Despite Predominance of Uropathogenic/Extraintestinal Pathotypes Among Travel-acquired Extended-spectrum - Lactamase–producing Escherichia coli, the Most Commonly Associated Clinical Manifestation Is Travelers’ Diarrhea

Anu Kantele, Tinja Lääveri, Sointu Mero, Inka M K Häkkinen, Juha Kirveskari, Brian D Johnston, James R Johnson
Despite Predominance of Uropathogenic/Extraintestinal Pathotypes Among Travel-acquired Extended-spectrum β-Lactamase–producing Escherichia coli, the Most Commonly Associated Clinical Manifestation Is Travelers’ Diarrhea

Anu Kantele,1,2 Tinja Lääveri,3 Sointu Mero,1 Inka M. K. Häkkinen,1 Juha Kirveskari,3 Brian D. Johnston,4 and James R. Johnson4

1Inflammation Center, Infectious Diseases, Helsinki University Hospital and University of Helsinki, 2Aava Travel Clinic, Medical Centre Aava, and 3Helsinki University Hospital Laboratory, Bacteriology, Finland, and 4Infectious Diseases, Veterans Affairs Medical Center, Minneapolis, Minnesota

**Background.** One-third of the 100 million travelers to the tropics annually acquire extended-spectrum β-lactamase (ESBL)-producing Enterobacteriaceae (ESBL-PE), with undefined clinical consequences.

**Methods.** Symptoms suggesting Enterobacteriaceae infections were recorded prospectively among 430 Finnish travelers, 90 (21%) of whom acquired ESBL-PE abroad. ESBL-PE isolates underwent polymerase chain reaction–based detection of diarrheagenic Escherichia coli (DEC) pathotypes (enteroaggregative E. coli [EAEC], enteropathogenic E. coli [EPEC], enterotoxigenic E. coli [ETEC], enteroinvasive E. coli, and Shiga toxin–producing E. coli), and extraintestinal pathogenic/uropathogenic E. coli (ExPEC/UPEC). Laboratory-confirmed ESBL-PE infections were surveyed 5 years before and after travel.

**Results.** Among the 90 ESBL-PE carriers, manifestations of Enterobacteriaceae infection included travelers’ diarrhea (TD) (75/90 subjects) and urinary tract infection (UTI) (3/90). The carriers had 96 ESBL-producing E. coli isolates, 51% exhibiting a molecular pathotype: 13 (14%) were DEC (10 EAEC, 2 EPEC, 1 ETEC) (12 associated with TD) and 39 (41%) ExPEC/UPEC (none associated with UTI). Of ESBL-PE, 3 (3%) were ExPEC/UPEC-EAEC hybrids (2 associated with diarrhea, none with UTI). Potential ESBL-PE infections were detected in 15 of 90 subjects (17%). The 10-year medical record survey identified 4 laboratory-confirmed ESBL-PE infections among the 430 travelers, all in subjects who screened ESBL-PE negative after returning home from their index journeys but had traveled abroad before their infection episodes.

**Conclusions.** Half of all travel-acquired ESBL-producing E. coli strains qualified molecularly as pathogens. Extraintestinal and uropathogenic pathotypes outnumbered enteric pathotypes (41% vs 14%), yet the latter correlated more closely with symptomatic infection (0% vs 92%). Despite more ESBL-PE strains qualifying as ExPEC/UPEC than DEC, travel-acquired ESBL-PE are more often associated with TD than UTI.

**Keywords.** extended-spectrum beta-lactamase; ESBL; travel; DEC; ExPEC.

International travel drives the global spread of antimicrobial-resistant organisms from high- to low-risk regions [1]. Visitors to the (sub)tropics have been well documented to acquire multidrug-resistant (MDR) intestinal bacteria: 20%–70% carry extended-spectrum β-lactamase (ESBL)–producing Enterobacteriaceae (ESBL-PE) at return [2–10].

Colonizing intestinal MDR bacteria may have considerable implications for both the community and colonized individuals. Such strains can spread silently to close contacts within a community, in healthcare settings, and eventually even worldwide [11]. Colonized travelers usually remain asymptomatic [1], but can develop severe, even life-threatening MDR infections [12, 13]. Infections with MDR strains result in greater morbidity, mortality, and economic losses than those caused by susceptible strains [12]. Although one report suggested that imported ESBL-PE strains have low pathogenicity [14], no prospective study has assessed the clinical infection risk such strains pose.

In studies exploring clinical ESBL-PE isolates, urinary tract infection (UTI) has appeared to be the most common type of infection caused by travel-acquired MDR bacteria [15]. In a recent study, 9% of travel-acquired ESBL-PE strains carried UTI-associated virulence factors; data on possible related symptoms...
of cystitis were not reported [14]. Additionally, retrospective studies have identified international travel as a risk factor for UTI caused by ESBL-PE [15, 16].

The most common health problem travelers encounter is travelers’ diarrhea (TD) [17]. In numerous recent studies utilizing multiplex quantitative polymerase chain reaction (qPCR) assays, the various diarrheagenic *Escherichia coli* (DEC) pathotypes have been the most frequently detected stool pathogens in TD; multiple pathogens often co-occur [18, 19]. Although DEC and other enteric pathogens are also found among asymptomatic travelers, the prevalence is higher among those with TD [18, 19]. Many studies have explored acquisition of ESBL-PE during travel [1–10, 20], and others have examined TD pathogens [18, 19] and their resistance profiles [21, 22]. However, none have investigated whether some travel-acquired ESBL-PE isolates actually are TD pathogens, as might be expected, given the rising prevalence of antibiotic resistance, including ESBL production, among TD pathogens [23].

Here, we studied the clinical significance of travel-acquired ESBL-PE. Specifically, we prospectively surveyed Finnish international travelers for symptoms suggesting Enterobacteriaceae infections (eg, UTI and TD), screened their feces for travel-acquired ESBL-PE, determined whether the colonizing ESBL-PE strains qualified molecularly as DEC or extraintestinal pathogenic/uropathogenic *E. coli* (ExPEC/UPEC), and compared clinical data with molecular data. We also reviewed subjects’ medical records for clinical ESBL-PE infections during the 5 years before and after the index travel episode.

**MATERIALS AND METHODS**

**Study Design, Volunteers, Samples, and Travel Destinations**

As reported elsewhere, 430 incipient travelers were recruited prospectively at the Travel Clinic of Aava Medical Centre [7] prior to visiting the tropics. Each subject provided a pre- and posttravel stool sample to be screened for ESBL-PE, completed a questionnaire before and after their journey, and kept a diary while abroad. The Helsinki University Hospital ethics committee approved the protocol. All subjects provided written informed consent.

Subjects whose posttravel but not pretravel stools yielded ESBL-PE were considered to have travel-acquired ESBL-PE. Subjects with vs without travel-acquired ESBL-PE constituted the ESBL⁺ and ESBL⁻ groups, respectively. We have previously reported risk factors for acquiring ESBL-PE [7]; all subjects but 1 were the same in both studies.

To identify all possible symptomatic ESBL-PE infections experienced during travel, we searched for symptoms suggesting Enterobacteriaceae infections (described below), then investigated whether subjects with such symptoms had travel-acquired ESBL-PE, and whether the strains carried pathotype-defining markers for DEC, ExPEC, or UPEC. A theoretical maximum rate of infections caused by travel-acquired ESBL-PE was calculated. Case criteria were as follows: (1) symptoms of an Enterobacteriaceae infection; (2) acquisition of ESBL-PE during travel; and (3) for TD, the travel-acquired ESBL-PE strain qualified as DEC. For UTI and other Enterobacteriaceae infections, no pathotype restrictions applied, since non-ExPEC/UPEC also can cause UTI. Despite uncertain causality, this approach should identify the maximum rate of infections caused by travel-acquired ESBL-PE (Figure 1).

In addition to exploring ESBL-PE infections during travel, we sought for relevant results in the subjects’ medical records. To assess whether the index travel increased the risk of clinical ESBL-PE infections, the search included 5 years before index travel and 5 years after it.

**Symptoms of Potential Enterobacteriaceae Infections**

As noted above, the posttravel questionnaire and diaries captured whether while abroad the subject had experienced symptoms suggesting Enterobacteriaceae infection (eg, TD or UTI). TD was defined as passage of ≥3 loose or liquid stools per day, or more frequently than normal for the individual [24]. Cystitis was defined as dysuria and/or frequency and/or urgency [25], and pyelonephritis as a febrile illness (with or without urinary symptoms) without diarrhea or respiratory symptoms or other obvious cause of fever.

**Molecular Identification of Diarrheal and Extraintestinal Pathogens**

As detailed previously, fecal ESBL-PE were isolated and characterized using established methods with culture on chromID ESBL (bioMérieux, Marcy-l’Étoile, France), followed by double-disk synergy (Oxoid, Thermo Fisher Scientific, Hampshire, United Kingdom) test for cefotaxime (30 µg), ceftazidime (30 µg), and cefpodoxime (30 µg), alone or with clavulanic acid (10 µg), and species identification by Vitek GN (bioMérieux) [7]. Susceptibility testing for ciprofloxacin, cotrimoxazole, nitrofurantoin, tobramycin, ertapenem, imipenem, and meropenem was performed with Etest (bioMérieux) according to criteria set by the European Committee on Antimicrobial Susceptibility Testing 5.0 (2018; www.eucast.org). β-Lactamase genes (TEM, OXA, SHV, CTX-M) and plasmid-mediated AmpC β-lactamase genes (DHA, CIT) were identified with multiplex PCR [26].

To identify DEC, isolates underwent multiplex qPCR screening for 5 DEC pathotypes—enteroaggregative *E. coli* (EAEC), enteropathogenic *E. coli* (EPEC), enterotoxigenic *E. coli* (ETEC), enteroinvasive *E. coli* (EIEC), and Shiga toxin-producing *E. coli* (STEC)—by a method described previously in detail [27]. To identify ExPEC and UPEC, isolates underwent multiplex PCR screening for pathotype-defining extraintestinal virulence genes [28]. Isolates were designated as ExPEC if positive for ≥2 of *pap*AH and/or *pap*C (*P. fimbriae*), *sfa*/*foc* (S and
F1C fimbriae), *afa/dra* (Dr-binding adhesins), *iutA* (aerobactin system), and *kpsM II* (group 2 capsule), and as UPEC if positive for ≥2 of *vat* (vacuolating toxin), *chuA* (heme uptake), *fyuA* (yersiniabactin system), and *yfcV* (adhesin). Isolates could exhibit multiple pathotypes (Table 1).

Major *E. coli* phylogenetic group was defined by multiplex PCR [29], and sequence type complex (STc; group of closely related sequence types [STs]) by a combination of STc-specific PCR assays and *fumC/fimH* typing [30].

### Additional Medical Record Screening for Severe Extraintestinal ESBL-PE Infections

To identify laboratory-confirmed ESBL-PE infections for 5 years both before and after travel, we surveyed the subjects’ laboratory records for ESBL-PE clinical isolates between 1 March 2004 and 30 February 2015, and interviewed each patient with positive findings for additional travel history. The Helsinki University Hospital medical records include results from the Helsinki University Hospital Laboratory (HUSLAB), which serves all public healthcare institutions in the Uusimaa region (1.6 million inhabitants). In Finland, nearly all patients who are severely ill or need blood cultures are treated in public hospitals. Since 2010, patients hospitalized outside the Nordic countries within the prior year have been screened for MDR organisms upon hospital admission.

### Statistical Methods

Statistical analyses were performed with SPSS software version 22.0 (SPSS Inc, Armonk, New York). The χ² test or Fisher exact test was used to compare categorical variables.
For comparison, in the ESBL− group, 213 of 340 (63%) subjects (specifically, cystitis), the latter including 2 of 49 (4%) females. Of these subjects, 75 of 90 (83%) had TD and 3 of 90 (3%) had UTI. The remaining 340 (79%) subjects constituted the ESBL+ group. Of these, 3 non–ESBL-PE–positive clinical sample (2, urine; 1, peritoneal fluid) were collected before study participation. These patients had visited Northern Africa (2 patients) and South Asia (1 patient) within 3 months before the ESBL-PE infection. The fourth developed ESBL-PE bacteremia and peritonitis (related to appendiceal perforation) 4 years after study participation, and 2 months after visiting sub-Saharan Africa.

## Table 1. Pathotypes of 96 Travel-acquired Extended-spectrum β-Lactamase–producing Escherichia coli Strains

| Category and Pathotype(s) | No. (% of 96) |
|---------------------------|---------------|
| DEC only (ie, not ExPEC/UPEC) | 10 (10) |
| EAEC | 7 (7) |
| EPEC | 2 (2) |
| ETEC | 1 (1) |
| ExPEC/UPEC only (ie, not DEC) | 36 (38) |
| ExPEC, +/- UPEC | 22 (23) |
| UPEC, +/- ExPEC | 34 (35) |
| ExPEC, not UPEC | 2 (2) |
| UPEC, not ExPEC | 14 (14) |
| ExPEC + UPEC | 20 (21) |
| DEC + ExPEC/UPEC | 49 (51) |
| DEC, +/- UPEC/ExPEC | 13 (13) |
| ExPEC/ExPEC, +/- DEC | 39 (40) |
| ExPEC, +/- DEC | 23 (24) |
| UPEC, +/- DEC | 37 (38) |
| No pathotype detected | 47 (49) |

Abbreviations: +/-, with/without; DEC, diarrheagenic Escherichia coli; EAEC, enteropathogenic Escherichia coli; EPEC, enteropathogenic Escherichia coli; ETEC, enterotoxigenic Escherichia coli; ExPEC, extraintestinal pathogenic Escherichia coli; UPEC, uropathogenic Escherichia coli.

Five DEC pathotypes (enteroaggregative, enteropathogenic, enterotoxigenic, enteroinvasive, and Shiga toxin–producing E. coli).

### RESULTS

#### Description of Travelers, Travels, and Travel-acquired ESBL-PE

The demographics, destinations, and ESBL-PE colonization status of the 430 prospectively recruited international travelers have been reported previously [7]. The 90 (21%) with travel-acquired ESBL-PE constituted the ESBL+ group. Of these, 61% were females and 66% were aged <50 years; the median age was 36.5 years (interquartile range [IQR], 28–58 years), and the median journey duration was 16.5 days (IQR, 12–27 days) (Table 2). The 90 posttravel stools yielded 99 travel-acquired ESBL-PE strains, of which 96 were E. coli, 2 Klebsiella oxytoca, and 1 Escherichia hermannii. The 3 non–E. coli isolates were from separate subjects, each also with ESBL-producing E. coli. The remaining 340 (79%) subjects constituted the ESBL− group.

#### Symptoms of Enterobacteriaceae Infection

The only symptoms suggesting Enterobacteriaceae infection reported by ESBL+ group members were those of TD and UTI. Of these subjects, 75 of 90 (83%) had TD and 3 of 90 (3%) had UTI (specifically, cystitis), the latter including 2 of 49 (4%) females. For comparison, in the ESBL− group, 213 of 340 (63%) subjects had TD and 13 of 340 (4%) had UTI.

### Diarrheal Pathotypes

Of the 96 ESBL-producing E. coli isolates, 13 (14%) qualified as DEC, including 10 EAEC, 2 EPEC, and 1 ETEC (no STEC or EIEC), each from a different subject (Table 1). Of the 13 subjects with ESBL-producing DEC, 12 (92%) had experienced TD while abroad. Eight (62%) had visited South Asia, for which the DEC fraction among ESBL-PE isolates was 8 of 29 (28%), vs 5 of 67 (7%) for all other regions combined (P = .02) (Table 2).

### Extraintestinal and Combined Pathotypes

Of the 96 ESBL-producing E. coli fecal isolates, 39 (41%) qualified as ExPEC and/or UPEC, that is, presumptive extraintestinal pathogens (Table 1). Of these, 3 (8%) carried markers of combined intestinal and extraintestinal pathotypes (2 EAEC + ExPEC, 1 EAEC + ExPEC + UPEC).

### Antibiotic Resistance Profiles

Among the 96 isolates, resistance was common to tobramycin, ciprofloxacin, and cotrimoxazole, but rare or absent to carbapenems and nitrofurantoin (Table 3). Resistance prevalence, which varied significantly by pathotype for tobramycin, ciprofloxacin, and cotrimoxazole, was lowest among DEC isolates, intermediate among pathotype-negative isolates, and highest among ExPEC/UPEC-only isolates (P < .01 for all comparisons of DEC isolates vs ExPEC/UPEC-only isolates).

### Phylogroups

Phylogroup distribution and extraintestinal virulence gene content varied significantly by pathotype (Table 4). Most non-ExPEC/UPEC isolates belonged to phylogroups A and B1, whereas nearly all ExPEC/UPEC isolates belonged to groups D and F. Likewise, STc10 predominated among non-ExPEC/UPEC isolates, whereas STc38, STc405, and STc648 predominated among ExPEC/UPEC isolates. Consistent with the virulence gene–based criteria for ExPEC and UPEC, all extraintestinal virulence genes studied were twice or more as prevalent among ExPEC/UPEC-only isolates than non-ExPEC/UPEC isolates.

### Clinical ESBL-PE Isolates From Medical Record Review

Our 10-year medical record review (5 years before and after travel) identified 4 ESBL-PE isolates in clinical samples collected from a presumed focus of infection during routine medical care. At the time of the present study, however, the corresponding 4 subjects all fell within the ESBL+ group. For 3, the ESBL-PE–positive clinical sample (2, urine; 1, perianal abscess) had been collected before study participation. These patients had visited Northern Africa (2 patients) and South Asia (1 patient) within 3 months before the ESBL-PE infection. The fourth developed ESBL-PE bacteremia and peritonitis (related to appendiceal perforation) 4 years after study participation, and 2 months after visiting sub-Saharan Africa.
Additionally, during the 5-year follow-up, routine stool screening cultures upon hospital admission detected ESBL-PE colonization in 3 subjects (2 patients, ESBL− group; 1 patient, ESBL+ group). All 3 had recently revisited the tropics.

**DISCUSSION**

Despite growing concern about the surge of antimicrobial resistance among clinical isolates, many clinicians overlook the potential risks asymptomatic intestinal colonization by ESBL-PE.

**Table 2. Demographics, Destinations, and Symptoms in Relation to Pathotype Among 90 International Travelers**

| Host Characteristic | Subjects<sup>ab</sup> | ESBL-producing E. coli Strains<sup>c</sup> | Not DEC or ExPEC/UPEC<sup>c</sup> | DEC Only<sup>c</sup> | ExPEC/UPEC Only<sup>c</sup> | DEC and ExPEC/UPEC<sup>c</sup> | 4-Group Comparison | P Value |
|---------------------|-----------------------|------------------------------------------|-------------------------------|-----------------------|---------------------------------|-------------------------------|----------------|---------|
| Total               | 90 (100)              | 96 (100)                                 | 47 (49)                       | 10 (11)               | 36 (37)                         | 3 (3)                         | NA             |         |
| Female gender       | 49 (54)               | 54 (56)                                 | 24 (51)                       | 5 (50)                | 24 (47)                         | 1 (33)                        | NA             |         |
| Age <50 y           | 59 (66)               | 64 (67)                                 | 29 (62)                       | 7 (70)                | 2 (67)                          | 24 (73)                       | NA             |         |
| Travel destination  |                       |                                          |                               |                       |                                 |                               | NA             |         |
| South Asia          | 28 (31)               | 29 (30)                                 | 12 (26)                       | 8 (80)                | 9 (25)                          | 0 (0)                         | NA             |         |
| Southeast Asia      | 33 (37)               | 37 (38)                                 | 18 (38)                       | 1 (10)                | 16 (44)                         | 2 (67)                        | NA             |         |
| Sub-Saharan Africa  | 23 (26)               | 24 (25)                                 | 14 (29)                       | 0 (0)                 | 10 (28)                         | 0 (0)                         | NA             |         |
| East Asia           | 2 (2)                 | 2 (2)                                   | 0 (0)                         | 1 (10)                | 1 (3)                           | 0 (0)                         | NA             |         |
| North Africa and Middle East | 4 (4) | 4 (4) | 3 (6) | 0 (0) | 3 (6) | 0 (0) | 1 (33) |         |
| TD during travel<sup>de</sup> | 75 (83) | 81 (84) | 36 (77) | 10 (100) | 33 (92) | 2 (67) | NA |         |
| UTI during travel   | 3 (3)                 | 3 (3)                                   | 3 (6)                         | 0 (0)                 | 0 (0)                           | 0 (0)                         | NA             |         |
| Antibiotic use during travel<sup>e</sup> | 28 (31) | 32 (34) | 15 (33) | 2 (20) | 15 (42) | 0 (0) | NA |         |

Data are presented as no. (%). Abbreviations: DEC, diarrheagenic *Escherichia coli*; ESBL-PE, extended-spectrum β-lactamase producing Enterobacteriaceae; ExPEC, extraintestinal pathogenic *Escherichia coli*; TD, travelers’ diarrhea; UPEC, uropathogenic *Escherichia coli*; UTI, urinary tract infection.

aThe 90 volunteers acquired a total of 99 ESBL-producing Enterobacteriaceae strains, of which 96 were *E. coli*, 1 was *Escherichia hermannii*, and 2 were *Klebsiella oxytoca*. Each of the carriers of the latter 3 strains also had an ESBL-producing *E. coli* strain. Six volunteers had 2 different ESBL-producing *E. coli* strains.

bData are given per subject.

cData are shown per strain.

dTypical of TD, many volunteers had concomitant stool pathogens other than the ESBL/DEC shown here.

eInformation missing for 1 traveler who had 2 ESBL-PE strains.

**Table 3. Antibiotic Resistance Profile in Relation to Pathotype Among Extended-spectrum β-Lactamase–producing *Escherichia coli* Strains Acquired During International Travel**

| Resistance Genotype<sup>ab</sup> or Phenotype<sup>c</sup> | Trait Prevalence by Pathotype Category | Total | Not DEC or ExPEC/UPEC | DEC Only | ExPEC/UPEC Only | DEC + ExPEC/UPEC | 4-Group Comparison | P Value |
|-----------------------------------------------------------|--------------------------------------|-------|-----------------------|----------|-----------------|-----------------|-------------------|---------|
| Total                                                     | 96 (100)                             | 47 (49) | 10 (11)               | 36 (37)  | 3 (3)           | NA              | NA                |         |
| Resistance genes                                         |                                      |       |                       |          |                 |                 |                   |         |
| TEM                                                       | 48 (50)                              | 25 (53) | 3 (30)                | 18 (50)  | 2 (67)          | .55             |                   |         |
| OXA                                                      | 22 (23)                              | 11 (23) | 0 (0)                 | 11 (31)  | 0 (0)           | .17             |                   |         |
| SHV                                                      | 4 (4)                                | 2 (4)  | 1 (10)                | 1 (3)    | 0 (0)           | .78             |                   |         |
| CTX-M group 1                                             | 67 (70)                              | 30 (64) | 8 (80)                | 28 (78)  | 1 (33)          | .23             |                   |         |
| CTX-M group 9                                             | 26 (27)                              | 13 (28) | 1 (10)                | 10 (28)  | 2 (67)          | .28             |                   |         |
| Resistant or intermediate to antibiotic                  |                                      |       |                       |          |                 |                 |                   |         |
| Carbapenem                                               | 0 (0)                                | 0 (0)  | 0 (0)                 | 0 (0)    | 0 (0)           | NA              |                   |         |
| Tobramycin<sup>d</sup>                                   | 49 (51)                              | 23 (49) | 2 (20)                | 24 (67)  | 0 (0)           | .042<sup>d</sup> |                   |         |
| Ciprofloxacin                                            | 64 (67)                              | 33 (70) | 3 (30)                | 28 (78)  | 0 (0)           | .003<sup>d</sup> |                   |         |
| Nitrofurantoin                                           | 2 (2)                                | 2 (4)  | 0 (0)                 | 0 (0)    | 0 (0)           | .55             |                   |         |
| Cotrimoxazole                                            | 71 (74)                              | 32 (68) | 5 (50)                | 31 (86)  | 1 (33)          | .054<sup>d</sup> |                   |         |

Data are presented as no. (%) unless otherwise indicated. Abbreviations: DEC, diarrheagenic *Escherichia coli*; ExPEC, extraintestinal pathogenic *Escherichia coli*; NA, not applicable; TD, travelers’ diarrhea; UPEC, uropathogenic *Escherichia coli*.

aData are given per strain.

bNo strains carried CTX-M group 2, 8, or 25; 1 “DEC only” strain carried DHA; 2 ESBL-PE strains with a negative pathotype analysis; and 1 each “UPEC only” and “ExPEC + UPEC” strains carried CIT.

cIsolates testing as resistant or intermediate were recorded as resistant.

dFor DEC-only and DEC + ExPEC/UPEC strains combined vs ExPEC/UPEC-only strains: for tobramycin (2/13 vs 24/36), P = .003; for ciprofloxacin (3/13 vs 28/36), P = .001; and for cotrimoxazole (8/13 vs 31/36), P = .008 (Fisher exact test, 2-tailed).

eTobramycin data missing for 1 isolate.
may pose to travelers [31]. Our finding that a large proportion of imported ESBL-PE strains qualify molecularly as presumptive intestinal or extraintestinal pathogens suggests that more travel-acquired ESBL-PE may cause infections than has been recognized previously.

We found that 13 of 90 (14%) of the travel-acquired ESBL-PE strains were DEC, usually EAEC, a known cause of TD [18, 19, 32]. Most ESBL-producing EAEC strains originated in South Asia, which has the highest rates reported for both TD [17] and ESBL-PE acquisition [1], and here had the highest proportion of DEC among travel-acquired ESBL-PE strains (28% vs 7% for other regions combined).

Recent research on TD has documented stool pathogens among both symptomatic and asymptomatic travelers [18, 19], rendering causality ambiguous in individual cases. However, at the population level, in appropriately controlled studies, EAEC [18, 19, 32] and other intestinal pathogens [18, 19] have been more prevalent among symptomatic than asymptomatic travelers. Here, 12 of 13 (92%) volunteers with ESBL-producing DEC had TD, although (as is typical of TD) 10 of 13 had other potentially contributory stool pathogens (Supplementary Table 1). Thus, although the ESBL-producing DEC strain may not have accounted for the symptoms in all 12 TD cases, it probably contributed to some, especially the 3 in which ESBL-producing EAEC was the sole pathogen detected. Indeed, we have previously shown, within the same cohort, in a 382-subject subgroup without antibiotic use, DEC strains to be significantly associated with TD (for EAEC: odds ratio [OR], 2.6 [95% confidence interval {CI}, 1.5–4.4]); for ETEC: OR, 5.9 [95% CI, 2.9–11.9]; and for EPEC: OR, 1.7 [95% CI, 1.0–2.7]) [19].

Others have also recently identified ESBL production by DEC strains [33]. Given the increasing prevalence of antimicrobial resistance among diarrheagenic pathogens [21, 22, 34, 35], this phenotype strikes us as especially worrying, since it further narrows treatment options if antimicrobial therapy is indicated.

Almost half (41%) of all travel-acquired ESBL-producing E. coli exhibited the ExPEC and/or UPEC pathotype, suggesting an increased potential for causing extraintestinal disease. However, among ESBL* group subjects, the only documented extraintestinal infections typical of Enterobacteriaceae were 3 cases of cystitis, all in subjects whose ESBL-producing strain was not ExPEC/UPEC. Allowing for the possibility that these subjects’ non-ExPEC/UPEC ESBL-PE strains caused the cystitis episode, our data suggest a 3% (3/90) maximum incidence for extraintestinal infections due to travel-acquired ESBL-producing E. coli. Of note, this low rate appears...
applicable only to healthy travelers, since in our recent study of ill subjects hospitalized abroad, up to 11% of travelers carrying a resistant strain (either ESBL- or carbapenemase-producing Enterobacteriaceae) had an extraintestinal infection recorded [36].

In previous reports, most carriers of travel-acquired ESBL-PE strains have remained asymptomatic [1]; symptomatic/clinical ESBL-PE infections have been considered rare [7,14], with UTI as the most common clinical manifestation [15,37]. Our data suggest a theoretical maximum rate of 17% for any symptomatic ESBL-PE infection during travel (TD: 12 [13.3%]; UTI: 3 [3.3%]) among our 90 travelers with travel-acquired ESBL-PE.

We recognize we may have missed some cases in which ESBL-producing DEC caused TD but were no longer detected upon the subjects’ return to Finland, since travel-acquired ESBL-PE sometimes disappear during travel (A. Kantele, unpublished observation). This may be more likely for certain DEC types than others; for example, EAEC apparently persists longer than ETEC [18]. Likewise, because the posttravel stools were collected soon after return, we may have failed to capture some nonsevere cystitis and TD episodes, if symptoms developed only after stool sampling (ie, we may have detected the incipient pathogen during its latent/incubation phase, unaware that it later caused an infection). With respect to TD, since 12 of 13 subjects with ESBL-DEC had already reported TD during travel, data on (later-occurring) posttravel symptoms could theoretically have added at the most only the single subject who had not reported TD during travel.

Our theoretical maximum rate of 12 TD and 3 UTI cases caused by travel-acquired ESBL-PE among 90 colonized subjects contradicts the common view that UTI is the most frequent clinical consequence of such colonization [15]. Indeed, although this is evidently the case in developed countries [37], our results—a novel observation—suggest that for travelers visiting tropical regions, TD outnumbers UTI as a clinical correlate of travel-acquired ESBL-PE. This appears logical as TD pathogens directly reach their target—ie, the intestinal mucosa—while the pathogenesis of UTI requires that the colonizing strains access the urinary tract.

All 3 strains with overlapping intestinal and extraintestinal pathotype qualified as EAEC, an established association [38,39]. Of note, we have not encountered an overlap between ExPEC/UPEC and other DEC pathotypes than EAEC. Although the selective forces that favor such EAEC-ExPEC/UPEC hybrid strains remain undefined, EAEC-related adhesins may confer both uroepithelial and enterocyte adherence, and EAEC can cause UTI [32].

The observed phylogenetic segregation of colonizing DEC strains (groups A and B1; STc10 vs ExPEC/UPEC strains (groups D and F; STc38, STc405, STc648) accords with patterns identified among clinical isolates [40]. It supports the concept that these divergent pathotypes are mainly derived from distinct E. coli lineages, with the gut microbiota representing the (shared) primary reservoir.

Our 10-year medical record survey (spanning 5 pretravel and 5 posttravel years) was unlikely to have missed many severe posttravel infections treated at other institutions, since these medical records cover approximately 90% of all ESBL-PE isolates from clinical samples in the Uusimaa region (Martti Vaara, HUSLAB, personal communication). Furthermore, the 5-year posttravel survey window should suffice because of the transience of ESBL-PE colonization (median duration, 3 months), with a 1-year posttravel prevalence of only approximately 10% [10]. The laboratory records would, however, have missed mild diseases such as cystitis, which in most instances is treated empirically per international recommendations, without culture [25]. Accordingly, cystitis cases were reliably captured only during travel, by our prospective symptom surveillance.

Our medical record survey identified no more than 4 confirmed ESBL-PE infections unrelated to the index travel, all (paradoxically) among subjects in the ESBL − group. All 4 had traveled to a high-risk tropical area within 3 months before their clinical ESBL-PE infection, suggesting that they had contracted the causative strain during that journey. These data exemplify nicely the dynamic and paradoxical nature of the situation. Specifically: (1) each journey to an endemic area poses an appreciable risk of contracting ESBL-PE; (2) half of such strains are pathogenic E. coli (ExPEC/UPEC > DEC); (3) ESBL-PE colonization, although transient, confers a nonzero risk of clinical infection (TD more likely than extraintestinal infection); and (4) new travel involves new exposure and, thus, renewed risk of colonization and, possibly, infection.

CONCLUSIONS

In 13 of 90 (14%) returning Finnish travelers with travel-acquired ESBL-PE, the strains were DEC (mainly EAEC), with a high associated TD attack rate (92%). By contrast, in 40% the strains represent ExPEC and/or UPEC, yet with no associated clinical disease (extraintestinal infection attack rate, 0%). The only potential index travel–associated extraintestinal ESBL-PE infections were 3 cases of cystitis, involving subjects whose travel-acquired ESBL-producing E. coli strain was neither ExPEC/UPEC nor DEC. Thus, among travel-acquired ESBL-PE strains, the most frequent pathotype is ExPEC/UPEC, yet pathotype-consistent clinical manifestations are more common among hosts colonized by DEC than ExPEC/UPEC strains. The maximum total rate of ESBL-PE infections among colonized individuals was estimated at 17%; instead of UTI, the most commonly associated clinical manifestation of travel-acquired ESBL-PE was diarrhea.

Supplementary Data

Supplementary materials are available at Clinical Infectious Diseases online. Consisting of data provided by the authors to benefit the reader, the
posted materials are not copyrighted and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

Notes
Acknowledgments. The authors express a debt of gratitude to the late Dr Jukka Ruutta for recruiting the volunteers. The authors also thank the nurses at the Travel Clinic of Aava Medical Centre for their help; the personnel of Helsinki University Hospital Laboratory for assistance with the stool specimens; and Jukka Ollgren, MSc, for expert help with the statistical analyses.

Disclaimer. The funding sources had no involvement in study design, data collection, analysis, interpretation of data, writing of the report, and decision to submit the manuscript for publication.

Financial support. The work was supported by the Finnish Governmental Subsidy for Health Science Research; the Scandinavian Society for Antimicrobial Chemotherapy Foundation; the Paulo Foundation; the Sigrid Jusélius Foundation; and the Finnish Cultural Foundation. This work also was supported in part by the Office of Research Development, US Department of Veterans Affairs (grant numbers 1 I01 CX000920-01 and 2 I01 CX000920-04 to J. R. J.).

Potential conflicts of interest. A. K. has received honoraria for lectures from Valneva and Immunor, and investigator-initiated grants from Pfizer and Valneva. J. R. J. has had research contracts or consultancies with Achaogen, Allergan, Crucell/Janssen, IDGenomics, Merck, Shionogi, Syntiron, Melinta, and Tetraphase, and holds patent applications for tests to detect E. coli strains. J. K. is an employee of Mobidiag, which develops diagnostic tests for infectious diseases, none of which were used in the study. All other authors report no potential conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

References
1. Woerther PL, Andremont A, Kantele A. Travel-acquired ESBL-producing Enterobacteriaceae: impact of colonization at individual and community level. J Travel Med 2017; 24:29–34.
2. Tangdén T, Cars O, Melhus A, Löwdin E. Foreign travel is a major risk factor for colonization with Escherichia coli producing CTX-M-type extended-spectrum beta-lactamases: a prospective study with Swedish volunteers. Antimicrob Agents Chemother 2010; 54:3564–8.
3. Tham J, Olhausen T, Walder M, Brodun A, Ahl J, Melander E. Extended-spectrum beta-lactamase-producing Escherichia coli in patients with travellers’ diarrhoea. Scand J Infect Dis 2010; 42:275–80.
4. Ostholm-Balkhed A, Tännberg M, Nilsson M, Nilsson LE, Hanberger H, Häggren A; Travel Study Group of Southeast Sweden. Travel-associated faecal colonization with ESBL-producing Enterobacteriaceae: incidence and risk factors. J Antimicrob Chemother 2013; 68:2144–53.
5. Paltansing S, Vlot JA, Kraakman ME, et al. Extended-spectrum ß-lactamase-producing Enterobacteriaceae among travellers from the Netherlands. Emerg Infect Dis 2013; 19:1206–13.
6. Kuenzl E, Jaeger VK, Frei R, et al. High colonization rates of extended-spectrum ß-lactamase (ESBL)-producing Escherichia coli in Swiss travellers to South Asia—a prospective observational multicentre cohort study looking at epidemiology, microbiology and risk factors. BMC Infect Dis 2014; 14:528. doi: 10.1186/1471-2334-14-528.
7. Kantele A, Liäviäri T, Mero S, et al. Antimicrobials increase travellers’ risk of colonization by extended-spectrum betalactamase-producing Enterobacteriaceae. Clin Infect Dis 2015; 60:837–46.
8. Rupp E, Armand-Lefèvre L, Estellat C, et al. High rate of acquisition but short duration of carriage of multidrug-resistant Enterobacteriaceae after travel to the tropics. Clin Infect Dis 2015; 61:593–600.
9. Reutel EA, Sonder GJ, Stolte I, et al. Travel to Asia and traveler’s diarrhoea with antibiotic treatment are independent risk factors for acquiring ciprofloxacin-resistant and extended spectrum beta-lactamase-producing Enterobacteriaceae—a prospective cohort study. Clin Microbiol Infect 2016; 22:731.e1–7.
10. Arcilla MS, van Hattem JM, Haverkate MR, et al. Import and spread of extended-spectrum ß-lactamase-producing Enterobacteriaceae by international travellers (COMBAT study): a prospective, multicentre cohort study. Lancet Infect Dis 2017; 17:78–85.
11. Woerther PL, Bundet C, Chachaty E, Andremont A. Trends in human fecal carriage of extended-spectrum ß-lactamases in the community: toward the globalization of CTX-M. Clin Microbiol Rev 2013; 26:744–58.
12. Schwaber MJ, Navon-Venezia S, Kaye KS, Ben-Ami R, Schwartz D, Carmeli Y. Clinical and economic impact of bacteremia with extended-spectrum-beta-lacta- 

mase-producing Enterobacteriaceae. Antimicrob Agents Chemother 2006; 50:1257–62.
13. England R, Robert J, Toyrina-Kouyoundjian E, Laouria S, Meyeronnson V, Cuemes E. MDR-GNB Travel Working Group. High rate of multidrug-resistant gram-negative bacilli carriage and infection in hospitalized traveling returners: a cross-sectional cohort study. J Travel Med 2015; 22:292–9.
14. Vading M, Kabir MH, Kalin M, et al. Frequent acquisition of low-virulence strains of ESBL-producing Escherichia coli in travellers. J Antimicrob Chemother 2016; 71:5348–55.
15. Soraa A, Sandseth A, Sandven I, Brunborg C, Jenum PA. Risk factors for community-acquired urinary tract infections caused by ESBL-producing Enterobacteriaceae—a case-control study in a low prevalence country. PLoS One 2013; 8:e69581.
16. Osthoff M, McGuinness SL, Wagen AZ, Eise P. Urinary tract infections due to extended-spectrum-lactamase-producing gram-negative bacteria: identification of risk factors and outcome predictors in an Australian tertiary referral hospital. Int J Infect Dis 2015; 34:79–83.
17. Steffen R, Hill DR, Dupont HL. Traveler’s diarrhea: a clinical review. JAMA 2015; 313:71–80.
18. Liäviäri T, Antikainen J, Pakkanen SH, Kierveski J, Kantele A. Prospective study of pathogens in asymptomatic travellers and those with diarrhoea: aetiological agents revisited. Clin Microbiol Infect 2016; 22:535–41.
19. Lertsetthakarn P, Silapong S, Sukpaisal P, et al. Travelers’ diarrhoea in Thailand: a quantitative analysis using TaqMan array card. Clin Infect Dis 2018; 67: 120–7.
20. Kantele A, Mero S, Kierveski J, Liäviäri T. Fluoroquinolone antibiotic users select fluoroquinolone-resistant ESBL-producing Enterobacteriaceae (ESBL-PE)—data of a prospective traveller study. Travel Med Infect Dis 2017; 16:23–30.
21. Mender Arancibia E, Pitari C, Ruiz J, Marco E, Gascon J, Vila J. Evolution of antimicrobial resistance in enteroaggregative Escherichia coli and enterotoxigenic Escherichia coli causing traveller’s diarrhoea. J Antimicrob Chemother 2009; 64:343–7.
22. Ouyang-Latimer J, Jafari S, VanTassel A, et al. In vitro antimicrobial susceptibility of bacterial enteropathogens isolated from international travelers to Mexico, Guatemala, and India from 2006 to 2008. Antimicrob Agents Chemother 2011; 55: 874–8.
23. Margulieux KR, Srijan A, Ruekit S, et al. Extended-spectrum beta-lactamase prevalence and virulence factor characterization of enterotoxigenic Escherichia coli responsible for acute diarrhoea in Nepal from 2001 to 2016. Antimicrob Resist Infect Control 2018; 7:87. doi:10.1186/s13756-018-0377-2.
24. World Health Organization. Health topics: diarrhoea. Available at: http://www.who.int/topics/diarrhoea/en/. Accessed 28 October 2018.
25. Warren JW, Abrutyn E, Habel JR, Johnson JR, Schaeffer AJ, Stamm WE. Guidelines for antimicrobial treatment of uncomplicated acute bacterial cystitis and acute pyelonephritis in women. Infectious Diseases Society of America (IDSA). Clin Infect Dis 1999; 29:745–58.
26. Dallenbe C, Da Costa A, Decrè D, Favier C, Arlet G. Development of a set of multiplex PCR assays for the detection of genes encoding important beta-lactamases in Enterobacteriaceae. J Antimicrob Chemother 2010; 65:490–5.
27. Antikainen J, Kantele A, Pakkanen SH, et al. Quantitative polymerase chain reaction assay for rapid detection of 9 pathogens directly from stools of travelers with diarrhea. Clin Gastroenterol Hepatol 2013; 11:300–7.e3.
28. Johnson JR, Johnston BD, Porter S, Thuras P, Aziz M, Price LB. Accessory traits and phylogenetic background predict Escherichia coli extraintestinal virulence better than does ecological source. J Infect Dis 2019; 219:121–32.
29. Clermont O, Christenson JK, Denamur E, Gordon DM. The Clermont sequence type 131-H30 (ST131-H30), 2000 to 2014: Antimicrob Agents Chemother 2016; 27:614–30.
30. Johnson JR, Johnston BD, Porter S, Thuras P, Aziz M, Price LB. Accessory traits and phylogenetic background predict Escherichia coli extraintestinal virulence better than does ecological source. J Infect Dis 2019; 219:121–32.
31. Hebbelstrup Jensen B, Olsen KE, Struve C, Krogfelt KA, Petersen AM. Epidemiology and clinical manifestations of enteroaggregative Escherichia coli. Clin Microbiol Rev 2014; 27:614–30.
33. Guiral E, Mendez-Aracibia E, Soto SM, et al. CTX-M-15-producing enteroaggregative Escherichia coli as cause of travelers’ diarrhea. Emerg Infect Dis 2011; 17:1950–3.
34. Gunell M, Aulu L, Jalava J, et al. Cefotaxime-resistant Salmonella enterica in travelers returning from Thailand to Finland. Emerg Infect Dis 2014; 20:1214–7.
35. Post A, Martiny D, van Waterschoot N, et al. Antibiotic susceptibility profiles among Campylobacter isolates obtained from international travelers between 2007 and 2014. Eur J Clin Microbiol Infect Dis 2017; 36:2101–7.
36. Khawaja T, Kirveskari J, Johansson S, et al. Patients hospitalized abroad as importers of multiresistant bacteria—a cross-sectional study. Clin Microbiol Infect 2017; 23:673.e1–8.
37. Rodríguez-Baño J, Pascoal A. Clinical significance of extended-spectrum beta-lactamases. Expert Rev Anti Infect Ther 2008; 6:671–83.
38. Wallace-Gadsden F, Johnson JR, Wain J, Okeke IN. Enteroaggregative Escherichia coli related to uropathogenic clonal group A. Emerg Infect Dis 2007; 13: 757–60.
39. Olesen B, Scheutz F, Andersen RL, et al. Enteroaggregative Escherichia coli O78:H10, the cause of an outbreak of urinary tract infection. J Clin Microbiol 2012; 50:3703–11.
40. Johnson JR, Russo TA. Molecular epidemiology of extraintestinal pathogenic Escherichia coli. EcoSal Plus 2018; 8. doi:10.1128/ecosalplus.ESP.0004-2017.