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Sex-specific genetic effects in physical activity: results from a quantitative genetic analysis

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Abstract

Background: The objective of this study is to present a model to estimate sex-specific genetic effects on physical activity (PA) levels and sedentary behaviour (SB) using three generation families.

Methods: The sample consisted of 100 families covering three generations from Portugal. PA and SB were assessed via the International Physical Activity Questionnaire short form (IPAQ-SF). Sex-specific effects were assessed by genotype-by-sex interaction (GSI) models and sex-specific heritabilities. GSI effects and heterogeneity were tested in the residual environmental variance. SPSS 17 and SOLAR v. 4.1 were used in all computations.

Results: The genetic component for PA and SB domains varied from low to moderate (11 % to 46 %), when analyzing both genders combined. We found GSI effects for vigorous PA (p = 0.02) and time spent watching television (WT) (p < 0.001) that showed significantly higher additive genetic variance estimates in males. The heterogeneity in the residual environmental variance was significant for moderate PA (p = 0.02), vigorous PA (p = 0.006) and total PA (p = 0.001). Sex-specific heritability estimates were significantly higher in males only for WT, with a male-to-female difference in heritability of 42.5 (95 % confidence interval: 6.4, 70.4).

Conclusions: Low to moderate genetic effects on PA and SB traits were found. Results from the GSI model show that there are sex-specific effects in two phenotypes, VPA and WT with a stronger genetic influence in males.

Keywords: Sex effects, Heritability, Physical activity, Sedentary behaviour

Background

It has been widely observed that moderate to high physical activity (PA) levels are associated with lower risks for several chronic diseases, especially the major consequences of the metabolic syndrome, namely obesity, type 2 diabetes, and cardiovascular disease [1–3]. In spite of these observations, individuals in the general population tend to adopt less active or sedentary lifestyles [4–6]. A recent report showed that approximately 31.1 % of adults worldwide are physically inactive, with values ranging from 17 % (in southeast Asia) to about 43 % (America and eastern Mediterranean). Furthermore, approximately 80 % of adolescents between 13–15 years old do not reach the recommended daily PA values (60 min/day of moderate to vigorous PA) [7]. Moreover, there is a marked sex-difference in PA levels (regardless of how PA levels are measured), with males consistently having higher PA levels than females [4–8].

Here we treat sex as a composite internal environment [9, 10], and we ask if the sex environment has a significant effect on the genetic variation underlying PA levels. Although results on the genetics of PA have increased over the last decade [11–15], there is still little known about how sex affects the genetic determinants of different expressions of PA phenotypes [16]. Previous studies that employed family designs in the genetic analysis of PA levels have demonstrated that genetic effects can account for a significant proportion (as high as 60 %) of PA variability [17–21]. However, such studies did not test for possible genotype-by-sex interaction (GSI) underlying sex-specific effects in PA. One widely used approach towards addressing this issue is to make inferences on the basis of sex-specific heritabilities of PA. As we demonstrate in this...
report, however, sex-specific heritability is a potentially misleading measure of the importance of sex-specific genetic effects, and can in fact lead to inaccurate inferences. To see why sex-specific heritability is potentially misleading in regard to sex-specific genetic effects, consider the definition of the heritability as the ratio of the additive genetic variance to the total phenotypic variance, which in the absence of genetic marker data is simply the sum of the additive genetic and residual environmental variances. Because the heritability is a ratio, its value can change as a result of changes in the genetic effects or in the residual environmental effects, or in both. Thus, with respect to the objective of elucidating sex-specific genetic effects per se, sex-specific heritability estimates can be indecisive and/or misleading. Alternatively, we advocate a formal GSI model [22, 23] designed to detect sex-specific genetic effects sensu strictu in PA. We contrast this formal GSI model with a re-parameterized form of the model designed to detect sex-specific heritabilities in PA. In contrasting the two, we show that whereas the formal GSI model can efficiently detect sex-specific genetic effects, the sex-specific heritability model can and does lead to improper inferences. Thus, the primary aim of this study is to investigate potential sex-specific genetic effects influencing PA levels and sedentary behaviour (SB), and the secondary aim is to show that GSI modelling and sex-specific heritability modelling can lead to different conclusions.

Methods
Sample
The study used cross-sectional PA and SB data arising from three generation families comprising 1034 subjects from 100 families (517 females and 517 males), aged 7 to 85 years old from Portugal. The University of Porto Ethics committee approved the study and written informed consent was obtained from all subjects. The parents or legal guardian signed the written informed consent of the children and adolescents. The average family size was 10, with a range of 9 to 16 subjects. Due to missing data, only 724 individuals from 100 families were included in the analysis.

Physical activity and sedentary behaviour
PA and SB were measured with the International Physical Activity Questionnaire, short form (IPAQ-SF). This instrument has acceptable measurement properties in estimating PA levels with international validation results previously reported [8, 24]. Participants aged below 15 years were interviewed by a team previously trained according to the standardized IPAQ-SF approach. All other individuals self-reported their responses to all questionnaire items. A random sample of 10 families was retested two weeks apart and reliability values (ANOVA-based intraclass correlation coefficient) within each family member were around 0.80.

Participants answered about intensity, frequency and duration of their physical activities during one week, as well as about their SB. Derived phenotypes were: vigorous PA (VPA); moderate PA (MPA); walking (WK); total PA (TPA); time spent watching television (WT) and sitting time (ST). Standard procedures were adopted, using continuous scales of weekly energy expenditure to represent several PA levels, expressed by metabolic equivalent (MET/minutes/week). The reference values for the calculation of energy expenditure were: $VPA = 8.0$ METs; $MPA = 4.0$ METs; and $WK = 3.3$ METs. TPA is an index of the sum of the values reported for VPA, MPA and WK. SB values were maintained in minutes per day, according to the IPAQ protocol [25].

Statistical analysis
SPSS version 17.0 was used for exploratory data analysis in order to verify probable data entry errors as well as the presence of outliers. Due to high skewness and kurtosis, median values and interquartile range were used to describe all phenotypes. PA and SB phenotypes were adjusted for the following covariate effects: age, sex, age$^2$, sex$^*$age, sex$^*$age$^2$. The residuals obtained after accounting for the covariate effects were then transformed using an inverse normal transformation [26, 27].

We now describe the statistical genetic models employed in our analyses. Assuming that dominance and epistasis are negligible, comparisons of individual phenotype $y$ define the covariance of the basic polygenic model as:

$$
\text{Cov}(y_x, y_z) = 2 \phi_{xz} \sigma^2_g + \sigma^2_x \delta_{xz}
$$

(1)

where $x$ and $z$ index individuals, $2 \phi_{xz}$ gives the expected coefficient of relationship, $\sigma^2_g$ is the additive genetic variance, $\sigma^2_x$ is the environmental variance, and $\delta_{xz}$ is defined as 1 when individuals $x$ and $z$ are the same and 0 otherwise. This model is used to estimate heritability, which is given as: $h^2 = \frac{\sigma^2_g}{\sigma^2_g + \sigma^2_e} = \frac{\sigma^2_g}{\sigma^2_f}$ where $\sigma^2_g$ is the total phenotypic variance. The model can be extended to account for GSI effects by allowing for sex-specific additive genetic and environmental variances denoted by $\sigma^2_{gfh}$, $\sigma^2_{gmn}$, $\sigma^2_{fgh}$ and $\sigma^2_{mnh}$ where $f$ and $m$ index females and males, respectively, and for an across-sex genetic correlation denoted by $p_{gfh,mnh}$.

In principle, sex-specific heritabilities can be computed from the relevant parameter estimates under this GSI model. However, for formal statistical inferences under the model, we would have no recourse but to use crude approximations of the standard errors for the sex-specific heritabilities, and an associated Wald test statistic. Worse still, the Wald test statistic formed in this case from variance component estimates is known to be problematic [28, 29].
Therefore, in order to be able to estimate sex-specific heritabilities for which reliable statistical inferences can be made, we developed a reparameterized version of the GSI model by instead allowing for sex-specific phenotypic variances, heritabilities, and corresponding "environmentalities", respectively denoted by \( \sigma^2_{gf}, \sigma^2_{gm}, h^2_f, h^2_m, (1 - h^2_f) = \sigma^2_f, \) and \((1 - h^2_m) = \sigma^2_m,\) while maintaining the same parameterization for the across-sex genetic correlation.

On finding the maximum likelihood estimates (MLEs) of the parameters, defined as estimates that make the full likelihood function a maximum, and the model likelihoods under the null and alternative hypotheses, inferences are then made by way of the likelihood ratio test and its associated test statistic (LRT). The LRT is computed as minus twice the difference in ln-likelihoods estimated under the null and alternative hypotheses, and is in standard cases distributed as a chi-square with degrees of freedom (d.f.) given by the difference in the number of estimated or un-constrained parameters. However, as is often the case for variance components models, the non-standard case of testing a null hypothesis lying at a boundary of its acceptable parameter range yields a LRT distributed as a 50:50 mixture of a point-mass at 0 and a chi-square with 1 d.f., which we write as \( \left( \frac{1}{2} \chi^2_0 + \frac{1}{2} \chi^2_1 \right) \) [30, 31]. For example, the null hypothesis that the heritability equals zero lies at a boundary of its acceptable parameter range. In this case, the LRT is distributed as \( \left( \frac{1}{2} \chi^2_0 + \frac{1}{2} \chi^2_1 \right) \).

In this report we are chiefly concerned with addressing the existence of GSI effects and of sex-specific heritabilities. It can be shown that in order for there to be a GSI effect, there must be either heterogeneity in the additive genetic variance across environments (i.e. \( \sigma^2_g \neq \sigma^2_{gm} \)) or the genetic correlation across environments must be different from unity (i.e. \( \rho_{gf(m)} \neq 1 \)) or both. Thus, for GSI we have two null hypotheses to test, namely \( \sigma^2_g = \sigma^2_{gm} = \sigma^2_f \) and \( \rho_{gf(m)} = 1 \). When testing \( \sigma^2_g = \sigma^2_{gm} = \sigma^2_f \) we have a 1 d.f. difference between models compared, and so the LRT is distributed as \( \chi^2_1 \). When testing \( \rho_{gf(m)} = 1 \), we again have a 1 d.f. difference, but the relevant null hypothesis, namely \( \rho_{gf(m)} = 1 \), lies at a boundary of its acceptable parameter range, and so the LRT in this case is distributed as \( \left( \frac{1}{2} \chi^2_0 + \frac{1}{2} \chi^2_1 \right) \). Using the GSI model, we can also test the null hypothesis that the sex-specific residual environmental variances are equal, namely \( \sigma^2_g = \sigma^2_{gm} = \sigma^2_e \) for this case, the LRT is distributed as \( \chi^2_2 \). The next inferential step is to test if each of the sex-specific heritabilities are significantly different from 0, with LRTs distributed as \( \left( \frac{1}{2} \chi^2_0 + \frac{1}{2} \chi^2_1 \right) \).

Reports in the literature of sex-specific heritabilities are invariably presented as interval estimates. In order to make our results comparable with the previous studies, we constructed likelihood-based 95 % confidence intervals (CIs) following recommendations in the literature [32, 33]. A likelihood-based 95 % CI around a given maximum likelihood estimate (MLE) is one that satisfies the following general formula:

\[
\text{LRT} \leq \chi^2(a). \tag{2}
\]

where the LRT is computed by comparing a model for a given MLE effect size (ES) to a model with a parameter ES constrained in the direction of the lower or upper bound that satisfies the formula, and \( \chi^2(a) \) is the chi-square with the relevant d.f. under the comparison which corresponds to the nominal level of significance given by \( a \), which in our case is \( a = 0.05 \). Thus, to compute the 95 % CI with respect to the null hypothesis of equality of sex-specific heritabilities or that a given sex-specific heritability is different from zero we find the parameter estimates satisfying:

\[
\text{LRT} \leq \chi^2(a = 0.05) = 3.84 \tag{3}
\]

In one case in the ensuing it was not possible to compute a 95 % CI using this approach. Thus, we used the following formula for an approximate 95 % CI [34], where SE is the standard error:

\[
95\% \text{ CI} = \text{ES} \pm 1.96 \times \text{SE} \tag{4}
\]

To help shed light on our findings, we performed a post-hoc power analysis. Power is defined as the probability of correctly rejecting the null hypothesis, and is computed as the probability integral from the point on the alternative distribution corresponding to the nominal significance level or alpha (on the null distribution) to the upper limit of the alternative distribution at positive infinity. Since the total probability of any distribution is 1, power can be conveniently computed as:

\[
\text{Pr}(\sigma^2_{gf} \neq \sigma^2_{gm}) \quad \text{or} \quad \text{Pr}(\rho_{gf(m)} < 1) = \int_{X_{\alpha/2}}^{\infty} dX_{\alpha/2} - \int_{0}^{X_{\alpha/2}} dX_{\alpha/2} = 1 - \int_{0}^{X_{\alpha/2}} dX_{\alpha/2} = 1 - \beta, \tag{5}
\]

where the distribution under the alternative hypothesis is the non-central chi-square distribution, denoted by \( X^2_{\nu, \alpha} \), \( \nu \) is the degrees of freedom (d.f.) parameter, \( \xi \) is the non-centrality parameter (NCP), \( X_{\alpha/2, \xi = 0} \) is the point on the non-central chi-square distribution corresponding to the 100(1 - \( \alpha \)) percentage point on the distribution under the null hypothesis, and \( \beta = \int_{0}^{X_{\alpha/2}} dX_{\alpha/2} \) is the probability of making a type II error. We used a similar approach to that used by Blangero et al. [35].
Results and discussion

Main descriptive statistics for all PA and SB phenotypes are presented in Table 1. About half of first and second generation subjects did not report VPA. The TPA levels increase across generations. Reported SB has higher values in the first generation, as expected, although second and third generation subjects had similar values.

Heritability estimates for the five phenotypes are presented in Table 2. All traits are statistically significant ranging from 0.112 (WT) to 0.456 (WK). The covariates explained 2% to 30% of the phenotypic variance.

We found GSI effects for VPA (p = 0.0005) and WT (p = 0.02), and heterogeneity in the residual environmental variance for VPA (p = 0.006), MPA (p = 0.02) and TPA (p = 0.001) (Table 3). Sex-specific heritability estimates are reported in Table 4, showing significantly higher values in males for WT but not for VPA (Fig. 1).

We report power to detect GSI effects for each trait in Table 5. There was sufficient power to detect GSI due to heterogeneity in the additive genetic variance for only VPA but not for the other traits. Also, there was sufficient power to detect GSI due to a genetic correlation coefficient less than 1 for only WT but not for the other traits. That we found significant GSI due to additive genetic variance heterogeneity in VPA is not too surprising given that we had sufficient power for this trait. On the other hand, there are two related issues regarding the power analysis results for WT that need some explanation. The first issue is that we found significant GSI due to additive genetic variance heterogeneity for WT but the power to detect this GSI effect was not sufficient. The second issue is the converse in that we had maximum power to detect a GSI effect due to a genetic correlation coefficient less than 1 and yet we failed to reject the null hypothesis of $\rho_{(f,m)} = 1$. On these issues, we note the following observations. Low power does not entirely preclude the rejection of a null hypothesis. Power merely measures the probability at which rejection of a false null hypothesis may occur. Thus, it is possible (though perhaps not so probable) to have low power and yet reject a false null hypothesis. Conversely, maximum power simply means that if a null hypothesis is false, then it will be rejected every time, but it does not necessarily mean that the null hypothesis is in fact false. It is well known that complex morphological and physiological traits may often exhibit marked sexual dimorphism, especially traits associated with body fat distribution and physiological weight regulation [36–38]. Marked sex-specific variation is also evident in behavioral characteristics such as PA and SB, and may have a significant genetic component or may be modulated by genetic determinants through sex-specific effects [16]. To assess these sex-specific effects on several PA and SB phenotypes, we employed models of GSI effects and of sex-specific heritabilities. We tested for GSI effects and heterogeneity in the residual environmental variance. Then, to be able to compare our results with reports in the literature, which

Table 1 Descriptive statistics for age, physical activity levels and sedentary behaviour by generation and sex

| Trait     | 1st Generation | 2nd Generation | 3rd Generation |
|-----------|----------------|----------------|----------------|
|           | Grandfather    | Grandmother    | Father         | Mother         | Son            | Daughter       |
| n         | n = 104        | n = 104        | n = 147        | n = 147        | n = 111        | n = 111        |
| VARIABLES| Mean ± sd       | Mean ± sd      | Mean ± sd      | Mean ± sd      | Mean ± sd      | Mean ± sd      |
| Age (years) | 74.4 ± 10.7  | 76.4 ± 7.6     | 41.9 ± 5.2    | 40.9 ± 4.9     | 13.6 ± 2.4    | 13.3 ± 2.5    |
| VPA (mets/min/week) | 0 (0) | 0 (0) | 0 (320) | 0 (160) | 280 (960) | 80 (640) |
| MPA (mets/min/week) | 0 (80) | 0 (0) | 140 (400) | 160 (360) | 280 (620) | 240 (480) |
| WK (mets/min/week) | 330 (297) | 247.5 (256) | 743 (660) | 660 (561) | 718 (1068) | 743 (990) |
| SB ST (min/day) | 420 (180) | 420 (120) | 300 (120) | 300 (120) | 360 (120) | 360 (120) |
| WT (min/day) | 155 (138) | 180 (120) | 80 (75) | 80 (60) | 92.5 (80) | 105 (60) |
| TPA (mets/min/week) | 413 (291) | 264 (346) | 1058 (923) | 1015 (913) | 1722.25 (1571) | 1545 (1435) |

sd: standard deviation; IQR: interquartile range; VPA: vigorous physical activity; MPA: moderate physical activity; WK: walking; ST: sitting time; WT: watching television; TPA: total physical activity

Table 2 Heritability estimates ($h^2$), standard errors (se), p-values and explained variance by covariates

| Trait | $h^2$ ± se | p-value | Variance explained by covariates (%) |
|-------|------------|---------|-------------------------------------|
| VPA   | 0.280 ± 0.061 | $1.0 \times 10^{-7}$ | 2.0 |
| MPA   | 0.312 ± 0.060 | $2.4 \times 10^{-5}$ | 8.0 |
| WK    | 0.456 ± 0.058 | $3.1 \times 10^{-14}$ | 15.0 |
| ST    | 0.287 ± 0.059 | $4.1 \times 10^{-8}$ | 16.0 |
| WT    | 0.112 ± 0.061 | $2.5 \times 10^{-2}$ | 30.0 |
| TPA   | 0.284 ± 0.059 | $3.4 \times 10^{-8}$ | 10.0 |

VPA: vigorous physical activity; MPA: moderate physical activity; WK: walking; ST: sitting time; WT: watching television; TPA: total physical activity. Covariates in the regression models included age, sex, age$^2$, sex*age, sex*age$^2$. 

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are usually framed in terms of sex-specific heritabilities, we tested for sex-specific heritabilities in our data.

In the present study, low to moderate genetic contributions to PA and SB phenotypes were found, ranging from 11% to 46%. These results are in accord with previous quantitative genetic studies on nuclear or extended families, which have suggested considerable genetic influence in these traits, notwithstanding the methodological differences to measure PA and SB traits and the diversity in statistical procedures [39]. It should be noted, however, that studies in monozygotic and dizygotic twins have reported greater intraclass correlations for PA levels suggesting moderate to high genetic influences, from 13% to 98% [39].

We found evidence of significant GSI for VPA and WT. In particular, VPA and WT exhibited GSI via significant heterogeneity in the additive genetic variance across genders. For VPA and WT, the additive genetic variance was higher in males than in females. Further, evidence of significant heterogeneity in the residual environmental variance for WT, but not for VPA was shown. Our analysis shows that the male-specific heritabilities for VPA and WT and the female-specific heritability for VPA were significantly different from zero. As can be expected from the GSI results, the male- and female-specific heritabilities for WT were significantly different from each other, which is consistent with Brazilian families’ sedentarism trait shown by Horimoto et al. [16], although their models were parameterized differently, considered no covariates, and had a distinct definition of sedentarism. Surprisingly, however, we found that the male- and female-specific heritabilities for VPA were not significantly different from each other. The explanation for this apparent discrepancy between the GSI and sex-specific results for VPA is actually quite simple. For VPA, the difference across the sexes in the residual environmental variance was of similar magnitude and direction as the difference across genders in the additive genetic variance, and thus the sex-specific heritabilities were quite similar for both males and females. For WT, the additive genetic variance was significantly different across sexes but not the residual environmental variance. Consequently, the sex-specific heritability for WT was significantly lower in females compared to males, which is in opposition to the Brazilian data from Horomito et al. [16], in which the

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### Table 3 P-values for genotype-by-sex interaction effects

| Trait | Equal sex-specific additive genetic variances | Genetic correlation coefficient equal to 1 | Equal sex-specific residual environmental variances |
|-------|---------------------------------------------|--------------------------------------------|---------------------------------------------------|
| VPA   | 0.0003                                      | 0.08                                       | 0.006                                             |
| MPA   | 0.33                                        | 0.18                                       | 0.02                                              |
| WK    | 0.78                                        | 0.36                                       | 0.27                                              |
| ST    | 0.27                                        | 0.50                                       | 0.65                                              |
| WT    | 0.02                                        | 0.23                                       | 0.10                                              |
| TPA   | 0.24                                        | 0.20                                       | 0.001                                             |

VPA: vigorous physical activity; MPA: moderate physical activity; WK: walking; ST: sitting time; WT: watching television; TPA: total physical activity

---

### Table 4 Sex-specific heritabilities

| Trait | % h² (95% CI) | % h² (95% CI) | Δh² (95% CI)* |
|-------|--------------|--------------|---------------|
|       | Males        | Females      |                |
| VPA   | 51.364 (29.75, 72.25) | 32.004 (9.8, 55.59) | 19.3 (−13.4, 50.7) |
| MPA   | 35.477 (13.3, 58.6) | 39.938 (18.4, 62) | −4.461 (−36.1, 27.2) |
| TPA   | 32.931 (12.28, 54.4) | 37.909 (17.03, 59.44) | −4.978 (−35.8, 25.6) |
| WT    | 45.293 (18.9, 72.25) | 2.8 (−20.916, 26.516) | 42.485 (6.4, 70.4) |

*Δh² = male % h² − female % h². The null hypothesis that Δh² = 0 is equivalent to the null hypothesis that the sex-specific heritabilities are equal.

VPA: vigorous physical activity; MPA: moderate physical activity; WK: walking; ST: sitting time; TPA: total physical activity; WT: watching television
heritability was 22 % for females and 5 % for males. It is important to stress that in their study, if a family member did not take part in sports, they were classified as having a sedentarism phenotype. Furthermore, in their analysis no heterogeneity in variance by gender was found when they adjusted their models to age, sex and age-by-sex interaction covariates. Variance heterogeneity was only found when no covariates were included in the models.

Most studies on the relationship between PA levels and WT-like measures have found that the two variables tend to show either a weak relationship or no relationship at all [40–44]. Further, at least two studies have shown that PA levels and WT-like variables have independent effects on cardiovascular disease risk factors, which seems to suggest that there is a weak or even no relationship between the two variables [45, 46]. We should point out, however, that a few studies have found an inverse relationship between PA levels and WT-like measures [47–49]. Nonetheless, evidence from the majority of studies conversant on this issue supports the hypothesis that PA levels and WT-like measures are only weakly associated at best.

Our results introduce an interesting twist to this weak relationship between PA levels and WT-like measures. We showed that the residual environmental variance decreased from males to females for VPA but not for WT. A plausible two-fold explanation of these different patterns is given as follows. It may be that for whatever set of sociocultural reasons there are more opportunities for PA for males than for females (e.g. leisure time practice of team sports) and/or a greater emphasis placed on males relative to that placed on females to be physically active. Additionally, to explain the effectively similar levels of residual environmental variance in males and females for WT it may be that there is enough of a variety of TV shows to equally attract the attention of both sexes and/or that there are TV shows both sexes tend to watch. In the VPA case, the sex-specific heritabilities were rendered statistically indistinguishable by virtue of the residual environmental variance, even in the face of significant heterogeneity in the sex-specific additive genetic variance. These results serve as a sobering reminder that heritabilities are also significantly influenced by the residual environmental variance.

A number of twin studies have reported sex-specific heritabilities of several PA traits, including leisure time PA (LTPA), sports participation (SP), and SP index (SPI) (Table 6). As can be gathered from Table 6, the 95 % CIs on the heritability estimates overlap, which implies that

| Author              | Ano   | Country     | Trait    | % h² (95 % CI) | % h² (95 % CI) |
|---------------------|-------|-------------|----------|---------------|---------------|
| Boomsma et al. [55] | 1989  | Netherlands | SP       | 77 (NA)       | 35 (NA)       |
| Beunen & Thomis [51]| 1999  | Belgium     | SP       | 44 (21–91)    | 83 (66–91)    |
| Maia et al. [52]    | 2002  | Portugal    | LTPA     | 66 (48.9–73.3) | 32 (0.4–61.8) |
| Maia et al. [52]    | 2002  | Portugal    | SPI      | 68.4 (41.5–89.2) | 39.8 (04.73.0) |
| Carlsson et al. [56]| 2006  | Sweden      | LTPA     | 64 (55–72)    | 51 (48–60)    |
|                    |       |             |          | (14–28 yrs)   |               |
| Carlsson et al. [56]| 2006  | Sweden      | LTPA     | 40 (30–45)    | 41 (30–45)    |
|                    |       |             |          | (29–46 yrs)   |               |
| Stubbe et al. [57]  | 2006  | Australia   | EP       | 22.9 (0–56.1) | 31.1 (0.3–55.6) |
| Stubbe et al. [57]  | 2006  | Denmark     | EP       | 44 (24.2–55.7) | 50.1 (30.3–57.7) |
| Stubbe et al. [57]  | 2006  | Finland     | EP       | 55.8 (38.4–63.3) | 61 (44.5–66.3) |
| Stubbe et al. [57]  | 2006  | Netherlands | EP       | 68.1 (34.2–79.0) | 50.3 (21.3–70.3) |
| Stubbe et al. [57]  | 2006  | Norway      | EP       | 33.6 (6.7–61.7) | 56.6 (46.5–63.8) |
| Stubbe et al. [57]  | 2006  | Sweden      | EP       | 63.9 (52.1–68.6) | 59.5 (46.9–64.7) |
| Aaltonen et al. [50]| 2010  | Finland     | LTPA     | 53 (44–60)    | 38 (28–40)    |

h²: heritability; CI: confidence intervals; SP: sport participation; LTPA: leisure-time physical activity; SPI: sport participation index; EP: exercise participation

Table 5 Statistical power analysis for the trait-specific parameters

| Trait | Power or probability of rejecting a false null hypothesis for the following null hypotheses for genotype × sex interaction |
|-------|---------------------------------------------------------------------------------------------------------------|
| VPA   | 0.93558                                                                                                     |
| MPA   | 0.16386                                                                                                     |
| WK    | 0.05925                                                                                                     |
| ST    | 0.19644                                                                                                     |
| WT    | 0.67527                                                                                                     |
| TPA   | 0.22014                                                                                                     |

Table 6 Sex-specific heritabilities of different physical activity phenotypes in twin studies
the sex-specific heritabilities are not different. However, as we showed for VPA it is quite possible that there is significant GSI even in the face of effectively equal sex-specific heritabilities. Moreover, although we did not find evidence of the across-sex genetic correlation being different from one, it is in theory possible for the across-sex genetic correlation to be different than one in these studies. Thus, it may still be possible that, even though most of the studies in Table 6 report sex-specific heritabilities that are not significantly different from each other, there are still significant GSI effects influencing these traits. Conversely, our results logically imply that even if sex-specific heritabilities are significantly different, as in the study by Aaltonen et al. [50] in Table 6, it may not be due to heterogeneity in the additive genetic variance but instead to heterogeneity in the residual environmental variance. That is, it is possible that the sex-specific heritabilities are rendered statistically dissimilar by virtue of heterogeneity in the residual environmental variance, even in the face of an additive genetic variance that is stable across the sexes. These considerations show that in order to be able to make robust statistical inferences about sex-specific genetic determinants it is not enough to estimate sex-specific heritabilities, but rather a formal GSI model must be employed.

The GSI model showed the presence of sex-specific effects in two phenotypes, namely VPA and WT. In light of the power analysis results, these findings are somewhat surprising in that our sample had low power overall to detect GSI effects. Comparing the present results with previous studies is difficult because we could only find one report from a similar extended family design [16], which, as previously mentioned, used a different set of PA phenotypes. Studies with nuclear or extended families [19–21] typically just report mean values of different PA levels and tend to conclude that males are more active than females. However, GSI results are available from twin data analysis [50–52] and suggest the presence of a stronger genetic component in males than in females in some PA phenotypes. This dissimilarity between sexes may not be accurately explained by different sets of genes acting in males and females, which is consistent with our inability to reject the hypothesis that genetic correlation across the sexes is unity. However, it suggests a stronger genetic influence mainly in males. Besides a genetic predisposition towards being physically active or sedentary, genetic variation in PA levels may also be influenced by differences in body size and shape, motor performance, and psychological-drive to be physically active [51, 52].

Molecular genetic studies on PA levels and SB are scant, with little to offer in regard to sex-specific effects. The only association study reporting on this issue was by Simonen et al. [53] who found a consistent association between DNA sequence variation in the dopamine D2 receptor gene (DRD2) with PA levels only in females. To explain this finding they suggested that, because maternal-offspring correlations were consistently higher than paternal-offspring correlations, maternal genotype may have had a stronger impact on offspring's PA levels relative to the effect of paternal genotype. Alternatively, they suggested that the effect of DRD2 in males could have been diluted by a stronger influence of environmental factors relative to that influencing females.

The results and conclusions of the present study may be affected by two main limiting factors. The first refers to the PA assessment through a self-administered questionnaire for individuals over 15 years-old. However, the International Physical Activity Questionnaire’s short form has acceptable measurement properties to access PA levels with validation in several countries [8, 24]. The second limitation has to do with inadequately accounting for household effects. In the case of the Brazilian family data previously mentioned, no significant household effects were found in any PA phenotype. Controlling for the shared environmental effects has been a difficult task. These involve several non-measurable living aspects within families and data regarding their magnitude are still inconsistent [17, 18, 21].

Although there are limitations, this study presents strengths to be highlighted. The participation of a large sample size and age range as well as relationship among different kinship degrees allow structuring expected covariates to a polygenic model [54]. The adequacy of our sample size in regard to the polygenic model notwithstanding, our power analysis results revealed that overall our study had low power to detect GSI effects. We can increase our power in this respect by increasing our sample size for future investigations and by using more sensitive PA and SB measures.

Conclusions

In summary, our findings showed low to moderate genetic effects on PA and SB. Results from the GSI model show that there are sex-specific effects in two phenotypes, VPA and WT with a stronger genetic influence in males. This is consistent with reports in the literature suggesting a higher genetic component in males than females. We also found that inferences based on a GSI model can differ from those based on a sex-specific heritability model. Indeed, if our inferences were based solely on the latter approach we might have concluded that there are no statistically significant sex-specific genetic effects influencing VPA. This highlights the danger of relying on comparisons of sex-specific heritabilities to make such inferences.
The present results advance our understanding of the genetic determinants of PA levels and SB traits. We are optimistic that studies along these lines will one day elucidate the presence of distinct PA- and SB-related genes in males versus females.

Abbreviations
PA: Physical activity; GSI: Genotype-by-sex interaction; SB: Sedentary behaviour; IPAQ-SF: International Physical Activity Questionnaire short form; VPA: Vigorous physical activity; MPA: Moderate physical activity; WK: Walking; TPA: Total physical activity; WT: Time spent watching television; ST: Sitting time; MLEs: The maximum likelihood estimates; LRT: Likelihood ratio test; df: Degrees of freedom; CI: Confidence interval.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
JRM, RNC, MDS, DS, TNG, FKS, RG participated in the conception and design of the study, collected all the information, and cleaned the data set; VPD, RNC, JRM did the analysis and interpretation; VPD, RNC, JB, PK, JARM wrote the manuscript and revised it. All authors read and approved the final manuscript.

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References
1. Churilla JR, Fitzhugh EC. Relationship between leisure-time physical activity and metabolic syndrome using varying definitions: 1999–2004 NHANES. Diab Vac Dær Res. 2009;6(2):100–9.
2. Cho BR, Shin A, Kim J, Lee SH, Sung J. Leisure-time physical activity is associated with a reduced risk for metabolic syndrome. Ann Epidemiol. 2009;19(11):784–92.
3. Luke A, Dugas LR, Durazo-Arvizu RA, Cao G, Cooper RS. Assessing physical activity and its relationship to cardiovascular risk factors: NHANES 2003–2006. BMC Public Health. 2011;11:387.
4. Reddish C, Bo Andersen L, Wiederkopp N, Hano M, Klassen-Heggebe L, Sardinia LB, et al. Physical activity levels and patterns of 9- and 15-year-old European children. Int Sports Med Sci. 2004;34(6):86–92.
5. Troiano RP, Berrigan D, Dodd KD, Mâsse LC, Tilert T, McDowell M. Physical activity in the United States measured by accelerometer. Med Sci Sports Exerc. 2008;40(1):181–8.
6. Trost SG, Pate RR, Sallis JF, Freedson PS, Taylor WC, Dowda M, et al. Age and gender differences in objectively measured physical activity in youth. Med Sci Sports Exerc. 2002;34(2):350–5.
7. Hallal PC, Andersen LB, Bull FC, Guthold R, Haskell W, Ekelund U. Global physical activity levels: surveillance progress, pitfalls, and prospects. Lancet. 2012;380(9838):247–57.
8. Bauman A, Fiona B, Tien C, Craig CL, Ainsworth BE, Sallis JF, et al. The international prevalence study on physical activity: results from 20 countries. Int J Behav Nutr Phys Act. 2006;12(121).
9. Voruganti VS, Diego VP, Haack K, Cole SA, Blanergo J, Göring HH, et al. A QTL for genotype by sex interaction for anthropometric measurements in Alaskan Eskimos (GOCADAN Study) on chromosome 19q12-13. Obesity (Silver Spring). 2011;19(9):1840–6.
10. Comuzzie AG, Blanergo J, Mahaney MC, Mitchell BD, Hixson JE, Samollow PB, et al. Major gene with sex-specific effects influences fat mass in Mexican Americans. Genet Epidemiol. 1995;12(5):475–88.
11. Moore-Harrington T, Lighthart BT. Driven to be inactive?-The genetics of physical activity. Prog Mol Biol Transl Sci. 2010;94:271–90.
12. Blanergo J, Hunt MW. In: Bouchard C, Hoffman E, editors. Characterizing the extent of human genetic variation for performance-related traits, in Genetic and Molecular Aspects of Sports Performance: Blackwell Publishing Ltd; p. 33–45.
13. Rankinen T. In: Bouchard C, Katzmarzyk P, editors. Genetics of physical activity level, in Physical Activity and Obesity. Champaign, IL: Human Kinetics, 2010. p. 73–6.
14. Roth SM, Rankinen T, Hagberg JM, Loos RJ, Pérusse L, Sarzynski MA, et al. Advances in exercise, fitness, and performance genomics in 2011. Med Sci Sports Exerc. 2012;44(S4):S809–17.
15. Rankinen T, Bouchard C. Gene-exercise interactions. Prog Mol Biol Transl Sci. 2012;108:447–60.
16. Horimoto AR, Girão SR, Oliveira CM, Alvim RO, Soler JP, de Andrade M, et al. Heritability of physical activity traits in Brazilian families: the Baependi Heart Study. BMC Med Genet. 2011;12:155.
17. Mitchell BD, Rainwater DL, Hsu WC, Kennedy AJ, Stern MP, MacCluer JW. Familial aggregation of nutrient intake and physical activity: results from the San Antonio Family Heart Study. Annals of Epidemiology. 2003;13(2):128–35.
18. Perusse L, TrembLAY A, Leblanc C, Bouchard C. Genetic and environmental influences on level of habitual physical activity and exercise participation. Am J Epidemiol. 1989;129(5):1012–22.
19. Seabra AF, Mendonça DM, Goring HH, Thomis MA, Maia JA. Genetic and environmental factors in familial clustering in physical activity. Eur J Epidemiol. 2008;23(8):205–11.
20. Simonen RL, Perusse L, Rankinen T, Rice T, Rao DC, Bouchard C. Familial aggregation of physical activity levels in the Quebec family study. Med Sci Sports Exerc. 2002;34(7):1137–42.
21. Choh AC, Demerath EW, Lee M, Williams KD, Towne B, Siewogel RM, et al. Genetic analysis of self-reported physical activity and adiposity: the Southwest Ohio Family Study. Public Health Nutr. 2009;12(8):1052–60.
22. Diego V, et al. Genotype x sex interaction analyses identify a region on chromosome 20 contributing to anthropometric measures of obesity in the San Antonio Family Heart Study. 2006;14(9): p. 1267.
23. Voruganti VS, Diego VP, Haack K, Cole SA, Blanergo J, Göring HH, et al. A QTL for genotype by sex specific interaction for anthropometric measurements in Alaskan Eskimos on chromosome 19q12-13. The GOCADAN Study. Obesity. 2006;14:14.
24. Craig CL, Marshall AL, Sjöström M, Bauman AE, Booth ML, Ainsworth BE, et al. International physical activity questionnaire: 12-country reliability and validity. Med Sci Sports Exerc. 2003;35(8):1381–95.
25. IPAQ. Guidelines for Data Processing and Analysis of the International Physical Activity Questionnaire. 2005: Available from: https://sites.google.com/site/ipaq/scoring-protocol.
26. Diego V, Rainwater DL, Wang XL, Cole SA, Curran JE, Johnson MP, et al. Genotype x adiposity interaction linkage analyses reveal a locus on chromosome 1 for lipoprotein-associated phospholipase A2, a marker of inflammation and oxidative stress. Am J Hum Genet. 2007;80(1):168–77.
27. Peng B, Yu RK, Dehoff KL, Amos CI. Normalizing a large number of quantitative traits using empirical normal quantile transformation. BMC Proc. 2007;1Suppl:S156.
28. Fears TR, Bencichou J, Gail MH. A reminder of the fallibility of the Wald statistic. J Am Statist Assoc. 1996;90(430):226–7.
29. Rao CR. In: Balakrishnan N, Kannan N, Nagaraja HN, editors. Score test. Historical review and recent developments, in Advances in Ranking and Selection, Multiple Comparisons, and Reliability Methodology and Applications. Boston: Birkhäuser; 2005. p. 3–20.
30. Dominicus A, Skovdla A, Gesing HK, Pedersen NL, Palmeberg J. Likelihood ratio tests in behavioral genetics: problems and solutions. Behav Genet. 2006;36(2):331–40.
31. Self SG, Liang KY. Asymptotic Properties of Maximum-Likelihood Estimators and Likelihood Ratio Tests under Nonstandard Conditions. J Am Stat Assoc. 1987;82(398):605–10.
32. Neale MC, Miller MB. The use of likelihood-based confidence intervals in genetic models. Behav Genet. 1997;27(2):113–20.
33. Severini TA. Likelihood Methods in Statistics. Oxford: Oxford University Press; 2000.
34. Nakagawa S, Cuthill IC. Effect size, confidence interval and statistical significance: a practical guide for biologists. Biol Rev Camb Philos Soc. 2007;82(4):591–605.
35. Blangero J, Diego VP, Dyer TD, Almeida M, Peralta J, Kent Jr JW, et al. A kernel of truth: statistical advances in polygenic variance component models for complex human pedigrees. Adv Genet. 2013;81:1–31.

36. Lovejoy JC, Sainsbury A, Grp SCW. Sex differences in obesity and the regulation of energy homeostasis. Obes Rev. 2009;10(2):154–67.

37. Shi H, Seeley RJ, Clegg DJ. Sexual differences in the control of energy homeostasis. Front Neuroendocrinol. 2009;30(3):396–404.

38. Stevens J, Katz EG, Huxley RR. Associations between gender, age and waist circumference. Eur J Clin Nutr. 2010;64(1):16–15.

39. de Vilhena e Santos DM, Katzmarzyk PT, Seabra AF, Maia JA. Genetics of physical activity and physical inactivity in humans. Behav Genet. 2012;42(4):559–78.

40. Feldman DE, Barnett T, Shrier I, Rossignol M, Abenhaim L. Is physical activity differentially associated with different types of sedentary pursuits? Arch Pediatr Adolesc Med. 2003;157(8):797–802.

41. Marshall SJ, Biddle SJ, Gorely T, Cameron N, Murddy I. Relationships between media use, body fatness and physical activity in children and youth: a meta-analysis. Int J Obes Relat Metab Disord. 2004;28(10):1238–46.

42. Nilsson A, Andersen LB, Ommundsen Y, Froberg K, Sardinha LB, Pielbl-Aulin K, et al. Correlates of objectively assessed physical activity and sedentary time in children: a cross-sectional study (The European Youth Heart Study). BMC Public Health. 2009;9:322.

43. Smith NE, Rhodes RE, Naylor PJ, McKay HA. Exploring moderators of the relationship between physical activity behaviors and television viewing in elementary school children. Am J Health Promot. 2008;22(4):231–6.

44. Taveras EM, Feld AE, Berkey CS, Rifas-Shiman SL, Frazier AL, Colditz GA, et al. Longitudinal relationship between television viewing and leisure-time physical activity during adolescence. Pediatrics. 2007;119(2):e314–9.

45. Aaltonen S, Ortega-Alonso A, Kujala UM, Kaprio J. The heritability of consistent leisure time physical activity and inactivity in the Finnish Twin Cohort. Med Sci Sports Exerc. 2010;41(5, Suppl 1):519–20.

46. Beunen G, Thomis M. Genetic determinants of sports participation and daily physical activity. Int J Obes Relat Metab Disord. 1999;23 Suppl 3:S55–63.

47. Borecki IB, Rao DC, Province MA, editors. Familial Resemblance and Heritability, in Genetics Dissection of Complex. San Diego; 2001.

48. Boomsma DI, van den Bree MB, Orlebeke JF, Molenaar PC. Resemblances of parents and twins in sports participation and heart rate. Behav Genet. 1989;19(1):25–41.

49. Carlson S, Anderson T, Lichtenstein P, Michaelsson K, Ahlbom A. Genetic effects on physical activity: results from the Swedish Twin Registry. Med Sci Sports Exerc. 2006;38(8):1396–401.

50. Stubbe JH, Boomsma DI, Vink JM, Cornes RK, Martin NG, Skythte A, et al. Genetic influences on exercise participation in 37,051 twin pairs from seven countries. PLoS One. 2006;1:e222.