The Effect of Betel Quid Extract on Wound Healing Process in Male Wistar Rats (*Rattus norvegicus* L.)

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ABSTRACT— Wound can be occurred during dental treatment. In outline, there are several phases of wound healing; inflammatory phase, proliferative phase, and maturation phase. Various drugs in various forms are used to accelerate the healing process, but mostly they have side effects. Therefore, traditional medicine, such as betel quid consisted of betel leaf, areca nut, gambier, and calcium hydroxide, is developed. The aim of this study was to investigate the effect of wound healing process in male Wistar rats. A total of 30 male Wistar rats were taken and divided into 5 groups: Group 1, 2, and 3 (K1,K2, K3) were given 5%, 10% and 15% concentrations of betel quid extract oinments; Group 4 (K4) was positive control (hyaluronic acid 0.2% oinment); Group 5 (K5) was negative control (placebo oinment). One mm-diameter of lower lip mucosal wounds on rats were created by using cylinder diamond bur. The oinments were applied twice daily for 10 days. The number of neutrophils on first and third day were measured and the thickness of epithelium on 10 days were determined. All groups of betel quid extracts exhibited the reduction of the number of neutrophils on inflammatory phase. Group 3 shown as the highest effect and had no significant different with positive control. Betel quid extracts in all groups also improved epithelial thickness on proliferative phase, in which group II and group III had no significant different with positive control. Betel quid extract had effect on wound healing process in male Wistar rats due to its ability in supressing inflammation and in increasing reepithelization.

Keywords— anti-inflammatory, betel quid, epithelial thickness, wound healing

1. INTRODUCTION

Wounds or injuries to the oral cavity often occur due to some dental procedures such as scaling, root planning, and trauma. Wound is a disruption of the cellular and anatomical tissue continuity resulting from physical, mechanical, chemical, thermal, or radioactive substance [1,2]. Those wounds or injuries stimulate biological process as a response through a complex and dynamic recovery process causing continuous recovery of anatomy and function called wound healing [3,4]. A completely healed wound is interpreted that the tissue has been backed to normal anatomical structure, functional, and its appearance [5].

Basically, the wound healing process is divided into several phases; inflammatory phase, proliferative or fibroplasial phase, and remodelling or maturation phase [6]. The inflammatory phase is characterized by hemostasis [4]. Hemostasis refers to platelet aggregation and platelet plug formation [7]. Platelet activity is linked to the initiation of clotting cascades. The aggregation of thrombocytes and platelets in fibrin network forms coagulation. This activity associated with minimizing blood loss [8]. Insoluble fibrin generated by proteolytic coagulation cascade stabilizes the blood clot.7 In another site, wound and activation of clotting factor induces the release of inflammatory mediators such as thromboxane A2, prostaglandin, histamine. Those mediators promote clinical signs of inflammation, such as rubor, kalor, dolor, tumor and functiolaesa [9,10]. After several hours of wounding, neutrophils appear into injury’s area. These phagocytic cells have responsibility in debris scavenging, killing bacteria, and decontaminating the wound from foreign organism. Neutrophils eliminate microbes and dead cells through phagocytosis, or by releasing free radicals. Neutrophils perform the process of marginization, adhesion, and migration before starting their phagocytosis function [11,12]. Neutrophils together with another anti inflammatory cells induce angiogenesis in proliferative phase.
Proliferative phase begins on the third day of injury and ends on the 14th day thereafter. This phase is responsible for the closure of wound. It is characterized by the process of angiogenesis, fibroplasia, and reepithelization [5,13]. Keratinocytes were migrated in a few hours after injury. Keratinocyte promotes protein secretion to produce basement membrane, stimulate reepithelization and wound closure [14]. Nutrients and oxygen are needed in wound repair, so that formation of blood vessels or angiogenesis are induced [15]. Cytokines, produced by neutrophils, together with growth factors (PDGF, TGF-b, bFGF), present the migration of fibroblast. Fibroblasts proliferate and form granulation tissue [16]. The tissue will complete wound closure, provide cell adhesion, proliferation and differentiation during wound healing [14,16]. The last phase of wound healing is remodeling or maturation phase, which starts from two weeks after lesion up to one year depending on the severity of the wound. The phase is marked by the maturing of extracellular matrix through restructuring, degradations and reshynthesis of matrix [6]. Extracellular matrix has many functions, such as delivering cytokines and growth factor, tissue repair, controlling morphogenesis, regulating cell activities and bioavailability [17,18].

Many drugs are used to heal the wound, such as topical hyaluronic acid. Dechet et al. stated that 0.2% hyaluronic acid gel has important role in regulating the cell proliferation. However, some adverse effects may occur, such as allergic reaction, tissue necrosis, infection, and etc [20]. Therefore, natural substances as alternative medicine are develop as substitute, i.e betel quid.

Chewing betel quid has been commonly practiced by many people all over the world, like Indonesia, Malaysia, India, Bangladesh, and Taiwan. Chewing betel quid habit has many impacts on social, cultural, religious and economic life, and received special attention in the past. In addition, people used to believe that it can strengthen their teeth and relieve toothache. Betel quid is a mixture of areca nut (Areca catechu L.), gambier (Uncaria gambir Roxb.), mineral slaked lime (calcium hydroxide) wrapped in betel leaves (Piper betle L.) [21]. All ingredients of betel quid are capable of accelerating wound healing process.

Betel leaf has long been used as a household remedy for oral mucosal inflammation [22]. Previous study has been reported that betel leaf has active compound including essensial oil (hydroxyl chavicol, eugenol, chavicol, chavibethol, estragol, terpene, sequiterpene, triterpenoid, β-cytosterol) and tannin [23]. Lin et al mentioned that phytochemical compounds contained in betel leaf was effective to inhibit inflammatory reaction and stimulate reepithelization process on wound healing [24].

The areca nut is the dark red seed of Areca catechu that has reported to have several therapeutic properties, including analgesic, anti-inflammation and antioxidant, and has a role in wound healing. The areca nut extract contains procyanidin, the main condensed tannin which widely disperses to the polyphenolic compounds and has pharmacological effects [25]. Vonna et al mentioned that the ethanol extract of areca nut had wound healing activities in Mus musculus albinus mice [26].

Gambier plants (Uncaria gambir Roxb.) also have a role in wound healing, i.e antioxidant, antiseptic, anti-inflammation, and cell proliferation, as they have the main content of catechin [27-31]. In addition, gambier contains polyphenol groups such as alkaloids, terpenoids, flavonoids, and other polyphenolic compounds. Another components contained in the gambier are catechu tannat, pyricatechol, red catechu, quercetin and fixed oil. The content of flavonoid takes part on wound healing process by increasing collagen formation, decreasing macrophages and tissue edema and increasing the amount of fibroblasts [32].

Mineral slaked lime has chemical formula of CaOH (calcium hydroxide). Calcium hydroxide has been shown to have antimicrobial properties and reduce periapical inflammation, so it is increasingly being used. Calcium hydroxide paste has a high pH of 12.5 to 12.8. The high alkaline nature of the calcium hydroxide is able to neutralize the acid (buffer), reduce the inflammatory reaction and stimulate healing [31,33,34].

The purpose of this study was to determine the effect of wound healing extract of betel quid on male rats Wistar strain. Wound healing process was investigated by evaluating the anti-inflammatory effect and reepithelization thickness of wound in vivo.

2. METHODOLOGY

The study was true experimental in vivo with pretest-posttest control group design design to know anti-inflammatory effect and posttest only control group design to know the effect of reepitileization on wound healing of betel quid extract. The study protocol had been approved by Research Ethical Commission of Mohammad Hoesin General Hospital (RSMH) Palembang and Medical Faculty of Sriwijaya University with ethical certificate No. 391/kepkrsmhfkunsri/2017.

Animals

The study was carried out using Wistar ((Rattus norvegicus L.) rats weighing 150-200 grams, 8-12 weeks old, obtained from Pharmacy School of Bandung Institute of Technology (SF-ITB), with certificate no. 519.2/3691-Dispangtan/2017. Rats were raised in the Animal House of the Medical Faculty of Sriwijaya University. They were maintained under standar housing conditions.
conditions at room temperature of 20\(^\circ\)-25\(^\circ\) C with dark-light cycle 12h/12h. The animals were acclimatized to laboratory condition for eight days before study [35]. During acclimatization process, rats were fed with standard pellet diet and water ad libitum.

Thirty male Wistar rats were divided into 5 groups, Group I (K1) was treated with 5% betel quid extract ointment, Group II (K2) was treated with 10% betel quid extract ointment, Group III (K3) was treated with 5% betel quid extract ointment, Group IV (K4) was positive control, treated with 0.2% hyaluronic acid ointment (purchased from Ricefarma Pharm.Co, Surabaya, Indonesia), and Group V (K5) was negative control, treated with placebo ointment.

**Plant materials**

Betel quid components, including betel leaf (*Piper betle* L.), areca nut (*Areca catechu* L.), gambier (*Uncaria gambir* Roxb.) and mineral slaked lime (calcium hydroxide) were collected from Babatoman Village, Sekayu Subdistrict, Musi Banyuasin District, South Sumatra Province, Indonesia. All the components and material were identified and authenticated by Faculty of Agriculture, Sriwijaya University, Indonesia.

**Extraction of betel quid**

Betel quid was extracted by using soxhletation method. Betel quid components consisting of 8 g of betel leaf, 3.5 g of areca nut, 2.5 g of gambier and 2 g of betel lime were mixed and were placed in thimble made from cellulose. Soxhlet apparatus was filled with 96% ethanol solvent at ratio of 1:4 and heated at temperature of 50\(^\circ\) C for five hours. The extract was filtered and concentrated in vacuum reduced pressure using rotary flash evaporator (IKA RV10, Staufen, Germany) for 48 hours, until solvent had been removed.

**Preparation of ointment**

Betel quid extract was formulated as ointments by different concentration of 5%, 10% and 20%. The dose given to experimental rats was 50 mg. The basic materials for ointment were lipid based material consisted of 15% *adeps lanae* and 85% *vaselin album*. The base was prepared by weighing accurately. The hot mortar and pestle was taken out from oven (Cole-Palmer Ltd, UK) and adeps lane was poured into mortar and stirred by using pestle until melt. Subsequently, vaseline album was poured into mortar and was stirred by using pestle gently until homogenous and form ointment base. Betel quid extract was added according to formulation of respective groups and was stirred until homogenous. In preparing placebo ointment, the base was taken and treated in the same manner to formulate ointment without any active ingredients. Ointment extract was flattened on surface of glass base for subsequent homogenity test. An ointment was considered homogenous if particles mixture was evenly distributed. Homogenous ointment preparations extract was put into ointment pot and was labeled according to treatments.

**Preparation of 1% carrageenan**

0.05 grams of carrageenan was weighed and suspended in 5 ml of 0.9% NaCl solution in the measuring flask.

**Induction of mucosal wound**

Lower lip mucosal wound was induced by using 1-mm diameter of cylinder diamond bur (Microdont, USA). Prior to wound induction, animals were anesthetized with 0.2 mL ketamine by i.m. injection. The lower lip of rats was withdrawn by using tweezer (Fisher brand\textsuperscript{TM}, Thermofisher Co, UK) and was swabbed with aquadest-wetted sterile cotton. Mucosal wound was made by using cylinder diamond bur at depth of about 1 mm in accordance to bur diameter used in this process. Blood was cleaned with aquadest-wetted sterile cotton and dried. Carrageenan was injected intraorally in another side of lower lip of rat. Five hours after injection, blood samples were taken and sent to Province’s Health Laboratory of South Sumatera, Indonesia, to count the number of neutrophil.

**Topical wound application**

Ointments were applied respectively by using sterile cotton bud on lower lip mucosal wound of rats twice daily, every 12 hours, and lasts for 10 days. On the first day, ointments were given immediately after induction of wound. To avoid contact with the wound, rats were isolated from eating and drinking for 1 hour after treatment.

**Counting the number of Segmented neutrophil cells**

The number of neutrophil cell was performed using a hematology analyzer (Yumizen H500\textsuperscript{®}, Horiba Medical Eqiupment Co., Japan). Blood samples were taken on the first day post induction of carrageenan and the third day, after 6 times application of ointments for 3 days. Blood samples were taken from sinus orbita of rat eyes, then placed on a tube containing anti-coagulant
ethylene diamine tetra acetic acid (EDTA) purchased from Province’s Health Laboratory of South Sumatera, Palembang, Indonesia. Furthermore, the number of segmented neutrophil cells in rat blood were calculated.

**Histological Assessment**

Wistar rats were euthanized by using chloroform in the 11th days. The mucosal wound were taken by using surgical scissor (Surgipro Inc, Germany). The specimens was fixed in 10% formalin solution and stained with haematoxylin and eosin (H and E) prepared by Anatomical Pathology Laboratory of Dyaat Natalis, Palembang, Indonesia. To reduce subjective bias, all assessment were made with histological slides coded. Epithelial thickness of all samples was evaluated by a histo-pathologist.

**Microscopic assessment**

The specimens were observed through optical microscope with 400x magnification in 5 fields of view. The observed structure was the epithelial layer from stratum corneum to stratum basal. The pictures of specimens was taken by using Micro-ocular MD130 electron eyepiece (Ome-Top System Co., Ltd, Taiwan). The newly formed epithelial thickness from the picture was measured and analyzed.

**Statistical analysis**

The data of the neutrophil cell numbers and reepithelization thickness were collected and analyzed statistically using SPSS vers 22 (IBM® inc.pvt ltd, US). Both data were performed with Levene’s test to know the homogeneity of samples and normality test of Shapiro Wilk to know the distribution of samples. P value of > 0.05 was considered that data were homogen and normal. To compare the changes of the neutrophil cells between before and after study, we used paired t-test. One-way Anova was used for analyzing the significant differences in all groups, followed by Tukey’s post hoc test for multiple comparisons. The p-value less than 0.05 was regarded as statistically significant.

### 3. RESULTS AND DISCUSSION

**Evaluation of the number of neutrophil cells in the inflammatory phase of wound healing**

Normality and homogeneity test had been carried out and the result showed p value>0.05, so it meant that the data were normally distributed and homogen. The mean of neutrophil cells in betel quid groups after treatment reduced significantly compare to before treatment (table 1). The higher concentration of betel quid extract, the lower number of neutrophil cells in blood. The lowest number of neutrophils cells was exhibited in hyaluronic acid group. The decreasing on the number of neutrophil was also shown in placebo. It was happened because on the 3rd day, neutrophil cells began to decrease and were replaced by macrophages on the 4rd day.

| Table 1. Comparison of the number of neutrophils before and after treatment |
|-----------------------------|-----------------------------|-----------------------------|
| **Group**                  | **Means of neutrophil cells (%)** | **P Value** |
|                            | Pre-test                     | Post-test                   | *
| K1 (5% betel quid extract) | 28.83 ± 3.66                 | 16.83 ± 3.06                | 0.003* |
| K2 (10% betel quid extract)| 29.17 ± 5.38                 | 14.33 ± 3.14                | 0.000* |
| K3 (20% betel quid extract)| 29.50 ± 2.58                 | 9.17 ± 3.60                 | 0.000* |
| K4 (positive control)      | 29.67 ± 5.46                 | 6.33 ± 3.14                 | 0.000* |
| K5 (negative control)      | 28.17 ± 7.73                 | 19.00 ± 6.93                | 0.002* |

*Paired t test, p=0.050

Result of one-way ANOVA of p value<0.000 implied that betel quid extract had significant effect in reducing the number of neutrophils, so it was able to accelerate the wound healing process.

Post Hoc Tukey’s were done to compare betel quid extract with positive and negative control groups. Table 3 showed that 20% betel quid extract had ability in decreasing neutrophil cells compare to negative control and had no significant difference with positive control group (0.2% hyaluronic acid).
Figure 1. The number of neutrophils. Betel quid extract of 20% concentration displayed anti-inflammatory effect significantly and data were expresses as mean ± SD, *p<0.05 versus positive control group; **p<0.05 versus negative control group.

**Evaluation of epithelial thickness on proliferation phase of wound healing**

Normality and homogeneity test were done, and the results showed p>0.05. That meant data were normal and homogen. One-way ANOVA was performed and the result was p value of <0.05.

Figure 2. Epithelial thickness of wound. Betel quid extract of 10% concentration stimulated re-epithelization significantly and data were expresses as mean ± SD, *p<0.05 versus positive control group; **p<0.05 versus negative control group.
The epithelial thickness of wound assay was used to evaluate the epithelial cell production in the proliferation phase on wound treated with 5%, 10% and 20% betel quid extracts. Re-epithelization of wound increased at concentration of 10% and 20% when compared to negative control (p value<0.05) and had no significant difference with positive control. It revealed that 10% betel quid extract had similar proliferative effect to 0.2% hyaluronic acid.

From the results, it has been proved experimentally that betel quid extract had potential effect in accelerating wound healing process. The effectiveness of betel quid extract was according to our findings that the extract had ability in reducing the number of neutrophils on the 3rd day of wound and boosting the re-epithelization on the 11th day of wound. The role of extract in inflammation and proliferation phase hastened the wound healing.

20% betel quid extract had the same effectiveness as 0.2% hyaluronic acid in decreasing inflammatory cells. The reduction of the cells expedited the healing to the next phase, namely proliferation phase. This study also revealed that after 11 days, re-epithelization of 15% betel quid extract was similar to 0.2% hyaluronic acid on mucosal wound. The presence of new epithel on 20% betel quid extract was thicker than 0.2% hyaluronic acid. The findings underlined the ability of betel quid extract to accelerate the proliferation contributing to fast the healing process.

The effect of wound healing is due to its phytoconstituents of betel quid extract. Betel quid are consisted of betel leaves (Piper betle L), areca nut (Areca catechu), gambier leaves (Uncaria gambier) and calcium hydroxide. Previous study has explained that each compositions had their own active compound that help in stimulating wound healing process. These findings exhibit that all ingredients of betel quid has worked sinergistically for the healing process.

Piper betle contained in betel quid possess anti inflammatory properties, such as eugenol and hydroxychavicol [36]. Eugenol is potential as an anti inflammation, because it suppresses the expression of cyclooxygenase-2 enzyme (Cox-2), TNF signaling, cytokines expression and production, and NF-kappaB. Eugenol inhibits prostaglandin synthesis and neutrophil chemotaxis [37,38]. Pin et al evaluated anti inflammatory activities of extract Betel leaves by using hyalurondiase (HYA), xantine oxide (XOD) and lipoxygenase (LOX) inhibition assays, which plays important role in the leukotriene biosynthesis pathway and biological regulator systems of inflammatory disorders. The results showed that Betel leaves extract represented high inhibition in XOD and LOX assays. Those activities were related to eugenol and hydroxychavicol contained in Betel leaves [39]. Eugenol in Piper betle has ability in inhibiting leukotrien C4 (LTC4) formation in the pathophysiology of inflammation [40]. Hydroxychavicol (HC) found in Piper betle is also potential as an anti inflammatory agents. HC induces the inhibition of COX-1/COX-2, the phrase of the pro inflammatory cytokines TNF-a, platelet calcium signaling, and ROS scavenging [33,41].

In proliferation phase, Piper betle contained in betel quid increases hydroxyproline content, a major part of collagen and essensial in collagen stability. Ghazali et al reported that Piper betle leaves extract improved re-epithelization, fibroblast, collagen synthesis and deposition, and exhibited the decreasing of the oxidative stress markers [42]. Keat et al investigated that Betel extract increased wound contraction rate and protein content and created wound closure [43]. Lien et al stated that Piper betle Linn extract promoted wound healing by stimulating NIH3T3 fibroblast proliferation [44]. Nilugal et al reported that Piper betle leaves and stem extracts enhanced the rate of wound contraction and reduced healing time, highlighted by the full thickness coverage of organized tissue and collagen fibers on the wound [45].

Another composition of betel quid is Areca nut. Areca nut grows in several country such as Indonesia, Malaysia, India, Bangladesh, China, etc. Traditionally, areca nut are used for some medical use [46,47]. The major chemical properties of Areca nut are polyphenols (flavonoids and tannin), polysaccarides, proteins, fats, fibers, alkaloids, minerals, vitamin B and C [48]. The anti inflammatory effect of areca is related to the inhibition of cyclooxygenase-2, the reduction of the number of neutrophils, the release of prostaglandins (PGEs), IL-1, IL-6, leukotrienes, histamins, the suppression of T cell activities, and the induction of the release of IL-4 [46,49,50]. Areca nut also consists of procyanidin, a condensed tannin from flavonoid group, that plays important role in hampering NFkB, phospholipase, lipoxygenase, and cyclooxygenase [48,51]. Anthikat and Michael reported that Areca catechu was significantly inhibited oedema, induced by chronic inflammatory agents [52]. Areca nut posses antimicrobial constituents that keep the wound sterile, so that the wound healing process will be hastened. Alkaloids and polyphenols in areca nut accelerates wound healing by increasing wound contraction and breaking strength, and also enhancing hydroxyproline level in tissue granulation [53]. Verma et al revealed that Areca catechu promoted burn wound healing, by increasing wound contraction, reepithelization, as well as dexamethason [54]. Bharat et al also stated that Areca catechu was effective in expediting of wound healing process [55].

Gambier (Uncaria gambier Roxb.) as one of betel quid composition, has also been used in Indonesia for some medical reason, such as anti bacteria, antioxidant, anti-inflammation, analgesic, and antihypercholesteremia. The main ingredient consisted in gambier that has important role in traditional medicine is catechins. Catechins are type of polyphenol and part of the flavonoid groups, that strongly has antioxidant effect [56]. The anti-inflammatory effect of catechins is due to its ability in
reducing cyclooxygenase, lipoxygenase, and phospholipase enzyme activities. Once those enzymes are inhibited, the mediators of inflammations including prostaglandin and leukotriene synthesis are disrupted [57]. Catechins decrease XO enzyme activities, NO production, NF-kB activation, pro inflammatory cytokines, TNF-a, IL-1, IL-2, IL-6, IL-8, IL-12 and phospholipase A2 [58,59]. Riswana et al revealed that gambier leaf acted as anti inflammatory because it created XO inhibitory activities [60].

The antibacterial and anti-inflammatory effect of gambier accelerate proliferation and remodelling process of wound healing. By keeping the wound sterile, the irritant is removed, the number of neutrophils are reduced, the inflammatory phase is quicken, and the next phase of wound healing including proliferation and maturation phase are stimulated [61]. Sumosa et al reported that gambier in various concentrations influenced the time and percentage of wound healing process [62]. Septiani et al stated that ethanol gambier leaves lowered the width and length of wound, so the time of healing process was shortened [63].

Calcium hydroxide (Ca(OH)₂) is commonly use in dental procedure as analgesic, anti-inflammation, and regeneration of dental and periapical tissue. Ca(OH)₂ is an inorganic compound that widely uses in many formulation of pulp capping, root canal medication and filling [64]. Ca(OH)₂ reduces TNF-a protein and mRNA level, initiates vascular and inflammatory cell migration and proliferation to handle and remove irritating agents. It will stimulate fibronectin gene expression, mineralization, inhibit bacterial LPS, so that it can induce tissue regeneration, including migration and proliferation of mesenchymal and endothelial pulp cells, followed by collagen formation [65,66]. Kasaj et al evaluated periodontal wound healing of non-surgical periodontal patients and proved that the application of oily calcium hydroxide suspension (Osteoinductal) mediated the wound healing process, by leading superficial necrosis. Thus necrosis caused inflammatory reaction, then the inflammatory layer is replaced by fibroblast [67]. Its biological and antiseptic effects are due to alkalinity and calcium ion release. The alkaline pH activates alkaline phosphatase and reinforces biomeneralization process. Slight necrosis on the surface layer induced by alkaline pH will precipitate wound healing [68].

4. CONCLUSION

Betel quid extract presents anti-inflammatory effect and wound re-epithelization on mucosal wound healing process and characterized as dose-dependant effect. This finding also contribute to find another mechanism of action of active ingredients from those plants for further study.

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