Diversity and abundance of nematodes in soil treated with solarization treatments

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Abstract. Putri A H, Indarti S, Harjaka T. 2021. Diversity and abundance of nematodes in soil treated with solarization treatments. Biodiversitas 22: 2612-2618. Solarization is a tillage cultivation technique that influences the presence of nematodes in agricultural land. This research aims to determine the solarization effect on soil temperature and nematode abundance and diversity in the soil. Observations on species, diversity, and abundance of the nematodes were performed at the Nematology Laboratory, Department of Plant Pests and Diseases, Faculty of Agriculture, Gadjah Mada University. A completely randomized design (CRD) was used with six treatments and three replications. Meanwhile, the treatment groups were based on solarization duration (in days) on the soil which includes A(3), B(7), C(14) D (30), and K0 (control, without solarization) and K1 (control, farmer treatment). Field observations were performed to measure soil temperature, moisture, and pH. Nematodes were analyzed in the laboratory to determine the genus, abundance, and diversity. Furthermore, extracted plant-parasitic nematodes were isolated by using the whitehead tray method while the entomopathogenic group was isolated using the baiting method. The results showed the differences in tillage treatment and solarization affect soil temperature (diversity and abundance). A polynomial regression model explained the relationship between increased solarization time and soil temperature. The highest soil temperature was found in cultivated land and solarization land for 14 days (41.0°C) and the lowest was in the treatment of farmers (33.67°C). The lowest abundance was found at 14 days solarization, namely 286 individuals/100g, and the diversity index value of 1.37. The highest diversity index was found in the treatment of farmers with an index value of 1.75. The solarization treatment is best applied in the field for 14 days.

Keywords: Soil solarization, temperature, diversity and abundance of nematodes.

INTRODUCTION

Nematodes have an important role in soil agriculture. Nematodes have great economic benefits of both harmful and useful effects (Hailu and Hailu 2020). Plant-parasitic nematodes can cause massive damage to crops (Cotton et al. 2014). Plant-parasitic nematodes are one of the pests that can affect production in horticultural, food, and plantation crops. Plant-parasitic nematodes live and develop in plant tissue. Parasite nematodes can migrate from infected plants to other plants through the water around their host plants. Some examples of nematodes that are plant parasites are Meloidogyne sp. also called root-knot nematodes. One of the host plants of the nematode Meloidogyne sp. is a tomato plant. The specific symptoms of the attack can cause a knot or swelling of the roots (Mutalali’iah et al. 2019).

Plant parasitic nematodes cause damage to plant parts such as leaves, stems, and roots. Most nematodes are found to attack the roots of plants. Nematodes in the roots caused the growth of plants is abnormal and the density of the nematode population affects plant growth (Kayani et al. 2017). Citrus nematode (Tylenchulus semipenetrans) causes severe damage to citrus. The nematode build-up was the maximum at the lower inoculum level and an inverse relationship was observed between the reproduction factor and inoculum densities of the nematode. Roots function as transporters of nutrients to the plant tissue above the soil surface. If nematodes attack the roots, the normal root system will be reduced, causing the transport file network to experience interference. Roots that are disturbed by the presence of nematodes cause the plant to wither easily (Irshad et al. 2012).

Farmers use various control techniques to solve the problem of nematodes in the soil. One of them is solarization method. According to Candido et al. (2011), soil solarization is a technique that utilizes solar radiation and aims to increase soil temperature by using clear polyethylene sheets. When combined with an aerobic disinestation of soil, solarization can be used as an alternative to control pathogens and parasitic nematodes in the soil (Butler et al. 2012). Furthermore, Candido et al. (2008) stated that nematode recolonization could be hampered provided soil solarization is being carried out every two to three years. Hence, sustainable solarization treatment progressively reduces the population densities and prevents soil recolonization by nematodes.

Apart from parasitic nematodes, entomopathogenic nematodes were also found in agricultural land. The presence of entomopathogenic nematodes will benefit agricultural ecosystems because they can function as a biological control agent. The diversity of types of parasitic nematodes and entomopathogenic nematodes in an ecosystem is influenced by temperature. Solarization treatment on tillage will affect temperature. Several types of nematodes can survive and reproduce at certain
temperatures. The research results of Glazer and Salame (2000) proved entomopathogenic nematodes can survive temperatures up to 45°C. Solarization treatment is thought to affect the presence of species, population, and diversity of nematodes in agricultural land. Research has been carried out to determine the effect of tillage treatment using solarization techniques on soil temperature, population abundance, and nematode diversity in the soil.

MATERIALS AND METHODS

Research location
The study was conducted from July to November 2020 in Banyudono Village, Dukun Sub-District, Magelang Regency, Central Java. Observations on nematode genus, diversity, and abundance were carried out at the Nematology Laboratory, Department of Plant Pests and Diseases, Faculty of Agriculture, Gadjah Mada University.

Tillage and solarization treatment
Land preparation was done by cultivating the soil and loosening it until it was ready for planting. Transparent plastic with a thickness of 0.25mm was installed by covering all the soil plots as an experimental unit. The soil was turned over and watered before were covered with clear plastic. Solarization time duration of the soil was 3, 7, 14, and 30 days. Meanwhile, control without solarization and farmer treatment (without treatment, covered with clear plastic, but silver plastic mulch is used, directly before planting until harvest).

A completely randomized design (CRD) was used as the research method while treatment was according to variable solarization duration. The 0 day solarization treatment and tillage were used by farmers as controls. Meanwhile, each treatment was replicated three times. The notation for each treatment was allotted as K0 for control without solarization and K1 for farmers' treatment. Also, treatments were carried out in groups denoted as A, B, C, and D with the duration of 3, 7, 14, and 30 days, respectively, while the plot (1x2 m²) were randomly executed on the field. Solarization was made via loosened soil, irrigated, and covered with transparent polyethylene plastic. The closure duration was adjusted to the duration of solarization treatment.

Temperature, moisture, and pH were measured after solarization treatments. These parameters were measured with a soil survey instrument (Mediatech Soil Survey Instrument Digital 4 in 1 Backlight) by sticking it in the treated soil for 10 seconds. The tools are first fixed and then stuck into the ground vertically; hence, the temperature, moisture, and pH will appear.

Extraction, isolation, identification, analysis of nematode diversity and abundance
The nematode diversity and abundance were observed from solarized soil samples. The soil was taken from the treatment field randomly with approximately 200 g at a 10-20 cm depth from the surface. Also, samples were placed in a labeled clear plastic bag measuring 20x34cm² and afterward taken to the laboratory for extraction. Plant-parasitic nematodes were extracted using the whitehead tray method (Kaya and Stock 1997), while entomopathogenic nematodes were isolated using the baiting method (Cowles et al. 2005).

Whitehead tray method for the extraction and isolation of parasitic nematodes
Soil samples from the field were isolated and extracted using a modified whitehead tray method (Kaya and Stock 1997). In this procedure, a whitehead tray was prepared while tissue paper was placed on it. Furthermore, the soil from the field (100g) was afterward placed on the filter tray and leveled with water until it touched the surface of the filter paper. Soil that had been submerged with water was left for 24 hours at room temperature to isolate resident nematodes. Moreover, the soaked water resulting from nematode isolation was made into a suspension and deposited for 15 minutes. The suspension result was deposited in water, which contains nematodes with a gradually reduced volume until the suspension was set at ±55ml. A total of 5mL suspension was taken and observed under a microscope binocular (Olympus CX-22), to identify the nematode genus and abundance.

Baiting method for entomopathogenic nematode isolation
Soil samples were isolated using the baiting method to obtain entomopathogenic nematodes. The soil was placed into a bottle until it reached half of its volume (Cowles et al. 2005). Bait insects (Tenebrio molitor) wrapped in gauze were placed in a bottle and gradually filled to the brim with soil. Afterward, the soil and bait insect larvae contained in the bottle were left for 7 days. The larvae were then transferred to a petri dish and left for 3 days after which the dead ones were filtered and placed on a buffer in a closed jar. The jar was filled with water until it touched the paper's edge and then incubated for 14 days. Entomopathogenic nematodes that moved into distilled water are those ready to be harvested. Nematodes were identified based on morphology and their abundance was then calculated.

Nematode identification was done based on morphological characteristics. The main characters were the body shape, the body length, part of the oral cavity (presence or absence of stylet, stylet type), the shape and length of the esophagus, the tail shape. The recovered nematodes were observed under microscope binocular (Olympus CX-22) and then identified by using a key on the web nematode.unl.edu and book Key to Genera of Plant-Parasitic Nematodes (Mai and Lyon 1975; Hazir et al. 2004).

Data analysis
The data for soil temperature with the abundance and diversity of nematode were analyzed using the analysis of variance ANOVA test. Meanwhile, the LSD test was performed at a 5% level whenever the result was significantly different. The analysis was performed using the Stat 8.0 program (a statistical program for windows), while the relationship between solarization, temperature, and abundance was depicted in graphical form.
Furthermore, regression analysis was used to determine the relationship between solarization treatments and soil temperature. This regression model was based on the R² value hence, an R² value close to 1 was the most appropriate model used to determine the formula of the relationship. Regression analysis was performed using the Excel program (Excel for Windows 2019).

Nematode abundance and diversity found at sample locations were analyzed by using a diversity index (H index). The diverse genus of nematode genus can be measured by using a diversity index that refers to the Shannon-Wiener (Krebs 1989). Meanwhile, the Shannon-Wiener diversity index equation is as follows:

\[ H' = -\sum_{i=1}^{n} \left( \frac{n_{i}}{N} \right) \ln \left( \frac{n_{i}}{N} \right) \]

Where:
- \( H' \) : Shannon-Wiener Diversity Index
- \( S \) : Number of species
- \( n_{i} \) : number of individual species-i
- \( N \) : Number of individuals of all species.

**RESULTS AND DISCUSSION**

**Effect of solarization treatment on soil temperature**

The observations on soil temperature treated periodically with solarization as well as the control and farmers treatment showed significantly different results (P = 0.0012) (Table 1). The treated solarized soil moisture was the same (40%). This was due to the simultaneous performance of the watering treatment. The level of soil acidity (pH) was also the same (7.0). This was due to the simultaneous application of solarization as well as the control and farmers treatments.

The highest temperature in the solarized soil was at 14 days (41°C). However, this was not significantly different with 7 days (39.67°C) but was significantly different from other treatments. Therefore, it can be seen that there was an increase in soil temperature when soil solarization duration was increased. Also, this increasing temperature trend progressed from 3, 7 to 14 days, but the temperature decreased with further solarization duration. As observed, the temperature was lower for soil solarized for 30 days than those for 3, 7, and 14 days.

There was a relationship between solarization duration, temperature, and abundance, which was described in terms of the three parameters. The relationship between solarization duration and temperature based on regression analysis is illustrated in Figure 1 and Table 2.

The relationship between solarization duration and soil temperature was modeled as \( y = \text{temperature} \) and \( x = \text{solarization duration} \). Furthermore, the polynomial relationship was polynomial with the value \( R^2 = 0.9 \) or close to 1, and the equation was \( y = -0.0288x^2 + 0.8973x + 34.322 \). Compared with other forms of relationships, the \( R^2 \) value was lesser than 1 and close to 0 indicating no relationship. But in this observation, the polynomial regression equation with a value of \( R^2 = 0.9077 \) means that there is a relationship between solarization treatment time and temperature (Table 2).

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**Table 1. The temperature and pH of solarized soil with different duration of time, control, and farmer treatment**

| Treatment solarization | Soil temp. (°C) | pH |
|------------------------|----------------|----|
| 14 days                | 41.00 a        | 7.0|
| 7 days                 | 39.67 a        | 7.0|
| 3 days                 | 37.00 b        | 7.0|
| 30 days                | 35.33 bc       | 7.0|
| Control (without solarization) | 35.33 bc | 7.0|
| Farmer treatment       | 33.67 c        | 7.0|

**Figure 1.** The regression form of the relationship between temperature and solarization duration. A. Linear, B. Polynomial, C. Exponential
Temperature affects the presence and abundance of nematodes in the soil. The higher the temperature, the lower the abundance of the nematode population (Figure 2). The highest temperature was at 14 days of solarization (41.0°C), with the lowest nematode population (286 individuals/100g), followed by 7 days (39.67°C; 300.68 individuals/100g). While, the solarization treatment for 30 days and the control had the same temperature of 35.33°C, and the nematode population between the two treatments were not significantly different. The lowest temperature was found in the treatment of farmers with 33.67°C and a nematode population of 363.01/100g.

Population abundance and nematode diversity in solarized soil, without solarization, and farmer treatment

In different solarization treatments, different genus and abundance of nematodes were found. There are several genera of nematodes that were not found in solarization treatments but were found in the control and treatment of farmers. The varieties of nematodes found were identified down to their genus based on morphological characteristics (type of stylet, posterior). Furthermore, the abundance of each genus in all treatments can be seen in Table 3.

The population of nematode found in each solarized soil at different durations showed significant differences at the 5% level (P = 0.000). All treatments, with or without solarization including those by the farmers had a population that ranged from 286 to 579.34 nematodes (Figure 3).

The highest nematode abundance was in the solarized soil for 3 days (579.34 individuals /100g) significantly different between treatments. Also, the population was seemingly lower in soil treated for 14 days (286 individuals/100g). Meanwhile, the abundance of nematodes in the land is thought to affect plants especially when there are more resident parasitic genus than other breeds such as saprophages or non-parasitic nematodes. Population abundance and diversity of nematode genus determined the diversity index value.

The diversity of nematodes in solarized soil treated at intervals with or without solarization and those treatments by farmers showed significant differences. Therefore, results analysis of soil diversity treated with solarization at intervals indicated that the diversity index was significantly different (P = 0.0034) (Table 3).

Table 3. Genus and abundance of nematode populations found in soil solarization treatments

| Genus          | 3 days  | 7 days  | 14 days | 30 days | Control | Farmers treatment |
|----------------|---------|---------|---------|---------|---------|-------------------|
| Aphelenchoides | 36.67   | 3.67    | 3.67    | 3.67    | 69.67   | 99.00             |
| Helicotylenchus| 80.67   | 124.67  | 91.67   | 201.67  | 0.00    | 11.00             |
| Meloidogyne   | 172.33  | 73.33   | 44.00   | 58.67   | 157.67  | 36.67             |
| Paratylenchus | 0.00    | 0.00    | 0.00    | 0.00    | 3.67    | 25.67             |
| Pratylenchus  | 0.00    | 3.67    | 29.33   | 0.00    | 18.33   | 33.00             |
| Tylenchus     | 55.00   | 14.67   | 44.00   | 25.67   | 69.67   | 80.67             |
| Heterorhabditis* | 201.67 | 77.00   | 73.33   | 22.00   | 3.67    | 29.33             |
| Steinernema*  | 33.00   | 3.67    | 0.00    | 25.67   | 84.33   | 47.67             |

Note: *entomopathogenic nematode
The diversity index of nematodes on solarized soils without solarization and farmer treatment.

| Treatment solarization | Diversity index (H') |
|------------------------|----------------------|
| 3 days                 | 1.18 b               |
| 7 days                 | 1.38 ab              |
| 14 days                | 1.37 ab              |
| 30 days                | 1.05 b               |
| Without solarization   | 1.50 ab              |
| Farmer treatment       | 1.75 a               |

Note: Figures followed by the same lowercase letters in the same column indicate insignificant differences based on the LSD test at the 5% level.

In Table 4, it can be seen that the nematode diversity index was significantly different in the 3 days treatment from the farmer's treatment. The highest diversity index was found in the treatment of farmers (1.75), while the lowest diversity is at 30 days of treatment (1.05). These values were determined by the number of genus present and the abundance of nematodes in the soil.

Discussion

Soil treatment with solarization is one of the cultivation techniques in an agroecosystem. Solarization or soil tillage treatment mostly uses clear plastic. The application of this technique at different intervals is expected to improve soil temperature. While the moisture levels observed for the soil treated were the same. Watering was carried out simultaneously on all treated fields. Similarly, the soil pH shows the same value for each cultivated and solarized soil (pH 7.0). The difference was in the soil temperature that had been confirmed from five research treatments. The highest soil temperature occurred at 14 days of solarization. However, it was not significantly different from the 7 days treatment. This suggests that solarization treatment for 1 to 2 weeks can increase soil temperature. Simmons et al. (2013) previously proved that solarization alters soil temperature. Solarization treatment using a clear plastic trap can increase soil temperature from passive solar heating.

The regression relationship between solarization duration and soil temperature was polynomials (Fig. 1). Meanwhile, polynomial regression in this research is different from linear regression. Furthermore, the analysis was carried out by adding up the effect of the variables, including the duration of solarization as x to predict the value or soil temperature. The value of y increased as the value of x increases, but the increase was only up to a certain point. Also, the increase in soil temperature was followed by increasing solarization duration. However, this increase only proceeded to a certain point after which temperature began to decrease. It is assumed that in the solarization treatment 3, 7, and 14 days the soil conditions are still loose so that the reception of the sun's temperature in these conditions is still effective. In contrast to the 30 days solarization treatment, the soil conditions have begun to solidify. Soil temperature factors affect the life cycle and behavior of nematodes living in the soil (Munteanu 2017). Solarization can increase soil temperature causing death to resident nematodes. Parasitic nematodes die when the highest temperature during solarization is achieved at or nearby the soil surface. However, soil temperature decreases with increasing depth (Stapleton 2000).

In this research, the temperature increased to 41°C with solarization treatment for 14 days and gradually decreased afterward. The relationship between temperature and solarization duration was polynomial ($R^2 = 0.98$). According to Nelly et al. (2005), predictions or models expressed by polynomial regression show that the value of y is determined by x and the power x. Furthermore, modeling the effect of solarization duration on temperature as described by the polynomial regression equation showed that the increase in temperature occurred by increasing the solarization duration to a certain point, it remains constant or decreases gradually.

The differences in soil temperature are thought to have contributed to the presence of nematodes. All genera of parasitic nematodes were found in the soil treated with different solarization. Except for Pratylenchus parasite nematodes were not present in the 3 days solarization treatment, and Paratylenchus nematodes were only in the control treatment and the farmer's treatment. According to Triman and Mulyadi (2001), parasitic nematode plants can live in temperatures between 15-30°C whereas at low temperatures around 5-15°C and very high temperatures ranging from 30-40°C these organisms become inactive. Similarly, entomopathogenic nematodes can also live and develop at temperatures above 25°C because these particular genera become inactive resulting in reduced effectiveness in controlling the host. Temperature affects lipids in entomopathogenic nematodes. Lipids represent the main source of energy in entomopathogenic nematodes (Andalo et al. 2011).

Based on Gaugler and Kaya (1990), the suitable water temperature for nematodes was 20.9 + 5.9°C, while the soil temperature was 20.1 + 4.5°C. In the solarization treatment, entomopathogenic nematodes were found, namely Steinernema and Heterorhabditis. Heterorhabditis nematodes were found in all solarization treatments, while Steinernema nematodes were not found in the 14 days solarization treatment. Soil temperature in the 14 days solarization treatment was 41°C, higher than the other solarization treatments. The study by Glazer and Salame (2000) showed the ability of entomopathogenic nematodes to survive and withstand the highest temperature conditions.

The abundance of nematode populations had different results in each solarization treatment. As shown in Table 3, some nematode genera were not found in certain treatments. Pratylenchus nematodes were not found in solarization treatments of 3, 7, 14, and 30 days. These nematodes were found in the control and treatment of farmers. In the treatment of farmers, all types of nematodes were found. The highest nematode diversity was found in the treatment of farmers and it was not different from the control. The lowest nematode diversity was found in the 30 days solarization treatment but it was not significantly different from the 3, 7, and 14 days solarization treatment. Biotic and abiotic factors influence the diversity of
nematodes in the land. Among these factors that influence it are soil temperature and humidity (Sagita et al. 2017).

In conclusion, differences in tillage treatment and solarization affect soil temperature. The relationship between increasing solarization time and soil temperature is described by a polynomial regression model. The highest soil temperature was found in cultivated and solarized land for 14 days at 41.0°C and the lowest in the treatment of farmers, namely 33.67°C. Solarization treatment can best be applied in the field for 14 days. The nematode genera found in the observation field include; *Aphelenchoides, Helicotylenchus, Meloidogyne, Paratylenchus, Pratylenchus, Tylenchus, Heterorhabditis*, and *Steinernema*.

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