Bioactive compounds from *Holothuria atra* of Indian ocean

Devaraj Isaac Dhinakaran¹* and Aaron Premnath Lipton²

**Abstract**

The sea cucumber (*Holothuria atra*) extracts have been evaluated for the presence of bioactive compounds and various biological activities. The methanol extracts showed anti proliferative activities against the Hela and MCF-7 cell lines. Similarly the inhibitory effects of Herpes simplex virus 1 and 2 cells were detected using the plaque reduction assay. The extracts of *H. atra* were purified using the silica gel column chromatography. The active fractions collected were observed for antimicrobial activity. The GC-MS analysis showed the availability of 59 compounds. The active bioactive compounds found in the *H. atra* were analyzed and their structure was identified using the $^1$HNMR and $^{13}$C NMR experiments.

**Keywords:** *Holothuria atra*; Bioactive compounds; Cell lines Herpes simplex virus MCF-7

**Introduction**

The marine holothurians are spiny skinned invertebrates, which form important commercial group among the echi- noderms. In India, about 200 species of holothurians are known of which 75 species are from shallow waters within 20 m depths (Cuvillier 2002). (James 1994) suggests that great potential exists for the extraction of valuable bioactive compounds from the sea cucumbers in the Indian coast. A new immunomodulatory lead Cumaside a complex of monosulfated triterpene glycosides from the sea cucumber *Cucumaria japonica* possesses cytotoxic activity against Ehrlich carcinoma cells (Aminin et al. 2004). Sphingoid base composition of cerebrosides from sea cucumber *Stichopus variegatus* exhibited cytotoxicity against human colon cancer cell and induced apoptosis. They are major constituents of c-17, c-19 alkyl chain and 1-3 double bonds (Sugwara et al. 2006). Triterpene glycosides are the predominant secondary metabolites of the sea cucumber *Hemoeidea spectabilis* which exhibited wide spectra of biological activities, including antifungal, cytotoxic, hemolytic, cytostatic and immunomodulatory functions Chludil et al. (2002). A new lanostane-type triterpene glycoside, impatiensiens A and bivittoside D were isolated from the sea cucumber *Holothuria impatients* Sun et al. (2007). The potential angiogenesis inhibitors, a novel sulfated saponin philinnopside A, isolated from the sea cucumber *Pentacta quandrangulari*, possessed dual antiangiogenic and antitumour effects (Tong et al. 2005).

Fuscocineroside C bioactive compound obtained from sea cucumber *Holothuria fuscocinerea* a triterpene glycoside showed cytotoxic nature against human cancer cells Zhang et al. (2006). Hillaside C a triterpene derived from sea cucumber *Holothuria hilla* inhibited the growth of human leukemia, breast and colon cancer cells in vitro in a dose and time-dependent manner by a mechanism that required induction of apoptosis and the concomitant reduction of the apoptosis-suppressing protein Bcl-effect Wu et al. (2006). Intercedenside D–I isolated a cytotoxic triterpene glycoside from the sea cucumber *Mensamaria intercedens* a marine natural product inhibited proliferation of several human cancer cell lines Zou et al. (2005). Steroid glycosides are a class of widespread natural products having marine origins. Spirostan and furostan steroid saponins, pregnane glycosides have a potential to be used as cancer therapies. Structurally, these glycosides exhibit a moderate cytotoxicity against human leukemia cell lines (Prassas and Diamandis 2008). Linhardt et al. (1990) found that low molecular weight sulphated polysaccharides are noted from sea cucumbers with efficient anticoagulant activities and several pharmacological properties. The chondroitin and glucosamine components
of holothuria were reported to be important cartilage building blocks and other bioactivities including anti-inflammatory and anti tumor activity properties (Heredia and Ubeda 1998). The extract LPS obtained from *Stichopus japonicus* induced inflammatory response via blocks the MAPK signaling pathway in murine macrophages, showed *in vitro* with anti-inflammatory potential Himayaa et al. (2010). The sea cucumber *Telenata ananas* derived bioactive compounds were reported to act as the chemokine receptor subtype-5 (CCR5) with possible anti-HIV activity Hegde et al. (2002). Potential use of sea cucumber *S. liouvillei* isolated compound chondroitin sulfate (the polysaccharides) are reported to exhibit antiviral activity Hegde et al. (2002). Considering this as an evidence in the present study, attempts were made to find out the bioactive compounds from marine invertebrates such as the Holothuria.

**Materials and methods**

**Sample collection and extract preparation**

*Holothuria atra* specimens with a size range of 10 to 30 cm in length and 30 to 180 g weight were collected from fishing nets operated off Kanyakumari (8° 03′ and 8° 35′ of the north latitudes and 77° 15′ and 77° 36′ of the east longitudes) in the Indian Ocean. Immediately upon collection, they were dissected to remove the internal organs and packed using ice prior and kept at -80°C or extraction. The skin portion was peeled off and stored in methanol in separate containers. The biologically active compounds were extracted as a function of their polarity using water and organic solvents. About 200 g of frozen samples were homogenized with deionized water and methanol. The mixture was continuously stirred in the dark at 4°C for 24 h. Then it was centrifuged at 5000 rpm for 15 min. The supernatant was collected and filtrated. The collected organic extracts were freeze-dried and kept at -80°C, while the insoluble solid materials were re-extracted with methanol (100%) (Chen 2003).

**MTT assay using Hela cell lines and MCF-7 cancer cell lines**

The cells were preincubated at a concentration of 1 × 10^6 cells/ml in culture medium for 3 h at 37°C and 6.5% CO_2_. Then, the cells were seeded at a concentration of 10^6 cells/ml in culture medium for 3 h at 37°C and 6.5% CO_2. Then, the cells were preincubated at a concentration of 1 × 10^4 cells/well in 100 μl culture medium and at various concentrations of extracts (dissolved in 2% DMSO dimethylsulphoxide solution) into microplates (tissue culture grade, 96 wells, flat bottom) and incubated for 24h at 37°C and 6.5% CO_2. Then, 10 μl MTT labelling mixture was added and incubated for 4 h at 37°C and 6.5% CO_2. Each experiment was conducted as triplicates sets. Then 100 μl of solubilization solution was added into each well and incubated for overnight. The spectrophotometric absorbance of the samples was measured using a microplate (ELISA) reader. The wavelength to measure absorbance of the formazan product in 570 nm according to the filters available for the ELISA reader was used. The reference wavelength was more than 650 nm. IC_{50} values were calculated Percentage inhibition of novel compounds against all cell lines was calculated using the following formula:

\[
\text{(At}-\text{Ab) } \% \text{ cell survival} = \frac{\text{Ab} - \text{Ac}}{\text{Ab}} \times 100
\]

**Trypan blue dye exclusion test**

Being an essential dye, Trypan blue was used in estimating the number of viable cells present in a population. The culture sample was mixed to resuspend cells. 20 μl of cell culture sample was taken and filled into sterile microfuge tube. To this 20 μl of 0.4% Trypan blue solution was added and mixed well by gently aspirating and dispensing the solution with the help of micropipette. The coverslip was fixed on the centre top of the hemocytometer. To the 10 μl mixture of the cell culture the Trypan Blue mixture taken from the microfuge tube was added and kept in the hemocytometer assembly on microscope stage using 100 X magnification. The number of live and dead cells were recorded. 

\[
\% \text{viability} = \frac{(\text{live cell count}/\text{total cell count})}{\times 100}
\]

**Plaque reduction assay**

Vero monolayer cells grown in 24 well tissue culture plates were infected with HSV -1 and HSV-2. Virus dilutions were made from 10^2 to 10^7 using 0.1 ml of viral suspension. Virus adsorption was carried out for 1h at 37°C in the presence of test extract. Virus dilutions were prepared in Eagles minimum essential medium. Prior to incubation, an overlay medium comprising of 0.8% carboxy methyl cellulose with 2% FBS was added. It was done to avoid formation of secondary plaques. Infected cell cultures were incubated at 37°C at 5.0% CO_2 incubator for 2 to 3 days. The infected cells were stained and observed for plaque reduction. The infectivity titers were expressed as the number of plaque forming units per ml (pfu ml^{-1}). After incubation, cultures were stained with 1% (w/v) crystal violet solution. The plaques were counted by visual examination and the percentage of plaque inhibition was calculated. The Pfu = Plaque number × reciprocal of dilution × reciprocal of volume in ml.

The antiviral activity was defined as the percentage of plaque inhibition as follows:

\[
\% \text{ Plaque inhibition} = \frac{1 - (\text{Number of plaque in test})}{\text{Number of plaque in control}} \times 100
\]
Column chromatography
Silica (230-400 mesh) gel slurry was prepared using methanol. The column was packed with silica gel. After washing the column with same solvents, the sample (3 to 5 ml) was poured and then eluted with methanol: water on different percentages (10 to 100% methanol). The active fractions were collected and used for NMR analysis.

Antibacterial activity
Disc diffusion method was employed to test the active fractions of *H. atra* obtained from the column chromatography Bauer et al. (1966). Antibacterial activity was determined using Muller Hinton agar (Hi Media). The bacterial cultures were obtained from the Microbial type culture collection and gene bank (MTCC), Institute for Microbial Technology, Chandigarh, India. They were *Staphylococcus aureus* MTCC 737, *E.coli* MTCC 443, *Klebsiella pneumonia* MTCC 109, *Listeria monocytogenes* MTCC 1143, and *Serratia liquefaciens* MTCC 3039. The plates were aseptically streaked with the test microorganism using a sterile swab and allowed to dry for a few minutes. Sterilized filter paper discs (Whatman no.1; 6 mm diameter) were used. The fractions were collected from 10 to 100% levels of methanolic extracts and were evaluated at 100 μl concentration. The plates were then incubated for 24 h at 37°C. Controls were blank discs impregnated with solvent. The diameter of the inhibition zone formed around the disc was measured.

GC-MS analysis
The methanol extract of the sea cucumber *H. atra* was analyzed by GC-MS (Make: Fisons GC8000 series and MS: md800). The GC column dimension was: 30 mm, 0.25 mm, 0.5 mm AB-35MS fused silica capillary column. The GC conditions were as follows: injector temperature 250°C column temp isothermal at 100°C then programmed to rise up to 250°C at 6°C/min and held at this temperature for 10 minutes. The ion source temperature was 200°C and the interface temperature was 250°C. Helium gas was engaged for carrier gas at the rate of 1ml/min. Spectra was obtained in the EI mode with 70eV ionization energy. The compounds were identified by comparison with the standards. If not available, the mass spectra was matched with inbuilt library like wileys, NIST (Stonik et al. 1998).

NMR analysis
The active fractions obtained from column chromatography were analysed for Nuclear magnetic resonance spectroscopy (NMR) analysis. Optical rotations were measured on a Perkin- Elmer Model 341 LC polarimeter. ¹H NMR and ¹³C NMR experiments were performed on Bruker Unity 400 and 600 MHz spectrometers. NMR spectra were referenced to the CD3OD solvent signals at δ 3.30 (1H) and 49.00 (13C), respectively. The spectra were obtained using the standard Bruker software. The samples were dissolved in different solvents (i.e. DMSO-d₆, CDC13, and CD3OD), the choice of which was dependent on the solubility of the samples. The observed chemical shift (δ) values were given in ppm and the coupling constants (J) in Hz.

Results and discussion
The results in Table 1 show that the amount of sea cucumber *H. atra* extract required to inhibit 50% of the antitumor activity against the Hela cell lines in 96 well plates could be determined. Antitumor activity was measured by using the IC₅₀ values. The anti proliferative effect (IC₅₀) value exhibited by the *Holothuria atra* was 468.0 against the cervical cancer cell line (Hela). The cell inhibition was determined from the extract concentration ranged from 0.078 mg/ml to 10 mg/ml. The absorbance values were measured at 570 nm. Percentage of growth inhibition was

### Table 1 Cytotoxicity analysis of *Holothuria atra* extracts against Hela cell lines using MTT assay

| Concentration (mg/ml) | Absorbance (570 nm) | % inhibition | IC₅₀ |
|-----------------------|---------------------|--------------|------|
| 0.078                 | 0.125               | 40.56        |      |
| 0.156                 | 0.103               | 52.12        |      |
| 0.3125                | 0.10                | 60.42        |      |
| 0.625                 | 0.093               | 68.30        |      |
| 1.25                  | 0.085               | 75.34        | 468.0|
| 2.5                   | 0.070               | 83.26        |      |
| 5                     | 0.069               | 88.32        |      |
| 10                    | 0.052               | 90.56        |      |
| Cell control          | 0.45                | 100          |      |

### Table 2 Cytotoxicity analysis of *Holothuria atra* extracts against MCF-7 cell lines using MTT assay

| Concentration (mg/ml) | Absorbance (570 nm) | % inhibition | IC₅₀ |
|-----------------------|---------------------|--------------|------|
| 0.078                 | 0.120               | 30.22        |      |
| 0.156                 | 0.102               | 35.64        |      |
| 0.3125                | 0.10                | 48.72        |      |
| 0.625                 | 0.088               | 55.60        |      |
| 1.25                  | 0.085               | 60.62        | 3520 |
| 2.5                   | 0.075               | 65.20        |      |
| 5                     | 0.062               | 70.22        |      |
| 10                    | 0.052               | 75.60        |      |
| Cell control          | 0.45                | 100          |      |

### Table 3 Percentage cell inhibition and characterization of cell line exposed to *Holothuria atra* using tryphan blue

| Cell line | % cell inhibition | Dead cell count | Total cell count | pH |
|-----------|-------------------|-----------------|------------------|----|
| Hela      | 81.81             | 1.80 × 10⁵      | 2.20 × 10⁵       | 6.9|
| MCF-7     | 72.72             | 1.76 × 10⁵      | 2.42 × 10⁵       | 7.2|
identified at different concentration of the extracts. The gradual decrease in absorbance values showed increase in inhibition effect of the extracts against the Hela cell lines. The findings suggest that *H. atra* showed cell inhibition to the tune of 90% to the maximum in Hela cells and 75% of cell inhibition in MCF-7 cells. Five cerebrosides, PA-0-1, PA-0-5, PA-2-5, PA-2-6 and CE-2c were reported from the Japanese sea cucumber *Pentacta australis* Higuchi et al. (1994). A ganglioside molecular species SJG-1, isolated from the sea cucumber *Stichopus japonicus*. SJG-1 possessed a sialic acid, nonhydroxy fatty acids and phytosphingosine-type long chain bases as major ceramide components. SJG-1 exhibited neurotogenic activity towards the rat pheochromocytoma cell line PC12 cells in the presence of nerve growth factor Kaneko et al. (1999). Additionally, a cerebroside isolated from the sea cucumber *Stichopus japonicas* showed effective antitumor activity (Hayashi et al. 1990). The data presented in the Table 2 suggested the antitumor activity of the methanol extracts of *H. atra* against the breast cancer cell lines MCF-7 in 96 well microtitre plates. The susceptibility of cells to the extract exposure was characterized by IC\textsubscript{50} values. Results indicated that the anti proliferative effect increased with the increase in concentration of the extracts. The IC\textsubscript{50} value for the sea cucumber *Holothuria atra* was 352.0 A decrease in number of viable cells with the increase in concentration of the extracts was noted. From Table 3 the results of cell counting and viability of cells using tryphan blue staining were indicators for the influence of extracts. The percentage cell inhibition of Hela was 81.81% and MCF-7 was 72%. The cell proliferation and inhibition measurement of the sea cucumber *Holothuria atra* showed that it can be developed as an antitumor agent. The cytotoxic effects of extracts against the Hela and MCF-7 cell lines are observed through the inverted microscope (Figure 1). Promising *in vitro* cytotoxic compounds such as the Calcigeroside B, C1 and C2 identified from the holothurians included triterpene glycosides. They showed antiproliferative action against the human and murine tumour cell lines (Alejandro and Gustafson 2003). In the present study, the tumor growth was inhibited by *Holothuria atra* extract and dosage was an important criteria which influenced the efficacy of the cell death in Hela, and MCF-7 cell lines. These suggest the possibility of apoptotic cell death through the activation of Bax a proapoptotic protein.

**Figure 1** Cytotoxic effects of *H. atra* extracts on A) Hela and B) MCF-7 cell lines.

**Figure 2** *In vitro* antiviral activity of *Holothuria atra* extracts against A) HSV-1 and B) HSV-2 using plaque reduction assay.
Angiogenesis inhibitors and aromatase inhibitors present in sea cucumbers play a major role in reducing the growth of breast cancer and prostate cancers, especially the solid tumors. Research results showed that angiogenesis inhibitors effectively block the growth of tumors by cutting off their nutrient and blood supply. The mechanism by which they block tumor growth is driven by the inhibition of receptor tyrosine kinases (RTKs) that are over expressed by cancer cells (Chi 2006). Using MTT assay it could be inferred that H. atra extracts can block the growth of breast cancer cells (MCF-7) by inducing apoptosis. In H. atra extract there is a possibility of blocking the receptors such as the tyrosine kinases (RTKs) in cancer cells of Hela and MCF-7. The methanol extracts of Holothuria atra showed maximum inhibition of antitumor cells and it was observed by cell inhibition using trypan blue.

The concentration of extracts for H. atra was from 10 μg/ml to 70 μg/ml, respectively. The tested viruses were affected with the increase in concentration of extracts. The H. atra exhibited significant antiviral activity, and suggested the potential role of extracts. The effect of inhibition in plaque formation was evaluated based on the 101 to 107 dilutions of HSV-1 and HSV-2. In H. atra the highest plaque inhibition rate was as at 75% with \(2.4 \times 10^3\) pfu ml\(^{-1}\). Less inhibition rate was observed at 33% for \(6.0 \times 10^9\) pfu ml\(^{-1}\). The results of (Figure 2)

### Table 4 Inhibitory action of Holothuria atra extracts against HSV strains (\%)

| Dilution of virus | Concentration of H. atra extracts | Plaque forming units pfu/ml | % Plaque inhibition [HSV-1 and HSV-2] |
|-------------------|-----------------------------------|-----------------------------|---------------------------------------|
| Control           | Nil                               | 9.2 × 10\(^1\)              | 8.7 × 10\(^1\)                        | -                                 |
| \(10^{-1}\)       | 10 μg                             | 6.0 × 10\(^9\)              | 6.4 × 10\(^3\)                        | 33                                |
| \(10^{-2}\)       | 20 μg                             | 5.6 × 10\(^8\)              | 5.6 × 10\(^4\)                        | 40                                |
| \(10^{-3}\)       | 30 μg                             | 5.2 × 10\(^7\)              | 4.7 × 10\(^5\)                        | 44                                |
| \(10^{-4}\)       | 40 μg                             | 4.4 × 10\(^6\)              | 4.3 × 10\(^6\)                        | 53                                |
| \(10^{-5}\)       | 50 μg                             | 3.9 × 10\(^5\)              | 2.3 × 10\(^9\)                        | 58                                |
| \(10^{-6}\)       | 60 μg                             | 3.3 × 10\(^4\)              | 2.8 × 10\(^8\)                        | 65                                |

Figure 3 Anti bacterial activity of the column purified fractions of sea cucumber, Holothuria atra against A) Serratia liquefaciens B) Escherichia coli C) Klebsiella pneumoniae D) Staphylococcus aureus.
suggest the effects of *H. atra* extracts on the inhibition of virus replication after attachment of HSV-1 and 2 on Vero cells. In *H. atra*, the plaque inhibition obtained was high at 74% with $2.3 \times 10^9$ pfu ml$^{-1}$ whereas less effect was seen in *H. atra* was 27% with $6.4 \times 10^3$ pfu ml$^{-1}$ units (Table 4). Saponins the secondary metabolites which are triterpene glycosides present in sea cucumbers like *H. forskali* are reported to have antiviral property by *in vitro* and *in vivo* methods (Kerr and Chen 1995). It was observed that the *H. atra* extracts exhibited antiviral activity on plaque reduction assay in which maximum effect was seen against the HSV-1 to the tune of 74%. Thus

![Figure 4 GC-MS Analysis of the methanolic extracts of the sea cucumber Holothuria atra.](image)

Figure 4 GC-MS Analysis of the methanolic extracts of the sea cucumber *Holothuria atra*. 
Holothuria atra extracts have the ability to arrest the multiplication of virus and suppress its growth by influencing the growth factors. This could have resulted in appearance plaques in the plaque reduction assay. Bioactive peptides and hemolytic lectins have been reported from sea cucumber as a source of antiviral activity. Among Holothuroidea genera, Cucumaria echinata and C. frondosa contained lectin and peptide, respectively. They have been found in the body wall mucus Hisamatsu et al. (2008). Fucoidans, polysaccharides containing substantial percentages of L-fucose and sulfate ester groups are generally considered as constituents of sea cucumbers. Fucoidan can inhibit the development of cytopathic effect (CPE).

| No | Name of the compound | Molecular formula | M.W |
|----|----------------------|------------------|-----|
| 1 | Ethane 11dichloro(CAS)| C2H4Cl2          | 98  |
| 2 | Carbamic acid/hydroxyl, ethyl ester | C3H7O3N      | 105 |
| 3 | Ethane sulphonyl chloride,2 chloro | C2H4O2Cl2    | 162 |
| 4 | Methyl 40-methyl-beta-D-xylopyranoside | C7H14O5    | 178 |
| 5 | Propylhexedrine       | C10H21N         | 155 |
| 6 | 2-methyl-1,3-dithiacyclpentane | C4H8S2      | 120 |
| 7 | Ethane dithioic acid  | C2H4S2          | 92  |
| 8 | 4 ketopinolic         | C7H10O5         | 174 |
| 9 | 2-ethylomethyl-1,3-dithiacyclpentane | C8H12O2N    | 162 |
| 10| Carbomic chloride,methoxymethyl | C3H6O2NCl   | 123 |
| 11| Xylene,ethyldimethyl  | C4H12SL         | 88  |
| 12| D-pseudo ephedrine or ephedrine or pholedrine | C10H15ON    | 165 |
| 13| Ethanol,1-methoxy-benzoate | C10H21O3    | 180 |
| 14| Methanamine hippurate artefact | H4O         | 0   |
| 15| 2,6,10,14 tetra methylpentadecane-2-ol | C19H34O     | 284 |
| 16| 2,8-heptadecatrien-1-ol | C17H30O      | 250 |
| 17| Methyl 19-hydroxyicosan-5(z),8(z),11(z),14(z)-tetra-tetra. | C21H34O3   | 334 |
| 18| 3,4epoxy-6,9-octadecadiene | C18H32O     | 264 |
| 19| (14z,17z)-3,20-dibromo-21-ethyl-2,6-epoxy-1-oxacyclo | C22H34O2   | 496 |
| 20| (1)-ema-1,3,11(13)-trien-12-ol or Beta –costol | C15H24O     | 220 |
| 21| Phenylpropanolamine   | C9H13ON        | 187 |
| 22| 3,3-dibromotricyclo(2,4)undecane | C11H16B12   | 306 |
| 23| Transcanyophyllene or Alpha far neose or epsilon cadinene. | C15H24     | 204 |
| 24| 2,3-2poxy-5,8-hectadecadien-1-ol | C9H12O2     | 266 |
| 25| (++)-3-methylidenetricyclo(2,9)undecan-8-one | C12H16O    | 176 |
| 26| 7-methylbicyclo oct-7-one | C9H14       | 122 |
| 27| Methylareachidonate 5,8,11,14 eicosatetraenoic | C21H34O2   | 318 |
| 28| 2 –naphthalenemethanol-decahydro-5-methylene-8 | C14H22O2   | 206 |
| 29| Tricyclo decanedimethanol | C12H20O2    | 196 |
| 30| 2-3-29Methoxy carbonyl) ethyl bicyclo 1-1-1pent-1 | C13H18O4   | 238 |
| 31| Transcanyophyllene.   | C15H24        | 204 |
| 32| 2xo,2endo3endo,3exo-bis (epoxymethane) bicyclo | C9H12O2    | 152 |
| 33| 2-pyrrolidinarbocyclic acid,5-methyl, phenylne | C13H17O2N   | 219 |
| 34| Cyclopropane,1-ethyl-2-hexeny1,1apha,1e Bet | C11H18      | 150 |
Figure 5 NMR analysis of active fractions obtained from Column chromatography (A) $^1$H NMR Spectrum of Holothuria atra (B) $^1$H NMR Spectrum of Holothuria atra (C) $^{13}$C NMR Spectrum of Holothuria atra.
and protect cultural cells from infection caused by viruses (Hemmingson et al. 2006). This can induce the antiviral effects against the Herpes simplex viruses.

Figure 3 shows the antibacterial activity for the active fractions obtained from column chromatography of H. atra against various Gram positive and Gram negative bacteria viz., Klebsiella pneumonia, Serratia liquefaciens, Staphylococcus aureus, Listeria monocytogenes, and Escherichia coli. The fractions collected from 50 to 100% showed maximum effect whereas the fractions from 10 to 40% did not show any activity. In H. atra the E. coli had less effect at 50 and 60% of fractions which showed the zone diameter of 2 and 3mm range. Figure 4 shows the GC-MS graphical representation of the extracts of sea cucumber. The interpretation on mass spectrum GC-MS was carried out and the spectra of the unknown component were compared to the spectrum of the known components stored in the NIST library. The name, molecular weight of the components was ascertained. A total of 59 natural compounds were identified from the extracts of sea cucumber (H. atra). The active principles with their retention time (RT), molecular formula, molecular weight (MW) were ascertained (Table 5).

Fucoidan of sea cucumber Laminaria japonica has anti RNA and DNA virus functions. The antiviral effects of fucoidan on infection was against poliovirus III, adeno-virus III, ECHO6 virus, coxsackie B3 virus and coxsackie A16. Fucoidan inhibited the development of cytopathic effect (CPE) and protected the cultural cells from infection caused by the viruses Li et al. (1995). Sulfated polysaccharides from sea cucumbers such as the Cucumaria japonica, Holothuria impatiens are reported to exhibit antiviral activity. Based on this fact, Japanese scientists have patented their scientific findings regarding the potential use of sea cucumber chondroitin sulfate to inhibit human immunodeficiency virus (HIV) infection Beutler et al. (1993). Triterpene glycosides, namely holothuriniosides A, B, C and D as well as desholothurin A from sea cucumber (Holothuria forskali), have considerable antitumour activity against P388 cell lines. The saponins isolated from the aqueous and methanolic extract of sea cucumber (Holothuria forskali) have showed considerable antiviral activities Mulloy et al. (2000). Considering these as well as the results of present investigations, the methanolic extracts of Holothuria atra could form effective antitumour and antiviral agents. Previous work showed that sulphated polysaccharides such as glycosaminoglycans an inhibitor of human immunodeficiency virus binds to T lymphocytes and showed antiviral activity Toido et al. (2003). In low concentrations, the extract showed potent inhibitory effect towards Herpes simplex virus and thus has got significant drug value. The antiviral activity using the sea cucumber Ludwigothurea grisea and Thelenota ananas derived fucosylated chondroitin sulfates (FCS), was recognized as the sulfated polysaccharides. It inhibited human immunodeficiency virus (HIV) infection Mc Clure et al. (1992). The present work suggested that Holothuria atra extracts showed virucidal action through reduction in number of plaques formed during plaque reduction assay against the HSV-1 and HSV-2. The strong growth inhibitory activity found in the extract of Holothuria atra might be the source for the development of antitherpetic compound.

The structure of bioactive compounds was elucidated by using NMR spectra from the active fractions. In addition, the methyl groups were observed in the 1H NMR spectra including singlets and doublets which were integrated relatively for olefinic proton at δ position. The 13C NMR spectrum showed the presence of a carbon—carb on double and indicated the presence of two conjugated carbonyls. It also showed the appearance of two carbon signals. Figure 5 represent the 1H and 13C NMR data of Holothuria atra. Some of the bioactive compounds identified with their structures are given below. Sea cucumber derived fucosylated chondroitin sulfates (FCS) which inhibited the growth of human immunodeficiency virus and also acted as a cytotoxic agent was initially obtained from Stichopus badionotus Kaswandi et al. (2004). It could be predicted that high molecular weight compounds present in Holothuria atra detected by NMR analysis could form a potent antiviral sources (Additional file 1).

Conclusions

The H. atra extract had various compounds such as the flavonoids, phenolic components, terpenoids, saponins, alkaloids etc. The GC-MS analysis revealed the presence of 59 compounds. It was found that H. atra extracts showed anti proliferative activities against the Hela and MCF-7 cell lines. Similarly the inhibitory action of extracts were found against the HSV-1 and HSV-2 strains was analyzed by plaque reduction assay. From NMR analysis the structural elucidation of the active compounds were studied. These results will direct future efforts to optimize the anti proliferative activity of these bioactive compounds.

Additional file

Additional file 1: Structures of bioactive compounds as based on NMR spectrum [Figure 5].

Competing interests

The authors declare that they have no competing interests.

Authors’ contributions

DID carried out the studies on antitumor and antiviral activities using the H. atra extracts. The bioactive compounds were identified from the purified extracts APL drafted the manuscript. All authors read and approved the final manuscript.
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Author details
1Centre for Marine Science and Technology, Manonmaniam Sundaranar University, Rajakkamangalam, Kanyakumari District, Tamil Nadu 629502, India.
2Marine Biotechnology Laboratory, Central Marine Fisheries Research Institute, Vizhinjam 695521, Thiruvananthapuram, Kerala, India.

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