Detection of human case of dengue virus 2 belonging to sylvatic genotype during routine surveillance of fever in Senegal, Kolda 2021

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Dengue virus 2 (DENV-2) was detected in a febrile patient living in Saré Yoba in the Kolda region of southern Senegal. Phylogenetic analysis based on the full coding region revealed that the virus belongs to the DENV-2 sylvatic genotype and is closely related to a strain (JF260983/99.66% identity) detected in Spain in a tourist who traveled to Guinea-Bissau (which borders the Kolda region) in 2009. This highlights a potential recent under-reported circulation of sylvatic dengue in the southern part of Senegal and calls for reinforced integrated surveillance among humans, non-human primates, and arboreal mosquitoes through a one-health approach.

KEYWORDS
fever, DENV 2, sylvatic, Senegal, one health

Background

Dengue is the most prevalent arboviral disease in tropical and subtropical areas. Dengue is caused by the dengue virus (DENV) and the etiological agents exist in four antigenically and phylogenetically distinct serotypes (DENV 1–4) (1). Infection with any DENV serotypes causes diseases ranging from flu-like illness (i.e., dengue fever) to a life-threatening disease known as severe dengue (2). The WHO estimates that one-third of the world's population is at risk of dengue infection (2). Each of the existing dengue...
serotypes is maintained in two different ecologically and evolutionary distinct transmission cycles, namely the human cycle and the sylvatic cycle. The human cycle, in which only humans act as a reservoir, is maintained between domestic and peridomestic mosquitoes; in contrast, the sylvatic cycle involves non-human primates and arboreal mosquitoes (3). Despite the central and basal role that sylvatic strains of DENV play in evolution and emergence, there is no report of continuous and sustained transmission (4).

In Senegal, mainly in the south of the country (i.e., the Kédougou area), the landscape of DENV circulation was long dominated by the occurrence and maintenance of sylvatic cycles (5). In 2009, a shift occurred, with the first reported urban DENV epidemic in Dakar, and this was followed by the recurrent and yearly multifocal and multiserotype circulation of human cycle strains (6). Here, we report a case of dengue virus 2 (DENV-2) infection. A phylogenetic analysis, based on full coding region, revealed that the DENV-2 infection was closely related to a strain circulating in Guinea-Bissau in 2009.

The study

In collaboration with the Ministry of Health and Social Action (Dakar, Senegal) and the Unit of Epidemiology Clinical Research and Data Sciences at the Institut Pasteur de Dakar (IPD) (Dakar, Senegal), our laboratory (i.e., the Virology Unit at IPD) is conducting syndromic surveillance of fever around the country through a program named the 4S (Syndromic Sentinel IPD) is conducting syndromic surveillance of fever around the country through a program named the 4S (Syndromic Sentinel IPD). As part of this nationwide surveillance project, samples from febrile patients are collected and shipped on a weekly basis to the WHO collaborating center for suspected arbovirus infection diagnosis. In November 2021, a patient suspected of arbovirus infection presented in the Sare Yoba health district, located in the Kolda region (southern Senegal). The patient was male, aged 28 years, and presented with symptoms that included headaches, myalgia, asthenia, arthralgia, and chills. A malaria rapid diagnostic test yielded a negative result. Following 2 days of fever, a venous blood sample was collected from the patient and shipped to the Virology Unit at IPD for diagnosis. At IPD, the blood sample was centrifuged at 2000 rpm for 5 minutes, and serum was harvested and aliquoted into 2-ml tubes. RNA extraction was performed using the QIAGEN viral RNA kit (QIAGEN, Hilden, Germany), using 140 µl of input serum, in accordance with the manufacturer’s recommendations. Extracted nucleic acid was subjected to screening for seven arboviruses, as previously recommended by Dieng and colleagues (8), of which only DENV gave a positive result. To define the incriminating serotype, DENV-positive RNA was subjected to a multiplex quantitative reverse transcription PCR assay using a TIB Molbiol Modular Dx Dengue typing kit (cat. no. 40-0700-24; TIB Molbiol, Berlin, Germany) (5). Surprisingly, the multiplex quantitative reverse transcription (qRT) PCR assay failed to define the dengue serotype. To define the virus serotype/genotype, we successfully amplified a partial NS5 gene sequence using FU1/FD3 (9), and the obtained amplicon was approximately 1 kb. The amplicon was purified using AMPure (Beckman Coulter Inc., Brea, CA, USA) beads at a ratio of 1:0.8. A sequencing library was prepared for the Oxford Nanopore MinION (Oxford Nanopore Technologies plc, Oxford, UK) using the rapid barcoding kit (SQK RBQ110.96), loaded onto a R9 flow cell and sequenced using a MinION MK1C device.

Raw data were collected and basecalled using guppy (https://community.nanoporetech.com). Adapters were trimmed using NanoFilt (10) (options -headcrop 50 and -tailcrop 50) and reads were mapped to a DENV reference genome (NC_001474.2) using Minimap2 (11). A National Center for Biotechnology Information (NCBI) BLASTn search of the obtained sequence resulted in no hits above the 70% identity threshold.

Based on results from the NCBI BLASTn (12) search, we downloaded full genome sequences of closely related sylvatic DENV-2 sequences and designed a tiling PCR primal scheme, generating amplicons of around 900 bp and covering the coding region of sylvatic DENV-2 strains. PCR amplification was performed using Q5® High-Fidelity 2X Master Mix (New England Biolabs, Ipswich, MA, USA) in accordance with a protocol previously described by Dieng and colleagues (13). The sequencing and data analysis were the same as previously used for NS5 gene sequencing.

To determine the evolutionary history, we download representative sequences of described DENV-2 genotypes. The obtained dataset was aligned using MAFFT (14) and a maximum likelihood (ML) tree was constructed using IQ-TREE (15).

The obtained ML (Figure 1) tree clearly shows that, based on a nearly full genome sequence, our strain fell within the West African DENV-2 sylvatic genotype and is closely related to a strain linked to hemorrhagic DENV detected in Spain from a tourist who traveled to Guinea-Bissau through Senegal in 2009 (16), and not to strains of the DENV-2 cosmopolitan genotype, which was responsible for the latest DENV-2 epidemic in Senegal (6, 13). This is the first identification of circulating sylvatic DENV-2 in Senegal since 2000 (17). Interestingly, the Kolda region borders the Niokolo-Koba National Park, which is home to monkey species (Papio papio, Erythrocebus patas) known to be a reservoir of sylvatic DENV-2 (3, 18). In addition, experimental findings from the surrogate human models of infection and from cultured cells suggest that there is little or no adaptive barrier for the emergence of sylvatic DENV in human populations, possibly reflecting the evolution of DENV as an opportunistic virus that is capable of infecting a wide range of primate species (3).

This finding, in addition to the present detection of this sylvatic genotype in the Kolda area (which shares a border with Guinea-Bissau), highlights a potential unnoticed circulation of...
sylvatic DENV-2 in the south of Senegal, and this is corroborated by the fact that the patient was not traveling, confirming that the case was autochthonous.

In 2021, a national seroprevalence study conducted in 14 administrative regions in Senegal led to the detection of five DENV immunoglobulin M-positive samples in the Kolda region (unpublished data). No samples have been reported to be qRT PCR positive prior to this case.

Interestingly, clinical infection with sylvatic strains is indistinguishable from human transmission cycle strains, and this can lead to under-reporting of sylvatic DENV (16) because genomic surveillance of the circulating DENV strain in Senegal, as well as in Africa, is limited (19). All parameters mentioned above support the high likelihood of spillover of sylvatic DENV from Africa or Asia in the human transmission cycle. In addition, many studies report that people with African ancestry confer some level of protection against severe dengue infection (20), and this, in addition to increased tourism in Africa (21), raises concerns about the risk of increased sylvatic dengue infection in Africa among non-indigenous people in near future.

This calls for determining the genetic diversity of circulating DENV strains, which is crucial before any vaccination policy can be implemented. Indeed, determining which contemporary genotypes are in circulation in a given area is crucial for ensuring effective diagnostics and for developing preventive and therapeutic countermeasures.

**Conclusion**

DENV is now hyperendemic in Senegal, with the co-circulation of DENV 1–3 belonging to human transmission cycles, and it is marked by yearly epidemics that may constrain the identification of sylvatic DENV. Nevertheless, as human DENV has the potential to enter human cells and cause hemorrhagic disease, an integrated one-health approach between humans, mosquitoes, and non-human primates is urgently needed in regions, other than the Kedougou area, located in the south of Senegal. A one-health approach could improve dengue fever surveillance through active, existing human malaria-like illness surveillance within the 4S Network.
Finally, to be able the discriminate between sylvatic and epidemic DENV strains, real-time genomic surveillance of DENV could play a key role in virus surveillance around the country. This real-time genomic surveillance will help us to better understand the evolutionary history, transmission, and spread, with complex transmission dynamics involving both urban and sylvatic DENV cycles.

Data availability statement

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

Ethics statement

In this study, we used samples collected as part of approved ongoing surveillance conducted by the IPD (i.e., a WHO collaborating center for arboviruses and hemorrhagic fever reference and research). The Senegalese National Ethical Committee approved the protocol as a less than minimal risk research, and written consent forms were not required. All samples from humans were deidentified before we performed virus detection, characterization, and analysis.

Author contributions

Conceptualization: OusF, AS, BD, MB, CL, CT, and OumF; methodology: ID and MN; software: ID; original draft: ID; final draft and review: all authors. All authors contributed to the article and approved the submitted version.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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