Early View

Task force report

Epidemic and pandemic viral infections: impact on tuberculosis and the lung. A consensus by the World Association for Infectious Diseases and Immunological Disorders (WAidid), Global Tuberculosis Network (GTN) and members# of ESCMID Study Group for Mycobacterial Infections (ESGMYC)

Catherine Wei Min Ong, Giovanni Battista Migliori, Mario Raviglione, Gavin MacGregor-Skinner, Giovanni Sotgiu, Jan-Willem Alffenaar, Simon Tiberi, Cornelia Adlhoch, Tonino Alonzi, Sophia Archuleta, Sergio Brusin, Emmanuelle Cambau, Maria Rosaria Capobianchi, Concetta Castilletti, Rosella Centis, Daniela M. Cirillo, Lia D’Ambrosio, Giovanni Delogu, Susanna M.R. Esposito, Jose Figueroa, Jon S. Friedland, Benjamin Ho Choon Heng, Giuseppe Ippolito, Mateja Jankovic, Hannah Yejin Kim, Senia Rosales Klintz, Csaba Ködmön, Eleonora Lalle, Yee Sin Leo, Chi-Chiu Leung, Anne-Grete Märtsön, Mario Melazzini, Saeid Najafi Fard, Pasi Penttinen, Linda Petrone, Elisa Petrucciolì, Emanuele Pontali, Laura Saderi, Miguel Santin, Antonio Spanevello, Reinout van Crevel, Marieke J. van der Werf, Dina Visca, Miguel Viveiros, Jean Pierre Zellweger, Alimuddin Zumla, Delia Goletti.

Please cite this article as: Min Ong CW, Migliori GB, Raviglione M, MacGregor-Skinner G, et al. Epidemic and pandemic viral infections: impact on tuberculosis and the lung. A consensus by the World Association for Infectious Diseases and Immunological Disorders (WAidid), Global Tuberculosis Network (GTN) and members# of ESCMID Study Group for Mycobacterial Infections (ESGMYC). *Eur Respir J* 2020; in press (https://doi.org/10.1183/13993003.01727-2020).

This manuscript has recently been accepted for publication in the *European Respiratory Journal*. It is published here in its accepted form prior to copyediting and typesetting by our production team. After these production processes are complete and the authors have approved the resulting proofs, the article will move to the latest issue of the ERJ online.

Copyright ©ERS 2020. This article is open access and distributed under the terms of the Creative Commons Attribution Non-Commercial Licence 4.0.
Epidemic and pandemic viral infections: impact on tuberculosis and the lung. A consensus by the World Association for Infectious Diseases and Immunological Disorders (WAidid), Global Tuberculosis Network (GTN) and members of ESCMID Study Group for Mycobacterial Infections (ESGMYC).

Authors:
Catherine Wei Min Ong1,2#, Giovanni Battista Migliori3*, Mario Raviglione4,5, Gavin MacGregor-Skinner6, Giovanni Sotgiu7, Jan-Willem Alffenaar8,9,10#, Simon Tiberi11,12#, Cornelia Adlhoch13˄, Tonino Alonzi14, Sophia Archuleta1, Sergio Brusin13˄, Emmanuelle Cambau15#, Maria Rosaria Capobianchi16, Concetta Castilletti16, Rosella Centis3, Daniela M. Cirillo17#, Lia D'Ambrosio18, Giovanni Delogu19,20#, Susanna M.R. Esposito21, Jose Figueroa22, Jon S. Friedland23#, Benjamin Ho Choon Heng24, Giuseppe Ippolito25, Mateja Jankovic26#, Hannah Yejin Kim8,9,10, Senia Rosales Klintz23˅, Csaba Ködmön13˄, Eleonora Lalle16, Yee Sin Leo27, Chi-Chiu Leung28, Anne-Grete Mårtson29, Mario Melazzini30, Saeid Najafi Fard14, Pasi Pentinen13˄, Linda Petrone14, Elisa Petruccioli14, Emanuele Pontali31, Laura Saderi7, Miguel Santin32,33#, Antonio Spanevello34,35, Reinout van Crevel36,37#, Marieke J. van der Werf13˄, Dina Visca34,35, Miguel Viveiros38#, Jean Pierre Zellweger39, Alimuddin Zumla40, Delia Goletti14˄.

*Equally contributed
˄ The European Centre for Disease Prevention and Control Public Health Emergency (ECDC PHE) team co-Authors did not participate to the Delphi process and development of Table 4

Affiliations:
1. Department of Medicine, Yong Loo Lin School of Medicine, National University of Singapore, Singapore
2. Institute for Health Innovation & Technology (iHealthtech), National University of Singapore, Singapore
3. Servizio di Epidemiologia Clinica delle Malattie Respiratorie, Istituti Clinici Scientifici Maugeri IRCCS, Tradate, Italy
4. Centre for Multidisciplinary Research in Health Science, University of Milan, Milan, Italy
5. Global Studies Institute, University of Geneva, Geneva, Switzerland
6. Department of Public Health Sciences, Penn State College of Medicine, Hershey, Pennsylvania, USA
7. Clinical Epidemiology and Medical Statistics Unit, Department of Medical, Surgical and
Experimental Sciences, University of Sassari, Sassari, Italy
8. University of Sydney, Sydney Pharmacy School, Sydney, New South Wales, Australia
9. Westmead Hospital, Sydney, Australia
10. Marie Bashir Institute of Infectious Diseases and Biosecurity, University of Sydney, Sydney, Australia.
11. Blizard Institute, Barts and The London School of Medicine and Dentistry, Queen Mary University of London, London, United Kingdom.
12. Division of Infection, Royal London Hospital, Barts Health NHS Trust, London, United Kingdom
13. Public Health Emergency team, European Centre for Disease Prevention and Control, Stockholm, Sweden
14. Translational Research Unit, Epidemiology and Preclinical Research Department, “L. Spallanzani” National Institute for Infectious Diseases (INMI), IRCCS, Rome, Italy
15. APHP-Lariboisiere, Bacteriologie, laboratory associated to the National reference center for mycobacteria; IAME UMR1137, INSERM, University of Paris, Paris, France
16. Laboratory of Virology, Epidemiology and Preclinical Research Department, “L. Spallanzani” National Institute for Infectious Diseases (INMI), IRCCS, Rome, Italy
17. Emerging Bacterial Pathogens Unit, IRCCS San Raffaele Scientific Institute, Milan, Italy
18. Public Health Consulting Group, Lugano, Switzerland
19. Università Cattolica Sacro Cuore, Roma, Italy
20. Mater Olbia Hospital, Olbia, Italy
21. Pediatric Clinic, Pietro Barilla Children’s Hospital, University of Parma, Parma, Italy
22. National Health Service (NHS), London, United Kingdom
23. St George’s, University of London, London, United Kingdom
24. Tuberculosis Control Unit, Department of Respiratory and Critical Care Medicine, Tan Tock Seng Hospital, Singapore
25. Scientific Direction, “L. Spallanzani” National Institute for Infectious Diseases (INMI), IRCCS, Rome, Italy
26. University of Zagreb, School of Medicine, Clinic for Respiratory Diseases, University Hospital Center Zagreb, Zagreb, Croatia
27. National Centre for Infectious Diseases, Singapore
28. Hong Kong Tuberculosis, Chest and Heart Diseases Association, Wanchai, Hong Kong, China
29. University of Groningen, University Medical Center Groningen, Department of Clinical Pharmacy and Pharmacology, Groningen, the Netherlands
30. Scientific Direction, Istituti Clinici Scientifici Maugeri IRCCS, Pavia, Italy
31. Department of Infectious Diseases, Galliera Hospital, Genova, Italy
32. Department of Infectious Diseases, Bellvitge University Hospital-Bellvitge Biomedical Research Institute (IDIBELL), L’Hospitalet de Llobregat, Spain
33. Department of Clinical Science, University of Barcelona, L’Hospitalet de Llobregat, Spain.
34. Division of Pulmonary Rehabilitation, Istituti Clinici Scientifici Maugeri, IRCCS, Tradate, Italy
35. Department of Medicine and Surgery, Respiratory Diseases, University of Insubria, Varese-Como, Italy
36. Radboudumc Center for Infectious Diseases, Radboud Institute for Health Sciences, Radboudumc, Nijmegen, the Netherlands
37. Centre for Tropical Medicine and Global Health, Nuffield Department of Medicine, University of Oxford, Oxford, United Kingdom
38. Global Health and Tropical Medicine, Institute of Hygiene and Tropical Medicine, NOVA University of Lisbon, Lisbon, Portugal
39. TB Competence Centre, Swiss Lung Association, Berne, Switzerland
40. Department of Infection, Division of Infection and Immunity, University College London and NIHR Biomedical Research Centre, UCL Hospitals NHS Foundation Trust, London, United Kingdom

**Corresponding Author:**
Delia Goletti, Translational Research Unit, Epidemiology and Preclinical Research Department, “L. Spallanzani” National Institute for Infectious Diseases (INMI), IRCCS, Via Portuense 292, 00149, Rome, Italy. Email: delia.goletti@inmi.it

**Co-corresponding Author:**
Giovanni Battista Migliori, Servizio di Epidemiologia Clinica delle Malattie Respiratorie, Istituti Clinici Scientifici Maugeri IRCCS, Via Roncaccio 16, Tradate, Varese, 21049, Italy. E-mail: giovannibattista.migliori@icsmaugeri.it
**Key words:** viral infections, lung, TB, COVID-19, immunology, diagnosis, treatment, prevention, infection control, workplace safety

**Running title:** Epidemics, pandemic viral infections and interactions with TB and the lung

**Take-home message:** In this consensus we describe the effects of the viral infections resulting in epidemics and pandemics affecting the lung (MERS, SARS, HIV, influenza A (H1N1)pdm/09 and COVID-19) and their interactions with tuberculosis, the top infectious disease killer.
Abstract

Major epidemics including some that qualify as pandemics, such as Severe Acute Respiratory Syndrome (SARS), Middle-Eastern Respiratory Syndrome (MERS), Human Immunodeficiency Virus, pandemic H1N1/09 and most recently COVID-19 affect the lung. Tuberculosis (TB) remains the top infectious disease killer but apart from the TB-HIV syndemic, little is known regarding the interaction of viral epidemics and pandemics with TB. The aim of this consensus-based document is to describe the effects of the viral infections resulting in epidemics and pandemics that affect the lung (MERS, SARS, HIV, influenza A (H1N1)pdm/09 and COVID-19) and their interactions with TB. A search of the scientific literature was performed. A writing committee of international experts including the European Centre for Disease Prevention and Control Public Health Emergency (ECDC PHE) team, the World Association for Infectious Diseases and Immunological Disorders (WAidid), the Global Tuberculosis Network (GTN) and members of ESCMID Study Group for Mycobacterial Infections (ESGMYC) was established. Consensus was achieved after multiple rounds of revisions between the writing committee and a larger expert group. A Delphi process involving the core group of authors, excluding the ECDC PHE team identified the areas requiring review/consensus, followed by a second round to refine the definitive consensus elements. The epidemiology, immunology of these viral infections and their interactions with TB are discussed with implications on diagnosis, treatment and prevention of airborne infections (infection control, viral containment and workplace safety). This consensus document represents a rapid and comprehensive summary on what is known on the topic.
1. **Introduction.**

The 21st century has been marked by major epidemics, including some that qualify as pandemics, caused by old diseases such as cholera, plague, and yellow fever as well as emerging ones such as Severe Acute Respiratory Syndrome (SARS), Ebola, Zika, Middle-Eastern Respiratory Syndrome (MERS), Human Immunodeficiency Virus (HIV, although technically endemic), influenza A (H1N1)pdm/09 and most recently COVID-19. Several of these viruses affect the lung. Tuberculosis (TB) remains the top infectious disease killer caused by a single organism responsible for 1.5 million deaths in 2018 [1]. Apart from the TB-HIV syndemic, little is known regarding the interaction of other viral epidemics with TB. This consensus-based document describes the effects of the main viral epidemics which predominately affect the lungs or cause systemic immunosuppression (MERS, SARS, HIV, influenza A(H1N1)pdm09 and COVID-19), and their interactions with TB at a diagnostic, treatment and public health level. It is fruit of a collaborative project involving the ECDC PHE team (European Centre for Disease Prevention and Control, Public Health Emergency), the World Association for Infectious Diseases and Immunological Disorders (WAidid), the GTN (Global Tuberculosis Network) and members of ESGMYC.

2. **Methods**

We did a rapid and non-systematic search of the literature using the key-words ‘COVID-19’, ‘tuberculosis’, ‘viral infection’, ‘HIV infection’, ‘SARS’, ‘lung’, ‘immunology’, ‘diagnosis’, ‘prevention’, ‘infection control’, ‘workplace’ to identify a minimum set of references from an electronic database (PUBMED), existing guidelines on TB, viral diseases, airborne diseases, and grey literature from their inception until 29 April 2020. Guidelines were retrieved from the websites of the main international health-related centres, whereas grey literature was accessed using Google search engine.

A writing committee composed of international experts was established, including the ECDC PHE team, WAidid, GTN and ESGMYC.

Consensus on the content was achieved after multiple rounds of revisions between the writing committee and the larger group of experts [2].

A Delphi process involving the core group of Authors, excluding the ECDC PHE team, identified the areas requiring review/consensus, followed by a second round to refine the definitive consensus elements.

As this review is not aimed at duplicating World Health Organization (WHO), ECDC and other existing guidelines, the GRADE methodology was not used, and no formal recommendations are provided. The available information on prevention, diagnosis, and treatment of TB and pulmonary viral diseases was selected by the experts and summarised and country examples were provided to critically discuss the public health response.
3. Viral diseases of the lung

3.1 Epidemiology.

Viral respiratory infections are a major public health concern due to the capacity of viruses to spread from person to person directly via aerosols/droplet nuclei, small droplets or virus laden secretions from larger droplets; or indirectly by contact with contaminated surfaces [3]. Large respiratory droplets are generated primarily during coughing, sneezing and talking, and during procedures such as suctioning and bronchoscopy, which can also generate droplet nuclei. Transmission occurs when droplets containing microorganisms from an infected person are expelled a short distance through the air and deposited on another individual’s conjunctivae, nasal mucosa or mouth. Large droplets fall quickly onto surfaces close to the infected person, increasing the risk of contact transmission. Moreover, viral infections can also be transmitted via aerosol particles of small size (<5-10 µm) which may be infectious at a distance of several meters [4-7]. Recent evidence suggests the SARS-CoV-2 virus may be present in exhaled air while talking and breathing [8], being detected for several hours [9] on different surfaces.

Respiratory infections can be classified by the causative virus such as influenza or clinically according to the clinical syndrome. Symptoms may include fever, non-productive cough, coryza, sneezing, dyspnoea, myalgia, fatigue and non-exudative pharyngitis [10].

The clinical spectrum can encompass asymptomatic infection, upper respiratory tract infection, and lower respiratory tract infection that can result in pneumonia or acute respiratory distress [11], and systemic infection [12].

The severity of viral respiratory illness varies widely, and severe disease is more likely in older patients with or without comorbidities. Infants may have more severe disease for some organisms. Morbidity may result directly from viral infection or may be due to exacerbation of other chronic medical conditions, or bacterial super-infection [13, 14].

The spread of respiratory virus infections varies between countries and regions, depending on differences in population, geography, climate, immunization coverage, and socioeconomic status [15-17].

3.2 Immunology.

The first line of defence against respiratory viral infections includes intrinsic defences such as mucous and antiviral peptides. When these are circumvented, viruses enter the epithelial cells by recognizing viral components via Toll-like receptors (TLRs) and intracellular receptors [18] (Figure 1), and initiate the inflammatory response. Innate cells such as dendritic cells, alveolar macrophages, natural killer cells and neutrophils are recruited. All these cells promote an anti-viral response and are important for the
establishment of adaptive responses. Concurrently, these inflammatory cells may be important in driving innate immune-mediated tissue damage, a process which also occurs in tuberculosis [19].

T-cells contribute to the generation of the B-cell response, and cell-mediated immunity leading to viral clearance. In particular, B-cells produce antibodies that may neutralise the respiratory viruses directly by binding to viral surface proteins or activating the complement cascade (Figure 1)[20, 21]. Viral clearance is also mediated by CD8+-specific T-cells with cytolytic activity. Protective anti-viral T-cell response is a Th1 response mainly mediated through interferon (IFN)-γ production [22-25]. Moreover, to prevent lung tissue damage, all these responses are finely modulated; regulatory mechanisms adopted by T-cells such as cytokine secretion, upregulation of inhibitory receptors [26], or expansion of the T-cell regulatory subset lead to a balance between tissue damage and clearance of the virus. The immune systems of neonates, infants, children and adults are different, in their composition and functional responsiveness to infectious diseases [27, 28].

Regarding the response to Mycobacterium tuberculosis (Mtb), after mycobacterial dissemination to the lymph nodes, dendritic cells present bacterial antigens to T-cells and prime them [29, 30]. Priming occurs around 10 days post-infection in the mediastinal lymph nodes and is followed by generation of effector T-cells [31], and Th1 CD4+ T-cells that lead to the formation of granulomas. Granulomas are organized structures where T-cells and B-cells surround innate immune cells (macrophages and neutrophils) with a fibrotic capsule to generate a hypoxic environment to prevent Mtb growth [32], with hypoxia potentially worsening tissue destruction in TB [33]. CD4 Th1 host responses are crucial [34], especially at the beginning for TB control [35, 36]. Regarding the CD8 T-cell component, mouse studies have revealed a relatively smaller role of CD8+ T-cells in protection against Mtb infection [37], and an even smaller contribution of B-cells and humoral immunity [38]. Differently, in human studies Mtb-specific CD8 T-cells have been associated with active TB [39-44], both in HIV-uninfected and -infected patients [35, 42, 45], and in response to recent infections [43, 44]. Increased CD8 T-cell responses is associated with Mtb load and longitudinal studies have shown a decrease of this CD8 T-cell response during anti-TB treatment [39, 40]. It has been shown that they are differently modulated over the course of TB disease suggesting a role in the TB pathogenesis that is not yet fully elucidated [46-48].

Bacille-Calmette Guerin (BCG) vaccination as a potential intervention against COVID-19

BCG, the current vaccine against TB has important protective effects against other infections. In randomised trials, BCG reduced infant mortality by around 40% [49], and respiratory infections other than TB by 70% in adolescents [50]. These ‘non-specific effects’ of BCG-vaccination are explained by epigenetic and metabolic reprogramming of innate immune cells, a process termed ‘trained immunity’ [51]. Clinical evidence suggests that BCG may protect against viruses [52], and BCG-vaccinated healthy adults re-challenged with the live yellow fever vaccine showed improved anti-viral immunity and decreased viral
loads [53]. In mice, BCG-vaccination protects against influenza A, lowering viral replication and lung injury [54, 55].

A recent ecological analysis has suggested that BCG-vaccination may also protect against COVID-19 [56]. Countries without universal policies of BCG-vaccination (Italy, Netherlands, USA) seem more severely affected by COVID-19 compared to countries with universal BCG policies. However, such ecological studies that relate country aggregate and individual data should be interpreted with caution. Also, COVID-19 showed a recent increase since publication in low- and middle-income countries and may still be underreported, confounders such as age were not taken into account, and variable BCG policies over time affect individual BCG coverage [57-59]. Several large RCTs currently evaluating the effect of BCG-vaccination against COVID-19 in thousands of health care workers and elderly, in the Netherlands, Australia and other countries, will provide evidence to support or refute BCG as a cheap and rapidly scalable preventive measure against COVID-19 and other viral respiratory infections.

3.3 Influenza H1N1 and lung disease

The two most serious impacts of influenza virus on the lung are the development of pneumonia and exacerbation of pre-existing pulmonary disease [60]. Such events seem rare and variable during most seasonal influenza periods but may be more frequent and severe in pandemics. During the 2018-2019 season, there was an estimated 32 million cases of influenza resulting in 32,000 deaths in the USA [61]. H1N1, the virus behind the 1918 and 2009 pandemics, appears to cause more rapid and severe pneumonia than other strains, with higher rates of bacterial super-infection [62]. H1N1 also affects the paediatric population [63, 64].

Primary viral pneumonia is characterised by rapid onset of non-productive cough, headache, myalgias, dyspnoea, tachypnea, hypoxia and ground-glass opacities on CT scans. Secondary bacterial pneumonia, which may occur concurrently or following the development of viral pneumonia, is a frequent complication. Bacterial super-infection occurs through direct damage of the respiratory epithelium with modification of local and systemic immune defense. Bacterial super-infection, mainly due to *Staphylococcus aureus* and *Streptococcus pneumoniae* [65], is reported in 20-47% of influenza patients admitted to intensive care units (ICU). Its global prevalence varies between 0.1-10% according to ancient surveys and 1.59% according to a recent Korean survey during the 2009 influenza A/(H1N1)pdm09 pandemic [66]. Influenza and bacterial super-infection are two distinct clinical-pathological syndromes which have been described by Morens and Fauci [67]. The first with 10 to 15% fatal cases is similar to acute respiratory distress (ARDS)-like syndrome while the latter with poorer prognosis with 85 to 90% fatal cases manifesting as acute bronchopneumonia, with pathogenic bacteria cultured on autopsy [60, 62, 68].
3.4 **Other viral infections and lung disease (SARS, MERS)**

In recent decades, previously unknown zoonotic respiratory tract infections with epidemic potential such as SARS and MERS emerged. Human coronaviruses are usually classified into low and highly pathogenic [69]. The low pathogenic coronaviruses infect the upper respiratory tract and cause ‘flu-like’ mild respiratory illness, while highly pathogenic ones (SARS and MERS) predominantly infect lower airways often causing fatal pneumonia [69].

Severe coronavirus pneumonia is often associated with rapid virus replication, massive inflammatory cell infiltration and elevated pro-inflammatory cytokine/chemokine responses producing acute lung injury and ARDS. Recent studies in experimentally infected animals strongly suggest a crucial role for virus-induced immune-pathological events in causing fatal pneumonia following coronavirus infection [69].

High initial viral titres in the airways, age and comorbidities (hypertension, diabetes, obesity, heart failure, renal failure, etc.) are associated with worse outcomes [70-74].

SARS-CoV, which enters the human cell by the angiotensin-converting enzyme-2 (ACE2) receptor [75], presents usually with three phases [76]: 1) rapid viral replication with fever, cough, and other non-specific symptoms, disappearing in a few days; 2) high fever, hypoxemia, and progression to pneumonia-like symptoms, despite a progressive decline of viral replication; 3) development of ARDS in ~20% of patients with mortality [77-79].

MERS, which enters the human cell by dipeptidyl peptidase four (DPP4) receptor [80], usually starts with flu-like symptoms: fever, sore throat, non-productive cough, myalgia, shortness of breath, and dyspnoea, often progressing to pneumonia (ICU admission necessary) [73, 81]. It can also cause gastrointestinal symptoms (abdominal pain, vomiting, and diarrhoea).

3.5 **COVID-19 and lung disease**

According to a recent report from China, COVID-19, the disease caused by SARS-CoV-2, is characterised by three clinical patterns: absence or paucity of symptoms, mild to moderate disease, severe pneumonia requiring admission to ICU [82]. Dyspnoea develops after a median time of 8 days from illness onset, with a median time to ICU admission of 5 days. Up to 20% of the patients require the transfer to ICU [83, 84], with consequent overwhelming of healthcare capacity. The evidence suggests that while 25% of COVID-19 patients have co-morbidities including chronic obstructive pulmonary disease (COPD), diabetes mellitus, hypertension, coronary heart disease, cerebrovascular disease and malignancies, the proportion amounted to more than 90% among those who died [83-87]. In children, COVID-19 symptoms are usually milder with better outcome than adults [88-92]. Frequency of clinical presentations and outcome appear different in Europe with higher lethality compared to China, although this figure may change overtime due to
better estimation of the total number of infections [93]. Besides virologic diagnosis, imaging by chest radiography, ultrasound and computer tomography (CT) are important for diagnosis and management. Main CT abnormalities include ground-glass opacity and consolidation [94]. The combination of CT scan findings, respiratory parameters (peripheral capillary oxygen saturation (SpO2) and PaO2/FiO2), and blood tests (C-reactive proteins, lymphocyte count, LDH, triglycerides, ferritin, fibrinogen, D-dimer, IL-6) [84, 95], are important features to identify those at highest risk for ICU transfer. Lungs of dead COVID-19 patients showed oedema, proteinaceous exudate, focal reactive hyperplasia of pneumocytes with patchy inflammatory cellular infiltration, and multinucleated giant cells with fibroblastic plugs in airspaces [96, 97]. A recent study reported autopsy cases contained diffuse alveolar damage, with mononuclear response (CD4+ aggregates) surrounding thrombosed vessels, in the presence of associated haemorrhages [98-100].

3.6 HIV and lung disease

The spectrum of HIV-associated pulmonary diseases is broad and the lungs are one of the most frequently affected organ systems in HIV-infected persons regardless of age [101]. The absolute CD4 T-cell count, used as a surrogate marker of immunodeficiency, is important in guiding the aetiological evaluation of lung infections [102]. Pulmonary TB infection and reactivation are more likely with a CD4 count of <500 cells/mL. Opportunistic infections, *Pneumocystis jirovecii*, bacterial infections, Kaposi sarcoma, and extrapulmonary/disseminated forms of TB occur mainly in patients with CD4 T-cell counts < 200 cells/mL. Cytomegalovirus (CMV) infection, *Mycobacterium avium* complex (MAC) and aspergillosis occur usually at CD4 counts < 50 cells/mL. Risk factors to consider are the geographical origin that predisposes to specific disease (such as TB, coccidioidomycosis, paragonimiasis and histoplasmosis), adherence to antiretroviral therapy, prescription of *Pneumocystis jirovecii* prophylaxis, and presence of comorbidities. Community-acquired bacterial pneumonia occurs at all stages of HIV infection but is more frequent in patients with profound CD4 T-cell depletion and decreases with antiretroviral therapy. Community acquired pneumonia accounts for 35% to 50% of all hospital admission cases due to respiratory failure and is the main reason for ICU admission [103].

4. TB and respiratory viral diseases

4.1 TB and influenza

The association of TB and influenza could be bidirectional: TB may increase the susceptibility to influenza and the risk of complications and influenza may increase the susceptibility to TB. The susceptibility to influenza appears to be greater in patients with pre-existing pulmonary disease (such as asthma and COPD). As a large proportion of post-TB treatment patients have long-term functional impairment, mainly COPD, which can be severe [104, 105], patients with such pulmonary sequelae may be predisposed and more susceptible to influenza infection and its complications including mortality [106]. Furthermore, the temporary immunosuppression induced by TB may increase the susceptibility of patients to influenza infection. An excess mortality associated with influenza has been described among TB patients in
South Africa [107]. TB patients have a similar prevalence of viral and bacterial co-infection as their household contacts but TB patients often have more severe disease if they are co-infected [108].

As early as 1919, the occurrence of TB among patients surviving influenza or pneumonia, but without clear distinction between both diseases was reported [109]. Influenza induces a temporary increase in the susceptibility to bacterial infections, exemplified by the frequent occurrence of bacterial pneumonia following viral pneumonitis [110]. Because influenza impairs the immune response, it may be expected that influenza could also promote the development of active TB among patients with latent TB infection [111], but the occurrence of TB may occur much later than the occurrence of influenza, thus making the temporal association difficult to demonstrate. There was an excess mortality from pulmonary TB during the influenza pandemics of 1889 and 1918 in Switzerland [112]. It has also been reported that summer influenza epidemics in Wuhan, China, may have contributed to the increase in reported TB cases [113]. Conversely, a report from Thailand did not demonstrate a worse outcome for patients with concurrent influenza and TB [114] and another report from Indonesia did not demonstrate a correlation between antibodies against influenza and the presence of TB but there was an association between the level of antibody titres against influenza virus and the stage of TB [115]. Interestingly, influenza vaccination is reported to be a protective factor against TB in elderly persons in Taiwan [116]. The exact impact of concurrent influenza and TB remain uncertain.

4.2 TB and HIV

The interaction between TB and HIV is well known. Without anti-retroviral treatment, the risk of latent TB infection progressing to active TB disease in people living with HIV and AIDS (PLWHA) is greater than immunocompetent hosts. In PLWHA the risk of developing TB is in the order of 10% per year [117, 118]. This elevated risk is behind the WHO recommendation of TB screening and/or preventive treatment for all PLWHA [119, 120]. New regimens as short as 1 month (daily rifapentine plus isoniazid) to 3 months (weekly rifapentine plus isoniazid) were recently recommended by WHO, and are well tolerated and effective [119, 120]. Important programmatic implications for collaboration between TB and HIV/AIDS services exist: TB services should test for HIV (allowing treatment of TB patients with anti-retrovirals and cotrimoxazole preventive therapy in patients with HIV-TB co-infection) and HIV/AIDS services should screen for latent TB infection (LTBI), using Tuberculin skin test/TST or interferon-γ release assay (IGRA), and initiate prompt treatment of TB or LTBI in PLWHA [119, 120]. Based on this rationale (two diseases, one patient) WHO promotes TB/HIV collaborative activities focused on 3 main pillars [121]: 1) establish TB/HIV collaborative mechanisms; 2) decrease burden of TB among PLWHA; 3) decrease burden of HIV among TB patients. Moreover, testing for other infections in addition to HIV in TB clinics may be indicated during epidemics and pandemics.
4.3 TB, SARS and MERS

TB co-infection with SARS is rare. A study of 83 patients with SARS found 3 patients with TB co-infection, where one patient with SARS subsequently developed TB, while the other two had TB and then developed SARS [122]. All three patients were on steroid therapy, which may have decreased viral- and/or TB-specific immunity and increased the risk of co-infection. In a different cohort of 236 SARS patients, two were diagnosed with pulmonary TB [123]. The development of TB in the presence of SARS may be due to CD4 lymphopaenia during the viral infection [124], as CD4+ T-cells are crucial for TB-specific immunity [41, 125-129]. Lastly, one TB patient developed SARS co-infection because of a wrong admission to a cohort of SARS patients [130]. This highlights the importance of remaining vigilant to other communicable diseases including TB when epidemic or pandemic infections dominate media headlines [131]. TB with MERS Co-V coinfection is also rare. A report of 295 MERS-CoV patients found 2 TB patients [132], although it was unclear which the initial infection was.

4.4 TB and COVID-19

There may be interaction between COVID-19 and TB [133], but long-term observations are lacking [134]. Only two studies investigated the interactions so far. In the first ever cohort of 49 patients from 8 countries COVID-19 was diagnosed before, simultaneously or after TB [135]. In the second study including 69 patients mortality was investigated [136]. In a separate anecdotal report, Liu et al reported an increased prevalence of latent TB in patients with severe COVID-19 infection and concluded that infection with Mtb may influence the progression and outcome of COVID-19 [133]. Evidence on the interactions between TB and COVID-19 is needed.

5. Respiratory viral diseases and TB in the elderly, prisoners and other vulnerable groups

The elderly (age ≥ 65 years old), prisoners and other vulnerable groups such as forced migrants may reside in high density communal settings which can perpetuate rapid infectious disease transmission during an epidemic or pandemic. Immunosenescence is an additional risk factor in elderly [137].

Clinical presentation of these infections in the elderly can be subtle, with atypical manifestation such as delirium, and may present with complications. SARS, and COVID-19 respiratory failure are well-documented in the elderly [138, 139]. Conversely to influenza A (2009 H1N1)pdm09, a study of 4,962 patients found elderly patients having less risk of respiratory failure, ICU admission or mechanical ventilation [140]. For TB, old age is a risk factor for active TB with poorer treatment outcomes [1, 141], while TB symptoms are indistinguishable from symptoms of malignancies. Moreover, the elderly may also suffer from abnormal drug absorption and/or drug toxicities due to polypharmacy for comorbidities. Similarly, immunocompromised and pregnant women may also present with complications including respiratory failure when infected with pandemic H1N1 influenza, or TB [142-144].
Diagnostic tests for each group depend on available resources and on accessibility, which may be limited to none for the homeless and the incarcerated, although there should be equity in the availability of testing and treatment. The elderly may further have technical difficulties in providing quality respiratory samples for testing, such as for TB, when they have an impaired cough response. A poor-quality respiratory sample inevitably delays diagnosis and contact tracing efforts.

Treatment and management of viral infections and TB include prompt isolation of presumptive cases and of microbiological confirmed ones depending on available resources, or even controlled release of prisoners [145]. This is part of the comprehensive strategy to mitigate transmission in nursing homes for elderly, in refugee camps and correctional facilities [146]. Prognosis of TB, SARS and COVID-19 tend to be worse, with higher mortality in the elderly [77, 138, 147].

6. Diagnostic challenges in viral diseases and TB

For prompt diagnosis of viruses causing severe acute respiratory infections (SARI)[148], such as SARS-CoV, MERS-CoV and SARS-CoV-2 [149-152], and differentiation from other common bacterial infections, a strategic laboratory approach is needed. This approach requires integrating conventional virology assays, molecular platforms combining nucleic acid extraction and PCR or RT-PCR, and rapid molecular tests (RDTs) used in point-of-care (POC) minilabs (Table 1). Positive results using single or multiplex RDTs may lead to adequate cohorting and management of infected patients [153]. Negative results are often less conclusive because of a lack of sensitivity and non-standardised collection of specimens. Using metagenomic next-generation sequencing (mNGS), pathogens not included in the tests can be detected including both known and novel viruses. Genomic data gives information on virulence genes [154], resistance mutations and clusters using phylogenetic approaches [155, 156]. Specific antibody detection remains useful for seroprevalence studies in selected populations and in vaccine studies. The recurrence of old pathogens and emergence of new ones like SARS-CoV-2 underline the importance of worldwide virus surveillance systems [157]. For this purpose, developing protein microarrays to respiratory viruses’ serology is useful [153].

Diagnosis of active TB relies on direct detection of Mtb, most often in respiratory specimens. Although culture remains the “gold standard” in terms of sensitivity and specificity, effective molecular assays to detect the Mtb DNA are also used on platforms and in POCs [158]. Moreover, these Mtb molecular assays can detect mutations associated with resistance, rapidly detecting multi-drug resistant (MDR-)TB strains resistant to rifampin and isoniazid, allowing appropriate therapies and curbing transmission of these strains [159]. Testing immune memory to a previous TB exposure (i.e latent TB infection), performed with the Mantoux test and IGRAs, cannot be used as surrogate of protection but identifies persons who have been previously infected and can be useful to guide the TB diagnostic algorithm [160].
The massive use of molecular assays to diagnose COVID-19 introduced similar molecular platforms that could be used for detecting pathogens in respiratory specimens. The challenge lies in sample processing and RNA/DNA extraction protocols, rather than cross-reactivity resulting in false-positive results. This is an opportunity to strengthen the diagnostic potential of microbiological laboratories, producing an invaluable asset to improve diagnosis against other infections including TB. At the same time, the expertise, tools and networks developed for TB diagnosis could aid the rapid implementation of molecular diagnosis of COVID-19 and other viral infections. Tests (lateral flow assay and ELISA-based) to rapidly detect the antigens in swabs or respiratory secretions and to determine serological evidence of recent and past infection and evidence of neutralizing antibodies are currently being evaluated.

7. Impact of new, potential and existing drugs for viral diseases and COVID-19 on TB therapy.

Eighty percent of COVID-19 cases are generally mild and self-limiting and may require no treatment. Lacking licensed drugs for SARS-CoV-2, therapeutic approaches in severely ill patients are limited to supportive care and empirical use of antibiotics to prevent or treat secondary infections [84, 86, 161, 162]. To provide active treatment for SARS-CoV-2, drugs potentially inhibiting viral replication (Figure 2) are of interest [163].

TB patients with COVID-19 may require an adapted therapeutic approach compared to patients without COVID-19. Switching to intravenous anti-TB drugs is recommended for patients in intensive care in order to optimize drug exposure in critically ill patients [164]. Therapeutic solutions for COVID-19 in TB patients need to be considered in the perspective of anti-TB treatment. However, more evidence ideally from clinical trials is necessary.

7.1 Lopinavir/ritonavir (Kaletra)

Lopinavir/ritonavir is a widely studied treatment for COVID-19 although evidence of efficacy is still limited [165]. In non-TB patients the combination, studied in an open label randomised controlled trial, did not show any virologic or clinical response compared to standard of care [166]. For the treatment of TB-HIV co-infected patients, lopinavir/ritonavir is not recommended in combination with rifampicin due to cytochrome P450 (CYP) induction. Super boosting of lopinavir/ritonavir by additional ritonavir in children on rifampicin-based TB treatment could be attempted as comparable drug exposure was achieved in situations without rifampicin [167]. Alternatively, rifabutin in a dose of 150 mg once daily has been used in combination with lopinavir/ritonavir [168].
7.2 Chloroquine phosphate, Hydroxychloroquine

Chloroquine and hydroxychloroquine are evaluated in several clinical trials for therapy and prophylaxis against SARS-CoV-2 following promising *in vitro* results [163, 169]. In the absence of results from well-designed clinical studies, clinical benefit is currently unknown [170, 171]. Both drugs have immunomodulatory properties [172, 173]. Chloroquine has shown to reduce tumor necrosis factor-alpha production and receptor-mediated signaling in monocytes, which could prevent SARS-CoV-2 induced severe inflammatory response [174]. TB physicians should be careful when combining these drugs with TB drugs like moxifloxacin, bedaquiline, delamanid and clofazimine due to risk of increased QTc prolongation. An ongoing trial was halted due to irregular heart rates and increased risk of fatal heart arrhythmia [175]. The Food and Drug Administration (FDA) has also issued a warning on these 2 drugs [176]. In addition, rifampicin increase chloroquine CYP3A4 and CYP2D6-mediated metabolism to desethylchloroquine and bisdesethylchloroquine [177]. It is unclear whether these metabolites are active against SARS-CoV-2.

7.3 Steroids

Intravenously administered steroids have been recommended for selected non-TB patients with ARDS preferably in trial setting [86, 161, 162]. However, the role of steroids to reduce ARDS in TB patients is limited as data of good quality to support the use of steroids outside the treatment of TB meningitis is scarce [164]. Evidence on the use of steroids in COVID-19 is awaited.

7.4 Drug interactions

Potential drug-drug interactions (DDI) are presented in Table 2. The summary of interactions are largely based on evaluations made from pharmacokinetics and toxicity profiles of drugs given alone and comparatively, when co-administered with other drugs in a separate study, in the absence of real dedicated DDI studies. The summary includes effects on drug exposure, monitoring/action and potential mechanisms.

7.5 Immunomodulatory drugs

To reduce the inflammatory response, inhibitors of interleukin (IL)-1 and IL-6 and JAK1/JAK 2 inhibitor baricitinib are being studied [178, 179]. Azithromycin may be of potential interest for immunomodulatory effect [180], although data on its efficacy are lacking and its effect antimicrobial resistance should be considered. Interferons are being tested because of their stimulatory activities of innate antiviral responses [163].
7.6 Novel drugs

Antiviral candidates like azvudine, baloxavir marboxil, favipiravir, remdesivir, ribavirin and umifenovir are being tested for COVID-19 [163, 181-183] (Figure 2). Remdesivir is a nucleotide analogue showing in vitro activity against SARS-CoV-2 [183]. Remdesivir is being studied in two large phase 3 RCTs (NCT04252664, NCT04257656), of which one multi-centre trial conducted in Hubei, China in severe COVID-19 showed no difference in time to clinical improvement or mortality benefit [184]. Umifenovir has shown in vitro activity against SARS-CoV-1 [185] and improved radiological findings when added to lopinavir/ritonavir in a small RCT [186] and seems suitable for further development.

8. Controlling viral diseases and TB: strengths and opportunities.

8.1 Principles of viral containment

Globalisation, increased urbanisation resulting in large vastly populated and overcrowded cities and the development of fast mass transit networks and consequent ease of travelling, means that a virus can spread across a country or a continent in just a few hours. In the absence of a vaccine or an effective treatment, the tools to control a new viral infection have remained the same as during the 1918 Spanish influenza pandemic; namely early public health interventions designed to reduce the risk of transmission and spread of infection such as increased respiratory hygiene, cough etiquette and hand washing, voluntary isolation of infected individuals or households as well as quarantine of their contacts, followed by voluntary or mandatory physical distancing measures, restrictions on travel and transportation and dissemination of basic infection prevention and control messages and advice to the general population [93]. National Lockdown is an extreme measure that, while potentially reducing transmission, may also result in the collapse of the economy of a country. These non-pharmaceutical countermeasures aim at reducing the impact of COVID-19 by minimising the number of contacts that result in disease transmission and, thus, reducing the effective reproduction number $R_0$ to below 1. The reduction of the number of cases during the epidemic peak is crucial to reduce the burden on the health care services and other related sectors and aims to flatten the curve by spreading cases over a longer period of time. This approach, while not necessarily reducing the total number of cases, gains time necessary for the development, production and distribution of effective and safe pharmaceuticals (i.e. vaccines and antiviral drugs), implementation of adequate hospital response and obtaining necessary ICU equipment as well as more sensitive diagnostic tests.

Infection Control refers to the different methods and strategies deployed to reduce or prevent the incidence and/or transmission of infections (see section 8.2). Containment, through early detection, investigation and reporting of cases, together with contact tracing with self-isolation, aimed at containing, preventing or
Delaying the spread of the disease in the community. Geographical containment in a defined area relies on measures to restrict the virus spreading beyond the ‘Containment Zone’ or ‘Cordon sanitaire’, including pharmacological and public health interventions such as intensified surveillance and laboratory testing, movement restrictions in and out of the containment zone, and monitoring the area immediately surrounding the containment zone (buffer zone) for secondary infections [187].

Delaying the spread of infection can be achieved by early identification and treatment of cases, monitoring and follow-up of contacts, physical distancing measures such as proscribing public or religious gatherings, closing schools impacting working parents, sports events or businesses reducing the contact between people. It aims at lowering the epidemic peak reducing the burden of cases at a given time maintaining it below health care capacity.

A systematic analysis of the responses to the 1918 Influenza pandemic showed that in the USA, cities that introduced early social isolation measures experienced a significant reduction of viral spread, approximately 50% lower peak death rates and nearly 20% lower cumulative excess mortality than cities that did not, with a consequent reduction on health care pressures [188].

### 8.2 Airborne infection control and workplace safety

Airborne infection control in healthcare settings uses a hierarchy of control measures based on elimination of sources of infection, engineering controls, administrative controls, and personal protective equipment (i.e., surgical masks for infectious patients and respirators for healthcare workers and visitors)[189-191]. This approach is described in detail in the TB guidelines [189, 190], but can be extended also for viral infections including COVID-19 [192, 193]. As presently under discussion in countries under a post-lockdown phase perspective, the concept is valid also to ensure workplace safety in non-healthcare settings.

While N95/N99 and FFP2/FFP3 respirators or higher level respirators including disposable filtering face piece respirators, powered air purifying respirators, elastomeric respirators (defined as per USA and European standards, respectively) and eye protection are recommended to protect healthcare staff and other exposed individuals at workplace level (after adequate training), the use of surgical masks is debated [194, 195]. Although there is agreement on the use of surgical masks to limit the spread of droplet nuclei for isolated symptomatic patients, the potential mass use of surgical masks to limit the community spread of COVID-19 during the early stages of infection and from asymptomatic individuals is strongly discussed [192, 194, 195]. Arguments against their wide use are based on the false sense of protection (e.g., the individual feels the surgical mask protects him/her from acquiring infection) as well as the potential risks of moisture retention, long mask re-use, and limited filtration capacity [196]. While WHO is revising its recommendations, the use of masks among community members has been re-evaluated [195, 197, 198]. Recent ECDC guidance states that face masks used by the general population may reduce the spread of the
infection in the community by minimising the excretion of respiratory droplets from infected individuals who have not yet developed symptoms or who remain asymptomatic [199]. In general, all infection control measures are important to prevent infections and render workplaces safe.

The stability of SARS-CoV-2 is similar to SARS-CoV-1 and studies indicate that aerosol and fomite transmission of SARS-CoV-2 is plausible [9], and can be associated with nosocomial spread and superspreading events since it can remain viable and infectious in aerosols for hours and on surfaces up to days [200].

Current evidence provides support for direct contact and respiratory droplets as predominant routes of SARS-CoV-2 transmission [201], and highlights the importance of environmental surface cleaning with a hospital grade disinfectant and meticulous hand hygiene.

SARS-CoV-2 is inactivated by common disinfection measures such as a 5-minute contact with household bleach [202]. The following disinfectants kill the virus: ice-cold acetone (90 seconds), ice-cold acetone/methanol mixture (40:60, 10 minutes), 70% ethanol (10 minutes), 100% ethanol (5 minutes), paraformaldehyde (2 minutes), and glutaraldehyde (2 minutes). Commonly used brands of hand disinfectants also inactivate SARS-CoV (30 seconds) [203]. The ECDC guidance on disinfection of environments in healthcare and non-healthcare settings potentially contaminated with SARS-CoV-2 recommends products with virucidal activity licensed in the national markets or 0.05% sodium hypochlorite (NaClO) (dilution 1:100, if household bleach is used, which is usually at an initial concentration of 5%). For surfaces that can be damaged by sodium hypochlorite, products based on ethanol of at least 70% can be used [204]. The virus is sensitive to heat (60°C for 30 minutes) [203], and UV radiation (60 minutes) [205].

Outside the host, the virus can survive for 4 days in diarrheal stool samples with an alkaline pH [203], more than 7 days in respiratory secretions at room temperature, for at least 4 days in undiluted urine, faeces and human serum at room temperature[202], up to 9 days in suspension, 60 hours in soil/water, more than a day on hard surfaces such as glass and metal [203], up to 48 hours on plastic surfaces [206], and 6 days in dried state [203].

The virus does not survive well after drying on paper, but lasts longer on disposable gowns compared to cotton, ones [202]. Human coronavirus 229E can remain infectious on high-touch environmental surfaces (polyvinylchloride, laminate, wood, stainless steel) for at least 7 days at ambient temperature (24°C) and relative humidity conditions (~50%) [207].
The specific features of COVID-19, which spreads very rapidly with a short incubation period and infects exponentially thousands of individuals in all age groups [208], calls for the implementation of specific containment measures as discussed in section 8.1.

8.3 Human resources, equipment and new approaches to clinical management

The COVID-19 pandemic is, first and foremost, a health crisis [209]. However, it is rapidly becoming also an economic crisis. In a vicious circle, the reduction in economic activities reduces money circulation, tax revenues and finances available for establishing the public-health countermeasures needed to control the pandemic. At the same time, social protection measures to ensure a minimum salary to the many workers who cannot be supported by their employers increases the financial constraints at the government level. The poverty generated by the economic crisis is likely to have medium and long-term consequences, particularly in resource-limited countries, with increase in malnutrition and poverty-related diseases which include TB.

To mitigate the consequences of this or future pandemics it is important countries develop specific plans with adequate human and financial resources [210]. This will prevent or limit resources currently reserved for other purposes (e.g. for TB programmes) which then get diverted to the emergency [211], including the shortage of PCR reagents being used for COVID-19 diagnosis which may impact on molecular TB diagnosis. Moreover, the emergency plan should be able to ensure rapid procurement and distribution of diagnostics, drugs, ventilators, masks, personal protective equipment and respirators needed to ensure an adequate response and adequate human resources [210, 212]. Telemedicine would be an important approach to deliver care, especially as a means to reduce the risk of cross-contamination caused by close contact [213].

To be effective, as part of an emergency response, telehealth would require to be routinely used in the health system. This would imply a change in the management and the redesign of existing models of care. Moreover a central system of controlling the pandemic is needed, for example in Italy, the Italian Civil Protection Department [214], which is normally dedicated to seismic hazard or natural disaster assessment and intervention. A central system will ensure prompt coordination of the emergency response and implementation of the emergency plan [212].

8.4 Impact of ‘fear’ of viral infections on health services and TB services

There are many factors affecting access to health care: affordability and physical/geographical accessibility are essential; socio-demographic factors (ethnicity, gender, age, marital and socioeconomic status) and psychological factors such as fear can significantly hinder or delay access to TB diagnosis and care.
Fear, defined as an instinctive emotional reaction to a specific, identifiable and immediate threat such as a dangerous animal, infection, deportation or imprisonment has a protective function associated with defensive behaviours such as hiding, fight or flight responses. Fear of TB itself, fear of discrimination – either self-stigma or fear of being stigmatised by others, including health care providers; fear of factors associated with health care such as the fear that receiving a diagnosis of TB or TB treatment could affect the way they are perceived by society and even lead to deportation or exclusion are well recognised barriers to timely access to care [215].

In addition, physical distancing measures imposed to reduce the transmission of COVID-19 (SARS-CoV-2) such as self-quarantine, closure of all non-essential services including small clinics, movement restrictions and limited access to public transport, police patrol and enforced isolation measures all have a potential deleterious impact on access to TB care. These factors affect all groups but disproportionately impact those minorities more often afflicted by TB such as migrants, refugee and asylum seekers, ethnic minorities or the poor. The effect on management of non-pandemic conditions including TB, strokes and myocardial infections is not only because of unwillingness of individuals to attend healthcare facilities for fear of catching infection, but also because anything with fever may wrongly be assumed to be caused by the pandemic organism.

9. Lessons on COVID-19 and TB: lessons learned and common solutions

9.1 Country response to COVID-19

The response of European Union (EU) and European Economic Area (EEA) countries and the UK to the COVID-19 epidemic is provided in Table 3[216]. By 3 April 2020, 25/31 (81%) countries had closed all educational institutions, higher education and secondary schools, primary schools, and day-care/nurseries; in some countries primary schools (2/31 [6%]) and/or day-care/nurseries (6/31 [19%]) remained open. Public spaces were closed in 30/31 (97%) countries, and includes closure of museums, theatres, cafes and restaurants, and gyms and sport facilities. Mass gatherings have been cancelled in all 31/31 countries. In some countries this means that indoor and outdoor gatherings of > 5 persons are forbidden and in others only larger gatherings of more than 1000 persons are banned. Finally, more than half of EU/EEA countries and the UK (18/31, 58%) implemented a full national lockdown, i.e. stay-at-home orders for regions or entire countries. Of the 13 countries without national lockdown, one issued stay-at-home recommendations to the general population, and nine issued stay-at-home recommendations for risk groups. The assessment of the response shows that there has been a lack of uniformity in the implementation of different measures. An analysis of the efficacy of those at the end of the emergency should guide future policies.
9.2 TB and COVID-19: interactions

There are multiple interactions at the different levels of prevention, diagnosis and treatment between the responses to TB and COVID-19 [135, 136, 217]. Similarities and differences related to infection control and workplaces safety have been discussed.

For diagnosis, the possibility of using a platform like Xpert offers synergies, although the risk is that during the COVID-19 emergency the existing equipment is diverted from TB diagnosis. Personal protective equipment is needed to protect laboratory personnel handling viral specimens [135, 136].

Cough is the pivotal symptom to diagnose TB. In the absence of adequate diagnostic tools, the presence of cough, fever and other non-specific symptoms complicates differential diagnosis of TB, COVID-19 and other respiratory infections [135].

The necessary physical distancing policies are likely to negatively affect active case finding as well as community-based activities which are important to manage TB (and HIV) in high TB burden countries [217]. However, they may reduce transmission of TB, which has already been shown for influenza [218, 219]. Both clinical and programmatic approaches to TB treatment are also affected in high–burden countries and drug procurement (particularly for second-line drugs) is likely to suffer as well.

Last but not least, the expected major economic crisis which will follow the COVID-19 pandemic may result in increased poverty, social disturbances, and malnutrition with a profound impact increasing TB incidence and mortality [220-222].

9.3 What we can and cannot learn from TB programmes in the response to COVID-19 pandemic

Political commitment, strategic planning, community mobilisation, research and development are key elements in the battle against either TB or COVID-19. While a sustainable and effective treatment programme is the key to successful control of TB, good pandemic preparation with adequate emergency response capacity is needed when confronted with viruses with pandemic potential such as SARS-CoV-2 or influenza. Early and determined action to contain the COVID-19 epidemic at its localized or importation stage will give more time for reinforcement of care facilities, manpower and supplies [223]. Triage of suspects and patients according to their isolation and treatment needs will help to maximize the healthcare throughput with limited resources. Simultaneous measures to contain the spread of the epidemic is necessary to limit the mounting patient load and avoid major breaches in nosocomial infection control and total collapse of the healthcare system.
Breaking the transmission link is the main method for containing an infectious disease in the absence of an effective vaccine. Controlling a respiratory infection at source is often more cost-effective than targeting multiple intervention points downstream for environmental control or personal protection [224]. For TB, wearing a surgical mask can reduce the transmission by 56% [225]. The evidence for the protective effect of cloth masks is limited and contradicting [196, 226]. For both diseases, early case detection is important for source control, but case finding may be delayed by inconspicuous or non-specific symptoms [135]. TB can be stopped at source by effective treatment. Isolation for the whole infectious period is needed to contain the spread of COVID-19. Like TB, contact tracing may only pick up a small portion of COVID-19 cases when widespread local transmission occurs with unclear transmission links [227]. Physical distancing [228], and adjunctive use of face masks for unavoidable person-to-person contact are then the only effective measures to slow the otherwise exponential growth of the COVID-19 epidemic [194]. The current physical distancing and stay-at-home measures due to the COVID-19 pandemic will pose challenges to TB programmes to provide the necessary diagnosis, treatment and care for the people and communities affected by TB. To avoid disruption of TB services innovative approaches (such as virtual care, digital health, and community-monitoring solutions, etc.) will be necessary [197].

10. Consensus statements

The consensus statements derived from the Delphi process and the level of agreement achieved are summarised in Table 4. The ECDC PHE did not participate in the Delphi.

The scores, from the 37 questionnaires received, exceeded 3 (from intermediate to highest score) in all the 18 statements, ranging from 3.1 to 4.8 out of a maximum of 5. The lowest scores were found for the statements on hydroxychloroquine/chloroquine and possible protective effect of BCG (Statements 5 and 12), where less evidence is available. The highest scores were found for the statements on risk factors, outcomes and transmission (Statements 6 and 7) and for public health and prevention measures (Statements 13 and 14).

The majority of experts scored 4 or 5 (high scores) in all statements except Statements 5 and 12. The Delphi process suggests the experts tended to agree on the relevance of the statements where more solid and high-quality evidence is available.

11. Conclusions

Altogether in this consensus-based document we describe the effects of epidemic and pandemic viral infections (SARS, MERS, influenza A(H1N1)pdm09, HIV and COVID-19) and the interactions with TB and the lung, the majority of these diseases are droplet-borne. The diagnostic tools range from nucleic acid detection and molecular techniques, to immunoassays, and traditional cultures for Mtb. Core management
issues were discussed, including drugs, drug-drug interactions, novel therapies, and principles of infection control and workplace safety. COVID-19 and TB interactions were discussed in-depth based on the scanty information available so far, which requires broad and in-depth research into the subject. New approaches to clinical management and country responses specifically to COVID-19 were elaborated by the expert panel, as well as opportunities and lessons for future responses.
Acknowledgements

The co-Authors of the World Association of Infectious Diseases and Immunological Disorders (WAidid) and Global Tuberculosis Network (GTN) are: Jan-Willem Alffenaar, Emmanuelle Cambau, Rosella Centis, Daniela M. Cirillo, Lia D'Ambrosio, Masoud Dara, Giovanni Delogu, Susanna M.R. Esposito, Jose Figueroa, Delia Goletti, Catherine Wei Min Ong, Giovanni Battista Migliori, Emanuele Pontali, Mario Raviglione, Giovanni Sotgiu, Antonio Spanevello, Simon Tiberi, Martin van den Boom, Dina Visca, Jean Pierre Zellweger.

Out of 40 experts responding to the invitation, 38 (95%) endorsed the document as follows:

a) From the Global Tuberculosis Network (GTN):
Onno W. Akkerman (The Netherlands); Francois-Xavier Blanc (France); Sergey Borisov (Russian Federation); Anna Cristina Carvalho (Brazil); Muhwa J. Chakaya (Kenya); Margareth Dalcolmo (Brazil); Martin Enwerem (South Africa); Alberto Garcia-Basteiro (Spain); José-María García-García (Spain); Rafael Laniado-Laborín (Mexico); Selene Manga (Peru); Ekaterina Manika (Greece); Alessandro V. Mariani (Brazil); Andrey Maryandyshev (Russian Federation); Alberto Matteelli (Italy); Fernanda Mello (Brazil); Moschos Charalampos (Greece); Luis Adrian Rendon (Mexico); Apostolos Papavasiliou (Greece); Alberto Pibello (Niger); Denise Rossato Silva (Brazil); Isabel Saraiva (Portugal); Barbara Seaworth (USA); Ivan Solovic (Slovakia); Simon Tiberi (United Kingdom); Zarir Udwadia (India); Andrey Zagorski (USA); Richard Zaleskis (Latvia).

b) From the ESCMID Study Group for Mycobacterial Infections (ESGMYC):
Ana-Gil Brusola (Spain), Danilo Buonsenso (Italy), Paola Di Carlo (Italy), Francis Drobniewski (United Kingdom), Sally Hargreaves (United Kingdom), Laura Nellums (United Kingdom), Cristina Russo (Italy), Thomas Schön (Sweden), Ulrika Simonsson (Sweden), Cristina Vilaplana (Spain).

Sir Alimuddin Zumla is a co-PI of the Pan-African Network on Emerging and Re-Emerging Infections (PANDORA-IDNET; https://www.pandora-id.net/) funded by the European and Developing Countries Clinical Trials Partnership the EU Horizon 2020 Framework Programme for Research and Innovation. Sir Zumla is also in receipt of a National Institutes of Health Research senior investigator award.

Catherine Wei Min Ong is funded by Singapore National Medical Research Council (NMRC/TA/0042/2015, CSAINV17nov014); National University Health System (NUHS/RO/2017/092/SU/01, CFGFY18P11, NUHSRO/2020/042/RO5+5/ad-hoc/1), Singapore; National Centre for Infectious Diseases, Singapore (STPRG-FY19-003); and recipient of the Young Investigator Award, Institut Merieux, Lyon, France.

Masoud Dara, Martin van den Boom, A.b.m. Tauhidul Islam from the World Health Organization read and critically reviewed the manuscript.
Anne-Grete Märtson was funded by Marie Skłodowska-Curie Actions, Grant Agreement number: 713660—PRONKJEWAIL—H2020-MSCA-COFUND-2015.

Conflict of interest: All other authors declare no conflict of interest.

The article is part of the activities of the Global Tuberculosis Network (GTN); and of the WHO Collaborating Centre for Tuberculosis and Lung Diseases, Tradate, ITA-80, 2017-2020- GBM/RC/LDA.).

Part of the work was supported by the Italian Ministry of Health, Ricerca Corrente (Linea 1 and Linea 3), Italian Ministry of Health GR-2018-12367178, Italian Ministry of Health GR-2016-02364014.
**Figure legend**

**Figure 1.** The lungs and gut are exposed to environmental substances and pathogens. Early protection response to respiratory viruses includes mucous, surfactants and antiviral peptides that can prevent initial attachment and viral entry. Respiratory viruses enter via the respiratory epithelium. Epithelial cells have a key role in initiating the immune response by recognizing viral components -Pathogen Associated Molecular Patterns (PAMPs)- via TLRs and intracellular receptors. These cellular sensors trigger a signaling cascade resulting in the upregulation of Type I and III interferons and the inflammatory response. This leads to differentiation of dendritic cells (DCs) that mediate the induction of the adaptive immunity, and promote the recruitment of innate immunity cells, in particular neutrophils and natural killer (NK) cells. NK cells have the ability to kill virus-infected cells via perforin-granzyme–dependent mechanisms or by the Fas-FasL pathway. Moreover, alveolar macrophages, recruited monocytes and macrophages as well as DCs, pick pathogen components and contribute to the immune response. All of these cells produce cytokines and chemokines that are important for the establishment of the adaptive responses and of the anti-viral state. The adaptive response to respiratory viruses is mediated by both T and B-cell compartments. T-cells contribute to the generation of the B-cell response. B-cells produce antibodies that may neutralise the respiratory viruses directly by binding to viral surface proteins that are essential for entry of the virus into host cells or through the ligation of Fc receptors to trigger the complement cascade and antibody-dependent cell-mediated cytotoxicity. Antibodies are in the form of IgA, mainly in the upper respiratory tract, or IgG in the lower respiratory tract. Viral clearance is also mediated by CD8+-specific T-cells with cytolitic activity. Protective anti-viral T-cell response is mainly mediated by the IFN-γ production and is therefore biased toward a Th-1 response whereas other T-cell subsets as Th2 cells and Th17 cells play a minor role and they may be responsible for lung tissue damage. Moreover, regulatory mechanisms adopted by T-cells such as IL-10 secretion, or upregulation of inhibitory receptors such as programmed cell death protein 1 (PD-1) or expansion of the T-regulatory cell subsets work to balance tissue damage and viral clearance.

**Figure 2.** Proposed mechanism of action of drugs used for SARS-CoV-2. SARS-CoV-2 can enter the cell through ACE2 and TMPRSS2. Camostat mesylate acts as an inhibitor of TMPRSS2 and umifenovir can inhibit the viral entry to the cell [181, 229, 230]. Chloroquine, hydroxychloroquine and baricitinib mechanism of action is not fully understood, however it is proposed that these drugs affect the viral entry. Baricitinib also inhibits the AP-2-associated protein kinase [174, 181, 231]. Lopinavir/ritonavir and ASC-09/ritonavir as protease inhibitors inhibit the proteolysis. Lopinavir/ritonavir inhibits specifically the proteinase 3CLpro [232]. Ribavirin and favipiravir both have wide antiviral activity and have the potential to inhibit SARS-CoV-2 RNA replication [233-235]. Azvudine, a nucleoside reverse transcriptase inhibitor, also inhibits the RNA replication [236]. A probable mechanism of action for baloxavir marboxil is the inhibition of transcription through inhibiting cap-dependent endonuclease [237]. Favipiravir and remdesivir, inhibit the RNA-dependent RNA polymerase, which results in reducing RNA synthesis [181, 234, 235, 238].
|                          | Respiratory viruses                                                                 | M. tuberculosis complex                                      |
|--------------------------|-------------------------------------------------------------------------------------|-------------------------------------------------------------|
|                          | Sensitivity | Specificity | Time to result* | User-friendly | Unknown or uncommon viruses | Sensitivity | Specificity | Time saving | User-friendly | Drug susceptibility testing |
| **Molecular diagnosis**  |             |             |                |               |                             |             |             |             |               |                          |
| Manual NAAT              | +++         | +++         | -              | -             | -                            | +++         | ++          | +           | -             | +                        |
| Automated NAAT           | +++         | +++         | +              | +             | -                            | +++         | +++         | ++          | ++            | ++                       |
| POCT-NAAT                | +           | ++          | +++            | +++           | -                            | +++         | +++         | +++          | +             | +                        |
| NGS (metagenomics and WGS)| -           | -           | -              | -             | ++                           | +           | +++         | -           | -             | +++                      |
| **Microscopy**           |             |             |                |               |                             |             |             |             |               |                          |
| IFA or ZN/AR¹            | -           | ++          | -              | -             | -                            | +           | +           | ++          | +             | -                        |
| **Culture²**             |             |             |                |               |                             |             |             |             |               |                          |
| POCT Ag                  | -           | +           | +++            | ++            | -                            | +¹          | ++          | +++         | +++           | NA                       |
| ELISA                    | ++          | ++          | -              | -             | -                            | NA          | NA          | NA          | NA            | NA                       |
| **Immunodiagnosis**      |             |             |                |               |                             |             |             |             |               |                          |
| Serology                 | NRU         | NRU         | -              | -             | -                            | Not recommended | Not recommended | NRU | NRU | NA |
| IFNγ release assays      | ES          | ES          | -              | -             | -                            | ++          | +           | ++          | +             | NA                       |
| Skin test⁴               | NA          | NA          | NA             | NA            | NA                           | ++          | +           | ++          | ++            | NA                       |

Abbreviations: NAAT: Nucleic Acid Amplification Tests; POCT: Point of Care Test; NGS: Next Generation Sequencing; WGS: whole genome sequencing; NA: Not applicable; NRU: Not routinely used; ES: Experimental settings only.¹ IFA: Immunofluorescence microscopy on respiratory samples to detect the most common viruses; ZN/AR: Ziehl-Neelsen or Auramine/Rhodamine staining to detect acid fast bacilli; ²Viral culture are established in several eukaryotic cell lines. Mycobacterial culture in liquid or solid media; ³The only approved POCT Ag for TB detects LAM in urine samples and has been licensed to diagnose TB in HIV-infected patients and to monitor therapy; ⁴Mantoux test; Quantitation: - very poor; + poor; ++: good; +++: excellent. °Considering only the time of the procedure: less than 2 hours.
Table 2. Drug interactions between TB and potential COVID-19 medications.

| WHO First line TB drugs | Second line TB drugs: Group A | Group B | Group C |
|--------------------------|-------------------------------|---------|---------|
| INH | RIF | EMB | PZA | LFX | MFX | BDQ | LZD | CFZ | Cs | DLM | IMI/CIS | MEM | AMI | STR | ETO | PTO | PAS |

**Antivirals**

| Atazanavir [239] | — | ▼ X | — | — | — | ▼ ▼ | ▼ ▼ | — | — | ▼ ▼ | — | — | — | — | — | — |
| Baloxavir marboxil [240] | — | — | — | — | — | △ △ | △ △ | — | — | △ △ | — | — | — | — | — | — |
| Favipiravir [241, 242] | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — |
| Galidesivir n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a | n/a |
| Lopinavir/ Ritonavir [243, 244] | — | ▼ X | — | — | — | ▼ ▼ | ▼ ▼ | — | — | ▼ ▼ | — | — | — | — | — | — |
| Oseltamivir | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — |
| Remdesivir [243] | — | ▼ X | — | — | — | — | — | — | — | — | △ △ | — | — | — | — | — | — |
| Ribavirin [243] | — | — | — | — | — | — | — | — | — | △ △ | — | — | — | — | — | — | — |
| Umifenovir [245, 246] | — | ▼ X^* | — | — | — | △ △ | △ △ | — | — | △ △ | — | — | — | — | — | — |

**Antibacterials**

| Azithromycin [247] | — | — | — | — | — | ▼ ▼ | ▼ ▼ | — | — | ▼ ▼ | — | — | — | — | — | — |

**Antiprotozoal**

| Chloroquine [248-250] | — | ▼ X | — | — | — | ▼ ▼ | ▼ ▼ | — | — | ▼ ▼ | — | — | — | — | — | — |
| Nitazoxanide | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — |

**Immunomodulating drugs**

| Anakinra | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — |
| Baricitinib | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — |
| Hydroxychloroquine [248, 251] | — | ▼ X | — | — | — | ▼ ▼ | ▼ ▼ | — | — | ▼ ▼ | — | — | — | — | — | — |
| Interferons [252-254] | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — |
| Tocilizumab [243, 255] | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — | — |
Interaction symbols:
- Decreased exposure to the TB drug + Action required (action: dose adjustment or monitoring).
- Increased exposure to the TB drug + Action required.
- Decreased exposure to the COVID-19 drug + Action required.
- Increased exposure to the COVID-19 drug + Action required.

-no significant interaction predicted based on metabolic pathway (does not mean absence of interaction).
Δ possible interaction based on metabolism and clearance, but no specific data available.

n/a no available pharmacokinetic data.

Monitoring/action symbols:
- ♥ requires ECG monitoring due to the risk of QT and/or PR prolongation, or other cardiac abnormalities.
- ♦ requires full blood count (FBC) monitoring
- U uric acid monitoring
- Monitor for potential seizures (rare)
- Monitor for ototoxicity
- X should not be administered together.

Mechanism symbols:
- Cytochrome P450 mediated mechanism
- UGT enzyme-mediated glucuronidation

Additional information:
a. recommended based on predicted interaction.
b. UGT 1A1 is involved in moxifloxacin metabolism and could be involved in umifenovir metabolism (mainly UGT 1A9).
c. Both drugs are metabolized by CYP3A4.
d. CYP3A4 is involved in the metabolism of baloxavir (minor extent) and umifenovir, and clofazimine is a CYP3A4 inhibitor.
e. both drugs primarily undergo renal excretion.

Abbreviations:
WHO: World Health Organization, INH: isoniazid, RIF: rifampicin, EMB: ethambutol, PZA: pyrazinamide, LFX: levofloxacin, MFX: moxifloxacin, BDQ: bedaquiline, LZD: linezolid, CFZ: clofazimine, Cs: cycloserine, DLM: delamanid, IMI/CIS: imipenem/cilastin, MEM: meropenem, AMI: amikacin, STR: streptomycin, ETO: ethionamide, PTO: prothionamide, PAS: p-aminosalicylic acid, LPV/r: lopinavir/ritonavir, HCLQ: hydroxychloroquine, CLQ: chloroquine, RDV: remdesivir
Table 3: Response measures undertaken in EU/EEA Member States and the UK at the national level as of 3 April 2020*

| Country          | Higher education/Secondary School | Primary School | Day-care or nursery | Closure of public spaces | Mass gathering cancellations | Stay-at-home order (enforced) | Stay-at-home recommendation (general population) | Stay-at-home recommendation (risk group) |
|------------------|----------------------------------|----------------|---------------------|--------------------------|----------------------------|-------------------------------|-----------------------------------------------|-----------------------------------|
| Austria          | X                                 | X              | X                   | X                        | X                          | X                             |                                 | X                                 |
| Belgium          | X                                 | X              | X                   | X                        | X                          | X                             |                                 | X                                 |
| Bulgaria         | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Croatia          | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Cyprus           | X                                 | X              | X                   | X                        | X                          | X                             |                                 | X                                 |
| Czech Republic   | X                                 | X              | X                   | X                        | X                          | X                             |                                 | X                                 |
| Denmark          | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Estonia          | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Finland          | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| France           | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Germany          | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Greece           | X                                 | X              | X                   | X                        | X                          | X                             |                                 | X                                 |
| Hungary          | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Iceland          | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Ireland          | X                                 | X              | X                   | X                        | X                          | X                             |                                 | X                                 |
| Italy            | X                                 | X              | X                   | X                        | X                          | X                             |                                 | X                                 |
| Latvia           | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Liechtenstein    | X                                 | X              | X                   | X                        | X                          | X                             |                                 | X                                 |
| Lithuania        | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Luxembourg       | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Malta            | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Netherlands      | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Norway           | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Poland           | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Portugal         | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Romania          | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Slovakia         | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Slovenia         | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| Spain            | X                                 | X              | X                   | X                        | X                          | X                             |                                 | X                                 |
| Sweden           | X                                 | X              | X                   | X                        | X                          |                               |                                 | X                                 |
| United Kingdom   | X                                 | X              | X                   | X                        | X                          | X                             |                                 | X                                 |

* The data on response measures are based on information available from official public sources as of Friday 3 April 2020 at 18:00 and may not capture measures being taken by countries that are not reported on publicly available websites. The situation is evolving rapidly and this represents a
A snapshot of the measures that countries in the EU/EEA and the UK have reported to date. The response measures displayed are national measures, reported on official public websites. Response measures collected include: mass gathering cancellations (for specific events or a ban on gatherings of a particular size); closure of public spaces (including restaurants, entertainment venues, non-essential shops and so on); closure of educational institutions (including day-care or nursery, primary schools, and secondary schools and higher education); stay-at-home recommendations for risk groups or vulnerable populations (such as the elderly, people with underlying health conditions, physically disabled people etc.); stay-at-home recommendations for the general population (which are voluntary or not enforced); and stay-at-home orders for the general population (these are enforced and also referred to as ‘lockdown’). The data on response measures has several limitations. Firstly, there is substantial heterogeneity in physical distancing policies and their implementation between countries. For instance, the level of enforcement of measures may vary between countries and there may be specific rules and exceptions to the measures, making interpretation of the data challenging. The measures displayed in these figures are measures reported at national level and it should be noted that due to the evolution of the outbreak in certain regions, regional or local measures often preceded national ones. The exact dates of introduction were often available from official sources but delays in their implementation may have occurred. Additionally, availability of public data from official government sources varies among countries. For some countries, data are no longer available on official websites concerning measures that are no longer in force, which may result in the data for more recent measures being more complete.
Table 4. Consensus statements derived from the Delphi process

| Consensus statements                                                                 | 1 (lowest relevance) | 2       | 3       | 4       | 5 (highest relevance) | Mean (SD) |
|--------------------------------------------------------------------------------------|----------------------|---------|---------|---------|-----------------------|-----------|
| 1. Large droplets increase the risk of respiratory viral infection through direct transmission. | 0 (0.0)              | 6 (16.2)| 6 (16.2)| 10 (27.0)| 15 (40.5)             | 3.9 (1.1) |
| 2. Respiratory viral infections are more likely to occur in older patients (with or without comorbidities) and infants. | 1 (2.7)              | 5 (13.5)| 10 (27.0)| 12 (32.4)| 9 (24.3)              | 3.6 (1.1) |
| 3. Elderly patients are more likely to develop acute respiratory distress syndrome and there is an age-related death risk. | 0 (0.0)              | 0 (0.0) | 4 (10.8)| 10 (27.0)| 23 (62.2)             | 4.5 (0.7) |
| 4. Antibodies might neutralize respiratory viruses and, then, decrease the risk of recurrent infections. | 0 (0.0)              | 4 (10.8)| 7 (18.9)| 17 (46.0)| 9 (24.3)              | 3.8 (0.9) |
| 5. BCG-vaccination might offer protection against COVID-19. RCTS are needed.         | 4 (10.8)             | 7 (18.9)| 10 (27.0)| 10 (27.0)| 6 (16.2)              | 3.2 (1.2) |
| 6. Severe COVID-19 is associated with rapid virus replication, massive inflammatory cell infiltration in the lung, and elevated pro-inflammatory cytokine/chemokine response. | 0 (0.0)              | 0 (0.0) | 1 (2.7) | 7 (18.9) | 29 (78.4)             | 4.8 (0.5) |
| 7. High initial SARS-CoV-2 load in the airways, older age (≥65 years), and comorbidities of the infected individual are associated with worse COVID-19 outcome and thus patients with these risk factors need close attention. | 0 (0.0)              | 0 (0.0) | 3 (8.1) | 7 (18.9) | 27 (73.0)             | 4.7 (0.6) |
| 8. The combination of CT scan findings (ground-glass opacity and consolidation), clinical presentation respiratory parameters (peripheral capillary oxygen saturation (SpO2) and PaO2/FiO2 ), and blood tests (C-reactive proteins, lymphocyte number, fibrinogen, D-dimers, IL-6 ) helps identifying COVID-19 patients at highest risk for ICU transfer. | 0 (0.0)              | 1 (2.7) | 0 (0.0) | 10 (27.0)| 26 (70.3)             | 4.6 (0.6) |
| 9. CD4 T-cell counts is key to guide the aetiological evaluation of lung infections in HIV-infected individuals. | 0 (0.0)              | 3 (8.1) | 7 (18.9)| 18 (48.7)| 9 (24.3)              | 3.9 (0.9) |
| 10. Temporary immunosuppression induced by TB might increase the susceptibility to influenza viruses. | 2 (5.4)              | 6 (16.2)| 8 (21.6)| 18 (32.5)| 9 (24.3)              | 3.5 (1.2) |
| 11. An excess mortality associated with influenza is found among TB patients.        | 1 (2.7)              | 5 (13.5)| 8 (21.6)| 15 (40.5)| 8 (21.6)              | 3.7 (1.1) |
| 12. Chloroquine and hydroxychloroquine have potential to improve the treatment success rate of COVID-19 patients. RCTS are needed. | 4 (10.8)             | 8 (21.6)| 10 (27.0)| 9 (24.3) | 6 (16.2)              | 3.1 (1.3) |
| 13. Public and social distancing reduce the risk of SARS-CoV-2 transmission.         | 0 (0.0)              | 0 (0.0) | 3 (8.1) | 6 (16.2) | 28 (75.7)             | 4.7 (0.6) |
| 14. Appropriate use of facial masks (surgical masks in the general population; N95 for HCWs performing aerosol- | 0 (0.0)              | 0 (0.0) | 1 (2.7) | 9 (24.3) | 27 (73.0)             | 4.7 (0.5) |
producing activities) on symptomatic patients and their contacts can reduce the risk of SARS-CoV-2 infection by limiting the spread of droplet nuclei from isolated symptomatic patients.

15. SARS-CoV-2 virus remain infectious in the environment on different surfaces for days.

|   | 0 (0.0) | 5 (13.5) | 7 (18.9) | 5 (13.5) | 20 (54.1) | 4.1 (1.1) |
|---|---------|----------|----------|----------|-----------|-----------|

16. Social protection measures and specific national centralized emergency plans can reduce the healthcare and socio-economic burden of respiratory viral infections resulting in epidemics/pandemics.

|   | 0 (0.0) | 0 (0.0) | 4 (10.8) | 14 (37.8) | 19 (51.4) | 4.4 (0.7) |
|---|---------|---------|----------|-----------|-----------|-----------|

17. Stigma and social discrimination affect all virus-infected population groups but disproportionately the minorities.

|   | 0 (0.0) | 5 (13.5) | 7 (18.9) | 12 (32.4) | 13 (35.1) | 3.9 (1.1) |
|---|---------|----------|----------|-----------|-----------|-----------|

18. Late implementation of national lockdown can itself alone be effective in reducing the burden of COVID-19 but it has serious impact on the society and the economy.

|   | 3 (8.1) | 3 (8.1) | 6 (16.2) | 8 (21.6) | 17 (46.0) | 3.9 (1.3) |
References

1. WHO. Global tuberculosis report 2019. 2019.
2. Migliori GB, Tiberi S, Zumla A, Petersen E, Chakaya JM, Wejse C, Munoz Torrico M, Duarte R, Alffenaar JW, Schaff HS, Marais BJ, Cirillo DM, Alagna R, Rendon A, Pontali E, Piubello A, Figueroa J, Ferlazzo G, Garcia-Basteiro A, Centis R, Visca D, D’Ambrosio L, Sotgiu G, members of the Global Tuberculosis N. MDR/XDR-TB management of patients and contacts: Challenges facing the new decade. The 2020 clinical update by the Global Tuberculosis Network. Int J Infect Dis 2020: 92S: S15-S25.
3. Yan J, Grantham M, Pantelic J, Bueno de Mesquita PJ, Albert B, Liu F, Ehrman S, Milton DK, Consortium E. Infectious virus in exhaled breath of symptomatic seasonal influenza cases from a college community. Proc Natl Acad Sci U S A 2018: 115(5): 1081-1086.
4. Alonso C, Raynor PC, Goyal S, Olson BA, Alba A, Davies PR, Torremorell M. Assessment of air sampling methods and size distribution of virus-laden aerosols in outbreaks in swine and poultry farms. J Vet Diagn Invest 2017: 29(3): 298-304.
5. Blachere FM, Lindsley WG, Weber AM, Beezhold DH, Thewlis RE, Mead KR, Noti JD. Detection of an avian lineage influenza A(H7N2) virus in air and surface samples at a New York City feline quarantine facility. Influenza Other Respir Viruses 2018: 12(5): 613-622.
6. Leung NH, Zhou J, Chu DK, Yu H, Lindsley WG, Beezhold DH, Yen HL, Li Y, Seto WH, Peiris JS, Cowling BJ. Quantification of Influenza Virus RNA in Aerosols in Patient Rooms. PLoS One 2016: 11(2): e0148669.
7. Milton DK, Fabian MP, Cowling BJ, Grantham ML, McDevitt JJ. Influenza virus aerosols in exhaled breath: particle size, culturability, and effect of surgical masks. PLoS Pathog 2013: 9(3): e1003205.
8. Lewis D. Is the coronavirus airborne? Experts can’t agree. [News] 2020 [cited 18 April 2020]; Available from: https://www.nature.com/articles/d41586-020-00974-w
9. van Doremalen N, Bushmaker T, Morris DH, Holbrook MG, Gamble A, Williamson BN, Tamin A, Harcourt JL, Thornburg NJ, Gerber SI, Lloyd-Smith JO, de Wit E, Munster VJ. Aerosol and Surface Stability of SARS-CoV-2 as Compared with SARS-CoV-1. N Engl J Med 2020: 382(16): 1564-1567.
10. Arnold FW, Fuqua JL. Viral respiratory infections: a cause of community-acquired pneumonia or a predisposing factor? Curr Opin Pulm Med 2020: 26(3): 208-214.
11. Jain S. Epidemiology of Viral Pneumonia. Clin Chest Med 2017: 38(1): 1-9.
12. Varga Z, Flammer AJ, Steiger P, Haberecker M, Andermatt R, Zinkernagel AS, Mehra MR, Schuepbach RA, Ruschitzka F, Moch H. Endothelial cell infection and endotheliitis in COVID-19. Lancet 2020: 395(10234): 1417-1418.
13. Jain S, Self WH, Wunderink RG, Fakhran S, Balk R, Bramley AM, Reed C, Grijalva CG, Anderson EJ, Courney DM, Chappell JD, Qi C, Hart EM, Carroll F, Trabue C, Donnelly HK, Williams DJ, Zhu Y, Arnold SR, Ampofo K, Waterer GW, Levine M, Lindstrom S, Winchell JM, Katz JM, Erdman D, Schneider E, Hicks LA, McCullers JA, Pavia AT, Edwards KM, Finelli L, Team CES. Community-Acquired Pneumonia Requiring Hospitalization among U.S. Adults. N Engl J Med 2015: 373(5): 415-427.
14. Jain S, Williams DJ, Arnold SR, Ampofo K, Bramley AM, Reed C, Stockmann C, Anderson EJ, Grijalva CG, Self WH, Zhu Y, Patel A, Hymas W, Chappell JD, Kaufman RA, Kan JH, Dansie D, Lenny N, Hillyard DR, Haynes LM, Levine M, Lindstrom S, Winchell JM, Katz JM, Erdman D, Schneider E, Hicks LA, Wunderink RG, Edwards KM, Pavia AT, McCullers JA, Finelli L, Team CES. Community-acquired pneumonia requiring hospitalization among U.S. children. N Engl J Med 2015: 372(9): 835-845.
15. Saha S, Chadha M, Al Mamun A, Rahman M, Sturm-Ramirez K, Chittaganpitch M, Pattamadilok S, Olsen SJ, Sampurono OD, Setiawaty V, Pangesthi KN, Samaan G, Archkhawongs S, Vongphrachanh P, Phoneko D, Corwin A, Touch S, Buchy P, Chea N, Kitsutani P, Mai le Q, Thiem VD, Lin R, Low C, Kheong CC, Ismail N, Yusof MA, Tandoc A, 3rd, Roque V Jr., Mishra A, Moen AC, Widdowson MA, Partridge J, Lal RB. Influenza seasonality and vaccination timing in tropical and subtropical areas of southern and southern-eastern Asia. *Bull World Health Organ* 2014: 92(5): 318-330.

16. Chadha MS, Potdar VA, Saha S, Koul PA, Broor S, Dar L, Chawla-Sarkar M, Biswas D, Gunasekaran P, Abraham AM, Shrikhande S, Jain A, Anukumar B, Lal RB, Mishra AC. Dynamics of influenza seasonality at sub-regional levels in India and implications for vaccination timing. *PLoS One* 2015: 10(5): e0124122.

17. Madhi SA, De Wals P, Grijalva CG, Grimwood K, Grossman R, Ishiwada N, Lee PI, Nascimento-Carvalho C, Nohynek H, O'Brien KL, Vergison A, Wolter J. The burden of childhood pneumonia in the developed world: a review of the literature. *Pediatr Infect Dis J* 2013: 32(3): e119-127.

18. Vareille M, Kieninger E, Edwards MR, Regamey N. The airway epithelium: soldier in the fight against respiratory viruses. *Clin Microbiol Rev* 2011: 24(1): 210-229.

19. Ong CW, Elkington PT, Friedland JS. Tuberculosis, pulmonary cavitation, and matrix metalloproteinases. *Am J Respir Crit Care Med* 2014: 190(1): 9-18.

20. Jegaskanda S, Reading PC, Kent SJ. Influenza-specific antibody-dependent cellular cytotoxicity: toward a universal influenza vaccine. *J Immunol* 2014: 193(2): 469-475.

21. Gupta N, LeGoff J, Chamat S, Mercer-Delarue S, Touzelet O, Power UF, Kazatchkine MD, Simon F, Lacroix-Desmazes S, Bayry J, Kaveri SV. Affinity-purified respiratory syncytial virus antibodies from intravenous immunoglobulin exert potent antibody-dependent cellular cytotoxicity. *PLoS One* 2013: 8(7): e69390.

22. Openshaw PJ, Chiu C. Protective and dysregulated T cell immunity in RSV infection. *Curr Opin Virol* 2013: 3(4): 468-474.

23. Krishnamoorthy N, Khare A, Oriss TB, Raundhal M, Morse C, Yarlagadda M, Wenzel SE, Moore ML, Peebles RS, Jr., Ray A, Ray P. Early infection with respiratory syncytial virus impairs regulatory T cell function and increases susceptibility to allergic asthma. *Nat Med* 2012: 18(10): 1525-1530.

24. Byström J, Al-Adhoubi N, Al-Bogami M, Jawad AS, Mageed RA. Th17 lymphocytes in respiratory syncytial virus infection. *Viruses* 2013: 5(3): 777-791.

25. Chiu C, Openshaw PJ. Antiviral B cell and T cell immunity in the lungs. *Nat Immunol* 2015: 16(1): 18-26.

26. Abril-Rodriguez G, Ribas A. SnapShot: Immune Checkpoint Inhibitors. *Cancer Cell* 2017: 31(6): 848-848 e841.

27. Simon AK, Hollander GA, McMichael A. Evolution of the immune system in humans from infancy to old age. *Proc Biol Sci* 2015: 282(1821): 20143085.

28. Olin A, Henckel E, Chen Y, Lakshmikanth T, Pou C, Mikes J, Gustafsson A, Bernhardsson AK, Zhang C, Bohlin K, Brodin P. Stereotypic Immune System Development in Newborn Children. *Cell* 2018: 174(5): 1277-1292 e1214.

29. Chackerian AA, Alt JM, Perera TV, Dascher CC, Behar SM. Dissemination of Mycobacterium tuberculosis is influenced by host factors and precedes the initiation of T-cell immunity. *Infect Immun* 2002: 70(8): 4501-4509.

30. Wolf AJ, Desvignes L, Linas B, Banaiee N, Tamura T, Takatsu K, Ernst JD. Initiation of the adaptive immune response to Mycobacterium tuberculosis depends on antigen production in the local lymph node, not the lungs. *J Exp Med* 2008: 205(1): 105-115.

31. Reiley WW, Calayag MD, Wittmier ST, Huntington JL, Pearl JE, Fountian JJ, Martino CA, Roberts AD, Cooper AM, Winslow GM, Woodland DL. ESAT-6-specific CD4 T cell
responses to aerosol Mycobacterium tuberculosis infection are initiated in the mediastinal lymph nodes. *Proc Natl Acad Sci U S A* 2008: 105(31): 10961-10966.

32. Kumar A, Farhana A, Guidry L, Saini V, Hondalus M, Steyn AJ. Redox homeostasis in mycobacteria: the key to tuberculosis control? *Expert Rev Mol Med* 2011: 13: e39.

33. Ong CWM, Fox K, Ettorre A, Elkinson PT, Friedland JS. Hypoxia increases neutrophil-driven matrix destruction after exposure to Mycobacterium tuberculosis. *Sci Rep* 2018: 8(1): 11475.

34. Dwivedi VP, Bhattacharya D, Chatterjee S, Prasad DV, Chattopadhyay D, Van Kaer L, Bishai WR, Das G. Mycobacterium tuberculosis directs T helper 2 cell differentiation by inducing interleukin-1beta production in dendritic cells. *J Biol Chem* 2012: 287(40): 33656-33663.

35. Petruccioli E, Scriba TJ, Petrone L, Hatherill M, Cirillo DM, Joosten SA, Ottenhoff TH, Denkinger CM, Goletti D. Correlates of tuberculosis risk: predictive biomarkers for progression to active tuberculosis. *Eur Respir J* 2016: 48(6): 1751-1763.

36. Goletti D, Lindestam Arlehamn CS, Scriba TJ, Anthony R, Cirillo DM, Alonzi T, Denkinger CM, Cobelens F. Can we predict tuberculosis cure? What tools are available? *Eur Respir J* 2018: 52(5).

37. Lin PL, Flynn JL. CD8 T cells and Mycobacterium tuberculosis infection. *Semin Immunopathol* 2015: 37(3): 239-249.

38. Chan J, Mehta S, Bharrhan S, Chen Y, Achkar JM, Casadevall A, Flynn J. The role of B cells and humoral immunity in Mycobacterium tuberculosis infection. *Semin Immunol* 2014: 26(6): 588-600.

39. Day CL, Abrahams DA, Lerumo L, Janse van Rensburg E, Stone L, O’Rie T, Pienaar B, de Kock M, Kaplan G, Mahomed H, Dheda K, Hanekom WA. Functional capacity of Mycobacterium tuberculosis-specific T cell responses in humans is associated with mycobacterial load. *J Immunol* 2011: 187(5): 2222-2232.

40. Day CL, Moshi ND, Abrahams DA, van Rooyen M, O’Rie T, de Kock M, Hanekom WA. Patients with tuberculosis disease have Mycobacterium tuberculosis-specific CD8 T cells with a pro-apoptotic phenotype and impaired proliferative capacity, which is not restored following treatment. *PLoS One* 2014: 9(4): e94949.

41. Chiacchio T, Petruccioli E, Vanini V, Cuzzi G, Pinnetti C, Sampaolesi A, Antinori A, Girardi E, Goletti D. Polyfunctional T-cells and effector memory phenotype are associated with active TB in HIV-infected patients. *J Infect* 2014: 69(6): 533-545.

42. Rozot V, Vigano S, Mazza-Stalder J, Idrizi E, Day CL, Perreau M, Lazor-Blanchet C, Petruccioli E, Hanekom W, Goletti D, Bart PA, Nicod L, Pantaleo G, Harari A. Mycobacterium tuberculosis-specific CD8+ T cells are functionally and phenotypically different between latent infection and active disease. *Eur J Immunol* 2013: 43(6): 1568-1577.

43. Nikolova M, Markova R, Drenskaya R, Muhtarova M, Todorova Y, Dimitrov V, Taskov H, Saltini C, Amicosante M. Antigen-specific CD4+ and CD8-positive signatures in different phases of Mycobacterium tuberculosis infection. *Diagn Microbiol Infect Dis* 2013: 75(3): 277-281.

44. Lancioni C, Nyendak M, Kiguli S, Zalwango S, Mori T, Mayanja-Kizza H, Balyeysa S, Null M, Baseke J, Mulindwa D, Byrd L, Swarbrick G, Scott C, Johnson DF, Malone L, Mudido-Musoke P, Boom WH, Lewinsohn DM, Lewinsohn DA, Tuberculosis Research U. CD8+ T cells provide an immunologic signature of tuberculosis in young children. *Am J Respir Crit Care Med* 2012: 185(2): 206-212.

45. Chiacchio RG, Prioste FE, Vanstreels RE, Knobl T, Kolber M, Miyashiro SI, Matushima ER. Health evaluation and survey of zoonotic pathogens in free-ranging capybaras (Hydrochoerus hydrochaeris). *J Wildl Dis* 2014: 50(3): 496-504.
46. Joosten SA, van Meijgaarden KE, Del Nonno F, Baiocchini A, Petrone L, Vanini V, Smits HH, Palmieri F, Goletti D, Ottenhoff TH. Patients with Tuberculosis Have a Dysfunctional Circulating B-Cell Compartment, Which Normalizes following Successful Treatment. *PLoS Pathog* 2016: 12(6): e1005687.

47. Li H, Javid B. Antibodies and tuberculosis: finally coming of age? *Nat Rev Immunol* 2018: 18(9): 591-596.

48. Jones A, Pitts M, Al Dulayymi JR, Gibbons J, Ramsay A, Goletti D, Gwenin CD, Baird MS. New synthetic lipid antigens for rapid serological diagnosis of tuberculosis. *PLoS One* 2017: 12(8): e0181414.

49. Biering-Sorensen S, Jensen KJ, Monterio I, Ravn H, Aaby P, Benn CS. Rapid Protective Effects of Early BCG on Neonatal Mortality Among Low Birth Weight Boys: Observations From Randomized Trials. *J Infect Dis* 2018: 217(5): 759-766.

50. Nemes E, Geldenhuys H, Rozot V, Rutkowski KT, Ratangee F, Bilek N, Mabwe S, Makhethe L, Erasmus M, Toefy A, Mulenga H, Hanekom WA, Self SG, Bekker LG, Ryall R, Gurunathan S, DiazGranados CA, Andersen P, Kromann I, Evans T, Ellis RD, Landry B, Hokey DA, Hopkins R, Ginsberg AM, Scriba TJ, Hatherill M, Team CS. Prevention of *M. tuberculosis* Infection with H4:IC31 Vaccine or BCG Revaccination. *N Engl J Med* 2018: 379(2): 138-149.

51. Netea MG, Joosten LA, Latz E, Mills KH, Natoli G, Stunnenberg HG, O'Neill LA, Xavier RJ. Trained immunity: A program of innate immune memory in health and disease. *Science* 2016: 352(6284): aaf1098.

52. Moorlag S, Arts RJW, van Crevel R, Netea MG. Non-specific effects of BCG vaccine on viral infections. *Clin Microbiol Infect* 2019: 25(12): 1473-1478.

53. Arts RJW, Moorlag S, Novakovic B, Li Y, Wang SY, Oosting M, Kumar V, Xavier RJ, Wijmenga C, Joosten LAB, Reusken C, Benn CS, Aaby P, Koopmans MP, Stunnenberg HG, van Crevel R, Netea MG. BCG Vaccination Protects against Experimental Viral Infection in Humans through the Induction of Cytokines Associated with Trained Immunity. *Cell Host Microbe* 2018: 23(1): 89-100 e105.

54. Mukherjee S, Subramaniam R, Chen H, Smith A, Keshava S, Shams H. Boosting efferocytosis in alveolar space using BCG vaccine to protect host against influenza pneumonia. *PLoS One* 2017: 12(7): e0180143.

55. Spencer JC, Ganguly R, Waldman RH. Nonspecific protection of mice against influenza virus infection by local or systemic immunization with Bacille Calmette-Guerin. *J Infect Dis* 1977: 136(2): 171-175.

56. WHO. Bacille Calmette-Guérin (BCG) vaccination and COVID-19. 2020.

57. Miller A, Reandelar MJ, Faschigione K, Roumenova V, Li Y, Otazu GH. Correlation between universal BCG vaccination policy and reduced morbidity and mortality for COVID-19: an epidemiological study. medRxiv, 2020.

58. Hensel J, McGrail DJ, McAndrews KM, Dowlatshahi D, LeBleu VS, Kalluri R. Exercising caution in correlating COVID-19 incidence and mortality rates with BCG vaccination policies due to variable rates of SARS CoV-2 testing. medRxiv, 2020.

59. Kirov S. Association Between BCG Policy is Significantly Confounded by Age and is Unlikely to Alter Infection or Mortality Rates. medRxiv, 2020.

60. Daoud A, Laktineh A, Macrander C, Mushtaq A, Soubani AO. Pulmonary complications of influenza infection: a targeted narrative review. *Postgrad Med* 2019: 131(5): 299-308.

61. CDC. Estimated Influenza Illnesses, Medical visits, Hospitalizations, and Deaths in the United States — 2018–2019 influenza season. 2020 [cited 6th April 2020]; Available from: https://www.cdc.gov/flu/about/burden/2018-2019.html
62. Rice TW, Rubinson L, Uyeki TM, Vaughn FL, John BB, Miller RR, 3rd, Higgs E, Randolph AG, Smoot BE, Thompson BT, Network NA. Critical illness from 2009 pandemic influenza A virus and bacterial coinfection in the United States. Crit Care Med 2012: 40(5): 1487-1498.

63. Esposito S, Molteni CG, Daleno C, Tagliabue C, Picciolli I, Scala A, Pelucchi C, Fossali E, Principi N. Impact of pandemic A/H1N1/2009 influenza on children and their families: comparison with seasonal A/H1N1 and A/H3N2 influenza viruses. J Infect 2011: 63(4): 300-307.

64. Marchisio P, Baggi E, Bianchini S, Principi N, Esposito S. Clinical and socioeconomic impact of pediatric seasonal and pandemic influenza. Hum Vaccin Immunother 2012: 8(1): 17-20.

65. Metersky ML, Masterton RG, Lode H, File TM, Jr., Babinchak T. Epidemiology, microbiology, and treatment considerations for bacterial pneumonia complicating influenza. Int J Infect Dis 2012: 16(5): e321-331.

66. Song JY, Cheong HJ, Heo JY, Noh JY, Yong HS, Kim YK, Kang EY, Choi WS, Jo YM, Kim WJ. Clinical, laboratory and radiologic characteristics of 2009 pandemic influenza A/H1N1 pneumonia: primary influenza pneumonia versus concomitant/secondary bacterial pneumonia. Influenza Other Respir Viruses 2011: 5(6): e535-543.

67. Morens DM, Taubenberger JK, Fauci AS. Predominant role of bacterial pneumonia as a cause of death in pandemic influenza: implications for pandemic influenza preparedness. J Infect Dis 2008: 198(7): 962-970.

68. Morens DM, Fauci AS. The 1918 influenza pandemic: insights for the 21st century. J Infect Dis 2007: 195(7): 1018-1028.

69. Channappanavar R, Perlman S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. Semin Immunopathol 2017: 39(5): 529-539.

70. Peiris JS, Lai ST, Poon LL, Guan Y, Yam LY, Lim W, Nicholls J, Yee WK, Yan WW, Cheung MT, Cheng VC, Chan KH, Tsang DN, Yung RW, Ng TK, Yuen KY, group Ss. Coronavirus as a possible cause of severe acute respiratory syndrome. Lancet 2003: 361(9366): 1319-1325.

71. Assiri A, Al-Tawfiq JA, Al-Rabeeah AA, Al-Rabieh FA, Al-Hajjar S, Al-Barrak A, Flenb坦 H, Al-Nassir WN, Balkhy HH, Al-Hakeem RF, Makhdoom HQ, Zumla AI, Memish ZA. Epidemiological, demographic, and clinical characteristics of 47 cases of Middle East respiratory syndrome coronavirus disease from Saudi Arabia: a descriptive study. Lancet Infect Dis 2013: 13(9): 752-761.

72. Saad M, Omrani AS, Baig K, Bahloul A, Elzein F, Matin MA, Selim MA, Al Mutairi M, Al Nakhli D, Al Aidaroos AY, Al Sherbeeni N, Al-Khashan HI, Memish ZA, Albarrak AM. Clinical aspects and outcomes of 70 patients with Middle East respiratory syndrome coronavirus infection: a single-center experience in Saudi Arabia. Int J Infect Dis 2014: 29: 301-306.

73. Al-Tawfiq JA, Hinedi K, Ghandour J, Khairalla H, Musleh S, Ujayli A, Memish ZA. Middle East respiratory syndrome coronavirus: a case-control study of hospitalized patients. Clin Infect Dis 2014: 59(2): 160-165.

74. Zumla A, Hui DS, Perlman S. Middle East respiratory syndrome. Lancet 2015: 386(9997): 995-1007.

75. Li W, Moore MJ, Vasilieva N, Sui J, Wong SK, Berne MA, Somasundaran M, Sullivan JL, Luzuriaga K, Greenough TC, Choe H, Farzan M. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. Nature 2003: 426(6965): 450-454.
76. WHO. Emergency preparedness, response: Preliminary Clinical Description of Severe Acute Respiratory Syndrome. [cited 9th May 2020]; Available from: https://www.who.int/csr/sars/clinical/en/

77. Peiris JS, Chu CM, Cheng VC, Chan KS, Hung IF, Poon LL, Law KI, Tang BS, Hon TY, Chan CS, Chan KH, Ng JS, Zheng BJ, Ng WL, Lai RW, Guan Y, Yuen KY, Group HUSS. Clinical progression and viral load in a community outbreak of coronavirus-associated SARS pneumonia: a prospective study. *Lancet* 2003: 361(9371): 1767-1772.

78. Nicholls J, Dong XP, Jiang G, Peiris M. SARS: clinical virology and pathogenesis. *Respirology* 2003: 8 Suppl: S6-8.

79. van den Brand JM, Haagmans BL, van Riel D, Osterhaus AD, Kuiken T. The pathology and pathogenesis of experimental severe acute respiratory syndrome and influenza in animal models. *J Comp Pathol* 2014: 151(1): 83-112.

80. Raj VS, Mou H, Smits SL, Dekkers DH, Muller MA, Dijkman R, Muth D, Demmers JA, Zaki A, Fouchier RA, Bellemann E, Rottier PJ, Osterhaus AD, Bosch BJ, Haagmans BL. Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC. *Nature* 2013: 495(7440): 251-254.

81. Arabi YM, Arifi AA, Balkhy HH, Najm H, Aldawood AS, Ghabashi A, Hava H, Alothman A, Khalidi A, Al Raiy B. Clinical course and outcomes of critically ill patients with Middle East respiratory syndrome coronavirus infection. *Ann Intern Med* 2014: 160(6): 389-397.

82. Guan WJ, Ni ZY, Hu Y, Liang WH, Ou CQ, He JX, Liu L, Shan H, Lei CL, Hui DSC, Du B, Li LJ, Zeng G, Yuen KY, Chen RC, Tang CL, Wang T, Chen PY, Xiang J, Li SY, Wang JL, Liang ZJ, Peng XY, Wei L, Liu Y, Hu YH, Peng P, Wang JM, Liu JY, Chen Z, Li G, Zheng ZJ, Qiu SQ, Luo J, Ye CJ, Zhu SY, Zhong NS, China Medical Treatment Expert Group for C. Clinical Characteristics of Coronavirus Disease 2019 in China. *N Engl J Med* 2020.

83. Xu XW, Wu XX, Jiang XG, Xu KJ, Ying LJ, Ma CL, Li SB, Wang HY, Zhang S, Gao HN, Sheng JF, Cai HL, Qiu YQ, Li LJ. Clinical findings in a group of patients infected with the 2019 novel coronavirus (SARS-CoV-2) outside of Wuhan, China: retrospective case series. *BMJ* 2020: 368: m606.

84. Zhou F, Yu T, Du R, Fan G, Li Y, Liu Z, Xiang J, Wang Y, Song B, Gu X, Guan L, Wei Y, Li H, Wu X, Xu J, Tu S, Zhang Y, Chen H, Cao B. Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. *Lancet* 2020: 395(10229): 1054-1062.

85. Wu Z, McGoogan JM. Characteristics of and Important Lessons From the Coronavirus Disease 2019 (COVID-19) Outbreak in China: Summary of a Report of 72314 Cases From the Chinese Center for Disease Control and Prevention. *JAMA* 2020.

86. Alhazzani W, Moller MH, Arabi YM, Loeb M, Gong MN, Fan E, Oczkowski S, Levy MM, Derde L, Dzierba A, Du B, Aboodi M, Wunsch H, Cecconi M, Koh Y, Chertow DS, Maitland K, Alshamsi F, Belley-Cote E, Greco M, Laudy M, Morgan JS, Kesecioglu J, McGeer A, Mermel L, Mammen MJ, Alexander PE, Arrington A, Centofanti JE, Citerio G, Baw B, Memish ZA, Hammond N, Hayden FG, Evans L, Rhodes A. Surviving Sepsis Campaign: Guidelines on the Management of Critically Ill Adults with Coronavirus Disease 2019 (COVID-19). *Crit Care Med* 2020.

87. Vitacca M NS, Santus P, Harari S. Early consensus management for non-ICU ARF SARS-CoV-2 emergency in Italy: from ward to trenches. *Eur Respir J* 2020.

88. Cai J, Xu J, Lin D, Yang Z, Xu L, Qu Z, Zhang Y, Zhang H, Jia R, Liu P, Wang X, Ge Y, Xia A, Tian H, Chang H, Wang C, Li J, Wang J, Zeng M. A Case Series of children with 2019 novel coronavirus infection: clinical and epidemiological features. *Clin Infect Dis* 2020.
89. Lu X, Zhang L, Du H, Zhang J, Li YY, Qu J, Zhang W, Wang Y, Bao S, Li Y, Wu C, Liu H, Liu D, Shao J, Peng X, Yang Y, Liu Z, Xiang Y, Zhang F, Silva RM, Pinkerton KE, Shen K, Xiao H, Xu S, Wong GWK, Chinese Pediatric Novel Coronavirus Study T. SARS-CoV-2 Infection in Children. *N Engl J Med* 2020.

90. Cui Y, Tian M, Huang D, Wang X, Huang Y, Fan L, Wang L, Chen Y, Liu W, Zhang K, Wu Y, Yang Z, Tao J, Feng J, Liu K, Ye X, Wang R, Zhang X, Zha Y. A 55-Day-Old Female Infant infected with COVID-19: presenting with pneumonia, liver injury, and heart damage. *J Infect Dis* 2020.

91. Liu W, Zhang Q, Chen J, Xiang R, Song H, Shu S, Chen L, Liang L, Zhou J, You L, Wu P, Zhang B, Lu Y, Xia L, Huang L, Yang Y, Liu F, Semple MG, Cowling BJ, Lan K, Sun Z, Yu H, Liu Y. Detection of Covid-19 in Children in Early January 2020 in Wuhan, China. *N Engl J Med* 2020: 382(14): 1370-1371.

92. Shen K, Yang Y, Wang T, Zhao D, Jiang Y, Jin R, Zheng Y, Xu B, Xie Z, Lin L, Shang Y, Lu X, Shu S, Bai Y, Deng J, Lu M, Ye L, Wang X, Wang Y, Gao L. China National Clinical Research Center for Respiratory D, National Center for Children's Health BC, Group of Respirology CPSCMA, Chinese Medical Doctor Association Committee on Respirology P, China Medicine Education Association Committee on P, Chinese Research Hospital Association Committee on P, Chinese Non-government Medical Institutions Association Committee on P, China Association of Traditional Chinese Medicine CoCsH, Medicine R, China News of Drug Information Association CoCsSM, Global Pediatric Pulmonology A. Diagnosis, treatment, and prevention of 2019 novel coronavirus infection in children: experts' consensus statement. *World J Pediatr* 2020.

93. ECDC. Guidelines for the use of non-pharmaceutical measures to delay and mitigate the impact of 2019-nCoV, Stockholm 2020., 2020.

94. Wang Y, Dong C, Hu Y, Li C, Ren Q, Zhang X, Shi H, Zhou M. Temporal Changes of CT Findings in 90 Patients with COVID-19 Pneumonia: A Longitudinal Study. *Radiology* 2020: 200843.

95. Wang W, Xu Y, Gao R, Lu R, Han K, Wu G, Tan W. Detection of SARS-CoV-2 in Different Types of Clinical Specimens. *JAMA* 2020.

96. Liu Q, Wang RS, Qu GQ, Wang YY, Liu P, Zhu YZ, Fei G, Ren L, Zhou YW, Liu L. Gross examination report of a COVID-19 death autopsy. *Fa Yi Xue Za Zhi* 2020: 36(1): 21-23.

97. Tian S, Hu W, Niu L, Liu H, Xu H, Xiao SY. Pulmonary Pathology of Early-Phase 2019 Novel Coronavirus (COVID-19) Pneumonia in Two Patients With Lung Cancer. *J Thorac Oncol* 2020.

98. Fox SE, Akmatbekov A, Harbert JL, Li G, Brown JQ, Vander Heide RS. Pulmonary and Cardiac Pathology in Covid-19: The First Autopsy Series from New Orleans. medRxiv, 2020.

99. ISARIC. ISARIC: COVID-19 Report: 27 April 2020. 2020.

100. Rello J, Storti E, Belliato M, Serrano R. Clinical phenotypes of SARS-CoV-2: Implications for clinicians and researchers. *Eur Respir J* 2020.

101. Tokman S, Huang L. Evaluation of respiratory disease. *Clin Chest Med* 2013: 34(2): 191-204.

102. Azoulay E, de Castro N, Barbier F. Critically Ill Patients With HIV: 40 Years Later. *Chest* 2020: 157(2): 293-309.

103. Segal LN, Methe BA, Nolan A, Hoshino Y, Rom WN, Dawson R, Bateman E, Weiden MD. HIV-1 and bacterial pneumonia in the era of antiretroviral therapy. *Proc Am Thorac Soc* 2011: 8(3): 282-287.
104. Marais BJ, Chakaya J, Swaminathan S, Fox GJ, Ehtesham NZ, Ntoumi F, Zijenah L, Maurer M, Zumla A. Tackling long-term morbidity and mortality after successful tuberculosis treatment. *Lancet Infect Dis* 2020.

105. Pasipanodya JG, Miller TL, Vecino M, Munguia G, Garmon R, Bae S, Drewyer G, Weis SE. Pulmonary impairment after tuberculosis. *Chest* 2007: 131(6): 1817-1824.

106. Walaza S, Tempia S, Dawood H, Variava E, Wolter N, Dreyer A, Moyes J, Von Mollendorf C, McMorrow M, Von Gottberg A, Haffejee S, Venter M, Treurnicht FK, Hellferssee O, Martinson NA, Ismail N, Cohen C. The Impact of Influenza and Tuberculosis Interaction on Mortality Among Individuals Aged >/=15 Years Hospitalized With Severe Respiratory Illness in South Africa, 2010-2016. *Open Forum Infect Dis* 2019: 6(3): ofz020.

107. Walaza S, Cohen C, Nanoo A, Cohen AL, McNerney J, von Mollendorf C, Moyes J, Tempia S. Excess Mortality Associated with Influenza among Tuberculosis Deaths in South Africa, 1999-2009. *PLoS One* 2015: 10(6): e0129173.

108. Mhimbira F, Hiza H, Mbuba E, Hella J, Kamwela L, Sasamalo M, Ticlla M, Said K, Mhlu G, Chiryamkubi M, Schindler C, Reither K, Gagneux S, Fenner L. Prevalence and clinical significance of respiratory viruses and bacteria detected in tuberculosis patients compared to household contact controls in Tanzania: a cohort study. *Clin Microbiol Infect* 2019: 25(1): 107 e101-107 e107.

109. anon. Tuberculosis after Influenza. *Cal State J Med* 1919: 17(3): 85.

110. Ballinger MN, Standiford TJ. Postinfluenza bacterial pneumonia: host defenses gone awry. *J Interferon Cytokine Res* 2010: 30(9): 643-652.

111. Park Y, Chin BS, Han SH, Yun Y, Kim YJ, Choi JY, Kim CO, Song YG, Kim JM. Pandemic Influenza (H1N1) and Mycobacterium tuberculosis Co-infection. *Tuberc Respir Dis (Seoul)* 2014: 76(2): 84-87.

112. Zurcher K, Zwahlen M, Ballif M, Rieder HL, Egger M, Fenner L. Influenza Pandemics and Tuberculosis Mortality in 1889 and 1918: Analysis of Historical Data from Switzerland. *PLoS One* 2016: 11(10): e0162575.

113. Luo T, Sumi A, Zhou D, Kobayashi N, Mise K, Yu B, Kong D, Wang J, Duan Q. Seasonality of reported tuberculosis cases from 2006 to 2010 in Wuhan, China. *Epidemiol Infect* 2014: 142(10): 2036-2048.

114. Roth S, Whitehead S, Thamthitiwat S, Chittaganpitch M, Maloney SA, Baggett HC, Olsen SJ. Concurrent influenza virus infection and tuberculosis in patients hospitalized with respiratory illness in Thailand. *Influenza Other Respir Viruses* 2013: 7(3): 244-248.

115. de Paus RA, van Crevel R, van Beek R, Sahiratmadja E, Alisjahbana B, Marzuki S, Rimmelzwaan GF, van Dissel JT, Ottenhoff TH, van de Vosse E. The influence of influenza virus infections on the development of tuberculosis. *Tuberculosis (Edinb)* 2013: 93(3): 338-342.

116. Yen YF, Pan SW, Su VY, Chuang PH, Feng JY, Su WJ. Influenza Vaccination and Incident Tuberculosis among Elderly Persons, Taiwan(1). *Emerg Infect Dis* 2018: 24(3): 498-505.

117. McShane H. Co-infection with HIV and TB: double trouble. *Int J STD AIDS* 2005: 16(2): 95-100; quiz 101.

118. Lawn SD, Harries AD, Williams BG, Chaisson RE, Losina E, De Cock KM, Wood R. Antiretroviral therapy and the control of HIV-associated tuberculosis. Will ART do it? *Int J Tuberc Lung Dis* 2011: 15(5): 571-581.

119. WHO. WHO consolidated guidelines on tuberculosis: module 1: prevention: tuberculosis preventive treatment. 2020.

120. WHO. WHO operational handbook on tuberculosis: module 1: prevention: tuberculosis preventive treatment. 2020.
121. WHO. WHO policy on collaborative TB/HIV activities: guidelines for national programmes and other stakeholders 2012.

122. Liu W, Fontanet A, Zhang PH, Zhan L, Xin ZT, Tang F, Baril L, Cao WC. Pulmonary tuberculosis and SARS, China. Emerg Infect Dis 2006: 12(4): 707-709.

123. Low JG, Lee CC, Leo YS, Low JG, Lee CC, Leo YS. Severe acute respiratory syndrome and pulmonary tuberculosis. Clin Infect Dis 2004: 38(12): e123-125.

124. Cui W, Fan Y, Wu W, Zhang F, Wang JY, Ni AP. Expression of lymphocytes and lymphocyte subsets in patients with severe acute respiratory syndrome. Clin Infect Dis 2003: 37(6): 857-859.

125. Goletti D, Lee MR, Wang JY, Walter N, Ottenhoff THM. Update on tuberculosis biomarkers: From correlates of risk, to correlates of active disease and of cure from disease. Respiratio 2018: 23(5): 455-466.

126. Lindestam Arlehamn CS, McKinney DM, Carpenter C, Paul S, Rozot V, Makgotlho E, Gregg Y, van Rooyen M, Ernst JD, Hatherill M, Hanekom WA, Peters B, Scriba TJ, Sette A. A Quantitative Analysis of Complexity of Human Pathogen-Specific CD4 T Cell Responses in Healthy M. tuberculosis Infected South Africans. PLoS Pathog 2016: 12(7): e1005760.

127. Petruccioli E, Petrone L, Vanini V, Sampaolesi A, Gualano G, Girardi E, Palmieri F, Goletti D. IFNgamma/TNFalpha specific-cells and effector memory phenotype associate with active tuberculosis. J Infect 2013: 66(6): 475-486.

128. Chiachio T, Petruccioli E, Vanini V, Cuzzi G, La Manna MP, Orlando V, Pinnetti C, Sampaolesi A, Antinori A, Caccamo N, Goletti D. Impact of antiretroviral and tuberculosis therapies on CD4(+) and CD8(+) HIV/M. tuberculosis-specific T-cell in co-infected subjects. Immunol Lett 2018: 198: 33-43.

129. Petruccioli E, Chiachio T, Navarra A, Vanini V, Cuzzi G, Cimaglia C, Codecas LR, Pinnetti C, Riccardi N, Palmieri F, Antinori A, Goletti D. Effect of HIV-infection on QuantiFERON-plus accuracy in patients with active tuberculosis and latent infection. J Infect 2020.

130. Wong CY, Wong KY, Law TS, Shum TT, Li YK, Pang WK. Tuberculosis in a SARS outbreak. J Chin Med Assoc 2004: 67(11): 579-582.

131. Ong CWM, Goletti D. Impact of the global COVID-19 outbreak on the management of other communicable diseases. Int J Tuberc Lung Dis 2020.

132. Alfaraj SH, Al-Tawfiq JA, Altuwaijri TA, Memish ZA. Middle East Respiratory Syndrome Coronavirus and Pulmonary Tuberculosis Coinfection: Implications for Infection Control. Intervirology 2017: 60(1-2): 53-55.

133. Liu Y, Bi L, Chen Y, Wang Y, Fleming J, Yu Y, Gu Y, Liu C, Fan L, Wang X, Cheng M. Active or latent tuberculosis increases susceptibility to COVID-19 and disease severity. medRxiv, 2020.

134. Maciel EL, Goncalves EJ, Dalcolmo MMP. Tuberculosis and coronavirus: what do we know? Epidemiol Serv Saude 2020.

135. Tadolini M, Codecas LR, García-García JM, Blanc FX, Borisov S, Alffenaar J, Andréjak C, Bachez PB, P. A., Belilovsky E, Cardoso-Landivar J, Centis R, D’Ambrosio L, De Souza-Galvão ML, Dominguez-Castellano A, Dourmane S, Jackyn MF, Frosiart A, Giacomet V, Goletti D, Grard S, Gualano G, Izadifar A, Du DL, Royo MM, Mazza-Stalder J, Motta I, Ong CWM, Palmieri F, Rivière F, Rodrigo T, Silva DR, Sánchez-Montalvá A, Saporiti M, Scarpellini P, Schlemmer F, Spanevello A, Sumarokova E, Tabernerio E, Tambyah PA, Tiberi S, Torre A, Visca D, Murguiondo MZ, Sotgiu G, Migliori GB. Active tuberculosis, sequelae and COVID-19 co-infection: first cohort of 49 cases. Eur Respir J 2020: In Press.
136. Motta I, Entis I, D’Ambrosio L, Garcia-Garcia J, Goletti D, Gualano G, Lipani F, Palmieri F, Sanchez-Montalva A, Pontali E, Sotgiu G, Spanevillo A, Stochino C, Tabernero E, Tadolini M, van den Boom M, Villa S, Visca D, Migliori GB. Tuberculosis, COVID-19 and migrants: preliminary analysis of deaths occurring in 69 patients from two cohorts. Pulmonology 2020: In Press.

137. Ault R, Dwivedi V, Koivisto E, Nagy J, Miller K, Nagendran K, Chalana I, Pan X, Wang SH, Turner J. Altered monocyte phenotypes but not impaired peripheral T cell immunity may explain susceptibility of the elderly to develop tuberculosis. Exp Gerontol 2018: 111: 35-44.

138. Novel Coronavirus Pneumonia Emergency Response Epidemiology T. [The epidemiological characteristics of an outbreak of 2019 novel coronavirus diseases (COVID-19) in China]. Zhonghua Liu Xing Bing Xue Za Zhi 2020: 41(2): 145-151.

139. Wang JT, Sheng WH, Fang CT, Chen YC, Wang JL, Yu CJ, Chang SC, Yang PC. Clinical manifestations, laboratory findings, and treatment outcomes of SARS patients. Emerg Infect Dis 2004: 10(5): 818-824.

140. Reed C, Chaves SS, Perez A, D’Mello T, Daily Kirley P, Aragon D, Meek JJ, Farley MM, Ryan P, Lynfield R, Morin CA, Hancock EB, Bennett NM, Zansky SM, Thomas A, Lindegren ML, Schaffner W, Finelli L. Complications among adults hospitalized with influenza: a comparison of seasonal influenza and the 2009 H1N1 pandemic. Clin Infect Dis 2014: 59(2): 166-174.

141. WHO. Systematic screening for active tuberculosis: Principles and recommendations. 2013.

142. Jamieson DJ, Honein MA, Rasmussen SA, Williams JL, Swerdlow DL, Biggerstaff MS, Lindstrom J, Krist JD, Christ CM, Bohm SR, Fonseca VP, Ritger KA, Kuhles DJ, Eggers P, Bruce H, Davidson HA, Lutterloh E, Harris ML, Burke C, Cocoros N, Finelli L, MacFarlane KF, Shu B, Olsen SJ, Novel Influenza APWG. H1N1 2009 influenza virus infection during pregnancy in the USA. Lancet 2009: 374(9688): 451-458.

143. Cordero E, Aydillo T, Farinas MC, Pano-Pardo JR, Pachon J, Riera M, Lopez-Medrano F, Payares A, Moreno A, Rodriguez-Bano J, Oteo JA, Martinez-Montauti J, Torre-Cisneros J, Segura F, Carratala J, Novel Influenza ASGotSfRiID. Immunosuppressed patients with pandemic influenza A 2009 (H1N1) virus infection. Eur J Clin Microbiol Infect Dis 2012: 31(4): 547-556.

144. Mathad JS, Gupta A. Tuberculosis in pregnant and postpartum women: epidemiology, management, and research gaps. Clin Infect Dis 2012: 55(11): 1532-1549.

145. Akiyama MJ, Spaulding AC, Rich JD. Flattening the Curve for Incarcerated Populations - Covid-19 in Jails and Prisons. N Engl J Med 2020.

146. Kinner SA, Young JT, Snow K, Southalan L, Lopez-Acuna D, Ferreira-Borges C, O’Moore E. Prisons and custodial settings are part of a comprehensive response to COVID-19. Lancet Public Health 2020: 5(4): e188-e189.

147. Negin J, Abimbola S, Marais BJ. Tuberculosis among older adults--time to take notice. Int J Infect Dis 2015: 32: 135-137.

148. Pinsky BA, Hayden RT. Cost-Effective Respiratory Virus Testing. J Clin Microbiol 2019: 57(9).

149. Almekhlafi GA, Albarrak MM, Mandourah Y, Hassan S, Alwan A, Abudayah A, Altayyar S, Mustafa M, Aldaghistani T, Alghamedi A, Talag A, Malik MK, Omrani AS, Sakr Y. Presentation and outcome of Middle East respiratory syndrome in Saudi intensive care unit patients. Crit Care 2016: 20(1): 123.

150. Booth CM, Matukas LM, Tomlinson GA, Rachlis AR, Rose DB, Dwosh HA, Walmsley SL, Mazzulli T, Avendano M, Derkach P, Ephtimios IE, Kitai I, Mederski BD, Shadowitz SB, Gold WL, Hawryluck LA, Rea E, Chenkin JS, Cescon DW, Poutanen SM,
Detsky AS. Clinical features and short-term outcomes of 144 patients with SARS in the greater Toronto area. *JAMA* 2003: 289(21): 2801-2809.

151. Midgley CM, Jackson MA, Selvarangan R, Turabelidze G, Obringer E, Johnson D, Giles BL, Patel A, Echols F, Oberste MS, Nix WA, Watson JT, Gerber SI. Severe respiratory illness associated with enterovirus D68 - Missouri and Illinois, 2014. *MMWR Morb Mortal Wkly Rep* 2014: 63(36): 798-799.

152. Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, Zhao X, Huang B, Shi W, Lu R, Niu P, Zhan F, Ma X, Wang D, Xu W, Wu G, Gao GF, Tan W, China Novel Coronavirus I, Research T. A Novel Coronavirus from Patients with Pneumonia in China, 2019. *N Engl J Med* 2020: 382(8): 727-733.

153. Wumkes ML, van der Velden AM, de Bruin E, Meerveld-Eggink A, Koopmans MP, Rimmelzwaan GF, Rijkers GT, Biesma DH. Microarray profile of the humoral immune response to influenza vaccination in breast cancer patients treated with chemotherapy. *Vaccine* 2017: 35(9): 1299-1305.

154. Goodwin S, McPherson JD, McCombie WR. Coming of age: ten years of next-generation sequencing technologies. *Nat Rev Genet* 2016: 17(6): 333-351.

155. Li CX, Li W, Zhou J, Zhang B, Feng Y, Xu CP, Lu YY, Holmes EC, Shi M. High resolution metagenomic characterization of complex infectomes in paediatric acute respiratory infection. *Sci Rep* 2020: 10(1): 3963.

156. Kufner V, Plate A, Schmutz S, Braun DL, Gunthard HF, Capaul R, Zbinden A, Mueller NJ, Trkola A, Huber M. Two Years of Viral Metagenomics in a Tertiary Diagnostics Unit: Evaluation of the First 105 Cases. *Genes (Basel)* 2019: 10(9).

157. Gillim-Ross L, Subbarao K. Emerging respiratory viruses: challenges and vaccine strategies. *Clin Microbiol Rev* 2006: 19(4): 614-636.

158. Machado D, Couto I, Viveiros M. Advances in the molecular diagnosis of tuberculosis: From probes to genomes. *Infect Genet Evol* 2019: 72: 93-112.

159. Evans D, Sineke T, Schnippel K, Berhanu R, Govathson C, Black A, Long L, Rosen S. Impact of Xpert MTB/RIF and decentralized care on linkage to care and drug-resistant tuberculosis treatment outcomes in Johannesburg, South Africa. *BMC Health Serv Res* 2018: 18(1): 973.

160. Goletti D, Sanduzzi A, Delogu G. Performance of the tuberculin skin test and interferon-gamma release assays: an update on the accuracy, cutoff stratification, and new potential immune-based approaches. *J Rheumatol Suppl* 2014: 91: 24-31.

161. WHO. Clinical management of severe acute respiratory infection when COVID-19 is suspected: Interim guidance. 2020 [cited 8th April 2020]; Available from: https://www.who.int/publications-detail/clinical-management-of-severe-acute-respiratory-infection-when-novel-coronavirus-(ncov)-infection-is-suspected

162. Alhazzani W, Moller MH, Arabi YM, Loeb M, Gong MN, Fan E, Oczkowski S, Levy MM, Derde L, Dzierba A, Du B, Aboodi M, Wunsch H, Cecconci M, Koh Y, Chertow DS, Maitland K, Alshamsi F, Belley-Cote E, Greco M, Laundy M, Morgan JS, Kesecioglu J, McGeer A, Mermel L, Mammen MJ, Alexander PE, Arrington A, Centofanti JE, Citerio G, Baw B, Memish ZA, Hammond N, Hayden FG, Evans L, Rhodes A. Surviving Sepsis Campaign: guidelines on the management of critically ill adults with Coronavirus Disease 2019 (COVID-19). *Intensive Care Med* 2020.

163. McCrery EK, Pogue, J.M.; on behalf of the Society of Infectious Diseases Pharmacists. COVID-19 Treatment: A Review of Early and Emerging Options *Open Forum Infectious Diseases* 2020.

164. Hagan G, Nathani N. Clinical review: tuberculosis on the intensive care unit. *Crit Care* 2013: 17(5): 240.
Dorward J, Gbinigie K. Lopinavir/ritonavir: A rapid review of effectiveness in COVID-19. 2020 14 April 2020 [cited 16 April 2020]; Available from: https://www.cebm.net/covid-19/lopinavir-ritonavir-a-rapid-review-of-the-evidence-for-effectiveness-in-treating-covid/

Cao B, Wang Y, Wen D, Liu W, Wang J, Fan G, Ruan L, Song B, Cai Y, Wei M, Li X, Xia J, Chen N, Xiang J, Yu T, Bai T, Xie X, Zhang L, Li C, Yuan Y, Chen H, Li H, Huang H, Tu S, Gong F, Liu Y, Wei Y, Dong C, Zhou F, Gu X, Xu J, Liu Z, Zhang Y, Li H, Shang L, Wang K, Li K, Zhou X, Dong X, Qu Z, Lu S, Hu X, Ruan S, Luo S, Wu J, Peng L, Cheng F, Pan L, Zou J, Jia C, Wang J, Liu X, Wang S, Wu X, Ge Q, He J, Zhan H, Qiu F, Guo L, Huang C, Jaki T, Hayden FG, Horby PW, Zhang D, Wang C. A Trial of Lopinavir-Ritonavir in Adults Hospitalized with Severe Covid-19. N Engl J Med 2020.

Rabie H, Denti P, Lee J, Masango M, Coovadia A, Pillay S, Liberty A, Simon F, McIlleenon H, Cotton MF, Lallemant M. Lopinavir-ritonavir super-boosting in young HIV-infected children on rifampicin-based tuberculosis therapy compared with lopinavir-ritonavir without rifampicin: a pharmacokinetic modelling and clinical study. Lancet HIV 2018.

Lan NT, Thu NT, Barrail-Tran A, Duc NH, Lan NN, Laureillard D, Lien TT, Borand L, Quillet C, Connolly C, Lagarde D, Pym A, Lienhardt C, Dung NH, Taburet AM, Harries AD. Randomised pharmacokinetic trial of rifabutin with lopinavir/ritonavir-antiretroviral therapy in patients with HIV-associated tuberculosis in Vietnam. PLoS One 2014: 9(1): e84866.

Principi N, Esposito S. Chloroquine or hydroxychloroquine for prophylaxis of COVID-19. Lancet Infect Dis 2020.

Chen Z, Hu J, Zhang Z, Jiang S, Han S, Yan D, Zhuang R, Hu B, Zhang Z. Efficacy of hydroxychloroquine in patients with COVID-19: results of a randomized clinical trial. medRxiv, 2020.

Gautret P, Lagier JC, Parola P, Hoang VT, Meddeb L, Mailhe M, Doudier B, Courjon J, Giordanengo V, Vieira VE, Dupont HT, Honore S, Colson P, Chabriere E, La Scola B, Rolain JM, Brouqui P, Raoult D. Hydroxychloroquine and azithromycin as a treatment of COVID-19: results of an open-label non-randomized clinical trial. Int J Antimicrob Agents 2020: 105949.

Jang CH, Choi JH, Byun MS, Jae DM. Chloroquine inhibits production of TNF-alpha, IL-1beta and IL-6 from lipopolysaccharide-stimulated human monocytes/macrophages by different modes. Rheumatology (Oxford) 2006: 45(6): 703-710.

Ben-Zvi I, Kivity S, Langevitz P, Shoenfeld Y. Hydroxychloroquine: from malaria to autoimmunity. Clin Rev Allergy Immunol 2012: 42(2): 145-153.

Savarino A, Boelaert JR, Cassone A, Majori G, Cauda R. Effects of chloroquine on viral infections: an old drug against today's diseases? Lancet Infect Dis 2003: 3(11): 722-727.

Borba MGS, Val FF, Sampaio VS, Alexandre MAA, Melo GC, Brito M, Mourao MPG, Brito-Sousa JD, Baia-da-Silva D, Guerra MVF, Hajjar BA, Pinto RC, Balieiro AAS, Pacheco AGF, Santos JDO, Jr., Naveca FG, Xavier MS, Siqueira AM, Schwarzbolz A, Croda J, Nogueira ML, Romero GAS, Bassat Q, Fontes CJ, Albuquerque BC, Daniel-Ribeiro CT, Monteiro WM, Lacerda MVG, CloroCovid T. Effect of High vs Low Doses of Chloroquine Diphosphate as Adjunctive Therapy for Patients Hospitalized With Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Infection: A Randomized Clinical Trial. JAMA Netw Open 2020: 3(4): e208857.

FDA. FDA cautions against use of hydroxychloroquine or chloroquine for COVID-19 outside of the hospital setting or a clinical trial due to risk of heart rhythm problems. 2020 [cited 4th May 2020]; Available from: https://www.fda.gov/drugs/drug-safety-and-availability/fda-cautions-against-use-hydroxychloroquine-or-chloroquine-covid-19-outside-hospital-setting-or
177. Ducharme J, Farinotti R. Clinical pharmacokinetics and metabolism of chloroquine. Focus on recent advancements. *Clin Pharmacokinet* 1996: 31(4): 257-274.
178. Toniati P, Piva S, Cattalini M, Garrafa E, Regola F, Castelli F, Franceschini F, Foca E, Andreoli L, Latronico N, Brescia International R, Training HUB. Tocilizumab for the treatment of severe COVID-19 pneumonia with hyperinflammatory syndrome and acute respiratory failure: A single center study of 100 patients in Brescia, Italy. *Autoimmun Rev* 2020: 102568.
179. Cantini F, Niccoli L, Matarrese D, Nicastri E, Stobbione P, Goletti D. Baricitinib therapy in COVID-19: A pilot study on safety and clinical impact. *J Infect* 2020: In Press.
180. Weng D, Wu Q, Chen XQ, Du YK, Chen T, Li H, Tang DL, Li QH, Zhang Y, Lu LQ, Zhou NY, Song JC, Wang C, Li HP. Azithromycin treats diffuse panbronchiolitis by targeting T cells via inhibition of mTOR pathway. *Biomed Pharmacother* 2019: 110: 440-448.
181. Li H, Zhou Y, Zhang M, Wang H, Zhao Q, Liu J. Updated approaches against SARS-CoV-2. *Antimicrob Agents Chemother* 2020.
182. Li G, De Clercq E. Therapeutic options for the 2019 novel coronavirus (2019-nCoV). *Nat Rev Drug Discov* 2020: 19(3): 149-150.
183. Wang M, Cao R, Zhang L, Yang X, Liu J, Xu M, Shi Z, Hu Z, Zhong W, Xiao G. Remdesivir and chloroquine effectively inhibit the recently emerged novel coronavirus (2019-nCoV) in vitro. *Cell Res* 2020: 30(3): 269-271.
184. Wang Y, Zhang D, Du G, Du R, Zhao J, Jin Y, Fu S, Gao L, Cheng Z, Lu Q, Hu Y, Luo G, Wang K, Lu Y, Li H, Wang S, Ruan S, Yang C, Mei C, Y. W, Ding D, Wu F, Tang X, Ye X, Ye Y, Liu B, Yang J, Yin W, Wang A, Fan G, Zhuo F, Liu Z, Gu X, Xu J, Shang L, Zhang Y, Cao L, Guo T, Wan Y, Qin H, Jiang Y, Jaki T, Hayden FG, Horby PW, Cao B, Wang C. Remdesivir in adults with severe COVID-19: a randomised, double-blind, placebo-controlled, multicentre trial. *Lancet* 2020.
185. Khamitov RA, Loginova S, Shchukina VN, Borisevich SV, Maksimov VA, Shuster AM. [Antiviral activity of arbidol and its derivatives against the pathogen of severe acute respiratory syndrome in the cell cultures]. *Vopr Virusol* 2008: 53(4): 9-13.
186. Deng L, Li C, Zeng Q, Liu X, Li X, Zhang H, Hong Z, Xia J. Arbidol combined with LPV/r versus LPV/r alone against Corona Virus Disease 2019: A retrospective cohort study. *J Infect* 2020.
187. WHO. WHO Interim Protocol: Rapid operations to contain the initial emergence of pandemic influenza., 2012.
188. Hatchett RJ, Mecher CE, Lipsitch M. Public health interventions and epidemic intensity during the 1918 influenza pandemic. *Proc Natl Acad Sci U S A* 2007: 104(18): 7582-7587.
189. Migliori GB, Nardell E, Yedilbayev A, D’Ambrosio L, Centis R, Tadolini M, van den Boom M, Ehsani S, Sotgiu G, Dara M. Reducing tuberculosis transmission: a consensus document from the World Health Organization Regional Office for Europe. *Eur Respir J* 2019: 53(6).
190. WHO. WHO guidelines on tuberculosis infection prevention and control: 2019 update, Geneva. 2019.
191. Thorne CD, Khozin S, McDiarmid MA. Using the hierarchy of control technologies to improve healthcare facility infection control: lessons from severe acute respiratory syndrome. *J Occup Environ Med* 2004: 46(7): 613-622.
192. Bai Y, Yao L, Wei T, Tian F, Jin DY, Chen L, Wang M. Presumed Asymptomatic Carrier Transmission of COVID-19. *JAMA* 2020.
193. Zou L, Ruan F, Huang M, Liang L, Huang H, Hong Z, Yu J, Kang M, Song Y, Xia J, Guo Q, Song T, He J, Yen HL, Peiris M, Wu J. SARS-CoV-2 Viral Load in Upper Respiratory Specimens of Infected Patients. *N Engl J Med* 2020: 382(12): 1177-1179.

194. Leung CC, Lam TH, Cheng KK. Mass masking in the COVID-19 epidemic: people need guidance. *Lancet* 2020: 395(10228): 945.

195. Leung CC, Lam TH, Cheng KK. Let us not forget the mask in our attempts to stall the spread of COVID-19. *International Journal of Tuberculosis and Lung Diseases* 2020.

196. MacIntyre CR, Seale H, Dung TC, Hien NT, Nga PT, Chughtai AA, Rahman B, Dwyer DE, Wang Q. A cluster randomised trial of cloth masks compared with medical masks in healthcare workers. *BMJ Open* 2015: 5(4): e006577.

197. Leung CC, Cheng KK, Lam TH, Migliori GB. Masking to complement social distancing in saving lives in COVID-19 pandemic. *Int J Tuberc Lung Dis* 2020.

198. Esposito S, Principi N, Leung CC, Migliori GB. Universal use of face masks for success against COVID-19: evidence and implications for prevention policies. *Eur Respir J* 2020.

199. ECDC. Using face masks in the community: Reducing COVID-19 transmission from potentially asymptomatic or pre-symptomatic people through the use of face masks 2020.

200. Chen YC, Huang LM, Chan CC, Su CP, Chang SC, Chang YY, Chen ML, Hung CC, Chen WJ, Lin FY, Lee YT, Medicine SRGoNTUCo, National Taiwan University H. SARS in hospital emergency room. *Emerg Infect Dis* 2004: 10(5): 782-788.

201. WHO. Modes of transmission of virus causing COVID-19: implications for IPC precaution recommendations. 2020 29 March 2020 [cited 9th May 2020]; Available from: https://www.who.int/news-room/commentaries/detail/modes-of-transmission-of-virus-causing-covid-19-implications-for-ipc-precaution-recommendations

202. Lai MY, Cheng PK, Lim WW. Survival of severe acute respiratory syndrome coronavirus. *Clin Infect Dis* 2005: 41(7): e67-71.

203. Rabenau HF, Cinatl J, Morgenstern B, Bauer G, Preiser W, Doerr HW. Stability and inactivation of SARS coronavirus. *Med Microbiol Immunol* 2005: 194(1): 1-6.

204. ECDC. ECDC Technical report: Disinfection of environments in healthcare and nonhealthcare settings potentially contaminated with SARS-CoV-2. 2020.

205. Duan SM, Zhao XS, Wen RF, Huang JJ, Pi GH, Zhang SX, Han J, Bi SL, Ruan L, Dong XP, Team SR. Stability of SARS coronavirus in human specimens and environment and its sensitivity to heating and UV irradiation. *Biomed Environ Sci* 2003: 16(3): 246-255.

206. Berger A, Drosten C, Doerr HW, Sturmer M, Preiser W. Severe acute respiratory syndrome (SARS)--paradigm of an emerging viral infection. *J Clin Virol* 2004: 29(1): 13-22.

207. Bonny TS, Yezli S, Lednicky JA. Isolation and identification of human coronavirus 229E from frequently touched environmental surfaces of a university classroom that is cleaned daily. *Am J Infect Control* 2018: 46(1): 105-107.

208. Dara M SG, Reichler MR, Chiang CY, Chee CBE, Migliori GB. New diseases and old threats: lessons from tuberculosis for the COVID-19 response. *Int J Tuberc Lung Dis* 2020.

209. McKee M, Stuckler D. If the world fails to protect the economy, COVID-19 will damage health not just now but also in the future. *Nat Med* 2020.

210. WHO. COVID-19 Strategic Preparedness and Response Plan: OPERATIONAL PLANNING GUIDELINES TO SUPPORT COUNTRY PREPAREDNESS AND RESPONSE. . 2020.

211. Qian X, Ren R, Wang Y, Guo Y, Fang J, Wu ZD, Liu PL, Han TR, Members of Steering Committee SoGHCPMA. Fighting against the common enemy of COVID-19: a practice of building a community with a shared future for mankind. *Infect Dis Poverty* 2020: 9(1): 34.
212. WHO. Critical preparedness, readiness and response actions for COVID-19. Technical Guidance 2020 [cited 13 April 2020]; Available from: https://www.who.int/emergencies/diseases/novel-coronavirus-2019/technical-guidance/critical-preparedness-readiness-and-response-actions-for-covid-19

213. Smith AC, Thomas E, Snoswell CL, Haydon H, Mehrotra A, Clemensen J, Caffery LJ. Telehealth for global emergencies: Implications for coronavirus disease 2019 (COVID-19). J Telemed Telecare 2020: 1357633X20916567.

214. Civile P. Coronavirus emergency. 2020 [cited 13 April 2020]; Available from: http://www.protezionecivile.gov.it/home

215. Dubayova T, van Dijk JP, Nagyova I, Rosenberger J, Havlikova E, Gdovinova Z, Middel B, Groothoff JW. The impact of the intensity of fear on patient's delay regarding health care seeking behavior: a systematic review. Int J Public Health 2010: 55(5): 459-468.

216. ECDC. Coronavirus disease 2019 (COVID-19) in the EU/EEA and the UK – eighth update. 2020.

217. WHO. WHO Information Note: Tuberculosis and COVID-19. 2020.

218. Soo RJJ, Chiew CJ, Ma S, Pung R, Lee V. Decreased Influenza Incidence under COVID-19 Control Measures, Singapore. Emerg Infect Dis 2020: 26(8).

219. Kuo SC, Shih SM, Chien LH, Hsiung CA. Collateral Benefit of COVID-19 Control Measures on Influenza Activity, Taiwan. Emerg Infect Dis 2020: 26(8).

220. Saunders MJ, Evans CA. Fighting poverty to prevent tuberculosis. Lancet Infect Dis 2016: 16(4): 395-396.

221. Carter DJ, Glaziou P, Lonroth K, Siroka A, Floyd K, Weil D, Raviglione M, Houben R, Boccia D. The impact of social protection and poverty elimination on global tuberculosis incidence: a statistical modelling analysis of Sustainable Development Goal 1. Lancet Glob Health 2018: 6(5): e514-e522.

222. Migliori GB, Garcia-Basteiro AL. Predicting the effect of improved socioeconomic health determinants on the tuberculosis epidemic. Lancet Glob Health 2018: 6(5): e475-e476.

223. WHO. Report of the WHO-China Joint Mission on Coronavirus Disease 2019 (COVID-19) 2020.

224. CDC. Guidelines for Environmental Infection Control in Health-Care Facilities (2003). 2003.

225. Dharmadhikari AS, Mphahlele M, Stoltz A, Venter K, Mathebula R, Masotla T, Lubbe W, Pagano M, First M, Jensen PA, van der Walt M, Nardell EA. Surgical face masks worn by patients with multidrug-resistant tuberculosis: impact on infectivity of air on a hospital ward. Am J Respir Crit Care Med 2012: 185(10): 1104-1109.

226. ECDC. Cloth masks and mask sterilisation as options in case of shortage of surgical masks and respirators. 2020.

227. Li Q, Guan X, Wu P, Wang X, Zhou L, Tong Y, Ren R, Leung KSM, Lau EHY, Wong JY, Xing X, Xiang N, Wu Y, Li C, Chen Q, Li D, Liu T, Zhao J, Liu M, Tu W, Chen C, Jin L, Yang R, Wang Q, Zhou S, Wang R, Liu H, Luo Y, Liu Y, Shao G, Li H, Tao Z, Yang Y, Deng Z, Liu B, Ma Z, Zhang Y, Shi G, Lam TTY, Wu JT, Gao GF, Cowling BJ, Yang B, Leung GM, Feng Z. Early Transmission Dynamics in Wuhan, China, of Novel Coronavirus-Infected Pneumonia. N Engl J Med 2020: 382(13): 1199-1207.

228. Prem K, Liu Y, Russell TW, Kucharski AJ, Eggo RM, Davies N, Centre for the Mathematical Modelling of Infectious Diseases C-WG, Jit M, Klepac P. The effect of control strategies to reduce social mixing on outcomes of the COVID-19 epidemic in Wuhan, China: a modelling study. Lancet Public Health 2020.

229. Blaising J, Polyak SJ, Pecheur EJ. Arbidol as a broad-spectrum antiviral: an update. Antiviral Res 2014: 107: 84-94.
230. Hoffmann M, Kleine-Weber H, Schroeder S, Kruger N, Herrler T, Erichsen S, Schiergens TS, Herrler G, Wu NH, Nitsche A, Muller MA, Drosten C, Pohllmann S. SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell* 2020.

231. Richardson P, Griffin I, Tucker C, Smith D, Oechsle O, Phelan A, Stebbing J. Baricitinib as potential treatment for 2019-nCoV acute respiratory disease. *Lancet* 2020: 395(10223): e30-e31.

232. Morse JS, Lalonde T, Xu S, Liu WR. Learning from the Past: Possible Urgent Prevention and Treatment Options for Severe Acute Respiratory Infections Caused by 2019-nCoV. *Chembiochem* 2020: 21(5): 730-738.

233. Jordan PC, Stevens SK, Deval J. Nucleosides for the treatment of respiratory RNA virus infections. *Antivir Chem Chemother* 2018: 26: 2040206618764483.

234. Furuta Y, Gowen BB, Takahashi K, Shiraki K, Smee DF, Barnard DL. Favipiravir (T-705), a novel viral RNA polymerase inhibitor. *Antiviral Res* 2013: 100(2): 446-454.

235. Shiraki K, Daikoku T. Favipiravir, an anti-influenza drug against life-threatening RNA virus infections. *Pharmacol Ther* 2020: 107512.

236. Wang RR, Yang QH, Luo RH, Peng YM, Dai SX, Zhang XJ, Chen H, Cui XQ, Liu YJ, Huang JF, Chang JB, Zheng YT. Avudine, a novel nucleoside reverse transcriptase inhibitor showed good drug combination features and better inhibition on drug-resistant strains than lamivudine in vitro. *PLoS One* 2014: 9(8): e105617.

237. Abraham GM, Morton JB, Saravolatz LD. Baloxavir: A Novel Antiviral Agent in the Treatment of Influenza. *Clin Infect Dis* 2020.

238. Gordon CJ, Tchesnokov EP, Feng JY, Porter DP, Gotte M. The antiviral compound remdesivir potently inhibits RNA-dependent RNA polymerase from Middle East respiratory syndrome coronavirus. *J Biol Chem* 2020.

239. Busti AJ, Hall RG, Margolis DM. Atazanavir for the treatment of human immunodeficiency virus infection. *Pharmacotherapy* 2004: 24(12): 1732-1747.

240. Shionogi & Co. Ltd. Review Report: XofluxaTM (baloxavir). 2018 [cited 2020 6th April]; Available from: https://www.pmda.go.jp/files/000225380.pdf

241. Madelain V, Nguyen TH, Olivo A, de Lamballerie X, Guedj J, Taburet AM, Mentre F. Ebola Virus Infection: Review of the Pharmacokinetic and Pharmacodynamic Properties of Drugs Considered for Testing in Human Efficacy Trials. *Clin Pharmacokinet* 2016: 55(8): 907-923.

242. Louthrenoo W, Hongsongkiat S, Kasitanon N, Wangkaew S, Jatuworapruk K. Effect of Antituberculous Drugs on Serum Uric Acid and Urine Uric Acid Excretion. *J Clin Rheumatol* 2015: 21(7): 346-348.

243. Liverpool Drug Interaction Group. Interactions with experimental COVID-19 therapies. 2020 [cited 2020 1st April]; Available from: https://www.covid19-druginteractions.org/

244. Pandie M, Wiesner L, McIlerson H, Hughes J, Siwendu S, Conradie F, Variava E, Maartens G. Drug-drug interactions between bedaquiline and the antiretrovirals lopinavir/ritonavir and nevirapine in HIV-infected patients with drug-resistant TB. *J Antimicrob Chemother* 2016: 71(4): 1037-1040.

245. Deng P, Zhong D, Yu K, Zhang Y, Wang T, Chen X. Pharmacokinetics, metabolism, and excretion of the antiviral drug arbidol in humans. *Antimicrob Agents Chemother* 2013: 57(4): 1743-1755.

246. Song JH, Fang ZZ, Zhu LL, Cao YF, Hu CM, Ge GB, Zhao DW. Glucuronidation of the broad-spectrum antiviral drug arbidol by UGT isoforms. *J Pharm Pharmacol* 2013: 65(4): 521-527.
247. Lu ZK, Yuan J, Li M, Sutton SS, Rao GA, Jacob S, Bennett CL. Cardiac risks associated with antibiotics: azithromycin and levofloxacin. Expert Opin Drug Saf 2015: 14(2): 295-303.

248. Chatre C, Roubille F, Vernhet H, Jorgensen C, Pers YM. Cardiac Complications Attributed to Chloroquine and Hydroxychloroquine: A Systematic Review of the Literature. Drug Saf 2018: 41(10): 919-931.

249. Krzeminski P, Lesiak A, Narbutt J. Seizures as a rare adverse effect of chloroquine therapy in systemic lupus erythematosus patients: a case report and literature survey. Postepy Dermatol Alergol 2018: 35(4): 429-430.

250. Bortoli R, Santiago M. Chloroquine ototoxicity. Clin Rheumatol 2007: 26(11): 1809-1810.

251. Malcangi G, Fraticelli P, Palmieri C, Cappelli M, Danielli MG. Hydroxychloroquine-induced seizure in a patient with systemic lupus erythematosus. Rheumatol Int 2000: 20(1): 31-33.

252. Zamorano JL, Lancellotti P, Rodriguez Munoz D, Aboyans V, Asteggiano R, Galderisi M, Habib G, Lenihan DJ, Lip GYH, Lyon AR, Lopez Fernandez T, Mohty D, Piepoli MF, Tamargo J, Torbicki A, Suter TM. 2016 ESC Position Paper on cancer treatments and cardiovascular toxicity developed under the auspices of the ESC Committee for Practice Guidelines: The Task Force for cancer treatments and cardiovascular toxicity of the European Society of Cardiology (ESC). Eur Heart J 2016: 37(36): 2768-2801.

253. Shakil AO, Di Bisceglie AM, Hoofnagle JH. Seizures during alpha interferon therapy. J Hepatol 1996: 24(1): 48-51.

254. Sharifian MR, Kamandi S, Sima HR, Zaringhalam MA, Bakhshaee M. INF-alpha and ototoxicity. Biomed Res Int 2013: 2013: 295327.

255. Genovese MC, Rubbert-Roth A, Smolen JS, Kremer J, Khaishi M, Gomez-Reino J, Sebba A, Pilson R, Williams S, Van Vollenhoven R. Longterm safety and efficacy of tocilizumab in patients with rheumatoid arthritis: a cumulative analysis of up to 4.6 years of exposure. J Rheumatol 2013: 40(6): 768-780.
