Plasmodium falciparum antigenic variation: relationships between widespread endothelial activation, parasite PfEMP1 expression and severe malaria

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Abstract

Background: Plasmodium falciparum erythrocyte membrane protein 1 (PfEMP1) is a family of variant surface antigens (VSA) that mediate the adhesion of parasite infected erythrocytes to capillary endothelial cells within host tissues. Opinion is divided over the role of PfEMP1 in the widespread endothelial activation associated with severe malaria. In a previous study we found evidence for differential associations between defined VSA subsets and specific syndromes of severe malaria: group A-like PfEMP1 expression and the "rosetting" phenotype were associated with impaired consciousness and respiratory distress, respectively. This study explores the involvement of widespread endothelial activation in these associations.

Methods: We used plasma angiopoietin-2 as a marker of widespread endothelial activation. Using logistic regression analysis, we explored the relationships between plasma angiopoietin-2 levels, parasite VSA expression and the two syndromes of severe malaria, impaired consciousness and respiratory distress.

Results: Plasma angiopoietin-2 was associated with both syndromes. The rosetting phenotype did not show an independent association with respiratory distress when adjusted for angiopoietin-2, consistent with a single pathogenic mechanism involving widespread endothelial activation. In contrast, group A-like PfEMP1 expression and angiopoietin-2 maintained independent associations with impaired consciousness when adjusted for each other.

Conclusion: The results are consistent with multiple pathogenic mechanisms leading to severe malaria and heterogeneity in the pathophysiology of impaired consciousness. The observed association between group A-like PfEMP1 and impaired consciousness does not appear to involve widespread endothelial activation.

Keywords: Group A-like, Angiopoietin-2, Rosetting, Sequestration, Impaired consciousness, Cerebral malaria, Respiratory distress

Background

Severe life threatening P. falciparum malaria is a major cause of mortality and morbidity in young children in sub-Saharan Africa. In endemic areas, severe malaria is most prevalent in children under the age of five years, before they acquire immunity to severe disease as a result of repeated exposure. Severe malaria manifests in children in three partly overlapping syndromes; impaired consciousness (IC), respiratory distress (RD), and severe malarial anemia (SMA) [1]. Of these, IC and RD were found to be the key indicators of life threatening malaria in a hospital setting in sub-Saharan Africa [1].

The pathophysiological process underlying each of these syndromes is still not understood. Disturbed microcirculation is thought to play a major role [2]. In cerebral malaria (CM) (severely impaired consciousness), sequestration of parasite infected erythrocytes (IE) in the microvasculature of the brain is thought to be important [3-6].

PfEMP1, a parasite encoded protein expressed on the surface of the IE, interacts with host receptors on the microvascular endothelia as well as unparasitized erythrocytes (a phenotype referred to as rosetting) leading to
sequestration of the IE in organs. PfEMP1 is therefore thought to play a central role in parasite virulence. PfEMP1 is encoded by about 60 var genes per parasite genome and undergoes antigenic variation. Switches in the expression of the repertoire of var genes results in a high degree of plasticity in the antigenic and adhesive properties of the infecting parasite population.

Epidemiological studies have shown a subset of var genes preferentially expressed in young and non-immune children to be associated with severe malaria especially in children with IC [7-10]. This is consistent with 1) the hypothesis that some PfEMP1 variants have growth advantage in immunologically naïve children as the result of exhibiting a superior ability to sequester [11-18] and 2) the observed relationship between the density of sequestration in vital organs such as the brain and fatal malaria [4,5,18]. Beside sequestration of IE, severe malaria is characterised by systemic endothelial activation and widespread release of activation markers such as von Willebrand factor (vWF) [19], soluble ICAM-1(sICAM-1) [20] and angiopoietin-2 (ang-2) [21,22]. As sequestration occurs in the endothelial cells (ECs) of the microvasculature, it is believed that PfEMP1 mediated adhesion of parasitized red blood cells to the host microvasculature induces endothelial activation compromising the vascular integrity [23,24]. Recently, endothelial activation markers such as ang-2, soluble Tie-2 receptor, vWF have been shown to be associated with severe malaria [22,25]. Furthermore ang-2 is associated with retinopathy [25], a feature identified as a surrogate marker for cerebral sequestration [5,26] and a recent study found fibrin deposition in the brain to be associated with sequestration of IE [27]. If there is a connection between parasite var expression patterns and disease severity, through mechanisms involving sequestration and endothelial activation we would expect to observe a relationship between the expression of the var subset associated with severe malaria and markers of endothelial activation.

Previously, we showed through var expression profiling of 217 clinical P. falciparum isolates, that expression of a specific subset of “group A-like” PfEMP1 types is associated with severe malaria [8]. This was based on PCR amplification of a region within the DBLα domain of PfEMP1 and sequencing. Moreover, we showed that expression of these group A-like PfEMP1 types is associated primarily with the severe syndromes of IC, while the parasite rosette phenotype was associated primarily with RD [8,10]. In previous post-mortem studies of Thai adults [3,4] and African children [5], the severity of IC was shown to correlate with sequestration of IE by direct binding to the vascular endothelia [18]. The subgroup of vars expressed by the infecting parasites on the surface of IE may therefore determine the level of endothelial activation as suggested by [28].

In this study, we explored whether a relationship exists between widespread endothelial activation (represented by plasma ang-2 levels), parasite VSA expression and severe malaria. Specifically, we investigated whether widespread endothelial activation could provide a causal link between the expression of the group A-like vars and IC on the one hand and rosette frequency and RD on the other. To this end we measured ang-2 levels in the plasma of children from Kilifi, Kenya, presenting with either severe or non-severe malaria.

**Methods**

**Ethics statement**

Ethical approval for this study was obtained from Kenya Medical Research Institute (KEMRI) Ethical Review Committee (SSC 1131), and written, informed consent was obtained from parents/guardians of the study participants.

**Sample collection and clinical classification of patients**

The method of sample collections was described in detail in [8,10]. Severe malaria cases include children with microscopically confirmed P. falciparum infection and admitted to the hospital ward with impaired consciousness (Blantyre coma score (bcs) ≤ 5 in patients aged ≥ 8 months or ≤ 3 in patients aged under 8 months) [29], respiratory distress (deep “Kussmaul” breathing) [30], severe malarial anemia (haemoglobin < 5 g/dl) [1]. The severe form of impaired consciousness was defined as cerebral malaria (bcs ≤ 2).

**Sampling of DBLα-tag sequence and classification**

The method for amplification of DBLα-tag from cDNA has been described in [31] and sequence classification for the dataset used in this study is described in [8,10]. Briefly, a blood sample was taken from all the children at admission. After removal of the white blood cells (WBC), an aliquot (~100 µl) of the erythrocyte portion was directly re-suspended in TRIzol and kept at −80 degrees till use. RNA was extracted from the frozen TRIzol and cDNA synthesized using random hexamers. Using degenerate primers a highly conserved region within the DBLα-domain of vars was PCR amplified, cloned into TOPO T-cloning vector, transformed into E. coli, and 100 colonies from each sample sequenced. The sequences were classified as described in [31,32]. Briefly they were classified based on the number of cysteine residues present (majority containing either two cysteine (cys2) or four cysteine (cys4) [31]. The sequences were also classified using network analysis according to whether they fall into groups that tend not to share polymorphic regions [33]. Group A-like vars belong to cys2 and block sharing group 1 [33]. Each subgroup was expressed as a percentage of the total sequences.
Rosetting, IE surface antibody, and ang-2

Rosette frequency data was obtained as described previously [10]. IE surface antibodies data was obtained as described in [10]. Briefly we measured each participant’s IgG antibody levels (acquired as mean fluorescent intensity (MFI)) to the infected erythrocyte surface of eight ex vivo clinical isolates grown to the trophozoite stage using flow cytometry. The eight isolates were selected on the basis of their var expression (ranging from high to low cys2 expression). The median MFI value of this IE surface recognition by IgG against the eight isolates was calculated for each participant [10].

Ang-2 level in the acute plasma was determined by a commercial ELISA Kit from R & D (cat no; DANG20) using the manufacturer’s protocol.

PfHRP2 ELISA

Plasma PfHRP2 level was determined from the acute plasma using ELISA. Three plasma samples from patients with high P. falciparum parasitemia were used to determine both the dynamic range and the concentration of PfHRP2 using recombinant PfHRP2 protein obtained from MyBioSource (Cat no MBS232321). These plasma samples were then used to construct a standard curve. The ELISA assay was performed in duplicates in dilutions ranging from 1: 50 to 1:2000 and concentration (ng/ml) calculated from the linear range of the standard curve. Samples with OD readings greater than 20% standard deviation in the duplicate well was repeated or excluded from the analysis.

Statistical analysis

Stata version 11 was used for all statistical analyses and P ≤0.05 was considered significant. Correlations between variables were evaluated using Spearman’s rank correlation coefficient or the 2-sample Wilcoxon rank-sum (Mann–Whitney) test. Linear regression was used when the outcome is a continuous variable and logistic regression when binary. All the regression analyses were adjusted for age. All independent variables that were significant in a univariate analysis were included in models with, either IC, or RD variable. Before use in a multivariable linear regression model that considered host explanatory variables to the fit of a simpler regression model.

Results

Patients characteristics

The samples used were from a published study [8] that involved 217 study subjects. Of these, 213 had sufficient plasma available to be included (see Additional file 1: Figure S1 for patient characteristics). Previously, the var expression profiles of the infecting parasites from these children were determined [10] together with a measure of the breadth of antibodies against antigens on the surface of the IE [8] present at the time of disease. Rosette frequency was determined for parasites from 130 of the samples [10].

We used ang-2 as a marker of widespread endothelial activation because unlike sICAM-1 [34] and vWF [35,36] that can be released from non-ECs or ang-1 which is entirely from non-endothelial source, ang-2 is only released from activated ECs [37,38]. Moreover, ang-2 is relatively stable under freeze-thaw cycling [39].

Ang-2 levels are associated with the severe malaria syndromes; impaired consciousness and respiratory distress

Consistent with previous studies [21,25], analysis of 213 children’s plasma showed that ang-2 levels were higher in children with severe malaria compared to those who were non-severe (Figure 1A: Mann–Whitney U test Z = 6.5, P < 0.0001, N = 213).

Since severe malaria in African children presents in three overlapping syndromes [1], these syndromes may be differentially associated with markers of endothelial activation. To explore this, we used ang-2 as the dependent variable in a multivariable linear regression model that considered host age, IC, and RD as explanatory variables (SMA was not included due to few samples with this clinical phenotype). Both RD and IC displayed significant and independent associations with ang-2 (Figure 1B) suggesting that endothelial activation may play a role in the pathophysiology of both syndromes.

The association of IE surface antigen profiles with plasma angiopoietin-2

Given the potential link between cytoadherence and endothelial activation we explored two measures of variant surface antigen (VSA) expression: rosetting and group-A like var, in relation to ang-2 levels. A significant positive correlation was found between ang-2 and both
group A-like var expression (Figure 2A) and rosetting (Figure 2B). To explore whether these relationships between the measures of VSA and ang-2 were due to direct parasite:host interactions, we tested whether the associations were maintained when we split the analysis into severe and non-severe cases (Figure 2C-F). Neither group A-like expression nor rosetting showed evidence for an association with ang-2 among non-severe cases (Figure 2E-F). This suggests that disease severity is a confounder, i.e. the observed associations between VSA expression and ang-2 are due to the fact that both are associated with disease severity rather than their direct associations with each other.

Group A-like expression and ang-2 are independently associated with impaired consciousness

We explored further whether the previously observed relationship between the group A-like var subgroup expression and IC [10] may involve widespread endothelial activation. We did not consider rosetting here because we showed previously that it has no association with IC within this dataset [10].

We used age-adjusted logistic regression models predicting IC with group A-like var expression and ang-2 as independent variables. First, each of them was used as the only explanatory variable in an age-adjusted model and then in combination to adjust for one another. As shown in Figure 3A and Table 1, model 6A, despite a moderate reduction in the odds ratio (OR) when adjusted for ang-2, the group A-like var subgroup expression remained significantly associated with IC. Similarly, ang-2 remained significantly predictive of IC when adjusted for group A-like var expression (Figure 3B and Table 1, model 6A). A positive improvement to the fit of the model was observed when both variables were considered compared to either considered alone (likelihood ratio \( \chi^2 = 22.5, P < 0.001 \) and \( \chi^2 = 6.5, P < 0.01 \) respectively, compared to group A-like and ang-2 considered alone). These results suggest that the group A-like var expression and ang-2 are independently associated with IC.

Next we examined the relationship between ang-2 levels and carriage at the time of disease of antibodies to the IE surface. Previously [8,10] we measured levels of these antibodies against 8 parasite isolates by flow cytometry and the median value of this IE surface recognition by IgG was calculated (henceforth called “IE surface antibodies”). IE surface antibodies showed a negative association with group A-like expression and IC [10]. We asked whether this negative association with IC might involve a causal network that includes ang-2. This would be expected if, for example, IE surface antibodies, by inhibiting cytoadherence mediated by group A-like PfEMP1 protect against endothelial activation and subsequently IC. We therefore tested the independence of IE antibodies, ang-2 and IC using logistic regression. As illustrated in Figure 3A and Table 1 model 7A, adjusting for IE surface antibodies reduced the estimated relationship between group A-like and IC, but showed no effect on the relationship between ang-2 and IC (Figure 3B). We did not alter the relationship between IE surface antibodies and IC (Figure 3C). Thus unlike group A-like var expression, the relationship between IE antibodies and IC appears to be independent of ang-2. Overall, a similar observation was made when CM (bcs \( \leq 2 \)) was considered as the outcome (Table 1 model 1B, 6B, 7B and 8B).

Overall, the independence between group A-like var expression and IE antibodies on the one hand and ang-2 on the other, in relation to their associations with IC suggest that distinct pathways may link group-A like var expression and widespread endothelial activation to IC.
Previous analysis of this dataset suggested that the association between group A-like var expression and IC is independent of peripheral parasite density [10]. We explored this further with another plasma marker, PfHRP2 (thought to be a marker of parasite burden within the host [40]). PfHRP2 has recently been found to have a strong relationship with retinopathy positive cerebral malaria [41]. To test the effect of PfHRP2 on the relationship between group A-like expression and IC, we measured plasma PfHRP2 and tested its relation with group A-like expression and ang-2 using Spearman’s rank correlation. Consistent with the observation made with the peripheral parasitemia (Figure 3D), group A-like var expression showed no evidence for an association with PfHRP2 (Figure 3E). Furthermore, both group A-like expression and PfHRP2 showed independent associations with IC in an age-adjusted logistic regression analysis (Table 1 model 10). In contrast PfHRP2 and ang-2 showed evidence for non-independence in their associations with IC (Table 1, model 12). These results suggest that group A-like expression and ang-2 have contrasting relationships with parasite load and the latter seems to be more important for ang-2 release.

**Associations between Rosette frequency, parasitemia, endothelial activation, and respiratory distress**

Previously, we found that RD showed evidence for an association with peripheral parasitaemia and rosetting frequency within this dataset [10]. Above, we showed ang-2
level is associated with RD (Figure 1B and Table 2), but that appeared not to be a direct relationship between rosetting and ang-2 within the non-severe cases (Figure 2F) suggesting a lack of direct causal relationship. To further explore whether the relationship between rosetting and RD involves endothelial activation measured by plasma ang-2, we used rosette frequency and ang-2 as explanatory variables in age-adjusted logistic regression models predicting RD. We then tested the independence of rosetting frequency and ang-2 in their associations with RD. When ang-2 and rosette frequency were included together in the analysis the OR of the relationship between rosetting and RD dropped by almost a half and the association was no longer statistically significant (Figure 4A and Table 2 model 7). In contrast ang-2 retained its significant associations with RD (Figure 4B, Table 2 model 7). Adding rosetting into a model that considered ang-2 alone showed no evidence of improving the fit ($\chi^2 = 2.2$, $P = 0.14$). Taken together these and the results shown in Figure 2F do not support a strong causal link between rosetting and endothelial activation, though Figure 4A is consistent with some level of non-independence. This is potentially well explained by the association between both endothelial activation and rosetting with parasite density (Figure 4C-F) and the non-independent associations of rosetting frequency with RD when adjusted for ang-2 and parasitaemia in age-adjusted logistic regression analysis (Table 2 models 12 and 13).

**Discussion**

In this study we tested whether a marker of widespread endothelial activation, plasma ang-2 levels throw light on our previously reported associations between expression of group A-like var genes and malaria with IC on the one hand and rosetting frequency with RD on the other [10].

Previously, a link between in vivo endothelial dysfunction and severe malaria has been established [21]. In addition, a further study in Malawi revealed that plasma ang-2 is higher in children with retinopathy positive cerebral malaria [25]. As retinopathy is a surrogate marker of cerebral sequestration [5,26,42], this supports the idea that parasite...
sequestration causes endothelial activation. The recent identification of endothelial protein C receptor (EPCR) as a receptor for severe disease-associated PfEMP1 and the hypothesis that this interaction may drive inflammation [27,28] gives further support to this notion. Following these observations, we explored whether group A-like PfEMP1 subsets (i.e. DC8 and DC13) in P. falciparum isolates sampled from children with severe and non-severe malaria have been shown to be associated with severe malaria [9]. Therefore, in the presence of low host IE surface antibodies, the expression of group A-like PfEMP1 (such as DC13) may give the parasite growth advantage in the initial infection, dominating the sequestered biomass, and causing impaired consciousness before widespread endothelial activation and inflammation occurs.

Our converse observation was that the relationship between ang-2 and IC is not altered by adjustment for group A-like var expression (Figure 3 and Table 1). This suggests that parasites expressing group A-like PfEMP1 do not form part of an explanatory link between ang-2 and IC. This in turn suggests that endothelial activation does not confer a growth advantage to parasites expressing group A-like PfEMP-1 through cytoadherence to inducible adhesives molecules such as ICAM-1. ICAM-1 mediated IE cytoadhesion is known to be mediated by a

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**Table 1 Age-adjusted logistic regression models predicting (A) impaired consciousness and (B) cerebral malaria (N = 213)**

| Models | A: impaired consciousness | B: cerebral malaria |
|--------|---------------------------|---------------------|
|        | Explanatory variables     | OR (95% CI)         | OR (95% CI)         | P value | P value |
| 1      | Group A-like              | 4.5 (1.7, 11.4)     | 2.97 (1.4, 7.76)    | 0.026   |
| 2      | ang-2                     | 3.0 (1.9, 4.7)      | 2.52 (0.57, 4.03)   | 0.0012  |
| 3 $    | Parasite density (peripheral) | 1.37 (1.12, 1.67)  | 1.46 (1.16, 1.84)   | 0.001   |
| 4      | PfHRP2                    | 1.34 (1.13, 1.60)   | 1.43 (1.17, 1.74)   | 0.0004  |
| 5      | IE surface antibodies     | 0.96 (0.94, 0.98)   | 0.96 (0.94, 0.99)   | 0.001   |
| 6      | Group A-like              | 3.5 (1.32, 9.44)    | 2.38 (1.48, 3.82)   | 0.0003+ |
| 7 $    | ang-2                     | 2.82 (1.78, 4.46)   | 2.38 (1.48, 3.82)   | 0.0003+ |
| 8      | Group A-like              | 2.3 (0.86, 6.3)     | 2.35 (1.43, 3.85)   | 0.001+  |
| 9 $    | IE surface antibodies     | 0.96 (0.94, 0.98)   | 0.97 (0.95, 0.99)   | 0.003+  |
| 10     | ang-2                     | 2.67 (1.76, 4.66)   | 2.35 (1.43, 3.85)   | 0.001+  |
| 11     | IE surface antibodies     | 0.96 (0.94, 0.98)   | 0.97 (0.95, 0.99)   | 0.003+  |
| 12     | Group A-like              | 4.15 (1.6, 10.7)    | 2.65 (0.99, 7.08)   | 0.05    |
| 13     | Parasite density (peripheral) | 1.3 (1.1, 1.65)    | 1.43 (1.14, 1.81)   | 0.002+  |
| 14     | Group A-like              | 4.24 (1.6, 11.21)   | 2.68 (0.98, 7.37)   | 0.05    |
| 15     | PfHRP2                    | 1.32 (1.12, 1.57)   | 1.40 (1.15, 1.71)   | 0.001+  |
| 16     | ang-2                     | 2.7 (1.7, 4.3)      | 2.17 (1.33, 3.53)   | 0.002+  |
| 17     | Parasite density (peripheral) | 1.21 (0.97, 1.5)   | 1.32 (1.04, 1.68)   | 0.023+  |
| 18     | ang-2                     | 2.6 (1.60, 4.24)    | 2.07 (1.24, 3.44)   | 0.005+  |
| 19     | PfHRP2                    | 1.18 (0.98, 1.41)   | 1.28 (1.04, 1.58)   | 0.019+  |

The analysis in this table considers the outcomes IC and CM as factors, which means non-IC/CM severe cases are included in the control group and hence the analysis is conservative. The association of group A-like with impaired consciousness is independent of ang-2 (model 6). Similarly the IE surface antibodies and ang-2 show independent association with impaired consciousness (Model 8A, compare with model 7A). In Model 6B, the association of group A-like expression with CM is not statistically significant when ang-2 was considered. However, the odds ratios of both the unadjusted and ang-2 adjusted are similar in magnitude and direction. $ indicate models that have been published in [8,10] and repeated here for comparison. † = variables that improved the fit of the model using LR χ² improvement test. PfHRP2 data was obtained for 207 out of the 213 samples.
Table 2 Age-adjusted logistic regression models predicting respiratory distress (N = 130)

| Models | Explanatory variables | Respiratory distress OR (95% CI) | P |
|--------|-----------------------|-------------------------------|---|
| 1 $    | Rosette frequency     | 5.7(1.6, 20.6)               | 0.008 |
| 2 $    | Group A-like          | 1.76(0.49, 6.34)             | 0.39 |
| 3      | ang-2                 | 4.1(2.0, 8.55)               | 0.0001 |
| 4 $    | parasite density (peripheral) | 2.59(1.56, 4.3) | 0.0002 |
| 5      | PfHRP2                | 1.58(1.18, 2.11)             | 0.002 |
| 6 $    | IE surface antibodies | 0.98(0.96, 1.0)             | 0.1 |
| 7      | Rosette frequency     | 3.0(0.7, 12.9)              | 0.14 |
| 8 $    | Rosette frequency     | 2.0(0.51, 8.3)              | 0.3 |
| 9      | Rosette frequency     | 3.54(0.93-13.42)            | 0.06 |
| 10     | ang-2                 | 3.2(1.5, 6.99)              | 0.003‡ |
| 11     | parasite density (peripheral) | 2.17(1.30, 3.62) | 0.003‡ |
| 12     | Rosette frequency     | 1.25(0.26, 6.0)             | 0.78 |
| 13     | Rosette frequency     | 3.1(1.4, 6.9)               | 0.005‡ |
| 14     | PfHRP2                | 2.18(0.49, 9.70)            | 0.31 |
| 15     | PfHRP2                | 1.34(0.08, 1.83)            | 0.07 |
| 16     | ang-2                 | 3.0(1.38, 6.56)             | 0.005‡ |

The analysis in this table is similar to that in Table 1 except the outcome of interest is respiratory distress. Ang-2 seems to be in the causal pathway linking rosetting frequency to respiratory distress (models 7, 12 and 13). $ variables that improved the fit of the model using LR $χ^2$ improvement test. The analysis in this table is based on 130 samples with rosette frequency data. PfHRP2 data was obtained for 126 out of the 130 samples. $ indicate models published in [16] and repeated here for comparison.

It is important to note that that sequestration per se does not need to play a direct role in endothelial activation for it to be important in disease pathology. Since all IE are thought to sequester, high parasite burden, whatever the underlying cause could lead to endothelial activation independent of the PfEMP1 cytoadhesive profile. This is consistent with the findings of two in vitro studies that used human brain microvascular endothelial cell lines (HBMEC [48,49] and hCMEC/D3 [50] to test whether cytoadherence to ECs is important for endothelial activation. Although they differ in their conclusion on what may be responsible for EC activation, they both agree that binding of IE to the ECs is not necessary for endothelial activation. The study by zougbede et al. further demonstrate that contact between the IE and ECs is not required for endothelial activation [50]. Again in a post-mortem study, Silamut et al. [4] observed, generalised endothelial activation, not confined to sites of parasite sequestration in brain samples from Thai and Vietnamese adults who died from severe malaria, further supporting the possibility of endothelial activation occurring independently of IE binding [4]. Moreover, sera from patients with falciparum malaria were found to induce increased expression of substance P, an observation that was not made with sera from healthy controls [51]. These results raise the possibility that widespread endothelial activation is independent of direct cytoadherence of the IE to the vascular endothelia and that parasite derived circulating factors may be responsible for endothelial activation.

Clearly, parasite density (in the context of falciparum malaria) is likely to be an important determinant of endothelial activation. Parasite density may contribute to endothelial activation by increasing the amount of released microparticles, free haemoglobin, and other molecules. Recently, levels of plasma PfHRP2 (thought to represent parasite load [40,52], though found not useful as a marker of parasite load in a study in Kilifi [53]), was found to differentiate between retinopathy positive CM from retinopathy negative CM [41]. Considering this, the finding that ang-2 is higher in retinopathy positive CM cases compared to retinopathy negative cases [25] is consistent with parasite load being an important determinant of endothelial activation. Moreover, previous work suggests that high parasite burden would lead to an increase in lactic acid production [54], causing acidosis and RD. In this context we found ang-2 to be significantly correlated with parasite density (Figure 4E-F) and base-excess (Additional file 1: Figure S2). Parasite density was shown to be associated with respiratory distress (Table 2) which is a manifestation of metabolic acidosis [30], measured by base excess.

Our var gene expression measurements and assessment of rosetting phenotype come from circulating parasites rather than sequestered ones. Since sequestration and microvascular obstruction occur in multiple small areas of the microvasculature and not in a uniform manner [4,55,56], we cannot rule out the possibility of group A-like PfEMP1 involvement in localised EC activation in the brain micro-vascular network of the brain.
vasculature which is not reflected in the measured plasma ang-2 levels. The latter possibility has been suggested by a recently published study that found fibrin deposition occurring more commonly in CM cases than fatal encephalopathic controls that is restricted to microvessels with sequestered IE [27].

**Conclusion**

Further studies are needed to determine whether systemic endothelial activation observed in malaria is causally related to cytoadherence or invoked by circulating factors such as microparticles and other IE derived toxins. In future studies it will be important to explore the role of cytoadhesion to endothelial protein C receptor and use var expression assays that more directly measure the levels of domain cassettes known to bind to this receptor.

**Additional file**

Additional file 1: Figure S2. Patients characteristics. Figure S2. The relationship between ang-2 and base-excess.
Abbreviations
PEMP1: Plasmodium falciparum erythrocyte membrane protein one; IC: Impaired consciousness; RD: Respiratory distress; SMA: Severe malarial anaemia; IE: Infected erythrocyte; CM: Cerebral malaria; EC: Endothelial cell; HBMEC: Human brain microvascular endothelial cell.

Competing interests
The authors declare that they have no competing interest.

Authors’ contributions
PCB, KM: designed the study and provided overall study supervision. GMW, JNNM, MO, and PCB: processed samples, performed var sequencing, rssetting and IE surface antibody measurements. AA, EK, and MM performed the plasma Ang-2 ELISA: AA, GF and PCB performed the statistical analysis. AA and PCB wrote the manuscript with input from all authors. All authors read and approved the final manuscript.

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