Investigation of Genetic Causes in Patients with Congenital Heart Disease in Qatar: Findings from the Sidra Cardiac Registry

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Abstract: Congenital heart disease (CHD) is one of the most common forms of birth defects worldwide, with a prevalence of 1–2% in newborns. CHD is a multifactorial disease partially caused by genetic defects, including chromosomal abnormalities and single gene mutations. Here, we describe the Sidra Cardiac Registry, which includes 52 families and a total of 178 individuals, and investigate the genetic etiology of CHD in Qatar. We reviewed the results of genetic tests conducted in patients as part of their clinical evaluation, including chromosomal testing. We also performed whole exome sequencing (WES) to identify potential causal variants. Sixteen patients with CHD had chromosomal abnormalities that explained their complex CHD phenotype, including six patients with trisomy 21. Moreover, using exome analysis, we identified potential CHD variants in 24 patients, revealing 65 potential variants in 56 genes. Four variants were classified as pathogenic/likely pathogenic based on the American College of Medical Genetics and Genomics and the Association for Molecular Pathology (ACMG/AMP) classification; these variants were detected in four patients. This study sheds light on several potential genetic variants contributing to the development of CHD. Additional functional studies are needed to better understand the role of the identified variants in the pathogenesis of CHD.

Keywords: congenital heart defect; Qatar; genetic investigation; whole exome sequencing; single-nucleotide variant; chromosomal abnormalities

1. Introduction

Congenital heart disease (CHD) is one of the most common birth defects worldwide, with a general prevalence of 1–2% in newborns [1]. The prevalence of specific manifestations of CHD varies significantly in different populations, and their genetic etiology is incredibly complex [2]. For example, sporadic cases account for 5–10% of all cases of CHD with a
genetic component, and only a small fraction of CHD cases is described as familial [2]. Moreover, isolated cases of CHD are more likely to be caused by mutations in single genes compared to chromosomal aberrations. To date, ~400 genes have been shown to be associated with this disease, but ~60% of CHD cases remain unexplained.

Epidemiological studies show a higher incidence of CHD in Arab countries due to high consanguinity and endogamy [3,4]. For instance, it was reported that the incidence of CHD in the province of Al Qassim in Saudi Arabia doubled compared to that in the United States of America (USA) [5]. The annual incidence of CHD among live births is estimated to be ~9–15/1000 in Qatar, ~12/1000 in Jordan, ~11/1000 in Lebanon, ~7/1000 in Oman, and ~10/1000 in Gaza city in Palestine [6–10]. In Egypt, consanguinity was reported in 44% of CHD cases, whereas it was around 30.5% in Tunisia [11,12]. Moreover, in 2015, 65% of fetuses affected by CHD in the United Arab Emirates came from consanguineous families [4].

Here, we established the Sidra Cardiac Registry in Qatar to collect research data and samples from patients with CHD, cardiomyopathies, and other cardiac phenotypes, as well as from their relatives. We also studied the genetic architecture of 52 patients with CHD, identifying the potential variants and genes that may explain their clinical phenotype. Our findings may help shed light on the possible increased prevalence of hereditary forms of CHD in Arabs and the importance of genetic research among the Arab and Middle Eastern populations, which remain insufficiently studied.

2. Materials and Methods

2.1. Ethical Approval

Ethical approval was obtained from the Institutional Review Boards of Sidra Medicine (IRB #1500769) and Qatar University (IRB #1666-E/22) in accordance with the Declaration of Helsinki. Informed consent/assent was obtained from all study participants. The criteria for inclusion in the Sidra Cardiac Registry were patients from Sidra Medicine with cardiac phenotypes (e.g., CHD, cardiomyopathy, channelopathy), first- and second-degree relatives of patients with cardiac phenotypes, Qatari and non-Qatari participants, as well as children and adults. Eligible participants who could not provide informed consent/assent due to medical reasons or language barriers were excluded.

2.2. Study Participants and Data Collection

The study included 52 patients with CHD enrolled in the Sidra Cardiac Registry at Sidra Medicine in Qatar, in addition to 126 family members (2 affected, 124 unaffected). The classification of patients’ congenital malformations corresponded to the nomenclature of the International Classification of Diseases, Tenth Revision (ICD-10) [13]. Demographic, clinical, and family history data were collected from the study participants along with 1–10 mL of whole blood, which was required for genetic investigation. Family pedigrees were generated using HaploPainter (http://haplopainter.sourceforge.net/) (accessed on 1 January 2022). Data about cytogenetic tests, such as karyotyping, chromosomal microarray, Fluorescence In Situ Hybridization (FISH), performed as part of a patient’s evaluation, were obtained along with the pathogenicity interpretation report that was provided by the laboratory where the test was performed.

2.3. Whole Exome Sequencing (WES)

Patients who did not undergo chromosomal testing (n = 25) and patients who had a negative result for chromosomal abnormalities (n = 11) were included in the whole exome sequencing (WES) analysis together with their family members who were enrolled in the study (n = 77). Genomic DNA was extracted from blood, and WES libraries were prepared using the SureSelect kit (Agilent, Santa Clara, CA, USA), followed by sequencing using a HiSeq 4000 (Illumina, San Diego, CA, USA) at Integrated Genomic Services, Sidra Medicine.
2.4. Sequence Quality, Alignment, and Variant Calling

The WES pipeline involved the following: multiplexed raw sequencing files (.bcl files) were converted into separate FastQ files, and the quality of the sequencing reads was assessed using FASTQC [14]. Adaptor sequences were trimmed using trimadap (https://github.com/lh3/trimadap) (accessed on 1 January 2022) and high-quality reads were mapped to the National Center for Biotechnology Information (NCBI) human reference genome GRCh37/hg19 (National Institute of Health, New York, USA) using Burrows-Wheeler Aligner (BWA) v0.7.8 (Broad Institute, Cambridge, Massachusetts, USA), in particular, the BWA-MEM algorithm (arXiv:1303.3997[q-bio-GN]). Duplicate reads were marked after alignment using SAMBLASTER [15], and BAM files were sorted using SAMtools [16]. These steps were accomplished using the BWA-kit package. Picard and Mosdepth tools were used to perform the alignment quality control. After alignment, the reads were recalibrated, and variants were called using the Genome Analysis Toolkit (GATK) v4.1 [17]. The Exome-Agilent-V6.bed file was fed into the variant calling step to limit the variant calling to exome regions. The individual g.VCF files were joined to generate a single merged VCF file for all samples. We also ran peddy on the merged VCF file to ensure the pedigree assignment was correct before performing segregation analysis. The joined VCF file was split, decomposed, and normalized using VT software v0.57 (University of Michigan, Michigan, USA) [18]. The joint normalized VCF file was annotated using Snpeff v4.3 (http://pcingola.github.io/SnpEff/) (accessed on 1 January 2022) [19], classifying variants into high, moderate, and low impact, based on their predicted pathogenic effect on the gene [19]. Variants were also annotated with the following:

(a) Allele frequencies, obtained from the Genome Aggregation Database (gnomAD) (https://gnomad.broadinstitute.org/) (accessed on 1 January 2022), Qatar 1000 Genomes [20,21], and Greater Middle East (GME) Variome Project (http://igm.ucsd.edu/gme/) (accessed on 1 January 2022) [22].

(b) Variant pathogenicity prediction scores, including Sorting Intolerant From Tolerant (SIFT), Polymorphism Phenotyping (PolyPhen), and Combined Annotation Dependent Depletion (CADD).

(c) Variant phenotypic and abnormality-related information using the available databases, mainly the Human Gene Mutation Database (HGMD®), ClinVar (https://www.ncbi.nlm.nih.gov/clinvar/) (accessed on 1 January 2022) and Genome-Wide Association Study (GWAS) Catalog.

(d) Information on the phenotypic abnormalities of the captured genes, obtained from Human Phenotype Ontology (HPO|) (https://hpo.jax.org/app/) (accessed on 1 January 2022) [23].

2.5. Single-Nucleotide Variant (SNV) Segregation Analysis and Filtration

Familial segregation analysis was performed using genome mining (Gemini) v0.30.2 (https://gemini.readthedocs.io/en/latest/) (accessed on 1 January 2022). The final annotated VCF file was imported into the Gemini database along with the pedigree (PED) file using vcftools utilities. Gemini provided various segregation patterns (i.e., de novo, autosomal dominant, autosomal recessive, compound heterozygous, X-linked recessive, and X-linked dominant), and a custom Python script was used to write the Gemini output data to Excel sheets. Then, the obtained variants were filtered by removing those with a CADD value of <13, minor allele frequency (MAF) of >1% in the control databases (gnomAD, Qatar 1000 genome, GME), and low impact according to the SnpEff annotation [24] (Figure 1).

Prioritization of the genes was conducted by searching PubMed (https://pubmed.ncbi.nlm.nih.gov/), Google (http://google.com), and Google Scholar (https://scholar.google.com/) (accessed on 1 February to March 2022). For search queries, we combined the gene code AND “cardiac” OR “heart” OR “cardiac defect” OR “congenital cardiac defect” OR “malformation” OR “defect”. We extracted data on the molecular role of the gene in cardiogenesis or cardiac pathology, in vitro or in vivo experiments, as well as case reports of an association between the gene or its variant and CHD.
Whole Exome Analysis approach

Figure 1. A flowchart of the whole exome sequencing analysis (WES) approach performed in this study.

The American College of Medical Genetics and Genomics and the Association for Molecular Pathology (ACMG/AMP) guidelines [25] were used to classify variants into “benign”, “likely benign”, “uncertain significance”, “likely pathogenic”, and “pathogenic” using the InterVar online tool (https://wintervar.wglab.org/) (accessed on 1 February to 1 March 2022). The novelty of the variant was established if it had not been reported in the literature or publicly available databases.

2.6. Statistical Analysis

Fisher’s exact tests were used to examine the association between obtaining a positive result in chromosomal testing or WES analysis and the demographic data of patients. Correlations were considered significant if the $p$-value was $<0.05$ across the demographic data (i.e., survival, ethnicity, gender, family history of CHD, family history of congenital disease, and consanguinity). Statistical tests were performed separately for patients who underwent chromosomal testing ($n = 27$) and for patients included in the WES analysis ($n = 52$). All statistical analyses were performed using IBM-SPSS v.28.

3. Results

3.1. Demographic and Clinical Characteristics

The Sidra Cardiac Registry included 52 patients suffering from CHD, as well as a total of 126 family members. While 43 out of 52 families contained at least a trio (i.e., a proband and both parents), the remaining 9 had a proband with only a mother or father. Of all patients, 61.50% ($n = 32$) were Arab from Qatar, Syria, Yemen, Tunisia, Morocco, Egypt, Sudan, Lebanon, and Kuwait, followed by 36.50% ($n = 19$) South Asian from India, Pakistan, Sri Lanka, and Bangladesh, and one European from Serbia. Of our patients, 48% ($n = 25$) were females and 52% ($n = 27$) were males. All 52 patients belonged to the pediatric population with the mean age of 1 year and 2 months ($SD = \pm 3.6$). The maximum age of...
patients was 18 years, while the minimum was 2 days. The consanguinity rate was 2.30% (n = 22). Of 52 patients with CHD, 44.20% (n = 23) had several cardiac defects. In addition, 32.70% (n = 17) had a combination of several CHD types (Figure 3).

Eleven types of CHD phenotypes were identified in 52 patients, distributed as follows: 19.23% (n = 10) of cases presented with tetralogy of Fallot (TOF), 17.30% (n = 9) had septal defects (arterial septal defect, ventricular septal defect), 7.69% (n = 4) had hypoplastic left heart syndrome (HLHS), 9.61% (n = 5) had transposition of the great arteries (TGA), 1.92% (n = 1) had valve stenosis (pulmonary valve stenosis), 3.84% (n = 2) had coarctation of the aorta (COA), 1.92% (n = 1) had arterial tortuosity syndrome (ATS), 1.92% (n = 1) had double outlet right ventricle (DORV), 1.92% (n = 1) had polyvalvular disease, 1.92% (n = 1) had total anomalous pulmonary venous return (TAPVR), and 32.69% (n = 17) had a combination of several CHD types (Figure 3).
Genes 2022, 13, x FOR PEER REVIEW... tion, two patients (cardio-32.A and cardio-37.A) presented with two different chromosomal aberrations: one patient had a deletion of chromosome 6, 16p11.2 deletion, and 8p11.21 duplication. In addition, two patients (cardio-32.A and cardio-37.A) presented with two different chromosomal aberrations: one patient had a deletion of Adam Oliver Syndrome, mosaic trisomy 14, chromosome 6 deletion, 16p11.2 deletion, and 8p11.21 duplication. In addition, two patients (cardio-32.A and cardio-37.A)

Figure 3. Cardiac phenotypes reported in patients with CHD (n = 52).

3.2. Clinical Cytogenetic Testing

As part of the clinical evaluation, 27 out of 52 patients (52%) with CHD underwent cytogenetic testing for the presence of chromosomal abnormalities, and 16 out of 27 patients (59.2%) had positive results for chromosomal abnormalities (Figure 4). No significant relationship was found between the demographic characteristics studied in 27 patients and chromosomal abnormalities (p > 0.05).

Figure 4. A flowchart illustrating genetic investigations conducted in patients with CHD and the outcomes obtained from chromosomal testing and whole exome sequencing analysis. LB: likely benign; LP: likely pathogenic; P: pathogenic; US: uncertain significance. wo variants could not be classified due to limited data availability.

Six patients had trisomy 21 (Down Syndrome), three had a 22q11.2 deletion (DiGeorge Syndrome), two had a 7q11.23 deletion (William Syndrome), and five had other chromosomal abnormalities (Table 1, Table S1). Additional chromosomal abnormalities were a 9q34.3 deletion (Adam Oliver Syndrome), mosaic trisomy 14, chromosome 6 deletion, 16p11.2 deletion, and 8p11.21 duplication. In addition, two patients (cardio-32.A and cardio-37.A) presented with two different chromosomal aberrations: one patient had a deletion of
10q26.3 and 7q11.23, and the other had a 22q11.2 deletion with mosaic trisomy 8 (Table S1). In all patients, extracardiac phenotypes were identified along with the cardiac phenotype, as shown in Table S1. The clinical laboratory provided interpretation of all chromosomal aberrations, with the exception of cardio-3.A (chromosome 6 deletion) and cardio-45.A (8p11.21 duplication), where no interpretation was provided (Table S1).

Parental testing was performed in seven patients with positive results for chromosomal abnormalities. Three out of seven patients had negative results for any aberrations in both parents (Table S1). In three cases, the mother had normal chromosomal testing results, while the father either had no records, was not enrolled, or had inconclusive results. In one case, there was the same copy number variation (CNV) (16p11.2 deletion) in the father (cardio-56.C) as in the proband (cardio-9.A), and the aberration was classified as likely benign (Table S1). All patients with positive results for chromosomal abnormalities (n = 16) had syndromic CHD with extracardiac manifestations, and 62.50% (n = 10) of the 16 patients manifested with septal defects.
Table 1. Results of the chromosomal testing conducted in 27 patients with CHD as part of their clinical evaluation.

| Patient | Cardiac Phenotype | Extracardiac Phenotype | Chromosomal Abnormality | Genes Encompassed | Associated Condition | Interpretation of Test Results | Parental Testing |
|---------|------------------|------------------------|-------------------------|-------------------|---------------------|-------------------------------|-----------------|
| Cardio-1.A | ASD, VSD | Hypotonia, dysmorphic features, developmental delay | 47, XY, +21 | Gain of one full copy of chromosome 21 | Down Syndrome | Pathogenic | No record |
| Cardio-3.A | TGV | Vertebral abnormalities, anal atresia, cardiac abnormalities, tracheoesophageal fistula, renal anomalies, limb defects | Chromosome 6 deletion * | Unknown | Unknown | Unknown | No record |
| Cardio-4.A | TOF, PA | Thymus hypoplasia, compromised immune system, absent left kidney, idiopathic left club foot, global developmental delay with central hypotonia | 22q11.2 deletion | 40 genes (TBX1 and COMT) | DiGeorge Syndrome | Pathogenic | Mother is normal; father has no records |
| Cardio-6.A | VSA | Congenital nasolacrimal duct obstruction, esotropia, failure to thrive, hypothyroidism, supraventricular tachycardia, thrombocytopenia | 22q11.2 deletion | 40 genes (TBX1 and COMT) | DiGeorge Syndrome | Pathogenic | No records |
| Cardio-9.A | ASD, DORV, PA, Hypoplastic mitral valve and left ventricle | Hypotension, acidosis, bradycardia, severe developmental delay, seizures | 9q34.3 deletion | SNAPC4, PMPCA, INPP5E, SEC16A, NOTCH1 | Adams Oliver Syndrome | Likely Pathogenic | Mother is normal; father is inconclusive |
| Cardio-28.A | TOF | Acute renal failure, fluid overload, skin pigmentation, undescended testicles | Mosaic 47, XY, +14 | Gain of one full copy of chromosome 14 in some somatic cells | Mosaic trisomy 14 | Pathogenic | Parents are normal |
| Patient   | Cardiac Phenotype | Extracardiac Phenotype                                                                 | Chromosomal Abnormality | Genes Encompassed                                      | Associated Condition          | Interpretation of Test Results | Parental Testing |
|-----------|-------------------|---------------------------------------------------------------------------------------|-------------------------|-------------------------------------------------------|------------------------------|-------------------------------|------------------|
| Cardio-30.A | TOF               | Bilateral hydronephrosis, hypotonia, dysmorphic features                               | 47, XY, +21             | Gain of one full copy of chromosome 21                | Down Syndrome                | Pathogenic                    | No records       |
| Cardio-32.A | VSD               | Dysphagia, gastroesophageal reflux disease, failure to thrive, global developmental delay, hypotonia, central sleep apnea, right ankle contracture, asymmetric leg length | 7q11.23 deletion, 10q26.3 deletion, | ELN, LIMK1, BAZ1B, CLIP2, GTF2IRD, NSUN5, CLDN4, EIF4H, LAT2, MLXIPL, TBL2, WBSCR18, WBSCR22, WBSCR27 — Not defined for 10q26.3 deletion | William Syndrome and 10q26.3 deletion | Pathogenic                    | Parents are normal |
| Cardio-33.A | ASD               | Polydactyly, dysmorphic features, delayed motor development                            | 47, XY, +21             | Gain of one full copy of chromosome 21                | Down Syndrome                | Pathogenic                    | Parents are normal |
| Cardio-37.A | TOF, PS           | Failure to thrive, hypothyroidism                                                     | 47, XY, +8 22q11.21 deletion | Gain of one full copy of chromosome 8                | DiGeorge syndrome and mosaic trisomy 8 | Pathogenic                    | No records       |
| Cardio-39.A | VSD               | Dysmorphic features, hypotonia                                                        | 47, XY, +21             | Gain of one full copy of chromosome 21                | Down Syndrome                | Pathogenic                    | No records       |
| Cardio-44.A | AS, PS            | Anal stenosis, dysmorphic features                                                    | 7q11.23 deletion                             | ELN, LIMK1, BAZ1B, CLIP2, GTF2IRD, NSUN5, CLDN4, EIF4H, LAT2, MLXIPL, TBL2, WBSCR18, WBSCR22, WBSCR27  | William Syndrome             | Pathogenic                    | No records       |
| Cardio-45.A | HLHS              | Depressed nasal bridge, developmental delay                                            | 8p11.21 duplication                      | Not reported                                         | Not specified                | Unknown                       | Mother is normal |
| Cardio-56.A | ASD, PA, TAPVD, Heterotaxy | Bowel obstruction, osteomyelitis, asplenia                                           | 16p11.2 deletion                     | SH2B1 gene                                           | Not specified                | Likely benign                 | Mother is normal; father has the same deletion |
**Table 1. Cont.**

| Patient  | Cardiac Phenotype | Extracardiac Phenotype                                | Chromosomal Abnormality | Genes Encompassed | Associated Condition | Interpretation of Test Results | Parental Testing |
|----------|-------------------|-----------------------------------------------------|-------------------------|-------------------|----------------------|-----------------------------|------------------|
| Cardio-60.A | ASD              | Laryngomalacia, swallowing dysfunction             | 47, XX, +21           | Gain of one full copy of chromosome 21 | Down Syndrome         | Pathogenic                  | No records       |
| Cardio-62.A | ASD              | Hypotonia, dysmorphic features                     | 47, XY, +21           | Gain of one full copy of chromosome 21 | Down Syndrome         | Pathogenic                  | No records       |

ASD: atrial septal defect; DORV: double outlet right ventricle; HLHS: hypoplastic left heart syndrome; PA: pulmonary atresia; PS: pulmonary stenosis; TAPVD: total anomalous pulmonary venous drainage; TGV: transposition of the great arteries; TOF: tetralogy of Fallot; VSD: ventricular septal defect. *: the detailed laboratory report is not available for this patient.
3.3. Variants Identified by Whole Exome Sequencing Analysis

Thirty-six patients with CHD, along with 77 family members, were included in the WES analysis, conducted to identify potential gene variants that could cause the disease (Figure 4). Of the 36 families, 28 were trios and included a proband and both parents; three included a proband, both parents, and at least one sibling; four included a proband and only one parent; and one included a proband, one parent, and proband siblings. Across 24 patients, the mean depth of coverage for all genes was 74.31X (SD = ±15.4).

Initially, 5606 variants were detected, while only 65 remained after we applied a series of filtrations (Figure 4). These 65 variants were detected in 56 genes associated with CHD pathogenesis, cardiogenesis, or cardiac pathology; they were present in 24 out of 36 patients. In the remaining 12 patients with CHD, no variants within the prioritized genes were detected.

Based on the ACMG/AMP guidelines, 51 variants (78.46%) were classified as uncertain significant (US), eight (12.30%) were likely benign, two (3.07%) were pathogenic, two (3.07%) were likely pathogenic, and two (3.07%) variants (c.1437-6delT and c.4154-7dupT) could not be classified due to limited data availability (Figure S1). Of the 65 variants, 23 were novel variants that had not been previously identified or reported; they were located in 21 different genes (Table S2).

Our segregation analysis using Gemini showed that 12 (18.46%) variants followed an autosomal recessive inheritance pattern, 2 (3.07%) were autosomal dominant, 20 (30.76%) were compound heterozygous, 2 (3.07%) were X-linked recessive, and 1 (1.53%) showed a double-inheritance autosomal dominant and compound heterozygous pattern. One variant (1.53%) followed uniparental disomy (UPD), eighteen (27.69%) were de novo, and nine (13.84%) could not be attributed to a certain inheritance pattern, as the parents were not enrolled in the study (Figure S1).

Of the 65 variants, 46 were found to be inherited: 16 variants (autosomal dominant, compound heterozygous, or X-linked recessive) were inherited from the mother, 9 were inherited from the father (autosomal dominant or compound heterozygous), and 4 were inherited from both parents (autosomal recessive). In 17 cases, the origin of the variant could not be accurately determined, as one of the parents was not enrolled in the study. Therefore, the origin of the variant in those 17 cases could have been either autosomal dominant or compound heterozygous (Table S4). We identified four pathogenic or likely pathogenic variants in four patients. A patient from Yemen (cardio-27.A), who presented with syndromic CHD (TOF, long QT interval, and hearing loss), had a pathogenic de novo variant c.6292C>T in the CHD7 gene. A patient from Sri Lanka (cardio-12.A), who had multiple cardiac defects, including COA, VSD, and PDA (Table 2), was found to have a pathogenic de novo variant c.2883-1G>T in the ROBO1 gene. In addition, a patient from Pakistan (cardio-15.A) with Shone’s complex had a likely pathogenic variant c.817+1G>C in the SMAD6 gene, and a patient from Yemen (cardio-15.A) with ATS had a possible pathogenic variant c.243C>G in the SLC2A10 gene (Tables 2 and S4).

We also identified eight likely benign variants in 20.8% (5/24) of patients, which were inherited from either one (n = 3) or both parents (n = 5) (Table S4). Compound heterozygous variants, such as c.98390A>G in TTN, were detected in a patient from Qatar (cardio-2.A) with multiple cardiac defects and an extracardiac phenotype. Other examples were c.3401G>T in IFT172, seen in a patient from Sudan (cardio-36.A) with TOF, and c.2867G>C in COL5A2, seen in a patient with HLHS from Sudan (cardio-31.A). Moreover, we identified two patients, cardio-15.A from Pakistan and cardio-18.A from Qatar, who were diagnosed with multiple cardiac defects. Those patients had homozygous likely benign variants such as p.Val1457Ile in CCDC141 and c.6035C>T and c.1751A>C in CMYAS, respectively. HLHS was present in two patients from Sudan (cardio-31.A and cardio-45.A), with likely benign variants. Three variants were detected in cardio-31.A, including two homozygous variants in DNAAF3 (c.1606G>A and c.131G>T) and one variant c.2867G>C in COL5A2.
### Table 2. Pathogenic/likely pathogenic variants detected in CHD patients.

| Gene    | Amino Acid Change | Nucleotide Change | Variant Type | Variant Impact | Familial Segregation | Zygosity | Inheritance | ACMG/AMP Classification | CHD Phenotype | Extra Phenotype | Patient       |
|---------|-------------------|-------------------|--------------|----------------|----------------------|----------|-------------|--------------------------|---------------|-----------------|---------------|
| ROBO1   | -                 | c.2883-1G>T SNP   | splice acceptor | de novo        | Heterozygous         | None     | Pathogenic  | Multiple CHD             | CHD           | None            | Cardio-12.A   |
| SMAD6   | -                 | c.817+1G>C SNP    | splice site donor | de novo     | Heterozygous         | None     | Likely pathogenic | Shone’s complex           | None          | Cardio-15.A     |
| SLC2A10 | p.Ser81Arg        | c.243C>G SNP      | Missense     | AR             | Homozygous           | Both parents | Likely pathogenic | ATS                  | None          | Cardio-5.A     |
| CHD7    | p.Arg2098 *       | c.6292C>T SNP     | stop gained  | de novo        | Heterozygous         | None     | Pathogenic  | TOF                   | Prolonged QT interval, hearing loss | Cardio-27.A   |

AR: autosomal recessive; ATS: arterial tortuosity syndrome; CHD: congenital heart disease; TOF: tetralogy of Fallot. * means that the amino acid Arginine2098 is predicted to change to a stop codon.
Moreover, we identified 51 different variants classified as US. Most of these variants had been reported previously (n = 30), while others were novel (n = 21) (Table S4). Among the novel US variants, 8 were de novo: c.*16G>A in CCNC, c.49G>C in GGN, c.2054C>T in NUP133, c.123C>G in FOXP1, c.584G>A in LRRC14B, c.343G>A in HIST1H1E, c.2957G>T in ZFPM2, and c.1817A>C in FLTL1. Other novel US variants were inherited from the mother in an X-linked recessive pattern: c.10570T>A in HUWE1 and c.463C>T in TBC1D25, detected in patients cardio-12.A and cardio-23.A (Table S4). Other novel US variants were passed on to a patient from only one parent in an autosomal dominant or compound heterozygous inheritance pattern: c.848G>C in CRELD1, c.2204T>A in ADAMTS8, and c.66A>T in SLC24A4 (Table S4). The presence of 30 US variants identified here had been previously reported by other studies or identified in either control or clinical databases. Nine US variants had been reported among patients with other diseases, for example c.5695G>A in DYNC2H1 (Table S3). The remaining 25 US variants have been reported in control databases at a low frequency but have not been found in the published literature, for example c.5437G>A in PRR12 (Table S3).

4. Discussion

In this study, we established the Sidra Cardiac Registry in Qatar and investigated the genetic etiology of 52 patients with CHD and their 126 relatives. As for chromosomal testing, out of 52 patients, 30.7% (n = 16) had confirmed chromosomal abnormalities. In previous studies, it was estimated that 11–13% of CHD cases are associated with chromosomal abnormalities [26–28]. Our prevalence rates appear to be higher than the reported prevalence; however, our results may not be representative of the general population due to the small sample size. In 6 of the 16 positive cases (37%), trisomy 21 was detected, with septal defects being the most prominent cardiac finding. These results support former studies that have identified septal defects as the most common form of cardiac abnormality in patients with trisomy 21 [29,30]. Other chromosomal aberrations encountered in our sample were William syndrome and DiGeorge syndrome, both of which have a strong association with CHD [31,32]. Two patients with 22q11.2 deletion had conotruncal heart defects (TOF) along with valve stenosis; however, in one of these cases (cardio-37.A), another aberration was encountered, which was a mosaic nondisjunction in chromosome 8. Previous studies have reported conotruncal defects, such as TOF and DORV, often present in patients with 22q11.2 deletion.

Although discussion of the cardiac phenotype of mosaic trisomy 8 is limited due to a paucity of published data, cardiovascular [33], laterality [34], and septal [35] defects have been reported in several cases. Mosaic trisomy 14 is another chromosomal aberration that was observed in a patient with syndromic CHD (cardio-28.A); it is typically associated with cardiac defects [36], such as patent ductus arteriosus [37], and TOF [38,39]. However, CHD does not seem to always co-occur with trisomy 14. In addition, we encountered four cases with CNVs, such as a 9q34.3 deletion in cardio-9.A, who had multiple cardiac defects along with extracardiac manifestations. The deleted region houses a group of genes, such as NOTCH1, associated with Adams–Oliver syndrome. This condition is often accompanied by cardiac malformations, including septal defects, vascular defects, and TOF [40,41]. Deletion of 10q26.3 and chromosome 6 was observed in two patients (cardio-32.A and cardio-3.A) with septal defects and TGA, respectively. 10q26 deletion syndrome has been reported in cases with septal defects [42] and patent ductus arteriosus [43]. In total, 16 patients with CHD had chromosomal abnormalities, mainly as part of a syndrome with multiple systemic and developmental comorbidities. Most had trisomy 21, a 7q11.23 deletion, and a 22q11.2 deletion, while rare CNVs were less common.

From the WES analysis, we identified four pathogenic or likely pathogenic variants in five different genes in six patients with CHD. A de novo pathogenic variant c.2883-
Variant c.243C>G in SLC2A10 was found in a homozygous state in a patient from Yemen coming from a consanguineous family. The variant c.243C>G has been previously identified in a homozygous state in 15 unrelated patients from Qatar belonging to Bedouin tribes with the same clinical manifestation of ATS [46], as well as in several Mediterranean families [47]. The variant c.243C>G has not been detected in the gnomAD or GME datasets; however, it has been detected in the Qatar 1000 genome project, with a MAF of 0.007. To our knowledge, this is the first time this variant is encountered in a patient from Yemen. Given its recessive pattern of inheritance and the high consanguinity rate among Arabs compared to other populations [3], it is possible that this variant is more prevalent in Arabs and is a founder mutation that causes disease in homozygosity.

Both variants c.817+1G>C in SMAD6 and c.6292C>T in CHD7 were likely pathogenic and were identified in a heterozygous state in cardio-15.A and cardio-27.A, respectively. It has been reported that SMAD6 plays a role in the cardiac development in chick models, where the induction of SMAD6 and Nkx2-5 contributed to the pathogenesis of CHD. Nonsynonymous variants in SMAD6 have been observed in cardiovascular malformations, such as a bicuspid aortic valve with mild aortic stenosis and aortic coarctation [48]. Considering the supposed role of the gene in cardiovascular malformation, it is possible that c.817+1G>C in SMAD6 had influenced the manifestation of Shone’s complex in our patient.

Variant c.6292C>T in CHD7 was detected in cardio-27.A, who had syndromic CHD. Variants of loss of function in CHD7 are associated with CHARGE syndrome (OMIM# 214800), a combination of multiple congenital malformations in which 70–92% of cases have cardiac defects [49]. Variable expressivity has been observed in patients with positive CHD7 variants, where patients did not meet the diagnostic criteria for CHARGE syndrome [50]. This finding highlights the difficulty in establishing a clear genotype–phenotype correlation among patients positive for CHD7 variants, as well as the possibility of missing CHD7-positive cases who fail to meet the diagnostic criteria for CHARGE syndrome.

We detected eight variants classified as likely benign in five patients with CHD. Five variants followed an autosomal recessive pattern of inheritance; they were detected in a homozygous state in three patients (cardio-15.A, cardio-18.A, and cardio-31.A) coming from consanguineous families. Variant p.Val1457Ile in CCDC141 was observed in one patient (cardio-15.A) with Shone’s complex, who also had a pathogenic variant in SMAD6. Variants in CCDC141 have been linked to cardiovascular malformations and were observed to play a role in the cardiac development in zebrafish [51]. Variants c.6035C>T and c.1751A>C were present in the CMYA5 gene in one patient (cardio-18.A). Both variants were seen in homozygosity and were inherited from both parents. These variants were also detected in unaffected siblings, but in a heterozygous state. It was noted that all the likely benign variants found in our cohort had a higher MAF in the GME and Qatar 1000 genome control databases compared to the gnomAD database. This may be influenced by the high consanguinity rate in the Middle Eastern population, considering the fact that five
variants followed a recessive pattern of inheritance. Consequently, there is a need for further assessments of the role of c.6035C>T and c.1751A>C in CMYA5 and c.3401G>T in IFT172 to determine their effect on the function of the corresponding genes, given the contradictory results of in silico analyses and their relatively high frequency in the Middle Eastern population.

We reported 51 variants classified as US in 49 genes of 23 patients with CHD. Fourteen variants were de novo, while the rest were inherited from either one parent or both. We reported five US variants that were inherited from both parents in an autosomal recessive pattern, including c.43A>T in BUB1, c.926A>G in ITGAL, c.884A>G in SMYD4, c.4867C>A in SHROOM3, and c.937G>T in GLA. Homozygous variant c.43A>T in BUB1 was detected in a patient from Qatar (cardio-18.A) coming from a consanguineous family, who had aortic stenosis and left ventricular hypertrophy. Interestingly, this variant has been exclusively reported in the Qatar 1000 genome project among individuals from Qatar, but not in the gnomAD or GME datasets. In silico variant effect predictions, including SIFT and PolyPhen, suggested the possibility of a benign effect, but the CADD score for this variant was 21.2. Using bioinformatic computational analysis, it was shown that BUB1 plays a potential role in CHD. However, its exact role in the pathogenesis of CHD remains unknown, and no reported cases have supported this claim [52].

Variant c.926A>G in ITGAL was detected in a homozygous state in a patient from Qatar (cardio-38.A) coming from a consanguineous family. This variant is novel since it has not been previously reported in any control and clinical datasets. SIFT and PolyPhen consider the variant deleterious and possibly damaging; however, limited published data explain the role of ITGAL in the context of cardiac defects. One study has suggested that it plays an indirect role in cardiovascular diseases [53]. Moreover, the variant c.884A>G in SMYD4 was seen in homozygosity in a patient from Sudan (cardio-31.A) with HLHS. The variant was considered deleterious and damaging based on SIFT and PolyPhen, as well as the CADD of 28.1. Nevertheless, the variant was detected in all control datasets that we analyzed. In the gnomAD dataset, no “probably healthy” subject was homozygous for this variant, which increases the possibility that it may cause HLHS.

Variant c.937G>T in the GLA gene, located in the X chromosome, was observed in a patient (cardio-50.A) with TOF; it was inherited from the mother in a UPD pattern. This variant has been detected in other patients with hypertrophic cardiomyopathy, Fabry disease, angiookeratoma corporis diffusum, and sudden unexplained death (Table S3). Other published case studies have shown that loss-of-function variants in GLA are associated with hypertrophic cardiomyopathy [54]. The difference in phenotype between our patient and previously reported cases with the same variant highlights the difficulty of establishing genotype–phenotype correlations and, consequently, establishing a genetic diagnosis. Overall, many US variants were not fully able to explain the CHD phenotype in our patients, despite the fact that they were located in genes that play a potential role in the pathogenesis of CHD.

In our study, explaining the CHD phenotype in the context of SNVs was challenging. We identified 13 variants shared between our patients and patients from other populations with a variable range of clinical phenotypes; none of them showed a phenotype similar to our patients. Possible explanations may be the variable expressivity and penetrance of the variant, the influence of modifier genes, and the effect of ethnic background. Around 400 genes have been linked to the pathogenesis of CHD, and the American Heart Association (AHA) has issued a statement [1] discussing the genetic architecture of CHD in the context of more than 200 genes and loci. Of them, 27 genes were classified as very commonly associated, 16 as associated, and 21 as occasionally associated. In the Sidra registry cohort, we were able to only capture variants in six genes (KRAS, CHD7, KMT2D, DYNC2H1, SMAD6, CRELD1) [1].

This limited number of fully explained genes is of no value for clinical practice and genetic counseling, where many other genes are frequently seen, especially in patients with an isolated CHD phenotype or those from understudied populations. We observed several
variants inherited in a homozygous pattern among Arab consanguineous families, such as c.243C>G in SLC2A10, c.6035C>T and c.1751A>C in CMYA5, c.1606G>A and c.131G>T in DNAAF3, c.43A>T in BUB1, c.926A>G in ITGAL, and c.884A>G in SMYD4. MAF values indicate a relatively higher prevalence of those variants in the Arab population. For example, c.243C>G in SLC2A10 has a MAF of 0.0071 in the Qatar 1000 genome, while this variant was not detected in other control datasets. Similar observations were noted for c.6035C>T and c.1751A>C in CMYA5, as well as for c.1606G>A and c.131G>T in DNAAF, all of which have a higher MAF in the GME and Qatar 1000 genome dataset compared to gnomAD. Given the endogamous nature of the Middle Eastern population, it is expected that such a variant may have a higher allele frequency than reported. Therefore, the role of these variants in the pathogenesis of CHD must be rigorously assessed.

Our study has some limitations that may have biased our results and their interpretation, including a small sample size, the inability to determine the inheritance of some variants due to the fact that parents were not included in the WES analysis component of the study, as well as the inability to access the original reports for some patients. On the other hand, our study focused on examining nuclear DNA rather than mitochondrial DNA; therefore, there is a possibility that mitochondrial variants were missed in unexplained cases [55,56]. Considering the above, we emphasize the importance of expanding research efforts in the field of CHD genetic architecture.

5. Conclusions

We identified a potential genetic etiology of CHD in 40 out of 52 studied patients with CHD. Sixteen of them were explained by cytogenetic abnormalities, while 24 patients had one or more variants identified by our WES analysis that may have contributed to the pathogenesis of CHD. We identified four pathogenic/likely pathogenic SNVs that could explain the phenotype of CHD in four patients. We detected some recessive variants, such as c.884A>G in SMYD6, which appear to have higher frequencies in Arab and Middle Eastern populations compared to others. However, the exact role of these variants in cardiac pathogenesis has not yet been established. Future in vitro and in vivo functional assessments are needed to better understand the role of these variants in the pathogenicity of CHD.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/genes13081369/s1; Figure S1. Stratification of variants identified in 24 patients with CHD. Table S1: Positive results of chromosomal investigation done for the 27 of our patient’s cohort as part of their clinical evaluation. Table S2: Summary of the novel variants identified in 24 CHD patients through whole exome sequencing (WES) analysis. Table S3: Summary of the shared variants identified in 24 CHD patients through whole exome sequencing (WES) analysis. Table S4: Summary of the variants identified which were classified based on ACMG/AMP guideline as (a) pathogenic/likely pathogenic, (b) likely benign and (c) variants of uncertain significance. References [57–71] are cited in the Supplementary Materials.

Author Contributions: S.O. analyzed the data and wrote the initial draft of the manuscript; D.V., A.E.J., M.K., S.U. and H.A.-S. were involved in participant recruitment and clinical data collection; H.K.-E.-k. and M.A.-S. participated in the supervision of S.O., data interpretation, and manuscript writing; N.S. provided bioinformatics support and assisted in writing the Materials and Methods section of the manuscript; J.U. assisted in the identification of study participants and study management; L.R.S. provided funding for the APC and proofreading, supervision, and critically reviewed the manuscript; K.N.A.-S. designed and managed the study, supervised S.O., interpreted the data, and reviewed the manuscript. All authors have read and agreed to the published version of the manuscript.

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Data Availability Statement: For reasons of privacy and confidentiality, the data from this study are available from the corresponding authors upon reasonable request.

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