CASE REPORT

Castleman’s Disease with Cutaneous Involvement Manifesting as Multiple Violaceous Plaques on Entire Body

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Castleman’s disease (CD) is an uncommon B-cell lymphoproliferative disorder characterized by lymph node hyperplasia with vascular proliferation. Cutaneous involvement in CD is rare. A 65-year-old man presented with a 7-year history of gradually developing multiple reddish to violaceous indurated plaques on the scalp, trunk, and legs. On physical examination, there were palpable enlarged cervical, axillary, and inguinal lymph nodes. Laboratory examination revealed anemia, thrombocytosis, hyperproteinemia, hypoalbuminemia, and polyclonal hypergammaglobulinemia. An inguinal lymph node biopsy and a skin biopsy were performed and the patient was diagnosed with the plasma cell type of CD. Chemotherapy was started and the lesions have responded to treatment. (Ann Dermatol 23(S2) S169 ~ S174, 2011)

Keywords-
Angiofollicular lymphoid hyperplasia, Castleman’s disease, Giant lymph node hyperplasia, Skin

INTRODUCTION

Castleman’s disease (CD) is an angiofollicular lymphoid hyperplasia, or giant lymph node hyperplasia. It is characterized by lymph node hyperplasia and vascular proliferation. It was first described in 1956 by Castleman et al. as a localized lymphadenopathy that usually involves mediastinum or pulmonary hilus resembling thymoma. Keller et al. divided CD histologically into three types: hyaline-vascular, plasma cell, and mixed type. Several associated diseases of CD have been described, including paraneoplastic pemphigus, thrombotic thrombocytopenic purpura, amyloidosis, and Kaposi sarcoma. A few cases of CD with cutaneous involvement have been reported. However, there are no reports in the Korean literature. Here, we report a case of CD which showed a characteristic cutaneous manifestation of multiple violaceous plaques with multiple lymphadenopathy that responded well to chemotherapy.

CASE REPORT

In May 2008, a 65-year-old man presented with multiple violaceous plaques which had developed over the previous 7 years. Lesions started as small violaceous macules on left axilla, and then spread to the scalp, trunk, and legs. They slowly formed confluent indurated plaques. He had pulmonary tuberculosis and tuberculosis lymphadenitis which were diagnosed by open lung biopsy 4 years prior and had been treated with oral medication. His family history was non-contributory. On physical examination, variable sized reddish to violaceous indurated plaques were observed on his scalp, chest, back,
Fig. 1. Multiple reddish to violaceous indurated plaques on both flanks (A, B) and back (C).

Fig. 2. (A) Lymph node biopsy shows multiple follicles with vascular proliferation (H&E, original magnification ×100), (B) Many plasma cells in the interfollicular area with Russell bodies (arrow) (H&E, ×400).

axilla, and both legs (Fig. 1). Superficial lymph nodes were palpable in his neck, axilla, and inguinal areas. A previous operative scar was observed on his right side of chest. He has been suffered from night sweats for several years. However, there were no other systemic symptoms such as fever, weight loss, or asthenia. On laboratory examination, we found anemia (Hgb 9.4 g/dl, normal range: 13.5~17.5), thrombocytosis (platelet $555 \times 10^3$/mm³, normal range: 150~350×10³), increased erythrocyte sedimentation rate (ESR) (105 mm/hr, normal range: 0~17), hyperproteinemia (total protein 11.4 g/dl, normal range: 5.5~8.0), and polyclonal hypergammaglobulinemia (IgG 4,030 mg/dl, normal range: 614~1,293; IgM

Fig. 3. Skin biopsy shows multiple lymphoid follicles as nodular infiltration in the dermis (H&E, ×20).
Fig. 4. (A) Lymphoid follicles are composed of numerous lymphocytes and plasma cells (H&E, ×400). Immunohistochemical stains are positive for CD 20 (B, ×100), CD 79a (C, ×200), and bcl-2 (D, ×200).

547 mg/dl, normal range: 53 ~ 334). Hepatitis B antigen and anti-human immunodeficiency virus (HIV) antibody were negative. A peripheral blood smear showed a mild degree of microcytic hypochromic anemia with rouleaux formation and thrombocytosis without definite abnormal cells. A bone marrow biopsy revealed a small increase of mature plasma cells (approximately 7.3%). There were multiple hypermetabolic lymph nodes on his neck, chest, and abdomen. Splenomegaly was determined by positron emission tomography-computed tomography but there was no other internal solid organ involvement. An excisional biopsy from the left inguinal lymph node showed follicular formation with many lymphocytes and dense plasma cells (Fig. 2). A skin biopsy taken from a right flank lesion revealed variable sized lymphoid follicles in the dermis (Fig. 3) and nodular infiltrates of lymphocytes, lymphoplasmacytoid cells, and many plasma cells (Fig. 4A). In the immunohistochemical stain, lymphocytes and plasma cells stained positively for CD 20, CD 79a, CD 5, CD 45RO, and bcl-2 (Fig. 4B × D). CD3 was weakly positive in the reactive T cells. CD10 and bcl-6 were negative. Kappa and lambda light chain were equally stained. These findings were consistent with multicentric plasma cell type of CD. He was treated with 8 cycles of CHOP chemotherapy (cyclophosphamide, adriamycin, vincristine, and prednisone). The size and hypermetabolic activities of lymph nodes decreased and skin lesions were much improved with mild hyperpigmentation. He remains healthy 2 years after treatment.

**DISCUSSION**

CD is an uncommon B-cell lymphoproliferative disorder.
Table 1. Summary of reported cases of Castleman’s disease with cutaneous involvement

| Author                | Age (yrs) | Clinical features                              | Location    | Histologic type      |
|-----------------------|-----------|-----------------------------------------------|-------------|----------------------|
| Kubota et al.         | 57/M      | Multiple brownish indurated nodules            | Face, trunk | Plasma cell type     |
| Skelton and Smith     | 72/F      | Multiple nodules                               | Trunk       | Plasma cell type     |
| Klein et al.          | 44/F      | Multiple erythematous and violaceous nodules   | Trunk, extremity | Plasma cell type     |
| Kayasut et al.        | 71/M      | Multiple reddish macules and plaques           | Trunk       | Plasma cell type     |
| Ahmed et al.          | 42/F      | Multiple erythematous nodules and plaques      | Trunk       | Plasma cell type     |
| Okuyama et al.        | 45/F      | Multiple brownish infiltrated nodules and plaques | Trunk       | Plasma cell type     |
| Higashi et al.        | 39/F      | Multiple reddish to brownish plaques           | Face, trunk | Plasma cell type     |
| Tomasini et al.       | 56/F      | Multiple reddish to violaceous indurated plaques | Face, trunk, extremity | Plasma cell type     |
| Our case              | 65/M      | Multiple reddish to violaceous indurated plaques | Scalp, trunk, extremity | Hyaline vascular type |
|                       |           |                                               |             |                      |

M: male, F: female.

The time to diagnosis the disease is often long, which is attributable to clinical polymorphism and poor awareness of the disease. CD can occur at any age, even during childhood, with peak frequency during adulthood. There are two clinical forms of CD: unicentric and multicentric. The onset age of multicentric CD is older than that of unicentric CD. There are no gender differences in unicentric CD but males have multicentric CD more often than females. The unicentric (solitary) form commonly presents as a mass in mediastinum. It can also be found outside the thorax (both nodal and extranodal), for example on the neck, arm, abdomen, vulva, and pericardium. The multicentric form shows generalized lymphadenopathy and various systemic symptoms. It may also be associated with POEMS syndrome.

Histologic variants of CD are composed of hyaline-vascular (90%), plasma cell (10%), and a mixed type. The hyaline-vascular type represents 80 to 90% of all cases of the solitary form. The multicentric form consists almost entirely of plasma cell type. The hyaline-vascular type lacks the systemic signs and/or symptoms which are commonly associated with plasma cell types. Histopathologically, it is characterized by small, concentrically whorled, lymphoid follicles surrounded by small lymphocytes arranged in a concentric, onion-skin pattern. Extensive capillary proliferation and hyalinized sclerotic collagen are present between follicles. The plasma cell type shows many systemic signs and/or symptoms including fever, anemia, weight loss, loss of appetite, elevated ESR, hypergammaglobulinemia, and hypoalbuninemia. It reveals as well developed hyperplastic secondary lymphoid follicles and many plasma cells exist predominantly in the interfollicular areas. Occasionally, there are numerous Russell bodies, deposition of an amorphous acidophilic material that probably contains fibrin, and immune complexes in the center of follicles.

Cutaneous involvement of CD has rarely been reported in the multicentric form. Among ten reported cases, three patients were male and seven patients were female. Their mean age was 50.8 ± 12.7 yrs (range: 36 ~ 72 yrs). All skin lesions manifested as multiple erythematous to brownish nodules and plaques predominantly on the back but also on the face, trunk, and extremities. Histologic features were plasma cell type except in one case. Results are summarized in the Table 1. In our case, the patient showed multiple reddish to violaceous indurated patches on his scalp, chest, back, axilla, and both legs, which is similar to other cases. A skin biopsy and a lymph node biopsy showed nodular aggregates of many lymphocytes and plasma cells with Russell bodies in the dermis. Immunohistochemical staining for kappa and lambda light chains confirmed the polyclonal nature of the plasma cell infiltrate. These findings were compatible with plasma cell type CD.

The pathogenesis of CD is still not understood. Interleukin (IL)-6 is an important factor in the pathogenesis of CD, especially in the multicentric form. IL-6 contributes to proliferation and maturation of B lymphocytes into mature plasma cells and proliferation of blood vessels. Yoshizaki et al. showed that the cells present in the germinal center of the enlarged lymph nodes produce IL-6 and there is close relationship between the serum levels of IL-6 and the clinical symptoms of patients with CD. Recently, the incidence of CD in HIV patients has increased. Epidemiological and polymerase chain reaction studies have suggested that human herpes virus (HHV)-8 is associated with multicentric CD in all HIV positive patients and in almost 50% of HIV negative patients. HHV-8, known as Kaposi sarcoma associated herpes virus, also produces a functional analogue of IL-6. Although the exact pathogenesis of CD is unknown, immune dysregulation of IL-6 may play a key role.
addition, CD is associated with high levels of IL-10, tumor necrosis factor-β, interferon-γ, vascular endothelial growth factor, and macrophage colony-stimulating factor. In our patient, we could not check IL-6 expression or HHV-8 DNA sequences.

The differential diagnosis of CD includes angiolymphoid hyperplasia with eosinophilia (ALHE), Kimura disease, sinus histiocytosis with massive lymphadenopathy, cutaneous plasmacytoma, angioimmunoblastic T cell lymphoma, primary cutaneous marginal zone lymphoma, and primary cutaneous follicular center cell lymphoma. CD can be distinguished from these diseases by combined generalized lymphadenopathy, clinical manifestations, type and patterns of infiltrate cells, and immunohistochemical stain, especially polyclonal restriction of kappa and lambda light chains.

The treatment differs depending on clinical types. Surgical excision is effective for unicentric CD. This treatment allows full recovery without relapse in almost all cases. On the other hand, there is no consensus for the treatment of the multicentric type. A variety of approaches, including surgery, chemotherapy, radiation therapy, and anti-viral therapies, have been attempted. The treatment of choice is a monthly combination chemotherapy such as CHOP (cyclophosphamide, vincristine, doxorubicin, prednisone) or AVBD (etoposide, ifosfamide, cisplatin alternating with doxorubicin, vincristine, bleomycin and dacarbazine). Other therapies, such as interferon-α, retinoid acid or rituximab or anti-IL-6 antibodies, have been used and gave promising results. Generally, the prognosis of multicentric CD is poor, but our patient had 8 cycles of CHOP chemotherapy and responded well to treatment. In conclusion, we report a rare case of cutaneous involvement of multicentric CD which had a good prognosis. However, further long-term follow up for detection of recurrence is needed.

REFERENCES

1. McCarthy MJ, Vukelja SJ, Banks PM, Weiss RB. Angiofollicular lymph node hyperplasia (Castleman’s disease). Cancer Treat Rev 1995;21:291-310.
2. Keller AR, Hochholzer L, Castleman B. Hyaline-vascular and plasma-cell types of giant lymph node hyperplasia of the mediastinum and other locations. Cancer 1972;29:670-683.
3. Castleman B, Iversen L, Menendez VP. Localized mediastinal lymph node hyperplasia resembling thymoma. Cancer 1956;9:822-830.
4. Chorzelski T, Hashimoto T, Maciejewska B, Amagai M, Anhalt GJ, Jablonska S. Paraneoplastic pemphigus associated with Castleman tumor, myasthenia gravis and bronchiolitis obliterans. J Am Acad Dermatol 1999;41:393-400.
5. Couch WD. Giant lymph node hyperplasia associated with thrombotic thrombocytopenic purpura. Am J Clin Pathol 1980;74:340-344.
6. Altiparmak MR, Pamuk GE, Pamuk ON, Doğusoy G. Secondary amyloidosis in Castleman’s disease: review of the literature and report of a case. Ann Hematol 2002;81:336-339.
7. Liu W, Lacouture ME, Jiang J, Kraus M, Dickstein J, Soltani K, et al. KSHV/HHV8-associated primary cutaneous plasmablastic lymphoma in a patient with Castleman’s disease and Kaposis’ sarcoma. J Cutan Pathol 2006;33 Suppl 2:46-51.
8. Kubota Y, Noto S, Takakuwa T, Tadokoro M, Mizoguchi M. Skin involvement in giant lymph node hyperplasia (Castleman’s disease). J Am Acad Dermatol 1993;29:778-780.
9. Skelton HG, Smith KJ. Extranasal multicentric Castleman’s disease with cutaneous involvement. Mod Pathol 1998;11:93-98.
10. Klein WM, Rencic A, Munshi NC, Nousari CH. Multicentric plasma cell variant of Castleman’s disease with cutaneous involvement. J Cutan Pathol 2004;31:448-452.
11. Hayasat K, Le Tourneau A, Rio B, Bost-Bezeaud F, Heuberger L, Diebold J, et al. Multicentric Castleman’s disease with cutaneous plasmacytosis and systemic plasmacytosis: the same entity? Histopathology 2006;49:557-558.
12. Ahmed B, Tschen JA, Cohen PR, Zaki MH, Rady PL, Tying SK, et al. Multicentric castleman’s disease responds to anti interleukin-6 treatment. Mol Cancer Ther 2007;6:2386-2390.
13. Okuyama R, Harigae H, Moriya T, Tagami H, Ichinohasama R, et al. Indurated nodules and plaques showing a dense plasma cell infiltrate as a cutaneous manifestation of Castleman’s disease. Br J Dermatol 2007;156:174-176.
14. Higashi Y, Kanekura T, Sakamoto R, Mochitomi Y, Kanzaki T. Multicentric Castleman disease with cutaneous manifestations: report of 2 cases and comparison with systemic plasmacytosis. Dermatology 2007;214:170-173.
15. Tomasini D, Zampatti C, Serio G. Castleman’s disease with numerous mantle zone lymphocytes with clear cytoplasm involving the skin: case report. J Cutan Pathol 2009;36:887-891.
16. Sleater J, Mullins D. Subcutaneous Castleman’s disease of the wrist. Am J Dermatopathol 1995;17:174-178.
17. Dham A, Peterson BA. Castleman disease. Curr Opin Hematol 2007;14:354-359.
18. Casper C. The aetiology and management of Castleman disease at 50 years: translating pathophysiology to patient care. Br J Haematol 2005;129:3-17.
19. Hiran T, Yasukawa K, Harada H, Taga T, Watanabe Y, Matsuda T, et al. Complementary DNA for a novel human interleukin (BSF-2) that induces B lymphocytes to produce immunoglobulin. Nature 1986;324:73-76.
20. Yoshizaki K, Matsuda T, Nishimoto N, Kuritani T, Taeo H, Aozasa K, et al. Pathogenic significance of interleukin-6 (IL-6/BSF-2) in Castleman’s disease. Blood 1989;74:1360-1367.
21. Soulier J, Grollet L, Oksenhendler E, Cacoub P, Ca
Hatem D, Babinet P, et al. Kaposi’s sarcoma-associated herpesvirus-like DNA sequences in multicentric Castleman’s disease. Blood 1995;86:1276-1280.

22. Gessain A, Sudaka A, Brière J, Fouchard N, Nicola MA, Rio B, et al. Kaposi sarcoma-associated herpes-like virus (human herpesvirus type 8) DNA sequences in multicentric Castleman’s disease: is there any relevant association in non-human immunodeficiency virus-infected patients? Blood 1996;87:414-416.

23. Pavlidis NA, Briassoulis E, Klouvas G, Bai M. Is interferon-α an active agent in Castleman’s disease? Ann Oncol 1992; 3:85-86.

24. Rieu P, Droz D, Gessain A, Grünfeld JP, Hermine O. Retinoic acid for treatment of multicentric Castleman’s disease. Lancet 1999;354:1262-1263.