Constructing Biological Pathways by a Two-Step Counting Approach

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Abstract

Networks are widely used in biology to represent the relationships between genes and gene functions. In Boolean biological models, it is mainly assumed that there are two states to represent a gene: on-state and off-state. It is typically assumed that the relationship between two genes can be characterized by two kinds of pairwise relationships: similarity and prerequisite. Many approaches have been proposed in the literature to reconstruct biological relationships. In this article, we propose a two-step method to reconstruct the biological pathway when the binary array data have measurement error. For a pair of genes in a sample, the first step of this approach is to assign counting numbers for every relationship and select the relationship with counting number greater than a threshold. The second step is to calculate the asymptotic p-values for hypotheses of possible relationships and select relationships with a large p-value. This new method has the advantages of easy calculation for the counting numbers and simple closed forms for the p-value. The simulation study and real data example show that the two-step counting method can accurately reconstruct the biological pathway and outperform the existing methods. Compared with the other existing methods, this two-step method can provide a more accurate and efficient alternative approach for reconstructing the biological network.

Introduction

One great challenge of postgenomic research is to explore complex biological pathways from genomic data such as DNA sequences, protein sequences, and gene expression profiles. The network building method is widely used throughout biology to reconstruct complex biological pathways.

We take MAPK pathway as an example. The MAPK/ERK pathway is a signal transduction pathway that couples intracellular responses to the binding of growth factors to cell surface receptors. Robert et al. [1] and related studies [2–5] based on biology experiments provide the MAPK pathway (Figure 1).

It would be interesting if Figure 1 can be reconstructed in terms of their expression profile of Wsc1/2/3, Mid2,…, etc. To reduce the cost of experiments, one possibility is to predict the activation status of these genes through their microarray expression data for inferring the pathway.

There have been methods proposed in literature for reconstructing genetic regulatory networks in terms of microarray data. For instance, the Bayesian network model is an important technique that has been studied in the last two decades [6–8]. In addition, Wei and Li [9] proposed a hidden spatial-temporal Markov random field model to identify genes that are related to biological pathway. Allocco et al. [10] provided a variety of methods to find the gene-pairs with similarity relationship. Moreover, other algorithms using linear models [11,12], differential equation [11,13], neural network [14] and structural equation modeling [15] have been proposed to explore gene regulatory networks based on genomewide data. However, most of these methods have limitations in dealing with large-scale gene regulatory network because of their complex model structures. Also, careful discretization can be used to denoise high-throughput data. One such example can be found in Xing and Karp [16].

To overcome the disadvantage of the mentioned methods, we consider a simple model based on the Boolean network to reconstruct a large scale gene network in this study. Boolean networks have been proposed and investigated for a long time in literature. Kauffman [17,18] considered a dynamic version of Boolean networks. Liang et al. [19] proposed the algorithm REVEAL to infer gene regulatory network by calculating the Shannon entropy. Akutsu and Miyano [20] proposed an identification algorithm to reconstruct the Boolean network by comparing the collected data with all possible Boolean functions and input datasets. In order to make Boolean network more comprehensive, Shmulevich et al. [21] proposed the model of probability Boolean network (PBN). Moreover, for large-scale gene regulatory networks, Kim et al. [22] have used Boolean network with chi-square test on the yeast cell cycle microarray gene expression datasets. Markowetz et al. [23] proposed the nested effects model to infer the genetic network. Li et al. [24] made a comparison between the approaches of probabilistic Boolean network and dynamic Bayesian network. More recent developments are referred to Ay, Xu and Kahveci [25] and Davidich and Bornholdt [26].

In this article, we consider the directed acyclic Boolean (DAB) network as a tool for exploring biological pathways. Our goal is to...
construct a DAB network from the noisy array data. Since it involves noisy data, the reconstruction of the pathways cannot employ a deterministic inference. Instead, we need to establish a statistical model to capture its random characteristics. A DAB network is characterized by two kinds of pairwise relationships: similarity and prerequisite. The former represents a pair of elements with coherent on-off states. The latter is a partial order relationship, namely, the on-status of one element is a prerequisite for the on-status of another element. More specifically, if one element is a prerequisite to another element, the off-status of one element will restrict another element’s off-status. A DAB network is uniquely determined by its state space: all possible on-off states subjected to the pairwise relationships.

Recently, a Boolean implication network is proposed with similar aspect as the DAB network, which investigates all Boolean implications between pairs of genes for large-scale genome microarray datasets [27]. For any pair of elements, they use two statistics to test whether there is any specific relationship between the pair of elements. However, the methods are more applicable for dealing with mass information of datasets.

The approach of building a DAB network based on the expectation-maximization (EM) algorithm to derive the maximum likelihood estimator [28] for a statistical model is established in Li and Lu [29]. Their strategy is to build up a statistical model with measurement error and assign scores for the possible relationships between two genes, and then use the scores to select the true relationship. This method involves more computation and cannot provide a simple closed-form statistic to recover the true relationships between genes.

In this study, we propose a simple method to estimate pairwise relationships between elements from noisy array data. The approach is based on two steps: the first one is to count the numbers of different pairwise relationships in a sample, and the second one is to test the relationship hypotheses according to their asymptotic p-values. Compared with the Li and Lu [29] method, this new approach has a simple closed form and it is not time-consuming. In addition, the proposed counting approach shows substantial improvement compared to the Sahoo et al. [27] method. We conduct a simulation study to an example used in Li and Lu [29]. It is shown that the proposed method can recover all of the true relationships. A simulation study for a larger scale network is given in the supplementary material. In addition, the proposed method is implemented on the MAPK pathway example. It can recover true relationships among seven relationships, however, Li and Lu’s method only recovers one true relationship in this example. In this real data example, the new method shows a significant improvement in adopting a DAB network for exploring the pathway.

Methods

To describe the model and notations, we adopted a simple example used in Li and Lu [29] to illustrate the model assumption. Figure 2 shows the relationships of the seven elements in this example derived from the 13 states of Table 1. In the diagram, the notation $A \rightarrow B$ denotes that $A$ is a prerequisite of $B$ and the notation $E \sim B$ denotes that $B$ and $E$ are similar. Note that $A, \ldots, G$ in Figure 2 are called elements. The definitions of prerequisite and similar relationships for any two elements $A$ and $B$ are defined as follows.

Assume that an element only has two levels, on or off. We use “0” and “1” to represent “off” and “on” states respectively. For two elements $A$ and $B$, $A$ is a prerequisite for $B$ if the on-state of $A$ is necessary for the on-state of $B$, and we denote it by $A \rightarrow B$. When $A$ and $B$ are on and off simultaneously, the relationship between $A$ and $B$ is called similar and is denoted by $A \sim B$. We define $A$ to be the dual state of $A$. It means that $A = 0$ when $A = 1$.

![Diagram of a directed acyclic Boolean network with seven elements and twelve pair relationships. Only arrows between covering pairs are shown.](image_url)
There are 4 possible situations for the prerequisite relationship, and 2 possible situations for the similar relationship, see Table 2. Totally, there are 6 possible relationships for any two genes. The prerequisite relationship is a partial order. It is transitive on the ground-set, namely, \( A \prec B \) and \( B \prec C \) implies \( A \prec C \). The notations \( \sim_{+} \) and \( \sim_{-} \) in Table 2 denote the possible states of \( A \) and \( B \) and the impossible states of \( A \) and \( B \) under the relationship, respectively.

Let \((m_{00}, m_{01}, m_{10}, m_{11})\) and \((q_{00}, q_{01}, q_{10}, q_{11})\) denote the counts and probabilities corresponding to states \((A,B)=(0,0),(0,1),(1,0)\) and \((1,1)\) without measurement error. From the possible relationships shown in Table 2, we can propose a hypothesis corresponding to each relationship. For example, the first similar relationship in Table 2 is \( A \sim B \), which means that the two situations \((A,B)=(0,0)\) and \((A,B)=(1,1)\) hold. In this case, the probability of the two situations, \((A,B)=(0,1)\) and \((A,B)=(1,0)\), should be zero. Thus, its corresponding hypothesis is \( q_{01} = q_{10} = 0 \). Other situations follow a similar argument. The hypotheses for the 6 relationships are presented in Table 3. Under the measurement error model assumption, let \((n_{00}, n_{01}, n_{10}, n_{11})\) and \((r_{00}, r_{01}, r_{10}, r_{11})\) denote the counts and probabilities corresponding to states \((A,B)=(0,0),(0,1),(1,0)\) and \((1,1)\) with misclassification probability \(p\).

Because of the misclassification error, \(n_{00}\) may be split up into four categories. We use the notations \(m_{00,00}, m_{00,01}, m_{00,10}\) and \(m_{00,11}\) to represent the counts of four cells split from \(m_{00}\). Analogous notations are defined for \(m_{01}, m_{10}\) and \(m_{11}\). Consequently, their generating probabilities are calculated as follows: \(q_{k,l}=p^{\left(k^*-k\right)}\left(1-p\right)^{\left(l^*-l\right)}q_{k,l}\). Here, we adopt the notation \(q_{k,l}\) analogous to \(m_{k,l}\). The splitting counts and probabilities implied by misclassification error are given in Tables 4 and 5.

Now we go back to the example of Figure 2 which includes 7 elements. There are a total of \(2^7=128\) states for a seven-element network. Only thirteen of these states in Table 1 are compatible with the twelve pairwise relationships in the above example. From Figure 2, there are 12 true relationships between the elements, which are

\[
C < G, A < G, A < C, A < D, A < E, \\
B \sim E, A < F, F < G, C < D, A < B, C < F, D < G.
\]

Under the measurement error model assumption, we do not directly observe the 13 states but observe states with measurement error. We aim to reconstruct the true pathway. A proposed method is given in the following.

The two-step counting method

Suppose we have a sample \(S=(S_1,\ldots,S_n)\) of size \(n\) for \(m\) genes where \(S_i=(o_1, o_2, \ldots, o_m)\), \(o_i=0\) or 1. For example, in Table 1, there is a sample of size 13 for seven genes. We propose a two-step approach to recover their relationships.

The first step: counting

For a pair of genes, say \(A\) and \(B\), we can count the numbers for 6 relationships in Table 2 for the \(n\) states. The relationships with a counting number greater than a given threshold are regarded as potential relationships.

If there are no measurement errors, it is reasonable to expect that the counting number of two elements, say \(A\) and \(B\), satisfying the true relationship is exactly equal to \(n\). However, since it involves measurement errors, the counting number with respect to the true relationship may not be exactly equal to \(n\). For each pair
of elements, we count the numbers satisfying the 6 relationships respectively, say \(c_1, \ldots, c_6\). Since we expect that the misclassification probability is low, the counting number \(\leq n\) corresponding to the true relationship should be close to \(n\). Thus, we can select the relationships with a counting number greater than a threshold. The threshold selection is suggested as follows.

**Threshold Selection.** The suggested thresholds for the similar and prerequisite relationships are

\[
 n((1-w)^2 + w^2)
\]

and

\[
 n((1-w)^2 + w(1-w))
\]

respectively, where \(w = p + z_{\alpha/2}(p(1-p)/n)^{1/2}\). Here, the misclassification error probability \(p\) can be assumed to be known from empirical experiences. If \(p\) is unknown, the maximum likelihood approach for estimating \(p\) is given in Appendix D in the materials section.

The argument for the threshold selection is given in Appendix A in the materials section. It is based on a confidence bound approach associated with the counting number formulas. The approach is to derive the formulas for the two kinds of relationships, and then uses a confidence interval approach to obtain a lower bound for the counting number formulas. The forms \(n((1-w)^2 + w^2)\) and \(n((1-w)^2 + w(1-w))\) are inferred by the counting number formulas with misclassification probability \(p\), where \(w\) value is derived by a confidence bound approach.

The second step: asymptotic p-value

Besides directly counting the relationships’ numbers, the second step is to test the relationships in Table 3 using an asymptotic p-value. Then we combine both steps to estimate the true relationship between two elements.

The following simulation study shows that the two steps are both essential for selecting the true relationship. If any one of the steps is used solely in selecting the true relationship, the simulation shows that it cannot select the true relationships very accurately.

The p-values derived for the 6 hypotheses with misclassification probability \(p\) corresponding to the 6 relationships are listed as follows. The derivations are given in Appendix B in the materials section.

For testing \(H_0 : q_{10} = q_{01} = 0\) vs \(H_1 : q \neq H_0\), the asymptotic p-value for large sample size \(n\) is

\[
 1 - 2\Phi\left(\frac{(n_{10} + n_{01})/n - 2p(1-p)}{\sqrt{2(1-p)p(1-2p^2)/n}}\right),
\]

where \(\Phi\) is the cumulative distribution of the standard normal random distribution. The asymptotic p-value for testing \(H_0 : q_{10} = q_{11} = 0, H_0 : q_{01} = 0, H_0 : q_{10} = 0, H_0 : q_{00} = 0, H_0 : q_{11} = 0\), are the forms of (10), (13), (14), (15) and (16), respectively, which are given in Appendix B in the materials section.

The extremeness of the observed value for the test statistic under the null hypothesis leads to a small p-value, which would imply rejection of the null hypothesis. Thus, if the null hypothesis is the true relationship, we expect to obtain a higher p-value. In the second step, we also set a threshold for the asymptotic p-value such that the relationships with asymptotic p-value greater than or equal to the threshold are selected.

A large p-value indicates a larger possibility that the null hypothesis holds. Note that the p-value is less than or equal to 1. In this study, we use the threshold 1 for the p-value criterion in the examples because the largest p-value for each relationship is one. From the simulation study and the real data example discussed in this study, setting 1 to be a threshold for p-value criterion can lead to very accurate results. Note that for other examples, it is possible that the largest p-value is not 1. In this case, we need to observe the p-values to select a suitable threshold.

It is worth noting that the hypothesis testing procedure corresponds to a confidence interval approach [30]. From the confidence interval viewpoint, when the p-value is large enough (close to 1) or small enough (close to 0), we have confidence to accept or reject the null hypothesis. Therefore, in this study, when p-value is 1, we have confidence to accept the null hypothesis.

The two-step method is described as follows.

**Procedure for selecting the true relationship of \(m\) elements**

**Step 1.** For a sample of size \(n\) for \(m\) elements, calculate the counting numbers for the 6 relationships of each pair of the elements. Set a threshold for the counting numbers. Select the relationships with a counting number greater than the threshold.

**Step 2.** For each pair of elements, derive the asymptotic p-values for each relationship and set a threshold for the p-value. Select the relationships with a asymptotic p-value greater than or equal to the threshold.

| A/B | 0          | 1          |
|-----|------------|------------|
| 0   | \(m_{00,00}\) | \(m_{01,00}\) | \(m_{01,01}\) | \(m_{00,10}\) | \(m_{00,11}\) | \(m_{01,11}\) |
| 1   | \(m_{10,00}\) | \(m_{10,01}\) | \(m_{11,00}\) | \(m_{11,01}\) | \(m_{11,11}\) |

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**Table 5.** Splitting probabilities caused by misclassification error.

| A/B | 0          | 1          |
|-----|------------|------------|
| 0   | \(q_{00,00} = (1-p)^2 q_{00}\) | \(q_{01,01} = p(1-p) q_{00}\) | \(q_{00,01} = p(1-p) q_{01}\) | \(q_{00,10} = (1-p)^2 q_{00}\) |
|     | \(q_{01,00} = p(1-p) q_{00}\) | \(q_{00,11} = p^2 q_{00}\) | \(q_{01,11} = p(1-p) q_{00}\) | \(q_{01,01} = p^2 q_{01}\) |
| 1   | \(q_{10,00} = (1-p)^2 q_{00}\) | \(q_{10,01} = p(1-p) q_{00}\) | \(q_{10,10} = (1-p)^2 q_{10}\) | \(q_{10,11} = p(1-p) q_{11}\) |
|     | \(q_{11,00} = p^2 q_{10}\) | \(q_{11,01} = p(1-p) q_{10}\) | \(q_{11,10} = (1-p)^2 q_{11}\) | \(q_{11,11} = (1-p)^2 q_{11}\) |

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Step 3. For each pair of elements, select the relationships satisfying both criteria of Step 1 and Step 2. This relationship is the estimated relationship for the two elements.

Note that it is possible to have more than one relationship satisfying both criteria for two elements. But from a simulation result and a real data application, it shows that in most situation, there is only one relationship satisfying both criteria.

The asymptotic p-value has a closed form which can be easily calculated and the counting number can also be easily calculated. This shows that this method can provide a convenient way to recover the biological pathway.

An Example

We revisit the example of Figure 2 to illustrate the counting step. Assume that we only have a sample of the states for the 7 elements and we want to recover the 12 true relationships. Note that there are totally $C_7^2 = 21$ pairs of the 7 elements and there are only 12 pairs with relationships in this example. When considering the case without measurement error, we can reconstruct the pathway from a sample using the counting number method if the sample size is large enough. We can construct the Boolean network for the example by identifying prerequisite or similar relationships. From Table 1, we list the relationship corresponding to the highest counting number for each pair as follows:

$(A < B), c_{AB} = 13, (A < C), c_{AC} = 13, (A < D), c_{AD} = 13, (A < E), c_{AE} = 13$,

$(A < F), c_{AF} = 13, (A < G), c_{AG} = 13, (B < C), c_{BC} = 12, (B < D), c_{BD} = 10$,

$(B < E), c_{BE} = 13, (B < F), c_{BF} = 10, (B < G), c_{BG} = 12, (C < D), c_{CD} = 13$,

$(C < E), c_{CE} = 12, (C < F), c_{CF} = 13, (C < G), c_{CG} = 13, (D < G), c_{DG} = 13$,

$(D < E), (D < F), (D < G), c_{DE} = 10, (D < F), (D < G), c_{DG} = 9$,

$(E < F), (E < G), c_{EG} = 12, (F < G), c_{FG} = 13$,

where $c_{AB}$ denotes the counting number corresponding to the indicating relationship $(A < B)$.

If there is only one relationship corresponding to the highest counting number, we list that one, such as $(A < B)$; if there are more than one relationship corresponding to the highest counting number, we list all of the relationships, such as $(D < E), (D < F), (D < G)$. In the 21 pairs, the relationships corresponding to the highest counting number 13 is the 12 true relationships, and the relationships with the counting number less than 13 are not the true relationships.

Comparison

We consider two existing methods for detecting the pairwise relationships between any two elements. A simulation study is conducted to compare the proposed method with the existing methods for the measurement error case.

Existing methods

Li and Lu [29] proposed the directed acyclic Boolean network to recover the genetic network. For any pair of element, they use the EM algorithm to calculate the maximum likelihood estimator of misclassification rate $p$ under the multinomial distribution model structure and adopt a criterion that requires a true relationship to correspond to a small estimator of $p$ in order to select a relationship. Besides the disadvantage that the EM algorithm is time-consuming, this method is also shown to be less accurate than the counting method from a simulation study.

Another method for inferring the relationship of any two elements is proposed by Sahoo et al. [27]. For any two genes $A$ and $B$, let $n_{00}, n_{01}, n_{10}$ and $n_{11}$ denote the numbers of the four states $(0,0), (0,1), (1,0)$ and $(1,1)$ of $(A,B)$, respectively from a sample. For example, to infer whether the relationship $A < B$ is true, they use the following two statistics to test if the relation $(A,B) = (0,1)$ is true:

\[
\text{error rate} = \frac{1}{2} \left[ \frac{n_{01}}{n_{00} + n_{01}} + \frac{n_{01}}{n_{11} + n_{01}} \right]
\]

\[
\text{statistic} = \frac{\text{expected} - \text{observed}}{\sqrt{\text{expected}}}
\]

where “expected” and “observed” denote the values of $(n_{01} + n_{00}) \times (n_{01} + n_{11})/((n_{00} + n_{01}) + (n_{10} + n_{11}))$ and $n_{01}$, respectively.

The relationship $A < B$ in Sahoo et al. method is regarded as true when the “error rate” value is less than 0.1 and “statistics” value is greater than 3 [27]. However, from our calculation, the method may lead to inaccurate results when the sample size is not large. For instance, suppose the number of experiments we observed is 91 and the numbers of states corresponding to $(0,1), (0,0), (1,0)$ and $(1,1)$ are 1, 30, 30 and 30 respectively, resulting in a small “statistic” value of 2.94. Note that the state $(0,1)$ indicates that the relationship $A < B$ does not hold. However, since the state $(0,1)$ only occurs once, it may be due to a measurement error. In this case, the method does not select the relationship $A < B$. This shows that the criterion is too conservative to select a potential relationship when the sample size is not large enough.

Simulation

We conduct a simulation study using the example of Figure 2 with 13 compatible states (Table 1) in order to compare the proposed method with the two existing methods. With a misclassification probability 0.05, we generate 100 states for the simulation comparison. Tables 6 and 7 show the counting numbers and p-values for different relationships with a sample size of 100, respectively. Note that the notations $H_{0101}, H_{0011}, H_{01}, H_{10}, H_{00}, H_{11}$ in Tables 6 and 7 denote the relationships in order in Table 3.

In this case, the maximum value for the counting number is 100 because the sample size is 100. As discussed in above, we can set a threshold (2) for the counting number. In this case, the thresholds for the similar and prerequisite relationships are 86 and 93. And we set the threshold for the p-value to be 1 because the highest p-value for each pair is 1 in this case. The relationships corresponding to the hypotheses with a p-value 1 are the candidates for the true relationship.

For any pair of the 7 elements, there are 6 possible relationships of each pair. Since there are 21 pairs for the 7


### Table 6. The counting numbers for the 21 pairs in the 100 states under each relationship.

| hypothesis | \( H_{010} \) | \( H_{001} \) | \( H_{01} \) | \( H_{10} \) | \( H_{00} \) | \( H_{11} \) |
|------------|---------------|---------------|---------------|---------------|---------------|---------------|
| (A, B)     | 55            | 45            | 98            | 57            | 90            | 55            |
| (A, C)     | 72            | 28            | 95            | 77            | 93            | 35            |
| (A, D)     | 51            | 49            | 93            | 58            | 95            | 54            |
| (A, E)     | 56            | 44            | 98            | 58            | 90            | 54            |
| (A, F)     | 54            | 46            | 98            | 56            | 90            | 56            |
| (A, G)     | 26            | 74            | 99            | 27            | 89            | 85            |
| (B, C)     | 51            | 49            | 64            | 87            | 83            | 66            |
| (B, D)     | 46            | 54            | 70            | 76            | 77            | 77            |
| (B, E)     | 91            | 9             | 95            | 96            | 52            | 57            |
| (B, F)     | 51            | 49            | 76            | 75            | 71            | 78            |
| (B, G)     | 53            | 47            | 92            | 61            | 55            | 92            |
| (C, D)     | 35            | 65            | 76            | 59            | 94            | 71            |
| (C, E)     | 50            | 50            | 86            | 64            | 84            | 66            |
| (C, F)     | 72            | 28            | 98            | 74            | 72            | 56            |
| (C, G)     | 42            | 58            | 98            | 44            | 72            | 86            |
| (D, E)     | 43            | 57            | 74            | 69            | 79            | 78            |
| (D, F)     | 37            | 63            | 72            | 65            | 81            | 82            |
| (D, G)     | 37            | 63            | 87            | 50            | 66            | 97            |
| (E, F)     | 50            | 50            | 76            | 74            | 72            | 78            |
| (E, G)     | 52            | 48            | 92            | 60            | 56            | 92            |
| (F, G)     | 66            | 34            | 98            | 68            | 48            | 86            |

### Table 7. The p-values for the 21 pairs in the 100 states under each relationship.

| hypothesis | \( H_{010} \) | \( H_{001} \) | \( H_{01} \) | \( H_{10} \) | \( H_{00} \) | \( H_{11} \) |
|------------|---------------|---------------|---------------|---------------|---------------|---------------|
| (A, B)     | 0.8207        | 0.0065        | 1             | 0             | 0.1358        | 0             |
| (A, C)     | 0.0011        | 0.0005        | 1             | 0             | 0.0095        | 0.0680        |
| (A, D)     | 0.1330        | 0.1511        | 0.7931        | 0             | 1             | 0             |
| (A, E)     | 0.9228        | 0.0046        | 1             | 0             | 0.1201        | 0             |
| (A, F)     | 0.7200        | 0.0092        | 1             | 0             | 0.1527        | 0             |
| (A, G)     | 0.0010        | 0.6534        | 1             | 1             | 0.7599        | 0.3630        |
| (B, C)     | 0.9149        | 0.4654        | 0.0060        | 1             | 0.7636        | 0.0152        |
| (B, D)     | 0.4237        | 1             | 0.6569        | 0.6504        | 0.9855        | 0.9855        |
| (B, E)     | 0.0004        | 1             | 0.8094        | 1             | 0             | 0             |
| (B, F)     | 0.9994        | 0.8352        | 0.9990        | 0.9065        | 0.9118        | 0.9189        |
| (B, G)     | 0.4376        | 0.1168        | 1             | 0             | 0             | 0.8556        |
| (C, D)     | 0.0004        | 1             | 0.0268        | 0             | 0.0054        | 0.0025        |
| (C, E)     | 0.7452        | 0.6280        | 1             | 0.0058        | 0.9122        | 0.0165        |
| (C, F)     | 0.0010        | 1             | 0             | 0.0005        | 0.0014        | 0             |
| (C, G)     | 0.0724        | 0.1689        | 1             | 0             | 0.0070        | 0.3746        |
| (D, E)     | 0.1613        | 1             | 0.4171        | 0.4297        | 1             | 0.9031        |
| (D, F)     | 0.0093        | 1             | 0.1226        | 0.1554        | 0.8975        | 1             |
| (D, G)     | 0.0003        | 1             | 0.0377        | 0             | 0.0001        | 0             |
| (E, F)     | 0.9367        | 0.9716        | 0.9702        | 0.7834        | 0.9965        | 0.9993        |
| (E, G)     | 0.3662        | 0.1523        | 1             | 0             | 0             | 0.9043        |
| (F, G)     | 0.0010        | 1             | 0.0001        | 0             | 0             | 0.0110        |

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**Yeast expression data**

We revisit the MAPK pathway example from the Introduction. The datasets used in analyzing the MAPK pathway include 81 experimental data excluding two data with missing values, 57 from Spellman et al. [31] and 26 from Zhu et al. [32]. The datasets from Spellman et al. [31] include 18 data from the alpha factor experiments, 14 data set from the Elutrtation experiments and 24 data sets from cdc15 experiments. The datasets from Zhu et al. [32] include 25 data from Forkhead experiments. The raw data can be download from the Stanford Microarray Database [33]. We adopt values corresponding to the Log(base2) column in the raw dataset to reconstruct the MAPK pathway, which are log ratio values of red to green signal.

A gene state is regarded as on state or off state when the log ratio value of red to green signal is greater than or less than 0, respectively. The gene expression data for the 81 experimental data (Table S1) are given in the supplementary material.

In this study, we apply the two-step approach to explore the expression profiles, and show exploratory results on the pathway. The results are also compared with the Li and Lu’s method [29] and Shao et al. method [27].

We implement the proposed method to the yeast cell cycle data [31,32]. In our analysis, we assume that the level \( z = 0.1 \). According to the threshold selection formulas (2), the thresholds for the similar and prerequisite relationships are 61 and 69, respectively. And the threshold for the asymptotical p-value we selected is 1.

According to the network structure reconstructed using our proposed approach, we can see that Wsc2p and Mid2p activate Rho1p, Pkc1p and Bck1p which results in activation of the downstream of MAPK cascade, Mkk1p and Mlp1p. Activated
The proposed approach is designed to test the hypothesis of two-step counting. The number of elements is crucial in determining the time complexity of the method. Therefore, the time complexity for calculating the counting numbers and biological pathways from noisy array data is significant. This new method is particularly useful for reconstructing pathways from genetic interactions alone. Long chains of events may happen on the protein level (e.g., pathway comprises more than genetic interactions alone. Long chains of events may happen on the protein level (e.g., deactivation by phosphorylation) which does not necessarily have to be regulated via gene expression, but not other biological interactions.

In summary, we propose a two-step approach to test the biological pathways from noisy array data. This new method has the advantages of easy calculation for the counting numbers and simple closed forms of the p-value. From the simulation results, we can see that this method can precisely estimate the true relationships for most of the situations. Compared with the other existing methods, it can provide a more accurate and efficient alternative approach for reconstructing the biological network.

Materials and Methods

Appendix A: Threshold Selection

(i) Suppose the misclassification probability is p. For a similar relationship such as the case A ~ B, in this case, we have

\[ m_{01} = m_{10} = 0 \text{ and } m_{00} + m_{11} = n. \]  

By (3), the last equation is equal to \(n(p^2 + (1-p)^2)\), which is the mean of the counting number if this similar relationship holds. Since we cannot expect that the counting number is always equal to the threshold of two elements, A and B, a prerequisite relationship holds. In this case, we have

\[ m_{01} = 0 \text{ and } m_{00} + m_{11} + m_{10} = n. \]  

With misclassification error, the counting number corresponding to the relationship is

\[(1-p)^2 + p^2) m_{00} + 2p(1-p)m_{01} + 2p(1-p)m_{10} + ((1-p)^2 + p^2) m_{11} + \]

By (4), (5) is equal to

\[ ((1-p)^2 + p^2) m_{00} + 2p(1-p)m_{01} + ((1-p)^2 + p^2) m_{11} + \]

By a similar argument as in (i), we suggest \((1-w)^2 + w(1-w)n\) as a threshold for the prerequisite relationship.

Appendix B: Computational details

The methods for testing the 6 hypotheses in Table 3 are listed as follows.
(i) For deriving the p-value of the test:

\[ H_0 : q_{01} = q_{10} = 0 \] vs \[ H_1 : q \neq H_0, \]

we can consider the following two different situations. Note that the condition \( q_{01} = q_{10} = 0 \) in hypothesis \( H_0 \) is equivalent to \( q_{01} + q_{10} = 0 \) because \( q_{01} \geq 0 \) and \( q_{10} \geq 0 \).

(I) The misclassification probability \( p \) is zero.

The statistics

\[
\frac{(n_{01} + n_{10})/n - (q_{01} + q_{10})}{\sqrt{(n_{01} + n_{10})/n(1 - (n_{01} + n_{10})/n)}}
\]

has an asymptotic standard normal distribution under the null hypothesis \( q_{01} + q_{10} = 0 \).

(ii) The misclassification probability \( p \) is greater than zero.

In this case, the mean and variance of the random variable \( n_{01} + n_{10} \) is

\[
E(n_{01} + n_{10})/n = 2p(1-p)(q_{00} + q_{11}) + (p^2 + (1-p)^2)(q_{01} + q_{10})
\]

\[
= 2p(1-p)
\]

and

\[
Var(n_{01} + n_{10})/n = 2(1-p)p(q_{00} + q_{11})(1 - 2p(q_{00} + q_{11}) + 2p^2(q_{00} + q_{11}))/n
\]

\[
= 2(1-p)p(1-2p + 2p^2)/n
\]

under the null hypothesis.

Consequently, the asymptotic p-value is

\[
P\left(|Z| \geq \frac{(n_{01} + n_{10})/n - 2p(1-p)}{\sqrt{2(1-p)p(1-2p + 2p^2)/n}}\right),
\]

which can be rewritten as (3).

(iii) For deriving the p-value of the test:

\[ H_0 : q_{00} = q_{11} = 0 \] vs \[ H_1 : q \neq H_0, \]

by an argument similar to (i), for \( p > 0 \), the asymptotic p-value is

\[
1 - 2\Phi\left(-\frac{(n_{00} + n_{11})/n - 2p(1-p)}{\sqrt{2(1-p)p(1-2p + 2p^2)/n}}\right).
\]

(iv) For deriving the p-value of the test:

\[ H_0 : q_{10} = 0 \] vs \[ H_1 : q \neq H_0, \]

by an argument similar to (ii), the asymptotic p-value is

\[
1 - 2\Phi\left(-\frac{n_{01}/n - p(1-p)q_{01}}{\sqrt{p((1-p) - q_{10} + 2p q_{10})}}\right).
\]

(v) For deriving the p-value of the test:

\[ H_0 : q_{00} = 0 \] vs \[ H_1 : q \neq H_0, \]

by an argument similar to (iii), the asymptotic p-value is

\[
1 - 2\Phi\left(-\frac{n_{00}/n - p(1-p)q_{00}}{\sqrt{p((1-p) - q_{01} + 2p q_{01})}}\right).
\]

(vi) For deriving the p-value of the test:

\[ H_0 : q_{11} = 0 \] vs \[ H_1 : q \neq H_0, \]

by an argument similar to (i), the asymptotic p-value is

\[
1 - 2\Phi\left(-\frac{n_{11}/n - p(1-p)q_{11}}{\sqrt{p((1-p) - q_{10} + 2p q_{10})}}\right).
\]

The estimators \( \hat{q}_{ij} \) of \( q_{ij} \) in the above formulas of asymptotic p-values are given in Appendix C.

Appendix C: Frequency estimation

If the misclassification probability \( p \) is known, the methods for estimating the probability \( q_{00}, q_{01}, q_{00} \) and \( q_{00} \) are listed as follows.

According to Table 5, we have

\[
r_{00} = (1-p)^2 q_{00} + p(1-p)q_{01} + p(1-p)q_{10} + p^2 q_{11},
\]
The derived values are used as estimators for $q_{ij}$.

**Appendix D: Misclassification probability estimation**

If $p$ is unknown, we can apply the maximum likelihood approach to estimate $p$. By Table 5, we can rewrite the multinomial model for the observations $n_{ij}=0,1$ in terms of $p$ and other parameters. The maximum likelihood approach for deriving the maximum likelihood estimator of $p$ is based on the likelihood function

$$L(p) = \prod_{i,j} \frac{n_{ij}}{(n_{ij})^{n_{ij}} (1-n_{ij})^{1-n_{ij}}}$$

where $n_{ij} \neq 0,1$. This involves $p$ and other parameters, $q_{ij}, p = 0.1$, given in Appendix C. The maximum likelihood approach is to find the maximum likelihood estimators of $p$ and $q_{ij}$ such that the estimators can maximize the likelihood function (17) [29].

**Supporting Information**

**Figure S1** An example of Boolean network with 10 elements.

**Table S1** The 81 experimental yeast expression data.

**Author Contributions**

Conceptualized and designed the experiments: HW HH-SL. Performed the experiments: HW T-HC. Analyzed the data: HW T-HC. Wrote the paper: HW HH-SL T-HC.

**References**

1. Roberts CJ, Nelson B, Martin MJ, Stoughton R, Meyer MR, et al. (2000) Signaling and circuitry of multiple MAPK pathways revealed by a matrix of global gene expression profiles. Science 287: 873–880.

2. Buecheler BM, Errede B (1997) Coordination of the mating and cell integrity mitogen-activated protein kinase pathways in Saccharomyces cerevisiae. Molecular and Cellular Biology 17: 6537–6545.

3. O’Rourke SM, Herskovitz I (1998) The hog1 MAPK prevents cross talk between the hog and pheromone response MAPK pathways in Saccharomyces cerevisiae. Genes & Development 12: 2974–2986.

4. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

5. Jensen FV (2001) Bayesian Networks and Decision Graphs. New York: Springer.

6. Posas F, Saito H (1997) Osmotic activation of the HOG MAPK pathway between the hog and pheromone response MAPK pathways in Saccharomyces cerevisiae. Genes & Development 12: 2874–2886.

7. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

8. Pearl J (1988) Probabilistic Reasoning in Intelligent Systems: Networks of Plausible Inference. San Mateo: Morgan Kaufmann.

9. Wei Z, Li H (2008) A hidden spatial-temporal markov random field model for network-based analysis of time course gene expression data. Annals of Applied Statistics 1: 612–633.

10. Pearl J (1988) Probabilistic Reasoning in Intelligent Systems: Networks of Plausible Inference. San Mateo: Morgan Kaufmann.

11. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

12. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

13. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

14. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

15. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

16. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

17. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

18. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

19. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

20. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

21. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

22. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

23. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

24. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

25. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

26. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

27. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

28. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

29. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

30. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.

31. Madhani HD, Fink GR (1997) Combinatorial control required for the specificity of yeast MAPK signaling. Science 275: 1314–1317.
30. Bickel PJ, Doksum KA (2000) Mathematical statistics: basic ideas and selected topics. Prentice Hall 1.
31. Spellman PT, Sherlock G, Zhang MQ, Iyer VR, Anders K, et al. (1998) Comprehensive identification of cell cycle-regulated genes of the yeast saccharomyces cerevisiae by microarray hybridization. Molecular Biology of Cell 9: 3273–3297.
32. Zhu G, Spellman PT, Volpe T, Brown PO, Botstein D, et al. (2000) Two yeast forkhead genes regulate the cell cycle and pseudohyphal growth. Nature 406: 90–94.
33. Hubble J, Demeter J, Jin H, Mao M, Nitzberg M, et al. (2009) Implementation of GenePattern within the Stanford Microarray Database. [http://smd.stanford.edu/].
34. Agresti A, Coull BA (1998) Approximate is better than 'exact' for interval estimation of binomial proportions. The American Statistician 52: 119–126.
35. Wang H (2007) Exact confidence coefficients of confidence intervals for a binomial proportion. Statistica Sinica 17: 361–368.
36. Wang H (2008) Exact confidence coefficients of simultaneous confidence intervals for multinomial proportions. Journal of Multivariate Analysis 99: 896–911.
37. Wang H (2009) Exact average coverage probabilities and confidence coefficients of confidence intervals for discrete distributions. Statistics and Computing 19: 139–146.