Production of Endoglucanases by *Streptomyces thermocoprophilus* CP1 using Rice Straw as a Substrate

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**Abstract**

Rice straw is a major agricultural waste that can be used as an alternative substrate to expensive raw materials for endoglucanases (CMCase) production by microorganisms. This study aimed to search for a microorganism having the potential to produce endoglucanase from rice straw. From compost samples, 40 bacterial colonies were isolated on carboxymethylcellulose (CMC) agar. Among them, 16 isolates showed a hydrolysis zone on a CMC agar plate with hydrolysis (HC) values ranging from 1.15±0.02 to 4.40±0.52. Based on hydrolysis zone diameter and HC value, isolates CP1, CP2 and CP3 were further examined for their CMCase production in CMC broth. According to CMCase production and stability, isolate CP1 was selected for further study. The optimal pH and temperature for CMCase production of isolate CP1 were 5 and 45 °C, respectively. When using pre-treated rice straw as a substrate for semi-solid-state fermentation, the highest CMCase activity of 0.142 ± 0.008 U/mL was obtained in a medium containing pre-treated rice straw of 60 g/L. The sequence alignment analysis and phylogenetic analysis suggested that the isolate CP1 was likely to be *Streptomyces thermocoprophilus*. The microorganism obtained from this study may be not only industrially important but also beneficial to the environment.

**Keywords:** Endoglucanases, *Streptomyces thermocoprophilus*, rice straw, actinomyces, semi-solid-state fermentation

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INTRODUCTION

Cellulase enzymes are a group of complex enzymes converting cellulose to glucose and other sugars. The cellulase enzymes comprise three major types of enzymes: endoglucanases (E.C. 3.2.1.4), exoglucanases, including cellobiohydrolases (CBHs) (E.C. 3.2.1.91) and β-glucosidase (E.C. 3.2.1.21). Endoglucanases randomly breakdown β-1,4 glycosidic bonds in the cellulose strand releasing glucose and oligosaccharides. Cellobiohydrolases act at termini of cellulose and oligosaccharides produced by endoglucanases to convert them to cellobiose which is further digested by β-glucosidase to glucose. Synergetic activities of all these three enzymes provide the best result of complete hydrolysis of raw cellulose to glucose.

Cellulases are important enzymes with many applications in many industries and agriculture, especially in bioethanol production, textile and food industries. Although the enzymes are found to be produced by fungi and bacteria, those from bacteria including actinomycetes are become exploited because of their high growth rate and availability in diverse habitats, including extreme environments that are sources of thermophilic, psychrophilic, alkaliphilic, acidophilic, and halophilic strains. Actinomycetes become an attractive microorganism for cellulase production. Various strains of actinomycetes have been evaluated for their ability to degrade the lignocellulosic biomass, which can be potentially implemented in the lignocellulolytic enzymes production from different value-added products. Lignocellulosic biomass (LCB) is the most abundant organic compound found in nature, especially in plant cell walls. Recently, LCB has gained increasing attention as economic feedstock for second-generation (2G) biofuels. LCB conversion to bioethanol consists of four major steps including pretreatment, saccharification, fermentation and distillation. The pretreatment process is performed to remove or redistribute the lignin, to reduce the cellulose crystallinity, and to increase the porosity significantly. Subsequent saccharification or hydrolysis is done by acids or enzymes to hydrolyze cellulose and hemicellulose into fermentable sugars (hexoses and pentoses). Enzymatic hydrolysis is preferred over acid hydrolysis due to low energy requirement and by-product formation. The enzymes required for efficient deconstruction of polysaccharides into monomeric sugars include modular and non-modular glycosyl hydrolases (GHs) comprising of cellulases and hemicellulases, carbohydrate esterases (CEs), and auxiliary activity (AA) proteins.

Rice straw is a major agricultural biomass waste produced in Southeast Asia and particularly in Thailand, and yet most of this biomass is left unutilized and is often burned in the field causing severe environmental problems at both local and global levels. It is estimated that over 20 million ton of rice straw are produced in Thailand every year. Similar to other plant residues, rice straw is mainly composed of cellulose, hemicellulose, and lignin. Cellulose is a complex carbohydrate with a high molecular weight and a maximum of 10,000 monomeric units of D-glucose, linked by β-1,4-glycosidic bonds. Hemicellulose is strongly linked to the surface of cellulose microfibrils. It entirely consists of pentose (xylose and arabinose) and hexose sugars (glucose, galactose, mannose, and rhamnose). Lignin denotes a complex amorphous aromatic polymer with a three-dimensional network, composed of phenylpropane units linked together. Having a complex structure consisting mainly of cellulose and hemicellulose, rice straw is difficult to be degraded and utilized as a bio-resource. To overcome such problem, the invention of efficient, practical and cost effective approaches to rice straw degradation is necessary. The attractive of using lignocellulosic biomass as a sustainable feedstock for biofuel and others valuable bioactive compound productions, many research works related to isolating cellulase producing actinomycetes and their applications were conducted during the last five years as summarized in Table 1. In the present study, various actinomycetes were isolated from compost made in Ubon Ratchathani, Thailand and screened for their potential to produce cellulolytic enzymes. The actinomycetes with the most efficient cellulase production was further evaluated for its ability to produces cellulase when using rice straw as a substrate. The production of cellulase enzymes from biomass is a way to increase the value of agricultural waste and reduce raw material cost of enzyme production process.
Table 1. Research works related to isolating cellulase producing actinomycetes and their applications during the last five years

| Source of actinomycetes | Pre-treatment method | Enzymes production conditions | Maximum enzyme activity | Selected strains | Future application | Ref. |
|-------------------------|----------------------|-------------------------------|-------------------------|------------------|-------------------|------|
| soil                    | without pretreatment | 0.5% CMC pH 6.5 Temp 45°C 160 rpm, 4 d | endoglucanase 27 IU/mL | *Streptomyces* sp. DSK59 | sorghum stover hydrolysis incombinaion with commercial cellulase animal feed | 20   |
| soil of an olive pressing factory | mashing (Ø <1 mm) | 3% olive pomace powder pH 7.4 Temp 40°C 150 rpm, 30 d | cellulase 1.44 ±0.02 U/mL | *Streptomyces* sp. S1M3I | in use in composting process of rubber bark | 21   |
| agricultural and agro-industrial residues/soil | grinding to an average size of 0.5 mm | 10% of decomposed rubber bark without pH adjustment Temp 30°C 150 rpm, 4 d | CMCase 0.58 U/mL xylanase 0.72 U/mL | *S. albogriseolus* A2 | | 22   |
| soil                    | without pretreatment | 1% CMC without pH adjustment Temp 37°C 120 rpm 5 d | endoglucanase 4.9 U/mL | *S. macrosporeus* KRC 21.D | endoglucanase production | 23   |
| lignocellulosic compost | grinding/soaking in 15% (w/v) NaOH with 3% H₂O₂ for 4 h and autoclaved at 121°C, 5 min | 2% of empty fruit bunch pH 6.5 Temp 45°C 150 rpm, 5 d | endoglucanase 925 U/g substrate | *Thermocaprophilus* TC13W | endoglucanase production from low cost agriculture waste | 24   |
MATERIALS AND METHODS

Isolation of endoglucanases-producing actinomycetes from compost

The samples of composting agricultural waste were collected from the Faculty of Agriculture, Ubon Ratchathani University, Thailand. One gram of compost sample was subjected to ten-fold serial dilution with sterilized peptone water containing peptone 1.0, NaCl 8.5 (g/L). One mL of each dilution (for 10⁻⁴ to 10⁻⁶) was spread on carboxymethylcellulose (CMC) agar and then incubated at 37°C until colonies were observed. Colonies with different morphological characteristics were picked up. The selected colonies were purified by cross streak technique on CMC agar and stored at 4°C in CMC slant and 20% (v/v) glycerol at -20°C. All of them were subjected to gram staining and cell morphology investigation under a light microscope.

The rice straw pretreatment

Rice straw locally grown in Warin Chamrap district, Ubon Ratchathani province, Thailand was collected, washed, cut, dried at 60°C and crushed to 0.5 mm particles. The resulting particles were treated with 2% (w/v) NaOH at room temperature for 24 h and washed with water till neutral pH and dried at 60°C.

Primary screening for endoglucanases-producing actinomycetes

The primary screening of endoglucanases-producing actinomycetes was carried out on an agar medium (CMC plate assay method). All of the isolated actinomycetes cells were pre-cultured on CMC broth at 37°C for 3 d. After that 5 µL of cultures were dropped on CMC agar and then incubated at 37°C. After 7 d incubation, a lugol-iodine solution was poured over CMC plates for 15 min and then rinsed off. A clear zone formation around the actinomycetes colonies indicated the hydrolysis of CMC. The highest endoglucanases activity is assumed by the highest ratio of the clear zone diameter to colony diameter. Hydrolysis capacity (HC) value was calculated as described by Lu et al.27

Table 1. Cont....

| Source of actinomycetes | Pre-treatment method | Enzymes production conditions | Maximum enzyme activity | Selected strains | Future application | Ref. |
|-------------------------|---------------------|-------------------------------|-------------------------|------------------|--------------------|-----|
| soil, rotten wood, leaf and rice straw | soaking rice straw in 1% (w/v) NaOH for 24 h | 1% rice straw pH 7.0 Temp 30°C 100 rpm, 12 d | endoglucanase 0.406 U/mL | S. roseofulvus | cellulase production from agricultural waste treatment | 25 |
| rice straw | soaking sugarcane bagasse in 1% (w/v) NaOH for 24 h | 1% sugarcane bagasse pH 7.0 Temp 30°C 100 rpm, 12 d | | | | |
| soil | mashing (800 and 2000 µm) | | endoglucanase 13.85±0.22 U/g amyrase 20.97±0.16 U/g pectinase 11.24±0.14 U/g xylanase 15.22±0.11 U/g endoglucanase 0.142 ± 0.008 U/mL | S. fulvissimus CK7 | production ethanol from agriculture residues | 26 |
| compost | milling and soaking in 1%(w/v) NaOH during autoclaving at 121°C, 15 min | 6% rice straw solid state fermentation pH 5.0 Temp 45°C 5 d | | S. thermocoprophilus CP1 | use in ethanol or bioactive natural products production from lignocellulosic biomass | This study |
Secondary screening for endoglucanases-producing actinomyces

Three isolates with high HC values or large clear zones were evaluated for their endoglucanases production potential in CMC broth. The actinomyces cells were activated on CMC agar at 37°C until sporulation. The spore suspension was prepared by scratching of each agar with 10 mL distilled water containing 0.1% tween 80. For endoglucanases production, 2 mL of the spore suspension was used to inoculate into a flask containing 50 mL CMC broth and then incubated at 37°C for 10 d. Samples were taken every 2 d for cellulase activity assay. The samples were centrifuged at 5,000 rpm for 20 min and the resulting supernatants were used for enzyme activity assay.

Effect of pH and temperatures on endoglucanases production

The optimal pH and temperature for endoglucanases production of the selected isolate CP1 were determined. To determine the optimal pH, the isolate CP1 was pre-cultured on CMC broth at 37°C for 3 d. After that 5 µL of cultures were dropped on CMC agar with different pH ranging from 3 to 8 and incubated at 37°C for 5 d. To determine the optimal temperature, five plates of CMC agar with pH adjusted to 5 were inoculated and incubated at different temperatures including 30, 35, 40, 45, and 50°C for 5 d. After incubation, a clear zone diameter was determined and HC values were calculated.

Endoglucanases production by using pretreated rice straw

To evaluate the ability of isolate CP1 to produce endoglucanases when using rice straw as a substrate, semi-solid-state fermentation was performed. Two mL of the spore suspension with the initial spore concentration of 107 spores/mL was used as an inoculum. Fermentation medium containing 1 g of pretreated rice straw moistened with 15 mL mineral solution containing CaCl₂ 0.1, MgSO₄.7H₂O 0.1, K₂HPO₄ 1.0, NaCl 0.2, yeast extract 5.0, tween 80 1.5, (NH₄)₂SO₄ 0.5, FeSO₄ 0.1 and NaNO₃ 1.0 (g/L) in 250 ml Erlenmeyer and culture condition at 37°C for 5 d were used. At the end of the fermentation process, the supernatant obtained from centrifugation at 5000 rpm for 20 min was used for endoglucanases activity assay.

CMCase activity assay

CMCase or endoglucanase activity was assayed followed the method of Ghose²⁸ with slightly modification. The reaction mixture of substrate (pH 4.8) and crude enzyme was incubated at 50°C for 10 min. The reducing sugar was analyzed by the method described by Miller.²⁹ One enzyme unit (U) is defined as the amount of enzyme required to produce 1 µmoles of glucose per min under the assay conditions. CMCase activity was expressed as units per mL of a crude enzyme (U/mL).
Molecular identification of isolate CP1

Molecular techniques were conducted to identify the isolate CP1. The 16S rRNA gene was amplified from the isolate CP1 genomic DNA by the conventional PCR method using universal primer pairs, 27F and 1492R. The amplicon was sent to Macrogen for sequencing. The sequence was subjected to sequence alignment analysis using NCBI BLASTN program. Multiple alignments by ClustalW and Neighbor Joining tree construction of the CP1 16S rRNA gene and those of other Streptomyces species available on GenBank database were conducted on MEGA 7.0 software.

RESULTS

From the compost samples, a total of 40 isolates grown on CMC agar were collected. Among them, only 16 isolates produced a zone of hydrolysis indicating the production of extracellular endoglucanases (Table 2). Colony morphological observation showed that all of them had similar characteristics such as cream color, lobate margin, and convex surface except for shape being observed as round or irregular (Table 2). Observation under a light microscope indicated that all of them grew as filamentous and were found to be gram-positive bacteria.

Based on the result of the CMC plate assay method, out of the total 40 isolates, 16 isolates showed zone of hydrolysis indicating production of extracellular endoglucanases. The isolate CP1 grew on CMC agar and produced the largest hydrolysis zone (Table 3). However, the highest HC values of 4.40±0.52, 3.79±0.18 were observed in isolates CP2 and CP3, respectively (Table 3). Therefore, isolates CP1, CP2 and CP3 were further investigated for their ability to produce endoglucanases enzyme in liquid CMC medium.

To compare endoglucanases production of isolates CP1 CP2 and CP3, CMCase activity profiles were followed every 2 d until 14 d. In the first 6 d of observation, CMCase activities of all 3 isolates were somewhat stable. After day 6, CMCase activities of CP2 and CP3 substantially decrease whereas that of CP1 continuously increased towards the end of fermentation (Fig. 1). Based on CMCase production and stability, isolate CP1 was selected for further investigation.

The optimal pH and temperature for isolate CP1 to provide the highest endoglucanases activity were determined. To determine the optimal pH, hydrolysis zones produced by isolate CP1 grown on CMC agar adjusted pH between 4 and 8 were observed. It was revealed that the optimal pH for endoglucanases production was

| Isolate | Colony diameter (cm) | Hydrolysis zone diameter (cm) | HC value |
|---------|----------------------|------------------------------|----------|
|         | 1   | 2   | 3   | average | 1   | 2   | 3   | average |             |
| CP1     | 2.35 | 2.35 | 2.35 | 2.35±0.00 | 5.40 | 5.40 | 5.20 | 5.33±0.09 | 2.24±0.04 |
| CP2     | 0.90 | 0.70 | 0.90 | 0.83±0.12 | 3.65 | 3.35 | 3.55 | 3.52±0.12 | 4.40±0.52 |
| CP3     | 0.90 | 0.90 | 1.05 | 0.95±0.09 | 3.55 | 3.60 | 3.65 | 3.60±0.04 | 3.79±0.18 |
| CP5     | 1.20 | 1.10 | 1.35 | 1.22±0.13 | 3.90 | 3.80 | 3.95 | 3.88±0.06 | 3.24±0.25 |
| CP7     | 2.00 | 2.00 | 2.05 | 2.02±0.03 | 3.05 | 2.95 | 2.95 | 2.98±0.05 | 1.50±0.03 |
| CP8     | 1.90 | 1.90 | 1.80 | 1.87±0.06 | 2.35 | 2.45 | 2.35 | 2.38±0.05 | 1.26±0.02 |
| CP9     | 1.00 | 1.00 | 1.20 | 1.07±0.12 | 3.55 | 3.50 | 3.80 | 3.62±0.13 | 3.42±0.16 |
| CP17    | 1.30 | 1.40 | 1.35 | 1.35±0.05 | 1.55 | 1.50 | 1.55 | 1.53±0.02 | 1.15±0.02 |
| CP18    | 2.00 | 1.90 | 2.00 | 1.97±0.06 | 2.95 | 2.95 | 2.85 | 2.92±0.05 | 1.48±0.05 |
| CP25    | 1.50 | 1.60 | 1.45 | 1.52±0.08 | 3.10 | 3.05 | 3.10 | 3.08±0.02 | 2.05±0.08 |
| CP26    | 1.10 | 1.00 | nd   | 0.70±0.07 | 3.55 | 3.35 | nd   | 2.30±0.10 | 3.39±0.08 |
| CP28    | 1.20 | 1.20 | 0.75 | 1.05±0.26 | 3.60 | 3.70 | 3.20 | 3.50±0.22 | 3.42±0.52 |
| CP29    | 1.70 | 1.80 | 1.80 | 1.80±0.10 | 2.35 | 2.45 | 2.35 | 2.38±0.05 | 1.31±0.03 |
| CP38    | 2.10 | 2.00 | 2.10 | 2.07±0.06 | 3.10 | 3.30 | 3.20 | 3.20±0.08 | 1.52±0.01 |
| CPY     | 0.90 | 1.20 | nd   | 0.70±0.21 | 3.45 | 3.85 | nd   | 3.65±0.20 | 3.35±0.48 |
| CPZ     | 1.30 | 1.30 | 1.20 | 1.27±0.06 | 4.15 | 4.05 | 4.05 | 4.08±0.05 | 3.25±0.07 |

nd = not determine
between pH 4 and 5 where the largest hydrolysis zones were observed (Fig. 2). At pH 6 and 7, hydrolysis zones were decreased in size and not observed at pH 8 (Fig. 2). However, the growth of isolate CP1 at pH 6-8 was better than at pH 4-5 indicating that isolate CP would prefer acidic conditions for endoglucanases production and neutral or weak acidic or alkali conditions for their growth.

The influence of temperature on endoglucanases activity was determined by incubating the CMC agar plates at a range of temperatures of 30, 35, 40, 45, and 50°C. It was observed that HC values increased with increasing temperature and the optimal temperature for endoglucanases production was 45°C (Fig. 3). However, endoglucanases production decreased when temperature was increased up to 50°C.

To apply rice straw as a substrate for endoglucanases production by isolate CP1, substrate concentration is essential to support the proper growth of the bacteria. Therefore, the effect of rice straw content on endoglucanases production was studied. Different amounts of pre-treated rice straw (1, 2, 3, 4 and 5 g) were moist with 50 mL of distilled water and spores with the initial concentration of 1 x 107 spores/mL were inoculated to the rice straw. Although, the optimal temperature for endoglucanases production of isolate CP1 was 45°C but incubating at that temperature caused reduction of water content. Therefore, semi-solid-state fermentation was carried out at 40°C. When using rice straw contents of 20-60 g/L, a continuous increase of CMCase activity was observed (Fig. 4). However, increasing rice straw contents up to 80-100 g/L lowered the CMCase activity (Fig. 4).

The partial sequence of the isolate CP1 16S rRNA gene obtained from PCR amplification was deposited to the GenBank sequence database under the accession number MZ413512. The sequence was subjected to sequence alignment analysis and phylogenetic analysis. The sequence alignment analysis using NCBI BLASTN program

**Fig. 1.** CMCase activities of isolates CP1 ( ), CP2 ( ), and CP3 ( ) when grown in CMC broth at 37°C.

**Fig. 2.** Effect of pH on endoglucanases production of isolate CP1 when grown on CMC agar and incubated at 37°C for 5 d.
revealed that the sequence of CP1 16S rRNA gene was 99.57% identical to that of *Streptomyces thermocoprophilus* strain NBRC 100771 (accession number NR_112594.1). The phylogenetic tree constructed from 16S rRNA genes of the isolate CP1 and other *Streptomyces* species available on the GenBank database revealed that the isolate CP1 was closely related to *Streptomyces thermocoprophilus* strain NBRC 100771 with high bootstrap support of 91% (Fig. 5). The concordance of the results from sequence alignment analysis and phylogenetic analysis suggested that the isolate CP1 was likely to be *Streptomyces thermocoprophilus*.

**DISCUSSION**

An important step for the utilization of lignocellulosic biomass is the conversion of the biomass into smaller and simple structures. This step requires the catalytic activity of lignocellulolytic enzymes including cellulases, hemicellulases, and lignolytic enzyme. Therefore, attempts have been spent to isolate new microorganisms producing high activity of cellulolytic enzymes. Many different habitats such as soil, compost, forest waste, animal feces, paper mills, and hot springs have been used as the sources of isolating cellulase producing microorganisms. In the present study, composts

![Fig. 3. Effect of temperature on endoglucanases production of CP1 when grown on CMC agar for 5 d.](image)

![Fig. 4. The CMCase activity of isolate CP1 when using various content of pretreated rice straw as a substrate for semi-solid-state fermentation and incubating at 40°C for 5 d.](image)
were used to isolate endoglucanases producing microorganisms and the result showed that all of our endoglucanases producing isolates were gram positive and filamentous in shape that are characteristics of actinomycetes. In agreement with our study, actinomycetes are well known to be components of the microflora of composts.\textsuperscript{34} Most of actinomycetes capable of producing cellulolytic enzyme belonging to the genus \textit{Streptomyces}, such as \textit{S. albogriseolus},\textsuperscript{35} \textit{S. flavogriseus},\textsuperscript{36} \textit{S. albaduncus},\textsuperscript{37} \textit{S. cellulolyticus},\textsuperscript{38} and \textit{S. albospinus}.\textsuperscript{39}

In our study, \textit{S. thermocoprophilus} CP1 could produce endoglucanases at a pH range of 3.0 to 7.0. The enzyme production was drastically reduced at pH 6.0-7.0 and was not observed at pH 8.0. The initial pH has been reported to influence enzymes transport across the cell membrane.\textsuperscript{40} The optimal pH for endoglucanases production of \textit{Streptomyces} spp. was varied from 5.0 to 7.0. Similar to our results, the optimal pH for cellulase production of \textit{Streptomyces} sp. MDS was 5.0.\textsuperscript{41} In contrast, the optimal pH for cellulase and xylanase productions of \textit{Streptomyces} sp. AMT-3,\textsuperscript{42} \textit{S. transformant} T3-1\textsuperscript{43} and \textit{S. drozdowiczii}\textsuperscript{44} were 6.5-7.0.

Temperature is one of the key factors that significantly impacts microbial growth and enzyme production. In this study, we investigated a temperature range of 30-50°C for the optimal temperature for endoglucanases production by \textit{S. thermocoprophilus} CP1. This strain gave the highest HC value of 2.49±0.01 at 45°C (Fig. 3). Several studies have shown that optimal temperatures for cellulase production of \textit{S. drozdowiczii},\textsuperscript{44} \textit{Streptomyces} T3-1\textsuperscript{43} and \textit{Streptomyces} sp. SLBA-08\textsuperscript{8} were more than 45°C. However, some \textit{Streptomyces} strains such as \textit{S. albidosavus} SAMRC-UFH5\textsuperscript{8} and \textit{Streptomyces} sp. MDS\textsuperscript{41} optimally produced cellulases at lower temperature (40°C). The ability to grow and produce cellulolytic enzymes at high temperature (35-50°C) in acidic condition makes \textit{S. thermocoprophilus} CP1 suitable for co-cultivation with other thermotolerant yeast such as \textit{K. marxianus} and \textit{S. cerevisiae} (thermotolerant strains) to produce some valuable products in simultaneous saccharification and fermentation (SSF).

A suitable pretreatment method is a key in breaking down the recalcitrant lignin structure leading to the accessibility of cellulose towards hydrolytic enzymes for its conversion into monosaccharides.\textsuperscript{15} There are various pretreatment processes applied for lignocellulosic biomass. The choice of pretreatment methods relies on economic factor, type of lignocellulosic feedstock, and environmental impacts. In present study, a rice straw was milled and followed by alkali pretreatment carried out at high temperatures (121°C) for 15 min. Milling can increase the specific surface area for enzymatic hydrolysis and degree of

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\textbf{Fig. 5.} Neighbor-joining tree of the isolate CP1 and other \textit{Streptomyces} species. Values on nodes represent percentage bootstraps out of 1000 bootstrap samples; values less than 50% are not shown. Scale bar represents the number of mutations per sequence position.
polymerization, but at the same time can reduce cellulose crystallinity. Alkali application is able to alter the structure of lignin by degrading ester and glycosidic side chains, cellulose swelling, and partial cellulose recrystallization. However, only alkali pretreatment would not be enough to remove hemicellulose. Therefore, to enhance cellulase productivity by increasing cellulose accessibility to cellulolytic enzymes, other pretreatment methods should be investigated. Ionic liquid pretreatment is a novel and emerging pretreatment technologies. Recently, the application of ionic liquid (IL) for lignocellulose pretreatment has attracted considerable attention. These compounds have been exploited extensively due to their green properties, high thermal stabilities and negligible vapor pressures preventing the release of toxic gases. Pretreatment of rice straw with cholinium amino acids ionic liquids ([Ch][AA] ILs), a novel type of bio-ILs, resulted in the improvement of the initial saccharification rates of rice straw residues and polysaccharide digestion.45

Submerged and solid state fermentation has been used for cellulases production.40 Generally, the production of cellulases by microorganisms has been reported to utilize the submerged fermentation.20-25,47,48 However, for cellulases production, the use of lignocellulosic biomass along with the application of cost effective fermentation strategies such as solid-state fermentation has been suggested.49 The solid-state fermentation usually used for fungi cultivation but not for Streptomyces species. However, we found that S. thermocoprophilus CP1 could grow and produce cellulase using semi-solid state fermentation. Therefore, our result would be useful for further investigation on using semi-solid state fermentation for cellulases production from others lignocellulosic biomass by Streptomyces spp.

To make concept of biorefineries economically feasible, genetic engineering techniques have been used recently to improve the enzymatic expression of microorganisms.55 Unlike Escherichia coli, Streptomyces spp. have not been a popular microorganisms for metabolic engineering because they are not well-characterized. However, during the last decade, metabolic engineering strategies have been applied to more microorganism species other than E. coli.53 CelStrep, a 379-amino-acid endoglucanase encoded by celstrep gene, was identified in Streptomyces sp. strain G12 and classified as a family 12 glycoside hydrolase. By using recombinant technology, the recombinant CelStrep was successfully produced by E. coli.54 Since molecular genetic tools are available for Streptomyces spp., cellulase gene cloning and sequencing could be further carried out in our strain.

Streptomyces is considered to be a very large genus consisting of more than 500 species with the number increasing every year. Among closely related Streptomyces species, their 16S rRNA gene sequences are very similar. This sometimes can cause ambiguity in molecular identification of Streptomyces spp. For example, the BLASTN analysis showed that 16S rRNA gene sequence of the isolate GMKU 937 was found to be 99% identical to that of Streptomyces coelicoflavus NBRC 15399 but the phylogenetic analysis demonstrated that Streptomyces diastaticus NRRL B-1773 15399 was the closest species of the isolate GMKU 937.55 Sometimes results from the BLASTN analysis and phylogenetic analysis were in agreement but the bootstrap supports in phylogenetic trees were very low (less than 50%).56 Fortunately, those problems were not found in this study. The isolate CP1 was concordantly identify as Streptomyces thermocoprophilus by both BlastN analysis and phylogenetic analysis with high percentage bootstrap support.

CONCLUSIONS AND FUTURE RESEARCH PERSPECTIVES

S. thermocoprophilus CP1 isolated from the compost was shown to produce endoglucanases using rice straw as a substrate. This finding may be beneficial not only for industries but also environments. The endoglucanases produced by S. thermocoprophilus CP1 may convert environmental wastes such as rice straw and other cellulosic biomass to produce sugars for various industries including bioethanol and other bioactive natural products production. Future studies should also consider investigating the sustainability features of the developed platform using advanced sustainability assessment tools such as LCA, exergy and their combination as mention in several articles.57 In addition, enzyme
characterization, productions of exoglucanase and beta-glucosidase, improving enzyme productivity by using genetic tools should be further studied in *S. thermocoprophilus* CP1.

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**CONFLICT OF INTEREST**
The authors declare that there is no conflict of interest.

**AUTHORS’ CONTRIBUTION**
All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

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**DATA AVAILABILITY**
All datasets generated or analyzed during this study are included in the manuscript.

**ETHICS STATEMENT**
This article does not contain any studies with human participants or animals performed by any of the authors.

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