A Facile Activation Method for Improving the Wettability of Polyurethane Surfaces

LUIZA MADALINA GRADINARU*, STELIAN VLAD1, MIOARA DROBOTA1, MARIA SPIRIDON1, IOAN ISTRATE2

1 Petru Poni Institute of Macromolecular Chemistry, 41-A Grigore Ghica Voda Alley, 700487, Iasi, Romania
2 Polymer Adhesive Tapes SRL, 96-B Timisoara Blvd., Bucharest, Romania

This work reports the activation of polyurethane film surfaces in order to enrich them with polar groups such as -NH2, -OH, -COOH or radicals, which further allows immobilization of several bioactive compounds. UV-activation was used to introduce new groups on the polymer surface without affecting the bulk properties. The current arising species improve the wettability of the PU surfaces as it was observed from the static contact angle measurements. The structure and composition of the new PU surfaces were analyzed by using ATR-FTIR spectroscopy. The results suggested the possibility of modifying the PU surfaces in a shorter time period, in order to provide many sites to attach other biomacromolecules by polar interaction or hydrogen bonding.

Keywords: polyurethane surfaces, UV-activation, wettability, bioactive compounds immobilization

Polyurethanes (PUs) are one of the most versatile and promising families of polymers because they can be prepared from a wide variety of materials with different properties leading to a high variety of applications. They offer the possibility to tailor the final properties through the variation in the compositions by changing the ratio of soft and hard segments. Thus, PUs have been extensively used in modern life, in different fields such as coatings, adhesives, packaging, furniture, automotive, textile etc. [1-3]. Moreover, they can be obtained in various forms, such as: films, foams, gels or nanofibers. Over the past decades, PUs have been used for several biomedical applications due to their good biocompatibility, biodegradability and mechanical properties [4]. There was also a high demand of PUs based materials in manufacturing of various implantable or extracorporeal medical devices, drug delivery systems, in tissue engineering, wound dressings, etc. [5-7].

When biomaterials come into contact with the body, the biological response is greatly dependent by the chemistry and structure of the material surfaces, and therefore, the functionality of a polymeric surface is of utmost importance. Generally, surface modification of polymers aims to tailor the surface characteristics of the material for a specific application without affecting the bulk properties. Several physical and chemical techniques have been developed to improve the surface wetting, leading to blood compatibility or cellular adhesion [8, 9]. Among them, UV-treatment is one of the most suitable surface modification techniques to activate the polymeric surfaces because it exhibits several advantages, such as: low cost processing, simple equipment etc [2].

Various strategies have been developed, over the time, for polyurethane surfaces modification, because the first interaction of the biomaterials and living system is at the interface. Moreover, the blood compatibility of polymers is strongly influenced by chemical groups present at their surfaces [10]. As shown in literature, these methods involve surface modification and further grafting a hydrophilic component, such as poly(ethylene oxide) [11], 2-hydroxyethylmethacrylate [12], peptides [13], or heparin [14].

During UV treatment, some bond scission of existing groups occur at the surface of polyurethane, creating new functional groups, such as -NH2, -OH, radicals etc [15,16]. These new species can further provide an anchor for attaching bioactive natural or synthetic compounds, like: proteins, peptides, enzymes or antimicrobial agents depending on the desired application.

The aim of this study was to increase the hydrophilicity/wettability of PU surface using a facile method because the enriched polar groups on the polymer surface could provide many sites to immobilize different bioactive molecules by polar interaction and hydrogen bonding. In order to confirm the required modification at surface level, some indirect or direct methods are successful used for their characterization [17]. Two experimental techniques were chosen for the present study: Attenuated total reflectance-Fourier transform infrared (ATR-FTIR) and contact angles measurement. The first one gives important information on composition and structure of PU surfaces and the second one provides quantitative information concerning the surface free energies. All the measurements were carried out before and after UV-treatment of the PU surfaces.

Experimental part

Materials
Poly(caprolactone) diol (Mn = 2000 Da), poly(1,4-butylene adipate) diol (Mn = 2000 Da), 1,4-butanediol, dimethylformamide (DMF) and ethylene glycol were purchased from Sigma-Aldrich (Germany) and used as received. Methylene diphenyl 4,4’-disocyanate was freshly distilled before synthesis. Other chemicals were of analytical grade and used without further purification.

Preparation of PU films

The synthesis method of this PU, using a two-step solution polymerization has been previously reported in detail [15, 16, 18]. Thus, the PUs was obtained from poly(caprolactone) diol and poly(1,4-butylen adipate) diol as soft segment, 1,4-butanediol as chain extender, and methylene diphenyl 4,4’-disocyanate as hard segment. After polymer synthesis, the dried PU was redissolved in
DMF at a concentration of 30% (w/w). The PU solution was degassed under vacuum (10-15 mmHg) and then it was casted onto a glass slide using a doctor blade with a gap of 0.6 mm. The films were precipitated in distilled water at 45°C. The resulted PU films were intensively washed with distilled water and dried at room temperature and low pressure (1-2 mmHg). The films were finally cut into rectangular shapes of 20 x 60 mm and used for further modification.

**Surface modification**

The PU samples were activated in air at room temperature with a Herolab UV Analysis Lamp (Germany), Model: UV-8 S/L: 254/365 nm, UV tube type: je 1 x 8 W, Intensity: 680 (S)/950 (L) µW/cm². The distance between the samples and light source was kept constant at 20 cm. The rectangular PU samples were UV-irradiated for different time points at 2, 4, 6 and 8 h.

**Surface analysis**

**ATR-FTIR analysis**

Attenuated total reflectance-Fourier transform infrared (ATR-FTIR) spectra of the PU samples were recorded using a BrukerLUMOS - FT-IR Microscope (Germany) equipped with a diamond crystal with single reflection at 45° angle and OPUS 8 spectral processing software. The sample surface was scanned in the 4000-600 cm⁻¹ range, with a resolution of 2 cm⁻¹. All ATR-FTIR spectra were recorded at room temperature.

**Static contact angle measurements**

Static contact angle measurements with the sessile drop method were recorded and analyzed at room temperature on a CAM-101 contact angle system from KSV Instruments (Finland). The contact angle system was equipped with a liquid dispenser, video camera, and drop-shape analysis software. For this study, double distilled water and ethylene glycol were used as solvents. The reported contact angle values are the average of at least 3 measurements on different regions of the surface, for each kind of liquid.

**Results and discussions**

**ATR-FTIR analysis**

The spectroscopic methods are key tools in determining the presence of functional groups on the polymeric surfaces. Thus, the UV-activation process was monitored by ATR-FTIR analysis and the spectra were collected before and after exposure to UV light for 2, 4, 6 and 8 h, respectively.

The ATR-FTIR spectrum of non-irradiated PU shows the main absorption bands, characteristic of the PU structure: free -NH and -OH stretching vibrations (3500 cm⁻¹), hydrogen-bonded -NH stretching vibrations (3330 cm⁻¹), C-H asymmetric and symmetric stretching (2968 cm⁻¹ and 2856 cm⁻¹ respectively), C=O stretching (1727 and 1702 cm⁻¹), and C-O-C stretching (1223 and 1075 cm⁻¹).

It is well known that the UV-irradiation affects the chemical structures of the PU surfaces at longer activation time, leading to degradation [19]. However, shorter UV-irradiation time is a preferred method to activate the PU surface, in order to improve its wettability, providing further anchors for attaching some bioactive compounds, without reduction of polymer molar mass [20]. The mechanism is very complex and involves, as previously was shown, some cleavage of the urethane group (Photo-Fries rearrangement), photooxidation of the central -CH₂ group flanked by aromatic rings, Norrish type mechanism, etc (fig. 1) [15, 16, 19, 21]. Thus, after UV-irradiation several changes were noticed in the ATR-FTIR spectra (fig. 2).

UV treatment induced the appearance of some hydrophilic groups on the PU surfaces. Thus, the -NH and -C=O groups from the PU structure display a fingerprint in important infrared regions because these groups can interact by inter- and intramolecular hydrogen bonding. The presence of the absorption bands in the 3700-3200 cm⁻¹ region has been assigned to the -OH and -NH stretching vibration. The PU surfaces were characterized by a peak at around 3330 cm⁻¹ due to the absorption of bonded N-H stretching vibration, which decreases with irradiation time.

**Fig. 2. ATR-FTIR spectra of the PU samples before and after UV-irradiation at different time points**
At longer exposure time the shoulder intensity, around 3500 cm$^{-1}$, slightly increases as displayed in figure 2. This peak corresponds to the free-hydrogen stretching vibrations of free -NH-COO-, -NH$_2$-, -OH that arises after irradiation of PU surfaces and is the result of both Photo-Fries rearrangements of the urethane groups and photo oxidation of ester groups [15, 16].

The peaks at 2959 cm$^{-1}$ and 2854 cm$^{-1}$ can be assigned to the asymmetric and symmetric C–H stretching vibration of the -CH$_2$- groups.

Two peaks of carbonyl absorption can also be identified, which can be due to a free carbonyl stretching vibration at 1727 cm$^{-1}$ and a hydrogen - bonded carbonyl at 1702 cm$^{-1}$ (fig. 3). Moreover, a shoulder around 1781 cm$^{-1}$, characteristic to the C=O stretching vibration of carboxyl groups was distinguished and increases with the irradiation time. These carboxyl groups are originated from the ester functional groups in the PU structure [22]. This is consistent with the broadening of the -OH stretching vibration from the 3500 cm$^{-1}$.

Like -NH region, the C=O stretching vibration region has a major concern because provides useful information on the mode of hydrogen bonding. The vibration band in the carbonyl region can be considered as a sum of peaks, obtained by its deconvolution, based on the Gaussian function. For example, the deconvolution spectra of the non-irradiated and 8h-activated PU samples were performed. For these samples different peaks varying in height, wavenumber and wavelength were resulted by deconvolution. Thus, the non-irradiated PU sample has the peaks maximum at 1740, 1731, 1723, 1706, 1698 and 1689 cm$^{-1}$, respectively, whereas the 8hours activated PU sample at 1782, 1735, 1731, 1726, 1712, 1699 and 1691 cm$^{-1}$, respectively. Therefore, the deconvolution peaks from the higher frequency are assigned to the free C=O stretching vibration and those from lower frequency are attributed to the H-bonded C=O stretching vibration [23].

In figure 5 the ATR-FTIR spectra of PU samples before and after UV-irradiation, between 1600 and 600 cm$^{-1}$ are presented. Generally, the PU structures are not stable to light and are susceptible to degradation when they are exposed to the UV radiation [24]. Thus, the signal at 1599 cm$^{-1}$ is characteristic to the stretching vibration of the C=C in the aromatic rings [25] and shows a tendency of decreasing with increase of the UV-exposure time. This phenomenon is due to the oxidation of the -CH$_2$- groups derived from the aromatic disiocyanate structures, leading to some conjugated quinone products [19, 24].

The absorbance at around 1530 cm$^{-1}$ in the ATR-FTIR spectrum of PU sample could be attributed to the coupling of N–H bending vibration with C-N stretching vibration in the -C-NH group (amide II band) and is more complex than amide I [19]. The decrease in the absorbance region of the amide II indicates that some degradation have occurred in the urethane groups via a free radical mechanism [26] resulting in the appearance of some radicals that could later interact with bioactive molecules. The peak at 1414 cm$^{-1}$ is due to the symmetric bent vibration of the -CH$_2$- groups derived from the polyester structures used in PU synthesis.

The absorption band at 1310 cm$^{-1}$ corresponds to the combination of N–H bending vibration and C-N stretching vibration (amide III band). Between 1300-1100 cm$^{-1}$ are the bands characteristic to the C-O stretching vibrations in esters. The stretching vibration of C-O-C groups from the urethane structures can be observed at 1073 cm$^{-1}$ and the symmetric stretching vibration at 1022 cm$^{-1}$.

Finally, the finger print infrared region between 900–600 cm$^{-1}$ was investigated. The UV-irradiated PU samples display a decrease of the bands centered at 856, 813, 772 and 664 cm$^{-1}$, respectively, corresponding to the aromatic C-H out-of-plane or in-plane deformation vibration in p-disubstituted rings [21, 27].
UV-activation exhibited an average contact angle of 115°, before and after UV-activation. The PU film before interfacial tension, three interfaces and can be described by Young’s equation: where: $\gamma_{SV}$ is the energy of surface, $\gamma_d$ is the solid - drop interfacial tension, $\gamma_{LV}$ is the liquid-vapor surface tension and $\cos \theta$ is the drop - surface contact angle [29-31].

Figure 6 shows the average contact angles data of PU films before and after UV-activation. The PU film before UV-activation exhibited an average contact angle of 115°, suggesting that PU surface is hydrophobic. After 2 h of UV-activation of PU surfaces, the contact angle decreased to 114°, and further decreased after 8 h of irradiation until 95°. The increase in hydrophilicity and better wettability behavior of the 8 h UV-irradiated PU surface compared to untreated PU are attributed to a complex reaction mechanism like Photo-Fries rearrangement of the urethane groups or Norrish I reaction [15, 16]. Therefore, after UV-irradiation, the PU surfaces become hydrophilic due to the arising of some polar groups such as -NH$_2$, -OH, -COOH etc. (fig. 1), as will be further seen in calculation of polar component of free energy. These polar groups on the surface, and as a consequence, the surface free energy increased.

Conclusions

The results presented in this work suggest that the modification of PU surfaces by using UV-irradiation may be obtained without affecting the bulk properties. The UV-treatment of the PU surfaces, in a shorter time of irradiation, results in breaking of chemical bonds and generation of new functional groups on the surface which can be controlled by the irradiation time. These new groups are generally polar groups, such as -NH$_2$, -OH, -COOH etc., that could provide sites to immobilize proteins or other molecules by polar interaction or hydrogen bonding. UV-activation process was monitored by ATR-FTIR spectroscopy which is a key tool in determining the presence of functional groups on the PU surfaces. Moreover, the UV-modifications induced an increase of hydrophilicity which was evaluated by static contact angle measurements, a convenient and simple method of surface analysis. The wettability and surface energy measurements of the PU surfaces are important parameters that could influence the cellular adhesion and biocompatibility. In conclusion, this method was proven to be a suitable surface modification technique to activate the PU films in order to enhance the biomacromolecules absorption.

| Sample time points | $W_a$ (mN/m) | $\gamma_{SV}$ (mN/m) | $\gamma_{LV}$ (mN/m) | $\gamma_{SL}$ (mN/m) | $\gamma_{SE}$ (mN/m) |
|-------------------|--------------|----------------------|----------------------|----------------------|----------------------|
| 0 h               | 41.34        | 5.87                 | 4.17                 | 1.69                 | 37.32                |
| 2 hrs             | 43.13        | 6.24                 | 4.48                 | 1.53                 | 36.86                |
| 4 hrs             | 47.33        | 7.81                 | 6.22                 | 1.49                 | 29.28                |
| 6 hrs             | 53.98        | 10.50                | 8.20                 | 1.30                 | 29.31                |
| 8 hrs             | 66.33        | 17.17                | 16.31                | 0.85                 | 23.63                |
References

1. AKINDOYO, J.O., BEG, M.D.H., GHAZALI, S., ISLAM, M.R., JERYARATNAM, N. YUVARAJ, A.R., RSC Adv., 6, 2016, p. 114453.
2. ALVES, P., FERREIRA, P., GIL, M.H., Polyurethane: Properties, Structure and Applications, Cavaco, L.I., Melo, J.A. (Ed.), Nova Publishers, 2012, p. 25.
3. SOMARATHNA, H.M.C., RAMAN, S.N., MOHOTTI, D., MUTALIB, A.A., BADRI, K. H., Constr. Build. Mater., 190, 2018, p. 995.
4. KHATOON, H., AHMAD, S., Significances Bioeng. Biosci., 2, 2018, 000536.
5. SHELKE, N.B., NAGARALE, R.K., KUMBAR, S.G., Natural and synthetic biomedical polymers, Kumbar, S.G., Laurencin, C.T., Deng, M. (Ed.), Elsevier, Amsterdam, 2014, p. 123.
6. WANG, W., WANG, C., The design and manufacture of medical devices, Woodhead Publishing, 2012, p. 115.
7. CITU, I.M., BORCAN, F., ZAMBORI, C., TITA, B., PAUNESCU, V., ARDELEAN, S., Rev. Chim. (Bucharest), 66, no.1, 2015, p. 233.
8. THEVENOT, P., HU, W., TANG, L., Curr. Top. Med. Chem., 8, 2008, p. 270.
9. STAMM, M., Polymer Surfaces and Interfaces. Characterization, Modification and Applications, Springer, Berlin, 2008.