Inclusion of Limited Amounts of Extruded Legumes Plus Cereal Mixes in Normocaloric or Obesogenic Diets for Rats: Effects on Lipid Profile

Luis A. Rubio 1,*, Isabel Aranda-Olmedo 1 and Mercedes Martín-Pedrosa 2

1 Department of Physiology and Biochemistry of Animal Nutrition, Estación Experimental del Zaidín (EEZ, CSIC), Profesor Albareda, 1, 18008 Granada, Spain; isabel.aranda@eez.csic.es
2 SGIT-INIA, Tecnología de los Alimentos, Ctra. de la Coruña Km 7, 28040 Madrid, Spain; mmartin@inia.es

* Correspondence: lrubio@eez.csic.es; Tel.: +34-958-572757; Fax: +34-958-572753

Received: 2 April 2020; Accepted: 26 May 2020; Published: 1 June 2020

Abstract: Overweight and obesity are regarded as world epidemics and are major risk factors for a number of chronic diseases, including diabetes, cardiovascular diseases, and cancer. Two new highly palatable extruded mixes based on rice and pea (Pisum sativum) or kidney bean (Phaseolus vulgaris) meals were incorporated into normocaloric or obesogenic diets for rats at a low inclusion level (25%). Our purpose was to evaluate the effects of dietary incorporation of this new food ingredient on lipid profile. Organs (heart, liver, kidneys, spleen, small intestine, colon, cecum) and visceral fat relative weights were different (p < 0.01) from controls for animals fed the obesogenic diets and in rats fed extruded diets with respect to controls. Faecal excretion of bile acids was higher (p < 0.01) for rats fed extruded mixes compared with controls. The inclusion of extruded mixes replacing part of the casein in the control diet lowered liver cholesterol and triglycerides (p < 0.001) and plasma low-density lipoprotein (LDL; p < 0.01) values, although plasma high-density lipoprotein (HDL) was unaltered. Both the inclusion of extruded mixes and the use of obesogenic diets resulted in significantly (p < 0.001) different long chain fatty acid (LCFA) profiles in liver and visceral fat. Incorporating extruded legume plus cereal mixes beneficially influenced lipid metabolism, and would therefore deserve closer attention in human intervention studies, particularly with adolescents. To our knowledge, this is the first report on the nutritional and physiological effects of extruded legume plus cereal mixes.

Keywords: extrusion; legume; lipid metabolism; obesity; rat

1. Introduction

Overweight and obesity are defined as abnormal or excessive fat accumulation that presents a risk to health. According to the WHO [1], overweight and obesity are regarded as world epidemics and are major risk factors for a number of chronic diseases, including diabetes, cardiovascular diseases, and cancer. Once considered a problem only in high-income countries, overweight and obesity are now dramatically on the rise in low- and middle-income countries, particularly in urban settings. Even more worrying is the fact that the prevalence is steadily increasing in children and adolescents, both in developed and developing countries [2]. In 2011, more than 40 million children under five years of age suffered from or were overweight [3].

Obesity is associated with metabolic derangements in multiple tissues, which contribute to the progression of insulin resistance and metabolic syndrome. This has been defined as a cluster that requires at least three of five factors: increased waist circumference, hypertriglyceridemia, low high-density lipoprotein (HDL) cholesterol, hypertension, and a fasting glucose of 110 mg/dL or
higher [4]. Obesity is also associated with the Western diet [5]. Eating patterns in Western industrialised countries are characterised by high energy consumption and chronic overconsumption of saturated fat, cholesterol, sugar, and salt, which is also related to the development of other pathologies such as diabetes, cardiovascular and degenerative disorders, and cancer [6]. In this context, it has been proposed that weight loss could be achieved with controlled energy diets and a high content of cereals and legumes [7]. Legumes have long been an important component of the human diet because of their content in protein, carbohydrates (mainly in the form of starch), and many other nutrients such as proteins, and fibre [8], but in addition, the consumption of legumes can have therapeutic effects, being an important element of the Mediterranean diet pattern [9]. Additionally, legume carbohydrates, known as “slow-release carbohydrates”, when added to the diet are an effective tool in the treatment of obesity, diabetes, and hyperlipidemia [10]. The substitution of energy-rich foods for pulses has been reported to have beneficial effects on the prevention and treatment of obesity and related disorders such as cardiovascular diseases, type 2 diabetes, and metabolic syndrome [10–12].

Recommendations to increase the fibre intake in the child/adolescent population emphasise the greater consumption of fruits, vegetables, pulses, and whole grains. Given the difficulties of changing the dietary habits of the population, one way to optimise fibre intake would be through the consumption of other types of fibre-supplemented foods [13]. Alternative foods with high palatability would be particularly useful in this context. Accordingly, the present work aimed to add some more information to the study of the effects that the consumption of legumes and cereals may have on health through the evaluation of new products derived from these foodstuffs that have been adapted to the present consumer demands. These demands in Western societies no longer respond so much to the need to satisfy hunger or the need of energy, but to the “hunger of health” or absence of disease.

In this context, new highly palatable [14] extruded mixes of legumes plus cereals similar to a snack food have been produced [15]. Two of these extruded mixes have been incorporated in the current work into normocaloric or obesogenic diets for rats to evaluate the effects on lipid profile and intestinal microbiota composition [16]. Obesogenic diets are those that promote obesity by presenting high calorific contents mainly from a high proportion of carbohydrates, usually simple sugars and/or fats [17]. As the intake of this new snack type food in practical conditions would be necessarily limited, we included it in the diet at a low inclusion level (25%) to provide no more than 20% of the total dietary protein. Our purpose was to evaluate the effects of dietary incorporation of this new food ingredient on lipid profile.

2. Materials and Methods

All management and experimental procedures carried out in this rat trial were in strict accordance with current European regulations (86/609 E.E.C.) regarding laboratory animals. The Bioethics Committee for Animal Experimentation at our institution (EEZ-CSIC) approved the study protocol.

2.1. Preparation of the Extruded Mixes and Diets

Two extruded legumes plus cereal mixes were produced by using a Clextral Evolum25 twin-screw extruder (Clextral, Riez 42702 Firminy Cedex, France). These were rice (Oryza sativa) + pea (Pisum sativum, cv Cartouche), or kidney bean (Phaseolus vulgaris, cv Almonga) + carob tree (Ceratonia siliqua) fruits. Extruded mixes (PEM and KEM) composition was rice meal/pea or kidney bean meal/carob fruits meal, respectively (50/40/10, w:w:w) and contained no casein. All procedures were carried out as described previously [15]. The chemical composition of casein, and extruded mixes are shown in Table 1.

The diets (Table 2) were based on casein (CAS) or casein plus pea or kidney bean extruded mixes (PEM and KEM) and were formulated to contain the same amount of digestible energy and protein. In order not to be too far from practical conditions in human nutrition, PEM and KEM addition was limited to a low inclusion level (25%) to provide no more than 20% (two servings) of the total recommended daily protein intake. Another relevant point on formulation in the present work was that diets contained no added cholesterol. Appropriate amounts of synthetic amino acids were added
to the extruded mix-based diets, taking into account their amino acid composition (Table 1) to equalise them to control (casein) values. The diets were supplemented with vitamins and minerals to target requirements [18]. Obesogenic diets (CAS-OB, PEM-OB, and KEM-OB) were the same diets described above (CAS, PEM, and KEM) with added amounts (5.48 g/day) of IDEAL commercial condensed milk (composition in Table 2).

Table 1. Analysed composition (g/kg) of casein and the legume + cereal extruded mixes.

|                          | Casein  | Extruded Pea Mix 1 | Extruded Kidney Bean Mix |
|--------------------------|---------|--------------------|--------------------------|
| Dry matter               | 920.0   | 944.4              | 947.6                    |
| Protein                  | 813.3   | 128.2              | 132.3                    |
| Ether extract            | ND 2    | 1.2                | 1.4                      |
| Ash                      | ND      | 34.6               | 38.4                     |
| Crude fibre              | ND      | 32.1               | 24.6                     |
| Dietary fibre            | ND      | 94.1               | 114.5                    |
| α-galactosides           | ND      | 32.8               | 25.8                     |
| Total carbohydrates 3    | ND      | 653.5              | 635.2                    |
| Crude energy (Kcal/g)    | 3.940   | 3.600              | 3.620                    |

Amino acids

|                | Casein | Extruded Pea Mix 1 | Extruded Kidney Bean Mix |
|----------------|--------|--------------------|--------------------------|
| Aspartate      | 58.9   | 15.1               | 15.8                     |
| Glutamate      | 184.1  | 25.4               | 26.2                     |
| Serine         | 49.1   | 6.0                | 7.3                      |
| Histidine      | 16.5   | 3.3                | 4.1                      |
| Glycine        | 27.5   | 6.1                | 6.1                      |
| Threonine      | 39.6   | 4.4                | 5.0                      |
| Arginine       | 37.4   | 12.5               | 11.4                     |
| Alanine        | 32.6   | 6.8                | 7.0                      |
| Tyrosine       | 49.0   | 5.5                | 5.7                      |
| Cystine        | 3.4    | 1.5                | 1.4                      |
| Valine         | 53.3   | 7.0                | 7.5                      |
| Methionine     | 24.5   | 1.8                | 2.5                      |
| Pheny lalanine | 46.9   | 7.2                | 8.1                      |
| Isoleucine     | 40.9   | 5.9                | 6.3                      |
| Leucine        | 74.9   | 10.9               | 11.9                     |
| Lysine         | 97.9   | 7.2                | 7.5                      |
| Proline        | 82.8   | 6.6                | 6.1                      |

1 Pea (*Pisum sativum*) extruded mix (rice/pea/carob tree bean, 50/40/10); kidney bean (*Phaseolus vulgaris*) extruded mix (rice/kidney bean/carob tree bean, 50/40/10). 2 ND, not determined. 3 Total solids excluding protein, fat, ash, dietary fibre, and α-galactosides.

2.2. Biological Assays

A total of 72 male weaned Wistar rats (Charles River Laboratories), matched by weight (80.0 ± 2.7 g, mean ± SE) and with 4 weeks of age, were individually housed in metabolism cages throughout the experiment. Rats were fed a control casein diet between weaning and the start of the experiment. Animals were then randomly distributed into six groups (n = 12), and each group was assigned to one of the dietary treatments (see above). The animals were individually housed in metabolic cages in an environmentally controlled room under standard conditions (temperature: 20–22 °C with a 12 h light–dark cycle and 55–70% humidity). The rats had free access to their diets and tap water, and were fed the different diets for 21 days. Feed offered was calculated to be 95% of ad libitum intake for this age and species, and was progressively increased according to rats’ age. Rats ate all feed offered and there were no leftovers. Faeces were collected daily and stored separately as a 1-week pool for each animal. The faeces were weighed, lyophilised, powdered, and then homogenised. On day 21, after an overnight fast, they were refed at defined time intervals with 11 g of diet and euthanized 60 min after refeeding so that all rats were in the same feeding situation. Animals were anaesthetised with sodium pentobarbital (5 mg/100 g of body weight) (Abbott Laboratories, Granada, Spain) and terminal
Exsanguination was performed by cannulation of the carotid artery. Organs and visceral fat (defined as the sum of the mesenteric, epididymal, and perirenal fat depots) were removed and weighed, and intestinal contents were immediately frozen at −20 °C and then freeze-dried until analysis.

Table 2. Composition (g/kg) of the diets.

|                      | CAS 1 | PEM  | KEM  |
|----------------------|-------|------|------|
| Extruded mix         | -     | 250  | 250  |
| Casein               | 209   | 160  | 160  |
| Maize starch         | 261.28| 209.58| 209.58|
| Potato starch        | 150   | -    | -    |
| Cellulose            | 50    | 50   | 50   |
| Sunflower oil        | 70    | 70   | 70   |
| Sucrose              | 150   | 150  | 150  |
| DL-Methionine        | 3     | 3    | 3    |
| Tryptophane          | -     | 0.70 | 0.70 |
| Cystine-HCl          | 1.80  | 1.80 | 1.80 |
| Vitamins + minerals  | 45    | 45   | 45   |
| Ca diposphate        | 55    | 55   | 55   |
| Citric acid          | 0.12  | 0.12 | 0.12 |
| Iron sulfate heptahydrate | 0.20 | 0.20 | 0.20 |
| Cr2O3                | 3     | 3    | 3    |

|                      |       |      |      |
|----------------------|-------|------|------|
| Calculated composition |      |      |      |
| Energy (kcal/g)      | 3.23  | 3.22 | 3.21 |
| Protein              | 170   | 170  | 170  |
| Fat                  | 70    | 70   | 70   |
| Carbohydrates        | 486.3 | 531.3| 525.8|
| Dietary fibre        | 50.0  | 73.5 | 78.6 |

1 CAS, casein diet; KEM, kidney bean (Phaseolus vulgaris) extruded mix diet; PEM, pea (Pisum sativum) extruded mix diet.

2.3. Chemical and Biochemical Analysis

All chemicals were obtained from Sigma-Aldrich Química S.L. (Madrid, Spain), unless otherwise stated. Proximate analysis (Table 1) of casein and extruded mixes was carried out following AOAC (Association of Official Agricultural Chemists) procedures as in Arribas et al. [15], and amino acids were determined by the Pico Tag method in an HPLC analyser (Waters, Milford, MA, USA) as in Rubio et al. [19]. α-Galactosides were determined by HPLC equipped with a refractive index detector according to Pedrosa et al. [20].

Plasma cholesterol and triglycerides analysis was performed in a BS-200 Biochemical Analyzer (Shenzhen Mindray Bio-medical Electronics Co., Ltd., Mindray Medical España S.L., Madrid, Spain) with chemicals from Spinreact (Spinreact, S.A.U., Gerona, Spain). Liver cholesterol and triglycerides were determined by using colorimetric/fluorometric methods from Abcam kits (ab65359 and ab65336 for cholesterol and triglycerides, respectively).

Long chain fatty acids (LCFA) in liver and visceral fat were determined as in Palmquist and Jenkins [21]. Briefly, samples (0.3 g) were taken in test tubes with Teflon screw caps and kept at 4 °C. Internal standard (C23:0, Larodan’s methyl tricosanoate ref. 20-2300; 4 mg in 2.0 mL n-heptane) were added, followed by 1.8 mL of 10% methanolic HCl. The tubes were tightly capped, vortexed, heated in a water bath at 90 °C for 2 h, cooled, and then 0.6 mL of heptane and 6 mL of 6% K2CO3 were added. Tubes were then vortexed and centrifuged at 4 °C for 5 min at 500×g. The top layer (organic solvent) was transferred to a tube with a screw Teflon cap containing 0.6 g of sodium sulphate. After vortexing again and centrifuging at 4 °C for 5 min at 500×g, the solvent layer was transferred to 2 mL GLC (gas liquid chromatography) sample vials, which were analysed by GLC.
Bile acids in faeces were determined as in Hagio et al. [22]. Briefly, faeces were freeze-dried, ground, extracted in ethanol, sonicated, heated twice in a water bath, and centrifuged. This procedure was repeated twice, and the pooled extracts were evaporated and dissolved in methanol. The methanol extracts were purified and bile acids were analysed by liquid chromatography (LC) using an Acquity UPLC system (Waters) with a gradient elution from a BEH C18 column (1.7 mm, 100 mm × 2.0 mm ID; Waters).

2.4. Statistical Analysis

Statistical significance of data was tested by two-way (+/- extruded mix addition, +/- obesogenic diet) analysis of variance (ANOVA) with Tukey’s post-hoc test with statistical significance set at \( p < 0.05 \). Evaluation of the relationship between the different variables of interest was carried out by computing the relevant correlation coefficient (Pearson’s linear correlation). For qPCR microbial counts, discriminant analysis (DA) was used to check if the groups to which bile acids or LCFA observations belonged to were distinct, and principal component analysis (PCA) was used to study the relationships between bacterial groups [23].

3. Results

3.1. Growth and Feed Intake

Food consumption (Table 3) and final body weight (Figure 1) was the same among non-obesogenic groups and among obesogenic groups, and different \( (p < 0.01) \) between non-obesogenic and obesogenic groups.

![Figure 1](image_url)

**Figure 1.** Final body weight of rats fed casein or casein + extruded mix diets either obesogenic or not. CAS, casein diet; KEM, kidney bean \((Phaseolus vulgaris)\) extruded mix diet; PEM, pea \((Pisum sativum)\) extruded mix diet. Diets CAS-OB, KEM-OB, and PEM-OB were the same diets described above with added 5.48 g/day condensed milk. Values are means \((n = 12)\) with their standard error of the mean (SEM) in bars. \(^{ab}\) Means with different superscript differ \((p < 0.01)\).
Table 3. Total daily feed intake (g/day) of rats fed casein or casein plus extruded mix diets either obesogenic or not for 21 days.

|                  | CAS       | CAS-OB    | PEM/KEM   | PEM-OB/KEM-OB |
|------------------|-----------|-----------|-----------|---------------|
| Intake (g)       | 15.71     | 21.48     | 15.71     | 21.48         |
| Energy (kcal)    | 48.30     | 66.15     | 48.00     | 65.52         |
| Protein (g)      | 2.54      | 2.95      | 2.54      | 2.95          |
| Fat (g)          | 1.05      | 1.51      | 1.05      | 1.51          |
| Carbohydrates (g)| 7.27      | 10.34     | 7.86/7.94 | 11.16/10.97   |

3.2. Organs and Visceral Fat Relative Weights

Carcass, organs (heart, liver, kidneys, spleen, stomach, small intestine, cecum, and colon), and visceral fat relative weights (g/100 g fresh weight) of rats fed the different diets are recorded in Table 4. Carcass weight was higher ($p < 0.01$) than controls for rats fed extruded diets, and liver weight was higher ($p < 0.01$) for rats fed obesogenic diets compared with non-obesogenic diets. Visceral fat relative weights were higher ($p < 0.01$) for the animals fed the obesogenic diets in comparison with non-obesogenic diets, and in rats fed extruded diets respect to the control (CAS or CAS-OB) diets. A significant interaction ($p = 0.034$) was found between extrusion and obesogenic effects for visceral fat. Rats fed extruded mixes had lower ($p < 0.01$) small intestine, colon, and cecum relative weights compared with controls.

Table 4. Organs relative weight (g/100 g fresh body weight) of rats fed casein or casein plus extruded mix diets either obesogenic or not.

|                  | Carcass | Heart | Liver | Kidneys | Visceral Fat | Spleen | Stomach | Small Intestine | Colon | Cecum |
|------------------|---------|-------|-------|---------|--------------|--------|---------|-----------------|-------|-------|
| CAS              | 76.474  | 0.310 | 4.022 | 0.950   | 1.398        | 0.350  | 0.596   | 3.775           | 0.483 | 1.186 |
| CAS-OB           | 77.685  | 0.316 | 4.226 | 0.891   | 2.127        | 0.350  | 0.560   | 3.544           | 0.405 | 1.175 |
| PEM              | 79.701  | 0.304 | 4.018 | 0.949   | 1.606        | 0.376  | 0.605   | 3.322           | 0.406 | 0.490 |
| PEM-OB           | 79.535  | 0.306 | 4.413 | 0.964   | 2.922        | 0.382  | 0.556   | 3.272           | 0.411 | 0.455 |
| KEM              | 79.884  | 0.318 | 3.982 | 0.925   | 1.601        | 0.386  | 0.600   | 3.210           | 0.374 | 0.490 |
| KEM-OB           | 78.924  | 0.328 | 4.193 | 0.943   | 2.615        | 0.409  | 0.620   | 3.268           | 0.386 | 0.492 |
| Pooled SD        | 1.744   | 0.024 | 0.377 | 0.071   | 0.500        | 0.065  | 0.073   | 0.278           | 0.070 | 0.214 |

$E$, effect of inclusion of extruded legume/rice/carob tree bean; $O$, effect of the obesogenic diet; $E \times O$, interaction of both effects.

3.3. Bile Acids in Feces

Fecal excretion of cholic, chenodeoxycholic and ursodeoxycholic acids was lower ($p < 0.05$) and deoxycholic acid and total bile acids higher ($p < 0.01$) for rats fed extruded mixes compared with CAS and CAS-OB controls (Table 5). Obesogenic diets did not induce any significant change in bile acids fecal excretion. However, there was a significant ($p < 0.05$) interaction between extrusion and obesogenic effects for total bile acids. As shown in Figure 2, values for rats fed diets based in casein (CAS and CAS-OB) were different ($p < 0.05$) from those fed diets based in extruded mixes.
Table 5. Bile acids concentrations (µmol/g) in the feces of rats fed casein or casein plus extruded mix diets either obesogenic or not.

| Diet      | CAS  | CAS-OB | KEM  | KEM-OB | PEM  | PEM-OB | Pooled SD | E   | O   | E × O |
|-----------|------|--------|------|--------|------|--------|-----------|------|------|-------|
| Cholic acid | 0.088 | 0.134  | 0.072 | 0.082  | 0.093| 0.055  | 0.058     | 0.021| 0.414| 0.043 |
| Deoxycholic acid | 0.112 | 0.124  | 0.784 | 0.805  | 0.946| 0.614  | 0.208     | <0.001| 0.168| 0.114 |
| Chenodeoxycholic acid | 0.005 | 0.007  | 0.003 | 0.003  | 0.003| 0.002  | 0.004     | 0.002| 0.391| 0.177 |
| Ursodeoxycholic acid | 0.004 | 0.005  | 0.001 | 0.003  | 0.003| 0.003  | 0.002     | <0.001| 0.279| 0.800 |
| Total     | 0.206 | 0.245  | 0.892 | 0.837  | 1.062| 0.673  | 0.244     | <0.001| 0.129| 0.041 |

Values are means (n = 12). 1 CAS, casein diet; KEM, kidney bean (Phaseolus vulgaris) extruded mix diet; PEM, pea (Pisum sativum) extruded mix diet. Diets CAS-OB, KEM-OB and PEM-OB were the same diets described above with added 5.48 g/d condensed milk. 2 E, effect of inclusion of extruded legume/rice/carob tree bean; O, effect of the obesogenic diet; E × O, interaction of both effects.

Figure 2. Discriminant analysis of the expression levels of bile acid concentration (µM/g) in the faeces of rats fed CAS (yellow); CAS-OB (grey); KEM (orange); KEM-OB (dark blue); PEM (light blue); PEM-OB (green). For details on the diets, see the Materials and Methods section and Table 2. Values for rats fed diets based on casein (CAS and CAS-OB) were different (p < 0.05) from those fed diets based on extruded mixes, although not different to each other.

3.4. Liver and Plasma Cholesterol and Triglycerides

Results on liver and plasma cholesterol and triglycerides are reported in Table 6. The inclusion of the extruded mix replacing part of the casein in the control diet (CAS) lowered liver cholesterol and triglycerides (p < 0.001) and plasma low-density lipoprotein (LDL; p < 0.01) values, although plasma HDL was unaltered. The obesogenic diets induced lower (p < 0.017) liver cholesterol values with respect to non-obesogenic diets only in the diet with the extruded legumes plus cereal mix. The obesogenic diets increased (p < 0.05) plasma triglyceride values with respect to the non-obesogenic diets.
Table 6. Cholesterol and triglyceride concentrations in liver (mg/g fresh weight) and plasma (mg/dL) of rats fed casein or casein plus extruded mix diets either obesogenic or not.

|                | Cholesterol | Triglycerides |
|----------------|-------------|---------------|
|                | Liver       | Plasma        |
|                | LDL         | HDL           |
|                | Liver       | Plasma        |
| CAS            | 2.016       | 16.77         | 32.13         | 6.573         | 125.08 |
| CAS-OB         | 1.911       | 15.20         | 31.50         | 7.880         | 203.46 |
| PEM            | 1.771       | 11.48         | 29.53         | 5.364         | 144.19 |
| PEM-OB         | 1.465       | 9.17          | 36.41         | 5.449         | 207.42 |
| KEM            | 1.684       | 9.46          | 38.84         | 5.027         | 78.13  |
| KEM-OB         | 1.475       | 12.05         | 37.02         | 6.231         | 146.71 |
| Pooled SD      | 0.297       | 6.44          | 9.65          | 1.853         | 56.86  |

|                | p-Values    |
|----------------|-------------|
| CAS            | 0.001       | 0.01         | 0.191        | 0.001         | 0.240  |
| PEM            | 0.017       | 0.776        | 0.730        | 0.050         | <0.001 |
| Pooled SD      | 0.308       | 0.626        | 0.566        | 0.382         | 0.811  |

Values are means (n = 12). 1 CAS, casein diet; KEM, kidney bean (*Phaseolus vulgaris*) extruded mix diet; PEM, pea (*Pisum sativum*) extruded mix diet. Diets CAS-OB, KEM-OB, and PEM-OB were the same diets described above with added 5.48 g/day condensed milk. 2 E, effect of inclusion of extruded legume/rice/carob tree bean; O, effect of the obesogenic diet; E × O, interaction of both effects.

3.5. Long Chain Fatty Acid Composition of Liver and Visceral Fat

As shown in Table 7, the livers of rats fed diets containing extruded mixes (KEM or PEM) had higher proportions of C10, C15:1, C16, C17:1, C18, C18:1 11c, C18:2, C20, and C20:3, and lower proportions of C20:4 and C22:6 than CAS controls. Obesogenic diets increased the proportions of C16:1, C18, C18:1 9c, and C18:1 11c, and lowered those of C18:2 n6 9c12c, C20, C18:3 n6, C20:2 n6, C24, and C24:1. The inclusion of the extruded mixes induced an increase in the proportions of monounsaturated fatty acid (MUFA) in the liver, while the obesogenic diets caused increases of both MUFA and polyunsaturated fatty acid (PUFA) in the liver. No significant interactions between extruded mix and obesogenic diets were found in LCFA liver composition.

The visceral fat (Table 8) of rats fed the extruded mixes had higher proportions of C16, C16:1, and C18:1 9c, and lower proportions of C18:2 n6. The obesogenic diets induced increases of C10, C12, C14, C16, C16:1, C17, C18:1 9c, and C18:1 11c, and decreases of C18:2 n6, C20:2 n6, and C20:4. Extruded mixes and obesogenic diets induced an increase in the proportions of SFA and MUFA, and a decrease of PUFA proportions in the visceral fat.
Table 7. Effect of the diet on liver long chain fatty acid (LCFA) composition of rats fed control diets (CAS) or diets supplemented with two extruded mixes of legumes plus cereals, normocaloric or obesogenic.

| Diet                  | CAS    | CAS-OB  | PEM    | PEM-OB  | KEM    | KEM-OB  | Pooled SD | E       | O       | E × O     |
|-----------------------|--------|---------|--------|---------|--------|---------|-----------|---------|---------|-----------|
| C10, capric acid      | 0.430  | 0.410   | 0.568  | 0.448   | 0.638  | 0.596   | 0.222     | 0.021   | 0.405   | 0.614     |
| C14, myristic acid    | 3.210  | 2.228   | 3.052  | 2.816   | 2.752  | 2.012   | 0.209     | 0.929   | 0.218   | 0.639     |
| C15:1, cis-10-pentadecenoic acid | 1.820 | 2.906   | 4.889  | 3.672   | 4.164  | 5.238   | 1.873     | <0.001 | 0.289   | 0.182     |
| C16, palmitic acid    | 17.194 | 19.199  | 23.474 | 21.570  | 21.395 | 23.237  | 3.586     | <0.001 | 0.147   | 0.091     |
| C16:1, palmitoleic acid | 1.282 | 2.863   | 2.216  | 2.776   | 2.093  | 3.014   | 1.053     | 0.096   | <0.001 | 0.121     |
| C17, heptadecanoic acid | 0.234 | 0.216   | 0.260  | 0.240   | 0.212  | 0.262   | 0.050     | 0.179   | 0.882   | 0.270     |
| C17:1, cis-10-heptadecenoic acid | 0.103 | 0.113   | 0.169  | 0.173   | 0.198  | 0.124   | 0.108     | 0.050   | 0.748   | 0.514     |
| C18, stearic acid     | 11.132 | 10.027  | 16.108 | 13.345  | 15.229 | 14.576  | 2.989     | <0.001 | 0.024   | 0.599     |
| C18:1 9t, elaidic acid | 0.203  | 0.196   | 0.214  | 0.211   | 0.188  | 0.289   | 0.068     | 0.171   | 0.287   | 0.143     |
| C18:1 9c, oleic acid  | 11.702 | 15.513  | 12.890 | 13.486  | 12.866 | 15.230  | 3.164     | 0.991   | 0.003   | 0.167     |
| C18:1 11c, ?          | 3.026  | 3.752   | 3.678  | 3.638   | 4.571  | 0.899   | 0.050     | 0.025   | 0.534   |           |
| C18:2 n6 9c12c, linoleic acid | 13.683 | 11.581  | 16.293 | 11.952  | 15.385 | 12.727  | 2.697     | 0.015   | <0.001  | 0.224     |
| C20, arachidonic acid | 0.048  | 0.061   | 0.043  | 0.055   | 0.046  | 0.041   | 0.012     | 0.025   | <0.001  | 0.291     |
| C20:1 n9, cis-11-eicosenoic acid | 0.193 | 0.179   | 0.196  | 0.121   | 0.159  | 0.160   | 0.060     | 0.098   | 0.113   | 0.453     |
| C20:2 n6, cis-11,14-eicosadienoic acid | 0.446 | 0.284   | 0.471  | 0.293   | 0.455  | 0.313   | 0.123     | 0.505   | <0.001  | 0.985     |
| C22, behenic acid     | 0.032  | 0.032   | 0.030  | 0.030   | 0.035  | 0.039   | 0.013     | 0.391   | 0.747   | 0.796     |
| C20:3 n6, cis-8,11,14-eicosatrienoic acid | 0.375 | 0.311   | 0.554  | 0.427   | 0.4865 | 0.395   | 0.167     | 0.007   | 0.053   | 0.613     |
| C20:4, arachidonic acid | 4.907  | 2.610   | 0.011  | 0.022   | 0.003  | 0.093   | 3.589     | <0.001  | 0.207   | 0.189     |
| C24, lignoceric acid  | 0.044  | 0.040   | 0.046  | 0.034   | 0.044  | 0.062   | 0.047     | 0.252   | 0.041   | 0.169     |
| C24:1 n9, nervonic acid | 0.043  | 0.035   | 0.035  | 0.028   | 0.045  | 0.033   | 0.020     | 0.540   | 0.001   | 0.280     |
| C22:6 n3, cis-4,7,10,13,16,19-docosahexaenoic acid | 0.401  | 0.554   | 0.198  | 0.177   | 0.295  | 0.357   | 0.618     | <0.001  | 0.185   | 0.223     |

Values are means (n = 12). ¹ CAS, casein diet; KEM, kidney bean (Phaseolus vulgaris) extruded mix diet; PEM, pea (Pisum sativum) extruded mix diet. Diets CAS-OB, KEM-OB, and PEM-OB were the same diets described above with added 5.48 g/day condensed milk. ² E, effect of inclusion of extruded mix; O, effect of the obesogenic diet; E × O, interaction of both effects. ³ SFA, saturated fatty acids; MUFA, monounsaturated fatty acids; PUFA, polyunsaturated fatty acids.
Table 8. Effect of the diet on visceral fat long chain fatty acid (LCFA) composition of rats fed control diets (CAS) or diets supplemented with two extruded mixes of legumes plus cereals, normocaloric or obesogenic.

| Diet 1 | CAS   | CAS-OB | PEM   | PEM-OB | KEM   | KEM-OB | Pooled SD | E     | O     | E × O |
|--------|-------|--------|-------|--------|-------|--------|-----------|-------|-------|-------|
| C10, capric acid | 0.071 | 0.200  | 0.051 | 0.195  | 0.082 | 0.179  | 0.073     | 0.410 | <0.001| 0.679 |
| C12, lauric acid | 0.217 | 0.556  | 0.209 | 0.478  | 0.275 | 0.478  | 0.189     | 0.469 | <0.001| 0.167 |
| C14, myristic acid | 1.574 | 2.382  | 1.442 | 2.271  | 1.878 | 2.155  | 0.877     | 0.848 | 0.005 | 0.575 |
| C14:1, myristoleic acid | 0.173 | 0.203  | 0.134 | 0.230  | 0.164 | 0.179  | 0.100     | 0.676 | 0.162 | 0.700 |
| C15, pentadecanoic acid | 0.046 | 0.146  | 0.182 | 0.175  | 0.136 | 0.108  | 0.146     | 0.195 | 0.344 | 0.127 |
| C16, palmitic acid | 20.513| 25.447 | 23.092| 25.948 | 23.163| 26.239 | 2.844     | 0.005 | <0.001| 0.090 |
| C16:1, palmitoleic acid | 4.812 | 6.281  | 5.164 | 7.011  | 5.541 | 6.305  | 1.105     | 0.074 | <0.001| 0.683 |
| C17, elaidic acid | 0.108 | 0.168  | 0.101 | 0.157  | 0.101 | 0.148  | 0.045     | 0.255 | <0.001| 0.654 |
| C17:1, cis-10-heptadecenoic acid | 0.034 | 0.012  | 0.015 | 0.012  | 0.009 | 0.013  | 0.018     | 0.068 | 0.077 | 0.068 |
| C18, stearic acid | 3.341 | 3.109  | 3.528 | 3.400  | 3.181 | 3.316  | 0.509     | 0.401 | 0.449 | 0.453 |
| C18:1 9c, oleic acid | 31.051| 31.865 | 32.531| 32.163 | 31.158| 32.765 | 1.348     | 0.050 | 0.045 | 0.819 |
| C18:1 11c, ? | 1.737 | 2.651  | 1.952 | 2.528  | 1.778 | 2.277  | 0.687     | 0.696 | <0.001| 0.255 |
| C18:2 n6, linoleic acid | 33.343| 23.807 | 28.625| 22.592 | 29.967| 23.246 | 4.962     | 0.006 | <0.001| 0.066 |
| C18:3 n6, γ-linolenic acid. | 0.115 | 0.082  | 0.173 | 0.126  | 0.081 | 0.095  | 0.073     | 0.461 | 0.307 | 0.786 |
| C20:1, cis-11-eicosenoic acid | 0.193 | 0.162  | 0.278 | 0.163  | 0.175 | 0.248  | 0.118     | 0.272 | 0.481 | 0.864 |
| C20:3 n3, linolenic acid | 0.417 | 0.344  | 0.361 | 0.341  | 0.534 | 0.469  | 0.233     | 0.455 | 0.403 | 0.778 |
| C20:2 n6, cis-11,14-eicosadienoic acid | 0.213 | 0.135  | 0.258 | 0.162  | 0.163 | 0.194  | 0.092     | 0.459 | 0.049 | 0.301 |
| C20:4, arachidonic acid | 0.671 | 0.423  | 0.565 | 0.411  | 0.375 | 0.212  | 0.290     | 0.082 | 0.027 | 0.695 |

SFA 3 | 26.104 | 31.269 | 28.896 | 32.876 | 32.025 | 32.838 | 3.414 | 0.002 | <0.001| 0.356 |
MUF A | 37.991 | 41.281 | 40.091 | 42.185 | 38.876 | 41.823 | 1.974 | 0.004 | <0.001| 0.289 |
P UFA | 34.602 | 24.588 | 29.634 | 25.374 | 30.939 | 24.104 | 5.197 | 0.006 | <0.001| 0.057 |

Values are means (n = 12). 1 CAS, casein diet; KEM, kidney bean (*Phaseolus vulgaris*) extruded mix diet; PEM, pea (*Pisum sativum*) extruded mix diet. Diets CAS-OB, KEM-OB, and PEM-OB were the same diets described above with added 5.48 g/day condensed milk. 2 E, effect of inclusion of extruded legume/rice/carob tree bean; O, effect of the obesogenic diet; E × O, interaction of both effects. 3 SFA, saturated fatty acids; MUF A, monounsaturated fatty acids; PUFA, polyunsaturated fatty acids.
Discriminant analysis (Figures 3 and 4) of the data showed that both the inclusion of extruded mixes and the use of obesogenic diets resulted in significantly ($p < 0.001$) different LCFA profiles in the liver and in the visceral fat.

![Figure 3](image1.png)

**Figure 3.** Discriminant analysis of liver fatty acid composition of rats fed CAS (yellow); CAS-OB (grey); KEM (orange); KEM-OB (dark blue); PEM (light blue); PEM-OB (green). For details on the diets, see the Materials and Methods section and Table 2. All groups were different ($p < 0.001$) from each other.

![Figure 4](image2.png)

**Figure 4.** Discriminant analysis of visceral fat fatty acid composition of rats fed CAS (yellow); CAS-OB (grey); KEM (orange); KEM-OB (dark blue); PEM (green); PEM-OB (light blue). For details on the diets, see the Materials and Methods section and Table 2. Groups CAS-OB, PEM-OB, and KEM-OB were different ($p < 0.01$) from CAS, PEM, and KEM, but not different to each other.

4. **Discussion**

According to the United States Department of Agriculture (USDA) food pattern recommendations, for a 2000 kcal diet, one serving (40 g) of extruded blends similar to those used here would provide (on average) 11% of daily protein intake, 0.1% of daily fat intake, and 25% of daily carbohydrate intake [15]. The inclusion of either of the extruded mixes of legumes plus cereals described above in casein-based diets for rats replacing no more than 20% of the protein had significant effects on lipid profile. As a preliminary indication, we must point out here that most of the discussion below will lie
on the legume portion of the extruded mix for two main reasons: (i) legumes are usually mixed with starch of different sources, mainly corn or rice starch, in order to obtain puffed products, and (ii) very scant information is found in the literature on the nutritional effects of extruded rice in vivo.

Health professionals and agencies such as the USDA and Health Canada are promoting dietary changes including increased consumption of grain legumes (or pulses) such as peas, kidney beans, lentils, and chickpeas (www.pulsecanada.com). In the current work, pea and kidney bean seeds were used to produce the extruded composites, which were incorporated into non-obesogenic diets (PEM and KEM) formulated to be equivalent in energy and protein to a control diet (CAS). Condensed milk was added to the daily food intake to produce the obesogenic diets (CAS-OB, PEM-OB, KEM-OB) (Table 2). This ingredient has been previously utilised in well-characterised rat models of hyperphagia-induced obesity [24,25]. As expected, rats fed the obesogenic diets showed substantially higher intakes of food (36.73%), energy (36.50–36.95%), protein (16.40%), fat (51%), and carbohydrates (41.98–42.23%) (Table 3). This resulted in higher final body weights (Figure 1) and visceral fat relative weights (Table 4) with respect to non-obesogenic controls in rats fed the obesogenic diets, which is similar to previous results with this type of diets [25]. However, the incorporation of the extruded mixes at the relatively low levels used here (250 g/kg) had a significant effect on liver fat composition (Table 6) and on the amount and composition of visceral fat (Tables 4 and 7). This is not in agreement with previous work, where lower fat deposition has usually been reported in rats fed obesogenic diets based on legume (particularly soy) protein [26], albeit with all or most of the control protein replaced by legume protein in most cases. As explained above, our aim in the current work was to evaluate the effects of incorporating the extruded mix in the diet to replace only a limited amount of the protein in the control diet (closer to practical consumption). The literature on the effects of limited amounts of legume seed meals, either heat treated or not, on growth or physiological parameters in rats is quite scarce. At this level of inclusion, legumes in general, and kidney beans or peas in particular, have not been reported to affect growth rate or lipid deposition, particularly after heat treatment and supplementation with limiting amino acids [27,28].

Legume-based diets have been associated with changes in organs relative weights in rats [29]. In the current study, rats fed extruded mixes had lower small intestine, colon, and cecum relative weights compared with controls (Table 4). This is contrary to previous works, where higher relative weights of the intestinal sections have usually been reported and ascribed to the presence of higher amounts of indigestible material (particularly dietary fibre) and/or an increased production of short chain fatty acids (SCFA), which have a stimulating effect on gut epithelial proliferation [30,31]. This discrepancy is likely due to the lower dietary inclusion levels in the current study, which resulted in smaller differences in dietary fibre (Table 2) as compared to other studies. However, in a previous work, we found lower cecum, rectum, and large intestine relative weights, as well as a tendency for small intestine relative weight, in rats fed diets based on isolated lupin (Lupinus albus) protein [32]. It has previously been reported that soy protein reduce colon weight in mice in comparison to casein, an effect ascribed to reduced cell proliferation [33]. Since legumes form an important part of the “prudent” diet that may be associated with reduced colon cancer risk in humans [34], the effect of legume protein consumption on the reduction of large intestinal weight and the relationship between intestinal weight changes and intestinal cell proliferation/carcinogenesis is worthy of further investigation.

Liver and visceral fat relative weights were as expected higher for the animals fed the obesogenic diets in comparison with non-obesogenic diets. However, carcass and visceral fat weight were also higher than controls in rats fed the extruded diets (Table 4). This was unexpected since there are no indications on fat retention in animals fed diets containing extruded legumes. Three explanations may be adduced for higher energy absorption and thus higher lipid accumulation in extruded supplemented diets. Firstly, extruded mix-supplemented diets used herein contained between 47–57% higher amounts of fibre than controls (Table 2). According to McBurney and Sauer [35], hind gut fermentation of fibre would provide as much as 13 kJ/g fermentable dietary fibre coming from SCFA absorption, and 40–57 g non-starch polysaccharides (NSP)/100 g ingested was fermented in rats fed faba bean- or
chick pea-based diets [36]. Thus, the diets containing the extruded mixes would have resulted in a greater energy utilisation of the diet due to the higher amounts of SCFA produced and absorbed [37–39]. Secondly, bile acid excretion was higher in rats fed diets containing extruded mixes (Table 5). Dietary lipids, phospholipids, and cholesterol are solubilised by way of the detergent capacity of bile acids [40], and thus higher intestinal bile acids concentrations would facilitate lipids absorption. Finally, proximate analysis of extruded materials does not lack specific complications [15], which makes it more difficult to accurately establish their protein, carbohydrate, and lipid concentrations, and consequently their energy content for dietary formulation.

Pulses have traditionally been consumed mostly after some kind of heat treatment for gastronomic reasons, but also to reduce the effect of a number of bioactive compounds that in some cases have deleterious effects [41,42]. Compared with the traditional processing methods, extrusion processing, which involves a combined thermo-mechanical treatment with moisture and temperature control, is a quicker and more consistent way to cause thermal/chemical breakdown of deleterious bioactive compounds and at the same time could alter the physical, chemical, and nutritional nature of nutrients in a desirable manner [43]. In fact, pea and kidney bean extrusion treatment has shown a positive effect on mineral, in vitro protein, and starch digestibilities and in vivo nutritional utilisation [44–46]. Even more, extrusion cooking improved the nutritional quality of pea seeds without reducing their hypocholesterolemic and triglyceride-lowering properties [28,30], which is in keeping with our current results (Table 6). However, those previous results were obtained with a more than double (570 g/kg) dietary level of inclusion than those used here. Another relevant point in this investigation was that we used no manipulation of cholesterol metabolism by feeding dietary cholesterol, which is in contrast to what has been previously reported in the literature, where the effects of bean feeding were generally assessed on diet-induced hypercholesterolemia. Additionally, in working with low inclusion levels (100 g/kg) and no cholesterol addition, McPherson [47] found a lowering effect of cooked kidney bean on blood cholesterol. This indicated that legume intake is likely to be a very effective means for keeping low circulating cholesterol values, and that it is able to disturb endogenous cholesterol metabolism, which was also confirmed in the current work by the lowering of liver cholesterol values (Table 5). The mechanisms related with this effect have been linked to the combined effects of a number of bioactive substances and to the protein and fibre [48] or starch [49] fractions of the seed. Legume seed meals and protein isolates are known to modulate plasma amino acid concentrations [29,30,50,51] and induce other physiological effects including hypocholesterolaemia mediated by hormonal (insulin, glucagon) shifts [52,53]. Other reported mechanisms include up-regulation of hepatic mRNA levels of Sterol Regulatory Element-Binding Protein 2 (SREBP-2), a major transcriptional regulator of intracellular cholesterol levels, and Cytochrome P450 7A1 (CYP7A1), the rate-limiting enzyme in bile acid biosynthesis [54], or downregulation of fatty acids synthesis [55,56]. The hypocholesterolemic effect has also been linked to differences in the dietary Lys/Arg and Met/Gly ratios [51,57,58]. In the current work, these ratios for the different diets (Lys/Arg: CAS 2.62, KEM 1.99, PEM 1.92; Met/Gly: CAS 2.35, KEM 0.85, PEM 0.81) were quite different due to differences in amino acid contents in casein and extruded mixes (Table 1). However, these ratios have been found to be more effective in regulating triglyceride than cholesterol values [50,57]. In a laboratory trial using non-overweight and diet-induced obese rats, short-term dietary intake of kidney beans reduced the total plasma cholesterol and low-density lipoprotein (LDL) cholesterol, without affecting high density lipoprotein (HDL) cholesterol or plasma total triglycerides levels [59], which is in line with the current results (Table 6). This would be also in line with the increased faecal bile acids here reported (Table 5), since increased bile acid excretion has also been claimed as a mechanism to explain the cholesterol lowering effect of legume protein-based diets through removal of blood cholesterol via the LDL receptor [48] or bile acids binding within the intestine [60]. Finally, bile acid metabolism is known to be modulated by gut microbiota, affecting the biotransformation, reabsorption, and excretion of bile acids by catalysing a range of biochemical reactions [61]. Nakatani et al. [62] have recently found an elevated cecal and faecal bile acid pool in mice that had consumed mung bean protein, and those changes were linked
to dramatic changes in the gut microbiome, such as such as changes in the Firmicutes/Bacteroidetes proportions (see Rubio et al. [16]).

The dietary inclusion of extruded kidney bean or pea plus rice mixes resulted in significant changes in the proportions of LCFA both in liver and visceral fat. Thus, the inclusion of the extruded mixes induced an increase in the proportions of MUFA in the liver, and of SFA and MUFA in the visceral fat, while the obesogenic diets gave place to an increase in MUFA and a decrease in PUFA in the liver and increases of SFA, MUFA, and a decrease in PUFA in the visceral fat (Tables 7 and 8). Moreover, dietary inclusion of extruded legumes plus cereal mixes gave place to significantly different fatty acid profiles in both liver and visceral fat (Figures 3 and 4) when compared with the casein control diet. Very scant information can be found in the literature thus far on the effects of legume feeding on long chain fatty acid metabolism. In the current work, it is noteworthy that the fat composition of the diets was almost identical, as all of them contained sunflower in the same proportions (Table 2), and the amounts of fat in extruded mixes was lower than 0.2% (Table 1). Therefore, the differences in protein and/or fibre composition of KEM and PEM with respect to CAS are likely to be the ultimate reason for the differences found in this work. In 1992, Ogawa et al. [63] reported that the fatty acid composition of liver phospholipids was influenced by the type of soybean protein used. Quite interestingly, these authors found an increase respect to CAS in C18:2 n-6, and a decrease in C22:5 n-6 in serum and adipose tissue, which is in keeping with the results reported here in liver fat (Table 7) but not in visceral fat (Table 8), albeit with a much lower amount of legume protein. Legume feeding to cows has been reported to induce changes even in milk fatty acids composition [64], most likely due to changes in ruminal fermentation. Finally, obesogenic diets also gave place to increases in SFA and MUFA and decreases in PUFA, presumably due to the much higher amounts of SFA and MUFA in milk fat [65], which is in line with our current results.

5. Conclusions

The substitution of energy-rich foods for pulses has been reported to have beneficial effects on the prevention and treatment of obesity and related disorders such as cardiovascular diseases, type 2 diabetes, and metabolic syndrome. Alternative foods with high palatability such as extruded mixes would be particularly useful particularly in the context of child/adolescent population. The dietary use of limited amounts (closer to practical conditions) of an extruded legumes plus cereal mix did modulate carcass, visceral fat, small intestine, colon, and cecum relative weights in rats. Faecal excretion of bile acids was also affected in rats fed extruded mixes, while obesogenic diets did not induce any significant change in bile acids faecal excretion. The inclusion of extruded mixes lowered liver cholesterol and triglycerides and plasma LDL values, and resulted in different LCFA profiles in liver and visceral fat. To our knowledge, this is the first report on the nutritional and physiological effects of extruded legume plus cereal mixes. The results reported here reinforce the contention on the health benefits of incorporating these kinds of food ingredients into the human diet, and would therefore deserve closer attention in human intervention studies, particularly with adolescents.

**Author Contributions:** Conceptualisation, L.A.R.; methodology, L.A.R. and I.A.-O.; resources, M.M.-P.; data curation, I.A.-O. and L.A.R.; writing—original draft preparation, L.A.R., M.M.-P., and I.A.-O.; writing—review and editing, L.A.R. and M.M.-P.; funding acquisition, M.M.-P. and L.A.R. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was supported by Instituto Nacional de Investigaciones Agrarias (INIA), Consejo Superior de Investigaciones Científicas (CSIC), and the Spanish Ministry of Science and Innovation under the projects RTA2012-00042-C02-01, CSIC201540E083, and AGL 2017-83772-R. The work was also partially supported by the Fondo Europeo de Desarrollo Regional (FEDER) and Fondo Social Europeo (FSE) funds from the European Union.

**Conflicts of Interest:** The authors declare no conflict of interest.

**Abbreviations**

CAS, casein diet; PEM and KEM, diets containing pea or kidney bean extruded mixes, respectively; CAS-OB, PEM-OB, and KEM-OB, obesogenic diets containing casein or pea or kidney bean extruded mixes, respectively.
Foods 2020, 9, 704

References

1. WHO. Global Action Plan for the Prevention and Control of Non-Communicable Diseases 2013–2020; World Health Organization: Geneva, Switzerland, 2013; Available online: http://apps.who.int/iris/bitstream/10665/94384/1/9789241562363_eng.pdf?ua=1 (accessed on 19 March 2020).

2. Ng, M.; Fleming, T.; Robinson, M.; Thomson, B.; Graetz, N.; Margono, C.; Mullany, E.C.; Biryukov, S.; Abbafati, C.; Afera, A.; et al. Global, regional, and national prevalence of overweight and obesity in children and adults during 1980–2013: A systematic analysis for the Global Burden of Disease Study. Lancet 2014, 384, 766–781. [CrossRef]

3. Owen, J.; Reisin, E. Non-communicable disease: A welcome and long needed addition to the WHO’s 2012 world health statistics. Curr. Hypert. Rep. 2012, 14, 475–477. [CrossRef]

4. Haffner, S.M.; Taegtmeyer, H. Epidemic Obesity and the Metabolic Syndrome. Circulation 2003, 108, 1541–1545. [CrossRef]

5. Tilg, H.; Moschen, A.R.; Kaser, A. Obesity and the microbiota. Gastroenterology 2009, 136, 1476–1483. [CrossRef]

6. Engbers, L.H.; van Poppel, M.N.; Paw, M.C.A.; van Mechelen, W. The effects of a controlled worksite environmental intervention on determinants of dietary behaviour and self-reported fruit, vegetable and fat intake. BMC Public Health 2006, 6, 1. [CrossRef]

7. Williams, P.G.; Grafenauer, S.J.; O’Shea, J.E. Cereal grains, legumes, and weight management: A comprehensive review of the scientific evidence. Nutr. Rev. 2008, 66, 171–182. [CrossRef]

8. Dahl, W.J.; Foster, L.M.; Tyler, R.T. Review of the health benefits of peas (Pisum sativum L.). Br. J. Nutr. 2012, 108, S3–S10. [CrossRef]

9. Salas-Salvadó, J.; Guasch-Ferré, M.; Lee, C.-H.; Estruch, R.; Clish, C.B.; Ros, E. Protective effects of the Mediterranean diet on type 2 diabetes and metabolic syndrome. J. Nutr. 2016, 146, 9205–9275. [CrossRef]

10. Rebello, C.J.; Greenway, F.L.; Finley, J.W. A review of the nutritional value of legumes and their effects on obesity and its related co-morbidities. Obes. Rev. 2014, 15, 392–407. [CrossRef]

11. Ruscica, M.; Pavanello, C.; Gandini, S.; Gomaraschi, M.; Vitali, C.; Macchi, C.; Morlotti, B.; Aiello, G.; Bosisio, R.; Calabresi, L.; et al. Effect of soy on metabolic syndrome and cardiovascular risk factors: A randomized controlled trial. Eur. J. Nutr. 2018, 57, 499–511. [CrossRef]

12. Abete, I.; Parra, D.; Martinez, J.A. Legume-, fish-, or high-protein-based hypocaloric diets: Effects of weight loss and mitochondrial oxidation in obese men. J. Med. Food 2012, 15, 100–108. [CrossRef] [PubMed]

13. Marinangeli, C.P.; Jones, P.J. Pulse grain consumption and obesity: Effects on energy expenditure, substrate oxidation, body composition, fat deposition and satiety. Br. J. Nutr. 2012, 108 (Suppl. 1), S46–S51. [CrossRef] [PubMed]

14. Arribas, C.; Cabellos, B.; Cuadrado, C.; Guillamón, E.; Pedrosa, M.M. Bioactive compounds, antioxidant activity, and sensory analysis of rice-based extruded snacks-like fortified with bean and carob fruit flours. Foods 2019, 8, 381–394. [CrossRef]

15. Arribas, C.; Cabellos, B.; Sánchez, C.; Cuadrado, C.; Guillamón, E.; Pedrosa, M.M. The impact of extrusion on the nutritional composition, dietary fiber and in vitro digestibility of gluten-free snacks based on rice, pea and carob flour blends. Food Funct. 2017, 8, 3654–3663. [CrossRef]

16. Rubio, L.A.; Aranda-Olmedo, I.; Contreras, S.; González, G.; Peralta-Sánchez, J.M.; Martin-Pedrosa, M. Inclusion of limited amounts of extruded legumes plus cereal mixes in normocaloric or obesogenic diets for rats: Effects on intestinal microbiota composition. J. Sci. Food Agric. 2020, in press.

17. Shen, W.; Gaskins, R.H.; McIntos, M.K. Influence of dietary fat on intestinal microbes, inflammation, barrier function and metabolic outcomes. J. Nutr. Biochem. 2014, 25, 270–280. [CrossRef]

18. Grant, G.; Dorward, P.M.; Pusztai, A. Pancreatic enlargement is evident in rats fed diets containing raw soybeans (Glycine max) or cowpeas (Vigna unguiculata) for 800 days but not in those fed diets based on kidney beans (Phaseolus vulgaris) or lupin seed (Lupinus angustifolius). J. Nutr. 1993, 123, 2207–2215. [CrossRef]

19. Rubio, L.A.; Muzquiz, M.; Burbano, C.; Cuadrado, C.; Pedrosa, M.M. High apparent ileal digestibility of amino acids in raw and germinated faba bean (Vicia faba)-and chickpea (Cicer arietinum)-based diets for rats. J. Sci. Food Agric. 2002, 82, 1710–1717. [CrossRef]
20. Pedrosa, M.M.; Cuadrado, C.; Burbano, C.; Allaf, K.; Haddad, J.; Gelencsér, E.; Takács, K.; Guillamón, E.; Muzquiz, M. Effect of instant controlled pressure drop on the oligosaccharides, inositol phosphates, trypsin inhibitors and lectins contents of different legumes. *Food Chem.* 2012, 131, 862–868. [CrossRef]

21. Palmquist, D.L.; Jenkins, T.C. Challenges with fats and fatty acids methods. *J. Anim. Sci.* 2003, 81, 3250–3254. [CrossRef]

22. Hagio, M.; Matsumoto, M.; Fukushima, M.; Hara, H.; Ishizuka, S. Improved analysis of bile acids in tissues and intestinal contents of rats using LC/ESI-MS. *J. Lipid Res.* 2009, 50, 173–180. [CrossRef]

23. XLSTAT (Addinsoft, 2017). Available online: [https://www.xlstat.com/es/soluciones/base](https://www.xlstat.com/es/soluciones/base) (accessed on 19 March 2020).

24. Huisamen, B.; Dietrich, D.; Bezuidenhout, N.; Lopes, J.; Flepis, B.; Blackhurst, D.; Lochner, A. Early cardiovascular changes occurring in diet-induced, obese insulin-resistant rats. *Mol. Cell. Biochem.* 2012, 368, 37–45. [CrossRef]

25. Chu, H.-F.; Pan, M.-H.; Ho, C.-T.; Tseng, Y.-H.; Wang, W.W.; Chau, C.-F. Variations in the eicosanoid and fatty acid compositions in different parts of the tigernut (*Cyperus esculentus*) plant. *Chem. Nat. Comp.* 2014, 50, 179–182. [CrossRef]

26. Torre-Villalvazo, I.; Tovar, A.R.; Ramos-Barragán, V.E.; Cerbón-Cervantes, M.A.; Torres, N. Soy protein ameliorates metabolic abnormalities in liver and adipose tissue of rats fed a high fat diet. *J. Nutr.* 2008, 138, 462–468. [CrossRef]

27. Thompson, M.D.; Thompson, H.J.; Brick, M.A.; McGinley, J.N.; Jiang, W.; Zhu, Z.; Wolfe, P. Mechanisms Associated with Dose-Dependent Inhibition of Rat Mammary Carcinogenesis by Dry Bean (*Phaseolus vulgaris*, L.). *J. Nutr.* 2008, 138, 2091–2097. [CrossRef]

28. Alonso, R.; Grant, G.; Marzo, F. Thermal treatment improves nutritional quality of pea seeds (*Pisum sativum* L.) without reducing their hypocholesterolemic properties. *Nutr. Res.* 2001, 21, 1067–1077. [CrossRef]

29. Rubio, L.A.; Grant, G.; Daguid, T.; Brown, D.; Pusztai, A. Organs relative weight and plasma amino acid concentrations in rats fed diets based in legume (faba bean, lupin, chickpea, soybean) seed meals or their fractions. *J. Sci. Food Agric.* 1999, 79, 187–194. [CrossRef]

30. Wang, Y.H.A.; McIntosch, G.H. Extrusion and boiling improve rat body weight gain and plasma cholesterol lowering ability of peas and chickpeas. *J. Nutr.* 1996, 126, 3054–3062. [CrossRef]

31. Olivera, L.; Rodríguez Canul, R.; Pereira-Pacheco, F.; Cockburn, J.; Soldani, F.; Mckenzie, N.H.; Duncan, S.; Olvera-Novoa, M.A.; Grant, G. Nutritional and physiological responses of young growing rats to diets containing raw cowpea seed meal, protein isolate (globulins), or starch. *J. Agric. Food Chem.* 2003, 51, 319–325. [CrossRef]

32. Caligari, S.; Chiesa, G.; Camisassi, D.; Johnson, S.K.; Glio, D.; Marchesi, M.; Parollini, C.; Rubio, L.A.; Sirtori, C.R. Lupin (*Lupinus albus*) protein isolate has adequate nutritional value and reduces large intestinal weight in rats after restricted and ad libitum feeding. *Ann. Nutr. Metab.* 2006, 50, 528–537. [CrossRef]

33. Guo, J.Y.; Li, X.S.; Browning, J.D.; Rottinghaus, G.E.; Lubahn, D.B.; Constantinou, A.; Bennink, M.; MacDonald, R.S. Dietary soy isoflavones and estrone protect ovariectomized ER alpha KO and wild-type mice from carcinogen-induced colon cancer. *J. Nutr.* 2003, 134, 179–182. [CrossRef]

34. Patterson, C.A.; Maskus, R.; Dupasquier, C. Pulse crops for health. *Cereal Foods World* 2009, 54, 108–112. [CrossRef]

35. McBurney, M.I.; Sauer, W.C. Fibre and large bowel energy absorption: Validation of the integrated ileostomy-fermentation model using pigs. *J. Nutr.* 1993, 123, 721–727. [CrossRef]

36. Rubio, L.A. Carbohydrates digestibility and faecal nitrogen excretion in rats fed raw or germinated faba bean (*Vicia faba*)- and chickpea (*Cicer arietinum*)-based diets. *Br. J. Nutr.* 2003, 90, 301–309. [CrossRef]

37. Yang, J.; Keshavarzian, A.; Rose, D.J. Impact of dietary fibre fermentation from cereal grains on metabolite production by the fecal microbiota from normal weight and obese individuals. *J. Med. Food* 2013, 16, 862–867. [CrossRef]

38. Flint, H.J. Obesity and the gut microbiota. *J. Clin. Gastroenterol.* 2011, 45, S128–S132. [CrossRef]

39. Chen, J.; He, X.; Huang, J. Diet effects in gut microbiome and obesity. *J. Food Sci.* 2014, 79, R442–R451. [CrossRef]

40. Ros, E. Intestinal absorption of triglyceride and cholesterol. Dietary and pharmacological inhibition to reduce cardiovascular risk. *Atherosclerosis* 2000, 151, 357–379. [CrossRef]
41. Muzquiz, M.; Varela, A.; Burbano, C.; Cuadrado, C.; Guillamón, E.; Pedrosa, M.M. Bioactive compounds in legumes: Pronutritive and antinutritive actions. Implications for nutrition and health. *Phytochem. Rev.* 2012, *11*, 227–244. [CrossRef]

42. Gulewicz, P.; Martínez-Villaluenga, C.; Kasprowicz-Potocka, M.; Frias, J. Non-Nutritive compounds in fabaceae family seeds and the improvement of their nutritional quality by traditional processing—A review. *Pol. J. Food Nutr. Sci.* 2014, *64*, 75–89. [CrossRef]

43. Nikmaram, N.; Ying Leong, S.; Koubaa, M.; Zhu, Z.; Barba, F.J.; Greiner, R.; Oey, I.; Roohinejad, I. Effect of extrusion on the anti-nutritional factors of food products: An overview. *Food Control* 2017, *79*, 62–73. [CrossRef]

44. Alonso, R.; Aguirre, A.; Marzo, F. Effects of extrusion and traditional processing methods on antinutrients and in vitro digestibility of protein and starch in faba and kidney beans. *Food Chem.* 2000, *68*, 159–165. [CrossRef]

45. Alonso, R.; Grant, G.; Dewey., P.; Marzo, F. Nutritional assessment in vitro and in vivo of raw and extruded peas (*Pisum sativum* L.). *J. Agric. Food Chem.* 2000, *48*, 2286–2290. [CrossRef]

46. Alonso, R.; Rubio, L.A.; Muzquiz, M.; Marzo, F. The effect of extrusion cooking on mineral availability in pea and kidney bean seed meals. *Anim. Feed Sci. Technol.* 2001, *94*, 1–13. [CrossRef]

47. McPherson, L. Effects of the consumption of fully cooked red kidney beans (*Phaseolus vulgaris*) on the growth rate of rats and the morphology of the gut wall. *J. Sci. Food Agric.* 1991, *57*, 611–621. [CrossRef]

48. Potter, S.M. Overview of proposed mechanisms for the hypocholesterolemic effect of soy. *J. Nutr.* 1995, *125*, 606S–611S.

49. Krupa-Kozak, U.; Juskiewicz, J.; Wrókowska, M.; Soral-Smietana, M.; Zdunczyk, Z. Native and microwaved bean and pea starch preparations: Physiological effects on the intestinal ecosystem, caecal tissue and serum lipids in rats. *Br. J. Nutr.* 2010, *103*, 1118–1126. [CrossRef]

50. Ruβio, L.A.; Grant, G.; Scislowski, P.W.O.; Brown, D.; Bardocz, S.; Pusztaı, A. The utilization of lupin (*Lupinus angustifolius*) and faba bean globulins by rats is poorer than of soybean globulins or lactalbumin but the nutritional value of lupin seed is lower only than that of lactalbumin. *J. Nutr.* 1995, *125*, 2145–2155. [CrossRef]

51. Fernández-Quintela, A.; del Barrio, A.S.; Macarulla, M.T.; Martínez, J.A. Nutritional evaluation and metabolic effects in rats of protein isolates obtained from seeds of three legume species. *J. Sci. Food Agric.* 1998, *78*, 251–260. [CrossRef]

52. Ruβio, L.A. Physiological effects of legume storage proteins. *Nutr. Abs. Rev. Ser. A Hum. Exp.* 2000, *70*, 197–204.

53. Torres, N.; Torre-Villalvazo, I.; Tovar, A.R. Regulation of lipid metabolism by soy protein and its implication in diseases mediated by lipid disorders. *J. Nutr. Biochem.* 2006, *17*, 365–373. [CrossRef] [PubMed]

54. Parolini, C.; Rigamonti, E.; Marchesi, M.; Busnelli, M.; Cinquanta, P.; Sirtori, C.R.; Chiesa, G. Cholesterol-lowering effect of dietary *Lupinus angustifolius* proteins in adult rats through regulation of genes involved in cholesterol homeostasis. *Food Chem.* 2012, *132*, 1475–1479. [CrossRef] [PubMed]

55. Rigamonti, E.; Parolini, C.; Marchesi, M.; Diani, E.; Brambilla, S.; Sirtori, C.R.; Chiesa, G. Hypolipidemic effect of dietary pea proteins: Impact on genes regulating hepatic lipid metabolism. *Mol. Nutr. Food Res.* 2010, *54*, S24–S30. [CrossRef] [PubMed]

56. Watanabe, H.; Inaba, Y.; Kimura, K.; Asahara, S.-I.; Kido, Y.; Matsumoto, M.; Motoyama, T.; Tachibana, N.; Kaneo, S.; Kohno, M.; et al. Dietary mung bean protein reduces hepatic steatosis, fibrosis, and inflammation in male mice with diet-induced, nonalcoholic fatty liver disease. *J. Nutr.* 2017, *147*, 52–60. [CrossRef] [PubMed]

57. Sugano, M.; Ishiwaki, N.; Nagata, Y.; Imaizumi, K. Effects of arginine and lysine addition to casein and soya-bean protein on serum lipids, apolipoproteins, insulin and glucagon in rats. *Br. J. Nutr.* 1982, *48*, 211–221. [CrossRef]

58. Tanaka, K.; Sugano, M. Effects of addition of sulfur containing amino acids and glycine to soyabean protein and casein on serum cholesterol levels of rats. *J. Nutr. Sci. Vitaminol.* 1989, *35*, 323–332. [CrossRef]

59. Zhu, Z.; Jiang, W.; Thompson, H.J. Edible dry bean consumption (*Phaseolus vulgaris* L.) modulates cardiovascular risk factors and diet-induced obesity in rats and mice. *Br. J. Nutr.* 2012, *108*, S66–S73. [CrossRef]
60. Ngoh, Y.-Y.; Choi, S.B.; Gan, C.-Y. The potential roles of Pinto bean (*Phaseolus vulgaris* cv. Pinto) bioactive peptides in regulating physiological functions: Protease activating, lipase inhibiting and bile acid binding activities. *J. Funct. Foods* 2017, 33, 67–75. [CrossRef]

61. Wahlström, A.; Sayin, S.; Marschall, H.U.; Backhed, F. Intestinal crosstalk between bile acids and microbiota and its impact on host metabolism. *Cell. Metab.* 2016, 24, 41e50.

62. Nakatani, A.; Li, X.; Miyamoto, J.; Igarashi, M.; Watanabe, H.A.; Watanabe, K.; Motoyama, T.; Tachibana, N.; Kohno, M.; Inoue, H.; et al. Dietary mung bean protein reduces high-fat diet-induced weight gain by modulating host bile acid metabolism in a gut microbiota-dependent manner. *Biochem. Biophys. Res. Commun.* 2018, 501, 955e961. [CrossRef]

63. Ogawa, T.; Gatchalian-Yee, M.; Sugano, M.; Kimoto, M.; Matsuo, T.; Hashimoto, Y. Hypocholesterolemic effect of undigested fraction of soybean protein in rats fed no cholesterol. *Biosci. Biotechnol. Biochem.* 1992, 11, 1845. [CrossRef]

64. Woods, V.B.; Fearon, A.M. Dietary sources of unsaturated fatty acids for animals and their transfer into meat, milk and eggs: A review. *Livest. Sci.* 2009, 126, 1–20. [CrossRef]

65. Benbrook, C.M.; Butler, G.; Latif, M.A.; Leifert, C.; Davis, D.R. Organic production enhances milk nutritional quality by shifting fatty acid composition: A United States–wide, 18-month study. *PLoS ONE* 2013, 8, e82429. [CrossRef] [PubMed]

© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).