PHARMACEUTICALS AND ENDOCRINE-DISRUPTING COMPOUNDS: ACCUMULATION IN THE TISSUE OF SKADAR LAKE FISH

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ABSTRACT

Determining the presence of pharmaceuticals and endocrine-disrupting compounds in fish tissues has been carried out in Montenegro for the first time in Moraca river and Skadar lake. Skadar lake, Montenegro, is the largest of the Balkan lakes and has a surface area which fluctuates seasonally from approximately 370 to 600 km². During the spring time of 2016, in the time of high water levels of the lake, fish were fetched in triplets and identified. Determining the presence of pharmaceuticals and endocrine-disrupting compounds in fish tissues has been carried out in Montenegro for the first time. Their muscle tissue was separated, grinded, freeze-dried, and then lyophilized. Prepared samples were analyzed with liquid chromatography coupled to tandem mass spectrometry to determine the presence of pharmaceuticals and endocrine-disrupting compounds. Obtained results are under the border of detection, out of 38 tested distributes, only the tissue of Alburnus scoranza showed the presence of triclosan.

Keywords: Endocrine-disrupting compounds, Lyophilized, Liquid chromatography coupled to tandem mass spectrometry, Alburnus scoranza, Triclosan.

INTRODUCTION

In recent years, potential risks associated with the release of Endocrine Disrupting Compounds (EDCs) into the aquatic environment have become an increasingly important issue for environmental regulators. Several investigations have shown that the substances of pharmaceutical origins are often not eliminated during wastewater treatment, and are not biodegradable as well (Ternes, 1998; Daughton et al., 1999; Zwiener et al., 2004). Due to the conservative nature of physiological processes, chemicals affect several aquatic species (e.g. algae, invertebrate and fish) in a manner similar to their affect on humans due to comparable target molecules (Fent et al., 2006). Many contaminants have been detected in wastewater effluents and surface waters in recent years, including pharmaceuticals (PhACs) and (EDCs). Many of these compounds are not completely removed in the wastewater treatment plants (WWTPs) and are released to the environment (Gros et al., 2009). EDCs are designed to modify physiological or biochemical functions in target organisms but can have severe consequences when non-target organisms are under their exposure.

Better understanding of how these compounds are transferred from water to the biota could help to determine potential ecosystem damage associated with the WWTP effluent discharge contaminants. It is generally accepted that substances with octanol-water partition coefficient (log KOW) values higher than or equal to 3 have the potential to bioaccumulate in biological tissues. Many PhACs and EDCs have a log KOW < 3, and thus are not expected to bioaccumulate. However, some of these compounds are lipid soluble and therefore potentially bioaccumulative in the environment (Huerta et al., 2013) and fish tissue. Moreover, when considering bioaccumulation of PhACs and EDCs in aquatic organisms, one must take other factors into consideration, such as the different rates of metabolism in various organisms, how their metabolites accumulate, and the uptake and depuration kinetics.

Situated along the Montenegro-Albanian border, Lake Skadar is the largest lake of the Balkan Peninsula with unique characteristics, wide range of endemic and rare or endangered plant and animal species, recognized as a wetland site of international significance according to the Ramsar Convention (2002). In spite of its importance, inflowing waters from the Moraca river and other regional rivers contaminated by the industry, municipal and agricultural activities in the area influence the Lake. Therefore, the Lake has been the subject of various physical, chemical, biological and toxicological examinations (Beeton & Karaman, 1981; Rastall et al., 2004; Rakocevic-Nedovic & Hollert, 2005; Stesevic et al., 2005; Perovic et al., 2010; Kastratovic et al., 2012; Stesevic et al., 2014; Vemic et al., 2014; Petrovic et al., 2016, Perovic et al., 2019). In this study, we assessed the level of bioaccumulation of ECDs and PhACs in aquatic organisms (fish tissues) Rutilus prespensis (Karaman, 1924), Squalius platyceps (Zupanic et al., 2010), Scardinius krenevici (Bianco & Kottelat, 2005), Chondrostoma ohridanum (Karaman, 1924), Cyprinus carpio (Linnaeus, 1758) and Alburnus scoranza (Bonaparte, 1845).
EXPERIMENTAL

Area of study and sampling

The research location belongs to the area of Skadar lake and its tributaries. Locations are marked on the Figure 1.

Figure 1. Map of Lake Skadar with sampling locations.

Lake Skadar is the largest lake on the Balkan Peninsula. It is located in karst area on the Montenegro-Albanian border and covers the surface from approximately 370 to 600 km², depending on seasonal fluctuations. This area is classified as an internationally important RAMSAR site (Lake Skadar, Site, 3YU003, Ramsar Convention 2002) and promoted as a National Park IUCN Category II. In spite of its importance, inflowing waters from the Moraca River and other regional rivers contaminated by the industry, municipal and agricultural activities in the area affect the lake. River Moraca (Mediterranean basin), near the town Podgorica, is relatively intact but still receives the treatment of waste water discharge, produced by 300,000 citizens. River Moraca emerges at the topographic point of 975 m above the creeks which disembogue under the Javorje and Zebalci slopes, with largest of them being Javor and Rzav creeks. River Moraca is 113 km long, which makes it the longest river in Montenegro. The researched locations with the coordinates.

Vukovec 42°20’02”N; 19°11’60” E, "Left branch of river Moraca" 42°25’84” N; 19°13’46”E "Right branch of river Moraca" 42°27’70” N; 19°12’32”E "Kraljeva glavica"-Vranjina 42°27’57”N; 19°12’15”E "Tanki rt"- Under the bridge 42°27’90” N, 19°13’49”E.

Throughout decades, this locality has been under the influence of industrial waters originating from Kombinat Aluminijuma (CAP) and Machine Factory "Radoje Dakić". Therefore, all researches were conducted in order to determine the concentration of PCB, aluminium, heavy metals and oil in water, land and biota. However, this industry slowly stopped working during the 90s. Podgorica and its surroundings make up for more than a half of the population of Montenegro. Therefore, waste waters are seen as the main pollutants of river Moraca.

Animals and experimental design

Samples were fetched in triplets using fishnets in spring of 2016, during the time of high water level of the lake, we separated muscle tissues and repeated the process 3 times, grinded them separately in a hand blender, put them into special sample boxes and froze them. Two days later, they were transported to the Marine Institute of Montenegro where they were lyophilized.

Sample purification optimization

Fish homogenate extracts were spiked with a mixture of the target analytes and a comparison was made between three purification techniques: SPE Florisil cartridges (1 g, 6 ml cartridges), Gel permeation chromatography (GPC) in an Agilent 1260 Infinity high pressure liquid chromatography system in tandem with a diode array detector (HPLC-DAD), SPE Oasis HLB (200 mg, 6 ml cartridges) followed by GPC. For the clean-up method based on SPE Florisil, fish extracts were redissolved in 10 ml of acetonitrile prior SPE, which was performed in a J.T.Baker® system.

Liquid chromatography and mass spectrometry analysis

The chromatographic tandem mass spectrometry method applied for the analysis of fish extracts was adapted from Gros et al. (2009). Briefly, chromatographic separations were carried out with a Waters Acquity Ultra-PerformanceTM liquid chromatography system, using an Acquity HSS T3 column (50 mm×2.1 mm i.d., 1.8 µm particle size) for the compounds analyzed in positive electrospray ionization (PI) mode and Acquity BEH C18 column (50 mm×2.1 mm i.d., 1.7 µm particle size) for the ones analyzed under negative electrospray ionization (NI) mode. For the analysis in PI mode, separation conditions were as follows: solvent (A) methanol, solvent (B) 10mM formic acid/ammonium format (pH 3.2) at a flow rate of 0.5 mL/min. The analysis in NI mode was performed by using acetonitrile (A) and (B) 5mM ammonium acetate/ammonia (pH=8) at a flow rate of 0.6 mL/min. The sample volume injected was 5_L in both cases. The UPLC instrument was coupled to a 5500 OTRAP hybrid triple quadrupole-linear ion trap mass spectrometer (Applied Biosystems): nitrogen collision gas (CAD) medium; source gases GS1 and GS2 were set at 60 and 50 psi, respectively. Besides the monitoring of the SRM transitions, the relative abundance of the two SRM transitions in the sample were compared with those in the standards, and the relative abundances in the samples must be within ±20% of the two SRM ratios in the analytical standards. Prepared samples were sent to Catalan Institute for Water Research (ICRA)-Gerona in order to determine the existence of certain EmS and EDCs in the fish tissues using the (Huerta et al., 2012) and (Huerta et al., 2013) method.
NUMERICAL RESULTS

Occurrence of PhACs and EDCs in freshwater fish is presented in table 1.

Obtained results are under the border of detection, or in the frames of the method detection and quantification limits (MDL, MQL). Out of 38 tested distributes, only one (triclosan) was founds in the muscle tissue of *Alburnus scoranza* in concentration 14,1±1,2 ng/g (Table 1).

Triclosan, phenol, are commonly used commercial microbicides found in toothpastes and soaps. Triclosan (is widely used as an antibacterial agent in various industrial products, such as textile goods, soap, shampoo, liquid toothpaste and cosmetics, and often detected in wastewater effluent. However, there is a paucity of data on the toxicity of triclosan and its effects on aquatic organisms. For example, (Tatarazako et al., 2004) in his observation found that concentration of fish (*Danio rerio* and *Oryzias latipes*) was from 0.07 to 0.29 mg/l. Other scientists have studied the toxicity of triclosan too, especially because of its wide spread in chemical industry. For example, Ximei et al. (2013), researched assessment of toxic effects of triclosan on swordtail fish (*Xipophorus helleri*) using a multi-biomarker approach. Another example is the research of triclosan in Fresh Water Fish Gibelion Catla from the Kaveri River, India, and its Consumption Risk Assessment (Govindarej et al., 2014).

Of note, while fish can be exposed to higher levels of triclosan from their surroundings, algae and invertebrates are often considered more sensitive Chalew & Halden (2009). A number of different aquatic toxicity thresholds for triclosan are available in the literature. Colgate-Palmolive scientists used an unconventional method to develop a PNEC of 1,550ng/L (Capdevielle et al., 2008). In contrast, a more traditional and conservative method based on acute algal toxicity has led to the use of PNEC of 4.7 ng/L (e.g., von der Ohe et al., 2000). Should the latter threshold be considered more appropriate for San Francisco Bay, the few recent ambient Bay surface water measurements available (up to 68±26 ng/L) may suggest cause for concern. However, existing data are too few to trigger reclassification of triclosan within the RMP’s CEC risk and management action framework (Sutton et al., 2016). Until more data are generated, triclosan may remain classified as a low concern (Tier II) for San Francisco Bay.

| Sample name | Ratilus prespensis | Squalius platycps | Scardinius knezveci | Chondrostoma ohridanum | Ciprinus carpio | Alburnus scoranza | MDL | MQL |
|-------------|--------------------|-------------------|---------------------|-----------------------|----------------|-------------------|------|-----|
| Caffeine    | <MDL               | <MDL              | <MDL                | <MDL                  | <MDL           | <MDL              | 0,3255 | 1,0849 |
| Progesterone| <MDL               | <MDL              | <MDL                | <MDL                  | <MDL           | <MDL              | 0,0158 | 0,0526 |
| Levonorgestrel| <MDL              | <MDL              | <MDL                | <MDL                  | <MDL           | <MDL              | 0,0002 | 0,0006 |
| TCEP        | <MDL               | <MDL              | <MDL                | <MDL                  | <MDL           | <MDL              | 0,25   | 0,84 |
| TBEP        | <MQL               | <MQL              | <MQL                | <MQL                  | <MQL           | <MQL              | 0,07   | 0,27 |
| TCCP        | <MQL               | <MQL              | <MQL                | <MQL                  | <MQL           | <MQL              | 25,4   | 84,8 |
| Estrone     | <MDL               | <MDL              | <MDL                | <MDL                  | <MDL           | <MDL              | 0,003  | 0,0099 |
| 17β-Estradiol| <MDL              | <MDL              | <MDL                | <MDL                  | <MDL           | <MDL              | 0,01   | 0,05 |
| Estriol     | <MDL               | <MDL              | <MDL                | <MDL                  | <MDL           | <MDL              | 0,0002 | 0,0007 |
| 17α-Ethinylestradiol| <MDL | <MDL              | <MDL                | <MDL                  | <MDL           | <MDL              | 0,0004 | 0,0014 |
| Estrone-3-sulfate| <MDL | <MDL              | <MDL                | <MDL                  | <MDL           | <MDL              | 0,0007 | 0,0022 |
| Bisphenol A | <MQL               | <MQL              | <MQL                | <MQL                  | <MQL           | <MQL              | 0,02   | 0,07 |
| Triclosan   | <MQL               | <MQL              | <MQL                | <MQL                  | <MQL           | <MQL              | 14,1±1,3 | 0,0018 | 0,0061 |
| Methylparaben| <MDL               | <MDL              | <MDL                | <MDL                  | <MDL           | <MDL              | 0,01   | 0,03 |

Table 1. Occurrence of EDCs and PhACs in freshwater fish in Lake Skadar lake.
| Sample name | Rutillus prespensis | Squalius platyceps | Scardinius knezevici | Chondrostoma ohridanum | Ciprinus carpio | Alburnus scoranza | MDL | MQL |
|-------------|---------------------|---------------------|---------------------|-----------------------|----------------|------------------|------|------|
| Atenolol    | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,01 | 0,02 |
| Carazolol   | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,01 | 0,02 |
| Metropolol  | <MQL                | <MDL                | <MQL                | <MQL                  | <MQL           | <MQL             | 0,1  | 0,35 |
| Nadolol     | <MQL                | <MQL                | <MQL                | <MQL                  | <MQL           | <MQL             | 0,31 | 1,04 |
| Propanolol  | <MDL                | <MDL                | <MQL                | <MQL                  | <MQL           | <MQL             | 0,09 | 0,29 |
| Sotalol     | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,08 | 0,26 |
| Carbamazepine | <MDL              | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,05 | 0,17 |
| Citalopram  | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,12 | 0,4  |
| Diazepam    | <MDL                | <MDL                | <MQL                | <MQL                  | <MQL           | <MQL             | 0,03 | 0,1  |
| 10,11-EpoxyCBZ | <MDL              | <MDL                | <MQL                | <MDL                  | <MQL           | <MQL             | 0,02 | 0,05 |
| 2-HydroxyCBZ | <MDL               | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,01 | 0,03 |
| Lorazepam   | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,06 | 0,2  |
| Sertraline  | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,25 | 0,83 |
| Venlafaxine | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,01 | 0,03 |
| Clopidrogel | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,01 | 0,04 |
| Codeine     | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,02 | 0,06 |
| Levamisole  | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,02 | 0,05 |
| Salbutamol  | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,01 | 0,04 |
| Diclofenac  | <MDL                | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 1,73 | 5,77 |
| Hydrochlorothiazide | <MDL              | <MDL                | <MDL                | <MDL                  | <MDL           | <MDL             | 0,01 | 0,02 |

**CONCLUSION**

After the detailed research of PhACs and EDCs in the fish tissues (Rutilus prespensis, Squalius platyceps, Scardinius knezevici, Chondrostoma ohridanum, Cyprinus carpio and Alburnus scoranza) it is determined that the concentration of researched distributors are found in MDL and MQL. Examined fish are a part of Skadar lake fauna, and are typical representatives. Economically, they are the most important fish. Exception to this research is detected concentration of triclosan (14,1±1,3 ng/g) in the tissue of fish Alburnus scoranza. It is considered that the specific habitat and diet of this fish caused these results in its muscle tissue. Further detailed research is needed to establish the source of the PhACs and EDCs.

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