Overexpression of the PPAR-γ protein in primary Ta/T1 non-muscle-invasive urothelial carcinoma

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Abstract. Peroxisome proliferator-activated receptor-γ (PPAR-γ) is a well-known nuclear receptor that is activated in the nucleus to regulate several transcription factors. Its expression patterns have been examined in various types of cancer. The present study investigated the expression patterns of PPAR-γ in non-muscle-invasive urothelial carcinoma. The expression rates of PPAR-γ, p53 and Ki-67 were compared to determine whether PPAR-γ may be considered as an immunobiomarker for bladder cancer. The intensity and extent of PPAR-γ expression were evaluated in 79 cases of non-muscle-invasive urothelial carcinoma (30 cases of papillary carcinoma low-grade, 30 cases of high-grade and 19 cases of carcinoma in situ) and 30 non-malignant cases. The nuclear overexpression of PPAR-γ was frequently observed in non-muscle-invasive urothelial carcinoma (63/79 cases) but was rarely detected in non-malignant cases (2/30 cases). The histological proliferation types of non-muscle-invasive urothelial carcinoma revealed that PPAR-γ was more frequently overexpressed in papillary carcinoma (54/60 cases) than in carcinoma in situ (9/19 cases). Immunohistochemical staining demonstrated that PPAR-γ was more useful as an immunobiomarker than p53 or Ki-67 (diagnostic odds ratios; 55.13, 16.82 and 11.13, respectively). In summary, this study demonstrated that the expression patterns of PPAR-γ were associated with histological proliferation type and that PPAR-γ was expressed in the nuclei of papillary carcinoma cells. These findings suggested that immunohistochemical staining for PPAR-γ may be used to comprehensively detect non-muscle-invasive urothelial carcinoma.

Introduction

Bladder cancer is the ninth most common cancer worldwide that ranks 13th for mortality rate, and ~430,000 cases of bladder cancer are reported each year (1). Smoking is the highest risk factor for bladder cancer, accounting for 50% of all cases (2). Urothelial carcinoma (UC) is a common histological type of bladder cancer that includes non-muscle-invasive UC (pathological stages Ta, T1 and Tis) and muscle-invasive UC (pathological stages T2 and higher) (3). Treatment guidelines for UC recommend the assessment of muscle invasiveness. Patients with Ta and T1 non-muscle-invasive UC undergo transurethral resection of bladder tumor (TURBT) for diagnostic and therapeutic purposes (4). Bacillus Calmette-Guerin (BCG) therapy is conducted for Tis UC, while a combination of cystectomy, chemoradiotherapy and radiation therapy is used to treat muscle-invasive UC (4). Non-muscle-invasive UC is separated into two distinct categories based on tumor growth as follows: Papillary and flat non-muscle-invasive UC. Furthermore, ~70-75% of primary UC cases are papillary carcinoma, whereas ~1-3% are the pure form of flat carcinoma. Non-muscle-invasive papillary UC can be distinguished as low grade (NMIPUC-L) and high grade (NMIPUC-H) based on architectural and cytological features (3).

Peroxisome proliferator-activated receptors (PPARs) are nuclear hormone receptors that comprise three subtypes: PPAR-α, PPAR-γ and PPAR-δ. The PPAR-γ protein has been detected in adipose tissue (5). The relationship between the expression of PPAR-γ and colon cancer has attracted increasing attention (6-8). It was demonstrated that PPAR-γ induces cell differentiation, arrests cell growth and reduces tumor growth rate in colon cancer. PPAR-γ ligands are expected to become promising therapeutic agents for chemoprevention and treatment. However, previous studies on PPAR-γ activation using

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several agonists have not provided consistent findings, such as the tumor suppressive or oncogenic role of PPAR-\(\gamma\), in a heterogeneous nature in bladder cancer (9,10). Furthermore, limited information is currently available on the expression of PPAR-\(\gamma\) in carcinoma in situ (CIS), such as flat carcinoma. Therefore, the relationship between PPAR-\(\gamma\) expression and the histological proliferation type in UC remains unclear. The present study aimed to investigate the expression of PPAR-\(\gamma\) in non-muscle-invasive UC, including CIS, and to compare it with that in normal urinary epithelial cells to clarify whether PPAR-\(\gamma\) may be used as an immunobiomarker in urothelial carcinoma.

**Materials and methods**

**Subjects.** Tissue samples, including TURBT and biopsy samples, were collected from 30 non-malignant cases and 79 non-muscle-invasive UC cases (NMIPUC-L 30 cases, NMIPUC-H 30 cases, and CIS 19 cases) at the Shikoku Cancer Center between April 2013 and March 2018. Samples from patients with inflammation in the bladder urothelium or those who recovered from UC following TURBT and BCG therapy were included as non-malignant cases. A histological diagnosis was based on the World Health Organization (WHO) classification of specimens stained using hematoxylin and eosin (3). The present study was approved by the Institutional Research Ethics Committee of the Shikoku Cancer Center (Ehime, Japan) and Kagawa Prefectural University of Health Sciences (Kagawa, Japan).

**Immunohistochemical staining.** Formalin-fixed paraffin-embedded tissue samples were cut into 4-μm-thick sections. All samples were rehydrated and deparaffinized using EZ buffer (Roche Diagnostics). Antigen masking was removed using pH 8.5 CC1 buffer (Roche Diagnostics) at 95°C for 64 min. All samples were then incubated with H\(2\)O\(_2\) (Roche Diagnostics) to block endogenous peroxidase activity. Sections were incubated with primary antibody against PPAR-\(\gamma\) (mouse monoclonal; Santa Cruz Biotechnology, Inc.; cat. no. sc-7273; 1:200) at 36°C for 32 min. The I-VIEW DAB Benchmark ULTRA (Roche Diagnostics) was used as an automatic immunostainer for immunohistochemical processes. To demonstrate the diagnostic utility of PPAR-\(\gamma\) in non-muscle-invasive UC cases, staining for p53 (mouse monoclonal; Agilent Technologies, Inc.; cat. no. M7001; Ready-to-use) and Ki-67 (mouse monoclonal; Agilent Technologies, Inc.; cat. no. M7240; 1:100) was performed on the same non-muscle-invasive UC and non-malignant samples. The p53 and Ki-67 immunohistochemical protocols were the same as the PPAR-\(\gamma\) immunohistochemical protocol. The expression pattern of PPAR-\(\gamma\) in the urinary bladder. The following PPAR-\(\gamma\) expression patterns were observed in the urinary bladder: Nuclear expression (Fig. 1A), nuclear-cytoplasmic expression (Fig. 1B), cytoplasmic expression (Fig. 1C) and no expression (Fig. 1D). The localization of PPAR-\(\gamma\) protein was significantly different between non-muscle-invasive UC cases and non-malignant cases (P<0.0001; Table I). PPAR-\(\gamma\) was mainly expressed in the nuclei of tumor cells in non-muscle-invasive UC, including NMIPUC-L (Fig. 2A), NMIPUC-H (Fig. 2B) and CIS (Fig. 2C). Conversely, PPAR-\(\gamma\) was partly expressed in the cytoplasm of urinary epithelial cells in non-malignant cases (Fig. 2D). Statistical analyses demonstrated that the nuclear overexpression of PPAR-\(\gamma\) was significantly higher in non-muscle-invasive UC cases compared with non-malignant cases (P<0.0001; Table II). The PPAR-\(\gamma\)-positive expression was significantly lower in CIS cases as flat carcinoma compared with NMIPUC-L and -H as papillary carcinoma (P=0.0002; Table III). In addition, PPAR-\(\gamma\) intensity in CIS with PPAR-\(\gamma\)-positive tended to be weak (Fig. 3). The PPAR-\(\gamma\)-positive expression was associated with the pathological stage (P=0.0003), but not with age, sex or histological grades (Table III).
Genomic alteration of PPAR-γ in bladder cancer. Using the TCGA datasets of bladder cancer and cBioPortal online tool to analyze PPAR-γ gene mutations or copy number alterations, alteration rates were 16.99% (70/412 cases; TCGA, Cell 2017), 16.79% (22/131 cases; TCGA, Nature 2014) and 16.3% (67/411 cases; TCGA, PanCancer Atlas; Fig. 4). In addition, PPAR-γ gene amplification accounted for most changes, with amplification rates of 13.59% (56/412 cases; TCGA, Cell 2017), 14.50% (19/131 cases; TCGA, Nature 2014), and 12.65% (52/411 cases; TCGA, PanCancer Atlas; Fig. 4).

Comparison of PPAR-γ, p53 and Ki-67 values as immunobiomarkers for non-muscle-invasive UC. Fig. 5 shows representative immunohistochemical staining for PPAR-γ (Fig. 5A, D, G and J), p53 (Fig. 5B, E, H and K) and Ki-67 (Fig. 5C, F, I and L). Immunohistochemical staining for PPAR-γ showed the highest sensitivity (79.7%). The diagnostic odds ratio (DOR) was 55.13 (Table IV). The expression of these immunobiomarkers was significantly higher in non-muscle-invasive UC cases compared with non-malignant cases. However, the PPAR-γ-positive expression did not significantly differ among the histological grades of non-muscle-invasive papillary UC (NMIPUC-L; 86.7%, NMIPUC-H; 93.3%. P=0.3980; Table V). Conversely, the aberrant p53 and high Ki-67 expression were significantly lower in NMIPUC-L than in NMIPUC-H. Furthermore, only the PPAR-γ positivity clearly distinguished NMIPUC-L from non-malignant cases (P<0.0001; Table V). Five out of 19 CIS cases were positive for PPAR-γ or had wild-type p53, whereas 17 were positive for PPAR-γ or had aberrant p53 (Table VI).

Discussion

The present study demonstrated that the localization, intensity, extent and genomic alteration of PPAR-γ expression significantly differed between non-muscle-invasive UC and non-malignant cases. The expression pattern of PPAR-γ in CIS suggested a relationship with the histological proliferative type but not the histological grade. Furthermore, the present study evaluated the usefulness of PPAR-γ, p53
Table II. Nuclear expression and cytoplasmic expression of PPAR-γ in non-muscle-invasive papillary urothelial carcinoma (low and high-grades), flat carcinoma and non-malignant cases.

| PPAR-γ expression | Nuclear score | Cytoplasmic score |
|-------------------|---------------|-------------------|
|                   | ≥4 | <4 | P-value | ≥4 | <4 | P-value |
| Non-muscle-invasive urothelial carcinoma cases (n=79) | 63 | 16 | <0.0001 | 6 | 73 | 0.0007 |
| NMIPUC low-grade and high-grade cases (n=60) | 54 | 6 | <0.0001 | 2 | 58 | <0.0001 |
| Flat carcinoma\textsuperscript{a} cases (n=19) | 9 | 10 | 0.0009 | 4 | 15 | 0.3575 |
| Non-malignant cases (n=30) | 2 | 28 | | 10 | 20 | |

\textsuperscript{a}Flat carcinoma is carcinoma in situ. NMIPUC, non-muscle-invasive papillary urothelial carcinoma; PPAR-γ, peroxisome proliferator-activated receptor-γ.

Figure 2. Representative images of the expression of PPAR-γ in urinary bladder tissues. (A) Non-muscle-invasive papillary urothelial carcinoma (low-grade), (B) non-muscle-invasive papillary urothelial carcinoma (high-grade), (C) carcinoma in situ and (D) non-malignant cases. Magnification, x400. Scale bar, 50 µm.

and Ki-67 as immunobiomarkers. The nuclear expression of PPAR-γ was significantly higher in non-muscle-invasive UC compared with non-malignant cases. In addition, PPAR-γ was more efficient for the detection of non-muscle-invasive UC than p53 and Ki-67. These results provided evidence for the potential role of PPAR-γ as an immunobiomarker in non-muscle-invasive UC.

In the present study, PPAR-γ showed different expression patterns in non-muscle-invasive UC and non-malignant cases. We considered PPAR-γ protein as being overexpressed in nuclei of urinary bladder tissues with malignant transformation. A previous study reported the nuclear PPAR-γ positive staining in colorectal tissues regardless of whether the tissue was malignant or not (27). In ovarian tumors, Zhang et al (11) reported a significant difference in the expression of PPAR-γ between the normal epithelium and malignant tumors and demonstrated that PPAR-γ was overexpressed in nuclei along with disease progression. In the present study, tumor cells of NMIPUC cases showed moderate cytoplasmic expression and high nuclear expression. The cytoplasmic expression of PPAR-γ was inversely associated with nuclear expression, with that in non-malignant cases being significantly higher than that in non-muscle-invasive UC. These results indicated that PPAR-γ was overexpressed in nuclei with malignant transformation. These differences in the expression patterns of PPAR-γ between normal urinary epithelial cells and tumor cells reflected the malignant transformation. PPAR-γ is a nuclear receptor that is activated in the nucleus to regulate...
several transcription factors. PPAR-γ ligands induce apoptosis in various carcinomas (28,29). In colon cancer, the PPAR-γ ligand 15-deoxy-Δ12,14-prostaglandin J2 (15d-PGJ2) was shown to inhibit the activity of nuclear factor-κB (NF-κB) and reduce the expression of the Bcl-2 protein, ultimately leading to apoptosis (30‑32). Furthermore, 15d-PGJ2, which is one of the natural ligands for PPAR-γ, can inhibit the growth of neoplastic urothelial cells (33). Bcl-2 and its transcription factor, NF-κB, have been suggested to inhibit tumor proliferation in UC by activating PPAR-γ. Previous studies demonstrated that PPAR-γ activation can arrest the cell cycle; however, most of the study materials examined were muscle‑invasive UC (9,34). The relationships between the expression of PPAR-γ, apoptosis, cell cycle and malignant transformation remain controversial. Further investigation is therefore required to elucidate the PPAR-γ pathway in UC.

Table III. Association between patient clinicopathological characteristics and the nuclear expression of PPAR-γ in non‑muscle‑invasive urothelial carcinoma.

|                          | Cases no. | PPAR-γ expression |                  | P-value   |
|--------------------------|-----------|-------------------|------------------|-----------|
|                          |           | Positive          | Negative         |           |
| Mean age ± standard deviation, years |           | 74.0±8.11         | 70.9±8.33        | 0.3926    |
| Sex                      |           |                   |                  |           |
| Male                     | 63        | 50                | 13               | 0.866     |
| Female                   | 16        | 13                | 3                |           |
| Histological grade       |           |                   |                  |           |
| Non-muscle‑invasive urothelial carcinoma, low‑grade | 30        | 26                | 4                | 0.2204    |
| Non-muscle‑invasive urothelial carcinoma, high‑grade<sup>a</sup> | 49        | 37                | 12               |           |
| Histological proliferation type |           |                   |                  |           |
| Papillary carcinoma<sup>b</sup> | 60        | 54                | 6                | 0.0002    |
| Flat carcinoma<sup>c</sup> | 19        | 9                 | 10               |           |
| Pathological stage       |           |                   |                  |           |
| Tis                      | 19        | 9                 | 10               | 0.0003    |
| Ta                       | 45        | 41                | 4                | 0.619     |
| T1                       | 15        | 13                | 2                |           |

<sup>a</sup>High‑grade includes the non‑muscle‑invasive papillary urothelial carcinoma, high‑grade (n=30) and carcinoma in situ (n=19). <sup>b</sup>Papillary carcinoma includes the non‑muscle‑invasive papillary urothelial carcinoma, low‑grade (n=30) and high‑grade (n=30). <sup>c</sup>Flat carcinoma is carcinoma in situ. PPAR‑γ, peroxisome proliferator‑activated receptor‑γ.

Figure 3. Intensity and extent of PPAR‑γ immunohistochemical staining in non‑muscle‑invasive papillary urothelial carcinoma and carcinoma in situ with positivity for PPAR‑γ. NMIPUC, non‑muscle‑invasive papillary urothelial carcinoma; CIS, carcinoma in situ; PPAR‑γ, peroxisome proliferator‑activated receptor‑γ.
expression assay. Regarding the relationship between PPAR-γ expression and cancer histological grades, Mylona et al (37) and Nakashiro et al (33) indicated that the nuclear expression of PPAR-γ was inversely correlated with histological grades in UC. However, as presented in Table III, a relationship was not observed between the nuclear overexpression of PPAR-γ in non-muscle-invasive UC and histological grades. Histological grades are diagnosed from morphology based on architectural and cytological features. Furthermore, the WHO classification defines low-grade UC as papillary carcinoma, whereas high-grade UC includes papillary, flat and infiltrating types. Since flat carcinoma was not sufficiently examined in previous studies, the relationship between PPAR-γ and histological proliferation types was not investigated in detail. The results from the present demonstrated that PPAR-γ expression significantly differed among pathological stages. CIS corresponded to Tis pathological stage while papillary carcinoma corresponded to Ta or T1 pathological stages. Therefore, the results from statistical analyses appeared to be dependent on the histological proliferation type and not on the pathological stage. Statistical analyses of PPAR-γ expression in Ta and T1 did not reveal any significant differences. Regarding the PPAR-γ expression in muscle-invasive UC, a previous study reported that PPAR-γ expression levels are lower in muscle-invasive UC cases than in non-muscle-invasive UC cases (37). This finding supports our results showing that PPAR-γ expression might be associated with the proliferation type.

![Figure 4. Genetic alteration analysis of peroxisome proliferator-activated receptor-γ in patients with bladder cancer using cBioPortal. TCGA, The Cancer Genome Atlas.](image-url)
An immunohistochemical method to detect UC has not yet been established. Previous studies reported a relationship between UC and certain immunobiomarkers, such as p53, a tumor suppressor protein, and Ki-67, a cell proliferation marker. However, the sensitivity of p53 as an immunobiomarker was 26-59%, while that of Ki-67 was 16-58% for non-invasive papillary UC, including low and high grades, with the former grade not being detected by these immunobiomarkers (38-42). The sensitivities of p53 and Ki-67 in the present study were consistent with previous findings; however, the sensitivity and DOR of p53 and Ki-67 were dependent on the low frequency of the aberrant type/high expression in NMIPUC-L. Therefore, PPAR-γ as an immunobiomarker may be useful for detecting non-muscle-invasive UC despite the histological grade. In the present study, p53 showed the highest sensitivity as an immunobiomarker for CIS (63.2%, 12/19). The aberrant type of p53 has been widely investigated and used as an immunobiomarker to detect CIS (43). Based on the data shown in the present study, we considered CIS to have been comprehensively detected using a combination of PPAR-γ and p53 as immunobiomarkers rather than using p53 alone.

A limitation of the present study was that the molecular biological analysis did not include muscle-invasive UC cases or UC cell lines. Therefore, an association was not observed between UC invasiveness and PPAR-γ expression. In addition, we did not obtain clinical data on recurrence because of the limited number of PPAR-γ-negative cases in UC presenting with recurrence following TURBT. However, non-muscle-invasive UC frequently relapses, and we speculated that PPAR-γ may serve an important role in this process. Although further investigation is required, we herein attempted to clarify the usefulness of PPAR-γ as an immunobiomarker in samples from patients with non-muscle-invasive UC.

A routine and less invasive method to detect UC is urinary cytology; however, it has not yet been established as a useful screening method for UC due to its low sensitivity. Meuleman and Delaere (44) reported that the diagnostic findings of urinary cytology were subject to UC differentiation level and infiltrating stage. In cytological samples, difficulties are associated with distinguishing NMIPUC-L from normal urinary epithelial cells based on a morphological diagnosis under a microscope because morphologically, NMIPUC-L negligibly exhibits nuclear atypia and pleomorphism (45,46). According to NCCN Clinical Practice Guidelines in Oncology (4), ~70-75% of primary UC cases are NMIPUC-L or NMIPUC-H. Therefore, NMIPUC-L and NMIPUC-H are frequently encountered in urinary cytology but are not

Figure 5. Representative immunohistochemical staining for PPAR-γ, p53 and Ki-67. Immunohistochemical staining showing (A) PPAR-γ, (B) p53 and (C) Ki-67 in non-muscle-invasive papillary urothelial carcinoma (low-grade), (D) PPAR-γ, (E) p53 and (F) Ki-67 in non-muscle-invasive papillary urothelial carcinoma (high-grade), (G) PPAR-γ, (H) p53 and (I) Ki-67 in carcinoma in situ, and (J) PPAR-γ, (K) p53 and (L) Ki-67 in non-malignant cases. Magnification, x400. Scale bar, 50 µm. PPAR-γ, peroxisome proliferator-activated receptor-γ.
The results from the present study demonstrated that PPAR-\(\gamma\) immunohistochemical staining could detect more cancer cells than other immunobiomarkers for NMIPUC-L and NMIPUC-H. An ancillary diagnostic test using PPAR-\(\gamma\) immunocytochemical staining may effectively increase the accuracy of urinary cytology.

In summary, the present study demonstrated that expression patterns of PPAR-\(\gamma\) were associated with histological proliferation type and that PPAR-\(\gamma\) was expressed in the nuclei of papillary carcinoma cells. Immunohistochemical staining for PPAR-\(\gamma\) appeared to be more useful as an immunobiomarker for non-muscle-invasive UC than the other biomarkers examined. Although further investigation is needed, this study suggested that PPAR-\(\gamma\) immunobiomarker may be considered as a promising tool for UC early detection.

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Availability of data and materials

The datasets used during the present study are available from the corresponding author upon reasonable request.
Authors' contributions

ST, YT and EH designed the study. ST, YT, SH, HO, TM, TY and NT performed the experiments. ST, YT, SH and EH analyzed all data. ST and YT wrote the manuscript. HO, NT and EH confirm the authenticity of all raw data. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The present study was performed in accordance with the Declaration of Helsinki and was approved by the Ethics Committee of the Shikoku Cancer Center (Ehime, Japan; approval no. 2018-95) and Kagawa Prefectural University of Health Sciences (Kagawa, Japan; approval no. 291). In this retrospective study, the Institutional Review Board previously granted a waiver for written informed consent by publishing information on the study on the Home Page and providing the option to opt-out.

Patient consent for publication

Not applicable.

Competing interest

The authors declare that they have no competing interests.

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