The H19/let-7 Feedback Loop Contributes to Developmental Dysplasia and Dislocation of the Hip

Bo NING¹, Rui JIN², Dahui WANG¹, Jun SUN²

¹Department of Pediatric Orthopaedics, Children’s Hospital of Fudan University, Shanghai, China, ²Department of Pediatric Orthopaedics, Children’s Hospital of Anhui Medical university, Hefei, Anhui, China.

Received March 29, 2018
Accepted October 16, 2018
Epub Ahead of Print January 10, 2019

Summary
Developmental dysplasia and dislocation of the hip (DDH) is the most common type of lower limb deformity in pediatric orthopedics. The mechanism of the signaling pathway has been studied in depth. However, the role of epigenetic regulation, such as lncRNA, is still far from clear. In this study, we successfully established a rat model of DDH and demonstrated that H19 was down-regulated in the development of DDH. Further, we constructed H19 knockdown (KD) and overexpression chondrocytes. H19 KD suppressed the proliferation of normal chondrocytes, while overexpression of H19 promoted cell proliferation of DDH chondrocytes. Finally, we revealed that H19 bound to let-7 and inhibited its function, acting as a competing endogenous RNA. Down-regulation of H19 is closely associated with DDH progression and H19 is an important epigenetic factor that regulates the proliferation of chondrocytes. H19 may thus be a potential clinical marker for DDH diagnosis and treatment.

Key words
Developmental dysplasia and dislocation of hip • H19 • Proliferation • let-7

Corresponding author
Jun Sun, Department of Pediatric Orthopaedics, Children’s Hospital of Anhui Medical university, Hefei, Anhui, China. E-mail: sunjun507@126.com

Introduction
Developmental dysplasia and dislocation of the hip is the most common type of lower limb deformity in pediatric orthopedics (Senaran et al. 2007). The vast majority of patients can be diagnosed early, and with application of a brace or surgical treatment can be completely cured. However, there are still a few patients who suffer re-dislocation, residual hip dysplasia, residual acetabular dysplasia or osteoarthritis (Carney et al. 2005, Thomas et al. 2007). Most of these patients have to undergo total hip arthroplasty before the age of 50, causing the patients themselves and their family great psychological and economic burden (Ashraf et al. 2014). In-depth analysis of the molecular mechanism underlying development of this disease and study of the proteins or genes that play key roles in this process should help with screening and identification of these patients.

Hip cartilage is mainly composed of chondrocytes and their extracellular matrix. Chondrocytes differentiate from mesenchymal stem cells. Extracellular matrix, which is secreted by chondrocytes, mainly consists of type II collagen and polysaccharide. These components together fight against joint stress and maintain normal joint shape and function (Bo et al. 2012). It has been shown that dysplasia of cells and changes in extracellular matrix components occur in the articular cartilage in DDH. These changes are closely related to cell proliferation, apoptosis and metabolism in chondrocytes (Zhang et al. 2015, Wei et al. 2016). The Wnt/β-catenin pathway plays a critical role in the development of osteoarthritis (Krase et al. 2014). Previous studies also show that overexpression of β-catenin induces chondrocyte dedifferentiation, while significantly inhibiting chondrocyte proliferation and promoting...
apoptosis (Pei et al. 2014). Although the mechanism of DDH and the signaling pathways involved has been studied in depth, research into the epigenetic regulation, such as lncRNA, is still lacking.

Long non-coding RNAs (lncRNAs), which act as epigenetic regulators, are widely involved in the regulation of gene expression at the levels of chromosome modification, transcription, post-transcriptional processing and translation (Li et al. 2014). Normal expression of lncRNA is necessary for normal cell function and individual development, while its abnormal expression is often associated with disease (Kazemzadeh et al. 2015). For example, tissue-specific expression of lncRNA PCAT-1, which regulates expression of a series of tumor-related genes such as CENPF, BRCA2 and CENPE, plays a key role in the oncogenesis of prostate cancer (Prensner et al. 2014). Recently, there have been increasing reports of the effects of lncRNA in osteoblast and chondrocyte proliferation and differentiation. For example, lncRNA UFC1 has been shown to promote the proliferation of chondrocytes by inhibiting the expression of miR-34a (Zhang et al. 2016). High expression of lncRNA HOTAIR promotes the expression of MMP family proteins such as MMP2 / MMP7 / MMP9 and further induces chondrocyte apoptosis (Zhang et al. 2016). Although much research on lncRNA and chondrocytes has been reported, the relationship between lncRNA and DDH has not been studied in sufficient detail. Due to the difficulty of obtaining clinical cartilage, we therefore studied DDH using an animal model. In this study, we successfully constructed a rat DDH model and used this to analyze lncRNA expression. We found that H19 is down-regulated in our DDH model and we used this to explore the function and mechanism of H19 in DDH.

**Methods**

**Rat model of developmental dysplasia of the hip and hip joint collection**

All the animals used in experiments were maintained according to the National Institutes of Health guide for the care and use of laboratory animals (NIH Publications No. 8023, revised 1978) and protocols were approved by the Institutional Ethics Committee of the Children’s Hospital of Fudan University (Shanghai, China). Twenty pregnant Wistar rats (8 weeks, 200-250 g) were purchased from Shanghai Laboratory Animal Co., Ltd., Shanghai, China. Forty male neonates and 1-day old pups were used in this study and were randomly divided into two groups. The DDH model was created by fixation of the hind limbs in an extension and adduction position with medical tape from day 1 to day 12, and normal rats of the same age constituted the control group. During the period of fixation, the model rats were fed by their mothers along with the control group. At different days, sacrifice was performed by means of overdose of general anesthetic. Total hips were excised immediately for subsequent procedures.

**Safranin O staining**

For histochemical staining, the isolated hips were fixed in 4 % paraformaldehyde for 24 h. The hips were then decalcified in 10 % ethylene diamine tetraacetic acid until completely demineralized and embedded in paraffin. After sectioning, the samples were deparaffinized with xylene and the safranin O staining process was performed according to a standard protocol. All the chemical reagents used in this experiment were bought from Sinopharm Chemical Reagent Company, Shanghai, China.

**Isolation of primary chondrocytes**

Primary rat chondrocytes were isolated from rat hips. The acetabulum cartilage from normal rats was isolated and rinsed twice with PBS. The isolated cartilage pieces were incubated in a 10 cm dish with 10 ml of collagenase solution at 0.5 mg/ml overnight at 37 °C under 5 % CO2. Then the suspension of isolated cells was filtered through a sterile cell strainer (BD Falcon™, San Jose, CA, USA) by setting over a 50 ml tube and centrifuging for 5 min. Finally, the rat chondrocytes were cultured in high-glucose Dulbecco’s modified Eagle’s medium (Gibco, Grand Island, NY, USA) supplemented with 10 % fetal bovine serum (FBS) (Gibco) and 1 % penicillin/streptomycin (HyClone, Logan, UT, USA) in a cell incubator at 37 °C with 5 % CO2 (Thermo Fisher Scientific, Waltham, NJ, USA).

**Immunofluorescence staining**

The chondrocytes were fixed in 4 % paraformaldehyde and then permeabilized with 0.1 % Triton X-100. The cells were blocked with 10 % FBS in PBS and incubated with BrdU antibody overnight at 4 °C (1:1000, ab8152, Abcam, Cambridge, UK). Next day the cells were rinsed, then incubated with anti-rabbit secondary antibody (1:1000, ab150116, Abcam) for 1 h at room temperature in the dark. After washing, the cells
were stained with DAPI for 5 min and then images were captured using a fluorescence microscope (TE2000, Nikon, Tokyo, Japan).

Plasmids, lentivirus supernatants and infection of chondrocytes

The shH19 (H19 short hairpin RNA (shRNA) knockdown) sequences were as follows: Shscramble-F, gatccCAGCGCTGACAACAGTTTCATCGTCGAGATGAAACTGTTGTCAGCGCTGTTTTTg; shscramble-R, aattcAAAAACAGCGCTGACAACAGTTTCATATATCTCATGAAACTGTTGTCAGCGCTGg; shH19 -1F, gatccGACTGAGGGGCTAGCTCGGTCAAGAGACCGAGCTAGCCCCTCAGTCTTTTTg; shH19 -1R, aattcAAAAAGACTGAGGGGCTAGCTCGGTCTCTTGAAAGGTCAGCTGCTTCTTCTCg; shH19 -2F, gatccGAGAAGAAGCAGCTGACCTTTCAAGAGAAGGTCAGCTGCTTCTTCTTTTTg; shH19 -2R, aattcAAAAAGAGAAGAAGCAGCTGACCTCTTCTTCTCg.

The shH19 sequences were ligated into the PLVX-shRNA2 vector which expressed a classic scrambled shRNA and green fluorescent protein (ZsGreen). The rat H19 sequences were obtained from NCBI. H19 PF: CCGGAATTCGGGGGTGGGGGGTAATGGGGAAAC; H19 PR: CCGTCAGGTCTCTGCGGTCTCTTGGT. The H19 sequences were ligated into the pLVX-IRES-ZsGreen1 vector which expressed the green fluorescent protein (ZsGreen). The lentivirus was packaged with 293T cells and then infection the chondrocytes. Next, the cells were selected through Flow cytometry using FACS Calibur (BD Biosciences). Then, ZsGreen positive cells expressing ZsGreen protein were observed using fluorescence microscopy (TE2000, Nikon, Japan).

Cell proliferation assay

The Cell Counting Kit-8 (CCK-8) reagent detects the number of living cells and is widely used for cell proliferation assays. For the CCK-8 assay, the chondrocytes were seeded into 96-well plates at a density of 1 × 10^3 cells/well. After 12, 24, 36, 60 or 72 h, 10 µl of the reagent (Dojindo, Kumamoto, Japan) was added to each well of the 96-well plate and the plate was incubated for 2 h at 37 °C. Finally, the absorbance at 450 nm was recorded using a SpectraMax M5.

For cell cycle analysis, cells were fixed in 70 % ice-cold ethanol and followed by RNase A treatment and staining with 50 µg/ml of propidium iodide for DNA content analysis in a FACS Calibur BD flow cytometer. The data were collected using the BD FACSuite analysis software. The acquired FACS data were analyzed by ModFit LT software (Verity Software House, Inc., Topsham, ME, USA).

Western Blotting

Cultured cells were lysed in strong RIPA buffer containing Halt Protease Inhibitor Cocktail (Thermo Fisher Scientific, Waltham, MA, USA). The obtained protein concentrations were measured using a BCA protein assay kit (Pierce, Rockford, IL, USA).

Primary antibodies targeting COL2A (sc-28887, Santa Cruz Biotechnology, Santa Cruz, CA, USA), Caspase-3 (ab90437, Abcam), active Caspase 3 (ab2302, Abcam), or GAPDH (ab70699, Abcam) were incubated with the blots overnight at 4 °C, following by incubation with the appropriate horseradish peroxidase (HRP)-conjugated secondary antibodies. Detection of HRP was performed using the Super Signal West Pico Chemiluminescent Substrate (Pierce).

Reverse Transcription, qRT-PCR

Total RNA was isolated using Trizol reagent (Invitrogen, Carlsbad, CA, USA) following the manufacturer’s instructions. For each sample, 500 ng of RNA was reverse transcribed to cDNA using the PrimeScript RT reagent kit (TaKaRa, Dalian, China). Then the obtained cDNA was amplified with the Takara Ex Taq PCR kit (TaKaRa), and quantitative real time polymerase chain reaction (qRT-PCR) amplification was conducted with the Stratagene Mx3000 QPCR system (Stratagene, La Jolla, CA, USA) and analyzed by the 2^{-ΔΔct} method. The primer sequences used in these assays are as follows: H19-F:ACGGGGAGCCAGACATTCATC, H19-R:TGAGAAGAAGCAGCTGACCTC; Gapdh-F:TGGAGTCTACTGGCGTCTT, Gapdh-R:TGGAGTCTACTGGCGTCTT.

Statistical Analysis

The error bars represent the SEM of three independent experiments. Data are represented as mean ± SEM; n = 3. *, **, and *** indicate P<0.05, P<0.01 and P<0.001, respectively (Student’s t test).

Results

LncRNA H19 is down regulated in DDH development.

Safranin O staining showed that cartilage cells in the control group were arranged in an orderly manner and
Fig. 1. H19 is down-regulated in a rat DDH model. a. Safranin O staining of the Control group and DDH group. Scale bars = 60 μm. b. Western blot analysis of COL2A in Control and DDH groups. c. qRT-PCR analysis of the IncRNA H19 in the Control and DDH groups. d. Morphology of GFP-positive lentiviral-infected chondrocytes. Scale bars = 60 μm. e. qRT-PCR analysis of the RNA level of H19 in Control and H19 KD chondrocytes. The data are displayed relative to the results of the Control and represented as mean ± SEM; n=3.

The intercellular matrix was continuous (Fig. 1a). However, cells in the DDH group were disordered and the main cells were spindle-shaped. There were a large number of gaps in the cell matrix in the DDH model group (Fig. 1a). It has been shown that expression of the collagen II component Col2A1 is reduced in the development of DDH (Vonk et al. 2014). Herein, we analyzed the expression level of Col2A1 and found that it was significantly down-regulated in DDH (Fig. 1b). We further examined the expression of IncRNA H19 in the DDH model. The results showed that, compared with the control group, the expression of H19 was significantly decreased in DDH group. To investigated the role of H19, we constructed two shRNAs targeting H19: shH19-1 and shH19-2 (Fig. 1c). Then we packaged the lentivirus and infected normal chondrocytes which were isolated from the control group of rats. Successfully-infected chondrocytes carry the GFP protein and the infected cells were selected by flow cytometry (Fig. 1d). After confirming successful infection, we further validated the expression of the gene by qRT-PCR. Compared with the control (scrambled shRNA), shH19-1 and shH19-2 both efficiently down-regulated H19 IncRNA (Fig. 1e).

**H19 KD suppresses the proliferation of normal chondrocytes**

H19 is closely involved in the lung cancer cell proliferation, therefore we studied its function in chondrocyte proliferation. We used the CCK-8 assay to evaluate cell numbers of the control group and the H19 knockdown (H19 KD) group at 12, 24, 36, 60 and 84 h. The results showed that H19 KD caused a significant decrease in chondrocyte number (Fig. 2a). To confirm the function of H19 in chondrocyte proliferation, we compared BrdU incorporation in the control and H19 KD groups. Compared with the control group, BrdU incorporation in the H19 KD group was significantly reduced, suggesting that H19 KD decreases the proliferation capacity of chondrocytes (Fig. 2b). Consistently, H19 KD in chondrocytes dramatically increased the cell population at G0/G1 phase, whereas it reduced the cell population at G2/M phase (Fig. 2c, d). These data suggest that the proliferative ability of H19 KD chondrocytes is significantly inhibited.
Fig. 2. H19 KD suppresses cell proliferation. a. CCK-8 assay analysis of the cell proliferation of Control and H19 KD chondrocytes isolated from our rat model. ** indicates P < 0.01. b. BrdU immunostaining analysis of the cell proliferation of Control and H19 KD chondrocytes. Scale bars = 60 μm. c and d. Flow cytometric analysis of the cell cycle of Control and H19 KD chondrocytes.

H19 affects chondrocyte proliferation but not apoptosis.

Cell apoptosis is closely related to DDH progression. Therefore we examined apoptosis by flow cytometry. We found no significant difference in the number of apoptotic cells between the H19 KD and control chondrocytes (Fig. 3a). Caspase-3, an important terminal cleavage enzyme, is a crucial mediator of apoptosis, and activated Caspase-3 is a key indicator of apoptosis. H19 KD did not alter the protein levels of Caspase-3 or of activated Caspase-3 (Fig. 3b, c), indicating that H19 does not affect chondrocyte apoptosis.

H19 overexpression promotes chondrocyte proliferation.

To further confirm the function of H19 in DDH progression, we over-expressed H19 in chondrocytes isolated from DDH model rats. qRT-PCR analysis showed that H19 was successfully over-expressed (Fig. 4a). Overexpression of H19 promoted the proliferation of chondrocytes from the DDH model rats (Fig. 4b). Similarly, BrdU immunostaining revealed that overexpression of H19 enhanced the proportion of dividing chondrocytes (Fig. 4c). We then performed flow cytometry to further identify the relationship between
H19 and cell cycle regulation. The results showed that overexpression of H19 significantly decreased the cell population at G0/G1 phase and increased the cell population at G2/M phase (Fig. 4d and 4e). These data suggest that overexpression of H19 promotes the proliferation of chondrocytes isolated from our DDH model.

**H19 regulates proliferation of chondrocytes through direct binding to let-7.**

The molecular mechanism of lncRNA includes RNA–protein interactions as well as acting as a competing endogenous RNA that binds to a miRNA and inhibits its function. It is reported that H19 interacts with miRNA let-7 and functions as a let-7 inhibitor (Kallen et al. 2013). We next explored the role of let-7 in the effects of H19 in chondrocytes. Interestingly, the level of let-7 was significantly up-regulated in the development of DDH in our model while H19 was down-regulated in this model (Fig. 4a). Similarly, when H19 was overexpressed, let-7 was down-regulated (Fig. 4b). Thus it appears that the function of H19 is associated with let-7 in this disease. It has been reported that let-7 interacts with H19 directly (Kallen et al. 2013). Together, these results suggest that H19 down-regulation impairs proliferation of chondrocytes through regulating the expression of let-7.

**a**

![Flow cytometric analysis of apoptosis of Control and H19 KD chondrocytes.](image)

**b**

![Western blotting analysis of apoptosis-related genes in Control and H19 KD chondrocytes.](image)

**c**

![Relative expression of Caspase-3 and Active Caspase-3.](image)

**Fig. 3.** H19 does not affect apoptosis but does promote cell proliferation of chondrocytes. *a.* Flow cytometric analysis of apoptosis of Control and H19 KD chondrocytes. *b.* and *c.* Western blotting analysis of apoptosis-related genes in Control and H19 KD chondrocytes. The data are displayed relative to the results of Control.

**Discussion**

DDH is one of the most frequent congenital abnormalities in newborns. It is associated with degenerative lesions of cartilage in the joint, but the molecular mechanism of DDH is still far from clear. We found that the formation and development of DDH may be closely related to H19. These results reveal, for the first time, the role of the epigenetic factor lncRNA in DDH, which helps to deepen the understanding of the pathogenesis of DDH as well as providing a potential molecular marker for the diagnosis of DDH. At present,
the treatment of DDH is mainly dependent on surgical methods for infants more than 18 months old, such as hip replacement and associated bony osteotomy (Wenger, et al. 2014). Although the vast majority of patients can achieve a satisfactory cure, there are a small number of patients who remain disabled, leaving a heavy burden on the patients and their family (Anuar et al. 2015). A deeper understanding of the molecular mechanisms, such as lncRNA, will help with the development of targeted drugs, thereby increasing treatment options for these patients. In addition to lncRNA, epigenetic regulators include DNA methylation, histone acetylation and deacetylation, as well as miRNA and many other regulators (Wee et al. 2014). It may be very interesting to investigate the function and mechanism of these epigenetic regulators in DDH.

H19, an imprinted oncofetal gene, has a central role in carcinogenesis. However, its function in DDH has not been explored. Here, we found that H19 was closely related to the formation of DDH in rats. Its

![Graphs and figures](image.png)

**Fig. 4.** H19 overexpression promotes chondrocytes proliferation. 

|                | G0/G1 phase(%) | S phase(%) | G2/M phase(%) |
|----------------|---------------|------------|---------------|
| Control        | 80.06 ± 2.18  | 7.0 ± 0.96 | 9.02 ± 1.27   |
| H19            | 72.23 ± 2.26  | 8.86 ± 1.42| 16.14 ± 1.84  |

The data are displayed relative to the results of the Control and are represented as mean ± SEM; n=3. *** indicates P<0.001. CCK-8 assay analysis of the cell proliferation of Control and H19 chondrocytes isolated from Control rats. ** indicates P<0.01. BrdU immunostaining analysis of the cell proliferation of Control and H19 chondrocytes. Scale bars = 60 μm. Flow cytometric analysis of the cell cycle of Control and H19 chondrocytes.
expression was significantly down-regulated as the hip condition deteriorated; further we found that the effect of H19 in the DDH process may be associated with its role in regulating cell proliferation. In the process of hip development, the proliferative ability of chondrocytes is very important, so lack of H19 should lead to limitation of the cell proliferation capacity, leading to hip articular cartilage dysfunction and hip dysplasia. In one previous study, it was reported that H19 is linked to diseases such as cancer and cardiovascular disease (Liu et al. 2016, Zhou et al. 2015). Our findings reveal the role of H19 in DDH. These results suggest that H19 may be useful as an important disease indicator, and further can be used as a target for developing new drugs or treatment in DDH.

At the molecular level, one of the main functions of lncRNAs is its action as a ceRNA, which can bind to miRNAs and inhibit their function (Zhou et al. 2016). Previous studies have found that let-7, which is closely related to cell proliferation in human cells, is an important regulatory factor of H19 (Yan et al. 2015). We found that let-7 is a downstream target of H19, through which H19 regulates cell proliferation in rat chondrocytes. Further, we confirmed this result by luciferase assay. This study demonstrated that a similar let-7/H19 regulatory feedback loop is present not only in the human but also in the rat. Therefore, this loop is conserved in these two species. For ethical reasons, it is difficult to obtain human chondrocytes, but because of this conserved pathway, we estimate that the let-7/H19 regulatory pathway may also play an important role in the process of human DDH.

In summary, our study has shown that expression of H19 is significantly down-regulated during DDH development and that H19 may be an important epigenetic regulatory factor in DDH formation (Fig. 5c). We also identified let-7 as an H19 target gene. Our study is the first to reveal that H19 is an epigenetic regulator of DDH and the challenge for future studies will be to better understand this regulation in other epigenetic regulators, such as miRNA and chromatin modifiers.
Conflict of Interest
There is no conflict of interest.

Acknowledgements
This work was supported by the Natural Science Foundation of Anhui Province (project no. 1408085QH153).

Abbreviations
DDH, developmental dysplasia and dislocation of hip; KD, knockdown; lncRNA, long non-coding RNA; miRNA, microRNA; PBS, phosphate buffer saline; FBS, fetal bovine serum; shRNA, short hairpin RNA; qRT-PCR, quantitative real time polymerase chain reaction; CCK-8, Cell Counting Kit-8.

References

ANUAR R, MOHD-HISYAMUDIN HP, AHMAD MH, ZULKIFLEE O: The Economic Impact of Managing Late Presentation of Developmental Dysplasia of Hip (DDH). *Malays Orthop J* **9**: 40-43, 2015.

ASHRAF A, LARSON AN, MARADIT-KREMERS H, KREMERS WK, LEWALLEN DG: Hospital costs of total hip arthroplasty for developmental dysplasia of the hip. *Clin Orthop Relat Res* **472**: 2237-2244, 2014.

BO N, PENG W, XINGHONG P, MA R: Early cartilage degeneration in a rat experimental model of developmental dysplasia of the hip. *Connect Tissue Res* **53**: 513-520, 2012.

CARNEY BT: Acetabular dysplasia following closed reduction of developmental dislocation of the hip. *J Surg Orthop Adv* **14**: 122-124, 2005.

KALLEN AN, ZHOU XB, LU J, QIAO C, MA J, YAN L, LU L, LIU C, YI JS, ZHANG H, MIN W, Bennett AM, Gregory RI, Ding Y, Huang Y: The imprinted H19 lncRNA antagonizes let-7 microRNAs. *Mol Cell* **52**: 101-112, 2013.

KAZEMZADEH M, SAFARALIZADEH R, ORANG AV: LncRNAs: emerging players in gene regulation and disease pathogenesis. *J Genet* **94**: 771-784, 2015.

KRASE A, ABEDIAN R, STECK E, HURSCHLER C, RICHTER W: BMP activation and Wnt-signalling affect biochemistry and functional biomechanical properties of cartilage tissue engineering constructs. *Osteoarthritis Cartilage* **22**: 284-292, 2014.

LI X, WU Z, FU X, HAN W. LncRNAs: insights into their function and mechanics in underlying disorders. *Mutat Res Rev Mutat Res* **762**: 1-21, 2014.

LIU L, AN X, LI Z, SONG Y, LI L, ZUO S, LIU N, YANG G, WANG H, CHENG X, ZHANG Y, YANG X, WANG J: The H19 long noncoding RNA is a novel negative regulator of cardiomyocyte hypertrophy. *Cardiovasc Res* **111**: 56-65, 2016.

PEI X, MO Y, NING B, YUAN Z, PENG L, MA R: The role of TGFβ1 stimulating ROCK I signal pathway to reorganize actin in a rat experimental model of developmental dysplasia of the hip. *Mol Cell Biochem* **391**: 1-9, 2014.

PRENSNER JR, CHEN W, HAN S, IYER MK, CAO Q, KOTHARI V, EVANS JR, KNUDSEN KE, PAULSEN MT, LJUNGMAN M, LAWRENCE TS, CHINNAIYAN AM, FENG FY: The long non-coding RNA PCAT-1 promotes prostate cancer cell proliferation through cMyc. *Neoplasia* **16**: 900-908, 2014.

SENARAN H, BOWEN JR, HARCKE HT: Avascular necrosis rate in early reduction after failed Pavlik harness treatment of developmental dysplasia of the hip. *J Pediatric Orthop* **27**: 192-197, 2007.

THOMAS SR, WEDGE JH, SALTER RB: Outcome at forty-five years after open reduction and innominate osteotomy for late-presenting developmental dislocation of the hip. *J Bone Joint Surg Am* **89**: 2341-2350, 2007.

VONK LA, KRAGTEN AH, DHERT WJ, SARIS DB, CREAMERS LB: Overexpression of hsa-miR-148a promotes cartilage production and inhibits cartilage degradation by osteoarthritic chondrocytes. *Osteoarthritis Cartilage* **22**: 145-153, 2014.

WEE S, DHANAK D, LI H, ARMSTRONG SA, COPELAND RA, SIMS R, BAYLIN SB, LIU XS, SCHWEIZER L: Targeting epigenetic regulators for cancer therapy. *Ann N Y Acad Sci* **1309**: 30-36, 2014.

WEI YS, LI DH, LIU WL, JIANG DM: Altered Chondrocyte Apoptosis Status in Developmental Hip Dysplasia in Rabbits. *Balkan Med J* **33**: 639-644, 2016.

WENGER DR: Surgical treatment of developmental dysplasia of the hip. *Instr Course Lect* **63**: 313-323, 2014.
YAN L, ZHOU J, GAO Y, GHAZAL S, LU L, BELLONE S, YANG Y, LIU N, ZHAO X, SANTIN AD, TAYLOR H, HUANG Y: Regulation of tumor cell migration and invasion by the H19/let-7 axis is antagonized by metformin-induced DNA methylation. *Oncogene* **34**: 3076-3084, 2015.

ZHANG C, WANG P, JIANG P, LV Y, DONG C, DAI X, TAN L, WANG Z: Upregulation of IncRNA HOTAIR contributes to IL-1β-induced MMP overexpression and chondrocytes apoptosis in temporomandibular joint osteoarthritis. *Gene* **586**: 248-253, 2016.

ZHANG G, WU Y, XU D, YAN X: Long Noncoding RNA UFO1 Promotes Proliferation of Chondrocyte in Osteoarthritis by Acting as a Sponge for miR-34a. *DNA Cell Biol* **35**: 691-695, 2016.

ZHANG X, MENG Q, MA R, CHEN G, CHENG L, SHEN J: Early acetabular cartilage degeneration in a rabbit model of developmental dysplasia of the hip. *Int J Clin Exp Med* **8**: 14505-14512, 2015.

Zhou M, Diao Z, Yue X, Chen Y, Zhao H, Cheng L, Sun J: Construction and analysis of dysregulated IncRNA-associated ceRNA network identified novel IncRNA biomarkers for early diagnosis of human pancreatic cancer. *Oncotarget* **7**: 56383-56394, 2016.

Zhou X, Yin C, Dang Y, Ye F, Zhang G: Identification of the long non-coding RNA H19 in plasma as a novel biomarker for diagnosis of gastric cancer. *Sci Rep* **5**: 11516, 2015.