Dynamics of DNA Methylation Reprogramming Influenced by X Chromosome Dosage in Induced Pluripotent Stem Cells

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ABSTRACT: How the epigenome of one cell type is remodeled during reprogramming into another unrelated type of cell remains unclear. Overexpression of transcription factors in somatic cells enables the induction of induced pluripotent stem cells (iPSCs). This process entails genome-wide remodeling of DNA methylation, chromatin, and transcription. Recent work suggests that the number of active X chromosomes present in a cell influences remodeling of DNA methylation during somatic cell reprogramming to mouse iPSCs. Female iPSCs with 2 active X chromosomes display global DNA hypomethylation, whereas male XY iPSCs show DNA methylation levels similar to the somatic cells they are derived from. Global DNA methylation erasure in female iPSCs takes place genome-wide and involves repression of DNA methyltransferases. However, on loss of one X chromosome, female iPSCs acquire a DNA methylation landscape resembling that of XY iPSCs. Therefore, it is the X chromosome dosage that dictates global DNA methylation levels in iPSCs. Here, we discuss the evidence that links X chromosome dosage with the regulation of DNA methylation in pluripotent stem cells. We focus on iPSCs reprogramming studies, where X chromosome status is a novel factor impacting our understanding of epigenetic remodeling.

KEYWORDS: Induced pluripotent stem cells (iPSCs), X chromosome inactivation, DNA methylation, reprogramming, epigenetics

Introduction

A fundamental discovery in biology is that differentiated cells can be reprogrammed into induced pluripotent stem cells (iPSCs) by transcription factor overexpression. Induced pluripotent stem cells resemble embryonic stem cells (ESCs) derived from the inner cell mass (ICM) of preimplantation embryos. They possess the capacity for indefinite self-renewal and differentiation into any cell type of the body. Therefore, reprogramming to iPSCs is a promising technology for regenerative medicine and a powerful tool for studying cellular identity. In a stepwise manner, reprogramming to iPSCs involves silencing of somatic genes, followed by sequential activation of pluripotency genes. Such rewiring of transcription involves remodeling of the epigenome.

The specialized identity of somatic cells is maintained by a variety of epigenetic mechanisms that ensure its stability. For reprogramming to succeed, these mechanisms must be reversed. This is an important point because failure to properly reset the epigenome has been associated with incomplete and aberrant reprogramming. DNA methylation is one example of modification associated with epigenetic mechanisms, which is remodeled during reprogramming. However, when and how DNA methylation is reset during transcription factor–induced pluripotency remains poorly understood.

Global and Sex-Specific Changes in DNA Methylation During Reprogramming to iPSCs

Efforts over the past decade have shed light onto the dynamics and the molecular mechanisms underlying DNA methylation reprogramming. Previous studies suggested that DNA methylation changes occur late during reprogramming. The iPSCs have also been shown to tolerate global DNA hypomethylation. Inhibition of the DNA methyltransferase 1, DNMT1, enhances the efficiency of iPSCs' derivation, suggesting that DNA methylation acts as a barrier to reprogramming.

Although the sex of cells is often ignored in reprogramming studies, it was recently revealed to strongly influence the resetting of global DNA methylation in iPSCs. The iPSCs derived from male cells show levels of global DNA methylation comparable with somatic cells, such that the global level of DNA methylation does not change dramatically during reprogramming. However, when female cells are reprogrammed, the resulting iPSCs display global erasure of DNA methylation (Figure 1). The global hypomethylation takes place after the intermediate reprogramming stage marked by SSEA1. In support of sex-specific DNA hypomethylation in iPSCs, studies in ESCs have reported female-specific DNA hypomethylation. In summary, changes in global DNA methylation level are sex specific and take place late in reprogramming to iPSCs.
Global DNA Hypomethylation in iPSCs Correlates With the Presence of Two Active X Chromosomes

Reprogramming somatic cells to pluripotency induces reaction of the silent X chromosome (XCR) in female cells. Therefore, early passage female iPSCs have 2 active X chromosomes, whereas male iPSCs have only 1 active X chromosome. This skewed ratio of X-to-autosome gene expression (X dosage) underlies DNA hypomethylation in female iPSCs. This is supported by experiments where on high passage, female iPSCs lose 1 of the 2 X chromosomes and as a result, global DNA methylation reverts to levels similar to XY iPSCs (Figure 1). Therefore, X dosage dictates global methylation levels in iPSCs.

Mechanistically, it is likely that, as a result of XCR in female iPSCs, the expression of X-linked gene Dusp9 increases and drives global hypomethylation in female iPSCs. This is supported by experiments where on high passage, female iPSCs lose 1 of the 2 X chromosomes and as a result, global DNA methylation reverts to levels similar to XY iPSCs (Figure 1). Therefore, X dosage dictates global methylation levels in iPSCs.

Influence of X Dosage on DNA Methylation of Key Regulatory Regions Associated With the Control of Cell Identity

Comprehensive genome-wide DNA methylation analyses have revealed that the dynamics of DNA methylation during reprogramming depend on the type of genetic elements considered and their location in the genome.

Imprinting is an epigenetic mechanism in mammals, required for proper development, in which differential expression of the maternal and paternal alleles of certain genes has been attributed to DNA methylation. The global erasure of DNA methylation in female iPSCs leads to the loss of this mark at imprint control regions (Figure 1). Moreover, these imprints are not reestablished after genome remethylation in XO female iPSCs. Thus, increased X dosage induces irreversible changes in DNA methylation associated with key mammalian epigenetic mechanisms.

Transcriptional programs are controlled by a set of key regulatory regions that include enhancers and promoters. Although the expression of somatic genes is downregulated early during reprogramming, somatic enhancers remethylation is initiated at intermediate reprogramming stages and completed only late during reprogramming in male iPSCs. In addition, the early wave of focal DNA methylation at ESC enhancers is initiated early during reprogramming, independent of sex, before global DNA methylation erasure in female iPSCs. Thus, increased X dosage induces irreversible changes in DNA methylation associated with key mammalian epigenetic mechanisms.

Repetitive elements form a large portion of mammalian genomes. Processes such as DNA methylation and repressive histone modifications help to maintain repression of these elements, including potentially mobile transposable elements. Interestingly, several classes of repetitive elements such as LINEs, SINEs, and LTRs are demethylated in XX iPSCs, but not in XY iPSCs (Figure 1), indicating that also these elements are sensitive to X dosage in pluripotent stem cells.

The overall picture that emerges is that distinct regulatory elements are dynamically methylated and unmethylated during reprogramming depending on their usage, with global erasure of DNA methylation as a result of increased X dosage in female iPSCs affecting most genomic regions tested.

Implications for Studying Reprogramming

These findings bring a set of important questions. Whether the pattern of DNA methylation in iPSCs reflects the state of
pluripotent cells in the embryo and irrespective of that, what are the consequences of sex-specific DNA methylation for reprogramming studies and conclusions drawn from them?

X chromosome inactivation is a developmentally regulated process used in mammals to mediate gene dosage compensation between XX and XY cells. Early in development, pluripotent cells of the female ICM transiently acquire 2 active X chromosomes. However, it does not lead to clear differences in DNA methylation between female and male ICMs. Hence, we speculate that global DNA hypomethylation in iPSCs results from the long-term maintenance of what normally is a transient developmental state in the ICM.

To study DNA methylation in reprogramming, it is imperative to consider the sex of the cells used. For example, using only male cells would bypass female-specific global DNA demethylation in the pluripotent state. Hence, DNA methylation changes seen would mistakenly be attributed to reprogramming in general rather than X dosage effects. Conversely, pooling male and female cells in unknown ratios masks DNA methylation heterogeneity. Likewise, the sex of control cells should match the experimental ones, to avoid biased interpretations. Moreover, comparing male and female cells is not sufficient. One must also define the number of active X chromosomes, as, strictly speaking, it is X dosage that is important in female iPSCs for dictating global DNA methylation, and not the sex in its entirety. Therefore, comparing male and female iPSCs requires the knowledge of the number of active X chromosomes. Recent work by our group also shows that X dosage modulates the transcriptional and open chromatin landscape, as well as growth properties, of iPSCs. Altogether, these studies link sex chromosomes with DNA methylation and pluripotency and as a result bring novel, nonnegligible considerations to studying and understanding cell fate reprogramming.

Implications for Human Pluripotency

Are X dosage–dependent DNA methylation dynamics also present in human ESCs and iPSCs? Sex-specific differences in gene expression, but not in the global level of DNA methylation, have been reported in conventional primed human pluripotent stem cells. Currently, however, human naive pluripotent stem cells are thought to reside in an incomplete naive pluripotent state and have reduced X-linked gene expression compared with female cells of the early preimplantation embryo. We speculate that naive human pluripotent stem cells cultured in conditions that would properly induce X dosage increase may show additional sex-specific molecular and functional differences. With the emergence of improved culture conditions to induce and maintain human naive pluripotency, it will be interesting to assess potential X dosage effects.

Concluding Remarks

DNA methylation is a paradigmatic mark for the study of epigenetic processes. X dosage can have important effects on DNA methylation in iPSCs. Understanding the mechanisms by which DNA methylation is established and maintained is important to gain insights into health and diseases. Somatic cell reprogramming to iPSCs provides a useful system to study the reversal of epigenetic memory and the molecular mechanisms involved in cell state transitions.

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Author Contributions

AJ and VP wrote the manuscript with input from all authors.

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