Five tumor-associated autoantibodies expression levels in serum predict lung cancer and associate with poor outcome

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Background: This study aims to evaluate diagnostic feasibility of the tumor-associated autoantibodies and their clinical significance in lung cancer.

Methods: Blood samples were collected from 352 lung cancer patients and 226 healthy individuals. Immunohistochemical staining was used to evaluate the expression of autoantibodies in 69 cases of lung cancer tissues. Enzyme-linked immunosorbent assay (ELISA) was performed to measure the five autoantibodies levels in serum from lung cancer patients and healthy controls.

Results: Higher expression of GAGE7, MAGEA1, PGP9.5, CAGE and p53 were found in lung cancer tissues than normal tissues from the cancer genome atlas (TCGA) database. The expression levels of the five tumor-associated autoantibodies in lung cancer tissues were positively associated with those statuses in serum. There were significantly higher levels of each autoantibody in lung cancer patients than healthy individuals. The area under curve (AUC) for GAGE7, MAGEA1, PGP9.5, CAGE and p53 in diagnosis of lung cancer were 0.787, 0.641, 0.873, 0.825 and 0.749, respectively. With combination of these five autoantibodies by the logistic regression model, 0.891 of AUC was shown. In addition, the level of each autoantibody was increased in the advanced stage. Particularly, the level of PGP9.5 was significantly higher in the advanced T stage, N stage and TNM stage and CAGE was significantly elevated in the advanced N stage.

Conclusions: Summarily, autoantibodies levels of GAGE7, MAGEA1, PGP9.5, CAGE and p53 in serum could be used for the diagnosis of lung cancer and PGP9.5 may predict poor prognosis for the patients with lung cancer.

Keywords: Tumor-associated autoantibodies; lung cancer; diagnosis

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Introduction

Lung cancer is the most common cause of death from malignant cancer worldwide (1,2). Surgical resection is the primary and effective option for lung cancer, but only 20% to 25% of patients are eligible for the resection (3). The majority of patients with lung cancer are diagnosed with advanced disease owing to the paucity of early diagnostic methods. Biopsy and pathologic examination were widely used to diagnose lung cancer when lesion was discovered by chest X-ray or computerized tomography (CT). A number of biomarkers such as carcinoembryonic antigen (CEA) in blood have been identified in the clinic to assist diagnosis of lung cancer (4,5). Due to low expression of these biomarkers, they had limited sensitivity to be considered as
appropriately screening method for lung cancer (6,7).

Autoantibody which can reflect a host’s immune response to tumor-associated antigens (TAAs) have been validated for early detection of lung cancer and subsequent treatment (8-10). Currently, we aimed to explore the autoantibodies for screening lung cancer. We extracted TAAs for lung cancer from the cancer genome atlas (TCGA) database and further describe how expression status of the corresponding tumor-associated autoantibodies in serum to assess the malignance of pulmonary nodules and their significant in the clinic.

**Methods**

**Patients and specimens**

In current study, 352 sera were consecutively collected from the patients with primary lung cancer which were diagnosed by pathological examination through thoracotomy or needle biopsy and 69 of tumor tissues were also got among these patients from January 2015 to January 2017 in the Department of Thoracic Surgery II of Peking University Cancer Hospital. Those patients with lung benign lesion were excluded. In the meantime, 226 cases normal individuals with healthy physical examination were included in this study from the Department of Thoracic Surgery of Jilin Guowen Hospital. Those individuals with malignant tumor history were excluded. The stage was classified according to tumor-lymph node-metastasis (TNM) classification system and the American Joint Committee on Cancer (AJCC). Three hundred and forty-nine cases of patients were detected with carcino-embryonic antigen (CEA) in our department of clinical laboratory. Two hundred and eighty-six patients were examined by positron emission computed tomography (PET)-CT. In addition, 226 cases of sera from normal individuals with healthy physical examination. The participants signed informed consent and this study has been approved with the Ethics Committee of Peking University Cancer Hospital.

**Immunohistochemical staining and score evaluation**

According to our previous study (11), immunohistochemistry assay was done. Briefly, formalin-fixed sections were dehydrated using the graded serial ethanol solution. Antigen retrieval was performed in the 10 mM citrate buffer at 98–100 °C for 30 min and 10% normal goat serum was used to block the unspecific binding for 30 min. The primary antibodies including mouse monoclonal anti-GAGE7 antibody (1:100 dilution; Abcam, UK), mouse monoclonal anti-MAGEA1 antibody (1:100 dilution; Absin, China), mouse monoclonal anti-PGP9.5 antibody (1:200 dilution; Abcam, UK), mouse monoclonal anti-CAGE antibody (1:100 dilution; Absin, China) and mouse monoclonal anti-p53 antibody (1:200 dilution; Abcam, UK) were added on the section overnight at 4 °C, followed by incubation with HRP-conjugated goat anti-mouse secondary antibody for 30 min at room temperature (Sigma-Aldrich, Poole, Dorset, UK).

Immunohistochemical evaluation was graded based on the intensity of staining (1+, weak; 2+, moderate; 3+, strong) and the percentage of stained cells (0, for <5%; 1, for 5–25%; 2, for 26–50% and 3, >50%) (12). The score used for the analyses was the average values. When combining these two parameters, 0–1 and >1 were considered negative and positive staining, respectively. Scoring was reviewed in parallel by two experienced pathologists who were blinded to all clinical data.

**Enzyme-linked immunosorbent assay (ELISA)**

Serum samples from patients and healthy individuals were tested by ELISA assay. Purified recombinant proteins of GAGE7, MAGEA1, PGP9.5 CAGE and p53, were coated onto 96-well microtiter plates (CancerProbe, China). Serum samples from patients and healthy individuals were diluted at 1:100 and added in the plated for 1h at room temperature. After the samples were washed with PBS, HRP-conjugated secondary antibody was used to incubating the samples for 30 min. The optical density value of each well was read at 450 nm, and the relative expression levels of the five autoantibodies were calculated based on the standard curve of each plate. The value for concentration of the 5 autoantibodies in serum were defined as unit (U). This experiment for all the samples were tested in duplicate.

**Statistical analysis**

Pearson’s correlation method was used to analyze the correlation of autoantibodies expression between serum and lung cancer tissues. Associations between the autoantibodies’ expressions and patients’ clinical variables were assessed using t-test. The receiver operating characteristic (ROC) analysis and logistic regression were used to evaluate the diagnostic value of 5 autoantibodies. When we constructed the logistic model to evaluate the combination of five autoantibodies, we used the multivariate regression analysis to calculate the prediction probability of combination of five autoantibodies for each patient and then used the ROC analysis to evaluate
the diagnostic value. All statistical analyses were performed using SPSS 23.0 statistical software. The data are presented as the means ± standard error of the mean (SEM). P<0.05 was considered significant difference.

**Results**

**Screening of TAAs for lung cancer patients**

Firstly, we selected the commonly reported TAAs including AnnexinA1 (13,14), AnnexinA2 (15), CathepsinD (16), CyclinD1 (17), p53 (18), PGP9.5 (19), Recoverin (20), Paxillin (21), MALAT1 (22), GAGE7 (23), CAGE (24,25), MAGEA1 (26) and Myc (27) and analyzed these autoantibodies expressions in the 403 cases of lung cancers (241 cases of adenocarcinoma and 162 squamous cell carcinoma) and 57 cases of normal lung tissues from TCGA database of RNA-sequence (RNA-SEQ) (Figure 1A). GAGE7, MAGEA1, PGP9.5, CAGE and p53 were observed to be highly expressed in the lung cancer tissues compared to normal tissues. The correlation analysis for five autoantibodies between tumor tissues and serum from lung cancer patients was done (Figure 1B-G).
normal tissues with more than 1.5-fold increase.

**Correlation of tumor-associated autoantibodies between tumor tissues and serum of lung cancer patients**

Then, we detected GAGE7, MAGEA1, PGP9.5, CAGE and p53 protein expression in 69 cases of lung cancer tissues by immunohistochemistry (Figure 1B). Moreover, these five autoantibodies were also investigated in the corresponding serum samples from these 69 lung cancer patients. As shown in Figure 1C,D,E,F,G and Table 1, positive association were shown in lung cancer tissues and serum samples for GAGE7, MAGEA1, PGP9.5, CAGE and p53 detection (r=0.4296, P=0.0002; r=0.4634, P<0.0001; r=0.4731, P<0.0001; r=0.3352, P=0.0049; r=0.2653, P=0.0276; respectively). The results indicated that expression level of each single autoantibody in serum were consistent with their protein expression degrees in lung cancer tissues.

**Diagnostic potential of tumor-associated autoantibodies in serum for lung cancer**

We asked how expression levels of the autoantibodies such as GAGE7, MAGEA1, PGP9.5, CAGE and p53 varied in serum between the patients with lung cancer and normal individuals. We tested the concentration of these five autoantibodies in serum from 352 lung cancer patients and 226 normal individuals. For GAGE7, MAGEA1, PGP9.5, CAGE and p53 expression, there were 25.3±4.6, 12.8±2.3, 14.8±2.2, 11.7±1.5, 19.6±2.2 U/mL in lung cancer patients and 3.8±0.2, 5.0±1.5, 5.0±0.3, 4.6±0.7, 8.2±1.4 U/mL in normal individuals, respectively (Figure 2A). A t-test showed, there were significantly higher levels of these five autoantibodies in lung cancer patients compared to the normal individuals (P<0.0001, 0.005, <0.0001, <0.0001, <0.0001 for GAGE7, MAGEA1, PGP9.5, CAGE and p53, respectively).

**Table 1** The correlation analysis for autoantibodies expression between tumor tissues and serum from lung cancer patients.

| Autoantibodies | Negative, N (%) | 1+, N (%) | 2+, N (%) | 3+, N (%) | r value | P value |
|----------------|-----------------|-----------|-----------|-----------|---------|---------|
| GAGE7          | 17 (24.6)       | 30 (43.5) | 18 (26.1) | 4 (5.8)   | 0.4296  | 0.0002  |
| MAGEA1         | 11 (15.9)       | 37 (53.6) | 19 (27.5) | 2 (2.9)   | 0.4634  | <0.0001 |
| PGP9.5         | 38 (55.1)       | 17 (24.6) | 11 (15.9) | 3 (4.3)   | 0.4731  | <0.0001 |
| CAGE           | 27 (39.1)       | 17 (24.6) | 22 (31.9) | 3 (4.3)   | 0.3352  | 0.0049  |
| p53            | 25 (36.2)       | 18 (26.1) | 11 (15.9) | 15 (21.7) | 0.2653  | 0.0276  |
We further analyzed whether these five autoantibodies could discriminate the patients with lung cancer and normal individuals. The sensitivity, specificity, optimal cut-off value and area under curve (AUC) of each autoantibody were demonstrated in Table 2. To estimate the risk of individuals being diagnosed with lung cancer, ecoefficiency value was generated for each autoantibody by logistic regression model (Table 2). The combinational predictive probability was then created as following Eq.: \[ \text{PRE} = \frac{1}{1 + \exp(-3.054 + 0.103 \times \text{GAGE7} + 0.001 \times \text{MAGEA1} + 0.406 \times \text{PGP9.5} + 0.006 \times \text{CAGE} + 0.002 \times \text{p53})} \]. For combinational autoantibodies, we could observe 90.1%, 71.7% and 0.891 for the sensitivity, specificity and AUC (Table 2), showing better predictive performance than each signal autoantibody (Figure 2B).

### Clinical significance of five tumor-associated autoantibodies for lung cancer patients

We further analyzed relationship between expression status of GAGE7, MAGEA1, PGP9.5, CAGE and p53 in serum from 352 lung cancer patients and their clinical characteristics like age, gender, pathological type, T stage, N stage and TNM stage, SUV\(_{\text{max}}\) of primary tumor and CEA level. The clinical features for these patients are summarized in Table 3. There was no statistically significant difference between age and any autoantibody (Figure 3A). The p53 level was higher in male than female (Figure 3B, \(P=0.031\)). The level of MAGEA1 and p53 were highly expressed in patients of squamous cell carcinoma than adenocarcinoma and other pathological types (Figure 3C, \(P=0.026\) and 0.014, respectively). Furthermore, increased levels for each autoantibody was found, when the T stage, N stage and TNM stage got more advanced (Figure 3D,E,F). Among these five autoantibodies, PGP9.5 level was significantly elevated in serious T stage, N stage and TNM stage (\(P=0.004, 0.005,\) and 0.032, respectively). The CAGE was significantly higher in serious N stage (\(P=0.017\)). Increase of every autoantibody concentration in serum was demonstrated in the lung cancer patients with SUV\(_{\text{max}}\) of tumor \(>4.0\) than SUV\(_{\text{max}}\) \(\leq 4.0\), and there were statistically significant differences for GAGE7 and CAGE (Figure 3G, \(P=0.008\) and 0.028, respectively). The expression level for any autoantibody was higher in the patients with positive CEA (CEA \(>5.0\) ng/mL) than the patients with negative CEA, although the difference was not statistically significant (Figure 3H). These results indicated that these autoantibodies were associated with gender, pathological type, T stage, N stage and TNM stage and SUV\(_{\text{max}}\) of primary tumor for the lung cancer patients.

### Discussion

For lung cancer, diagnosis often occurs so late that approximately two-thirds of patients have lost the opportunity for radical surgery (28,29). Lung cancer screening using low-dose CT is recommended in select high-risk smokers, and there are 24.2% positive screening tests. It is necessary to identify the malignant lung nodules from positive screening results. Detection of autoantibody in serum to be potential biomarker for diagnosis of cancer has been developed (30,31). With analysis of the TCGA database, we selected the highly expressed TAAs including GAGE7, MAGEA1, PGP9.5, CAGE and p53 genes in lung cancer tissues at mRNA level. Autoantibodies against GAGE7, MAGEA1 and CAGE which are members of the cancer testis antigen family, have also been reported in lung cancer (10,32). Antibodies recognizing p53 protein were first detected in breast cancer patients (33), and were later reported in the

| Autoantibodies | Sensitivity | Specificity | Cut-off value (U/mL) | AUC (95% CI) | Coefficiency |
|---------------|-------------|-------------|---------------------|--------------|--------------|
| GAGE7         | 0.756       | 0.739       | 3.7                 | 0.787 (0.749–0.825) | 0.103        |
| MAGEA1        | 0.364       | 0.85        | 3.4                 | 0.641 (0.596–0.686) | 0.001        |
| PGP9.5        | 0.818       | 0.757       | 6.2                 | 0.873 (0.844–0.902) | 0.406        |
| CAGE          | 0.739       | 0.832       | 3.8                 | 0.825 (0.789–0.861) | 0.006        |
| p53           | 0.724       | 0.704       | 6.1                 | 0.749 (0.708–0.789) | 0.002        |
| Combination   | 0.901       | 0.717       | 0.5008*             | 0.891 (0.864–0.917) | –            |

*, predictive probability. ROC, receiver operating characteristic; AUC, area under curve.
Table 3 Clinicopathological of 352 lung cancer patients

| Variables         | Lung cancers, N (%) |
|-------------------|---------------------|
| **Age**           |                     |
| ≤60               | 168 (47.7)          |
| >60               | 184 (52.3)          |
| **Gender**        |                     |
| Male              | 184 (52.3)          |
| Female            | 168 (47.7)          |
| **Smoking history**|                    |
| Yes               | 143 (40.6)          |
| No                | 209 (59.4)          |
| **Drinking history**|                    |
| Yes               | 75 (21.3)           |
| No                | 277 (78.7)          |
| **Pathological type**|                  |
| Adenocarcinoma    | 275 (78.1)          |
| Squamous cell carcinoma | 66 (18.8) |
| Others            | 11 (3.1)            |
| **CEA (ng/mL)**   |                     |
| ≤5.0              | 259 (74.2)          |
| >5.0              | 90 (25.8)           |
| **Tumor size (cm)**|                    |
| ≤3.0              | 240 (68.2)          |
| >3.0              | 112 (31.8)          |

Table 3 (continued)

| Variables | Lung cancers, N (%) |
|-----------|---------------------|
| **SUVmax**|                     |
| ≤4.0      | 134 (38.1)          |
| >4.0      | 152 (43.2)          |
| Unknown   | 66 (18.8)           |
| **EGFR status** |               |
| Wild      | 57 (16.2)           |
| Mutation  | 136 (38.6)          |
| Unknown   | 193 (54.8)          |
| **T stage**|                    |
| T1        | 152 (43.2)          |
| T2        | 159 (45.2)          |
| T3–4      | 41 (11.6)           |
| **N stage**|                    |
| N0        | 249 (70.7)          |
| N1        | 48 (13.6)           |
| N2        | 55 (15.6)           |
| **TNM stage**|                 |
| I         | 216 (61.4)          |
| II        | 50 (14.2)           |
| III–IV    | 86 (24.4)           |

CEA, carcinoembryonic antigen; EGFR, epidermal growth factor receptor.

Early stages of lung cancer as well (10). The antigen of PGP9.5 was a tumor suppressor gene that is inactivated by promoter methylation or gene deletion in several types of human cancers (34). In addition, PGP9.5 was previously identified as an antigen that induces an antibody response in lung cancer (35,36). We further analyzed the correlation between the expression of five autoantibodies against these TAAs in tumor tissues and the serum from lung cancer patients and found that the relationship was statistically significant. It was proved that autoantibodies against GAGE7, MAGEA1, PGP9.5, CAGE and p53 in serum could reflect the expression level of these antigens in primary tumors.

To discriminate lung cancer and healthy individuals, we firstly found that significantly higher level of GAGE7, MAGEA1, PGP9.5, CAGE and p53 in lung cancer patient than in individuals. Next, we could see high AUC value of 0.787, 0.641, 0.873, 0.825 and 0.749 for GAGE7, MAGEA1, PGP9.5, CAGE and p53 in ROC analysis, respectively. Previous studies have shown that single or combinations of multiplex autoantibodies can be used as a tool for the diagnosis of lung cancer patients at all stages or early-stage, and the combination of multiplex autoantibodies shows a higher detection capacity (37). Jia et al. also found multiply autoantibodies were more accurate to predict lung cancer than single autoantibody (38). With the encouraging results, we provided a hypothesis that whether a panel of these five autoantibodies could help...
Figure 3 Association between five tumor-associated autoantibodies expression level and clinical characteristics. (A,B,C,D,E,F,G,H)

Associated analyses of GAGE7, MAGEA1, PGP9.5, CAGE and p53 level in 325 with clinical features like age, gender, pathological type, T stage, N stage and TNM stage, SUVmax of primary tumor and CEA level, respectively. *, P<0.05 was considered significant difference. CEA, carcinoembryonic antigen. Ad, adenocarcinoma. SCC, squamous cell carcinoma.
diagnose lung cancer. ROC analysis indicated the accuracy of diagnosis was up to exciting 89.1% with sensitivity 90.1% and specificity 71.7% when combining the five antibodies. Compared to the CEA with 25.8% sensitivity in all 352 lung cancer patients of current study, five autoantibodies are suggested to be favorable biomarkers for detection of lung cancer. Maybe the five antibodies can be used to monitor the occurrence of lung cancer at an early stage by using a test kit to be developed.

Murray et al. proved acceptable concentration level of autoantibodies including p53, SOX2, NY-ESO-1, CAGE, GBU4-5, Annexin1 in serum that could identify 40% of primary lung cancer (9). Chapman et al. used ELISA assay to display significance of autoantibodies such as p53, c-Myc, HER2, NY-ESO-1, CAGE, MUC1 and GBU4-5 in diagnosis and prognosis for lung cancer patients (10). Currently, we selected higher levels tumor-associated autoantibodies of GAGE7, MAGEA1, PGP9.5, CAGE and p53 in lung cancer tissues from TCGA database. We also discovered the same autoantibodies of p53 and CAGE with the above reports in lung cancer. Furthermore, we clarify correlation between the expression of the autoantibody levels in serum and clinical characteristics for the 352 lung cancer patients. It was found that there was higher expression of each single autoantibody in CEA positive, SUV$_{\text{max}}$ >4.0, advanced T stage and N stage group compared to CEA negative, SUV$_{\text{max}}$ ≤4.0, early T stage and N stage, although the differences were not all significant. Especially, the PGP9.5 level was positively correlated with T stage, N stage and TNM stage with significant differences. Additionally, MAGEA1 and p53 highly expressed in the patients with pathological type of squamous cell carcinoma and increased level of GAGE7 and CAGE were observed in the high level of SUV$_{\text{max}}$ for lung cancer patients with PET-CT examination. All in all, the autoantibody may reflect the prognosis of lung cancer and it is necessary to design further study.

**Conclusions**

In sum, our data demonstrate that the serum antibody levels of GAGE7, MAGEA1, PGP9.5, CAGE and p53 were apparently higher in lung cancer patients than normal individuals and in view of that they could be used as early diagnosing lung cancer with satisfactory sensitivity and accuracy. Therefore, detection kits for early diagnosis of lung cancer may be developed in the future. In addition, the serum levels of autoantibodies were correlated with the stage of lung cancer and the prognosis value would be evaluated in further study.

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**Footnote**

**Conflicts of Interest:** All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/tcr.2019.07.25). The authors have no conflicts of interest to declare.

**Ethical Statement:** The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. This study has been carried out in accordance with the Declaration of Helsinki (as revised in 2013) and has been approved with the Ethics Committee of Peking University Cancer Hospital and the number of ethics approval is 2015KT71. The participants signed informed consent prior to sample collection.

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