Small-Molecule Inhibition of BRDT for Male Contraception

Martin M. Matzuk, Michael R. McKeown, Panagis Filippakopoulos, Qinglei Li, Lang Ma, Julio E. Agno, Madeleine E. Lemieux, Sarah Picaud, Richard N. Yu, Jun Qi, Stefan Knapp, and James E. Bradner

INTRODUCTION

Although ~4% of the mammalian genome encodes genes expressed in male germ cells during spermatogenesis (Schultz et al., 2003), contraceptive drugs for men have remained elusive. To date, the only drugs in clinical trials are testosterone analogs that alter endogenous androgen production, although there is a short list of other possible targets (e.g., GAPDH) and drugs (e.g., gamendazole) (Aitken et al., 2008). This lack of contraceptive alternatives for men is partially responsible for the high rate of unplanned pregnancies, especially in teenagers, and contributes to the maternal mortality, ethical, social, and financial costs associated with abortions and deliveries to single mothers. To approach this dearth of contraceptive alternatives for men, we have undertaken to develop small molecules that could target spermatogenic-specific proteins that have been shown to be essential for both spermatogenesis and fertility in mammals. One such contraceptive target is the testis-specific and bromodomain-containing protein BRDT.

BRDT is a tissue-restricted, chromatin-associated protein expressed in pachytene spermatocytes, diploteine spermatocytes, and round spermatids (Shang et al., 2007). During postmeiotic maturation, BRDT localizes to the nucleus and reorganizes hyperacetylated histones through twin acetyl-lysine recognition modules, or bromodomains (Berkovits and Wolgemuth, 2011; Morinière et al., 2009; Shang et al., 2007). The essential role of BRDT in spermatogenesis is mediated by the first bromodomain (BRDT(1); Figure 1A), which binds the tetra-acetylated amino-terminal tail of histone 4 (H4Kac4) with moderate potency (20 μM) (Morinière et al., 2009). Structural studies of murine BRDT have demonstrated that BRDT(1) binds a diacetylated histone 4 peptide (H4Kac8ac) in part through a conserved asparagine (Morinière et al., 2009), akin to other bromodomain coactivator proteins (Dhalluin et al., 1999). Genetic studies of BRDT have demonstrated that selective deletion of the BRDT(1)-encoding region is sufficient to confer sterility in...
homzygous hypomorphic male mice (Shang et al., 2007), and a recently published genome-wide association study of idiopathic male infertility identified single-nucleotide polymorphisms of BRDT as significantly associated with oligozoospermia or azoospermia in European men (Aston et al., 2010). These insights establish a compelling rationale to target BRDT for a contraceptive effect.

Recently, we established the feasibility of targeting human bromodomains with acetyl-lysine competitive small molecules (Filippakopoulos et al., 2010). Our index study described a potent thienodiazepine inhibitor ((+)-JQ1; Figure 1B; $K_I = 90$ nM) of the BET family coactivator protein BRD4, which is implicated in the pathogenesis of cancer via transcriptional control of the MYC oncogene (Delmore et al., 2011; Zuber et al., 2011). Protein sequence alignment of human BRD4(1) to BRD4(2) reveals 81% identity and 89% similarity, including all surface residues predicted to contact (+)-JQ1 (Figure 1C and Data S1 and S2 available online). Based on these insights and preliminary evidence of binding to BRDT(1) established by differential scanning fluorimetry (Filippakopoulos et al., 2010), we endeavored to characterize the biochemical and functional effects of (+)-JQ1 on BRDT(1).

We report here that direct inhibition of BRDT by a small-molecule bromodomain inhibitor exerts a dose- and time-dependent inhibitory effect on spermatogenesis. Structural studies of BRDT(1) bound to JQ1 reveal ligand-receptor shape complementarity and complete occlusion of the acetyl-lysine recognition cavity. These and further biochemical studies establish the molecular mechanism of potent BRDT inhibition observed in vivo, where JQ1 targets meiotic and postmeiotic male germ cells, leading to impaired spermatogenesis and compromised motility.

In mating studies, JQ1 accomplishes a complete and reversible inhibitory effect on spermatogenesis. Structural studies of (+)-JQ1 and BRDT(1) are depicted with a black star. Contacts between (+)-JQ1 and hBRDT(1) is depicted with a blue star. Contacts between (+)-JQ1 and mBRDT(1) is depicted with a black star.
testosterone levels or mating behaviors and without prompting obvious teratogenic effects in offspring. These results indicate that targeting a developmental epigenetic reader protein with an orally bioavailable small molecule can modulate male fertility for a contraceptive effect.

RESULTS

Molecular Recognition of BRDT by JQ1
To assess competitive binding to BRDT(1), we devised homogeneous, luminescence proximity assays that are capable of quantifying binding of a synthetic, biotinylated tetra-acetylated histone 4 peptide (H4Kac4, residues 1–20) to recombinant epitope-tagged murine or human BRDT(1). Dose-ranging studies of (+)-JQ1 demonstrated potent inhibition of H4Kac4 binding with a half-maximum inhibitory concentration (IC50) value of 10 nM for human BRDT(1) and 11 nM for human BRDT(1) (Figure 1D). In contrast, the (−)-JQ1 enantiomer was inactive for either ortholog, establishing stereospecific, ligand-competitive binding. To confirm competitive inhibition of the acetyl-lysine binding module, isothermal titration calorimetry (ITC) was performed on human BRDT(1), employing a synthetic H4Kac4 peptide in the presence and absence of (+)-JQ1 (Figure 1E). This peptide was found to bind two protein modules concomitantly with a Kd of 25.5 μM, in good agreement with published data (Morinière et al., 2009). This interaction was directly inhibited in the presence of (+)-JQ1 that partially saturated the acetyl-lysine binding (ratio 1.0:8) due to solubility limits of the inhibitor. The remaining unoccupied sites bound the H4Kac4 peptide with identical affinity and expected stoichiometry (Table S1). Binding free energies and dissociation constants (Kd) determined by ITC for each bromodomain of human and murine BRDT confirm potencies, specific (ratio 1:1) interactions with all domains (Kd = 44–190 nM; Figure 1F). Notably, (+)-JQ1 binds murine BRDT(1) with high ligand efficiency (LE = 0.30), compared to reported inhibitors of protein–protein interactions (Wells and McClendon. 2007).

The mode of binding of (+)-JQ1 to BRDT(1) was established by X-ray crystallography at high-resolution (Figure 2 and Table S2). Similar to the structure of murine BRDT(1) (Morinière et al., 2009), human BRDT(1) exhibits the canonical structural features of a bromodomain, namely a left-handed bundle of four antiparallel α helices (αZ, αA, αB, and αC) and a hydrophobic acetyl-lysine binding pocket defined by interhelical ZA and BC loops (Figures 2A–2C) (Dhalluin et al., 1999). The observed, extraordinary shape complementarity between (+)-JQ1 and BRDT(1) explains the increased potency of (+)-JQ1 compared to the physiologic H4Kac4 ligand. In addition, (+)-JQ1 engages all surface residues analogous to those on murine BRDT(1) mediating molecular recognition of H4Kac8ac (Figures 2D and 2E) (Morinière et al., 2009). Notable examples include a hydrogen bond linking the triazole ring and the evolutionarily conserved asparagine (Asn109), as well as extensive hydrophobic contacts with the ZA and BC loop regions (Figure 2B). These biochemical and structural studies confirm and explain potent, acetyl-lysine competitive inhibition of BRDT(1) by (+)-JQ1 (hereafter referred to as JQ1).

Testis Bioavailability of JQ1
To test the ability of JQ1 to reach its contraceptive target cells, the testicular bioavailability of JQ1 was assessed by serum and tissue pharmacokinetic analysis. Following a single intraperitoneal (i.p.) dose of JQ1 (50 mg/kg) in male mice, measurements of JQ1 concentration were made in serum, testis, and brain (Figure 3A and Table S3). Overall, JQ1 exhibits excellent testicular bioavailability (AUCtestis/AUCplasma = 259%), suggesting preferential distribution into this tissue compartment with rapid (Tmax = 0.25 hr) and pronounced exposure (Cmax = 34 μg/ml). Corroborating barrier permeability by JQ1, nearly uniform blood-brain barrier permeability was observed after single-dose pharmacokinetic studies (AUCbrain/AUCplasma = 98%).

JQ1 Impairs Sperm Count and Motility
To determine the possible consequences of blocking BRDT function in vivo, we tested the spermatogenic effects of JQ1 administered to male mice. First, juvenile or adult male mice were administered daily i.p. injections of JQ1 (50 mg/kg once daily [QD]) or vehicle control over a 3 or 6 week period. After drug or vehicle treatment, mice were either sacrificed or mated to females while continuing to receive JQ1. All JQ1-treated males had a significant reduction in testis volume (Figures 3B and 3C). With daily injections of 50 mg/kg JQ1, males treated from 3–6 weeks of age showed a reduction to 75.4% of control, males treated from 6–9 weeks of age showed a reduction to 54.7%, and males treated for 6 weeks with JQ1 (6–12 weeks of age) showed a reduction to 40.6% (Figure 3C). Thus, longer treatment with JQ1 has the most prominent effect on testicular volume.

To examine the effect on sperm production, sperm counts were determined after JQ1 or vehicle treatment. Epididymal sperm number was reduced to 28% of control after 3 weeks of 50 mg/kg daily exposure (3–6 weeks of age) and 11% of control after 6 weeks of treatment (6–12 weeks of age) (Figure 3D). Analysis of sperm motility after 3 weeks of JQ1 (50 mg/kg QD) revealed a significant, 4.5-fold reduction in sperm motility (16.0% for the JQ1-treated versus 72.4% for the control; Figure 3E). Following 6 weeks of daily JQ1, only 5% of the sperm demonstrated progressive motility compared to 85% of sperm from controls (Figure S1). Thus, daily exposure to JQ1 quantitatively reduced sperm number and motility.

Recently, the tolerability of twice-daily administration of JQ1 was reported in a confirmatory study of JQ1 in hematologic malignancies (Mertz et al., 2011). Pharmacokinetic study of twice-daily (BID) JQ1 (50 mg/kg) confirmed excellent penetration of the blood:testis barrier and demonstrated a marked increase in drug exposure in the testis (AUC0–24hr = 96,640 hr*ng/ml; Figure 3F and Table S4), accompanied by apparent partitioning into this compartment (AUCtestis/AUCplasma = 236%). With BID injections of JQ1 (JQ1 50 mg/kg), adult males treated for 3 weeks (9–12 weeks) showed reduction in testis weight to 63.3% of controls (Figure 3G), Paralleling the reduction in testis volume, BID treatment resulted in reduction in sperm in the cauda epididymis to 36.6% of the control (Figure 3H), a dramatic 5.8-fold reduction in sperm motility (11.2% for JQ1-treated versus 64.5% for controls; Figure 3I), firmly establishing the tolerability and antispermatogenic effect of BID JQ1 in mice. Altogether,
these findings are similar to defects observed in mice deficient in BRDT(1) (Shang et al., 2007).

JQ1 Selectively Targets Male Germ Cells without Affecting Hormone Levels

Based on the observed inhibition of spermatogenesis, we performed further mechanistic studies of BRDT inhibition. Consistent with the reduction in testis volume, the cross-sectional area of seminiferous tubules from testes of males treated daily from 3–6 weeks with JQ1 were 84.9% of controls, males treated from 6–12 weeks with JQ1 were 56.4%, and males treated from 9–12 weeks with JQ1 BID were 65.0% of controls (Figure 3J). Histological examination established a JQ1-dependent decrease in the amount and number of tubules that had obvious and abundant round spermatids and spermatozoa in the lumen (Figures 4A–4H). This was particularly apparent in stage VII tubules, in which a marked reduction in round spermatids was observed in JQ1-treated mice. Of note, mice treated with JQ1 BID for 3 weeks had a 79% reduction in round spermatids in stage VII tubules (112.7 ± 7.4 [controls] versus 23.9 ± 3.6 [JQ1-treated]; p < 0.0001). Analysis of epididymides of JQ1-treated males also showed a similar finding in which fewer sperm were observed in the epididymal lumen compared to their abundance in controls (Figure S1). Detailed microscopic analysis of the testes of mice treated with JQ1 revealed seminiferous tubule degeneration, sloughing of germ cells, and multinucleated syncytiasts (Figures 4F and 4H; black arrows), findings also described in cyclin A1 (CcnA1) knockout mice, which are infertile due to a spermatocyte block (Liu et al., 1998). Thus, JQ1 treatment causes a block in spermatogenesis and a resultant reduction in testicular production of spermatozoa.

Because absence of glycoprotein hormone and abrogation of androgen signaling pathways cause male infertility (Matzuk and Lamb, 2008), we measured the serum levels of follicle-stimulating hormone (FSH), luteinizing hormone (LH), and testosterone after treatment with JQ1. Differences in the levels of FSH,
producing Leydig cells in JQ1-treated males (Figures 4A and 4B; red arrows). Thus, JQ1 effects are specific to germ cells and do not alter hormone-dependent processes.

**Figure 3. BRDT Inhibition with a Testis-Permutable Small Molecule Reduces Testis Size and Sperm Counts and Motility**

(A) Pharmacokinetic analysis of JQ1 in plasma (black lines) and testis (red lines) following a single administration of JQ1 (50 mg/kg IP) to male mice. Data represent the mean ± SD.

(B) Gross analysis of testes from 9-week-old mice that received control or JQ1 (50 mg/kg daily) for 3 weeks.

(C) Testis weights from mice treated with control or JQ1 (50 mg/kg QD) for 3–6 weeks, 6–9 weeks, or 6–12 weeks of age. Data represent the mean ± SEM and are annotated with P values obtained from a two-tailed t test (*p < 0.05).

(D) Graphical representation of sperm counts obtained from the entire epididymides of males treated with JQ1 or control from 6–9 weeks of age or the tail of the epididymis of males treated from 6–12 weeks of age with vehicle or JQ1 (50 mg/kg QD). Data represent the mean ± SEM (*p < 0.05).

(E) Motility of mature sperm obtained from the cauda epididymis of adult males treated with JQ1 (50 mg/kg daily) from 6–12 weeks of age. Data represent the mean ± SEM (*p < 0.0001).

(F) Pharmacokinetic analysis of JQ1 in plasma (black lines) and testis (red lines) following twice-daily (BID) administration of JQ1 (50 mg/kg IP) to male mice. Data represent the mean ± SD.

(G) Sperm counts (mg) from mice treated with control or JQ1 (50 mg/kg BID) from 9–12 weeks of age. Data represent the mean ± SEM (*p < 0.05).

(H) Sperm counts obtained from the cauda epididymis of males treated from 9–12 weeks of age with vehicle or JQ1 (50 mg/kg BID). Data represent the mean ± SEM (*p < 0.05).

(I) Motility of mature sperm obtained from the cauda epididymis of adult males treated with JQ1 (50 mg/kg BID) or vehicle from 9–12 weeks. Data represent the mean ± SEM (*p < 0.001).

(J) Motility of mature sperm obtained from the cauda epididymis of adult males treated with JQ1 (50 mg/kg BID) or vehicle from 9–12 weeks, as shown) or control (*p < 0.05).

(K, L, and M) Male mice treated with JQ1 or vehicle from 6–12 weeks exhibit statistically similar serum levels of (K) testosterone, (L) luteinizing hormone (LH), and (M) follicle-stimulating hormone (FSH). See also Figure S1 and Tables S3 and S4.

LH, and testosterone were not statistically significant between control and JQ1-treated males (Figures 3K–3M). Likewise, weights of the seminal vesicles, a major androgen-responsive tissue, were similar (Figure S1). These results are consistent with the presence of histologically normal testosterone-producing Leydig cells in JQ1-treated males (Figures 4A and 4B; red arrows). Thus, JQ1 effects are specific to germ cells and do not alter hormone-dependent processes.

**Molecular Analysis of the Germ Cell Defects in JQ1-Treated Testes**

To molecularly define the consequences of BRDT inhibition by JQ1, we performed genome-wide expression analysis of males treated with 50 mg/kg JQ1 daily for 6 weeks (6–12 weeks of age). Hierarchical clustering separated samples by treatment assignment and identified 1,685 unique genes enriched or upregulated and 675 unique genes depleted or downregulated (2-fold or greater; Q value < 0.05) with JQ1 exposure (Figure 5A). The broad transcriptional events observed—more than an order of magnitude greater than prior studies of JQ1 (Delmore et al., 2011)—suggested a pronounced effect on multistage differentiation more than selective effects on discrete transcriptional programs. Multiple germ-cell-expressed genes were suppressed upon treatment with JQ1, including Ccna1 (expressed in pachytene spermatocytes), Msy2 (pachytene spermatocytes and postmeiotic germ cells), polo-like kinase 1 (Plk1; diplotene and diakinesis
stage spermatocytes, secondary spermatocytes, and round spermatids), aurora kinase C (Aurkc; chromocenter clusters in diplo- tene spermatocytes), and A-kinase anchoring protein 4 (Akap4; postmeiotic germ cells). These results are significant because knockout mice lacking cyclin A1 have a spermatocyte block (Liu et al., 1998); Msy2 knockout mice are sterile due to postmeiotic germ cell block and multinucleated spermatids (Yang et al., 2005); absence of AKAP4 prevents progressive motility (Miki et al., 2002); humans with aurora kinase C mutations demonstrate polyploid nuclei secondary to meiosis I arrest (Dieterich et al., 2007); and the mammalian PLK1 protein, similar to fruit fly polo kinase, is believed to function in the completion of meiotic division.

To determine the stages of spermatogenesis at which JQ1 functions, we then queried these data with functionally defined gene sets reflecting early (pachytene spermatocyte) or late (spermatid) transcriptional signatures (Schultz et al., 2003) and observed coordinate depletion (normalized enrichment score [NES] = −2.5 to −2.1; p < 0.001) in three independent modules by gene set enrichment analysis (GSEA).

Our expression profiling results were confirmed and extended by gene-specific RT-PCR (Figure 5D and Table S6). Genes expressed early in spermatogenesis such as Plzf, a spermatogonial marker, and Stra8, expressed in differentiating spermatogonia and preleptotene spermatocytes, are 2.0- and 1.3-fold enriched, respectively, in testes of JQ1-treated mice compared to controls. In addition to Ccna1, Msy2, Plk1, Aurkc, and Akap4, other key genes expressed during meiosis or spermiogenesis, including Brdt (mid- to late-spermatocytes), Papoll (step 1–7 round spermatids), Kif17 (step 4–7 spermatids), and Prm1 (step 7–16 spermatids), are 2.1- to 7.3-fold lower in testes of mice treated with JQ1 versus control. Unlike the Brdt mutant mouse (Shang et al., 2007), in which pachyten e spermatocyte-expressed gene Hist1h1t is upregulated, JQ1 treatment leads to a 2.6-fold downregulation of this gene, in line with suppression of Brdt, Ccna1, Msy2, and Plk1. This difference is likely secondary to enhanced blockade of BRDT function by JQ1 in spermatocytes compared to the Brdt hypomorphic mouse.

To further characterize the consequences of these molecular changes, we performed histological analysis of zona pellucida...
BRDT Inhibition Confers a Reversible Contraceptive Effect in Males

To determine the effects of alternative JQ1 treatment regimens on the fertility of male mice, JQ1 (50 mg/kg/day) and vehicle...
control were each delivered to 10 male mice for 6 weeks. To ensure that JQ1 and vehicle were having proper effects, three males from each group were sacrificed and were shown to have reduced testis size (control: 107.4 ± 8.8; JQ1: 62.8 ± 7.5 mg; p < 0.05), sperm counts (control: 12.3 ± 0.6 × 10^6; JQ1: 3.8 ± 0.3 × 10^6; p < 0.0001), and sperm motility (control: 72.4 ± 2.5%; JQ1: 16.0 ± 0.1%; p < 0.0001). After this pretreatment period, the remaining seven JQ1-treated and seven control males were mated continuously to two adult females per male while continuing to receive JQ1 or vehicle. After the first month of breeding, the seven control males had sired 14 litters of offspring. However, only four of the seven JQ1-treated males sired offspring (only six litters of offspring and a reduced number of pups). For the three JQ1-treated males that had not yet sired offspring, these mice demonstrated normal mating behavior as evident by the presence of copulation plugs. Thus, even at a low dose of JQ1, there is a partial contraceptive effect.

As the mice appeared to be healthy at 50 mg/kg/day of JQ1, we escalated the dosage of JQ1 in the second mating month to 75 mg/kg/day. The male mice appeared to fall into two groups: those that show a contraceptive effect with 50 mg/kg/day followed by 75 mg/kg/day (Figure 7A) and those that still produced at least one litter under this lower-dose regimen (Figure 7B). For the three mice that were most responsive to the low-dose regimen, only one was able to sire a litter of two offspring (born on the first day of month 2). For the latter group, the four males that sired offspring were treated in mating month 3 with 50 mg/kg twice per day. These males were not observed to sire any offspring in month 3 (Figure 7B). Thus, depending on individual variability of each of the males, total daily dosages between 50 and 100 mg/kg can produce a complete contraceptive effect in male mice.

To assess the reversibility of the JQ1-induced contraceptive effect, we examined whether fertility returned after JQ1 treatment was halted. Among the three infertile mice treated with the low-dose regimen, infertility remained complete at 1 month of recovery (mating month 3), indicating a durable effect of JQ1 on spermatogenesis. However, in mating month 4, all three males sired offspring with a statistically similar number of pups per female as observed for the controls (Figure 7A). The average days to effective copulation was estimated to be 31.7 ± 6.0 days.
In mice, the recovery of fertility was assessed following the cessation of JQ1 treatment. After a total mating period of 7 months, the weights and sperm parameters approached normal levels (Figure S4), consistent with fully recovered fertility. Thus, mice treated with JQ1 at 50 mg/kg/day and then 75 mg/kg/day recover fertility within 6 weeks of withholding the drug and show no long-term effects of JQ1 treatment. For the mice that required escalation to the high-dose regimen (50 mg/kg BID), JQ1 was halted after the first month of this treatment (i.e., treatment month 3). In months 4 and 5, these males failed to sire offspring (Figure 7B). However, all JQ1-treated males sired offspring in month 6 and sired similar litter sizes as controls in months 6–8. The average days to effective copulation for this cohort was estimated to be 65.7 ± 7.7 days (range: 58–81 days). Thus, longer JQ1 treatment and/or the increased dosage of JQ1 resulted in an extended (~1 month) period of infertility.

To more precisely determine the integrity and function of the testes and spermatozoa following withdrawal of JQ1 therapy, two additional independent experiments were performed. In the first experiment, JQ1-treated adult male mice were examined at the end of 6 weeks of treatment (50 mg/kg/day) or after 2 or 4 months following cessation of therapy. After 2 months, the testis weight and sperm counts and motility were increased, and after 4 months, the weights and sperm parameters approached normal levels (Figure S4). Histologically, seminiferous tubules of JQ1-treated mice are indistinguishable from controls (Figures 7I and 7J). In the second experiment, a cohort of mice was treated with vehicle or 3 weeks of high-dose JQ1 (50 mg/kg BID). Mice were sacrificed at 3 weeks of treatment and 10, 30, and 60 days after JQ1 withdrawal. Males treated with high-dose JQ1 continue to show defects in testicular parameters 30 days after halting treatment but exhibit complete recovery of all parameters within 60 days (Figures 7F–7H), consistent with the documented recovery of fertility. Thus, depending on duration and overall dosage of JQ1, testicular and semen parameters normalize within 1–3 months to allow productive copulation and return of fertility.

Figure 7. BRDT Inhibition with JQ1 Causes a Reversible Contraceptive Effect in Male Mice

(A) Adult males were pretreated for 6 weeks with vehicle control (n = 7) or JQ1 (50 mg/kg QD; n = 3) and then caged continuously with two females each while continuing 50 mg/kg QD for month 1 and escalating to 75 mg/kg QD for month 2. JQ1 treatment was stopped at the end of month 2 of mating. Graphical representation of pups born in each month to the caged females reveals a contraceptive effect evident in months 1–3 (data represent mean ± SEM; *p < 0.001) and durable restoration of fertility at month 4.

(B) Adult males were pretreated for 6 weeks with vehicle (n = 7) or JQ1 (50 mg/kg QD; n = 4) and then caged continuously with two females each while continuing 50 mg/kg QD for month 1, escalating to 75 mg/kg QD for month 2, and further escalating to 50 mg/kg BID for month 3. JQ1 treatment was stopped at the end of month 3 of mating. Graphical representation of pups born in each month to the caged females reveals a complete contraceptive effect evident in months 3–5 (data represent mean ± SEM; *p < 0.001) and durable restoration of fertility at month 6.

(C–E) Graphical representation of (C) testis mass (mg), (D) sperm count, and (E) sperm motility of adult control males (n = 3) and JQ1-treated males (n = 3) following a 4 month recovery from the low-dose regimen shown in (A) (data represent mean ± SEM; NS, not significant).

(F–H) Graphical representation of (F) testis mass, (G) sperm count, and (H) sperm motility of adult control males (n = 4) and JQ1-treated males after 3 weeks of 50 mg/kg BID treatment (n = 2), as well as following 10 days (n = 2), 30 days (n = 2), and 60 days (n = 2) of recovery from the high-dose (BID) regimen (data represent mean ± SEM; *p < 0.05).

(I and J) Microscopic analysis of seminiferous tubules from (I) control and (J) JQ1-treated males following 4 months of recovery reveals complete histological recovery.

(K) Litter sizes and pups born following JQ1 cessation are normal. See also Figure S4.

(range: 26–38 days) based on the birth of the first litters after treatment was stopped and the 19 day gestation period of mice. After a total mating period of 7 months (i.e., 4 months after halting the low-dose regimen), testis volume, seminiferous tubule area, testis histology, sperm motility, and sperm counts were statistically similar to the control group (Figures 7C–7E and S4), consistent with fully recovered fertility. Thus, mice treated with JQ1 at 50 mg/kg/day and then 75 mg/kg/day recover fertility within 6 weeks of withholding the drug and show no long-term effects of JQ1 treatment. For the mice that required escalation to the high-dose regimen (50 mg/kg BID), JQ1 was halted after the first month of this treatment (i.e., treatment month 3). In months 4 and 5, these males failed to sire offspring (Figure 7B). However, all JQ1-treated males sired offspring in month 6 and sired similar litter sizes as controls in months 6–8. The average days to effective copulation for this cohort was estimated to be 65.7 ± 7.7 days (range: 58–81 days). Thus, longer JQ1 treatment and/or the increased dosage of JQ1 resulted in an extended (~1 month) period of infertility.
Lastly, offspring that were born from JQ1-treated males showed normal size, activity, and behavior as offspring born from controls (Figure 7K and data not shown). When offspring from vehicle and JQ1-treated fathers were allowed to mate with adult females (2 females/cage) at 6 weeks of age, both sets of males (n = 6 for each group) displayed normal mating behavior, sired offspring from each female, and produced statistically similar litter sizes from matings over the first month (control: 7.73 ± 0.70 pups/litter; JQ1: 7.50 ± 0.34 pups/litter). At 11 weeks of age, the testis weights (control: 98.7 ± 2.8 mg; JQ1: 95.8 ± 3.2 mg), sperm counts (control: 11.8 ± 0.3 × 10⁶; JQ1: 11.2 ± 0.6 × 10⁶), and sperm motility (control: 69.5 ± 1.7%; JQ1: 69.8 ± 1.3%) of the offspring were statistically similar. Together, these findings indicate that JQ1 did not have any long-term transgenerational effects on testis physiology or reproductive capacity.

DISCUSSION

Using a highly potent and selective chemical probe, our studies provide pharmacologic validation of the amino-terminal bromodomain of BRDT as a target for male contraception. JQ1 emerges as a lead compound for a new class of drugs that can cross the blood-testis boundary, inhibit bromodomain activity during spermatogenesis, impair sperm generation and motility, and produce a reversible contraceptive effect in mammals. These data support JQ1 as the first contraceptive agent that selectively and reversibly targets the male germ cell.

Modulation of chromatin structure during spermatogenesis by an inhibitor of an epigenetic reader protein, such as BRDT, raises concern for epigenetic developmental abnormalities in progeny born to animals on treatment or following cessation. To date, we have not identified any anomalous phenotypes in offspring born from vehicle or JQ1-treated fathers. We have only observed full recovery of fertility in treated males; testis size, seminiferous tubule diameter, histological appearance, sperm number and motility, and fertility of JQ1-treated males recover to the levels of controls (Figures 7 and S4). These observations are consistent with the restricted expression of the BRDT target in pachytene spermatocytes through round spermatids and with the function of BRDT in these stages to promote spermatogenesis. Indeed, the phenotype of mice treated with JQ1 closely resembles the seminal findings by Wolgemuth and colleagues in the characterization of BRDT(1) hypomorphic mice (Shang et al., 2007). We cannot rule out that some of effects of JQ1 are through inhibition of other bromodomain proteins possessing selectivity for BRDT to extend the favorable pharmacologic profile of BRDT inhibition for male contraception. Recently described tool compounds of BET bromodomains provide divergent putative scaffolds for lead optimization (Chung et al., 2011; Dawson et al., 2011; Nicodeme et al., 2010). Compounds that demonstrate higher affinity and specificity for BRDT would be expected to reduce any possible long-term, adverse effects of a pan-BET bromodomain inhibitor. Structural and sequence comparisons of BRDT(1) and BRD4(1) reveal differences in surface residues that may be explored to develop further isoform-selective BET inhibitors. Because human and mouse BRDT proteins are highly conserved and have nearly identical bromodomain pockets based on our structural predictions, we envision that our discoveries can be completely translated to men, providing a novel and efficacious strategy for a male contraceptive.

This chemical genetic study of bromodomain inhibition for male contraception provides pharmacologic target validation, encourages further drug development efforts, and offers insights guiding and enabling pharmaceutical BRDT inhibitor development. As this index study closely examines effects on human and murine BRDT, further study of additional species may be relevant for drug development. Toward this objective, we have completed preliminary studies of JQ1 in rats in which we have observed excellent pharmacologic exposure to JQ1 i.p. injection (Figure S5A and Table S7) and have identified a tolerated dose (10 mg/kg i.d.), which is effective in reducing testis mass, sperm count, and motility (Figures S5B–S5D), accompanied by comparable histological evidence of impaired spermiogenesis (Figure S5E). For a chemical probe intended for laboratory research, JQ1 has excellent drug-like properties yet possesses a short terminal elimination half-life. Pharmacokinetic studies performed herein demonstrate that excellent drug exposure can be accomplished with twice-daily administration. The observed complete contraceptive effect of JQ1 when given twice daily suggests that more continuous inhibition of BRDT-dependent effects on spermatogenesis may be required for clinical translation of this research.

We are currently creating and testing the effects of JQ1 derivatives possessing selectivity for BRDT to extend the favorable therapeutic window of BRDT inhibition for male contraception. Studies performed herein demonstrate that excellent drug exposure can be accomplished with twice-daily administration. The observed complete contraceptive effect of JQ1 when given twice daily suggests that more continuous inhibition of BRDT-dependent effects on spermatogenesis may be required for clinical translation of this research.

EXPERIMENTAL PROCEDURES

(±)-JQ1
The direct-acting, small-molecule bromodomain inhibitor was prepared as described (Filippakopoulos et al., 2010).

Protein Cloning
cDNA-encoding BRDT (human: NCBI accession number AAB87862.1, obtained from FivePrime; murine: NCBI accession number EDL20168.1, obtained from Dr. Saadi Khochbin) was used as template to PCR amplify the N-terminal bromodomain regions of human and murine BRDT. PCR products were isolated and subcloned into a pMCSG7-derived expression vector (pNIC28-BsaI), using ligation-independent cloning. The constructs (hBRDT(1) 21–137, hBRDT(2) 257–382, mBRDT(1) 23–133, and mBRDT(2) 257–383) were transformed into competent Mach1 cells (Invitrogen, UK) to yield the final plasmid.

BRDT Proximity Assay
AlphaScreen assays were performed on an Envision 2104 instrument with minor modifications from the manufacturer’s protocol (PerkinElmer, USA).
Isothermal Titration Calorimetry

Isothermal titration calorimetry (ITC) was carried out on a VP-ITC titration microcalorimeter from MicroCal, LLC (Northampton, MA). Dissociation constants and thermodynamic parameters are listed in Figure 1.

Crystallization of BRDT and (+)-JQ1

BRDT(1) crystallizations were carried out using the sitting drop vapor diffusion method at 4°C using a mosquito crystallization robot (TTP Labtech, Royston, UK). Data collection and refinement statistics can be found in Table S2. The model and structure factors have been deposited with PDB accession code: 4FLP.

Mouse Contraceptive Studies

(+)-JQ1 was dissolved in DMSO at 50 mg/ml, 75 mg/ml, or 100 mg/ml and then diluted 1:10 in 10% (2-Hydroxypropyl)β-cyclodextrin (Sigma-Aldrich) or 20% Captisol (CYDEX, Lenexa, KS). The mixture was injected i.p. into male mice at 1% of the body weight (50 mg/kg or 75 mg/kg) or 0.5% of the body weight for the 50 mg/kg BID injections. The control was DMSO dissolved 1:10 in 10% (2-Hydroxypropyl)β-cyclodextrin or 20% Captisol and injected similarly. Juvenile or adult C57BL6/J129S5 hybrid mice for these studies were weighed daily before injections and fed ad libitum.

For mating studies, males were pretreated for 6 weeks with vehicle or JQ1 and caged consecutively with two females each while continuing 50 mg/kg QD for month 1 and escalating to 75 mg QD for month 2. The JQ1-treated males were responsive to this “low-dose” treatment regimen were allowed to recover untreated while continuing to breed to 2 females per cage through 7 months of continuous breeding. Males that continued to sire offspring in month 2 were treated in month 3 of breeding with 50 mg/kg BID (“high-dose” regimen), and JQ1 treatment was stopped for these males and controls while continuing to breed to 2 females per cage through 8 months of continuous breeding. Cages were checked daily, and offspring born per each female were recorded.

For the recovery studies, males were sacrificed post-JQ1 and control treatment, and testis and sperm parameters were determined. These studies were approved by the Administrative Committee on Laboratory Animal Care at Baylor College of Medicine, and all experiments were conducted in accordance with the NIH guide for the Care and Use of Laboratory Animals.

Histology and Measurement of Testis Seminiferous Tubule Area

Testes and epididymides were fixed in Bouin’s solution and processed for sectioning. Digital images of cross-sections were captured, and 25 tubules were randomly selected for each sample. Seminiferous tubule area was determined using NIH Image J software. The mean ± SEM of the tubule area for each treatment group was calculated, and comparisons between control and JQ1 treatment groups were made using a t test.

Epididymal Sperm Counts and Motility

Counts were performed on spermatozoa isolated from the entire epididymis or azoospermia in a large cohort of men of European descent. Hum. Reprod. 25, 1383–1397.

REFERENCES

Atkinson, R.J., Baker, M.A., Doncel, G.F., Matzuk, M.M., Mauck, C.K., and Harper, M.J. (2008). As the world grows: contraception in the 21st century. J. Clin. Invest. 118, 1330–1343. 358–368.

Dawson, M.A., Prinjha, R.K., Dittmann, A., Giotopoulos, G., Bantscheff, M., Chan, W.I., Robson, S.C., Chung, C.W., Hopf, C., Savitski, M.M., et al. (2011). Inhibition of BET recruitment to chromatin as an effective treatment for MLL-fusion leukemia. Nature 478, 529–533.

Delmore, J.E., Issa, G.C., Lemieux, M.E., Rahl, P.B., Shi, J., Jacobs, H.M., Kastritis, E., Gilpatrich, T., Paranal, R.M., Qi, J., et al. (2011). BET bromodomain inhibition as a therapeutic strategy to target c-Myc. Cell 146, 904–917.

SUPPLEMENTAL INFORMATION

Supplemental Information includes Extended Experimental Procedures, five figures, two data files, and seven tables and can be found with this article online at http://dx.doi.org/10.1016/j.cell.2012.06.045.

ACKNOWLEDGMENTS

We are grateful to Drs. J. Brown and J. Plutzky for pharmacodynamic data; Dr. A. Roy for advice on the spermatozoa assays; Y. Vasquez and Dr. Z. Yu for technical help; J. Marineau, K. Shaw, and W. Smith for support of peptide synthesis; Dr. S. Kistler for the generous gift of the anti-TNP2 antibody; and Dr. S. Krocobin for helpful comments. These studies were supported, in part, by the Eunice Kennedy Shriver NICHD/NIH through cooperative agreement U01-HD060496 as part of the Cooperative Program in Male Contraception; an Alkek Award for Pilot Projects in Experimental Therapeutics; and a grant from the Burroughs Wellcome Fund, the NIH (K08CA128972), the Smith Family Foundation, and Damon-Runyon Cancer Research Foundation (to J.E.B.). The SGC is a registered charity (number 1097737) that receives funds from the CIHR, Canada Foundation for Innovation, Genome Canada, GlaxoSmithKline, Pfizer, Eli Lilly, Abbott, Takeda, Novartis Research Foundation, Ontario Ministry of Research and Innovation, and Wellcome Trust. P.F. is supported by a Wellcome Trust Career Development Fellowship (095751/Z/11/Z). Serum FSH, LH, and testosterone analyses were performed at the University of Virginia Center for Research in Reproduction Ligand Assay and Analysis Core supported by NICHD (SCCPARR) grant U54-HD28934. Baylor College of Medicine and Dana-Farber Cancer Institute have filed patent applications on technologies relating to this work, which have been licensed to Tensha Therapeutics.

M.M.M. and J.E.B. designed the study, performed experiments, analyzed data, prepared the manuscript, and supervised the research. M.R.M. designed the BRDT biochemical assays, analyzed tissue histology, and analyzed data. P.F., S.P., and S.K. prepared purified protein, performed crystallographic studies, and analyzed data. Q.L., L.M., and J.E.A. performed mating studies and collected tissues for histologic, sperm, and RNA analyses. M.E.L. performed gene expression analyses. R.N.Y. performed developmental studies and sperm phenotyping. J.Q. performed the scaling synthesis of (+)-JQ1. All authors edited the manuscript.

Received: December 16, 2011

Accepted: June 30, 2012

Published: August 16, 2012
Dey, A., Chitsaz, F., Abbasi, A., Misteli, T., and Ozato, K. (2003). The double bromodomain protein Brd4 binds to acetylated chromatin during interphase and mitosis. Proc. Natl. Acad. Sci. USA 100, 8758–8763.

Dhalluin, C., Carlson, J.E., Zeng, L., He, C., Aggarwal, A.K., and Zhou, M.M. (1999). Structure and ligand of a histone acetyltransferase bromodomain. Nature 399, 491–496.

Dieterich, K., Soto Rifo, R., Faure, A.K., Hennebicq, S., Ben Amar, B., Zahi, M., Perrin, J., Martinez, D., Sèle, B., Jouk, P.S., et al. (2007). Homozygous mutation of AURKC yields large-headed polyoid spermatozoa and causes male infertility. Nat. Genet. 39, 661–665.

Filippakopoulos, P., Qi, J., Picaud, S., Shen, Y., Smith, W.B., Fedorov, O., Morse, E.M., Keates, T., Hickman, T.T., Felletar, I., et al. (2010). Selective inhibition of BET bromodomains. Nature 468, 1067–1073.

Katoh, K., and Toh, H. (2008). Recent developments in the MAFFT multiple sequence alignment program. Brief. Bioinform. 9, 286–298.

Kumar, T.R., Wang, Y., Lu, N., and Matzuk, M.M. (1997). Follicle stimulating hormone is required for ovarian follicle maturation but not male fertility. Nat. Genet. 15, 201–204.

Lin, Y.N., Roy, A., Yan, W., Burns, K.H., and Matzuk, M.M. (2007). Loss of zona pellucida binding proteins in the acrosomal matrix disrupts acrosome bio-membrane transitions during spermiogenesis in mice with transition nuclear protein Tnp1and Tnp2mutations. Proc. Natl. Acad. Sci. USA 104, 16669–16674.

Miki, K., Willis, W.D., Brown, P.R., Goulding, E.H., Fulcher, K.D., and Eddy, E.M. (2002). Targeted disruption of the Akap4 gene causes defects in sperm flagellum and motility. Dev. Biol. 248, 331–342.

Moliné, J., Rousseaux, S., Steuernwald, U., Soler-López, M., Curtet, S., Vitte, A.L., Govin, J., Gaucher, J., Sadoul, K., Hart, D.J., et al. (2009). Cooperative binding of two acetylation marks on a histone tail by a single bromodomain. Nature 461, 664–668.

Nicodeme, E., Jeffrey, K.L., Schaefer, U., Beinke, S., Dewell, S., Chung, C.W., Chandwani, R., Marazzi, I., Wilson, P., Coste, H., et al. (2010). Suppression of inflammation by a synthetic histone mimic. Nature 468, 1119–1123.

Roy, A., Lin, Y.N., Agno, J.E., DeMayo, F.J., and Matzuk, M.M. (2007). Absence of tektin 4 causes asthenozoospermia and subfertility in male mice. FASEB J. 21, 1013–1025.

Schultz, N., Hamra, F.K., and Garbers, D.L. (2003). A multitude of genes expressed solely in meiotic or postmeiotic spermatogenic cells offers a myriad of contraceptive targets. Proc. Natl. Acad. Sci. USA 100, 12201–12206.

Shang, E., Salazar, G., Crowley, T.E., Wang, X., Lopez, R.A., Wang, X., and Wolgemuth, D.J. (2004). Identification of unique, differentiation stage-specific patterns of expression of the bromodomain-containing genes Brd2, Brd3, Brd4, and Brdt in the mouse testis. Gene Expr. Patterns 4, 513–519.

Shang, E., Nickerson, H.D., Wen, D., Wang, X., and Wolgemuth, D.J. (2007). The first bromodomain of Brdt, a testis-specific member of the BET sub-family of double-bromodomain-containing proteins, is essential for male germ cell differentiation. Development 134, 3507–3515.

Wells, J.A., and McClendon, C.L. (2007). Reaching for high-hanging fruit in drug discovery at protein–protein interfaces. Nature 450, 1001–1009.

Wolfes, H., Kogawa, K., Millette, C.F., and Cooper, G.M. (1989). Specific expression of nuclear proto-oncogenes before entry into meiotic prophase of spermatogenesis. Science 245, 740–743.

Yang, J., Medvedev, S., Yu, J., Tang, L.C., Agno, J.E., Matzuk, M.M., Schultz, R.M., and Hecht, N.B. (2005). Absence of the DNA-/RNA-binding protein MSY2 results in male and female infertility. Proc. Natl. Acad. Sci. USA 102, 5755–5760.

Zeller, K.I., Jegga, A.G., Aronow, B.J., O’Donnell, K.A., and Dang, C.V. (2003). An integrated database of genes responsive to the Myc oncogenic transcription factor: identification of direct genomic targets. Genome Biol. 4, R69.

Zhao, M., Shirley, C.R., Mounsey, S., and Meistrich, M.L. (2004). Nucleoprotein transitions during spermiogenesis in mice with transition nuclear protein Tnp1 and Tnp2 mutations. Biol. Reprod. 71, 1016–1025.

Zuber, J., Shi, J., Wang, E., Rappaport, A.R., Herrmann, H., Sison, E.A., Magoon, D., Qi, J., Blatt, K., Wunderlich, M., et al. (2011). RNAi screen identifies Brd4 as a therapeutic target in acute myeloid leukaemia. Nature 478, 524–528.