Immunotherapy for Diffuse Large B-Cell Lymphoma: Current Landscape and Future Directions

Dipenkumar Modi 1,*, Bindu Potugari 2 and Joseph Uberti 1

1 Department of Oncology, Barbara Ann Karmanos Cancer Institute, Wayne State University, 4100 John R, HW04HO, Detroit, MI 48201, USA; ubertij@karmanos.org
2 Department of Hematology and Oncology, St Joseph Mercy Health System, Ann Arbor, MI 48201, USA; Bindu.Potugari@stjoeshealth.org
* Correspondence: modid@karmanos.org; Tel.: +1-313-576-8782

Simple Summary: Immunotherapy has played a pivotal role in the management of relapsed DLBCL. Stem cell transplant and CAR T-cell therapy are curative treatment modalities for relapsed disease. Despite this, a subset of patients continues to progress, and their outcomes remain dismal. Newer therapeutic options to optimize outcomes as well as minimize toxicity are warranted.

Abstract: Diffuse large B-cell lymphoma (DLBCL) is a heterogeneous disease. B-cell receptor (BCR) pathway is essential for malignant B-cell growth, survival, and proliferation. Various immune cells, including T-cells and macrophages in the tumor microenvironment (TME) contribute to tumor cell survival and pathogenesis of chemo-resistance. The presence of many targets on the malignant B-cells and in the TME has led to emergence of novel therapeutic agents. Stem cell transplant is the oldest treatment modality leveraging immune system in DLBCL. Subsequently, CD20 targeting monoclonal antibody and chimeric antigen receptor (CAR) T-cell therapy changed the treatment landscape of DLBCL. Recently, multiple novel immunotherapeutic agents have been added in the armamentarium for the management of DLBCL, and many are under development. In this review article, we will review latest updates of immunotherapeutic agents in the management of DLBCL.

Keywords: diffuse large B-cell lymphoma (DLBCL); autologous stem cell transplant (autoSCT); allogeneic stem cell transplant (alloSCT); CAR T-cell therapy; bispecific T-cell engager antibody; immunotherapy; nivolumab; pembrolizumab; checkpoint inhibitors

1. Introduction

Diffuse large B-cell lymphoma (DLBCL) is the most common type of non-Hodgkin’s lymphoma (NHL), representing approximately 30–40% of new cases each year. Using gene expression profiling (GEP), it can be classified into two subtypes according to the cell of origin (COO): germinal center B-cell (GCB) and non-germinal center B-cell (non-GCB) [1]. GCB and non-GCB subtypes account for 40% and 60% of de-novo DLBCL, respectively. Patients with the GCB subtype have better outcomes when compared to those with the non-GCB subtype [1–3]. Recently, a number of molecular abnormalities have identified subsets of DLBCL which carry unfavorable prognoses. MYC translocation is reported in 10–15% of DLBCL. In the CORAL (Collaborative Trial in Relapsed Aggressive Lymphoma) study, relapsed 8q24/MYC positive DLBCL had an inferior 4-year progression-free survival (PFS) (18% vs. 42%) and overall survival (OS) (29% vs. 62%), with salvage chemotherapy followed by autologous stem cell transplant (autoSCT), when compared to MYC negative DLBCL [4]. Double or triple hit lymphoma (DHL/THL) is a subtype of DLBCL which is characterized by MYC and BCL2 and/or BCL6 gene rearrangements by FISH, and represents 6–14% of DLBCL cases [5]. It carries the worst prognosis among all DLBCL cases [6–9]. Even if a DHL is deemed chemo-sensitive to salvage chemotherapy, the 4-year PFS after autoSCT is 28% [10]. Double-expressor lymphomas (DEL: Myc positive ≥40%
and Bcl2 + in >50% by IHC) account for 40% of newly diagnosed DLBCL and 50% of relapsed refractory DLBCL. DELs have a better prognosis than DHL, however, this is worse than the “standard” DLBCL [7,11]. Relapsed chemo-sensitive DEL patients have a 4-year PFS of 48% after autoSCT when compared to 58% of “standard” DLBCL [10]. DELs are associated with the non-GCB subtype, while DHL/THL are typically associated with the GCB subtype. In a study involving 893 patients with de-novo DLBCL, 66% of DEL patients were of the ABC subtype, and 39% of non-DEL patients were of the ABC subtype [12]. Additionally, 27% of GCB–DLBCL cases express a 104-gene double-hit signature (DHITsig), and only half of these patients have concurrent MYC and BCL2 rearrangements. Patients with DHITsig experience inferior outcomes after R-CHOP than those without DHITsig (5 year time to progression, 57% and 81%, respectively). Moreover, these patients carry poor outcomes regardless of their MYC/BCL2 rearrangement status [13].

With the increased availability of deep genomic sequencing, novel, molecularly defined subsets of DLBCL are being identified. Schimtz et al. identified four genetic subtypes of DLBCL using exome and transcriptome sequencing: MCD (co-occurrence of MYD88L265P and CD79B mutations), BN2 (BCL6 fusions and NOTCH2 mutations), N1 (NOTCH1 mutations), and EZB (EZH2 mutations and BCL2 translocations). The BN2 and EZB subtypes were associated with favorable survival, while the MCD and N1 subtypes carry inferior outcomes when treated with chemoimmunotherapy [14]. Similarly, Chapuy et al. classified DLBCL into five different subgroups according to the presence of low-frequency alterations, somatic copy number alterations (SCNA), and structural variants (SV). These included low-risk ABC–DLBCLs of extrafollicular/marginal zone origin, (C1), high-risk GCB–DLBCLs with BCL2 SVs and alterations of PTEN and epigenetic enzymes, (C3), low-risk GCB–DLBCLs with distinct alterations in BCR/PI3K and JAK/STAT, as well as BRAF pathway components and multiple histones, (C4), and a COO-independent group of tumors with a biallelic inactivation of TP53, 9p21.3/CDKN2A and associated genomic instability, (C2) [15]. Further refinement of the genetic subtypes of DLBCL was performed, which included the MCD, N1, A53, BN2, ST2, MYC+, and MYC− subsets [16]. Furthermore, another study demonstrated five distinct subtypes of DLBCL using genomic sequencing: MYD88, BCL2, SOCS1/SGK1, TET2/SGK1, and NOTCH2 [17]. Although our understanding of genetic subtypes and their association with clinical features and treatment outcomes is increasing with time, it is not routinely used clinically, and currently remains under investigation.

2. CD20 Directed Monoclonal Antibody (mAb)

Rituximab is the first immunotherapy used in the treatment of DLBCL. The addition of rituximab to chemotherapy has been shown to be superior in terms of complete remission, as well as event-free, progression-free, and overall survival when compared to chemotherapy alone in newly diagnosed DLBCL patients [18–23]. Obinutuzumab is a type II glycoengineered anti-CD20 mAb, and has a superior antibody-dependent cellular cytotoxicity (ADCC) and phagocytosis than rituximab [24]. A randomized phase III trial, comparing addition of obinutuzumab or rituximab to CHOP chemotherapy in previously untreated advanced DLBCL, did not demonstrate any difference in the progression-free and overall survival [25]. Furthermore, rituximab is typically combined with salvage chemotherapy and other therapeutic regimens for the treatment of relapsed refractory DLBCL [26–29]. We routinely use rituximab in newly diagnosed and relapsed refractory settings.

3. Autologous Stem Cell Transplant

Rituximab plus an anthracycline-based chemo-immunotherapy (R-CHOP) is the standard treatment for newly diagnosed DLBCL, and provides long-term remissions in up to 70% of patients [20,23,30]. Approximately 30% of patients eventually relapse. Salvage chemotherapy followed by autoSCT offers durable remissions in approximately half of patients with relapsed DLBCL. In the pre-rituximab era, the PARMA trial compared autoSCT
with consolidative chemotherapy in 215 chemo-sensitive relapsed NHL patients. After two cycles of salvage DHAP chemotherapy, 109 patients with chemo-sensitive disease were randomized either to autoSCT or to four additional cycles of DHAP consolidative chemotherapy. AutoSCT resulted in superior response rates (84% vs. 44%), event-free survival (EFS) (46% vs. 12%), and OS (53% vs. 32%) [31]. In the post-rituximab era, the CORAL and the NCIC-CTG LY.12 study were performed to evaluate the impact of the incorporation of rituximab in the salvage setting. The CORAL study randomized relapsed refractory CD20 positive DLBCL patients to either the R-ICE or R-DHAP salvage regimens, and responding patients proceeded to autoSCT. Response rates were similar after three cycles of R-ICE and R-DHAP (63.5% vs. 62.8%, respectively), and no significant difference for the three-year EFS or OS was noted between R-ICE and R-DHAP treatments [28]. Similarly, the Canadian study NCIC-CTG LY.12 compared GDP to DHAP in transplant eligible patients with relapsed refractory aggressive NHL, and reported no difference in the overall response rates (ORR) (45.2% vs. 44%, respectively), EFS, and OS following autoSCT [26,27].

The CORAL study randomized patients with relapsed DLBCL to rituximab maintenance or observation alone following autoSCT and did not observe any difference in the EFS, PFS, or OS [28]. The BMT CTN 0401 study substituted rituximab with radioimmunotherapy, as a part of the BEAM conditioning regimen, and did not observe an improvement in outcomes [32]. The Center for International Blood and Marrow Transplant Research (CIBMTR) registry study did not observe any difference in OS, PFS, relapse, or non-relapse mortality (NRM) with the addition of rituximab to the BEAM conditioning regimen in 862 relapsed chemo-sensitive DLBCL patients undergoing autoSCT [33].

Factors Affecting Outcomes of AutoSCT

The age adjusted international prognostic index (IPI) score incorporates clinical variables such as age, serum LDH, ECOG performance status, Ann Arbor stage, and extra-nodal involvement. In the CORAL trial, patients with a high IPI score (2-3) experienced a lower EFS when compared to those with a low IPI score (0-1) (18% vs. 40%) [28]. The second line age-adjusted IPI (sAAIPI) score, consisting of LDH, stage III or IV disease, and performance status, predicts PFS and OS in relapsed refractory DLBCL treated with ICE chemotherapy followed by autoSCT. Three-year PFS and OS were 70% and 74% for low-risk sAAIPI (0 factor), 39% and 49% for intermediate risk sAAIPI (1 factor), and 16% and 18% for high-risk sAAIPI (two to three factors) [34].

The timing of disease relapses affects the post-transplant outcomes. Three-year PFS was significantly lower for patients experiencing an early relapse (<12 months) when compared to late relapse (>12 months) in the CORAL trial (20% vs. 45%) [28]. Primary refractory disease portends poor outcomes with salvage chemotherapy followed by autoSCT [27,35,36]. The SCHOLAR-1 study evaluated the outcomes of refractory DLBCL (progressive or stable disease to first-line or salvage chemotherapy, or relapse within 12 months of autoSCT), and revealed a poor response rate to the next line of therapy (ORR 26% and CR 7%), a median survival of 6.3 months and a one-year survival of 28% [37]. The MSKCC study reviewed the outcomes of primary refractory DLBCL patients who underwent salvage chemotherapy followed by autoSCT. Patients were grouped into primary partial responders (partial response (PR) to initial therapy) or primary progressors (minimal or no response to initial therapy) [38]. The Three-year PFS was 49% in primary partial responders compared to 17% in primary progressors. Moreover, the overall response rate (complete response (CR), PR) was 68% in the primary partial responders versus 40% in the primary progressors. Patients with a Deauville response from one to three had a 3-year PFS of 68% when compared to 30% in Deauville score four patients and 0% in Deauville score five patients [38]. Recently, phase III studies comparing CAR T-cell therapy with the standard of care (autoSCT) in relapsed refractory DLBCL patients have completed accrual and results are awaiting (NCT03391466, NCT03570892, NCT03575351).

The PET scan response after salvage chemotherapy is predictive of the autoSCT outcomes [39]. Patients with a Deauville score of one to three in response to salvage
chemotherapy experienced a higher three-year PFS and OS of 77% and 86%, respectively, compared to 49% and 54% [40]. Another study evaluated the impact of pre-transplant PET scans following salvage chemotherapy. PET-negative CR after salvage was associated with an improved four-year PFS (64% vs. 32%) and OS (75% vs. 56%) when compared to PET-positive patients [41]. Patients with PR from salvage chemotherapy are at higher risk of failure than CR [40]. A recent CIBMTR registry analysis showed that patients with a chemo-sensitive PET-positive PR following salvage chemotherapy experienced a five-year PFS of 40% after autoSCT [42]. Based on the above-mentioned evidence, we routinely perform PET scans after salvage chemotherapy. We consider autoSCT in chemo-sensitive relapsed DLBCL patients who have achieved partial or complete remission (Deauville score one to three) to salvage chemotherapy, and reserve CAR T-cell therapy to patients with stable or progressive disease (Deauville score four to five). We do not routinely offer rituximab maintenance therapy after autoSCT.

4. Allogeneic Stem Cell Transplant

Allogeneic stem cell transplant (alloSCT) is one of the curative treatment modalities in DLBCL. However, its utility is limited partly due to a high non-relapse mortality (NRM), and the availability of newer therapeutic options, including CAR T-cell therapy. Although CRS and ICANS caused by CAR T-cell therapy can be life threatening, NRM is typically low. Moreover, the responses to CAR T-cell therapy are independent to the disease burden, which makes it an attractive treatment option in chemo-refractory disease when compared to alloSCT. Thus, CAR T-cell therapy has essentially replaced alloSCT as a preferred treatment modality in those patients who have failed multiple lines of therapy, and those with post-autoSCT relapse. A recent CIBMTR analysis demonstrated a remarkable decline in the use of alloSCT for DLBCL and a sharp increase in the use of CAR T-cell therapy. Although CAR T-cell therapy offers durable remissions in relapsed DLBCL patients, approximately 50% of these patients still experience disease relapse, and their outcomes are dismal [43,44]. AlloSCT has a role in such circumstances [45,46]. Moreover, alloSCT has been shown to provide durable remissions in high-risk DLBCL, such as DHL and DEL [47], or in disease that has been unresponsive to multiple prior therapies. AlloSCT can be considered in situations where CAR T-cell therapy is not feasible, such as refractory cytopenia or the presence of myelodysplastic syndrome. Unlike CAR T-cell therapy, which specifically targets the CD19 antigen on lymphoma cells, alloSCT offers a strong immune response against multiple unknown tumor antigens through the graft-versus-lymphoma effect that is exerted by the donor stem cells [48,49].

Table 1 summarizes the outcomes of alloSCT for DLBCL. The CIBMTR study evaluated the outcomes of 503 DLBCL patients who underwent alloSCT after disease progression, following prior autoSCT [46]. The three-year NRM, relapse, PFS, and OS were 30, 38, 31, and 37%, respectively. A Karnofsky performance status (KPS) of <80%, chemoresistance, an autoSCT to alloSCT interval of <1 year, and myeloablative conditioning were associated with an inferior PFS. Similarly, the European Society for Blood and Marrow Transplantation (EBMT) analysis reported the outcomes of alloSCT in 101 DLBCL patients who progressed after autoSCT [50]. At 3 years, the NRM was 28.2%, the relapse rate was 30.1%, the PFS was 41.7%, and the OS was 53.8%. A high NRM was noted in older patients (>45 years) and in those with an early relapse (<12 months) after autoSCT. The relapse rate was high in refractory patients, and a time interval to relapse after autoSCT of <12 months was associated with a lower PFS. In addition, recent studies have shown a higher rate of NRM and a lower relapse rate with myeloablative conditioning regimens (MAC) when compared to reduced intensity conditioning regimens (RIC) in alloSCT treatment for NHL, offsetting the survival advantage associated with MAC regimens [49,51]. Our practice corroborates with previous studies, and we typically reserve alloSCT for post-CAR T-cell relapse, the presence of concurrent myelodysplasia, or the failure of CAR T-cell therapy production.
### Table 1. Outcomes of allogeneic stem cell transplants in DLBCL.

| Study                | No of Pts | Conditioning Regimen | Acute GVHD (Grade 2–4) | Chronic GVHD (1-yr) | NRM (1-yr) | Relapse | PFS (1-yr) | OS (1-yr) |
|----------------------|-----------|----------------------|------------------------|---------------------|------------|---------|------------|----------|
| Thomson et al. [52], 2009 | 48        | RIC 100%             | 17%                    | 22%                 | 29%        | 33% (4-yr) | 48% (4-yr) | 47% (4-yr) |
| Sirvent et al. [53], 2010 | 68        | RIC 100%             | 39%                    | 41%                 | 23%        | 41% (2-yr) | 44% (2-yr) | 49% (2-yr) |
| Lazarus et al. [54], 2010 | 79        | MAC 100%             | 42% (100-day)          | 23%                 | 41%        | 30% (1-yr) | 29% (1-yr) | 33% (1-yr) |
| Van Kampen et al. [50], 2011 | 101     | MAC (37%) vs. RIC (63%) | 33%                    | 42%                 | 24.5%      | 24% (1-yr) | 51.5% (1-yr) | 64.7% (1-yr) |
| Bacher et al. [51], 2012 | 396     | MAC (n = 165) vs. RIC (n = 143) vs. NMA (n = 88) | 43% vs. 43% vs. 44% | 35% vs. 39% vs. 33% | 47% vs. 31% vs. 29% | 23% vs. 32% vs. 37% (1-yr) | 30% vs. 37% vs. 34% (1-yr) | 38% vs. 46% vs. 45% (1-yr) |
| Fenske et al. [46], 2016 | 503    | MAC (25%) vs. RIC (75%) | 36%                    | 40%                 | 23%        | 33% (1-yr) | 44% (1-yr) | 54% (1-yr) |
| Modi et al. [49], 2020 | 70      | MAC (67%) vs. RIC (23%) | 36.2% vs. 8.7% (Grade 3–4) | 27.7% vs. 43.5% (Extensive) | 39.7% vs. 39.1% (3-yr) | 25.5% vs. 17.4% (3-yr) | 34.2% vs. 34.7% (3-yr) | 34.4% vs. 34.4% (3-yr) |

RIC: Reduced intensity conditioning regimen; MAC: Myeloablative conditioning regimen; NMA: Non-myeloablative; NRM: Non-relapsed mortality; PFS: Progression-free survival; OS: Overall survival; Yr: Year; GVHD: Graft-versus-host-disease.

## 5. Chimeric Antigen Receptor T-Cell Therapy

The chimeric antigen receptor (CAR) consists of an extracellular antigen recognition domain, which binds to cell surface antigens, and an intracellular signaling domain that provides an activation signal. T-cell receptor activation requires two signals: the first signal is delivered through the T-cell receptor, and the second signal is provided by a costimulatory molecule that allows the proliferation of T-cells. The universal presence of CD19, CD20, and CD22 antigens on malignant B-cells make them the perfect targets for cellular therapies. CD19-directed autologous chimeric antigen receptor T-cell (CAR T-cell) therapy has revolutionized the treatment paradigm for DLBCL (Figure 1). Currently, axicabtagene ciloleucel (axi-cel, Yescarta®), tisagenlecleucel (tisa-cel, Kymriah®), and lisocabtagene maraleucel (liso-cel, Breyanzi®) have been approved by the US Food and Drug Administration (FDA) for the treatment of relapsed refractory DLBCL. The ZUMA-1 multicenter phase II single arm trial evaluated axi-cel in DLBCL, primary mediastinal B-cell lymphoma (PMBCL), and transformed follicular lymphoma (FL), and those which were refractory to prior therapy or had relapsed following autoSCT. The overall response rate (ORR) was 83% and the CR rate was 58%. At a median follow-up of 27.1 months, the median duration of response was 11.1 months, the median OS was not obtained, and the median PFS was 5.9 months [55]. Moreover, one third of the patients with (11/33) PR and half of the patients (11/24) with stable disease at 1 month achieved CR at 6 months. Similarly, the JULIET trial evaluated tisa-cel in relapsed refractory DLBCL patients who were ineligible for autoSCT, or who had disease progression following autoSCT and demonstrated an ORR of 52%, and a CR of 40% [56]. Approximately 54% of patients with PR achieved CR. One-year survival was 49%. Liso-cel is a third CD-19-directed autologous CAR T-cell product with a 4-1BB co-stimulatory domain, which is administered as a sequential infusion of two components (CD8+ and CD4+ CAR T-cells) at equal target doses (1:1). The TRANSCEND NHL 001 study evaluated liso-cel in relapsed refractory DLBCL, PMBCL, transformed DLBCL from indolent lymphoma, and FL grade 3B. At a median follow-up of 18.8 months, the ORR was 73%, and the CR rate was 53% [57]. Various large, retrospective reports of real-world experiences in using CAR T-cell therapy have shown a similar efficacy [58–62] (Table 2). Given the comparable efficacy and toxicity profiles of approved CAR T-cell therapies, currently the choice of therapy is largely guided by patient’s insurance.
**Table 2.** Clinical trials of CAR T-cell therapy in lymphoma.

| Trial                          | CAR-T Product | Costimulatory Domain | No of Pts | Bridging Chemotherapy | Lymphodepleting Chemotherapy | Patient Population | ORR/CR (%) | CRS/ Grade ≥ 3 (%) | ICANS/ Grade ≥ 3 (%) |
|-------------------------------|---------------|----------------------|-----------|-----------------------|------------------------------|---------------------|-------------|-------------------|-----------------------|
| ZUMA-1 [55]                  | Axi-cel       | CD28                 | 111       | No                    | Fludarabine (Flu) 30 mg/m² and Cyclophosphamide (Cy) 500 mg/m² × 3 days | DLBCL ≥ 2 lines, transformed FL (tFL), PMBCL | 82/58       | 93/13             | 64/28                 |
| JULIET [56]                  | Tisa-cel      | 4-1BB                | 165       | Yes (92%)             | Flu 25 mg/m² and Cy 250 mg/m² × 3 days, or bendamustine 90 mg/m² × 2 days | DLBCL ≥ 2 lines, tFL | 52/40       | 58/23             | 21/12                 |
| TRANSCEnd-NHL 001 [57]       | Liso-cel      | 4-1BB                | 342       | Yes (59%)             | Flu 30 mg/m² and Cy 300 mg/m² × 3 days | DLBCL ≥ 2 lines, tFL, PMBCL | 73/53       | 42/2              | 30/10                 |
| Nastoupil et al. [60]        | Axi-cel       | CD28                 | 165       | Yes (53%)             | Flu 30 mg/m² and Cy 500 mg/m² × 3 days | DLBCL, PMBCL, tFL | 82/64       | 92/7              | 69/31                 |
| Jacobson et al. [58]         | Axi-cel       | CD28                 | 122       | Yes (45%)             | Not available | DLBCL, PMBCL, tFL, transformed marginal zone lymphoma (tMZL), transformed CLL | 70/50       | 93/16             | 70/35                 |
| Sesques et al. [62]          | Axi-cel/Tisa-cel | CD28/4-1BB          | 70        | Yes (97%)             | Bendamustine (2%); Flu and Cy | DLBCL, PMBCL, tFL, tMZL | 63/48       | 85/8              | 28/10                 |
| Jaglowski et al. (CIBMTR) [59]| Tisa-cel      | 4-1BB                | 70        | Not available         | Not available | DLBCL, tFL | 59.6/38.3 | NA/4.3             | NA/4.3                |
| Pasquini et al. (CIBMTR) [61]| Axi-cel       | CD28                 | 453       | Not available         | Not available | Large B-cell lymphoma | 70/52       | 83/14             | 61/NA                 |

ORR: Overall response rate; CR: Complete response; CRS: Cytokine release syndrome; ICANS: Immune effector cell-associated neurotoxicity syndrome; NA: Not available; DLBCL: Diffuse large B-cell lymphoma; FL: Follicular lymphoma; PMBCL: Primary mediastinal B-cell lymphoma.
It is important to mention that above-mentioned studies differ in their efficacy and toxicities mainly due to the differences in the study design, population, and lymphodepleting chemotherapy. The ZUMA-1 study reported higher ORR and CR rates when compared to other studies, where bridging chemotherapy was not allowed. It is likely that the ZUMA-1 study may have included patients with a relatively indolent disease biology when compared to others [55–57]. Furthermore, the UK study reported a lower ORR when compared to other studies, which could be secondary to the prolonged time from the patient selection to CAR T-cell infusion (median 63 days) [62]. The CD28 costimulatory domain results in rapid T-cell expansion, which leads to an earlier and more severe cytokine release syndrome (CRS), while the use of the 4-1BB costimulatory domain results in a slower expansion of T-cells, as well as a lower incidence and severity of both CRS and immune effector cell-associated neurotoxicity syndrome (ICANS). CAR T-cell therapy is well tolerated in patients with an advanced age [63], multiple comorbidities or a borderline performance status [60], and side effects are, in general, manageable. Several strategies have been employed to mitigate the toxicities that are associated with CAR T-cell therapy. Recently, the use of prophylactic corticosteroids (oral dexamethasone 10 mg daily day zero through day two) has shown to reduce the incidence and severity of CRS (incidence 80%, all grade ≤ 2) and ICANS (incidence 58%, grade 3 ≥ 13%) following axi-cel administration [64]. No impact on the response rate was noted. CAR T-cell therapy is known to induce granulocyte-macrophage colony-stimulating factor (GM-CSF) production, which mediates inflammatory reactions and neurotoxicity. The ZUMA-19 trial, investigating lenzilumab (an anti-GM-CSF monoclonal antibody) in combination with axi-cel in relapsed refractory large B-cell lymphoma, is undergoing [65]. Given the improvement in CRS and ICANS with prophylactic corticosteroids, we routinely offer dexamethasone 10 mg daily for 3 days following axi-cel infusion.

Outcomes of the patients who progress following CAR T-cell therapy are poor [43]. The US lymphoma CAR T-cell consortium reported the outcomes of 136 patients (out of 275, 49%) who had experienced disease progression after axi-cel treatment. The median

![Figure 1. Overview of the interaction between malignant B-cells and other immune cells in the tumor microenvironment, and the therapeutic targets in diffuse large B-cell lymphoma. CAR: Chimeric antigen receptor; BiTE: Bispecific T-cell engager; SIRP: Signal regulatory protein.](image-url)
OS from disease progression was 6 months. Among 74% who received further therapy, the ORR was 29%, with a CR of 17%, and a median PFS of 55 days. CD19 losses were found in approximately 30% of patients at progression [44]. Antigen escape is one of the mechanisms of CAR T-cell resistance, where the malignant cells that are treated with CAR T-cells display the partial or complete loss of the CD19 antigen. To overcome this problem, dual CAR T constructs or tandem CAR T, which is a single CAR construct that contains two single-chain fragment variables (scFvs) to concomitantly target multiple target tumor antigens, have been used [66]. In addition, dual-targeted CAR T-cells (CD19/CD22, CD19/CD20) have shown promising activity in acute lymphoblastic leukemia (ALL), as well as B-cell malignancies [67–69]. AUTO3, anti-CD19/CD22 CAR-T treatment, with or without pembrolizumab, has demonstrated an ORR of 64% and a CR of 55% in a phase I/II study of relapsed refractory DLBCL [70]. CAR T-cell exhaustion and immunosuppressive TME are other plausible etiologies of CAR T-cell failure. PDI blockade with pembrolizumab has demonstrated an ORR of 27% in patients with a progression of NHL after CD19 CAR T-cell therapy [71]. The ZUMA-6 investigated the safety and efficacy of axi-cel in combination with atezolizumab (four doses every 3 weeks) in relapsed refractory DLBCL patients and noted an ORR of 90% with a CR of 60%. CAR T-cell expansion was greater than two-fold higher than observed in the ZUMA-1 trial [72].

A proportion of patients with a proliferative disease could not receive CAR T-cell therapy in a timely manner; this was either due to a manufacturing failure or delays. The median time from enrollment to infusion was 54 days in the JULIET trial [56], and the median time from leukapheresis to axi-cel delivery was 17 days in the ZUMA-1 study [55]. Approximately 31% of enrolled patients in the JULIET trial could not receive CAR T-cell infusion secondary to disease progression, and around 7% had a manufacturing failure. Similarly, approximately 10% of patients in the ZUMA-1 trial did not receive CAR T-cell infusion. Moreover, T-cell dysfunction with a decrease in functional T-cells is more prevalent in patients who have had multiple lines of prior therapy. Allogeneic CAR T-cells (off the shelf CAR T) can overcome these factors, limiting the access of CAR T-cell therapy to high-risk populations. The off-the-shelf, allogeneic CAR T-cell product, PBCAR0191, has shown a CR of 33% (standard lymphodepletion) and 71% (escalated lymphodepletion) in 13 patients with CD19-positive relapsed refractory NHL. Neither GVHD nor the presence of a CRS or ICANS grade of over three was noted [73]. Table 3 shows the selected ongoing studies of allogeneic and dual-construct CAR T-cell therapy.

| Intervention | Molecular Target | Trial | Phase | Indication | Primary Endpoint | Status |
|--------------|-----------------|-------|-------|------------|-----------------|--------|
| R-CHOP/PoV-R-CHP plus Glofitamab | BiTE (CD3/CD20) | NCT04914741/COALITION | I/II | Untreated DLBCL | Safety | Recruiting |
| Glofitamab plus R-CHOP | BiTE (CD3/CD20) | NCT04980222 | II | Untreated DLBCL | CR | Recruiting |
| Mosunetuzumab or in combination with PoV | BiTE (CD3/CD20) | NCT03677154 | I/II | Untreated DLBCL | Safety, ORR, CR | Recruiting |
| Tafasitamab plus lenalidomide plus R-CHOP vs. R-CHOP | Anti-CD19 mAb | NCT04824092/FrontMIND | III | Untreated DLBCL | PFS | Recruiting |
| Venetoclax plus PoV plus R-CHP | Anti-CD79b ADC | NCT04790903 | I | Untreated DLBCL | Safety | Recruiting |
| Induction and Maintenance Avelumab | Anti-FD-L1 antibody | NCT03244176 | I | Untreated DLBCL | Safety | Recruiting |

**Table 3.** Selected ongoing trials of Immunotherapy in DLBCL.
| Intervention                                                                 | Molecular Target                        | Trial                      | Phase | Indication                        | Primary Endpoint | Status     |
|------------------------------------------------------------------------------|----------------------------------------|---------------------------|-------|-----------------------------------|------------------|------------|
| Loncastutixmab plus rituximab vs. R-GemOx                                   | Anti-CD19 ADC                          | NCT04384484/LOTIS-5       | III   | RR DLBCL                          | PFS              | Recruiting |
| Loncastuximab Tesirine and Ibrutinib                                       | Anti-CD19 ADC                          | NCT03684694/LOTIS-3       | I/II  | RR DLBCL, MCL                     | Safety           | Recruiting |
| PoV plus R-GemOx vs. R-GemOx                                                | Anti-CD79b ADC                         | NCT04182204/POLARGO       | III   | RR DLBCL                          | OS               | Recruiting |
| PoV plus R-ICE vs. R-ICE                                                    | Anti-CD79b ADC                         | NCT04833114               | III   | RR DLBCL                          | EFS              | Recruiting |
| Nivolumab plus ipilimumab and adaptive T-cell therapy                       | Anti-PD-1 and CTLA-4 antibody          | NCT03305445               | Ib/Ii | RR DLBCL                          | Safety, CR       | Recruiting |
| Copanlisib and nivolumab                                                    | Anti-PD-1 antibody                     | NCT03484819               | II    | RR DLBCL, PMBCL                   | ORR              | Recruiting |
| Pembrolizumab plus anti-CD20 antibody                                        | Anti-PD-1 antibody                     | NCT03401853               | II    | RR DLBCL, FL                      | ORR              | Recruiting |
| Atzolizumab plus R-GemOx                                                    | Anti-PD-L1 antibody                    | NCT03321643               | I     | RR DLBCL                          | Safety           | Recruiting |
| Pembrolizum Plus Vorinostat                                                 | Anti-PD-1 antibody                     | NCT03150329               | I     | RR DLBCL, FL, HL                  | Safety           | Recruiting |
| Camrelizumab plus Apatinib                                                  | Anti-PD-1 antibody                     | NCT04476459               | I/II  | RR DLBCL                          | ORR              | Recruiting |
| Tislelizumab plus Lenalidomide                                               | Anti-PD-1 antibody                     | NCT04796857               | I/II  | RR DLBCL                          | ORR              | Recruiting |
| Varilimumab plus Nivolumab                                                  | Anti-CD27/anti-PD-1 antibody           | NCT03038672               | II    | RR NHL                            | ORR              | Recruiting |
| Nivolumab plus lenalidomide                                                 | Anti-PD-1 antibody                     | NCT03015896               | I/II  | RR NHL, HL                        | Safety           | Recruiting |
| DPX-Survivac along or with pembrolizum with or without low-dose cyclophosphamide | Anti-PD-1 antibody                     | NCT04920617               | II    | RR DLBCL                          | ORR              | Recruiting |
| AUTO3 (CD19/CD22 CAR T) with Pembrolizumab                                 | CAR T/Anti-PD-1 antibody               | NCT03287817/ALEXANDER     | I/II  | RR DLBCL                          | Safety, ORR      | Recruiting |
| CD19 CAR-T Expressing IL7 and CCL19 Combined with PD1 mAb                   | CAR T/Anti-PD-1 antibody               | NCT04381741               | I     | RR DLBCL                          | ORR              | Recruiting |
| C-CAR066 (anti-CD20 CAR T-cell therapy)                                     | CAR T-cell                             | NCT04316624               | I     | RR DLBCL who failed CD19 CAR T-cell therapy | Safety           | Recruiting |
| anti-CD19 and anti-CD20 dual specific CAR T-Cells                            | CAR T-cell                             | NCT04848672               | I     | RR DLBCL                          | Safety           | Recruiting |
| MB-CART2019.1 (CD19/CD20 CAR T) vs. SOC                                     | CAR T-cell                             | NCT04844866               | II    | RR DLBCL                          | PFS              | Recruiting |
| LUCAR-20S (Anti-CD20 CAR T)                                                  | CAR T-cell                             | NCT04176913               | I     | RR DLBCL, FL, MCL, CLL            | Safety           | Recruiting |
| Autologous Anti-CD20 CAR-T                                                  | CAR T-cell                             | NCT03277729               | I/II  | RR NHL                            | Safety           | Recruiting |
| Autologous Anti-CD22 CAR-T                                                  | CAR T-cell                             | NCT04088890               | I/Ib  | RR NHL                            | Safety           | Recruiting |
| Autologous anti-CD19/CD20 CAR T                                            | CAR T-cell                             | NCT04215016               | I     | RR DLBCL                          | Safety           | Recruiting |
| Autologous anti-CD19/CD20 CAR T                                            | CAR T-cell                             | NCT04007029               | I     | RR NHL, CLL                       | Safety           | Recruiting |
| Autologous anti-CD19/CD22 CAR T                                            | CAR T-cell                             | NCT03233854               | I     | RR NHL                            | Safety           | Recruiting |
| Acalabrutinib with Anti-CD19 CAR T                                        | CAR T-cell                             | NCT04257578               | I/II  | RR NHL                            | Safety           | Recruiting |

PoV: Polatuzumab Vedotin; DLBCL: Diffuse large B-cell lymphoma; RR: Relapsed Refractory; NHL: Non-Hodgkin’s lymphoma; MCL: Mantle cell lymphoma; PMBCL: Primary Mediastinal B-cell lymphoma; FL: Follicular lymphoma; HL: Hodgkin’s lymphoma; CLL: Chronic lymphocytic leukemia; OS: Overall survival; EFS: Event-free survival; ORR: Overall response rate; CR: Complete remission; PFS: Progression-free survival; BiTE: Bispecific T-cell Engager; mAb: Monoclonal antibody; ADC: Antibody drug conjugate.
6. Immune Checkpoint Inhibitor Therapy

The programmed death-1 (PD-1) receptor is expressed by activated T-cells, B-cells, NK cells, and macrophages. It regulates the T-cell-mediated immune response through binding to its ligands, programmed cell-death protein-1 ligand-1 (PD-L1) and PD-L2. Tumor cells express PD-L1 and PD-L2, which subsequently down-regulates T-cell activation, and indirectly helps them to escape the immune response. Upregulation of CTLA-4 in T-cells is another mechanism through which tumor cells suppress the immune response. (Figure 1). In Hodgkin’s Lymphoma (HL), an amplification of 9p24.1 has been shown to increase the expression of PD-L1 and PD-L2, and is associated with a shorter PFS [74]. Subsequently, PD-1 blockade by the anti-PD-1 antibodies nivolumab and pembrolizumab has shown promising results in relapsed [75–78] and newly diagnosed [79,80] HL.

Unlike HL, DLBCL cells do not frequently express PD-L1. PD-L1 gene alteration has been reported in approximately 25–31% of DLBCL patients [81,82]. A study evaluated the impact of PD-L1 positivity on tumor cells and their microenvironment (mPD-L1) in DLBCL. The expression of PD-L1 on tumor cells and mPD-L1 was observed in 11% and 15.3% of cases, respectively. PD-L1 and mPD-L1-positive DLBCL were significantly associated with the non-GCB subtype and Epstein-Barr virus positivity. Patients with PD-L1-positive DLBCL had a poor OS when compared to PD-L1-negative DLBCL patients, while no difference in OS was observed between mPD-L1-positive and mPD-L1-negative DLBCL patients [83]. Likewise, another study reported that DLBCL with PD-L1 alterations experienced an inferior PFS following front-line chemoimmunotherapy; however, the in relapsed setting, PD-L1 alterations were associated with a response to anti-PD-1 antibodies [82].

Checkpoint inhibitors (CPI) have shown disappointing results in DLBCL. Nivolumab was evaluated in relapsed refractory lymphoma and multiple myeloma in a phase I, multicenter study. Among eleven patients with DLBCL, the ORR was 36% (CR = 18% and PR = 18%), and the median PFS was only 7 weeks. At a median follow-up of 22.7 weeks for DLBCL patients, one of the four patients had a continued response [84]. Checkmate 036, the combination of nivolumab plus ipilimumab, yielded an ORR of 20% in relapsed refractory NHL patients (10/15 with DLBCL), with a median PFS of only 1.5 months [85]. Pidilizumab, an-anti-PD-1 monoclonal antibody, was evaluated in a phase II study of DLBCL patients undergoing autoSCT. In patients without progressive disease, pidilizumab was offered at every 42 days for 3 cycles, beginning 30 to 90 days after autoSCT. The ORR among patients with a measurable disease post-autoSCT was 51%, and the CR and PR rates were 34% and 17%, respectively. The 16 month PFS from the first treatment was 72%. Among those patients who had remained PET-positive at the conclusion of salvage therapy, the 16 month PFS was 70% [86]. Furthermore, pembrolizumab treatment, as a maintenance therapy in chemo-sensitive DLBCL patients undergoing autoSCT, revealed an 18 month PFS of 59% [87]. Recently, a phase II study evaluating the effect of treatment with nivolumab, 3 mg/kg every 2 weeks, in relapsed refractory DLBCL patients who were ineligible for autoSCT, or had failed autoSCT, was conducted. At a median follow-up of 9 months in the autoSCT-failed cohort and 6 months in the autoSCT-eligible cohort, the ORR values were 10% and 3%, respectively. The median PFS and OS were 1.9 and 12.2 months in the autoSCT-failed cohort, and 1.4 and 5.8 months in the autoSCT-eligible cohort, respectively. Of the evaluable samples in the analysis of 9p24.1, 16% had a low-level copy gain and 3% contained amplifications. The low response rates in DLBCL were attributed to infrequent genetic alterations in 9p24.1 [88]. The combination of pembrolizumab and R-CHOP was evaluated in 30 newly diagnosed DLBCL cases, and the ORR and CR were 90% and 77%, respectively. At a median follow-up of 25.5 months, the two-year PFS was 83% [89].

Atezolizumab, a humanized IgG1 anti-PD-L1 antibody, in combination with 6 cycles of R-CHOP followed by 12 months of consolidation, was evaluated in 42 untreated advanced DLBCL patients. The ORR was 87.5%, and the two-year PFS and OS were 74.9% and 86.4%, respectively [90]. A number of early clinical trials have shown a modest activity of atezolizumab in combination with various therapeutic agents in relapsed refractory DLBCL patients [91,92]. Durvalumab, a humanized IgG1 anti-PD-L1 antibody, in combination
with R-CHOP, has demonstrated a CR rate of 54% in treatment-naïve DLBCL patients [93], and has shown a fair activity in combination with ibrutinib in relapsed refractory DLBCL patients [94]. Ipilimumab, an anti-CTLA-4 antibody, has shown insignificant activity in relapsed refractory B-cell NHL [95–97]. Altogether, these data suggest that immune CPI therapies have failed to yield a clinically significant efficacy in DLBCL patients, while the addition of immune CPI therapies to first-line chemoimmunotherapeutic regimens has demonstrated an improvement in responses when compared to historical control rates. However, the long-term survival data are immature, and larger randomized trials are warranted. Ongoing studies of CPI with CAR-T and other agents are shown in Table 3.

7. Anti-CD47 Antibody

CD47 or “Do Not Eat Me” is an antiphagocytic signal that is expressed by cancer cells and enables immune evasion by macrophages. Its expression is associated with a poor prognosis. Anti-CD47 antibodies can induce the phagocytosis of tumor cells by the blockade of CD47 and its ligand, SIRPα. Anti-CD47 antibodies induce an antitumor T-cell response. Hu5F9-G4, an IgG4 humanized anti-CD47 monoclonal antibody (mAb), is associated with an ORR of 40% and a CR of 33% in relapsed refractory DLBCL patients [98]. Given the encouraging activity of anti-CD47 antibodies, various trials that incorporate this agent in combination with other regimens are ongoing.

8. Bispecific T-Cell Engager (BiTE) Antibody

BiTE antibodies are a newer form of immunotherapy for B-cell NHL. BiTE antibodies target antigens on tumor cells, while another end targets T-cells. It binds to tumor antigen and T-cells simultaneously, and facilitates the T-cell killing of cancer cells (Figure 1). The antigen-experienced T-cell subsets drive cancer cell death, while naïve T-cells are not activated. BiTE antibodies also increase the secretion of cytokines, which lead to changes in the tumor microenvironment. Blinatumomab was the first CD3/CD19 BiTE evaluated in a phase I study of relapsed refractory NHL in 76 patients, and 14 DLBCL patients were included. Among 35 patients who were treated with the maximum tolerated dose of 60 µg/m²/day, the ORR was 69%, and 55% for DLBCL patients (CR 36%) [99]. In a phase II study of blinatumomab in relapsed refractory DLBCL patients, the ORR after one cycle was 43%, the CR rate was 19%, and the median PFS was 3.7 months [100]. Blinatumomab was evaluated as a second salvage therapy, following platinum-based first salvage chemotherapy for relapsed refractory NHL, and the ORR at 12 weeks was 37% with a CR rate of 22% [101]. Based on these encouraging results, research into the combination of blinatumomab with lenalidomide (NCT02568553) or pembrolizumab (NCT03340766) is ongoing.

Several next generation BiTE antibodies are under development (Table 4). Notably, these antibodies have shown encouraging activity in relapsed refractory NHL patients, including those who fail CAR T-cell therapy [102–105]. Moreover, they are available off-the-shelf, and have a prolonged half-life, which allows a more convenient, once-weekly dose. Glofitamab is a BiTE antibody with a 2:1 configuration, allowing for the bivalent binding to CD20 on B-cells, and monovalent binding to CD3 on T-cells [105]. BiTE antibodies have shown encouraging activity and have offered complete remission in patients who have received multiple prior lines of therapy, including CAR T-cell therapy. Currently BiTE antibodies are not FDA approved, and therefore their use remains under investigation. Several trials incorporating these agents in first-line or in salvage settings are ongoing (Table 3).
## Table 4. Trials of BiTE antibodies in B-cell non-Hodgkin’s lymphoma.

| BiTE Antibody               | No of Pts | Pt Population                  | Dosing                                                                 | ORR (%) | CR (%) | Median PFS (Months) | CRS                  | ICANS               |
|-----------------------------|-----------|--------------------------------|-----------------------------------------------------------------------|----------|--------|---------------------|----------------------|---------------------|
| Epcoritamab [104]           | 68        | R/R NHL (DLBCL = 46; FL = 12; MCL = 4) | SC: 0.0128–60 mg R2PD = 48 mg DLBCL 12–60 mg = 68%; 48–60 mg = 91%      | FL 12–48 mg = 80% | DLBCL 12–60 mg = 46%; 48–60 mg = 55% | ≥ 12 mg = 9.1; ≥ 48 mg = NR | Grade 1–2 = 58%; Grade 3 = 3% |
| Mosunetuzumab + Polatuzumab [103] | 22 | R/R NHL (DLBCL = 12; FL3B = 3; tFL = 4; FL 1-3A = 3) | IV: 1-2-60 mg; PoV 1.8 mg/kg every 3 weeks Aggressive NHL = 63.2%; Post-CAR T = 57.1% FL = 100% Aggressive NHL = 47.4%; Post-CAR T = 28.6% FL = 100% | | | | Grade 1 = 9.1% None |
| Odronextamab (REGN1979) [102] | 136 | R/R NHL (DLBCL = 78) | 0.03–320 mg weekly × 12, then every 2 weeks DLBCL ≥ 80 mg, no Car-T = 55% DLBCL ≥ 80 mg, prior Car-T = 33% | | | | 61% all grades; Grade 3 ≥ 7%; Grade 3 = 3.7% |
| Glofitamab [105]            | 171       | R/R NHL (DLBCL = 73; FL1-3A = 44; tFL = 29; Richter’s = 10; PMBCL = 3) | R2PD = 2.5/10/30 Aggressive NHL = 48%; DLBCL = 41.4% DLBCL ≥ 10 mg = 55.3%; tFL ≥ 10 mg = 64.3% Aggressive NHL = 33.1%; DLBCL = 28.8% DLBCL ≥ 10 mg = 42.1%; tFL ≥ 10 mg = 64.3% | | | | 50.3% all grades; grade 3–4 = 3.5% 43.3% |

NHL: Non-Hodgkin’s Lymphoma; R/R: Relapsed refractory; DLBCL: Diffuse large B-cell lymphoma; FL: Follicular lymphoma; MCL: Mantle cell lymphoma; ORR: Overall response rate; CR: Complete remission; CRS: Cytokine release syndrome; ICANS: Immune effector cell-associated neurotoxicity syndrome; R2PD: Recommended phase 2 dose.
9. Antibody Drug Conjugates (ADC)

Polatuzumab Vedotin (PoV) is an anti-CD79b-directed ADC, and delivers monomethyl auristatin E (MMAE) inside malignant B-cells. The combination of PoV with bendamustine and rituximab (BR) has been shown to improve the CR (40% vs. 17.5%), PFS (median, 9.5 vs. 3.7 months), and OS (median, 12.4 vs. 4.7 months) when compared to BR in transplant-ineligible relapsed refractory DLBCL patients [29]. PoV in combination with lenalidomide and rituximab was evaluated in transplant-ineligible or failed relapsed refractory DLBCL patients. At a median follow-up of 9.5 months, the ORR and CR were 39% and 27%, respectively, and the median PFS and OS were 6.3 and 10.9 months, respectively [106]. Most recently, a phase III study (POLARIX), comparing PoV plus R-CHOP with R-CHOP alone in newly diagnosed DLBCL patients, has finished accrual and results are awaiting.

Loncastuximab tesirine (ADCT-402) is a humanized anti-CD19-directed ADC, delivering a pyrrolobenzodiazepine dimer cytotoxin, SG3199. It has shown an ORR of 48.3%, a CR of 24.1%, and a median PFS of 4.9 months in a phase II trial (LOTIS-2) involving relapse refractory DLBCL patients [107]. Currently, loncastuximab tesirine is being evaluated in combination with R-CHOP in untreated DLBCL patients.

Tafasitamab (MOR208) is an Fc-modified humanized anti-CD19 mAb. In a phase II trial (L-MIND) of relapsed DLBCL patients who were ineligible for transplant, the combination of tafasitamab and lenalidomide demonstrated an ORR of 57.5% and a CR of 40%. At a median follow-up of ≥35 months, the median PFS was 11.6 months, and the median OS was 33.5 months. The median PFS, duration of response, and OS were significantly longer in patients receiving tafasitamab and lenalidomide as second-line therapy when compared to those receiving third- or later-line treatment (23.5 months, 43.9 months, and 45.7 months when compared to 7.6 months, not reached, and 15.5 months, respectively) [108]. A phase II study of tafasitamab in combination with bendamustine when compared to BR in RR DLBCL is ongoing.

All the above-mentioned therapeutic options are non-curative and are often used in relapsed refractory DLBCL to bridge to stem cell transplant or CAR T-cell therapy. Given the impact of BR plus PoV on lymphocyte recovery and peripheral T-cell collection, we typically use this regimen following T-cell collection. Moreover, CD19 expression is typically maintained after the cessation of tafasitamab or loncastuximab tesirine treatments, and prior treatment with either of these agents does not preclude CAR T-cell therapy [109,110]. A study involving 14 DLBCL patients with their disease progressing after loncastuximab tesirine and subsequently undergoing CD19-directed CAR T-cell therapy revealed an ORR of 50% (CR 43% and PR 7%) [110]. Thus, we often consider CD19-directed agents in chemo-refractory patients, to control their disease prior to CAR T-cell therapy.

10. Conclusions

Tremendous progress has been made in understanding the molecular pathways that are involved in the pathogenesis of DLBCL. Immunotherapy approaches for the treatment of DLBCL have transformed the therapeutic landscape of the relapsed disease state. Beginning with traditional immunotherapy approaches, such as rituximab and stem cell transplant, the current treatment for DLBCL includes CART-cell therapy and immune checkpoint inhibitors for a subset of patients. Despite the groundbreaking advances in this area, there remain a number of challenges, including the emergence of resistance and toxicity. A multitude of clinical trials utilizing novel treatment strategies to overcome these challenges are underway, with bispecific antibodies being at the forefront. As these new approaches to leverage the host immune response unfold, the next decade is likely to bring a revolution to this area.

Author Contributions: D.M., B.P.: Literature search and data analysis; D.M. Wrote the first draft of the manuscript; D.M., B.P., J.U.: Approval of the final draft of the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.
Conflicts of Interest: Dipenkumar Modi serves in the advisory board in the MorphoSys and Seagen.

References
1. Alizadeh, A.A.; Eisen, M.B.; Davis, R.E.; Ma, C.; Lossos, I.S.; Rosenwald, A.; Boldrick, J.C.; Sabet, H.; Tran, T.; Yu, X.; et al. Distinct types of diffuse large B-cell lymphoma identified by gene expression profiling. Nature 2000, 403, 503–511. [CrossRef]
2. Lenz, G.; Wright, G.; Dave, S.; Xiao, W.; Powell, J.; Zhao, H.; Xu, W.; Tan, B.; Goldschmidt, N.; Iqbal, J.; et al. Stromal Gene Signatures in Large-B-Cell Lymphomas. N. Engl. J. Med. 2008, 359, 2313–2323. [CrossRef]
3. Scott, D.W.; Mottok, A.; Ennishi, D.; Wright, G.W.; Farinha, P.; Ben-Neriah, S.; Kridel, R.; Barry, G.S.; Hother, C.; Abrisqueta, P.; et al. Prognostic Significance of Diffuse Large B-Cell Lymphoma Cell of Origin Determined by Digital Gene Expression in Formalin-Fixed Paraffin-Embedded Tissue Bi-opsies. J. Clin. Oncol. 2015, 33, 2848–2856. [CrossRef]
4. Cuccuini, W.; Briere, J.; Mounier, N.; Voelker, H.U.; Rosenwald, A.; Sundstrom, C.; Cogliatti, S.; Hirchau, E.; Ysebaert, L.; Bron, D.; et al. MYC+ diffuse large B-cell lymphoma is not salvaged by classical R-ICE or R-DHAP followed by BEAM plus autologous stem cell transplantation. Blood 2012, 119, 4619–4624. [CrossRef]
5. Scott, D.W.; King, R.L.; Staiger, A.M.; Ben-Neriah, S.; Jiang, A.; Horn, H.; Mottok, A.; Farinha, P.; Slack, G.W.; Ennishi, D.; et al. High-grade B-cell lymphoma with MYC and BCL2 and/or BCL6 rearrangements with diffuse large B-cell lymphoma morphology. Blood 2018, 131, 2060–2069. [CrossRef]
6. Cheah, C.Y.; Oki, Y.; Westin, J.R.; Turturro, F. A clinician’s guide to double hit lymphomas. Br. J. Haematol. 2014, 168, 784–795. [CrossRef]
7. Green, T.M.; Young, K.H.; Visco, C.; Xu-Monette, Z.Y.; Orazi, A.; Go, R.S.; Nielsen, O.; Gadeberg, O.V.; Mourtis-Andersen, T.; Frederiksen, M.; et al. Immunohistochemical Double-Hit Score Is a Strong Predictor of Outcome in Patients with Diffuse Large B-Cell Lymphoma Treated With Rituximab Plus Cyclophosphamide, Doxorubicin, Vincristine, and Prednisone. J. Clin. Oncol. 2012, 30, 3460–3467. [CrossRef]
8. Landsburg, D.J.; Petrich, A.M.; Abramson, J.; Sohani, A.R.; Press, O.; Cassaday, R.D.; Chavez, J.C.; Song, K.; Zelenetz, A.D.; Gandhi, M.; et al. Impact of oncogene rearrangement patterns on outcomes in patients with double-hit non-Hodgkin lymphoma. Cancer 2015, 122, 559–564. [CrossRef]
9. Oki, Y.; Noorani, M.; Lin, P.; Davis, R.E.; Neelapu, S.S.; Ma, L.; Ahmed, M.; Rodriguez, M.A.; Hagegeister, F.B.; Fowler, N.; et al. Double hit lymphoma: The MD Anderson Cancer Center clinical experience. Br. J. Haematol. 2014, 166, 891–901. [CrossRef]
10. Herrera, A.F.; Mei, M.; Low, L.; Kim, H.T.; Griffin, G.K.; Song, J.Y.; Merryman, R.W.; Bedell, V.; Pak, C.; Sun, H.; et al. Relapsed or Refractory Double-Expressor and Dou-blle-Hit Lymphomas Have Inferior Progression-Free Survival After Autologous Stem-Cell Transplantation. J. Clin. Oncol. 2017, 35, 24–31. [CrossRef]
11. Staiger, A.M.; Ziepert, M.; Horn, H.; Scott, D.W.; Barth, T.F.E.; Bernd, H.W.; Feller, A.C.; Klapper, W.; Szczepanowski, M.; Hummel, M.; et al. German High-Grade Lymphoma Study Group. Clinical Impact of the Cell-Of-Origin Classification and the MYC/BCL2 Dual Expression Status in Diffuse Large B-Cell Lymphoma Treated Within Prospective Clinical Trials of the German High-Grade Non-Hodgkin’s Lymphoma Study Group. J. Clin. Oncol. 2017, 35, 2515–2526.
12. Hu, S.; Xu-Monette, Z.Y.; Tzankov, A.; Green, T.; Wu, L.; Balasubramanyam, A.; Liu, W.M.; Visco, C.; Li, Y.; Miranda, R.N.; et al. MYC/BCL2 protein coexpression con-tributes to the inferior survival of activated B-cell subtype of diffuse large B-cell lymphoma and demonstrates high-risk gene expression signatures: A report from The International DLBCL Rituximab-CHOP Consortium Program. Blood 2013, 121, 4021–4031. [CrossRef]
13. Ennishi, D.; Jiang, A.; Boyle, M.; Collinge, B.; Grande, B.M.; Ben-Neriah, S.; Rushton, C.; Tang, J.; Thomas, N.; Slack, G.W.; et al. Double-Hit Gene Expression Signature De-fines a Distinct Subgroup of Germinal Center B-Cell-Like Diffuse Large B-Cell Lymphoma. J. Clin. Oncol. 2019, 37, 190–201. [CrossRef]
14. Schmitz, R.; Wright, G.W.; Huang, D.W.; Johnson, C.A.; Phelan, J.D.; Wang, J.Q.; Roulland, S.; KasbeKar, M.; Young, R.M.; Shaffer, A.L.; et al. Genetics and Pathogenesis of Diffuse Large B-Cell Lymphoma. N. Engl. J. Med. 2018, 378, 1396–1407. [CrossRef]
15. Chapuy, B.; Stewart, C.; Dunford, A.J.; Kim, J.; Kamburov, A.; Redd, R.A.; Lawrence, M.S.; Roemer, M.G.M.; Li, A.J.; Ziepert, M.; et al. Molecular subtypes of diffuse large B cell lymphoma are associated with distinct pathogenic mechanisms and outcomes. Nat. Med. 2018, 24, 679–690. [CrossRef]
16. Wright, G.W.; Huang, D.W.; Phelan, J.D.; Coulilbaly, Z.A.; Roulland, S.; Young, R.M.; Wang, J.Q.; Schmitz, R.; Morin, R.; Tang, J.; et al. A Probabilistic Classification Tool for Genetic Subtypes of Diffuse Large B Cell Lymphoma with Therapeutic Implications. Cancer Cell 2020, 37, 551–568.e14. [CrossRef]
17. Lacy, S.E.; Barrans, S.L.; Beer, P.A.; Painter, D.; Smith, A.G.; Roman, E.; Cooke, S.L.; Ruiz, C.; Glover, P.; Van Hoppe, S.J.L.; et al. Targeted sequencing in DLBCL; outcomes and mechanisms: A Haematological Malignancy Research Network report. Blood 2020, 135, 1759–1771. [CrossRef]
18. Coiffier, B.; Lepage, E.; Briere, J.; Herbrecht, R.; Tilly, H.; Bouabdallah, R.; Morel, P.; Van Den Neste, E.; Salles, G.; Gaulard, P.; et al. Gisselbrecht CCHOP chemotherapy plus rituximab com-pared with CHOP alone in elderly patients with diffuse large-B-cell lymphoma. N. Engl. J. Med. 2002, 346, 235–242. [CrossRef]
19. Feugier, P.; Van Hoof, A.; Sebban, C.; Solal-Celigny, P.; Bouabdallah, R.; Fermé, C.; Christian, B.; Lepage, E.; Tilly, H.; Morschhauser, F.; et al. Long-term results of the R-CHOP study in the treatment of elderly patients with diffuse large B-cell lymphoma: A study by the Groupe d’Etude des Lym-phomes de l’Adulte. J. Clin. Oncol. 2005, 23, 4117–4126. [CrossRef]
20. Habermann, T.M.; Weller, E.A.; Morrison, V.A.; Gascoyne, R.D.; Cassileth, P.A.; Cohn, J.B.; Dakhil, S.R.; Woda, B.; Fisher, R.I.; Peterson, B.A.; et al. Rituximab-CHOP Versus CHOP Alone or With Maintenance Rituximab in Older Patients With Diffuse Large B-Cell Lymphoma. *J. Clin. Oncol.* **2006**, *24*, 3121–3127. [CrossRef]

21. Pfreundschuh, M.; Schubert, J.; Ziepert, M.; Schmits, R.; Mohren, M.; Lengfelder, E.; Reiser, M.; Nickenic, C.; Clemens, M.; Peter, N.; et al. Six versus eight cycles of bi-weekly CHOP-14 with or without rituximab in elderly patients with aggressive CD20+ B-cell lymphomas: A randomised controlled trial (RICOVER-60). *Lancet Oncol.* **2008**, *9*, 105–116. [CrossRef]

22. Pfreundschuh, M.; Trümper, L.; Österborg, A.; Pettengell, R.; Trneny, M.; Imrie, K.; Ma, D.; Gill, D.; Walewska, J.; Zinzani, P.L.; et al. CHOP-like chemotherapy plus rituximab versus CHOP-like chemotherapy alone in young patients with good-prognosis diffuse large-B-cell lymphoma: A randomised controlled trial by the MabThera International Trial (MInT) Group. *Lancet Oncol.* **2006**, *7*, 379–391. [CrossRef]

23. Pfreundschuh, M.; Kuhnt, E.; Trümper, L.; Österborg, A.; Trneny, M.; Shepherd, L.; Gill, D.S.; Walewska, J.; Pettengell, R.; Jaeger, U.; et al. CHOP-like chemotherapy with or without rituximab in young patients with good-prognosis diffuse large-B-cell lymphoma: 6-year results of an open-label randomised study of the MabThera International Trial (MInT) Group. *Lancet Oncol.* **2011**, *12*, 1013–1022. [CrossRef]

24. Mössner, E.; Brünker, P.; Moser, S.; Püntener, U.; Schmidt, C.; Herter, S.; Grau, R.; GERDES, C.; van Puijenbroek, E.; et al. Increasing the efficacy of CD20 antibody therapy through the engineering of a new type II anti-CD20 antibody with enhanced direct and immune effector cell-mediated B-cell cytotoxicity. *Blood* **2010**, *115*, 4393–4402. [CrossRef]

25. Vitolo, U.; Trneny, M.; Belada, D.; Burke, J.M.; Carella, A.M.; CHUA, N.; ABRISQUETA, P.; Demeter, J.; FLENN, I.; HONG, X.; et al. Obinutuzumab or Rituximab Plus Cyclophosphamide, Doxorubicin, Vincristine, and Prednisone in Previously Untreated Diffuse Large B-Cell Lymphoma. *J. Clin. Oncol.* **2017**, *35*, 3529–3537. [CrossRef]

26. Baetz, T.; Chen, B.E.; COUBAN, S.; Tom Kouroukis, C.; Buckstein, R.; Kuruvilla, J.; Howson-Jan, K.; Szwajcer, D.; Federico, M.; Meyer, R.M.; et al. Effect of the addition of rituximab to sal-vage chemotherapy prior to autologous stem cell transplant in aggressive CD20+ lymphoma: A cohort comparison from the NCIC Clinical Trials Group Study LY.12. *Leuk. Lymphoma* **2017**, *58*, 64–69. [CrossRef]

27. Crump, M.; Kuruvilla, J.; COUBAN, S.; MacDonald, D.A.; Kukreti, V.; Kouroukis, C.T.; Rubinger, M.; Buckstein, R.; IMRIE, K.R.; Federico, M.; et al. Randomized Comparison of Gemicitabine, Dexamethasone, and Cisplatin Versus Dexamethasone, Cytabine, and Cisplatin Chemotherapy Before Autologous Stem-Cell Transplantation for Relapsed and Refractory Aggressive Lymphomas: NCIC-CTG LY.12. *J. Clin. Oncol.* **2014**, *32*, 3490–3496. [CrossRef]

28. Gisselbrecht, C.; Glass, B.; Mounier, N.; Gill, D.S.; Linch, D.C.; TRNENY, M.; Bosly, A.; Ketterer, N.; Shipilberg, O.; Hagberg, H.; et al. Salvage Regimens With Autologous Transplantation for Relapsed Large B-Cell Lymphoma in the Rituximab Era. *J. Clin. Oncol.* **2010**, *28*, 4184–4190. [CrossRef]

29. Sehn, L.H.; Herrera, A.F.; Flowers, C.R.; Kamdar, M.K.; McMillan, A.; Hertzig, B.; Assouline, S.; Kim, T.M.; Kim, W.S.; Ozcan, M.; et al. Polatuzumab Vedotin in Relapsed or Primary Refractory Diffuse Large B-Cell Lymphoma. *J. Clin. Oncol.* **2020**, *38*, 155–165. [CrossRef]

30. Collière, B.; Thieblement, C.; Van Den Neste, E.; Lepeu, G.; Plantier, I.; Castaigne, S.; Lefort, S.; Marit, G.; MACRO, M.; Sebban, C.; et al. Long-term outcome of patients in the LHNY-98.5 trial, the first randomized study comparing rituximab-CHOP to standard CHOP chemotherapy in DLBCL pa-tients: A study by the Groupe d’Etudes des Lymphomes de l’Adul. *Blood* **2010**, *116*, 2040–2045. [CrossRef]

31. Philip, T.; Guglielmi, C.; Hagenbeek, A.; Somers, R.; Van der Leie, H.; Bron, D.; Sonneveld, P.; Gisselbrecht, C.; Cahn, J.Y.; Harousseau, J.L.; et al. Autologous bone marrow transplantation as compared with salvage chemotherapy in relapses of chemotherapy-sensitive non-Hodgkin’s lymphoma. *N. Engl. J. Med.* **1995**, *333*, 1540–1545. [CrossRef]

32. Vose, J.M.; Carter, S.; Burns, L.J.; Ayala, E.; Press, O.W.; Moskowitz, C.H.; Stadtmueller, E.A.; Mineshi, S.; Ambinder, R.; Fenske, T.; et al. Phase III randomized study of rituximab/carmustine, etoposide, cytarabine, and melphalan (BEAM) with autologous hematopoietic cell transplantation for relapsed diffuse large B-cell lymphoma: Results from the BMT CTN 0401 trial. *J. Clin. Oncol.* **2013**, *31*, 1662–1668. [CrossRef]

33. Jagadeesh, D.; Majhail, N.S.; He, Y.; Ahn, K.W.; Ms, C.L.; Ahmed, S.; Aljurf, M.; Bacher, U.; Badawy, S.M.; Bejanyan, N.; et al. Outcomes of rituximab-BEAM versus BEAM conditioning regimen in patients with diffuse large B-cell lymphoma undergoing autologous transplantation. *Cancer* **2020**, *126*, 2279–2287. [CrossRef]

34. Hamlin, P.A.; Zelenetz, A.D.; Kewalramani, T.; Qin, J.; Satogapan, J.M.; Verbel, D.; Noy, A.; Portlock, C.S.; Straus, D.J.; Yahalom, J.; et al. Age-adjusted International Prognostic Index predicts autologous stem cell transplantation outcome for patients with relapsed or primary refractory diffuse large B-cell lymphoma. *Blood* **2003**, *102*, 1989–1996. [CrossRef]

35. Guglielmi, C.; Gomez, F.; Philip, T.; Hagenbeek, A.; Martelli, M.; Sebban, C.; Melpied, N.; Bron, D.; Cahn, J.Y.; Somers, R.; et al. Time to relapse has prognostic value in patients with aggressive lymphoma enrolled onto the PARMA trial. *J. Clin. Oncol.* **1998**, *16*, 3264–3269. [CrossRef]

36. Vose, J.M.; Koster, J.; Loberiza, F.R.; Arevalo, A.; Bast, M.; Armitage, J.; Bierman, P.J.; Bociek, R.G.; Armitage, J.O. Late relapse in patients with diffuse large B-cell lymphoma. *Br. J. Haematol.* **2010**, *151*, 354–358. [CrossRef]

37. Crump, M.; Neeapu, S.S.; Farooq, U.; Neste, E.V.D.; Kuruvilla, J.; Westin, J.; Link, B.K.; Hay, A.; Cerhan, J.R.; Zhu, L.; et al. Outcomes in refractory diffuse large B-cell lymphoma: Results from the international SCHOLAR-1 study. *Blood* **2017**, *130*, 1800–1808. [CrossRef]
38. Vardhana, S.A.; Sauter, C.S.; Matasar, M.J.; Zelenetz, A.D.; Galasso, N.; Woo, K.M.; Zhang, Z.; Moskowitz, C.H. Outcomes of primary refractory diffuse large B-cell lymphoma (DLBCL) treated with salvage chemotherapy and intention to transplant in the rituximab era. Br. J. Haematol. 2016, 176, 591–599. [CrossRef]

39. Terasawa, T.; Dahabreh, I.J.; Nihashi, T. Fluorine-18-fluorodeoxyglucose positron emission tomography in response as-sessment before high-dose chemotherapy for lymphoma: A systematic review and meta-analysis. Oncologist 2010, 15, 750–759. [CrossRef]

40. Sauter, C.S.; Matasar, M.J.; Meikle, J.; Schoder, H.; Ulner, G.A.; Migliacci, J.C.; Hilden, P.; Devlin, S.M.; Zelenetz, A.D.; Moskowitz, C.H. Prognostic value of FDG-PET prior to autologous stem cell transplantation for relapsed and refractory diffuse large B-cell lymphoma. Blood 2015, 125, 2579–2581. [CrossRef]

41. Armand, P.; Welch, S.; Kim, H.T.; LaCasce, A.S.; Jacobsen, E.D.; Davids, M.S.; Jacobson, C.; Fisher, D.C.; Brown, J.R.; Coughlin, E.; et al. Prognostic factors for patients with dif-fuse large B cell lymphoma and transformed indolent lymphoma undergoing autologous stem cell transplantation in the positron emission tomography era. Br. J. Haematol. 2013, 160, 608–617. [CrossRef] [PubMed]

42. Shah, N.N.; Ahn, K.W.; Litovich, C.; Fenske, T.; Hamadani, M. Is autologous transplantation (autoHCT) in relapsed diffuse large B-cell lymphoma (DLBCL) patients achieving only a PET/CT positive partial remission (PR) appropriate in the CAR-T cell era? J. Clin. Oncol. 2020, 38, 8000. [CrossRef]

43. Chow, V.A.; Gopal, A.K.; Maloney, D.G.; Turtle, C.J.; Smith, S.D.; Ujiani, C.S.; Shadman, M.; Cassaday, R.D.; Till, B.G.; Tserg, Y.D.; et al. Outcomes of patients with large B-cell lymphomas and progressive disease following CD19-specific CAR T-cell therapy. Am. J. Hematol. 2018, 94, E209–E213. [CrossRef]

44. Spiegel, J.Y.; Dahiya, S.; Jain, M.D.; Tamasiress, J.; Nastoupiil, L.J.; Jacobs, M.T.; Ghobadi, A.; Lin, Y.; Lunnung, M.; Lekakis, L.; et al. Outcomes of patients with large B-cell lymphoma progressing after axicabtagene ciloleucel therapy. Blood 2021, 137, 1832–1835. [CrossRef] [PubMed]

45. Shadman, M.; Gauthier, J.; Hay, K.A.; Voutsinas, J.M.; Milano, F.; Li, A.; Hirayama, A.V.; Sorror, M.L.; Cherian, S.; Chen, X.; et al. Safety of allogeneic hematopoietic cell transplant in adults after CD19-targeted CAR T-cell therapy. Blood Adv. 2019, 3, 3062–3069. [CrossRef] [PubMed]

46. Fenske, T.S.; Ahn, K.W.; Graff, T.M.; DiGilio, A.; Bashir, Q.; Kamble, R.T.; Ayala, E.; Bacher, U.; Brammer, J.E.; Cairo, M.; et al. Allogeneic transplantation provides durable remission in a subset of DLBCL patients relapsing after autologous transplantation. Br. J. Haematol. 2016, 174, 235–248. [CrossRef]

47. Herrera, A.F.; Rodig, S.J.; Song, J.Y.; Kim, Y.; Griffin, G.K.; Yang, D.; Nikolaenko; L.; Mei, M.; Bedell, V.; Dal Cin, P.; et al. Outcomes after Allogeneic Stem Cell Transplantation in Patients with Double-Hit and Double-Expressor Lymphoma. Biol. Blood Marrow Transpl. 2018, 24, 514–520. [CrossRef] [PubMed]

48. Ghosh, N.; Ahmed, S.; Ahn, K.W.; Khanal, M.; Litovich, C.; Aljurf, M.; Bacher, V.U.; Bredeson, C.; Epperla, N.; Farhadfar, N.; et al. Association of Reduced-Intensity Conditioning Regimens With Overall Survival Among Patients With Non-Hodgkin Lymphoma Undergoing Allogeneic Transplant. JAMA Oncol. 2020, 6, 1011–1018. [CrossRef]

49. Modi, D.; Kim, S.; Surapaneni, M.; Ayash, L.; Alavi, A.; Ratanatharthorn, V.; Deol, A.; Uberti, J.P. R-BEAM versus Reduced-Intensity Conditioning Regimens in Patients Undergoing Allogeneic Stem Cell Transplantation for Relapsed Refractory Diffuse Large B Cell Lymphoma. Biol. Blood Marrow Transpl. 2020, 26, 683–690. [CrossRef]

50. van Kampen, R.J.; Canals, C.; Schouten, H.C.; Nagler, A.; Thomson, K.J.; Vernant, J.P.; Buzyn, A.; Boogars, M.A.; Luan, J.J.; Maury, S.; et al. Allogeneic stem-cell transplantation as salvage therapy for patients with diffuse large B-cell non-Hodgkin’s lymphoma relapsing after an autologous stem-cell transplantation: An analysis of the European Group for Blood and Marrow Transplantation Registry. J. Clin. Oncol. 2011, 29, 1342–1348. [CrossRef]

51. Bacher, U.; Klyuchnikov, E.; Le-Rademacher, J.; Carreras, J.; Armand, P.; Bishop, M.R.; Bredeson, C.N.; Cairo, M.S.; Fenske, T.S.; Freytes, C.O.; et al. Conditioning regimens for al-lotransplants for diffuse large B-cell lymphoma: Myeloablative or reduced intensity? Blood 2012, 120, 4256–4262. [CrossRef]

52. Thomson, K.J.; Morris, E.C.; Bloom, A.; Cook, G.; Milligan, D.; Parker, A.; Clark, F.; Yung, L.; Linch, D.C.; Chakraverty, R.; et al. Favorable long-term survival after re-duenced intensity allogeneic transplantation for multiple-relapse aggressive non-Hodgkin’s lymphoma. J. Clin. Oncol. 2009, 27, 426–432. [CrossRef]

53. Sirvent, A.; Dhedin, N.; Michallet, M.; Mounier, N.; Faucher, C.; Yakoub-Agha, I.; Mothy, M.; Robin, M.; Tabrizi, R.; Clement, L.; et al. Low nonrelapse mortality and pro-longed long-term survival after reduced-intensity allogeneic stem cell transplantation for relapsed or refractory diffuse large B cell lymphoma: Report of the Societe Francaise de Greffe de Moelle et de Therapie Cellulaire. Biol. Blood Marrow Transpl. 2020, 16, 78–85. [CrossRef]

54. Lazarus, H.M.; Zhang, M.-J.; Carreras, J.; Hayes-Lattin, B.M.; Ataergin, A.S.; Bitran, J.D.; Bolwell, B.J.; Freytes, C.O.; Gale, R.P.; Goldstein, S.C.; et al. A Comparison of HLA-Identical Sibling Allogeneic versus Autologous Transplantation for Diffuse Large B Cell Lymphoma: A Report from the CIBMTR. Biol. Blood Marrow Transpl. 2010, 16, 35–45. [CrossRef]

55. Locke, F.L.; Ghobadi, A.; Jacobson, C.A.; Miklos, D.B.; Lekakis, L.J.; Oluwole, O.O.; Lin, Y.; Braunischweig, I.; Hill, B.T.; Timmerman, J.M.; et al. Long-term safety and activity of axicabtagene ciloleucel in refractory large B-cell lymphoma (ZUMA-1): A single-arm, multicentre, phase 1–2 trial. Lancet Oncol. 2019, 20, 31–42. [CrossRef]

56. Schuster, S.J.; Bishop, M.R.; Tam, C.S.; Waller, E.K.; Borchmann, P.; McGuirk, J.P.; Jäger, U.; Jaglowski, S.; Andreadis, C.; Westin, J.R.; et al. Tisagenlecleucel in Adult Relapsed or Refractory Diffuse Large B-Cell Lymphoma. N. Engl. J. Med. 2019, 380, 45–56. [CrossRef] [PubMed]
76. Armand, P.; Shipp, M.A.; Ribrag, V.; Michot, J.-M.; Zinzani, P.L.; Kuruvilla, J.; Snyder, E.S.; Ricard, A.D.; Balakumar, A.; Rose, S.; et al. Programmed Death-1 Blockade With Pembrolizumab in Patients With Classical Hodgkin Lymphoma After Brentuximab Vedotin Failure. J. Clin. Oncol. 2016, 34, 3733–3739. [CrossRef] [PubMed]
77. Chen, R.; Zinzani, P.L.; Fanale, M.A.; Armand, P.; Johnson, N.A.; Brice, P.; Radford, J.; Ribrag, V.; Molin, D.; Vassilakopoulos, T.P.; et al. Phase II Study of the Efficacy and Safety of Pembrolizumab for Relapsed/Refractory Classic Hodgkin Lymphoma. J. Clin. Oncol. 2017, 35, 2125–2132. [CrossRef]
78. Younes, A.; Santoro, A.; Shipp, M.; Zinzani, P.L.; Timmerman, J.M.; Ansell, S.; Armand, P.; Fanale, M.; Ratanatharathorn, V.; Kuruvilla, J.; et al. Nivolumab for classical Hodgkin’s lymphoma after failure of both autologous stem-cell transplantation and brentuximab vedotin: A multicentre, multicohort, single-arm phase 2 trial. Lancet Oncol. 2016, 17, 1283–1294. [CrossRef]
79. Bröckelmann, F.P.; Goergen, H.; Kuehl, U.; Meissner, J.; Ordemann, R.; Halbguth, T.V.; Sasse, S.; Söker, M.; Kerkhoff, A.; Mathas, S.; et al. Efficacy of Nivolumab and Avelumab in Early-Stage Unfavorable Classic Hodgkin Lymphoma: The Randomized Phase 2 German Hodgkin Study Group NIVAH Trial. JAMA Oncol. 2020, 6, 872–880. [CrossRef]
80. Ramchandren, R.; Domingo-Domènech, E.; Rueda, A.; Treneny, M.; Feldman, T.A.; Lee, H.J.; Provençio, M.; Sillaber, C.; Cohen, J.B.; Savage, K.J.; et al. Nivolumab for Newly Diagnosed Advanced-Stage Classic Hodgkin Lymphoma: Safety and Efficacy in the Phase II CheckMate 205 Study. J. Clin. Oncol. 2019, 37, 1997–2007. [CrossRef] [PubMed]
81. Menter, T.; Bodmer-Haecki, A.; Dinnhöfer, S.; Tzankov, A. Evaluation of the diagnostic and prognostic value of PD-L1 expression in Hodgkin and B-cell lymphomas. Hum. Pathol. 2016, 54, 17–24. [CrossRef] [PubMed]
82. Godfrey, J.; Tumuluru, S.; Bao, R.; Leukam, M.; Venkataraman, G.; Phillip, J.; Fitzpatrick, C.; McElherne, J.; MacNabb, B.W.; Orlowski, R.; et al. PD-L1 gene alterations identify a subset of diffuse large B-cell lymphoma harboring a T-cell–inflamed phenotype. Blood 2019, 133, 2279–2290. [CrossRef] [PubMed]
83. Kiyasu, J.; Miyoshi, H.; Hirata, A.; Arakawa, F.; Ichikawa, A.; Niino, D.; Sugita, Y.; Yufu, Y.; Choi, I.; Abe, Y.; et al. Expression of programmed cell death ligand 1 is associated with poor overall survival in patients with diffuse large B-cell lymphoma. Blood 2015, 126, 2193–2201. [CrossRef]
84. Lesokhin, A.M.; Ansell, S.M.; Armand, P.; Scott, E.C.; Halwani, A.; Gutierrez, M.; Millenson, M.M.; Cohen, A.D.; Schuster, S.J.; Lebovic, D.; et al. Nivolumab in Patients With Relapsed or Refractory Hematologic Malignancy: Preliminary Results of a Phase Ib Study. J. Clin. Oncol. 2016, 34, 2698–2704. [CrossRef]
85. Ansell, S.; Gutierrez, M.E.; Shipp, M.A.; Gladstone, D.; Moskowitz, A.; Borello, I.; Popa-McKiver, M.; Farsaci, B.; Zhu, M.L.; Lesokhin, A.M.; et al. A Phase 1 Study of Nivolumab in Combination with Ipilimumab for Relapsed or Refractory Hematologic Malignancies (CheckMate 039). Blood 2016, 128, 183. [CrossRef]
86. Armand, P.; Nagler, A.; Weller, E.A.; Devine, S.M.; Avigan, D.E.; Chen, Y.B.; Kaminski, M.S.; Holland, H.K.; Winter, J.N.; Mason, J.R.; et al. Disabling immune tolerance by pro-grammed death-1 blockade with pidilizumab after autologous hematopoietic stem-cell transplantation for diffuse large B-cell lymphoma: Results of an international phase II trial. J. Clin. Oncol. 2013, 31, 4199–4206. [CrossRef] [PubMed]
87. Frigault, M.J.; Armand, P.; Redd, R.A.; Jeter, E.; Merryman, R.; Coleman, K.C.; Herrera, A.F.; Dahli, P.; Nieto, Y.; LaCasce, A.S.; et al. PD-1 blockade for diffuse large B-cell lymphoma after autologous stem cell transplantation. Blood Adv. 2020, 4, 122–126. [CrossRef]
88. Ansell, S.M.; Minnema, M.C.; Johnson, P.; Timmerman, J.M.; Armand, P.; Shipp, M.A.; Rodig, S.J.; Ligon, A.H.; Roemer, M.G.M.; Reddy, N.; et al. Nivolumab for Relapsed/Fractory Diffuse Large B-Cell Lymphoma in Patients Ineligible for or Having Failed Autologous Transplantation: A Single-Arm, Phase II Study. J. Clin. Oncol. 2019, 37, 481–489. [CrossRef]
89. Smith, S.D.; Till, B.G.; Shadman, M.S.; Lynch, R.C.; Cowan, A.J.; Wu, Q.V.; Voutsinas, J.; Rasmussen, H.A.; Blue, K.; Ujjani, C.S.; et al. Pembrolizumab with R-CHOP in previously untreated diffuse large B-cell lymphoma: Potential for biomarker driven therapy. Br. J. Haematol. 2020, 189, 1119–1126. [CrossRef] [PubMed]
90. Younes, A.; Burke, J.M.; Cheson, B.D.; Diefenbach, C.; Ferrari, S.; Hahn, U.H.; Hawkes, E.A.; Khan, C.; Losssos, I.S.; Musuraca, G.; et al. Safety and Efficacy of Atezolizumab in Combination with Rituximab Plus CHOP in Previously Untreated Patients with Diffuse Large B-Cell Lymphoma (DLBCL): Updated Analysis of a Phase I/II Study. Blood 2019, 134, 2274. [CrossRef]
91. Palomba, M.L.; Cartron, G.; Popplewell, L.; Ribrag, V.; Westin, J.; Chitra, S.; Huw, L.; Newberry, K.; Raval, A.; Xu, J.; et al. Safety and clinical activity of atezolizumab in combination with gemcitabine plus nab-paclitaxel in patients with locally advanced or metastatic triple-negative breast cancer: A phase 2 trial. Blood 2020, 135, 3733–3739. [CrossRef] [PubMed]
92. Herbaux, C.; Chessequieres, H.; Bouabdallah, R.; Guidez, S.; Gyan, E.; Gressin, R.; Morineau, N.; Ysebaert, L.; Le Guillou, S.; Laurent, C.; et al. Atezolizumab + nab-trastuzumab + venetoclax in patients with relapsed or refractory indolent non-Hodgkin’s lymphoma (R/R INHL): Primary analysis of a phase 2 trial from LYSA. J. Clin. Oncol. 2021, 39, 7544. [CrossRef]
93. Nowakowski, G.; Willenbacher, W.; Greil, R.; Larson, T.; Patel, K.; Jäger, U.; Manges, R.; Trümpfer, L.; Haidoun, C.; Everaerts, H.; et al. Safety and efficacy of the pd-1 inhibitor durvalumab with r-chop or r2-chop in subjects with previously untreated, high-risk dlbcl. Hematol. Oncol. 2019, 37, 132–134. [CrossRef]
94. Herrera, A.F.; Goy, A.; Mehta, A.; Ramchandren, R.; Pagel, J.M.; Svboda, J.; Guan, S.; Hill, J.S.; Kwei, K.; Liu, E.A.; et al. Safety and activity of ibritumomab tiuxetan in combination with durvalumab in patients with relapsed or refractory lymphoma or diffuse large B-cell lymphoma. Am. J. Hematol. 2020, 95, 18–27. [CrossRef]
95. Ansell, S.M.; Hurvitz, S.A.; Koenig, P.A.; LaPlant, B.R.; Kabat, B.F.; Fernando, D.; Habermann, T.M.; Inwards, D.J.; Verma, M.; Yamada, R.; et al. Phase I Study of ipilimumab, an Anti-CTLA-4 Monoclonal Antibody, in Patients with Relapsed and Refractory B-Cell Non–Hodgkin’s Lymphoma. *Clin. Cancer Res.* 2009, 15, 6446–6453. [CrossRef] [PubMed]

96. Tuscano, J.M.; Maveralakis, E.; Groschen, S.; Tsao-Wei, D.; Luxardi, G.; Merleev, A.A.; Beaven, A.; DiPersio, J.F.; Popplewell, L.; Chen, R.; et al. A Phase I Study of the Combination of Rituximab and Ipilimumab in Patients with Relapsed/Refractory B-Cell Lymphoma. *Clin. Cancer Res.* 2019, 25, 7004–7013. [CrossRef] [PubMed]

97. Khouri, I.F.; Caballero, J.; Bassett, R.L.; Allison, J.P.; Gulbis, A.M.; Sharma, P.; Turturro, F.; Jabbour, E.J.; Milton, D.R.; Vence, L.M. Ipilimumab plus Lenalidomide after Allogeneic and Autologous Stem Cell Transplantation for Patients with Lymphoid Malignancies. *Clin. Cancer Res.* 2017, 24, 1011–1018. [CrossRef]

98. Advani, R.; Flinn, I.; Popplewell, L.; Forero, A.; Bartlett, N.L.; Ghosh, N.; Kline, J.; Roschewski, M.; LaCasce, A.; Collins, G.P.; et al. CD47 Blockade by Hu5F9-G4 and Rituximab in Non-Hodgkin’s Lymphoma. *N. Engl. J. Med.* 2018, 379, 1711–1721. [CrossRef]

99. Goebeler, M.-E.; Knop, S.; Viardot, A.; Kufer, P.; Topp, M.S.; Einsele, H.; Noppeney, R.; Hess, G.; Kallert, S.; Mackensen, A.; et al. Bispecific T-Cell Engager (BiTE) Antibody Construct Blinatumomab for the Treatment of Patients With Relapsed/Refractory Non-Hodgkin Lymphoma: Final Results From a Phase I Study. *J. Clin. Oncol.* 2016, 34, 1104–1111. [CrossRef]

100. Viardot, A.; Goebeler, M.-E.; Hess, G.; Neumann, S.; Pfreundschuh, M.; Adrian, N.; Zettl, F.; Libicher, M.; Sayehli, C.; Stieglmaier, R.; et al. Phase 2 study of the bispecific T-cell engager (BiTE) antibody blinatumomab in relapsed/refractory diffuse large B-cell lymphoma. *Blood* 2016, 127, 1410–1416. [CrossRef]

101. Coyle, L.; Morley, N.J.; Rambaldi, A.; Mason, K.D.; Verhoef, G.; Furness, C.L.; Zhang, A.; Jung, A.S.; Cohan, D.; Franklin, J.L. Open-Label, phase 2 study of blinatumomab as second salvage therapy in adults with relapsed/refractory aggressive B-cell non-Hodgkin lymphoma. *Leuk. Lymphoma* 2020, 61, 2103–2112. [CrossRef]

102. Budde, L.E.; Ghosh, N.; Chavez, J.C.; Lossos, I.S.; Mehta, A.; Dorritie, K.A.; Kamdar, M.K.; Negricea, R.; Pham, S.; Hristopoulos, J.; et al. Promising tolerability and efficacy results from dose-escalation in an ongoing phase IIb/II study of mosunetuzumab (M) with polatuzumab vedotin (Pol) in patients (pts) with relapsed/refractory (R/R) B-cell non-Hodgkin’s lymphoma (B-NHL). *J. Clin. Oncol.* 2020, 38, 3913–3922. [CrossRef]

103. Clausen, M.R.; Luchi, D.; Armstrong, P.; Johnson, P.W.; Linton, K.M.; Lewis, D.J.; Chamuleau, M.; Balari, A.S.; Cunningham, D.; Elliott, B.; et al. Subcutaneous epcoritamab in patients with relapsed/refractory B-cell non-Hodgkin lymphoma: Safety profile and antitumor activity. *J. Clin. Oncol.* 2019, 37, 5954–5962. [CrossRef]

104. Buckingham, C.; Aribisala, S.; Lele, P.; Xiong, J.; Callander, S.; Harrington, K.; Oldham, J.; Miceli, S.; Farewell, V.T.; et al. Long-term follow-up of patients treated on the phase II L-MIND study of tafasitamab (MOR208) plus lenalidomide in patients with relapsed or refractory diffuse large B-cell lymphoma. *Haematologica* 2020, 105, 828–839. [CrossRef]

105. Chen, R.; et al. A Phase I Study of the Combination of Lenalidomide and Rituximab in Patients with Relapsed/Refractory B-Cell Lymphoma. *J. Clin. Oncol.* 2021, 39, 5708–5717. [CrossRef]

106. Diefenbach, C.S.M.; Abrisqueta, P.; Gonzalez-Barca, E.; Panizo, C.; Perez, J.M.A.; Miall, F.; Bastos-Oreiro, M.; Lopez-Guillermo, A.; Banerjee, L.; McMillan, A.; et al. Polatuzumab vedotin (Pol) + rituximab (R) + lenalidomide (Len) in patients (pts) with relapsed/refractory (R/R) diffuse large B-cell lymphoma (DLBCL): Primary analysis of a phase Ib/2 trial. *J. Clin. Oncol.* 2021, 39, 7512. [CrossRef]

107. Caimi, P.F.; Ai, W.; Alderuccio, J.P.; Ardestan, M.; Hamadani, M.; Hess, B.; Kahl, B.S.; Radford, J.; Solh, M.; Stathis, A.; et al. Loncastuximab tesirine in relapsed or refractory diffuse large B-cell lymphoma (LOTIS-2): A multicentre, open-label, single-arm, phase 2 trial. *Lancet Oncol.* 2021. [CrossRef]

108. Oliai, C.; De Vos, S. Case Report: Sustained Remission Achieved from Anti-CD19 CAR T Cell Therapy Despite Prior Treatment with Anti-CD19 Antibody Tafasitamab (MOR208) in a Patient with Relapsed and Refractory Diffuse Large B-Cell Lymphoma. *Blood* 2019, 134, 5360. [CrossRef]

109. Thapa, B.; Caimi, P.F.; Ardestan, M.; Solh, M.; Carlo-Stella, C.; Kahl, B.S.; Hamadani, M. CD19 antibody-drug conjugate therapy in DLBCL does not preclude subsequent responses to CD19-directed CAR T-cell therapy. *Blood Adv.* 2020, 4, 3850–3852. [CrossRef] [PubMed]