From Biorefinery to Food Product Design: Peach (*Prunus persica*) By-Products Deserve Attention

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Abstract
There is an increasing demand for functional foods to attend the consumers preference for products with health benefits. Peach (*Prunus persica*), from Rosaceae family, is a worldwide well-known fruit, and its processing generates large amounts of by-products, consisting of peel, stone (seed shell + seed), and pomace, which represent about 10% of the annual global production, an equivalent of 2.4 million tons. Some studies have already evaluated the bioactive compounds from peach by-products, although, the few available reviews do not consider peach by-products as valuable materials for product design methodology. Thereby, a novelty of this review is related to the use of these mostly unexplored by-products as alternative sources of valuable components, encouraging the circular bioeconomy approach by designing new food products. Besides, this review presents recent peach production data, compiles briefly the extraction methods for the recovery of lipids, proteins, phenolics, and fiber from peach by-products, and also shows in vivo study reports on anti-inflammatory, anti-obesity, and anti-cerebral ischemia activities associated with peach components and by-product. Therefore, different proposals to recover bioactive fractions from peach by-products are provided, for further studies on food-product design.

Keywords Design thinking · Circular economy · Bioactive ingredients · Sustainable processes

Introduction
The search for sustainable industrial processes has guided the production of chemicals and fuels from biomass-based economy (Arora et al., 2018). However, millions of ton of by-products from food processing industry are currently generated worldwide, being disposed of in landfills, used in composting or for animal feed (Rico et al., 2020). The industry is dealing with the increasing interest in naturalness as current and future trends in global trades. The companies, aware of the consumers demands, are continuously looking for process innovations to maintain the natural characteristics of the products. Therefore, understanding the concept of product design is highly relevant for the development of food products, warranting naturalness attributes. Firstly, the highest challenge for companies is understanding the customer’s necessities and capture their attention, transforming the consumer’s ideas into chemical and physical parameters of the final product, improving its appeal (Taifouris et al., 2020). Therefore, in order to attend these demands, the companies must follow modern and innovative technologies to provide competitive and sustainable products, resulting in optimized food production with less water and energy consume, and also better use of edible and non-edible parts of food raw materials and supplies. Consequently, the companies need to improve the design, optimization, and development of different formulations and techniques, before introducing new food products to the market (Granato et al., 2020).

Besides, it is essential to change the form of production, converting the linear economy into a sustainable circular bio-economy. The philosophy of the dominant economic model of “take, make and dispose” must be urgently transformed into a sustainable model (Maina et al., 2017). For that, the concept of upcycled foods, which is the use of food ingredients or processed food materials, that would, otherwise, be discarded, must be properly understood (Goodman-Smith...
et al., 2021). Applying the concepts of circular economy and biorefinery is decisive to recover high-added value molecules since the growing demand for processed foods increases the generation of residues or by-products from the processed foods (Banerjee et al., 2018, 2019). Mostly, these biomasses are not fully exploited, with insufficient knowledge about their nutritional and economic values (Banerjee et al., 2018). Some fruit residues have already been studied considering the biorefinery concept, such as pineapple, orange, mango, and banana (Arora et al., 2018; Banerjee et al., 2019; Naranjo et al., 2014; Torre et al., 2019). However, no similar data have been found for peach by-products. Therefore, a search of Scopus database (http://www.scopus.com) was performed using a different set of Booleans concerning peach and its residues or by-products from studies published from 2010 to 2021. The search considered the title, abstract, and keywords of the published works, and the results are presented in Fig. 1.

Regarding documents, the first search included the terms “peach” AND Prunus AND residue OR waste OR “by-product”, showing 236 documents, represented by research articles (93.6%), book chapters (0.4%), review articles (2.5%), book chapter (0.4%) and other (3.4%). In order to expand the search and comparing with other fruits of the same genus such as nectarine, plum, apricot, and cherry, and using the same Booleans (Fig. 1), the results showed the peach fruit represented highest quantity of article (93.6%), followed cherry (88.5%), and plum (88.2%). These searches demonstrate the growing interest in peach by-products, although, there is still a lack of reviews to support the researches in this area. Those studies found from this search are related to wood residues from the peach plantation and the removal of pharmaceutical compounds using systems with different stone residues. Therefore, the present review deals with the contribution of peach by-products bringing a combined product design approach to the biorefinery concept considering its use as biomass, and as a potential source of bioactive compounds, highlighting in vivo studies.

Food production suffered major transformations in recent years, mostly stimulated by the increasing demand for sustainably processed foods, which must be considered safe, fresh, natural and with high-nutritional value (Granato et al., 2020). Due to the COVID-19 pandemic, the search for natural components with bioactivities such as anti-inflammatory has deeply increased because they may play a crucial role in many diseases, including viral infections. This is explained by the use of products enriched with bioactive compounds, which can contribute to human health and the immune system (Benvenutti et al., 2021).

Although there are few literature data related to the extraction of bioactive compounds from peach residues and by-products associated with biological activities, additional information on how to improve the use of these residues is still in need. In this context, this review focuses on peach industrial by-products, production data, chemical composition, and biorefinery and product design approaches for the recovery and use of these compounds.

**Peach Fruit and Its Industrial Processing**

Customers widely appreciate peach fruits (*Prunus persica*) for their taste, texture, juiciness, and nutritional value. According to genomic and phenotypic evidence, some reports accredit peach origin to southwest China (Patra & Baek, 2016). Besides, this fruit belongs to the *Rosaceae* family (Nowicka & Wojdylo, 2019), and nowadays it is estimated that there are currently more than 400 cultivars

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**Fig. 1** Publications about peach, nectarine, cherry, apricot and plum from 2010 to 2022 according to SCOPUS Database Platform (www.scopus.com).

Source: the author
worldwide (Li & Wang, 2020). Therefore, peach is one of the most variable fruit species, with diverse germplasms, and different shapes, sizes, seeds, peel, and pulp colors (red, white, or yellow flesh). The fruit also presents a climacteric peak, where the respiration and evolution of ethylene show a drastic increase, compared to non-climacteric fruits. Therefore, because it is highly perishable, cold storage after harvesting is an important ally in reducing respiration rates and maturity associated to metabolic changes during senescence (Minas et al., 2018). The peach fruit is divided into three parts. The first is the pulp or mesocarp, corresponding to 75.2% of the fruit weight. The pulp is juicy, yellow and usually has an acid and sweet taste, varying widely, and with pH on average from 3.50 to 4.00 (Featherstone, 2015). The second part is the peel (exocarp), which represents 22.5% of the fruit. The last part is the stone, the endocarp, which is formed by the seed (inside), covered by a hard shell, denominated seed shell or kernel shell. The seed represents 5 to 12.5% of the fruit weight, depending on the peach specie (De França Sousa et al., 2018; Nowicka & Wojdylo, 2019). In general, the stones consist an average of 6% of seed and 94% of seed shell (Uysal et al., 2014). According to the Food and Agriculture Organization Corporate Statistical Database (FAOSTAT, 2020), the world production of peaches and nectarines in 2020 was over 24.5 million tones. Peaches and nectarines are very similar, and belong to the same specie (*Prunus persica*) and family (Rosaceae). For that reason, the data are presented combined (Carrasco et al., 2013). In 2020 production of the five continents were Asia 73.6%, Europe 14.9%, Americas 7%, Africa 4.3%, and Oceania 0.3%, where China, Spain, Italy, Türkiye, Greece, Iran, and the USA are the world leader countries in peach production (FAOSTAT, 2020). In 2020, Brazil occupied the 15th world position, with peaches and nectarines production of 201,880 ton.

Besides the economic importance, peaches have several nutritional benefits due to their high content of vitamin A, potassium and organic acids, sugars, and minerals, conferring interesting nutritional value (Manzoor et al., 2012). This fruit is highly consumed and destined for different food products such as jellies, sweets, and tea, among others. Besides, peaches are also used in formulations of various cosmetic products (Vásquez-Villanueva et al., 2015). Canned peaches in syrup cover 93% of the total processed peach products, while peach jam covers 6% and peach juice only 1% (Kamenidou et al., 2002).

The peach industrial process depends on the final product, with the most popular represented by peach in syrup (canned or glass vessels) and concentrated peach purée, where the last is used as an ingredient for formulations such as baby food, juice, jams, pulp and yogurt (Aktağ & Gökmen, 2021). Figure 2 shows a flowchart of peach processing for these featured products and their respective generated by-products.

Basically, the processing of peaches in syrup consists of harvesting, selection, peel chemical removal (sodium

![Diagram](image-url)
hydroxide solutions from 1.5 to 2% concentration, near boiling temperature) (Featherstone, 2015), cutting stone removal and steam blanching. It is followed by filling the syrup at the glass or canned packaging, for final pasteurization. Peach purée concentrate is obtained by washing/selecting, removing the leaves and the stones (formed by seed and seed shell) (Fig. 2), crushing, heating (90–95 °C), evaporation (60–75 °C), deaeration, filling and sterilization (105–120 °C), and purée concentration (Aktag & Gokmen, 2021), which is ready for packaging and commercialization.

The main residues generated from peach processing are peel, pomace, and stone. According to Plazzotta et al. (2020), around 15 million metric ton of peach are processed worldwide each year to produce juices, with approximately 10% of discard, depending on the fruit’s ripeness. Consequently, based on the annual global production and considering 10% of residues or by-products (Plazzotta et al., 2020), above 2.4 million ton of peach by-products are globally generated each year. Besides, considering the Brazilian production in 2020, about 20 thousand ton of peach by-products are generated each year in Brazil. The increase in fruit harvesting and processing, and the lack of proper handling methods and infrastructure, also increase the generation of by-products, which are poorly explored worldwide. Studies demonstrate that peach by-products contain oils (seed), anthocyanins (pomace), pectin (peel and pomace), and proteins (seed) (Faravash & Ashtiani, 2008; Pagán et al., 2001; Vásquez-Villanueva et al., 2015; Wu et al., 2011).

Normally, peach processing by-products are disposed in landfills, used as animal feed, burned, or used in steam production. Therefore, considering the possibility to improve the use of these by-products, new strategies have been developed for the recovery of relevant substances from these biomasses, such as bioactive compounds, vitamins, pectin, and phenolics, which could be recovered for further use in pharmaceutical, food, and cosmetic industry. Consequently, the following topics discuss relevant aspects for better use of peach by-products, and are valuable for applying the product design methodology to obtain derived products from the peach processing chain.

**Chemical Composition**

Considering that peach by-products are represented mostly by peel, pomace, and stone (seed + seed shell), it is important to investigate the components from these constituents to evaluate their potential uses. Different studies have shown that fruit peels present several beneficial substances, such as flavonoids, hydroxycinnamic acids, flavonols, anthocyanins, and carotenoids, which are highly accumulated in the peel compared to the pulp of various fruits (Michailidis et al., 2021). Patra and Baek (2016) evaluated the peach peel composition and detected components such as chlorogenic acid, catechin, epicatechin, rutin, and cyanidin-3-glycoside. Additionally, Saidani et al. (2017) detected the following phenolic compounds from peach peel: Flavonols (quercetin-3-galactoside; quercetin-3-O-glucoside plus quercetin-3-O-rutinoside; kaempferol-3-O-glucoside), Hydroxycinnamic acid (neochlorogenic acid; p-coumaroylquinic acid; chlorogenic acid; 4-cafeoylquinic acid; caffeoylquinic acid derivative) and anthocyanin (cyanidin-3-O-glucoside). The content of flavonoids from peach peel ranged from 39.3 to 245 mg equivalent of catechins/100 g of fresh weight (fw) while for peach pulp these values ranged from 8.18 to 112 catechins/100 g of fresh weight, considering nine different peach cultivars. Besides, Saidani et al. (2017) indicated chlorogenic acid as the main hydroxycinnamic acid from peach peel (from 6.74 to 31.2 mg/100 fw), followed by neochlorogenic acid (1.02–7.98 mg/100 fw) and anthocyanins (from 0.24 to 17.6 mg cyanidine-3-glycoside/100 g fw). These compounds were more representative from peach peel compared to peach pulp. In addition, Redondo et al. (2021) detected high amounts of quercetin from peach peel (7.1 mg 100/g fw), compared to other fruits such as plum (5.8 mg 100/g fw) and apricot (6.4 mg 100/g fw).

Nowicka and Wójdyło (2019) evaluated seeds composition of 20 different peach varieties. The authors detected (+)-catechin content ranging from 49.49 to 250.79 mg/100 g dry matter (dm). The flavan-3-ols dimers, procyandin B1 and procyandin B2, were detected in most seeds, with average content of 150.65 and 28.12 mg/100 g dm, respectively. The group of hydroxycinnamic acids was the second main polyphenolic group detected in peach seeds, after flavan-3-ols. These phenolic acids ranged from 130.94 mg to 2275.95 mg/100 g. In addition, the main carotenoids found were β-carotene and xanthophylls (mono- or di-hydroxylated carotenoids), zeaxanthin, β-cryptoxanthin, and violaxanthin (Lara et al., 2020).

The peach pomace is a by-product from the juice processing and contains the fruit peel and pulp. Several studies have shown that peach pomace is rich in pectin (Faravash & Ashtiani, 2007), while peach pulp has phenolic compounds and anthocyanins, with contents varying with peach species (Cevallos-Casals et al., 2006). Caffeic acid, protocatechuic acid, and gallic acid were also detected in peach pomace (El Darra et al., 2018). For instance, the methanolic extract presented from 100 to 449 mg of chlorogenic acid equivalent (CGA)/100 g of fresh pulp weight, while the main identified anthocyanin, the cyanidin 3-glycoside, varied from 6 to 37 mg of cyanidin 3-glucoside equivalents/100 g fresh pulp weight. Liu et al. (2018), evaluated the antioxidant potential from peel and pulp of four varieties of peach from China. Methanol/acetone extracts showed that peels have higher phenolic content, from 45.5 to 64.8%, compared to
the pulp. Abidi et al. (2015) evaluated the aroma and phenolic compounds from nectarine *Prunus persica* (L. Batsch) genotypes, with results indicating this fruit, from the same family as peach, as a good source of phenolic compounds, especially ascorbic acid. The authors identified more than 60 volatile compounds, including 10 carboxylic acids, 10 aldehydes, 5 alcohols, 3 esters, 12 ketones, 8 lactones, and 12 terpenoids.

Finally, the lignocellulosic composition of peach stone consists mainly of lignin, cellulose, and hemicellulose, forming complex bonds on cell walls, where the lignin provides physical protection around the sugar fraction (Buratti et al., 2018). Uysal et al. (2014) indicated peach stone (seed shell + seed) is composed of 46% cellulose, 14% hemicellulose, and 33% lignin. For Instance, Hong et al. (2021) characterized the phenolic acids from the ethanolic extract (70%, 20 mL) from peach seed shell, identifying the main hydroxycinnamic acid components as p-Coumaric acid 4-O-glucoside, 3-Caffeoylquinic acid, 3-Feruloylquinic acid, 3-p-Coumaroylquinic acid, m-Coumaric acid, Hydroxycaffeic acid. The second group was hydroxybenzoic acids, formed by: Ellagic acid acetyl-xyloside, Protocatechuic acid 4-O-glucoside, 2-Hydroxybenzoic acid, 2,3-Dihydroxybenzoic acid, 3-O-Methylgallic acid. The third group was the hydroxyphenylpropanoic acids: 3-Hydroxy-3-(3-hydroxyphenyl) propionic acid. The fourth group was hydroxyphenylpropanoic acids: Dihydrocaffeic acid 3-O-glucuronide and 3-Hydroxy-3-(3-hydroxyphenyl) propionic acid. The main anthocyanin detected was Cyanidin 3-O- (2-O- (6-O- (E) -caffeoyl-D-glucoside) -D-glucoside) -5-O-D-glucoside. These results indicate the high-value of the components present in the peach seed shell, which suggest possible uses in food, nutraceutical, and pharmaceutical industries.

The main phenolic compounds detected from the peach by-products (peel, pomace and seed shell) are highlighted in Fig. 3, based on the works by El Darra et al. (2018); Hong et al. (2021); Nowicka and Wojdyło (2019); Saidani et al. (2017). The main groups of detected compounds were hydroxycinnamic acids, flavonols, hydroxybenzoic acids, and procyanidins, which were detected by various researchers on peach by-products.

Besides, some studies have evaluated that peach seeds are also rich in fatty acids and protein. For instance, Hao et al. (2019) used gas chromatography–mass spectrometry (GC–MS) to define the fatty acid composition from peach seed oil, indicating 86% of unsaturated fatty acids, within the total fatty acids content. The main unsaturated fatty acids were oleic (55.2%) and linoleic (30.8%) acids, while the main saturated fatty acids were palmitic (7.97%), stearic (2.37%), and α-linolenic (0.11%) acids. Peach seed extracts were obtained by Shukla and Kant (2020) using Soxhlet with different solvents, petroleum ether, chloroform, ethyl...
acetate, ethanol, and water. The results provided 29.36% of protein and 7.48% of crude fat from the dry seed. In addition to the high protein and fat contents, peach seeds are also rich in phenolic compounds (González-García et al., 2016; Sánchez-Vicente et al., 2009).

Peach by-products are considerable sources of nutrients such as lipids, proteins, fibers, and carbohydrates, and the centesimal composition for peach by-products (pomace, peel and stone) is summarized in Table 1. For instance, the oil and protein content from peach seeds represents 48.4% and 26.7%, respectively (Gettens, 2016; Lazos, 1991; Rahma & El-Aal, 1988) while the fiber content from the peach peel represents 57% (Pla et al., 2012; Yangilar, 2016), which makes seed and peel as interesting by-products. Besides, the most recent and relevant works about the chemical composition of peach by-products are summarized in Table 2. It shows the highest relevance of peach peel as a research subject, compared to other peach by-products, with most compounds evaluated by various chromatography methods.

### Recovery of Valuable Components from Peach By-Products

Extraction methods are the most common operations used to obtain natural extracts from raw materials, for further use as a food ingredient (Zhang et al., 2018). Several extraction techniques can be used, from classical, or conventional procedures, to emergent or alternative methods. Maceration and Soxhlet (SOX) are within the conventional and well established most used methods. Otherwise, pressurized liquid extraction (PLE), supercritical fluid extraction (SFE), microwave-assisted extraction (MAE), and ultrasound-assisted extraction (UAE) are within the alternative procedures (Rudke et al., 2020). Nowadays, various studies reported the use of these emergent methods due to their advantages over conventional techniques, such as high yields or selectivity, and use of solvents generally recognized as safe (GRAS). Besides, these alternative methods are fast, have low energy and solvent consumption, and can be very selective towards target compounds. Then, these combined advantages can designate these methods as ecological processes with less environmental impact than traditional procedures (Martins et al., 2020). Various studies involving different extractions methods for peach by-products are analyzed as follows.

Plazzotta et al. (2020) studied the pomace from peach juice processing. Microwave-assisted extraction (MAE) and ultrasound-assisted extraction (UAE), both methods using ethanol: water (70:30) as solvent at 80 °C, were applied for the recovery of antioxidant compounds from the peach juice pomace. The optimization of the processes indicates the best extraction conditions as 540 W during 50 s for MAE, and using 23% amplitude during 120 s for UAE. The extract samples recovered from the frozen peach residues provided higher flavonoids, anthocyanins, and vitamin C, compared to extracts from dry residues. Plazzotta et al. (2021) used two extraction methods with ethanol (70%) to obtain unpurified bioactive compounds for peach pomace, a conventional thermal extraction, at 50 °C for up to 90 min, and the pulsed electric field (PEF) technology with an energy input of 0.0014 to 2.88 kJ/kg. The processing time required for PEF was much lower, in the order of microseconds (μs), compared to conventional thermal extraction (average time of 40 min, reaching similar extraction efficiency than PEF). The bioactive fraction recovered by PEF from peach pomace reached total flavonoid content (TFC) of 86 mg QE/100 dm, total phenolics content (TPC) of 386 mg GAE/100 g dm, and DPPH value of 80 mg TE/100 g dm. Otherwise, the conventional thermal treatment, set at 50 °C, provided maximum values for TPC, TFC and DPPH of 524 ± 27 mg GAE/100 g dm, 108 ± 1 mg QE/100 g dm and 143 ± 7 mg TE/100 g dm, respectively. Although, the traditional extraction method shows samples with the highest values of TPC, TFC, and DPPH, the PEF method is a promising technology because it has lower energy costs than traditional methods, and recovers valuable compounds.

Adil et al. (2007) recovered polyphenols from peach pomace by subcritical extraction with CO₂ added with ethanol. The effect of the process parameters, pressure (20–60 MPa), temperature (40–60 °C), ethanol concentration (14–20% by weight), and extraction time (10–40 min) were studied. The results indicate that the increase in all parameters significantly contributes to the recovery of total phenolics content (TPC), and DPPH performance to evaluate the antioxidant activity. The optimum conditions were 51 MPa, 52.3 °C, 20% ethanol content and 40 min of extraction. TPC values were up to 0.26 mg of gallic acid equivalent/g of dried

### Table 1 Average centesimal composition of peach by-products, in percentage (%) of dry samples

| Fruit by-product | Carbohydrate | Fiber | Protein | Lipids | Ashes | Reference |
|------------------|--------------|-------|---------|--------|-------|-----------|
| Pomace           | 13.06        | 38.85 | 5.32    | 0.37   | 2.13  | Pagán and Ibarz (1999), Pagán et al. (2001), Sarkar and Choudhury (2014) |
| Peel             | 31.00        | 57.48 | 9.01    | 2.88   | 4.30  | Pla et al. (2012), Yangilar (2016) |
| Seed             | 12.91        | 4.0   | 26.77   | 48.41  | 3.82  | Gettens (2016), Lazos (1991), Rahma and El-Aal (1988) |
Table 2 Summarizes chemical composition data from peach by-products and the identification methods used

| Source          | Work proposal                                                                                                                                                                                                                                                                                                                                 | Identification technique * | Chemical composition                                                                                                                                                                                                                                                                                                                                 | References | Source Work proposal Identification technique * | Chemical composition References |
|-----------------|-----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-----------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|------------|-----------------------------------|----------------------------------|
| Peel            | Identify anthocyanins from peach peel; the regulatory role of 1-methylcyclopropene and ethylene on anthocyanin accumulation, and the mechanism of ethylene-mediated inhibition of anthocyanin biosynthesis in the fruit                                                                                                                          | HPLC-TOF–MS                 | Cyanidin 3-Glucoside, Cyanidin 3-Galactoside, Cyanidin 3-Rutinoside, Delphinidin-3-Glucoside, Petunidin-3-Galactoside and Petunidin-3-Glucoside                                                                                                                                                                      | Zhang et al. (2022) | Identify and quantify the phenolics profile                                                                                                                   | Quercetin-3-Galactoside (FO1), Quercetin-3-O-Glucoside Plus Quercetin-3-O-Rutinoside (FO2) And Kaempferol-3-O-Glucoside | Saidani et al. (2017) |
| Peel            | Identify and quantify the phenolics profile                                                                                                                                                                                                                                                                                                      | UPLC-MS                     |                                                                                                                                                                                                                                                                                                                                                       |            | Determine the polyphenols profile and antioxidant capacity from peach peel and pulp, of 6 different peach cultivars and one nectarine cultivar                                                                                       | Flavanols (Quercetin-3-Rutinoside, Quercetin-3-Glucoside, Quercetin-3-Rhamnoside and Kaempferol-3-Rutinoside) | Stojanovic et al. (2016) |
| Peel            | Determine a time course of UV-B-stimulated transcription of genes involved in UVR8 signaling, in phenolics biosynthesis and their transcriptional regulators; phenolics quantification from peach peel                                                                                                                                                                                      | UHPLC-ESI-QTOF-MS           | Cyanidin (2-(3,4-Dihydroxyphenyl) Chromenyl-3,5,7-Triol; Anthocyanins), (+)-Catechin (Flavanols), Luteolin (3′, 4′, 5, 7-Tetrahydroxylavone; Resveratrol (3′, 4′, 5-Trihydroxy-Trans-Stilbene; Stilbenes), 5-Pentadecylresorcinol (Alkylphenols), Hydroxycinnamic Acids, Sesamine (Furofuran Lignans) and Mataresinol (Dibenzybutyrolactone and Dihydroxydibenzoylbutane Lignan) | Santin et al. (2019) | Peel Report the phenolic and vitamin C composition, in vitro antioxidant potencies and metal chelating activity of pulp and peel for five peach cultivars                                                                 | Hydroxybenzoic Acid (Protocatechuic Acid), Two Hydroxycinnamates (Chrologenic Acid, Neo-Chlorogenic Acid), One Flavan 3-Oh ((+)-Catechin) And One Flavonol (Quercetin-3-Rutinoside) | Liu et al. (2015) |
| Peel            | Determine a time course of UV-B-stimulated transcription of genes involved in UVR8 signaling, in phenolics biosynthesis and their transcriptional regulators; phenolics quantification from peach peel                                                                                                                                                                                      | UHPLC-ESI-QTOF-MS           | Cyanidin (2-(3,4-Dihydroxyphenyl) Chromenyl-3,5,7-Triol; Anthocyanins), (+)-Catechin (Flavanols), Luteolin (3′, 4′, 5, 7-Tetrahydroxylavone; Resveratrol (3′, 4′, 5-Trihydroxy-Trans-Stilbene; Stilbenes), 5-Pentadecylresorcinol (Alkylphenols), Hydroxycinnamic Acids, Sesamine (Furofuran Lignans) and Mataresinol (Dibenzybutyrolactone and Dihydroxydibenzoylbutane Lignan) | Santin et al. (2019) | Peel Report the phenolic and vitamin C composition, in vitro antioxidant potencies and metal chelating activity of pulp and peel for five peach cultivars                                                                 | Hydroxybenzoic Acid (Protocatechuic Acid), Two Hydroxycinnamates (Chrologenic Acid, Neo-Chlorogenic Acid), One Flavan 3-Oh ((+)-Catechin) And One Flavonol (Quercetin-3-Rutinoside) | Liu et al. (2015) |
| Peel            | Peel Report the phenolic and vitamin C composition, in vitro antioxidant potencies and metal chelating activity of pulp and peel for five peach cultivars                                                                                                                                                                                                  | HPLC–DAD                   | Hydroxybenzoic Acid (Protocatechuic Acid), Two Hydroxycinnamates (Chrologenic Acid, Neo-Chlorogenic Acid), One Flavan 3-Oh ((+)-Catechin) And One Flavonol (Quercetin-3-Rutinoside)                                                                                                                                                  | Liu et al. (2015) | Peel Report the phenolic and vitamin C composition, in vitro antioxidant potencies and metal chelating activity of pulp and peel for five peach cultivars                                                                 | Hydroxybenzoic Acid (Protocatechuic Acid), Two Hydroxycinnamates (Chrologenic Acid, Neo-Chlorogenic Acid), One Flavan 3-Oh ((+)-Catechin) And One Flavonol (Quercetin-3-Rutinoside) | Liu et al. (2015) |
| Seeds           | Recover phenolic compounds from the stone from peach (Prunus persica L.), nectarine (Prunus mucipersica L.), plum (Prunus domestica L.) and apricot (Prunus armeniaca L.)                                                                                                                                                                                                  | HPLC–PDA; LC–ESI-QTOF-MS/MS | Gallic acid, Protocatechuic acid, p-Hydroxybenzoic acid, Chlorogenic acid, Caffeic acid, Catechin, Epicatechin, Epicatechin gallate, Quercetin and Kaempferol                                                                                                                                              | Hong et al. (2021) | Seeds Recover phenolic compounds from the stone from peach (Prunus persica L.), nectarine (Prunus mucipersica L.), plum (Prunus domestica L.) and apricot (Prunus armeniaca L.) | Gallic acid, Protocatechuic acid, p-Hydroxybenzoic acid, Chlorogenic acid, Caffeic acid, Catechin, Epicatechin, Epicatechin gallate, Quercetin and Kaempferol | Hong et al. (2021) |
| Seeds           | Separate components from Prunus persica kernel for possible development of anti-inflammatory, analgesic, and antipysic medicinal agents from natural resources                                                                                                                                                                                                                       | LC–ESI–MS/MS                | Amagdal, Prunasin, Apigenin O-pentoside, Methylated flavonoid haxoside and Naringenin O-hexoside                                                                                                                                                                                                                                                      | Eshamy et al. (2019) | Separate components from Prunus persica kernel for possible development of anti-inflammatory, analgesic, and antipysic medicinal agents from natural resources                                                                 | Amagdal, Prunasin, Apigenin O-pentoside, Methylated flavonoid haxoside and Naringenin O-hexoside | Eshamy et al. (2019) |
| Seeds           | Identify the peptides from peach seeds and their relation with the protective effect of genotypes in which they were identified                                                                                                                                                                                                                                           | RP-HPLC-ESI-Q-TOF           | Peptides contained high amounts of hydrophobic amino acids and imidazole-containing amino acids                                                                                                                                                                                                                                                                                                   | Hernández-Coroto et al. (2018) | Identify the peptides from peach seeds and their relation with the protective effect of genotypes in which they were identified                                                                 | Peptides contained high amounts of hydrophobic amino acids and imidazole-containing amino acids | Hernández-Coroto et al. (2018) |
| Seeds           | Evaluate the oxidative stability, thermal behavior, antioxidant activity, phenolic content, and physicochemical properties of Prunus persica kernel oil                                                                                                                                                                                                                | GC                          | Palmitic acid, Palmitoleic acid, Stearic acid, Oleic acid and Linoleic acid                                                                                                                                                                                                                                                                                     | Sodeifian and Sajadian (2021) | Evaluate the oxidative stability, thermal behavior, antioxidant activity, phenolic content, and physicochemical properties of Prunus persica kernel oil | Palmitic acid, Palmitoleic acid, Stearic acid, Oleic acid and Linoleic acid | Sodeifian and Sajadian (2021) |
| Pomace          | Peptides in fraction PSH-3 kilodaltons (kDa) from peach seeds                                                                                                                                                                                                                                                                                                                                                  | LC-Q-TOF–MS/MS              | Peptide isoleucine–tyrosine–serine–proline–histidine (IYSPH)                                                                                                                                                                                                                                                                                                    | Vásquez-Villanueva et al. (2019) | Peptides in fraction PSH-3 kilodaltons (kDa) from peach seeds | Peptide isoleucine–tyrosine–serine–proline–histidine (IYSPH) | Vásquez-Villanueva et al. (2019) |
| Pomace          | To assess the binding capacity of the soluble peach fiber (SPF) as influenced by the microfluidization pretreatment and cellulase hydrolysis                                                                                                                                                                                                                  | HPAEC-PAD                  | Lactose, glucose, xyloose,mannose, and fructose, rhamnose and arabinose                                                                                                                                                                                                                                                                                       | Xu et al. (2015) | To assess the binding capacity of the soluble peach fiber (SPF) as influenced by the microfluidization pretreatment and cellulase hydrolysis | Lactose, glucose, xyloose,mannose, and fructose, rhamnose and arabinose | Xu et al. (2015) |

*HPLC-TOF–MS high-performance liquid chromatography coupled mass spectrometry time-of-flight, UPLC–MS ultrahigh-pressure liquid chromatography coupled mass spectrometry, HPLC–DAD high-performance liquid chromatography diode array detection, UHPLC/QTOF ultrahigh-pressure liquid chromatography coupled quadrupole-time-of-flight, UHPLC–ESI–QTOF–MS ultrahigh-pressure liquid chromatography photodiode array and liquid chromatography electrospray ionization quadrupole-time-of-flight mass/mass spectrometry, HPLC–PDA LC–ESI–QTOF–MS/MS high-performance liquid chromatography photodiode array and liquid chromatography electrospray ionization quadrupole-time-of-flight mass/mass spectrometry, LC–ESI–MS/MS liquid chromatography electrospray ionization mass/mass spectrometry, RP-HPLC–ESI–Q-TOF reversed-phase high-performance liquid chromatography coupled to electrospray-ionization quadrupole-time-of-flight mass spectrometry, CG gas chromatography, LC-Q-TOF–MS/MS liquid chromatography quadrupole-time-of-flight mass/mass spectrometry, HPAEC-PAD high performance anion exchange chromatography coupled with a pulse amperometric detector. The data from Table 2 comprises recent publications from 2015-2021.
sample, with AE of 1.5 mg of DPPH /mg dried sample. Faravash and Ashtiani (2008) recovered pectin from peach pomace in an acid medium (with HCL at 60 °C) followed by precipitation with ethanol (96%). The experiments were carried out using different HCl volumes (35, 65, 105, and 130 mL), acid washing times of 30, 60, and 180 min and ethanol:extract ratios of 0.5, 1, and 1.5. The maximum pectin recovery yield was 9.94 ± 0.2%, obtained using 65 mL of HCl, the extraction yield of 1.5, and the acid washing time of 120 min.

Mezzomo et al. (2010) recovered the oily fraction from peach seed (also called peach almond) by different methods: Soxhlet with different solvents (n-hexane, dichloromethane, ethyl acetate and ethanol), hydrodistillation, and ethanol maceration followed by partitioning with the solvents n-hexane, dichloromethane, ethyl acetate and ethanol. The Supercritical Fluid Extraction (SFE) was conducted, to compare with the traditional low-pressure methods, at the conditions of 30, 40 and 50 °C and at 100, 200 and 300 bar, carried out with CO₂ and ethanol as co-solvent in concentrations of 2% and 5% (w/w). The Soxhlet with ethanol provided higher yield values compared to SFE. The main fatty acids detected from the oily fraction were oleic and linoleic acids, particularly for the samples recovered by Soxhlet with ethanol: water mixture (50:50 v/v) and SFE with 5% ethanol at 50 °C/300 bar. The authors concluded that peach seeds are an important source of edible oil with high content of unsaturated fatty acids that have health benefits.

Peach seeds were used by González-García et al. (2016) for the recovery of protein. The seeds were first defatted with hexane and washed with MeOH/H₂O and acetone/H₂O. The extraction, using a native buffer and a denatured buffer as solvents, was conducted by applying sonication for 10 min, followed by gentle shaking overnight. The recovered protein was precipitated using acetone at 1:2 ratio (extract:acetone), and stored in freezer overnight. The method of Combinatorial Peptide Ligand Libraries (CPLLs) aids the reduction of the dynamic range of protein concentration, contributing to identify higher number of proteins compared to conventional methods. The results from CPLLs enable the identification of 97 unique genetic products from peach seeds. In addition, 14 bioactive peptides were also detected within the protein sequence from peach seeds. The results from this work suggest that the bioactive peptides recovered from peach seeds are valuable and worth investigating, as proteins are important for human nutrition and present high added value.

Several extraction methods have been studied to obtain different valuable fractions from peach by-products. These procedures are summarized in Table 3, which contains the extraction methods applied to peach by-products, the solvents and operating conditions used, and the main results obtained. The data from Table 3 represent the most recent publications on this subject. However, these available works have not explored the concept of biorefinery applied to peach by-products, nor discussed the food design approach to value the recovered fractions from peach by-products or residues. These relevant aspects, addressed to improve the peach processing chain, are presented in the next sections.

**Biorefinery Concept for the Processing of Peach By-Products**

The biorefinery concept arises from the need to reduce the existing dependence on fossil-based resources to cover the rising demand for energy, fuels, chemicals, polymers, and oil (Torre et al., 2019). Biomass and food by-products are also considered a potential source of high-added value chemicals such as fermentable sugar, furfural, citric acid, pectin, proteins, lipids, phenolic compounds, among other substances (Maciel-Silva et al., 2019; Ortiz-Sanchez et al., 2021). The food by-products are abundant, renewable and inexpensive bio-residues, or biomasses, which can be fully processed to obtain valuable bioactive compounds, as an interesting alternative from economic and environmental aspects (Gullón et al., 2020). Besides, the adequate processes for the recovery of these valuable compounds, allied with the appropriate pre-treatments to modify the cellulose, lignin, and hemicellulose, converting the lignocellulosic biomasses, are of utmost relevance, contributing to efficient conversion of underestimated by-products, such as peach residues (Manara et al., 2014). These methods can be applied, individually or combined, in a biorefinery approach, preferably represented by ecological and sustainable processes to attend the Sustainable Development Goals (SDG) of the United Nations (UN). The growing interest in this area expands the application of new technologies, enabling the development of innovative products and businesses, expanding the portfolio of the existing companies, and contributing to achieving new markets.

Newest approaches have been showing the use of a deep eutectic solvent (DES) as a relevant alternative for the recovery of valuable phytochemicals from food products (Benvenuti et al., 2019). Şahin and Bilgin (2022) used a lactic acid/glycerol mixture (1/1) and the addition of water (10, 30 and 50% v/v) to recover bioactive compounds from the peach peel. The results showed that with the use of DES with the addition of 50% water, at the optimized condition of 90 s and 8400 rpm, the recovered extract presented a polyphenol content of 10.68 mg-GAE/g-dried and chlorogenic acid 2.69 mg/g-dried. Skiba and Vorobyova (2022) used a green solvent named NADES (natural deep eutectic solvent) composed of lactic acid, glucose, and water (5:1:3) to obtain phenolic compounds from peach pomace, and the main identified compounds were chlorogenic acid (1.04 mg/g extract), gallic acid (1.04 mg/g extract), and ellagic acid (0.54 mg/g extract). These results indicate the potential of NADES as a green solvent for the recovery of phenolic compounds from peach pomace.
Table 3  Summarized data from extraction procedures applied for peach by-products: methods, solvents, operational conditions; main results

| Source        | Extraction                     | Solvent                           | Extraction parameters                                                                 | Compound                                      | References            |
|---------------|--------------------------------|-----------------------------------|----------------------------------------------------------------------------------------|-----------------------------------------------|-----------------------|
| Maceration    | Methanol and acetone           | One gram of sample and 20 mL of acidic methanol–water (50:50, v/v, pH 2) at room temperature/1 h. 20 mL of 70% aqueous acetone added to the residue, followed by stirring, shaking and centrifugation | Phenolic compounds, flavonoid and L-ascorbic acid and antioxidant assay | Liu et al. (2015)                   |
| Ultrasound    | HCl concentrations (0, 0.1, 1 or 2%) | 30 mL of 80% (v/v) acetone solution/60 min | Polyphenolic compounds | Stojanovic et al. (2016) |
| Ultrasound    | HCl concentration 0.01%        | Independent variables: potency range (10–50%), sample mass (0.1–0.5 g) and aqueous ethanol (EtOH) concentration (20–80%, v/v) and fixed 15 min | Total phenolic content, anthocyanin, p-hydroxybenzoic acid and p-coumaric acid | Karrulbaş and Şahin (2022) |
| Peel          | Homogenizer-assisted extraction | Deep eutectic solvent (lactic acid/glycerol mixture (1/1)) | Independent variables: water (10, 30, and 50%, v/v), time (30, 60, and 90 s), speed (6000, 8000, and 10,000 rpm) | Total phenolic content and chlorogenic acid | Şahin and Mehmet (2022) |
| Microwave     | HCl 0.01% (v/v)                | Independent variables: Power (350–500 W), time (30–90 s) and aqueous ethanol (EtOH) concentration (20–80%, v/v) | Total phenolics content, total anthocyanins content, p-hydroxybenzoic acid and p-coumaric acid | Kurtulbaş et al. (2022) |
| Microwave     | With 5 mL of 5% HPO₃; 10 mL MeOH/ H₂O/formic acid (60:38:2 v/v/v); 10 mL water | Adding 5 mL of 5%; (60:38:2 v/v/v); 10 mL | Ascorbic acid, total phenolics content, flavonoids, polyphenols and antioxidant capacity and sugar | Saidani et al. (2017) |
| Maceration    | 1 mL + 1 mL 95% EtOH centrifuged 4500 RPM 10 min diluted with 50% EtOH | Extracted at 25 °C by ethanol (absolute, ≥ 99.5%) (400, 1500 and 250 mL) (3 × 72 h) | Total phenol, flavonoids and carotenoids | Loizzo et al. (2015) |
| Extraction    | Petroleum ether, chloroform, ethyl acetate, ethanol, and water | 60 cycles of siphoning, completed with each solvent and extraction continued until siphon tube become colorless | Alkaloids, carbohydrate, glycosides, inulin, protein, steroids/triterpenoids, fixed oils and fats, flavonoids, phenolic, compound and tannins, gums and mucilage | Shukla and Kant (2020) |
| Seed          | Sonication (10 min)            | MeOH/H₂O₃ (80:20), acetone/H₂O (80:20) | Extract/aceton ratio of 1.2 and centrifuged (30 min, 13,400 rpm) | Bioactive peptides | González-García et al. (2016) |
| Ultrasound    | 5 mL of extraction buffer (100 mM Tris–HCl (pH 7.5) | 1 min at 30% of amplitude, after centrifugation at 4000 g/10 min, Supernatant collected, added to 10 mL of cold acetone, and kept in the fridge for 15 min | Peptides | Vásquez-Villanueva et al. (2015) |
| Pomace        | Ethanol:water (70:30)          | Optimal MAE (540 W, 50 s) and UAE (23%, 120 s) | Polyphenols, flavonoids, anthocyanins, and vitamin C | Plazzotta et al. (2020) |
| Pomace        | Ethanol:water (70:30)          | PEF at 50 °C and 90 min and PP dispersions were thus treated at 0.8–10 kV/cm, using 4–30 monopolar pulses of 4 μs | Bioactive extracts | Plazzotta et al. (2021) |

The data from Table 3 comprises recent publications from 2015-2021.
extract), and ferulic acid (0.115 mg/g extract). This extract was compared with plasma-assisted extraction with water, which provided higher antioxidant activity in terms of 50% inhibition concentration (IC50 of 1.05 mg/mL) compared to the extract recovered by NADES (IC50 of 1.3 mg/mL). Mármol et al. (2021) evaluated the extraction of bioactive compounds from peach pomace using ethanol: water (w/w) ratio (70:30) as a solvent, and followed by homogenization at 1600 rpm for 2 min. The results showed that the most representative phenolic component recovered was the ellagic acid (28.26 mg/100 g).

These works indicate that peach presents interesting phytochemical compounds that were detected in peach peel and pomace. Using green solvents at different extraction methods is an alternative to promote the valorization of these biomasses, which is in line with the biorefinery concept. Nevertheless, the optimization of a selected extraction procedure, preferably performed on a laboratory scale, is high relevance for the industrial processing of agro-industrial by-products (Martins et al., 2020).

In Brazil, there is a huge generation of residual biomasses, mostly agro-industrial residues or by-products, which has not yet received much attention as a direct energy source, such as biodiesel. Biodiesel is a fuel obtained from vegetable oils, animal fats, or residual vegetable oils (Longati et al., 2020). According to Ribeiro et al. (2018), the highest concentration of biodiesel processing plants in Brazil is located in the South region, and the most used raw materials are soybean oil (70%), beef fat (17%), and used frying oil (1%). These examples show the limited use of other biomasses, although several studies have been indicating other abundant and renewable plant-based biomasses as valuable alternatives for the application of the integrated biorefinery concept. Among these commodities and important fruits, we can mention: soy, orange, corn, tamarind and mango (Longati et al., 2020; Ma et al., 2021; Martins et al., 2020; Monteiro et al., 2021; Ortiz-Sanchez et al., 2021). As already mentioned, because peach is an important worldwide commodity, some researchers have already proposed the use of peach residues as a source of high-value products. However, there is a lack of studies related to the use of integrated paths, in a biorefinery concept, for the valorization of this residue. Therefore, besides biofuel production, the biorefinery concept involves biomass conversion to produce biofuels, energy, and also valuable chemicals (Cherubini, 2010). Consequently, following the concept of circular bioeconomy (principles related to reuse, reduction and recycling), the process integration, allows the recovery of different products from the original biomass and also improves sustainability. Yet, industrial units still have a considerable challenge in combining these two concepts.

### Bioactivities Associated to Peach By-Products

Most literature studies about peach by-products are related to the bioactive compounds and antioxidant activity associated with peel, pomace, seeds, and seed shell. To justify the significance of these peach by-products, as sources of biological active components, the most recent and relevant studies concerning this subject are discussed below, highlighting biological activities such as antimicrobial, antioxidant, antidiabetic, anti-obesity, anti-inflammatory, and anti-cerebral ischemia.

### Antioxidant Activity

Hong et al. (2021) evaluated the antioxidant activity and the phenolics content of the ethanolic extract (70%, 20 mL) from the shell of the peach seed. The results show the total phenolics content (TPC) of 0.47 (mg gallic acid equivalents (GAE)/g), total flavonoids content (TFC) of 0.18 (mg quercetin equivalents (QE)/g), total tannins content (TTC) of 0.07 (mg catechin equivalents (CE)/g), while the antioxidant activity by 2,2-Diphenyl-1-picrylhydrazyl antioxidant assay (DPPH) was 0.98 (mg ascorbic acid equivalents (AAE)/g), while by ferric reducing-antioxidant power (FRAP) assay was 0.54 (mg AAE/g), and by 2,2′-Azino-bis-(3-ethylbenzothiazoline-6-sulfonic acid (ABTS) assay was 0.43 (mg AAE/g), with total antioxidant capacity (TAC) of 0.27 (mg AAE/g), estimated by phosphomolybdate method. According to Hong et al. (2021), these results show higher radical scavenging capacity compared to other seed shells such as nectarine (0.23 mg AAE/g), plum (0.21 mg AAE/g), and apricot (0.25 mg AAE/g).

Another relevant study, by Nowicka and Wojdyło (2019), shows that the total polyphenols content of peach seeds, from 20 different cultivars, ranged from 3.8 to 12.7 g/100 dry matter, while the cyanogenic glycoside content ranged from 17.4 to 245.7 mg/100 of dry matter. The main identified polyphenols were from Flavan-3-ols group, (+)-catechin, (−)-epicatechin, (−)-epicatechin gallate; and from Polymeric procyanidins group (B1, B2 and procyanidin A2). The main phenolic acids were Hydroxycinnamic acids (chlorogenic acid, neochlorogenic acid, 3-O-p-coumaryloquinic acid and cis-5-p-coumárolyquinic acid, 2-O-cafeoyl-L-malate, phenylpropanoid o-diphenol phaselic acid) and Hydroxybenzoic acids (ellagic acid, dilactone of hexahydroxydiphenic acid). The peach seeds were also characterized by high antioxidative potential and the ability to inhibit enzymes linked to obesity, type 2 diabetes and Alzheimer’s disease. Generally, confirmed that peach seed could be a valuable source of polyphenols used by the pharmaceutical industry.
Redondo et al. (2021) detected and quantified the polyphenols and evaluated the antioxidant activity from peach pulp, peel and seeds, from fruits in commercial maturity. The evaluated extracts were recovered by methanol: water solution (80:20; v:v). The results from peel extract showed the presence of quercetin 6.2 mg/100 g, chlorogenic acid 3.9 mg/100 g, and neochlorogenic acid 7.1 mg/100 g, while from seed extract, the detected and quantified compounds were quercetin (5.5 mg/100 g), chlorogenic acid (4.0 mg/100 g), neochlorogenic acid (4.9 mg/100 g), and ferulic acid (5.7 mg/100 g). As for the pulp, the results detected were quercetin (5.4 mg/100 g), chlorogenic acid (1.2 mg/100 g), and neochlorogenic acid (4.8 mg/100 g). These values suggest that the peach seeds present relevant content of phenolic compounds compared to other fruit parts, and the antioxidant potential provided by the seed extracts confirm this behavior. For instance, the results by FRAP method provided the highest antioxidant capacity for peach seed (3.3 mmol Trolox/100 FW), while peach peel was 2.2 mmol Trolox/100 FW and pulp 0.2 mmol Trolox/100 FW. However, it is essential to note that the values vary with the conditions used to recover the extracts, such as type of solvent, extraction technique, fruit maturity, and harvest conditions (soil, geographical aspects and plant variety). The study by Redondo et al. (2021) reveals the potential of peach by-products as a source of bioactive compounds with different potential uses, such as in pharmaceutical, cosmetic, food, or animal feed formulations. It is also important to highlight that the peach pomace contains pulp and peel. El Darra et al. (2018) recovered polyphenols from peach pomace by solid–liquid ratio 1:10 (w:v) extraction using β-cyclodextrin and ethanol as solvent, and identified the components gallic acid, caffeic acid, and protocatechin from the recovered extract. The mixture of β cyclodextrin and ethanol selectively enhanced the extraction of gallic and caffeic acids with yields 220 and 328 µg/g dry matter, respectively, while ethanol was more selective to protocatechic (4899 µg/g dry matter).

Mokrani and Madani (2016) investigated the effect of the following extraction parameters (solid/liquid) of peach fruit: (Prunus persica L.): solvent type (ethanol, methanol, acetone and water), time (30–450 min), and temperature (25–70 °C). The recovered extracts were evaluated in terms of total phenolics content (TPC), total flavonoids content (TFC), and antioxidant potential. Optimized extraction condition provided a peach extract with high values of TPC, TFC, DPPH radical scavenging activity and FRAP activity, referent of 363 mg GAE/100 g, 81 mg QE/100 g, 48% inhibition percentage and 317 mg AAE/100 g, respectively. With these results, and comparing with data presented by Hong et al. (2021), Nowicka and Wojdylo (2019) and by Redondo et al. (2021), it is possible to highlight that peach by-products have relevant bioactive compounds, comparable to the edible part of the fruit. Thus, further studies should be carried out to enhance the use of these by-products as a source of relevant bioactive compounds, and add value to these biomasses.

Antimicrobial Activity

Patra and Baek (2016) evaluated the biological synthesis of gold nanoparticles (AuNPs) generated from peach peel aqueous extract. The extract showed strong antibacterial synergistic activity when combined with the positive control substances kanamycin (inhibition zone from 9.38 to 20.45 mm) and rifampicin (inhibition zone from 9.52 to 25.23 mm), and also synergistic anticandidal activity when combined with amphotericin B (inhibition zone from 10.09 to 15.47 mm) for five pathogenic Candida species. In addition, the extract presented strong antioxidant potential by elimination of 1,1-diphenyl-2-picrilhidroxil radical, nitric oxide, 2,2′-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) radical. Although the significant antimicrobial potential detected for peach peel, there is a lack of information related to this important bioactivity associated with peach by-products, which should be further investigated.

Antidiabetic and Anti-obesity

Kan et al. (2020) investigated the influence of peach peel phenolic extract in preventing lipid accumulation and intestinal microbiota composition of mice with high-fat diet-induced obesity. Extracts were obtained with methanol by ultrasound technique (400 W, 40 °C) for 3 h. The recovered extracts showed that peach peel significantly inhibited lipid accumulation in mice fed with a high-fat diet. Meanwhile, treatment with peach peel phenolic extract regulated the expression levels of mRNA involved in lipid metabolism. Furthermore, the results showed that the extract regulated the intestinal microbiota composition, increasing the relative abundance of Lactobacillus, Bacteroides, Lachnospiraceae, Prevotellaceae Alloprevotella, Akkermansia, Roseburia, and Ruminococcus. This effect might be attributed to the phenolic content and composition of the peach peel extracts, showing the relevance of this by-product to provide a functional food additive to improve lipid metabolic abnormalities associated with obesity. Based on what has been discussed so far, few studies have evaluated the potential of by-products activities. Thus, some studies are still focused on the edible part of the fruit. For instance, Nowicka et al. (2018) provided important insights on properties of the extract from the pulp of Prunus persica L. Batsch, obtained by sonication with methanol:H₂O (80:20%, v/v) + 1% HCl at 20 °C for 15 min. Results from in vitro analysis show the potential to inhibit enzymes relevant for the management of type 2 diabetes (α-amylase, α-glucosidase) and obesity (pancreatic lipase). The yellow peach extract, with high carotenoids content.
(varieties Harrow Diamond and Harrow Beauty), showed high inhibitory activity towards porcine pancreatic lipase.

Sharma et al. (2018) evaluated the antidiabetic activity of extract from leaves of *Prunus persica*. The extract prepared by percolation with 90% ethanol at room temperature was concentrated under reduced pressure at 50–55 °C and used at in vivo assays with adult male rats. The results show that 200 mg extract/kg provided significant anti-hyperglycemic activity. Improvement in rat body weight and lipid profile was also observed by streptozotocin induced diabetic rat model. The extract good performance suggests its possible use as a natural drug candidate for diabetes mellitus treatment.

Song et al. (2019) investigated the anti-obesity effect of extracts from flowers of *Prunus persica* L. Batsch, obtained with distilled water at 100 °C for 2 h under reflux. The results from in vivo assays using peach flower extract show a significant reduction in body weight, abdominal fat mass, serum glucose, alanine transaminase and aspartate aminotransferase levels, and liver and spleen weights from mice. Besides, the extract also reduced the levels of glucose, cholesterol, alanine aminotransferase (ALT), aspartate aminotransferase (AST), and leptin, while the levels of AST/ALT and adiponectin/leptin ratios and adiponectin were increased. The results suggest an anti-obesity effect of the extract in obese mice, which may improve the hepatic lipid metabolism by reducing lipogenesis and increasing fatty acid oxidation.

Wang et al. (2017) evaluated the antidiabetic activity potential of peach gum polysaccharides, from peach tree trunks and fruits. The peach gum exudate was dissolved in boiling water at a ratio of 1:100 (w/v), and filtered to obtain the supernatant solution. The concentrated solution was precipitated overnight in four folds of ethanol, for purification, and the resulting polysaccharides were dried. The peach gum polysaccharide reduced significantly (42.76%) the postprandial blood glucose in streptozotocin-induced diabetic mice. Histology and immunohistochemistry results confirmed that the polysaccharide, consisting of arabinose, xylose, and galactose, partially restored the pancreatic islets in diabetic mice, and enhanced the expression of pancreatic duodenal homeobox-1, insulin and hexokinase-1. The results may suggest the peach gum-derived polysaccharide as a relevant non-insulin therapy for diabetes treatment, probably due to the presence of the detected compounds.

**Anti-inflammatory Potential**

Elshamy et al. (2019) prepared seed extract from nectarine (*Prunus persica*, variety *nectarina*) by maceration with methanol 70%, a fruit from the same family as peach. The acute anti-inflammatory effect of the extract was evaluated by a carrageenan-induced rat hind paw edema test. The nectarine seed extract was administrated in rats at doses of 50 and 100 mg/kg reducing significantly the paw edema by 11 and 47%, respectively. The results suggest that alcoholic nectarine seed extract is a safe natural anti-inflammatory, providing analgesic activity through central and peripheral mechanisms. Furthermore, the authors evaluated the content of total phenols (55.91 ± 5.78 mg/g) and flavonoids (29.89 ± 0.55 mg/g), indicating that the extract is a promising source of these constituents, which are possibly related to the anti-inflammatory effect.

Another interesting study concerning the edible part of the fruit was conducted by Kim et al. (2017) which prepared white-flesh peach extract (WFPE) using ethanol 80% for 48 h at room temperature. The study evaluated the effect of the extract on the excretion of nicotine and 1-hydroxypyrene metabolites in smokers and chronic nicotine-induced tissue damage in mice. The WFPE inhibited nitrotyrosine expression and inflammatory responses in liver, kidney, and lung tissues of nicotine-treated mice. The results showed that WFPE increased the metabolism of toxic smoke components of tobacco in smokers and protected normal tissues against nicotine toxicity in mice. Although this research has been conducted for peach edible parts, and no anti-inflammatory studies were conducted for peach by-products, the peach flesh is also part of the pomace, therefore, some of these activities could be also associated to the pomace (by-products), and should be further investigated to confirm this active behavior.

Seo et al. (2020) evaluated the anti-inflammatory potential of the methanolic extract from *P. persica* on lipopolysaccharide stimulated glial cells. Glial cells are immune cells from the central nervous system, with a key role in numerous neurodegenerative diseases, including Alzheimer’s, Parkinson’s and Huntington’s disease. We used different parts of the peach fruit (leaves, fruits, and twigs) to obtain the methanolic extracts, prepare by sonication (1,500 W, 40 kHz) frequency for 15 min at 2 h intervals for 3 days. The results indicated that the combined methanolic extracts reduced the transcription of several pro-inflammatory enzymes (nitric oxide synthase and cyclooxygenase-2) and cytokines (tumor necrosis factor-α, interleukin (IL)-1β and IL-6) in lipopolysaccharide-stimulated cells. Furthermore, the extract inhibited the activation of nuclear factors and several mitogen-activated protein kinases necessary for the transcription of the pro-inflammatory mediator. Therefore, it can be used as a potential therapeutic agent for neurodegenerative diseases and neurotoxicity by suppressing glial cell activation. The evaluated extract presented chlorogenic acid, catechin, quercetin, and quercetin-3-O-glycoside, and the anti-inflammatory effect on PPB on BV2 cells may be due to the presence of phenolics and flavonols.
Cerebral Ischemia

Li et al. (2020) studied aqueous extract mixtures from peach seeds and rhubarb (Rheum palmatum L) rhizome, and its effect on cerebral ischemia/reperfusion injury (I/R), using different mixture proportions (peach seeds: rhubarb) (m/v), 1:0, 2:3, 1:1, 3:2 and 0:1. The results demonstrate that the extracts effectively reduced the neurobehavioral defect scores, the areas of cerebral infarction caused by cerebral ischemia and pathological brain tissue injury, and inhibited abnormal increases in inflammatory factors. Also, the extracts alleviated the I/R damage by altering the uncontrolled expression of the adenosine protein (ADORA2A). The extract that provided the best regulatory effect was obtained from a 1:1 ratio of peach seed: dried rhubarb root.

Other Applications

Different applications from peach by-products have been studied and presented in the literature. For instance, applications in pharmaceuticals, cosmetics, food products, and energy industries were discussed. Fratebianchi et al. (2017) used peach pomace as a source of carbon and energy to produce polygalacturonase in submerged cultures using Aspergillus Sojae. Papaioannou and Liakopoulou-Kyriakides (2012) studied peach peels as carbon source for Blakeslea trispora culture for carotenoid production. The results indicate 76% of β-carotene production, and this high performance suggests the use of this residue as substrate culture for carotenoids production on industrial scale.

Li et al. (2018) evaluated the chemical characteristics of peach seed shells, the sugar yields, and the impact of Deep Eutectic Solvent (DES), formed by choline chloride 1:2: lactic acid, as biomass (seed shell) pre-treatment, affecting physical and chemical properties and lignin extraction. The results from this biomass, compared to typical woody and herbaceous biomass, exhibit significantly higher bulk density and hardness. Enzymatic saccharification of peach seed shells treated with DES provided a high glucose yield using Aspergillus Sojae. The results indicate 76% of β-carotene production, and this high performance suggests the use of this residue as substrate culture for carotenoids production on industrial scale.

Product Design: Functional Food Product

Functional foods are natural or industrially processed food products that, when regularly consumed at satisfactory levels in a diversified diet, have positive health effects beyond basic nutrition (Granato et al., 2020). The Functional Food Center (FFC) from USA (Texas, USA) is committed to leading research on functional foods as well as bringing the functional foods to the mainstream worldwide food markets, and introducing a new approach for the definition of functional foods (Martirosyan & Singh, 2015). This new concept describes functional foods as products containing biologically active compounds in effective and non-toxic quantities, derived from natural or processed food sources, and clinically tested, based on specific biomarkers, providing health benefits for prevention, management or treatment of chronic diseases or their symptoms (Alongi & Anese, 2021). Therefore, functional foods must be regulated by the Food and Drug Administration (FDA) before approval, and are suitable if they are constituted by compounds Generally Recognized as Safe (GRAS).

Given the above assumptions about functional foods, the increase in various types of chronic diseases has motivated the food industry to develop food products that help consumers to prevent these diseases and provide other health benefits (Alongi & Anese, 2021). According to Micheli et al. (2019), the design thinking concept could be utilized in different areas including public policy, education, healthcare, politics, and social and economic development. This concept has emerged in sustainability literature to generate positive economic, social, and environmental impacts, in association with business transition, food innovation, and applied business innovation (Massari et al., 2021). Meanwhile, the use of design thinking, double diamond, and multi-criteria reverse engineering methodologies (Fig. 4) is fundamental to reach the consumer’s new needs, and also to elucidate the industrial challenges for the use of by-products as a source of bioactive and functional compounds. Therefore, the health benefits from peach by-products (discussed previously) may suggest its industrial use to provide functional foods. Although, the long way that has to be crossed to produce these new products can be facilitated by the product design methodology, as discussed below.
The design thinking method is based on solving problems and has become an important approach for food design. According to Olsen (2015), this methodology can contribute to food innovation using empathy, visualization, rapid prototyping, and collaboration (Fig. 4). According to Tkaczewska et al. (2021), the method to create products must be economically and technologically viable, following the aspects: (I) observation and synthesis, (II) visualization and rapid prototyping, and (III) revising and refining. For Batat and Addis (2021), the design thinking concept is a high contribution to food innovation, when designing sustainable food products and services that respond to the needs of consumers.

Another methodology proposed for food design is Multi-Criteria Reverse Engineering (MCRE). This technique simultaneously considers several criteria of the product’s properties, increasing the complexity of the problem. For instance, MCRE enables the selection of adequate process conditions for designing or redesigning food, food processes, and food-related systems (Thomopoulos et al., 2019). According to Dima et al. (2020), the MCRE as a design method for functional foods considers the following aspects: to develop a nutritional, sensorial, and functional evaluation of the food product, attending the consumer’s needs; to select target bioactive components, its nutraceuticals distribution systems, encapsulation technique, food matrix, or the excipient; to choose the packaging method, analyze costs, impacts on the environment; to analyze the perception of the food product, and, finally, to test the bioavailability of nutraceuticals incorporated into the food product (Fig. 4); therefore, combining several specific criteria for functional food design.

The Double Diamond model, another product design methodology also known as 4D (Sijtsema et al., 2020), consists of four steps: discover, define, develop, and deliver (Fig. 4). The first step (Discover) is essential to establish new opportunities, markets and processes, and is performed with the aid of literature data. The “Define” step is fundamental to select insights to demarcate and redefine the study topic, accompanied by the definition of the adequate technologies, ingredients, and packages. The third step is to “Develop” the design refine it, and teste it with multidisciplinary teams, which involves developing samples/concepts with different processing, ingredients, packaging, target groups, and sensory research. The last step, “Deliver”, is the final prototype (food design), packaging and distribution, which are tested, produced and launched.

These models can help the product design by combining different tools that can be a solution to obtain innovative products aligned with sustainability, reducing agro-industrial by-products or improving their use and value. Therefore, based on what has been discussed so far, peach

Fig. 4 Methodologies design thinking, double diamond, and multi-criteria reverse engineering approach in food design. Source: the author
by-products (pomace, peel, seed shell, and seed) are relevant biomasses with high application potential (components with valuable bioactivities associated). For instance, the recovered products could be used in food, pharmaceutical, and cosmetic formulations. Besides, the use of these bioactive compounds implies in several processing steps, such as recovery, purification, isolation, identification and protection. Moreover, it is necessary to evaluate the production chain and the different sectors, such as logistics, transport, and packaging, to direct their by-products for further processing. For instance, by-product processing consists of drying, milling, extraction, fractionation, and others, which provide different products that must be evaluated in terms of quality and processing yield (efficiency). Therefore, the “product design” may provide the adequate feasibility for the definition of the “new process”.

Because of the strong relevance of the 17 SDG from ONU, there is a growing search for greener processes and protocols to obtain bioactive substances from industrial food by-product. To achieve these objectives, it is still necessary to study the extraction methods capable of reducing solvent and energy consumption and, also increasing the overall eco-sustainability of the food life cycle.

Some existing business models can serve as inspiration. For instance, the company Rubian Extracts, a Brazilian startup, recently developed natural extracts by supercritical CO2 technology, recovering biocompounds from by-products. Combining biorefinery and circular economy concepts is the key for the sustainable recovery of by-products, for application as food ingredients, with functional properties. Therefore, governmental policies may incentive partnerships between private companies, and researchers as a fast solution for food-product design, seeking benefits for society.

Conclusion and Perspectives

Peach processing by-products (pomace, seeds, and seed shells) are valuable renewable biomasses due to their content in lipids, proteins, phenolic compounds, fibers, and relevant bioactive phytochemicals including neochlorogenic acid, gallic acid, caffeic acid, and procyanidin, which are associated to bioactivities such as anti-inflammatory, anti-obesity, and anti-cerebral ischemia. The recovery of these valuable phytochemicals from peach by-products by using emerging green technologies (Pulsed electric field, supercritical fluid extraction, microwave-assisted extraction, and ultrasound-assisted extraction) may contribute to the development of new products for food or cosmetic industries. The valorization of peach by-products represents an opportunity to develop food products by combining the biorefinery concept with methodologies of design thinking, double diamond, and multi-criteria reverse engineering. These methodologies may improve the peach processing chain by optimizing the use of underestimated by-products, contributing to a sustainable industry. Therefore, peach by-products are promising sources of several molecules of high value, which contribute to rise the interest in these biomasses.

Author Contribution Carla Roana Monteiro Rudke (carla.roana@posgr ad.ufsc.br) conceived and structured the review. Acácio Antonio Ferreira Zielinski (acacio.zielinski@ufsc.br) and Sandra Regina Salvador Ferreira (s.ferreira@ufsc.br) contributed to the conception of the work, supervision, revising, and editing the manuscript.

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Data Availability All data sharing and material are results from research and properly referenced along with the manuscript.

Declarations

Competing Interests The authors declare no competing interests.

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