Supplemental information

Head-to-tail polymerization by VEL proteins underpins cold-induced Polycomb silencing in flowering control

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Supplementary Fig. 1 (related to Fig. 1)

**VEL-dependent formation of nuclear condensates**

(A-E) Representative confocal images of COS-7 (A-C) or HeLa (D-E) cells transfected with wt or mutant GFP-VIN3, GFP-VEL1 or GFP-VRN5, as indicated in panels; scale bars 10 µm. (F-H) Representative confocal images of *N. benthamiana* leaves infiltrated with wt or mutant GFP-VIN3, as indicated in panels; scale bars 5 µm. (I) Western blot showing comparable levels of wt and mutant GFP-VIN3 proteins; arrow, internal control (large subunit of Rubisco, visualized by Ponceau staining).
Supplementary Fig. 2 (related to Fig. 2)

**Deep conservation of VEL domains in plants**

(A) Sequence alignment of VEL domains from diverse plant species, as indicated (At, Arabidopsis thaliana; MYA, million years ago); point mutations disabling self-association of VIN3 are indicated above. (B-D) SEC-MALS of purified Lip-tagged VEL domains from various species as indicated in inset; curves, elution profiles; line traces, molar masses as derived from MALS.
Supplementary Fig. 3 (related to Fig. 4)

**Conserved VEL folds and confirmation of self-interacting surface by NMR**

Overlays of ribbon diagrams of different VEL domains; black balls, N-termini; (A) VIN3VEL RR>AD (7O6U, light blue) and VIN3VEL R556D I575D (7O6T, green); RMSD 1.69 Å; (B) VIN3VEL RR>AD (7O6U, light blue) and VEL1VEL I664D (7O6W, blue), RMSD 1.63 Å; (C) VIN3VEL R556D I575D (7O6T, green) and VEL1VEL I664D (7O6W, blue), RMSD 1.77 Å; (D) VEL1VEL I664D (7O6W, blue) and VEL1VEL RK>AD I664D (7O6V, gray), RMSD 0.41 Å. (E) 15N-labeled RR>AD probed with unlabeled I575D; top panel, relative peak heights from HSQC of unbound (maroon) versus HSQC of bound (blue); bottom panel, differences in percent. (F) Percent differences in (E) plotted onto the surface of the RR>AD NMR structural model, confirming that the tail surface as defined by crystallography also serves as the polymerisation interface in solution. (G) ITC profiles (top panel) of VIN3VEL RR>AD binding to VIN3VEL I575D. Fitted data (bottom panel) show a Kd value of 1.16 µM for the dimeric interface.
Abnormal filaments of double-mutant VEL domains

Abnormal head-to-tail interactions in (A) VEL1VEL (R643A K645D I664D; 7O6V) or (B) VIN3VEL (R556D I575D; 7O6T) crystals bearing point mutations in both head and tail surfaces; magenta, R643A K645D (A), or R556D (B); red, I664D (A), or I575D (B).
Supplementary Fig. 5 (related to Fig. 6)

**NMR spectroscopy of VIN3 \_VEL RR>AD**

(A) VIN3 \_VEL RR>AD sequence, with four \(\alpha\)-helices above (magenta, R554A and R556D); RCI-S\(^2\) showing the propensity of each residue (green dots) to be within a turn (<0.85) or an \(\alpha\)-helix (>0.85); bottom, RDC plot further confirming ‘H4 tucked under’ conformation (see main text).

(B) Full assignments of VIN3 \_VEL 529-603, with RR>AD residues marked (magenta); spectrum acquired at 300 \(\mu\)M.
Supplementary Fig. 6 (related to Fig. 6)

VEL-related 4HB domains in DNAJ co-chaperones

(A) Table of top hits from DALI searches for VIN3VEL (left) and VEL1VEL (right), with 4HB from Zuotin (orange) and DAXX (gray) highlighted.

(B, C) Overlays of (B) VIN3VEL RR>AD (wheat) with 4HB domain of DAXX (5Y6O; gray), RMSD 3.24 Å, (C) VEL1VEL (blue) with 4HB domain of Zuotin (6CGH; orange), RMSD 2.69 Å; black balls, N-termini.

(D) SEC-MALS of human DAXX (gray trace) and human ZRF1 (orange trace), both corresponding to monomers; void volume of column at 8 ml.

|   | VIN3_NMR                | Z     | VEL1_crystal | Z     |
|---|-------------------------|-------|--------------|-------|
| 1 | 40MG - RIBOSOME-BINDING DOMAIN OF ZU01 | 6.1 | 6HPN - ANTIGEN, P35 | 6.4 |
| 2 | 6HPN - ANTIGEN, P35     | 6.0 | 6CGH - SOLUTION STRUCTURE OF THE FOUR-HELIX BUNDLE REGION OF HUMAN J-PROTEIN ZUOTIN | 5.3 |
| 3 | 5Y6O - CRYSTAL STRUCTURE OF DAXX N-TERMINAL FOUR-HELIX BUNDLE DOMAIN (4HB) IN COMPLEX WITH ATROX | 5.9 | 4GMQ - RIBOSOME-BINDING DOMAIN OF ZU01 | 5.0 |
| 4 | 3TDW - GENTAMICIN RESISTANCE PROTEIN | 5.2 | 5Y6O - CRYSTAL STRUCTURE OF DAXX N-TERMINAL FOUR-HELIX BUNDLE DOMAIN (4HB) IN COMPLEX WITH ATROX | 5.0 |
| 5 | 6CGH - SOLUTION STRUCTURE OF THE FOUR-HELIX BUNDLE REGION OF HUMAN J-PROTEIN ZUOTIN | 5.0 | 1OKS - RNA POLYMERASE ALPHA SUBUNIT | 4.8 |
| 6 | 2IPC - PREPROTEIN TRANSLOCASE SECA SUBUNIT | 4.9 | 3FW7 - PUTATIVE FRUCTOSAMINE-3-KINASE | 4.7 |
| 7 | 1OKS - RNA POLYMERASE ALPHA SUBUNIT | 4.7 | 1K30 - GLYCEROL-3-PHOSPHATE ACYLTRANSFERASE | 4.7 |
| 8 | 3FW7 - PUTATIVE FRUCTOSAMINE-3-KINASE | 4.7 | 6N9Y - NON-STRUCTURAL PROTEIN 1 OF BLUE TONGUE VIRUS | 4.6 |
| 9 | 2LWX - SOLUTION STRUCTURE OF THE C-TERMINAL PDH1-ACTIVATING DOMAIN OF THE J-PROTEIN ZU01 | 4.8 | 3TDW - GENTAMICIN RESISTANCE PROTEIN | 4.6 |
| 10| 5LJX - PHOSPHOPROTEIN | 4.5 | 2IPC - PREPROTEIN TRANSLOCASE SECA SUBUNIT | 4.6 |
Supplementary Fig. 7 (related to Fig. 7)

**Nuclear localization of wt and RI>DD mutant VIN3-GFP**

(A) SEC-MALS of purified Lip-VIN3VEL RI>DD, at increasing concentrations as indicated in panels; *line traces*, molar masses as derived from MALS. Highest concentration is still monomeric (calculated 23.4 kDa, observed 24 kDa); void volume of column at 8 ml. (B-F) Confocal images of Arabidopsis root tips expressing wt VIN3-GFP (homozygous) or polymerization-deficient RI>DD mutant (independent first generation transgenic lines) at 6WT0. Scale bar 50 μm. Transgene copy numbers in individual RI>DD lines are 3 (#31), 11 (#44) and 3 (#61). Note that all images were taken with the same microscopy settings and on the same day from the same individual plants as those shown in main Fig. 7 whose flowering is delayed compared to wt VIN3-GFP, revealing comparable levels of fluorescence in the nuclei of these plants.
| PDB accession codes | VE1_VEL | VIN3_VEL |
|---------------------|---------|----------|
| I664D               | 7O6W    | 7O6V     |
| R643A K645D I664D   |         | 7O0Q     |
| I575D               | 7O6U    | 7O6T     |

**Crystal data**

- **Wavelength (Å)**: 0.97950 (0.97950)
- **Resolution (Å)**: 27.47 – 2.64 (2.77 – 2.64)
- **Space group**: P 63, P 2 1 2, I 4 1, P 2 1 2
- **Unit cell dimensions**:
  - a, b, c (Å): 79.26, 79.26, 79.26, 59.96, 59.96, 52.07, 58.05, 52.07, 119.99, 119.99, 119.99
  - α, β, γ (°): 90, 90, 120, 90, 90, 90

**Total reflections**

- 239198 (22200)
- 338001 (38598)
- 101791 (10799)
- 82212 (4592)
- 349471 (22221)

**Unique reflections**

- 6335 (769)
- 13183 (1481)
- 11740 (1191)
- 5941 (354)
- 9677 (677)

**Refinement**

- **Resolution**: 27.47 – 2.64, 29.07 – 2.4, 28.82 – 1.84, 29.47 – 2.02
- **Number of reflections**: 6281, 13134, 11698, 5913, 9609
- **R-work / R-free (%)**: 21.16 – 24.92, 21.36 – 24.07, 22.34 – 26.95, 22.13 – 24.44, 20.24 – 23.66
- **Nº of atoms**: 1151, 2308, 2357, 604, 1623
- **Average B Factors (Å²)**:
  - Wilson/overall: 80.42, 53.72, 45.12, 38.71, 41.25
  - Protein: 66.86, 45.62, 60.33, 33.78, 34.11
  - Ligand: 161.50, - , - , - , 52.82
  - Water molecules: 79.40, 53.22, 52.33, 48.46, 50.39
  - All atoms: 67.33, 45.65, 60.20, 34.17, 34.78

**RMSDs deviations**

- Bonds lengths (Å): 0.006, 0.008, 0.007, 0.010, 0.009
- Bond angles (°): 1.01, 1.16, 1.53, 1.50, 1.22

**Ramachandran plot Statistics (%)**

- Favored regions: 97.76, 100, 98.89, 100, 100
- Allowed regions: 2.24, 0, 1.11, 0, 0
- Disallowed regions: 0, 0, 0, 0, 0

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**Table S1**

Crystal data collection and refinement statistics