Effect of Doublesynch and Estradoublesynch protocols on estrus induction, conception rate, plasma progesterone, protein, and cholesterol profile in anestrous Gir heifers

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Abstract

Aim: This study aimed to evaluate the efficacy of Doublesynch and Estradoublesynch protocols on estrus induction, conception rates, plasma progesterone, protein, and cholesterol profile in anestrous Gir heifers.

Materials and Methods: In this study, 50 pubertal anestrous Gir heifers were selected from the field and farm conditions. The heifers were dewormed (injection ivermectin, 100 mg, s/c) and supplemented with minerals and vitamins (injection organic phosphorus 800 mg and injection Vitamin AD₃E and Biotin 10 ml i/m) and multi-mineral bolus at 1 bolus daily for 7 days. The heifers were randomly divided into three groups: Doublesynch (n=20), Estradoublesynch (n=20), and control (n=10). The animals were monitored for estrus response, estrus interval, behavioral signs, and conception rates after induced/first, second, and third cycle post-treatment. Blood samples were obtained on day 0, day 9, day 12, and on day 12 post-artificial insemination (AI) for determination of plasma progesterone, protein, and cholesterol profile.

Results: The estrus response rate between Doublesynch and Estradoublesynch protocols was similar between treated heifers (85% and 95%). The interval from the second prostaglandin F2α (PGF₂α) injection to estrus induction did not differ between the groups (63.87±4.19 vs. 58.27±3.83 h). The conception rates following induced estrus (20% vs. 30%), at the second cycle (23.07% vs. 16.66%), at the third cycle (22.22% vs. 30.00%), and the overall conception rate (45% and 55%) within 27.89±5.75 and 26.45±5.48 days were the same across the treatment groups. The mean plasma progesterone concentrations were significantly (p<0.01) higher on day 9 (second PGF₂α injection) and day 12 post-AI compared to day 0 (first PGF₂α injection) and the day of fixed-timed artificial insemination. The concentrations were also significantly (p<0.05) higher in conceived than non-conceived heifers on day 9 of treatment and day 12 post-AI in both the protocols. The mean plasma cholesterol concentrations were significantly higher during peak follicular and luteal phases compared to the initial anestrus phase in both the protocols. The values were also higher in non-conceived than conceived animals in both the protocols. The plasma protein profile was not influenced by the sampling days or conceived and non-conceived status.

Conclusion: The results showed that both Doublesynch and Estradoublesynch protocols resulted in similar estrus induction and conception rates with modulation of plasma progesterone and cholesterol profile in anestrous Gir heifers.

Keywords: cholesterol, conception rate, estrus synchronization, Gir heifers, progesterone, proteins, pubertal anestru.
reported to affect fertility status of bovines [10]. The progesterone hormone is responsible for stimulation of cyclicity, follicular development, and maintenance of pregnancy. Protein deficiency retards the development of reproductive organs and is considered to be a factor responsible for failure or delay in the onset of postpartum estrus [11]. Cholesterol, the most important sterol, is an essential precursor of steroid hormones in the body.

However, studies on the use of recently developed estrus synchronization protocols, namely Ovsynch, controlled internal drug release (CIDR), Doublesynch, and Estradoublesynch in pubertal anestrus heifers of zebu cattle breeds are meager in the literature [12]. Even whether plasma protein and cholesterol levels are altered by these protocols of estrus synchronization is also scarce in cattle [13]. Moreover, the estrus response and conception rates with the use of Doublesynch and Estadoublesynch protocols in both cyclic and acyclic bovines were quite encouraging in earlier studies [14-17]. Hence, this study was aimed to evaluate if Doublesynch and Estradoublesynch protocols induce successful ovulatory estrus, modulate plasma progesterone, protein, and cholesterol profile, and enhance fertility in pubertal anestrus Gir heifers under field conditions.

Materials and Methods

Ethical approval

The prior approval from the Institutional Animal Ethics Committee was obtained for the use of farm animals in this study.

Selection and pre-synchronization treatment of animals

The study was carried out during August 2016 to May 2017 on 50 pubertal anestrus Gir heifers of Baroda District Milk Union, Vadodara, as well as from villages of Junagadh district in Gujarat. The anestrus heifers selected were in the age group of 30-42 months and having average body score (BCS, 2.5-3.5) with smooth, small inactive ovaries that were confirmed by twice rectal palpation 10 days apart. All these animals were initially injected once with 100 mg ivermectin s/c, injection organic phosphorus 800 mg, and multivitamins AD,E 10 ml i/m, and were supplemented with multiminerals bolus at 1 bolus daily for 7 days only to control parasitism, improve nutritional status, and thereby to improve response to hormone therapy. They were then randomly divided into 2 equal groups of 20 heifers each and were subjected to the following two estrus synchronization protocols, keeping one group of 10 animals as an untreated control.

Synchronization protocols

Under Doublesynch protocol, the heifers received i/m injection of Cloprostenol sodium 500 µg on day 0, injection busserelin acetate, a GnRH analog 20 µg on day 2, second injection of Cloprostenol sodium 500 µg on day 9 and 10 µg GnRH on day 11, followed by fixed-time artificial insemination (FTAI) twice at 16 and 24 h later, while in Estradoublesynch protocol, the cows received an injection of estradiol benzoate 1 mg on day 10, in place of the second GnRH injection on day 11 in Doublesynch with FTAI twice at 48 and 60 h post-estradiol injection. Ten anestrus heifers kept without any hormonal intervention and followed for spontaneous estrus served as control. Animals inseminated at induced/spontaneous estrus, if not conceived, were followed for the next two cycles. In non-return cases, pregnancy was confirmed by per-rectal examination 60 days of the last AI. Among non-pregnant animals, the ovarian status was also checked as to whether the animal was cyclic or has turned out to be anestrus again.

Blood sampling and assay procedure

Blood samples were collected from jugular veins in heparinized vacutainers on day 0 – just before treatment, day 9 – at the time of PGF₂α administration, day 12 – induced estrus/FTAI, and on day 12 post-AI. The blood samples were centrifuged at 3000 rpm for 15 min. The plasma separated out was stored in a deep freezer at −20°C with a drop of sodium merthiolate (0.1%). The plasma progesterone concentrations were measured by employing standard radioimmunoassay technique of Kubasic et al. [18]. Labeled antigen (125I), antibody-coated tubes, and standards were procured from Immunotech-SAS, Marseilles-13009, Cedex, France. The plasma total protein and total cholesterol concentrations were determined by Biuret and CHOD/PAP method, respectively, using standard procedures and assay kits with the help of a chemistry analyzer (Mindray, BS 120, Nanshan, Shenzhen-518057, China).

Statistical analysis

The data on estrus induction response and conception rates were analyzed using Chi-square test, and those of plasma progesterone, total cholesterol, and total protein profile using ANOVA and Duncan’s multiple range test or “t-” test employing Statistical Package for Social Sciences (SPSS, USA) software version 20.00 to know the variations between sampling days, treatment groups, and conceived/non-conceived status [19].

Results and Discussion

Estrus induction response and conception rates

The behavioral estrus induced in pubertal anestrus Gir heifers under Doublesynch and Estradoublesynch protocols (85 vs. 95%) and the mean intervals from the second PGF₂α injection to estrus (63.87±4.19 vs. 58.27±3.83 h) did not differ significantly. The intensity of signs of induced estrus under Doublesynch and Estradoublesynch protocols was prominent in 50% and 65%, moderate in 25% and 25%, and weak in 10% and 5% heifers, respectively, while 15% and 5% animals under respective protocols did not show any behavioral estrus signs. Among the control group, only two animals (20%) exhibited spontaneous estrus.
during the period of 90-day follow-up (Table-1). The estrus synchronization rates achieved with Doublesynch and Estradoublesynch protocols concurred well with previous reports in crossbred anestrus cattle [13] and in acyclic buffaloes [14,20], but were higher than 70% each reported by Parida et al. [21] in buffaloes. Moreover, the mean estrus induction intervals recorded with both the protocols corroborated well with earlier reports on anestrus cattle [13] and buffaloes [20,21].

The conception rates in heifers at induced/first, second, third cycle, and overall of 3 cycles following Doublesynch (20.00%, 23.07%, 22.22%, and 45.00%, respectively) and Estradoublesynch protocols (30.00%, 16.66%, 30.00%, and 55.00%, respectively) were statistically same across the treatments (Table-1). Moreover, among the 11 and 9 non-conceived Gir heifers under Doublesynch and Estradoublesynch protocols, 7 and 6 remained cyclic, while 4 and 3 turned out to be anestrus by 60 days of estrus induction/FTAIm as revealed by ovarian status. Overall 50% success rate of fertility was achieved in just average 27 days from the start of two treatment protocols, which was much higher than only 10% found in control group.

The conception rates obtained at induced estrus with Doublesynch and Estradoublesynch protocols were in accordance with the reports of Roodbari et al. [16] as 18.7% and 26.2% in Holstein Friesian cows and with the reports of Prajapati et al. [13] as 40% and 30% in crossbred cows. However, other researchers found much higher conception rates of 43.00% and 59.45% at induced estrus in anestrus cattle following the use of Doublesynch [17] and Estradoublesynch [22] protocols, respectively. Further, the present conception rate at FTAI under Doublesynch protocol is comparable to 28.57% recorded in cattle [15], but it is much lower than 55-58% reported in acyclic buffaloes [20,23].

The present overall conception rate of 45% with Doublesynch protocol is also much lower than 80.00%, 72.80% and 71.43% obtained in anestrus cattle by Prajapati et al. [13], Abubaker et al. [15], and Ozturk et al. [17], respectively. The present overall conception rate of 55% with Estradoublesynch protocol is, however, comparable with 55% and 60% obtained with Doublesynch and Estradoublesynch protocols in an earlier study in cattle [24]. In two more studies, the conception rate with Estradoublesynch protocol was found to be 60%, and 64% in anestrus crossbred cattle and buffaloes, respectively [13-14]. Our results for the first service and overall 3 cycles’ conception rates with Doublesynch and Estradoublesynch protocols were also comparable with those of Patel et al. [25] in anestrus buffaloes from middle Gujarat. The present relatively lower and same conception rates found across treatments could be attributed to identical estrus response, the subject being heifers of just average BCS with small cervix, frightening and struggle while catching for AI from loose housing paddock, and AI being performed by semi-skilled inseminators under field conditions.

| Treatment groups | Estrus induction response (%) | PGF 2α injection to estrus induction interval (h) | Conception rate (%) | Status of NP Gir heifers at 60-day post-AI | Initiation of treatment to conception (days) |
|------------------|-----------------------------|-------------------------------------|---------------------|---------------------------------|----------------------------------------|
| Doublesynch      | 85.00                       | 63.87±4.14                          | 20.00 (4/20)        | Cyclic                          | 27.89±4.75                             |
| Estradoublesynch | 95.00                       | 58.27±5.12                          | 30.00 (6/20)        | Cyclic                          | 26.45±5.48                             |
| Anestrus control | 20.00                       | -                                   | 0.00 (0/1)          | Anestrus                        | 83.00*                                 |

NP=Non-pregnant, PGF 2α: Prostaglandin F2α. *From the initiation of the experiment. Statistically, the values were similar between protocols.
**Plasma progesterone profile**

In bovines, corpus luteum (CL) or luteinized follicle is the principal source of progesterone hormone *in vivo*, and it is responsible for the stimulation of cyclicity, follicular development, and maintenance of pregnancy. Its estimation in blood or milk reflects the ovarian response to gonadotropins and/or prostaglandins and thereby ovarian dynamics or pregnancy. The overall mean plasma progesterone concentrations were low or basal on the day of initiation of treatment in both the protocols. This suggested the true anestrus status of Gir heifers selected for the study. Further, the mean concentrations on day 9 of treatment, i.e., just before the second PGF2α injection, were found to be significantly higher than on day 0 under Doublesynch (1.23±0.12 ng/ml) and Estradoublesynch (1.40±0.20 ng/ml) protocols. This might be due to luteinization of some of the growing follicles and/or ovulation of dominant follicle and formation of CL under the influence of the first GnRH injection. Thereafter, within 2-3 days of the second PGF2α injection, the concentrations dropped significantly to the basal levels with induced estrus, when FTAs were done. These levels again increased significantly (p<0.05) on day 12 post-AI in both the groups with mean values of 2.41±0.38 and 2.55±0.41 ng/ml, respectively. This could be due to ovulatory-induced estruses with development and maintenance of CL and establishment of pregnancy in varying number of animals in each group (Table-2). Further, in Estradoublesynch protocol, the estradiol benzoate injected on day 10 might have triggered positive feedback effect on hypothalamus and pituitary glands resulting in ovulatory luteinizing hormone surge and thereby improved conception rate in that group. The present trend of plasma progesterone compared well with the earlier reports on cattle [26-27] and buffaloes [25].

The mean plasma progesterone concentrations were higher (p<0.05) in conceived than non-conceived cows on day 12 post-AI in both Doublesynch and Estradoublesynch protocols with values of 5.30±0.56 versus 1.69±0.21 and 5.02±0.49 versus 1.49±0.17 ng/ml, respectively. Further, significant differences were also found in conceived and non-conceived animals on day 9 of treatment (Table-2), indicating better luteal activity before induced estrus in conceived animals. Significantly lower plasma progesterone profile seen on day 12 post-AI in non-conceived cows under both the protocols proved that these were the cases of anovulatory estrus and/or luteal insufficiency. Earlier workers [12,28-29] observed significantly (P<0.05) higher plasma progesterone values on day 20-21 post-AI in conceived than non-conceived anestrus zebu cattle with Ovsynch and CIDR protocols. Oyedipse et al. [30] observed nonsignificantly higher plasma progesterone values for the pregnant than non-pregnant heifers following synchronization of estrus and AI. The trend and mean plasma progesterone concentrations found in the present study for the effect of sampling days and conceived and nonconceived statuses of animals closely collaborated with previous reports [4,25,29,31] following use of various estrus induction/synchronization protocols in anestrus cattle and buffaloes.

**Plasma total protein profile**

In heifers under Doublesynch protocol, the overall mean plasma protein concentrations did not differ significantly between sampling days. A similar non-significant variation in mean plasma protein levels in anestrus Gir [28] and crossbreds [32] cattle treated with Ovsynch and CIDR protocols have been reported earlier. In Estradoublesynch protocol, the mean plasma protein levels on the day of FTAI and day 12 post-AI were significantly (p<0.05) higher compared to day 0 and 9 of the protocol. Recent studies [13,33-34] also reported similar findings with different synchronization protocols in postpartum anestrus crossbred cows. Further, the overall mean protein concentrations did not differ significantly between the two protocols at day 0 and day 9. However, heifers under Estradoublesynch protocol had significantly (p<0.05) higher values on the day of FTAI and day 12 post-AI as compared to Doublesynch. Similar findings were reported by Ammu et al. [28] in Ovsynch- and CIDR-treated anestrus Gir cows and by Patel et al. [32] in crossbred cows. The overall pooled mean plasma total protein concentration was significantly lower in anestrus heifers under Doublesynch than Estradoublesynch protocol (7.22±0.09 vs. 7.58±0.07 g/dl). However, in a recent study [24], an inverse trend was observed with higher value in crossbred cattle under Doublesynch protocol than Estradoublesynch protocol (8.19±0.27 vs. 7.31±0.69 g/dl).

Further, the protein profile did not differ significantly between conceived and non-conceived groups at any of the days in any of the protocols. These observations were in close agreement with those in anestrus crossbred cattle with the same protocols [13], and in normal cows [35] that conceived than those did not conceive (6.86±0.10 vs. 6.04±0.10 g/dl). Plasma protein levels change with different stages of reproduction, depending on the feed intake of the animal. Protein deficiency retarded the development of reproductive organs and was considered to be a factor responsible for failure or delay in the onset of postpartum estrus [11]. However, Gentile et al. [36] reported that serum protein level was not related to fertility in dairy cows.

**Plasma total cholesterol profile**

Significant differences were observed in the overall mean plasma cholesterol concentrations between sampling days in heifers under Doublesynch protocol. The values on day 12 post-AI and day of FTAI were significantly higher as compared to those of day 0 and day 9. Similar results were also found.
Table 2: Plasma progesterone, total protein, and total cholesterol concentrations in pubertal anestrus Gir heifers on different days of various estrus induction/synchronization protocols and day 12 post-AI.

| Plasma profile | Synchronization protocol | Status | No. of heifers | Days from treatment/AI |
|----------------|--------------------------|--------|----------------|------------------------|
|                |                          |        |                | D-0        | D-9        | D-12, FTAI | D-12 post-AI | Overall  |
| Progesterone (ng/ml) | Doublesynch              | Conceived | 4              | 0.72±0.08  | 1.80±0.31  | 0.48±0.02  | 5.30±0.56  | 2.08±0.52  |
|                  |                          | Non-concd | 16             | 0.53±0.06  | 1.09±0.11  | 0.56±0.05  | 1.69±0.21  | 0.96±0.08  |
|                  |                          | Overall   | 20             | 0.56±0.05  | 1.23±0.12  | 0.54±0.04  | 2.41±0.36  | 1.19±0.13  |
| Estradoublesynch | Conceived                | 6       | 0.44±0.04      | 2.46±0.40  | 0.53±0.05  | 5.02±0.49  | 2.11±0.42  |
|                  | Non-concd                | 14      | 0.58±0.05      | 0.95±0.07  | 0.54±0.05  | 1.49±0.17  | 0.89±0.07  |
|                  | Overall                  | 20      | 0.54±0.04      | 1.40±0.20  | 0.53±0.04  | 2.55±0.41  | 1.25±0.15  |
| Protein (g/dl)   | Doublesynch              | Conceived | 4              | 7.20±0.09  | 7.27±0.10  | 7.32±0.12  | 7.25±0.12  | 7.26±0.05  |
|                  | Non-concd                | 16      | 7.02±0.24      | 7.27±0.27  | 7.28±0.22  | 7.25±0.22  | 7.21±0.12  |
|                  | Overall                  | 20      | 7.06±0.19      | 7.27±0.21  | 7.29±0.18  | 7.26±0.18  | 7.22±0.09  |
| Estradoublesynch | Conceived                | 6       | 7.41±0.26      | 7.47±0.25  | 7.85±0.20  | 8.09±0.28  | 7.70±0.13  |
|                  | Non-concd                | 14      | 7.17±0.20      | 7.44±0.18  | 7.72±0.17  | 7.76±0.16  | 7.52±0.09  |
|                  | Overall                  | 20      | 7.24±0.16      | 7.45±0.14  | 7.76±0.13  | 7.86±0.14  | 7.58±0.07  |
| Cholesterol (mg/dl) | Doublesynch              | Conceived | 4              | 117.88±8.56 | 120.10±7.40 | 137.68±5.68 | 142.98±10.31 | 129.67±4.63 |
|                  | Non-concd                | 16      | 127.45±3.68    | 134.83±3.33 | 145.81±13.42 | 149.97±3.92 | 139.51±2.17 |
|                  | Overall                  | 20      | 125.53±3.41  | 131.88±3.25  | 144.19±3.61  | 148.58±3.68  | 137.54±2.00 |
| Estradoublesynch | Conceived                | 6       | 120.88±4.41  | 131.75±3.79  | 143.79±2.34  | 137.41±1.24  | 133.46±2.30 |
|                  | Non-concd                | 14      | 134.40±3.17  | 147.47±3.54  | 158.84±4.57  | 160.88±4.91  | 150.39±2.45 |
|                  | Overall                  | 20      | 130.34±2.89  | 142.76±3.14  | 154.32±3.60  | 153.83±4.21  | 145.32±2.03 |

D-0=Day of starting the treatment, D-9=Day of PG injection, While D-12=Fixed-timed artificial insemination, Non-concd=Non-conceived. Means of a trait bearing uncommon superscripts within the row (x, y, z), column (a, b), and conceived and non-conceived (p, q) subgroups within the protocol differ significantly (p<0.05). FTAI=Fixed-timed artificial insemination.
in Estradoublesynch protocol with significantly higher value on day 9 compared to day 0. The mean plasma cholesterol concentration was lowest at initial anestrus phase (day 0), but reached a peak on the day of induced estrus/FTAI (follicular phase) and day 12 post-AI (luteal phase) in both the protocols (Table-2). Bora et al. [33] observed higher plasma cholesterol concentrations on the day of induced estrus and on day 20 than day 10 or 0 of treated respondent postpartum anestrus crossbred cows. However, in earlier studies employing Ovsynch, Cosynch, PRID, etc., the plasma cholesterol levels were not influenced significantly between sampling days in anestrus cattle [4,32,34].

There were no significant differences in protein profile between conceived and non-conceived heifers at any of the sampling days, although the values were apparently higher in non-conceived than conceived animals with significant difference in pooled values (139.51±2.17 vs. 129.67 ±4.63 mg/dl) in heifers under Doublesynch protocol, while in heifers under Estradoublesynch protocol, the concentrations were significantly (p<0.01) higher in non-conceived than conceived animals at all days including overall pooled mean (Table-2). Similar findings were also noted earlier [13] with both these protocols in anestrus cows. The increase in circulatory cholesterol level at estrus induction was opined to be due to the mechanism by which estrogens affect the complex interrelationships of pituitary–thyroid–adrenal functions and the estrogenic agents had an effect on the carbohydrate metabolism in turn caused an increased production of cholesterol in endocrine gland tissue from acetate [37]. Steroid hormones have a direct relationship with cholesterol metabolism. The higher cholesterol level in the cycling animals is indicative of more secretion of steroids during estrus induction due to increased ovarian activity [38]. Kavani et al. [39] opined that the low cholesterol level might have resulted in inadequate synthesis of sex steroid hormones leading to anestrus condition.

Conclusion

From the study, it can be concluded that both Doublesynch and Estradoublesynch protocols resulted in almost similar estrus induction and conception rates, with modulation of plasma progesterone and cholesterol profile. It is, therefore, suggested to use Estradoublesynch protocol due to its cost-effective nature in the successful treatment of anestrus in pubertal Gir heifers.

Authors’ Contributions

DMP and AJD planned and designed the study. The experiment was conducted by NJC, KBV, KKH, and JAP. NJC did laboratory work. AJD and DMP carried out interpretation of results. All authors participated in data analysis, preparation of a draft of the manuscript, and read and approved the same. All authors read and approved the final manuscript.

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Competing Interests

The authors declare that they have no competing interests.

References

1. Kumar, P. and Singhal, L.K. (2006) Gir: Important milch cattle of Western India. Indian Cow, 1: 67-68.
2. Gaur, G.K., Kaushik, S.N. and Garg, R.C. (2003) The Gir cattle breed of India—Characteristics and present status. Anim. Gen. Resources Info., 33: 21-30.
3. Singh, U., Khurana, N.K. and Inder, J. (1998) Plasma progesterone profiles and fertility status of anoestrous zebu cattle treated with norgestomet response estradiol-crg regimen. Theriogenology, 50: 1191-1199.
4. Dhani, A.J., Nakrami, B.B., Hadiya, K.K., Patel J.A. and Shah, R.G. (2015) Comparative efficacy of different estrus synchronization protocols on estrus induction response, fertility and plasma progesterone and biochemical profile in crossbred anestrus cows. Vet. World, 8: 1310-1316.
5. Cirit, U., Ak, K. and Ileri, I.K. (2007) New strategies to improve the efficiency of the Ovsynch protocol in primiparous dairy cows. Bull. Vet. Inst. Pulawy, 51: 47-51.
6. Sirmour, S., Nema, S.P., Singh, B.K. and Shukla, S.P. (2006) Induction of estrus in delayed pubertal crossbred heifers. Indian J. Anim. Reprod., 27: 55-58.
7. Masoumi, R., Badiei, A., Shahneh, A., Kohram, H., Dirandeh, E., Colazo, M.G. (2017) A short presynchronization with PGFα and GnRH improves ovarian response and fertility in lactating Holstein cows subjected to a Heatsynch protocol. Ann. Anim. Sci., 17: 169-177.
8. Dirandeh, E., Roodbari, A.R., Golizadeh, M., Deldar, H., Masoumi, R., Kazemifard, M. and Colazo, M.G. (2015) Administration of prostaglandin Fα 14 d before initiating a G6G or a G7G timed artificial insemination protocol increased circulating progesterone prior to artificial insemination and reduced pregnancy loss in multiparous Holstein cows. J. Dairy Sci., 98: 5414-5421.
9. Mario, B., Roberto, S., Jose, L.M., Leopoldo, J.M., Marcos, H. and Roney, S.R. (2016) Evolution in Fixed-time, from Synchronization of Ovulation to Improve Fertility. Available from: http://www.researchgate.net/publication/268506071. Last accessed on 14-07-2017.
10. Jain, G.C. (1994) Mineral profiles during anoestrus and repeat breeding in bovines. Int. J. Anim. Sci., 9: 241-245.
11. Roberts, S.J. (1986) Veterinary Obstetrics and Genital Diseases. 3rd ed. Ithaca, New York, USA.
12. Khade, N.B., Patel, D.M., Naikoo, M., Dhami, A.J., Sarvaiya, N.P. and Gohel, M.M. (2011) Estrus induction in pubertal anoestrus Gir heifers using different hormone protocols. Indian J. Field Vet., 7: 4-8.
13. Prajapati, A.R., Dhami, A.J., Hadiya, K.K. and Patel, J.A. (2018) Influence of estrus synchronization protocols on plasma profile of progesterone, protein and cholesterol in acyclic Holstein Friesian crossbred cows. Indian J. Vet. Sci. & Biotech., 13(4): 5-11.
14. Mirmahmoudi, R., Souri, M. and Prakash, B.S. (2014) Comparison of endocrine changes, timing of ovulations, ovarian follicular growth, and efficacy associated with
Estradoublesynch and Heatsynch protocols in Murrah buffalo cows. Theriogenology, 82: 1012-1020.

15. Abubaker, P.S., Kurien, M.O., Ghosh, K.N., Simon, S., Anil, K.S. and Becha, B.B. (2013) Ovulation synchronization for improving fertility in postpartum cows. J. Vet. Anim. Sci., 44: 42-45.

16. Roodbari, A.R., Dirandeh, E. and Colazo, M.G. (2015) A New Ovulation Synchronization Protocol (Estradoublesynch) Improves Fertility in Heat-stressed Lactating Holstein Cows. 3rd Int. Congr. Large Animal Practitioners, Tehran, Iran, Feb 2015.

17. Ozturk, O.A., Cirit, U., Baran, A. and Ak, K. (2010) Is Doublesynch protocol a new alternative for timed artificial insemination in anoestrous dairy cows. Theriogenology, 73: 568-576.

18. Kubasic, N.P., Hallauer, G.D. and Brodows, R.G. (1984) Evaluation of direct solid-phase RIA for progesterone, useful for monitoring luteal function. Clin. Chem., 30: 284-286.

19. Sneedcor, G.W. and Cochran, W.G. (1966) Statistical Methods. 8th ed. Iowa State University Press, Ames, Iowa, USA.

20. Mirmahmoudi, R. and Prakash, B.S. (2012) The endocrine changes, the timing of ovulation and the efficacy of the Doublesynch protocol in the Murrah buffalo (Bubalus bubalis). General Comp. Endocrinol., 177: 153-159.

21. Parida, P.K., Mishra, P.C., Mohanty, D.N., Swain, R.K., Barik, A.K. and Das, S. (2015) Successful use of Different Synch Protocols for Estrus Induction in Buffaloes. Proc. XXXI Annual Convention of ISSAR, Veterinary College, Hebbal, Bengaluru, Dec., 3-5. p28.

22. Shrvanan, V., Kulaskekar, K., Krishnakumar, K. and Venkataramanna, S. (2016) Synchronization of Ovulation using Ovsynch and Estradoublesynch Protocols in Crossbred Cattle. Proc. 32rd Annual Convention of ISSAR and National Seminar. Dec. 6-8, Tirupati (AP), India. p29.

23. Dhindsa, S.S., Honparkhe, M., Grewal, R.S. and Brar, P.S. (2016) Fertility enhancement through Doublesynch protocol in buffalo during summer season. Indian Vet. J., 93: 17-19.

24. Sahoo, J.K., Das, S.K., Sethy, K., Mishra, S.K., Swain, R.K., Mishra, P.C. and Sahoo, S.P. (2017) Comparative evaluation of hormonal protocol on the performance of crossbred cattle. Trop. Anim. Health Prod., 49: 259-263.

25. Patel, A.J., Patel, J.A., Dhami, A.J., Prajapati, J.P. and Parmar, S.C. (2018) Estrus induction, fertility and biochemical profile in true anoestrous Surti buffaloes primed with different estrus synchronization protocols. Indian J. Anim. Reprod., 39(2): 36-39.

26. Diaz, T. (1986) Plasma progesterone levels during the estrous cycle of Holstein and Brahman cows, Carora type and crossbred heifers. Theriogenology, 26: 419-432.

27. Bhoraniya, H.L., Dhami, A.J., Naikoo, M., Parmar, B.C. and Sarvaiya, N.P. (2012) Effect of oestrus synchronization protocols on plasma progesterone profile and fertility in postpartum anoestrous Kankrej cows. Trop. Anim. Health Prod., 44: 1191-1197.

28. Ammu, R., Dhami, A.J., Ankita, K. and Pande, A.M. (2012) Postpartum plasma progesterone and metabolic profile in pregnant and non-pregnant Gir cows following estrus synchronization. Indian J. Anim. Prod. Mgmt., 24: 40-44.

29. Desmukh, Y.D., Markandeya, N.M., Chaudhari, R.J. and Deshmukh, V.V. (2010) Progesterone profile in two different protocols of estrus synchronization in postpartum Red Kandhari cows. Indian J. Anim. Reprod., 31: 37-39.

30. Oyedipe, E.O., Vojh, A.A., Marire, B.N. and Pathiraja, N. (1986) Plasma progesterone concentrations during the oestrous cycle and following fertile and non-fertile inseminations of zebu heifers. Br. Vet. J., 142: 41-46.

31. Naikoo, M., Dhami, A.J. and Ramakrishnan, A. (2016) Effect of estrus synchronization on plasma progesterone profile and fertility response in postpartum suckled anoestrous Kankrej cows. Indian J. Anim. Res., 50: 460-465.

32. Patel, K.R., Dhami, A.J., Hadiya, K.K., Savalia, K.K. and Sarvaiya, N.P. (2013) Effect of CIDR and Ovsynch protocols on estrus response, fertility and plasma progesterone and biochemical profile in true anoestrous crossbred cows. Indian J. Anim. Prod. Mgmt., 29: 50-58.

33. Bora, B., Perumal, P., Bonia, K. and Biswas, R.K. (2014) Effect of non-hormonal treatments on postpartum true anoestrous crossbred dairy cows. Int. J. Bio-Resour. Stress Mgmt., 5: 249-255.

34. Borakhatariya, D.N., Panchal, M.T., Dhami, A.J., Hadiya, K.K. and Kalasartya, R.M. (2017) Efficacy of estrus synchronization protocols during summer and winter seasons together with biochemical and minerals profile in anoestrous cows. Indian J. Vet. Sci. Biotech., 13: 9-16.

35. Srivastava, S.K. and Sahni, K.L. (2000) Blood mineral level affecting pregnancy rates in cows and buffaloes. Indian. J. Anim. Sci., 70: 33-34.

36. Gentile, G., Moretti, M., Gaiani, R. and Giordani, L. (1978) Correlation between some biochemical constituents and low fertility in Moderice province. Clin. Vet., 101: 17-24.

37. Purohit, M.K. and Kohli, I.S. (1977) Variations in the blood serum cholesterol level in Rathi cow during estrus. Indian Vet. J., 54: 268-270.

38. Pal, S.K., Mohanty, B.N., Ray, S.K.H. and Mohanty, S.N. (1991) Studies on serum protein, cholesterol and certain enzymes in relation to reproductive status in bovine females. Indian J. Anim. Reprod., 12: 28-29.

39. Kavani, F.S., Sharma, V.K., Siddiquee, G.M. and Vadodaria, V.P. (1987) Serum proteins, ascorbic acid and total cholesterol in anoestrous Kankrej heifers. Indian J. Anim. Reprod., 8: 148-150.

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