Higher aggrecan 1-F21 epitope concentration in synovial fluid early after anterior cruciate ligament injury is associated with worse knee cartilage quality assessed by gadolinium enhanced magnetic resonance imaging 20 years later

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Research article

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Abstract

Background. To investigate if cartilage related biomarkers in synovial fluid are associated with knee cartilage status 20 years after an anterior cruciate ligament (ACL) injury.

Methods. We studied 25 patients with a complete ACL rupture without subsequent ACL reconstruction or radiographic knee OA. All had a delayed gadolinium-enhanced magnetic resonance imaging of cartilage (dGEMRIC) 20 years after the ACL injury, using the T1 transverse relaxation time in the presence of gadolinium (T1Gd) which estimates the concentration of glycosaminoglycans in hyaline cartilage. Synovial fluid samples were aspirated acutely (between 0 and 18 days) and during 1 to 5 follow up visits between 0.5 and 7.5 years after injury. We quantified synovial fluid concentrations of aggrecan (epitopes 1-F21 and ARGs), cartilage oligomeric matrix protein, matrix metalloproteinase-3 and tissue inhibitor of metalloproteinase-1 by immunoassays, and sulfated glycosaminoglycans by Alcian blue precipitation. Western blot was used for qualitative analyses of aggrecan fragments in synovial fluid and cartilage samples.

Results. Western blot indicated that the 1-F21 epitope was located within the chondroitin sulfate 2 region of aggrecan. Linear regression analyses (adjusted for age, sex, body mass index and time between injury and sampling) showed that acute higher synovial fluid 1-F21-aggrecan concentrations were associated with shorter T1Gd values 20 years after injury, i.e. inferior cartilage quality (standardized effects between -0.67 and -1.0). No other statistically significant association was found between molecular biomarkers and T1Gd values.

Conclusion. Higher acute synovial fluid 1-F21-aggrecan concentrations in ACL injured patients, who managed to cope without ACL reconstruction and were without radiographic knee OA, were associated with inferior knee cartilage quality assessed by dGEMRIC 20 years after injury.

Background

Post-traumatic osteoarthritis (OA) is common after an anterior cruciate ligament (ACL) injury and is manifested by radiographic structural knee joint changes with osteophytes and decreased cartilage height, and with patients experiencing knee pain and stiffness (1-6). Concomitant acute traumatic knee cartilage injuries are very common in ACL injured knees (7). The mechanical damage is usually evidenced by superficial cartilage fibrillation and sometimes also with visible cracks down to the subchondral bone, and bone marrow lesions are present in almost every magnetic resonance imaging (MRI) after an acute ACL injury (8, 9). Even if there is no visual damage to the cartilage surfaces at the time of arthroscopy there may be micro-damage to cartilage matrix and cell death especially in the superficial regions (10). The ACL injury with cartilage damage triggers an immediate inflammatory response which acts in combination with an abnormal long-term mechanical loading of the injured knee believed to generate post-traumatic OA (11-13).
We lack means to diagnose and treat early microscopic joint changes in cartilage; radiography is limited by its insensitivity in detecting these early joint changes, and they are not visible until years after disease onset when the cartilage might be beyond repair (14, 15). Different molecular markers or combinations of biomarkers in synovial fluid, serum and urine have been suggested to be useful as prognostic OA-markers (16-22). Altered turnover and loss of cartilage sulfated glycosaminoglycans (sGAG) is a recognized and important early event of the development of OA (23). The delayed gadolinium-enhanced MRI of cartilage (dGEMRIC) is a non-invasive quantitative MRI technique that reflects the content of highly negatively charged macromolecules, such as sGAG, in the cartilage (24). A strong correlation between dGEMRIC estimated cartilage sGAG content and histological scores has been found (25). The dGEMRIC technique and study protocol have been validated (26), and clinically relevant associations between the dGEMRIC and risk factors for OA have been presented (27, 28). The dGEMRIC technique has also proved to have a prognostic value for OA development (29-31).

Studies of associations between molecular biomarkers and MRI cartilage findings have been called for (32). Only a couple of studies on association between synovial fluid molecular biomarkers and MRI cartilage findings 3 to 5 years after an ACL injury have been published (33, 34), and studies with longer follow-up time are lacking.

The aim of the present study was to examine if the concentration of molecular biomarkers in synovial fluid taken 0 to 7.5 years after ACL-injury were associated with knee cartilage quality assessed by dGEMRIC 20 years later.

**Methods**

*Subjects and visits*

Patients were from a well characterized cohort of 100 consecutive ACL-injured subjects prospectively recruited at the Lund University Hospital between 1985 and 1989 (35). All 100 subjects had a complete ACL tear and were within 18 days after initial trauma assessed by arthroscopy and x-ray with no significant signs of pre-existing knee OA (Figure 1a and 1b). The participants were treated with early physiotherapeutic knee rehabilitation without primary ACL reconstruction. Synovial fluid was collected early after injury (called acute visit; 0 to 18 days) and prospectively at 1 to 5 visits during the following 7.5 years (Figure 1b). For another study with the purpose to examine the association between knee cartilage quality and knee function, 32 subjects without ACL reconstruction or radiographic signs of OA at the 16-year follow up (described below) were examined with dGEMRIC 20 years after their ACL injury (36). Since the dGEMRIC method is reliant on the presence of joint cartilage, only subjects having Osteoarthritis Research Society International (OARSI, (37)) atlas grades of ≤ 1 were included in the study. Twenty-five of the 32 subjects examined with dGEMRIC had one or more available synovial fluid sample aspirated following their injury and were included in this study (Figures 1a and 1b, Table 1).

*Radiography at the 16 year follow up*
Radiographs at the 16 year (range 11-18 years) follow up were obtained in standardized standing anteroposterior knee position with both knees in 20 degrees of flexion and weight bearing on a tilt table; a fluoroscopically positioned x-ray beam was used to optimize medial tibial plateau alignment. The radiographs were independently read by two observers blinded to clinical details. Joint space narrowing (JSN) and osteophytes were graded independently on frontal images on a 4-point scale (range 0-3, 0 = no evidence of JSN or bony change) according to the OARSI atlas (14, 15, 37). The interrater reliability (kappa statistic) was $\kappa = 0.78$ for JSN and $\kappa = 0.52$ for osteophytes (38).

Synovial fluid sampling

Twenty-five subjects were included in this study with any kind of synovial fluid samples, i.e. either from first and/or following visit(s) as follows: 20 subjects had their synovial fluid aspirated at the acute visit within 18 days (median 6 days) after injury, and 22 subjects had their synovial fluids collected at between one and five visits during the subsequent 7.5 years of follow-up (median 4 years); these synovial fluids are called chronic samples (Figure 1b, Table 1). The subjects visited the orthopedic outpatient ward only for study purposes (35, 38). All synovial fluids were collected without joint lavage, and the samples were centrifuged at 3000xg for 10 minutes in room temperature and supernatants were stored at -80ºC.

Molecular marker analyses in synovial fluid

sGAG, in synovial fluid mainly chondroitin and keratan sulfate (CS and KS), was quantified by Alcian Blue precipitation (39). Two different aggrecan epitopes were quantified using immunoassays and the monoclonal antibodies (mAb) 1-F21 and OA-1. According to previous publications, mAb 1-F21 is suggested to recognize a protein sequence within or close to the KS region of aggrecan (18, 40). mAb OA-1 recognizes the ARGs neoeptope generated by aggrecanase cleavage at the TEGE$^{392/393}$ARGS site in the interglobular domain of aggrecan (41). Cartilage oligomeric matrix protein (COMP) was quantified using a commercial assay from AnaMar AB/IDS (cat. no. AN-14-1006-71); the AnaMar COMP-epitope has not been published. Matrix metalloproteinase-3 (MMP-3) and tissue inhibitor of metalloproteinase-1 (TIMP-1) were quantified using monoclonal and polyclonal antibodies; the MMP-3 immuno-assay recognizes both the pro- and active form of the protease and the complex with TIMP; the TIMP-1 immuno-assay detects only free TIMP-1 (42-44). Data on ARGs-aggrecan was generated for this study, all other biomarker data were available from previous studies on the described ACL cohort (45, 46).

The ratio MMP-3/TIMP-1 was used to investigate differences in these biomarkers alone or as a ratio between the enzyme and its inhibitor. We further investigated the ratios of sGAG/COMP, ARGs-aggrecan/COMP and 1-F21 aggrecan/COMP as biomarkers; ratios like these have been suggested to minimize the influence of varying amounts of obtainable synovial fluid (47).

Assessment with dGEMRIC at the 20 year follow up

Subjects were investigated with dGEMRIC on average 20.6 years (range between 18 and 23 years) after the ACL injury (Figure 1b, Table 1). Briefly, Gd-DTPA$^{2-}$ (Magnevist$,^6$ Schering AG, Berlin, Germany) was
injected intravenously at a dose of 0.3 mmol/kg body weight. To optimize the uptake of Gd-DTPA$^{2-}$ into the cartilage, subjects exercised by walking up and down the stairs for approximately ten minutes, starting five minutes after injection. Two hours after injection, post-contrast imaging of the cartilage was performed using a standard 1.5 T MRI system with a dedicated knee coil (Magnetom Vision; Siemens Medical Solutions, Erlangen, Germany). Central parts of the weight-bearing lateral and medial femoral cartilage were identified, and quantitative relaxation time calculations were performed in a 3 mm thick sagittal slice on each condyle, using sets of six turbo inversion recovery images with different inversion times: TR = 2000 ms, TE = 15 ms, FoV 120 x 120 mm$^2$, matrix = 256 x 256, TI = 50, 100, 200, 400, 800 and 1600 ms. A full-thickness region of interest (ROI) in the cartilage was examined. T1Gd was calculated using the mean signal intensity from each ROI (48), and the dGEMRIC images were analyzed and ROIs were drawn using the MATLAB-based Mokkula software (26). An orthopaedic surgeon performed the ROI measurements. All MRI data was available from a previous study (36).

Western blot of aggrecan

Aggrecan fragments from synovial fluid (pooled from 47 subjects with knee OA or knee injury) were purified by mini-preparations of cesium-chloride density-gradient centrifugation in absence or presence of guanidinium chloride, collecting the associative A1 and dissociative D1 fractions, as described (49). Purified aggrecan (i.e. A1D1 fraction prepared from pooled knee cartilage from ten subjects with OA) was in vitro digested using aggrecanase-1 (ADAMTS-4, a disintegrin and metalloproteinase with thrombospondin motifs-4) or MMP-3 as described (50). The samples were deglycosylated and separated by SDS-PAGE on 3-8% Tris-acetate mini-gels and transferred to PVDF-membranes (39). For the immune-reaction we used antibodies against aggrecan G1-domain (Affinity BioReagents no. PA1-1747, polyclonal IgG diluted 1:400), 1-F21 aggrecan epitope (IgG monoclonal antibody diluted 1:75000), ARGS-aggrecan epitope (IgG monoclonal neoeptiote antibody OA-1 diluted to 5.3 µg/ml) and chondroitin sulfate clone 3B3 (Seikagaku no. 270789 IgM monoclonal antibody against chondroitinase treated chondroitin 6-sulfate diluted to 0.33 µg/ml). Secondary antibodies were peroxidase-conjugated horse anti-mouse IgG (CST no. 7076S diluted to 10 ng/ml), goat anti-mouse IgM (Sigma no. 8786 diluted to 10 ng/ml) and goat anti rabbit IgG (KPL no. 074-1516 diluted to 13 ng/ml). The immunobands were visualized using Pierce ECL Plus Western Blotting Substrate (no. 32132) and film (Amersham Hyperfilm ECL) or luminescence image analyser Bio-Rad ChemiDoc MP.

Statistical analysis

Associations between the molecular biomarkers and dGEMRIC T1Gd values were investigated using linear regression models with adjustments for age at injury, sex, body mass index at dGEMRIC examination and time between injury and biomarker sampling. Results from crude (without adjustments) linear regression analyses are presented as a supplement (Table S1). Mann-Whitney tests were used for comparison of biomarker values between acute and chronic subject groups. For correlation analysis Spearman's rank ($r_S$) was used. For subjects with more than one chronic sample, the average biomarker concentration and the average time after injury were used in the linear regression model. The dGEMRIC
values were normally distributed. Biomarker data were log10 transformed to obtain normal distribution. To be able to compare effect sizes between biomarkers, we report standardized effects from the linear regression analyses. The reported effects estimate how many standard deviations the dependent variable (dGEMRIC) will change per standard deviation increase in the predictor variable (biomarker concentration). All tests were 2-tailed and \( P \leq 0.05 \) was considered statistically significant. The statistical analysis was performed with SPSS 24.0 for Windows software package.

**Results**

**dGEMRIC (T1Gd) and synovial fluid biomarker values**

The mean (standard deviation, SD) T1Gd dGEMRIC values at 20 years post injury for the 25 subjects was 397 ms (53) for the medial femoral cartilage, 431 ms (81) for the lateral femoral cartilage and 414 ms (58) for the medial and lateral femoral cartilage. For all biomarkers measured in synovial fluid, the concentrations were higher in the acute samples compared to chronic samples (Table 2).

**Associations between synovial fluid biomarkers and dGEMRIC at 20 years**

Of all investigated biomarkers, the only statistically significant associations found were between dGEMRIC and 1-F21 aggrecan and 1-F21 aggrecan/COMP ratio in the acute samples (Figure 2). These biomarker values were inversely associated with T1Gd values in the medial, lateral and combined compartments (Figure 2). The standardized effect sizes ranged from -0.67 to -1.0, and were similar between 1-F21 aggrecan alone or as a ratio of 1-F21 aggrecan/COMP. Crude linear regression analyses between molecular biomarkers and dGEMRIC showed similar associations as the adjusted analyses (Supplementary Table S1).

**Investigation of aggrecan assay specificity**

There was a positive correlation between the aggrecan markers (1-F21 aggrecan, sGAG and ARGS-aggrecan) detected in the acute samples (\( r_S = \) between 0.697 and 0.789, \( p \leq 0.006, n = 14-16; \) Figure S1). Since only 1-F21 aggrecan of the three different aggrecan assays showed associations with subsequent cartilage quality, we investigated what type of aggrecan and proteoglycans the different quantitative aggrecan and proteoglycan assays detected in synovial fluid. In Western blots we used the same aggrecan antibodies as in the immunoassays (i.e. against ARGS-aggrecan and 1-F21 aggrecan) and as a control for Alcian Blue detected proteoglycans we used the 3B3 antibody. Samples used in these experiments were two different density-gradient centrifuge fractions (A1 and D1) of aggrecan purified from pooled synovial fluid. The result showed clear differences in the type of aggrecan fragments detected by the antibodies in synovial fluid (Figure 3A). The ARGS-aggrecan antibody (mAb OA-1) detected three distinct protein fragments of aggrecan approximated to be ARGS-CS2, ARGS-CS1 and ARGS-KS. The 3B3 antibody detected the widest spectrum of aggrecan species, including fragments of the sizes of ARGS-CS2 and ARGS-CS1, but showed no, or very week reactivity against fragments around 64 kDa where ARGS-KS migrates. The 1-F21 antibody detected only high molecular weight species of
sizes above 170 kDa, thus likely detecting the ARG5-CS2 species but not the ARG5-CS1 and ARG5-KS
species (Figure 3A).

To further determine the location of the 1-F21 epitope, we made Western blots using samples of aggrecan
which had been in vitro digested with ADAMTS-4 or MMP-3. The 1-F21 antibody detected high molecular
aggrecan fragments of sizes corresponding to ARG5-CS2 and FFGV-CS2 in ADAMTS-4 or MMP-3
digested material, respectively (Figure 3B). However, no reactivity was noted against the corresponding
G1-TEGR and G1-IPEN fragments, or against ARG5-CS1 that is present in the ADAMTS-4 digested
aggrecan sample (Figure 3B). These results suggest that the 1-F21 epitope is located within the CS2
region of aggrecan (Figure 4).

Discussion

This study presents a long-term follow-up of an ACL-injury cohort where patients were treated with knee
rehabilitation without ACL reconstruction and were without definite radiographic signs of radiographic OA
16 years after their injury. We found that in this patient group higher acute synovial fluid concentrations
of large aggrecan fragments detected with the 1-F21 antibody were associated with lower T1Gd values
measured by dGEMRIC 20 years later. None of the other investigated biomarkers measured acutely after
injury or up to 7.5 years after injury were associated with dGEMRIC T1Gd at the follow up. Similar
findings have been observed in rheumatoid arthritis, where subjects with destructive disease (that
required joint replacement) had higher initial levels of 1-F21 aggrecan compared to subjects with non-
destructive disease when evaluated up to 12 years later (47). In accordance with previous studies
evaluating knee injured subjects (27, 30, 31, 36) a slightly higher dGEMRIC value in the lateral than in the
medial femoral cartilage was found also in this study. Medial and lateral dGEMRIC values in this study
were not statistically different from control values in healthy uninjured subjects, indicating a still rather
well preserved knee cartilage (36).

Using an ex vivo biomechanical cartilage injury model culturing explants in the presence of inflammatory
cytokines, Wang et al. showed that large size aggrecan fragments were released from the injured
cartilage momentarily and during the first 14 days (51). Based on a similar cartilage explant model
exposing the cartilage for cyclic loading, Orozco et al. showed a decrease in aggrecan concentration and
presence of chondrocyte death around the cartilage cracks, which was not observed in the intact cartilage
(52). The same authors suggested that the early decrease of aggrecan in cartilage extracellular matrix
following injury and subsequent tissue loading, without the addition of inflammatory drive, might be
caused by the release of aggrecan through the damaged cartilage surface into the synovial cavity by high
pressure fluid outflow. The cartilage leakage of structural proteins such as aggrecan into the synovial
fluid is most likely dependent on the amount of compression and the shear forces on the joint surfaces at
the trauma situation, but also on the quality of the affected cartilage. High quality knee cartilage of well-
trained athletes is densely packed with proteoglycans, and higher synovial fluid concentrations of
proteoglycans were found after an ACL injury in well-trained athletes compared to levels in less well-
trained individuals with ACL injured knees (53). However, in the patients from this cohort we found no
association between the measured molecular biomarkers or T1Gd values and their rather uniform activity levels (data not shown).

Previous reports have suggested that the 1-F21 epitope resides within or close to the KS-region of aggrecan (40). However, since neither the N-terminal fragments G1-TEGE and G1-IPEN, nor ARGS-KS-CS1 or the shorter ARGS-KS fragments were detected by the 1-F21 antibody in the Western blots, the position of the 1-F21 epitope is further distal and most likely resides within the CS2 region (Figure 4).

Using the same assays as herein for the detection of aggrecan fragments in the synovial fluid we have shown that the concentration of 1-F21 aggrecan, ARGS-aggrecan and sGAG were increased directly after a knee injury (18, 20, 46, 54). This increase is most likely caused by the knee trauma and subsequent inflammation as a part of the repair mechanism in the joints during the acute phase after injury (11).

From the Western blot investigation in this study it is evident that there are differences in which aggrecan fragments these three aggrecan assays detect. While the ARGS-aggrecan assay detects specific aggrecanase generated ARGS-fragments, the sGAG and 1-F21 assays detect a variety of similar broad range large aggrecan fragments, concordant with the strong correlation between the sGAG and 1-F21 biomarkers (18). Although there was a strong positive correlation between the aggrecan markers for the acute samples in this study, only 1-F21 aggrecan was associated with dGEMRIC values.

There are limitations in this study. Although the study design planned for repeated sampling of synovial fluid from the injured knee over several years, we do not have a complete set of data from every subject (Table 1). The study cohort is a selected subgroup that managed to cope well with their ACL injury without ACL reconstruction and had no radiographic knee OA at long-term follow-up (i.e. OARSI atlas grades ≤ 1), and the results may thus not be generalizable to all ACL injured subjects. On the other hand, the selection of investigated patients could be an important factor to explain our results in this study. These ACL-injured subjects had few subsequent knee injuries that would blur the association between the magnitude of the first traumatic cartilage injury and dGEMRIC values 20 years later. Other knee injury studies are more variable regarding inclusion, sampling time, age of subjects and highly variable knee pathologies and surgeries which might influence the results from these cohorts (18, 20, 55).

A study showed that cartilage pre-contrast T1 and thickness are sources of variation in dGEMRIC indicating that well-trained elite runners with a thicker deep knee cartilage than sedentary volunteers achieve a higher dGEMRIC value (ms) just because of a thicker cartilage and not due to differences in cartilage structure (56). This might be a limitation with the dGEMRIC method but is probably of less importance in our studied cohort which had a uniform low to medium high activity level.

**Conclusion**

In conclusion, higher synovial fluid concentrations of large aggrecan fragments detected by the 1-F21 antibody early after ACL injury were associated with worse knee cartilage quality estimated by dGEMRIC 20 years later. High synovial fluid concentrations of large sized aggrecan fragments in acutely ACL
injured knees may reflect the magnitude of the acute concomitant knee cartilage trauma, associated with later joint cartilage quality.

**Abbreviations**

OA: osteoarthritis; ACL: anterior cruciate ligament; MRI: magnetic resonance imaging; sGAG: sulfated glycosaminoglycans; dGEMRIC: delayed gadolinium-enhanced MRI of cartilage; OARSI: Osteoarthritis Research Society International; JSN: joint space narrowing; CS: chondroitin sulfate; KS: keratan sulfate; mAb: monoclonal antibody; COMP: cartilage oligomeric matrix protein; MMP-3: matrix metalloproteinase-3; TIMP-1: tissue inhibitor of metalloproteinase-1; ROI: region of interest; ADAMTS-4: a disintegrin and metalloproteinase with thrombospondin motifs-4; SD: standard deviation; ARGS agcan: ARGS neoepitope of aggrecan; IGD: interglobular domain; SF: synovial fluid.

**Declarations**

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**AUTHORS’ CONTRIBUTIONS**

All authors have substantially contributed to either the conception and/or design of the study (PN, SL, LSL, AS), acquisition of data (PN, AS), or analyses and interpretation of data (PN, SL, LSL, AS). All authors have participated in the writing process and approved the final version of the manuscript. Paul Neuman (paul.neuman@skane.se) takes responsibility for the integrity of the work.

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**AVAILABILITY OF DATA AND MATERIALS**

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

**ETHICS APPROVAL AND CONSENT TO PARTICIPATE**
This study was approved by the Lund University Medical Faculty Research Ethics Committee (Dnr 38-1986, LU 506-02). Written informed consent for inclusion in the study was obtained from all patients.

CONSENT TO PUBLISH

Not applicable.

COMPETING INTERESTS

A Struglics is a member of the journal’s editorial board.

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Tables

Table 1. Characteristics of the study subjects with dGEMRIC examination at the 20 years follow-up and available acute and/or chronic synovial fluid samples.

| Time after injury to SF sampling | Age at injury mean (SD) | Men, % | BMI at injury, mean (SD) | BMI 20 years post injury, mean (SD) |
|----------------------------------|------------------------|-------|------------------------|----------------------------------|
| Total study group | 25 | 0 days to 7.5 years | 24.5 (6.2) | 52 | 23.6 (3.0) | 25.3 (3.5) |

| Subjects with SF samples, n | Time after injury |
|-----------------------------|-------------------|
| Acute samples | 20 | 0-18 days (median 6 days) |
| Chronic samples | 22 | 0.5-7.5 years (median 4 years) |

- 4 | 0.5-1.5 years |
- 17 | 1.5-2.5 years |
- 12 | 2.5-3.5 years |
- 11 | 3.5-4.5 years |
- 10 | 4.5-5.5 years |
- 3 | 5.5-6.5 years |
- 1 | 6.5-7.5 years |

1) 25 subjects with any kind of SF-samples (i.e. acute and/or one or more chronic samples): 3 subjects had only acute samples, 5 subjects had only chronic samples and 17 subjects had both acute and chronic samples.
Delayed gadolinium enhanced MRI of cartilage (dGEMRIC) examination: mean = 20.6 years (range = 18 to 23 years) after injury. SF = synovial fluid. SD = standard deviation.

Table 2. Concentration of biomarkers, expressed as median values (25th and 75th percentiles), in acute and chronic samples.

| Biomarkers          | Acute samples          | Chronic samples         | P-values |
|---------------------|------------------------|-------------------------|----------|
|                     | Concentration          | N                       | Concentration | N |<0.001 |
| sGAG, µg/ml         | 181.8 (116.8, 323.8)   | 16                      | 58.1 (42.8, 75.1) | 21 |<0.001 |
| 1-F21 agcan, µg/ml  | 679.0 (413.4, 873.6)   | 16                      | 117.0 (85.6, 168.3) | 14 |<0.001 |
| ARG5 agcan, nM      | 11.5 (6.8, 21.1)       | 18                      | 1.5 (1.1, 2.1)   | 22 |<0.001 |
| COMP, µg/ml         | 180.0 (146.5, 214.5)   | 13                      | 57.0 (52.3, 73.0) | 12 |<0.001 |
| MMP-3, nM           | 37.5 (21.4, 56.8)      | 18                      | 5.2 (1.5, 8.5)   | 17 |<0.001 |
| TIMP-1, nM          | 52.5 (43.1, 67.8)      | 18                      | 7.3 (5.7, 9.3)   | 17 |<0.001 |
| sGAG/COMP           | 1.08 (0.7, 1.6)        | 11                      | 0.9 (0.8, 1.1)   | 12 | 0.295 |
| 1-F21 agcan/COMP    | 4.9 (1.7, 5.7)         | 11                      | 1.5 (1.2, 2.5)   | 9  | 0.038 |
| ARG5 agcan/COMP     | 0.07 (0.04, 0.10)      | 12                      | 0.02 (0.02, 0.03)| 12 | 0.002 |
| MMP-3/TIMP-1        | 0.7 (0.4, 1.1)         | 18                      | 0.6 (0.3, 1.0)   | 17 | 0.642 |

1) Statistical analyses using Mann Whitney.
sGAG = sulfated glycosaminoglycans, 1-F21 agcan = 1-F21 epitope of aggrecan, ARG5 agcan = ARG5 neoepitope of aggrecan, COMP = cartilage oligomeric matrix protein, MMP-3 = matrix metalloproteinase 3, TIMP-1 = tissue inhibitor of metalloproteinase 1.
Figure 1

(a) Flow diagram of study subjects. (b) Timeline showing synovial fluid sampling and imaging and arthroscopic acquisitions. The 16-year x-ray examinations were done between 11 and 18 years after the ACL injury, while the 20-year dGEMRIC assessments were done 18 to 23 years after injury.
Figure 2

Adjusted linear regression analyses between molecular biomarkers and dGEMRIC. Molecular biomarkers in acute and chronic synovial fluid samples were used as prognostic variables for cartilage quality assessed by dGEMRIC 20 years post ACL injury. Squares: mean effect with size being proportional to number of available biomarker data. Grey area: highlights statistical significance with an alpha level of 0.05. Standardized effect: the estimate of the average change in dGEMRIC T1Gd (expressed as standard deviation) that corresponds to a 1 standard deviation change in the prognostic factor. 1-F21 agcan = 1-F21 epitope of aggrecan, ARGs agcan = ARGs neoepitope of aggrecan, COMP = cartilage oligomeric matrix protein, MMP-3 = matrix metalloproteinase 3, sGAG = sulfated glycosaminoglycans, TIMP-1 = tissue inhibitor of metalloproteinase 1. dGEMRIC medial + lateral = the sum of medial and lateral dGEMRIC values divided by 2.
Figure 3

Western blot of synovial fluid and cartilage samples. (A) Synovial fluid A1 and D1 samples on membranes probed with antibodies against 6-sulfated chondroitin sulfate stubs (3B3), aggrecan epitope 1-F21 and ARG5-aggrecan. (B) ADAMTS-4 or MMP-3 in vitro digested cartilage A1D1 aggrecan samples on membranes probed with antibodies against aggrecan epitope 1-F21 and G1-domain of aggrecan. The position of Mw markers (left side) and the immunobands are indicated. The images are from different experiments showing representative signals. The original images from full size blotted gels are shown in Figure S2. Keratan sulfate region (KS), chondroitin sulfate region (CS) and globular domains (G1, G2 and G3) are illustrated in Figure 4. One to three µg sGAG was loaded per well. IGD = interglobular domain.
Figure 4

Schematic figure of aggrecan showing MMP (IPEN/FFGV) and aggrecanase (TEGE/ARGS) cleavage sites in the inter-globular domain (IGD). The amino acid numberings are based on the full-length human aggrecan amino acid sequence starting with the N-terminus 1MTTL and finishing with the C-terminus STAH2415 (NCBI accession no. P16112). The positions for recognition of 3B3 and aggrecan 1-F21 antibodies are shown by dashed lines. IGD = interglobular domain; KS = keratan sulfate region; CS = chondroitin sulfate region; G = globular domains.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- Additionalfile1TableS1.pdf
- Additionalfile2FigureS1.pdf
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