Preferential Tumor Localization in Relation to 18F-FDOPA Uptake for Lower-Grade Gliomas

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Abstract

Purpose

Although tumor localization and 3,4-dihydroxy-6-\(^{18}\)F-fluoro-L-phenylalanine (FDOPA) uptake may have an association, preferential tumor localization in relation to FDOPA uptake is yet to be investigated in lower-grade gliomas (LGGs). This study aimed to identify differences in the frequency of tumor localization between FDOPA hypometabolic and hypermetabolic LGGs using a probabilistic radiographic atlas.

Methods

Fifty-one patients with newly diagnosed LGG (WHO grade II, 29; III, 22; isocitrate dehydrogenase wild-type, 21; mutant 1p19q non-codeleted, 16; mutant codeleted, 14) who underwent FDOPA positron emission tomography (PET) were retrospectively selected. Semiautomated tumor segmentation on FLAIR was performed. Patients with LGGs were separated into two groups (FDOPA hypometabolic and hypermetabolic LGGs) according to the normalized maximum standardized uptake value of FDOPA PET (a threshold of the uptake in the striatum) within the segmented regions. Spatial normalization procedures to build a 3D MRI-based atlas using each segmented region were validated by an analysis of differential involvement statistical mapping.

Results

Superimposition of regions of interest showed a high number of hypometabolic LGGs localized in the frontal lobe, while a high number of hypermetabolic LGGs was localized in the insula, putamen, and temporal lobe. The statistical mapping revealed that hypometabolic LGGs occurred more frequently in the superior frontal gyrus (close to the supplementary motor area), while hypermetabolic LGGs occurred more frequently in the insula.

Conclusion

Radiographic atlases revealed preferential frontal lobe localization for FDOPA hypometabolic LGGs, which may be associated with relatively early detection.

Introduction

Tumor localization is a key factor in the care of patients with lower-grade glioma (LGG) as it correlates with their demographic characteristics, molecular status, clinical presentation, surgical management, and survival time. For example, LGGs were more likely to invade the insula in elderly patients than in young patients, where they tended to localize in the temporal lobe [1]. From a molecular standpoint, only 20% of isocitrate dehydrogenase (IDH) wild-type gliomas were localized in the frontal lobe, while IDH mutant LGGs, especially with 1p19q co-deletion, were more likely to occur in the frontal lobe [2–4]. LGGs in the frontal lobe were associated with a higher risk of preoperative seizure episodes [5]. With regards to surgical intervention, LGGs involved in the insular region were associated with decreased extent of
resection and unfavorable prognosis, while patients with the frontal lobe LGGs experienced more thorough resection and exhibited more favorable prognosis than patients with LGGs in the other lobes [6]. For the evaluation of tumor distribution, several studies employed a probabilistic magnetic resonance imaging (MRI)-based brain atlas to specify the probability of anatomical tumor localization associated with the patients’ characteristics (age and sex) [7], clinical presentation (symptoms and Karnofsky performance) [8], and molecular status (IDH, epidermal growth factor receptor, O6-methylguanine methyltransferase, and phosphatase and tensin homolog) [9, 10].

Amino acid positron emission tomography (PET), including 3,4-dihydroxy-6-^{18}F-fluoro-L-phenylalanine (FDOPA), is often used as a clinical tool in neuro-oncology to identify metabolically active tumor tissue. Higher amino acid tracer uptake was reported to be associated with higher tumor grade and shorter overall or progression-free survival [11–13]. Molecular status, including IDH mutation and 1p19q codeletion, may also have an association with amino acid tracer uptake, although they are still debated [14–19].

Tumor localization and FDOPA uptake may have an association with each other and cooperatively influence the prognosis of patients with LGGs; however, preferential tumor localization in correlation with varying FDOPA uptake has not yet been investigated. The purpose of this study was to identify differences in frequency of localization between FDOPA hypometabolic and hypermetabolic LGGs using a probabilistic MRI-based brain atlas, and to evaluate overall survival (OS) with different FDOPA metabolic status. Delineation of such spatial patterns may improve understanding of underlying tumor pathophysiology and may also lead to appropriate subsequent management.

**Materials And Methods**

**Patient Selection**

A total of 51 adult patients (age > 20 years) with newly diagnosed and histologically confirmed LGGs (World Health Organization [WHO] grade II or III) who underwent FDOPA PET and MRI between 2007 and 2019 were retrospectively selected. All patients were diagnosed with LGGs according to the WHO 2007 or 2016 classification, and were classified based on IDH mutational status and 1p19q codeletion status by conventional techniques [20]. No patients underwent stereotactic biopsy prior to FDOPA PET or MRI. OS was measured from the time of the PET scan until death or the censored date (maximum, 2000 days) with median term of 740 days. All patients signed institutional review board-approved consent forms to have their imaging, clinical, and molecular data included in our research database (IRB IRB#10-000655). The patient cohort in this study was partly overlapped with a previous study [21].

**FDOPA PET Image Acquisition**

FDOPA PET images were acquired with a high-resolution full-ring PET/CT scanner (ECAT-HR; CTI/MIMVista; Siemens, Knoxville, TN, USA) after the subjects fasted for more than four hours. Following previously reported procedures, FDOPA was synthesized and injected intravenously [22, 23]. Computed
tomography images were acquired prior to the PET scan for attenuation correction. Three-dimensional FDOPA emission data were acquired for a total of 30 minutes, and the data were integrated between 10–30 minutes following the injection to obtain 20-minute static FDOPA images after reconstruction. FDOPA PET images were reconstructed using an ordered-subset expectation maximization iterative reconstruction algorithm consisting of six iterations with eight subsets [24, 25]. Finally, a Gaussian filter with a full width at half maximum of 4 mm was applied. The resulting voxel sizes were 1.34 mm × 1.34 mm × 3 mm for FDOPA PET images. Standardized uptake value (SUV) maps of FDOPA were calculated based on the radioactive activity divided by the decay-corrected injected dose per body mass [26]. Resulting SUV maps were subsequently normalized relative to the median value of the normal-appearing striatum (nSUV) [20, 27].

**Magnetic Resonance Image Acquisition**

Anatomical MRI consisted of standard T1-weighted pre- and post-contrast images (2D axial turbo spin echo with 3-mm slice thickness and no interslice gap, or 3D inversion-prepared gradient echo images with 1–1.5 mm isotropic voxel size) and T2-weighted fluid-attenuated inversion recovery (FLAIR) images acquired at 3-mm slice thickness with no interslice gap using a 1.5-T or 3-T clinical MRI scanner.

**Postprocessing and ROI Analysis**

The processing procedures are described in Fig. 1. A single tumor region of interest (ROI) was segmented based on the regions of hyperintensity on T2-weighted FLAIR images by a board-certificated neuroradiologist (H.T. with 14 years of clinical experience) with Analysis of Functional NeuroImages software (AFNI; NIMH Scientific and Statistical Computing Core; Bethesda, MD, USA; https://afni.nimh.nih.gov) using a semi-automatic procedure as previously described [28, 29]. FLAIR and PET images, as well as FLAIR hyperintense ROIs were registered to the post-contrast T1-weighted images for each patient using a six-degree of freedom rigid transformation and a mutual information cost function using FSL software (flirt, FMRIB, Oxford, UK; http://www.fmrib.ox.ac.uk/fsl/). Each registered ROI was applied to the corresponding PET images. Maximum nSUV (nSUV_{max}) within the FLAIR hyperintense ROI and biological tumor volume (BTV), which included the voxels within the ROI higher than the median uptake value in the striatum, were calculated. The LGG patients were stratified into FDOPA hypometabolic (nSUV_{max} < 1) and hypermetabolic (nSUV_{max} > 1) groups according to the nSUV_{max} with a cut-off value of one relative to the striatum. This cut-off value, one, is determined according to the previous suggestion [30]. Anatomical FLAIR volume and BTV are reported as milliliters.

Each post-contrast T1-weighted images was registered to a 1.0 mm isotropic T1-weighted brain atlas (MNI152; Montreal Neurological Institute [MNI]) using a 12-degree of freedom affine transformation with statistical parametric mapping 12 software (SPM12; Wellcome Trust Centre for Neuroimaging, London, UK; https://www.fil.ion.ucl.ac.uk/spm/software/spm12/), and applied the transform matrix to each FLAIR ROI. The registered ROIs in the left hemisphere were flipped to the right hemisphere in the MNI space. All tumor ROIs in the MNI space were superimposed to create a voxel-wise frequency map of tumor.
occurrence in the hypometabolic and hypermetabolic groups separately, and used for the following analysis of differential involvement (ADIFFI) statistical mapping technique [9, 10].

Statistical Analyses

The demographic (sex, age, WHO grade), molecular (IDH/1p19q) status, initial symptoms (seizure; focal neurological deficit, i.e. aphasia, hemiparesis, and muscle weakness; neurological symptoms, i.e. optic/olfactory/hearing/tasting abnormalities, dizziness, and vertigo; headache; other symptoms, i.e. mental/personality changes, unusual feelings; or incidental), and imaging metrics (nSUV_{max}, FLAIR volume, and BTV) were compared between the FDOPA hypometabolic and hypermetabolic LGGs using the Fisher's exact or Mann–Whitney U test.

ADIFFI consisted of first constructing a 2 × 2 contingency table comparing two differential phenotypes (e.g. phenotypes A and B) and tumor versus non-tumor for each image voxel. Next, a two-tailed Fisher's exact test was performed on a voxel-wise basis. According to the Fisher's exact test, the probability of obtaining an observed pattern in the 2 × 2 contingency table is given by

\[ p = \frac{(a + b)!(c + d)! (a + c)! (b + d)!}{a! b! c! d! n!}, \]

where \( a \) is the frequency of tumor occurrence in a particular voxel for phenotype A; \( b \) the frequency of tumor occurrence in a particular voxel for phenotype B; \( c \) the frequency of no tumor occurring in a particular voxel for phenotype A; \( d \) the frequency of no tumor occurring in a particular voxel for phenotype B; \( n \) the total number of patients; and the exclamation point represents the factorial operation. To calculate the significance of the observed pattern, the contingency table corresponded to the total probability of observing a pattern in the contingency table as extreme or more extreme. Then, the \( p \) value was recalculated from each voxel for all cases in which the marginal totals were the same as the observed tables, and only for cases in which the arrangement was as extreme as the observed pattern. We performed this iteratively so that values were incremented to calculate a more extreme pattern, adding the previous \( p \) value in each image voxel each time until the most extreme pattern was achieved, which varied from voxel to voxel. The final \( p \) value represents the probability of observing the given pattern in the contingency table by chance. The \( p \) values less than .05 were considered significant. Additional details are presented in a previous publication [9].

For the cluster-based permutation correction outlined by Bullmore et al., [31] a total of 500 random permutations were performed, the resulting ADIFFI-defined clusters with a connection of 18 directions were retained, and the 95% confidence interval (CI) for significant cluster size occurring by chance were documented. The cluster-size thresholds had a 5% probability of occurring by chance.

ADIFFI and cluster-based correction were additionally performed between different age, FLAIR volume, or BTV with cut-off of the median value, and also among different molecular statuses.
Kaplan-Meier curves were used to depict differences in the OS, and the log-rank test was employed to compare OS between the FDOPA hypometabolic and hypermetabolic LGGs. Cox univariate regression analyses were conducted to investigate the association of OS with the age, nSUV\textsubscript{max}, FLAIR volume, and BTV.

Statistical analyses were performed using MATLAB (R2019b; MathWorks, Natick, MA, USA) and GraphPad Prism (Version 8.3; GraphPad Software, La Jolla, CA, USA).

**Results**

Table 1 summarizes patient demographics and molecular information, while Supplemental Table 1 describes each patient in more detail. The current study included 51 newly diagnosed patients with gliomas with a median age of 52 years at the time of the PET examination. The patient cohort consisted of 31 males and 20 females. According to 2007 or 2016 WHO criteria, 29 patients had grade II, and 22 had grade III gliomas. A total of 21 gliomas were IDH wild-type, 16 were IDH mutant 1p19q non-codeleted, and 14 were IDH mutant 1p19q codeleted. When comparing between different FDOPA metabolic status, patients with FDOPA hypometabolic LGGs (n = 14, nSUV\textsubscript{max} < 1) were significantly younger than patients with hypermetabolic LGGs (n = 37, nSUV\textsubscript{max} > 1) (p = .014). There were no significant differences between FDOPA hypometabolic and hypermetabolic LGGs in the ratio of the WHO grade (p = .54), IDH/1p19q mutation status (p = .57), nor initial symptoms (p = .43). The volumes of the FLAIR ROIs were significantly smaller in FDOPA hypometabolic than hypermetabolic LGGs (p = .012).
### Table 1: Patient demographics and imaging metrics

|                                      | Total | FDOPA hypometabolic LGGs | FDOPA hypermetabolic LGGs | \( p \) value |
|--------------------------------------|-------|--------------------------|---------------------------|---------------|
| Number of patients                   | 51    | 14                       | 37                        |               |
| Female                               | 20 (39%) | 8 (57%)                   | 12 (32%)                  | .12\textsuperscript{a} |
| Age (median [IQR], year)             | 52 (37, 61) | 47 (27, 51)               | 57 (39, 63)               | .014\textsuperscript{b*} |
| WHO classification grade             |       |                          |                           | .54\textsuperscript{a} |
| II                                   | 29 (57%) | 9 (64%)                   | 20 (54%)                  |               |
| III                                  | 22 (43%) | 5 (36%)                   | 17 (46%)                  |               |
| IDH mutation and 1p19q codeletion status |       |                          |                           | .57\textsuperscript{a} |
| Wild-type                            | 21 (41%) | 5 (36%)                   | 16 (43%)                  |               |
| Mutant 1p19q non-codeleted           | 16 (31%) | 6 (43%)                   | 10 (27%)                  |               |
| Mutant 1p19q codeleted               | 14 (27%) | 3 (21%)                   | 11 (30%)                  |               |
| Initial symptom                      |       |                          |                           | .43\textsuperscript{a} |
| Seizure                              | 13 (25%) | 3 (21%)                   | 10 (27%)                  |               |
| Focal neurological deficit           | 6 (12%)  | 1 (7%)                    | 5 (14%)                   |               |
| Neurological symptoms                | 10 (20%) | 3 (21%)                   | 7 (19%)                   |               |
| Headache                             | 6 (12%)  | 0 (0%)                    | 6 (16%)                   |               |
| Other symptoms                       | 8 (16%)  | 3 (21%)                   | 5 (14%)                   |               |
| Incidental                           | 8 (16%)  | 4 (28%)                   | 4 (11%)                   |               |
| \( nSUV_{\text{max}} \) (median [IQR]) | 1.16 (0.97, 1.48) | 0.94 (0.86, 0.96)         | 1.28 (1.13, 1.64)         | < .001\textsuperscript{b*} |
| Volume of FLAIR hyperintense regions (median, IQR, mL) | 29.7 (13.2, 63.9) | 12.3 (6.0, 31.5)          | 44.0 (18.2, 82.5)         | .012\textsuperscript{b*} |
| Biological tumor volume (median, IQR, mL) | 0.22 (0, 3.16) | 0 (0, 0)                  | 1.69 (0.17, 5.97)         | < .001\textsuperscript{b*} |
Superimposition of tumor ROIs of FDOPA hypometabolic or hypermetabolic LGGs on the MNI template brain suggested a high number of FDOPA hypometabolic LGGs localized in the frontal lobe (Fig. 2a). Meanwhile, a high number of FDOPA hypermetabolic LGGs was found to be localized in the insula, putamen, and temporal lobe (Fig. 2b). ADIFFI statistical analysis of LGGs based on the FDOPA metabolic status identified two spatially distinct clusters. One was localized in the frontal lobe, especially in the superior frontal gyrus, more frequently associated with FDOPA hypometabolic LGGs (Fig. 3a), and the other was localized in the insula more frequently associated with FDOPA hypermetabolic LGGs (Fig. 3b).

The preferential tumor localization in different age, FLAIR volume, BTV, or molecular status are shown in Supplemental Fig. 1–6. In brief, LGGs in younger patients (age < 52 years) preferentially localized in the frontal lobe (especially in the superior frontal gyrus and pars orbitalis to pars triangularis) and in the temporal lobe (especially in the fusiform gyrus). LGGs in older patients (age ≥ 52 years) did not have preferential localization. LGGs with smaller FLAIR volume (< 29 mL) or BTV (< 0.22 mL) did not show preferential localization. There was no preferential localization between either pair of different IDH/1p19q molecular status.

The Kaplan-Meier curves and log-rank tests showed a significant difference in OS between FDOPA hypometabolic and hypermetabolic LGGs (Fig. 4, p = .046), with longer survival in FDOPA hypometabolic LGGs. The Cox univariate analysis demonstrated a significant increase in hazard associated with the patient's age (hazard ratio [HR] = 1.075, 95% CI = [1.003, 1.152], p = .042), but not with the nSUV_{max} (HR = 1.403, 95% CI = [0.670, 2.937], p = .37), FLAIR volume (HR = 0.997, 95% CI = [0.977, 1.017], p = .74), or BTV (HR = 1.056, 95% CI = [0.991, 1.125], p = .089).

**Discussion**

In the current study, frequency of localization for gliomas with different FDOPA metabolic status was evaluated using a probabilistic MRI-based brain atlas. The FDOPA hypometabolic LGGs occurred more frequently in the frontal lobe, especially in the superior frontal gyrus, while the FDOPA hypermetabolic LGGs occurred more frequently in the insula. The log-rank test identified significantly longer survival in FDOPA hypometabolic LGGs compared with hypermetabolic LGGs.

The frontal lobe, especially the supplementary motor area (SMA), and the insular area have previously been reported as predominant localizations for LGGs [32, 33]. Cytoarchitectonic and chemoarchitectonic similarities or similar functional roles representing an interface between multimodal areas were suspected to affect the preferential localization in such areas [33]. Our findings are consistent with these
Previous studies, while revealing more frequent localization in the frontal lobe (close to the SMA) for the FDOPA hypometabolic LGGs and more frequent localization in the insula for the FDOPA hypermetabolic LGGs. As no significant differences were detected in the ratio of tumor grades nor IDH/1p19q molecular status between the different FDOPA metabolic status, other factors are likely to influence the associations of FDOPA uptake and preferential localization for LGGs.

Relatively early detection of the frontal lobe LGGs provides one possible hypothesis to explain preferential frontal lobe localization for FDOPA hypometabolic LGGs. The frontal lobe, especially the SMA, represents a functional interface between the prefrontal cortex and primary sensorimotor areas [33]. Specifically, the SMA plays an important role in the control and coordination of complex motor processes and takes part in programming sequential movement patterns [34]; thus, the frontal lobe LGGs close to such functionally eloquent areas may easily cause noticeable symptoms. A previous meta-analysis also indicated that LGGs in the frontal lobe were associated with a higher risk of preoperative seizure episodes [5]. Dysfunction of brain regions close to such eloquent areas and seizure episodes may result in early imaging examinations and, subsequently, contribute to early detection of gliomas.

This hypothesis was also supported by our results that patients with FDOPA hypometabolic LGG were significantly younger and represented smaller tumor volume in anatomical images compared to patients with hypermetabolic LGG. Incidentally detected LGGs at a non-symptomatic phase were reported to show younger patient age and smaller tumor volume [35–37]. The duration of the clinically non-symptomatic phase of LGGs was 4–15 years with velocity of diametric expansion at 3.5 mm per year [37, 38]. Meanwhile, gliomas often increased their amino acid tracer uptake throughout the course of the disease [39]. These results suggested that LGGs gradually increased their FDOPA uptake and volume from a non-symptomatic phase, and that FDOPA hypometabolic LGGs were detected at a relatively early phase due to their localization close to the symptomatic regions or by chance.

In contrast, the reason why the hypermetabolic LGGs preferentially localized to the insular region was unclear. Epilepsy, cognitive disturbance, and autonomic dysfunction are known symptoms in gliomas involved in the insula [40]. Meanwhile, a homotopic contralateral plasticity due to the slow-growing nature of LGGs may compensate the function of the insula, masking any noticeable symptoms until a relatively late phase [41].

The current study demonstrated that patients with FDOPA hypometabolic LGGs had longer OS than patients with hypermetabolic LGGs, although Cox regression did not show significant association of the OS with the nSUV_{max} or BTV. This was not surprising because amino acid tracer uptake is believed to reflect tumor activities, and several studies have reported better prognosis in patients with lower amino acid tracer uptake gliomas [12, 13]. According to our hypothesis, some hypometabolic gliomas may have been detected at a relatively early phase. On the other hand, some studies have reported a better prognosis in patients with LGGs in the frontal lobe than the other lobes, partly due to the improved rate of complete resection [6, 32, 42, 43]. Thus, the preferential frontal lobe localization and metabolic hyp-
activities of LGGs along with a possibility of early detection may interactively contribute to a better prognosis.

The major limitation of this study was a relatively small population size. Hence, all ROIs in the left hemisphere were flipped to the right hemisphere. It was not possible to evaluate the laterality of LGGs, while asymmetry in human brain structure, function, and gene expression has been documented [44]. Previous studies also described substantial lateralization in specific molecular subtypes of gliomas [9, 10]. A study with a larger population investigating the association of tumor lateralization and amino acid tracer uptake is required. The FDOPA uptake in the normal brain structures may vary in relation to the patient age [45], which may have affected the stratification of gliomas. Because of retrospective nature the acquisition parameters and scanners of MRI could not be exactly matched across patients, and the Karnofsky performance status were not obtained from all subjects.

Conclusion

Radiographic atlas analysis revealed that FDOPA hypometabolic LGGs preferentially localized in the frontal lobe. The frontal lobe LGGs in functionally eloquent areas may be detected at a relatively early phase due to symptomatic episodes, which may be associated with tumor hypometabolism, smaller volume, and younger patient age.

Declarations

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Conflicts of interest

Ellingson—Advisory Board—Hoffman La-Roche; Siemens; Nativis; Medicenna; MedQIA; Bristol-Myers Squibb; Imaging Endpoints; Agios. Paid Consultant—Nativis; MedQIA; Siemens; Hoffman La-Roche; Imaging Endpoints; Medicenna; Agios. Grant Funding—Hoffman La-Roche; Siemens; Agios; Janssen. Ellingson also holds a patent on this technology (US Patent #15/577,664; International PCT/US2016/034886). Cloughesy—Advisory Board—Roche/ Genentech, Amgen, Tocagen, NewGen, LPath, Proximagen, Celgene, Vascular Biogenics Ltd, Insys, Agios, Cortice Bioscience, Pfizer, Human Longevity, BMS, Merck, Notable Lab, MedQIA.

Ethics approval/Consent to participate/Consent for publication
All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards. Written informed consent was obtained from all individual participants to have their imaging, clinical, and molecular data included in our research database (IRB IRB#10-000655).

**Availability of data and material**

Not applicable

**Code availability**

Not applicable

**Author contribution:**

Hiroyuki Tatekawa: study design, data analysis, drafting the manuscript, approving the final content

Hiroyuki Uetani: study design, data analysis, revising the manuscript, approving the final content

Akifumi Hagiwara: study design, data analysis, revising the manuscript, approving the final content

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Noriko Salamon: study design, revising the manuscript, approving the final content

Benjamin M. Ellingson: study design, data analysis, revising the manuscript, approving the final content
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