Effects of ivabradine hydrochloride combined with trimetazidine on myocardial fibrosis in rats with chronic heart failure

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Abstract. Effects of ivabradine hydrochloride (Iva) and trimetazidine on myocardial fibrosis (MF) in rats with chronic heart failure (CHF) were explored. Fifty Wistar rats were randomly divided into sham operation, model, Iva, trimetazidine and combined drug group with 10 rats each. All rats except those in sham operation group were subjected to establish CHF model by constricting the abdominal aorta. After successful modeling, rats in the sham operation and model group received normal saline (10 mg/kg) gavage daily, the Iva group received Iva (10 mg/kg) gavage, the trimetazidine group received trimetazidine (10 mg/kg) gavage, and the combined drug group were given Iva (10 mg/kg) and trimetazidine (10 mg/kg) gavage for 12 weeks. The changes of hemodynamic indexes and heart rate, connective tissue growth factor (CTGF) and superoxide dismutase (SOD) levels as well as transforming growth factor β1 (TGF-β1) and collagen I (COL-I) expression levels in myocardial tissue of each group were detected. Compared with sham operation group, the left ventricular end-diastolic pressure (LVEDP) level, CTGF expression, TGF-β1 mRNA and COL-I mRNA expression levels in model group increased significantly, but the ± dp/dt_{max} and the SOD content in myocardial tissue decreased significantly. Compared with model group, the LVEDP level, CTGF expression, TGF-β1 mRNA and COL-I mRNA expression levels in Iva group, trimetazidine group and combined drugs decreased significantly, but the ± dp/dt_{max} and the SOD content in myocardial tissue increased significantly (P<0.05). Changes in the combined drug group were the most notable (P<0.05). Iva combined with trimetazidine reduces LVEDP in rat with CHF, increases SOD content, and inhibits CTGF expression and TGF-β1 and COL-I expression levels in myocardial tissues, thus achieving the inhibitory effect on MF.

Introduction

Heart failure is a clinical syndrome with pathophysiological changes in heart caused by impaired ventricular diastolic or systolic function due to damaged cardiac function or abnormal internal structure of heart (1). Chronic heart failure (CHF), as one of the leading causes of cardiovascular disease death, has a serious impact on health and quality of life of patients (2). Studies have shown that myocardial fibrosis (MF) is the most important pathological basis of CHF (3). Other studies (4,5) show that MF actually refers to a process in which a large amount of collagen fibers in the heart matrix aggregate or collagen compositions change.

Ivabradine hydrochloride (Iva), a highly specific If channel drug with controlling effect on sinus heart rate (6), also delay heart failure and improves heart function (7). Due to its unique dual action mechanism, Iva can control spontaneous diastolic depolarization in sinoatrial node and regulate heart rate by selectively and specifically inhibiting cardiac pacing If current, providing a new therapeutic idea for heart failure (8). Trimetazidine, as a myocardial metabolic drug, is also widely used in cardiovascular therapy due to its anti-oxidation and anti-ischemia effects (9). It has been shown that trimetazidine has inhibitory effects on the activities of myocardial fatty acids and oxidation-related enzymes, which is conducive in increasing myocardial productivity and thus improving ventricular function (10). Although these two drugs are currently used in CHF, studies on their effects on MF are still rare. Therefore, CHF rat models were established and the effects of Iva and trimetazidine on MF were observed in this
study in order to provide more theoretical data for the treatment of CHF.

Materials and methods

Animals and experimental materials. A total of 50 Wistar male rats weighing 220.51±10.24 g were selected and purchased from Shanghai Slaccas Experimental Animal Co., Ltd., with a production license of SCXK (Shanghai) 2012-0002, fed at a constant temperature of 22°C with normal circadian rhythm and free diet. Iva was purchased from Servier Laboratories. Trimetazidine was purchased from Beijing Wansheng Pharmaceutical Co., Ltd. Sodium pentobarbital was purchased from Hubei Hongyun Long Biological Technology Co. Ltd. Connective tissue growth factor (CTGF) polyclonal antibody was purchased from Invitrogen; Thermo Fisher Scientific, Inc. Superoxide dismutase (SOD) kit was purchased from Dojindo Molecular Technologies, Inc. Primary mouse anti-rat CTGF (cat. no. AMAB91366; dil, 1:1,000) and β-actin monoclonal antibodies (cat. no. A1978; dil, 1:400), secondary rabbit anti-mouse polyclonal antibody (cat. no. SAB3701212; dil, 1:100) used in western blot analysis were purchased from Sigma-Aldrich; Merck KGaA. BCA quantitative kit was purchased from Beijing Solarbio Science & Technology Co., Ltd. RT-qPCR kit and minScript reverse transcription kit were purchased from Takara Biotechnology Co., Ltd. Biological signal recorder system was purchased from Anhui Zhenghua Biological Instrument Equipment Co., Ltd.

The study was approved by the Ethics Committee of The Affiliated Hospital of Jining Medical University (Jining, China).

Establishment and grouping of animal models. Ten rats were randomly selected as sham operation group, and the other rats were all used to construct CHF rat model by constricting the abdominal aorta. The specific steps were: All rats were fasted for more than 12 h before surgery, then 3% pentobarbital sodium solution was prepared and injected intraperitoneally (0.15 ml/100 g) to anesthetize the rats. After anesthesia, a 2.5 cm long incision was made in the abdominal skin along the anterior midline at the costal arch, then the left renal artery was freed from the abdominal aorta at the upper part by the anterior midline at the costal arch, then the left renal artery was freed from the abdominal aorta at the upper part by

Table I. Primer sequences.

| Factor          | Upstream primer | Downstream primer          |
|-----------------|-----------------|----------------------------|
| TGF-β1          | 5'-TGACCGCAACAACGCAATCTA-3' | 5'-CACCTCGAGTTTGGGACTGATC-3' |
| COL-I           | 5'-ATGCCTGGTTGGGAGAGCA-3'   | 5'-GAGGAGCAGGACCTTGTGAG-3'   |
| β-actin         | 5'-GAGAGGAAATCGTGCGTGAC-3'  | 5'-CATCTGCTGGAGGCTGCA-3'     |

of continuous feeding, 10 CHF rats were randomly selected for hemodynamic detection. Left ventricular end-diastolic pressure (LVEDP) ≥15 mmHg indicated the modeling was successful. Forty rats with successful modeling were randomly divided into model, Iva, trimetazidine and combined drug group (Iva + trimetazidine) with 10 rats each. Rats in the sham operation and model group were given 10 mg/kg of normal saline daily, rats in the Iva group were given 10 mg/kg of Iva daily, rats in the trimetazidine group were given 10 mg/kg of trimetazidine daily, and rats in the combined drug group were given Iva (10 mg/kg) and trimetazidine (10 mg/kg) daily for 12 weeks of continuous treatment. The changes of hemodynamic indexes, heart rate, CTGF and SOD levels as well as transforming growth factor β1 (TGF-β1) and collagen I (COL-I) expression levels in myocardial tissue in each group were detected.

Index detection method

Hemodynamic index detection. After the treatment, the right common carotid artery was retrograde intubated to the left ventricle, and then the other end was connected to the pressure transducer containing multimedia biological signal recorder system in order to record maximum rising and decreasing rate of the left ventricular end-diastolic pressure (LVEDP) and the left ventricular pressure (∆dp/∆t_{max}).

Detection of SOD and CTGF. The myocardial tissue was homogenized by a high-speed homogenizer, then the expression of SOD was detected using enzyme-linked immunosorbent assay (ELISA), and the specific operation strictly followed the kit instructions. The content of CTGF in the myocardial tissue was detected by western blot analysis. The specific method was as follows: Total protein in the myocardial tissue was extracted, and separated with 10% SDS-PAGE, then transferred to PVDF membrane. The membrane was blocked with 5% skimmed milk at room temperature for 1 h, then incubated overnight at 4°C with primary mouse anti-rat CTGF (cat. no. AMAB91366; dil, 1:1,000) and β-actin monoclonal antibodies (cat. no. A1978; dil, 1:400) both from Sigma-Aldrich; Merck KGaA and incubated at 37°C for 1 h with secondary rabbit anti-mouse polyclonal antibody (cat. no. SAB3701212; dil, 1:100; Sigma-Aldrich; Merck KGaA and incubated at 37°C for 1 h with secondary rabbit anti-mouse polyclonal antibody (cat. no. SAB3701212; dil, 1:100; Sigma-Aldrich; Merck KGaA) and secondary goat anti-rabbit polyclonal antibody (cat. no. SAB3701212; dil, 1:100; Sigma-Aldrich; Merck KGaA). The protein bands on the membrane were developed with DAB developer.

Detection of TGF-β1 mRNA and COL-I mRNA expression by RT-qPCR. The myocardial tissue of rats was cut and then added with TRIzol reagent to extract total RNAs. The purity and concentration of RNAs were detected by an ultraviolet spectrophotometer. According to the instructions of reverse transcription kit, 1 µg of total RNA was reverse transcribed to cDNA, with reaction parameters of 30°C for 10 min, 42°C
for 30 min and 95˚C for 5 min. The transcribed cDNA was used for qPCR amplification, and the system was as follows: 10 µl of Taqman PCR Master Mix Ⅱ, 0.25 µl of Takara Ex Taq, 0.5 µl of each upstream and downstream primer, ddH2O complemented to 20 µl.

β-actin was used as an internal reference, and the primer sequences are shown in Table Ⅰ.

qPCR reaction conditions: Pre-denaturation at 94˚C for 2 min, then 95˚C for 30 sec and 60˚C for 30 sec for 40 circles, then extension at 72˚C for 1 min. Real-time quantitative PCR detection was carried out with qPCR instrument, and the experiment was repeated 3 times. The results were analyzed using the 2^(-ΔΔCq) method (11).

Statistical analysis. SPSS 19.0 (IBM Corp., Armonk, NY, USA) was used to analyze the experimental data. The measurement data were expressed as mean ± standard deviation (SD). One-way analysis of variance (ANOVA) was used to compare the differences among groups, and the Dunnett’s t-test was used for the following pairwise comparison. The difference was statistically significant with P<0.05.

Results

Expression of hemodynamic indexes in each group. Compared with the sham operation group, the LVEDP in the model group increased significantly, while the ±dp/dt_max decreased significantly (P<0.05). Whereas, compared with the model group, the LVEDP in the Iva, trimetazidine and combined drug group decreased significantly, while the ±dp/dt_max increased significantly (P<0.05). Changes in the combined drug group were the most notable (P<0.05), and the indexes in the Iva and trimetazidine group had no significant difference (P>0.05) (Table Ⅱ).

Expression of CTGF and SOD in myocardial tissue of each group. Compared with the sham operation group, SOD content decreased and CTGF expression increased in myocardial tissue of the model group. While compared with the model group, the content increased in myocardial tissue of the Iva, trimetazidine and combined drug group (P<0.01). The changes in the combined drug group were the most notable (P<0.05), and the expression of SOD in the Iva group and trimetazidine group had no significant difference (P>0.05). CTGF, connective tissue growth factor.

Expression of CTGF and SOD in myocardial tissue of each group. Western blot analysis showed that compared with the sham operation group, CTGF expression increased in myocardial tissue of the model group. Compared with the model group, the expression decreased in myocardial tissue of the Iva, trimetazidine and combined drug group (P<0.01). The changes in the combined drug group were the most notable (P<0.05), and the expression of CTGF in the Iva and trimetazidine group had no significant difference (P>0.05). CTGF, connective tissue growth factor.

Expression levels of TGF-β1 and COL-I in myocardial tissue of each group. Compared with the sham operation group, the expression levels of TGF-β1 mRNA and COL-I mRNA in myocardial tissue of model group increased, while compared with the model group, the expression levels decreased in myocardial tissues of the Iva, trimetazidine and combined drug group (P<0.05). Changes in the combined drug group
**Table III. Expression of CTGF and SOD in myocardial tissue of each group.**

| Index     | Sham operation group (n=10) | Model group (n=10) | Iva group (n=10) | Trimetazidine group (n=10) | Combined drug group (n=10) | F value | P-value |
|-----------|-----------------------------|--------------------|------------------|---------------------------|---------------------------|---------|---------|
| CTGF      | 0.162±0.023^a               | 0.488±0.022        | 0.276±0.025^b,a   | 0.281±0.031^b,a          | 0.174±0.026^a             | 261.0   | <0.001  |
| SOD (U/g) | 189.3±11.71^a               | 102.6±14.23        | 141.8±15.31^b,a   | 139.7±14.52^b,a          | 185.6±11.39^a             | 71.30   | <0.001  |

^aP<0.05, compared with model group; ^bP<0.05, compared with drug group. CTGF, connective tissue growth factor; SOD, superoxide dismutase.

**Table IV. Expression of TGF-β1 mRNA and COL-I mRNA in myocardial tissue of each group.**

| Index       | Sham operation group (n=10) | Model group (n=10) | Iva group (n=10) | Trimetazidine group (n=10) | Combined drug group (n=10) | F value | P-value |
|-------------|-----------------------------|--------------------|------------------|---------------------------|---------------------------|---------|---------|
| TGF-β1 mRNA | 0.369±0.038^a               | 0.875±0.046        | 0.631±0.039^b,a   | 0.657±0.035^b,a          | 0.439±0.042^a             | 328.4   | <0.001  |
| COL-I mRNA  | 0.635±0.051^a               | 1.211±0.049        | 0.812±0.055^b,a   | 0.826±0.048^b,a          | 0.723±0.056^a             | 178.5   | <0.001  |

^aP<0.05, compared with model group; ^bP<0.05 compared with drug group. TGF-β1, transforming growth factor-β1; COL-I, collagen I.

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**Discussion**

The high incidence of hypertension, diabetes and coronary heart diseases caused by accelerated aging of the population and the improvement of living standards leads to a rising number of heart failure patients (12). In recent years, diuretics, β receptor blockers are mostly used in clinical treatment of heart failure but achieving unsatisfactory efficacy and still high mortality rates (13,14). A study pointed out that the decisive mechanism of CHF was ventricular remodeling, of which MF is the most significant (15). MF is reported to be an important pathological basis of heart failure, mainly referring to the remodeling of intermyocardial collagen network (ICN) (16). In this study, the CHF rat models were prepared by constricting the abdominal aorta, and the effects of Iva and trimetazidine alone and in combination on MF in CHF rats were explored.

LVEDP is often used to express cardiac volume load clinically. Some studies show that its increase leads to increased mitochondria and decreased adenosine triphosphate (ATP) in cardiac myocytes, promotes the synthesis of collagen fibers and eventually accelerates the process of MF in CHF rats (16,17). Our experimental results showed that, compared with the sham operation group, the LVEDP in the model...
group increased significantly while the ±dp/dt max decreased significantly (P<0.05), which indicated that the model group rats developed MF. Compared with the model group, the LVEDP in the Iva group, trimetazidine group and combined drug group decreased significantly while the ±dp/dt max increased significantly (P<0.05). Moreover, the changes in the combined drug group were the most notable (P<0.05). The results suggested that both Iva and trimetazidine improved MF in rats, and the combined medication achieved a more obvious improvement, which was suspected to be related to the maximum rising and decreasing rate of left ventricular pressure in rats. A previous study (18) also reached the same conclusion as ours when discussing the effect of trimetazidine on MF in CHF rats, but there is no relevant study to explain it for the time being. SOD is an important antioxidant enzyme and a major substance for eliminating free radicals in vivo. CTGF is a factor that can induce fibroblast proliferation with obvious mitogen and chemotaxis. Both SOD and CTGF are important factors for evaluating MF (19,20). Our results showed that, compared with the sham operation group, SOD content decreased and CTGF expression increased in myocardial tissue of the model group. Compared with the model group, SOD content increased and CTGF expression level decreased in myocardial tissue of the Iva group, trimetazidine group and combined drug group (P<0.05), but the changes in the combined drug group were the most notable (P<0.05). The results suggested that Iva and trimetazidine affected MF process through CTGF and SOD in myocardial tissue of CHF rats. A study (21) has shown that Iva could inhibit the oxidative stress reaction of myocardial cells in CHF rats, and another study (22) also showed that trimetazidine, as a new anti-myocardial ischemia drug, also had the function of oxygen free radical inhibition, oxidation resistance and apoptosis, which partially explained the changes in SOD expression in our conclusion. It was considered that (23) TGF-β1, a multifunctional regulatory factor, could affect MF through Smads-dependent and Smads-independent pathways. There is also a study (24) showing that excessive synthesis and deposition of COL-I promoted MF in patients with hypertension and heart failure. Therefore, the expression levels of TGF-β1 mRNA and COL-I mRNA in myocardial tissues were also detected in this study. The results showed that compared with the sham operation group, the expression levels of both TGF-β1 mRNA and COL-I mRNA in myocardial tissues of the model group were increased. Compared with the model group, the expression levels decreased in myocardial tissues of the Iva group, trimetazidine group and combined drug group (P<0.05), and changes in the combined drug group were the most notable (P<0.05). The results suggested that both Iva and trimetazidine could inhibit the expression levels of TGF-β1 and COL-I, and the inhibition effect was most pronounced when used in combination. The above results all suggest that Iva and trimetazidine can effectively inhibit MF in CHF rats, and the effect of combined administration is greater than that of the single drug. However, at present, there is no related research on the synergistic effect of the two drugs through different mechanisms.

Collectively, Iva combined with trimetazidine can inhibit MF in CHF rats by regulating LVEDP, SOD content and CTGF expression in myocardial tissue, and by inhibiting TGF-β1 mRNA and COL-I expression, and the inhibition effect is most pronounced when used in combination. However, in our study, the specific mechanism of Iva and trimetazidine in regulating MF and the possible toxic and side effects of combined medication were not fully addressed, thus further studies are anticipated.

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Availability of data and materials

The datasets used and/or analyzed during the present study are available from the corresponding author on reasonable request.

Authors’ contributions

DM wrote the manuscript. TX, GC and XW performed qPCR and ELISA. ZL and XL were responsible for western blot analysis. JL and NY contributed to analysis of observation indexes. All the authors read and approved the final manuscript.

Ethics approval and consent to participate

The study was approved by the Ethics Committee of The Affiliated Hospital of Jining Medical University (Jining, China).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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