Genetic polymorphisms at SIRT1 and FOXO1 are associated with carotid atherosclerosis in the SAPHIR cohort

Lyudmyla Kedenko†, Claudia Lamina†, Igor Kedenko1, Barbara Kollerits2, Tobias Kiesslich1,3, Bernhard IglSeder4, Florian Kronenberg2* and Bernhard Paulweber1

Abstract

Background: SIRT1 and FOXO1 interact with each other in multiple pathways regulating aging, metabolism and resistance to oxidative stress and control different pathways involved in atherosclerotic process. It is not known, if genetic polymorphisms (SNPs) at the SIRT1 and FOXO1 have an influence on carotid atherosclerosis.

Methods: Intima-media thickness (IMT) was measured on the common and internal carotid arteries. Morphological alterations of the carotid arteries and size of these alterations were included in the B-score grading on a five point scale. Eleven SNPs at SIRT1 and FOXO1 gene loci were genotyped in the SAPHIR cohort (n = 1742). The association of each SNP with common carotid IMT, internal carotid IMT and B-score was analyzed using linear regression models.

Results: A significant association was found between common carotid IMT and two SNPs at FOXO1 - rs10507486, rs2297627 (beta = -0.00168, p = 0.0007 and beta = -0.00144, p = 0.0008 respectively) and at least a trend for rs12413112 at SIRT1 (beta = 0.00177, p = 0.0157) using an additive model adjusting for age and sex. Additional adjustment for traditional cardiovascular risk factors and markers (BMI, smoking status, hypertension, total cholesterol, HDL-cholesterol, hsCRP) even improved the strength of this association (p = 0.0037 for SIRT1 and p = 0.0002 for both SNPs at FOXO1). Analysis for internal carotis IMT and B-score did not reveal any significant association. One haplotype in FOXO1 showed a moderate effect on common carotid IMT and B-score in comparison to the reference haplotype of this gene. Several SNPs within SIRT1 showed differential effects for men and women with higher effect sizes for women: rs3740051 on all three investigated phenotypes (interaction p-value < 0.0069); rs2236319 on common and internal carotid IMT (interaction p-value < 0.0083), rs10823108, rs2273773 on common carotid IMT and rs1467568 on B-score (interaction p-value = 0.0007). The latter was significant in women only ($\beta_{women} = 0.111$, $p_{women} = 0.00008$; $\beta_{men} = -0.009$, $p_{men} = 0.6464$).

Conclusions: This study demonstrated associations of genetic variations at the SIRT1 and FOXO1 loci with carotid atherosclerosis and highlighted the need for further investigation by functional studies.

Background

SIRT1 and FOXO1 are evolutionary conserved regulators of aging, metabolic processes and resistance to oxidative stress. Recently, their role in vascular homeostasis has been suggested. SIRT1 seems to have protective properties against atherosclerosis [1]. It controls vascular tone and endothelial function by deacetylation of endothelial nitric oxide synthase and by stimulating its activity [2,3]. In endothelial cells and macrophages SIRT1 has anti-inflammatory functions by down-regulating the expression of various pro-inflammatory cytokines, interfering with the NFκB signaling pathway, preventing macrophage foam cell formation and suppressing of endothelial tissue factor (coagulation factor III) expression [1]. In vascular smooth muscle cells SIRT1 enhances the activity of tissue inhibitor of metalloproteinase 3 that might stabilize atherosclerotic plaques [4]. Moreover, SIRT1 is a key component in
several stress-responsive pathways involved in apoptotic cell death and cellular senescence [3].

FOXO1 is strongly expressed in atherosclerotic plaques, regulates expression of different cell cycle regulators and is involved in multiple atherogenic pathways in endothelial cells. FOXO1 activity is regulated through processes of phosphorylation by Akt, acetylation at multiple lysine residues and deacetylation by several deacetylases, including the NAD+-dependent deacetylase SIRT1 [5-9].

FOXO1 is deacetylated in response to oxidative stress and hyperglycemia. Deacetylation of FOXO1 leads to its activation and enhanced expression of FOXO1 target genes [10]. Mice bearing constitutively deacetylated alleles of Foxo1 develop larger atherosclerotic lesions despite improved plasma lipid levels [11]. Ablation of the three genes encoding isoforms of FOXO in endothelial cells prevents atherosclerosis in LDL receptor knockout mice [12]. The atheroprotective effect of FOXO1 deletion was associated with a decrease of insulin-dependent Akt phosphorylation in endothelial cells. Growth factors induce phosphorylation and nuclear exclusion of forkhead proteins in vascular smooth muscle cells that inhibits proliferation of these cells and neo-intimal hyperplasia [13].

Findings from previous studies showed that SIRT1 and FOXO1 interact with each other in regulating the response to oxidative stress, ageing and longevity. SIRT1 has multiple effects on FOXO-induced gene expression that depresses processes of apoptosis and induces cell-cycle arrest and survival [7,14]. Until now, associations of genetic polymorphisms at SIRT1 and FOXO1 with body mass index (BMI), visceral obesity, insulin sensitivity, type 2 diabetes mellitus (T2DM) and all-cause mortality risk have been described [15-19]. Taking into consideration that atherosclerosis is an aspect of the aging process we hypothesized that genetic polymorphisms at SIRT1 and FOXO1 may also play a role in carotid atherosclerosis.

Methods
Study population
The Salzburg Atherosclerosis Prevention Program in subjects at High Individual Risk (the SAPHIR study) has been initiated in 1999 as a prospective study that investigates genetic and environmental factors contributing to atherosclerotic vascular diseases [20]. Study participants were recruited by health screening programs in large companies in and around the city of Salzburg, Austria. All study participants provided a signed informed consent and the study was approved by the Ethical Committee of Salzburg. All clinical investigations were conducted according to the principles expressed in the Declaration of Helsinki. The study comprises 1770 healthy unrelated Caucasian subjects (663 females and 1107 males aged 39-67 years). At baseline, all participants were subjected to a comprehensive examination — detailed personal and family history, physical, instrumental and laboratory investigations. All laboratory parameters were determined in the fasting state. Participants were diagnosed as having T2DM if their fasting plasma glucose concentration was ≥7 mmol/L and/or they were being treated with anti-diabetic therapy. The participants were placed into hypertension group if the systolic blood pressure was ≥140 mm Hg and/or diastolic blood pressure ≥90 mm Hg or they received antihypertensive medication.

Ultrasongraphy
Intima-media thickness (IMT) was measured end-diastolic according to the leading edge method for both the near and far walls on the common and internal carotid arteries by high resolution B-mode ultrasound using the ATL HDI 3000 CV (Philips Medical Systems, Bothell, WA, USA) according to the Asymptomatic Carotid Artery Plaque Study (ACAPS) protocol [21]. The protocol included multiple longitudinal and transverse imaging planes of the common carotid artery (CCA) on the 8 mm distance proximal to the bifurcation at height of the flow divider and within the proximal 8 mm of the internal carotid artery (ICA). The mean values of left and right arteries for CCA and ICA were used in the statistical analyses. Because of the technical difficulties in some cases of ultrasonography common carotid IMT and internal carotid IMT were available in 1689 and 1687 participants, respectively. All measurements were conducted and analyzed by a single experienced ultrasound operator who was blinded to all clinical and laboratory data.

The protocol for B-score included multiple longitudinal and transverse imaging planes of the common and internal carotid arteries. Morphological alterations of the carotid arteries and size of these alterations were included in the B-score grading on a 5-point scale: 0 - no alteration, 1 - wall thickness >1 mm, 2 - plaque < 2 mm, 3 - plaque 2 to 3 mm, 4 - plaque >3 mm, and 5 - total obstruction of the lumen. Adding the B-score of all segments divided by the number of segments resulted in a mean B-score. The mean values of the mean B score were used in the statistical analyses. B-score data were available in 1759 participants.

Selection of single nucleotide polymorphisms and genotyping
Genotype data of the SIRT1 and FOXO1 genes were uploaded from HapMap Data (Phase III/ Rel.#2, Feb 09) (http://hapmap.ncbi.nlm.nih.gov/cgi-perl/gbrowse/hapmap3r2_B36/) and were transferred to SNP tagger (http://www.broad.mit.edu/mpg/tagger/server.html) to identify haplotype-tagging single nucleotide polymorphisms (SNPs). Using the haplotype block structure, we selected a maximally informative subset of validated SNPs with minor allele frequency of >5% for SIRT1 and >10% for FOXO1, which...
pairwise tag other SNPs within that gene regions with \( r^2 > 0.8 \). The difference in selected minor allele frequencies between the SIRT1 and FOXO1 genes was based on less polymorphic genetic structure of the SIRT1 gene. Two SNPs at the SIRT1 (rs2236319 and rs3740051) were additionally included due to their association with the aging diseases – Alzheimer [22].

Genomic DNA was isolated from whole blood by QIAamp DNA Blood Mini Kit according to manufacture protocol (Qiagen, Germany). DNA samples from whole blood were available for 1742 participants. Genotyping was performed using 5` nuclease allelic discrimination TaqMan genotyping method and pre-designed assays from Applied Biosystems (Foster City, CA, USA) according to the manufacturer’s instructions.

**Statistical analysis**

The association of each SNP was analyzed with log-transformed mean common carotid IMT, mean internal carotid IMT and mean B-score using linear regression models adjusted for a) age and sex and b) age, sex and traditional cardiovascular risk factors and markers (BMI, total cholesterol, HDL-cholesterol, current smoking status, hypertension, hsCRP). For better interpretability of effects, the beta estimates that are reported are based on the outcome-variables on the original scale, whereas \( \beta \)-values are derived from the respective log-transformed models to ensure linear model assumptions. An additive genetic effect was assumed. The analyses were also conducted stratified for sex. In this case analyses were adjusted for age, BMI, total cholesterol, HDL-cholesterol, current smoking status, hypertension, and hsCRP. The occurrence of differential effects between men and women has been evaluated by including SNP*sex interaction terms in the fully adjusted regression model. Additional analyses were performed including pairwise SNP-SNP interaction terms between SNPs of both genes, using the same adjustment models and inheritance assumptions as above.

The Bonferroni correction for multiple testing was applied on an independent number of tests for the main analyses, which are the association analyses of the single SNPs with the three phenotypes. This number was calculated using the effective number of loci [23], which accounts for the correlation structure between the SNPs. Since all three phenotypes are highly correlated, no additional adjustment was made with regard to the number of tested phenotypes or secondary analyses as the interaction analyses. The effective number of loci was estimated to be 6 altogether, three SNPs at both FOXO1 and SIRT1. Therefore, the significance threshold was set to 0.0083 (0.05 divided by 6) for all SNP-based analyses.

Also the haplotypes for SIRT1 and FOXO1 genes were estimated by the expectation maximization algorithm using the haplo.stats package (http://CRAN.R-project.org/package=haplo.stats) in the R software environment [24]. Subsequent association analysis of the number of haplotype copies on log-transformed IMT and B-score was adjusted for age and sex. Linkage disequilibrium (LD)-plots were performed using the program Haploview [25]. All other analyses were performed using R software version 2.14.2.

**Results**

**Description of the SAPHIR cohort**

Clinical and main laboratory characteristics of the SAPHIR cohort are presented in Table 1. Females were older than males and also had higher levels of total cholesterol and HDL-cholesterol in comparison to males.

**Genotype characteristics, linkage disequilibrium and association analysis**

We genotyped 6 SNPs at SIRT1 and 5 SNPs at FOXO1.

Table 2 shows the corresponding descriptive statistics of the genotypes. Two SNPs at FOXO1 (rs10507486 and rs17446614) showed a deviation from the Hardy-Weinberg equilibrium which was not as extreme to justify exclusion of these SNPs. The pairwise LD in terms of \( r^2 \)-values is given in Figure 1. \( r^2 \) is rather high between four SNPs in SIRT1 (>0.87).

Association analysis results for IMT and B-score are presented in Table 3. Significant associations were found between log-transformed mean common carotid IMT and two variants at FOXO1 (rs10507486, rs2297627) based on an additive model and adjustment for age and sex. These effects are even enhanced after adjustment for traditional cardiovascular risk factors and markers (\( p = 0.0002 \)). In this multivariate model, a significant effect was also observed for one variant at SIRT1 (rs12413112, \( p = 0.0037 \)). The variant allele of rs12413112 at SIRT1 was associated with an increase of about 0.002 \( \mu m \) in mean common carotid IMT (fully-adjusted model). At the same time variant alleles at the FOXO1 had an opposite effect and were associated with decreasing IMT (decrease of about 0.0018 \( \mu m \) for rs10507486 or 0.0016 \( \mu m \) for rs2297627).

**Haplotype analysis**

Only haplotypes with a frequency > 1% were included in the haplotype analysis. Genomic structure of the SIRT1 and FOXO1 and their haplotypes are presented in Figure 2.

There was no difference in effects between any haplotypes in SIRT1 (Table 4). The haplotype ATGTC in FOXO1 (frequency 5.1%) was associated with CCA IMT and mean B-score in comparison to the reference haplotype (ATGCT) for this gene (\( \beta = -0.00326, p = 0.0006 \) for common carotid IMT and \( \beta = -0.09123, p = 0.0077 \) for B-score). For mean CCA IMT, this haplotype effect primarily reflects the single SNP effects of rs10507486.
and rs2297627, since this haplotype differs by alleles at these SNPs from the reference haplotype. For B-score, however, this haplotype analysis detected an association that was not identified on the SNP level.

**Sex-stratified analyses**

SNP*sex interaction terms and sex-stratified analyses revealed additional information on the association of the investigated SNPs with atherosclerosis in the carotid arteries. Differential effects for men and women were observed for several SNPs within *SIRT1* (Table 5): for rs3740051 on all three investigated phenotypes (p interaction <0.0069); for rs2236319 on CCA IMT and ICA IMT (p interaction <0.0083), for rs10823108 and rs2273773 on common carotid IMT only and for rs1467568 on B-score only (p interaction = 0.0007). For all of these SNPs, effect directions differed between men and women with decreasing IMT and/or B-score for each minor allele for men, and increasing IMT and/or B-score for the minor allele for women. In men, these decreasing effects were not significant. In women, all of these differentially effective SNPs were at least nominally significantly associated with their respective phenotypes (all women-specific p-values < 0.013) with about 3-4 times higher effect sizes for women than for men. The highest difference was observed for the rs1467568 at *SIRT1* on the B-score with a significant effect in women only ($\beta_{\text{women}} = 0.111, p_{\text{women}} = 0.00008; \beta_{\text{men}} = -0.009, p_{\text{men}} = 0.6464$).

There were no differential effects between men and women in any of the *FOXO1* SNPs.

### Table 1 Clinical and laboratory characteristics of the SAPHIR cohort (mean ± SD or n, %)

| Variables          | All (n = 1770) | Males (n = 1107) | Females (n = 663) |
|--------------------|---------------|-----------------|------------------|
| Age, years         | 51.39 ± 6.02  | 48.84 ± 5.42    | 55.66 ± 4.32     |
| BMI, kg/m²         | 26.80 ± 4.12  | 26.92 ± 3.72    | 26.58 ± 4.70     |
| Hypertension, n (%)| 986 (55.7)    | 990 (53.3)      | 964 (59.7)       |
| Smoking, n (%)     | 340 (19.2)    | 254 (22.9)      | 86 (13)          |
| T2DM, n (%)        | 60 (3.4)      | 34 (3.1)        | 26 (3.9)         |
| Total cholesterol, mg/dl | 228.77 ± 40.13 | 225.98 ± 39.79 | 233.46 ± 40.29  |
| HDL-cholesterol, mg/dl | 59.62 ± 15.72 | 55.04 ± 13.39  | 67.30 ± 16.33   |
| hs-CRP, mg/dl      | 0.289 ± 0.625 | 0.255 ± 0.696   | 0.347 ± 0.480    |
| Mean B score       | 0.447 ± 0.504 | 0.421 ± 0.495   | 0.490 ± 0.518    |
| CCA IMT, μm        | 0.0763 ± 0.0128 | 0.0758 ± 0.0125 | 0.0769 ± 0.0131 |
| ICA IMT, μm        | 0.0827 ± 0.0133 | 0.0827 ± 0.0131 | 0.0826 ± 0.0135 |

### Table 2 Characteristics of the 11 single nucleotide polymorphisms and their genotype quality in the SAPHIR cohort

| SNP        | Alleles Minor/Major* | SAPHIR (n = 1742) | Genotype counts** | HWE p*** | Call rate |
|------------|----------------------|-------------------|-------------------|----------|-----------|
| **FOXO1**  |                      |                   |                   |          |           |
| rs17446593 | G/A                  | 0.178             | 1186/513/55       | 1.000    | 0.991     |
| rs2721068  | C/T                  | 0.260             | 958/666/121       | 0.755    | 0.986     |
| rs17446614 | A/G                  | 0.155             | 1259/430/55       | 0.017    | 0.985     |
| rs10507486 | A/G                  | 0.203             | 1135/529/91       | 0.006    | 0.992     |
| rs2297627  | G/A                  | 0.304             | 867/710/180       | 0.055    | 0.993     |
| **SIRT1**  |                      |                   |                   |          |           |
| rs3740051  | G/A                  | 0.063             | 1546/204/8        | 0.682    | 0.993     |
| rs2236319  | G/A                  | 0.063             | 1543/207/7        | 1.000    | 0.993     |
| rs12413112 | A/G                  | 0.120             | 1365/363/29       | 0.367    | 0.993     |
| rs10823108 | A/G                  | 0.063             | 1546/203/9        | 0.414    | 0.993     |
| rs2273773  | C/T                  | 0.063             | 1544/205/8        | 0.683    | 0.993     |
| rs1467568  | A/G                  | 0.340             | 769/768/299       | 0.424    | 0.986     |

*Minor and Major alleles based on the plus-strand.
**Number of homozygotes for the major allele/heterozygotes/homozygotes for the rare allele.
***Based on exact test of HWE.
Discussion

Our study is the first to report a significant association of genetic polymorphisms at SIRT1 and FOXO1 with carotid atherosclerosis. More pronounced effects were present for two SNPs (rs10507486 and rs2297627) at FOXO1 and common carotid IMT. Also we found one haplotype in FOXO1 with a moderate effect on common carotid IMT and B-score in comparison to the reference haplotype. Finally, we detected a sex-specific effect for the SNPs at SIRT1 with a protective role of the minor alleles in males and an opposite effect in females.

The pathophysiological functions of SIRT1 and FOXO1 in vascular homeostasis and carotid atherosclerosis have been described within the last years and exert protective roles in case of SIRT1 and controversial roles for FOXO1. Also, the interaction between SIRT1 and FOXO1 in different organisms has been demonstrated. SIRT1 deacetylates FOXO1 and regulates its transcriptional activity, in turn, FOXO1 exerts a positive feedback mechanism regulating SIRT1 expression. FOXO1 can directly bind to SIRT1 promoter region and increase SIRT1 transcription and expression [26]. Phosphorylation of FOXO1 makes it retention in the cytoplasm and inhibition of its transcription activity [27,28]. In response to oxidative stress, FOXO1 translocates to the nucleus, interacts with SIRT1 resulting in the deacetylation of FOXO1 [10]. This depresses the sensitivity of FOXO1 to insulin-induced phosphorylation, resulting in gain function [29]. At the same time, acetylation of FOXO1 facilitates their phosphorylation and nuclear translocation.

Oxidative stress or hyperglycemia induce FOXO1 deacetylation in vascular endothelial cells, that contributes to the increased risk of atherosclerosis in diabetic patients. Experiments with isolated vascular endothelial cells from mice indicated that deacetylated FOXO1 affects its gene expression response to a variety of pathogenic stimuli and increases interaction of monocytes with endothelium [11].

Previous publications have shown an association of genetic variations at SIRT1 with BMI and visceral obesity [16], acute insulin secretion [30], basal energy expenditure and the respiratory quotient [31]. Carriers of the mutant allele were resistant against lifestyle-induced improvement of fasting plasma glucose, had less increase in insulin sensitivity and an attenuated decline in liver fat [31]. Also it was shown that expression of the SIRT1 gene was negatively correlated with carotid IMT [32]. However, no genetic polymorphisms in the SIRT1 gene were associated with expression level of the SIRT1 transcript [16].

There are also findings about the role of genetic variations in the FOXO1 and the susceptibility for T2DM. Müssig et al. showed the nominally significant association of rs2297627 with β-cell dysfunction (p = 0.0387) and impaired glucose tolerance (p = 0.0221). Carries of the minor allele had reduced insulin secretion and elevated glucose levels during an oral glucose tolerance test [17]. Kuningas et al. revealed the role of different haplotypes in the human FOXO1a gene in glucose metabolism, all-cause and T2DM mortality risk [33].

Association of genetic polymorphisms at the SIRT1 and FOXO1 loci with carotid atherosclerosis have not been reported before. However, significant associations between genetic variants in other SIRT gene - the SIRT6 and atherosclerotic plaque have been observed [34]. The last meta-analysis of genome-wide association studies by the CHARGE consortium identified some variants associated
| SNP    | Mean common carotid IMT (n = 1689) | Mean internal carotid IMT (n = 1687) | Mean B-score (n = 1759) |
|--------|-----------------------------------|--------------------------------------|-------------------------|
|        | Age-and sex-adjusted | Fully adjusted | Age-and sex-adjusted | Fully adjusted | Age-and sex-adjusted | Fully adjusted |
|        | β (se) | p-value | β (se) | p-value | β (se) | p-value | β (se) | p-value | β (se) | p-value |
| **FOXO1** | | | | | | | | | | | |
| rs17446593 | -0.00046 (0.00052) | 0.3934 | -0.00047 (0.00052) | 0.3821 | 0.00041 (0.00057) | 0.5654 | 0.00053 (0.00056) | 0.4289 | 0.00391 (0.02081) | 0.7763 | 0.00648 (0.02066) | 0.6889 |
| rs2721068 | -0.00089 (0.00045) | 0.0412 | -0.00106 (0.00045) | 0.0145 | -0.00049 (0.00049) | 0.2739 | -0.00055 (0.00048) | 0.2224 | -0.01029 (0.01803) | 0.6473 | -0.01329 (0.01786) | 0.5229 |
| rs17446614 | -0.00118 (0.00054) | 0.0272 | -0.00125 (0.00053) | 0.0180 | -0.00032 (0.00058) | 0.4625 | -0.00027 (0.00058) | 0.5278 | -0.00860 (0.02131) | 0.7509 | -0.00725 (0.02118) | 0.7789 |
| rs10507486 | -0.00168 (0.00042) | 0.0007 | -0.000184 (0.00047) | 0.0002 | -0.00083 (0.00052) | 0.1080 | -0.00079 (0.00052) | 0.1278 | -0.02940 (0.01907) | 0.1178 | -0.03007 (0.01898) | 0.1007 |
| rs2297627 | -0.00144 (0.00042) | 0.0008 | -0.000158 (0.00042) | 0.0002 | -0.00100 (0.00046) | 0.0333 | -0.00104 (0.00045) | 0.0253 | -0.03310 (0.01681) | 0.0516 | -0.03620 (0.01664) | 0.0301 |
| **SIRT1** | | | | | | | | | | | |
| rs3740051 | 0.00048 (0.00082) | 0.5530 | 0.000053 (0.00081) | 0.5012 | 0.00044 (0.00088) | 0.7644 | 0.000068 (0.00088) | 0.5590 | 0.00096 (0.03260) | 0.9972 | 0.00816 (0.03250) | 0.8322 |
| rs2236319 | 0.00033 (0.00082) | 0.7001 | 0.000039 (0.00081) | 0.6382 | 0.00016 (0.00088) | 0.9579 | 0.000039 (0.00088) | 0.7410 | -0.00578 (0.03272) | 0.8706 | 0.00484 (0.03254) | 0.8668 |
| rs12413112 | 0.00177 (0.00061) | 0.0157 | 0.000206 (0.00061) | 0.0037 | 0.00100 (0.00066) | 0.1375 | 0.000108 (0.00066) | 0.1064 | 0.05019 (0.02423) | 0.0847 | 0.04818 (0.02411) | 0.0989 |
| rs10823108 | 0.00031 (0.00081) | 0.6926 | 0.000038 (0.00080) | 0.6102 | -0.00002 (0.00087) | 0.9096 | 0.000024 (0.00087) | 0.8477 | -0.01326 (0.03240) | 0.6779 | -0.00217 (0.03222) | 0.9476 |
| rs2273773 | 0.00052 (0.00081) | 0.5360 | 0.000059 (0.00081) | 0.4716 | 0.00023 (0.00088) | 0.9143 | 0.000045 (0.00088) | 0.7025 | -0.00549 (0.03257) | 0.8698 | 0.00502 (0.03239) | 0.8692 |
| rs1467568 | 0.00034 (0.00042) | 0.5594 | 0.000045 (0.00041) | 0.3747 | 0.00063 (0.00045) | 0.2329 | 0.00072 (0.00045) | 0.1563 | 0.03433 (0.01659) | 0.0775 | 0.03577 (0.01645) | 0.0643 |

β-estimate and se: based on original scale of phenotype variables; p-value: based on log-transformed phenotype variables.
Full-adjusted model: age, sex, BMI, total cholesterol, HDL-cholesterol, current smoking status, hypertension, hsCRP.
In bold: Significant p-values after correction for multiple testing.
with common carotid IMT and plaques. However, no SNP achieved significant thresholds on a genome-wide scale for internal carotid IMT [35].

The genetic effect of SIRT1 and FOXO1 on common carotid IMT shown in the current study might be explained by their direct involvement in the pathophysiological mechanisms of atherosclerosis. FOXO1 integrates diverse cellular signals through its post-translational modifications, such as phosphorylation, acetylation, deacetylation resulting in changes in DNA binding properties, transcriptional activity, protein stability and subcellular localization. At the same time, SIRT1 is one of multiple regulators of FOXO1 activity in endothelial and smooth muscle cells.

It has to be noted that the definition of the carotid atherosclerotic phenotype consists of site-dependent IMT measurement and plaque-score. Although IMT and plaques are highly correlated with each other [36,37], we found in our study an association of genetic polymorphisms at the SIRT1 and FOXO1 only with common carotid IMT. A haplotype specific effect was also found for mean B-score, though, which could not be detected at the single SNP level. The site-specific differences in atherosclerosis phenotype might be based on the important role of blood flow and genetics in manifestation and progression of carotid atherosclerosis [38-40]. Elevated carotid IMT may be the result of an increased intima layer as a consequence of atherosclerotic process, but also as an

Table 4 Results of linear regression model for additive haplotype effects (the most common haplotype has been defined as the reference haplotype)

| Haplotype | Mean common carotid IMT | Mean internal carotid IMT | Mean B-score |
|-----------|-------------------------|---------------------------|--------------|
|           | Age-and sex-adjusted    | Age-and sex-adjusted      | Age-and sex-adjusted |
|           | β (se)  | p-value   | β (se)  | p-value   | β (se)  | p-value   |
| FOXO1     |         |           |         |           |         |           |
| 2         | -0.00011 (0.00061) | 0.0777 | -0.00003 (0.00066) | 0.8915 | -0.00724 (0.02420) | 0.8421 |
| 3         | -0.00040 (0.00068) | 0.4764 | -0.00066 (0.00074) | 0.3924 | -0.00199 (0.02748) | 0.5289 |
| 4         | -0.00326 (0.00089) | **0.0006** | -0.00250 (0.00096) | 0.0185 | -0.00913 (0.03574) | **0.0077** |
| 5         | 0.00119 (0.00106) | 0.2412 | 0.00164 (0.00114) | 0.1446 | 0.00082 (0.06020) | 0.7220 |
| 6         | -0.00231 (0.00147) | 0.0981 | -0.00269 (0.00159) | 0.0816 | -0.00619 (0.05776) | 0.2777 |
| SIRT1     |         |           |         |           |         |           |
| 2         | -0.00067 (0.00056) | 0.2232 | -0.00040 (0.00060) | 0.6033 | 0.00765 (0.02199) | 0.2618 |
| 3         | 0.00163 (0.00063) | 0.0363 | 0.00102 (0.00068) | 0.1593 | 0.05442 (0.02509) | 0.0765 |
| 4         | 0.00022 (0.00087) | 0.8573 | 0.00026 (0.00094) | 0.9468 | -0.00035 (0.03497) | 0.8911 |

β-estimate and se: based on original scale of phenotype variables; p-value: based on log-transformed phenotype variables.
All models are age-and sex-adjusted.
In bold: Significant p-values after correction for multiple testing.
Table 5 Sex-specific effects for the genetic polymorphisms in *FOXO1* and *SIRT1* genes, fully adjusted model

| SNP          | CCA IMT (n = 1689) | ICA IMT (n = 1687) | B score carotis (n = 1759) |
|--------------|-------------------|--------------------|---------------------------|
|              | β (se) men        | β (se) women       | p-value interaction       | β (se) men        | β (se) women       | p-value interaction |
| *FOXO1*      |                   |                    |                           |                   |                    |                           |
| rs17446593   | -0.00139 (0.00062)| 0.00114 (0.00091) | 0.0147                    | -0.00005 (0.00068)| 0.00149 (0.00097) | 0.2658                   |
| rs2721068    | -0.00125 (0.00055)| -0.00093 (0.00076) | 0.4421                    | -0.00077 (0.00060)| -0.00035 (0.00081) | 0.5616                   |
| rs17446614   | -0.00164 (0.00063)| -0.00057 (0.00094) | 0.2277                    | -0.00052 (0.00070)| 0.00017 (0.00100) | 0.5361                   |
| rs10507486   | -0.00218 (0.00057)| -0.00138 (0.00082) | 0.3032                    | -0.00098 (0.00063)| -0.00058 (0.00088) | 0.7306                   |
| rs2297627    | -0.00151 (0.00051)| -0.00187 (0.00071) | 0.9872                    | -0.00105 (0.00056)| -0.00120 (0.00076) | 0.9945                   |
| *SIRT1*      |                   |                    |                           |                   |                    |                           |
| rs3740051    | -0.00122 (0.00094)| 0.00433 (0.00152) | 0.0019                    | -0.00104 (0.00104)| 0.00458 (0.00162) | 0.0024                   |
| rs2236319    | -0.00133 (0.00094)| 0.00408 (0.00152) | 0.0025                    | -0.00108 (0.00103)| 0.00372 (0.00162) | 0.0083                   |
| rs12413112   | 0.00140 (0.00073) | 0.00309 (0.00106) | 0.2166                    | 0.00129 (0.00080) | 0.00066 (0.00113) | 0.7856                   |
| rs10823108   | -0.00113 (0.00092)| 0.00383 (0.00153) | 0.0063                    | -0.00118 (0.00101)| 0.00359 (0.00164) | 0.0101                   |
| rs273773     | -0.00107 (0.00093)| 0.00426 (0.00153) | 0.0030                    | -0.00097 (0.00102)| 0.00378 (0.00163) | 0.0091                   |
| rs1467558    | -0.00027 (0.00050)| 0.00160 (0.00070) | 0.0329                    | -0.00021 (0.00056)| 0.00215 (0.00075) | 0.0124                   |

β-estimate and se: based on original scale of phenotype variables; p-value: p-value from the SNP*genender interaction term of the full-adjusted model, based on log-transformed phenotype variables.

Full-adjusted model: age, sex, BMI, total cholesterol, HDL-cholesterol, current smoking status, hypertension, hsCRP.

In bold: Significant p-values after correction for multiple testing.
increased media layer due to vascular hypertrophy in hypertension or increasing of both layers. Nevertheless, IMT in carotid arteries is an established marker of the early onset of atherosclerosis and has been included in recent guidelines as a potentially useful marker for cardiovascular disease prediction in clinical and epidemiological studies [41,42].

It is evident, that there is a sex-specific aspect of cardiovascular disease. In the Tromso Study, carotid atherosclerosis was a stronger predictor of myocardial infarction in women than in men [43]. Also this study showed that only in women triglyceride levels were associated with an increase in IMT, while physical activity and smoking were predictors of IMT in men [44]. Previously we published that the metabolic syndrome is a stronger risk factor for early carotid atherosclerosis in women than in men. In females, blood glucose and triglyceride levels showed the strongest association with IMT, whereas HDL-cholesterol had a stronger effect in men [45]. The available literature describes sex-specific effects only for SIRT1 in relation to increased risk for T2DM in Pima Indians [30] and SIRT6 for atherosclerotic plaque [34]. We cannot explain the mechanism underlying the sex-related association shown in our study. The SNPs that showed sex-specific effects were not significant in the overall analysis. Different effect of genetic polymorphisms in the SIRT1 on carotid atherosclerosis in female and male populations might be partial explained by the different role of sex hormones in regulation of gene expression or important role of sex-specific risk factors for atherosclerosis.

The strength of this investigation is that all ultrasound investigations were done by a single experienced ultrasound operator who was blinded to all clinical and laboratory measurements. It allowed us to reduce inter-observer variability and obtain data independent from the clinical phenotype. Moreover, the gene targeting approach used in this study gave not only the possibility to find association of genetic variations with carotid IMT and B-score, but also brought a new information about role of genetics of SIRT1 and FOXO1 in the atherosclerotic process. There are also some limitations of this investigation. First, replication of the findings in an independent cohort is missing. Due to power issues it was also not possible to perform an internal validation approach. Secondly, some of the included SNPs were in linkage disequilibrium although SNPs have been selected using SNPtagger to exclude such a situation. In the analysis, the LD structure was accounted for by calculating the number of independent SNPs. Therefore, further replication studies and also functional studies are warranted to elucidate the molecular mechanism of this association and the role of these genetic variations at SIRT1 and FOXO1 in carotid atherosclerosis.

Conclusions
Atherosclerosis is a clinically latent process for many years before its manifestation as coronary, cerebrovascular and/or peripheral arteries diseases. The identification of patients with high cardiovascular risk in subclinical phase of atherosclerotic process is critical for primary prevention of cardiovascular diseases. Genetics in combination with clinical parameters and carotid phenotypes may help us to understand pathogenesis of atherosclerotic diseases which might improve the prediction of cardiovascular diseases. However, the functional consequences of SIRT1 and FOXO1 genetics in regulation of carotid atherosclerosis remain incompletely understood and should be investigated in further functional studies.

Abbreviations
BMI: Body mass index; CCA: Common carotid artery; DNA: Deoxyribonucleic acid; FOXO1: Forkhead box protein O1 (protein); FOXO1: Forkhead box protein O1 (gene, human); Foxo1: Forkhead box protein O1 (gene, mouse); HDL: High-density lipoprotein; hsCRP: High-sensitivity C-reactive protein; HWE: Hardy-Weinberg equilibrium; IC: Internal carotid artery; IMT: Intima-media thickness; LD: Linkage disequilibrium; LDL: Low-density lipoprotein; MAF: Minor allele frequency; NAD+: Nicotinamide adenine dinucleotide; NFκB: Nuclear factor ‘kappa-light-chain-enhancer’ of activated B-cells; SAPHIR: Salzburg Atherosclerosis Prevention Program in subjects at High Individual Risk; SIRT1: Sirtuin 1 (protein); SIRT7: Sirtuin 1 (gene, human); SNP: Single nucleotide polymorphism; T2DM: Type 2 diabetes mellitus.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
LK designed and supervised the study, carried out the genotyping and drafted the manuscript. CL performed the statistical analysis and drafted the manuscript. FK carried out the genotyping and managed the SAPHIR study. BK performed the statistical analysis and helped to draft the manuscript. TK carried out the genotyping and helped to draft the manuscript. Bi performed ultrasonography and managed the SAPHIR study. FK designed and supervised the study, drafted the manuscript. BP designed and supervised the study, managed the SAPHIR study and drafted the manuscript. All authors read and approved the final manuscript.

Acknowledgements
The SAPHIR population was supported by Kamillo-Eisner Stiftung and Salzburger Forschungsgesellschaft. Current study was funded by the Forschungsförderungsfond of the Paracelsus Medical University (grant Nr. E-09/09/055-PAU). The authors would like to thank the field investigators of the SAPHIR cohort for their assistance with data collection. We appreciate the technical assistance of Fabienne Buchsteiner from the University Clinic for Internal Medicine I for TaqMan genotyping. Above all, the authors thank the study participants.

Author details
1 First Department of Internal Medicine, Paracelsus Medical University/ Salzburger Landeskliniken, Mullner Hauptstrasse 48, 5020 Salzburg, Austria.
2 Division of Genetic Epidemiology, Innsbruck Medical University, Schöpfstrasse 41, 6020 Innsbruck, Austria.
3 Institute of Physiology and Pathophysiology, Paracelsus Medical University, Strubergasse 21, 5020 Salzburg, Austria.
4 Department of Geriatric Medicine, Paracelsus Medical University/Christian-Doppler-Klinik, Ignaiz-Hanner-Straße 79, 5020 Salzburg, Austria.

Received: 28 June 2014 Accepted: 24 September 2014 Published online: 02 October 2014
Insulin stimulates phosphorylation of the Forkhead transcription factors. J Biol Chem 2008, 283:2392–2396.

14. Mostoslavsky R, Cohen HY, Hu LS, Cheng HL, Jedrychowski MP, Gygi SP, Kenedko i et al. Increased FoxOs integrate pleiotropic actions of insulin in Mammalian SIRT1 represses forkhead transcription factors. J Biol Chem 2005, 280(20):20599–20605.

15. Wang L, Cabral D, Beecham A, McClendon MS, Bonithon-Kopp C, Touboul PJ, Berr C, Leroux C, Mainard F, Courbon D, Ducimetiere P: Rationale and design for the Asymptomatic Carotid Artery Plaque 21. Genetic architecture of the APM1 gene and its influence on adiponectin plasma levels and parameters of the metabolic syndrome in 1,727 healthy Caucasians. Diabetologia 2006, 49(5):375–384.

16. Halldinsalmi S, Vepsäläinen S, Hiltunen M, Kivistö AM, Salminen A, Laakso M, Soininen H: Genetic study between SIRT1, PPARD, PGC-1α genes and Alzheimer's disease. J Neural 2008, 255:5668–673.

17. Li J, Li L: Adjusting multiple testing in multilocus analyses using the eigenvalues of a correlation matrix. Heredity (Edinb) 2005, 95(3):221–227.

18. Sinnwell JP: SD. Haplo.stats:Statistical Analysis of Haplotypes with Traits and Covariates when Linkage Phase is Ambiguous. R package version 1.4.4, 2009.

19. Barrett JC, Fry B, Maller J, Daly MJ: Haploview: analysis and visualization of LD and haplotype maps. Bioinformatics 2005, 21(2):263–265.

20. Xiong S, Salazar G, Patrushev N, Alexander RW: FoxO1 mediates an autofeedback loop regulating SIRT expression. J Biol Chem 2011, 286(7):5289–5299.

21. Maiese K, Cheng ZZ, Chang YC, Hou J: Clever cancer strategies with FoxO transcription factors. Cell Cycle 2008, 7:2838–2839.

22. Maiese K, Cheng ZZ, Chang YC, Hou J: A “FOXO” in sight: targeting FoxO proteins from conception to cancer. Med Res Rev 2009, 29(3):395–418.

23. Qiang L, Banks AS, Accili D: Uncoupling of acetylcyanation from phosphorylation regulates FoxO1 function independent of its subcellular localization. J Biol Chem 2010, 285(26):23796–23791.

24. Dong Y, Guo T, Traurig M, Mason C, Kubes S, Perez J, Knowler WC, Bogardus C, Hanson RL, Bailer JR: SIRT1 is associated with a decrease in acute insulin secretion and a sex specific increase in risk for type 2 diabetes in Pima Indians. Mol Genet Metab 2011, 104(4):661–665.

25. Weyrich P, Machicao F, Reinhart J, Machann J, Schick F, Tschritter O, Stefan N, Fritsche A: Haring HU: SIRT1 genetic variants associate with the metabolic response of Caucasians to a controlled lifestyle intervention–the TULIP Study. BMC Med Genet 2008, 9:100.

26. de Kreutzenberg SV, Ceolotto G, Papparella I, Bortoluzzi A, Semplicini A, Luca CC, Blanton SH, Sacco RL, Rundek T: Association of the sirtuin and Alzheimer pathways in men and women: heritability of carotid intima-media thickness as an indicator of atherosclerosis at other sites of the carotid. Stroke 2003, 34(2):279–283.

27. Clark SJ, Fajlach M, Olsson B, Jacobson P, Cauchy S, Balkau B, Marre M, Lantieri O, Andersson JC, Jernas M, Altman TJ, Richardson S, Sjoorp M, Wong HY, Carlsson LM, Farghely M, Winter AJ: Association of sirtuin 1 (SIRT1) gene SNPs and transcript expression levels with severe obesity. Obesity (Silver Spring) 2012, 20(1):178–185.

28. Mussig K, Staiger H, Machicao F, Stancakova A, Kuustio J, Laakso M, Thamer C, Schick F, Clausen CD, Stefan N, Fritsche A: Haring HU: Association of common genetic variation in the FOXO1 gene with beta-cell dysfunction, impaired glucose tolerance, and type 2 diabetes. J Clin Endocrinol Metab 2009, 94(1):1353–1360.

29. Shimoyama Y, Suzuki K, Hamaajima N, Niiwa T: Sirtuin 1 gene polymorphisms are associated with body fat and blood pressure in Japanese. J Cardiol Res 2013, 15(3):339–347.

30. Zillikens MC, van Meurs JB, Rivadeneira F, Amin N, Hofman A, Oostra BA, Sijbrands EJ, Witteman JC, Pols HA, van Duijn CM, Uitterlinden AG: SIRT1 genetic variation is related to BMI and risk of obesity. Diabetes 2009, 58(12):2828–2834.

31. Heid M, Wagner R, Gohlke H, Isgleider B, Mueller JC, Cip P, Ladunauer G, Renter R, Stadtmayr A, Mackevics V, Illig T, Kronenberg F, Faulweber B.
41. Greenland P, Alpert JS, Beller GA, Benjamin EJ, Budoff MJ, Fayad ZA, Foster E, Hlatky MA, Hodgson JM, Kushner FG, Lauer MS, Shaw LJ, Smith SC Jr, Taylor AJ, Weintraub WS, Wenger NK, Jacobs AK: 2010 ACCF/AHA guideline for assessment of cardiovascular risk in asymptomatic adults: a report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines. Circulation 2010, 122(25):e584–e636.

42. Stein JH, Korcarz CE, Hurst RT, Lonn E, Kendall CB, Mohler ER, Najjar SS, Rembold CM, Post WS: Use of carotid ultrasound to identify subclinical vascular disease and evaluate cardiovascular disease risk: a consensus statement from the American Society of Echocardiography Carotid Intima-Media Thickness Task Force. Endorsed by the Society for Vascular Medicine. J Am Soc Echocardiogr 2008, 21(2):93–111. quiz 189–190.

43. Johnsen SH, Mathiesen EB, Joakimsen O, Stensland E, Wilsgaard T, Lochen ML, Njolstad I, Arnesen E: Carotid atherosclerosis is a stronger predictor of myocardial infarction in women than in men: a 6-year follow-up study of 6226 persons: the Tromso Study. Stroke 2007, 38(11):2873–2880.

44. Stensland-Bugge E, Bonaa KH, Joakimsen O, Njolstad I: Sex differences in the relationship of risk factors to subclinical carotid atherosclerosis measured 15 years later: the Tromso study. Stroke 2000, 31(3):574–581.

45. Iglseder B, Cip P, Mallamare I, Ladurner G, Paulweber B: The metabolic syndrome is a stronger risk factor for early carotid atherosclerosis in women than in men. Stroke 2005, 36(6):1212–1217.

doi:10.1186/s12881-014-0112-7

Cite this article as: Kedenko et al.: Genetic polymorphisms at SIRT1 and FOXO1 are associated with carotid atherosclerosis in the SAPHIR cohort. BMC Medical Genetics 2014 15:112.