Effects of marking methods and fluorescent dusts on Aedes aegypti survival

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Abstract

Background: Tracking the movement of mosquitoes and understanding dispersal dynamics is essential for the control and prevention of vector-borne diseases. A variety of marking techniques have been used, including dusts and dyes.

Methods: In this study, Aedes aegypti were marked using fluorescent dusts ('DayGlo' A-19 Horizon Blue & A-13-N Rocket Red; 'Brian Clegg': pink, blue & red), fluorescent paints ('Brian Clegg': blue, red & yellow) and metallic gold dust ('Brian Clegg'). Dusting methods were those previously used in mark-release-recapture experiments, including application with a bulb duster, creation of a dust storm or shaking in a bag.

Results: Results showed marking mosquitoes using a dust storm allowed relatively high survival, compared to unmarked controls (Males: \(\chi^2 = 3.24, df = 4, p = 0.07\); Females: \(\chi^2 = 3.24, df = 4, p = 0.04\)), and high marking efficiency. Using a bulb duster showed high survival in male mosquitoes (\(\chi^2 = 12.59, df = 4, p < 0.000\)), but low survival in female mosquitoes during the first 15 days of the study (\(\chi^2 = 5.17, df = 4, p < 0.05\)). The bulb duster also had the lowest marking efficiency compared to other dry marking techniques. The bag method showed low survival in males during the first 15 days of the study (\(\chi^2 = 5.77, df = 4, p < 0.05\)). Applying paints had an overall negative impact on survival for males (\(\chi^2 = 5.03, df = 3, p < 0.05\)), but not for females (\(\chi^2 = 0.19, df = 3, p = 0.661\)). Males dusted with DayGlo Horizon Blue dust, and females dusted with DayGlo Rocket Red dust, had the most significant reduction in survivorship in comparison to the control (Males: \(\chi^2 = 15.70, df = 6, p < 0.000\); Females: \(\chi^2 = 24.47, df = 6, p < 0.000\)). Mosquitoes marked with Brian Clegg gold dust showed mortality rates similar to controls within male mosquitoes (\(\chi^2 = 0.18, df = 6, p = 0.674\)), but significantly lower in females (\(\chi^2 = 16.59, df = 6, p < 0.000\)).

Conclusions: This study showed that marking technique and colour can have a significant impact on the survival and marking coverage of a mosquito.

Keywords: Aedes aegypti, Fluorescent dust, Fluorescent paint, Survivorship

Background

Dispersal is an essential component of understanding insect biology, behaviour, life history and population dynamics [1]. Understanding dispersal dynamics and flight ranges of mosquito vectors is essential for the mitigation of disease, successful implementation of protection against infection and an important step in understanding the ecology of a vector [2]. Mark-release-recapture (MRR) techniques can be used to estimate mosquito population densities, feeding behaviour, duration of gonotrophic cycles and their dispersal behaviour [2]. An ideal marking technique should be cost-effective, easily applied, visible, non-toxic and should not affect the behaviour, development, longevity or reproduction of mosquitoes [1,3]. Marking techniques have the potential to affect the mortality and dispersal rates of marked individuals in a way that could bias the results arising from MRR experiments, so preliminary experiments on marking methodologies are required prior to carrying out MRR experiments [2].

A variety of methods have been used to mark mosquitoes, including dusts [4-9], dyes [10,11], paints [12,13], trace elements [14] and radioactive isotopes [15,16] (see reviews in [1-3]). One of the most common methods of marking mosquitoes externally is to apply micronized particles of dust (also known as powder or pigments), particularly fluorescent dust, to a large number of mosquitoes [1].
Dusts are cost-effective, available in a range of colours, easily applied and very detectable.

Several types of fluorescent dusts have been used to mark insects, including ‘Helecon’ and Radiant dusts, but the ‘DayGlo’ series A and AX are now the most commonly used as no adhesive is needed for mosquitoes to retain marks [2]. Dusts can be applied using a syringe [7] or bulb duster [17], putting them in a bag containing dust and shaking them gently [18], or by creating a dust storm within a cage [19]. Many shaking methods can cause high mortality by applying too much dust to mosquitoes. This can increase mortality, decrease mobility and affect the sensory organs [1], giving biased results in MRR studies. There is also the constraint of dusts not persisting long enough for long-term studies and transference of dusts to unmarked individuals [1,20].

Paints can be applied individually [12] or to large groups of individuals [11,13]. Applying paints individually can be time-consuming, but mass marking by spraying paints can be easy and quick. Usually paints are diluted with acetone or alcohol before being sprayed from hand atomizers or spray guns [1]. Myles and Grace [21] experimented with spray paints as an adhesive for borate dusts on termites, claiming they were non-toxic. Fluorescent paints adhere to body parts, can achieve 100% coverage and are readily identifiable under UV light [22]. Marking with paints is usually believed to cause little mortality, but it is likely that applying paint spots to mosquito wings affects behaviour and possibly survival [2]. Droplet size and visibility needs to be balanced in order to ensure that the mosquito is unaffected and able to be seen.

The effects of marking adults with powders or stains, or any other substance, should always be carefully evaluated by comparing mortalities of marked and unmarked mosquitoes of the same species, sex and if possible same age over the lifetime of the insect. Unfortunately it appears from the literature that in many experiments where marking was carried out, the effects of the marking were not evaluated statistically, or inadequately so [2]. In order to improve marking efficiency for future dispersal and population studies, methods should be rigorously compared. This study was conducted to determine the best method and colour for marking mosquitoes for MRR experiments by comparing the mortalities of marked and unmarked individuals, whether immobilising the mosquitoes had adverse effects on survival and calculating how efficient each method is in marking.

Methods
Mosquitoes
Aedes aegypti (Linnaeus) were used for all laboratory experiments. For the rearing of mosquitoes, second instar larvae were placed in plastic trays and fed daily with fish food (TetraMinBaby®). Pupae were transferred to a cup of water in an insect rearing cage. Larvae and adults were maintained under insectary conditions (27°C, 70% RH and a photoperiod of 12:12 [L:D] h) and provided with 10% sucrose solution.

Marking of mosquitoes
Two or three day old Aedes aegypti mosquitoes were randomly aspirated into 90 mm plastic containers (1 L tumblers) with gauze tops, until there were 15–18 females and 15–18 males in each container. After applying the marking technique, all experimental containers had cotton wool soaked in 10% sucrose solution placed on top of the gauze, which was refreshed daily. A plastic lid was placed over the top of the container to keep the humidity high. Dead mosquitoes were recorded daily and removed.

Mosquitoes were immobilised prior to the marking methods to allow a better coverage, and to prevent accidental release. To check that survival was not affected by immobilising them, an experiment was set up to compare briefly immobilised mosquitoes to control mosquitoes. Two containers were placed in a freezer (−18°C) for one minute, and then changed over to a container held at room temperature (22°C) until the mosquitoes had recovered. Two containers of immobilised mosquitoes and two containers of control mosquitoes were then placed back in the insect rearing room (27°C) until all mosquitoes died.

Dusts from two companies were used in this experiment; A-19 Horizon Blue and A-13-N Rocket Red ‘DayGlo’ series A fluorescent pigments (DayGlo Color Corp, Cleveland, OH, USA), which will be referred to as ‘D Blue’ and ‘D Red’, gold metallic dust (Brian Clegg, UK), pink, blue and red fluorescent dusts (Brian Clegg, UK), which will be referred to as ‘BC Gold’, ‘BC Pink’, ‘BC Blue’ and ‘BC Red’. Yellow, blue and red fluorescent paints (Brian Clegg, UK) were also used in this experiment. DayGlo dusts are manufactured from organic dyes, incorporated into a melamine formaldehyde resin and grounded into a fine powder [2]. Brian Clegg dusts are composed of calcium/magnesium carbonate, for use as powder paints.

The marking methods used in this experiment were; placing mosquitoes in a bag with dust at the bottom and gently shaking (hereafter known as ‘bag’ method), using a bulb duster to create a small puff of dust (hereafter known as ‘bulb duster’ method), using a fan to create a small dust storm within a cage (hereafter known as ‘dust storm’ method) and lightly spraying paint in small droplets (hereafter known as ‘paint’ method). The paint solution was made by mixing 2 g dust, 200 ml paint of the same colour and 200 ml distilled water. This was repeated for each paint colour. The paint control was made up of 200 ml distilled water. After the mosquitoes had been immobilised, they were transferred to a tray and the paint solution was finely sprayed, using an atomiser, three times
over the mosquitoes. Mosquitoes were then transferred to a container to recover, and placed in the insect rearing room. All DayGlo and Brian Clegg dusts were used for the bulb duster, bag and dust storm method. The bulb duster was loaded with 0.3 g dust per container; mosquitoes were transferred to a tray and sprayed four times. The bag method had 0.3 g dust placed in the bottom of the bag; immobilised mosquitoes were placed in the bag and gently shaken in the bag. Mosquitoes for the dust storm method were also placed in a bag with 0.3 g dust, a fan was used to create a dust storm within the bag. After all dusting methods, immobilised mosquitoes were gently placed back in their original container. Each method had a control, where mosquitoes were handled similarly to marked mosquitoes, but dusts or paints were not added. Each method, control and colour had three repeats, each containing at least 30 mosquitoes. The survival experiment continued until all mosquitoes died.

‘Marking efficiency’ scores were given to each colour and method used. Scores were calculated by whether dust could be seen from 20 cm away, and if dust was present on their head, thorax, abdomen, legs or wings when observed using a dissection microscope. A score of ‘1’ was given if present, or ‘0’ if not present, for each category. The sum of these categories gave the overall marking efficiency score, out of a maximum of six.

Data analysis
The survival data was analysed using the log rank test in SPSS Version 20.0 [23] across 15, 30 and 61 days. Using different time scales allowed comparison with other studies which have examined survival over short time periods and consequently may have observed different trends. Across all experimental treatments there was a clear difference between male (N = 1260), and female (N = 1235) survival ($\chi^2 = 1080.80$, df = 1, $p < 0.0001$), therefore analysis was performed separately for each sex. For comparing methods; bag, bulb duster, dust storm and paint methods were compared to unmarked control mosquitoes. For comparing colours, D Blue, D Red, BC Blue, BC Pink, BC Red and BC Gold were compared to unmarked method controls. Yellow, blue and red paints were analysed separately to the paint controls for comparison of colours.

A generalised linear model (GLM) with quasi-poisson errors was used to test marking efficiency in relation to marking method and colour. A GLM with poisson errors showed that the data was over-dispersed. Interactions were tested for, but were not present. Mosquitoes that died early (days 1–20) and late (days 21–61) within the experiment were compared separately for marking efficiency. Males and females were combined within the model because their marking efficiencies were not significantly different (df = 1, $p = 0.897$). All models were plotted to see how well the model fitted the data. All graphs were plotted using R version 2.15.3 [24].

Results
The experiment ran for 61 days, at which point all mosquitoes had died. Median longevity was 12 and 28 days for males and females, respectively. Overall, 716 (58%) of females survived beyond 30 days, but only 21 (2%) male individuals survived beyond 30 days. There were no significant differences in longevity between immobilised mosquitoes and their controls for males ($\chi^2 = 0.82$, df = 1, $p = 0.364$) or females ($\chi^2 = 2.21$, df = 1, $p = 0.137$).

The bag method for males showed a dramatic decline in survivorship during the first 15 days of the experiment (Figure 1a), and was the only method that was significantly different to the control (Table 1). Bulb duster, dust storm and paint methods had significantly higher longevity in males than the control, but the bag did not (Table 1). Contrary to males, females showed a dramatic decline in survivorship in the first 15 days of the experiment for the bulb duster method (Figure 1b), which was the only method significantly different to the control (Table 1). Pairwise comparisons of treatments showed that marking method was the dominant determinant of survival in comparison to other treatments (Figure 2).

With the exception of BC Gold, the survival and longevity of mosquitoes was significantly lower for all colours compared to the control in males (Figure 1c). In females, the survival of mosquitoes was significantly lower for all colours (Figure 1d). The same trend was observed for the 15 and 30 periods in males (Table 1). For females, the first 15 days did not show a significantly different survivorship to the control, but did for the 30 day periods (Table 1). DayGlo dusts showed a higher mortality rate than the Brian Clegg dusts, but both were significantly different to the control for males and females (Table 1). For the paint colours, significant differences were not observed in the first 15 days for males and the first 15 and 30 days for females (Table 1). Blue paint significantly decreased the longevity of female mosquitoes over 61 days, but red and yellow paint increased the longevity (Table 1). Red, blue and yellow paint increased the survivorship of males for 30 days (Table 1). The 61 and 30 day periods showed similar trends for all treatments for both males and females.

For marking efficiency, no statistical difference was observed between early (days 1–20) and late (days 21–61) survival, indicating that those individuals who showed signs of being marked in the beginning of the experiment remained so for the rest of the experiment (df = 1, $p < 0.0001$). All methods of application and colours were visible ($p \leq 0.002$ for all dusting combinations), but some more visible than others (Figure 3). The paint method was the least visible, followed by the bulb duster (Figure 3a).
D Blue, D Red, BC Red and BC Pink showed the greatest marking efficiency (Figure 3b). DayGlo dusts had a higher marking efficiency than Brian Clegg dusts.

**Discussion**

The effect of dusts on insect survival and behaviour varies with species [25] thus the amount of dust used and application technique is likely to have an effect. Although some studies have found no significant differences between the survival rates of marked insects to unmarked controls [25-39], others have documented adverse dusting effects [40-47], with factors including brand and colour often associated with dramatic differences between marked and unmarked controls.

This study found that males and female *A. aegypti* were adversely affected by the bag and the bulb duster methods. It is possible that the bag method was more damaging to males due to their fragility and that physical shaking damaged their extremities. Shaking procedures have shown to be detrimental to delicate insects because they place too much dust on the insects, and cause high mortality immediately after marking [1,48]. It is less certain as to why the bulb duster method gave greater mortality for females but it may also be due to an...
Table 1 Log-rank tests for *Aedes aegypti* marked using dusts, paints and different marking methods

| Method      | N  | df | Males: 15 days | Males: 30 days | Females: 15 days | Females: 30 days |
|-------------|----|----|----------------|----------------|------------------|------------------|
|             |    |    | $\chi^2$       | p-value        | $\chi^2$         | p-value          |
| Bag         | 326| 4  | 5.77           | 0.016*         | 0.31             | 0.577            |
| Bulb duster | 335|    | 1.66           | 0.197          | 10.65            | 0.001***         |
| Dust storm  | 338|    | 0.02           | 0.886          | 3.11             | 0.078            |
| Paint       | 195|    | 1.19           | 0.275          | 5.03             | 0.025*           |
| Colour (dust) |    |    |                |                |                  |                  |
| D Blue      | 144| 6  | 16.493         | 0.000***       | 15.70            | 0.000***         |
| D Red       | 144|    | 14.689         | 0.000 ***      | 8.72             | 0.003***         |
| BC Blue     | 139|    | 4.42           | 0.039*         | 0.05             | 0.830            |
| BC Pink     | 148|    | 9.824          | 0.002**        | 14.84            | 0.000***         |
| BC Red      | 142|    | 6.403          | 0.011*         | 0.49             | 0.484            |
| BC Gold     | 138|    | 3.343          | 0.068          | 0.03             | 0.857            |
| Colour (paint) |    |    |                |                |                  |                  |
| BC Blue     | 47 | 3  | 0.37           | 0.543          | 9.843            | 0.002***         |
| BC Red      | 51 |    | 0.393          | 0.531          | 2.793            | 0.095            |
| BC Yellow   | 48 |    | 1.942          | 0.163          | 5.575            | 0.018*           |
| Company     |    |    |                |                |                  |                  |
| DayGlo      | 288| 2  | 18.609         | 0.000***       | 14.74            | 0.000***         |
| Brian Clegg | 281|    | 6.77           | 0.005***       | 0.292            | 0.589            |

***indicates $p<0.001$, **p<0.01, *p<0.05, . p<0.1.

Figure 2 Pairwise treatment comparison of significance. Surface plot showing pairwise patterns of significance between different treatments for the first 15 days of **a**) male and **b**) female survival.
excess of dust that female mosquitoes were unable to groom off. Too much dust can decrease mobility, interfere with sensory organs and increase mortality [49]. Excessive moisture whilst marking can cause insects to become ‘gummed up’ with dust [26], but this was not an issue in this study as relative humidity was constant and there were no observations of any water droplet formation or gumming of dust.

The commonly used dust storm method had less impact on survival and thus is better than the other two dusting methods. This is perhaps due to dust storms atomising the dust better than the bag and bulb dusting methods. As for marking efficiency for each method, the dust storm and bag provided the greatest coverage of mosquitoes. Spraying mosquitoes with fluorescent paint had no marked effect on survival, possibly due to the small droplet use in the study, which was designed to balance survival and coverage. The paint method showed a low marking efficiency in comparison to the dry marking methods. This could be increased by extra sprays during marking, but too much moisture can affect behaviour and survival [2]. Whilst relative humidity prolongs mosquito survival, excess wetting mosquitoes can be a source of mortality [26], so this method must be used with caution.

Colour choice appears to be important for MRR studies. The reds and pinks used had intermediate values for both survival and marking efficiency. Although D Blue could be seen using the naked eye, the significant negative effect on survival makes it unsuitable for studies where an assumption is made that the marked individuals are of equal fitness to the unmarked. Other shades of blue which have a different chemical make-up and/or different concentrations of agents may not create this effect. This was observed in *Reticulitermes* which had a significant mortality rate over 15 days for Sudan Red 7B but not Neutral Red [50]. This may contribute to the success and failure of any MRR studies which use different shades of D Blue or other colours, although many other factors should be considered such as the scale, location, season, duration of the study and marking method.

The low impact of BC Blue on survival could be used as evidence to argue this but owing to its poor marking efficiency, it is possible that the dust was unable to adhere to individuals and was thereby spuriously associated with high survivorship. A colour change occurred in BC Gold tests with blue webbing across appendages at 15 days and it additionally had greater longevity for those marked with this colour for reasons unknown.

Mosquitoes marked with the two particular shades of DayGlo dust had reduced survival, but further testing with more colours, shades and chemical compositions is required to conclude whether one brand has lower mortality. Other studies have also observed reduced survival rates that are dependent on the colour of dust used. Coviella *et al.* [45] observed reduced survival in cicadellids, but only for one of the six colours of DayGlo dusts tested. One of 14 colours of powders tested by Toepfer *et al.* [51] reduced the survival in corn rootworm adults.
Two concerns in scaling the results from this laboratory study up to a semi-field or field trial are establishing a practical immobilisation method and preventing contact-transfer among marked and unmarked individuals. Firstly, using a freezer to immobilise mosquitoes in the field would prove difficult in certain situations, but diethyl ether is a suitable replacement. Even though freezing was chosen here over a chemical agent, provided the knock-out time is similar and the chemical has no lasting effects, it is likely to have no significant effect in the survival of the mosquito [2]. As it is well established that many species and strains vary in environmental resilience and excessive immobilising methods have the potential to cause significant mortality, immobilisation and marking technique should be tested in the laboratory and suitable concentrations established before field experiments. It should also be noted that marking under a laboratory setting without the uncontrolled weather and environmental conditions of a field study are likely to mean that marking efficiency will be high for a longer duration than can be expected in the field.

Secondly, contact-transfer can bias results when using methods which heavily rely on dust coverage, potentially marking 1-3% of the unmarked mosquitoes after 24 hours if exposed to DayGlo mosquitoes in a confined space [20]. Dust particles can potentially be transferred to unmarked insects in traps and sweep nets used for sampling [52], where individuals are forced to come into contact with each other, but is not an issue with sticky traps. It should be noted that same-sex transfer is likely to be of little concern and most transfer would occur during mating. Crumpacker et al. [26] observed no dust transfer during mating of Drosophila pseudoobscura or following the crowding of heavily marked and unmarked individuals after they were allowed to clean themselves before mixing. The implication of this for field studies is that they should mark individuals then give them time to clean themselves in relatively uncrowded conditions before releasing them.

We expect it is likely that the effects we observed here of dusting methods on survival of Ae. aegypti will be similar to the effects on Anopheles species, although other experiments have shown little difference in survival of marked and unmarked An. punctulatus, An. maculatus, An. sinensis, An. subpictus and An. stephensi [6,31,39,52-54]. The survival of a mosquito is dependant on many variables, including its size and resistance to environmental stress [55-57]. The effects of marking represents an additional stress on the body of the insect, making it necessary to carry out this experiment on other medically-important mosquito subfamilies, including the malaria vector species in Anopheles, where a range of body sizes and survival rates have been reported [2,58,59].

We found that application method had a significant effect on mosquito survival, and that the dust storm method caused the least mortality. Of the colours, the two shades of blue we tested should be avoided. For increased survival and marking efficiency, BC Pink or BC Red appear to be the most viable options. Care should be taken before using new colours (e.g. BC Gold) and when assuming there are no significant effects on mosquito survival with a chosen method, as our results suggest they may strongly bias the results of a MRR study. Choosing the best technique for use in the field will be essential to the success of field-based studies in mosquito dispersal.

Conclusions
Few studies have addressed the implications of marking efficiency and survivorship on male and female mosquitoes following marking, and even fewer have compared marking methods. Aedes aegypti is the primary vector of dengue, and its spatial movement is of interest to many policymakers and modellers. With no specific treatment, and the increased global incidence of dengue, it is becoming paramount to understand the vector's dispersal through studies such as MRR. Successful MRR studies require a benign technique that adheres to the mosquito for a defined duration. This study showed that treatments not only affected males and females differently across 15, 30 and 61 days, but also particular colours and methods were significantly different to controls. Males dusted with D Blue and females with D Red had the most significant reduction in survivorship in comparison to the control. Dusting using BC Red or BC Pink showed both reasonable performance in marking and impact on overall survival across males and females. Overall, the dust storm method provided the best trade-off between survival and marking efficiency.

Abbreviations
MRR: Mark-release-recapture study; D (colour): DayGlo colour; BC (colour): Brian clegg colour.

Competing interests
The authors declare that they have no competing interests.

Authors' contributions
Both authors participated in the design of the study, performed the statistical analyses, read and approved the final manuscript.

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