Taxonomic revision of *Mordellistena hirtipes* species complex with new distribution records (Insecta, Coleoptera, Mordellidae)

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Abstract
A taxonomic revision of species related to *Mordellistena hirtipes* Schilsky, 1895 is presented. Five species among the *M. hirtipes* complex are recognised: *M. hirtipes* Schilsky, 1895, *M. pseudohirtipes* Ermisch, 1965, *M. purpurascens* Costa, 1854, *M. balearica* Compte, 1985, and *M. irritans* Franciscolo, 1991. Descriptions, differential diagnoses, an identification key, and new distributional records are provided. Principal Component Analysis is performed for visualisation of differentiation between taxa. The following taxonomic acts are proposed: *Mordellistena podlussanyi* Czető, 1990 and *M. aega* Franciscolo, 1949 are proposed as junior subjective synonyms of *M. hirtipes* Schilsky, 1895; *M. fageli* Ermisch, 1969 and *M. pseudohirtipes krotosensis* Czető, 1990 are proposed as junior subjective synonyms of *M. pseudohirtipes pseudohirtipes* Ermisch, 1965; *M. geronensis* Ermisch, 1977 and *M. istrica* Ermisch, 1977 are proposed as junior subjective synonyms of *M. purpurascens* Costa, 1854.

Keywords
taxonomy, identification key, distribution, new synonym, morphometry, Principal Component Analysis (PCA)
**Introduction**

*Mordellistena* Costa, 1854 is the largest genus within the family Mordellidae Latreille, 1802, comprising approximately 800 described species (Horák 2011). Adults are commonly found on flowers where they feed on pollen and nectar. Larvae are found in the stems of herbaceous plants or in decaying wood.

The Western Palaearctic species are conventionally assigned to species groups proposed by Ermisch (1956, 1969b), based on combinations of morphological characters. There are no studies focused on the phylogeny of Palaearctic species of genus *Mordellistena*, and thus the phylogenetic relationships among its species remain unresolved.

The present study is focused on nine morphologically related taxa, which represent a complex within *M. confinis* species group (Ermisch 1956, 1969b, Batten 1977). The unique character shared by all species of the complex is the expanded second segment of maxillary palpi in males, bearing very long setae on the ventral surface (Figs 4A, 6C, 7C). Additional characters are a strongly convex body and unique shape of parameres (Figs 5D–G, 6E–H, 7G–J, 8, 9).

Species belonging to this complex were described by Costa (1854), Schilsky (1895), Ermisch (1965, 1969a, 1977), Compte (1985), and Franciscolo (1949, 1991). Most of descriptions appeared to be insufficient for proper identification and after examination of type material, it became clear that some of the taxa are conspecific. In the present paper, we provide redescriptions of *M. hirtipes* Schilsky, 1895, *M. pseudohirtipes* Ermisch, 1965, and *M. purpurascens* Costa, 1854. Important diagnostic characters are visualised in drawings and photographs. We also performed Principal Component Analysis (PCA) based on morphometric measurements. This method is widely used in taxonomical research of invertebrates to help separate putative species in difficult species-complexes, to visualise differentiation between species and to evaluate the importance of peculiar morphometric characters (Kucharczyk et al. 2012, Przybycień and Waclawik 2015).

Examination of material from several localities in Western Palaearctic revealed new distribution records and new biological information for *M. hirtipes* Schilsky, 1895, *M. pseudohirtipes* Ermisch, 1965, and *M. purpurascens* Costa, 1854.

**Materials and methods**

Dried specimens were relaxed in water with a few drops of acetic acid to allow for the dissection. Specimens were observed using Leica MZ16 stereomicroscope with magnification up to 120×, illuminated with diffuse light (neon bulb, 6400 K). Dissected body parts for drawings were temporarily mounted on slides in glycerine. Drawings were prepared using Leica drawing tube attached to Leica DM 1000 microscope, scanned and traced in Adobe Illustrator CS6. Dissected body parts were after examination mounted on the same card as respective specimen using dimethyl hydantoin formaldehyde (DMHF) or put to genitalia microvials filled with glycerine and pinned with the respective
specimen. Measurements were taken using ocular micrometre. Intervals of measured values are followed by data in parentheses: arithmetic mean ± standard deviation, n = number of measured specimens. Total length (TL) was measured from the anterior margin of pronotum to the apices of elytra; elytral length (EL) was measured from the apex of scutellar shield to the apices of the elytra; elytral width (EW) was measured at the widest point of elytra. Digital photographs were taken using Canon EOS 5D mark II camera attached to Zeiss Axio Zoom.V16 stereomicroscope. Image stacks were produced manually, combined using Zerene Stacker software and edited in Adobe Photoshop CC.

Terminology used in morphological descriptions follows Lawrence and Ślipiński (2010) and is supplemented by terminology used by Lu et al. (1997) for the genitalia.

Specimen data are given in the following format: number of specimens and sex, depository: exact data from labels in quotation marks; slash indicates separate labels; author’s remarks are given in square brackets.

Principal Component Analysis (PCA) was conducted in PAST 3.12 software (Hammer et al. 2001), based on variance-covariance matrix of 12 morphometric characters: HL, HW, PL, PW, EL, EW, PTiL, MsTiL, MrTiL, RPrL, BRPr, and LPrL. Measurements were taken from 59 male specimens (including holotypes / lectotypes) assigned to eight taxa (M. hirtipes Schilsky, 1895, M. podlussanyi Czető, 1990, M. aegera Franciscolo, 1949, M. pseudohirtipes pseudohirtipes Ermisch, 1965, M. pseudohirtipes krotosensis Czető, 1990, M. jageli Ermisch, 1969, M. geronensis Ermisch, 1977, and M. istrica Ermisch, 1977). Lectotype of M. purpurascens was not measured because of its bad condition. Plot created in PAST 3.12 was subsequently edited in Adobe Illustrator CS6.

All nomenclatorial acts follow regulations of ICZN (1999).

Overall 149 specimens from following depositories were examined:

CSB collection of Dávid Selnekovič, Bratislava, Slovakia
HNHM Hungarian Natural History Museum, Budapest, Hungary
MCST Museo Civico di Storia Naturale, Trieste, Italy
MNCN Museo Nacional de Ciencias Naturales, Madrid, Spain
MNHU Museum für Naturkunde der Humboldt Universität, Berlin, Germany
MZFN Museo Zoológico dell’Università Federico II, Naples, Italy
SNSD Senckenberg Naturhistorische Sammlungen, Dresden, Germany

Abbreviations of measured characters:

| Abbreviation | Description                        |
|--------------|------------------------------------|
| BLPr         | basal part of left paramere length |
| BRPr         | basal part of right paramere length|
| EL           | length of elytra                   |
| EW           | width of elytra (combined)         |
| HL           | length of head                     |
| HW           | width of head                      |
| LabL         | length of labrum                   |
| LabW         | width of labrum                    |
| LPrL         | length of left paramere            |
| MsTiL        | length of mesotibiae               |
| MsTrL        | length of mesotarsi                |
| MtTiL        | length of metatibiae               |
Data resources

The data underpinning the analyses reported in this paper are deposited at GBIF, the Global Biodiversity Information Facility, https://doi.org/10.15468/pkhkul

Taxonomy

*Mordellistena* Costa, 1854

*Mordellistena* Costa, 1854: 16 [type species: *Mordellistena confinis* Costa, 1854: 18]

*Mordellistena birtipes* species complex

**Diagnosis.** Integument including legs and maxillary palpi completely black; metatibial spurs black; pubescence of dorsum yellowish, sometimes darkened in apical portions of elytra but never completely dark. Antennomeres I–IV shorter and narrower than following ones (Figs 5A, B, 6A, B, 7A, B). Maxillary palpomere II expanded in males, bearing very long setae on ventral surface (Figs 4A, 6C, 7C). Metatibiae at least with three lateral ridges, all parallel to apical margin of tibia. Metatarsomeres I and II with ridges.

*Mordellistena (s. str.) birtipes* Schilsky, 1895

Figs 1, 4A, B, 5A–J

*Mordellistena birtipes* Schilsky, 1895: 46 (original description); Heyden et al. 1906: 455 (catalogue); Csiki 1915: 35 (catalogue); Schaufuss 1916: 766 (distribution); Ermisch 1963: 62 (distribution); Ermisch 1965: 268 (distribution); Batten 1976: 168 (distribution); Batten 1977: 171–173 (distribution, figures, key); Horák 1990: 136 (lectotype and paralectotypes designation, figures); Odnosum 2003: 36–37, 40, 46 (key, figures, distribution); Horák 2008: 98 (catalogue, distribution); Odnosum 2010: 153, 192–194 (key, description, figures, distribution); Samin et al. 2016: 24 (distribution); Ruzzier et al. 2017: 152 (distribution).

| Abbreviation | Description |
|--------------|-------------|
| MtTrL        | length of metatarsi |
| PL           | length of pronotum |
| PTiL         | length of protibiae |
| PTrL         | length of protarsi |
| PW           | width of pronotum |
| PygL         | length of pygidium |
| RPrL         | length of right paramere |
| St8L         | length of sternite VIII |
| St8W         | width of sternite VIII |
| TVtL         | length of terminal abdominal ventrite |
| TL           | total length |
| TPalL        | length of terminal segment of maxillary palpi |
| TPalW        | width of terminal segment of maxillary palpi |
Figures 1–3. 1 Mordellistena hirtipes Schilsky, 1895, male 2 M. pseudohirtipes Ermisch, 1965, male 3 M. geronensis Ermisch, 1977, male. Scale bar: 0.50 mm.

*Mordellistena aegea* Franciscolo, 1949: 90, 93 syn. nov. (original description); Batten 1977: 169 (remarks); Horák 2008: 96 (catalogue).

*Mordellistena podlussanyi* Czető, 1990: 26–29 syn. nov. (original description); Horák 2008: 100 (catalogue).

**Type locality.** Attalia [Turkey].

**Type material examined.** *M. hirtipes*: LECTOTYPE (by designation of Horák (1990: 136)): 1 ♂, MNHU: “Attalia Reitter [hand written] / hirtipes Schils. [hand written] / Type [red label] / Zool. Mus. Berlin / [card with dissected genitalia] / LECTOTYPUS [red label] / Mordellistena hirtipes Schils. J. Horák det. 1985”; PARALECTOTYPES (by designation of Horák (1990: 136)): 4 ♂♂, 3 ♀♀, MNHU: “Attalia Reitter [hand written] / Coll. Schilsky / Type [red label] / Zool. Mus. Berlin / PARALECTOTYPUS [red label] / Mordellistena hirtipes Schils. J. Horák det. 1985”; 17 ♂♂, 9 ♀♀, MNHU: “♂ [or] ♀ / Coll. Schilsky / Type [red label] / Zool. Mus. Berlin / PARALECTOTYPUS [red label] / Mordellistena hirtipes Schils. J. Horák det. 1985”; 1 ♀, MNHU: “Syrien Kaifa.
Reitter. / Coll. Schilsky / Type [red label] / Zool. Mus. Berlin / PARALECTOTYPUS [red label] / Mordellistena hirtipes Schils. J. Horák det. 1985”; 1 ♀, MNHU: “Morea Hagios Wlassis Brenske / hirtipes [hand written] / Type [red label] / Zool. Mus. Berlin / PARALECTOTYPUS [red label] / Mordellistena hirtipes Schils. J. Horák det. 1985”.

*M. aegea*: HOLOTYPE: 1 ♂, MCST: “Pod. Sper. Coo 7. VII.–VIII. [Podere sperimentale, Kos Island; hand written] / 7. VII.–VIII. Pod. Sper. Coo [hand written] / 19 [blue label] / [card with dissected median lobe] / Olo [hand written] Typus / [cover slides with dissected parameres and sternite VIII] / Mordellistena aegea n. sp. DET. FRANCESCOLO / HOLOTPUS Mordellistena aegea Franciscolo, 1949 D. Selnekovič labelled 2018 / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selnekovič det. 2018”.

*M. podlussanyi*: HOLOTYPE: 1 ♂, HNHM: “♂ / [transparent plastic board with dissected genitalia] / Krotos KRÉTA / 1981. V. 12. leg. Podlussány / Holotypus Mordellistena podlussanyi Czető, 1988 [red label, hand written] / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selnekovič det. 2017”.

**Additional material examined. Croatia:** 1 ♂, HNHM: “Dalmatia leg. Endrödy-Younga / Dubrovnik Ins. Lokrum / 1958. VIII. 7. Kätscher / Mordellistena hirtipes Schils. det. R. Batten 1979 / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selnekovič det. 2017”.

**Cyprus:** 1 ♂, HNHM: “Cyprus Laranka Glaszner / Mordellistena hirtipes Schils. det. R. Batten 1979 / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selnekovič det. 2017”; 12 ♂♂, 2 ♀♀, CSB: “Cyprus W, Limassol env., Germasogeia Reservoir 34°45’19”N, 33°05’36”E, 27. IV. 2018 D. Selnekovič leg. / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selnekovič det. 2018”.

**Greece:** 1 ♂, SNSD: “♂ / Insel Rhodos 24.5.–5.8.62 /
Stadt Rhodos Umg. Dr Mand / Genitalpräparat / MORDELLISTENA pseudohirtipes Erm. K. Ermisch det. 19 / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selneković det. 2017”; 1 ♂, HNHM: “Crete Biró / Ins. Dia 25.–29. V. / Mordellistena hirtipes Schils. det. R. Batten 1980 / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selneković det. 2017”. Montenegro: 3 ♂♂, 3 ♀♀, CSB: “Montenegro SW Bar city env. 42°07’56”N, 19°07’33”E, 22. VI. 2011 D. Selneković / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selneković det. 2012”. 2 ♂♂, 3 ♀♀, CSB: “Montenegro SW Bar city–Volujica hill 242°04’16”N, 19°06’10”E, 20. VI. 2011 D. Selneković / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selneković det. 2012”. 2 ♂♂, 2 ♀♀, CSB: “Montenegro SW Bar city–Stari Bar 42°05’31”N, 19°07’58”E, D. Selneković 19. VI. 2011 / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selneković det. 2012”. 1 ♂, CSB: “Montenegro SW Bar city, on Daucus 42°06’N, 19°06’E, 19. VI. 2011 D. Selneković 2011 / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selneković det. 2012”.

Spain: 1 ♂, 1 ♀, SNSD: “Spanien, Prov. Gerona Tossa de mar A. Kampf, VII–VIII 35 / Paratypus Mordellistena geronensis Ermisch / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selneković det. 2017”. 1 ♂, SNSD: “Nordostspanien Costa brava 27. 7. 53 Dr David / Paratypus / PARATYPUS Mordellistena (s. str.) geronensis Ermisch, 1977 Selneković labelled 2017 [red label] / Mordellistena (s. str.) hirtipes Schilsky, 1895 D. Selneković det. 2017”.

Differential diagnosis. Parameres of *M. hirtipes* are shorter in proportion to the body dimensions than in *M. pseudohirtipes* and *M. purpurascens* (EL/LPrL ratio in *M. hirtipes*: 7.87–9.17 (8.48 ± 0.40, n = 14), *M. pseudohirtipes*: 4.65–7.17 (5.89 ± 0.71, n = 25) and *M. purpurascens*: 4.42–5.84 (4.98 ± 0.35, n = 19); EL/RPrL ratio in *M. hirtipes*: 10.07–11.89 (11.10 ± 0.50, n = 14), *M. pseudohirtipes*: 5.91–8.63 (7.42 ± 0.72, n = 25) and *M. purpurascens*: 5.57–6.94 (6.19 ± 0.41 n = 19). Ventral branch of the right paramere is in *M. hirtipes* (Fig. 5D–G) usually distinctly shorter than the dorsal one whereas in *M. pseudohirtipes* (Fig. 6E–H) and *M. purpurascens* (Fig. 7G–J) it is subequal or longer. Basal part of the left paramere is in *M. hirtipes* (Fig. 5D–G) distinctly shorter than in *M. purpurascens* (Fig. 7G–J). Sides of elytra less convergent apically than in *M. pseudohirtipes* and *M. purpurascens*. Terminal maxillary palpomere in females shorter and broader, its inner angle is more acute (Fig. 4B) than in *M. pseudohirtipes* (Fig. 6D) and *M. purpurascens* (Fig. 7D).

Redescription. Measurements: TL: ♂♂ 3.21–3.95 mm (3.51 ± 0.24 mm, n = 13), ♀♀ 2.79–4.68 mm (3.44 ± 0.52 mm, n = 9); HL: ♂♂ 0.72–0.93 mm (0.81 ± 0.06 mm, n = 14), ♀♀ 0.67–0.80 mm (0.74 ± 0.04 mm, n = 9); HW: ♂♂ 0.87–1.02 mm (0.94 ± 0.05 mm, n = 14), ♀♀ 0.77–0.96 mm (0.87 ± 0.06 mm, n = 9); PL: ♂♂ 1.04–1.33 mm (1.17 ± 0.11 mm, n = 14), ♀♀ 0.94–1.20 mm (1.09 ± 0.08 mm, n = 9); PW: ♂♂ 1.06–1.37 mm (1.23 ± 0.10 mm, n = 14), ♀♀ 0.98–1.29 mm (1.16 ± 0.10 mm, n = 9); EL: ♂♂ 2.34–2.96 mm (2.65 ± 0.19 mm, n = 14), ♀♀ 2.08–2.79 mm (2.48 ± 0.21 mm, n = 9); EW: ♂♂ 1.10–1.43 mm (1.25 ± 0.11 mm, n = 14), ♀♀ 1.10–1.36 mm (1.23 ± 0.09 mm, n = 9); PTiL: ♂♂ 0.70–0.87 mm (0.77 ± 0.05 mm, n = 14), ♀♀ 0.56–0.73 mm (0.65 ± 0.06 mm, n = 9); PTrL: ♂♂ 0.65–0.74 mm (0.71 ± 0.03 mm, n = 11), ♀♀ 0.57–0.66 mm (0.63 ± 0.03 mm, n = 7); MsTiL: ♂♂ 0.83–1.10 mm (0.97 ± 0.08 mm, n = 14), ♀♀ 0.78–1.01 mm (0.87 ± 0.07 mm, n = 9); MsTrL: ♂♂ 1.06–1.30 mm (1.16 ± 0.06 mm, n = 11), ♀♀ 0.91–1.13 mm (1.05
Figure 5. *Mordellista hirtipes* Schilsky, 1895: A antenna, male B antenna, female C labial palpus, female D parameres, lectotype E parameres, holotype of *M. podlussanyi* F parameres, Greece G parameres, Spain H aedeagal median lobe I sternite VIII, male J sternite VIII, female.
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± 0.07 mm, n = 9); MtTiL: ♂♂ 0.69–0.91 mm (0.81 ± 0.05 mm, n = 14), ♀♀ 0.66–0.86 mm (0.76 ± 0.06 mm, n = 9); MtTrL: ♂♂ 1.48–1.87 mm (1.67 ± 0.11 mm, n = 10), ♀♀ 1.33–1.69 mm (1.51 ± 0.11 mm, n = 9); PygL: ♂♂ 1.42–1.85 mm (1.58 ± 0.12 mm, n = 13), ♀♀ 1.17–1.56 mm (1.37 ± 0.12 mm, n = 9); TTrL: ♂♂ 0.54–0.87 mm (0.68 ± 0.10 mm, n = 13), ♀♀ 0.44–0.79 mm (0.63 ± 0.11 mm, n = 9); LPrL: 0.29–0.35 mm (0.31 ± 0.02 mm, n = 14); RPrL: 0.22–0.26 mm (0.24 ± 0.01 mm, n = 14); St8L: ♂♂ 0.57–0.63 mm (n = 2); St8W: ♂♂ 0.38–0.40 mm (n = 2).

Habitus illustrated in Fig. 1. Body slender, widest at the end of anterior third of elytra. Integument black. Head and pronotum covered with yellowish pubescence; pubescence on elytra yellowish in proximal half, gradually darkened towards apices, sometimes with reddish or violet metallic sheen; venter covered with yellowish pubescence, darkened along posterior margins of ventrites 3–5.

Head moderately convex dorsally, wider than long, widest before middle, HW/HL ratio: ♂♂ 1.10–1.23 (1.17 ± 0.04, n = 14), ♀♀ 1.13–1.23 (1.17 ± 0.03, n = 9). Dorsal surface with microreticulation and small round punctures bearing short setae; ventral surface with transverse microreticulation and sparse, small punctures bearing short setae; small medial triangular part before gula without punctures. Occipital margin rounded in dorsal aspect, straight or slightly concave if seen from behind. Eyes oval, finely faceted with short interfacetal setae. Anterior margin of clypeus straight. Labrum transverse, approximately two times as wide as long, anterior margin straight; surface with microreticulation and small, round punctures bearing short setae. Antennae rather long, slightly serrate (Fig. 5A, B); antennomeres I–IV subequal in length, slightly shorter and slenderer than following ones; antennomeres V–X in males 1.40–1.60×, in females 1.20–1.30× as long as wide; antennomere XI oval, ~2.30× as long as wide. Mandibles symmetrical, bidentate, lateral portions microreticulated with short setae, outer distal portion with group of seven long sensilla; mola well developed, minutely dentate; prostheca well developed, setose. Galea gradually expanded toward apex, covered with apically expanded sensilla; lacinia setose medio-apically, reaching half of length of galea. Maxillary palpomere II distinctly expanded with long setae on ventral side in males (Fig. 4A); not expanded, without long setae in females (Fig. 4B); maxillary palpomere III short, ~1.50× as long as wide; terminal maxillary palpomere broadly secuiform, inner angle situated around middle, TPalL/TPalW ratio: ♂♂ 1.95–2.20 (2.09 ± 0.09, n = 14), ♀♀ 1.95–2.20 (2.10 ± 0.08, n = 9). Terminal labial palpomere fusiform, bearing sparse long sensilla on whole surface, and group of short sensilla at apex (Fig. 5C).

Pronotum moderately convex, approximately as long as wide, widest just behind middle, PW/PL ratio: ♂♂ 1.00–1.09 (1.05 ± 0.03, n = 14), ♀♀ 1.02–1.07 (1.06 ± 0.02, n = 9). Surface finely microreticulate, with small, rasp-like punctures bearing flat seta. Anterior margin rounded, slightly produced mesally, anterior angles broadly rounded; lateral carinae emarginated in lateral aspect; posterior margin forming short mesal lobe, emarginated before posterior angles; posterior angles rectangular in lateral aspect. Posterior marginal bead interrupted before posterior angles. Hypomeron triangular with round concavity posteriorly. Prosternum in front of procoxae narrow, expanded laterally; prosternal process incomplete, narrow, slightly constricted in the
middle. Scutellar shield small, triangular, covered with small, round punctures bearing short setae. Metaventrite strongly convex in the middle; surface weakly microreticulated with small, transversally confluent, rasp-like punctures; posterior margin in the produced mesally; discrinem rather indistinct. Metanepisternum trapezoidal, narrowed posteriorly, dorsal margin emarginated, ventral margin straight.

Elytra long, narrow, widest in anterior 1/3, EL/EW ratio: ♂♂ 2.02–2.26 (2.12 ± 0.08, n = 14), ♀♀ 1.88–2.07 (2.01 ± 0.05, n = 9). Surface with weak transverse microreticulation and rasp-like punctures bearing flat setae. Lateral margins regularly rounded, apices separately rounded.

Protibiae in males expanded basally, bearing fringe of long setae in basal 1/3; PTiL/PTrL ratio: ♂♂ 1.02–1.17 (1.08 ± 0.05, n = 11), ♀♀ 0.95–1.12 (1.03 ± 0.05, n = 7). Protarsomere I as long as two following tarsomeres combined; protarsomere IV simple, parallel-sided, shallowly emarginate at apex. Claws on protarsi with three, on meso and metatarsi with four denticles. Mesotibiae slightly bent inwards; mesotarsus longer than tibia, MsTiL/MsT rL ratio: ♂♂ 0.78–0.89 (0.82 ± 0.03, n = 11), ♀♀ 0.79–0.90 (0.83 ± 0.03, n = 9). Metacoxae large, anterior margin straight, posterior margin broadly rounded. Metatibiae bearing short subapical ridge and 3–4 lateral ridges parallel with apical margin of tibia, reaching 1/3 of tibial width. Metatibial spurs black, inner one ~1.30× as long as outer one. Metatarsomere I bearing 4–5 short ridges, metatarsomere II bearing 2–3 ridges, metatarsomeres III and IV without ridges. Metatarsus ~2.00× as long as metatibia, MtTiL/MtTrL ratio: ♂♂ 1.97–2.18 (2.07 ± 0.06, n = 10), ♀♀ 1.89–2.11 (1.98 ± 0.06, n = 9).

Pygidium long, slender, narrowly truncate at apex, PygL/TVtL ratio: ♂♂ 2.00–3.04 (2.36 ± 0.26, n = 13), ♀♀ 1.76–3.14 (2.22 ± 0.37, n = 9). Apical margin of terminal abdominal ventrite arcuate.

Male genitalia: sternite VIII with long setae in apical part, apical margin produced and weakly bilobed mesally (Fig. 5I), St8L/ST8W ratio: ♂♂ 1.49–1.59 (n = 2). Sternite IX long, slender, arrow-shaped, with medial longitudinal keel at apex. Parameres (Fig. 5D–G) rather short, EL/LPrL ratio: 7.87–9.17 (8.48 ± 0.40, n = 14); EL/RPrL ratio: 10.07–11.89 (11.10 ± 0.50, n = 14); LPrL/RPrL ratio: 1.24–1.37 (1.31 ± 0.04, n = 14). Left paramere with short basal part, LPrL/BLPr ratio: 1.88–2.12 (1.98 ± 0.08, n = 14); dorsal branch expanded and obliquely truncate apically; ventral branch slender, slightly bent medially, pointed at apex. Right paramere rather short, ventral branch distinctly shorter than dorsal one, dorsally bent; dorsal branch expanded and rounded apically. Median lobe long, slender, apical part slightly expanded (Fig. 5H). Phallobase with short tubular process (approximately 1/6 of total length) and long, slender furca.

Female genitalia: sternite VIII with apical protuberance and long setae alongside apical and lateral margins, spiculum ventrale short, broadly clavate (Fig. 5J); St8L/ST8W ratio: ♀ 1.36 (n = 1).

**Sexual dimorphism.** Females are usually more robust; with shorter antennae. Maxillary palpomere II is not expanded in females and without long setae on ventral side. Terminal maxillary palpomere is shorter in females, with angles more rounded. Protibiae are not expanded in females, without long setae in basal portion.
**Distribution.** Croatia, Cyprus, France, Greece, Iran, Israel, Jordan, Macedonia, Montenegro, Romania, Spain, Syria, Turkey, Turkmenistan, Ukraine. *Mordellistena hirtipes* is reported here for the first time from Croatia, Montenegro, and Spain. Csiki (1915) mentioned also “Österreich” (Austria); however, this information is probably based on a misidentification.

**Biology.** Adults were found on the flowers of *Daucus* sp. (Apiaceae) and *Helichrysum* sp. (Asteraceae) on dry grasslands and in urban environment.

**Remarks.** Franciscolo (1949) described *M. aegea* based on three specimens from Kos island (Greece). Batten (1977) mentioned that this species does not belong to the *micans* group because the antennomere IV and V are equal in length. Examination of holotype revealed that this specimen belongs to *M. hirtipes*. We consider this taxon as a junior synonym of *M. hirtipes*.

Czető (1990) described *M. podlussanyi* based on a single male specimen from Crete. He mentioned in the original description that the maxillary palpomere II is not dilated. Examination of the holotype actually revealed, that the palpomere is expanded, and any other differences which could separate this taxon from *M. hirtipes* were found. This interpretation is also supported by the results of PCA analysis (Fig. 10A). We propose *M. podlussanyi* as a junior synonym of *M. hirtipes*.

In HNHM collections, there are three specimens of *M. hirtipes*, labelled by Reitter as holotype and paratypes. However, these specimens are not mentioned in original description and labels were probably added subsequently, after the description. These specimens are not parts of the type series.

**Mordellistena (s. str.) pseudohirtipes** Ermisch, 1965
Figs 2, 6A–J

*Mordellistena pseudohirtipes* Ermisch, 1965: 268 (original description); Batten 1976: 168 (distribution); Batten 1977: 171–173 (distribution, figures, key); Plaza 1983: 574 (distribution, biology); Czető 1990: 28–29 (description, figure); Franciscolo 1995: 12 (distribution); Horák 2008: 100 (distribution); Odnosum 2010: 153, 194–195 (key, description, distribution); Ruzzier 2013: 109 (distribution).

*Mordellistena fageli* Ermisch, 1969: 112 syn. nov. (original description); Batten 1976: 168 (distribution); Plaza 1983: 575–576 (distribution, biology); Horák 2008: 97 (distribution).

*Mordellistena pseudohirtipes krotosensis* Czető, 1990: 28 syn. nov. (original description); Horák 2008: 100 (distribution).

**Type locality.** Nessebar env., Bulgaria.

**Type material examined.** *M. pseudohirtipes pseudohirtipes*: HOLOTYPE: 1 ♂, SNSD: “♂ / Genitalpräparat / Bulgariä Ümg. Nessebar Juli 1961 leg. BECH / Holotypus [red label] / MORDELLISTENA pseudohirtipes Erm. K Ermisch det. 19 / Coll. ERMISCH Leipzig Ankauf 1970 / Staatl. Museum für Tierkunde Dresden”; PARATYPE: 1 ♀,
Figure 6. Mordellistena pseudohirtipes Ermisch, 1965: A antenna, male B antenna, female C maxillary palpus, male D maxillary palpus, female E parameres, holotype F parameres, holotype of M. pseudohirtipes ssp. krotosensis G parameres, holotype of M. fageli H parameres, France I aedeagal median lobe J sternite VIII, male.
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SNSD: “♀ / Bulgaria Umg. Nessebar Juli 1961 leg. BECH / Allotypus [red label] / MORDELLISTENA pseudohirtipes Erm. K. Ermisch det. 19 / Coll. ERMISCH Leipzig Ankauf 1970 / Staatl. Museum für Tierkunde Dresden”. M. pseudohirtipes krotosensis: HOLOTYPE: 1 ♂, HNHM: “[transparent plastic board with dissected genitalia] / ♂ / Krotos KRÉTA / 1981. V. 12. leg. Podlussány / Holotypus Mordellistena pseudohirtipes Ermisch, 1965 ssp. krotosensis, Czető 1988 [red label] / Mordellistena (s. str.) pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2017”. M. fageli: HOLOTYPE: 1 ♂, SNSD: “♂ / Genitalpräparat / Portugal: Algrave Caldas de Monchique V–1960 G. Fagel / R. I. Sc. N. B. I. G. 22.145 / Holotypus [red label] / coll. ERMISCH, Leipzig, Ankauf 1970 / Staatl. Museum für Tierkunde Dresden / HOLOTYPUS Mordellistena (s. str.) fageli Ermisch, 1969 D. Selnekovič labelled 2017 [red label] / Mordellistena (s. str.) pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2017”; PARATYPE: 1 ♀, SNSD: “♀ / Portugal: Algarve Caldas de Monchique V–1960 G. Fagel / R. I. Sc. N. B. I. G. 22.145 / Allotypus [red label] / coll. ERMISCH, Leipzig, Ankauf 1970 / Staatl. Museum für Tierkunde Dresden / ALLOTYPUS (PARATYPUS) Mordellistena (s. str.) fageli Ermisch, 1969 D. Selnekovič labelled 2017 [red label] / Mordellistena (s. str.) pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2017”.

Additional material examined. Algeria. 1 ♂, SNSD: “Algérie: Algérois, Kaddous 3–V–1954 G. Fagel / R. I. Sc. N. B. I. G. 19.867 / coll. ERMISCH, Leipzig, Ankauf 1970 / Staatl. Museum für Tierkunde Dresden / “Mordellistena (s. str.) pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2017” [in collection as M. fageli]. Bulgaria. 1 ♂, SNSD: “♂ / Nessebar, Bulgaria 28. 5. – 10. 6. 1963 Karl Bleyl / MORDELLISTENA pseudohirtipes Erm. K. Ermisch det. 19”; 1 ♂, CSB: “Bulgaria mer. occ. Sandanski (→ Liljanovo) 5. – 10. 1976 Karel Majer lgt. / Mordellistena (s. str.) pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2016”. France. 2 ♂♂, SNSD: “♂ / Genitalpräparat / France Basses Alpes St. Michel l’Observat. 24. 7 – 10. 8. 63 Rudkjöb. / MORDELLISTENA pseudohirtipes Erm. K. Ermisch det. 19”; 1 ♂, SNSD: “♂ / Ardèche 10. 7. 65 Banne Balazuc / MORDELLISTENA pseudohirtipes Erm. K. Ermisch det. 19; 1 ♂ SNSD: “♂ / Genitalpräparat / Südfrankreich Camargue, 13. 6. 1952, leg. Freude / Mordellistena Lopezi Ermisch det. K. Ermisch 63” [in collection as M. lopezi]; 1 ♂, SNSD: “♂ / Genitalpräparat / Pyrenées or. Umg. Banyuls 30. 5.–10. 6. 53 / Mordellistena Lopezi Ermisch det. K. Ermisch 63” [in collection as M. lopezi]; 1 ♂, SNSD: “♀ / Gall. mer. Agay (Var) 18. 7. 58 W. Liebmam / Genitalpräparat” [in collection as M. lopezi]; 1 ♂, SNSD: “♀ / Genitalpräparat / Fr. Ardèche Bois de Paiolive 1. 7. 66 Balazuc [hand written] / Paratypus / Staatl. Museum für Tierkunde Dresden / PARATYPUS Mordellistena (s. str.) geronensis Ermisch, 1977, Selnekovič labelled 2017 [red label] / Mordellistena (s. str.) pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2017” [in collection as M. geronensis]. Georgia. 1 ♂, SNSD: “♀ / Genitalpräparat / SSSR–Gruzie Tbilisi 7.57 R. Dvořák / MORDELLISTENA pseudohirtipes Erm. K. Ermisch det. 19 / Coll. ERMISCH Leipzig Ankauf 1970 / Staatl. Museum für Tierkunde Dresden”; 2 ♂♂, SNSD: “♀ / SSSR–Gruzie Tbilisi 7.57 R. Dvořák / MORDELLISTENA pseudohirtipes Erm. K. Ermisch det. 19”. Greece. 1 ♂, SNSD: “♀ / Genitalpräparat / Athos Daphni A. Schatzmayr / MORDELLISTENA pseudohirtipes Erm. K Ermisch det. 19”; 1 ♂,
SNSD: “♂ / Genitalpräparat / Ephesus / J. Sahlb. / MORDELLISTENA pseudohirtipes Erm. K. Ermisch det. 19”; 1 ♂, HNHM: “Creta Biró / Amari 4. VII. 06 / Mordellistena pseudohirtipes Erm. det. R. Batten 1980 / Mordellistena (s. str.) pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2017”. Israel. 1 ♂, CSB: “Israel Jerusalem 25. III. 2001, ?? leg. / Mordellistena pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2014”; 1 ♂, HNHM: “Israel Rehovot 1965. V. 20. Dr. Erdős / coll. Dr. J. Erdős / Mordellistena pseudohirtipes Erm. det. R. Batten 1980 / Mordellistena (s. str.) pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2017”. Italy. 1 ♂, SNSD: “♂ / Genitalpräparat / Sicilia Magara d. V. 16. 5. 61 W. Liebmann” [in collection as M. lopezi]. Macedonia. 1 ♂, SNSD: “♂ / Veles, Mac. 23. – 25. 5. 55. leg. F. Schubert / Genitalpräparat / MORDELLISTENA pseudohirtipes Erm. K Ermisch det. 19”. Montenegro. 2 ♂♂, 1 ♀, CSB: “Montenegro S Skadarske jazero lake N, Virpazar village env. D. Selnekovič 21. VI. 2011 / Mordellistena pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2012”. Spain. 1 ♂, SNSD: “♂ / Genitalpräparat / Son Españalet 1–VI–1958 R. López / Paratypus [red label] / Mordellistena Lopezi Ermisch det. K. Ermisch 63 / Mordellistena (s. str.) pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2017”; 1 ♂, SNSD: “♂ / Genitalpräparat / SEVILLA Hi. m. Marismas, V. 1943 G. Frey, C. Koch / Mordellistena Lopezi Ermisch det. K. Ermisch 63 / Mordellistena (s. str.) pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2017”. Ukraine. 2 ♂♂, SNSD: “♂ / Genitalpräparat / Umgeb. Jalta Krim, Ende Juli 1965 leg. F. Hieke / MORDELLISTENA pseudohirtipes Erm. K Ermisch det. 19”; 1 ♂, HNHM: “Krim Jaila 17. VI. 1956 leg. L. Horváth / Mordellistena pseudohirtipes Ermisch, 1965 D. Selnekovič det. 2017”.

Differential diagnosis. From M. purpurascens it differs in shorter parameres (EL/LPrL ratio: M. pseudohirtipes: 4.65–7.17 (5.89 ± 0.71, n = 25), M. purpurascens: 4.42–5.84 (4.98 ± 0.35, n = 19); EL/RPrL ratio: M. pseudohirtipes: 5.91–8.63 (7.42 ± 0.72, n = 25), M. purpurascens: 5.57–6.94 (6.19 ± 0.41, n = 19)). Basal part of the left paramere (Fig. 6E–H) is shorter than in M. purpurascens (Fig. 7G–J). Body usually smaller (TL: M. pseudohirtipes: ♂♂ 2.47–4.05 mm (3.20 ± 0.43 mm, n = 25) ♀♀ 3.26–4.47 mm (3.68 ± 0.56 mm, n = 3), M. purpurascens: ♂♂ 3.10–4.42 mm (3.75 ± 0.35 mm, n = 19), ♀♀ 3.31–4.42 mm (3.82 ± 0.36 mm, n = 14)). Differences between M. hirtipes are mentioned above.

Redescription. Measurements: TL: ♂♂ 2.47–4.05 mm (3.20 ± 0.43 mm, n = 25) ♀♀ 3.26–4.47 mm (3.68 ± 0.56 mm, n = 3); HL: ♂♂ 0.64–0.93 mm (0.75 ± 0.09 mm, n = 25), ♀♀ 0.77–1.04 mm (0.87 ± 0.12 mm, n = 3); HW: ♂♂ 0.70–1.06 mm (0.87 ± 0.11 mm, n = 29), ♀♀ 0.77–1.04 mm (0.87 ± 0.12 mm, n = 3); PL: ♂♂ 0.83–1.29 mm (1.05 ± 0.14 mm, n = 25), ♀♀ 1.06–1.42 mm (1.19 ± 0.16 mm, n = 3); PW: ♂♂ 0.83–1.39 mm (1.07 ± 0.17 mm, n = 29), ♀♀ 1.12–1.67 mm (1.31 ± 0.25 mm, n = 3); EL: ♂♂ 1.88–3.02 mm (2.37 ± 0.31 mm, n = 25), ♀♀ 2.44–3.35 mm (2.75 ± 0.42 mm, n = 3); EW: ♂♂ 0.81–1.46 mm (1.09 ± 0.18 mm, n = 25), ♀♀ 1.17–1.69 mm (1.35 ± 0.24 mm, n = 6); PTiL: ♂♂ 0.57–0.91 mm (0.70 ± 0.10 mm, n = 25), ♀♀ 0.65–0.90 mm (0.74 ± 0.11 mm, n = 6); PTrL: ♂♂ 0.52–0.84 mm (0.63 ± 0.08 mm, n = 24), ♀♀ 0.60–0.80 mm (0.68 ± 0.09, n = 3); MsTiL: 0.71–1.17 mm (0.87 ± 0.11 mm, n = 25).
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Habitus given in Fig. 2. Body strongly elongate, slender, widest just behind humeri. Integument black, mouthparts sometimes paler. Pubescence on head pale yellowish, on pronotum yellowish to dark grey, on elytra yellowish in anterior 1/2, darkened towards apices, or completely dark grey, sometimes with reddish or purplish metallic sheen, on pygidium dark grey, on venter yellowish, darkened along posterior margins of abdominal ventrites.

Head moderately convex dorsally, wider than long, widest just before middle, HW/HL ratio: ♂♂ 1.08–1.23 (1.15 ± 0.03, n = 25), ♀♀ 1.11–1.16 (1.13 ± 0.02, n = 3). Dorsal surface weakly microreticulated with small, round punctures bearing short setae. Occipital margin rounded in dorsal aspect, straight, or slightly concave seen from behind. Eyes oval, completely reaching occiput, not expanded onto ventral surface, finely faceted, with short interfacetal setae. Anterior margin of clypeus straight. Labrum transverse, anterior margin straight or very slightly emarginate; surface microreticulation with small, round punctures and setae. Antennae slightly serrate (Fig. 6A, B); antennomeres I–IV subequal in length; antennomeres V–X longer and wider, in males ~1.30×, in females ~1.20× as long as wide; terminal antennomere elongate oval, ~1.90× as long as wide. Galea gradually expanded toward apex, covered with apically expanded sensilla. Maxillary palpi (Fig. 6C–D) black; palpomere I very short; palpomere II distinctly expanded, with long setae on ventral side in males, not expanded, rather long and narrow in females; palpomere III short, ~1.50× as long as wide; terminal palpomere in males broadly securiform, with inner angle situated around the middle, in females slenderer, with inner angle situated in apical 1/3; TPalL/TPalW ratio: ♂♂ 1.80–2.30 (2.07 ± 0.11, n = 25), ♀♀ 2.05–2.31 (2.21 ± 0.12, n = 3).

Pronotum moderately convex, approximately as long as wide, PW/PL ratio: ♂♂ 0.97–1.12 (1.02 ± 0.03, n = 25), ♀♀ 1.04–1.18 (1.10 ± 0.06, n = 3). Surface finely transversally microreticulate, covered with rasp-like punctures, distance between punctures 2.00–4.00 times as long as the diameter, each puncture bears flat, pointed seta. Anterior margin rounded, slightly produced mesally, anterior angles broadly rounded; lateral carinae emarginated in lateral aspect; posterior margin forming short mesal lobe, emarginated before posterior angles; posterior angles in lateral aspect rectangular, acute. Posterior marginal bead interrupted before posterior angles. Scutellar shield small, triangular, with small, rasp-like punctures bearing setae. Metanepisternum trapezoidal, narrowed posteriorly, ventral margin straight, dorsal margin emarginate.

Elytra long and narrow, moderately convex, widest at the end of anterior 1/4, EL/EW ratio: ♂♂ 2.02–2.40 (2.18 ± 0.10, n = 25), ♀♀ 1.99–2.08 (2.04 ± 0.04, n = 3). Surface with weak transverse microreticulation and rasp-like punctures, these are larger and more densely arranged than those on pronotum, each puncture...
bears flat seta. Lateral margins rather strongly convergent, regularly rounded; apices separately rounded.

Profemora slender, in males somewhat stouter than in females. Protibiae straight, in males distinctly expanded in basal half, here with fringe of long, thick setae; PTiL/PTrL ratio: ♂♂ 1.02–1.24 (1.11 ± 0.05, n = 25), ♀♀ 1.02–1.13 (1.09 ± 0.05, n = 3). Protarsomere I in females as long as two following tarsomeres combined, in males slightly longer; protarsomere IV simple, slightly shorter than previous one, shallowly emarginate at apex; terminal protarsomere slightly shorter than previous two tarsomeres combined. Claws on protarsi with three denticles, on meso- and metatarsi with four denticles. Metotibiae slightly bent medially; metotarsus longer than tibia, MsTiL/MsTrL ratio: ♂♂ 0.79–0.90 (0.83 ± 0.03, n = 13), ♀♀ 0.85–0.92 (n = 2). Metacoxae large, anterior margin slightly emarginated, posterior margin broadly rounded. Metatibia bearing short subapical ridge and 3–4 lateral ridges parallel to apical margin of tibia, reaching 1/3 of tibial width. Metatibial spurs black, long, inner one ~1.30× as long as outer one. Metatarsomere I bearing 3–5 short ridges, metatarsomere II bearing 2–3 ridges, metatarsomere III without ridges. Metatarsus ~2.00× as long as metatibia, MtTrL/MtTiL ratio: ♂♂ 2.02–2.32 (2.18 ± 0.07, n = 17), ♀♀ 1.98–2.00 (n = 2).

Pygidium long and slender, PygL/TVtL ratio: ♂♂ 1.82–2.47 (2.19 ± 0.15, n = 23), ♀♀ 1.88–2.79 (2.22 ± 0.32, n = 6). Apical margin of terminal ventrite arcuate.

Male genitalia: sternite VIII rather short, setae present in apical 1/3, apical protuberance short, slightly bilobed at apex (Fig. 6J); St8L/St8W ratio: ♂♂ 1.50–1.53 (n = 2). Sternite IX long, slender, arrow-shaped, with medial longitudinal keel in apical part. Parameres (Fig. 6E–H) rather long, EL/LPrL ratio: 4.65–7.17 (5.89 ± 0.71, n = 25), EL/RPrL ratio: 5.91–8.63 (7.42 ± 0.72, n = 25), LPrL/RPrL ratio: 1.12–1.39 (1.26 ± 0.06, n = 25). Left paramere: dorsal branch expanded apically, obliquely truncate at apex; ventral branch narrow, slightly bent medially, pointed at apex, LPrL/BLPr ratio: 1.73–2.24 (1.95 ± 0.11, n = 25). Right paramere: dorsal branch rather narrow, slightly expanded, rounded at apex; ventral branch as long as or slightly shorter than dorsal branch, bent dorsally in apical part, pointed at apex. Median lobe long, slender, slightly expanded in apical part (Fig. 6I). Phallobase with short tubular process (approximately 1/6 of total length) and long, slender furca.

**Sexual dimorphism.** Females are more robust than males, their protibiae are not expanded in basal 1/3 and without fringe of long setae. Maxillary palpomere II is not expanded in females and without long setae on ventral side. Terminal maxillary palpomere is slenderer in females and its inner angle is situated more distally than in males. Antennae are somewhat shorter in females.

**Distribution.** Algeria, Azerbaijan, Bulgaria, Croatia, Cyprus, France, Georgia, Greece, Israel, Italy, Macedonia, Montenegro, Morocco, Portugal, Spain, Turkey, Ukraine. *Mordellistena pseudohirtipes* is reported here for the first time from Israel and Montenegro.

**Biology.** Adults were collected on the flowers of Apiaceae plants on dry grasslands. Plaza (1983) mentioned that *M. pseudohirtipes* was collected in Spain on following plant species: *Thapsia villosa, Daucus carota* (both Apiaceae) and *Ruta montana* (Rutaceae).
Remarks. Mordellistena fageli was placed in the pentas–group in the original description, based on the dark pubescence and three ridges on the metatarsomere II. In fact, as Horák (1983) already mentioned, this species belongs to M. hirtipes complex, based on the strongly convex body and expanded maxillary palpomere II. Examination of type material did not reveal any characters which could separate this taxon from M. pseudohirtipes. In the plot from PCA analysis (Fig. 10A), M. fageli is placed just next to the cluster of M. pseudohirtipes, in the same plane along the PC 1 axis. We consider these taxa as conspecific and propose M. fageli as a junior synonym of M. pseudohirtipes.

Czető (1990) described M. pseudohirtipes krotosensis based on two male specimens from Crete island. Characters such as length of the body, length of pygidium and colouration of pubescence, that he used for differentiation of the subspecies, are subjects of individual variability. Results of PCA (Fig. 10A) show, that holotype of M. pseudohirtipes krotosensis is placed within the cluster of the nominotypical subspecies. After examination of holotype we consider this subspecies as a junior synonym of M. pseudohirtipes pseudohirtipes.

In Ermisch’s collection, there is a series of specimens named Mordellistena lopezi. Such species has not been described, and in fact, all the specimens belong to M. pseudohirtipes, except the one labelled as “Type”, which belongs to M. purpurascens.

Mordellistena (s. str.) purpurascens Costa, 1854
Figs 3, 7A–K

Mordellistena purpurascens Costa, 1854: 17 + Plate XXI (original description, figure); Gemminger and Harold 1870: 2113 (catalogue, as syn. of Mordellistena pumila (Gyllenhal, 1810)); Emery 1876: 95 (as syn. of Mordellistena micans (Germar, 1817)); Baudi di Selve 1877: 827 (as syn. of M. micans); Heyden et al. 1883: 142 (catalogue, as syn. of M. micans); Schilsky 1898: 77 (as syn. of M. micans); Heyden et al. 1906: 456 (catalogue, as syn. of M. micans); Csiki 1915: 37 (catalogue, as syn. of M. micans); Ermisch 1977: 169 (misidentification); Batten 1977 (misidentification); Kaszab 1979: 72, 74 (misidentification); Franciscozolo 1991: 172–173 (misidentification); Odnosum 2003: 36–46 (misidentification); Odnosum 2005: 199 (misidentification); Ruzzier 2013: 110 (misidentification).

Mordellistena geronensis Ermisch, 1977: 169 syn. nov. (original description in the key); Kaszab 1979: 72–73 (figures, key); Franciscozolo 1991: 171–172 (key, figures); Horák 2008: 98 (distribution).

Mordellistena istrica Ermisch, 1977: 169 syn. nov. (original description in the key); Kaszab 1979: 73 (key); Horák 2008: 98 (distribution).

Type locality. Naples, Italy.

Type material examined. M. purpurascens: LECTOTYPE, here designated, glued, genitalia in separate microvial, right metatarsus missing: 1 ♂, MZFN: “Mordellistena
Figure 7. *Mordellistena purpurascens* Costa, 1845: **A** antenna, male **B** antenna, female **C** maxillary palpus, male **D** maxillary palpus, female **E** sternite VIII, male **F** sternite VIII, female **G** parameres, lectotype **H** parameres, holotype of *M. geronensis* **I** parameres, holotype of *M. istrica* **J** parameres, France **K** aedeagal median lobe, holotype.
Taxonomic revision of Mordellistena hirtipes species complex...

purpurascens n. Napoli [hand written by Costa] / LECTOTYPE Mordellistena purpurascens Costa, 1854 D. Selneković des. 2019” [red label]. M. geronensis: HOLOTYPE: 1 ♂, SNSD: “♂/Genitalpräparat/[card with dissected genitalia, right antenna and right protarsus]/Spanien, Prov. Gerona, Tossa de mar, A. Kampf. VII–VIII 35 / Holotypus [red label]/Holotypus Mordellistena geronensis Ermisch/Staat. Museum für Tierkunde Dresden”; PARATYPES: 1 ♀, SNSD: “Spanien, Prov. Gerona, Tossa de mar, A. Kampf, V–VI 35 / sp.? grupe micans, det. Ermisch 1940 / Paratypus / PARATYPUS Mordellistena (s. str.) geronensis Ermisch, 1977, Selneković labelled 2017 [red label] / Mordellistena (s. str.) purpurascens Costa, 1854, D. Selneković det. 2019”; 2 ♂♂, 2 ♀♀♀, SNSD: “♂[or]♀/Ardèche, 10. 7. 65, Banne, J. Balazuc / Paratypus / PARATYPUS Mordellistena (s. str.) geronensis Ermisch, 1977, Selneković labelled 2017 [red label] / Mordellistena (s. str.) purpurascens Costa, 1854, D. Selneković det. 2017”; 1 ♂, SNSD: “♂/Ardèche, Sammzon, 8. 7. 65, Balazuc / Paratypus / PARATYPUS Mordellistena (s. str.) geronensis Ermisch, 1977, Selneković labelled 2017 [red label] / Mordellistena (s. str.) purpurascens Costa, 1854, D. Selneković det. 2017”. M. istrica: HOLOTYPE: 1 ♂, SNSD: “♂/[card with dissected genitalia]/Pola, Istr. F. Lang/[blank red circular label]/MORDELLISTENA istrian [illegible handwriting] det. Ermisch 1952 / Type [red label] / Holotypus Mordellistena istrica Ermisch / Mordellistena (Mordellistena) geronensis Ermisch det. P. Leblanc 2007 / Mordellistena (s. str.) purpurascens Costa, 1854 D. Selneković det. 2019”; PARATYPES: 1 ♀, SNSD: “Plomin, Warmehang 14. 6. 1965 / Istrien K. Wellschmeld / Paratypus / PARATYPUS Mordellistena (s. str.) istrica Ermisch, 1977, Selneković labelled 2017” [red label]; 1 ♂, SNSD: “♂/Corsica / Paratypus / PARATYPUS Mordellistena (s. str.) istrica Ermisch, 1977, Selneković labelled 2017 [red label] / Mordellistena (Mordellistena) geronensis Ermisch det. P. Leblanc 2007 / Mordellistena (s. str.) purpurascens Costa, 1854 D. Selneković det. 2019”; 1 ♂, 1 ♀, SNSD: “12. 7. 14 Gallia m. Agay Rapp / Paratypus / PARATYPUS Mordellistena (s. str.) istrica Ermisch, 1977, Selneković labelled 2017” [red label].

Additional material examined. Greece: 1 ♂, 2 ♀♀♀, CSB: “Greece N, Corfu – Kavos, 39°24′16″N, 20°05′53″E, F. Repta leg., 28. VIII. 2011 / Mordellistena purpurascens Costa, 1854, D. Selneković det. 2019”. Italy: 1 ♂, CSB: “IT–Sicilia, Madonia, Termini, Sciarà, M San Calogero, ex. l, 2–3. 6. 2011, M. Šárovec, 3. 8 / Mordellistena purpurascens Costa, 1854, D. Selneković det. 2019”; 1 ♂, SNSD: “♂/Gavoi Sard. 750m 21.–26. 8. 55 J. Kless 78 / Mordellistena Lopezi Ermisch det. K. Ermisch / Mordellistena (s. str.) purpurascens Costa, 1854 D. Selneković det. 2019” [in collection as M. lopezi]; 1 ♂, SNSD: “♂/Genitalpräparat / ITALIA mer. Capaccio Hüdepohl VI. 64 / Mordellistena (s. str.) purpurascens Costa, 1854 D. Selneković det. 2019” [in collection as M. lopezi]. Montenegro: 4 ♂♂, 4 ♀♀♀ CSB: “Montenegro SE, 42°06′N, 19°06′E, Bar–centrum, on Daucus sp., D. Selneković 19. VI. 2011 / Mordellistena (s. str.) purpurascens Costa, 1854, D. Selneković det. 2019”; 1 ♀ CSB: “Montenegro SE, BAR env., 42°07′56″N, 19°07′33″E, 22. VI. 2011 / Mordellistena (s. str.) purpurascens Costa, 1854, D. Selneković det. 2019”; 1 ♂ HNHM: “Dalmatia Horváth / Zelenika 906. VIII. / Mordellistena pseudohirtipes Erm. det. R. Batten
1979 / Mordellistena (s. str.) purpurascens, Costa, 1854 D. Selnekovič det. 2019”.

**Morocco:** 1 ♂, CSB: Morocco Moyen Atl, Khenifra 15km E M. Šárovec 11. VII. 2007 / Mordellistena purpurascens Costa, 1854 D. Selnekovič det. 2019”; 4 ♂♂, 3 ♀♀, CSB: “Morocco Moyen Atl, Khenifra 10km I M. Šárovec 30. V. 2007 / Mordellistena purpurascens Costa, 1854 D. Selnekovič det. 2019”.

**Spain:** 1 ♂, SNSD: “♂ / Genitalpräparat / Son Españolet 1–VI–1958 R. López / Typus [red label] / Mordellistena Lopezi Ermisch det. K. Ermisch / Mordellistena (s. str.) purpurascens Costa, 1854 D. Selnekovič det. 2019” [in collection as *M. lopezi*].

**Differential diagnosis.** *M. purpurascens* closely resembles *M. hirtipes* and *M. pseudohirtipes*. The differences are described under these species.

**Redescription.**

Measurements: TL: ♂♂ 3.10–4.42 mm (3.75 ± 0.35 mm, n = 19), ♀♀ 3.31–4.42 mm (3.82 ± 0.36 mm, n = 14); HL: ♂♂ 0.77–0.97 mm (0.85 ± 0.06 mm, n = 19), ♀♀ 0.78–0.96 mm (0.86 ± 0.06 mm, n = 14); HW: ♂♂ 0.91–1.17 mm (1.01 ± 0.07 mm, n = 19), ♀♀ 0.84–1.12 mm (0.98 ± 0.09 mm, n = 14); PL: ♂♂ 1.06–1.44 mm (1.23 ± 0.10 mm, n = 19), ♀♀ 1.04–1.44 mm (1.25 ± 0.11 mm, n = 14); PW: 1.10–1.56 mm (1.30 ± 0.13 mm, n = 19), ♀♀ 1.13–1.58 mm (1.35 ± 0.14 mm, n = 13); EL: ♂♂ 2.44–3.35 mm (2.81 ± 0.27 mm, n = 19), ♀♀ 2.50–3.38 mm (2.88 ± 0.27 mm, n = 14); EW: ♂♂ 1.15–1.59 mm (1.35 ± 0.13 mm, n = 19), ♀♀ 1.19–1.66 mm (1.43 ± 0.15 mm, n = 14); ATiL: ♂♂ 0.71–0.93 mm (0.81 ± 0.07 mm, n = 19), ♀♀ 0.65–0.91 mm (0.75 ± 0.08 mm, n = 14); ATrL: ♂♂ 0.64–0.84 mm (0.72 ± 0.07 mm, n = 15), ♀♀ 0.62–0.80 mm (0.69 ± 0.06 mm, n = 13); ITiL: ♂♂ 0.91–1.23 mm (1.03 ± 0.10 mm, n = 19), ♀♀ 0.86–1.23 mm (1.01 ± 0.12 mm, n = 14); ITiL: ♂♂ 1.12–1.64 mm (1.27 ± 0.15 mm, n = 9), ♀♀ 1.02–1.34 mm (1.16 ± 0.10 mm, n = 14); PTrL: 0.78–1.08 mm (0.88 ± 0.08 mm, n = 19), ♀♀ 0.75–1.05 mm (0.88 ± 0.08 mm, n = 14); PTrL: 1.64–2.18 mm (1.87 ± 0.19 mm, n = 9), ♀♀ 1.48–2.16 mm (1.77 ± 0.21 mm, n = 11); PygL: ♂♂ 1.50–2.12 mm (1.86 ± 0.18 mm, n = 19), ♀♀ 1.35–1.98 mm (1.67 ± 0.20 mm, n = 14); TVrL: ♂♂ 0.58–0.87 mm (0.77 ± 0.10 mm, n = 18), ♀♀ 0.60–0.92 mm (0.77 ± 0.08 mm, n = 14); RPrL: 0.52–0.64 mm (0.56 ± 0.03 mm, n = 19); LPrL: 0.41–0.51 mm (0.45 ± 0.03 mm, n = 19); St8L: ♂♂ 0.65–0.80 mm (0.70 ± 0.07 mm, n = 3); St8W: ♂♂ 0.49–0.52 mm (0.50 ± 0.01 mm, n = 3).

Habitus illustrated in Fig. 3. Body strongly elongate, slender, widest behind anterior 1/4 of elytra. Integument black, anterior margin of clypeus and mandibles somewhat paler. Pubescence on head and thorax yellowish; on elytra yellowish in anterior half, gradually darkened apically; on venter yellowish, darkened along posterior margins of ventrites 3 and 4 and completely dark grey on terminal ventrite and pygidium.

Head convex dorsally, wider than long, widest about middle, HW/HL ratio: ♂♂ 1.11–1.23 (1.19 ± 0.03, n = 19), ♀♀ 1.06–1.19 (1.14 ± 0.03, n = 14). Dorsal surface weakly microreticulated, with small, round punctures, each bearing short seta. Ventral surface with weak transverse microreticulation and sparsely arranged, round punctures, each bearing short seta. Occipital margin rounded in dorsal aspect, straight if seen from behind. Eyes oval, completely reaching occiput, not expanded onto ventral surface, finely faceted, with short interfacial setae. Anterior margin of
clypeus straight. Labrum transverse, LabW/LabL: ♂ 2.04–2.27 (2.15 ± 0.10, n = 5), ♀ 1.88–2.38 (2.21 ± 0.20, n = 5), anterior margin straight or very shallowly emarginate mesally; surface covered with small, round punctures, each bearing seta. Antennae slightly serrate, expanded from antennomere V (Fig. 7A, B); antennomeres I and II short, subequal in length and width; antennomere III equal in length and slightly slenderer than previous two; antennomeres IV slightly longer and wider than previous one; antennomeres V–X wider than previous four, in males ~1.60×, in females ~1.30× as long as wide; antennomere XI elongate oval, ~2.20× as long as wide. Galea gradually expanded apically, covered with apically expanded sensilla. Maxillary palpi (Fig. 7C–D) black; palpomere I very short; palpomere II in males expanded with long setae on ventral side, in females slenderer, without long setae; palpomere III short, ~1.80× as long as wide, in males with long setae on ventral side; terminal palpomere securniform, in males wider than in females, inner angle situated around middle in males, in terminal 1/3 in females; TPalL/TPalW ratio: ♂ 1.72–2.16 (1.96 ± 0.10, n = 17), ♀ 2.08–2.34 (2.21 ± 0.09, n = 14).

Pronotum moderately convex, slightly wider than long, PW/PL ratio: ♂ 0.97–1.10 (1.05 ± 0.03, n = 19), ♀ 1.00–1.15 (1.07 ± 0.04, n = 13). Surface weakly microreticulated with small, rasp-like punctures, distance between punctures 1.50–2.00× as long as puncture diameter, each puncture bearing flat seta. Anterior margin rounded, slightly produced mesally, anterior angles broadly rounded; lateral carinae rounded in dorsal aspect, shallowly but distinctly emarginate in lateral aspect; posterior margin forming short mesal lobe, emarginated laterally before posterior angles; posterior angles rectangular, pointed in lateral aspect. Posterior marginal bead interrupted before posterior angles. Prosternum in front of procoxae narrow, laterally expanded. Scutellar shield small, triangular, with small punctures bearing short setae. Mesoventral process ca. half as wide as mesofemora. Metaventrite large, posterior margin produced mesally between metacoxae; longitudinal discrime rather indistinct. Metaneepisternum trapezoidal, slightly narrowed posteriorly, dorsal margin emarginate, ventral margin straight.

Elytra long, narrow, widest at end of anterior 1/4, EL/EW ratio: ♂ 1.97–2.23 (2.08 ± 0.07, n = 19), ♀ 1.83–2.15 (2.02 ± 0.07, n = 14). Dorsal surface covered with weak transverse microreticulation and rasp-like punctures, distance between punctures ~1.50× as long as puncture diameter; each puncture bearing flat seta. Lateral margins regularly rounded, apices separately rounded. Protibiae straight, basal part in males slightly expanded and bearing distinct fringe of long setae; PTiL/PTiL ratio: ♂ 0.98–1.24 (1.13 ± 0.06, n = 15), ♀ 0.98–1.17 (1.10 ± 0.05, n = 13). Protarsomere I in females as long as two following tarsomeres combined, in males slightly longer; protarsomere IV simple, parallel-sided, very shallowly emarginated at apex. Claws on protarsi rather long, slender, with three denticles, on meso- and metatarsi with four denticles. Mesotibiae slightly bent medially; mesotarsus longer than tibia, MsTiL/MsT rL ratio: ♂ 0.75–0.90 (0.84 ± 0.04, n = 9), ♀ 0.82–0.95 (0.88 ± 0.04, n = 14). Metacoxae large, anterior margin straight, posterior margin broadly rounded. Metatibiae bearing short subapical ridge and 3–4 lateral ridges parallel to apical tibial margin, reaching 1/3 of tibial width. Metatibial spurs black, inner one
~1.30× as long as outer one. Metatarsomere I bearing in males 5, in females 3–4 short lateral ridges; metatarsomere II bearing 2–3 ridges; metatarsomeres III and IV without ridges. Metatarsus ~2.00× as long as metatibia, MtTrL/MtTiL ratio: ♂♂ 2.00–2.33 (2.13 ± 0.10, n = 10), ♀♀ 1.90–2.18 (2.01 ± 0.07, n = 11).

Pygidium long, slender, PygL/TVtL ratio: ♂♂ 1.88–3.32 (2.43 ± 0.35, n = 18), ♀♀ 1.88–2.64 (2.18 ± 0.17, n = 14). Apical margin of terminal abdominal ventrite arcuate.

Male genitalia: sternite VIII rather short, with long setae in apical part, apical protuberance rather short, slightly bilobed at apex (Fig. 7E); St8L/St8W ratio: ♂♂ 1.26–1.32 (1.28 ± 0.02, n = 4). Sternite IX long, slender, arrow-shaped, with medial longitudinal keel apically. Parameres (Fig. 7G–J) rather long, EL/LPrL ratio: 4.42–5.84 (4.98 ± 0.35, n = 19), EL/RPrL ratio: 5.57–6.94 (6.19 ± 0.41, n = 19); LPrL/RPrL ratio: 1.16–1.30 (1.24 ± 0.03, n = 19). Left paramere with very long basal part, LPrL/BLPr ratio: 1.50–1.91 (1.76 ± 0.09, n = 19); dorsal branch strongly expanded apically, obliquely truncate at apex; ventral branch slender, slightly bent medially. Right paramere rather long with long branches; ventral branch longer than dorsal one, pointed at apex; dorsal branch rather narrow, slightly expanded apically, rounded at apex. Median lobe (Fig. 7K) long, slender, apical part narrow or slightly expanded. Phallobase with short tubular process (approximately 1/6 of total length) and long, slender furca.

Female genitalia: sternite VIII (Fig. 7F) with slightly bilobed apical protuberance, long setae situated at apex and alongside lateral margins; spiculum ventrale short, broadly clavate; St8L/St8W ratio: 1.62 (n = 1).

**Sexual dimorphism.** Females are more robust, with protibiae not expanded and without fringe of long setae in basal part. Maxillary palptomere II not expanded in females and without long setae on ventral side. Terminal maxillary palptomere is wider in males, with its inner angle situated approximately in the middle (Fig. 7C), in females it is generally slenderer, with its inner angle situated in terminal 1/3 (Fig. 7D). Antennae are shorter in females; antennomeres V–X ~1.60× as long as wide in males, ~1.30× in females.

**Distribution.** Croatia, France, Greece, Italy, Montenegro, Morocco, Spain. *Mordellistena purpurascens* is reported here for the first time from Greece and Montenegro. Odnosum (2003, 2005, 2010) reported *M. purpurascens* from Kazakhstan, Turkmenistan, Tajikistan, and Ukraine. However, based on the figures of parameres provided in all the three mentioned studies, it is obvious that he referred to a different species (see Discussion).

**Biology.** Adults were collected by the first author in Montenegro, in urban environment of Bar on the flowers of *Daucus* sp. (Apiaceae).

**Remarks.** *Mordellistena purpurascens* was described by Costa (1854) and referred to be found in several localities in former “Regno di Napoli” (southern parts of present Italy). Series of *M. purpurascens* in Costa’s collection in MZFN contains only two specimens. One of them with the original label “Mordellistena purpurascens n. Napoli” is designed here as a lectotype. The other specimen labelled “S. Severina” without identification label belongs to a different species from the *gemellata*-group (sensu Ermisch 1956). Genitalia of the lectotype were examined for the first time for the purposes of the present study (Fig. 7G).
Ermisch (1977) briefly described two new species *M. geronensis* and *M. istrica* as a part of an identification key. He differentiated these species from each other based on the shape of the apical part of the median lobe (expanded in *M. istrica*, not expanded in *M. geronensis*). Shape of the apical part of median lobe depends on the observation method (dry/wet, card mounted/slide mounted). After examining the series of slide mounted median lobes of both taxa, we did not find any differences in the shape. Examination of the male genitalia from type specimens of *M. purpurascens* (Fig. 7G), *M. geronensis* (Fig. 7H) and *M. istrica* (Fig. 7I) revealed that these taxa are conspecific. We thus propose *M. istrica* and *M. geronensis* as the junior synonyms of *M. purpurascens*.

Type series of *M. istrica* includes a female paratype (Pola, Croatia), which we were not able to assign to *M. purpurascens* or *M. pseudohirtipes*. Type series of *M. geronensis* includes a male paratype (Bois de Paiolive, Ardèche, France), which in fact belongs to *M. pseudohirtipes*, and three paratypes (Tossa de mar, Spain; Costa Brava, Spain), which belong to *M. hirtipes*. In Ermisch’s collection, there is a series of specimens named *Mordellistena lopezi*. Such species has not been described. Specimen labelled as “Typus”, in fact, belongs to *M. purpurascens*, the rest of the specimens belong to *M. pseudohirtipes*.

*Mordellistena (s. str.) balearica* Compte, 1985

*Fig. 8*

*Mordellistena balearica* Compte, 1985: 63–64 (original description); Horák 2008: 96 (distribution).

**Type locality.** Palma de Mallorca, Majorca.

**Type depository.** According to the original description (Compte 1985), holotype should be deposited in MNCN. However, despite of the effort of the curator, the specimen was not found.

**Diagnosis.** *Mordellistena balearica* was described based on a single male specimen from Mallorca. According to the original description, this species closely resembles *M. pseudohirtipes* and can be distinguished from this species by longer antennae (antennomeres V–X two times longer than wide) and different shape of parameres (Fig. 8) (Compte 1985). All characters mentioned in the original description suggest that this taxon is conspecific with *M. pseudohirtipes*; unfortunately, the authors did not have the opportunity to study the type.

**Distribution.** Known only from type locality.

**Remarks.** Compte (1985) mentioned following information: “This specimen, together with other specimens collected by P. López in Majorca, which current location I don’t know, was studied by the specialist Mr Ermisch, who considered it as a new species for science, called *M. balearica*, a name that seems to have remained *in litteris*. There are several specimens collected by P López in Mallorca in Ermisch’s collection (SNSD) labelled by Ermisch as *M. balearica* which in fact all belong to *M. thuringiaca* Ermisch, 1963.
**Mordellistena (s. str.) irritans** Franciscolo, 1991

Fig. 9

*Mordellistena irritans* Franciscolo, 1991: 168–173 (original description); Franciscolo 1995: 12 (distribution); Horák 2008: 98 (distribution).

**Type locality.** Lampedusa Is., Italy.

**Type depository.** Museo d’Aumale, Terrasini, Palermo, Italy: 1 ♀ holotype, 1 ♂ paratype (Franciscolo 1991). Not examined.

**Diagnosis.** *Mordellistena irritans* can be assigned to *M. hirtipes* complex based on the expanded maxillary palpomere II in males and the shape of parameres. This species can be distinguished from all other species in the complex by the characteristic shape of the left paramere with dorsal branch parallel-sided and rounded at apex (Fig. 9).

**Distribution.** Known only from the type locality.

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**Key to the species of *M. hirtipes* species complex**

1. Dorsal branch of the left paramere not expanded, parallel-sided, rounded at apex (Fig. 9)................................................................................................. *M. irritans*

   – Dorsal branch of left paramere expanded apically, obliquely truncate at apex...

2. Parameres shorter, EL/LPrL ratio: 7.87–9.17 (8.48 ± 0.40, n = 14); EL/RPrL ratio: 10.07–11.89 (11.10 ± 0.50, n = 14); basal part of left paramere short; ventral branch of the right paramere usually distinctly shorter than the dorsal one (Fig. 5D–G). Terminal segment of maxillary palpi in females shorter and broader, inner angle is more acute (Fig. 4B). Pubescence on pronotum and elytra yellowish, somewhat darkened posteriorly.................. *M. hirtipes*

   – Parameres longer, EL/LPrL ratio: 4.42–7.17, EL/RPrL ratio: 5.57–8.63; basal part of left paramere longer; ventral branch of right paramere equally long or longer than the right one (Figs 6E–H, 7G–J). Terminal segment of maxillary palpi in females slenderer, its inner angle is rounded (Figs 6D, 7D). Pubescence on pronotum and elytra sometimes completely dark greyish...............................3

3. Parameres shorter, EL/LPrL ratio: 4.65–7.17 (5.89 ± 0.71, n = 25), EL/RPrL ratio: 5.91–8.63 (7.42 ± 0.72, n = 25); basal part of left paramere shorter (Fig. 6E–H). Pubescence on pronotum and elytra sometimes almost completely dark greyish ................................. *M. pseudohirtipes*

   – Parameres longer, EL/LPrL ratio: 4.42–5.84 (4.98 ± 0.35, n = 19), EL/RPrL ratio: 5.57–6.94 (6.19 ± 0.41 n = 19); basal part of left paramere longer (Fig. 7G–J). Pubescence on pronotum and elytra yellowish, darkened posteriorly but not completely dark greyish .......................... *M. purpurascens*
Results of PCA analysis

Principal Component Analysis (PCA) was conducted based on following morphometric characters: HL, HW, PL, PW, EL, EW, PTiL, MsTiL, MtTiL, RPrL, BRPr, and LPrL. Characters were measured in 59 male specimens, including holotypes / lectotype of every taxon. The first two principal components describe 91.03% (PC 1) and 6.32% (PC 2) of variation. PC 1 correlates mostly with elytral length (loading 0.66), elytral width (loading 0.37) and pronotal width (loading 0.35); PC 2 correlates with characters measured on parameres: left paramere length (loading 0.68), right paramere length (loading 0.56), basal part of left paramere length (loading 0.43).

Visualisation of the results of PCA analysis (Fig. 10A) shows clear distinction of *M. hirtipes*, *M. pseudohirtipes*, and *M. purpurascens* along the PC 2 axis. Cluster of *M. istrica* overlaps with cluster of *M. geronensis*; *M. pseudohirtipes krotosensis* is placed within the cluster of *M. pseudohirtipes pseudohirtipes*; *M. fageli* is placed next to the cluster of *M. pseudohirtipes* in one plane along the PC 1 axis, and *M. podlussanyi* and *M. aegea* are placed next to cluster of *M. hirtipes*. Results of PCA correspond with hypotheses based on observations of morphological characters.

Length of elytra and length of parameres are characters, that reach the highest loadings in PCA analyses. Ratios of these characters (EL/RPrL, EL/LPrL) are useful for identification and are used in diagnoses. Differences in values of selected ratios are presented in Fig. 10B.
Figure 10. A Results of Principal Component Analysis (PCA) B Box-plots showing differences between species in selected ratios. Top and bottom of the boxes represent first and third quartiles, transverse band represents the median and whiskers represent maximum and minimum.

Discussion

The family Mordellidae is taxonomically very challenging and thus rather poorly known. Most of the original descriptions are insufficient for proper identification and differentiation of the species, especially those published before the 1950s (before K Ermisch provided a more precise method of description). There are still some species which were described as several different taxa, sometimes even by the same author (e.g., *M. pseudohirtipes* Ermisch, 1965 = *M. fageli* Ermisch, 1969). Characters used for the differentiation of these taxa were usually misinterpreted (e.g., the shape of the
median lobe in *M. geronensis* Ermisch, 1977 and *M. istrica* Ermisch, 1977) or they are subjects of the intraspecific variability (e.g., the dark coloration of the pubescence in *M. fageli* Ermisch, 1969). In other cases, the insufficient descriptions in combination with overlooking of the type specimens led to a misinterpretation of the taxa. It can be seen for example in some species described by Achille Costa (1854). Revision of the type specimens in his collection deposited in MZFN revealed that several species described by him were incorrectly interpreted by the subsequent authors as completely different species, one of them is *M. purpurascens* Costa, 1854 treated in the present paper. This species was considered by the subsequent authors as a synonym of either *M. pumila* (Gyllenhal, 1810) (Gemminger and Harold 1870) or *M. micans* (Germar, 1817) (Emery 1876; Baudi di Selve 1877; Heyden et al. 1883, 1906; Schilsky 1898; Csiki 1915). Later it was treated again as a valid species by Ermisch (1977), Ermisch in Kaszab (1979) and Batten (1977) but none of these authors had studied the types and it is obvious, based on their figures of the genitalia that the specimens considered by them as *M. purpurascens* belong to a different species. Their misinterpretations were later followed by Odnosum (2003, 2005, 2010) who published several new distribution records for *M. purpurascens* which were then included in the catalogue by Horák (2008). Only the examination of the lectotype of *M. purpurascens* Costa, 1854 done by the first author revealed that it is conspecific with the types of *M. geronensis* Ermisch, 1977 and *M. istrica* Ermisch, 1977. As it can be seen from this example, examination and redescriptions of the type specimens are essential for the future studies, especially in such taxonomically difficult and species-rich family as Mordellidae.

We live in the era of the global biodiversity crisis caused by the anthropogenic interventions in the natural ecosystems. But how does these changes affect the diversity and distribution patterns of Mordellidae beetles is not known. Despite of the great effort of the authors such as Ermisch (e.g., 1956, 1965, 1969a), Horák (e.g., 1990, 2008) and Odnosum (e.g., 2003, 2005, 2010) who have summarised and published a vast number of distributional records, the information about distribution and ecology of many Palearctic species is still very poor and several species are reported only from a single locality stated in the original description. It is very important to gather and provide new distributional and ecological records, however, it is also essential to pay effort to correct identification of the specimens to guarantee the accuracy of the published biological data.

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