Heparin improves organ microcirculatory disturbances in caerulein-induced acute pancreatitis in rats

Marek Dobosz, Lucjanna Mionskowska, Stanislaw Haæ, Sebastian Dobrowolski, Dariusz Dymecki, Zdzislaw Wajda

INTRODUCTION
Pathophysiology of acute pancreatitis (AP) is heterogeneous and involves a complex cascade of events. Impaired tissue perfusion within the pancreatic gland is one of the early disorders accompanying the disease[1,2]. Prolonged ischaemia and impaired tissue oxygenation, resulting from vasoconstriction, hemoconcentration, hypercoagulation, leukocyte adherence, increased vascular permeability, and interstitial oedema, play an important role in the progression of the disease, leading to the development of pancreatic necrosis[2-4]. Besides the pancreas, the microcirculatory disturbances, as a consequence of hypovolemia and systemic inflammatory response, are also observed in other organs[5,6].

Although the hypovolemia and impairment of cardiac output can be normalized easily by vigorous fluid replacement, microcirculatory disturbances may be prevented merely by adequate fluid therapy, even though the cardiovascular parameters are stabilized at the baseline level[9]. Various vasoactive mediators, as bradykinin, endothelin, thromboxan, platelet activating factor, and nitric oxide participate in the development of microcirculatory failure[10,11]. In the last decade, the beneficial effect of therapeutic strategies in acute pancreatitis, affecting vasoactive mediators, is confirmed in several experimental studies.

One of the possible therapeutic agents, influencing microcirculatory disturbances by means of its anticoagulant and antiinflammatory properties, which could be applied in acute pancreatitis, is heparin. Some reports underlined beneficial impact of this well known polysaccharide on the course of the disease[11-13]. In this study we examined whether heparin influenced splanchic malperfusion in experimental caerulein-induced acute pancreatitis in rats.

MATERIALS AND METHODS
Animal models
The study was carried out on 34 male Wistar rats weighing 180-200 g. The rats were kept on a standard rat chow and fasted overnight before the experiment with water ad libitum. Acute pancreatitis was induced by four intraperitoneal injections of caerulein (Cn) (Sigma, St. Louis, USA) (15 µg/kg) in 1 mL of saline at 1h intervals: at the beginning, and consecutively after the first, second, and third hour of the experiment. Five hours after the first caerulein injection, rats were anaesthetized with pentobarbital sodium (40 mg/kg). Following the anaesthesia, a laparotomy was performed, and a fiberoptic probe of laser Doppler flowmeter (Periflux 4 001, Perimed Jarfalla, Sweden) was positioned against the surface of the pancreas, liver, kidney, stomach, colon, and skeletal muscle of thigh in order to estimate the organs’ perfusion. Blood flow was measured in three different portions of each organ, the mean values were calculated, and presented as percent changes from basal values obtained in control rats (100%). After the measurements, blood was aspirated from the inferior caval vein for hematocrit and interleukin 6 tests, the pancreas was removed for microscopic evaluation, and the animals were exsanguinated.

The animals were randomly allocated into three groups: Group I (n=10), control; Group II (n=12), Cn-induced pancreatitis without treatment; and Group III (n=12), Cn-induced pancreatitis treated with heparin (Polfa, Warszawa, Poland) 2×2.5 mg/kg, given in the 3rd and 4th after the first Cn injection.

This study was approved by the Ethical Committee of the Medical University of Gdañsk.

Interleukin-6 functional assay
The IL-6 bioassay was performed using the IL-6 dependent
Control rats. The heparin administration significantly diminished Cn-induced acute pancreatitis caused a significant increase of serum interleukin 6 assay.

**Statistical analysis**

Data are presented as mean±SD. The differences between the groups were analysed by means of ANOVA test. Probability values less than 0.05 were considered significant.

**RESULTS**

Four intraperitoneal caerulein injections resulted in marked pancreatic oedema in all the rats. The microscopic examination revealed oedematous form of acute pancreatitis, with inter-and intralobular oedema, vacuolisation of parenchymal cells, and leukocyte infiltration within the pancreatic gland. No parenchymal necrosis was detected.

**Microcirculatory values**

Caerulein induced acute pancreatitis resulted in a significant drop of pancreatic microperfusion to 36.9±4% of basal values. Administration of heparin significantly improved the microcirculation of the pancreas up to 75.9±13%. Hepatic perfusion in rats with pancreatitis receiving no treatment was decreased to 56.6±6%. Heparin injection raised this parameter to 75.2±16%. Renal blood flow in group II with pancreatitis was diminished to 45.1±6%, which was improved significantly with heparin treatment up to 79.3±15%. Microcirculatory values of stomach and colon wall in acute pancreatitis group were reduced to 65.2±8% and 69.8±6% respectively. Intraperitoneal injection of heparin caused improvement in the blood flow in stomach to 78.1±19%, and especially in colon to 102.5±19%. Similarly, skeletal muscle perfusion in rats with pancreatitis was significantly ameliorated after heparin administration from 59.2±3% to 77.9±13% (Figure 1).

**Serum interleukin 6 assay**

Cn-induced acute pancreatitis caused a significant increase of serum IL-6 activity from 37.7±21 U/mL up to 359.0±66 U/mL in control rats. The heparin administration significantly diminished IL-6 level to 288.5±58 U/mL (Table 1).

**Hematocrit measurements**

The mean hematocrit level of the control group without pancreatitis was 41±3%. In group II with Cn-induced pancreatitis the hematocrit was significantly increased to 53±4%. Heparin administration in animals with pancreatitis of group III significantly reduced the hematocrit to 46±3% (Table 1).

| Group                                      | Interleukin 6 (U/mL) | Hematocrit (%) |
|--------------------------------------------|----------------------|----------------|
| Control                                    | 37.7±21              | 41±3           |
| Acute pancreatitis                         | 359.0±66             | 53±4           |
| Acute pancreatitis + heparin               | 288.5±58             | 46±3           |

*p<0.05 vs control, †p<0.05 vs acute pancreatitis group.

**DISCUSSION**

Caerulein-induced experimental pancreatitis was adopted by many authors to investigate microcirculatory changes of AP. The present study indicated that in Cn-induced acute pancreatitis the microperfusion of the pancreas decreased to 36.9% of basal values. These observations are in accordance with other investigations, which revealed the drop of local pancreatic blood flow in Cn induced pancreatitis[2,13,14], although a majority of microcirculatory studies did not report such an obvious deterioration of the pancreatic perfusion as observed in the current study. Such a sharp decline of the pancreatic microperfusion might be influenced by the fact that the animals did not receive i.v. fluid administration, and could be elucidated also by an additional stress resulted from six i.p. injections of the drugs in conscious animals. In Liu’s study[16], the water immersion stress in rats with Cn-induced pancreatitis caused further pancreatic ischemia, diminishing the pancreatic blood flow to 37% of the normal value, and was considered as an aggravating factor in AP.

In contrast to these observations, the Heidelberg group described increased pancreatic capillary flow in experimental oedematous pancreatitis[15,16]. The discrepancy of the results may be explained by various techniques of microcirculatory measurements and i.v. fluid administration. On the other hand, in the model of necrotizing pancreatitis, the authors observed a significant decrease of the pancreatic capillary flow[16].

The disturbances of microcirculation in acute pancreatitis were not only confined to pancreatic capillary bed but were also observed in other organs[3-5]. It was suggested that diffuse microcirculatory disorders might play a crucial role in the development of the pancreatitis-associated multiorgan dysfunction syndrome[7,12], some authors even defined severe AP as a systemic dysfunction syndrome[7,12].

The present study confirmed these data. Besides the pancreas, reduced capillary perfusion was observed in liver, kidney, stomach, colon, and skeletal muscles, however the drop in perfusion of other organs was not so remarkable as in the pancreas. This suggests that in pancreatitis, pancreatic gland is especially susceptible to microcirculatory disorders. Kinmal et al. found that splanchic malperfusion seemed to begin with pancreatic hypoperfusion before disturbances in gut microcirculation[11]. On the other hand, Hotz et al. noted that in mild pancreatitis, pancreatic capillary perfusion remained unchanged, whereas mucosal and subserosal colonic capillary blood flow was significantly reduced. It was also shown in their experiment that severe pancreatitis was associated with a marked reduction in both pancreatic and colonic capillary perfusion[11].

Microcirculatory disturbances in AP comprised many
components: decreased capillary blood flow and capillary density, increased capillary permeability, and enhanced leukocyte-endothelial interaction[3]. It still remains unclear which of these factors is the initiating one or the most important. It seems to be logical that any effort to improve the microcirculation may be beneficial for all organs, irrespective of the underlying initiating factors.

Many studies documented positive impacts of various treatments on improving microcirculation in acute pancreatitis. In studies by Konturek et al. and Liu et al. pancreatic microperfusion was ameliorated with L-arginine (nitric oxide donor) treatment[13,14]. Improvement of impaired pancreatic microcirculation by isovolumic hemodilution with dextran was observed by Klar et al.[19]. Rheologic properties of pentoxifylline, its antiinflammatory effect, reduction of thrombus formation, and lowering of blood viscosity had a positive influence on Cn-induced pancreatitis in study of Gomez-Cambreron et al.[20]. An advantageous effect of ICAM-1 antibodies on microcirculation in AP was shown by Foitzik et al.[21], and Werner et al.[22]. A single i.v. injection of bovine hemoglobin in an experiment by Strate et al. reduced microcirculatory dysfunction and pancreatic tissue damage in rodent pancreatitis[23]. In a study by Zhao et al. Chinese natural medicines: Yuanhu (YHI) and Huoxuehuayu (HHI-1) had a positive impact on acute pancreatitis development, including pancreatic malperfusion and serum IL-6 level. Recently, it was shown that endothelin receptor blockade might be a promising therapeutic approach in AP, resulting in improvement of microcirculation, decreased capillary permeability and tissue injury[2,23,24].

Intraperitoneal heparin administration in the current study significantly improved blood perfusion of examined organs. The beneficial effect of heparin on pancreatic microcirculation may in theory have an influence on severity of the disease, it could prevent or restrict the development of pancreatic necrosis. It was shown in our recent publication that heparin treatment in oedematous pancreatitis ameliorated the microscopic alterations of the pancreas: reduced leukocyte infiltration and diminished pancreatic oedema[25].

By improving hepatic capillary blood flow, heparin could influence phagocytic Kupffer cell function in the liver which is important for eliminating antigens and toxic substances in acute pancreatitis. The study of Forcacs et al. showed a close relationship between liver phagocytic function and hepatic microperfusion in the early stage of experimental pancreatitis[26]. On the other hand, Chen et al. revealed that a significant number of adherent leukocytes was observed in hepatic microcirculation 2 h after the induction of AP[27]. This could impair hepatic perfusion, and might be a potential target for heparin treatment. In another study, Chen et al. observed beneficial effects of protease inhibitor gabexate mesilate on pancreatic and hepatic microcirculation[28].

The pathophysiology of renal insufficiency, often observed as a complication of acute pancreatitis, is heterogeneous. The improvement of renal blood flow observed after heparin treatment in the rats with pancreatitis could also prevent this complication. Foitzik et al. using endothelin receptor blockers in severe acute pancreatitis, besides the enhancement of pancreatic perfusion, observed improved renal function[7]. It was shown that decreased capillary blood flow in the colonic mucosa was associated with impaired gut barrier function and increased translocation of live bacteria through the morphologically intact colonic wall[29]. The present study revealed that heparin treatment in the group with pancreatitis led the altered microperfusion of the colonic wall to the normal level. It suggests that heparin administration may play a role in prevention of secondary pancreatic infection.

Besides the organ microcirculatory improvement, heparin administration significantly decreased interleukin 6 activity, and reduced hematocrit level. IL-6 was a reliable predictor of acute pancreatitis severity[29]. Diminished IL-6 activity in the group with AP treated with heparin may indicate that this therapy could limit inflammatory response observed in AP. Beneficial influence of heparin on hematocrit level, observed in group III, may also suggest that heparin reduces capillary permeability, not just in the pancreatic gland, and prevent the fluid to the third space. Therefore heparin therapy may reduce the need for vigorous fluid resuscitation as applied in patients with AP.

The improvement of systemic microperfusion after heparin administration in rats with AP might be explained by its anticoagulant effect which diminished blood viscosity and facilitated capillary blood flow. Besides its anticoagulant properties, heparin may play a positive role in AP treatment due to its antiinflammatory effects: TNF inhibition[30], protection from oxygen free radicals[31], the blockade of complement activity[22] and histamine release[32]. It was also shown that heparin prevented post-ischemic endothelial cell dysfunction[33], neutrophil adhesion[34], and had an inhibitory effect on the biosynthesis and release of endothelin-1, a very potent endogenous vasoconstrictor[35,36].

In conclusion, heparin counteracts microcirculatory disorders, not only within the pancreas but also in other organs. The preservation of organ microperfusion with this treatment may be significant for diminishing and treating local and systemic complications in AP.

REFERENCES

1. Kinnala PJ, Kuttila KT, Gronroos JM,HAVia TV, Nevalainen TJ, Niinikoski JH. Splanchnic and pancreatic tissue perfusion in experimental acute pancreatitis. Scand J Gastroenterol 2002; 37: 845-849
2. Zhou ZG, Chen YD, Sun W, Chen Z. Pancreatic microcirculatory impairment in experimental acute pancreatitis in rats. World J Gastroenterol 2002; 8: 933-936
3. Zhou ZG, Chen YD. Influening factors of pancreatic microcirculatory impairment in acute pancreatitis. World J Gastroenterol 2002; 8: 406-412
4. Strate T, Mann O, Kleinhaus H, Schneider C, Knoefel WT, Yekebas E, Standl T, Bleechle C, Izicki JR. Systemic intravascular infusion of bovine hemoglobin significantly reduces microcirculatory dysfunction in experimentally induced pancreatitis in the rat. Ann Surg 2003; 238: 765-771
5. Foitzik T, Eibl G, Hotz B, Hotz H, Kahrau S, Kasten C, Schneider P, Buhr HJ. Persistent multiple organ microcirculatory disorders in severe acute pancreatitis: experimental findings and clinical implications. Dig Dis Sci 2002; 47: 130-138
6. Skoromnyi AN, Starosek VN. Hemodynamic changes in the liver, kidney, small intestine and pancreas in experimental acute pancreatitis. Klin Khir 1998; 12: 46-48
7. Foitzik T, Eibl G, Hotz HG, Faulhaber J, Kirchengast M, Buhr HJ. Endothelin receptor blockade in severe acute pancreatitis leads to systemic enhancement of microcirculation, stabilization of capillary permeability, and improved survival rates. Surgery 2000; 128: 399-407
8. Chen HM, Hwang TL, Chen MF. The effect of gabexate mesilate on pancreatic and hepatic microcirculation in acute experimental pancreatitis in rats. J Surg Res 1996; 66: 147-153
9. Knoll JA, Imman MG, Strodel WE, Eckhauser FE. Pancreatic response to crystalloid resuscitation in experimental pancreatitis. J Surg Res 1987; 43: 387-392
10. Gabryelewicz A, Kosidlo S, Prokopowicz J, Podkowicz K. Does heparin modify protease-antiprotease balance in acute experimental pancreatitis in rats. Hepatogastroenterology 1986; 33: 79-82
11. Gibrilbourne IA, Watson H, Davies GC. 111In-platelet and 125I-fibrinogen deposition in the lungs in experimental acute pancreatitis. J Surg Res 1987; 43: 521-526
12. Rabenstein T, Roggenbuck S, Framke B, Martin M, Fischer B, Nusko G, Muehlendorf S, Hochberger J, Eil C, Hahn EG, Schneider HT. Complications of endoscopic sphincterotomy:
Can heparin prevent acute pancreatitis after ERCP? Gastrointest Endosc 2002; 55: 476-483

Konturek SJ, Szlachcic A, Dembinski A, Warzocha Z, Jaworek J, Stachura J. Nitric oxide in pancreatic secretion and hormone-induced pancreatitis in rats. Int J Pancreatol 1994; 15: 19-28

Liu X, Nakano I, Yamaguchi H, Ito T, Goto M, Koyanagi S, Kinjoh M, Nawata H. Protective effect of nitric oxide on development of acute pancreatitis in rats. Dig Dis Sci 1995; 40: 2162-2169

Klar E, Schraut W, Foitzik T, Buhr H, Herfarth C, Messmer K. Impact of microcirculatory flow pattern changes on the development of acute edematous and necrotizing pancreatitis in rabbit pancreas. Dig Dis Sci 1994; 39: 2639-2644

Schmidt J, Ebeling D, Ryschik E, Werner J, Gebhard MM, Klär E. Pancreatic capillary blood flow in an improved model of necrotizing pancreatitis in the rat. Surg Res 2002; 106: 335-341

Gullo A, Berlot G. Ingredients of organ dysfunction or failure. World J Surg 1996; 20: 430-436

Hotz HG, Foitzik T, Rohweder J, Schulze JD, Fromm M, Runkel NS, Buhr HJ. Intestinal microcirculation and gut permeability in acute pancreatitis: early changes and therapeutic implications. J Gastrointest Surg 1998; 2: 518-525

Klar E, Mall G, Messmer K, Herfarth C, Rattner DW, Warshaw AL. Improvement of impaired pancreatic microcirculation by isovolemic hemodilution protects pancreatic morphology in acute biliary pancreatitis. Surg Gynecol Obstet 1993; 176: 144-150

Gomez-Cambronero L, Camps B, de La Asuncion JG, Cerda M, Pellin A, Pallardo FV, Calvette J, Sweery JH, Mann GE, Vina J, Sastre J. Reduction of local and systemic complications of acute pancreatitis by monoclonal antibody to ICAM-1. Arch Chir Suppl Kongressbd 1998; 115(Suppl I): 427-429

Plusczynk T, Witzel B, Menger MD, Schilling M. ETA and ETB receptor function in pancreatitis-associated microcirculatory failure, inflammation, and parenchymal injury. Am J Physiol Gastrointest Liver Physiol 2003; 285: G145-153

Dobosz M, Wajda Z, Hac S, Myśliwski J, Bryl E, Mionskowska L, Roszkiewicz A, Myśliwski A. Nitric oxide, heparin and proacine treatment in experimental ceruline-induced acute pancreatitis in rats. Arch Immunol Ther Exp 1999; 47: 155-160

Forgacs B, Eibl G, Wudel E, Franke J, Faulhaber J, Kahrau S, Buhr HJ, Foitzik T. RES function and liver microcirculation in the early stage of acute experimental pancreatitis. Hepatogastroenterology 2003; 50: 861-866

Chen HM, Sunamura M, Shibuya K, Yamauchi JI, Sakai Y, Fukuyama S, Mikami Y, Takeda K, Matsuo S. Early microcirculatory derangement in mild and severe pancreatitis models in mice. Surg Today 2001; 31: 634-642

Foitzik T, Stufler M, Hotz HG, Kliinert J, Wagner J, Warshaw AL, Schulzke JD, Fromm M, Buhr HJ. Glutamine stabilizes intestinal permeability and reduces pancreatic infection in acute experimental pancreatitis. J Gastrointest Surg 1997; 1: 40-47

Bhatia M, Brady M, Shokuhi S, Christmas S, Neoptolemos JP, Slavin J. Inflammatory mediators in acute pancreatitis. J Pathol 2000; 199: 117-125

Lantz M, Thysell H, Nilsson E, Olsson I. On the binding of tumor necrosis factor (TNF) to heparin and the release in vivo of the TNF-binding protein I by heparin. J Clin Invest 1991; 88: 2026-2031

Hiebert LM, Liu JM. Heparin protects cultured arterial endothelial cells from damage by toxic oxygen metabolites. Atherosclerosis 1990; 83: 47-51

Weiler JM, Edens RE, Linhardt RJ, Kapelanski DP. Heparin and modified heparin inhibit complement activation in vivo. J Immunol 1992; 148: 3210-3215

Lucio J, D’Brot J, Guo CB, Abraham WM, Lichtenstein LM, Kagey-Sobotka A, Ahmed T. Immunologic mast cell-mediated responses and histamine release are attenuated by heparin. J Appl Physiol 1992; 73: 1093-1101

Sternbergh WC 3rd, Makhoul RG, Adelman B. Heparin prevents postischemic endothelial cell dysfunction by a mechanism independent of its anticoagulant activity. J Vasc Surg 1993; 17: 318-327

Leculier C, Benzerara O, Couprie N, Francina A, Lasne Y, Archimbaud E, Fiere D. Specific binding between human neutrophils and heparin. Br J Haematol 1992; 81: 81-85

Imai T, Hirata Y, Emori T, Marumo F. Heparin has an inhibitory effect on endothelin-1 synthesis and release by endothelial cells. Hypertension 1993; 21: 353-358

Inoue K, Hirota M, Kimura Y, Kuwata K, Ohmuraya M, Ogawa M. Further evidence for endothelin as an important mediator of pancreatic and intestinal ischemia in severe acute pancreatitis. Pancreas 2003; 26: 218-223

Edited by Zhu LH and Xu FM