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Molecular cloning and transcriptional activity of a new Petunia calreticulin gene involved in pistil transmitting tract maturation, progamic phase, and double fertilization

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Abstract Calreticulin (CRT) is a highly conserved and ubiquitously expressed Ca\(^{2+}\)-binding protein in multicellular eukaryotes. As an endoplasmic reticulum-resident protein, CRT plays a key role in many cellular processes including Ca\(^{2+}\) storage and release, protein synthesis, and molecular chaperoning in both animals and plants. CRT has long been suggested to play a role in plant sexual reproduction. To begin to address this possibility, we cloned and characterized the full-length cDNA of a new CRT gene (PhCRT) from Petunia. The deduced amino acid sequence of PhCRT shares homology with other known plant CRTs, and phylogenetic analysis indicates that the PhCRT cDNA clone belongs to the CRT1/CRT2 subclass. Northern blot analysis and fluorescent in situ hybridization were used to assess PhCRT gene expression in different parts of the pistil before pollination, during subsequent stages of the progamic phase, and at fertilization. The highest level of PhCRT mRNA was detected in the stigma–style part of the unpollinated pistil 1 day before anthesis and during the early stage of the progamic phase, when pollen is germinated and tubes outgrow on the stigma. In the ovary, PhCRT mRNA was most abundant after pollination and reached maximum at the late stage of the progamic phase, when pollen tubes grow into the ovules and fertilization occurs. PhCRT mRNA transcripts were seen to accumulate predominantly in transmitting tract cells of maturing and receptive stigma, in germinated pollen/growing tubes, and at the micropylar region of the ovule, where the female gametophyte is located. From these results, we suggest that PhCRT gene expression is up-regulated during secretory activity of the pistil transmitting tract cells, pollen germination and outgrowth of the tubes, and then during gamete fusion and early embryogenesis.

Keywords Calcium homeostasis · Calreticulin · Chaperone activity · Gene expression · Nucleolus · Pollen–pistil interactions

Abbreviations
Ca\(^{2+}\) Calcium ions
CRT Calreticulin gene
CRT Calreticulin protein
ER Endoplasmic reticulum
FISH Fluorescent in situ hybridization
M-MLV Moloney murine leukemia virus
MOPS 3-(N-morpholino)propanesulfonic acid
PP1 First stage of the progamic phase
PP2 Second stage of the progamic phase
PP3 Third stage of the progamic phase
Sitt Stigma transmitting tract
Sytt Style transmitting tract
UPI Unpollinated immature pistil
UPM Unpollinated mature pistil
Introduction

In double fertilization, a reproductive system unique to flowering plants, two immotile sperm cells are delivered to the female gametophyte (the embryo sac) by a pollen tube. The male gametes are released into the embryo sac, usually within one of the two synergid cells commonly known as the receptive synergid. One sperm cell fuses with the egg cell to generate a zygote/embryo, whereas the other fuses with the central cell to form the nutritive endosperm. Ca\(^{2+}\) plays essential signaling, physiological, and regulatory roles during this multi-step process, which comprises three successive phases: pollination, progamic phase, and gamete fusion (see review by Ge et al. 2007).

A good candidate protein to facilitate this Ca\(^{2+}\) homeostasis is CRT, a key Ca\(^{2+}\)-binding/buffering ER-resident protein that is highly conserved and extensively expressed in all eukaryotic organisms investigated, with the exception of yeast (see reviews by Jia et al. 2009; Michalak et al. 2009). In animals, CRT is involved in many different intra- and extracellular processes, such as Ca\(^{2+}\) storage and signaling, molecular chaperone activity in the ER, regulation of gene expression, control of cell adhesion and migration, immune regulation, apoptosis, and pathogenesis (see reviews by Michalak et al. 2009; Gold et al. 2010; Wang et al. 2012). Plant CRT has the same molecular structure as the animal protein and shares its chaperone and Ca\(^{2+}\) binding activities (see reviews by Crofts and Dencke 1998; Mariani et al. 2003; Jia et al. 2009; Thelin et al. 2011; Li and Yang 2012).

Two CRT genes (CRT1 and CRT2) exist in the human, mouse, pig, and rat genomes (Persson et al. 2002). The expression profiles of these two mammalian genes have led to the suggestion that CRT1 is the main protein isoform, whereas CRT2 is testis specific. More than 120 plant CRT sequences are described in current gene databases, and CRT genes have been isolated from barley (Chen et al. 1994), tobacco (Denecke et al. 1995), maize (Kwiatkowski et al. 1995; Dresselhaus et al. 1996), Chinese cabbage (Lim et al. 1996), Ricinus (Coughlan et al. 1997), Arabidopsis (Nelson et al. 1997), rice (Li and Komatsu 2000), and wheat (Jia et al. 2008; An et al. 2011). In higher plants, the CRT family consists of three members, which are classified into two distinct subclasses: CRT1/CRT2 (also designated CRT1a/CRT1b) and CRT3 (Nelson et al. 1997; Persson et al. 2003; Christensen et al. 2010). Sequence homology of plant CRTs suggests that CRT1 and CRT2 are similar to each other and correspond with the animal CRT1, whereas the plant-specific CRT3 genes are more highly conserved across species (see review by Jia et al. 2009).

CRT expression in plants has been correlated with tissue regeneration, immunity, abiotic stress responses, and cell-to-cell communication by plasmodesmata (see reviews by Jia et al. 2009; Thelin et al. 2011). There are also some indications that CRT may be involved in reproductive events in plants. CRT is expressed abundantly in different flower organs (Denecke et al. 1995; Hassan et al. 1995; Coughlan et al. 1997; Nelson et al. 1997; Borisjuk et al. 1998; Navazio et al. 2002; Hsieh and Huang 2005; Nardi et al. 2006; Christensen et al. 2010) and during pollen–pistil interactions, fertilization, embryogenesis, and seed germination (Chen et al. 1994; Denecke et al. 1995; Dresselhaus et al. 1996; Coughlan et al. 1997; Nelson et al. 1997; Williams et al. 1997; Borisjuk et al. 1998; Lenartowska et al. 2001, 2002, 2009; Christensen et al. 2010; Li et al. 2011). CRT’s expression pattern suggests that it could play a role in regulation of Ca\(^{2+}\) homeostasis during pollen–pistil interactions and thus contribute to successful fertilization. Some reports indicate that plant CRT expression is modulated during sexual reproduction, and in some plants a higher level of CRT transcripts has been detected in gametes and zygotes than in vegetative cells (Denecke et al. 1995; Dresselhaus et al. 1996; Nelson et al. 1997). Our previous work (Lenartowska et al. 2001, 2009) examined the possibility that CRT is expressed during pollination in Petunia and Haemanthus. However, at the time of those studies, we were limited to examining CRT mRNA expression using probes derived from other plants. Here we report cloning of the Petunia CRT homolog, PhCRT. We identify a full-length cDNA sequence and describe its molecular characteristics and phylogenetic relationships to other plant CRT genes. We show that PhCRT is highly expressed during pistil transmitting tract maturation, pollen germination and tube outgrowth on the stigma, and then during fertilization and early embryogenesis. We discuss the timing of expression and subcellular localization of PhCRT transcripts with respect to the possible functions of CRT in plant sexual reproduction.

Materials and methods

Plant material

Commercial cultivars of Petunia hybrida were grown at room temperature (Laboratory of Developmental Biology, Faculty of Biology and Environment Protection, Nicolaus Copernicus University, Toruń, Poland). Whole Petunia pistils were dissected from closed flowers 1 day before anthesis, from unpollinated flowers at anthesis, and from flowers pollinated with compatible pollen at anthesis. To examine pollen tube growth during the progamic phase, pollinated pistils were dissected from the flowers at different time points after pollination, cut along the longitudinal axis, stained with 0.1 % aniline blue according to the standard...
protocol, and observed by fluorescence microscopy. For Northern blot analysis, whole pistils or pistils divided into stigma–style fragments and ovaries were frozen in liquid nitrogen and stored at −80 °C until they were used. For fluorescent in situ hybridization, samples of unpollinated and pollinated pistils (fragments of stigmas and ovaries) were fixed, dehydrated in ethanol, and embedded as previously described (Lenartowska et al. 2009).

RNA extraction and RT-PCR analysis

Plant material (100 mg) was ground in liquid nitrogen. Total RNA was extracted in TRI Reagent® (Sigma) according to the manufacturer’s protocol. First-strand CRT cDNA synthesis was performed with 1 μg of total RNA, Super-Script III reverse transcriptase, and an oligo(dt)20 primer according to the manufacturer’s instruction (Life Technologies). Nested PCR was done to amplify the internal part of the PhCRT cDNA. In brief: 2 μl of first-strand cDNA was used as template for PCR amplification with Pfx50™ DNA polymerase and outer gene-specific primers (Table 1). A 1 μl aliquot of the first PCR mixture served as the template in a second PCR using the inner gene-specific primers (Table 1). PCR cycles were as follows: 95 °C for 15 s followed by 35 cycles of 95 °C for 30 s, 57 °C for 1 min, 72 °C for 1 min, followed by a final extension step of 72 °C for 4 min. A 1-μl aliquot of the outer 3′-RACE PCR mixture served as the template in the inner 3′-RACE PCR using 3′-RACE Inner Primer and 3′-RACE gene-specific inner primer (Table 1). PCR cycles were as above.

For 5′ RACE, total RNA was dephosphorylated with calf intestinal phosphatase and treated with tobacco acid pyrophosphatase to remove the 5′ cap structure from the full-length mRNA. 5′-RACE Adapter was ligated to treated mRNA with T4 RNA ligase (see the full-length sequence

| Amplified sequence/type of reaction | Sense primer | Antisense primer |
|-----------------------------------|-------------|-----------------|
| 3′ RACE reverse transcription | 3′-RACE gene-specific outer primer (RL035) | 3′-RACE adapter |
|                                  | 5′-GGAAAGCAACCTATGATGTGACAACC-3′ | 5′-GCGACAGAAATATTACGACCTCAC TATAAGT12VN-3′ |
| 3′ CRT cDNA end/out 3′-RACE PCR | 3′-RACE gene-specific inner primer (RL036) | 3′-RACE outer primer |
|                                  | 5′-TCAAGGATGACCCCGATCTC-3′ | 5′-GCGACAGAAATATTACGACCTC-3′ |
| 3′ CRT cDNA end/in 3′-RACE PCR  | 3′-RACE gene-specific inner primer (RL036) | 3′-RACE inner primer |
|                                  | 5′-GCGCCGATGCTTAATACGACCTC-3′ | 5′-GCGACAGAAATATTACGACCTC-3′ |
| 5′ CRT cDNA end/out 5′-RACE PCR  | 5′-RACE outer primer | 5′-RACE gene-specific outer primer (RL037) |
|                                  | 5′-GCTGATGCGATGGAATGAAACCTG-3′ | 5′-CCACACAGTCAGTCTGACT-3′ |
| 5′ CRT cDNA end/in 5′-RACE PCR  | 5′-RACE inner primer | 5′-RACE gene-specific inner primer (RL038) |
|                                  | 5′-CGCGGATCAGAACACTGCTTGGTCTC-3′ | 5′-GCTGATGCGATGGAATGAAACCTG-3′ |
| Internal part of Petunia CRT/first PCR | CRT outer primer (RL039) | CRT outer primer (RL040) |
|                                  | 5′-GAGGGTTGGCAAATGGCTACTCAA-3′ | 5′-GAGGGTTGGCAAATGGCTACTCAA-3′ |
| Internal part of Petunia CRT/second PCR | Nested gene-specific primer for CRT (RL041) | Nested gene-specific primer for CRT (RL042) |
|                                  | 5′-GCTGATGCGATGGAATGAAACCTG-3′ | 5′-GCTGATGCGATGGAATGAAACCTG-3′ |
| Full-length Petunia CRT cDNA | RL068 | 5′-TGGTAAATTTCGATCATCAC-3′ |
|                                  | 5′-AGCTCGATATTTCCATCACATCAGCTG-3′ | 5′-TGGTAAATTTCGATCATCAC-3′ |
| Internal part of Petunia 18S rRNA | RL097 | 5′-ACTAGGACGTTATCGATCGCT-3′ |
|                                  | 5′-GAGCCATATGGTCCAACCAC-3′ | 5′-GAGCCATATGGTCCAACCAC-3′ |

Sequence of the adapter ligated to treated mRNA 5′-GCUGAAGGCGGAUAGGAACACUGCGUUUGCGGCUUUGAGAAA-3′ in the 5′ RACE reaction.
under the Table 1). Reverse transcription was performed with random decamers in the presence of the M-MLV enzyme and RNase inhibitor at 42 °C for 1 h. The 5' RACE primers were designed according to the cloned internal fragment sequence of the PhCRT cDNA. A 1-μl aliquot of the reverse transcription reaction was used in the outer 5'-RACE PCR in the presence of the 5' RACE Outer Primer, 5'-RACE gene-specific outer primer and PfX50™ DNA polymerase (Table 1). PCR cycles were as follows: 95 °C for 15 s followed by 35 cycles of 95 °C for 30 s, 68 °C for 1 min, 72 °C for 1 min, followed by a final extension step of 72 °C for 4 min. A 1-μl aliquot of the outer 5'-RACE PCR mixture served as the template in an inner 5'-RACE PCR using the 5'-RACE Inner primer and 5' RACE gene-specific inner primer (Table 1). PCR cycles were as above. The 3' and 5' RACE PCR products were visually inspected on a 1 % agarose gel in TBE buffer.

Plasmid construction and molecular probe synthesis

PhCRT RT-PCR or RACE products were cloned into the pJET1.2 vector using the CloneJET™ PCR Cloning Kit (Thermo Scientific) according to the protocol provided with the kit. The insert was verified by sequencing for its correct amplification and orientation in pJET. To prepare the full-length PhCRT antisense probe, the plasmid containing the PhCRT insert was cut with Xba I and purified on CHROMA SPIN™ + TE – 100 columns. A 1-μg aliquot served as template to generate the digoxigenin (DIG)-labeled probe using the DIG RNA Labeling Kit (SP6/T7) according to the manufacturer’s instructions (Roche). An RT-PCR cDNA containing the internal part of the Petunia 18S rRNA sequence was cloned in pJet 1.2 vector. DIG-labeled 18S probe was generated by PCR according to the manufacturer’s instructions (PCR DIG Synthesis Kit, Roche) with uncut plasmid DNA as the template (Table 1).

Northern blot analysis

Total RNA concentrations were measured with a NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies), and RNA quality was assessed by visualization on a 1 % agarose gel in 1× TBE buffer. A 15-μg aliquot of total RNA was dried under vacuum, resuspended in 5 μl of RNase-free H2O, supplemented with two volumes of denaturing buffer containing 50 % formamide, 6.1 % formaldehyde, 1× MOPS, 1× loading dye buffer, and 40 mg/dm−3 ethidium bromide, and heat denatured. RNA was resolved on a 1.2 % formaldehyde-agarose gel, capillary transferred onto a nylon membrane (Hybond-N+, Amersham) in 20× SSC buffer, rinsed with distilled water, and UV cross-linked in the G-S Gene Linker® UV Chamber (BioRad). Membranes were prehybridized for 40 min in DIG Easy Hyb buffer (Roche) at 65 °C, and then hybridized overnight to the DIG-labeled full-length PhCRT antisense probe at a final concentration 150 ng/ml. Chemiluminescence detection was performed according to the manufacturer’s instruction (DIG Luminescent Detection Kit, Roche). To reprobe the membrane with Petunia 18S DIG-labeled probe, the blot was stripped twice at 80 °C in 50 % formamid, 5 % SDS, 50 mM Tris–HCl, pH 7.5, for 60 min, and then rinsed with 2× SSC. Rehybridization was performed in the same conditions as hybridization described above. Northern hybridization was performed a minimum of three times for each experiment, and representative data are shown. Quantification of signals was done with Image Gauge 3.4 software (Science Lab99). All data obtained from the Northern blot experiments were subjected to one-way ANOVA test.

Fluorescent in situ hybridization (FISH)

Samples of plant material were fixed with freshly prepared 4 % formaldehyde in phosphate-buffered saline (PBS), pH 7.2, overnight at 4 °C. After fixation, specimens were washed in PBS, dehydrated through a graded series of ethanol, embedded in LR Gold resin (Fluka), sectioned with a diamond knife into semi-thin sections (longitudinal sections of stigmas and ovary fragments), and transferred onto microscope slides covered with Biobond (BioCell). The DIG-labeled antisense PhCRT RNA probe generated above was used at a final concentration of 0.5 μg/μl. Prehybridization and hybridization were carried out in 50 % formamide, 4× SSC, 5× Denhardt’s, and 50 mM sodium phosphate buffer for 1 h at 42 °C and overnight at 37 °C, respectively. Signals were detected using primary mouse anti-DIG and secondary goat-anti-mouse IgG-Alexa Fluor 488 antibodies (Roche and Life Technologies, respectively). A no-probe control was also performed. In the final step, DNA was stained with 2 μg/ml 4', 6-diamidino-2-phenylindole (DAPI, Fluka). Images were acquired using an Olympus BX50 fluorescence microscope, Olympus XC50 digital color camera, and CellB software (Olympus Soft Imaging Solutions GmbH).

In silico sequence analysis

The Neighbor-joining tree estimated from distances between the plant CRT amino acid sequences was built using Mega version 4 [http://www.megasoftware.net/], using Tajima-Nei distance and 1000 bootstrap (Tamura et al. 2007). Plant CRTs were aligned in ClustalW2 [http://www.ebi.ac.uk/Tools/msa/clustalw2/] (Larkin et al. 2007). PhCRT molecular mass and phosphorylation sites were predicted by ProtParam and NetPhos programs, respectively (Blom et al. 1999; Gasteiger et al. 2007).
CRT amino acid sequences from different species were retrieved from GenBank [http://www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov). *Petunia* Expressed Sequence Tags (EST) database sequences showing homology to the *Arabidopsis CRT* cDNA clone (NM_001036122) were identified with the BLAST program [http://blast.ncbi.nlm.nih.gov/Blast.cgi](http://blast.ncbi.nlm.nih.gov/Blast.cgi) (Altschul et al. 1990). Primer3Plus was used to design gene-specific primers [http://www.bioinformatics.nl/cgi-bin/primer3plus/primer3plus.cgi/](http://www.bioinformatics.nl/cgi-bin/primer3plus/primer3plus.cgi/) (Untergasser et al. 2007). Statistical analysis was carried out by one-way ANOVA [http://www.danielsoper.com/statcalc3/calc.aspx?id=43](http://www.danielsoper.com/statcalc3/calc.aspx?id=43).

**Results**

*PhCRT* gene cloning and sequence analysis

Full-length *Arabidopsis CRT* cDNA sequence (NM_001036122) was used to search the *Petunia* EST database. Three *Petunia* EST sequences (CV296977.1, CV296202.1, and DC240877.1) showed homology to the 5' end of the *Arabidopsis CRT* cDNA, and one (DW177206.1) showed homology to the 3' end. Because none of the 5' EST sequences overlapped with the 3' EST sequence, we designed primers to amplify the *PhCRT* cDNA inner sequence. The obtained 982-bp inner sequence was then used to design gene-specific primers that were used to clone the *PhCRT* 3' and 5' ends by rapid amplification of cDNA ends. The resulting fragments were sequenced and used to design a set of primers to amplify a full-length *PhCRT* cDNA, which was cloned into the pJET 1.2 vector. The *PhCRT* cDNA sequence is 1,550 bp and consists of a 30-bp 5' untranslated region (UTR) upstream of the ATG initiation codon, a 1,254-bp open reading frame (ORF) that terminates with a TAA stop codon, and a 266-bp 3' UTR (Fig. 1). Two putative polyadenylation signal sequences (ATTAAA and TTAAT) (Nevins 1983) were mapped to the 3'-end of the *PhCRT* sequence, 17 and 16 bp upstream of the poly(A) tail, respectively. (The full-length *PhCRT* cDNA sequence was submitted to the European Nucleotide Archive (accession number HG738129)).

Deduced amino acid sequence analysis of the *PhCRT* cDNA clone

The putative *PhCRT* ORF encodes a relatively acidic (pI 4.43, as predicted by ProtParam software (Gasteiger et al. 2005) 417-amino acid polypeptide with a calculated molecular mass of 47.77 kDa. The predicted molecular mass is smaller than that of native CRT isolated from *Petunia*, which migrates at 58 kDa on an SDS-PAGE gel (data not shown). The NetPhos program (Blom et al. 1999) predicts that *PhCRT* has eight to ten putative phosphorylation sites for serine, tyrosine, or threonine kinases. Similar to other plant CRTs, *PhCRT* has three evolutionarily conserved domains: N, P and C. The globular N-domain starts with a predicted signal peptide and contains an N-glycosylation consensus sequence (NHTS) at amino acids 59–62 and a microbodies targeting site at amino acids 162–164 (Fig. 1). The location of the N-glycosylation site is species-specific within the plant CRT family (see review by Jia et al. 2009). A search of structural motifs revealed two conserved CRT-family signature regions, calreticulin 1 and calreticulin 2, in the N-domain (Fig. 1). Three cysteine residues were found in this region (Fig. 1); the second and third cysteine residues could form a disulphide bridge that is essential for proper folding of the N-terminal region (see review by Jia et al. 2009). The N-domain is also enriched for histidine residues, which most likely mediate Zn$^{2+}$ binding (see review by Michalak et al. 2009). A highly conserved proline-rich region, the P-domain, extends from amino acid 208 to amino acid 309. This region contains three repeats each of two proline-rich sequences, M1 (PXXIXDPXXKKPeXWDe) and M2 (GXWXAXXNPXYK) (Fig. 1). Both motifs contain a high percentage of acidic amino acids, which are required for low capacity, but relatively high-affinity Ca$^{2+}$-binding; this region is responsible for the CRT lectin-like chaperone activity (see review by Michalak et al. 2009). The C-terminus of CRT is enriched in negatively charged residues that are responsible for binding Ca$^{2+}$ with relatively high capacity, but low affinity. This domain ends with an HDEL sequence, which is required for targeting and retention of CRT in the ER lumen (see review by Jia et al. 2009).

The *PhCRT* sequence was compared to several of the full-length CRT1/CRT2 sequences found in the GenBank database. As shown Fig. 2, the ClustalW2 program revealed that the deduced amino acid sequence of *PhCRT* shared high identity with CRTs from *Nicotiana* (91.83 %), *Arabidopsis* CRT1a and CRT1b (80.1 and 76.74 %, respectively), *Oryza* CRT1a and CRT1b (71.22 and 74.1 %, respectively), and maize CRT1 and CRT2 (74.58 and 72.66 %, respectively). We used MEGA 4.1 and the neighbor-joining method to generate a phylogenetic tree of the plant CRT amino acid sequences. *Human CRT*, which shares 48.4 % identity with *Petunia CRT* (Fig. 2) was used as an outgroup. The resulting tree (Fig. 3), groups the eleven plant CRTs into two large clusters of CRT1a/b (or CRT1/CRT2) and CRT3. Cluster analysis suggests that the *PhCRT* gene belongs to the CRT1/CRT2 group (Fig. 3). The CRT1/CRT2 group contains the CRTs that play general chaperone functions in plants (Christensen et al. 2010).
Expression pattern of the PhCRT gene before pollination and during the progamic phase

The Petunia pistil is composed of a solid style filled with transmitting tissue and a wet stigma that is covered with exudate at the receptive stage. As shown in Fig. 4, three main stages of the progamic phase have been defined: a first stage, when pollen germinates and tubes outgrow on the stigma (PP1, Fig. 4b); a second stage, when pollen tubes penetrate the stigma/style transmitting tract (PP2, Fig. 4d, e); and a third stage, when pollen tubes grow into the ovules (PP3, Fig. 4f). Figure 4a and c shows fragments of the stigma and the ovary, respectively, of unpollinated mature pistils at anthesis (UPM pistil with exudate on the stigma). The embryo sac (female gametophyte) of Petunia develops according to the Polygonum-type (Willemse and Van Went 1984) and consists of two synergids, the egg cell, the central cell, and three antipodals (Van Went and Kwee 1990). A schematic representation of the Petunia ovule during the PP3 stage and at fertilization is shown in Fig. 4g–i.

To analyze the expression pattern of PhCRT in relation to anthesis, pollination, and pollen tube growth from the stigma to the ovary, we first investigated PhCRT mRNA transcript levels in whole pistils at 1 day before anthesis (unpollinated immature pistil without exudate on the stigma, UPI), at anthesis (UPM), and during the PP1, PP2, and PP3 stages after pollination. Northern blot analysis of total Petunia pistil RNA revealed that the highest level of PhCRT mRNA was detected during the PP3 stage after pollination.
PhCRT expression was at the UPM stage. Expression then decreased gradually through the PP1, PP2, and PP3 stages (Fig. 5a). This result was somewhat surprising because some previous studies in other plant species reported that the highest levels of CRT mRNA transcripts were correlated with fertilization, embryogenesis, and seed development (Chen et al. 1994; Nelson et al. 1997; Borisjuk et al. 1998; Navazio et al. 2002). However, most other studies have been carried out on ovaries, and the transcriptional activity of CRT genes in whole pistils before pollination and during subsequent stages of the progamic phase has never been examined. Therefore, we compared PhCRT mRNA transcript levels in unpollinated pistils before and at anthesis (UPI and UPM pistils, respectively). In addition, to assess developmental and tissue-specific expression of PhCRT, we divided the pistils into stigma–style fragments (Fig. 5b) and ovaries (Fig. 5c). Whereas comparable levels of PhCRT mRNA transcripts were observed in the ovaries dissected from UPI and UPM pistils (Fig. 5c), levels of PhCRT mRNA were about threefold higher in stigma–style fragments of UPI pistils than in the same fragments of UPM pistils (Fig. 5b). These results suggest that elevated expression of PhCRT before pollination correlates with maturation of the stigma–style transmitting tract.

**Proline-rich domain**

**C-terminal domain**

**Fig. 2** Sequence alignment of PhCRT with other known plant CRTs: *Arabidopsis* (At CRT1a, At CRT1b), *Nicotiana* (Nt CRT), *Oryza* (Os CRT1a, Os CRT1b), and *Zea* (Zm CRT1, Zm CRT2). PhCRT amino acid sequence is given in bold. Sequence data were obtained from GenBank with the following accession nos.: *Arabidopsis thaliana* (crt1a, aac49695; crt1b, aee28414); *Nicotiana tabacum* (acH72686); *Oryza sativa* (crt1a, BaF13713; crt1b, BaF21197); *Zea mays* (crt1, caa86728 and crt2, aaF01470). *Petunia hybrida* crt1 (Hg738129) was deposited in the European Nucleotide Archive. The globular N-domain and C-terminal domain are indicated by lines under the amino acid sequence. The numbers on the right indicate the amino acid position. Asterisks denote positions that have a single, fully conserved residue. Colon or dot indicates conservation between groups of strongly similar properties (>0.5 in the Gonnet PaM 250 matrix) or weakly similar properties (<0.5 in the Gonnet PaM 250 matrix), respectively (Larkin et al. 2007).
of PhCRT or stabilization of the transcript. PhCRT mRNA levels dropped at PP2, and PP3, when the entire stigma–style transmitting tract is penetrated by growing pollen tubes. By contrast, in the ovaries, a progressive increase of PhCRT mRNA level was observed after pollination and through the progamic phase. The PhCRT mRNA level was highest at PP3, when pollen tube ingrowth into the ovule and fertilization is highly probable (Fig. 5c). These results indicate that PhCRT transcriptional activity or mRNA stabilization in ovaries might be stimulated by progressive growth of the pollen tubes in the pistil. Moreover, the highest level of PhCRT mRNA transcripts in the ovary at the PP3 stage could be indicative of a response to fertilization.

Subcellular localization of the PhCRT mRNA transcripts in stigmas and ovaries before and after pollination

To determine the spatial distribution of PhCRT mRNA transcripts in the somatic cells of the stigma transmitting tract (sitt), germinated pollen, growing pollen tubes, and various cells of the ovaries (somatic cells of placenta and ovule and female gametophyte cells), samples of stigmas and ovaries dissected from unpollinated and pollinated pistils were fixed, embedded, sectioned, and processed for FISH and visualized by fluorescence microscopy before pollination and at the beginning and end of the progamic phase. Longitudinal sections of stigmas dissected from UPI pistils showed accumulation of PhCRT mRNA transcripts in the cytoplasm of the sitt secretory cells (Fig. 6a). In those cells, mRNAs were also found to be abundant in the nuclei (Fig. 6b, b'). After anthesis, when stigma exudate was intensively secreted (Fig. 6d, stars), PhCRT mRNA transcripts were observed in the cytoplasm of sitt cells (Fig. 6d). However, at this stage (UPM pistil) the level of fluorescence was lower than observed at the UPI stage, and signal was occasionally observed in close proximity to the sitt cell nuclei (Fig. 6d). No labeling was observed when the probe was omitted (Fig. 6c).

After pollination, the strongest hybridization signals were observed at the PP1 stage in hydrated (Fig. 7a, a') and germinated (Fig. 7b, c') pollen as well as in outgrowing pollen tubes (Fig. 7d, d'). Pollen hydrated and germinated on the stigma surface showed uniform fluorescence throughout the cytoplasm and associated with the active aperture region of germinating pollen (Fig. 7b, b'). As shown in Fig. 7d, e, at the PP1 stage, pollen tubes of various lengths were visible on the stigma sections. In short tubes, PhCRT mRNA transcripts were almost uniformly distributed in the cytoplasm with the exception of an extra-apical zone of the tip-growing cell, where signal was weaker (Fig. 7d). In elongated tubes, hybridization signals were detected predominantly along the cytoplasm...
of the tip-growing cells, and only small regions (probably vacuoles) were devoid of fluorescence (Fig. 7e). In images showing cross-sections of growing pollen tubes, mRNA enrichment was observed at the sub-apical region, where no vacuoles are present (Fig. 7f). At the PP1 stage, sitt cells showed weaker labeling than they did before pollination.

Fig. 4 Aniline blue staining (glaucous fluorescence) of germinated pollen and pollen tubes growing in planta (a–f) and schematic representations of mature Petunia ovules before and at fertilization (g–i). All pictures from a to f show longitudinal sections of stigma, style, or ovary fragments. a Stigma of UPM pistil; autofluorescence (red) of starch in amyloplasts is visible in the stigma transmitting tract (sitt). b Stigma of pollinated pistil at the PP1 stage; germinated pollen and outgrowing tubes are visible in the sitt. c Ovary of UPM pistil; similar images showing lack of pollen tubes were observed in pollinated pistils at the PP1 and PP2 stages (data not shown). Stigma (d) and style (e) at the PP2 stage; whole sitt and style transmitting tract (sytt) are penetrated by pollen tubes; some callose plugs (bright points) are visible in long tubes (e). f Ovary of pollinated pistil at the PP3 stage; pollen tubes growing on the placenta (pl) and ingrowing into ovules (ov) are visible at this stage. Black stars on a and b show vascular strand, white star on f shows pollen tube ingrowth into an ovule. g Unfertilized ovule at the PP3 stage; the embryo sac contains the egg apparatus, which consists of two synergids (sy), the egg cell (ec), the central cell (cc), and three antipodals (an). h Ovule at the end of the PP3 stage (moment of sperm cells release into the receptive synergid). i Fertilized ovule with the zygote (zy) and endosperm (en); the receptive synergid is degenerated. cx cortex, es embryo sac, fn funicle, in inner integument, mi micropyle, nu nucellus, oin outer integument, pt pollen tube, sn sperm nuclei, vn vegetative nucleus. Bars 100 μm
In these cells, PhCRT mRNA transcripts were localized in peripheral cytoplasm and in close proximity to the cell nucleus (Fig. 7f). At the PP3 stage, hybridization signals were only detected in residual cytoplasm along the edge of the vacuolated pollen grains/tubes and crushed sitt cells (Fig. 7g). These results are consistent with our northern blot analysis and confirm that the highest level of PhCRT mRNA accumulation correlates with sitt maturation (which results in the presence of exudates on the receptive stigma), and with pollen germination and pollen tube outgrowth in the stigma.

To gain insight into possible differential transcriptional activity of PhCRT, we paid special attention to FISH signals in the nuclei, as these may indicate nascent transcription. Hybridization signals were never detected in generative and vegetative nuclei of hydrated pollen (Fig. 7a, a'). By contrast, label was observed in vegetative nuclei in early germinating pollen (Fig. 7c, c') and in pollen with very short tubes (Fig. 7d, d'). No signal was detected in vegetative nuclei of elongated pollen tubes (Fig. 7e).

We next examined PhCRT FISH signals in longitudinal sections of the ovary before pollination (Fig. 8). At this stage, somatic cells of the placenta and the ovule (nucellus and integuments cells) exhibited diffuse signal throughout the cytoplasm (Fig. 8a, b, respectively). The transcripts were also found in the nuclei of these cells (Fig. 8a, b, arrows). Similar localization patterns were observed in the cytoplasm of the female gametophyte cells, particularly at the micropylar end of the embryo sac. Within each embryo sac, both synergids (Fig. 8c), the egg cell (Fig. 8d, e), and the central cell (Fig. 8f) were all positive for PhCRT mRNA transcripts. By contrast, antipodals localized on the chalazal end of the embryo sac showed weaker labeling in the cytoplasm (Fig. 8g). Before pollination, hybridization signals were associated only with the nuclei of the egg apparatus cells (Fig. 8c, d, arrows), and no labeling was detected in the nuclei of the central and antipodal cells.

Detailed observations of the ovary samples dissected from pollinated pistils at the PP3 stage revealed that PhCRT mRNA transcripts were enriched at the micropylar region of the ovule, where the female gametophyte is located and fertilization occurs inside the embryo sac (Fig. 9, 10). Just before gamete fusion, the strongest signals were detected in pollen tubes, which penetrated the placenta surface (Fig. 9a), and in the micropylar canal of the ovule (Fig. 9b). Preferential accumulation of PhCRT mRNA transcripts was also observed in the cytoplasm of the receptive synergid just before gamete fusion (Fig. 9c), when two sperm cells were released (Fig. 9c, boxed region), and during fertilization (Fig. 10b). A PhCRT mRNA transcript gradient
was visible in the cytoplasm of the receptive synergid at the moment of sperm cells release, at which point signal accumulated in the micropylar pole of the cell (Fig. 9c, arrows). Labeling was also detected in both sperm cells (Fig. 9c’, c’’). During fertilization, high levels of \( \text{PhCRT} \) mRNA transcripts were observed in the zygote (Fig. 10a–c) and endosperm cells (Fig. 10a, d). In these cells, hybridization signals were detected not only throughout the cytoplasm, but also in nuclei (Fig. 10a) and nucleoli of the zygote (Fig. 10a, c, stars and c’, dotted line) and endosperm cells (Fig. 10a, d, arrows and d’, dotted line). At the PP3 stage, high labeling was seen in the cytoplasm of the nucellus cells within both the micropylar and chalazal regions of the ovule (Figs. 10d, 9d, respectively), but very little signal was detected in degenerating synergid (Fig. 10a) or antipodal cells (Fig. 9d). These results are consistent with our Northern blot analysis and confirm that the highest accumulation of \( \text{PhCRT} \) mRNA in the ovary correlates with the late stage of the progamic phase, when pollen tube grows into a receptive synergid, and with gamete fusion and early embryogenesis.

**Discussion**

\( \text{PhCRT} \) is a highly conserved gene belonging to the CRT1/CRT2 subfamily

Here we report the first cloning of a \( \text{CRT} \) gene, \( \text{PhCRT} \), from the flowering plant \( \text{Petunia} \). Alignment of the deduced amino acid sequence indicates that \( \text{PhCRT} \) shares homology with other plant CRTs, from 71 to 91 % identity. \( \text{PhCRT} \) appears to be most similar to \( \text{Nicotiana} \) CRT and \( \text{Arabidopsis} \) CRT1a and CRT1b isoforms, whereas the similarity between \( \text{PhCRT} \) and human CRT is lower, only about 48 %. Such low identity level has also been observed between mammalian CRT and other plant CRTs (Li and Komatsu 2000; Jia et al. 2008). Although multiple plant CRT isoforms exist, several regions in their sequences are conserved. Our in silico analysis of the deduced \( \text{PhCRT} \) amino acid sequence revealed that it has all the typical domains and motifs of a CRT: an N region that contains an ER-targeting sequence, a NHTS consensus sequence, calreticulin 1 and calreticulin 2 motifs, three cysteine residues, a microbodies targeting site, and six histidine residues; a P-domain containing three repeats each of the proline-rich sequences M1 and M2, and a putative nuclear targeting sequence; and a C domain containing an HDEL motif.

Generally, plants contain three CRT isoforms: CRT1 and CRT2 (in \( \text{Arabidopsis} \) CRT1a and CRT1b) represent one subgroup, and CRT3 is a divergent member (see reviews by Jia et al. 2009; Thelin et al. 2011). Some recent reports revealed that the CRT1 and CRT2 family members are components of a general ER chaperone network, whereas the CRT3 gene is co-expressed with pathogen- and signal transduction-related genes, suggesting functional specialization (Jin et al. 2009; Li et al. 2009; Saijo et al. 2009; Christensen et al. 2010). Our phylogenetic analysis indicates that the \( \text{PhCRT} \) cDNA clone that we identified belongs to the CRT1/CRT2 subclass. This is consistent with the presence of an N-glycosylation site at amino acids 59–62. Location of this site near position 50–60 in the N-domain is shared among some other plant CRT1/CRT2 isoforms; in CRT3, the N-glycosylation site is usually closer to amino acid 96 (see review by Jia et al. 2009). Moreover, it is thought that tissue-dependent expression differs among members of the CRT family. Whereas CRT1 and CRT2 are most abundant in floral, root, and leaf tissues in both \( \text{Arabidopsis} \) and maize, CRT3 shows highest expression in leaves and roots, but not in flowers. Given the sequence features and our observations that the \( \text{PhCRT} \)
Fig. 7 Localization of PhCRT mRNA (green) in stigmas of pollinated pistils at PP1 (a–f) and PP3 (g) stages. a, a’ Pollen hydrated on the stigma. b–c’ Early germinated pollen. d, d’ Germinating pollen grain with a very short tube. e Longitudinal section of a pollen tube elongated on the stigma. f Cross-section of the sub-apical zone of a pollen tube growing between sitt cells. g, g’ Vacuolated pollen grain/tube on the stigma. Positions of sitt cell nuclei and generative/vegetative nuclei of the male gametophyte (pollen grain/tube) are stained with DAPI (blue in a’, b, c’, d’, e, f, g’). aa active aperture, gn generative nucleus, pg pollen grain, pt pollen tube, sitt stigma transmitting tract cell, vn vegetative nucleus. Bars 10 μm
gene that we identified is expressed in the pistil, we conclude that it encodes a CRT1/CRT2 homolog and most likely plays general roles associated with protein folding and Ca\(^{2+}\) homeostasis.

The highest expression of PhCRT in the ovary correlates with pollen tube entry into the embryo sac, fertilization, and early embryogenesis.

We observed high levels of PhCRT mRNA in the micropylar region of the ovule before and after pollination and dramatic up-regulation of PhCRT expression at the PP3 stage, when transcript accumulation was observed in receptive synergids, and then in zygotes and early endosperms. Hybridization signals were detected in both the cytoplasm and nuclei of these cells, suggesting high transcriptional activity. We did not observe such dramatic increase in PhCRT expression level at the PP3 stage in somatic cells of the placenta and ovule, nor in the antipodal cells, which do not participate in double fertilization.

Our Northern blot and FISH data indicated that the maximum level of PhCRT expression in the ovary is at the late stage of the proagamic phase, when the pollen tube has released sperm cells into the receptive synergid and fertilization has occurred. These results are consistent with findings in several other plant species. Elevated levels of CRT mRNA transcripts were observed at the time of fertilization in whole barley ovaries (Chen et al. 1994) in maize zygotes and immature embryos after in vitro fertilization (Dresselhaus et al. 1996), and during somatic and zygotic embryogenesis in Nicotiana (Borisjuk et al. 1998). Although, the function of CRT during gamete fusion and embryo development remains unclear, our detailed in situ studies on CRT transcriptional activity in the embryo sac during pollen tube entry and early embryogenesis suggest three likely possibilities. One explanation is that enhanced expression of PhCRT after fertilization is required for rapid cell divisions of the zygote and early endosperm, as was suggested by Chen et al. (1994) and Dresselhaus et al. (1996). Alternatively, Borisjuk et al. (1998) postulated that the increased CRT mRNA level during Nicotiana embryogenesis indicates that the chaperone function of CRT is required for proper embryo development and endosperm function. CRT, which is mainly localized in the ER and Golgi, may be required for the proper folding of proteins that are synthesized on the ER and then transported to the Golgi for sorting to their final destinations. Finally, given that precise regulation of Ca\(^{2+}\) is required for gamete fusion and activation of the fertilized egg cell (see review by Ge et al. 2007), CRT may be required to maintain Ca\(^{2+}\) homeostasis.

We report the first evidence of PhCRT mRNA localization in the nucleoli of the zygote and endosperm. In addition to its well-known function in ribosome biogenesis, the plant nucleolus is thought to function in mRNA biogenesis (see review by Shaw and Brown 2011). mRNAs from a wide range of genes were identified in a cDNA library.
from purified Arabidopsis nucleoli. Several hypotheses have been put forth to explain this phenomenon. First, aberrantly spliced mRNAs might localize to the nucleolus for degradation. This could be particularly important in extremely active cells such as those of the early embryo and endosperm. Second, specific mRNAs could have a nucleolar phase preceding their export from the nucleus. Finally, the presence of small regulatory RNAs in the nucleolus suggests that this structure may be involved in regulation of gene expression, possibly in response to cellular conditions (see review by Shaw and Brown 2011). The examined early division stage of Petunia embryogenesis represent cell stages with a gene expression program that results in high metabolic activities for cell proliferation and biosynthesis.

A micropylar-chalazal gradient of PhCRT in the receptive synergid

One of our most striking, and novel, observations was that PhCRT mRNA formed a gradient in the receptive synergid cell at the PP3 stage. The highest concentration of PhCRT mRNA was at the micropylar pole of the cell, above the site of released sperm cells. One possibility is that this expression pattern reflects a role for PhCRT in functional differentiation of sister synergids. In all flowering plants examined (except Plumbago, which lacks synergids), the highest level of Ca$^{2+}$ in the embryo sac is in the receptive synergid, where Ca$^{2+}$ is also distributed in a micropylar-chalazal gradient (see review by Ge et al. 2007). Generally, only one pollen tube enters each embryo sac in angiosperms, and precise regulation of the Ca$^{2+}$ level at the micropylar region of the penetrated ovule determines its receptivity. Recently, Iwano et al. (2012) showed in Arabidopsis that, upon pollen tube arrival at the synergid, Ca$^{2+}$ oscillation begins at the micropylar pole and spreads toward the chalazal pole. Ca$^{2+}$ in the synergid cell reaches a maximum at pollen tube rupture. In Petunia and tobacco, the synergid that accepts the pollen tube has high levels of loosely bound Ca$^{2+}$ (Tian and Russel 1997) and membrane-bound Ca$^{2+}$ (Huang and Russel 1992; Tirlapur et al. 1993), which represent exchangeable Ca$^{2+}$ associated with Ca$^{2+}$-binding proteins such as calmodulin or CRT. We postulate that the micropylar-chalazal gradient of PhCRT mRNA within the receptive synergid reflects a role of CRT in modulating the local concentration of Ca$^{2+}$ to prevent extra pollen tubes from entering the embryo sac.

Alternatively, an accumulation of PhCRT mRNA in the cytoplasm of the receptive synergid after release of the male gametes may reflect a role for Ca$^{2+}$ regulation in...
positioning of the male gametes for fusion with the female gametes. Beautiful experiments in which fluorescent phalloidin was injected into the embryo sac of *Torenia* clearly showed dramatic changes in the actin organization from anthesis to post-fertilization (Fu et al. 2000). Actin structures called coronas form after anthesis in the receptive synergid and at the interface between egg apparatus cells and vanish after fertilization. These coronas appear to be involved in reception of the pollen tube and migration and deposition of non-motile sperm cells to the fusion site with the female gametes. Studies by Ye et al. (2002) revealed that the two sperm cells deposited in the receptive synergid also have actin filaments oriented along their long axes. Thus, it is likely that transport of the sperm cells to the fusion site depends on actin-myosin interactions within the embryo sac, which in turn requires $\text{Ca}^{2+}$. Actin dynamics is a $\text{Ca}^{2+}$-dependent process, and the activities of many actin-binding proteins are regulated by $\text{Ca}^{2+}$. Therefore, maintenance of an optimal $\text{Ca}^{2+}$ environment within the embryo sac is essential for fertilization; the $\text{Ca}^{2+}$-buffering protein CRT is an excellent candidate to fulfill this role.

*PhCRT* mRNA transcripts were also detected within the two sperm cells. Although enhancement of CRT expression was previously revealed in maize sperm cells (Williams et al. 1997), the possible role of CRT in plant male gametes remains unclear, and further experiments are needed to explain this phenomenon.

*PhCRT* is highly expressed during pistil transmitting tract maturation and pollen tube outgrowth on the stigma.

This paper reports the first link between elevated CRT expression and pistil transmitting tract maturation. Both our Northern blots and FISH revealed that the highest level of *PhCRT* expression was in the unpollinated pistil 1 day before anthesis, and that the levels of mRNA were higher in stigma–style fragments of UPI pistils than in the same parts of UPM pistils. In addition, we observed higher abundance of *PhCRT* transcripts in the cytoplasm and nuclei of sitt secretory cells before anthesis. Together, these results suggest that *PhCRT* mRNA transcription starts before the exudate appears on the stigma surface. Similarly, CRT is highly abundant in other plant tissues that are secretory in nature, such as the secreting nectaries in *Arabidopsis* flowers (Nelson et al. 1997), the inner layer of the fertilized ovule (called endothelium) in *Nicotiana* (Borisjuk et al. 1998), and the active tapetum in *Nicotiana* anthers (Nardi et al. 2006). Both the tapetum and the endothelium are involved in synthesis of nutrients for the developing pollen or embryo, whereas the pistil transmitting tract cells produce nutrients for growing pollen tubes. Therefore, we postulate that the secretory sitt (and probably also sytt)
cells in *Petunia*, like the endothelium cells and the tapetum cells, require an active biosynthetic and secretory apparatus and rely on CRT as a molecular chaperone to achieve high secretory activity.

We observed a high level of *PhCRT* expression in the upper part of the pistil at the PP1 stage, when pollen germinate and tubes outgrow on the receptive stigma. FISH labeling indicated that the mRNA was mainly localized uniformly in the cytoplasm of the hydrated and germinated pollen as well as in growing pollen tubes. Previously, we showed a similar distribution of *CRT* mRNA in *Haemanthus* germinating pollen and growing tubes in vitro (Lenartowska et al. 2009). The signal was weaker and less specific than in *Petunia* however, most likely owing to the fact that we did not have a *Haemanthus*-specific probe. One explanation for the high level of *CRT* mRNA in germinated pollen and growing tubes might be that the chaperone activity of CRT facilitates the high rate of protein synthesis required for this fast tip-growing cell. Alternatively, CRT’s Ca\(^{2+}\)-buffering properties may contribute to the maintenance of cytoplasmic Ca\(^{2+}\) homeostasis, which is critical for pollen tube growth (see review by Ge et al. 2007).

An important question is whether transcription is required in a pollen tube while it is growing. It is commonly accepted that most plants accumulate mRNAs and proteins in their mature pollen until the time of germination and thus do not require transcription during elongation. For example, recent data show that most of the RNAs required for germination and pollen tube growth are stored in the mature pollen grain in *Petunia*, so de novo transcription may not be necessary (Ishimizu et al. 2010). However, in vitro studies have suggested that de novo mRNA synthesis does occur during pollen germination and tube elongation (see review by Huang et al. 2011). We observed *PhCRT* mRNA in the vegetative nucleus of germinating pollen and very short pollen tubes, but not *PhCRT* mRNA in the nuclei of hydrated pollen and longer tubes. This result is consistent with our previous in vitro studies in *Haemanthus* (Lenartowska et al. 2009) and suggests that transcription is active during pollen germination and the start of tube outgrowth, but then is silent during pollen tube elongation.

Taken together, we conclude that *PhCRT* is expressed during multiple steps of plant reproduction: pistil transmitting tract maturation, pollen germination and tube outgrowth, fertilization, and early embryogenesis. We speculate that CRT’s molecular chaperone and Ca\(^{2+}\)-buffering activities facilitate these processes, which require high rates of protein synthesis and careful regulation of Ca\(^{2+}\) homeostasis.

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Conflict of interest The authors declare that they have no conflict of interest.

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