Case description
An 11-year-old female spayed domestic medium-hair cat was presented for dental prophylaxis, at which time no oral mass was appreciated. Fifteen days after a dental cleaning, a mass expanding the oral mucosa of the rostral mandible was identified. An incisional biopsy revealed that the oral mucosa was infiltrated by neoplastic round-to-spindloid mesenchymal cells arranged in streams and small, dense aggregates consistent with an undifferentiated sarcoma. The patient was managed medically for approximately 6 months following the diagnosis, but, owing to declining health, euthanasia was elected and a post-mortem examination was performed. On post-mortem examination, the previously described neoplastic cells were infiltrating the rostral mandible and had metastasized to the right submandibular lymph node. Immunohistochemistry performed during the postmortem examination found that neoplastic cells were positive for Iba-1, CD18 and CD204, and negative for MUM-1, S100, Melan-A and E-cadherin, favoring a diagnosis of oral histiocytic sarcoma. Although recently recognized in cats, feline oral histiocytic sarcoma is rare, the tumor’s immunohistochemical profile is unstandardized, and the tumor’s behavior and prognosis are unclear. The diagnosis is challenging if small incisional biopsies are submitted and the neoplasm is poorly differentiated. This case report discusses the clinical, macroscopic, microscopic and immunohistochemical features of oral histiocytic sarcoma in a cat with mandibular invasion and submandibular lymph node metastasis.

Keywords: Histiocytic sarcoma; immunohistochemistry; metastasis; oral neoplasia; undifferentiated sarcoma

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were extracted or had crown amputations, leaving four upper incisors and two upper molars remaining. No masses or other abnormalities were noted on dental radiographs. Robenacoxib (Onsior; Elanco) and buprenorphine (Simbadol; Zoetis) were prescribed for 3 days for analgesia. Fifteen days post-procedure, the owner noticed a small, red swelling extending from the gingiva to the mucocutaneous junction on the rostral portion of the mandible. At the recheck appointment 21 days post-procedure, the cat was sedated for a more thorough examination of the swelling. Dental radiographs were performed, and a fine-needle aspirate and an incisional biopsy were obtained. Dental radiographs showed a soft tissue swelling of the rostral mandible.

Cytology of the aspirate identified large numbers of spindloid-to-rounded mesenchymal cells with moderate anisocytosis and anisokaryosis. A few of the mesenchymal cells were described as having a plasmacytoid appearance. The incisional biopsy of the rostral mandibular soft tissue swelling was 0.5 × 0.2 × 0.1 cm, homogeneously tan in color and firm. Microscopically, abutting intact lamina epithelialis mucosa, the lamina propria was infiltrated by neoplastic round-to-spindloid mesenchymal cells arranged in streams and small, dense aggregates that were unencapsulated and supported by pre-existing collagen. Cells had variably distinct cell borders, an oval-to-indentated, occasionally eccentrically located, nucleus with vesicular chromatin and 1–2 variably distinct nucleoli, and small-to-moderate amounts of eosinophilic, vacuolated cytoplasm. Additionally, some cells had subtle paranuclear clearing. The neoplastic cells exhibited mild nuclear pleomorphism, moderate anisocytosis and anisokaryosis, and the mitotic count was 10 mitoses per 10 high power (×400) fields. Immunohistochemistry for cytokeratin (AE1/AE3) was negative, ruling out an epithelial origin of the neoplastic cells. The final diagnosis was sarcoma, and plasmacytoma and lymphoma were listed as possible differential diagnoses. In lieu of additional immunohistochemistry, a second opinion by a board-certified pathologist specializing in oral pathology was requested. The second-opinion, final diagnosis was round cell neoplasm (favored plasmacytoma). Further immunohistochemistry was declined.

Owing to the poor prognosis for long-term survival and potential complications of mandibulectomy in cats, the owner elected for palliative care. Prednisolone (2mg/kg per day PO [Compounded; Roadrunner Pharmacy]) was prescribed and the patient was given buprenorphine (Simbadol; Zoetis) as needed for pain management. The mass grew rapidly over 6 months, with moderate amounts of hemorrhage. Euthanasia was elected when the cat was dysphagic and appeared uncomfortable 195 days after the initial dental procedure (180 days after the owner’s initial observation of the mass).

On gross examination of the oral cavity, there was an irregular 3.0 × 2.5 × 1.5 cm, firm, ulcerated, tan-to-dark red (previous hemorrhage) infiltrative mass arising from the right rostral mandibular gingiva, invading and expanding the rostral portion of the right mandible, and extending to the right ventral aspect of the tongue (Figure 1). All teeth were absent except the right and left maxillary molars and incisors. The right submandibular lymph node was firm and on cut section there was a focal 0.3 cm diameter tan nodule obliterating the parenchyma. Aside from the oral mass, additional postmortem findings included peritoneopericardial hernia and unilateral renal lymphoplasmacytic interstitial nephritis and fibrosis. All other organ systems were grossly unremarkable. Samples of the oral mass, right mandible, tongue, mucocutaneous junction of the mass, right submandibular lymph node, liver, lungs, adrenal glands, stomach, small and large intestines, kidneys, brain, urinary bladder, heart, spleen and pancreas were placed in 10% neutral buffered formalin, processed routinely, paraffin embedded, sectioned at 4µm and stained with hematoxylin and eosin.

Microscopically, the oral neoplasm was invasive and consisted of poorly differentiated, spindloid-to-round cells arranged in streams, bundles and occasionally sheets. Within the sections of the mandible and right
submandibular lymph node, the previously described neoplastic cells were invading and replacing the bone (Figure 2a) and were effacing and replacing the normal architecture of the right submandibular lymph node (Figure 2b). Neoplastic cells were not identified in any of the other examined tissues.

Discussion

Histiocytic sarcomas in cats are considered uncommon and have been sporadically reported in a wide variety of anatomic locations, including the tarsus, mediastinum, disseminated, eye, nasopharynx and brain.1–13 Oral histiocytic sarcomas, in particular, are considered rare in any species.14–16 To date, there has only been one published report in a cat.17 The most common differential diagnosis for an oral neoplasm in a cat is squamous cell carcinoma followed by fibrosarcoma.14,18 Our case report describes the second known case of oral histiocytic sarcoma in a cat. The novel features of this case include the presence of extensive mandibular invasion and regional (right submandibular) lymph node metastasis. Additionally, this report is the first to use CD204 as one of the diagnostic steps in differentiating histiocytic sarcoma from the other, more common, poorly differentiated feline oral tumors, and is the first to use immunohistochemistry to determine the histiocytic lineage of oral histiocytic sarcoma in cats.

The tumor’s neoplastic cells had large, indented-to-reniform nuclei and abundant eosinophilic cytoplasm; multinucleation was also occasionally observed (Figure 3a). Based on the microscopic features, the differential diagnoses included fibrosarcoma, amelanotic melanoma, plasmacytoma and histiocytic sarcoma. Accordingly, immunohistochemistry for Melan-A, S-100, MUM-1, Iba-1 and CD18 was performed. The diagnosis of histiocytic sarcoma was then made based on the presence of strong membranous staining for ionized calcium-binding adapter molecule 1 (Iba1) (Figure 3b) and CD18. Both Iba-1 and CD18 occur in all subpopulations of the macrophage/monocyte lineage (including Langerhans and interstitial dendritic cells [DCs]), in inflammatory and neoplastic histiocytes, as well as in normal histiocytes.19 The neoplastic cells were also frequently positive for CD204 (Figure 3c); this is a scavenger receptor that is expressed by macrophages, and is a reliable marker for histiocytic sarcomas in dogs.20,21 Neoplastic cells were diffusely negative for Melan-A, S100 and MUM-1 (Figure 3d), ruling out melanoma and plasma cell tumor. Spleen was used as a positive control for all histiocytic markers (CD18, Iba-1 and CD204) and MUM-1.

In dogs, immunostaining of most histiocytic sarcomas suggests that they are derived from interstitial DCs (CD1a+, MHC class II+ and CD11c/CD18+).1,22 Interstitial DCs are resident DCs typically found within most organs except the brain.23 Further classification of the histiocytic lineage of this neoplasm was attempted by performing immunohistochemistry for E-cadherin. E-cadherin staining, if present, suggests that neoplastic cells are likely of Langerhans DC origin. The neoplastic cells, in this case, were diffusely negative for E-cadherin; while this is suggestive of interstitial DC origin, it is important to note that a lack of E-cadherin staining does not completely rule out Langerhans DC origin because, after activation, Langerhans DCs can downregulate their expression of E-cadherin as they migrate to the dermis.1,24,25 To definitively determine that histiocytes are of Langerhans DC origin, electron microscopy is needed to identify cytoplasmic Birbeck granules. Birbeck granules are induced when Langerhans DCs are exposed to transforming growth factor beta.1,24–26 Further classification of the histiocytes with immunohistochemistry indicates that these neoplastic cells are most likely of interstitial DC origin. As previously mentioned, this finding is consistent with the current literature, as histiocytic sarcomas are predominantly of interstitial DC origin.1,22
While histiocytic sarcomas of interstitial DC origin are most common, there are rare reports of neoplasms of macrophage–myeloid origin. For example, a nasal histiocytic sarcoma of macrophage–myeloid type was reported in a cat.\textsuperscript{13} There was infiltration of the bone marrow but no nodal metastasis and neoplastic cells were predominantly round and demonstrated phagocytic activity. The cells were immunohistochemically positive for lysozyme, CD18 and MHC class II, and cytochemically positive for alpha naphthyl acetate esterase.\textsuperscript{13}

**Figure 3** Oral histiocytic sarcoma in a cat: microscopic and immunohistochemical features. (a) Neoplastic cells are pleomorphic, ranging from round to spindloid and occasionally are multinucleated (inset, arrowheads); hematoxylin and eosin × 400. Immunohistochemistry for (b) ionized calcium-binding adaptor molecule 1 (Iba-1), (c) CD204 and (d) multiple myeloma 1 (MUM-1); hematoxylin and eosin × 400
Conclusions
In cats, histiocytic sarcoma is a rare differential diagnosis for primary oral neoplasia; however, this entity should be considered if, microscopically, the neoplasm is consistent with a poorly differentiated, pleomorphic sarcoma. Immunohistochemistry to confirm histiocytic origin and to exclude more common diagnoses is strongly recommended and may include CD18, Iba-1 and CD204. In this case, within 6 months, this aggressive tumor had invaded the mandible and metastasized to a regional lymph node. To our knowledge, while this is the second report of oral histiocytic sarcoma in a cat, it is the first to elucidate the tumor’s behavior, predict patient prognosis and reaffirm what the literature purports; that is, histiocytic sarcomas typically originate from interstitial DCs.

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Ethical approval
This work involved the use of non-experimental animals only (including owned or unowned animals and data from prospective or retrospective studies). Established internationally recognized high standards (‘best practice’) of individual veterinary clinical patient care were followed. Ethical approval from a committee was therefore not specifically required for publication in JFMS Open Reports.

Informed consent
Informed consent (either verbal or written) was obtained from the owner or legal custodian of all animal(s) described in this work (either experimental or non-experimental animals) for the procedure(s) undertaken (either prospective or retrospective studies). No animals or humans are identifiable within this publication, and therefore additional informed consent for publication was not required.

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