Gastrin-releasing Peptide Receptor Imaging in Breast Cancer Using the Receptor Antagonist $^{68}$Ga-RM2 And PET

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Abstract

Introduction: The gastrin-releasing peptide receptor (GRPR) is overexpressed in breast cancer. The present study evaluates GRPR imaging as a novel imaging modality in breast cancer by employing positron emission tomography (PET) and the GRPR antagonist $^{68}$Ga-RM2.

Methods: Fifteen female patients with biopsy confirmed primary breast carcinoma (3 bilateral tumors; median clinical stage IIB) underwent $^{68}$Ga-RM2-PET/CT for pretreatment staging. In vivo tumor uptake of $^{68}$Ga-RM2 was correlated with estrogen (ER) and progesterone (PR) receptor expression, HER2/neu status and MIB-1 proliferation index in breast core biopsy specimens.

Results: 13/18 tumors demonstrated strongly increased $^{68}$Ga-RM2 uptake compared to normal breast tissue (defined as PET-positive). All PET-positive primary tumors were ER- and PR-positive (13/13) in contrast to only 1/5 PET-negative tumors. Mean SUV$_\text{MAX}$ of ER-positive tumors was 10.6±6.0 compared to 2.3±1.0 in ER-negative tumors ($p=0.016$). In a multivariate analysis including ER, PR, HER2/neu and MIB-1, only ER expression predicted $^{68}$Ga-RM2 uptake (model: $r^2=0.55$, $p=0.025$). Normal breast tissue showed inter- and intraindividually variable, moderate GRPR binding (SUV$_\text{MAX}$ 2.3±1.0), while physiological uptake of other organs was considerably less except pancreas. Of note, $^{68}$Ga-RM2-PET/CT detected internal mammary lymph nodes with high $^{68}$Ga-RM2 uptake (n=8), a contralateral axillary lymph node metastasis (verified by biopsy) and bone metastases (n=1; not detected by bone scan and CT).

Conclusion: Our study demonstrates that $^{68}$Ga-RM2-PET/CT is a promising imaging method in ER-positive breast cancer. In vivo GRPR binding assessed by $^{68}$Ga-RM2-PET/CT correlated with ER expression in primary tumors of untreated patients.

Key words: GRPR, gastrin-releasing peptide receptor, bombesin, PET, positron emission tomography, breast cancer, ER, estrogen receptor.
Introduction

The gastrin-releasing peptide receptor (GRPR) is a promising target for molecular imaging and targeted therapy, showing an overexpression in several human tumors such as prostate carcinoma, breast cancer (BC), and peritumoral vessels in ovarian cancer [1, 2]. Using autoradiography Reubi et al found a GRPR expression of high density in 72-74 % of ductal BC specimens [2, 3], while Dalm et al. observed an even higher fraction of GRPR-positive samples of 96 % [4]. GRPR is a subtype of the bombesin receptor family with the physiologic ligand gastrin releasing peptide (GRP). GRP has various physiologic functions, including the release of gastrin and regulation of enteric motor function. GRP and GRPR also appear to play a (so far poorly characterized) role in human carcinogenesis and tumor proliferation [5-8]. Several GRPR agonists have been radiolabeled and tested preclinically [9, 10] and clinically [11, 12]. However, recent studies have indicated that GRPR antagonists show superior tumor uptake and image contrast, when compared to GRPR agonists [13, 14]. Furthermore, GRPR antagonists allow for a safer clinical use since no acute adverse effects (e.g., gastrointestinal) are expected.

In the past years, there have been several preclinical and clinical studies in prostate cancer targeting the GRPR [12, 15-18]. In contrast, there are only few studies focusing on GRPR imaging of BC. In a recent preclinical study Dalm et al. successfully imaged BC in mouse xenografts with small animal single-photon emission computed tomography (SPECT/CT) using a GRPR antagonist [4]. Prignon et al. visualized BC xenografts in mice using a 68Ga-labeled GRPR positron emission tomography (SPECT/CT) using a GRPR antagonist [5]. In a consecutive study, 68Ga-RM2-PET/CT showed a good diagnostic accuracy for the diagnosis of primary prostate carcinoma [25]. On BC, no studies using this tracer have been reported to date.

The different molecular subtypes of BC include luminal A, luminal B, luminal B-like, Her2-neu positive and triple negative subtypes. While the luminal subtypes show high estrogen receptor (ER) expression and represent the most common subtypes with an excellent prognosis, the ER-low or ER-negative subtypes have higher recurrence rates and poorer prognosis [26]. Therefore, novel molecular diagnostic biomarkers may be helpful to optimize subtype-specific treatment in BC. Moreover, in vitro studies suggest a positive correlation between ER and GRPR expression [4, 27]. Thus, GRPR-PET/CT imaging may also provide information about ER receptor status of BC.

The aim of the present analysis was to evaluate the feasibility and possible value of GRPR-PET/CT in BC in a clinical setting and to determine if GRPR expression in BC is associated with typical prognostic parameters such as ER, progesterone receptor (PR) and HER2-neu expression, MIB-1 proliferation index and patient age.

Material & Methods

Patients

Fifteen female patients with newly diagnosed primary unilateral or bilateral BC in neoadjuvant and adjuvant treatment situation underwent 68Ga-RM2-PET/CT for staging purposes between July 2014 and February 2015 on compassionate-use basis. We intentionally only included patients with suspected locally advanced breast cancer as these patients presumably profit most from an extended staging. Furthermore, only patients were recruited that did not undergo any preceding local or systemic therapies that might interfere with GRPR binding. All patients gave written informed consent and the present data analysis was approved by the institutional ethical review board (496/14). The patient age (mean ± standard deviation) was 54.5 ± 12.5 years (range 33 – 75 years).

Breast Core Biopsy

BC diagnosis was confirmed by core needle biopsy prior to 68Ga-RM2-PET/CT. In the case of clinically suspicious axillary lymph nodes an evaluation by core needle biopsy of lymph nodes was also performed. Time span between biopsy and 68Ga-RM2-PET/CT was 14.9 ± 8.1 days (range 7 – 37 days).

In addition, suspicious findings on PET/CT
were re-assessed clinically by certified BC specialists; possibly affected axillary lymph nodes were verified by core needle biopsy, whereas internal mammary lymph nodes are not routinely assessed by ultrasound and biopsy at our institution.

**Immunohistochemistry**

Tissue obtained by breast core biopsy was analyzed for ER and PR status, HER2/neu expression and MIB-1 proliferation index. Therefore, five serial tissue slices of 2 µm thickness were prepared for immunohistochemistry (IHC) using a Leica RM2255 Microtome and stained afterwards for ER, PR, HER2/neu and MIB-1. Ready-to-use antibodies for ER (IR657, Clone 1D5), PR (IR068, Clone 636) and HER2/neu (A0485) were used for antigen detection. All slides were stained with Dako Real® Detection System (Dako K5001) according to the instructions of the manufacturer. Omission of primary antibody served as the negative control. For ER, PR and MIB-1, nuclear staining of non-tumoral mammary glands was used as internal positive control. For HER2/neu external controls form part of every run. A nuclear reactivity of ER and PR in > 1 % of the tumor cells was rated as a positive reaction. HER2/neu overexpression (HER 3+) was defined as an intense and complete membrane staining in more than 10 % of the tumor cells.

**Tracer synthesis**

The RM2 precursor was provided by Piramal Imaging (Berlin, Germany). Radiolabelling of RM2 with 68GaCl3 was accomplished using a fully automated synthesis module (Pharmtracer, Eckert & Ziegler, Berlin, Germany). The automated preparation was done according to Good Manufacturing Practice under sterile conditions. Briefly, the 68Ge/68Ga generator (IGG 100, Eckert & Ziegler) was eluted with 0.1 M HCl. Chemical purification and concentration of the generator eluate was carried out using a cation exchange resin (Strata x-c, Phenomenex). 68Ga (III) was eluted from the cartridge into the reaction vial using a 97.6 % acetonitrile/0.02 N HCl solution. The reaction vial contained 60 µg RM2 in 2 mL sodium acetate buffer (0.2 M, pH 4.0) and 200 µL ethanol. Labelling was accomplished by heating the reaction mixture at 95°C for 10 min. For further purification, the solution was passed over a C18 light cartridge (Waters, USA), washed with 3 mL saline and eluted with 1 mL 50 % ethanol. The final product was constituted by addition of 7 mL saline and sterilized by filtration using Millex 0.22 µm filter (Millipore, USA). Quality control was performed using an Agilent 1260 HPLC system with a Gina radioactivity detector (Raytest, Germany) in combination with a ACE 3 C18 (150 x 4.6 mm) column. The solvent system was A H2O (0.1 % TFA) and B acetonitrile (0.1 % TFA). The gradient was 0 – 8 min 28 % B, 8 – 9 min 60 % B, 9 – 14 min 60 % B, 14 – 15 min 28 % B at a flow rate of 0.6 mL/min. The decay-corrected yield was > 90 % and the radiochemical purity of the final product was > 98 %. The product was sterile and pyrogen-free (< 0.5 EU/mL).

**68Ga-RM2-PET/CT**

All patients received an intravenous injection of 118 – 213 MBq (3.19 – 5.76 mCi) 68Ga-RM2 (peptide dose 26 – 56 µg). Whole-body PET/CT scans were performed 1 h p.i. (2 min per bed position, contrast-enhanced diagnostic CT – 120 keV, 100 – 250 mAs, dose modulation, FOV 600 mm², 512 x 512 matrix, slice thickness 2 mm) from the base of the skull to the proximal femur (64-slice GEMINI TF PET/CT, Philips Healthcare, Cleveland, USA). PET data were fully corrected for attenuation, scatter, decay and randoms and expressed as standardized uptake values (SUV; i.e., local radioactivity concentration normalized to injected dose per body weight).

**PET/CT analysis**

PET images were analyzed by two experienced nuclear medicine physicians in consensus. PET-positivity was defined as focal tracer uptake of the primary tumor beyond local background in correlation with CT. Volume-of-interest (VOI) analysis was performed using PMOD (Ver. 3.205, PMOD Technologies, Zürich, Switzerland). The primary tumor was delineated under CT guidance on all transaxial slices and maximum SUV (SUVMAX) was calculated. Circular regions of interest (ROI) were placed on normal breast tissue (NBT) (diameter 30 mm), liver (diameter 40 mm), pancreas (diameter 20 mm), muscle (diameter 30 mm) and adipose tissue (diameter 30 mm) and mean SUV (SUVMEAN) as well as SUVMAX were extracted. Lymph node sizes were assessed on contrast-enhanced diagnostic CT (CT portion of the PET/CT) measuring the long- and short-axis diameters on transaxial slices (voxel size 1.17 x 1.17 x 2 mm³).

**Statistical Analysis**

Analyses were performed using JMP (Ver. 11; SAS Institute, Cary, USA). PET positivity and SUV values were correlated with ER, PR, HER2/neu status and MIB-1 proliferation index in univariate analysis (Fisher’s exact test for nominal and Spearman’s rho (p) for ordinal/continuous data). In addition, a multivariate analysis was performed with ER and PR expression, MIB-1 proliferation index (each expressed...
as percent) and HER2/neu status (binary categorical) as independent variables to predict the observed SUVMAX value (dependent variable).

**Results**

Three patients presented with bilateral tumors. Histological tumor types were 14 no special type (NST; formerly classified as invasive ductal carcinoma), 3 invasive lobular carcinoma (ILC) and 1 mucinous carcinoma (MC). Patient characteristics and imaging findings, including lymph node (LN) findings are summarized in Table 1. No adverse effects were observed during or following the administration of 68Ga-RM2.

NBT showed an intra- and interindividually variable, moderately strong 68Ga-RM2 binding with a SUVMEAN 1.1 ± 0.5 and an SUVMAX 2.3 ± 1.0. Physiological biodistribution demonstrated little 68Ga-RM2 uptake in fat (SUVMEAN 0.2 ± 0.04) and muscle (SUVMEAN 0.4 ± 0.10), moderate uptake in liver (SUVMEAN 1.1 ± 0.3) and intense uptake in pancreas (SUVMEAN 40.8 ± 9.8), which corresponds to earlier studies [11, 16]. Furthermore a weak negative association between 68Ga-RM2 uptake of NBT and patient age was observed, which however did not reach a level of statistical significance on univariate analysis (Spearman’s ρ = -0.36; p = 0.14).

13/18 tumors were clearly visualized by increased 68Ga-RM2 uptake compared to NBT and thus rated as PET-positive (Figure 1). Mean SUVMAX of PET-positive tumors was 11.2 ± 5.8 (range 3.9 – 22.5). 5/18 tumors could not be differentiated from NBT and thus were defined as PET-negative. Mean SUVMAX of PET-negative tumors was 2.4 ± 0.9 (range 1.4 – 3.5). However, in these cases, metastatic manifestations (i.e., metastatic axillary lymph nodes) could still be identified clearly in regions with low physiological tracer uptake (Figure 2). SUVMAX of the primary tumor did not correlate with patient age (p = 0.71).

All tumors rated PET-positive were also positive for ER and PR expression, in contrast to only 1/5 tumors visually rated PET-negative (Patient No. 13; SUVMAX 3.0). The single PET-negative, ER-positive tumor was a mucinous carcinoma with an immunohistochemical ER expression of 30% and a PR expression of 40%. Mean SUVMAX of ER-positive tumors was 10.6 ± 6.0 compared to 2.3 ± 1.0 in ER-negative tumors (p = 0.016). NST and ILC did show comparable SUVMAX values (NST 9.2 ± 6.9, ILC 8.9 ± 4.1; p = 0.92). All ER-positive tumors were also positive for PR status. PET positivity was significantly associated with ER status (Fisher’s exact test, p = 0.0016) and PR status (p = 0.0016), but not with HER2/neu status (p = 0.07). For quantitative PET measures (i.e., SUVMAX) and relative biomarker expressions (i.e., ER, PR, MIB-1 in percent), significant correlations were found for ER (Spearman’s ρ = 0.72, p = 0.0013) and PR (ρ = 0.50, p = 0.0003). As may be expected, ER and PR expression was correlated (ρ = 0.72, p = 0.0007). In a multivariate analysis including ER, PR, HER2/neu and MIB-1 as predictor variables, only ER expression predicted 68Ga-RM2 uptake (model: r² = 0.55, p = 0.025; parameter estimates: ER p = 0.0075, all other parameters p > 0.15) (Figure 3).

Table 1. Patient biopsy characteristics and imaging findings.

| Patient No | Type/Side | Immunohistochemistry | Image findings |
|------------|-----------|----------------------|----------------|
|            |           | ER | PR | HER2/neu | MIB-1 Index | PET Rating | LN No. and location | LN size /mm (CT) |
| 1          | NST / bilateral | + | + | - | 40% | pos | left: > 5 ax / 3 IMLN | 3x3 – 11x7 |
| 2          | NST / left | + | + | - | 40% | pos | > 5 ax / 4 IMLN | 15x10 – 11x6 |
| 3          | ILC / right | + | + | - | 22% | pos | 1 ax | 10x9 |
| 4          | NST / right | - | - | - | 90% | neg | 2 IMLN | 5x3 – 24x20 |
| 5          | NST / right | + | + | - | 8% | pos | 1 ax | 14x6 |
| 6          | ILC / right | + | + | - | 30% | pos | / | / |
| 7          | NST / bilateral | right: + | right + | left: + | right: 18% | right: pos | / | / |
| 8          | NST / left | - | - | - | 80% | neg | 5 IMLN | 7x7 – 11x14 |
| 9          | NST / bilateral | + | + | - | right: 18% | pos | right: 2 ax / 1 IMLN | 3x4 – 11x5 |
| 10         | ILC / left | + | + | - | 15% | pos | / | / |
| 11         | NST / right | + | + | - | 16% | pos | / | / |
| 12         | NST / right | - | - | - | 60% | neg | > 5 ax / 1 IMLN / 1 sc / 1 spc | 6x6 – 9x9 |
| 13         | MC / right | + | + | - | 20% | neg | > 5 ax | 9x8 – 42x24 |
| 14         | NST / right | + | + | - | 23% | pos | > 5 ax / 5 IMLN / 2 med / 1 hil | 3x4 – 22x15 |
| 15         | NST / left | + | + | - | 38% | pos | > 5 ax / 4 IMLN / 9 spc / 1 cl 3x3 – 14x19 |

Abbreviations: pos, positive; neg, negative; LN, lymph node; ax, axillary; IMLN, internal mammary lymph node; med, mediastinal; hil, hilar; spc, subpectoral; scl, supraclavicular; cl, contralateral axillary; for additional abbreviations see text.
Figure 1. 74-year-old patient (No. 7) with a bilateral NST with an ER/PR-positive tumor on the right side (PET-positive; SUV_{MAX} 8.32) and an ER/PR-negative tumor on the left side (PET-negative; SUV_{MAX} 2.68). Maximum intensity projection (left); CT (upper row); \(^{68}\)Ga-RM2-PET (lower row); primary tumors indicated by red arrows. Note the physiological uptake in the pancreas, esophagus and rectum. For abbreviations see text.

Figure 2. 50-year-old patient (No. 13) with a mucinous carcinoma of the right breast with low ER/PR-expression. Note that the primary tumor could not be distinguished from normal breast tissue, while an ipsilateral axillary lymph node metastasis (histologically proven; SUV_{MAX} 2.9) was identified by increased \(^{68}\)Ga-RM2-uptake relative to the surrounding tissue. CT (left); \(^{68}\)Ga-RM2-PET (middle); fusion images (right); primary tumor indicated by white arrow; lymph node metastasis indicated by red arrows. For abbreviations see text.

Figure 3. Result of the multivariate analysis. Observed SUV_{MAX} values (y-axis) were plotted over predicted SUV_{MAX} values (x-axis; included predictor variables: ER and PR expression, MIB-1 proliferation index (each expressed as percent) and HER2/neu status (binary categorical)). ER expression is the only statistically significant predictor of SUV_{MAX}. Symbols: filled circles, no special type; open squares, invasive lobular carcinomas; open triangle, mucinous carcinoma. For abbreviations see text.

Given the paucity of data on the clinical diagnostic value of GRPR-PET, suspicious findings of possible therapeutic relevance were only accepted as true findings if confirmed by CT or biopsy. \(^{68}\)Ga-RM2-PET/CT revealed a possible upstaging compared to conventional clinical staging (i.e. breast, axillary and abdominal ultrasound, chest x-ray and bone scan; conventional clinical staging does not include ultrasound of internal mammary lymph nodes or FDG-PET/CT) in 7/15 (47 %) of the examined BC patients including 6 to stage III and 1 to stage IV (Table 2). In 8 patients internal mammary lymph nodes (IMLN), strongly positive on \(^{68}\)Ga-RM2-PET and rated as suspicious for malignancy by CT (size 3 to 19 mm), were detected, in line with an possible upstaging to stage IIIA and IIIC in 6 patients (Figure 4A). Upstaging to stage IV
occurred in one patient due to the detection of a contralateral axillary lymph node metastasis verified by biopsy (Figure 4B).

In one patient several bone metastases were detected on bone scan and CT. All lesions also showed high $^{68}$Ga-RM2 uptake. However, several additional lesions were depicted by increased focal $^{68}$Ga-RM2 uptake in PET, but not detected by CT or bone scan (Figure 4C).

Table 2. Primary clinical and $^{68}$Ga-RM2-PET/CT stages 1.

| Primary clinical stage | Total cases | $^{68}$Ga-RM2-PET/CT stage | Upstaged cases n (%) |
|------------------------|-------------|-----------------------------|---------------------|
| IA                     | 2           | IIA                         | 0 (0)               |
| IIA                    | 5           | IIB                         | 3 (60)              |
| IIIB                   | 3           | IIIA                        | 2 (66)              |
| IIIA                   | 3           | IIIB                        | 1 (33)              |
| IIIB                   | 1           | IV                          | 0 (0)               |
| IIIC                   | -           | -                           | -                   |
| IV                     | 1           | Total                       | 7 (47)              |

1 According to the 7th Ed. of the AJCC Cancer Staging Manual [49].
*Patient with a bilateral tumors.

Discussion

The present study demonstrates that $^{68}$Ga-RM2-PET/CT is capable of in vivo visualizing GRPR expression with high contrast in the majority of the examined BC patients (73 %; 11/15). Our data are in accordance with the in vitro results obtained by Reubi et al. [2, 3]. Apart from NST, we were also able to clearly visualize all ILC, which may be missed by conventional FDG-PET/CT due to often limited FDG avidity [28]. While univariate analyses revealed a strong association between GRPR binding and ER as well as PR status, a multivariate regression analysis identified ER as the primary predictor of $^{68}$Ga-RM2 uptake. Therefore, $^{68}$Ga-RM2-PET/CT may provide a promising imaging modality with high image quality in patients with ER-positive tumors, representing the majority of BC cases (81 % in a recent study examining a diverse patient population [29]).
The positive correlation between GRPR binding and ER expression, which we found in our in vivo study, is in line with previous in vitro studies: Halmos et al. showed a significant association between high-affinity binding of the GRPR agonist $^{125}$I-Tyr$^4$-bombesin on isolated cell membranes and estrogen receptor expression in BC biopsy specimens ($r = 0.671, p < 0.005$) [27]. Similarly, Dalm et al. found a significant positive correlation ($p = 0.026$) between the extent of GRPR expression detected by autoradiography with the GRPR agonist $^{111}$In-AMBA (5-step score) and ER status in BC tissue specimens [4]. This was confirmed by a subsequent study by the same group, demonstrating a significant positive association ($p < 0.001$) between GRPR messenger RNA (mRNA) and ER positivity in BC [30]. Interestingly, GRPR mRNA levels correlated significantly with prolonged progression-free survival in metastatic ER-positive BC patients undergoing tamoxifen treatment [30]. These findings underline the potential clinical relevance of GRPR imaging in BC patients.

To the best of our knowledge the mechanism for the correlation between ER expression and the expression of GRPR and other G protein couple receptors are not well understood are not well understood and needs further studies [31] [32].

Albeit our findings warrant verification by larger patient series, it is tempting to speculate about potential clinical indications of GRPR PET in BC patients. For instance, in clinical practice the hormone receptor status is usually already known at time of the initial diagnosis by biopsy. This information could be used for a personalized selection of the staging strategy that may include GRPR PET/CT in ER-positive tumors. Furthermore, GRPR PET/CT may allow investigators to assess ER expression levels non-invasively in whole-body imaging studies. This could provide unique opportunities to study the heterogeneity of ER expression. Of note, tumor heterogeneity between the primary tumor and synchronous metastases seems to be less than in the situation of metachronous metastases [33]. On one hand, this supports the use of GRPR PET for initial staging of known ER-positive primary tumors. On the other, if the correlation between GRPR binding and ER expression holds true also in the course of the disease and its various treatments, GRPR PET may also be helpful to uncover a possibly increasing heterogeneity of the ER status over time (e.g., loss of ER expression or occurrence of new ER-negative manifestations). This may be of important therapeutic and prognostic implications (e.g., for endocrine therapy or targeted radionuclide therapy).

ER status of BC can be images with the ER ligand $^{16}$α-[18F]-fluoro-17β-oestradiol (FES) PET imaging of in vivo ER expression by employing ER-specific ligands. In a meta-analysis of 4 studies, van Kruchten et al. calculated an overall sensitivity of FES-PET of 84 % for the detection of ER-positive BC [34]. The correlation coefficients for the relationship of $^{18}$F-FES uptake and ER expression ranged from 0.56 – 0.96 [34-36]. In our study, $^{68}$Ga-RM2 binding to GRPR correlated well with ER expression (Spearman’s $\rho = 0.70, p = 0.0013$), which is in the same range as for $^{18}$F-FES. Possible advantages of $^{68}$Ga-RM2-PET/CT compared to $^{18}$F-FES-PET/CT include a more favorable biodistribution for tumor staging due to much lower liver and intestinal radiotracer uptake. In addition, $^{68}$Ga-RM2 binding to GRPR is less likely to be affected by endogenous estrogen levels (e.g., differences between pre- and postmenopausal patients), sex hormone-binding globulin serum levels and anti-estrogen treatment. However, it remains to be seen if and to what extent these factors affect GRPR expression and thus $^{68}$Ga-RM2 uptake as a prerequisite for the possible use of GRPR PET in pre-treated patients and for treatment monitoring.

FDG-PET/CT is a valuable diagnostic tool for the detection of distant metastases [37, 38]. Its role for evaluation of locoregional lymph node status on the other hand is controversial. According to a recent review the sensitivity of FDG-PET/CT for the detection of axillary lymph nodes ranges between 24 and 82 % [39]. There are some limitations of FDG-PET imaging that may lead to reduced diagnostic accuracy. Inflammatory reactions may mimic tumor infestation and thus lead to false positive results [40]. Small tumor manifestations might be missed by PET due to the limited spatial resolution and contrast compared to background. In the present study $^{68}$Ga-RM2-PET/CT allowed us to visualize affected lymph nodes less than 5 mm in maximum diameter due the very low radiotracer uptake in muscles and fat tissue. The low background also allows for the detection of lymph node metastases of primary tumors with little GRPR expression.

In our preliminary analysis, we found a potential clinical upstaging of 47 % of the examined patients (7/15) after $^{68}$Ga-RM2-PET/CT due to the detection of suspicious lymph nodes. $^{68}$Ga-RM2-PET/CT showed a high detection rate for suspicious IMLN (53 %; 8/15), leading to potential upstaging of 40 % of the patients (6/15). A recently published study using FDG-PET/CT found suspicious IMLN in 62/216 patients (29 %) [41]. We hypothesize that the high detection rate observed in our study is due to the intense uptake of $^{68}$Ga-RM2-PET/CT by ER-positive BC and the very low physiologic uptake of $^{68}$Ga-RM2-PET/CT in fat and muscle tissue. However, this hypothesis needs to be tested in studies.
performing an intra-individual comparison of FDG-PET/CT and $^{68}$Ga-RM2-PET/CT.

The clinical relevance of suspicious internal mammary lymph nodes on patient outcome has been discussed controversially in literature. Hennequin et al. for example show no apparent survival benefit when performing external beam irradiation of IMLNs [42], while Chang et al. suggest a significant longer disease-free survival [43]. Regarding the detection of distant bone metastases, $^{68}$Ga-RM2-PET not only identified metastases also detected by CT and bone scan, but also showed suspicious bone lesions solely positive on $^{68}$Ga-RM2-PET. Interestingly an earlier study using a GRPR agonist and scintigraphic imaging in tamoxifen-resistant BC (n = 5) could not identify known bone metastases [21]. While these results are highly promising for imaging ER-positive tumors, further evaluation in larger patient series and systematic histologic validation are obviously needed.

In addition to diagnosis, GRPR targeting could provide a new therapeutic approach via peptide receptor radionuclide therapy (PRRT) in patients with GRPR-positive BC, as already practiced for neuroendocrine tumors, where somatostatin receptor-targeting PRRT has become a very important asset [44-47]. GRPR antagonists, such as RM2, labeled with a therapeutic radioisotope (e.g., lutetium-177) could be used to treat BC patients while using Ga-68 labeled RM2 as a diagnostic companion in a theranostic treatment concept. We observed high physiological uptake in pancreatic tissue, which according to GRPR blockade studies in mice is most likely due to specific GRPR binding [48]. Given the faster washout of GRPR antagonists from the pancreas than from prostate cancer tissue observed in previous preclinical [15, 18] and clinical [16] studies, the radiation dose to the pancreas is expected to be significantly lower than the radiation dose to tumor manifestations. Dosimetric studies employing long-lived nuclides (e.g., Lu-177) and dynamic data acquisition are needed to explore in how far pancreatic uptake may limit the therapeutic application of GRPR antagonists compared to other small molecules (e.g., ER-targeting agents). However, in a murine prostate cancer model long-term tumor regression could be achieved by treatment with $^{177}$Lu-DOTA-RM2 without evidence for damage to the pancreas [15].

Limitations of our study are the relatively low number of patients, which renders our multivariate analysis preliminary. Albeit our preliminary results suggest a diagnostic value of $^{68}$Ga-RM2-PET/CT in ER-positive BC (i.e., sensitivity of 93% for ER-positive tumors, upstaging in 47% of patients), this needs to be verified by future studies. For calculation of diagnostic parameters like sensitivity, specificity and accuracy, studies need to include larger patient cohorts with various suspicious breast lesions and to pursue a comprehensive pathological verification of detected lesions (i.e., with and without increased $^{68}$Ga-RM2 uptake, breast and metastatic lesions). The present patient sample comprises only patients with known BC, while possible false-positive lesions (e.g., fibroadenoma [22] were not observed. Furthermore, not all suspicious lymph nodes (in particular IMLN) detected by $^{68}$Ga-RM2-PET/CT were histologically confirmed. Thus, it has to be emphasized that aforementioned results concerning the potential upstaging of patients need further validation. Finally, we did not pursue a detailed head-to-head comparison of FDG-PET with $^{68}$Ga-RM2-PET/CT. Such studies are needed to define possible benefits of one method over the other for a given clinical condition (e.g., tumor types NST vs. ILM, ER status positive vs. negative) and method-specific drawbacks (e.g., physiological GRPR expression in breast tissue or increased FDG uptake in inflammatory lymph nodes).

**Conclusion**

Our study demonstrates that $^{68}$Ga-RM2-PET/CT is a promising imaging method in BC patients, especially for ER-positive tumors. *In vivo* GRPR binding assessed by $^{68}$Ga-RM2-PET/CT correlated with ER expression in primary tumors of untreated BC patients.

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**Competing Interests**

Philipp T Meyer and Wolfgang A Weber receive financial support from Piramal Imaging for an ongoing research project (outside the submitted work). Helmut R Maecke is a co-inventor of a patent for bombesin receptor antagonists.

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