Numb inhibits epithelial-mesenchymal transition via RBP-Jk-dependent Notch1/PTEN/FAK signaling pathway in tongue cancer

Jin-Yun Li*, Wen-Xiao Huang, Xiao Zhou, Jie Chen and Zan Li*

Abstract

Background: Oral cancer has been estimated as the sixth most frequent solid cancer all over the world, in which tongue squamous cell carcinoma (TSCC) is the most common type of oral cancers. However, the mechanism of TSCC metastasizing to lymph node and distant sites has not been completely understood.

Methods: In this study, RT-qPCR method was used to detect the mRNA level of Numb, PTEN and Notch1 genes, as well as EMT-associated genes. Western blot assay was utilized to detect protein level of these genes. In addition, we determined cell proliferation by MTT assay and employed transwell invasion assay and wound healing assay to probe the abilities of invasion and migration, respectively. To investigate the role of PTEN, its inhibitor VO-Ohpic trihydrate was used to treat SCC-4 and CAL27 cells.

Results: We found that Numb expression was downregulated in SCC-9 and CAL-27 cells compared to NHOK cells. Instead, Notch1 level in SCC-9 and CAL-27 cells were higher than that in NHOK cells. Furthermore, the results showed that Numb overexpression significantly suppressed proliferation, migration and invasion of SCC-9 and CAL-27 cells via regulating Notch1 signaling and EMT-related genes expression. By contrast, we observed that RBP-Jk knockdown had an inhibitory role in proliferation, migration and invasion of SCC-9 and CAL-27 cells. In cells with Numb overexpression or RBP-Jk knockdown, p-FAK and EMT-related genes were remarkably regulated.

Conclusions: Our findings provide new mechanism of understanding the metastasis of TSCC and help develop therapeutic strategies for treating tongue cancer.

Keywords: Tongue squamous cell carcinoma (TSCC), Numb, Notch1 signaling, PTEN, Epithelial-mesenchymal transition (EMT)

Background

Oral cancer accounts for about 1–3% of all human cancer cases, which is the 6th most occurred cancer in the world. Tongue squamous cell carcinoma (TSCC) is the most common type of oral cancer [1–3]. Most importantly, lymph node and distant metastasis are the most adverse prognostic factors and will cause the death of TSCC patients [4, 5]. In spite of large advances in understanding how tongue cancer initiates and progresses, the number of deaths of TSCC patients increased by approximately 10% over the past 5 years [6, 7]. Thus, it remains urgent to further clarify the molecular mechanism of carcinogenesis and metastasis of tongue cancer.

The Notch1 signaling pathway is an evolutionarily conserved pathway, which has been involved in a wide variety of physiological and pathological processes, including cell fate determination, cell differentiation, tissue patterning and morphogenesis, and various types of cancer [8–10]. It has been studied that Notch1 signaling pathway participates in invasion of TSCC via regulating matrix metalloproteinases (MMPs) [11]. Therefore, we...
attempted to figure out the detailed mechanism of how Notch1 signaling and its regulators (like Numb) contributes to progression and metastasis of tongue cancer.

Phosphatase and tensin homolog (PTEN), commonly regarded as the tumor suppressor, is reported to regulate cancer cell invasion through blocking a variety of signaling pathways [12, 13]. In TSCC tissue, PTEN expression was lower than that in noncancerous counterparts, which suggested PTEN might act as tumor suppressor of TSCC [14, 15]. More intriguingly, Siming Xie et al. reported that PTEN upregulation suppresses invasion of tongue cancer cells through repression of EMT, which provided direct evidence that PTEN played tumor inhibitory role in progression and metastasis of tongue cancer [16].

Here, we reveal Numb, negative regulator of Notch1 signaling pathway, plays an inhibitory role in EMT of tongue cancer cells via regulating Notch1/RBP-Jκ/PTEN/p-FAK axis. In this study, we confirmed the crucial role of RBP-Jκ in EMT of tongue cancer cells. We believed that these findings will facilitate the development of targeted drugs used for treating tongue cancer.

**Methods**

**Cell lines and reagents**

Tongue squamous cancer cell lines (SCC-9 and CAL-27) and 293 T cell were from American Tissue Culture Collection (ATCC, USA) and cultured in Dulbecco’s Modified Eagle’s Medium (DMEM, Hyclone) with 10% fetal bovine serum (FBS) and 100 U/mL penicillin/streptomycin at 37°C, 5% CO2. Of which, SCC-9 cells were grown in 1:1 Hams F-12, DMEM (modified). Primary normal human oral keratinocytes (NHOK), purchased from PriCells, were isolated from human gingival tissues and cultured as described previously [17]. PTEN inhibitor VO-Ohpic trihydrate was purchased from MedChemExpress (USA) and worked at the concentration of 35 nM.

**Plasmid and lentivirus packaging**

Numb cDNA sequence was cloned and ligated into pCDH-MCS-EF1-puro vector. RBP-Jκ short hairpin RNA (shRNA-RBP-Jκ) or negative control shRNA (shRNA-NC) were inserted into PLKO.1 vector. In order to make lentiviruses, pPAX2 and pVSVG vectors plus transfer vector were introduced into 293 T cells. Then, we harvested the supernatant at 48 h post transfection, which was filtered with 0.45 μm membrane and concentrated by a centrifugal filter (EMD Millipore, Amicon Ultra 100 k). When subject to virus infection, the cells medium was treated with virus supernatant at 1:5 ratios, post 24 h, we used ~ 2 μg/mL puromycin to select the Numb-overexpressed stable cell lines. The shRNA sequences are synthesized as previously described [18].

**Western blot**

We performed this experiment as standard procedure in accordance with previous descriptions [8]. The following antibodies were used: anti-E-cadherin (cell signaling technology, USA), anti-N-cadherin (cell signaling technology, USA), anti-Snail (cell signaling technology, USA), anti-MMP-9 (cell signaling technology, USA), anti-Numb (cell signaling technology, USA), anti-Notch1 (cell signaling technology, USA), anti-FAK (cell signaling technology, USA), anti-p-FAK (cell signaling technology, USA), anti-PTEN (cell signaling technology, USA), anti-RBP-Jκ (millipore, USA), anti-GAPDH (Proteintech, USA).

**Invasion assay**

To perform this assay, TSCC (1 × 105 cells) were mixed with about 200 μL of FBS-free medium. Chambers containing 8.0 μm pore membranes (Millipore) with matrix was used in this assay. The TSCC cells were seeded into the top chamber. Afterwards, about 500 μL complete medium was added to bottom chamber. Roughly 48 h post incubation, the invaded cells were fixed and stained with crystal violet, and finally were counted using the inverted microscope.

**Wound healing assay**

We measured migration of cells in vitro through wound healing assay. In brief, 2 × 105 SCC-9 and CAL-27 cells were passaged to 6-well plates. The cells were incubated in the complete culture medium under normal conditions for 16 h. then, we scratched the monolayer and incubated cells in fresh medium without FBS for 24 h. Eventually, we measured the wound width. we visualized and photographed 3 different locations under the inverted microscope.

**RT-qPCR**

We harvested cells and extracted total RNAs by Trizol method. Then, chloroform was added to the solution. The sample was subject to centrifuge at 12,000 rpm for 10 min, and was transferred to new RNase-free EP tube. Resulting solution was mixed with equal volume of isopropanol and subject to centrifuge at 12,000 rpm for 10 min. After that, removing the supernatant and adding 70% ethanol to wash pellet. Eventually, discarding the ethanol and drying the RNA pellet. And, we used 35 μL Rnase-free H2O to dissolve RNA.

We employed ~ 1 μg of RNA for reverse transcription. SYBR dye was used to detect signaling, the GAPDH serves as internal control. The primers used in this study as follows:

Numb-F: TCTGCTCCGATGACCAAACC
Numb-R: GCACCAGAAGATTGACCCCCA
Notch1-F: CAACTGCCAGAACCTTGTGC
Notch1-R: GGCAACGTCAACACCTTGTC
GAPDH-F: GAGTCAACGGATTTGGTCGT
GAPDH-R: TTGATTTTGGAGGGATCTCG

Statistical analysis
All experiments were conducted for three replicates, all values were represented as mean ± SD, comparisons of two groups were done using two-tailed unpaired student’s t-test. *P < 0.05 was considered statistically significant.

Results
Numb and Notch1 expressions were altered in tongue squamous cancer cell lines
Recently, it has been studied that Notch1 signaling is implicated in progression of tongue cancer [8]. To validate, we examined the expression of Numb (Notch1 regulator) and Notch1 in tongue squamous cancer cell lines (SCC-9 and CAL-27) and normal human oral keratinocytes (NHOK) by RT-qPCR method. We found that Numb expression level was significantly decreased in SCC-9 and CAL-27 cells compared with that in NHOK cells (Fig. 1a). By contrast, the level of Notch1 was increased by about 2 folds in SCC-9 and CAL-27 cells (Fig. 1b). These data suggested that, indeed, Numb and Notch1 signaling were implicated into tongue cancer progression.

Numb overexpression inhibits proliferation, invasion and migration of SCC-9 and CAL-27
To further confirm the anti-tumor role of Numb in tongue cancer, we constructed Numb-overexpressed SCC-9 and CAL-27 cell lines. First, we used MTT assay to evaluate SCC-9 and CAL-27 cells growth and found that cell proliferation ability of cells were markedly suppressed upon Numb overexpression compared to negative control (NC) (Fig. 2a).

Next, we assessed cell migration by wound healing assay when the cells transfected with empty vector or Numb at the time points of 0, 24, 48 h. The results demonstrated that Numb-overexpressed SCC-9 and CAL-27 cells displayed impaired migration ability relative to NC group cells (Fig. 2b and c). In addition, we also investigated the role of Numb in invasion of tongue cancer cells (SCC-9 and CAL-27) using transwell invasion experiment at 24 h. Our data revealed that the number of invaded tongue cancer cells expressing Numb was

---

Fig. 1 Expression of Numb and Notch1 in tongue cancer cell lines (CAL-27 and SCC-9) and normal human oral keratinocytes (NHOK). a RT-qPCR analysis shows the level of Numb in SCC-9, CAL-27 and NHOK cells. b RT-qPCR analysis shows mRNA level of Notch1 in SCC-9, CAL-27 and NHOK cells. The data were represented as mean ± SD from 3 biological replicates (*p < 0.05; **p < 0.01)
remarkably lower than NC group cells (Fig. 2d and e). Collectively, these results indicated that Numb had an inhibitory role in tongue cancer proliferation and metastasis.

**Numb regulates Notch1 signaling and EMT-associated genes expression**

In order to dissect the molecular mechanism Numb-suppressed proliferation and tongue cancer cells EMT, we probed expression of Notch1 and EMT-associated genes (Fig. 3a and b). Western blot analyses demonstrated that Notch1 expression level was decreased in Numb-overexpressed SCC-9 and CAL-27 cells. More importantly, we observed that EMT-inhibited genes (E-cadherin) expression level was upregulated, whereas RBP-Jκ and EMT-promoted genes (N-cadherin, Snail, MMP-9) expression level was downregulated. These data showed Numb had a critical role in regulation of Notch1 signaling, RBP-Jκ and EMT-associated genes expression, which is more likely to mediate Numb-inhibited EMT of tongue cancer cells.

**PTEN inhibition leads to increased expression of EMT-associated genes expression**

To explore the role of PTEN in regulation of EMT-associated genes expression in tongue cancer cell lines (SCC-9 and CAL-27), we utilized PTEN inhibitor

---

Fig. 2 Numb overexpression inhibits proliferation, invasion and migration of SCC-9 and CAL-27 cells. a Cell proliferation assay and quantification of NC (mock) or Numb-overexpressed SCC-9 and CAL-27 cells by MTT assay at 48 h after transfection. NC denotes cells transfected with empty vector, Numb denotes cells transfected with pCDH-Numb. b and c Wound healing assay and quantification shows the migration ability of NC (mock) or Numb-overexpressed SCC-9 and CAL-27 cells. The photographs were taken at the time points of 0, 24, 48 h. Magnification ×10(5000 μm) in wound healing assay. d and e Transwell assay and quantification shows the invasion ability of NC (mock) or Numb-overexpressed SCC-9 and CAL-27 cells. The photographs were taken at 24 h. Magnification× 50(2000 μm) in transwell assay. The data were represented as mean ± SD from 3 biological replicates (*p < 0.05; **p < 0.01; ***p < 0.001)
VO-Ohpic trihydrate to abolish the function of PTEN and then detected expression level of EMT-associated genes (Fig. 4a and b). As shown in the western blot, PTEN expression was suppressed by VO-Ohpic trihydrate in Numb-overexpressed SCC-9 and CAL-27 cells. However, phosphorylated form of FAK was elevated in the presence of VO-Ohpic trihydrate. In addition, the result showed VO-Ohpic trihydrate treatment disrupted EMT-related genes expression level modulated by Numb. Together, our data suggested that PTEN inhibitor VO-Ohpic trihydrate could effectively block Numb-inhibited EMT genes expression in SCC-9 and CAL-27 cells.

Next, we attempted to assess whether RBP-Jk was key for proliferation and metastasis of tongue cancer cells. We generated RBP-Jk-depleted tongue cancer cell lines (SCC-9 and CAL-27) using short hairpin RNA approach. Then, MTT experiment was carried out to detect cell proliferation of SCC-9 and CAL-27 cells. We found that RBP-Jk knockdown SCC-9 and CAL-27 cells grew more slowly than NC cells (Fig. 5a).

To test whether RBP-Jk had a role in invasion and migration of tongue cancer cells, we carried out transwell
invasion assay and wound healing assay to evaluate the invasion and migration ability of the cells, respectively. We observed that RBP-Jκ knockdown resulted in impaired migration capability of SCC-9 and CAL-27 cells (Fig. 5b and c). In parallel, fewer numbers of invaded cells (decreased by 30–40%) were stained by crystal violet when the cells were transfected with RBP-Jκ shRNA compared to NC cells at 24 h (Fig. 5d and e). In summary, all these results revealed that RBP-Jκ was important for proliferation, migration and invasion of tongue cancer cells (SCC-9 and CAL-27).

RBP-Jκ knockdown regulates PTEN and EMT-associated genes expression
To clarify the underlying molecular mechanism of RBP-Jκ knockdown-caused proliferation and metastasis change of tongue cancer cells, we attempted to probe PTEN, FAK and EMT-related genes expression after RBP-Jκ knockdown in SCC-9 and CAL-27 cells (Fig. 6a and b). Western blot analyses demonstrated that PTEN and EMT-inhibited gene (E-cadherin) expression level were upregulated in RBP-Jκ knockdown cells, on the other hand, p-FAK and EMT-promoted genes (N-cadherin, Snail, MMP-9) expression level was downregulated upon RBP-Jκ depletion. Therefore, we concluded that RBP-Jκ was a key regulator of PTEN/FAK/EMT-related genes pathway.

PTEN inhibition leads to increased expression of EMT-associated genes expression with abatement of RBP-Jκ
We next attempted to figure out whether PTEN inhibitor also influenced expression of p-FAK and EMT-related genes in the tongue cancer cells with RBP-Jκ knockdown (Fig. 7a and b). Western blot analyses showed PTEN and E-cadherin was modestly downregulated in RBP-Jκ knockdown cells treated with VO-Ohpic trihydrate. Nonetheless, p-FAK and EMT-promoted genes (N-cadherin, Snail, MMP-9) expression level were modestly
upregulated in response to VO-Ohpic trihydrate treatment. Our data indicated that PTEN inhibition had the ability to partially rescue RBP-Jκ knockdown-induced changes in expression of p-FAK and EMT-regulated genes.

Discussion
Recently, Notch1 has been shown to exert its oncogenic role in various types of cancers, including lung, colorectal, T-cell acute lymphoblastic leukemia, breast and prostate carcinomas [19, 20]. However, the detailed mechanism by which Notch1 regulates progression and metastasis of tongue cancer remains unclear. Indeed, Notch1 signaling is activated in human tongue carcinoma [21]. In our study, we reveal the critical role of Numb and RBP-Jκ in regulating EMT of SCC-9 and CAL-27 cells, thereby highlighting the existence of Numb/Notch1/RBP-Jκ/PTEN/FAK/EMT axis in tongue cancer cells.

Epithelial-mesenchymal transition (EMT), a key prerequisite for individual development and cancer cells metastasis, can be initiated or controlled by developmental and various environmental cues [22]. In breast and other
cancers, Notch1 signaling induced EMT phenotype by up-regulating EMT-promoted genes (Slug and Snail) expression, then downregulated E-cadherin [23, 24]. Consistent with previous studies, we observed that Notch1 also had the ability to regulate EMT-related genes expression in tongue cancer cells. In addition, we found Notch1 influenced EMT of tongue cancer through its downstream effector PTEN and RBP-\( \kappa \). These results provided more insight into the mechanism of Notch1-induced EMT.

Numb is the human homologue of the protein numb, initially discovered in \textit{Drosophila melanogaster}, well-known for its multifaceted role in neurogenesis [25]. The antagonistic role of Numb in the Notch1 pathway led researchers to investigate the potential role of Numb in tumorigenesis in a number of tumors [26]. However, to date, little is known about whether Numb had an effect on EMT of tongue cancer cells. Our study, for the first time, investigate and clarify the inhibitory role of Numb in proliferation, invasion and migration of tongue cancer cells through negatively regulating Notch1 signaling and modulating EMT-related genes expression.

In order to elucidate the downstream mechanism of Notch1-induced EMT of tongue cancer, we focused on tumor suppressor PTEN, a reported target of Notch1 via CBP-1 binding to PTEN DNA promoter [27]. It is well established that PTEN loss is a prognostic marker and contributes to development of tongue cancer [28]. We also confirmed the critical role of PTEN in Numb overexpression- and RBP-\( \kappa \) knockdown-caused changes in proliferation and metastasis of tongue cancer cells by employing PTEN inhibitor VO-Ohpic trihydrate. Besides, we further explored and confirmed that PTEN exerted its role through regulating the activity of FAK (p-FAK level), thereby affecting expression of EMT-associated genes.

**Conclusions**

In summary, we propose that Numb, negative regulator of Notch1 signaling, plays a suppressive role in proliferation and metastasis of tongue cancer cells. Mechanistically,
Notch1 further regulates PTEN via in a RBP-Jκ-dependent manner to impact activity of FAK that is essential for EMT phenotype of tongue cancer cells. Nonetheless, the further investigation of FAK importance in EMT of tongue cancer cells requires to be undertaken by using FAK inhibitor or shRNA.

Abbreviations
EMT: Epithelial-mesenchymal transition; FBS: Fetal bovine serum; MMPs: Matrix metalloproteinases; NC: Negative control; NHOK: Normal human oral keratinocytes; PTEN: Phosphatase and tensin homolog; TSCC: Tongue squamous cell carcinoma

Acknowledgements
We would like to give our sincere gratitude to the reviewers for their constructive comments.

Funding
Not applicable.

Availability of data and materials
All data generated or analysed during this study are included in this published article [and its supplementary information files].

Authors' contributions
LJY designed the study, prepared and edited the manuscript. HWX and ZX performed experimental studies and acquired the data. CJ did literature research and analyzed the data. LZ prepared and reviewed the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate
Not applicable.

Consent for publication
Not applicable.
Competing interests
The authors declare that they have no competing interests.

Publisher's Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 19 July 2018 Accepted: 12 April 2019
Published online: 25 April 2019

References
1. Jemal A, Bray F, Center MM, et al. Global cancer statistics. CA Cancer J Clin. 2011;61:69–90.
2. Warnakulasuriya S. Global epidemiology of oral and oropharyngeal cancer. Oral Oncol. 2009;45:309–16.
3. Kawakita A, Yamamoto S, Yamada S, et al. MicroRNA-21 promotes oral cancer invasion via the Wnt/beta-catenin pathway by targeting DKK2. Pathol Oncol Res. 2014;20:253–61.
4. Ho CM, Lam KH, Wei W, et al. Occult lymph node metastasis in small oral tongue cancers. Head Neck. 1992;14:359–63.
5. Sprio RH, Huvos AG, Wong GY, et al. Predictive value of tumor thickness in squamous carcinoma confined to the tongue and floor of the mouth. Am J Surg. 1986;152:345–50.
6. Yu X, Li Z. MicroRNA expression and its implications for diagnosis and therapy of tongue squamous cell carcinoma. J Cell Mol Med. 2016;20:10–6.
7. Ko SY, Lin SC, Wong YK, et al. Increase of disintegrin metalloprotease 10 (ADAM10) expression in oral squamous cell carcinoma. Cancer Lett. 2007;245:33–43.
8. Yu B, Wei J, Qian X, et al. Notch1 signaling pathway participates in cancer invasion by regulating MMPs in lingual squamous cell carcinoma. Oncol Rep. 2012;27:547–52.
9. Yoshida R, Nagata M, Nakayama H, et al. The pathological significance of Notch1 in oral squamous cell carcinoma. Lab Invest. 2013;93:1068–81.
10. Upadhyay P, Nair S, Kaur E, et al. Notch pathway activation is essential for maintenance of stem-like cells in early tongue cancer. Oncotarget. 2016;7:50437–50449.
11. Delbosc S, Glorian M, Le Port AS, et al. The benefit of docosahexaenoic acid on the migration of vascular smooth muscle cells is partially dependent on Notch regulation of MMP-2/9. Am J Pathol. 2008;172:1480–40.
12. Song MS, Salmena L, Pandolff PP. The functions and regulation of the PTEN tumour suppressor. Nat Rev Mol Cell Biol. 2012;13:283–96.
13. Chu EC, Tamawski AS. PTEN regulatory functions in tumor suppression and cell biology. Med Sci Monit. 2004;10:RA235–41.
14. Xie SM, Shen LJ, Yin C, et al. Expression of tumor suppressor gene PTEN, PIP3 and cyclin D1 in oral squamous cell carcinoma and their correlations. Zhonghua Kou Qiang Yi Xue Za Zhi. 2006;41:407–10.
15. Squarize CH, Castillo RH, Abrahao AC, et al. PTEN Deficiency Contributes to the Development and Progression of Head and Neck Cancer. Neoplasia. 2013;15:461–71.
16. Xie S, Lu Z, Lin Y, et al. Upregulation of PTEN suppresses invasion in Tca8113 tongue cancer cells through repression of epithelial-mesenchymal transition (EMT). Tumour Biol. 2016;37:6681–9.
17. Min BM, Lee G, Kim SH, et al. Electrospinning of silk fibroin nanofibers and its effect on the adhesion and spreading of normal human keratinocytes and fibroblasts in vitro. Biomaterials. 2004;25:1289–97.
18. Lake RJ, Tsai PF, Choi I, et al. RBPJ, the major transcriptional effector of Notch signaling, remains associated with chromatin throughout mitosis, suggesting a role in mitotic bookmarking. PLoS Genet. 2014;10:e1004204.
19. Allemann K, Maillard J, Aster JC, Pear WS. Notch signaling in cancer. Cancer Biol Ther. 2002;1:465–76.
20. Nickoloff BJ, Osborne BA, Miele L. Notch signaling as a therapeutic target in cancer: a new approach to the development of cell fate modifying agents. Oncogene. 2003;22:6598–608.
21. Zhang TH, Liu HC, Zhu LJ, et al. Activation of Notch signaling in human tongue carcinoma. J Oral Pathol Med. 2011;40:37–45.
22. Moustakas A, Heldin CH. Signaling networks guiding epithelial-mesenchymal transitions during embryogenesis and cancer progression. Cancer Sci. 2007;98:1512–20.
23. Zhang X, Zhao X, Shao S, et al. Notch1 induces epithelial-mesenchymal transition and the cancer stem cell phenotype in breast cancer cells and STAT3 plays a key role. Int J Oncol. 2015;46:1141–8.