Effects of residual hydrocarbons on the reed community after 10 years of oil extraction and the effectiveness of different biological indicators for the long-term risk assessments

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1. Introduction

Increasing oil exploration, manufacture and transportation of petroleum products has resulted in numerous environmental issues (Lin et al., 2002; Peng et al., 2009; Peterson et al., 2003; Ribeiro et al., 2013). Petroleum Hydrocarbons (PH) generally have...
a direct toxic effect on most organisms (Lin et al., 2002; Peng et al., 2009; Ribeiro et al., 2013), have long persistence in the environment and long-term adverse effects on ecosystems (Culbertson et al., 2008; Li and Boufadel, 2010; Peterson et al., 2003; Reddy et al., 2002). Furthermore, PH may eventually affect human health (Ha et al., 2012; Lu et al., 2012) and associated activities, such as agriculture (Anoliefo and Vwioko, 1995) and tourism (Mendelssohn et al., 2012). Therefore, the ecological risk assessment of oil pollution is of interest to researchers, environmental regulators and legislators. The long-term persistence of PH in the environment highlights the need for further environmental impact studies across a similar or longer period of time.

The development of effective procedures to both assess ecological risk and predict environmental damage from oil pollution depends on the selection of key indicators (Niemeijer and de Groot, 2008). Ideally, key indicators should represent information about the composition, structure and function of the ecological system (Bremner et al., 2006; Dale and Beyeler, 2001), and these indicators should be easily and routinely monitored (Bremner et al., 2006; Dale and Beyeler, 2001; Miller et al., 2006; Niemeijer and de Groot, 2008), be sensitive to oil pollution, respond to oil pollution in a predictable manner and match the spatial and temporal scales of the investigations (Dale and Beyeler, 2001; Niemeijer and de Groot, 2008; Niemi and McDonald, 2004). The predictive power and sensitivity of indicators actually depend on the spatial and temporal scales of variation. Indicators whose scales match that of the investigation will yield higher predictive power and sensitivity, and empirical evidences are needed to support this principle (Wiens, 1989).

A number of studies have investigated the effects of PH on plant individuals, population and the community that contains them (Hester and Mendelsohn, 2000; Meudic et al., 2007; Rosso et al., 2005), based on the premise that plants are the foundation of ecosystem structure and function and are susceptible to environmental perturbations (Miller et al., 2006).

To investigate the PH effects on plants, many studies focus on acute short-term exposure to PH in the laboratory or greenhouse (Lin et al., 2002; Meudic et al., 2007; Rosso et al., 2005; Yu et al., 2012). Under controlled conditions, acute toxicity tests using plant physiological and organismal indicators allow detailed, rapid and cost-effective measures for characterizing the effects of PH on individual plants (Forbes et al., 2006; van Gestel et al., 2001). PH damage the chloroplast, decrease chlorophyll concentration (Rosso et al., 2005; Yu et al., 2012), and inhibit photosynthesis (Chaineau et al., 2003; Pezeshki et al., 2001; Rosso et al., 2005). Further, PH can reduce plant transpiration when they coat the foliage and block the stomata. Recovery of transpiration begins when the PH dissipate from the leaf surfaces (Pezeshki et al., 2000). Reduced leaf growth due to PH exposure was observed in a variety of plant species (Anoliefo and Vwioko, 1995; Zhang et al., 2007). The Leaf Area Index (LAI) usually declines as the level of PH increase (Zhang et al., 2007). This reduction in LAI is due to decreases in the size and number of leaves (Zhang et al., 2007). PH also can inhibit the height (Lin et al., 2002; Peng et al., 2009; Rosso et al., 2005; Zhang et al., 2007), stem density (Culbertson et al., 2008), biomass and coverage (Culbertson et al., 2008; Lin and Mendelsohn, 2012; Peng et al., 2009; Rosso et al., 2005; Zhang et al., 2007) of plants. These tests allow explorations of the underlying mechanisms for the PH effects under controlled conditions, but they are not practical for the analysis and prediction of plant community behavior in all its complexity in field situations (Forbes et al., 2006; van Gestel et al., 2001; Pezeshki et al., 2000; Zhu et al., 2012).

Community-level indicators are more ecologically relevant to changes in the ecosystem (Attrill and Depleège, 1997). However, few studies have investigated the potential long-term effects of elevated PH levels on plant community in the field (Culbertson et al., 2008; Hester and Mendelsohn, 2000). PH have been found to lower plant community productivity (Kinako, 1981; Mishra et al., 2012; Zhu et al., 2013), simplify their structure (Burk, 1977; Collins et al., 1994; Mendelssohn et al., 1990; Mishra et al., 2012) and reduce their biological diversity (Osuji et al., 2004). This change in community composition depends on the differences in species’ sensitivities to PH (Lin and Mendelsohn, 1996; Pezeshki et al., 2000). Usually some species with low tolerance to PH were absent in the following growing season (Burk, 1977; Kinako, 1981). Considering the long-term effects of oil pollution on plant communities (especially the biological diversity), the predictive power and sensitivity of community indicators should be directly compared with indicators at other levels to improve risk assessment (Niemeijer and de Groot, 2008).

To facilitate the selection of indicators and improve the monitoring of effects of elevated PH levels, we investigated long-term impacts of oil pollution on natural community dominated by the reed species (Phragmites australis), and compared the predictive power and sensitivity of different indicators from the physiological, organismal, and community levels. We hypothesized that: (1) combining indicators at the different biological levels may improve our understanding of the effects of elevated PH levels; (2) community indicators may carry more predictive power and be more sensitive to elevated PH levels than indicators below the community level.

2. Materials and methods

2.1. Study area and sample design

The Yellow River Delta in eastern China has undergone long-term disturbance due to intensive oil exploitation since 1964 (Bi et al., 2011; Liang et al., 2012), and provided a suitable site for the ecological risk assessment of oil pollution and consequent elevated soil Total Petroleum Hydrocarbon (TPH) levels. This region is a litoral wetland ecosystem that provides a habitat for a number of plant species and is highly valued for both agricultural and tourism development. In this delta, the reed species (P. australis) was widely distributed and often dominates the plant community (Chen et al., 2011; Zhu et al., 2014).

The field work was conducted in August 2009 within the Chengdong Oilfield (Shengli Oilfield Company, 118°34′38.35″E–118°38′08.53″E, 37°57′34.07″N–38°01′28.50″N) in the Yellow River Delta. The study area is adjacent to the National Natural Reserve of the Yellow River Delta, with a straight distance of approximate 32 km to the Yellow River (Fig. 1). The site is sparsely populated and oil extraction continues to be the major production activity (Bureau of Statistics of the Dongying City, 2013). The soil in this area is saline, and its hydrology is affected by rainfall (Yu et al., 2012). The elevation, soil bulk density and water content of the sample plots were listed in Table A.1 of the supplementary data. Reeds dominate these sample plots with several plant species growing under the reeds, such as Suaeda salsa, Scorzonera mongolica, Limonium bicolor and Aeluropus sariscus (Table A.2 in the supplementary data).

Based on the information provided by the Hekou Oil Production Plant, Shengli Oilfield Company, SINOPEC, sample plots around each of the five oil wells were established (Fig. 1). The duration of pumping from the oil wells was 7, 8, 10, 12 and 13 years. All wells produced heavy oils.

To obtain a gradient of oil pollution and associated changes of plant parameters, seven distances (0, 5, 10, 20, 30, 50 and 100 m) were determined for each oil well. At each distance, three sampling points were located closely. For each sampling point, a 1 m × 1 m quadrat was established (Silliman and Bertness, 2004). Except for net photosynthetic and transpiration rates of reed leaves, other parameters were measured at three sampling points for each
Fig. 1. The location of the study site in eastern China and the sampling design of this investigation. The spatial extent of the panel 3 is approximately 7 km × 8 km. The sample lines and the quadrats in the panel 3 are not in their actual sizes. For the simplicity, only the sampling design of sample plot 1 is fully shown. The sampling design is described in the last paragraph of the Section 2.1.

distance of each well. The net photosynthetic and transpiration rates of reed leaves were measured at the sampling points at distances between 5 m and 100 m in three sample plots because of the availability of time and instrument. For each well, observed values from quadrats with the same distance to oil well center were averaged. Therefore, net photosynthetic and transpiration rates of reed leaves had six averaged data for each of three wells with a total of 18 data, and other parameters had seven averaged data for each of five wells with a total of 35 data.

2.2. Physiology, growth and reproduction of reed

The net photosynthetic and transpiration rates of reed leaves were measured using a portable photosynthesis system (LI-6400, LI-COR Biosciences, Lincoln, NE, United States) between 9:00 and 11:00 am. The LI-6400 was equipped with an LED light source and a CO2 injection system, which maintained constant light intensity (1000 µmol quanta m−2 s−1) and fixed CO2 concentration (360 µmol CO2 mol−1) during the measurements. The light intensity and CO2 concentration were determined based on a one-day test that represented the common condition at that time (Xiao and Wang, 2005). Ten reed individuals were randomly chosen from each sampling point. Gas exchange measurements were made in situ on the widest part of the leaf using the fifth leaf from the reed’s apex. The net photosynthetic and transpiration rates of reed leaves narrower than 2 cm were calculated based on their actual leaf area (Rosso et al., 2005). After measuring foliar gas exchange, the concentration of chlorophyll was measured using a chlorophyll meter (SPAD-502, Konica Minolta Optics, Inc., Japan) from the same leaf position on the reed as the gas exchange measurements (Markwell et al., 1995).

Leaf traits of the reed species, including leaf number, width and length, were measured on thirty reed individuals from each sampling point, along with the reed height. Reed stem density was determined by directly counting the number of stems. The vertical projected coverage (%) of reed was visually estimated. The aboveground parts of reed were harvested and dried at 80 °C for 72 h, then weighed to obtain the aboveground biomass (dry weight) of reed plants (Lin and Mendelssohn, 2012).

2.3. Composition and productivity of the reed community

The coverage and aboveground biomass of the reed community were measured as those of reed plants. The number of plant species in each quadrat was recorded for calculating three diversity indices: Simpson’s diversity index, Shannon–Wiener index and Pielou evenness index. All three indices are widely used in the ecological research (Magurran, 2004). The Simpson index is one of the best known, and earliest, dominance measures. This index is also one of the most meaningful and robust diversity measures available. The Shannon–Wiener index tends to emphasize the species richness component of diversity and is used as a benchmark measure of biological diversity in many long-term ecological researches. The calculation of Shannon index made it possible to calculate a widely used evenness measure, the Pielou evenness index (Johnston and Roberts, 2009; Magurran, 2004). Evenness measures are different from the richness measure, and may be expected to respond to changes in community composition or structure even when there is no change to absolute species richness (Johnston and Roberts, 2009).

Simpson’s diversity index ($D$) was calculated using the following formula:

$$D = 1 - \sum_{i=1}^{S} P_i^2 = 1 - \sum_{i=1}^{S} \left( \frac{N_i}{N} \right)^2$$ (1)

where $P_i$ is the proportion of species $i$ individuals in all individuals in the sample, $S$ is the number of species, $N_i$ is the number of species $i$ individuals, and $N$ is the number of all the individuals.

Shannon–Wiener index ($H$) was calculated using the following formula:

$$H = -\sum_{i=1}^{S} P_i \log_{10} P_i$$ (2)
Pielou evenness index \( (E) \) was calculated using:

\[
E = \frac{H}{H_{\text{max}}}
\]

where \( H \) is the Shannon–Wiener information function and \( H_{\text{max}} \) equals \( \ln S \).

### 2.4. Soil TPH concentration

Spilled oil is usually concentrated in the surface soil (Zhu et al., 2012), especially the top 30 cm of soils (Benka-Coker and Ekundayo, 1995; Zhu et al., 2012). Therefore, three samples from the top 30 cm of soils were taken from each quadrat (Benka-Coker and Ekundayo, 1995). Soil TPH concentrations in the samples were measured using the infrared spectrophotometry method (Chaîneau et al., 2003; Ji et al., 2004). PH were extracted from 10 g dried (60 °C/24 h) soil samples using 20 mL of carbon tetrachloride for 30 min by ultrasonication (40kHz) at 40 °C. The infrared absorption of the extract was measured at the bandwidth of 2930 cm\(^{-1}\). The soil TPH concentration was calculated as the difference between the total TPH in the polluted soil and the biogenic TPH content in the control (Chaîneau et al., 2003).

### 2.5. Data analysis

Soil TPH concentrations and other indicators at the same distance from the each well were averaged, resulting in 18 data points for photosynthetic and transpiration rates and 35 data points for other indicators. To evaluate the predictive power of indicators, simple linear regression models between each indicator and soil TPH concentration were established using IBM SPSS Statistics 19. The Predictive power was represented by the coefficient of determination \( (R^2) \) or proportion of total variance explained by the linear model.

The slope of regression line expresses the change of dependent variable caused by a unit change of the independent variable (Zar, 2010). Therefore, the slope can be used to represent the indicators' sensitivities to soil TPH concentrations. The first step in this analysis is to eliminate the difference in measure scale for different indicators. This step was completed by dividing observed values of each indicator by its largest value, to bring all values in the range \([0,1]\) (Legendre and Legendre, 2012). After the linear regression models were established using standardized data, the sensitivity analysis was finally implemented by comparing slopes of regression lines pairwise. This comparison was accomplished by using the Student's \( t \) statistic in a fashion analogous to that of testing for differences between two population means (Zar, 2010).

### 3. Results

#### 3.1. Presence of petroleum hydrocarbons

Leaks from approximately 10 years of oil extraction resulted in 18 data points for photosynthetic and transpiration rates and 35 data points for other indicators. This step was completed by dividing observed values of each indicator by its largest value, to bring all values in the range \([0,1]\) (Legendre and Legendre, 2012). After the linear regression models were established using standardized data, the sensitivity analysis was finally implemented by comparing slopes of regression lines pairwise. This comparison was accomplished by using the Student's \( t \) statistic in a fashion analogous to that of testing for differences between two population means (Zar, 2010).

#### 3.2. Physiology, growth and reproduction of reed

Residual hydrocarbons significantly inhibited leaf photosynthesis of reed plants (Fig. 2), whereas hydrocarbons did not significantly affect the leaf chlorophyll concentration \( (p = 0.78) \). Residual hydrocarbons also had no significant effect on leaf transpiration \( (p = 0.33) \).

Residual hydrocarbons significantly inhibited the formation and growth of reed leaves. The decrease in leaf number, width and length were significantly and negatively related to increasing soil TPH concentration (Fig. 3). The increasing TPH concentration significantly reduced the reed height (Fig. 4A). The asexual reproduction of reed was not significantly affected by residual hydrocarbons, and the stem density of reed was not significantly correlated with soil TPH concentration \( (p = 0.54) \).

The effect of reduced photosynthetic rates and leaf growth resulted in a significant decrease in plant coverage and aboveground biomass with increasing soil TPH concentration (Fig. 4B and C).

#### 3.3. Composition and productivity of the reed community

The vertical projective coverage and aboveground biomass of the community dominated by reeds significantly decreased with increasing soil TPH concentration (Fig. 5). The species richness also decreased with increasing soil TPH concentration (Fig. 6A). The composition of the community was affected by residual hydrocarbons, as shown by the significant negative association between the three diversity indices (Simpson's diversity index, Shannon–Wiener index, and Pielou evenness index) and soil TPH concentration (Fig. 6B–D).

#### 3.4. Predictive power of indicators

Among the indicators of physiology, growth and reproduction, the three indicators (leaf chlorophyll concentration, transpiration, and stem density of reeds) did not respond to the variation in soil TPH concentration, four indicators (leaf net photosynthetic rate with \( n = 18 \), height, coverage and aboveground biomass of reeds) responded to soil TPH concentration with coefficients of determination \( (R^2 < 0.30) \) and three indicators (leaf numbers, width and length of reed) responded to TPH concentration with \( R^2 > 0.30 \) (Table 1). Among six indicators of the reed community, four indicators (Simpson’s diversity index, Shannon–Wiener index, Pielou evenness index and community aboveground biomass) responded to oil pollution with \( R^2 > 0.30 \). The \( R^2 \) of community vertical projective coverage was 2.0 times as large as that of reed vertical projective coverage, and that of community aboveground biomass was 1.72 times as large as that of reed aboveground biomass. Therefore, the predictive powers of community indicators were better than other indicators, except leaf number, width and length. For all indicators, leaf width yielded the highest predictive power or proportion of total variance explained \( (R^2 = 0.46) \).
Fig. 3. Effects of soil total petroleum hydrocarbons (TPH) concentration on leaf number (A), width (B), and length (C) of reed.

Fig. 4. Effect of soil total petroleum hydrocarbons (TPH) concentration on the reed height (A), foliar projective coverage (B), and aboveground biomass (C).

Table 1
Difference among slopes of regression lines calculated using transformed data (dividing original observed values of each indicator by its largest value). The second column presents the slopes of regression lines, and the minuses before the slopes indicate the negative correlations between indicators and soil TPH concentration. The third column shows the coefficients of determination ($R^2$) of regression lines. The upper/right part shows $t$ values calculated pairwisely, and the lower/left part contains the corresponding $p$ values. $E$ (Pielou evenness index), $D$ (Simpson's diversity index), $H$ (Shannon–Wiener index), $Cover$ (Community coverage), $Richness$ (Species richness), $Bio$ (Community aboveground biomass), $LN$ (Leaf number per reed ramet), $LL$ (Leaf length), $LW$ (Leaf width), $RCover$ (Reed coverage), $RBio$ (Reed aboveground biomass), $Height$ (Reed height), $Photo$ (Net photosynthetic rate of reed leaf).

| Indicators | Slopes   | $R^2$ | $E$     | $D$     | $H$     | Cover | Richness | Bio | LN | LL | LW | RCover | RBio | Height | Photo |
|------------|----------|-------|---------|---------|---------|-------|----------|-----|----|----|----|--------|------|---------|-------|
| $E$        | $-0.63$  | $0.34$ | $0.097$ | $0.287$ | $0.830$ | $0.963$ | $1.051$ | $1.371$ | $1.517$ | $1.641$ | $1.627$ | $1.765$ | $2.115$ | $2.378$ |
| $D$        | $-0.61$  | $0.34$ | $0.46$  | $0.187$ | $0.730$ | $0.867$ | $0.948$ | $1.264$ | $1.407$ | $1.533$ | $1.534$ | $1.668$ | $2.015$ | $2.307$ |
| $H$        | $-0.57$  | $0.36$ | $0.39$  | $0.43$  | $0.572$ | $0.721$ | $0.799$ | $1.132$ | $1.286$ | $1.424$ | $1.426$ | $1.568$ | $1.943$ | $2.362$ |
| Cover      | $-0.47$  | $0.29$ | $0.20$  | $0.23$  | $0.28$  | $0.173$ | $0.206$ | $0.492$ | $0.619$ | $0.777$ | $0.891$ | $0.997$ | $1.345$ | $1.913$ |
| Richness   | $-0.43$  | $0.24$ | $0.17$  | $0.19$  | $0.24$  | $0.43$  | $0.017$ | $0.271$ | $0.378$ | $0.533$ | $0.692$ | $0.775$ | $1.088$ | $1.625$ |
| Bio        | $-0.43$  | $0.31$ | $0.15$  | $0.17$  | $0.21$  | $0.42$  | $0.49$  | $0.285$ | $0.410$ | $0.584$ | $0.735$ | $0.837$ | $1.201$ | $1.906$ |
| LN         | $-0.39$  | $0.38$ | $0.18$  | $0.11$  | $0.13$  | $0.31$  | $0.39$  | $0.39$  | $0.125$ | $0.339$ | $0.551$ | $0.652$ | $1.061$ | $2.106$ |
| LL         | $-0.38$  | $0.46$ | $0.07$  | $0.08$  | $0.10$  | $0.27$  | $0.35$  | $0.34$  | $0.242$ | $0.487$ | $0.592$ | $1.037$ | $2.394$ |
| LW         | $-0.35$  | $0.38$ | $0.05$  | $0.07$  | $0.08$  | $0.22$  | $0.30$  | $0.28$  | $0.37$  | $0.40$  | $0.310$ | $0.387$ | $0.787$ | $1.989$ |
| RCover     | $-0.30$  | $0.14$ | $0.05$  | $0.06$  | $0.08$  | $0.19$  | $0.25$  | $0.23$  | $0.29$  | $0.31$  | $0.38$  | $0.031$ | $0.293$ | $0.971$ |
| RBio       | $-0.30$  | $0.18$ | $0.04$  | $0.05$  | $0.06$  | $0.16$  | $0.22$  | $0.20$  | $0.26$  | $0.28$  | $0.35$  | $0.49$  | $0.289$ | $1.104$ |
| Height     | $-0.26$  | $0.19$ | $0.02$  | $0.02$  | $0.03$  | $0.09$  | $0.14$  | $0.12$  | $0.15$  | $0.15$  | $0.22$  | $0.39$  | $0.39$  | $1.015$ |
| Photo      | $-0.12$  | $0.23$ | $0.01$  | $0.01$  | $0.01$  | $0.03$  | $0.06$  | $0.03$  | $0.02$  | $0.01$  | $0.03$  | $0.14$  | $0.14$  | $0.16$  |
3.5. Sensitivities of indicators

The six community indicators (Pielou evenness index, Simpson’s diversity index, Shannon–Wiener index, community coverage, species richness, and community aboveground biomass) showed higher sensitivities (regression equation slopes) to soil TPH concentration than other indicators (Table 1), and the three diversity indices were the most sensitive. The three leaf indicators (leaf number, length and width) were moderately sensitive. Leaf length was significantly less sensitive than the Pielou evenness index and Simpson’s diversity index. Leaf width, reed coverage and aboveground biomass were significantly less sensitive than all three diversity indexes. Reed height was significantly less sensitive than all three diversity indices as well as community coverage. Net photosynthetic rate was significantly less sensitive than other indicators, except reed coverage, biomass and height.

4. Discussion

4.1. Physiology, growth and reproduction of reed

Reduced plant growth has usually been attributed to hydrocarbons’ effects on plant–soil–air relationships. The short-term and long-term effects are often different. Short-term effects include plant death and growth inhibition when leaves or soil are coated with hydrocarbons (Pezeshki et al., 2000). Long-term effects tend to be sublethal (Peterson et al., 2003) and are usually associated with soil degradation (Mendelssohn et al., 2012). In this study, soil TPH effects on reed plants were sublethal, although the acute effects may have occurred immediately after an increase in soil TPH concentration.

Short-term studies have found that elevated soil TPH concentration was associated with decreased leaf chlorophyll concentration and inhibition of photosynthesis (Chaineau et al., 2003; Pezeshki et al., 2001; Yu et al., 2012). In our study, residual hydrocarbons inhibited photosynthesis in reed leaves (Fig. 2), but had no effect on its leaf chlorophyll concentration. ‘Escravos’ petroleum also decreased the CO₂ assimilation of Salicornia virginica, but had no significant effect on total pigment concentration (Rosso et al., 2005). These results suggested that elevated soil TPH concentration may affect chloroplast function but not its structure, especially in long-term studies, where chloroplast structure often recovers (Pezeshki et al., 2000). In this study, residual petroleum hydrocarbons did not significantly affect leaf transpiration in reeds, because the oil coating on leaves was not observed. This was consistent with the recovery of leaf structure.

Our study found that elevated soil TPH concentration affected the formation and growth of leaves (Fig. 3). The inhibiting effects of elevated soil TPH concentration on reed height, coverage and aboveground biomass were also observed (Fig. 4). Previous field studies indicate that the detrimental effects of elevated soil TPH concentration on biomass continue for several months (Lin and Mendelssohn, 2012) or decades (Culbertson et al., 2008) after initial contamination. Ji et al. (2004) demonstrate experimentally that the addition of soil TPH significantly reduces the aboveground biomass of reeds in the first year but this was reversed during the second year relative to the control (no oil addition). This is likely because petroleum hydrocarbons were broken down by the mature reed wetlands resulting in an improvement in soil nutrients (Ji et al., 2004). However, the results are different in another experiment in which oil addition was repeated in the second year (Ji et al., 2007). These results indicated that the concentration of petroleum hydrocarbons determines the duration of effects, and the long-term effects of elevated soil TPH concentration differed from the short-term effects because of cumulative soil TPH (Lin and Mendelssohn, 2012).

4.2. Composition and productivity of the reed community

If elevated soil TPH concentration reduces individual plant performance, the coverage and aboveground biomass of the community tend to decrease because of the cumulative effects. The reduction in vertical foliar projective coverage of plant community as soil TPH concentration increases was observed in the field from three months (Mendelssohn et al., 1990) to many years (Burk, 1977; Collins et al., 1994) after initial contamination. Controlled experiments and field investigations reveal that the productivity of plant communities is reduced for several months after an oil pollution event (Lin and Mendelssohn, 2012), as was evident in our study of the long-term effects (Fig. 5).

As shown by our study and previous findings (Osuji et al., 2004), species with low soil TPH tolerance tend to die (Lin and Mendelssohn, 1996; Pezeshki et al., 2000), and consequently measures of species richness and diversity change (Osuji et al., 2004; Fig. 6). In this case, the death or disappearance of some species means that those species cannot be used for measuring physiological, organismal, and population indicators. Lethal effects of elevated soil TPH concentration is better reflected by community indicators. Therefore, indicators of species richness or diversity yield additional information about the effects of elevated soil TPH concentration.

4.3. Selection of indicators for the long-term risk assessment

This study indicated that elevated soil TPH concentration had multiple effects on reed community. Indicators from different levels of biological organization responded to long-term elevated soil TPH concentration with different predictabilities and sensitivities. Indicators obtained using standard measurement methods selected in this study were commonly used in previous studies. However, the measurement of coverage, aboveground biomass,
stem density and morphological indicators of plants or leaves require less technology and time than measurements needed to calculate diversity indices of community and physiological functions of individual plants or organs. The biomass measurement was destructive, whilst non-destructive methods (such as remote sensing) still require verification with ground-truth data, which involves destructive sampling. While conducting measurements to calculate diversity indexes required additional time to identify all the species and some species were unfamiliar to the authors, these diversity indexes can help improve our understanding of the effects of elevated soil TPH concentration on the community composition. The measurement of plant and organ physiological processes, such as photosynthesis and transpiration, requires specialized instruments, with which the researcher must be familiar, making such measurements more intensive.

Our study indicated that indicators based on levels below that of community (e.g. population, organism, organ, physiological) had less predictive power than community level indicators for long-term responses to elevated soil TPH concentration, with the exception of leaf number, width and length (Table 1). The predictive power of community aboveground biomass and coverage was higher than those of reed aboveground biomass and coverage (Table 1). In a controlled experiment, Zhang et al. (2007) showed that the decrease of leaf width was more strongly related to elevated soil TPH concentration than decreases in leaf length, stem biomass or height. Furthermore, aboveground biomass of S. alterni-flora has a stronger response to elevated soil TPH concentration than does stem density or height (Lin et al., 2002). Our study found that predictability of leaf length response to elevated soil TPH concentration was more predictable than the response of reed stem density (Table 1).

The sensitivity analysis indicated that community indicators (especially the three diversity indices) were highly sensitive to elevated soil TPH concentration, the leaf number, width and length were moderately sensitive, and the net photosynthetic rate had low sensitivity (Table 1). Zhang et al. (2007) show that leaf length is more sensitive to elevated soil TPH concentration and sediment treatment than are plant height, stem biomass or leaf width. An investigation of the effects on plants of a 14-day ultraviolet-B (UV-B) irradiation show that final accumulated biomass is a better measure of plants' UV-B sensitivity than are levels of chlorophyll or UV-absorbing compounds (Smith et al., 2000). When water stress is imposed on grapevines (Vitis vinifera L.), the final ratio of the number of leaves on lateral branches II to the number of leaves on lateral branches I is more sensitive than is stomatal conductance (Pellegrino et al., 2005). The latter two studies together with our results suggested that the biochemical and physiological processes were less sensitive to environmental changes when investigations were conducted at longer temporal scales, which are far away from the scales these biochemical and physiological processes occur at. Indicators' ability to predict environmental changes depends on the spatial and temporal scales of variation (Wiens, 1989). Indicators that change near the spatial and temporal scales of investigations have high predictabilities and sensitivities (Wiens, 1989). In other words, changing with the environmental stresses at the same scales was an important prerequisite for indicators to show high predictabilities and sensitivities. However, the rankings of indicators may be different when indicators were ordered by their predictive power ($R^2$) and sensitivities (slope) separately, as was shown by the current and previous studies (Zhang et al., 2007).
divergence of predictive power and sensitivity would challenge the selection of indicators, and this issue deserves further investigation.

The scale principle would facilitate the selection of indicators. The spatial and temporal scales of the investigations would give cues for the effectiveness of indicators (Dale and Beyeler, 2001; Niemeijer and de Groot, 2008; Niemi and McDonald, 2004). The short-term controlled experiments examining the individual plants’ response and underlying mechanisms in the laboratory and greenhouse would give priorities to organismal and physiological indicators (Forbes et al., 2006; Zhu et al., 2012). At those scales, the changes of organismal and physiological indicators were more obvious, and the changes of population and community were less so. The early monitoring in the field after oil pollution should consider the extent of the elevated soil TPH concentration effects. If the elevated soil TPH concentration only affects the organs of some plants, the plant community may recover after a period. In this situation, community indicators such as diversity indices may not be appropriate, because community indicators may not change over time. The long-term monitoring in the field should first consider community indicators (Attrill and Depledge, 1997; Zhu et al., 2012), because alternations in the composition and structure of communities constitute an integrated long-term response (including growth, reproduction and mortality) of constituent species (Attrill and Depledge, 1997). Meanwhile, some physiological processes and morphological indicators might no longer change after long-term adaptation of plants. Therefore, a flexible indicator set should be established according the context of the investigation and the progress of the monitoring.

5. Conclusions

Community indicators responded to long-term oil pollution with higher predictive power and sensitivities compared with other indicators, except leaf length, number and width, which yielded the highest predictive power. Determining the scale of indicators may facilitate their selection, especially when there is a lack of available information. However, the criterion of indicator will change with the objectives of the assessment. To better assess the ecological risk of oil pollution, the methodology and analyses at different levels should be combined to describe real-world biological systems under oil pollution at multiple spatial and temporal scales.

Acknowledgements

This work was supported by the National Key Technology R&D Program of China (2008BA43B01). This funding had no involvement in the study design; in the collection, analysis and interpretation of data; in the writing of the report; and in the decision to submit the article for publication. The authors appreciate the two anonymous reviewers improving this manuscript with their careful and insightful reviews. The authors appreciate Prof. Guangsheng Zhou providing the photosynthesis system (LI-6400) and Dr. Bingrui Jia guiding the use of this equipment. The authors declare that no competing interest exists.

Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at http://dx.doi.org/10.1016/j.ecolind.2014.08.017.

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