Effect of extremely low-concentration gaseous chlorine dioxide against surface 
Escherichia coli, Pseudomonas aeruginosa and Acinetobacter baumannii in wet conditions on glass dishes

Hirofumi Morino1,2*, Masafumi Futatsukame1,2, Takanori Miura1,2 and Takashi Shibata1,2

Abstract
Objective: Healthcare-associated infections due to Gram-negative bacteria (GNB) are a major cause of mortality and morbidity throughout the world. The purpose of the research described here was to evaluate the possibility of using an extremely low-concentration gaseous chlorine dioxide (ClO2, 0.01 ppmv, 0.028 mg/m³) as a technique to reduce the risk of environmental infection by GNB. In this study we set up an exposure chamber (1 m³) and used three types of GNB, namely Escherichia coli, Pseudomonas aeruginosa and Acinetobacter baumannii.

Results: The extremely low-concentration gaseous ClO2 inactivated E. coli (> 2 log₁₀ reductions, within 2 h), P. aeruginosa (> 4 log₁₀ reductions, within 2 h) and A. baumannii (> 2 log₁₀ reductions, within 3 h) in wet conditions on glass dishes. Treatment of moist environments with extremely low-concentration gaseous ClO2 may help to reduce the risk of environmental infection by GNB without harmful effects.

Keywords: Chlorine dioxide, Gas, Bacteria, Escherichia coli, Pseudomonas aeruginosa, Acinetobacter baumannii

Introduction
Healthcare-associated infections (HAIs) due to Gram-negative bacteria (GNB) are a major cause of mortality and morbidity throughout the world [1]. Many GNB cause respiratory tract infections; these include Pseudomonas aeruginosa, Escherichia coli, Acinetobacter baumannii and Klebsiella pneumonia [2]. In a previous study, we showed that gaseous chlorine dioxide (ClO₂, CAS No. 10049-04-4) in a low concentration (mean 0.05 ppmv, 0.14 mg/m³) inactivates E. coli in wet conditions on a glass surface [3]. Recently, Ogata et al. reported that extremely low-concentration gaseous ClO₂ (0.01 ppmv, 0.028 mg/m³) inactivated airborne bacteria and viruses [4]. This concentration is only 1/10 of the threshold limit value (0.1 ppmv, 0.28 mg/m³) for gaseous ClO₂ defined by the American Occupational Safety and Health Administration (OSHA) as an 8 h time-weighted average [5]. Based on that report, we speculated that extremely low-concentration gaseous ClO₂ can also inactivate bacteria on surfaces. To investigate the efficacy of extremely low-concentration gaseous ClO₂ against surface bacteria, we set up an exposure chamber (1 m³), which can maintain a constant, extremely low-concentration of the gas. In this study, we used three types of bacterium, namely E. coli, P. aeruginosa and A. baumannii. The purpose of this study is to evaluate the usefulness of this treatment with extremely low-concentration gaseous ClO₂ as a
technique to reduce the risk of infection by environmental GNB without harmful effects.

**Main text**

**Materials and methods**

**Test bacteria and preparation**

Preparation of the test bacteria followed our previous report with some modifications [3]. *E. coli* NBRC 3972, *P. aeruginosa* NBRC 13275 and *A. baumannii* NBRC 110494 were obtained from the Biological Resource Center (NITE, Japan). Each bead from the stock vials (PL.170/M, Microbank™, Pro-Lab Diagnostics Inc., U.S.A.) of *E. coli* and *P. aeruginosa* stored at −80 °C was transferred into 5 ml of Soybean Casein Digest (SCD) broth (393-00185, Nihon Seiyaku, Japan). Incubation was carried out in a shaking incubator at 200 rpm for 18 h at 37 °C. One bead from a stock vial of *A. baumannii* was transferred onto a SCD agar plate (51048, Nissui Pharmaceutical Co., LTD., Japan) and incubated at 37 °C for 18 h. Cells of *E. coli* and *P. aeruginosa* were collected by centrifugation at 1400×g for 15 min at room temperature. Cells of *A. baumannii* on the agar plate were collected with an inoculating loop. These bacteria were washed three times with Dulbecco’s phosphate buffer saline (D-PBS). The bacteria were resuspended in D-PBS and adjusted to an OD660 of 0.1 (1×10⁸ cells/ml). These bacteria were used for inoculation.

A 100 µl aliquot of the bacterial suspension (1×10⁸ cells/ml in D-PBS) was placed on 5 cm diameter glass dishes. In this study, bacteria on the glass dishes did not undergo a drying process. The dishes were placed in the 1 m³ exposure chamber described below. The preparations were exposed to 0.01, 0.03 ppmv ClO₂ gas or air (control) for 0, 1, 2 and 3 h. Subsequently, 120 µl of SCD medium with 1 mM Na₂S₂O₃ as neutralizer was added to the glass dishes after the exposure to air or ClO₂ gas. The bacteria on the glass dishes were collected using a cell scraper (179693, Thermo Fisher Scientific, U.S.A.).

**Determination of viable counts**

The bacteria samples were serially diluted tenfold and 100 µl of diluted bacteria was inoculated on SCD agar. The bacteria were cultured at 35 °C for 24 to 48 h and the number of colonies on SCD agar was counted. The viable cell counts were determined as colony forming units (CFU)/dish.

**Exposure chamber**

We prepared a cubic exposure chamber (1 m³) to evaluate the bactericidal effect of the extremely low concentration gaseous ClO₂ (Fig. 1a). The ClO₂ gas was introduced into the exposure chamber by a ClO₂ gas generator which was made in our laboratory. The humidity in the exposure chamber was constantly controlled by a humidity controller (ADPAC-N1000-AH, ADTEC Corporation, Japan). To maintain a homogeneous concentration of ClO₂ gas in the exposure chamber, the contained air was constantly circulated using two fans (MU1225S-11, ORIX, Japan). The glass dishes inoculated with bacteria were placed in the exposure chamber. The concentration of gaseous ClO₂ in the exposure chamber was measured by a ClO₂ gas analyzer (Midas Gas Detector, MIDAS-E-BR2, Honeywell Analytics, IL, U.S.A.). Furthermore, the determination of concentration of ClO₂ gas was performed using an established protocol (OSHA Method ID-202) with some modifications. Briefly, the ClO₂ gas was collected in a midget impinger (1448-02, SOGORIKAGAKU GLASS WORKS, Japan) containing 20 ml of 1.2 mM potassium iodide (KI) in a 1.5 mM Na₂CO₃/1.5 mM NaHCO₃ buffer solution at 0.5 L/min for 20 min. The collected ClO₂ (as ClO₂⁻) was analyzed by an ion chromatograph (ICS-3000, Thermo Fisher Scientific, U.S.A.).

![Fig. 1 Illustration of the exposure chamber setup and changes in the concentration of gaseous ClO₂ in the exposure chamber. a Schematic diagram of bacteria on glass dishes in an exposure chamber. The bacteria were placed in 1 m³ exposure chamber in wet conditions on glass dishes. b Changes in the concentration of gaseous ClO₂ in the exposure chamber. The error bar in the graph indicates S.D. (n=4). The inset shows correlation between gaseous ClO₂ concentrations (ppbv) by ion chromatography versus measurements using ClO₂ gas analyzer](image)
Scientific (DIONEX), U.S.A.). The temperature and relative humidity in the exposure chamber were measured by a thermo-hygrometer (TR-72 wf, T&D, Japan; accuracy of ±0.3 °C and ±2.5% RH).

**Statistical analysis**

The presence of any significant difference was determined by Student’s *t* test (two-tailed).

**Results**

**Determination of gaseous ClO₂ concentration in the exposure chamber**

The concentration of gaseous ClO₂ measured by the gas analyzer in the exposure chamber is shown in Fig. 1b. As shown in Fig. 1b (inset), there was a linear correlation between gaseous ClO₂ concentrations determined by ion chromatography and measurements by the ClO₂ gas analyzer; the line was then fitted to a linear function by regression analysis. The values obtained by the ClO₂ gas analyzer were corrected using the regression equation. The coefficient of determination *R*² of this fitting was 0.994. The average value of concentration of gaseous ClO₂ was 0.01 ± 0.001 ppmv. The temperature and relative humidity in the exposure chamber were 24.4 ± 0.2 °C and 56.6 ± 1%, respectively.

**Bactericidal activity of gaseous ClO₂ at extremely low concentrations**

We evaluated the bactericidal activity of gaseous ClO₂ at extremely low concentrations against *E. coli*, *P. aeruginosa* and *A. baumannii* in wet conditions on glass dishes. The bactericidal activity of gaseous ClO₂ against *E. coli* exhibited reductions of > 2 log₁₀ (mean 0.01 ppmv, *p* < 0.05) and > 4 log₁₀ (mean 0.03 ppmv, *p* < 0.05) after 2 h as compared to the control values (Fig. 2a). The bactericidal activity of gaseous ClO₂ against *P. aeruginosa* exhibited reductions of > 4 log₁₀ (mean 0.01 ppmv, *p* < 0.01) after 2 h as compared to the control values and achieved undetectable levels (< 1 CFU/dish) after 3 h (mean 0.03 ppmv, *p* < 0.05) (Fig. 2b). The bactericidal activity of gaseous ClO₂ against *A. baumannii* showed reductions of > 2 log₁₀ (mean 0.01 ppmv, *p* < 0.01) and > 4 log₁₀ (mean 0.03 ppmv, *p* < 0.01) after 3 h as compared to the control values (Fig. 2c).

**Bactericidal activity of gaseous ClO₂ under dirty conditions**

Efficacy of gaseous ClO₂ against microbes in wet conditions was decreased in the presence of fetal bovine serum (FBS) as an organic substance [3]. Hence, we examined the effect of an FBS load (0, 3, 5 and 10%) on the inactivation of *P. aeruginosa*, which had the highest sensitivity to gaseous ClO₂ of the three evaluated bacteria, with a ClO₂ gas of concentration of 0.01 ppmv. The extremely low concentration gaseous ClO₂ (0.01 ppmv) inactivated *P. aeruginosa* with 5% FBS (2.9 log₁₀ reductions) within 3 h (Table 1).

**Discussion**

Gaseous agents and disinfectant vapors have excellent diffusive characteristics. Such agents can disinfect areas where liquid agents are difficult to use. However, many procedures using formaldehyde, peracetic acid, etc. have disadvantages, for example, skin inflammation...
The extremely low-concentration gaseous ClO₂ (0.01 ppmv) used to inactivate GNB in this research was only 1/10 of that concentration. In a previous study, we demonstrated that gaseous ClO₂ at 0.1 ppmv was not toxic when whole bodies of rats were exposed to the gas for 6 months [6]. Furthermore, Ogata et al. reported that the no observed adverse effect level (NOAEL) of gaseous ClO₂ was 1 ppmv [7]. From these results, we think that use of extremely low-concentration gaseous ClO₂ (0.01 ppmv) is a feasible method for inactivating GNB in the presence of humans, without adverse effects.

In this study, the bactericidal activity against multidrug-resistant (MDR)-GNB of gaseous ClO₂ of extremely low-concentration was not determined. However, a ClO₂ solution of 10 mg/L drastically reduced the number of MDR-P. aeruginosa and MDR-A. baumannii bacteria within 60 s under conditions of a mixture containing a high concentration of bovine serum albumin and sheep erythrocytes [8]. Furthermore, the effect of inactivation by a ClO₂ solution of 10 mg/L against drug-sensitive P. aeruginosa (1.4 log₁₀ reductions, within 15 s) was lower than that against MDR-P. aeruginosa (3.6 log₁₀ reductions, within 15 s). In other words, MDR-P. aeruginosa are more sensitive to ClO₂ than drug-sensitive P. aeruginosa. These data suggest that extremely low-concentration gaseous ClO₂ may have a bactericidal effect against MDR-GNB.

In a previous study, we discussed the fact that moisture plays an important role in the inactivation of feline calicivirus on glass dishes by < 0.1 ppmv ClO₂ gas [9]. Hence, in this study we evaluated bactericidal activity against GNB of extremely low-concentration gaseous ClO₂ in wet conditions, but not in the dry state. Previous studies are not sufficient to clarify the route of infection for GNB in wet environments such as sinks and bathroom. However, several studies suggest that a watery environment as occurs in kitchens, drains, bathrooms, sinks and faucets serves as a reservoir for microorganisms [10, 11]. Barker et al. showed that flushing a toilet produces aerosols containing microbes that are capable of causing surface contamination within the toilet [12]. In this study, gaseous ClO₂ at an extremely low-concentration inactivated GNB in wet conditions on glass dishes as a model of microbes on surfaces in a wet environment. In addition, the extremely low-concentration gaseous ClO₂ inactivated P. aeruginosa with 5% FBS (2.9 log₁₀ reductions) within 3 h. The percent FBS contained in a microbial suspension (minimum 5% FBS) was designed by the United States Environmental Protection Agency (US EPA) for virucidal effectiveness testing using feline calicivirus [13]. It should be noted that the effect of inactivation of P. aeruginosa by extremely low-concentration gaseous ClO₂ was considerably decreased in the presence of 10% FBS. Therefore, its inactivating effect may be limited under dirty conditions (> 5% FBS). In addition, we examined the effect of an erythrocytes load (final 0.3% (w/v) BSA and 0.3% (v/v) erythrocytes) on the inactivation of P. aeruginosa by extremely low-concentration gaseous ClO₂. The extremely low-concentration gaseous ClO₂ (0.01 ppmv) inactivated P. aeruginosa with erythrocytes (1.0 log₁₀ reductions) within 3 h. Taken together, treatment of moist environments with extremely low-concentration gaseous ClO₂ may help to reduce the risk of environmental infection by GNB without harmful effects.

### Limitations

We were not able to determine the bactericidal activity of gaseous ClO₂ of extremely low-concentration against multidrug-resistant (MDR)-GNB. We were also not able to evaluate the effect of organic substance load on the inactivation of E. coli and A. baumannii by extremely low-concentration gaseous ClO₂. We were also not able to evaluate bactericidal activity of gaseous ClO₂ at extremely low-concentrations against GNB in environments.

### Abbreviations

GNB: Gram-negative bacteria; ClO₂: Chlorine dioxide; SCD: Soybean Casein Digest; HAI: Healthcare-associated infections; OSHA: Occupational Safety and Health Administration; NITE: National Institute of Technology and Evaluation; D-PBS: Dulbecco’s phosphate buffer saline; CFU: Colony forming units; MDR: Multidrug-resistant; OD: Optical Density; BSA: Bovine serum albumin.

### Acknowledgements

Not applicable.

### Authors’ contributions

HM, TM and TS contributed to the conception and study design. HM and MF performed experiments and statistical analyses. HM wrote the draft manuscript and prepared figures. All authors read the draft manuscript and
provided feedback about the draft manuscript. All authors read and approved the final manuscript.

Funding
This work was supported by Taiko Pharmaceutical Co., Ltd.

Availability of data and materials
All data generated or analyzed during this study are included in this published article.

Ethics approval and consent to participate
Not applicable.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

Author details
1 Taiko Pharmaceutical Co., Ltd., Uchihonmachi 3-34-14, Suita, Osaka 564-0032, Japan. 2 Kyoto Plant/R&D Center, 1-2-1 Hikanai, Seikacho, Soraku-gun, Kyoto 619-0237, Japan.

Received: 25 December 2019 Accepted: 28 January 2020
Published online: 12 February 2020

References
1. Tacconelli E, Cataldo MA, Dancer SJ, De Angelis G, Falcone M, Frank U, Kahlmeter G, Pan A, Petrosillo N, Rodríguez-Baño J, Singh N, Venditti M, Yokoe DS, Cookson B. ESCMID guidelines for the management of the infection control measures to reduce transmission of multidrug-resistant Gram-negative bacteria in hospitalized patients. Clin Microbiol Infect. 2014;20:1–55.
2. Rodrigo-Troyano A, Sibila O. The respiratory threat posed by multidrug resistant Gram-negative bacteria. Respology. 2017;22:1288–99.
3. Morino H, Fukuda T, Miura T, Shibata T. Effect of low-concentration chlorine dioxide gas against bacteria and viruses on a glass surface in wet environments. Lett Appl Microbiol. 2011;53:628–34.
4. Ogata N, Sakasegawa M, Miura T, Shibata T, Takigawa Y, Taura K, Taguchi K, Matsumura K, Nakahara K, Kato D, Sagawa K, Oka H. Inactivation of airborne bacteria and viruses using extremely low concentrations of chlorine dioxide gas. Pharmacology. 2016;97:301–6.
5. Occupational Safety and Health Administration (OSHA), United States Department of Labor. OSHA Annotated Table Z-1, https://www.osha.gov/dsg/annotated-pels/tablez-1.html.
6. Akamatsu A, Lee C, Morino H, Miura T, Ogata N, Shibata T. Six-month low level chlorine dioxide gas inhalation toxicity study with two-week recovery period in rats. J Occup Med Toxicol. 2012;7:1–8.
7. Ogata N, Koizumi T, Ozawa F. Ten-week whole-body inhalation toxicity study of chlorine dioxide gas in rats. J Drug Metab Toxicol. 2013;4:1–5.
8. Hinenoya A, Awasthi SP, Yasuda N, Shima A, Morino H, Koizumi T, Fukuda T, Miura T, Shibata T, Yamasaki S. Chlorine dioxide is a better disinfectant than sodium hypochlorite against multi-drug resistant Staphylococcus aureus, Pseudomonas aeruginosa, and Acinetobacter baumannii. Jpn J Infect Dis. 2015;68:276–9.
9. Morino H, Fukuda T, Miura T, Lee C, Shibata T, Sanekata T. Inactivation of feline calicivirus, a norovirus surrogate, by chlorine dioxide gas. Biocontrol Sci. 2009;14:147–53.
10. Kizny Gordon AE, Mathers AJ, Cheong EYL, Gottlieb T, Kotay S, Walker AS, Peto TEA, Crook DW, Stoesser N. The hospital water environment as a reservoir for carbapenem-resistant organisms causing hospital-acquired infections: a systematic review of the literature. Clin Infect Dis. 2017;64:1435–44.
11. Kagan LJ, Aiello AE, Larson E. The role of the home environment in the transmission of infectious diseases. J Community Health. 2002;27:247–67.
12. Barker J, Jones MV. The potential spread of infection caused by aerosol contamination of surfaces after flushing a domestic toilet. J Appl Microbiol. 2005;99:339–47.
13. United States Environmental Protection Agency (US EPA), virucidal effectiveness testing using feline calicivirus as surrogate for Norovirus, https://www.epa.gov/pesticide-registration/virucidal-effectiveness-testing-using-FELINE-calicivirus-surrogate-norovirus.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.