Treatment and Outcomes among Patients with Cryptococcus gattii Infections in the United States Pacific Northwest

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Abstract

Background: Cryptococcus gattii is a fungal pathogen causing an emerging outbreak in the United States Pacific Northwest (PNW). Treatment guidelines for cryptococcosis are primarily based on data from Cryptococcus neoformans infections; applicability to PNW C. gattii infection is unknown. We evaluated the relationship between initial antifungal treatment and outcomes for PNW C. gattii patients.

Methods: Cases were defined as culture-confirmed invasive C. gattii infections among residents of Oregon and Washington States during 2004–2011. Clinical data were abstracted from medical records through one year of follow-up. Recommended initial treatment for central nervous system (CNS), bloodstream, and severe pulmonary infections is amphotericin B and 5-flucytosine; for non-severe pulmonary infections, recommended initial treatment is fluconazole. Alternative initial treatment was defined as any other initial antifungal treatment.

Results: Seventy patients survived to diagnosis; 50 (71%) received the recommended initial treatment and 20 (29%) received an alternative. Fewer patients with pulmonary infections [21 (64%)] than CNS infections [25 (83%)] received the recommended initial treatment (p = 0.07). Among patients with pulmonary infections, those with severe infections received the recommended initial treatment less often than those with non-severe infections (11% vs. 83%, p<0.0001). Eight patients with severe pulmonary infections received alternative initial treatments; three died. Four patients with non-severe pulmonary infections received alternative initial treatments; two died. There was a trend towards increased three-month mortality among patients receiving alternative vs. recommended initial treatment (30% vs. 14%, p = 0.12), driven primarily by increased mortality among patients with pulmonary disease receiving alternative vs. recommended initial treatment (42% vs. 10%, p = 0.07).

Conclusions: C.gattii patients with pulmonary infections – especially severe infections – may be less likely to receive recommended treatment than those with CNS infections; alternative treatment may be associated with increased mortality. Reasons for receipt of alternative treatment among C.gattii patients in this area should be investigated, and clinician awareness of recommended treatment reinforced.

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Introduction

Cryptococcus spp. are basidiomycetous yeast, with two species, C. gattii and C. neoformans, causing nearly all human cryptococcal infections [1–3]. C. neoformans typically causes disease in immunocompromised individuals [4,5], and is an important and common cause of opportunistic infections in HIV/AIDS patients worldwide [6–8]. In contrast, C. gattii has historically been characterized as a rare pathogen, with disease confined to tropical and subtropical climates, particularly the highly endemic regions of Australia and Papua New Guinea [9,10]. Until recently, C. gattii was thought to affect primarily immunocompetent persons living in these regions [11–15].

Since 2004, an outbreak of C. gattii infections has been documented in the United States Pacific Northwest states of Oregon and Washington [16]. The outbreak in these states is thought to have originated in, and spread from, British Columbia, Canada, where infections caused by the same C. gattii genetic types have been documented since 1999 [17,18]. Genetic sequencing has demonstrated that C. gattii can be divided into four molecular types, denoted as VGI, VGII, VGIII and VGIV [19]; these molecular types can be distinguished by various genetic techniques.
and have different geographic distributions [20–23]. The emergence of *C. gattii* infections in Oregon, Washington State, and British Columbia is primarily due to the clonal expansion of three genetic subtypes belonging to the molecular type VGII, called VGIIa, VGIIb, and VGIIc [24–26]; these have been referred to as ‘outbreak-strain’ subtypes. Outbreak-strain subtypes are different from those found in historically endemic Australia and Papua New Guinea (and other areas of the world), where infections are most frequently caused by nonclonal strains of *C. gattii*, most commonly VG1 [11,27–29]. Clinical differences between *C. gattii* infections in the United States Pacific Northwest and historically endemic areas have been described. While *C. gattii* in historically endemic areas has been reported to infect primarily immunocompetent persons, causing meningocerebralitis [11,27,28,30–32], *C. gattii* infections in Oregon and Washington State occur frequently in immunocompromised persons and present most often as respiratory illness [33].

Current guidelines for the management and treatment of cryptococcal disease from the Infectious Disease Society of America (IDSA) recommend antifungal treatment that varies by site and severity of infection [34]. The recommended initial treatment (RIT) for severe pulmonary disease, central nervous system (CNS) disease, and other disseminated disease (such as cryptococcosis) is amphotericin B (AMB) and 5-flucytosine (5FC); for non-severe pulmonary disease, the RIT is fluconazole, with itraconazole and posaconazole as acceptable second-line agents [34]. These recommendations are unchanged from previous IDSA guidelines for cryptococcosis, released in 2000, which were available when the majority of patients in this report were diagnosed [35].

Although IDSA guidelines for treatment of cryptococcal disease are based primarily on data from *C. neoformans* infections in HIV and solid organ transplant patients, these guidelines are intended to apply to patients with *C. neoformans* or *C. gattii* infections. A limited number of *C. gattii*-specific recommendations were included for the first time in the 2010 IDSA guidelines and are based on data from *C. gattii* infections in historically endemic areas, the only data available at the time of the guideline-writing [34]. These recommendations pertain mainly to patients with cryptococcomas, which previous data have suggested are more common in patients infected with *C. gattii* than *C. neoformans* [31], and include consideration of surgery for patients with large cryptococcomas, increased radiologic and follow-up evaluations for those with cryptococcomas or hydrocephalus, and possible use of AMB/5FC in patients with large and/or multiple pulmonary cryptococcomas [34].

Given the genotypic and clinical differences between *C. gattii* infections in the United States Pacific Northwest and cryptococcal infections due to either *C. gattii* from historically endemic areas or *C. neoformans*, applicability of the current IDSA guidelines to *C. gattii* patients in Oregon and Washington State is unknown. We conducted a retrospective cohort study of *C. gattii* infections reported in these states to evaluate the relationship between IDSA guideline-recommended initial antifungal treatment and clinical outcomes.

**Methods**

**Case Definition and Case-finding**

A case was defined as culture-confirmed *C. gattii* infection reported to CDC during January 1, 2004 to October 1, 2011, in a person residing in Oregon or Washington State. Patients were identified from existing passive laboratory-based surveillance systems that capture culture-confirmed *C. gattii* infections in these states. Although reporting is passive, *C. gattii* has been notifiable in Oregon since 2011, and in Washington State (as a rare disease of public health importance) since 2006. This investigation was conducted as part of routine public health practice in response to an ongoing outbreak of *C. gattii* in the United States Pacific Northwest. This investigation was reviewed and designated as non-research by a CDC ethics liaison, informed consent was not obtained from patients, and the study was deemed exempt from formal institutional review board evaluation.

**Data Collection**

We abstracted data from case-patient medical records using a standardized form. Information about demographics, underlying medical conditions, current medications, signs and symptoms, laboratory and radiologic studies, and treatments and procedures performed secondary to *C. gattii* diagnosis were recorded. Data were collected from case-patients’ initial visits and at two, six, 12, 24 and 52 weeks of follow-up. Death databases were searched to identify any deaths that occurred during follow-up. All data were entered into a Microsoft Access database.

**Definition of Terms**

We limited our analyses to case-patients with invasive *C. gattii* disease, which we defined as infection of any of the deep organs or body tissues, including blood. We excluded superficial infections, including skin, throat and urinary tract infections without evidence of involvement of other organs, due to the small number and absence of specific treatment recommendations for these infections [34]. Additionally, we excluded children <13 years, as the IDSA guidelines for children differ to some extent from those for adults [34,35].

Infections were categorized in a hierarchy, based on site of infection. Infections were categorized as ‘bloodstream’ if the patient had at least one positive blood culture for *C. gattii*, regardless of other positive cultures. Infections were categorized as ‘CNS’ if cerebrospinal fluid (CSF) or brain tissue cultures yielded *C. gattii* in the absence of documented bloodstream infection. Infections were also categorized as ‘CNS’ if the case-patient had a positive CSF cryptococcal antigen result or had brain tissue histopathology consistent with cryptococcal disease and a positive culture for *C. gattii* from a body site outside the CNS (not including blood). Infections were categorized as ‘pulmonary’ if respiratory specimens (sputum, bronchoalveolar lavage) or lung tissue cultures yielded *C. gattii* in the absence of documented bloodstream or CNS infection. Bloodstream infections were categorized separately from other invasive *C. gattii* infections (e.g. CNS infections) due to the high mortality that was observed in patients with fungemia due to *C. gattii*. Pulmonary infections were further categorized as either ‘non-severe’ or ‘severe’. Severe pulmonary infections were defined as those in which the patient required intensive care unit admission for treatment of pulmonary disease. Non-severe pulmonary infections included all other pulmonary infections.

We defined immunocompromise as the presence of any of the following documented conditions or medications in a patient at initial presentation for cryptococcal disease: active hematologic malignancy, recent neutropenia (defined as absolute neutrophil count <500 cells/µL in the 30 days before cryptococcal diagnosis), history of allogeneic or autologous stem cell transplant, solid organ transplant, cytotoxic chemotherapy, HIV infection, autoimmune disease, oral or parenteral steroid use at a dose >5 mg/day during the past year, or current use of other drugs that suppress the immune system (i.e. methotrexate, tumor necrosis factor-α inhibitor). We defined major medical comorbidities as...
existence of pulmonary, cardiac, liver or renal disease, documented diabetes, or immunocompromise as defined above.

RIT was defined, based on 2010 and 2000 IDSA guidelines, as the administration of AMB/5FC for CNS infections, severe pulmonary infections, and bloodstream infections, and administration of an azole drug (itraconazole, itraconazole, or posaconazole) for patients with non-severe pulmonary infections [34,35]. Alternative initial treatment (AIT) included any other initial antifungal treatment for the respective infections. Recognizing that clinical information obtained during the days following a patient’s diagnosis with cryptococcosis might impact clinical decision-making, we assessed whether treatment was RIT or AIT at four days after a diagnosis of C.gattii was made. For example, for patients with severe pulmonary disease for whom RIT included AMB/5FC, an alternate treatment during days 1–4 after diagnosis did not result in an AIT classification if the patient was switched to AMB/5FC by day five. However, continued AIT past the four-day mark would result in a patient being designated as receiving AIT.

Data Analysis
Comparisons of proportions were evaluated with the χ² test; the Fisher’s exact test was used when one or more cell counts were <5. Comparisons of medians was done with the Wilcoxon-rank-sum test. All analysis was done in SAS version 9.3 (SAS Institute Inc., Cary, NC).

Results
Demographics
We identified 74 patients with invasive C. gattii infections: 19 (26%) in Washington and 55 (74%) in Oregon. Four patients died before diagnosis of C. gattii infection (3, 7, 7, and 12 days before diagnosis); two had bloodstream infections and two had pulmonary infections (both severe).

Seventy (95%) patients survived to diagnosis and were included in further analysis. Median time from symptom onset to diagnosis was 34 (range: 3–351) days. Median patient age was 54 (range: 15–96) years; 36 (51%) were female. Sixty-five (93%) patient isolates were identified as outbreak-strain VGII subtypes, with 43 (61%) VGIIa, 17 (24%) VGIIc, and five (7%) VGIIb; of the remaining isolates, four (6%) were molecular type VGI and one (1%) was VGIII. Fifty-seven (81%) patients were hospitalized at the time of cryptococcal diagnosis. Of the 69 patients with immune status documented, 35 (51%) were immunocompromised at presentation. The most common immunocompromising conditions were systemic steroid use (24 patients; 69%) and autoimmune disease (13 patients; 37%). Among all 70 patients who survived to diagnosis, 3 (4%) patients had documented HIV infection; 36 (51%) additional patients had documented testing for HIV infection at the time of diagnosis of C. gattii infection and were found to be negative. Non-immunocompromising comorbid conditions were also common: 29 (41%) patients had cardiovascular disease, 16 (23%) had diabetes, and 14 (20%) had underlying respiratory disease. Nine (13%) patients were otherwise healthy (no immunocompromise and no comorbid conditions). Thirteen (19%) patients died within three months of diagnosis (Table 1). Table 2 shows all documented sites of C. gattii infection for all patients and the infection-type categorization used during this analysis.

While most immunocompromised patients had pulmonary infections (20/35, 57%), most immunocompetent patients had CNS infections (18/34, 53%). Time from symptom onset to diagnosis of C. gattii infection was significantly longer among patients with pulmonary infections (50 days) than those with either CNS (24 days) (p = 0.005) or bloodstream infections (27 days) (p = 0.02). There were no differences in immune status between patients with bloodstream infections and either pulmonary or CNS infections.

Treatment and Outcomes
Of the 70 patients surviving to diagnosis, 50 (71%) received RIT and 20 (29%) received AIT. Three (43%) patients with bloodstream infections received AIT, compared with 12 (36%) patients with pulmonary infections and five (17%) patients with CNS infections (Table 3). More patients with pulmonary (36%) than CNS (17%) infections received AIT, though this difference was borderline significant (p = 0.07) (Table 3). Patients with bloodstream infections were not significantly more likely than those with either pulmonary (p = 1.0) or CNS (p = 0.16) infections to receive AIT; however, the small number of patients with bloodstream infections likely limited our ability to compare these groups.

Among patients with pulmonary infections, those with severe infections were more likely to receive AIT than those with non-severe infections (89% vs. 17%; p<0.001) (Table 3). Of the eight patients with severe pulmonary infections receiving AIT, seven (88%) received an azole only and one (12%) received AMB monotherapy (no 5FC given) (Table 4). Of the four patients with non-severe pulmonary infections who received AIT, one (25%) received AMB monotherapy, one (25%) received caspofungin and voriconazole, and two (50%) received no treatment (Table 4). All five patients with CNS infections and all three patients with bloodstream infections who received AIT received AMB monotherapy (no 5FC given) (Table 4).

Thirteen (19%) of the 70 patients surviving to diagnosis died within three months. Three-month mortality was highest for patients with bloodstream infections (3/7; 43%), next-highest for patients with pulmonary infections (7/33; 21%), and lowest for patients with CNS infections (3/30; 10%). Overall, three-month mortality was non-significantly higher among patients receiving AIT compared with those receiving RIT (30% vs. 14%, p = 0.12), driven primarily by a trend in increased mortality among patients with pulmonary infections receiving AIT versus RIT (42% vs. 10%, p = 0.07) (Table 3). The increased three-month mortality among patients with pulmonary disease receiving AIT versus RIT was observed both for patients with severe (36% vs. 0%, p = 1.0) and non-severe pulmonary disease (50% vs. 10%, p = 0.12) (Table 3). RIT was not associated with increased mortality among patients with CNS or bloodstream infections (p = 1.0 for both). No association was found between three-month mortality and immunocompromising conditions; nine (26%) immunocompromised patients died within three months of their diagnosis, compared with four (12%) immunocompetent patients (p = 0.22). Similarly, no association was found between three-month mortality and presence of any pre-existing major medical comorbidity; ten (16%) patients with pre-existing comorbidities died within three months of their diagnosis, compared with three (33%) without any pre-existing condition (p = 0.35). No association was found between three-month mortality and time to diagnosis (p = 0.68).
Discussion

We describe the initial antifungal treatments utilized in United States Pacific Northwest C. gattii infections and subsequent patient outcomes. Patients in this analysis, as previously described [16,33], were frequently immunocompromised or had serious comorbid conditions and most commonly presented with pulmonary disease. The overall case-fatality rate for this cohort was high (19%). We found that while a substantial minority of patients did not receive IDSA guideline-recommended initial therapy, the receipt of alternative initial treatments was not equally distributed across all C. gattii infections. Fewer patients with pulmonary infections compared with central nervous system infections received IDSA guideline-recommended initial therapy. Among patients with isolated pulmonary infections, fewer with severe pulmonary infections received recommended initial therapy compared with those persons with non-severe infections. Among the patients who received alternate initial treatment, most were ‘under-treated’, either through failure to receive 5-flucytosine with amphotericin B (for patients with CNS, bloodstream and severe pulmonary infections), or failure to receive any treatment (for patients with non-severe pulmonary infections). Receipt of an alternative initial therapy was associated with a non-significant trend towards increased mortality (approximately 30% excess mortality) in the three months after diagnosis, particularly among patients with pulmonary infections.

There are a number of reasons why IDSA-recommended initial therapy might not have been used with patients in this cohort. While infectious disease clinicians are likely to be aware of the

Table 1. Characteristics of patients with invasive Cryptococcus gattii infection in United States Pacific Northwest who survived to diagnosis (N = 70).

| Characteristic | Sub-category | N (%) |
|---------------|--------------|-------|
| Female        |              | 36 (51) |
| VGII molecular type isolates* |              | 65 (93) |
| Median age (range) in years |              | 54 (15–96) |
| Immunocompromiseb |              | 35 (51) |
| Systemic steroid usec |              | 24 (69) |
| Autoimmune diseasec |              | 13 (37) |
| HIVc |              | 3 (4) |
| Hospitalized at cryptococcal diagnosis |              | 57 (81) |
| Medical co-morbidityc |              | 29 (41) |
| Cardiovascular disease |              | 16 (23) |
| Respiratory disease |              | 14 (20) |
| Otherwise healthy (no immunocompromise or comorbid conditions) |              | 9 (13) |
| Site of infection |              | 33 (47) |
| Pulmonary |              | 30 (43) |
| CNS |              | 7 (10) |
| Severity of pulmonary infection |              | 9 (27) |
| Severe |              | 24 (73) |
| Median time from symptom onset to diagnosis in days |              | 34 (3–351) |
| Died within 3 months of diagnosis |              | 13 (19) |

*VGII molecular type isolates include isolates from the three outbreak genotypes, VGIIa, VGIIb, and VGIIc.

1n=69.

Categories not mutually exclusive.

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Table 2. All documented sites of Cryptococcus gattii infection and categorization for analysis*.

| Body sites found to have Cryptococcus gattii infection during clinical workup | Categorization of infection type for analysis | Number of patients |
|-------------------------------|---------------------------------------------|-------------------|
| Lungs | Pulmonary | 33 |
| Blood | Bloodstream | 2 |
| Blood/Central Nervous System | Bloodstream | 4 |
| Blood/Central Nervous System/Lungs | Bloodstream | 1 |
| Central Nervous System/Lungs | CNS | 5 |
| Central Nervous System | CNS | 25 |

*Total patients in analysis with pulmonary infection, 33; with bloodstream infection, 7; with CNS infection, 30.

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IDSA guidelines for cryptococcal disease, many patients are initially treated by clinicians without formal infectious disease training who may not be aware of the IDSA guidelines. Specifically, they may not be aware that severe pulmonary cryptococcosis should be treated in the same way as central nervous system cryptococcosis, leading to under-treatment of patients with severe pulmonary infections. Additionally, as C. gattii infections in the United States Pacific Northwest appear to be clinically different from C. gattii infections in other areas of the world, some clinicians who are aware of the IDSA guidelines in Oregon and Washington State may initially deviate from IDSA-recommended therapy due to concerns about the generalizability of the guidelines to their patients with C. gattii infection. Finally, clinicians may not have used guideline-recommended initial therapy due to matters beyond their control, such as patient contraindications to medications, insurance restrictions, or drug shortages. While we were unable to evaluate why clinicians chose, in a minority of patients, to pursue alternative treatments, our data suggests that there might be some benefit in adhering to IDSA guideline-recommended initial treatment in United States Pacific Northwest C. gattii patients, particularly those with pulmonary disease. Further research is needed for the reasons for use of alternative initial treatment regimens is needed.

Pulmonary cryptococcosis presents a number of clinical challenges in diagnosis and treatment. Unlike cryptococcal meningitis, a common HIV-related opportunistic infection, pulmonary cryptococcosis is much less-commonly-recognized clinical entity, even among HIV-infected persons [36]. Diagnostic delays for patients with pulmonary cryptococcosis, as seen in this cohort, have been documented previously [37–40]. In terms of treatment, while IDSA guidelines do specify use of antifungal treatment for pulmonary cryptococcal infections – even mild disease - and the use of amphotericin B and 5-flucytosine in ‘severe’ pulmonary disease [34,35], the quality of the evidence for both recommendations is limited and based on “…opinions of respected authorities…clinical experience, descriptive studies, or reports of expert committees” [34]. Unlike for cryptococcal meningitis [41–44], no randomized controlled trials evaluating best treatments for pulmonary cryptococcosis have been published, and divergent opinions exist in the literature on the utility and optimal type of antifungal treatment for these patients. Some clinicians have suggested that asymptomatic or minimally symptomatic pulmonary cryptococcosis in immunocompetent persons requires no antifungal treatment at all [45,46], while others have recommended azole drugs or amphotericin B in all cases [47,48]. In Australia, where guidelines recommend amphotericin B and 5-flucytosine for all but mild/asymptomatic pulmonary cryptococcosis, Chen et al recently published outcomes data on ten patients with isolated pulmonary C. gattii infection. The majority of patients were treated with amphotericin B and 5-flucytosine and only one death was reported [49], raising the question of whether a more aggressive approach might be warranted among patients with pulmonary C. gattii infections. Larger-scale evaluations of patients with pulmonary cryptococcal infections, including patients with C. gattii infections from both previously-recognized endemic areas and the United States Pacific Northwest, are needed to identify the most appropriate treatment and improve outcomes.

We chose to evaluate initial antifungal treatment, and not treatment later in the course of disease, for several reasons. First, initial treatment, termed induction therapy, for cryptococcal disease has been shown to have a strong impact on mortality. Studies of HIV-infected patients during the early years of the HIV epidemic demonstrated that induction therapy for cryptococcal meningitis with fluconazole resulted in worse outcomes than

### Table 3. Three-month mortality among patients by site of infection and by initial therapy receiveda.

| Sites of infection          | n   | Received recommended initial treatment | 3-month mortality among those receiving recommended initial therapy | Received alternative initial therapy | 3-month mortality among those receiving alternative initial therapy |
|----------------------------|-----|----------------------------------------|---------------------------------------------------------------|------------------------------------|---------------------------------------------------------------|
| All                        | 70  | 50 (71%)                               | 7 (14%)                                                      | 20 (29%)                          | 6 (30%)                                                      |
| **By site of infection**   |     |                                        |                                                              |                                    |                                                              |
| Pulmonary                  | 33  | 21 (64%)                               | 2 (10%)                                                      | 12 (36%)                          | 6 (30%)                                                      |
| CNS                       | 30  | 25 (83%)                               | 3 (12%)                                                      | 5 (17%)                           | 0 (0%)                                                       |
| Bloodstream                | 7   | 4 (57%)                                | 2 (40%)                                                      | 3 (43%)                           | 1 (50%)                                                      |
| **By severity of pulmonary infection** |     |                                        |                                                              |                                    |                                                              |
| Severe pulmonary           | 9   | 1 (11%)                                | 0 (0%)                                                       | 8 (89%)                           | 4 (17%)                                                      |
| Non-severe                 | 24  | 20 (83%)                               | 2 (2%)                                                       | 4 (17%)                           | 2 (50%)                                                      |

*aMortality measured from date of diagnosis; 4 patients died prior to diagnosis and receipt of antifungal therapy and are not included in this table.

**Table 4. Initial therapy received by Cryptococcus gattii patients.**

| Site of infection   | n   | Recommended initial therapy (n) | Alternative initial therapy (n) |
|---------------------|-----|---------------------------------|--------------------------------|
| Severe pulmonary    | 9   | Amphotericin B/5-flucytosine (1) | Azole only (7); Amphotericin B only (1) |
| Non-severe pulmonary| 24  | Fluconazole (20)                | No treatment (2); Amphotericin B only (1); caspofungin/voriconazole (1) |
| CNS                 | 30  | Amphotericin B/5-flucytosine (25)| Amphotericin B only (5)     |
| Bloodstream         | 7   | Amphotericin B/5-flucytosine (4) | Amphotericin B only (3)     |
induction with amphotericin B and 5-flucytosine [46]. Similarly, treatment with amphotericin B alone (i.e., without 5-flucytosine) has been shown to be inferior to combination amphotericin B and 5-flucytosine therapy for induction [44,45]. In addition, we believed that selection of the initial antifungal drug is less likely than subsequent treatment choices to be influenced by outside factors (e.g., patient contraindications, adverse reactions) and therefore more indicative of physician preference and knowledge. However, it is possible that consolidation and maintenance drug choices as well as duration of therapy also may influence patient outcomes. Unfortunately, we were not able to evaluate that relationship in this study.

This analysis included several limitations. First, this patient group included those with C. gattii infections that were severe enough to required hospitalization; thus, our findings regarding treatment and outcomes are likely not applicable to mild, self-limited pulmonary C. gattii infections. However, few of these mild infections have been identified in this cohort and it is unclear how frequently they occur. Second, due to the retrospective nature of this study, not all patients received identical diagnostic testing (e.g., lumbar puncture, chest imaging, bronchoalveolar lavage); this may have led to incomplete ascertainment of all sites of infection. Third, these results are specific to patients with C. gattii infection in the United States Pacific Northwest, and may not be generalizable to patients with C. gattii infection in other areas. Finally, the number of patients in our evaluation was small, particularly in subgroup analyses. More data, ideally from prospective studies or clinical trials, is needed to understand the relationship, if any, between site of infection, initial antifungal treatment, and outcomes in this population.

This is the first evaluation of the effect of initial antifungal treatment on patient outcomes in the North American outbreak of C. gattii. We show that a substantial minority of patients are not getting the current guideline-recommended initial antifungal therapy, which may be associated with improved outcomes. Timely diagnosis and appropriate treatment for patients with C. gattii infection will continue to be a clinical question as the emergence of C. gattii in Oregon, Washington State, British Columbia, and elsewhere continues. C. gattii infections in persons with and without recent travel history to the United States Pacific Northwest or British Columbia are increasingly being reported throughout the United States [14,29,33,30,51]; this increased visibility will raise more questions about the best treatment for patients with C. gattii. As our identification of C. gattii infections improves and diagnoses increase, as they are likely to do, careful collection of treatment-related data from patients with these infections will be vital to improving outcomes.

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Author Contributions

Conceived and designed the experiments: RMS JRH ED NM-H. Performed the experiments: RMS AM-J MT TS JRH NM-H. Analyzed the data: RMS JRH. Contributed reagents/materials/analysis tools: MT ED NM-H. Wrote the paper: RMS JRH. Revised JRH RMS.

References

1. Casadevall A, Perfect JR, editors (1998) Cryptococcus neoformans. Washington, DC: ASM Press. 542 p.
2. Perfect JR, Casadevall A (2011) The History of Cryptococcus and Cryptococcosis. In: Heitman J, Kozel TR, Kwon-Chung J, Perfect J, Casadevall A, editors. Cryptococcus: From Human Pathogen to Model Yeast. Washington, DC: ASM Press. 17–26.
3. Heitman J, Kozel TR, Kwon-Chung J, Perfect J, Casadevall A, editors (2011) Cryptococcus: from pathogen to model yeast. Washington, DC: ASM Press. 17–26.
4. Bennett JE, Dismukes WE, Duma RJ, Medoff G, Sandle MA, et al. (1979) A comparison of amphotericin B alone and combined with fluconazole in the treatment of cryptococcal meningitis. N Engl J Med 301: 126–131.
5. Butler WT, Alling DW, Spickard A, Uz JP (1964) Diagnostic and Prognostic Value of Clinical and Laboratory Findings in Cryptococcal Meningitis, a Follow-up Study of Forty Patients. N Engl J Med 270: 59–67.
6. Committee UKCH/CHSS, Garvey L, Winston A, Walsh J, Post F, et al. (2011) HIV-associated central nervous system diseases in the recent combination antiretroviral therapy era. Eur J Neurol 18: 527–534.
7. Aeschliman V, Thiemenmann F, Pepper DJ, Boule A, Wilkinson RJ, et al. (2010) Central nervous system disorders after starting antiretroviral therapy in South Africa. AIDS 24: 2171–2176.
8. Wadhwa A, Kaur R, Bhatta P (2008) Profile of central nervous system disease in HIV/AIDS patients with special reference to cryptococcal infections. Neurologist 14: 247–251.
9. Kwon-Chung KJ, Bennett JE (1984) Epidemiologic differences between the two varieties of Cryptococcus neoformans. Ann J Epidemiol 120: 123–130.
10. Kwon-Chung KJ, Bennett JE (1984) High prevalence of Cryptococcus neoformans var. gattii in tropical and subtropical regions. Zentralbl Bakteriol Mikrobiol Hyg A 257: 213–218.
11. Speed B, Dunt D (1995) Clinical and host differences between infections with the two varieties of Cryptococcus neoformans. Clin Infect Dis 21: 28–34; discussion 35–36.
12. Lalloo D, Fisher D, Naraqi S, Laurenson I, Temu P, et al. (1994) Cryptococcal meningitis (C. neoformans var. gattii) leading to blindness in previously healthy Melanesian adults in Papua New Guinea. Q J Med 87: 343–349.
13. Meyer W, Castaneda A, Jackson S, Huynh M, Castaneda E, et al. (2003) Molecular typing of IberoAmerican Cryptococcus neoformans isolates. Emerg Infect Dis 9: 189–195.

Bowers M, Hagen F, Boekhout T (2008) Diversity of the Cryptococcus neoformans-Cryptococcus gattii species complex. Rev Iberoam Micol 25: S4–26.
14. Byrnes EJ, 3rd, Li W, Lewis Y, Perfect JR, Carter DA, et al. (2009) First reported case of Cryptococcus gattii in the Southeastern USA: implications for travel-associated acquisition of an emerging pathogen. PLoS One 4: e5851.
15. Lopez-Martinez R, Soto-Hernandez JL, Osotrysky-Zechnner L, Castanon-Oliveares LR, Angeles-Morales V, et al. (1996) Cryptococcus neoformans var. gattii among patients with cryptococcal meningitis in Mexico. First observations. Mycopathologia 134: 61–64.
16. MacDougall L, Kidd SE, Galanis E, Mak S, Leslie MJ, et al. (2007) Spread of Cryptococcus gattii in British Columbia, Canada, and detection in the Pacific Northwest, USA. Emerg Infect Dis 13: 42–50.
17. Fye M, MacDougall L, Romney M, Starr M, Pearce M, et al. (2008) Cryptococcus gattii infections on Vancouver Island, British Columbia, Canada: emergence of a tropical fungus in a temperate environment. Can Commun Dis Rep 34: 1–12.
18. Galanis E, MacDougall L (2010) Epidemiology of Cryptococcus gattii, British Columbia, Canada, 1999–2007. Emerg Infect Dis 16: 251–257.
19. Bowers M, Hagen F, Boekhout T (2008) Diversity of the Cryptococcus neoformans-Cryptococcus gattii species complex. Rev Iberoam Micol 25: S4–12.
20. Bowers M, Hagen F, Kuramae EE, Boekhout T (2008) Six monophyletic lineages identified within Cryptococcus neoformans and Cryptococcus gattii by multi locus sequence typing. Fungal Genet Biol 45: 400–421.
21. Fraser JA, Giles SS, Wemunk EC, Geunes-Boyer SG, Wright JR, et al. (2005) Same-sex mating and the origin of the Vancouver Island Cryptococcus gattii outbreak. Nature 437: 1360–1364.
22. Subrin JJ, Costa AK, Cordoviro RA, Brilhaente RS, Moura FE, et al. (2010) Molecular methods for the diagnosis and characterization of Cryptococcus: a review. Can J Microbiol 56: 445–458.
23. Harris J, Lockhart S, Chiller T (2011) Cryptococcus gattii: where do we go from here? Med Mycol.
24. Kidd SE, Guo H, Bartlett KH, Xu J, Kronstad JW (2005) Comparative gene genealogies indicate that two clonal lineages of Cryptococcus gattii in British Columbia resemble strains from other geographical areas. Eukaryot Cell 4: 1629–1638.
25. Kidd SE, Hagen F, Tscharke RL, Huynh M, Bartlett KH, et al. (2004) A rare genotype of Cryptococcus gattii caused the cryptococcosis outbreak on Vancouver Island (British Columbia, Canada). Proc Natl Acad Sci U S A 101: 17258–17263.
38. Taniguchi D, Sawada T, Ryu C, Nagayasu T (2012) Post-renal transplant pulmonary cryptococcosis mimicking primary lung cancer with multiple lung metastases. Tuberc Respir Dis (Seoul) 73: 182–186.

37. Fujimoto H, Suito T, Oyama T (2012) Pulmonary cryptococcosis mimicking primary lung cancer with multiple lung metastases. Tuberc Respir Dis (Seoul) 73: 182–186.

36. Harris JR, Lindsley MD, Henchaichon S, Poonwan N, Naorat S, et al. (2012) Cryptococcus neoformans var. gattii infections in multiple states outside the us pacific northwest. Emerg Infect Dis 19: 1621–1627.

35. Saag MS, Graybill RJ, Larsen RA, Pappas PG, Perfect JR, et al. (2000) Practice guidelines for the management of cryptococcal disease. Infectious Diseases Society of America. Clin Infect Dis 30: 710–718.

34. Perfect JR, Dismukes WE, Dromer F, Goldman DL, Graybill JR, et al. (2010) Cryptococcus neoformans in Australia and New Zealand. Australasian Cryptococcal Study Group. Clin Infect Dis 51: 499–508.

33. Harris JR, Lockhart SR, Debess E, Marsden-Haug N, Goldoft M, et al. (2011) Cryptococcus gattii genotype VGI infection in New England. Pediatr Infect Dis J 30: 1111–1114.