A multi-parameter evaluation of the neuroprotective and cognitive-enhancing effects of *Origanum onites* L. (Turkish Oregano) essential oil on scopolamine-induced amnestic rats

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Abstract
Alzheimer’s disease (AD) is a neurodegenerative disorder characterized by progressive deterioration of cognitive functions (dementia) and represents a growing public health concern since the population in the age groups at risk is increasing. The latter raises an urgent need to translate research findings in the basic brain and behavioral sciences into anti-AD drugs and disease-modifying therapies. *Origanum onites* (L.), also called Turkish oregano, is a perennial and herbaceous plant species grown for centuries for medicinal, cosmetic and culinary purposes. This is the first study to investigate the putative neuroprotective and pro-cognitive activities of *O. onites* essential oil (OEOO) against scopolamine-induced amnesia of AD-type in Wistar albino rats. The results of behavioral tests revealed that OEOO administration was able to significantly alleviate learning and memory impairments induced by scopolamine in vivo. The observed effects could be attributed to inhibition of acetylcholinesterase activity, attenuation of oxidative stress and prevention of neuronal apoptosis in the hippocampus and frontal cortex of AD rats. Modulation of pro-inflammatory enzymes, including cyclooxygenase-2, inducible nitric oxide synthase and myeloperoxidase, might further contribute to the neuroprotective properties of OEOO, as predicted by our in silico models. These findings offer novel insights into the therapeutic potential of OEOO in patients with AD.

Keywords Scopolamine-induced amnesia · Alzheimer’s disease · Oxidative stress · Neuroinflammation · Apoptosis · *Origanum onites*

Introduction
Alzheimer’s disease (AD) is an aging-related neurodegenerative disorder with progression, which is manifest in both cognitive and neuropsychiatric symptoms (Soria Lopez et al. 2019). The progression of the AD continuum causes deficits in cognitive functions to become more perceptible. It is commonly believed that a loss of cholinergic neurons from the forebrain and a corresponding decrease in acetylcholine levels, both of which represent hallmarks of AD pathology, predispose to dementia (Ferreira-Vieira et al. 2016). Changes in cholinergic transmission, oxidative stress, inflammation or monoaminergic disorders can be counted among the different causes of memory loss in dementia. With these effects, neuronal apoptosis and consequent memory disorders develop (Deng et al. 2019).

Today, the administration of acetylcholinesterase (AChE) inhibitors appears to provide the most promising results in the symptomatic treatment of mild to moderately severe
AD. While the patient’s learning and memory functions are improved by increasing the acetylcholine levels by means of AChE inhibitors, they also possess limitations due to the side effects caused by the activation of the cholinergic system (Deng et al. 2019). Furthermore, there is compelling evidence showing that chronic inflammation (neuroinflammation) significantly contributes to the pathogenesis of AD (Kinney et al. 2018). A number of pro-inflammatory mediators, including inducible enzyme systems, are known to be associated with this sustained immune response in the brain. Therefore, it is no surprise that anti-inflammatory interventions may show efficacy in treating AD. However, efforts to prevent dementia by using aspirin and other non-steroidal anti-inflammatory drugs (NSAIDs) have generally been unsuccessful to date (Jordan et al. 2020).

The science of natural medicine, which has been around for centuries, greatly supports the need for both different drug applications and clinical drugs. Precious compounds in natural products serve to form the basis of modern drug research as well as to guide research which leads to the best possible cure (Howes et al. 2003). Due to the limitations of current therapy, treatment regimens that can benefit from AD modifying and/or preventive agents in daily tasks that require cognitive functions have come into focus (Farlow et al. 2008). In this context, medicinal plants known or researched to have anti-inflammatory, antioxidant, neuroprotective effects have become the focus (Perry et al. 2003; Rabiei et al. 2015; Xu et al. 2017; Capatina et al. 2020).

Scopolamine (Sco) is an alkaloid drug that produces antagonism at muscarinic acetylcholine receptors (mAChRs) by interfering with cholinergic transmission, impairing learning and short-term memory in both rodents and humans (Zhou et al. 2018). Therefore, Sco has been used in many preclinical studies to screen drugs with potential therapeutic values in dementia patients with AD to produce an amnesia model (Rezvani et al. 2011; Zhou et al. 2018). The literature provides important evidence stating that Sco has numerous effects in the brain, such as oxidative stress, which eventually cause cognitive impairment (Rahnama et al. 2015). It has been reported that natural products or plant components, some of which are currently in use as potent AChE inhibitors, can reverse memory deficits in studies using various animal models, including Sco-induced models (Savelev et al. 2004; Rabiei et al. 2015).

*Origanum onites* L. (also known as Turkish oregano) is used as a raw drug for the treatment of respiratory tract diseases, hypertension, and high cholesterol, as well as an analgesic, sedative, and general oral antiseptic (Tepe et al. 2016). Thymol and carvacrol, which are the two major components in the essential oils of many aromatic plants as in *Origanum onites* essential oil (OOEO), have antioxidant, anti-inflammatory and AChE-inhibitory properties that could be effective in treating the cognitive symptoms of AD (Baser 2008; Azizi et al. 2012). The current study, which is the first of its kind, aims at evaluating the influence of OOEO on the acquisition and consolidation of memory and learning processes impaired by Sco treatment in Wistar albino rats using behavioral tests. It also attempts to investigate the underlying pathophysiological mechanisms of cognitive dysfunction through assaying the activity of AChE and determining the levels of the oxidative stress markers malondialdehyde (MDA) and reduced glutathione (GSH) as well as of the mitochondria-mediated apoptosis markers bcl-2, bax, caspase (casp)-3 and casp-9 in the frontal cortex and hippocampus of the animals. Finally, in order to estimate the anti-inflammatory activity of OOEO, the major component of the essential oil is docked in a computational setting onto three selected pro-inflammatory enzymes, namely cyclooxygenase-2 (COX-2), inducible nitric oxide synthase (iNOS), and myeloperoxidase (MPO).

**Materials and methods**

**OOEO and chemicals**

OOEO was procured from TÜRER, Inc. (İzmir, Turkey). The identity of the source plant was confirmed by comparison with a sample stored in the Herbarium of Anadolu University Faculty of Pharmacy Başer Library of Essential Oil Constituents (ESSE 14,567, Eskişehir, Turkey). Unless stated otherwise, all chemicals and all antibodies were purchased from Sigma-Aldrich Co. (St. Louis, Missouri, USA) and Santa Cruz Biotechnology, Inc. (Santa Cruz, CA, USA), respectively.

**Analysis of OOEO and identification of its components**

OOEO was analyzed by gas chromatography/mass spectrometry (GC/MS) using the Agilent 5977B GC/MSD system, and the relative percentages of successfully separated compounds were computed from the corresponding flame ionization detector peak areas according to previously described methods (Demirci et al. 2004; Aykac et al. 2020). Retention indices were determined using n-alkane standards as reference, while retention indices were compared with those of authentic samples. Primarily, Baser Library of Essential Oils Constituents was used in the characterization of components.

**Animals**

Female rats (Wistar albino), weighing 200–250 g, were obtained from Marmara University Experimental Animals Research and Implementation Center (İstanbul, Turkey). All experiments were approved by Research and Local Committee on Animal Research Ethics of Marmara University (approval no. 81.2019.mar) and executed in accordance with
the “Guide for the Care and Use at Laboratory Animals. All rats were housed in cages (n = 3 rats per cage) in a controlled 12-h light/dark cycle at a humidity of 50 ± 5% and a temperature of 22 ± 2 °C. Rats were allowed to access water and food ad libitum. The rats housed in the above-mentioned habituation conditions one week before the behavioral experiments started were randomly divided into five groups (n = 6 rats in each group).

**Modeling amnesia and construction of experimental groups**

Groups of rats received i.p. injections of scopolamine either alone (group Sco, 1 mg/kg) (Ghasemi et al. 2019; Aykac et al. 2019) or in combination with galantamine (Gal, 1.5 mg/kg, i.p.) (Geerts et al. 2005) or with OOEO (Sco + OOEO, 0.25 ml/kg, p.o.) (Amiresmaeili et al. 2018). The control group received either saline (0.9% w/v NaCl, i.p.) or OOEO (0.25 ml/kg, p.o.) instead of Sco. All treatments were given at 9 a.m. each day, starting from the end of the isolation period until rats were sacrificed (Fig. 1). Locomotor activity, MWM and novel object recognition (NOR) tests were performed to evaluate cognition, learning, recognition and spatial memory function in rats.

**Locomotor activity test**

For each rat, the number of total transitions between the squares and rearing frequencies were recorded for 5 min with the help of a video camera hanging above the apparatus consisting of a total of 25 squares, with the side length of each square being 15 cm (Rogóz et al. 2012).

**NOR test**

The NOR test was carried out for each rat in three stages as follows: habituation, familiarization, and test. The rat was placed in a box (65 cm × 45 cm × 65 cm) for 10 min in the habituation phase. The next day, the behavior of the rat placed in the box with two identical objects of the same shape and color was recorded for 10 min in the familiarization phase. After the familiarization phase, the rat was allowed to stay in its cage for 1 h. The behavior of the rat in the box containing one of the sample objects used in the familiarization phase (old object) and a novel (new) object is observed for 3 min and record using a video camera (Antunes and Biala 2012). After each animal completed the NOR test, all used boxes and objects were cleaned with a solution containing 70% (v/v) ethanol and the next rat’s behavior experiments were started. The results obtained...
from the NOR test are expressed as the index of discrimination with results ranging from −1 to +1 (Aubele et al. 2008).

Discrimination index ($s$) = (Time spent with the novel object – Time spent with the sample object) / (Time spent with the novel object + Time spent with the sample object).

**MWM test**

The maze was an open circular pool (diameter of 1.2 m, a height of 0.47 m and depth of 30 cm) approximately half-filled with water made opacified with milk powder (water temperature is 24–26°C). MWM task consists of the training period and probe trials. While there was a platform in the MWM tests from day 1 to day 4 (training period), there was no platform in the tank in day 5 experiments (probe trial). The pool was divided into four equal quadrants (quarter-circles) and a platform was hidden 2 cm below the surface in the northeast (NE) direction on the first day of the experiments (Vorhees and Williams 2006). In the first 4 days, each rat received three training sessions per day. In these 4-day training experiments, rats were randomly released from one of the four quadrants (NE, SE, SW, or NW direction) into the maze and allowed to swim for 75 s to find the platform. Platform finding times of rats in 60 s are recorded. The animals that failed to find the platform within this period are directed to the platform allowed to stay there for 20 s and their platform finding times are saved as 60 s. The swimming activity of each rat, which will be used to evaluate the escape delay and escape rate, was recorded using a fixed video camera hung from the ceiling. On the day of the probe test, in the NE direction, the platform was lifted and the rat was allowed to swim for 60 s by releasing it into the tank from SW (the opposite direction of NE). The time spent in the target quadrant was recorded for each rat.

**Euthanasia and removal of brain tissue samples**

The rats were deep anesthetized with thioventral sodium at 50 mg/kg (Actavis; Archimedes Pharma, Reading, UK) immediately after the last cognitive test session, and their brains were dissected into different anatomical regions. The hippocampus (anteroposterior plane: between 6.70–4.70 mm anterior to the inter-aural line) and frontal cortex (anteroposterior plane: between 13.20–11.20 mm anterior to the inter-aural line) were sliced according to the coordinates stated in Paxinos and Watson’s Rat Brain Atlas (2007).

**Determination of AChE activity and measurement of MDA and GSH levels in different brain tissues**

The activity of AChE, which is responsible for the degradation of acetylcholine in tissue samples, was determined using an acetylcholinesterase colorimetric commercial kit by Abcam (ab138871) and was expressed in U/mg protein. The assay was performed according to the manufacturer’s instructions, and the activity was measured at 410 nm using a spectrophotometer. In cortex and hippocampal tissue samples, MDA levels (expressed as nmol/mg protein) were measured using a commercial test kit (ab118970, Abcam), while GSH levels (expressed as µg/mg protein) were measured according to the method described previously (Wang et al. 2019). All readings were made on a universal microplate spectrophotometer (MDA at 532 nm, GSH at 420 nm).

**Immunoblotting analysis**

The immunoblotting analysis, which is used to determine the total amount of protein present in the tissues, was briefly performed as follows: the protein content of the prefixes was determined by the Lowry method (Lowry et al. 1951) after centrifuging the tissues in Tris–HCl (pH 7.2) buffer. Samples containing 100 µg protein were loaded onto a 12% sodium dodecyl sulfate–polyacrylamide gel (SDS–PAGE) and then transferred onto the membranes (Schleicher and Schuell, 0.45 µm, Germany). All membranes were incubated with polyclonal primary antibodies for 12 h. All antibodies were used at a dilution of 1:100, except bax (1:200). The β-actin expression levels were used as a standard for all membranes. Protein bands were visualized by NBT/BCIP on membranes incubated with secondary antibody for 1 h. Densitometric analysis of all membranes was performed using Bio-Rad Molecular Analyst software (www.totallab.com). Molecular weights for β-actin, bax, bcl-2, casp-3 and casp-9 are 43, 26, 20 and 35 kDa respectively.

**Protein–ligand docking**

The 3D conformer of carvacrol (PubChem entry: 10,364) in SDF format was retrieved from the public chemical database PubChem (Kim et al. 2019) and separately docked onto the crystal structures of three selected human pro-inflammatory enzymes — namely, COX-2 (PDB entry: 5KIR) (Orlando and Malkowski 2016), inducible iNOS; (PDB entry: 3E7G) (Garcin et al. 2008) and MPO (PDB entry: 5QJ2) (Wurtz et al. 2018) — using the fully automated protein–ligand docking tool JAMDA (Schellhammer and Rarey 2007; Henzl et al. 2014; Flachsenberg et al. 2020) available at https://proteins.plus (Fährrolfes et al. 2017; Schöning-Stierand et al. 2020). Each protein was prepared through the Protoss optimization routine, in which cofactors and structurally relevant water molecules were maintained. Each binding site was defined by the corresponding bona fide inhibitor, with a site radius of 6.5 Å. Molecular docking was executed with high precision.
Statistical analysis

All data were analyzed using the GraphPad Prism 6 tool (GraphPad Software Inc., San Diego, CA, USA) and expressed as means ± SEM. Indexes in acquisition of MWM trials, such as escape latency and escape rate, were analyzed by repeated-measure two-way ANOVA. One-way ANOVA followed by a Bonferroni post hoc test was used in the analysis of the other behavior parameters, biochemical and molecular experiments. For all statistical analyses, \( p < 0.05 \) was considered statistically significant.

Results

Characterization of OOEO

In the chemical analysis of OOEO, 32 individual components were determined, four of which constituted 90.8% of the said essential oil (Aykac et al. 2020; Table 1). The two major components of OOEO were found to be carvacrol and \( \gamma \)-terpinene (78.7% and 6.9%, respectively).

Effect of OOEO on locomotor activity in Sco-induced amnestic rats

The locomotor activity test was used to evaluate the negative effects on cognitive behavior such as spontaneous exploratory activities and arousal that may be caused by the administrated treatments. Our results suggest that Gal, OOEO, Sco and Sco + OOEO treatments did not affect the overall locomotor activities of rats \( [F(4,32) = 0.491 \text{ and } F(4,32) = 0.714, \text{ respectively}; \ p > 0.05] \). It was determined that Sco-induced amnesia group had less rearing frequency and shorter total distance travelled in 5 min compared to the Sco + OOEO groups, however, there was no significant change in the total distance between all groups (Fig. 2a,b).

OOEO ameliorated cognitive function and spatial memory in amnestic rats induced by Sco

Significant differences were identified between the treatment groups with the NOR test, which was designed to assess short-term recognition memory \( (F = 8.649, \ p < 0.001) \). The discrimination index was significantly reduced in Sco-induced amnesia rats compared to control rats \( (p < 0.001) \) and showed defective cognitive abilities (Fig. 2c). The administration of OOEO treatment to the amnestic rats \( (\text{Sco} + \text{OOEO group}) \) increased the discrimination index compared to the Sco group \( (p < 0.001) \). The discrimination index was significantly increased in Gal and OOEO groups compared to the Sco-induced amnesia group \( (p < 0.001, \text{ in both groups}) \).

### Table 1

| LRI   | RRI  | Compound name          | Relative percentage amounts (%) in the essential oil of *Origanum onites* |
|-------|------|------------------------|--------------------------------------------------------------------------|
| 1020  | 1032 | \( \alpha \)-Pinene     | 0.4                                                                      |
| 1024  | 1035 | \( \alpha \)-Thujene     | 1.1                                                                      |
| 1072  | 1076 | Camphene               | 0.3                                                                      |
| 1119  | 1118 | \( \beta \)-Terpinene   | 0.1                                                                      |
| 1172  | 1174 | Myrcene                | 0.6                                                                      |
| 1177  | 1179 | \( \alpha \)-Phellandrene | 0.2                                                                  |
| 1191  | 1188 | \( \alpha \)-Terpinene  | 1.3                                                                      |
| 1213  | 1203 | Limonene               | 0.2                                                                      |
| 1222  | 1218 | \( \beta \)-Phellandrene | 0.2                                                                  |
| 1260  | 1255 | \( \gamma \)-Terpinene  | 6.9                                                                      |
| 1287  | 1280 | \( p \)-Cymene          | 4.1                                                                      |
| 1298  | 1290 | Terpinolene            | 0.1                                                                      |
| 1457  | 1452 | 1-Octen-3-ol           | 0.1                                                                      |
| 1478  | 1474 | *trans*-Sabinene hydrate | 0.4                                                                  |
| 1555  | 1553 | Linalool               | 1.4                                                                      |
| 1564  | 1556 | *cis*-Sabinene hydrate | 0.2                                                                      |
| 1569  | 1565 | Linalyl acetate        | 0.1                                                                      |
| 1624  | 1609 | Terpinene-4-ol         | 0.7                                                                      |
| 1628  | 1612 | \( \beta \)-Caryophyllene | 0.5                                                                  |
| 1638  | 1628 | Aromadendrene          | 0.1                                                                      |
| 1717  | 1706 | \( \alpha \)-Terpineol  | 0.2                                                                      |
| 1728  | 1719 | Borneol                | 0.4                                                                      |
| 1748  | 1741 | \( \beta \)-Bisabolene | 1.1                                                                      |
| 1770  | 1751 | Carvone                | Tr                                                                       |
| 1786  | 1773 | \( \delta \)-Cadinene  | Tr                                                                       |
| 1793  | 1776 | \( \gamma \)-Cadinene  | 0.1                                                                      |
| 1899  | 1890 | Carvacryl acetate      | 0.1                                                                      |
| 2033  | 2008 | Caryophyllene oxide    | 0.1                                                                      |
| 2159  | 2144 | Spathunelol            | 0.1                                                                      |
| 2205  | 2187 | T-Cadinol              | 0.2                                                                      |
| 2210  | 2198 | Thymol                 | 0.2                                                                      |
| 2243  | 2239 | Carvacrol              | 78.4                                                                     |
| Total |      |                        | 100.0                                                                   |

LRI: linear retention indices against the \( n \)-alkane series. RRI: relative retention indices calculated against \( n \)-alkanes on a polar column (Demirci et al. 2004). Tr: trace (<0.1%)
day when compared with the Sco-induced amnesia group ($p < 0.001$ for both group).

Analysis of the escape rate displayed significant differences in treatment groups ($F = 446.8, p < 0.001$) and training days ($F = 137.8, p < 0.001$) (Fig. 3b). The rats in the Sco-induced amnesia group maintained statistically significant lower escape rate than control rats from the second day ($p < 0.001$; Fig. 3b). It was determined that the escape rate of the Sco+OOEO group increased significantly on the second day ($p < 0.001$). The escape rate of Gal and OOEO treatment groups was increased from the second day when compared with the Sco-induced amnesia group ($p < 0.001$ for both groups).

Significant differences were determined between the time spent in the target quadrant between treatment groups in the probe experiments without a platform on the fifth day ($F = 8.473, p < 0.001$; Fig. 3c). In the research trial to determine if they remembered platform localization, the
Fig. 3 Effects of *Origanum onites* essential oil on scopolamine-induced impairment of cognition, learning and recognition and spatial memory function. 

**a** Latency to find platform(s), **b** escape rate (%) and **c** time spent in the target quadrant(s) in the Morris water maze. Six rats per group. *p < 0.05 vs. control and **p < 0.05 vs. scopolamine-induced amnesia group.*
time spent in the target quarter was significantly reduced in the Sco-induced amnesia group compared to the control group ($p < 0.001$). OOEO treatment applied to the amnesia group significantly increased the time spent in the target quadrant compared to the Sco-induced amnesia group ($p < 0.001$). It was determined that the time spent in the target quadrant increased significantly in the Gal and OOEO groups compared to the Sco-induced amnesia group ($p < 0.001$).

OOEO reversed the progressive loss of cholinergic function and oxidative stress parameters in amnestic rats induced by Sco

The MDA levels of frontal cortex and hippocampus regions, an indicator of lipid peroxidation, was increased in the Sco-induced amnesia group as compared to the control group ($p < 0.001$) (Fig. 4a, b). In Gal, OOEO and Sco + OOEO groups, MDA level significantly decreased in both regions.

![Graphs showing MDA levels in different groups](image)

**Fig. 4** a–b Acetylcholinesterase (AChE) activity, c–d malondialdehyde (MDA) levels and d–e reduced glutathione (GSH) levels in the frontal cortex and hippocampal regions of scopolamine-induced amnestic rats. Six rats per group. * $p < 0.05$ vs. control and † $p < 0.05$ vs. scopolamine-induced amnesia group.
comparing to Sco-induced amnesia group ($p < 0.001$). The GSH levels of both regions was decreased in the Sco-induced amnesia group as compared to the control group ($p < 0.001$). In Gal, OOEO and Sco + OOEO groups, GSH level significantly increased in cortex comparing to Sco-induced group ($p < 0.001$, $p < 0.001$ and $p < 0.01$, respectively) (Fig. 4c), as in the hippocampus region ($p < 0.01$, $p < 0.01$, and $p < 0.05$, respectively) (Fig. 4d). AChE activity was increased in both regions in the Sco-induced group compared to the control group ($p < 0.001$, Fig. 4e and $p < 0.01$, Fig. 4f). Gal and OOEO treatment significantly reduced AChE activity compared to the Sco-induced group in cortex ($p < 0.001$ in both groups), as in the hippocampus ($p < 0.05$ in both groups). Thus, OOEO expressed an anti-AChE feature, which parallels improving memory function in rats, as observed in behavior tests.

**OOEO reduced apoptosis in the two different brain regions of Sco-induced amnestic rats**

Intrinsic apoptosis, a type of mitochondria-centered cell death, is characterized by the formation of apoptosomes, activation of casp-9 and subsequent activation of effector caspases. Essentially, the entire process is regulated by the BCL-2 family of proteins and maintained by the caspase family of proteins (Tang, 2019).

The ratio of bax to bcl-2 in the frontal cortex and hippocampal regions, an indicator of apoptosis, was increased in the Sco-induced amnesia group as compared to the control group ($p < 0.001$ and $p < 0.01$) (Fig. 5a,b). In Gal, OOEO and Sco + OOEO groups, the bax/bcl-2 ratio was significantly decreased in cortex ($p < 0.001$, $p < 0.01$ and $p < 0.05$) and hippocampus regions ($p < 0.001$, $p < 0.05$ and $p < 0.05$, respectively) comparing to Sco-induced group.

The expression levels of casp-3 and -9 in the frontal cortex and hippocampus, an indicator of intrinsic apoptosis, were increased in the Sco-induced amnesia group as compared to the control group ($p < 0.001$ for frontal cortex in both expression level; for hippocampus $p < 0.001$ and $p < 0.01$, respectively) (Fig. 5d,f). In Gal, OOEO and Sco + OOEO groups, casp-3 levels significantly decreased in frontal cortex ($p < 0.001$, $p < 0.01$ and $p < 0.05$, respectively) and hippocampus regions ($p < 0.01$, $p < 0.001$ and $p < 0.05$, respectively) comparing to Sco-induced group. In Gal, OOEO and Sco + OOEO groups, casp-9 levels significantly decreased in frontal cortex ($p < 0.01$ in all groups).

**Fig. 5** The density of immunoblotting of a–b bax/bcl-2 ratio, c–d casp-3 levels, and e–f casp-9 levels of the hippocampal and frontal cortex regions of rats in scopolamine-induced AD rat model ($n = 6$). Six rats per group. * $p < 0.05$ vs. control and + $p < 0.05$ vs. scopolamine-induced amnesia group. Sco: scopolamine; Gal: galantamine; OOEO: *Origanum onites* essential oil; Sco + OOEO: scopolamine plus *Origanum onites* essential oil.
and hippocampus regions (p < 0.001, p < 0.001 and p < 0.05, respectively) comparing to Sco-induced group.

**Carvacrol could be accommodated within the inhibitor-binding pockets of selected pro-inflammatory enzymes**

In an attempt to speculate about the possible anti-inflammatory effects of OOEO in the human body, we docked its major bioactive component carvacrol onto three of the known enzymes of inflammation — namely, COX-2, iNOS and MPO. The results of redocking calculations demonstrated that JAMDA was able to reproduce the crystallographic binding modes of the *bona fide* enzyme inhibitors well, with root-mean-square deviations (RMSDs) less than 1 Å (Table 2). In cross-docking experiments, carvacrol was found to be able to occupy the inhibitor-binding sites of COX-2, iNOS and MPO, albeit with lower docking scores compared to those for the *bona fide* inhibitors. The binding of carvacrol at the said sites of COX-2, iNOS and MPO appeared to be stabilized mainly by van der Waals forces and hydrophobic interactions (Fig. 6). In the case of the predicted MPO–carvacrol complex, π–σ (at 3.48 Å) and π–alkyl (at 4.07 Å) interactions were found to occur between the ligand and the heme prosthetic group of the protein.

**Discussion**

With a rapidly growing elderly population worldwide, a focus on finding efficient preventive and therapeutic strategies in the management of AD should prove timely and topical to researchers. The available evidence suggests that culinary herbs, such as *Melissa officinalis* L. (Lamiaceae), can potentially provide a natural treatment for AD (Howes et al. 2003; Gürbüz et al. 2019). In the relevant scientific literature, there also exist other studies indicating various clinical, cholinesterase inhibitory and neurobiological activities of different Lamiaceae plants (Perry et al. 2003; Orhan et al. 2008). All these as well as previous promising yet limited work on the bioactivity of traditionally consumed *O. onites* L. have enabled us in designing and executing the current study, in which the previously underdetermined neurobiological activity of the essential oil from this herb was investigated thoroughly. Putative neuroprotective and cognitive-ameliorating effects of OOEO on Sco-induced learning and memory impairments in rats were examined by behavioral tests, biochemical assessment, and computational simulations.

Here, we evaluated the cognitive effects of OOEO on memory and learning as well as the possible mechanisms underlying these effects in a rat model of Sco-induced amnesia. Initially, we tested whether OOEO treatment caused changes in behavioral parameters, such as locomotory activity and learning and memory abilities, in rats. Our results suggest that OOEO administered at a dose of 0.25 ml/kg enhanced memory functions previously impaired by Sco, corrected the negative impact of Sco on memory acquisition without affecting the locomotor activity, and also improved Sco-induced amnesia. It has been suggested that Sco at high doses (>0.03 mg/kg) exerts cognitive and non-cognitive effects on learning and memory performance in rats and mice, involving (i) primary or specific effects (e.g., inattention and sensory indiscrimination), (ii) secondary or non-specific effects (e.g., locomotor activity and anxiety), and (iii) peripheral side effects (e.g., pupil dilatation and salivation) (Klinkenberg and Blokland 2010). In fact, administration of Sco at 0.056 mg/kg and more increases locomotor activity (as a non-cognitive effect) in rats (Gholamrezal et al. 2002; Chintoh et al. 2003). In our experiments, we used a Sco concentration of 1 mg/kg which is ~18-fold higher than the minimal effective dose; however, no change was observed in the locomotor activities in different experimental groups. Therefore, OOEO treatment is likely to suppress the non-cognitive effects of Sco. The same also applies to the known anti-AChE drug Gal. Memory impairment caused by Sco administered i.p. to rats at a dose of 1 mg/kg was assessed based on the NOR test, which provides a rapid performance assessment for rodents (Van Morktazi–Borojeni et al. 2017). Rats treated with Sco were unable distinguish between the novel and familiar objects, which ultimately showed us that recognition was significantly deficient in this group of animals. Upon OOEO treatment, however, Sco-induced deterioration of cognitive abilities in rats was found to be abolished, and improvements in memory performance were noted. The results of the MWM test demonstrated that the administration of Sco significantly reduced the discrimination index, escape rate and time spent in the target quadrant and increased latency to find the platform, all of which are indicators of learning skills and memory in rodents. OOEO administration, however, was found to ameliorate Sco-induced impairments on the discrimination

| **Table 2** Derived docking scores relating to the best-selected binding pose for carvacrol in the inhibitor-binding pockets of pro-inflammatory enzymes |
|-----------------|-----------------|------------------|-----------------|
| **Bona fide inhibitor** | **RMSD** | **Docking score** | **Carvacrol** | **Docking score** |
| COX-2a | 0.532 Å | -2.52030 | -1.65951 |
| iNOSb | 0.682 Å | -2.62362 | -1.66283 |
| MPOc | 0.632 Å | -3.2663 | -1.64903 |
| aPDB entry 5KIR | bPDB entry 3E7G | cPDB entry 5QJ2 |
index, latency to find the platform, escape rate and time spent in the target quadrant. Collectively, these results suggest that OOEO may prevent Sco-induced learning and memory impairments in rats.

Due to the important role of the cholinergic system in learning and memory, maintaining acetylcholine levels are critical for brain functions (Blake et al. 2014). The action of AChE breaks down the naturally occurring neurotransmitter acetylcholine into acetate and choline at synaptic sites, which terminates or suspends cholinergic transmission (Ballard et al. 2005). Accordingly, an agent promoting the activity of AChE should ideally impair memory via lowering acetylcholine levels as Sco achieved in the amnestic group. On the other hand, in the present study, the standard drug Gal and OOEO significantly reduced the activity of AChE and allowed the rats to retain the memory of the tasks they learned as observed in our rich array of behavioral experiments. This AChE inhibition could be attributed to one or more components of OOEO. Indeed, AChE from electric eel has previously been shown to be inhibited by carvacrol (IC_{50}: 0.063 mg/ml) and its derivative thymohydroquinone (IC_{50}: 0.04 mg/ml) in an in-vitro setting (Jukic et al. 2007). Considering that impairments in the human brain cholinergic pathways are among the underlying causes of AD and that the use of AChE inhibitors that increase acetylcholine levels in the CNS constitutes the essential remedy for the disease (Terry and Buccafusco 2003), the inhibition of AChE in vivo by OOEO suggests that this essential oil may represent a novel candidate therapeutic agent for AD.

The CNS is particularly susceptible to pro-oxidants because of its unique architecture and operation, exemplified by a high oxygen consumption concomitant with a high unsaturated lipid content and low antioxidant protection (Kowalczyk et al. 2020). Therefore, it is no surprise that the overproduction of reactive oxygen species (ROS) in the CNS is associated with several neurodegenerative diseases including AD. The most commonly affected brain regions are believed to be the hippocampus, substantia nigra, and striatum (Phaniendra et al. 2015). Our study confirms that oxidative stress is involved in Sco-induced dementia in rats as revealed by an increase in the concentration of MDA, a secondary oxidation product of polyunsaturated fatty acids, and a decrease in the concentration of GSH, a major cellular antioxidant buffer molecule, in both hippocampus and frontal cortex. In contrary, the standard drug Gal and OOEO were able to significantly improve this oxidative stress profile in the single brain structures. ROS can also induce neuronal death in multiple forms including apoptosis (Fricker et al. 2018). Animal studies have shown that the activation and execution of apoptosis in neurons in the cortex and hippocampus impairs learning and memory (Kuhn et al. 2005; Sun et al. 2009). While encounter with Sco increased the bax/bcl-2 ratio and caspase levels in the frontal cortex and hippocampus of amnestic rats, Gal and OOEO treatments were effective in decreasing the production of apoptotic protein factors in both regions. Taken together, it is plausible to assume that the antioxidant and pro-survival properties of OOEO at a cellular scale may contribute to the reversal of Sco-induced memory deficits in rats.

Neuroinflammation can be defined as an inflammatory response within the CNS, which is usually caused by one of a variety of pathological insults including trauma, ischemia, infection, and toxins. The production of pro-inflammatory molecules (e.g., cytokines, chemokines, NO, and ROS) by innate immune cells, such as microglia/macrophages and astrocytes, in the course of neuroinflammation may ultimately result in synaptic dysfunction, impaired neurogenesis, and neuronal death (Leng and Edison 2021). A major contribution to neuronal death is ROS generation by the induction or activation of inflammation-related enzyme systems including COX-2, iNOS, and MPO. For instance, MPO has been shown to be aberrantly expressed in astrocytes in human AD brain (Maki et al. 2009). The involvement of MPO in disease pathogenesis is further supported by the presence of 3-chlorotyrosine, a specific biomarker of MPO-catalyzed oxidation, in the hippocampus from patients with AD (Green et al. 2004). In addition, activated macrophages/microglia are known to induce the expression of other downstream pro-inflammatory enzymes such as COX-2 and iNOS (Choi et al. 2009). It has been demonstrated that COX-2 levels are elevated in AD brain (Pasinetti and Aisen 1998; Yasojima et al. 1999; Earley et al. 2000) and that they correlate with the clinical progression of the disease (Ho et al. 2001). Similarly, iNOS expression has been found in the hippocampal region of the brain affected by AD and linked to neuronal degeneration in the disease (Sunhee et al. 1999). Our computational predictions illustrate that carvacrol can be housed in the inhibitor-binding pockets of MPO, COX-2 and iNOS. In fact, COX-2 from sheep has previously been shown to be inhibited by carvacrol in an in-vitro setting (Landa et al. 2009). Furthermore, the potency of COX-2 inhibition by carvacrol (IC_{50}: 0.8 μM) has been found to be comparable to that by an NSAID. It is therefore plausible to assume that the in vivo observed anti-inflammatory action of oregano (Taleb et al. 2018) and other traditionally used carvacrol-rich plant drugs (Sosa et al. 2005) could be partly due to the modulation of pro-inflammatory enzyme activities.

We acknowledge several limitations to our study, such as the inability to extrapolate animal data to human data and the difficulty of studying mood and behavior in experimental animals. Given the fact that AD is a chronic progressive disorder with a complex multifactorial etiology, the favorable findings of this study may need to be replicated in other disease models, such as transgenic animal models of AD, to adequately assess the potential for OOEO in AD.
Also, further research is needed to investigate the probable involvement of cholinergic receptors/channels or neurotransmitters other than acetylcholine (e.g., glutamate, gamma aminobutyric acid, and catecholamines) in the cognitive-enhancing activity of OEOE. It is to be noted, however, that any treatment that reverses symptoms, improves quality of life, slows neurodegeneration, and saves vast amounts of money marks a significant step forward in the quest to cure AD.

Author contributions  Asli Aykac, Kerem Terali, Dilek Özbeylı, Kemal Hüsnü Can Başer and Gökşel Şener conceived the study design. Aslı Aykac, Dilek Özbeylı, Seren Ede, Ömercan Albayrak and Gökşel Şener were responsible for animal experiments. Aslı Aykac and Kerem Terali were responsible for molecular experiments and statistical analysis. Aslı Aykac, Kerem Terali and Dilek Özbeylı wrote the original draft manuscript. Aslı Aykac, Kerem Terali, Kemal Hüsnü Can Başer and Gökşel Şener reviewed and edited final version of the manuscript. Aslı Aykac, Kerem Terali, Dilek Özbeylı, Seren Ede, Ömercan Albayrak, Kemal Hüsnü Can Başer and Gökşel Şener have seen and read the final version of the article. The corresponding author attests that all listed authors meet authorship criteria and that no others meeting the criteria have been omitted.

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Data availability  All data generated or analyzed during this study are included in this published article.

Declarations

Ethics approval  All procedures were carried out in accordance with the National Institute of Health Guide for the Care and Use of Laboratory Animals and the protocols were approved by the Marmara University Animal Care and Use Committee (MUHDEK approval no: 112.2016. mar).

Conflict of interest  The authors declare that they have no conflicts of interest.

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