Inter- and intraspecific genetic and morphological variation in a sibling pair of carabid species

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Background: Pogonus littoralis and Pogonus chalceus are very close related species with quite different ecological preferences within salt marshes. We study the evolutionary processes in and between these presumably young species. Therefore, we compare the variation in ecologically relevant characters and the genetic variation within one of the species (intraspecific differentiation) with the variation of the two types of characters between the two species (interspecific variation).

Data are compared between two independent sets of populations, one set at a small geographical scale (the ecologically diverse Guérande area in France) and the other set at an Atlantic-Mediterranean scale.

Results: Body and relative wing size and IDH1 allozyme data show that the intraspecific variation in P. chalceus is high and in the same range as the interspecific variation (P. chalceus versus P. littoralis). Based on neutral markers (other allozymes and mitochondrial DNA) on the other hand, the intraspecific variation in P. chalceus is much lower in comparison to the interspecific variation.

Conclusion: The different ecotypes in the highly polytypic species P. chalceus are as highly differentiated in ecological characters as true species, but are not recognised as such by screening neutral DNA polymorphisms. This can be interpreted as a case of ongoing speciation driven by natural selection adapting each ecotype to its respective ecological niche. The same ecological process can be recognised in the differentiation between the two sister species, where en plus reproductive isolation between the two gene pools occurred, allowing independent drift and mutation accumulation in neutral genetic characters.
distinguishable ecotypes, respectively adapted to canal and pond habitat. Comparisons between the Guérande region (microscale) and populations along the Atlantic coast (macroscale) confirmed the generality of the hypothesis regarding ecological processes responsible for this differentiation: habitat stability [2]. The Guérande ecotypes are also slightly differentiated based on neutral molecular markers (microsatellites and allozymes), suggesting that partial barriers to gene flow between the two ecotypes are present. Previous work on a wide range of taxa has demonstrated that strong natural selection can lead to divergence in spite of gene flow [3-7]. Our Guérande results can therefore be interpreted as a case of ongoing speciation driven by natural selection adapting each ecotype to its respective ecological niche, i.e. species in *status nascenti* (see also [8,9]).

In the same Guérande region and along the European Atlantic and Mediterranean coast, another *Pogonus* species, *P. littoralis* (Duftschmid, 1812) lives in a third kind of microhabitat: unvegetated, temporary dry salt marsh ponds or creeks, where it lives between cracks in humid sea clay. This species is, in contrast to *P. chalceus*, constantly macropterous, always with maximally developed wings and functional flight musculature [10]. The beetle is highly mobile because it regularly has to move between temporarily dry salt marsh ponds and creeks during its life cycle. Both species can be hardly distinguished by external morphology (for example large individuals of *P. chalceus* versus small *P. littoralis*) but have clearly distinguishable genitalia.

The data in this article are to some extent compiled from previous works [2,11,12]. Nevertheless, the novelty of this study lies in the fact for the first time the two carabid sister species are analyzed jointly allowing for valuable comparisons to be made. In this study, we will first compare the two ecotypes of *Pogonus chalceus* with the closely related species, *Pogonus littoralis* at a microscale (Guérande region). Therefore, we will use population data on wing and body size, *IDH1* allozyme polymorphism as well as apparently neutral markers (other allozymes and mtDNA). We will also test if the microscale results are valid at a larger scale across Europe by means of an independent data set of different populations of both *Pogonus* species. In all of these cases, we will evaluate the contribution of intra population, inter population, inter ecotype and interspecific variation to the total variance.

**Results**

**Body size**

Fig. 1 also shows male and female body sizes for the *P. chalceus* ecological groups and for the *P. littoralis* populations on a European scale [see also additional file 2]. Mean male body size is small for the stable (3.68; range: 3.2–4.3) and intermediate *P. chalceus* populations (3.68; range 3–4.3), somewhat higher for the temporary populations (mean: 3.92; range: 3.4–4.6) and high for the *P. littoralis* (mean: 4.32; range: 3.6–4.8) ones. Body size values of females show again a similar pattern and are always larger than male body sizes. Mean female body size is 4.04 for the stable *P. chalceus* populations (range: 3.2 to 4.6) compared to 4.11 for the intermediate populations (range: 3.1 to 4.7) and 4.3 for the temporary *P. chalceus* populations (range: 3.4 to 4.8) and 4.6 for the *P. littoralis* populations (4.1–5.2).

In the Guérande region and considering the two species (nested design ANOVA; six *P. chalceus* populations (canals and ponds pooled) versus three *P. littoralis* populations, the major part of variance (based on body size) is found among species (Table 1; 74.24% for males and 51.96% for females). If we consider three groups (three canal populations (*P. chalceus*), three pond populations (*P. chalceus*) and three *P. littoralis* populations, the major part of variance is even more pronouncedly found among groups (84.96% for males and 72.08% for females). Variance among populations within groups considering three groups instead of two drops from 17.5 to 2.35% for males and from 29.57 to 4.28% for females. This indicates that this variance was almost completely due to the differences in body size between populations of *P. chalceus* from different microhabitats. All variance components are statistically significant.

On a European scale and considering the two species (25 *P. chalceus* populations versus six *P. littoralis*), the major part of variance (based on body size) is found among species (Table 1; 68.37% for males and 48.37% for females). If we consider four ecological groups (14 temporary (*P. chalceus*), five intermediate (*P. chalceus*), six stable (*P. chalceus*) and five *P. littoralis*), the variance among groups drops (49.39% for males and 33.87% for females) and the variance within populations augments (45.62% for males and 58.53% for females). Variance among populations within groups considering four groups instead of two drops a little from 10.13 to 5% for males and from 12.29
Relative wing size

Fig. 2 shows male and female relative wing sizes for both P. chalceus microhabitats and for the P. littoralis populations in the Guérande region [see also additional file 1]. Mean male relative wing size for the canal populations is small (28.19; range: 20–35%), intermediate for the ponds (mean: 64.24; range: 25–82.5%) and high for the P. littoralis populations (mean: 103.59; range: 92.5–112.5%). Relative wing size values of females show a similar pattern and are not larger than male relative wing sizes. Mean female relative wing size for the canals is 26.93 (range: 17.5–32.5%), 62.31 for the ponds (range: 25–80%) and 103.04 for the P. littoralis populations (range: 92.5–110%).

Fig. 2 also shows male and female relative wing size in ecological groups of P. chalceus and of P. littoralis populations on a European scale [see also additional file 2]. Mean male relative wing size is small for the populations of the old, highly stable salt marsh areas (35.28; range: 22.5–62.5%), some higher for the populations of the salt marshes of intermediate stability (mean: 51.07; range: 25–85%), higher for the populations of the small, unstable areas (mean: 82.23, range: 40–112.5%) and very high for the P. littoralis populations (mean: 106.16; range: 90–112.5%). Relative wing size values of females show a similar pattern and are not larger or smaller than male relative wing sizes. Mean female relative wing size for the stable populations is 33.43 (range: 20–82.5%) compared to 49.91 for the populations of intermediate stability situations (range: 25–85%), 80.41 for the temporary populations of the highly unstable salt marshes (range: 40–
97.5%) and 107 for the *P. littoralis* populations (range: 92.5–115%).

In the Guérande region and considering the two species (nested design ANOVA; six *P. chalceus* populations (canals and ponds pooled) and three *P. littoralis* populations), the major part of variance (based on relative wing size) is found among species (Table 2; 78.84% for males and 80.37% for females). If we consider three groups (three canal populations (*P. chalceus*), three pond populations (*P. chalceus*) and three *P. littoralis* populations), the major part of variance is even more clearly found among groups (95.48% for males and 93.92% for females). Variance among populations within groups considering three groups instead of two drops from 18.64 to 0.64% for males and from 16.21 to 0.8% for females. This indicates that this variance is almost completely due to the differences in relative wing size between populations of *P. chalceus* from different microhabitats. All variance components are statistically significant.

On a European scale and considering the two species (25 *P. chalceus* populations versus six *P. littoralis* populations), the major part of variance (based on relative wing size) is found among species (Table 2; 57.61% for males and 60.16% for females). If we consider four ecological groups (14 temporary (*P. chalceus*), five intermediate (*P. chalceus*), six stable (*P. chalceus*) and six *P. littoralis* populations), the major part of variance is again even more pronounced among groups (82.22% for males and 82.58% for females). Variance among populations within groups considering four groups instead of two drops from 36.11 to 6.34% for males and from 32.8 to 6.89% for females. This indicates that this variance is again almost completely due to the differences in relative wing size between populations of *P. chalceus* from different ecological or salt marsh area stability groups. All variance components are statistically significant.

### IDH1 allozyme marker

In Guérande, both *Idh1*-2 and *Idh1*-4 alleles are frequent in ponds, whereas canals are nearly fixed at *Idh1*-4 (Fig. 3A) [see also additional file 1]. *P. littoralis* populations are fixed at the *Idh1*-6 allele. Allele *Idh1*-1, *Idh1*-3, and *Idh1*-5 are very rare in *P. chalceus* and therefore not shown in Figure 4. Considering the two species (AMOVA; six *P. chalceus* populations (canals and ponds pooled) and three *P. littoralis*), the major part of variance (based on *IDH1*) is found among groups (Table 3; 61.93%). If we consider three groups (three canal populations (*P. chalceus*), three pond populations (*P. chalceus*) and three *P. littoralis* populations, the major part of variance is still found among groups (64.25%). Variance among populations within groups considering three groups instead of two drops from 11.49 to 0.1%. This indicates that this variance is almost completely due to differences in *IDH1* between populations of *P. chalceus* from different microhabitats. All variance components are statistically significant.

At a European scale, both *Idh1*-2 and *Idh1*-4 alleles are frequent in the intermediate stability populations, whereas the temporary populations are nearly fixed at the *Idh1*-2 allele and the stable populations at the *Idh1*-4 allele (Fig. 3B) [see also additional file 2]. *P. littoralis* populations are fixed at the *Idh1*-6 allele. Considering two groups (25 *P. chalceus* populations versus six *P. littoralis* populations, the major part of variance (based on *IDH1*) is found among groups (Table 3; 62.29%). If we consider four ecological groups (14 temporary (*P. chalceus*), five intermediately stable (*P. chalceus*), six highly stable (*P. chalceus*) and six *P. littoralis* populations, the major part of variance is somewhat lower but is still found among groups (53.26%). Variance among populations within groups, considering four groups instead of two, drops from 12.26 to 2.07%. This indicates that this variance is almost completely due to differences in *IDH1* between populations of *P. chalceus* from different ecological

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Table 1: Analysis of variance (nested design ANOVA) based on male or female body size in two regions: Guérande microscale and Europe macroscale

| region       | groups                          | source of variation | % var male | % var female |
|--------------|--------------------------------|---------------------|------------|-------------|
| Guérande     | *P. chalceus*/P. littoralis     | among groups        | 74.24      | 51.96       |
|              |                                 | among populations within groups | 17.50 | 29.57       |
|              |                                 | within populations within groups | 8.26 | 18.47       |
|              | ponds/canals/P. littoralis       | among groups        | 84.96      | 72.08       |
|              |                                 | among populations within groups | 2.35 | 4.28        |
|              |                                 | within populations within groups | 12.69 | 23.63       |
| Europe       | *P. chalceus*/P. littoralis     | among groups        | 68.37      | 48.37       |
|              |                                 | among populations within groups | 10.13 | 12.29       |
|              |                                 | within populations within groups | 21.51 | 39.35       |
|              | stable/intermediate/temporary/P. littoralis | among groups | 49.39 | 33.87       |
|              |                                 | among populations within groups | 5.00 | 7.61        |
|              |                                 | within populations within groups | 45.62 | 58.53       |
groups, as *P. littoralis* is fixed in a different allele. All variance components are statistically significant.

**Other allozymes**

The number of studied individuals and allozyme allele frequencies for each population in the Guérande region is given in additional file 3. In the Guérande and considering two groups (AMOVA; six *P. chalceus* populations (canals and ponds) versus three *P. littoralis* populations), the major part of variance (based on four neutral allozymes) is found between species (Table 3; 58.43%). If we consider three groups (three canal populations (*P. chalceus*), three pond populations (*P. chalceus*) and three *P. littoralis* populations), the variance among groups drops to 41.99% and the major part of variance is now found within populations (53.02%). Variance among populations within groups considering three groups instead of two remains in the same range (3.39% for two groups compared to 4.98% for three groups).

The number of studied individuals and allozyme allele frequencies for each population at a European scale is given in additional file 4. At a European scale and considering two groups (AMOVA; 25 *P. chalceus* populations and six *P. littoralis* populations), the major part of variance (based on four neutral allozymes) is found within populations (Table 3; 49.81%) and among groups (36.49%). If we consider four ecological groups (14 temporary (*P. chalceus*), five intermediate stable (*P. chalceus*), six highly stable (*P. chalceus*) and six *P. littoralis* populations), the variance among groups drops to 19.05% and the major part of variance is still found within populations (65.24%). Variance among populations within groups considering four groups instead of two remains in the same range.
(13.70% for two groups compared to 15.71% for four groups). All variance components are statistically significant.

Mitochondrial DNA

The 459-bp COI mitochondrial sequences revealed two haplotypes in the Guérande region. Haplotype one was shared by individuals of both canal and pond ecotype. Haplotype two was exclusive to the canal ecotype (Table 4). The 497-bp 16S sequences revealed only one haplotype in the Guérande region (Table 5).

The 459-bp COI sequences included 32 variable sites on a European scale (29 parsimony informative) and revealed nine unique haplotypes (four for P. chalceus and five for P. littoralis, with no haplotype shared by the two species). Most haplotypes were exclusive to a particular sampling site, with the exception of haplotype one and three which appeared in eight different localities, and haplotype five, which was found in three localities (Table 4). Selective neutrality was confirmed for this gene ($P > 0.1$ for all test statistics with $D^*$ and $F^*$).

The neighbour joining tree in Figure 5 shows that both species form clearly separated entities (high bootstrap values). Differences are found in 28 positions between haplotypes 5,6,8,9 (P. littoralis) and haplotypes 1 and 2 (P. chalceus; Table 4). 27 base differences are found between haplotypes 5,6,8,9 and haplotypes 3 and 4 (P. chalceus). 27 base differences are found between haplotype 7 (P. littoralis) and haplotypes 1,2 (P. chalceus) and 26 base differences with haplotypes 3,4 (P. chalceus). Intrapopulation differences in both P. chalceus and P. littoralis are very small (only a limited number of individuals studied) and between each haplotype there are only one to at most two bases different.

The 497-bp 16S sequences included 11 variable sites (9 parsimony informative) and revealed three unique haplotypes (Table 5; two for P. chalceus and one for P. littoralis, Table 3: Analysis of molecular variance (AMOVA) based on IDH1 allozyme or 4 neutral allozymes in two regions: Guérande microscale and Europe macroscale

| source of variation | % var IDH1 | % var allo |
|---------------------|------------|-----------|
| Guérande             |            |           |
| P. chalceus/P. littoralis among groups | 61.93      | 58.43     |
| among populations within groups | 11.49      | 3.39      |
| within populations | 26.57      | 39.18     |
| ponds/canals/P. littoralis among groups | 64.25      | 41.99     |
| among populations within groups | 0.1        | 4.98      |
| within populations | 35.65      | 53.02     |
| Europe              |            |           |
| P. chalceus/P. littoralis among groups | 62.29      | 36.49     |
| among populations within groups | 12.26      | 36.49     |
| within populations | 25.44      | 32.80     |
| stable/intermediate/temporary/P. littoralis among groups | 53.26      | 19.05     |
| among populations within groups | 2.07       | 15.71     |
| within populations | 44.67      | 65.24     |
with no haplotype shared by the two species). Only one haplotype was exclusive to a particular sampling site (haplotype two). Haplotype one appeared in all 11 *P. chalceus* localities, and haplotype three appeared in all three *P. littoralis* sites. Selective neutrality was confirmed for this gene (*P > 0.1* for all test statistics with *D* and *F*). Both species form, as in the case for COI, clearly separated entities (Table 5). 10 base differences are found between haplotype 3 (*P. littoralis*) and haplotypes 1 and 2 (*P. chalceus*). Interpopulation differences in both *P. chalceus* are very small (between the two haplotypes there is only one base different). There were no interpopulation differences found in *P. littoralis*.

**Discussion**

*Pogonus littoralis* and *Pogonus chalceus* are closely related species, sometimes relatively hard to identify without dissection of the genitalia. We are interested to study the evolutionary processes in and between these presumably young species. We therefore compare the degree of intraspecific variation (in ecological groups of *P. chalceus*) and the degree of interspecific variation (*P. chalceus* versus

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**Figure 3**

Allele frequencies for *IDH1* in the Guérande region (part A; canals (*P. chalceus*) and ponds (*P. chalceus*) and *P. littoralis*). Allele frequencies for *IDH1* on a European scale (part B; stable (*P. chalceus*) and intermediate (*P. chalceus*), temporary and *P. littoralis*). *Idh1*-2: black, *Idh1*-4: light gray, *Idh1*-6: dark gray.
relative wing size, very large part of the total variance (based on body size, sister species size differences as well as genetic differences between the found between species (summary in Table 6). The study of populations versus ancestry among populations within groups drops drastically ecotypes in \((At\) both geographical scales and considering two groups \((P. chalceus\) populations versus \(P. littoralis\) populations), a very large part of the total variance (based on body size, relative wing size, \(IDH1\) and four neutral allozymes) is found between species (summary in Table 6). The study of the two mitochondrial genes also shows that both species form clearly separated entities. It is clear that relative wing size differences as well as genetic differences between the sister species \(P. chalceus\) and \(P. littoralis\) (interspecific) in this study are very marked and allow an easy species recognition.

\(P. littoralis\) between a variety of morphological characteristics and molecular markers. In all of these cases, we did this with an ANOVA splitting up the total variance among groups, among populations within groups and within populations (Table 6).

At both geographical scales and considering two groups \((P. chalceus\) populations versus \(P. littoralis\) populations), a very large part of the total variance (based on body size, relative wing size, \(IDH1\) and four neutral allozymes) is found between species (summary in Table 6). The study of the two mitochondrial genes also shows that both species form clearly separated entities. It is clear that relative wing size differences as well as genetic differences between the sister species \(P. chalceus\) and \(P. littoralis\) (interspecific) in this study are very marked and allow an easy species recognition.

Body size, relative wing size and \(IDH1\) allozyme data in the beetle \(P. chalceus\) are also strongly divergent between contrasting microhabitats (intraspecific: two ecotypes in Guérande) as well as between three ecological groups at macroscale (highly stable versus temporarily stable and temporary populations; \([2]\) and this study). If we consider four groups on a macroscale (3 groups in \(P. chalceus\) + 1 group of \(P. littoralis\)) or three groups on a microscale (2 ecotypes in \(P. chalceus\) + 1 group of \(P. littoralis\)), the variance among populations within groups drops drastically as compared to the analysis of two groups (all \(P. chalceus\) populations versus \(P. littoralis\); based again on body size, relative wing size and \(IDH1\); summary in Table 6). This study clearly shows that the intraspecific variation based on those three characteristics in \(P. chalceus\) is very high and in the same order of magnitude as the degree of interspecific variation (\(P. chalceus\) versus \(P. littoralis\)). We have suggested earlier that this huge phenotypic and \(IDH1\) divergence in \(P. chalceus\) has been driven by divergent natural selection \([2]\). As relative wing size is to a large extent genetically determined \([1]\), this indeed suggests divergent selection between populations. And the observation that the \(IDH1\) locus screened within our samples shows allelic differences between habitats strongly suggests a locus undergoing evolution through natural selection. Moreover, the canal and pond microhabitats differ from each other with respect to temperature, salinity and water level fluctuations \([2]\). Numerous studies based on allozymes have revealed patterns of allelic distribution associated with environmental factors, such as temperature and salinity \([13,14]\). Regarding the function of \(IDH1\), the enzyme catalyses the rate-limiting step of the tricarboxylate cycle. Possible links with growth, however, are not direct and could be associated with the energy that is produced from the reaction. Divergent selection can lead to reproductive isolation and assortative mating and ultimately to speciation \([8,15]\).

On the other hand, in a previously study was shown that \(P. chalceus\) ecotypes in the Guérande region were only slightly differentiated (based on allozyme and microsatellite markers) compared to the results based on adaptive characteristics \([2]\). The smaller degree of intraspecific divergence is also reflected in the mitochondrial data from this study. Moreover, allozyme and mtDNA data from this study show that the populations of \(P. chalceus\) are much more related to each other than to their sister species \(P. littoralis\) both on a micro- and macroscale. Often, little or no genetic divergence is found in neutral markers between ecologically and morphologically differentiated populations \([3-5,7,16-18]\). Our results can be interpreted as a case of ongoing speciation in \(P. chalceus\) where divergence reflects a balance between selection and gene flow (see also \([2]\)). Several studies suggest that tinal marshes may be an appropriate ecotone in which to search for instances of ecological speciation. The studied species show, as is the case in our study, distinct morphological differences despite little divergence in molecular markers \([7,19-21]\).

In view of the above shown analogy between intra- and interspecific variation, it seems reasonable to assume that the same ecological adaptive bifurcation was also the first step in the speciation process of \(P. chalceus\) and \(P. littoralis\). The speciation process was here fully accomplished by the reproductive isolation between the two groups, allowing independent drift and mutation accumulation in neutral genetic characters.

**Methods**

**Sampling**

\(P. chalceus\) populations from three different sites in the Guérande region are analysed (macroscale; Fig. 5; see also \([2]\)). Each site consists of two drastically differing micro-
Table 4: 459 bp of COI sequenced for 90 individuals of *P. chalceus* and 22 of *P. littoralis*

Haplotype sequence information

| Haplotype No. | 1   | 2   | 3   | 4   | 5   | 6   | 7   | 8   | 9   | 10  | 11  | 12  | 13  | 14  | 15  | 16  | 17  | 18  | 19  | 20  | 21  | 22  | 23  | 24  | 25  | 26  | 27  | 28  | 29  | 30  | 31  | 32  |
|---------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| 1             | A   | T   | A   | C   | T   | C   | A   | A   | A   | T   | T   | T   | T   | T   | T   | C   | G   | T   | A   | A   | A   | C   | T   | A   | A   | T   | T   | C   | A   | C   |
| 2             | A   | T   | A   | C   | T   | C   | A   | A   | A   | T   | T   | T   | T   | T   | T   | C   | G   | T   | A   | C   | A   | A   | A   | C   | T   | A   | A   | T   | T   | C   | A   |
| 3             | A   | T   | A   | C   | T   | C   | A   | A   | A   | T   | T   | T   | T   | T   | T   | C   | G   | T   | A   | T   | A   | A   | A   | T   | T   | A   | A   | T   | T   | C   | A   |
| 4             | A   | T   | C   | C   | T   | C   | A   | A   | A   | T   | T   | T   | T   | T   | T   | C   | G   | T   | A   | T   | A   | A   | A   | T   | T   | A   | A   | T   | T   | C   | A   |
| 5             | A   | A   | A   | T   | A   | T   | T   | T   | T   | C   | C   | C   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   |
| 6             | A   | A   | A   | T   | A   | T   | T   | T   | T   | C   | C   | C   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   |
| 7             | A   | A   | A   | T   | A   | T   | T   | T   | T   | C   | C   | C   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   |
| 8             | G   | A   | A   | T   | A   | T   | T   | T   | T   | C   | C   | C   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   |
| 9             | A   | A   | A   | A   | T   | A   | T   | T   | T   | C   | C   | C   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   | A   |

FREQUENCY OF HAPLOTYPES

| Haplotype No. | CA | PO | ND | M | Z | HEI | OS | NIE | CA | SO | M | S | M | GA | R | CA | AL | B | GU | EI | Z | W | RO | U |
|---------------|----|----|----|---|---|-----|----|-----|----|----|---|---|---|---|---|---|----|---|---|---|---|---|---|---|---|---|---|---|---|---|
| 1             | 8  | 6  | 6  | 7  | 7  | 6  | 2  | 7  |    |    |   |   |   |   |   |   |    |   |   |   |   |   |   |   |   |   |   |   |   |   |
| 2             | 2  | 3  |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
| 3             | 1  | 7  | 5  | 6  | 4  | 1  | 6  |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
| 4             |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
| 5             |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
| 6             |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
| 7             |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
| 8             |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |
| 9             |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |    |

Bold lower case letters show variable positions numbered from 1 to 32 which are not position numbers in the gene. Dots indicate invariable positions. The table shows the variable sites of the haplotypes. The table below shows the haplotype frequency in each population.
Table 5: 497 bp of 16S sequenced for 62 individuals of *P. chalceus* and 15 of *P. littoralis*.

| Haplotype No. | 1   | 2   | 3   | 4   | 5   | 6   | 7   | 8   | 9   | 10  | 11  |
|---------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| 1             | G   | T   | G   | T   | A   | T   | A   | A   | T   | T   |     |
| 2             | G   | T   | G   | T   | A   | T   | A   |     | T   | T   |     |
| 3             | A   | A   | A   | A   | A   | T   | A   | G   | A   | G   |     |

Haplotype sequence information

Frequency of haplotypes

| Haplotype No. | CANALI | POND1 | MOK | ZWC | HEI | OOS | NIE | SOM | MSM | CAMA | ALB | GUE1 | ZWC | ROU |
|---------------|--------|-------|-----|-----|-----|-----|-----|-----|-----|------|-----|------|-----|-----|
| 1             | 6      | 6     | 5   | 6   | 7   | 4   | 5   | 5   | 7   | 5    | 5   |     |     |     |
| 2             |        |       |     |     |     |     |     |     |     |      |     |     |     | 1   |
| 3             |        |       |     |     |     |     |     |     |     |      |     |     |     |     |

Bold lower case letters indicate variable positions numbered from 1 to 11 which are not position numbers in the gene. Dots indicate invariable positions. The table shows the variable sites 1–11 of the haplotypes. The table below shows the haplotype frequency in each population.
habitats, situated only 10–20 metres from each other and separated by one or two dikes. We compare *P. chalceus* populations from three canals (CANAL1; CANAL2; CANAL3; Fig. 5) to three adjacent pond populations (POND1; POND2; POND3; see also [2]). Furthermore, we analyse Guérande *P. littoralis* populations from three different sites (GUE1, GUE2, GUE3; Fig. 5; see also [12]) nearby the aforementioned *P. chalceus* population couples.

The two related species are also studied on a macroscale with completely independent population samples (Guérande populations not included). Data on *P. chalceus* populations from the Netherlands (FRI), Belgium (BRA, WAT, MOK, ZWC, HEI, LIS, OOS, NIE, MOE), England (SEA), France (CAN, AUT, SOM, MSM, VEY, GAC, GIR, TOU, CAM, ROU), and Spain (IBI, ALB, MUJ, ALM; Fig. 5; see also [11,22]. For *P. littoralis*, six populations are analysed here, five of them from France (AUT, MSM, TOU, CAM, ROU; [12]). From these sites in France, we also sampled *P. chalceus* populations (see above). In Belgium, *P. littoralis* is critically endangered and the previous record went back to 1956 and was from Ostend [23]. Recently, a supposed new *P. littoralis* population has been discovered in Belgium and is also included here (Fig. 5; ZWC). Populations of *P. chalceus* on a European scale were assigned...
to belong to one of three different salt marsh area stability types: temporary (BRA, WAT, MOK, HEI, LIS, OOS, MOE, TOU, CAM, ROU, IBI, ALB, MUR, ALM), intermediate (ZWC, NIE, MSM, FRI, AUT) and stable (SEA, CAN, SOM, VEY, GAC, GIR; see also [2,11]. Temporary populations of *P. chalceus* are situated in the Mediterranean part of Europe or occur in small (<4 ha) and young (<400 years) Atlantic salt marshes. Stable and intermediate populations live in larger marshes situated along the Atlantic coast. The only difference between both salt marsh areas is their estimated age (Stable: >1000 years; Intermediate: between 400–1000 years). The age of salt marshes was estimated using historical information [24-27].

### Morphological analysis

Body size (elytral length) and wing size were measured by means of a calibrated ocular under a binocular microscope. Generally, carabid wing size follows an allometric relationship with body size. [28] developed an index that corrects for this allometry, i.e. percentage of maximal realisable relative wing size. Relative wing size is wing length × width divided by elytral length × width. Relative wing size is then expressed as a percentage of the maximal wing size for a beetle of a given size. This index was shown to be an unbiased estimator for comparing different individuals, populations and species of carabid beetles [28]. In ground beetles, females are generally larger than males. Therefore, we analyse male and female body sizes separately. To be complete, we analyse female and male relative wing size also separately. Body size and relative wing size are compared between species and populations with ANOVA's. Total variance is partitioned among groups (species or species and ecotypes), among populations within groups, and within populations by carrying out a nested design ANOVA using STATISTICA (version 7.1; StatSoft Inc., Tulsa, UK) on both a micro- and macroscale.

### Genetic divergence

Data are used from five polymorphic enzymes: aldehyde oxidase (*AO*, E.C. 1.2.3.1), glucose-6-phosphate isomerase (*GPI*, E.C. 5.3.1.9), isocitrate dehydrogenase 1 and 2 (*IDH1*, *IDH2*, E.C. 1.1.1.42), phosphoglucomutase (*PGM*, E.C. 2.7.5.1.). Protocols of electrophoresis are provided by [29]. Earlier work showed that one locus (*IDH1*) was non-neutral and we will always analyse it separately [11].

Departures from Hardy-Weinberg equilibrium expectation were tested with an exact test using the GENEPOP software (Version 3.2; [30]). Significance levels were adjusted by using sequential Bonferroni correction. Similarly as in the analyses for body and wing size, total genetic variance is partitioned among groups (species, ecotypes), among populations within groups, and within populations by carrying out a hierarchical analysis of molecular variance (AMOVA) using ARLEQUIN (version 3.000; [31]) on both a micro- and macroscale.

### Table 6: Summary of analysis of variance for body size, wing size, *IDH1* and allozymes

| region         | source of variation | % var body size male | % var wing size male | % var *IDH1* | % var allo |
|----------------|---------------------|----------------------|----------------------|--------------|------------|
| Guérande       | among groups (P. chalceus vs P. littoralis) | 74.24                | 78.84                | 61.93        | 58.43      |
|                 | among populations within groups              | 17.50                | 18.64                | 11.49        | 3.39       |
|                 | within populations                            | 8.26                 | 2.52                 | 26.57        | 39.18      |
| Europe         | among groups (P. chalceus vs P. littoralis)   | 68.37                | 57.61                | 62.29        | 36.49      |
|                 | among populations within groups              | 10.13                | 36.11                | 12.26        | 13.70      |
|                 | within populations                            | 21.51                | 6.28                 | 25.44        | 49.81      |
| Guérande       | among groups (2 ecotypes vs P. littoralis)    | 84.96                | 95.48                | 64.25        | 41.99      |
|                 | among populations within groups              | 2.35                 | 0.64                 | 0.1          | 4.98       |
|                 | within populations                            | 12.69                | 3.88                 | 35.65        | 53.02      |
| Europe         | among groups (3 ecolog groups vs P. littoralis)| 49.39              | 82.22                | 53.26        | 19.05      |
|                 | among populations within groups              | 5                   | 6.34                 | 2.07         | 15.71      |
|                 | within populations                            | 45.62                | 11.45                | 44.67        | 65.24      |

PCRs for nucleotide sequencing of COI utilized primers C1-J-1718 and C1-N-2191 and for 16S we utilized primers LR-J-1307 and LR-N-13398 [32]. DNA amplification reactions were performed in a 25 µL final volume. Each reaction mix contained 5 µL of extract, 1× buffer (Sigma), 1.5 mM MgCl₂, 200 µM of each dNTP, 0.4 µM of each primer and 0.6 U RedTaq DNA polymerase (Sigma). Initial denaturation was for 2 min at 95°C, followed by 35 cycles of 1 min at 95°C, 1 min 30 s at 48°C (and 46°C for 16S), and 2 min at 72°C; 9 min at 72°C completed the program. The reaction was purified with columns following manufacturer's recommendations. Sequencing was done by BigDye Terminator version 3.1 kits on an ABI 3130 sequencer (Applied Biosystems). Sequences were aligned using BioEdit version 5.0.6 [33]. We tested for
neutrality of mutations following Fu & Li's method with $D^*$ and $F^*$ test statistics using DNASP 4.0 [34,35]. A phylogeny of unique haplotypes was constructed from the calculated Kimura two-parameter distances using the neighbour-joining approach within MEGA ([36]; 1000 bootstrap replicates).

**Competing interests**

The author(s) declare that they have no competing interests.

**Authors’ contributions**

HD collected the majority of the data, and carried out most of the calculations and drafted the text. Y-PM assisted the field work, advised regarding ANOVA’s and final edited the text. KD supplied historical information, assisted the field work, advised regarding ANOVA’s and final edited the text. All three authors read and approved the final text.

**Additional material**

**Additional file 1**
Number of males and females for which body size and wing size is measured and number of individuals used for IDH1 allozyme electrophoresis in the Guérande populations. Click here for file

[http://www.biomedcentral.com/content-supplementary/1746-1448-3-4-S1.doc](http://www.biomedcentral.com/content-supplementary/1746-1448-3-4-S1.doc)

**Additional file 2**
Number of males and females for which body size and wing size is measured and number of individuals used for IDH1 allozyme electrophoresis in the different populations on a European scale. Click here for file

[http://www.biomedcentral.com/content-supplementary/1746-1448-3-4-S2.doc](http://www.biomedcentral.com/content-supplementary/1746-1448-3-4-S2.doc)

**Additional file 3**
Allele frequencies from four allozymes (AO, IDH2, PGI, PGM) studied in the Guérande populations. N: number of studied individuals. Click here for file

[http://www.biomedcentral.com/content-supplementary/1746-1448-3-4-S3.doc](http://www.biomedcentral.com/content-supplementary/1746-1448-3-4-S3.doc)

**Additional file 4**
Allele frequencies from four allozymes (AO, IDH2, PGI, PGM) studied in the European populations. N: number of studied individuals. Click here for file

[http://www.biomedcentral.com/content-supplementary/1746-1448-3-4-S4.doc](http://www.biomedcentral.com/content-supplementary/1746-1448-3-4-S4.doc)

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