**In Silico** Modeling of Human α2C-Adrenoreceptor Interaction with Filamin-2

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### Abstract

Vascular smooth muscle α2C-adrenoceptors (α2C-ARs) mediate vasoconstriction of small blood vessels, especially arterioles. Studies of endogenous receptors in human arteriolar smooth muscle cells (referred to as microVSM) and transiently transfected receptors in heterologous HEK293 cells show that the α2C-ARs are perinuclear receptors that translocate to the cell surface under cellular stress and elicit a biological response. Recent studies in microVSM unraveled a crucial role of Rap1A-Rho-ROCK-F-actin pathways in receptor translocation, and identified protein-protein interaction of α2C-ARs with the actin binding protein filamin-2 as an essential step in the process. To better understand the molecular nature and specificity of this interaction, in this study, we constructed comparative models of human α2C-AR and human filamin-2 proteins. Finally, we performed in silico protein-protein docking to provide a structural platform for the investigation of human α2C-AR and filamin-2 interactions. We found that electrostatic interactions seem to play a key role in this complex formation which manifests in interactions between the C-terminal arginines of α2C-ARs (particularly R454 and R456) and negatively charged residues from filamin-2 region between residues 1979 and 2206. Phylogenetic and sequence analysis showed that these interactions have evolved in warm-blooded animals.

### Introduction

The α2-adrenoceptors (α2-ARs) are members of the G protein-coupled family of receptors (GPCRs), which is one of the largest families of proteins in the human genome [1,2]. GPCRs are known to mediate important physiological functions and therefore, are targets for many current drugs; It has been estimated that 30% of major drugs target these receptors [3]. Three human α2-AR subtypes have been cloned and designated α2C10, α2C2 and α2C4 based on their human chromosomal localization, and subsequently renamed to α2A-ARs, α2B-ARs and α2C-ARs, respectively [4]. Phylogenetic classification of α2-ARs shows that they belong to the biogenic amine receptor cluster of the α-Group of Rhodopsin receptors [5].

Within the three α2-ARs subtypes, α2C-ARs have unique regulation, cellular localization and trafficking profile in the human and murine microvasculature. The α2C-ARs modulate blood flow and are preferentially expressed in the smooth muscle cells of the microcirculation, particularly arterioles [6]. The α2C-ARs mediate vasoconstriction upon stimulation by the endogenous agonist norepinephrine (noradrenaline) and therefore, have a unique and crucial role in physiology and pathophysiology involving the peripheral circulation [7,8]. It is therefore important to understand mechanisms of receptor expression and trafficking for a clear understanding of α2C-AR biology. The α2C-ARs are intracellular receptors that are known to translocate to the cell surface under stress conditions such as cold temperature. They play a vital role in skin thermoregulation [3,9]. In heterologous HEK 293 cells, α2C-ARs are present in the transGolgi at physiological 37°C temperature. Moderate cooling to 28°C leads to cell surface translocation of functional α2C-ARs [10]. The mechanism of cooling-triggered translocation involves release of mitochondrial reactive oxygen species, activation of RhoA-ROCK signaling, and receptor cell surface translocation [11,12]. Recent
studies have identified a temperature-independent (i.e. physiological 37°C coupled), and cyclic AMP (cAMP)-dependent mechanism of receptor expression and translocation coupled to the cAMP receptor Epac and Rap1A-Rho-ROCK signaling pathway [13]. Cyclic AMP leads to increased transcription of α2C-ARs through JNK-c-jun nuclear signaling and increased cell surface translocation of mature receptors through RhoA-ROCK signaling and F-actin coupled pathway [13,14]. Therefore, divergent signaling pathways, including cooling-triggered or cAMP-triggered converge on a common pathway, are necessary for receptor translocation to the cell surface.

More recent studies have identified protein-protein interaction between the α2C-AR carboxyl terminus and the actin-binding protein filamin-2 in mediating cell surface translocation of intracellular receptors [15]. In this study we performed computational modeling of α2C-AR to filamin-2 binding in order to better understand protein-protein specificity of this interaction. Our studies show that this approach complements and supports the experimental approaches utilized in previous studies [15].

**Materials and Methods**

**Sequence analyses**

α2C-adrenoceptors (ADRA2C). Searches of human α2C-adrenoceptor homologs were carried out using a locally installed version of PSI-BLAST algorithm [16] against the non-redundant (nr) version of the NCBI sequence database (as of June, 2014). The gapped blast algorithm (blastingp) with the expectation value (E-value) threshold for the retrieval of related sequences set to 0.001. Three iterations of PSI-BLAST were run, and all sequences from hits with an expectation value better than 0.001 were retrieved.

**Sequence clustering.** α2C-adrenoceptors belong to a large family of G protein-coupled receptors [1,2]. Hence, a homology search, yielded 72,730 proteins. To facilitate further analysis, we applied sequence clustering using CLANS [17], to group these sequences into families. CLANS (Cluster ANalysis of Sequences), is a Java program that applies a version of the Fruchterman-Reingold graph layout algorithm for visualizing protein families based on pairwise similarity. This algorithm helps to represent the force between any two nodes, where each node represents a pair of proteins. In order to draw graphs in an aesthetically pleasing way, the algorithm has to minimize the energy of the system by moving the nodes and changing the forces between them. CLANS uses the P-values of high-scoring segment pairs (HSPs) obtained from an N×N BLAST search, to compute attractive and repulsive forces between each sequence pair in a user-defined dataset. Two or three dimensional representation is achieved by randomly seeding sequences in space. The sequences are then moved within this environment according to the force vectors resulting from all pairwise interactions and the process is repeated to achieve convergence.

**Multiple Sequence alignment.** All sequences classified as members of the α2-adrenoceptor superfamily were aligned using MUSCLE [18]. Incomplete sequences were discarded (if the deletion spanned >30% of the alignment). BioEdit program [19] was used to manually optimize the alignments to preserve the continuity of secondary structure elements, including transmembrane helices.

**Phylogenetic analyses.** The phylogenetic tree of the α2-adrenoceptor superfamily was inferred for all members of this family. Based on the multiple sequence alignment provided by MUSCLE, MEGA 5 [20] was used to construct a minimum evolution phylogenetic tree, with pairwise gaps deletion and JTT matrices [21]. The stability of individual nodes was calculated using the bootstrap test (with 100 replicates) and additionally confirmed by the interior branch test (IBT).

**Protein structure prediction**

Human α2C-adrenoceptor and has been experimentally shown to interact with the filamin-2 (FLN2) region [15]. In the absence of experimentally determined structure that shows this interaction, we constructed comparative models of human FLN2 protein (gi number: 3914602) and human ADRA2C protein (GI number: 3914602). In the following sections, structure prediction of these two proteins are discussed in detail.

**Modeling of filamin-2 (FLN2) region.** According to the HHpred [22] program, three protein domains were found within the FLN2 fragment that had been shown to be responsible for ADRA2C binding [15]. Based on the predicted domain boundaries we redefined the filamin-2 region that binds ADRA2C, to 202 amino acid residues that are located between residue 1982 and 2183. This region was investigated by using the state-of-the-art structure prediction servers, that include GeneSilico metaserver [23], Zhang-Server [24], Robetta [25], HHpred [22], and Multicom [26] server. The initial models provided by these servers were submitted to the QA-RecombineIt server [27], which operates in two stages. In the first stage, the server predicts both global and local accuracy of models. In the second stage, the server runs an algorithm that performs a ‘recombination’ of the best ranked parts of the input models into new hybrid structures that are likely to be better than the input models themselves. By using recombination of the initial models, the QA-RecombineIt generated 100 additional consensus models. From these models, the final model was selected by using Model Quality Assessment Programs, such as MetaMQAP [28], ProQ2 [29], GOAP [30], DFIRE [31] and MQAPmulti (M Pawlowski, unpubl.).

**Modeling of α2C-adrenoceptor.** To model the structure of human α2C-adrenoceptor, its sequence was submitted to GeneSili-co metaserver [23], Zhang-Server [24], Robetta [25], HHpred [22], and Multicom [26] server. Noteworthy, in contrast to FLN2 protein, the α2C-adrenoceptor is a transmembrane protein. Hence, in addition to these aforementioned protein structure prediction servers, we used servers optimized to predict 3D structure of transmembrane proteins. Among these servers were: GPCRMM [32], GPCR-ITASSER [33] and GPCR-SSFE [34]. These servers created 145 initial models in total, which were used as input for the QA-RecombineIt server [27]. By using recombination of the initial models, the QA-RecombineIt generated 100 additional consensus models. From these models, the final model was selected by using MQAPmulti (M Pawlowski, unpubl.), and ProQ3M [35] programs. Notably, ProQ3M is the only MQAP that has been created to predict the correctness of transmembrane proteins.

**Docking between ADRA2C and FLN2 region between amino acid residues 1982 and 2183**

Docking models of the ADRA2C and FLN2 complex were generated with the HADDOCK server [36,37]. Docking by HADDOCK is driven by predictions of likely residues involved in protein-protein interface (ambiguous interaction restraints (AIRs)). Such residues may be active (interacting residue) or passive (solvent-accessible neighbor of interacting residue). AIRs for both ADRA2C and FLN2 (residues 1982 to 2183) were predicted by using CPORT [38]. Then, two hundred complexes were generated by the HADDOCK program and clustered. Selection of the best complex was based on cluster size, HADDOCK score and electrostatic energy. Among the ten best clusters, we selected a cluster that was the second most populated cluster, but was characterized by highest HADDOCK score and lowest electro-
static energy. PISA [39] was used to analyze the protein-protein docking and binding interfaces. The illustrations and visualizations of the final model were produced in PyMOL (version 1.4.1) [40].

Results

Sequence database searches and retrieval of members of the \( \alpha_2 \)-adrenoceptor family

To identify a complete set of \( \alpha_2C \)-adrenoceptor sequences, including sequences of \( \alpha_2 \)-AR, \( \alpha_2B \)-AR, and \( \alpha_2C \)-adrenoceptors, we used full-length sequences of representatives of these three types of \( \alpha_2 \)-adrenoceptors (GI numbers: 191539970, 33590960 and 3914602) from \( H. sapiens \) to carry out homology search. We removed identical proteins retrieved in different searches and finally, we obtained 72,730 proteins homologous to the \( \alpha_2 \)-adrenoceptors. Notable, all members of \( \alpha_2 \)-adrenoceptors were find during the first iteration of PSI-BLAST.

Extracting \( \alpha_2 \)-adrenoceptors from G protein-coupled receptors

Since the performed homology searches had provided not only the \( \alpha_2 \)-adrenoceptors, but also other G protein-coupled receptors (GPCRs), we performed the clustering of these all proteins to identify clusters that contain \( \alpha_2 \)-adrenoceptors only. We clustered the G protein-coupled receptors based on the pair-wise BLAST similarity scores by using the CLANS program [17]. We tried different P-value thresholds and found that the value of \( 10^{-11} \) produced best resolved sequence “clans” corresponding to different highly homologous subtypes of \( \alpha_2 \)-adrenoceptors, including \( \alpha_2 \)-A, \( \alpha_2 \)-B, and \( \alpha_2 \)-C-adrenergic (with strong connections within each clan and preferred connections between a few, but not all clans) (Figure 1, panel A). Figure 1, panel B focuses only on \( \alpha_2 \)-adrenoceptor family. Even though, fraction of \( \alpha_2 \)-adrenoceptor proteins were clustered clearly as one of the \( \alpha_2 \)-adrenoceptor subfamilies, the classification of some \( \alpha_2 \)-adrenoceptors was still unsolved.

Multiple sequence alignment and phylogenetic analysis of \( \alpha_2 \)-adrenoceptors

Based on the results of preliminary clustering, we extracted only members of \( \alpha_2 \)-adrenoceptor family. Then we calculated family-specific multiple sequence alignments using MUSCLE [18] and adjusted them manually (as described in Methods) to remove truncated sequences and redundant, nearly identical versions of the same protein, and to improve the placement of insertions and deletions. The refined multiple sequence alignment was used to infer the phylogenetic tree of the \( \alpha_2 \)-adrenoceptor family by using Minimum Evolution (ME) analysis carried out with MEGA 5 [20]. To calculate the stability of individual nodes, the bootstrap test and the interior branch test were applied. Noteworthy, for this phylogenetic tree, for all branches with bootstrap support \( >50\% \), the ITB support was equal or higher 50.

The Minimum Evolution tree (Figure 1, panel C) provides significant support for main branches, allowing us to resolve the deep branching pattern. This compressed tree indicates the division of \( \alpha_2 \)-adrenoceptors family into 4 subfamilies. This approach clearly showed that the \( \alpha_2C \)-ARs are relatively distinct and form a separate branch, while the \( \alpha_2A \)-ARs and \( \alpha_2B \)-ARs are the most closely related to one another. These mutual orientations of the \( \alpha_2A \)-AR, \( \alpha_2B \)-ARs and \( \alpha_2C \)-ARs are in agreement with the previously published phylogenetic analysis of these subtypes [41]. The tree also supports the findings of Ruuskanen et al. that there is another subtype of \( \alpha_2 \)-adrenoceptors, named \( \alpha_2D \)-adrenoceptors, and originally identified in Zebrafish by comparison of ligand binding characteristics of \( \alpha_2 \)-adrenoceptors, but not by phylogenetic analysis [42]. The \( \alpha_2D \)-adrenoceptors form the separate branch showing that this family is relatively distinct from other subfamilies. Noteworthy, since the rodent orthologue of the human \( \alpha_2A \) is occasionally misleadingly called \( \alpha_2D \), we want to emphasize that in our work we follow the naming proposed by Ruuskanen et al.

Detailed analysis of the evolution of the C-terminus of \( \alpha_2C \)-adrenoceptors is presented in Figure 2, panel A. The \( \alpha_2C \)-adrenoceptors appears in nearly all sequenced Vertebrata. The \( \alpha_2C \)-adrenoceptors are divided into 5 subfamilies containing members found in: 1) Mammals, excluding Marsupials, 2) only Marsupials, 3) Birds, 4) Amphibians, 5) Reptile, and 6) Fish. Interestingly, the C-terminus of \( \alpha_2C \)-adrenoceptors is either Arginine- or Lysine-rich only in Mammals and Birds. This finding may be connected with the fact that these warm-blooded animals need systems to control temperature of the most peripheral parts of their bodies. We postulate that the \( \alpha_2C \)-adrenoceptor may be involved in the process, in which the receptor’s highly positively charged C-terminal helix may be responsible for the receptor translocation.

To investigate this hypothesis, we decided to build a computational model of the complex of \( \alpha_2C \)-adrenoceptor and filamin-2, which is presented in the following paragraphs.

Computational simulation of \( \alpha_2C \)-AR-filamin-2 binding domains

We performed computational modeling predictions of full-length \( \alpha_2C \)-AR and filamin-2 (amino acids 1979-2206) structure to better understand the specificity of \( \alpha_2C \)-AR-filamin-2 protein-protein interactions. In the absence of a crystal structure for \( \alpha_2C \)-AR and filamin-2 region, we utilized amino acid homology searches, domain predictions, followed by protein-protein docking, to identify the residues that play a key role in \( \alpha_2C \)-AR-filamin-2 recognition and binding as described below.

Modeling of filamin-2. In the absence of experimentally determined structure for functionally characterized human filamin-2, we constructed a comparative model of a human filamin-2 region (amino acids 1979-2206) found to bind \( \alpha_2C \)-adrenoceptor. First, to perform initial sequence analysis the sequence of FLN2 (amino acids 1979-2206) was submitted to GeneSilico metaserver [23]. This analysis revealed that this region is composed of three domains (roughly residues 1982-2100, 2101-2178 and 2179-2183). Both the N-terminal and C-terminal domains of FLN2 were found to exhibit significant similarity to Filamin/ABP280 repeat family, whose members have been found to interact with such proteins like: \( \beta \)-Integrin, Rho, Rho-associated kinase (ROCK), and many others [43]. In contrast to the N-terminal and C-terminal domains of FLN2, the domain in the middle (2101-2178 residues) exhibited no evident similarity to any known protein family. Nearly all individual fold-recognition methods (e.g., HHSEARCH: score 100, FFAS score -50.1, COMPASS score: 2.72e-59, PHYRE score: 1e-19) reported the structure of the protein with PDB code 2j3s [crystal structure of filamin A Ig domains 19-21] as the potentially best template to model FLN2 region 1982-2183 (i.e., its closest homologs among proteins of known structure); the sequences of 2j3s and the target proteins share 54% sequence identity. In the next step, the sequence of FLN2 (amino acids 1979-2206) was submitted also to Zhang-Server [24], Robetta Server [25], HHpred [22], and Multicom [26] server; these servers have been shown to be the best automatic methods for proteins structure modeling [44]. In total we collected 145 initial models, which were submitted to the QA-
RecombineIT [27] server that operates through following two stages. In the first stage (QA-mode), the server predicts both the global quality of input models and provides estimates of local quality as the deviation between C-\(\alpha\) atoms in the models and corresponding atoms in the unknown native structure. The input models and the predictions of the models’ correctness become the input for the second stage (RecombineIt-mode), in which fragments predicted to be better than others are judiciously combined to generate hybrid (consensus) models. Finally, hybrid models are scored by the MQAPs implemented in the QA-mode and then presented to the user. By using recombination of the initial models, the QA-RecombineIt generated 100 additional consensus models for the filamin-2 region between residue 1982 and 2183. From these models, the final model was selected by using Model Quality Assessment Programs, such as MetaMQAP [28], ProQ2 [29], and MQAPmulti (M Pawlowski, unpubl.). These methods predict GDT_TS-score of a protein model without the knowledge about the true native structure of the protein. Global Distance Test (GDT_TS) corresponds to the average value of fractions of C-\(\alpha\) atoms in the model that are placed within the distances of 1, 2, 4 or 8 Å from corresponding C-\(\alpha\) atoms in the experimentally determined structure. This metric has values in the [0,1] range, where 1 corresponds to the highest quality model. In contrast to RMSD (root-mean-square-deviation) score, GDT_TS-score is insensitive to local structure variation. In general, two structures with GDT_TS-score lower than 0.3 correspond to random similarity and those with GDT_TS-score at least 0.5 indicate high similarity between the predicted model and native structure. Model Quality Assessment Programs, may be divided into two main classes: 1) single-model MQAPs - methods capable of assessing quality for single models, without using any alternative models (decoys) generated for the same protein; 2) clustering MQAPs – methods that perform all against-all structural comparisons to obtain mean similarity scores for ranking models. Moreover, it was shown that a linear combination of scores provided by clustering and single model MQAP perform well for selection of the most accurate model from a set of alternative models for the target protein [29]. Thus, to select the final model of filamin-2 region (amino acids 1979–2206) we applied a linear combination of MQAPmulti (a clustering MQAP, weight: 0.8) and two single model MQAPs MetaMQAP, and ProQ2 (weight: 0.1 each), then the model with the highest score was selected as the final model. The selection procedure was inspired by the findings that a linear combination of scores provided by clustering and

Figure 1. Initial clustering of GPCR proteins and phylogenetic tree of all \(\alpha_2\)-adrenoceptors. Panel A presents two-dimensional projection of the CLANS clustering results obtained for the GPCR proteins, a clan corresponding to \(\alpha_2\)-adrenoceptors is indicated by an ellipse. Panel B presents two-dimensional projection of the CLANS clustering of \(\alpha_2\)-adrenoceptors. Panel C presents the postulated phylogenetic tree of the \(\alpha_2\)-adrenoceptor family. Only the major branches corresponding to subfamilies are shown. Values at the nodes indicate the statistical support for the particular branches, according to the bootstrap test. The human rhodopsin sequence was used for rooting the tree.

doi:10.1371/journal.pone.0103099.g001
Figure 2. The Minimum evolution tree and multiple sequence alignment of C-terminal tail of the a2C-adrenoceptor family. Panel A - proteins are indicated by the species name and the NCBI GI number. Values at the nodes indicate the statistical support for the particular branches, according to the bootstrap test. For each protein also its C-terminal sequence is presented. Sequences were aligned by MUSCLE program. Amino acids are colored according to the chemical properties of their side-chains (negatively charged: red, positively charged: blue, polar: magenta, hydrophobic: green). Only the alignment that corresponds to the C-terminal helix and flanking residues is shown. The helix was predicted by GeneSilico metaserver. Panel B - the last 14 amino acids of a2C-AR C-terminus highlighting the arginine-rich stretch (underlined). This region is conserved in mammals and in human arteriole-derived vascular smooth muscle cells (microVSM) interacts with the actin-binding protein filamin-2, shown in experimental studies to be necessary for receptor translocation to the cell surface. The numbers denote amino acids in the full-length a2C-AR polypeptide. The arrows point to amino acid residues identified by in-silico modeling to be involved in interaction with filamin-2.

doi:10.1371/journal.pone.0103099.g002

Figure 3. Predicted models of filamin-2 (FLN2) and a2C-adrenoceptor (ADRA2C) proteins, and their complex. Panels A and B present cartoon diagram of FLN2 (region between residues 1982 and 2183) and ADRA2C protein models. Positively and negatively charged regions are indicated by blue and red colors, respectively. Panel C presents whole protein-protein complex predicted by HADDOCK program. Panel D shows the interaction between receptor’s C-terminal helix and the filamin-2 region that is responsible for binding the receptor.

doi:10.1371/journal.pone.0103099.g003
clustered. The resulting clusters were analyzed and ranked according to the HADDOCK score which consists of a weighted sum of energies that include intermolecular electrostatic, van der Waals, desolvation and AIR (ambiguous interaction restraints) and a buried surface area term. HADDOCK clustered 146 structures in 10 clusters, which represents 73.0% of the water-refined models HADDOCK generated. The largest cluster had 41 structures, the 5th best HADDOCK score (−91.2) and the 6th best (lowest) electrostatic energy (−508.8) among all 10 clusters. However, the protein-protein interfaces among those structures did not involve any interactions between the C-terminal helix of ADRA2C and FLN2 between residues 1982 and 2183, as previously proven to occur by Motawea and coworkers [15].

Thus, among the ten most populated clusters, we searched for clusters that had receptor-filamin complexes having C-terminal helix of the receptor molecule involved in interactions with the filamin molecule. Cluster number 7 was the only one that satisfied this criterion. This cluster was the 7th most populated one (8 members), had the 4th highest HADDOCK score (−126.8), but was characterized by the best electrostatic energy among all clusters (−938.1). The medoid of this cluster was selected as the final solution of protein-protein docking.

Protein-protein interface between ADRA2C and FLN2 region between amino acid residues 1982 and 2183. Figure 3 panel D presents the protein-protein interface most likely to be involved in the recognition and binding of $\alpha_{2C}$-adrenoceptor by human filamin-2. The interface area, measured by PISA server [39], occupied 1277.6 Å². Three arginines (R454, R456 and R461) are stabilized by negatively charged residues in the filamin-2 structure: E2004, E2059 and D2060, respectively. Another interaction involved in the complex stabilization is lysine K449 that is stabilized by aspartic acid at position 2032 (D2032) in filamin-2 sequence.

Multiple sequence alignments of ADRA2C (presented in Figure 2, panel A) and FLN2 (region between amino acid residues 1982 and 2183, no data shown) reveals that the residues found to be involved in the complex stabilization are conserved between the homologs of human ADRA2C and FLN2, which is typical for protein-protein interaction sites [46]. Noteworthy, in the case of ADRA2C the conservation is observed only for Mammals and Birds, that is in contrast to that observed for FLN2, where the conservation is observed for all members of this family. Taken together, these findings suggest that these two genes have not coevolved, but the genes of ADRA2C animals have evolved in order to interact with the filamin-2 in warm-blooded animals.

Discussion

Recently it was shown that the C-terminal helix of human $\alpha_{2C}$-adrenoceptor binds to filamin-2 region between residues 1979 and 2206 [15]. To study this interaction in the context of evolution, we have carried out extensive bioinformatics analyses and proposed a structural model of this complex. The approach used in the present study complements and supports the experimental approach described in the previous study [15]. The results of multiple sequence alignment of $\alpha_{2C}$-adrenoceptor family combined with the phylogenetic analysis showed that among all animals studied here, only the warm-blooded ones have $\alpha_{2C}$-adrenoceptor C-termini that are either Arginine- or Lysine-rich. We postulate that this highly positively charged helix is involved in the binding of the $\alpha_{2C}$-adrenoceptor to the filamin-2, in which satisfying the electrostatic energy is the driving force. The last 14 amino acids, unique to the $\alpha_{2C}$-AR subtype (Figure 2, panel B), play a critical role in protein-protein interaction with filamin-2.

Such binding, as shown by Motawea et al., 2013 [15] is responsible for translocation of functional receptors to the cell surface. Motawea et al. 2013 also performed site-directed mutagenesis of the arginine-rich region (R454 to R458) and replaced all five arginines with non-polar alamines (A454 to A458). The receptor with these alamines was not able to interact with filamin-2. This finding, together with the fact that the $\alpha_{2C}$-adrenoceptors with a non-positively charged C-terminal helices occur only in cold-blooded animals, supports our postulate that the positive charge is critical for the binding and translocation. The molecular docking reveals the interactions involved in the creation of the protein-protein interface between these two proteins, particularly R454 and R456 in stabilizing this interaction. We found that there are four interactions that stabilize the positive charge of the C-terminal helix, including three arginines (R454, R456 and R461) that are stabilized by negatively charged residues in the filamin-2 structure: E2004, E2059 and D2060, respectively. Another interaction involves lysine K449 (K449) that is stabilized by aspartic acid at position 2032 (D2032) in filamin-2. We postulate also, that the arginines numbered as R455, R457 and R458 are also important for the creation of the protein-protein interface, although they were not shown by the protein-protein docking study (see Fig. 3) to create any important interactions within the protein-protein interface. However, they can act as O-ring residues [47] whose role is to occlude bulk water molecules from the hot spots. Exclusion of water from the binding interface is thought to be entropically favorable. In addition, removing of solvent dipoles lowers the local dielectric constant for the hotspot, increasing the energetic contribution of electrostatic interactions [47].

Indeed, experimental studies performed by Motawea et al show that the receptor having arginines (R454–R458) replaced with alanines (A454–A458) does not associate with filamin-2 [15]. Experimental studies also suggest the role of the arginine-rich region (R454–R458) in retaining mature receptors in the Golgi compartment. In transiently transfected HEK293 cells the mature glycosylated receptor (the ~70 kDa form that has passed through the ER, cis/medial Golgi and is endoglycosidase H resistant) is retained in the transGolgi, and translocates to the cell surface in response to stimulus including cold temperature [10]. The receptor having arginines replaced with alanines however, is no longer retained and is localized on the cell surface [15]. The studies therefore suggest that $\alpha_{2C}$-AR interaction with filamin-2 enables stimulus-dependent regulated cell surface delivery and function compared with constitutive presence on the cell surface.

It remains to be determined why the C-terminal helix is arginine-rich in Mammals (not including Marsupials) and lysine-rich in the rest of warm-blooded animals. As shown in figure 2, panel A, the C-terminal helices of the $\alpha_{2C}$-ARs in Fish are both lysine- and arginine-rich. It may suggest that in the common ancestor of all warm-blooded animals the $\alpha_{2C}$-AR could have had both arginine and lysine rich C-terminal helix, and during the species speciation the lysine-rich variant has been kept among Birds and Marsupials, in contrast to the arginine-rich variant that has been kept among the rest of Mammals. Taking this hypothesis into account, it would be interesting to see what will happen if the human $\alpha_{2C}$-AR has its C-terminal helix replaced by the Birds/ Marsupials lysine-rich variant. Could it function the same way as the wild-type variant of the receptor in skin thermoregulation in humans? Future experimental studies will allow examination of this hypothesis.

It has been shown that $\alpha_{2C}$-ARs are intracellular receptors that are translocated to the cell surface in response to cellular stress including cold temperatures and play a vital role in skin
thermoregulation [8,9]. However, it is estimated that in 5–10% of the general US population, this system is overregulated and leads to Raynaud’s phenomenon, an exaggerated vasospastic response to cold or to emotional stress [48–50]. One of the possible attempts to find therapeutics for Raynaud’s would be to develop small molecules that are antagonists of human α2C-ARs [51]. However, since it has been shown that α2C-ARs are responsible for, among others, sympathetic neurotransmission - [52], the usage of such an antagonist would be likely to be associated with many serious side-effects in humans. Notably, in the present study we showed the possible interface between the α2C-AR and filamin-2. We believe that this finding may contribute to the development of new therapeutics for Raynaud’s phenomenon that target the protein-protein interface between those two proteins, selectively inhibiting cell surface translocation of intracellular receptors. Our optimism is based on the fact that targeting protein-protein interface has been very successful in drug design, for example in identifying inhibitors of the Bcl-2 protein [53] or inhibitors of the binding of S100B, a calcium binding protein, and p53 [54]. We believe that in the case of Raynaud’s syndrome such therapeutics can target the protein-protein interface between the filamin-2 and intracellular α2C-AR, but sparse surface receptors expressed in other tissues, reducing side-effects. It is also interesting if targeting the drug design towards the protein-protein interface instead of α2C-ARs itself would help to avoid some issues associated with recent GPCR drug discovery. One of these issues arises from the observation that many of possible small molecules that target GPCRs, but not the protein-protein interface in which GPCRs are involved, are generally at the upper limits of Lipinski’s rules in terms of molecular weight and/or lipophilicity [55,56]. This suggests that they would have been “high risk” in terms of both toxicity and cross-reactivity giving a low success rate in the clinic [57,58]. Thus, we hope that drugs interfering with the protein-protein interface of α2C-ARs and filamin-2 will be less prone to these negative side effects.

Conclusions

Here, we showed extensive bioinformatics analyses aimed to study the binding of α2C-AR to filamin-2 region between residues 1979 and 2206, which has lead us to the following findings and conclusions. First, by protein-protein docking, we characterized the protein-protein interface, in which the C-terminal helix of α2C-AR is involved in the complex creation. Second, the electrostatic interactions seem to play a key role in this complex formation which manifests in interactions between the C-terminal arginines of α2C-ARs (particularly R1454 and R1456) and negatively charged residues from filamin-2 region between residues 1979 and 2206. Finally, multiple sequence alignments and phylogenetic analysis showed that these interactions are conserved in warm-blooded animals.

According to the 3did database [59], a catalog of domain-based interactions of known three-dimensional structure, there is no crystal structure where the C-terminal helix of a GPCR protein was involved in protein-protein interface. Thus, we believe that this model of the α2C-AR-filamin-2 complex will help in the further investigation of the mechanism of the GPCR protein translocation to any cell compartment, including the α2C-AR translocation to the cell surface in the context of cellular pathology and pathophysiology.

Author Contributions

Conceived and designed the experiments: MP MAC AK. Performed the experiments: MP SS. Analyzed the data: MP MAC AK. Contributed to the writing of the manuscript: MP SS HRBM MAC AK.

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