TBX5-AS1, an enhancer RNA, is a potential novel prognostic biomarker for lung adenocarcinoma

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Abstract

Background: Enhancer RNAs (eRNAs) are demonstrated to be closely associated with tumourigenesis and cancer progression. However, the role of eRNAs in lung adenocarcinoma (LUAD) remains largely unclear. Thus, a comprehensive analysis was constructed to identify the key eRNAs, and to explore the clinical utility of the identified eRNAs in LUAD.

Methods: First, LUAD expression profile data from the Cancer Genome Atlas (TCGA) dataset and eRNA-relevant information were integrated for Kaplan-Meier survival analysis and Spearman's correlation analysis to filter the key candidate eRNAs that was associated with survival rate and their target genes in LUAD. Then, the key eRNA was selected for subsequent clinical correlation analysis. KEGG pathway enrichment analyses were undertaken to explore the potential signaling pathways of the key eRNA. Data from the human protein atlas (HPA) database were used to validate the outcomes and the quantitative real time-polymerase chain reaction (qRT-PCR) analysis was conducted to measure eRNA expression levels in tumour tissues and paired normal adjacent tissues from LUAD patients. Finally, the eRNAs were validated in pan-cancer.

Results: As a result, TBX5-AS1 was identified as the key eRNA, which has T-box transcription factor 5 (TBX5) as its regulatory target. KEGG analysis indicated that TBX5-AS1 may exert a vital role via the PI3K/AKT pathway. Additionally, the qRT-PCR results and the HPA database indicated that TBX5-AS1 and TBX5 were significantly downregulated in tumour samples compared to matched-adjacent pairs. The pan-cancer validation results showed that TBX5-AS1 was associated with survival in four tumours, namely, adrenocortical carcinoma (ACC), LUAD, lung squamous cell carcinoma (LUSC), and uterine corpus endometrial carcinoma (UCEC). Correlations were found between TBX5-AS1 and its target gene, TBX5, in 26 tumour types.

Conclusion: Collectively, we supposed that TBX5-AS1 is a tumor suppressor, and could be a new prognostic biomarker for LUAD patients and promote the targeted therapy of LUAD.

Background

Lung cancer is recognized as one of the most common malignant carcinomas, and ranks first for morbidity and mortality among all malignant tumours in the world. Nonsmall cell lung cancer (NSCLC) is the most frequently diagnosed type of lung cancer, and comprises two major types—adenocarcinoma and squamous carcinoma (1,2). Because the early symptoms of lung cancer aren't clear and are confused easily with those of other respiratory diseases, the rates of misdiagnosis and missed diagnosis are high, and the majority of patients are diagnosed at an intermediate or late stage, at which the effective treatment options are limited and the prognosis is usually poor. In recent years, with the development of gene detection technology and targeted therapies, great strides have been made in the treatment and prognosis of LUAD. However, the 5-year overall survival is still less than 20% (3,4), highlighting the urgent need to identify effective prognosis-related biomarkers for LUAD; evaluating the
prognosis of LUAD patients through biomarkers in the early stage of the disease is of great importance to improving the prognosis and survival rate of these patients.

Over the past decade, the rapid application and development of high-throughput sequencing technology has opened a new window for noncoding RNAs, and accumulating evidence has shown that the occurrence and development of a series of major diseases are related to noncoding RNA dysregulation (5-7). Among these, RNAs generated from enhancers (enhancer RNAs; eRNAs) have gained substantial interest owing to their underlying roles in the mediation of gene transcription and enhancer functions, and their frequent overlap with noncoding risk loci associated with diseases (8-11). Enhancers are key cis-regulatory elements that promote the expression of eukaryotic genes (12,13). eRNAs are 500–5,000 bp-long RNAs produced through enhancer transcription (14). They can activate enhancer activity independently, and can also combine with other protein factors to promote the formation of the enhancer-promoter loop, thereby activating the expression of downstream genes (15-17). eRNA dysregulation can affect biological processes such as the cell cycle and the growth of cancer cells, or change the expression of target genes, suggesting that eRNAs may serve as novel targets for tumour treatment (15,18,19). A recent study found that, mutations in the eRNA element of ACTRT1 in basal cell carcinoma can weaken the enhancer's activity as well as the expression of ACTRT1, resulting in the aberrant activation of Hedgehog signaling and, consequently, the promotion of carcinogenesis and tumour development (20). In T-cell acute lymphoblastic leukemia (T-ALL), it was also reported that the eRNA ARIEL could recruit mediator proteins to the ARID5B enhancer, thereby boosting enhancer-promoter interactions and activating the expression of ARID5B. These activities led to the aberrant activation of the TAL1-induced transcriptional program and myc oncogenic signals (21). NET1 is an oncogene of key importance in breast cancer. Related research has demonstrated that the NET1-associated eRNA (NET1e) could upregulate the expression of its target gene and promote breast cancer progression. In addition, overexpression of NET1e in situ can lead to the resistance of breast cancer cells to BEZ235 and octreotide (22). Additionally, a separate research discovered an eRNA, AP001056.1, is able to serve to be a prognostic marker for head and neck squamous cell carcinoma (HNSCC) (23). However, to date, no study has investigated the potential of using eRNAs as biomarkers to predict the survival of patients with LUAD.

Herein, we aimed at identifying prognostic eRNAs as well as their target genes within LUAD for predicting LUAD patients' prognosis. Our results may facilitate the understanding of the potential value of the clinical application of LUAD-associated eRNAs in prognosis stratification and promote the targeted therapy of LUAD.

**Materials And Methods**

**Identification of Predictive eRNAs in LUAD by Comprehensive Data Analysis**

Clinical data, RNA expression profile data, and survival information for 33 cancer types were downloaded from the TCGA database. All RNA expression profile data were transformed to log2(FPKM+1). Ensemble transcript IDs were con-verted to their corresponding GENCODE using GTF annotation files for humans.
We searched and comprehensively analysed the eRNA source literature to obtain the relevant eRNA information. The human GTF files were used to transform eRNA transcript IDs into gene symbols and the expression profiles of the eRNAs were extracted from the RNA expression profile of lung adenocarcinomal. Then, we combined the eRNA expression matrix with the lung adenocarcinomal survival data using the limma R software package.

The survival-associated eRNAs were screened using Kaplan–Meier methodology. The patients were classified into low- and high-expression groups according to the median expression of each eRNA. Differences in survival were compared between the two groups using p<0.05 as the cut-off and a survival curve for the eRNAs that met the conditions was drawn. Then, Spearman's correlation analysis was performed to obtain candidate key eRNAs related to survival and associated with target genes with roles in LUAD; statistical significance was considered at cor >0.4 and p<0.001.

**Gene Enrichment Analysis**

Besides the predicted targets, Pearson's correlation analysis was performed to identify other genes in LUAD that were significantly associated with the key eRNA (cor >0.4, and p<0.001). The target genes of the key eRNA were converted into their corresponding gene IDs for further analysis using the org.Hs.eg.db R software package. To investigate which functions and pathways were affected by the eRNA in tumourigenesis and metastasis, the clusterProfiler package in R was used for the pathway enrichment analyses of Kyoto Encyclopedia of Genes and Genomes (KEGG) and Gene Ontology (GO) of the key eRNA target genes. Enriched GO terms with an adjusted p-value <0.001 were considered as relevant biological processes.

**Clinical LUAD Sample Collection**

A total of 10 LUAD tumor tissues and para-tumor samples were collected from patients diagnosed with LUAD according to pathological analysis and who were undergoing surgery at the Second Xiangya Hospital of Central South University. After tumor resection, the tumor and adjacent tissues were immediately frozen in liquid nitrogen and stored at -80°C until total RNA extraction. All patients gave their informed consent for inclusion before surgery. The study was conducted in accordance with the Declaration of Helsinki, and the protocol was approved by the Ethics Committee of the Second Xiangya Hospital of Central South University. (protocol code 2020038 and 07/01/2020)

**Quantitative Real-Time Polymerase Chain Reaction (qRT-PCR) Validation**

TRIzol reagent was used to extract total RNA from tissue samples (Invitrogen, GrandIsland, NY, USA). The quality and quantity of RNA were estimated by NanoDropND-1000 (Thermo Scientific, Waltham, MA, USA), while the integrity of RNA were assessed by standard denaturing agarose gel electrophoresis. Then, the RNA was reverse-transcribed into cDNA by adopting SuperScript III Reverse Transcriptase (Invitrogen) in accordance with the instructions of the manufacturer. A 2× PCR Master Mix (Arraystar) and an Applied Biosystems ViiA 7 Real-Time PCR System were adopted for qRT-PCR following the instructions of the
manufacturer. The relative eRNA and mRNA expression levels were calculated using the $2^{-\Delta\Delta Ct}$ method. The primers used and the sequences are listed in Table 1 and Table 2. The data represent the means of three experiments.

**Human Protein Atlas Analysis**

The human protein atlas (HPA; https://www.Proteinatlas.org/) comprises an atlas of human protein expression patterns in normal and tumour tissues. In this study, we assessed the expression of T-box transcription factor 5 (TBX5) in the protein expression module of the HPA database and analysed the immunohistochemical staining results for TBX5 in both tumour and normal tissues (antibody: HPA008786).

**Verification in Pan-Cancer**

First, the expression data of TBX5-AS1 and its target gene TBX5 in pan-cancer were obtained using the R limma package, and then the expression matrix was combined with the pan-cancer survival data. The samples were classified into low- and high-expression groups according to the median value of the expression of TBX5-AS1, and then the Kaplan–Meier method was used to compare the survival difference between the two groups. $p<0.05$ was considered statistically significant. A survival curve was plotted for TBX5-AS1 in tumours that met the criteria. Spearman's coefficient was used to test the correlation between TBX5-AS1 and its corresponding target genes TBX5 in pan-cancer. A correlation coefficient $>0.4$ and a $p$-value $< 0.001$ were considered statistically significant.

**Results**

**Screening the Key eRNA in LUAD**

A total of 174 survival-related eRNAs in LUAD ($p<0.05$; Additional file 1: Table S1) were identified through the Kaplan-Meier method. Spearman's correlation was then used to screen these 174 eRNAs to identify those with a significant correlation with their target genes that were associated with LUAD. Only 76 eRNAs met the condition (Spearman's rank correlation coefficient $r>0.40$, $p<0.001$; Additional file 1: Table S2). Of these, TBX5-AS1 exhibited the highest cor value, and was therefore considered to be the most relevant eRNA associated with its target gene. Kaplan–Meier curve results for TBX5-AS1 showed that patients in the low-expression group survived longer than those in the high-expression group ($p=0.02998$; Figure 1a). In addition, there was a positive correlation between TBX5-AS1 and its target gene TBX5 ($r=0.89973$, $p=2.2\times10^{-15}$; Figure 1b). We further in-vestigated the connections between the clinical features of LUAD patients and TBX5-AS1 expression and found that the expression level of TBX5-AS1 in LUAD was significantly linked to T stage (T1 vs. T2, $p=0.00011$), N stage (N0 vs. N2, $p=0.05$), sex ($p=0.0061$), and cancer status ($p=0.018$) (Figure 2a–d); however, other clinical characteristics, such as age and M stage, had no clear correlation with TBX5-AS1 expression ($p>0.05$) (Figure 2e, f).

**Gene Enrichment Analysis**
A total of 1,407 transcripts presented a significant correlation with TBX5-AS1 (p<0.05), including TBX5. GO enrichment analysis and KEGG pathway analysis of the 1,407 target genes provided the basis for the biological study of these genes. The top 10 terms for molecular function (MF), biological process (BP) and cellular component (CC) are summarized in Figure 3a. In BP, the terms were mainly related to extracellular matrix organization and extracellular structure organization; in CC, they were related to extracellular matrix, collagen-containing extracellular matrix, postsynaptic membrane, and adherens junction, among others. In MF, term enrichment mainly involved extracellular matrix structural constituent and glycosaminoglycan binding. The intensity of the colors indicates the p-value (stronger colors indicate lower p-values)—the lower the p-value, the more significant the GO term. The top 20 pathways are summarized in Figure 3b. KEGG enrichment analysis showed that the most significantly enriched biological process was "PI3K/AKT signaling pathway" (adjusted p < 0.001). The PI3K/AKT signaling pathway-related genes with a Spearman's correlation coefficient >0.6 are depicted in Table 3.

Data Validation

qRT-PCR was used to measure the expression levels of TBX5-AS1 and TBX5 in 10 LUAD samples and paired adjacent samples. The outcomes indicated that the expression of TBX5-AS1 and TBX5 in LUAD tissues was significantly downregulated when comparing it with adjacent tissues, which was consistent with its expression trend in the TCGA data set (p<0.05; Figure 4a-b). and a significantly positive correlation between TBX5-AS1 and TBX5 were seen in the samples (Spearman's rank correlation coefficient r = 0.6893, p = 0.008) (Figure 4c). Furthermore, immunohistochemistry data extracted from the HPA indicated that the protein expression of TBX5 was higher in non-tumour tissues compared with that in tumour tissues (Figure 4d-e).

Pan-Cancer Verification

To determine the prognostic role of our selected eRNA in pan-cancer and the correlation with its target gene, we conducted survival and correlation analyses. The results showed that TBX5-AS1 was associated with survival in four tumours, namely, adrenocortical carcinoma (ACC), LUAD, uterine corpus endometrial carcinoma (UCEC) and lung squamous cell carcinoma (LUSC). The survival curves for TBX5-AS1 in these four tumours are shown in Figure 5a–d. In addition, we found that TBX5-AS1 and its target gene were associated with 26 types of tumours (Additional file 1: Table S3).

Discussion

eRNAs are noncoding RNAs that are transcribed from enhancers. Studies have shown that enhancers usually regulate the expression of target genes by forming a chromatin loop with the target promoter (8,24). For example, Rosenfeld et al. found that estrogen-regulated eRNAs can bind cohesin complex proteins, including RAD21 cohesin complex component (RAD21) and structural maintenance of chromosomes 3 (SMC3), and the authors proposed that eRNA/cohesin interactions serve to stabilize chromatin looping in breast cancer cells (25). Additionally, mediator complex subunit 1 (MED1) in prostate cancer cells (26), heterogeneous nuclear ribonucleoprotein U (hnRNPU) in gastric cancer cells
(27), CCCTC-binding factor (CTCF) in colon cancer cells (28), and mediator complex subunit 12 (MED12) in T-cell acute lymphoblastic leukaemia cells (29) have been shown to regulate chromatin loop stability through interactions with eRNAs, thereby modulating the expression of downstream target genes. Although increasing evidence has indicated the critical roles of eRNAs in the control of gene expression, the potential role of eRNAs in LUAD remains largely unexplored.

In the present study, we obtained RNA expression profile data and clinical information for LUAD from the TCGA database and integrated these data with eRNA-related information. Using Spearman’s correlation and Kaplan–Meier analyses, we identified 76 candidate eRNAs related to the prognosis of LUAD. We further identified TBX5-AS1 as the most critical eRNA candidate sequence in LUAD, and TBX5 as its regulatory target. Clinical correlation analysis showed that TBX5 was differentially expressed according to T stage, N stage, gender, and cancer status. KEGG pathway enrichment results suggested that TBX5 may affect the survival outcome of LUAD patients through the PI3K/AKT pathway. Our pan-cancer validation results revealed that TBX5-AS1 was associated with survival in four types of tumours (ACC, LUAD, LUSC, and UCEC). Furthermore, TBX5-AS1 expression was correlated with that of its target gene, TBX5, in 26 tumour types. Finally, we validated the expression of TBX5-AS1 in surgical specimens by qRT-PCR and examined the expression levels of TBX5 in LUAD using the HPA databases. Together, our results indicated that TBX5-AS1 could be used as an independent predictor of LUAD.

Our results further suggest that TBX5-AS1 may regulate chromatin loop stability by binding to specific protein factors, which in turn promotes the expression of TBX5, its target gene. TBX5, a member of the T-box gene family, plays an important role in heart development (30-32). Members of this family are involved in a variety of processes, including cell–cell signalling, proliferation, apoptosis, and migration (33). Studies have shown that TBX5 dysregulation plays an important role in breast cancer (33), gastric cancer (34), oesophageal carcinoma (35,36), and colon cancer (37). For example, the TBX5 mRNA expression level is significantly downregulated in colorectal cancer cells; its methylation rate is significantly higher in cancer tissues than in normal tissues; and overexpressing TBX5 can inhibit the growth of colorectal cancer cells, promote the apoptosis of cancer cells, and reduce the migration rate of cancer cells (37). In this study, we found that TBX5 was highly expressed in normal paracancerous tissues and cell lines, whereas its expression was downregulated in LUAD tissue and cells. These findings revealed that the level of TBX5 is negatively correlated with lung cancer progression, which is consistent with the regulation of TBX5 in colon cancer (37). To date, whether TBX5 has a role in LUAD remains unknown. However, Ma et al. (2017) found that TBX5 may induce early and late apoptosis in NSCLC cells by activating key molecules in the apoptotic pathway, including cleaved caspase-3, cleaved caspase-8, CDKN2, and PARP. Moreover, the overexpression of TBX5 can inhibit cell proliferation, clone formation, and invasion, as well as induce apoptosis, in a NSCLC cell line (38). eRNA TBX5-AS1 might exert its antitumour effects through promoting the expression of TBX5. In summary, the present findings suggested that TBX5-AS1 has potential as a diagnostic biomarker and therapeutic target for LUAD.

Although we identified the key eRNA in LUAD and investigated its possible role in this disease, our study had several limitations. For instance, a larger clinical cohort is needed to further assess the diagnostic
and prognostic potential of TBX5-AS1 in LUAD, while the potential function of TBX5-AS1 in LUAD also requires further investigation. Moreover, we demonstrated that TBX5-AS1 has roles in multiple cancer types, including ACC, LUAD, LUSC, and UCEC, and the emerging roles of TBX5-AS1 in these cancers also need to be characterized.

Conclusion

In conclusion, we showed for the first time that TBX5-AS1 is a prognostic eRNA for LUAD, and our results have highlighted the functional importance of TBX5-AS1 in the pathophysiology of this disease.

Declarations

Ethics approval and consent to participate:

The study was conducted according to the guidelines of the Declaration of Helsinki, and approved by the Ethics Committee of the Second Xiangya Hospital of Central South University (protocol code 2020038 and 07/01/2020).

Consent for publication:

Informed consent was obtained from all subjects involved in the study.

Availability of data and materials

The data supporting the findings of this study are available from the corresponding author on reasonable request.

Competing interest:

The authors declare no conflict of interest.

Funding:

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Authors’ contributions:

Conception and design: WP; Administrative support: KN, BC; Provision of study materials or patients: TH; Collection and assembly of data: LC, BC; Data analysis and interpretation: LC, TH; Manuscript writing: LC; Final approval of manuscript: All authors.

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## Tables

### Table 1 Primers designed for qRT-PCR validation of eRNA.

| Name         | Bidirectional primer sequence                  | Tm (℃) | Product length (bp) |
|--------------|-----------------------------------------------|--------|---------------------|
| β-actin      | F:5' GTGGCCGAGGACTTTGATTG3’ R:5’ CCTGTAACAACGCATCTCATATT3’ | 60     | 73                  |
| TBX5-AS1     | F:5’ CGCAGTGGTGGATGCTCG 3’ R:5’ CTCGGCTCAGAGGTCAAGTAGG 3’ | 60     | 189                 |

### Table 2 Primers designed for qRT-PCR validation of mRNA.

| Name | Bidirectional primer sequence                  | Tm (℃) | Product length (bp) |
|------|-----------------------------------------------|--------|---------------------|
| β-actin | F:5’ GTGGCCGAGGACTTTGATTG3’ R:5’ CCTGTAACAACGCATCTCATATT3’ | 60     | 73                  |
| TBX5  | F:5’ GGTCTCTTTTTGCTGCTCCTTTT 3’ R:5’ TCAGGCTCCAGAGGTGTG 3’ | 60     | 195                 |

### Table 3 List of the PI3K/AKT signaling pathway-related genes associated with TBX5-AS1 expression (r>0.600, p<0.001)
| Gene   | cor  | Gene   | cor  | Gene   | Cor  |
|--------|------|--------|------|--------|------|
| ITGA8  | 0.817| TNXB   | 0.574| FGF1   | 0.458|
| LAMA2  | 0.785| VWF    | 0.566| THBS1  | 0.443|
| PDGFRA | 0.699| F2R    | 0.56 | RBL2   | 0.442|
| HGF    | 0.696| COL6A3 | 0.533| JAK1   | 0.438|
| GHR    | 0.696| ITGA4  | 0.531| FGFR2  | 0.438|
| ANGPT1 | 0.693| NTF3   | 0.53 | ITGA10 | 0.432|
| FGF10  | 0.692| VEGFD  | 0.527| PIK3R5 | 0.428|
| PDGFRA | 0.675| IL7R   | 0.518| PTEN   | 0.42 |
| COL6A5 | 0.646| ITGA1  | 0.517| ERBB4  | 0.419|
| GNG2   | 0.623| PIK3CG | 0.508| NGFR   | 0.418|
| FGF7   | 0.622| TLR4   | 0.491| COL4A2 | 0.408|
| ITGA9  | 0.619| TNR    | 0.486| FGFR1  | 0.407|
| TEK    | 0.6  | ANGPT4 | 0.482| PPP2R2B| 0.407|
| COL6A6 | 0.598| MAGI2  | 0.478| FLT4   | 0.405|
| LAMA4  | 0.597| FLT3   | 0.474| COL4A4 | 0.401|
| TNN    | 0.589| AKT3   | 0.47 | PIK3R6 | 0.401|
| PIK3R1 | 0.584| GNG7   | 0.463|       |      |
| FGF2   | 0.575| CCND2  | 0.462|       |      |
Figure 1

(a) Kaplan–Meier survival curves for TBX5-AS1. (b) A scatter plot showing the correlation between the TBX5 proportion and TBX5-AS1 expression (p<0.05).
Figure 2

The relationship between TBX5-AS1 and clinical features; (a) cancer status (p=0.018); (b) gender (p=0.0061); (c) N stage (N0 vs. N2, p=0.05); (d) T stage (T1 vs. T2, p=0.00011); (e) age (p>0.05); and (f) M stage (p>0.05).
Figure 3

(a) Biological process (BP), cellular component (CC), and molecular function (MF) in Gene Ontology (GO) enrichment analysis. (b) The top 20 enriched KEGG pathways.

Figure 4

Quantitative real-time polymerase chain reaction results for RNAs: (a) TBX5, (b) TBX5-AS1; (c) Correlation between TBX5 and TBX5-AS1; (d) TBX5 expression in lung adenocarcinoma (LUAD) tissues and normal tissues from the Human Protein Atlas (HPA) database.
Figure 5

(a–d) Kaplan–Meier survival curves for TBX5-AS1 in pan-cancer (p<0.05).

Supplementary Files

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