Case Report

NGS Analysis Revealed Digenic Heterozygous GCK and HNF1A Variants in a Child with Mild Hyperglycemia: A Case Report

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Abstract: Monogenic diabetes (MD) represents a heterogeneous group of disorders whose most frequent form is maturity-onset diabetes of the young (MODY). MD is predominantly caused by a mutation in a single gene. We report a case of a female patient with suspected MD and a positive family history for diabetes and obesity. In this patient, two gene variants have been identified by next-generation sequencing (NGS): one in the Glucokinase (GCK) gene reported in the Human Gene Mutation Database (HGMD) and in the literature associated with GCK/MODY, and the other in the hepatocyte nuclear factor 1A (HNF1A) gene not previously described. The GCK variant was also identified in the hyperglycemic father, whereas the HNF1A variant was present in the mother. This new case of digenic GCK/HNF1A variants identified in a hyperglycemic subject, evidences the importance of NGS analysis in patients with suspected MD. In fact, this methodology will allow us to both increase the number of diagnoses and to identify mutations in more than one gene, with a better understanding of the genetic cause, and the clinical course, of the disease.

Keywords: hyperglycemia; digenic variants; GCK/MODY; HNF1A/MODY

1. Introduction

Monogenic diabetes (MD) accounts for 1–6% of all cases of diabetes in patients of pediatric-adolescent age, but its prevalence is probably underestimated [1,2]. The most common type of MD is maturity-onset diabetes of the young (MODY), a clinically and genetically heterogeneous group of monogenic, non-insulin-dependent, and nonketotic diabetes mellitus characterized by early onset (commonly before age 25), the absence of autoimmunity, and beta-cell dysfunction [3].

Up to now, mutations in 14 genes involved in glucose homeostasis and pancreatic development have been associated with this form of diabetes [4]. Although it is an autosomal dominant disorder, de novo mutations should be considered in patients without a family history of diabetes [5]. The most common forms of MODY involve mutations in hepatocyte nuclear factor 1A (HNF1A), and glucokinase (GCK) genes, representing ~70% of all MODY subtypes [6,7]. The prevalence of GCK/MODY is higher in some countries (United States, Germany, Italy, France, and Spain) [8–11]; this may be because this form of diabetes is more easily identified in childhood in these countries.

Clinical manifestations of GCK/MODY are characterized by a slight increase in blood fasting glucose (5.5–8.0 mmol/L; 99–144 mg/dL), which is often present at birth [12].
Affected individuals are usually asymptomatic and the hyperglycemia is often discovered during routine medical examinations, such as during pregnancy or family screening when MODY is suspected. Micro- or macrovascular complications are extremely rare and no pharmacological treatment is recommended, but diet and regular physical activity are sufficient to maintain good glycemic control. Only in pregnancy may there be a need for treatment with insulin [13].

HNF1A/MODY is characterized by a progressive reduction in insulin secretion and, consequently, hyperglycemia. This requires treatment with drugs, and is associated with a tendency to develop microvascular complications [5]. The onset of the disease usually occurs during adolescence with an initial post-prandial hyperglycemia, which is then generalized in the patient.

Recognition of GCK/ and HNF1A/MODY, particularly when the patient is of pediatric age, is of fundamental importance, because this allows us to select the appropriate clinical management and to conserve healthcare resources. In fact, diabetic patients with HNF1A/MODY can avoid insulin and be treated with oral hypoglycemic agents such as sulfonylureas, to which they are particularly sensitive, whereas GCK/MODY patients, due to their mild phenotype, require less frequent clinical monitoring than patients with other forms of diabetes [14,15]. Therefore, molecular diagnosis of MODY may provide a perceptible impact on both the patient’s quality of life and the healthcare costs [16,17].

Herein, we report a new case of a digenic variant, identified by next-generation sequence analysis (NGS), in a hyperglycemic pediatric patient with suspected MD.

2. Case Presentation

We report the case of a female patient, the only child of non-consanguineous parents. The family history showed fasting hyperglycemia (135 mg/dL-7.5 mmol/L) and elevated hemoglobin-glycosylated HbA1C (6.3%-45 mmol/mol) in the father, and similar values in the paternal grandmother. In both cases, the condition was not treated. In the maternal family, the great-grandparents had a history of type 2 diabetes mellitus (T2D). The father and mother were both obese, with a body mass index (BMI) of 36 and 39, respectively. The mother and the mother’s sisters had thrombophilia (Figure 1A).

![Pedigree of the analyzed family. The pedigree shows the segregation of the variants detected in the proband and her family.](image)

**Figure 1.** (A) Pedigree of the analyzed family. The pedigree shows the segregation of the variants detected in the proband and her family. In the subject III.1, the variant, carried in the heterozygous status, is the c.868 G>T; p.Glu290*, in the glucokinase (CGK) gene; the III.2 subject carried the c.872 C>G; p.Pro291Arg, in the HNF1A gene. The proband of the study (IV.1) shows all the two variants cited above in the parents. I.1 and I.2 proband’s maternal great-grandparents: diabetes; II.2 proband’s paternal grandmother: diabetes; III.1 proband’s father: diabetes, obesity; III.2 proband’s mother: obesity, thrombophilia; III.3 proband’s aunt: thrombophilia; IV.1: diabetes (B) GCK and HNF1A Sanger sequences of the proband, the mother, and the father.

The patient came to our attention at 7 years of age, as a result of several episodes of fasting hyperglycemia (maximum glucose value detected 134 mg/dL-7.44 mmol/L)
occasionally detected with a glucometer. Her medical history was characterized by birth at full-term from normal pregnancy, with her weight appropriate for gestational age (AGA). She did not present episodes of hypoglycemia at birth but she showed moderate to severe pulmonary valve stenosis, which was successfully treated with percutaneous valvuloplasty, and a speech delay that was diagnosed at 3 years of age.

Laboratory investigations showed glycemia 105 mg/dL (5.8 mmol/L), HbA1c 6.3% (45 mmol/mol). Her autoimmune diabetes autoantibodies (GAD, IAA, IA2, and ZnT8) were negative and her thyroid hormones (FT3, FT4, TSH) with autoantibodies (anti-tyroglobulin, anti-peroxidase) were normal. An oral glucose tolerance test (OGTT) showed impaired glucose tolerance (IGT) without insulin resistance, and her liver and renal function tests were normal, with an adequate lipid profile: total cholesterol 167 mg/dL (4.32 mmol/L), LDL cholesterol 111 mg/dL (2.87 mmol/L), HDL cholesterol 50 mg/dL (1.29 mmol/L), triglycerides 91 mg/dL (1.03 mmol/L). Her BMI was 22.7 kg/m² (BMIz-score: 2; >97th percentile), according to the curves of the World Health Organization (WHO), adjusted for age and sex. The patient was not undergoing any drug therapy.

Based on the early onset of the diabetes, the absence of beta-cell autoantibodies, and on the family history of diabetes, genetic tests for MD were performed.

3. Diagnostic Assessment

Peripheral blood samples for DNA analysis were collected after obtaining written informed consent from both the patient and her parents. Genomic DNA was extracted from leukocytes with a QiAamp DNA Blood Mini Kit (Qiagen, Hilden, Germany), according to the manufacturer’s instructions, and quantified by a NanoDrop ND-1000 spectrophotometer (ThermoFisher Scientific, Waltham, MA, USA). A next-generation sequencing (NGS) analysis, including 42 genes associated with non-autoimmune diabetes, was performed (Table 1). For each gene, we analyzed the coding regions, 50 bp in each of the intronic boundaries, the promoter, and the 3′UTR, for a total target size of about 1 Mb. A total of 50 ng of gDNA was processed through the SureSelectQXT Target Enrichment system (Agilent Technologies, Santa Clara, CA, USA) for Illumina multiplexed sequencing. Sequencing reactions were carried out on the MiSeq instrument (Illumina, San Diego, CA, USA). The sequence reads were aligned to the human reference genome (hg38) using the Alissa Align & Call v1.0.2.10 tool (Agilent Technologies, Santa Clara, CA, USA). The evidenced sequence variants were evaluated by Alissa Interpret v5.2.6 CE IVD software (Agilent Technologies, Santa Clara, CA, USA), using GRCh38.p2 and several databases of genomic variants [18].

Table 1. Panel of genes associated with monogenic diabetes, analysed in the proband.

| GENE PANEL | ABCC8 | CEL | GCK | HNF4A | KLF11 | PDX1 | SLC16A1 |
|------------|-------|-----|-----|-------|-------|-------|---------|
| AKT2       | CISD2 | GLIS3| IER3IP1| LMNA  | PLIN1 | SLC19A2|
| APPL1      | EIF2AK3| GLUD1| INS  | MAFA  | POLD1 | SLC9B1 |
| ASB14      | FOXP3 | HADH| INS-IGF2| NEUROD1| PPARG | TRMT10A|
| BLK        | GATA4 | HNF1A| INSR | NEUROG3| PTF1A | WFS1   |
| C12orf43   | GATA6 | HNF1B| KCNJ11| PAX4  | RFX6  | ZFP57  |

The Alamut software ([http://www.interactive-biosoftware.com/alamut-visual/](http://www.interactive-biosoftware.com/alamut-visual/)) Alamut Visual v2.11.0, accessed on 23 March 2021)) was used to determine the pathogenicity of all missense variants. Bioinformatic tools available online, such as Sorting Intolerant From Tolerant (SIFT) ([http://sift.jcvi.org/](http://sift.jcvi.org/), accessed on 8 March 2021) and Polymorphism Pheno-type v2 (PolyPhen-2) ([http://genetics.bwh.harvard.edu/pph2/](http://genetics.bwh.harvard.edu/pph2/), accessed on 8 March 2021) were utilized to classify identified variants. Further predictions were assessed with the Mutation Taster tool ([http://www.mutationtaster.org](http://www.mutationtaster.org), accessed on 8 March 2021) [19] and other tools included on the VarSome website ([https://varsome.com/variant/hg38](https://varsome.com/variant/hg38), accessed on 8 March 2021) [20,21]. All software programs were used with their default parameters. Finally, we classified the variants as pathogenic, likely pathogenic, variants of
uncertain significance (VOUS), likely benign, or benign, in accordance with the standards and guidelines of the American College of Medical Genetics and Genomics (ACMG) [22].

Two nucleotide variants in exon 8 (c.868 G > T; p.Glu290*) of the GCK gene and in exon 4 (c.872 C > G; p.Pro291Arg) of the HNF1A gene were identified. These variants were confirmed with standard Sanger sequencing. Molecular sequencing extended to the diabetic parents showed that the GCK variant was present in the father and the HNF1A variant was present in the mother (Figure 1B).

4. Discussion

Monogenic diabetes comprises a group of heterogeneous genetic disorders characterized by the early onset of diabetes, the absence of autoimmunity, and beta-cell dysfunction [8,23].

Recognition of these forms of diabetes is crucial for reducing both the complications and the treatment costs associated with the disease, and to improve glycemic control with the most appropriate treatment and follow-up for patients [17,24]. In particular, HNF1A/MODY patients are generally responsive to sulfonylureas, whereas, for GCK/MODY patients, no pharmacological treatment is recommended and diet and regular physical activity are sufficient to maintain good glycemic control [6,14].

In this case report, we described an unusual molecular diagnosis performed by NGS analysis in a hyperglycemic child, which revealed two distinct genetic variants in the GCK and HNF1A genes. The GCK variant, reported in the Human Gene Mutation Database (HGMD) (http://www.hgmd.cf.ac.uk/ac, accessed on 8 March 2021) and in the literature as associated with GCK/MODY [12], was also identified in the diabetic father. The patient presented a moderate hyperglycemia consistent with a GCK/MODY diagnosis.

The HNF1A variant, not reported in HGMD or in any known genomic variants database, has been classified as novel and its pathogenicity has been evaluated. In particular, according to ACMG classification criteria [21], we added fundamental parameters other than those by default in the VarSome database and therefore classified p.Pro291Arg as “likely pathogenic” because it has a low frequency (combined minor allele frequency, MAF < 1%), it is located in a mutational hotspot (PM1), it is a missense variant in a gene that has a low rate of benign missense variation (PP2), other different changes in 291 amino acid residues have been described as pathogenic (PM5), and a reputable source (NCBI ClinVar) (https://www.ncbi.nlm.nih.gov/, accessed on 8 March 2021) reports that the variant is likely pathogenic (PP5). Moreover, the variant is located in a conserved region among species, indicating its functional relevance, and wild-type amino acid Proline has physicochemical differences with respect to Arginine. Furthermore, because computational prediction tools were not all in accord, they were not considered as criteria for classifying the variant.

The p.Pro291Arg is also present in the mother (40 years) who has not shown glycemic alterations until now.

It is known that the clinical expression of HNF1A/MODY is highly variable, even within the same family [3,25]. In fact, HNF1A mutation carriers may be normoglycemic while their mutated siblings may be hyperglycemic, as reported by Miedzybrodzka et al. [26]. The authors described a family with a missense mutation in the HN1A gene that was not fully penetrant, because it was present in all diabetic family members even if with considerable variations in severity and age of diagnosis (ranging from 16 to 67 years), and in two non-diabetic family members (aged 87 and 46) [26]. This case confirms that, among the HNF1A variants, some are sufficient for diabetes onset; however, not all carriers of these variants have early-onset diabetes and some do not have diabetes at all. It was hypothesized that multiple loci, inherited independently of diabetes, are modifiers of the HNF1A/MODY phenotype [25,27]. Some of these loci may exert their effect at the level of the beta-cell, enhancing the insulin secretion deficit caused by HNF1A mutations; others may influence the onset of diabetes by affecting the amount of insulin that peripheral tissues require to maintain glycemia at the normal level [25].
This phenotypic variability and incomplete penetrance have been also described in HNF4A/MODY [28,29] and were found in our clinical experience too (unpublished data).

As HNF1A/MODY is usually onset at a post-pubertal age, we could hypothesize that our patient, thus far showing a mild phenotype compatible with GCK/MODY, due to the progressive beta-cell function failure caused by the HNF1A mutation, will develop a more severe phenotype later in life. However, studies on phenotypic effects in the general population of rare variants in MODY genes, previously reported as pathogenic, have provided evidence that the majority of variant carriers remain euglycemic through middle age, highlighting the limitations of disease variant databases, as well as bioinformatics criteria, in assigning pathogenicity to rare variants [30]. Genetic and functional studies have also demonstrated that missense variants in HNF1A can have a spectrum of effects, ranging from weak effects, predisposing patients to T2D, to highly penetrant effects, causing MODY [30,31].

Few reports are present in the literature describing individuals with digenic variants in different MODY genes [32–36]. One of these reported, for the first time, the co-inheritance of HNF1A and GCK mutations in two patients of the same family with the coexistence of two phenotypes, GCK/MODY and HNF1A/MODY [35]. In particular, the adult patient showed typical HNF1A/MODY characteristics and the presence of the second GCK mutation did not modify the diabetic phenotype, whereas the younger patient with the same genotype showed, as did our patient, only impaired fasting glucose (IFG), which was likely due to a shorter disease evolution.

This new case of digenic GCK/HNF1A variants, evidenced in a hyperglycemic pediatric patient by NGS analysis, confirms that the eventuality of two mutations in different MODY genes should not be excluded in patients with suspect of MD. In particular, in our proband, because we cannot exclude a possible evolution of the disease, a more careful clinical follow-up is necessary for accurate management and treatment. In addition, considering the relevance of HNF1A variants in the contribution of T2D risk, a greater attention to lifestyle and a correction of the weight is strongly suggested for the mother carrier of the novel HNF1A variant.

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References

1. Mozzillo, E.; Salzano, G.; Barbetti, F.; Maffeis, C.; Lombardo, F.; Franzese, A.; Delvecchio, M.; Marigliano, M. Survey on Etiological Diagnosis of Diabetes in 1244 Italian Diabetic Children and Adolescents: Impact of Access to Genetic Testing. *Diabetes Res. Clin. Pract.* 2015, 107, e15–e18. [CrossRef]

2. Vaxillaire, M.; Bonnefond, A.; Liatis, S.; Ben Salem Hachmi, L.; Jotic, A.; Boissel, M.; Gaget, S.; Durand, E.; Vaillant, E.; Derhourhi, M.; et al. Monogenic Diabetes Characteristics in a Transnational Multicenter Study from Mediterranean Countries. *Diabetes Res. Clin. Pract.* 2021, 171, 108553. [CrossRef]

3. Fajans, S.S.; Bell, G.I.; Polonsky, K.S. Molecular Mechanisms and Clinical Pathophysiology of Maturity-Onset Diabetes of the Young. *N. Engl. J. Med.* 2001, 345, 971–980. [CrossRef]
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4. Delvecchio, M.; Pastore, C.; Giordano, P. Treatment Options for MODY Patients: A Systematic Review of Literature. *Diabetes Ther.* 2020, 11, 1667–1685. [CrossRef]

5. Nkonge, K.M.; Nkonge, D.K.; Nkonge, T.N. The Epidemiology, Molecular Pathogenesis, Diagnosis, and Treatment of Maturity-Onset Diabetes of the Young (MODY). *Clin. Diabetes Endocrinol.* 2020, 6, 20. [CrossRef] [PubMed]

6. Ellard, S.; Bellanné-Chantelot, C.; Hattersley, A.T. European Molecular Genetics Quality Network (EMQN) MODY group Best Practice Guidelines for the Molecular Genetic Diagnosis of Maturity-Onset Diabetes of the Young. *Diabetologia* 2008, 51, 546–553. [CrossRef]

7. Capuano, M.; Garcia-Herrero, C.M.; Tinto, N.; Carluccio, C.; Capobianco, V.; Coto, I.; Cola, A.; Iafusco, D.; Franzese, A.; Zagari, A.; et al. Glucokinase (GCK) Mutations and Their Characterization in MODY2 Children of Southern Italy. *PLoS ONE* 2012, 7, e38906. [CrossRef] [PubMed]

8. Delvecchio, M.; Mozzillo, E.; Salzano, G.; Iafusco, D.; Frontino, G.; Patera, P.I.; Rabbone, I.; Cherubini, V.; Grasso, V.; Tinto, N.; et al. Monogenic Diabetes Accounts for 6.3% of Cases Referred to 15 Italian Pediatric Diabetes Centers During 2007 to 2012. *J. Clin. Endocrinol. Metab.* 2017, 102, 1826–1834. [CrossRef]

9. Estalella, I.; Rica, I.; Perez de Nanclares, G.; Bilbao, J.R.; Vazquez, J.A.; San Pedro, J.I.; Busturia, M.A.; Castaño, L. Spanish MODY Group Mutations in GCK and HNF-1α-Deficient Explain the Majority of Cases with Clinical Diagnosis of MODY in Spain. *Clin. Endocrinol.* 2007, 67, 538–546. [CrossRef] [PubMed]

10. Schober, E.; Rami, B.; Grabert, M.; Thon, A.; Kapellen, T.; Holl, R.W. DPV-Wiss Initiative of the German Working Group for Paediatric Diabetology and Phenotypical Aspects of Maturity-Onset Diabetes of the Young (MODY Diabetes) in Comparison with Type 2 Diabetes Mellitus (T2DM) in Children and Adolescents: Experience from a Large Multicentre Database. *Diabet. Med.* 2009, 26, 466–473. [CrossRef] [PubMed]

11. Carmody, D.; Naylor, R.N.; Bell, C.D.; Berry, S.; Montgomery, J.T.; Tadie, E.C.; Hwang, J.L.; Greeley, S.A.W.; Philipson, L.H. GCK-MODY in the US National Monogenic Diabetes Registry: Frequently Misdiagnosed and Unnecessarily Treated. *Acta Diabetol.* 2016, 53, 703–708. [CrossRef] [PubMed]

12. Tinto, N.; Zagari, A.; Capuano, M.; De Simone, A.; Capobianco, V.; Daniele, G.; Giugliano, M.; Spadaro, R.; Franzese, A.; Sacchetti, L. Glucokinase Gene Mutations: Structural and Genotype-Phenotype Analyses in MODY Children from South Italy. *PLoS ONE* 2008, 3, e1870. [CrossRef] [PubMed]

13. Bitterman, O.; Tinto, N.; Franzese, A.; Iafusco, F.; Festa, C.; Mozzillo, E.; Napoli, A.; Iafusco, D. Glucokinase Deficit and Birthweight: Does Maternal Hyperglycemia Always Meet Fetal Needs? *Acta Diabetol.* 2018, 55, 1247–1250. [CrossRef] [PubMed]

14. Delvecchio, M.; Salzano, G.; Bonurà, C.; Cauvin, V.; Cherubini, V.; d’Annunzio, G.; Franzese, A.; Giglio, S.; Graziano, V.; et al. Can HbA1c Combined with Fasting Plasma Glucose Help to Assess Priority for GCK-MODY vs HNF1A-MODY Genetic Testing? *Acta Diabetol.* 2018, 55, 981–983. [CrossRef] [PubMed]

15. Iafusco, F.; De Sanctis, P.; Pirozzi, D.; Capone, S.; Lombardo, B.; Gambale, A.; Confetto, S.; Zanfardino, A.; Iolascon, A.; Pastore, L.; et al. Molecular Diagnosis of MODY3 Permitted to Reveal a de Novo 12q24.31 Deletion and to Explain a Complex Phenotype in a Young Diabetic Patient. *Clin. Chem. Lab. Med.* 2019, 57, e306–e310. [CrossRef]

16. Pinelli, M.; Acquaviva, F.; Barbetti, F.; Caredda, E.; Coccozza, S.; Delvecchio, M.; Mozzillo, E.; Pirozzi, D.; Prisco, F.; Rabbone, I.; et al. Identification of Candidate Children for Maturity-Onset Diabetes of the Young Type 2 (MODY2) Gene Testing: A Seven-Item Clinical Flowchart (7-IF). *PLoS ONE* 2013, 8, e79933. [CrossRef]

17. Szopa, M.; Matejko, B.; Ucieklak, D.; Uchman, A.; Hodenhoff, J.; Mrozinska, S.; Glodzicki, W.; Zapala, B.; Platek, T.; Solecza, I.; et al. Quality of Life Assessment in Patients with HNF1A-MODY and GCK-MODY. *Endocrine* 2019, 64, 246–253. [CrossRef]

18. Limongelli, G.; Nunziato, M.; D’Argenio, V.; Esposito, M.V.; Monda, E.; Mazzuccara, C.; Caiazza, M.; D’Aponte, A.; D’Andrea, A.; Bossone, E.; et al. Yield and Clinical Significance of Genetic Screening in Elite and Amateur Athletes. *Europ. J. Prev. Cardiol.* 2020, 2047487320934265. [CrossRef]

19. Schwarz, J.M.; Cooper, D.N.; Schuelke, M.; Seelow, D. MutationTaster2: Mutation Prediction for the Deep-Sequencing Age. *Nat. Methods* 2014, 11, 361–362. [CrossRef]

20. Pion, S.E.; Eccles, D.M.; Easton, D.; Foulkes, W.D.; Genuardi, M.; Greenblatt, M.S.; Hogervorst, F.B.L.; Hoogerbrugge, N.; Spurdle, A.B.; Tavtigian, S.V.; et al. Sequence Variant Classification and Reporting: Recommendations for Improving the Interpretation of Cancer Susceptibility Genetic Test Results. *Hum. Mutat.* 2008, 29, 1282–1291. [CrossRef]

21. Kopanos, C.; Tsioikas, V.; Kouris, A.; Chapple, C.E.; Albarca Aguilara, M.; Meyer, R.; Massouras, A. VarSome: The Human Genomic Variant Search Engine. *Bioinformatics* 2019, 35, 1978–1980. [CrossRef] [PubMed]

22. Richards, S.; Aziz, N.; Bale, S.; Bick, D.; Was, S.; Gastier-Foster, J.; Grody, W.W.; Hegde, M.; Lyon, E.; Spector, E.; et al. Standards and Guidelines for the Interpretation of Sequence Variants: A Joint Consensus Recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet. Med.* 2015, 17, 405–424. [CrossRef]

23. Mazzuccara, C.; Iafusco, D.; Liguori, R.; Ferrigno, M.; Galderisi, A.; Vitale, D.; Simonelli, F.; Landolfo, P.; Prisco, F.; Masullo, M.; et al. Mitochondrial Diabetes in Children: Seek and You Will Find It. *PLoS ONE* 2012, 7, e34956. [CrossRef] [PubMed]

24. Bennett, J.T.; Vasta, V.; Zhang, M.; Narayanan, J.; Gerrits, P.; Hahn, S.H. Molecular Genetic Testing of Patients with Monogenic Diabetes and Hyperinsulinism. *Mol. Genet. Metab.* 2015, 114, 451–458. [CrossRef]

25. Kim, S.H.; Ma, X.; Klupa, T.; Powers, C.; Pezzolesi, M.; Warram, J.H.; Rich, S.S.; Krolevets, A.S.; Doria, A. Genetic Modifiers of the Age at Diagnosis of Diabetes (MODY3) in Carriers of Hepatocyte Nuclear Factor-1alpha Mutations Maps to Chromosomes 5p15, 9q22, and 14q24. *Diabetes* 2003, 52, 2182–2186. [CrossRef] [PubMed]
26. Miedzybrodzka, Z.; Hattersley, A.T.; Ellard, S.; Pearson, D.; de Silva, D.; Harvey, R.; Haites, N. Non-Penetrance in a MODY 3 Family with a Mutation in the Hepatic Nuclear Factor Ialpha Gene: Implications for Predictive Testing. *Eur. J. Hum. Genet.* 1999, 7, 729–732. [CrossRef]

27. Locke, J.M.; Saint-Martin, C.; Laver, T.W.; Patel, K.A.; Wood, A.R.; Sharp, S.A.; Ellard, S.; Bellanné-Chantelot, C.; Hattersley, A.T.; Harries, L.W.; et al. The Common HNF1A Variant I27L Is a Modifier of Age at Diabetes Diagnosis in Individuals with HNF1A-MODY. *Diabetes* 2018, 67, 1903–1907. [CrossRef] [PubMed]

28. Spiro, A.J.; Vu, K.N.; Warnock, A.L. An Atypical HNF4A Mutation Which Does Not Conform to the Classic Presentation of HNF4A-MODY. *Case Rep. Endocrinol.* 2018, 2018, 1560472. [CrossRef] [PubMed]

29. Laver, T.W.; Colclough, K.; Shepherd, M.; Patel, K.; Houghton, J.A.L.; Dusatkova, P.; Pruhovala, S.; Morris, A.D.; Palmer, C.N.; McCarthy, M.I.; et al. The Common p.R114W HNF4A Mutation Causes a Distinct Clinical Subtype of Monogenic Diabetes. *Diabetes* 2016, 65, 3212–3217. [CrossRef] [PubMed]

30. Flannick, J.; Beer, N.L.; Bick, A.G.; Agarwala, V.; Molnes, J.; Gupta, N.; Burtt, N.P.; Florez, J.C.; Meigs, J.B.; Taylor, H.; et al. Assessing the Phenotypic Effects in the General Population of Rare Variants in Genes for a Dominant Mendelian Form of Diabetes. *Nat. Genet.* 2013, 45, 1380–1385. [CrossRef] [PubMed]

31. Najmi, L.A.; Aukrust, I.; Flannick, J.; Molnes, J.; Burtt, N.; Molven, A.; Groop, L.; Alshuler, D.; Johansson, S.; Bjørkhauge, L.; et al. Functional Investigations of HNF1A Identify Rare Variants as Risk Factors for Type 2 Diabetes in the General Population. *Diabetes* 2017, 66, 335–346. [CrossRef] [PubMed]

32. Shankar, R.K.; Ellard, S.; Standiford, D.; Pihoker, C.; Gilliam, L.K.; Hattersley, A.; Dolan, L.M. Digenic Heterozygous HNF1A and HNF4A Mutations in Two Siblings with Childhood-Onset Diabetes. *Pediatr. Diabetes* 2013, 14, 535–538. [CrossRef]

33. Karges, B.; Bergmann, C.; Scholl, K.; Heinze, E.; Rasche, F.M.; Zerres, K.; Debatin, K.-M.; Wabitsch, M.; Karges, W. Digenic Inheritance of Hepatocyte Nuclear Factor-Ialpha and -Ibeta with Maturity-Onset Diabetes of the Young, Polycystic Thyroid, and Urogenital Malformations. *Diabetes Care* 2007, 30, 1613–1614. [CrossRef] [PubMed]

34. Forlani, G.; Zucchini, S.; Di Rocco, A.; Di Luzio, R.; Scipione, M.; Marsac, E.; Romeo, G.; Marchesini, G.; Mantovani, V. Double Heterozygous Mutations Involving Both HNF1A/MODY3 and HNF4A/MODY1 Genes: A Case Report. *Diabetes Care* 2010, 33, 2336–2338. [CrossRef] [PubMed]

35. López-Garrido, M.P.; Herranz-Antolin, S.; Alija-Merillas, M.J.; Giralt, P.; Eiscrbo, J. Co-Inheritance of HNF1a and GCK Mutations in a Family with Maturity-Onset Diabetes of the Young (MODY): Implications for Genetic Testing. *Clin. Endocrinol.* 2013, 79, 342–347. [CrossRef] [PubMed]

36. Yang, Y.; Zhou, T.-C.; Liu, Y.-Y.; Li, X.; Wang, W.-X.; Irwin, D.M.; Zhang, Y.-P. Identification of HNF4A Mutation p.T130I and HNF1A Mutations p.I27L and p.S487N in a Han Chinese Family with Early-Onset Maternally Inherited Type 2 Diabetes. *J. Diabetes Res.* 2016, 2016, 3582616. [CrossRef]