ABSTRACT

Objectives Vasohibin-1 (VASH1) is an endogenous angiogenesis regulator expressed in activated vascular endothelial cells. We previously reported that high VASH1 expression is a predictor of progression in acinar adenocarcinoma of the prostate. In this study, we evaluated the characteristics of ductal adenocarcinoma of the prostate by comparing the level of VASH1 expression between ductal and acinar adenocarcinoma specimens.

Design and setting A retrospective cohort study at two centres in Japan.

Participants Among the 1495 patients who underwent radical prostatectomy or transurethral resection for the past 15 years, a total of 14 patients diagnosed with ductal adenocarcinoma and 20 patients diagnosed with acinar adenocarcinoma with a Gleason score of 4+4 were included.

Interventions We immunohistochemically examined the CD34 expression as the microvessel density (MVD) and activated endothelial cells as the VASH1 density (vessels per mm²).

Primary and secondary outcome measures The primary outcome was the association of MVD and VASH1 density between ductal and acinar adenocarcinoma, and the secondary outcome was their oncological outcomes.

Results Nine patients (64.3%) with ductal adenocarcinoma were diagnosed at an advanced clinical stage, and five patients (35.7%) died from cancer during a median follow-up of 56.0 months. The VASH1 densities (mean±SD) in ductal and acinar adenocarcinoma were 45.1±18.5 vs 16.1±21.0 (p<0.001), respectively, while the MVD (mean±SD) in ductal and acinar adenocarcinoma were 65.3±21.9 vs 80.8±60.7 (p=0.666), respectively. The 5-year cancer-specific survival rates for high and low VASH1 expression were 70.0% and 100.0% (p=0.006), respectively. High VASH1 expression and a diagnosis of ductal adenocarcinoma were significant predictors of cancer-specific survival.

Conclusions Ductal adenocarcinoma was more aggressive and had higher VASH1 expression than acinar adenocarcinoma, although MVD was equivalent. These results indicate that VASH1 expression may serve as a novel biomarker for the aggressive nature of ductal adenocarcinoma.
However, ductal adenocarcinoma is often diagnosed at an advanced clinical stage with local or distant metastasis, making prostate-specific mortality significantly worse than that of acinar adenocarcinoma. Due to its rarity, ductal adenocarcinoma is poorly characterised immunohistochemically and genetically, and specific immunohistochemical biomarkers have not been established. The discovery of novel biomarkers for diagnostic significance or prognostic prediction and the identification of new targets for pharmacotherapy for ductal adenocarcinoma are, therefore, the focus of much research.

Angiogenesis is a critical biological process, which consists of neovascularisation. Angiogenesis is induced under certain pathological conditions such as wound healing, inflammation and cancer. Under normal conditions, angiogenesis is maintained by a complex balance of endogenous angiogenesis-stimulating and angiogenesis-inhibiting factors. The microenvironment of prostate cancer includes reactive stroma, comprised of multiple cell types which have been altered from their normal state to become permissive of cancer cell progression. During tumourigenesis, the angiogenic switch entails new blood vessels sprouting either directly by induction of growth factors, such as vascular endothelial growth factor (VEGF) and transforming growth factor-β, or indirectly by recruitment of host immune cells. Newly formed microvessels provide oxygen and nutrients, promoting tumour growth, malignant progression and metastasis formation.

One of the biomarkers that could reflect intratumoral angiogenic aggressiveness is the microvessel density (MVD); however, MVD may not reflect the angiogenic activity alone because it corresponds to the number of formed vessels, including quiescent ones. Vashibinin-1 (VASH1) is a novel endogenous angiogenesis regulator, which is specifically expressed in activated vascular endothelial cells (ECs). VEGF induces VASH1 via the VEGF receptor 2 and its downstream protein kinase inhibitor, which is specifically expressed in activated vascular endothelial cells (ECs). VEGF induces VASH1 via the VEGF receptor 2 and its downstream protein kinase inhibitor (VEGF) and transforming growth factor-beta, or indirectly by recruitment of host immune cells. Newly formed microvessels provide oxygen and nutrients, promoting tumour growth, malignant progression and metastasis formation.

In the present study, we examined the expression of VASH1 and MVD using transurethral resection of the prostate (TURP) or radical prostatectomy (RP) samples in patients with rare ductal adenocarcinoma and acinar adenocarcinoma with a Gleason score of 4+4. This study aimed to compare the level of VASH1 expression between ducal and acinar adenocarcinoma and investigate whether ductal adenocarcinoma possesses an aggressive angiogenesis behaviour and if it is related to clinical outcomes.

**METHODS**

**Patients**

We retrospectively analysed the clinical records of 1495 patients who underwent RP or TURP at two of our hospitals for the past 15 years. All pathological specimens were reviewed by dedicated uropathologists to unify the reproducibility of the diagnosis. At first, we excluded the intraductal carcinoma component for ductal adenocarcinoma. We exclusively extracted acinar adenocarcinomas with a Gleason score of 4+4; even adenocarcinomas with a minor amount of Gleason pattern 3 or 5 were excluded. In addition, we excluded the three following groups of patients: First, patients that had received neoadjuvant hormonal treatment before RP or TURP. Second, patients that were followed up at a different hospital. Third, patients who missed the follow-up appointment within the 6-month window after the primary treatment. The remaining 14 patients diagnosed with ductal adenocarcinoma and 20 patients diagnosed with acinar adenocarcinoma with a Gleason score of 4+4 were included. The clinical characteristics of patients are shown in table 1.

**Immunohistochemistry**

Tissue microarray blocks were constructed from formalin-fixed paraffin-embedded RP and TURP specimens from all 34 patients. For each specimen, two cores with a diameter of 4 mm were obtained from the two target areas of the tissue microarray blocks. We performed immunohistochemical staining for VASH1 and CD34 as previously reported. First, tissue sections were deparaffinised in xylene, hydrated by immersion in graded alcohols, and then distilled water. After antigen retrieval, endogenous peroxidase activity was blocked using 0.3% hydrogen peroxidase. The tissue sections were then incubated with a blocking solution containing 6% dry milk in phosphate-buffered saline (PBS). The primary antibodies used were mouse monoclonal antibodies (mAbs): anti-human VASH1 mAb diluted at a concentration of 4 µg/mL and anti-CD34 (Nichirei Biosciences, Tokyo, Japan). Mouse mAb was used against a synthetic peptide corresponding to the 286–299 amino acid sequence of VASH1, as suggested by Watanabe et al. Next, the tissue sections were incubated with secondary antibodies (Histofine Simple Stain MAX PO (M); Nichirei Biosciences) after washing with PBS. The colour was developed with 3, 3-diaminobenzidine tetra-hydrochloride in 50 mM Tris-HCl (pH 7.5) containing 0.005% hydrogen peroxide. Finally, the sections were counterstained with haematoxylin. The positive control slide CD34 antigen was prepared from paraffin-embedded
bladder cancer tissues with high MVD. Appropriate negative control slides for CD34 antigen and VASH1 were prepared by substituting the primary antibody with the immune globulin fraction of non-immune mouse serum at the same concentration in each staining run.

**Evaluation of immunostaining**

Two authors (HK and TKo) independently evaluated the immunoreactivity by counting the microvessels in proximity to the tumour using an Olympus IX71 microscope (Olympus, Tokyo, Japan). These authors were blinded to the clinical course of the patients, and the average of the numbers counted by the two investigators was used for subsequent analyses. The microvessels were identified based on their architecture, lumen lined by ECs and complemented by positivity of the ECs for anti-CD34 after scanning the immunostained section at low magnification (×40 and ×100). The areas with the highest number of distinctly highlighted microvessels were selected and counted at high magnification (×200). Any immunostained EC or cluster separated from the adjacent vessels was counted as a single microvessel, even in the absence of the vessel lumen. Each count was defined as the highest number of microvessels identified at the hotspot. The highest number of microvessels in the hotspot was counted to determine MVD. The VASH1-positive signals were counted in the hotspot area, in which the highest number of vessels positive for anti-CD34 was identified. We defined the number of VASH1-positive signals per mm² as the VASH1 density (vessels per mm²).

**Statistical analysis**

The $\chi^2$ or Mann-Whitney U test was used to compare the VASH1 density or MVD of each location between patients with ductal and acinar adenocarcinoma. We used a median VASH1 density of ≥30/mm² and a MVD of ≥60/mm² as the cut-off level. The probability of cancer-specific survival (CSS) was estimated using the Kaplan-Meier method, and the median follow-up was calculated for patients who were alive at their last follow-up session. Differences among groups were considered significant at $p<0.05$. These analyses were conducted using SPSS V.26.0 (IBM).

**Patient and public involvement**

Patients were not directly involved in the design or evaluation of the current report.

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Table 1  Clinicopathological characteristics in 34 patients with ductal adenocarcinoma and acinar adenocarcinoma

|                          | Total | Ductal adenocarcinoma | Acinar adenocarcinoma | P value |
|--------------------------|-------|-----------------------|-----------------------|---------|
| Patients (n)             | 34    | 14                    | 20                    | –       |
| Median age (range)       | 72 (48–85) | 76 (48–85)            | 71 (60–76)            | NS      |
| ≤70                      | 15    | 5                     | 10                    | NS      |
| >70                      | 19    | 9                     | 10                    |         |
| Median iPSA (range)      | 8.0 (1.1–129.0) | 5.6 (1.1–129.0)      | 9.0 (4.1–22.2)        | NS      |
| ≤4.0                     | 6     | 6                     | 0                     | 0.002   |
| >4.0                     | 28    | 8                     | 20                    |         |
| cT stage                 |       |                       |                       |         |
| <T2                      | 15    | 3                     | 12                    | 0.038   |
| >T3                      | 19    | 11                    | 8                     |         |
| Ductal component         |       |                       |                       |         |
| Pure/mixed type          | 9/5   |                       |                       |         |
| Metastasis at diagnosis  |       |                       |                       |         |
| No                       | 27    | 7                     | 20                    | 0.001   |
| Yes                      | 7     | 7                     | 0                     |         |
| MVD (mean±SD)            | 74.4±48.7 | 65.3±21.9            | 80.8±60.7            | NS      |
| <60                      | 13    | 5                     | 8                     | NS      |
| ≥60                      | 21    | 9                     | 12                    |         |
| VASH1 density (mean±SD)  | 28.0±24.5 | 45.1±18.5            | 16.1±21.0            | <0.001  |
| <30                      | 17    | 1                     | 16                    | <0.001  |
| ≥30                      | 17    | 13                    | 4                     |         |
| VASH1 density/MVD ratio (mean±SD) | 0.436±0.370 | 0.724±0.265        | 0.236±0.294          | <0.001  |

cT stage, clinical T stage; iPSA, initial prostate-specific antigen; MVD, microvessel density; NS, not significant; VASH1, vasohibin-1.
RESULTS

Patient characteristics and evaluation of VASH1 density and MVD

The clinicopathological parameters and VASH1 density or MVD of 14 ductal adenocarcinoma and 20 acinar adenocarcinoma cases are shown in table 1. The median patient age was 72 years (range 48–85 years) and the median pretreatment prostate-specific antigen (PSA) level was 8.0 ng/mL. The pretreatment PSA level in patients with ductal adenocarcinoma (median 5.6 ng/mL, range 1.1–129.0 ng/mL) was lower than that of patients with acinar adenocarcinoma (median 9.0 ng/mL, range 4.1–22.2 ng/mL), and the levels varied widely. According to the pathology reports, nine patients (64.3%) with ductal adenocarcinoma had a pure type, while five patients (35.7%) had mixed ductal and acinar adenocarcinoma type. No statistically significant differences in clinical characteristics were noted between patients with pure and mixed types. Table 1 shows that the tumour of patients with ductal adenocarcinoma had a more aggressive behaviour than that of those with acinar adenocarcinoma. Nine patients (64.3%) with ductal adenocarcinoma were diagnosed at an advanced clinical stage with local and distant metastases, and five patients (35.7%) died of cancer during a median follow-up of 56.0 months.

To elucidate the biological significance of VASH1 in ductal adenocarcinoma, we examined the immunohistochemical expression of VASH1 and CD34 in the tumour tissue (figure 1). High VASH1 expression was observed in vascular ECs around the papillary architecture of tumours in ductal adenocarcinoma (figure 1A), while it was only partially observed in acinar adenocarcinoma lesions with a Gleason score of 4+4 (figure 1B). The mean±SD VASH1 densities of ductal and acinar adenocarcinoma samples were 45.1±18.5 vs 16.1±21.0 (p<0.001), respectively (table 1). On the other hand, the expression of CD34 in vascular ECs was at the same level (figure 1C,D), and the mean MVD of ductal and acinar adenocarcinoma samples were 65.3±21.9 vs 80.8±60.7 (p=0.666), respectively.

We previously evaluated VASH1 expression levels in patients with acinar adenocarcinoma in two studies: one with 167 patients using RP specimens and the other with 104 patients using prostate needle biopsy specimens. The mean VASH1 densities were 9.9±7.3 and 9.7±9.5, respectively. Compared with previous data, the VASH1 density of acinar adenocarcinoma in the present study cohort was relatively higher because of the high Gleason score. Nevertheless, the VASH1 density of ductal adenocarcinoma samples was higher than that of acinar adenocarcinoma samples in this study.

Prognostic significance of VASH1 expression

Univariate analyses were performed to determine indicators of CSS. Figure 2 shows that ductal adenocarcinoma and high VASH1 expression were significant predictors of CSS (figure 2A,B), while high MVD was not a significant predictor (figure 2C). The 5-year CSS rates for high and low VASH1 expression were 70.0% and 100.0%, respectively (p=0.006). In addition, a low PSA level (PSA <4.0 ng/mL) at the initial diagnosis (online supplemental figure 1) was a significant predictor of CSS. Multivariate analysis revealed no independent predictors of CSS.

Next, we focused on the differences in the VASH1 density/MVD ratio. The mean VASH1 density/MVD ratios for ductal and acinar adenocarcinoma samples were 0.72±0.265 and 0.23±0.294 (p<0.001), respectively (table 1). Then, we re-evaluated the univariate analysis of

![Figure 1](https://example.com/figure1.png)

**Figure 1** Immunostaining for VASH1 (vasohibin-1) and CD34 in ductal adenocarcinoma (A and C, respectively) and in acinar adenocarcinoma with the Gleason score of 4+4 (B and D, respectively). Scale bar=0.1 mm.

![Figure 2](https://example.com/figure2.png)

**Figure 2** Kaplan-Meier curves of cancer-specific survival according to (A) histology, (B) microvessel density (MVD) and (C) vasohibin-1 (VASH1) density in all patients (n=34).
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VASH1 density separately for patients with low or high MVD values (figure 3). Six (46.2%) and 11 (55.0%) patients showed high VASH1 density in the low MVD group and high MVD group, respectively. The 5-year CSS rates for high and low VASH1 densities were 83.3% and 100.0% in the low MVD group (p=0.317) (figure 3A) and 63.5% and 100.0% in the high MVD group (p=0.012) (figure 3B). Together, these results demonstrated that MVD in ductal adenocarcinoma is not a predictor of CSS, but it could act as a complementary subdivision of the VASH1 density.

DISCUSSION

To the best of our knowledge, this study was the first to evaluate the level of neovascularisation around the tumour in patients with ductal adenocarcinoma using a novel angiogenesis regulator termed VASH1 and compared the oncological outcome between ductal and acinar adenocarcinoma. While the biological features along with the clinical and prognostic significance of ductal adenocarcinoma remain unclear, some studies have reported that ductal adenocarcinoma is usually detected at an advanced clinical stage, resulting in a higher rate of prostate cancer-specific mortality than acinar adenocarcinoma in general but similar to acinar adenocarcinoma with a Gleason score of 4+4.3 23 24 Ranasinghe et al reported that 112 (25.7%) patients had metastasis at diagnosis among the largest cohort of 435 patients who were newly diagnosed with ductal adenocarcinoma and concluded that the rate of de novo metastatic ductal adenocarcinoma was much higher than that previously reported by other studies (12.0%-13.6%).25

In the era of PSA screening, early diagnosis and early radical treatment became increasingly possible, even in patients with high Gleason score or high-risk acinar adenocarcinoma in Japan. However, PSA screening tests are often ineffective in patients with ductal adenocarcinoma because they often present with only macroscopic haematuria and/or urinary obstruction without elevated serum PSA levels. This difficulty in early diagnosis may affect prognosis. In fact, 6 patients (42.9%) with ductal adenocarcinoma showed a low PSA level (<4.0 ng/mL), 11 patients (78.6%) had a cT3 stage or worse, and 7 patients (50.0%) showed distant metastasis at the time of diagnosis in this study. Moreover, the oncological outcome of ductal adenocarcinoma was predominantly worse than that of acinar adenocarcinoma with a Gleason score of 4+4.

We hypothesised that increased angiogenesis is one of the main factors causing malignant progression and poor prognosis of ductal adenocarcinoma. In our previous study, we reported that since MVD corresponds to the number of formed vessels and included vessels without any potential for neovascularisation in acinar adenocarcinoma, VASH1 was expressed almost selectively in tumour-associated ECs.13 15 We also revealed that high VASH1 expression was associated with tumour malignancy as evaluated by the Gleason score and pT staging, and it was an independent predictor of tumour progression in acinar adenocarcinoma.13 22 No previous studies quantitatively analysed the immunohistochemical evaluation of neovascularisation in ductal adenocarcinoma. Therefore, whether VASH1 expression and/or MVD offer prognostic information may add value to the diagnosis of ductal adenocarcinoma.

This contribution has, however, been studied in other lesions. For instance, in hepatocellular carcinoma (HCC), the prognostic role of MVD is controversial. Several studies have suggested that increased MVD could serve as an independent prognostic factor of decreased overall survival.26 27 On the other hand, several studies have demonstrated significantly lower MVD in patients with advanced-stage HCC.28 29 Accordingly, Murakami et al reported that VASH1-positive vessels had higher angiogenic activity than the CD34-positive ones in HCC. In addition, they found that the VASH1/CD34 ratio

Figure 3 Kaplan-Meier curves of cancer-specific survival among (A) low microvessel density (MVD) groups (n=21) or (B) high MVD groups (n=21) according to the vasohibin-1 (VASH1) density.

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increased according to tumour progression and was associated with poor prognosis.\(^3\)

In our VASH1 and CD34 staining, the ductal adenocarcinoma specimens showed high VASH1 expression irrespective of the serum PSA level, and this expression was predominantly higher than that of acinar adenocarcinoma specimens, while MVD showed no differences between the two lesions. In addition, the VASH1 density/MVD ratio was higher in ductal adenocarcinoma than in acinar adenocarcinoma, and patients presenting with high MVD, and high VASH1 density likely have a worse prognosis. These results indicated that ductal adenocarcinoma showed more activated ECs with aggressive neovascularisation than acinar adenocarcinoma with a Gleason score of 4+4, which had a pronounced growth and poor prognosis among all prostate cancers. Additionally, when we examined VASH1 expression in normal tissues adjacent to the cancer in several ductal adenocarcinoma patients, we found that VASH1 expression tended to be higher in ductal adenocarcinoma than in acinar adenocarcinoma. This result may reflect the cancer microenvironment of increase angiogenesis throughout the prostate in ductal adenocarcinoma. Since aggressive neovascularisation in ductal adenocarcinoma does not depend on the PSA level, the invasion or progression pattern of ductal adenocarcinoma may be different from that of acinar adenocarcinoma; in any case, the expression of VASH1 could serve as a novel biomarker for the aggressive nature of ductal adenocarcinoma.

In contrast, in patients with oesophageal squamous cell carcinoma, which is a fast-growing cancer with poor prognosis, Ninomiya et al reported that high VASH1 and VASH2 expression are predictors of poor prognosis. Subsequently, Yamamoto et al demonstrated that the plasma concentrations of VASH1 and VASH2 are associated with poor oncological outcomes in patients with oesophageal squamous cell carcinoma.\(^19\)\(^,\)\(^21\) These new approaches and viewpoints can be considered for application to other cancers. In recent years, targeted drug therapy based on genomic sequencing or liquid biopsy has been widely used for several cancers. Liquid biopsy is the real-time analysis of tumour cells or tumour cell products released into the blood or other body fluids by primary or metastatic tumour lesions.\(^32\) These techniques can enable the development of new methods for the early detection of primary cancer, determination of therapeutic targets and discovery of progression mechanisms. Currently, although there are no established guidelines for ductal adenocarcinoma and its genomic or immunohistochemical profile remains unclear, large-scale immunohistochemical and/or genetic analyses as well as aggressive neovascularisation and its biomarker, VASH1, are expected to help determine ductal adenocarcinoma aggressiveness.

This study has several limitations. First, it was performed in a retrospective manner with a limited number of patients due to the rarity of ductal adenocarcinoma. The reason for the small number of patients with acinar adenocarcinoma with a Gleason score of 4+4 is that the VASH1 density is much lower than that of ductal adenocarcinoma, as shown by our results, so we assumed that increasing the number of patients would not change the results. Second, the patient population was heterogeneous, with a short follow-up period. A larger multicentre study is, thus, warranted to confirm our findings. Finally, there were only a few treatment options or accurate imaging studies performed, such as multislice CT scan or multiparametric MRI, since the retrospective data collection involved a period over 10 years. This factor may adversely affect the analysis of the prognostic factors.

In conclusion, ductal adenocarcinoma specimens showed predominantly stronger VASH1 staining than those of acinar adenocarcinoma, and higher VASH1 expression was a significant predictor of CSS in patients with ductal adenocarcinoma. Overall, these results indicate that ductal adenocarcinoma possesses aggressive characteristics and that VASH1 could serve as a novel prognostic biomarker.

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