3D Printing of Artificial Blood Vessel: Study on Multi-Parameter Optimization Design for Vascular Molding Effect in Alginate and Gelatin

Huanbao Liu 1, Huixing Zhou 1,2,*, Haiming Lan 1, Tianyu Liu 1, Xiaolong Liu 1 and Hejie Yu 1

1 College of Engineering, China Agricultural University, Beijing 100083, China; leaderliu@cau.edu.cn (H.L.); bs20153070217@cau.edu.cn (H.L.); perc_lty@cau.edu.cn (T.L.); perc_bxl@cau.edu.cn (X.L.); yuhejie@cau.edu.cn (H.Y.)
2 School of Mechanical-Electronic Vehicular Engineering, Beijing University of Civil Engineering and Architecture, Beijing 100044, China

* Correspondence: zhouhuixing@bucea.edu.cn; Tel.: +86-010-6273-7742

Received: 30 June 2017; Accepted: 29 July 2017; Published: 31 July 2017

Abstract: 3D printing has emerged as one of the modern tissue engineering techniques that could potentially form scaffolds (with or without cells), which is useful in treating cardiovascular diseases. This technology has attracted extensive attention due to its possibility of curing disease in tissue engineering and organ regeneration. In this paper, we have developed a novel rotary forming device, prepared an alginate–gelatin solution for the fabrication of vessel-like structures, and further proposed a theoretical model to analyze the parameters of motion synchronization. Using this rotary forming device, we firstly establish a theoretical model to analyze the thickness under the different nozzle extrusion speeds, nozzle speeds, and servo motor speeds. Secondly, the experiments with alginate–gelatin solution are carried out to construct the vessel-like structures under all sorts of conditions. The experiment results show that the thickness cannot be adequately predicted by the theoretical model and the thickness can be controlled by changing the parameters. Finally, the optimized parameters of thickness have been adjusted to estimate the real thickness in 3D printing.

Keywords: cardiovascular disease; 3D printing; alginate–gelatin; optimized parameters

1. Introduction

Cardiovascular disease is a common disease that seriously affects the health of human beings, especially those over 50 years old. Many scientists [1–6] have adopted a lot of measures to treat cardiovascular disease, but more than 50% of survivors cannot completely take care of themselves. The number of deaths from cardiovascular diseases is up to 15 million people each year globally, which top killer of human beings. Therefore, it is significant to develop an effective method to fabricate vessel-like structures to treat cardiovascular disease.

3D printing is a novel technology to be applied to regenerative medicine that has attracted extensive attention due to its potential to cure disease through tissue engineering and organ regeneration [7–12]. In the past few years, many functional tissues and organs were manufactured by 3D printing systems such as inkjet-based printing [13–15], micro-extrusion printing [16], and laser-assisted printing [17–19], which have different forming parameters in different 3D printing system. One important challenge is the material viscosity characteristic which can determine the types of 3D printers. The inkjet-based printing has viscosity a limit ranging from 3.5 to 12 mPa·s and the viscosity limit of laser-assisted printing ranges from 1 to 300 mPa·s, but the viscosity limit of extrusion-based printing ranges from 30 to >6 × 10⁷ mPa·s. The different working mechanisms and parameters of nozzles results in different viscosity limits.
Many research teams have adopted the above 3D printing strategies to fabricate vessel-like structures with deposition modeling methods [20–22]. Cui, X. et al. [23] have used thermal inkjet printing to fabricate the human microvasculature. Then Hoch, E. et al. [24] have shown that inkjet-based 3D bioprinting can be expected to be of great use for vessel-like structures. Gaebel, R. et al. [25] have adopted laser printing to distribute the endothelial cells to fabricate blood vessels. Kucukgul, Can et al. [26] have adopted a deposition modeling method to construct vessel-like structures by extrusion-based 3D bioprinting. In those methods, the vessel-like structures have poor modeling effect and material accumulation phenomena. However, there is little research into adopting the rotary forming method to fabricate vessel-like structures. Gao, Q. et al. [27] have adopted the rotary printing method to fabricate vessel-like structures; it can be proven that this method can improve the cell survival rate. Meanwhile, Liu, H. et al. [28,29] have adopted the rotary printing method to obtain vessel-like structures and analyzed the synchronization among nozzle extrusion, nozzle speed, and rotating speed based on an extrusion-based 3D bioprinter, which could improve the modeling effect in this forming method.

In this paper, we present our 3D printing device for fabricating vessel-like structures, and further optimize the thickness parameters of nozzle extrusion speed nozzle speed and rotating speed.

The nozzle moving speed parameter $\delta$ and the pressure parameter $\nabla$ have been adjusted to estimate the real thickness in the experiment.

2. System and Process

Figure 1 shows the extrusion-based 3D printing system, which is mainly composed of a motion control system, multinozzle distribution system, rotary forming system, and temperature control system. The multinozzle 3D printing system is a solid free-form fabrication system that performs extrusion-based processes. Two nozzles with different biomaterials can work effectively in this 3D printing system which can be controlled by computer. The specification of the 3D printing system is shown in Table 1.

Figure 1. The extrusion-based 3D printing system with rotary printing device.
When the nozzle speed is less than \( V_1 \) which were sliced to generate the G-code in the computer, and then each region was filled with different biomaterials. The biomaterials can be squeezed by pressure control system, the rotating rod is controlled by the motor, and the nozzle speed is controlled by linear motor. In the Figure 2a, the biomaterials can be liquefied in suitable temperatures (35–40 °C). Secondly, the temperature control system can control the material temperature to achieve the conversion between solid and liquid states. In addition, there is no heating damage for bioactivity in this temperature control system.

3. Theory

Figure 2 shows the schematic diagram of rotary printing device under the different nozzle speed. The biomaterials can be squeezed by pressure control system, the rotating rod is controlled by the motor, and the nozzle speed is controlled by linear motor. In the Figure 2a, \( V_1 \), \( V_2 \), and \( V_3 \) denote the nozzle speed, the extrusion speed, and the motor speed respectively. When the nozzle speed is greater than \( V_1 \), the biomaterials will demonstrate the dispersion phenomenon, as shown in Figure 2b. When the nozzle speed is less than \( V_1 \), the biomaterials will demonstrate accumulation, as shown in Figure 2c.

In this 3D printing system, 3D physical models (STL) were divided into different regions, which were sliced to generate the G-code in the computer, and then each region was filled with different materials. The STL files can be obtained by CAD/CAM software or 3D scanners.

| Parameters                  | Value                  |
|-----------------------------|------------------------|
| Dimensions \((X \times Y \times Z)\) | 150 × 150 × 150 mm³   |
| Position resolution         | ±5 μm                  |
| Temperature range           | 0–60 °C                |
| Print speed                 | 0.1–50 mm/s            |
| Pressure range              | 0–1 MPa                |

Table 1. The specification of 3D printing system.

In order to realize the reconstruction of vessel-like structures and avoid the phenomenon of material accumulation and material dispersion, the theoretical model can be obtained by this rotary printing device, which is given that [28]

\[
V_3 = V_1 + V_2
\] (1)

Figure 2. The schematic diagram of rotary printing device under the different nozzle speed: (a) ideal state; (b) nozzle moving speed greater than \( V_1 \); (c) nozzle moving speed less than \( V_1 \).
Because equal time is consumed in the building process of vessel-like structures, the relationship among nozzle speed, extrusion speed, and motor speed is calculated as

\[
\frac{S_1}{V_1} = \frac{S_2}{V_2} = \frac{S_3}{V_3}
\]  

(2)

where \( S_1, S_2, \) and \( S_3 \) refer to linear motor displacement, rotating rod perimeter, and material extrusion displacement.

The thickness can be derived by Equations (1) and (2)

\[
H = \alpha \sqrt{(V_3L/2V_1)^2 - L^2 - r} \\
H = \alpha \sqrt{(V_3C/2V_2)^2 - L^2 - r}
\]  

(3)

where \( H \) is the thickness, \( L \) is the displacement of nozzle, \( \alpha \) is an invariant constant, and \( r \) is the rotating rod radius.

The extrusion forming principle under the different nozzle moving speeds is shown in Figure 3. The thickness can be changed under the different control parameters. When the diameter of rotating rod and motor rotation speed remain unchanged, the displacement relation under the different nozzle moving speed can be calculated as

\[
\begin{align*}
S_{1''} & > S_{1'} > S_1 \\
S_{2''} & = S_{2'} = S_2 \\
S_{3''} & > S_{3'} > S_3
\end{align*}
\]  

(4)

The goal is to predict displacement changes which have different forming effects under different nozzle moving speeds. In order to obtain the real thickness, the new thickness coefficient can be adjusted to measure the thickness range.

**Figure 3.** The relationship between the displacement and thickness under the different nozzle moving speed due to surface wettability.

### 4. Experiment Procedure

#### 4.1. Materials

In order to optimize the above mentioned theoretical model, we designed the experiment to fabricate the vessel-like structures. The rotating rods with different diameters of 3, 4.26, and 6.9 mm were used in this experiment. We selected alginate and gelatin as experiment materials. In the experiment, alginate and gelatin were premixed, extruded, and post crosslinked after depositing on the rotating rod in the 3D printing system. The alginate and gelatin solution was cross-linked with calcium chloride (5% \( w/w \)). The parameters of experiment materials are shown in Table 2. All of the above materials were purchased from Xilong Scientific Co., Ltd. (Guangdong, China).
Therefore, the alginate solution with gelatin can improve the material characteristics such as viscosity, structures. Compared with the deposition modeling, this method does not build the scaffold to sustain mechanical properties, and printable characteristics. The viscosity value increased gradually along with the increase of gelatin concentration. In Figure 4b, we can see that the 3% alginate with different concentrations of gelatin (4%, 6%, and 8%) shows an increase in viscosity with the increase of alginate concentration. In Figure 4a, we observe that the viscosity of alginate solution increases with the increase of alginate concentration.

5. Results and Discussion

5.1. Material Viscosity

Viscosity is an important factor which plays a major role in fabricating the vessel-like structures. Figure 4 shows the relationship between viscosity and material concentration. In Figure 4a, the viscosity of alginate solution increases with the increase of alginate concentration. In Figure 4b, we can see that the 3% alginate with different concentrations of gelatin (4%, 6% and 8% w/w) have different viscosity characteristics. The viscosity value increased gradually along with the increase of gelatin concentration. Therefore, the alginate solution with gelatin can improve the material characteristics such as viscosity, mechanical properties, and printable.

![Figure 4](image_url)

**Figure 4.** Dependence of material concentration on viscosity: (a) the relationship between viscosity and concentration of alginate; (b) the relationship between viscosity and concentration of gelatin under the concentration of alginate (3% w/w).

In this experiment, the extrusion-based nozzle has been developed to fabricate the vessel-like structures. Compared with the deposition modeling, this method does not build the scaffold to sustain...
the vessel-like structures, which can reduce time consumption in the build process of vessel-like structure. In order to improve the forming effect, the adaptive parameters should be set to avoid the problems of material dispersion and material accumulation, as shown in Figure 5. Those problems will reduce the forming accuracy and mechanical properties.

![Figure 5](image_url)  
**Figure 5.** The problems of material dispersion and material accumulation.

Dependence of the extrusion pressure on molding effect: In this experiment, we have adopted a solution with 3% alginate and 8% gelatin for initial research. When the rotating speed was 4.25 r/s and nozzle moving speed was 1.62 mm/s, the vessel-like structures exhibited different molding effects under different pressures, as shown in Figure 6. The different external diameters or thicknesses can be achieved under the different extrusion pressures. Figure 6a shows the relationship between thickness and extrusion pressure. We can see that the thickness is on the increase with the increase of pressure. The molding effects are shown in Figure 6b. When the pressure exceeds the 0.31 Mpa, the molding effect appears as material accumulation, which leads to a poor forming effect. Conversely, when the pressure is below the 0.16 MPa, the vessel-like structures will not be fabricated by this 3D printing system.

![Figure 6](image_url)  
**Figure 6.** The molding relationship and effect: (a) dependence of the extrusion pressure on thickness under different diameters; (b) the relationship between molding effect and diameter.

Figure 7 shows the relationship between the printable area and extrusion pressure under the different diameters. We can see that the relatively stable thickness region can be derived to analyze the
forming state of vessel-like structures. Based on this, the parameter of the theoretical model can be readjusted to estimate the thickness of vessel-like structures. Based on these analysis results, we can draw a conclusion that the optimized model parameter $\nabla$ is derived

$$H = \nabla \beta$$ (5)

where $\beta$ is $a \sqrt{(V_3 C / 2V_2)^2 - L^2 - r}$, $\nabla$ is the optimized parameter ranged from 0.45 to 2.45.

**Figure 7.** The relationship between the printable area and diameter under the different extrusion pressure.

Dependence of the moving speed of nozzle on molding effect: The controllability of materials plays a significant role in the construction of vessel-like structures. In the case of high pressure, the materials are difficult to be controlled in this 3D printing system. Meanwhile, the materials are difficult to be extruded under low pressure circumstances. To overcome the above-mentioned drawbacks, the pressure and the rotating speed were set to 0.224 MPa and 4.25 r/s, respectively. The vessel-like structures have the different thicknesses under the different nozzle moving speed. The relationship between thickness and moving speed of nozzle is shown in Figure 8. The conclusion is that the thickness is on the decline with the increase of nozzle moving speed. When the nozzle moving speed is less than 0.75 mm/s, the vessel-like structure will present material accumulation. Conversely, when the nozzle moving speed exceeds the certain values, vessel-like structures will not form.

**Figure 8.** The molding relationship and effect: (a) dependence of the nozzle moving speed on thickness under the different diameters; (b) the relationship between molding effect and diameter.
Parameter optimization design: Figure 9 shows the relationship between the printable area and the different diameters under different nozzle moving speeds. Similarly to the Figure 7, the vessel-like structures have a relatively stable thickness region. Based on this, the parameters of the theoretical model can be readjusted to estimate the thickness of vessel-like structures.

\[ H = \delta \mu \]  

(6)

where \( \mu \) is \( \sqrt{(V_3L/2V_1)^2 - L^2 - r} \). \( \delta \) is the optimized parameter ranged from 0.447 to 2.44.

![Figure 9. The relationship between the printable area and diameter under the nozzle moving speed.](image)

Based on the optimized analysis results under the different pressure and nozzle moving speed, the optimized parameters of the theoretical model can be derived from the rotating forming device. The parameter of pressure is similar to the parameter of nozzle moving speed, namely \( \delta \approx \nabla \). It is proven that the rotating rods with different diameters have a similar range under constant viscosity. Although the thickness characteristics of vessel-like structures are similar between the different pressures and nozzle moving speeds, we will intend to control the nozzle moving speed under a constant pressure due to the time delay in the process of material extrusion.

6. Conclusions

In conclusion, we have designed a novel 3D printing system using a rotary forming device to fabricate vessel-like structures for theoretical analysis, and its feasibility has been shown in this study. Compared with the deposition modeling method, those vessel-like structures produced by rotary forming method not only have higher forming accuracy, but also have faster molding speed. Using this rotary forming device, we have proposed a theoretical model to analyze motion synchronization. The experiments with alginate–gelatin solution are carried out to verify the theoretical results. The experiment results show that the thickness cannot be adequately predicted by the theoretical model and the thickness can be controlled by changing the correlation parameters. We have optimized the parameters under different pressures and nozzle moving speeds in the process of 3D printing, in which the parameter of pressure is similar to the parameter of nozzle moving speed, namely \( \delta \) and \( \nabla \). This optimized theoretical model can be used to predict the thickness of the alginate–gelatin material. Meanwhile, it has been proven that extrusion-based 3D printing with alginate and gelatin could afford an opportunity to treat cardiovascular disease due to the possibility of angiogenesis in tissue engineering and organ regeneration.
Acknowledgments: This work is supported by the national high technology research and development program of China (863 program) under Grant 2015AA020305-01.

Author Contributions: Huanbao Liu conceived the problem and designed the solution; Huixing Zhou conceived and designed the experiments; Haiming Lan and Tianyu Liu prepared the biological materials; Xiaolong Liu revised and improved the manuscript for the final submission; Huanbao Liu and Hejie Yu analyzed the data.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Jia, W.; Gungor-Ozkerim, P.S.; Zhang, Y.S.; Yue, K.; Zhu, K.; Liu, W.; Pi, Q.; Byambaa, B.; Dokmeci, M.R.; Shin, S.R.; et al. Direct 3D bioprinting of perfusable vascular constructs using a blend bioink. *Biomaterials* 2016, 106, 58–68. [CrossRef] [PubMed]
2. Zhang, Y.; Yu, Y.; Akkouch, A.; Dababneh, A.; Dolati, F.; Ozbolat, I.T. In Vitro Study of Directly Bioprinted Perfusable Vasculature Conduits. *Biomater. Sci.* 2015, 3, 134–143. [CrossRef] [PubMed]
3. Can, K.; Ozler, S.B.; Inci, I.; Karakas, E.; Irmak, S.; Gozuacik, D.; Taralp, A.; Koc, B. 3D bioprinting of biomimetic aortic vascular constructs with self-supporting cells. *Biotechnol. Bioeng.* 2015, 112, 811–821. [CrossRef]
4. Tae-Yun, K.; Jung Ho, L.; Bum Jin, K.; Jo, A.K.; Jung Min, H.; Byoung Soo, K.; Hyung Joon, C.; Jong-Won, R.; Dong-Woo, C. In vivo endothelization of tubular vascular grafts through in situ recruitment of endothelial and endothelial progenitor cells by RGD-fused mussel adhesive proteins. *Biofabrication* 2015, 7, 015007. [CrossRef]
5. Poldervaart, M.T.; Gremmels, H.; van Deventer, K.; Fledderous, J.O.; Oner, F.C.; Verhaar, M.C.; Dhert, W.J.; Alblass, J. Prolonged presence of VEGF promotes vascularization in 3D bioprinted scaffolds with defined architecture. *J. Control. Release* 2014, 184, 58–66. [CrossRef] [PubMed]
6. Yu, Y.; Zhang, Y.; Martin, J.A.; Ozbolat, I.T. Evaluation of Cell Viability and Functionality in Vessel-like Bioprintable Cell-Laden Tubular Channels. *J. Biomach. Eng.* 2013, 135, 1–9. [CrossRef] [PubMed]
7. Murphy, S.V.; Atala, A. 3D bioprinting technology for engineering complex tissues. *Biotechnol. Adv.* 2016, 34, 422–434. [CrossRef] [PubMed]
8. Dhakshinamourthy, S.; Rauf, S.; Hauser, C. 3D bioprinting technology for regenerative medicine applications. *Int. J. Bioprint.* 2016, 2. [CrossRef]
9. Falguni, P.; Gantelius, J.; Svahn, H.A. 3D Bioprinting of Tissue/Organ Models. *Angew. Chem. Int. Ed.* 2016, 55, 4650–4665. [CrossRef]
10. Poulos, S.J.; Miller, J.S. Tissue vascularization through 3D printing: Will technology bring us flow? *Dev. Dyn.* 2015, 244, 629–640. [CrossRef] [PubMed]
11. Liu, H.; Zhou, H.; Lan, H.; Liu, T. Organ regeneration: Integration application of cell encapsulation and 3D bioprinting. *Virtual Phys. Prototyp.* 2017, 1–11. [CrossRef]
12. Nishiyama, Y.; Nakamura, M.; Henmi, C.; Yamaguchi, K.; Mochizuki, S.; Nakagawa, H.; Takiura, K. Development of a three-dimensional bioprinter: Construction of cell supporting structures using hydrogel and state-of-the-art inkjet technology. *J. Biomach. Eng.* 2009, 131, 035001. [CrossRef] [PubMed]
13. VanOsch, T.H.J.; Peraelaer, J.; de Laat, A.W.M.; Schubert, U.S. Inkjet Printing of Narrow Conductive Tracks on Untreated Polymeric Substrates. *Adv. Mater.* 2008, 20, 343–345. [CrossRef]
14. Saunders, R.E.; Brian, D. Inkjet printing biomaterials for tissue engineering: Bioprinting. *Int. Mater. Rev.* 2014, 59, 430–448. [CrossRef]
15. Zhao, Y.; Li, Y.; Mao, S.; Sun, W.; Yao, R. The influence of printing parameters on cell survival rate and printability in microextrusion-based 3D cell printing technology. *Biofabrication* 2015, 7, 045002. [CrossRef] [PubMed]
16. Lothar, K.; Brandt, O.; Deiwick, A.; Chichkov, B. Laser assisted bioprinting at different wavelengths and pulse durations with a metal dynamic release layer: A parametric study. *Int. J. Bioprint.* 2017, 3. [CrossRef]
17. He, P.; Katis, I.N.; Eason, R.W.; Sones, C.L. Laser direct-write for fabrication of three-dimensional paper-based devices. *Lab Chip* 2016, 16, 3296–3303. [CrossRef] [PubMed]
19. Bourget, J.M.; Kerouredan, O.; Medina, M.; Remy, M.; Thebaud, N.B.; Bareille, R.; Chassande, O.; Amedee, J.; Catros, S.; Devillard, R. Patterning of Endothelial Cells and Mesenchymal Stem Cells by Laser-Assisted Bioprinting to Study Cell Migration. BioMed Res. Int. 2016, 3569843. [CrossRef] [PubMed]

20. Norotte, C.; Marga, F.S.; Niklason, L.E.; Forgacs, G. Scaffold-free vascular tissue engineering using bioprinting. Biomaterials 2009, 30, 5910–5917. [CrossRef] [PubMed]

21. Duan, B.; Hockaday, L.A.; Kang, K.H.; Butcher, J.T. 3D bioprinting of heterogeneous aortic valve conduits with alginate/gelatin hydrogels. J. Biomed. Mater. Res. A 2013, 101, 1255–1264. [CrossRef] [PubMed]

22. Kolesky, D.B.; Truby, R.L.; Gladman, A.S.; Busbee, T.A.; Homan, K.A.; Lewis, J.A. 3D bioprinting of vascularized, heterogeneous cell-laden tissue constructs. Adv. Mater. 2014, 26, 3124–3130. [CrossRef] [PubMed]

23. Cui, X.; Boland, T. Human microvasculature fabrication using thermal inkjet printing technology. Biomaterials 2009, 30, 6221–6227. [CrossRef] [PubMed]

24. Hoch, E.; Tovar, G.E.; Borchers, K. Bioprinting of artificial blood vessels: Current approaches towards a demanding goal. Eur. J. Cardio-Thorac. Surg. 2014, 46, 767–778. [CrossRef] [PubMed]

25. Gaebel, R.; Ma, N.; Liu, J.; Guan, J.; Koch, L.; Klopsch, C.; Gruene, M.; Toelk, A.; Wang, W.; Mark, P.; et al. Patterning human stem cells and endothelial cells with laser printing for cardiac regeneration. Biomaterials 2011, 32, 9218–9230. [CrossRef] [PubMed]

26. Kucukgul, C.; Ozler, S.B.; Inci, I.; Karakas, E.; Irmak, S.; Gozuacik, D.; Taralp, A.; Koc, B. 3D bioprinting of biomimetic aortic vascular constructs with self-supporting cells. Biotechnol. Bioeng. 2015, 112, 811–821. [CrossRef] [PubMed]

27. Gao, Q.; Liu, Z.; Lin, Z.; Qiu, J.; Liu, Y.; Liu, A.; Wang, Y.; Xiang, M.; Chen, B.; Fu, J.; et al. 3D Bioprinting of Vessel-like Structures with Multilevel Fluidic Channels. ACS Biomater. Sci. Eng. 2017, 3, 399–408. [CrossRef]

28. Liu, H.; Zhou, H.; Chairinnas. The synchronization among nozzle extrusion, nozzle speed and rotating speed based on 3D vessel bioprinter. In Proceedings of the 2016 International Conference on Instrumentation, Control and Automation (ICA), Bandung, Indonesia, 29–31 August 2016.

29. Liu, H.; Zhou, H.; Lan, H.; Liu, F.; Wang, X. Multinozzle Multichannel Temperature Deposition System for Construction of a Blood Vessel. SLAS Technol. 2017. [CrossRef] [PubMed]