Occurrence of anti-\textit{Toxoplasma gondii} and anti-\textit{Neospora caninum} antibodies in pigs in the State of Pará, Brazil

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Abstract

We investigated the occurrence of \textit{Toxoplasma gondii} and \textit{Neospora caninum} antibodies in pigs raised in the Northeast of Pará, Brazil. At Study I, convenience sampled 151 pigs at two slaughterhouses, with and without state inspection; and Study II, which assessed 159 pigs with probabilistic sampling from nine pig farms. Serological analysis was performed using indirect fluorescent antibody test for \textit{T. gondii} and \textit{N. caninum} with a cutoff of 64 and 50, respectively. Overall, 6.77% pigs were seropositive for \textit{T. gondii} and 5.16% for \textit{N. caninum}. In Study I, pigs slaughtered with and without state inspection presented similar occurrence for both coccidia (p>0.05). Study II found an association between \textit{N. caninum} seropositivity and sludge discarded into the soil, feeding pigs with animal-based protein, subsistence system, and absence of nipple drinkers. No association was found for \textit{T. gondii}. Pigs from Pará are a potential source of \textit{T. gondii} infection to humans. To our best knowledge, this is the first study to report anti-\textit{N. caninum} antibodies in the serum of pigs in Pará State, Brazilian Amazon.

Keywords: Swine, coccidian, Brazilian Amazon, \textit{Toxoplasma gondii}, \textit{Nespora caninum}.

Resumo

Foi investigada a ocorrência de anticorpos contra \textit{Toxoplasma gondii} e \textit{Neospora caninum} em suínos criados no nordeste do Pará, Brasil. No Estudo I, foram amostrados 151 porcos em dois matadouros, com e sem inspeção estadual. O Estudo II avaliou 159 suínos com amostragem probabilística de nove granjas de suínos. Para sorologia, utilizou-se o teste de imunofluorescência indireta para \textit{T. gondii} e \textit{N. caninum} com ponto de corte de 1:64 e 1:50, respectivamente. No geral, 6,77% dos suínos foram soropositivos para \textit{T. gondii} e 5,16% para \textit{N. caninum}. No Estudo I, suínos abatidos em matadouros com e sem inspeção estadual apresentaram ocorrência semelhante para ambos os coccídios (p>0,05). Os animais amostrados de Belém, Benevides, Marituba, Bujaru, Castanhal e Igarapé-Mirí foram positivos para \textit{T. gondii}, enquanto os soropositivos para \textit{N. caninum} foram encontrados em Belém, Bujaru, Castanhal e Santo Antônio do Tauá. O Estudo II encontrou associação entre soropositividade de \textit{N. caninum} e esterco descartado no solo, alimentação dos suínos com proteína de origem animal, criação de subsistência e ausência de bebedores tipo “nipple”. Não foi encontrada associação para \textit{T. gondii}. A carne suína apresenta potencial risco de transmissão de \textit{T. gondii} para os habitantes da região. De acordo com nosso conhecimento, este é o primeiro relato de anticorpos anti-\textit{N. caninum} em suínos no estado do Pará, Amazônia brasileira.

Palavras-chave: Suínos, coccídios, Amazônia, \textit{Toxoplasma gondii}, \textit{Nespora caninum}.
Introduction

Data from the latest agricultural census demonstrates the robustness of Brazilian pig farming, with the production of pig meat having almost doubled between 1996 and 2006, from 1.2 to 2.3 million tons. This improvement has been boosted by the increase in the exports that quadrupled in the same period, from 4.5% to 21.1% (IBGE, 2017). In contrast, in the northern region, pig farming did not show the same technological improvement and was characterized by subsistence production (self-consumption) or as a complementary source of income through regular or sporadic marketing in local consumer markets. In the State of Pará, the expansion of cattle farming has coincided with a sharp decline in the number of pig farms, which has practically not evolved over the years, being restricted to small properties (Silva et al., 2017).

Toxoplasma gondii is an intracellular protozoan that is distributed worldwide and is capable of infecting birds and mammals, including humans, with an estimated global seroprevalence of 19% (95% confidence interval [CI]: 17–22%) in pigs (Foroutan et al., 2019). In the Amazon region, serological studies have already demonstrated the prevalence of *T. gondii* in wild animals (Minervino et al., 2010; Soares et al., 2011; Vitaliano et al., 2015) including primates (Minervino et al., 2017), dogs (Valadas et al., 2010; Minervino et al., 2012), horses (Moreira et al., 2019) and pigs (Cavalcante et al., 2006; Freitas et al., 2009; Sousa et al., 2014). To our best knowledge, there is only one report on the occurrence of *T. gondii* antibodies in pigs in Pará State, which was limited to the municipality of Belém (Freitas et al., 2009), and reported that 50% (55/110) of the tested animals were seropositive.

In pigs, the main form of *T. gondii* infection is horizontal transmission; however, transplacental transmission is responsible for clinical manifestations of fetal death and mummification, preterm births, and varied symptoms (Dubey, 2009).

Neospora caninum is an intracellular protozoan that causes multisystem infection, and has domestic and wild canids (dingo, coyote, and gray wolf) as definitive hosts, and several warm-blooded animals as intermediate hosts (Donahoe et al., 2015; Cerqueira-Cézar et al., 2017). Studies in the State of Pará found *N. caninum* antibodies in cattle (Minervino et al., 2008; Chiebao et al., 2015) and dogs (Paz et al., 2019) however, there are no reports on the occurrence of *N. caninum* in pigs in this region.

Considering the lack of studies regarding the occurrence of *T. gondii* in the region and that infection by this parasite in pigs continues to be of public health concern (Dubey et al., 2020), and due pigs being a possible source of infection to *N. caninum* definitive hosts, this study aimed to determine the presence of antibodies and the associated risk factors against both parasites in pigs reared in the State of Pará, Brazil.

Material and Methods

This study was approved by the Animal Ethics Commission of the Federal Rural University of Pará (protocol 038/2013, and process 23084.010923/2013-15).

Study area

The study was conducted in the Northeast of Pará state, Northern Brazil, located in the Amazon Biome (Figure 1). The climate of the region is characterized by an annual mean temperature of 25.9 °C and an annual mean rainfall of 2,150 mm, with a strong concentration from January to June, and more rarely, from July to December (Martorano et al., 2017).

Two different serological studies were conducted during the study period: Study I was performed as initial screening for the occurrence of *T. gondii* and *N. caninum* in the region, and Study II was a cross-sectional epidemiological survey.

Study I sampled a total of 151 pigs by convenience sampling method at two slaughterhouses - one with State Inspection Service (SIS) (n=96) and the other without SIS (n=55). At the initial screening, all animals were adults. The only information available was the origin of the pigs. Animals from three distinct regions of Pará State, Northeast, Marajó, and Belém Metropolitan area, were included in this study. Pigs where from the following cities Bujaru, Castanhal, Belém, Marituba and from the Marajó island. (unknown city).

Study II was a prospective cross-sectional epidemiological survey of pig farms from the Northeast and Belém metropolitan areas of Pará State. The number of animals to be sampled was calculated using the Epi Info™ software.
Coccidia antibodies in pigs from Pará, Brazil

(U.S. Centers for Disease Control and Prevention, Atlanta, USA) with the following parameters: herd population (41,443,594 pigs); expected prevalence of 9.3 (based on the T. gondii results of Study I, which presented the highest occurrence), expected error of 5%, and confidence interval of 97%, which indicated the requirement for 129 samples. Thus, blood samples from 159 pigs were collected from nine pig farms. A epidemiological questionnaire was applied to the individuals responsible for the farms, which had questions regarding facilities, type of management, production characteristics, hygiene–sanitary features, and the presence/absence of cats and dogs. The age of the pigs varied from one to seven months. Pigs were sampled in the following cities: Castanhal, Igarapé-miri, Santo Antônio do Tauá, Belém and Benevides.

For Study I, blood samples were obtained during the slaughter of the pigs. Samples for Study II were drawn by cephalic puncture of the pigs in the farms. The samples were kept at room temperature until clotting and then transported to the laboratory in an ice cooler. The serum was collected, centrifuged, and stored at −20 °C until analysis.

Figure 1. Study site with the municipalities included in Study I and Study II. In the study I samples were from Marajó island but the specific municipality could not be confirmed.
Coccidia antibodies in pigs from Pará, Brazil

Serological analysis

The presence of antibodies to *T. gondii* and *N. caninum* was determined by an indirect fluorescent antibody test (IFAT). The test antigens were derived from whole tachyzoites of the RH strain of *T. gondii*, maintained in Swiss albino mice, and from whole tachyzoites of the NC-1 strain of *N. caninum*, maintained in an M-617 monocyte culture (André et al., 2010).

The IFAT was performed according to previously described protocols using a 1:64 cutoff for *T. gondii* (Vidotto et al., 1990) and 1:50 cutoff for *N. caninum* (Azevedo et al., 2010). Positive and negative controls were added to each slide. Positive sera were subsequently diluted two-fold serially and the sample titer was the highest dilution which provided a positive result.

Statistical analysis

The frequency of positive results for each parasite was analyzed separately for Study I and Study II. Study I evaluated the association of seropositive animals with the municipality of origin and the presence or absence of SIS in the abattoir using the χ² test or the Fisher exact test.

For Study II, the associations between the possible epidemiological risk factor variables and the prevalence of *T. gondii* and *N. caninum* antibodies were estimated using the χ² test by the Mantel-Haenszel method, considering the serological status (positive or negative) of the animals as a dependent variable for each parasite studied. The odds ratios and respective 95% CIs were also calculated. The SAS statistical program was used, with at least 5% (p <0.05) significance level.

Results

The total number of pigs sampled in Studies I and II was 310, which presented an occurrence of 11.3% (35/310) and 5.2% (16/310) for *T. gondii* and *N. caninum*, respectively. The occurrence of antibodies to *T. gondii* and *N. caninum* in pigs from Studies I and II are presented in Table 1. Animals sampled from Belém, Benevides, Marituba, Bujaru, Castanhal, and Igarapé-Miri were positive for *T. gondii*, whereas those seropositive for *N. caninum* were found in Belém, Bujaru, Castanhal, and Santo Antônio do Tauá.

|       | *Toxoplasma gondii* |       | *Neospora caninum* |
|-------|---------------------|-------|--------------------|
|       | Positive/tested | %    | Positive/tested | %    |
| I     | 14/151         | 9.3  | 7/151              | 4.6  |
| II    | 21/159         | 13.2 | 9/159              | 5.7  |
| Overall | 35/310        | 11.3 | 16/310             | 5.2  |

Table 1. Frequency of antibodies to *T. gondii* and *N. caninum* in the sera of pigs from slaughterhouses (Study I) or free reared (Study II) in the State of Pará.

No association was observed between the frequency of pigs positive for *T. gondii* and *N. caninum* and abattoir with and without SIS (p>0.05). Six of 96 (6.25%) and 8 of 55 (14.54%) pigs were seropositive for *T. gondii* in slaughterhouses with and without SIS, and 3 of 96 (3.13%) and 4 of 55 (7.27%) pigs were seropositive for *N. caninum* in the slaughterhouses with and without SIS, respectively.

The number of pigs positive for *T. gondii* and *N. caninum*, stratified by age groups, in Study II are presented in Tables 2 and 3, respectively. Antibodies against *N. caninum* were observed in piglets aged less than 1, 3, 4, and 6 months of age. No statistical difference was observed in pigs for both *T. gondii* and *N. caninum* when stratified by their age group (p>0.05).

In Study II, the associations between the presence of *T. gondii* and *N. caninum* and the following variables in seropositive animals were analyzed: presence of cats, presence of dogs, sludge discarded into the soil, feeding pigs with animal-origin protein, subsistence system or intensive confinement system, type of drinker, and breeding together with ruminant or poultry. There was no association between seropositivity to *T. gondii* and the variables evaluated. The presence of antibodies to *N. caninum* was associated with the following variables: sludge discarded into the soil, feeding pigs with animal-origin protein, and absence of suction (nipple) drinkers (Table 4).
Discussion

*T. gondii* occurrence (6.77%) in pigs from Pará was higher than that found in growing-finishing pigs in Paraná: 2.60% (Carletti et al., 2005), and in pigs slaughtered in São Paulo and Minas Gerais: 0% (Pezerico et al., 2007); in the later study, the absent of positive animals was probably due improvements in hygienic and sanitary management adopted in large-scale pig farming. It is noteworthy that infection with *T. gondii* is widespread in our study area, which was verified in 6 of the 7 municipalities investigated (85.71%).
In contrast, the frequency of infection was lower than that observed in slaughter pigs in the state of Paraiba, with occurrence of 36.2% (Azevedo et al., 2010) and 19.5% (Feitosa et al., 2014). Although infection rates may vary in finishing and slaughter pigs, in the breeding stock, the infection is generally high, as was verified in Alagoas (26.9%) (Valença et al., 2011) and Paraná (35%) (Tsutsui et al., 2003), probably due to the longer time of exposure to the agent (Carletti et al., 2005).

In the present study, 22.2% of the piglets (<1 month of age) had antibodies to *T. gondii*, and positive animals were found at 5 months of age. Pigs with 2, 3, and 4 months of age were all seronegative, reinforcing the presence of colostral antibodies in the first month of age. This was also observed in Spain (García-Bocanegra et al., 2010), where a higher seroprevalence was found in pigs from 1–3 weeks of age born to positive mothers. In piglets born from sows with low antibody titers, the persistence of colostral antibodies is short, ending as soon as weaning occurs (Solaymani-Mohammadi & Petri, 2006).

*T. gondii*-positive pigs were found at 5 and 6 months of age, indicating a possible horizontal infection from that age. The rate of infection is influenced by the length of stay on the property, with the prevalence of *T. gondii* antibodies higher in boars and sows due more time for horizontal transmission to occur (Vidotto et al., 1990; Dubey, 2016).

Limited reports deal with *N. caninum* in pigs, and this is the first study from Para State and one of the few worldwide that evaluated risk factors for parasite prevalence. Since *N. caninum* infection is restricted to animals, its presence in pig farms does not pose a human health concern, and the pigs are restricted to being a possible source of infection for the parasite definite host (Cerqueira-Cézar et al., 2017). This may explain the lack of more surveys of *N. caninum* in pigs worldwide, especially in Brazil.

Among pigs, little is known about *N. caninum* infection and reproductive problems. Jensen et al. (1998) experimentally infected pregnant pigs with *N. caninum* and observed disease caused by the parasite in the sows and also transplacental infection. However, Helmick et al. (2002) analyzed serum samples from 454 breeding sows with a history of abortion and infertility, all of which were negative.

The frequency of infection by *N. caninum* in both Study I (4.6%) and Study II (5.7%) was higher than that found in a confinement system in Paraiba state (3.2%) (Azevedo et al., 2010). However, they were lower than those recorded in the Bahia state (7.5%), Brazil (Almeida, 2004) and in wild pigs from Mato Grosso do Sul State (10.8%) (Soares et al., 2016), respectively, in the northeastern and central areas of Brazil. It is noteworthy that our survey involved animals of all ages, including newborn piglets less than 1 month of age. The positive results obtained from these animals were most likely due to colostrum consumption, resulting from immune system defense against an early life infection. Serological surveys in Brazil did not include newborn animals; hence, our results are difficult to compare.

Several risk factors were found to be associated with the presence of *N. caninum* antibodies, with a higher odds ratio to sewage disposal in the area, feeding the animals with slaughterhouse residues and subsistence system. Pigs reared in the subsistence system (with a familiarly workforce) were 10.5 times more likely to be *N. caninum* seropositive (95% CI: 2.5-43.0), whereas pigs with feed that included animal-origin protein, such as blood meal, were 7.9 more likely to be positive for *N. caninum* (95% CI: 1.6-39.6). This may be related to the poor sanitary conditions associated with subsistence farms, with intensive systems having greater hygiene control. Feeding pigs with animal protein was associated with *N. caninum* antibodies, which is explained by the possible presence of parasite oocysts in the animal meat, which, depending on the processing, can act as a source of infection to pigs through horizontal transmission (Dubey et al., 2017).

A positive association of antibodies to *N. caninum* in farms that discard their sludge in the soil is another factor that can be associated with poor farm hygiene present mostly in subsistence farms. The absence of nipple drinkers was associated with pig infection and can be explained by the increase in water source contamination in other types of drinkers.

To the best of our knowledge, this is the first study to determine the occurrence of *N. caninum* in piglets of different ages. The occurrence of seropositive animals with less than 1 month of age was 11.11%, with titers of 50 and 100, probably indicating colostral antibodies. Among weaned animals, seropositivity to *N. caninum* was only observed from 3 to 4 months of age, in which the titers varied between 50 and 400, remaining low (50) in the following months. This pattern is similar to that of *T. gondii* infection described by García-Bocanegra et al. (2010), in which seropositive piglets from the maternity showed a decrease in antibody titers after weaning, only developing seroconversion when entering the termination phase, where contact with contaminated food and water is greater.
Animals slaughtered from abattoir with and without state inspection did not present different seroprevalence for both *T. gondii* and *N. caninum*. Slaughterhouse without inspection may receive animals from dubious origin which could have higher prevalence of diseases due to poor hygiene conditions, which was not observed in the present study. Such data is difficult to compare since most of the studies deal only with pigs from abattoir with inspection (Carletti et al., 2005; Feitosa et al., 2014), although in Pará state previous report show 50% prevalence from animals slaughtered without inspection (Freitas et al., 2009).

*T. gondii* and *N. caninum* infection were associated with the presence of its respective definitive host in the farms (Dubey, 2009; Gui et al., 2020). We fail to detect any association with the presence of dogs and cats with the occurrence of antibodies to *T. gondii* and *N. caninum* in pigs from Northeast Pará. This may be related with the limited sample size or due to the access to the farm site by surrounding cats and dogs without the knowledge of the farmer, however this lack of association was also found in backyards pigs from Mato Grosso state, Brazil (Minetto et al., 2019).

In conclusion, reagent pigs to antibodies against *T. gondii* and *N. caninum* were found the State of Pará, Brazilian Amazon. No associations with the investigated risk factors was found for the prevalence of *T. gondii* antibodies in pigs. However, for *N. caninum*, subsistence farms, sludge discharge in the soil, absence of nipple drinkers, and feeding pigs with animal-origin protein were associated with a higher occurrence of seropositive pigs. Our analysis with piglets showed that newborn animals can acquire antibodies, probably due to colostral ingestion of the pathogen, and that seroconversion started at 5 months of age for *T. gondii* and at 4 months of age for *N. caninum*.

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**References**

Almeida M. Epidemiologia de *Neospora caninum*. Rev Bras Parasitol Vet 2004; 13(Supl. 1): 1.

André MR, Adania CH, Teixeira RHF, Silva KF, Jusí MMG, Machado STZ, et al. Antibodies to *Toxoplasma gondii* and *Neospora caninum* in Captive Neotropical and Exotic Wild Canids and Felids. *J Parasitol* 2010; 96(5): 1007-1009. http://dx.doi.org/10.1645/GE-2502.1. PMid:20950109.

Azevedo SS, Pena HF, Alves CJ, Guimarães AA Fo, Oliveira RM, Maksimov P, et al. Prevalence of anti-*Toxoplasma gondii* and anti-*Neospora caninum* antibodies in swine from Northeastern Brazil. *Rev Bras Parasitol Vet* 2010; 19(2): 80-84. http://dx.doi.org/10.1590/S1984-29612010000200002. PMid:20624342.

Carletti RT, Freire RL, Shimada MT, Ruffolo BB, Begale LP, Lopes FMR, et al. Prevalência da infecção por *Toxoplasma gondii* em suínos abatidos no Estado do Paraná, Brasil. *Semina: Ciênc Agrár* 2005; 26(4): 563. http://dx.doi.org/10.5433/1679-0359.2005v26n4p563.

Cavalcante GT, Aguiar DM, Chiebao D, Dubey JP, Ruiz VLA, Dias RA, et al. Seroprevalence of *Toxoplasma gondii* Antibodies in Cats and Pigs From Rural Western Amazon, Brazil. *J Parasitol* 2006; 92(4): 863-864. http://dx.doi.org/10.1645/GE-830R.1. PMid:16995406.

Cerqueira-Cézar CK, Calero-Bernal R, Dubey JP, Gennari SM. All about neosporosis in Brazil. *Rev Bras Parasitol Vet* 2017; 26(3): 253-279. http://dx.doi.org/10.1590/s1984-29612017045. PMid:28876360.

Chiebao DP, Valadas SYOB, Minervino AHJ, Castro V, Romaldini AHCN, Calhau AS, et al. Variables Associated with Infections of Cattle by *Brucella abortus*, *Leptospira* spp. and *Neospora* spp. in Amazon Region in Brazil. *Transbound Emerg Dis* 2015; 62(5): e30-e36. http://dx.doi.org/10.1111/tbed.12201. PMid:26302373.

Donahoe SL, Lindsay SA, Krockenberger M, Phalen D, Šlapeta J. A review of neosporosis and pathologic findings of *Neospora caninum* infection in wildlife. *Int J Parasitol Parasites Wildl* 2015; 4(2): 216-238. http://dx.doi.org/10.1016/j.ijppaw.2015.04.002. PMid:25973393.

Dubey JP, Cerqueira-Cézar CK, Murata FHA, Kwok OCH, Hill D, Yang YR, et al. All about *Toxoplasma gondii* infections in pigs: the past decade. *Vet Parasitol* 2020; 288: 109185. http://dx.doi.org/10.1016/j.vetpar.2020.109185. PMid:33271424.

Dubey JP, Hemphill A, Calero-Bernal R, Schares G. *Neosporosis in animals*. Boca Raton: CRC Press; 2017. http://dx.doi.org/10.1201/9781315152561.

Dubey JP. Toxoplasmosis in pigs: the last 20 years. *Vet Parasitol* 2009; 164(2-4): 89-103. http://dx.doi.org/10.1016/j.vetpar.2009.05.018. PMid:19559531.

Dubey JP. *Toxoplasmosis of animals and humans*. 2nd ed. Boca Raton, Florida, USA: CRC Press; 2016. http://dx.doi.org/10.1201/9781420092370.
Feitosa TF, Vilela VL, de Melo LR, de Almeida JL No, Souto DV, de Morais DF, et al. *Toxoplasma gondii* and *Neospora caninum* in slaughtered pigs from Northeast, Brazil. *Vet Parasitol* 2014; 202(3-4): 305-309. http://dx.doi.org/10.1016/j.vetpar.2014.03.015. PMid:24703253.

Foroutan M, Fakhri Y, Riahi SM, Ebrahimpour S, Namroodi S, Taghjipour A, et al. The global seroprevalence of *Toxoplasma gondii* in pigs: A systematic review and meta-analysis. *Vet Parasitol* 2019; 269: 42-52. http://dx.doi.org/10.1016/j.vetpar.2019.04.012. PMid:31079827.

Freitas JA, Oliveira JP, Ramos OS, Ishizuka MM. Frequência de anticorpos anti-*Toxoplasma gondii* em suínos abatidos sem inspeção em Belém. *Arq Bras Med Vet Zootec* 2009; 61(5): 1230-1232. http://dx.doi.org/10.1590/S0102-09352009000500030.

García-Bocanegra I, Simon-Grifé M, Dubey JP, Casal J, Martín GE, Cabezón O, et al. Seroprevalence and risk factors associated with *Toxoplasma gondii* in domestic pigs from Spain. *Parasitol Int* 2010; 59(3): 421-426. http://dx.doi.org/10.1016/j.parint.2010.06.001. PMid:20601110.

Gui BZ, Lv QY, Ge M, Li RC, Zhu XQ, Liu GH. First report of *Neospora caninum* infection in pigs in China. *Transbound Emerg Dis* 2020; 67(1): 29-32. http://dx.doi.org/10.1111/tbed.13358. PMid:31538409.

Helmick B, Otter A, McGarry J, Buxton D. Serological investigation of aborted sheep and pigs for infection by *Neospora caninum*. *Res Vet Sci* 2002; 73(2): 187-189. http://dx.doi.org/10.1053/rvsc.2002.0093-0. PMid:12204640.

Instituto Brasileiro de Geografia e Estatística – IBGE. *Produção da Pecuária Municipal* [online]. 2017 [cited 2020 Aug 1]. Available from: https://www.ibge.gov.br/estatisticas/economicas/agricultura-e-pecuaria/9107-producao-da-pecuaria-municipal.html?edicao=22651&t=resultados

Jensen L, Jensen TK, Lind P, Henriksen SA, Uggla A, Bille-Hansen V. Experimental porcine neosporosis. *APMIS* 1998; 106(4): 475-482. http://dx.doi.org/10.1111/j.1699-0463.1998.tb01374.x. PMid:9637720.

Martorano LG, Vitorino MI, Silva BPPC, Moraes JRSC, Lisboa LS, Sotta ED, et al. Climate conditions in the eastern amazon: rainfall variability in Belém and indicative of soil water deficit. *Afr J Agric Res* 2017; 12(21): 1801-1810. http://dx.doi.org/10.5897/AJAR2016.11801.

Minervino AH, Soares HS, Barrêto-Júnior RA, Neves KA, Pena HF, Ortolani EL, et al. Seroprevalence of *Toxoplasma gondii* antibodies in captive wild mammals and birds in Brazil. *J Zoo Wildl Med* 2010; 41(3): 572-574. http://dx.doi.org/10.1638/2010-0046.1. PMid:20945665.

Minervino AH, Cassinelli ABM, De Lima JTR, Soares HS, Malheiros AF, Marcili A, et al. Prevalence of anti-*Neospora caninum* and anti-*Toxoplasma gondii* antibodies in dogs from two different indigenous communities in the Brazilian Amazon Region. *J Parasitol* 2012; 98(6): 1276-1278. http://dx.doi.org/10.1645/S0022-3395-0012. PMid:22551468.

Minervino AH, Cassinelli ABM, De Souza AJS, Alves MM, Soares MCP, Ferreira DAC, et al. Detection of *Toxoplasma gondii* antibodies in captive non-human primates in the Amazon region, Brazil. *J Med Primatol* 2017; 46(6): 343-346. http://dx.doi.org/10.1111/jmp.12314. PMid:28972656.

Minervino AH, Ragozo AMA, Monteiro RM, Ortolani EL, Gennari SM. Prevalence of *Neospora caninum* antibodies in cattle from Santarém, Pará, Brazil. *Res Vet Sci* 2008; 84(2): 254-256. http://dx.doi.org/10.1016/j.rvsc.2007.05.003. PMid:17619028.

Minetto MK, Witter R, Oliveira ACS, Minetto JA, Barros ML, Aguilar DM, et al. Antibodies anti-*Toxoplasma gondii* and anti-*Neospora caninum* in backyard pigs from the state of Mato Grosso, Brazil. *Rev Bras Parasitol Vet* 2019; 28(3): 403-409. http://dx.doi.org/10.5897/RBPV.1903. PMid:31390435.

Moreira TR, Sarturi C, Stelmachchuk FN, Andersson E, Norlander E, De Oliveira FLC, et al. Prevalence of antibodies against *Toxoplasma gondii* and *Neospora* spp. in equids of Western Pará Brazil. *Acta Trop* 2019; 189: 39-45. http://dx.doi.org/10.1016/j.actatropica.2018.09.023. PMid:30267659.

Paz GS, Colhado BS, Anton MM, Rocha KS, Silva DB, Moraes CCG, et al. Infecção por *Toxoplasma gondii*, *Neospora caninum*, *Leishmania major* e *Trypanosoma cruzi* em cães do estado do Pará. *Cienc Anim Bras* 2019; 20: e33566. http://dx.doi.org/10.1590/1809-6891v20e33566.

Pezzer GB, Pezerico SB, Silva RC, Hoffmann JL, Camargo LB, Langoni H. Ocorrência de anticorpos anti-*Toxoplasma gondii* e anti-*Leptospira* spp. em suínos abatidos em três abatedouros dos estados de Minas Gerais e São Paulo. *Arq Inst Biol (Sao Paulo)* 2007; 74(3): 267-270.

Silva ICM, Bremm B, Teixeira JL, Costa NS, Barcellos JOJ, Braccini J, et al. Spatialization of Brazilian pig production: relationship between productive, physical, environmental, and socio-economic variables. *Trop Anim Health Prod* 2017; 49(5): 951-958. http://dx.doi.org/10.1007/s11250-017-1281-0. PMid:28365820.

Soares HS, Minervino AH, Barrêto-Júnior RA, Neves KAL, Oliveira MF, Santos JR, et al. Occurrence of *Toxoplasma gondii* Antibodies in *Dasyprocta aguti* from Brazil: Comparison of Diagnostic Techniques. *J Zoo Wildl Med* 2011; 42(4): 763-765. http://dx.doi.org/10.1638/2011-0136.1. PMid:22040480.
Coccidia antibodies in pigs from Pará, Brazil

Soares HS, Ramos VN, Osava CF, Oliveira S, Szabó MPJ, Piovezan U, et al. Occurrence of antibodies against *Neospora caninum* in wild pigs (*Sus scrofa*) in the Pantanal, Mato Grosso do Sul, Brazil. *Braz J Vet Res Anim Sci* 2016; 53(1): 112-116. http://dx.doi.org/10.11606/issn.1678-4456.v53i1p112-116.

Solaymani-Mohammadi S, Petri WA Jr. Zoonotic implications of the swine-transmitted protozoal infections. *Vet Parasitol* 2006; 140(3-4): 189-203. http://dx.doi.org/10.1016/j.vetpar.2006.05.012. PMid:16828229.

Sousa RA, Lemos JF, Farias LA, Lopes CD, Santos KR. Seroprevalence and risk factors for *Toxoplasma gondii* infection in pigs in southern Piauí. *Rev Bras Parasitol Vet* 2014; 23(1): 98-100. http://dx.doi.org/10.1590/1984-29612014015. PMid:24728369.

Tsutsui VS, Navarro IT, Freire RL, Freitas JC, Prudencio LB, Delbem ACB, et al. Soroepidemiologia e fatores associados à transmissão do *Toxoplasma gondii* em suínos do norte do Paraná. *Arch Vet Sci* 2003; 8(2): 27-34. http://dx.doi.org/10.5380/avs.v8i2.4030.

Valadas S, Minervino AHH, Lima VMF, Soares RM, Ortolani EL, Gennari SM. Occurrence of antibodies anti-*Neospora caninum*, anti-*Toxoplasma gondii*, and anti-*Leishmania chagasi* in serum of dogs from Pará State, Amazon, Brazil. *Parasitol Res* 2010; 107(2): 453-457. http://dx.doi.org/10.1007/s00436-010-1890-2. PMid:20445991.

Valença RMB, Mota RA, Anderlini GA, Faria EB, Cavalcanti ÉFSTF, Albuquerque PPF, et al. Prevalência e fatores de risco associados à infecção por Toxoplasma gondii em granjas suinícolas tecnificadas no Estado de Alagoas. *Pesq Vet Bras* 2011; 31(2): 121-126. http://dx.doi.org/10.1590/S0100-736X2011000200005.

Vidotto O, Navarro IT, Mitsuka R, Freire RL, Freire RL. Estudos epidemiológicos da toxoplasmose em suínos da região de Londrina – PR. *Semina: Ciênc Agrár* 1990; 11(1): 53-59. http://dx.doi.org/10.5433/1679-0359.1990v11n1p53.

Vitaliano SN, de Mendonça GM, de Sandres FA, Camargo JS, de Tarso P, Basano SA, et al. Epidemiological aspects of *Toxoplasma gondii* infection in riverside communities in the Southern Brazilian Amazon. *Rev Soc Bras Med Trop* 2015; 48(3): 301-306. http://dx.doi.org/10.1590/0037-8682-0040-2015. PMid:26108008.