Antiplasmodial and Cytotoxic Activities of Toad Venoms from Southern Amazon, Brazil

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Abstract: The drug-resistance of malaria parasites is the main problem in the disease control. The huge Brazilian biodiversity promotes the search for new compounds, where the animal kingdom is proving to be a promising source of bioactive compounds. The main objective of this study was to evaluate the antiplasmodial and cytotoxic activity of the compounds obtained from the toad venoms of Brazilian Amazon. Toad venoms were collected from the secretion of Rhinella marina and Rhaebo guttatus in Mato Grosso State, Brazil. The powder was extracted at room temperature, yielding 2 extracts (RG and RM) and a substance (‘1’) identified as a bufadienolide, named telocinobufagin. Growth inhibition, intraerythrocytic development, and parasite morphology were evaluated in culture by microscopic observations of Giemsa-stained thin blood films. Cytotoxicity was determined against HepG2 and BGM cells by MTT and neutral red assays. The 2 extracts and the pure substance ‘1’ tested were active against chloroquine-resistant Plasmodium falciparum strain, demonstrating lower IC50 values. In cytotoxic tests, the 2 extracts and substance ‘1’ showed pronounced lethal effects on chloroquine-resistant P. falciparum strain and low cytotoxic effect, highlighting toad parotoid gland secretions as a promising source of novel lead antiplasmodial compounds.

Key words: Plasmodium falciparum, antiplasmodial effect, toad venom, bufadienolide, malaria chemotherapy

INTRODUCTION

The 2015 Nobel Prize in Physiology or Medicine was awarded to 3 scientists who made pioneering discoveries of drugs currently used to treat neglected tropical diseases [1]. A portion of the award was to Youyou Tu (China), guided by ancient Chinese texts, isolated a component from the plant Artemisia annua, called artemisinin, which proved to be highly effective against the malaria parasite [2]. Artemisinin has now become the drug of choice for the treatment of malaria throughout most of the tropical world.

Malaria remains an important cause of illness and death in children and adults in underdeveloped countries in which it is endemic. The drug-resistant malaria parasite is the main problem in the disease control. Artemisinin resistance has been described in malaria endemic areas, threatening malaria control, treatment, and elimination efforts worldwide [3]. World Health Organization (WHO) updated their treatment policy in countries where P. falciparum parasite is endemic from use of monotherapy with drugs such as chloroquine, amodiaquine, and sulfadoxine–pyrimethamine (SP) to currently recommended artemisinin-based combination therapies (ACT) [4]. However, resistance to ACTs has arisen recently in P. falciparum in South East Asia [4,5]. Therefore, it is necessary to develop novel drugs for malaria therapy, particularly in regions where resistant Plasmodium strains are present.

The use of natural products has provided a prospective strategy for identifying novel antimalarial drugs. In this context, the bioprospecting of secondary metabolites can be an important tool to new antimalarial drugs discovery. The use of animals, plants, fungi, and bacteria are important sources of biologically active substances with structural diversity and novel mechanisms of action, which can possibly provide patentable products [6-12]. An example is the development of Captopril®,
this medicine was based on research on small peptides from the 
vomit of the South American snake (*Bothrops jararaca*) that 
were known to potentiate the action of bradykinin [13-15].

In the class of Amphibians are the family Bufonidae that pos-
sesses about 471 species [16]. An important genus of this family 
is *Rhinella* (formerly *Bufo* in the New World), which consists of 
about 258 species. In Latin America, they are found in Amazon 
regions of Brazil, Bolivia, Colombia, Peru, Suriname, Guiana, 
and Venezuela [17]. The skin secretions and venon of amphibi-
ans are rich sources of bioactive compounds, such as peptides, 
alkaloids, bufadienolides, biogenic amines, and proteins. These 
molecules play a crucial role in the physiological functions of 
these animals, especially for predation and protection against 
microorganisms [18,19]. Previous studies with *Rhinella marina* 
venom resulted in the identification by LC-MS of the 4 bufadi-
enolides; telocinobufagin, marinobufagin, bufalin, and resibu-
fogenin, and in *Rhaebo guttatus* venom marinobufagin was iden-
tified [20]. Bufadienolides are an important group of steroid 
hormones that has shown vasoconstriction [21], antiviral 
[22,23], antitumoral [24-26], cytotoxic [8,20,27,28], antileish-
manial, and antitypanosomal activities [29].

The potential of natural products to provide the development 
of antimalarial compounds is evident [30]. Animals contain a 
large assortment of structurally unique secondary metabolites 
that can be useful as new chemical templates for drug discovery 
[6,8]. Although amphibian skin secretions have proved to be a 
rich source of exclusive molecules, no studies have evaluated 
the potential antimalarial activity of these compounds. Our ob-
jectives were to conduct bioprospecting of the extracts and pure 
compound from *R. marina* (synonymy *Bufo marinus*) and 
*R. guttatus* toad venom, occurring in the Southern Amazon of 
Mato Grosso, Brazil, with antiplasmodial and cytotoxic activi-
ties against tumor and normal cell lines.

**MATERIALS AND METHODS**

**Sample collection**

Toad venom samples were collected from the secretion of *R. marina* 
and *R. guttatus* in Mato Grosso State, Brazil. The animals were 
identified by one of the authors (D. J. Rodrigues, IBAMA, SIS-
BIO: no. 30034-1). Voucher specimens (*R. marina* ABAM-H 
1262 and *R. guttatus* ABAM-H 1538) were deposited in the 
Acervo Biológico da Amazônia Meridional (Sinop, Mato Gros-
so, Brazil).

**Extraction and isolation**

Samples of toad venom of *R. marina* and *R. guttatus* were 
dried, powdered, and extracted 3 times (5 ml) with CHCl₃/
MeOH by ultrasonication for 10 min at room temperature 
[20]. The extracts were identified by the following codes: RM 
(*R. marina*) and RG (*R. guttatus*). The extract (1.1 g) from 
*R. marina* venom was chromatographed on Sephadex LH-20 col-
umn (GE Healthcare, Freiburg, Germany) using MeOH as elu-
tant. The subfraction 140 yielded the compound ‘1’ (25.7 mg).
The compound ‘1’ (Fig. 1) was identified by spectrometric 
methods (mass, NMR [H and ¹³C]) and by comparison with lit-
erature [18], as a bufadienolide, named telocinobufagin.

**Parasites**

Chloroquine (CQ)-resistant W2 strain of *P. falciparum* was 
used for in vitro blood stage culture to test the antiplasmodial 
efficacy of toad venom extracts and ‘1’. *P. falciparum* continuous 
culture was maintained according to the method described 
previously [31] with minor modifications. Cultures were 
maintained at 5% hematocrit using type O human erythrocytes 
in RPMI 1640 medium (Sigma-Aldrich, St. Louis, Missouri, 
USA) supplemented with 25 mM NaHCO₃, 1.0% albumax, 45 
mg/L hypoxanthine, 40 μg/ml gentamycin and incubated at 
37°C under approximately 5% of CO₂. The parasites were 
synchronized at ring stage by sorbitol treatment [32]. Initial 
parasitemia was adjusted to 0.5% with 2% hematocrit in 
experiments.
Antiplasmodial activity

Growth inhibition of intraerythrocytic forms and parasite morphology were evaluated in culture by the microscopic observation of Giemsa-stained thin blood films. After synchronisation by sorbitol treatment, parasite culture at ring stage (0.5% parasitemia and 2% hematocrit) was added to each well of 96-well microculture plates. The toad venom extracts (RG and RM) and ‘1’ were dissolved in DMSO and diluted to appropriate concentrations using complete medium (10 mg/ml) and stored at 4°C. The final concentration in the culture was 1%. After incubation at 37°C for 48 hr, *P. falciparum* growth inhibition was assessed by Giemsa-stained smears by observing 5,000 erythrocytes per 1 thin blood film in triplicate. The culture medium was replaced with fresh medium with or without test samples/control drugs. Chloroquine (CQ) was used as a reference antimalarial. The activity of the extracts and ‘1’ was expressed as the percentage reduction in parasitemia relative to controls without drugs. All experiments were performed in triplicate. The results were expressed as the mean of the IC₅₀ (the lethal drug concentration that reduced parasite viability in 50%).

Cells culture to cytotoxic assays

The cytotoxicity testing was performed in 2 cell lines; human hepatoma cells (HepG2) and normal kidney glomerular cells (BGM). Both of these cell lines were cultured in 75 cm² sterile flasks. RPMI 1640 (Sigma-Aldrich, St. Louis, Missouri, USA) medium supplemented with 10% heat-inactivated fetal calf serum and 40 mg/L gentamycin in a 5% CO₂ atmosphere at 37°C. When confluent, the cell monolayer was washed with culture medium, trypsinized, distributed in a flat-bottomed 96-well plate (5 × 10⁵ cells/well), and incubated for 18 hr at 37°C for cell adherence [33]. The compounds (20 μl) were diluted to various Log concentrations (0.2-200 μg/ml) and incubated with the cells for 24 hr in a 5% CO₂ atmosphere at 37°C.

Cytotoxic activity

A 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) solution (5 mg/ml; 20 μl/well) was added to evaluate mitochondrial viability; after a further 3 hr incubation, the supernatants were carefully removed, 100 μl of DMSO was added to each well, and the reactions were mixed to solubilize the formazan crystals. The optical density was determined at 540 nm to measure the signal and background, respectively (Spectra Max340PC, Molecular Devices, Sunnyvale, California, USA). The cell viability was expressed as a percentage of the control absorbance in the untreated cells after subtracting the appropriate background.

The neutral red assay (NR) [34] was also performed in triplicate. After addition of the samples and 3 hr incubation, the supernatants were carefully removed and 200 μl of neutral red solution (40 μg/ml) were added to each well. The microplates were incubated again at 37°C in a humidified atmosphere with 5% CO₂ for 3 hr. Then, the supernatant was removed and added to 200 μl of formaldehyde solution (0.5%, v/v) and CaCl₂ (1%) to each well of the test plate. After 5 min, the supernatant was again removed, and 100 μl acid alcohol solution (50%, v/v ethanol in 1% v/v acetic acid) were added to each well. The absorbance of the microplate was read in an ELISA reader (Spectra Max340PC, Molecular Devices, Sunnyvale, California, USA) using 540 nm filter.

The minimum lethal dose for 50% of the cells (LD₅₀) was determined as described [35]. RM and RG extracted compounds, ‘1’, and chloroquine concentrations used to calculate LD₅₀ were 2-2,000 μg/ml.

Selectivity index (SI)

A selectivity index (SI) corresponding to the ratio between the cytotoxic and antiparasitic activities of each sample tested. The values greater than 10 were considered indicative of lack of toxicity; however, the substances with values below 10 were considered toxic [36]. The SI index was calculated as follow: SI = LD₅₀ Cell/IC₅₀ P. falciparum.

Statistical analysis

The concentrations of compounds able to inhibit 50% of parasite growth (IC₅₀) were determined based on the equation of the curve obtained by plotting the % of parasitemia regression vs the log of the concentration of compound. The coefficients of regression of these curves were calculated using the method of least squares. The LD₅₀ were determined based on the equation of the curve obtained by plotting the % of cellular death versus the concentration of compound (GraphPad Prism Software®, version 5.0 for Windows, San Diego, California, USA). The average IC₅₀ and LD₅₀ were compared using ANOVA. Statistical significance was defined at the 5% level (P < 0.05).

RESULTS

In vitro anti-plasmodial activity

The extracts RG, RM, and ‘1’ diluted in DMSO were assayed
for antiplasmodial activity against chloroquine-resistant \( P. falciparum \) W2. Table 1 shows the antiplasmodial activity of RG, RM, and ‘1’ in 2 different experiments. When evaluated the venom extracts and ‘1’ in concentrations of 100 µg/ml, all of them showed the parasitemia reduction above 60%. The compound ‘1’ showed high potential of parasitemia reduction in vitro and the lower standard deviation (Fig. 2). Despite the high reduction of parasitemia of compound ‘1’, RG extract had a lower IC\(_{50}\) value. Starting from 100 µg/ml, the compounds were diluted to various concentrations (0.78-100 µg/ml) to calculate the IC\(_{50}\) values. All samples (RG, RM, and ‘1’) showed values lower than 2 μg/ml, thus considered active against \( P. falciparum \) W2 strain (Fig. 3).

**Cytotoxic activity on HepG2 and BGM cells**

To evaluate the cytotoxic activity of the extracts and compound ‘1’, cytotoxic assays were performed by MTT and neutral red assays in human hepatoma cells (HepG2) and normal kidney glomerular cells (BGM). When analyzed the cytotoxic activity of the samples by MTT assay in HepG2 cells, it was observed that all samples (RG, RM, and ‘1’) showed high LD\(_{50}\) values, being low or not cytotoxic to this cell line (57 > 100 µg/ml) (Table 1). In MTT assay in BGM cells, RG extract showed LD\(_{50}\) values of 34.8 µg/ml. This value to RG extract can be considered low cytotoxic to BGM cells. The other extracts (RM and ‘1’)

![Fig. 2. Parasitemia reduction (%) of \( P. falciparum \) W2 strain showing the mean± SD of the RG, RM, and ‘1’ compound. RM, \( R. marina \); RG, \( R. guttatus \); ‘1’, telocinobufagin at a concentration of 100 µg/ml; CQ, chloroquine, the antimalarial used as the positive control at a concentration of 100 µg/ml.](image)

![Fig. 3. Dose-response curves of RM, RG, and ‘1’ in function of linear regression. Effects of different concentrations of RM, RG, and ‘1’ (0.78-100 µg/ml) on the activity of \( P. falciparum \) W2 strain. Each point represents the mean± SD of 2 different samples performed in triplicates. IC\(_{50}\) values are presented in graphic.](image)

Table 1. IC\(_{50}\), LD\(_{50}\), and SI values obtained from in vitro tests with extracts (RG and RM), 1 obtained from toads venom, and chloroquine (CQ) against \( P. falciparum \) W2 strain

| Samples | IC\(_{50}\) Value (µg/ml) | R | MTT | LD\(_{50}\)a | NR | MTT | LD\(_{50}\)a | NR |
|---------|-----------------|---|-----|-------|---|-----|-------|---|
|         | HepG2 | BGM | HepG2 | BGM | HepG2 | BGM | HepG2 | BGM |
| RG      | 0.050   | 0.5247 | 100.91 | 34.83 | >100 | 22.62 | >2,000 | >500 |
| RM      | 0.534   | 0.9396 | 57.00 | 38.90 | >200 | >200 | 72.84 | >100 |
| ‘1’     | 1.280   | 0.9646 | 102.16 | 133.86 | >200 | >200 | 84.50 | >100 |
| CQ      | 0.234   | 0.9894 | >1,000 | >1,000 | >1,000 | >1,000 | >1,000 | >1,000 |

\( a \)LD\(_{50}\) and SI values were obtained with MTT and neutral red (NR) cytotoxic tests in human hepatoma cells (HepG2) and normal kidney glomerular cells (BGM).

\( b \)Chloroquine: antimalarial used as a positive control at a concentration of 2-2,000 µg/ml.
showed high LD₅₀ values (38.9–100 μg/ml), not being cytotoxic to BGM cells.

When the cytotoxic activity of the samples was analyzed in the neutral red assay, it was observed that all compounds showed high LD₅₀ values in both the cell line HepG2 and is therefore considered not cytotoxic. In BGM cells, the RG showed LD₅₀ values of 22.6 μg/ml (Table 1).

Evaluating the selectivity index (SI), all compounds showed high selectivity for the parasites when analyzed by MTT assay in the HepG2 cells (SI > 72.8). In BGM lineage, it was observed SI > 100 for the all compounds tested. In neutral red assays, all samples showed high selectivity for the parasites when analyzed in HepG2 and BGM cells (> 100) (Table 1).

**DISCUSSION**

Several extensive molecules and natural products screens against *P. falciparum* blood stages have been published annually, emerging as new candidates for antimalarial drugs. However, no antiplasmodial activity of molecules and aqueous and organic extracts obtained from toad skin glands, whose secretions exhibit bufadienolides, was previously described.

Secretions from 2 toad species, *R. marina* and *R. guttatus*, were chemically investigated previously, and the isolation of telocinobufagin was described in *R. marina* venom [20]. When analyzed the IC₅₀ values of extracts and compound 1 from the toad venom, it was noted that all samples showed values lower than 5 μg/ml can be considered highly active against *P. falciparum* W2 strain. According to WHO guidelines and studies by Pink et al. [37], the rating for the antimalarial activity of a drug is as follows: highly active (IC₅₀ < 5 μg/ml), promising active (5-15 μg/ml), lowest active (15-50 μg/ml), and inactive (> 50 μg/ml).

The telocinobufagin (1) showed a high reduction of parasitemia in vitro. One study demonstrated the biological activity of steroids telocinobufagin and hellebrigenin against *Leishmania chagasi* promastigotes, but only hellebrigenin was active against *Trypanosoma cruzi* trypomastigotes [29]. Ferreira et al. [20] showed the differences in composition between *R. marina* and *R. guttatus* venoms, in terms of the number and type of constituents. The telocinobufagin is found only in *R. marina* venom, together with another 3 bufadienolides: marinobufagin, bufalin, and resibufogenin. However, only bufadienolide (marinobufagin) was identified in *R. guttatus* venom. Thus, the lowest IC₅₀ value in antiplasmodial test can be explained by the presence of only on bufadienolide in the constitution of *R. guttatus* venom. This suggests a synergistic effect of bufadienolides present in *R. marina* venom.

Venom extracts from *R. marina* and *R. guttatus* showed moderate cytotoxic activities against HepG2 and BGM cells in MTT assay, respectively. Besides, when the cytotoxic activity of the extracts was analyzed by neutral red assay, it was observed that all samples showed high LD₅₀ values in the HepG2 cells, and RG showed moderate cytotoxicity in BGM cells. A study in seaweed in northeastern Brazil showed greater susceptibility of BGM cells to crude extracts of the studied algae when compared to macrophages [37]. However, in our study, the purified substance telocibufaginin did not show cytotoxic activities for cell line HepG2. The low cytotoxicity of the telocinobufagin against cancer cell lines (HL-60, SF-295, MDA-MB-435, and HCT-8) was demonstrated in bufadienolides isolated from the Brazilian toad *Rhinella schneideri* [8].

In our study, antiplasmodial and cytotoxic activities in HepG2 cells were best observed in RG extract. The presence of 4 bufadienolides in RM extract displayed less in vitro efficacy against *P. falciparum* compared to the RG extract. These results differ from previous studies that have reported a higher cytotoxic activity of venom extracts from *R. marina* in comparison to those from *R. guttatus* due the presence of 3 other bufadienolides (telocinobufagin, bufalin, and resibufogenin), as well as marinobufagin, only bufadienolide identified in *R. guttatus* venom [20].

According to Bézivin et al. [36], values higher than 10 (SI > 10) is indicative of high selectivity values, whereas values below 10 (SI < 10) are considered as low selectivity. In this study, for all tested samples (RG, RM, and 1), the SI was considered as high selectivity. The high selectivity index of the extracts and 1 indicate either an absence or large difference in the structure of the target molecule between *P. falciparum* and the mammalian cell. Therefore, these extracts could be exploited to identify the molecular target, which subsequently could be helpful to design novel therapeutics to combat malaria.

In this work, it was important to assess the cytotoxic activity of extracts and pure compound 1 in 2 different cell lines; tumoral and non-tumoral cells. An important article showed recently that the same chemical agent or compound can have different mechanisms of action in the cells when tested in different cell types [39]. The RM and 1 compounds did not have the LD₅₀ values determined in BGM cells used in both cytotoxic assays. Thus, it can be suggested that the mechanism of ac-
tion of these compounds in BGM cells can be different from HepG2 cells. These results demonstrate the importance of the cytotoxicity test and calculating the selectivity index for natural products with possible antimalarial activity. If only IC₅₀ values for parasite were analyzed, it may be concluded that all the samples would have potential antimalarial activities. In a study by Cenzi [40] to assess the antimalarial activity of sulfonamides derived from 4-metoxichalcona, all molecules were considered highly active against P. falciparum W2. However, when analyzing the selectivity index values of sulfonamides derived from 4-metoxichalcona, it was concluded that none of molecules was considered to have high selectivity for the parasite.

In summary, all assayed samples (extracts and ‘1’) showed antiplasmodial activity. The extracts of R. marina and R. guttatus venoms and ‘1’ isolated compound from R. marina showed pronounced lethal effects in P. falciparum chloroquine-resistant strain, highlighting toad parotoid gland secretions as a promising source of novel lead antimalarial compounds. The compound ‘1’ can serve as a prototype molecule for the development of more active compounds.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

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