STUDY PROTOCOL

The effects of green cardamom supplementation on blood glucose, lipids profile, oxidative stress, sirtuin-1 and irisin in type 2 diabetic patients: a study protocol for a randomized placebo-controlled clinical trial

Mohadeseh Aghasi 1, Shohreh Ghazi-Zahedi 1, Fariba Koohdani 2, Fereydoun Siassi 1, Ensieh Nasli-Esfahani 3, Ali Keshavarz 4*, Mostafa Qorbani 5, Hoorieh Khoshamal 1, Asma Salari-Moghaddam 1 and Gity Sotoudeh 1*

Abstract

Background: It has been suggested that the antioxidant, anti-inflammatory and hypolipidemic activities of cardamom may improve diabetes. However, the effect of this spice has not been investigated in diabetic subjects. This study was planned to determine the effects of green cardamom on blood glucose, lipids and oxidative stress status in type 2 diabetic patients.

Methods/design: Eighty overweight or obese patients with type 2 diabetes will be selected. They will be randomly assigned to receive 3 g/d green cardamom or placebo for 10 weeks. The socio demographic, physical activity and 24-h food recall questionnaires will be collected for each subject. Weight, height and waist circumference will be measured. Determination of blood glucose, lipid profile, and oxidative stress biomarkers including serum levels of total antioxidant capacity (TAC), malondialdehyde (MDA), and glutathione peroxidase (GPx) and superoxide dismutase (SOD) in red blood cells will be performed. The homeostasis model assessment-estimated insulin resistance (HOMA-IR) index and the quantitative insulin-sensitivity check index (QUICKI) will be calculated. Also, serum levels of irisin, and Sirtuin1 (SIRT1) will be measured.

Discussion: This trial will be the first study to explore the effects of green cardamom supplementation on glycemic control, lipid profile and oxidative stress in patients with type 2 diabetes mellitus. The results from this trial will provide evidence on the efficacy of green cardamom in type 2 diabetes mellitus.

Trial registration number: (http://www.irct.ir, identifier: IRCT2016042717254NS), Registration date: 23.11.2016.

Keywords: Trial protocol, Diabetes, Green cardamom
**Background**

Diabetes Mellitus (DM) is a non-communicable disease that affects many persons annually. In 2000, 171 million people suffered from DM globally, and it has been estimated to reach 366 million people in 2030 [1]. DM increases the risk of cardiovascular disease (CVD) by 2 to 4-fold. Dyslipidemia plays an important role in the pathogenesis of CVD in type 2 diabetes mellitus (T2DM) patients [2].

Studies have shown that high glucose level by stimulating reactive oxygen species (ROS) production damages β-cells and leads to impaired insulin release and insulin resistance [3, 4]. The antioxidant defense system is responsible for the neutralization of ROS; however, β-cells have a poor antioxidant defense system. Therefore, antioxidant supplementation by elevating antioxidant defense capacity can protect against β-cells dysfunction [5].

Many investigations have been executed regarding the relationship between sirtuin-1 (SIRT1) and irisin with the occurrence of DM. SIRT1 is a class III protein deacetylase that is associated with aging, inflammation and CVD [6]. Moreover, it has been suggested that SIRT1 plays an important role in glucose metabolism by stimulating pancreatic insulin release and insulin signaling pathway [7]. Investigations have stated that activation of SIRT1 leads to deacetylation of nuclear factor (NF)-κB, peroxisome proliferator activated receptor (PPAR)-γ and PPAR-γ coactivator 1α (PGC-1α) which have beneficial effect on glycemic indices, obesity and mitochondrial function [6, 7]. PGC-1α is a transcriptional coactivator that regulates energy balance and expression of fatty acids oxidation genes. In addition, it is involved in the improvement of mitochondrial function, insulin sensitivity and alleviation of oxidative stress. PPAR-γ and NF-κB are transcriptional factors that regulate fat metabolism and inflammation, respectively. Therefore, the inhibition of PPAR-γ and NF-κB activity by SIRT1 deacetylation suppresses fat accumulation and inflammatory processes in the human body [6, 8]. Studies have shown that lower SIRT1 activity is involved in the pathogenesis of DM and insulin resistance. Therefore, interventions that increase SIRT1 activity may be beneficial in improvement of insulin resistance and control of nuclear factor [7, 8]. Studies have shown that dietary polyphenols by stimulation of SIRT1 activity result in the suppression of NF-κB and activation of PGC-1α; hence improving insulin resistance and lipid metabolism [6].

Irisin is a recently known exercise-mediated myokine that regulates glucose hemeostasis [9]. Irisin contributes to the browning of white adipose tissue by increasing uncoupling protein-1 (UCP-1) in these cells, which results in the improvement of insulin sensitivity and glucose metabolism. UCP-1 is a key molecule that regulates energy expenditure [10]. Studies have indicated that a lower level of irisin is associated with DM and insulin resistance [9]. The activation of PGC-1α is the main stimulator of irisin release, therefore the elevation of irisin level may be a therapeutic approach in the management of insulin resistance and DM. It has been suggested that flavonoids such as quercetin and resveratrol by activation of PGC-1α and the SIRT1-related signaling pathway contribute to mitochondrial biogenesis [11]. Mitochondria regulates oxidative metabolism in muscle, hence increase in mitochondrial biogenesis results in higher insulin-related glucose uptake by muscle which subsequently improves insulin sensitivity [12].

Green cardamom (*Elettaria cardamomum*) is a dietary spice (in the ginger family) with nutraceutical effects that has antioxidant, anti-inflammatory and anti-carcinogenic properties [13, 14]. In a recent study, Ahmed et al. indicated that cardamom supplementation by suppression of α-amylase and α-glucosidase enzymes has anti-diabetic effects and may regulate glucose metabolism [15]. Cardamom volatile oil is composed of terpenes, esters, flavonoids, 1,8-cineole, alpha-terpinyl acetate and limonene [16]. Previous studies have indicated that 1,8-cineole has antioxidant and lipid-lowering effects [17, 18]. Cardamom is high in polyphenolic compounds such as quercetin, kaempferol, luteolin and pelargonidin which possess antioxidant properties [19]. It has been reported that resveratrol and quercetin can increase the deacetylation effect of SIRT1 by 13-fold and 4.6-fold, respectively [20]. Therefore, it has been hypothesized that cardamom antioxidant and anti-inflammatory activities may improve DM. Kandikattu et al. in an experimental model has shown that cardamom can stimulate in vitro superoxide dismutase (SOD), catalase and glutathione activity [21]. However, in two randomized clinical trials, cardamom (3 g/d) had no significant effect on oxidative stress biomarkers in prediabetic and diabetic patients [22, 23]. To the best of our knowledge, there is no clinical trial that has investigated the effects of cardamom on SIRT1 secretion and subsequently PGC-1α, irisin and insulin sensitivity in diabetic patients. Therefore, this study has designed to investigate the effect of *Elettaria cardamomum* on blood glucose, lipid profile and oxidative stress biomarkers in T2DM patients. To determine the mechanism of cardamom effect on blood glucose and lipid levels, blood irisin, and SIRT1 will be assessed.

**Methods and design**

**Study design**

This is a randomized, double blind, clinical trial. The flow chart of the study protocol is presented in Fig. 1. This study was conducted at the Diabetes Research Center, Endocrinology and Metabolism Clinical Sciences Institute, Tehran University of Medical Sciences, Tehran, Iran.
Sample size:
The sample size was calculated using the following equation:

\[ n = \frac{(Z_{1-\alpha/2} + Z_{1-\beta})^2 (S_1^2 + S_2^2)}{(\mu_1 - \mu_2)^2} \]

A total of 40 diabetic patients were calculated in each group (cardamom and placebo groups) based on a significant mean difference of TAC [24], with 95% confidence level, 80% power and additional drop-out rate of 20%.

Study population
T2DM patients were recruited from the outpatient department of Diabetes Research Center, Endocrinology and Metabolism Clinical Sciences Institute, Tehran University of Medical Sciences, Tehran, Iran. The diagnosis of diabetes will be based on medical records, using the American Diabetic Association (ADA), (fasting blood sugar (FBS) ≥126 mg/dl or 2-h postparndial (2HP) ≥200 mg/dl or HbA1c ≥6.5%) [25]. Eligible diabetic patients will be enrolled in the study and written informed consent was obtained at the baseline.

Inclusion and exclusion criteria
For the present study, T2DM patients, aged 30–60 years, with a diagnosis of T2DM for at least 2 years, HbA1C value greater than 7%, body mass index (BMI) between 25 and 35 kg/m², who will be treated with a stabilized dose of oral anti-diabetic drugs and statins will be included in the study.

Patients will be excluded if: 1) they are pregnant or lactating; 2) have gastrointestinal disorders that interfere with the bowel function, severe hepatic, renal (dialysis), inflammatory and thyroid diseases; 3) being on insulin therapy or need insulin based on expert physician’s opinion 4) have diabetes complications including micro and macrovascular complications; 5) take warfarin, fibrates (PPAR- α ligand), TZD (PPAR-gammaligand) and anti-depressant agents; 6) adhering to a specific diet during the past 3 months; 7) have
history of smoking or alcohol intake at least once a week in the past month; 8) receive (at least once a week) herbals, antioxidant, multivitamin/mineral supplements in the past 3 months; 9) they were unstable on the current dose of medications. Patients, who consume less than 90% of their intervention, will also be excluded.

Randomization
Participants will be randomly allocated into two groups: cardamom and placebo groups and will be followed-up for 10 weeks. Randomizations will be conducted by an assistant using permuted block randomization method and stratified randomization will be used to match participants based on age and gender distribution. The intervention allocation will be blinded for both investigators and participants.

Intervention
Fruits of *Elettaria Cardamom* will be purchased from Samex Agency (India), by the Traditional Medicine and Research Centre (TMRC), Shahid Beheshti University of Medical Sciences, Tehran, Iran. Cardamom (green cardamom powder) and placebo (rusk powder) capsules will be prepared by TMRC. Each capsule will contain 0.5 g of whole green cardamom powder or rusk powder. Shape, size and color of placebo capsules will be completely similar to the cardamom capsules. All placebo capsules will be placed in the same bag containing the cardamom capsules, in order to have the smell of cardamom.

The voucher number of green cardamom is *E. cardamomum* (L.) Maton, Family: Zingiberaceae, PMP-669. Essential oil, as well as the phenolic and flavonoid contents of whole green cardamom will be measured using gas chromatography–mass spectrometry (GC-MS) and high-performance liquid chromatography (HPLC) at the Institute of Medicinal Plants, Shahid Beheshti University of Medical Sciences. Some of the polyphenolic compounds of green cardamom such as caffeic acid, gallic acid, quercetin and luteolin, which were mentioned in other articles [26, 27] will be determined by HPLC. Cardamom and placebo capsules will be provided to both groups monthly for 10 weeks. Participants will be required to consume 2 capsules (3 g daily) per meal. The last packages of capsules will be checked at the end of the month and the number of remaining capsules will be counted; thereafter, new packages will be delivered to patients. All capsules will be given simultaneously with the usual diabetes care that includes maintenance of the oral anti-diabetic drugs dosage.

Participants will be asked to keep their usual lifestyle including medical nutrition therapy and physical activity level.

Adherence
To assess patients’ compliance during the 10 weeks, a researcher will be assigned to check them weekly using telephone to discern whether they were consuming the supplements.

Patient safety
Patients will be monitored weekly during the study period and any occurrence of adverse events will be recorded.

Study outcomes
**Primary outcomes** The primary outcomes of this clinical trial are the changes in HbA1C, FBS, insulin, lipid profile (triglyceride, total cholesterol, HDL-cholesterol, LDL-cholesterol), oxidative stress biomarkers such as TAC, SOD, GPx, MDA, SIRT1 and irisin at the end of the study in comparison with the baseline values.

**Secondary outcomes** The secondary outcomes of this clinical trial are changes in weight, BMI and waist circumferences (WC).

Procedure After obtaining the informed consent, both groups will be visited at the clinical research center twice: at baseline, and after 10 weeks. Subjects will be interviewed regarding their socio-demographic background at baseline. At each visit, anthropometric indices (weight, height and waist circumference) will be measured, fasting blood sample will be collected and a 24-h food recall and international physical activity questionnaire (IPAQ) will be obtained.

Height will be measured using the SECA stadiometer to the nearest 0.1 cm and weight will be measured using a SECA electronic scale to the nearest 0.1 kg. Waist circumference will be measured midway between the lowest rib and the iliac crest using a non-stretchable measuring tape to the nearest 0.1 kg. BMI will be calculated as weight (kg) divided by height squared (m²). Venous blood sample (10 ml) will be drawn after 12-h overnight fasting by trained nurses in seated position to measure biochemical markers. Then blood samples will be centrifuged at 3000 rpm for 10 min at 4 °C to obtain the serum and plasma and will be stored at ~80 °C until biochemical analyses. Lipid profiles (TC, HDL, LDL, TG) and glucose level will be measured by the enzymatic colorimetric method using kits. Serum insulin will be measured using enzyme-linked immunosorbent assay (ELISA) kit. Insulin sensitivity and insulin resistance will be determined using the QUICKI and HOMA-IR equation, respectively.
Assessment of dietary intake
To assess participants’ dietary intake, a 24-h food recall will be collected 3 times during the study (at baseline, middle and end of study). Patients will complete food descriptions including food and drinks (brand names), food preparation (ingredients) in detail as much as possible in the last day. Pictures of food commonly consumed in Iran, together with a set of common household measurement tools (glass, cup, soup bowls, plates, teaspoon and tablespoon) will be provided to assist subjects in estimating the portion sizes of the food. Dietary intake will be analysed with Nutritionist version 4.

Assessment of physical activity levels
IPAQ will be applied to assess the physical activity level of participants [29]. The IPAQ form comprises walking, moderate-intensity and vigorous-intensity activity and will be expressed as metabolic equivalents per minute (MET-min) per week. The levels of physical activity will be categorized into low, moderate and high, based on the IPAQ criteria.

Statistical analysis
All statistical analyses will be performed using SPSS version 21 (SPSS Inc., Chicago, USA). Data will be expressed as mean ± SD for continuous variables and percentage for non-continuous variables. Normality tests will be assessed through Shapiro–Wilk tests carried out on each parameter before considering the normality of data. A multivariate regression analysis will be used to assess time effects and time-by-treatment interactions effect on all outcome variables. A p-value of less than 0.05 will be considered as significant.

Ethical considerations
This clinical trial will be conducted after approval by the Ethics Committee of Tehran University of Medical Sciences and after obtaining informed consent from the participants. Whenever a person is unable to continue supplementation, he/she will be excluded from the study.

The limitations
One of the limitations of this study is the lack of cooperation of some patients who were replaced with other patients.

Discussion
T2DM is a common chronic disease worldwide. Recently, herbal medicine has become more popular due to their beneficial effects [30]. Cardamom (E. cardamomum) is an expensive spice with an old history [31]. Today, cardamom is grown in India, Thailand, Guatemala, Ceylon and Malay Archipelago [26]. The main characteristics of cardamom include sweet odor, warm nature and mild strong taste that make it a popular spice [31]. Cardamom is rich in antioxidant compounds that cause elevation and activation of the antioxidant defense system [32].

To the best of our knowledge, this is the first randomized controlled trial that will determine the effect of cardamom on glycemic status, lipid profile, oxidative stress biomarkers, SIRT1 and irisin in T2DM. The selection of patients with T2DM is the main strength of the present study. The results of this trial will provide clinical evidence on the effectiveness and safety of cardamom supplementation in patients with T2DM.

Abbreviations
BMI: Body Mass Index; CVD: Cardiovascular Disease; DM: Diabetes Mellitus; GPx: Glutathione Peroxidase; HOMA-IR: Homeostasis Model Assessment-estimated Insulin Resistance; IPAQ: International Physical Activity Questionnaire; MDA: Malondialdehyde; QUICKI: Quantitative Insulin-sensitivity Check Index; ROS: Reactive Oxygen Species; SIRT1: Sirtuin1; SOD: Superoxide Dimutase; T2DM: Type 2 Diabetes Mellitus; TAC: Total Antioxidant Capacity

Funding
This research has been supported by Tehran University of Medical Sciences and Health services grant no. 94-04-161-31,133.

Availability of data and materials
Not applicable.

Authors’ contributions
GS designed the initial idea of this work, which was further develope by SAK and MA. FK and FS and EN coordinated the study. MQ advised on statistical analysis. MA and SGZ and HKA organized participant management and data-collection. GS and MA and ASM drafted the manuscript. The manuscript has been read and approved by all authors.

Ethics approval and consent to participate
The Ethics Committee of Tehran University of Medical Sciences has approved study protocol (IRTUMS.REC.1395.27/00). A written informed consent form will be signed and dated by subjects and investigators at the beginning of the study.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.
Publisher's Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details
1. Department of Community Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences, Tehran, Iran. 2. Department of Cellular and Molecular Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences, Tehran, Iran. 3. Department of Clinical Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences, Tehran, Iran. 4. Department of Community Medicine, School of Medicine, Alborz University of Medical Sciences, Karaj, Iran. 5. Department of Community Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences, Tehran, Iran. 6. Department of Clinical Nutrition, School of Nutritional Sciences and Dietetics, Tehran University of Medical Sciences, Tehran, Iran. 7. Department of Community Medicine, School of Medicine, Alborz University of Medical Sciences, Karaj, Iran.

Received: 12 February 2017 Accepted: 19 December 2017

Published online: 17 January 2018

References
1. Wild S, Roglic G, Green A, Sicree R, King H. Global prevalence of diabetes: estimates for the year 2000 and projections for 2030. Diabetes Care. 2004; 27(5),1047–1053.[Pubmed].
2. Ejtahed HS, Mohtadi-Nia J, Homayouni-Rad A, Niafar M, Asghari-Jafarabadi M, Mofid V, et al. Effect of probiotic yogurt containing Lactobacillus acidophilus and Bifidobacterium lactis on lipid profile in individuals with type 2 diabetes mellitus. J Dairy Sci. 2011;94(4), 3288–3294.[Pubmed].
3. Sakuraba H, Mizukami H, Yagihashi N, Wada R, Hanyu C, Yagihashi S. Reduced beta-cell mass and expression of oxidative stress-related DNA damage in the islet of the Japanese Type II diabetic patients. Diabetologia. 2002;45(1), 85–96.[Pubmed].
4. Mohamed AK, Bierhaus A, Schleicker S, Tittscher H, Ziegler R, Nawroth PP. The role of oxidative stress and NF-kappaB activation in late diabetic complications. BioFactors. 1999;10(2–3),157–167.[Pubmed].
5. Karanakanar U, Pask KG. A systematic review of oxidative stress and safety of antioxidants in diabetes: Focus on islets and their defense. Diabetes Metab J. 2013;37(2),106–112.[Pubmed].
6. Chung S, Yao H, Hwang JW, Arunachalam G, Rahman I. Regulation of SIRT1 in cellular functions: role of polyphenols. Arch Bioche Biophys. 2010;1; 501(1),199–216.[Pubmed].
7. Kitada M and Koya D. SIRT1 in Type 2 Diabetes: Mechanisms and Therapeutic Potential. Diabetes Metab J. 2013;37(5), 315.[Pubmed].
8. Cao Y, Jiang X, Ma H, Wang Y, Xue P, and Liu Y. SIRT1 and insulin resistance. Arch Bioche Biophys. 2010;1; 501(1),79–86.[Pubmed].
9. Koh, S, Dostbil N & Alemdar S. Antimicrobial Effect of Seed Extract of Maton (Zingiberaceae), Elettaria cardamomum. YÜ Vet Fak Derg. 2006;16(2),99–101.[Pubmed].
10. Ağaçoğlu, S, Doşbil N & Alemdar S. Antimicrobial Effect of Seed Extract of Cardamom (Elettaria cardamomum Maton). YÜ Vet Fak Derg. 2006;16(2),99–101.[Pubmed].
11. Davis JM, Murphy EA & Carmichael MD. Effects of the dietary flavonoid quercetin upon performance and health. Curr Sports Med Rep. 2013;8(4), 206–213.[Pubmed].
12. Joseph AM and Hood DA. Relationships between Exercise, Mitochondrial Biogenesis and Type 2 Diabetes. Med Sport Sci. 2014;60, 48–61.[Pubmed].
13. Bhatacharjee B, Chatterjee J. Identification of proapoptotic, anti-inflammatory, anti-proliferative, anti-invasive and anti-angiogenic targets of essential oils in human breast cancer cell lines: dual reverse virtual screening and binding pose analysis. Asian Pac J Cancer Prev. 2013;14(6), 3735–3742.[Pubmed].
14. Jin S, Cho KH. Water extracts of cinnamon and clove shows potent inhibition of protein glycation and anti-atherosclerotic activity in vitro and in vivo hypolipidemic activity in zebrafish. Food Chem Toxicol. 2011; 49(7),1521–1526.[Pubmed].
15. Cho KH, 1,8-Cineole Protected Human Lipoproteins From Modification By Oxidation and Glycation and Exhibited Serum Lipid-Lowering and Anti-Inflammatory Activity in Zebrafish. BMC Rep. 2012;45(10),565–570.[Pubmed].
16. Deepa G, Ayesha S, Nishtha K, Thankamani M. Comparative evaluation of various total antioxidant capacity assays applied to phytochemical compounds of indian culinary spices. Int Food Res J. 2013;20(4), 1711–1716. View article.
17. De boer VCI, de Goffau MC, Arts ICW, Hollman PCH, Keijer J. SIRT1 stimulation by polyphenols is affected by their stability and metabolism. Mech Ageing Dev. 2006;127(7–8),618–627.[Pubmed].
18. Cho KH. 1,8-Cineole Protected Human Lipoproteins From Modification By Oxidation and Glycation and Exhibited Serum Lipid-Lowering and Anti-Inflammatory Activity in Zebrafish. BMC Rep. 2012;45(10),565–570.[Pubmed].
19. Deepa G, Ayesha S, Nishtha K, Thankamani M. Comparative evaluation of various total antioxidant capacity assays applied to phytochemical compounds of indian culinary spices. Int Food Res J. 2013;20(4), 1711–1716. View article.
20. Azimi P, Ghasvand R, Feizi A, Hatri M, and Abbas B. Effects of cinnamon, cardamom, saffron and ginger consumption on markers on glycemic control, lipid profile, oxidative stress and inflammation in type 2 diabetes patients. Rev Diabet Stud. 2014; 11(3), 258–266.[Pubmed].
21. kazemi S, Yaghoobi-Khouli F, Sassi F, Rahimi A, Ghavipour M, Koohdani F, Sotoudeh G. Cardamom supplementation improves inflammatory and oxidative stress biomarkers in hyperlipidemic, overweight, and obese pre-diabetic women: A randomized double-blind clinical trial. J. Sci Food Agric. 2017;97(15),5296–5301.[Pubmed].
22. Verma SK, Jain V & Singh DP. Effect of Greater cardamom (Amomum subulatum Roxb) on blood lipids, fibrinolysis and total antioxidant status in patients with ischemic heart disease. Asian Pac J Trop Dis. 2012;5379–5473.[Science Direct].
23. American Diabetes Association. Diagnosis and classification of diabetes mellitus. Diabetes Care. 2012;35(Suppl 1),564–571.[Pubmed].
24. Ekdolinnova OV, Tarlak I, Nenelova EV, Glažková IF. Testing phenol compounds in spices. European Journal of Applied Sciences. 2013;5(1),142–147.[Pubmed].
25. M C, Kaefer & Milner J A. Herbs and Spices in Cancer Prevention and Treatment. Herbal Medicine: Biomolecular and Clinical Aspects. 2011; 196;347–361.[Pubmed].
26. Miller NJ, Rice-Evans C, Davies MJ, Gopinathan Y, Milner A. A novel method for measuring antioxidant capacity and its application to monitoring the antioxidant status in premature neonates. Clin Sci (Lond). 1993;84(4),407–412.[Pubmed].
27. Craig CL, Marshall AL, Sjostrom M, Bauman AE, Booth ML, Ainsworth BE, et al. International physical activity questionnaire: 12-country reliability and validity. Med Sci Sports Exerc. 2003;35(8),1381–1389.[Pubmed].
28. Chen J, Huang Y, Guo W, Jang W, Lin WC & Huang CY. Cardamom (Elettaria cardamomum Maton) Crude and Its Main Compounds: Antioxidant, anti-influenza, and anti-diabetic activities in vitro. Food Chem Toxicol. 2011;49(7),1521–1526.[Pubmed].
29. Amma KPAP, Rani MP, Sasidharan I, Nisha VNP. Chemical composition, antimicrobial and antioxidant activities of Cardamom (Elettaria cardamomum Maton). J Sci Food Agric. 2013;93(9),407–412.[Pubmed].
30. Chen J, Huang Y, Guo W, Jang W, Lin WC & Huang CY. Cardamom (Elettaria cardamomum Maton) Crude and Its Main Compounds: Antioxidant, anti-influenza, and anti-diabetic activities in vitro. Food Chem Toxicol. 2011;49(7),1521–1526.[Pubmed].
31. Amma KPAP, Rani MP, Sasidharan I, Nisha VNP. Chemical composition.