Research Article

Network Pharmacological Analysis and Experimental Study of the Antipharyngitis Mechanism of the Chaiqin Qingning Capsule

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Received 14 September 2021; Accepted 17 March 2022; Published 28 April 2022

Academic Editor: Ali Nokhodchi

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Objective. The study aimed to explore the active composition and mechanism of the Chaiqin Qingning capsule (CQQN) against pharyngitis based on the network pharmacology and through using a pharyngitis rat model.

Methods. The active ingredients and targets of CQQN were queried using the TCMSP database. Disease-related target genes were queried in the DrugBank, GeneCards, OMIM, and DisGeNET databases using “pharyngitis” as the search term. The STRING database was used to establish a protein-protein interaction (PPI) network. GO function enrichment and KEGG pathway enrichment analyses were performed to identify active components and key targets. Cytoscape software (version 3.7.2) was used to construct an active component/target gene/enrichment pathway network. AutoDock software was used to select the best binding target for molecular docking. The effect of CQQN was verified in the pharyngitis rats. Results. Network pharmacology showed 30 active compounds in CQQN with 240 targets, including 54 for the treatment of pharyngitis. Potential active ingredients included quercetin, kaempferol, stigmasterol, saikosaponin D, and isorhamnetin. The key targets involved were AKT1, TNF, IL-6, and IL-1β. Signaling pathways included virus infection, TNF, IL-17, and cancer pathways. The molecular docking results showed that the critical components in CQQN had good potential for binding to key target genes. Animal experiments showed that CQQN could significantly reduce the expression of TNF-α, IL-1β, IL-6, and IL-17 in the serum of rats with pharyngitis (P < 0.05). Hematoxylin and eosin staining showed that the inflammatory state of pharyngeal tissue in rats was significantly reduced compared to that in the model group.

Conclusion. CQQN can improve pharyngitis by regulating the TNF and IL-17 signaling pathways. The study makes a positive exploration and provides a new idea for a more comprehensive and in-depth excavation of CQQN with an intervention effect on pharyngitis and other upper respiratory diseases in the future.

1. Introduction

Pharyngitis is inflammation of pharyngeal mucosa and surrounding lymphoid tissue, characterized by congestion and edema [1]. The main symptom of pharyngitis is a sore, dry, or itchy throat. Pharyngitis is usually caused by various viral or bacterial infections. The virus is the most common cause, accounting for about 40%-60% of cases. The second cause is a bacterial infection, accounting for about one-third of children’s pharyngitis, but only about 5%-10% of that in adults [2, 3]. Signs and symptoms of pharyngitis caused by bacteria, viruses, and other microorganisms usually overlap and are difficult to distinguish. This also creates difficulties in the proper use of drugs in practice. Anti-infection treatments (penicillins, erythromycins, and sulfonamides) are the first choice for acute bacterial pharyngitis. Although there is a specific therapeutic effect, long-term drug use significantly increases bacterial resistance, resulting in a poor therapeutic effect and affecting patients’ adherence [4].

Chaiqin Qingning capsule (CQQN) is a Chinese patent medicine preparation composed of Radix Bupleuri, Baicalin, and Artificial Calculus Bovis. Radix Bupleuri has antipyretic, anti-inflammatory, immune enhancement, antibacterial, and antitumor pharmacological effects. Baicalin has antipyretic, antibacterial, anti-inflammatory, antiviral, antiendotoxin, antioxidant, and antitumor effects. Artificial Calculus Bovis has the pharmacological effects of clearing away heat, detoxifying, and regulating immunity [5]. CQQN is clinically used
for the treatment of acute pharyngitis and sore throat. It has the efficacy of shortening the course of the disease, improving clinical symptoms and signs such as coughing, nasal obstruction, runny nose, and pharyngeal redness with high safety and effectiveness, and it has no apparent adverse reactions [6, 7]. In several multicenter randomized controlled trials, the clinical efficacy evaluation results of CQQN showed that it could effectively relieve fever and sore throat caused by pharyngitis [8]. However, the pharmacological mechanism of CQQN is unclear [9].

The clinical effect of traditional Chinese medicine (TCM) results from the comprehensive action between its active component group and the complex biological network of the body. Integration analysis between the active component group and the target group of the body is a challenge for studying the pharmacological mechanism of TCM [10, 11]. Through the theory and method of integrated system biology and bioinformatics, network pharmacology has carried out an overall analysis of the interaction between drugs and the body at the macro-level, bringing new research ideas to clarify the pharmacological mechanism of TCM [12, 13]. The potential active components of drugs and their possible targets can be expressed as a nested topological interaction network. By analyzing the topological properties of the network, influential molecular groups, target groups, and disease-related signaling pathways of drugs can be analyzed as a whole. Through the continuous development of predecessors, network pharmacology has been widely used to study the pharmacological mechanism of TCM and its compound, providing a rapid and effective overall analysis for pharmacological research of TCM [14, 15].

This study aimed to explore the intervention mechanism of CQQN in pharyngitis by network pharmacology and to verify the mechanism in a rat pharyngitis model. The results can provide references for follow-up research.

2. Materials and Methods

2.1. Reagents and Animals. CQQN capsules were purchased from Yangzijiang Pharmaceutical Group Co., Ltd. (China), batch number 21022312. Ammonia water was purchased from Nanjing Chemical Reagent Co., Ltd. (China), batch number 200303196X. The enzyme-linked immunosorbent assay (ELISA) kit was purchased from Jiangsu Enzyme Immunoassay Industry Co., Ltd. (China), batch number 202106.

The rats were purchased from Hangzhou Medical College, animal license number SCXK (Zhejiang) 2019-0002 and certificate number 20210628Aazz010000604. The rats were raised under controlled humidity (55-65%), temperature (22 ± 2°C), and daily light intensity (12-h/12-h light/dark cycles). The rats had free access to water and standard diets. The experimental protocols were approved by the Committee on Laboratory Animal Care of China Pharmaceutical University, and all animals were treated with humane care according to the National Institutes of Health (USA) guidelines.

2.2. Database and Analysis Platform/Software. TCMSP platform (http://lsp.nwu.edu.cn/tcmsp.php/), SwissTargetPrediction (http://www.swistargetprediction.ch/), DrugBank database (https://go.drugbank.com/), GeneCards database (https://www.genecards.org/), OMIM database (https://www.omim.org/), DisGeNet database (https://www.disgenet.org/home/), UniProt database (http://www.uniprot.org/), PubChem database (http://pubchem.ncbi.nlm.nih.gov/), STRING database (https://string-db.org/), protein structure database (http://www.rcsb.org/), Cytoscape (version3.7.2), ChemBioDraw (version 14.0), Venny plot production platform (http://bioinformatics.psb.ugent.be/webtools/Venn/), PyMOL software, AutoDockTools 1.5.6 software, and R 4.1.0 software were used.

2.3. Screening of Active Compounds and Target Genes. The components of CQQN “Radix Bupleuri,” “Baicalin,” and “Artificial Calculus Bovis” were used as search terms to screen active compounds from the TCMSP and the TCM system pharmacology databases. Oral bioavailability (OB) ≥ 30% and drug-likeness (DL) ≥ 0.18 were used as screening criteria for active compounds, and related target genes were collected [16]. The standardized targets were obtained from the UniProt database.

2.4. Screening of Targets Related to Pharyngitis. The relevant pharyngitis targets were queried in the DisGeNET, GeneCards, OMIM, and the DrugBank databases, and duplicate targets were deleted after collection. The remaining targets were used as pharyngitis disease targets.

2.5. Screening Core Targets. The potential targets of CQQN on pharyngitis were obtained by matching the target genes of the active ingredients with the target genes related to pharyngitis. The Venny map was used to draw the intersection map of the active drug component and disease targets.

2.6. Construction of the Protein-Protein Interaction Network. The protein-protein interaction (PPI) network between targets was created by selecting “Multiple proteins” in the STRING database interface, copying the intersection targets into “List of Names,” and selecting “Homo sapiens” in “Organism.” Other parameters remain as default. The obtained PPI network was imported into the Cytoscape software and visualized using the network analyzer function.

2.7. GO and KEGG Enrichment Analysis. R packages including “colorspace” and “string” were installed. A Bioconductor package (http://org.Hs.eg.db) that includes “DOSE,” “clusterProfiler,” and “pathview” was used to perform the Gene Ontology (GO) biological processes and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis on the common target of CQQN and pharyngitis. According to $P < 0.05$, the top ten biological processes (BP), molecular functions (MF), cellular components (CC), and the top 20 KEGG pathways were selected. Subsequently, experimental data such as active ingredients, targets, and the top 20 KEGG pathways were imported into the Cytoscape software to construct a network of “active ingredient/target/enrichment pathways.”
2.8. Molecular Docking Verification. The Mol2 format file of the active components of the drug was downloaded from the PubChem database, and the 3D structure of the target protein was downloaded from the PDB database. PyMOL software was used to remove water and phosphate from the protein. Then AutoDockTools and AutoDock Vina software were used for molecular docking. The binding free energy was used as the evaluation criterion for the binding degree of compounds.

2.9. Acute Pharyngitis Induced by Ammonia in Rats. Ten of the 50 healthy SD rats were randomly selected as the blank group. The rest were sprayed with 15% ammonia water on the throat with a throat sprayer, with a daily schedule of one spray in the morning and one at noon, 0.2-0.4 mL each spray for three days. The pharyngeal mucosa was swollen due to acute stimulation, thus forming an acute pharyngitis model [17].

| Herb                  | ID       | Compound                                                   | OB (%) | DL |
|-----------------------|----------|------------------------------------------------------------|--------|----|
| Radix Bupleuri        | MOL001645| Linoleyl acetate                                          | 42.1   | 0.2|
| Radix Bupleuri        | MOL002776| Baicalin                                                  | 40.12  | 0.75|
| Radix Bupleuri        | MOL000449| Stigmasterol                                              | 43.83  | 0.76|
| Radix Bupleuri        | MOL000354| Isorhamnetin                                              | 49.6   | 0.31|
| Radix Bupleuri        | MOL000422| Kaempferol                                               | 41.88  | 0.24|
| Radix Bupleuri        | MOL004598| 3,5,6,7-tetramethoxy-2-(3,4,5-trimethoxyphenyl)chromone   | 31.97  | 0.59|
| Radix Bupleuri        | MOL004609| Areapilllin                                               | 48.96  | 0.41|
| Radix Bupleuri        | MOL13187 | Cubebin                                                   | 57.13  | 0.64|
| Radix Bupleuri        | MOL004624| Longikaurin A                                            | 47.72  | 0.53|
| Radix Bupleuri        | MOL004628| Octalupine                                                | 47.62  | 0.28|
| Radix Bupleuri        | MOL004644| Sainfururan                                               | 79.91  | 0.23|
| Radix Bupleuri        | MOL004648| Troxerutin                                               | 31.6   | 0.28|
| Radix Bupleuri        | MOL004653| (+)-Anomalin                                              | 46.06  | 0.66|
| Radix Bupleuri        | MOL004702| Saikosaponin c_qt                                        | 30.5   | 0.63|
| Radix Bupleuri        | MOL004718| Go Spinasterol                                            | 42.98  | 0.76|
| Radix Bupleuri        | MOL000490| Petunidin                                                | 30.05  | 0.31|
| Radix Bupleuri        | MOL000098| Quercetin                                                | 46.43  | 0.28|
| Radix Bupleuri        | MOL004635| Saikosaponin A                                            | 32.39  | 0.09|
| Radix Bupleuri        | MOL004636| Saikosaponin B                                           | 6.70   | 0.13|
| Radix Bupleuri        | MOL004701| Saikosaponin C                                           | 5.12   | 0.05|
| Radix Bupleuri        | MOL004637| Saikosaponin D                                           | 34.39  | 0.09|
| Artificial cow-bezoar | MOL008838| Methyl (4R)-4-[(3R,5S,7S,8R,9S,10S,12S,13R,14S,17R)-3,7,12-trihydroxy-10,13-dimethyl-2,3,4,5,6,7,8,9,11,12,14,15,16,17-tetradecahydro-1H-cyclopenta[al]phenanthrene-17-yl]pentanoate | 32.32 | 0.76|
| Artificial cow-bezoar | MOL008839| Methyl desoxycholate                                     | 34.63  | 0.73|
| Artificial cow-bezoar | MOL008845| Deoxycholic acid                                          | 40.72  | 0.68|
| Artificial cow-bezoar | MOL008846| ZINC01280365                                             | 46.38  | 0.49|
| Artificial cow-bezoar | MOL000953| CLR                                                       | 37.87  | 0.68|
| Artificial cow-bezoar | MOL008841| Taurocholate                                             | 10.25  | 0.87|
| Artificial cow-bezoar | MOL008842| Chenodeoxycholic acid                                     | 27.17  | 0.69|
| Baicalin               | MOL002913| Dihydrobaicalin_qt                                        | 40.04  | 0.21|
| Baicalin               | MOL002935| Baicalin                                                  | 40.12  | 0.75|

The rats in the pharyngitis model were randomly divided into the model groups, which were the small-dose (138.83 mg/kg, equal to half of the clinical equivalent CQQN dose), the medium-dose (277.65 mg/kg, equal to the clinical equivalent CQQN dose), and the large-dose (555.30 mg/kg, equal to twice the clinical CQQN equivalent dose) CQQN groups. Each CQQN group was administered CQQN once a day for five days. The blank and model groups received an equal volume of 0.9% sodium chloride (NS) solution. After the last administration of CQQN or 0.9% NS, the pharyngeal mucosa of the animals was observed for quantitative rating evaluation. The pharyngeal scoring standard was based on the color of the pharynx tissue, secretion, degree of hyperemia, and swelling. The following four grades were assigned: grade (-): the rat’s pharyngeal tissue was light red, moist, smooth with no congestion; grade (+): the rat’s pharyngeal tissue was close to grade (-), but there were visible mild chronic congestion, poor mucosal gloss, and pharyngeal secretions; grade (++): the pharyngeal tissue of rats...
Table 2: The 54 potential antipharyngitis target genes of active ingredients.

| UniProt ID | Protein name                                           | Gene name   |
|------------|--------------------------------------------------------|-------------|
| P08254     | Stromelysin-1                                          | MMP3        |
| P27338     | Amine oxidase [flavin-containing] B                    | MAOB        |
| P05177     | Cytochrome P450 1A2                                     | CYP1A2      |
| Q9Y2R2     | Tyrosine-protein phosphatase non-receptor type 22       | PTPN22      |
| P09488     | Glutathione S-transferase Mu 1                         | GSTM1       |
| P22301     | Interleukin-10                                         | IL10        |
| Q08209     | Serine/threonine-protein phosphatase 2B catalytic subunit alpha isoform | PPP3CA   |
| P00533     | Epidermal growth factor receptor                       | EGFR        |
| P00441     | Superoxide dismutase [Cu-Zn]                           | SOD1        |
| P21397     | Amine oxidase [flavin-containing] A                     | MAOA        |
| P60568     | Interleukin-2                                          | IL2         |
| P04626     | Receptor tyrosine-protein kinase erbB-2                 | ERBB2       |
| P12830     | Cadherin-1                                              | CDH1        |
| P38484     | Interferon gamma                                        | IFNG        |
| P10415     | Apoptosis regulator Bcl-2                              | BCL2        |
| P08913     | Alpha-2A adrenergic receptor                            | ADRA2A      |
| P05362     | Intercellular adhesion molecule 1                       | ICAM1       |
| P04798     | Cytochrome P450 1A1                                     | CYP1A1      |
| Q14790     | Caspase-8                                               | CASP8       |
| Q9NWT6     | Hypoxia-inducible factor 1-alpha                        | HIF1A       |
| P10145     | Interleukin-8                                           | CXCL8       |
| P16581     | E-selectin                                              | SELE        |
| P06401     | Progesterone receptor                                   | PGR         |
| O95863     | Zinc finger protein SNAIL                               | SNAI1       |
| P04637     | ETS domain-containing protein Elk-1                    | TP53        |
| P08588     | Beta-1 adrenergic receptor                              | ADRB1       |
| P01375     | Tumor necrosis factor                                   | TNF         |
| P55211     | Caspase-9                                               | CASP9       |
| P14778     | Interleukin-1 alpha                                     | IL1A        |
| P38936     | Cyclin-dependent kinase inhibitor 1                     | CDKN1A      |
| P05164     | Myeloperoxidase                                         | MPO         |
| P31749     | RAC-alpha serine/threonine-protein kinase               | AKT1        |
| P40763     | Signal transducer and activator of transcription 3      | STAT3       |
| P13866     | Sodium/glucose cotransporter 1                          | SLC5A1      |
| P35554     | Prostaglandin G/H synthase 2                            | PTGS2       |
| P00326     | Alcohol dehydrogenase 1C                                | ADH1C       |
| P01584     | Interleukin-1 beta                                      | IL-1β       |
| P24385     | G1/S-specific cyclin-D1                                 | CCND1       |
| P47869     | Gamma-aminobutyric acid receptor subunit alpha-2        | GABRA2      |
| P13726     | Tissue factor                                           | F3          |
| P28223     | 5-hydroxytryptamine 2A receptor                         | HTR2A       |
| P07550     | Beta-2 adrenergic receptor                              | ADRB2       |
| P15692     | Vascular endothelial growth factor A                    | VEGFA       |
| P01106     | Myc proto-oncogene protein                              | MYC         |
| P20248     | Cyclin-A2                                               | CCNA2       |
| P22303     | Acetylcholinesterase                                    | AChE        |
| P29965     | CD40 ligand                                             | CD40LG      |
| P42224     | Signal transducer and activator of transcription 1-alpha/beta | STAT1     |
showed chronic congestion, dark red, and mild swelling with a small amount of secretion; and grade (+++): the pharyngeal tissue was in a state of chronic hyperemia, dark red, increased mucus secretion, and noticeable swelling [18].

2.10. The Expression of Inflammatory Cytokines in Serum Determined by ELISA. Upon the completion of CQQN or 0.9% NS administration, the serum of rats was taken, and the blood samples were placed at 4°C for 30 min. Serum was separated by 3000 rpm centrifugation for 10 min. Serum levels of IL-1β, IL-6, TNF-α, and IL-17 were measured by ELISA according to the kit instructions.

2.11. Observation of Pharyngeal Tissue Lesions. Pharyngeal mucosa and submucosa were extracted and fixed with 10% formalin solution. After hematoxylin and eosin (HE) staining, the histomorphological changes were observed under a light microscope.

2.12. Statistical Analysis. Excel was used for data entry, SPSS 25.0 software for data analysis, and GraphPad Prism 9.0 was used for mapping. Measurement data were expressed as mean ± standard deviation (mean ± SD). One-way analysis of variance was used for pairwise comparison between groups. P < 0.05 was considered statistically significant, and P < 0.01 indicated that the difference was extremely significant.

3. Results

3.1. Screening of Active Drug Ingredients and Target Genes. A total of 374 ingredients were obtained from the TCMSP database (353 in Radix Bupleuri, 19 in Artificial Calculus Bovis, and 2 in Baicalin). The active compounds were then screened with DL ≥ 0.18 and OB ≥ 30% as the condition. Saikosaponins A, B, C, and D were reported to be the main active components of Radix Bupleuri [19]. Finally, 30 active components of CQQN were obtained, including 21 in Radix Bupleuri, 2 in Baicalin, and 7 in Artificial Calculus Bovis (Table 1). Active components were searched, screened, and standardized by the UniProt database and SwissTargetPrediction. There were 375 Radix Bupleuri, 3 Baicalin, and 46 Artificial Calculus Bovis (Supplementary Table 1). A total of 226 drug targets were obtained after removing duplicate targets.

3.2. Acquisition and Screening of Targets for Pharyngitis. With the keywords “pharyngitis,” 46 pharyngitis-related targets were found in the DrugBank database, 536 in the GeneCards database, 38 in the OMIM database, and 30 in the DisGent database. After removing duplicate targets, 618 pharyngitis targets were obtained (Supplementary Table 2). The 54 common targets of the CQQN component and pharyngitis were considered potential key targets (Table 2). Targets were imported into the Venny mapping software, and the Wayne diagram was drawn, as shown in Figure 1.

3.3. GO and KEGG Analysis. The Bioconductor package in R software was used to perform the GO enrichment and KEGG pathway analyses on common targets of CQQN and pharyngitis. A total of 1,590 GO items were obtained (P < 0.05), including 1,506 BP items, 14 CC items, and 70 MF items. BP is mainly related to the response to reactive oxygen species (ROS) in the metabolic process, the response to drugs, the regulation of ROS, and the T cell activation. CC is related to the membrane raft, the side of the membrane, and the outer membrane. MF is primarily related to binding cytokines, protein phosphatase, kinase, and tumor necrosis factor (TNF) receptor superfamily (Supplementary Table 3). According to the enrichment results, the top ten BP, CC, and MF items were selected for the bubble graph, as shown in Figure 2.

The KEGG enrichment analysis resulted in 139 pathways (P < 0.05). KEGG analysis showed that the relevant CQQN signaling pathways in the treatment of pharyngitis included virus infection, TNF, IL-17, and cancer pathways (Supplementary Table 4). The top 20 pathways with the highest number of genes for the treatment of pharyngitis were selected for the bubble chart, as shown in Figure 3. The significantly enriched genes were AKT1, IL-6, TNF, STAT3, TP53, PTGS2, EGFR, CCND1, CASP3, BCL2, STAT1, MMP9, and ICAM1.

| UniProt ID | Protein name                              | Gene name  |
|-----------|-------------------------------------------|------------|
| Q9NS23    | Ras association domain-containing protein 1 | RASSF1     |
| P08684    | Cytochrome P450 3A4                        | CYP3A4     |
| P05231    | Interleukin-6                              | IL-6       |
| Q15722    | Leukotriene B4 receptor 1                 | LTB4R      |
| P42574    | Caspase-3                                  | CASP3      |
| P14780    | Matrix metalloproteinase-9                | MMP9       |

Table 2: Continued.
3.4. Construction of Active Ingredient/Target/Enrichment Pathway Network. The active ingredients of CQQN, the potential targets for treating pharyngitis, and the top 20 KEGG-enriched pathways \( (P \leq 0.05) \) were imported into the Cytoscape software to construct a network diagram of active components/target/enrichment pathways. There were 93 nodes (22 compounds, 51 targets, and 20 pathways) and 347 interacting edges in the network, as shown in Figure 4. Each compound corresponds to multiple targets in the network diagram, indicating that multiple targets may synergistically treat pharyngitis with CQQN. The topological analysis of the results was carried out using Cytoscape software, as shown in Figure 4. The larger the circle label, the higher the degree, the more associated targets, and the greater the importance of the research. A total of 54 nodes and 566 interaction lines were obtained (Figure 5(a)). After calculation, the average degree of the freedom of nodes in PPI was 21. The top 15 targets in the PPI network were selected, which were AKT1, TNF, IL-6, TP53, EGFR, IL-1β, CASP3, PTGS2, VEGFA, CXCL8 STAT3, MMP9, HIF1A, IL10, and MYC (Figure 5(b)). These targets are suggested to be the potential key targets of CQQN in treating pharyngitis.

3.5. PPI Network Construction. The intersection target of components and diseases was imported into the STRING database for analysis to obtain the PPI network. The species were restricted to “Homo sapiens” and filtered with confidence ≥0.4. The results were imported into the Cytoscape software for visual analysis. Among the top ten components with a higher degree in the network diagram, there were four flavonoids (quercetin, kaempferol, isorhamnetin, and 3,5,6,7-tetramethoxy-2-(3,4,5-trimethoxyphenyl) chromone), three saponins (saikosaponin D, saikosaponin A, and saikosaponin B), one sterol component (stigmasterol), one lignan component (cubebin), and a steroid compound (chenodeoxycholic acid). The results show that these compounds may be the key components of CQQN in the treatment of pharyngitis, as shown in Table 3.

3.6. Molecular Docking Analysis. The molecular docking technology was used to predict the binding ability of the active ingredients of CQQN with potential targets. The top ten active ingredients with higher degrees obtained by the topological analysis were used as ligands, and the top 15 key targets with higher degrees selected from the results of the PPI network and the KEGG analysis were receptors for the docking calculations.
the prediction of binding ability. The docking result retained only the highest absolute value of predicted binding affinity (kcal/mol) for each pair of molecular docking. It is generally believed that binding energy of less than -4.25 kcal/mol indicates that the ligand has a specific binding activity to the receptor, less than -5.0 kcal/mol has better binding activity, and less than -7.0 kcal/mol has strong binding activity.

The ten active pharmaceutical ingredients were bound to 14 target proteins (TP50 was deleted because the ligand structure could not be found), and the data obtained were analyzed by a heat map, as shown in Figure 6. Among them, the ingredients with a docking score $\leq -9.0$ kcal/mol accounted for 12.1% (17/140); the ingredients with a docking score between -9.0 and -7.0 kcal/mol accounted for 51.4% (72/140), and the ingredients with a docking score $\geq -7.0$ kcal/mol accounted for 36.4% (51/140). The results show that the active ingredients of CQQN have strong binding ability with key targets. Five typical components (saikosaponin D, isorhamnetin, chenodeoxycholic acid, saikosaponin A, and saikosaponin B) and four key targets (TNF, IL-6, IL-1$\beta$, and AKT1) were selected for the analysis of the binding mode. As shown in Figure 7, the binding mode is mainly hydrogen bonding, and the hydrophobic interaction further stabilizes the compound-protein structure.

3.7. Effect of CQQN on the Rat Model of Acute Pharyngitis

3.7.1. Observation of the General State of an Acute Pharyngitis Rat Model. All rats in the blank group were in good physical condition, had bright fur color, and had normal eating and drinking patterns. After modeling, all rats had swelling and congestion of the pharynx, and oral secretions gradually increased. Some rats had local ulceration of the mouth and lips, and superficial ulcers were formed in the pharynx. After continuous administration of CQQN for five days, the redness and swelling of the small-, medium-, and large-dose groups of rats in the CQQN were reduced to varying degrees. The amount of secretion was significantly reduced compared to that of the model group (Table 4).

3.7.2. Effect on Pathological Changes in Pharynx Tissue in Rats with Acute Pharyngitis. In the structure of the blank group, the rat pharynx tissue and the submucosal glands were normal, and there were no inflammatory cells. In the structure of the model group, the rat pharynx tissue changed with the congested mucosal layer of blood vessels. The submucosal layer of connective tissue proliferated, and many inflammatory cells infiltrated. The pharyngeal mucosa of rats in the high-dose CQQN group was basically restored.
to normal, and the submucosal inflammatory cells and mucosal glands were significantly reduced. In the medium-dose CQQN group, a small number of inflammatory cells were observed in the mucosa and submucosa in the pharyngeal tissue. In the small-dose CQQN group, the pharyngeal mucosa of rats became thinner. Inflammatory cells were significantly reduced, and the submucosal mucosal glands were basically normal, as shown in Figure 8.

The histopathological grade of the pharynx in the model group was significantly higher than that of the blank group ($P < 0.01$). Compared to the model group, the pharyngeal tissues of the CQQN small-, medium-, and large-dose groups improved significantly ($P < 0.01$), as shown in Table 5.

### 3.7.3. Serum Levels of IL-6, IL-1β, TNF-α, and IL-17 in Rats of Each Group

Compared to the blank group, the levels of TNF-α, IL-1β, IL-6, and IL-17 in the model group increased significantly ($P < 0.01$). Compared to the model group, the CQQN large-dose group significantly reduced the levels of

| Ingredient   | Chemical name                                           | Degree |
|--------------|---------------------------------------------------------|--------|
| Flavonoid    | Quercetin                                               | 38     |
| Flavonoid    | Kaempferol                                              | 16     |
| Sterol       | Stigmastanol                                            | 9      |
| Saponin      | Saikosaponin D                                          | 4      |
| Flavonoid    | Isorhamnetin                                            | 4      |
| Steroid      | Chenodeoxycholic acid                                   | 3      |
| Saponin      | Saikosaponin A                                          | 3      |
| Saponin      | Saikosaponin B                                          | 2      |
| Lignans      | Cebubin                                                 | 2      |
| Flavonoid    | 3,5,6,7-tetramethoxy-2-(3,4,5-trimethoxyphenyl)chromone | 2      |
IL-6, TNF-α, and IL-1β (P < 0.01) and significantly reduced the level of IL-17 (P < 0.05). The medium-dose CQQN group significantly reduced the levels of IL-1β and IL-6 (P < 0.01) and significantly reduced the TNF-α level (P < 0.05), as shown in Table 6 and Figure 9.

4. Discussion

Pharyngitis is an inflammation of the upper respiratory tract. Sore throat and dysphagia in the disease process will seriously affect patients’ daily lives and even cause serious
complications, such as rheumatic heart disease, glomerulonephritis after streptococcal infection, sepsis, or autoimmune diseases [22]. CQQN is a TCM preparation containing complex ingredients and is often used clinically to treat upper respiratory tract diseases such as sore throat and pharyngitis.

Network pharmacology results show that quercetin, kaempferol, stigmasterol, saikosaponin A, saikosaponin C, saikosaponin D, isorhamnetin, and other ingredients play a vital role in the treatment of pharyngitis by CQQN. Quercetin is a flavonoid compound. It is considered the most effective ROS scavenger and inhibits the production of several proinflammatory factors, such as TNF-α and NO [23]. In vitro treatment of activated T cells with quercetin blocks IL-12-induced tyrosine phosphorylation of JAK2, TYK2, STAT3, and STAT4, resulting in a decrease in IL-12-induced T cell proliferation and Th1 differentiation [24]. Several in vitro studies using different cell lines have shown

### Table 4: Rat pharyngeal tissue scores in each group.

| Group       | Dose (mg/kg) | N  | — | + | ++ | +++ |
|-------------|--------------|----|----|---|----|-----|
| Blank group | —            | 10 | 10 | — | —  | —   |
| Model group | 138.83       | 10 | 10 | — | 2  | 8   |
| CQQN group  | 277.65       | 10 | 3  | 5 | 1  | 3   |
| CQQN group  | 555.30       | 10 | 8  | 2 | —  | —   |

Figure 7: Molecular docking patterns of the active ingredients of the target CQQN protein in the treatment of pharyngitis.
that quercetin inhibits lipopolysaccharide (LPS)-induced TNF-α production in macrophages [25, 26] and LPS-induced IL-8 production in lung A549 cells [27]. Kaempferol has anti-inflammatory, antioxidant, and immunomodulatory pharmacological properties. Kaempferol can significantly improve lung ischemia-reperfusion injury, inhibit the release of inflammatory factors including IL-6 and TNF-α in bronchoalveolar lavage fluid, and reduce the reaction of oxidative stress [28]. Kaempferol can regulate the phosphorylation of IκB-α and p65 and inhibit inflammation in vitro and in vivo by regulating the MAPK and NF-κB pathways. NF-κB, which is involved in the immune response and the inflammatory response, is a familiar molecular target for the treatment of pharyngitis [29]. Saikosaponin is a triterpene saponin that has a variety of pharmacological activities, including anti-inflammatory and antioxidant effects. Saikosaponin can reduce inflammation and regulate autophagy by inhibiting the PI3k/Akt/mTOR signaling pathway [30]. Stigmasterol has antioxidant, anti-inflammatory, antitumor, and other pharmacological activities and

![Figure 8: Results of histopathological HE staining results of pharynx of rats in each group (HE, ×200).](image)

### Table 5: Histopathological grading of pharyngeal tissue of rats in each group.

| Group      | Dose (mg/kg) | n  | - | + | ++ | +++ |
|------------|--------------|----|---|---|----|-----|
| Blank group| —            | 10 | 10| 0 | 0  | 0   |
| Model group| —            | 10 | 0 | 0 | 0  | 10  |
|            | 138.83       | 10 | 1 | 6 | 2  | 1   |
| CQQN group | 277.65       | 10 | 5 | 1 | 0  | 0   |
|            | 555.30       | 10 | 8 | 2 | 0  | 0   |

Note: *P < 0.05 and **P < 0.01 vs. model group. "-" means that the mucosa of the pharynx and submucosa is normal. "+" indicates that the scaly epithelium of the pharynx has little hyperplasia and a small number of inflammatory cells infiltrated under the mucosa. "++" means scaly epithelium hyperplasia of the pharynx mucosa, and inflammatory cell infiltration is seen in the lower layer. "+++" means the presence of the pharynx mucosa scaly epithelium hyperplasia with a large number of inflammatory cells infiltrated in the mucosa [21].

| Group      | Dose (mg/kg) | IL-6 | IL-1β | TNF-α | IL-17 |
|------------|--------------|------|-------|-------|-------|
| Blank group| —            | 54.49 ± 9.50** | 13.24 ± 3.84** | 116.67 ± 31.72** | 18.01 ± 7.41** |
| Model group| —            | 116.23 ± 5.58  | 29.89 ± 3.94   | 213.99 ± 29.40  | 33.27 ± 12.26 |
|            | 138.83       | 108.25 ± 9.61  | 26.79 ± 2.72   | 200.02 ± 41.13  | 26.71 ± 9.44  |
| CQQN group | 277.65       | 96.91 ± 7.23** | 17.83 ± 2.32** | 170.34 ± 30.73* | 23.90 ± 6.53  |
|            | 555.30       | 78.39 ± 7.68** | 14.37 ± 2.79** | 127.13 ± 32.95* | 21.66 ± 6.78* |

Note: *P < 0.05 and **P < 0.01 vs. model group.
can significantly suppress the expression of proinflammatory mediators (TNF-α, IL-6, IL-1β, iNOS, and COX-2) and increase the expression of anti-inflammatory cytokines (IL-10) through downregulating the expression of NF-kBp65 (inhibiting p-IKB-α activation) and p38MAPK [31]. Isorhamnetin has various effects such as protecting cardiovascular and cerebrovascular, antitumor, anti-inflammatory, and antiviral activities. Treatment with isorhamnetin inhibits the phosphorylation of the mitogen-activated protein kinase (MAPK) and NF-κB pathways induced by TNF-α [32]. The above shows that the key active ingredients of CQQN all have good anti-inflammatory, antioxidant, or immunomodulatory effects.

The GO analysis result shows that the cytokine receptor binding and TNF receptor binding are at the top. It indicates that these receptors may be the primary drug targets of CQQN in the treatment of pharyngitis. TNF is a multifunctional cytokine. It plays an essential role in the pathophysiology of several diseases [33]. TNF-α is an inflammatory cytokine produced by macrophages/monocytes during acute inflammation and is responsible for a wide range of signaling events within cells, leading to necrosis or apoptosis [34]. The results of the KEGG pathway enrichment analysis show that the targets of CQQN are mainly concentrated in virus infection and TNF, IL-17, and cancer pathways. Studies have found that the TNF signaling pathway plays a key role in the inflammatory response. TNF-α is the most important cytokine in this pathway by upregulating MAPK, ERK, NF-κB, and other signaling pathways to induce apoptosis and plays a proinflammatory role [35]. MAPK is related to cell proliferation and immune regulation. It can manipulate key host cell signals to induce IL-1β, IL-6, TNF-α, and other inflammatory mediators [36]. IL-17 is the signature cytokine of the Th17 cell and is secreted as a homodimer or as a heterodimer with IL-17F [37]. IL-17 is a crucial mediator of mucosal surveillance and barrier integrity. The most prominent function of IL-17 is to provide a protective inflammatory response against pathogens at boundary tissues, such as the skin, intestine, and lungs. In pharyngitis or tonsillitis caused by streptococcus, peripheral T cells can be induced to differentiate into Th17 cells. IL-17 secreted by Th17 cells has a proinflammatory effect [38]. In addition, IL-17 is involved in epithelial cell and neutrophil-mediated immune responses against extracellular microbes and autoimmune diseases’ pathogenesis [39, 40]. Consequently, CQQN is likely to treat pharyngitis by participating in the IL-17 signal pathway.

The PPI topological analysis shows 16 strongly associated proteins, among which AKT1, TNF, IL-6, and IL-1β are the predicted targets of CQQN. The molecular docking results showed that the active components of CQQN had a strong affinity with the key targets, a stable hydrogen bond at the binding site, and a stable conformation and good binding activity. It can provide a theoretical basis for CQQN treating pharyngitis at the molecular level.

Through the animal experiment, we found that the pharyngeal mucosa of pharyngitis rats in each dose group improved to various degrees after CQQN was administered.
for five days. Under a light microscope, it was found that the pathological changes of the pharyngeal tissue in CQQN were less than that in the model group. Inflammatory cells in the submucosa were significantly reduced, and mucosal glands were also decreased.

IL-6, IL-1β, TNF-α, and IL-17 in the medium and large-dose CQQN groups were significantly lower than those in the model group. It is suggested that CQQN inhibits the development of inflammation by reducing the production of proinflammatory factors and mediators and blocking the transmission of proinflammatory signals and playing a role in the treatment of pharyngitis.

Saikosaponin A has been reported to reduce serum IL-6, TNF-α, and IL1β after anorectal surgery in rats [41], and quercetin can reduce levels of TNF-α and IL-6 in mouse nasal mucosa tissue [42]. Stigmasterol has been reported to be related to reducing critical cytokines in the pain and inflammation (TNF-α, IL-1β, and IL-6) [43]. This shows that the active ingredients in CQQN have potential therapeutic anti-inflammatory and immunomodulatory effects.

This study screened potential active ingredients, key targets, and CQQN pathways of CQQN in the treatment of pharyngitis through network pharmacology. The results show that CQQN can intervene in the expression of inflammatory factors through multiple components, targets, and pathways. The results of animal experiments indicate that CQQN exerts a therapeutic effect by downregulating proinflammatory cytokines (TNF-α, IL-6, IL-1β, and IL-17) to inhibit the TNF and IL-17 signaling pathways. However, this study only conducted a single efficacy test through the pharyngitis rat model; more studies of CQQN need to be conducted. This study provides ideas and a basis for a more comprehensive and in-depth exploration of TCM with intervention effects on pharyngitis in the future.

Data Availability
The data used to support the findings of this study are available in the supplementary information of the manuscript.

Conflicts of Interest
The authors declare that there is no conflict of interest with respect to the publication of this article.

Authors’ Contributions
Design of the study was contributed by Xuansheng Ding and Can Wang. Data collection was performed by Can Wang, Hongjin Gao, Lianzhan Huang, and Zhen Wang. Data analysis and interpretation was performed by Can Wang and Hongjin Gao. Writing and revision of the article was contributed by Xuansheng Ding, Can Wang, and Lianzhan Huang.

Acknowledgments
The authors thank China Pharmaceutical University for providing the experimental platform.

Supplementary Materials

Supplementary Table 1: information of predicted CQQN targets. Supplementary Table 2: information of known pharyngitis-related targets. Supplementary Table 3: top 10 GO enrichment of biological processes (BP), cellular components (CC), and molecular functions (MF) with their representative enriched terms (one per cluster). Supplementary Table 4: top 20 KEGG pathway. (Supplementary Materials)

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