Bacterial colonization of the upper airways of children positive and negative for SARS-CoV-2 during the COVID-19 pandemic

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Abstract

Background: Our understanding of the influence of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection on bacterial colonization in the children’s upper nasopharyngeal tract during the coronavirus infectious disease (COVID-19) pandemic is limited. This study aimed to determine whether there were any differences in bacterial colonization between asymptomatic children with or without a positive SARS-CoV-2 quantitative reverse transcriptase-polymerase chain reaction (RT-qPCR) results in the community setting.

Methods: A cross-sectional community-based exploratory study was conducted from March to May 2021 in Semarang, Central Java Province, Indonesia. Using stored nasopharyngeal swabs collected from children under 18 years as a contact tracing program, we performed a real-time quantitative (qPCR) for the most important bacterial colonizing pathogens: Streptococcus pneumoniae, Haemophilus influenzae, Staphylococcus aureus, and Klebsiella pneumoniae.

Results: Swabs from a total of 440 children were included in this study, of which 228 (51.8%) were RT-qPCR-confirmed SARS-CoV-2 positive. In the 440 children, colonization rates were highest for H. influenzae (61.4%), followed by S. pneumoniae (17.5%), S. aureus (12.0%), and K. pneumoniae (1.8%). The co-occurrence of both S. pneumoniae and H. influenzae in the upper respiratory tract was significantly associated with a SARS-CoV-2 negative RT-qPCR. In contrast, colonization with only S. aureus was more common in SARS-CoV-2-positive children.

Conclusion: Overall, this exploratory study concludes that there is a significant difference in the bacterial nasopharyngeal colonization pattern between SARS-CoV-2 positive and negative in asymptomatic children in the community in Indonesia.

Keywords: Bacterial colonization, COVID-19 pandemic, Children, Indonesia

Background

The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is a highly transmissible and pathogenic coronavirus that first emerged in late 2019 and has since caused a pandemic of coronavirus disease 2019 (COVID-19), which poses a threat to human health and public safety globally [1]. A variety of symptoms, ranging from mild to severe respiratory failure, indicate the pathophysiology of SARS-CoV-2 infection in people. SARS-CoV-2 replicates primarily in the epithelial cells of the upper respiratory tract, from where it spreads and penetrates the lungs [2]. However, as the body of knowledge about the continuing pandemic grows, it seems that young
children are less vulnerable to severe symptoms of SARS-CoV-2 infection [3].

Several studies have been conducted to determine the existence of SARS-CoV-2 co-infection with respiratory bacteria in the majority of adult patients who have contracted the virus. Gram-negative bacilli: *Acinetobacter baumannii*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *Stenotrophomonas maltophilia* and *Staphylococcus aureus* were commonly found as hospital-acquired superinfections, while *Streptococcus pneumoniae*, *Staphylococcus aureus*, and *Haemophilus influenzae* were found as community-acquired co-infection [4–6].

*S. pneumoniae*, *H. influenzae*, *S. aureus*, and *K. pneumoniae* are significant colonizers of children’s upper respiratory tracts that lead to lower respiratory tract infections, the leading cause of death in children under five years [7–10]. *S. pneumoniae* and *H. influenzae* are the most common bacterial causes of pneumonia, with *S. aureus* and *K. pneumoniae* associated with certain severe cases [11]. *K. pneumoniae*, along with other Gram-negative bacilli, is a more prevalent cause of pneumonia in Asia than in other regions worldwide [10]. By systematically searching the literature, we found that *K. pneumoniae* was the most frequently reported bacterial agent of lower respiratory tract infection in children under the age of five in Indonesia [12].

There is very little information about the association between the SARS-CoV-2 virus and bacterial colonization in asymptomatic children during public health measures during the COVID-19 pandemic. Therefore, this exploratory research aimed to determine whether there were any differences in *S. pneumoniae*, *H. influenzae*, *S. aureus*, and *K. pneumoniae* colonization between asymptomatic children with positive and negative SARS-CoV-2 RT-qPCR results in the community setting in Indonesia.

**Methods**

**Study design and setting**

A cross-sectional community-based exploratory study was conducted from March to May 2021 in Semarang, Central Java Province, Indonesia.

**Study population**

The inclusion criteria in this study were the availability of nasopharyngeal swabs of children below the age of 18 years that were recently (less than two weeks before the visit) in contact with a confirmed COVID-19 patient from which swab samples were collected for contact tracing and were asymptomatic during the sample collection. We excluded swabs with an insufficient sample volume or incomplete personal data (date of birth, sex, national identification number). The Ethical Committee of the Faculty of Medicine Universitas Diponegoro approved the study protocol (No.64/EC/KEPK/FK-UNDIP/III/2021).

**Clinical data collection**

We extracted age and sex data from the patient registry in the laboratory information system.

**Sample collection**

The nasopharyngeal swabs were collected by trained nurses from Primary Health Centres (Puskesmas) in viral transport medium across Central Java and were sent to the Microbiology Laboratory, Faculty of Medicine, Universitas Diponegoro, Semarang, Indonesia. This laboratory is the designated reference laboratory for SARS-CoV-2 PCR testing under standardized clinical laboratory conditions.

In order to provide a reliable comparison between SARS-CoV-2 positive and negative swab samples, we selected all positive samples together with a similar number of randomly selected negative samples from the same day. If the SARS-CoV-2 positive sample was higher than the number of negative samples on a respective day, all available negative samples were selected on that particular day. All samples were aliquoted (1 ml) and stored in a −80 °C freezer until further analysis.

**Quantitative real-time PCR (qPCR)**

DNA for the positive controls was extracted from *S. pneumoniae* (ATCC® 49,619™), *S. aureus* (ATCC® 25,923™), *K. pneumoniae* (ATCC® 33,495™), and *H. influenzae* (ATCC® 49,247™) using PureLink® Genomic DNA Mini Kit (cat.no.K182001, Invitrogen™, USA) based on the protocol for Gram-positive and Gram-negative bacteria. For real-time PCR standards, sequential dilution (tenfold) of the DNA spanning seven orders of magnitude starting at 10 ng/µl was prepared in two sets on each plate to validate PCR efficiency across the sample.

Each sample was thawed, vortexed, and aliquoted (100 µl) into a microcentrifuge tube (cat.no.1210-SoS, SSbio®, USA). The aliquoted samples were incubated at 95 °C for 15 min to lyse the bacteria and extract their DNA. Based on previous studies, four sets of primers (Table 1) based on highly conserved gene-specific for each bacterium were used [13–16]. The PCR was all performed using RealQ Plus 2 × Master Mix for Probe without ROX™ (cat.no.A313402, Ampliqon, Denmark) two-step PCR program in CFX Connect™ Real-Time PCR Detection System (Bio-Rad, USA). The PCR was done in a 25 µl reaction containing 1 µl of template DNA. Cycling conditions were 95 °C for 15 min, followed by 49 cycles of 95 °C for 15 s, and 20 s at 61 °C for *S. pneumoniae* and *K. pneumoniae*, 60 °C for *S. aureus*, and 55 °C for *H. influenzae*, respectively. Samples in which the
quantification cycle (Cq) values < 40 were measured were considered positive.

**Statistical analysis**
We computed the Mann–Whitney U test to determine the statistical difference in the median age between those that tested positive and negative for COVID-19. Using Pearson's chi-square test, we compared age groups and sex between SARS-CoV-2 infected and uninfected children. To determine the statistical significant differences in the mean rank of the Cq value among four bacteria, we used the Kruskal–Wallis H test. The effect size of the association between two nominal variables was determined using Odds Ratio (95% confidence interval (CI)). All tests were performed on a two-tail hypothesis, and a *p* value below 0.05 was considered statistically significant. All data collected were analysed using SPSS® Version 26.0 (IBM Corp. Release 2019. IBM® SPSS® Statistics for Macintosh, Version 26.0. Armonk, NY).

**Results**

**Demographic characteristic**
A total of 440 samples from healthy and asymptomatic children were collected in this study. Two hundred twenty-eight (51.8%) children had SARS-CoV-2 positive RT-qPCR results. The majority were children aged twelve to seventeen (50.5%) and females (50.9%). The median (IQR) age of the children with positive or negative SARS-CoV-2 RT-qPCR results was 12 (7) and 11 (8) years, respectively. A marginally significant association was found for SARS-CoV-2 positivity within the age group (*p* = 0.055), and no statistically significant association was found for SARS-CoV-2 positivity within gender (*p* = 0.44) (Table 2).

**Bacterial colonization in the upper respiratory tract**
The Cq values from all samples, indicative of the density of bacterial colonization, were determined for *S. pneumoniae*, *H. influenzae*, *S. aureus*, and *K. pneumoniae* (Fig. 1). *S. pneumoniae* had the lowest Cq value median (33.4), followed by *H. influenzae* (35.9), *S. aureus* (36.9), and *K. pneumoniae* (38.0) (*p* < 0.001). *H. influenzae* had the highest colonization rate (61.4%), followed by *S. pneumoniae* (17.5%), *S. aureus* (12.0%), and *K. pneumoniae* (1.8%) (Table 3). There was a significant association between age and overall colonization with *S. aureus* (*p* = 0.003) and *S. pneumoniae* (*p* = 0.003), with the highest colonization rates in 6–11 years (Table 3).

![Table 1](https://example.com/table1.png)

| Organism (target gene) | Primer name | Nucleotide sequence of primers and probes |
|------------------------|-------------|-------------------------------------------|
| *S. pneumoniae* (lytA) | lytA-CDC-f  | 5'-ACGAATTCATGAGATGAGA3'                  |
|                        | lytA-CDC-r  | 5'-CTGGGCTATCCGCT-3'                     |
|                        | lytA-CDC-pr | 5'-FAM-GCGAAAACGGTCGATACAAGGAG3'-BHQ1     |
| *S. aureus* (nuc)      | nucF        | 5'-GTTCACTTTGAATCTTTAGTTGTA3'            |
|                        | nucR        | 5'-AATGCGAGGTCTTATGTTAATT3'              |
|                        | nucPr       | 5'-FAM-AAATCTAGTACTGACGAAATGCA3'-BHQ1     |
| *K. pneumoniae* (mdh)  | mdhF        | 5'-GGGGCCGATGGGCTA-3'                    |
|                        | mdhR        | 5'-GATACCCGATTCCATTTAAGACG-3'            |
|                        | mdhP        | 5'-FAM-GCCGCCGATTGGATCCGA-3'-BHQ1        |
| *H. influenzae* (hpd)  | hpdF729     | 5'-AGATTGGAAGAAACACAAAGAA3'-              |
|                        | hpdR819     | 5'-CAGCACCGAGATTTAACACT3'-               |
|                        | hpdPbr762i  | 5'-FAM-AAAATCCATGGT-BHQ1-AATATAGTTA      |

![Table 2](https://example.com/table2.png)

| Variables analysed using the Mann–Whitney U test, *p* < 0.05 | SARS-CoV-2 negative (*N* = 212) | SARS-CoV-2 positive (*N* = 228) | *p* |
|--------------------------------------------------------------|---------------------------------|---------------------------------|-----|
| Age (median; IQR)                                           | 12; 7                           | 11; 8                           | 0.023* |
| Age groups (years)                                          | 0–5                             | 35 (15.4)                       | 0.055+ |
| 6–11                                                        | 67 (31.6)                       | 67 (29.4)                       |     |
| 12–17                                                       | 96 (45.3)                       | 126 (55.3)                      |     |
| Sex                                                         | Male                            | 116 (50.9)                      | 0.44 |
|                                                            | Female                          | 112 (49.1)                      |     |

*Variables analysed using the Mann–Whitney U test, *p* < 0.05
+ Variables analysed using Pearson's chi-square test, marginally significant

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**Table 1** Primers and probes used for the qPCR assays to detect *S. pneumoniae, H. influenzae, S. aureus,* and *K. pneumoniae*

**Table 2** Demographic characteristics of the study participants
Table 4 summarizes the colonization prevalence in children positive or negative for SARS-CoV-2. In most nasopharyngeal swab samples, only a single species was detected (Table 4). Although *H. influenzae* had the highest detection rate in children, those who were only colonized with *S. aureus* had higher odds of being SARS-CoV-2 positive (OR 3.9, 95% CI 1.1 to 13.9). In this population, co-occurrence of *S. pneumoniae* and *H. influenzae* dominates (61.5%) the other two bacterial species combinations. More importantly, this specific combination was significantly associated with SARS-CoV-2 negative RT-qPCR results (OR 0.5, 95% CI 0.3 to 0.9).

**Discussion**

This study was conducted during the second wave of the COVID-19 pandemic, when the delta variant was most prevalent [17] in Indonesia, to determine any differences in the colonization patterns of *S. pneumoniae*, *H. influenzae*, *S. aureus*, and *K. pneumoniae* between SARS-CoV-2 positive and negative children. In this study, it was found that *H. influenzae* had the highest colonization rate. Interestingly, co-colonization of *S. pneumoniae* and *H. influenzae* in the nasopharynx was associated with the absence of SARS-CoV-2 infection, while children only colonized with *S. aureus* had higher odds of having SARS-CoV-2 positive results. These results suggest a difference in bacterial colonization patterns between SARS-CoV-2 RT-qPCR positive and negative in asymptomatic children.

In this study, we found that from all 440 children included, *H. influenzae* had the highest colonization rate, followed by *S. pneumoniae*, *S. aureus*, and *K. pneumoniae*. This result differs from pre-COVID-19 studies, which found the highest colonization rates for *S. pneumoniae*, followed by *H. influenzae* and *S. aureus* [12]. A pre-COVID-19 study among HIV-infected children in Jakarta and healthy children in three provinces in Indonesia (West Java, West Nusa Tenggara, West Sumatra) showed 18% and 27.5% *H. influenzae* carrier rates [18, 19].
Compared to other Southeast Asia countries, the colonization rate of *H. influenzae* in our study is also considerably higher. A study in Malaysia showed 14.3% and 4.9% *H. influenzae* carrier rates in healthy unvaccinated and vaccinated children, respectively [20]. The Indonesian ministry of health started to include the Hib vaccination in the national vaccination program in 2013; this might have caused the higher carrier rate of *H. influenzae* in this study since most of our participants were older than 5 years old.

While an earlier study suggested that bacterial co-infection with *H. influenzae, S. pneumoniae*, increased the risk of morbidity and mortality in patients hospitalized due to community-acquired pneumoniae and SARS-CoV-2 infection [21, 22], we found that this specific combination of pathogenic bacterial species was associated with reduced occurrence of SARS-CoV-2 infection in children. Co-occurrence of *H. influenzae* and *S. pneumoniae*, leading to increased density of *H. influenzae* in the presence of *S. pneumoniae* in a neonatal rat model, was previously shown and suggested synergy between these species [23]. A possible explanation is a nutritional dependency in which *S. pneumoniae* provides nutrients to *H. influenzae*, as observed for *S. aureus* and *H. influenzae*, described as the ‘satellite phenomenon’ [23]. It is tempting to speculate that this synergistic interaction in the nasopharynx elicits mucosal immune responses leading to inhibition of infection with SARS-CoV-2, although a causal link has not been demonstrated in this study.

The association of SARS-CoV-2 infection and *S. aureus* colonization, as found in this study, is in line with other studies [22] showing that although bacterial co-infection in COVID-19 patients was low, *S. aureus* was the most commonly found species. Regarding *K. pneumoniae*, we found 1.8% carriage in all samples. This result is lower than a study held eight years ago in Indonesia that found 7% of *K. pneumoniae* carriage in children under five years of age [24]. However, as proposed by Farida and colleagues, exposure to unsafe food and water was likely the culprit of the previously high carrier rate [24]; during mobility reduction and school closure, children were less exposed to street food, thus reducing the change of *K. pneumoniae* transmission in the pandemic era.

We acknowledge some limitations in this study. First, we had no access to information on the history of the children’s vaccination and current antibiotic use; thus, we cannot compare the bacterial colonization among children who have received a vaccination and have used antibiotics and those who have not. Second, there is no exact data related to when the children were exposed to SARS-CoV-2 infection. Despite these limitations, to our knowledge, this is the first study to examine the differences in bacterial colonization between SARS-CoV-2 RT-qPCR positive and negative in asymptomatic children. Further studies using 16 s rRNA sequencing and metagenomics in both symptomatic and asymptomatic children would improve our understanding of global changes in the microbiome of the upper respiratory as a consequence of SARS-CoV-2 infection.

### Table 4 Co-colonization of bacterial species in SARS-CoV-2 infected and uninfected children

| Variables analysed using Pearson’s chi-square test, \(p < 0.05\) | SARS-CoV-2 negative \((N = 212)\) | SARS-CoV-2 positive \((N = 228)\) | Odd ratio \((95\% CI)\) |
|---|---|---|---|
| No colonization | 70 (33.0) | 70 (30.7) | 0.9 (0.6 to 1.3) |
| Single bacterial species | | | |
| *S. aureus* | 3 (3.3) | 12 (11.1) | 3.8 (1.1 to 13.9)* |
| *S. pneumoniae* | 4 (4.4) | 7 (6.5) | 1.6 (0.5 to 5.7) |
| *K. pneumoniae* | 2 (2.2) | 0 (0.0) | NA |
| *H. influenzae* | 81 (90.0) | 89 (82.4) | 1.0 (0.7 to 1.5) |
| Two bacterial species | | | |
| *S. aureus* and *S. pneumoniae* | 0 (0.0) | 2 (4.3) | NA |
| *S. aureus* and *H. influenzae* | 10 (20.0) | 20 (43.5) | 1.9 (0.9 to 4.3) |
| *S. pneumoniae* and *H. influenzae* | 37 (74.0) | 22 (47.8) | 0.5 (0.3 to 0.9)* |
| *K. pneumoniae* and *H. influenzae* | 3 (6.0) | 2 (4.3) | 0.6 (0.1 to 3.7) |
| Three bacterial species | | | |
| *S. aureus* and *S. pneumoniae* and *H. influenzae* | 2 (100.0) | 3 (75.0) | 1.4 (0.2 to 8.5) |
| *S. aureus* and *K. pneumoniae* and *H. influenzae* | 0 (0.0) | 1 (25.0) | NA |
Conclusion
This exploratory study shows differences in the bacterial nasopharyngeal colonization pattern between asymptomatic SARS-CoV-2 positive and negative children during public health measures in the community in Indonesia. Differences in the colonization pattern might influence the epidemiology of bacterial respiratory tract infections in children, as well as modulate the risk for SARS-CoV-2 infection.

Supplementary Information
The online version contains supplementary material available at https://doi.org/10.1186/s12879-022-07851-z.

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Author contributions
VRC, MIdJ, and QdM conceived the study. All authors contributed to the study design. VRC and RH contributed to data collection and data analysis. VRC, RH, ESL, and HF drafted the manuscript. VRC, MIdJ, QdM critically revised the manuscript. All authors have agreed to both to be personally accountable for the author’s contributions and to ensure that questions related to the accuracy or integrity of any part of the work, even ones in which the author was not personally involved, are appropriately investigated, resolved, and the resolution documented in the literature. All authors read and approved the final manuscript.

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Availability of data and materials
All data generated or analysed during this study are included in this published article and its supplementary files (Additional file 1).

Declarations
Ethics approval and consent to participate
The research was performed in accordance with the Declaration of Helsinki. Data were collected for the purpose of public health response, and therefore, informed consent was not obtained. Informed consent was waived by the Ethical Committee of the Faculty of Medicine Universitas Diponegoro (No.64/EC/KEPK/UK-UNDIP/III/2021). The dataset used for this research was de-identified. The Ethical Committee of the Faculty of Medicine Universitas Diponegoro approved the study protocol (No.64/EC/KEPK/UK-UNDIP/III/2021).

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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References
1. Hu B, Guo H, Zhou P, Shi ZL. Characteristics of SARS-CoV-2 and COVID-19. Nat Rev Microbiol. 2021;19(3):141–54.
2. Wolff R, Corman VM, Guggemos W, Seilmaier M, Zange S, Muller MA, et al. Viral nucleic acid testing in hospitalised patients with COVID-2019. Nature. 2020;581(7809):465–9.
3. Bhuyan MU, Stibby E, Hassan MZ, Chan M, Islam MS, Haider N, et al. Epidemiology of COVID-19 infection in young children under five years: a systematic review and meta-analysis. Vaccine. 2021;39(4):667–77.
4. Langford BJ, So M, Raybarhan S, Leung V, Westwood D, MacFadden DR, et al. Bacterial co-infection and secondary infection in patients with COVID-19: a living rapid review and meta-analysis. Clin Microbiol Infect. 2020;26(12):1622–9.
5. Rawson TM, Moore LS, Zhu N, Ranganathan N, Skolimowska K, Gilchrist M, et al. Bacterial and fungal coinfection in individuals with coronavirus: a rapid review to support COVID-19 antimicrobial prescribing. Clin Infect Dis. 2020;71(9):2459–68.
6. Garcia-Vidal C, Sanjuan G, Moreno-Garcia E, Puerta-Alcalde P, Garcia-Pouton N, Chumbita M, et al. Incidence of coinfections and superinfections in hospitalized patients with COVID-19: a retrospective cohort study. Clin Microbiol Infect. 2021;27(1):e13–8.
7. Weiser JN, Ferreira DM, Paton JC. Streptococcus pneumoniae: transmission, colonization and invasion. Nat Rev Microbiol. 2018;16(6):355–67.
8. Giufre M, Daprai L, Cardines R, Bernaschi P, Rava L, Accogli M, et al. Carriage of Haemophilus influenzae in the oropharynx of young children and molecular epidemiology of the isolates after fifteen of years. Int J Med Microbiol. 2015;336(4):622–34.
9. Groud JA, Rich HE, Alcorn JF. Host-pathogen interactions in gram-positive bacterial pneumonia. Clin Microbiol Rev. 2019;32(3).
10. Juan CH, Fang SY, Chou CH, Tsai TY, Lin YT. Clinical characteristics of patients with pneumonia caused by Klebsiella pneumoniae in Taiwan and prevalence of antimicrobial-resistant and hypervirulent strains: a retrospective study. Antimicrob Resist Infect Control. 2020;9(1):4.
11. le Roux DM, Zar HJ. Community-acquired pneumonia in children - a changing spectrum of disease. Pediatr Radiol. 2017;47(11):1392–8.
12. Ciptaningtyas VR, De Mast Q, De Jonge MI. The burden and etiology of lower respiratory tract infections in children under five years of age in Indonesia. J Infect Dev Ctries. 2021;15(5):603–14.
13. Carvalho Mda G, Tondella ML, McCaustland K, Weidlich L, McGee L, Mayer LW, et al. Evaluation and improvement of real-time PCR assays targeting lytA, ply, and psaA genes for detection of pneumococcal DNA. J Clin Microbiol. 2007;45(8):2460–6.
14. Wang X, Mair R, Hatcher C, Theodore MJ, Edmond K, Wu HM, et al. Detection of bacterial pathogens in Mongolia meningitis surveillance with a new real-time PCR assay to detect Haemophilus influenzae. Int J Med Microbiol. 2011;301(4):303–9.
15. Kilic A, Muldrew KL, Tang YW, Basoutoglou AC. Triplex real-time polymerase chain reaction assay for simultaneous detection of Staphylococcus aureus and coagulase-negative staphylococci and determination of methicillin resistance directly from positive blood culture bottles. Diagn Microbiol Infect Dis. 2010;66(4):349–55.
16. Fukushima H, Sato Y, Hasegawa H, Saeki H, Katano H. Development of a new real-time PCR assay to detect Haemophilus influenzae. Int J Med Microbiol. 2020;26(12):1622–9.
17. Litbang.Kemkes.go.id. Map of Sequence Distribution and Variants of SARS-CoV-2 in Indonesia Indonesia: National Institute of Health Research and Development, Ministry of Health of the Republic of Indonesia 2021 [updated 2022 Nov 14]. https://www.litbang.kemkes.go.id/ peta-sebaran- varian-covid-19/. Accessed 21 Feb 2022.
18. Dunne EM, Murad C, Sudigdoad S, Fadlyana E, Tariqan R, Indriyani SAK, et al. Carriage of Streptococcus pneumoniae, Haemophilus influenzae,
Moraxella catarrhalis, and Staphylococcus aureus in Indonesian children: a cross-sectional study. PLoS ONE. 2018;13(4): e0195098.
19. Safari D, Lestari AN, Khoeri MM, Tafroji W, Gin-Rachman EA, Harimurti K, et al. Nasopharyngeal carriage and antimicrobial susceptibility profile of Haemophilus influenzae among patients infected with HIV in Jakarta, Indonesia. Access Microbiol. 2020;2(12): acmi000165.
20. Palaniappan PA, Mohamed Sukur S, Liow YL, Maniam S, Sherina F, Ahmad N. Carriage of Haemophilus influenzae among children attending childcare centres in Kuala Lumpur, Malaysia in the post vaccination era: a cross-sectional study. Vaccine. 2020;38(51):8232–7.
21. Ngocho JS, Minja L, van der Gaast-de Jongh CE, Rahamat-Langendijk JC, Langereis JD, Mmbaga BT, et al. Viral-bacterial (co-) occurrence in the upper airways and the risk of childhood pneumonia in resource-limited settings. J Infect. 2020;81(2):213–20.
22. Westblade LF, Simon MS, Satlin MJ. Bacterial coinfections in coronavirus disease 2019. Trends Microbiol. 2021;29(10):930–41.
23. Margolis E, Yates A, Levin BR. The ecology of nasal colonization of Streptococcus pneumoniae, Haemophilus influenzae and Staphylococcus aureus: the role of competition and interactions with host’s immune response. BMC Microbiol. 2010;10:59.
24. Farida H, Severin JA, Gasem MH, Keuter M, van den Broek P, Hermans PW, et al. Nasopharyngeal carriage of Klebsiella pneumoniae and other Gram-negative bacilli in pneumonia-prone age groups in Semarang, Indonesia. J Clin Microbiol. 2013;51(5):1614–6.

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