RESEARCH ARTICLE

Cerebral Hemodynamic Changes of Mild Traumatic Brain Injury at the Acute Stage

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Abstract

Mild traumatic brain injury (mTBI) is a significant public health care burden in the United States. However, we lack a detailed understanding of the pathophysiology following mTBI and its relation to symptoms and recovery. With advanced magnetic resonance imaging (MRI), we can investigate brain perfusion and oxygenation in regions known to be implicated in symptoms, including cortical gray matter and subcortical structures. In this study, we assessed 14 mTBI patients and 18 controls with susceptibility weighted imaging and mapping (SWIM) for blood oxygenation quantification. In addition to SWIM, 7 patients and 12 controls had cerebral perfusion measured with arterial spin labeling (ASL). We found increases in regional cerebral blood flow (CBF) in the left striatum, and in frontal and occipital lobes in patients as compared to controls (p = 0.01, 0.03, 0.03 respectively). We also found decreases in venous susceptibility, indicating increases in venous oxygenation, in the left thalamostriate vein and right basal vein of Rosenthal (p = 0.04 in both). mTBI patients had significantly lower delayed recall scores on the standardized assessment of concussion, but neither susceptibility nor CBF measures were found to correlate with symptoms as assessed by neuropsychological testing. The increased CBF combined with increased venous oxygenation suggests an increase in cerebral blood flow that exceeds the oxygen demand of the tissue, in contrast to the regional hypoxia seen in more severe TBI. This may represent a neuroprotective response following mTBI, which warrants further investigation.
Introduction

Over 1.7 million Americans suffer a traumatic brain injury (TBI) each year, most of which are classified as mild TBI (mTBI), also called concussion [1]. Despite its name, mTBI is not inconsequential [2–5]. The prolonged post-concussive symptoms significantly affect the quality of life of both patients and their families. However, the majority of patients with mTBI lack structural imaging correlates [6] thus making the clinical detection and outcome prediction a challenge. Following the initial traumatic insult, a series of vascular responses and perfusion changes is set in motion. Insults to the cerebral vasculature, such as microbleeds, local perfusion reduction, or oxygen metabolism changes are known to cause devastating secondary complications after moderate to severe TBI [7,8]. Further, perfusion studies using either arterial spin labeling (ASL) [9] or contrast-enhanced perfusion weighted imaging (PWI) [10] have demonstrated reduced cerebral blood flow in the chronic stages of mTBI.

Cerebral perfusion and oxygenation are of particular interest as markers of brain function, as neurons rely nearly completely on aerobic metabolism for their energy production. However, decreases in perfusion or venous oxygenation in the brain could indicate either a potentially at-risk area with unmet energy demands or a change in energy demands (a result of a change in function), which represent two entirely different injury responses with different implications for outcome. These hypoperfused areas can be identified by decreased CBF and oxygenation [11]; in patients with moderate to severe TBI ischemic volume has been shown to correlate with negative outcomes [12]. Additionally, early impairment of CBF in patients with severe injury correlates with poor brain tissue oxygenation [13]. Because of this, Zwienenberg and colleagues [14] have suggested optimizing cerebral perfusion and blood flow in the treatment of head injury patients. In mTBI, however, there is a lack of published data on the relationship between CBF and prognosis, as well as a lack of methods to distinguish primary alterations in CBF from primary alterations in brain metabolic demand. In particular, there is a lack of investigation of CBF changes in patients with mTBI in the acute stage (within 48 hours) of injury.

Further, there is paucity of data on blood oxygenation investigation in TBI patients and the extent of neurovascular compromise in mTBI, particularly of the early hemodynamic response in the acute stage, is still largely unknown. Due to the decoupling of cerebral blood supply and metabolic demand, measurement of CBF alone may not reflect the metabolic status of brain tissue. Ischemic/hypoxic brain tissue will have increased oxygen extraction, which results in decreased blood oxygenation in draining veins. Together with CBF, a measurement of venous blood oxygenation may help to paint a comprehensive picture of the hemodynamics of the brain tissue.

There have been attempts to extract oxygen saturation using T2, T2+ or T2’ approaches [15–31]. MR phase-based methods [32–37] have also been used on individual veins in an attempt to extract venous susceptibility and oxygen saturation. Another method is quantitative susceptibility mapping (QSM) which provides measures of oxygen extraction fraction [38,39]. Recently, Haacke et al have developed a method called susceptibility weighted imaging and mapping (SWIM), which is a form of QSM, for quantification of venous blood oxygenation [40]. Unlike other phase-based methods, the application of a regularized inverse filter to the frequency domain of the phase images results in susceptibility maps that are unaffected by the orientation of the blood vessels [40], providing a strong advantage over other phase-based measurements of susceptibility.

By pairing ASL with SWIM, it is possible to assess both perfusion and oxygenation of the injured brain. Thus, in the present study, we have assessed both CBF and venous oxygenation of several brain regions in which abnormalities are known to correlate with symptoms, including frontal, parietal, and temporal gray matter [41], cingulate cortex [42], and the basal ganglia.
as well as occipital gray matter, thalamus, and striatum. We sampled the susceptibility of major veins that drain these and other areas including septal, thalamostriate, internal cerebral, and the basal veins of Rosenthal. We hypothesized that, after concussion, patients with mTBI may demonstrate reduced cerebral blood flow and decreased venous blood oxygenation in the acute stage. Additionally, we administered a short neurocognitive assessment, the standardized assessment of concussion (SAC) [43], to examine the correlation between the cognitive deficits and the imaging abnormalities.

Methods

Subject selection

This study was approved by both the Human Investigation Committee of Wayne State University and the Institutional Review Board of Detroit Medical Center. Written informed consent was obtained from each subject before enrollment.

A cohort of patients was prospectively recruited from the Detroit Receiving Hospital Emergency Department (ED), which is part of the Level-1 trauma center in the Detroit Medical Center. Patient eligibility was based on the mTBI definition by the American Congress of Rehabilitation Medicine [44] with the following inclusion criteria: Patients aged 18 or older with the initial Glasgow Coma Scale (GCS) score of 13 – 15 in ED with any period of loss of consciousness less than 30 minutes or any post traumatic amnesia less than 24 hours, or recorded change of mental status (confused, disoriented or dazed). All patients required a CT scan as part of their clinical evaluation. All patients were be able to communicate in English. The exclusion criteria included patients under the age of 18 years, pregnancy, medically documented history of previous brain injury, neurological disorders or psychoactive medications, history of substance abuse, CT indication of any metal in the brain and body, known contraindication to MRI (such as a pacemaker or other non-MR compatible implanted device) as identified by safety screening, or presentation without a clear history of trauma as their primary event (e.g., seizure, epilepsy, etc.). The patients’ records were retrospectively screened as well to exclude any patient who does not fit our inclusion criteria.

A total of 14 patients with mTBI were recruited from the ED and 18 healthy controls were recruited via local advertising and from patients’ friends and relatives, as part of a study sponsored by the Department of Defense. Among them, SWI and structural images were acquired for all subjects and ASL data were obtained for 7 patients and 12 controls (Table 1), as the scanning protocol was amended to include the ASL sequence shortly after the initial study began. ASL data from two patients were excluded from the final analysis due to excessive motion during the scan. For mTBI patients, neurocognitive status was measured using the SAC questionnaire, which includes measures of attention, orientation and memory status.

Data on patients’ blood alcohol levels, drug use, ED-administered medications, and blood pressure (BP) were collected from clinical records for inclusion in the final analyses as possible confounders.

Imaging

All imaging was performed on a Siemens 3T VERIO magnet with a 32 channel head coil. The imaging protocol included a standard 3D T1 MPRAGE, T2 Fluid-attenuated inversion recovery (FLAIR), diffusion tensor imaging (DTI), SWI, ASL, and resting state functional MRI. This paper focuses on the hemodynamic analysis of brain injury using SWI and ASL data. All structural MRI images, including SWI images, were reviewed by two board-certified neuroradiologists to identify other conditions that could confound the results of the study. The
Table 1. ASL-assessed patients’ and controls’ demographic data and cause of injury.

| Case ID | Age (Years) | Sex | Race       | Delay to Scan | ER GCS | Injury Mech. | ASL | SWI | Structural findings                      |
|---------|-------------|-----|------------|---------------|--------|--------------|-----|-----|------------------------------------------|
| **Patients** |          |     |            |               |        |              |     |     |                                           |
| 001     | 27         | F   | Caucasian  | 41 hr         | MVA    | X            |     |     | X                                        |
| 2       | 29         | M   | Caucasian  | 10 d          | 15     | MVA          | X   |     |                                           |
| 002     | 24         | M   | Caucasian  | 46 hr         | Fall   | X            |     |     | X                                        |
| 4       | 20         | M   | Caucasian  | 27 hr         | 15     | Fall         | X   |     | Moderate bleed, scalp edema              |
| 6       | 21         | M   | Black      | 4 d           | MVA    | X            |     |     | X                                        |
| 7       | 25         | M   | Black      | 17 hr         | 15     | Assault      | X   | X   | X                                        |
| 9       | 56         | M   | Indian     | 26 hr         | 15     | MVA          | X   |     | X                                        |
| 11      | 35         | M   | Black      | 36 hr         | Fall   | X            |     |     | X                                        |
| 14      | 30         | M   | Caucasian  | 7 d           | 15     | Fall         | X   | X   | X                                        |
| 15      | 36         | F   | Black      | 9 hr          | 15     | MVA          | X   |     | X                                        |
| 16      | 19         | M   | Black      | 3 hr          | 15     | MVA          | X   |     | X                                        |
| 17      | 23         | M   | Black      | 9 hr          | 15     | MVA          | X   |     | X                                        |
| 18      | 21         | F   | Caucasian  | 2 d           | 15     | MVA          |     |     | X                                        |
| 19      | 30         | F   | Asian      | 8 hr          | 15     | SBV          | X   |     | X                                        |
| **Mean** | 27.14      |     |            |               | 55.29  |              |     |     |                                          |
| **SD**  | 5.52       |     |            |               | 68.82  |              |     |     |                                          |
| **Median** | 26       |     |            |               | 31.5 hr|              |     |     |                                          |
| **Range** | 19–56     |     |            |               | 3 hr—10 d|              |     |     |                                          |
| **Controls** |          |     |            |               |        |              |     |     |                                           |
| 1       | 24         | F   | Asian      |               | X      | X            |     |     | X                                        |
| 2       | 23         | M   | Indian     |               | X      |              |     |     | X                                        |
| 3       | 45         | M   | Caucasian  |               | X      | Nonsp hyperint|     |     |                                           |
| 5       | 22         | M   | Caucasian  |               | X      | Capillary telangiectasia | |     |                                           |
| 6       | 27         | M   | Asian      |               | X      |              |     | X   | X                                        |
| 7       | 23         | F   | Asian      |               | X      | X            |     |     | X                                        |
| 8       | 22         | F   | Asian      |               | X      | X            |     |     | X                                        |
| 9       | 65         | F   | Asian      |               | X      | Pineal gland cyst, Nonsp hyperint | |     |                                           |
| 36      | 52         | F   | Caucasian  |               | X      | X            |     |     | X                                        |
| 37      | 44         | M   | Caucasian  |               | X      | X            |     |     | X                                        |
| 38      | 41         | M   | Caucasian  |               | X      | X            |     |     | X                                        |
| 39      | 28         | F   | Caucasian  |               | X      |              |     |     | X                                        |
| 40      | 27         | F   | Arabic     |               | X      | X            |     |     | X                                        |
| 41      | 29         | M   | Caucasian  |               | X      | X            |     |     | X                                        |
| 42      | 33         | M   | Caucasian  |               | X      | X            |     |     | X                                        |
| 43      | 66         | F   | Caucasian  |               | X      | Nonsp hyperint|     |     | X                                        |
| 44      | 38         | M   | Hispanic   |               | X      | Nonsp hyperint|     |     | X                                        |
| 45      | 28         | M   | Caucasian  |               | X      |              |     |     | X                                        |
| 46      | 21         | M   | Black      |               | X      |              |     |     | X                                        |
| **Mean** | 30.08      |     |            |               |        |              |     |     |                                          |
| **SD**  | 10.24      |     |            |               |        |              |     |     |                                          |
| **Median** | 28       |     |            |               |        |              |     |     |                                          |
| **Range** | 21–66     |     |            |               |        |              |     |     |                                          |

hr = hours, d = days, nonsp hyperint = nonspecific hyperintensity, SBV = pedestrian struck by vehicle.

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neuroradiologists were blinded to the medical history and clinical condition of the subjects to avoid bias.

Arterial spin labeling. A pulsed ASL sequence was used, with a repetition time (TR) of 2830 milliseconds and echo time (TE) of 11 milliseconds, flip angle of 90 degrees, field of view of 384x384, and in-plane resolution of 4x4x4. After acquisition, the perfusion data were processed automatically by the Siemens online software to produce motion corrected relative CBF (rCBF) images for off-line post-processing.

SWI. The SWI sequence was a T2$^*$ Gradient Echo (GRE) based sequence with long TE and 3D flow compensation. The SWI data were collected with a TR/TE of 39/20 milliseconds, flip angle of 20 degrees, field of view of 256x256, and in-plane resolution of 0.5x0.5x2. It was later interpolated to 0.5x0.5x2 in-plane resolution.

Image processing

ASL processing and analysis. A semi-automated process was used to select the regions of interest (ROIs) to avoid errors in manual selection of the ROIs (Fig. 1). Untagged T2-weighted images in ASL dataset and rCBF images were skull-stripped. T2 images were normalized to the T1-weighted International Consortium for Brain Mapping (ICBM) template, and a transformation matrix was applied to the relative rCBF images to bring them into the same standard space. A MATLAB-based Statistical Parametric Mapping 8 (SPM8) package was used for normalization [45–47].

After bringing all the images into the ICBM standard space, ROIs were automatically selected using the Wake Forest University PickAtlas (WFU pickatlas) [48]. Using these predefined ROIs in the ICBM space, regional rCBF values from the striatum, caudate nucleus, thalamus, globus pallidus, putamen, and frontal, occipital, parietal and temporal lobes of each subject were recorded.

SWIM processing and analysis

Based on the previously described procedure [40,49], the following steps were taken for SWIM processing: a) skull stripping to remove the artifacts caused by skull and brain tissue interface by using the software package MRICro (MRICro, Version 1.40) with the Brain Extraction Tool (BET) [50]; b) phase unwrapping to extract SWI phase signal; c) background field removal to correct the background field inhomogeneity according to the method by Pandian et al. in 2008 [51]; d) inverse filtering to extract susceptibility signal; and e) iterative reconstruction to remove remnant ringing of potential microhemorrhages. This resulted in a high resolution susceptibility map of the venous structures of the brain.

The SWIM data provided a means by which oxygen saturation ($Y$) in the veins can be measured from the bulk susceptibility difference ($\Delta X$) between tissues [33] ($\Delta X$ is expressed in parts per billion; ppb). The susceptibility difference $\Delta X$ of major veins is directly related to the venous blood oxygen saturation ($Y_v$) through the following relationship:

$$\Delta X = \Delta X_{do} \times Hct \times (1 - Y_v)$$

where $\Delta X$ is the susceptibility measurement of major veins, $Y_v$ is venous oxygenation, $\Delta X_{do}$ is the susceptibility difference per unit hematocrit between fully deoxygenated blood and fully oxygenated blood ($4\pi \times 0.27$ ppm [52]), and Hct is the fractional hematocrit in the vein, which is approximately 40% [33].
Fig 1. Diagram of the ASL analysis workflow. T2 images were skull-stripped and normalized to the ICBM T1-weighted template. A transformation matrix was applied to skull-stripped rCBF images with SPM8, to bring them into the same template. ROIs were selected automatically in the lobes and in deep brain structures using the Wake Forest University (WFU) PickAtlas.

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The relative changes of oxygen saturation in mTBI patients can be calculated using the following simplified equation where all constants have cancelled out:

\[
\frac{\Delta Y}{1 - Y_n} = \frac{\Delta \chi_p}{\chi_n}
\] (2)

where \(\Delta Y\) is the oxygenation difference between mTBI patients and normal controls, \(\Delta \chi_p\) is the susceptibility difference between mTBI patients and normal controls, and \(\chi_n\) is the susceptibility value of normal controls. If the oxygen saturation of veins in the normal healthy controls \((Y_n)\) is assumed to be 0.7 [53,54], then relative oxygenation change \(\Delta Y\) could be found from this relationship.

Major veins were selected for analysis (Fig. 2). These included the left, right and central septal veins, the left and right thalamostriate veins, the internal cerebral vein, and the left and right basal veins of Rosenthal. A semi-automated approach was adapted to obtain the relative susceptibility values. Baseline values were selected for each subject by taking the average of 3 measures of means of background brain tissue intensities. ROIs were manually selected for each vessel with a low pass filter cut off of 25% of the baseline value. For each ROI, minimum, maximum, mean, standard deviation and total number of pixels were recorded.

Neurocognitive testing

At the acute setting, once a patient was conscious and deemed to be clinically capable of participating in the study, neurocognitive testing and a survey with a focus on post-concussion symptoms (PCS) were administered. Given the emergency care setting, it was not feasible to perform a full battery of neuropsychological assessment. Instead, a short instrument called Standardized Assessment of Concussion (SAC) [43] was used to assess the patients’ neurocognitive status. The SAC instrument was originally developed for onsite testing of subject’s neurocognitive performance after sports concussion [55]. SAC is sensitive to the acute changes following concussion and it only requires limited training to administer [56]. The SAC assesses 4 cognitive domains including orientation, attention, immediate memory and delayed recall, and the resulting scores are added to give a patient score between 0 and 30, with a lower score indicating greater cognitive deficit. Previous studies reported SAC to be sensitive to brain injury in the emergency care setting, particularly the delayed recall [57].

![Fig 2. Major veins selected for susceptibility analysis.](https://example.com/fig2)

Fig 2. Major veins selected for susceptibility analysis. 1) Left septal vein; 2) right septal vein; 3) central septal veins; 4) left thalamostriate vein; 5) right thalamostriate vein; 6) internal cerebral vein; 7) left basal vein of Rosenthal; and 8) right basal vein of Rosenthal.

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Edition of the SAC instrument also has a graded symptom checklist with all PCS symptoms listed. Patients were asked to grade each symptom from none, mild, moderate, to severe, (rated from 0 to 3, respectively). The points were added to give the overall PCS score.

**Results**

Structural MRI findings

Most patients and controls had no findings on T1 and T2 FLAIR imaging. Two patients and three controls had non-trauma-related findings: arachnoid cyst, pericoallosal lipoma, pineal gland cyst, capillary telangiectasia, and calcification of the falx, respectively. A moderate sized bleed was identified in the gray matter of the right parietal lobe in patient number 4, opposite of scalp edema, and patient 9 showed two small bleeds in the ventrical and cortical parenchyma. Nonspecific hyperintensities were found in 2 patients and 5 controls. These results are summarized in Table 1.

Susceptibility and blood oxygenation differences between patients and controls

A group comparison between the patients with mTBI and the controls was conducted. A two-tailed student’s t-test was performed. The left thalamostriate vein (p = 0.03) and right basal vein of Rosenthal (p = 0.05) showed significantly decreased susceptibility values in patients with mTBI when compared to the control group. Fig. 3 shows the comparison of mean values with standard error bars for mTBI patients and controls. Table 2 shows the group mean values for each vein and student’s t-test comparison between control and patient groups.

![Fig 3. Mean susceptibility values and standard error in major veins. * indicates statistically significant difference between controls and acute visit; † indicates statistically significant difference between acute visit and one-month follow up. R: right, L: left, Int: internal.](https://example.com/fig3.png)

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Inter-rater reliability

Intra-rater and inter-rater correlations were performed for SWIM analysis in 8 randomly chosen patients, to assess the reliability and reproducibility of the method. For inter-rater reliability, a fresh rater was trained on the same process and unaware of the previous results. The correlation ($R^2$ value) was 0.98, demonstrating that this study can be repeated with consistency across users.

The intra-reader consistency was checked by the same user one week after the first analysis. The intra-reader tests showed correlation of 0.99 indicating excellent reliability of the analysis method.

ASL differences between patients and controls

We observed significantly higher rCBF values in left striatum of mTBI patients as compared to controls ($p = 0.01$, student t-test). In particular, the caudate, putamen, and pallidum rCBF were significantly increased. We also observed elevated rCBF values in left frontal and occipital lobes ($p = 0.03$ for both, student t-test; Table 3, Fig. 4). No significant changes in the thalamus, globus pallidus, or temporal or parietal lobes were found (all $p > 0.05$ for student t-tests).

Table 2. Group mean values for each vein and student’s T-test comparison between control & patient groups and between control and follow-up groups.

| Region          | Controls mean (SD) | Patients mean (SD) | T-Test patient vs. control | Patient $Y_p$ (%) |
|-----------------|--------------------|--------------------|----------------------------|-------------------|
| R Septal        | 86.58 (23.87)      | 77.36 (22.31)      | 0.272                      | 3.19              |
| L Septal        | 78.33 (25.86)      | 68.77 (22.71)      | 0.294                      | 3.66              |
| C Septal        | 68.85 (17.34)      | 58.91 (13.26)      | 0.098                      | 4.33              |
| R Thalamostriate| 144.79 (33.10)     | 135.65 (28.80)     | 0.423                      | 1.89              |
| L Thalamostriate| 136.70 (30.58)     | 115.06 (22.83)     | 0.037*                     | 4.75              |
| Int Cerebral    | 123.58 (18.68)     | 122.06 (27.77)     | 0.874                      | 0.37              |
| R Basal         | 137.78 (34.46)     | 114.46 (20.58)     | 0.039*                     | 5.08              |
| L Basal         | 140.38 (38.50)     | 125.80 (35.85)     | 0.279                      | 3.12              |

* and bold font indicates significant difference. L = left, R = right.

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Table 3. Comparison of mean CBF values (in mL/100g/min) between control and patient groups.

| Region       | Controls mean (SD) | Patients mean (SD) | $p$ value |
|--------------|--------------------|--------------------|----------|
| Left thalamus| 30.03 (6.25)       | 41.61 (6.71)       | 0.22     |
| Right thalamus| 37.33 (6.17)     | 49.44 (8.77)       | 0.28     |
| Left striatum| 17.49 (4.01)       | 32.27 (3.72)       | 0.01*    |
| Right striatum| 21.4 (4.31)       | 33.69 (6.38)       | 0.14     |
| Frontal lobe | 24.38 (6.83)       | 37.54 (13.09)      | 0.03*    |
| Temporal lobe| 33.88 (10.81)      | 47.31 (17.97)      | 0.11     |
| Occipital lobe| 31.27 (5.02)      | 52.42 (19.63)      | 0.03*    |
| Parietal lobe| 42.14 (9.8)        | 53.66 (15.33)      | 0.10     |

$p$ value shows level of significance for student’s t-test.

* and bold font indicates significant difference.

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Neurocognitive testing

Given the small sample size, mTBI patients’ SAC scores were compared with a normalized dataset reported by McCrea et al., of 568 healthy controls [43]. mTBI patients’ delayed recall scores were significantly lower than the normalized dataset on the delayed recall score (p = 0.04; Table 4, Fig. 5). Other measures were not significantly different from controls.

No significant correlations were found between SAC/Post Concussion Syndrome (PCS) scores and the rCBF values of all of the above mentioned regions.

Potential confounders

Patients’ blood alcohol levels were tested at the discretion of the attending emergency physicians in any case of suspected alcohol intoxication. Of the 14 patients in this study, one patient out of five tested had detectable alcohol in the blood in the ED, at 46 mg/dl. Eight patients received opiates as part of their ED treatment, four of whom had tested positive for opiates or benzodiazepines and two of whom tested positive for cannabinoids, and one patient received...
benzodiazepines. No other patients tested positive for opiates or benzodiazepines. Three patients received intravenous (IV) normal saline, and another received IV potassium chloride. Three reported a diagnosis of hypertension, while five had a systolic BP over 140 in the ED, including two patients with a systolic BP over 160.

Because of the small sample size, there was exceedingly low power for detecting effects of any of these as confounders, but there were no significant differences in susceptibility between patients with mTBI who did (n = 8) or did not receive (n = 6) opiates in ER (left thalamostriate vein p = 0.84, right basal vein of Rosenthal p = 0.70), between subjects who did (n = 3) or did not (n = 11) receive normal saline (left thalamostriate vein p = 0.46, right basal vein of Rosenthal p = 0.07), or between subjects who did (n = 4) or did not have (n = 10) positive opiate or benzodiazepine drug screen (left thalamostriate vein p = 0.24, right basal vein of Rosenthal p = 0.67). For the patients who received ASL imaging, there were no significant differences in rCBF between subjects who did (n = 2) or did not receive (n = 5) opiates in ER (left striatum p = 0.60, frontal lobe p = 0.73, and occipital lobe p = 0.96) or between subjects who did (n = 2) or did not receive (n = 5) normal saline (left striatum p = 0.29, frontal lobe p = 0.61, and occipital lobe p = 0.45). In summary, none of these possible confounders were found to have a significant effect on measured rCBF or susceptibility.

Discussion

To the best of our knowledge, this is the first study reporting cerebral hemodynamic changes using a non-invasive MRI method in the acute stage of mTBI. Our data have demonstrated that, as a result of concussion, the brain has increased rCBF and consequently higher venous oxygenation. This finding is in opposition to our original hypothesis that the brain may have decreased rCBF and decreased venous oxygenation. This is a novel finding in patients with mTBI in the acute post-TBI stage.
Results of ASL data analysis indicate that the left striatum has significantly higher rCBF in the patient than in the control group. This is in contrast to several studies indicating decreased rCBF after moderate-severe TBI [57–60]. This is likely the result of the differences between these studies and ours in both severity of the injury and timing of the scanning. All of our patients were classified as having mTBI and most were scanned at the acute stage (within 48 hours after injury), which may be expected to show different pathophysiology than the sub-acute or chronic stages of moderate or severe TBI. To our knowledge, such ultra-early scanning in this population has not been attempted in the past.

Although the increase in rCBF contradicts some of the earlier publications, there are a few studies indicating higher CBF after trauma [61–67]. Marion et al., observed that CBF peaked to a significant high at the 24 hour time point after injury [61], but studied a diverse range of injury pathologies at a wide range of times after injury. Obrist et al., studied the cerebral metabolic rate of oxygen (CMRO₂) and CBF in 75 severely-injured patients and observed a positive correlation between GCS score and CBF in patients with far lower GCS scores than ours. They also found that CBF reached its highest peak at 24 hours after injury [68], supporting the same timeline as seen in Marion’s study. Muizelaar et al., suggested that CMRO₂ is significantly positively correlated with GCS scores [64], though the study was limited to severe cases and therefore did not include patients with GCS scores as high as our patients had. Bouma et al., also observed that relative CBF reaches its peak between 24 to 48 hours after injury [62,65], which was supported by another study by the same group [63] and also by the results of the study by Mendelow et al., [69]. In an animal study, Prat et al., observed that the animals impacted with less weight (producing a less severe injury) showed an increase in CBF between one and three hours after injury, which was followed by a decrease in CBF thereafter [60].

SWI analysis indicates that relative susceptibility values in the veins of patients are lower than in the control group. In particular, the left thalamostriate vein and the right basal vein of Rosenthal had significantly lower susceptibility. This implies that there is more oxygen left in the veins in patients as compared to the control group, in contrast to earlier animal studies [70]. A study by Shen et al., found increased susceptibility induced by head injury using the Marmarou rat model [71]. However, the severity and mechanism of the injury in Shen’s study differs from ours. Considering this, there could be several explanations for the decreased relative susceptibility. One is that the decreased susceptibility could represent a deficit in oxygen consumption at tissue level after injury. In several studies it has been shown that the coupling between CMRO₂ and CBF has been disrupted following injury [72–75]. Decreased CMRO₂ has been observed in these studies, suggesting that either the tissue is unable to absorb the optimal amount of oxygen or that there is a reduction in oxygen demand. Another proposition that might explain the lower relative susceptibility value is that there might be increased rCBF in the regions drained by those veins; an increase in rCBF with stable CMRO₂ would result in decreased oxygen extraction and more oxygen left in the veins.

A fourth option is that the brain is utilizing another source of energy, such as lactate, instead of glucose. Several studies have shown this to occur in extreme conditions such as starvation and trauma [76], using substrates such as b-hydroxybutyrate (bHB) [77] or lactate [78]. A possible mechanism behind this could be efforts by the brain to maintain energy production in the presence of mitochondrial dysfunction and decreased CMRO₂. There have been few studies showing decreased CMRO₂ after TBI [79]. In the absence of any structural damage visible on CT or MRI, this could represent a temporary inability of the brain to use oxygen efficiently via glucose uptake. It is possible that the brain is attempting to overcome mitochondrial dysfunction and lowered CMRO₂ by increasing CBF.

It is also possible that this increase in CBF is a neuroprotective mechanism that is hindered by or lost in more severe injuries in which CBF is found to be decreased. Mild mitochondrial
dysfunction leads to an increase in anaerobic ATP production in order to meet demands. The brain attempts to maintain the energy equilibrium in the acute stage, which requires an increase in CBF to offset a decrease in efficiency of ATP production in affected areas of the brain. This decoupling of CBF and CMRO₂ explains the combination of increased CBF and increased venous oxygenation.

Unfortunately, the strong similarity between patient and control group SAC scores led to relatively low sensitivity to detect a correlation between rCBF or SWIM abnormalities and symptom presence or severity. Despite the advantage in ease of administering the SAC questionnaire, a more sensitive questionnaire or neuropsychological test should be applied in the future to more accurately characterize the deficits experienced by patients with mTBI.

There are several approaches to extracting oxygen saturation, including T2, T2* or T2' approaches [15–31,33,80–86], phase-based methods [32–37,52], and susceptometry [49]. All these methods have advantages and disadvantages. All of these methods are interesting, but they all meet with difficulties, such as with phase wrapping. Additionally, none of these are in common clinical use today except for susceptibility weighted imaging (SWI) and the phase approach leading to SWIM. By evaluating each major vein inside the brain parenchyma, we will be able to determine the blood oxygenation at a regional level. Similar to a brain catheter probe measuring surrounding brain tissue oxygenation, SWIM uses the major veins of the brain like embedded catheter probes for detecting surrounding brain tissue oxygenation, as well.

Veins vary in size from roughly 1 cm in diameter for the dural sinuses, to a few millimeters for pial veins, to only several hundred microns for medullary veins [87]. Our susceptibility measurements of major veins (between 70 and 150ppm) are significantly lower than that in superior sagittal sinus (SSS) measured in susceptometry approach. The different susceptibilities in veins with different sizes are due to its partial volume effects [88]. In looking at the relative changes of venous oxygenation between patients and controls, this partial volume effect may downplay the group difference instead of exaggerating the confounding factors. In comparison, a susceptometry approach using the SSS does not suffer partial volume effect but it only offers global oxygenation instead of regional information. Furthermore, since the SSS is located at the interface of brain tissue and skull, the strong artifact on phase images makes the SSS not ideal to evaluate on the current 3D SWI data. Additionally, the brain extraction step during SWIM processing often removes a small portion of the outer cortex along with the skull, as this area is generally noisy in the phase images. Part of the SSS is usually removed along with this, while the deeper veins of the brain are unaffected. For these reasons, we chose to analyze the deeper veins instead of the SSS; while they offer slightly lower SNR, we have greater confidence in the accuracy of the values obtained.

Our current study has several limitations that need to be discussed. First, our relatively small sample size provides low power. With a larger sample size, some of the relationships that are present in the data may turn out to be not significant. We are currently in the process of expanding this data set, and intend to investigate these relationships further. In addition, while the recruitment of the first few patients had a wide range delays between injury and MRI (with one subject having their first scan 10 days after injury, potentially confounding the results), subjects recruited later had their MRI within a much shorter time window, which should reduce some of the variability. The initial patients are still being followed up, as well, which will allow for a more longitudinal analysis of their recovery. Additionally, we will acquire more detailed neuropsychological data in our control group for a more relevant comparison. We also acknowledge that the SAC is considered as a sideline test to supplement a full battery of neuropsychological test. In a typical acute setting, it is not possible to hold the patient for an hour-long full battery neuropsychological test, which is also a confounding factor of the study. Finally, we must consider that the increase in rCBF could be a result of some other difference...
between the patients with mTBI and the control group like, for example, the stress related to injury present in patients with mTBI but not healthy controls enrolled in the study. Further, because of the difficulties in recruiting orthopedically injured controls from the ED, as well as increased difficulties with follow up as compared to control subjects recruited from the surrounding community, the patients may have received interventions which affect CBF that the controls have not. While none of the possible confounders identified were found to have a significant effect in this dataset, this possibility should be considered in a larger dataset that would provide more power for detecting or disproving the presence of such differences. One notable possible confounder is IV administration of normal saline, commonly given to most patients in the ED. Some studies have shown this to have an effect on CBF [89,90], but these studies have been performed in humans and animals with far different conditions than the patients in this study, including subarachnoid hemorrhage and cardiac arrest. What we are currently lacking are studies of the effects of normal saline on rCBF in normovolemic, non-hypoxic patients with normal intracranial pressure. We do not anticipate that standard, IV-administered saline will have a significant effect on CBF, which is supported by the changes in CMRO2; if the CBF changes were artifacts from volume expansion or increased cardiac output, we would not expect the CMRO2 to change.

In the future, we plan to take these analyses a step further and to calculate the cerebral metabolic rate of oxygen (CMRO2) for all patients scanned. This will require the addition of several 2D SWI sequences with higher resolution, in order to obtain absolute quantification while avoiding partial volume effects in very small veins. This measurement will allow us to better describe the metabolic status of the tissue.

In summary, imaging of changes in hemodynamics and metabolism following mTBI can clearly provide us with more information about the pathophysiology that underlies the symptoms, progression, and recovery of brain injury. In this study, we demonstrated that patients with mTBI have increased cerebral blood supply as a potential compensatory mechanism to cope with brain injury at the acute stage. Further investigation in larger cohort is needed to establish the relationship between rCBF and tissue oxygenation and its neurocognitive underpinnings.

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Author Contributions
Conceived and designed the experiments: ZK RW BO EMH. Performed the experiments: HD NW ZK VM CZ RW BO. Analyzed the data: HD NW JL WW XW EMH ZK. Wrote the paper: HD NW JL WW XW JS ZK.

References
1. Faul M, Xu L, Wald M, Coronado V (2010) Traumatic Brain Injury in the United States: Emergency Department Visits, Hospitalizations, and Deaths, 2002–2006. Atlanta (GA): Centers for Disease Control and Prevention, National Center for Injury Prevention and Control. PMID:25506974
2. Levin HS, Robertson CS (2013) Mild traumatic brain injury in translation. J Neurotrauma 30: 610–617. doi: 10.1089/neu.2012.2394 PMID: 23046349
3. McCrea M, Iverson GL, McAllister TW, Hammeke TA, Powell MR, et al. (2009) An integrated review of recovery after mild traumatic brain injury (MTBI): implications for clinical management. Clin Neuropsychol 23: 1368–1390. doi: 10.1080/13854040903074652 PMID: 19882476
4. Pomschar A, Koerte I, Lee S, Laubendörfer R, Straub A, et al. (2013) MRI evidence for altered venous drainage and intracranial compliance in mild traumatic brain injury. PLoS One 8: e55447. doi: 10.1371/journal.pone.0055447 PMID: 23405151

5. Zumstein MA, Moser M, Mottini M, Ott SR, Sadowski-Cron C, et al. (2011) Long-Term Outcome in Patients With Mild Traumatic Brain Injury: A Prospective Observational Study. Journal of Trauma and Acute Care Surgery 71: 120–127 110.1097/TA.1090b1013e3181f1092d1670. doi: 10.1097/TA.0b013e3181f2d670 PMID: 21045743

6. McAllister TW, Spalinger MB, Flashman LA, Saykin AJ (2001) Neuroimaging findings in mild traumatic brain injury. J Clin Exp Neuropsychol 23: 775–791. PMID: 11910544

7. Barnes SR, Haacke EM (2009) Susceptibility-weighted imaging: clinical angiographic applications. Magn Reson Imaging Clin N Am 17: 47–61. doi: 10.1016/j.mric.2008.12.002 PMID: 19364599

8. Tong KA, Ashwal S, Holshouser BA, Nickerson JP, Wall CJ, Shutter LA, et al. (2004) Diffuse axonal injury in children: clinical correlation with hemorrhagic lesions. Ann Neurol 56: 36–50. PMID: 15236400

9. Ge Y, Patel MB, Chen Q, Grossman EJ, Zhang K, et al. (2009) Assessment of thalamic perfusion in patients with mild traumatic brain injury by true FISP arterial spin labelling MR imaging at 3T. Brain Injury 23: 666–674. doi: 10.1080/02699050903014899 PMID: 19557570

10. Liu W, Wang B, Wolfowitz R, Yeh P-H, Nathan DE, et al. (2013) Perfusion deficits in patients with mild traumatic brain injury characterized by dynamic susceptibility contrast MRI. NMR in Biomedicine 26: 651–663. doi: 10.1002/nbm.2910 PMID: 23456696

11. Abate MG, Trivedi M, Fryer T, Smielewski P, Chatfield D, et al. (2008) Early Derangements in Oxygen and Glucose Metabolism Following Head Injury: The Ischemic Penumbra and Pathophysiological Heterogeneity. Neurocritical Care 9: 319–325. doi: 10.1007/s12028-008-9119-2 PMID: 18563636

12. Coles JP, Fryer TD, Smielewski P, Chatfield DA, Steiner LA, et al. (2004) Incidence and mechanisms of cerebral ischemia in early clinical head injury. J Cereb Blood Flow Metab 24: 202–211. PMID: 14747747

13. Zauner A, Daugherty WP, Bullock MR, Warner DS (2002) Brain oxygenation and energy metabolism: part I-biological function and pathophysiology. Neurosurgery 51: 289–301; discussion 302. PMID: 12182767

14. Zwienenberg M, Muizelaar JP (2001) Cerebral perfusion and blood flow in neurotrauma. Neurol Res 23: 167–174. PMID: 11320595

15. An H, Lin W (2000) Quantitative measurements of cerebral blood oxygen saturation using magnetic resonance imaging. J Cereb Blood Flow Metab 20: 1225–1236. PMID: 10950383

16. An H, Lin W (2002) Cerebral oxygen extraction fraction and cerebral venous blood volume measurements using MRI: effects of magnetic field variation. Magn Reson Med 47: 958–966. PMID: 11979575

17. An H, Lin W, Celik A, Lee YZ (2001) Quantitative measurements of cerebral metabolic rate of oxygen utilization using MRI: a volunteer study. NMR Biomed 14: 441–447. PMID: 11746936

18. Chien D, Levin DL, Anderson CM (1994) MR gradient echo imaging of intravascular blood oxygenation: T2* determination in the presence of flow. Magn Reson Med 32: 540–545. PMID: 7997123

19. Golay X, Silvennoinen MJ, Zhou J, Clingman CS, Kauppinen RA, et al. (2001) Measurement of tissue oxygen extraction ratios from venous blood T2(2): increased precision and validation of principle. Magn Reson Med 46: 282–291. PMID: 11477631

20. He X, Zhu M, Yablonskiy DA (2008) Validation of oxygen extraction fraction measurement by qBOLD technique. Magn Reson Med 60: 882–886. doi: 10.1002/mrm.21719 PMID: 18816808

21. He X, Yablonskiy DA (2007) Quantitative BOLD: mapping of human cerebral deoxygenated blood volume and oxygen extraction fraction: default state. Magn Reson Med 57: 115–126. PMID: 17191227

22. Hoppel BE, Weisskoff RM, Thulborn KR, Moore JB, Kwong KK, et al. (1993) MR gradient echo imaging of intravascular blood oxygenation and cerebral hemodynamics. Magn Reson Med 30: 715–723. PMID: 8139453

23. Li D, Waight DJ, Wang Y (1998) In vivo correlation between blood T2* and oxygen saturation. J Magn Reson Imaging 8: 1236–1239. PMID: 9848734

24. Li D, Waight Y, Waight DJ (1998) Blood oxygen saturation assessment in vivo using T2* estimation. Magn Reson Med 39: 685–690. PMID: 9581597

25. Lu H, Ge Y (2008) Quantitative evaluation of oxygenation in venous vessels using T2-Relaxation-Under-Spin-Tagging MRI. Magn Reson Med 60: 357–363. doi: 10.1002/mrm.21627 PMID: 18666116

26. Oja JM, Gillen JS, Kauppinen RA, Kraut M, van Zijl PC (1999) Determination of oxygen extraction ratios by magnetic resonance imaging. J Cereb Blood Flow Metab 19: 1289–1295. PMID: 10598932

27. Qin Q, Gracik K, van Zijl PC (2011) Determination of whole-brain oxygen extraction fractions by fast measurement of blood T2(2) in the jugular vein. Magn Reson Med 65: 471–479. doi: 10.1002/mrm.22556 PMID: 21264936
van Zijl PC, Eleff SM, Ulatowski JA, Oja JM, Ulug AM, et al. (1998) Quantitative assessment of blood flow, blood volume and blood oxygenation effects in functional magnetic resonance imaging. Nat Med 4: 159–167. PMID: 9461188

Wright GA, Hu BS, Macovski A (1991) 1991 I.I. Rabi Award. Estimating oxygen saturation of blood in vivo with MR imaging at 1.5 T. J Magn Reson Imaging 1: 275–283. PMID: 1802140

Weiskoff R, Kihne S, Cohen M, Thuiborn K. Quantitative in vivo blood oxygenation measurements by echo planar imaging at 1.5 Tesla; 1991. pp. 307.

Xu F, Ge Y, Lu H (2009) Noninvasive quantification of whole-brain cerebral metabolic rate of oxygen (CMRO2) by MRI. Magn Reson Med 62: 141–148. doi: 10.1002/mrm.21994 PMID: 19526517

Wright GA, Hu BS, Macovski A (1991) 1991 I.I. Rabi Award. Estimating oxygen saturation of blood in vivo with MR imaging at 1.5 T. J Magn Reson Imaging 1: 275–283. PMID: 1802140

Weiskoff R, Kihne S, Cohen M, Thuiborn K. Quantitative in vivo blood oxygenation measurements by echo planar imaging at 1.5 Tesla; 1991. pp. 307.

Xu F, Ge Y, Lu H (2009) Noninvasive quantification of whole-brain cerebral metabolic rate of oxygen (CMRO2) by MRI. Magn Reson Med 62: 141–148. doi: 10.1002/mrm.21994 PMID: 19526517

Wright GA, Hu BS, Macovski A (1991) 1991 I.I. Rabi Award. Estimating oxygen saturation of blood in vivo with MR imaging at 1.5 T. J Magn Reson Imaging 1: 275–283. PMID: 1802140

Weiskoff R, Kihne S, Cohen M, Thuiborn K. Quantitative in vivo blood oxygenation measurements by echo planar imaging at 1.5 Tesla; 1991. pp. 307.

Xu F, Ge Y, Lu H (2009) Noninvasive quantification of whole-brain cerebral metabolic rate of oxygen (CMRO2) by MRI. Magn Reson Med 62: 141–148. doi: 10.1002/mrm.21994 PMID: 19526517

Wright GA, Hu BS, Macovski A (1991) 1991 I.I. Rabi Award. Estimating oxygen saturation of blood in vivo with MR imaging at 1.5 T. J Magn Reson Imaging 1: 275–283. PMID: 1802140

Weiskoff R, Kihne S, Cohen M, Thuiborn K. Quantitative in vivo blood oxygenation measurements by echo planar imaging at 1.5 Tesla; 1991. pp. 307.

Xu F, Ge Y, Lu H (2009) Noninvasive quantification of whole-brain cerebral metabolic rate of oxygen (CMRO2) by MRI. Magn Reson Med 62: 141–148. doi: 10.1002/mrm.21994 PMID: 19526517

Wright GA, Hu BS, Macovski A (1991) 1991 I.I. Rabi Award. Estimating oxygen saturation of blood in vivo with MR imaging at 1.5 T. J Magn Reson Imaging 1: 275–283. PMID: 1802140

Weiskoff R, Kihne S, Cohen M, Thuiborn K. Quantitative in vivo blood oxygenation measurements by echo planar imaging at 1.5 Tesla; 1991. pp. 307.

Xu F, Ge Y, Lu H (2009) Noninvasive quantification of whole-brain cerebral metabolic rate of oxygen (CMRO2) by MRI. Magn Reson Med 62: 141–148. doi: 10.1002/mrm.21994 PMID: 19526517

Wright GA, Hu BS, Macovski A (1991) 1991 I.I. Rabi Award. Estimating oxygen saturation of blood in vivo with MR imaging at 1.5 T. J Magn Reson Imaging 1: 275–283. PMID: 1802140

Weiskoff R, Kihne S, Cohen M, Thuiborn K. Quantitative in vivo blood oxygenation measurements by echo planar imaging at 1.5 Tesla; 1991. pp. 307.
51. Pandian DS, Ciulla C, Haacke EM, Jiang J, Ayaz M (2008) Complex threshold method for identifying pixels that contain predominantly noise in magnetic resonance images. J Magn Reson Imaging 28: 727–735. doi: 10.1002/jmri.21487 PMID: 18777533

52. Fernández-Seara MA, Techawiboonwong A, Detre JA, Wehrli FW (2006) MR susceptometry for measuring global brain oxygen extraction. Magnetic Resonance in Medicine 55: 967–973. PMID: 16598726

53. Abdul-Khaliq H, Troitzsch D, Berger F, Lange PE (2000) [Regional transcranial oximetry with near infrared spectroscopy (NIRS) in comparison with measuring oxygen saturation in the jugular bulb in infants and children for monitoring cerebral oxygenation]. Biomed Tech (Berl) 45: 328–332. PMID: 11155535

54. Macmillan CS, Andrews PJ (2000) Cerebrovenous oxygen saturation monitoring: practical considerations and clinical relevance. Intensive Care Med 26: 1028–1036. PMID: 11030158

55. McCrea M, Guskiewicz KM, Marshall SW, Barr W, Randolph C, et al. (2003) Acute effects and recovery time following concussion in collegiate football players: the NCAA Concussion Study. JAMA 290: 2556–2563. PMID: 14625332

56. Naunheim RS, Matero D, Fucetola R (2008) Assessment of patients with mild concussion in the emergency department. J Head Trauma Rehabil 23: 116–122. doi: 10.1097/HTR.0b013e318177b026 PMID: 18362765

57. Fieschi C, Battistini N, Beduschi A, Boselli L, Rossanda M (1974) Regional cerebral blood flow and intraventricular pressure in acute head injuries. J Neurol Neurosurg Psychiatry 37: 1378–1388. PMID: 4375173

58. Overgaard J, Tweed WA (1974) Cerebral circulation after head injury. 1. Cerebral blood flow and its regulation after closed head injury with emphasis on clinical correlations. J Neurosurg 41: 531–541. PMID: 4418221

59. Barclay L, Zemcov A, Reichert W, Blass JP (1985) Cerebral blood flow decrements in chronic head injury syndrome. Biol Psychiatry 20: 146–157. PMID: 3970995

60. Prat R, Markiv V, Dujovny M, Misra M (1997) Evaluation of cerebral autoregulation following diffuse brain injury in rats. Neurol Res 19: 393–402. PMID: 9263220

61. Marion DW, Darby J, Yonas H (1991) acute regional cerebral blood flow changes caused by severe head injuries. J Neurosurg 74: 407–414. PMID: 1899694

62. Bouma GJ, Muizelaar JP (1992) Cerebral blood flow, cerebral blood volume, and cerebrovascular reactivity after severe head injury. J Neurotrauma 9 Suppl 1: S333–348. PMID: 1588625

63. Bouma GJ, Muizelaar JP (1995) Cerebral blood flow in severe clinical head injury. New Horiz 3: 384–394. PMID: 7496746

64. Muizelaar JP, Marmarou A, DeSalles AA, Ward JD, Zimmerman RS, et al. (1989) Cerebral blood flow and metabolism in severely head-injured children. Part 1: Relationship with GCS score, outcome, ICP, and PVI. J Neurosurg 71: 63–71. PMID: 2739843

65. Bouma GJ, Muizelaar JP (1993) Evaluation of regional cerebral blood flow in acute head injury by stable xenon-enhanced computerized tomography. Acta Neurochir Suppl (Wien) 59: 34–40. PMID: 8310860

66. Kelly DF, Martin NA, Kordestani R, Counelis G, Hovda DA, et al. (1997) Cerebral blood flow as a predictor of outcome following traumatic brain injury. J Neurosurg 86: 633–641. PMID: 9120627

67. Kelly DF, Kordestani RK, Martin NA, Nguyen T, Hovda DA, et al. (1996) Hyperemia following traumatic brain injury: relationship to intracranial hypertension and outcome. J Neurosurg 85: 762–771. PMID: 8893712

68. Obrist WD, Langfitt TW, Jaggi JL, Cruz J, Gennarelli TA (1984) Cerebral blood flow and metabolism in comatose patients with acute head injury. Relationship to intracranial hypertension. J Neurosurg 61: 241–253. PMID: 6737048

69. Mendelow AD, Teasdale GM, Russell T, Flood J, Patterson J, et al. (1985) Effect of mannitol on cerebral blood flow and cerebral perfusion pressure in human head injury. J Neurosurg 63: 43–48. PMID: 3925092

70. Shen Y, Kou Z, Kreipke CW, Petrov T, Hu J, et al. (2007) In vivo measurement of tissue damage, oxygen saturation changes and blood flow changes after experimental traumatic brain injury in rats using susceptibility weighted imaging. Magn Reson Imaging 25: 219–227. PMID: 17275617

71. Marmarou A, Foda MA, van den Brink W, Campbell J, Kita H, et al. (1994) A new model of diffuse brain injury in rats. Part I: Pathophysiology and biomechanics. J Neurosurg 80: 291–300. PMID: 8263269

72. Chen SF, Richards HK, Smielewski P, Johnstrom P, Salvador R, et al. (2004) Relationship between flow-metabolism uncoupling and evolving axonal injury after experimental traumatic brain injury. J Cereb Blood Flow Metab 24: 1025–1036. PMID: 15366423
73. Harris NG, Mironova YA, Chen SF, Richards HK, Pickard JD (2012) Preventing flow-metabolism uncoupling acutely reduces axonal injury after traumatic brain injury. J Neurotrauma 29: 1469–1482. doi: 10.1089/neu.2011.2161 PMID: 22321027

74. Vink R, Faden AI, McIntosh TK (1988) Changes in cellular bioenergetic state following graded traumatic brain injury in rats: determination by phosphorus 31 magnetic resonance spectroscopy. J Neurotrauma 5: 315–330. PMID: 3249310

75. Hovda DA, Yoshino A, Kawamata T, Katayama Y, Becker DP (1991) Diffuse prolonged depression of cerebral oxidative metabolism following concussive brain injury in the rat: a cytochrome oxidase histochemistry study. Brain Res 567: 1–10. PMID: 1667742

76. Prins ML, Lee SM, Fujima LS, Hovda DA (2004) Increased cerebral uptake and oxidation of exogenous betaHB improves ATP following traumatic brain injury in adult rats. J Neurochem 90: 666–672. PMID: 15255945

77. McKenna MC, Tildon JT, Stevenson JH, Hopkins IB (1994) Energy metabolism in cortical synaptic terminals from weanling and mature rat brain: evidence for multiple compartments of tricarboxylic acid cycle activity. Dev Neurosci 16: 291–300. PMID: 7768208

78. Brandt RB, Waters MG, Rispler MJ, Kline ES (1984) D- and L-lactate catabolism to CO2 in rat tissues. Proc Soc Exp Biol Med 175: 328–335. PMID: 6420798

79. Vespa P, Bergsneider M, Hattori N, Wu HM, Huang SC, et al. (2005) Metabolic crisis without brain ischemia is common after traumatic brain injury: a combined microdialysis and positron emission tomography study. J Cereb Blood Flow Metab 25: 763–774. PMID: 15716852

80. Weiskoff R, et al. Quantitative in vivo blood oxygenation measurements by echo planar imaging at 1.5 Tesla; 1991.

81. Lu H, Ge Y (2008) Quantitative evaluation of oxygenation in venous vessels using T2-Relaxation-Under-Spin-Tagging MRI. Magnetic Resonance in Medicine 60: 357–363. doi: 10.1002/mrm.21627 PMID: 18661116

82. Sedlacik J, Rauscher A, Reichenbach JR (2009) Quantification of modulated blood oxygenation levels in single cerebral veins by investigating their MR signal decay. Z Med Phys 19: 48–57. PMID: 19459858

83. Sedlacik J, Reichenbach JR (2010) Validation of quantitative estimation of tissue oxygen extraction fraction and deoxygenated blood volume fraction in phantom and in vivo experiments by using MRI. Magn Reson Med 63: 910–921. doi: 10.1002/mrm.22274 PMID: 20373392

84. Yablonisky DA, Haacke EM (1994) Theory of NMR signal behavior in magnetically inhomogeneous tissues: the static dephasing regime. Magn Reson Med 32: 749–763. PMID: 7869897

85. Yablonisky DA, Haacke EM (1997) An MRI method for measuring T2 in the presence of static and RF magnetic field inhomogeneities. Magn Reson Med 37: 872–876. PMID: 9178238

86. Yablonisky DA, Reinus WR, Stark H, Haacke EM (1997) Quantitation of T2' anisotropic effects on magnetic resonance bone mineral density measurement. Magn Reson Med 37: 214–221. PMID: 9001145

87. Nonaka H, Akima M, Hatori T, Nagayama T, Zhang Z, et al. (2003) Microvasculature of the human cerebrum: Arteries of the deep white matter. Neuropathology 23: 111–118. PMID: 12777099

88. Zheng W, Nichol H, Liu S, Cheng YC, Haacke EM (2013) Measuring iron in the brain using quantitative susceptibility mapping and X-ray fluorescence imaging. Neuroimage 78: 68–74. doi: 10.1016/j.neuroimage.2013.04.022 PMID: 23591072

89. Jost SC, Diringer MN, Zazulia AR, Videen TO, Aiyagari V, et al. (2005) Effect of normal saline bolus on cerebral blood flow in regions with low baseline flow in patients with vasospasm following subarachnoid hemorrhage. Journal of Neurosurgery 103: 25–30. PMID: 16121969

90. Krep H, Breil M, Sinn D, Hagendorff A, Hoeff A, et al. (2004) Effects of hypertonic versus isotonic infusion therapy on regional cerebral blood flow after experimental cardiac arrest cardiopulmonary resuscitation in pigs. Resuscitation 63: 73–83. PMID: 15451589