Characterization of pharmacological properties of methanolic seed and stem bark extracts of *Ziziphus mauritiana* (BAU Kul) using in-vitro and in-vivo animal (*Swiss albino* male mice) model

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Abstract

**Background:** *Ziziphus mauritiana* belongs to family of Rhamnaceae, is an improved fast-growing variety of plum and seasonal fruit which is very popular and commonly known as "BAU Kul" in Bangladesh. This study was designed to evaluate the medicinal values of seed and stem bark extracts of *Ziziphus mauritiana*.

**Methods:** Antioxidant, thrombolytic, membrane stabilizing, cytotoxic and antimicrobial activities were characterized using in-vitro methods and anti-diarrheal, analgesic and hypoglycemic activities were evaluated in *Swiss albino* male mice. The coarse powder of the plant parts were extracted with methanol and methanol portion of the extract was evaporated naturally under room temperature. Then the dried plant extracts were undergone for subsequent investigation.

**Results:** Total phenolic and flavonoid content, and antioxidant capacity of stem bark extract were higher than seed extract. Both extracts had similar DPPH (IC$_{50}$ of seed 4.53 μg/ml and stem bark 4.13 μg/ml) and NO (IC$_{50}$ of seed 5.92 μg/ml and stem bark 5.47 μg/ml) free radicals scavenging potential and seed extract was most cytotoxic (LC$_{50}$ = 1.467 μg/ml). Stem bark extract showed higher thrombolytic and anti-inflammatory potential, and also exhibited 14 mm zone of inhibition against *S. aureus*. Stem bark extract showed highest anti-diarrheal (75.68% defecation inhibition, *p* < 0.001), analgesic (68.63% writhing inhibition, *p* < 0.001) and hypoglycemic activity (44.27% blood glucose reduction after 3 h, *p* < 0.001) at 400 mg/kg body weight dose.

**Conclusion:** This study confirms good medicinal properties of selected plant parts. Further studies on isolation and characterization of its bioactive compounds are highly required.

**Keywords:** *Z. mauritiana*, Antioxidant, Anti-microbial, Thrombolyis, Anti-inflammatory, Cytotoxicity, Anti-diarrheal, Anti-diabetic
Introduction

The biological resource of nature such as plants, animals and microorganisms can be used for social, environmental and therapeutic purposes. Wide ranges of pharmacological activities are recorded for many medicinal plants globally. Medicinal plants are the nature’s gift to human beings to help them pursue a disease-free healthy life [1]. “BAU Kul” is known as *Ziziphus mauritiana* and belongs to family of Rhamnaceae. “BAU Kul” is a newly improved fast-growing variety of plum and sweet seasonal fruit which is very popular to the consumers of Bangladesh. The name “BAU” indicates its hybridization source, Bangladesh Agricultural University (BAU) which derived from cross-breeding local fruit from Bangladesh with jujube fruits from the Philippines, Somalia, India, Taiwan and China. The average weight of fruit is 90 g and seeds are small in size [2, 3]. The seed and stem bark parts of *Ziziphus mauritiana* were included in our present study because there are very limited reports on the selected plant parts.

Methods

**Ethnomedicinal survey of Ziziphus mauritiana**

Local folk medicine practitioners were included to know the traditional usage pattern of *Ziziphus mauritiana* either single or combination with other plant parts in different ailments. Both male and female kavirajes of Gaibandha district of Bangladesh were included in this survey. Written Informed consent was obtained and those who refused to provide information were excluded from this study. A total of 10 kavirajes (folk medicine practitioner) were interviewed and information was recorded. All the questions were asked in native language (Bengali) for their easy understanding.

**Collection and identification of the plant sample**

The seeds and stem barks part of *Ziziphus mauritiana* was collected from the BAU Kul garden of Gaibandha district of Bangladesh and identified by a taxonomist of Department of Botany, Jahangirnagar University, Savar, Dhaka.

**Preparation and extraction of the sample**

The plant parts were washed thoroughly in distilled water and sundried for 15 days and oven dried for 7 days at 40 °C and then it was grinded into coarse powder. Then plant parts were extracted by maceration extraction process using methanol [4]. 600 g of powder was taken and submersed into 2.5 l of methanol and went through regular shaking for 14 days to prevent clump formation in order to ease of filtration. Then cotton filtration process was performed to get supernatant. The methanol portions from the extract evaporate naturally under the room temperature. The extracts were kept away from sunlight in a air conditioned room and preserved in a refrigerator at 2-8 °C.

**Phytochemical screening**

Phytochemical screening of the extracts were carried out for the detection of active components like carbohydrates, glycosides, saponins, steroids, flavonoids and alkaloids according to the standard protocols [5, 6].

**Antioxidant properties**

**Determination of total phenolic content**

Total phenolic contents of the extracts were determined by the Folin-Ciocalteu reagent method [7]. All of extracts and standard were diluted by serial dilutions as (50 μg/ml to 250 μg/ml) then, on each test tube containing 1 ml of diluted solution of sample and standard, following reagent solutions were added 5 ml folin-ciocalteu reagent (previously diluted with water 1:10 v/v) and 4 ml (7.5% sodium carbonate) of sodium carbonate. Samples were incubated at 20 °C for 60 min and standard diluted solution reagent mixture was incubated at 20 °C for 30 min. Absorbance of samples and standard were measured at 765 nm using UV-VIS spectrophotometer against blank. A typical blank solution contained the solvent used to dissolve the plant extract. The total content of phenol compounds in plant extracts in gallic acid equivalents (GAE) was calculated using the following equation:

\[
C = \frac{(c \times V)}{m}
\]

Where,

- C = total content of phenol compounds, mg/gm plant extract, in GAE
- c = the concentration of Gallic acid established from the calibration curve (mg/ml)
- V = the volume of extract in ml
- m = the weight of crude plant extract in gm

**Determination of total flavonoid content**

Aluminum chloride colorimetric method was used for flavonoids determination [8]. 1 ml of plant extract or standard of different diluted (25 μg/ml to 250 μg/ml) concentrations solutions was taken in a test tube and 3 ml of methanol, 0.2 ml of 10% aluminum chloride, 0.2 ml of 1 M potassium acetate and 5.6 ml of distilled water were added. It was incubated at room temperature for 30 min then absorbance of the reaction mixture was measured at 415 nm using UV-VIS spectrophotometer against blank (methanol). The total flavonoid contents of the plant extracts were expressed as mg/g quercetin equivalent using the equation obtained from the reference standard quercetin.
Determination of total antioxidant capacity

The total antioxidant capacity was evaluated by the phosphomolybdenum method [9]. 0.3 ml of extract and sub-fraction in methanol, ascorbic acid used as standard (25–400 μg/ml) and blank (methanol) were combined with 3 ml of reagent mixture separately and incubated at 95 °C for 90 min. After cooling to room temperature, the absorbance of each sample was measured at 695 nm against the blank. The antioxidant capacity is expressed as the mg/g of equivalents of ascorbic acid and was calculated using the equation obtained from the reference standard ascorbic acid.

DPPH free radical scavenging assay

The free radical scavenging capacity of the extracts was determined using DPPH [10]. 1 ml of plant extract or standard of different diluted concentration (25 μg/ml to 400 μg/ml) solutions were taken in test tubes and freshly prepared 2 ml of 0.004% DPPH solution was added to each test tube to make the final volume of 3 ml. The mixture was incubated at room temperature for 30 min; the absorbance was read at 517 nm using UV-VIS spectrophotometer. Ascorbic acid was used as standard. Control sample was prepared containing the same volume without any extract or standard. Methanol was served as blank. Free radical scavenging activity of the extracts was evaluated as % inhibition and/or IC50 using the following equation:

\[
\%\text{inhibition} = \left(1 - \frac{A_1}{A_0}\right) \times 100\%
\]

Here,

\(A_1\) = Absorbance of the extract or standard
\(A_0\) = Absorbance of the control

Nitric oxide (NO) scavenging assay

Nitric oxide scavenging assay was carried out using sodium nitroprusside [11]. This can be determined by the use of the Griess Illosvoy reaction. 4 ml of each plant extracts or standard of different concentration solutions were taken and 1 ml of Sodium nitroprusside, (5 mM) solution was added into the test tubes. The test tubes were incubated for 2 h at 30 °C to complete the reaction. 2 ml solution was withdrawn from the mixture and mix with 1.2 ml of griess reagent and the absorbances of the solutions were measured at 550 nm using a spectrophotometer against blank. Typical control solutions contain the same solution mixture without plant extract or standard. Percentage inhibition of the NO free radical was measured by using the equation of % inhibition of the DPPH free radical.

Cytotoxicity

The cytotoxic property of the extract was evaluated using brine shrimp lethality test [12]. This investigation was carried out on Artemia salina (Brine shrimp). One spoon of cyst was hatched for 48 h in saline water, prepared by dissolving 38 g NaCl into 1 L water and the cyst became living nauplii. Different concentrations of 800 μg/ml, 400 μg/ml, 200 μg/ml, 100 μg/ml, 50 μg/ml, 25 μg/ml, 12.5 μg/ml and 6.25 μg/ml of both seed and stem bark extract were prepared using dimethyl sulfoxide as solvent. Ten nauplii were taken in each test tube and finally volume was adjusted by saline water. In the present study vincristine sulfate was used as positive control. Vincristine is a very cytotoxic alkaloid and it was evaluated at very low concentrations eg. 25 μg/ml, 12.5 μg/ml, 5 μg/ml, 1 μg/ml, 0.5 μg/ml, 0.25 μg/ml, 0.125 μg/ml and 0.06 μg/ml. 50 μl of DMSO was added to each of three pre marked test tubes containing 4.9 ml of simulated sea water and 10 shrimp nauplii was added to use as negative control groups. All the test tubes were kept in rest for 24 h and then counted for living and dead nauplii by using a magnifying glass against a black background. From this data, the percent (%) of lethality was calculated for each concentration using the following equation-

\[
Pt = \left[\frac{(Po - Pc)}{(100 - Pc)}\right] \times 100
\]

Where,

\(Po\) = Observed mortality
\(Pc\) = Control mortality

The effectiveness or the concentration-mortality relationship of plant product is usually expressed as a median lethal concentration (LC50) and determined by linear regression method from plotting % mortality against correspondent log of concentration.

Thrombolytic activity

In-vitro thrombolytic activity method was described by Daginawala [13]. 1 ml of blood was transferred to each of the 8 previously weighed vials and incubated them 37°C for 45 min. After incubation, blood clots remain in the bottom of the vials and the serum was removed from upper portion. Then the weight of the clots containing vials was taken.100 μl of sample extract solutions was taken in four vials (vials 1, 2, 3, 4) and 100 μl distilled water was taken in vial two vials (Vial 5, 6) and 100 μl streptokinase was taken in last two vials (vial 7 and 8). All the vials were incubated for 90 min at 37°C and again the serum was removed upper portion and the clots remained at the bottom. Then the weight of the vials was taken again. After incubation the lysis of the blood clot was calculated by following equation-
\[
\% \text{ of clot lysis} = \left( \frac{\text{weight of the clot after lysis}}{\text{weight of the clot before lysis}} \right) \times 100
\]

Membrane stabilizing potential
The hypotonic and heat induced membrane stabilizing activity of the extracts was determined on human erythrocytes by using the method of Shahriar [14]. In hypotonic solution-induced hemolysis, the test sample made of stock erythrocyte (RBC) suspension (30 μL) with 5 ml of hypotonic solution (50 mM NaCl) in 1 ml, 10 mM sodium phosphate buffer saline (pH 7.4) containing either the seed and stem bark extracts (1 mg/ml) or acetyl salicylic acid (0.10 mg/ml). The acetyl salicylic acid was used as a reference standard. The mixtures was incubated for 10 min at room temperature, centrifuged for 10 min at 1500 rpm and the absorbance (O.D.) of the supernatant was measured at 540 nm using UV spectrophotometer. The percentage inhibition of either hemolysis or membrane stabilization was calculated using the following equation

\[
\% \text{inhibition of haemolysis} = 100 \times \left( \frac{(\text{OD}_1 - \text{OD}_2)}{\text{OD}_1} \right)
\]

Where,
\[
\text{OD}_1 = \text{Optical density of hypotonic-buffered saline solution alone (control) and}
\]
\[
\text{OD}_2 = \text{Optical density of test sample in hypotonic solution}
\]

In heat induced hemolysis, 5 ml of the isotonic buffer, containing 0.5 ml of seed and stem bark extracts (dissolved in distilled water) was put into two duplicate sets of falcon tubes. The vehicle, in the same amount was added to another tube as control. Erythrocyte suspension (30 μL) was added to each tube & mixed gently by inversion. One pair of the tubes was incubated at 54 °C for 20 min in a water bath. The other pair was maintained at 0-5 °C in an ice bath. The reaction mixture was centrifuged for 10 min at 3000 rpm & the absorbance (O.D.) of the supernatant was measured at 540 nm. The percentage inhibition or acceleration of hemolysis in tests was calculated according to the following equation-

\[
\% \text{Inhibition of haemolysis} = 100 \times \left[ 1 - \left( \frac{\text{OD}_2 - \text{OD}_3}{\text{OD}_1 - \text{OD}_3} \right) \right]
\]

Where,
\[
\text{OD}_1 = \text{test sample unheated,}
\]
\[
\text{OD}_2 = \text{test sample heated &}
\]
\[
\text{OD}_3 = \text{control sample heated}
\]

Evaluation of antimicrobial activity
Disc diffusion method is a classical method used to test antimicrobial susceptibility. This method was first described by Bauer who had attempted the method against a large number of bacteria [15]. In this method, the test substance is soaked in 6 mm diameter filter paper disc and placed in Mueller-Hinton agar media. The test substance will out of the disc into the media based on the solubility of the substance in the media and size. The area where the concentration of the test substance is sufficient to inhibit bacterial growth, there will be clear zones. By measuring the diameter of the clear zone, it is possible to predict the antibacterial activity of the test substance. The same principle can be applied to the measurement of antifungal activity. Standard disc of Kanamycin antibiotic (5 μg/disc), Fluconazole antifungal discs (25 μg), blank discs and solvent discs were used as positive and negative control, respectively. The antimicrobial activity of the test agents was determined by measuring the diameter of zone of inhibition expressed in mm.

Experimental animal
For the experiment Swiss albino male mice, 4–5 weeks of age, weighing between 25 and 30 g were collected from ICDDR, B, Mohakhali, Dhaka. Animals were maintained under standard environmental conditions (temperature: (25.0 ± 2.0) °C, relative humidity: (55–65) % and 12 h light/12 h dark cycle) and free access to feed and water. The animals were acclimatized to laboratory condition for 1 week prior to experiments. All protocols for animal experiment were approved by the institutional ethical committee.

Anti-diarrheal activity by castor oil induced method
The experiment was carried out by the slightly modified procedure previously described by Umer [16]. The anti-diarrheal activity of the methanolic extract of seed and stem bark of Ziziphus mauritiana was evaluated using the method of castor oil induced diarrhea in mice. The animals were divided into control, positive, and test groups containing four mice in each group. Control group received vehicle (plain distilled water) at dose 10 ml/kg orally. The positive control group received loperamide at the dose of 50 mg/kg orally. The test group received methanolic extract of seed and stem bark at the doses of 200 mg/kg and 400 mg/kg body weight. Each animal was placed in an individual jar of the floor surface was covered with absorbent tissue paper. The floor covering was changed at every hour and their stool counted. After 60 min of administration of test samples the mice of all groups were orally treated with 0.5 ml of castor oil. Then the mice were placed in transparent cages to observe for consistency of fecal matter and frequency of detection for 4 h. The activity was expressed as percent inhibition of diarrhea.
Peripheral analgesic activity by acetic acid induced writhing test

The acetic acid writhing test in mice as described by Koster was employed with slight modification [17]. Mice were randomly divided into six groups of four mice in each group. Each group received a particular treatment. The first group was given 10 ml/kg of 1% Tween 80 orally and served as control. Group 2 was served as standard where diclofenac sodium has given to mice as dose of 50 mg/kg of body weight. Groups 3, 4 received 200 mg/kg and 400 mg/kg of body weight of seed extract. Groups 5, 6 received 200 mg/kg and 400 mg/kg of body weight of stem bark extract. Thirty minutes later each mouse was injected intra peritoneally with 0.7% acetic acid at doses of 10 ml/kg of body weight. Full writhing was not always completed by the mice. Accordingly, two half writhing were considered as one full writhing. The number of writhing responses was recorded for each mouse during a subsequent 5 min period after 15 min intra peritoneal administration of acetic acid and the mean abdominal writhing for the each group was obtained and recorded.

Anti-diabetic activity

Mice were kept fasting overnight with free access to water. Before administration of drug and extract solutions fasting blood glucose levels were estimated by glucose oxidase method [18, 19]. Twenty four fasted mice were divided into 6 groups had 4 mice in each group. Group I served as control and received only Tween-80 (a suspending agent) and DMSO mixed with normal saline water. Group II was administered glibenclamide orally at dose of 5 mg/kg body weight. The crude methanolic extract as a dose of 200 mg/kg body weight and 400 mg/kg body weight as fine aqueous suspension were administered orally to group III to group VI. After 60 min of administration of test samples, the mice of all groups were orally treated with 20 mg/ml glucose solution (200 mg/kg body wt). Blood samples were collected from tail vein just prior to glucose administration and at 1, 2 and 3 h after glucose loading. Blood glucose level of the experimental animals was measured by using a glucometer and glucose-oxidase- peroxidase reactive strips.

Acute toxicity study

Acute toxicity describes the adverse effects of a substance which results either from a single exposure or multiple exposures in a short period of time, usually less than 24 h. Acute toxicity of seed and stem bark extract of Ziziphus mauritiana was determined as described by Walum [20]. Mice were randomly divided into eleven groups of four mice in each group. Each group received a particular treatment. Group I served as control and received only 10 ml/kg normal saline water. Group II to VI served as seed extract and group VII to XI served as stem bark extract and received single oral dose each of 500 mg/kg, 1000 mg/kg, 1500 mg/kg, 2000 mg/kg and 4000 mg/kg body weight respectively. Animals were observed for behavior and mortality rate.

Statistical analysis

Data was expressed as Mean ± SEM (Standard error of Mean). The results were analyzed statistically by using Microsoft Excel 2010. T-test of two equal variance were done and results below *p < 0.05, **p < 0.01 and ***p < 0.001 are considered statistically significant. IC_{50} values for scavenging of free radicals, LC_{50} and LC_{90} for cytotoxicity were calculated from the dose response curve.

Results

Ethno-medicinal survey of Ziziphus mauritiana

All the kavirajes interviewed were male, secondary level educated and professional (full time) healer of Gaibandha district of Bangladesh. All of them were belongs to the age in between 40 and 50 years. Ziziphus mauritiana is found almost all over the country and considered as an effective herbal remedy. According to this study, it was found that the various parts of this plant are used in the treatment of in fever, indigestion, liver disease, diarrhoea, wound healing and jaundice (Table 1).

Phytochemical screening

The preliminary phytochemical screening of selected parts of Ziziphus mauritiana confirmed the presence of some secondary metabolites like carbohydrates, glycosides, steroids, flavonoids, alkaloids summarized in Table 2.

Antioxidant properties

Determination of total phenol content

The stem bark extract had highest amount of total phenol content (69.591 ± 0.92 mg/g of GAE) than seed extract (62.475 ± 0.39 mg/g GAE), which was calculated from the regression equation of the calibration curve (y = 0.0069x-0.0341 and R^2 = 0.9961) of reference standard gallic acid (Table 3).

Determination of flavonoid content

The total flavonoid content of both extracts was lower than the total phenolic content, which is often the case for most plants. Quercetin was used as a reference standard to estimate total flavonoid content of the plant extracts. Flavonoid content of seed and stem bark extracts were 15.500 ± 0.50 mg/g and 33.000 ± 2.00 mg/g of quercetin equivalent which was estimated from the regression equation (y = 0.0002x - 0.0002 and R^2 = 0.9965) of quercetin (Table 3).
**Determination of total antioxidant capacity**
The total antioxidant capacities of the seed and stem bark extracts of were determined from the calibration curve \(y = 0.0008x - 0.0002\) and \(R^2 = 0.9906\) established by reference standard ascorbic acid. Total antioxidant capacity of seed and stem bark extract were 13.675 ± 0.87 mg/g and 39.000 ± 0.87 mg/g of ascorbic acid equivalent (Table 3).

**DPPH free radical scavenging assay**
The antioxidant activity of the extractives of BAU Kul was evaluated by the widely used and most reliable DPPH radical scavenging assay method. This antioxidant assay is based on the ability of the extractives to scavenge the stable DPPH radical that contains an odd electron. The calculated \(IC_{50}\) value for the seed and stem bark extract were 4.53 μg/ml \((y = 13.814x - 12.615\) and \(R^2 = 0.9811\)) and 4.13 μg/ml \((y = 14.244x - 8.7692\) and \(R^2 = 0.9606\)) compared against the reference standard ascorbic acid \((4.77 \mu g/ml, y = 13.256x - 13.186\) and \(R^2 = 0.996\)) (Fig. 1 and Table 4).

**Nitric oxide (NO) scavenging assay**
Free radical scavenging activity of Ziziphus mauritiana extracts was also measured by using nitric oxide method. The stem bark extract had higher \(IC_{50}\) of 5.47 μg/ml \((y = 11.66x - 13.725\) and \(R^2 = 0.9949\)) than seed extracts \((5.92 \mu g/ml, y = 10.903x - 14.51\) and \(R^2 = 0.9854\)) compared against the standard ascorbic acid \((4.77 \mu g/ml, y = 13.256x - 13.186\) and \(R^2 = 0.996\)), (Fig. 2 and Table 4).

**Cytotoxicity**
In brine shrimp lethality bioassay, the crude extracts were screened for probable cytotoxic activity. The concentration at which 50% and 90% mortality of brine shrimp nauplii occurred were determined. In this study, seed extract was found most cytotoxic as evidenced from its \(LC_{50}\) and \(LC_{90}\) values (Calculated from regression equation, \(y = 39.29x - 7.6485\) and \(R^2 = 0.9633\)) against brine shrimp \((LC_{50} = 1.46 \mu g/ml\) and \(LC_{90} = 2.48 \mu g/ml\)) compared against reference standard cytotoxic drug vincristine sulphate (Table 4).

**Thrombolytic activity**
Several thrombolytic drugs obtained from various sources are used for the treatment of thrombosis. Thrombolytic agents are used to disrupt already formed blood clots in clinical settings where ischemia may be fatal. As a part of discovery of cardio protective drugs from natural resources the extractives of Z. mauritiana were assessed for thrombolytic activity and the results are presented in Table 4.

**Membrane stabilizing potential**
The different parts of crude extracts of Z. mauritiana protected the haemolysis of RBC, induced by hypnotic solution and heat as compared to the standard acetylsalicylic acid (ASA). In hypotonic solution induced and heat induced hemolysis, ASA inhibited the haemolysis of RBC \(64.24 \pm 0.44\%\) and \(67.59 \pm 1.462\%\), seed inhibited \(25.00 \pm 1.56\%\) and \(34.79 \pm 2.553\%\), stem bark inhibited \(41.88 \pm 1.58\%\) and \(46.0 \pm 2.951\%\) respectively, summarized in Table 4.

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**Table 1** Traditional usage of Ziziphus mauritiana

| Plant Parts | Disease | Method of Use |
|-------------|---------|---------------|
| Fruits      | Improved the digestion and liver function | Juice of fruit and fresh fruit |
|             | Fever   | Prepared juice combined with Chalta (Dellenia indica) by mixing with sugar |
|             | Wound healing | Dried ripe fruit given orally |
| Leaves      | Liver function and jaundice | Prepared juice by mixing with sugar |
| Barks       | Wound healing | Topical application of paste |
| Roots       | Wound healing | Topical application of paste |
| Seeds       | Not Used | |
| Flowers     | Not Used | |

**Table 2** Phytochemical screening of Ziziphus mauritiana extractives

| Phytochemical Tests | Seed | Stem Bark |
|---------------------|------|-----------|
| Carbohydrates       | ++   | +++       |
| Glycosides          | +++  | ++        |
| Glucosides          | –    | +         |
| Saponins            | +    | +++       |
| Steroids            | +++  | +         |
| Tannins             | –    | +         |
| Flavonoids          | +    | +         |
| Alkaloids           |      |           |
| Hager’s reagent     | ++   | +         |
| Wagner’s reagent    | +    | ++        |
| Dragendorff’s reagent | +   | +         |

[Note, – = Not present, + = Present in mild amount, ++ = Present in moderate amount, +++ = Present in large amount]
Evaluation of antimicrobial activity
In this present study, two gram positive, two gram negative and a fungus microorganism were evaluated for anti-microbial activities and only the stem bark extract showed antibacterial activity (Fig. 3), but lacked of anti-fungal activity at 400 μg/disk (Table 5).

Anti-diarrheal activity by castor oil induced method
The effect of the methanolic extract of *Z. mauritiana* on castor oil induced diarrhoea in mice showed a dose dependent decrease in the number of defected pellets. The seed extract at a dose 200 mg/kg and 400 mg/kg reduced defected pellet by 37.84% and 62.16% whereas the stem bark extract showed 54.05% and 75.68% inhibition of defection respectively, shown in Fig. 4.

Peripheral analgesic activity by acetic acid induced writhing test
In the analgesic activity performed using acetic acid-induced writhing model in mice, the seed extract produced 45.10% and 64.71% writhing inhibition at the doses of 200 mg/kg and 400 mg/kg body weight respectively. At same dose, the others sample methanol stem bark extract inhibited the writhing 49.02% and 68.63% respectively, shown in the Fig. 5.

Anti-diabetic activity
The hypoglycemic activity is an important parameter to evaluate anti-diabetic activity and it was showed by dose dependent manner. In this study, upon increment of the dose, the plant extracts showed increased level of hypoglycemic activity. According to this study, stem bark extract showed higher % reduction of blood glucose level than seed extract at both dose (Fig. 6).

Acute toxicity study
Safety of plant extract is evaluated mostly by acute oral toxicity study. Among the 11 mice group, no death or toxic reaction was observed in mice during the test period. This indicated the absence of any toxic material in the extracts of *Z. mauritiana*. The seed and stem bark plant extract of this plant was found safe up to 4000 mg/kg body weight dose.

Discussion
Herbal medicines are the staple of medical treatment in many developing countries and used for virtually all minor ailments. Individual herbal medicines in developing regions vary considerably, healers in each region have learned over centuries which local herbs have medicinal worth [21]. Traditional botanical knowledge and experience can be useful not only for the biologists, but also these are equally relevant for anthropologist,
archaeologists, environmental scientists, foresters, sociologists, literatures, geographers, pharmacists and many others who take interests bioresources [22]. The fresh fruit and juice of fruit of *Ziziphus mauritiana* improved the digestion and liver disease. Sometime the combination fruit of BAU Kul (*Ziziphus mauritiana*) and Chalta (*Dellenia indica*) are given in treatment of fever prepared by juice mixing with sugar. Dried ripe fruit given as orally in the treatment of wound healing. The paste of bark and root also effective on wound healing and in the treatment of diarrhea, dried bark and young roots are blended and combined/single, prepared juice mixed with sugar and water administered orally. It is reported that the juice of leaves improves liver function and jaundice. There were no use found of flower and seed. Due to a lack of research, little is known about the safety of consuming any type of *Ziziphus mauritiana* plant in dietary supplement form, but in our present study, there were no side effect reported by the folk medicine practitioners. Seed contained carbohydrate, glucoside, saponin, steroid, flavonoid, alkaloid but it also indicates the absence of glycoside and tannin. The stem bark showed the presence of all secondary metabolites which are carbohydrate, glycoside, glucoside, saponin, steroid, tannin, flavonoid and alkaloid. Phenolics are important constituents that contribute to the functional quality, color and flavor of plants and they have significant roles both as singlet oxygen quenchers and free radical scavengers that help to minimize molecular damage [23]. Both extract showed higher phenolic content, moderate flavonoid content and antioxidant capacity. The antioxidant activity results showed increased activity by dose

| Extracts | Scavenging capacity assay, IC\textsubscript{50} (μg/ml) | Brine shrimp lethality bioassay (μg/ml) | Thrombolytic activity | % Inhibition of hemolysis ± SEM (membrane stabilizing potential) |
|----------|-------------------------|--------------------------------------|----------------------|-------------------------------------------------------------|
|          | DPPH | NO | LC\textsubscript{50} | LC\textsubscript{90} | % of clot lysis | Hypotonic Solution | Heat Induce |
| SE       | 4.53\textsuperscript{a, b} | 5.92\textsuperscript{b} | 1.46\textsuperscript{a} | 2.48 | 24.95 | 25.00 ± 1.56 | 34.79 ± 2.55 |
| SBE      | 4.13\textsuperscript{a, b} | 5.47\textsuperscript{b} | 1.92\textsuperscript{a} | 3.25 | 34.65 | 41.88 ± 1.58 | 46.03 ± 2.95 |
| AA       | 3.43 | 4.77 | – | – | – | – | – |
| VS       | – | – | 0.39 | 1.12 | – | – | – |
| SK       | – | – | – | – | 66.38 | – | – |
| DL       | – | – | – | – | 6.91% | – | – |
| ASA      | – | – | – | – | – | 64.24 ± 0.44 | 67.59 ± 1.46 |

\textsuperscript{a} SE Seed Extract, \textsuperscript{b} SBE Stem Bark Extract, \textsuperscript{AA} Ascorbic Acid, \textsuperscript{VS} Vincristine Sulphate, \textsuperscript{SK} Streptokinase, \textsuperscript{DL} Distilled Water, \textsuperscript{ASA} Acetyl Salicylic Acid. Data was expressed as Mean ± SEM (Standard error of Mean). t-test of two equal variance was done to analyze the data set. Values in the same column with different superscripts are significantly different from another, \(p<0.05\).
dependent manner. Both extracts showed excellent free radicals scavenging potential. In this study, stem bark extract was more potential antioxidant potential than seed extract. Both extracts showed very good cytotoxic activity compared to the standard vincristine sulphate. The lower the LC$_{50}$ and LC$_{90}$ values of a sample indicate the higher bioactivity. A similar result was found in ethanol extracts of leaves of this plant [24].

In present study, methanolic seed and stem bark extracts of *Z. mauritiana* were evaluated for possible thrombolytic potential. The thrombolytic activities of the extracts were evaluated against reference standard streptokinase. 100 μl streptokinase (SK) as a positive control (30,000 IU) showed 66.38% lysis of clot on the other hand sterile distilled water was treated as negative control which exhibited a negligible percentage of lysis of clot 6.91%. In vitro thrombolytic activity study revealed that seed and stem bark extract exhibited 24.95% and 34.65% clot lysis respectively. This study revealed that the stem bark extract of *Z. mauritiana* exhibited highest thrombolytic activity. The seed and stem bark extracts had lower clot lysis potential than leaves extract [25]. Inflammation is due to release of lysosomal constituent’s that also causes cell death. Rupturing of lysosomal membrane releases lysosomal constituents and the stabilization of lysosomal membrane inhibits the release of lysosomal constituents [26]. Thus. Membrane stabilizing activity is related to anti-inflammatory activities. Inhibition of hypotonic solution and heat induced erythrocyte membrane lysis can be taken for the in-vitro determination of anti-inflammatory activity of drugs or plant extracts [27]. Both seed and stem bark extracts showed potential anti-inflammatory activity. A similar data were found of different extractives of leaves of *Z. mauritiana* on hypotonic solution and heat induces hemolysis of erythrocyte membrane [26]. A human pathogen is a kind of disease causing microorganism which induces infection by invasion and multiplication within human being. Sometimes, one’s immune system alone cannot fight against these virulent pathogens when an individual exposed to them. In that case antimicrobial agents as therapeutics are used. Among all the microbes, bacteria is responsible for terrible infectious diseases in humans. Microorganism grows very fast by multiplication and induces infection through endotoxins and exotoxins [28]. To kill bacteria, there huge numbers of antibiotics are available. But unfortunately, these pathogens can easily be mutated and become less susceptible or resistant towards latest generation antibiotics by changing their genetic information very quickly. Whereas the side effects of such antibiotics can cause serious organ damage like liver and kidney [29]. As a result, in critical clinical situations, it is seen that patients are dying in infectious diseases due to the inactivity of antibiotics. In the present study, among the five tested organisms, the stem bark extract of *Ziziphus mauritiana* resulted in variable zone of inhibitions but seed extract showed resistance to all tested microorganism at concentration 400 μg/ml. Stem bark extract displayed its effectiveness against gram-positive strain *Staphylococcus aureus*, *Bacillus megaterium* and gram negative strain *Escherichia coli, Salmonella typhi* to a potential extent compared with kanamycin shown in Table 3. A similar activity was found for different parts of *Ziziphus mauritiana* [3, 24, 30].

Table 5 Antimicrobial activity of *Z. mauritiana* extractives

| Test Microorganisms Zone of inhibitions (mm) | Methanol | Seed | Stem Bark | Standard |
|---------------------------------------------|----------|------|-----------|----------|
| **Gram positive bacteria**                  |          |      |           |          |
| *Staphylococcus aureus*                     | 0        | 0    | 14        | 27       |
| *Bacillus megaterium*                       | 0        | 0    | 12        | 25       |
| **Gram negative bacteria**                  |          |      |           |          |
| *Escherichia coli*                          | 0        | 0    | 10        | 24       |
| *Salmonella typhi*                          | 0        | 0    | 10        | 20       |
| **Fungi**                                   |          |      |           |          |
| *Aspergillus niger*                         | 0        | 0    | 0         | 20       |

Fig. 3 Zone of inhibition of *Ziziphus mauritiana* extracts and standard in selected microorganism.
In anti diarrheal study, significant reduction of defected pellet was found compared to the standard loperamide. Stem bark extract showed higher activity than seed extract at same dose. At a dose 400 mg/kg body weight both stem bark and seed extract exhibited highly reduction of the defected pellet compared with the standard loperamide at dose 50 mg/kg body weight. Similar result was also reported previously of different part of this plant [31, 32]. The diarrhea inducing properties is as a result of ricinoleic acid; a castor oil active metabolite, which is liberated by the action of lipases in the upper part of the small intestine. Ricinoleic acid exerts its effect by production of local irritation and inflammation of the intestinal mucosa, causing the release of
prostaglandins that eventually increases gastrointestinal motility and net secretion of water and electrolytes [33]. Thus, the anti diarrheal effect of the seed and stem bark could be attributed to inhibition of castor oil-induced prostaglandin synthesis. The anti-diarrheal activity might also be due to inhibition of active secretion of ricinoleic acid, resulting in the activation of Na\(^+\), K\(^+\) ATPase activity that promotes absorption of Na\(^+\) and K\(^+\) in the intestinal mucosa. This effect could probably be linked to the presence of terpenoids, tannins and flavonoids in the seed and bark extract, which are shown to promote colonic absorption of water and electrolytes [34]. The acetic acid induced abdominal constriction test is used frequently for peripherally acting drugs. The pain induction occurs by liberating endogenous substances as well as some other pain mediators such as arachidonic acid metabolites via cyclooxygenases, such as prostaglandins [35]. The stem bark extract showed more significant analgesic activity than seed extract. At the dose of 400 mg/kg body weight, the both extract seed and stem bark showed that higher writhing inhibition than the standard drug diclofenac sodium at the dose of 50 mg/kg body weight. In this study, the positive standard diclofenac sodium inhibited about 60.78% at the dose of 50 mg/kg body weight. Similar activity is found in previous study of different parts of this plant [36, 37]. At higher dose (400 mg/kg), for both extracts, % reduction of blood glucose level almost similar to the standard drug glibenclamide at a dose 5 mg/kg body weight. The similar hypoglycemic activity was found in previous study of different parts of this plant. The fruit extracts of *Ziziphus mauritiana* was found effective in reducing both hyperlipidemia and hypoglycaemia accompanying diabetes on alloxan induced diabetic rats. The most effective dose was 400 mg/kg body weight [38]. The aqueous extract and the non-polysaccharide fraction of the aqueous extract were found to exhibit significant anti-hyperglycemic and hypoglycemic activities. The petroleum ether extract was found to exhibit only an anti hyperglycemic effect [39]. Another study showed seed extract alone or in combination with glyburide reduced the blood glucose level in all the alloxan diabetic mice after acute and sub-acute (28 days) administration [40]. Drug safety is one of the important topics in daily medical practice. In this study, no mortality showed even at a maximum dose up to 4000 mg/kg body weight for both extracts. This observation is agreed with who assessed acute and sub-acute toxicity of hydro-alcoholic fruit extracts of this plant using higher dose 5000 mg/kg and they reported no behavioural changes and no mortality was observed in rats [38]. There are rare reports on chemical composition on the different parts of *Z. mauritiana*. Ashraf et al., investigated the chemical composition of *Z. mauritiana* leave extracts by GC-MS analysis and found methyl stearate and α-linolenic acid as the major components in methanol and hexane extracts. They also confirmed the presence of palmitic acid, phytol methanol, hexane and chloroform extracts of the leaves of *Z. mauritiana* [41]. Extensive analysis about different phyto-constituents in different parts of *Z. mauritiana* along with their pharmacological properties is highly required.
Conclusion
The presence of different phytochemical may be responsible for in-vitro and in-vivo pharmacological properties of BAU Kul. Further investigations are in progress to identify the active constituents responsible for pharmacological activity of Ziziphus mauritiana. In this present study, the data was satisfactory. The difference of pharmacological activity of BAU Kul (Z. mauritiana) may be varying with other Z. mauritiana due to genetic variations. In ethnobotanical survey of this study, the data were collected only Gaibandha district in Bangladesh. These data do not include the area of all over the country; therefore regional variation may occur. However, more detailed phytochemical analysis will be necessary to isolate and characterize the active compounds responsible for the pharmacological activities as well as to understand the exact mechanisms of action of their underlying pharmacological activities.

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Authors’ contributions
Authors Md. Khokon Miah Akanda and A. H. M. Nazmul Hasan were involved in the laboratory work. Author A.H.M. Nazmul Hasan designed this study and acted as a supervisor of this study. The author(s) read and approved the final manuscript.

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Availability of data and materials
All the data are preserved by the authors and ready to provide at any stage if any question arise.

Ethics approval and consent to participate
This study was approved in a meeting of Committee for Advanced Studies Ethics approval and consent to participate if any question arise. All the data are preserved by the authors and ready to provide at any stage if any question arise.

Consent for publication
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