MEGANTE: A Web-Based System for Integrated Plant Genome Annotation

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The recent advancement of high-throughput genome sequencing technologies has resulted in a considerable increase in demands for large-scale genome annotation. While annotation is a crucial step for downstream data analyses and experimental studies, this process requires substantial expertise and knowledge of bioinformatics. Here we present MEGANTE, a web-based annotation system that makes plant genome annotation easy for researchers unfamiliar with bioinformatics. Without any complicated configuration, users can perform genomic sequence annotations simply by uploading a sequence and selecting the species to query. MEGANTE automatically runs several analysis programs and integrates the results to select the appropriate consensus exon–intron structures and to predict open reading frames (ORFs) at each locus. Functional annotation, including a similarity search against known proteins and a functional domain search, are also performed for the predicted ORFs. The resultant annotation information is visualized with a widely used genome browser, GBrowse. For ease of analysis, the results can be downloaded in Microsoft Excel format. All of the query sequences and annotation results are stored on the server side so that users can access their own data from virtually anywhere on the web. The current release of MEGANTE targets 24 plant species from the Brassicaceae, Fabaceae, Musaceae, Poaceae, Solanaceae, Rosaceae and Vitaceae families, and it allows users to submit a sequence up to 10 Mb in length and to save up to 100 sequences with the annotation information on the server. The MEGANTE web service is available at https://megante.dna.affrc.go.jp/.

Keywords: Gene prediction • Plant genome annotation • Web service.

Abbreviations: CDS, coding sequence; EST, expressed sequence tag; FLCDNA, full-length cDNA; GO, gene ontology; ORF, open reading frame; Sn, sensitivity; Sp, specificity.

Introduction

With the advent of high-throughput sequencing technologies, plant genome sequencing has been accelerated, and the data are being utilized for crop improvement (Bevan and Uauy 2013). The accumulation of the large amount of plant genome sequences led to constructions of comparative genomics databases (Mihara et al. 2010, Nagamura et al. 2011, Rouard et al. 2011, Goodstein et al. 2012) and development of a plant-specific controlled vocabulary for effective data integration (Cooper et al. 2013). However, the costs of the data management and analyses are increasing because of the need for high-spec computers, huge amounts of data storage and expertise in both computer science and molecular biology. In these data analyses, genome annotation is one of the most fundamental and indispensable steps (Yandell and Ence 2012), directly affecting further studies such as molecular evolutionary analyses, transposon tagging and microarray experiments. The annotation procedures require a higher level of bioinformatics skill, as several analysis programs must be conducted followed by the integration of the results to predict gene structures and assign gene functions. Thus, an easy-to-use annotation platform, which does not require any expertise in bioinformatics, would be essential for researchers to perform genome annotation and to visualize the results on a graphical viewer to interpret the annotation.

Currently, several types of analysis tools are available online for plant genome annotation. For example, online versions of ab initio gene prediction programs, such as AUGUSTUS (Stanke and Waack 2003), FgenesSH (Salamov and Solovyev 2000) and GeneMark.hmm (Lukashin and Borodovsky 1998), can be used to find open reading frames (ORFs) from genomic sequences. FPGP (Amano et al. 2010) aligns full-length cDNA (FLcDNA) sequences of dicot and monocot plants to a query sequence. Gramene (Youens-Clark et al. 2011) and PlantGDB (Duvick et al. 2008) provide a web service for a similarity search against plant nucleotide or protein databases. For graphical representation of the analysis results, WebGBrowse (Podicheti et al. 2009) is a good candidate. Although such web services are useful for genome annotation, it is time-consuming for researchers to access multiple web sites and interpret their results one by one. Moreover, such an annotation procedure is difficult for non-bioinformaticians to select the appropriate tools and parameter sets for the input sequences. Therefore, an integrated analysis tool to execute a series of analysis
programs automatically is required to support genome analyses such as positional cloning of plant genes (Chen et al. 2009, Xu et al. 2011). Several web-based annotation pipelines are available for plant genome sequences. Some of them are designed for specific plant genome annotation; RiceGAAS (Sakata et al. 2002) is for rice and TriAnnot (Leroy et al. 2012) is for wheat. There are also more versatile genome annotation tools that can be adapted not only for plants but also for other species. DNA subway (Goff et al. 2011) provides parameter sets for both animals and plants. MAKER (Cantarel et al. 2008) has a highly configurable web interface to select reference databases and parameters for analysis programs. However, there are few plant species that are supported in the existing annotation pipelines.

Here we describe a new plant genome annotation web service called MEGANTE that runs several analysis programs against query sequences, integrates the results and visualizes the annotation information on a genome browser. Compared with the existing tools, one of the notable features of MEGANTE is its simple interface, which is easy to use, even for non-experts. In addition, the service targets a wide variety of plant species and is able to accept large query sequences up to 10 Mb in length.

Results

Features of the MEGANTE web service

At the time of first use, MEGANTE requires an e-mail address and password to create an account. MEGANTE stores all data including query sequences submitted by users and the analysis results on the server side, and stores the data until users explicitly remove them via a web interface (Fig. 1A). Several analysis programs are automatically conducted in the system, but users do not need to specify any parameters or reference databases for the annotation process. Users can start annotation simply.

Fig. 1 Screenshots of the MEGANTE web interface. (A) Uploaded queries are listed. The list allows users to see the statuses of annotation jobs, download analysis results and jump to an annotation viewer. Clicking the sequence ID shows or hides detailed information about the query sequence. (B) Users can submit query sequences through this interface. (C) Annotation viewer with GBrowse. Users can select which data tracks to show or hide on the annotation map with the ‘Select Tracks’ tab. (D) Detailed annotation information of the predicted genes linked from the data tracks in GBrowse. ORF and amino acid sequences are also shown on this page.
between web browsers and the server is protected by SSL. Users upon completion of the annotation. The data transfer when submitting a query, an e-mail is sent to notify the system archives the annotation results in a single ZIP file for each species listed in Table 1, we run a SeqClean script (http://sourceforge.net/projects/seqclean/) to remove poly(A) tails, vectors, low complexities and short sequences from the transcripts.

### Table 1 Species supported in MEGANTE

| Families   | Species                      |
|------------|------------------------------|
| Brassicaceae | Arabidopsis thaliana  |
|            | Brassica napus              |
|            | Brassica rapa               |
|            | Raphanus sativus           |
| Fabaceae   | Glycine max                 |
|            | Lotus japonicus            |
|            | Medicago truncatula        |
|            | Vigna unguiculata          |
| Musaceae   | Musa acuminata             |
| Poaceae    | Brachypodium distachyon    |
|            | Hordeum vulgare            |
|            | Oryza sativa               |
|            | Phyllostachys edulis       |
|            | Sorghum bicolor            |
|            | Triticum aestivum          |
|            | Zea mays                   |
| Salicaceae | Populus trichocarpa        |
| Solanaceae | Nicotiana tabacum          |
|            | Solanum lycopersicum       |
|            | Solanum melongena          |
|            | Solanum tuberosum          |
| Rosaceae   | Malus × domestica          |
|            | Prunus persica             |
| Vitaceae   | Vitis vinifera             |

### Reference databases used in the system and pre-processing for annotation

MEGANTE uses several reference databases for the genome annotation, which include FLcDNAs and expressed sequence tags (ESTs) obtained from INSDC (Nakamura et al. 2013), protein sequences from Swiss-Prot and the TrEMBL plant division of UniProtKB (Magrane and UniProt Consortium 2011), and a protein family and domain database, InterPro (Hunter et al. 2011). We update the databases on a regular basis. Up-to-date details of the databases, such as the number of sequences, are described on the MEGANTE web site. After retrieving FLcDNAs and ESTs for each species listed in Table 1, we run the sim4db alignment, only interspecies FLcDNA alignment. In the sim4db alignment, only interspecies FLcDNAs from the same class (monocot or dicot) of plants are used to reduce the calculation time. The cut-off identity and coverage of the protein alignment are set to 90%. For gene prediction of the Musaceae and Rosaceae families, which have a relatively small number of sequences in protein databases, the values are relaxed to 80% to increase the number of proteins that could be mapped to the queried sequences. It was confirmed that the relaxed condition did not decrease the gene prediction accuracy (data not shown). The system runs sim4db with an identity and coverage cut-off of 50% and then finds the longest ORFs in each locus for a downstream analysis. All the results, which contain genes predicted by the four ab initio gene finders, protein alignments and ORFs generated from the sim4db alignments, are merged to create consensus gene structures using JIGSAW (Allen et al. 2006). Simultaneously, PASA (Haas et al. 2003) generates EST assemblies by mapping intraspecies ESTs to the query sequence. To achieve a more accurate prediction, the system runs PASA again to incorporate the EST assemblies into the consensus gene structures generated by JIGSAW. Consequently, predicted genes are classified into three categories: (i) genes inferred from intraspecies FLcDNAs; (ii) genes

### Annotation workflow

The overall annotation workflow is shown in Fig. 2. The annotation process begins with filtering out repeat elements detected by RepeatMasker (http://repeatmasker.org) with the MIPS Repeat Element Database (Nussbaumer et al. 2013). Next, to predict the exon–intron structures, the system aligns intraspecies FLcDNAs to a query sequence using BLAT (Kent 2002) with a cut-off of ≥98% identity and coverage. Although intraspecies FLcDNAs are effective for accurate gene prediction, in many cases the number of the sequences is not sufficient to cover entire genes. For this reason, we also use AUGUSTUS (Stanke and Waack 2003), GeneZilla (Allen et al. 2006), GlimmerHMM (Allen et al. 2006) and SNAP (Korf 2004) for ab initio gene prediction. For protein alignment, ProSplign (Sayers et al. 2012) for protein alignment with SwissProt and TrEMBL, and sim4db (Walenz and Florea 2011) for interspecies FLcDNA alignment. The uploaded sequences are first queued, and then the queries are processed on an application server in a round-robin fashion to schedule the processes fairly. In the current system, five annotation processes can run in parallel. The whole annotation process can be completed within approximately 150 min for a 1 Mb sequence and approximately 15 h for a 10 Mb sequence. After finishing the annotation process, the following results are reported: repeat elements; alignments of transcript and protein sequences; predicted gene structures; similarities to known proteins; functional domains; and Gene Ontology (GO) terms (Ashburner et al. 2000). All the results are visualized with a widely used genome browser, GBrowse (Stein et al. 2002), which is integrated into the system. Furthermore, the system archives the annotation results in a single ZIP file for download. The file contains the annotation information in both Microsoft Excel and GFF3 (http://www.sequenceontology.org/gff3.html) formats. If users select an option for e-mail notification when submitting a query, an e-mail is sent to notify the users upon completion of the annotation. The data transfer between web browsers and the server is protected by SSL encryption.
successfully incorporated with EST assemblies; and (iii) genes that did not overlap with EST assemblies or genes that failed to be incorporated with EST assemblies because of inconsistencies between the exon–intron structures. ORFs of class (i) genes are determined by selecting the longest ORFs. This process is not required for classes (ii) and (iii) because they already contain ORF information. If no ORFs are found, those sequences are treated as non-protein-coding genes. Lastly, sequence similarity against SwissProt and TrEMBL proteins is examined with blastp in BLAST+ (Camacho et al. 2009), and InterProScan (Quevillon et al. 2005) is conducted to identify functional domains and assign GO terms to the ORFs.

**Visualization of annotation results**

After all the annotation procedures are completed on an application server, the results are returned back to a web server for graphical representation with GBrowse. In addition to the three classes of predicted genes that were previously mentioned, gene loci and repeat regions are displayed on the annotation map in GBrowse (Fig. 1C). MEGANTE also provides other data tracks as follows: (i) protein alignments of the SwissProt and TrEMBL plant division generated by ProSplign; (ii) intraspecies FLcDNA aligned by BLAT; (iii) EST assemblies generated by PASA; (iv) interspecies FLcDNA aligned by sim4db; and (v) repetitive elements detected by Repeat Masker. Users can select the tracks with the ‘Select Tracks’ tab in the window. Details of gene attributes and function annotation, such as top 10 BLAST hits to Swiss-Prot and TrEMBL, InterPro domains and GO terms, are linked from the predicted gene track (Fig. 1D). ORF and protein sequences can also be retrieved from the same page. For secure data management, we added authentication and authorization mechanisms into the original GBrowse so that user data are not disclosed to any others.

**Application to plant genome sequences**

To show the efficiency of MEGANTE, we applied this web service to genomic sequences from *Arabidopsis thaliana*, *Glycine max*, *Musa acuminate*, *Oryza sativa*, *Populus trichocarpa*, *Solanum lycopersicum*, *Musca domestica* and *Vitis vinifera*. Each sequence consisted of 1,000 fragments of a genome sequence, and each one contained one transcript sequence. Details of the data sets are described in the Materials and Methods. To evaluate the performance of MEGANTE, we examined the predictive accuracies for the coding sequence (CDS) coordinates by sensitivity (Sn) and specificity (Sp) that are commonly used for the evaluation of gene prediction programs (Pavy et al. 1999, Rogic et al. 2001, Yao et al. 2005). Sn is defined as the proportion of actual positives that are correctly predicted, and Sp is defined as the proportion of predicted positives that are true positives. We calculated the Sn and Sp at both the exon and gene levels. The exon
level means that the start and end positions of a CDS are checked at each exon. At gene level evaluation, it is necessary to identify all CDS coordinates in a transcript correctly. All of the results are summarized in Table 2. For comparison, the results of individual ab initio gene predictions employed in the system are also shown in the table. Differences in predictive accuracies can be observed among the species. For instance, approximately 80% of CDSs for A. thaliana and O. sativa were correctly identified, while the Sns at gene level for M. acuminate and M. domestica were much lower, approximately 10–20%. However, MEGANTE exhibited higher Sn and Sp in almost all categories in comparison with the ab initio gene finders.

Furthermore, we ran MEGANTE against 13 genome contigs from wheat chromosome 3B (Choulet et al. 2010), which were used in the evaluation of a wheat genome annotation pipeline, TriAnnot (Leroy et al. 2012). The overall size of the contigs is approximately 18 Mb, and they contain 172 CDSs. We used the same evaluation as previously described. The results revealed that the Sn and Sp were 77.6% and 88.2% at the exon level and 64.5% and 63.5% at the gene level, respectively. The results could not be directly compared with the values described in the study on TriAnnot because the numbers of contigs and genes used for the evaluation were not identical between these two. However, both the Sn and Sp of MEGANTE were comparable with those of TriAnnot.

**Discussion**

In this article, we introduced MEGANTE, a web service for integrated plant genome annotation. The interface of MEGANTE is designed mainly for non-bioinformatics researchers. Complex configurations for annotation procedures are not required; therefore, users can perform genome annotation simply by copying and pasting genomic sequences and selecting the species they want to query. Graphical representation is important for quickly interpreting the analysis results. We utilized GBrowse (Stein et al. 2002) for data visualization because this viewer is widely used in several plant genome databases (Goodstein et al. 2012, Lamesch et al. 2012, Sakai et al. 2013) and users should be familiar with its interface.

### Table 2: Predictive accuracies of MEGANTE and individual gene prediction programs used in the system

| Test gene sets         | Evaluation categories | MEGANTE | AUGUSTUS | GeneZilla | GlimmerHMM | SNAP |
|------------------------|-----------------------|---------|----------|-----------|------------|------|
| A. thaliana            | Exon level            | Sn (%)  | 95.2     | 84.4      | 71.8       | 80.9 | 75.5 |
|                        |                       | Sp (%)  | 87.9     | 75.6      | 66.5       | 72.7 | 59.2 |
|                        | Gene level            | Sn (%)  | 84.2     | 56.4      | 43.6       | 49.1 | 38.7 |
|                        |                       | Sp (%)  | 58.8     | 43.4      | 28.0       | 35.2 | 22.4 |
| G. max                 | Exon level            | Sn (%)  | 78.8     | 75.3      | 57.5       | 56.3 | 61.8 |
|                        |                       | Sp (%)  | 86.7     | 71.1      | 54.2       | 59.1 | 54.3 |
|                        | Gene level            | Sn (%)  | 51.4     | 35.9      | 22.1       | 28.2 | 18.2 |
|                        |                       | Sp (%)  | 48.8     | 32.5      | 13.6       | 16.0 | 12.1 |
| M. acuminate           | Exon level            | Sn (%)  | 46.1     | 48.3      | 19.9       | 34.7 | 30.5 |
|                        |                       | Sp (%)  | 62.3     | 54.4      | 28.7       | 31.5 | 36.4 |
|                        | Gene level            | Sn (%)  | 12.8     | 12.2      | 5.9        | 7.6  | 5.8  |
|                        |                       | Sp (%)  | 12.7     | 11.2      | 4.1        | 3.4  | 4.0  |
| O. sativa              | Exon level            | Sn (%)  | 91.9     | 52.9      | 57.0       | 74.0 | 45.4 |
|                        |                       | Sp (%)  | 86.7     | 67.4      | 46.4       | 59.0 | 50.6 |
|                        | Gene level            | Sn (%)  | 78.0     | 29.5      | 21.9       | 37.8 | 19.7 |
|                        |                       | Sp (%)  | 57.1     | 29.3      | 12.2       | 21.6 | 15.6 |
| P. trichocarpa         | Exon level            | Sn (%)  | 76.7     | 73.7      | 60.8       | 55.0 | 63.4 |
|                        |                       | Sp (%)  | 81.1     | 71.0      | 57.8       | 59.3 | 57.4 |
|                        | Gene level            | Sn (%)  | 32.3     | 26.8      | 19.6       | 21.4 | 14.4 |
|                        |                       | Sp (%)  | 32.0     | 25.8      | 13.4       | 13.2 | 10.7 |
| S. lycopersicum        | Exon level            | Sn (%)  | 85.7     | 69.4      | 49.0       | 49.3 | 57.0 |
|                        |                       | Sp (%)  | 91.1     | 74.4      | 49.1       | 52.4 | 49.3 |
|                        | Gene level            | Sn (%)  | 62.6     | 29.5      | 22.0       | 26.8 | 19.3 |
|                        |                       | Sp (%)  | 60.0     | 34.0      | 13.0       | 14.5 | 12.3 |
| M. × domestica         | Exon level            | Sn (%)  | 59.0     | 59.1      | 47.1       | 49.8 | 42.3 |
|                        |                       | Sp (%)  | 68.5     | 62.5      | 47.3       | 50.1 | 46.1 |
|                        | Gene level            | Sn (%)  | 22.1     | 22.5      | 13.2       | 19.1 | 11.2 |
|                        |                       | Sp (%)  | 19.0     | 18.4      | 7.7        | 9.7  | 7.8  |
| V. vinifera            | Exon level            | Sn (%)  | 61.0     | 51.4      | 46.8       | 35.8 | 41.7 |
|                        |                       | Sp (%)  | 83.7     | 51.2      | 38.5       | 31.0 | 36.0 |
|                        | Gene level            | Sn (%)  | 22.7     | 10.5      | 7.1        | 6.6  | 5.0  |
|                        |                       | Sp (%)  | 27.3     | 7.9       | 3.5        | 2.6  | 2.6  |

Gene prediction parameters we used for each target species are described in the Materials and Methods.
MEGANTE has unique features that are not found in similar services. For instance, the service is able to accept a query sequence with a length of 10 Mb, which is larger than the other services. Another prominent feature of MEGANTE is that it targets a wide variety of plant species including 24 species from eight families. This was made possible by adapting common parameter sets for gene structure prediction for all species in the same family. For example, the system creates consensus gene structures for the Poaceae family by using JIGSAW (Allen et al. 2006) with a parameter matrix generated from a reference gene set from *O. sativa*. Although it is generally preferable to optimize gene prediction parameters for a particular species, the number of reference genes with high-quality annotation is not large enough for parameter optimization, and enriched annotation of closely related species shows much better performance for this application. In fact, our evaluation of wheat genome sequence annotation using MEGANTE revealed that the prediction parameters for *O. sativa* were sufficient for wheat in our annotation workflow.

One important point to be considered is the updates of data, including reference databases and parameter files for gene prediction in the system. We plan to update the transcript and protein databases at least twice a year, and regenerate gene prediction parameters when new reference annotation data are released. Most of the methods used to generate parameter files for gene prediction are automated in the system, and thus the overall procedure for one species can be completed within a week. Furthermore, it is possible to adapt MEGANTE to any other species by collecting transcript sequences or optimizing gene prediction parameters with reference annotation data for the species of interest.

**Materials and Methods**

**Compiling reference gene sets for gene prediction**

To optimize gene prediction parameters for each species, annotated genes are required as reference gene sets. We use the term [training] to refer to the optimization. We collected *A. thaliana* gene sets from the Brassicaceae family, *G. max* from Fabaceae, *M. acuminata* from Musaceae, *O. sativa* from Poaceae, *P. trichocarpa* from Salicaceae, *S. lycopersicum* from Solanaceae, *M. domestica* from Rosaceae, and *V. vinifera* from Vitaceae. The system uses the same parameter sets for gene prediction of all species in the same family, while transcript sequences used for alignment are distinct from each other. The *A. thaliana* gene set was retrieved from representative genes in TAIR10 (Lamesch et al. 2012); *G. max* and *P. trichocarpa* from Phytozone v9.0 (Goodstein et al. 2012); *M. acuminata* from The Banana Genome Hub version 1 (Droc et al. 2013); *O. sativa* from representative genes in RAP-DB IRGSP 1.0 (Sakai et al. 2013); *S. lycopersicum* from ITAG2.3 in SGN (Bombarely et al. 2011); *M. domestica* from v1.0p assembly and annotation in GDR (Jung et al. 2008); and *V. vinifera* from the 12X version of genome assembly and annotation in Grape Genome Browser (Jaillon et al. 2007). First, we excluded genes that did not begin with an initiation codon or did not end with a stop codon in each gene set. Then, we randomly selected 1,000 genes for training of ab initio gene prediction programs, 10,000 for training of JIGSAW and 1,000 for an evaluation of the overall performance of MEGANTE. The data sets for training and evaluation did not overlap with each other so that valid evaluation between independent sets was possible. All of the gene sequences extracted from genome assemblies contain CDSs and their 1 kb upstream and downstream sequences.

**Training of gene prediction programs**

We initially trained ab initio gene finders, AUGUSTUS (Stanke and Waack 2003), GeneZilla (Allen et al. 2006), GlimmerHMM (Allen et al. 2006) and SNAP (Korf 2004), with each gene set. However, the gene finders had pre-trained parameter files for some plant species; thus, we did not train the programs for those species. The pre-trained parameters we used were AUGUSTUS for Arabidopsis, maize and tomato; GlimmerHMM for Arabidopsis and rice; and SNAP for Arabidopsis and rice. AUGUSTUS for maize is used for the *Poaceae* family. For GeneZilla, we employed an automatic training program, GRAPE (Majoros and Salzberg 2004). To train the other programs, we followed the instructions provided by the software developer. Subsequently, training of JIGSAW with all of the ab initio gene finders was conducted using 10,000 CDSs. In addition, interspecies FLcDNA alignment with sim4db and protein alignment with ProSplign were also conducted against the same data sets. These procedures were the same as previously described for the annotation workflow. All the results were fed to JIGSAW as sources of evidence. The training of JIGSAW was performed by the train_jigsaw.pl script in the package with default options.

**Availability and implementation of the system**

MEGANTE was implemented in a web application framework, Catalyst, with MySQL as the backend database. The frontend web interface was built with HTML5, CSS3 and JavaScript, and was tested on the following web browsers: Safari 6, Chrome 28, Firefox 23 and Internet Explorer 9 and 10. This service is available for free at https://megante.dna.affrc.go.jp/.

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**Disclosures**

The authors have no conflicts of interest to declare.
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