Huang-Lian-Jie-Du-Tang Protects Rats from Cardiac Damages Induced by Metabolic Disorder by Improving Inflammation-Mediated Insulin Resistance

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Abstract

Huang-lian-jie-du-tang (HLJDT), a traditional Chinese medicine, has been shown to improve insulin resistance (IR) induced by inflammation, a key event in the development of metabolic syndrome (MS). The present study aimed to investigate the protective effects of HLJDT on MS and explore the underlying mechanism. MS rats were established with obese-diets and treated with normal saline, aspirin or HLJDT. The myocardial lesions were identified by echocardiogram, transmission electron microscope, and Sirius-red staining. The inflammatory cytokines were measured by ELISA and real-time PCR. The activation of NF-κB, JNK, SOCS3, IRS1 and AKT in the heart was detected by immunohistochemistry and Western blot analysis. Compared with the controls, MS rats developed obvious obesity, hypertension, dyslipidemia, IR, inflammation, and cardiac damage. Moreover, phosphorylated IRS-1 at Ser307 was correlated with the activation of NF-κB, JNK and SOCS3 and the inhibition of AKT in the heart from MS rats. These data suggest that serine phosphorylation of IRS-1 in response to inflammation is mediated, in part, by NF-κB, JNK and SOCS3. Notably, HLJDT inhibited the activation of NF-κB and reduced serine phosphorylation of IRS-1. In summary, HLJDT protects myocardium from IR-mediated injury by inhibiting serine phosphorylation of IRS-1 in MS rats.

Materials and Methods

Composition and Preparation of Huang-Lian-Jie-Du-Tang (HLJD)

HLJDT were purchased from Jianlian Company of Traditional Crude Drugs (Jinan, China), and carefully authenticated by Dr. Xiang-Hong Liu [Pharmaceutical Preparation Section, Shandong University Qilu Hospital, Jinan, China]. Voucher specimens (numbers were listed in Table 1) were deposited at the Herbarium.
of Shandong University (Jinan, China). Rhizoma coptidis, Radix scutellariae, Cortex phellodendri and Fructus gardeniae were mixed in the ratio of 3:2:2:3 and the total weight was 500 g. The mixture was decocted twice by refluxing with water (1:10 and then 1:5 w/v) for 1 h, and the solution obtained was concentrated to give an extract of 330 g. The yield of HLJD T extract was about 66.7% (w/w) according to the original herbs and the whole processes were manipulated by Pharmaceutical Preparation Section, Shandong University Qilu Hospital, Jinan, China. For the extract, the content of berberine hydrochloride was 18 mg/g measured by the lamellar scanning method. The resulting products were stored at 4°C and diluted to the desired concentrations with PBS (pH 7.4) and filtered before use.

Animals

The animal studies were approved by the Animal Ethics Committee of Shandong University. Six-week-old 130–150 g Wister male rats (n = 75) were obtained from Shandong University (Jinan, China). Rats were randomized into two diet groups: one group (n = 10) was fed normal diet and the other group (n = 65) was fed with obese-diet (2% fat, 10% sucrose, 6% salt and 8% defatted milk powder) and drinking water (20% sucrose solution) at libitum. After 16 weeks, 24-h fasting blood samples, final body weight (BW) and systolic blood pressure (SBP) were obtained. Compared with the controls, the obese-fed rats with MS were identified, using criteria analogous to ATP III as follows: hypertriglyceridemia, low HDL-C, high fasting glucose, excessive waist circumference, and hypertension. 32 MS rats were randomly divided into three groups: MS+HLJD T group (MS+H, n = 11), MS+Aspirin group (MS+A, n = 11), and MS group (n = 10). All of the rats were fed obese diet before and after treatment, and dosed orally with HLJD T (1.04 g/100 g), aspirin (120 mg/kg), and equal volume of PBS, respectively, every day for 12 weeks. Age matched control Wistar rats (n = 10) were randomly selected (NC group), which were fed normal diet throughout the experiments and dosed orally with equal volume of PBS every day for 12 weeks. HLJD T was dissolved in PBS and the rats were monitored to ensure HLJD T was completely consumed.

Analysis of BW, SBP and Heart Rate

BW was recorded every two weeks in the morning throughout the experiment, together with SBP and Heart Rate (HR) by the tail-cuff method using a Rat Tail Manometer provided by Japanese and Chinese Friendly Hospital (RBT-1; Beijing, China).

Analysis of Fasting Insulin, Glucose and Lipid Profiles in the Serum

Samples of venous blood were collected, fasting insulin (FINS), glucose (FBG), and lipid profiles (triglyceride, cholesterol, HDL-C, LDL-C) were measured after 12-hour fasting every two weeks.

Table 1. Recipe of Huang-Lian-Jie-Du-Tang (HLJD T) formulation.

| Components       | Voucher specimens number | Part used | Amount used (g) |
|-------------------|--------------------------|-----------|----------------|
| Coptis chinensis  | SU00186                  | Root      | 150            |
| Scutellaria baicalensis | SU00174              | Root      | 100            |
| Phellodendron amurense | SU00165             | Bark      | 100            |
| Gardenia jasminoides | SU00201            | Fruit     | 150            |

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Insulin resistance index was calculated as follows: IRI = (FBG*-FINS)/22.5.

Echocardiogram Analysis

Echocardiogram was performed on left ventricular (LV) at baseline (6 week), after 16 week, and at the end of the study, using Hewlett-Packard equipment, Sonos 7500 model, and an electronic 10-MHz transducer. All the animals were anesthetized with 10% chloral hydrate (0.3 ml/100 g) by intraperitoneal injection, and, after eplation of the anterior region of the thorax, they were positioned in the left lateral decubitus position. Two-dimensional and M-mode images from the parasternal long and short axis were obtained and recorded on videotape for later review. The cardiac structures were measured in at least three consecutive heart cycles. The left ventricle diastolic and systolic diameter (LVDd and LVSd), interventricular septum (IVS), and the left ventricle posterior wall thickness (LVPPW) were measured. The left ventricular fractional shortening (FS) was calculated using the following equation: FS (%) = [(LVDd – LVSd)/LVDd] × 100%. Pulse-wave Doppler spectra (E and A wave) velocity of mitral inflow were recorded from the apical 4-chamber view, with the sample volume placed near the tips of the mitral leaflets. The peak velocities in the rapid ventricular filling phase (E wave) and during atrial contraction (A wave) were measured from the videotape in 3 consecutive cardiac cycles.

Siris Red Staining

At the end of the experiment, the rats were executed. Then, the heart was isolated at the aortic root and both atria were removed. A part of LV was cut transversely parallel to the atrio-ventricular groove at the equatorial plane and embedded in paraffin, then 5 µm-thick sections of LV were cut and stained with the collagen-specific Sirius red for the measurement of the interstitial fibrosis.

Immunohistochemical Staining

The sections were dewaxed and rehydrated, then soaked in 1% citric acid buffer, pH 6.0 at 92–98°C for 15 min for antigen retrieval. The slides were cooled at room temperature for 30 min, rinsed with PBS, and incubated in 5% bovine serum albumin for 20 min to block nonspecific binding. The slides were incubated with 1:200 rabbit polyclonal anti-rat NF-κB p65 antibody (Abcam, USA) or 1:200 rabbit polyclonal anti-rat ICAM-1 antibody (Santa Cruz Biotechnology, USA) overnight at 4°C, then incubated with biotinylated anti-rabbit IgG secondary antibodies. DAB substrate kits (Vector Labs) were used to reveal the immunohistochemical reaction. PBS was used instead of primary antibody as a negative control.

Transmission Electron Microscope (TEM)

A part of the myocardium, about 0.5×1×5 mm³, was fixed with 2.0% glutaraldehyde for electron microscopic examination. The
tissues were examined by using an H-7000FA transmission electron microscope (Hitachi Co. Ltd., Tokyo, Japan) for ultrastructural changes in cardiomyocytes.

**Quantitative Reverse Transcription–polymerase Chain Reaction (qRT–PCR)**

Total RNA was prepared from myocardium samples using Trizol reagent (Invitrogen, Inc., Carlsbad, CA, USA), according to the manufacturer’s instructions. Total RNA was treated with D N a s e (D N A r e e r e , Ambion Inc., Austin, TX, USA) to remove genomic DNA and quantified using a spectrophotometer (Eppendorf Co. German). Three microgram of total RNA was reverse transcribed into cDNA using Murine Moloney Leukemia virus (M-MLV) reverse transcriptase (Promega, USA). qRT–PCR was performed using SYBR Green reagent (TaKaRa Corp., Kyoto, Japan) and primers specific for collagen I/III, TGF-

**Gene rimer sequence(5’–3’)** | **Product length (bp)** | **Annealing temp( C)** | **Cycle**
--- | --- | --- | ---
IL-6 | GTCAACTCCATCTGCCCCTTC | 164 | 60 | 40
ACTGGTCGTTTGGGTTGGTTG | | | |
ICAM-1 | TTTCGATCTTCCGACTAGGG | 112 | 60 | 40
AGCTTCAAGGGCAGGAAACAA | | | |
TNF-α | AGTCCGGGCAAGTCTACCTT | 175 | 60 | 40
TGAACCAAATCTCCCTTCT | | | |
TGFβ1 | GGTGACTTTCCTGTTGACGAGCA | 106 | 55 | 40
TTGAAATATACACACGCAAGGC | | | |
Collagen I | TTACCCTACAGCAGCTGT | 196 | 57 | 40
TTGGATGAGGGGATTTAC | | | |
Collagen III | GGTCACTTTCCCTGTTGACAGCA | 201 | 55 | 40
TTGTAATATACACACGCAAGGC | | | |
β-actin | CGTTGACATCCGTTAAGACC | 176 | 60 | 40
TAGAGCCACCAATCCCAACA | | | |

**Enzyme-linked Immunosorbent Assay (ELISA)**

Serum samples were collected from the rats every two weeks and kept frozen at -80°C until analysis of TNF-α level by a commercially available ELISA kit (Bender MedSystem, Austria, Vienna) according to the manufacturer’s instructions. The reaction was terminated by the addition of acid and the absorbance was measured at 450 nm. A standard curve was prepared from seven standard dilutions and serum TNF-α level was determined.

**Western Blot Analysis**

The frozen LV tissues were extracted and equal amounts of protein (50 μg) were fractionated on 10% SDS-polyacrylamide gels and then transferred to nitrocellulose membranes. Membrane was blocked at 4°C with 5% nonfat milk in Tris-buffered saline (25 mM Tris, 137 mM NaCl, and 2.7 mM KCl) containing 0.05% Tween-20 and then incubated overnight at 4°C with the following primary antibodies: rabbit polyclonal anti-rat NF-kB antibody (Abcam, USA; dilution: 1:500), rabbit polyclonal anti-rat SOCS3 antibody (Abcam, USA; dilution: 1:400), rabbit polyclonal anti-rat phospho-JNK (Thr183/Tyr185) antibody (Cell Signaling Technology, USA; dilution: 1:1000), rabbit polyclonal anti-rat phospho-IRS-1 (Ser307) antibody (Cell Signaling Technology, USA; dilution: 1:1000), rabbit polyclonal anti-rat phospho-Akt (Ser473) antibody (Cell Signaling Technology, Germany). Primer sequences and optimal PCR annealing temperatures and cycle numbers were listed in Table 2, and transcript levels were normalized to β-actin, which was used as an endogenous control. The 2^−ΔΔCt relative quantification method was used to calculate fold difference in transcript levels between samples.

**Results**

**HLJDT Affects Baseline Measurements for BW, SBP and HR**

At baseline, mean BW and SBP were similar between the normal-fed and obese-fed rats (P>0.05). After 16 weeks, the obese-fed rats were significantly overweight and had higher SBP (7% and 36% increase, respectively) compared with the normal-fed rats (P<0.05) (Fig. 1). After 12-week treatment with aspirin or HLJDT, BW was markedly reduced by 18% or 17%, respectively (Table 3). SBP was reduced by 8% in HLJDT-treated rats but not significantly changed in aspirin-treated rats (Table 3). There was no significant difference in HR among all groups (Table 3).
HLJDT Affects Baseline Measurements for Serum Values

At baseline, all the measured values, including FBG, FINS, TG, CHO, HDL-C and LDL-C, were similar between the normal-fed and obese-fed rats \( (P > 0.05) \). 16 weeks later, the obese-fed rats tended to have higher CHO, FBG and FINS levels (21%, 69% and 54% increase, respectively), and lower HDL-C (17% decrease) compared with the normal-fed rats \( (P < 0.05) \). At the end of the experiment, MS rats had the greatest decrease in HDL-C levels and increase in FINS and FBG levels compared with NC group (Table 3). Furthermore, there was significant decrease of FINS level in both aspirin and HLJDT-treated groups compared with MS group (Table 3). HDL-C levels were increased by 11% and 22% in aspirin-treated and HLJDT-treated rats, respectively (Table 3).

HLJDT Improves Cardiac Function

Characteristics of echocardiogram at baseline showed no significant differences between the normal-fed and obese-fed rats \( (P > 0.05) \). After 16 weeks, IVS and LVPW in the obese-fed rats obviously increased (9% and 9% compared with the controls, respectively), indicating the presence of LV concentric hypertrophy \( (P < 0.05) \). FS, representing LV systolic function, was not different between the obesity-fed and normal-fed rats \( (P > 0.05) \). Furthermore, E/A was decreased by 23% in the MS rats \( (P < 0.05) \) (Fig. 2). At the end of the study, there were more marked changes in IVS, LVPW and E/A (Table 4). Additionally, compared with the MS rats, treatment with either aspirin or HLJDT significantly slowed the progression of cardiac hypertrophy and protected the diastolic function, as estimated by IVS, LVPW and E/A (Table 4). Moreover, there were no significant differences between aspirin-treated and HLJDT-treated groups.
HLJDT Improves Ultrastructure of Cardiomyocytes

TEM analysis showed the derangement of myofibers, and swollen mitochondria in cardiomyocytes in MS rats compared with the NC rats (Fig. 3). Moreover, after aspirin or HLJDT treatment, the cardiac ultrastructure was obviously improved (Fig. 3).

HLJDT Affects Collagen Contents

Sirius red staining revealed an increase of interstitial fibrosis in the myocardium of MS rats compared to the NC group. Furthermore, a fraction of interstitial fibrosis was prevented by both aspirin and HLJDT (Fig. 4).

HLJDT Decreases Serum TNF-α Level

At baseline, the mean value of TNF-α was similar in the obese-fed and normal-fed rats. Serum TNF-α level of MS rats was 4 fold higher than in the controls at 16 weeks (Fig. 5). The difference persisted till the end of the study. However, the elevated TNF-α level was decreased by 36% in aspirin group and 26% in HLJDT group (Fig. 5).

HLJDT Decreases NF-κB p65 and ICAM-1 Levels in the Heart

The expression of NF-κB p65 and ICAM-1 in the rat hearts was evaluated by immunohistochemistry staining. In the MS rats, the expression of NF-κB p65 and ICAM-1 was significantly increased in the myocardial tissues compared to the NC group. However, both NF-κB p65 and ICAM-1 staining were decreased in either aspirin or HLJDT-treated rats compared to MS rats (Fig. 6).

HLJDT Downregulates the Expression of Inflammatory Factors in the Heart

The mRNA expression of IL-6, TNF-α, ICAM-1, collagen types I and III, TGF-β1 and IKKβ were significantly increased in MS rats compared to NC rats (P<0.05, Fig. 7). However, aspirin and HLJDT significantly attenuated the increased expression of IL-6, TNF-α, ICAM-1, collagen I, collagen III and TGF-β1 mRNA (P<0.05, Fig. 7). The interclass analysis showed that these

Table 3. General characteristics in the four groups after administration.

| Parameter          | NC (n = 10) | MS (n = 10) | MS+A (n = 11) | MS+H (n = 11) |
|--------------------|-------------|-------------|--------------|--------------|
| BW (g)             | 565.58±26.27 | 662.87±25.79** | 545.98±34.87#* | 554.29±35.26** |
| SBP (mmHg)         | 101.83±4.45   | 136.89±7.02** | 133.51±7.13   | 126.43±7.52** |
| HR (beats/minute)  | 412.89±38.65  | 407.69±53.82  | 408.58±40.79  | 419.78±45.39  |
| FBG (mg/dL)        | 5.29±0.52     | 8.72±0.96**   | 8.11±0.79     | 6.59±0.76**   |
| FINS (μU/mL)       | 17.89±5.36    | 36.15±9.86**  | 25.85±7.88#*  | 24.89±8.46#*  |
| IRI (μU/mL)        | 5.28±1.31     | 13.96±3.19**  | 8.05±2.16#    | 6.89±2.12#    |
| TG (mg/dL)         | 0.75±0.27     | 0.92±0.25     | 0.75±0.18     | 0.74±0.36     |
| HDL-C (mg/dL)      | 1.38±0.21     | 1.06±0.15**   | 1.19±0.20     | 1.35±0.21#*   |
| LDL-C (mg/dL)      | 0.36±0.06     | 0.40±0.06     | 0.37±0.05     | 0.36±0.05     |

All data were mean±SD. BW (g): body weight; SBP: blood pressure (mmHg); HR (beats/minute): heart rate; FBG (mmol/L): fasting blood glucose; FINS (μU/mL): fasting insulin; IRI: insulin resistance index; TG (mmol/L): triglyceride; HDL-C (mmol/L): HDL-cholesterol; LDL-C (mmol/L): LDL-cholesterol.

*P<0.05 vs. NC group; **P<0.01, #P<0.05 vs. MS group.

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Table 4. Parameters measured by echocardiogram.

| Parameter          | NC (n = 10) | MS (n = 10) | MS+A (n = 11) | MS+H (n = 11) |
|--------------------|-------------|-------------|--------------|--------------|
| LVDD (mm)          | 5.36±0.39   | 5.43±0.42   | 5.39±0.43    | 5.33±0.45    |
| LVSd (mm)          | 2.19±0.19   | 2.31±0.26   | 2.23±0.24    | 2.09±0.23    |
| IVS (mm)           | 1.73±0.06   | 2.16±0.17** | 1.96±0.13**  | 1.89±0.12**  |
| LVPW (mm)          | 1.86±0.11   | 2.19±0.12** | 1.99±0.06**  | 1.93±0.07**  |
| FS (%)             | 0.59±0.02   | 0.58±0.05   | 0.57±0.05    | 0.59±0.04    |
| A wave (cm/s)      | 35.89±8.47  | 64.76±12.76** | 51.67±10.35** | 52.36±11.79** |
| E/A                 | 2.13±0.19   | 1.36±0.26** | 1.67±0.23#   | 1.69±0.22#   |

All data were mean±SD. LVDD (mm): left ventricular end-diastolic dimension; LVSd (mm): left ventricular end-systolic dimension; IVS (mm): intraventricular septal wall; LVPW (mm): left ventricular posterior wall thickness; FS: fractional shortening; A wave (cm/s): peak late diastolic flow velocity, E/A: ratio of peak early diastolic filling velocity to peak velocity at atrial contractions.

**P<0.01, *P<0.05 vs. NC group; #P<0.05 vs. MS group.

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Figure 2. Transmitral inflow patterns (E and A wave) for MS rat at 22 and 34 weeks. Note increased A waves and decreased E/A in the MS group.

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values were not significantly different between aspirin-treated and HLJDT-treated groups (\(P>0.05\), Fig. 7).

**HLJDT Downregulates the Phosphorylation of IRS-1 in the Heart**

Western blot analysis revealed a profound up-regulation of the levels of SOCS3 and phospho-JNK by inflammation. At the same time, serine phospho-IRS1 was increased and phospho-AKT was decreased in the MS rats compared to the NC group (\(P<0.05\), Fig. 8). Treatment with aspirin markedly attenuated the increases of phospho-JNK and phospho-IRS1 levels (\(P<0.05\), Fig. 8), while the level of phospho-JNK was not affected by HLJDT treatment (\(P>0.05\), Fig. 8). Moreover, compared with the MS rats, phospho-AKT level was increased in both aspirin and HLJDT-treated rats (\(P<0.05\), Fig. 8).

**Figure 3. Cardiac lesions measured by transmission electron microscopy.** A: NC group (n = 10). Note the regular membrane and intercalated disc between adjacent myocytes. B: MS group (n = 10). Interrupted capillary membrane, and derangement and increase of myofibers and mitochondria in cardiomyocytes were visible. Swollen mitochondria were shown by the arrows. C: MS+A group (n = 11). D: MS+H group (n = 11). Compared with MS group, the ultrastructural changes of drug-fed groups were obviously improved. Original magnification: \(\times10,000\). doi:10.1371/journal.pone.0067530.g003

**Figure 4. Cardiac fibrosis measured by Sirius Red staining.** The top panels showed representative images of Sirius Red-stained LV sections. A: NC group (n = 10); B: MS group (n = 10); C: MS+A group (n = 11); D: MS+H group (n = 11). The bottom bar graph showed quantitative interstitial fibrosis ratio. Each bar represented the mean ±SD. **\(P<0.01\), *\(P<0.05\) vs. NC; ###\(P<0.01\), ##\(P<0.05\) vs MS. Original magnification, \(\times400\). doi:10.1371/journal.pone.0067530.g004

**Figure 5. Serum concentration of TNF-α measured by ELISA.** The left bar graph indicated serum concentration of inflammatory cytokines before administration, while the right one indicated serum concentration of inflammatory cytokines after the administration of aspirin or HLJDT for 12 weeks. Each bar represented the mean ±SD. **\(P<0.01\), *\(P<0.05\) vs. NC; ###\(P<0.01\), ##\(P<0.05\) vs. MS. NC group (n = 10); MS group (n = 10); MS+A group (n = 11); MS+H group (n = 11). doi:10.1371/journal.pone.0067530.g005
Discussion

In this study, we succeeded in producing obesity and metabolic disturbances similar to those seen in human MS by feeding obese-diet to rats. Based on this model we showed that MS caused significant myocardial lesions, increased the production of inflammatory cytokines and reduced insulin signaling in the heart. In previous study, we demonstrated that HLJDT treatment could inhibit inflammation and ameliorate MS-induced cardiac damage [15]. In the present study, we further investigated the mechanism by which HLJDT could improve IR, and compared the therapeutic effect between HLJDT and aspirin.

The obese-diet used in the present study was a high-fat, high-glucose and high-salt diet and sucrose solution, similar to the content of Western diets. We developed a MS model of rats with the following features: (1) Significant differences in BW between the obese-fed and normal-fed rats were developed from 20 weeks of age and persisted till 34 weeks. (2) Increased SBP was found in MS rats from 12 weeks of age, consistent with the classical features of MS [16]. (3) Low HDL-C was found in MS rats, while high TG and LDL-C were not found, suggesting that the lipid profile in MS rats is not exactly same as in human MS. (4) Elevated FBG, INS and IRI are regarded as a distinctive measure of IR. (5) The expression of inflammatory cytokines was increased and associated with increasing BW. These trends resembled those reported in human with obesity and IR.

MS is defined by the cluster of several classic cardiovascular risk factors (such as hypertension, obesity, type 2 diabetes, high triglycerides and low HDL-C), and the underlying pathophysiology is thought to be related to IR [17,2]. In the present study, the obese-fed rats developed obesity with low HDL-C as well as elevated BP, plasma glucose and insulin, which are considered as MS-related features. Therefore, this animal model is regarded as a model that mimics human MS and could be used to investigate the pathophysiology of cardiac damage and develop novel pharmacotherapy and preventative measures for MS.

IR is defined as a state of reduced responsiveness to normal circulating levels of insulin and plays a major role in the development of MS [18,2,19,20]. Thus we propose that IR is a key therapeutic target for the treatment and prevention of MS. Recent reports have linked the relation between obesity and IR in two different ways. First, ectopic lipid accumulation is a potential mechanism for this relationship. Second, a systemic chronic inflammatory response in obesity, characterized by altered

Figure 6. Immunohistochemistry staining of ICAM-1 and NF-κB p65 in the heart. A,E: NC group (n = 10); B,F: MS group (n = 10); C,G: MS+A group (n = 11); D,H: MS+H group (n = 11). The panels showed representative images of micrographs of immunohistochemistry staining in 4 groups. Original magnification, ×400.
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Figure 7. Expression of IL-6, TNF-α, ICAM-1, Collagen I/III and TGF-β1 mRNA in the heart. mRNA levels were detected by real-time PCR and each bar represented the mean ±SD. **P<0.01, *P<0.05 vs. NC; ***P<0.01, **P<0.05 vs. MS. NC group (n = 10); MS group (n = 10); MS+A group (n = 11); MS+H group (n = 11).
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cytokine production and activation of inflammatory signaling pathways, is another mechanism [21]. In a chronic inflammatory response of MS, inflammatory cytokines such as TNF-α, IL-1 and IL-6 were activated [4]. In addition, in adipose tissue of obese rodents and humans, TNF-α expression is increased [22], and reducing TNF-α expression could reduce IR in obese rodents [23].

Inflammation-mediated IR implicates several pathways including IKKα/β/NF-κB, JNK and SOCS3, which are activated by inflammatory cytokines. Many studies demonstrated that inhibiting the activity of IKK, JNK and SOCS3 by pharmacological inhibition or gene knockout could improve IR [8,24–27,9]. In our study, the expression of inflammatory cytokines such as TNF-α, IL-1, and IL-6 was inhibited by pharmacological inhibitions, resulting in improving IR. Taken together, these results suggest that aberrant inflammation pathways may contribute to the onset of IR in MS rats.

Insulin signaling pathway in metabolism is regulated by the insulin-stimulated tyrosine phosphorylation of IRS-1 and the activity of PI3-kinase/AKT signaling pathway. The PI3K-dependent activation of the AKT kinase is known to control programmed cell death and cellular metabolism [28]. Moreover, activated AKT could preserve cardiac function because in rodent models of myocardial infarction, bone marrow–derived mesenchymal stem cells expressing constitutively activated AKT could enhance cardiomyocytes survival and organ function [29]. In our study, there were significant differences of left ventricle (LV) between the obese-fed and normal-fed rats, including LV structure (IVS and LVPW), diastolic function (A wave and E/A), and myocardial composition (the amount of collagen I/III). These differences were associated with reduced PI 3-kinase/AKT pathway. These results suggest that a long exposure to the obese-diet could inhibit insulin signaling, thereby inducing myocardial damage on both cardiac structure and function.

The efficacy of aspirin (acetylsalicylic acid) on reducing serine phosphorylation of IRS-1 associated with decreasing inflammatory cytokines has been reported [13]. In our study, by using aspirin as positive control we confirmed these findings. Thus we conclude that aspirin may enhance the insulin sensitivity by protecting IRS proteins from serine phosphorylation catalyzed by NF-κB and JNK.

HLJDT, a combination of herbs used in traditional Chinese medicine, consists of four medicinal compositions. Coptidis rhizome is the principal drug in HLJDT and Berberine hydrochloride, a key component of coptidis rhizome, has shown inhibitory effect on inflammation [30]. Although the effects of HLJDT on anti-inflammation and IR have been reported [31,32], the underlying mechanism remains elusive. In our study, we found that HLJDT could reduce inflammatory mediators and improve IR, and this is associated with the inhibition of NF-κB activation induced by inflammatory cytokines. Compared with aspirin, the effects of HLJDT on reducing inflammation and increasing insulin-sensitivity were not significantly different.

In conclusion, insulin resistance induced by inflammation in MS could impair cardiac structure and function. Thus, increasing insulin sensitivity could contribute to MS therapy. The traditional Chinese medicine such as HLJDT could be an effective strategy to improve IR and protect MS patients from IR-mediated cardiac injury.

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Author Contributions

Conceived and designed the experiments: YGC XPJ. Performed the experiments: CBL XXL PLB. Analyzed the data: XXL. Contributed reagents/materials/analysis tools: HQG YZ. Wrote the paper: CBL.

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Figure 8. The activation of SOCS3, phospho-JNK, phospho- AKT and serine phospho- IRS1 in the heart. The left panels showed representative blots of the left ventricular. The right bar graph showed relative protein levels. Each bar represented the mean ±SD. **P<0.01, *P<0.05 vs. NC; ##P<0.01, *P<0.05 vs. MS. NC group (n = 10); MS group (n = 10); MS+A group (n = 11); MS+H group (n = 11). doi:10.1371/journal.pone.0067530.g008
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