Abstract. Background/Aim: Patients with stage IIIA (N2) non-small cell lung cancer (NSCLC) with no progression after induction chemotherapy are usually selected for surgery. Nowadays, response to chemotherapy is not predictable. We aimed to identify genomic predictive markers for response to induction chemotherapy in stage IIIA (N2) NSCLC patients.

Patients and Methods: Whole-exome sequencing (WES) was performed on samples from 11 patients with no response after induction chemotherapy and 6 patients with documented pathological response, admitted to the Hotel Dieu Hospital, Paris or Allegemeines Krakenhaus University, Vienna. Results: A higher alternative allele frequency was found on SENP5, rs63736860, rs1602 and NCBP2, rs553783 in the non-responder group, and on RGP1, rs1570248, SLFN12L, rs2304968, rs9905892, and GBA2, rs3833700 in the responder group. Conclusion: These polymorphisms contribute to individual sensibility to chemotherapy response. Interrogation of these genetic variations may have potential applicability when deciding the treatment strategy for patients with stage III NSCLC (N2).

Patients with non-small cell lung cancer (NSCLC) stage IIIA, with invasion of mediastinal lymph nodes (N2), represent about 25% of total NSCLC diagnosed cases (1). Chemotherapy is one of the main treatment options for these patients, followed by surgery if patients can tolerate it and are at least stabilized by the induction chemotherapeutic regimen. In patients with unresectable disease and without progression after chemotherapy the more recently addition of durvalumab demonstrated in the PACIFIC trail showed improved progression-free survival and overall survival (2) proving the benefit of checkpoint inhibitors in NSCLC stage III patient therapy.

The most common type of platinum salts used in NSCLC chemotherapy are cisplatin and carboplatin. They have different toxicity profiles, but cisplatin showed a better rate of objective response (3).

The mechanism of action of cisplatin is mediated by cisplatin-DNA adducts that interfere with the DNA repair mechanism of the cell eventually leading to cell apoptosis (4). The response to chemotherapy cannot be predicted by clinical methods, although pharmacogenomics studies are identifying SNPs that might estimate drug therapeutic effects. So far, studies that focused on finding predictive biomarkers for NSCLC stage IIIA (N2) patients’ response to platinum chemotherapy, were mainly limited to genetic variations in genes known to be associated with platinum metabolism or DNA repair. These two mechanisms are known to be primarily affected by chemotherapy treatment. Fewer studies were directed to identifying polymorphisms in other genes, that might predict the sensitivity or resistance to therapy.

In this study, we aimed to identify differences in the molecular profiles between NSCLC stage IIIA (N2) responders and non-responders to chemotherapy (cisplatin/carboplatin) with emphasis in finding predictive variants in new genes, using whole-exome sequencing (WES).

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Correspondence to: Madalina Grigoroiu, MD, Ph.D., Fundeni Clinical Institute, Fundeni Ave, No. 258, 022328 Bucharest, Romania. Tel: +40 213180417, Fax: +40 213180417, e-mail madalinalidia@yahoo.com

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Patients and Methods

The design of the study was previously published in extenso (5). Below, we describe its most important points.

Patient groups. The study flow chart is shown in Figure 1A. Briefly, 73 patients admitted to Hotel Dieu Hospital in Paris and Algemeines Krankenhaus in Vienna, with diagnosis of or suspected to have NSCLC, clinical stage II A (N2) were prospectively included in the study, between January 2011 and December 2012, based on a written informed consent. All patients had either mediastinoscopy or thoracoscopy in order to confirm the mediastinal lymph nodes invasion. After histological examination of the lymph node status, 44 patients were excluded from the study. Causes of exclusion are shown in Figure 1A. Only samples with more than 70% cellularity were considered for the genomic analysis. The remaining 29 patients received induction chemotherapy, based on one platinum salt (cisplatin or carboplatin) in association with a second chemotherapeutic agent. We excluded one patient who died after the first cure of chemotherapy, because of chemotherapy toxicity. After induction chemotherapy, the patients with clinical response according to WHO criteria were referred to surgical treatment.

“Non-responder” patients (group A) were considered those with no clinical response to induction chemotherapy and who finally were not referred for surgical resection and patients with clinical response to induction chemotherapy according to WHO criteria, but persistent N2 status on pathological examination of the resected lymph nodes after lung surgery. The “responders” patients (group B) were patients with no residual disease in the mediastinal lymph nodes on pathological examination after radical lymphadenectomy performed at lung surgery (down-staging). After surgical treatment, there were 21 patients in the “non-responders” group (group A) and 7 patients in the “responders” group (group B). Only tumour mediastinal lymph node (N2) samples before induction chemotherapy were analyzed in this study. Finally, 17 patients (11 patients in “non-responders” group and 6 patients in “responders” group) had tumor samples that fulfill the quality criteria required for whole exome sequencing (WES). The average sequencing depth on target and the total number of SNP obtained on each sample are shown in Table 1.

DNA extraction. DNA was extracted from fresh-frozen tissue with a DNA extraction kit (Stratagene, San Diego, CA, USA) or QIAamp DNA Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer’s instructions. DNA concentration and integrity was checked with a Nanodrop ND 1000 and Qubit 2.0 fluorimeter and by gel electrophoresis in 1% agarose.

DNA sequencing. Whole-exome sequencing (WES) was performed at the Beijing Genomics Institute (BGI, PR China). Genomic DNA was hybridized using Sure Select Target Enrichment system (Agilent, Santa Clara, CA, USA) at 50X on an Illumina HiSeq2000 platform according to manufacturer protocol. BWA (Burrow-Wheeler Aligner) alignment and genome build 37 (hg19) was used to generate Bam files for data processing and finally for variant calling. Picard was used for removal of PCR duplicates. SNPs were called with Soaps, InDels with Samtools/GATK and SNVs with Varscan. Variant annotation was done using ANNOVAR for the confident variants.

### Table 1. Parameters of whole exome sequencing (WES) showing the average sequencing depth on target (at least 50X) and number of SNPs detected in each sequenced sample.

| Group | Sample   | Average sequencing depth on target | Total number of SNPs |
|-------|----------|------------------------------------|----------------------|
| A     | 11B00790 | 50.72                              | 79,698               |
| A     | 11B02967 | 57.72                              | 78,338               |
| A     | LYN84    | 65.56                              | 84,592               |
| A     | LYN77_1  | 60.65                              | 81,344               |
| A     | 12B05696 | 58.44                              | 80,321               |
| A     | 11B04855 | 57.23                              | 87,236               |
| A     | 11B02702 | 66.19                              | 80,081               |
| A     | 11B3023  | 68.76                              | 78,202               |
| A     | 11B2941  | 66.86                              | 84,410               |
| A     | LYN72_1  | 56.25                              | 74,755               |
| A     | 11B05089 | 68.14                              | 81,449               |
| B     | 11B00602 | 65.67                              | 74,452               |
| B     | 12B3843  | 64.8                               | 84,547               |
| B     | LYN43_1  | 63.28                              | 76,863               |
| B     | LYN48_1  | 58.82                              | 81,829               |
| B     | LYN101_1 | 65.66                              | 86,017               |
| B     | 11B01622 | 65.85                              | 78,490               |

Data analysis. Our analysis aimed in identifying differences between group A and group B of patients looking at all identified variants. All variants were merged for all analyzed samples and the frequencies of reference allele and variant allele were calculated. Next, two by two tables were generated for the reference and variant allele in group A and group B and Fisher’s exact test was used to calculated significance value (R package). Analysis of linkage disequilibrium was done using the LDmatrix module from LDlink (6), in European populations. Statistical analysis and graphs were done with R Studio 3.5.2 or GraphPad Prism 7 (GraphPad Software Inc., San Diego, CA, USA).

Results

Patient characteristics. Seventeen patients were included in our study (Figure 1A), eleven in group A (patients with evolution of the disease after induction chemotherapy or persistent N2+ on pathological examination after surgical resection) and six in group B (patients with down-staging disease after induction therapy). The cohort average age was 61.7±9.2 years (60.2±10.4 years for group A and 64.2±6.0 years for group B, p=0.42, Mann-Whitney test). In total, ten male and seven female patients were analyzed, five male and six female patients in group A and five male and one female patient in group B (p=0.3, Fisher’s exact test) (Figure 1B).

Identification of SNPs that associate with response to therapy in NSCLC stage II A (N2). Single-nucleotide polymorphisms (SNPs) are a source of human genetic variation, and pharmacogenetic studies have revealed that
Nastase et al: Sensitivity-associated Polymorphisms in Stage III NSCLC (N2)

Figure 1. (A) Study workflow showing number of patients in each group and exclusion reasons. (B) Diagram showing patient distribution in response groups, type of response to treatment and gender (http://sankeymatic.com).
Figure 2. Continued
SNPs could influence the pharmacokinetics of the drugs and impact patients’ response to treatment (7), proving their importance in drug selection process. Analysis of WES data between non-responders (group A) and responders (group B) to chemotherapy led to identification of 216 variants (p < 0.05, Fisher’s exact test) that segregated between the two groups. By lowering the p to 0.01, seven polymorphism in five genes were selected to be strong predictors of response: Senrin/SUMO-specific peptidase 5 (SENP5), rs63736860 (OR=0.027, 95%CI=0.0013-0.55, p=0.0016), rs1602 (OR=0.052, 95%CI=0.005-0.56, p=0.009) and nuclear cap-binding protein subunit 2 (NCBP2), rs553783 (OR=0.0052, 95%CI=0.005-0.53, p=0.006). Gene variants had a higher alternative allele frequency in non-responder group, while RGP1 homolog, RAB6A GEF complex partner 1 (RGP1), rs1570248 (OR=25, 95%CI=1.28-485.9, p=0.004), RGP1 homolog, RAB6A GEF complex partner 1 (RGP1), rs1570248 (OR=25, 95%CI=1.28-485.9, p=0.004), Schlafen family member 12 (SLFN12), rs9905892 (OR=21, 95%CI=1.11-398.9, p=0.006), and glucosylceramidase beta 2 (GBA2), rs3833700 (OR=20.24, 95%CI=1.04-393.9, p=0.01) gene variants had a higher alternative allele frequency in responder group (Figure 2A, B and C and Table II).
Analysis of linkage disequilibrium for the markers located on the same chromosome (chromosomes 3, 9 and 17), showed a strong association between pairs of SNPs in European populations, thus suggesting a lack of genetic recombination and their co-inheritance (Figure 3A, B and C).

Among all the 216 SNPs (that were statistically significant, \( p < 0.05 \)), 64 were predicted to affect an exon, with two of those predicted to be frameshift variants: rs34595082, MRPS34 (\( p = 0.028 \)) and rs11467417, DEFB126 (\( p = 0.035 \)). The rest were either missense, affecting the 3' or 5'UTR sequence of the gene or were synonymous mutations.

Searching for variants in previously reported cancer genes among the 216 SNPs, we identified rs1407179 in ROS1 gene, an intronic variant that was more frequent in group A, and thus predicting a negative response to therapy (\( p = 0.0144 \)) and rs4077170 in POLE that predicted a favorable response to therapy (\( p = 0.0237 \)) (Table II).

**Discussion**

NSCLC is the most common type of lung cancer and the leading cause of oncological death among men and women worldwide (8, 9). It comprises of two histological subtypes: adenocarcinoma, squamous cell carcinoma, with adenocarcinoma subtype being the most widely diagnosed, representing about 40% of cases (10). Smoking is the major risk factor for the disease (11) that acts by inducing DNA damage and increasing the risk for accumulation of driver mutations in cancer genes (12), most notably the C>A (G>T) transversion in the TP53 gene (13) located on the non-transcribed strand. Nevertheless, patients that were never smokers might develop the disease probably due to other environmental exposures. Research in NSCLC facilitated an improved understanding of the molecular mechanisms that lead to the onset and development of the disease that reflected in a better therapeutic management (14). About two-sixths of NSCLC patients harbor a targetable oncogenic addiction that makes them candidates to targeted therapy (14), but the other patients, without evident alterations will be treated with frontline standard platinum-based chemotherapy. Even so, tumors that initially have a targetable alteration will develop resistance and so the response rate of the patients will be modest.

Patients with NSCLC stage IIIA, with invasion of mediastinal lymph nodes (N2), represent a heterogenous prognostic category of patients. Bulky N2 or functionally impaired patients will not be eligible for surgery, but for the rest of N2 patients, a treatment by induction chemotherapy followed by surgical resection if the disease is at least stabilized, is the most adopted strategy. The patients who will be down-staged by the induction chemotherapy (no residual disease on the mediastinal lymph nodes after radical lymphadenectomy) are those who will benefit most from this strategy (15). The response to induction chemotherapy is one of the most important prognostic factors (16) for patients with NSCLC stage IIIA (N2), but it is currently impossible to predict this response.

In this study, we identified new markers (polymorphisms) that might predict response to therapy, in patients with NSCLC stage IIIA (N2) that were prospectively followed-up and had samples collected from mediastinal lymph nodes prior to chemotherapy, by using whole-exome sequencing.

We sequenced samples from seventeen patients and detected 216 SNPs that discriminated between responders and non-responders to platinum therapy, at a \( p < 0.05 \) by the Fischer’s \( t \)-test. Seven of these polymorphisms segregated between non-responder and responder patients with a \( p < 0.01 \) and were located on chromosome 3, in SENP5 and NCBP2 genes, chromosome 9 in RGPl and GBA2 genes and chromosome 17, SLFN12L gene. To the best of our knowledge, these polymorphisms have not been described before as being associated with response to therapy of NSCLC patients.

Sentrin/SUMO-specific peptidases 5 (SENP5) belongs to a family of peptidases involved in maturation of small ubiquitin-like modifiers (SUMO) residues and their removal (17) from protein targets. SUMOylation/deSUMOylation are post-translational modifications (PMT) that can regulate biological processes like DNA damage, immune response, apoptosis and tumorigenesis (18, 19) as well as T cell activation and differentiations (20). SENP5 has C-terminal hydrolase and isopeptidase activities and an important role in mitosis (21). It shares sequence and substrate homology with SENP3 both being located in the nucleus and usually classified as a subfamily of SENP proteins (22-24). SENP5 was identified as a candidate driver gene at the locus 3q26-29 in squamous cell carcinoma of the lung, a type of NSCLC, together with SENP2, DCUN1D1 and DVL3 (25), this whole region being a biomarker of differentiation between different histological subtypes of NSCLC (26). SENP5 is mapped at 3q29 locus that was suggested to be a susceptibility region in NSCLC (27, 28). Amplification of the distal regions of 3q, including 3q29 is considered a parameter of neoplastic transformation in lung cancer (29).

In hepatocellular carcinoma, drug activation of SENP5 inhibited hypoxia that has been shown to be link to resistance to chemotherapy in part by activation of the Sonic Hedgehog (SHH) pathway (30). Nuclear cap-binding protein subunit 2 (NCBP2) component of the nuclear cap-binding protein complex (CBC) was identified as a target gene of miR-193a-5p in lung cancer (31). In prostate cancer, silencing of miR-193a-5p increased sensitivity to docetaxel (32), suggesting a possible new possible therapeutic approach.

Schlafen family member 12-like (SLFN12L) is a human Schlafen protein that was reported to promote differentiation in human prostate cancer and is known to be down-regulated during T cell activation and up-regulated by type I interferons (IFNs) (33, 34).
Figure 3. Heatmap matrix of pairwise linkage disequilibrium in 5 European populations for SNPs on chromosome 3 (A); chromosome 9 (B) and chromosome 17 (C) (heatmaps and D’ and R2 were generated using the LDmatrix module from LDlink (6); the shade of red represents the degree of linkage disequilibrium between the pairs.)
Glucosylceramidase beta 2 (GBA2) non-lysosomal glucosylceramidase catalyzes the hydrolysis of glucosylceramide (GlcCer) to free glucose and ceramide (34). Mutations are associated with the progression of neurological disorders (35). It is highly expressed in the liver and mutations were associated with deficient liver regeneration mainly due to changes in STAT3 signaling pathway (36). RGP1 homolog, RAB6A GEF complex partner 1 (RGP1) together with RIC1 forms a complex that acts as a guanine nucleotide exchange factor (GEF), which activates RAB6A by exchanging bound GDP for free GTP and may thereby be required for efficient fusion of endosome-derived vesicles with the Golgi compartment (37).

Although limited in sample size, mainly due to the complexity of the disease and difficulty of achieving longitudinal samples, our study identified variants that might be important in the therapy selection process. It also accentuates on the essential role of inter-individual variability associated with chemotherapy response in NSCLC stage IIIA (N2) patients and supports the need of more similar studies, on bigger cohorts, to be undertaken.

Conflicts of Interest
The Authors declare no conflicts of interest.

Authors’ Contributions
AN processed the samples, extracted DNA, assessed sample’s quality, prepared samples for shipping and analyzed the results received from BGI; AL and DD were involved in histopathological review of the samples; VL was involved in sample processing; SD, IP and MG were involved in the implementation and coordination of the study; WK, MA and EC were involved in sample collection, MG conceived the study, AN and MG wrote the manuscript with the assistance and final approval of all Authors.

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