Synthesis and Antitumor Activities of Phenanthrene-Based Alkaloids

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Abstract: A series of phenanthrene-based tylophorine derivatives (PBTs) were synthesized and their cytotoxic activities against the H460 human large-cell lung carcinoma cell line were evaluated. Among these compounds, N-(3-hydroxy-2,6,7-trimethoxyphenanthr-9-ylmethyl)-L-prolinol (5a), and N-(3-hydroxy-2,6,7-trimethoxyphenanthr-9-ylmethyl)-L-valinol (9) exhibited good activities, with IC₅₀ values of 11.6 and 6.1 μM, respectively.

Keywords: PBTs; synthesis; cytotoxicity

1. Introduction

Phenanthroindolizidine alkaloids have been of considerable interest as anticancer agents because of their exceptionally potent antitumor activity [1]. Based on recent studies, the molecular target of phenanthroindolizidine analogues may be novel and different from the targets of known anticancer drugs [2]. Over the past decades, the synthesis and biological activities of varied 2,3,6,7-functionalized
phenanthroindolizidine alkaloids have been reported. The substituents mainly include functionalities such as hydroxyl, methoxyl, and hydrogen [3–5]. The structure activity relationships of phenanthroindolizidine alkaloids were summarized by Gao et al. in 2007 [3]. The degree of cytotoxicity was dependent on the type and pattern of substitution on the phenanthrene ring. In 2006, Wei et al. [6] found that phenanthrene-based tylophorine derivatives (PBTs), which were phenanthroindolizidine derivatives resulting from opening of the indolizidine ring, exhibited significant cytotoxic activity. Further work from the same group resulted in the synthesis of a series of PBTs with different substituents at C-9, which indicated that a five- or six-carbon distance between the nitrogen and a terminal polar substituent in the C-9 chain is quite favorable for cytotoxic activity [7]. They also reported that antitumor activities of PBTs were induced by inhibition of the activation of Akt and NF-κB signaling pathway in tumor cells [8,9].

As part of our studies on antitumor agents derived from natural products, we have designed and synthesized a series of PBT analogues to investigate the effects of oxygenic functional groups on the phenanthrene ring and C-9 side chain on cytotoxic activity. The present paper reports the synthesis of these PBTs (Scheme 1) and evaluation of their cytotoxic activities against the H460 human large-cell lung carcinoma cell line in vitro (Table 1).

**Scheme 1.** The synthetic routes to PBTs.
Table 1. The structures of 5a, 5b, 8-12 and their cytotoxicities against H460 cell line.

| Compound | R₁   | R₂          | IC₅₀ (μM) |
|----------|------|-------------|-----------|
| 5a       | H    | N(CH₂OH)    | 11.6      |
| 5b       | OMe  | N(CH₂OH)    | 53.8      |
| 8        | H    | N(OH)       | 68.1      |
| 9        | H    | N(OH)       | 6.1       |
| 10       | H    | N(CH₂OAc)   | 46.8      |
| 11       | H    | N(CH₂OSO₃H) | 53.4      |
| 12       | H    | N(CH₂Cl)    | 62.9      |

Adriamycin used as positive control, IC₅₀ = 1.72 μM.

2. Results and Discussion

2.1. Chemical synthesis

The synthetic routes to the PBTs are depicted in Scheme 1. Perkin condensation [10] involving 4-hydroxy-3-methoxy- or 3,4-dimethoxybenzaldehyde and 3,4-dimethoxyphenylacetic acid in the presence of Ac₂O and Et₃N afforded the cinnamic acid derivatives 2a (R = Ac) or 2b (R = Me) [11–14]. By treatment with oxalyl chloride [15], the acids 2a or 2b gave an acid chloride intermediate which could be converted to the corresponding aromatic amides 3a (R = Ac) or 3b (R = Me) [10] by using L-proline methyl ester hydrochloride. Through oxidation-coupling with ferric trichloride [11] as the catalyst, the amides were efficiently cyclized to deliver 9-amido-substituted phenanthrenes 4a (R = Ac) and 4b (R = Me) [10,16], respectively. Finally, the two amides were reduced with LiAlH₄ [17] to give 5a (R = OH) and 5b (R = Me) [18,19]. By treatment with concentrated H₂SO₄/HOAc [20]
or SOCl₂/CH₂Cl₂ [21], 5a was converted to 10 and 11, or 12. Compound 7 was synthesized from 2a and L-valine methyl ester hydrochloride following the same synthetic route as described for 4a.

In our synthetic route, the key step is the reduction of the aromatic amides with aluminum and boron hydrides. In the reduction process we tried LiAlH₄/THF [17], NaBH₄-MeSO₃H/DMSO [22] and NaBH₄-I₂/THF [23] systems to reduce compound 7. First, reduction with the NaBH₄-MeSO₃H/DMSO system did not afford the target product, but rather a series of small amounts of unidentified products. Then, by using LiAlH₄ as the catalyst, the 3-acetoxyl on the phenanthrene ring and the ester group in the amido side chain of 7 were reduced to hydroxyl and hydroxymethyl groups, respectively, as shown for compound 8, but the carbonyl linkage was not reduced. Finally, under the catalysis of NaBH₄-I₂/THF system, 8 was successfully reduced to 9 in a satisfactory yield of 78%. However, compounds 4a and 4b were completely reduced using the LiAlH₄/THF system in one step to give 5a and 5b in consistently high yields of 90% and 93%, respectively.

2.2. Biological activity

The synthesized PBTs were screened for in vitro cytotoxic activity against the H460 human large-cell lung carcinoma cell line with an MTT assay procedure [24]. Adriamycin was used as the reference compound. Table 1 summarizes the structures of 5a, 5b, 8-12 and their cytotoxicities (IC₅₀) against the H460 cell line. All of them exhibited diminished cytotoxic activities to some extent, in comparison to their parent natural products (e.g., tylophorine, IC₅₀ 0.5~1.7 μM) [25].

The results indicate that cytotoxicities of PBTs were influenced by the presence of oxygenic functional substituents on the phenanthrene, and amino acid side chains at the C-9 position of phenanthrene and the linkage between the nitrogen and the phenanthrene. Replacing the 3-methoxyl on the phenanthrene skeleton with a hydroxyl increased the cytotoxic activity, as shown in the comparison of 5b (3-methoxyl, IC₅₀ 53.8 μM) and 5a (3-hydroxyl, IC₅₀ 11.6 μM). Thus, a hydroxyl at the phenanthrene C-3 is quite favorable for cytotoxic activity. In the prolinol side chain, when the terminal hydroxyl was converted to an acetate ester, sulfate ester and chloride, the cytotoxic activities were significantly diminished, it may relate to the removal of the terminal hydrogen bond effect on the assumed biological target [7]. Conserving the C-3 hydroxyl substitution on the phenanthrene skeleton of 5a and opening the pentacyclic prolinol resulted in a more active compound 9 (IC₅₀ 6.1 μM). This result indicates that changing the cyclic side chain to an acyclic one may increase the flexibility of the C-9 side chain, which favors attaining an optimal conformation for binding to an assumed biological target. The amide derivative 8 exhibited very low activity (IC₅₀ 68.1 μM), in agreement with the report of Wei et al. [6].

3. Experimental

3.1. General

Melting points of the synthesized compounds were determined on a Digital Melting Point Apparatus XT4A and are uncorrected. IR spectra were recorded with a Perkin-Elmer model 298 spectrometer (KBr). 1D NMR spectra were recorded on Bruker ARX-300 or Bruker AV-600 spectrometers. ESIMS were recorded with a Finnigan LCQ mass spectrometer and EIMS were measured by an Agilent/HP
3.2. Preparation of PBT derivatives

3.2.1. Synthesis and spectroscopic data of 4-acetoxy-α-(3,4-dimethoxyphenyl)-3-methoxycinnamic acid (2a) and 3,4-dimethoxy-α-(3,4-dimethoxyphenyl)cinnamic acid (2b)

A mixture of 4-hydroxy-3-methoxybenzaldehyde (1a, 0.15 g, 1 mmol), 3,4-dimethoxyphenylacetic acid (0.19 g, 1 mmol), acetic anhydride (2 mL) and triethylamine (0.3 mL) was heated to reflux at 140 °C for 7 h using a CaCl₂ tube. Then the mixture was cooled to room temperature, diluted with water (10 mL) and refluxed at 120 °C for 30 min. The cooled mixture was extracted with EtOAc (3 × 50 mL). The combined organic extracts were washed with saturated brine (150 mL), dried over anhydrous Na₂SO₄, filtered, concentrated in vacuo and the residue recrystallized from methanol to afford 2a (0.32 g, 86%) as pale-yellow crystals. m.p. 166-168 °C; ¹H-NMR (300 MHz, CDCl₃) δ: 2.28 (s, 3H, CH₃CO), 3.47 (s, 3H, OCH₃), 3.82 (s, 3H, OCH₃), 3.90 (s, 3H, OCH₃), 6.67 (d, 1H, J = 1.5 Hz, Ar-H), 6.79 (d, 1H, J = 1.5 Hz, Ar-H), 6.83 (d, 1H, J = 8.2, 1.5 Hz, Ar-H), 6.84 (dd, 1H, J = 8.1, 1.5 Hz, Ar-H), 6.91 (d, 1H, J = 8.1 Hz, Ar-H), 6.92 (d, 1H, J = 8.2 Hz, Ar-H), 7.87 (s, 1H, C=CH); IR (KBr) cm⁻¹ ν: 3421, 3062, 2964, 2839, 1764, 1674, 1619, 1516; EIMS m/z: 372 [M]⁺ (C₂₀H₂₀O₇), 330 (100%) [M-acyl]⁺. Compound 2b (0.31 g, 90%) was synthesized as a deep yellow solid from 3,4-dimethoxy-benzaldehyde (1b, 0.17 g, 1 mmol) and 3,4-dimethoxyphenylacetic acid (0.19 g, 1 mmol) in the same synthetic route as that for 2a. m.p. 217-219 °C; ¹H-NMR (300 MHz, CDCl₃) δ: 3.47 (s, 3H, OCH₃), 3.81 (s, 3H, OCH₃), 3.84 (s, 3H, OCH₃), 3.89 (s, 3H, OCH₃), 6.55 (d, 1H, J = 1.8 Hz, Ar-H), 6.70 (d, 1H, J = 1.8 Hz, Ar-H), 6.72 (d, 1H, J = 8.5 Hz, Ar-H), 6.84 (dd, 1H, J = 8.2, 1.8 Hz, Ar-H), 6.85 (dd, 1H, J = 8.5, 1.8 Hz, Ar-H), 6.92 (d, 1H, J = 8.2 Hz, Ar-H), 7.86 (s, 1H, C=CH); IR (KBr) cm⁻¹ ν: 3448, 3043, 2938, 2835, 1668, 1593, 1516; ESI MS m/z: [M-H]⁻ 343 (C₁₉H₂₀O₆).

3.2.2. Synthesis and spectroscopic data of methyl N-[4-acetoxy-α-(3,4-dimethoxyphenyl)-3-methoxycinnamyl]pyrrolidine-2-carboxylate (3a) and methyl N-[3,4-dimethoxy-α-(3,4-dimethoxyphenyl)cinnamyl]pyrrolidine-2-carboxylate (3b)

To a stirred solution of 2a (0.74 g, 2 mmol) and DMF (0.1 mL) in anhydrous THF (10 mL), oxaly chloride (2 mL, 22.4 mmol) in anhydrous THF (2 mL) was added dropwise. After the addition was complete, the mixture was stirred at room temperature for 4 h. Excess oxaly chloride was removed under reduced pressure. The yellow residue was dissolved in anhydrous CH₂Cl₂ (20 mL), to which was added dropwise a solution of L-proline methyl ester hydrochloride (0.35 g, 2.1 mmol) and pyridine (1 mL, 12.4 mmol) in anhydrous CH₂Cl₂ (4 mL) with stirring. The mixture was stirred overnight at room temperature. Then 1 N HCl (15 mL) was added to the reaction system with vigorous stirring to quench the reaction. The organic layer was seperated and the aqueous layer was extracted with CH₂Cl₂.
The combined organic extracts were washed with saturated brine (90 mL) and dried over anhydrous Na₂SO₄. After filtration and evaporation in vacuo, the residue was purified by column chromatography on silica gel (hexane/EtOAc, 1:1.25 v/v) to afford 3a (0.92 g, 95%) as a colorless solid; m.p. 53-55 °C; ¹H-NMR (300 MHz, CDCl₃) δ: 1.81-1.94 (m, 3H overlapped, 1.5×CH₂), 2.23 (m, 1H, 0.5×CH₂), 2.26 (s, 3H, CH₃CO), 3.30 (m, 2H, CH₂N), 3.53 (s, 3H, OCH₃), 3.73 (s, 3H, OCH₃), 3.76 (s, 3H, OCH₃), 3.85 (s, 3H, OCH₃), 4.55 (m, 1H, CHN), 6.76-6.92 (m, 7H overlapped, 7×Ar-H); IR (KBr) cm⁻¹ ν: 3041, 2954, 2836, 1764, 1745, 1633, 1600, 1513; EIMS m/z: 483 [M⁺] (C₂₆H₂₉NO₈), 441 [M-acyl]⁺, 424 [M-CΟ-C₆H₁₀NO₂-acyl]⁺.

Compound 3b (1.27 g, 96%) was synthesized as a light yellow solid from 2b (1 g, 2.9 mmol) and L-proline methyl ester hydrochloride (0.49 g, 3.0 mmol) by the same synthetic route as that for 3a; m.p. 44-46 °C; ¹H-NMR (300 MHz, CDCl₃) δ: 1.81-1.98 (m, 3H overlapped, 1.5×CH₂), 2.24 (m, 1H, 0.5×CH₂), 3.34 (m, 2H, CH₂N), 3.57 (s, 3H, OCH₃), 3.78 (s, 3H, OCH₃), 3.85 (s, 3H, OCH₃), 3.88 (s, 3H, OCH₃), 4.56 (m, 1H, CHN), 6.70-6.95 (m, 7H overlapped, 7×Ar-H); IR (KBr) cm⁻¹ ν: 3047, 2953, 2836, 1743, 1632, 1600, 1514; EIMS m/z: 455 [M⁺] (C₂₅H₂₉NO₇), 327 [M-CΟ-C₆H₁₀NO₂]+, 299 (100%) [M-CΟ-C₆H₁₀NO₂-acyl]⁺.

3.2.3. Synthesis and spectroscopic data of methyl N-(3-acetoxy-2,6,7-trimethoxyphenanthr-9-ylcarbonyl)pyrrolidine-2-carboxylate (4a) and methyl N-(2,3,6,7-tetramethoxyphenanthr-9-ylcarbonyl)-pyrrolidine-2-carboxylate (4b)

To a solution of 3a (0.48 g, 1 mmol) in anhydrous CH₂Cl₂ (15 mL) was added anhydrous FeCl₃ (0.68 g, 4.3 mmol), and the mixture was stirred at room temperature for 2 h under nitrogen. Then 1 N HCl (1 mL) diluted with saturated brine (10 mL) was added to the stirred reaction system. The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 30 mL). The combined organic extracts were washed with saturated brine (80 mL) and dried over anhydrous Na₂SO₄. After filtration and evaporation, the residue was purified by column chromatography on silica gel (hexane/EtOAc, 1:1.5 v/v) to afford 4a (0.41 g, 85%) as an off-white solid; m.p. 229-231 °C; ¹H-NMR (300 MHz, CDCl₃) δ: 1.88 (m, 1H, 0.5×CH₂), 1.95 (m, 1H, 0.5×CH₂), 2.09 (m, 1H, 0.5×CH₂), 2.37 (m, 1H, 0.5×CH₂), 2.43 (s, 3H, CH₃CO), 2.32 (m, 1H, 0.5×CH₂N), 3.56 (m, 1H, 0.5×CH₂N), 3.84 (s, 3H, CH₃O), 3.97 (s, 3H, CH₃O), 4.00 (s, 3H, CH₃O), 4.01 (s, 3H, CH₃O), 4.84 (dd, 1H, J = 8.4, 4.8 Hz, CHN), 7.29 (s, 1H, Ar-H), 7.59 (s, 1H, Ar-H), 7.70 (s, 1H, Ar-H), 7.77 (s, 1H, Ar-H), 8.14 (s, 1H, Ar-H); IR (KBr) cm⁻¹ ν: 3083, 2953, 2836, 1763, 1743, 1631, 1508; ESI MS m/z: [M+H]+ 482 (C₂₆H₂₇N₂O₈). Compound 4b (1.33 g, 84%) was synthesized as a light yellow solid from 3b (1.6 g, 3.5 mmol) and anhydrous FeCl₃ (2.27 g, 14 mmol) by a similar synthetic route as that of 4a; m.p. 200-202 °C; ¹H-NMR (300 MHz, CDCl₃) δ: 1.77-1.98 (m, 2H overlapped, CH₂), 2.06 (m, 1H, 0.5×CH₂), 2.32 (m, 1H, 0.5×CH₂), 3.22 (m, 1H, 0.5×CH₂N), 3.36 (m, 1H, 0.5×CH₂N), 3.81 (s, 3H, CH₃O), 3.98 (s, 3H, CH₃O), 4.06 (s, 3H, CH₃O), 4.09 (s, 6H, 2×CH₃O), 4.81 (m, 1H, CHN), 7.14 (s, 1H, Ar-H), 7.52(s, 1H, Ar-H), 7.64 (s, 1H, Ar-H), 7.73 (s, 1H, Ar-H), 7.74 (s, 1H, Ar-H); IR (KBr) cm⁻¹ ν: 3077, 2952, 2836, 1741, 1628, 1509; EIMS m/z: 453 [M⁺] (C₂₅H₂₇NO₇), 325 (100%) [M-C₆H₁₀NO₂⁺].
3.2.4. Synthesis and spectroscopic data of N-(3-hydroxy-2,6,7-trimethoxyphenanthr-9-ylmethyl)-L-prolinol (5a) and N-(2,3,6,7-tetramethoxyphenanthr-9-ylmethyl)-L-prolinol (5b)

To a stirred, ice-salt bath cooled solution of 4a (3.1 g, 6.4 mmol) in anhydrous THF (100 mL) was added LiAlH₄ (1.3 g, 39 mmol) in one portion. After being stirred for 30 min kept at 0 °C, the mixture was heated to reflux at 100 °C for 2 h using a CaCl₂ tube. To the cooled mixture were added ice and saturated NaHCO₃ solution (100 mL). After stirring for 10 min, the THF layer was separated and the aqueous layer was extracted with THF (2 × 50 mL). The combined organic extracts were washed with saturated brine (200 mL) and dried over anhydrous Na₂SO₄. After filtration and evaporation, the residue was purified by column chromatography on silica gel (CH₂Cl₂/MeOH, 20:1 v/v) to afford 5a (2.3 g, 90%) as an off-white solid. m.p. 178-180 °C; ¹H-NMR (600 MHz, CDCl₃) δ: 1.70 (m, 1H, 0.5×CH₂), 1.74 (m, 1H, 0.5×CH₂), 1.87 (m, 1H, 0.5×CH₂), 2.02 (m, 1H, 0.5×CH₂), 2.46 (m, 1H, 0.5×CH₂), 2.95 (m, 1H, 0.5×CH₂), 2.97 (m, 1H, CHN), 3.52 (d, 1H, J = 10.8 Hz, 0.5×CH₂O), 3.74 (d, 1H, J = 12.6 Hz, 0.5×ArCH₂N), 7.18 (s, 1H, Ar-H), 7.50 (s, 1H, Ar-H), 7.57 (s, 1H, Ar-H), 7.82 (s, 1H, Ar-H), 7.95 (s, 1H, Ar-H); IR (KBr) cm⁻¹ ν: 3388, 3050, 2953, 2839, 1617, 1513; ESI MS m/z: 398 [M+H]⁺ (C₂₃H₂₇NO₅).

Compound 5b (0.84 g, 93%) was synthesized as a white solid from 4b (1 g, 2.2 mmol) and LiAlH₄ (0.7 g, 21 mmol) by the same synthetic route as that described for 5a; m.p. 217-218 °C; ¹H-NMR (300 MHz, CDCl₃) δ: 1.57-1.67 (m, 2H overlapped, CH₂), 1.79 (m, 1H, 0.5×CH₂), 1.91 (m, 1H, 0.5×CH₂), 2.38 (m, 1H, 0.5×CH₂), 2.80 (m, 1H, 0.5×CH₂), 2.87 (m, 1H, CHN), 3.42 (dd, 1H, J = 10.5, 2.4 Hz, 0.5×CH₂), 3.65 (d, 1H, J = 12.6 Hz, 0.5×ArCH₂N), 3.68 (dd, 1H, J = 10.5, 3.6 Hz, 0.5×CH₂O), 3.96 (s, 3H, CH₃O), 4.00 (s, 3H, CH₃O), 4.02 (s, 3H, CH₃O), 4.04 (s, 3H, CH₃O), 4.39 (d, 1H, J = 12.6 Hz, 0.5×ArCH₂N), 7.12 (s, 1H, Ar-H), 7.44 (s, 1H, Ar-H), 7.52 (s, 1H, Ar-H), 7.70 (s, 1H, Ar-H), 7.74 (s, 1H, Ar-H); IR (KBr) cm⁻¹ ν: 3423, 3050, 2958, 2836, 1617, 1513; ESI MS m/z: 398 [M+H]⁺ (C₂₃H₂₇NO₅).

3.2.5. Synthesis and spectroscopic data of N-[4-acetoxy-α-(3,4-dimethoxyphenyl)-3-methoxy-cinnamyl]-L-valine methyl ester (6)

Compound 6 (1.90 g, 91%) was synthesized as a colorless solid from 2a (1.6 g, 4.3 mmol) and L-valine methyl ester hydrochloride (0.75 g, 4.5 mmol) by a similar synthetic route as that for 3a; m.p. 46-48 °C; ¹H-NMR (300 MHz, CDCl₃) δ: 0.76 (d, 3H, J = 6.9 Hz, CH₃), 0.91 (d, 3H, J = 6.8 Hz, CH₃), 2.13 (m, 1H, CH), 2.27 (s, 3H, CH₃CO), 3.48 (s, 3H, OCH₃), 3.71 (s, 3H, OCH₃), 3.84 (s, 3H, OCH₃), 3.93 (s, 3H, OCH₃), 4.65 (brdd, 1H, J = 8.7, 4.9, CHN), 6.10 (d, 1H, J = 8.7 Hz, NH), 6.62 (d, 1H, J = 1.7 Hz, Ar-H), 6.75 (dd, 1H, J = 8.1, 1.7 Hz, Ar-H), 6.81 (d, 1H, J = 1.7 Hz, Ar-H), 6.85 (d, 1H, J = 8.2 Hz, Ar-H), 6.88 (dd, 1H, J = 8.2, 1.7 Hz, Ar-H), 6.98 (d, 1H, J = 8.2 Hz, Ar-H), 7.77 (s, 1H, C=CH); IR (KBr) cm⁻¹ ν: 3420, 3050, 2961, 2836, 1767, 1740, 1670, 1616, 1513; EIMS m/z: 485 [M]⁺ (C₂₆H₃₁NO₈), 443 (100%) [M-acyl]⁺, 328[M-CO-C₆H₁₂NO₂]⁺, 285[M-acyl-CO-C₆H₁₂NO₂]⁺.
3.2.6. Synthesis and spectroscopic data of N-(3-acetoxy-2,6,7-trimethoxyphenanthr-9-ylcarbonyl)-L-valine methyl ester (7)

Compound 7 (335 mg, 86%) was synthesized as an off-white solid from 6 (390 mg, 0.80 mmol) and anhydrous FeCl₃ (0.53 g, 3.24 mmol) using the same synthetic route as that for 4a; m.p. 205-206 °C; ¹H-NMR (300 MHz, CDCl₃): δ: 1.02 (d, 3H, \( J = 6.9 \) Hz, CH₃), 1.12 (d, 3H, \( J = 6.8 \) Hz, CH₃), 2.36 (m, 1H, CH), 2.43 (s, 3H, CH₃CO), 3.83 (s, 3H, OCH₃), 3.98 (s, 3H, OCH₃), 4.01 (s, 3H, OCH₃), 4.09 (s, 3H, OCH₃), 4.93 (brdd, 1H, \( J = 8.8 \) Hz, 4.9Hz, CHN), 6.64 (d, 1H, \( J = 8.8 \) Hz, NH), 7.29 (s, 1H, Ar-H), 7.73 (s, 1H, Ar-H), 7.76 (s, 1H, Ar-H), 7.79 (s, 1H, Ar-H), 8.11(s, 1H, Ar-H); IR (KBr) cm⁻¹: 3288, 3050, 2961, 2834, 1760, 1742, 1635, 1598, 1508; ESI MS m/z: 482[M-H]⁻ (C₂₆H₂₈NO₈).

3.2.7. Synthesis and spectroscopic data of N-(3-hydroxy-2,6,7-trimethoxyphenanthr-9-ylcarbonyl)-L-valinol (8)

A 100 mL dry three-necked flask was charged with a stirred suspension of LiAlH₄ (61 mg, 1.6 mmol) in anhydrous THF (15 mL) kept at −10~0 °C for 10 min. Compound 7 (150 mg, 0.31 mmol) dissolved in anhydrous THF (15 mL) was added dropwise to the suspension with stirring at −10~0 °C. After the addition was complete, stirring was continued for 10 min, then the warmed reaction system was refluxed at 100 °C for 2 h using a CaCl₂ tube. To the cooled mixture were added ice and saturated NaHCO₃ solution (30 mL). After being stirred for 10 min, the THF layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 30 mL). The combined organic extracts were washed with saturated Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by column chromatography on silica gel (CH₂Cl₂/MeOH, 30:1 v/v) to afford 8 (106 mg, 83%) as a white solid; m.p.182-183 °C; ¹H-NMR (300 MHz, DMSO) δ: 0.94 (d, 3H, \( J = 6.8 \) Hz, CH₃), 0.99 (d, 3H, \( J = 6.8 \) Hz, CH₃), 1.98 (m, 1H, CH), 3.56 (m, 2H, CH₂O), 3.81 (s, 3H, OCH₃), 3.92(s, 3H, OCH₃), 3.96(m, 1H, CHN), 3.98(s, 3H, OCH₃), 7.43 (s, 1H, Ar-H), 7.69 (s, 1H, Ar-H), 7.70 (s, 1H, Ar-H), 7.85 (s, 1H, Ar-H), 7.98 (s, 1H, Ar-H), 8.12(d, 1H, \( J = 9.15 \) Hz, NH); IR (KBr) cm⁻¹: 3296, 3050, 2961, 2834, 1760, 1742, 1635, 1593, 1508; ESI MS m/z: 412 [M-H]⁻ (C₂₃H₂₆NO₆).

3.2.8. Synthesis and spectroscopic data of N-(3-hydroxy-2,6,7-trimethoxyphenanthr-9-ylmethyl)-L-valinol (9)

Compound 8 (41 mg, 0.1 mmol) dissolved in anhydrous THF (5 mL) was added dropwise to a stirred solution of NaBH₄ (20 mg, 0.5 mmol) in anhydrous THF (20 mL) kept at −5~0 °C. Then I₂ (59 mg, 0.23 mmol) dissolved in anhydrous THF (7 mL) was added dropwise to the stirred reaction system at −5~0 °C over 2 h. After the addition was complete, the mixture was warmed to room temperature and heated to reflux at 100 °C for 4 h. Then to the cooled mixture, 1 N HCl (8 mL) was added dropwise with stirring to quench the excess hydride. After the gas evolution ceased, saturated NaHCO₃ solution (8 mL) was added dropwise to neutralize the excess HCl. The THF layer was separated and dried over anhydrous Na₂SO₄. After filtration, a solution of BF₃·Et₂O (2 mL) in THF (5 mL) was added dropwise to the filtrate, then saturated NaHCO₃ solution (10 mL) was added to dissociate the target amine. The THF layer was separated, and the aqueous layer was extracted with
THF (2 × 30 mL). The combined extracts were washed with saturated brine (100 mL) and dried over anhydrous Na₂SO₄. After filtration and evaporation in vacuo, the residue was purified by column chromatography on silica gel (CH₂Cl₂/MeOH, 15:1 v/v) to afford 9 (31 mg, 78%) as an off-white solid; m.p. 224-225 °C; ¹H-NMR (300 MHz, DMSO) δ: 0.91 (d, 3H, J = 6.7 Hz, CH₃), 0.96 (d, 3H, J = 6.7 Hz, CH₃), 2.15 (m, 1H, CH), 2.98 (m, 1H, CHN), 3.60 (m, 1H, O.5×OCH₂), 3.73 (m, 1H, 0.5×OCH₂), 3.92 (s, 3H, OCH₃), 3.93 (s, 3H, OCH₃), 3.98 (s, 3H, OCH₃), 4.45 (m, 2H, ArCH₂N), 7.33 (s, 1H, Ar-H), 7.59 (s, 1H, Ar-H), 7.68 (s, 1H, Ar-H), 7.86 (s, 1H, Ar-H), 7.97 (s, 1H, Ar-H); IR (KBr) cm⁻¹ ν: 3423, 3050, 2927, 2836, 1618, 1508; ESI MS m/z: [M+H]+ 400 (C₂₃H₂₉NO₅).

3.2.9. Synthesis and spectroscopic data of [N-(3-hydroxy-2,6,7-trimethoxyphenanthr-9-ylmethyl)-pyrrolidin-2-yl]methyl acetate (10) and [N-(3-hydroxy-2,6,7-trimethoxyphenanthr-9-ylmethyl)pyrrolidin-2-yl]methyl hydrogen sulfate (11)

To a dry 50 mL three-necked flask charged with 5a (1 g, 2.52 mmol) was added glacial acetic acid (5 mL) at −15 °C. The mixture was stirred for 5 min. Then frozen concentrated sulfuric acid (10 mL) was added dropwise to the stirred system at −15 °C. After the addition was complete, stirring was continued for 20 min. Then the solution was warmed to room temperature and gently heated to 35 °C. Thirty min later, the reaction mixture was cooled to room temperature, poured into water (100 mL) and adjusted to pH 10 with 1N KOH solution. The resulting mixture was extracted with EtOAc (3 × 80 mL). The combined extracts were washed with saturated brine (250 mL), dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The residue was purified by column chromatography on silica gel (CH₂Cl₂/MeOH, 15:1 v/v) to afford 10 and 11. Compound 10 (0.3 g, 27%), a white solid; m.p. 136-137 °C; ¹H-NMR (600 MHz, CDCl₃) δ: 1.66 (m, 3H overlapped, 1.5×CH₂), 2.01 (m, 1H, 0.5×CH₂), 2.06 (s, 3H, CH₃CO), 2.35 (m, 1H, 0.5×CH₂N), 2.84 (m, 1H, 0.5×CH₂N), 2.96 (m, 1H, CHN), 3.68 (d, 1H, J = 13.2 Hz, 0.5×ArCH₂N), 4.03 (s, 3H, OCH₃), 4.05 (s, 3H, OCH₃), 4.09 (s, 3H, OCH₃), 4.13 (m, 1H, 0.5×CH₂O), 4.29 (m, 1H, 0.5×CH₂O), 4.61 (d, 1H, J = 13.2 Hz, 0.5×ArCH₂N), 7.18 (s, 1H, Ar-H), 7.49 (s, 1H, Ar-H), 7.81 (s, 1H, Ar-H), 7.84 (s, 1H, Ar-H), 7.96 (s, 1H, Ar-H); IR (KBr) cm⁻¹ ν: 3430, 3050, 2953, 2834, 1736, 1618, 1509; ESI MS m/z: 438 [M-H]⁻, 440 [M+H]+ (C₂₅H₂₉NO₆).

Compound 11 (0.2 g, 17%), a white solid; m.p. 216-218 °C; ¹H-NMR (600 MHz, DMSO) δ: 1.78 (m, 1H, 0.5×CH₂), 1.80 (m, 1H, 0.5×CH₂), 2.02 (m, 1H, 0.5×CH₂), 2.23 (m, 1H, 0.5×CH₂), 3.10 (m, 1H, 0.5×CH₂N), 3.35 (m, 1H, 0.5×CH₂N), 3.94 (s, 3H, OCH₃), 4.00 (s, 3H, OCH₃), 4.03 (s, 3H, OCH₃), 4.14 (m, 2H, CH₂O), 4.34 (d, 1H, J = 13.2 Hz, 0.5×ArCH₂N), 4.57 (m, 1H, CHN), 5.33 (d, 1H, J = 13.2 Hz, 0.5×ArCH₂N), 7.35 (s, 1H, Ar-H), 7.56 (s, 1H, Ar-H), 7.82 (s, 1H, Ar-H), 7.91 (s, 1H, Ar-H), 8.01 (s, 1H, Ar-H); IR (KBr) cm⁻¹ ν: 3423, 3050, 2924, 2850, 1624, 1614, 1514; ESI MS m/z: 476 [M-H]⁻, 478 [M+H]+ (C₂₃H₂₇NO₈S).

3.2.10. Synthesis and spectroscopic data of N-(3-hydroxy-2,6,7-trimethoxyphenanthr-9-ylmethyl)-2-chloromethylpyrroloidine (12)

A 250 mL dry three-necked flask fitted with a dropping funnel and a condenser was charged with 5a (2 g, 5.04 mmol) and anhydrous CH₂Cl₂ (70 mL). The condenser was fitted with a trap to remove the vapors of hydrogen chloride and sulfur dioxide. Freshly distilled SOCl₂ (5 mL) in anhydrous CH₂Cl₂ (30 mL) was added dropwise to the stirred system at 0 °C. After the addition was complete,
the mixture was warmed to room temperature and heated to reflux at 45 °C for 1.5 h, after which the reaction system was cooled to room temperature and adjusted to pH 10 with saturated Na₂CO₃ solution (100 mL) at 0 °C. The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂ (2 × 50 mL), the combined extracts were washed with saturated brine (200 mL) and dried over anhydrous Na₂SO₄. After filtration and evaporation in vacuo, the residue was purified by column chromatography on silica gel (PE/EtOAc, 3:1 v/v) to afford 12 (1.78 g, 85%) as an off-white solid; m.p. 184-185 °C; ¹H-NMR (300 MHz, CDCl₃) δ: 1.67 (m, 2H overlapped, CH₂), 1.82 (m, 1H, 0.5×CH₂), 2.12 (m, 1H, 0.5×CH₂), 2.26 (m, 1H, 0.5×CH₂N), 2.41 (m, 1H, 0.5×CH₂Cl), 2.72 (m, 1H, 0.5×CH₂N), 3.08 (m, 1H, 0.5×CH₂Cl), 3.83 (d, 1H, J = 13.2 Hz, 0.5×ArCH₂N), 3.97 (d, 1H, J = 13.2 Hz, 0.5×ArCH₂N), 3.98 (m, 1H, CHN), 4.03 (s, 3H, OCH₃), 4.05 (s, 3H, OCH₃), 4.10 (s, 3H, OCH₃), 7.17 (s, 1H, Ar-H), 7.44 (s, 1H, Ar-H), 7.82 (s, 1H, Ar-H), 7.86 (s, 1H, Ar-H), 7.96 (s, 1H, Ar-H); IR (KBr) cm⁻¹ ν: 3442, 3050, 2947, 2832, 1619, 1508; ESI MS m/z: 416 [M+H]+ (C₂₃H₂₆ClNO₄).

3.3. Cytotoxic assay

Cytotoxicities were determined by MTT method [24] using human large-cell lung carcinoma cell line (H460) grown in RPM1-1640 medium plus 10% heat-inactivated fetal bovine serum. The assays were performed in 96-well microtiter plates. Compounds 5a, 5b, 8-12 were dissolved in DMSO and diluted to six different concentrations (10, 3.3, 1.0, 0.33, 0.10, and 0.033 mM) respectively, and each solution was ten-fold diluted to six different concentrations using culture medium (1.0, 0.33, 0.10, 0.033, 0.010, and 0.0033 mM), then 10 µL of each solution was added to 90 µL (about 5,000 cells) culture medium wells. After incubation at 37 °C for 72 hours, 10 µL of MTT (5 mg/mL) was added to each well and incubated for four hours, and then liquid in the wells was removed. DMSO (150 µL) was added to each well. The absorbance was recorded on a microplate reader at wavelength of 590 nm, and the IC₅₀ was defined as 50% reduction of absorbance in the control assay. Compound 5a, 5b, 8-12 showed cytotoxicity against human tumor cell line H460 with IC₅₀ value of 11.6, 53.8, 68.1, 6.1, 46.8, 53.4 and 62.9 µM, respectively. Adriamycin, with an IC₅₀ value of 1.72 µM, was used as positive control.

4. Conclusions

In conclusion, we have synthesized a series of PBT analogues by a facile route and evaluated their in vitro cytotoxic activities against human large-cell lung carcinoma cell line (H460). On this basis, the SAR analysis and potential antitumor activities of these alkaloids are under investigation. Compounds 5a and 9 with a substitution of hydroxyl at position 3 and a hydroxyl at C-9 side chain terminus were the most effective cytotoxic compounds. These results may be helpful for the design of future PBT antitumor reagents, and offer potential application in the discovery of antitumor drugs.

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Sample Availability: Samples of compounds 2–12 are available from the authors.

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