Constituents and Anthelmintic Activity Evaluation of Albizia Adiantifolia (Schumach) W.F. Wright Essential Oils From Nigeria

Akinsola Akande¹, Sherifat Aboaba¹, Guido Flamini²

¹Department of Chemistry, University of Ibadan, Ibadan, Nigeria
²Dipartimento di Farmacia, Universita di Pisa, Pisa, Italy

Correspondence: Sherifat Aboaba, Department of Chemistry, University of Ibadan, Ibadan, Nigeria.
E-mail: saboaba@gmail.com

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Abstract

Albizia adiantifolia (Schumach) W.F. Wright (Fabaceae) is a plant used traditionally in treating different health ailments which includes worm infections. The essential oils (EOs) were obtained by hydrodistillation in an all glass Clevenger apparatus, and characterized by gas chromatography (GC) and gas chromatography-mass spectrometry analysis (GC-MS). In vitro petri-dish anthelmintic activity was carried out using adult earthworm, Eudrilus eugeniae. The leaves, stem bark and root bark EOs afforded a total of 40, 26 and 20 constituents representing 90.9%, 94.1% and 90.9% of the total oil fractions respectively. The classes of compounds identified in the leaves, stem bark and root bark are oxygenated monoterpenes (4.1%, 1.7% and 4.0% respectively), sesquiterpene hydrocarbons (39.5%, 67.3% and 42.6% respectively), oxygenated sesquiterpenes (18.7%, 22.3% and 30.1% respectively), non-terpene derivatives (12.1%, 2.6% and 14.2% respectively) and apocarotenoids (16.5% and 0.2% in the leaves and stem-bark). β-caryophyllene (23.0%), E-geranylacetone (7.4%), acorenone (6.4%), viridiflorol (6.4%), α-zingiberene (6.3%) and ar-curcumene (4.6%) were the major constituents in the leaves oil, β-caryophyllene (39.3%), selin-11-en-4-α-ol (10.4%), α-zingiberene (9.6%), ar-curcumene (7.2%), carophyllene oxide (6.4%) and α-humulene (5.6%) were the major constituents in the stem bark oil, while β-caryophyllene (32.1%), selin-11-en-4-α-ol (13.1%), carophyllene oxide (8.4%), pentadecanal (6.1%) and α -humulene (4.4%) were the major constituents in the root bark oil. β -caryophyllene dominated the oils. The leaf EO was the most active against E. eugeniae worm. All the oils showed a relatively higher activity compared to Albendazole, in a concentration dependent manner. There was significant difference (p<0.001) in activity.

Keywords: Albizia adiantifolia, Clevenger apparatus, β -caryophyllene, Eudrilus eugeniae, Albendazole

1. Introduction

Albizia adiantifolia (Schumach) W.F. Wright is a large deciduous tree commonly known as “the West African Albizia or rough-bark flat-crown” in English (Orwa et al., 2009) and locally referred to as Ayinreta, igbabo (Yoruba) and kawo (Hausa) in Nigeria (Lawal et al., 2010). A. adiantifolia is a tree in the Fabaceae family distributed majorly from Senegal, Kenya, Angola, South Africa, Swaziland and extending to Eastern Madagascar. Domestically it has found use as firewood, furniture, as well as in vehicle body, cabinet works, and locally valued as a shade tree for some crops such as cocoa and coffee. It can also be used for soil improvement and conservation while the gum from the bark is used in cosmetics (Lemmens, 2007). In traditional medicine, various parts are used in treating different ailments such as toothache, bronchitis, diarrhoea, tapeworm infection (anthelmintic), abdominal pains, typhoid fever, urinary and respiratory tracts infections, Alzheimer’s disease and as an antidote against poison. It is also used as vermifuge, purgative, irregular menstruation and even administered to women in child labour (Orwa et al., 2009, Lawal et al., 2010, Lemmens, 2007, Tamokou et al., 2012, Beppet et al., 2014, Abubakar and Majinda, 2015). A. adiantifolia administered alone or in combination with Trichilia dregeana Sond., can be used to treat Gonorrhoea and Syphilis (De Wet et al., 2012).

Extracts from different parts of the plant possess activities such as antioxidant (Tamokou et al., 2012 and Beppet et al., 2014) anxiety, depression and oxidative stress activities (Beppet et al., 2015) antimicrobial, haemolytic, in vitro immunomodulatory, anti-inflammatory and anticholinesterase activities (Tamokou et al., 2012 and Abubakar and Majinda, 2015). Lupeol, aurantiamide, D-pinitol, protocatechuic acid as well as triterpenoidal saponins and several flavonoids which are reported to be contained in different parts, have been isolated from the plant (Tamokou et al., 2012 and Abubakar and Majinda, 2015).
The objective of this study is to extract and characterize the essential oils (EOs) from *A. adiantifolia* leaves, stem bark and root bark, and further determine their *in vitro* anthelmintic activity using *Eudrilus eugeniae* adult earthworm.

2. Materials and Methods

2.1 Sample Collection and Essential Oil Isolation

Samples from the leaves, stem bark and root bark of the plant were collected fresh from a forest vegetation at Awotan area in Ibadan, Oyo state, Nigeria in July 2013 and identified at the herbarium of Forest Research Institute of Nigeria (FRIN), Jericho Ibadan where voucher specimens were deposited with herbarium number FHI 109922. Pulverized leaves (300 g), stem bark (350 g) and root back (350 g) samples were used in an all-glass Clevenger apparatus designed according to British Pharmacopoeia specifications (MHRA, 1980) to obtain the essential oils by hydrodistillation method in 4 hours. The oils were dried over anhydrous sodium sulphate (Na2SO4) and stored inside the refrigerator at 4°C prior to use.

2.2 Gas Chromatography - Mass Spectrometry (GC-MS) Analysis

Gas Chromatographic (GC) analyses of the essential oils were performed on a HP-5890 Gas chromatograph equipped with a HP-Wax and HP-5 capillary columns (30 m x 0.25 mm, film thickness of 0.25 mm). The GC oven temperature which was programmed at 60 °C was held for 10 min and heated to 220 °C at 5 °C/min. The temperature for both the injector and the detector was maintained at 250 °C. The carrier gas used was Helium at a flow rate of 2 mL/min. The Gas Chromatographic-Mass Spectrometry (GC-MS) analyses were carried out on a Varian CP-3800 gas chromatograph interfaced to a Varian Saturn 2000 ion trap Mass Detector operated at 70 eV. The injector and transfer line temperatures were 220 °C and 240 °C, respectively. The GC oven temperature was programmed from 60 °C to 240 °C at 3 °C/min. Helium was used as a carrier gas at a flow rate of 1 mL/min. The constituents of the oils were identified on the basis of comparison of the retention times with those of the authentic samples, comparing their retention indices relative to the series of n-hydrocarbons, and by comparison of their mass spectra with published spectra and those of reference compounds from NIST, 2002. The relative concentration of each constituent was calculated by integration of GC peak areas (Adams, 2007).

2.3 Anthelmintic Assay

Preliminary evaluation of *in vitro* anthelmintic activity was carried out according to the method by Priya et al., 2012 with slight modifications. Indigenous in Africa, the adult earthworm (*Eudrilus eugeniae*) commonly known as the West African night crawler (Monebi and Ugwumba, 2013, Obohet al., 2007 and Dominguez et al., 2001) was used owing to its anatomical and physiological resemblance with the intestinal roundworm parasites of human beings (Lakshananam et al., 2011, Thomas and Devi, 2013 and Pueblos et al., 2015). These adult earthworms are common species in our region of study. Average sizes of the earthworms, 6 – 8 cm in length and 0.2-0.3 cm in width, were collected from moist soils and identified in the department of Zoology, University of Ibadan, Ibadan. They were washed with distilled water to remove any faecal matter. Albendazole, (ALBZ) (brand name – ZENTEL, by ©SmithKline Beecham Lab. Pharm.), often extensively administered clinically as an anthelmintic drug, was the test standard. EOs are relatively insoluble in water but were made soluble in Tween-80 (10% v/v in distilled water). The EOs and ALBZ were dissolved in 10 mL Tween-80 and diluted up to 30 mL to prepare five concentrations (1, 2, 3, 4 and 5% v/v for each of EO and ALBZ) in five petri-dishes. Five worms each were introduced into each concentration. 10% v/v Tween-80 in distilled water was used as negative control. Observations were made to determine the time for paralysis (when no movement could be observed except when shaken vigorously) and death (when worms neither wriggled when shaken nor moved when pinched with a needle, followed with fading away of body colour) of worms to have taken place. Activity was achieved by comparing results obtained for the EO extracts with that of the reference standard drug Albendazole.

Values for time of paralysis and death were expressed as mean ± standard error of mean (SEM). Analysis of variance (ANOVA), followed by student’s t test was carried out using Graph Pad Prism, version 5.01 statistical software to determine the significance of differences between experimental and control groups. At 95% confidence interval, p values < 0.001 were considered statistically significant.

3. Results and Discussion

The hydrodistilled leaves, stem bark and root bark essential oils (EOs) of *Albizia adiantifolia* obtained by Gas Chromatography (GC) and Gas Chromatography-Mass Spectrometry (GC-MS) analyses yielded 0.26%, 0.23% and 0.25% (v/w) respectively. As presented in table 1, a total of 40, 26 and 20 constituents representing 90.9%, 94.1% and 90.9% of the total oil fractions in the leaves, stem bark and root bark were identified. Based on the class of compounds present, complex mixtures in the leaves, stem-bark and root-bark were identified, such as oxygenated monoterpenes (4.1%, 1.7% and 4.0% respectively), sesquiterpene hydrocarbons – the most dominant class (39.5%, 67.3% and 42.6% respectively), oxygenated sesquiterpenes (18.7%, 22.3% and 30.1% respectively), non-terpene derivatives (12.1%, 2.6% and 14.2% respectively) and apocarotenoids (67.3% and 42.6%) in the leaves and stem-bark. Interestingly, none of the constituents present are in the class of monoterpene hydrocarbons.
The leaves essential oil constitutes majorly β-caryophyllene (23.0%), E-geranylacetone (7.4%), acorenene (6.4%), viridiflorol (6.4%), α-zingiberene (6.3%) and ar-curcumene (4.6%), while the stem bark oil is dominated by β-caryophyllene (39.3%), selin-11-en-4-α-ol (10.4%), -zingiberene (9.6%), ar-curcumene (7.2%), caryophyllene oxide (6.4%) and -humulene (5.6%). Also, the root bark oil contained majorly β-caryophyllene (32.1%), selin-11-en-4-α-ol (13.1%), caryophyllene oxide (8.4%), pentadecanal (6.1%) and α-humulene (4.4%). In the pattern of oil composition, -zingiberene and ar-curcumene were the dominant compounds in the leaves and stem bark, while selin-11-en-4-α-ol, caryophyllene oxide and α-humulene altogether dominated the stem bark and root bark oils. β-caryophyllene is the most abundant constituent in the three EOs. Other significant constituents include 1,8-cineole (2.2%, 1.5%, 1.9% in the leaves, stem bark and root bark respectively), β-sesquiphellandrene (2.5% and 2.8% in the leaves and stem bark respectively), valerianol (2.9% in the root bark) and δ-cadinene (3.8% in the root bark).

β-caryophyllene, the dominant constituent of Commiphora gileadensis EO (which is also one of the major oil constituents identified in this study), possesses many pharmacological properties such as anti-inflammatory, antifungal, local anesthetic and has been reported that the EO exhibited antiproliferative pro-apoptotic effects in tumor cells (Eitan et al., 2012). Also, it was identified as a dominant component of Garcinia mangostana Linn. leaves and stem bark EOs. G. mangostana oils exhibited high toxicity (LC50 values of 1.70 and 5.15 µg/mL, leaves and stem bark respectively) against brine shrimp (Artemia salina) and also showed antibacterial activities against some clinical isolates (Aboaba et al., 2014). Furthermore, β-caryophyllene and β-caryophyllene oxide are documented to occur in a large number of plants and possess significant anticancer activities by affecting growth and proliferation of numerous cancer cells (Fidy et al., 2016).

Murraya paniculata (L.) Jack, is a traditional medicinal plant for the treatment of abdominal pain, diarrhea, stomach ache, headache, edema, thrombosis, and blood stasis. Three of the major constituents (β-caryophyllene, α-zingiberene and selin-6-en-4-o-ol) identified in the EOs of M. Paniculata leaves are similar to the major constituents found in this study. The oil was reported to show moderate activity in the brine shrimp lethality test (LC50 = 41µg/mL) and a high nematicidal activity against Caenorhabditis elegans (LC50 = 37µg/mL) (Dosoky et al., 2016). Eugenol and β-caryophyllene dominated Ocimum sanctum Linn. In addition, the essential oil showed potent in vitro anthelmintic activity against the nematode C. elegans according to Mali and Mehta, 2008. Zingiberene is the dominant constituent of ginger. As a medicine, ginger essential oil and its oleoresin are used to aid digestion, as an expectorant and as a cure for stomach ache, toothache, diarrhea and asthmatic respiratory disorders (Kamaliroosta et al., 2013). Zingiberene has also been suggested as an important ingredient in stomachic medications (Malhotra and Singh 2003).

The result in table 2 shows the anthelmintic activity of A. adiantifolia essential oils. It was observed that the time of paralysis and death of worms decreases as concentration increases. The leaf essential oil shows activity higher than both the stem bark and root bark oils with time of paralysis and death at 12.60 and 60.20 minutes, respectively for the highest concentration. At all concentrations, the essential oils were more active than the standard drug (Albendazole) used. The effect of varying the concentrations of A. adiantifolia essential oils and albendazole were significantly different from one another. All values were also significantly different from the reference standard (p<0.001).

Many of the in vitro investigations on anthelmintic efficacy of plant extracts and their essential oils have been based on the effects they pose on organisms such as earthworms (Sutar et al., 2010; Akhtar et al., 2000; Bairagi et al., 2011; Mali and Mehta, 2008) as well as various gastro-intestinal nematodes (roundworms), cestodes (tapeworms) and nematodes (flukes) of human and livestock (Akhtar et al., 2000; Mali and Mehta, 2008; Ferreira et al., 2013). Due to the availability and easy access to earthworms, suitable models for anthelmintic drug screening have been established (Subash et al., 2012; Mali and Mehta, 2008). Many substances toxic to earthworms tend to cause irritation which eventually often lead to withdrawal of the worm from the environment of such substance, or cause flaccid paralysis. However, by virtue of these effects, very probably, anthelmintics would act by a way of expelling parasitic worms from their host’s gastrointestinal tract through peristalsis (Subash et al., 2012; Akhtar et al., 2000).

4. Conclusions

The compositional patterns and components of A. adiantifolia essential oils of Nigeria origin are reported for the first time. Sesquiterpene hydrocarbons dominated the leaves, stem bark and root bark essential oils. Result from the anthelmintic assay displayed the activity of A. adiantifolia oils against the worm used and their potency were inversely proportional to the time taken for paralysis and death of the worms to occur. The three essential oils show a relatively higher activity compared to Albendazole in concentration dependent manner which could be attributed to one or more components of the oil. This result could be a preliminary inference to the traditional medicinal usage of the plant in treating tapeworm infection, abdominal pains and as purgative. Also, owing to known activities demonstrated by some of the compounds identified in the oils, further suggests the potential pharmacological activities of the plant. Since the discovery and treatment of diseases with herbs has been on the increase, and the attention which their active principles have attracted as sources for new drugs, A. adiantifolia essential oils could be a good natural product for many pharmacological activities.
Table 1. Essential oil constituents of *A. adiantifolia* from GC/GC-MS analysis

| Constituents                                  | L.R.I | L.R.I* | AAL  | AASB  | AARB  |
|-----------------------------------------------|-------|--------|------|-------|-------|
| 2-heptanone                                    | 891   | 889    | 0.8  | -     | -     |
| Benzaldehyde                                   | 962   | 952    | -    | -     | 1.4   |
| 1-octen-3-one                                   | 980   | 972    | -    | -     | 0.9   |
| 6-methyl-5-hepten-2-one                        | 987   | 981    | 1.0  | -     | -     |
| 2-octanone                                     | 993   | 988    | 1.0  | -     | -     |
| Mesitylene                                     | 996   | 994    | -    | -     | 1.4   |
| 1,8-cineole                                    | 1034  | 1026   | 2.2  | 1.5   | 1.9   |
| Seudalone                                      | 1063  | -      | 0.4  | -     | -     |
| Linool                                         | 1101  | 1095   | 0.8  | -     | 2.1   |
| Nonanal                                        | 1104  | 1100   | 1.6  | -     | -     |
| 2,4-dimethylbenzaldehyde                       | 1179  | -      | 0.4  | -     | -     |
| Naphthalene                                    | 1181  | 1178   | 0.6  | -     | 1.1   |
| (Z,E)-undeca-1,3,5-triene                      | 1182  | -      | 0.5  | -     | -     |
| α-terpineol                                    | 1191  | 1186   | 0.4  | -     | -     |
| Safranal                                       | 1197  | 1196   | 0.7  | -     | -     |
| Decanal                                        | 1206  | 1201   | 0.4  | -     | -     |
| β-cyclocitral                                   | 1222  | 1217   | 1.4  | -     | -     |
| P-methyl-4-en-3-one                            | 1251  | -      | 0.7  | -     | -     |
| β-cyclocitralmethylalcohol                     | 1256  | -      | 0.7  | -     | -     |
| 5-methyltetralin                               | 1264  | -      | 0.5  | -     | -     |
| Thymol                                         | 1292  | 1289   | 0.2  | -     | -     |
| n-tridecane                                    | 1300  | 1300   | 0.5  | -     | -     |
| α-ionone                                       | 1352  | -      | 0.6  | -     | -     |
| Dehydro-ar-ionone                              | 1353  | -      | 0.5  | -     | -     |
| (E)-β-damascenone                              | 1382  | 1383   | 0.7  | -     | -     |
| 1,4-dimethyltetralin                          | 1391  | -      | 0.5  | -     | -     |
| β-elemene                                      | 1392  | 1389   | 0.5  | -     | -     |
| Cyperene                                       | 1398  | 1398   | -    | 2.3   | -     |
| n-tetradecane                                  | 1400  | 1400   | 1.3  | -     | -     |
| Isoaracylophylene                              | 1405  | 1408   | 0.4  | -     | -     |
| Italicene                                      | 1405  | 1405   | 0.7  | -     | -     |
| β-caryophyllene                                | 1419  | 1417   | 23.0 | 39.3  | 32.1  |
| (E)-α-ionone                                   | 1428  | 1428   | 0.5  | 0.2   | -     |
| cis-α-ambrinol                                 | 1437  | 1439   | 0.4  | -     | -     |
| 2-phenylethyl butanoate                       | 1440  | 1439   | -    | -     | 0.9   |
| α-humulene                                     | 1455  | 1452   | 2.4  | 5.6   | 4.4   |
| (E)-geranyl acetone                            | 1457  | 1453   | 7.4  | -     | -     |
| Sesquisabinene                                 | 1460  | 1457   | -    | 0.8   | -     |
| γ-murolene                                     | 1478  | 1478   | 0.3  | -     | -     |
| ar-curcumene                                   | 1483  | 1479   | 4.6  | 7.2   | -     |
| (E)-β-ionone                                   | 1487  | 1487   | 4.7  | -     | -     |
| α-zingiberene                                  | 1496  | 1493   | 6.3  | 9.6   | -     |
| α-bulnesene                                    | 1507  | 1509   | -    | 0.2   | -     |
| β-bisabolene                                   | 1508  | 1505   | 0.3  | -     | -     |
| trans-γ-cadinene                               | 1514  | 1513   | -    | 0.3   | -     |
| δ-cadinene                                     | 1524  | 1522   | -    | 3.8   | -     |
| β-sesquiphellandrene                           | 1525  | 1521   | 2.5  | 2.8   | -     |
| Occidental                                     | 1548  | 1550   | -    | 1.7   | -     |
| (E)-nerolidol                                  | 1564  | 1561   | 0.7  | 0.5   | -     |
| Caryophyllene oxide                            | 1582  | 1582   | 3.6  | 6.4   | 8.4   |
| Viridiflorol                                   | 1591  | 1592   | 6.4  | 2.7   | 3.0   |
| trans-β-elemene                                | 1601  | 1601   | -    | 0.3   | -     |
| Humulene epoxide II                            | 1607  | 1608   | -    | 0.5   | 1.0   |
| Tetradecanal                                   | 1614  | 1611   | -    | 0.8   | 1.0   |
| Caryophyllene-9(14),8(15)-dien-5-ol            | 1636  | 1639   | 0.8  | 0.8   | -     |
| T-cadinol                                      | 1641  | 1638   | 0.8  | 0.5   | -     |
| Selin-11-en-4-α-ol                             | 1655  | 1658   | -    | 10.4  | 13.1  |
| Valerianol                                     | 1656  | 1656   | -    | 2.9   | -     |
| Acorenone                                      | 1688  | 1692   | 6.4  | 0.2   | -     |
| 2-pentadecane                                  | 1699  | 1697   | -    | 1.4   | -     |
| Pentadecanal                                   | 1716  | -      | 1.5  | 1.8   | 6.1   |
| *Monoterpene hydrocarbons                     |       |        | 0.0  | 0.0   | 0.0   |
| *Oxigenated monoterpenes                      |       |        | 4.1  | 1.7   | 4.0   |
| *Sesquiterpene hydrocarbons                   |       |        | 39.5 | 67.3  | 42.6  |
| *Oxigenated sesquiterpenes                    |       |        | 18.7 | 22.3  | 30.1  |
| *Apocarotenoids                               |       |        | 16.5 | 0.2   | 0.0   |
| *Non-terpene derivatives                      |       |        | 12.1 | 2.6   | 14.2  |
| **Total identified**                          | **90.9** | **94.1** | **90.9** | **90.9** | **90.9** |

Major constituents are represented in bold

LRI = Linear retention index; LRI* = Linear retention index values from Adams, 2007; AAL = *Albizia adiantifolia* leaves; AASB = *Albizia adiantifolia* stem bark; AARB = *Albizia adiantifolia* root bark.
Table 2. Anthelmintic activity of *Albizia adiantifolia* essential oils

| EO Conc. (% v/v) | Time of Paralysis (Mins) expressed as Mean±SEM (N=5) | AAL | AASB | AARB | ALBZ |
|------------------|------------------------------------------------------|-----|-----|-----|-----|
| 1.00             | 30.40±1.36                                           | 32.80±2.01 | 34.00±1.64 | 97.20±1.39 |
| 2.00             | 25.80±1.71                                           | 27.20±1.43 | 28.60±1.60 | 94.20±1.77 |
| 3.00             | 20.20±1.28                                           | 24.40±1.50 | 24.80±1.71 | 89.60±1.29 |
| 4.00             | 17.00±1.73                                           | 18.20±1.28 | 20.00±1.22 | 87.40±1.08 |
| 5.00             | 12.60±1.21                                           | 15.60±0.93 | 16.60±1.08 | 82.80±1.28 |

| EO Conc. (% v/v) | Time of Death (Mins) expressed as Mean±SEM (N=5) | AAL | AASB | AARB | ALBZ |
|------------------|---------------------------------------------------|-----|-----|-----|-----|
| 1.00             | 96.20±3.51                                          | 98.20±3.22 | 101.40±3.97 | 154.60±1.86 |
| 2.00             | 85.60±2.66                                          | 89.80±2.87 | 98.00±3.18 | 149.20±2.35 |
| 3.00             | 78.60±2.54                                          | 80.80±2.60 | 87.40±3.36 | 140.60±1.72 |
| 4.00             | 70.40±3.43                                          | 72.80±2.96 | 76.80±2.65 | 135.00±1.92 |
| 5.00             | 60.20±3.09                                          | 61.20±2.73 | 69.60±2.94 | 130.20±1.77 |

AAL = *Albizia adiantifolia* leaves; AASB = *Albizia adiantifolia* stem bark; AARB = *Albizia adiantifolia* root bark; ALBZ = Albendazole (Standard); SEM = Standard error of mean; N = number of worms in each petri-dish.

The time taken for paralysis and death of worms to occur in distilled water (negative control) was observed to be >> 200 minutes. This is due to the body cells eventually absorbing water by osmosis.

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