New mechanisms of antiplatelet activity of nifedipine, an L-type calcium channel blocker

Tz-Chong Chou

1. Introduction

Platelets are unucleated fragments derived from bone marrow megakaryocytes. Traditionally, the most well-known function of platelets is that they are responsible for hemostasis in response to vascular injury and endothelial disruption. Recent studies have indicated that platelets also have an immunomodulatory activity through production of several pro-inflammatory mediators promoting pathogenic thrombi formation and inflammatory responses [1, 2]. Platelets perform their functions mainly through secretion of several proteins stored in various cytoplasmic granules. There are at least three different types of granules (α-granules, dense core granules, lysosomes), and a complex membranous system in platelets. The α-granules contain hemostatic factors (factor V, von Willebrand factor (vWF) and fibrinogen) and other cytokines, mitogenic factors (PDGF and bFGF) and proteases (MMP2, MMP9) [3]. The mediators stored in α-granules can be selectively released in response to the activation of different receptors. Dense granules store small non-protein molecules such as ADP, ATP, serotonin, calcium and pyrophosphate, which all play a central role in the amplification of platelet aggregation. Lysosomes contain glycosidases, proteases, and cationic proteins with bactericidal activity.

Excessive platelet activation has been regarded as a key pathological factor in the development of many vascular diseases such as acute coronary syndromes, myocardial infarction and atherosclerosis [4, 5]. Endothelial dysfunction/injury initially induces platelet aggregation and thrombus formation in mice, which is considerably inhibited when the PPAR-β/γ antagonists are administrated simultaneously. Collectively, these results provide important information regarding the mechanism by which nifedipine inhibits platelet aggregation and thrombus formation through activation of PPAR-β/γ-mediated signaling pathways. These findings highlight that PPARs are novel therapeutic targets for preventing and treating platelet-hyperactivity-related vascular diseases.

Keywords: Nifedipine; Peroxisome proliferator-activated receptors; Nuclear factor-xB; Platelet aggregation; Protein kinase Ca; Nitric oxide

Abstract

Platelet hyperactivity often occurs in hypertensive patients and is a key factor in the development of cardiovascular diseases including thrombosis and atherosclerosis. Nifedipine, an L-type calcium channel blocker, is widely used for hypertension and coronary heart disease therapy. In addition, nifedipine is known to exhibit an antiplatelet activity, but the underlying mechanisms involved remain unclear. Several transcription factors such as peroxisome proliferator-activated receptors (PPARs) and nuclear factor kappa B (NF-κB) exist in platelets and have an ability to regulate platelet aggregation through a non-genomic mechanism. The present article focuses on describing the mechanisms of the antiplatelet activity of nifedipine via PPAR activation. It has been demonstrated that nifedipine treatment increases the activity and intracellular amount of PPAR-β/γ in activated platelets. Moreover, the antiplatelet activity of nifedipine is mediated by PPAR-β/γ-dependent upon the up-regulation of the PI3K/AKT/NO/cyclic GMP/PKG pathway, and inhibition of protein kinase Ca (PKCa) activity via an interaction between PPAR-β/γ and PKCa. Furthermore, suppressing NF-xB activation by nifedipine through enhanced association of PPAR-β/γ with NF-xB has also been observed in collagen-stimulated platelets. Blocking PPAR-β/γ activity or increasing NF-xB activation greatly reverses the antiplatelet activity and inhibition of intracellular Ca2+ mobilization, PKCa activity, and surface glycoprotein IIb/IIIa expression caused by nifedipine. Thus, PPAR-β/γ-dependent suppression of NF-xB activation also contributes to the antiplatelet activity of nifedipine. Consistently, administration of nifedipine markedly reduces fluorescein sodium-induced vessel thrombus formation in mice, which is considerably inhibited when the PPAR-β/γ antagonists are administrated simultaneously. Therefore, platelets are an important link between tissue damage and monocytes leads to the pathogenesis of atherosclerosis.

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concept, several lines of evidence have demonstrated that platelet hyperactivity often occurs in hypertensive or cardiovascular patients [6, 7]. Thus, agents with inhibiting platelet hyperactivity may be potential therapeutic drugs for platelet-related vascular diseases.

2. Platelet activation

Platelet adhesion to the extracellular matrix is the initial step in haemostasis [8]. When vascularity is damaged, the immobilized vWF on exposed collagen becomes a strong adhesive substrate. The vWF, a multimeric adhesive glycoprotein, contains binding sites for collagen glycoprotein (GP)Ib and integrin GPIb/IIIa (αIIbβ3) [9]. The adhesion is mediated by the interaction between the GPIb-V-IX-V receptor complex on the platelet surface to vWF, and GPV1 and GPIa to collagen at sites of vascular injury. The action of vWF and GPIb-IX-V complex is required for the adhesion of platelets to the subendothelium, which enables GPV1 binding to collagen [10]. In addition, collagen serves as a binding site for vWF in the subendothelial matrix, and therefore contributes to the adhesion of unactivated platelets via GPIb-IX-V (Figure 1) [11]. The adhesion is mediated by the interaction of the GPIb-V-IX-V complex with vWF via the activated GPIIb–IIIa receptors. This triggers platelet aggregation by crosslinking adjacent platelets through binding to fibrinogen and vWF via the activated GPIb–IIIa receptors.

There are multiple pathways regulating platelet aggregation. The platelet agonists including ADP, thrombin, collagen and serotonin perform their functions through their specific receptors of platelets. ADP stored in dense granules is released during platelet activation. ADP promotes platelet activation through its receptors (P2Y1 and P2Y12). P2Y1 is a G-protein-coupled seven-transmembrane domain receptor that stimulates platelet shape change and mobilizes calcium from intracellular stores by activating phospholipase C (PLC) [13]. Activation of the P2Y12 receptor inhibits the adenylate cyclase activity of platelets and seems to be responsible for a positive feedback mechanism for platelet stimulation especially by weak agonists [14]. There are two receptors (protease activated receptors 1 (PAR1) and 4 (PAR4)) on platelet surfaces for thrombin, the most potent physiological platelet activator [15]. PAR1 mediates platelet activation at low concentrations of thrombin, while PAR4 is activated at higher thrombin concentration. PARs are also expressed in other cells in the vasculature, such as leukocytes, endothelial cells and smooth muscle cells. It has been reported that PAR-1 and PAR-2 expressed in the vessel wall are involved in contractility, inflammation, proliferation, and repair [16]. Collagen is also a strong platelet activating agent because it stimulates platelet adhesion and is mediated by the binding of vWF to it at the sites of vascular injuries. Moreover, other glycoprotein receptors such as GPIb, GPIb/IIIa, GPIa/IIa, and GPVI all contribute to collagen-mediated platelet activation [1]. Serotonin expresses its actions through the serotonin receptor 5-HT2A to enhance procoagulant activity via retention of fibrinogen and thrombospondin on platelet surfaces. Furthermore, epinephrin and other catecholamines stimulate the platelet α2A-adrenergic receptor, which coupled with a G-protein leads to the inhibition of adenylate cyclase activity, which in turn induces platelet aggregation [17].

When platelets are activated by collagen or thrombin, arachidonic acid (AA) is liberated from membrane phospholipids. This liberation is caused by Ca2+-dependent phospholipase A2 (PLA2), diglyceride lipase and/or phosphatidic acid specific phospholipase A2 [18]. Then, AA is converted to thromboxane A2 (TXA2), a potent activator for the release reaction and aggregation of platelets by cyclooxygenase (COX) and thromboxane synthase. TXA2, when released from the platelets, binds to the G-protein coupled thromboxane receptor and functions as an agonist for platelet activation.
[19]. In addition, TXA₂ also participates in a positive feedback loop further increasing [Ca²⁺], and AA-induced ATP release and platelet aggregation [20]. Phosphoinositides breakdown is another critical mechanism accounting for agonist-induced platelet activation. Simulation by agonists such as thrombin or collagen results in PLC-catalyzed hydrolysis of plasma membrane phospholipid phosphatidylinositol 4,5-bisphosphate, with concomitant formation of inositol 1,4,5-trisphosphate (IP₃) and diacylglycerol [21]. The IP₃ binds to the IP₃ receptor on sarcoplasmic reticulum (SR) triggering a significant release of Ca²⁺ from SR, leading to an increase of intracellular Ca²⁺ concentration [22]. It has been demonstrated that an increase of intracellular Ca²⁺ concentration, as a result of either calcium influx and/or calcium release from intracellular stores, is fundamental to platelet activation [23]. Importantly, diacylglycerol acts as an activator for protein kinase C (PKC) that is essential for agonist-induced platelet aggregation and granule secretion [24]. At least seven PKC isoforms (α, β, δ, η, ε, θ, and ζ) are found in platelets. Among these PKC isoforms, activation of PKCa through Syk-dependent phosphorylation is crucial for platelet activation [25]. Moreover, TXA₂ has an ability to activate PKC by activating PLC-dependent pathways [26], suggesting that blocking TXA₂ formation may inhibit PKCα-related signaling pathways. Mitogen-activated protein kinases (MAPKs) containing extracellular signal-regulated kinase (ERKs), the c-Jun N-terminal kinase (JNK) and p38 MAPK have been identified in platelets [27]. The roles of JNKs and ERKs in platelet mechanisms are still unclear. On the other hand, activation of p38 MAPK by PKC can phosphorylate cPLA₂ on Ser505, leading to the production of AA and TXA₂ synthesis [28]. Thus, cPLA₂ activation is regulated, at least in part, by PKC through p38 MAPK. These platelet-activating mechanisms ultimately up-regulate surface GPIIb/IIIa expression, thereby promoting the binding of fibrinogen and platelet aggregation.

### 3. Antiplatelet mechanisms

#### 3.1. NO/cyclic GMP

Nitric oxide (NO), synthesized from L-arginine by nitric oxide synthase (NOS), activates intracellular soluble guanylyl cyclase (sGC) and guanosine 3′,5′-cyclic monophosphate (cGMP) formation, which subsequently activates cGMP-dependent protein kinase (PKG). The NO-dependent signaling pathway is known to play an important modulatory role both in physiological and pathological conditions [29]. Up-regulation of the NO/cGMP/PKG1 cascade reportedly inhibits platelet activation by regulating actin filament dynamics, integrin activation, and intracellular Ca²⁺ mobilization, which in turn suppresses PLC and PKC activity [30, 31]. Moreover, PKG promotes sarcoplasmic reticulum ATPase (SERCA)-dependent refilling of intraplatelet Ca²⁺ stores and inhibits inositol-1,4,5-trisphosphate-stimulated Ca²⁺ release from the sarcoplasmic reticulum, which in turn decreases intracellular Ca²⁺ level and platelet activation [30]. PKG is capable of phosphorylating the TXA₂ receptor, thereby inhibiting its function. cGMP also indirectly increases intracellular cAMP through inhibition of phosphodiesterase type 3 to synergistically inhibit platelet aggregation [33]. Interestingly, previous studies have indicated that the actions of NO on platelet function are also mediated by a cGMP-independent mechanism that inhibits exocytosis of platelet granules (dense, lysosomal, and α-granules) by S-nitrosylation of N-ethylmaleimidesensitive factor (NSF) [34].

#### 3.2. Cyclic AMP

It has been demonstrated that the elevation of cyclic AMP formation reduces platelet functions including adhesion, aggregation, the release of granule contents as well as the rise of intracellular Ca²⁺ mobilization [33, 35]. The steady-state level of cyclic AMP is maintained by a balance between the rate of synthesis by adenylyl cyclase and the rate of degradation by cyclic AMP phosphodiesterase. Research has shown that several cyclic AMP-elevating agents exert antiplatelet activity through a cyclic AMP-dependent protein kinase (PKA)-dependent signal pathway [36, 37].

#### 3.3. GPIIb/IIIa Inhibitors

When platelets are activated by physiological agonists such as thrombin, ADP, or collagen, the intracellular signal pathways for platelet activation are stimulated and thereby induce conformational changes of αIIbβ₃ leading to the formation of an activated state with high affinity for fibrinogen and numerous other ligands [1, 38]. As a result of the enhancement of αIIbβ₃-mediated binding to the bivalent molecule, fibrinogen may cause platelets aggregation. To date, two binding sites have been well characterized in αIIbβ₃: an Arg-Gly-Asp (RGD)-binding site and a Lys-Glu-Ala-Gly-Asp-Val (KAGDV)-binding site. Fibrinogen binds via the KAGDV-binding site. Agents that bind within the ligand-binding region of αIIbβ₃ and block the binding of its natural ligands have been developed and termed GPIIb/IIIa inhibitors. There are three FDA-approved integrin αIIbβ₃ inhibitors, and they include abciximab (ReoPro; Lilly), eptifibatide (Integrilin; millennium Pharmaceuticals/Schering–Plough), and tirofiban (Aggrastat; merck). Abciximab is a murine human chimeric fab fragment that was derived from the murine monoclonal antibody 7E3. Eptifibatide is a KDG-containing cyclic heptapeptide. And tirofiban is a non-peptide derivative based on the RGD sequence [39]. Clinical studies have indicated that a blockade of the glycoprotein IIb/IIIa receptors limits the inflammatory responses secondary to coronary intervention, suggesting that inhibition of inflammatory marker expression by GPIIb/IIIa inhibitors may contribute to its clinical benefit [40]. Our previous study tested the effect of the synthesized RGRHGD with the highest local hydrophilicity region of B chain of β-bungarotoxin on platelet aggregation. The RGRHGD holds parts of both RGD and KDG peptides that have been reported to exhibit a high binding affinity to GPIIb/IIIa. Moreover, the inhibitory effect of RGRHGD on platelet aggregation is associated with attenuation of TXA₂ formation and intracellular calcium mobilization. These findings may, at a later date, provide a useful method for finding potential therapeutic agents through molecular modeling analysis [41].

### 4. Peroxisome proliferator-activated receptors (PPARs) and platelet activation

PPARs belonging to ligand-activated transcription factors modulate several important biological effects, including lipid, glucose homeostasis, energy metabolism, and inflammation [42, 43]. A variety of compounds can serve as PPAR ligands and activate the receptor. PPARα ligands are fatty acids, and their derivatives as well as eicosanoids include 8-S-hydroxyeicosatetraenoic acid (8SHETE) and leukotriene B₄ (LTB₄) [44]. Moreover, fibrates, the synthetic ligands for PPARα, are widely used in the treatment of hypertriglyceridemia and hyperlipidemia [44]. Other phar-
macological compounds such as nonsteroidal anti-inflammatory drugs (NSAIDs) are confirmed as PPARs ligands [45]. PPARγ is activated by 15-deoxy-12, 14-prostaglandin J2 (PG-J2) and 15-hydroxyeicosatetraenoic acid (15-HETE) [46, 47] that are AA metabolites derived from COX and lipoxygenase pathways. In addition, fatty acid-derived compounds of oxidised LDL, including 9- and 13-hydroxyoctadecadienoic acid (9- and 13-HODE), and glitazones [48] (an anti-diabetic drug), indomethacin, and ibuprofen all function as ligands for PPARγ [45]. In response to their ligands, PPARs undergo a conformational change leading to the recruitment of distinct coactivators and corepressors. Subsequently, these changes result in PPAR heterodimerization with cис-retinoid X receptor (RXR) and in turn regulate downstream gene expression by binding to the peroxisome proliferator response element (PPRE) that exist in the promoter of target genes of nucleated cells. Although platelets are anuclear cells, they also contain transcription factors such as PPARs. The existence of three PPAR isoforms (α, β/δ, and γ) in human platelets has been demonstrated, and activation of PPARs inhibits platelet aggregation through a nongenomic mechanism [49, 50]. It has been reported that the inhibitory effect of PPAR agonists on platelet aggregation is associated with the modulation of GPVI, PKCa and calcium mobilization signals [51]. Our recent study indicated that the antplatelet activity of alpha-lipoic acid is mediated by PPAR-α/γ-dependent processes [50]. Therefore, reagents exerting PPAR-activating activity have been regarded as a new class of antplatelet drugs.

5. PPARs and atherosclerosis

Atherosclerosis is a complex process characterized by lipid accumulation in the arterial wall resulting in heart and brain infarction. The atherogenesis is intiated via the attraction of various cells such as monocyte/macrophages, T lymphocytes, endothelial cells, and smooth muscle cells (SMCs). This cellular activation promotes local inflammatory responses and migration and proliferation of SMCs, which in turn leads to the formation of foam cells. PPARs are expressed in the vascular wall and atherosclerotic lesions, suggesting that they may modulate the atherogenic processes. Clinical studies have indicated that PPARγ ligand troglitazone inhibits SMC proliferation and decreases the intima and media thickness of carotid arteries [52]. The inhibitory effects of PPARs (α and γ) on the expression of inflammatory genes, such as interleukin-6, cyclooxygenase-2, inducible nitric oxide synthase (iNOS), matrix metalloproteinase-9, endothelin-1, lip accumulation within the plaque, and thrombogenesis [53] have been proposed to be the underlying mechanisms for their anti-atherosclerotic effects. Furthermore, PPARα activators also can induce apoptosis of activated macrophages by inhibiting the antiapoptotic NF-κB pathway [54] and reducing monocytic recruitment to early atherosclerotic lesions by inhibition of monocyte-recruiting proteins such as vascular cell adhesion molecules (VCAM)-1 expression in endothelial cells [55]. These findings suggest that up-regulation of PPAR expression/activation may prevent the progress of atherosclerotic disease.

6. Nifedipine, a dihydropyridine calcium channel blocker (CCB)

The calcium channel blockers (CCB) are a group of drugs used to treat cardiovascular diseases including hypertension, angina, and peripheral vascular disorders. CCBs were approved for the treatment of hypertension in the 1980s. Since then, CCBs have increased markedly because of their effective lowering of blood pressure with few side effects. In addition to the cardiovascular effects of CCBs, other beneficial functions of CCBs, such as anti-oxidative, anti-inflammatory, anti-atherosclerotic, bone-remodeling, and immunomodulating properties have been reported [56]. It is well known that inflammation is a fundamental basis of atherosclerosis. Several reports have revealed that dihydropyridine CCBs exert anti-inflammatory effect by suppressing the tumor necrosis factor (TNF), monocyte chemoattractant protein-1 (MCP-1), and pro-inflammatory cytokine expression accompanied by the reduction of NF-κB activation in various vascular cells including endothelial cells, macrophages, and smooth muscle cells [56]. These actions of CCBs may contribute to the anti-atherosclerotic effect in vascular cells.

CCBs can be classified into 3 main classes according to their different structure. The three classes are the phenylalkylamines (e.g., verapamil), the benzothiazepines (e.g., diltiazem), and the dihydropyridines (e.g., nifedipine, amlodipine, isradipine). It is known that different classes of CCBs have differing pharmacologic actions. With their relative potency of lowering blood pressure, the dihydropyridine-type compounds such as nifedipine are the most potent subclass. Nifedipine, a dihydropyridine-based L-type CCB, is widely used in the treatment of hypertension and coronary heart diseases. Clinical studies have shown that a significant reduction of new coronary lesions and the intima-media thickness in the carotid artery were observed in patients treated with nifedipine [57]. These findings confirmed that nifedipine has an anti-atherosclerotic effect beyond its blood pressure-lowering effect. The protection may be associated with suppressing reactive oxygen species (ROS) formation and subsequent inflammatory responses, as well as smooth muscle cell proliferation, migration and differentiation [56]. Importantly, nifedipine can activate PPAR-γ by inhibiting ERK1/2 activity in macrophages [58], suggesting that PPARs may involve the pharmacological effects of nifedipine. As increased intracellular Ca2+ concentration ([Ca2+]i) is essential for platelet activation, therefore, reagents that attenuate platelet [Ca2+]i may have an antplatelet activity. As expected, nifedipine is capable of inhibiting platelet aggregation, though platelets lack L-type calcium channels. To date, the underlying molecular mechanisms remain unclear; as although the activation of NO/cGMP-dependent signaling pathway [59, 60] has been proposed as a possible mechanism that contributes to its antplatelet activity of CCBs.

7. Nifedipine-mediated anti-aggregatory effect via activation of PPAR-β/γ

PPARs play an important role in the modulation of metabolism and inflammatory processes, and exhibit a protective effect against the development of atherosclerosis and cardiovascular diseases [61, 62]. Traditionally, the actions of transcription factors are thought to be mainly attributed to their regulation in gene expression. Recently, accumulating evidence supports that there are nongenomic actions of these receptors [63]. Because a number of transcription factors including PPARs, estrogen receptors (ER), and nuclear factor kappa B (NF-kB) have been found in platelets, many studies have focused on the role of cytoplasmic PPARs in platelet function. It has been reported that the activation of
Pepers (α, β/δ, and γ) by respective agonists inhibits platelet aggregation [49, 50] and slows intraarterial thrombus formation due to increased NOS expression [64]. Thus, selective ligands for PPARs may negatively regulate platelet activation. Based on the finding that nifedipine induces PPAR-γ activity in macrophages and smooth muscle cells [58], it is possible that PPARs may involve nifedipine-mediated regulation of platelet and vascular functions.

Our recent study has confirmed that treatment with nifedipine significantly increases PPAR-β and PPAR-γ activity due to the inhibition of phosphorylation of ERK1/2 and PPAR-γ without affecting PPAR-α activity in collagen-stimulated platelets [65]. These results indicate that nifedipine is a dual PPAR-β-γ activator in platelets. However, other CCBs like amlodipine are PPAR-β activators, and lacidipine has no significant effect on PPARs activity in human platelets, suggesting that the activation of PPARs is not a common effect of all CCBs. Accordingly, the effects of different CCBs on platelet PPARs activity and the role of PPARs on CCBs-mediated antplatelet activity are diverse and chemical structure specific. Upon activation by inducers such as collagen, a rapid release of PPAR-β/γ from the α-granules of platelets into extracellular regions results in a marked reduction of the intracellular amount of PPAR-β/γ, which may have a systemic effect. A novel finding is that nifedipine greatly inhibits the release of PPAR-β/γ from activated platelets, thereby increasing the intracellular availability of PPAR-β/γ which may enhance its cellular functions like the regulation of platelet activation. However, the underlying mechanisms accounting for the phenomenon require further investigation. Blocking PPAR-β/γ activity with their specific antagonists significantly reverses the inhibitory effect of nifedipine on platelet aggregation, supporting that PPAR-β/γ is involved in the antiplatelet activity of nifedipine. In addition, nifedipine-mediated up-regulation of the PI3K/Akt NO/cGMP/PKG cascade that results in a reduction of Ca^2+ mobilization is regulated by a PPAR-β/γ-dependent signaling pathway (Figure 1). The activation of PKCa is crucial for platelet secretion and aggregation [24]. Consistent with our previous findings is the discovery that the inhibitory effect of PPAR-α-γ on platelet PKCa activity is associated with its association with PKCa [50]. In collagen-stimulated platelets, nifedipine also induces an interaction between PPAR-β/γ and PKCa accompanied by decreased PKCa activity evidenced by reduced PKCa phosphorylation in the complex. Similarly, an addition of PPAR-β-γ antagonists abrogated the attenuation of PKCa activity by nifedipine, suggesting that direct interaction between PPAR-β/γ and PKCa is a possible way to suppress PKCa activity.

Furthermore, administration of nifedipine markedly inhibited fluorescein sodium and irradiation-induced vessel thrombus formation in vivo. However, the antithrombotic effect of nifedipine was considerably reduced when PPAR-β/γ antagonists were administered simultaneously. Taken together, the antplatelet and antithrombotic effects of nifedipine are mediated by activation of PPAR-β/γ leading to up-regulation of the NO/cGMP/PKG cascade, as well as inhibition of PKCa activity and intracellular Ca^2+ mobilization.

8. NF-κB and platelet activation

NF-κB, a transcription factor, normally exists as an inactive cytoplasmic complex heterodimer complex composed of p50 and p65 subunits bound to the inhibitory protein, IκB-α. Upon stimula-
Treatment with nifedipine decreases NF-κB activation by inhibiting IKK-β/IκBα phosphorylation in collagen-stimulated platelets. The inhibition of NF-κB activation is significantly reversed by specific PPAR-β/γ antagonists, supporting the notion that PPAR-β/γ negatively regulates NF-κB activation in platelets. Additionally, activation of NF-κB with betulinic acid (BetA) abolishes the nifedipine’s inhibition of intracellular Ca2+ mobilization and platelet aggregation, indicating that PPAR-β/γ-mediated NF-κB activation involves the antiplatelet activity of nifedipine [70]. Notably, our research demonstrated for the first time that in activated platelets, nifedipine also induces an interaction of PPAR-β/γ with NF-κB leading to decreased p65NF-κB phosphorylation in the complex. These results may provide a novel mechanism by which PPAR-β/γ suppresses NF-κB activation in platelets through a direct interaction with NF-κB. Furthermore, the suppression of NF-κB activation by nifedipine is at least partly attributed to PPAR-β/γ-dependent up-regulation of the NO/cyclic GMP/PKG1 pathway (Figure 3).

The binding of fibrinogen to the surface GPIIb/IIIa complex is a critical final step for platelet aggregation by crosslinking platelets and by the stabilization of aggregates. It has been reported that inhibiting NF-κB activation by NF-κB inhibitors reduces the outside-in/inside-out signaling of GPIIb/IIIa and fibrinogen binding in activated platelets [67]. Furthermore, GPIIb/IIIa is also required for NF-κB activation in human neutrophils [71], suggesting that there is a mutual activation between NF-κB and GPIIb/IIIa. Thus, the decreased surface GPIIb/IIIa expression by nifedipine may be a consequence of PPAR-β/γ-down-regulated NF-κB activation as evidenced by blocking PPAR-β/γ-activity or enhancing NF-κB activation resulting in an elevated expression of GPIIb/IIIa.

In conclusion, nifedipine significantly increases the activity and intracellular levels of PPAR-β/γ in activated platelets, which subsequently up-regulates the PI3K/Akt/eNOS/NO/cyclic GMP/PGC1 cascade leading to the suppression of intracellular calcium mobilization, surface GPIIb/IIIa expression, and PKC activity via association of PPAR-β/γ with PKC. In addition, nifedipine is capable of inhibiting NF-κB activity through direct interaction of PPAR-β/γ with NF-κB. These effects of nifedipine ultimately inhibit platelet aggregation and thrombosis formation. All told, nifedipine may be a potential drug for alleviating atherothrombosis and vascular diseases by targeting PPAR-β/γ-dependent signaling pathways in platelets.

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