Low Serum Neutrophil Gelatinase-associated Lipocalin Level as a Marker of Malnutrition in Maintenance Hemodialysis Patients.

AUTHOR(S):
Imamaki, Hirotaka; Ishii, Akira; Yokoi, Hideki; Kasahara, Masato; Kuwabara, Takashige; Mori, Keita P; Kato, Yukiko; ... Mukoyama, Masashi; Yanagita, Motoko; Mori, Kiyoshi

CITATION:
Imamaki, Hirotaka ...[et al]. Low Serum Neutrophil Gelatinase-associated Lipocalin Level as a Marker of Malnutrition in Maintenance Hemodialysis Patients.. PloS one 2015, 10(7): e0132539.

ISSUE DATE:
2015-07-10

URL:
http://hdl.handle.net/2433/202581

RIGHT:
© 2015 Imamaki et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.
RESEARCH ARTICLE

Low Serum Neutrophil Gelatinase-associated Lipocalin Level as a Marker of Malnutrition in Maintenance Hemodialysis Patients

Hirotaka Imamaki1, Akira Ishii1, Hideki Yokoi1, Masato Kasahara2, Takashige Kuwabara1,9, Keita P. Mori1, Yukiko Kato1, Takashi Kuwahara3, Masugi Satoh4, Kimihiko Nakatani5, Yoshihiko Saito5, Naohisa Tomosugi6, Akira Sugawara7, Kazuwa Nakao6, Masashi Mukoyama1,9, Motoko Yanagita1,10, Kiyoshi Mori10*

1 Department of Nephrology, Kyoto University Graduate School of Medicine, Kyoto, Japan, 2 Institute for Advancement of Clinical and Translational Science, Kyoto University Hospital, Kyoto, Japan, 3 Department of Nephrology, Saiseikai Ibaraki Hospital, Osaka, Japan, 4 Misugikai Satou Hospital, Osaka, Japan, 5 First Department of Internal Medicine, Nara Medical University, Nara, Japan, 6 Division of Advanced Medicine, Medical Research Institute, Kanazawa Medical University, Kanazawa, Japan, 7 Department of Nephrology, Osaka Red Cross Hospital, Osaka, Japan, 8 TK Project, Medical Innovation Center, Kyoto University Graduate School of Medicine, Kyoto, Japan, 9 Department of Nephrology, Kumamoto University Graduate School of Medical Sciences, Kumamoto, Japan, 10 TMK Project, Medical Innovation Center, Kyoto University Graduate School of Medicine, Kyoto, Japan

* mori@tmk.med.kyoto-u.ac.jp

Abstract

Background
Neutrophil gelatinase-associated lipocalin (NGAL or LCN2) is an iron-transporting factor which possesses various activities such as amelioration of kidney injury and host defense against pathogens. Its circulating concentrations are elevated in acute and chronic kidney diseases and show a positive correlation with poor renal outcome and mortality, but its clinical significance in maintenance hemodialysis (HD) patients remains elusive.

Methods
Serum NGAL levels were determined by enzyme-linked immunosorbent assay in outpatient, Japanese HD subjects. Their correlation to laboratory findings and morbidity (as development of severe infection or serum albumin reduction) was investigated using linear regression analysis and \( \chi^2 \) test.

Results
Pre-dialysis serum NGAL levels in HD patients were elevated by 13-fold compared to healthy subjects (n=8, \( P<0.001 \)). In a cross-sectional study of 139 cases, serum NGAL concentrations were determined independently by % creatinine generation rate (an indicator of muscle mass, standardized coefficient \( \beta=0.40, P<0.001 \)), peripheral blood neutrophil count (\( \beta=0.38, P<0.001 \)) and anion gap (which likely reflects dietary protein intake, \( \beta=0.16, P<0.05 \)). Iron administration to anemic HD patients caused marked elevation of peripheral
blood hemoglobin, serum ferritin and iron-regulatory hormone hepcidin-25 levels, but NGAL levels were not affected. In a prospective study of 87 cases, increase in serum albumin levels a year later was positively associated to baseline NGAL levels by univariate analysis ($r=0.36, P<0.01$). Furthermore, within a year, patients with the lowest NGAL tertile showed significantly increased risk for marked decline in serum albumin levels ($>0.4$ g/dl; odds ratio 5.5, 95% confidence interval 1.5–20.3, $P<0.05$) and tendency of increased occurrence of severe infection requiring admission (odds ratio 3.1, not significant) compared to the middle and highest tertiles.

**Conclusion**

Low serum NGAL levels appear to be associated with current malnutrition and also its progressive worsening in maintenance HD patients.
patients [24, 25]. We also investigated whether serum NGAL levels are altered by iron administration, HD session and passage of blood through the kidneys.

**Results**

**Baseline characteristics of patients**

Baseline characteristics of 139 out-patient, Asian-Japanese, maintenance HD subjects are shown in Table 1, S1 and S2 Tables. Clinical parameters studied include routine laboratory data and indices of nutrition, iron status and HD efficiency. Pre-dialysis serum NGAL levels in HD patients (mean±SD, 916±345 ng/ml) were elevated approximately by 13-fold compared to healthy subjects (68±15 ng/ml, n = 8, P<0.001). When patients were divided into tertiles of serum NGAL levels, several clinical parameters showed significant difference among groups (Table 1), which were further characterized below. Gender and presence of diabetes mellitus (DM) did not significantly affect serum NGAL levels (male, 899±354, n = 79; female, 937±334, n = 60; cases with DM, 841±291, n = 38; without DM, 944±360 ng/ml, n = 101).

**Univariate analysis of serum NGAL levels**

In a cross-sectional study, we examined association of baseline serum NGAL levels with above mentioned parameters, using univariate linear regression analysis (S3 Table). Among the parameters examined, peripheral blood neutrophil count, white blood cell (WBC) count, anion gap, serum creatinine and % creatinine generation rate (%CGR) exhibited the strongest correlation coefficients (r) with serum NGAL levels, which were >0.48, respectively (P<0.001, Table 2). None of indices for iron homeostasis, including red blood cell count, blood hemoglobin, hematocrit, reticulocyte count, serum ferritin, iron, total iron binding capacity, unsaturated iron binding capacity and transferrin saturation were significantly associated with NGAL levels (P>C21, respectively, S3 Table), suggesting that systemic iron abundance does not affect serum NGAL levels.

**Multivariate analysis of serum NGAL levels**

To carry out multivariate linear regression analysis for baseline serum NGAL levels, we entered not only age and gender but also all the above 5 variables tightly correlated to serum NGAL levels, since it was difficult to pick up representative variables among them. Through this analysis, %CGR, neutrophil count and anion gap were selected as independent variables determining serum NGAL levels (Table 3, Fig 1A–1C).

Next, we investigated clinical implication of the above findings (Fig 1 and S4 Table). Blood neutrophil count showed strong correlation with WBC count (r = 0.92, P<0.001) and serum CRP level (r = 0.35, P<0.001), suggesting that neutrophil count is a marker of bone marrow myeloid activity and inflammatory status (S4 Table, Fig 1D). %CGR, which is known as an indicator of muscle mass [23], exhibited the strongest correlation to serum creatinine (r = 0.84, P<0.001). Anion gap and normalized protein catabolic rate (nPCR) have been reported to be highly correlated to dietary protein intake in HD patients [22, 26], and they were associated well to each other (r = 0.50, P<0.001)[26]. Furthermore, serum NGAL levels were significantly correlated to other markers of nutrition such as serum albumin (r = 0.29, P<0.001), choline esterase (r = 0.23, P = 0.01) and phosphorus levels (r = 0.29, P = 0.001) (Table 2). These findings suggest that muscle mass, protein intake, myeloid activity and inflammatory status are the major determinants of serum NGAL levels in steady-state maintenance HD patients.
Table 1. Baseline characteristics of 139 maintenance HD patients in correlation with tertiles of serum NGAL concentrations.

| Baseline characteristics | Pre-dialysis NGAL tertile |   |   |   |
|-------------------------|---------------------------|---|---|---|
|                         | Low (ng/ml) | Medium (ng/ml) | High (ng/ml) | P for trend |
| N                       | 46 (500–663) | 47 (826–943) | 46 (1130–1423) | <0.001 |
| Pre-dialysis NGAL (ng/ml) | 583 (331–490) | 639 (513–773) | 878 (768–1013) | <0.001 |
| NGAL removal ratio (%) | 24±16 | 28±17 | 29±18 | 0.37 |
| Age (year) | 69±10 | 64±13 | 62±11 | 0.02 |
| Male gender [n (%)] | 31 (67%) | 20 (42%) | 28 (60%) | <0.001 |
| Diabetes [n (%)] | 14 (30%) | 14 (30%) | 10 (22%) | 0.34 |
| HD period (month) | 29 (13–68) | 74 (42–140) | 102 (53–156) | <0.001 |
| Na (mEq/l) | 140±3 | 139±3 | 138±3 | <0.001 |
| Cl (mEq/l) | 105±4 | 103±3 | 102±4 | <0.001 |
| K (mEq/l) | 4.8±0.9 | 5.0±0.7 | 5.0±0.8 | 0.37 |
| Ca (mg/dl) | 8.5±0.9 | 8.7±0.9 | 8.9±0.8 | 0.04 |
| Normalized Ca (mg/dl) | 8.9±0.8 | 9.1±0.8 | 9.1±0.7 | 0.25 |
| Phosphorus (mg/dl) | 4.8±1.4 | 5.2±1.4 | 5.8±1.8 | 0.01 |
| Intact PTH (pg/ml) | 162 (97–232) | 183 (92–287) | 215 (81–336) | 0.19 |
| BUN (mg/dl) | 61±16 | 65±13 | 72±17 | 0.01 |
| Creatinine (mg/dl) | 9.1±2.4 | 11.0±2.4 | 12.2±2.6 | <0.001 |
| UA (mg/dl) | 6.4±1.4 | 6.7±1.2 | 7.2±1.3 | 0.03 |
| Total protein (g/dl) | 6.2±0.4 | 6.4±0.5 | 6.6±0.4 | <0.001 |
| Albumin (g/dl) | 3.7±0.4 | 3.7±0.4 | 3.9±0.3 | <0.01 |
| Choline esterase (IU/l) | 197±55 | 219±58 | 227 ±71 | 0.06 |
| Total cholesterol (mg/dl) | 146±33 | 156±30 | 164±50 | 0.06 |
| Triglyceride (mg/dl) | 76 (63–101) | 96 (63–172) | 106 (78–146) | 0.05 |
| CRP (mg/dl) | 0.10 (0.05–0.37) | 0.10 (0.05–0.26) | 0.12 (0.07–0.30) | 0.21 |
| Base excess (mEq/l) | -3.9±3.3 | -4.4±3.0 | -6.4±3.7 | <0.01 |
| Anion gap (mEq/l) | 19.7±2.3 | 20.4±2.7 | 22.4±3.0 | <0.001 |
| White blood cell (/μl) | 491±1456 | 534±1639 | 714±2054 | <0.001 |
| Neutrophil (/μl) | 3187±1108 | 3691±1246 | 5109±1892 | <0.001 |
| Red blood cell (x10^6/μl) | 3.3±0.56 | 3.42±0.50 | 3.51±0.47 | 0.28 |
| Hemoglobin (g/dl) | 10.2±1.4 | 10.0±1.4 | 10.7±1.4 | 0.03 |
| Hematocrit (%) | 32.0 (29.9–35.2) | 31.9 (28.3–34.7) | 33.9 (30.8–35.9) | 0.31 |
| Platelet (x10^9/μl) | 152±47 | 175±63 | 197±59 | <0.01 |
| Ferritin (ng/ml) | 126 (43–225) | 80 (33–184) | 148 (52–120) | 0.18 |
| Iron (mg/dl) | 68±31 | 65±28 | 63±38 | 0.70 |
| TIBC (mg/dl) | 235±33 | 258±49 | 248±57 | 0.08 |
| UIBC (mg/dl) | 168±45 | 191±64 | 184±63 | 0.15 |
| TSAT (%) | 29.0±14.6 | 26.2±13.1 | 26.4±14.6 | 0.53 |
| %CGR (%) | 81±27 | 106±23 | 110±24 | <0.001 |
| nPCR (g/kg/day) | 0.83±0.20 | 0.91±0.15 | 0.96±0.20 | <0.001 |
| Kt/Vsp | 1.31±0.32 | 1.49±0.29 | 1.41±0.29 | 0.02 |

Biochemical analysis was carried out using pre-dialysis serum, unless specified. See **S1 Table** for other clinical parameters. Values are expressed as means±SD, median (interquartile range) or number (%). Trend for differences among serum NGAL tertiles was compared by ANOVA for continuous variables and by Kruskal-Wallis test for dichotomous and non-normal variables, respectively. PTH, parathyroid hormone; BUN, blood urea nitrogen; UA, uric acid; CRP, C-reactive protein; TIBC, total iron binding capacity; UIBC, unsaturated iron binding capacity; TSAT, transferrin saturation; CGR, creatinine generation rate; nPCR, normalized protein catabolic rate; Kt/Vsp, single-pool Kt/V.

doi:10.1371/journal.pone.0132539.001
Impact of HD efficiency, HD period and age upon serum NGAL levels

Since serum NGAL levels were positively correlated to serum creatinine levels (r = 0.51, P < 0.001) and HD period (r = 0.47, P < 0.001), we examined whether elevated NGAL was caused by low dialysis efficiency (Table 2, Fig 2A and 2B). Oppositely, HD efficiency calculated as single-pool Kt/V (Kt/Vsp) was positively associated with NGAL levels (r = 0.20, P < 0.05, Fig 2C). Furthermore, serum NGAL levels were negatively correlated to age (r = -0.29, P < 0.001).

### Table 2. Clinical parameters showing significant correlations with baseline pre-dialysis serum NGAL levels by univariate linear regression analysis.

| Parameter         | r    | P     |
|-------------------|------|-------|
| Neutrophil        | 0.53 | < 0.001 |
| White blood cell  | 0.52 | < 0.001 |
| Anion gap         | 0.51 | < 0.001 |
| Creatinine        | 0.51 | < 0.001 |
| %CGR              | 0.49 | < 0.001 |
| HD period         | 0.47 | < 0.001 |
| pH                | -0.37 | < 0.001 |
| Platelet          | 0.35 | < 0.001 |
| Na                | -0.33 | < 0.001 |
| nPCR              | 0.33 | < 0.001 |
| BUN               | 0.33 | < 0.001 |
| Cl                | -0.32 | < 0.001 |
| Base excess       | -0.32 | < 0.001 |
| Albumin           | 0.29 | < 0.001 |
| Age               | -0.29 | < 0.001 |
| Phosphorus        | 0.29 | 0.001 |
| Total protein     | 0.29 | 0.001 |
| HCO₃⁻             | -0.28 | 0.001 |
| Ca                | 0.27 | 0.002 |
| UA                | 0.26 | 0.003 |
| Choline esterase  | 0.23 | 0.01 |
| Normalized Ca     | 0.22 | 0.01 |
| Total cholesterol | 0.20 | 0.02 |
| Kt/Vsp            | 0.20 | 0.02 |

Among clinical parameters shown in Table 1, parameters which had significant correlations to serum NGAL levels are shown (P < 0.05, respectively). N = 139. Correlations of the rest of parameters are shown in S3 Table.

doi:10.1371/journal.pone.0132539.t002

### Table 3. Determinants of baseline serum NGAL levels by multivariate linear regression analysis.

| Variable   | Standardized coefficient (β) | P     |
|------------|-----------------------------|-------|
| %CGR       | 0.40                        | <0.001 |
| Neutrophil | 0.38                        | <0.001 |
| Anion gap  | 0.16                        | <0.05  |

The above 3 variables explained 42% of the expected NGAL value (adjusted r²). Variables entered but excluded by stepwise analysis were age, gender, serum creatinine, and blood WBC count. N = 139.

doi:10.1371/journal.pone.0132539.t003
Fig 1. Linear correlation of baseline pre-dialysis serum NGAL levels with clinical parameters. Correlations of serum NGAL level with (A) peripheral blood neutrophil count, (B) % creatinine generation rate (%CGR) and (C) anion gap are shown. (D) Correlation between neutrophil count and serum C-reactive protein (CRP) level is also shown. Linear line in each graph shows regression line. N = 139.

doi:10.1371/journal.pone.0132539.g001

Fig 2. Linear correlation of baseline pre-dialysis serum NGAL levels with serum creatinine, dialysis efficiency, age and HD period. Correlations with (A) creatinine (Cr), (B) HD period, (C) single-pool Kt/V (Kt/Vsp), and (D) age are shown. (E) Correlation between HD period and age. N = 139.

doi:10.1371/journal.pone.0132539.g002
Fig 2D) and age was negatively correlated to HD period (r = -0.28, P = 0.001, Fig 2E). These findings suggest that patients with long HD history had elevated serum NGAL levels because they contained relatively younger subjects whose nutritional conditions were good.

Impact of iron administration upon serum NGAL

To directly test whether iron homeostasis is linked to NGAL regulation, we examined the effects of iron administration in 6 anemic HD patients who had low serum ferritin levels (33 ±20 ng/ml). With 10 doses of 50 mg intravenous iron injection (twice a week for 5 weeks), blood hemoglobin, serum ferritin and hepcidin-25 levels were significantly elevated at 3 or 8 weeks (Fig 3, S5 Table). Hepcidin-25 is a circulating hormone which inhibits intestinal absorption of iron and release of iron from macrophages [5, 27–29]. By contrast, serum NGAL levels were not affected during the observation period, indicating that abundance of systemic iron does not alter serum NGAL levels in HD patients.

NGAL clearance by HD session

NGAL is a secretory protein whose molecular weight (MW) is 25 kilo dalton (Da) as a monomer, but it may exist in larger molecular forms, such as NGAL:matrix metalloproteinase-9 (MMP-9 or gelatinase B) heterodimer (MW 130 kDa) [1]. We examined whether circulating NGAL and other substances are removed by HD procedure (S6 Table). As shown in Fig 4A, serum NGAL level showed a rapid decrease at 10 min after initiation of HD (by 15±9%, n = 9, not significantly), which may have been caused by adsorption of NGAL to HD circuit and dialysis membrane, and it slowly decreased during the rest of time in HD. At the end of HD, NGAL levels were decreased by 27±17% (n = 139, P <0.001 comparing before and after HD; Fig 4A, Table 1). Since blood neutrophils are one of important sources of circulating NGAL [1, 30], we investigated the kinetics of blood WBC and neutrophil counts (n = 9), and observed temporal drop at 10 min after initiation of HD but it recovered to baseline by 60 min [31], indicating that neutrophil count cannot simply explain the alteration of serum NGAL levels during HD session (Fig 4B). In comparison, the arterial blood concentrations of low MW substances such as potassium ion (39 Da), BUN (60 Da), creatinine (113 Da), and β2-microglobulin (12 kDa) decreased constantly during HD session (Fig 4B). On the other hand, albumin (60 kDa) and IgG (150 kDa) levels increased slightly as water removal proceeded during HD. The kinetics of arterial blood NGAL levels were intermediate of low and high MW substances, suggesting that NGAL is constantly removed during HD but by low efficiency.

As described above, serum NGAL levels were decreased by 27% on average after HD session, but NGAL levels were rather increased in 7% of HD patients: their NGAL removal ratios were -6.2±5.4%, n = 10). NGAL removal ratio was not significantly correlated to pre-dialysis NGAL (r = 0.11, P = 0.19, n = 139) but significantly correlated to post-dialysis NGAL levels (r = -0.44, P <0.001). Univariate analysis indicated that serum HCO₃⁻ (r = 0.37, P <0.001), base excess (r = 0.35, P <0.001), Kt/Vsp (r = 0.35, P <0.001), pCO₂ (r = 0.30, P <0.001) and pH (r = 0.20, P <0.05) show significant correlation to NGAL removal ratios. On the other hand, unlike pre-dialysis NGAL levels, NGAL removal ratios were not significantly correlated to % CGR (r = -0.01), neutrophil count (r = 0.12) or anion gap (r = 0.00; P >0.05, respectively). These findings suggest that metabolic acidosis (low HCO₃⁻, low base excess and low pH), secondary respiratory alkalosis (low pCO₂) and low HD efficiency potently inhibit reduction in serum NGAL levels during HD session, independently from pre-dialysis serum NGAL levels.
NGAL clearance by renal circulation

To study whether circulating NGAL is removed during renal passage, we collected plasma (instead of serum) from aorta and renal vein in a separate group of 30 patients who underwent coronary angiography (S7 Table). As shown in Table 4, in 15 patients not receiving HD, plasma creatinine levels in renal vein were 19±10% less compared to those in aorta (P<0.001), and plasma NGAL levels in renal vein were 13±12% less as compared to aorta (P<0.001). On the other hand, in 15 maintenance HD subjects, plasma creatinine and NGAL levels were not different between renal vein and aorta, suggesting that appreciable amount of NGAL was neither removed at nor secreted from kidneys with end-stage renal disease. As negative controls, plasma albumin (60 kDa) and choline esterase (340 kDa) [32] concentrations in aorta and renal vein were similar both in non-HD and HD subjects. These findings suggest that circulating NGAL is partially eliminated during renal passage in subjects with preserved renal function, likely by glomerular filtration [6], but not in patients with end-stage renal disease.

Prospective association of baseline NGAL level with serum albumin reduction and development of infection

Findings so far indicate that HD patients with low serum NGAL concentrations are characterized to have reduced muscle mass, low protein intake and mild neutropenia (Tables 1 and 3), implying morbid and infection-prone prognosis. To further elucidate clinical significance of serum NGAL concentrations in maintenance HD patients, we longitudinally investigated development of severe infection (requiring admission) and changes in serum albumin levels during a year of observation period among 95 patients in two of the three dialysis centers.

Within a year, two patients died due to congestive heart failure and lung cancer, respectively, who belonged to the middle tertile group of baseline NGAL levels (Table 5). Follow-up data could not be obtained in 6 cases (S1 Fig). Of the rest of 87 patients (S8 Table), 30 subjects (34%) underwent one to three admissions within a year. In 7 cases, treatment of infectious disease was the major purpose, at least, in one admission (among them, 4 cases belonged to the lowest NGAL tertile). Pathogens identified were Candida albicans (causing pneumonia), Campylobacter fetus (sepsis), Enterobacter aeruginosa (pyelonephritis), Enterobacter cloacae.
Fig 4. Removal of circulating substances by HD session. Blood was serially drawn from the arterial and venous ends of HD circuit (closed circle and open box, respectively) during HD session. End points varied from 3.0 to 4.5 h. Mean±SD. (A) Comparison of arterial blood NGAL levels between before and after HD session (left, n = 139). Time course of arterial and venous NGAL levels are also shown (right, n = 9). (B) Serum levels of K, BUN, creatinine (Cr), β2-microglobulin (MG), albumin (Alb) and IgG, as well as blood WBC and neutrophil counts are shown (n = 9). Arterial levels of K, BUN, Cr, β2-MG were always higher than venous levels, indicating active removal by HD. On the other hand, venous levels of albumin and IgG tended to be higher than arterial levels, reflecting hemoconcentration by HD. *P<0.05, significant difference from time 0 by repeated-measures ANOVA. **P<0.05, significant difference between arterial and venous blood.

doi:10.1371/journal.pone.0132539.g004

(sepsis), *Pseudomonas aeruginosa* (pneumonia) and *Streptococcus aureus* (lower extremity gangrene). In subjects with the lowest NGAL tertile, odds ratio (OR) [95% confidence interval] for development of severe infection requiring admission was 3.1 [0.6–15.0] compared to the middle and highest tertiles, but it did not reach statistical significance (S9 Table). Presence of diabetes mellitus also showed a tendency to be associated with increased occurrence of severe infection, but again not significantly (S9 Table). On the other hand, OR for admission due to
non-infectious causes (23 cases, 26%) was not elevated in the lowest NGAL tertile group: 0.7
[0.2–2.0]. Non-infectious causes included diagnosis and treatment of malignancy (n = 5), car-
diovascular (14), pulmonary (1), and gastrointestinal disorders (3), fixation of internal shunt
(6), and ophthalmic (4) or orthopedic surgery (2), including overlaps. These findings revealed
that HD patients with reduced serum NGAL concentrations show a tendency to develop severe
infection.

Next, we studied correlation of baseline serum NGAL levels with changes in serum albumin
levels after a year. As shown in Fig 5, serum albumin levels were stable in the highest and mid-
dle NGAL tertiles. In the lowest NGAL tertile, serum albumin levels showed a significant
decrease by 0.22±0.17 g/dl (P < 0.05). Consistently, when we defined marked serum albumin
decline by ≥0.4 g/dl per year (which occurred in 12 cases), its OR was 5.5 [1.5–20.3, P < 0.05]
in the lowest NGAL tertile group compared to the middle and highest tertile groups (Table 6).
Event numbers were too little to carry out multivariate analysis for severe infection or marked
albumin loss.

We further evaluated relative importance of baseline serum NGAL levels among various
indices of nutritional condition (Table 7). By univariate analysis, follow-up albumin levels were
positively correlated to numerous nutritional markers such as baseline NGAL, albumin, geriat-
cric nutritional risk index (GNRI) [33, 34], creatinine, %CGR, anion gap, choline esterase, tri-
glyceride levels and neutrophil counts, and negatively to age. If not completely, similar findings

Table 4. Comparison of plasma levels of markers in aorta and renal vein among patients who underwent coronary angiography.

|                     | Creatinine (mg/dl) | NGAL (ng/ml) | Albumin (g/dl) | Choline esterase (IU/l) |
|---------------------|-------------------|--------------|---------------|------------------------|
|                     | non-HD            | HD           | non-HD        | HD                     | non-HD | HD                     |
| Aorta               | 0.90±0.47         | 7.60±1.62    | 92±62         | 765±183                | 4.0±0.5 | 3.7±0.3                | 274±69 | 216±47                |
| Renal vein          | 0.73±0.39*        | 7.43±1.60    | 81±59*        | 768±213                | 4.0±0.5 | 3.6±0.3                | 275±69 | 212±43                |
| (mean of relative levels) | (81±10%)*         | (98±4%)      | (87±12%)*     | (100±10%)              | (100±4%) | (99±6%)                | (100±2%) | (99±5%)              |

Blood was collected from aorta and renal vein during coronary angiography in 15 patients not receiving HD (non-HD) and 15 receiving maintenance HD (HD).

*P<0.001 between aorta and renal vein.

Table 5. Clinical course of HD patients during a year of follow-up.

| Baseline NGAL          | Lowest (n = 32) | Middle and Highest (n = 63) | Sum (n = 95) |
|------------------------|-----------------|----------------------------|--------------|
| Death                  | 0               | 2                          | 2            |
| Lost follow-up         | 3               | 3                          | 6            |
| Follow-up available    | 29              | 58                         | 87           |
| Albumin loss           | 8               | 4                          | 12           |
| Severe infection       | 4               | 3                          | 7            |
| Admission without infection | 6               | 17                         | 23           |
| No admission           | 19              | 38                         | 57           |

Albumin Loss, reduction in serum albumin levels by 0.4 g/dl or larger after a year. Severe infection, occurrence of at least one admission within a year, in which infection was the major cause. Admission without infection, one to three times of admissions within a year, among which infection was never the major cause.
were obtained for baseline albumin levels. Correlation of NGAL with follow-up or baseline albumin levels was not necessarily stronger compared to other nutritional markers. On the other hand, albumin increase was positively correlated to baseline NGAL and neutrophil but negatively correlated to baseline albumin levels. Importantly, correlation of NGAL with albumin increase \((r = 0.36, P < 0.01)\) was the strongest among markers studied in this study, suggesting that serum NGAL levels may be especially useful for the prediction of alteration in nutritional conditions among HD patients. These findings elucidated that HD patients with reduced serum NGAL levels may have increased risk to develop severe infection and large reduction in serum albumin concentrations.

Subanalysis after exclusion of patients with inflammatory disorders

Since inflammation, infection and malignancy have been recently reported to be associated with elevated circulating NGAL levels \([9, 14, 15, 35]\), we performed a subanalysis by excluding such patients (S2 Table, S1 Fig). We excluded 31 subjects who had either infection (under treatment for mycobacterium tuberculosis, \(n = 3\)), malignancy (lung, colon, prostate, tonsill cancer, multiple myeloma or malignant lymphoma; \(n = 6\)) or elevated serum CRP levels (>0.5 mg/dl).

Table 6. Prediction of marked serum albumin reduction in a year.

| Variable     | Odds ratio | 95% CI       | P    |
|--------------|------------|--------------|------|
| Low NGAL     | 5.5        | 1.5–20.3     | 0.02 |
| DM           | 2.2        | 0.6–8.3      | 0.26 |
| Male         | 2.2        | 0.6–7.8      | 0.23 |

Marked serum albumin reduction was defined as ≥0.4g/dl. Low NGAL, lowest serum NGAL tertile at baseline. DM, presence of diabetes mellitus. CI, confidence interval. \(N = 87\).
n = 26), with some overlaps. Mean serum NGAL level of excluded cases (990±454 ng/ml, n = 31) were slightly (11%, P = 0.17) higher compared to the rest of patients included in subanalysis (894±305 ng/ml, n = 108). Multivariate linear regression analysis showed that baseline serum NGAL levels were determined by %CGR ($\beta = 0.50$, $P < 0.001$) and neutrophil count ($\beta = 0.36$, $P < 0.001$). Furthermore, the lowest NGAL tertile exhibited OR of 7.1 [1.6–29.8, $P < 0.01$, $n = 73$] for marked reduction in serum albumin levels in a year, and OR of 2.9 [0.6–13.9, $P = 0.22$, $n = 73$] for development of severe infection requiring admission within a year, as compared to the middle and highest NGAL tertiles. Serum albumin increase after a year in these cases was significantly associated with baseline NGAL levels ($r = 0.37$, $P < 0.01$, $n = 73$) and neutrophil counts ($r = 0.27$, $P < 0.05$). These findings show that excluding patients in inflammatory status did not basically change determinants of NGAL levels and clinical outcomes associated with low NGAL concentrations in our study.

**Discussion**

In the present study, using univariate and multivariate analyses, we have shown that serum NGAL levels in out-patient, Japanese, maintenance HD subjects were independently determined by muscle mass (represented by %CGR), nutritional status (by anion gap) and peripheral blood neutrophil counts (which presumably reflect bone marrow myeloid activity and inflammatory status). Furthermore, after one year of observation, patients in the lowest tertile of baseline serum NGAL levels were at a significant risk to experience marked decline in serum albumin levels and showed an increased tendency to develop severe infection, as compared to

| Baseline variable | Baseline albumin | Follow-up albumin | Albumin increase |
|-------------------|------------------|------------------|------------------|
|                   | $r$   | $P$   | $r$   | $P$   | $r$   | $P$   |
| NGAL              | 0.19  | 0.09  | 0.41  | $< 0.001$ | 0.36  | $< 0.01$ |
| Albumin           | NA    | NA    | 0.77  | $< 0.001$ | -0.25 | $< 0.05$ |
| GNRI              | 0.71  | $< 0.001$ | 0.54  | $< 0.001$ | -0.17 | 0.13 |
| Choline esterase  | 0.42  | $< 0.001$ | 0.43  | $< 0.001$ | 0.05  | 0.68 |
| Creatinine        | 0.51  | $< 0.001$ | 0.53  | $< 0.001$ | 0.08  | 0.49 |
| %CGR              | 0.42  | $< 0.001$ | 0.46  | $< 0.001$ | 0.06  | 0.57 |
| BUN               | 0.10  | 0.35  | 0.14  | 0.19  | 0.04  | 0.69 |
| nPCR              | 0.12  | 0.30  | 0.16  | 0.14  | 0.09  | 0.41 |
| Anion gap         | 0.27  | $< 0.05$ | 0.32  | $< 0.01$ | 0.14  | 0.21 |
| Total cholesterol | 0.12  | 0.25  | 0.18  | 0.10  | 0.02  | 0.83 |
| Triglyceride      | 0.37  | $< 0.01$ | 0.40  | $< 0.01$ | 0.10  | 0.49 |
| Phosphorus        | 0.22  | $< 0.05$ | 0.19  | 0.08  | -0.03 | 0.81 |
| Neutrophil        | 0.11  | 0.35  | 0.24  | $< 0.05$ | 0.26  | $< 0.05$ |
| CRP               | -0.16 | 0.16  | -0.12 | 0.29  | 0.10  | 0.39 |
| post-BW           | 0.08  | 0.47  | 0.14  | 0.19  | 0.07  | 0.56 |
| post-BMI          | 0.04  | 0.73  | 0.04  | 0.75  | -0.03 | 0.82 |
| Age               | -0.43 | $< 0.001$ | -0.46 | $< 0.001$ | -0.06 | 0.56 |
| HD period         | 0.45  | $< 0.001$ | 0.25  | $< 0.05$ | 0.13  | 0.23 |
| Gender            | 0.06  | 0.58  | 0.05  | 0.67  | -0.04 | 0.74 |

GNRI, geriatric nutritional risk index; post-BW, post-dialysis body weight; post-BMI, post-dialysis body mass index (calculated as the weight in kilograms divided by the square of the height in meters). N = 87.

doi:10.1371/journal.pone.0132539.t007
the middle and highest tertiles. These findings reveal for the first time, to our knowledge, that a kidney injury biomarker NGAL reflects the current nutritional status, and may be useful to predict its progressive worsening in HD patients.

In obese humans [35] and mice [36], serum NGAL levels are reportedly elevated compared to lean ones. In the present study, we propose that nutrition (by anion gap) and physical constitution (by %CGR) potently and independently affect serum NGAL levels in maintenance HD patients. In obese mice, NGAL gene expression is enhanced in the liver and adipose tissue as compared to lean mice [37]. Of note, neither pre- nor post-dialysis body mass index (BMI) was associated with pre-dialysis serum NGAL levels in HD patients of our study (S3 Table), likely because only few obese subjects were included in this study (mean BMI was 22.2 kg/m²), and also because congestive heart failure, hypotension and malnutrition might have caused inappropriate water retention and superficial elevation of BMI in some patients. Very recently, in the absence of kidney injury, we reported that approximately 70% of circulating NGAL derives from neutrophils in humans and mice [30]. Nutritional status is known to positively affect blood neutrophil counts [38]. As inducers of NGAL, inflammatory cytokines such as interleukin (IL)-1β and IL-6 have an activity to stimulate NGAL expression [15, 39]. Indeed, serum IL-1β levels are elevated in long-term HD patients [40], and skeletal muscle synthesizes and secretes IL-6 during HD session [41], which may play a role in massively elevated serum NGAL levels in HD patients. Through these possibilities, we speculate that neutrophils, adipocytes, liver and muscle are potential sources of circulating NGAL in HD patients. Furthermore, we elucidated that circulating NGAL is partially removed from the blood by renal passage in subjects with preserved renal function and by HD in end-stage renal disease patients.

Previous studies showed correlation between serum NGAL and CRP levels by multivariate analysis among HD patients, but significant correlation was lost in our multivariate analysis. The reasons may include (1) blood neutrophil (or WBC) counts were evaluated only in our study and contribution of CRP was incorporated into neutrophil counts, and (2) our patients had much lower CRP levels (median 0.1 mg/dl) compared to reports by Bolignano et al. (median 0.8 mg/dl) [42] and Malyszko et al. (mean 0.7 mg/dl) [43].

Metabolic acidosis (low pH) compensated by respiratory alkalosis (low pCO²) was a typical pattern of acid-base balance in HD patients studied (S1 Table). Metabolic acidosis associated with increased anion gap may be caused by underdialysis and anorexia. On the other hand, however, high dietary protein intake (and, therefore, high nPCR) may result in greater net acid gain and acidosis (thus, high anion gap), which is not necessarily harmful in well-dialyzed HD patients [26, 44]. In this work, serum NGAL levels were positively associated with anion gap, nPCR, and Kt/Vsp [42] (Table 2), favoring the latter effect. As can be predicted from the formula for anion gap calculation (see Materials and Methods section), Cl⁻ concentration was negatively associated with anion gap (r = -0.36, P<0.001) and, thus, also with serum NGAL levels (r = -0.32, P<0.001).

Of note, serum NGAL levels were increased after HD session in 7% of patients, and metabolic acidosis was associated with elevation in serum NGAL levels. NGAL was originally identified as exocytosed material from phorbol myristate acetate-stimulated neutrophils [1]. Importantly, mild acidic environment at levels typically observed in HD patients (pre-dialysis plasma HCO₃⁻ levels < 21 mEq/l) enhances degranulation (or oxidative burst reaction) of activated neutrophils [45], and HD procedure potentially induces neutrophil degranulation [46]. These findings raise a possibility that NGAL may be secreted from activated neutrophils during HD session.

Since NGAL is an iron carrier protein [2–4, 10, 47], it is important to test whether iron status directly affects circulating NGAL concentrations. Here we show that serum NGAL levels were not significantly altered by repeated iron administration, at the amount sufficient to cause
4.3-fold elevation in serum ferritin levels. Bolignano et al. recently reported that NGAL levels are elevated after iron injection in maintenance HD patients but only by 9% (which was accompanied with 1.8-fold ferritin elevation) [42]. The iron formula used was sodium ferric gluconate which may cause leukocyte activation by generation of reactive oxygen species [42, 48]. Indeed, there are iron formulas which reportedly do not increase circulating NGAL levels after injection [49, 50]. Taking these findings into account, we would like to conclude that iron is not a major determinant of serum NGAL levels, at least, in maintenance HD patients. Bolignano et al. also reported marginal correlation between serum NGAL and TSAT levels (r = 0.29, P = 0.04) but it may be an indirect effect as the authors suggested [42].

In the present study, we show that maintenance HD patients with reduced serum NGAL levels were prospectively associated with decrease in serum albumin levels, and had an increased likelihood to require admission for the treatment of infectious diseases. These findings can be caused by malnutrition and neutropenia, and by reduced NGAL’s bacteriostatic activity. Growth inhibition of several but not all pathogens by iron depletion is an established activity of NGAL [10–12, 51]. Serum NGAL levels in our maintenance HD patients were approximately 10–20 fold higher compared to levels reported in healthy subjects [21, 52], but iron-chelating activity of NGAL might be partly inhibited in HD patients, since these patients are replete with exogenous iron [10] to avoid anemia (Table 1).

We further studied why baseline NGAL levels can predict future changes in albumin levels (S2 Fig). Among 87 patients who were enrolled in longitudinal analysis, correlation between baseline NGAL and increase in albumin after a year (r = 0.36, P<0.01) was much stronger than that between baseline NGAL and albumin (r = 0.19, P = 0.09). By reviewing medical records of typical cases, we realized that cases with hypoalbuminemia and high NGAL levels may experience elevation in serum albumin levels after recovery from infection (or inflammatory disorders). On the other hand, in cases with preserved albumin levels and low NGAL levels, serum albumin levels may decrease once severe infection develops and reduced albumin levels may persist even after recovery from infection. These characteristics of serum NGAL levels may make NGAL an interesting biomarker which has a robust power to predict alteration in serum albumin levels in a year.

Circulating NGAL level is an early biomarker for AKI and high NGAL concentrations predict progression of renal damage (or serum creatinine elevation) [16] and mortality in AKI [18]. On the other hand, we show here that high NGAL levels in maintenance HD patients were closely associated with good nutritional status at present and preserved serum albumin levels after a year, and therefore potentially with reduced morbidity and mortality [53]. These findings may, at first glance, appear confusing. However, similar paradoxical observations have been found for circulating creatinine level, the gold standard for renal function evaluation, and for circulating homocysteine level [54–56]. These phenomena are called reverse epidemiology or risk factor paradox.

There are several limitations in the current work. First, this is a short (one year of observation) and small scale study involving only three dialysis centers. To study clinical outcomes associated to blood NGAL levels, we examined marked changes in serum albumin levels and occurrence of severe infection, but we had to refrain from multifactor adjustments because of little event numbers. Second, in a prospective study, we examined serum albumin levels at a single point of one year after enrollment. Furthermore, hospitalized patients at enrollment were excluded in this study, which might have affected association of baseline clinical parameters with outcomes in a year. Third, mortality rate was low and a number of patients (34%) were admitted to hospitals at least once. These facts may reflect a health insurance system in Japan, in which end-stage renal disease patients are given an intense health care.
Therefore, carefulness is required to extrapolate the findings obtained here into patients in other countries.

In conclusion, here we show evidence indicating that nutritional status, muscle mass and WBC counts are important determinants of serum NGAL in maintenance HD patients. It is likely that similar association also occurs in subjects with normal or nearly-normal renal function, just as serum creatinine is affected by muscle mass in patients with various renal functions [57]. Since blood and urinary NGAL levels are now vigorously measured worldwide to evaluate a broad spectrum of kidney diseases, especially in the setting of AKI, it is important to elucidate the regulatory mechanism of NGAL concentration for precise interpretation of the results from NGAL measurements. Large-scale, long-term clinical studies may be warranted to investigate whether low serum NGAL levels in maintenance HD patients are associated with higher incidence of infectious disorders, morbidity and mortality.

**Materials and Methods**

**Patients**

Out-patient, maintenance, Japanese HD subjects were enrolled between December 2008 through March 2009 at two dialysis centers, Saiseikai Nakatsu Hospital (n = 79) and Misugikai Satou Hospital (n = 16), in Osaka, Japan (S1 Fig). Patients were consecutively enrolled and prospectively followed for a year. New patients were added in February and March 2015 at Misugikai Otokoyama Hospital (n = 44) in Osaka, but follow up data were not available for those patients. Of overall 155 patients recruited, cases who did not agree to participate (n = 8), who were admitted during the recruitment period (n = 5), and who were under hemodiafiltration (n = 3) were omitted. Thirty-one patients under inflammatory status were excluded in a subanalysis (as shown in the last paragraph of Results section). All patients were on HD with standard bicarbonate buffer and type III or IV high-flux membranes for 3.0–4.5 h, three times a week.

Healthy volunteers were also enrolled (6 males and 2 females; age 35±5). Presence of DM was defined by taking any oral anti-DM medication or insulin injection. A separate group of 30 patients were enrolled at Nara Medical University Hospital, who underwent elective coronary angiography for evaluation of ischemic heart disease [58]. Only plasma was available for these patients. The protocol was approved by ethical committees in participating dialysis centers, Kyoto University Graduate School of Medicine (No. E-541), and Nara Medical University (No. 2002–009), and the study was conducted according to the Declaration of Helsinki. All participants gave written informed consent.

**Laboratory Analyses**

Blood samples were collected at the beginning and end of HD session after the longest interdialytic period (on Mondays or Tuesdays). To examine serial changes in blood concentrations of NGAL and other parameters, blood was obtained also at 10, 60 and 120 min after beginning of HD in 9 patients. In 6 cases who started to be treated with intravenous iron injection (Cideferron, macromolecular complex of ferric hydroxide with dextrin and citric acid; Nippon-zoki, Osaka, Japan), blood collection was carried out at 0, 1, 2, 3 and 8 weeks after initiation of iron injection. Serum or plasma was separated immediately and kept frozen at -80°C until analysis. NGAL was measured by sandwich ELISA (BioPorto; Gentofte, Denmark) usually after 4,000-fold dilution. We studied same-patient-variability of blood NGAL levels in 10 HD patients by comparing 2 points which were 1 week apart. By paired analysis of the cases, the second measurement gave 99.6±10.6% value (mean±SD) of the first, indicating that serum NGAL levels were considerably stable, at least, for short term. Hepcidin-25 was determined at Medical Research Institute, Kanazawa Medical University by a proteomic method using
surface-enhanced laser desorption ionization time of flight mass spectrometry (SELDI-TOF MS) [28]. Routine laboratory measurements were carried out in each clinical institute. Serum CRP levels were measured by an improved latex aggregation method (Nittobo Medical, Tokyo), and concentrations below 0.03 mg/dl were given a value of 0.03 (see Fig 1D). Creatinine was measured by an enzymatic method.

**Calculation of clinical indices**

Calcium levels were normalized with Payne’s formula, when serum albumin was less than 4 g/dl [59]. Anion gap was determined as (Na⁺ + K⁺) - (Cl⁻ + HCO₃⁻). Calculation as Na⁺ - Cl⁻ - HCO₃⁻ gave very similar findings (not shown). Kt/Vsp [60], nPCR [61] and %CGR [23] were determined by formulas previously described. Replacement of Kt/Vsp with equilibrated Kt/V gave almost identical results (not shown). GNRI was calculated as 14.89 x serum albumin (g/dl) + 41.7 x [(post-dialysis body weight)/(ideal body weight)] [33, 34]. When post-dialysis body weight exceeded ideal body weight, the ratio was given a value of 1.

**Statistical analyses**

All variables were expressed as mean±SD, median (25–75 percentile) or number (percentage). Difference in two groups was examined either by paired t-test or repeated-measures ANOVA with Dunnett post test. Comparison among three groups was carried out by ANOVA for continuous variables and Kruskal-Wallis test for dichotomous and non-normal variables. Univariate linear regression analysis was performed with Spearman rank test. Multivariate linear regression analysis was done with stepwise method using entrance/exit tolerances of 0.05/0.10. Odds ratio was calculated by χ² test. All analyses were conducted using SPSS Version 17.0 (SPSS Inc., Chicago, IL, USA). The P values reported were two-sided and considered significant at < 0.05.

**Supporting Information**

**S1 Fig. Overall study population.**
(TIF)

**S2 Fig. Correlation of increase in serum albumin levels after a year with baseline clinical parameters among 87 patients.** Linear lines show regression lines. sAlb increase, increase in serum albumin after a year; sNGAL, baseline serum NGAL; sAlb, baseline serum albumin; sCRP, baseline serum C-reactive protein; NS, not significant. A red point indicates a 72-year-old man who had baseline concentrations of NGAL 2082 ng/ml, albumin 3.0 g/dl, CRP 4.39 mg/dl, and WBC 6120 /μl. He had mild pneumonia at enrollment. His albumin level was elevated by 0.4 g/dl in a year. A green point indicates a 72-year-old woman having baseline NGAL 414 ng/ml, albumin 4.0 g/dl, CRP 0.07 mg/dl, and WBC 2948 /μl. She developed septic shock and was admitted twice at 2 and 6 months after enrollment. She survived but hypoalbuminemia of 3.5 g/dl persisted after a year.
(TIF)

**S1 Table. Comprehensive baseline clinical parameters of 139 maintenance HD patients.**
Body mass index (BMI) was calculated as the weight in kilograms divided by the square of the height in meters. MCV, mean corpuscular volume; MCH, mean corpuscular hemoglobin; MCHC, mean corpuscular hemoglobin concentration.
(XLSX)
S2 Table. Raw data of 139 HD patients at baseline.
(XLSX)

S3 Table. Correlations of clinical parameters with baseline pre-dialysis serum NGAL levels by univariate linear regression analysis. N = 139.
(XLSX)

S4 Table. Mutual association among 5 clinical parameters which were closely correlated to baseline serum NGAL level. Numbers indicate correlation coefficients (r) for statistically significant association by univariate linear regression analysis (P<0.05). Strong correlations are highlighted in gray (P<0.001). NS, not significant. N = 139.
(XLS)

S5 Table. Raw data of 6 anemic patients treated with iron.
(XLSX)

S6 Table. Raw data of blood concentrations of circulating substances during HD session in 9 patients.
(XLSX)

S7 Table. Raw data of blood concentrations of NGAL, creatinine, albumin and choline esterase in aorta and renal vein in 30 patients. AO, aorta; RV, renal vein; Cr, creatinine; ChE, choline esterase.
(XLSX)

S8 Table. Raw data of clinical events which occurred during one year follow-up in 87 HD patients.
(XLSX)

S9 Table. Prediction for occurrence of severe infection within a year. N = 87.
(XLSX)

Acknowledgments
We gratefully acknowledge Ms. Y. Ogawa, S. Ogino and other lab members for assistance and encouragement.

Author Contributions
Conceived and designed the experiments: HI KM. Performed the experiments: HI AI KPM T. Kuwahara MS K. Nakatani NT KM. Analyzed the data: HI AI HY MK T. Kuwabara KPM YK T. Kuwahara MS K. Nakatani YS NT AS K. Nakao MM MY KM. Wrote the paper: HI MM MY KM.

References
1. Kjeldsen L, Johnsen AH, Sengelov H, Borregaard N. Isolation and primary structure of NGAL, a novel protein associated with human neutrophil gelatinase. The Journal of biological chemistry. 1993; 268 (14):10425–32. PMID: 7683678.
2. Goetz DH, Holmes MA, Borregaard N, Bluhm ME, Raymond KN, Strong RK. The neutrophil lipocalin NGAL is a bacteriostatic agent that interferes with siderophore-mediated iron acquisition. Molecular cell. 2002; 10(5):1033–43. PMID: 12453412.
3. Mori K, Nakao K. Neutrophil gelatinase-associated lipocalin as the real-time indicator of active kidney damage. Kidney international. 2007; 71(10):967–70. doi: 10.1038/sj.ki.5002165 PMID: 17342180.
4. Bao G, Clifton M, Hoette TM, Mori K, Deng SX, Qiu A, et al. Iron traffics in circulation bound to a siderocalin (Ngal)-catechol complex. Nature chemical biology. 2010; 6(8):602–9. doi: 10.1038/nchembio.402 PMID: 20581821; PubMed Central PMCID: PMC2907470.
5. Hingorani S, Molitoris BA, Himmelfarb J. Ironing out the pathogenesis of acute kidney injury. American journal of kidney diseases: the official journal of the National Kidney Foundation. 2009; 53(4):569–71. doi: 10.1053/j.ajkd.2009.01.002 PMID: 19324244; PubMed Central PMCID: PMC2693097.

6. Mori K, Lee HT, Rapoport D, Drexler IR, Foster K, Yang J, et al. Endocytic delivery of lipocalin-siderophore-iron complex rescues the kidney from ischemia-reperfusion injury. The Journal of clinical investigation. 2005; 115(3):610–21. doi: 10.1172/JCI23056 PMID: 15711640; PubMed Central PMCID: PMC548316.

7. Yang J, Goetz D, Li JY, Wang W, Mori K, Sellik D, et al. An iron delivery pathway mediated by a lipocalin. Molecular cell. 2002; 10(5):1045–56. PMID: 12453413.

8. Hanai J, Mamamoto T, Seth P, Mori K, Karumanchi SA, Barasch J, et al. Lipocalin 2 diminishes invasiveness and metastasis of Ras-transformed cells. The Journal of biological chemistry. 2005; 280(14):13641–7. doi: 10.1074/jbc.M413047200 PMID: 15691834.

9. Bolignano D, Donato V, Lacquaniti A, Fazio MR, Bono C, Coppolino G, et al. Neutrophil gelatinase-associated lipocalin (NGAL) in human neoplasias: a new protein enters the scene. Cancer letters. 2010; 288(1):10–6. doi: 10.1016/j.canlet.2009.05.027 PMID: 19540040.

10. Flo TH, Smith KD, Sato S, Rodriguez DJ, Holmes MA, Strong RK, et al. Lipocalin 2 mediates an innate immune response to bacterial infection by sequestering iron. Nature. 2004; 432(7019):917–21. doi: 10.1038/nature03104 PMID: 15531878.

11. Saiga H, Nishimura J, Kuwata H, Okuyama M, Matsumoto S, Sato S, et al. Lipocalin 2-dependent inhibition of mycobacterial growth in alveolar epithelium. Journal of immunology. 2008; 181(12):8521–7. PMID: 19050270.

12. Chan YR, Liu JS, Pociask DA, Zheng M, Mietzner TA, Berger T, et al. Lipocalin 2 is required for pulmonary host defense against Klebsiella infection. Journal of immunology. 2009; 182(8):4947–56. doi: 10.4049/jimmunol.0803282 PMID: 19342674; PubMed Central PMCID: PMC2708928.

13. Paragas N, Qiu A, Zhang Q, Samstein B, Deng SX, Schmidt-Ott KM, et al. The Ngal reporter mouse detects the response of the kidney to injury in real time. Nature medicine. 2011; 17(2):216–22. doi: 10.1038/nm.2290 PMID: 21240264; PubMed Central PMCID: PMC3059503.

14. Xu SY, Pauksen K, Venge P. Serum measurements of human neutrophil lipocalin (HNL) discriminate between acute bacterial and viral infections. Scandinavian journal of clinical and laboratory investigations. 1995; 55(2):125–31. doi: 10.3109/00365519509089604 PMID: 7667605.

15. Cowland JB, Muta T, Borregaard N. IL-1beta-specific up-regulation of neutrophil gelatinase-associated lipocalin (NGAL) in human neoplasias: a new protein enters the scene. Cancer letters. 1995; 94(66):1231–8. doi: 10.1016/S0140-6736(05)74811-X PMID: 15811456.

16. Mishra J, Dent C, Tarabishi R, Mitsnefes MM, Ma Q, Kelly C, et al. Neutrophil gelatinase-associated lipocalin (NGAL) as a biomarker for acute renal injury after cardiac surgery. Lancet. 2005; 365(9466):1231–8. doi: 10.1016/S0140-6736(05)74811-X PMID: 15811456.

17. Wagener G, Jan M, Kim M, Mori K, Barasch JM, Sladen RN, et al. Association between increases in urinary neutrophil gelatinase-associated lipocalin and acute renal dysfunction after adult cardiac surgery. Anesthesiology. 2006; 105(3):485–91. PMID: 16931980.

18. Dent CL, Ma Q, Dastrala S, Bennett M, Mitsnefes MM, Barasch J, et al. Plasma neutrophil gelatinase-associated lipocalin predicts acute kidney injury, morbidity and mortality after pediatric cardiac surgery: a prospective uncontrolled cohort study. Critical care. 2007; 11(6):R127. doi: 10.1186/cc6192 PMID: 18070344; PubMed Central PMCID: PMC2246223.

19. Haase-Fielitz A, Bellomo R, Devarajan P, Story D, Matalanis G, Dragun D, et al. Novel and conventional serum biomarkers predicting acute kidney injury in adult cardiac surgery—a prospective cohort study. Critical care medicine. 2009; 37(2):555–60. doi: 10.1097/CCM.0b013e318195846e PMID: 19114878.

20. Bolignano D, Lacquaniti A, Coppolino G, Campo S, Arena A, Buemi M. Neutrophil gelatinase-associated lipocalin reflects the severity of renal impairment in subjects affected by chronic kidney disease. Kidney & blood pressure research. 2008; 31(4):255–8. doi: 10.1053/j.ajkd.2009.01.002 PMID: 19324244; PubMed Central PMCID: PMC2693097.

21. Bolignano D, Lacquaniti A, Coppolino G, Donato V, Campo S, Fazio MR, et al. Neutrophil gelatinase-associated lipocalin (NGAL) and progression of chronic kidney disease. Clinical journal of the American Society of Nephrology. CJASN. 2009; 4(2):337–44. doi: 10.2215/CJN.03530708 PMID: 19176795; PubMed Central PMCID: PMC2637601.

22. Borah MF, Schoenfeld PY, Gotch FA, Sargent JA, Wolfsen M, Humphreys MH. Nitrogen balance during intermittent dialysis therapy of uremia. Kidney international. 1978; 14(5):491–500. PMID: 750694.

23. Shinzato T, Nakai S, Miwa M, Iwayama N, Takai I, Matsumoto Y, et al. New method to calculate creatinine generation rate using pre- and postdialysis creatinine concentrations. Artificial organs. 1997; 21(8):864–72. PMID: 9247178.
24. Chandra RK. Nutrition, immunity, and infection: present knowledge and future directions. Lancet. 1983; 1(8326 Pt 1):688–91. PMID: 6132048.

25. Bergstrom J. Nutrition and mortality in hemodialysis. Journal of the American Society of Nephrology: JASN. 1995; 6(5):1329–41. PMID: 8589306.

26. Dumler F, Falla P, Butler R, Wagner C, Francisco K. Impact of dialysis modality and acidosis on nutritional status. ASAIO journal. 1999; 45(5):413–7. PMID: 10503617.

27. Nemeth E, Tuttle MS, Powelson J, Vaughn MB, Donovan A, Ward DM, et al. Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. Science. 2004; 306(5704):2090–3. doi: 10.1126/science.1104742 PMID: 15514116.

28. Tomosugi N, Kawabata H, Wakatabe R, Higuchi M, Yamaya H, Urnêhara H, et al. Detection of serum hepcidin in renal failure and inflammation by using ProteinChip System. Blood. 2006; 108(4):1381–7. doi: 10.1182/blood-2005-10-4043 PMID: 16621968.

29. Nakashima T, Kuragano T, Kaibe S, Nagasawa Y, Hasuike Y. Should we reconsider iron administration based on prevailing ferritin and hepcidin concentrations? Clinical and experimental nephrology. 2012; 16(6):819–26. doi: 10.1007/s10157-012-0694-3 PMID: 23053592; PubMed Central PMCID: PMC3521641.

30. Kanda J, Mori K, Kawabata H, Kuwabara T, Mori KP, Imamaki H, et al. An AKI biomarker lipocalin 2 in the blood derives from the kidney in renal injury but from neutrophils in normal and infected conditions. Clinical and experimental nephrology. 2014. doi: 10.1007/s10157-014-0952-7 PMID: 24599361.

31. Bouillanne O, Morineau G, Dupont C, Coulombel I, Vincent JP, Nicolis I, et al. Geriatric Nutritional Risk Index: a new index for evaluating at-risk elderly medical patients. The American journal of clinical nutrition. 1999; 45(5):413–7. PMID: 10503617.

32. Craddock PR, Fehr J, Dalmasso AP, Brighan KL, Jacob HS. Hemodialysis leukopenia. Pulmonary vascular leukostasis resulting from complement activation by dialyzer cellophane membranes. The Journal of clinical investigation. 1977; 59(5):879–88. doi: 10.1172/JCI108710 PMID: 856872; PubMed Central PMCID: PMC372296.

33. Lockridge O, Bartels CF, Vaughan TA, Wong CK, Norton SE, Johnson LL. Complete amino acid sequence of human serum cholinesterase. The Journal of biological chemistry. 1987; 262(2):549–57. PMID: 3542989.

34. Boullanne O, Morineau G, Dupont C, Coulombel I, Vincent JP, Nicolis I, et al. Geriatric Nutritional Risk Index: a new index for evaluating at-risk elderly medical patients. The American journal of clinical nutrition. 1999; 45(5):413–7. PMID: 10503617.

35. Yamada K, Furuya R, Takita T, Maruyama Y, Yamaguchi Y, Ohkawa S, et al. Simplified nutritional screening tools for patients on maintenance hemodialysis. The American journal of clinical nutrition. 2008; 87(1):106–13. PMID: 18175743.

36. Wang Y, Lam KS, Kraegen EW, Sweeney G, Zhang J, Tso AW, et al. Lipocalin-2 is an inflammatory marker closely associated with obesity, insulin resistance, and hyperglycemia in humans. Clinical chemistry. 2007; 53(1):34–41. doi: 10.1373/clinchem.2006.075614 PMID: 17040956.

37. Yan QW, Yang Q, Mody N, Graham TE, Hsu CH, Xu Z, et al. The adipokine lipocalin 2 is regulated by obesity and promotes insulin resistance. Diabetes. 2007; 56(10):2533–40. doi: 10.2337/db07-0007 PMID: 17639021.

38. Zhang J, Wu Y, Zhang Y, Leroith D, Bernlohr DA, Chen X. The role of lipocalin 2 in the regulation of inflammation in adipocytes and macrophages. Molecular endocrinology. 2008; 22(6):1416–26. doi: 10.1210/me.2007-0420 PMID: 18292240; PubMed Central PMCID: PMC2422824.

39. Drenick EJ, Alvarez LC. Neutropenia in prolonged fasting. The American journal of clinical nutrition. 1971; 24(7):859–63. PMID: 5091528.

40. Zhang Y, Foncea R, Deis JA, Guo H, Bernlohr DA, Chen X. Lipocalin 2 expression and secretion is highly regulated by metabolic stress, cytokines, and nutrients in adipocytes. PloS one. 2014; 9(5):e96997. doi:10.1371/journal.pone.0096997 PMID: 24818605; PubMed Central PMCID: PMC4018437.

41. Goldstein SL, Leung JC, Silverstein DM. Pro- and anti-inflammatory cytokines in chronic pediatric dialysis patients: effect of aspirin. Clinical journal of the American Society of Nephrology: CJASN. 2006; 1(5):979–86. doi: 10.2215/CJN.02291205 PMID: 17699316.

42. Raj DS, Dominic EA, Pai A, Osman F, Morgan M, Pickett G, et al. Skeletal muscle, cytokines, and oxidative stress in end-stage renal disease. Kidney international. 2005; 68(5):2338–44. doi: 10.1111/j.1523-1755.2005.00695.x PMID: 16221238.

43. Bolignano D, Coppolino G, Romeo A, De Paola L, Buemi A, Lacquaniti A, et al. Neutrophil gelatinase-associated lipocalin (NGAL) reflects iron status in haemodialysis patients. Nephrology, dialysis, transplantation: official publication of the European Dialysis and Transplant Association—European Renal Association. 2009; 24(11):3398–403. doi: 10.1111/j.1440-1746.2009.05962.x PMID: 19546966.

44. Myśliwiec M, Myśliwiec J, Koc-Ziolkowska E, Kozinski P, Myśliwiec M. Neutrophil gelatinase-associated lipocalin in dialyzed patients is related to residual renal function, type of renal replacement therapy...
and inflammation. Kidney & blood pressure research. 2009; 32(6):464–9. doi: 10.1159/000274048 PMID: 20068352.

44. Kalantar-Zadeh K, Mehrrota R, Fouque D, Kopple JD. Metabolic acidosis and malnutrition-inflammation complex syndrome in chronic renal failure. Seminars in dialysis. 2004; 17(6):455–65. doi: 10.1111/j.0953-6766.2004.00196.x PMID: 15660576.

45. Wann JG, Hsu YH, Yang CC, Lin CS, Tai DW, Chen JS, et al. Neutrophils in acidic haemodialysed patients have lower intracellular pH and inflamed state. Nephrology, dialysis, transplant: official publication of the European Dialysis and Transplant Association—European Renal Association. 2007; 22(9):2613–22. doi: 10.1093/ndt/gfm227 PMID: 17504845.

46. Horl WH. Hemodialysis membranes: interleukins, biocompatibility, and middle molecules. Journal of the American Society of Nephrology: JASN. 2003; 13 Suppl 1:S62–71. PMID: 11972764.

47. Barasch J, Mori K. Cell biology: iron thievery. Nature. 2004; 432(7019):811. PMID: 15602535.

48. Gupta A, Zhuo J, Zha J, OLP J, Pai A. Effect of different intravenous iron preparations on lymphocyte intracellular reactive oxygen species generation and subpopulation survival. BMC nephrology. 2010; 11:16. doi: 10.1186/1471-2369-11-16 PMID: 20716362; PubMed Central PMCID: PMC2933673.

49. Jiang W, Constante M, Santos MM. Anemia upregulates lipocalin 2 in the liver and serum. Blood cells, molecules & diseases. 2008; 41(2):169–74. doi: 10.1016/j.bcmd.2008.04.006 PMID: 18519167; PubMed Central PMCID: PMC2909274.

50. Johnson AC, Becker K, Zager RA. Parenteral iron formulations differentially affect MCP-1, HO-1, and NGAL gene expression and renal responses to injury. American journal of physiology Renal physiology. 2010; 299(2):F426–35. doi: 10.1152/ajprenal.00248.2010 PMID: 20504881; PubMed Central PMCID: PMC2928522.

51. Fluckinger M, Haas H, Merschak P, Glasgow BJ, Redi B. Human tear lipocalin exhibits antimicrobial activity by scavenging microbial siderophores. Antimicrobial agents and chemotherapy. 2004; 48 (9):3367–72. doi: 10.1128/AAC.48.9.3367-3372.2004 PMID: 15328098; PubMed Central PMCID: PMCS147377.

52. Kuwabara T, Mori K, Mukoyama M, Kasahara M, Yokoi H, Saito Y, et al. Urinary neutrophil gelatinase-associated lipocalin levels reflect damage to glomeruli, proximal tubules, and distal nephrons. Kidney international. 2009; 75(3):285–94. doi: 10.1038/ki.2008.499 PMID: 19148153.

53. Lowrie EG, Lew NL. Death risk in hemodialysis patients: the predictive value of commonly measured variables and an evaluation of death rate differences between facilities. American journal of kidney diseases: the official journal of the National Kidney Foundation. 1993; 15(5):458–82. PMID: 2333868.

54. Fleischmann EH, Bower JD, Salahudeen AK. Risk factor paradox in hemodialysis: better nutrition as a partial explanation. ASAIO journal. 2001; 47(1):74–81. PMID: 11193620.

55. Kalantar-Zadeh K, Block G, Humphreys MH, Kopple JD. Reverse epidemiology of cardiovascular risk factors in maintenance dialysis patients. Kidney international. 2003; 63(3):793–808. doi: 10.1046/j.1523-1755.2003.00803.x PMID: 12631061.

56. Kalantar-Zadeh K, Block G, Humphreys MH, McAllister CJ, Kopple JD. A low, rather than a high, total plasma homocysteine is an indicator of poor outcome in hemodialysis patients. Journal of the American Society of Nephrology: JASN. 2004; 15(2):442–53. PMID: 14747392.

57. Heymsfield SB, Arteaga C, McManus C, Smith J, Moffitt S. Measurement of muscle mass in humans: validity of the 24-hour urinary creatinine method. The American journal of clinical nutrition. 1983; 37 (3):478–94. PMID: 6829490.

58. Iwama H, Uemura S, Naya N, Imagawa K, Takemoto Y, Asai O, et al. Cardiac expression of placent growth factor predicts the improvement of chronic phase left ventricular function in patients with acute myocardial infarction. Journal of the American College of Cardiology. 2006; 47(8):1559–67. doi: 10.1016/j.jacc.2005.11.064 PMID: 16630991.

59. Payne RB, Little AJ, Williams RB, Milner JR. Interpretation of serum calcium in patients with abnormal serum proteins. British medical journal. 1973; 4(5893):643–6. PMID: 4758544; PubMed Central PMCID: PMC1587636.

60. Daugirdas JT. Second generation logarithmic estimates of single-pool variable volume Kt/V: an analysis of error. Journal of the American Society of Nephrology: JASN. 1993; 4(5):1205–13. PMID: 8305648.

61. Shinzato T, Nakai S, Fujita Y, Takai I, Morita H, Nakane K, et al. Determination of Kt/V and protein catabolic rate using pre- and postdialysis blood urea nitrogen concentrations. Nephron. 1994; 67(3):280–90. PMID: 7936017.