Collagenase-2 (MMP-8) as a Point-of-care Biomarker in Periodontal Disease in Patients with or Without Fixed Prosthesis Therapeutic Response to Doxycycline

ANDRA AUNGURENCEI1, IONUT LUCHIAN2*, ANCUTA GORIUC3*, DANA CONSTANTINESCU1, IOANA MARTU1, DIANA DIACONU POPA1, ANCA VITALARIU1, DIANA LUCHIAN1, KAMEL EARAR6, MONICA TATARCIUC1

1University of Medicine and Pharmacy Grigore T. Popa Iasi, Faculty of Medical Dentistry, Department of Prosthetic Dentistry, 16 Universitatii Str., 700115, Iasi, Romania
2University of Medicine and Pharmacy Grigore T. Popa Iasi, Faculty of Medical Dentistry, Department of Periodontology, 16 Universitatii Str., 700115, Iasi, Romania
3University of Medicine and Pharmacy Grigore T. Popa Iasi, Faculty of Dentistry, Department of Biochemistry, 16 Universitatii Str., 700115, Iasi, Romania
4University of Medicine and Pharmacy Grigore T. Popa Iasi, Faculty of Dentistry, Department of Internal Medicine, 16 Universitatii Str., 700115, Iasi, Romania
5University of Medicine and Pharmacy Grigore T. Popa Iasi, Faculty of Dentistry, Department of Biochemistry, 16 Universitatii Str., 700115, Iasi, Romania

Collagenase-2 (MMP-8) as a Point-of-care Biomarker in Periodontal Disease in Patients with or Without Fixed Prosthesis Therapeutic Response to Doxycycline

Periodontal diseases ranges from simple gum inflammation to serious disease that consists in major damage to the soft tissue and bone teeth support. Periodontal diseases affects the marginal and apical periodontium and results from the interaction between bacterial biofilm and the host response. To determine the concentrations of MMP-8, as a disease marker, in saliva in prosthetic and nonprosthetic, aggressive (AP) and chronic (CP) periodontitis, doxycycline treated patients. 40 patients were distributed into 3 groups: 12 diagnosed with aggressive (AgP), 18 with chronic (CP) periodontitis. Each of these groups was subdivided into 2 subgroups with and without fixed prosthesis. 10 patients were in the normal group. Matrix metalloproteinase 8 (MMP-8) was evaluated before and after systemic doxycycline (Dox) treatment. Mean MMP-8 value into the control group was 0.57 ng/mL with a standard deviation (STD) of 0.094 ng/mL. Highest MMP-8 value was established for the nonprosthesis AgP subgroup, before Dox treatment. The highest reduction in MMP-8 levels (40.8%) was between nonprosthesis AgP before Dox treatment and the same group after Doxycycline treatment. MMP-8 saliva levels are lower than GCF levels, mostly through a dilution mechanism as previous studies had shown. Our study revealed that saliva MMP-8 level is reliable marker for AgP but not for CP. Doxycycline treatment, in terms of lowering MMP-8 levels is most effective in patients that have AP and are also wearing fixed.

Keywords: periodontal disease, fixed prosthesis, biomarker, doxycycline treatment

It has been established that periodontal disease results from the imbalance between oral microbiota with its antigens (necessary but not sufficient condition) on one side and the patient's mainly neutrophil and macrophage involved nonspecific inflammatory reaction (NIR) on the other [1-4].

NIR leads to alterations of the tooth supporting structures, most important alveolar bone and periodontal ligament, the major damage being represented by the lost of the type I collagen in the periodontium with matrix metalloproteinase (MMPs) mediated, osteoclastic, phagocytic and plasminogen dependent destructive pathways being well recognised [5-9].

The tests that are used in periodontal diagnosis, laborious and time consuming, are: probing depth (PD), clinical attachment level (CAL), bleeding on probing, plaque index and radiographic recordings and tend to express the consequences of the disease not the current status of the periodontium, these tests cannot predict the future destruction of the P or the response to treatment [10-13].

Neutrophil mainly but also other major human cell types (fibroblasts, keratinocytes, macrophages and endothelial cells) released MMPs, are a family of zinc dependent metalloproteinases, the only class of enzymes in mammals that can degrade collagens, MMP-8 and MMP-9 having the most collagen cleavaging potency and a steep correlation between periodontal disease and their concentrations in the gingival crevicular fluid (GCF) and doxycyclin or tetracycline at subantimicrobial doses as a pharmacological ribosomal 30S unit level inhibitor and also being considered useful indicators for the severity of NIR [14,15].

In the domain of periodontal diagnostics, oral fluid-based biomarkers have been studied mainly in the gingival crevicular fluid (GCF) and saliva [6,7,16,17].

Gingival crevicular fluid (GCF) is a variant of serum transudate secreted by the gum in small amounts in healthy tissue and large quantity in periodontal disease that contains components of connective tissues, epithelial cells, bacteria, cytokines, enzymes, used for periodontal disease diagnosis and its treatment monitoring [18].

Saliva provides an easily available, noninvasive diagnostic tool for a rapidly widening range of diseases and clinical situations. Saliva, as a mirror of oral and systemic health, is a valuable source for clinically relevant information because it contains biomarkers specific for

* email: ionut_luchian@yahoo.com; ancuta.goriuc@yahoo.com In this paper, all authors have an equal contribution as the first author.
the unique physiologic aspects of periodontal diseases [3,6,7].

The mainstay for periodontitis treatment is represented by the mechanical debridement followed by local or systemic use of antibiotics, doxycycline (Dox) at just 20 mg twice daily dose inducing a significant reduction in neutrophil collagenase GCF concentrations, with the local administration route being the best way in terms of gastrointestinal side effects, microbial antibiotic resistance, patient cooperation and other medication interaction [19].

Dental prosthesis (dental bridge) remains one of the most important treatment procedure with well recognised benefits in periodontal disease through the mechanism of improving the teeth mobility [20-24].

Our study focused on saliva MMP8 levels surveyed in patients with aggressive (AP) and chronic periodontitis (CP) before and after Dox treatment.

**Experimental part**

**Mathenial and method**

The study was conduced in Grigore.T.Popa University, the Periodontology Clinic. Patients saliva samples were procesed in to the Saint Spiridon Hospital Laboratory. The study included patients that adressed the Periodontology Clinic in the 2018-2019 period.

Three main groups were established. The first group included 12 patients with aggressive periodontitis (AgP) diagnosed according to clinical criteria: pocket depths over 4-5 mm, gingival bleeding on probing and radiological signs that proved osteolysis. In this group, 2 subgroups were established one of eight patients with fixed prosthesis, the second one of 4, without fp. Specific AgP exclusion criteria were: chronic periodontitis (CP) present and previous 6 months antinflammatory treatment.

The second group included 18 patients with CP diagnosed according to clinical criteria: pocket depths over 6 mm, gingival inflamation, radiological proven horizontal osteolysis. In this group, 2 subgroups were established one for 10 patients with fixed prosthesis and another one for 8 patients without. Specific group inclusion criteria: age over 35, more than 20 teeth present, no severe parodontitis episode present. Specific group exclusion criteria: presence of severe periodontitis.

Third group included 10 patients with normal clinical peridontal status.

General inclusion criteria for the patients were: the signing of informed consent, chronic (CP) or AgP present according to criteria.

General exclusion criteria were: smoking habit present, other chronic disease present, antibiotic treatment 3 months prior to the study, pregnancy, periodontal treatment 6 months prior to the study, allergies.

Fixed prosthesis patients were alreadly wearing their fp when entering the study, no fp was applied as treatment during the study.

The study’s purpouse was to determine and compare MMP-8 saliva concentrations in groups and subgroups of patients before and after Dox treatment.

MMP-8 levels, for each patient, were evaluated using Elabscience provided ELISA testing kit before and after Dox treatment.

Saliva was sampeled from all patients in Eppendorf tubes, centrifuged for 2 minutes at 1000 rpm, with the resulting supernantant being between 400 µl and 2000 µl. ELISA was performed using the supernatant to determine the concentration of MMP-8. Reactives used were adjusted to room temperature before use. Standard solution was achieved by adding 1 mL of reference standard diluent on the human liophilised standard MMP8 left for 10 minutes incubation time at room temperature. Eight standards were prepared at halfed concentrations starting from 10 ng/mL, seventh dilution at 0.16 ng/mL and the eight being blank. In each ELISA plate well 100 microL from standard and probes were added. ELISA plate was then sealed and incubated at 37 C for 90 minutes. 100 microLof substrate solution 0.01 dilution was added in each cell followed again by sealing and incubation for 60 minutes at 37 degrees. Buffer solution was than used for plate washing. 3 times at 1 minute interval. 100 mL of enzym Hrp (horseradish peroixdaze conjugate) in each ELISA plate well, followed by incubation at 37 degrees celsius for 30 minutes. Aspiration of each ELISA plate well with buffer solution, 5 times at 1 minutes interval. 90 microLof substrate solution was then added in each ELISA plate well and than incubated for 15 minutes at 37 degrees celsius. Coloration was observed. Color intensity dictated the necessity of increasing or decreasing the incubation time up till 30 minutes. 50 microLof Stopping solution was than added in each ELISA plate well. Absorbance was then read at 450 nm using a ELISA plate reader ELISA BIO-RAD and Magellan software was used for analysis.

After saliva prevaleation, all patients had conventional periodontial therapy consisting in removal of dental plaque and calculus, followed by smothing and planing of exposed surface of the roots in teeth with deep periodontal pokets.

Doxycycline was administered oraly in subantibacterial concentrations of 20 mg twice daily for 7 days. No adverse effects were aknowledeged following antibiotic treatment.

Reevaluation of patient after Doxycycline included all examinations that were made at the begining of the study: periodontal status evaluation clinic and paraclinic. Saliva was resampeled and same assay protocol used for MMP-8 evaluation.

**Results and discussions**

All patients enroled completed the study. Patients were distributed into 3 main groups: AgP CP and normal (control group). Each of the first two groups were split into with and withoutfp. All 4 subgrups thus obtained were further analysed before and after administration of Dox. Doxycycline was not administered to the control group.

Average, standard deviation and significance tests were performed on groups and subgroups. Intrasubgroup variation was analysed between values of MMP-8 before and after Dox administration. Mean value of MMP -8 in control group was 0.257 ng/mL with standard deviation of 0.094 ng/mL and is the group that by far had the best concentration of values around the average.

Highest average MMP-8 value has been aknowledged in the AgP without fixed prosthesis subgroup and before Dox treatment (mean value: 14.562 ng/mL, SD: 2.651 ng/mL) followed by the values, in the same subgroup after Dox treatment (mean value: 9.320 ng/mL, SD: 1.708 ng/mL). Lowest average MMP-8, was in the fixed prosthesis CP subgroup after Dox treatment (mean value: 1.681 ng/mL, std: 0.904 ng/mL).

The highest MMP-8 level variation was in the fixed prosthesis AP subgroup, before and after Dox treatment with a drop of 40.8%from the initial MMP-8 level. The lowest variation between before and after Doxycycline treatment was in the non prosthesis CP subgroup (a 14.1% drop in MMP8 levels) (see Table 1). There was also significant MMP-8 level difference in the CP group between prosthetic and without prosthesis patients (mean value: 1.986 ng/mL and 5.938 ng/mL respectively).

Because the MMP-8 assay was conducted on saliva instead of GCF the values (expressed in ng/mL) were lower.
than the average values in GCF in other published findings through a dilution mechanism most likely [25-27].

When statistical analysis was made for AgP fp subgroup, a statistical significant difference was established between before and after Doxycycline treatment average MMP-8 values (6.04 ng/dL and 3.57 ng/dL respectively) and control group, p<0.001.

The same statistical analysis was made for AgP non fp subgroup, significant difference was established between before and after Dox treatment average MMP-8 values (14.56 ng/dL and 6.039 ng/dL respectively) and control group, p<0.001.

Statistical analysis on CP fixed prothesis subgroup did not establish significant difference between before and after Dox treatment average MMP-8 values (1.98 ng/dL and 1.68 ng/dL respectively) (p=0.38); same analysis made in CP non fixed prothesis subgroup also did not establish significant difference between before and after Doxycycline treatment.

| Groups and subgroups | Average (ng/mL) | Standard Deviation (ng/mL) | In Group variation (the drop in value between before and after Dox treatment) in % |
|----------------------|-----------------|----------------------------|----------------------------------------------------------------------------------|
| C                    | 0.257           | 0.09                       |                                                                                  |
| AgP                  | 6.015           | 1.915                      |                                                                                  |
| A                    | 14.562          | 2.851                      |                                                                                  |
| C                    | 1.986           | 1.057                      |                                                                                  |
| C                     | 1.811           | 0.904                      |                                                                                  |
| C                    | 5.358           | 1.297                      |                                                                                  |
| CDox                 | 2.087           | 1.115                      |                                                                                  |

C-control group; AgP-aggressive periodontitis (AgP) with fixed prothesis (fp) before doxycycline treatment (Dox); A-AgP without fp before Dox; Cfp-Chronic periodontitis (CP) with fp before Dox; CfpDox-Chronic periodontitis (CP) with fp after Dox; Co-control group
treatment average MMP-8 values (5.94 ng/dL and 5.09 ng/dL respectively) (p=0.25). This may be because CP as a whole has different lesions dominant triggering pathways than MMP-8 mediated.

When statistical analysis was performed between fixed prosthesis and non fixed prosthesis patients it has been established that wearing fp significantly decreased mean MMP-8 values (p<0.001) both in AgP and CP groups before and after Dox treatment.

From the point of view of the doxycycline administration impact, only in AgP group Dox treatment significantly lowered the mean MMP-8 levels (p<0.001) while in CP group it didn’t. Doxycycline treatment should be associated with fixed prosthesis treatment for maximum therapeutic effect.

Regarding host modulation therapies Doxycycline is one many possibilities, acting on one component of the inflammation cascade, on matrix metalloproteinases, however, another class of immunomodulatory drugs are DMARDS, which influence the release and action of certain biomolecules and cytokines such as IL 1, IL-6, IL 17, TNF alfa [28].

However, the detection of salivary MMP-8 can be considered a biomarker of periodontitis and could be used as a valuable indicator of health and pathologic process in patients with and without fixed prosthesis.

Conclusions
Salivary MMP-8 represents a reliable marker for AgP but not so reliable for CP. Doxycycline treatment administration is most effective in patients that have AgP and are also wearing fixed prosthesis (dental bridges) and it is well proven through MMP-8 levels.

References
1. GENCO, R.J., Host responses in periodontal diseases: Current concepts. J Periodontol 1992;63(4 Suppl):338-55.
2. LUCHIAN, I., MARTU, I., MARTU, C., GORIUC, A., BELDIMAN, A., MARTU, S. Rev Chim. (Bucharest) 67, no. 6, 2016, p. 1073-1075.
3. NITESCU-KAPPENBERG, D.C., MIHAI, C., OANTA, C., MARTU, I., VOLOVAT, S.R., MARTU, S. Rev. Chim. (Bucharest), 68, no. 3, 2017, p. 549-552.
4. GEORGESCU, M., VRINCEANU, D., RADULESCU, L., TUSALIU, M., MARTU, C., CURUTIU, C., HUSSIEN, M.D., BUDU, V. et al. Rom Biotechnological Letters, 22, no. 4, 2017, p. 12681-12686.
5. PAGE, RC, J Periodontal Res 1991; 26(3 Pt 2):230-42.
6. SUFARU, I.G., SOLOMON, S.M., PASARIN, L., MARTU-STEFANACHE, M.A., OANTA, A.C., MARTU, I., CIOCAN PENDEFUNDA, A., MARTU, S. Rom J of Oral Rehab, 8, no. 1, 2016, p. 97-103.
7. MURARIU, A., FORNA, D.A., MANOLACHE, F., FORNA, N.C. Rom. J of Oral Rehab, 11, no. 2, 2019, p. 274-278.
8. NICOLAE, V., CHISCOP, I., CIORANU IBRIC, V.S., MARTU, M.A., SIUSTIS, I., IOVAN, A., MARTU, S. Rev Chim. (Bucharest), 66, no. 12, 2015, p. 2121-2123.
17. SOLOMON, S., URSARESCU, I., MARTU, A., LUCHIAN, I., AGOP-FORNA, D., MARTU, S., FORNA, N.C. Rev. Chim. (Bucharest). 66, no. 8, 2015, p. 1166-1168.
18. KINNEY, J.S., MORELLI, T., OH, M., et al. J Clin Periodontol. 2014, 41(2):113-120.
19. FARAMARZI, M., MARAMI, Z., SHIRMOHMMADI, A., CHITSAZI, M., RAHBAR, M., SADIGHI, M., J. of Int. Oral Health 2016; 8(7):781-786.
20. TATARCUIUC, M., VITALARIU, A., LUCA, O., AUNGURENCEI, O., FRATILA, D., DIACONU-POPA, D. Rev. Chim. (Bucharest). 69, no. 2, 2018, p. 407-410.
21. DIACONU-POPA, D., VITALARIU, A., HOLBAN-CIOLOCA, C., AUNGURENCEI, A., LUCA, O., TATARCUIUC, M. Rev Chim (Bucharest) 68, no. 10, 2017, p. 2382-2385.
22. DIACONU, D., TATARCUIUC, M.S., MELINTE, A., VIPALARIU, A.M., Rom. J. of Oral Rehab. 5, no. 3, 2013, p. 84-90.
23. BOSINCEANU, D.G., SANDU, I.G., BOSINCEANU, D.N., MARTU, I., SURLARI Z., FORNA, N.C. Mat. Plast., 55, no. 3, 2018, p. 423-425.
24. MARTU, I., LUCHIAN, I., DIACONU-POPA, D., BOSINCEANU, D.G., VITALARIU, A., LUCA, O., TATARCUIUC, M. Rom. J. of Oral Rehab. 9, no. 1, 2017, p. 27-31.
25. SURLIN, P., OREA, B., SOLOMON, S.M. et al. ROMANIAN JOURNAL OF MORPHOLOGY AND EMBRYOLOGY Rom. J. of Morphology and Embryology, 55, no. 3, suppl. S, 2014, p. 1137-1141.
26. SURLIN, P., RAUTEN, A.M., MATEESCU, G.O. et al. Rom. J. of Morphology and Embryology, 50, no. 2, 2009, p. 181-184.
27. SURLIN, P., RAUTEN, A.M., MOGOANTA, L., et al. Rom. J. of Morphology and Embryology, 51, no. 3, 2010, p. 515-519.
28. MARTU, M.A., REZUS, E., POPA, C., SOLOMON, S.M., LUCHIAN, I., CIOCAN PENDEFUND, A., SIOUSTIS, I., ANTON, D., MARTU, S., FOIA, L. Rom. J. of Oral Rehab. 10, no. 4, 2018, p. 161-165.

Manuscript received: 6.11.2019