Rediscovery and phylogenetic relationships of the scolopendromorph centipede *Mimops orientalis* Kraepelin, 1903 (Chilopoda): a monotypic species of Mimopidae endemic to China, for more than one century

Chao Jiang¹, Yunjun Bai¹, Mengxuan Shi², Juan Liu¹

¹ State Key Laboratory of Dao-di Herbs Breeding Base, National Resource Center for Chinese Materia Medica, China Academy of Chinese Medical Sciences, Beijing, 100700, China ² School of Life Sciences, Henan University, Kaifeng, 475001, China

Corresponding author: Chao Jiang (jiangchao0411@126.com)

Abstract

*Mimops orientalis* Kraepelin, 1903 is a monotypic species of Mimopidae endemic to China. The species is known only from a single specimen, the holotype. Little is known about its biology, habitat associations, or phylogenetic relationships. It was rediscovered on Qinling Mountain in Shaanxi and Henan provinces, China, 117 years after its last record. Detailed descriptions and colour photographs of living specimens are provided along with its ecology, updated conservation notes, and data on sexual dimorphism. A genetic analysis (COI, 16S rRNA, and 28S rRNA) was conducted to assess the phylogenetic relationships among Mimopidae, Cryptopidae, Scolopendridae, Scolopocryptopidae, and Plutoniumidae. The results support classifying Mimopidae as a valid family.

Keywords

morphology, new distribution, Qinling Mountain
Introduction

*Mimops* Kraepelin, 1903 is one of the least-known genera of scolopendromorph centipedes. The type species, *Mimops orientalis* Kraepelin, 1903, described from a single specimen from Shaanxi Province, China, has a single white ocellus on each side of the cephalic plate. Since the original description, the exact type locality of this species has been unknown, and no additional findings of *M. orientalis* have been reported. The genus *Mimops* was initially placed in the Cryptopidae and subsequently redescribed by Lewis (2006) from the holotype of *M. orientalis*, establishing a new monotypic family, Mimopidae Lewis, 2006.

However, the only material of *M. orientalis*, the unsexed holotype specimen, did not allow Lewis (2006) to clarify the behaviour or information on the distribution, ecology, habitat, or phylogenetic relationships of *Mimops*. Because *M. orientalis* possesses characteristics from the families Plutoniumidae, Cryptopidae, Scolopocryptopidae, and Scolopendridae (e.g., forcipules and coxosternal plates without teeth as in Scolopocryptopidae, ultimate legs with numerous small spines as in Cryptopidae, and unpigmented ocellus-like pale areas as in Plutoniumidae), the taxonomic status of Mimopidae is still confusing, as are the phylogenetic relationships among Mimopidae, Cryptopidae, Scolopendridae, Scolopocryptopidae, and Plutoniumidae.

Throughout 2018 and 2019, we made intensive biodiversity surveys in the Taiping National Forest Park and Longyuwan National Forest Park, China. During this fieldwork, fresh material of *M. orientalis* was collected at the edge of Qinling Mountain. It is a region that has only few previous surveys for centipedes but has been highlighted as one of 16 biodiversity hotspots (Huang et al. 2016). Our rediscovery of this species has allowed us to reassess its taxonomic status, as well as that of the family Mimopidae, and to give fuller details of this species’ morphological variations and live coloration. In this paper, we redescribe the morphological characteristics of *M. orientalis* and construct a phylogeny based on COI and 16S mitochondrial DNA fragments and 28S nuclear DNA fragments.

Materials and methods

Taxonomic sampling

The studied individuals of *Mimops orientalis* were collected in Taiping National Forest Park, Shaanxi, China, and Funiushan National Nature Reserve, Henan, China. Other representative samples of Cryptopidae, Scolopendridae, Scolopocryptopidae, and Plutoniumidae were collected more generally in China. During the field study, some individuals were photographed with an Olympus E-M10 II camera to record their living colour pattern. The colour description is based on that of living centipedes. Specimens are deposited in the Institute of Chinese Materia Medica, China Academy of Chinese Medical Sciences, China (*CMMI*).
The taxonomic characteristics of the specimens were observed under an Olympus SZ16 stereomicroscope. Multifocus montage images were produced using Helicon Focus 6.7.1 software from a series of source images taken by a Canon 50D digital camera attached to the stereomicroscope. Terminology applied to the external anatomy follows Bonato et al. (2010).

All newly collected centipede specimens were stored in 75% ethanol. Legs 15 and 17 of the newly collected specimens were removed and stored in 100% ethanol for DNA extraction.

Phylogenetic sampling and DNA sequence analyses

Genomic DNA was extracted from leg tissue using a DNeasy Blood and Tissue Kit (Qiagen, Hilden, Germany). Two fragments of mitochondrial DNA (mtDNA), the encoded parts of the COI and 16S ribosomal RNA genes, and a fragment of 28S nuclear ribosomal RNA were amplified according to previously literature (Vahtera et al. 2012, 2013; Siriwut et al. 2016). PCR products were bidirectionally sequenced using an ABI 3730 DNA analyser (Applied Biosystems, CA, USA). New sequences determined in this study were aligned using ClustalW in BioEdit with default settings.

Maximum likelihood and Bayesian inference approaches were employed using MEGA X (Kumar et al. 2018) and PhyloSuite (Zhang et al. 2018), respectively, to determine the phylogenetic positions of *M. orientalis* based on the new sequences. Standard statistical tests were applied to evaluate branch support (bootstrap support and posterior probability). The best-fit substitution models and partitioning strategies were inferred for these three datasets using ModelFinder (Kalyaanamoorthy et al. 2017) integrated into PhyloSuite (Zhang et al. 2020). Based on Akaike Information Criterion, GTR+I+G4 was chosen as the best-fitting model for ML analyses and GTR+G for MrBayes analyses. Individual gene trees were constructed in MEGA using the maximum likelihood algorithm with 500 bootstrap replicates. Bayesian analyses were performed using MrBayes 3.2.6 (Ronquist et al. 2012) integrated into PhyloSuite with the default settings, and 10,000,000 MCMC generations, sampling every 100 generations. Sequences were deposited in GenBank with accessions of MT093838–MT093846, MT084401–MT084409, and MT084368–MT084375.

**Results**

*Mimops orientalis* Kraepelin, 1903

**Type material.** Holotype: China, Süd Schensi, August 1903, kept in Zoologisches Institut und Zoologisches Museum der Universität, Hamburg, Germany (ZMUH).

**Type locality.** Shaanxi, China. Perhaps referring to Xi’an, Shaanxi, China.

**Specimens examined.** Seven *Mimops orientalis* specimens were collected near a river in the Taiping National Forest Park, Hu county, Shaanxi, China (33.98N, 108.69E,
and another *M. orientalis* were collected in 2019 from Funiushan National Nature Reserve, Luanchuan, Henan, China. *Mimops orientalis* (*n* = 8). CMMI 20190714001, adult, under a stone near a river, Longyuwan National Forest Park, Luanchuan, Henan, China, (33.713N, 111.775E, 1110 m alt.) collected by Mengxuan Shi, on 14 Jul. 2019. CMMI 20190908001, adult male, under a stone near a ditch, Tai ping National Forest Park, Hu county, Shaanxi, China (33.98N, 108.69E, 630 m alt.), collected by Chao Jiang, on 08 Sept. 2019. CMMI 20190908002, juvenile, under a stone in some brushwood, Tai ping National Forest Park, Hu county, Shaanxi, China (33.98N, 108.69E, 620 m alt.), collected by Chao Jiang, on 08 Sept. 2019. CMMI 20190908003 and CMMI 20190908004, juvenile, under stones in grass halfway up a steep hill, Tai ping National Forest Park, Hu county, Shaanxi, China (33.98N, 108.69E, 640 m alt.), collected by Chao Jiang, on 08 Sept 2019. CMMI 20190908005, adult, under a trash bin near the road, Tai ping National Forest Park, Hu county, Shaanxi, China (33.98N, 108.69E, 626 m alt.), collected by Chao Jiang, on 08 Sept. 2019. CMMI 20190908006, subadult, under a stone near the collection site of specimen CMMI 20190908005, collected by Chao Jiang, on 08 Sept. 2019. CMMI 20190908007, juvenile, under a stone of brushwood, Tai ping National Forest Park, Hu county, Shaanxi, China (33.98N, 108.69E, 620 m alt.), collected by Chao Jiang, on 08 Sept. 2019.

**Other specimens for DNA sequencing.** Plutoniumidae: *Theatops chuanensis* Di et al., 2010, CMMI 20190405013, Tianzishan National Reserve, Zhangjiajie, Hunan province, China. CMMI 20190606004, Wen county, Longnan, Gansu province, China.

Cryptopidae: *Cryptops* sp., CMMI 20190413007, Longjin road, Huoshan county, Liu’an, Anhui province, China.

Scolopendridae: *Scolopendra mutilans* L. Koch, CMMI 20190331001, Qianfodong National Forest Park, Duodao distinct, Jinmen, Hubei province, China. CMMI 20190702006, Xiangshan, Dinghai distinct, Zhoushan, Zhejiang province, China.

Scolopocryptopidae: *Scolopocryptops nigrimaculatus* Song, Song & Zhu, 2004, CMMI 20181207002, Jinxihu hotel, West Lake, Hangzhou, Zhejiang province, China.

**Distribution.** Currently, we have confirmed that *M. orientalis* occurs on Qinling Mountain (Shaanxi and Henan provinces) and inhabits forests near rivers above 500–1200 m elevation (Fig. 1).

**Ecology.** Specimens of *M. orientalis* were collected from two locations on Qinling Mountain. We captured all of our new material at midday (11:00–13:00), always in forest edges near a river and under microhabitat refuges (rocks, bushes, leaf litter, and garbage). Two adult specimens were hiding under stones coated with moss, and another adult was encountered under some garbage containing rainwater. Most juveniles were encountered under small stones in bushes or moving over leaf litter in a fragmented patch of *Liquidambar formosana* Hance. Natural vegetation in the surrounding areas is deciduous forest, composed mainly of *Juglans regia* L., *Cotinus coggygria* Scop., and *L. formosana* Hance. Vegetative cover was ornamented by *Urtica* spp., *Viola* spp., *Cyperus iria* L., and *Oplismenus undulatifolius* (Arduino) Beauv (Figs 2, 3). We fed captive centipedes with mealworms, fish moths, and cockroaches, the latter two typically occurring in the centipedes’ microhabitat. The centipedes became active at approximately 20:00 most nights and would quickly escape to shelter when disturbed by noise.
Rediscovery of Mimopidae centipede in China

Figure 1. The current known distribution of *Mimops orientalis*. Two main record localities were in Shaanxi and Henan provinces, China, indicated by a red dot and a green pentagon, respectively; the red dot perhaps indicates the type locality.

**Live colouration.** Cephalic plate brownish red to orange-red, more deeply coloured in the anterior part. Coxosternites, forcipules, and tergites pale yellow to pale brown, ultimate segment and ultimate legs pale orange. Antennae, all sternites, and dorsal aspects of the legs light yellow. The live colour of a specimen from Shaanxi is slightly different from others. Its ultimate leg prefemur was nearly brown, which is more deeply coloured than that of the femur and tibia. The juvenile individual was uniformly light yellow on the cephalic plate, antennae, tergites, sternites, and all legs (Figs 4, 5).

**Redescription.** Lewis (2006) examined and described the holotype of *M. orientalis*. The majority of features of the newly collected specimens are in agreement with the holotype, but with some differences.

Adult length 42–56 mm. Cephalic plate smooth, about as long as it is wide, very finely punctate with the posterior margin overlying tergite 1. Cephalic plate lacks paramedian sulci in adults (two sulci in juvenile). A pale area instead of lateral ocelli at the base of each antenna (Lewis [2006] stated that is a single ocellus, whereas Kraepelin [1903] treated this as an eyespot). The animals show no response when the pale areas are illuminated with searchlights. Antenna extend to the posterior end of tergite 8, usually with 18 articles (holotype with 17 articles on the right antenna). Articles approximately 1.20–1.42 times as long as they are wide (based on article 4). Amount of hair on the antenna gradually increases, with the presence of glabrous articles variable (Lewis [2006] stated that the basal six articles were glabrous, while Kraepelin (1903) stated that the basal seven articles were glabrous). In our collected specimens, basal articles 4 and 5 were dorsally nearly glabrous (Figs 6–9), and articles 5–7 (Fig. 8, speci-
Figures 2–5. Habitat of *Mimops orientalis* 2, 3 Habitats at locality of *M. orientalis* in Taiping National Forest Park, Hu county, Shaanxi, China 4, 5 living specimens of *M. orientalis*.

men CMMI 20190908003) or 6 and 7 (Fig. 9, specimen CMMI 20190908004) had sparse hair; articles 8–18 were densely hirsute.

Tooth plates are short but wide, ridged and have approximately 25 ridges on each side and a prominent seta behind the anterior margin (Fig. 10). Forcipular trochanteroprefemur processes are very short, apex truncated, and with a long and acute process spine. Forcipular medial tibia and femur also have a prominent acute spine. Articles 1–3 of the second maxillary telopodite show hair and apical claw form a spine with long comb hair (Fig. 10).

Tergite 1 with anterior transverse (ring) sulcus (Fig. 6). Tergites 2–20 with complete paramedian sutures. Two longitudinal sulci lateral to the paramedian sutures on the five posterior tergites except the ultimate tergite. Tergites 2–20 without margination. Posterior part of the ultimate leg-bearing segment with complete margination. Sternites with complete paramedian sulci from 3 to 19, almost complete on sulcus 2. Nine round spiracles are present with one each on segments 3, 5, 8, 10, 12, 14, 16, 18, and 20. The spiracles protrude out of the segments (Figs 11, 12), and are cup-shaped, with a simple structure, and without humps (Fig. 13) (Lewis [2006] stated that the spiracles were filled with humps).

All legs with two-segmented tarsi. Two tibial spurs on legs 1–18, 19 with one, 20 without. Legs 1–20 each with two tarsal spurs (Fig. 14). Legs 1–5 with one outside femur spur dorsally, legs 1–3 also with one prefemur spur dorsally (Fig. 15). Legs 1–14 with
very few spines ventrally on the prefemur and femur and a distal transverse row dorsomedially on the prefemur, femur, and tibia (Fig. 14). Legs 15 and 16 with few spines ventrally on the prefemur and sparse spines ventrally on the femur. Legs 17–19 with prefemur, femur, and tibia thickly spined dorsally, medially, and ventrally, the spines on prefemur gradually increase from legs 17–19. Leg 20 with prefemur and femur thickly spined on all surfaces, tibia on all but the medial surface and tarsus 1 spined dorsally and medially.

Tergite 21 with small spines and a narrow posterior median depression (Fig. 17). Sternite 21 with sides converging posteriorly, without depression. Central zone of sternite 21 with sporadic small spines dorsal and ventral and peripheral zone with two to four rows of small spines (Fig. 16, 19). Coxopleuron with an oval pore field of many small pores and small scattered spines (Fig. 19). The coxopleural process was moderately long and digitiform, with small spines (Figs 19, 20). The prefemur and femur of the ultimate legs (Fig. 18) are covered with numerous small spines dorsally, medially, and ventrally, without grooves or strips (Lewis [2006] stated with the presence of a strip on the median ventral). Tibia spined on all but the median ventral and medial surfaces, and tarsus 1 has a few dorsomedial spines. Pretarsal accessory spurs are absent.

Figures 6–9. Head of *Mimops orientalis* Kraepelin, 1903 6, 7 Head and tergite 1 and 2, dorsal and ventral views 8, 9 dorsal views of the right antenna.
Genital segments well developed, reaching the distance between the posterior margin of sternite 21 and the distal part of the coxopleural process. In males, sternite of genital segment 1 round, with short setae ventrally, genital segment 2 round and convex, also with short setae ventrally. Penis columnar, with long setae dorsally. Gonopod present in males with seven or eight long setae (Fig. 19). Anal valve well developed, composite by two hemispheres, with a yellow strip near the posterior margin. Genital segments short in female, posterior margin of sternite of genital segment 1 with several spines. Anal valve composed of two hemispheres, with a cavity in the centre.

Conservation status. Although this species was found from two localities separated by 400 km, there is not yet enough information about the distribution, abundance, or threats to this species, and so further surveys are needed; we consider it Data Deficient for now (IUCN 2020).

Molecular analyses. We obtained 887 bp sequences of COI, 537 bp of 16S and 992 bp of 28S of *M. orientalis*. The complete matrix included sequences from 44 centipede species (Table 1), which consists of all five families of Scolopendromorpha. *Theatops chuanensis* Di et al., 2010, *Cryptops* sp., *Scolopendra mutilans* L. Koch, and *Scolopocryptops nigrimaculatus* Song, Song & Zhu, 2004 were also sequenced and subjected to phylogenetic analysis. These species represent the most common species of the families Plutoniumidae, Cryptopidae, Scolopendridae, and Scolopocryptopidae,
Figures 16–18. Ultimate segment and ultimate legs of *Mimops orientalis* 16 Ventral view of the ultimate segment 17 dorsal view of the ultimate segment 18 dorsal view of ultimate legs.
respectively, in China. For the class Chilopoda, which is commonly divided into five orders, namely Scolopendromorpha, Lithobiomorpha, Scutigeromorpha, Craterostigmomorpha, and Geophilomorpha, four representative species, *Scutigera coleoptrata* (Scutigeromorpha), *Craterostigmus tasmanianus* (Craterostigmomorpha), *Lithobius forficatus* (Lithobiomorpha), and *Bothriogaster signata* (Geophilomorpha) were selected as outgroups to assess the phylogenetic of *M. orientalis* as well as Mimopidae.

The results of the phylogenetic analysis are presented in Figures 21 and 22. Bayesian inference (BI) and maximum likelihood (ML) analyses yielded trees based on the combined alignments of COI + 16S + 28S, which demonstrated essentially consistent topologies. All Scolopendromorpha species used in this study could be considered as two main clades, the ocellate scolopendromorphs and the blind scolopendromorphs, both well supported in the ML tree. Mimopidae was positioned as sister group to the other families of blind scolopendromorphs. However, Bayesian inference analyses identified Scolopendromorpha species as belonging to three clades, the ocellate scolopendromorphs, the blind scolopendromorphs sensu Pocock, 1896 (comprising Plutoniumidae, Cryptopidae, and Scolopocryptopidae), and the monophyletic family Mimopidae with quite strong support (PP 0.94). Both methods obtained from the molecular data place Scolopendridae as a monophyletic family, comprising two subfamilies, Scolopendrinae and Otostigmidae. Likewise, Scolopocryptopidae comprises two subfamilies, Scolopocryptopinae and Newportiinae. Cryptopidae, and Scolopocryptopidae are sister groups together with Plutoniumidae in the molecular analysis.
Rediscovery of Mimopidae centipede in China

Figures 21, 22. Phylogenetic trees obtained from the COI, 16S, and 28S sequences 21 ML tree from the analysis of the concatenated genes 22 tree from the Bayesian analysis of all genes combined. Numbers at the nodes are bootstrap percentages obtained from the ML analyses and posterior probabilities obtained from BI.
**Table 1.** List of the 44 selected sequences obtained from this study GenBank nucleotide database used in phylogenetic analyses.

| Taxonomy          | Species                                  | Voucher number | Taxon locality         | GenBank accession number          | Reference |
|-------------------|------------------------------------------|----------------|------------------------|-----------------------------------|-----------|
|                   | **Mimopidae**                            |                |                        | COI                               |           |
|                   | *Mimops orientalis*                      | CMMI 20190908002 | Shaanxi, China         | MT093838 MT084401 MT084368        | This study|
|                   | *Mimops orientalis*                      | CMMI 20190908003 | Shaanxi, China         | MT093839 MT084402 MT084369        | This study|
| Cryptopidae       | *Mimops orientalis*                      | CMMI 20190908006 | Shaanxi, China         | MT093840 MT084403 MT084370        | This study|
|                   | *Cryptops (Cryptops) trisulcatus*        | IZ-130578      | Italy                  | HQ402544 HQ402493 AF000783        | 1         |
|                   | *Cryptops (Cryptops) doriae*             | IZ-130581      | Thailand               | KF676500 KF676458 KF676354        | 1         |
|                   | *Cryptops (Trigonocryptops) galathaeae*  | AM KS 58457    | Australia              | KY288743 KY288724 KY288709        | 1         |
|                   | *Cryptops (Paracryptops) indicus*        | IZ-130608      | Vietnam                | KF676505 KF676463 KF676357        | 1         |
|                   | *Cryptops sp.*                           | CMMI 20190908002 | Liaoning, China       | MT093841 MT084404 MT084371        | This study|
| Plutoniumidae     | *Theatops erythrocephalus*               | IZ-130611      | Portugal               | HM453313 HM453222 AF000784        | 1         |
|                   | *Theatops posticus*                      | IZ-131448      | USA                    | AY288766 AY288727 –              | 1         |
|                   | *Theatops chuanaensis*                   | CMMI 20190908002 | Hunan, China          | MT093842 MT084405 MT084372        | This study|
|                   | *Theatops chuanaensis*                   | CMMI 20190908002 | Gansu, China          | MT093843 MT084406 MT084373        | This study|
|                   | *Plutonium zwierleni*                    | PD-0-0221      | Italy                  | LN890292 LN890289 LN890291        | 1         |
| Scolopocryptopidae| *Newportia divergens*                    | IZ-130770      | Mexico                 | JX422667 JX422690 KF676359        | 1         |
|                   | *Newportia adisi*                        | IZ-130770      | Brazil                 | KF676506 KF676465 JX422586        | 1         |
|                   | *Scolopocryptops nigridius*              | IZ-130806      | USA                    | JX422680 JX422704 JX422594        | 1         |
|                   | *Scolopocryptops rubiginosus*            | IZ-130823      | Taiwan                 | JX422682 JX422706 –              | 1         |
|                   | *Scolopocryptops nigriculatus*           | CMMI 20181207002 | Hangzhou, China       | MT093844 MT084407 –              | This study|
|                   | *Scolopocryptops sexipunctatus*          | IZ-131450      | USA                    | AM288745 AM288726 AM288710        | 1         |
| Scolopendridae    | *Scolopendra subspinipes*                | IZC 00146521   | Martinique             | HQ402554 HQ402502 HQ402538        | 1         |
|                   | *Scolopendra mutilans*                   | CMMI 20190908002 | Hubei, China          | MT093845 MT084408 MT084374        | This study|
|                   | *Scolopendra mutilans*                   | CMMI 20190908002 | Zhejiang, China       | MT093846 MT084409 MT084375        | This study|
|                   | *Scolopendra viridis*                    | IZ-130727      | USA                    | DQ201431 DQ201425 DQ222134        | 1         |
|                   | *Cormocephalus hartmeyeri*               | IZ-130632      | Australia              | KF676531 KF676491 KF676391        | 1         |
|                   | *Cormocephalus monteithi*                | IZ-130638      | Australia              | DQ201430 AF370861 AF173280        | 1         |
|                   | *Cormocephalus pseudopunctatus*          | IZ-130646      | South Africa           | KF676534 KF676493 KF676398        | 1         |
|                   | *Hemiscolopendra marginata*              | IZ-130659      | USA                    | HQ402548 HQ402496 –              | 1         |
| Taxonomy        | Species                        | Voucher number | Taxon locality | GenBank accession number | Reference |
|-----------------|--------------------------------|----------------|----------------|--------------------------|-----------|
| Scolopendridae  | *Arthrorhabdus formosus*       | AMNH LP6656    | South Africa   | HQ402539 HQ402488 HQ402522 | 1         |
|                 | *Scolopendrina orientalis*     | IZ-130623      | New Caledonia  | HQ402542 HQ402491 KF676404 | 1         |
|                 | *Notiasemus glauerti*          | WAMT124343     | Australia      | KF676539 KF676498 KF676405 | 1         |
|                 | *Oeostigmus astenus*           | IZ-130669      | Fiji           | HM453312 HM453221 HQ402532 | 1         |
|                 | *Oeostigmus politus politus*   | IZ-130674      | China          | KF676512 KF676470 KF676368 | 1         |
|                 | *Digitipes barnabasi*          | IZ-130664      | Taiwan         | KF676513 KF676471 KF676369 | 1         |
|                 | *Digitipes coonoensis*         | CES 091355     | India          | JX531905 JX531775 JX531826 | 2         |
|                 | *Ethmostigmus curtipes*        | CES 08960      | India          | JX531850 JX531720 JX531793 | 2         |
|                 | *Ethmostigmus rubripes rubripes* | IZ-130653   | Australia      | KF676515 KF676474 KF676372 | 1         |
|                 | *Rhysida atra*                | IZ-130677      | South Africa   | HQ402552 HQ402500 HQ402536 | 1         |
|                 | *Rhysida polyacantha*          | IZ-130665      | Australia      | KF676518 KF676476 KF676376 | 1         |
| Lithobiomorpha  | *Lithobius forficatus*         |               |               | AJ270997 AJ270997 EF199984 | 3         |
| Craterostigmomorpha | *Craterostigmus tasmanianus* | IZ-132143      | Australia      | EU024611 EU024597 HM453265 | 1         |
| Geophilomorpha  | *Bothriogaster signatus*       | IZ-131562      | Greece         | AY288749 AY288730 HM453290 | 1         |
| Scutigeromorpha | *Scutigera coleoptrata*        | IZ-130904      | South Africa   | DQ222170 DQ222156 EF199983 | 1         |

1 = Vahtera et al. 2013; 2 = Joshi and Karanth 2012; 3 = Hwang et al. 2001.
Discussion

The taxonomic position of Mimops has been controversial for many years. Attems (1930) divided scolopendromorphs into blind scolopendromorphs and ocellate scolopendromorphs, the latter containing only a single family, Scolopendridae, whereas the former includes three eye-less scolopendromorph families (Cryptopidae, Plutoniumidae, and Scolopocryptopidae), hence, the name blind clade. The genus Mimops was first classified into the subfamily Cryptopinae of the family Cryptopidae by Attems (1930) and then removed from Cryptopinae in a cladistic analysis by Schileyko and Pavlinov (1997). Shelley (2002) moved Mimops to the Scolopendridae, and subsequently Lewis (2006) restudied the holotype and placed it in a new family, Mimopidae. Most recently, Edgecombe and Giribet (2019) also treated Mimops as an ocellate scolopendromorph in the monotypic family Mimopidae. A critical controversy focused on the presence or absence of ocelli in M. orientalis. Attems (1930) and Schileyko treated the “Augenfleck” not as a real eye and placed the species in the blind clade, whereas Lewis, Shelley, and Edgecombe considered it as a single ocellus. With careful inspection, we found that M. orientalis has two round pale areas, rather than ocelli, at the base of the antenna. The pale area of M. orientalis is identical to that of Theatops chuanensis of the family Plutoniumidae, but absent in Scolopocryptopidae (Figs 23–27). However, ocelli are not an arbitrary family characteristic in scolopendromorphs. Tonkinoden- tus Schileyko, 1992, an eye-less scolopendrid-like genus, was unequivocally supported inside Scolopendridae by morphological and molecular data (Schileyko et al. 2019).

Three recently described scolopendrid centipedes, Cormocephalus sagmus Edgecombe et al. 2019, C. pyropygus Edgecombe et al. 2019, and C. delta Edgecombe et al. 2019, form a new blind species group in the genus Cormocephalus Newport, 1844 which otherwise presents four ocelli. Mimops also shares many scolopendrid-like characteristics, but distinct differences from Tonkinodentus and Cormocephalus imply that eye loss may have occurred several times in the evolution of Scolopendromorpha.

Mimops orientalis also possesses several unusual characteristics among Plutoniumidae, Cryptopidae, Scolopocryptopidae, and Scolopendridae. It shares many characteristics with scolopendrid centipedes, including a smooth cephalic plate without setae or spines, several glabrous basal antennal articles, the presence of sternite and tergite paramedian sutures, the presence of spines on the prefemora of the ultimate legs, and an anterior transverse sulcus on tergite 1. These characteristics render it similar to Scolopendra species, particularly many New World varieties. However, M. orientalis lacks lateral ocelli and instead possesses a pale area, which is only present in plutoniumids, and the spines on the ultimate legs are very small, which is similar to some Trigonocryptops species, e.g., Cryptops (Trigonocryptops) camouwealensis Edgecombe, 2006. This renders the taxonomic position of Mimops puzzling.

In this study, we confirmed Mimopidae as a valid family based on both morphological and molecular data. We also place Mimops as the basal taxa of scolopendromorphs for the following reasons: 1) M. orientalis is resolved as monophyletic both in maximum likelihood and Bayesian inference analyses under all parameter sets that were
Rediscovery of Mimopidae centipede in China

explored; 2) the ocelli of Mimops are regressed and have left two pale areas, whereas the outgroups Lithobiomorpha, Scutigeromorpha, and Craterostigmomorpha all have ocelli (or a compound eye with ommatidia in the case of Scutigeromorpha), indicating that ocelli may be a plesiomorphic characteristic; 3) Mimops bears 21 trunk segments, whereas 21 to 23 segments is an unreversed apomorphy of Scolopocryptopidae; 4) the long and acute spines on the forcipular trochanteroprefemoras of Mimops may be a primitive characteristic of trochanteroprefemur process; and 5) the small homologous spines on the ultimate legs, coxopleural process and ultimate sternites seem to be an intermediate state between setae and spines. However, although BI and ML analyses both support the validity of Mimopidae, the precise position of Mimops is not delimited due to the unresolved trichotomy in BI tree. Further research based on complete mitochondrial genome or transcriptomic data may help to clarify phylogenetic relationships among Mimopidae and other families.

Figures 23–27. Ocelli characteristics of Scolopendromorpha 23 Mimops orientalis 24 Theatops chuanensis 25 Scolopendra mutilans 26 Scolopocryptops nigrimaculatus 27 Cryptops sp.
Acknowledgements

The authors sincerely thank Mr Jiazhou Lu for help collecting specimens and taking photographs of living specimens. This work was supported by National Key R & D Program of China (2019YFC1711000) and the Fundamental Research Funds for the Central public welfare research institutes (ZZXT201908).

References

Bonato L, Edgecombe GD, Lewis JG, Minelli A, Pereira LA, Shelley RM, Zapparoli M (2010) A common terminology for the external anatomy of centipedes (Chilopoda). ZooKeys 69: 17–51. https://doi.org/10.3897/zookeys.69.737

Edgecombe GD (2006) A troglobitic cryptopid centipede (Chilopoda: Scolopendromorpha) from western Queensland. Records of the Western Australian Museum 23 (2): 193–198. https://doi.org/10.18195/issn.0312-3162.23(2).2006.193-198

Edgecombe GD, Giribet G (2019) The molecularization of centipede systematics. In: Fusco G (Ed.) ‘Perspectives on Evolutionary and Developmental Biology Essays for Alessandro Minelli’. Padova University Press, London, 154–165.

Edgecombe GD, Huey JA, Humphreys WF, Hillyer M, Burger MA, Volschenk ES, Waldock JM (2019) Blind scolopendrid centipedes of the genus Cormocephalus from subterranean habitats in Western Australia (Myriapoda: Scolopendromorpha: Scolopendridae). Invertebrate Systematics 33 (6): 807–824. https://doi.org/10.1071/IS19015

Huang J, Huang J, Liu C, Zhang J, Lu X, Ma K (2016) Diversity hotspots and conservation gaps for the Chinese endemic seed flora. Biological Conservation 198: 104–112. https://doi.org/10.1016/j.biocon.2016.04.007

Hwang UW, Friedrich M, Tautz D, Park CJ, Kim W (2001) Mitochondrial protein phylogeny joins myriapods with chelicerates. Nature 413(6852): 154–157. https://doi.org/10.1038/35093090

Joshi J, Karanth KP (2012) Coalescent method in conjunction with niche modeling reveals cryptic diversity among centipedes in the Western Ghats of South India. PLoS ONE 7 (8): e42225. https://doi.org/10.1371/journal.pone.0042225

Kalyaanamoorthy S, Minh BQ, Wong TKF, von Haeseler A, Jermiin LS (2017) ModelFinder: fast model selection for accurate phylogenetic estimates. Nature Methods 14: 587–589. https://doi.org/10.1038/nmeth.4285

Kraepelin K (1903) Revision der Scolopendriden. Mitteilungen aus dem Naturlhistorischen Museum, Hamburg, 62 pp.

Kumar S, Stecher G, Li M, Knyaz C, Tamura K (2018) MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. Molecular Biology and Evolution 35(6): 1547–1549. https://doi.org/10.1093/molbev/msy096

Lewis JG (2006) On the scolopendromorph centipede genus Mimops Kraepelin, with a description of a new family (Chilopoda: Scolopendromorpha). Journal of Natural History 40(19–20): 1231–1239. https://doi.org/10.1080/00222930600861231
Ronquist F, Teslenko M, van der Mark, P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. System Biology 61: 539–542. https://doi.org/10.1093/sysbio/sys029

Schileyko AA, Pavlinov IJ (1997) A cladistic analysis of the order Scolopendromorpha (Chilopoda). Entomologia Scandinavica 51 (Suppl.): 33–40.

Schileyko, AA, Solovyeva EN (2019) On the taxonomic position of the enigmatic genus *Tōkinodentus* Schileyko, 1992 (Chilopoda, Scolopendromorpha): the first molecular data. ZooKeys 840: 133–155. https://doi.org/10.3897/zookeys.840.33635

Shelley RM (2002) A synopsis of the North American centipedes of the order Scolopendromorpha (Chilopoda). Virginia Museum of Natural History 5: 1–108.

Siriwut W, Edgecombe GD, Sutchart C, Tongkerd P, Panha S (2016) A taxonomic review of the centipede genus *Scolopendra* Linnaeus, 1758 (Scolopendromorpha, Scolopendridae) in mainland Southeast Asia, with description of a new species from Laos. ZooKeys 590: 1–124. https://doi.org/10.3897/zookeys.590.7950

Vahtera V, Edgecombe GD, Giribet G (2012) Evolution of blindness in scolopendromorph centipedes (Chilopoda: Scolopendromorpha): insight from an expanded sampling of molecular data. Cladistics 28 (1): 4–20. https://doi.org/10.1111/j.1096-0031.2011.00361.x

Vahtera V, Edgecombe GD, Giribet G (2013) Phylogenetics of scolopendromorph centipedes: can denser taxon sampling improve an artificial classification? Invertebrate Systematics 27(5): 578–602. https://doi.org/10.1071/IS13035

Zhang D, Gao F, Jakovlić I, Zou H, Zhang J, Li WX, Wang GT (2020) PhyloSuite: an integrated and scalable desktop platform for streamlined molecular sequence data management and evolutionary phylogenetics studies. Molecular Ecology Resources 20(1): 348–355. https://doi.org/10.1111/1755-0998.13096