Resistin-Like Molecule-β Promotes Invasion and Migration of Gastric Carcinoma Cells

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Background: Resistin-like molecule-β (RELMβ) is a novel secretory protein from intestinal goblet cells and participates in epithelial differentiation, tumor occurrence, and immune response. RELMβ is absent in normal gastric mucosa tissue, but is abundantly expressed in gastric carcinoma tissues, and is correlated with tumor invasion and metastasis. Epithelial-mesenchymal transition (EMT) is an important mechanism governing tumor cell invasion. This study thus investigated the modulation of RELMβ in gastric cancer metastasis and its correlation with EMT.

Material/Methods: We used RELMβ-low expression AGS cell line of gastric cancer and normal mucosa cell line GES1 as in vitro models, on which RELMβ-expressing vector was transfected. The invasion and migration of cells were quantified by Transwell assay. EMT-related protein including E-cadherin, N-cadherin, Snail, and Vimentin were detected by Western blotting in transfected AGS cells.

Results: RELMβ transfection significantly potentiated invasion and migration abilities of AGS cells, whose RELMβ protein level was significantly elevated compared to those in untransfected AGS or GES1 cells. After RELMβ transfection, EMT-related proteins, including N-cadherin, Snail, and Vimentin levels, were elevated, but E-cadherin expression was depressed.

Conclusions: RELMβ-overexpression can facilitate invasion and migration of gastric carcinoma cells and it increases the expression of EMT-related proteins, such as N-cadherin, Snail, Vimentin, but decreases E-cadherin level, thus promoting the progression of EMT.

MeSH Keywords: Activated-Leukocyte Cell Adhesion Molecule • Carcinoma, Acinar Cell • Precursor Cells, B-Lymphoid

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Background

The members of the resistin-like molecules (RELMs) family share the same cysteine structure. As an important family member, RELMb is secreted by intestinal goblet cells [1]. RELMb has been shown to be abundantly expressed in gastric carcinoma tissues and is closely correlated with biological behaviors and clinical features of tumors [2]. Endothelial-mesenchymal transition (EMT), an important process underlying tumor invasion and metastasis, requires the participation of multiple cytokines for the transformation of epithelial cell genes [3]. This study transfected RELMb-expressing vector into gastric carcinoma cell line and normal gastric mucosa epithelial cells, on which cell invasion and metastatic abilities, along with EMT-related factors, including E-cadherin, N-cadherin, Snail, Vimentin, were measured.

Material and Methods

Cell culture and transfection

Gastric carcinoma cell line AGS and normal gastric mucosa epithelial cell line GES1 were purchased from Baili Biotech (Shanghai, China). Cells were cultured in RPMI1640 medium (Gibco, USA) in a humidified chamber at 37°C perfused with 5% CO₂.

Recombinant vector pClR1- RELMb and expressing vector peDNA3.1/Zeo(+) were digested by EcoRI enzyme, followed by gel purification and de-phosphorylation. Log-phased cells were seeded into 6-well plates in 3 groups: blank control (no transfection), empty vector pcDNA3.1/Zeo(+) transfection, and pcDNA3.1- RELMb transfected groups. The transfection was performed using the Lipofealmine2000 kit (Invitrogen, USA) following the manufacturer’s instructions.

Real-time PCR

Log-phased AGS and GES1 cells were collected, along with transfected AGS cells. Total RNA was extracted by use of the Trizol kit (Invitrogen, USA) following the manual instruction. After quantification by A260/A280 ratio, 200 ng RNA was used as the template for synthesizing cDNA using polyA tail. PCR amplification was performed using a fluorescent quantitative PCR kit (Sangon, China). Specific primers were designed as follows: " RELMb: 5’-CACCATGAAG CCTACACTGTGTTTCC-3’ (Forward), 5’-TTAACCTCGCGCAAGGCGG-3’ (Reverse); GAPDH: 5’-GCAAGCTATCCATGACAACTTTGCC-3’ (Forward), 5’-GCCTGCTTACACAC TCCTGATGTC-3’ (Reverse)". PCR amplification consisted of 40 cycles each containing 95°C denaturation for 30 s, 60°C annealing for 30 s and 72°C elongation for 45 s. The relative mRNA expression was calculated by comparative Ct method.

Transwell chamber assay

For evaluating invasion ability, A Transwell chamber (Millipore, USA) was pre-coated with Matrigel at 4°C overnight, followed by incubation of serum-free medium on the artificial membrane at 37°C for 1 h. Transfected AGS cells were seeded in the upper chamber, while RPMI1640 medium was added into the lower chamber. At the endpoint, cells were stained by Giemsa and observed under an inverted microscope. For evaluating cell migration ability, cells were seeded in the chamber in the same manner without an artificial basal membrane. Cell invasion and migration ability was assessed through counting the numbers of positive-staining cells under a microscope.

Western blotting

Transfected AGS cells were collected for total protein extraction. After quantification, proteins were separated in 8% SDS-PAGE (40 μg per well) and were transferred to the PVDF membrane. The

| Group          | RELMb mRNA | AGS cell  | GES1 cell  |
|----------------|------------|-----------|------------|
| Transfected AGS cell | 3.25±0.81*# | 2.79±0.61 | 0.97±0.08  |

* p<0.05 compared to AGS cells; # p<0.05 compared to GES1 cells.
membrane was blocked for 1 h at room temperature. Mouse anti-human RELMβ monoclonal antibody (1:200, Santa Cruz, USA), mouse anti-human vimentin, N-cadherin, E-cadherin polyclonal antibodies (1:1 2000, Santa Cruz, USA), or mouse anti-human Snail monoclonal antibody (1:700, Cell Signaling, USA) was applied for 4°C overnight incubation. After being cultured in secondary antibody and development, the membrane was exposed and the image was captured and analyzed. Intensity of positive bands was quantified using Image J software. Protein expression was quantified as a ratio to loading control (β-actin or GADPH).

Statistical analysis

We used the SPSS 17.0 software package to process all collected data. Measurement data are presented as mean ± standard deviation (SD), while enumeration data were compared by chi-square test. The t test was used for between-group-comparisons. Statistical significance was defined when p<0.05.

Results

RELMβ expression in transfected AGS cells

Using real-time PCR to detect the mRNA level of RELMβ in AGS cells after transfection, we found significantly elevated RELMβ mRNA level after transfection (p<0.05, Table 1, Figure 1).

Cell migration and invasion assay

Using Transwell assay to detect migration and invasion ability of RELMβ-transfected AGS cells, results showed significantly
potentiated invasion and migration ability of AGS cells after transfection (p<0.05, Figure 2).

**Table 2. RELMβ protein expression.**

| Group          | RELMβ transfected | AGS cell | GES1 cell |
|----------------|-------------------|----------|-----------|
| RELMβ protein  | 5.47±0.89*#       | 2.61±0.03| 0.38±0.02 |

* p<0.05 compared to AGS cells; # p<0.05 compared to GES1 cells.

**Figure 3. RELMβ protein expressions. β-actin was used as a loading control.**

**Table 3. EMT-related protein expression levels.**

| Group     | RELMβ transfected | AGS cell | GES1 cell |
|-----------|-------------------|----------|-----------|
| E-cadherin| 0.14±0.03*        | 0.26±0.05| 0.44±0.07 |
| N-cadherin| 0.76±0.09*        | 0.46±0.05| 0.15±0.07 |
| Snail     | 0.65±0.04*        | 0.37±0.08| 0.18±0.06 |
| Vimentin  | 1.16±0.12*        | 0.28±0.02| 0.13±0.07 |

* p<0.05 compared to AGS cells; # p<0.05 compared to GES1 cells.

**Discussion**

The RELMs family is a group of secretory proteins containing abundant cysteine. It consists of 4 major family members: RELMα/HIMF, RELMβ/FIZZ2, Resistin/FIZZ3, and RELMγ/FIZZ4 [4,5]. RELMβ was first discovered in epithelial cells of the mouse intestine, while human RELMβ is mainly distributed in pulmonary vessels, bronchial mucosa epithelial cells, intestinal epithelial cells, kidney, and adrenal glands [6]. Studies have confirmed the elevated RELMβ secretion from intestinal goblet cells in mice infected with nematodes, and such secretion was mediated by cytokines, including IL-4, IL-13, and STAT6 for specific mucosal immunity and gastrointestinal defense [7]. People with elevated plasma RELMβ levels normally have smoking history or deficits of exercise, both of which are independent risk factors for gastric carcinoma [8].

In this study we transfected RELMβ-expressing vector into gastric cancer AGS cells, in parallel with blank AGS and normal gastric mucosal epithelial GES1 cells as controls. Real-time PCR and Western blotting showed significantly elevated...
expression level of RELMβ in those cells after transfection. Transwell assay demonstrated increased migration and invasion abilities of transfected AGS cells. A further comparison showed that, even in intact AGS cells, RELMβ expression level was still higher than in GES1 cells. These data collectively show that over-expression of RELMβ can facilitate the migration and invasion of gastric carcinoma cells. It has been discovered that RELMβ is up-regulated in colorectal cancer tissues compared to those in normal intestine epithelial cells. Moreover, serum RELMβ in gastrointestinal tumor patients was positively correlated with tumor stage and malignancy [9,10]. A systematic study of 156 gastric cancer patients also revealed the correlation between higher RELMβ level and unfavorable prognosis, possibly due to the elevated expression of RELMβ in gastric carcinoma tissues and further induction of matrix metalloproteinase-2 (MMP-2) and MMP-9 from mononuclear cells for tumor progression [11]. Esophagus carcinoma and Barrett esophageal atypical proliferation cells also had elevated serum RELMβ levels with advancement of TNM stage, making it a potential marker for esophageal squamous carcinoma. RELMβ has also been suggested to facilitate mitosis, proliferation, and migration of fibroblasts [12]. The binding of CDX-2 onto human RELMβ can facilitate proliferation of intestinal epithelial cells, modulate cell cycle, and potentiate cell-to-cell adhesion, making the RELMβ level negatively correlated with tumor differentiation stage [13,14].

EMT has been shown to be closely related with invasion and migration of malignant tumors, mainly due to the acquisition of mesenchymal cells in epithelial-derived cells [15]. As a tumor-suppressor gene, E-cadherin can limit the peripheral infiltration of tumor cells, along with the invasive migration [16]. N-cadherin, however, mainly inhibits cell apoptosis and is positively expressed in the formation and differentiation of embryonic tissues [17]. Vimentin is expressed in mesenchymal tissues and is related with tumor invasion and metastasis, making it an important cytokine in aggravating tumor malignancy [18]. The incidence of gastric carcinoma in China is increasing. Due to its insidious onset, gastric cancer is usually diagnosed at its late stage. With the progression of disease, there will be aggravated tumor cell invasion, which can be reflected by EMT as decreased epithelial marker E-cadherin and increased mesenchymal markers N-cadherin, Snail, and Vimentin [19,20]. In this study, we found elevated protein levels of N-cadherin, Snail, and Vimentin in RELMβ-transfected AGS cells, in addition to decreased expression of E-cadherin, all of which suggest the progression of EMT. We thus propose that the down-regulation of RELMβ may impede the occurrence of EMT, although further studies are required for substantiation.

Conclusions

RELMβ is abundantly expressed in gastric carcinoma cells, and can facilitate the invasion and migration of tumor cells via facilitating EMT, as it increased expression of EMT-related proteins, including N-cadherin, Snail, and Vimentin, in addition to inhibiting E-cadherin expression. The early and timely detection of RELMβ, therefore, may provide a novel diagnostic index and/or drug target for gastric cancer.

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