Research Article

Ginsenoside Rb1 Preconditioning Enhances eNOS Expression and Attenuates Myocardial Ischemia/Reperfusion Injury in Diabetic Rats

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1. Introduction

Diabetes mellitus has long been identified as the leading cause of the development of ischemic heart disease within the diabetic population [1]. Many epidemiological findings unanimously support the fact that cardiovascular diseases (CVD) are the primary mortality factor among diabetic patients [2]. The diabetic state is associated with increased oxidative stress, and the hyperglycemia further stimulates the production of advanced glycosylated end products, which increase microvascular permeability. Moreover, the abnormal lipid metabolism increases oxidized low-density lipoprotein (ox-LDL) formation, resulting in oxidative stress which exacerbates myocardial and vascular endothelial cell insult due to myocardial ischemia/reperfusion injury. Hearts from diabetic subjects are less resistant to ischemic insults [3, 4]. We have recently demonstrated that postischemic myocardial infarct sizes were significantly larger in diabetic rats 8 weeks after STZ-injection as compared to non-diabetic controls [5], and we have found that Rb1 can confer cardioprotection in diabetic rats. However, the mechanism by which Rb1 confers cardioprotection in diabetes has not been explored. Therefore, in the current study, we explored whether or not enhancing nitric oxide bioavailability plays a critical role in Rb1-mediated cardioprotection in diabetic subjects.

Nitric oxide (NO) is a fundamental endothelium-derived relaxing factor (EDRF), which at physiological concentration possesses vasodilatory properties inhibits leukocyte and platelet adherence, suppresses smooth muscle cell hyperplasia [6], and is also beneficial in limiting myocardial ischemia/reperfusion (MI/R) injury. Numerous studies have already demonstrated the cardioprotective effects of NO during ischemia/reperfusion events [7, 8]. However, in patients with long-standing diabetes, the availability of NO decreases progressively [9]; in addition, eNOS expression is reduced as well as mediators for the synthesis of NO. Hence, such situations aggravate vascular endothelial dysfunction and
atherosclerosis in diabetes [10, 11]. In animal studies, it has been found that eNOS provides protection to some extent against I/R injury to extracorporeal hearts [12], and bioavailability of NO and availability of eNOS decrease I/R injury and promote reperfusion of myocardial infarction and after-heart failure in left ventricular function recovery [13].

Ginsenoside Rb1 is the main bioactive component of Ginseng and Shenfu injection. It has various beneficial effects on the cardiovascular system, such as scavenging free radicals, blocking calcium overinflux into neurons, inhibiting Na⁺ channel activities, improving energy metabolism, and preserving the structural integrity of the neurons, among many others [14]. Recent studies have demonstrated that Ginsenoside Rb1 has protective effects against myocardial ischemic injury in nondiabetic animals [15] and the molecular mechanism of this beneficial effect is still unidentified.

The present study aims to determine whether Ginsenoside Rb1 preconditioning can protect against I/R injury in diabetic rats by enhancing the expression of eNOS and increasing the content of NO as well as inhibiting oxidative stress.

2. Materials and Methods

2.1. Experimental Animals. The experimental procedures and protocols used in this research work were approved by the Animal Use Committee of Wuhan University. Healthy male adult Sprague-Dawley (SD) rats (weighing 220–280 g), procured from the Experimental Animal Centre of Wuhan University. After one week of adaption period, they were kept fasting for 12 hr and injected with streptozotocin (STZ; Sigma, USA; pH 4.5) intraperitoneally at a dosage of 65 mg/kg. Five days after STZ injection, the rats were kept fasting for 5 hr, and the blood samples from the tails were collected and fasting blood sugar was measured using SureStrep glucometer (Johnson & Johnson Company). Rats with blood glucose level ≥16.7 mmol/L (300 mg/L) were considered diabetic models [16].

2.2. Myocardial Ischemia: Reperfusion Model. After a 12 hour period of fasting, the animals were anesthetized with 3% pentobarbital sodium injection (50 mg/kg) administered intraperitoneally and then fixed on the operating table for surgical procedures. ECG electrodes were placed subcutaneously in both front limbs and left back limb and lead II was continuously monitored. A 14-gauge angiocatheter was inserted into the trachea through a midline neck incision and then connected to a volume-controlled ventilator (DW-2000, Jiapeng Keji, Shanghai, China). The left femoral vein was catheterized for intravenous administration of drugs. The physiological saline or drugs (Ginsenoside Rb1, L-NAME) was slowly administered through the femoral catheter. Heparin (500 U/kg) was administered intravenous and the right carotid artery was catheterized for the continuous monitoring of mean arterial pressure (MAP) and for blood sampling. The electrocardiogram and blood pressure signal was displayed on an M3 monitor (Siemens, Germany). A left thoracotomy in the fourth intercostal space was performed and the pericardium was opened. A 6–0 silk suture was passed with a tapered needle under the left anterior descending coronary artery, 2 mm from the tip of the left auricle. The ends of the suture were threaded through a piece of plastic tubing, forming a snare that occluded the artery when tightened and clamped. The coronary artery was occluded for 30 min and then allowed reperfusion (by releasing the clamp) for 2 hr.

2.3. Experimental Protocol. The diabetic rats were randomly assigned to five groups, and each group 8 rats: group 1, sham-operated rats (Sham); group 2, ischemia/reperfusion group (30 min of myocardial ischemia and 120 min of reperfusion, I/R); group 3, Ginsenoside Rb1 preconditioning group, Ginsenoside Rb1 40 mg/kg was administered intravenously 10 min before coronary ischemia and reperfusion (Rb1) [14]; group 4, L-NAME + I/R group, L-NAME was given at a dose of 10 mg/kg 25 min before ischemia (L-NAME) [17]; group 5, L-NAME + Ginsenoside Rb1-treated group (L-NAME + Rb1). In the second set of experiments, seven similar experimental groups of rats (n = 4 in each group) were subjected to the same experimental procedures. At the end of reperfusion, the animals were sacrificed by an intravenous injection of 10% potassium chloride solution, and myocardial samples were collected from ischemic left ventricle regions for protein analysis. All animals survived after ischemia reperfusion until being terminated at the completion of the experiment. The samples were frozen in liquid nitrogen, and stored at −80°C for later analysis.

2.4. Determination of the Area at Risk and Infarct Size. At the end of the 2 hr of reperfusion, the rats were given heparin (1 U/g, intraperitoneally). The coronary artery was reoccluded, and 1 mL of 1.5% solution of Evans blue dye was injected via the left femoral vein to identify the ischemic risk area. 2 mL of 10% potassium chloride solution was injected into the femoral vein to stop the heart, and then, the heart was excised. The presence of Evans blue dye indicated nonischemic area and its absence indicated area at risk (AAR). After resecting the right ventricle, the left ventricle (LV) was cut into five transverse slices from the apex to the base. The slices were incubated in 1% triphenyltetrazolium chloride (TTC) solution at 37°C for 25 min, and they were then photographed with a digital camera (Canon, PowerShot AS1000, Japan) and weighed. For each slice, the total LV area, the area lacking Evans blue staining (AAR) and the area lacking TTC staining (infarct area, IA) were determined using a light electron microscope (Olympus, Japan). The ratios of IA/AR and AAR/LV were calculated for each slice and then summed over all slices.

2.5. Detection of Plasma Creatine Kinase (CK) and Lactate Dehydrogenase (LDH). The cardiac damage was evaluated by measuring plasma creatine kinase (CK) and lactate dehydrogenase (LDH). The blood samples were drawn from the right carotid artery at the end of reperfusion period. The plasma concentrations of CK and LDH were measured by automatic biochemical analyzer (AU-2700, Olympus, Japan).
2.6. Histopathological Examination. At the end of the experiments, the formalin-fixed, paraffin-embedded sections of myocardial tissues were fixed in 10% buffered formalin and sections were prepared from paraffin-embedded tissues. The level of histological tissue injury was assessed by hematoxylin and eosin (H&E) staining using light microscopy (400x magnification).

2.7. Immunohistochemical Staining for eNOS. The expression of eNOS was determined by immunohistochemistry. Paraffin-embedded left ventricular tissue blocks were sectioned at 3 μm. The sections were deparaffinized, rehydrated, treated with target retrieval buffer, blocked with 3% hydrogen peroxide, washed with phosphate-buffered saline (PBS), and blocked with 5% normal goat serum in PBS for 30 min. The sections were then incubated overnight with the polyclonal antienothelial nitric oxide synthase (eNOS) antibody (rabbit antienothelial nitric oxide synthase, 1:500, pH 7.2, with 0.1% bovine serum albumin (Boster Bio-Tech, Wuhan, China), followed by biotin-conjugated secondary antibody at 1:1000 dilutions. Finally, the sections were incubated with avidin-biotin complex kit (Boster Bio-Tech, Wuhan, China) and detected by using a dianinobenzidine (DAB) reagent (Boster Bio-Tech Wuhan, China). The slides were examined in 400-fold magnification by light microscopy (Olympus BX50 Microphotographic System, Japan). For each animal, three random tissue sections (five fields per section) were examined. Quantitative immunohistochemical assessments for myocardial eNOS expression were performed as previously described. A mean optical density (OD), which related to immunohistochemical staining intensity, was measured by image cytometry with HIPAS-2000 image analysis software (Qianli Technical Imaging, Wuhan China).

2.8. Determination of SOD and MDA. The myocardial oxidative stress contents were assayed by the measurement of SOD and MDA. At the end of reperfusion, the myocardial supernatant was isolated from ischemia heart tissue samples by centrifugation at 4000 rpm for 10 min at 4°C. The MDA level in the supernatant was determined by the measurement of the thiobarbituric acid (TBA) reaction using a commercial kit (Jiancheng Biological, Nanjing, China) [18]. And the SOD activity in the supernatant was evaluated by inhibition of nitroblue tetrazolium (NBT) reduction by O\textsubscript{2} generated by the xanthine/xanthine oxidase system using a commercial kit (Jiancheng Biological, Nanjing, China) [19].

2.9. Assay of NO Production. The level of NO in myocardial tissue was measured by the Griess method and according to the indication on the NO assay kit (Jiancheng Biological, Nanjing, China). The myocardium was lysed and centrifuged and then 50 μL of Griess reagent was added to 50 μL of supernatant. Nitrite concentration was determined by a spectrophotometer (721-100; Tianpu Analytical Instrument Co., Shanghai, China) at 550 nm from a standard curve (0–100 μmol/L) derived from NaNO\textsubscript{2}.

2.10. Statistical Analysis. Values were expressed as means ± SEM. All biochemical parameters including plasma LDH, CK, SOD, and MDA were assayed in duplicate. Therefore, these data are themselves means of duplicate assays. Statistical analysis was performed using SPSS 13.0 for Windows software. The statistical significance was determined by ANOVA. A value of $P < 0.05$ was considered to be statistically significant.

3. Results

3.1. Baseline and Model Characteristics. For each diabetic animal model, a 4-hour fasting plasma glucose (FPG) was measured every 14 days. The diabetic rats with fasting plasma glucose concentrations ≥16.7 mmol/L showed signs of polydipsia, polyphagia and polyuria (Table 1).

3.2. Infarct Size Assessment of the Operated Groups. The ratio of area at risk of left ventricular mass (AAR/LV) did not differ significantly among the groups. In I/R group, the infarct size was 51.7 ± 4.34% of the area at risk (IS/AAR). Pretreatment with Ginsenoside Rb1 yielded an infarct size of 36.9 ± 2.34% ($P < 0.05$ versus I/R group). Administration of L-NAME or I/R + Rb1 + L-NAME did not affect infarct size ($P > 0.05$ versus I/R group). Pretreatment with L-NAME hindered the cardioprotective effect of Ginsenoside Rb1 preconditioning ($P < 0.05$ versus Rb1 group) (Figure 1).

3.3. Effects of Ginsenoside Rb1 on Plasma CK and LDH. To investigate whether Ginsenoside Rb1 limited cardiomyocyte necrosis, the activity of serum CK and LDH were measured at the end of reperfusion period. Comparing with Sham group, serum CK and LDH of I/R group were markedly
Table 1: The fasting plasma glucose and body weight of diabetic rats (*n* = 60).

| Parameter | 5d | 2W | 4W | 6W | 8W |
|-----------|----|----|----|----|----|
| FPG (mmol/L) | 19.9 ± 2.4 | 23.0 ± 2.4 | 22.9 ± 1.9 | 22.4 ± 3.8 | 25.2 ± 2.8 |
| BW (g) | 258.5 ± 35.4 | 273.6 ± 23.2 | 283.6 ± 28.4 | 266.0 ± 21.5 | 242.8 ± 22.3 |

Figure 2: Effects of Rb1 with or without L-NAME pretreatment on plasma levels of CK, LDH after 30 min of myocardial ischemia and 120 min of reperfusion. Results are expressed as mean ± SEM. *P* < 0.01 versus Sham group, *#* P < 0.01 versus I/R group, *Δ* P < 0.01 versus Rb1 group.

Figure 3: Effect of Ginsenoside Rb1 preconditioning on SOD and MDA. Results are expressed as mean ± SEM. *P* < 0.01 versus Sham group, *#* P < 0.01 versus I/R group, *Δ* P < 0.01 versus Rb1 group.

Increased (both *P* < 0.01). After preconditioning with Ginsenoside Rb1, the CK and LDH levels were significantly lower compared with the I/R group (both *P* < 0.05). Moreover, pretreatment with L-NAME completely abolished Ginsenoside Rb1-induced decrease in plasma CK, LDH (Figure 2).

3.4. Effects of Ginsenoside Rb1 on Myocardial SOD and MDA Levels. To examine whether Ginsenoside Rb1 decreased the diabetic myocardial oxidative stress, SOD and MDA levels in the ischemia heart tissue were measured. Comparing with Sham group, MDA level in I/R group was markedly increased, while SOD activity was lower (*P* < 0.05). After preconditioning with Ginsenoside Rb1, MDA levels in Rb1 and Rb1 + L-NAME groups were decreased significantly compared with the I/R group, while the activity of SOD were higher (*P* < 0.05), (Figure 3).

3.5. Effect of Ginsenoside Rb1 on Myocardial NO Production. To further investigate the biochemical events associated with Ginsenoside Rb1 pretreatment, NO production in ischemic heart was evaluated. NO production, which was indicated as the nitrite formation, was found to be decreased in I/R group. We found that Ginsenoside Rb1 induced a significant raise in nitrite level (*P* < 0.05); however, this effect of elevated nitrite formation was inhibited with the presence of L-NAME (*P* < 0.05), (Figure 4).

3.6. Expression of eNOS in the Diabetic Heart. As shown in Figure 5, a very small amount of eNOS was detected in myocardial tissue of the Sham group. Significantly reduced expression of eNOS was observed in group I/R (*P* < 0.05,
versus Sham). When compared with group I/R, group Rb1 had significantly higher expression of eNOS ($P < 0.05$). Pretreatment with L-NAME almost completely abolished the induction of the expression of eNOS ($P < 0.05$, L-NAME + Rb1 versus Rb1 group).

3.7. Heart Histopathologic Changes. The histopathological changes in the ischemic heart tissue at the end of reperfusion were assessed by standard H&E staining. As shown in Figure 6, the myocardium of Sham group maintained normal tissue structure and shape, and the myocardial fibers were arranged in an orderly manner. I/R group showed acute heart injury characterized by areas of necrosis, neutrophilic inflammation, myocardial, and interstitial edema. The myocardial fibers were partially ruptured and lysed. Rb1 group revealed markedly reduced neutrophilic inflammation and interstitial edema with preservation of myocardium compared with I/R group. More importantly, L-NAME was administered prior to Rb1, the destruction of heart tissue was more severely as compared with I/R group.

4. Discussion

In this study, Ginsenoside Rb1 preconditioning limited the heart infarct size and MDA concentrations, while increasing SOD activity and attenuating cellular damage. The protective effects induced by Ginsenoside Rb1 preconditioning were related to the expression of eNOS and NO. These protective effects were blocked by an eNOS inhibitor (L-NAME), suggesting that eNOS activity and endothelium-derived NO mediated the protective effects of Ginsenoside Rb1 pretreatment.

Several mechanisms have been proposed for the oxidative damage during chronic hyperglycemia, such as mitochondrial reactive oxygen species (ROS) overproduction [20], glucose auto-oxidation [21], synthesis of AGEs [22], and hyperglycemia reduced NO availability. Endothelial dysfunction has been documented in various forms of diabetes and even in prediabetic individuals [23, 24]. The pathogenesis of this endothelial dysfunction involves many components including increased polyol pathway flux, altered cellular redox state, increased formation of diacylglycerol, and the subsequent activation of specific protein kinase C isoforms, and accelerated nonenzymatic formation of AGEs. There are many clinical and experimental evidences suggesting that increased sympathetic activities, activated cardiac renin-angiotensin system, myocardial ischemia/functional hypoxia, and elevated circulating levels of glucose result in oxidative stress in cardiovascular system of diabetic subjects. Oxidative stress associated with an impaired antioxidant defense status may play a critical role in subcellular remodeling, calcium-handling abnormalities, and subsequent diabetic cardiomyopathy [25]. A clinical study indicated that systemic oxidative stress occurred during cardiopulmonary bypass and postcardiopulmonary bypass periods and also in patients with diabetes mellitus, and the systemic oxidative stress was higher [26]. So, we hypothesized that Ginsenoside Rb1 pretreatment may attenuate myocardial ischemia reperfusion injury through an increase in the activity of eNOS, enhancing the availability of NO, and decreasing the oxidative stress.

NO released from endothelial cells, is a pivotal vaso-protective molecule, which maintains normal vascular tone, regulates leukocyte-endothelial cell interactions, inhibits platelet aggregation, limits smooth muscle cell proliferation, and may affect cardiac myocyte function, neutrophil activation, and free-radical production [11, 27, 28]. Previous studies clearly demonstrated that the deficiency of eNOS exacerbates myocardial I/R injury [29], and myocardial ischemia and reperfusion are associated with marked impairment of endothelium-dependent coronary relaxation, reduced contractility, and cardiac arrhythmias. In many studies, reperfusion injury has been attributed to a reduced formation or activity of NO because the NO precursor, L-arginine, NO donors of different chemical structures, and interventions resulting in stimulated NO production involving endothelial receptors have all been found to protect the heart [30]. Moreover, the overexpression of eNOS, the administration of NO donors, and inhaled NO gas therapy all significantly protect the myocardium [31]. Yang et al. [32] also showed that NO released by endothelial NO synthase (eNOS) is important in blood pressure regulation and also has an impact on cardiac function and remodeling.

In addition, there are evidences strongly suggesting that Ginsenoside Rb1 bears various beneficial effects on the cardiovascular system. In cell culture model, Ginsenoside Rb1 enhanced NO production and the expression of eNOS mRNA in TNF-α stimulated HUVECs. [33]. In vitro, Ginsenoside Rb1 has been shown to reduce homocysteine-induced endothelial dysfunction and free radical production as well as eNOS downregulation in porcine coronary arteries [34–36]. Study has shown that Ginsenoside Rb1 significantly inhibited cardiocyte apoptosis and reduced ischemia-reperfusion in rats [15]. Recent studies demonstrated that administration of Ginseng berry significantly improved systemic insulin sensitivity and glucose homeostasis in the rats. Others have shown that the Ginsenoside compounds attenuated inhibitory inflammatory response after cardiopulmonary bypass in patients with congenital heart diseases, which played an
that Ginsenoside plays a significant role in antihyperglycemic action [38]. Therefore, it was important to determine whether pretreatment with Ginsenoside Rb1 would reduce myocardial infarction after ischemia reperfusion events. In addition, a previous study showed that Ginsenoside Rb1, a major active component of Ginseng, could actually induce NO production and regulate acute eNOS activation in human aortic endothelial cells [39].

In this study, by using an in vivo rat model, we found that Ginsenoside Rb1 preconditioning significantly decreased infarct size and plasma CK, LDH after 30 min of regional myocardial ischemia and 120 min of reperfusion, and Ginsenoside Rb1 pretreatment attenuated the increase in necrosis, neutrophilic inflammation respond, and myocardial and interstitial edema. The myocardial MDA level, reflecting the lipid peroxide and oxidative stress, was also significantly suppressed in Rb1 group, while the SOD activity was increased. Ginsenoside Rb1 upregulated eNOS and NO contents, while eNOS inhibitor (L-NAME) reversed the protective effect of Ginsenoside Rb1 pretreatment. Therefore, our study demonstrated the protective effect of Ginsenoside Rb1 pretreatment, at least in part, by eNOS and NO. Certain limitations of the present study need to be noted. The current study did not investigate mRNA and protein expressions of eNOS. This will be the goal of future study.
In summary, our study suggests that Ginsenoside Rb1 preconditioning attenuated the severity of heart injury induced by myocardial ischemia/reperfusion injury in diabetic rats. The protective effect of Ginsenoside Rb1 preconditioning was mediated in part through the induction of endogenous NO released by eNOS.

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