Expression pattern of tumour-associated antigens in hepatocellular carcinoma: association with immune infiltration and disease progression

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Background: The distinct expression pattern of tumour-associated antigens (TAAs) might be a critical reason for the inefficacy of immunity-based treatments and heterogeneous postsurgical recovery in patients with solid tumours, including hepatocellular carcinoma (HCC). However, little is known about the clinical value of the coexpression patterns of multiple TAAs.

Methods: We determined the expression of multiple TAAs with identified immunogenicity (GPC3, AFP, SSX-2, NY-ESO-1, EpCAM, midkine) and the density of tumour-infiltrating immune cells by immunohistochemistry in a panel of 362 primary HCC patients. We evaluated the association between the TAAs, immune cell infiltration, clinicopathological parameters, and prognosis.

Results: Patients who coexpressed more TAAs had better prognosis (P<0.00001, overall survival). The integrated pattern of TAA was associated with good differentiation and small tumour size, and with more CD57⁺ natural killer and CD20⁺ B-cell infiltration (P<0.05). Multivariate Cox proportional hazards analysis identified the TAA index as an independent prognostic indicator (hazard ratio 0.625; 95% confidence interval 0.467–0.837; P=0.002), and could further predict patient prognosis in collaboration with local immune infiltration.

Conclusion: Our results could provide new evidence for the improvement of prognostic molecular signatures in HCC, and a novel rationale for patient enrolment in future immunotherapeutic trials and/or clinical treatments.

Hepatocellular carcinoma (HCC) is among the world’s leading cancer threats, and its incidence is increasing (Fan et al, 2011; Jemal et al, 2011). Despite improved diagnostic and treatment strategies, the overall survival (OS) of patients with HCC remains poor (Bruix and Llovet, 2009; Villanueva et al, 2012). Hepatocellular carcinoma is a highly complex disease that is generally resistant to commonly used chemotherapy and radiotherapy. Only a small proportion of newly diagnosed patients are eligible for potential curative therapies, including resection and liver transplantation. However, these therapeutic procedures most often do not provide a complete cure, and half of the treated patients experience tumour recurrence within 3 years (Villanueva et al, 2010; Yang and Roberts, 2010; Gao et al, 2012).

Hepatocellular carcinoma is usually present in inflamed fibrotic and/or cirrhotic liver with extensive leukocyte infiltration. Thus, the immune status at a tumour site can largely influence the...
biologic behaviour of HCC (Grivennikov et al., 2010; Fridman et al., 2012; Qin, 2012). Although it is commonly believed that innate and adaptive immunity would inhibit cancer growth, solid tumour cells, which often arise from chronic inflammation and survive immunoediting, can escape from immunosurveillance through different mechanisms (Disis, 2010; Vesely et al., 2011). After decades of substantial efforts, the success of several recent proof-of-concept clinical trials targeting immune regulators (e.g., ipilimumab and MDX-1106) suggests that active immunotherapy represents a path to obtain a durable and long-lasting response in cancer patients. However, their effects on the regression of human tumours remain limited, and only a relatively small fraction of patients derives clinical benefits (Mellman et al., 2011; Topalian et al., 2011). The reconstruction of immune surveillance for aberrant cells requires adequate tumour-associated antigen (TAA) expression, and the distinct expression pattern of TAAs in different tumours may be a critical reason for heterogeneous therapeutic effectiveness. Severe deficiency in TAAs is a common trick of tumour immune escape in primary cancer and/or during postsurgical relapse (Schreiber et al., 2011; Fu, 2012). However, little is currently known about the clinical significance of the coexpression pattern of multiple TAAs in solid tumours, including HCC.

In view of the present situation, we aimed to determine the expression of several TAAs with identified immunogenicity and potential therapeutic value in HCC and to evaluate their clinicopathological roles. In general, our results showed that the expression patterns of these TAAs (TAA index) were associated with local immune infiltration, disease progression, and prognosis.

### MATERIALS AND METHODS

**Patients and tissue samples.** Archived formalin-fixed, paraffin-embedded tissues were obtained at the Cancer Center of Sun Yat-sen University, from 362 patients who had all undergone curative resection for HCC between January 2002 and December 2005. There were 317 (87.6%) patients with hepatitis B virus infection. We defined curative resection for HCC as a resection margin of at least 1 cm, complete resection of all tumour nodules, and leaving the cut surface free of tumour based on histological examination. Intra-operative ultrasound and postsurgical contrast-enhanced computed tomography (CT) were routinely used to ensure the complete removal of the HCC (Gao et al., 2007; Kuang et al., 2011; Xu et al., 2012b). No patient received anti-cancer therapies or had distant metastasis prior to the surgery. The clinical stage of tumours was determined according to the tumor-nodes-metastasis (TNM) classification system of the International Union Against Cancer (edition 6). The Institutional Review Boards of the Cancer Center approved the study. Written informed consent was obtained from all patients. Clinicopathological characteristics are summarised in Table 1.

**Follow-up of patients and postoperative treatment.** Patients were followed postoperatively at the outpatient clinic with regular surveillance for recurrence using the serum alpha-fetoprotein (AFP) level, abdominal ultrasonography, and chest radiography at 2- to 4-month intervals (Gao et al., 2007; Ding et al., 2011; Kuang et al., 2011; Xu et al., 2012b). When tumour recurrence or metastasis was suspected, further examinations, including CT and hepatic angiography, were performed. Biopsies were obtained when necessary. Patients with confirmed recurrence received further treatment, including a second surgical resection, transcatheter arterial chemoembolisation, radiofrequency ablation or percutaneous ethanol injection. The median follow-up was 34.5 months (range 1–95 months). Of the 362 patients examined during the follow-up period, 190 patients (52.5%) died, 193 (53.3%) were diagnosed with tumour recurrence, and 106 (29.3%) remained alive without recurrence. Overall survival was defined as the interval between surgery and death or between surgery and the last observation for surviving patients. The time to recurrence was defined as the interval between surgery and recurrence or between surgery and the last observation for patients without recurrence.

**Tissue microarray and immunohistochemistry.** The tissue microarray (TMA) was constructed as described previously (Xu et al., 2012b). Briefly, blocks containing the advancing edges of tumoural and peri-tumoural HCC tissue were used for TMA construction. Haematoxylin and eosin (H&E)-stained slides were reviewed without the knowledge of the patient clinical characteristics and outcomes. Duplicate 1.0-mm tissue cores were obtained from

| Characteristics | No. of patients (%) |
|-----------------|---------------------|
| **Age, year**   |                     |
| Median          | 48.5                |
| Range           | 20–78               |
| **Gender**      |                     |
| Male            | 324 (89.5)          |
| Female          | 38 (10.5)           |
| **Hepatitis B virus infection** |       |
| No              | 45 (12.4)           |
| Yes             | 317 (87.6)          |
| **Hepatitis C virus infection** |       |
| No              | 356 (98.3)          |
| Yes             | 6 (1.7)             |
| **Alpha-fetoprotein** |       |
| ≤25 ng ml⁻¹     | 107 (29.6)          |
| >25 ng ml⁻¹     | 255 (70.4)          |
| **Child-Pugh class** |       |
| A               | 339 (93.6)          |
| B               | 23 (6.4)            |
| **Differentiation** |                 |
| I–II            | 289 (79.8)          |
| III–IV          | 73 (20.2)           |
| **Tumour number** |                     |
| Single          | 274 (75.7)          |
| Multiple        | 88 (24.3)           |
| **Tumour size** |                     |
| ≤5 cm           | 128 (35.4)          |
| >5 cm           | 234 (64.6)          |
| **Vascular invasion** |             |
| Absent          | 309 (85.4)          |
| Present         | 53 (14.6)           |
| **TNM stage**   |                     |
| I               | 259 (71.5)          |
| II–III          | 103 (28.5)          |

Abbreviation: TNM = tumour-lymph node metastasis.

**Table 1. Patient characteristics**
two regions (total, four punches) in the paraffin-embedded tissue blocks. Tissue microarrays containing the tissue cores were then cut into 5 \( \mu m \) sections for immunohistochemistry (IHC) staining.

The IHC of the paraffin sections was carried out using a two-step protocol (DakoCytomation, Glostrup, Denmark). Briefly, 5 \( \mu m \) sections were deparaffinised, and endogenous peroxidase activity was blocked with 0.3% hydrogen peroxide containing double-distilled water for 10 min. Antigen retrieval was performed by microwave treatment in 0.01 mol \( \text{L}^{-1} \) citrate buffer, pH 6.0. Primary antibodies (Abs) were applied and incubated at 4°C overnight. Antigen retrieval was developed with peroxidase and 3,3'‐diaminobenzidine tetrahydrochloride. The sections were then counterstained with haematoxylin (Zymed Laboratories Inc.) and mounted in nonaqueous mounting medium. Antibodies information is

Figure 1. Tumour‐associated antigen (TAA) expression in hepatocellular carcinoma (HCC). (A) Immunohistochemical detection of GPC3, AFP, NY‐ESO‐1, SSX‐2, EpCAM, MDK in HCC tumour tissue (× 400 magnification). (B) Frequency of expression of the six TAAs in HCC tumour tissue determined by immunohistochemistry. (C) Frequency of coexpression of the indicated numbers of TAA (in any combination) in HCC tumour tissue. (D) TAA expression in each sample. Each column represents the TAA expression in an individual patient. Blue bar, positive expression. (E) The inter‐relationship between each TAA in HCC tumour tissue. Values denote the Pearson correlation coefficients; values closer to 1 indicate a better correlation. * \( P < 0.05 \); ** \( P < 0.001 \).
summarised in Supplementary Table 1. The appropriate negative controls were used, in which the primary Abs were replaced by irrelevant, isotype-matched Abs at the same concentration. If necessary, positive control tissues were applied as suggested by the manufacturers.

Evaluation of immunohistochemical variables. All TMA cores were screened and evaluated with a computerised image analysis platform constructed using the TMAJ Image application (http://tmaj.pathology.jhmi.edu). The expression of each antigen was scored according to the proportion of expression, as previously described (Xu et al., 2012b).

We initially investigated eight TAAs with identified immunogenicity and potential therapeutic function (Breous and Thimme, 2011): AFP (Thimme et al., 2008), GPC3 (Sawada et al., 2012), SSX-2 (Bricard et al., 2005), NY-ESO-1 (Korangy et al., 2004; Xu et al., 2012a), EpCAM (Yamashita et al., 2008), MDK (Jia et al., 2007; Kerzerho et al., 2010), melanoma antigen gene-A (MAGE-A) (Bricard et al., 2005), and telomerase reverse transcriptase (hTERT) (Mizukoshi et al., 2011). However, the expression of MAGE-A was below 10% and the immunodetection of hTERT was unsatisfactory due to nonspecific IHC staining in HCC (data not shown). For the six TAAs we eventually examined (AFP, GPC3, SSX-2, NY-ESO-1, EpCAM, and MDK), any proportion of any positive degree of intensity was considered positive. For the densities of lymphocytic infiltration, median cut-off points were used for the definition of subgroups.

Statistical analysis. Kaplan–Meier estimates were calculated and compared using the log-rank test. A multivariate Cox proportional hazard regression model was applied to estimate the adjusted hazard ratio (HR) and 95% confidence interval (CI) and to identify independent prognostic factors. The association between variables was evaluated using the χ² test or Fisher’s exact test when appropriate. P<0.05 was considered to indicate statistical significance. Statistical analyses were performed with SPSS 13.0 (SPSS Inc., Chicago, IL, USA).

RESULTS

Expression of TAAs in HCC tissue. Immunohistochemistry was performed to analyse the expression of TAAs in HCC parenchyma. Of eight initial TAAs, six (GPC3, AFP, SSX-2, NY-ESO-1, EpCAM, and MDK; Figure 1A) were eventually included and evaluated in a panel of 362 HCC patients. Most of the TAAs were frequently expressed and were detected in over 50% of the samples, except for NY-ESO-1, which was detected in only 14.6% (53/362) of the samples (Figure 1B). The frequency for coexpression of two to five TAAs was 11.9% (43/362), 26.5% (96/362), 29.8% (108/362), and 20.4% (74/362), respectively. Only 1.4% (5/362) of the samples revealed no expression of any TAA, and 5% (18/362), respectively. Only 1.4% (5/362) of the samples revealed no expression of any TAA, and 5% (18/362), respectively. Only 1.4% (5/362) of the samples revealed no expression of any TAA, and 5% (18/362), respectively.

Association between TAAs and disease progression. Patients were categorised into three groups according to the number of positive TAAs (TAA index) in the tumour tissue. As summarised in Table 2, tumour differentiation and tumour size were significantly associated with TAA expression (P = 0.009 and 0.018, respectively). In cases with less TAA expression, patients tended to have larger tumours and poor differentiation. For example, only 4.8% (14/289) of patients with grade I-II tumours expressed none or one of the six TAAs as compared with 12.3% (9/73) of patients with grade III-IV tumours, indicating that TAA deficiency might be another special feature of poorly differentiated tumours.

Kaplan–Meier survival curves were plotted to investigate the correlation between the expression of each TAA and patient survival. The log-rank statistic was used to compare survival rates. The expression of SSX-2, EpCAM, and MDK was positively correlated with better OS (P = 0.024, 0.0001, and 0.017, respectively; Figure 2). Interestingly, survival was better in the patients with a higher TAA index (Figure 2G). The OS of patients who expressed more than four TAAs (median survival, 60 months) was prolonged as compared with patients who expressed zero or one TAA (median survival, 18 months). The five-year OS rate in patients with 0-1 TAA expression was only 13.6% as compared with the 37.2% of those who expressed 4–6 TAAs. However, there was no significant association between TAA expression and recurrence in this cohort (Supplementary Figure 1).

| Table 2. Association between TAA expression and patients’ clinical characteristics |
|-------------------------------|-------------------------------|-----------------|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| Characteristics               | 0–1                          | 2–3             | 4–6               | P-value           |
| Age, year                     |                               |                 |                   |                   |
| Median                        | 43                           | 50              | 48                | 0.115             |
| Range                         | 25–62                        | 20–77           | 21–78             |                   |
| Gender                        |                               |                 |                   |                   |
| Male                          | 22                           | 124             | 178               | 0.609             |
| Female                        | 1                            | 15              | 22                |                   |
| Hepatitis B virus infection   |                               |                 |                   |                   |
| No                            | 4                            | 23              | 18                | 0.089             |
| Yes                           | 19                           | 116             | 182               |                   |
| Hepatitis C virus infection   |                               |                 |                   |                   |
| No                            | 22                           | 137             | 197               | 0.579             |
| Yes                           | 1                            | 2               | 3                 |                   |
| Alpha-fetoprotein             |                               |                 |                   |                   |
| < 25 ng ml⁻¹                  | 9                            | 48              | 50                | 0.097             |
| > 25 ng ml⁻¹                  | 14                           | 91              | 150               |                   |
| Child-Pugh class              |                               |                 |                   |                   |
| A                             | 20                           | 133             | 186               | 0.242             |
| B                             | 3                            | 16              |                   |                   |
| Tumour number                 |                               |                 |                   |                   |
| Single                        | 18                           | 101             | 155               | 0.568             |
| Multiple                      | 5                            | 38              | 45                |                   |
| Tumour size                   |                               |                 |                   |                   |
| < 5 cm                        | 2                            | 49              | 77                | 0.018             |
| > 5 cm                        | 21                           | 90              | 123               |                   |
| Vascular invasion             |                               |                 |                   |                   |
|Absent                         | 17                           | 118             | 174               | 0.239             |
|Present                        | 6                            | 21              | 26                |                   |
|TNM stage                      |                               |                 |                   | 0.366             |
| I–II                          | 15                           | 95              | 149               |                   |
| III                           | 8                            | 44              | 51                |                   |

Abbreviations: TAA — tumour-associated antigen; TNM — tumour-lymph node metastasis. Bold values indicate statistical significance.
We next assessed whether the TAA index could serve as an independent predictor of OS. A multivariate Cox proportional hazards analysis was performed, and the variables that were associated with survival by univariate analysis were adopted as covariates (summarised in Table 3). Serum AFP level, tumour differentiation, tumour size, and vascular invasion remained associated with OS in the multivariate Cox proportional hazards analysis ($P = 0.019, 0.002, 0.001,$ and $<0.0001$, respectively). The TAA index could still predict OS independent of these clinical factors (HR, 0.625; 95% CI, 0.467–0.837; $P = 0.002$; Table 3).

Association between TAA expression and local immune cell infiltration. Recent studies by our and other groups have shown that local immune cell status could influence HCC progression (Gao et al., 2007; Ding et al., 2009; Zhou et al., 2009; Wu et al., 2013). Therefore, we also assessed the infiltration of different type of immune cells (Supplementary Figure 2) in the tumour microenvironment, and their relationship with TAA expression. As shown in Figure 3 and Supplementary Table 2, there was a positive correlation between AFP or EpCAM and CD20$^+$ B cell or CD57$^+$ natural killer (NK) cell densities, but a negative correlation between SSX-2 and FoxP3$^+$ regulatory T (Treg) cells. However, NY-ESO-1 expression was significantly positively correlated with CD8$^+$ T, B, NK, and Treg cells. Moreover, patients who coexpressed more TAAs tended to have more B and NK cells, but not CD15$^+$ neutrophils, CD68$^+$ macrophages, or Treg cells infiltrating in the tumour. In addition, there was no correlation between the TAA index and immune cell infiltration in the peri-tumour region. Taken together, a higher TAA index could indicate a microenvironment with more anti-tumour immune cell infiltration but not tumour-educated regulatory cells in the HCC nest.

Prognostic values with combination of TAA index and immune infiltration. We next evaluated the combined influence of the TAA index and immune cell densities (CD3$^+$, CD4$^+$, CD8$^+$, CD20$^+$, CD57$^+$, and FoxP3$^+$ cells; Figure 4). Patients with a high TAA index (4–6 TAA coexpression) and high density of T (including CD3$^+$, CD4$^+$, and CD8$^+$ cells) and CD20$^+$ B or
had shorter survival time (median survival ≤28 months; Figure 4).

Supporting the general view that Treg cells suppress tumour immunity, low FoxP3þ Treg cell infiltration with a high TAA index predicted better OS (median survival >60 months) as compared with any other subgroup (P<0.01, Figure 4F). Patients with high Treg density and a low TAA index had the shortest survival time (median survival, 29 months). These results demonstrated that the TAA index and immune cell infiltration could together determine tumour progression and patient survival.

**DISCUSSION**

It is commonly believed that poorly immunogenic transformed cells that escape immune surveillance would lead to the primary appearance of overt cancer and/or post-treatment relapse; however, little is known about the clinical value of the immunogenic features of malignant cells in solid tumours, including HCC. This study provides the first evidence that the expression pattern of multiple TAAs is associated with cancer progression and postsurgical prognosis in HCC patients.

Several other groups have reported the expression frequencies of single TAA, mostly on an mRNA level (Yamashita et al, 2008; Wang et al, 2009). In the present study, we further demonstrated the expression and clinical value of multiple antigens on a protein level, which should be the critical reason for heterogeneous effectiveness of immunotherapy and provide supportive evidence for ‘immunoediting’ in human cancers. The host immune system not only controls tumour quantity, but also shapes tumour immunogenicity. The finding that the coexpression of more TAAs was associated with better-differentiated and/or smaller tumours and the association between the expression of several antigens can aid in our understanding of the nature of immunoedited tumour progression.

Therefore, the expression patterns of TAAs in human tumours should be a result of long-term co-evolution between the immune system and malignant cells, and would be closely related to patient prognosis after surgical interventions and/or other treatments such as immunotherapies. Further analysis of the immune cell infiltration provided more evidence for the association between TAAs and tumour progression. Generally, the expression of single TAAs was positively associated with the infiltration of different lymphocyte subsets, including CD3þ (Galon et al, 2006), CD8þ

**Table 3. Cox proportional Hazard regression analysis of patients’ overall survival**

| Variables                              | Hazard ratio | 95% CI     | P-value | Hazard ratio | 95% CI     | P-value |
|----------------------------------------|--------------|------------|---------|--------------|------------|---------|
| Gender (male vs female)                | 1.073        | 0.842–1.368 | 0.568   | NA           |            |         |
| Hepatitis B virus infection (absent vs | 1.218        | 0.781–1.901 | 0.384   | NA           |            |         |
| Hepatitis C virus infection (absent vs | 1.852        | 0.687–4.994 | 0.223   | NA           |            |         |
| Alpha fetoprotein (≤25 ng ml⁻¹ vs >25  | 1.66         | 1.189–2.316 | 0.003   | 1.505        | 1.070–2.118| 0.019   |
| Child–Pugh class (A vs B)              | 1.17         | 0.679–2.016 | 0.572   | NA           |            |         |
| Differentiation (I–II vs III–IV)       | 1.749        | 1.254–2.441 | 0.001   | 1.716        | 1.219–2.416| 0.002   |
| Tumour number (single vs multiple)     | 1.233        | 0.890–1.707 | 0.208   | NA           |            |         |
| Tumour size (≤5 cm vs >5 cm)           | 2.109        | 1.516–2.934 | <0.0001 | 1.796        | 1.266–2.547| 0.001   |
| Vascular invasion (absent vs present)  | 3.935        | 2.770–5.591 | <0.0001 | 3.399        | 2.061–5.605| <0.0001 |
| TNM stage (I–II vs III)                | 2.132        | 1.583–2.871 | <0.0001 | 0.926        | 0.595–1.442| 0.735   |
| TAA index (0–3 vs 4–6)                 | 0.626        | 0.471–0.833 | 0.001   | 0.625        | 0.467–0.837| 0.002   |

Abbreviations: CI = confidence interval; NA = not applicable; TAA = tumour-associated antigen; TNM = tumour-lymph node metastasis. Note: Cox proportional hazards regression model, variables associated with survival by univariate analysis were adopted as covariates in multivariate analyses. Bold values indicate statistical significance.

**Figure 3. The inter-relationship between tumour-associated antigen (TAA) expression and immune cell infiltration in tumoural and peri-tumoural regions.** Values denote the Pearson correlation coefficients; values closer to 1 indicate a better correlation. *P<0.05; **P<0.001.

CD57þ NK cells all had longer survival time (median survival >60 months). Patients with a low TAA index (0–3 TAA coexpression) and low density of these effective immune cells...
(Gao et al., 2007), CD20⁺ (Nielsen et al., 2012), and CD57⁺ cells (Wu et al., 2013), which have all been reported to facilitate anti-tumour immunity, and was negatively associated with FoxP3⁺ Treg cells (Zhou et al., 2009). Recently, Chew et al. (2012) reported that a given set of chemokines was correlated with lymphocyte infiltration and prognosis in HCC, which also support the protective role of anti-tumour immune milieu in HCC progression. Tumours coexpressing more TAAs tended to have more CD20⁺ B

Figure 4. The combination of tumour-associated antigen (TAA) expression and immune cell infiltration correlates with overall survival (OS). Kaplan–Meier curves illustrate the duration of OS according to the TAA index (TI) and the density of (A) CD3⁺, (B) CD4⁺, (C) CD8⁺, (D) CD20⁺, (E) CD57⁺, or (F) FoxP3⁺ cells in the tumour region. Red values indicate statistical significance.
and CD57⁺ NK cells, but not FoxP3⁺ Treg cells or other inflammatory cells, including CD15⁺ neutrophils (Kuang et al., 2011) and CD68⁺ macrophages (Ding et al., 2009), which are often exploited by solid tumours to establish a tumour-promoting microenvironment. Interestingly, the unexpected result of relationship between NY-ESO-1 and FoxP3 indicated that self-TAAs might also be involved in immune homeostasis and limiting acute inflammation. Taken together, the expression of more TAAs might promote anti-tumour immune reaction or surveillance and facilitate the postsurgical recovery of HCC patients.

It is conventionally believed that the adaptive immune response mediated by tumour-infiltrating lymphocytes (TILs) influences the behaviour of human cancers (Abastado, 2012). High densities of CD3⁺ T cells in the centre of a tumour and the invasive margin of colorectal tumours predict better clinical outcomes (Galon et al., 2006). For HCC, however, results by ours (Supplementary Figure 3) and other have shown that the infiltration of total T lymphocytes was not associated with postsurgical prognosis (Gao et al., 2007). In the present study, we stratified the patients according to the TAA index, and the density of CD3⁺ T cells was positively associated with the OS in patients with a low TAA index (Figure 4A). Subsequently, apart from the type and density of immune cells, features of the tumour itself, such as TAA expression, should also be important factors of immune regulation in HCC. Therefore, the combination of the TAA index and the densities of immune cells could further predict HCC prognosis. Recently, we developed an immune-based in situ molecular classifier that could aid in the identification of patients who are at greatest risk for postsurgical recurrence of HCC (Xu et al., 2012b). The predictive values of TAAs could provide more parameters to optimise molecular classifiers for HCC outcomes. Of course, other tumour cell features (such as proliferation) should also be considered important during early cancer evolution and later progression. In tumours with weak proliferation (low Ki-67), the TAA index was closely associated with better prognosis, while all of the patients with intensive proliferation had poor prognosis (Supplementary Figure 4). In general, the coactions of immunoe-diting and the vital power of tumour cells could continue shaping malignancies and influence patient survival after treatments, including resections and/or biological therapies.

Although clinical trials involving immunotherapy with T-cell clones specific for a single antigen have provided a foundation for proof-of-principle studies, reduced clinical efficacy has been encountered in contrast to the substantial therapeutic impact of transfer with polyclonal TIL cultures. The outgrowth of antigen-loss tumour variants in treated patients indicates the ability of rapidly adaptable tumour cells to evade narrowly focused therapies (Mellman et al., 2011; He et al., 2012). Recently, new therapies based on sophisticated knowledge of the suppressive tumour immune microenvironment were designed to overcome tolerance and reactivate anti-tumour immunity to induce potent, long-lasting responses (Mellman et al., 2011). For example, in early-phase clinical trials involving patients with advanced solid tumours such as metastatic melanoma, renal cell carcinoma, colorectal cancer, and non-small-cell lung cancer, monoclonal Abs against immune-checkpoint proteins (such as ipilimumab, tremelimumab, and MDX-1106) could induce a state of equilibrium between the immune system and cancer, resulting in prolonged disease stabilisation. Nevertheless, only a relatively small fraction of patients exhibited an objective response and derived clinical benefits (Topalian et al., 2011). In view of this, the discrepancies in the TAA profiles should be a critical reason for heterogeneous therapeutic efficacy. At present, immunotherapies that interrupt the tolerogenic pathways and reactivate endogenous immunity are being evaluated, appearing to be a promising HCC treatment option (primary or adjuvant for chemotherapy and/or surgery). To prevent overtreatment and to achieve more convincing results, molecular classification based on TAA expression patterns should also be an important strategy in clinical trials of immunotherapy.

In brief, TAA expression patterns could serve as important prognostic factors in HCC. Tumour-associated antigen expression should be associated with anti-tumour immune infiltration, and particularly, involved in disease progression and the reconstitution of immune surveillance after surgical intervention. Moreover, our results could provide a new evidence for improvement of the prognostic molecular signatures in HCC, and a potential rational consideration for patient enrolment in future immunotherapeutic trials and/or clinical treatments.

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CONFLICT OF INTEREST

The authors declare no conflict of interest.

REFERENCES

Abastado JP (2012) The next challenge in cancer immunotherapy: controlling T-cell traffic to the tumor. Cancer Res 72: 2159–2161.
Bécourt E, Thimme R (2011) Potential of immunotherapy for hepatocellular carcinoma. J Hepatol 54: 830–834.
Bricard G, Bouzourene H, Martinet O, Rimoldi D, Halkic N, Gillet M, Chaubert P, Macdonald HR, Romero P, Cerottini JC, Speiser DE (2005) Naturally acquired MAGE-A10- and SSX-2-specific CD8⁺ T cell responses in patients with hepatocellular carcinoma. J Immunol 174: 1709–1716.
Bruix J, Llovet JM (2009) Major achievements in hepatocellular carcinoma. Lancet 373: 614–616.
Chew V, Chen J, Lee D, Loh E, Lee J, Lim KH, Weber A, Slankamenac K, Poon RT, Yang H, Ooi LL, Toh HC, Heikenwalder M, Ng IO, Nardin A, Abastado JP (2012) Chemokine-driven lymphocyte infiltration: an early intratumoural event determining long-term survival in resectable hepatocellular carcinoma. Gut 61: 427–438.
Ding T, Xu J, Wang F, Shi M, Zhang Y, Li SP, Zheng L (2009) High tumour-infiltrating macrophage density predicts poor prognosis in patients with primary hepatocellular carcinoma after resection. Hum Pathol 40: 381–389.
Ding T, Xu J, Zhang Y, Guo RP, Wu WC, Zhang SD, Qian CN, Zheng L (2011) Endothelium-coated tumour clusters are associated with poor prognosis and micrometastasis of hepatocellular carcinoma after resection. Cancer 117: 4878–4889.
Disis ML (2010) Immune regulation of cancer. J Clin Oncol 28: 4531–4538.
Fan ST, Mau LoC, Poon RT, Yeung C, Leung Liu C, Yuen WK, Ming Lam C, Ng KK, Ching Chan S (2011) Continuous improvement of survival outcomes of resection of hepatocellular carcinoma: a 20-year experience. Ann Surg 253: 745–758.
Fridman WH, Pagès F, Sautès-Fridman C, Galon J (2012) The immune contexture in human tumours: impact on clinical outcome. Nat Rev Cancer 12: 298–306.
Fu YX (2012) New immune therapy targets tumor-associated environment: from bone marrow to tumor site. Cell Mol Immunol 9: 1–2.
Gao Q, Qiu SJ, Fan J, Zhou J, Wang XY, Xiao YS, Xu Y, Li YW, Tang ZY (2007) Intratumoral balance of regulatory and cytotoxic T cells is associated with prognosis of hepatocellular carcinoma after resection. J Clin Oncol 25: 2586–2593.
Gao Q, Shi Y, Wang X, Zhou J, Qiu S, Fan J (2012) Translational medicine in hepatocellular carcinoma. Front Med 6: 122–133.
Galon J, Costes A, Sanchez-Cabo F, Kirilovsky A, Mlecnik B, Lagorce-Pagès C, Tosolini M, Camus M, Berger A, Wind P, Zinzindohoué F, Bruneval P,
Cugnenc PH, Trajanoski Z, Fridman WH, Pagès F (2006) Type, density, and location of immune cells within human colorectal tumors predict clinical outcome. Science 313: 1960–1964.

Grivennikov SI, Greten FR, Karin M (2010) Immunity, inflammation, and cancer. Cell 140: 883–899.

He J, Tang XF, Chen QY, Mai HQ, Huang ZF, Li J, Zeng YX (2012) Ex vivo expansion of tumor-infiltrating lymphocytes from nasopharyngeal carcinoma patients for adoptive immunotherapy. Chin J Cancer 31: 287–294.

Jemal A, Bray F, Center MM, Ferlay J, Ward E, Forman D (2011) Global cancer statistics. CA Cancer J Clin 61: 69–90.

Korangy F, Ormandy LA, Bleck JS, Klempnauer J, Wilkens L, Manns MP, Greten TF (2004) Spontaneous tumor-specific humoral and cellular immune responses to NY-ESO-1 in hepatocellular carcinoma. Clin Cancer Res 10: 4332–4341.

Kuang DM, Zhao Q, Wu Y, Peng C, Wang J, Xu Z, Yin XY, Zheng L (2011) Peritumoral neutrophils link inflammatory response to disease progression by fostering angiogenesis in hepatocellular carcinoma. J Hepatol 54: 948–955.

Kerzerho J, Adotevi O, Castelli FA, Dosset M, Berardeau K, Szely N, Lang F, Tartour E, Maillere B (2010) The angiogenic growth factor and biomarker midkine is a tumor-shared antigen. J Immunol 185: 418–423.

Korany F, Ormandy LA, Bleck JS, Klempnauer J, Wilkens L, Manns MP, Greten TF (2004) Spontaneous tumor-specific humoral and cellular immune responses to NY-ESO-1 in hepatocellular carcinoma. Clin Cancer Res 10: 4332–4341.

Xu H, Gu N, Liu ZB, Zheng M, Xiong F, Wang SY, Li N, Lu J (2012a) NY-ESO-1 expression in hepatocellular carcinoma: A potential new marker for early recurrence after surgery. Oncol Lett 3: 39–44.

Xu J, Ding T, He Q, Yu XJ, Wu WC, Jia WH, Yun JP, Zhang Y, Shi M, Shao CK, Pan WD, Yin XY, Min J, Zhuang SM, Zheng L (2012b) An in situ molecular signature to predict early recurrence in hepatitis B virus-related hepatocellular carcinoma. J Hepatol 57: 313–321.

Yamashita T, Forgues M, Wang W, Kim JW, Ye Q, Jia H, Budhu A, Zanetti KA, Chen Y, Qin LX, Tang ZY, Wang XW (2008) EpCAM and alpha-fetoprotein expression defines novel prognostic subtypes of hepatocellular carcinoma. Cancer Res 68: 1451–1461.

Yang JD, Roberts LR (2010) Hepatocellular carcinoma: A global view. Nat Rev Gastroenterol Hepatol 7: 448–458.

Zhou J, Ding T, Pan W, Zhu LY, Li L, Zheng L (2009) Increased intratumoral regulatory T cells are related to intratumoral macrophages and poor prognosis in hepatocellular carcinoma patients. Int J Cancer 125: 1640–1648.

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