Apolipoprotein E polymorphism modulation of asymmetric dimethylarginine in hypertensive patients is determined by renal function

Mauro Sergio Martins Marrocos, Andrei Alkmin Teixeira, Beata Marie Quinto, Silmara de Melo Carmona, Mariana Kuniyoshi, Cassio Jose Rodrigues, Maria Aparecida Dalboni, Silvia Manfredi, Maria Eugênia Canziani and Marcelo Costa Batista

Abstract

Background: Endothelial dysfunction is considered an early step of atherosclerotic vascular disease. Asymmetric dimethylarginine (ADMA), the main endogenous inhibitor of nitric oxide synthase (NOS), plays a critical role in the process of atherosclerosis in a uremic environment. Increased plasma ADMA not only works as a cardiovascular morbidity biomarker but it is also involved in the genesis of atherosclerosis in renal disease. Considering the relationships of apolipoprotein E (ApoE) polymorphism with LDL cholesterol (LDL-C) levels and coronary risk, it is possible that it brings on susceptibility to endothelial dysfunction and atherogenesis seen on uremia.

Methods: Six hundred twenty patients were stratified according to glomerular filtration rate (GFR) estimated by Chronic Kidney Disease Epidemiology Collaboration (CKDEPI) formula: group I > 60 mL/min, group II ≤ 60 mL/min and > 15 mL/min, and group III ≤ 15 mL/min or in hemodialysis. Polymorphic ApoE analysis was performed by polymerase chain reaction amplification (PCR). Plasma ADMA levels were measured by high performance liquid chromatography (HPLC). Groups were compared on clinical and laboratory characteristics as well as allele and genotype distribution towards.

Results: The ε2 allele of ApoE was present in 62 (10.3%) patients, ε3 allele in 581 (96.2%), and ε4 allele in 114 (18.9%). Their distribution among the 3 groups was uniform. Such uniformity was not observed when we considered endothelial function measured by asymmetric dimethylarginine. In group III, the frequency of ε4 allele was significantly lower in the third tertile compared with the first tertile (14.7 versus 53.3%, P = 0.000; Pearson chi-square). In groups I and II, there was no difference in allele frequency according to ADMA levels. This association remained significant even after confounding factors corrections (OR 0.329, 95% CI 0.155 - 0.699, P = 0.004).

Conclusions: The results of this study shows that the frequency of ε4 allele of ApoE is significantly lower among hypertensive patients on hemodialysis with the highest levels of ADMA. Uremia is capable of determining lower plasma ADMA levels in hypertensive ε4 allele carriers.

Keywords: Asymmetric Dimethylarginine, Apolipoprotein E, Polymorphism, Atherogenesis, Chronic Kidney Disease
**Background**

As stated in the 2014 United States Renal Data System Annual Data Report, the burden of cardiovascular disease (CVD) in chronic kidney disease (CKD) is still considerable, where CVD in CKD individuals is 2–3 more frequent when compared with the general population. There is a stepwise decrease in survival related to advanced CKD stage, and mortality after congestive heart failure (CHF) or myocardial infarction (MI) in patients with stages 4–5 CKD is similar to that of dialysis patients [1].

It was first reported in 1992 that nitric oxide (NO) synthesis can be inhibited by ADMA, which is substantially elevated in plasma of patients on hemodialysis. It was suggested that the accumulation of ADMA and concomitant inhibition of NO synthesis may contribute to hypertension, immune dysfunction, and CVD in these patients [2]. ADMA, as the main endogenous inhibitor of NOS, plays a critical role in the process of atherosclerosis in a uremic environment [3]. Endothelial dysfunction resulting from reduced NO activity is considered an early step of atherosclerotic vascular disease [4]. Increased plasma ADMA concentrations not only works as a cardiovascular morbidity biomarker but is causally involved in the genesis of atherosclerosis in renal disease [5–7].

Also related to atherosclerosis, ApoE is a multifunctional protein that plays a major role in the metabolism of cholesterol and triglycerides by binding to receptors in the liver to help mediate the clearance of chylomicrons and very low-density lipoproteins from the bloodstream. ApoE polymorphism is also implicated in the burden of CVD. Boerwinkle et al. reported that up to 17 % of the genetic variability in total plasma cholesterol may be attributable to the ApoE polymorphism [8].

Since ApoE polymorphism is a key player in lipid metabolism, it is possible that it leads to susceptibility to endothelial dysfunction and atherogenesis. Aim of study was to evaluate the interaction between ADMA levels and ApoE polymorphism in patients stratified according 3 different degrees of renal function.

**Results**

Demographic and laboratory characteristics of the patients are summarized in Table 1. Eight subjects were excluded because of missing ApoE polymorphism data, leaving 612 subjects for analysis. Mean age of the patients was 60.1 ± 14.1 years, 311 (50.7 %) were male, mean body mass index (BMI) was 28.0 ± 5.3, 386 (63.2 %) were Caucasians, 238 (38.9 %) had Diabetes Mellitus (DM), and 218 (35.6 %) had CVD. Groups I, II

| Table 1 Demographic and laboratory characteristics of patients stratified by renal function |
|---------------------------------|---------------------------------|---------------------------------|---------------------------------|------------------|
| Group I(N = 189)               | Group II(N = 222)               | Group III(N = 202)              | p                      |
| Age (years)            60.0 ± 11.7 | 66.8 ± 11.6 | 52.4 ± 14.9 | 0.000* |
| Male                  57 (30.5 %) | 136 (61.3 %) | 117 (58.2 %) | 0.000*** |
| Caucasian             105 (56.1 %) | 145 (65.9 %) | 138 (68.9 %) | 0.000*** |
| BMI (kg/cm²)          30.3 ± 6.2 | 28.6 ± 5.1 | 25.2 ± 2.6 | 0.000** |
| Waist circumference (cm) 98.3 ± 13.9 | 101.9 ± 13.5 | 95.6 ± 10.8* | 0.004* |
| Diabetes mellitus      59 (32.6 %) | 118 (53.4 %) | 65 (32.5 %) | 0.000*** |
| Metabolic Syndrome     118 (64.1 %) | 158 (71.8 %) | 146 (73.0 %) | 0.000*** |
| Cardiovascular Disease 44 (24.6 %) | 91 (41.9 %) | 86 (43.7 %) | <.000*** |
| o Cerebrovascular disease 20 (11.2 %) | 26 (12.0 %) | 21 (10.7 %) | 0.912*** |
| o Coronary disease     17 (9.5 %) | 51 (23.5 %) | 44 (22.3 %) | 0.001*** |
| o Peripheral vascular disease 11 (6.1 %) | 18.9 (41.0 %) | 46 (23.4 %) | 0.000*** |
| o Congestive heart failure 17 (9.5 %) | 33 (15.3 %) | 31 (15.7 %) | 0.000*** |
| Statin use             77/181 (40.7 %) | 125/222 (54.3 %) | 107/200 (53.2 %) | 0.017*** |
| Cholesterol(mM)        4.78 ± 0.93 | 4.65 ± 1.12 | 3.72 ± 1.00* | 0.000* |
| HDL-C(mM)              1.27 ± 0.34 | 1.16 ± 0.33* | 0.99 ± 0.37* | 0.000* |
| LDL-C(mM)              2.78 ± 0.82 | 2.64 ± 0.97* | 1.89 ± 0.74* | 0.000* |
| Triglycerides(mM)      1.61 ± 0.90 | 1.80 ± 0.98 | 1.91 ± 1.31 | 0.023** |
| Creatinine( μM)        73.4 ± 15.9 | 175.0 ± 69.8 | 930.8 ± 284.6 | 0.000*** |
| ADMA ( μM)             0.48 ± 0.12 | 0.75 ± 0.31* | 1.34 ± 0.90 | 0.000** |
| CRP (mg/dL)            0.49 ± 0.73 | 0.62 ± 0.83 | 1.47 ± 3.78 | 0.004** |

*ANOVA, **Kruskal-Wallis-one way- ANOVA, ***Pearson chi-square

By post hoc Bonferroni test: *groups III and II, *groups III and I, *groups II and I
and III were composed respectively of 199, 206 and 207 patients. In comparison with group I, groups II and III included more males and Caucasians, while group III had a lower mean age (52.4 ± 14.9 years). Hemodialysis vintage in group III was 37.36 ± 26.84 months. BMI and waist circumference decreased from group I to group III. Prevalence of DM was highest in group II (53.4 %); metabolic syndrome and CVD increased in prevalence from group I to group III. Use of statins was predominately in groups II and III (54.3 % and 53.2 %).

HDL cholesterol (HDL-C), LDL-C and triglyceride concentrations significantly decreased with decrease in estimated GFR. Plasma concentrations of ADMA and C reactive protein (CRP) significantly increased with decrease in estimated GFR.

The ε2 allele was present in 62 (10.3 %) patients, ε3 allele in 581 (96.2 %), and ε4 allele in 114 (18.9 %). The ε3/2 genotype was present in 57 (9.3 %) patients, ε3/3 genotype in 438 (71.7 %), ε4/2 genotype in 10 (1.6 %), ε4/3 genotype in 93 (15.2 %) and ε4/4 genotype in 13 (2.1 %). Table 2 depicts ApoE genotype and allele distribution among the 3 renal function groups. There was evidence of a significant association (P = 0.018) with Caucasians for the ε4 allele, when distribution of genotypes and alleles were stratified by race (Table 3).

Table 4 depicts the statistically significant difference in plasma ADMA levels between patients on renal replacement therapy (RRT) based on ApoE ε4 allele presence; Table 5 presents demographic and laboratory characteristics of patients according to ADMA tertile. ApoEε4 allele predominated in the first tertil of ADMA (Table 6). As also shown in Fig. 1, in group III, the frequency of ε4 allele of ApoE was significantly lower in the third tertile of ADMA compared with the first tertile. In groups I and II, there was no difference in allele frequency according to ADMA level. This association remained significant even after controlling for confounders (OR 0.329,95 % CI 0.155 – 0.699; P = 0.004), namely age, sex, BMI, DM, CVD, CRP, LDL-C and dialysis vintage (Table 7).

Analyses of clinical and laboratory characteristics of ε2 and ε3 allele carriers showed no statistical difference between allele (+) versus allele (−) groups and stratified for RRT allocation or not (data not shown).

Discussion

According to the literature, our data confirm that increased plasma ADMA levels are associated with decreasing GFR. We did not identify a difference in the distribution of ApoE genotypes and alleles between groups, but among patients on hemodialysis and with highest ADMA levels, the frequency of ε4 allele of ApoE was significantly lower when compared with that in patients in the first ADMA tertile. The observed difference did not occur in patients with normal renal function or in earlier stages of CKD.

In fact, ADMA levels are intrinsically related with renal function and this association has been studied in the past few years. The 3 groups in our study showed progressively higher plasma ADMA levels following the evolution of CKD. In accordance with our data, Fliser et al. found that ADMA was significantly associated with progression of nondiabetic kidney diseases [9]. Ravani et al. found that in patients with mild CKD plasma ADMA was inversely proportional to GFR and represented a strong and independent risk marker for progression to end stage renal disease (ESRD) and mortality [10]. Initially, the elevation of ADMA in uremia was credited only to the decrease in GFR. Thereafter, elevations in plasma ADMA concentrations were also reported in incipient renal diseases, such as autosomal-dominant polycystic kidney disease [11] and IgA nephropathy [12], and in normal renal function such hypercholesterolemia or atherosclerosis [13]. ADMA is renally excreted to some extent,
but its rise in CKD is due to increased activity of protein arginine methyltransferases [14], a family of enzymes involved in the process of post-translational methylation of arginine residues, a common mechanism of protein modification. In addition, accumulation of ADMA in patients with CKD is also related to decreased activity of the enzyme dimethyl arginine dimethyl aminohydrolase (DDAH) [14], part of the main pathway for ADMA degradation, which hydrolyzes ADMA to dimethylamine and L-citrulline.

So far, two isoforms of DDAH have been characterized and cloned: DDAH I is predominately found in tissues that express neuronal NOS, whereas DDAH II is predominately found in tissues expressing endothelial NOS [14, 15]. It is estimated that healthy humans produce approximately 300 μmol of ADMA per day, of which approximately 250 μmol are metabolized by DDAHs [16].

ADMA is an endogenous inhibitor of nitric oxide synthases (NOS), which may in part explain the impaired vasorelaxation, elevated inflammation, and reduced angiogenesis reported in CKD patients and animal models of CKD [17]. The role of ADMA in atherogenesis is not restricted to its inhibitory action on NOS. It upregulates the expression of acyl-coenzyme A cholesterol acyltransferase 1 present in macrophages and is implicated in the formation of foam cells [7]. ADMA also impairs the migratory capacity of angiogenic progenitor cells (APCs) in patients with coronary artery disease through a micro RNA-21-dependent mechanism, inhibiting superoxide dismutase 2 in APCs [18].

Accumulation of ADMA is a risk factor and is causally related to the development of endothelial dysfunction and cardiovascular disease in patients with CKD [19–22]. Zoccali et al. demonstrated in patients undergoing hemodialysis with initially normal carotid intima-media thickness, that ADMA and CRP are interacting factors in the progression of carotid intimal lesions [23].

ApoE gene contains three potential alleles: ε2, ε3 and ε4, forming six genotypes and determining diversity in clinical expression. Allele frequency from high to low is ε3, ε2 and ε4, and ε3ε3 is the most common phenotype in humans [24], as seen in our study. In NHANES III [25], the ε2 allele was the rarest for each ethnicity examined, and the frequency of the ε4 allele was 10.8 %

### Table 4 Demographic and laboratory characteristics of patients according to RRT allocation and ApoE ε4 allele presence

|                      | In RRT       | Not in RRT  |
|----------------------|--------------|-------------|
|                      | ε4 allele (+) (N = 40) | ε4 allele (-) (N = 158) | P   | ε4 allele (+) (N = 74) | ε4 allele (-) (N = 330) | P   |
| Age (years)          | 52.47 ± 14.97 | 53.60 ± 14.36 | 0.669* | 62.80 ± 13.36 | 63.81 ± 11.72 | 0.511* |
| Dialysis vintage     | 38.95 ± 24.24 | 37.22 ± 27.69 | 0.697* | - | - | - |
| Male                 | 25 (21.6 %) | 91 (78.4 %) | 0.574** | 37 (19.7 %) | 151 (80.3 %) | 0.508** |
| Caucasian            | 31 (22.8 %) | 105 (72.2 %) | 0.403** | 51 (20.6 %) | 196 (79.4 %) | 0.440** |
| BMI (kg/cm²)         | 25.15 ± 11.12 | 25.57 ± 2.44 | 0.356* | 29.17 ± 6.40 | 29.33 ± 5.61 | 0.844* |
| Waist circumference (cm) | 97.25 ± 15.96 | 99.30 ± 9.13 | 0.283* | 99.81 ± 14.73 | 100.27 ± 13.59 | 0.798* |
| Diabetes mellitus    | 15 (23.1 %) | 50 (76.9 %) | 0.418** | 32 (18.8 %) | 138 (81.2 %) | 0.895** |
| Metabolic syndrome   | 28 (19.4 %) | 116 (80.6 %) | 0.621** | 48 (17.8 %) | 222 (82.2 %) | 0.733** |
| Cardiovascular disease | 19 (22.6 %) | 65 (77.4 %) | 0.547** | 26 (19.8 %) | 105 (80.2 %) | 0.653** |
| oCerebrovascular disease | 0 (0.0 %) | 20 (100.0 %) | 0.016** | 13 (28.3 %) | 33 (71.7 %) | 0.073** |
| oCoronary disease    | 13 (29.5 %) | 31 (70.5 %) | 0.096** | 11 (16.9 %) | 54 (83.1 %) | 0.703** |
| oPeripheral vascular disease | 9 (20.5 %) | 35 (79.5 %) | 0.976** | 10 (20.4 %) | 39 (79.6 %) | 0.697** |
| oCongestive Heart Failure | 6 (19.4 %) | 25 (80.6 %) | 0.850* | 7 (14.9 %) | 40 (85.1 %) | 0.480** |
| Statin use           | 23 (21.9 %) | 78.1 % | 0.551** | 38 (19.4 %) | 158 (80.6 %) | 0.602** |
| Cholesterol(mmol/L)  | 3.75 ± 1.01 | 3.62 ± 0.90 | 0.470* | 4.58 ± 1.06 | 4.75 ± 1.06 | 0.175* |
| HDL-C(mmol/L)        | 0.97 ± 0.34 | 1.08 ± 0.46 | 0.108* | 1.15 ± 0.31 | 1.22 ± 0.35 | 0.126* |
| LDL-C(mmol/L)        | 1.92 ± 0.74 | 1.82 ± 0.76 | 0.469* | 2.62 ± 0.85 | 2.79 ± 0.92 | 0.151* |
| Triglycerides(mmol/L)| 1.98 ± 1.36 | 1.66 ± 1.14 | 0.160* | 1.84 ± 1.06 | 1.69 ± 0.90 | 0.237* |
| ADMA (μM)            | 0.95 ± 0.45 | 1.44 ± 0.96 | 0.000* | 0.58 ± 0.17 | 0.63 ± 0.28 | 0.134* |
| CRP (mg/dL)          | 1.67 ± 3.52 | 1.42 ± 3.88 | 0.711* | 0.50 ± 0.72 | 0.57 ± 0.76 | 0.468* |

*Student’s t test  **Pearson chi-square
among Mexican Americans, compared to 15% in whites and 22% in non-Hispanic blacks. Similarly to Chu et al. [25], we demonstrated that the ε2 allele was the less prevalent in all groups in our analysis of allele and genotype distribution by race. ApoE ε4 allele showed evidence of significant association with Caucasians.

Several previous studies reported that the ε2 allele is a genetic risk factor for all-cause CKD [26] and related to increased risk of ESRD. [27, 28] In contrast, the ε4 allele may be associated with a lower risk of diabetic nephropathy [29]. Furthermore, lipoprotein glomerulopathy, a rare inherited renal disease characterized by proteinuria...

### Table 5
Demographic and laboratory characteristics of patients according to ADMA tertile

|                        | First tertile (0.12 - 0.54) (N = 199) | Second tertile (0.55 - 0.79) (N = 206) | Third tertile (0.80 - 4.91) (N = 207) | P       |
|------------------------|--------------------------------------|---------------------------------------|--------------------------------------|---------|
| Age (years)            | 62.37 ± 10.53                        | 66.01 ± 11.95                        | 65.64 ± 11.70                       | 0.000** |
| Dialysis vintage (months) | 2.48 ± 24.24                        | 7.90 ± 1.48                          | 25.62 ± 1.92c                       | 0.000** |
| Male                   | 25 (21.6 %)                          | 91 (78.4 %)                          | 37 (19.7 %)                         | 0.508***|
| Caucasian              | 31 (22.8 %)                          | 105 (72.2 %)                         | 51 (20.6 %)                         | 0.440***|
| BMI (kg/m²)            | 29.74 ± 5.55                         | 29.11 ± 5.24                         | 27.63 ± 13.43                       | 0.000** |
| Waist circumference (cm) | 99.46 ± 13.43                       | 101.45 ± 11.40                       | 101.88 ± 14.62                      | 0.798   |
| Diabetes mellitus      | 15 (23.1 %)                          | 50 (76.9 %)                          | 32 (18.8 %)                         | 0.895***|
| Metabolic syndrome     | 28 (19.4 %)                          | 116 (80.6 %)                         | 48 (17.8 %)                         | 0.733   |
| Cardiovascular disease | 66 (30.1 %)                          | 77 (35.2 %)                          | 76 (34.7 %)                         | 0.646** |
| ε3/ε2                  | 19 (9.9 %)                           | 21 (10.3 %)                          | 16 (7.8 %)                          | NS      |
| ε3/ε3                  | 123 (64.4 %)                         | 147 (72.4 %)                         | 158 (77.5 %)                        | NS      |
| ε4/ε2                  | 7 (3.7 %)                            | 2 (1.0 %)                            | 1 (0.5 %)                           | NS      |
| ε4/ε3                  | 37 (19.4 %)                          | 3 (1.0 %)                            | 25 (12.3 %)                         | NS      |
| ε4/ε4                  | 5 (0.8 %)                            | 3 (0.5 %)                            | 4 (0.7)                             | NS      |
| ε2 allele              | 26 (13.6 %)                          | 23 (11.3 %)                          | 17 (8.3 %)                          | NS      |
| ε3 allele              | 179 (93.7 %)                         | 197 (97.0 %)                         | 199 (97.5 %)                        | NS      |
| ε4 allele              | 49 (25.7 %)                          | 35 (17.2 %)                          | 29 (14.2 %)                         | 0.011   |
| Adjusted residual      | 2.9                                  | 0.7                                  | 2.1                                 |         |

\*ANOVA, **Kruskal-Wallis-one way- ANOVA, ***Pearson chi-square
By post hoc Bonferroni test: * groups III and II, bgroups III and I, cgroups II and I

### Table 6
ApoE genotypes and alleles among ADMA tertiles

|                        | First tertile (0.12 - 0.54) (N = 199) | Second tertile (0.55 - 0.79) (N = 206) | Third tertile (0.80 - 4.91) (N = 207) | P       |
|------------------------|--------------------------------------|---------------------------------------|--------------------------------------|---------|
| ε3/ε2                  | 19 (9.9 %)                           | 21 (10.3 %)                          | 16 (7.8 %)                          | NS      |
| ε3/ε3                  | 123 (64.4 %)                         | 147 (72.4 %)                         | 158 (77.5 %)                        | NS      |
| ε4/ε2                  | 7 (3.7 %)                            | 2 (1.0 %)                            | 1 (0.5 %)                           | NS      |
| ε4/ε3                  | 37 (19.4 %)                          | 3 (1.0 %)                            | 25 (12.3 %)                         | NS      |
| ε4/ε4                  | 5 (0.8 %)                            | 3 (0.5 %)                            | 4 (0.7)                             | NS      |
| ε2 allele              | 26 (13.6 %)                          | 23 (11.3 %)                          | 17 (8.3 %)                          | NS      |
| ε3 allele              | 179 (93.7 %)                         | 197 (97.0 %)                         | 199 (97.5 %)                        | NS      |
| ε4 allele              | 49 (25.7 %)                          | 35 (17.2 %)                          | 29 (14.2 %)                         | 0.011   |

\*Pearson chi-square
and progression to CKD, has a strong relation with mutations in the ApoE gene [30]. An association between ESRD and ApoE polymorphism has been found in some but not all studies. Hubacek et al. found more carriers of the ε2 allele in ESRD patients (15.9 %) than in controls [28]. However, Roussos et al. [31] found that patients with ESRD showed no difference in ε2, ε3 and ε4 distribution compared with the control group. Feussner et al. also failed to find an association between ε2 and ESRD. [32] We did not find statistical differences between groups stratified by renal function, analyzing allele and genotype distribution.

When we accessed the relation between ADMA level and ApoE allele frequency among those patients in RRT, we observed that patients on hemodialysis and with the highest ADMA levels exhibited a significantly lower frequency of the ε4 allele when compared with that of patients in the sum of first and second tertiles of ADMA. This association remained significant even after correction for confounding factors. An interrelationship of ADMA and ApoE in atherogenesis was assessed in the work of Jacobi et al., in which mice deficient in ApoE and overexpressing DDAH showed reduced plaque formation in the aorta. A functional analysis of aortic ring preparations revealed improved endothelial function in mice overexpressing DDAH [33]. This association is interesting because it provides the patient with potentially greater endothelial dysfunction, represented by higher ADMA, and risk of cardiovascular complications the potential protection associated with the absence of the ε4 allele. Medina-Urrutia et al. [34] showed that the presence of the ε4 allele was associated with elevations in LDL-C, while the presence of ε2 was associated with decreased levels of LDL-C. Compared with individuals with the ε3/ε3 genotype, ε2 carriers have a 20 % lower risk of coronary heart disease and ε4 carriers have a slightly higher risk [35].

A recent study by our group in a cohort of hypertensive patients with metabolic syndrome, revealed a higher prevalence of CVD among ε4 allele carriers [36]. On the other hand, this association suggests that uremia is capable of determining lower plasma ADMA levels among ε4 allele carriers. ApoE ε4 allele expression could be limited by the presence of certain clinical variables, as can be seen in our study.

This study had some limitations. First, it was a cross-sectional study. The results of a study composed exclusively of hypertensive patients, conducted at a single center, cannot be extrapolated to the general population. It is descriptive and the mechanisms underlying this association cannot be inferred here. A greater number of participants in each renal function group could have allowed us to determine if this association between lower frequency of ApoE ε4 allele with higher level of ADMA also occurs in earlier stages of CKD. Genetic analysis methods (single nucleotide polymorphism) have limitations in the comprehension of the genotype - phenotype interface: these methods do not enable the simultaneous analysis of

Table 7 Logistic regression analysis of the association between ApoE ε4 allele frequency and third ADMA tertile, among patients in RRT

|                         | OR   | CI          | p     |
|-------------------------|------|-------------|-------|
| Adjusted for ApoE ε4 allele                        | 0.329| 0.155 - 0.699| 0.004|
| Adjusted for ApoE ε4 allele, age                       | 0.330| 0.155 - 0.700| 0.004|
| Adjusted for ApoE ε4 allele, age, race                  | 0.341| 0.162 - 0.719| 0.005|
| Adjusted for ApoE ε4 allele, age, race, sex             | 0.321| 0.148 - 0.697| 0.005|
| Adjusted for ApoE ε4 allele, age, race, sex, BMI         | 0.317| 0.146 - 0.689| 0.004|
| Adjusted for ApoE ε4 allele, age, race, sex, BMI, DM     | 0.263| 0.111 - 0.623| 0.002|
| Adjusted for ApoE ε4 allele, age, race, sex, BMI, DM, CVD | 0.270| 0.114 - 0.638| 0.003|
| Adjusted for ApoE ε4 allele, age, race, sex, BMI, DM, CVD, CRP | 0.271| 0.114 - 0.640| 0.003|
| Adjusted for ApoE ε4 allele, age, race, sex, BMI, DM, CVD, CRP, LDL-C | 0.283| 0.118 - 0.680| 0.005|
| Adjusted for ApoE ε4 allele, age, sex, BMI, DM, CVD, CRP, LDL-C, dialysis vintage | 0.283| 0.118 - 0.681| 0.005|

Fig. 1 Comparison of frequency of ApoE ε4 allele between ADMA tertiles 3 and 1 in renal function groups

* P = 0.008; Pearson chi-square

Marrocos et al. Lipids in Health and Disease (2016) 15:14 Page 6 of 9
multiple genes, like in genome wide association, hampering the search for a genetic signature for polygenetic diseases.

**Conclusions**
The presented results show, for the first time, that uremia is capable of determining lower plasma ADMA levels in hypertensive ε4 allele carriers. This association between lower frequency of ApoE ε4 allele and higher ADMA levels in ESRD may indicate a new way to approach atherogenesis in the inflammatory state of uremia. Confirmation of such observations needs to be validated in a longitudinal observation with a larger population study.

**Methods**
The study was approved by the Ethics and Research Committee (no. 311.413/06.21.2013, Ministry of Health, Brazil). All patients received relevant explanations and signed Ethics and Research Committee Informed Consent.

Six hundred twenty patients were stratified into 3 groups according to estimated GFR by CKD-EPI formula: group I > 60 mL/min, group II ≤ 60 mL/min and > 15 mL/min, and group III ≤ 15 mL/min or in hemodialysis. Exclusion criteria were as follows: aged under 18 years, clinical or laboratory suspicion of acute renal failure, pregnancy and cancer. Patient’s data were collected by clinical evaluation, physical examination, file reviews and laboratory tests. Presence of CVD was considered when identified in office or registry files: history of stroke, peripheral vascular disease, coronary artery disease or CHF. [37] Identification of DM was determined according to the American Diabetes Association Guidelines [38].

Plasma concentrations of glucose, total cholesterol and triglycerides were determined by automated enzymatic assays. HDL was measured in serum by the homogeneous method after precipitation of VLDL and LDL with phosphotungstate and magnesium ions, centrifugation and measurement of the supernatant absorbance at 500 nm wavelength, following the manufacturer’s instructions. LDL was calculated according to the Friedewald equation. Renal function was assessed by plasma creatinine level, determined by the Jaffé method with calibration traceable to an isotope dilution mass spectrometry reference measurement procedure [39]. CRP was measured by ultrasensitive immunoturbidimetry, following the manufacturer’s instructions.

Blood samples from subjects with ESRD undergoing hemodialysis was always obtained immediately before treatment.

**ADMA levels**
Plasma ADMA levels were measured by HPLC, as described by Teerlink et al. [40] Briefly, samples were prepared as follows: 200 μL of plasma containing EDTA or heparin were transferred to an Eppendorf tube (1.5 mL), and 100 μL of internal standard solution (40 μM monomethyl-larginine) were then added. PBS was added to complete the volume to 1 mL. This mixture was introduced into an OASYS extraction cartridge (Waters) coupled to a vacuum system previously equilibrated with 1 mL of methanol and 1 mL of deionized water. Next, the cartridge was rinsed with 1 mL of 100 mM HCl, followed by 1 mL of methanol, to elute neutral compounds and acids, and elution was performed with 1 mL of ammonia/water/methanol (10/40/50) solvent. The eluate recovered was dried at 60 °C in a speed-vacuum system, and the residue obtained was dissolved in 100 μL of water, followed by the addition of 100 μL of ortho-phthalaldehyde. After 15 min of reaction, the samples were transferred to appropriate HPLC vials.

We used a Symmetry C18 column (3.9 x 150 mm; 4 μm) coupled to a pre-column equilibrated with the same stationary phase. The mobile phase A consisted of 50 mM potassium phosphate buffer (pH 6.5) and mobile phase B (acetonitrile/water; 1/1, v/v). Samples (20 μL) were separated using an HPLC system with automatic injector. Standard solutions containing arginine (25, 50, 75, 100 and 150 μM) and ADMA (0.25, 0.50, 1.00, 2.5, 5.0 μM) and 40 μM of internal standard solution were extracted as described above to be injected before and after the injection of samples. The flow rate was 1.1 mL/min. The time interval between each injection was 30 min, and fluorescence was measured at emission and excitation wavelength of 340 and 455 nm, respectively.

**Analysis of ApoE gene polymorphisms**
The sequence of 244 base pairs of the ApoE gene was amplified by PCR, using the primers: sense 5’-TCCAAGGACCTGCAGGCCGCA-3’ and antisense 5’ACAGAATTCCGCCCGCCCTGTTACCTGCCA-3’. The PCR products were digested with the restriction enzyme HhaI, and the fragments were separated by 10 % polyacrylamide gel electrophoresis. Afterwards, the gel was incubated with ethidium bromide (0.5 μL/mL) for 90 min at 120 V in 1xTBE solution, and the DNA fragments were visualized with UV light. The ε2, ε3 and ε4 alleles were analyzed and the corresponding genotypes determined.

Groups were analyzed and compared with regard to epidemiological, clinical and laboratory characteristics as well as ApoE allele and genotype distribution. The prevalence of different alleles and genotypes were analyzed in relation
to race, where Hardy-Weinberg equilibrium was observed. Plasma ADMA levels were compared on the basis of RRT allocation ApoE polymorphism.

Statistical analysis
Results were expressed as mean ± SD or percentage. Comparisons of continuous variables among the three renal function groups were carried out with ANOVA and Kruskal-Wallis-one way-ANOVA, followed by post hoc Bonferroni tests. Comparisons of frequency between groups were performed using Pearson chi-square test.

Binary logistic regression analysis was used to test the association between ApoE ε4 allele frequency and endothelial dysfunction, in patients in RRT. The level of nullity was fixed at 0.05 or 5 % for all tests.

Abbreviations
ADMA: Asymmetric dimethylarginine; NOS: Nitric oxide synthase; ApoE: Apolipoprotein E; LDL-C: LDL cholesterol; GFR: Glomerular filtration rate; CKD-EPI: Chronic kidney disease epidemiology collaboration; PCR: Polymerase chain reaction amplification; HPLC: High performance liquid chromatography; CVD: Cardiovascular disease; CKD: Chronic kidney disease; CHF: Congestive heart failure; NQ: Nitric oxide; DM: Diabetes mellitus; CRP: C reactive protein; RRT: Renal replacement therapy; ESRD: End stage renal disease; DDAH: Dimethyl arginine dimethyl aminohydrolase; APCs: Angiogenic progenitor cells.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
MSMM and AAT contributed with clinical data collection; BMRQ, SdeMC and MCB contributed with laboratory experiments; CJOR and MCB contributed with study design; MCB, SM and MEC participated in its coordination; MSMM and MCB did statistical analysis and wrote the paper. All the authors read and approved the final manuscript.

Acknowledgements
We thank all the patients who very sympathetic and kindly participated in this study.

Financial support
This study was supported by Fundo de Amparo à Pesquisa do Estado de São Paulo/APESP.

Author details
1Nephrology Division, Universidade Federal de São Paulo, R. Pedro de Toledo, 781 14 andar, Vila Clementino, São Paulo, CEP 04039-032, Brazil.
2Nephrology Division-New England Medical Center, Tufts University, Boston, MA, USA. 3Research and Education Institute, Hospital Israelita Albert Einstein, São Paulo, São Paulo, Brazil.

Received: 21 September 2015 Accepted: 13 January 2016

References
1. Saran R, Li Y, Robinson B, Ayanian J, Barkshain R, Bragg-Gresham J, et al. U S renal data system 2014 annual data report: epidemiology of kidney disease in the United States. Am J Kidney Dis. 2015;65(Suppl 1):S1–S306. doi:10.1053/j.ajkd.2015.05.001.
2. Leone A, Moncada S, Vallance P, Calver A, Collier J. Accumulation of an endogenous inhibitor of nitric oxide synthesis in chronic renal failure. Lancet. 1992;339:572–5.
3. Kielstein JT, Frölich JC, Haller H, Fliser D. ADMA (asymmetric dimethylarginine): an atherosclerotic disease mediating agent in patients with renal disease? Nephrol Dial Transplant. 2001;16:1742–5. doi:10.1093/ndt/16.9.1742.
4. Kajimoto H, Kai H, Aoki H, Yasuoka S, Anegawa T, Aoki Y, et al. Inhibition of eNOS phosphorylation mediates endothelial dysfunction in renal failure: new effect of asymmetric dimethylarginine. Kidney Int. 2012;81:762–8. doi:10.1038/ki.2011.476.
5. Lu TM, Huang MY, Lin CC, Hsu CP, Lin SJ. Asymmetric dimethylarginine and clinical outcomes in chronic kidney disease. Clin J Am Soc Nephrol. 2011;6:1566–72. doi:10.2215/CINJ08409910.
6. Zoccali C, Bode-Böger S, Mallamaci F, Benedetto F, Tripepi G, Malatino L, et al. Plasma concentration of asymmetrical dimethylarginine and mortality in patients with end-stage renal disease: a prospective study. Lancet. 2001;358:2113–7.
7. Zhu ZD, Jia JQ, Zhang X, Wang YJ, Wang DH. Asymmetric dimethylarginine upregulates the expression of ACAT-1 in THP-1 macrophage-derived foam cells. J South Med Univ. 2010;30:2613–8.
8. Boerwinkle E, Visvikis S, Welsh D, Steinmetz J, Hanash SM, Sing CF. The use of measured genotype information in the analysis of quantitative phenotypes in man. II. The role of the apolipoprotein E polymorphism in determining levels, variability, and covariability of cholesterol, betalipoprotein, and triglycerides in a sample of unrelated individuals. Am J Med Genet. 1987;27:567–82.
9. Fiser D, Krennberg F, Kielstein JT, Morath C, Bode-Böger SM, Haller H, et al. Asymmetric dimethylarginine and progression of chronic kidney disease: the mild to moderate kidney disease study. J Am Soc Nephrol. 2005;16:2456–61. doi:10.1681/ASN.2005020179.
10. Ravani P, Tripepi G, Malberti F, Testa S, Mallamaci F, Zoccali C. Asymmetrical dimethylarginine predicts progression to dialysis and death in patients with chronic kidney disease: a competing risks modelling approach. J Am Soc Nephrol. 2005;16:2449–55.
11. Kielstein JT, Böger RH, Bode-Böger SM, Frölch JC, Haller H, Ritze E, et al. Marked increase of asymmetric dimethylarginine in patients with incipient primary chronic renal disease. J Am Soc Nephrol. 2002;13:170–8.
12. Uchida HA, Nakamura Y, Kihara M, Nori H, Harayama Y, Sugiyama H, et al. Steroid pulse therapy impaired endothelial function while increasing plasma high molecule adiponectin concentration in patients with lgA nephropathy. Nephrol Dial Transplant. 2006;21:3475–80. doi:10.1093/ndt/gfl423.
13. Ito A, Tsao PS, Adimoolam S, Kimoto M, Ogawa T, Cooke JP. Novel mechanism for endothelial dysfunction: dysregulation of dimethylarginine dimethylaminohydrolase. Circulation. 1999;99:3092–5. doi:10.1161/01.CIR.99.24.3092.
14. Matusuoka K, Ueda S, Yamagishi S, Matsumoto Y, Kaneyuki U, Shibata R, et al. Molecular mechanism for elevation of asymmetric dimethylarginine and its role for hypertension in chronic kidney disease. J Am Soc Nephrol. 2006;17:2176–83. doi:10.1681/ASN.2005121379.
15. Schwedhelm E, Böger RH. The role of asymmetric and symmetric dimethylarginines in renal disease. Nat Rev Nephrol. 2011;7:275–85. doi:10.1038/nrneph.2011.31.
16. Achan V, Broadhead M, Malak M, Whitley G, Leiper J, MacAllister R, et al. Asymmetric dimethylarginine causes hypertension and cardiac dysfunction in humans and is actively metabolized by dimethylarginine dimethylaminohydrolase. Arterioscler Thromb Vasc Biol. 2003;23:1455–9. doi:10.1161/01.ATV.0000081742.92006.59.
17. Sibal L, Agarwal SC, Home PD, Böger RH. The role of asymmetric dimethylarginine (ADMA) in endothelial dysfunction and cardiovascular disease. Curr Cardiol Rev. 2010;6:82–90. doi:10.2174/157340310791162659.
18. Fleissner F, Jazbutyte V, Fiedler J, Gupta SK, Yin X, Xu Q, et al. Short communication: asymmetric dimethylarginine impairs angiogenic progenitor cell function in patients with coronary artery disease through a microRNA-21–dependent mechanism. Circ Res. 2010;107:138–43. doi:10.1161/CIRCRESAHA.110.216770.
19. Kielstein JT, Böger RH, Bode-Böger SM, Schäffer J, Barbey M, Koch KM, et al. Marked increase of asymmetric dimethylarginine in patients with kidney disease. Nephrol Dial Transplant. 2006;21:2176–83. doi:10.1093/ndt/gfl423.
20. Lu TM, Chang MY, Lin CC, Hsu CP, Lin SJ. Asymmetric dimethylarginine: an atherosclerotic disease mediating agent in patients with renal disease? Nephrol Dial Transplant. 2001;16:1742–5. doi:10.1093/ndt/16.9.1742.
early diabetic nephropathy in type 1 diabetes. Diabetes Care. 2004;27:765–9. doi:10.2337/diabetes.27.3.765.

23. Zoccali C, Benedetto FA, Maas R, Mallamaci F, Tripepi G, Malatino LS, et al. Asymmetrical dimethylarginine, C-reactive protein, and carotid intima-media thickness in end-stage renal disease. J Am Soc Nephrol. 2002;13:490–6.

24. Zhou TB, Qin YH, Xu HL. Association of apoE gene expression and its gene polymorphism with nephrotic syndrome susceptibility: a meta-analysis of experimental and human studies. Mol Biol Rep. 2012;39:9347–54. doi:10.1007/s11033-012-1751-4.

25. Chu AY, Parekh RS, Astor BC, Coresh J, Berthier-Schaad Y, Smith MW, et al. Association of apoE polymorphism with chronic kidney disease in a nationally representative sample: a Third National Health and Nutrition Examination Survey (NHANES III) Genetic Study. BMC Med Gen. 2009;10:108. doi:10.1186/1471-2350-10-108.

26. Liberopoulos E, Siamopoulos K, Elsaf M. Apolipoprotein E and renal disease. Am J Kidney Dis. 2004;43:223–33.

27. Xue C, Nie W, Tang D, Yi L, Mei C. Apolipoprotein E gene variants on the risk of end stage renal disease. PLoS ONE. 2013;8(12), e83367. doi:10.1371/journal.pone.0083367.

28. Hubacek JA, Bloudickova S, Kubinova R, Pikhart H, Viklicky O, Bobak M. Apolipoprotein E polymorphism in hemodialysed patients and healthy controls. Biochem Genet. 2009;47:688–93. doi:10.1007/s10528-009-9266-y.

29. Zhou TB, Qin YH, Xu HL. Association of apoE gene expression and its gene polymorphism with nephrotic syndrome susceptibility: a meta-analysis of experimental and human studies. Mol Biol Rep. 2012;39:9347–54. doi:10.1007/s11033-012-1751-4.

30. Hu Z, Huang S, Wu Y, Liu Y, Liu X, Su D, et al. Hereditary features, treatment, and prognosis of the lipoprotein glomerulopathy in patients with the APOE Kyoto mutation. Kidney Int. 2014;85:416–24. doi:10.1038/kj.2013.335.

31. Roussos L, Ekstrom U, Ehle PN, Oqvist B, Floren CH. Apolipoprotein E gene expression and its gene polymorphism in 385 patients on renal replacement therapy in Sweden. Scand J Urol Nephrol. 2004;38:504–10.

32. Feussner G, Wey S, Bommer J, Deppermann D, Grützmacher P, et al. Apolipoprotein E phenotypes and hyperlipidemia in patients under maintenance hemodialysis. Hum Genet. 1992;88:307–12.

33. Rouslos L, Ekstrom U, Ehle PN, Oqvist B, Floren CH. Apolipoprotein E polymorphism in 385 patients on renal replacement therapy in Sweden. Scand J Urol Nephrol. 2004;38:504–10.

34. Jacobs J, Maas R, Cardounel AJ, Arend M, Pope AJ, Cordasic N, et al. Dimethylarginine dimethylaminohydrolase overexpression ameliorates atherosclerosis in apolipoprotein E-deficient mice by lowering asymmetric dimethylarginine. Am J Pathol. 2010;176:2559–70. doi:10.2353/ajpath.2010.090614.

35. Medina-Urrutia AX, Cardoso-Saldana GC, Zamora-Gonzalez J, Liria YK, Posadas-Romero C. Apolipoprotein E polymorphism is related to plasma lipids and apolipoproteins in Mexican adolescents. Hum Biol. 2004;76(4):605–14.

36. Bennett AM, Di Angelantonio E, Maas R, Mallamaci F, Tripepi G, Malatino LS, et al. Asymmetric dimethylarginine, C-reactive protein, and carotid intima-media thickness in end-stage renal disease. J Am Soc Nephrol. 2002;13:490–6.

37. Levey AS, Stehman-Breen CM, Kusek JW, Liang JM, Beck GJ, Coresh J, et al. A new equation to estimate glomerular filtration rate. Ann Intern Med. 2009;150(9):604–12.

38. Cannon CP, Brindis RG, Chair CM, Cohen DJ, Cross Jr. JT, Drozda JP, et al. 2013 ACCF/AHA key data elements and definitions for measuring the clinical management and outcomes of patients with acute coronary syndromes and coronary artery disease: A report of the American College of Cardiology Foundation/American Heart Association Task Force on Clinical Data Standards (Writing Committee to Develop Acute Coronary Syndromes and Coronary Artery Disease Clinical Data Standards). J Am Coll Cardiol. 2013;61:992–1025. doi:10.1016/j.jacc.2012.10.005.

39. American Diabetes Association. Diagnosis and classification of diabetes mellitus. Diabetes Care. 2014;37 Suppl 1:S18–90. doi:10.2337/dc13-S002.

40. Levey AS, Stevens LA, Schmid CH, Zhang YL, Castro AF, Feldman HI, et al. A new equation to estimate glomerular filtration rate. Ann Intern Med. 2009;150(9):604–12.