Biosurfactants’ Potential Role in Combating COVID-19 and Similar Future Microbial Threats

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Abstract: During 2020, the world has experienced extreme vulnerability in the face of a disease outbreak. The coronavirus disease 2019 (COVID-19) pandemic discovered in China and rapidly spread across the globe, infecting millions, causing hundreds of thousands of deaths, and severe downturns in the economies of countries worldwide. Biosurfactants can play a significant role in the prevention, control and treatment of diseases caused by these pathogenic agents through various therapeutic, pharmaceutical, environmental and hygiene approaches. Biosurfactants have the potential to inhibit microbial species with virulent intrinsic characteristics capable of developing diseases with high morbidity and mortality, as well as interrupting their spread through environmental and hygiene interventions. This is possible due to their antimicrobial activity, ability to interact with cells forming micelles and to interact with the immune system, and compatibility with relevant processes such as nanoparticle synthesis. They, therefore, can be applied in developing innovative and more effective pharmaceutical, therapeutics, sustainable and friendly environmental management approaches, less toxic formulations, and more efficient cleaning agents. These approaches can be easily integrated into relevant product development pipelines and implemented as measures for combating and managing pandemics. This review examines the potential approaches of biosurfactants as useful molecules in fighting microbial pathogens both known and previously unknown, such as COVID-19.

Keywords: biosurfactants; COVID-19; antiviral; antimicrobial

1. Introduction

It is generally accepted that a pandemic occurs when an infectious disease agent spreads to multiple borders or continents; the WHO declares a pandemic when a disease spreads worldwide. Some characteristics have been developed to describe what a pandemic is, mostly drawn from two factors: the intrinsic characteristics of the microbe, and the way the microbe interacts with humans. These characteristics include novelty, little or no immunity in the population, explosiveness in relation to the type of transmission, speed of transmission, a widespread common source, contagiousness, and severity [1]. This is the case with COVID-19 which began spreading across the world in early 2020.

The COVID-19 pandemic has affected many economies in the world. Huge spending, debt and loss of human lives are just some of the heavy losses affected countries have suffered. Most developed countries which were otherwise thought to be prepared to face outbreaks have the highest infection rates and number of deaths. Many countries were apparently not adequately prepared, despite lessons learnt from previous outbreaks. The COVID-19 pandemic provides another opportunity to evaluate outbreak or pandemic preparedness and take into consideration all the resources that could help prepare better responses in future pandemics. At a time when the UN sustainable development
goals have galvanized governments around the world to implement transitions towards more sustainable green and knowledge-based economies, preparing for future outbreaks should integrate sustainable bio-based products. One such group of products is biosurfactants. They have many advantages, such as better biodegradability, suitable for a circular economy, are safer and more sustainable compared to synthetic petroleum-based products.

There are growing research interests in biosurfactant-based products and their applications. Large-scale commercialization and effective integration into industrial processes will require multisectoral collaboration of various stakeholders such as policymakers, funding agencies, industry, and academia. Interdisciplinary research involving collaborations between clinicians, industry and researchers is also required as the main factor to kick-start innovation to provide transformative and disruptive solutions for fighting disease outbreaks. Stakeholders are therefore challenged by this opportunity to lead research and innovation for the fight against future pandemics [2]. Outbreaks can be caused by xenobiotics or microorganisms of bacterial, fungal, or viral origin. Other secondary elements can help the start outbreaks or facilitate the propagation of already existing ones such as drug resistance, use of chemicals or human manipulation. Infectious agents can be rapidly transmitted from person-to-person through three main routes: airborne, vector-borne, and waterborne.

The diversity of these factors shows that for a truly effective response system to be developed, innovative approaches in the development of therapeutics, environmental management and hygiene must be integrated. As the challenges of large-scale production are overcome, more specific applications are expected to be revealed in future. The aim of this review article, therefore, is to present relevant approaches that could be used to develop innovative sustainable solutions using biosurfactants applicable to the management of COVID-19 and preparedness for possible outbreak.

1.1. Biosurfactants

Biosurfactants are amphiphilic compounds synthesized by bacteria, fungi, or plants, capable of lowering the surface tension of liquids. Their hydrophilic moieties can be made of acid, cationic peptide, anion, sugar (monosaccharide, disaccharide, or polysaccharide) and hydrophobic moieties made of hydrocarbon or fatty acid chains. They are environmentally friendly and have been shown to have many industrial applications sometimes performing better than synthetic surfactants [3–7]. Their physico-chemical properties include; high stability in a wide range of environmental conditions such as extreme pH, temperature and also salt concentration [8,9], high biodegradability with a high rate of mineralization by soil microcosms [10], low toxicity, surface tension reduction, foaming capacity and antimicrobial activity against pathogens [11–14].

Well known classifications of biosurfactants are based on their charges and molecular structures. Biosurfactants obtained from microbes are generally anionic or neutral, while a few are cationic [7]. Long-chain fatty acids generally characterize the hydrophobic moiety, while organic acid, alcohol, amino acid or carbohydrate functional groups characterize the hydrophilic moiety [7]. They are also broadly grouped based on their chemical structures as high molecular weight and low molecular weight molecules. Low molecular weight molecules are the glycolipids, phospholipids and lipopeptides, while the high molecular weight groups are the polymeric and particulate biosurfactants [15] (Figure 1). In addition to their environmental friendliness and wide industrial applications, biosurfactants are more sustainable because they can be produced from cheap feedstock and industrial and urban waste, and can be recycled [16,17]. However, there are still challenges such as the pathogenicity of the main biosurfactant producer strains Pseudomonas and Bacillus and the relatively higher cost of large-scale production that make biosurfactants less commercially competitive than petroleum-derived synthetic surfactants [18]. Nevertheless, there is a growing scientific research interest in the improvement of the commercial competitiveness of biosurfactants. Bacterial strains are being engineered, new feedstocks are tested, and fermentation designs investigated to develop innovative ways to improve production effi-
ciency. Amidst these challenges, it is expected that biosurfactants will occupy a significant market share, which is projected to be about USD 5.52 billion by 2022, growing at a CAGR (compound annual growth rate) of 5.6% [19].

Figure 1. Representative biosurfactants produced by microorganisms utilizing water-soluble and/or water-insoluble substrates [3].

1.2. Pathogens and Other Agents with Outbreak or Pandemic Potential

RNA viruses are the class with the highest risks of an outbreak especially those that are airborne such as severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), because the mere act of breathing spreads the virus. Other organisms could potentially evolve or be engineered through drug resistance or human manipulation. With these factors in mind, the development of pipelines for both broad-spectrum and specific therapeutics could be important to add resilience against outbreak-causing pathogens. The first line of treatment
for these emerging diseases can be broad-spectrum antimicrobials, while specific treatment options could benefit from modifications that diversify the mechanism of action of such therapeutic molecules. This can also be integrated with other environmental approaches that disrupt transmission from the common source of spread in the population [20]. According to a report by the Intergovernmental Platform on Biodiversity and Ecosystem Services, there are over 1.7 million viruses unknown to man, with 540–850 thousand that can potentially infect humans. The cost of risk mitigation for these pandemics could be 100 times less than the cost of pandemic management. If the status quo on the fight of infectious diseases is not changed, we may see more outbreaks in the future with more damaging effects [21]. Some currently known diseases with high pandemic or outbreak importance include; Crimean–Congo hemorrhagic fever, chikungunya, cholera, Ebola virus disease, influenza (seasonal, zoonotic, pandemic), Hendra virus infection, Lassa fever, meningitis, Marburg virus disease, MERS-CoV, Nipah virus infection, monkeypox, novel coronavirus (2019-nCoV), SARS, smallpox, Tularaemia, plague, Rift Valley fever, Yellow fever and Zika virus disease [22]. Important bacterial-resistant strains include Escherichia coli, Staphylococcus aureus, Klebsiella pneumoniae and Mycobacterium tuberculosis, among others [20].

2. Approaches That Can Use Biosurfactants in Outbreak Prevention and Management

There are various areas where biosurfactants can be used as alternative sustainable solutions or as innovative components applicable in outbreak prevention and management. These include uses in therapeutics (vaccine development, immune system enhancers and drug delivery and developments), environmental applications as agricultural and pest biopesticides, and various industrial sectors as surface cleaning agents, disinfectants, detergents, and the packaging industries. All these applications are summarized below in Figure 2, and further discussed in the following subsections.

![Figure 2. A summary of potential approaches for applications of biosurfactants in future outbreak prevention and control.](image)

2.1. Therapeutics

Treatment availability is the most important element for the elimination of an outbreak, because the pathogen can be eliminated, thereby interrupting the transmission cycle and impeding the spread. However, the development of an effective drug against a disease takes years due to the required clinical trials and approval processes. Using already-approved low toxicity molecules or repurposing existing drugs to target a new pathogen can contribute to the development of effective therapy in a shorter time. Biosurfactants are one of the most promising biomolecules in the pharmaceutical industry because of their structural versatility, stability, micelle forming ability, biological compatibility and
low toxicity that are useful in the design of therapeutics. They can be safely used for oral, nasal, or dermal applications [23]. Another important characteristic of biosurfactants is their ability to interact with surfaces such as the membranes of organisms or with their surrounding environment, which can be useful in intracellular targeting.

Lipopeptides, glycolipids, and Mannosylerythritol lipids produced from Candida species are the most common microbial surfactants that have been investigated for pharmaceutical applications. They are relevant in pharmaceuticals for the delivery of drugs and genes to target cells, the design of molecules to interact with components of the immune system, and as antimicrobial agents. Biocompatibility, low toxicity, and environmental friendliness are advantages that make them better options over synthetic surfactants [18,24–26]. Biosurfactants have various innovative applications in the development of effective therapeutics which can come in handy at times of urgent need, such as during outbreaks or pandemics.

2.1.1. Delivery Systems

Delivery systems are structures to improve the effectiveness of drugs, but challenges such as poor delivery due to dilution by biological fluids and drug precipitation are amongst the main reasons for various failures in drug delivery system designs. Challenges faced when surfactants are used as delivery systems include the stability, packaging and formulations providing proper solubility in lipids, low drug loading capacity and risks of gastrointestinal irritations due to high amounts used [27].

Biosurfactants can form self-aggregating structures called micelles (Figure 3), which can act as good emulsifiers and have antimicrobial properties useful in the design of drug delivery systems. Biosurfactant-based microemulsion drug delivery systems can be used to make existing therapeutics more effective by either increasing the loading capacity, bioavailability, or provide more control in the release. They can encapsulate and solubilize hydrophobic and hydrophilic drugs in emulsions [25]. Microemulsion drug delivery systems are bio-compatible, thermodynamically stable, and usually highly surface active. Glycolipids and lipopeptides have been commonly used for this purpose and are a potential replacement for contemporary synthetic options [28]. These versatile microemulsion systems are emerging as novel drug delivery systems that can enable the modification of drug formulations for topical, oral, nasal, ocular, intravenous, or other routes of drug administration [25,27].

![Figure 3](image_url) (A) Biosurfactant molecule, (B) Reverse micelle structure, (C) Micelle structure.

Glycolipid sugar chains can also be used to target specific cells because they can be recognizable by carbohydrate-binding proteins on cell surfaces [29]. Through precise
modifications, this can be applied in targeting specific cells with intracellular pathogens such as viruses, thereby reducing spread to other less-affected cells. Glycolipid micelles can form structures such as liposomes, gels, niosomes, cubosomes and hexosomes [29] which can serve as a vehicle for the delivery of a drug to a target site while maintaining its integrity and that of the molecule carried in the body fluid environment. In the case of COVID-19, an aerosol formulation containing biosurfactant drug delivery system has been suggested to be a likely mode to deliver drugs, because the virulence is mainly observed in the lungs. In addition to delivery, another advantage is their antiviral activity which can inhibit viruses at the locus of infection [30]. In nanobiotechnology, applications of rhamnolipids as microemulsion stabilizers of different nanoparticles in drug delivery systems have also been reported. These include nickel oxide nanoparticles [31,32] and silver nanoparticles [33–36]. It is clear that biosurfactant-based drug delivery systems have the potential for better efficiency in drug delivery systems; research has increased around the exploration of the effects on virulent microorganism species that have outbreak potential.

2.1.2. Vaccines and Immunity

Coronavirus diseases such as severe acute respiratory syndrome (2002–2003) and Middle East respiratory syndrome (2011) were completely new agents of zoonotic origin. With COVID-19 infections, a normally functioning immune system clears the virus completely without the person developing symptoms. The innate and adaptive immunity play a primary role in this process because the immune system is still unfamiliar with the pathogen and is unable to work properly in some cases. Priming or inducing immune cells (T cells, B cells, macrophages, neutrophils, etc.) is crucial for an effective adaptive response. When T cells are primed, they clear viral infected cells through a cellular immune response as well as mediating the differentiation of B cells into plasma cells and memory cells which, respectively, produce viral-specific antibodies and store a memory of the response in case the infection returns [37]. Therefore, for new infections, adaptive immunity plays an important role through T cell activation. One way to safely activate T cells is by using vaccines. Peptide antigen vaccines are effective because they can be obtained at high purity compared to whole organisms or protein vaccines. However, they are limited by low immunogenicity.

Bacterial lipopeptides can be potent nontoxic, nonpyrogenic immunological adjuvants when coupled with antigens. They activate the immune system by signaling through toll-like receptor 2 (TLR2). Activity can vary considerably with different structures of lipopeptides providing great flexibility in the design of vaccine systems [38]. Deres and colleagues used lipopeptides as adjuvants with viral peptides to prime viral-specific cytotoxic T cells which are a component of the cellular immune response against viral infections. For activation to occur, T cells must recognize peptides from the virus coupled with the MHC (major histocompatibility complex) class I molecules displayed by cells infected by the virus. Tripalmitoyl-S-glycerylcysteinyl-seryl-serine lipopeptide has been covalently attached to a synthetic viral peptide which produced the same cytotoxic T cells mediated immune response observed with a live and infectious virus [39]. These formulations can prove to be effective in cases where there is no initial immunity to a pathogen, or as a resort to boost immunity in combination with other therapy.

The development of effective formulations with adjuvants that can safely boost immunity and enhance activity is a major challenge in vaccine development [40]. Some previous studies have investigated the role of surfactants in defense using bioactive peptides for the inactivation of enveloped viruses. The viral cycle of the influenza virus, for example, was reportedly inhibited by Cyclosporin A biopeptide produced by a fungus known as Tolypocladium inflatum through impeding viral assembly, or the budding (exiting step), after protein synthesis [41]. It was hypothesized that targeting these latter stages in the viral life cycle can overcome the problem of resistance for antiviral drugs, and hence limit disease spread.
2.1.3. Inflammation

COVID-19 infections are characterized by pulmonary and systemic inflammatory responses which cause cytokine storms. This results in hypersensitivity and the death of other healthy cells, leading to the manifestation of severe acute respiratory distress syndrome and disseminated intravascular coagulation, which are the main causes of mortality during infection. These storms may be because of an increase in the levels of inflammatory molecules such as IL-2, TNF-α and IL-6. Meanwhile, a low inflammatory response may promote a moderate T cell response [42]. The anti-inflammatory and antiviral role of biosurfactants has been demonstrated through cytokines (TNF-α, IL-6, IL-8, IL-12, IL-18 and IL-1β) and toll-like receptors-2 (TLR-2). After an inflammatory response, these factors can cause the secretion of proteins with cationic charge and other reactive oxygen species, including lysozyme, which can be used for therapeutic purposes. The reactive oxygen species produced have anti-inflammatory and antiviral properties which can be used as therapeutic agents against viral diseases. These can be used in the management of the cytokine storm, which is a cause of damages in the lungs as found in many COVID-19 patients. Although this hypothesis has not yet been tested or confirmed, it has been advanced as a possible mechanism which could be applied for the development of therapeutics against viral infections including COVID-19 [43].

2.1.4. Enzymes and Biocatalysts

There is a growing use of colloids in chemistry and biotechnology, particularly enzyme-containing reversed micellar systems. Biosurfactants are molecules with both hydrophobic and hydrophilic moieties, which, when introduced in nonpolar organic solvents, can self-aggregate when their concentration is greater than the critical micelle concentration. This aggregated structure contains an inner polar core made from polar structures and an outer nonpolar part consisting of hydrocarbon tails. The inner core provides a structure in which nano-sized particles can be contained and thermodynamically stabilized. Additionally, these structures are transparent enough to be seen under microscopes. It was also discovered that this inner environment mimics the internal environment of cells, which could be an explanation for the increase in enzyme activity. DNA cleavage enzymes have been tested both in vivo and in vitro, showing disruption in essential genes required for replication in Hepatitis B, HIV, HPV, HSV, HTLV viral infections [44]. The increasing use of CRISPR Cas technology in therapy shows that gene therapy will increasingly be used in many diseases. Enzyme delivery systems can be useful options in the treatment or delivery of enzyme components to the required target site. This approach could provide an opportunity to diversify the portfolio and pipelines for the treatment options considered for COVID-19 or other serious infections. There are numerous other applications in medicine, especially for protein extraction and bioactive molecule separation and purification, as well as enzyme drug delivery [45].

2.1.5. Probiotics

Probiotics are living bacteria that have health benefits to the human gut microbiota, either by improving or restoring it. Besides improving the gut microbiota, they can contribute to improving the immune system. Probiotic lactic acid bacteria have been reported to produce various biosurfactants with important applications in biotherapeutics due to their important antimicrobial and anti-adhesive activity. They can significantly improve the defense of the gastrointestinal microbiota in outbreaks that cause gastrointestinal diseases. The main producers of probiotics include Lactobacillus and Bifidobacterium. Other lactic acid bacteria include Enterococcus, Leuconostoc, Pediococcus, Sporolactobacillus, Streptococcus, Lactococcus, and non-lactic acid bacteria include Propionibacterium, Bacillus cereus, Saccharomyces, Bifidobacterium lactis, and Escherichia coli strain nissle [46].

Probiotic bacteria producing biosurfactants have shown anti-microbial activity against Gram-negative and Gram-positive bacteria, including yeast and other possible human pathogens [46]. These include Lactobacillus paracasei [47], glycolipid from Lactococcus lactis
with activity against multidrug-resistant (MDR) pathogens [48], *Lactobacillus helveticus* [49], lipopeptide from *Bacillus cereus* NK1 [50], *L. paracasei* ssp *paracasei* A20 [51], *Lactobacillus casei* MRTL3 [52] and *Lactobacillus acidophilus* [53,54].

There is some evidence that probiotics may be able to decrease the risk and duration of viral respiratory infections. This may, therefore, be helpful for the management of COVID-19 infections. Although the actual mechanism of action is undefined, clear benefits were demonstrated in some initial trials. It was hypothesized that this may be through direct interaction with the virus or stimulation of the immune system. Research to investigate specific viral effects, especially on RNA viruses and other respiratory viruses, is recommended because this could provide a safe method to fight related infections in emergencies [55].

2.1.6. Drug Development

Chemical modifications of biomolecules, especially lipids, have numerous applications in chemistry and pharmacokinetics. The hydrogen atom deuterium is a stable hydrogen isotope, having both a proton and a neutron within its nucleus. Deuterium oxide (D$_2$O) is well known in medicinal chemistry, and it is being used in the modification of various biomolecules for pharmaceutical purposes [56]. Deuterated biosurfactants are produced by bacteria strain AD7 of *Pseudomonas aeruginosa* with varying levels of D$_2$O and carbon substrates. These molecules are safer and can be used to monitor drug metabolism within biological systems. These are used as substrates, therefore the level of deuteration of the biosurfactant can be manipulated. The flexibility of a biosurfactant used as an adjuvant can enable modifications on antibiotics and other drugs, which can result in the improved performance of an existing antibiotic, or even restrict the development of antimicrobial resistance with the possibility of using a small dose of the drug [57]. Besides pharmacokinetics, the antimicrobial properties of biosurfactants can be exploited in drug development. *Rhodococcus fascians* BD8 isolated from artic soil was found to produce a trehalose lipid biosurfactant with antimicrobial activity against the drug-resistant bacteria *Vibrio harveyi* and *Proteus vulgaris*. Partial inhibition at 11–34% was observed on other Gram-positive and Gram-negative bacteria, as well as 30% inhibition of *Candida albicans* at 0.5 mg/mL concentration [58].

In combination with existing drugs, synergistic effects between biosurfactants and antibiotics have also been reported. Methicillin-resistant *Staphylococcus aureus* was inhibited by using a joint sophorolipid and tetracycline treatment in vitro. Bacteria inhibition was observed at concentrations below the minimum inhibitory concentration. Understanding the underlying mechanisms of synergy between biosurfactants and antibiotics could prove useful in the development of effective treatments against drug-resistant pathogens [59]. Another way to develop effective treatments is the use of precision antimicrobials. Their applications in personalized medicine as a way of reducing the effects of drugs in the body are rapidly expanding. This novel approach to treating infectious diseases can use biosurfactants [60].

Bacaucin is a peptide biosurfactant isolated from *Bacillus subtilis* strain CAU21, which was reported to have broad-spectrum antimicrobial properties against Gram-positive bacteria but with haemolytic and cytotoxic effects. When the lipid portion is removed and the ring of the heptapeptide opened, bacaucin-1 is produced, which has more hydrophilic portions exposed, resulting in an overall decrease in the hydrophobicity of the molecule. Bacaucin-1 was selectively active against antibiotic-resistant *S. aureus* strains through cell membrane disruption, and demonstrated no bacterial resistance and cytotoxicity to mammalian cells in vitro and in vivo [61]. Structural modification of biosurfactants could lead to the development of highly precise drugs which can be used against pathogens sharing many similarities with human cells.

Biosurfactants from *Bacillus subtilis* have also shown in vitro antiviral properties against the enveloped virus species such as herpes, retroviruses, and other non-enveloped viruses. These groups share structural similarity with pathogens such as HIV, MERS, SARS, and Hepatitis. *Bacillus subtilis* surfactin inactivated the viruses at concentrations of 25 µm–80 µm. The mechanism of action was through the destruction of viral lipid membranes and capsid [62]. The formation of ion channels in the viral capsids and lipid
envelopes leading to the loss of proteins involved in the process of membrane attachment, fusion and penetration has been reported [63]. Other results have described the effects of surface-active lipopeptide mixtures and surfactin analogues against Newcastle disease virus and Porcine epidemic diarrhea virus, emphasizing the idea of potential use as new antiviral drugs [63–65]. The wide antiviral properties show their possible applications in various pharmaceutical formulations against enveloped viruses such as SARS-CoV-2.

2.2. Applications in Diagnostics

2.2.1. Nanomaterials and Nanotechnology

Biosensors (typically simple, sensitive, robust, and cost-effective) combined with nanomaterials, also known as nanobiosensors, can serve as a bridge between advanced diagnostics/detection and routine testing. It is essential that the production processes for nanoparticles become cleaner, less toxic, and more environmentally safe so that the negative impact on the environment from waste can be minimized. Microorganisms are capable of synthesizing inorganic molecules that can be deposited either intracellularly or excreted extracellularly. Sophorolipid-capped cobalt nanoparticles can be used to generate biocompatible particle surfaces by the attachment of bioactive molecules such as glycosidases or lectins for diagnostic and medicinal applications. An important issue of biocompatibility properties is the accessibility of the sophorose group at the surface of the nanoparticles [66]. Silver nanoparticles synthesized with purified rhamnolipids from P. aeruginosa BS-161R strain demonstrated broad-spectrum antimicrobial activity against Gram-negative and Gram-positive bacteria, and against Candida albicans [35].

Nanotechnology is a relatively new field which is growing very rapidly, and nanoparticles are being used in sophisticated devices and in treatment. As the need for nanoparticles grows, it will be necessary to make production environmentally friendly and sustainable. This can be achieved using the biosurfactant-mediated production of nanoparticles. Diagnostics using nanotechnology are the basis for highly sensitive and specific diagnostic devices that require very small sample amounts. During outbreaks, the availability of highly accurate and specific diagnostics is important in the proper identification of the causative agent, and subsequently building a response.

2.2.2. Contrasting Agents

Microbubbles synthesized with biosurfactants can offer an option for the synthesis or the development of non-invasive, low-cost, and highly specific diagnostics. They can be applied in diagnoses using molecular imaging. For the diagnosis of specific diseases, the bubble surface can be chemically modified or conjugated to a disease-specific ligand. When retained in the targeted tissue and detected with ultrasound by using it as a contrast, it can be a means for specific and sensitive diagnosis in early disease detection and progression [67,68].

2.3. Environmental Approaches

2.3.1. Environmental Control and Management against Potential Outbreaks

Wastewater remains a significant transmission route of pathogens that have an outbreak or pandemic potential. This poses a significant challenge, especially for health authorities when it comes to testing and understanding the transmission dynamics of pathogens. In the case of the current COVID-19 pandemic, the role of wastewater treatment in the transmission of the virus remains unknown [69]. Detecting pathogen DNA or RNA in wastewater samples can provide an early warning system in the transmission of the pathogen in the community. Therefore, early warning systems in wastewater treatment facilities using optimized protocols for sampling, sample storage and recommended concentrations of genetic material can be an important tool for the early detection, response, and management of an outbreak [70]. Biosurfactants have been shown to have broad-spectrum antimicrobial activity, therefore their applications in the treatment of wastewater can serve as an early and safe method for managing sewage in sewage treatment facilities [71].
Wastewater pollution can be a cause of thalassogenic infectious diseases when poorly treated wastewater is disposed into the sea. The WHO estimates that, globally, 120 million cases of gastrointestinal diseases and about $5 \times 10^7$ cases of severe respiratory diseases are caused by swimming in wastewater polluted waters [72]. *Bacillus amyloliquefaciens* ST34 and *Pseudomonas aeruginosa* ST5 are biosurfactant-producing bacteria that have been isolated from wastewaters, producing surfactin and rhamnolipids with antimicrobial activity and a broad spectrum of other pathogens including drug-resistant *Staphylococcus aureus*, *Escherichia coli* strains and *Candida albicans* [73]. This indicates that they could have an important role in the initial biological treatment stages of wastewater. Poorly treated sewage from hospitals and quarantine facilities can be a potential source of the spread of pathogens. Wastewater treatment was shown to be enhanced when a lipopeptide was applied to a lignocellulosic biocomposite. This resulted in a boost of the adsorption characteristics of the biocomposite through an increase in its stability, roughness, sharpness, and roundness [71]. By trapping compounds such as heavy metals and xenobiotics, and reducing the BOD in water treatment process, this contributes to a decrease in microbial pathogen populations. Various combinations of biosurfactant-producing bacteria have been isolated from wastewater. A combination of biosurfactant-producing bacteria with very broad spectrum activity could be considered as additional treatment options in wastewater facilities.

### 2.3.2. Vector Control

Infectious disease transmission can be from person-to-person or through a biological intermediate organism called a vector. Vectors to most important infectious diseases are insects which are commonly found in our environment. Important viral diseases such as dengue, chikungunya and Zika have all been causes of serious outbreaks in many regions worldwide. Their vector, *Aedes* sp. mosquito, is in contact with over half of the world’s population [74]. Protozoans are thought to have an unlimited pandemic potential; they are the only species which vector infectious diseases that have caused the extinction of a mammalian species. This was observed with *Trypanosoma lewisi*, a vector-borne disease that made the Christmas Island rat (*Rattus macleari*) in Australia extinct around the early 20th century [75]. Furthermore, the malaria-causing protozoan *Plasmodium* was also thought to have killed 50% of all humans that existed on earth [74]. Trypansomiasis and other vector-borne pathogens can be confined to a geographical region because of the limitation of movement of the vector pathogen and the inability to survive in various earth climatic regions [74]. Nevertheless, their impact on human health cannot be overlooked.

Biosurfactant-containing biopesticides are biodegradable, show lower resistance in insect populations, have higher selectivity and biological safety compared to non-target species, in addition to being highly effective at lower concentrations [76]. Biosurfactant lipopeptides have been used as bioactive components in biopesticide formulations containing *Bacillus thuringiensis*. These formulations are lethal to the pupal and larval stages of insect vectors, with mosquitocidal activity against *Aedes aegypti*, the vector of dengue fever [77], *Anopheles stephensi*, the primary mosquito vector of malaria in India [78], and *Culex quinquefasciatus*, a vector for arboviruses and avian malaria [79]. ZnO nanoparticles synthesized with *B. licheniformis* EPS showed toxicity against the larvae of malaria and Zika virus vectors *Anopheles stephensi* and *Aedes aegypti*, with high biocompatibility and non-toxicity demonstrated on hemolysis potential tests [80]. Insect control through outdoor and indoor spraying is an important control approach in disease control programs in regions affected by vector-borne diseases. Conventional pesticides such as DDT have issues of environmental toxicity and developing insect resistance. Biosurfactants could be the solution to more effective and environmentally friendly biopesticides use.

### 2.4. Hygiene and Personal Protective Equipment

Outbreak prevention and management requires the application of integrated approaches in combination with therapeutics. For vector-borne diseases, environmental control is certainly important. In the case of infectious diseases that can be transmitted
from person-to-person directly or indirectly through fomites, hygiene and personal protection is also an important factor to consider. The environmental and biological safety of the products to be used is not to be overlooked. Protective equipment such as masks are widely recommended to limit the spread of COVID-19, although gradually constitute an environmental problem especially with single usage masks that are discarded everywhere. Although conventional facemasks offer protection against pathogens in aerosols by acting as a barrier, they are not lethal to the pathogens. This implies there is still a risk of infection which is the reason for their one-time usage. Masks from biodegradable silk or biodegradable polymers conjugated with cotton can facilitate degradability and user-friendliness. Biosurfactants are biodegradable and less biologically toxic; masks with additional silk layers and impregnated with biosurfactant material can offer both effective filtering and lethality to pathogens [2].

Biosurfactants on the surface of materials can confer many benefits. The antimicrobial and antiadhesive and antibiofilm properties of biosurfactants have been reported [81]. Trehalose lipids specifically were investigated using modified polystyrene and silicone surfaces and shown to inhibition colonization on polystyrene and silicone surfaces [58,82]. Biosurfactants produced by *Lactobacillus rhamnosus* and *Lactobacillus jensenii* also showed antimicrobial, antiadhesive and antibiofilm activity against MDR bacteria *Acinetobacter baumannii* and *E. coli* on surfaces, at 25–100 mg/mL, inhibition of biofilm formation at 25–50 mg/mL, and dispersed already-formed biofilms of *S. aureus* and *A. baumannii* at 50–100 mg/mL [83].

Antimicrobial activity has also been observed with polyvinyl alcohol, as well as a polyvinyl alcohol–biosurfactant mixture in plastic and glassware [84]. This can enable the development of re-usable consumables and disinfectants for hospital equipment. Biosurfactants with higher molecular mass are generally considered to be better emulsifiers, forming long-lasting stable emulsions which are highly desirable properties in cleaning [85]. For detergency and cleaning, the amphiphilic nature of biosurfactants allows them to bind simultaneously to hydrophobic moieties of microbes with the fatty acid chains and water with their hydrophilic moieties. This results in emulsification that removes dirt from the surface followed by solubilization into small droplets [30].

Nosocomial infections could cause hospital-bound outbreaks within the hospital environment. Hospital areas are often characterized by rapid recontamination of disinfected areas, multi-drug resistance development potential, and limited time for the action of cleaning agents. Additionally, the use of synthetic surfactants does not prevent reinfection, can lead to increased pollution, and may provoke chemical sensitivity reactions in patients. Surfactant-based cleaning agents used in hospital environment showed promising results as alternatives to chemical-based surfactants, reducing >80% of *S. aureus*, *Pseudomonas* sp., *Candida* sp. and coliforms with bio-stabilization of the microbial load over time [86]. Additionally, biosurfactants isolated from psychrophiles can be used effectively at cold temperatures. Viral particles and pathogens often become inactive in cold conditions but reactivate when conditions become favorable. Most additives are also inactive at cold temperatures, which may make such environments a potential reservoir for pathogens. A more environmentally friendly and bactericidal effect can be obtained using biosurfactants in washing or storage requiring cold temperatures [9].

Furthermore, biosurfactants are structurally versatile and can be combined with enzymes in detergent formulations as well as to possibly make the formulation fully renewable [87]. Common household cleaning products and detergents contain typically 15–40% surfactants, which can often cause skin irritations as compared to biosurfactants which cause little or no irritation [30]. Initial commercialization of biosurfactant-based products for household and personal care are already seen with sophorolipids produced by Evonik [88]. Skin tenderness, biodegradability, good cleaning, and environmental friendliness are amongst the properties reported for these products [88]. Chemical companies are partnering to produce a new brand of renewable and biodegradable biobased household cleaning products, based on rhamnolipids currently used in Chile [89]. The ap-
Applications of biosurfactants in cleaning are many and fully unexplored. As next-generation green molecules, they can play a role in limiting the spread of both known and unknown pathogens during outbreaks through effective hygiene.

2.5. Food Outbreak Control

There are suggestions of the possible transmission of COVID-19 through frozen foods. Viable SARS-CoV-2 pathogens have been found in frozen food storage areas, food, and its packaging [90]. Despite evidence that cold temperatures can prolong the shelf life of most pathogens; this has not received much attention in the case of COVID-19. As a matter of fact, there is no conclusive information available regarding the duration of COVID-19 persistence in different environments and surfaces [91,92]. However, more specific studies have shown that SARS-CoV-2 can survive at 4 to −80 °C on refrigerated foods such as meat, fish, poultry, or pork for 14–21 days. News reports from the Chinese CDC also mentioned findings of traces of COVID-19 on frozen cod packaging [90]. There could be an even wider threat through retail cold stores, where people travel to purchase daily groceries and then disperse to different regions through transportation. Although the pathogenicity and infectivity of the virus at those conditions have not yet been ascertained, it remains a possible route for spread and propagation of the virus [93].

Other known serious food-borne outbreak causative pathogens include *Norovirus*, *Clostridium perfringens*, *Salmonella*, *Staphylococcus aureus* and *Campylobacter* [94]. Biosurfactants could provide another opportunity for the interruption of infectious disease spread through food, mainly due to their previously mentioned relevant applications in packaging and their potential use in cold environments and as additives to the food industry [95]. In addition, as a subsidiary use to food industry applications and due to biosurfactants’ abilities to act as effective detergents with the added benefit of antimicrobial activity, their use as surface cleaning products could be highly advantageous.

3. Conclusions and Future Perspectives

Pandemic preparedness and management are integrated approaches involving a wide range of measures that can be applied simultaneously to build resilience against the causative agent. Recognizing the characteristics and potential for certain microorganisms that can cause a pandemic and developing pipelines for drugs, vaccines, and other pharmaceuticals against these potential agents for both broad-spectrum and specific pathogens will be of significant benefit to the development of resilience against these agents in the future. Our environment is also a major transmission vehicle for various pathogens and agents capable of disease outbreaks. We need to understand the role it plays in these events, and develop warning and response systems while maintaining a healthy environment. Exponential disease spread through direct and indirect contact with people can be limited using sustainable biosurfactants in hygiene and cleaning product formulations. There are many opportunities for biosurfactants to be applied as direct agents on virulent microbes as well as in other relevant interventions. Future technologies such as nanobiotechnology and various drone applications are also seen to be compatible with biosurfactants. For the disinfection or biopesticide spraying of large surface areas, drones could be used to deploy biosurfactant-based products, or identify nanoparticles for laboratory diagnostics. This could be highly useful in outbreak emergencies. Although biosurfactants are increasingly showing innovative applications in various areas, being environmentally friendly and often more effective than synthetic surfactants, production is still relatively uncompetitive. More research directed towards reducing the cost of production, development of applications in unexplored areas, and to explore the effects of specific biosurfactants on specific pathogens to provide more conclusive evidence for further applications.

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References

1. Pitlik, S.D. COVID-19 Compared to Other Pandemic Diseases. Rambam Maimonides Med. J. 2020, 11. [CrossRef] [PubMed]
2. Chakhalan, D.; Shultz, R.B.; Miles, C.E.; Kohn, J. Opportunities for biomaterials to address the challenges of COVID-19. J. Biomed. Mater. Res. A 2020, 108, 1974–1990. [CrossRef] [PubMed]
3. Banat, I.M.; Franzetti, A.; Gandolfi, I.; Bestetti, G.; Martinotti, M.G.; Fracchia, L.; Smyth, T.J.; Marchant, R. Microbial biosurfactants: applications and future potential. Appl. Microbiol. Biotechnol. 2010, 87, 427–444. [CrossRef] [PubMed]
4. Banat, I.M.; Makkar, R.S. Potential commercial applications of microbial surfactants. Appl. Microbiol. Biotechnol. 2000, 53, 495–508. [CrossRef]
5. Çakmak, H.; Güngörmedi, G.; Dikmen, G.; Çelik, P.A.; Çabuk, A. The true methodology for rhamnolipid: Various solvents affect rhamnolipid characteristics. Eur. J. Lipid Sci. Technol. 2017, 119, 1700002. [CrossRef]
6. Marchant, R.; Banat, I.M. Microbial biosurfactants: Challenges and opportunities for future exploitation. Trends Biotechnol. 2012, 30, 558–565. [CrossRef]
7. Sobrinho, H.; Luna, J.M.; Rufino, R.D.; Porto, A.; Sarubbo, L.A. Biosurfactants: Classification, properties and environmental applications. Recent Dev. Biotechnol. 2013, 11, 1–29.
8. Ben Ayed, H.; Jridi, M.; Maahej, N.; Nastri, M.; Hmiedet, N. Characterization and stability of biosurfactant produced by Bacillus mojavensis A21 and its application in enhancing solubility of hydrocarbon. J. Chem. Technol. Biotechnol. 2014, 89, 1007–1014. [CrossRef]
9. Perfumo, A.; Banat, I.M.; Marchant, R. Going Green and Cold: Biosurfactants from Low-Temperature Environments to Biotechnology Applications. Trends Biotechnol. 2018, 36, 277–289. [CrossRef]
10. Lima, T.M.S.; Procópio, L.C.; Brandão, F.D.; Carvalho, A.M.X.; Tótola, M.R.; Borges, A.C. Biodegradability of bacterial surfactants. Biodegradation 2011, 22, 585–592. [CrossRef]
11. Akbari, S.; Abdurahman, N.H.; Yunus, R.M.; Fayaz, F.; Alara, O.R. Biosurfactants—A new frontier for social and environmental safety: A mini review. Biotechnol. Res. Innov. 2018, 2, 81–90. [CrossRef]
12. Hirata, Y.; Ryu, M.; Oda, Y.; Igarashi, K.; Nagatsuaka, K.; Furuta, T.; Sugiuira, M. Novel characteristics of sophorolipids, yeast glycolipid biosurfactants, as biodegradable low-foaming surfactants. J. Biosci. Bioeng. 2009, 108, 142–146. [CrossRef] [PubMed]
13. Liu, J.-F.; Mbadinga, S.M.; Yang, S.-Z.; Gu, J.-D.; Mu, B.-Z. Chemical structure, property and potential applications of biosurfactants produced by Bacillus subtilis in petroleum recovery and spill mitigation. Int. J. Mol. Sci. 2015, 16, 4814–4837. [CrossRef] [PubMed]
14. Toptay, Y.; Çelikdemir, M.; Tuncer, C.; Şahin, Y.B.; Çelik, P.A.; Burnak, N.; Çabuk, A.; Bütün, V. Optimization of a biosurfactant production from bacteria isolated from soil and characterization of the surfactant. Turk. J. Biochem. 2016, 41. [CrossRef]
15. Shoeb, E.; Akhlaq, F.; Badar, U.; Akhter, J.; Imtiaz, S. Classification and industrial applications of biosurfactants. Acad. Res. Int. 2013, 4, 243–252.
16. Banat, I.M.; Satpute, S.K.; Cameotra, S.S.; Patil, R.; Nyaynait, N.V. Cost effective technologies and renewable substrates for biosurfactants’ production. Front. Microbiol. 2014, 5. [CrossRef]
17. Henkel, M.; Müller, M.M.; Kügler, J.H.; Lovaglio, R.B.; Contiero, J.; Syldatk, C.; Hausmann, R. Rhamnolipids as biosurfactants from renewable resources: Concepts for next-generation rhamnolipid production. Process. Biochem. 2012, 47, 1207–1219. [CrossRef]
18. Naughton, P.J.; Marchant, R.; Naughton, V.; Banat, I.M. Microbial Biosurfactants: Current trends and applications in Agricultural and Biomedical industries. J. Appl. Microbiol. 2019, 127, 12–28. [CrossRef]
19. Markets and Markets Biosurfactants Market Analysis Recent Market Developments Industry Forecast to 2016–2022. Available online: https://www.marketsandmarkets.com/Market-Reports/biosurfactant-market-163644922.html?gclid=CjwKCAiApzXxBRAPEiwAiM3DQmrdr778fPDUjk3XChXXZD0tAKDIQHs7eTX44QDmAHOOo_t91YemOSRoC0n8QAvD_BwE (accessed on 26 January 2020).
20. WHO. Antimicrobial Resistance. Available online: https://www.who.int/news-room/fact-sheets/detail/antimicrobial-resistance (accessed on 24 October 2020).
21. IPBES. Media Release: IPBES #PandemicsReport: Escaping the “Era of Pandemics” IPBES. Available online: https://ipbes.net/pandemics (accessed on 2 November 2020).
22. WHO. WHO Disease outbreaks. Available online: http://www.who.int/emergencies/diseases/en/ (accessed on 24 October 2020).
23. Elshikh, M.; Marchant, R.; Banat, I.M. Biosurfactants: Promising bioactive molecules for oral-related health applications. FEMS Microbiol. Lett. 2016, 363. [CrossRef]
24. Cameotra, S.S.; Makkar, R.S. Recent applications of biosurfactants as biological and immunological molecules. Curr. Opin. Microbiol. 2004, 7, 262–266. [CrossRef]
25. Gudina, E.J.; Rangarajan, V.; Sen, R.; Rodrigues, L.R. Potential therapeutic applications of biosurfactants. *Trends Pharmacol. Sci.* 2013, 34, 667–675. [CrossRef] [PubMed]

26. Rawat, G.; Dhamasa, A.; Kumar, V. Biosurfactants: The next generation biomolecules for diverse applications. *Environ. Sustain.* 2020. [CrossRef]

27. Ohadi, M.; Shahrvash, A.; Dehghannoudeh, N.; Eslaminejad, T.; Banat, I.M.; Dehghannoudeh, G. Potential Use of Microbial Surfactant in Microemulsion Drug Delivery System: A Systematic Review. *Drug Des. Devel. Ther.* 2020, 14, 541–550. [CrossRef]

28. Ohadi, M.; Amir-Heidari, B.; Mosaffa, M.H.; Mirparizi, A.; Basir, M.; Dehghan-Noudeh, G. Encapsulation of Biosurfactant-Producing Bacillus licheniformis (PTCC 1320) in Alginate Beads. *Biotechnology* 2014, 13, 239–244. [CrossRef]

29. Faivre, V.; Rosilio, V. Interest of glycolipids in drug delivery: From physicochemical properties to drug targeting. *Expert Opin. Drug Deliv.* 2010, 7, 1031–1048. [CrossRef] [PubMed]

30. Smith, M.L.; Gandolfi, S.; Coshall, P.M.; Rahman, P.K.S.M. Biosurfactants: A Covid-19 Perspective. *Front. Microbiol.* 2020, 11. [CrossRef] [PubMed]

31. Palanisamy, P. Biosurfactant mediated synthesis of NiO nanorods. *Mater. Lett.* 2008, 62, 743–746. [CrossRef]

32. Palanisamy, P.; Raichur, A.M. Synthesis of spherical NiO nanoparticles through a novel biosurfactant mediated emulsion technique. *Mater. Sci. Eng. C* 2009, 29, 199–204. [CrossRef]

33. Farias, C.B.B.; Ferreira Silva, A.; Diniz Rufino, R.; Moura Luna, J.; Gomes Souza, J.E.; Sarubbo, L.A. Synthesis of silver nanoparticles using a biosurfactant produced in low-cost medium as stabilizing agent. *Electron. J. Biotechnol.* 2014, 17, 122–125. [CrossRef]

34. Kiran, G.S.; Sabu, A.; Selvin, J. Synthesis of silver nanoparticles by glycolipid biosurfactant produced from marine *Brevibacterium casei* MSA19. *J. Biotechnol.* 2010, 148, 221–225. [CrossRef]

35. Kumar, C.; Mamidyala, S.K.; Das, B.; Sridhar, B.; Devi, G.; Karuna, M. Synthesis of Biosurfactant-Based Silver Nanoparticles with Purified Rhamnolipids Isolated from *Pseudomonas aeruginosa* BS-161R. *J. Microbiol. Biotechnol.* 2010, 20, 1061–1068. [CrossRef]

36. Xie, Y.; Ye, R.; Liu, H. Synthesis of silver nanoparticles in reverse micelles stabilized by natural biosurfactant. *Colloid Surf. A* 2006, 279, 175–178. [CrossRef]

37. Chowdhury, M.A.; Hossain, N.; Kashem, M.A.; Shahid, M.A.; Alam, A. Immune response in COVID-19: A review. *J. Infect. Public Health* 2020. [CrossRef]

38. Zaman, M.; Toth, I. Immunostimulation by Synthetic Lipopeptide-Based Vaccine Candidates: Structure-Activity Relationships. *Front. Immunol.* 2013, 4. [CrossRef] [PubMed]

39. Deres, K.; Schild, H.; Wiesmüller, K.H.; Jung, G.; Rammensee, H.G. In vivo priming of virus-specific cytotoxic T lymphocytes with synthetic lipopeptide vaccine. *Nature* 1989, 342, 561–564. [CrossRef] [PubMed]

40. Kischkel, B.; Rossi, S.A.; Santos, S.R.J.; Travassos, L.R.; Taborda, C.P. Therapies and Vaccines Based on Nanoparticles for the Treatment of Systemic Fungal Infections. *Front. Cell Infect. Microbiol.* 2020, 10. [CrossRef] [PubMed]

41. Garoff, H.; Hewson, R.; Opstelten, D.-J.E. Virus Maturation by Budding. *Nature* 1989, 342, 454–455. [CrossRef] [PubMed]

42. Garcia, I.F. Immune Response, Inflammation, and the Clinical Spectrum of COVID-19. *Front. Immunol.* 2020, 11. [CrossRef]

43. Subramaniam, M.D.; Venkatesan, D.; Iyer, M.; Subbarayan, S.; Govindasami, V.; Roy, A.; Narayanasamy, A.; Kamalakannan, S.; Gopalakrishnan, A.V.; Thangarasu, R.; et al. Biosurfactants and anti-inflammatory activity: A potential new approach towards COVID-19. *Curr. Opin. Environ. Sci. Health* 2020, 7, 667–675. [CrossRef] [PubMed]

44. Weber, N.D.; Aubert, M.; Dang, C.H.; Stone, D.; Jerome, K.R. DNA cleavage enzymes for treatment of persistent viral infections: Recent advances and the pathway forward. *Virology* 2014, 454–455, 353–361. [CrossRef]

45. Liang, Y.; Yuan, X.; Zeng, G.; Zhong, H.; Li, H.; Wang, W. Effects of surfactants on enzyme-containing reversed micellar system. *Sci. China Chem.* 2011, 54, 715–723. [CrossRef]

46. Hajfarajollahi, H.; Eslami, P.; Mohktarani, B.; Moghhabi, K.A. Biosurfactants from probiotic bacteria: A review. *Biotechnol. Appl. Biochem.* 2018, 65, 768–783. [CrossRef] [PubMed]

47. Gudina, E.J.; Teixeira, J.A.; Rodrigues, L.R. Isolation and functional characterization of a biosurfactant produced by *Lactobacillus paracasei*. *Colloid Surf. B* 2010, 76, 298–304. [CrossRef] [PubMed]

48. Saravanakumari, P.; Mani, K. Structural characterization of a novel xylolipid biosurfactant from *Lactococcus lactis* and analysis of antibacterial activity against multi-drug resistant pathogens. *Bioresour. Technol.* 2010, 110, 8851–8854. [CrossRef]

49. Sharma, D.; Saharan, B.S. Functional characterization of biomedical potential of biosurfactant produced by *Lactobacillus helveticus*. *Biotechnol. Rep.* 2016, 11, 27–35. [CrossRef]

50. Sriram, M.I.; Kalishwaralal, K.; Deepak, V.; Gracerosopat, R.; Srisakthi, K.; Gurunathan, S. Biofilm inhibition and antimicrobial action of lipopeptide biosurfactant produced by heavy metal tolerant strain *Bacillus cereus* NK1. *Colloid Surf. B* 2011, 85, 174–181. [CrossRef] [PubMed]

51. Gudina, E.J.; Rocha, V.; Teixeira, J.A.; Rodrigues, L.R. Antimicrobial and antiadhesive properties of a biosurfactant isolated from *Lactobacillus paracasei* ssp. *paracasei* A20. *Lett. Appl. Microbiol.* 2010, 50, 419–424. [CrossRef]

52. Sharma, D.; Singh Saharan, B. Simultaneous production of biosurfactants and bacteriocins by probiotic *Lactobacillus casei* MRTL3. *Int. J. Microbiol.* 2014. [CrossRef]

53. Satpute, S.K.; Mone, N.S.; Das, P.; Banat, I.M.; Banpurkar, A.G. Inhibition of pathogenic bacterial biofilms on PDMS based implants by *L. acidophilus* derived biosurfactant. *BMC Microbiol.* 2019, 19, 1–15.
54. Satpute, S.K.; Mone, N.S.; Das, P.; Banpurkar, A.G.; Banat, I.M. *Lactobacillus acidophilus* derived biosurfactant as a biofilm inhibitor: A promising investigation using microfluidic approach. *Appl. Sci.* **2018**, *8*, 1535. [CrossRef]

55. Lehtoranta, L.; Pitkäranta, A.; Korpeila, R. Probiotics in respiratory virus infections. *Eur. J. Clin. Microbiol. Infect. Dis.* **2014**, *33*, 1289–1302. [CrossRef] [PubMed]

56. Yang, J. *Deuterium: Discovery and Applications in Organic Chemistry*; Elsevier: Amsterdam, The Netherlands, 2016; p. 116.

57. Smyth, T.J.; Perfumo, A.; Marchant, R.; Banat, I.M.; Chen, M.; Thomas, R.K.; Penfold, J.; Stevenson, P.S.; Parry, N.J. Directed microbial biosynthesis of deuterated biosurfactants and potential future application to other bioactive molecules. *Appl. Microbiol. Biotechnol.* **2010**, *87*, 1347–1354. [CrossRef] [PubMed]

58. Janek, T.; Krasowska, A.; Czyżnikowska, Z.; Łukaszewicz, M. Trehalose Lipid Biosurfactant Reduces Adhesion of Microbial Pathogens to Polystyrene and Silicone Surfaces: An Experimental and Computational Approach. *Front. Microbiol.* **2018**, *9*. [CrossRef] [PubMed]

59. Juma, A.; Lemoine, P.; Simpson, A.B.J.; Murray, J.; O’Hagan, B.M.G.; Naughton, P.J.; Dooley, J.G.; Banat, I.M. Microscopic Investigation of the Combined Use of Antibiotics and Biosurfactants on Methicillin Resistant *Staphylococcus aureus*. *Front. Microbiol.* **2020**, *11*. [CrossRef] [PubMed]

60. Brooks, B.D.; Brooks, A.E. Therapeutic strategies to combat antibiotic resistance. *Adv. Drug Deliv. Rev.* **2014**, *78*, 14–27. [CrossRef] [PubMed]

61. Liu, Y.; Ding, S.; Dietrich, R.; Martlbauer, E.; Zhu, K. A Biosurfactant-Inspired Heptapeptide with Improved Specificity to Kill MRSA. *Angew. Chem.* **2017**, *129*, 1508–1512. [CrossRef]

62. Vollenbroich, D.; Özel, M.; Vater, J.; Kamp, R.M.; Pauli, G. Mechanism of inactivation of enveloped viruses by the biosurfactant surfactin from *Bacillus subtilis*. *Biologicals* **1997**, *25*, 289–297. [CrossRef]

63. Basit, M.; Rasool, M.H.; Naqvi, S.A.R.; Waseem, M.; Aslam, B. Biosurfactants production potential of native strains of *Bacillus subtilis* subsp. *subtilis* strain. *Appl. Microbiol. Biotechnol.* **2010**, *86*, 1737–1744. [CrossRef] [PubMed]

64. Yuan, L.; Zhang, S.; Peng, J.; Li, Y.; Yang, Q. Synthetic surfactin analogues have improved anti-PEDV properties. *PLoS ONE* **2019**, *14*, e0215227. [CrossRef]

65. Yuan, L.; Zhang, S.; Wang, Y.; Li, Y.; Wang, X.; Yang, Q. Surfactin Inhibits Membrane Fusion during Invasion of Epithelial Cells by Enveloped Viruses. *J. Virol.* **2018**, *92*. [CrossRef]

66. Plaza, G.A.; Chojniak, J.; Banat, I.M. Biosurfactant mediated biosynthesis of selected metallic nanoparticles. *Int. J. Mol. Sci.* **2014**, *15*, 13720–13737. [CrossRef]

67. Katariya, H.B. The concept of microbubble as a drug delivery system: An overview. *J. Pharm. Sci. Res.* **2012**, *3*, 3058–3063. [CrossRef]

68. Xu, Q.; Nakajima, M.; Liu, Z.; Shiina, T. Biosurfactants for Microbubble Preparation and Application. *Int. J. Mol. Sci.* **2011**, *12*, 462–475. [CrossRef]

69. Al Huraimel, K.; Alhosani, M.; Kunhabdulla, S.; Stitiyia, M.H. SARS-CoV-2 in the environment: Modes of transmission, early detection and potential role of pollutions. *Sci. Total Environ.* **2020**, *744*, 140946. [CrossRef]

70. Michael-Kordatou, I.; Karaolia, P.; Fatta-Kassinos, D. Sewage analysis as a tool for the COVID-19 pandemic response and management: The urgent need for optimised protocols for SARS-CoV-2 detection and quantification. *J. Environ. Chem. Eng.* **2020**, *8*, 104306. [CrossRef]

71. Perez-Ameneiro, M.; Vecino, X.; Cruz Freire, J.M.; Moldes, A. Wastewater treatment enhancement by applying a lipopeptide biosurfactant to a lignocellulosic biocomposite. *Carbohydr. Polym.* **2015**, *131*, 186–196. [CrossRef]

72. Shuval, H. Estimating the global burden of thalassogenic diseases: Human infectious diseases caused by wastewater pollution of the marine environment. *J. Water Health* **2003**, *1*, 53–64. [CrossRef]

73. Ndlovu, T.; Khan, S.; Khan, W. Distribution and diversity of biosurfactant-producing bacteria in a wastewater treatment plant. *Environ. Sci. Pollut. Res.* **2016**, *23*, 9993–10004. [CrossRef]

74. Adalja, A.A.; Watson, M.; Toner, E.S.; Cicero, A.; Inglesby, T.V. Characteristics of microbes most likely to cause pandemics and global catastrophes. In *Global Catastrophic Biological Risks*; Adalja, A., Eds.; Springer: Cham, Switzerland, 2019; Volume 424, pp. 1–20. [CrossRef]

75. Wyatt, K.B.; Campos, P.F.; Kolokotronis, S.-O.; Hynes, W.H.; DeSalle, R.; Daszak, P.; MacPhee, R.D.E.; Greenwood, A.D. Historical Mammal Extinction on Christmas Island (Indian Ocean) Correlates with Introduced Infectious Disease. *PLoS ONE* **2008**, *3*, e3602. [CrossRef]

76. Mnif, I.; Ghribi, D. Potential of bacterial derived biopesticides in pest management. *Crop. Protect.* **2015**, *77*, 52–64. [CrossRef]

77. Manonmani, A.M.; Geetha, I.; Bhuvaneswari, S. Enhanced production of mosquitocidal cyclic lipopeptide from *Bacillus subtilis* ssp. *subtilis*. *Indian J. Med. Res.* **2011**, *134*, 476–482. [PubMed]

78. Geetha, I.; Manonmani, A.M.; Paily, K.P. Identification and characterization of a mosquito pupicidal metabolite of a *Bacillus subtilis* subsp. *subtilis* strain. *Appl. Microbiol. Biotechnol.* **2010**, *86*, 1737–1744. [CrossRef] [PubMed]

79. Das, K.; Mukherjee, A.K. Assessment of mosquito larvicidal potency of cyclic lipopeptides produced by *Bacillus subtilis* strains. *Acta Trop.* **2006**, *97*, 168–173. [CrossRef] [PubMed]

80. Abinaya, M.; Vaseeharan, B.; Divya, M.; Sharmili, A.; Govindarajan, M.; Alharbi, N.S.; Kadaikunnan, S.; Khaled, J.M.; Benelli, G. Bacterial exopolysaccharide (EPS)-coated ZnO nanoparticles showed high antibiofilm activity and larvicidal toxicity against malaria and Zika virus vectors. *J. Trace Elem. Med. Biol.* **2018**, *45*, 93–103. [CrossRef]
81. Banat, I.M.; De Rienzo, M.A.D.; Quinn, G.A. Microbial biofilms: Biosurfactants as antibiofilm agents. Appl. Microbiol. Biotechnol. 2014, 98, 9915–9929. [CrossRef]
82. Ceresa, C.; Fracchia, L.; Williams, M.; Banat, I.M.; Díaz De Rienzo, M.A. The effect of sophorolipids against microbial biofilms on medical-grade silicone. J. Biotechnol. 2020, 309, 34–43. [CrossRef]
83. Sambanthamoorthy, K.; Feng, X.; Patel, R.; Patel, S.; Paranavitana, C. Antimicrobial and antibiofilm potential of biosurfactants isolated from lactobacilli against multi-drug-resistant pathogens. BMC Microbiol. 2014, 14, 197. [CrossRef]
84. Salman, J.A.S.; Al Kadhemy, M.; Jaleel, M.; Abdal, A. Effect of PVA, PVA/biosurfactant on some pathogenic bacteria in glass and plastic plates. Int. J. Curr. Microbiol. Appl. Sci. 2014, 3, 301–309.
85. Nitschke, M.; Costa, S.G.V.A.O. Biosurfactants in food industry. Trends Food Sci. Technol. 2007, 18, 252–259. [CrossRef]
86. Vandini, A.; Frabetti, A.; Antonioli, P.; Platano, D.; Branchini, A.; Camerada, M.; Lanzoni, L.; Balboni, P.; Mazzacane, S. Reduction of the microbiological load on hospital surfaces through probiotic-based cleaning procedures: A new strategy to control nosocomial infections. J. Microbiol. Exp. 2014, 1, 00027.
87. Onaizi, S.A.; He, L.; Middelberg, A.P.J. Rapid screening of surfactant and biosurfactant surface cleaning performance. Colloids Surf. B Biointerfaces 2009, 72, 68–74. [CrossRef] [PubMed]
88. Focus on Surfactants Evonik Commercializes Biosurfactants. Focus Surfactants 2016, 2016, 3–4. [CrossRef]
89. Unilever Unilever and Evonik Partner to Launch Green Cleaning Ingredient. Available online: https://www.unilever.com/news/press-releases/2019/unilever-and-evonik-partner-to-launch-green-cleaning-ingredient.html (accessed on 22 October 2020).
90. The Guardian. Live Coronavirus Found on Frozen Food Packaging in China. Available online: http://www.theguardian.com/world/2020/oct/19/live-coronavirus-found-on-frozen-food-packaging-in-china (accessed on 24 October 2020).
91. Fiorillo, L.; Cervino, G.; Matarese, M.; D’Amico, C.; Surace, G.; Paduano, V.; Fiorillo, M.T.; Moschella, A.; La Bruna, A.; Romano, G.L.; et al. COVID-19 Surface Persistence: A Recent Data Summary and Its Importance for Medical and Dental Settings. Int. J. Environ. Res. Public Health 2020, 17, 3132. [CrossRef] [PubMed]
92. Cervino, G.; Fiorillo, L.; Surace, G.; Paduano, V.; Fiorillo, M.T.; De Stefano, R.; Laudicella, R.; Baldari, S.; Gaeta, M.; Cicciù, M. SARS-CoV-2 Persistence: Data Summary up to Q2 2020. Data 2020, 5, 81. [CrossRef]
93. Han, J.; Zhang, X.; He, S.; Jia, P. Can the coronavirus disease be transmitted from food? A review of evidence, risks, policies and knowledge gaps. Environ. Chem. Lett. 2020. [CrossRef]
94. CDC. Foodborne Illnesses and Germs. Available online: https://www.cdc.gov/foodsafety/foodborne-germs.html (accessed on 24 October 2020).
95. Campos, J.M.; Stamford, T.L.M.; Sarubbo, L.A.; de Luna, J.M.; Rufino, R.D.; Banat, I.M. Microbial biosurfactants as additives for food industries. Biotechnol. Prog. 2013, 29, 1097–1108. [CrossRef]