Phase II study of intrabone single unit cord blood transplantation for hematological malignancies

Makoto Murata,1 Yoshinobu Maeda,2 Masayoshi Masuko,2 Yasushi Onishi,4 Tomoyuki Endo,5 Seitaro Terakura,1 Yuichi Ishikawa,3 Chisako Iriyama,1 Yoko Ushijima,1 Tatsunori Goto,2 Nobuharu Fujii,2 Mitsune Tanimoto,2 Hironori Kobayashi,6 Yasuhiko Shibasaki,6 Noriko Fukuhara,4 Yoshihiro Inamoto,3 Ritsuro Suzuki,2 Yoshihisa Kodera,9 Tadashi Matsushita,10 Hitoshi Kiyoi,1 Tomoki Naoe1,11 and Tetsuya Nishida1

1Department of Hematology and Oncology, Nagoya University Graduate School of Medicine, Nagoya; 2Department of Hematology and Oncology, Okayama University, Okayama; 3Department of Stem Cell Transplantation, Niigata University Hospital, Niigata; 4Department of Hematology and Rheumatology, Tohoku University Hospital, Sendai; 5Department of Hematology, Hokkaido University Graduate School of Medicine, Sapporo; 6Department of Hematology, Endocrinology and Metabolism, Niigata University Graduate School of Medical and Dental Science, Niigata; 7Department of HSCT Data Management and Biostatistics, Nagoya University Graduate School of Medicine, Nagoya; 8Department of Oncology and Hematology, Shimane University Cancer Center, Izumo; 9Department of Promotion for Blood and Marrow Transplantation, Aichi Medical University School of Medicine, Nagakute; 10Department of Transfusion Medicine, Nagoya University Hospital, Nagoya; 11National Hospital Organization Nagoya Medical Center, Nagoya, Japan

Key words
Cord blood transplantation, engraftment, hematological malignancy, intrabone, non-myeloablative conditioning

Correspondence
Makoto Murata, Department of Hematology and Oncology, Nagoya University Graduate School of Medicine, 65 Tsurumai, Showa, Nagoya, Aichi 466-8550, Japan. Tel: +81-52-744-2141; Fax: +81-52-744-2157; E-mail: mmurata@med.nagoya-u.ac.jp

Funding information
Practical Research Project for Allergic Diseases and Immunology (15ek010100h0003), Japan Agency for Medical Research and Development and a Grant-in-Aid for Scientific Research (KAKENHI 15k09498); Japan Society for the Promotion of Science.

Received March 27, 2017; Revised May 26, 2017; Accepted June 1, 2017

Cancer Sci 108 (2017) 1634–1639
doi: 10.1111/cas.13291

The outcomes of cord blood transplantation with non-irradiated reduced-intensity conditioning for hematological malignancies need to be improved because of graft failure and delayed engraftment. Intrabone infusion of cord blood cells has the potential to resolve the problems. In this phase II study, 21 adult patients with hematological malignancy received intrabone transplantation of serological HLA-A, B, and DR ≥4/6 matched single cord blood with a median number of cryopreserved total nucleated cells of 2.7 × 10^7/kg (range, 2.0–4.9 × 10^7/kg) following non-irradiated fludarabine-based reduced-intensity conditioning. Short-term methotrexate and tacrolimus were given as graft-versus-host disease prophylaxis, and granulocyte colony-stimulating factor was given after transplantation. No severe adverse events related to intrabone injection were observed. The cumulative incidences of neutrophils ≥0.5 × 10^9/L, reticulocytes ≥1%, and platelets ≥20 × 10^9/L recoveries were 76.2%, 71.4%, and 76.2%, respectively, with median time to recoveries of 17, 28, and 32 days after transplantation, respectively. The probability of survival with neutrophil engraftment on day 60 was 71.4%, and overall survival at 1 year after transplantation was 52.4%. The incidences of grade II–IV and III–IV acute graft-versus-host disease were 44% and 19%, respectively, with no cases of chronic graft-versus-host disease. The present study showed the safety of direct intrabone infusion of cord blood. Further analysis is required to confirm the efficacy of intrabone single cord blood transplantation with non-irradiated reduced-intensity conditioning for adult patients with hematological malignancy. This study was registered with UMIN-CTR, number 000000865.

Cord blood transplantation is a treatment option for patients with hematological malignancy.1 The comparison of outcomes of CBT and HLA-A, B, C and DRB1 allele-matched unrelated bone marrow transplantation for adult patients with acute leukemia showed no significant difference in overall survival between these groups.2 Additionally, the recent development of RIC allows elderly patients and patients with comorbidities to benefit from CBT.3,4 However, CBT with RIC, especially a non-irradiated conditioning regimen, has yet to be optimized, due in part to graft failure or delayed engraftment.4–7

In terms of successful engraftment, higher cell dose and better HLA-matching have an advantage.8 Donor-specific anti-HLA antibodies should also be considered.9 Double-unit CBT was established in the early 2000s.10 However, a recent prospective randomized study comparing single-unit and double-unit CBT after myeloablative conditioning for children and adolescents indicated that neutrophil recoveries were similar, and double-unit CBT was associated with lower and slower platelet recovery and a higher incidence of GVHD.11 Expansion of cord blood cells ex vivo before transplantation has been developed,12,13 and further clinical studies to confirm efficacy for reduction of transplant-related mortality are in progress. However, these strategies require specialized techniques, drugs, or cells, and cannot be adopted everywhere. Direct intrabone transplantation of cord blood cells, established by Frassoni et al.,14 potentially ensures engraftment and shortens the time for hematological recovery in single CBT. Neither special devices nor particular skills are required for this technique. Fortunately, no severe intrabone injection-related complications have been reported so far.

Based on these data, a phase II study was undertaken to assess the efficacy of intrabone transplantation of a single cord blood unit following non-irradiated RIC for adult patients with hematological malignancy.

© 2017 The Authors. Cancer Science published by John Wiley & Sons Australia, Ltd on behalf of Japanese Cancer Association. This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.
Materials and Methods

Patients. Patients were eligible for the study if they had hematological malignancy, needed CBT, were ≥55 years or 16–54 years old with a hematopoietic stem cell transplantation-specific comorbidity index ≥1.(15) and had available cord blood with serological HLA-A, B, and DR ≥4/6 matched and with cryopreserved TNCs at least 2 × 10^9/kg. Cord blood was obtained from the Japanese Red Cross Cord Blood Banks (Hokkaido, Kanto-Koshinetsu, Kinki, and Kyushu), Chubu Cord Blood Bank, Hyogo Cord Blood Bank, and Japan Cord Blood Bank Network (defunct) in Japan. This study was approved by the ethics committees of the following institutes: Nagoya University, Okayama University, Niigata University, Tohoku University, and Hokkaido University (all Japan). Patients were enrolled at these institutes between September 2007 and July 2015.

Transplantation. All patients received non-irradiated fludarabine-based RIC on the basis of patient characteristics and pre-transplant therapies. All patients received tacrolimus (initial dose 0.02–0.03 mg/kg) and short-term methotrexate (15 mg/m^2 on day 1, and 10 mg/m^2 on days 3 and 6; or 10 mg/m^2 on day 1, and 7 mg/m^2 on days 3 and 6) as GVHD prophylaxis. All patients received granulocyte-colony-stimulating factor starting from 5–7 days after transplantation until neutrophil engraftment.

Intrabone injection. Intrabone injection was carried out as described previously.(14) Cord blood was thawed, washed with a saline solution plus dextran and human albumin,(16) resuspended in approximately 10 mL of the solution, and aliquoted in two to four syringes. After local anesthesia, standard bone marrow aspiration needles were inserted into iliac bone. Aspiration of <0.5 mL bone marrow was done to assess that the needle was securely inserted into the bone marrow cavity. Then approximately 5 mL cord blood cell suspension was gently infused. This procedure was repeated for all remaining aliquots across the iliac crest.

Definitions. Neutrophil engraftment was defined as neutrophil recovery ≥0.5 × 10^9/L before day 60 with donor chimerism ≥90%. Donor chimerism was determined by quantitative PCR for informative short tandem repeats using DNA extracted from peripheral blood T cells.(17) The time to neutrophil recovery was defined as the first of 3 consecutive days of absolute neutrophil count ≥0.5 × 10^9/L. The time to reticulocyte and platelet recoveries was defined as the first of 3 consecutive days of reticulocytes ≥1%, platelet count ≥20 × 10^11/L, and platelet count ≥50 × 10^11/L without transfusion support. Acute GVHD and chronic GVHD were diagnosed and graded according to established criteria.(18,19)

Statistical analysis. The primary end-point was the probability of survival with neutrophil engraftment on day 60 after transplantation. Twenty-one patients were required to provide at least 80% power to differentiate the primary end-point of 80% with the width of the 95% CI being 25%. Assuming a drop-out rate of one patient, 22 patients were required. The probabilities of hematopoietic recovery and relapse were estimated on the basis of cumulative incidence curves. Overall survival after transplantation was estimated according to the Kaplan–Meier method. The groups were compared using the log–rank test. All tests were two-sided, and P < 0.05 was considered significant. The data were analyzed by Stata statistical software (StataCorp, College Station, TX, USA).

Results

Patient and transplantation characteristics. Twenty-two patients with hematological malignancy were enrolled in this study, but 1 patient did not receive intrabone CBT due to general deterioration after enrollment. The characteristics of 21 patients who received intrabone CBT are summarized in Table 1. The median age was 57 years (range, 38–66 years). Fourteen patients (66%) had leukemia (six in first complete remission, three in third complete remission, and five in non-complete remission), six (29%) had malignant lymphoma (two in first complete remission, one in second complete remission, and two in non-complete remission), and one (5%) had myelodysplastic syndromes in refractory anemia with excess blasts-1. The median number of cryopreserved TNCs in a cord blood unit was 2.7 × 10^9/kg (range, 2.0–4.9 × 10^9/kg), and the median number of cryopreserved CD34+ cells in it was 0.92 × 10^9/kg (range, 0.44–3.14 × 10^9/kg). Serological HLA-A, B, and DR 6/6, 5/6, and 4/6 matched cord blood in the host-versus-graft direction were used for two (10%), six (29%), and 13 (61%) patients, respectively. In one patient (no. 1), HLA-antibody against cord blood HLA was detected. All patients received fludarabine 150–180 mg/m^2 (150 mg/m^2 for one patient and 180 mg/m^2 for 19 patients) and cyclophosphamide 60–120 mg/kg (60 mg/kg for one patient and 120 mg/kg for 19 patients), with the exception of one patient who received fludarabine 125 mg/m^2 and melphalan 140 mg/m^2. No patient received TBI.

Intrabone injection. Intrabone injection of cord blood was carried out with local anesthesia in the patient’s room. No patient required i.v. sedative drugs, such as propofol. No severe adverse events were observed. Mild swelling of the skin at the injection site was observed in one patient, but it resolved spontaneously.

Hematopoietic recovery. Patient no. 1 showed no signs of any hematopoietic recovery and received transplantation with other cord blood on day 42, but died of multiorgan failure without a sign of engraftment of the second cord blood. Four patients (nos. 15, 19, 20, and 21) showed autologous hematopoietic recovery. Patient no. 20 received 150 mg/m^2 fludarabine and 120 mg/kg cyclophosphamide as preconditioning, and patient no. 21 received 180 mg/m^2 fludarabine and 60 mg/kg cyclophosphamide, whereas patients nos. 15 and 19 received 180 mg/m^2 fludarabine and 120 mg/kg cyclophosphamide, which was the preconditioning used in patients with successful engraftment (Table 1). Donor HLA-specific antigen was not detected in these four patients. Sex mismatch in host-versus-graft direction (combination of female patient and male donor) was not associated with a higher incidence of autologous hematopoietic recovery. The cumulative incidences of neutrophils ≥0.5 × 10^9/L on day 60, reticulocytes ≥1% on day 60, platelets ≥20 × 10^11/L on day 100, and platelets ≥50 × 10^11/L on day 100 recoveries were 76.2% (95% CI, 56.9–91.3%), 71.4% (51.8–88.3%), 76.2% (56.9–91.3%), and 66.7% (46.9–85.1%), respectively (Fig. 1). For those who achieved hematopoietic recovery, the median time to neutrophils, reticulocytes, platelets ≥20 × 10^11/L, and platelets ≥50 × 10^11/L recoveries were 17, 28, 32, and 38 days, respectively (Table 1). ABO major and/or minor mismatch did not affect reticulocyte recovery.

Neutrophil recovery was evaluated according to the numbers of TNCs and CD34+ cells, and HLA matching in the host-versus-graft direction (Table 2). There was a significant difference.
Table 1. Characteristics of patients with hematological malignancies and outcomes of intrabone single unit cord blood transplant

| Pt no. | Age, years | Diagnosis | Disease stage | Sex Pt/Do | ABO Pt/Do | DSA Preconditioning | Cord blood | Time to recovery, days | Acute GVHD, grade | Chronic GVHD Status at day 365 | Cause of death |
|--------|-------------|-----------|---------------|-----------|-----------|---------------------|------------|----------------------|-----------------|-----------------------------|-----------------|
| 1      | 53          | AML       | Non-CR        | Ma/Ma     | O/AB      | Pos (125) + M (140) | 2.7        | 0.69                 | 4/6             | NE                          | Dead (72)†       |
| 2      | 38          | AML       | 3CR           | Fe/Ma     | B/O       | Neg (180) + C (120) | 4.9        | 3.14                 | 4/6             | NE                          | Dead (244) VAHS |
| 3      | 56          | ATLL      | Non-CR        | Fe/Fe     | A/B       | Neg (180) + C (120) | 2.3        | 0.83                 | 5/6             | NE                          | Alive           |
| 4      | 62          | AML       | Non-CR        | Ma/Ma     | A/A       | Neg (180) + C (120) | 2.4        | 0.70                 | 6/6             | NE                          | Alive           |
| 5      | 59          | AML       | Non-CR        | Ma/Fe     | O/O       | Neg (180) + C (120) | 3.4        | 0.75                 | 6/6             | NE                          | Alive           |
| 6      | 55          | DLBCL     | Non-CR        | Ma/Fe     | AB/O      | Neg (180) + C (120) | 2.9        | 0.95                 | 5/6             | NE                          | Alive           |
| 7      | 59          | NK/T Leu | 1CR           | Ma/Fe     | AB/O      | Neg (180) + C (120) | 3.1        | 1.29                 | 5/6             | NE                          | Alive           |
| 8      | 64          | NK/T Lym | 1CR           | Ma/Fe     | AB/A      | Neg (180) + C (120) | 2.8        | 2.05                 | 4/6             | NE                          | Alive           |
| 9      | 56          | Ph+ ALL  | 1CR           | Ma/Fe     | B/C       | Neg (180) + C (120) | 2.4        | 0.73                 | 4/6             | NE                          | Alive           |
| 10     | 60          | T-LBL     | 1CR           | Ma/Fe     | A/A       | Neg (180) + C (120) | 2.9        | 0.96                 | 5/6             | NE                          | Alive           |
| 11     | 51          | ATLL      | 1CR           | Ma/Fe     | B/O       | Neg (180) + C (120) | 3.7        | 0.63                 | 5/6             | NE                          | Alive           |
| 12     | 61          | ALL       | 1CR           | Fe/Fe     | AB/A      | Neg (180) + C (120) | 3.8        | 1.35                 | 5/6             | NE                          | Alive           |
| 13     | 55          | AML       | 3CR           | Ma/Fe     | AB/A      | Neg (180) + C (120) | 4.8        | 2.60                 | 4/6             | NE                          | Alive           |
| 14     | 66          | MDS       | RAE1          | Ma/Fe     | O/O       | Neg (180) + C (120) | 2.0        | 1.10                 | 4/6             | NE                          | Alive           |
| 15     | 41          | NK/T Lym | 2CR           | Fe/Ma     | F/A       | Neg (180) + C (120) | 2.6        | 0.54                 | 4/6             | NE                          | Alive           |
| 16     | 60          | AML       | Non-CR        | Ma/Fe     | O/B       | Neg (180) + C (120) | 2.9        | 0.51                 | 4/6             | NE                          | Alive           |
| 17     | 63          | DLBCL     | Non-CR        | Ma/Fe     | O/AB      | Neg (180) + C (120) | 2.3        | 0.44                 | 5/6             | NE                          | Alive           |
| 18     | 47          | AML       | 3CR           | Ma/Fe     | B/A       | Neg (180) + C (120) | 2.5        | 1.60                 | 4/6             | NE                          | Alive           |
| 19     | 60          | Ph+ ALL  | 1CR           | Fe/Fe     | AB/O      | Neg (180) + C (120) | 2.1        | 0.72                 | 5/6             | NE                          | Alive           |
| 20     | 57          | T-LBL     | Non-CR        | Ma/Fe     | A/A       | Neg (150) + C (120) | 1.2        | 1.02                 | 4/6             | NE                          | Alive           |
| 21     | 50          | ALL       | 1CR           | Ma/Fe     | A/A       | Neg (180) + C (60)  | 2.2        | 1.02                 | 4/6             | NE                          | Alive           |

†Number in parentheses after “dead” indicates survival days after transplantation. ‡Number in parentheses after “relapse” indicates the day when the patient had a relapse. 1CR, first CR; 2CR, second CR; 3CR, third CR; ABO, ABO blood type; ALL, acute lymphoblastic leukemia; AML, acute myeloid leukemia; ATLL, adult T-cell leukemia/lymphoma; C, cyclophosphamide (mg/kg); CR, complete remission; DLBCL, diffuse large B-cell lymphoma; Do, donor; DSA, donor HLA-specific antigen; F, fludarabine (mg/m²); Fe, female; GVH, graft-versus-host direction; GVHD, graft-versus-host disease; HLA-m, human leukocyte antigen-matching; HVG, host-versus-graft direction; M, melphalan (mg/m²); Ma, male; MDS, myelodysplastic syndromes; N, neutrophils ≥0.5 x 10⁹/L; NA, not available; NE, not evaluable; Neg, negative; NK/T Leu, natural killer/T-cell leukemia; NK/T Lym, extranodal natural killer/T-cell lymphoma, nasal type; P2, platelets ≥20 x 10⁹/L; P5, platelets ≥50 x 10⁹/L; Ph+, Philadelphia chromosome-positive; PML, progressive multifocal leukoencephalopathy; Pos, positive; Pt, patient; R, reticulocytes ≥1%; RAEB1, refractory anemia with excess blasts-1; RMF, remission failure; T-LBL, T lymphoblastic lymphoma; TNC, total nuclear cells; VAHS, virus-associated hemophagocytic syndrome; VOD, veno-occlusive disease.
in the incidence of neutrophil recovery between patients receiving ≤2.7 × 10^7/kg TNCs (n = 11) and >2.7 × 10^7/kg TNCs (n = 10) (54.5% vs 100%, P = 0.0046).

Other outcomes. Other outcomes are summarized in Table 1. Of 16 patients who showed neutrophil recovery, seven (43.8%) and three (18.8%) developed grade II–IV and III–IV acute GVHD, respectively. No patients developed chronic GVHD. Thirteen (81.3%) patients developed cytomegalovirus antigenemia within 1 year of transplantation (data not shown). The probability of relapse was 75.0% (95% CI, 12.5–91.2%). Patient no. 16 obtained neutrophil recovery with full donor chimera, but died of veno-occlusive disease on day 42. The probability of survival with neutrophil engraftment on day 60 after transplantation was 71.4% (95% CI, 51.8–88.3%). Overall survival at 1 year after transplantation was 52.4% (95% CI, 29.7–70.9%). Six patients died of their original hematological malignancies. Four other patients died of graft failure, virus-associated hemophagocytic syndrome, leukoencephalopathy, and veno-occlusive disease, respectively. Patient no.16, who developed veno-occlusive disease, had advanced disease (acute myeloid leukemia with 33% blasts in the bone marrow just prior to preconditioning) and iron overload (red blood cell transfusion ≥30 times before transplantation, with approximately 1000 ng/mL ferritin) as risk factors for this condition.

Discussion

The present study indicated the feasibility of intrabone single CBT with non-TBI RIC regimens for adult patients with hematological malignancy. Since Rizzieri et al. (20) reported successful engraftment in two cases, several studies have confirmed the feasibility of CBT with TBI-containing RIC. Barker et al. (21) reported that the cumulative incidence of neutrophil recovery was 76% for adult patients with hematological malignancy preconditioned with fludarabine + busulfan + TBI 2Gy and 94% for adult patients with fludarabine + cyclophosphamide + TBI 2Gy. Miyakoshi et al. (22) reported that the cumulative incidence of neutrophil recovery was 93% for adult patients with advanced hematological malignancy preconditioned with fludarabine + melphalan + TBI 4 Gy. The most recently published paper from the Japanese Society for Hematopoietic Cell Transplantation reported that TBI regimens were significantly associated with a higher rate of neutrophil engraftment in CBT with RIC. (23) However, non-TBI RIC regimens in CBT often result in dismal outcomes. In a study from Duke University (Durham, NC, USA), in which 10 patients received fludarabine and cyclophosphamide without TBI followed by single CBT, only three (30%) engrafted with donor hematopoietic cells. (4) Another study from Duke University, in which 10 patients received fludarabine and busulfan without

![Fig. 1. Hematopoietic recoveries after intrabone single cord blood transplantation.](image)

Table 2. Neutrophil recovery according to the numbers of total nucleated cells and CD34+ cells, and HLA matching in the host-versus-graft (HVG) direction

| Neutrophil recovery, % | P-value |
|-----------------------|---------|
| Total nucleated cells |         |
| ≤2.7 × 10^7/kg (n = 11) | 54.5 | 0.0046 |
| >2.7 × 10^7/kg (n = 10) | 100 | |
| CD34+ cells |       |
| ≤0.92 × 10^5/kg (n = 11) | 81.8 | 0.6850 |
| >0.92 × 10^5/kg (n = 10) | 70.0 | |
| HLA-matching in HVG direction |     |
| 4/6 (n = 13) | 69.2 | 0.2720 |
| 5/6 or 6/6 (n = 8) | 87.5 | |

Cancer Sci | August 2017 | vol. 108 | no. 8 | 1637
© 2017 The Authors. Cancer Science published by John Wiley & Sons Australia, Ltd on behalf of Japanese Cancer Association.
TBI followed by double CBT, showed that only two (20%) achieved donor engraftment. A study from Japan, in which fludarabine and busulfan were used as the preconditioning for single CBT, reported a cumulative incidence of engraftment on day 60 of 53%. Given this result, we regarded as clinically relevant if at least 55% of patients achieved a primary endpoint. We also reported that only four (40%) of 10 patients were alive with donor engraftment on day 60 after preconditioning consisting of fludarabine + melphalan. In the present study, the cumulative incidence of neutrophil recovery was 76.2%, suggesting that intrabone transplantation can be an option to improve engraftment in CBT with non-TBI RIC regimens.

Nonetheless, it is not clear whether direct intrabone infusion of cord blood can overcome engraftment failure in CBT. Frasoni et al. undertook a phase II study in which 32 adult patients were treated by intrabone single CBT, mostly with myeloablative preconditioning, and the cumulative incidence of neutrophil recovery on day 44 was 85%. Okada et al. conducted a phase I study in which 10 adult patients were treated by intrabone transplantation of unwashed cord blood with TBI-containing RIC, and nine (90%) patients successfully achieved neutrophil recovery. These results are better than previous studies of i.v. CBT for adult patients, but they may be inconsistent with animal experiments showing that intrabone transplantation is 10–15 times more efficient in the engraftment of cord blood cells than i.v. transplantation. In fact, a retrospective analysis comparing outcomes of intrabone single CBT with i.v. double CBT after myeloablative preconditioning did not show a significant difference in the incidence of neutrophil recovery on day 60 (84% vs 91%, P = 0.62). Interestingly, the present study showed that there was a correlation between the number of TNCs and the engraftment rate, suggesting that the success or failure of neutrophil engraftment after intrabone CBT might also depend on the number of TNCs. Further study is required to determine whether intrabone injection truly improves the engraftment rate in CBT for adult patients.

In the present study, four patients showed autologous hematopoietic recovery. Two patients (nos. 20 and 21) received less intensive preconditioning than that used in patients with successful engraftment. This could be one of the causes of their autologous hematopoietic recovery. However, more importantly, another two patients (nos. 15 and 19) received the same preconditioning as patients with successful engraftment. Thus, the preconditioning regimen in intrabone CBT needs to be much optimized, and it seems interesting to consider whether the combination of intrabone infusion with more intensive non-TBI or TBI-containing regimens could result in much better engraftment rates in CBT.

The fact common to the present study and previous reports, except for one, is rapid engraftment in intrabone CBT. The median time to neutrophil recovery was 17–23 days, and platelet recovery was 32–41 days, which are much faster than i.v. CBT. A retrospective comparative study showed that neutrophil recovery was significantly faster in intrabone single CBT than in i.v. double CBT (23 vs 28 days, P = 0.001). This was the same for platelet recovery (36 vs 49 days, P = 0.003). Rapid engraftment may be the greatest benefit of intrabone transplantation because it may contribute to reducing infection, use of antibacterial drugs, blood transfusion, and medical costs.

Although a low incidence of acute GVHD may be a feature of intrabone transplantation, the present study demonstrated incidences of grade II–IV and III–IV acute GVHD of 43.8% and 18.8%, respectively, which were comparable to those in i.v. CBT for Japanese patients. Although a low incidence of relapse after intrabone transplantation is reported, the present study showed that the probability of relapse was 75.0%, which may be due to inclusion of many cases with advanced or high-risk hematological malignancy, or it may be due to weakness of conditioning regimens.

In conclusion, intrabone transplantation of a single cord blood unit using a non-TBI RIC regimen provides an opportunity for patients with hematological malignancy who are unable to be exposed to irradiation at the time of preconditioning for several reasons, such as preservation of fertility, second transplantation following first transplantation with high-dose TBI, and lack of availability of TBI equipment in the hospital. Even in hospitals where TBI is available, if a non-TBI regimen is selected, a transplant schedule can be made at the best time for patients, without dependence on the radiation schedule. In addition, non-TBI RIC may be especially useful for patients receiving urgent CBT for engraftment failure. Prospective randomized studies with homogeneous preconditioning and GVHD prophylaxis regimens are required to determine whether intrabone CBT provides definitively better outcomes for patients with hematological malignancy than i.v. CBT.

Acknowledgments

The authors would like to thank the medical staff at each transplantation center. This study was supported in part by a Practical Research Project for Allergic Diseases and Immunology (15ek0510010h0003 to M. Murata) from the Japan Agency for Medical Research and Development and a Grant-in-Aid for Scientific Research (KAKENHI 15K09498 to M. Murata) from the Japan Society for the Promotion of Science.

Disclosure Statement

The authors have no conflict of interest.

Abbreviations

- CBT: cord blood transplantation
- CI: confidence interval
- GVHD: graft-versus-host disease
- RIC: reduced-intensity conditioning
- TBI: total body irradiation
- TNC: total nuclear cell

References

1. Utsumoniya A, Choi I, Chihara D, Seto M. Recent advances in the treatment of adult T-cell leukemia-lymphomas. Cancer Sci 2015; 106: 344-51.
2. Terakura S, Atsuta Y, Tsukada N et al. Comparison of outcomes of 8/8 and 7/8 allele-matched unrelated bone marrow transplantation and single-unit cord blood transplantation in adults with acute leukemia. Biol Blood Marrow Transplant 2016; 22: 330–8.
3. Cutler C, Ballen K. Reduced-intensity conditioning and umbilical cord blood transplantation in adults. Bone Marrow Transplant 2009; 44: 667–71.
4. Chao NJ, Koh LP, Long GD et al. Adult recipients of umbilical cord blood allografts who received nonmyeloablative preparative regimens. Biol Blood Marrow Transplant 2004; 10: 569–75.
5. Komatsu T, Narimatsu H, Yoshimi A et al. Successful engraftment of mismatched unrelated cord blood transplantation following reduced intensity preparative regimen using fludarabine and busulfan. *Ann Hematol* 2007; 86: 49–54.

6. Horwitz ME, Morris A, Gasparetto C et al. Myeloablative intravenous busulfan/fludarabine conditioning does not facilitate reliable engraftment of dual umbilical cord blood grafts in adult recipients. *Biol Blood Marrow Transplant* 2008; 14: 591–4.

7. Narimatsu H, Watanabe M, Kohno A et al. High incidence of graft failure in unrelated cord blood transplantation using a reduced-intensity preparative regimen consisting of fludarabine and melphalan. *Bone Marrow Transplant* 2008; 41: 753–6.

8. Terakura S, Azuma E, Murata M et al. Hematopoietic engraftment in recipients of unrelated umbilical cord blood is affected by the CD34+ and CD8+ cell doses. *Biol Blood Marrow Transplant* 2007; 13: 822–30.

9. Hanajiri R, Murata M, Sugimoto K et al. Integration of humoral and cellular HLA-specific immune responses in cord blood allograft rejection. *Bone Marrow Transplant* 2015; 50: 1187–94.

10. Barker JN, Weisdorf DJ, Wagner JE. Creation of a double chimera after the transplantation of umbilical-cord blood from two partially matched unrelated donors. *N Engl J Med* 2001; 344: 1870–71.

11. Wagner JE Jr, Eapen M, Carter S et al. One-unit versus two-unit cord-blood transplantation for hematologic cancers. *N Engl J Med* 2014; 371: 1685–94.

12. de Lima M, McNiece I, Robinson SN et al. Cord-blood engraftment with ex vivo mesenchymal-cell coculture. *N Engl J Med* 2012; 367: 2305–15.

13. Wagner JE Jr, Brunstein CG, Boitano AE et al. Phase II/I trial of StemRegen-1 expanded umbilical cord blood hematopoietic stem cell supports testing as a stand-alone graft. *Cell Stem Cell* 2016; 18: 144–55.

14. Frassoni F, Gualandi F, Podestà M et al. Direct intrabone transplantation of unrelated cord-blood cells in acute leukemia: a phase I/I study. *Lancet Oncol* 2008; 9: 831–9.

15. Sorror ML, Giralt S, Sandmaier BM et al. Hematopoietic cell transplantation specific comorbidity index as an outcome predictor for patients with acute myeloid leukemia in first remission: combined FHCRC and MDACC experiences. *Blood* 2007; 110: 4606–13.

16. Rubinstein P, Dobrila L, Rosenfield RE et al. Processing and cryopreservation of placental/umbilical cord blood for unrelated bone marrow reconstitution. *Proc Natl Acad Sci U S A* 1995; 92: 10119–22.

17. Imahashi N, Ohashi H, Terakura S et al. Chimerism status after unrelated donor bone marrow transplantation with fludarabine-melphalan conditioning is affected by the melphalan dose and is predictive of relapse. *Ann Hematol* 2015; 94: 1139–48.

18. Przepiorka D, Weisdorf D, Martin P et al. 1994 Consensus conference on acute GVHD grading. *Bone Marrow Transplant* 1995; 15: 825–8.

19. Filipovich AH, Weisdorf D, Pavletic S et al. National Institutes of Health consensus development project on criteria for clinical trials in chronic graft-versus-host disease: I. Diagnosis and staging working group report. *Blood Marrow Transplant* 2005; 11: 945–56.

20. Rizzieri DA, Long GD, Vredenburgh JJ et al. Successful allogeneic engraftment of mismatched unrelated cord blood following a nonmyeloablative preparative regimen. *Blood* 2001; 98: 3486–8.

21. Barker JN, Weisdorf DJ, DeFor TE, Blazar BR, Miller JS, Wagner JE. Rapid and complete donor chimerism in adult recipients of unrelated donor umbilical cord blood transplantation after reduced-intensity conditioning. *Blood* 2003; 102: 1915–9.

22. Miyakoshi S, Yui K, Kami M et al. Successful engraftment after reduced-intensity umbilical cord blood transplantation for adult patients with advanced hematological diseases. *Clin Cancer Res* 2004; 10: 3586–92.

23. Nakasone H, Fuji S, Yakushijin K et al. Impact of total body irradiation on successful neutrophil engraftment in unrelated bone marrow or cord blood transplantation. *Am J Hematol* 2017; 92: 171–8.

24. Okada M, Yoshihara S, Taniguchi K et al. Intrabone marrow transplantation of unwashed cord blood using reduced-intensity conditioning treatment: a phase I study. *Biol Blood Marrow Transplant* 2012; 18: 633–9.

25. Terakura S, Wake A, Inamoto Y et al. Exploratory research for optimal GVHD prophylaxis after single unit CBT in adults: short-term methotrexate reduced the incidence of severe GVHD more than mycophenolate mofetil. *Bone Marrow Transplant* 2017; 52: 423–30.

26. Castello S, Podestà M, Menditto VG et al. Intra-bone marrow injection of bone marrow and cord blood cells: an alternative way of transplantation associated with a higher seeding efficiency. *Exp Hematol* 2004; 32: 782–7.

27. Rocha V, Labopin M, Ruggeri A et al. Unrelated cord blood transplantation: outcomes after single-unit intrabone injection compared with double-unit intravenous injection in patients with hematological malignancies. *Transplantation* 2013; 95: 1284–91.

28. Saglio P, Berger M, Vassallo E et al. Intrabone cord blood hematopoietic stem cell transplantation in a subset of very high-risk pediatric patients: a safety and feasibility pilot study. *J Pediatr Hematol Oncol* 2012; 34: 359–63.

29. Frassoni F, Varaldo R, Gualandi F et al. The intra-bone marrow injection of cord blood cells extends the possibility of transplantation to the majority of patients with malignant hematopoietic diseases. *Best Pract Res Clin Haematol* 2010; 23: 237–44.

30. Brunstein CG, Barker JN, Weisdorf DJ et al. Intra-BM injection to enhance engraftment after myeloablative umbilical cord blood transplantation with two partially HLA-matched units. *Bone Marrow Transplant* 2009; 43: 935–40.

31. Kanda J, Morishima Y, Terakura S et al. Impact of graft-versus-host disease on outcomes after unrelated cord blood transplantation. *Leukemia* 2017; 31: 663–8.