Research Article

Causal Association of Thyroid Signaling with C-Reactive Protein: A Bidirectional Mendelian Randomization

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Received 8 July 2022; Revised 23 July 2022; Accepted 27 July 2022; Published 13 August 2022

Academic Editor: Jincheng Wang

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Context. Existing literature demonstrated that thyroid-stimulating hormone (TSH) and free thyroid hormone (fT4) were associated with the C-reactive protein (CRP), an inflammatory risk factor of cardiovascular diseases (CVD), but the causal relationship between them remained unclear. Methods. Based on the latest genome-wide association study summary data, bidirectional two-sample Mendelian randomization (MR) was employed to detect the causal relationship and effect direction between TSH, fT4, and CRP. Furthermore, in view of obesity being an important risk factor of CVD, obesity trait waist-hip ratio (WHR) and body mass index (BMI) were treated as the research objects in MR analyses for exploring the causal effects of TSH and fT4 on them, respectively. Results. Genetically increased CRP was associated with increased TSH ($\beta = -0.02$, $P = 0.011$) and with increased fT4 ($\beta = 0.043$, $P = 0.001$), respectively, but there was no evidence that TSH or fT4 could affect CRP. In further analyses, genetically increased TSH was associated with decreased WHR ($\beta = -0.081$, $P = 0.002$). Genetically increased BMI was associated with increased TSH ($\beta = 0.03$, $P = 0.028$) and with decreased fT4 ($\beta = -0.078$, $P = 1.05e-4$). Causal associations of WHR and BMI with thyroid signaling were not supported by weighted median analysis in sensitivity analyses. Conclusion. TSH and fT4 were increased due to the higher genetically predicted CRP. WHR was decreased due to the higher genetically predicted TSH. These findings will provide reference for the prevention and treatment of inflammation and metabolic syndrome.

1. Introduction

Subclinical thyroid disease is a common public health issue. In hypothyroidism due to thyroid dysfunction, serum thyroid-stimulating hormone (TSH) levels are appropriately elevated while serum free thyroxine (fT4) levels are within normal range [1, 2]. Subclinical hypothyroidism affects up to 10% of the adult population [3]. A lot of previous studies showed variation in TSH or fT4 may increase the risk of future cardiovascular diseases (CVD) [4–6]. Recently, many investigations indicated that even in normal thyroid function individuals, variation in TSH and fT4 was associated with an increased risk of CVD and metabolic diseases [7–9], including obesity [1]. Therefore, it is important and urgent to pay more attention to TSH and fT4.

C-reactive protein (CRP) is an acute-phase inflammatory protein, which has been traditionally utilized as a clinical marker of inflammation, infection, and tissue damage [10]. Generally, CRP exhibits elevated expression during inflammatory disorders, such as CVD, rheumatoid arthritis, and some acute or chronic infection [11, 12]. Recently, research outputs showed that minor CRP elevation could contribute to an increased future risk of major cardiovascular events [13, 14]. In addition, there was growing evidence that elevated CRP levels are associated with cancer disease risk [15, 16]. Hence, CRP measurements have potential
utility as a clinical tool in assessing disease status and progression, including CVD, some infections, and cancer. Therefore, with the important role of CRP, more studies are needed to understand the complex mechanism of CRP production.

Existing literature demonstrated that subclinical hypothyroidism may be associated with elevated high-sensitive CRP, although the clinical implications were uncertain [17–19]. Some researchers found that there was a significant positive correlation between TSH and CRP [17, 20]. Meanwhile, a Brazilian longitudinal study of adult health also investigated the association between TSH and CRP, but this study showed that TSH was not associated with CRP because of the existence of confounders [21]. So, it was controversial about the relationship between thyroid signaling and CRP. Similarly, there were researches investigating the association of fT4 with CRP [18, 22, 23], and their conclusions were also controversial. Therefore, the association between thyroid signaling and CRP is hard to uncover.

Furthermore, as an important public health problem, obesity is also an important risk factor for CVD [11]. Previous investigations revealed a significant relationship between CRP and obesity [24–26]. In obese and overweight adults, CRP levels are significantly increased [27]. So it is meaningful to study obesity traits, such as waist-hip ratio (WHR) and body mass index (BMI).

Correlation describes whether two variables "go together." However, the fact that two variables change together does not necessarily mean that we know whether one variable causes the other to change or vice versa [28]. Therefore, it is necessary to study causal association. To this end, one powerful method is Mendelian randomization (MR) [29], which uses genetic variants as instrumental variants (IVs) and has been widely used [30]. MR can minimize the influence of confounding factors on the causal association between two variables, exposure and outcome. Note both individual data and publicly available genome-wide association study (GWAS) summary statistics are applicable in MR analyses. Moreover, bidirectional two-sample MR can explore the nature and direction of the links between them.

To date, as far as we know, no studies investigated the causal associations of TSH and fT4 levels with CRP levels. In this paper, we studied the causal association between thyroid signaling and CRP level. To further detect the possible causes of CVD, we also studied two obesity traits, WHR and BMI. For this, we utilized summary data from the latest and largest GWASs [31–33] and inferred causality in bidirectional two-sample MR analyses.

2. Materials and Methods

2.1. Data Sources. The first is the source of summary data related to thyroid signaling. Summary data for TSH within reference range were obtained from a GWAS meta-analysis that is the largest GWAS on thyroid function to date, including 120000 subjects, with more than 22 million single nucleotide polymorphisms (SNPs) [31]. These data are accessed through the GWAS Catalog (https://www.ebi.ac.uk/gwas). Summary data for fT4 within reference range were obtained from a GWAS meta-analysis in up to 72167 individuals with 8 million SNPs [32], which can be downloaded on dbGap website under the accession number phs000930 (https://www.ncbi.nlm.nih.gov/gap).

Second is the source of summary data on inflammatory factor CRP. Summary data for CRP were obtained from a GWAS meta-analysis which is the largest data set on inflammatory factors lately, including 49839 subjects (CRP: mean = 4.114 (SD = 4.836)) [33]. These summary data can be available through the GWAS Catalog (https://www.ebi.ac.uk/gwas). Summary data for WHR were obtained from a GWAS meta-analysis in 694649 individuals of European ancestry with 2.7 million SNPs combining UK Biobank and GIANT [34]. Summary data for BMI were obtained from a GWAS meta-analysis which included about 700000 participants of European ancestry with 2.3 million SNPs from GIANT [35].

2.2. Two-Sample MR. We conducted bidirectional two-sample MR analyses using data published by GWAS (Figure 1). Because the data is public, there is no need of ethical review.

2.3. Selection of SNPs. Based on the GWAS results [31–33] on TSH, fT4, and CRP, we used independent SNPs which are strongly associated at a genome-wide significant level (P < 5 × 10−8) with TSH, fT4, and CRP, respectively. The selected SNPs were used as IVs in using MR method.

2.4. Statistical Analysis. In order to avoid the estimator bias caused by weak IVs as much as possible, we calculated the F statistic (F = R²exposure/SE²exposure) as a measure of strength for each SNP. According to the existing literature, criterion of F ≥ 10 was adopted for screening strong IVs (F statistic was in 30.01–1231.188 for TSH, 30.25–455.33 for fT4, and 27.94–528.51 for CRP) [36]. The primary analysis used to examine the causality between exposure and outcome was inverse-variance weighted (IVW) method [37]. IVW
The results of MR analyses using various analysis methods (IVW, MR-Median, MR-Egger) are presented for comparison. The number of SNPs indicates the number of genetic variants used as instrument variables for MR analysis. MR: Mendelian randomization; SNPs: single nucleotide polymorphisms; CI: confidence interval; IVW: inverse-variance weighted; MR-Median: weighted median method; MR-Egger: Egger regression.

For the estimated causal effect of the exposure on the outcome, a $P$ value of less than 0.05 was considered as statistically significant. Statistical analysis was performed with R package “MendelianRandomization” version 0.5.1 in R version 4.1.0.

3. Results

Use the MR analysis method in previous sections to explore the causal relationships between thyroid signaling, CRP, and obesity traits. The MR-Egger intercepts were insignificant ($P > 0.05$) in all analyses. The result diagram is shown in Figure 2. The diagram showed whether there was a causal relationship between two subjects and showed the magnitudes and directions of the causal relationships. All causal relationships are significant at $P \leq 0.05$.

3.1. Causal Relationships between Thyroid Signaling and CRP

The results of MR analyses between genetically predicted TSH and $fT4$ levels (exposure) and CRP levels (outcome) are presented in Figure 3. 87 SNPs were in consideration when we investigated the causal association between TSH and CRP. Based on this analysis, we found neither serum TSH nor $fT4$ levels could cause changes in CRP ($TSH: \hat{\beta} = 0.003, 95\% CI = -0.032–0.039, P = 0.856$; $fT4: \hat{\beta} = 0.003, 95\% CI = -0.084–0.089, P = 0.953$) (also, see details in Supplementary Table S1).

Method aggregated two or more IVs to minimize the variance of the weighted average, and the weight given to each IV was the inverse of the variance of the estimate [38]. Note that the estimated effect obtained by IVW may be biased, which may be due to the violation of one assumption of IV. Specifically, IVs and outcome are not only related through exposure but also directly related, which is termed as pleiotropy. We addressed the problem of pleiotropy in sensitivity analyses.

In sensitivity analyses, we assessed the robustness of IVW in two complementary sensitivity analyses with different assumptions about horizontal pleiotropy: weighted median (MR-Median) [38] and MR-Egger regression [39]. MR-Median yielded consistent causal effect estimates compared with IVW method. Egger intercept in MR-Egger represented the average horizontal pleiotropic effect across the IVs. We used $I^2$ statistic and Cochran’s $Q$ test to quantify heterogeneity across all SNPs. If the results indicated the presence of horizontal pleiotropy or significant heterogeneity suggesting pleiotropy [40], we calculated individual Q statistic for each SNP, and SNPs were identified as potential pleiotropic variants if their individual Q statistics exceeded the 95th percentile of the chi-square distribution with one degree of freedom [41–43]. After excluding these potential pleiotropic IVs, the IVW, MR-Median, and MR-Egger methods were performed on the remaining IVs.

Figure 2: Causal effects between thyroid signaling, CRP, and obesity traits. TSH and $fT4$ are shown in pink ovals, CRP is shown in gray ovals, and BMI and WHR are shown in yellow ovals. The arrows’ direction denotes causal direction. The solid line and the dotted line, respectively, indicate whether the causal relationship is robust or not. The red and green arrows denote positive and negative causal relationships, respectively, and the number beside each arrow is the causal effects. All causal relationships are significant at $P \leq 0.05$. The results of MR analyses using various analysis methods (IVW, MR-Median, MR-Egger) are presented for comparison. The number of SNPs indicates the number of genetic variants used as instrument variables for MR analysis. MR: Mendelian randomization; SNPs: single nucleotide polymorphisms; CI: confidence interval; IVW: inverse-variance weighted; MR-Median: weighted median method; MR-Egger: Egger regression.

Figure 3: Causal effects of variation in TSH and $fT4$ levels on CRP. Presented $\hat{\beta}$s and CIs (horizontal lines and their corresponding numerical interval form on the right side) correspond to the effects of a one SD change in TSH or $fT4$ levels on the outcome CRP levels. The red and green arrows denote positive and negative causal relationships, respectively, and the number beside each arrow is the causal effects. All causal relationships are significant at $P \leq 0.05$. The results of MR analyses using various analysis methods (IVW, MR-Median, MR-Egger) are presented for comparison. The number of SNPs indicates the number of genetic variants used as instrument variables for MR analysis. MR: Mendelian randomization; SNPs: single nucleotide polymorphisms; CI: confidence interval; IVW: inverse-variance weighted; MR-Median: weighted median method; MR-Egger: Egger regression.

method aggregated two or more IVs to minimize the variance of the weighted average, and the weight given to each IV was the inverse of the variance of the estimate [38]. Note that the estimated effect obtained by IVW may be biased, which may be due to the violation of one assumption of IV. Specifically, IVs and outcome are not only related through exposure but also directly related, which is termed as pleiotropy. We addressed the problem of pleiotropy in sensitivity analyses.

In sensitivity analyses, we assessed the robustness of IVW in two complementary sensitivity analyses with different assumptions about horizontal pleiotropy: weighted median (MR-Median) [38] and MR-Egger regression [39]. MR-Median yielded consistent causal effect estimates compared with IVW method. Egger intercept in MR-Egger represented the average horizontal pleiotropic effect across the IVs. We used $I^2$ statistic and Cochran’s $Q$ test to quantify heterogeneity across all SNPs. If the results indicated the presence of horizontal pleiotropy or significant heterogeneity suggesting pleiotropy [40], we calculated individual Q statistic for each SNP, and SNPs were identified as potential pleiotropic variants if their individual Q statistics exceeded the 95th percentile of the chi-square distribution with one degree of freedom [41–43]. After excluding these potential pleiotropic IVs, the IVW, MR-Median, and MR-Egger methods were performed on the remaining IVs.
However, exchanging the exposure and outcome of interest in MR yielded different results (Figure 4). 27 SNPs were considered as IVs when we investigated the causal association between CRP and TSH. There was some evidence that higher CRP levels might cause higher TSH levels ($\beta = 0.020$, 95% CI = 0.005–0.036, $P = 0.011$), which was confirmed in sensitivity analyses using the MR-Median and MR-Egger method (see Supplementary Table S2).

3.2. Causal Relationships between Thyroid Signaling and Obesity Traits. Due to elevated CRP levels in overweight and obese adults [27], we also wanted to know whether there exist causal associations between thyroid signaling and obesity traits. Therefore, we conducted MR analyses of TSH and $fT4$ on obesity traits, respectively (Figure 5). MR analyses

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\begin{align*}
\text{TSH (Outcome)} & \quad \text{SNPs, N} \\
\text{IVW} & \quad 27 \quad 0.020 (0.005, 0.036) \\
\text{MR-Median} & \quad 27 \quad 0.030 (0.006, 0.054) \\
\text{MR-Egger} & \quad 27 \quad 0.040 (0.005, 0.084)
\end{align*}
\]

\[
\begin{align*}
\text{fT4 (Outcome)} & \quad \text{SNPs, N} \\
\text{IVW} & \quad 35 \quad 0.043 (0.018, 0.068) \\
\text{MR-Median} & \quad 35 \quad 0.055 (0.025, 0.086) \\
\text{MR-Egger} & \quad 35 \quad 0.085 (0.028, 0.143)
\end{align*}
\]
showed higher genetically predicted TSH could cause decreased WHR levels ($\beta = -0.02$, 95% CI $=-0.030$ to $-0.009$, $P = 3.99 \times 10^{-4}$), and MR-Median also led to similar results (see Supplementary Table S3 for more information).

For the association between obesity traits and thyroid signals (Figure 6), MR analyses showed higher genetically predicted BMI could cause higher TSH levels ($\beta = 0.081$, 95% CI $=-0.133$ to $-0.029$, $P = 0.002$), and higher genetically predicted BMI could cause higher TSH levels ($\beta = 0.030$, 95% CI $= 0.003$ to $0.057$, $P = 0.028$) and lower $t$T4 levels ($\beta = 0.02$, 95% CI $= 0.018$ to $0.035$, $P = 1.05 \times 10^{-4}$). This causal relationship was not robust because it is not supported by MR-Median (see Supplementary Table S4).

### 4. Discussion

In this study, the bidirectional two-sample MR analyses between thyroid signaling (TSH and $t$T4) and CRP levels were accomplished based on the current largest GWAS summary statistics. We studied the causal relationships between TSH and $t$T4 levels and CRP levels and found TSH and $t$T4 levels could be affected by CRP, whereas TSH and $t$T4 levels could not affect CRP levels. Furthermore, we found some evidence that there were associations between obesity traits (BMI and WHR) and $t$T4 levels. We also found that TSH levels could be significantly affected by BMI.

CRP responds quickly to inflammatory processes and is utilized as one of the best inflammatory markers. Various research results showed that there was a significant positive correlation between TSH and CRP [17, 44, 45]. However, their underlying causality was still unclear. A prospective study indicated that patients with subclinical hypothyroidism had increased levels of signs of low-grade inflammation (CRP levels) [20, 46]. On the other hand, the conclusions of many studies were not consistent with this prospective study. For example, some authors believed that serum CRP was not significantly affected by the thyroid dysfunction’s degree [47]. An observational study found that CRP was not correlated with $t$T4 and TSH [48]. These studies indicated that further evidence was needed to determine the causal link between TSH and $t$T4 levels and CRP levels. In this study, based on MR analysis results, we found that there was a causal association between CRP and thyroid signaling (TSH and $t$T4). TSH and $t$T4 levels could be positively affected by CRP levels, but not vice versa. The underlying cause of CRP affecting thyroid signaling is still unclear, possibly because severe inflammation may significantly affect the thyroid gland, leading to changes in thyroid signaling. Besides, we thought there were some potential effects of inflammation on deiodinase activity. Inflammation (elevated in CRP levels) which was related to infection or injury led to a reduction in deiodinase activity. This results in decreased conversion of $t$T4 to $t$T3, leading to high $t$T4 [17]. In the future, the causal relationship between CRP levels and TSH and $t$T4 levels may be confirmed with larger populations and more precise statistical methods.

Interestingly, in a Brazilian longitudinal study of adult health, obesity was considered as one of the most important confounders in the association study between TSH and CRP [21]. Some researches showed that CRP was correlated with obesity and the role of obesity in inflammation cannot be ignored [49]. This promoted us to study further the obesity traits.
In the subsequent MR analyses, we found that increased TSH could cause decreased WHR. In reverse MR analyses, increased WHR and BMI could cause decrease in $fT4$, and increased BMI could cause increase in TSH. Previous literature showed that lower $fT4$ was consistently associated with obesity in healthy euthyroid people [50, 51]. One research indicated that serum $fT4$ levels were negatively correlated with BMI and serum TSH levels were positively correlated with WHR and BMI [52]. It was suggested that the increase in $fT3$ levels in obese people may be a compensatory mechanism for the fat accumulation increase [53]. In obese people, thyroxine 5-deiodinase increased activity, inducing the increased peripheral conversion of $fT4$ to $fT3$ [49, 54]. The lower $fT4$ in obese and overweight people might partially result from this cause. These were consistent with the results of our MR study. Conclusion of a recent MR analysis was also consistent with our study; i.e., genetically predicted BMI was inversely associated with $fT4$ levels [55].

Another MR analysis pointed out that TSH could be significantly elevated by the genetically driven BMI, while $fT4$ could not be affected by BMI [56]. Notice in our study, we used the latest and largest GWAS summary data, where $fT4$ cohorts included nearly 70000 participants. Moreover, we performed sensitivity analyses to exclude pleiotropic and heterogeneous IVs, because these heterogenous SNPs could partially result in bias in MR analysis. In our analysis, TSH and $fT4$ both could be affected by BMI.

Advantages of this study design were that (1) GWAS data were freely available obtained from the largest recent GWAS on TSH, $fT4$, and CRP, respectively; (2) sensitivity analyses were performed in order to reduce potential bias resulting from potential pleiotropic and heterogeneous IVs. It is the first time to reach a conclusion based on MR analysis that higher genetically predicted CRP may induce an increase in TSH and $fT4$. However, this study has certain limitations. (1) Due to the accession of the public databases, we used people of diverse ancestry for CRP and people of European ancestry for thyroid signaling and obesity traits. MR analysis for population-stratification and other populations should be considered if related data can be available; (2) generally speaking, thyroid function is sex-specific; due to the limitation of TSH and $fT4$ summary data, we did not perform the sex-specific MR analyses.

Taken together, the bidirectional MR study demonstrated that higher TSH and $fT4$ levels were causally affected by higher CRP levels, but not vice versa. Further MR analyses provided evidence that higher obesity traits could cause lower $fT4$ and higher BMI could cause higher TSH.

Data Availability

All data generated or analyzed during this study are included in this published article or in the data repositories listed in References. Summary statistic data for genetic associations with thyroid signaling have been contributed by the thyroid GWAS meta-analysis of Hunt and the ThyroidOmics consortium. Summary statistic data for genetic association with body mass index and waist-hip ratio have been contributed by the GIANT consortium and MEGASTROKE consortium. Summary statistic data for genetic association with C-reactive protein have been contributed by the Population Architecture Using Genomics and Epidemiology study.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

T.L., H.G., and Y.H. conceptualized the study. T. L., Y. W. and Y. H. were responsible for the methodology. Y.H. was responsible for the funding acquisition. T.L. was responsible for the investigation. Y.H. was responsible for the project administration. T.L. was responsible for the resources. T.L. was responsible for the software. Y.W. and Y.H. supervised the study. T.L. and S.Y. validated the study. T.L. was responsible for the writing—original draft. H. G., Z. W., S. Y., and Y. H. were responsible for the writing, review and editing. All authors have read and agreed to the published version of the manuscript. Tingting Li, Haigang Geng, and Yuquan Wang contributed equally to this work.

Acknowledgments

This study was supported by grants to YH from the National Natural Science Foundation of China (grants nos. 11971117 and 11571082).

Supplementary Materials

Supplementary Table S1: results of Mendelian randomization analyses between genetically predicted thyroid-stimulating hormone (TSH) and free thyroid hormone ($fT4$) levels (exposure) and C-reactive protein (CRP) levels (outcome). Supplementary Table S2: results of Mendelian randomization analyses between genetically predicted CRP levels (exposure) and TSH and $fT4$ levels (outcome). Supplementary Table S3: results of Mendelian randomization analyses between genetically predicted TSH and $fT4$ levels (exposure) and obesity traits (outcome). Supplementary Table S4: results of Mendelian randomization analyses between genetically predicted obesity traits (exposure) and TSH and $fT4$ levels (outcome). (Supplementary Materials)

References

[1] M. A. Michalaki, A. G. Vagenakis, A. S. Leonardou et al., “Thyroid function in humans with morbid obesity,” Thyroid, vol. 16, no. 1, pp. 73–78, 2006.
[2] B. Biondi, A. R. Cappola, and D. S. Cooper, “Subclinical hypothyroidism,” Journal of the American Medical Association, vol. 322, no. 2, pp. 153–160, 2019.
[3] J. G. Hollowell, N. W. Staehling, W. D. Flanders et al., “Serum TSH, T4, and thyroid antibodies in the United States population (1988 to 1994): National Health and Nutrition Examination Survey (NHANES III),” The Journal of Clinical Endocrinology & Metabolism., vol. 87, no. 2, pp. 489–499, 2002.
[4] N. Rodondi, W. P. J. den Elzen, D. C. Bauer et al., “Subclinical hypothyroidism and the risk of coronary heart disease and mortality,” Journal of the American Medical Association, vol. 304, no. 12, pp. 1365–1374, 2010.

[5] C. Floriani, B. Gencer, T.-H. Collet, and N. Rodondi, “Subclinical thyroid dysfunction and cardiovascular diseases: 2016 update,” European Heart Journal, vol. 39, no. 7, pp. 503–507, 2018.

[6] A. E. Hak, H. A. Pols, T. J. Visser, H. A. Dreuxage, A. Hofman, and J. C. Witteman, “Subclinical hypothyroidism is an independent risk factor for atherosclerosis and myocardial infarction in elderly women: the Rotterdam study,” Annals of Internal Medicine, vol. 132, no. 4, pp. 270–278, 2000.

[7] L. J. van Tienhoven-Wind and R. P. Dullaart, “Low–normal thyroid function and the pathogenesis of common cardiometabolic disorders,” European Journal of Clinical Investigation, vol. 45, no. 5, pp. 494–503, 2015.

[8] B. O. Åsvold, T. Bjøro, C. Platou, and L. J. Vatten, “Thyroid function and the risk of coronary heart disease: 12-year follow-up of the HUNT study in Norway,” Clinical Endocrinology, vol. 77, no. 6, pp. 911–917, 2012.

[9] N. Takamura, A. Akiizhanova, N. Hayashida et al., “Thyroid function is associated with carotid intima-media thickness in euthyroid subjects,” Atherosclerosis, vol. 204, no. 2, pp. e77–e81, 2009.

[10] M. A. Albert, E. Danielson, N. Rifai, P. M. Ridker, and PRINCE Investigators, “Effect of statin therapy on C-reactive protein levels: the pravastatin inflammation/CRP evaluation (PRINCE): a randomized trial and cohort study,” Journal of the American Medical Association, vol. 286, no. 1, pp. 64–70, 2001.

[11] N. R. Sproston and J. J. Ashworth, “Role of C-reactive protein at sites of inflammation and infection,” Frontiers in Immunology, vol. 9, no. 754, 2018.

[12] T. P. Erlinger, E. A. Platz, N. Rifai, and K. J. Helzlsouer, “C-reactive protein and the risk of incident colorectal cancer,” Journal of the American Medical Association, vol. 291, no. 5, pp. 585–590, 2004.

[13] P. M. Ridker, M. Cushman, M. J. Stampfer, R. P. Tracy, and C. H. Hennekens, “Inflammation, aspirin, and the risk of cardiovascular disease in apparently healthy men,” New England Journal of Medicine, vol. 336, no. 14, pp. 973–979, 1997.

[14] P. M. Ridker, C. H. Hennekens, J. E. Buring, and N. Rifai, “C-reactive protein and other markers of inflammation in the prediction of cardiovascular disease in women,” New England Journal of Medicine, vol. 342, no. 12, pp. 836–843, 2000.

[15] P. C. Hart, I. M. Rajab, M. Alabraheem, and L. A. Potempa, “C-reactive protein and cancer—diagnostic and therapeutic insights,” Frontiers in Immunology, vol. 11, article 595835, 2020.

[16] K. H. Allin and B. G. Nordestroegaard, “Elevated C-reactive protein in the diagnosis, prognosis, and cause of cancer,” Critical Reviews in Clinical Laboratory Sciences, vol. 48, no. 4, pp. 155–170, 2011.

[17] R. Sharma, T. K. Sharma, G. Kaushik, S. Sharma, S. Vardey, and M. Sinha, “Subclinical hypothyroidism and its association with cardiovascular risk factors,” Clinical Laboratory, vol. 57, no. 9-10, pp. 719–724, 2011.

[18] M. Christ-Crain, C. Meier, M. Guglielmetti et al., “Elevated C-reactive protein and homocysteine values: cardiovascular risk factors in hypothyroidism? A cross-sectional and a double-blind, placebo-controlled trial,” Atherosclerosis, vol. 166, no. 2, pp. 379–386, 2003.

[19] N. Boulman, Y. Levy, R. Leiba et al., “Increased C-reactive protein levels in the polycystic ovary syndrome: a marker of cardiovascular disease,” The Journal of Clinical Endocrinology & Metabolism, vol. 89, no. 5, pp. 2160–2165, 2004.

[20] A. Tuzcu, M. Bahceci, D. Gokalp, Y. Tuzun, and K. Gunes, “Subclinical hypothyroidism may be associated with elevated high-sensitive C-reactive protein (low grade inflammation) and fasting hyperinsulinemia,” Endocrine Journal, vol. 52, no. 1, pp. 89–94, 2005.

[21] E. J. F. P. de Miranda, M. S. Bittencourt, I. S. Santos, P. A. Lotufo, and I. M. Benseñor, “Thyroid function and high-sensitivity C-reactive protein in cross-sectional results from the Brazilian longitudinal study of adult health (Elsa-Brasil): effect of adiposity and insulin resistance,” European Thyroid Journal, vol. 5, no. 4, pp. 240–246, 2016.

[22] C. Jublanc, E. Bruckert, P. Giralt et al., “Relationship of circulating C-reactive protein levels to thyroid status and cardiovascular risk in hyperlipidemic euthyroid subjects: low free thyroxine is associated with elevated hsCRP,” Atherosclerosis, vol. 172, no. 1, pp. 7–11, 2004.

[23] S. Singh, “Serum lipids, tHcy, hs-CRP, MDA and PON-1 levels in SCH and overt hypothyroidism: effect of treatment,” Acta Biomedica Atenei Parmensis, vol. 85, no. 2, pp. 127–134, 2014.

[24] J. R. Greenfield, K. Samaras, A. B. Jenkins et al., “Obesity is an important determinant of baseline serum C-reactive protein concentration in monozygotic twins, independent of genetic influences,” Circulation, vol. 109, no. 24, pp. 3022–3028, 2004.

[25] M. Hiura, T. Kikuchi, K. Nagasaki, and M. Uchiyama, “Elevation of serum C-reactive protein levels is associated with obesity in boys,” Hypertension Research, vol. 26, no. 7, pp. 541–546, 2003.

[26] M. Ebrahimi, A. R. Heidari-Bakavoli, S. Shoebili et al., “Association of serum hs-CRP levels with the presence of obesity, diabetes mellitus, and other cardiovascular risk factors,” Journal of Clinical Laboratory Analysis, vol. 30, no. 5, pp. 672–676, 2016.

[27] M. Visser, L. M. Bouter, G. M. McQuillan, M. H. Wener, and T. B. Harris, “Elevated C-reactive protein levels in overweight and obese adults,” Journal of the American Medical Association, vol. 282, no. 22, pp. 2313–2315, 1999.

[28] P. W. Holland, “Statistics and causal inference,” Journal of the American Statistical Association, vol. 81, no. 396, pp. 945–960, 1986.

[29] I. Postmus, J. Deelen, S. Sedaghat et al., “LDL cholesterol still a problem in old age? A Mendelian randomization study,” International Journal of Epidemiology, vol. 44, no. 2, pp. 604–612, 2015.

[30] J. R. Thompson, C. Minelli, K. R. Abrams, M. D. Tobin, and R. D. Riley, “Meta-analysis of genetic studies using Mendelian randomization—a multivariate approach,” Statistics in Medicine, vol. 34, no. 14, pp. 2241–2254, 2015.

[31] W. Zhou, B. Brumpton, O. Kabil et al., “GWAS of thyroid stimulating hormone highlights pleiotropic effects and inverse association with thyroid cancer,” Nature Communications, vol. 11, no. 1, pp. 1–13, 2020.

[32] A. Teumer, L. Chaker, S. Groenevweg et al., “Genome-wide analyses identify a role for SLC17A4 and AADAT in thyroid hormone regulation,” Nature Communications, vol. 9, no. 1, pp. 1–14, 2018.

[33] G. L. Wojcick, M. Graff, K. K. Nishimura et al., “Genetic analyses of diverse populations improves discovery for complex traits,” Nature Communications, vol. 570, no. 7762, pp. 514–518, 2019.
[34] S. L. Pulit, C. Stoneman, A. P. Morris et al., “Meta-analysis of genome-wide association studies for body fat distribution in 694,649 individuals of European ancestry,” *Human Molecular Genetics*, vol. 28, no. 1, pp. 166–174, 2019.

[35] L. Yengo, J. Sidorenko, K. E. Kemper et al., “Meta-analysis of genome-wide association studies for height and body mass index in ~700,000 individuals of European ancestry,” *Human Molecular Genetics*, vol. 27, no. 20, pp. 3641–3649, 2018.

[36] B. L. Pierce, H. Ahsan, and T. J. Vander Weele, “Power and instrument strength requirements for Mendelian randomization studies using multiple genetic variants,” *International Journal of Epidemiology*, vol. 40, no. 3, pp. 740–752, 2011.

[37] S. Burgess, A. Butterworth, and S. G. Thompson, “Mendelian randomization analysis with multiple genetic variants using summarized data,” *Genetic Epidemiology*, vol. 37, no. 7, pp. 658–665, 2013.

[38] J. Bowden, G. Davey Smith, P. C. Haycock, and S. Burgess, “Consistent estimation in Mendelian randomization with some invalid instruments using a weighted median estimator,” *Genetic Epidemiology*, vol. 40, no. 4, pp. 304–314, 2016.

[39] J. Bowden, G. Davey Smith, and S. Burgess, “Mendelian randomization with invalid instruments: effect estimation and bias detection through Egger regression,” *International Journal of Epidemiology*, vol. 44, no. 2, pp. 512–525, 2015.

[40] M. F. D. Greco, C. Minelli, N. A. Sheehan, and J. R. Thompson, “Detecting pleiotropy in Mendelian randomisation studies with summary data and a continuous outcome,” *Statistics in Medicine*, vol. 34, no. 21, pp. 2926–2940, 2015.

[41] S. Burgess, J. Bowden, T. Fall, E. Ingelsson, and S. G. Thompson, “Sensitivity analyses for robust causal inference from Mendelian randomization analyses with multiple genetic variants,” *Epidemiology*, vol. 28, no. 1, pp. 30–42, 2017.

[42] W. G. Cochran, “The comparison of percentages in matched samples,” *Biometrika*, vol. 37, no. 3–4, pp. 256–266, 1950.

[43] B. Baujat, C. Mahé, J. P. Pignon, and C. Hill, “A graphical method for exploring heterogeneity in meta-analyses: application to a meta-analysis of 65 trials,” *Statistics in Medicine*, vol. 21, no. 18, pp. 2641–2652, 2002.

[44] G. Gupta, P. Sharma, P. Kumar, and M. Itagappa, “Study on subclinical hypothyroidism and its association with various inflammatory markers,” *Journal of Clinical and Diagnostic Research: JCDR*, vol. 9, no. 11, pp. BC04–BC06, 2015.

[45] J. Wu, Y. Tao, H. Gu, and J. Sui, “Association between cardiovascular risk factors and serum thyrotropin concentration among healthy Chinese subjects and subjects with unsuspected subclinical hypothyroidism,” *Clinical Laboratory*, vol. 62, no. 5, pp. 807–814, 2016.

[46] J. Kvetny, P. Heldgaard, E. M. Bladbjerg, and J. Gram, “Subclinical hypothyroidism is associated with a low-grade inflammation, increased triglyceride levels and predicts cardiovascular disease in males below 50 years,” *Clinical Endocrinology*, vol. 61, no. 2, pp. 232–238, 2004.

[47] W.-Y. Lee, J.-Y. Suh, E.-J. Rhee, J.-S. Park, K.-C. Sung, and S.-W. Kim, “Plasma CRP, apolipoprotein A-1, apolipoprotein B and Lp(a) levels according to thyroid function status,” *Archives of Medical Research*, vol. 35, no. 6, pp. 540–545, 2004.

[48] V. Lubrano, A. Pingitore, A. Carpi, and G. Iervasi, “Relationship between triiodothyronine and proinflammatory cytokines in chronic heart failure,” *Biomedicine & Pharmacotherapy*, vol. 64, no. 3, pp. 165–169, 2010.