Exosomes: New players in cancer (Review)

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Abstract. The past decade has witnessed an exponential increase in research on exosomes. For many years considered to be extracellular debris, exosomes are now considered important mediators in intercellular communication. The capability of exosomes to transfer proteins, DNA, mRNA, as well as non-coding RNAs has made them an attractive focus of research into the pathogenesis of different diseases, including cancer. Increasing evidence suggests that tumor cells release a large sum of exosomes, which may not only influence proximal tumor cells and stromal cells in local microenvironment, but also can exert systemic effects when participating in blood circulation. In this study, we review the current understanding on this topic. The literature outlines two broad facets of exosomes in cancer: 1) promotion of tumor growth, tumorigenesis, tumor angiogenesis, tumor immune escape, drug resistance, and metastasis and 2) their role as promising biomarkers for cancer diagnosis and even as potential treatment targets for cancer patients.

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1. Introduction

In recent years, the function of exosomes has evoked increased interest, particularly in cancer research. Described to harbor and deliver functional molecules to recipient cells, exosomes seemingly add a new layer of complexity to our understanding of molecular biology in multicellular organisms.

The definition of exosomes has evolved over time; the term was coined by Trams et al to describe the release of extracellular vesicles (EVs) with 5'-nucleotidase activity from various normal and neoplastic cell lines (1). In the late 1980s, the usage of the term was adapted to describe small vesicles of endosomal origin that are released during reticulocyte differentiation following the fusion of multivesicular bodies (MVBs) with the plasma membrane (2). Initially ascribed a cell-scavenging function, exosomes are now known to perform a much wider range of biological functions (3-5). However, since exosomes are difficult to discriminate and purify from EVs of other origins, several EV-induced functions reported in the literature may not necessarily be attributable to exosomes alone.

Exosomes are small, lipid bilayer membrane vesicles (30-100 nm) derived from the luminal membrane of multivesicular bodies (MVBs), which are constitutively released by fusion with the cell membrane (6-10). In this review, we highlight the recent advances in our understanding of the biological and functional role of exosomes in cancer, with an emphasis on the potential use of exosomes as biomarkers and therapeutic agents.

2. Biogenesis, secretion, and uptake of exosomes

Biogenesis of exosomes initiates as an endocytic event at the plasma membrane. The mechanisms by which exosomes sort their cargo into the MVBs are not yet fully understood. The best-described mechanism for exosome biogenesis is driven by the endosomal sorting complex required for transport (ESCRT) and the multimolecular machinery is recruited to the endosomal membrane where the individual steps of the exosome biogenesis are orchestrated (11). The second pathway of MVB formation is independent of the ESCRT machinery and is based on the specific lipid composition of the endosomal membrane. It has been reported that ceramides were involved in exosomes biogenesis (12,13). Moreover, formation of MVBs has also been shown to be controlled by the syndecan heparin sulfate proteoglycans and their cytoplasmic adaptor syntenin (14).
Following the formation of MVBs, Rab GTPases govern their degradation as well as their secretion (13,15). The final release of exosomes occurs upon fusion of MVBs with the cellular plasma membrane, a process which is probably mediated, at least in part, by soluble N-ethylmaleimide-sensitive factor attachment protein receptors (SNAREs) (16). Furthermore, there is evidence that enhanced expression of p53, Pyruvate kinase type M2 (PKM2) and tumor suppressor-activated pathway 6 (TSAP6) upregulates the secretion of exosomes in tumor cells (17-19). Of note, the accumulation of intracellular Ca²⁺ and change in microenvironmental pH have been shown to affect the secretion of exosomes (20,21).

Cells appear to take up exosomes by several molecular mechanisms. Most experimental evidence suggested that exosomes are internalized into recipient cells via endocytosis (22,23). Endocytosis is a general statement for a range of molecular pathways, including calthrin-mediated endocytosis, caveolin-mediated endocytosis, phagocytosis and macrophagocytosis (24-26). Moreover, the uptake of exosomes by direct cell surface membrane fusion has also been studied. Palolini et al demonstrate that at least a part of exosomes are able to fuse with the recipient cells directly and low pH facilitates this process (21). Additionally, there is also evidence to support the role of lipid rafts in exosome
internalization (27,28). Noteworthy, several research groups have shown that the uptake of exosomes by recipient cells is an energy-dependent process. For instance, the internalization of exosomes by ovarian cancer cells via several endocytic pathways were strongly inhibited at 4˚C (24). Fig. 1 summarizes the complicated process of biogenesis, release, content and uptake of exosomes.

3. Contents of exosomes

During the biogenesis of exosomes and prior to their secretion, various molecules are uploaded into the lumen of exosomes. Exosomes contain several types of biomolecules, including lipids, proteins and nucleic acids (6) (Fig. 1). The exosomal content is heterogeneous and in a dynamic state, depending on the cell's origin, its physiological and pathological state, and even on the cellular release site (29).

When comparing exosomes with the total cell membranes, several studies observed that exosomes have a higher expression of sphingomyelin, cholesterol, phosphatidylserine, and generally of saturated fatty acids (6). The protein content in exosomes includes endosomal, plasma, and nuclear proteins (30). Proteins enriched in exosomes include those relevant for individual exosomal biogenesis pathways and for exosome secretion (31). Factors found in exosomes of different cell types include: TSG101, Alix, Rab GTPases, heat shock proteins (HSP70, HSP90), integrins, tetraspanins (CD9, CD63, CD81) and MHC class II proteins (6). A number of reviews have described the protein and lipid composition of exosomes, and various databases, including ExoCarta (http://www.exocarta.org/) and Vesiclepedia (http://microvesicle.org/), have cataloged the protein, lipid, and RNA content of exosomes (32,33). In addition, exosomes can contain genetic material such as mRNA, long noncoding RNA (IncRNA), microRNA (miRNA) and even double-stranded DNA (34-36). Moreover, the composition of exosomes can be different from the cells of their origin due to the selective sorting of cargo into exosomes. However, not much is known about these selective shuttling mechanisms.

4. Biological function of exosomes in cancer

Emerging evidence suggests that exosomes derived from cancer cells are involved in tumor growth, tumorigenesis, angiogenesis, tumor immune escape, drug resistance and metastasis. Exosomes are also involved in the intercellular communication between cancer cells and stromal cells, especially with cancer-associated fibroblasts (CAFs). In this section, we reviewed recently published articles reporting the roles of exosomes in cancer and their underlying molecular mechanisms (Fig. 2).

**Tumorigenesis.** Exosomes derived from malignant cells have shown the potential to induce cell transformation. They can

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**Figure 2.** Biological function of exosomes in cancer. Exosomes are involved in tumor growth, tumorigenesis, angiogenesis, tumor immune escape, drug resistance and metastasis. EMT, epithelial-mesenchymal transition.
also affect other cells in the heterogeneous tumor population, and be involved in the transfer of metastatic capacity. For instance, exosomes released by malignant breast cancer cells are taken up by less malignant tumor cells located within the same tumor, and thereby promote their migratory and metastatic potential (37).

In addition, exosomal miRNA from cancer cells can contribute to tumorigenesis. Both mature miRNA and premiRNA transcripts are present in exosomes along with other key components of the machinery for miRNA biogenesis such as DICER, TRBP, and AGO2 (35). Melo et al demonstrated that exosomes derived from breast cancer cells and sera of patients with breast cancer could instigate nontumorigenic epithelial cells to form tumors (35). Although much of the evidence has emanated from in vitro studies, the exchange of exosomes between tumor cells was recently demonstrated in vivo in a study which combined high-resolution imaging with a Cre-LoxP system. Exosomes derived from malignant breast cancer cells are taken up by less malignant tumor cells within the same tumor and distant tumors. The mRNA content in the exosomes promotes migratory behavior and metastatic capacity of the recipient cancer cells (38).

CAFs are major components of the tumor microenvironment. Exosomes derived from CAFs can mediate horizontal transfer of miRNAs and proteins to affect breast cancer progression (39,40). Webber et al demonstrated exosomal TGF-β1 participated in the activation of myofibroblast, which is a rate-limiting step in cancer progression (41). Hoshino et al reported the effect of cancer cell-derived exosomes on multiple steps of invadopodia life cycle, including invadopodia formation and stabilization. Release of exosomal proteinases were also shown to enhance degradation of extracellular matrix (ECM) associated with invadopodia maturation (42). By inducing ECM degradation, cancer cell-derived exosomes promote tumor cell invasiveness and motility. Of note, Lazar et al showed that adipocytes secrete exosomes in abundance, which are then taken up by tumor cells, leading to increased migration and invasion (43). Collectively, these findings point towards a role of exosomes in tumorigenesis.

**Tumor growth.** The effect of tumor-derived exosomes on tumor growth has been widely reported in the past decade. Exosomes from sera of glioblastoma patients are enriched with EGFRvIII mRNA. The proliferation potential of recipient cells was shown to be greatly enhanced on co-culture with EGFRvIII-containing exosomes (44). Peinado et al demonstrated that melanoma exosomes containing MET oncoprotein can support tumor growth as well (4). In colorectal cancer, tumor-derived exosomes are enriched in cell cycle-related mRNAs, promoting proliferation of endothelial cells and tumor growth (45). Another study by Kogure et al showed that exosomes derived from hepatocellular carcinoma (HCC) cells can modulate TAK1 expression and associated signaling pathways to enhance cell growth in recipient cells (46). In addition, exosomes from stromal cells can promote proliferation through other signaling pathways (47,48). For instance, Au Yeung et al demonstrated that CAF secreted exosomes to regulate survival and proliferation of pancreatic cancer cells, thus may serve as a potential target for overcoming resistance of chemotherapy. Similarly, miRNA-21 enriched in CAF exosomes may profoundly impact ovarian cancer growth by suppressing apoptosis through binding to APAF1 (49). Zhang et al reported that the loss of exosomal miRNA-320a from CAFs contributed to HCC proliferation (50).

While the majority of research evidence pertains to the pro-tumorigenic effect of tumor-derived exosomes, it is important to remember that the function of stromal-derived exosomes may differ from, and, perhaps, be opposite to that of cancer exosomes. The presence of a competitive biological process was well-characterized in multiple myeloma (MM). Bone marrow mesenchymal stromal cells (BM-MSCs) from patients with MM were shown to have a high expression of oncogenic proteins, which facilitated the growth of MM cells in vivo (51). In contrast, the level of miRNA-15a, a known suppressor of MM growth (52), was significantly higher in exosomes derived from BM-MSCs of normal individuals, thus suggesting a tumor suppressive role of MSC-derived miRNA-15a. Thus, exosomes from cancer and stromal cells may modulate tumor growth.

**Angiogenesis.** Angiogenesis is a fundamental physiological process involved in wound healing and carcinogenesis. Angiogenesis involves a close interaction between endothelial cells and their surrounding microenvironment. Uptake of exosomes by the endothelial cells (ECs) stimulates angiogenesis. Several groups reported the pro-angiogenic effect of tumor-derived exosomes on endothelial cells in various types of cancers such as glioblastoma, leukemia, multiple myeloma, melanoma, ovarian cancer and breast cancer (53-57). In a study by Skog et al, angiogenic-protein rich exosomes released from glioblastoma tumor cells were shown to stimulate tubule formation in ECs (53). Gopal et al demonstrated communication between oncogenic cells undergoing epithelial-mesenchymal transition (EMT) and endothelial cells via exosomes containing Rac1/PAK2 proteins as the angiogenic promoters (58).

Some miRNAs found in exosomes are thought to be specifically involved in tumor angiogenesis (59). For example, in colorectal cancer, miRNA-9 in tumor-derived exosomal vesiculars showed pro-angiogenic effects through inhibiting the expression of suppression of cytokine signaling 5 (SOCS5), promoting the migration of endothelial cells. In addition, miRNA-210 has been observed to suppress the expression of specific genes in endothelial cells, resulting in enhanced pro-angiogenic activity (60-62). Despite the direct pro-angiogenic effect of tumor-derived exosomes on endothelial cells, tumor-derived exosomes also demonstrated indirect effects on other stromal cells, such as CAFs. For example, in leukemia, tumor-derived exosomes induced a CAF phenotype in stromal cells in the surrounding microenvironment, hence leading to increased expression of pro-angiogenic factors in tumor (63). Of note, transfer of miRNA-125a from MSC-derived exosomes to ECs promoted angiogenesis, both in vitro and in vivo (64). The effect of tumor-derived exosomes on vascular remodeling may affect both tumor growth and metastasis. For instance, melanoma-derived exosomes can induce vascular leak at pre-metastatic sites and can also reprogramme bone marrow progenitors towards a pro-vasculogenic phenotype (4).
Exosomes have been shown to contain different kinds of angiogenic factors; it is necessary to understand the exact contributions of these factors to cancer development.

_Tumor immune escape._ Currently available evidence indicates a dual role of exosomes in mediating crosstalk between immune cells and cancer cells. On the one hand, various immune-stimulatory genes, such as mesothelin, and carcinoembryonic antigen (CEA), are packed in tumor-derived exosomes (65,66). On the other hand, several recent studies suggested that the cancer cells utilized exosomes containing nucleic acids and proteins to enact an immune escape.

An earlier study showed that the dendritic cell (DC)-derived exosomes stimulated an antitumor immune response (67). However, recent research showed tumor-derived exosomes may aid in immune evasion by impairing differentiation and maturation of dendritic cells, which changed their role from effective antigen presenting cells into negative modulators of immune response (68-70).

Exosomes may also play a role in the biology of cytotoxic T cells and regulatory T cells. For instance, in nasopharyngeal carcinoma, tumor-derived exosomes impaired T cell proliferation, differentiation and cytokine secretion _in vivo_ and _in vitro_. The effect appeared to be mediated via down-regulation of the MAPK1 and JAK/STAT pathways by exosomal miRNAs (71). Clayton _et al_ demonstrated an exosome-mediated mechanism that skewed the IL-2 responsiveness in favor of regulatory T cells and away from cytotoxic cells; this coordinated effect on cellular immunity strongly implicates the role of tumor-derived exosomes in immune escape by tumor cells (72).

Tumor-associated macrophages (TAMs) are currently the most widely studied inflammatory cell component of tumor microenvironment (TME) and may also be activated by tumor-derived exosomes. For instance, Fabbri _et al_ identified a new mechanism of communication between TAMs and cancer cells via exosomal miRNAs. Specifically, exosomal miRNA-21 and miRNA-29a were recruited in the TME by TAMs and bind to their Toll-like receptor 8, triggering the Nuclear factor kappa-light-chain-enhancer of activated B cells (NF-κB) pathway and the secretion of interleukin-6 (IL-6) (73).

Moreover, exosomes derived from human prostate cancer cells were shown to express ligands for NKG2D, which induced downregulation of NKG2D in NK cells and impaired NK cell cytotoxic function (74). Ying _et al_ demonstrated that ovarian tumor-derived exosomes play a crucial role in regulating the polarization of tumor-promoting M2 macrophages (75).

While the above examples demonstrate that tumor-derived exosomes may suppress the immune response, it appears that exosomes derived from immune cells can also influence the cancer cells. For instance, exosomes from activated CD8+ T cells increased tumor immunogenicity by activating ERK and NF-κB signaling, which can promote the metastatic potential of tumor cells (76). Collectively, these studies suggest that exosomes may mediate immunosuppression in tumor-bearing host in different ways.

_Drug resistance._ Drug resistance has long been a major obstacle in the management of cancers. Recently, exosomes are of great interest in drug resistance studies. Drug-resistant cancer cells may spread resistance to hitherto sensitive ones by releasing exosomes; such effects could be partly attributed to the intercellular transfer of specific proteins, miRNAs, and even long noncoding RNAs (77-80).

Stromal cells were found to initiate cross-talk with cancer cells via exosomes. CAF-derived exosomes were shown to contribute to proliferation and chemoresistance of pancreatic cancer cells (81). Boelens _et al_ demonstrated the transfer of exosomes from stromal cells to breast cancer cells activated antiviral retinoic acid-inducible gene 1 enzyme (RIG-1) signaling to regulate the expansion of therapy-resistant tumor-initiating cells (82). Further study by Au Yeung _et al_ showed that in ovarian cancer, exosomal transportation of CAF-derived miRNA-21 conferred paclitaxel resistance in ovarian cancer cells through targeting APAF1 (49). Moreover, a recent study by Sun _et al_ demonstrated that suppression of miRNA-122 resulted in taxol resistance by upregulating Septin-9 in HCC (29). Qu _et al_ reported that exosome-transmitted LncRNA promoted Sunitinib resistance in renal cancer by acting as a competing endogenous RNA for miRNA-34 and miRNA-449 to promote AXL and c-MET expression (79).

In addition, exosomes may also affect tumor chemotherapy by mediating drug efflux. Shedden _et al_ found that doxorubicin can be encapsulated and exported by tumor-derived exosomes (83). Moreover, exosomes may play a role in lowering the therapeutic effect of antibodies by modulating their binding to cancer cells. For instance, breast cancer cell-derived exosomes were shown to have a high expression of HER2 which interfered with the activity of the monoclonal antibody trastuzumab _in vitro_ (84). Collectively, exosomes derived from both cancer cells and stromal cells can contribute to the development of chemoresistance in tumor cells.

_Metastasis._ Exosomes not only affect cells in the location where they are produced, but may also influence cells in distant tissues. Exosomes are involved in both the initiation of metastasis and the preparation of a pre-metastatic niche. Exosomes may assist cancer cells in acquiring migratory and invasive properties through EMT. For instance, Aga _et al_ demonstrated that treatment of EBV-negative cells with LMP1-exosomes increases migration and invasiveness of nasopharyngeal cancer cells in functional assays, which correlates with the phenotype associated with EMT (85). Recently, _in vitro_ imaging experiment also showed the importance of exosome-mediated exchange of molecules for metastasis among tumor cells (38).

The pre-metastatic niche is a prerequisite of tumor metastasis. Exosomes derived from the primary tumor can act as potential mediators for priming the pre-metastatic niche. Pancreatic tumor-derived exosomes enriched in macrophage migration inhibitory factors recruited macrophages to establish pre-metastatic niche in the liver, which resulted in increased hepatic macrometastatic burden (86). Moreover, breast cancer cell-derived exosomal miRNA-122 was shown to suppress glucose uptake by niche cells to promote metastasis, both _in vitro_ and _in vivo_ (3). Liu _et al_ recently reported the Toll-like receptor 3 (TLR3) mediated crosstalk between pulmonary epithelial cells and tumor exosomal RNAs as being critical to the initiation of neutrophil recruitment and lung metastatic niche formation (87). In addition, tumor-derived exosomes were shown to express unique
integrins which prepare the pre-metastatic niche by fusing with resident cells and by activating Src phosphorylation and pro-inflammatory S100 expression at the eventual site of metastasis (88).

Recently, Bliss et al reported the nature of regulatory interactions between breast cancer cells and MSC mediated by exosomes and MSC-derived exosomes may stimulate cycling quiescence and early breast cancer dormancy in bone marrow (89). Moreover, tumor-derived exosomes carrying miRNA-181c may trigger the breakdown of blood-brain-barrier (BBB) to promote brain metastasis (90). Zhang et al demonstrated that astrocyte-derived exosomes mediate intercellular transfer of PTEN-targeting miRNAs to metastatic tumor cells, which results in PTEN silencing in these tumor cells. This loss of PTEN expression leads to increased chemokine CCL2 level that recruits myeloid cells to reciprocally promote brain metastasis (91). These findings show that the intercellular crosstalk mediated by exosomes is a crucial mechanism for tumor metastasis.

5. Exosomes as cancer biomarkers

Exosomes have been detected in nearly all kinds of body fluids, including blood, urine, saliva, amniotic fluid, cerebrospinal fluids, bile, ascites, tears, breast milk and semen (9,92-96). Almost all types of cells can secrete exosomes containing specific proteins, lipids, RNA and even DNA into their microenvironment and circulation (97).

The discovery of elevated level of exosomal miRNAs in the plasma of certain cancer patients was the first indication of the potential use of circulating exosomes as cancer biomarkers. Taylor et al reported that circulating exosomal miRNAs from patients with ovarian cancer were significantly distinct from profiles observed in benign disease and healthy volunteers (98). Microarray analyses of miRNAs derived from circulating exosomes derived from 88 patients with primary colorectal cancer (CRC) and 11 healthy donors, revealed significantly higher levels of 7 miRNAs in the former (99). Recently, Zhu et al demonstrated that the levels of miRNA-19a-3p, miRNA-21-5p and miRNA-425-5p were significantly elevated in exosomes from colorectal cancer patients’ serum samples (100).

In addition, Li et al reported that LncRNA can be detected in plasma, and one of the possible mechanisms of its stable existence in blood was its protection by exosomes (101). Li et al recently reported that high levels of exosomal circular RNAs (circRNAs) and serum exosomal circRNAs can distinguish cancer patients from healthy individuals (102).

Proteins associated with tumor-derived exosomes are also seen as potential biomarkers. Exosomes double-positive for CD63 or caveolin-1 and Rab-5b were significantly increased in plasma from melanoma patients as compared to that from healthy donors (103). Noteworthy, Melo et al showed that Glypican-1-positive circulating exosomes was a reliable biomarker for early detection of pancreatic cancer, with superior prognostic value than CA19-9 (104). Similarly, in a study by Lea et al provided proof-of-concept data that supported the high diagnostic power of phosphatidyserine-positive exosomes detected in the blood of women with ovarian malignancies (105).

For prognostic evaluation, circulating exosomes may enhance the stratification of cancer patients with high-risk factors. Manier et al reported that two circulating exosomal miRNAs, namely let-7b and miRNA-18a, improved survival prediction in patients with multiple myeloma (106). Similar results have been found in multiple types of cancers, including non-small cell lung cancer, esophageal squamous cell carcinoma, colorectal cancer, HCC and nasopharyngeal carcinoma (107-112).

For monitoring treatment response, exosomes have been demonstrated to accurately reflect the levels of proteins and mRNAs in parental cells throughout treatment and therefore can serve as potential biomarker of chemotherapy response (113,114). Skog et al identified a protein signature in circulating exosomes linked with clinical stages of melanoma patients. As a predictor of TKI treatment response, exosomal EGFRvIII splice variant was detectable in the serum of glioblastoma multiforme patients, but not the 30 matched controls (53).

Additionally, some newly developed technologies exist to sort and enrich exosomes. Wunsch et al recently demonstrated that nanoscale deterministic lateral displacement pillar arrays, an efficient technology to sort exosomes, may open up the potential for on-chip separation and diagnosis (115).

Collectively, the development of exosome-based novel biomarkers may provide benefits to cancer patients in a wide variety of ways (Table I).

6. Exosomes as cancer therapeutic targets

Thus far, not much progress has been made in the use of exosomes as therapeutic agents in clinical practice. However, their importance in carcinogenesis envisages their use in individualized cancer therapeutics in the future.

First, complete depletion of circulating exosomes may potentially result in major benefit for cancer patients. Ciravolo et al reported that exosomes purified from HER2-positive early-stage breast cancer patient sera and HER2-overexpressing breast cancer cells can inhibit trastuzumab-induced anti-proliferative activity (84). Removal of such exosomes from advanced breast cancer patients is likely to improve response to trastuzumab therapy (116). Next, dendritic cell-derived exosomes have been used as tumor vaccine to trigger host antitumor immune response and inhibit cell growth and proliferation. Phase I clinical trials of dendritic exosomes in patients with metastatic melanoma and advanced non-small cell lung cancer have shown modest therapeutic effects (117,118).

Exosomes may serve as delivery vehicle. Use of exosomes as nanocarriers for therapeutic agents is being actively explored. For instance, Alcarez et al demonstrated the use of exosomes for targeted delivery of siRNA after systemic administration, which indicates their potential use as vehicle for gene therapies (22). Tian et al reported that exosomes modified by targeting ligands can be used for delivery of doxorubicin to tumor cells (119). Kim et al reported good efficacy of an exosome-based system as a delivery vehicle for PTX to multi-drug resistant cancer cells (120).

More recently, MSC-derived exosomes are being examined for their role in MSC-based cellular therapy. For instance, Katakowski et al demonstrated that exosomes from
MSCs expressing miRNA-146b significantly reduced glioma xenograft growth in rat model of primary brain tumor (121). Similarly, a study reported by Ono et al suggested that exosomal transfer of miRNAs from MSCs may promote breast cancer dormancy in a metastatic niche (122). Additionally, use of exosomes derived from BM-MSCs as nanocarriers for targeted delivery of antitumor drugs such as paclitaxel (PTX) has also been investigated (123).

Table I. Exosomes from distinct body fluids of cancer patients as biomarkers.

| Exosomal cargos                  | Cancer types                         | Clinical value                                                                 | Biofluids          | Refs. |
|---------------------------------|--------------------------------------|--------------------------------------------------------------------------------|--------------------|-------|
| Phosphatidylserine              | Ovarian cancer                       | Elevated level of phosphatidylserine positive exosomes in ovarian cancer patients than healthy controls | Plasma             | (108) |
| TRPC5                           | Breast cancer                        | Elevated level of exosomal TRPC5 in cancer patients and higher levels may predict poorer progress | Plasma             | (124) |
| miRNA-1246, 21                  | Breast cancer                        | Elevated levels of these two exosomal miRNAs in breast cancer patients than healthy controls | Plasma             | (125) |
| CRNDE-h                         | Colorectal cancer                    | Elevated level of exosomal CRNDE-h in cancer patients and higher levels predict poorer progress | Serum              | (110) |
| miRNA-4772-3p                   | Colon cancer                         | Lower level of exosomal miRNA-4772-3p predict tumor recurrence                | Serum              | (126) |
| miRNA-19a-3p, 21-5p, 425-5p     | Colorectal cancer                    | Elevated levels of the three exosomal miRNAs in colorectal cancer patients than healthy controls | Serum              | (100) |
| miRNA-17-92a                    | Colorectal cancer                    | Elevated level in colorectal cancer patients and higher levels may predict poorer progress | Serum              | (127) |
| miRNA-19-3p, 21-5p, 221-3p      | Lung adenocarcinoma                  | Elevated levels of the three miRNAs in lung adenocarcinoma patients than healthy controls | Plasma             | (128) |
| miRNA-302a, 302-c, 126          | Non-small cell lung cancer (NSCLC)   | Elevated levels of the three miRNAs in NSCLC patients than healthy controls   | Plasma/Broncho-alveolar lavage | (129) |
| EML4-ALK fusion                 | NSCLC                                | EML4-ALK fusion transcripts have been identified in the exosomal RNA of NSCLC patients | Plasma             | (130) |
| Glypican-1                      | Pancreatic cancer                    | Higher level of Glypican-1 positive exosomes in patients with early- and late-stage pancreatic cancer than healthy controls | Serum              | (107) |
| miRNA-1246, 4644, 3976, 4306    | Pancreatic cancer                    | Elevated levels of these 4 exosomal miRNAs in pancreatic cancer patients compared to healthy controls | Serum              | (131) |
| LncRNA-p21                      | Prostate cancer                      | Elevated level of exosomal LncRNA-p21 in patients with prostate cancer than healthy controls | Plasma             | (132) |
| miRNA-21, 375                   | Prostate cancer                      | Elevated levels of urinary exosomal miRNA-21 and miRNA-375 in patients with prostate cancer than healthy controls | Urine              | (133) |
| miRNA-375, 1290                 | Prostate cancer                      | Castration-resident prostate cancer patients with elevated levels of both exosomal miRNA-375 and miRNA-1290 may predict poorer progress | Serum              | (134) |
| miRNA-718                       | HCC                                  | Lower level of exosomal miRNA-718 in patients with HCC patients with recurrence after liver transplantation than those without recurrence | Serum              | (135) |
| miRNA-21                        | HCC                                  | Elevated level of exosomal microRNA-21 in HCC cancer patients than healthy controls | Serum              | (136) |
| miRNA-211, 222, 224             | HCC                                  | Elevated levels of these three exosomal microRNAs in HCC cancer patients than healthy controls | Serum              | (137) |
| CD34                            | Acute myeloid leukemia (AML)         | Higher level of CD34 positive exosomes in AML patients than healthy control | Plasma             | (138) |
| Let-7b, miRNA-18a               | MM                                   | Elevated expression in cancer patients and higher levels of these two exosomal miRNAs may predict poorer progress | Serum              | (106) |
7. Conclusion

The rapid increase in the number of published studies, especially in oncology, clearly reflects the enthusiasm on research of exosome biology and function. Cancer cells communicate with surrounding and distant cells via exosomes. Exosomes constitute a bi-directional interaction network that mediates the intercellular cross talk between cancer cells and the microenvironment to promote cancer development, progression, metastasis, and drug resistance. However, most pieces of the aforementioned findings are gathered from cell culture experiments. Therefore, it is vital to substantiate these results in more rigorous in vitro settings. Moreover, exosomes released by cancer cells and TME are highly heterogeneous. But there are few reports explaining the role of each of these subtypes. The difficulties in elucidating the roles of specific exosomes include lack of unique molecules to distinguish each exosome subtype and appropriate exosomes isolation methods. Future technical advances may lead to significant progress in the understanding of the heterogeneity of exosomes.

Exosomes have been proven to be stable carriers of genetic materials, and been nominated as promising tumor biomarkers for cancer diagnosis and prognosis. Moreover, during cancer treatment, exosomes may switch their contents and may therefore exhibit traits for the monitoring of therapeutic efficiency. Furthermore, as vectors for drugs and tumor vaccines, exosome-based delivery is rigorously evaluated as an emerging therapeutic strategy for cancer. Additional studies are much needed to better elucidate their role and mechanism of action in cancer to reduce the risk of off-target effects and therapeutic failures. The several aforementioned results still require further validation in clinical settings, especially in independent prospective cohorts.

Altogether, with the emergence of precision medicine, future studies on exosomes should not only emphasize their roles in cancer biology, but may also open new avenues for cancer diagnostics and therapeutics.

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