Is the Phenotype Designation by PSP-MDS Criteria Stable Throughout the Disease Course and Consistent With Tau Distribution?

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Introduction: The MDS-PSP criteria have shown high sensitivity for the PSP diagnosis, but do not discriminate the phenotype diversity. Our purpose was to search for anatomopathological differences among PSP phenotypes resulting from the application of the MDS-PSP criteria comparing with the previous ones.

Methods: Thirty-four PSP cases from a single brain bank were retrospectively classified according to the criteria used by Respondek et al. in 2014 and the PSP-MDS criteria at 3 years (MDS-3y), 6 years (MDS-6y) and at the last clinical evaluation before death (MDS-last). Semiquantitative measurement of total, cortical and subcortical tau load was compared. For comparative analysis, PSP-Richards on syndrome and PSP postural instability were grouped (PSP-RS/PI) as well as the PSP atypical cortical phenotypes (PSP-Cx).

Results: Applying the Respondek’s criteria, PSP phenotypes were distributed as follow: 55.9% PSP-RS/PI, 26.5% PSP-Cx, 11.8% PSP-Parkinsonism (PSP-P), and 5.9% PSP-Cerebellum. PSP-RS/PI and PSP-Cx had a higher total tau load than PSP-P; PSP-Cx showed a higher cortical tau load than PSP-RS/PI and PSP-P; and PSP-RS/PI had a higher subcortical tau load than PSP-P. Applying the MDS-3y, MDS-6y and MDS-last criteria; the PSP-RS/PI group increased (67.6, 70.6 and 70.6% respectively) whereas the PSP-Cx group decreased (8.8, and 8.8 and 11.8%). Then, only differences in total and subcortical tau burden between PSP-RS/PI and PSP-P were observed.

Interpretation: After the retrospective application of the new MDS-PSP criteria, total and subcortical tau load is higher in PSP-RS/PI than in PSP-P whereas no other differences in tau load between phenotypes were found, as a consequence of the loss of phenotypic diversity.

Keywords: tauopathies, progressive supranuclear palsy (PSP), phenotypes, PSP-MDS criteria, tau protein load, clinical-pathological correlation
INTRODUCTION

Progressive supranuclear palsy (PSP) is a neurodegenerative disease characterized pathologically by the accumulation of hyperphosphorylated four-repeat (4R) tau protein within neurons (neurofibrillary tangles) and glial cells (tufted astrocytes and coiled bodies) (1).

The clinical diagnostic criteria established in 1996 by the National Institute of Neurological Disorders and Stroke (NINDS) (2) have high specificity for the diagnosis of the most frequent clinical presentation and original description of the disease, Richardson syndrome (PSP-RS), which is characterized by gait instability and falls, ophthalmoplegia, pseudobulbar signs and cognitive impairment. However, NINDS-PSP criteria have low sensitivity for the diagnosis of the atypical PSP phenotypes that have been described throughout the last 20 years, especially in early stages of the disease (3–7). For this reason, the movement disorders society (MDS) has recently reformulated PSP diagnostic criteria (8). The new MDS-PSP criteria categorize PSP symptoms into four clinical domains: ocular motor, postural instability, akinesia and cognitive dysfunction. Different combinations of these symptoms have defined various PSP phenotypes: PSP-RS, PSP-predominant postural instability (PSP-PI); PSP-predominant ocular motor dysfunction (PSP-OM); PSP-predominant parkinsonism (PSP-P); PSP-coverting gait freezing (PSP-PGF); PSP-predominant corticobasal syndrome (PSP-CBS); PSP-predominant frontal presentation (PSP-F) and PSP-predominant speech/language disorder (PSP-SL).

The pathogenic basis underlying these PSP phenotypes seems to be due to the different load and distribution of tau protein in a variety of cell types across encephalic regions (9, 10). This hypothesis is derived from studies showing more severe and widespread tau deposits in PSP-RS than in the subcortical phenotypes (3) and higher tau accumulation in neocortical regions in PSP-CBS (11), PSP-F (6), and PSP-SL (10) than in PSP-RS. In addition, to differences in overall tau burden, clinical phenotypes can also be differentiated based on the different cell types involved (neurons, astrocytes, oligodendroglia) (10).

The application of the new MDS-PSP criteria in clinical setting has confirmed a higher sensitivity than the previous criteria (12). However, they have demonstrated a lower phenotype diversity with higher representation of PSP-PI (13) and poor accuracy in differentiating PSP-RS from PSP-P phenotypes (14), although some of these studies have not included autopsy-confirmed diagnosis (15).

Prior to the publication of the new MDS-PSP criteria, Respondek et al. (16) studied the phenotypic spectrum of PSP by retrospective chart review in a cohort of 100 autopsy-confirmed PSP patients, constituting the first quantitative description of the relative distribution of PSP clinical phenotypes defined in a large multicenter cohort.

After the MDS-PSP criteria description, the study of Kovacs et al. (10) was the first one comparing tau load among PSP phenotypes and no other similar studies have been published afterwards.

The aim of our study was to search for anatomopathological differences regarding tau load and distribution among PSP phenotypes of 34 definitive PSP cases from the same brain bank resulting from the application of the criteria applied by Respondek et al. (16) and the new MDS-PSP diagnostic criteria (8).

MATERIALS AND METHODS

Subjects

Thirty-four pathologically confirmed PSP cases from the NavarraBiomed Brain Bank (2005–2017) were included. The study was performed under the ethics guidelines issued by our institution and written informed consent was obtained from all participants or family caregivers for pathological brain studies.

Classification Into PSP Clinical Phenotypes

We performed a retrospective review of the clinical charts of all patients included with a clinical diagnosis of PSP. Experienced movement disorders neurologists examined all patients and the clinical items not specifically mentioned in the clinical records were considered as absent. The age of onset (age at first symptom related with PSP), disease duration (years), sex and core clinical features were recorded for each case. Patients were retrospectively allocated into the different predominant phenotypes according to the criteria applied by Respondek in 2014 (Respondek’s criteria, Supplementary Table 1) (16). Moreover, they were classified into PSP-RS, PSP-PI, PSP-P, PSP-SL, PSP-F and PSP-CBS at 3 and 6 years of disease evolution (MDS-3y and MDS-6y respectively) and at the last clinical evaluation before death (MDS-last) applying the new MDS-PSP criteria (8). If a patient was deceased before that period, the data from the last evaluation was considered. The MAX rules (17) were also implemented when multiple alternatives were present for a single patient. Cases, which did not meet the clinical criteria of any phenotype, were classified as unclassified (PSP-U).

PSP-RS and PSP-PI phenotypes were grouped as PSP-RS spectrum (PSP-RS/PI) considering PSP-PI as a transitional form of PSP-RS (18). The PSP-SL, PSP-F and PSP-CBS were grouped as PSP cortical predominant phenotypes (PSP-Cx) for comparative analysis.

Neuropathological Examination and Immunohistochemistry

Brain processing was made according to the recommendation guide proposed by BrainNet Europe (19). Routine workflow included immunohistochemical staining of 3–5 μm-thick paraffin-embedded sections, followed by counterstaining with hematoxylin-eosin. Tau pathology detection was performed with a mouse monoclonal antibody anti-human PHF-TAU, clone AT8, (Innogenetics, Ghent, Belgium). Presence of other protein deposits were analyzed using a mouse monoclonal antibody against α-synuclein (NCL-L-ASYN; LeicaBiosystems) or an anti-phospho TDP-43 monoclonal antibody (TIP, PTDMO1, Cosmo Bio). Following incubation with the primary antibody, the sections were incubated with EnVision + system peroxidase (Dako, Agilent Technologies, Santa Clara, CA, USA) for 30 min at room temperature. The peroxidase reaction was visualized with diaminobenzidine and H2O2. Control
of the immunostaining included omission of the primary antibody; no signal was obtained following incubation with only the secondary antibody. Antibody omission was used as a negative control for all staining. Neuropathological examination was completed following the revised NINDS criteria (20) for PSP diagnosis and the assessment of others co-pathologies (Alzheimer disease, Lewy bodies disease, Multiple System Atrophy, TDP 43 frontotemporal lobar degeneration and argyrophilic grain disease) were performed according to their specific recommendations (21–23).

**Regional tau Protein Density Quantification**

For each subject, 21 brain regions stained with an antibody anti-human PHF-TAU, were analyzed by an investigator blinded to clinical data. These areas included: cortical regions (primary motor cortex, middle frontal cortex, parietal cortex, and their adjacent white matters), subcortical regions (putamen, globus pallidus, basal nucleus of Meynert, medial thalamus, subthalamic nucleus, substantia nigra pars compacta, midbrain tectum, ventral part of the pons, locus coeruleus and inferior olivary nucleus) and cerebellar regions (dentate nucleus, cerebellar cortex and white matter). The hippocampus region was excluded to avoid AD-related bias.

The presence of neurofibrillary tangles, neuropil threads, tufted astrocytes and oligodendrogial coiled bodies were graded using four categories semiquantitative analysis (0 = absent, + = mild, ++ = moderate, +++ = severe) (Supplementary Figure 1). Total PSP-tau burden was obtained by averaging all scores. Cortical PSP-tau and a subcortical PSP-tau burden were obtained by averaging cortical and subcortical regions respectively, excluding cerebellar structures. In addition, tau staining in neurons (tangles and neuritophilic threads), astrocytes and oligodendrocytes (coiled bodies and neuropil threads in the white matter) were averaged separately.

**Analysis of Genetic Variants**

Genomic DNA was isolated from frozen frontal cortex tissue by phenol-chloroform method (24). APOE genotyping was performed using polymerase chain reaction restriction fragment length polymorphism (PCR-RFLP) (25). APOE genotypes were analyzed after digesting amplified sequences with 5 units of Hhal enzyme at 37°C overnight. H1/H2 haplotypes were assessed based on the MAPT

| bp deletion/insertion variant located at intron 9 as previously described (26).

**Statistical Analysis**

Data from continuous variables were expressed by mean ± standard deviation and categorical variables by frequencies and percentage. Cox's regression adjusted for age of onset and total tau burden were used to compare disease duration between PSP RS/PI and the other phenotypes (PSP-Cx + PSP-P). Following the recent findings of other authors (27) we also compared disease duration between PSP-P and the other phenotypes (PSP-RS/PI + PSP-Cx). Linear regressions were computed to evaluate group differences in tau burdens among PSP phenotypes adjusting by age of onset and Braak stage and to analyze the association between tau burdens and disease duration adjusting by age of onset and Braak stage. Linear regressions were also computed to evaluate the association between MAPT or APOE genotype and tau burdens or disease duration adjusting by age of onset and Braak stage. Significance level for all comparisons was set at p value < 0.05. Statistical analysis was performed with SPSS software version 21.0 (IBM, Inc., USA).

**RESULTS**

**PSP Phenotypes Distribution**

The distribution of the PSP clinical phenotypes is summarized in Figure 1 and Supplementary Table 2. When applying the Respondek's criteria for predominant PSP phenotypes (16) we found the following distribution. 55.9% of patients exhibited a PSP-RS/PI phenotype (20.6% PSP-RS and 35.3% PSP-PI), 26.5% displayed a PSP-Cx variant (11.8% PSP-Progressive Non-Fluent Aphasia [PSP-PPNFA], 8.8% of patients had a PSP-Frontotemporal Dysfunction phenotype [PSP-FTD], and 5.9% PSP-CBS), 11.8% PSP-P, and 5.9% PSP-Cerebellum (PSP-C, not recognized in the new MDS-PSP classification). After applying the MDS-3y criteria, PSP phenotypes were distributed as follow: 67.6% PSP-RS/PI (47.1% PSP-RS and 20.6% PSP-PI), 17.6% PSP-U, 8.8% PSP-Cx (8.8% PSP-SL) and 5.9% PSP-P. Using the MDS-6y criteria, a different phenotypes distribution was obtained: 70.6% PSP-RS/PI (61.8% PSP-RS and 8.8% PSP-PI), 11.8% PSP-P, 8.8% PSP-Cx (5.9% PSP-SL and 2.9% PSP-F) and 8.8% PSP-U. Finally, applying MDS-last criteria the distribution was: 70.6% PSP-RS/PI (64.7% PSP-RS and 5.9% PSP-PI), 17.6% PSP-P, 11.8% PSP-Cx (8.8% PSP-SL and 2.9% PSP-F) and no PSP-U cases. No case ever met the criteria for PSP-PGF or PSP-OM.

**Demographics and Disease Duration**

The individual patient demographic characteristics are summarized in Table 1. Briefly, 52.9% of the patients were female and the average age of onset (SD) and disease duration were 72.3 (7.3) and 8 (4) years respectively. The mean time to diagnosis (SD) was 4.6 (4.2) years and Premor tem diagnosis accuracy rate in the last clinical evaluation was 91.2%. Case 30 and 32, classified as PSP-P according to the Respondek's and MDS-last criteria, were misdiagnosed in life as Parkinson's disease (PD); and case 25 allocated to PSP-PPNFA or PSP-SL was misdiagnosed as frontotemporal dementia.

The PSP-RS/PI phenotype had shorter disease duration than the other phenotypes (PSP-P + PSP-Cx) in the resultant groups after applying Respondek's and MDS-PSP criteria (MDS-6y and MDS-Last), adjusting for age at onset and total tau burden (Supplementary Table 3). When comparing PSP-P with the other phenotypes (PSP-RS + PSP-Cx), the PSP-RS/PI + PSP-Cx group had shorter disease duration than PSP-P applying MDS-last criteria with similar hazard ratio and no other significant differences were found (Supplementary Table 3).

**Neuropathological Description**

The individual pathological features are summarized in Table 1. The 32.4% of the patients exhibited exclusively...
FIGURE 1 | PSP phenotype distribution. The figure shows the PSP phenotype distribution after applying the criteria used by Respondek et al. in (16) (Respondek) or the Movement Disorders Society criteria and MAX rules at 3 years of disease evolution (MDS-3y), at 6 years (MDS-6y) and at last clinical evaluation before to death (MDS-last). PSP-RS, Richardson’s syndrome; PSP-PI, PSP with postural instability; PSP-FTD or PSP-F, PSP with predominant frontal presentation; PSP-PNFA or PSP-SL, PSP with predominant speech/language disorder; PSP-CBS, PSP with corticobasal syndrome; PSP-P, PSP with predominant parkinsonism; PSP-C, PSP with cerebellar presentation; PSP-U, unclassified PSP.
pathological findings of PSP. Alzheimer disease (AD) was the most frequent co-existing pathology (47.1%), with moderate-advanced stages (Braak stage 3–6) in 14.7% of cases. α-synuclein deposits were found in 11.8% of PSP cases (4 cases), which belonged to the PSP-RS/PI phenotype independently of the moment or classifying criteria employed. One case (number 18) showed Lewy bodies limited to the brainstem, and the other 3 cases displayed a limbic-transitional distribution. In addition, in case 9, hippocampal TDP-43 protein deposits were also identified. Finally, amyloid angiopathy and limbic argyrophilic grains were found in 8 and 2 cases respectively.

### Tau Burden Distribution
The distribution of tau load across the different brain regions of each case is represented in a heat map in Figure 2. Considering all cases, tau pathology was found more abundantly in subcortical structures. Differences in total, cortical and subcortical tau burden among phenotypes adjusted by age of onset and AD Braak stage are shown in Figures 3A–D.

Applying the Respondek’s criteria (Figure 3A), PSP-RS/PI and PSP-Cx had higher total tau burden than PSP-P. Cortical tau load was significantly more abundant in PSP-Cx than in PSP-RS/PI and PSP-P whereas differences in subcortical tau burden where only found between PSP-RS/PI and PSP-P.

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**TABLE 1 | Demographic and pathological features of the sample.**

| Case ID | Sex | Age of onset (yr.) | Disease duration (yr.) | Premortem diagnosis | Time to diagnosis (yr.) | Copathology | Braak stage | APOE genotype | MAPT genotype |
|---------|-----|--------------------|------------------------|---------------------|------------------------|-------------|-------------|---------------|---------------|
| 34      | F   | 70, 8              | 17, 5                  | PSP                 | 7, 8                   | AD          | 1           | E3/E3         | H1/H1         |
| 33      | M   | 71, 9              | 8, 4                   | PSP                 | 3, 4                   | AD          | 1           | n.a.          | H1/H1         |
| 32      | M   | 68, 7              | 17                     | PD                  | –                      | AG          | 0           | E3/E3         | H1/H1         |
| 31      | M   | 61, 8              | 8, 3                   | PSP                 | 6                      | none        | 0           | E3/E3         | H1/H1         |
| 30      | M   | 60, 3              | 18, 3                  | PD                  | –                      | AD          | 3           | E2/E4         | H1/H2         |
| 29      | F   | 72, 4              | 9, 3                   | PSP                 | 3, 8                   | AD          | 2           | E3/E3         | H1/H1         |
| 28      | M   | 65                 | 12                     | PSP                 | 10                     | none        | 0           | E3/E3         | H1/H1         |
| 27      | M   | 68, 3              | 11, 4                  | PSP                 | 5, 3                   | AA          | 0           | E3/E3         | H1/H1         |
| 26      | F   | 68, 5              | 9, 9                   | PSP                 | 3, 5                   | none        | 0           | E3/E3         | H1/H1         |
| 25      | F   | 81, 3              | 10, 2                  | FTD                 | –                      | AD          | 3           | E3/E4         | H1/H2         |
| 24      | M   | 72, 6              | 9, 3                   | PSP                 | 6, 5                   | AA          | 0           | E3/E3         | H1/H1         |
| 23      | F   | 73, 1              | 6                      | PSP                 | 5, 2                   | AD          | 2           | E3/E4         | H1/H1         |
| 22      | M   | 73, 1              | 4, 1                   | PSP                 | 2, 1                   | LBD, AG     | 0           | E3/E3         | H1/H1         |
| 21      | M   | 64, 6              | 6, 2                   | PSP                 | 3, 8                   | none        | 0           | E3/E3         | H1/H1         |
| 20      | F   | 75, 2              | 10, 5                  | PSP                 | 4, 4                   | none        | 0           | E3/E3         | H1/H2         |
| 19      | F   | 66, 6              | 2, 6                   | PSP                 | 2, 2                   | AD          | 2           | E3/E4         | H2/H2         |
| 18      | M   | 77, 4              | 4, 1                   | PSP                 | 1, 7                   | LBD         | 0           | E3/E3         | H1/H1         |
| 17      | F   | 79, 4              | 9, 5                   | PSP                 | 7, 8                   | AD, AA      | 1           | E3/E3         | H1/H1         |
| 16      | F   | 64, 7              | 6, 9                   | PSP                 | 4, 2                   | none        | 0           | E3/E3         | H1/H1         |
| 15      | F   | 74                 | 11, 6                  | PSP                 | 5, 4                   | AD, AA      | 4           | E3/E3         | H1/H1         |
| 14      | F   | 83, 8              | 4, 3                   | PSP                 | 2, 6                   | AA          | 0           | E3/E3         | H1/H1         |
| 13      | M   | 64, 3              | 6                      | PSP                 | 3                      | none        | 0           | E3/E3         | H1/H1         |
| 12      | F   | 56, 4              | 7, 3                   | PSP                 | 2, 6                   | None        | 0           | E3/E3         | H1/H1         |
| 11      | M   | 64, 2              | 5, 8                   | PSP                 | 3                      | None        | 0           | n.a.          | n.a.          |
| 10      | F   | 78, 3              | 3, 8                   | PSP                 | 2, 4                   | AD, AA      | 1           | E3/E3         | H1/H1         |
| 9       | F   | 79, 6              | 11                     | PSP                 | 3, 1                   | AD, LBD, TDP43 | 2   | E2/E3         | H1/H2         |
| 8       | F   | 76, 6              | 8, 6                   | PSP                 | 2, 7                   | AD          | 2           | E3/E3         | H1/H1         |
| 7       | M   | 77, 5              | 4, 5                   | PSP                 | 2, 6                   | AA          | 0           | E3/E3         | H1/H1         |
| 6       | F   | 88, 6              | 3, 4                   | PSP                 | 0                      | AD, AA      | 1           | n.a.          | n.a.          |
| 5       | M   | 82                 | 4                      | PSP                 | 1                      | None        | 0           | E3/E3         | H1/H1         |
| 4       | M   | 73                 | 6, 2                   | PSP                 | 0, 9                   | AD          | 2           | E3/E3         | H1/H1         |
| 3       | F   | 62, 1              | 5, 2                   | PSP                 | 2, 2                   | none        | 0           | E3/E3         | H1/H1         |
| 2       | M   | 79                 | 5, 3                   | PSP                 | 1, 5                   | AD          | 4           | n.a.          | n.a.          |
| 1       | F   | 84                 | 4, 4                   | PSP                 | 0                      | AD, LBD     | 4           | E3/E3         | H1/H1         |

ID, identification; yr., years; F, female; M, male; PD, Parkinson disease; FTD, frontotemporal dementia; AD, Alzheimer disease; AG, argyrophilic grains; AA, amyloid angiopathy; LBD, Lewy bodies disease; TDP43, hippocampal TDP-43 protein deposits. n.a, not available.
Regarding cellular tau pathology (Supplementary Figure 2), the PSP-Cx group showed higher total and cortical neuronal tau burden than PSP-P ($p < 0.05$) and higher neuronal, astroglial and oligodendrocyte cortical tau burden than PSP-RS/PI ($p < 0.05$).

When applying the MDS-3y criteria (Figure 3B), only significant differences in total and subcortical tau burden between PSP-RS/PI and PSP-P were observed. Neuronal subcortical tau burden was more abundant in PSP-RS/PI than in PSP-P. Moreover, higher astroglial cortical tau burden was found in PSP-Cx than PSP-RS/PI (Supplementary Figure 3).

When applying MDS-6y and MDS-last criteria (Figures 3C,D), only subcortical tau load was found significantly higher in PSP-RS/PI than in PSP-P without differences in cellular tau burden.

In the simple linear regression adjusting by age of onset and AD Braak stage, there was no association between total, cortical or subcortical tau burden and disease duration in the overall sample.

**DISCUSSION**

The new MDS-PSP classification (8) resulted from the need to establish sensitive and specific clinical criteria for the diagnosis of the different PSP phenotypes that have been described after the 1996 definition of the NINDS-PSP criteria (2), which are very specific for the PSP-RS phenotype, but very insensitive for the atypical variants. To date, several retrospective studies validating the new MDS-PSP criteria have been published (12–15) confirming that these criteria are much more sensitive and will likely lead to better detection of PSP vs. other neurodegenerative diseases being also feasible to apply in real clinical settings. Regarding the accuracy of these sets of criteria for differentiating among the various PSP phenotypes, their usefulness has been shown to be limited since they may overestimate the presence of PSP-RS and PSP-P phenotypes in detriment of the cortical phenotypes (13). This is noteworthy, as one of the main goals of the new MDS-PSP criteria has been to allow earlier diagnosis of PSP in general, but specifically of the atypical variants of PSP. These studies, some of which have no pathological correlate, have settled the need to redefine or revise the new MDS-PSP criteria (13, 15). On the other hand, to date, there are no pathological criteria to differentiate the phenotypes of PSP, but it seems clear that they depend on the different tau load and distribution within encephalic regions and to the different patterns of cellular tau pathologies (10). Taking into account all these data, we studied the tau load and distribution among the PSP phenotypes of a series of 34 cases who were retrospectively classified following the criteria established by Respondek in 2014 and also according to phenotypes resulting from the new MDS-PSP criteria, at
FIGURE 3 | Tau burden distribution in PSP phenotypes. The figure shows the total, cortical and subcortical semi-quantitative tau loads in the different PSP phenotypes after applying Respondek’s (A), MDS-3y (B), MDS-6y (C) and MDS-last (D) criteria. Simple linear regressions were computed to evaluate group differences between PSP subtypes adjusting by age of onset and Braak stage. (p < 0.05), (p < 0.01). Non-significant differences were not displayed. PSP-RS/PI, Richardson Syndrome spectrum; PSP-Cx, PSP cortical predominant phenotypes; PSP-P, PSP with predominant parkinsonism; PSP-U, unclassified PSP.
3 years of disease progression, at 6 years and at the final visit before death.

Handling a small sample of patients, we have grouped all cortical phenotypes in a group (PSP-Cx) and also PSP-RS together with PSP-PI (PSP-RS-PI), since the latter has been shown to be a transitional form of PSP-RS in which it transforms in a high percentage of cases with disease progression (28).

The application of the MDS-3y criteria implies a reduction of cases fulfilling features for diagnosis of atypical phenotypes and 6 cases of PSP-U arise, mainly at the expense of cortical phenotypes, which are reduced to 3 (PSP-SL). One of the PSP-C cases also moves to the PSP-U group and the other one to the PSP-RS group. This fact may be due to a bias because of the retrospective nature of the study and the lack of clinical data on incipient involvement of oculomotor dysfunction or postural instability. The number of PSP-P cases is reduced to two, in contrast to other studies that find that MDS-3y criteria overestimates the diagnosis of this phenotype in early stages of the disease (14). The number of cases in the PSP-RS/PI group applying the PSP-MDS criteria is practically similar, regardless of whether applied at 3 years of evolution, at 6 years or at the last visit before death. Our findings are in line with other recent publications showing that the new PSP-MDS criteria tends to overestimate PSP-RS in detriment of cortical phenotypes (13). This is due to the fact that some cortical phenotypes (PSP-CBS and PSP-SL) are unable to reach a higher level of certainty than “possible” and, thus, converge to PSP-RS or PSP-P phenotypes after applying MAX rule 1 (17). Surprisingly, we found a high number of PSP-SL cases compared to other series (10, 12, 13, 16) and this fact possibly reflects a case selection bias for biobank study.

The largest differences in tau load between phenotypes are found when comparing cases resulting from the application of Respondek’s criteria and thus, not only are significant differences in total and subcortical tau load found between the PSP-RS/PI and PSP-P groups, but also more cortical tau is found in the PSP-Cx group with respect to the PSP-RS/PI and PSP-P groups. The loss of phenotypic diversity implied by the application of the PSP-MDS criteria leads in our study to a loss of significant differences in tau load between the different phenotypes, contrary to the findings of the first study comparing the tau load between the phenotypes resulting from the application of the PSP-MDS criteria (10). In this multicenter study the cortical phenotypes do show increased cortical tau load. Our work does not reproduce these findings, probably because it is a single-center study with a small sample. Regarding cellular pathology after applying MDS-3y criteria, differences between PSP-Cx and PSP-RS/PI in the astroglial cortical tau load and between PSP-RS and PSP-P in the subcortical neuronal tau load were observed. Although they might be in consonance with those described by Kovacs et al. (10), they should be taken cautiously because of the phenotype clustering performed for the analysis.

Our findings, and those of other postmortem studies, do not allow to establish what happens in the early stages of the disease. We hypothesize that in cortical phenotypes, tau protein deposits firstly in the cortex, causing a cortical/subcortical gradient greater in the initial phases of the disease, and this fact is what determines the clinical syndrome. Unlike other neurodegenerative diseases such as AD or PD (23, 29), where different stages of disease progression have been defined, the evolution by stages in PSP is not equally established. Prodromal symptoms (16, 30) or preclinical forms have not been studied in depth until recently (31, 32). It has been suggested that tau pathology is initially confined to the pallid-nigro-lusian system (10, 33) and from these nuclei it extends to basal ganglia, pontine nuclei, cerebellum dentate nucleus and finally to the frontal and parietal lobes. Different patterns of involvement of subcortical circuitry, suggesting different patterns of disease spread through the brain has been proposed as the basis of the differences of PSP phenotypes (34). Further developing this hypothesis, Kovacs et al. have proposed a pattern of tau pathology with a staging system in PSP-RS that might be applicable to other PSP phenotypes (10). The spread of tau protein along neural connections is a pathogenic hypothesis not yet fully confirmed in primary tauopathies (35). Even more, this mechanism could not explain the astroglial pathology of primary tauopathies (36). PET tau imaging studies may clarify where tau deposits locate in early stages of the disease (34, 37).

Regarding disease duration, it is significantly shorter in PSP-RS/PI phenotype comparing with the other phenotypes (PSP-Cx + PSP-P), independently of the criteria applied with the exception of MDS-3y, probably due to the high presence of PSP-U cases excluded from the analyses. Recently it has been found that there is more dichotomy in terms of disease duration if PSP-RS is grouped with atypical cortical phenotypes than PSP-RS compared vs. all atypical phenotypes (27). These results have not been reproduced in our sample neither has a correlation between disease duration and tau load been found. It might be related to the lack of statistical power due to the cohort size. A significant negative correlation between tau load and disease duration (3) has suggested that more fulminant disease affects more regions, more severely from disease onset and contributes to an earlier death though probably there are other factors influencing disease duration, such as neuronal death or co-pathology (38–40).

The exact pathogenic basis that determines PSP phenotypes is not known. We have found predominance in H1/H1 and E3/E3 genotypes of MAPT and APOE genes in our series. MAPT gene haplotype variation has been shown to influence the risk of PSP and also to play a role in the severity of the disease (41). Cellular vulnerability (neuronal or astroglial) in PSP seems to be determined by molecular mechanisms and thus neuronal deposits are associated with brain expression of synaptic genes while astroglial deposits are associated with microglial and immune system gene expression (42).

The present study has weaknesses. One limitation is its retrospective nature. The main limitation is the small sample, which implies that atypical phenotypes are underrepresented, and leads to the phenotype or the anatomical regions clustering. In addition, the percentage of cases with multiproteinopathy is high, but this fact, is in accordance with recent studies that confirm the less important impact of co-pathologies on PSP progression (39) rate and demonstrates that tau burden
is the strongest correlate with clinical manifestations (40). In contrast, this study analyzes cases from the same brain biobank, which confers the strength of the standardization of neuropathological procedures.

As a conclusion, our study shows that tau load remains different between the PSP-RS and the PSP-P phenotypes resulting from retrospective application of the new MDS-PSP criteria, whereas no other differences regarding cortical or subcortical tau load between phenotypes are found, as a consequence of the loss of phenotypic diversity. Further studies are needed to understand the pathogenic basis of the different PSP phenotypes and also to confirm the need of reformulating the new MDS-PSP criteria.

**DATA AVAILABILITY STATEMENT**

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

**ETHICS STATEMENT**

The studies involving human participants were reviewed and approved by Comité Ético de Investigación con medicamentos (CEIm) de Navarra. The patients/participants provided their written informed consent to participate in this study.

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**AUTHOR CONTRIBUTIONS**

JS-R, ME, and MM contributed to conception and design of the study. VC, MR, JS-R, and VZ developed brain processing, sample genotyping, and measurement of tau burdens. JS-R, CL-M, and AG organized the database, designed the figures, and performed the statistical analysis. JS-R and ME wrote the first draft of the manuscript. VZ, PT-A, and BA wrote sections of the manuscript. All authors contributed to manuscript revision, read, and approved the submitted version.

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**SUPPLEMENTARY MATERIAL**

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fneur.2022.827338/full#supplementary-material
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