Elution Profiles of Synthetic CaSO₄ Hemihydrate Beads Loaded with Vancomycin and Tobramycin

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Published online: 23 April 2020
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Abstract

Backgrounds and Objectives  The use of local antibiotic delivery vehicles is common in the management of biofilm-related infections as they provide high concentrations of local antibiotics while simultaneously avoiding complications from systemic toxicity. We present a 100% pure synthetic calcium sulfate hemi-hydrate mixed with 240 mg tobramycin and 500 mg vancomycin per 10 cc mixture for use in revision surgeries of periprosthetic joint infections (PJIs). The purified carrier demonstrates bioabsorbability, promotion of bone growth, a physiologically favorable pH, and hydrophilicity. These unique properties may alleviate persistent postoperative wound drainage seen in patients with PJI. Our questions consist of two parts: (1) does the novel calcium sulfate carrier provide therapeutic concentrations of antibiotic locally that can kill biofilm related infections? (2) Are serum concentrations of antibiotic significant to cause concern for systemic toxicity?

Methods  To address these questions, we assayed the elution of antibiotic concentrations obtained from surgical drains and serum among 50 patients in the first 5 postoperative days.

Results  The elution of vancomycin and tobramycin was greatest on day 1 compared with those concentrations obtained on days 2, 3, 4, and 5; serum concentrations were largely undetectable. Our findings demonstrate that this calcium sulfate preparation provides therapeutic delivery of vancomycin and tobramycin locally at log 2–3 above the minimum inhibitory concentration (MIC), while avoiding toxic serum concentrations.

Conclusions  When used in one-stage revision arthroplasties, the bioabsorbable, purified carrier delivers high concentrations of antibiotic while avoiding systemic toxicity.

Keywords  Biofilm · Antibiotics · Infected total joint prosthetics · Calcium sulfate carrier

1 Introduction

Chronic infection presents a serious complication following the surgical implantation of total joint prosthetics. Once a patient becomes contaminated, bacteria adhere to surfaces of implanted components and form colonies (biofilm) that enter a state of chronic latency [1–4]. Bacteria entering into this sessile form may produce biofilm matrices (glycocalyx); they proliferate, exchange genetic information, and proliferate up to a point and stop (quorum sensing), leading to asymptomatic infections [5, 6]. With biofilm development, pathogen genomes are altered through cell-to-cell signaling pathways, becoming phenotypically distinct from planktonic forms [6]. This change allows sessile forms to develop recalcitrance to immune regulatory mechanisms and antimicrobial therapy, causing much concern over the management of chronic infection [5, 7–9]. Ideally, drugs used in treating biofilm-mediated infections should be capable of disrupting the bacteria’s structure [5, 7, 10, 11]. However, parenterally administered antibiotics treat planktonic bacterial infections with more success than the sessile forms [5, 11].

In response, surgeons employ local antibiotic carriers as an adjunct to one- and two-stage revision protocols for infected total joint prosthetics [12–17]. The use of local
Key Points

Local antibiotic delivery can effectively reduce biofilm formation

Synthetic calcium sulfate’s unique properties optimize antibiotic delivery

Calcium sulfate carries therapeutic antibiotic levels without systemic toxicity

delivery mechanisms provides high concentrations of drugs at the infection site with minimal serum concentrations [12, 18–21]. Furthermore, maintaining local antibiotic concentrations many times above the minimum inhibitory concentration (MIC) discourages the development of bacterial resistance that prolonged exposure to antibiotics below inhibitory concentrations may cause [20]. Currently, the use of antibiotic-impregnated poly-methyl-methacrylate (PMMA) spacers sets the standard for antibiotic therapy in two-stage revisions of septic joints [5, 16, 17, 21–24]. As PMMA spacers are not bioabsorbable, additional surgery becomes necessary for removal, thus rendering them inadequate as soft tissue fillers in one-stage treatment of osteomyelitis [12, 16, 17]. Moreover, the surface roughness of PMMA may potentiate bacterial adhesion, possibly leading to secondary infection [18, 25–28]. Finally, the use of PMMA may be recovered in the serum [16, 17], correlating with the observation of sustained antibiotic serum concentrations that may result in allergic reactions or complications [16, 17, 29].

As an alternative, calcium sulfate carriers offer multiple significant advantages over PMMA. These advantages include the reabsorption of calcium sulfate within weeks, the carrier releases its entire antibiotic load when still visualized by x-ray, mixture with heat-sensitive antibiotics remains unproblematic because of minimal heat production on curing, the pelleted carrier obliterates dead space following debridement of bone and soft tissue, and calcium sulfate serves as a potential synthetic osteoconductive bone substitute when placed in the bone [30–34]. These advantages make the use of a calcium sulfate vehicle intriguing for application in several scenarios, including abscess or soft tissue infection treatment, prophylaxis in high-risk patients, use with physiologic constructs in one-stage procedures for osteomyelitis, and as an adjunct to antibiotic-loaded bone cement. Preparation of calcium sulfate beads provides a stable platform for the delivery antibiotics; however, problems associated with the use of calcium sulfate involve complications with persistent postoperative wound drainage [35–40]. We feel this drainage may be due to specific physical properties of gypsum-mined calcium sulfate preparations, including its hydrophobicity, acidic pH, and impurity due to the presence of trace minerals as seen by Fourier transform infrared (FTIR) analysis. FTIR analysis involves transmitting infrared radiation through a material with a detector on the other side of the material reading the resulting spectrum of infrared radiation. The spectrum picked up by the detector then represents the chemical makeup of the material that is unique to materials only with that specific molecular structure, making it a useful tool in analyzing materials for quality [41].

This study examines clinical elution profiles of a purified calcium sulfate preparation in contrast to other less pure forms. Thus, we ask the following: (1) does this calcium sulfate preparation provide local delivery of antibiotics at concentrations exceeding MIC of common infecting pathogens? (2) Will serum concentrations of antibiotics remain at safe concentrations?

2 Patients and Methods

For the purposes of this study, we chose Stimulan® (Bio-composites, Keele Science Park, Staffordshire, UK) as the purified antibiotic carrier. Since a broad-spectrum antibiotic preparation provides coverage against gram-positive and -negative species with few resistant bacterial strains, we selected vancomycin and tobramycin to treat gram-positive and -negative bacteria, respectively [42]. These antibiotics also act as an effective combination due to their synergistic bactericidal potential in treating serious infections involving Staphylococcus aureus and other pathogens [20].

Calcium sulfate hemi-hydrate powder reacts with water provided by liquid tobramycin and sets according to the following reaction:

\[
\text{CaSO}_4 \cdot \frac{1}{2}\text{H}_2\text{O} + \frac{1}{2}\text{H}_2\text{O} = \text{CaSO}_4 \cdot 2\text{H}_2\text{O}
\]

The addition of antibiotics affects this setting reaction. Tobramycin, in liquid form, accelerates setting rates as opposed to the effect of vancomycin powder, with higher additions having a greater effect for delayed setting. See Table 1 for a comparison of preparation notes for liquid and powder forms of tobramycin with the carrier.

The mixing of calcium sulfate with the antibiotics is performed in the sterile field of the operating room intraoperatively. Tobramycin preparation and dose were prepared identically for each patient, according to Table 1. Vancomycin was also prepared identically for each patient, using a 500 mg per 10 cc pellet mixture. For surgical placement, the calcium sulfate/antibiotic paste is pressed into a pelleting mold (supplied with the product, Fig. 1) to produce beads of different dimensions. The paste is allowed to set within the mold, and the resulting beads are removed by flexing the
Elution Profiles of Antibiotic-Loaded CaSO₄ Hemihydrate Beads

mold over a sterile container. The beads remain covered in the container until implantation.

The first 50 patients that consented to the procedure were included (Table 2). There was no differentiation based on gender or age. Patients with a GFR > 60 and a cleared preoperative evaluation were viable candidates. Both serum and local drain samples were collected every morning at 0600 and taken immediately to the laboratory for same-day analysis. Serum samples were collected in red-top tubes with 3-ml aliquots. The local drain samples were collected in sterile glass tubes with 5-ml aliquots.

Upon completion of surgical debridement and implantation of prosthetic components, the calcium sulfate pellets are applied to the wound for dead space management with high concentrations of antibiotic bleaching. Patients are then fitted with a drain prior to wound closure for collection of the local exudates. In the 5 postoperative days, serum and drain samples are assayed with fluorescence polarization immunoassay (FPIA) with the Abbott Architect C8000 Chemistry Analyzer to evaluate the eluting characteristics of the Stimulan® carrier impregnated with vancomycin and tobramycin at the above-mentioned concentrations. As FPIA executes a single step to assay samples, it offers high-throughput screening of large numbers of samples.

### Table 1 Preparation of antibiotic-loaded beads for liquid or powder forms of tobramycin

| Form of tobramycin | Ingredients | Instructions | Setting time (min) |
|--------------------|-------------|--------------|--------------------|
| Liquid             | 6 ml liquid tobramycin (40 mg/mL) | Combine ingredients | 10 |
|                    | 10 cc pack of rapid cure | Mix for 30 s to a creamy paste | |
| Powder             | 240 mg tobramycin powder | Premix rapid cure for 1 min prior to addition of tobramycin powder | 7 |
|                    | 10 cc pack of rapid cure | Add 1 g tobramycin powder | |
|                    |                          | Mix 30 more seconds | |

### Table 2 Patient demographics and laboratory data (n = 50)

| Characteristic                  | Value |
|---------------------------------|-------|
| Gender (n)                      |       |
| Male                            | 22    |
| Female                          | 28    |
| Age (years), mean (range)       | 61 (13–82) |
| No. of knee PJI cases           | 33    |
| No. of hip PJI cases            | 15    |
| No. of elbow PJI cases          | 1     |
| No. of shoulder PJI cases       | 1     |
| GFR (ml/min/1.73 m²)            | All patients: > 60 |

GFR glomerular filtration rate, PJI periprosthetic joint infection

### 2.1 Bioanalytical Assay

All samples were tested via the Abbott Architect C8000 Chemistry Analyzer in the same manner. Upon collection and delivery to the laboratory, the protocol provided by the Abbott User Manual was followed [43]. The internal standard was sterile water. The upper and lower limits of quantification were 15 and 5 times the concentration of the internal standard, respectively. The laboratory was in-house, Texas Health Presbyterian Dallas, and operated by board-certified pathologists and laboratory technicians who performed routine quality improvement/quality assurance with the device.

### 3 Results

We analyzed 50 patients undergoing revision arthroplasty for infected total joints or major multiple revisions. Cases included 33 knees (1 bilateral), 15 hips, 1 elbow, and 1 shoulder from 22 females and 28 males. Average patient age was 61 years (range 13–82). None of the patients experienced relapse of infection or wound complications secondary to persistent wound drainage.

We evaluated local antibiotic concentrations to determine whether the concentrations contained in the eluent exceeded the MIC of common infecting pathogens and concentrations in the serum were also evaluated for all subjects [44–47]. Table 3 lists the results.
Mean values obtained from the local exudate demonstrated therapeutic concentrations of vancomycin and tobramycin in each of the 5 postoperative days. Concentrations of both antibiotics peaked on day 1 (averages: vancomycin: 265 µg/ml; tobramycin: 31 µg/ml, and ranges: vancomycin: 28.3–736.4; tobramycin: 6.4–97.2). Table 4 lists the change in mean values over time for each of the 50 subjects, and Figs. 2 and 3 illustrate the trend in mean value reduction over time.

We evaluated drain and blood serum concentrations of vancomycin and tobramycin on a standard hospital assay for these drugs in the 5 postoperative days. The assay reported antibiotic concentrations that were detected within specific concentration limits (vancomycin 2–400 µg/ml, tobramycin 0.25–1.0 µg/ml). Additional specific maximum assayable values for each antibiotic were recorded for a selection of the results. The majority of patients exhibited antibiotic blood serum concentrations below the lower concentration limits of the standard assay (vancomycin < 2 µg/ml, tobramycin < 0.5 µg/ml). However, a few patients exhibited detectable serum concentrations of antibiotics as follows: 11 (of the 50 patients) at day 1, 4 patients at day 2, and 3 patients at days 3 and 4. Table 5 lists the antibiotic serum concentrations for each patient.

Vancomycin is primarily effective against gram-positive cocci, _S. aureus_ and _S. epidermidis_, both methicillin-susceptible (MSSA and MSSE) or resistant-species (MRSA and MRSE), are typically sensitive to vancomycin with MICs < 1.5 µg/ml. Most strains of _streptococcus_ are sensitive to vancomycin. Vancomycin is considered bactericidal (MBC/MIC < 4 µg/ml) except with _enterococci_ and some tolerant _staphylococci_ (MBC/MIC > 32 µg/ml).

NA not applicable, MIC minimum inhibitory concentration, MBC MIC minimum bactericidal concentration, MSSA methicillin-sensitive _S. aureus_, MRSA methicillin-resistant _S. aureus_.

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**Table 3** Antimicrobial spectrum for tobramycin and vancomycin

| Organism (tobramycin) | MIC range (µg/ml) (tobramycin) | Organism (vancomycin) | MIC range (µg/ml) (vancomycin) |
|------------------------|---------------------------------|------------------------|---------------------------------|
| *Enterococcus faecalis* | 8–32                            | *Enterococci*          | 4.0                             |
| *Escherichia coli*     | 0.25–1.0                        | *MSSA*                 | <2.0                            |
| *Pseudomonas aeruginosa* | 0.25–1.0                       | *MRSA*                 | <2.0                            |
| *Staphylococcus aureus* | 0.12–1.0                        | Coagulase-negative *Staphylococci* | 4.0                             |
| NA                     | NA                              | Streptococci other than *S. pneumonia* | ≤1.0                            |

**Table 4** Local antibiotic concentrations of vancomycin and tobramycin

| Postoperative day | Vancomycin (µg/ml)* | Tobramycin (µg/ml)* |
|-------------------|---------------------|---------------------|
|                   | Mean (range)        | Mean (range)        |
| 1                 | 265 (28.3–736.4)    | 31 (6.4–97.2)       |
| 2                 | 172 (14.2–466.7)    | 9.4 (2.4–19.6)      |
| 3                 | 146 (5.8–394.5)     | 6.4 (1.6–19.9)      |
| 4                 | 146 (19.5–352.5)    | 5.3 (1.3–18.6)      |
| 5                 | 104 (5.8–190.5)     | 4.6 (2.2–9.8)       |

*Assayable values of antibiotics obtained from local exudate of drain samples. In the majority of assays conducted, maximum assayable limits for vancomycin and tobramycin were reported as >400 µg/ml and >20 µg/ml, respectively. As a result, mean values reported are lower than actual values.

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**Reduction of Vancomycin Concentrations in Postoperatively Collected Eluent**

![Fig. 2](image-url) Mean eluent vancomycin concentrations in the 5 postoperative days. Error bars represent the range of mean local vancomycin concentrations for each day.

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Δ Adis
By controlling the source of infection in total joint prosthetics through the disruption of biofilm formation, the surgeon creates an acute wound milieu that helps activate host immune mechanisms for effective wound healing. This control may be accomplished by combining the strict adherence to wound management through radical debridement with the administration of a local antibiotic depot capable of eluting potent concentrations for killing exposed biofilm [48]. Antibiotic-laden pure calcium sulfate hemihydrate aids in treatment of infected avascular bone isolated from systemic antibiotics while avoiding complications of toxicity associated with parenteral administration. A mixture of broad-spectrum antibiotics perpetuates eradication of heterogeneous bacterial phenotypes. We prefer vancomycin and tobramycin for this purpose as the concentrations eluded locally kill biofilm-related infections as demonstrated in the center for the biofilm engineering drip reactor system [48]. The pure form of the calcium sulfate carrier distinguishes itself from other preparations by its higher purity, physiologic pH, and hydrophilic properties. This bead becomes soft after hydration, allowing for placement in joints without scratching the articular surfaces, and demonstrates complete dissolution within a few weeks as illustrated by the radiographic films in Fig. 5 [49].

Monitoring the concentrations of antibiotic eluate allows for confidence in their therapeutic benefits, which depend on sustained blood concentrations at a minimum concentration while local concentrations remain high for the duration of therapy. Furthermore, excessive serum concentrations of vancomycin and tobramycin must be avoided because high concentrations may result in serious side effects, specifically ototoxicity and nephrotoxicity. The purpose of our study was therefore twofold: to verify that concentrations of vancomycin and tobramycin eluted from calcium sulfate would still be capable of inhibiting bacterial growth; to determine whether the antibiotics eluted from the pellet would remain

### Table 5 Serum concentrations of vancomycin and tobramycin in postoperative days 1–5

| Pt. ID no. | Day 1 | Day 2 | Day 3 | Day 4 | Day 5 |
|------------|-------|-------|-------|-------|-------|
|            | Vanc  | Tobra | Vanc  | Tobra | Vanc  | Tobra | Vanc  | Tobra | Vanc  | Tobra |
| 04         | 2.4   | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 |
| 05         | < 2.0 | 0.7   | < 2.0 | 0.7   | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 |
| 06         | 2.5   | 0.7   | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 |
| 08         | 2.6   | 1.8   | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 |
| 16         | 3.1   | 1.1   | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 |
| 21         | 2.2   | 0.9   | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 |
| 23         | 3.7   | 4.1   | 3.3   | 1.5   | 3.0   | < 0.5 | 2.5   | < 0.5 | < 2.0 | < 0.5 |
| 26         | < 2.0 | 0.6   | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 |
| 28         | < 2.0 | 2.1   | < 2.0 | 0.8   | < 2.0 | 0.6   | < 2.0 | < 0.5 | < 2.0 | < 0.5 |
| 30         | < 2.0 | 0.7   | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 |
| 32         | < 2.0 | 1.7   | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 |
| 35         | < 2.0 | 1.7   | < 2.0 | 0.7   | < 2.0 | < 0.5 | < 2.0 | < 0.5 | < 2.0 | < 0.5 |

Detectable values are highlighted in bold. Only patients with detectable serum values were included. Total of 50 patients tested.

*Vanc* vancomycin, *Tobra* tobramycin

![Graph](image-url)
local without elevating serum concentrations, which may result in general toxicity.

Limitations of this study include the ceiling placed on assayable values of antibiotic concentrations (as determined by FPIA analysis), the duration of evaluation, and the comparison between different volumes placed into wounds. Despite the truncated measurements, this study still demonstrates expected local and systemic elution characteristics. Regarding the temporal limitations, we would have preferred continued measurement of the antibiotic concentrations. However, our decision to discontinue after 5 days was based on concern for contamination from prolonged placement of the drains. Some of the patients in which we observed detectable serum concentrations also had concurrent implantation of PMMA cement impregnated with vancomycin and/or tobramycin as part of the reconstructive surgery. This concurrent implantation of an additional antibiotic delivery system may be a contributory factor to the incidences of detectable serum concentrations. Furthermore, controlling for the patient population may have yielded different results. For example, GFR changes with age and gender; therefore, medication levels may be different based on the specific patient. Also, a comparative group with PMMA or a control was not measured. Inclusion of a comparative group would have improved the specificity of the results and provided more evidence for the superiority of the Stimulan beads. However, there is significant evidence of the inefficacy of the PMMA for antibiotic implantation [16–18]. Additionally, this study was not designed for comparison against another product, but to show that the Stimulan beads

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can produce antibiotic concentrations high enough to kill bacteria related to biofilm infections but not too high to be toxic to the patient.

The clinical data also had limitations due to inaccuracy of antibiotic concentrations in the eluent as a result of the standard laboratory assay used. The laboratory agency conducting the FPIA analysis established standard routine reporting of antibiotic concentrations within specific concentration limits (vancomycin 2–400 mg/ml, tobramycin 0.5–20 mg/ml). Additionally, without the drains, higher concentrations of antibiotic would be expected locally for longer periods of time; however, drains are standard of care for patients with severe osteomyelitis. The exact values for antibiotic concentrations above the upper limits were not recorded for all patients. Therefore, the data do not give an accurate evaluation of local antibiotic concentrations in these cases. However, the data do indicate that these reported upper limits were present usually for only 1–2 days post-implantation. Thus, the use of antibiotic concentrations from eluent to indicate the diminishing local antibiotic concentrations over the 5-day period remains possible. In addition, the blood serum concentrations indicate that these high local concentrations post-implantation do not manifest into high systemic antibiotic concentrations. Moreover, additional laboratory data such as hepatic function, coagulation studies, and comorbidities would have been useful in controlling for any other variables that could confound the systemic antibiotic concentration measurements. However, the data were collected at the time of the initial study, and retrieval of the other information is not possible because of the deletion of the hospital records given some patient records eclipsed the 8-year maximum storage rule by the hospital system. Another limitation is that this study did not control for variability of the amount of antibiotic based on the volumetric size of the surgical bed. For example, larger surgical beds with the same number of pellets as smaller surgical beds will have smaller areas of antibiotic diffusion.

5 Conclusions

Based on our results and clinical experience, the antibiotic-loaded pure calcium sulfate hemihydrate demonstrates adequacy as a platform for the local delivery of antibiotics at therapeutic concentrations as well as a stable vehicle for incorporation of both vancomycin and tobramycin. In each of the 5 postoperative days evaluated, mean local concentrations of antibiotics exceeded values capable of inhibiting common pathogens. We observed no adverse reaction based on the presence of elevated serum concentrations of antibiotics and no occurrence of persistent wound drainage associated with the antibiotic-loaded pure calcium sulfate hemihydrate.

Acknowledgements

Pamela Jean Jensen MD. PamelaJensen@texas-health.org.

Compliance with Ethical Standards

Funding No source of funding was used to conduct this study.

Conflict of interest Dr. Maale has received royalty rights with Bicomposites Ltd. until April 1, 2020 and royalty rights currently with Smith and Nephew on the legion hk system. All other authors have no conflict of interest to declare.

Ethics approval All procedures in this study were in accordance with the 1964 Helsinki Declaration (and its amendments) and the details of the Ethics Committee or institutional review board which approved the study. The study was approved by the Institutional Review Board at Medical City Dallas.
Informed consent  At the time of the study, no additional consents were obtained. All consent related to the study was incorporated into the operative consent form.

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