Total nephroureterocystectomy and urethrectomy due to urothelial carcinoma associated with the BK polyomavirus infection after kidney transplantation: a case report with literature review

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Abstract

Background: BK polyomavirus (BKPyV) infection after kidney transplantation is an important cause of graft failure among kidney transplant recipient and may cause malignant tumor, although the association between BKPyV infection and malignant tumor has been controversial yet.

Case presentation: We report a case of a 39-year-old-male kidney transplantation (KTx) recipient with urine BKPyV replication who developed a graft pelvic tumor with the positive Simian virus 40 large T antigen (SV40 TAg). The patients received a living-related KTx from his 65-year-old mother. A protocol biopsy at 14 months after KTx showed BKPyV-associated nephropathy. Therefore, the dose of immunosuppressants was reduced, resulting in improved BKPyV viremia, but viruria persisted. About 117 months after KTx, urine cytology showed atypical cells suspicious for malignancy. Cystoscopy revealed a tumor on the neck of the bladder. Transurethral resection of the bladder tumor (TUR-BT) was performed; however, the diagnosis of malignancy was not confirmed at that time. Six months after the TUR-BT, urine cytology showed atypical cells definite for malignancy. Computed tomography and retrograde pyelography showed no evidence of urinary tract tumor and metastasis. Subsequently, total nephroureterocystectomy and urethrectomy were performed. Histological examination of the graft ureter revealed a high-grade urothelial carcinoma, with glandular differentiation, pT1. Immunohistochemically, the tumor showed positivities for SV40 TAg and p53, along with increased Ki67 labeling cells were increased. By contrast, nonneoplastic cells were negative for SV40 TAg. At the time of writing the present manuscript, the patient is free from recurrence or residual tumor and being closely monitored without additional therapy, 32 months after the surgery.

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Background
In recent years, long-term graft survival for kidney transplantation (KTx) has improved with advances in immunosuppressive therapy. Currently, the widely used strategy for immunosuppression related with KTx is suppression of helper T lymphocyte activity such as tacrolimus (TAC)-based immunosuppression; viral infection is considered as a frequent complication after KTx. In particular, BK polyomavirus (BKPyV) is an important cause of nephropathy and graft failure for KTx recipients. BKPyV infection may cause nephropathy in 2–10% of KTx recipients [1–5] and lead to graft loss in approximately 15–80% [1, 5–9]. Furthermore, BKPyV has been known to have a tropism for the genitourinary tract and causes clinical disease. Thus, for immunocompromised patients such as KTx recipients, not only BKPyV-associated nephropathy but also ureteral stenosis and hemorrhagic cystitis would occur.

Recently, the relationship between BKPyV and genitourinary malignant tumor, such as urothelial carcinoma [10, 11] especially in bladder cancer [11–13] and renal cell carcinoma [14], has been reported. However, few studies suggested a potential role of BKPyV in the development of urologic tract malignancies in KTx recipients [15], and the association remains controversial. Herein, we describe a case of a KTx recipient with positive urine BKPyV replication who developed a graft pelvic tumor with positive immunohistochemical staining for simian virus 40 large T antigen (SV40 TAg).

Case presentation
A 38-year-old man was diagnosed with chronic kidney disease and underwent hemodialysis. The etiology of his renal failure was unknown. His past medical history included hypertension alone, without a history of malignancy or any exposure to carcinogens. At 39 years of age, the patient received ABO-incompatible living-donor KTx. The donor was his 65-year-old mother. Human leukocyte antigen typing revealed two mismatches, and donor-specific antibody tests were negative. Immunosuppressive therapy comprised TAC, mycophenolate mofetil (MMF), methylprednisolone (MP), basiliximab, and rituximab. Plasmapheresis was also performed thrice because of ABO incompatibility. Serum creatinine (sCr) level decreased to 1.76 mg/dL on postoperative day (POD) 5. A protocol biopsy on POD 17 showed no evidence of rejection, and he was discharged on POD 23. Thereafter, TAC, MMF, and MP were continued as immunosuppressive maintenance therapy. Furthermore, his sCr level ranged from 2.0 to 2.4 mg/dL. At 14 months after KTx, his graft function became stable (sCr, 2.49); however, the protocol biopsy showed BKPyV-associated nephropathy (BKPyVAN) (Fig. 1) incidentally. In addition, polymerase chain reaction (PCR) with urine sample detected BKPyV at 4.9 × 10^7 copies/mL; BKPyV was undetectable in serum PCR test. Therefore, we decreased the dosage of TAC and switched MMF to mizoribine (MZ). Thereafter, the PCR titer of urine BKPyV decreased but persisted (Fig. 2). 98 months after KTx, hematuria was suddenly observed and persisted (urine red blood cell level varied between 10–19 and 50–99/high-power field). Thus, urine cytology was repeated during the follow-up; the patient was noted to be suspicious of malignancy by urine cytology and cystoscopy revealed tumor on the neck of the bladder at 117 months after KTx (Fig. 3). Transurethral resection of the bladder tumor (TUR-BT) was performed; however, we failed to find the tumor resection and thereby definitive diagnosis was not reached. Thereafter, urine cytological examination was periodically continued, and 6 months after TUR-BT, bladder, and graft urine cytology showed malignant cells (Fig. 4). Although computed tomography and retrograde pyelography showed no significant evidence of urinary tract tumor and metastasis, the patient was finally diagnosed as malignancy. Thereafter, total nephroureterocystectomy and urethrectomy were performed to radically resect the malignant tumor at 124 months after KTx. Grossly, there was a papillary tumor in the uretero-pelvic junction on the kidney graft (Fig. 5). Histologically, the tumor was diagnosed as a high-grade urothelial carcinoma with glandular differentiation, pT1, in the ureter of the allograft. Immunohistochemically, SV40 TAg and p53 were positive in the tumor cells. Ki67 was increased. Non-neoplastic cells failed to show a positive reaction for SV40 TAg staining (Fig. 6). The patient resumed maintenance hemodialysis after the operation. To date, the patient is closely monitored without additional therapy and without recurrence or metastasis for 32 months.

Conclusion: The relationship between BKPyV infection after KTx and bladder carcinogenesis remains to be elucidated. However, when the KTx recipients who continue to have BKPyV infection for a long time are treated, the possibility of risk factors for renourinary carcinoma should always be carefully considered.

Keywords: BK polyomavirus, Urothelial carcinoma, Kidney transplantation, Total nephroureterocystectomy, Urethrectomy
Discussion and conclusions
BKPyV primary infection commonly occurred as an asymptomatic, upper respiratory tract infection, or flu-like infection at childhood, and approximately 65–90% of the general population may reach a seroprevalence state [5, 16–20]; thereafter, the virus is considered to remain mainly in the urinary tract. BKPyV infection in KTx recipient develops not only renal dysfunction due to nephropathy but also ureteral stenosis and hemorrhagic cystitis [21]. Moreover, BKPyV has been recently reported as a viral tumor that occurs in immunocompromised hosts such as KTx recipients, together with a long-term allograft survival. The exact effect of BKPyV in KTx recipients remains uncertain; however, the incidence of urinary tract tumor such as bladder tumor and renal cell carcinoma would increase [10–14] throughout BKPyV replication [22, 23].

In general, BKPyVAN showed a positive immunohistochemical staining for SV40 TAg, p53, and Ki67 throughout the upregulation of these markers [24]. However, the role of BKPyV in solid organ recipients is uncertain at present, but urinary tract tumors are among the best candidates for an etiologic association with BKPyV infection. In vitro studies, BKPyV infection could transform

Fig. 1 Protocol biopsy of transplanted kidney performed on postoperative day 426. a BK polyomavirus-associated nephropathy (arrow) and b no result of rejection findings [a Simian vacuolating virus 40 large T antigen stain, and b hematoxylin-eosin stain]

Fig. 2 Patient clinical course. TAC tacrolimus, MMF mycophenolate mofetil, MP methylprednisolone, Rit rituximab, Basi basiliximab, MZ mizoribine, CyA cyclosporine, Cre serum creatinine (mg/dL), BKPyV BK polyomavirus-associated nephropathy, Urine BKPyV urine BK polyomavirus DNA-PCR (copies/mL/day), s/o malignancy suspicious of malignancy

123mo Urine BKPyV was continue positive
123mo Cytology of graft urine malignancy
① No rejection
② BKPyV
Observation after reduced immunosuppression
Cytology atypical cells
39y.o. LRKTx 1mo Bx.① 14mo Bx.② 33mo Bx.③ 98mo Occult blood 117mo Cytology s/o malignancy

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cells and cause malignancy in immunosuppressed or immature rodents. BKPyV genomes are divided into three distinct regions: the early viral gene region (EVGR), the late viral gene region (LVGR), and the noncoding control region. EVGR encodes the regulatory proteins large tumor antigen (LTAg), which inactivates the tumor suppressors, pRB and p53 [23]. Kenan et al. also hypothesized that dysregulation of LTAg by gene integration of BKPyV with persistent overexpression promotes cell growth, genetic instability, neoplastic transformation, and tumorigenesis [25]. These onco genes occur mainly in renalourinary organs, including the ureter [23], as in our case. Furthermore, Monini et al. reported that the presence and physical state of BKPyV DNA LTAg gene sequences were amplified by PCR from the prostate, kidney, ureter, and bladder, including these carcinomas [26]. Taking this into consideration, SV40 TAg positivity in tissues indicated BKPyV infection status, and past BKPyV infection may influence malignancy development; thus, SV40 TAg positivity in tissues is associated with BKPyV infectious carcinoma, although not always.

In KTx recipients receiving strong immunosuppressive therapy, viral infection would be a serious problem. Thus, once BKPyV infection is found detected, the reduction of immunosuppressive drugs is recommended [17]. Especially, TAC is a current key drug mainly used as an immunosuppressant for KTx recipients [27]. TAC is identified as a risk factor for developing BKPyV viremia or BKPyVAN [3, 28–30], which may reduce the TAC dosage [31]. In this report, the recipient was managed with maintenance immunosuppression consisting of TAC, MMF, and MP, but a protocol biopsy performed at 14 months after KTx revealed subclinical BKPyV-associated nephropathy; thereafter, TAC reduction and anti-metabolite medicine conversion were performed as immunosuppression weakening. After these changes, the PCR titer in urine BKPyV was decreased but remained.

In this case, malignancy was detected about 10 years after KTx. Although during this long duration, the graft kidney might have been exposed to exogenous carcinogens, e.g., smoking or chemical dyes such as aromatic amines, ionizing radiation, and cyclophosphamide, only BKPyV viruria was suspected as the risk factor for malignancy. Urine cytology was routinely performed, and the cellular atypism got higher and finally diagnosed as bladder cancer based on cystoscopy findings. At first, TUR-BT on the neck of the bladder was performed; however, we failed to catch the tumor resection and characteristic evaluation was not performed. Thereafter, repeat urine cytology was performed after TUR-BT to detect malignancy. The grade of malignancy based on urine cytology worsened and finally diagnosing allograft–ureter cancer. Although intravesical Bacillus Calmette-Guerin instillation was considered at that time, the treatment had been abandoned because of the patient status under immunosuppressant. In order to improve the curability, total nephroureterectomy and urethrectomy were performed at 124 months after KTx in spite of stable graft function.

For immunochemical staining, urothelial carcinoma was stained positive with SV40 TAg; however, normal tissues, such as native kidney and ureter, and allograft kidney tissues were negative with SV40 TAg staining.
SV40 TAg staining is a reliable test for BKPyV infection [32]; therefore, these results suggested that BKPyV infection for a long-term made graft urothelium malignant, and the focus of malignancy was allograft ureter clearly.

As regards the study limitation, detail DNA analysis or fluorescence in situ hybridization was not performed; however, SV40 TAg-positive cells in this case were detected only in the carcinoma tissue, and other tissues were not stained with immunohistochemical staining. Furthermore, we unfortunately failed to catch the bladder tumor during the TUR-BT. Therefore, the relationship between bladder tumor and BKPyV infection remains unclear.

In conclusion, an interesting but challenging case of urothelial carcinoma in allograft is presented for KTx recipients after BKPyV-associated nephropathy with remaining viruria. When the KTx recipients who continue to have BKPyV infection for a long time are treated, the possibility of risk factors not only for BKPyV-associated nephropathy but also for renourinary carcinoma should always be carefully considered. Therefore, we continued careful follow-up by routine urine
cytology tests, and the patient was immediately diagnosed with urinary tract carcinoma and total nephroureterocystectomy and urethrectomy were performed.

Literature review
As treatment strategies for BKPyV infection, reducing or stopping immunosuppressive agents, administering intravenous immunoglobulin, cidofovir induction, and others may be considered.

Their treatment efficacies are unclear, and acute rejection may occur from reduced immunosuppressive therapy. There are no antiviral drugs; therefore, screening for BKPyV replication and timely reduction of immunosuppression as the preemptive intervention is currently recommended [31, 33, 34]. Significant BKPyV replication manifests as (a) probable BKPyVAN, defined by sustained BKPyV viremia of > 3 log10 copies/mL in two measurements within 3 weeks; or (b) presumptive BKPyVAN, defined by at least one of two measurements are > 4 log10 copies/mL; or (c) proven BKPyVAN, according to biopsy results [34].

For reduction immunosuppression, two strategies have been recommended by the American Society of Transplantation guidelines: (i) dose reduction of calcineurin inhibitor (CNI) by 25–50% in one or two steps, followed by reducing antimetabolites by 50%, followed by discontinuing the latter and (ii) reduction of antimetabolites by 50%, followed by reducing CNI by 25–50%, and then discontinuing antimetabolites. Bischof et al. reported that the “CNI first” immunosuppression reduction strategy leads to similar long-term outcomes in patients with and without BKPyV viremia and is associated with a low risk for clinical antibody-mediated rejection after clearance of BKPyV viremia [31]. In addition, prednisone is typically tapered to 10 mg or less per day [34].

In the standard therapy, the conversion from MMF to MZ is not included because MZ is not available worldwide. In Japan, immunosuppression therapy, including MZ, is used for a compromised host for viral infections such as cytomegalovirus [35, 36]. Therefore, MZ is sometimes administered instead of MMF for the inhibition of BKPyV infection [37, 38].

T cell- and antibody-mediated rejection can occur complicated with BKPyV infection in tandem with immunosuppression reduction. It is required to judge which one is mainly a factor of deterioration of allograft function and may change the treatment strategy, such as whether immunosuppression should be re-increased or not.

Abbreviations
KTx: Kidney transplantation; TAC: Tacrolimus; BKPyV: BK polyomavirus; SV40 TAg: Simian virus 40 large T antigen; MMF: Mycophenolate mofetil; MP: Methylprednisolone; sCre: Serum creatinine; POD: Postoperative day; PCR: Polymerase chain reaction; BKPyVAN: PyW-associated nephropathy; TUR-BT: Transurethral resection of the bladder tumor; CNI: Calcineurin inhibitor

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Authors’ contributions
RM, M.O, K.U., Y.N, and H.I. participated in research design, writing the article, and performance of the research. H.F., K.Y., T.T., J.I., T.S., H.A., K.T., and K.N. contributed to the design and editing of the article. The authors read and approved the final manuscript.

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Consent for publication
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Competing interests
The authors have no competing interests and do not have any relationships to disclose.

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