Research Article

Neuroprotective Effects of Jitai Tablet, a Traditional Chinese Medicine, on the MPTP-Induced Acute Model of Parkinson’s Disease: Involvement of the Dopamine System

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1. Introduction

Parkinson’s disease (PD) is a debilitating chronic and progressive neurodegenerative disease. Between 0.5 and 1% of the population aged 65–69 years and 1–3% of those over 80 years old suffer from PD worldwide [1]. The pathophysiology of PD results from the progressive and selective loss of dopaminergic neurons, which leads to the depletion of striatal dopamine (DA), dopamine transporter (DAT), and DA receptors levels. PD patients exhibit several major clinical symptoms, such as bradykinesia, muscle rigidity, and resting tremor [2]. It has been reported that clinical symptoms appeared when approximately 80% of the striatal DA is lost [3]. Therefore, a major treatment strategy for PD focuses on DA supplementation, through the use of DA precursors such as levodopa (L-DOPA) [4], DA receptor agonists, and monoamine oxidase B (MAO-B) inhibitors [1, 5]. Madopar, which is composed of the DA precursor L-DOPA and the decarboxylase inhibitor benzerazide, is commonly used to treat PD by increasing DA concentrations. Although it is used for the relief of PD motor symptoms, it does not prevent or slow the progressive degeneration of neuronal cells and its long-term use may induce various side-effects including fluctuations and abnormalities in involuntary movements [6]. DA receptor agonists and MAO-B inhibitors are both associated with the same issues, which greatly limit their clinical utility [1]. Thus, there is still an urgent need for efficacious therapies that not only act on the motor symptoms...
of PD but also protect neuronal cells and present low side-effect profiles.

Traditional Chinese medicine (TCM) may offer therapies with better efficacy and fewer side-effects [7]. TCM preparations usually contain more than one component and target multiple body systems for maximal therapeutic effects [7, 8]. As such, TCM may prove superior to other pharmacotherapies in the treatment of chronic and degenerative diseases such as PD [8]. The Jitai tablet (JTT) is a traditional herbal formula consisting of the following ingredients: Papaveraceae Corydalis (yan hu suo), 10.20%; Solanaceae Datura metel (yang jin hua), 2.18%; Lamiaeae Salvia Miltiorrhizae (Dan shen), 16.87%; Araliaceae Panax ginseng (Ren shen), 2.18%; Apiaceae Angelica sinensis (Dang gui), 10.20%; Apiaceae Ligusticum chuanxiong (Chuan xiong), 5.71%; Asteraceae Carthamus tinctorius (Hong hua), 10.20%; Ranunculaceae Aconitum (Fuzi), 2.18%; Myristicaceae Myristica cagayanensis (Dou kou), 2.18%; Asteraceae Aucklandia (Mu xiang), 5.71%; Thymelaeaceae Aquilaria, (Chen xiang), 4.35%; Zingiberaceae Zingiber (Gan jiang), 2.18%; Lauraceae Cinnamomum (Rou gui), 2.18%; Sapindaceae (Tao ren), 10.20%; Pearl powder (Zhen zhu fen), 13.47% [9, 10]. JTT has been long used clinically to treat neuropsychiatric disorders. Moreover, prior studies in our laboratory have demonstrated that JTT could upregulate DAT in patients [11]. It is well known that DAT level is a standard for diagnosis of PD and can be used for evaluating therapeutic efficacy in PD treatment. Therefore, it is very interesting to investigate the effects of JTT on PD.

The neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP) causes damage to dopaminergic neurons and has been widely used for generation of animal models of PD [12]. In the present study, we examined the effects of JTT on behavioral changes and characteristics of the DA system in an MPTP-induced PD mouse model. Madopar was used as a positive control drug for comparison.

2. Materials and Methods

2.1. Animals and Treatment Procedures. Male C57BL/6 mice aged 6–8 weeks (20–24 g) were randomly divided into the following groups: (1) control, (2) vehicle (distilled water), (3) high dose of JTT (0.50 g/kg/day, JTT-H), (4) middle dose of JTT (0.15 g/kg/day, JTT-M), (5) low dose of JTT (0.05 g/kg/day, JTT-L), and (6) Madopar (Roche) (0.12 g/kg/day) [13]. All groups except for the control group were given MPTP (Sigma) intraperitoneally four times at 2-hour interval at the dosage of 20 mg/kg on the 8th day of the 15-day experiment period [14]. JTT was dissolved in deionized water. The prepared suspension was intragastrically given to mice at dose of 0.50 g/kg/day, 0.15 g/kg/day, or 0.05 g/kg/day for 15 days (0.1 mL/10 g body weight). The Madopar group was given Madopar at a dose of 0.12 g/kg/day intraperitoneally for 7 days after injection of MPTP. All animals were maintained according to the international guidelines for care and use of laboratory animals. Experimental procedures involving animals were approved by the Ethics Committee of Beijing Normal University (BNU/EC/01/2011).

2.2. Behavioral Analyses and Brain Tissue Preparation. Six mice were randomly selected from each group and a number of behavioral tests were performed on the 9th day (1 day after MPTP) and the 15th day (7 days after MPTP).

2.2.1. Locomotor Activity Test. Locomotor activity was assessed in an automated activity chamber (30 cm diameter, 15 cm high) connected to an infrared tracking analyzer that transmitted the animals’ movement distance and velocity to a computer. The test was started by placing the mouse at the center of the chamber for two min for acclimatization, and then the total horizontal ambulatory route within the following 30 min was recorded by a computer analyzer.

2.2.2. Rotarod Test. To determine fore limb and hind limb motor coordination and balance, a rotarod test as described previously was used [15]. Mouse was placed on the rotating bar of the rotarod unit (DXP-3; Institute of Materia Medica, Chinese Academy of Medical Sciences) set to a rotation speed of up to 18 rpm during the test session. The time spent on the rotating bar, known as the latent period, was recorded. Performance was recorded as 120 s if the latent period exceeded 120 s.

2.2.3. Brain Tissue Preparation. Mice were anesthetized with sodium pentobarbital (50 mg/kg, i.p.) after the behavior test on the 15th day and rapidly perfusion fixed with saline, followed by 4% paraformaldehyde in 0.1 M phosphate buffer (pH 7.4). Then, the brains were removed and stored at ~80 °C until use. The frozen brains were cut into 18 μm coronal sections using a cryostat microtome (CM3000; Leica, Germany). Frozen sections of the striatum and substantia nigra were used for immunohistochemistry and autoradiography experiments.

2.3. DAT and Tyrosine Hydroxylase (TH) Immunohistochemical Staining. Analyses of DAT immunoreactivity in the striatum and TH immunoreactivity in the substantia nigra (SN) were conducted on brain sections as previously described [16]. Briefly, sections were incubated with either rat anti-DAT (Abcam) at 1:1000 dilutions or mouse anti-TH (Sigma) at 1:10000 dilutions at 4 °C overnight. Then, the slices were incubated with peroxidase-conjugated secondary antibody (ZSGB-BIO) for DAT and the VECTASTAIN ABC kit (VECTOR) for TH. All staining sections were visualized with 3,3′-diaminobenzidine tetrahydrochloride (DAB). The sections were counted using a bright-field microscope (M165FC, Leica) and optical densities (OD) of DAT and TH were calculated by Image-ProPlus software. TH-neurons were manually counted by researchers blinded to the treatment groups.

2.4. Autoradiography Experiments. The brains were divided into two hemispheres, one used for in vitro autoradiography and the other for analysis of striatal levels of DA and its metabolites [17]. Autoradiography was conducted on frozen cryostat-cut 18 μm thick brain slices to measure DAT and D2 binding according to the procedures reported previously [18].
Briefly, brain sections were incubated for 60 min with 50 pM of $^{[125I]}$-IBZM for $D_2$ receptor ($D_2$R) binding or with 50 pM of $^{[125I]}$-$\beta$-CIT and in the presence of 1 mM fluoxetine for DAT binding. Nonspecific binding was determined in the adjacent slices in the presence of 10 $\mu$M sulpiride ($D_2$R antagonist, Sigma-Aldrich Co., USA) for $D_2$R or 100 mM nomifensine (DAT antagonist, Sigma-Aldrich Co., USA) and 100 mM fluoxetine for DAT. Sections were then exposed to a super sensitive phosphor screen (PerkinElmer) for 2 h.

3. Results

3.1. Behavioral Analyses

3.1.1. Locomotor Activity. Locomotor activity testing is a commonly used technique for behavioral monitoring in PD animal models. In this study, it was used to evaluate motor impairment in the MPTP-treated animals. As shown

![Figure 1: JTT improved MPTP-induced locomotor activity deficits in PD mice. Test was done on the 9th day (1 day after injection of MPTP) and 15th day (7 days after injection of MPTP). All the data, total movement distance (a), and mean velocity (b), $n = 4 – 6$, were shown as mean $\pm$ S.E.M, $^*P < 0.05$, $^{**}P < 0.01$ versus vehicle group.](image1)

Densitometry analysis of autoradiographs was carried out with OptiQuant software (PerkinElmer). $^{[125I]}$-IBZM and $^{[125I]}$-$\beta$-CIT were prepared according to procedures described previously [17, 19].

2.5. Measurement of DA and Its Metabolites in the Striatum. Levels of DA and its metabolites 3,4-dihydroxyphenylacetic acid (DOPAC) and homovanillic acid (HVA) were measured by high performance liquid chromatography with an electrochemical detector (HPLC-ECD) as previously described [20]. Briefly, the striatum was dissected, weighed, and homogenized in 0.2 M ice-cold perchloric acid. The sample was then centrifuged twice at 14,000 g for 15 min at 4°C. The supernatant was collected and filtered through a 0.22 $\mu$m Millipore filter and injected into the HPLC system for analysis.

2.6. Statistical Analyses. Statistical analyses included one-way analysis of variance (ANOVA) followed by LSD for behavioral tests and Tukey’s HSD post hoc test for all other tests. The results of immunohistochemical staining and autoradiography were indicated as a ratio compared to the control group.
Figure 3: JTT protected against MPTP-induced reductions in TH expression in the mouse SN. C57BL/6 mice were treated with JTT for 7 days and then MPTP (20 mg/kg, 4 times, at 2-hour interval, i.p.) was injected on the 8th day. Mice were sacrificed on the 7th day after MPTP injection (15th day of the experiment period) and after behavioral tests. Photomicrographs of representative SN (a) sections stained with an antibody against TH. Reduced activity of TH-neurons was observed in MPTP treated mice, which was partially prevented by treatment with JTT. The number of TH-positive neurons in the SN (b) was expressed as the mean ± S.D, n = 5. *P < 0.05, **P < 0.01 versus vehicle group, ##P < 0.01 versus JTT-H.

in Figures 1(a) and 1(b), on the 9th day, all animals in MPTP-treated groups exhibited a significant reduction in total movement distance and mean velocity compared to those in the control group (P < 0.01). On the 15th day, there was a marked improvement in locomotor activity in mice treated with high dose JTT (JTT-H) compared to those treated with the vehicle, including both movement distance and mean velocity (P < 0.05). The group of mice treated with Madopar also demonstrated significant improvements compared with the vehicle group in movement distance and mean velocity (P < 0.01).

3.1.2. Rotarod Test. The restorative effects of JTT on MPTP-induced motor coordination deficiencies were tested using the rotarod test. As shown in Figure 2, on the 9th day, all mice injected with MPTP had a significant reduction in rotarod performance compared to control mice (P < 0.01). On the 15th day, a significant difference was observed between control and vehicle (P < 0.01) mice. The latent period significantly increased in the JTT-H and JTT-M groups (P < 0.01) and the Madopar group (P < 0.01) when compared with mice in the vehicle group. JTT effectively countered the MPTP-induced behavioral coordination deficiencies in a dose-dependent manner.

3.2. Effects of JTT on MPTP-Induced Decreases in TH Positive Neurons in the SN. Immunohistochemical staining was used to measure the levels of TH in the SN. As shown in Figures 3(a) and 3(b), MPTP exposure led to a remarkable loss of TH positive neurons in the SN (P < 0.01) compared to control mice. Treatment with JTT-H and JTT-M significantly halted TH-neuron reductions (JTT-H, P < 0.01; JTT-M, P < 0.05), while treatment with Madopar had no effect. This result showed that JTT significantly protected against MPTP-induced reduction in TH-neurons in the mouse SN while Madopar did not.

3.3. DAT in the Striatum Measured by Immunohistochemical Staining and Autoradiography. DAT is a key protein used in the clinical diagnosis of PD. In this study, immunohistochemical staining and autoradiography were used to detect DAT levels
in the striatum. As shown in Figures 4(a) and 4(b), MPTP treatment elicited a significant reduction in DAT levels in the striatum ($P < 0.01$). Treatment with JTT-H ($P < 0.01$) and JTT-M ($P < 0.05$) significantly protected against this reduction. A comparison between JTT and Madopar found that JTT-H was significantly superior in its protective/restorative effect ($P < 0.05$): DAT level increased by 18% in the JTT-H group, but only by 9% in the Madopar group when compared to the vehicle group.

Figures 4(c) and 4(d) illustrate the range of DAT levels after drug treatments via autoradiography using $^{[125]}$I-$\beta$-CIT. MPTP treatment led to a significant reduction in DAT binding in the striatum compared with control mice. This reduction was attenuated in mice treated with JTT, while treatment with Madopar showed no significant effect when compared to vehicle mice. These results indicate that JTT could significantly protect against MPTP-induced reductions in DAT levels in the mouse striatum and that JTT-H is more effective than Madopar.

3.4. Autoradiography Detected $D_2R$ in Striatum. DA receptors gradually decrease in the progression of PD over time and the $D_2R$ is one of the main receptors of the DA system. Autoradiography with $^{[125]}$I-IBZM was used to detect levels of $D_2R$ expression in the striatum. As shown in Figure 5, MPTP treatment elicited a significant reduction in $D_2R$ binding with $^{[125]}$I-IBZM in striatum compared with that of control mice ($P < 0.01$). This reduction in striatal $D_2R$ levels was attenuated in mice treated with JTT-H and JTT-M in a dose-dependent manner (JTT-H, $P < 0.01$; JTT-M, $P < 0.05$). Although Madopar also had a significant effect to increase $D_2$ levels ($P < 0.05$) in MPTP-treated animals, JTT-H had a stronger effect: JTT-H reversed 26% of the MPTP-induced reductions in $D_2$ binding, compared with 17% by Madopar.

3.5. Levels of DA and Its Metabolites in the Striatum. The levels of DA and its metabolites in the striatum were measured by HPLC-ECD. As shown in Figure 6(a), mice in
striatum is expressed as the mean ± S.D, \( n = 5 \). * \( P < 0.05 \), ** \( P < 0.01 \) versus vehicle group.

The MPTP vehicle group showed a remarkable depletion of DA, DOPAC, and HVA in the striatum compared with those in the control group (\( P < 0.01 \)). The ratio of DOPAC and HVA to DA (Figure 6(b)) was significantly higher (\( P < 0.01 \)) in the vehicle group than the control, which is indicative of increased DA metabolism in the vehicle group. Treatment with JTT effectively inhibited the increase in DOPAC and HVA to DA ratio (JTT-H, \( P < 0.01 \)) in a dose-dependent fashion, despite the fact that the three dosages of JTT did not significantly alter striatal levels of DA and its metabolites. The effect of inhibiting this increased ratio was stronger with JTT-H and JTT-M than with Madopar (\( P < 0.05 \)). However, Madopar appeared to increase the levels of DA, DOPAC, and HVA in the striatum (\( P < 0.05 \)).

4. Discussion

The present study investigated the ability of JTT to offer protection against the neurotoxicity of MPTP. Our findings indicated that high dose of JTT (JTT-H, 0.50 g/kg/day) effectively protected against the MPTP-induced deficiencies in behavioral activities, in an efficacy equal to that of Madopar. Moreover, we found that JTT could protect against the loss of TH-neurons in the SN. JTT also attenuated the reduction in DAT levels in the striatum induced by MPTP. Furthermore, JTT protected against the reduction in striatal D2 levels typically induced by MPTP in a dose-dependent manner.

We also found that JTT could attenuate the increase in DA metabolism rates in MPTP-treated mice. In general, these protective/restorative effects were stronger in animal treated with JTT-H than Madopar.

Movement deficits in PD result from dopaminergic dysfunction in the brain. In our study, JTT treatment clearly did improve MPTP-induced motor deficits. Notably, we found that JTT could inhibit the rate of DA metabolism while failing to significantly increase the concentrations of DA and its metabolites. One explanation for these findings may be that behavioral changes are not solely directly related to striatal DA levels, and multiple factors are likely to be at play [21]. As is well known, long-term treatment with Madopar induces many side-effects and these side-effects coincide with the rise and fall of DA levels in the brain that occur in the course of L-DOPA therapy [22]. As for JTT, preclinical toxicological studies have found no apparent toxicity even at high doses (e.g., 9.8 g/kg/day) in SD rats during long-term use (6 months) [23]. In the clinical setting JTT also demonstrated no obvious adverse effects [24–26]. In the present study, JTT was found to improve striatal DA levels only appreciably but effectively attenuate the increase in DA turnover rate. This may suggest that long-term treatment with JTT is effective and will not induce the same side-effects as Madopar.

PD is characterized by degeneration of DA neurons and corresponds to loss of DAT, a membrane protein important for DA reuptake. Decreasing DAT levels is the gold standard by which clinical diagnoses of PD are made using medical imaging methods [27, 28]. In the present study, we found that JTT protected against MPTP-induced DAT reduction in a dose-dependent manner. Relatively speaking, Madopar treatment has little effect on DAT levels. It has been reported that ginseng, which is an active ingredient found in JTT, may suggest that long-term treatment with JTT is effective and may result from the direct action of JTT on DAT expression.

The D2R is one of the key receptors associated with PD but remains controversial because levels of D2R expression may decrease in PD patients or not. Antonini et al. found a significant reduction in putamen D2 binding to \([^{11}C]\)raclopride in PD patients who had carried a diagnosis for 3–5 years [30]. Rinne et al. found that striatal D2 increased in early PD patients [31]. In this study, we found significant decline in striatal D2 binding to \([^{125}I]\)-IBZM in MPTP-treated mice. Meanwhile, JTT significantly inhibited this reduction, suggesting that JTT may play a role in reversing D2R decline, which may be beneficial to the treatment of PD.

In order to investigate the neuroprotective effect of JTT in MPTP-induced PD model, we treated mice with JTT before MPTP, in a paradigm similar to that used in other studies reviewed by More et al. [7]. JTT was found to inhibit the reduction in positive TH-neurons in SN typically observed in vehicle MPTP mice. This suggests that JTT might offer protection against dopaminergic neurodegeneration induced by the neurotoxicity of MPTP. Moreover, JTT upregulates the levels of DAT and D2R in the striatum. Some components in the JTT prescription have been reported to have protective
5. Conclusions

Evidence from the present study appears to indicate that JTT can play an important role in behavioral and motor function improvement in MPTP-induced PD model mice. This is related to the DA system, including DAT, D_2R, and TH. Therefore, JTT may be considered as a potential complementary therapeutic agent for PD treatment, because it not only provides relief of motor symptoms, but also protects neurons and involves the DA system to restore levels of DAT and D_2R.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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References

[1] A. Toulouse and A. M. Sullivan, “Progress in Parkinson’s disease—where do we stand?” Progress in Neurobiology, vol. 85, no. 4, pp. 376–392, 2008.

Figure 6: JTT inhibited the increase in the ratio of DOPAC and HVA to DA in the striatum of MPTP mice. MPTP (20 mg/kg, 4 times, at 2-hour interval, i.p.) was injected on the 8th day of 15-day experiment period. The contents of DA, DOPAC, and HVA were detected by HPLC-ECD and the ratio of (DOPAC + HVA) to DA was calculated. Data regarding DA, DOPAC, HVA (a), and (DOPAC + HVA)/DA ratios (b) are expressed as the mean ± S.D, n = 4-5. *P < 0.05, **P < 0.01 versus vehicle group.
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[2] S. Hunot, F. Boissière, B. Faucheux et al., “Nitric oxide synthase and neuronal vulnerability in Parkinson’s disease,” *Neuroscience*, vol. 72, no. 2, pp. 355–363, 1996.

[3] J. D. Elsworth and R. H. Roth, “Dopamine synthesis, uptake, metabolism, and receptors: relevance to gene therapy of Parkinson’s disease,” *Experimental Neurology*, vol. 144, no. 1, pp. 4–9, 1997.

[4] T. Nagatsu and M. Sawadab, “L-DOPA therapy for Parkinson’s disease: past, present, and future,” *Parkinsonism and Related Disorders*, vol. 15, no. 1, pp. S3–S8, 2009.

[5] C. A. Davie, “A review of Parkinson’s disease,” *British Medical Bulletin*, vol. 86, no. 1, pp. 109–127, 2008.

[6] S. Fahn, “Levodopa and the progression of Parkinson’s disease,” *The New England Journal of Medicine*, vol. 351, no. 24, pp. 2498–2566, 2004.

[7] S. V. More, H. Kumar, S. M. Kang et al., “Advances in neuroprotective ingredients of medicinal herbs by using cellular and animal models of Parkinson’s disease,” Evidence-Based Complementary and Alternative Medicine, vol. 2013, Article ID 957875, 15 pages, 2013.

[8] X. Z. Li, S. N. Zhang, S. M. Liu et al., “Recent advances in herbal medicines treating Parkinson’s disease,” *Fitoterapia*, vol. 84, pp. 273–285, 2013.

[9] S.-P. Wang, L. Liu, L.-L. Wang et al., “Screening and analysis of the multiple absorbed bioactive components and metabolites in rat plasma after oral administration of Jitai tablets by high-performance liquid chromatography/diode-array detection coupled with electrospray ionization tandem mass spectrometry,” *Rapid Communications in Mass Spectrometry*, vol. 24, no. 11, pp. 1641–1652, 2010.

[10] S. P. Wang, L. Liu, L. L. Wang et al., “Structural characterization and identification of major constituents in Jitai tablets by high-performance liquid chromatography/diode-array detection coupled with electrospray ionization tandem mass spectrometry,” *Molecules*, vol. 17, no. 9, pp. 10470–10493, 2012.

[11] Y. Liu, M. Han, X. D. Liu et al., “Dopamine transporter availability in heroin-dependent subjects and controls: longitudinal changes during abstinence and the effects of Jitai tablets treatment,” *Psychopharmacology*, vol. 230, no. 2, pp. 235–244, 2013.

[12] S. Przedborski and M. Vila, “MPTP: a review of its mechanisms of neurotoxicity,” *Clinical Neuroscience Research*, vol. 1, no. 6, pp. 407–418, 2001.

[13] X. M. Li, H. B. Ma, Z. Q. Ma et al., “Ameliorative and neuroprotective effect in MPTP model of Parkinson’s disease by Zhen-Wu-Tang (ZWT), a traditional Chinese medicine,” *Journal of Ethnopharmacology*, vol. 130, no. 1, pp. 19–27, 2010.

[14] L. V. Jackson and S. Przedborski, “Protocol for the MPTP mouse model of Parkinson’s disease,” *Nature Protocols*, vol. 2, no. 1, pp. 141–151, 2007.

[15] H. N. Zhang, C. N. An, H. N. Zhang, and X. P. Pu, “Protocatechuic acid inhibits neurotoxicity induced by MPTP in vivo,” *Neuroscience Letters*, vol. 474, no. 2, pp. 99–103, 2010.

[16] Y. C. Chung, S. R. Kim, J.-Y. Park et al., “Fluoxetine prevents MPTP-induced loss of dopaminergic neurons by inhibiting microglial activation,” *Neuropharmacology*, vol. 60, no. 6, pp. 963–974, 2011.

[17] Z. Chen, X. Zhou, C. Y. Wu et al., “Synthesis and iodine labelling of β-CIT as a dopamine transporter imaging agent,” *Journal of Isotopes*, vol. 13, no. 1, pp. 7–11, 2000.

[18] Z. Xiong, G. Xu, Z. P. Chen et al., “Regulation of a sapogenin from Rhizoma Anemarrhenae, ZMR on dopaminergic system in a chronic mude of Parkinson’s disease,” *Chinese Journal of Nuclear Medicine*, vol. 29, no. 5, pp. 331–334, 2009.

[19] M. P. Kung, B. L. Liu, Y. Y. Yang, J. J. Billings, and H. F. Kung, “A kit formulation for preparation of iodine-123-IBZM: a new CNS D-2 dopamine receptor imaging agent,” *Journal of Nuclear Medicine*, vol. 32, no. 2, pp. 339–342, 1991.

[20] D. Luo, Q. Zhang, H. Wang et al., “Fucoidan protects against dopaminergic neuron death in vivo and in vitro,” *European Journal of Pharmacology*, vol. 617, no. 1–3, pp. 33–40, 2009.

[21] E. Rousselet, C. Joubert, J. Callebert et al., “Behavioral changes are not directly related to striatal monoamine levels, number of nigral neurons, or dose of parkinsonian toxin MPTP in mice,” *Neurobiology of Disease*, vol. 14, no. 2, pp. 218–228, 2003.

[22] H. Iderberg, V. Francardco, and E. Y. Pioli, “Animal models of L-DOPA-induced dyskinesia: an updat on the current options,” *Neuroscience*, vol. 211, pp. 13–27, 2012.

[23] Y. L. Chen, B. Y. Yan, and Y. P. Deng, “Pharmacology of Jitai tablets, a traditional Chinese medicine for detoxification,” *Chinese Journal of New Drug*, vol. 19, no. 14, pp. 1229–1233, 2010.

[24] Z. Li, Y. Tang, S. Hua et al., “Efficacy of Jitai for protracted withdrawal symptoms in heroin dependent patients,” *Chinese Journal of Drug Dependence*, vol. 16, no. 4, pp. 284–287, 2007.

[25] G. Xu, L. Duan, Z. Wang et al., “A double-blind clinical trial of Jitai tablets vs Lofexidine in the treatment of heroin withdrawal symptoms,” *Chinese Journal of Drug Dependence*, vol. 9, no. 4, pp. 273–277, 2000.

[26] S. Q. Hao, M. Zhao, R. W. Zhang et al., “The effectiveness comparison of Jitai tablets versus methadone in community-based drug treatment: a 1 year follow-up study,” *Addictive Behaviors*, vol. 38, no. 10, pp. 2596–2600, 2013.

[27] A. Winogrodzka, P. Bergmans, J. Booij, E. A. Van Royen, J. C. Stoof, and E. C. Wolters, “[123]β-CIT SPECT is a useful method for monitoring dopaminergic degeneration in early stage Parkinson’s disease,” *Journal of Neurology Neurosurgery and Psychiatry*, vol. 74, no. 3, pp. 294–298, 2003.

[28] B. Ravina, K. Marek, S. Eberly et al., “Dopamine transporter imaging is associated with long-term outcomes in Parkinson’s disease,” *Movement Disorders*, vol. 27, no. 11, pp. 1392–1397, 2012.

[29] L. Jia, Y. Zhao, and X.-J. Liang, “Current evaluation of the millennium phytotherapy-ginseng (II): Collected chemical entities, modern pharmacology, and clinical applications emanated from traditional Chinese medicine,” *Current Medicinal Chemistry*, vol. 16, no. 22, pp. 2924–2942, 2009.

[30] A. Antonini, J. Schwarz, W. H. Oertel, O. Pogarell, and K. L. Leenders, “Long-term changes of striatal dopamine D2 receptors in patients with Parkinson’s disease: a study with positron emission tomography and [11C]raclopride,” *Movement Disorders*, vol. 12, no. 1, pp. 33–38, 1997.

[31] U. K. Rinne, A. Laihinen, J. O. Rinne, K. Nagren, J. Bergman, and U. Ruotsalainen, “Positron emission tomography demonstrates dopamine D2 receptor supersensitivity in the striatum of patients with early Parkinson’s disease,” *Movement Disorders*, vol. 5, no. 1, pp. 55–59, 1990.

[32] H. Kim, “Neuroprotective herbs for stroke therapy in traditional eastern medicine,” *Neurological Research*, vol. 27, no. 3, pp. 287–301, 2005.

[33] D. O. Kennedy and A. B. Scholey, “Ginseng: potential for the enhancement of cognitive performance and mood,” *Pharmacology Biochemistry and Behavior*, vol. 75, no. 3, pp. 687–700, 2003.
[34] B. G. Park, H. J. Jung, Y. W. Cho et al., “Potentiation of antioxidative and anti-inflammatory properties of cultured wild ginseng root extract through probiotic fermentation,” *Journal of Pharmacy and Pharmacology*, vol. 65, no. 3, pp. 457–464, 2013.

[35] J. Wang, H.-M. Xu, H.-D. Yang, X.-X. Du, H. Jiang, and J.-X. Xie, “Rg1 reduces nigral iron levels of MPTP-treated C57BL6 mice by regulating certain iron transport proteins,” *Neurochemistry International*, vol. 54, no. 1, pp. 43–48, 2009.

[36] H. Tsukada, N. Harada, S. Nishiyama, H. Ohba, and T. Kakiuchi, “Cholinergic neuronal modulation alters dopamine D2 receptor availability in vivo by regulating receptor affinity induced by facilitated synaptic dopamine turnover: positron emission tomography studies with microdialysis in the conscious monkey brain,” *Journal of Neuroscience*, vol. 20, no. 18, pp. 7067–7073, 2000.