ANTIOXIDANT POTENTIAL IN RELATION TO TOTAL PHENOLICS AND PIGMENT CONTENTS AMONG THE SELECTED LANDRACES OF *Dioscorea alata L.* IN KERALA, INDIA

Anumol Jose, Sajna Nizar, Vishnu MR, Anil Kumar M*

Department of Botany, Union Christian College, Aluva, Kerala, India. Pin-683102.

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ABSTRACT

The present study aims to evaluate and compare the phenolics, pigment content and antioxidant potential of the four landraces of *Dioscorea alata* prevalent in Kerala. *Dioscorea alata* (greater yam) is an underutilized, food security crop that exhibits wide variability in morphological, biochemical and agronomical traits. One of the distinct morphological features that varied among landraces of *D. alata* is tuber flesh colour which ranged from white, yellow, pink and purple. CIELAB coordinates of each tuber colour were estimated using reflectance spectroscopy and represented in L*a* b* values. Quantification of total phenolic compounds and pigments were done spectroscopically using standard protocols. Antioxidant activity of the tuber extracts were estimated using 1,1-Diphenyl-2-picryl hydrazyl (DPPH) quenching assay and ferric reducing antioxidant power (FRAP) assay. Total phenol and flavonoid content in yellow and purple-fleshed *D. alata* were significantly higher than white and pink fleshed landraces. Spectrophotometric assays revealed the differential expression of anthocyanins and carotenoids in tubers, which corresponds to the colour difference among different landraces. Antioxidant activity based DPPH and FRAP assay were significantly different among the four landraces. Yellow and purple-fleshed landraces shared high antioxidant activity and was positively correlated with the phenolic and pigment contents. The study also listed certain morphological features that aid in identifying the landraces of *D. alata*.

* Corresponding author
E-mail: drmakumar@gmail.com (Anilkumar M)

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1 Introduction

*Dioscorea alata*, commonly known as greater yam, is an underutilized and climate resilient crop widely cultivated in tropical and subtropical countries (Bressan et al., 2011). This plant belongs to the genus *Dioscorea* of family Dioscoreaceae and being a subsistent food crop it is well known for the close relationship with human civilization. But, changed food habits and agricultural practices in the modern world turned this crop as a less preferable one and subsequently its status shifted to the underutilized category (Siqueira, 2017). The edible underground tuber of greater yam is rich in carbohydrates and other nutritionally important secondary metabolites. Major bioactive components present in greater yams included proteins, polyphenols, carotenoids, alkaloids and phytosterols (Lubag et al., 2008). Owing to the presence of these bioactive molecules, greater yam was reported to possess anti-inflammatory, anticancerous, antiobesity and antibacterial properties (Das et al., 2014; Jesus et al., 2016; Chen et al., 2017). The pharmacological and biological properties of greater yams have been attributed to the chemoprotective antioxidants. Antioxidants are molecules that at low concentration delays oxidation of nucleic acids, carbohydrates, lipids, and proteins (Holick et al., 2002; Forbes-Hernández et al., 2017). Antioxidants as food supplements impart health benefits in various conditions like stress, aging, pathogen infestation, apoptosis, and neurodegenerative diseases (Sindhi et al., 2013). Phytoconstituents such as polyphenols, carotenoids and alkaloids were known to possess antioxidative properties. Quantitative analysis of phytochemicals present in the plants could be reliable measure of its antioxidant property (Tang et al., 2015). Yam species exhibits considerably high antioxidant properties, thus making them an economically important crop in the modern scenario (Ukom et al., 2014). New developments and promising investigations in this area can elevate the agricultural preference of this neglected crop.

In India, 50 species of *Dioscorea* were reported from southern and north eastern hill regions of which 17 were distributed in Western Ghats (Kumar et al., 2017). Kerala is geographically located at the southwest edge of India and is known for its tropical climate ideal for the growth and propagation of *Dioscorea* species. *D. alata* is the most common yam present in Kerala and exhibits a broad spectrum of variability in morphological, agronomical and biochemical traits (Jyothi et al., 2017). Such variable morphotypes were considered as landraces and were known by different vernacular names. Even though an integral part of traditional agriculture in tropical and subtropical countries, landraces of *D. alata* were least explored and remain neglected.

One of the distinct morphological feature useful to identify these landraces were their tuber flesh colour that ranged from white to yellow, pink and purple. It was reported that there were measurable variations in the starch and protein content of these landraces, but the data available regarding the bioactive compounds and the potential antioxidative properties in them were scanty. The present investigation is an attempt to correlate the antioxidant property of selected land races of *D. alata* with the total total phenolics, flavonoids, carotenoids, and anthocyanins.

2 Materials and Methods

TPTZ (2,4,6-tripyridyl-s-triazine), FeSO₄.7H₂O (iron (II) sulfateheptahydrate) and 1,1-Diphenyl-2-picrylhydrazyl (DPPH) were purchased from Sigma Chemicals, USA. Aluminum chloride, FolinCio-Calteus’s phenol reagent, sodium carbonate, FeCl₃, gallic acid, ascorbic acid and hydrochloric acid were purchased from Merck. All the chemicals used including the solvents were of analytical grade.

2.1 Plant materials

Landraces of greater yams, which showed distinct colour differences in their tuber flesh, were collected from local farmers of Kerala. Morphological characterization and species identification were done according to the norms of IPGRI (IPGRI, 1997). Forty-five accessions of *D. alata* were maintained in the field as germplasm and the tuber colour was observed and documented at the end of growing seasons. Four landraces, which expressed stable white, yellow, pink and purple flesh colours for three consecutive growing seasons were selected for the study. Details of the accessions were given in table 1.

| Accession number | Local name       | Place of collection | Flesh colour |
|------------------|------------------|---------------------|--------------|
| KLE DA 01        | Chorakachil      | Ernakulam           | Purple       |
| KLI DA 12        | Rosakamban kachil| Idukki              | Pink         |
| KLP DA 08        | Parakachil       | Pathanamthitta      | Yellow       |
| KLK DA 10        | Bharanikachil    | Wayanan             | White        |

2.2 Morphological Characterization of Accessions

Morphological characterization was done as per the methods given by International Plant Genetic Resources Institute’s descriptor list (IPGRI/ IITA, 1997). The accessions were analyzed for distinct qualitative and quantitative characters. Only those features that
discriminated between the accessions were used for the present analysis. Data were collected in triplicates from at least three different healthy plants of each accession and then averaged.

2.3 Extraction of phenolics compounds and pigments

Phenolics, flavonoids, anthocyanins and carotenoids were extracted from the tuber as per method given by Ferede et al. (2010) and Fang et al. (2011). For the extraction of phenolics and flavonoids, one gram tuber flesh devoid of skin was homogenized in 10 ml of 60% ethanol and the supernatant was collected. The residue was re-extracted thrice and all the supernatants were pooled. For anthocyanin isolation, 60% ethanol acidified with 0.2% HCl was used. Carotenoid extraction was carried out by using 90% ethanol.

2.4 Biochemical estimations

Estimation of phenol, flavonoid, total anthocyanins and carotenoids were carried out according to the standard protocols. Each experiments were triplicated and the average values taken for analysis.

2.4.1 Total Phenol and flavonoid Estimations

The total phenolic contents of tuber extracts were determined using the Folin-Ciocalteau method (Sellappan et al., 2002). Quantification was based on gallic acid calibration curve and the results were expressed as gallic acid equivalents per 100 gram of fresh tissue. The results were averages of triplicate analysis.

The total flavonoid content was determined using Aluminium chloride assay (Zhishen et al., 1999; Pekal & Pyrzynska, 2014). 0.5 ml of 2% AlCl₃- ethanol solution was added to 0.5 ml of the sample mixed with 150 µl of 5% sodium nitrite. After six minutes of incubation, 2 ml 1N NaOH solution mixed and further incubated for 15 minutes. The absorbance of the mixture was measured at 510nm. Total flavonoid content was determined with the help of the calibration curve of quercetin and results were expressed as quercetin equivalents per gram of fresh tissue. The experiment was repeated thrice and averages were taken.

2.4.2 Anthocyanin and carotenoid estimations

Total anthocyanin content (TAC) was estimated by spectrophotometric pH differential method (Guisti & Wrolstad, 2005). The absorbance of tuber extract at 520 and 700 nm was measured using a UV-Visible Spectrophotometer (Shimadzu, Japan). Pigment content was calculated as Cy-3-glu equivalents using the following formula

Total Anthocyanins (mg/L) = $A \times MW \times DF \times 1000 / \varepsilon \times I$

Where $A = \text{Absorption at 520}; MW = \text{Molecular Weight of cyanidin-3-glucoside (449.2)}; DF = \text{Dilution Factor}; \varepsilon = \text{molar extinction coefficient of Cyanidin-3-glucoside (20900 L/cm/mg)}$

Total carotenoid content was determined from the absorbance measured at 450 nm on a Vis-spectrophotometer and was quantitatively estimated using the formula suggested by Rodríguez-Amaya & Kimura (2014).

Total carotenoid content (mg/g) = $A \times \text{Volume of extract in ml} \times 1000 / \varepsilon \times \text{sample weight in g}$

Where $A = \text{absorbance}; \varepsilon = \text{absorption coefficient (2500 L/cm/mg)}$

2.5 Measurement of colour coordinates

Reflectance spectrum of tuber samples was taken using UV spectrometer with integrated sphere attachment (2800 plus, Schimadzu, Japan) and the L*a*b* coordinates calculated from the spectrum using colour analysis software (Shimadzu).

2.6 Antioxidant activity analysis

DPH radical scavenging activity was estimated as per previous reports (Yu et al., 2002; Sing et al., 2002; Tang et al., 2015). For this, 100 µl of the sample extract was added to 2.9 ml of DPPH reagent in ethanol and vortexed vigorously. It was incubated in dark for 30 min at room temperature and the discolouration of DPPH was measured at 517 nm. Percentage inhibition on the discolouration of DPPH by the sample extract was expressed as ascorbic acid equivalents.

Ferric reducing antioxidant power (FRAP) was determined in the sample extracts according to previous reports (Abu Bakar et al., 2016). 3.0 ml of FRAP reagent was added to the known concentration of the sample extract. After six minutes of incubation at room temperature, the absorbance was measured at 593 nm against ascorbic acid standard.

2.7 Statistical analysis

The experimental results were expressed as mean ± standard deviation of three replicates and was subjected to one-way analysis of variance (ANOVA) followed by post-hoc analysis by tukey t-test (SPSS statistics version 20). P values ≤ 0.05 were regarded as significant. The correlation coefficient (R-value) was calculated using Microsoft Excel software.

3 Results and Discussion

3.1 Analysis of morphological features

Morphological features which prevailed stable for three consecutive growing seasons were analyzed and documented. Distinct and prominent features observed among accessions were summarized in table 2. Young leaves were of purple in all the accessions, which exhibited colouration in tuber flesh while it was
Table 2 Morphological variations in selected *D. alata* accessions from Kerala

| Character                      | KLE DA 01 (Purple) | KLI DA 12 (Pink) | KLP DA 08 (Yellow) | KLY DA 10 (White) |
|--------------------------------|--------------------|-----------------|-------------------|------------------|
| Young leaf colour              | Purplish green     | Purple with green tips | Purplish green     | Brownish green   |
| Leaf shape                     | Cordate broad with cupping | Sagittate long | Cordate long with cupping | Cordate long without cupping |
| Tuber shape                    | Variegated         | Round           | Variegated         | Cylindrical      |
| Tuber cortex colour            | Dark purple        | Dark purple     | Light purple       | Light brown      |
| Tuber flesh colour             | Purple             | Pink            | Yellow with white patches | White |
| Tuber skin texture             | Highly warty       | Smooth          | Warty              | Warty            |
| Presence and nature of spine   | No spine           | No spine        | Purple short spines at the base of young stem | No spine |
| Presence of stem patches       | No stem patches    | No stem patches | Purple long patches on young stem | No stem patches |

Data were the average of at least three healthy plants per accession

Brownish green in the white fleshed tuber. Pink-fleshed *D. alata* plants showed a distinct green coloured tip on their purple young leaf. Cordate and sagittate leaf shapes were frequently observed in *D. alata* accessions. Among different accessions, cordate shape again varied due to the presence or absence of cupping formed by the overlapping of leaf lobes. Pink-fleshed *D. alata* were distinct among the selected accessions due to the presence of sagittate shaped leaves and round shaped tuber with smooth skin. Tuber cortex colour expressed shades of purple in three accessions but was brown in the case of the yellow-fleshed tuber. Presence of spin and purple patches in the stem differentiates yellow fleshed yam from other accessions. According to IPGRI norms, 47 descriptors were used for morphological characterization of *Dioscorea* species (Hasan et al., 2008). Being the most polymorphic species in the genus *Dioscorea*, landraces of *D. alata* showed many overlapping morphological characteristics (Anoyke et al., 2014). The quantitative features of *D. alata* landraces were found to be unstable compared to qualitative characters. Hence the present study was focused only on qualitative characters which were distinct among different accessions. Morphological features such as mature leaf colour, the presence of bulbil, colour of ridge, petiole colour, colour of leaf margin and shape of stem cross section were evaluated and found to be unstable over growing seasons. In this juncture, focus has been given only to 8 qualitative characters which were relevant in identifying landraces of *D. alata*. Results suggested that young leaf colour, leaf shape, colour of tuber cortex and flesh, shape of tuber and stem characteristics such as the presence of spine and patches could be useful in identifying the landraces of greater yam. These morphological features were reported to be important in analysing morphological diversity among landraces of *D. alata* distributed in Ivory Coast, Vietnam, Ethiopia and Brazil (Bressan et al., 2011; Michel et al., 2015; Tewodros et al., 2018).

### 3.2 Colour Characteristics of tubers

The colour values of greater yam tubers from five landraces were presented in Table 3. CIE L*a*b* colour system (CIELAB) based on reflectance spectroscopy was adopted for the quantification of colour. The L*a*b* coordinates described in the present study indicates L* = 0 as black and L* = 100 as white and the a* and b* represents position between red (+) and green (−), and yellow (+) and blue (−) respectively (Tang et al., 2015). Raw tubers of the five landraces showed distinct colour variations both visually and in terms of colour coordinates. These colour differences in landraces may occur due to the presence of several types of pigments, such as anthocyanins and carotenoids. Anthocyanins impart pink and purple colour while carotenoids were responsible for yellow colour (Davies et al., 1998; Berman et al., 2016). The flesh colour variations in the *D. alata* landraces from white to different shades of purple by visual observations were reported earlier (Jyothi et al., 2017). Colour quantification using CIELAB coordinates help to identify the exact colour tone of *D. alata* landraces with accuracy and repeatability.

Table 3 CIELAB Colour coordinates of yam tuber flesh

| Accessions and tuber flesh colour | L*       | a*       | b*       |
|----------------------------------|----------|----------|----------|
| KLY DA 10 (White)                | 93.9 ±0.03 | 0.11 ±0.3 | 7.3 ±0.5|
| KLP DA 08 (Yellow)               | 82.6± 0.03 | 2.21± 0.02 | 58 0±.5 |
| KLI DA 12 (Pink)                 | 70.33 ±0.4 | 36.8± 0.3 | 2.71± 0.3|
| KLE DA 01 (Purple)               | 45.5 ± 0.2 | 50.45 ±0.6 | 42.8± 0.3|

Data were the average of three tubers per accession
3.3 Total Phenolics and Flavonoid

The total phenolic and flavonoid contents were significantly varied among differentially coloured tubers of *D. alata* (Figure 1). White and pink-fleshed landraces expressed significantly lower phenolic and flavonoid contents compared to others. Phenol and flavonoid contents in yellow and purple-fleshed greater yam landraces were comparable and higher than pink and white landraces. This suggested that yellow and purple-fleshed greater yam landraces were nutritionally superior to others. Plant-derived phenols and flavonoids were considered as dietary antioxidants that impart beneficial implications in human health (Bravo, 1998). Previous studies established that *D. alata* contains a substantially increased amount of phenols and flavonoids in its tuber (Sajjaanantakul et al., 2014). In the present investigation, there were significant variations in the total phenolic content among the landraces of *D. alata* and it was positively correlated with the tuber colour imparted by different pigments. The extent of these variations was found to be almost three-fold which clearly indicated the nutritional superiority of coloured tubers of *D. alata*. Phenolic composition of yellow and purple yam was equivalent to that of drumstick, cabbage, and cluster beans (Sreeramulu & Raghunath, 2010). Present results also confirmed that the amount of phenols present in yams was equal and even higher than most of the vegetables consumed by people.

3.4 Total anthocyanin and carotenoid

Anthocyanins and carotenoids were the pigments in yams which impart colour to its tubers. Results from the present study indicated that purple fleshed yams accumulate significantly higher amount of both pigments (Figure 2). Purple-fleshed tuber contained 22.7mg of anthocyanins and 1.89mg of carotenoids per100g of fresh tissue. In yellow fleshed landraces amount of carotenoids were less than purple fleshed tubers but appeared yellow owing to the low anthocyanin content (12.2mg/100g of tissue). Pink-fleshed yam tubers contained both pigments in lower concentrations but due to the elevated amount of anthocyanins compared to carotenoids its colour shifted more towards red scale than yellow. White-fleshed tuber cumulates both pigment in very low concentrations and hence appeared white in colour. Both in vitro and in vivo studies have provided significant evidence on the
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The potential of anthocyanins as a functional food. Anthocyanins were well known for their anti-inflammatory and anti-carcinogenic activity as well as preventive effects on cardiovascular diseases, obesity and diabetes (He & Giusti, 2010). Carotenoids were also considered as a potential dietary component which provides health benefits in decreasing the risk of diseases, especially certain type of cancers and eye diseases (Holick et al., 2002). Being a source of both carotenoids and anthocyanins, greater yam possesses immense potential as a functional food. Our results suggested the potential of purple and yellow fleshed landraces of greater yam as a reservoir of functionally important phytoconstituents.

### 3.5 Antioxidant activity and correlation analysis

Different landraces of greater yam contained high amounts of phenol, flavonoids, anthocyanins and carotenoids which were well known for their antioxidant properties. In this context the knowledge on the extent of antioxidant activities imparted by these landraces were of prime importance. Literature suggested a number of assays that have been employed to evaluate the in vitro antioxidant activities of different plant materials, of which ferric reducing antioxidant power (FRAP) and 2, 2,0-diphenyl-1-picryl hydrazyl (DPPH) radical scavenging assays were the most common. Both of these assays rely on the use of a positive control as it clearly helps to represent the antioxidant properties of the samples. In this study, ascorbic acid was used as the positive control. In table 4 the antioxidant activity of different landraces expressed in micromolar concentrations of ascorbic acid equivalent per gram of yam fresh tissue. Yellow and purple fleshed landrace have comparatively higher antioxidant properties in terms of both assays while white and pink fleshed landraces showed lower antioxidant activities. Similar to other studies (Brad Williams et al., 1995) FRAP assay values were proportionally different from DPPH assay values, presumably due to variations in reaction kinetics and antioxidant potentials of various reductive substrates as they interact with the two radicals (Ozgen et al., 2010). Antioxidant property of yellow and purple fleshed tubers were comparable and could be attributed to the fact that they have similar phenolic and flavonoid contents.

Correlation analysis was performed to determine the contribution of polyphenols, flavonoids and pigments towards antioxidant activities of landraces. The R² values obtained in the analysis were listed in table 5. The results suggested that there was a strong positive correlation between the phenol content and antioxidant activity in terms of DPPH and FRAP assay values (r-value< 0.9). The total flavonoid content and antioxidant activity was also correlated positively. Even though values from the two antioxidant assay techniques were different, the data sets in this study were highly correlated. The positive correlation (r value <0.7) between antioxidant activity and pigment contents suggested their contribution towards the antioxidant activities.

### Table 5 Correlation coefficient (R² values) of DPPH and FRAP activity vs total phenol, flavonoid, anthocyanin and carotenoids

| Antioxidant activity | Total phenol | Total flavonoid | Total anthocyanin | Total carotenoid |
|----------------------|-------------|----------------|------------------|-----------------|
| DPPH activity        | 0.99        | 0.96           | 0.74             | 0.89            |
| FRAP activity        | 0.95        | 0.90           | 0.47             | 0.70            |

Conclusion

This study documented certain distinct morphological features exhibited by four different landraces of *D. alata* in Kerala. The CIELAB colour coordinates were helpful in identifying tuber colour variations in the selected landraces. Among four landraces of greater yams, yellow and purple fleshed yams showed significantly higher antioxidant activity which in turn was positively correlated to the amount of phenolics and pigments. White and pink fleshed yams showed comparatively lower antioxidant activity and their phenol and pigment compositions were also low. This study suggested the nutritionally important biochemical properties of yellow and purple fleshed yams. In Kerala, white fleshed yam is more acceptable and popular as a food crop. On the contrary, the results of our study proved the nutritional superiority of yellow and purple yams over white fleshed yams. Hence, we suggest the use and cultivation of purple and yellow fleshed yams as a nutritionally rich food crop for the people of tropical and subtropical countries. Further investigation on the genetic and metabolic profile of these landraces are in progress in our lab.

Conflict of Interest

Authors would hereby like to declare that there is no conflict of interests that could possibly arise.

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