Research Article

Effects of Hemodialysis on Thiol-Disulphide Homeostasis in Critically Ill Pediatric Patients with Acute Kidney Injury

Ganime Ayar,1 Sanliay Sahin,1 Mutlu Uysal Yazıcı,2 Salim Neselioglu,3 Ozcan Erel,3 and Umut Selda Bayrakçı4

1University of Health Sciences, Ankara Child Health and Diseases, Hematology Oncology Training and Research Hospital, Pediatric Intensive Care Unit, Ankara, Turkey
2Hacettepe University, Faculty of Medicine, Department of Pediatric Intensive Care, Ankara, Turkey
3University of Yıldırım Beyazıt, Ankara Atatürk Education and Research Hospital, Department of Biochemistry, Ankara, Turkey
4University of Yıldırım Beyazıt, Department of Pediatric Nephrology, Ankara, Turkey

Correspondence should be addressed to Ganime Ayar; ganimeayar@gmail.com

Received 27 June 2018; Revised 26 August 2018; Accepted 10 September 2018; Published 25 September 2018

Academic Editor: Koichiro Wada

Copyright © 2018 Ganime Ayar et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Aim. To evaluate thiol/disulphide homeostasis as a new indicator of oxidative stress in AKI patients and to determine the effect of HD on antioxidant balance and oxidative stress through plasma thiols. Methods. This study was performed in patients aged between 12 months and 18 years prospectively who underwent hemodialysis due to AKI and were followed up for a year in a 22-bed tertiary pediatric intensive care unit. 20 patients and 39 controls were included. Results. No difference was present between the groups in terms of age and gender. Median values of plasma native thiol, total thiol, and percent thiol were significantly lower in AKI group both before and after dialysis when compared to control group. The median dynamic disulphide values were significantly lower in the AKI group of predialysis compared to the controls. When pre- and postdialysis values were compared, disulphide values were statistically higher after dialysis. When pre- and postdialysis native thiol, dynamic disulphide, total thiol, and percent thiol median values were compared, postdialysis values were significantly higher than the predialysis values. There was a positive correlation between albumin, total thiol, and native thiol values before dialysis in the patient group. Conclusion. AKI patients have low levels of thiol species showing the presence of oxidative stress and hemodialysis has a positive effect on thiol/disulphide balance. This new method may be an inexpensive and simple tool suitable for clinical studies and can be used in routine screening as a useful indicator to show oxidative stress.

1. Introduction

Acute kidney injury (AKI) is a medical problem characterized by the accumulation of breakdown products in blood as a result of the decrease in glomerular filtration rate within hours/days, which causes mortality by 50% to 80% in paediatric intensive care unit (PICU) [1, 2]. Although the pathogenesis of AKI is not clear, free oxygen radicals and antioxidant balance are involved in the pathogenesis [3–5]. Proximal tubular cells and renal medulla are especially prone to hypoxia and ischemia and poor oxygenation causes the development of tubular damage [6].

Thiol is a compound which contains sulphydryl group (-SH) that plays a crucial role in preventing the occurrence of oxidative stress in the cells. Thiols are not only the redox tampon in the body but also a principal element of antioxidant defence system [7]. While the majority of plasma thiols mainly consist of albumin and other proteins, a minor part involves low molecular weight thiols such as cysteine, cysteinyl-glycine, glutathione, homocysteine, and γ-glutamylcysteine [8, 9]. Thiol groups of protein, low molecular weight compounds, cysteine residues, and the other thiol groups are oxidized by the oxidant molecules and change into reversible disulphide connective structures. The reversible disulphide connective structures can be reduced to thiol groups and therefore thiol disulphide balance is maintained. Abnormal thiol-disulphide balance levels are associated with oxidative stress [10–13]. Oxidative stress leads
to oxidative destruction of lipids and other macromolecules, results in the change of the cell membrane and other cellular elements, and causes cell death and thus tissue damage and chronic diseases [9, 10]. There is a balance between oxidative stress and antioxidant defence mechanisms. There is a strong evidence of increase in morbidity of hemodialysis (HD) patients due to impairment of this balance by multiple factors [14–16]. Although HD is a commonly used renal replacement treatment, the role of dialysis in the prognosis of AKI is not fully elucidated yet [1].

Therefore, we aimed in this study to determine the thiol/disulphide balance as a new indicator of oxidative stress in AKI patients and to evaluate the effects of HD on antioxidant balance and oxidative stress through plasma thiols.

2. Material and Methods

This cross-sectional study was conducted in accordance with the Declaration of Helsinki and was approved by the Hospital Research Ethics Committee. Informed consent was obtained from all participants and their parents before the study. Volunteers were selected from patients who were consecutively admitted to the PICU with AKI requiring hemodialysis. Twenty patients aged between 12 months and 18 years with AKI and 39 healthy subjects were included in the study.

This study was performed at the Ankara Children's Haematology and Oncology Training and Research Hospital between June 2015 and July 2017. The thiol/disulphide homeostasis parameters were studied in the Yıldırım Beyazıt University Biochemistry Laboratory. Exclusion criteria for patients were the existence of chronic kidney failure, any hepatic disease, any form of vitamin supplementation, chromosomal and genetic syndromes, amyloidosis, severe anaemia, and malnutrition.

Physically and mentally healthy volunteers who applied to the paediatrics outpatient clinic, with no acute or chronic disease, required blood draw for any reason, and did not take any antioxidants or supplements were selected as the control group.

During the study, we used bicarbonate dialysis with polysulfone dialyzer membrane for all patients. Two millimetres of blood was collected from the hemodialysis catheter before and after dialysis from the patients who received hemodialysis during the routine blood draw. Venous blood samples of control groups were collected in tubes containing Ethylenediaminetetraacetic acid (EDTA). As soon as the blood samples are collected by using vacutainer, they are centrifuged at 3600 rpm x for 10 minutes and stored in the freezer at −80°C.

2.1. Measurement of the Thiol-Disulphide Homeostasis Parameters. Thiol-disulphide homeostasis consists of native thiol (−SH), total thiol (−SH+SS), disulphide (−SS), and disulphide/native thiol percent ratio (−SS/−SH %) parameters. Serum native thiol and total thiol levels were measured with a recently developed, automatic measurement method by using an automated clinical chemistry analyzer (Roche Hitachi Cobas c501 automatic analyzer, Roche Diagnostics, USA) [17]. Disulphide level and disulphide/native thiol percent ratio (index) were generated from the measured values. The half value of the difference between total thiol and native thiol amounts gave the disulphide bond amount. The index parameter was calculated based on percent ratio of disulphide and native thiol levels.

3. Statistical Analysis

Statistics were expressed as numbers and percentages for categorical variables for descriptive analysis and as mean, standard deviation, median, and 25th percentile (Q1), 75th percentile (Q3) for numerical variables. Normality of distribution was evaluated using the Shapiro–Wilks test. For two-group comparisons of non-normally distributed numerical variables, Mann-Whitney U test was employed. Wilcoxon signed rank test was used for comparison of the parameters studied before and after the dialysis. The correlation was evaluated using Spearman’s correlation test. SPSS Statistics for Windows version 21.0 was used for the analysis. The statistical significance level was P<0.05.

4. Results

Demographics and the laboratory data of the patient group are given in Table 1. No difference was present between patient and control groups regarding age and gender. The diagnosis of patients is summarized in Table 2. Table 3 demonstrates the comparison of thiol/disulphide homeostatic parameters in hemodialysis group. When pre- and postdialysis total thiol, native thiol, and dynamic disulphide median values were compared, postdialysis values were found to be significantly higher than the predialysis values (p<0.05). When pre- and postdialysis percent thiol values were compared, there was no statistically significant difference.

Median values of plasma native thiol, total thiol, and percent thiol were meaningfully lower in AKI group both before and after dialysis in comparison to the control group (Table 4). Total thiol was 482.00 mmol/L (IQR= 465.20-505.20) in the control group, 256.45 mmol/L before dialysis (IQR= 179.75-318-73) p=0.0001, and 324.05 mmol/L after dialysis (IQR= 276.00-410.63) p=0.0001, while native thiol (IQR= 179.75-318-73) p=0.0001, and 324.05 mmol/L after dialysis (IQR= 276.00-410.63) p=0.0001, while native thiol was 453.40 mmol/L in the control group, (IQR=431.90-464.30), 230.70 mmol/L before dialysis (IQR=182.63-293.60), and 294.45 mmol/L after dialysis (IQR=244.58-359.95), p=0.0001. There was a positive correlation between albumin, total thiol, and native thiol values before and after dialysis in the patient group (Table 5).

5. Discussion

The dynamic thiol/disulphide status is important in diseases where oxidative stress plays a crucial role in the pathogenesis. Thiols can undergo oxidation reaction via oxidants and form disulphide bonds. A disulphide bond is a covalent bond; the linkage is also called a SS-bond or disulphide bridge. Under conditions of oxidative stress, the oxidation of Cys residues
important in the pathogenesis of oxidative stress, rates of -SS x 100 / -SH, -SS x 100 / -SS + -SH, and -SH x 100 / -SS + -SH were investigated. Our results indicated that blood thiols were significantly low in these pediatric patients with AKI. Protein thiols are the most important redox pool, and thus low redox rates show the presence of a high oxidative load. Many studies showed that the level of oxidant radicals increases in chronic renal failure (CRF) [23–26]. The increase of oxidant radical and the decrease of plasma antioxidant activity contribute to an increase in oxidative damage and development of renal complications [25]. In addition, oxidant radicals and antioxidant balance are involved in the pathogenesis of acute renal failure [3–5]; antioxidants such as superoxide dismutase have been proved to protect the person against acute renal injury related to endotoxemia and oxygen radicals causing tubule damage during ischemia [4, 5].

In patients with renal failure, the increase in oxidant radical level is multifactorial; even one of them is mentioned to be hemodialysis [23, 25]. Oxidative stress increased during hemodialysis due to numerous factors such as uremic toxicity, malnutrition, anemia, and clinical status of the patients. HD also stimulates inflammation, increasing the production of oxidant, and results in increased oxidative stress [23, 27]. It is known that when the production of oxidant agents and antioxidant defence mechanisms is imbalanced, an important source of morbidity arises [28].

In our results, there were significantly higher postdialysis values of native thiol, dynamic disulphide, and total thiol compared to predialysis values. There are similar studies in the literature showing the effect of hemodialysis on antioxidant homeostasis [10, 24, 26, 29]. In a study conducted by Colombo et al., it was found that lower plasma levels of total thiols in patients with end-stage kidney disease compared to the healthy controls were the same as those of the healthy persons’ after dialysis, and they attributed this result to hemodialysis being the first session [26]. Although in our results postdialysis plasma thiol levels were not the same as those of the healthy group, these values have increased compared to the predialysis. As in our study, hemodialysis sessions may restore plasma thiol redox status in the acute period, but antioxidant capacity may be insufficient as the status becomes chronic. There are different studies which demonstrated the total serum antioxidant levels in chronic hemodialyzed patients compared to the healthy group [30]. Furthermore, some studies have found higher total serum

| Parameters                        | Case Group (n=20) | Control Group (n=39) | P     |
|----------------------------------|------------------|----------------------|-------|
| Sex (females/males)              | 11/9             | 22/17                | 0.918 |
| Age, years*                      | 12 (7.5-15.8)    | 10 (7.0-13.0)        | 0.265 |
| Hemoglobin (g/dL) pre-dialysis*  | 9.2 (7.9-10.4)   | 11.8 (10.7-12.1)     | <0.001|
| Albumin (g/dL) pre-dialysis*     | 3.5 (3.0-3.9)    | 4.2 (3.9-4.4)        | <0.001|
| Creatinine (mg/dL) pre-dialysis* | 3.3 (2.2-5.7)    | 0.4 (0.3-0.5)        | <0.001|

* Median (Q1-Q3).
Table 3: Comparison of thiol/disulphide homeostatic parameters in hemodialysis group.

|                      | Pre-dialysis | Post-dialysis | P    |
|----------------------|--------------|---------------|------|
|                      | Median (Q1-Q3) | Median (Q1-Q3) |      |
| Native thiol (µmol/L) | 230.70(182.63-293.60) | 294.45(244.58-359.95) | 0.003* |
| Total thiol (µmol/L)  | 256.45(179.75-318.73) | 324.05(276.00-410.63) | 0.003* |
| Disulphide (µmol/L)   | 13.50(10.56-17.31) | 15.35(12.92-21.15) | 0.021* |
| Disulphide/native thiol (%) | 5.62(4.44-7.88) | 6.15(4.16-7.77) | 0.747 |
| Disulphide/total thiol (%) | 5.08(4.03-6.96) | 5.47(3.84-6.72) | 0.778 |
| Native thiol/total thiol (%) | 89.00(85.50-91.83) | 89.70(86.54-92.46) | 0.841 |

*p < 0.05 was considered significant for statistical analyses (Wilcoxon signed rank test).

Table 4: Comparison of thiol/disulphide homeostatic parameters between control and hemodialysis group.

|                      | Control | P Pre-dialysis vs. Control | P Post-dialysis vs. Control |
|----------------------|---------|---------------------------|----------------------------|
|                      | Median (Q1-Q3) |                          |                            |
| Native thiol (µmol/L) | 453.40(431.90-464.30) | 0.0001*                   | 0.0001*                    |
| Total thiol (µmol/L)  | 482.00(465.20-505.20) | 0.0001*                   | 0.0001*                    |
| Disulphide (µmol/L)   | 19.55(14.95-21.70)   | 0.0001*                   | 0.113                      |
| Disulphide/native thiol (%) | 4.15(3.26-4.82) | 0.001*                    | 0.001*                     |
| Disulphide/total thiol (%) | 3.83(3.06-4.40) | 0.002*                    | 0.001*                     |
| Native thiol/total thiol (%) | 92.33(91.20-93.87) | 0.001*                    | 0.002*                     |

*p < 0.01 was considered significant for statistical analyses (Mann-Whitney U test).

Table 5: Correlations between thiol parameters and other parameters.

|                      | Pre-dialysis | Post-dialysis | Control |
|----------------------|--------------|---------------|---------|
|                      | r            | p             | r       | p       | r       | p       |
| Native thiol         |              |               | Total thiol |         | Disulphide |         |
| Age                  | -0.291       | 0.214         | -0.303   | 0.195   | 0.124   | 0.602   |
| Hemoglobin           | -0.333       | 0.152         | -0.337   | 0.146   | -0.288  | 0.219   |
| Albumin              | 0.646        | 0.002*        | 0.643    | 0.002*  | 0.192   | 0.418   |
| Creatinine           | 0.087        | 0.715         | 0.113    | 0.636   | 0.361   | 0.118   |
| Disulphide           |              |               |          |         |         |         |
| Age                  | -0.132       | 0.580         | -0.110   | 0.643   | -0.203  | 0.391   |
| Hemoglobin           | -0.342       | 0.140         | -0.339   | 0.144   | 0.075   | 0.755   |
| Albumin              | 0.445        | 0.049*        | 0.464    | 0.039*  | 0.366   | 0.112   |
| Creatinine           | 0.057        | 0.811         | 0.042    | 0.860   | -0.299  | 0.200   |

*p < 0.05 was considered significant for statistical analyses (Spearman’s correlation test).

antioxidant levels at predialysis [31, 32], and some have found a significant decrease in antioxidants after dialysis [32]. Different results might be caused by different tests used for measurement of antioxidants and the results might be affected by many patient-related factors [7]. However, we consider our study important since the plasma thiols are the largest antioxidant components of serum.

The difference between the patient and control groups in our study was statistically significant. Lower plasma thiol levels in predialysis and postdialysis, compared to the control
group, were consistent with the literature [16, 33, 34], suggesting that these patients are susceptible to oxidative damage. On the other hand, postdialysis thiol values were significantly higher than those of the predialysis, so that hemodialysis may have a positive effect on the oxidative balance in patients with AKI.

Among other antioxidant molecules, albumin was also low in the study group, and lower levels of albumin were found in predialysis patients, when compared to the postdialysis which suggests us that there was a positive correlation between albumin and thiol values.

Our study has a few limitations. The most significant one of these was the limited number of patients. And we ignored the number of patients during the dialysis session while obtaining blood samples. The single centred study was another limitation of this study.

6. Conclusion

This study showed that HD may affect the thiol/disulfide homeostasis. According to the findings, AKI patients have low levels of thiol species showing the presence of oxidative stress; and hemodialysis has a positive effect on thiol/disulfide homeostasis. Although further studies with larger sample groups including recurring dialysis sessions are required, it must be underlined that this new method may be an inexpensive and simple instrument suitable for medical studies that can be applied to routine screening as a useful indicator to show thiol-specific oxidative stress.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare no conflicts of interest.

References

[1] R. W. Schrier, W. Wang, B. Poole, and A. Mitra, "Acute renal failure: definitions, diagnosis, pathogenesis, and therapy," The Journal of Clinical Investigation, vol. 114, no. 1, pp. 5–14, 2004.
[2] K. Makris and L. Spanou, "Acute kidney injury: definition, pathophysiology and clinical phenotypes," The Clinical Biochemical Reviews, vol. 37, no. 2, pp. 85–98, 2016.
[3] E. Y. Plotnikov, T. A. Pavlenko, I. B. Pevzner et al., "The role of oxidative stress in acute renal injury of newborn rats exposed to hypoxia and endotoxin," FEBS Journal, vol. 284, no. 18, pp. 3069–3078, 2017.
[4] W. Wang, S. Jittikanont, S. A. Falk et al., "Interaction among nitric oxide, reactive oxygen species, and antioxidants during endotoxemia-related acute renal failure," American Journal of Physiology-Renal Physiology, vol. 284, no. 3, pp. F532–F537, 2003.
[5] E. Noiri, A. Nakao, K. Uchida et al., "Oxidative and nitrosative stress in acute renal ischemia," American Journal of Physiology-Renal Physiology, vol. 281, no. 5, pp. F948–F957, 2001.
[6] G. P. Kaushal, A. G. Basnakian, and S. V. Shah, "Apoptotic pathways in ischemic acute renal failure," Kidney International, vol. 66, no. 2, pp. 500–506, 2004.
[7] J. R. Winther and C. Thorpe, "Quantification of thiols and disulfides," Biochimica et Biophysica Acta (BBA) - General Subjects, vol. 1840, no. 2, pp. 838–846, 2014.
[8] L. Turell, R. Radi, and B. Alvarez, "The thiol pool in human plasma: The central contribution of albumin to redox processes," Free Radical Biology & Medicine, vol. 65, pp. 244–253, 2013.
[9] D. P. Jones, "Redefining oxidative stress," Antioxidants & Redox Signaling, vol. 8, no. 9-10, pp. 1865–1879, 2006.
[10] P. Soysal, A. T. Isik, A. F. Carvalho et al., "Oxidative stress and frailty: A systematic review and synthesis of the best evidence," Maturitas, vol. 99, pp. 66–72, 2017.
[11] S. D. Rodrigues, G. B. Batista, M. Inghererman, R. Pecoits-Filho, and L. S. Nakao, "Plasma cysteine/cystine reduction potential correlates with plasma creatinine levels in chronic kidney disease," Blood Purification, vol. 34, no. 3-4, pp. 231–237, 2013.
[12] M. Rosas-Diaz, M. Camarillo-Cadena, A. Hernández-Arana, E. Ramón-Gallegos, and R. Medina-Navarro, "Antioxidant capacity and structural changes of human serum albumin from patients in advanced stages of diabetic nephropathy and the effect of the dialysis," Molecular and Cellular Biochemistry, vol. 404, no. 1-2, pp. 193–201, 2015.
[13] S. C. Trelervin and L. R. Lopes, "Protein disulfide isomerase and nox: New partners in redox signaling," Current Pharmaceutical Design, vol. 21, no. 41, pp. 5951–5963, 2015.
[14] A. V. Crowe, A. Mc Ardle, F. McArdle et al., "Markers of oxidative stress in the skeletal muscle of patients on haemodialysis," Nephrology Dialysis Transplantation, vol. 22, no. 4, pp. 1177–1183, 2007.
[15] L. Signorini, S. Granata, A. Lupo, and G. Zaza, "Naturally occurring compounds: New potential weapons against oxidative stress in chronic kidney disease," International Journal of Molecular Sciences, vol. 18, no. 7, 2017.
[16] F. Galli, M. Piroddi, D. Bartolini et al., "Blood thiol status and erythrocyte glutathione-S-transferase in chronickidneydisease," Nephrology Dialysis Transplantation, vol. 22, no. 4, pp. 273–281, 2014.
[17] O. Erel and S. Nerselioğlu, "A novel and automated assay for thiol/disulphide homeostasis," Clinical Biochemistry, vol. 47, no. 18, pp. 326–332, 2014.
[18] W. Dröge, "Aging-related changes in the thiol/disulfide redox state: implications for the use of thiol antioxidants," Experimental Gerontontology, vol. 37, no. 12, pp. 1333–1345, 2002.
[19] R. E. Hansen, D. Roth, and J. R. Winther, "Quantifying the global cellular thiol-disulfide status," Proceedings of the National Academy of Sciences of the United States of America, vol. 106, no. 2, pp. 422–427, 2009.
[20] G. Vural, H. Bektas, S. Gumusyayla, O. Deniz, M. Alışık, and O. Erel, "Impaired thiol-disulphide homeostasis in patients with axonalpolyneuropathy," Neurological Research, vol. 40, no. 3, pp. 166–172, 2018.
[21] R. S. Derde, M. Godemir, O. Erel, H. Buyukaslan, and H. Karabag, "The relationship between serum thiol levels and thiol/disulfide homeostasis with head trauma in children," Clinical Laboratory, vol. 64, no. 1, 2018.
[22] K. Ünal, G. Erzin, R. N. Yüksel, M. Alışık, and O. Erel, "Thiol/disulphide homeostasis in schizophreniapatients with positive symptoms," Nordic Journal of Psychiatry, pp. 1–4, 2018.
[23] M. Morena, S. Delbosc, A.-M. Dupuy, B. Canaud, and J.-P. Cristol, “Overproduction of reactive oxygen species in end-stage renal disease patients: a potential component of hemodialysis-associated inflammation,” *Hemodialysis International*, vol. 9, no. 1, pp. 37–46, 2005.

[24] T. Apeland, O. Kristensen, and M. A. Mansoor, “The aminothiol redox status in haemodialysis patients does not improve with folate therapy,” *Scandinavian Journal of Clinical & Laboratory Investigation*, vol. 69, no. 2, pp. 265–271, 2009.

[25] J. Stępniewska, E. Golembiewska, B. Dołęgowska, M. Domaniński, and K. Ciechanowski, “Oxidative stress and antioxidative enzyme activities in chronic kidney disease and different types of renal replacement therapy,” *Current Protein & Peptide Science*, vol. 16, no. 3, pp. 243–248, 2015.

[26] G. Colombo, F. Reggiani, M. A. Podestà et al., “Plasma protein thiolation index (PTI) as a biomarker of thiol-specific oxidative stress in haemodialyzed patients,” *Free Radical Biology & Medicine*, vol. 89, pp. 443–451, 2015.

[27] A. Y. Putri and M. Thaha, “Role of oxidative stress on chronic kidney disease progression,” *Acta Medica Indonesiana (The Indonesian Journal of Internal Medicine)*, vol. 46, no. 3, pp. 244–252, 2014.

[28] M. L. Wratten, C. Tetta, F. Ursini, and A. Sevanian, “Oxidant stress in hemodialysis: Prevention and treatment strategies,” *Kidney International Supplements*, vol. 58, no. 76, pp. S126–S132, 2000.

[29] F. A. Köse, M. Seziş, F. Akçıçek, and A. Pabuççuoğlu, “Oxidative and nitrosative stress markers in patients on hemodialysis and peritoneal dialysis,” *Blood Purification*, vol. 32, no. 3, pp. 202–208, 2011.

[30] P. Castilla, R. Echarri, A. Dávalos et al., “Concentrated red grape juice exerts antioxidant, hypolipidemic, and antiinflammatory effects in both hemodialysis patients and healthy subjects,” *American Journal of Clinical Nutrition*, vol. 84, no. 1, pp. 252–262, 2006.

[31] C. Erdoğan, Y. Ünlüçerçi, A. Türkmen, A. Kuru, Ö. Çetin, and S. Bekpinar, “The evaluation of oxidative stress in patients with chronic renal failure,” *Clinica Chimica Acta*, vol. 322, no. 1-2, pp. 157–161, 2002.

[32] T. Ruskovska, E. H. J. M. Jansen, and R. Antarorov, “Evaluation of assays for measurement of serum (Anti)oxidants in hemodialysis patients,” *BioMed Research International*, vol. 2014, 2014.

[33] D. Zwolińska, W. Grzeszczak, M. Szczepańska, K. Klisi-Pstrusińska, and K. Szprynger, “Lipid peroxidation and antioxidative enzymes in children on maintenance dialysis,” *Pediatric Nephrology*, vol. 21, no. 5, pp. 705–710, 2006.

[34] A. Haciçevki, “Effect of hemodialysis on oxidative stress in patients with chronic renal failure,” *Ankara Üniversitesi Eczacilik Fakultesi Dergisi*, vol. 37, no. 2, pp. 91–100, 2008.