Non-Mono-Exponential Analysis of Diffusion-Weighted Imaging for Treatment Monitoring in Prostate Cancer Bone Metastases

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Diffusion-weighted imaging quantified using the mono-exponential model has shown great promise for monitoring treatment response in prostate cancer bone metastases. The aim of this prospective study is to evaluate whether non-mono-exponential diffusion models better describe the water diffusion properties and may improve treatment response assessment. Diffusion-weighted imaging data of 12 treatment-naïve patients with 34 metastases acquired before and at one, two, and three months after initiation of antiandrogen treatment are analysed using the mono-exponential, the intravoxel incoherent motion, the stretched exponential, and the statistical model. Repeatability of the fitted parameters and changes under therapy are quantified. Model preference is assessed and correlation coefficients across times are calculated to delineate the relationship between the prostate-specific antigen levels and the diffusion parameters as well as between the diffusion parameters within each model. There is a clear preference for non-mono-exponential diffusion models at all time points. Particularly the stretched exponential is favoured in approximately 60% of the lesions. Its parameters increase significantly in response to treatment and are highly repeatable. Thus, the stretched exponential may be utilized as a potential optimal model for monitoring treatment response. Compared with the mono-exponential model, it may provide complementary information on tissue properties and improve response assessment.

Advanced prostate cancer metastasizes to the bones with a frequency of more than 90%1. Up to 80% of the patients that are treated with antiandrogen therapy initially respond well with a progression-free interval of 23–37 months2. After this time, patients develop resistance to androgen deprivation, which inevitably leads to tumour progression3–5. Standard imaging techniques such as technetium-99 m bone scintigraphy and computed tomography that are used to assess the presence of bone metastases fail to accurately evaluate biological activity over time6. As a consequence, the response evaluation criteria of solid tumours (RECIST) consider bone metastases without associated soft-tissue masses as non-measurable7. For this reason, evaluation of treatment response in clinical practice primarily relies on measurements of the prostate-specific antigen (PSA) level, which has not been proven to be a surrogate biomarker for improved survival8–10. Thus, there is an unmet need for non-invasive biomarkers that allow assessment of treatment response in bone metastases.

Several studies have demonstrated the potential of diffusion-weighted imaging (DWI) quantified by the apparent diffusion coefficient (ADC) from the mono-exponential diffusion model for evaluating response to therapy in bone metastases from prostate cancer11–15. Thereby, increased values of the mean and median ADCs were observed after commencement of therapy relative to pre-treatment values. Furthermore, the results of a pilot study11–13 showed that the ADC values increased in response to therapy, which suggested a potential role of DWI in monitoring treatment response in bone metastases.
The ADC is a simple and robust quantitative biomarker but it draws an incomplete picture of the molecular motion in both healthy and pathological tissue since signal attenuation in DWI is not completely described by a mono-exponential process. For this reason, several studies have recently investigated the use of more complex non-mono-exponential diffusion models for the characterization of tumour masses17−28 and the evaluation of treatment response29, 30. The additional parameters in these models may better describe the water diffusion behaviour in bone metastases and provide complementary information on tissue properties, thus enabling more sensitive evaluation of changes in response to treatment.

Table 1. Average pre-treatment values of the various diffusion parameters (all units mm²/s except where shown) as well as inter-patient variability, and coefficient of variation as percentage.

| Model       | Parameter | Average baseline value (10⁻³ mm²/s) | Inter-patient variability (%) | Coefficient of variation (%) |
|-------------|-----------|-------------------------------------|-------------------------------|-----------------------------|
| Mono-exponential | ADC       | 0.76                                | 15.1                          | 4.4                         |
|             | ADC*      | 0.62                                | 14.1                          | 5.0                         |
| IVIM        | D         | 0.57                                | 14.9                          | 7.8                         |
|             | D*        | 0.81                                | 60.0                          | 42.5                        |
|             | f (no units) | 0.14                                | 27.5                          | 19.8                        |
|             | fD*       | 1.34                                | 51.0                          | 20.4                        |
| Stretched exponential | DDC       | 0.65                                | 18.6                          | 5.1                         |
|             | α (no units) | 0.76                                | 12.3                          | 5.0                         |
| Statistical | D₁*       | 0.97                                | 19.9                          | 6.4                         |
|             | σ*        | 0.87                                | 24.1                          | 10.0                        |

Results

Pre-treatment values and measurement repeatability. The average pre-treatment values of the various diffusion parameters are summarized in Table 1. With regard to the mono-exponential model, the results corroborate well with literature values11−15, 41. Two-sided paired t-tests showed no systematic differences between the pre-treatment values (p > 0.3 for all diffusion parameters). Inter-patient variability of the parameters was below 20% for all diffusion parameters with the exception of σ* from the statistical model, and f, fD*, and D* from the IVIM model (Table 1).

Repeatability of the diffusion parameters calculated from two pre-treatment measurements indicated coefficients of variation (CVs) smaller than or equal to 10% for all parameters of the mono-exponential, the stretched exponential, and the statistical model. From the IVIM model, only D resulted in a CV below 10% with f, fD*, and in particular D* featuring larger CVs (Table 1). These results are in agreement with previous studies in other tumours25, 30. Repeatability of the diffusion parameters is illustrated in Fig. 1 using Bland-Altman plots.

Parameter changes under therapy. By way of example, Fig. 2 depicts diffusion parameter maps and curve fits from one voxel within a pelvic lesion before androgen deprivation and one, two, and three months.
after treatment onset. The manually defined region of interest (ROI) is also depicted. There is a high similarity between the maps of the various diffusion coefficients ADC, ADC*, D, DDC, and Ds*. Visual inspection reveals an increase in the diffusion coefficients, in $\alpha$ from the stretched exponential, and in $\sigma^*$ from the statistical model under therapy. Example curve fits from one voxel within the lesion feature clear non-mono-exponential signal attenuation.

Repeated measures analysis of variance (ANOVA) revealed a significant increase in the median ADC ($p < 1 \times 10^{-6}$) and ADC* values ($p < 1 \times 10^{-6}$) under therapy. Furthermore, a significant increase in D ($p < 1 \times 10^{-6}$), f ($p = 2 \times 10^{-6}$), and f-D* ($p = 2.7 \times 10^{-3}$) from the IVIM model, in DDC ($p < 1 \times 10^{-6}$) and $\alpha$ ($p = 0.028$) from the stretched exponential, and in $D_s^*$ ($p < 1 \times 10^{-6}$) from the statistical model was observed. The only parameter that showed no significant change under therapy was D* ($p = 0.463$) from the IVIM model.

Changes in PSA levels and correlations with diffusion parameters. Changes in serum PSA levels under therapy confirmed treatment response in all patients. Repeated measures ANOVA revealed a significant decrease of the PSA values under therapy ($p < 0.001$). Compared with pre-treatment values, the PSA decreased significantly after one ($p < 0.001$), two ($p < 0.001$), and three ($p < 0.001$) months. After commencement of androgen deprivation treatment, average PSA values decreased by 85.7% (range = 51.4%–98.8%) at one month, 95.2% (range = 72.4%–99.9%) at two months, and 96.6% (range = 83.5%–99.9%) at three months.

Bivariate correlation showed a moderate negative relationship between the serum PSA values and ADC, ADC* from the mono-exponential, f from the IVIM, and DDC from the stretched exponential model (Table 3). The strongest correlation thereby occurred between the PSA values and f from the IVIM model. A weak negative correlation was observed between the PSA levels and D from the IVIM and Ds* from the statistical model (Table 3).

Parameter correlations between the various diffusion parameters. Table 4 summarizes the Pearson’s correlation coefficients and corresponding $p$-values between the various diffusion parameters. A very strong positive correlation was found between the ADC and the diffusion parameters from the non-mono-exponential models (Table 4) which agrees well with the similarities of the parameter maps observed in Fig. 2. Within the diffusion models a very strong positive correlation was observed between Ds* and $\sigma^*$ from the statistical model. A moderate positive correlation was found between DDC and $\alpha$ from the stretched exponential model and a moderate positive correlation was observed between f and D* from the IVIM model. Finally, bivariate correlation showed a weak relationship between D and f-D* from the IVIM model.

Model preferences. Model preference at each time point is summarized in Table 5. Non-mono-exponential models were preferred by 85% of the tumours before treatment begin and by 97%, 97%, and 91% after commencement of antiandrogen treatment. BIC analysis revealed that the stretched exponential model was overall favoured
**Figure 2.** Parameter maps for an exemplary patient with a pelvic bone metastasis from prostate cancer: (a) ROI circumscribing the lesion superimposed onto the pre-treatment ADC map, (b) enlarged sections showing the parameter maps of the various diffusion parameters before therapy and one, two, and three months after treatment onset, and (c) measured signals and fitted curves of one voxel within the lesion at each time point.

| Model                | Parameter | One month after treatment begin | Two months after treatment begin | Three months after treatment begin |
|----------------------|-----------|----------------------------------|----------------------------------|-----------------------------------|
| Mono-exponential     | ADC       | 35.2 (<1·10^{-6})               | 36.3 (<1·10^{-6})               | 34.6 (<1·10^{-6})                |
|                      | ADC*      | 35.6 (<1·10^{-6})               | 35.0 (<1·10^{-6})               | 28.7 (<1·10^{-6})                |
| IVIM                 | D         | 31.0 (<1·10^{-6})               | 30.5 (<1·10^{-6})               | 22.3 (8.1·10^{-6})               |
|                      | D*        | 12.0 (0.293)                    | 18.2 (0.168)                    | 14.0 (0.368)                     |
|                      | f         | 29.5 (5·10^{-6})                | 29.5 (1.5·10^{-6})              | 37.9 (4.7·10^{-6})               |
|                      | Df*       | 14.0 (0.073)                    | 13.0 (0.119)                    | 29.1 (2.6·10^{-4})               |
| Stretched exponential| DDC       | 41.5 (<1·10^{-6})               | 42.9 (<1·10^{-6})               | 39.6 (<1·10^{-6})                |
|                      | α         | 5.0 (0.005*)                    | 4.6 (0.015*)                    | 1.6 (0.523)                      |
| Statistical          | Df*       | 38.2 (<1·10^{-6})               | 39.0 (<1·10^{-6})               | 37.2 (<1·10^{-6})                |
|                      | σ*        | 29.7 (<1·10^{-6})               | 29.0 (<1·10^{-6})               | 29.1 (<1·10^{-6})                |

**Table 2.** Percentage changes of the various diffusion parameters observed one, two, and three months after treatment begin. The values in parentheses are the p-values of the post-hoc comparisons of the baseline values with those one, two, and three months after anticancer treatment. Significant statistics have p < 0.017, incorporating a correction for multiple comparisons. Statistically significant values are designated by an asterisk.
by the majority of approximately 60% of tumours at all time points. When considering the tumours where each model was preferred, the percentage of voxels within the lesion that favoured that model ranged between 28% and 85% (Table 5). This shows that across the lesions a moderate to large percentage of voxels favoured models other than the dominant model.

### Discussion

Osteoblastic bone metastases without associated soft tissue, such as those commonly encountered in prostate cancer, are classified as non-measurable according to RECIST7. Thus, the establishment of a non-invasive, validated response assessment is warranted since 90% of patients with advanced prostate cancer have bone metastases as the only site of disease involvement1. ADC quantified using the mono-exponential model has shown promise as a potential biomarker for assessing treatment response in this patient group11–15. However, our current study shows that there is clear statistical preference for using non-mono-exponential diffusion models compared with the mono-exponential model in bone metastases, suggesting that these models may provide better descriptions of the biological changes with treatment. The additional parameters derived using non-mono-exponential models are sensitive to treatment changes and may provide complementary information to improve the treatment response evaluation. With the exception of D* from the IVIM model,

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**Table 3.** Pearson’s correlation coefficients and corresponding p-values between the PSA and each of the various diffusion parameters. Statistically significant values are designated by an asterisk.

| Model          | Parameters   | Pearson’s correlation coefficient | p-value |
|----------------|--------------|----------------------------------|---------|
| Mono-exponential | ADC, PSA    | −0.45                            | 0.001*  |
|                | ADC*, PSA   | −0.39                            | 0.006*  |
| IVIM           | D, PSA       | −0.29                            | 0.044*  |
|                | D*, PSA      | 0.13                             | 0.40    |
|                | f, PSA       | −0.48                            | 0.002*  |
|                | fD*, PSA     | −0.25                            | 0.12    |
| Stretched exponential | DDC, PSA   | −0.46                            | 0.001*  |
|                | α, PSA       | −0.13                            | 0.426   |
| Statistical    | D*, PSA      | −0.39                            | 0.006*  |
|                | σ*, PSA      | −0.21                            | 0.21    |

**Table 4.** Pearson’s correlation coefficients and corresponding p-values between the various diffusion parameters. Statistically significant values are designated by an asterisk.

| Model          | Parameters   | Pearson’s correlation coefficient | p-value |
|----------------|--------------|----------------------------------|---------|
| Mono-exponential | ADC*, ADC   | 0.96                             | <1·10⁻⁶* |
|                | D, ADC       | 0.86                             | <1·10⁻⁶* |
|                | D, f         | 0.10                             | 0.25    |
|                | D, D*        | 0.09                             | 0.29    |
|                | D*, f        | −0.40                            | 3·10⁻⁶* |
|                | D, D*        | 0.17                             | 0.049*  |
| IVIM           | DDC, ADC     | 0.99                             | <1·10⁻⁶* |
|                | DDC, α       | 0.50                             | <1·10⁻⁶* |
| Stretched exponential | DDC, ADC   | 0.98                             | <1·10⁻⁶* |
|                | DDC, α       | 0.50                             | <1·10⁻⁶* |
| Statistical    | D*, ADC      | 0.98                             | <1·10⁻⁶* |
|                | D*, σ*       | 0.82                             | <1·10⁻⁶* |

**Table 5.** Number of lesions at every time point where the respective diffusion models was preferred based on the majority of voxels. The values in parentheses are the median and range percentage of voxels where the respective model was preferred, considering only lesions where the tumour model was overall favoured.

| Model          | Percentage of lesions where model was preferred |
|----------------|-----------------------------------------------|
|                | Before treatment | One month after treatment begin | Two months after treatment begin | Three months after treatment begin |
| Mono-exponential | 5 (43; 32–50)   | 1 (53; 53–53) | 1 (83; 83–83) | 3 (44; 33–59) |
| IVIM           | 4 (33; 54–64)   | 5 (44; 37–79) | 4 (42; 34–47) | 5 (45; 33–53) |
| Stretched exponential | 21 (42; 28–63) | 21 (42; 35–68) | 20 (46; 36–73) | 21 (43; 29–74) |
| Statistical    | 0 (N/A)        | 7 (45; 40–62) | 9 (36; 33–55) | 5 (47; 35–85) |
all diffusion parameters increased significantly after initiation of anti-androgen therapy compared with pre-treatment values. The fact that the stretched exponential model provided better data fitting within the majority of tumours and that the derived parameters DDC and \( \alpha \) were highly repeatable and sensitive to treatment effects supports its wider use in this patient cohort.

Significant increases in diffusion coefficient values were accompanied by large contemporaneous decreases in the serum PSA levels, in keeping with disease response. In this regard, the highest percentage change after initiation of androgen deprivation treatment was observed for DDC from the stretched exponential model. Bivariate correlation showed a very strong positive relationship between ADC from the mono-exponential model and the diffusion coefficients from the non-mono-exponential models, indicating that they are sensitive to the same tissue characteristics and provide similar information. As pointed out in previous studies, the mono-exponential model can be considered a conditional case of other diffusion models (i.e. when \( \alpha = 1 \) and \( f = 0 \))\(^{29, 30} \). As a result, lesions that favour the mono-exponential model are not disadvantaged by the utilization of other models but the additional parameters of these models do not add value in such tumours. Interestingly, our study also showed an increase in f values from IVIM analysis following treatment. The mechanism for an increase in f is uncertain, but may reflect vascular normalization within tumours. However, this would require further investigations.

Previous studies in other tumour types observed no significant correlation between \( \alpha \) and DDC from the stretched exponential model\(^{25, 28} \). By contrast, a moderate positive correlation between these parameters was observed in our study. Although the biological underpinning of \( \alpha \) is unclear, it is likely to reflect tissue heterogeneity, which may also influence the DCC measurement. Our results suggest that these parameters may contain complementary information, which could be exploited to improve treatment monitoring in prostate cancer bone metastases. Similarly, Pearson correlation showed no significant relationship between \( D \) and either \( f \) or \( D^* \) from the IVIM model, indicating that these parameter report on different tissue properties. The very strong correlation of \( D^* \) and \( \sigma^* \) from the statistical model, however, suggests that these parameters largely provide the same information.

Only moderate correlations between the serum PSA values and ADC, \( \text{ADC}^* \) from the mono-exponential, f from the IVIM, and DDC from the stretched exponential model were observed. The known prostate cancer heterogeneity may at least partially account for the fact that only moderate relationships could be delineated. However, with respect to the ADC, the present results are in good agreement with previous work\(^{17} \).

The repeatability of the diffusion parameters is an important consideration when evaluating different diffusion models for assessing response to therapy. In agreement with previous studies in other tumours\(^{25, 30} \), our results show that the parameters from the stretched exponential model show high precision which is comparable to the repeatability of ADC and perfusion-insensitive \( \text{ADC}^* \) from the mono-exponential model. High repeatability was also observed for parameters derived from the statistical model. In agreement with previous studies in other tumours\(^{25, 30} \), the precision of the perfusion-related parameters from the IVIM model was significantly worse. This may be due to the larger number of parameters that are fitted by the model, rendering it more sensitive to measurement noise. Due to time constraints in the clinical setting, only eight different b-values were acquired, which may not provide sufficient data support to reliably estimate the perfusion-sensitive components of the IVIM model. However, the generally limited measurement repeatability of the perfusion-related estimates is a well-documented drawback of IVIM modelling\(^{33, 34} \). Our study showed that the composite parameter f-D\(^* \) showed a similar repeatability as f. Inversely correlated errors of f and D\(^* \) as described elsewhere\(^{30} \) were not observed in our study.

The kurtosis model\(^{37} \), which has gained popularity in recent years, was not investigated in our current work since the non-Gaussian diffusion behaviour that underlies the kurtosis model has negligible effect in our acquired b-value range\(^{37, 42} \). Kurtosis imaging relies on the acquisition of a maximum b-value of at least 1500 s/mm\(^2 \), which is usually not incorporated in clinical body imaging\(^{29} \). The inclusion of higher maximum b-values would prolong the minimum achievable echo time, thus lowering the SNR and in turn decreasing accuracy and precision of the parameter estimates. Beyond that, higher maximum b-values increase eddy current-induced image distortions in the data.

There are limitations to our study. First, we included a relatively small patient cohort and larger patient studies would be useful to validate our findings. However, as most patients in this disease setting would have received androgen deprivation, chemotherapy or radiation therapy prior to any imaging, the relatively low patient number from a single institution is not unexpected. Second, while our study findings are in agreement with other studies evaluating non-mono-exponential diffusion models in cancer\(^{18, 21, 25, 30} \), more b-values in particular at the lower end of the range may be needed to realize the full potential of IVIM modelling for treatment monitoring of bone metastases.

In conclusion, our study shows that there is a statistical model fit preference for using non-mono-exponential diffusion models in bone metastases from prostate cancer before and after initiation of antiandrogen treatment. The additional parameters derived using these models may provide additional information, which may improve the evaluation of treatment response and may permit detecting the onset of antiandrogen resistance at an earlier time point. Our results show that the stretched exponential model was favoured within the majority of tumours. Its derived parameters DDC and \( \alpha \) demonstrated very good measurement repeatability and were also sensitive to treatment effects. We observed a moderate correlation of the two parameters, which indicates that they contain a certain amount of complementary information, which may be exploited to improve treatment response assessment. Thus, the stretched exponential may be utilized as a potential optimal model for monitoring treatment response in prostate cancer bone metastases.

Methods

Patient population. This prospective, single-centre clinical study was approved by the Cantonal Research Ethics Committee (Zurich, Switzerland) and written informed consent was obtained from all patients. This
study was conducted in accordance with the Declaration of Helsinki. Seventeen treatment-naive patients with prostate cancer bone metastases who fulfilled all inclusion and exclusion criteria were included. The inclusion criteria were as follows: adults with histologically proven prostate cancer, evidence of bone metastases in the pelvis confirmed on skeletal scintigrams and no prior history of antiandrogen treatment, chemotherapy or radiation therapy. The exclusion criteria were as follows: history of another malignancy, contraindication to MRI or unwillingness to participate in the clinical study. Five patients had to be secondarily excluded: three patients deceased prior to termination of the study and two patients withdrew willingness to participate in the study after an acute deterioration in their health condition. Thus, twelve men (mean age = 76, range = 67–85) with a total of 34 pelvic bone metastases from prostate cancer (mean size = 7.64 cm³, range = 2.64–39.65 cm³) were included in the analysis.

**Diagnosis of bone metastases at skeletal scintigraphy.** Diagnosis of metastatic disease in the pelvis was made within seven days of the baseline MRI examination using whole-body skeletal radionuclide scintigraphy, which was performed three hours after injection of 550 M bq ⁹⁹mTc-labeled dicarboxypropane diphosphonate (Teceos, Cibio, Gif-sur-Yvette, France). Images were acquired using a dual-head whole-body scanner with a low energy high-spatial-resolution collimator (E-CAM, Siemens, Munich, Germany).

**Androgen deprivation or androgen ablation treatment.** All patients underwent androgen deprivation by means of either medical treatment (three patients received goserelin (Zoladex®), two patients leuprorelin acetate (Lucrin®), two patients goserelin (Zoladex®) + denosumab (Xgeva®), two patients leuprorelin acetate (Lucrin®) + denosumab (Xgeva®), and one patient goserelin (Zoladex®) + bicalutamide (Casodex®) + denosumab (Xgeva®)) or surgical orchiectomy (two patients). Antiandrogen treatment was commenced within two days of the baseline MRI examination.

**MRI acquisition.** Image acquisition was performed on a 1.5 T whole-body scanner (Achieva, release 3.2.3.4, Philips Healthcare, Best, the Netherlands) within two days before and repeated one, two, and three months after the commencement of antiandrogen treatment. Pelvic images were acquired with the patient in the supine position by using a four-element receive-only body coil array (Philips, Healthcare, Best, the Netherlands). The imaging protocol consisted of transversal T₁-weighted, T₂-weighted, and proton density-weighted sequences followed by axial DWI including eight b-factors (b = 0, 50, 100, 150, 200, 400, 600, 800 s/mm²). Details of the MRI sequences are summarized in Table 6. The total measurement time of the protocol amounted to approximately 20 minutes. At baseline, the DWI scan was acquired twice to permit assessment of measurement repeatability.

**Serum PSA measurements for treatment response assessment.** To assess treatment response, the serum PSA levels were determined in tandem with each MRI examination. Venous blood was therefore drawn from all patients via the right or left cubital vein before and at one, two, and three months after starting androgen withdrawal treatment. A decline in PSA levels by more than 50% confirmed by a second measurement four weeks later was accepted as a response to treatment.

**Diffusion data analysis.** Data analysis was performed using in-house software written in MATLAB (The MathWorks, release 2016a, Natick, MA, USA). Prior to image analysis, eddy-induced image warping and motion were corrected in the in-vivo data sets using a correlation-based affine registration algorithm. Parameter maps were calculated by least-squares fitting using the mono-exponential and three non-mono-exponential diffusion models.

Estimates of the ADC were computed by mono-exponential fitting of the DWI data using the following equation:

\[ S(b) = S_0 \exp(-bADC), \]  

where \( S(b) \) corresponds to the signal with and \( S_0 \) without diffusion weighting and \( b \) denotes the b-value. To diminish the influence of perfusion effects, a mono-exponential fit of all DWI data with \( b > 200 \text{ s/mm}^2 \) was also performed (ADC*).

Estimates of D, D*, and f were obtained by fitting the IVIM model to the DWI data:

\[ \frac{S(b)}{S(0)} = D\exp(-bD) + f \times D_0\exp(-bD_0), \]
\[ S(b) = S_0 (\exp(-bD^*) + (1-f)\exp(-bD)), \]

with \(0 \leq f \leq 1\). In addition, the composite parameter \(fD^*\) was also computed which was shown to be linked to the relative perfusion or blood flow\(^{39}\). Starting values of \(D, D^*\), and \(f\) were computed by a least-squares fit of a mono-exponential curve to the DWI data with \(b > 200 \text{s/mm}^2\) followed by another mono-exponential curve fitted to the remaining signal at the lower \(b\)-values\(^{38}\).

Estimates of DDC and \(\alpha\) were determined by fitting the stretched exponential model to the DWI data\(^{36}\):

\[ S(b) = S_0 \exp(-(b\text{DDC})^\alpha) \]

with \(0 \leq \alpha \leq 1\).

Estimates of \(D_s\) and \(\sigma\) were computed by fitting the statistical model to the DWI data\(^{36}\):

\[ S(b) = S_0 \left( \frac{1 + \phi \left( \frac{D_s}{\sigma \sqrt{2\pi}} + \frac{b\text{DDC}}{\sigma \sqrt{2\pi}} \right)}{1 + \phi \left( \frac{D_s}{\sigma \sqrt{2\pi}} \right)} \right) \exp \left( -bD_s + \frac{1}{2}b^2 \sigma^2 \right). \]

Based on previously published results\(^{37}\), the parameters were reformulated for statistical analysis to obtain the mean \((\bar{D}_s)\) and the standard deviation \((\sigma^*)\) of the distribution of diffusion coefficients within a voxel\(^{38}\).

The parameter maps acquired one, two, and three months after the commencement of androgen withdrawal therapy were co-registered to the corresponding pre-treatment parameter map for each patient. For this purpose, each post-treatment ADC map was co-registered to the corresponding pre-treatment ADC map using a robust multiresolution alignment algorithm\(^{48}\) that was previously extended to allow for affine transformations\(^{12}\). The resulting transformation matrix was subsequently used to co-register the corresponding post- and pre-treatment parameter maps of the other diffusion parameters.

In line with previous work\(^{12,16}\), ROIs were manually defined on the baseline ADC maps, taking into account the diagnostic information of the skeletal scintigrams and the corresponding conventional anatomical images to simplify lesion localization. For each metastasis, ROIs were circumscribed on the transversal plane of each tumour-bearing slice and subsequently summed up, resulting in a three-dimensional ROI enclosing the entire lesion. Thereafter, the ROIs were applied to subsequent scans and to the corresponding parameter maps obtained using the non-mono-exponential diffusion models. Median values of each diffusion parameter were calculated in order to reduce sensitivity to outlier values and since the parameters were not normally distributed, resulting in ten measures at each time point. In patients with multiple lesions, the weighted median value of each diffusion parameter was also calculated on a per-patient basis in order to permit assessment of correlations with the PSA values.

**Statistical analysis.** Statistical analysis of the data was performed with SPSS (Statistics for Windows, version 23.0, IBM, Armonk, NY, USA). All statistics was calculated on the log-transformed data, back transformations were performed where appropriate\(^{49}\).

Using the two pre-treatment measurements of each diffusion parameter, measurement repeatability was assessed with the method of Bland and Altman\(^{45}\). Repeatability of the parameters was quantified using the CV defined as \(CV = 100\% \cdot \sqrt{\exp(\sigma^2/2)} - 1\), where \(\sigma^2\) corresponds to the variance of the difference between the baseline parameters. Differences in the pre-treatment values of the parameters were assessed using two-sided paired t-tests.

Cohort baseline values were computed as the mean of the logarithm of the two baseline measurements. Inter-patient variability was assessed by computing the variance of the mean of the logarithm of the two baseline values across all patients. The data were back transformed to percentages using \(100\% \cdot \sqrt{\exp(\sigma^2)} - 1\). Changes in the PSA values and the diffusion parameters of the four diffusion models under therapy were analysed using repeated measures ANOVA followed by pairwise comparisons of the parameter values before and after beginning of treatment. Using the Bonferroni correction, the significance level was set at \(p < 0.017\) to adjust for multiple comparisons. In all other statistical analyses, a p-value of \(p < 0.05\) was considered statistically significant. In these cases, no corrections for multiple comparisons were performed and raw p-values are reported since this study is primarily descriptive and many of the parameters are expected to exhibit a high degree of correlation.

Pearson’s correlation coefficient across times was used to assess correlations between the PSA and the parameters of each diffusion model. Moreover, correlations between the ADC and the diffusion coefficients of the non-mono-exponential models and between the parameters within each non-mono-exponential model were evaluated.

Statistical preference for the four models in the lesions was quantified at each time point using the BIC which penalizes for model complexity\(^{51}\):

\[ BIC = -2\log(L(\theta)) + k\log(n), \]

where \(L(\theta)\) denotes the value of the maximized likelihood objective function for a model with \(k\) parameters and \(n\) corresponds to the number of data points. In each voxel, the preferred model was given by the model with the lowest value of the BIC. The majority vote across all voxels of each lesion was used as preferred model within each lesion.
References

1. Carlin, B. I. & Andriole, G. L. The natural history, skeletal complications, and management of bone metastases in patients with prostate carcinoma. Cancer 88, 2989–2994 (2000).

2. Schellhammer, P. F. et al. A controlled trial of bicalutamide versus flutamide, each in combination with luteinizing hormone-Releasing hormone analogue therapy, in patients with advanced prostate carcinoma: analysis of time to progression. Cancer 78, 2164–2169 (1996).

3. Pienta, K. J. & Bradley, D. Mechanisms underlying the development of androgen-independent prostate cancer. Clin Cancer Res 12, 1665–1671 (2006).

4. Harris, W. P., Mostaghel, E. A., Nelson, P. S. & Montgomery, B. Androgen deprivation therapy: progress in understanding mechanisms of resistance and optimizing androgen depletion. Nat Clin Pract Urol 6, 76–85 (2009).

5. Karantanos, T., Corn, P. G. & Thompson, T. C. Prostate cancer progression after androgen deprivation therapy: mechanisms of castrate resistance and novel therapeutic approaches. Oncogene 32, 5501–5511 (2013).

6. Jambor, J. et al. Prospective evaluation of planar bone scintigraphy, SPECT, SPECT/CT, 18F-NaF PET/CT and whole body 1.5 T MRI, including DWI, for the detection of bone metastases in high risk breast and prostate cancer patients: SKELETA clinical trial. Acta Oncol 55, 59–67 (2016).

7. Eisenhauer, E. A. et al. New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). Eur J Cancer 45, 228–247 (2009).

8. Scher, H. I. et al. Design and end points of clinical trials for patients with progressive prostate cancer and castrate levels of testosterone: recommendations of the Prostate Cancer Clinical Trials Working Group. J Clin Oncol 26, 1148–1159 (2008).

9. Berthold, D. R. et al. Treatment of hormone-refractory prostate cancer with docetaxel or mitoxantrone: relationships between prostate-specific antigen, pain, and quality of life response and survival in the TAX-327 study. Clin Cancer Res 14, 2763–2767 (2008).

10. Halabi, S. et al. Prostate-specific antigen changes as surrogate for overall survival in men with metastatic castration-resistant prostate cancer treated with second-line chemotherapy. J Clin Oncol 31, 3944–3950 (2013).

11. Lee, K. C. et al. A feasibility study evaluating the functional diffusion map as a predictive imaging biomarker for detection of treatment response in a patient with metastatic prostate cancer to the bone. Neoplasia 9, 1003–1011 (2007).

12. Reischauer, C. et al. Bone metastases from prostate cancer: assessing treatment response by using diffusion-weighted imaging and functional diffusion maps—initial observations. Radiology 257, 523–531 (2010).

13. Messiou, C. et al. Assessing response in bone metastases in prostate cancer with diffusion weighted MRI. Eur Radiol 21, 2169–2177 (2011).

14. Blackledge, M. D. et al. Assessment of treatment response by total tumour volume and global apparent diffusion coefficient using diffusion-weighted MRI in patients with metastatic bone disease: a feasibility study. PLoS One 9, e19779 (2014).

15. Perez-Lopez, R. et al. Diffusion-weighted Imaging as a Treatment Response Biomarker for Evaluating Bone Metastases in Prostate Cancer: A Pilot Study. Radiology 283, 168–177 (2017).

16. Reischauer, C. et al. Pilot study on the detection of antiandrogen resistance using serial diffusion-weighted imaging of bone metastases in prostate cancer. J Magn Reson Imaging 43, 1407–1416 (2016).

17. Riches, S. F., Hawtin, K., Charles-Edwards, E. M. & Souza, N. M. Diffusion-weighted imaging of the prostate and rectal wall: comparison of biexponential and monoexponential modelled diffusion and associated perfusion coefficients. NMR Biomed 22, 318–325 (2009).

18. Jansen, J. F., Stambuk, H. E., Koutcher, J. A. & Shukla-Dave, A. Non-gaussian analysis of diffusion-weighted MR imaging in head and neck squamous cell carcinoma: A feasibility study. AJNR Am J Neuroradiol 31, 741–748 (2010).

19. Rosenkrantz, A. B. et al. Prostate cancer: feasibility and preliminary experience of a diffusional kurtosis model for detection and assessment of aggressiveness of peripheral zone cancer. Radiology 264, 126–135 (2012).

20. Mazzuferi, Y. et al. Diffusion-weighted magnetic resonance imaging of the prostate: improved robustness with stretched exponential modeling. J Comput Assist Tomogr 36, 695–703 (2012).

21. Yuan, J. et al. Non-Gaussian analysis of diffusion weighted imaging in head and neck at 3T: a pilot study in patients with nasopharyngeal carcinoma. PLoS One 9, e87024 (2014).

22. Bourne, R. M. et al. Information theoretic ranking of four models of diffusion attenuation in fresh and fixed prostate tissue ex vivo. Magn Reson Med 72, 1418–1426 (2014).

23. Yan, R. et al. Non-Gaussian diffusion MRI imaging of glioma: comparisons of multiple diffusion parameters and correlation with histologic grade and MIB-1 (Ki-67 labeling) index. Neuroradiology (2015).

24. Paner, R. et al. Evaluation of diffusion models in breast cancer. Medical Physics 42, 4833–4839 (2015).

25. Winfield, J. M. et al. Modelling DW-MRI data from primary and metastatic ovarian tumours. European Radiology 25, 2033–2040 (2015).

26. Merisari, H. et al. Fitting methods for intravoxel incoherent motion imaging of prostate cancer on region of interest level: Repeatability and gleason score prediction. Magn Reson Med 77, 1249–1264 (2017).

27. Zhang, G. et al. Comparison of non-Gaussian and Gaussian diffusion models of diffusion weighted imaging of rectal cancer at 3.0 T MRI. Sci Rep 6, 38782 (2016).

28. Winfield, J. M. et al. Separation of type and grade in cervical tumours using non-mono-exponential models of diffusion-weighted MRI. Eur Radiol 27, 627–636 (2017).

29. Hauser, T. et al. Characterization and therapy monitoring of head and neck carcinomas using diffusion-imaging-based intravoxel incoherent motion parameters—preliminary results. Neuroradiology 55, 527–536 (2013).

30. Orton, M. R. et al. Diffusion-weighted MR imaging of metastatic abdominal and pelvic tumours is sensitive to early changes induced by a VEGF inhibitor using alternative diffusion attenuation models. Eur Radiol 26, 1412–1419 (2016).

31. Le Bihan, D. et al. MR imaging of intravoxel incoherent motions: application to diffusion and perfusion in neurologic disorders. Radiology 161, 401–407 (1986).

32. Le Bihan, D. et al. Separation of diffusion and perfusion in intravoxel incoherent motion MR imaging. Radiology 168, 497–505 (1988).

33. Le Bihan, D., Turner, R., Moonen, C. T. & Pekar, J. Imaging of diffusion and microcirculation with gradient sensitization: design, strategy, and significance. J Magn Reson Imaging 1, 7–28 (1991).

34. Pekar, J., Moonen, C. T. & van Zijl, P. C. On the precision of diffusion/perfusion imaging by gradient sensitization. Magn Reson Med 23, 122–129 (1992).

35. Koh, D.-M., Collins, D. J. & Orton, M. R. Intravoxel incoherent motion in body diffusion-weighted MRI: reality and challenges. AJR Am J Roentgenol 196, 1351–1361 (2011).

36. Bennett, K. M. et al. Characterization of continuously distributed cortical water diffusion rates with a stretched-exponential model. Magn Reson Med 50, 727–734 (2003).

37. Jensen, J. H., Helpert, J. A., Ramani, A., Lu, H. & Kaczynski, K. Diffusional kurtosis imaging: The quantification of non-gaussian water diffusion by means of magnetic resonance imaging. Magn Reson Med 53, 1432–1440 (2005).

38. Hall, M. G. & Barrick, T. R. From diffusion-weighted MRI to anomalous diffusion imaging. Magn Reson Med 59, 447–455 (2008).

39. Ozarslan, E. et al. Observation of anomalous diffusion in excised tissue by characterizing the diffusion-time dependence of the MR signal. J Magn Reson 183, 315–323 (2006).
40. Zhou, X. J., Gao, Q., Abdullah, O. & Magin, R. L. Studies of anomalous diffusion in the human brain using fractional order calculus. *Magn Reson Med* 63, 562–569 (2010).
41. Messiou, C., Collins, D. J., Morgan, V. A. & Desouza, N. M. Optimising diffusion weighted MRI for imaging metastatic and myeloma bone disease and assessing reproducibility. *Eur Radiol* 21, 1713–1718 (2011).
42. Rosenkrantz, A. B. *et al.* Body diffusion kurtosis imaging: Basic principles, applications, and considerations for clinical practice. *J Magn Reson Imaging* 42, 1190–1202 (2015).
43. Bubley, G. J. *et al.* Eligibility and response guidelines for phase II clinical trials in androgen-independent prostate cancer: recommendations from the Prostate-Specific Antigen Working Group. *J Clin Oncol* 17, 3461–3467 (1999).
44. Netsch, T. & Van Muiswinkel, A. Quantitative evaluation of image-based distortion correction in diffusion tensor imaging. *IEEE Trans Med Imaging* 23, 789–798 (2004).
45. Le Bihan, D. & Turner, R. The capillary network: a link between IVIM and classical perfusion. *Magn Reson Med* 27, 171–178 (1992).
46. Yablonskiy, D. A., Brethorst, G. L. & Ackerman, J. J. H. Statistical model for diffusion attenuated MR signal. *Magn Reson Med* 50, 664–669 (2003).
47. Barr, D. R. & Sherrill, E. T. Mean and Variance of Truncated Normal Distributions. *Am Stat* 53, 357–361 (1999).
48. Nestares, O. & Heeger, D. J. Robust multiresolution alignment of MRI brain volumes. *Magn Reson Med* 43, 705–715 (2000).
49. Keene, O. N. The log transformation is special. *Stat Med* 14, 811–819 (1995).
50. Bland, J. M. & Altman, D. G. Statistical methods for assessing agreement between two methods of clinical measurement. *Lancet* 1, 307–310 (1986).
51. Schwarz, G. Estimating the dimension of a model. *Ann Stat* 6, 461–464 (1978).

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**Author Contributions**

C.R., D.-M.K., J.M.F., A.G., participated in the study design. C.R. and A.G. designed the magnetic resonance scanning sequence; R.P., A.G. recruited patients; R.P., A.G. collected the data; C.R. analysed the data; all authors participated in the interpretation of the data, C.R., D.-M.K., A.G. prepared the first draft of the manuscript; all authors revised and approved the manuscript.

**Additional Information**

**Competing Interests:** The authors declare that they have no competing interests.

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