Evaluation of Chitin as a Biomarker of Pathogenic Fungal Isolates
(Penilaian Kitin sebagai Biopenanda Pencilan Kulat Patogen)

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ABSTRACT
Chitin is a polysaccharide component of the inner cell wall of fungi that has been used to estimate fungal invasion in plant products. However, its detection in major pathogenic fungal species has not been investigated. The present study aimed to determine the chitin contents of pathogenic fungal species in order to evaluate its diagnostic potential as a biomarker for fungal infections. High performance liquid chromatography (HPLC) was used to measure chitin content. Pure chitin was acid hydrolyzed and the fluorescence of 9-fluorenylmethylchloroformate (FMOC-CI) derivatives of glucosamine produced were measured. The chitin contents of ten pathogenic fungal isolates were determined per mycelial dry weight. They varied from 18.61 (± 0.09) to 47.12 (± 0.50) µg/mg dry mycelial weight. Candida albicans and Cryptococcus neoformans exhibited the highest and lowest levels of chitin, respectively. Based upon relative amounts of chitin produced, three groups namely: high (Candida albicans, Cryptococcus gattii, Aspergillus niger and Penicillium at 47.12, 46.98, 46.05, and 44.15 µg/mg respectively), medium (Rhizopus, Aspergillus fumigatus, Fusarium solani, and Mucor at 36.61, 36.30, 35.03, and 34.84 µg/mg, respectively), and low (Candida tropicalis and Cryptococcus neoformans at 20.78 and 18.61 µg/mg, respectively), were identified. Chitin was not detectable in bacterial isolates used as controls. The chitin detection method offers a sensitive and specific tool for the quantification of chitin in pathogenic fungal isolates. The detection of chitin may be a useful assay for the diagnosis of fungal infections in clinical samples.

Keywords: Biomarker; chitin; diagnosis; HPLC; pathogenic fungi

INTRODUCTION
The cell wall of fungi contains chitin which is a polysaccharide comprising monomers of N-acetylglucosamine (Latge 2007). Chitin is very abundant in nature and is found as structural components not only in fungi but also in arthropods such as insects, crustaceans, cephalopods, and mollusks (Tharanathan & Kittur 2003) but is not present in any known bacteria (Kim 2013). Hence, the presence of chitin in food or raw material (other than sea food that contains crustaceans such as crabs, lobsters, and shrimp)
Fungi: In this study, 10 pathogenic fungal isolates from clinical samples or laboratory stock cultures were selected for testing, namely: Ascomycota fungi: *Aspergillus fumigatus*, *Aspergillus niger*, *Candida albicans*, *Candida tropicalis*, *Penicillium* sp., and *Fusarium solani*; Basidiomycota fungi: *Cryptococcus gattii*, and *Cryptococcus neoformans*; Zygomycota fungi: *Mucor* sp. and *Rhizopus* sp. All of the fungal isolates were maintained on sterile 1.5% (w/v) potato dextrose agar (PDA) slants and then used to inoculate sterile PDA plates which were incubated for 7 days. After 7 days of incubation, 5 mm² pieces of the agar from the actively growing edge of the mycelial colonies were aseptically cut and used to inoculate 1 L flasks containing 500 mL of sterile potato dextrose broth (PDB). The flask cultures were incubated for 7 days on a rotary shaker at 125 rpm at 37 °C. To harvest the mycelial growth, the flask contents were filtered through cheesecloth to trap the mycelia which were further washed 3 times with 100 mL of sterile de-ionized water. The harvested mycelia was freeze-dried and then stored in clean capped sample bottles in a desiccator until further use.

Bacteria: Nutrient agar was used to subculture 4 bacterial isolates, namely: *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Bacillus subtilis*, and *Escherichia coli*. They were then used to inoculate 500 mL flasks containing 250 mL nutrient broth which were then incubated on a rotary shaker at 125 rpm, 37 °C for 2 days. After this incubation, the bacterial biomass was harvested and freeze-dried before being stored in clean capped sample bottles in a desiccator until further use.

Sample preparation: Commercially produced pure chitin of shrimp origin (Sigma, C9752) was used as the chitin standard (0 to 1.5 mg). In order to remove extraneous proteins and amino acids from the samples, 1.0 mL of 0.2 N NaOH was used to treat 100 mg of freeze-dried samples (fungi or bacteria) in 1.5 mL screw-capped microcentrifuge tubes for 6 h at 20 °C. This was followed by a further 17.5 h of incubation in fresh 0.2 N NaOH at a higher temperature of 100 °C. After the second incubation, the samples were washed 4 times in distilled water by resuspension and pelleting via centrifugation and were free of contaminants that could interfere with the glucosamine in the HPLC step. The NaOH-treated samples were then acid hydrolyzed using 1.0 mL of 6 N hydrochloric acid (HCl) in 1.5 mL screw-capped microcentrifuge tubes at 100 °C (Ekblad & Nasholm 1996). The optimal length of hydrolysis time was determined earlier to be 7 h (data not shown) and this was used as the standard procedure in this study.
HPLC-determination of Chitin: To produce a fluorescent product, standard chitin or hydrolyzed samples from the previous step was converted to 9-fluorenymethylchloroformate (FMOC-C1) derivatives. Briefly, 40 µL of acid hydrolysate was first evaporated under reduced pressure before being dissolved in 800 µL of 5 µM homocysteic acid, the latter also serving as internal standard. An aliquot of this solution (200 µL) was pipetted into a 1.5 mL microcentrifuge tube containing 50 µL of 1 M borate buffer (pH 6.3). To start the reaction, 250 µL of 15 mM FMOC-C1 (in acetone) was added followed by incubation at 10 min at room temperature. n-heptane (1 mL) was then used to partition the sample and remove excess reagents by discarding the upper n-heptane phase while retaining the lower phase which contained the FMOC-glucosamine derivatives for HPLC analysis. Reversed-phase HPLC was performed using a waters liquid chromatography system equipped with a 600E pump, WISP 700 auto-injector and a workstation installed with the Empower 3 software to collect and analyze the data. A reversed-phase 250 mm × 5 mm (I.D.) ODS-Hypersil (5 µm) column was used. The HPLC conditions were as follows: initial 10 min of 1 mL/min flow rate of elution in 40 to 50% gradient of methanol in buffer (8 mL acetic acid, 1 mL triethylamine to 1 L water, pH adjusted to 4.2 with NaOH); 10 min of a second isocratic phase; 10 min of a third phase of elution in 50 to 68% methanol; and a final 10 min of elution in 68 to 90% methanol. The eluents flowed through a Waters 2475 fluorescence detector (excitation 260 nm, emission 330 nm).

RESULTS AND DISCUSSION
The detection of chitin in different taxa of pathogenic fungi was evaluated in this study by using HPLC. The amount of chitin in each sample was quantified by comparing the HPLC chromatogram with the chitin standard. Chromatograms of chitin standard showed two peaks with retention times between 1.2 and 2 min. The calibration curve of HPLC peak area for the chitin standard was linear with the values of 2894555, 2703874, 2469531, and 2185492 over the chitin amounts of 100, 50, 20, and 10 mg, respectively (Figure 1, Table 1). The linear increase in HPLC peak area according to chitin amount present in the sample indicates that the chitin extraction method utilized in this study is reliable, enabling accurate measurement of chitin concentration in a given sample. Therefore, the calibration curve was used to calculate the chitin content of fungal and bacterial samples. The calibration curve range in this study is broader and at a higher range (10 - 100 mg) compared to a previous study by Ekblad and Nasholm (1996) who obtained a linear calibration range of 0 - 0.3 mg for hydrolyzed chitin, hence our results show that 9-fluorenymethylchloroformate (FMOC-C1) glucosamine derivatives are reliable indicators of chitin presence even at high sample concentrations.

![FIGURE 1. Calibration curve of chitin standard](image)

| Chitin (mg) | HPLC RT (minutes) | HPLC peak area | HPLC peak height |
|------------|-------------------|----------------|-----------------|
| 100        | 1.12              | 998621         | 2894555         |
| 50         | 1.12              | 443099         | 2703874         |
| 20         | 1.12              | 148212         | 2469531         |
| 10         | 1.12              | 28245          | 2185492         |

RT = retention time
The chitin contents of ten pathogenic fungal isolates are shown in Table 2 and Figure 2. The chitin content varied from 18.61 (± 0.09) to 47.12 (± 0.50) (µg/mg dry mycelial weight). In our tests, C. neoformans showed the highest level of chitin while C. albicans exhibited the lowest chitin level.

### TABLE 2. Chitin content of fungal and bacterial isolates determined by HPLC

| Sample no. | Sample            | HPLC result | Chitin yield* (µg) | HPLC RT (minutes) | HPLC peak area | HPLC peak height |
|-----------|-------------------|-------------|--------------------|-------------------|----------------|------------------|
| 1-Af      | Aspergillus fumigatus | positive    | 36.30              | 1.12              | 311382         | 2582695         |
| 2-An      | Aspergillus niger  | positive    | 46.05              | 1.12              | 415734         | 2998531         |
| 3-Ca      | Candida albicans   | positive    | 47.12              | 1.12              | 427253         | 2919629         |
| 4-Ct      | Candida tropicalis | positive    | 20.78              | 1.12              | 145277         | 3060166         |
| 5-Crg     | Cryptococcus gattii| positive    | 46.98              | 1.12              | 425694         | 3129652         |
| 6-Crn     | Cryptococcus neoformans | positive | 18.61              | 1.12              | 122067         | 2656481         |
| 7-Fs      | Fusarium solani    | positive    | 35.03              | 1.12              | 297855         | 2686931         |
| 8-Mu      | Mucor              | positive    | 34.84              | 1.12              | 235775         | 2546814         |
| 9-Pe      | Penicillium        | positive    | 44.15              | 1.12              | 395475         | 2826747         |
| 10-Rh     | Rhizopus           | positive    | 36.61              | 1.12              | 314694         | 2525958         |
| 11-Bs     | Bacillus subtilis  | negative    | 0                  | -                 | -              | -               |
| 12-Ec     | Escherichia coli   | negative    | 0                  | -                 | -              | -               |
| 13-Pse    | Pseudomonas aeruginosa | negative | 0                  | -                 | -              | -               |
| 14-Ste    | Staphylococcus aureus | negative | 0                  | -                 | -              | -               |

*Chitin yield calculated per mg mycelial dry weight

RT = retention time
The samples could be categorised into 3 groups (high, medium, low) based upon the relative amounts of chitin produced per mycelial dry weight. *C. albicans*, *C. gattii*, *A. niger* and *Penicillium* at 47.12, 46.98, 46.05, and 44.15 µg/mg, respectively, were placed in the high chitin content group. The medium group consisted of *Rhizopus*, *A. fumigatus*, *F. solani* and *Mucor* at 36.61, 36.30, 35.03, and 34.84 µg/mg, respectively, while the low group consisted of *C. tropicalis* and *C. neoformans* at 20.78 and 18.61 µg/mg, respectively. Bacterial isolates did not show any positive result for the HPLC analysis because they do not contain chitin. The bacterial isolates were included in this study to prove that false positive results from bacterial presence in clinical samples would not occur.

The chromatograms of chitin indicate that it was uniformly detected at the same retention time for all fungal isolates tested (Table 2). Two peaks were observed with retention times between 1.2 and 2 min, similar to the chitin standard (Figure 2). The uniformity of chitin detection in fungi from different taxa is a very useful parameter in the clinical situation as a single test may be applied to detect infections caused by different fungal pathogens.

The quantity of chitin in fungi has been estimated to range from 1-2% to up to 40% of the cell wall dry weight (Munro & Gow 2001) and varies in different fungal species or taxonomic groups (Muzarelli et al. 2011). Although chitin is essential for fungal cell viability, its role in fungal virulence is not clear (Munro & Gow 2001), therefore, the variation in chitin quantity observed in this study may not be an indicator of the pathogenicity of the fungal species studied. However, the quantity of chitin in fungal cells may have significance when echinocandin antifungal drugs that target glucan in the cell wall are prescribed. Fungi compensate for drug induced cell wall damage by increased production of chitin to strengthen the cell wall (Walker et al. 2008). Mutant strains of *C. albicans* that had elevated

![Chromatograms of chitin in pathogenic fungal isolates studied.](image-url)
levels of chitin (2-4 fold higher than reference strain) were less susceptible to caspofungin, an echinocandin drug, while a low level of chitin conferred hypersensitivity to the drug (Plaine et al. 2008). Hence, the quantity of chitin in fungal cell walls may play a role in susceptibility to caspofungin. Further studies may provide clues on whether a threshold level for chitin content in fungi correlates to caspofungin resistance.

The chitin content may be used as a convenient marker for the estimation of fungal growth in various samples (Phillips & Gordon 1989). The synthesis of the cell wall and production of its structural components (chitin and glucan) are closely related to the plasma membrane and other membranes (Feofilova et al. 2006). These membranes contain chitin synthases and glucan synthases that construct chitin and glucans to be extruded into the cell wall space. The cell walls of pathogenic fungi such as Candida albicans, Aspergillus fumigatus, and Cryptococcus neoformans contain chitin, chitosan, β-1,3-glucan, β-1,6-glucan, mixed β-1,3-/β-1,4-glucan, α-1,3-glucan, melanin, and glycoproteins as major constituents (Free 2013). Although the detection of chitin cannot be used as an absolute measure of fungal biomass it may be used in comparative studies to give an indication of changes, either an increase or decrease in fungal growth (Gay 1991). Therefore, in a clinical situation, the quantitative detection of chitin in patients’ samples could have both diagnostic and prognostic value.

Joergensen and Wichern (2008) concluded that chitin is the most stable biomarker to assess the total fungal contribution to microbial tissue in soil compared to ergosterol and phospholipid fatty acids. Furthermore, all true fungi contain chitin (Wallander et al. 2013) and the predominant Basidiomycetes, Ascomycetes and Zygomycetes fungal taxa contain a significant chitin concentration of 5% of dry matter (Appuhn & Joergensen 2006). Therefore, the detection of chitin may have advantages over other biomarkers of infections currently utilized to diagnose fungal infections in clinical practice. In the clinical situation, glucan and mannan which are also components of fungal cell wall have been used as diagnostic markers of fungal infection (Arvanitis et al. 2014). Early diagnosis together with correct antifungal treatments contributes to the favorable prognosis of patients with invasive fungal infections (IFI) (Garey et al. 2006). However, current methods such as conventional microbiology often have low sensitivity for the diagnosis of IFI, thus noninvasive, non-culture-based tests are increasingly being used for faster diagnosis (Racil et al. 2010). For example, galactomannan detection is used in the diagnosis of invasive Aspergillus infections (Maertens et al. 2004, 2002), although mannan detection for diagnosis of Candida is less useful (Sendid et al. 2002). Detection of 1,3-β-D-glucan (BG), a fungal-cell-wall polysaccharide, is another non-culture based method of fungal detection. In IFIs by Candida and Aspergillus, BG is released into the bloodstream which can then be detected as a biomarker. However, BG has a limited usefulness for detecting IFI caused by the Basidiomycetes fungi Cryptococcus and Zygomycetes fungi such as Rhizopus sp., both of which are important fungal pathogens (Kedzierska et al. 2007). The detection of chitin in pathogenic fungi from both these taxa (Basidiomycetes and Zygomycetes) in this study is an indication of its superiority as a potential biomarker of fungal presence compared to BG. Chitin has been detected in clinical samples obtained via surgical procedures such as brain tissue (Pisa et al. 2016) and mucin (Taylor et al. 2002) by utilizing chitin-specific proteins which bind in situ to the chitin present in fungal cells. In the fungal cell wall, chitin is linked to β 1,3/1,6 glucan (Latge 2007), hence, in patients with IFI, it is likely that like BG, chitin can also be detected in the bloodstream.

We have shown that chitin can be quantitatively detected in fungal pathogens from all human pathogenic taxa. The presence of chitin in humans with fungal infections needs to be explored further as it may be an alternative biomarker, especially for IFIs. Chitin may be detected in blood samples of patients in a similar way as other biomarkers such as galactomannan, mannan and 1,3-β-D-glucan are detected in serum samples of patients with IFIs. The detection of chitin in clinical samples using chitin binding proteins such as chitinase (Taylor et al. 2002) or anti-chitin antibody (Pisa et al. 2016) implies that commonly used diagnostic methods such as enzyme immunoassays may be applied to detect fungal chitin in a sizable number of clinical fluid samples to evaluate the usefulness of chitin as a biomarker of fungal infections.

CONCLUSION

This study showed that the chitin content of 10 pathogenic fungal isolates from different genera and taxa was clearly detectable in all of the fungi tested. As fungal infections are on the rise and there is a need for diagnostic methods that are able to detect a range of fungi, especially those causing invasive infections, the detection of chitin may provide a sensitive and specific means for the detection of fungal presence in clinical samples.

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