Research article

Synthesis and Antifungal Evaluation of Novel N-Alkyl Tetra- and Perhydroquinoline Derivatives

Jürgen KRAUSS *, Michal HORNACEK, Christoph MÜLLER, Verena STAUDACHER, Martina STADLER, Franz BRACHER

Department of Pharmacy – Center for Drug Research, Ludwig-Maximilian-University, Butenandtstr. 5–13, 81377 Munich, Germany.

* Corresponding author. E-mail: Juergen.Krauss@cup.uni-muenchen.de (J. Krauß)

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Abstract

A series of novel N-alkyl tetra- and perhydroquinoline derivatives and their hydrochlorides were prepared from tetrahydro- or trans-perhydroquinoline by direct alkylation with alkyl halides and subsequent precipitation with HCl gas. The antimicrobial activity of the resulting amines was evaluated in an agar diffusion assay. The minimal inhibitory concentrations (MIC) of the active compounds were determined by the microdilution method. In contrast to the tetrahydroquinolines, the perhydro analogues showed significant antifungal activity. In an assay for the detection of target enzymes in ergosterol biosynthesis, N-undecylperhydroquinoline was identified as an inhibitor of Δ8,7-isomerase.

Keywords

Quinoline • Antifungal activity • Ergosterol biosynthesis • Δ8,7-Isomerase • Enzyme inhibitor

Introduction

In the last few decades, a dramatic increase in fungal infections was observed in the Northern Hemisphere. Especially cancer patients, organ-engrafted patients, and immune-compromised patients (e.g. AIDS patients) are predisposed to systemic fungal infections by Candida or Aspergillus species with a high lethality. Only a few drugs from three classes can be used in the treatment of these life-threatening systemic infections: azoles (e.g. fluconazole, posaconazole, or voriconazole), polyene macrolides (e.g. amphotericin B), and echinocandins (e.g. caspofungin, anidulafungin, or micafungin) [1, 2] (Fig. 1).
Ergosterol biosynthesis is an extremely important target in the development of new antimycotic drugs [3]. Ergosterol and the enzymes in the post-squalene part of ergosterol biosynthesis are specific for fungi, so specific inhibitors should be selective for the fungal pathway. By now, only the four enzymes, squalene epoxidase (by allylamines), C14 demethylase (by azoles), and $\Delta_8,7$-isomerase and $\Delta_{14}$-reductase (both by morpholines), of this complex pathway are targeted by antimycotics used in human medicine.

The mimicry of carbocationic high energy intermediates of this biosynthesis pathway by protonated amines is an often-used approach towards inhibitors of $\Delta_{8,7}$-isomerase and $\Delta_{14}$-reductase. The most important drug in human medicine using this concept is the morpholine amorolfine (A), but this drug can only be used in topical formulations [4] (Fig. 3). Several morpholine antifungals like fenpropimorph or tridemorph, showing the same mechanism of action, are used in agrochemistry.

The same mechanism of action could be shown for $N$-alkylpiperidines like fenpropidin [5–7]. Rahier and coworkers found the same mechanism of action for complex $N$-alkylperhydroisoquinolines (B). In our group, $N$-n-undecyl-\textit{trans}-decahydroisoquinoline (C) was identified as a potent antifungal inhibiting the enzyme $\Delta_{8,7}$-isomerase [8] (Fig. 3). Related
imidazol-5-yl carbinols showed comparable antifungal activities, but surprisingly did not interfere with ergosterol biosynthesis [9].

![A](image1.png) ![B](image2.png) ![C](image3.png)

**Fig. 3.** Potent Δ14-reductase and Δ8,7-isomerase inhibitors.

The surprisingly high antifungal activities of simple $N$-alkyldecahydroisoquinolines and imidazol-5-yl carbinols, the observed outstanding role of the length of the alkyl side chain in both series, and the surprising differences in molecular mechanisms of actions prompted us to perform analogous investigations on a third heterocyclic scaffold, the quinoline ring system.

**Results and Discussion**

In continuation of our above-mentioned work [8, 9], we evaluated the antifungal potency of simple $N$-alkyl tetrahydro- and perhydroquinoline derivatives in the present work. As we found in previous work that $N$-n-alkyl substitutents with nine to twelve carbons led to the highest antifungal potency, we focused on side chains with a length of C$_9$ to C$_{12}$.

In a first series, 1,2,3,4-tetrahydroquinoline (1a), first deprotonated with NaH, was alkylated with unbranched C$_9$ to C$_{12}$ alkyl halides to give the tertiary amines 2a–d. The amines were dissolved in dry diethyl ether and precipitated with HCl gas to give the more stable hydrochlorides 3a–d. In an alternative approach, 2d was prepared from 1a and dodecanoyl chloride to give the amide 6, which was reduced with LiAlH$_4$ to give 2d (Scheme 1).

In a second series, (±)-trans-perhydroquinoline (1b) was alkylated with C$_9$ to C$_{12}$ alkyl halides in the same way as described above to give the tertiary amines 4a–d, which were precipitated with HCl gas to give the hydrochlorides 5a–d (Scheme 1). The trans stereochemistry was selected since it resembles the stereochemistry typically found in the connections of the rings in ergosterol intermediates. The trans configuration of 1b was confirmed by comparison of $^{13}$C-NMR data with literature data [10].

The resulting compounds and known compounds, 1-undecylpiperidine (7) [14] and 1-decylpiperazine (8a) [15], as well as 1-methyl-4-undecylpiperazine (8b) [16] (prepared in the same way), were tested in an agar diffusion assay [11] against Gram-positive (*Staphylococcus equorum* (DSMZ-Nr. 20675), *Streptococcus entericus* (DSZM-Nr. 14446)) and Gram-negative bacteria (*Escherichia coli* (DSMZ-Nr. 426), *Pseudomonas marginalis* (DSMZ-Nr. 7527)), as well as the fungi *Yarrowia lipolytica* (DSMZ-Nr. 1345), *Candida glabrata* (DSMZ-Nr.11226), *Aspergillus niger* (DSMZ-Nr. 1988), and *Hyphopichia burtonii* (DSMZ-Nr. 70663) (Table 1).
Sch. 1. Synthesis of N-alkyl tetrahydro- and decahydroquinoline derivatives

Tab. 1. Results of the agar diffusion assay [50 µg/disc, diameter of zones of total inhibition [mm], GI = growth inhibition]

| Compound | E. coli | P. marginalis | Staph. equorum | S. entericus | Y. lipolytica | C. glabrata | A. niger | H. burtonii |
|----------|---------|--------------|----------------|--------------|--------------|-------------|---------|-------------|
| 2a       | 0       | 0            | 0              | 0            | 0            | 0           | 0       | 0           |
| 2b       | 0       | 0            | 0              | 0            | 0            | 0           | 0       | 0           |
| 2c       | 0       | 0            | 0              | 0            | 0            | 0           | 0       | 0           |
| 2d       | 0       | 0            | 0              | 0            | 0            | 0           | 0       | 0           |
| 4a       | 13 (Gl) | 10           | 20             | 15           | 14           | 7           | 12      | 0           |
| 4b       | 15 (Gl) | 13           | 22             | 19           | 16           | 12 (Gl)    | 20      | 13          |
| 4c       | 11 (Gl) | 12           | 20             | 13           | 13           | 14 (Gl)    | 13      | 13          |
| 4d       | 14 (Gl) | 9            | 12             | 9            | 9            | 11 (Gl)    | 11      | 12          |
| 6        | 0       | 0            | 0              | 0            | 0            | 0           | 0       | 0           |
| 7        | 7       | 8            | 13 (Gl)        | 17 (Gl)      | 22           | 8           | 0       | 12          |
| 8a       | 7       | 8            | 13 (Gl)        | 22 (Gl)      | 8            | 20          | 25      | 25          |
| 8b       | 10      | 15           | 18             | 15           | 25           | 11          | 12 (Gl) | 14          |
| clotrimazole | n.t.   | n.t.         | n.t.           | n.t.         | 15           | 13          | 15      | 25          |
| terbinafine | n.t.   | n.t.         | n.t.           | n.t.         | 12           | 0           | 26      | 15 (Gl)     |
| tetracycline | 26     | 12           | 30             | 10           | 0            | 0           | 0       | 0           |
The minimal inhibitory concentrations (MIC) of the most active compounds from the agar diffusion assay were determined in a microdilution assay on Candida glabrata, Yarrowia lipolytica, and Saccharomyces cerevisiae [11]. For comparison, the N-undecylperhydroisoquinoline (C) [8] was also tested here. For solubility reasons, the hydrochlorides were used in this assay (Table 2).

### Tab. 2. Minimal inhibitory concentrations (MIC [µg/mL] values are the arithmetic average of two determinations)

| Compound | Candida glabrata | Yarrowia lipolytica | Saccharomyces cerevisiae |
|----------|------------------|---------------------|--------------------------|
| 5a       | 50               | 7.5                 | 2.5                      |
| 5b       | 37.5             | 2.5                 | 1                        |
| 5c       | 25               | 5                   | 0.25                     |
| 5d       | 5                | 2.5                 | 2.5                      |
| 7        | 12.5             | 7.5                 | 1.75                     |
| 8a       | 45               | 10                  | 5                        |
| 8b       | 25               | 10                  | 50                       |
| C        | 5                | 1.75                | 0.1                      |

clotrimazole 1 (MIC<sub>90</sub> Candida spp.) [17]
fluconazole 32 (MIC<sub>90</sub> Candida spp.) [17]
nystatine B 4 (MIC<sub>90</sub> Candida spp.) [17]

The cytotoxicity of the compounds 3a–d, 5a–d, and 8b was determined in an MTT test [13] against a human leukemia cell line (HL 60). All compounds tested showed a moderate cytotoxicity against this cell line with IC<sub>50</sub> values between 6 to 46 µM (Table 3).

### Tab. 3. IC<sub>50</sub> values against a HL 60 cell line (IC<sub>50</sub> values are the arithmetic average of three determinations)

| Compound | IC<sub>50</sub> [µM] | Compound | IC<sub>50</sub> [µM] |
|----------|----------------------|----------|----------------------|
| 3a       | 19                   | 5a       | 22                   |
| 3b       | 60                   | 5b       | 7                    |
| 3c       | 16                   | 5c       | 7                    |
| 3d       | 15                   | 5d       | 6                    |
| 8b       | 46                   |          |                      |

The compounds 5c and 7 were also subjected to our whole-cell assay for identification of the target enzyme in ergosterol biosynthesis [12]. In this assay, the strains Candida glabrata and Saccharomyces cerevisiae were incubated with the test compounds, and after cell lysis, the changes in the sterol pattern were analyzed by GLC-MS. The accumulation of the Δ8(9)-sterol lichesterol (ergosta-5,8,22-trien-3β-ol) clearly indicates an inhibition of the enzyme Δ8,7-isomerase. Both compounds 5c and 7 showed an accumulation of lichesterol, so one mechanism of action is an inhibition of Δ8,7-isomerase.
Conclusion
The N-alkyl tetrahydroquinoline compounds 2a–d showed no antibiotic or antimycotic activity against the tested microorganisms. Their corresponding hydrochlorides 3a–d showed weak cytotoxicity. In contrast, the (±)-trans-N-alkylperhydroquinolines showed high antimycotic activity comparable to the commonly used drug clotrimazole. The maximum of activity was found with the C_{10} alkyl chain in the agar diffusion assay and with the C_{12} alkyl chain in the MIC determination, shorter alkyl chains led to a decrease in activity, as already found for other N-alkyl heterocycles [8, 9]. Compared to the recently described [8] N-alkyl perhydroisoquinolines (e.g. C), the new perhydroquinoline compounds showed similar antifungal activity, but higher cytotoxicity against a human cell line. Both perhydroquinolines and perhydroisoquinolines target the same enzyme in ergosterol biosynthesis (Δ8,7-isomerase), but the latter chemotype seems to have benefits in selectivity.

Experimental
General
IR-spectra: Perkin-Elmer FT-IR Paragon 1000; MS: Hewlett Packard MS-Engine; electron ionisation (EI) 70 eV, chemical ionisation (CI) with CH₄ (300 eV); NMR: JNM-Eclipse+400 (400 MHz) (¹H: 400 MHz, ¹³C: 100 MHz), and Avance III HD 400 MHz Bruker BioSpin (400 MHz); melting points: Büchi Melting Point B-540 (not corrected); flash column chromatography (FCC): silica gel 60 (230–400 mesh, E. Merck, Darmstadt); GLC-MS: Shimadzu GC-17 A (carrier: He, oven temperature program: 100–280°C, 10°C/min, capillary column: Varian VF-5 ms 30 m × 0.25, split injector T = 250°C, detector T = 260°C).

General Procedure I (N-alkylation)
The quinoline derivative (about 4 mmol) was dissolved in 50 mL dry THF and 3 equiv. of NaH were added. The suspension was refluxed for 1 h. Then 1.4 to 2 equiv. of the alkyl halide in 5 mL dry THF was added and the mixture was refluxed for 6 h. The mixture was quenched with 50 mL 10% aqueous NaOH and was extracted with ethyl acetate (3 × 50 mL). The combined organic layers were dried over Na₂SO₄, the solvent was evaporated, and the residue was purified by flash column chromatography.

General Procedure II (Preparation of Hydrochlorides)
1.0 mmol of the N-alkyl quinoline derivative was dissolved in 30 mL of dry diethyl ether and the solution was flushed with HCl gas for five minutes. The solvent was evaporated and the residue was disperged in 30 mL of dry diethyl ether, the suspension was placed in a fridge for 3 h and the precipitate was separated to give the analytically pure hydrochlorides.

1-Nonyl-1,2,3,4-tetrahydroquinoline (2a)
The compound was prepared according to General Procedure I from 0.535 g (4.01 mmol) 1,2,3,4-tetrahydroquinoline and 1.56 g (7.51 mmol) 1-bromononane to give 0.81 g (78%) as a pale yellow oil. ¹H-NMR (400 MHz, CDCl₃, TMS): δ 0.88 (t, J = 7.2 Hz, 3 H, CH₃), 1.20–1.37 (m, 12 H, 6 CH₂), 1.57 (m, 2 H, CH₂), 1.93 (tt, J = 5.7 Hz, J = 6.4 Hz, 2 H, CH₂, CH₃, CH₂)}
3-H), 2.74 (t, J = 6.4 Hz, 2 H, CH2, 4-H), 3.21 (t, J = 7.6 Hz, 2 H, CH2), 3.27 (t, J = 5.7 Hz, 2 H, CH2, 2-H). 6.53 (dd, J = 7.3 Hz, J = 7.4 Hz, 1 H, 6-H), 6.55 (d, J = 8.2 Hz, 1 H, 8-H), 6.92 (d, J = 7.3 Hz, 1 H, 5-H), 7.03 (dd, J = 7.4 Hz, J = 8.2 Hz, 1 H, 7-H). 13C-NMR (125 MHz, CDCl3, TMS): δ 14.1 (CH3), 22.2 (CH2), 22.7 (CH2), 26.1 (CH2), 27.3 (CH2), 28.2 (CH2), 29.3 (CH2), 29.6 (CH2), 31.89 (CH2), 49.4 (CH2), 51.5 (CH2), 110.4 (arom. CH), 115.1 (arom. CH), 122.1 (quat. C), 127.0 (arom. CH), 129.1 (arom. CH), 145.3 (quat. C). IR (KBr), ν, cm⁻¹: 3409, 3065, 3017, 2926, 2854, 1655, 1602, 1505, 1457, 1346, 1215, 1195, 1108, 1059, 743. MS (Cl, m/z, %): 260 ([M+1]+, 100), 146 (28). MS (EI, m/z, %): 259 ([M]+, 12), 146 (100). HRMS: Calcd. for C18H29N: 259.2300. Found: 259.2293.

1-Decyl-1,2,3,4-tetrahydroquinoline (2b)
The compound was prepared according to General Procedure I from 0.525 g (3.94 mmol) 1,2,3,4-tetrahydroquinoline and 1.659 g (7.50 mmol) 1-bromodecane to give 0.91 g (84%) of 2b as a pale yellow oil. 1H-NMR (400 MHz, CDCl3, TMS): δ 0.88 (t, J = 7.2 Hz, 3 H, CH3), 1.26–1.31 (m, 14 H, 7 CH2), 1.57 (m, 2 H, CH2), 1.94 (tt, J = 5.7 Hz, J = 6.4 Hz, 2 H, CH2, 3-H), 2.74 (t, J = 6.4 Hz, 2 H, CH2, 4-H), 3.21 (t, J = 7.6 Hz, J = 6.4 Hz, 2 H, CH2, 2-H), 6.52 (dd, J = 7.3 Hz, J = 7.6 Hz, 1 H, 6-H), 6.56 (d, J = 8.1 Hz, 1 H, 8-H), 6.92 (d, J = 7.3 Hz, 1 H, 5-H), 7.02 (dd, J = 7.6 Hz, J = 8.1 Hz, 1 H, 7-H). 13C-NMR (100 MHz, CDCl3, TMS): δ 14.1 (CH3), 22.2 (CH2), 22.7 (CH2), 26.1 (CH2), 27.3 (CH2), 28.2 (CH2), 29.3 (CH2), 29.6 (CH2), 31.9 (CH2), 49.4 (CH2), 51.5 (CH2), 110.4 (arom. CH), 115.1 (arom. CH), 122.1 (quat. C), 127.0 (arom. CH), 129.1 (arom. CH), 145.3 (quat. C). IR (KBr), ν, cm⁻¹: 3408, 3064, 3019, 2925, 2853, 1661, 1602, 1505, 1457, 1346, 1212, 1196, 1108, 1059, 742. MS (Cl, m/z, %): 274 (M+1, 100), 146 (28). MS (EI, m/z, %): 273 (M+, 10), 146 (100). HRMS: Calcd. for C19H31N: 273.2456. Found: 273.2457.

1-Undecyl-1,2,3,4-tetrahydroquinoline (2c)
The compound was prepared according to General Procedure I from 0.504 g (3.78 mmol) 1,2,3,4-tetrahydroquinoline and 1.743 g (7.41 mmol) 1-bromoundecane to give 0.88 g (81%) of 2c as a pale yellow oil. 1H-NMR (400 MHz, CDCl3, TMS): δ 0.88 (t, J = 7.2 Hz, 3 H, CH3), 1.26–1.31 (m, 16 H, 8 CH2), 1.57 (m, 2 H, CH2), 1.93 (tt, J = 5.7 Hz, J = 6.3 Hz, 2 H, CH2, 3-H), 2.74 (t, J = 6.3 Hz, 2 H, CH2, 4-H), 3.21 (t, J = 7.6 Hz, 2 H, CH2), 3.26 (t, J = 5.7 Hz, 2 H, CH2, 2-H), 6.53 (dd, J = 7.1 Hz, J = 7.6 Hz, 1 H, 6-H), 6.56 (d, J = 7.5 Hz, 1 H, 8-H), 6.93 (d, J = 7.1 Hz, 1 H, 5-H), 7.03 (dd, J = 7.3 Hz, J = 7.5 Hz, 1 H, 7-H). 13C-NMR (100 MHz, CDCl3, TMS): δ 14.1 (CH3), 22.2 (CH2), 22.7 (CH2), 26.1 (CH2), 27.3 (CH2), 28.2 (CH2), 29.3 (CH2), 29.6 (CH2), 29.6 (CH2), 29.7 (CH2), 31.9 (CH2), 49.4 (CH2), 51.5 (CH2), 110.4 (arom. CH), 115.1 (arom. CH), 122.1 (quat. C), 127.0 (arom. CH), 129.1 (arom. CH), 145.3 (quat. C). IR (KBr), ν, cm⁻¹: 3408, 3064, 3019, 2925, 2853, 1661, 1602, 1505, 1457, 1346, 1212, 1196, 1108, 1059, 742. MS (Cl, m/z, %): 274 (M+1, 100), 146 (28). MS (EI, m/z, %): 273 (M+, 10), 146 (100). HRMS: Calcd. for C19H31N: 273.2456. Found: 273.2457.

1-Dodecyl-1,2,3,4-tetrahydroquinoline (2d)
The compound was prepared according to General Procedure I from 0.514 g (3.86 mmol) 1,2,3,4-tetrahydroquinoline and 2.218 g (7.49 mmol) 1-iodododecane to give 0.90 g (78%) of 2d as a pale yellow oil.
Alternative approach: 0.594 g (1.89 mmol) N-dodecanoyl-1,2,3,4-tetrahydroquinoline (6) was dissolved in 30 mL of dry THF and 270 mg (3.8 mmol) of LiAlH₄ were added. The suspension was refluxed for 5 h, then quenched with 30 mL aqueous 10% NaOH and extracted with ethyl acetate (3 × 30 mL). The combined organic layers were dried over Na₂SO₄, the solvent was evaporated, and the residue was purified by flash column chromatography to give 0.503 g (88%) of 2d as a pale yellow oil.

δ 0.88 (t, J = 7.2 Hz, 3 H, CH₃), 1.26–1.31 (m, 18 H, 9 CH₂), 1.57 (m, 2 H, CH₂), 1.93 (tt, J = 5.7 Hz, J = 6.4 Hz, 2 H, CH₂, 3-H), 2.74 (t, J = 6.4 Hz, 2 H, CH₂, 4-H), 3.21 (t, J = 7.6 Hz, 2 H, CH₂), 3.26 (t, J = 5.7 Hz, 2 H, CH₂, 2-H), 6.53 (dd, J = 7.8 Hz, J = 7.9 Hz, 1 H, 6-H), 6.55 (d, J = 7.8 Hz, 1 H, 8-H), 6.93 (t, J = 7.9 Hz, 1 H, 5-H), 7.03 (dd, J = 7.8 Hz, J = 7.8 Hz, 1 H, 7-H). ¹³C-NMR (100 MHz, CDCl₃, TMS): δ 14.2 (CH₃), 22.2 (CH₂), 22.7 (CH₂), 26.1 (CH₂), 27.3 (CH₂), 28.2 (CH₂), 29.4 (CH₂), 29.6 (CH₂), 29.6 (CH₂), 29.7 (CH₂), 29.7 (CH₂), 31.9 (CH₂), 49.4 (CH₂), 51.5 (CH₂), 110.4 (arom. CH), 115.1 (arom. CH), 122.1 (quat. arom. C), 127.0 (arom. CH), 129.1 (quat. CH). IR (KBr), ν, cm⁻¹: 3407, 3065, 3020, 2923, 2852, 1660, 1602, 1546, 1456, 1346, 1195, 1059, 742. MS (CI, m/z, %): 302 (M⁺+Cl, 100), 146 (31). MS (EI, m/z, %): 301 (M⁺, 9), 146 (100). HR-MS: Calcd. for C21H35N: 301.2769. Found: 301.2770.

1-Nonyl-1,2,3,4-tetrahydroquinoline hydrochloride (3a)
The compound was prepared according to General Procedure II from 0.700 g (2.70 mmol) 1-nonyl-1,2,3,4-tetrahydroquinoline (2a) to give 0.48 g (60%) of 3a as a yellow-orange solid. M.p.: 65°C (diethyl ether). ¹H-NMR (400 MHz, CDCl₃, TMS): δ 0.87 (t, J = 7.2 Hz, 3 H, CH₃), 1.18–1.39 (m, 12 H, 6 CH₂), 1.91 (m, 2 H, CH₂), 2.97 (t, J = 6.9 Hz, 2 H, CH₂, 4-H), 3.35 (t, J = 8.5 Hz, 2 H, CH₂), 3.59 (t, J = 5.9 Hz, 2 H, CH₂, 2-H), 7.23 (m, 1 H, arom. CH), 7.34 (m, 2 H, 2 arom. CH), 7.69 (m, 1 H, arom. CH), 14.0 (s, 1 H, NH). ¹³C-NMR (100 MHz, CDCl₃, TMS): δ 14.1 (CH₃), 16.3 (CH₂), 22.6 (CH₂), 24.4 (CH₂), 26.7 (CH₂), 29.0 (CH₂), 29.11 (CH₂), 29.35 (CH₂), 31.7 (CH₂), 48.0 (CH₂), 58.7 (CH₂), 124.2 (arom. CH), 127.9 (arom. CH), 129.4 (arom. CH), 130.5 (quat. C), 130.6 (arom. CH), 136.0 (quat. C). IR (KBr), ν, cm⁻¹: 3064, 2930, 2855, 2437, 2254, 1592, 1501, 1472, 1171, 1066, 760. MS (Cl, m/z, %): 260 (M⁺-Cl, 100), 146 (19). MS (EI, m/z, %): 259 (M⁺-HCl, 12), 146 (100). HR-MS: Calcd. for C₁₈H₂₉N: 259.2300. Found: 259.2306.

1-Decyl-1,2,3,4-tetrahydroquinoline hydrochloride (3b)
The compound was prepared according to General Procedure II from 0.790 g (2.89 mmol) 1-decyl-1,2,3,4-tetrahydroquinoline (2b) to give 0.43 g (49%) of 3b as a yellow-orange solid. M.p.: 61°C (diethyl ether). ¹H-NMR (400 MHz, CDCl₃, TMS): δ 0.87 (t, J = 7.2 Hz, 3 H, CH₃), 1.24–1.31 (m, 14 H, 7 CH₂), 1.91 (m, 2 H, CH₂), 2.21 (tt, J = 5.8 Hz, J = 6.9 Hz, 2 H, CH₂, 3-H), 2.96 (t, J = 6.9 Hz, 2 H, CH₂, 2-H), 3.34 (t, J = 8.5 Hz, 2 H, CH₂), 3.58 (t, J = 5.8 Hz, 2 H, CH₂), 7.22 (m, 1 H, arom. CH), 7.34 (m, 2 H, 2 arom. CH), 7.69 (m, 1 H, arom. CH), 14.06 (s, 1 H, NH). ¹³C-NMR (100 MHz, CDCl₃, TMS): δ 14.1 (CH₃), 16.3 (CH₂), 22.7 (CH₂), 24.4 (CH₂), 24.8 (CH₂), 26.8 (CH₂), 29.0 (CH₂), 29.2 (CH₂), 29.4 (2 CH₂), 31.8 (CH₂), 48.0 (CH₂), 58.7 (CH₂), 124.2 (arom. CH), 127.9 (arom. CH), 129.4 (arom. CH), 130.4 (quat. C), 130.6 (arom. CH), 136.0 (quat. C). IR (KBr), ν, cm⁻¹: 3062, 2922, 2853, 2438, 2260, 1592, 1501, 1471, 1170, 1067, 759. MS (Cl, m/z, %): 274 (M⁺-Cl, 100), 146 (28). MS (EI, m/z, %): 273 (M⁺-HCl, 11), 146 (100). HR-MS: Calcd. for C₁₉H₃₁N: 273.2456. Found: 273.2458.
1-Undecyl-1,2,3,4-tetrahydroquinoline hydrochloride (3c)

The compound was prepared according to General Procedure II from 0.763 g (2.66 mmol) 1-undecyl-1,2,3,4-tetrahydroquinoline (2c) to give 0.37 g (43%) of 3c as a yellow-orange solid. M.p.: 52°C (diethyl ether). ¹H-NMR (400 MHz, CDCl₃, TMS): δ 0.88 (t, J = 7.2 Hz, 3 H, CH₃), 1.24–1.31 (m, 16 H, 8 CH₂), 1.85 (m, 2 H, CH₂), 2.22 (m, 2 H, CH₂, 3-H), 2.97 (t, J = 6.9 Hz, 2 H, CH₂, 4-H), 3.35 (t, J = 8.5 Hz, 2 H, CH₂), 3.59 (m, 2 H, CH₂, 2-H), 7.23 (m, 1 H, arom. CH), 7.34 (m, 2 H, 2 arom. CH), 7.69 (m, 1 H, arom. CH), 14.05 (s, 1 H, NH). ¹³C-NMR (100 MHz, CDCl₃, TMS): δ 14.1 (CH₃), 16.3 (CH₂), 22.7 (CH₂), 24.4 (CH₂), 24.8 (CH₂), 26.8 (CH₂), 29.0 (CH₂), 29.3 (CH₂), 29.4 (CH₂), 29.5 (CH₂), 29.5 (CH₂), 31.9 (CH₂), 48.0 (CH₂), 58.7 (CH₂), 124.2 (arom. CH), 127.9 (arom. CH), 129.4 (arom. CH), 130.5 (quat. C), 130.6 (arom. CH), 136.0 (quat. C). IR (KBr), ν, cm⁻¹: 3062, 2921, 2852, 2442, 2261, 1597, 1500, 1470, 1069, 759. MS (CI, m/z, %): 288 (M⁺-Cl, 100), 146 (33). MS (EI, m/z, %): 287 (M⁺-HCl, 10), 146 (100). HR-MS: Calcd. for C₂₀H₃₃N: 287.2613. Found: 287.2650.

1-Dodecyl-1,2,3,4-tetrahydroquinoline hydrochloride (3d)

The compound was prepared according to General Procedure II from 0.788 g (2.62 mmol) 1-dodecyl-1,2,3,4-tetrahydroquinoline (2d) to give 0.47 g (53%) of 3d as a brown solid. M.p.: 49°C (diethyl ether). ¹H-NMR (400 MHz, CDCl₃, TMS): δ 0.88 (t, J = 7.2 Hz, 3 H, CH₃), 1.25–1.31 (m, 18 H, 9 CH₂), 1.77 (m, 2 H, CH₂), 2.12 (m, 2 H, CH₂, 3-H), 2.87 (t, J = 6.3 Hz, 2 H, CH₂, 4-H), 3.29 (t, J = 8.5 Hz, 2 H, CH₂), 3.46 (t, J = 6.1 Hz, 2 H, CH₂, 2-H), 7.23 (m, 1 H, arom. CH), 7.33 (m, 2 H, 2 arom. CH), 7.70 (m, 1 H, arom. CH), 14.14 (s, 1 H, NH). ¹³C-NMR (100 MHz, CDCl₃, TMS): δ 14.1 (CH₃), 16.3 (CH₂), 22.7 (CH₂), 24.4 (CH₂), 24.8 (CH₂), 26.8 (CH₂), 29.1 (CH₂), 29.3 (CH₂), 29.4 (CH₂), 29.5 (CH₂), 29.6 (2 CH₂), 31.9 (CH₂), 48.0 (CH₂), 58.7 (CH₂), 124.2 (arom. CH), 127.9 (arom. CH), 129.4 (arom. CH), 130.5 (quat. C), 130.6 (arom. CH), 136.0 (quat. C). IR (KBr), ν, cm⁻¹: 3419, 3064, 2924, 2853, 2336, 1601, 1504, 1458, 1346, 1195, 1110, 1061, 742. MS (CI, m/z, %): 302 (M⁺-Cl, 100), 146 (33). MS (EI, m/z, %): 301 (M⁺-HCl, 10), 146 (100). HR-MS: Calcd. for C₂₁H₃₅N: 301.2769. Found: 301.2742.

(±)-trans-1-Nonyldecahydroquinoline (4a)

The compound was prepared according to General Procedure I from 0.505 g (3.63 mmol) (±)-trans-decahydroquinoline and 1.025 g (4.95 mmol) 1-bromononane to give 0.62 g (64%) of 4a as a pale yellow oil. ¹H-NMR (400 MHz, CDCl₃, TMS): δ 0.88 (t, J = 7.2 Hz, 3 H, CH₃), 0.94 (m, 1 H, CH₂), 0.98 (m, 1 H, CH₂), 1.09 (m, 1 H, CH₂), 1.19 (m, 1 H, CH₂), 1.21 (m, 1 H, CH), 1.23 (m, 1 H, CH₂), 1.26 (m, 12 H, 6 CH₂), 1.30 (m, 1 H, CH₂), 1.41 (m, 1 H, CH₂), 1.56 (m, 1 H, CH₂), 1.60 (m, 1 H, CH₂), 1.62 (m, 2 H, CH₂), 1.65 (m, 1 H, CH₂), 1.75 (m, 1 H, CH), 1.78 (m, 1 H, CH₂), 2.06 (m, 1 H, CH₂), 2.20 (m, 1 H, CH₂), 2.45 (m, 1 H, CH₂), 2.65 (m, 1 H, CH₂), 2.92 (m, 1 H, CH₂). ¹³C-NMR (100 MHz, CDCl₃, TMS): δ 14.2 (CH₃), 22.8 (CH₂), 24.1 (CH₂), 25.9 (CH₂), 25.9 (CH₂), 26.1 (CH₂), 27.9 (CH₂), 29.4 (CH₂), 29.7 (CH₂), 29.7 (CH₂), 30.2 (CH₂), 32.0 (CH₂), 32.7 (CH₂), 33.3 (CH₂), 42.0 (CH), 53.2 (CH₂), 53.6 (CH₂), 65.8 (CH). IR (KBr), ν, cm⁻¹: 2924, 2854, 1459, 1448, 1378, 1366, 1239, 1095. MS (Cl, m/z, %): 266 (M⁺+1, 100). MS (EI, m/z, %): 265 (M⁺, 6), 264 (3), 222 (49), 152 (100). HR-MS: Calcd. for C₁₈H₃₅N: 265.2769. Found: 265.2748.

(±)-trans-1-Decyldecahydroquinoline (4b)

The compound was prepared according to General Procedure I from 0.512 g (3.68 mmol)
(±)-trans-decahydroquinoline and 1.104 g (4.99 mmol) 1-bromodecane to give 0.46 g (45\%) of 4b as a pale yellow oil. $^1$H-NMR (400 MHz, CDCl$_3$, TMS): δ 0.88 (t, $J = 7.2$ Hz, 3 H, CH$_3$), 0.96 (m, 1 H, CH$_2$), 0.98 (m, 1 H, CH$_2$), 1.09 (m, 1 H, CH$_2$), 1.21 (m, 4 H, 2 CH$_2$), 1.23 (m, 1 H, CH), 1.26 (m, 14 H, 7 CH$_2$), 1.41 (m, 1 H, CH$_2$), 1.56 (m, 1 H, CH$_2$), 1.58 (m, 1 H, CH$_2$), 1.61 (m, 2 H, CH$_2$), 1.73 (m, 1 H, CH$_2$), 1.77 (m, 1 H, CH), 2.06 (m, 1 H, CH$_2$), 2.20 (m, 1H, CH$_2$), 2.45 (m, 1 H, CH$_2$), 2.65 (m, 1 H, CH$_2$), 2.92 (m, 1 H, CH$_2$). $^{13}$C-NMR (100 MHz, CDCl$_3$, TMS): δ 14.2 (CH$_3$), 22.8 (CH$_2$), 24.1 (CH$_2$), 25.8 (CH$_2$), 25.9 (CH$_2$), 26.0 (CH$_2$), 27.9 (CH$_2$), 29.4 (CH$_2$), 29.7 (CH$_2$), 29.7 (CH$_2$), 30.1 (CH$_2$), 32.0 (CH$_2$), 32.7 (CH$_2$), 33.3 (CH$_2$), 41.9 (CH), 53.1 (CH$_2$), 53.5 (CH$_2$), 65.8 (CH). IR (film), ν, cm$^{-1}$: 2924, 2853, 1460, 1447, 1378, 1366, 1239, 1095. MS (Cl, m/z, %): 280 (M$^+$, 100). MS (EI, m/z, %): 279 (M +, 2), 237 (4), 236 (23), 152 (100). HR-MS: Calcd. for C$_{19}$H$_{37}$N: 279.2926. Found: 279.2901.

(±)-trans-1-Undecyldecahydroquinoline (4c)
The compound was prepared according to General Procedure I from 0.512 g (3.68 mmol) (±)-trans-decahydroquinoline and 1.211 g (5.14 mmol) 1-bromoundecane to give 0.44 g (41\%) of 4c as a pale yellow oil. $^1$H-NMR (400 MHz, CDCl$_3$, TMS): δ 0.88 (t, $J = 7.2$ Hz, 3 H, CH$_3$), 0.95 (m, 1 H, CH$_2$), 0.98 (m, 1 H, CH$_2$), 1.09 (m, 1 H, CH$_2$), 1.21 (m, 2 H, CH$_2$), 1.23 (m, 1 H, CH$_2$), 1.26 (m, 18H, 9 CH$_2$), 1.42 (m, 1 H, CH$_2$), 1.58 (m, 2 H, CH$_2$), 1.61 (m, 2 H, CH$_2$), 1.62 (m, 1 H, CH), 1.77 (m, 1 H, CH$_2$), 2.05 (m, 1 H, CH$_2$), 2.20 (m, 1 H, CH$_2$), 2.46 (m, 1 H, CH$_2$), 2.64 (m, 1 H, CH$_2$), 2.89 (m, 1 H, CH$_2$). $^{13}$C-NMR (100 MHz, CDCl$_3$, TMS): δ 14.22 (CH$_3$), 22.8 (CH$_2$), 24.0 (CH$_2$), 25.8 (CH$_2$), 25.9 (CH$_2$), 26.0 (CH$_2$), 27.9 (CH$_2$), 29.4 (CH$_2$), 29.6 (CH$_2$), 29.7 (CH$_2$), 29.7 (CH$_2$), 30.1 (CH$_2$), 32.0 (CH$_2$), 32.7 (CH$_2$), 33.3 (CH$_2$), 41.9 (CH), 53.1 (CH$_2$), 53.5 (CH$_2$), 65.8 (CH). IR (film), ν, cm$^{-1}$: 2923, 2853, 1460, 1448, 1378, 1366, 1239, 1094. MS (Cl, m/z, %): 293 (M$^+$+1, 100). MS (EI, m/z, %): 293 (M$^+$, 2), 250 (23), 152 (100). HR-MS: Calcd. for C$_{20}$H$_{39}$N: 293.3082. Found: 293.3081.

(±)-trans-1-Dodecyldecahydroquinoline (4d)
The compound was prepared according to General Procedure I from 0.535 g (3.84 mmol) (±)-trans-decahydroquinoline and 1.485 g (5.01 mmol) 1-iodododecane and to give 1.08 g (3.53 mmol) (92\%) of 4d as a pale yellow oil. $^1$H-NMR (400 MHz, CDCl$_3$, TMS): δ 0.88 (t, $J = 7.2$ Hz, 3 H, CH$_3$), 0.95 (m, 1 H, CH$_2$), 0.97 (m, 1 H, CH$_2$), 1.06 (m, 1 H, CH$_2$), 1.20 (m, 1 H, CH$_2$), 1.21 (m, 1 H, CH), 1.26 (m, 20 H, 10 CH$_2$), 1.41 (m, 1 H, CH$_2$), 1.56 (m, 1 H, CH$_2$), 1.60 (m, 1 H, CH$_2$), 1.62 (m, 2 H, CH$_2$), 1.73 (m, 1 H, CH), 1.77 (m, 1 H, CH$_2$), 2.05 (m, 1 H, CH$_2$), 2.17 (m, 1 H, CH$_2$), 2.20 (m, 1 H, CH$_2$), 2.44 (m, 1 H, CH$_2$), 2.65 (m, 1 H, CH$_2$), 2.91 (m, 1 H, CH$_2$). $^{13}$C-NMR (100 MHz, CDCl$_3$, TMS): δ 14.22 (CH$_3$), 22.78 (CH$_2$), 24.1 (CH$_2$), 25.9 (CH$_2$), 26.0 (CH$_2$), 27.9 (CH$_2$), 29.4 (CH$_2$), 29.7 (CH$_2$), 29.7 (CH$_2$), 30.1 (CH$_2$), 31.0 (CH$_2$), 32.0 (CH$_2$), 32.7 (CH$_2$), 33.3 (CH$_2$), 42.0 (CH), 53.1 (CH$_2$), 53.6 (CH$_2$), 65.8 (CH). IR (KBr), ν, cm$^{-1}$: 2923, 2853, 1460, 1448, 1378, 1366, 1239, 1094. MS (Cl, m/z, %): 294 (M$^+$+1, 100). MS (EI, m/z, %): 293 (M$^+$, 2), 250 (23), 152 (100). HR-MS: Calcd. for C$_{21}$H$_{41}$N: 293.3082. Found: 293.3081.

(±)-trans-1-Nonyldecahydroquinoline hydrochloride (5a)
The compound was prepared according to General Procedure II from 0.380 g (1.43 mmol) (±)-trans-decahydroquinoline and 1.485 g (5.01 mmol) 1-iodododecane and to give 0.39 g (91\%) of 5a as a white solid. M.p.: 130°C (diethyl ether). $^1$H-NMR (400 MHz, CD$_2$Cl$_2$, TMS): δ 0.88 (t, $J = 7.2$ Hz, 3H,
CH₃), 1.01 (m, 1 H, CH₂), 1.04 (m, 1 H, CH₂), 1.14 (m, 1 H, CH₂), 1.28 (m, 10 H, 5 CH₂), 1.36 (m, 2 H, CH₂), 1.58 (m, 1 H, CH₂), 1.65 (m, 1 H, CH₂), 1.71 (m, 1 H, CH₂), 1.76 (m, 2 H, CH₂), 1.78 (m, 2 H, CH₂), 1.80 (m, 1 H, CH₂), 1.92 (m, 1 H, CH₂), 2.00 (m, 1 H, CH), 2.05 (m, 1 H, CH₂), 2.31 (m, 1 H, CH₃), 2.51 (m, 1 H, CH), 2.78 (m, 1 H, CH₂), 2.99 (m, 2 H, CH₂), 3.38 (m, 1 H, CH₂). ¹³C-NMR: (100 MHz, CD₂Cl₂, TMS): δ 14.3 (CH₃), 21.9 (CH₂), 23.1 (CH₂), 23.1 (CH₂), 25.0 (CH₂), 25.7 (CH₂), 27.3 (CH₂), 27.4 (CH₂), 29.5 (CH₂), 29.6 (CH₂), 29.8 (CH₂), 30.9 (CH₂), 32.2 (CH₂), 33.2 (CH₂), 39.4 (CH), 52.2 (CH₂), 53.3 (CH₂), 66.9 (CH). IR (KBr), ν, cm⁻¹: 2924, 2858, 2516, 2492, 1468, 1452, 1376, 1084, 1047. MS (CI, m/z, %): 266 (M+-Cl, 100), 152 (45). MS (EI): m/z (%) = 265 (M+-HCl, 2), 222 (25), 152 (100). HR-MS: Calcd. for C₁₈H₃₆N: 266.2848. Found: 266.2846.

(±)-trans-1-Decyldecahydroquinoline hydrochloride (5b)
The compound was prepared according to General Procedure II from 0.214 g (0.77 mmol) (±)-trans-1-decyldecahydroquinoline (4b) to give 0.21 g (86%) of 5b as a white solid. M.p.: 128°C (diethyl ether). ¹H-NMR (400 MHz, CDCl₃, TMS): δ 0.88 (t, J = 7.2 Hz, 3 H, CH₃), 1.01 (m, 1 H, CH₂), 1.14 (m, 1 H, CH₂), 1.28 (m, 14 H, 7 CH₂), 1.34 (m, 2 H, CH₂), 1.58 (m, 1 H, CH₂), 1.65 (m, 1 H, CH₂), 1.75 (m, 2 H, CH₂), 1.77 (m, 2 H, CH₂), 1.80 (m, 1 H, CH₂), 1.91 (m, 1 H, CH₂), 2.00 (m, 1 H, CH), 2.05 (m, 1 H, CH₂), 2.30 (m, 1 H, CH₂), 2.50 (m, 1 H, CH), 2.79 (m, 1 H, CH), 3.00 (m, 2 H, CH₂), 3.37 (m, 1 H, CH₂). ¹³C-NMR: (100 MHz, CD₂Cl₂, TMS): δ 14.3 (CH₃), 21.9 (CH₂), 23.1 (CH₂), 23.1 (CH₂), 25.0 (CH₂), 25.7 (CH₂), 27.3 (CH₂), 27.4 (CH₂), 29.5 (CH₂), 29.6 (CH₂), 29.8 (CH₂), 30.9 (CH₂), 32.2 (CH₂), 33.2 (CH₂), 39.4 (CH), 52.2 (CH₂), 53.3 (CH₂), 66.9 (CH). IR (KBr), ν, cm⁻¹: 2924, 2855, 2520, 2493, 1464, 1453, 1377, 1072, 1046. MS (CI, m/z, %): 280 (M+-Cl, 100), 152 (49). MS (EI): m/z (%) = 279 (M+-HCl, 2), 236 (24), 152 (100). HR-MS: Calcd. for C₁₉H₄₀N: 280.3004. Found: 280.3000.

(±)-trans-1-Undecyldecahydroquinoline hydrochloride (5c)
The compound was prepared according to General Procedure II from 0.194 g (0.66 mmol) (±)-trans-1-undecyldecahydroquinoline (4c) to give 0.18 g (83%) of 5c as a white solid. M.p.: 119°C (diethyl ether). ¹H-NMR (400 MHz, CDCl₃, TMS): δ 0.88 (t, J = 7.2 Hz, 3 H, CH₃), 1.02 (m, 1 H, CH₂), 1.14 (m, 1 H, CH₂), 1.27 (m, 16 H, 8 CH₂), 1.33 (m, 2 H, CH₂), 1.56 (m, 1 H, CH₂), 1.65 (m, 1 H, CH₂), 1.76 (m, 2 H, CH₂), 1.78 (m, 2 H, CH₂), 1.80 (m, 1 H, CH₂), 1.91 (m, 1 H, CH₂), 2.00 (m, 1 H, CH), 2.04 (m, 1 H, CH₂), 2.31 (m, 1 H, CH₂), 2.50 (m, 1 H, CH), 2.78 (m, 1 H, CH), 3.01 (m, 2 H, CH₂), 3.36 (m, 1 H, CH₂). ¹³C-NMR: (100 MHz, CD₂Cl₂, TMS): δ 14.3 (CH₃), 21.9 (CH₂), 23.1 (CH₂), 23.1 (CH₂), 25.0 (CH₂), 25.7 (CH₂), 27.3 (CH₂), 27.4 (CH₂), 29.5 (CH₂), 29.7 (CH₂), 29.8 (CH₂), 29.9 (CH₂), 30.8 (CH₂), 33.2 (CH₂), 39.4 (CH), 52.2 (CH₂), 53.3 (CH₂), 66.9 (CH). IR (KBr), ν, cm⁻¹: 2924, 2855, 2520, 2493, 1464, 1453, 1377, 1072, 1046. MS (CI, m/z, %): 294 (M+-Cl, 100), 152 (49). MS (EI): m/z (%) = 293 (M+-HCl, 2), 236 (24), 152 (100). HR-MS: Calcd. for C₂₀H₄₀N: 294.3161. Found: 294.3164.

(±)-trans-1-Dodecyldecahydroquinoline hydrochloride (5d)
The compound was prepared according to General Procedure II from 0.796 g (2.59 mmol) (±)-trans-1-dodecyldecahydroquinoline (4d) to give 0.76 g (85%) of 5d as a white solid. M.p.: 113°C (diethyl ether). ¹H-NMR (400 MHz, CDCl₃, TMS): δ 0.88 (t, J = 7.2 Hz, 3 H, CH₃), 1.01 (m, 1 H, CH₂), 1.13 (m, 1 H, CH₂), 1.27 (m, 18 H, 9 CH₂), 1.32 (m, 2 H, CH₂), 1.56 (m, 1 H, CH₂), 1.65 (m, 1 H, CH₂), 1.76 (m, 2 H, CH₂), 1.79 (m, 2 H, CH₂), 1.80 (m, 1 H, CH₂), 1.91 (m, 1 H, CH₂), 2.00 (m, 1 H, CH), 2.04 (m, 1 H, CH₂), 2.31 (m, 1 H, CH₂), 2.50 (m, 1 H, CH), 2.78 (m, 1 H, CH), 3.01 (m, 2 H, CH₂), 3.36 (m, 1 H, CH₂). ¹³C-NMR: (100 MHz, CD₂Cl₂, TMS): δ 14.3 (CH₃), 21.9 (CH₂), 23.1 (CH₂), 23.1 (CH₂), 25.0 (CH₂), 25.7 (CH₂), 27.3 (CH₂), 27.4 (CH₂), 29.5 (CH₂), 29.7 (CH₂), 29.8 (CH₂), 29.9 (CH₂), 30.0 (CH₂), 30.8 (CH₂), 32.3 (CH₂), 33.17 (CH₂), 39.4 (CH), 52.2 (CH₂), 53.3 (CH₂), 66.9 (CH). IR (KBr), ν, cm⁻¹: 2923, 2855, 2519, 2493, 1469, 1454, 1376, 1074, 1046. MS (CI, m/z, %): 294 (M+-Cl, 100), 152 (57). MS (EI): m/z (%) = 293 (M+-HCl, 2), 250 (16), 152 (100). HR-MS: Calcd. for C₂₀H₄₀N: 294.3161. Found: 294.3164.
N-Dodecanoyl-1,2,3,4-tetrahydroquinoline (6)

For this compound's preparation, 0.514 g (3.86 mmol) 1,2,3,4-tetrahydroquinoline and 1.232 g dodecanoyl chloride (5.63 mmol) were dissolved in 30 mL of dry 1,2-dimethoxyethane. Five mL of dimethyl ethyl amine were added and the mixture was stirred for 10 h. The solvent was evaporated, the residue was dissolved in 30 mL aqueous 10% NaOH, and extracted with ethyl acetate (3 × 30 mL). The combined organic layers were dried over Na₂SO₄, the solvent was evaporated, and the residue was purified by flash column chromatography to give 0.88 g (72%) of 6 as a pale yellow solid. M.p.: 37°C (isohexane/ethyl acetate). ¹H-NMR (400 MHz, CDCl₃, TMS): δ 0.88 (t, J = 7.2 Hz, 3 H, CH₃), 1.23 (m, 16 H, 8 CH₂), 1.65 (m, 2 H, CH₂), 1.96 (tt, J = 6.6 Hz, J = 6.6 Hz, 2 H, CH₂, 3-H), 2.49 (t, J = 7.6 Hz, 2 H, CH₂), 2.71 (t, J = 6.6 Hz, 2 H, CH₂, 4-H), 3.79 (t, J = 6.6 Hz, 2 H, CH₂, 2-H), 7.15 (m, 4 H, 4 arom. CH). ¹³C-NMR (100 MHz, CDCl₃, TMS): δ 14.1 (CH₃), 22.7 (CH₂), 24.2 (CH₂), 25.9 (CH₂), 26.8 (CH₂), 29.3 (CH₂), 29.3 (CH₂), 29.4 (CH₂), 29.5 (CH₂), 29.6 (CH₂), 29.6 (CH₂), 31.9 (CH₂), 34.5 (CH₂), 42.8 (CH₂), 124.7 (quat. C), 125.1 (arom. CH), 126.0 (arom. CH), 128.4 (2 arom. CH), 139.3 (quat. C), 173.2 (quat. C). IR (KBr), v, cm⁻¹: 3273, 3056, 2921, 2849, 1655, 1605, 1579, 1493, 1463, 1388, 1244, 1169, 1111, 1069, 756. MS (Cl): m/z (%): 316 (M⁺+1, 100), 133 (47). MS (EI, m/z, %): 315 (M⁺, 4), 133 (100). HR-MS: Calcd. for C₂₁H₃₃NO: 315.2562. Found: 315.2561.

Authors’ Statement

The authors declare no conflict of interest.

References

[1] Vanden Bossche H, Engelen M, Rochette F. Antifungal agents of use in animal health – chemical, biochemical and pharmacological aspects. J Vet Pharmacol Therap. 2003; 26: 5–29.
http://dx.doi.org/10.1046/j.1365-2885.2003.00456.x

[2] van Minnebruggen G, Francois IEJA, Cammune BPA, Thevissen K, Vroome V, Borgers M, Shroot B. A General Overview on Past, Present and Future Antimycotics. Open Mycol J. 2010; 4: 22–32.
http://dx.doi.org/10.2174/1874437001004010022

[3] Benveniste P. Sterol biosynthesis. Ann Rev Plant Physiol. 1986; 37: 275–308.
http://dx.doi.org/10.1146/annurev.pp.37.060186.001423
[4] Polak AM. 
Preclinical data and mode of action of amorolfine. 
Clin Exp Dermatol. 1992; 17 (Suppl. 1): 8–12. 
http://dx.doi.org/10.1111/j.1365-2230.1992.tb00270.x

[5] Baloch RI, Mercer EI. 
Inhibition of sterol Δ8-Δ7 isomerase and Δ14 reductase by fenpropimorph, tridemorph and fenpropidin in cell free enzyme systems from Saccharomyces cerevisiae. 
Phytochemistry. 1987; 26: 663–668. 
http://dx.doi.org/10.1016/S0031-9422(00)84762-7

[6] Baloch RI, Mercer EI, Wiggins TE, Baldwin BC. 
Inhibition of ergosterol biosynthesis in Saccharomyces cerevisiae and Ustilago maydis by tridemorph, fenpropimorph and fenpropidin. 
Phytochemistry. 1984; 23: 2219–2226. 
http://dx.doi.org/10.1016/S0031-9422(00)80523-3

[7] Taton M, Benveniste P, Rahier A. 
Use of rationally designed inhibitors to study sterol and triterpenoid biosynthesis. 
Pure Appl Chem. 1987; 59: 287–294. 
http://dx.doi.org/10.1351/pac198759030287

[8] Krauss J, Müller C, Kießling J, Richter S, Staudacher V, Bracher F. 
Synthesis and biological evaluation of novel N-alkyl tetra- and decahydroisoquinolines: Novel antifungals that target ergosterol biosynthesis. 
Arch Pharm. 2014; 347: 283–290. 
http://dx.doi.org/10.1002/ardp.201300338

[9] Krauss J, Gratzl C, Sturm V, Müller C, Staudacher V, Schmidt CQ, Bracher F. 
Synthesis and Biological Evaluation of Novel Alkyl-Imidazolyl Carbinols and their Esters: Potent Antimycotics. 
Sci Pharm. 2013; 81: 641–650. 
http://dx.doi.org/10.3797/scipharm.1304-17

[10] Booth H, Griffiths V. 
Carbon-13 Magnetic Resonance Studies of Cyclic Compounds. Part I. Piperidines and Decahydroquinolines. 
J Chem Soc Perkin Trans 2. 1973; 842–844. 
http://dx.doi.org/10.1039/P29730000842

[11] DIN Deutsches Institut für Normung e.V. 
Methoden zur Empfindlichkeitsprüfung von mikrobiellen Krankheitserregern gegen Chemotherapeutika. 
DIN 58940, Teil 3 - Beiblatt 1 und Teil 4 - Beiblatt 1, Beuth Verlag, Berlin 1998.

[12] Müller C, Staudacher V, Krauss J, Giera M, Bracher F. 
A convenient cellular assay for the identification of the molecular target of ergosterol biosynthesis inhibitors and quantification of their effects on total ergosterol biosynthesis. 
Steroids. 2013; 78: 483–493. 
http://dx.doi.org/10.1016/j.steroids.2013.006

[13] Mosmann T. 
Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. 
J Immunol Methods. 1983; 65: 55–63. 
http://dx.doi.org/10.1016/0022-1759(83)90303-4

[14] Mengliev AS, Ganiev SS, Yunusov TK, Auelbekov SA, Kuchkarov S. 
Synthesis of N-alkyl and N-acyl derivatives of anabasine, salsoline, piperidine, and morpholine. 
Uzbekskii Khimicheskii Zhurnal. 1995; 1: 35–37.
[15] Baltzly R.  
Unsymmetrically N-substituted piperazines. IV. N-Alkyl derivatives.  
J Am Chem Soc. 1954; 76: 1164–1165.  
http://dx.doi.org/10.1021/ja01633a069

[16] Devinsky F, Lacko I, Mlynarcik D, Krasnec L.  
Amine oxides. VII. Synthesis, IR spectra, and antimicrobial activity of 1,4-dialkylpiperazine dioxides.  
Collect Czech Chem Commun. 1982; 47: 1130–1138.  
http://dx.doi.org/10.1135/cccc19821130

[17] Tobulic S, Kratzer C, Graninger W, Georgopoulos A.  
In vitro-Wirksamkeit von Akazid plus gegenüber Spross- und filamentösen Pilzen im Vergleich zu konventionellen Antimykotika und Chlorhexidin.  
Antibiotika Monitor. 2006; 13–20.