benedict et al., 2012; Brown et al., 1989; Spiegel et al., 2002). Further, we speculate that chronotype, which refers to the endogenous propensity of an individual to sleep during particular periods of the day, contributes to the heterogeneity of response to vaccinations. While there are no studies that clearly implicate chronotype in determining vaccine response, the effect of the time of day on vaccination response would likely be modified by the endogenous chronotype. Thus, although the approach to leveraging the principles of chronobiology in improving the efficacy of the vaccine response against SARS-CoV2 is multilayered, simply time stamping vaccination across the day would allow us to rapidly scale up to define the impact of time in observational studies. An additional consideration is that circadian rhythms tend to deconsolidate in the elderly, so the likelihood of a time-dependent response to vaccination may be conditioned by the age of the patient.

Further, these differences in the immune system based on time of day should be considered when working on animal models of COVID-19. That there are differences between the mouse and man, in terms of immune architecture and response, is well documented (Mestas and Hughes, 2004; Shay et al., 2013). However, specifically relevant here is the fact that mice are nocturnal and humans are not. Promising results from mice containing a humanized ACE2 (Hassan et al., 2020) and other models need to be interpreted in the light of the nocturnal habitus of mice. Appreciation of the differences in their circadian rhythms may improve the reproducibility of data and directly affect the translation of findings from animal to human studies.

4. Using circadian rhythms to optimize the effect of antivirals and anti-inflammatory agents: Just like the immune system, many drug targets, transporters, and metabolizing enzymes are subject to circadian variation under the control of the clock (Zhang et al., 2014). The response to many drugs, particularly those with shorter half-lives, is greatly influenced by time of dosing (Sulli et al., 2018). While targeting drugs to a particular time of day to enhance efficacy and reduce adverse effects is common practice for some medications, it is still not practiced for all drugs with shorter half-lives (Erkekoglu and Baydar, 2012). Similarly, chronotherapy may be relevant to antivirals such as remdesivir and anti-inflammatory drugs such as aspirin (Chen et al., 2018; Bonten et al., 2015; Buurma et al., 2019), which are being used to treat COVID-19, but might also be relevant to antithrombotic medications used to treat COVID-19 complications. Finally, clock genes (Kojetin and Burris, 2014) and melatonin (Farias et al., 2019; Prado et al., 2018) have been shown to have anti-inflammatory effects in other models, and the use of melatonin to reentrain the clock in cases of lifestyle-related misalignment may also be considered to reduce the risk of infection.

In conclusion, our molecular clocks may modulate viral cellular entry and the consequent immunological response to infection. Timing of administration may influence the response to administration of a vaccine or to the drugs used to attenuate the severity of COVID-19. The various intersections of the circadian clocks and immunobiology as well as the epidemiology of COVID-19 (Fig. 2) constitute biologically plausible hypotheses that are readily testable.
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REFERENCES

Adrover JM, Del Fresno C, Crainiciuc G, Cuartero MI, Casanova-Acebes M, Weiss LA, Huerga-Encabo H, Silvestre-Roig C, Rossaint J, Cossio I, et al. A neutrophil timer coordinates immune defense and vascular protection. Immunity 2019;50:390-402 e10.

Benedict C, Bryting M, Markstrom A, Broman JE, and Schioth HB. Acute sleep deprivation has no lasting effects on the human antibody titer response following a novel influenza A H1N1 virus vaccination. BMC Immunol 2012;13:1.

Bonten TN, Snoep JD, Assendelft WJ, Zwaginga JJ, Eikenboom J, Huisman MV, Rosendaal FR, and van der Bom JG. Time-dependent effects of aspirin on blood pressure and morning platelet reactivity: a randomized cross-over trial. Hypertension 2015;65:743-750.

Brown R, Fang G, Husband AJ, and King MG. Suppression of immunity to influenza virus infection in the respiratory tract following sleep disturbance. Reg Immunol 1989;2:321-325.

Buurma M, van Diemen JJK, Thijs A, Numans ME, and Bonten TN. Circadian rhythm of cardiovascular disease: the potential of chronotherapy with aspirin. Front Cardiovasc Med 2019;6:84.

Cobey S and Hensley SE. Immune history and influenza virus susceptibility. Curr Opin Virol 2017;22:105-111.

Curtis AM, Bellet MM, Sassone-Corsi P, and O’Neill LA. Circadian clock proteins and immunity. Immunity 2014;40:178-86.

Doherty A, Jackson D, Hammerla N, Plötz T, Olivier P, Granat MH, White T, van Hees VT, Trenell MI, Owen CG, et al. Large scale population assessment of physical activity using wrist worn accelerometers: the UK Biobank study. PLoS One 2017;12:e0169649.

Druzd D, Matveeva O, Ince L, Harrison U, He W, Schmal C, Herzl H, Tsang AH, Kawakami N, Leliavski A, et al. Lymphocyte circadian clocks control lymph node trafficking and adaptive immune responses. Immunity 2017;46:120-132.

Early JO, Menon D, Wyse CA, Cervantes-Silva MP, Zaslona Z, Carroll RG, Palsson-McDermott EM, Angiari S, Ryan DG, Corcoran SE, et al. Circadian clock protein BMAL1 regulates IL-1beta in macrophages via NRF2. Proc Natl Acad Sci USA 2018;115:E8460-E846E8.

Ehlers A, Xie W, Agapov E, Brown S, Steinberg D, Tidwell R, Sañol G, Schutz R, Weaver R, Yu H, et al. BMAL1 links the circadian clock to viral airway pathology and asthma phenotypes. Mucosal Immunol 2018;11:97-111.

Erkekoglu P and Baydar T. Chronopharmacodynamics of drugs in toxicological aspects: a short review for clinical pharmacists and pharmacy practitioners. J Res Pharm Pract 2012;1:41-47.

Farias T, Paixao RID, Cruz MM, Cavalcante da Cunha de Sa RD, de Jesus Simão J, Antraco VJ, and Cardoso Alonso-Vale MI. Melatonin supplementation attenuates the pro-inflammatory adipokines expression in visceral fat from obese mice induced by a high-fat diet. Cells 2019;8.

Gibbs JE, Blaikley J, Beesley S, Matthews L, Simpson KD, Boyce SH, Farrow SN, Else KJ, Singh D, Ray DW, et al. The nuclear receptor REV-ERBalpha mediates circadian regulation of innate immunity through selective regulation of inflammatory cytokines. Proc Natl Acad Sci USA 2012;109:582-587.

Hassan AO, Case JB, Winkler ES, Thackray LB, Kafai NM, Bailey AL, McCune BT, Fox JM, Chen RE, Alsoussi
WB, et al. A SARS-CoV-2 infection model in mice demonstrates protection by neutralizing antibodies. Cell 2020;182:744-753.e4.

Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, Schiergens TS, Herrler G, Wu N-H, Nitsche A, Müller MA, et al. SARS-CoV-2 cell entry depends on ACE2 and TMPRSS2 and is blocked by a clinically proven protease inhibitor. Cell 2020;181:271-280 e8.

Keller M, Mazuch J, Abraham U, Eom GD, Herzog ED, Volk H-D, Kramer A, and Maier B. A circadian clock in macrophages controls inflammatory immune responses. Proc Natl Acad Sci USA 2009;106:21407-21412.

Kojentin DJ and Burris TP. REV-ERB and ROR nuclear receptors as drug targets. Nat Rev Drug Discov 2014;13:197-216.

Loef B, Nanlohy NM, Jacobi RHH, van de Ven C, Mariman R, van der Beek AJ, Proper KI, van Baarle D. Immunological effects of shift work in healthcare workers. Sci Rep 2019;9:18220.

Long JE, Drayson MT, Taylor AE, Toellner KM, Lord JM, and Phillips AC. Morning vaccination enhances antibody response over afternoon vaccination: a cluster-randomised trial. Vaccine 2016;34:2679-2685.

Man K, Loudon A, and Chawla A. Immunity around the clock. Science 2016;354:999-1003.

Mathew D, Giles JR, Baxter AE, Greenplate AR Wu JE, Alanio C, Oldridge DA, Kuri-Cervantes L, Pampena MB, D’Andrea K, et al. Deep immune profiling of COVID-19 patients reveals patient heterogeneity and distinct immunotypes with implications for therapeutic interventions. bioRxiv 2020. doi:10.1101/2020.05.20.106401

Mestas J and Hughes CC. Of mice and not men: differences between mouse and human immunology. J Immunol 2004;172:2731-2738.

Nobis CC, Dubeau Laramee G, Kervezee L, Maurice De Sousa D, Labrecque N, and Cermakian N. The circadian clock of C58 T cells modulates their early response to vaccination and the rhythmicity of related signaling pathways. Proc Natl Acad Sci USA 2019;116:20077-20086.

Nonaka H, Emoto N, Ikeda K, Fukuya H, Rohman MS, Raharjo SB, Yagita K, Okamura H, and Yokoyama M. Angiotensin II induces circadian gene expression of clock genes in cultured vascular smooth muscle cells. Circulation 2001;104:1746-1748.

Ohashi N, Isobe S, Ishigaki S, and Yasuda H. Circadian rhythm of blood pressure and the renin-angiotensin system in the kidney. Hypertens Res 2017;40:413-22.

Prado NJ, Ferder L, Manucha W, and Diez ER. Anti-inflammatory effects of melatonin in obesity and hypertension. Curr Hypertens Rep 2018;20:45.

Sengupta S, Tang SY, Devine JC, Anderson ST, Nayak S, Zhang SL, Valenzuela A, Fisher DG, Grant GR, López CB, et al. Circadian control of lung inflammation in influenza infection. Nat Commun 2019;10:4107.

Shay T, Jovic V, Zuzk O, Rothamel K, Puyraimond-Zemmour D, Feng T, Wakamatsu E, Benoist C, Koller D, and Regev A ImmGen Consortium. Conservation and divergence in the transcriptional programs of the human and mouse immune systems. Proc Natl Acad Sci USA 2013;110:2946-2951.

Silver AC, Arjona A, Walker WE, and Fikrig E. The circadian clock controls toll-like receptor 9-mediated innate and adaptive immunity. Immunity 2012;36:251-261.

Skarke C, Lahens NF, Rhoades SD, Campbell A, Bittinger K, Bailey A, Hoffmann C, Olson RS, Chen L, Yang G, et al. A pilot characterization of the human circadian clock. Proc Natl Acad Sci USA 2017;114:17141.

Smith MR and Eastman CI. Shift work: health, performance and safety problems, traditional countermeasures, and innovative management strategies to reduce circadian misalignment. Nat Sci Sleep 2012;4:111-32.

Spiegel K, Sheridan JF, and Van Cauter E. Effect of sleep deprivation on response to immunization. JAMA 2002;288:1471-1472.

Stone EF, Fulton BO, Ayres JS, Pham LN, Ziauddin J, and Shirasu-Hiza MM. The circadian clock protein timeless regulates phagocytosis of bacteria in Drosophila. PLoS Pathog 2012;8:e1002445.

Sulli G, Manoogian ENC, Taub PR, and Panda S. Training the circadian clock, clocking the drugs, and dragging the clock to prevent, manage, and treat chronic diseases. Trends Pharmacol Sci 2018;39:812-827.

Suzuki K, Hayano Y, Nakai A, Furuta F, and Noda M. Adrenergic control of the adaptive immune response by diurnal lymphocyte recirculation through lymph nodes. J Exp Med 2016;213:2567-2574.

Takahashi JS. Transcriptional architecture of the mammalian circadian clock. Nat Rev Genet 2017;18:164-179.

Waltenburg MA, Victoroff T, Rose CE, Butterfield M, Jervis RH, Fedak KM, Gabel JA, Feldpausch A, Dunne EM, Austin C, et al. Update: COVID-19 among workers in meat and poultry processing facilities—United States, April–May 2020. MMWR Morb Mortal Wkly Rep 2020;69:887-892.

Willetts M, Hollowell S, Aslett L, Holmes C, and Doherty A. Statistical machine learning of sleep and physical activity phenotype data from sensor data in 96,220 UK Biobank participants. Sci Rep 2018;8:7961.

Witting W, Kwa IH, Eikelenboom P, Mirmiran M and Swaab DF. Alterations in the circadian rest-activity rhythm in aging and Alzheimer’s disease. Biol Psychiatry 2004;27:563-572.

Yang G, Paschos G, Curtis AM, Musiek ES, McLoughlin SC, and FitzGerald GA. Knitting up the raveled sleave of care. Sci Transl Med 2013;5:212rv3.

Yu X, Rollins D, Ruhn KA, Stubblefield JJ, Green CB, Kashiwada M, Rothman PB, Takahashi JS, and Hooper LV. TH17 cell differentiation is regulated by the circadian clock. Science 2013;342:727-730.

Zhang R, Lahens NF, Ballance HL, Hughes ME, and Hogeness JB. A circadian gene expression atlas in mammals: implications for biology and medicine. Proc Natl Acad Sci USA 2014;111:16219-16224.

Zhuang X, Magri A, Hill M, Lai AG, Kumar A, Rambhatla SB, Donald CL, Lopez-Clavijo AF, Rudge S, Pinglick K, et al. The circadian clock components BMAL1 and REV-ERBa regulate flavivirus replication. Nat Commun 2019;10:377.