Fragmentation modes and the evolution of life cycles

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Abstract

Reproduction is a defining feature of living systems. To reproduce, aggregates of biological units (e.g., multicellular organisms or colonial bacteria) must fragment into smaller parts. Fragmentation modes in nature range from binary fission in bacteria to collective-level fragmentation and the production of unicellular propagules in multicellular organisms. Despite this apparent ubiquity, the adaptive significance of fragmentation modes has received little attention. Here, we develop a model in which groups arise from the division of single cells that do not separate but stay together until the moment of group fragmentation. We allow for all possible fragmentation patterns and calculate the population growth rate of each associated life cycle. Fragmentation modes that maximise growth rate comprise a restrictive set of patterns that include production of unicellular propagules and division into two similar size groups. Life cycles marked by single-cell bottlenecks maximise population growth rate under a wide range of conditions. This surprising result offers a new evolutionary explanation for the widespread occurrence of this mode of reproduction. All in all, our model provides a framework for exploring the adaptive significance of fragmentation modes and their associated life cycles.

Author summary

Mode of reproduction is a defining trait of all organisms, including colonial bacteria and multicellular organisms. To produce offspring, aggregates must fragment by splitting into two or more groups. The particular way that a given group fragments defines the life cycle of the organism. For instance, insect colonies can reproduce by splitting or by producing individuals that found new colonies. Similarly, some colonial bacteria propagate by fission or by releasing single cells, while others split in highly sophisticated ways; in multicellular organisms reproduction typically proceeds via a single-cell bottleneck phase. The space of possibilities for fragmentation is so vast that an exhaustive analysis seems daunting. Focusing on fragmentation modes of a simple kind we parametrise all possible modes of group fragmentation and identify those modes leading to the fastest population growth.
rate. Two kinds of life cycle dominate: one involving division into two equal size groups, and the other involving production of a unicellular propagule. The prevalence of these life cycles in nature is consistent with our null model and suggests that benefits accruing from population growth rate alone may have shaped the evolution of fragmentation mode.

Introduction

A requirement for evolution—and a defining feature of life—is reproduction [1–4]. Perhaps the simplest mode of reproduction is binary fission in unicellular bacteria, whereby a single cell divides and produces two offspring cells. In more complex organisms, such as colonial bacteria, reproduction involves fragmentation of a group of cells into smaller groups. Bacterial species demonstrate a wide range of fragmentation modes, differing both in the size at which the parental group fragments and the number and sizes of offspring groups [5]. For example, in the bacterium Neisseria, a diplococcus, two daughter cells remain attached forming a two-celled group that separates into two groups of two cells only after a further round of cell division [6]. Staphylococcus aureus, another coccoid bacterium, divides in three planes at right angles to one another to produce grape-like clusters of about 20 cells from which single cells separate to form new clusters [7]. Magnetotactic prokaryotes form spherical clusters of about 20 cells, which divide by splitting into two equally sized clusters [8].

These are just a few examples of a large number of diverse fragmentation modes, but why should there be such a wide range of life cycles? Do fragmentation modes have adaptive significance or are they simply the unintended consequences of particular cellular processes underpinning cell division? If adaptive, what selective forces shape their evolution? Can different life cycles simply provide different opportunities to maximise population growth rate?

A starting point to answer these questions is to consider benefits and costs of group living in cell collectives. Benefits may arise for various reasons. Cells within groups may be better able to withstand environmental stress [9], escape predation [10, 11], or occupy new niches [12, 13]. Also, via density-dependent gene regulation, cells within groups may gain more of a limiting resource than they would if alone [14, 15]. On the other hand, cells within groups experience increased competition and must also contend with the build up of potentially toxic waste metabolites [16, 17]. Thus, it is reasonable to expect an optimal relationship between group size and fragmentation mode that is environment and organism dependent [18–21].

Here we formulate and study a matrix population model [22] that considers all possible modes of group fragmentation. By determining the relationship between life cycle and population growth rate, we show that there is, overall, a narrow class of optimal modes of fragmentation. When the process of fragmentation does not involve costs, optimal fragmentation modes are characterised by a deterministic schedule and binary splitting, whereby groups fragment into exactly two offspring groups. Contrastingly, when a cost is associated with fragmentation, it can be optimal for a group to fragment into multiple propagules.

Our results show that the range of life cycles observed in simple microbial populations are likely shaped by selection for intrinsic growth rate advantages inherent to different modes of group fragmentation. While we do not consider complex life cycles, our results may contribute to understanding the emergence of life cycles underpinning the evolution of multicellular life.
Methods

Group formation and fragmentation

We consider a population in which a single type of cell (or unit or individual) can form groups (or complexes or aggregates) of increasing size by cells staying together after reproduction [18]. We assume that the size of any group is smaller than $n$, and denote groups of size $i$ by $X_i$ (see the list of used variables in Table 1). Groups die at rate $d_i$ and cells within groups divide at rate $b_i$; hence groups grow at rate $ib_i$. The vectors of birth rates $b = (b_1, \ldots, b_{n-1})$ and of death rates $d = (d_1, \ldots, b_{n-1})$ make the costs and benefits associated to the size of the groups explicit, thus defining the “fitness landscape” of our model.

Groups produce new complexes by fragmenting (or splitting), i.e., by dividing into smaller groups. We further assume that fragmentation is triggered by growth of individual cells within a given group. Consider a group of size $i$ growing into a group of size $i + 1$. Such a group can either stay together or fragment. If it fragments, it can do so in one of several ways. For example, a group of size 4 can give rise to the following five “fragmentation patterns”: 4 (the group does not split, but stays together), 3+1 (the group splits into two offspring groups: one of size 3, and one of size 1), 2+2 (the group splits into two groups of size 2), 2+1+1 (the group splits into one group of size 2 and two groups of size 1), and 1+1+1+1 (the group splits into four independent cells). Mathematically, such fragmentation patterns correspond to the five partitions of 4 (a partition of a positive integer $i$ is a way of writing $i$ as a sum of positive integers without regard to order; the summands are called parts [23]). We use the notation $\kappa \vdash \ell$ to indicate that $\kappa$ is a partition of $\ell$, e.g., there are $\zeta_4 = 5$ partitions of 4.

We consider an exhaustive set of fragmentation modes (or “fragmentation strategies”) implementing all possible ways groups of maximum size $n$ can grow and fragment into smaller groups, including both pure and mixed modes (Fig 1). A pure fragmentation mode is characterised by a single partition $\kappa \vdash \ell$, i.e., groups of size $i < \ell$ grow up to size $\ell$ and then fragment according to partition $\kappa \vdash \ell$. The partition $\kappa$ can then be used to refer to the associated pure strategy. The total number of pure fragmentation strategies is $\sum_{i=2}^{n} (\zeta_i - 1)$, which grows quickly with $n$: There are 128 pure fragmentation modes for $n = 10$, but 1,295,920 for $n = 50$. A mixed fragmentation mode is given by a probability distribution over the set of pure fragmentation modes. The relationship between pure and mixed fragmentation modes is hence similar to the one between pure strategies and mixed strategies in evolutionary game theory [24]. One of our main results is that mixed fragmentation modes are always dominated by pure

Table 1. List of variables.

| Symbol | Description |
|--------|-------------|
| $X_i$  | a group of size $i$ cells |
| $x_i$  | abundance of groups of size $i$ |
| $x$    | vector of abundances $x_i$ |
| $b_i$  | birth rate of cells in a group of $i$ cells |
| $d_i$  | death rates of groups of $i$ cells |
| $n$    | maximal group size |
| $\zeta_i$ | the number of partitions of integer $i$ |
| $\pi_i(\kappa)$ | the number of parts equal to $i$ in the partition $\kappa$ |
| $\mathbf{A}$ | projection matrix |
| $\lambda_1$ | population growth rate |
| $M$ | maximal benefit under monotonic fitness landscapes |
| $\alpha$ | degree of complementarity under monotonic fitness landscapes |

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Biological reactions and population dynamics

Together with the fitness landscape given by the vectors of birth rates \( \mathbf{b} \) and death rates \( \mathbf{d} \), each fragmentation strategy specifies a set of biological reactions. Consider the pure mode \( \kappa \rightarrow \ell \), whereby groups grow up to size \( \ell \) and then split according to fragmentation pattern \( \kappa \). A set of reactions

\[
X_i \xrightarrow{d_i} 0, \quad i = 1, \ldots, \ell - 1
\]

models the death of groups; an additional set of reactions

\[
X_i \xrightarrow{b_i} X_{i+1}, \quad i = 1, \ldots, \ell - 2
\]
models the growth of groups (without splitting) up to size $\ell - 1$. Finally, one reaction of the type

$$X_{i-1} \overset{(\ell-1)b_{i-1}}{\rightarrow} \sum_{i=1}^{\ell-1} \pi_i(\kappa) X_i,$$

(3)

models the growth of the group from size $\ell - 1$ to size $\ell$ and its immediate fragmentation in a way described by fragmentation pattern $\kappa \vdash \ell$, where parts equal to $i$ appear a number $\pi_i(\kappa)$ of times. For instance, for the pure fragmentation mode $2 + 1 + 1 \vdash 4$, Eq (3) becomes

$$X_3 \overset{3b_3}{\rightarrow} X_2 + 2X_1,$$

which stipulates that groups of size 3 grow to size 4 at rate $3b_3$ and split into one group of size 2 and two groups of size 1; here, $\pi_1(2 + 1 + 1) = 2$, $\pi_2(2 + 1 + 1) = 1$, $\pi_3(2 + 1 + 1) = 0$.

The sets of reactions (1), (2) and (3) give rise to the system of differential equations

$$\dot{x}_1 = -(b_1 + d_1)x_1 + (\ell - 1)b_{\ell-1} \pi_1(\kappa)x_{\ell-1},$$

$$\dot{x}_i = (i - 1)b_{i-1}x_{i-1} - (ib_1 + d_1)x_1 + (\ell - 1)b_{\ell-1} \pi_1(\kappa)x_{\ell-1}, \quad i = 2, \ldots, \ell - 1$$

where $x_i$ denotes the abundance of groups of size $i$. This is a linear system that can be represented in matrix form as

$$\dot{x} = Ax,$$

(4)

where $x = (x_1, x_2, \ldots, x_{\ell-1})$ is the vector of abundances of the groups of different size and

$$A = \begin{pmatrix}
-b_1 - d_1 & 0 & \cdots & 0 & (\ell - 1)b_{\ell-1} \pi_1(\kappa) \\
b_1 & -2b_2 - d_2 & 0 & \vdots & (\ell - 1)b_{\ell-1} \pi_2(\kappa) \\
0 & 2b_2 & -3b_3 - d_3 & 0 & (\ell - 1)b_{\ell-1} \pi_3(\kappa) \\
\vdots & \vdots & \ddots & \ddots & \ddots \\
0 & 0 & \cdots & (\ell - 2)b_{\ell-2} & (\ell - 1)b_{\ell-1}(\pi_{\ell-1}(\kappa) - 1) - d_{\ell-1}
\end{pmatrix}$$

is the projection matrix determining the population dynamics.

**Population growth rate**

For any fragmentation mode and any fitness landscape, the projection matrix $A$ is “essentially non-negative” (or quasi-positive), i.e., all the elements outside the main diagonal are non-negative [25]. This implies that $A$ has a real leading eigenvalue $\lambda_1$ with associated non-negative left and right eigenvectors $v$ and $w$. In the long term, the solution of Eq (4) converges to that of an exponentially growing population with a stable distribution, i.e.,

$$\lim_{t \to \infty} x(t) = e^{\lambda_1}w.$$

The leading eigenvalue $\lambda_1$ hence gives the total population growth rate in the long term, and its associated right eigenvector $w = (w_1, \ldots, w_m)$ gives the stable distribution of group sizes so that, in the long term, the fraction $x_i$ of complexes of size $i$ in the population is proportional to $w_i$. 
Dominance and optimality

For a given fitness landscape \((\mathbf{b}, \mathbf{d})\), we can take the leading eigenvalue \(\lambda_1(\mathbf{k}; \mathbf{b}, \mathbf{d})\) as a measure of fitness of fragmentation mode \(\mathbf{k}\), and consider the competition between two different fragmentation modes, \(\mathbf{k}_1\) and \(\mathbf{k}_2\). Indeed, under the assumption of no density limitation, the evolutionary dynamics are described by two uncoupled sets of differential equations of the form (4): one set for \(\mathbf{k}_1\) and one set for \(\mathbf{k}_2\). In the long term, \(\mathbf{k}_1\) is not outcompeted by \(\mathbf{k}_2\) if \(\lambda_1(\mathbf{k}_1; \mathbf{b}, \mathbf{d}) \geq \lambda_1(\mathbf{k}_2; \mathbf{b}, \mathbf{d})\); we then say that fragmentation mode \(\mathbf{k}_1\) dominates fragmentation mode \(\mathbf{k}_2\). We also say that strategy \(\mathbf{k}_1\) is optimal for given birth rates \(\mathbf{b}\) and death rates \(\mathbf{d}\) if it achieves the largest growth rate among all possible fragmentation modes.

Two classes of fitness landscape: Fecundity landscapes and survival landscapes

Fitness landscapes capture the many advantages or disadvantages associated with group living. These advantages may come either in the form of additional resources available to groups depending on their size or as an improved protection from external hazards. For our numerical examples, we consider two classes of fitness landscape, each representing only one of these factors. In the first class, that we call “fecundity landscapes”, group size affects only the birth rates of cells (while we impose factors. In the particular case of a fecundity landscape given by

\[
\mathbf{d} = (d_1, d_2) = (0, 0),
\]

these growth rates reduce to \(\lambda_1^{1+1} = 1\), \(\lambda_2^{2+1} = 1\), \(\lambda_3^{1+1+1} = 3/2\) and \(\lambda_4^{1+1+1+1} = 5/4\), and we have \(\lambda_3^{2+1} > \lambda_1^{1+1+1} > \lambda_2^{1+1}\). We then say that ternary and binary fission are dominated by the unicellular propagule strategy.

Results

Mixed fragmentation modes are dominated

Although for simplicity we focus our exposition on pure fragmentation strategies, we also consider mixed fragmentation strategies, i.e., probabilistic strategies mixing between different
pure modes. A natural question to ask is whether a mixed fragmentation mode can achieve a faster growth rate than a pure mode. We find that the answer is no. For any fitness landscape and any maximum group size \( n \), mixed fragmentation modes are dominated by a pure fragmentation mode (S1 Text, Appendix B). Thus, the optimal fragmentation mode for any fitness landscape is pure.

As an example, consider fragmentation modes 1+1 and 2+1, and a mixed fragmentation mode mixing between these two so that with probability \( q \) splitting follows fragmentation pattern 2+1 and with probability \( 1-q \) it follows fragmentation pattern 1+1. For any mixing probability \( q \) and any fitness landscape, the growth rate of the mixed fragmentation mode is given by

\[
\lambda_i^q = \frac{b_1(1-2q) - (d_1 + d_2) + \sqrt{(d_1 + d_2 - (1-2q)b_1)^2 + 4b_1(2qb_2 + (1-2q)d_2)}}{2},
\]

which can be shown to always lie between the growth rates of the pure fragmentation modes, i.e., either \( \lambda_i^{1+1} \leq \lambda_i^q \leq \lambda_i^{2+1} \) or \( \lambda_i^{2+1} \leq \lambda_i^q \leq \lambda_i^{1+1} \) holds and the mixed fragmentation mode is dominated (S1 Text, Appendix C).

To further illustrate our analytical findings, consider groups of maximum size \( n = 4 \) and a fecundity landscape given by \( b = (1, 2, 1.4) \). We randomly generated \( 10^7 \) mixed fragmentation modes by drawing the probabilities for growth without splitting from an uniform distribution and letting the probabilities of splitting according to a given fragmentation pattern be proportional to exponential random variables with rate parameter equal to one. We then calculated the growth rate of these mixed strategies together with the growth rate of the seven pure fragmentation modes available for \( n = 4 \), i.e., 1+1, 2+1, 1+1+1, 3+1, 2+2, 2+1+1, and 1+1+1+1 (Fig 2A). In line with our analysis, a pure fragmentation mode (namely 2+2, whereby groups grow up to size 4 and then immediately split into two bicellular groups) achieves a higher growth rate than the growth rate of any mixed fragmentation mode, and the highest growth rate overall.

**Optimal fragmentation modes are characterised by binary splitting**

Having shown that mixed fragmentation modes are dominated, we now ask which pure modes might be optimal. We find that, within the set of pure modes, “binary” fragmentation modes (whereby groups split into exactly two offspring groups) dominate “nonbinary” fragmentation modes (whereby groups split into more than two offspring groups). To illustrate this result, consider the simplest case of \( n = 3 \) and the three modes 1+1, 2+1, and 1+1+1, out of which 1+1 and 2+1 are binary, and 1+1+1 is nonbinary. Comparing their growth rates (as given in Eq (5), we find that \( \lambda_i^{1+1} \geq \lambda_i^{1+1+1} \) holds if \( b_1 - b_2 \geq d_1 - d_2 \) and that \( \lambda_i^{2+1} \geq \lambda_i^{1+1+1} \) holds if \( b_1 - b_2 \leq d_1 - d_2 \). Thus, for any fitness landscape, 1+1+1 is dominated by either 2+1 or by 1+1. More generally, we can show that for any nonbinary fragmentation mode, one can always find a binary fragmentation mode achieving a greater or equal growth rate under any maximum group size \( n \) and fitness landscape (S1 Text, Appendices D and E). Taken together, our analytical results imply that the set of optimal fragmentation modes is countable and, even for large \( n \), relatively small. Consider the proportion of pure fragmentation modes that can be optimal, which is defined by the ratio between the number of binary fragmentation modes and the total number of pure fragmentation modes. While this ratio is relatively high for small \( n \) (e.g., 2/3 \( \approx \) 0.67 for \( n = 3 \) or 4/7 \( \approx \) 0.57 for \( n = 4 \)), it decreases sharply with increasing \( n \) (e.g., 25/128 \( \approx \) 0.20 for \( n = 10 \) and 625/1295920 \( \approx \) 0.00048 for \( n = 50 \)).
Fig 2B shows the growth rate of the seven pure modes for $n = 4$ for a fecundity landscape given by $b = (1, b_2, 1.4)$ as a function of $b_2$. In line with our analysis, only binary fragmentation modes (1+1, 2+1, 2+2, and 3+1) can be optimal depending on the value of $b_2$. Contrarily, nonbinary fragmentation modes (1+1+1, 1+1+1+1, and 2+1+1) are never optimal.
patterns dictating which fragmentation mode will be optimal. Consider first the optimality map for fecundity landscapes (Fig 3A). A sufficient condition for the unicellular life cycle 1+1 to be optimal is that the birth rate of single cells is larger than the birth rate of pairs and triplets of cells ($b_1 > b_2$ and $b_1 > b_3$). In this case, there is no apparent reason why a fragmentation mode different than 1+1 would be optimal. Perhaps less trivially, 1+1 can also be optimal in cases where single cells are less fertile than groups of three cells, i.e., even if $b_1 < b_3$ holds. This requires the birth rate $b_2$ to be so small that the fecundity benefits accrued when reaching the size of three cells are not enough to compensate for the unavoidable penalty of passing through the less prolific state of two cells. Turning now to fragmentation mode 2+1, a necessary condition for this mode to be optimal is that pairs of cells have the largest birth rate, i.e., that $b_2 > b_1$ and $b_2 > b_3$ holds. Similarly, mode 3+1 can only be optimal if $b_3 > b_1$ and $b_3 > b_2$, so that groups of three have the largest birth rate. In these two cases, the optimal fragmentation mode (either 2+1 or 3+1) keeps one of the two offspring groups at the most productive size. Finally, for fragmentation mode 2+2 to be optimal, it is necessary that single cells have the lowest birth rate, i.e., that $b_2 > b_1$ and $b_3 > b_1$ holds. In this case, the fragmentation mode ensures that the life cycle of the organism never goes through the least productive unicellular phase. Under survival landscapes, fitness increases as death rates decrease. Taking this qualitative difference into account, the map of optimal fragmentation modes under survival landscapes (Fig 3B) follows similar qualitative patterns as the one under fecundity landscapes.

**Costly fragmentation allows for optimal nonbinary fragmentation and multicellularity without group benefits**

So far we have assumed that fragmentation is costless. However, fragmentation processes can be costly to the parental group undergoing division. This is particularly apparent in cases...
where some cells need to die in order for fragmentation of the group to take place. Examples in simple multicellular forms include *Volvox*, where somatic cells constituting the outer layer of the group die upon releasing the offspring colonies and are not passed to the next generation [26], the breaking of filaments in colonial cyanobacteria [27], and the fragmentation of "snowflake-like" clusters of the yeast *Saccharomyces cerevisiae* [28]. Fragmentation costs may also be less apparent. For instance, fragmentation may cost resources that would otherwise be available for the growth of cells within a group.

To investigate the effect of fragmentation costs on the set of optimal fragmentation modes, we consider two cases: proportional costs and fixed costs. For proportional costs, we assume that \( \pi - 1 \) cells die in the process of a group fragmenting into \( \pi \) parts. This case captures the fragmentation process of filamentous bacteria, where filament breakage entails the death of cells connecting the newly formed fragments [27]. For fixed costs, we assume that exactly one cell is lost upon each fragmentation event. This scenario is loosely inspired by yeast colonies with a tree-like structure, where cells can be connected with many other cells, so the death of a single cell may release more than two offspring colonies [19, 28]. Mathematically, both cases imply that fragmentation patterns are described by partitions of a number smaller than the size of the parent group (S1 Text, Appendix G).

For both kinds of costly fragmentation, we can show that mixed fragmentation modes are still dominated by pure fragmentation modes (the proof given in S1 Text, Appendix B also holds in this case). Moreover, for proportional costs the optimal fragmentation mode is also characterised by binary fragmentation, as it is the case for costless fragmentation (S1 Text, Appendix H). This makes intuitive sense, as the addition of a penalty for splitting into many fragments should further reinforce the optimality of binary splitting (whereby only one cell per fragmentation event is lost). In contrast, we find that under fragmentation with fixed costs the optimal fragmentation mode can involve nonbinary fragmentation, i.e., division into more than two offspring groups. This result can be readily illustrated for the case of \( n = 4 \) where the nonbinary mode 1+1+1 is optimal for a wide range of fitness landscapes (Fig 4).

Another interesting feature of costly fragmentation (implemented via either proportional or fixed costs) is that fragmentation modes involving the emergence of large groups can be optimal even if being in a group does not grant any fecundity or survival advantage to cells. If fragmentation is costless, as we assumed before, fitness landscapes for which groups perform worse than unicells (that is, \( b_i/b_1 \leq 1 \) for fecundity landscapes or \( d_i - d_1 \geq 0 \) for survival landscapes) lead to optimal fragmentation modes where splitting occurs at the minimum possible group size \( i = 2 \), so that no multicellular groups emerge in the population (cf. Fig 3). In contrast, under costly fragmentation some of these fitness landscapes allow for the evolutionary optimality of fragmentation modes according to which groups split at the maximum size \( n = 4 \) (2+1 under proportional costs, and 1+1+1 under fixed costs), and hence for life cycles where multicellular phases are persistent. This seems paradoxical until one realises that by staying together as long as possible groups delay as much as possible the inevitable cell loss associated to a fragmentation event. Thus, even if groups are less fecund or die at a higher rate than independent cells, staying together might be adaptive if splitting apart is too costly.

**Synergistic interactions between cells promote the production of unicellular propagules, while diminishing returns promote equal binary fragmentation**

Next, we focus on fitness landscapes for which either the birth rate of cells increases with group size (fecundity landscapes where larger groups are always more productive) or the death rate of groups decreases with group size (survival landscapes where larger groups always live
In this case, and for a maximum group size \( n = 4 \), the set of optimal modes is given by \( 2+2 \) and \( 3+1 \) if there are no fragmentation costs (Fig 3), by \( 2+1 \) if fragmentation costs are proportional to the number of fragments (Fig 3A and 3B), and by \( 2+1 \) and \( 1+1+1 \) if fragmentation involves a fixed cost of one cell (Fig 4C and 4D).

To investigate larger maximum group sizes \( n \) in a simple but systematic way, we consider fecundity landscapes with birth rates given by \( b_i = 1 + Mg_i \) and survival landscapes with death rates given by \( d_i = M(1 - g_i) \), where \( g_i = [(i - 1)/(n - 2)]^\alpha \) [29] models the fecundity or survival benefits associated to group size \( i \) and \( M > 0 \) is the maximum benefit (Fig 5). The parameter \( \alpha \) is the degree of complementarity between cells; it measures how important the addition of another cell to the group is in producing the maximum possible benefit \( M \) [30]. For low degrees of complementarity (\( \alpha < 1 \)), the sequence \( g_i \) is strictly concave and each additional cell

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Fig 4. Optimal fragmentation modes for fecundity and survival landscapes (costly fragmentation). For proportional costs (panels A and B), splitting into \( n \) parts involves the loss of \( n - 1 \) cells. In this case, and for \( n = 4 \), only two pure modes are possible: \( 2+1 \) (whereby a 4-unit group splits into a pair of cells and a single cell and loses one cell) and \( 1+1 \) (whereby a group of three splits into two single cells and loses one cell). For fixed costs (panels C and D), splitting involves the loss of a single cell, no matter the kind of partition. In this case, and for \( n = 4 \), an additional mode is possible: \( 1+1+1 \) (whereby a 4-unit group splits into three single cells and loses one cell). This last nonbinary mode can be optimal under a wide range of parameters.

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contributes less to the per capita benefit of group living [31] and groups of all sizes achieve the same functionality as $\alpha$ tends to zero. If $\alpha = 1$, the sequence $g_i$ is linear, and each additional cell contributes equally to the fecundity or survival of the group. Finally, for high degrees of complementarity ($\alpha > 1$), the sequence $g_i$ is strictly convex and each additional cell improves the performance of the group more than the previous cell did. In the limit of large $\alpha$, the advantages of group living materialise only when complexes achieve the maximum size $n - 1$ [31].

We numerically calculate the optimal fragmentation modes for $n = 20$ (costless fragmentation) or $n = 21$ (costly fragmentation) and the fitness landscapes described above for parameter values taken from $0.01 \leq \alpha \leq 100$ and $0.02 \leq M \leq 50$ (Figs 6 and 7). In line with our general analytical results, optimal fragmentation modes are always characterised by binary splitting when fragmentation is costless or when it involves proportional costs, while nonbinary splitting can be optimal only if fragmentation involves a fixed cost. We also find that, for each value of $\alpha$ and $M$, and for both costless and costly fragmentation, the optimal fragmentation mode is one where fragmentation occurs at the largest possible size. This is expected since the benefit sequence is increasing in group size and thus groups of maximum size perform better, either by achieving the largest birth rate per unit (fecundity landscapes) or the lowest death rate (survival landscapes). Which particular fragmentation mode maximizes the growth rate depends nontrivially on whether fragmentation is costless or costly (and in the latter case also on how such costs are implemented), on the kind of group size benefits (fecundity or survival),

Fig 5. Advantages of group living. Group size benefit $g_i = [(i-1)/(n-2)]^\alpha$ as a function of group size for different values of the degree of complementarity $\alpha$. If $\alpha < 1$, $g_i$ is concave; if $\alpha = 1$, $g_i$ is linear; if $\alpha > 1$, $g_i$ is convex.
on the maximum possible benefit $M$, and on the degree of complementarity $\alpha$. Despite this apparent complexity, some general patterns can be identified.

Let us focus on the case of fecundity landscapes and first fasten attention on the scenario of costless fragmentation (Fig 6A). A salient feature of this case is the prominence of two

Fig 6. Optimal life cycles under monotonic fecundity landscapes. Birth rates are given by $b_i = 1 + Mg_i$ where $g_i = ((i - 1)/(n - 2))^\alpha$. A Costless fragmentation, $n = 20$. B Fragmentation with proportional costs, $n = 21$. C Fragmentation with fixed costs, $n = 21$. For costless fragmentation and fragmentation with proportional costs, only binary modes 19+1, 18+2, …, 10+10 are optimal. In these cases, diminishing returns ($\alpha < 1$) make equal binary fragmentation (10+10) optimal. Also, optimality of the unicellular propagule strategy (19+1) requires increasing returns ($\alpha > 1$). For fragmentation with fixed costs, nonbinary modes 7+7+6, …, 1+1+1 can also be optimal.

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Fig 7. Optimal life cycles under monotonic survival landscapes. Death rates are given by $d_i = M(1 - g_i)$ where $g_i = ((i - 1)/(n - 2))^\alpha$. A Costless fragmentation, $n = 20$. B Fragmentation with proportional costs, $n = 21$. C Fragmentation with fixed costs, $n = 21$. For costless fragmentation and fragmentation with proportional costs, only binary modes 19+1, 18+2, …, 10+10 are optimal. In these cases, diminishing returns to scale ($\alpha < 1$) make equal binary fragmentation (10+10) optimal. Also, optimality of the unicellular propagule strategy (19+1) requires increasing returns to scale ($\alpha > 1$). For fragmentation with fixed costs, nonbinary modes 7+7+6, …, 1+1+1 can also be optimal.

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qualitatively different fragmentation modes: the “equal binary fragmentation” strategy 10+10 (whereby offspring groups have sizes as similar as possible) and the “unicellular propagule” strategy 19+1 (whereby offspring groups have sizes as different as possible). A sufficient condition for equal binary fragmentation to be optimal is that increase in size is characterised by diminishing returns. The intuition behind this result is that, if the degree of complementarity is small, then groups (complexes of size \(i \geq 2\)) have similar performance, while independent cells (\(i = 1\)) are at a significant disadvantage. Therefore, the optimal strategy is to ensure that both offspring groups are as large as possible, and hence of the same size. However, equal binary fragmentation can be also optimal for synergistic interactions, provided that complementarity is not too high. In contrast, the unicellular propagule strategy is optimal only for relatively high degrees of complementarity. This is because when complementarity is high only the largest group can reap the benefits of group living; in this case, the optimal mode is to have at least one offspring of very large size. Compared to 19+1 and 10+10, other binary splitting strategies are optimal in smaller regions of the parameter space, and in all cases only for synergistic interactions between cells.

Consider now the effects of introducing fragmentation costs proportional to the number of fragments (Fig 6B). Here, the region where the unicellular propagule strategy is optimal shrinks to the corner of the parameter space where benefits of group living and degree of complementarity are maximum, while the region of optimality for equal binary fragmentation expands. This makes intuitive sense. With fragmentation costs, the largest offspring group resulting from fragmenting according to the unicellular propagule strategy is of size 19, and hence always on the brink of fragmentation (once it grows to size 21) and incurring one cell loss. When group benefits are not high and synergistic enough, the unicellular propagule strategy is dominated by fragmentation modes (in particular, equal binary fragmentation) having smaller offspring for which the costs of fragmentation are not so immediate.

Finally, if costs of fragmentation are not proportional but fixed (Fig 6C), then two classes of nonbinary splitting become optimal in regions of the parameter space where equal binary fragmentation was optimal under proportional costs: (i) "multiple fission" (1+1+. . .+1) which is in general favored for small maximum benefit and increasing returns, and (ii) "multiple groups" (modes 2+2+. . .+2, 3+3+3+3+2, 4+4+3+3+3+3, 4+4+4+4+4, 5+5+5+5, and 7+7+6) which are often optimal for diminishing returns.

Fig 7 show the results for survival landscapes. The main difference in this case is that the unicellular propagule strategy can be the optimal strategy even when group living is characterised by diminishing returns. In general, fecundity benefits make equal binary fragmentation optimal under more demographic scenarios, while survival benefits make the unicellular propagule strategy optimal under more demographic scenarios.

Discussion

Reproduction is such a fundamental feature of living systems that the idea that the mode of reproduction may be shaped by natural selection is easily overlooked. Here, we analysed a matrix population model that captures the demographic dynamics of complexes that grow by staying together and reproduce by fragmentation. The costs and benefits associated with group size ultimately determine whether or not a single cell fragments into two separate daughter cells upon cell division, or whether those daughter cells remain in close proximity, with fragmentation occurring only after subsequent rounds of division. We allowed for a vast and complete space of fragmentation strategies, including pure modes (specifying all possible combinations of size at fragmentation and fragmentation pattern) and mixed modes (specifying all probability distributions over the set of pure modes), and identified those modes
achieving a maximum growth rate for given fecundity and survival size-dependent rates. Our research questions and methodology thus resonates with previous studies in life history theory [32, 33]. In the language of this field, our fragmentation strategies specify both the size at first reproduction and clutch size, where the latter is subject to a very specific trade-off between the number and size of offspring mathematically given by integer partitions.

We found that for any fitness landscape, the optimal life cycle is always a deterministic fragmentation mode involving the regular schedule of group development and fragmentation. This makes intuitive sense given our assumption that the environment is constant. However, this result might not hold if the environment is variable so that the fitness landscape changes over time. In this case different pure fragmentation modes will be optimal at different times, and natural selection might favour life cycles that randomly express a subset of locally optimal fragmentation patterns. Indeed, the evolution of variable phenotypes in response to changing environmental conditions (also known as bet hedging [34, 35]) has been demonstrated in other life history traits, such as germination time in annual plants [36], and capsulation in bacteria [37]. The extent to which mixed fragmentation modes can evolve via a similar mechanism is beyond the scope of this paper, but it can be addressed in future work by applying existing theory on matrix population models in stochastic environments [22].

We found that when fragmentation is costless, only strategies involving binary splitting (i.e., fragmentation into exactly two parts) are optimal. This result holds for all possible fitness landscapes, and hence any specification of how fecundity or survival benefits might accrue to group living. In particular, the optimal fragmentation mode under monotonic fitness landscapes is generally one of two types: equal binary fragmentation, which involves fission into two equal size groups, and the unicellular propagule strategy, which involves the production of two groups, one comprised of a single cell. Equal fragmentation is favoured when there is a significant advantage associated with formation of even the smallest group, whereas production of a unicellular propagule is favoured when the benefits associated with group size are not evident until groups become large. This makes intuitive sense: when advantages arise when groups are small, it pays for offspring to be in groups (and not single cells). Conversely, when there is little gain until group size is large, it makes sense to maintain one group that reaps this advantage. Interestingly, two bacteria that form groups and are well studied from a clinical perspective, Neisseria gonorrhoeae and Staphylococcus aureus, both show evidence of the above binary splitting fragmentation modes: Neisseria gonorrhoeae divide into groups of two equal sizes [6], while Staphylococcus aureus divide into one large group plus a unicellular propagule [7]. This leads to questions concerning the nature of the fitness landscape occupied by these bacteria and the basis of any collective level benefit as assumed by our model.

Once cell loss upon fragmentation is incorporated as a factor in collective reproduction, a wider range of fragmentation patterns becomes optimal. When fragmentation costs are fixed to a given number of cells, optimal fragmentation modes include those where splitting involves the production of multiple offspring. Among these, a prominent fragmentation strategy is multiple fission, where a group breaks into multiple independent cells. Such a fragmentation mode is reminiscent of palintomy in the volvocine algae [38]. A key difference between our “multiple fission” and palintomy is that the former involves a group of cells growing up to a threshold size at which point fragmentation happens, while the latter involves a single reproductive cell growing to many times its initial size and then undergoing several rounds of division. However, reinterpreting birth rates of cells in groups as growth rates of unicells of different sizes allows us to use our analysis to determine conditions under which such a mode of fragmentation is more adaptive than, say, the more standard strategy of growing to twice the initial size and then dividing in two (which for arbitrary sizes of offspring groups is equivalent to our “equal binary fragmentation” mode). Our results suggest that palintomy is favored
over binary fission (and any other fragmentation mode) under a wide range of demographic scenarios (Fig 6C).

Many multicellular organisms are characterised by a life cycle whereby adults develop from a single cell [39]. Passing through such a unicellular bottleneck is a requirement for sexual reproduction based on syngamy, but life cycles with unicellular stages are also common in asexual reproduction modes such as those used by multicellular algae and ciliates [40], and colonial bacteria such as *S. aureus* [7]. If multicellularity evolved because of the benefits associated to group living, why do so many asexual multicellular organisms begin their life cycles as solitary (and potentially vulnerable) cells? Explanatory hypotheses include the purge of deleterious mutations and the reduction of within-organism conflict [39, 41]. Our results make the case for an alternative (and perhaps more parsimonious) explanation: often, a life cycle featuring a unicellular bottleneck is the best way to guarantee that the “parent” group remains as large as possible to reap maximum fecundity and/or survival advantages of group living. Indeed, our theoretical results resonate with previous experimental work demonstrating that single-cell bottlenecks can be adaptive simply because they constitute the life history strategy that maximises reproductive success [42].

Previous theoretical work has explored questions related to the evolution of multicellularity using matrix population models similar to the one proposed in this paper. In a seminal contribution, Roze and Michod [43] explored the evolution of propagule size in the face of deleterious and selfish mutations. In their model, multicellular groups first grow to adult size and then reproduce by splitting into equal size groups, so that fragmentation mode strategies can be indexed by the size of the propagule. In our terminology, this refers to either “multiple fission” or “multiple groups”. An important finding of Roze and Michod [43] is that, even if large groups are advantageous, small propagules can be selected because they are more efficient at eliminating detrimental mutations. We did not study the effects of mutations, but allowed for general fitness landscapes and fragmentation modes, including cases of asymmetric binary division (e.g., the unicellular propagule strategy) neglected by Roze and Michod [43]. Our results indicate that modes of fragmentation involving single cells can lead to growth rate maximisation even when small propagule sizes divide less efficiently or die at a higher rate. In particular, we have shown that if fragmentation is costly, a strategy consisting of a multiple fragmentation mode with a propagule size of one (i.e., the small propagule strategy studied by Roze and Michod [43]) can be adaptive for reasons other than the elimination of deleterious mutations.

Closer to our work, Tarnita et al. [18] investigated the evolution of multicellular life cycles via two alternative routes: “staying together” (ST, whereby offspring cells remain attached to the parent) and “coming together” (CT, whereby cells of different origins aggregate in a group). In particular, they studied the conditions under which a multicellular strategy that produces groups via ST can outperform a solitary strategy whereby cells always separate after division. The way they modelled group formation and analyzed the resulting population dynamics (by means of biological reactions and matrix models) is closely related to our approach. Indeed, their solitary strategy is our binary mode 1+1, while their ST strategy corresponds to a particular kind of binary mixed fragmentation mode. However, the questions we ask are different. Tarnita et al. [18] were concerned with the conditions under which (multicellular) strategies that form groups can invade and replace (unicellular) strategies that remain solitary. Contrastingly, we aimed to understand the optimal fragmentation mode out of the vast space of fragmentation strategies comprising all possible deterministic and probabilistic pathways by which complexes can stay together and split apart. A key finding is that, for any fitness landscape and if the environment is constant, mixed fragmentation modes such as
some of the ST strategies considered by Tarnita et al. [18] will be outperformed by at least one pure fragmentation mode.

More recently, Rashidi et al. [20] developed a conceptual framework to study the competition of life cycles that involved five different life cycles defined by fragmentation patterns of the form 1+1+. . .+1 and an associated genetic control. Their model, which explicitly considers growing cells of different size, showed that depending on the fitness landscape, each of their five life cycles could prevail. By extending the range of life cycles to encompass all possible fragmentation modes (albeit with less detailed attributes), we have shown that certain life cycles will be suboptimal for any given fitness landscape.

In line with many studies in life history theory [32, 33], we made the simplifying assumption that the phenotype consists of demographic traits (in our case, probabilities of fragmenting into given fragmentation patterns) linked by trade-offs which interact to determine fitness (growth rate). This allowed us to predict the optimal phenotype at equilibrium at the expense of leaving unspecified whether, due to genetic constraints, such an equilibrium will be possible in an actual biological system. The question that inevitably arises is whether, given a presumptive genotype-phenotype mapping, it is possible for evolution to fine tune life cycles with group-level properties (such as specific fragmentation patterns) so that optimal fragmentation modes will be obtained as the endpoint of an evolutionary process. While a complete answer requires a more sophisticated analysis, we see no conceptual obstruction preventing seemingly arbitrary fragmentation modes to evolve. Firstly, genotype-phenotype maps of existing organisms can be complex and offer opportunity for adaptation, involving important qualitative behavioral changes [44–46]. Secondly, small genotypic changes can produce major phenotypic changes. For instance, Hammerschmidt et al. [3] observed the emergence of collective-level properties in a previously unicellular organism that was caused by a small number of mutations. Thirdly, even if a current set of genes cannot provide an appropriate template for given phenotypic traits, new genes can emerge de novo [47–51]. Finally, theoretical arguments suggest that genetic constraints can be effectively overcome in phenotypic evolution provided there is a rich variety of new mutant alleles [52]. We thus think that, both in the field and in the laboratory, multicellular organisms will be able to evolve a phenotype close to the optimal fragmentation mode in the (very) long run.

Supporting information
S1 Text. Supplementary methods.
(PDF)

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References

1. Godfrey-Smith P. Darwinian populations and natural selection. Oxford University Press; 2009.
2. Libby E, Rainey PB. A conceptual framework for the evolutionary origins of multicellularity. Physical Biology. 2013; 10(3):035001. https://doi.org/10.1088/1478-3975/10/3/035001
3. Hammerschmidt K, Rose CJ, Kerr B, Rainey PB. Life cycles, fitness decoupling and the evolution of multicellularity. Nature. 2014; 515(7525):75–79. https://doi.org/10.1038/nature13884 PMID: 25373677
4. van Gestel J, Tarnita CE. On the origin of biological construction, with a focus on multicellularity. Proceedings of the National Academy of Sciences. 2017; 114(42):11018–11026. https://doi.org/10.1073/pnas.1704631114
5. Angert ER. Alternatives to binary fission in bacteria. Nature Reviews Microbiology. 2005; 3(3):214–224. https://doi.org/10.1038/nrmicro1096
6. Westling-Håggström B, Elmros T, Normark S, Winblad B. Growth pattern and cell division in Neisseria gonorrhoeae. Journal of Bacteriology. 1977; 129(1):333–342. PMID: 807582
7. Koyama T, Yamada M, Matsushashi M. Formation of regular packets of Staphylococcus aureus cells. Journal of Bacteriology. 1977; 129(3):1518–1523. PMID: 845124
8. Keim CN, Martins JL, Abreu F, Rosado AS, de Barros HL, Borojec R, et al. Multicellular life cycle of magnetotactic prokaryotes. FEMS Microbiology Letters. 2004; 240(2):203–208. https://doi.org/10.1016/j.femsle.2004.09.035 PMID: 15522508
9. de la Fuente-Núñez C, Rebuffe-Charleux F, Fernández L, Hancock REW. Bacterial biofilm development as a multicellular adaptation: antibiotic resistance and new therapeutic strategies. Current Opinion in Microbiology. 2013; 16(5):580–589. https://doi.org/10.1016/j.mib.2013.06.013 PMID: 23880136
10. Boraas ME, Seale DB, Boxhorn JE. Phagotrophy by a flagellate selects for colonial prey: A possible origin of multicellularity. Evolutionary Ecology. 1998; 12(2):153–164. https://doi.org/10.1023/A:1006572582063
11. Kapselaki SE, Fisher RM, West SA. Predation and the formation of multicellular groups in algae. Evolutionary Ecology Research. 2016; 17(5):651–669.
12. Bonner JT. Evolutionary strategies and developmental constraints in the cellular slime molds. The American Naturalist. 1982; 119(4):530–552. https://doi.org/10.1086/283930
13. Rainey PB, De Monte S. Resolving conflicts during the evolutionary transition to multicellularity. Annual Review of Ecology, Evolution, and Systematics. 2014; 45:599–620. https://doi.org/10.1146/annurev-ecolsys-120213-091740
14. Williams P, Bainton NJ, Swift S, Chhabra SR, Winson MK, Stewart GS, et al. Small molecule-mediated density-dependent control of gene expression in prokaryotes: bioluminescence and the biosynthesis of carbapenem antibiotics. FEMS microbiology letters. 1992; 100(1-3):161–167. https://doi.org/10.1111/j.1574-6968.1992.tb05698.x PMID: 1478452
15. Diggle SP, Griffin AS, Campbell GS, West SA. Cooperation and conflict in quorum-sensing bacterial populations. Nature. 2007; 450(7168), 411–414. https://doi.org/10.1038/nature06279 PMID: 18004343
16. Groebe K, Mueller-Klieser W. On the relation between size of necrosis and diameter of tumor spheroids. International Journal of Radiation Oncology*Biology*Physics. 1996; 34(2):411–414. https://doi.org/10.1016/0360-3016(95)00172-6 PMID: 8724367
17. Stewart PS, Franklin MJ. Physiological heterogeneity in biofilms. Nature Reviews Microbiology. 2008; 6(3):199–210. https://doi.org/10.1038/nrmicro1838 PMID: 18264116
18. Tarnita CE, Taubes CH, Nowak MA. Evolutionary construction by staying together and coming together. Journal of Theoretical Biology. 2013; 320(0):10–22. https://doi.org/10.1016/j.jtbi.2012.11.022 PMID: 23206384
19. Libby E, Ratcliff WC, Travisano M, Kerr B. Geometry shapes evolution of early multicellularity. PLoS Computational Biology. 2014; 10(9):e1003803. https://doi.org/10.1371/journal.pcbi.1003803 PMID: 25233196
20. Rashidi A, Shelton DE, Michod RE. A Darwinian approach to the origin of life cycles with group properties. Theoretical Population Biology. 2015; 102:76–84. https://doi.org/10.1016/j.tpb.2015.03.003 PMID: 25814207
21. Kaveh K, Veller C, Nowak MA. Games of multicellularity. Journal of Theoretical Biology. 2016; 403:143–158. https://doi.org/10.1016/j.jtbi.2016.04.037 PMID: 27179461
22. Caswell H. Matrix population models. 2nd ed. Sinauer Associates; 2001.
23. Andrews GE. The theory of partitions. Cambridge, UK: Cambridge University Press; 1998.
24. Hofbauer J, Sigmund K. Evolutionary games and population dynamics. Cambridge, UK: Cambridge University Press; 1998.
25. Cohen JE. Convexity of the dominant eigenvalue of an essentially nonnegative matrix. Proceedings of the American Mathematical Society. 1981; 81(4):657–658. https://doi.org/10.2307/2044180
26. Kirk DL. A twelve step program for evolving multicellularity and a division of labor. BioEssays. 2005; 27(3):299–310. https://doi.org/10.1002/bies.20197 PMID: 15714559
27. Rossetti V, Filippini M, Svercel M, Barbour AD, Bagheri HC. Emergent multicellular life cycles in filamentous bacteria owing to density-dependent population dynamics. Journal of The Royal Society Interface. 2011; 8(65):1772–1784. https://doi.org/10.1098/rsif.2011.0102
28. Ratcliff WC, Denison RF, Borrello M, Travisano M. Experimental evolution of multicellularity. Proceedings of the National Academy of Sciences USA. 2012; 109(5):1595–1600.
29. Fromhage L, Kokko H. Monogamy and haplodiploidy act in synergy to promote the evolution of eusociality. Nature Communications. 2011; 2:397. https://doi.org/10.1038/ncomms1410 PMID: 21772268
30. De Jaegher K. Harsh environments and the evolution of multi-player cooperation. Theoretical Population Biology. 2017; 113:1–12. https://doi.org/10.1016/j.tpb.2016.09.003 PMID: 27664440
31. Hirshleifer J. From weakest-link to best-shot: The voluntary provision of public goods. Public Choice. 1983; 41(3):371–386. https://doi.org/10.1007/BF00141070
32. Stearns SC. The evolution of life histories. Oxford University Press, Oxford: 1992.
33. Roff DA. Life history evolution. Sinauer Associates; 2002.
34. Slatkin M. Hedging one’s evolutionary bets. Nature. 1974; 250:704–705. https://doi.org/10.1038/250704b0
35. Bull JJ. Evolution of phenotypic variance. Evolution. 1987; 41(2):303–315. https://doi.org/10.2307/2409140 PMID: 2856756
36. Cohen D. Optimizing reproduction in a randomly varying environment. Journal of Theoretical Biology. 1966; 12(1):119–129. https://doi.org/10.1016/0022-5193(66)90188-3 PMID: 6015423
37. Beaumont HUE, Galilee J, Kost C, Ferguson GC, Rainey PB. Experimental evolution of bet hedging. Nature. 2009; 462:90–93. https://doi.org/10.1038/nature08504 PMID: 19890329
38. Michod RE. Evolution of individuality during the transition from unicellular to multicellular life. Proceedings of the National Academy of Sciences. 2007; 104(Suppl 1):8613–8618. https://doi.org/10.1073/pnas.0701489104
39. Grosberg RK, Strathmann RR. One cell, two cell, red cell, blue cell; the persistence of a unicellular stage in multicellular life histories. Trends in ecology & evolution. 1998; 13(3):112–116. https://doi.org/10.1016/S0169-5347(97)01313-X
40. Herron MD, Rashidi A, Shenton DE, Driscoll WW. Cellular differentiation and individuality in the ‘minor’ multicellular taxa. Biologival Reviews. 2013; 88(4):844–861. https://doi.org/10.1111/bvr.12031 PMID: 23448295
41. Maynard Smith J, Szathmáry E. The major transitions in evolution. Oxford: W. H. Freeman; 1995.
42. Ratcliff WC, Herron MD, Howell K, Pentz JT, Rosenzweig F, Travisano M. Experimental evolution of an alternating uni- and multicellular life cycle in Chlamydomonas reinhardtii. Nature Communications. 2013; 4(2742). https://doi.org/10.1038/ncomms3742 PMID: 24193369
43. Roze D, Michod RE. Mutation, multilevel selection, and the evolution of propageal size during the origin of multicellularity. The American Naturalist. 2001; 158(6):638–654. https://doi.org/10.1086/323590 PMID: 18707357
44. Rainey PB, Travisano M. Adaptive radiation in a heterogeneous environment. Nature. 1998; 394(6688):69–72. https://doi.org/10.1038/27900 PMID: 9665128
45. Anderson CL, Karr TL. Wolbachia: evolutionary novelty in a rickettsial bacteria. BMC Evolutionary Biology. 2001; 1(1):10. https://doi.org/10.1186/1471-2148-1-10 PMID: 11734058
46. Rainey PB, Rainey K. Evolution of cooperation and conflict in experimental bacterial populations. Nature. 2003; 425(6953):72–74. https://doi.org/10.1038/nature01906 PMID: 12955142
47. Tautz D, Domazet-Lošo T. The evolutionary origin of orphan genes. Nature Reviews Genetics. 2011; 12(10):692–702. https://doi.org/10.1038/nrg3053 PMID: 21878963
48. Donoghue MT, Keshavaiah C, Swamidatta SH, Spillane C. Evolutionary origins of Brassicaceae specific genes in Arabidopsis thaliana. BMC Evolutionary Biology. 2011; 11(1):47. https://doi.org/10.1186/1471-2148-11-47 PMID: 21332978
49. Sabath N, Wagner A, Karlin D. Evolution of viral proteins originated de novo by overprinting. Molecular biology and evolution. 2012; 29(12):3767–3780. https://doi.org/10.1093/molbev/mss179 PMID: 22821011

50. Schlotterer C. Genes from scratch—the evolutionary fate of de novo genes. Trends in Genetics. 2015; 31(4):215–219. https://doi.org/10.1016/j.tig.2015.02.007 PMID: 25773713

51. Farr AD, Remigi P, Rainey PB. Adaptive evolution by spontaneous domain fusion and protein relocalisation. Nature Ecology and Evolution. 2017; 1:1562–1568. https://doi.org/10.1038/s41559-017-0283-7

52. Hammerstein P. Darwinian adaptation, population genetics and the streetcar theory of evolution. Journal of Mathematical Biology. 1996; 34(5-6):511–532. https://doi.org/10.1007/BF02409748 PMID: 8691083