Combining des-gamma-carboxyprothrombin and alpha-fetoprotein for hepatocellular carcinoma diagnosing: an update meta-analysis and validation study

Huaping Chen¹,*, Siyuan Chen¹,*, Shan Li¹,*, Zhijian Chen¹, Xuan Zhu¹, Meiyu Dai¹, Lingxi Kong¹, Xiaodan Lv¹, Zhili Huang¹ and Xue Qin¹

¹Department of Clinical Laboratory, First Affiliated Hospital of Guangxi Medical University, Nanning 530021, Guangxi, China

*These authors contributed equally to this work and should be considered as co-first authors

Correspondence to: Xue Qin, email: qinxue919@126.com

Keywords: des-gamma-carboxyprothrombin, AFP, hepatocellular cancer, diagnosis, meta-analysis

Received: June 22, 2017 Accepted: July 30, 2017 Published: August 07, 2017

Copyright: Chen et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License 3.0 (CC BY 3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

ABSTRACT

Controversies about the combination of des-gamma-carboxyprothrombin (DCP) and alpha-fetoprotein (AFP) for hepatocellular carcinoma diagnosing still exist. Hence, we performed this updated meta-analysis to estimate the diagnostic value of DCP, AFP and DCP + AFP in HCC. In addition, we conducted a validation study to analyze the performance of the candidate makers. After a systematic literature review, 27 studies from 20 articles were identified from four major databases. The pooled sensitivity and specificity were 69% and 89%, respectively, for DCP; for AFP, they were 65% and 88%, respectively; and they were 82% and 85%, respectively, for DCP + AFP. The values of the area under the curve (AUC) for DCP, AFP, DCP + AFP, respectively, were 0.88, 0.75, and 0.90. The validation study confirmed that the performance of DCP + AFP (sensitivity = 84%, specificity = 86%; AUC = 0.887) was higher than that of DCP (sensitivity = 76%, specificity = 92%; AUC = 0.843) or AFP (sensitivity = 73%, specificity = 92%; AUC = 0.837) alone.

INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the most (70–90%) frequent types of primary carcinoma of the liver and it is the third dominating cause of cancer-related mortality among men worldwide [1]. The incidence rate of HCC rises in accordance with increased rates of hepatitis C virus and hepatitis B virus infection [2]. Aflatoxin exposure, heavy alcohol drinking, nonalcoholic fatty liver disease, and smoking also contribute to the occurrence and progression of HCC [2].

Serum alpha-fetoprotein (AFP) is commonly used to diagnose HCC. However, owing to its low sensitivity and specificity, AFP testing alone is not recommended in diagnostic assessments of HCC [3–10]. Instead, it is used to combine with other serum or plasma tumor markers, which have been shown to have superior diagnostic abilities [3, 6, 11–15].

Des-gamma-carboxyprothrombin (DCP), an abnormal form of the prothrombin protein, is induced by a deficiency of vitamin K or antagonist-II [16]. The role of DCP as a biomarker of HCC was first reported in a study published 1984 [17], which found that DCP was present in 91% of HCC patients but not detectable in other benign liver diseases. Subsequently, numerous studies [6–8, 13, 18–20] demonstrated that a combined analysis of DCP and AFP led to better prediction of HCC although there are some controversies about the diagnostic accuracy of DCP + AFP.

Thus, we performed an update meta-analysis to contrast the diagnostic performance of DCP alone, AFP alone, and DCP + AFP in the detection of HCC. In addition, we performed a validation study of 45 HCC patients, 42 liver cirrhosis patients, 43 patients with hepatitis virus infections, and 44 normal controls to determine the diagnostic efficacy of these candidate markers.
RESULTS

Selection and characteristics of the included studies

As shown in Figure 1, a search of four databases (PubMed, Embase, the Cochrane Library, and the ISI Web of Science) and a manual search identified 951 relevant articles. Of these, 176 were published before 2000, and 231 were duplicates. After sieving the titles and abstracts, 412 articles about reviews, case reports, letters, nonhuman studies, or unrelated to the topic were excluded. The rest of 132 studies were deemed to be satisfactory, and they were read and evaluated carefully for a full-text review. Of these, 112 articles were eliminated for the following reasons: did not contain adequate data to construct a $2 \times 2$ table, did not contain sufficient information on the criteria used to diagnose HCC or analyzed only serum DCP and AFP alone, or they were published in non-English language periodicals. In total, 27 studies from 20 articles [11, 12, 14–16, 18–32] measured up for this meta-analysis and all of them were reported from 2000 to 2016.

This meta-analysis was included 7507 HCC patients and 5399 controls and the features of eligible studies were showed in Supplementary Table 1. All the included patients had undergone separate tests of serum and plasma levels of DCP and AFP as well as the test results of candidate markers (DCP, AFP, and DCP + AFP) had been analyzed by statistics.

The quality of the studies on the basis of the assessment scores of QUADAS [33–35] is summarized in Supplementary Table 2. Four articles met 10 of the 14 QUADAS norms, nine articles met 11 of the 14 QUADAS norms, six articles met 12 of the 14 QUADAS norms, and one article met 13 of the 14 QUADAS norms. According to the results of the QUADAS assessment, all the included studies were great in quality.

Pooled diagnostic performance of DCP, AFP, DCP + AFP in HCC diagnosing

Figure 2A–2C shows the sensitivity and specificity of the biomarkers: DCP: sensitivity ($I^2 = 89.6\%, P = 0.0000$) and specificity ($I^2 = 92.1\%, P = 0.0000$); AFP: sensitivity ($I^2 = 69.0\%, P = 0.0000$) and specificity ($I^2 = 90.0\%, P = 0.0000$); DCP + AFP: sensitivity ($I^2 = 79.9\%, P = 0.0000$) and specificity ($I^2 = 92.8\%, P = 0.0000$). As all the data pointed to significant heterogeneity in the meta-analysis, a random-effects model was chosen. When the data were pooled, the sensitivity and specificity of DCP, AFP, and

![Flow diagram of study selection process for eligible studies.](image-url)
DCP + AFP were 0.69 (0.68–0.70) and 0.89 (0.88–0.90), 0.65 (0.63–0.66) and 0.88 (0.87–0.89), and 0.82 (0.81–0.83) and 0.85 (0.85–0.86), respectively.

The summary receiver operator characteristic (SROC) curves of DCP, AFP, DCP + AFP are shown in Figures 3A–3C. The results demonstrated that the area

Figure 2: Forest plots of the sensitivity and specificity of candidate makers in the diagnosis of HCC. (A) DCP; (B) AFP; (C) DCP + AFP.
under the curve (AUC) values for DCP, AFP, DCP + AFP were, respectively, 0.88, 0.75, and 0.90. The pooled positive likelihood ratios (PLRs), negative likelihood ratios (NLRs), and diagnostic odds ratios (DORs) were 7.28 (5.52–9.61), 0.33 (0.29–0.37), 24.59 (17.98–33.62), respectively, for DCP; 5.07 (3.98–6.45), 0.41 (0.38–0.44), and 12.96 (9.90–16.98), respectively, for AFP; and 5.48 (4.31–6.96), 0.22 (0.19–0.25), and 26.45 (19.61–35.68), respectively, for DCP + AFP. The diagnostic performance of the candidate biomarkers are presented in Table 1. The results suggested that the diagnostic value of DCP + AFP in the detection of HCC was better than that of either DCP or AFP alone.

Threshold effects and meta-regression analysis of heterogeneity

To identify potential sources of heterogeneity, tests for threshold effects were conducted by Meta-Disc software. Spearman’s correlation coefficient value was 0.295 ($P = 0.135$) for DCP. It was $-0.007 (P = 0.971)$ and $0.060 (P = 0.765)$ for AFP and DCP + AFP, respectively, which suggested there were no threshold effects in this meta-analysis.

Other than threshold effects, the diversity of study populations in different countries, methodology used, and size of the study population can be potential sources of heterogeneity. To analyze these factors, a meta-regression was performed (Table 2). With regard to DCP, the results revealed no significant heterogeneity in respect of country (coefficient $= -0.132, P = 0.1736$), methodology (coefficient $= 0.044, P = 0.7654$), or size of the study population (coefficient $= 0.000, P = 0.2863$). It proved other factors might lead to the high heterogeneity of DCP. For AFP, they showed no significant heterogeneity in respect of the size of the study population (coefficient $= -0.001, P = 0.0525$). Interestingly, the results of the meta-regression indicated that differences in countries (coefficient $= -0.088, P = 0.0356$) and methodology (coefficient $= -0.106, P = 0.0454$) might be the source of heterogeneity in AFP studies.

Sensitivity analysis and publication bias

Removal of individual studies included in this meta-analysis was evaluated each time to determine the impact of the remaining data set on the sensitivity and specificity.
The merged results were stable, and they were not altered substantially by individual studies.

As shown in Figure 4A–4C, Begg’s funnel plot asymmetry pointed to potential publication bias in the included studies. Thus, a quantitative Egger’s linear regression test was conducted to provide statistic evidence of funnel plot symmetry or asymmetry. The results pointed to potential publication biases were detected in DCP (\(P = 0.000\)), AFP (\(P = 0.000\)) and DCP + AFP (\(P = 0.000\)) studies.

### Table 1: Meta-analysis results of DCP, AFP, DCP+AFP

| Maker     | Se (95%CI) | Sp (95%CI) | PLR (95%CI) | NLR (95%CI) | DOR (95%CI) | AUC |
|-----------|------------|------------|-------------|-------------|-------------|-----|
| DCP       | 0.69 (0.68-0.70) | 0.89 (0.88-0.90) | 7.28 (5.52-9.61) | 0.33 (0.29-0.37) | 24.59 (17.98-33.62) | 0.88 |
|           | 89.6% | 92.1% | 91.6% | 87.4% | 84.2% |
| AFP       | 0.65 (0.63-0.66) | 0.88 (0.87-0.90) | 5.07 (3.98-6.45) | 0.41 (0.38-0.44) | 12.96 (9.90-16.98) | 0.75 |
|           | 69.0% | 90.0% | 89.0% | 75.2% | 81.2% |
| DCP+AFP   | 0.82 (0.81-0.83) | 0.85 (0.85-0.86) | 5.48 (4.31-6.96) | 0.22 (0.19-0.25) | 26.45 (19.61-35.68) | 0.90 |
|           | 79.9% | 92.8% | 92.3% | 81.6% | 84.9% |

Se: sensitivity, Sp: specificity, PLR: positive likelihood ratio, NLR: negative likelihood ratio, DOR: diagnostic odds ratio, CI: confidence interval, AUC: area under ROC curve.

Validation study of the diagnostic performance of DCP + AFP in HCC

In the present study, there were 45 HCC patients (G 1), 43 liver cirrhosis patients (G 2), 42 patients with hepatitis virus infections (G 3), and 44 healthy individuals (G 4) (Supplementary Table 3). There was no statistically significance difference in the sex of the individuals in the four groups (\(P = 0.311\)). There was no statistic difference in

Figure 4: Begg’s funnel plot of candidate makers for the studies included in the meta-analysis. (A) DCP; (B) AFP; (C) DCP+AFP.
the age of the patients in the different groups: G 1 and G 2 (P  = 0.386), G 1 and G 3 (P  = 0.410), G 1 and G 4 (P  = 0.709), G 2 and G 3 (P  = 1.000), G 2 and G 4 (P  = 0.328), G 3 and G 4 (P  = 0.289). As shown in Figure 5 (A, B), the serum levels of both DCP and AFP were observably higher in G 1 than in G 2 (P  = 0.000 for both). They were also observably higher in G 1 compared to that in G 3 (P  = 0.000 for both) and meanwhile in G 1 than G 4 (P  = 0.000 for both). As shown in Supplementary Figure 1, the AUC values of DCP, AFP, and DCP + AFP in distinguishing between HCC (G 1) and liver cirrhosis (G 2) were 0.860 (95% CI: 0.770–0.925), sensitivity [Se]: 0.76, specificity [Sp]: 0.86), 0.805 (95% CI: 0.707–0.882, Se: 0.73, Sp: 0.84), and 0.859 (95% CI: 0.768–0.924, Se: 0.84, Sp: 0.77), respectively. In distinguishing HCC (G 1) from non-HCC (G 2, 3, and 4), the AUC values of DCP, AFP, and DCP + AFP were 0.843 (95% CI: 0.781–0.894, Se: 0.76, Sp: 0.92), 0.837 (95% CI: 0.774–0.889, Se: 0.73, Sp: 0.92), and 0.887 (95% CI: 0.830–0.930, Se: 0.84, Sp: 0.86), respectively (Table 3).

**DISCUSSION**

The results of this meta-analysis including 27 studies from 20 articles demonstrated that the overall sensitivity and specificity of DCP was, respectively, 69% and 89%. For AFP, the overall sensitivity and specificity was 65% and 88%. For DCP + AFP, these values were 82% and 85%, respectively. Although the specificity of the biomarkers did not differ greatly, the sensitivity of DCP + AFP was markedly higher, signifying the superior diagnostic value of DCP + AFP in HCC. DOR is a specific value of PLR and NLR. It is a single indicator of the performance of a trial but independent of prevalence. DOR ranges from zero to infinity, and higher values suggest preferable diagnostic performance. In the present study, the diagnostic odds ratios were 0.87 (0.72 to 1.06) for DCP, 0.87 (0.86 to 0.88) to 0.89 (0.89 to 0.90) for AFP, and 0.83 (0.82 to 0.84) to 0.87 (0.86 to 0.88) for DCP + AFP. The pooled specificity ranged from 0.88 (0.87 to 0.89) to 0.90 (0.89 to 0.91) for DCP, 0.87 (0.86 to 0.88) to 0.89 (0.88 to 0.90) for AFP, and 0.84 (0.83 to 0.85) to 0.87 (0.86 to 0.88) for DCP + AFP. Thus, sensitivity analyses in this meta-analysis illuminated that the pooled result and conclusions did not fluctuate by the various decisions that could be made during articles reviewing. It signifies the results of our meta-analysis can be considered with a higher credibility. The results of Egger’s linear regression test of DCP, AFP and DCP + AFP in HCC diagnosing were consistent with Begg’s funnel plots, which demonstrated that publication bias may be present in this meta-analysis. In the present study, we used an optimal search strategy, which included a comprehensive search of four English databases and a manual search to ensure that no relevant articles were overlooked. However, studies with negative results may remain unpublished, with both authors.

**Table 2: Meta-regression analyses of the heterogeneity in DCP and AFP**

| Variable | Coeff. | Std.Err. | P-value | RDOR (95%CI) | Coeff. | Std.Err. | P-value | RDOR (95%CI) |
|----------|--------|----------|---------|-------------|--------|----------|---------|-------------|
| Country  | −0.132 | 0.0940   | 0.1736  | 0.88        | (0.72; 1.06) | −0.088 | 0.0394 | 0.0256 | (0.84; 0.99) |
| Method   | 0.044  | 0.1454   | 0.7654  | 1.04        | (0.77; 1.41) | −0.106 | 0.0454 | 0.0291 | (0.82; 0.99) |
| Population | 0.000  | 0.0004   | 0.2863  | 1.00        | (1.00; 1.00) | 0.001 | 0.0003 | 0.0525 | (1.00; 1.00) |

Std.Err: standard error, RDOR: ratio of diagnostic odds ratio, CI: confidence interval.
and editors tending to publish positive results of which these two inestimable reasons may lead to publication bias in this article. Therefore, we further performed the trim and fill method to adjust for publication bias in the present meta-analysis [36]. For DCP in HCC diagnosing studies, the overall analysis merging the theoretical 33 (27 + 6) studies had little variance with the unredressed 27 studies (variance = 0.217, \( P = 0.000 \) vs. variance = 0.230, \( P = 0.000 \)) and that was similar to DCP + AFP studies (variance = 0.297, \( P = 0.000 \) vs. variance = 0.341, \( P = 0.000 \)); for AFP researches, none hypothetical study was added and no data were changed between adjusted and unadjusted ones (variance = 0.135, \( P = 0.000 \) for both) (Supplementary Figure 2). All of these results implied that the absent articles with negative results did not influence significantly the overall accuracy in our meta-analysis.

In our literature search, there are three similar meta-analyses that also assessed DCP, AFP, and DCP + AFP for HCC diagnosing [3–5]. Compared with the previous studies [3–5], the advantages of the present meta-analysis

**Figure 5: Levels of DCP and AFP in four different groups.** DCP was measured in mAU/ml and AFP in ng/ml. Median of original data (minimum to maximum) was shown respectively in the figures.
are as follows: First, this meta-analysis evaluated the diagnostic performance of DCP + AFP, whereas the previous meta-analyses focused on multiple serum biomarkers for HCC. The present meta-analysis is less ambiguous than previous ones. Second, this meta-analysis included more studies on DCP and AFP and larger sample sizes than previous studies. Third, all the included HCC cases were diagnosed according to gold standard methods to further enhance the validity of the findings. Fourth, the data on DCP and AFP were derived from identical groups to strengthen the comparability of the diagnostic accuracy of the candidate markers (DCP, AFP, and DCP + AFP). Fifth, we included a number of factors (study populations from different countries, test methods, and sample size) not included in previous meta-analyses to explore the source of heterogeneity. Besides, the major orientation in our meta-analysis was similar with a systematic review [37] published in 2014 but different in topics. The present meta-analysis was aimed at the diagnostic performance of DCP + AFP when another aimed to compare the deference between DCP and AFP as biomarkers in diagnosing HCC. In that review [37], the overall evaluations were: sensitivity = 0.63, specificity = 0.91, AUC = 0.83 for DCP, sensitivity = 0.59, specificity = 0.86, AUC = 0.77 for AFP, sensitivity = 0.81, specificity = 0.83, AUC = 0.88 for DCP + AFP. All of the summary results in our meta-analysis were agreed with that in that review. Comparing the systematic review [37], the merits of our meta-analysis were listed below: First, the strict inclusive and exclusive criteria were constituted in this meta-analysis, an English-language restriction and included HCC patients must be diagnosed by histology, for instance. Second, high quality was performed in each of included articles according to evaluated tool but the precious review was not. Finally, we conducted the analyses of threshold effects and meta-regression to explore the heterogeneity.

Based on the results of our meta-analysis and validation study, the diagnostic performance of a combination of DCP + AFP is prominent to that of DCP or AFP alone in the detection of HCC. Further research with optimized designs, greater numbers of studies, and larger sample sizes are required to shed light on remaining inconsistencies in variant results of combining des-gamma-carboxyprothrombin and alpha-fetoprotein for hepatocellular carcinoma diagnosing.

**MATERIALS AND METHODS**

**Search strategy**

A systematic literature search was conducted for relevant articles published in PubMed, Excerpta Medica Database (Embase), the Cochrane Library, and ISI Web of Science before 27 December 2016. For AFP, the search terms were AFP and α-fetoprotein. For DCP, the search terms were acarboxyprothrombin, PIVKA II, DCP, decarboxyprothrombin, non-carboxylated factor II, protein induced by vitamin K absence, and antagonists. For HCC, the search terms were liver neoplasms, hepatic neoplasms, hepatocellular cancer, liver cell carcinomas, HCC, small hepatocellular carcinoma, and SHCC. Moreover, the

**Table 3: Diagnostic values of candidate markers for hepatocellular carcinoma**

| Makers | AUC (95% CI) | Cut-off value | Se   | Sp   | Youden index |
|--------|--------------|---------------|------|------|--------------|
| DCP    |              |               |      |      |              |
| HCC vs. cirrhosis | 0.860 | 0.770–0.925 | 40 mAU/ml | 0.76 | 0.86 | 0.62 |
| HCC vs. no-HCC | 0.843 | 0.781–0.894 | -     | 0.76 | 0.92 | 0.67 |
| AFP    |              |               |      |      |              |
| HCC vs. cirrhosis | 0.805 | 0.707–0.882 | 11 ng/ml | 0.73 | 0.84 | 0.57 |
| HCC vs. no-HCC | 0.837 | 0.774–0.889 | -     | 0.73 | 0.92 | 0.66 |
| DCP+AFP |          |               |      |      |              |
| HCC vs. cirrhosis | 0.859 | 0.768–0.924 | -     | 0.84 | 0.77 | 0.61 |
| HCC vs. no-HCC | 0.887 | 0.830–0.930 | -     | 0.84 | 0.86 | 0.70 |

AUC: area under the curve; CI: confidence interval; Se: sensitivity; Sp: specificity.
Eligibility criteria and study selection

To be involved in the meta-analysis, all the articles must fulfill the criteria as follows: (i) contain both individual and combined sensitivity and specificity data on serum or plasma DCP and AFP assays, (ii) consist of patients with a pathologically proven diagnosis of HCC. The exclusion criteria were as follows: (i) reviews, meta-analyses, case reports, or letters; (ii) nonhuman studies; (iii) unrelated to the topic; (iv) incomplete information; and (v) non-English. When study populations overlapped, only the largest sized study or most comprehensive study was incorporated in the concluding analysis.

Data extraction and quality assessment

The following information was collected from each paper: the name of author, year of publication, country of research, quantity of cases with HCC, quantity of contrasts which may included healthy individuals, patients with hepatitis, cirrhosis or other nonmalignant hepatopathy and non-HCC cancers, assay methods, cut-off points, and the sensitivity and specificity of DCP and AFP alone and their combination (DCP + AFP). The QUADAS (Quality Assessment of Diagnostic Accuracy Studies) tool [33–35] was applied to evaluate the quality of the contained studies. Using the 14-item tool, the evaluators rated each item as “Yes”, “No,” or “Unclear.” The reviewers assigned a mark of “1” for “Yes” and “0” for “No” or “Unclear” to each of the 14 items. A high-quality article was deemed one that attained a final Q score of 10 or more.

Validation study of the diagnostic value of DCP, AFP, and DCP + AFP in diagnosing HCC

The study consisted of HCC patients (G 1, n = 45), liver cirrhosis patients (G 2, n = 43), patients with hepatitis virus infections (G 3, n = 42), and healthy controls (G 4, n = 44) matched in sex and age from the First Affiliated Hospital of Guangxi Medical University. HCC was diagnosed based on two or more typical imaging modalities or histological examinations. Cirrhosis patients were excluded if they showed evidence of progression to HCC. Patients with hepatitis virus infections were determined by laboratory diagnoses. Healthy subjects who tested negative results of hepatitis B virus surface antigen and with the normal levels of both of aspartate transaminase and alanine transaminase, were included. Individuals with obstructive jaundice and those taking warfarin, as well as samples with hemolysis and lipemia, were also excluded.

Serum levels of DCP were determined using a chemiluminescent immunoassay assay, and AFP concentrations were resolved by electrochemiluminescence immunoassay, according to the manufacturer’s instructions.

Statistical Analysis

Meta-analysis

Heterogeneity was evaluated by the I² test. A value of \( P < 0.1 \) and \( I^2 > 50\% \) were considered indicative of significant heterogeneity. A random effects model was selected in cases of obvious heterogeneity \( (P < 0.1, I^2 > 50\%) \). Otherwise \( (i.e., in cases of \( P \geq 0.1, I^2 \leq 50\%) \), a fixed effects model was applied. To evaluate the performance of the diagnostic studies, Spearman’s correlation coefficient was applied to test for threshold effects and the overall sensitivity, specificity, likelihood ratios, and diagnostic odds ratios (DORs), together with their 95% confidence intervals (CIs), in addition to their summary receiver operating characteristic (SROCs) curves and area under the curve (AUCs) were calculated by the quantities of true positive, false positive, false negative, and true negative. Furthermore, a meta-regression analysis, sensitivity analysis, and analyses for publication bias were executed simultaneously. All the statistic analyses were conducted utilizing Meta-Disc software (version 1.4) and Stata (version 12.0).

Validation study

In the validation study, the statistical data analyses were performed with IBM SPSS (version 20), and sectional graphics were executed using GraphPad Prism (version 5.01). A chi-square test was applied to compare classified variables. The Mann–Whitney test was performed to compare differences between two groups. A two-tailed test of \( P \)-value \( < 0.05 \) was considered as statistical differences. ROC curve and binary logistic regression analyses were used to calculate the areas under the ROC (AUROC) of DCP + AFP. The AUC was served for comparing the performance of DCP, AFP, and DCP + AFP. The cutoff values for DCP and AFP were 40mAU/ml and 11ng/ml, in accordance with the manufacturer’s instructions. As hepatic cirrhosis is known to be a pathogenic factor that is closely related to HCC, in this study, we analyzed the AUC, sensitivity, specificity, and Youden index in patients with and without HCC (G 1 vs. G 2, 3, and 4) and in HCC patients and liver cirrhosis patients (G 1 vs. G 2).

Abbreviations

DCP: des-gamma-carboxyprothrombin; AFP: alpha-fetoprotein; HCC: Hepatocellular carcinoma; AUC: area under the curve; QUADAS: Quality Assessment of Diagnostic Accuracy Studies; Se: sensitivity; Sp: specificity; PLRs: positive likelihood ratios; NLRs: negative likelihood ratios; DORs: diagnostic odds ratios; CIs: confidence intervals.
Author contributions

XQ and SL conceived and designed the meta-analysis and validation study. HC, SC and ZC performed the literature search and quality assessment. LK and XL implemented the data extraction. ZH and ZC executed the validation study. MD, HC conducted the statistical analyses. HC and SC wrote the manuscript.

ACKNOWLEDGMENTS

We thank the authors of the primary articles included in our meta-analysis. We could not perform the review without their previous studies. This work is subsidized by National Natural Science Foundation of China (NO.81460431) and Guangxi natural science funds (2013GXNSFAA253001).

CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

FUNDING

This work is supported by National Natural Science Foundation of China (NO.81460431) and Guangxi natural science funds (2013GXNSFAA253001).

REFERENCES

1. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A. Global cancer statistics, 2012. CA Cancer J Clin. 2015; 65:87–108. https://doi.org/10.3322/caac.21262.
2. El-Serag HB. Hepatocellular carcinoma. N Engl J Med. 2011; 365:1118–27. https://doi.org/10.1056/NEJMra1001683.
3. Hussein TD. Serological tumor markers of hepatocellular carcinoma: a meta-analysis. Int J Biol Markers. 2015; 30:e32–42. https://doi.org/10.5301/jbm.5000119.
4. Huang TS, Shyu YC, Turner R, Chen HY, Chen PJ. Diagnostic performance of alpha-fetoprotein, lens culinaris agglutinin-reactive alpha-fetoprotein, des-gamma carboxyprothrombin, and glypican-3 for the detection of hepatocellular carcinoma: a systematic review and meta-analysis protocol. Syst Rev. 2013; 2:37. https://doi.org/10.1186/2046-4053-2-37.
5. Hu B, Tian X, Sun J, Meng X. Evaluation of individual and combined applications of serum biomarkers for diagnosis of hepatocellular carcinoma: a meta-analysis. Int J Mol Sci. 2013; 14:23559–80. https://doi.org/10.3390/ijms141223559.
6. Viggiani V, Palombi S, Gennarini G, D’Ettorre G, De Vito C, Angeloni A, Frati L, Anastasi E. Protein induced by vitamin K absence or antagonist-II (PIVKA-II) specifically increased in Italian hepatocellular carcinoma patients. Scand J Gastroenterol. 2016; 51:1257–62. https://doi.org/10.1080/00365521.2016.1183705.
7. Sultanik P, Ginguay A, Vandame T, Popovici T, Meritet JF, Cynober L, Pol S, Bories PN. Diagnostic accuracy of des-gamma-carboxy prothrombin for hepatocellular carcinoma in a French cohort using the Lumipulse(R) G600 analyzer. J Viral Hepat. 2016. https://doi.org/10.1111/jvh.12622.
8. Pote N, Cauchy F, Albuquerque M, Voitot H, Belghiti J, Castella L, Puy H, Bedossa P, Paradis V. Performance of PIVKA-II for early hepatocellular carcinoma diagnosis and prediction of microvascular invasion. J Hepatol. 2015; 62:848–54. https://doi.org/10.3103/j.jhep.2014.11.001.
9. Abd El Gawad IA, Mossaalam GI, Radwan NH, Elzawahry HM, Ellifnawy NM. Comparing prothrombin induced by vitamin K absence-II (PIVKA-II) with the oncofetal proteins glypican-3, Alpha feto protein and carinoembryonic antigen in diagnosing hepatocellular carcinoma among Egyptian patients. J Egypt Natl Canc Inst. 2014; 26:79–85. https://doi.org/10.1016/j.jenic.2014.01.001.
10. Baek YH, Lee JH, Jang JS, Lee SW, Han JY, Jeong JS, Choi JC, Kim HY, Han SY. Diagnostic Role and Correlation with Staging Systems of PIVKA-II compared with AFP. Hepato-Gastroenterology. 2009; 56:763–7.
11. Lim TS, Kim DY, Han KH, Kim HS, Shin SH, Jung KS, Kim BK, Kim SU, Park JY, Ahn SH. Combined use of AFP, PIVKA-II, and AFP-L3 as tumor markers enhances diagnostic accuracy for hepatocellular carcinoma in cirrhotic patients. J Hepatol. 2016; 51:344–53. https://doi.org/10.3109/00365521.2015.1082190.
12. Jang ES, Jeong SH, Kim JW, Choi YS, Leissner P, Brechot C. Diagnostic Performance of Alpha-Fetoprotein, Protein Induced by Vitamin K Absence, Osteopontin, Dickkopf-1 and Its Combinations for Hepatocellular Carcinoma. PLoS One. 2016; 11:e0151069. https://doi.org/10.1371/journal.pone.0151069.
13. Caviglia GP, Abate ML, Petrini E, Gaia S, Rizzetto M, Smedile A. Highly sensitive alpha-fetoprotein, Lens culinaris agglutinin-reactive fraction of alpha-fetoprotein and des-gamma-carboxyprothrombin for hepatocellular carcinoma detection. Hepatology Research. 2016; 46:E130–E5. https://doi.org/10.1111/hepr.12544.
14. Seo SI, Kim HS, Kim WJ, Shin WG, Kim DJ, Kim KH, Jang MK, Lee JH, Kim JS, Kim HY, Kim DJ, Lee MS, Park CK. Diagnostic value of PIVKA-II and alpha-fetoprotein in hepatitis B virus-associated hepatocellular carcinoma. World J Gastroenterol. 2015; 21:3928–35. https://doi.org/10.3748/wjg.v21.i13.3928.
15. Song P, Feng X, Inagaki Y, Song T, Zhang K, Wang Z, Zheng S, Ma K, Li Q, Kong D, Wu Q, Zhang T, Zhao X, et al. Clinical utility of simultaneous measurement of alpha-fetoprotein and des-gamma-carboxy prothrombin for diagnosis of patients with hepatocellular carcinoma.

www.impactjournals.com/oncotarget
1. Yoon YJ, Han KH, Kim DY. Role of serum prothrombin induced by vitamin K absence or antagonist-II in a Chinese population. Surgery. 2016; 160:748–54. https://doi.org/10.1016/j.surg.2016.03.048.

2. Hady AA, El Shanawany F, Hassan MM, Anas A, Mostafa I. Evaluation of Human Telomerase Activity as a Novel Tumor Marker for Hepatocellular Carcinoma. Life Science Journal-Acta Zhengzhou University Overseas Edition. 2010; 7:153–61. https://doi.org/10.11159/000346080.

3. Marrero JA, Su GL, Wei W, Emick D, Conjeevaram HS, Fontana RJ, Lok AS. Des-gamma carboxyprothrombin can differentiate hepatocellular carcinoma from nonmalignant chronic liver disease in american patients. Hepatology. 2003; 37:1114–21. https://doi.org/10.1053/jhep.2003.50195.

4. Cui R, Wang B, Ding H, Shen H, Li Y, Chen X. Usefulness of determining a protein induced by vitamin K absence in detection of hepatocellular carcinoma. Chin Med J (Engl). 2002; 115:42–5.

5. Ikoma J, Kaito M, Ishihara T, Nakagawa N, Adachi Y. Early diagnosis of hepatocellular carcinoma using a sensitive assay for serum des-gamma-carboxy prothrombin: a prospective study. Hepatogastroenterology. 2002; 49:235–8. https://doi.org/10.1111/j.1572-0241.2006.00681.x.

6. Shimizu A, Shiraki K, Ito T, Sugimoto K, Ohmori S, Murata K, Takeda, T, Nakano T. Sequential fluctuation pattern of serum des-gamma-carboxy prothrombin levels detected by high-sensitive electrochemiluminescence system as an early predictive marker for hepatocellular carcinoma in patients with cirrhosis. Int J Mol Med. 2002; 9:245–50.

7. Ishii M, Gama H, Chida N, Ueno Y, Shinzawa H, Takagi T, Adachi Y. Early diagnosis of hepatocellular carcinoma in patients with cirrhosis. Hepatol Int. 2002; 6:110–5. https://doi.org/10.1111/j.1572-0241.2006.00681.x.

8. Volk ML, Hernandez JC, Su GL, Lok AS, Marrero JA. Risk factors for hepatocellular carcinoma may impair the performance of biomarkers: a comparison of AFP, DCP, and AFP-L3. Cancer Biomark. 2007; 3:79–87.

9. Nakamura S, Nousu K, Sakaguchi K, Ito YM, Ohashi Y, Kobayashi Y, Toshikuni N, Tanaka H, Miyake Y, Matsumoto E, Shiratori Y. Sensitivity and Specificity of Des-Gamma-Carboxy Prothrombin for Diagnosis of Patients with Hepatocellular Carcinomas Varies According to Tumor Size. Am J Gastroenterol. 2006; 101:2038–43. https://doi.org/10.1111/j.1572-0241.2006.00681.x.

10. Wang CS, Lin CL, Lee HC, Chen KY, Chiang MF, Chen HS, Lin TJ, Liao LY. Usefulness of serum des-gamma-carboxy prothrombin in detection of hepatocellular carcinoma. World J Gastroenterol. 2005; 11:6115–9.
36. Duval S, Tweedie R. Trim and fill: A simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. Biometrics. 2000; 56:455–63.

37. Li C, Zhang Z, Zhang P, Liu J. Diagnostic accuracy of des-gamma-carboxy prothrombin versus alpha-fetoprotein for hepatocellular carcinoma: A systematic review. Hepatol Res. 2014; 44:E11–25. https://doi.org/10.1111/hepr.12201.

38. Yu R, Ding S, Tan W, Tan S, Tan Z, Xiang S, Zhou Y, Mao Q, Deng G. Performance of Protein Induced by Vitamin K Absence or Antagonist-II (PIVKA-II) for Hepatocellular Carcinoma Screening in Chinese Population. Hepat Mon. 2015; 15:e28806. https://doi.org/10.5812/hepatmon.28806v2.

39. Choi JY, Jung SW, Kim HY, Kim M, Kim Y, Kim DG, Oh EJ. Diagnostic value of AFP-L3 and PIVKA-II in hepatocellular carcinoma according to total-AFP. World J Gastroenterol. 2013; 19:339–46. https://doi.org/10.3748/wjg.v19.i3.339.

40. Beale G, Chattopadhyay D, Gray J, Stewart S, Hudson M, Day C, Tserotoli P, Giannelli G, Manas D, Reeves H, AFP, PIVKAII, GP3, SCCA-1 and follisatin as surveillance biomarkers for hepatocellular cancer in non-alcoholic and alcoholic fatty liver disease. Bmc Cancer. 2008; 8. https://doi.org/10.1186/1471-2407-8-200.