High Diversity of Rabies Viruses Associated with Insectivorous Bats in Argentina: Presence of Several Independent Enzootics

Carolina Piñero¹, Federico Gury Dohmen², Fernando Beltran², Leila Martinez¹, Laura Novaro³, Susana Russo³, Gustavo Palacios⁴, Daniel M. Cisterna¹*

¹ Servicio de Neurovirosis, Instituto Nacional de Enfermedades Infecciosas, Administración Nacional de Laboratorios e Institutos de Salud (ANLIS) “Dr. Carlos G. Malbran,” Buenos Aires, Argentina, ² Instituto de Zoonosis “Dr. Luis Pasteur,” Buenos Aires, Argentina, ³ Dirección de Laboratorio y Control Técnico, Servicio Nacional de Sanidad y Calidad Agroalimentaria, Buenos Aires, Argentina, ⁴ National Center for Biodefense and Infectious Diseases, George Mason University, Manassas, Virginia, United States of America

Abstract

Background: Rabies is a fatal infection of the central nervous system primarily transmitted by rabid animal bites. Rabies virus (RABV) circulates through two different epidemiological cycles: terrestrial and aerial, where dogs, foxes or skunks and bats, respectively, act as the most relevant reservoirs and/or vectors. It is widely accepted that insectivorous bats are not important vectors of RABV in Argentina despite the great diversity of bat species and the extensive Argentinean territory.

Methods: We studied the positivity rate of RABV detection in different areas of the country, and the antigenic and genetic diversity of 99 rabies virus (RABV) strains obtained from 14 species of insectivorous bats collected in Argentina between 1991 and 2008.

Results: Based on the analysis of bats received for RABV analysis by the National Rabies system of surveillance, the positivity rate of RABV in insectivorous bats ranged from 3.1 to 5.4%, depending on the geographic location. The findings were distributed among an extensive area of the Argentinean territory. The 99 strains of insectivorous bat-related sequences were divided into six distinct lineages associated with Tadarida brasiliensis, Myotis spp, Eptesicus spp, Histiotus montanus, Lasiurus blossevillii and Lasiurus cinereus. Comparison with RABV sequences obtained from insectivorous bats of the Americas revealed co-circulation of similar genetic variants in several countries. Finally, inter-species transmission, mostly related with Lasiurus species, was demonstrated in 11.8% of the samples.

Conclusions: This study demonstrates the presence of several independent enzootics of rabies in insectivorous bats of Argentina. This information is relevant to identify potential areas at risk for human and animal infection.

Introduction

Rabies is a fatal infection of the central nervous system primarily transmitted by rabid animal bites. Rabies virus (RABV) circulates through two different epidemiological cycles: terrestrial and aerial, where dogs, foxes or skunks and bats, respectively, act as most relevant reservoirs and/or vectors.

In Argentina, successful vaccination and control of canine rabies in the 1980s revealed the importance of bats in RABV transmission. Cases associated with the hematophagous vampire bat Desmodus rotundus are common in endemic areas of Argentina [1]. Two human rabies cases were associated with this species in 1997 and 2001 [2]. No cases have yet been associated with insectivorous bats, a stark contrast to the United States and Canada where these bats are the most common source of indigenous acquired human rabies infections [3]. Thus, it is widely accepted that insectivorous bats are not important vectors of RABV in Argentina despite the great diversity of bat species and the extensive Argentinean territory [4,5,6].

Monoclonal antibodies (N-Mabs) directed against the viral nucleoprotein (produced by the CDC, USA) have allowed for identification of antigenic variants (V) associated with insectivorous bats circulating in Argentina. Two variants were identified: V4 and V6, associated with Tadarida brasiliensis and Lasiurus cinereus, respectively [7,8]. Additionally, although a limited number of specimens were analyzed, partial sequencing of the viral nucleoprotein has revealed at least four genetic variants or lineages associated with other insectivorous bat species [9]. Here we present an extensive study of the geographical distribution of disease associated with insectivorous bats, to infer which species are involved in the maintenance and transmission cycle of RABV in Argentina and to identify possible interspecies transmission patterns.
Passive surveillance of rabies from insectivorous bats

Methods

Passive surveillance of rabies from insectivorous bats

In Argentina, successful vaccination and control of terrestrial rabies in the 1980s revealed the importance of the aerial route in RABV transmission. Current distribution of cases shows a predominance of rabies by hematophagous bats in the Northern regions where rabies is a major public health concern; in contrast, in Central and Southern regions where rabies is not a major public health concern, little surveillance is performed. Based on the analysis of insectivorous bats received for RABV analysis by the National Rabies system of surveillance, the positivity rate of RABV in insectivorous bats in these regions ranged from 3.1 to 5.4%. This rate is comparable to other nations such as the United States (9–10%) where insectivorous bats are an important cause of concern for RABV surveillance systems. Antigenic and genetic analysis of a wide collection of rabies strains shows the presence of multiple endemic cycles associated with six bat insectivorous species distributed among an extensive area of the Argentinean territory and several countries of the Americas. Finally, inter-species transmission, mostly related with Lasiurus species, was demonstrated in 11.8% of the samples. Increased public education about the relationship between insectivorous bats and rabies is essential to avoid human cases and potential spread to terrestrial mammals.

Virus isolates and sequences

A total of 99 rabies samples were obtained from insectivorous bats and bat related cases between 1991 and 2008 from throughout Argentina. All viruses were isolated by intracerebral inoculation in mice as described previously [9]. Eight other RABV isolates from Argentinean insectivorous bats were also analyzed [8]. The host species and geographic location of rabies isolates are shown in Table S1. Samples were identified as follows: The first letters indicate abbreviation for the bat species follow by the number for each sample in the virus repository at SENASA or Instituto de Zoonosis “Dr Luis Pasteur”. Taxonomic characterization of the bats is performed by local specialists at the collection sites.

Antigenic characterization

Antigenic characterization was performed by indirect immunofluorescence using a panel of eight monoclonal antibodies directed against the viral nucleoprotein (C1, C4, C9, C10, C12, C15, C18, C19) kindly provided by the Centers for Disease Control and Prevention, (Atlanta, GA, USA). Positivity reactivity results were analyzed using previously described antigenic variant patterns [7].

RT-PCR and DNA sequencing

Viral RNA was extracted from isolates using TRIzol® (Invitrogen, Carlsbad, CA, USA). Reverse transcription and PCR amplification were achieved with primers 10 g and 304, as previously described [10]. The amplified product was sequenced using a BigDye Terminator v3.1 cycle sequencing kit according to the manufacturer’s protocol with the ABI PRISM® 310 Genetic Analyzer (Applied Biosystems Inc. Foster City, California, USA).

Phylogenetic analysis

A 264-bp region corresponding to the nucleoprotein gene located between nucleotides 1157 and 1420 and amino acids 363 to 450 (numbered according to strain SAD B19) was analyzed [11]. Raw sequence data were first edited using CHROMAS software (version1.3, Mc Carthy 1996, Griffith University, Queensland, Australia). Complete alignment was performed with Clustal X 1.8 [12]. The alignment was analyzed using Kimura 2 parameters as a method of substitution and Neighbor-Joining model to reconstruct the phylogenetic tree (MEGA version 4.1) [13]. The statistical significance of the phylogenies constructed was estimated by bootstrap analysis with 1000 pseudoreplicate data sets [14,15].

GenBank accession numbers

Partial nucleoprotein gene sequences described in this study were deposited in the GenBank database under the following accession numbers JF738250–JF738348.

Results

Geographic distribution of rabies in insectivorous bats

Between 1995 and 2007, public health authorities notified 1096 cases of animal rabies: dogs (33.6%), cattle (45.9%) or insectivorous bats (15.4%). The success of vaccination programs meant that 200 rabies cases reported for 2008–09, canine rabies only represented 14.5%, while rabies transmitted by insectivorous bats increased to 38%, and cattle remained constant (45.5%) [16]. Geographic distribution of rabies in Argentina between 2008 and 2009 is shown in Table 1. Importantly, Northeastern (NE) and Northwestern (NW) regions show a predominance of rabies by vampires while in Central and South regions, almost all reported cases are associated with insectivorous bats.

Table 1. Regional distribution of rabies in Argentina, 2008–2009.

| Species             | Northeastern | Northwestern | Central | South | Total |
|---------------------|--------------|--------------|---------|-------|-------|
| Cattle              | 59           | 29           | 6       | 0     | 94    |
| Insectivorous bats  | 2            | 3            | 70      | 4     | 79    |
| Dog                 | 4            | 23           | 0       | 0     | 27    |
| Fox                 | 3            | 0            | 0       | 0     | 3     |
| Total               | 68           | 55           | 76      | 4     | 203   |

Argentina is divided into five epidemiological regions. Cuyo region has not reported cases of rabies in the period of this study. doi:10.1371/journal.pntd.0001635.t001
From 1991 to 2008, Laboratories of National Rabies Network tested 4536 insectivorous bats for RABV from Buenos Aires city (CABA) and 23 from provinces of Argentina. Of these, 207 were found to be positive. Three provinces accounted for 92.2% of all reported cases of rabies in bats: Buenos Aires province, 70 cases (33.7%); CABA, 63 cases (30.4%); and Santa Fe province, 38 cases (18.4%). National positivity rate of rabies caused by insectivorous bats could not be calculated since the total number of bat specimens received from all provinces for rabies investigation was unavailable. However, local positivity rate could be determined in CABA, 3.5% (63/1792), Buenos Aires province 5.4% (70/1307) and Santa Fe province 3.1% (38/1113).

Of the bats infected with rabies virus, 65.4% of RABV-positive cases of *T. brasiliensis* were detected in Buenos Aires province, and 60.0% of *T. brasiliensis* Antigenic variant 4 (AgV4) was identified in 57 bats: 53 (92.9%) in CABA, 5.5% (38/697) in Buenos Aires (nine species). Great diversity was observed in Santa Fe (10 species) and Buenos Aires (nine species). *T. brasiliensis* accounted for 92.7% of the total in CABA (four species).

### Antigenic typing

Antigenic typing was performed on 103 isolates (Table S1). Antigenic variant 4 (AgV4) was identified in 57 bats: 53 (92.9%) in *T. brasiliensis*; the rest (7.1%) in *M. molossus* (n = 2), *Eumops patagonicus* (n = 1) and unidentified bat (n = 1). This variant showed the greatest distribution, scattered throughout the provinces. Antigenic variant 6 (AgV6) was identified in 25 specimens: 11 *L. cinereus* (44.0%), 4 *L. rhabdodens* (16.0%), and the remaining 10 (40.0%) in *Eptesicus* spp (n = 1), *M. molossus* (n = 1), *M. levis* (n = 1), *M. nigricans* (n = 1), four unidentified bats, and one dog. This variant mainly circulated in central provinces. Finally, 22 isolates exhibited 11 different atypical reaction pattern (ARP) (Table 2). These isolates were from *Myotis* (n = 7), *Eptesicus* (n = 9), *Histiotus* genus (n = 3), *T. brasiliensis* (n = 1), *L. blossevillii* (n = 1) bats and one from a cat.

### Molecular characterization

A total of 107 insectivorous bat-related sequences segregated into six distinct lineages. This was well supported by significant bootstrap values and clearly differentiated from those related to rabies in terrestrial animals (Figure 1).

The first lineage (TB) included 51 samples from *T. brasiliensis*, two from *M. molossus*, one from *Eumops patagonicus* and one from an unidentified bat. All isolates were typed as V4. This lineage exhibited a high nucleotide and amino acid similarity (99.0% and 100.0%, respectively). It showed the amino acidic residue N394 in the nucleoprotein that is characteristic of this genetic variant (Table 3). Rabies isolates obtained from *Myotis*, *Eptesicus* and *Histiotus* bats grouped in three highly diverse lineages (nucleotide intra-lineage distance of 6.7%, 2.0% and 3.4%, respectively). All these isolates resulted in several atypical reaction patterns with N-Mabs (ARP). *Myotis* lineage (MY) was subsequently divided in five sublineages (MY1–5) less-well supported, scattered throughout central provinces (Figure 2). The *Eptesicus* lineage (EP) was also divided in three sublineages (named EP1–3). The three EP sublineages resulted in different ARP circulating in Santa Fe, Entre Rios and Buenos Aires. Both lineages MY and EP presented a similar coding signature containing the amino acids A385 and L419. Lastly, the *Histiotus montanus* lineage (HM) included three isolates from southern provinces (Santa Cruz and Chubut). It was further divided in two sublineages (HM1 and HM2) in correspondence with their antigenic and nucleotide sequence clustering with nucleotide and amino acid distance of 4.3% and 2.3%; amino acid difference at position 369 (Q369K). Furthermore, they showed different reaction with C12 N-Mab (Table 3).

**Table 2. Antigenic patterns of bat rabies viruses from Argentina.**

| Associated source or reservoir | Patterns of reaction (N-Mabs) | Antigenic Variant |
|-------------------------------|-------------------------------|-------------------|
| Dog/mongoose                  | C1  | C4  | C9  | C10 | C12 | C15 | C18 | C19 |
| Dog                           | +   | +   | +   | +   | +   | +   | -   | +   | V1  |
| Desmodus rotundus             | -   | +   | +   | +   | +   | -   | -   | -   | V3  |
| Tadarida brasiliensis         | -   | +   | +   | +   | +   | -   | -   | -   | V4  |
| Lasiurus cinereus             | +/- | +   | +   | +   | +   | -   | -   | -   | V6  |
| LB                            | +/- | +   | +   | +   | +   | -   | -   | -   | ARPb |
| MY1                           | +   | +   | +   | +   | -   | -   | -   | -   | ARP |
| MY2                           | -   | +   | +   | +   | +   | -   | -   | -   | ARP |
| MY3                           | -   | +   | +   | +   | +   | +   | -   | -   | ARP |
| MY4                           | -   | +   | +   | +   | -   | -   | -   | -   | ARP |
| MY5                           | -   | +   | +/- | +   | +   | -   | -   | -   | ARP |
| HM1                           | -   | +   | +   | +   | -   | -   | -   | -   | ARP |
| HM2                           | -   | +   | +   | +   | -   | -   | -   | -   | ARP |
| EP1                           | +   | +   | +   | +   | -   | -   | -   | -   | ARP |
| EP2                           | +/- | +/- | +/- | +   | +   | -   | -   | -   | ARP |
| EP3                           | -   | +   | +   | +   | -   | -   | -   | -   | ARP |

*LB, Lasiurus blossevillii reactivity pattern found for sample Lb658-BA03; MY1–5, Myotis spp. reactivity pattern found for samples Epf457-SF04 and My594-CHA05 (MY1), Epf457-SF02 (MY2); Myf10-SF03 (MY3); Stnmbt16 (MY4) and Myf140-SF07 (MY5); HM1–2, Histiotus montanus reactivity pattern found for samples Stchmb80 (HM1) and Hm620-CHU07 and Hm580CHU07 (HM2); EP1–3, Eptesicus spp. reactivity pattern found for samples Epf1288-SF08 and Epf787-SF07 (EP1), Epf1202-SF06 and Epf497-SF05 (EP2) and Epf062-BA03 and Epb458-SF05 (EP3).

**ARP**, atypical reaction pattern.

doi:10.1371/journal.pntd.0001635.t002

---

Rabies in Bats from Argentina
Figure 1. Phylogenetic relationships among Argentinean isolates. Analysis was based on 264 nt of the nucleoprotein gene. A tree was constructed based on Kimura and neighbor joining parameters. TB, Tadarida brasiliensis; DR, Desmodus rotundus; MY, Myotis spp.; HM, Histiotus montanus; EP, Eptesicus spp; LB, Lasiurus blossevillii; LA, Lasiurus cinereus; TR, Terrestrial.

doi:10.1371/journal.pntd.0001635.g001
Finally, the last two lineages (named LB and LC), were detected in three different species of *Lasiurus* bats. LB included a single sample of *L. blossevilli*, antigenically typed as ARP. LC included 30 samples, obtained from 13 *L. cinereus* (43.3%), four *L. ega* (13.3%) and others species: one *Eptesicus* spp., one *E. bonariensis*, one *M. molossus*, three *Myotis* spp., one dog, and four samples from unclassified bats. LC is one of the less divergent lineages (0.3%) and was formed from 30 samples. Virus of the lineage LC were mainly typed as V6 (n = 25, 83.3%); ARP (n = 4, 13.3%) and V4 (n = 1; 3.4%) Deduced amino acid sequences revealed a characteristic amino acid S414 in both LC and LB lineages. Moreover, a change at position 419 (S419T) was identified between LC and LB lineages.

Molecular relationships between rabies associated with insectivorous bats of Argentina and the Americas

Analysis of nucleoprotein RABV sequences associated with insectivorous bats in Argentina and the Americas revealed several monophyletic clusters associated with specific bat species. This was consistent with previous analyses [17,18,19] (Figure 3). RABV samples obtained from *T. brasiliensis* bats from Argentina, Brazil, Chile and Uruguay segregated into a monophyletic cluster. RABV sequences associated with *Myotis* species occurring in South America were divided in two heterogeneous lineages formed by samples from Argentina, Chile and Uruguay or Brazil. Instead, RABV sequences associated with *Myotis* species occurring in North America (USA, Canada) formed a different cluster. Similarly, RABV samples from *Eptesicus* bats of Argentina and Brazil, represented primarily by *f. furinalis* subspecies, were grouped in two clusters. These clusters did not show relationships with the genetic clusters associated with *f. fuscus* subspecies from USA and Canada. Strains that were recovered in Argentina and Chile from *Histiotus* bats circulated in both countries as at least two sublineages less-well supported. Finally, RABV sequences obtained from different species of *Lasiurus* from the Americas were divided in three monophyletic clusters associated with subspecies *blossevilli*, *borealis* and *cinererus/ega*. An Argentinean sample obtained from *L. blossevilli* (Lb658-BA03) grouped with two Brazilian *L. blossevilli*-related RABV strains (BR-BAT27 and BR-BAT13). RABV isolates obtained from *L. cinererus* and *L. ega* from Argentina, Brazil, Chile, Uruguay, USA, Mexico and Canada grouped in a cluster with the highest nucleotide homogeneity (99.5%).

### Discussion

The current understanding of the epidemiology of rabies in Argentina states that the vast majority of cases in the Northern region are associated with hematophagous bats. Very few cases of terrestrial rabies were also detected in the Northwestern (Salta and Jujuy) and Northeastern provinces (Formosa and Chaco) during 2010. The rest of the country has been considered terrestrial rabies-free since the early 1980s. This epidemiological situation makes it difficult to estimate the real impact of the insectivorous bat rabies in our country. In the north, very few cases have been detected probably masked by endemic bovine rabies. While in the center and south, the absence of terrestrial rabies has led to low level of awareness among general public, public health officials and health administrators. As a result, rabies associated with insectivorous bats and its potential consequent implications in public and animal health have been largely neglected in Argentina.

Our study reveals that the positivity rate of rabies in insectivorous bats received in the laboratory for analysis ranges from 3.1 to 5.4%. This proportion is comparable to other countries such as the United States (9–10%) where insectivorous bats are the only cause of concern for RABV surveillance systems, and other South American countries (Brazil (1.3%) and Chile (4.2%) [20,21,22]. Fortunately, <1% of natural bat population have been shown to be infected [23]. Thus, the risk of contracting rabies from insectivorous bats is low. However,
evidence indicates that many of the human cases of rabies resulted from exposures to bats that were not recognized or reported [3]. Consequently, prevention of human infection with bat rabies virus variants remains an important public health concern. On the other hand, emergence of rabies in terrestrial hosts after spillover from chiropteran reservoirs has been described but does not typically result in sustained transmission. However, if host switching of rabies virus variants occur, once established could become enzootic in new reservoir species [24]. Therefore, special attention should be paid to unusual epidemiological patterns of terrestrial rabies transmission in new geographic areas.

Figure 2. Geographic distribution of RABV sublineages associated with insectivorous bats from Argentina.
doi:10.1371/journal.pntd.0001635.g002
Antigenic characterization utilizing the eight monoclonal antibodies developed by the CDC is widely used in Latin America for RABV surveillance. However, in some instances, antigenic analysis is unable to identify RABV isolates obtained from several insectivorous bat species because these isolates produced atypical reaction patterns (unrelated to previously described virus reservoirs) [8]. In those cases, partial genetic analysis of the viral nucleoprotein sequence allowed further characterization [25] allowing the identification of lineages or genetic variants maintained by insectivorous bat species in an independent enzootic cycle. Indeed, in our work, we identified six RABV lineages that were specifically associated with specific bat species. Moreover, genetic analysis allowed us to differentiate some of the previously accepted antigenic variants in independent sublineages that appear to be related with different geographical or ecological niche behaviors. *Tadarida brasiliensis* maintains circulation of its own antigenic (AgV4) with high degree of nucleotide and amino acid homogeneity in Argentina, Chile, Brazil and Uruguay. In contrast, analysis of RABV isolates recovered from *Myotis* and *Eptesicus* species showed a high antigenic diversity that could be related to the gregarious and non-migratory habits of these species [17,18]. The elevated antigenic diversity of RABV sustained by *Eptesicus* or *Myotis* species can complicate definitive strain typing. This could be the case of the only *Myotis* associated strain from Chile, which was assigned to an apparent AgV3 using a reduced panel of eight N-MAbs by Yung et al. [26], but our genetic characterization revealed its real clustering into *Myotis* group.

All Argentinean isolates obtained from *Histiotus montanus*, clustered in a single genetic group along with strains from Chile confirming that this species is its own viral variant reservoir. There is little information about this species other than it lives a solitary life and migrates seasonally between Argentina and Chile [27]. Although both countries are separated by the Andes Chain, an important natural barrier, low-lying passages of the mountain allow different species of bats and terrestrial mammals to move between the countries and thus spreading this viral variant [28].

Members of the genus *Lasiana* typically are solitary and migratory bats. In our study, rabies samples from *L. blossevillii, cinerus* and *ege* were analyzed. A RABV isolate from Argentinean *L.blossevillii* bat was distant to others obtained from the Lasiusinus genus but clustered with others from the same bat species from Brazil. Indeed, it yields a previously unrecognized genetic lineage circulating in Argentina.

On the other hand, antigenic and molecular analysis of rabies isolates from *Lasiusinus cinerus* confirmed that this species maintains its own antigenic (AgV6) and genetic variant as previously reported [29]. Despite showing a wide geographical distribution (Canada to Argentina), RABV isolates from this species exhibited a high degree of nucleotide and amino acid homogeneity, which could be explained by its ability to transport its own specific strain during its long migration pattern. Two rabies samples of Chilean Lasiusinus have been assigned to AgV4 (*T.hisilensis* reservoir) by Yung et al. [26], but our phylogenetic analysis showed that they grouped with Lasiusinus strains (AgV6). This apparent discrepancy could be explained by the inadequate use of the N-Mabs panel which could lead to confusion between AgVs. The difference between both variants falls only in the reactivity with monoclonal C1. According to the N-Mabs manufacturer’s instructions, all negative or diminished reactions should be confirmed by furthers tests of the samples with a 10-fold less-dilute antibody [30].

Molecular characterization of RABV isolates revealed that inter-species transmission is a relatively common event. Eleven (11.8%) of the 93 bat samples tested showed to be infected by a variant supported by another bat species. Cross-species transmission is facilitated by several types of species life-history traits and perhaps environmental variables structuring communities [31]. In the case of Lasiusinus, these seem to have an important role in these events, since we have identified this variant nearly in all bat species studied. Although, *L.cinerus* generally roosts in isolation, it has been observed occasional aggressive encounters at share roosts or during flight, which could promote viral transmission.

The findings of this study demonstrate the presence of rabies in several species of insectivorous bats throughout Argentina. Phylogenetic analysis of an extensive collection of rabies strains obtained from 14 species over a 17-year period shows complex epidemiological patterns characterized by the presence of multiple endemic cycles and relatively frequent inter-species transmission that are affected by several ecological aspects such as migration patterns, roosting and habitat. The establishment of viral variants associated with specific bat species can assist in the epidemiological investigation of cases of human rabies associated with bats and potential events spread to terrestrial mammals.

Supporting Information

Table S1 Rabies isolates from insectivorous bats of Argentina.

| Species                | AgV     |
|-----------------------|---------|
| L. blossevillii        | AgV4    |
| L. cinerus             | AgV6    |

Acknowledgments

The authors wish to thank all the professionals and non-professionals from the National Rabies Laboratories Network of the Ministry of Health of Argentina.

Author Contributions

Conceived and designed the experiments: CP DMC. Performed the experiments: CP LM. Analyzed the data: FGD FB GP. Contributed reagents/materials/analysis tools: LN SR. Wrote the paper: CP DMC.

References

1. Delpietro HA, Russo RG (1996) Ecological and epidemiologic aspects of the attacks by vampire bats and paralytic rabies in Argentina and analysis of the proposals carried out for their control. Rev Sci Tech 15: 971–984.
2. Larghi OP, Delpietro H (2004) Casos de rabia humana transmitida por murciélagos, Argentina. In: Cacchione RA, Durcal R, Larghi OP, eds. Tema de Zoonosis II. Buenos Aires: Asociacion Argentina de Zoonosis, pp 135–138.
3. De Serres G, Dallaire F, Cote M, Skowronski DM (2008) Bat rabies in the United States and Canada from 1950 through 2007: human cases with and without bat contact. Clin Infect Dis 46: 1329–1337.
4. Barquez RM, Mares M, Braun JK The bats of Argentina; University Mo/TT, editor. Texas.
5. Mena Segura C (2008) Rabia. Ciudad de Buenos Aires: 1964–2007. 44 años de lucha, logros alcanzados y progresos. In: Cacchione RA, Durcal R, Martino P,
eds. Temas de Zoonosis IV. Buenos Aires: Asociacion Argentina de Zoonosis. pp 139–150.

6. Amasino CF, Gury Dohmen F, de Gaetano J, Mena Segura C, Palazzolo A (2003) A case of bat rabies in a cat in the province of Buenos Aires, Argentina. Rev Sci Tech 22: 1021–1027.

7. Delpietro HA, Gury-Dhomen F, Larghi OP, Mena-Segura C, Abramdo L (1997) Monoclonal antibody characterization of rabies virus strains isolated in the River Plate Basin. Zentralbl Veterinarmed B 44: 477–483.

8. Cisterna D, Bonaventura R, Caillou S, Pozo O, Andreau ML, et al. (2005) Antigenic and molecular characterization of rabies virus in Argentina. Virus Res 109: 139–147.

9. Koprowski H (1996) The mouse inoculation test. In: Merlin FX, Kaplan M, Koprowski H, eds. Laboratories techniques in rabies. 4th ed. Geneva: World Health Organization. pp 80–97.

10. Smith JS (1995) Rabies virus. In: Murray PR BE, Pfaller MA, Tenover FC, Yolken R, eds. Manual of clinical microbiology. Washington: American Society for Microbiology Press. pp 997–1003.

11. Konzelmann KK, Cox JH, Schneider LG, Thiel HJ (1990) Molecular cloning and complete nucleotide sequence of the attenuated rabies virus SAD B19. Virology 175: 485–499.

12. Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, et al. (2007) Clustal W and Clustal X version 2.0. Bioinformatics 23: 2947–2948.

13. Tamura K, Dudley J, Nei M, Kumar S (2007) MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. Mol Biol Evol 24: 1596–1599.

14. Hillis DM (1993) An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. Syst Biol 42: 102–192.

15. Zander R (2004) Minimal Values for Reliability of Bootstrap and Jackknife Proportions, Decay Index, and Bayesian Prior Proportionality. Phyloinformatics 2: 1–13.

16. Ministerio de Salud de la Nacion (2010) Boletin Epidemiologico Nacional, Buenos Aires, Argentina. http: http://msal.gov.ar/htm/site/sala_situacion/boletines_ultimos.asp.

17. Kobayashi Y, Sato K, Kato M, Itou T, Cunha EM, et al. (2007) Genetic diversity of bat rabies viruses in Brazil. Arch Virol 152: 1995–2004.

18. Velasco-Villa A, Oeciari LA, Juarez-Idas V, Gomez-Sierra M, Padilla-Medina I, et al. (2006) Molecular diversity of rabies viruses associated with bats in Mexico and other countries of the Americas. J Clin Microbiol 44: 1697–1710.

19. Davis PI, Bourhy H, Holmes EC (2006) The evolutionary history and dynamics of bat rabies virus. Infect Genet Evol 6: 464–473.

20. Cunha EM, Silva LH, Lara Mdo C, Nassar AF, Albas A, et al. (2006) Bat rabies in the north-northeastern regions of the state of Sao Paulo, Brazil: 1997–2002. Rev Saude Publica 40: 1082–1086.

21. Favi CM, Rodriguez AL, Espinosa MC, Yung PV (2008) Rabies in Chile: 1949–2005. Rev Chilena Infectol 25: S8–S13.

22. Blanton JD, Robertson K, Palmer D, Rupprecht CE (2009) Rabies surveillance in the United States during 2008. J Am Vet Med Assoc 235: 676–680.

23. Klieg HH, Turnfeld AS, Ellison JA, Barrowd EF, Barcay RM (2011) Rabies prevalence in migratory tree-bats in Alberta and the influence of roosting ecology and sampling method on reported prevalence of rabies in bats. J Wildl Dis 47: 64–77.

24. Leslie MJ, Messenger S, Rohde RE, Smith J, Cheshier R, et al. (2006) Bat-associated rabies virus in Skunks. Emerg Infect Dis 12: 1274–1277.

25. Nadin-Davis SA, Huang W, Armstrong J, Casey GA, Rahlod C, et al. (2001) Antigenic and genetic divergence of rabies viruses from bat species indigenous to Canada. Virus Res 74: 139–156.

26. Yang V, Favi M, Fernandez J (2002) Genetic and antigenic typing of rabies virus in Chile. Brief report. Arch Virol 147: 2197–2205.

27. Nowak RM (1994) Walker’s bats of the world. Baltimore: The Johns Hopkins University Press.

28. Mann-Fisher G (1978) Los pequen ˜os mamiferos de Chile (marsupiales, quiropteros, edentados y roedores); Artigas JN, ed. Concepcion: Editorial de la Universidad de Concepcion. 342 p.

29. Sheeler-Gordon LL, Smith JS (2001) Survey of bat populations from Mexico and Paraguay for rabies. J Wildl Dis 37: 592–593.

30. Díaz AM, Papo S, Rodriguez A, Smith JS (1994) Antigenic analysis of rabies-virus isolates from Latin America and the Caribbean. Zentralbl Veterinarmed B 41: 153–160.

31. Stricker DG, Turmelle AS, Vouhof MJ, Kuzmin IV, McCracken GF, et al. (2010) Host phylogeny constrains cross-species emergence and establishment of rabies virus in bats. Science 329: 676–679.