An overview of the multi-pronged approach in the diagnosis of Alport syndrome for 22 children in Northeast China

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Abstract

Background: Alport syndrome (AS) is a kind of progressive hereditary nephritis induced by mutations of different genes that encode collagen IV. The affected individuals usually develop hematuria during childhood, accompanying with gradual deterioration of renal functions. In this study, the multi-pronged approach was employed to improve the diagnosis of AS.

Methods: Twenty-two children were diagnosed and treated at the Department of Pediatric Nephrology of Jilin University First Hospital between January 2017 and January 2020 using the multi-pronged approach. The following information was collected from patients, including age of onset, age at diagnosis, clinical manifestations, family history, renal pathology and genotype.

Results: All these 22 children were diagnosed with Alport syndrome according to the diagnostic criteria formulated by the Japanese Society of Nephrology (2015), among them, only 13 children met the diagnostic criteria released in 1988. All the 22 patients presented with hematuria, and proteinuria to varying degrees was observed in some patients. Three children suffered from hearing loss, but no child in the cohort had any visual problem or renal failure. Meanwhile, five patients were estimated to be at Stage 2, whereas the remaining 17 cases were at Stage 0. Renal biopsies were performed in 18 patients, including 14 showing glomerular basement membranes (GBM)-specific abnormalities. Moreover, 13 children were detected with mutations of genes encoding collagen IV.

Conclusions: The multi-pronged approach helps to improve the diagnosis of AS. Most patients do not have renal failure during childhood, but close assessment and monitoring are necessary. Also, the advancements in treatment are reviewed.

Keywords: Alport syndrome, Children, Multi-pronged approach for diagnosis, Therapy

Background

Alport syndrome (AS) is a hereditary disease related to type IV collagen, which usually results in progressive renal fibrosis and end-stage renal disease (ESRD) [1]. AS arises from mutations in genes COL4A3, COL4A4, and COL4A5. Typically, the X-linked AS (XL-AS), which is induced by mutations in the gene encoding the α5 chain of COL4 (COL4A5) on chromosome X, is the most common. According to previous study, the frequencies of XL-AS, autosomal recessive AS (ARAS), and autosomal dominant AS (ADAS) are estimated to be 80–85, 15%, and 1–5%, respectively [2]. Recent reports suggest that, around 60% AS patients belong to the XL-AS, while...
ARAS accounts for about 15% and ADAS occupies 25% [3, 4]. However, many patients harboring heterozygous mutations of COL4A3 or COL4A4 gene remain undiagnosed due to the subclinical course of disease and incomplete penetrance, as a result, it is difficult to determine the accurate prevalence [5].

In addition to hematuria and progressive renal failure, the affected patients also frequently suffer from extrarenal illnesses involving ears (sensorineural deafness) and eyes (peri-macular flecks and lenticous) [6]. With the advancements in medical technology, apart from electron microscopy [7], other investigation modes, such as immunohistochemical (IHC) analysis on basement membrane type IV collagen expression through skin or renal biopsy and genetic testing [8], have broadened the clinical and research repertoire to detect the changes in AS. Typically, the commonly used diagnostic criteria include family history (FH) of hematuria, sensorineural hearing loss, characteristic eye signs, diffuse esophageal leiomyomatosis, ultrastructural changes and abnormal distributions of the α(IV) collagen chains detected by IHC staining of glomerular basement membranes (GBM), and mutations of COL4A3, COL4A4 or COL4A5 gene [9]. Nonetheless, some clinicians can still not detect this disease due to insufficient assessment or atypical presentations. At the same time, not all patients can afford the fees of all examinations. This study aimed to review the problem that, some cases were missed due to the excessively stringent criteria for AS diagnosis. To address this problem, this study tested the hypothesis that implementing the more relaxed criteria similar to those defined by the Japanese Society of Nephrology improved the diagnosis of AS cases among our patients. In addition, the risk of progression was estimated and the progress in treatment was reviewed.

Methods
Diagnostic criteria
In order to improve the diagnosis of AS, the criteria established by the Working Group for Alport Syndrome in the Japanese Society of Pediatric Nephrology (JSPN) in 2015 was adopted (Table 1) [10]. Meanwhile, the diagnostic criteria formulated in 1988 were also listed [11].

Risk evaluation criteria
In this study, three criteria (Tables 2, 3 and 4) [6, 12, 13] were adopted to estimate the risk of renal progression, including clinical estimate and genotype-phenotype correlation in XL-AS.

Clinical investigation
Clinical data were collected from our patients, including the age of onset, age at diagnosis, duration from onset of symptoms to diagnosis, hematuria, proteinuria, estimated glomerular filtration rate (eGFR), extrarenal symptoms and FH. Moreover, the renal histopathological findings obtained by light microscopy (LM) and electron microscopy (EM), immunofluorescence staining and IHC staining of type IV collagen were also extracted for subsequent analysis. Gene data were obtained from some children with highly suspected AS.

Statistical analysis
The SPSS 20.0 statistical software was employed for data processing. The abnormally distributed data were

| Table 1 | JSPN diagnostic criteria\(^a\) [10] and criteria described in 1988\(^b\) [11] |
|-----------------|---------------------------------|
| Diagnostic features of Alport syndrome |
| I. Primary feature: |
| I-1. Persistent hematuria |
| II. Secondary features: |
| II-1. Mutations in type IV collagen genes |
| II-2. Type IV collagen abnormal expression |
| II-3. Glomerular basement membrane (GBM) -specific abnormalities |
| III. Accessory features |
| III-1. Family history of kidney diseases |
| III-2. Bilateral sensorineural deafness |
| III-3. Ocular abnormalities |
| III-4. Diffuse leiomyomatosis |

\(^a\)Diagnostic criteria: In addition to the primary feature, patients should satisfy one or more secondary features or satisfy two or more of the accessory features.

\(^b\)If patients only have the primary feature and a family member diagnosed with Alport syndrome, the case is set as a "suspected case". If patients have any one feature of type IV collagen (II-1 or II-2) among the secondary features, the case is set as "asymptomatic carriers". If features caused by other diseases should be excluded, for example, a family history of kidney failure due to diabetes.

1. Positive FH of macroscopic hematuria or chronic renal failure
2. Electron microscopic evidence of AS on renal biopsy
3. Characteristic ophthalmic signs (anterior lenticonus and macular flecks)
4. High-tone sensorineural deafness
expressed as medians (range), whereas the enumeration data were expressed as percentages (%).

Results
Altogether 22 children were diagnosed with AS from January 2017 to January 2020 at the Department of Pediatric Nephrology of Jilin University First Hospital. All these patients met the diagnostic criteria, including the primary feature and at least one of the secondary features (Table 5, Table 6). The clinical characteristics, renal pathological characteristics and gene mutations of our patients were elaborated as follows. Despite of gene detection results, only 13 children were diagnosed with AS according to the 1988 criteria (Table 6).

Clinical characteristics
The age at diagnosis among the 22 patients (including 12 boys and 10 girls) ranged from 34 to 170 (median, 84) months. Meanwhile, the duration between symptom onset and diagnosis varied from 1 to 97 (median, 5.5) months. All children (100%) had hematuria with dysmorphic red cells, among them, 18 (81.8%) had paroxysmal macroscopic hematuria in the process of upper respiratory infection. Proteinuria within the non-nephrotic range was presented in ten children (45.5%), with the proteinuria levels of less than 30 mg albumin per g creatinine or per day (Stage 0). In addition, five children (22.7%) had nephrotic range proteinuria (P2, P9, P15, P19 and P20) and were estimated to be at Stage 2. Upon diagnosis, three children (13.6%) were confirmed to have mild to moderate sensorineural hearing loss (P2, P9 and P19), even though the eGFR and vision of all children were within the normal ranges. Moreover, positive FH was identified in 16 patients (72.7%). Table 5 depicts the full details of the clinical findings.

Renal pathological characteristics
Renal biopsies were performed in 18 children, among them, 16 (88.9%) showed minor glomerular abnormalities (MGA) through LM, while one had focal segmental glomerulosclerosis (FSGS) and one had mesangial proliferative glomerulonephritis (MsPGN), respectively. Immunofluorescence staining was negative in 6 children (33.3%), while the remaining presented with non-specific

| Table 2 | Stages in the development of Alport syndrome [6] |
|---------|-------------------------------------------------|
| Stage 0 | microscopic hematuria (< 30 mg albumin per g creatinine or per day) |
| Stage 1 | microalbuminuria (30–300 mg albumin per g creatinine or per day) |
| Stage 2 | gross proteinuria (> 300 mg albumin per g creatinine or per day) |
| Stage 3 | impaired renal function (GFR < 60 ml/min/1.73m²) |
| Stage 4 | end-stage renal disease |

GFR glomerular filtration rate

| Table 3 | New classification system for Alport syndrome and related disorders [12] |
|---------|---------------------------------------------------------------------|
| Inheritance | Affected gene(s) | Genetic state | Comments | Estimated risk of ESRD |
|-----------|-----------------|---------------|----------|-----------------------|
| X-linked  | COL4A5          | Hemizygous (male subjects) | Rate of progression to ESRD and timing of extrarenal manifestations strongly influenced by genotype | 100% |
|           |                 | Heterozygous (female subjects) | Risk factors for progression: gross hematuria, SNHL, proteinuria, GBM thickening and lamellation | Up to 25% |
| Autosomal | COL4A3 or COL4A4 | Recessive (homozygous or compound heterozygous) | Rate of progression to ESRD and timing of extrarenal manifestations strongly influenced by genotype | 100% |
|           |                 | Dominant | | |
|           |                 | Hematuria Includes patients previously diagnosed as TBMN/ BFH Risk factors for progression: proteinuria, FSGS, GBM thickening and lamellation, SNHL, or evidence of progression in patient or family, genetic modifiers | 20% or more among those with risk factors for progression, < 1% in absence of risk factors |
| Digenic   | COL4A3, COL4A4, and COL4A5 | COL4A3 and COL4A4 mutations in trans and COL4A3 and COL4A4 mutations in cis | Clinical findings and pedigree simulate autosomal recessive transmission | Up to 100% |
|           |                 | Clinical findings and pedigree simulate autosomal dominant transmission | | Up to 20% |
|           |                 | Mutations in COL4A5 and in COL4A3 or COL4A4 | Inheritance pattern does not simulate any Mendelian transmission | Up to 100% (affected male subjects) |

BFH benign familial hematuria, ESRD end-stage renal disease, FSGS focal segmental glomerulosclerosis, GBM glomerular basement membrane, SNHL sensorineural hearing loss, TBMN thin basement membrane nephropathy
deposition of immune complex. Further, GBM-specific abnormalities were detected in 14 cases (77.8%) by EM. Two cases showed atypical results, with extensive thinning of GBM in P8 and irregular thinning of GBM in P13, respectively. Additionally, the expression levels of type IV collagen α2 and α5 chains were tested in eleven children. According to our results, all patients showed normal positive staining of GBM and tubular basement membranes for type IV collagen α2 chain. Three children (27.3%) showed abnormal expression of type IV collagen, of them, P18 and P19 (male) presented with negative staining of α5 (IV), while P21 (female) showed discontinuous α5 chain, and the remaining 8 (72.7%) exhibited intact staining for α5 chain (Table 5).

Gene detection
Thirteen children and their parents were tested by the high throughput-targeted next generation sequencing (NGS) technologies at the Beijing Zhiyin Oriental Transforming Medical Research Center Co., Ltd, Beijing Jinzhun Gene Science or Centre of Genetic Diagnosis of Jilin University First Hospital. Whole exome sequencing (WES) was accepted in most patients, while “Panel” was adopted in some patients. The protein conservation was validated by the UGENE software and shown in Fig. 1a-c. Meanwhile, the pathogenicity was predicted by the online Polyphen2 and SIFT software.

Table 7 depicts the detailed mutations of genes encoding type IV collagen. Among them, ten mutations (76.9%) were inherited in the X linked manner (8 from maternal side, 1 from paternal side, 1 of indeterminacy). Additionally, five boys harbored the COL4A5 missense mutation, including one (P2) with premature stop (Type S), three (P1, P9 and P14) with glycine-X-Y substitutions involving exons 21–47 (Type MS), and one (P15) with glycine-X-Y substitutions involving exons 1–20 (Type M). In the meantime, five girls had COL4A5 mutation, including one (P4) with non-glycine-X-Y missense mutation (Type MS), two (P6 and P11) with glycine-X-Y substitutions involving exons 21–47 (Type MS), one (P10) with glycine-X-Y substitutions involving exons 1–20 (Type M), and one (P12) with compound heterozygous mutations of COL4A5, where one sequence variant led to premature stop (Type S). Besides, P3 was identified with compound heterozygous mutations of COL4A4, P8 with autosomal dominant mutation of COL4A3, and P13 with both sequence variants of COL4A3 and COL4A5.

**Discussion**
It has been well documented that, mutations of genes encoding type IV collagen leads to AS. Studies investigating the correlations of mutations with genotype and phenotype have been under way in China [14, 15]. However, as a developing country, there are still many economically less-developed regions in China, especially in Northeast China. Due to economic reasons, WES can not be extensively accepted in all families, while renal biopsy is not widely accepted for conservative idea as well. Given the above reasons, many patients can not get adequate assessment. Without sufficient evidence, the diagnosis of AS can be ambiguous and the treatment may be delayed. Therefore, the Japanese diagnostic criteria [10] were applied in this study to diagnose this disease by the multi-pronged approach, hoping to reduce the misdiagnosis rate and improper treatment, estimate the risk of progressive renal disease, provide timely intervention, and minimize the economic costs.

Using the JSPN diagnostic criteria, 22 children were diagnosed with AS. In addition to the primary feature, patients should satisfy at least one secondary feature or at least two accessory features. All patients (100%) in our cohort generally presented with persistent hematuria, which was identified as the primary feature in the criteria, and proteinuria was also detected in some patients. This was consistent with published articles [16]. As mentioned above, not all patients received NGS or renal biopsy. Only five patients (P3, P6, P10, P14, P15) received both NGS and renal biopsy. Only five patients (P3, P6, P10, P14, P15) received both NGS and renal biopsy, which showed positive results. In addition, six patients with positive FH, including four (P1, P9, P11 and P12) who did not receive renal biopsy and two (P2, P4) who refused to perform renal histopathology in EM, were confirmed with COL4A5 variants by NGS. Besides, P8 showed diffuse thinning of GBM, whereas P13 showed irregular thinning of GBM. Although they were not typical in EM, they were confirmed by WES. In the meantime, they also had positive FH. The remaining nine patients were

| Genotype | Phenotype |
|----------|-----------|
| Type S (Severe) | large rearrangements, premature stop, frameshift, donor splice site mutations, and mutations involving the NC 1-domain, 15% de novo mutations |
| Type M5 (Moderate-severe) | non-glycine-X-Y missense, glycine-X-Y involving exons 21–47, in-frame and acceptor splice site mutations, 15% de novo mutations (5% de novo glycine-X-Y mutations) |
| Type M (Moderate) | glycine-XY mutations involving exons 1–20, 5% de novo mutations |

Table 4 Genotype-phenotype correlation in XL-AS [13]
| Patient ID | Gender | Age at onset/ diagnosis age (months) | Persistent Hematuria | Macroscopic Hematuria | Proteinuria of nephrotic range | Stage | ESRD | Mutations in type IV collagen genes (II-1) | Type IV collagen abnormal expression (II-2) | LM | EM: GBM-specific abnormalities (II-3) | FH (II-1) | Hearing loss (III-2) | Ocular changes (III-3) | Diffuse leiomyomatosis (III-4) |
|------------|--------|------------------------------------|---------------------|---------------------|-------------------------------|-------|------|------------------------------------------|--------------------------------------------|-----|---------------------------|------------|-------------------|-----------------------------|-------------------------------|
| P1         | M      | 38/45                              | +                   | +                   | -                             | 0     | -    | COL4A5                                  | ND                                        | ND | ND                        | +           | -                 | -                          | -                             |
| P2         | M      | 36/132                             | +                   | +                   | +                             | 2     | -    | COL4A5                                  | ND                                        | MGA | ND                        | +           | +                 | -                          | -                             |
| P3         | M      | 108/120                            | +                   | -                   | -                             | 0     | -    | COL4A4/COL4A4                            | intact staining for α5                    | MGA | +                        | -           | -                 | -                          | -                             |
| P4         | F      | 108/109                            | +                   | +                   | -                             | 0     | -    | COL4A5                                  | intact staining for α5                    | MGA | ND                        | +           | -                 | -                          | -                             |
| P5         | M      | 96/97                              | +                   | +                   | -                             | 0     | -    | ND                                       | ND                                        | MGA | +                        | +           | -                 | -                          | -                             |
| P6         | F      | 96/97                              | +                   | +                   | -                             | 0     | -    | COL4A5                                  | intact staining for α5                    | MGA | +                        | +           | -                 | -                          | -                             |
| P7         | F      | 60/61                              | +                   | +                   | -                             | 0     | -    | ND                                       | ND                                        | MGA | +                        | NA          | -                 | -                          | -                             |
| P8         | F      | 48/49                              | +                   | -                   | -                             | 0     | -    | COL4A3                                  | intact staining for α5                    | MGA | A                         | +           | -                 | -                          | -                             |
| P9         | M      | 48/84                              | +                   | +                   | +                             | 2     | -    | COL4A5                                  | ND                                        | ND | ND                        | +           | -                 | -                          | -                             |
| P10        | F      | 60/84                              | +                   | -                   | -                             | 0     | -    | COL4A5                                  | ND                                        | MGA | +                        | -           | -                 | -                          | -                             |
| P11        | F      | 24/60                              | +                   | -                   | -                             | 0     | -    | COL4A5                                  | ND                                        | ND | ND                        | +           | -                 | -                          | -                             |
| P12        | F      | 33/34                              | +                   | +                   | -                             | 0     | -    | COL4A5/COL4A5                            | ND                                        | ND | ND                        | +           | -                 | -                          | -                             |
| P13        | M      | 60/61                              | +                   | +                   | -                             | 0     | -    | COL4A3/COL4A5                            | ND                                        | MGA | A                         | +           | -                 | -                          | -                             |
| P14        | M      | 72/73                              | +                   | +                   | -                             | 0     | -    | COL4A5                                  | intact staining for α5                    | MGA | +                        | -           | -                 | -                          | -                             |
| P15        | M      | 143/170                            | +                   | +                   | +                             | 2     | -    | COL4A5                                  | ND                                        | FSGS | +                         | -           | -                 | -                          | -                             |
| P16        | F      | 107/108                            | +                   | +                   | -                             | 0     | -    | ND                                       | ND                                        | MGA | +                        | -           | -                 | -                          | -                             |
| P17        | F      | 24/73                              | +                   | +                   | -                             | 0     | -    | ND                                       | intact staining for α5                    | MGA | +                        | -           | -                 | -                          | -                             |
| P18        | M      | 36/85                              | +                   | +                   | -                             | 0     | -    | ND                                       | loss of staining for α5                    | MGA | ND                        | +           | -                 | -                          | -                             |
| P19        | M      | 24/121                             | +                   | +                   | +                             | 2     | -    | ND                                       | loss of staining for α5                    | MGA | +                        | +           | -                 | -                          | -                             |
| P20        | M      | 59/61                              | +                   | +                   | +                             | 2     | -    | ND                                       | intact staining for α5                     | MGA | +                        | -           | -                 | -                          | -                             |
| P21        | F      | 59/81                              | +                   | +                   | -                             | 0     | -    | ND                                       | discontinuous for α5                      | MGA | +                        | -           | -                 | -                          | -                             |
| P22        | M      | 118/122                            | +                   | +                   | -                             | 0     | -    | ND                                       | intact staining for α5                    | MGA | +                        | -           | -                 | -                          | -                             |

Total 22 22 18 5 0 13 3 14 16 3 0 0

M male, F female, ESRD end-stage renal disease, LM light microscope, EM electron microscope, MGA minor glomerular abnormalities, FSGS focal segmental glomerulosclerosis, MsPGN mesangial proliferative glomerulonephritis, GBM glomerula basement membrane, FH family history, ND Not do, NA not available, A atypical
confirmed with AS due to the typical GBM abnormalities. According to the criteria, these patients satisfied the primary feature, at least one secondary feature, accompanying with or without at least one accessory feature. Although some of these patients did not receive renal biopsy, while some did not undergo genetic test, the diagnosis was credible. When it came to the original criteria, only 13 of our children were diagnosed with AS. In comparison with the previous general diagnostic criteria, the Japanese criteria improved our diagnosis and covered patients who were easy to be ignored or ambiguous.

Not all of our patients were correctly diagnosed with AS at presentation. The median duration between symptom onset and diagnosis was 5.5 (range, 1–97) months, suggesting that it took nearly half a year to get diagnosed since onset. For patients who were willing to receive necessary examinations, only 1 month was required to
| ID | Gene | Location of chr | Exon | Nucleotide Change | Amino acid Change | Conservation of the protein | Source of variation | ACMG | Mutation type | Inheritance | Type |
|----|------|-----------------|------|------------------|------------------|------------------------|------------------|------|--------------|-------------|------|
| P1 | COL4A5 | chrX:107858210 | exon30 | c.[2466G>A]     | p.(Gly822Glu)    | conserved              | Mo(het)          | VUS | Missense     | XL          | MS   |
| P2 | COL4A5 | chrX:107866028 | exon33 | c.[2890G>T]     | p.(Gly964X)      | conserved              | Mo(het)          | Likely pathogenic | Nonsense   | XL   | S   |
| P3 | COL4A4 | chr2:227875130 | exon46 | c.[4421C>T]     | p.(Thr1474Met)   | conserved              | Mo(het)          | VUS | Missense     | AR          | /    |
|    | COL4A4 | chr2:227896933–227,896,934 | exon39 | c.[3636_3637del] | p.(Arg1212fs)    | conserved              | Fa(het)          | Likely pathogenic | Frame shift | AR   | /    |
| P4 | COL4A5 | chrX:107865935 | exon33 | c.[2797C>T]     | p.(Leu933Phe)    | conserved              | Mo(het)          | VUS | Missense     | XL          | MS   |
| P6 | COL4A5 | chrX:107867547 | exon34 | c.[2999G>T]     | p.(Gly1000Val)   | conserved              | Mo(het)          | Pathogenic     | Missense   | XL   | MS   |
| P8 | COL4A3 | chr2:228159760 | exon40 | c.[3499G>A]     | p.(Gly1167Arg)   | conserved              | Mo(het)          | Pathogenic(PS1 + PM1 + PM2 + PP3 + PP1) | Missense   | AD   | /    |
| P9 | COL4A5 | chrX:107842091 | exon25 | c.[1939G>A]     | p.(Gly647Arg)    | conserved              | Mo(het)          | Likely pathogenic | Missense   | XL   | MS   |
| P10| COL4A5 | chrX:107824232 | exon16 | c.[911G>A]      | p.(Gly304Glu)    | conserved              | Fa(hemi)         | Likely pathogenic (PM1 + PM2 + PM + PP1) | Missense   | XL   | M    |
| P11| COL4A5 | chrX:107842014 | exon25 | c.[1862G>A]     | p.(Gly621Ala)    | conserved              | Mo(het)          | Likely pathogenic (PS + PM1 + PM5) | Missense   | XL   | MS   |
| P12| COL4A5 | chrX:107829932 | exon19 | c.[1120G>T]     | p.(Gly374X)      | conserved              | indeterminacy     | Likely pathogenic | Nonsense   | XL   | S    |
|    | COL4A5 | chrX:107829930 | exon19 | c.[1118G>A]     | p.(Arg373Gln)    | conserved              | indeterminacy     | VUS | Missense     | XL          | MS   |
| P13| COL4A3 | chr2:228,175,529 | exon51 | c.[479T>G]      | p.(Leu1598Arg)   | conserved              | Mo(het)          | Likely pathogenic | Missense   | A    | /    |
|    | COL4A5 | chrX:107815050 | exon8  | c.[448G>C]      | p.(Gly150Arg)    | conserved              | Mo(het)          | VUS | Missense     | XL          | M    |
| P14| COL4A5 | chrX:107900779 | exon39 | c.[3508G>A]     | p.(Gly1170Ser)   | conserved              | Mo(het)          | Likely pathogenic (PS1 + PM2 + PP3 + PP1) | Missense   | XL   | MS   |
| P15| COL4A5 | chrX:107815050 | exon8  | c.[448G>T]      | p.(Gly150Trp)    | conserved              | indeterminacy     | Likely pathogenic (PS2 + PM2 + PP3) | Missense   | XL   | M    |

Mo mother, Fa father, XL X-linked, AR autosomal recessive, AD autosomal dominant, ACMG The American College of Medical Genetics and Genomics, Het heterozygous, hemi hemizygote, VUS uncertain significance
identify the etiology. However, much longer time was needed for more patients, even 4–8 years. In that case, many patients might receive improper treatments with an ambiguous diagnosis.

P2 was initially diagnosed with glomerulonephritis at the age of 3 years in the local clinic due to hematuria and nephrotic range of proteinuria. Subsequent renal biopsy, which was done when he was 9 years old, was compatible with minimal change disease (MCD). Further FH revealed that his mother had persistent hematuria and proteinuria of unknown etiology and that his grandfather died of uremia, his parents refused to undergo further investigation (Fig. 2a). The patient was initially managed with corticosteroids, followed by cyclophosphamide and mycophenolate mofetil, due to steroid resistance. After Tacrolimus treatment, the patient achieved partial remission, with urine protein being controlled at below 1 g per day. The patient was later confirmed with XL-AS (a missense mutation of COL4A5 inherited from his mother) at the age of 11 years by genetic testing (Fig. 2b).

The clinical manifestations of AS can occasionally be confused with other clinical entities. For instance, P4 of our cohort, who had a missense mutation of COL4A5 inherited from her mother (Fig. 3b), presented with macroscopic hematuria during infection, and she did not suffer from any hearing loss or visual problem. The renal biopsy of this patient revealed MGA, along with mild IgA deposition that was compatible with IgA nephropathy. If not for the presence of FH (Fig. 3a) that made her as the “suspected case”, the patient would have been treated as IgA nephropathy, accordingly, genetic testing was not performed. Interestingly, a child with similar clinical manifestations to our patient was misdiagnosed with IgA nephropathy [18]. His renal biopsy did not reveal features of AS until he had a second renal biopsy at 4 years later. Similarly, in another recent Chinese report [19], the proband who presented with hematuria and proteinuria was initially diagnosed with IgAN by renal biopsy and immunofluorescence detection. Because of the poor treatment outcome, the patient was identified with a novel mutation of COL4A5 by the gene detection. By the time a definite diagnosis was made, the patient had been treated with prednisolone accompanied with mycophenolate mofetil and tacrolimus successively.

Different from the typical manifestations, P3 displayed isolated hematuria with negative FH. The patient accidentally discovered microscopic hematuria in a health check. During the eight-month follow-up
period, the urine red blood cell count fluctuated from 10/HPF to 30/HPF. GBM-specific abnormalities were observed in renal biopsy. Besides, WES revealed compound heterozygous mutations of COL4A4. The renal pathology, gene mutation and family pedigree of P3 are shown in Fig. 4a-g.

On the other hand, whether the characteristic changes of GBM can be present depends on multiple factors, such as the age of biopsy and different mutations. In our cohort, 14 out of the 18 renal biopsies revealed GBM-specific abnormalities, while two cases manifested as extensive thinning of GBM (P8) and irregular thinning of GBM (P13), respectively. Thin basement membrane nephropathy (TBMN) is a relatively common disease with the reported incidence rate of 1% in the general population [20]. Mutations in the Col(IV)A3/A4 and Col(IV)A5 coding genes may be responsible for TBMN and AS [21]. However, in children and females, the only evidence of AS may be the thinning of GBM, which can be misdiagnosed with TBMN [22]. According to the newest classification [12], TBMN is currently considered as a lesion description rather than a diagnosis. It was possible that some of our patients previously diagnosed with TBMN actually had AS. P8 had extensive thinning of GBM and would have been diagnosed with TBMN. Different from other cases, the patient got an autosomal dominant mutation of COL4A3. Her mother presented with asymptomatic hematuria, while her grandfather received regular dialysis for 8 years since the diagnosis of uremia. Considering the ominous outcome of AS, we hoped to diagnose her with AS rather than TBMN. The patient will be followed up for a long time and receive a second renal biopsy if necessary. The renal pathology, gene mutation and the family pedigree are shown in Fig. 5a-f. P13 had irregular thinning of GBM and digenic mutations of COL4A3 and
Meanwhile, his mother presented with persistent hematuria and proteinuria of unknown etiology, and his grandfather died of uremia. The detailed information is exhibited in Fig. 6a-g.

Type IV collagen, which is a component of the GBM, is a triple helix composed of three α chains. Specifically, the α3(IV), α4(IV) and α5(IV) chains are present in GBM, Bowman’s capsule and the basement membranes of distal and collecting tubules. In our cohort, P18 and P19 (male) showed negative staining of α5(IV), while P21 (female) showed discontinuous α5 chain, which satisfied the abnormal expression of type IV collagen. For P19, in addition to the manifestation of nephrotic range proteinuria, the patient also had hearing loss when he was diagnosed. Similarly, Samar et al. [23] stated that negative staining of α5 chain was correlated with the worse prognosis and more severe ultrastructural alterations in AS men. Additionally, eight patients had intact staining of α5 chain. The putative causes might include the type and location of sequence variants. Hashimura et al. [24] hypothesized that, some missense and in-frame mutations of XL-AS might affect the structure of this triple helix, but its degradation rate was low. They also suggested that mutations located between exons 1 and 25 might lead to a less critical disruption of triple helix-forming process. This might explain for the positive staining in male patients with XL-AS who had milder clinical manifestations. However, the mechanism of autosomal AS has not been fully understood yet.

In addition to the definite diagnosis, the disease severity was also assessed. Five of our patients manifested with nephrotic range proteinuria and were estimated to be at Stage 2, three had hearing loss, and 14 presented with GBM-specific abnormalities by EM, but none of them showed renal failure. This might because that they were diagnosed within their first or second decades of lifetime, which gave us the time and opportunity to estimate the risk of progression and provide appropriate treatment.

In this study, a total of ten patients had COL4A5 mutation, one had compound heterozygous mutations of COL4A4, one had autosomal dominant (AD) mutation of COL4A3, and one had digenic mutations of COL4A3 and COL4A5. In XL-AS, hemizygous male patients have a 100% risk of progression to ESRD, although the progression rate and timing of extrarenal manifestations are related to the COL4A5 genotype [12]. Heterozygous female patients have a 25% risk of progression to ESRD throughout their lifetime. But this depends on a variety of risk factors, including a history of gross hematuria in childhood, sensorineural hearing loss, proteinuria, and extensive GBM thickening and lamellation [25]. Few Gly substitutions are non-pathogenic. Gly substitutions with a charged residue, such as Arg, Glu or Asp, often result
in the early-onset renal failure and more extrarenal features [26]. However, it is much more difficult to distinguish the pathogenic from the benign variants for non-Gly substitutions [27]. Our patients with COL4A5 mutation were estimated with subtypes ranging from Type MS to Type S, and most of them had risk factors, so renal function should be monitored closely within the next decade.

Autosomal AS associated with biallelic mutations (homozygous or compound heterozygous) in COL4A3 or COL4A4 exhibits a recessive inheritance pattern, which is related to a 100% risk of ESRD, and the rate of progression and timing of extrarenal manifestations are affected by the genotype [12]. P3 in our study had the compound heterozygous mutations of COL4A4 with GBM-specific renal pathology, and he was thus estimated with a 100% risk of ESRD. Patients with heterozygous mutations in COL4A3 or COL4A4 are considered to be affected in the presence of hematuria or proteinuria, including patients who would have been previously diagnosed with TBMN. In these individuals, the risk of ESRD is as high as 20% among those with risk factors for progression, including proteinuria, sensorineural hearing loss, FH of progression to ESRD, and renal biopsy findings of FSGS, or GBM thickening and lamellation, or all of the above. Recent systematic review states that there is a striking difference in the percentage of patients progressing to ESRD [5]. For instance, in a large cohort, many patients are misdiagnosed, since heterozygous COL4A3/COL4A4 mutations (a cause of TBMN) are associated with FSGS [28]. These figures regarding the risk of ESRD are not always solid and they are dependent on different patients and diverse age ranges. In our cohort, P8 who might be diagnosed with TBMN was detected with COL4A3 dominant mutation. Since her mother presented with isolated hematuria without ESRD, it was hopeful to look forward to the benign progression. Also, the risk of digenic inheritance should be further studied. It is reported in literature [29, 30] that, COL4A3/A4 mutations in cis resemble an AD inheritance with a more severe phenotype, while COL4A3/A4 mutations in trans mimicks an autosomal recessive inheritance with a less severe phenotype, and COL4A5 combined with COL4A3 triggers a more severe phenotype. In our cohort, P13 was a bit different. He got glycine-XY substitutions involving exons 1–20 in COL4A5 and irregular thinning of GBM, in addition, he also got a COL4A3 mutation, which contributed to a high risk of renal progression. These children are actively followed up and their renal progression is closely monitored at present.

There is no radical cure for AS for the time being, and attempts to use various stem cell therapies in animal models have attained ambiguous success. The use of cyclosporine, a calcineurin inhibitor, remains controversial due to its possible long-term nephrotoxic effects [17]. With the exception of cyclosporine, the renin-angiotensin-aldosterone system (RAAS) inhibitors are reported to be efficient and well tolerated to delay the progression of chronic kidney disease (CKD) in AS [31]. Gross et al. [32] carried out a double-blind, randomized, placebo-controlled, multicentre phase III trial to clarify the safety and efficacy of ramipril in child patients with AS, and discussed the efficacy of ramipril when the patients present with microhematuria only. So far, the Alport Syndrome Classification Working Group recommends to use ACEI in the presence of hematuria and overt proteinuria [12]. In addition, future therapies, including stem cells, chaperon therapy, collagen receptor blockade and anti-microRNA therapy, will shed more lights on the protection of kidneys in AS patients from further damage [33]. Through different mechanisms, therapies such as Bardoxolone, anti-miRNA-21, paracil-tol, lipid-lowering agents and epidermal growth factor receptor inhibitor, may play a certain role in mitigating renal fibrosis [34]. Meanwhile, chaperone and stem-cell based therapies are expected to show therapeutic efficacy at the collagen chains and GBM level, respectively. Nonetheless, when renal failure is inevitable, AS patients who undergo renal transplantation will have generally excellent outcomes [35]. Despite the prominent genotype-phenotype correlation, severe mutations do not impact the survival of patient or graft after transplantation [36].

Conclusions
To sum up, given the importance of early diagnosis and economic factors, the multi-pronged approach is adopted in this study to diagnose AS and estimate the risk of progression. In condition-limited settings, it is important to follow a pragmatic approach. In addition, the Japanese criteria do improve our diagnosis. RAAS inhibitors have been testified to show safety and efficacy in delaying renal progression. Patients receiving renal transplantation have excellent outcomes, along with favorable graft survival rates. Future therapies are on the way to change the “inevitable” outcome of the disease.

Abbreviations
AS: Alport syndrome; FH: Family history; GBM: Glomerular basement membrane; ESRD: End-stage renal disease; ARAS: Autosomal recessive Alport syndrome; ADAS: Autosomal dominant Alport syndrome; XL-AS: X-linked Alport syndrome; eGFR: Estimated glomerular filtration rate; IM: light microscopy; EM: Electron microscopy; MGA: Minor glomerular abnormalities; FSGS: Focal segmental glomerulosclerosis; MxPGN: Mesangial proliferative glomerulonephritis; NGS: Next generation sequencing; WES: Whole exome sequencing; TBMN: Thin basement membrane nephropathy; MCD: Minimal change disease; RAAS: Renin-Angiotensin-Aldosterone System; CKD: Chronic kidney disease
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Authors’ contributions

ZL collected the clinical and gene information and wrote the manuscript. SBC contributed to data collection and interpretation. ZBG were in charge of Ethics approval and consent to participate. MQS was responsible for critically revising the manuscript for important intellectual contents. All authors read and approved the final manuscript.

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Availability of data and materials

The datasets used and analyzed in the present study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

This study was approved by the Ethics Committee of Jilin University First Hospital. All mentioned data were accessed with the approval of the Hospital.

Consent for publication

Written informed consents were provided by the parents of patients for the publication of case history and clinical results. A copy of the written consent was available for the review by the editor of this journal.

Competing interests

The authors declare that they have no competing interests.

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