TGF-beta 1 levels are associated with lymphocyte percentages in patients with lung cancer treated with radiation therapy

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Purpose: Plasma TGF-β1 protein levels reportedly may predict the treatment outcomes of lung cancer. We hypothesized that in patients with lung cancer treated with radiation therapy (RT), TGF-β1 levels may correlate with the percentages of CD4+ T cells, CD8+ T cells, and the CD4+/CD8+ T cell ratio in peripheral blood.

Patients and methods: Eighty-two lung cancer patients satisfied the inclusion criteria. Platelet-poor plasma was obtained before RT, at the second and fourth weeks during RT, and at the end of RT (pre-, during-, and post-RT, respectively). TGF-β1 was measured via ELISA, while recording the percentages of lymphocyte subsets in peripheral blood. Short-term efficacy was categorized as complete response, partial response, stable disease, or progressive disease.

Results: Patients who had significantly lower TGF-β1 protein levels after RT than pre-RT seemed to have a better short-term effect (*P*<0.05) than those who had higher TGF-β1 levels. There was a significant association between the TGF-β1 levels and percentages of CD4+ T cells, CD8+ T cells, or CD4+/CD8+ T cell ratio during and at the end of RT. Changes in CD3+ T cells, B cells, or natural killer cells were not statistically related to the changes in TGF-β1 levels.

Conclusion: Lung cancer patients with TGF-β1 levels in plasma after RT that are below pre-RT levels may experience better short-term efficacy. The underlying mechanism may be related to the influence of TGF-β1 on antitumor immunity.

Keywords: lung cancer, radiotherapy, TGF-β1, lymphocytes, prognosis

Introduction

Lung cancer is the leading cause of cancer-related death in the world. It has been reported that TGF-β1 levels in the lung cancer tissues of patients, or in plasma, may be associated with prognosis of the disease. In locally advanced non-small-cell lung cancer (NSCLC), an increase in TGF-β1 levels in plasma during radiation therapy (RT) indicates a poorer prognosis. This may relate to immune escape or suppression induced by TGF-β1 at the end of RT. To the best of our knowledge, no studies have investigated the possible associations between plasma TGF-β1 levels and the lymphocytes CD4+ T cells, CD8+ T cells, or CD4+/CD8+ T cell ratio.

This study investigated the prognostic significance of plasma TGF-β1 levels during RT in the treatment of lung cancer, and the possible correlations during the course of RT between TGF-β1 levels and CD4+ T cell or CD8+ T cell levels, or the CD4+/CD8+ T cell ratio.
Materials and methods

Study design
Ethical approval for this investigation was obtained from the Research Ethics Committee, Tianjin Cancer Hospital & Institute. In brief, 82 patients with lung cancer were included in the final analysis. Blood samples were collected at the following timepoints: within a week prior to receiving RT (pre-RT), at the second and fourth weeks during RT (2 weeks during-RT or 4 weeks during-RT), and within a week after RT (post-RT). The blood samples were analyzed for CD4+ T cells, CD8+ T cells, B cells, and natural killer (NK) cells, and the CD4+/CD8+ T cell ratio was calculated. TGF-β1 levels were investigated for associations with CD4+ T cell or CD8+ T cell levels, or CD4+/CD8+ T cell ratio.

The response to RT treatment was evaluated by chest computed tomography (CT) image 1 month after RT, and classified as a complete or partial response, or stable or progressive disease. The short-term efficacy of treatment was then categorized as effective (complete or partial response) or ineffective (stable or progressive disease). The response rates were evaluated to determine the prognostic value of plasma TGF-β1 levels during RT.

Patients
We identified all patients with lung cancer, confirmed and treated at Tianjin Cancer Hospital, between March 1, 2016, and May 1, 2017. After appropriate eligibility was established, 82 patients with lung cancer participated. All patients involved in this study were informed of the content of the research and provided written informed consent.

All the participating patients had lung cancer, confirmed by pathology or cytology in Tianjin Cancer Hospital. Excluded from the analysis was any patient with medical complications or serious infectious diseases that could affect immune function; obvious contraindications for intensity-modulated RT; malignant tumors other than lung cancer; receiving immune therapy or taking immunosuppressive drugs within the previous 3 months; incomplete medical records; or who did not comply with the criteria mentioned above, gave up the treatment for various reasons, or were otherwise lost to follow-up.

Radiotherapy
All patients received RT with or without sequential or concurrent chemotherapy. Radiation was given via consistent intensity-modulated RT. Treatment planning was performed with a Philips Pinnacle+ radiation treatment planning system (Philips Medical Systems, Amsterdam, Netherlands).

For all patients, gross tumor volume was identified based on the CT images. The gross tumor volume included the tumor and the metastatic lymph nodes. The clinical target volume was based on the gross tumor volume, but also included the primary tumor bed and metastatic lymph nodes before chemotherapy. The distance from the margin of the gross tumor volume to the clinical target volume was 5 mm. The distance from the margin of the clinical target volume to the planning target volume was 5–10 mm. The radiation dose was 50–66 Gy in 20–33 fractions, 1.8–3 Gy per fraction, 1 fraction per day.

Chemotherapy
Chemotherapy was given before, during, or after RT, or at combinations of these intervals. When chemotherapy was given sequentially with RT, small-cell lung cancer (SCLC) patients received chemotherapy of etoposide and cisplatin, or etoposide and carboplatin. NSCLC patients received chemotherapy of platinum-based doublets (carboplatin or cisplatin combined with vinorelbine, paclitaxel, or gemcitabine). When chemotherapy was given concurrently, the combination of carboplatin or cisplatin and etoposide or paclitaxel was commonly used. The dosage of all the chemotherapy drugs was within normal limits. A chemotherapy cycle was considered to be 21 days. The median number of chemotherapy cycles was 4.

Sample collection and TGF-β1 measurement
Blood samples were collected, with dipotassium EDTA as the anticoagulant, at 3 timepoints as described above in the study design. Blood samples were placed on ice immediately after collection. Blood samples were centrifuged within 2 hours of collection at 3,000×g for 30 minutes. The upper one-third of the supernatants was collected and stored at −80°C. Plasma TGF-β1 levels were then measured by enzyme-linked ELISA using a Quantikine ELISA Kit (R&D Systems, Inc., Minneapolis, MN, USA). The presence of CD4+ T cells, CD8+ T cells, CD4+/CD8+ T cell ratio, B cells, and NK cells was tested by the Department of Clinical Laboratory, Tianjin Cancer Hospital. FACSCanto™ II flow cytometer (BD Biosciences, San Jose, CA, USA) was used to finish the measurement of lymphocyte percentages.

Criteria for evaluating therapeutic effect
The response rate was evaluated by chest CT image, 1 month after RT. In accordance with the Response Evaluation Criteria in Solid Tumors version 1.1, short-term efficacy was
TGF-β1 are associated with lymphocyte percentages and radiotherapy

classified as a complete response, partial response, stable disease, or progressive disease. Immeasurable lesions (such as bone metastasis sites or malignant pleural effusion) were generally not evaluated, unless involved in disease progression. Effective treatment was defined as a complete or partial response after RT. Ineffective treatment was defined as stable disease or progressive disease after RT.

Statistical analysis
Clinical characteristics were compared using the chi-squared test. The Mann–Whitney U test was used to compare differences in TGF-β1 in plasma and lymphocytes, while two-tailed Student’s t-test was used to compare the same patients with different timepoints. Independent t-test and one-way ANOVA were used to compare differences in TGF-β1 between different clinical characteristics. Correlations were tested by Pearson’s correlation analysis. All P-values were two-sided. Data are presented as mean ± SD unless otherwise specified.

Results
Clinical characteristics of patients
In this prospective study, 104 patients with lung cancer were initially considered for inclusion, and subsequently 22 patients were excluded for the following reasons: 11 received a diagnosis of esophageal cancer; 5 were found with thymoma; and 6 were missing data. Thus, 82 patients were included in the final analysis; different clinical characteristics are listed in Table 1. In addition, patients with different clinical characteristics also had similar TGF-β1 levels (Table 1).

Association between TGF-β1 levels and short-term response to treatment
Of the whole group, 47 (57%) achieved an effective response (CR+PR), and 35 (43%) achieved an ineffective response (SD+PD; Table 2). At different timepoints, TGF-β1 levels varied. Furthermore, patients with SCLC and NSCLC also had different TGF-β1 levels (Table 2). The TGF-β1 levels of the patients before RT were comparable (P<0.05; Table 2). After RT, the mean TGF-β1 levels of the patients who had shown an effective response were significantly lower than that of the patients for whom RT was ineffective (P<0.05; Table 2).

Overall, 47 patients showed an effective response to RT, while treatment was ineffective for 35 patients (Table 3). Of those with an effective response, the majority (36/47) had TGF-β1 levels after RT that were lower than the pre-RT levels. The TGF-β1 levels of most of the patients who showed

| Table 1 Clinical characteristics and TGF-β1 levels (ng/mL) of the 82 patients |
|-----------------|-----------------|-----------------|-----------------|-----------------|
| Gender          | n   | TGF-β1<sup>a</sup> | P-value<sup>a</sup> | TGF-β1<sup>b</sup> | P-value<sup>b</sup> |
| Male            | 51  | 2.81±0.42           | 0.827            | 2.52±0.47         | 0.274         |
| Female          | 31  | 3.02±1.07           |                 | 3.81±1.37         |               |
| Age (years)     |     |                   |                 |                 |               |
| ≥65             | 30  | 3.32±0.83           | 0.483            | 4.13±1.30         | 0.093         |
| <65             | 52  | 2.69±0.45           |                 | 2.33±0.43         |               |
| Histological type |    |                   |                 |                 |               |
| SCLC            | 21  | 2.74±0.46           | 0.078            | 2.83±0.67         | 0.247         |
| Adenocarcinoma  | 30  | 1.57±0.18           |                 | 4.68±1.68         |               |
| SCC             | 30  | 3.41±0.83           |                 | 2.41±0.65         |               |
| Others          | 1   | 1.30                |                 | 1.48              |               |
| Stage           |     |                   |                 |                 |               |
| I               | 1   | 1.14                | 0.053            | 0.81              | 0.389         |
| II              | 36  | 3.31±1.68           |                 | 2.14±0.64         |               |
| III             | 41  | 2.55±0.36           |                 | 2.90±0.58         |               |
| IV              | 4   | 4.40±1.65           |                 | 3.22±1.70         |               |
| Radiation dose, Gy |     |                   |                 |                 |               |
| ≥60             | 76  | 2.80±0.40           | 0.362            | 2.80±0.48         | 0.858         |
| <60             | 6   | 5.08                |                 | 2.26              |               |
| Chemotherapy cycles |     |                   |                 |                 |               |
| ≥4              | 46  | 2.40±0.38           | 0.098            | 2.01±0.37         | 0.017         |
| <4              | 36  | 3.77±0.85           |                 | 4.35±1.10         |               |

Notes: <sup>a</sup>Pre-RT. <sup>b</sup>Post-RT. TGF-β1 levels did not show statistical significance of the 82 patients in different clinical characteristics.

Abbreviations: RT, radiation therapy; SCC, squamous cell carcinoma; SCLC, small-cell lung cancer.
an ineffective response (27/35) were higher after RT relative to pre-RT. In the SCLC group specifically, 12 and 9 patients, respectively, experienced effective and ineffective responses to RT (Table 3). Of those who demonstrated an effective response, the majority (11/12) had TGF-β1 levels after RT that were lower than the pre-RT levels. The TGF-β1 levels of most of the SCLC patients who showed an ineffective response (6/9) were higher after RT relative to pre-RT. In the NSCLC group, 35 and 26 patients experienced effective and ineffective responses to RT, respectively (Table 3). Of those who demonstrated an effective response, the majority (25/35) had TGF-β1 levels after RT that were lower than pre-RT levels. The TGF-β1 levels of most of the SCLC patients who showed an ineffective response (21/26) were higher after RT relative to pre-RT.

TGF-β1 levels and lymphocytes

Overall, the TGF-β1 levels decreased only slightly from pre-RT to post-RT, while the percentages of CD3+ T cells and B cells were significantly lower at post-RT (both, P<0.01; Table 4). In addition, compared with pre-RT levels, at post-RT the CD4+ T cells and the CD4+ /CD8+ ratio were only slightly higher; CD8+ T cells were higher (P<0.05; Table 4); and NK cells remained stable.

At the second and fourth weeks during RT, and at the end of RT, TGF-β1 levels were significantly associated with the percentages of CD4+ T cells; CD8+ T cells; and the CD4+ /CD8+ ratio (P<0.05, all; Table 5). There were no significant associations between TGF-β1 levels and percentages of CD3+ T cells, B cells, or NK cells (Table 5).

**Discussion**

TGF-β is a member of a multifunctional cytokine family, and TGF-β1 is considered to participate in carcinogenesis by promoting cell proliferation, differentiation, and extracellular matrix production. Studies have shown that TGF-β1 levels may be a marker of patient prognosis; patients with tumors who had higher TGF-β1 levels after treatment compared with TGF-β1 levels before treatment seemed to have a significantly poorer overall prognosis. This may be due to the immune suppression effect of TGF-β1. Huang et al observed that high TGF-β1 protein levels were associated with a poor prognosis. What is more, studies have reported that chemotherapy may affect TGF-β1 levels. Our present small-sample study suggests that patients with decreased TGF-β1 levels at the end of RT have a higher response rate, which is in accord with the reported literature. Patients who received more than 4 cycles of chemotherapy had lower TGF-β1 levels compared with those who received fewer than 4 cycles.

The changes of lymphocytes could also be a prognosis biomarker. Results from Yang showed that the CD4+ /CD8+ ratio counts were consistently higher in prostate cancer
patients with a better response to RT while CD3+ and CD8+ cell counts were lower. These results are in accord with our research and indicated that variations in peripheral lymphocyte subpopulations are predictive of outcome after RT. Another research from Spain\(^{19}\) indicated that in prostate cancer patients undergoing RT, in vitro radiation-induced apoptosis of CD4+ T lymphocytes assessed before RT was associated with the probability of developing chronic genitourinary toxicity, and radiation-induced apoptosis of CD8+ T lymphocytes was associated with overall survival. In our study, instead of analyzing the probability of T lymphocytes as a prognostic biomarker, we focused on the relationship between TGF-β1 levels and T lymphocytes and hypothesized that the underlying mechanism of TGF-β1 could reflect treatment outcome that may be related to the immunosuppression effect of T lymphocytes. Recent studies have indicated that the specific immunologic microenvironment of tumors may be crucial to carcinogenesis and anti-tumorigenesis,\(^{19}\) and T cell-mediated cellular immunity may be an important mechanism for killing tumor cells.\(^{20–22}\) Results from Sakaguchi et al\(^{23}\) showed that TGF-β1 and interleukin 2 (IL-2) induced tumor tissues to produce more Treg (regulatory T) cells, which can inhibit the cytotoxic effect of cells, thereby impairing the immune system.\(^{22–25}\) Strauss et al\(^{26}\) reported that Treg cells with a distinct phenotype in tumor-infiltrating lymphocytes could produce TGF-β, which contributed to local immune suppression. There can be no doubt that radiotherapy can induce the suppression of immunity,\(^{27}\) with significant decline in T lymphocyte levels.\(^{28}\) In the present study, we found that after RT the percentage of CD3+ T cells and B cells had significantly decreased, but the percentage of CD8+ T cells significantly increased from pre-RT values. This suggests that the immune system, especially cellular immunity, was inhibited after RT. As for the CD4+ T cells, in our results, after radiotherapy it had a slight rise but did not achieve a statistical significance; therefore, we cannot determine the influence of the change of CD4+ T cells on the changes of whole T lymphocyte subsets. Large-sample clinical researches are urgently needed.

Over the course of RT treatment, TGF-β1 levels significantly and negatively correlated with the percentages of the CD4+ T cells and the CD4+/CD8+ ratio, but significantly and positively correlated with the percentages of CD8+ T cells. Previously, it was demonstrated that a decrease in

| Table 4 TGF-β1 levels and lymphocyte percentages of 82 patients at 4 timepoints |
|---------------------------------|-----------------|-----------------|-----------------|
|                                | Pre-RT           | During RT       | Post-RT         |
|                                | 2 weeks          | 4 weeks         |                 |
| TGF-β1, ng/mL                  | 2.8±0.38         | 2.85±0.40       | 2.51±0.39       | 2.74±0.46       |
| Lymphocytes, %                 |                 |                 |                 |
| CD3+ T cells                   | 69.1±1.71        | 68.60±1.81      | 65.63±1.64a     | 62.55±1.82a     |
| CD4+ T cells                   | 38.88±1.45       | 39.44±1.67      | 40.78±1.81      | 40.70±2.18      |
| CD8+ T cells                   | 26.45±1.52       | 28.00±1.69      | 26.73±1.39      | 30.23±2.56a     |
| CD4+/CD8+                      | 1.67±0.11        | 1.65±0.13       | 1.75±0.12       | 1.89±0.19       |
| B cells                        | 9.33±0.64        | 6.27±0.41a      | 7.35±0.68b      | 4.21±0.35a      |
| NK cells                       | 20.73±1.54       | 21.41±1.59      | 21.04±1.71      | 20.41±1.58      |

**Notes:** *Compared with pre-RT, P<0.01; 1, compared with pre-RT, P<0.05. The TGF-β1 levels decreased only slightly from pre-RT to post-RT, and the percentages of CD3+ T cells and B cells were significantly lower at post-RT. In addition, compared with pre-RT levels, at post-RT the CD4+ T cells and the CD4+/CD8+ ratio were only slightly higher; CD8+ T cells were higher; and NK cells remained stable.

**Abbreviations:** NK, natural killer; RT, radiation therapy.

| Table 5 Associations between TGF-β1 levels and lymphocyte percentages |
|---------------------------------|-----------------|-----------------|-----------------|-----------------|
|                                | During RT       |                 | Post-RT         |                 |
|                                | 2 weeks         | 4 weeks         |                 |                 |
| CD3+ T cells                   | −0.117          | 0.478           | −0.006          | 0.973           | 0.250           | 0.125           |
| CD4+ T cells                   | −0.581          | 0.023           | −0.516          | 0.037           | 0.011           | −0.648          | <0.001          |
| CD8+ T cells                   | 0.558           | 0.041           | 0.545           | 0.011           | 0.003           | 0.598           | <0.001          |
| CD4+/CD8+                      | −0.615          | 0.024           | −0.648          | 0.003           | 0.132           | 0.424           |                 |
| B cells                        | 0.042           | 0.8             | 0.056           | 0.736           | 0.073           | −0.298          | 0.066           |
| NK cells                       | −0.301          | 0.063           | −0.292          | 0.073           |                 |                 |                 |

**Notes:** At the second and fourth weeks during RT, and at the end of RT, TGF-β1 levels were significantly associated with the percentages of CD4+ T cells; CD8+ T cells; and the CD4+/CD8+ ratio. There were no significant associations between TGF-β1 levels and percentages of CD3+ T cells, B cells, or NK cells.

**Abbreviations:** NK, natural killer; r, correlation coefficient; RT, radiation therapy.
the CD4+/CD8+ ratio was an independent negative prognostic factor for survival in NSCLC patients. We previously showed that TGF-β1 levels negatively correlated with the CD4+/CD8+ ratio, and an increase in TGF-β1 accompanied by a decline in the CD4+/CD8+ ratio indicates a poor prognosis. This is consistent with previous reports.

**Conclusion**

Our research indicated that lung cancer patients with TGF-β1 levels in plasma after RT that are below pre-RT levels may experience better short-term efficacy. The underlying mechanism may be related to the influence of TGF-β1 on antitumor immunity.

All these results support the hypothesis that the underlying mechanism of TGF-β1 may be related to an influence on antitumor immunity, especially cellular immunity. Studies have reported that Treg cells express membrane-bound TGF-β1, which directly inhibited the functions of NK effector cells and downregulated NK cell receptors on the NK cell surface. Our present research suggested a weak correlation between TGF-β1 and NK cells, but unfortunately, the correlation was not statistically significant. Besides, in our study, one patient with large-cell neuroendocrine carcinoma (LCNEC) was included, considering the current treatment for LCNEC to be the same as NSCLC; therefore, the patient was just classified under the NSCLC group. We failed refining the pathological types into four types (SCLC, adenocarcinoma, squamous cell carcinoma, LCNEC), and hence further exploration is required.

**Disclosure**

The authors report no conflicts of interest in this work.

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