**REVIEW ARTICLE**

The Human Immune System toward *Staphylococcus aureus*

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**Abstract**: The immune system is responsible for protecting the host from pathogens, and it has evolved to deal with these pathogens. On the other hand, the co-evolution of pathogenic bacteria with hosts has led to the rise of an array of virulence genes that enable pathogen bacteria to evade or modulate the immune system. *Staphylococcus aureus* is a significant pathogen of humans that encodes several virulence factors that can modulate or evade from the innate and adaptive arm of the immune system. Overall, the immune reaction toward *S. aureus* contributes to stimulate innate and adaptive reactions. A profound understanding of the immune response to *S. aureus* infections will be critical for the development of vaccines and novel therapies. In this review, we summarized and discussed the novel information about the human immune system against *S. aureus*.

**Keywords**: *Staphylococcus aureus*, Innate immunity, Adaptive immunity, Pathogen, Immune system, Methicillin-resistant *S. aureus*.

**1. INTRODUCTION**

*Staphylococcus aureus*, a Gram-positive anaerobic bacterium, is the most important medically species in the genus of *Staphylococcus* [1]. This organism develops a wide variety of infections ranging from simple skin infections (like corks, scabies, boils, and abscess) to life-threatening diseases (like endocarditis, pneumonia, osteomyelitis, meningitis, septicemia, toxic shock syndrome) [2]. Methicillin-resistant *S. aureus* (MRSA) that is resistant to most antibiotics is considered a threat to public health [3]. Despite their obvious pathogenic potential, *S. aureus* strains have the potential to co-exist with their host as a commensal, with approximately 30% of the human population colonized at the mucocutaneous section [4, 5]. The human immune system detects pathogens and destroys them [6, 7]. In this regard, the immune system includes innate and adaptive sections [6, 7]. The innate or nonspecific immune system or natural immunity is the first level of protection and consists of cells that commonly find and destroy pathogens in the body [8]. As the primary class of defense in bacterial infections, the innate immune reactions are rapidly induced by pattern recognition routes that recognize non-specific markers of bacterial infection [9]. A major consequence is the stimulation of phagocytic immune cells like neutrophils and macrophages [9]. The innate immune cells can be induced via Toll-like Receptors that enhance antibacterial reactions, stimulate inflammation, and trigger effector cells. This process finally results in the production of interferons, pro-inflammatory as well as cytokines. Pro-inflammatory cytokines are produced by resident host cells that cause the fever [10].

Acquired (adaptive) host immunity is the second type of immunity that is much more evolved than innate immunity and is induced with each encounter with infectious agents and with each repeated exposure to a particular pathogen that increases its defense capacity and capabilities [11]. An important feature of this type of immunity is that it has a specific response for each external factor and that it is also a faster and stronger response [11]. Via activation of T cells and antibodies formation by B cells, the adaptive host immune reaction targets particular microbial antigens and could be recalled in subsequent bacterial infections to offer memory toward that specific pathogen [11]. In this review, we summarized and discussed the novel information about the human immune system against *S. aureus*.

**2. INNATE IMMUNE RESPONSES TOWARD *S. AUREUS***

The skin is the largest organ in the body and the main physical barrier between the organism and its environment [12]. It also plays an active role in the defense of the host and, therefore, plays a role in the development and maintenance of local immune and inflammatory responses [12]. Many immune
responses originate from this tissue as the gateway to many alien antigens [12]. The skin contains important cells like melanocytes, keratinocytes, and Langerhans and T lymphocytes. Langerhans is immature Dendritic Cells (DCs) of the immune system and its function is to trap the antigens that enter the host skin [13]. The epidermis layer of skin is organized by keratinocyte cells in various maturation levels, T lymphocytes as well as Langerhans cells. The dermis layer comprises extracellular matrix ingredients, like connective tissues, elastin fibers, and collagen [14]. The fibers offer a building skeleton to human blood vessels, fibroblasts, adipocytes, macrophage cells, DCs, mast cells, T cells, B cells, and plasma as well as Natural Killer (NK) cells [14]. As such, inhabitant human immune cells are plentiful in the skin, and these cells are all contributed to the control of infection of S. aureus strains.

Keratinocyte cells are the central cells of the epidermis and have a protective function against external factors such as bacteria [15]. Keratinocyte cells are usually the first cells that encounter microbial pathogens, and detect Pathogen-Associated Molecular Patterns (PAMPs) through various Pattern-Recognition Receptors (PRRs), like nucleotide-binding oligomerization domain-1 (NOD-1), nucleotide-binding oligomerization domain -2 (NOD -2), TLRs as well as the scavenger receptors CD36 [15 - 17]. Signaling via these human receptors stimulates transcription factors like AP1, NF-kB as well as CREB to produce chemokines, cytokines, and antimicrobial effectors, like Antimicrobial Peptides (AMPs) and inducible Nitric Oxide synthase [16, 18, 19]. TLR-1, TLR-2, and TLR-6 detect the cell wall components S. aureus, particularly peptidoglycan and lipopeptides [20, 21]. These TLRs apply the signaling adapter MyD88 to stimulate efficient and robust transcriptional programs that trigger inflammatory reactions [22]. TLR-1, TLR-2, and TLR-6, contribute in different stages of infection caused by S. aureus. Primarily, TLR-2 on Keratinocyte cell detect bacterial pathogens to release neutrophil antimicrobial peptides and chemo-attractants, like the defensins and cathelicidin LL-37, which produce pores in the membrane of bacteria [19]. TLR-2 molecules are formed on inhabitant recruited neutrophils, monocytes, and macrophages, which immediately react to S. aureus and further activate cytokine formation and phagocytosis [23]. It is supposed that TLR2 molecules are essential for localized and systemic S. aureus infection. Mouse deficient in TLR-1, TLR-2, TLR-6, and MyD88 is very susceptible to intranasal infection of S. aureus, as documented by the raised load of bacteria, poor inflammatory reactions, and increased morbidity and mortality in different models of disease [24]. In human skin, the TLR2 activity is controversial due to variation in virulence of microbial strains, measured endpoints as well as infectious dose (Fig. 1).

Neutrophils are a type of phagocytic white cells that have chemotaxis and are in the second line of nonspecific defense in the human immune system [25]. The neutrophil is a crucial section of the acute reaction and centrally relevant toward S. aureus [26]. The NOD-1, NOD-2 and intracellular PRRs that recognized microbial peptidoglycan stimulate antimicrobial peptide formation, inflammation, and phagocytic effector activities [27]. NOD-2 molecules detect muramyl-dipeptide from the peptidoglycan of S. aureus [28, 29]. The NOD2-deficient mouse is very susceptible to systemic and skin infections of S. aureus when compared with wild type counterparts [29, 30]. Eventually, scavenger receptors SRBII, CD36, as well as MARCO are required for optimal skin host defense against S. aureus [31, 32]. As a result, CD36 2/2 mouse shows raised bacterial loads and enhances severe dermonecrosis induced by α-toxin [31].

Macrophages have spread throughout the body [33]. Immature macrophages in the bloodstream are called monocytes [33]. Skin-inhabitant macrophage cells assist in the first S. aureus clearance, and in conjunction with perivascular macrophages. Moreover, they control the recruitment of monocytes and neutrophils to the location of infection [34]. Dermal macrophage cells could phagocytose and kill S. aureus efficiently by forming antimicrobial peptides, reactive nitrogen, and oxygen species as well as chelating proteins that starve microbes of essential micronutrients [35, 36]. Additionally, dermal macrophage cells release various chemo-attractant molecules that offer signals for the recruitment of neutrophil cells in a manner mediated to MyD88, IL-1-R [37]. Moreover, these cells contribute to the dead cell-clearance at the site of bacterial infection, which is vital for the resolution of the disorder [35, 36]. When the neutrophil cell reaches the location of the infection, it engulf the S. aureus and attempts to control bacterial growth by forming various antimicrobial effector molecules [35]. The neutrophil is a short-lived immune cell that quickly triggers apoptosis and requires to be eradicated from the location of the infection. Nevertheless, S. aureus cells form different toxins, like Panton-Valentine leukocidin, phenol soluble modulins, α-toxin as well as γ-hemolysin, and that could hasten neutrophil death by stimulating necrosis and cause to liberate the danger-mediated molecular patterns [35, 38]. Danger-mediated molecular patterns freed in S. aureus infection like IL-1α, IL-33, HGBM1, ATP, as well as calprotectin [39]. DCs and Langerhans cells capture the microbial antigens before contact with skin-draining lymph nodes [40, 41]. The study has found that in skin S. aureus infection, Langerhans cells that engulf the bacteria are stimulated by PAMPs, and subsequently migrate to draining lymph nodes where Langerhans cells elicit adaptive reactions specific to S. aureus [42, 43]. Although there are various distinct sub-kinds of dendritic cells in the human skin, its activity against S. aureus infection in the skin is not well-defined [44].

S. aureus cells can be engulfed by receptors that detect non-opsonized and opsonized bacteria [35, 38, 45]. S. aureus cells trigger different antimicrobial effector activities, when coated with opsonins like C3b and IgG [44]. Reactive oxygen species are produced via the actions of myeloperoxidase and NADPH oxidase following phagocytosis, which can directly kill microbes [46]. Nitric oxide molecules are a main reactive nitrogen species that formed from nitric oxide synthase with immunomodulatory and antimicrobial activity [46]. Pharmacologic inhibition and genetic deletion of nitric oxide production render mice susceptible to S. aureus infection [47].
Nevertheless, high levels of nitric oxide could exert anti-inflammatory impacts. High concentrations of nitric oxide can, therefore, predispose to bacterial infection via inhibiting the proliferation of the cell, stimulating host cell death, and preventing TNF-α formation induced by phagocyte and presentation of Ags. Additionally, *S. aureus* cells use nitric oxide to proliferate and preclude stimulation of the stress regulon through the fermentation of lactic acid [48, 49].

Neutrophils kill pathogens by degranulation of toxic components [50]. Degranulation stimulates the formation of particular granules containing antimicrobial peptides, like LL37, defensins, and cathelicidin-related antimicrobial peptide. Moreover, degranulation diffuses cathepsin, azurocidin, lysozymes, lactoferrin, elastase, and proteinase-3 [51, 52]. Neutrophils liberate structures of DNA named NET (neutrophil extracellular trap), as an additional effector mechanism to control *S. aureus* infection, that formed in a mechanism dependent on a TLR2 and MyD88 and is necessary for containing *S. aureus* in the host skin to prevent bacteremia [53]. NET finites the spread of bacteria because it is rich in antimicrobial agents like cathepsin, antimicrobial peptides, histones, elastase, as well as proteases [54]. Nevertheless, *S. aureus* cells can destroy NET, and the degradation product 2’-deoxyadenosine that stimulates apoptosis in macrophage cells, which raise bacterial survival in the abscess [55].

Some AMPs exist in the human skin with activity toward *S. aureus* cells, for example, Alpha-defensins (named HNPs (human neutrophil peptides)), dermcidin, Beta-defensins, cathelicidin, as well as RNase7 [56, 57]. Neutrophil releases high rates of HNPs, which included nearly half of the peptides inside granules of neutrophils [58]. Amongst HNPs, HNP2 peptides have the highest degree of antibacterial role toward *S. aureus* cells [59]. In humans, there are four human β-defensins (hBDs), including hBD1, hBD2, hBD3, hBD4, which are formed by keratinocytes, macrophages, as well as DCs [60, 61]. Amongst hBDs, hBD3 has a potent activity toward *S. aureus* [62]. Cathelicidin is also called LL-37 as it has 37 amino acid released from the cationic antimicrobial protein 18 kDa [63] similar to hBD3, which has strong activity against *S. aureus* [64]. This AMP is formed in neutrophil cells and could be stimulated in keratinocyte cells [64]. Another AMP, RNase 7, is formed by human keratinocyte cells and has activity toward *S. aureus* [65]. A current work discovered that high levels of RNase7 could stop *S. aureus* skin colonization in the host stratum corneum [66]. Additionally, dermcidin is formed by eccrine sweat glands with activity toward *S. aureus* [67]. The keratinocyte cells form the LL-37, hBD2 as well as hBD3 in *S. aureus* infections for prompt clearance of the bacterial cells [68]. In addition, stimulation of the EGFR (epidermal growth factor receptor) by wounding of skin cause in enhanced hBD3 formation offers another strategy for increasing antibacterial role toward *S. aureus* [69].

Four PGRPs (peptidoglycan recognition proteins) exist in...
humans, including PGRP1-4 that secrete proteins [70]. PGRP1 molecules are formed in neutrophil cells, where they bind peptidoglycan S. aureus inside tertiary granules and apply their bactericidal activity [71]. PGRP2 molecules are formed by keratinocyte cells and are distinct that they have the active amidases that cleave the peptidoglycan of S. aureus [72, 73]. Finally, PGRP3 and PGRP4 produce in the hair follicles, epidermis, sebaceous glands, as well as sweat glands and their relevance to S. aureus infection is unknown [74]. Complement activation, as a crucial section of the innate immune defense, triggers cytolytic reactions and acute inflammatory responses. Furthermore, it involved in the regulation of host adaptive immune defenses [75, 76]. The complement is stimulated through particular recognition routes, including the classical and the alternative and the lectin pathway [75]. All pathways form the active protease complexes, the C3 convertase, on the bacterial surface [77]. C3 convertase activity leads the labeling of microbial antigens with a high amount of C3b, as well as iC3b [75]. This process is named opsonization that is crucial for impressive phagocytic uptake through complement receptors by phagocytic cells [78]. It must be noted that a highly particular factor called staphylococcal complement inhibitor produced by S. aureus cells attaches to and stops the role of the C3 convertases on the staphylococcus surface [79].

3. ADAPTIVE IMMUNITY AGAINST S. AUREUS

It is unclear whether the limited association of B cells and S. aureus affects the pathogenesis and/or host immune response, but, it has been shown that the binding of CR2 expressed by B lymphocyte with C3d on the antigen surface has several roles useful for the antibody action [80]. For cognitive B cells, the simultaneous binding of CR2 with C3b with the B lymphocyte antigen receptor complex reduces the B cell activation threshold by several orders of magnitude. It is required for proper B cell activation [80]. On the other hand, non-cognitive B cells in the lymph nodes and spleen of mice play an important role in the transfer of immune complexes containing Immunoglobulin M to follicle DCs [80]. This process is dependent on the receptor and complement that is essential for the efficient formation of the germination center and maturation of the antibody affinity [80]. Unrecognized CR2 B cells attached to antigen-containing immune complexes can also deliver these immune complexes to antigen-specific T cells to enable appropriate T-cell activation [80]. Thus, molecules secreted by S. aureus that inhibit the association of C3d-containing immune complexes with CR2 can influence the host’s adaptive immune response against this bacterium [80].

B lymphocytes are immune cells that produce antibodies against pathogens, such as viruses [81]. These lymphocytes are involved in humoral immunity, which, together with cellular immunity, form a specific defense mechanism [81]. B lymphocytes act in a specific way; that is, they identify and eliminate a particular type of alien agent [81]. These cells have a major role in humoral immunity [82]. The primary function of antibody-deficient lymphocytes against antigens is to function as Antigen-Presenting Cells (APCs), and eventually to evolve into memory cells after activation by antigenic interactions [82]. Memory cells produce more antibodies in a shorter time after re-encountering the same antigen [83]. The main activity of B lymphocytes is to form immunoglobulins that neutralize the action of target proteins (like toxin) or opsonize microbial pathogens to optimize clearance and phagocytosis [83]. The significance of antibody-mediated protection toward microbial infectious agents is found by demonstrating this event that the absence of B lymphocyte maturation causes susceptibility against infections with encapsulated bacteria and viruses [84]. The notable absence of raised susceptibility in this regard to S. aureus argues that antibody is un-significant in protection toward infection of S. aureus. Nevertheless, it has found that early cutaneous S. aureus infection could stimulate antibody-mediated support toward a subsequent microbial infection in certain mouse strains, and many pre-clinical works have found at least partial support from subsequent microbial infection subsequent stimulation of antibodies by vaccination [85].

In the opsonophagocytosis, T lymphocyte has a dual function: (1) it is important for the production of opposing antibodies because T cell is essential for maturation of antibody affinity as well as class switches, (2) T cells promote phagocytosis by absorbing neutrophils and macrophages from the bone marrow to the site of infection [86, 87]. As long as the S. aureus resides in the macrophage phagosome, its elimination by T-cell cytokines is more pronounced by IFN-γ [87]. The infected cell must now be lysed by Cytotoxic Cells (CT) cells and/or NK cells [88]. Finally, T cells are essential for the (re) establishment of immune homeostasis by blocking inflammatory processes [88]. Treg regulatory (T-reg) cells specialize in this critical immune function [88]. T lymphocytes proliferate upon binding to the antigen, producing a variety of T cells, including some Treg and some T memory [89, 90]. Lethal T cells directly invade virus-infected cells and cancer cells and kill them [90]. That is why this type of immune response to cellular immunity is known [90]. T lymphocyte is thymic-derived cells that form particular TCR receptors that detect peptides derived from antigen in terms of MHC molecules on antigen APC cells. Similar to antibody and B lymphocytes, a case could be formed for an activity for T lymphocytes in S. aureus infection according to the presence of detectable T lymphocytes reactions in humans and the capacity of the bacteria to modulate T lymphocytes as exemplified by their formation of a multitude of T lymphocyte super-antigens [91, 92]. Nevertheless, it has been found that T lymphocyte is not essential for protection toward S. aureus in mice [93]. Different sets of T lymphocyte have differing activities, and further nuanced activities for these sets have become a document in mouse evaluations and with the detected susceptibility to various staphylococcal infections of individuals with Human Immunodeficiency Viruses (HIV) disorder [94]. Most of T lymphocytes like CD4 (positive) and CD8 (positive) T lymphocytes have long been detected as a main cellular weapon of adaptive immunity of host [95]. The main activity of CD8 (positive) T lymphocytes is to kill intracellular microbial pathogens by the cytolytic killing of the infected human cell [96]. Consistent with S. aureus cells being an early extracellular microbial pathogen, the clear activities for CD8 (positive) T lymphocytes have not been found, although activation of CD8 positive T lymphocytes could be found in staphylococcal super-antigen exposure and S. aureus.
infection. Naive CD4 positive T lymphocytes are polarized against different effector activities mediated to the cytokine milieu in which stimulation of their TCR receptor occurs. These Th lymphocytes are functionally determined by their cytokine formation profiles. A rate of these polarized lymphocytes will exist in the human as memory lymphocytes awaiting re-stimulation by next exposure to antigen. More recently, in addition to CD4 positive and CD8 positive T lymphocytes, a set of T lymphocytes has been described, such as γδ T lymphocytes, NK cells, and innate lymphoid cells, that involve mostly in the innate immune reactions at mucosal locations instead of memory specific to the antigen, although current works have proposed the potential for γδ T lymphocytes to involve in a memory reaction under some conditions [97].

CONCLUSION

In summary, host immunity in S. aureus infections is tissue-specific and it significantly is mediated to PRRs, like NOD-2, TLR-2, and for particular cytokine signaling mechanisms, like IL-1. Nevertheless, adaptive host immunity (B and T lymphocyte reactions) can impact this defense. Overall, the immune reaction toward S. aureus contributes to the stimulation of both innate and adaptive reactions. Adaptive immunity of the host may impact susceptibility to infections of S. aureus and its ability has specific significance in determining the results of chronic persistent human infections. Finally, a profound understanding of the human immune response to S. aureus infections will be critical for the development of vaccines and novel therapies.

AUTHOR'S CONTRIBUTIONS

R.M, S.K, R.G, S.G prepared the manuscript, and R. R edited the manuscript and H.H designed the figure.

CONSENT FOR PUBLICATION

Not applicable.

FUNDING

None declared.

CONFLICT OF INTEREST

The authors declare no conflict of interest, financial or otherwise.

ACKNOWLEDGEMENTS

Thanks to guidance and advice from the “Clinical Research Development Unit of Bagiyatullah Hospital”. The authors also thank the Department of Microbiology of Hamadan, Hamadan University of Medical Sciences, Iran.

REFERENCES

[1] Taylor TA, Unakal CG. Staphylococcus aureus. StatPearls [Internet]; StatPearls Publishing 2019.
[2] Lowry FD. Staphylococcus aureus infections. N Engl J Med 1998; 339(8): 520-32. [http://dx.doi.org/10.1056/NEJM199808203390806] [PMID: 9709046]
[3] Levy SB. Antibiotic-resistance-the problem intensifies. Adv Drug Deliv Rev 2005; 57(10): 1446-50. [http://dx.doi.org/10.1016/j.addr.2005.04.001] [PMID: 15949867]
[4] Brown AF, Leech JM, Rogers TR, McLaughlin RM. Staphylococcus aureus Colonization: Modulation of Host Immune Response and Impact on Human Vaccine Design. Front Immunol 2014; 4: 507. [http://dx.doi.org/10.3389/fimmu.2013.00907] [PMID: 24469186]
[5] Tong SYC, Davis JS, Eichenberger E, Holland TL, Fowler VG Jr. Staphylococcus aureus infections: epidemiology, pathophysiology, clinical manifestations, and management. Clin Microbiol Rev 2015; 28(3): 603-61. [http://dx.doi.org/10.1128/CMR.00134-14] [PMID: 26016486]
[6] Chaplin DD. 1. Overview of the human immune response. J Allergy Clin Immunol 2006; 117(2)(Suppl Mini-Primer): S430-5. [http://dx.doi.org/10.1016/j.jaci.2005.09.034] [PMID: 16453341]
[7] Rasouli M, Rohshareh M, Mohammad SM, Sajad K, Ahmadrreza M. The Human Immune System against Staphylococcus epidermidis. Critical Reviews in Microbiology 2019; 39
[8] Müller U, Vogel P, Alber G, Schaub GA. The innate immune system of mammals and insects Trends in innate immunity. Karger Publishers 2008; pp. 21-44.
[9] Kawai T, Akira S. The role of pattern-recognition receptors in innate immunity: update on Toll-like receptors. Nat Immunol 2010; 11(5): 373-84. [http://dx.doi.org/10.1038/ni.1863] [PMID: 20404851]
[10] Mantovani A, Casatella MA, Costantini C, Jaillon S. Neutrophils in the activation and regulation of innate and adaptive immunity. Nat Rev Immunol 2011; 11(8): 519-31. [http://dx.doi.org/10.1038/nri3024] [PMID: 21785456]
[11] Bonilla FA, Oetgen HC. Adaptive immunity. J Allergy Clin Immunol 2010; 125(2)(Suppl. 2): S33-40. [http://dx.doi.org/10.1016/j.jaci.2009.09.017] [PMID: 20061006]
[12] Nestle FO, Di Meglio P, Qin J-Z, Nickoloff BJ. Skin immune sentinels in health and disease. Nat Rev Immunol 2009; 9(10): 679-91. [http://dx.doi.org/10.1038/nri2622] [PMID: 19763149]
[13] Di Meglio P, Perera GK, Nestle FO. The multitasking organ: recent insights into skin immune function. Immunol 2011; 35(6): 857-69. [http://dx.doi.org/10.1038/ni.2172] [PMID: 21957434]
[14] Matejcik A. Skin Immunity. Arch Immunol Ther Exp (Warsz) 2018; 66(1): 45-54. [http://dx.doi.org/10.1007/s00005-017-0477-3] [PMID: 28623375]
[15] Ibrahim F, Khan P, Pujalje GG. Bacterial Skin Infections. Prim Care 2015; 42(4): 485-99. [http://dx.doi.org/10.1016/j.pop.2015.08.001] [PMID: 26612370]
[16] Miller LS. Toll-like receptors in skin. Adv Dermatol 2008; 24: 71-87. [http://dx.doi.org/10.1016/j.adrd.2007.11.002] [PMID: 19256306]
[17] Amarante-Mendes GP, Adjenpen S, Branco LM, Zanetti LC, Weinlich R, Bortoluci KR. Pattern recognition receptors and the host cell death molecular machinery. Front Immunol 2018; 9: 2379. [http://dx.doi.org/10.3389/fimmu.2018.02379] [PMID: 30459758]
[18] Pasparakis M, Haase I, Nestle FO. Mechanisms regulating skin immunity and inflammation. Nat Rev Immunol 2014; 14(5): 289-301. [http://dx.doi.org/10.1038/nri3640] [PMID: 24722477]
[19] Bitsch K, Wolz C, Krismer B, Peschel A, Schittek B. Keratinocytes as sensors and central players in the immune defense against Staphylococcus aureus in the skin. J Dermatol Sci 2015; 78(3): 215-20. [http://dx.doi.org/10.1016/j.jdermsci.2017.06.003] [PMID: 28655473]
[20] Fournier B. The function of TLR2 during staphylococcal diseases. Front Cell Infect Microbiol 2013; 2: 167. [http://dx.doi.org/10.3389/fcimb.2012.00167] [PMID: 23316843]
[21] Ozinsky A, Underhill DM, Fontenot JD, et al. The repertoire for pattern recognition of pathogens by the innate immune system is defined by cooperation between toll-like receptors. Proc Natl Acad Sci USA 2000; 97(25): 13766-71. [http://dx.doi.org/10.1073/pnas.250476997] [PMID: 11097540]
[22] Verstak B, Jack S, Ve T, et al. The TLR signaling adaptor TRAM interacts with TRAF6 to mediate activation of the inflammatory response by TLR2. J Leukoc Biol 2014; 96(3): 427-36. [http://dx.doi.org/10.1189/jlb.2A0913-487R] [PMID: 24812060]
[23] Yimin KM, Kohnawan M, Zhao S, et al. Contribution of toll-like receptor 2 to the innate response against Staphylococcus aureus infection in mice. PLOS One 2013; 8(9):e74287. [http://dx.doi.org/10.1371/journal.pone.0074287] [PMID: 24058538]
[24] Feuerstein R, Sridh M, Prinz M, Hennemann P. MyD88 in macrophages is critical for abscess resolution in staphylococcal skin infection. Journal of immunology (Baltimore, Md : 1950) 2015; 194: 2735-45. [http://dx.doi.org/10.4049/jimmunol.1402566]
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[25] Smith JA. Neutrophils, host defense, and inflammation: a double-edged sword. J Leukoc Biol 1994; 56(6): 686.

[26] Rigby KM, DeLeo FR. Neutrophils in innate host defense against Staphylococcus aureus infections. Semin Immunopathol 2012; 34(2): 237-59.

[27] Takada H, Uehara A. Enhancement of TLR-mediated innate immune responses by peptidoglycans through NOD signaling. Curr Pharm Des 2006; 12(32): 4163-72.

[28] Girardin SE, Boneca IG, Viala J, et al. Nod2 is a general sensor of peptidoglycan through muramyl dipeptide (MDP) detection. J Biol Chem 2003; 278(11): 8889-97.

[29] Schäffler H, Demircioğlu DG, Kühner D, et al. NOD2 stimulation by Staphylococcus aureus-derived peptidoglycan is boosted by Toll-like receptor 2 costimulation with lipoproteins in dendritic cells. Infect Immun 2014; 82(11): 4681-8.

[30] Parker D, Prince A. Immunoregulatory effects of necroptosis in immune battles of history: neutrophils and Staphylococcus aureus. Curr Opin Immunol 2014; 26(1): 79-91.

[31] Feuerstein R, Kolter J, Henneke P. Dynamic interactions between Staphylococcus aureus and epidermal Langerhans cells. Microorganisms 2018; 6(3): 87.

[32] Schäffler H, Demircioğlu DG, Kühner D, et al. NOD2 stimulation by Staphylococcus aureus-derived peptidoglycan is boosted by Toll-like receptor 2 costimulation with lipoproteins in dendritic cells. Infect Immun 2014; 82(11): 4681-8.

[33] Ouchi T, Kubo A, Yokouchi M, et al. Induction of MyD88 mediates Neutrophil-mediated Neutrophil Extracellular Traps Degradation by Autologous Neutrophil Extracellular Traps. Front Immunol 2014; 5: 467.

[34] Abtin A, Jain R, Mitchell AJ, et al. Perivascular macrophages mediate neutrophil recruitment during bacterial skin infection. Nat Immunol 2014; 15(1): 45-53.

[35] Kobayashi SD, Malachowa N, DeLeo FR. Pathogenesis of Staphylococcus aureus abscesses. Am J Pathol 2015; 185(6): 1518-27.

[36] Prinz M, Piller M, Jürgen V. Microglia and brain macrophages in the molecular age: from origin to neuropsychiatric disease. Nat Rev Neurosci 2014; 15(5): 300-12.

[37] Miller LS, O’Connell RM, Gutierrez MA, et al. MyD88 mediates neutrophil recruitment initiated by IL-1R but not TLR2 activation in immunity against Staphylococcus aureus. Immunity 2006; 24(1): 79-91.

[38] Guerra FE, Borgoga TR, Patel DM, Sward EW, Vojich JM. Epic immune battles of history: neutrophils vs. Staphylococcus aureus. Front Cell Infect Microbiol 2017; 7: 286.

[39] Kashem SW, Haniffa M, Kaplan DH. Antigen-presentation cells in the skin. Annu Rev Immunol 2017; 35: 469-99.

[40] Kaplan DH. Ontogeny and function of murine epidermal Langerhans cells. Nat Immunol 2017; 18(10): 1088-75.

[41] Dejani NN, Brandt SL, Pilheros A, et al. Topical Prostaglandin E Analog Restores Defective Dendritic Cell-Mediated TlH7 Host Defense Against Methicillin-Resistant Staphylococcus Aureus in the Skin of Diabetic Mice. Diabetes 2016; 65(2): 3718-29.

[42] Ouchi T, Kubo A, Yokouchi M, et al. Langerhans cell antigen capture through tight junctions confers preemptive immunity in experimental staphylococcal scalded skin syndrome. J Exp Med 2011; 208(13): 2607-13.

[43] Glenthøj A, Nickles K, Cowland J, Borregaard N. Processing of Staphylococcus aureus by professional phagocytes. Front Immunol 2014; 21(1): 30.

[44] Boezaert S, Amini P, Anders H-J, et al. To NET or not to NET: current opinions and state of the science regarding the formation of neutrophil extracellular traps. Cell Death Differ 2013; 20(6): 941-48.

[45] van der Does AM, Hiemstra PS, Mookherjee N. Antimicrobial Host Requirement for Staphylococcus aureus Virulence in Immature Macrophages. Front Immunol 2014; 25(12): 104.

[46] van Kessel KP, Bestebroer J, van Strijp JA. Neutrophil-mediated phagocytosis of Staphylococcus aureus. Front Immunol 2014; 5: 467.

[47] van der Does AM, Hiemstra PS, Mookherjee N. Antimicrobial Host Requirement for Staphylococcus aureus Virulence in Immature Macrophages. Front Immunol 2014; 25(12): 104.

[48] Prasad SV, Fiedoruk K, Daniluk T, Piekel E, Buchi R. Expression and Function of Host Defense Peptides at Inflammation Sites. Int J Mol Sci 2019; 21(1): 104.

[49] Glenthøj A, Nickles K, Cowland J, Borregaard N. Processing of Staphylococcus aureus by professional phagocytes. Front Immunol 2014; 21(1): 30.

[50] van der Does AM, Hiemstra PS, Mookherjee N. Antimicrobial Host Requirement for Staphylococcus aureus Virulence in Immature Macrophages. Front Immunol 2014; 25(12): 104.

[51] Prasad SV, Fiedoruk K, Daniluk T, Piekel E, Buchi R. Expression and Function of Host Defense Peptides at Inflammation Sites. Int J Mol Sci 2019; 21(1): 104.

[52] Glenthøj A, Nickles K, Cowland J, Borregaard N. Processing of Staphylococcus aureus by professional phagocytes. Front Immunol 2014; 21(1): 30.

[53] Prasad SV, Fiedoruk K, Daniluk T, Piekel E, Buchi R. Expression and Function of Host Defense Peptides at Inflammation Sites. Int J Mol Sci 2019; 21(1): 104.

[54] Glenthøj A, Nickles K, Cowland J, Borregaard N. Processing of Staphylococcus aureus by professional phagocytes. Front Immunol 2014; 21(1): 30.

[55] Prasad SV, Fiedoruk K, Daniluk T, Piekel E, Buchi R. Expression and Function of Host Defense Peptides at Inflammation Sites. Int J Mol Sci 2019; 21(1): 104.

[56] Glenthøj A, Nickles K, Cowland J, Borregaard N. Processing of Staphylococcus aureus by professional phagocytes. Front Immunol 2014; 21(1): 30.

[57] Prasad SV, Fiedoruk K, Daniluk T, Piekel E, Buchi R. Expression and Function of Host Defense Peptides at Inflammation Sites. Int J Mol Sci 2019; 21(1): 104.

[58] Glenthøj A, Nickles K, Cowland J, Borregaard N. Processing of Staphylococcus aureus by professional phagocytes. Front Immunol 2014; 21(1): 30.

[59] Prasad SV, Fiedoruk K, Daniluk T, Piekel E, Buchi R. Expression and Function of Host Defense Peptides at Inflammation Sites. Int J Mol Sci 2019; 21(1): 104.

[60] Glenthøj A, Nickles K, Cowland J, Borregaard N. Processing of Staphylococcus aureus by professional phagocytes. Front Immunol 2014; 21(1): 30.

[61] Prasad SV, Fiedoruk K, Daniluk T, Piekel E, Buchi R. Expression and Function of Host Defense Peptides at Inflammation Sites. Int J Mol Sci 2019; 21(1): 104.

[62] Glenthøj A, Nickles K, Cowland J, Borregaard N. Processing of Staphylococcus aureus by professional phagocytes. Front Immunol 2014; 21(1): 30.

[63] Prasad SV, Fiedoruk K, Daniluk T, Piekel E, Buchi R. Expression and Function of Host Defense Peptides at Inflammation Sites. Int J Mol Sci 2019; 21(1): 104.
Critical Role of the Antimicrobial Peptide LL-37/CRAMP in Protection of Colon Microbiota Balance, Mucosal Homeostasis, Anti-Inflammatory Responses, and Resistance to Carcinogenesis. Critical Reviews® in Immunology 2019; 39

[64] Febrizia A, Hatta M, Natriz R, Kasim VN, Idrus HH. Activity of Antimicrobial Peptide; Cathelicidin, on Bacterial Infection. Open Biochem J 2019; 13.

[65] Radmacher F, Dreyer S, Kopfnagel V, Gläser R, Werfel T, Harder J. The antimicrobial and immunomodulatory function of RNase 7 in skin. Front Immunol 2019; 10: 2553.

[66] Simians M, Dressel S, Gläser R, Harder J. RNase 7 protects healthy skin from Staphylococcus aureus colonization. J Invest Dermatol 2010; 130(12): 2826-8.

[67] Liu Q, Mazhar M, Miller LS. Immune and Inflammatory Reponses to Staphylococcus aureus Skin Infections. Curr Dermatol Rep 2018; 7(4): 338-49.

[68] Herman A, Herman AP. Antimicrobial peptides activity in the skin. Skin Res Technol 2019; 25(2): 111-7.

[69] Cho JH, Fraser IP, Fukase K, et al. Human peptidoglycan recognition proteins and lysosome Molecular Immunology. Elsevier Inc. 2016; pp. 389-403.

[70] Wang Z-M, Li X, Cocklin RR, et al. Staphylococcus aureus. J Biol Chem 2005; 106(7): 2551-8.

[71] Dziarski R, Royet J, Gupta D. Peptidoglycan recognition proteins and plasticity: insights from epigenetics. Immunology 2011; 134(3): 235-45.

[72] Wagner C, Kotsougianni D, Pich M, Prior B, Wentzsen A, Hänsch B. T lymphocytes in acute bacterial infection: increased prevalence of CD11b(+) cells in the peripheral blood and recruitment to the infected site. Immunology 2008; 125(4): 503-9.

[73] Hidron AI, Kempker R, Moanna A, Rimland D. Methicillin-resistant Staphylococcus aureus. Clin Microbiol Rev 2013; 26(3): 422-47.

[74] Spaulding AR, Salgado-Pabón W, Kohler PL, Horswill AR, Leung DY, Schlievert PM. Staphylococcal and streptococcal superantigen enotoxins. Clin Microbiol Rev 2013; 26(3): 422-47.

[75] Schuler M, Jann NJ, Ferracin F, Landmann R. T and B cells are not required for clearing Staphylococcus aureus in systemic infection despite a strong TLR2-MyD88-dependent T cell activation. J Immunol 2011; 186(1): 443-52.

[76] Hidron AI, Kempter R, Moanna A, Rimland D. Methicillin-resistant Staphylococcus aureus in HIV-infected patients. Infect Drug Resist 2010; 3: 73-86.

[77] Hidron AI, Kempter R, Moanna A, Rimland D. Methicillin-resistant Staphylococcus aureus in HIV-infected patients. Infect Drug Resist 2010; 3: 73-86.

[78] McNair MJ, Rossi AG. Innate Immunity: Phagocytes, Natural Killer Cells, and the Complement System Nijkamp and Parham’s Principles of Immunopharmacology. Springer 2019; pp. 117-37.

[79] Merle NS, Church SE, Fremeaux-Bacchi V, Roumenina LT. Complement System Part 1 - Molecular Mechanisms of Activation and Regulation. Front Immunol 2015; 6: 262.

[80] Nyaaga TK, Kobayashi SD, Freedman B, Porter AR, Voyich JM, Otto M, et al. Interaction of Staphylococci with Human B cells. PloS one 2016; e0164410.

[81] Cooper MD, Alder MN. The evolution of adaptive immune systems. Cell 2006; 124(4): 815-22.