Oestrogen receptor-negative/progesterone receptor-positive phenotype of invasive breast carcinoma in Japan: re-evaluated using immunohistochemical staining

Hajime Kuroda¹ · Nozomi Muroi² · Mitsuhiro Hayashi³ · Oi Harada⁴ · Kazuei Hoshi⁴ · Eisuke Fukuma⁵ · Akihito Abe⁶ · Keiichi Kubota⁵ · Yasuo Imai¹

Received: 5 April 2018 / Accepted: 26 July 2018 / Published online: 31 July 2018
© The Author(s) 2018

Abstract

Background The existence of progesterone receptor (PgR) expression in oestrogen receptor (ER)-negative breast carcinoma is controversial. Here, we re-evaluated ER-negative/PgR-positive (ER−/PgR+) carcinoma cases by immunohistochemical staining (IHC).

Materials and methods We selected patients who underwent surgery for primary breast carcinoma from our databases at Dokkyo Medical University Hospital and Kameda General Hospital. Among the 9844 patients, the largest series in Japan, 27 (0.3%) were initially diagnosed as ER−/PgR+ breast carcinomas and we re-evaluated by IHC.

Results The re-evaluated IHC showed that of the 27 patients with the initial results of ER−/PgR+, 12 were ER+/PgR+, 8 were ER−/PgR+, and 7 were ER−/PgR−. ER was negative in 12 of 27 patients (44.4%), and PgR was positive in 8 of 27 patients (29.6%). In our seven re-evaluated and confirmed as ER−/PgR+ cases, the staining proportions of tumor cells were 0% in ER and 1–69% (average 15.8%) in PgR. The average staining proportion of PgR in the re-evaluated ER−/PgR+ phenotype was lower than the initial diagnosis. Histological grading was as follows: grade I, one case; grade II, two cases; grade III, four cases. There were two lymph-node-positive cases.

Conclusions The ER−/PgR+ phenotype was confirmed after re-evaluation of ER and PgR assessment by a different pathologist. We recommend that pathologists discuss with clinicians, or re-test and re-evaluate ER/PgR expression, particularly in low-grade carcinoma and with a high staining proportion of PgR in the ER−/PgR+ phenotype.

Keywords Oestrogen receptor · Progesterone receptor · Breast carcinoma · Immunohistochemical staining · Re-evaluation

Introduction

Steroid hormone receptors were shown to be prognostic and predictive markers for breast carcinoma endocrine therapy [1–11]. It is generally suggested that all pathological diagnosed primary breast carcinomas be examined for oestrogen receptor (ER) and progesterone receptor (PgR) protein expression by immunohistochemical staining (IHC). However, the existence of PgR expression in ER-negative breast carcinoma is controversial [2–8]. PgR is a downstream relative of ER, and is regulated by ER, which binds to the oestrogen-responsive element (ERE) located in the promoter region of the PgR gene [12]. Therefore, researchers suggested that ER−/PgR+ is an erroneous result due to a technical artifact, and others reported that such cases are too rare to consider as a true phenotype for ER and PgR assessments in IHC-based methodology [2–4]. However, it is also reported
that downregulation of PgR was not mediated by a reduction in ER levels or ER activity, suggesting that regulation of PgR is independent of ER under some conditions [13]. Thus, other studies suggested that ER−/PgR+ breast carcinomas show definite clinical and biological features [5–8]. Herein, we re-evaluated all the ER−/PgR+ carcinoma cases which we have encountered according to the standard IHC methods. The aim of our study was to determine the existence of the ER−/PgR+ breast carcinoma phenotype and, if found, to elucidate its clinicopathological features and discuss management.

Methods

Immunohistochemistry

We selected patients who underwent surgery for primary breast carcinoma following ER/PgR IHC evaluations from our databases at Dokkyo Medical University Hospital and Kameda General Hospital. Surgical and biopsy specimens were fixed in 10% buffered neutral formalin solution for IHC at our hospitals. The immunohistochemical procedures used for the initial staining at Dokkyo Medical University Hospital and Kameda Medical Center Hospital were as follows: in Kameda Medical Center Hospital, the sections were taken to an automated stainer (DAKO, AUTOSTAINER) following the manufacturer’s instructions before 2012. The sections were taken to an automated stainer (VENTANA, BENCHMARK XT) from 2012 to 2017. The evaluated IHC assays were ER (clone 1D5, Dako, 1:50, nuclear), and PgR (clone PgR636, 1:800, nuclear) before 2012, and ER (clone SP1, Ventana, prediluted, nuclear), and PgR (clone 1E2, prediluted, nuclear) from 2012 to 2017. In Dokkyo Medical University Hospital, the evaluated IHC assays were ER (clone 1D5, Dako, 1:100, nuclear), and PgR (clone PgR636, 1:200, nuclear) before 2006. The sections were then taken to an automated stainer (VENTANA, BENCHMARK XT) following the manufacturer’s instructions from 2006 to 2017. The evaluated IHC assays were ER (clone 6F11, Ventana, prediluted, nuclear), and PgR (clone 16, prediluted, nuclear) from 2006 to 2009 and ER (clone SP1, Ventana, prediluted, nuclear), and PgR (clone 1E2, prediluted, nuclear) from 2006 to 2009. Details of the initial cut-off points for ER and PgR are unknown for both hospitals.

Among the 9844 patients, 27 (0.3%) were initially diagnosed as ER−/PgR+ breast carcinomas. The clinical history, pathological reports, and haematoxylin and eosin (H&E) slides from all 27 patients were reviewed. ER−/PgR+ carcinomas were re-evaluated by nuclear staining of ER and PgR immunohistochemically at the Department of Diagnostic Pathology of Dokkyo Medical University. In each case, the same block was selected for the IHC re-evaluation. The paraﬃn-embedded tissue block was recut into 5 μm sections. The re-evaluated IHC assays for ER (clone 6F11, Novocastra, 1:40, nuclear) and PgR (clone 16, Novocastra, 1:100, nuclear) were performed by additional H&E staining. The slides were treated with methanol containing 0.3% hydrogen peroxide to block endogenous peroxidase activity. Antigen retrieval was achieved with microwave treatment for markers. After incubation with the primary antibody, incubation with a secondary biotinylated antibody was performed for 15 min. After washing, the sections were incubated with streptavidin–peroxidase for 20 min. Finally, the enzyme was visualized after a 5-min incubation with diaminobenzidine. Counterstaining was performed with haematoxylin. The methodology was the same in ER and PgR. The re-evaluated ER, PgR, and H&E slides (Fig. 1) were reviewed by two pathologists independently. To increase reproducibility, ER and PgR positivity was defined as 1% nuclear staining of tumor cells (Fig. 2a, b) [14, 15]. As HER2-positive breast carcinomas appear as a biologically distinct phenotype, we excluded HER2-positive cases from this study. The Chi-square test was used to assess associations among the variables, and the Mann–Whitney test was used to compare the means of clinicopathological data. The associations of ER and PgR expression were analysed using Chi-square test and Fisher’s exact test to assess whether there were significant differences in expression. Differences were considered significant when \( P \) was less than 0.05.

Results

The results of clinicopathological and re-evaluated IHC for ER and PgR are summarized in Table 1. The clinicopathological findings of the 27 patients with the initial results of ER−/PgR+ were as follows: the patients’ ages ranged from...
39 to 73 years (mean age 55.8 years), and tumor size ranged from 0.1 to 11.7 cm (mean size 1.8 cm). Histologic types were 25 invasive carcinomas of no special type (NST), and 2 other cases. Histological grading was as follows: grade I, 12 cases; grade II, 6 cases; grade III, 9 cases. There were 7 lymph-node-positive cases. Tumor staging was as follows: stage I, 16 cases; stage II, 8 cases; stage III, 3 cases. The carcinomas recurred in four cases within the follow-up period of 60 months. These included one case of local recurrence, and three cases of distant metastasis (lymph node, bone, lung, and liver). Two patients died of disease. The re-evaluated IHC showed that of the 27 patients with the initial results of ER−/PgR+, 12 were ER+/PgR+, 8 were ER−/PgR−, and 7 were ER−/PgR+. ER was negative in

**Table 1** Clinicopathologic findings of 27 patients with an initial ER−/PgR+ diagnosis

|                        | A. Initial diagnosis | B. Different from initial diagnosis | C. Confirmed initial diagnosis | B vs C | P value |
|------------------------|---------------------|------------------------------------|-------------------------------|--------|---------|
| **Mean age**           | ER−/PgR+ (27 cases) | ER+/PgR+ (12 cases)                | ER−/PgR− (8 cases)            | ER−/PgR+ (7 cases) |         |
|                        | 55.8 (39–79)        | 47.8 (39–64)                      | 65.1 (49–79)                 | 56.9 (42–72)       | 0.9679  |
| **Mean tumor size (cm)** | 1.8 (0.1–11.7)     | 1.0 (0.1–2.3)                     | 2.9 (0.3–11.7)               | 1.7 (0.5–3.5)      | 0.9993  |
| **Histology**          | NST 25 (92.6%)      | NST 12 (100.0%)                   | NST 6 (75.0%)                | NST 7 (100.0%)     | 0.3846  |
|                        | Others 2 (7.4%)     | Others 0 (0%)                     | Others 2(25.0%)              | Others 0 (0%)      |         |
| **Histological grading** |                      |                                    |                              |                    |         |
| I                      | 12 (44.4%)          | 9 (75.0%)                         | 2 (25.0%)                    | 1 (14.3%)          | 0.2800  |
| II                     | 6 (22.3%)           | 2 (16.7%)                         | 2 (25.0%)                    | 2 (28.6%)          |         |
| III                    | 9 (33.3%)           | 1 (8.3%)                          | 4 (50.0%)                    | 4 (57.1%)          |         |
| **Status of nodal metastasis** |                |                                    |                              |                    |         |
| I                      | 16 (59.3%)          | 8 (66.6%)                         | 5 (62.5%)                    | 3 (42.9%)          | 0.2500  |
| II                     | 8 (29.6%)           | 2 (16.7%)                         | 2 (25.0%)                    | 4 (57.1%)          |         |
| III                    | 3 (11.1%)           | 2 (16.7%)                         | 1 (12.5%)                    | 0 (0%)             |         |
| IV                     | 0 (0%)              | 0 (0%)                            | 0 (0%)                       | 0 (0%)             |         |
| 5 year follow up data available cases | |                                    |                              |                    |         |
| I                      | 26 of 27            | 11 of 12                          | 8 of 8                       | 7 of 7             |         |
| II                     | 8 (3.7%)            | 0 (0%)                            | 0 (0%)                       | 1 (14.3%)          | 0.0929  |
| III                    | 3 (11.1%)           | 2 (16.6%)                         | 0 (0%)                       | 1 (14.3%)          | 0.7901  |
| IV                     | 2 (7.4%)            | 1 (8.3%)                          | 0 (0%)                       | 1 (14.3%)          | 0.4438  |
| Re-evaluated proportion of positive cells | |                                    |                              |                    |         |
| ER                     | 19.3%               | 28.8%                             | 52.4%                        | 66.9%              | 0%      | 15.8%   |
| PgR                    |                      |                                   |                              |                    |         |

*ER* oestrogen receptor, *PgR* progesterone receptor, *NST* invasive carcinoma of no special type

**Fig. 2** a Oestrogen receptor (ER)-negative and b progesterone receptor (PgR)-positive breast carcinoma (immunohistochemical staining, ×100)
12 of 27 patients (44.4%), and PgR was positive in 8 of 27 patients (29.6%). In our seven re-evaluated and confirmed as ER−/PgR+ cases, the staining proportions of tumor cells were 0% in ER and 1–69% (average 15.8%) in PgR. The average staining proportion of PgR in re-evaluated ER−/PgR+ phenotype was lower than the initial diagnosis.

The clinicopathological findings of the seven patients with re-evaluated results of the ER−/PgR+ phenotype were as follows. The patients’ ages ranged from 42 to 72 years (mean age 56.9 years). Tumor size ranged from 0.5 to 3.5 cm (mean size 1.7 cm). Histological grading was as follows: grade I, one case; grade II, two cases; grade III, four cases. There were two lymph-node-positive cases. Tumor staging was as follows: stage I, three cases; stage II, four cases. Carcinomas recurred in two cases within the follow-up period of 60 months, including one case of local recurrence and one case of distant metastasis (bone). One patient died of disease.

**Discussion**

There are reports, suggesting that the ER−/PgR+ phenotype does not exist and may be a technical artifact [2–4]. Nadji et al. evaluated a large series of 5993 breast carcinomas for ER expression by IHC analysis and found that the ER−/PgR+ phenotype did not exist [2]. Furthermore, several studies suggested that there was no ER−/PgR+ phenotype using re-evaluated IHC (Table 2). De Maeyer et al. found that none of their 32 cases initially considered to be the ER−/PgR+ phenotype were found to be so following a re-evaluated IHC test [3]. Maleki et al. also found that none of their 43 cases initially diagnosed as ER−/PgR+ phenotype showed the same IHC phenotype upon re-evaluation [4]. Ahmed et al. investigated a large number of ER−/PgR+ phenotype samples by re-evaluating IHC using a tissue microarray (TMA) [14]. In contrast to the previous studies, 92 of 267 cases were confirmed as ER−/PgR+. One limitation of this study is that they used the TMA method. The tumor heterogeneity, the data from TMA analysis may not be identical to the results from whole sections.

These differences in the previous articles may be due to the diagnostic criteria or the methods used for preparing the IHC. The recommended cut-off points of IHC have changed over several decades. Ogawa et al. suggested a 10% staining proportion may be an acceptable cut-off point for both ER and PR status by IHC [17]. However, among the cases initially scored as negative for ER and PgR, the proportion of positive cells in our re-evaluation was 0 or nearly 0%. Furthermore, an important issue is selection of the most reliable antibody. In some of our initial cases, monoclonal anti-ER 1D5 was used for immunohistochemical assessment of ER. Bogina et al. demonstrated the higher sensitivity of anti-ER SP1 and 6F11 clones compared with the 1D5 clone [18]. This may be the cause of the discrepancy in the ER results.

However, we must start from the reality that the methods of preparing IHC specimens differ in each institution and cannot be unified. Thus, it is wiser to accept the existence of the ER−/PgR+ phenotype, and then discuss management. Furthermore, in our study, the ER−/PgR+ phenotype was confirmed after the re-evaluation of ER and PgR by different pathologists. Therefore, regardless of the diagnostic criteria or the process for preparing specimens, we may find ER−/PgR+ breast cancer cases in our routine practice. Moreover, there were several reports showing the gene-expression profile of ER−/PgR+ breast carcinomas [19, 20]. Itoh et al. reported that 25% of cases showed the same IHC phenotype after re-evaluation by gene-expression profiling [19]. Taken together, these results provide further evidence that some ER−/PgR+ cases do not reflect a technical artifact, but are a distinct group of breast carcinomas.

As the ER−/PgR+ phenotype is extremely rare and mostly non-reproducible, the majority of cases classified as ER−/PgR+ may appear as different classifications [14]. The difference between the results of the initial and the re-evaluated IHC raises a concern about possible errors in pathological diagnosis. In our study, most of the initially evaluated ER−/PgR+ cases were ER+/PgR+ low-grade carcinomas after re-evaluation by IHC. Furthermore, in our results, PgR was negative in 8 out of 27 patients (29.6%), which differed from the initial diagnosis. Moreover, in the confirmed ER−/PgR+ cases, the staining proportion of PgR+ was low. Therefore, we recommend that

### Table 2: Articles on ER−/PgR+ breast cancer

| Article            | Year | No.    | ER−/PgR+ cases in medical record | Re-evaluated ER−/PgR+ cases | Method        |
|--------------------|------|--------|---------------------------------|----------------------------|---------------|
| De Maeyer et al. [3] | 2007 | 2013   | 32                              | 0 (0%)                     | IHC           |
| Maleki et al. [4]   | 2012 | 2432   | 43                              | 0 (0%)                     | IHC           |
| Ahmed et al. [14]   | 2016 | 8315   | 267                             | 92 (34.6%)                 | IHC (TMA)     |
| Current             | 2017 | 9844   | 27                              | 7 (25.9%)                  | IHC           |

ER oestrogen receptor, PgR progesterone receptor, IHC immunohistochemical staining, TMA tissue microarray
pathologists discuss with clinicians, or re-test and re-evaluate ER/PgR expression, particularly under the following conditions. (1) Histological grade is low. (2) PgR-positive proportion is high.

Ng et al. reported that the ER−/PgR+ phenotype had no distinct histopathological characteristics compared to the ER+/PgR+ and ER−/PgR− phenotypes and no prognostic impact [7]. In contrast, Rakha et al. reported poorer survival of ER−/PgR+ cases compared with ER+/PgR+ cases [5]. Furthermore, Purdie et al. reported that PgR expression is an independent prognostic variable more powerful than ER [21]. PgR expression in breast carcinoma may potentially define a distinct subgroup with paired function in the ER pathway that will probably benefit from endocrine therapy [2–4]. However, we could not draw any conclusions regarding histological characteristics or prognostic impact from the current study due to the small number of cases. It is natural that there is a difference in prognosis outcomes among the previous papers. This is because not only is the ER−/PgR+ phenotype extremely rare, but also there are no unified diagnostic criteria and specimen preparation methods as described above. The ER−/PgR+ phenotype can result in difficulties in examining biological behavior and deciding on an appropriate treatment strategy.

In conclusion, among the 9844 patients, the largest series in Japan, 27 (0.3%) were initially diagnosed as ER−/PgR+ breast carcinomas, and the ER−/PgR+ phenotype was still present after the re-evaluation of ER and PgR using whole sections recut from the same tissue block. However, 20 of 27 patients with the initial results of ER−/PgR+ differed from the initial diagnosis. We recommend that pathologists discuss with clinicians, or re-test and re-evaluate the ER/PgR expression, particularly in low-grade carcinoma and with a high staining proportion of PgR in the ER−/PgR+ phenotype.

Acknowledgements The authors thank Masaru Kojima, Atsuko Takada, Yoshimasa Nakazato, Yuko Kaneko, Shuhei Noda, and Ken-suke Ohikata for their advice. The authors thank Chiaki Matsuyama and Ayako Shimizu for their advice and technical assistance with immunohistochemical staining.

Funding This study was supported in part by JSPS KAKENHI Grant number JP16K08695 from the Ministry of Education, Science, Sports, and Culture of Japan.

Compliance with ethical standards

Ethical approval Ethical Committee of the Dokkyo Medical University, Hospital, Tochigi, Japan (No. 28126) and Kameda General Hospital, Chiba, Japan (No. 16-160). For this type of study, formal consent is not required.

Conflict of interest The authors declare no potential conflicts of interest.

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

1. Hammond ME, Hayes DF, Wolff AC. Clinical notice for American Society of Clinical Oncology-College of American Pathologists guideline recommendations on ER/PgR and HER2 testing in breast cancer. J Clin Oncol. 2011;29:e458. https://doi.org/10.1200/JCO.2011.35.2245.

2. Nadji M, Gomez-Fernandez C, Ganjee-Azar P, Morales AR. Immunohistochemistry of estrogen and progesterone receptors reconsidered: experience with 5,993 breast cancers. Am J Clin Pathol. 2005;123:21–7. https://doi.org/10.1309/4WV79N2GHJ3X1841.

3. De Maeyer L, Van Limbergen E, De Nys K, Moerman P, Pochet N, Hendrickx W, et al. Does estrogen receptor-negative/progesterone receptor-positive breast carcinoma exist? J Clin Oncol. 2008;26:335–6. https://doi.org/10.1200/JCO.2007.14.8411.

4. Maleki Z, Shariat S, Moki A, Atri M, ER-negative /PR-positive breast carcinomas or technical artifacts in immunohistochemistry? Arch Iran Med. 2012;15:366–9. https://doi.org/10.1016/j. archim.2018.05.005.

5. Rakha EA, El-Sayed ME, Green AR, Paish EC, Powe DG, Gee J, et al. Biologic and clinical characteristics of breast cancer with single hormone receptor positive phenotype. J Clin Oncol. 2007;25:4772–8.

6. Rhodes A, Jasani B. The oestrogen receptor-negative/progesterone receptor-positive (ER−/PR+) breast tumour: a biological entity or a technical artefact? J Clin Pathol. 2009;62:95–6. https://doi.org/10.1136/jcp.2008.060723.

7. Ng CH, Pathy NB, Taib NA, Ho GF, Mun KS, Rhodes A, et al. Do clinical features and survival of single hormone receptor positive breast cancers differ from double hormone receptor positive breast cancers? Asian Pac J Cancer Prev. 2014;15:7959–64.

8. Yu KD, Di GH, Wu J, Lu JS, Shen KW, Liu GY, Shen ZZ, Shao ZM. Breast cancer patients with estrogen receptor-negative/progesterone receptor-positive tumors: being younger and getting less benefit from adjuvant tamoxifen treatment. J Cancer Res Clin Oncol. 2008;134:1347–54.

9. Osborne CK, Yochmowitz MG, Knight WA 3rd, McGuire WL. The value of estrogen and progesterone receptors in the treatment of breast cancer. Cancer. 1980;46:2884–8.

10. Ravdin PM, Green S, Dorr TM, McGuire L, Fabian C, Pugh RP, et al. Prognostic significance of progesterone receptor levels in estrogen receptor-positive patients with metastatic breast cancer treated with tamoxifen: results of a prospective Southwest Oncology Group study. J Clin Oncol. 1992;10:1284–91.

11. Bardou VJ, Arpino G, Elledge RM, Osborne CK, Clark GM. Progesterone receptor status significantly improves outcome prediction over estrogen receptor status alone for adjuvant endocrine therapy in two large breast cancer databases. J Clin Oncol. 2003;21:1973–9.

12. Yu KD, Jiang YZ, Hao S, Shao ZM. Molecular essence and endocrine responsiveness of estrogen receptor-negative, progesterone receptor-positive, and HER2-negative breast cancer. BMC Med. 2015;13:254. https://doi.org/10.1186/s12916-015-0496-z.

13. Cui X, Schif R, Arpino G, Osborne CK, Lee AV. Biology of progesterone receptor loss in breast cancer and its implications for endocrine therapy. J Clin Oncol. 2005;23:7721–35.
14. Hammond ME, Hayes DF, Dowsett M, Allred DC, Hagerty KL, Badve S, et al. American Society of Clinical Oncology/College of American Pathologists guideline recommendations for immunohistochemical testing of estrogen and progesterone receptors in breast cancer. Arch Pathol Lab Med. 2010;134:907–22. https://doi.org/10.1043/1543-2165-134.6.907.
15. Hammond ME, Hayes DF, Dowsett M, Allred DC, Hagerty KL, Badve S, et al. American Society of Clinical Oncology/College of American Pathologists guideline recommendations for immunohistochemical testing of estrogen and progesterone receptors in breast cancer. J Clin Oncol. 2010;28:2784–95. https://doi.org/10.1200/JCO.2009.25.6529.
16. Ahmed SS, Thike AA, Zhang K, Lim JC, Tan PH. Clinicopathological characteristics of oestrogen receptor negative, progesterone receptor positive breast cancers: re-evaluating subsets within this group. J Clin Pathol. 2017;70:320–6. https://doi.org/10.1136/jclinpath-2016-203847.
17. Ogawa Y, Moriya T, Kato Y, Oguma M, Ikeda K, Takashima T, et al. Immunohistochemical assessment for estrogen receptor and progesterone receptor status in breast cancer: analysis for a cut-off point as the predictor for endocrine therapy. Breast Cancer. 2004;11:267–75.
18. Bogina G, Zamboni G, Sapino A, Bortesi L, Marconi M, Lunardi G, et al. Comparison of anti-estrogen receptor antibodies SP1, 6F11, and 1D5 in breast cancer: lower 1D5 sensitivity but questionable clinical implications. Am J Clin Pathol. 2012;138:697–702. https://doi.org/10.1093/ajcpaqj0027.
19. Itoh M, Iwamoto T, Matsuoka J, Nomoto T, Motoki T, Shien T, et al. Estrogen receptor (ER) mRNA expression and molecular subtype distribution in ER-negative/progesterone receptor-positive breast cancers. Breast Cancer Res Treat. 2014;143:403–9. https://doi.org/10.1007/s10549-013-2763-z.
20. Hefti MM, Hu R, Knoblauch NW, Collins LC, Haibe-Kains B, Tamimi RM, et al. Estrogen receptor negative/progesterone receptor positive breast cancer is not a reproducible subtype. Breast Cancer Res. 2013;15:R68.
21. Purdie CA, Quinlan P, Jordan LB, Ashfield A, Ogston S, Dewar JA, Thompson AM. Progesterone receptor expression is an independent prognostic variable in early breast cancer: a population-based study. Br J Cancer. 2014;110:565–72. https://doi.org/10.1186/bcr3462.