Differential roles of Scavenger receptor class B type I: A protective molecule and a facilitator of atherosclerosis (Review)

BAITAO MA¹, JING JIA², XUEBIN WANG¹, RUI ZHANG¹, SHUAI NIU¹, LENG NI¹, XIAO DI¹ and CHANGWEI LIU¹

¹Department of Vascular Surgery, Peking Union Medical College Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100730; ²Department of Obstetrics and Gynaecology, Tongji Hospital of Tongji Medical College, Huazhong University of Science and Technology, Wuhan, Hubei 430030, P.R. China

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Abstract. The scavenger receptor class B type I (SR-BI) is a multi-ligand membrane protein receptor that binds to high-density lipoprotein (HDL) under physiological conditions, promoting the selective uptake of cholesterol esters from HDL into cells. SR-BI also promotes the reverse transport of excess cholesterol from peripheral tissues to the liver, contributing to the synthesis of bile acids for excretion and the removal of excess cholesterol from the body, thereby lowering the cholesterol load and exerting anti-atherosclerotic effects. Studies in mice and humans have demonstrated that a functional defect of SR-BI can cause atherosclerotic lesions and cardiovascular diseases, such as myocardial infarction and stroke. Additionally, SR-BI in vascular endothelial cells promoted the deposition of low-density lipoprotein under the endothelium. Although SR-BI is widely expressed in various tissues and cell types throughout the body, its expression level and function vary accordingly. The present review focuses on the biological functions and mechanisms of SR-BI in regulating atherosclerosis.

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1. Introduction

The gene sequence of scavenger receptor class B type I (SR-BI) was first determined by Calvo and Vega (1) in 1993 and named CLA-1 (now SCARB1). SR-BI shares close sequence identity with cluster determinant 36 and lysosomal integral membrane protein 2 (LIMP-2). Acton et al (2) proposed that SR-BI was a high-affinity high-density lipoprotein (HDL) physiological membrane protein receptor that mediated the selective uptake of cholesterol esters (CEs) in HDL particles via hepatocytes. Subsequent research has focused on the transcriptional regulation, structure and function of SR-BI. SR-BI is highly conserved across different species, and is widely expressed in different tissues, including liver, gonad, adrenal gland, intestinal tract, macrophages, endothelial cells (ECs), platelets, placenta, adipocytes and smooth muscle cells (3). SR-BI is a member of the scavenger receptor superfamily and can recognize a variety of ligands, including HDL, natural and modified low-density lipoprotein (LDL), very-low-density lipoprotein (VLDL), anion-containing phospholipids, lipoprotein a (Lpa), phosphatidylserine (PS) and apoptotic cells (4). SR-BI has also been demonstrated to exhibit a high affinity for native HDL (4,5).

The main physiological function of SR-BI is binding with HDL to mediate the selective uptake and reverse transport of cholesterol (3). When circulating LDL particles bind to the LDL receptor (LDLR), cholesterol is delivered to cells via endocytosis of the holo-particle (3). After endocytosis, CEs carried in LDL are hydrolysed by acid lipase in the lysosome, releasing free cholesterol (FC) that is transported to the endoplasmic reticulum, where it is either re-esterified to CEs and stored in lipid droplets in the cytoplasm, or transported to the
plasma membrane (3). In contrast, the mechanism by which CEAs in HDL particles enter the cell is completely different from internalisation of the CE-rich lipoprotein particles, which is mediated by LDLR. After HDL particles bind to SR-BI on the cell surface, CEAs, FC and triglycerides are selectively delivered to the inside of the cell through hydrophobic channels formed by the extracellular domain of the SR-BI molecule, while the whole lipoprotein particle remains outside the cell (2,6). This process results in the selective uptake of cholesterol. In steroi-dogenic cells, CEAs that enter the cell in this way are hydrolysed by hormone-sensitive lipases before being used (7).

HDL is an anti-atherosclerotic lipoprotein, possibly due to its role in reverse cholesterol transport (RCT) (6). A meta-analysis indicated that a decrease in plasma HDL was associated with increased occurrence of atherosclerosis and cardiovascular diseases (8). Excess cholesterol in macrophages or foam cells in atherosclerotic lesions can be transferred to HDL particles through unidirectional outflow mediated by ATP-binding cassette A1 and ATP-binding cassette G1 (9), or flow to mature HDL particles via passive diffusion through hydrophobic channels formed by the extracellular domain of SR-B1 molecules (9). Both mechanisms decrease the storage of CEAs in cells and reduce the incidence of atherosclerosis and the risk of cardiovascular diseases (9). Mature HDL particles circulate through the blood to the liver, where they interact with SR-BI on the hepatocyte's membrane (10). This reverse transport of cholesterol to the liver results in the synthesis of bile acids for secretion into the lumen of the intestine and the ensuing excretion of excess cholesterol from the body (10). Interactions between HDL and SR-BI on the hepatocyte membrane and the selective uptake of CEAs are crucial RCT steps, which serve a critical role in cardiovascular protection (11).

2. Structure of SR-BI

The human SCARBI gene encoding the SR-BI protein is located at 12q24 on chromosome 12, is ~87 kb in length and comprises 13 exons and 12 introns (3). SR-BI has a predicted molecular weight of 57 kDa and contains multiple glycosylation sites (3). However, due to post-translational modification, it migrates at ~82 kDa when assessed using SDS-PAGE (12).

The human SCARBI mRNA yields five protein variants through alternative splicing (3). Isoform 1 is the main gene product, consisting of 509 amino acid (aa) residues from exons 1-12. This was the first isoform identified and named SR-BI (3). Isoform 2 comprises 499 aa residues and has the same C-terminus as isoform 1 but lacks aa residues 43-142. Isoform 3 is the longest variant with 552 aa residues. Isoform 4 (also called SR-BII) is a polypeptide comprised of 474 aa, with the same C-terminus as isoform 1, but a different N-terminus (aa 1-42). Isoform 5 (also called SR-BIII) comprises 506 aa residues and has a unique C-terminus (aa 468-552).

SR-BI has two short intracellular domains at the N-terminus (aa 1-11) and C-terminus (aa 462-509), two transmembrane domains (aa 12-32 and 441-461) and a large extracellular domain containing multiple N-linked glycosylation sites (3). The C-terminal cytoplasmic domain is essential for SR-BI signal transduction and contains kinase phosphorylation sites and protein binding sites, as well as a serine that can be phosphorylated by the kinase SIK-1, which increases the efficiency of CEAs uptake mediated by SR-BI (3).

A common C-terminal sequence (aa 468-552) in isomers 1, 2 and 4 includes the VQLQAKL motif that is required for binding to the PDZ domain-containing protein PDZK1, which is essential for maintaining the stability and function of SR-BI (13). The other C-terminal sequence (aa 487-494), IQAIESL, is a motif required to bind the guanine nucleotide exchange factor inhibitor of cytokinesis 4 (DOCK4), which is essential for the uptake and transendoctysis of LDL in ECs (14). Determination of the crystal structure of LIMP-2, a homologue of SR-BI, provided a basic framework for the structure of this protein family (15), which revealed that the main extracellular domain of SR-BI contained an antiparallel β-barrel core and multiple short α-helical structures that facilitated the formation of a hydrophobic channel and promoted cholesterol transport.

3. SR-BI in hepatocytes

SR-BI on the hepatocyte membrane mediates the selective uptake of CEAs in plasma HDL particles and the RCT process. SR-BI is also known to regulate lipoprotein homeostasis, inhibit the development of atherosclerosis and reduce the incidence of cardiovascular diseases (16,17).

Van Eck et al (18) established SR-BI knockout (SR-BI-/-) mice and fed them a western-type diet for 20 weeks. Compared with wild-type mice, cholesterol levels in the liver and the expression of key regulatory factors related to cholesterol homeostasis [3-hydroxy-3-methyl-glutaryl-coenzyme A (HMG-CoA) reductase, LDLR and cholesterol 7α-hydroxylase] were not affected, but a large amount of cholesterol-rich HDL accumulated in the circulatory system along with abnormally large HDL particles, which indicated impaired cholesterol delivery to the liver. SR-BI-deficient mice displayed an ~40% reduction in biliary cholesterol content, lipid deposition in the aortic wall and the increased occurrence of atherosclerosis, which indicated that the RCT process was damaged and the cholesterol clearance capacity was impaired (18). Apolipoprotein E (ApoE) single-knockout mice did not usually suffer early atherosclerosis, severe cardiac dysfunction, cardiac infarction or premature death when fed a standard diet. However, SR-BI and ApoE double gene knockout mice experienced severe hypercholesterolemia after ingesting a standard diet, and atherosclerotic lesions appeared in the early stages (4-5 weeks), resulting in coronary artery occlusion, myocardial infarction and cardiac dysfunction, with most mice dying at 5-8 weeks of age (19,20). Moreover, after a period of high-fat and high-cholesterol consumption, SR-BI and LDLR double gene knockout mice exhibited higher cholesterol expression levels, more severe atherosclerotic lesions and a higher incidence of myocardial infarction and mortality, compared with LDLR knockout (LDLR-/-) mice in the control group (21).

These studies indicated that SR-BI serves an important role in the prevention of atherosclerosis and the reduction of cardiovascular diseases, partly due to selective cholesterol uptake and RCT mediated by SR-BI on the hepatocyte membrane.

SR-BI on hepatocytes is beneficial to cardiovascular fitness through the regulation of HDL cholesterol metabolism (22). Liver-specific SR-BI deficiency induced atherosclerotic lesions, while overexpression of SR-BI in hepatocytes decreased atherosclerosis (22). After transient overexpression of SR-BI in hepatocytes, LDLR-/- mice fed with a high-fat and high-cholesterol...
diet displayed a marked reduction in early and advanced atherosclerotic lesions, and also experienced significantly decreased HDL cholesterol but unchanged or only modestly reduced non-HDL cholesterol levels (23). Huby et al (24), produced a hypomorphic allele (hypomSR-BI), an SR-BI conditional knockout mouse model, by targeting the SR-BI gene through loxp site insertion, resulting in a decrease in SR-BI expression. After consuming an atherogenic diet for 11 weeks, plasma total cholesterol (TC) levels were increased 2-fold compared with the control group, and hypomSR-BI-KO (liver) and SR-BI mice developed severe hypercholesterolemia due to the accumulation of VLDL-sized particles in the circulation (24). The degree of atherosclerosis was more severe in hypomSR-BI mice, hypomSR-BI-KO (liver) mice and SR-BI mice compared with the control mice, by 2.5-fold, 32-fold and 48-fold, respectively. There was no difference in plasma lipid expression levels between hypomSR-BI-KO (liver) mice and SR-BI mice. However, compared with SR-BI mice, hypomSR-BI-KO (liver) mice had fewer atherosclerotic lesions (24). The aforementioned studies indicated that SR-BI expressed in hepatocytes serves a critical role in anti-atherosclerosis, and SR-BI expressed in other tissues or cells also perform anti-atherosclerotic functions to a limited extent.

In addition to the selective uptake of CEs from HDL, SR-BI expressed in the liver can also clear VLDL and Lpa. Van Eck et al (25) demonstrated that at 20 mins after the intravenous injection of (125)I-β-VLDL, the liver uptake ability of SR-BI-/- mice was decreased by 44% compared with that of wild-type mice, while VLDL in plasma was significantly increased (45.9±8.8 vs. 6.1±2.3 μg/ml). A study by Yang et al (26) demonstrated that SR-BI acted as a receptor for atherosclerotic particles containing Lpa and mediated the selective uptake of lipids in Lpa. SR-BI mice presented with the significantly reduced clearance rate of Lpa in the plasma, while SR-BI overexpression in the liver could significantly increase the clearance rate. Humans carry a number of different SCARBI gene mutations that reduce the ability of the liver to uptake CEs in HDL and Lpa, resulting in significantly elevated plasma HDL and Lpa expression levels (27), although this is accompanied by higher HDL levels and the risk of cardiovascular disease is significantly increased. The aforementioned studies in mice and humans indicated that in addition to mediating the reverse transport of cholesterol in surrounding tissues to liver cells via HDL particles, generating and excreting bile, liver SR-BI also had an anti-atherogenic role via regulating the homeostasis of lipoproteins in the blood.

4. SR-BI in endothelial cells

SR-BI in ECs not only binds with HDL to activate downstream signalling pathways (28) or transport HDL to the subintimal region via transcytosis (29) to exert an anti-atherosclerotic effect, but also mediates the delivery of LDL into the artery wall via transcytosis (14,30), thereby promoting atherosclerosis (Fig. 1). SR-BI regulates the function of ECs under the stimulation of HDL via activating Src kinases, phosophatidylinositol 3-kinase (PI3K), protein kinase B/mitogen-activated protein kinases and the Rac GTP enzyme, resulting in changes in the actin cytoskeleton, promotion of EC migration (Fig. 1), enhancement of EC repair and regeneration and improvement of endothelial monolayer integrity in vivo (31,32). The HDL/SR-BI interaction induced the phosphorylation of endothelial nitric oxide synthase and the synthesis of nitric oxide via phosphorylation of Src kinases, protected ECs from apoptosis and promoted the growth and migration of ECs (32). Moreover, phosphorylation of Src kinases induced by HDL/SR-BI required the assistance of the adaptor protein PDZK1 (3,33). In the aforementioned study, compared with the control group, EC-specific SR-BI-overexpressing C57BL/6N mice and ApoE-/- mice fed a high-fat and high-cholesterol diet displayed significantly decreased plasma levels of TC and HDL, while LDL was remarkably increased, and the severity of atherosclerosis was decreased. However, for C57BL/6N mice and ApoE-/- mice fed a normal chow diet, EC-specific SR-BI overexpression had no significant effect on plasma lipids and lipoproteins (34), suggesting that SR-BI expressed on ECs may serve an important role in the regulation of lipid and lipoprotein homeostasis and preventing atherosclerosis.

SR-BI on ECs also serve a role in transcytosis, transporting plasma lipoproteins to the endomembranous (29,30). Rohrer et al (29) indicated that SR-BI could mediate the transport of HDL in the circulation to the intima of vessels, receive cholesterol in foam cells or macrophages and reduce cholesterol deposition beneath the arterial endothelium, thereby exerting an anti-atherosclerotic effect. The diameter of translocated HDL particles was decreased, but protein moieties remained intact (29). A study by Fung et al (35) demonstrated that SR-BI on the brain microvascular endothelium could mediate HDL transcytosis, and this process did not depend on the adaptor protein PDZK1. Armstrong et al (30) revealed that SR-BI mediated the transcytosis of LDL in ECs. Overexpression of SR-BI enhanced the endocytosis of LDL in ECs, but the effect was significantly reduced when an excess of HDL (a high-affinity ligand for SR-BI) was administered or the expression of SR-BI was depleted by small interfering RNA-mediated silencing (30).

A recent study by Huang et al (14) demonstrated that SR-BI mediated LDL transcytosis associated with ECs via DOCK4 to promote the development of atherosclerosis (Fig. 1). Transcytosis of LDL particles through ECs required direct binding to SR-BI expressed on the apical membranes of ECs, and an 8 aa (IQYAYSSEl) cytoplasmic motif in C-terminus of SR-BI that recruited DOCK4 (36), which promoted the internalisation of SR-BI and transcytosis of LDL across EC monolayers by coupling LDL and SR-BI via Rho GTPases Ras-associated C3 botulinum toxin substrate 1 (Rac1) activation (14). Selective deletion of SR-BI in ECs in male and female wild-type, ApoE-/- and LDLR-/- mice had the protective effect against atherosclerosis but it did not alter circulating levels of TC, HDL or the lipoprotein profile (14). The aforementioned results indicated that the role of SR-BI expressed on ECs in atherosclerosis is complex, and diverse results from different studies might be related to the metabolic state of the body and the cholesterol load, which suggested that accurate individualised treatment in different populations could be achieved when formulating anti-atherosclerosis strategies targeting SR-BI.

5. SR-BI in macrophages

The SR-BI protein was also expressed on macrophages, such as Kupffer cells in the liver (37) and foam cells in atherosclerotic plaques (38). SR-BI expressed on macrophages has an important role in the regulation of lipid and lipoprotein homeostasis and preventing atherosclerosis.
role in selective cholesterol uptake (39), cholesterol efflux (40,41), HDL-induced anti-inflammatory responses (41,42) and the inhibition of atherosclerotic progression (43).

Tao et al (43) demonstrated that macrophage Sr-Bi bound to PS and induced Src phosphorylation, which led to the activation of downstream molecules PI3K and Rac1, phagocytosis and elimination of apoptotic cells in atherosclerotic lesions, thereby reducing necrosis, inhibiting inflammatory responses and exerting an anti-atherosclerotic effect. Moreover, Sr-Bi also mediated HDL-induced anti-inflammatory effects by inhibiting the activation of NF-κB and the production of tumor necrosis factor α (42).

Although Sr-Bi was expressed in many bone marrow-derived cells (BMDCs), including B and T lymphocytes, lymphocyte Sr-Bi did not affect the development of atherosclerosis (44). In order to investigate the role of macrophage Sr-Bi in atherosclerosis, Van Eck et al (45) used bone marrow transplantation technology to specifically regulate the expression of Sr-Bi in leukocytes of LDLR−/− mice, and indicated that macrophage Sr-Bi had a unique dual role in the process of atherosclerosis. After feeding LDLR−/− mice with a western-type diet for 9 or 12 weeks, the presence of Sr-Bi in BMDCs reduced the development of atherosclerosis. However, after only 4 weeks in LDLR−/− mice fed a western diet and wild-type mice fed a high-cholesterol and high-cholate diet, expression of Sr-Bi in BMDCs promoted the development of small fat streak lesions (45). In addition, Sr-Bi-null and apoE-hypomorphic (Sr-Bi−/−/ApoE−/−) mice transplanted with bone marrow derived from Sr-Bi−/−/ApoE−/− mice and fed a high-fat high-cholesterol and cholate-containing diet for 4 weeks revealed a significant reduction in the degree of coronary atherosclerosis and the incidence of myocardial infarction compared with mice transplanted with bone marrow derived from Sr-Bi−/− mice (46).

Sr-Bi expressed by macrophages has also been revealed to mediate cholesterol efflux from macrophages into mature HDL particles (40), which is an important stage in the process of anti-atherosclerosis. Compared with the wild-type group, the rate of cholesterol efflux from macrophages isolated from Sr-Bi−/− mice to mature HDL particles was reduced by 20%. In addition, Badeau et al (47) indicated that Sr-Bi inhibitors significantly reduced cholesterol efflux from THP-1 macrophages. A previous clinical study has demonstrated that cholesterol outflow to HDL in macrophages from human carriers of the SCARB1 gene P297S mutation was significantly reduced compared with control subjects (48). In conclusion, macrophage Sr-Bi exerted an anti-atherosclerotic effect by eliminating apoptotic cells, inducing anti-inflammatory responses and promoting cholesterol efflux.

6. SR-BI in platelets

Platelets are closely associated with the development of atherosclerosis (49), and their activation and aggregation can generate thrombus, accelerate the progression of atherosclerotic lesions and subsequently occlude arteries, which results in reduced blood/oxygen supply to organs or tissues, ultimately leading to the occurrence of cardiovascular events, such as myocardial infarction and stroke (49,50). Therefore, inhibiting the abnormal aggregation and activation of platelets could reduce the occurrence of cardiovascular events (50). Valiyaveetil et al (51) found that the Sr-Bi protein was present in both resting and activated platelets, and that oxidised HDL particles could bind to and interact with Sr-Bi to inhibit platelet activation and aggregation, thereby reducing the risk of thrombosis. In addition, Brodte et al (52) demonstrated that natural HDL, the main component of HDL, bound to
SR-BI on platelets and activated the signal pathways, which inhibited platelet activation. Furthermore, SR-BI<sup>-/-</sup> mice were more likely to suffer deep vein thrombosis (53), and the time required for arterial occlusion was shorter in a model of FeCl<sub>3</sub>-induced arterial thrombosis (54). The increase of circulating platelet and bone marrow megakaryocyte numbers was impaired in SR-BI<sup>-/-</sup> mice when challenged with thrombopoietin. However, proplatelets were normally produced by megakaryocytes from normolipidemic bone marrow-specific SR-BI<sup>-/-</sup> mice (55). Furthermore, studies in humans indicated that carriers of the P297S mutation of the SCARB1 gene had significantly increased FC levels in platelets and impaired platelet function (48).

7. Conclusion

In conclusion, SR-BI is a multi-ligand membrane receptor that has a high affinity for HDL and is believed to be the physiological receptor for HDL. Expression of SR-BI in different tissues and cell types exhibit different roles in the process of atherosclerosis. SR-BI in hepatocytes and platelets have been demonstrated to inhibit the development of atherosclerotic lesions and thrombosis, and act as a protective factor for cardiovascular diseases (16,17). However, SR-BI expressed in vascular ECs has been indicated to promote the deposition of LDL under the endothelium, aggravating atherosclerosis and representing a risk factor for cardiovascular diseases. The effects of macrophage SR-BI on the pathogenesis of atherosclerosis and cardiovascular diseases are complex and depend on the metabolic and inflammatory status at different stages of atherosclerosis. Therefore, the role of SR-BI in the cardiovascular system remains ambiguous, and all the previously reported cardioprotective and atherosclerotic effects of SR-BI must be considered when designing therapeutic agents to treat atherosclerosis and cardiovascular diseases.

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Authors' contributions

BTM and JJ wrote the manuscript. XBW, RZ, SN, LN, XD and CWL reviewed and edited the manuscript. All authors read and approved the final manuscript.

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Not applicable.

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Competing interests

The authors declare that they have no competing interests.

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