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Human Bocavirus in Saudi Arabia: Molecular Epidemiology and Co-Infections among children with acute respiratory tract infections

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Abstract

Background

During the last few years since HBoV discovery, the virus has attracted less attention and appeared to be one of the most common acute respiratory tract pathogens in children. However, few studies focused on genetic characterization and epidemiology of HBoV in Saudi Arabia, leading us to evaluate the molecular epidemiology of HBoV strains among children with ARI. This study was designed to study the epidemiological, genetic diversity and circulation pattern of HBoV among hospitalized children (< 7 years) due to acute respiratory tract infection (ARTI) during two consecutive years. Nasopharyngeal aspirate (NPA) samples (n=196) were collected. HBoV and co-infection with other respiratory pathogens were identified using qRT-PCR. Positive samples were subsequently subjected to amplify full-length of VP1/VP2 gene, followed by sequencing and phylogenetic analyses.

Results

Our data showed that 3.4% (2014/2015) and 11.3% (2015/2016) of children hospitalized due to acute respiratory infections (ARI) were infected by HBoV-1. We have shown that HBoV-1 was detected year-round. HBoV-1 also was co-detected with one or multiple other respiratory viruses. Phylogenetic analysis of complete HBoV VP1/2 sequences showed no nucleotide variation and the circulation of HBoV-1. The multisequence analysis showed complete identity between HBoV isolated from 2014/2015 and 2015/2016, suggesting no genetic variability among the local circulating HBoV-1 genotypes. Our data reveal that genetically conserved HBoV-1 is circulating in Saudi Arabia.

Conclusions

The multisequence analysis showed complete identity between HBoV-1 isolated from 2014/2015 and 2015/2016, suggesting no genetic variability among the local circulating HBoV-1 genotypes. Our data reveal that genetically conserved HBoV-1 is circulating year-round in Saudi Arabia.
Keywords: Molecular epidemiology; acute respiratory tract infections; human bocavirus; co-infections; children; genotyping; Saudi Arabia
Introduction

Viral respiratory tract infections (RTIs) are the major (50-90%) infection respiratory syndrome that affects human, particularly infants, children elderly, and the immunocompromised individuals. HBoV infections in infants has been associated with acute respiratory tract infection (ARTI). ARTI among children is a major cause of morbidity and mortality globally [1,2]. HBoV causes more than 21.5% RTIs among children worldwide [2]. HBoV causes lower respiratory tract symptoms includes viral bronchiolitis, wheezing pneumonia and respiratory distress [3,4]. A number of studies revealed that HBoV-1 infection was associated with severe lower respiratory tract diseases, leading to hospitalization and intensive care unit (ICU) admission [2,3,4,5]

HBoV is classified into Parvoviridae family, Parvovirinae subfamily and the genus Bocaparvovirus [6,7]. Human Bocavirus is a 5.3 kb, single stranded DNA genome, icosahedral non-envelope with approximately diameter 18–26 in nanometers [8]. The genome of HBoV contains three open reading frames (ORFs). The first ORF encodes nonstructural proteins (NS), the second ORF encodes (NP1) protein and the third ORF encodes (VP1 and VP2) proteins [9,10,11]. Based on genetic variability of VP1gene, HBoV are grouped into four genotypes (HBoV-1, HBoV-2, HBoV-3 and HBoV-4). Several studies have shown that HBoV1 is associated with respiratory tract infections, while HBoV-2, 3 and 4 are associated with gastrointestinal tract infection [8,9,10].

Although Saudi Arabia is considered a virgin soil for spread and evolution of respiratory viruses, there is substantial lack of data regarding epidemiology, circulation pattern and mechanisms adopted in diversification and evolution of HBoV. In a previous set of studies conducted by our laboratory the prevalence and circulation pattern of several respiratory viruses were studied. HBoV studies in Saudi Arabia have only reported in few studies [19,20,25]. There is no published data on the complete Vp1 and VP2 gene sequences of HBoV-1 in Saudi Arabia. This study was designed to study the epidemiological, genetic diversity, circulation pattern and molecular evolution of bocavirus among children.
Materials and Methods

Ethical considerations

The institutional review board at King Saud University- Collage of Medicine University reviewed and approved the study protocol (IRB register number E-14-1326). The parents of the patients participated in this study gave written informed consent before enrollment.

Study design, patients, respiratory samples collection and analysis

Nasopharyngeal aspirate (NPA) samples (n=196), from hospitalized neonates, infants and young children < 7 years of both sexes were collected from King Khalid University Hospital (KKUH) in Riyadh, Saudi Arabia, over two consecutive years (2014/2015 and 2015/2016). ARI cases were defined as an illness that showed one or more of the main symptoms of the clinical manifestations including: cough, fever (temperature, >38 °C), coryza, difficult or shortness of breath, dyspnea, nasal, sneeze and obstruction during admission time. NPA samples were collected from different wards pediatric intensive care unit (PICU), pediatric surgery, cardiology and medical wards). The respiratory samples were placed into 1 mL of universal transport medium (Kit Cat. No. 360c, Copan Italia, Brescia, Italy), transported on ice to the Virology Research Group (VRG) Laboratory, King Saud University, and were mixed via pulse-vortexing and divided into two aliquots. The samples were examined immediately or stored at -80 °C until further test. All methods in this study were preformed in accordance with international guidelines.

HBoV and other respiratory pathogens detection using real-time quantitative polymerase chain reaction

For viral nucleic acid extraction, 140 μl of respiratory clinical samples was used with QIAamp Viral RNA extraction kit (Qiagen, Valencia, CA) according to the manufacturer's instructions. This kit enables to extract DNA along with the RNA and no step for DNase digestion was included. The extracted viral nucleic acid was eluted in 60 μl of elution buffer and kept aliquotted at -80°C until used in qPCR. During the extraction process an internal RNA virus (extraction control), Equine arteritis virus (EAV), which was included in the
FTD kit (Fast-track Diagnostics, Luxembourg), was added to the lysis buffer for each sample and negative control for monitoring extraction, reverse transcription quality and possible PCR inhibition. The concentration (ng/μl) and purity (260/280) of the RNA were measured using NanoDrop2000 (Thermo- scientific). The extracted nucleic acid samples were directly subjected to multiplex RT-PCR analysis using the FTD® Respiratory Pathogens 21 kit (Fast-track Diagnostics, Junglinster, Luxemburg) using 7500 Applied Biosystem™ real-time machine (ThermoFisher Scientific Inc, MA, USA). This kit able to detect specific viral or bacterial DNA sequences in the reaction through increasing a fluorescence observed from the relevant dual-labelled probe which expressed as a cycle threshold value (Ct). The RT-PCR reaction was performed in a total of 25 μL, containing 12.5 μl Buffer, 1.5μl PPmix, 1μl enzyme mix (Fast-track mastermix) and 10 μl extracted viral nucleic acid. A negative and a positive control involved in the kit were used in all runs to observer the assay performance. Amplification was conducted using 7500 Applied Biosystem™ real-time machine (ThermoFisher Scientific Inc, MA, USA) under the following thermocycling conditions: One cycle of initial reverse transcription at 50 °C for 15 min, holding at 94 °C for 1 min, followed by 40 cycles at 94 °C for 8s s and 60 °C for 1 min. In each RT-PCR cycles the reading of fluorescence was obtained in the 60 °C /34 s step and the threshold value (Ct) were determined.

**PCR and sequencing of Human Bocavirus (HBoV)**

The qPCR-positive samples were used for viral DNA extraction by QIAamp DNA Mini Kit (Qiagen), and subjected to amplify full-length of VP1 and VP2 gene. The PCR protocol involved amplification of three fragments that cover the entire VP1 and VP2 genes (Figure 1). Designed oligonucleotide primers used for PCR showed in (Table 1). PCR reaction sequence was amplified using GeneAmp® High-Fidelity PCR Kit (Applied Biosystems, Foster City, CA) according to the manufacturer’s instructions. The total 50 μl reaction mix consisted of 1 μl GeneAmp PCR buffer II (100 mM Tris_HCl, pH 8.3_500 mM KCl) (Applied Biosystems), 2.5 mM MgCl2, each dNTP at 0.2 mM, 20 pmol each of the primers and 10 μl DNA. The reaction tubes were placed, in GeneAmp 9700 thermal cycler (Applied Biosystems, Foster City, CA), at 95 °C for 5 min (initial denaturation), followed by 45
cycles of denaturation at 94 °C for 15 s, annealing at 60 °C for 1 min and extension at 72 °C for 1 min, and one cycle at 72 °C for 10 min (final extension). All PCR products were separated, visualized by agarose gel and were purified with IllustraTM GFX PCR DNA and Gel Band Purification Kit (GE Healthcare). Sequencing of the purified products on both strands was conducted using the specific PCR primers at GTAC Biotech (Cologne, Germany).

Table I. The general nested PCR primer pairs for HBoV-1 VP1 and VP2 gene.

| Primer name     | Primer sequence                  |
|-----------------|----------------------------------|
| Boca-Seq1-FP    | 5′-GAA GAC GAG GGA GAG TAC ATC-3’ |
| Boca-Seq1-RP    | 5′-CCT CCA ATA CTT CCT GTT CCT C-3’ |
| Boca-Seq2-FP    | 5′-GTC TGA CAC TGA CAT TCA AGA CC-3’ |
| Boca-Seq2-RP    | 5′-GTT GGT GCC AGA CAT CCG CTT G-3’ |
| Boca-Seq3-FP    | 5′-GGA CCA CAG TCA TCA GAC-3’   |
| Boca-Seq3-RP    | 5′-CCA CTA CCA TCG GGC TG-3’   |

Phylogenetic and alignment analysis. The HBoV complete VP1 and VP2 gene sequence were downloaded from GenBank and edited and assembled using Bioedit program, version 7.0 (Ibis Biosciences, Carlsbad, CA). Phylogenetic analyses were performed using MEGA software. Briefly, HBoV sequences were aligned using Clustal W algorithm. A phylogenetic tree was constructed with the neighbor-joining algorithm with 1000 bootstrap resampling.
Figure 1. shows the primers design and overlapped amplified area of the HBoV-1 VP1 and VP2 genes that was used for PCR and sequencing.
Results

Prevalence and epidemiology of HBoV
During the two-year period of this study (2014–2016), we analysed a total of 196 respiratory samples from hospitalized children (age; 1 month to 7 years) due to lower respiratory tract infections. Overall, HBoV was detected in 13 samples (6.6%) by RT-PCR. Of these, HBoV was detected in 3.4% (4/116) and in 11.3% (9/80) of samples collected during 2014/2015 and 2015/2016, respectively. We have shown that HBoV was detected year-round without a marked seasonality. HBoV infection was higher in males (69.2%) than in females (30.8%). Among HBoV-infected children, 84.6% were between 1 and 8 months old and 15.3%, between 1 and 6 years.

HBoV and Co-Infections
In order to study HBoV co-respiratory viral infections, we examined the rate of the major respiratory viruses among the HBoV patients (n = 196). Surprisingly, we found all children with HBoV infection in this study were co-infected with other respiratory viruses (Table 2), including 3.6% patients with Rhinovirus, 3.1% with human respiratory syncytial virus (HRSV), 2.6% with parainfluenza virus (PIV), 1.5% with adenovirus (Adv), 1.0% with influenza A virus (IAV), 0.5% with coronavirus OC43 (HCoV-OC43) and 0.5% with metapneumovirus (HMPV). Multiple infection consisting of HBoV plus three other viruses (HRSV, IAV, Adv, and HPIV) was detected in one patient.

Table 2. Co-infection with other respiratory viruses among HBoV positive children.

| Patient No | Age   | Gender | Co-infection with other respiratory viruses |
|------------|-------|--------|---------------------------------------------|
| 1          | 8 months | Male       | HMPV                                        |
| 2          | 5 months | Male       | HRSV+Rhino+HPIV-3                           |
| 3          | 5 months | Male       | HRSV                                        |
| 4          | 5 months | Female     | HRSV+Rhino+Entero                           |
| 5          | 6 months | Female     | Rhino+Influenza A virus                     |
| No. | Age   | Gender | Co-infections                  |
|-----|-------|--------|--------------------------------|
| 6   | 4 months | Female | HRSV+Adv+IAV+HPIV-3+          |
| 7   | 19 months | Female | Rhino+ Adv+ HPIV-3            |
| 8   | 5 months  | Male   | Rhino+ Adv                    |
| 9   | 7 months  | Male   | Adv+ HPIV-3                   |
| 10  | 4 months  | Male   | Rhino+HPIV-1                  |
| 11  | 4 months  | Male   | HRSV                          |
| 12  | 7 months  | Male   | Rhino                         |
| 13  | 6 months  | Male   | HCoV-OC43                     |

**HBoV molecular characterization**

We successfully sequenced all HBoV positive samples. The results of sequences were compared with HBoV reference sequences registered in the GenBank database (https://www.ncbi.nlm.nih.gov/genbank/). Phylogenetic tree constructed showed that HBoV-positive specimens of this study clustered around HBoV-1 genotype sharing 97-100% identity with HBoV-1 reference sequences (Figure 2). We detected only HBoV-1 genotype, our results suggest that a single genetic lineage of HBoV-1 is circulating and causing respiratory infections among children in Saudi Arabia. Ten complete sequence of HBoV-VP1/VP2 genes were deposited in GenBank Accession number (MH427520, MH427519, MH427518, MH427517, MH427516, MH427515, MH427514, MH427513, MH427512 and MH427511).
Figure 2. Phylogenetic tree of HBoV-1 positive samples from children with acute respiratory tract infections from Saudi Arabia. Nucleotide-sequence-based phylogenetic analysis of complete HBoV-1 VP1 and VP2 genes (shown in red) and reference HBoV strains from the GenBank database. A neighbor-joining tree of complete VP1 and VP2 nucleotide sequences was conducted using MEGA software. The accession number, country and year of each strain are shown. Phylogenetic trees were constructed with the neighbor-joining method. Our HBoV strains were in the same cluster as the reference isolates and they belonged to HBoV-1.
Discussion

During the last few years since HBoV discovery, HBoV is appeared to be one of the most common acute respiratory tract pathogens in children (1). In the present study, we studied the prevalence and epidemiology of HBoV among children up to 6 years old in Saudi Arabia, as well as major respiratory viral co-infections and the characterization of circulating HBoV genotypes. Overall, HBoV was detected in 3.4% (4/116) and 11.3% (9/80) of respiratory samples collected during 2014/2015 and 2015/2016, respectively. Interestingly, we detected HBoV in one case age 41 days-old child, supporting the possibility that HBoV can infect a very young child. We have shown that HBoV-1 was detected year-round without a marked seasonality. Besides, co-infections with one or multiple respiratory viruses were detected in all HBoV-positive samples. Interestingly, all HBoV1 infected cases were associated with high rates of other respiratory viral co-infections. Several studies revealed that rates of detection of HBoV in acute respiratory infection (ARI) is about 3-19.0% [13,14]. In the present study, the overall detection rate of HBoV in among hospitalized children due to LRTI was 6.63% which was similar to other countries of the world.

In this study, co-infections with one or multiple respiratory viruses were detected all HBoV-positive samples. Several studies, have shown that HBoV-infected children were more likely to be co-infected with other respiratory viruses [12,15,16]. The higher rate of HBoV co-infection is in line with a higher HBoV co-infection which reported from different countries around the world [17,18,19]. In addition, previous study from Saudi Arabia found only one child diagnosed with HBoV as a single virus entity while most of children (17/18) showed coinfection with other viral pathogens. The most commonly detected viruses were HRSV (72.2%), IAV (66.66%), adenovirus (33.33%) and HPIV-3 (5.5%) [20]. Likewise, we found 3.6% patients co-infected with Rhinovirus, (3.1%) with human respiratory syncytial virus (HRSV), 2.6 % with parainfluenza virus (PIV), 1.5% with adenovirus (Adv), 1.0% with influenza A virus (IAV), 0.5% with coronavirus OC43 (HCoV-OC43) and 0.5% with metapneumovirus (HMPV). Multiple infection consisting
of HBoV plus three other viruses (HRSV, IAV, Adv, and HPIV) was detected in one patient.

Seasonal distribution of HBoV is not well defined, and might be affected by geographic areas. Some studies showed that high prevalence of HBoV infections occurred in the winter and spring [21,22,23,24], while other studies showed high prevalence in the early summer and late spring [23,24,25]. However, clear seasonal activity was not observed in some studies. In Saudi Arabia, the seasonality of HBoV infection has not been clearly reported. In one study conducted in Eastern province Saudi Arabia, HBoV was detected in August, October and December [19,25]. Nevertheless, seasonal variations of respiratory viruses detected from children with respiratory tract infections was studied. Respiratory viruses were detected year-round with a peak in winter (December to March) in Saudi Arabia [19,25]. In this study, we have shown that HBoV was detected year-round without a marked seasonality, thus, we cannot draw any conclusions about seasonality based on our data. HBoV seasonality should be investigated carefully, an overlapping in seasonality of HBoV may be due to the HBoV- coinfection and different respiratory viruses co-circulating at the same time. Establishing the national surveillance system is needed to identify a clear picture of HBoV seasonality in Saudi Arabia.

Based on Vp1/ and VP2 complete gene sequences, all HBoV strains isolated in this study were identified as HBoV-1. Although no variability exists in HBoV-1 genomic region encoding Vp1/ and VP2, all our HBoV strains were in the same cluster as the reference isolates and they belonged to HBoV-1. The multisequence analysis showed complete identity between HBoV isolated from 2014/2015 and 2015/2016, suggesting minor genetic variability among the local circulating HBoV-1 strains over short period of time. Together with several studies from different geographical regions [20,21,22,23], our results provide support evidence for a global circulation of HBoV-1 among children causing ARTI. Several HBoV-1 molecular epidemiology studies have been conducted previously in Saudi Arabia [20,25], however, the complete viral sequences published were infrequent and inadequate to draw any chronological or spatial characteristics. Molecular characterization
of multiple HBoV-1 strains from all regions of Saudi Arabia might help in understanding
the evolution of this virus. In a previous set of studies conducted by our laboratory
(Virology research group), the prevalence and circulation pattern of several respiratory
viruses were studied. Although the results were promising, tracking the epidemiology and
genetic variation among Saudi isolates for an extended period is strongly recommended.

In conclusion, over a period of two years (2014–2016), we detected HBoV in 3.4% (4/116)
and 11.3% (9/80) of samples collected in 2014/2015 and 2015/2016, respectively. We
detected HBoV year-round without a marked seasonality with high rates of viral co-
infections. Our data reveal that genetically conserved HBoV-1 is circulating in the Saudi
Arabia. **Further** epidemiological and molecular characterization are required of multiple
HBoV-1 strains from all regions of Saudi Arabia are required to understand the evolution
of this virus.
References

1- Abdel-Moneim, A. S., Kamel, M. M., Al-Ghamdi, A. S., & Al-Malky, M. I. Detection of bocavirus in children suffering from acute respiratory tract infections in Saudi Arabia. *PloS one*, 8(1), 2013; e55500.

2- Allander T, Jartti T, Gupta S, Niesters HG, Lehtinen P, Osterback R et al. Human bocavirus and acute wheezing in children. Clin Infect Dis 44:2007; 904–910.

3- Allander, T., Tammi, M. T., Eriksson, M., Bjerkner, A., Tiveljung-Lindell, A., & Andersson, B. Cloning of a human parvovirus by molecular screening of respiratory tract samples. *Proceedings of the National Academy of Sciences of the United States of America*, 102(36), 2005;12891–12896.

4- Bagasi AA, Howson-Wells HC, Clark G, Tarr AW, Soo S, Irving WL, McClure CP. Human Bocavirus infection and respiratory tract disease identified in a UK patient cohort. J Clin Virol. 2020; 129:104453. doi: 10.1016/j.jcv.2020.104453.

5- Brieu, N., Guyon, G., Rodiere, M., Segondy, M. & Foulongne, V. Human bocavirus infection in children with respiratory tract disease. Pediatr Infect Dis J 27, 2008; 969–973.

6- Cashman O, O’Shea H. Detection of human bocaviruses 1, 2 and 3 in Irish children presenting with gastroenteritis. Arch Virol. 2012;157(9):1767–73.

7- Deniz Kesebir, Marietta Vazquez, Carla Weibel, Eugene D. Shapiro, David Ferguson, Marie L. Landry, Jeffrey S. Kahn, Human Bocavirus Infection in Young Children in the United States: Molecular Epidemiological Profile and Clinical Characteristics of a Newly Emerging Respiratory Virus, *The Journal of Infectious Diseases*, Volume 194, Issue 9, 2006; 1276–1282.

8- Eskola, Vesa MD, PhD*; Xu, Man MSc†; Söderlund-Venermo, Maria PhD† Severe Lower Respiratory Tract Infection Caused by Human Bocavirus 1 in an Infant, The Pediatric Infectious Disease Journal. 36; 2017;1107-1108 doi: 10.1097/INF.0000000000001681.
9- García-García ML, Calvo C, Falcón A, Pozo F, Pérez-Breña P, De Cea JM, Casas I. Role of emerging respiratory viruses in children with severe acute wheezing. Pediatr Pulmonol. 2010; 45:585–91.

10- Ghietto LM, Majul D, Ferreyra Soaje P, Baumeister E, Avaro M, Insfrán C, Mosca L, Cámara A, Moreno LB, Adamo MP. Comorbidity and high viral load linked to clinical presentation of respiratory human bocavirus infection. Arch Virol. 2015;160(1):117-27. doi: 10.1007/s00705-014-2238-5.

11- Guido, M., Tumolo, M. R., Verri, T., Romano, A., Serio, F., De Giorgi, M., De Donno, A., Bagordo, F., & Zizza, AHuman bocavirus: Current knowledge and future challenges. World journal of gastroenterology, 22(39); 2016; 8684–8697. https://doi.org/10.3748/wjg.v22.i39.8684.

12- Jiang, W., Yin, F., Zhou, W. et al. Clinical significance of different virus load of human bocavirus in patients with lower respiratory tract infection. Sci Rep 6, 2016;20246. https://doi.org/10.1038/srep20246.

13- Kaida, A., Kubo, H., Takakura, K. & Iritani, N. Detection and quantitative analysis of human bocavirus associated with respiratory tract infection in Osaka City, Japan. Microbiol Immunol 54, 2010; 276–281.

14- Kenmoe, S., Vernet, MA., Njankouo-Ripa, M. et al. Phylogenic analysis of human bocavirus detected in children with acute respiratory infection in Yaounde, Cameroon. BMC Res Notes. 2017;10, 293. https://doi.org/10.1186/s13104-017-2620-y.

15- Koseki, N., Teramoto, S., Kaiho, M., Gomi-Endo, R., Yoshioka, M., Takahashi, Y., Nakayama, T., Sawada, H., Konno, M., Ushijima, H., Kikuta, H., Ariga, T., & Ishiguro, N. Detection of human bocaviruses 1 to 4 from nasopharyngeal swab samples collected from patients with respiratory tract infections. Journal of clinical microbiology, 50(6), 2012;2118–2121. https://doi.org/10.1128/JCM.00098-12.

16- Liu, W. K. et al. Detection of human bocavirus from children and adults with acute respiratory tract illness in Guangzhou, southern China. BMC Infect Dis 11; 2011;345.
17-Ljubin-Sternak S, Meštrović T, Ivković-Jureković I, Tešović G, Mlinarić-Galinović G, Lukšić I, et al. High detection rates of human bocavirus in infants and small children with lower respiratory tract infection from Croatia. Clin Lab. 2019.

18-Moesker FM, van Kampen JJ, van der Eijk AA, van Rossum AM, de Hoog M, Schutten M, Smits SL, Bodewes R, Osterhaus AD, Fraaij PL. Human bocavirus infection as a cause of severe acute respiratory tract infection in children. Clin Microbiol Infect. 2015;21(10): 964.e1-8. doi: 10.1016/j.cmi.2015.06.014.

19-Petrarca, L., Nenna, R., Frassanito, A. et al. Human bocavirus in children hospitalized for acute respiratory tract infection in Rome. World J Pediatr 16, (2020); 293–298. https://doi.org/10.1007/s12519-019-00324-5.

20-Petrarca, L., Nenna, R., Frassanito, A. et al. Human bocavirus in children hospitalized for acute respiratory tract infection in Rome. World J Pediatr 16, 2020; 293–298. https://doi.org/10.1007/s12519-019-00324-5

21-Pilger DA, Cantarelli VV, Amantea SL, Leistner-Segal S. Detection of human bocavirus and human metapneumovirus by real-time PCR from patients with respiratory symptoms in Southern Brazil. Mem Inst Oswaldo Cruz 106(1):2011; 56–60.

22-Pozo F, Garcia-Garcia ML, Calvo C, Cuesta I, Perez-Brena P, Casas I. High incidence of human bocavirus infection in children in Spain. J Clin Virol. 40:2007;224–228.

23-Rikhotso, M. C., Kabue, J. P., Ledwaba, S. E., Traoré, A. N., & Potgieter, N. (2018). Prevalence of Human Bocavirus in Africa and Other Developing Countries between 2005 and 2016: A Potential Emerging Viral Pathogen for Diarrhea. Journal of tropical medicine, 7875482. https://doi.org/10.1155/2018/7875482.

24-Silva, P.E., Figueiredo, C.A., Luchs, A. et al. Human bocavirus in hospitalized children under 5 years with acute respiratory infection, São Paulo, Brazil, 2010. Arch Virol 163, 2018; 1325–1330.
25- Sun H, Sun J, Ji W, Hao C, Yan Y, Chen Z, et al. Impact of RSV coinfection on human bocavirus in children with acute respiratory infections. J Trop Pediatr. 2019; 65:342-51.
Declarations

Ethics approval and consent to participate

The institutional review board at King Saud University- Collage of Medicine reviewed and approved the study protocol (IRB register number E-14-1326). The parents of the patients participated in this study gave written informed consent before enrollment.

Consent for publication

Not applicable

Availability of data and materials

The datasets used during the current study are available from the corresponding author on reasonable request.

Complete sequence of HBoV-VP1/VP2 genes deposited in GenBank from this study, Accession numbers:

1. MH427520: https://www.ncbi.nlm.nih.gov/nuccore/MH427520.1
2. MH427519: https://www.ncbi.nlm.nih.gov/nuccore/MH427519.1
3. MH427518: https://www.ncbi.nlm.nih.gov/nuccore/MH427518.1
4. MH427517: https://www.ncbi.nlm.nih.gov/nuccore/MH427517.1
5. MH427516: https://www.ncbi.nlm.nih.gov/nuccore/MH427516.1
6. MH427515: https://www.ncbi.nlm.nih.gov/nuccore/MH427515.1
7. MH427514: https://www.ncbi.nlm.nih.gov/nuccore/MH427514.1
8. MH427513: https://www.ncbi.nlm.nih.gov/nuccore/MH427513.1
9. MH427512: https://www.ncbi.nlm.nih.gov/nuccore/MH427512.1
10. MH427511: https://www.ncbi.nlm.nih.gov/nuccore/MH427511.1

Competing interests

The authors declare that they have no competing interests

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**Authors' contributions**

Study design, respiratory samples collection and performed the experiments: Maaweya E Hamed and Haitham M Amer. Molecular analysis: Mohamed A Farrag. Clinical data collection and manuscript revision: Fahad N Almajhdi, Ibrahim M. Aziz and Rauf Bhat. Wrote the paper: Maaweya E Hamed. All authors read and approved the final manuscript.

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Figure 1 shows the primers design and overlapped amplified area of the HBoV-1 VP1 and VP2 genes that was used for PCR and sequencing.
Phylogenetic tree of HBoV-1 positive samples from children with acute respiratory tract infections from Saudi Arabia. Nucleotide-sequence-based phylogenetic analysis of complete HBoV-1 VP1 and VP2 genes (shown in red) and reference HBoV strains from the GenBank database. A neighbor-joining tree of complete VP1 and VP2 nucleotide sequences was conducted using MEGA software. The accession number, country and year of each strain are shown. Phylogenetic trees were constructed with the neighbor-joining method. Our HBoV strains were in the same cluster as the reference isolates and they belonged to HBoV-1.