Potential Role of Probiotics in Ameliorating Psoriasis by Modulating Gut Microbiota in Imiquimod-Induced Psoriasis-Like Mice

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Abstract: Psoriasis is an immune-mediated systemic disease that may be treated with probiotics. In this study, probiotic strains that could or could not decrease interleukin (IL)-17 levels were applied to imiquimod (IMQ)-induced psoriasis-like mice via oral administration. Bifidobacterium adolescentis CCFM667, B. breve CCFM1078, Lacticaseibacillus paracasei CCFM1074, and Limosilactobacillus reuteri CCFM1132 ameliorated psoriasis-like pathological characteristics and suppressed the release of IL-23/T helper cell 17 (Th17) axis-related inflammatory cytokines, whereas B. animalis CCFM1148, L. reuteri CCFM1147, and L. reuteri CCFM1040 neither alleviated the pathological characteristics nor reduced the levels of inflammatory cytokines. All effective strains increased the contents of short-chain fatty acids, which were negatively correlated with the levels of inflammatory cytokines. By performing 16S rRNA gene sequencing, the diversity of gut microbiota in psoriasis-like mice was found to decrease, but all effective strains made some specific changes to the composition of gut microbiota compared to the ineffective strains. Furthermore, except for B. breve CCFM1078, all other effective strains decreased the abundance of the family Rikenellaceae, which was positively correlated with the levels of inflammatory cytokines. These findings demonstrated effects of strain-specificity, and how probiotics ameliorated psoriasis and provide new possibilities for the treatment of psoriasis.

Keywords: psoriasis; probiotics; IL-23/Th17 axis; gut microbiota

1. Introduction

Psoriasis is a skin disease with erythema and scales as the main clinical manifestations. Psoriasis was originally thought to be a disease of epidermal keratinocytes, but is now considered one of the most common immune-mediated diseases [1]. The prevalence of psoriasis worldwide is approximately 2%, but varies according to different regions [2]. Although psoriasis occurs at any age, its incidence is highest between 18 and 39 years of age, or between 50 and 69 years of age [3]. The typical clinical manifestations of psoriasis are scales and erythema, localised or widely distributed [4]. According to the different clinical manifestations of diseased skin, psoriasis is classified as plaque-type psoriasis, guttate psoriasis, inverse psoriasis, pustular psoriasis, palmoplantar psoriasis, and erythrodermic psoriasis [5]. Although there have been a large number of studies on psoriasis, the pathogenesis of psoriasis is not fully understood. It has been recently confirmed that the pathogenesis of psoriasis is not due to a single cause, and involves many aspects, including genetic, immunological, environmental, and other factors [6]. Additionally, psoriasis is an innate and adaptive immune system disease in which keratinocytes, dendritic cells,
and T cells play a central role [7]. Several abnormalities occur during the development of psoriasis, including in antigen presentation, activation of the nuclear factor kappa-B signalling pathway, T helper (Th) cells population differentiation (especially Th17 cells), and enhancement of IL-17 responses [6,8]. Recently, more attention has been paid to the interleukin (IL)-23/Th17 axis in psoriasis. IL-23 drives the differentiation of Th17 cells, and Th17 cells subsequently produce IL-22 and IL-17 [9]. These inflammatory cytokines, especially IL-17, accelerate the development of psoriasis [10]. Due to the complexity of psoriasis pathogenesis, there are no methods to completely cure and not relapse after the drug is stopped. Biologics are currently the most effective method for treating psoriasis and having certain safety. However, biological preparations are usually expensive. Many kin of biologics, including anti-IL-12/23p40 antibody and IL-17 inhibitors [11], have been used to treat psoriasis. The application of IL-17 inhibitors, including secukinumab, ixekizumab, and brodalumab [11], has confirmed that IL-17 is a therapeutic target for psoriasis. Therefore, we hypothesised that probiotics that suppress the release of IL-17 may be used to treat psoriasis.

The gut is an important digestive organ, and a microbiota niche. The gut microbiota population is in dynamic equilibrium, with many functions, including resistance to pathogen invasion, maintenance of gut homeostasis [12], metabolism of carbon and nitrogen [13], and nourishment of the host [14]; therefore, human health is closely associated with gut microbial alteration. In addition, the gut microbiota regulates immunity through its metabolites; short-chain fatty acids (SCFAs), products of carbohydrate digestion by gut microbiota, promote T cell differentiation into regulatory T (Treg) cells [15], and this in turn blocks the differentiation of other T cells, such as Th17 cells [16]. A large number of studies have reported that there are gut microbial disorders in patients with psoriasis. Hidalgo et al. [17] and Scher et al. [18] have both reported a significant decrease in gut microbial diversity. At the phylum level, the percentage of Firmicutes significantly increases while that of Bacteroidetes significantly decreases, contributing to an elevated ratio of Firmicutes/Bacteroidetes; additionally, there is a positive correlation between the ratio and psoriasis severity [19,20]. At the genus level, the relative abundance of Akkermansia and Ruminococcus significantly decrease; these genera may produce SCFAs and then induce Treg differentiation or maintain intestinal homeostasis [18,21]. At the species level, the relative abundance of Akkermansia muciniphila and Faecalibacterium prausnitzii significantly decreased; both are known as beneficial gut microbiota [21,22]. Therefore, changes in gut microbiota are important for psoriasis development, and regulating gut microbiota may be an alternative treatment for psoriasis.

Severe psoriasis has been related to nutrient deficiency, because hyperproliferation and desquamation of the skin epidermis will accelerate nutrient loss [23]. Besides this, reports in the literature have proven that certain dietary patterns have anti-inflammatory effects, and thus have the effect of alleviating psoriasis [24]. Therefore, it is feasible to improve psoriasis by adjusting diet. Probiotics are defined as living microorganisms that have a beneficial effect on the health of the host when ingested in sufficient quantities. The main food source of a probiotic is dairy products. Probiotics also have the effect of improving gut microbial homeostasis and have potential immunomodulatory effects [25,26]. Chen et al. have applied Lactobacillus pentosus GMNL-77 to imiquimod (IMQ)-induced psoriasis-like mice through intragastric administration, resulting in reduced pathological features in mice [27]. Rather et al. have applied L. sakei proBio-65 extract to IMQ-induced psoriasis-like mice through topical application, which also resulted in reduced pathological features in mice [28]. Although the mechanism was not spelled out, both studies found that levels of IL-23/Th17 axis-related inflammatory cytokines, including IL-23, IL-22, and IL-17, decreased after probiotic treatment. Therefore, we hypothesize that increasing the intake of foods with anti-inflammatory probiotics can ameliorate psoriasis.

In this study, probiotic strains that could or could not decrease IL-17 levels in other animal pathological models, including mice with asthma [29], constipation, and rheumatoid arthritis (as measured in other experiments, some data are not shown), were applied
to IMQ-induced psoriasis-like mice. The purpose of this study was to (1) compare the ameliorating effects of different probiotic strains on the pathological characteristics of psoriasis; (2) compare suppressing effects of different probiotic strains on inflammatory cytokine levels associated with psoriasis; (3) compare modulating effects of different probiotic strains on gut microbiota; (4) perform correlation analysis to explore how probiotics ameliorate psoriasis.

2. Materials and Methods

2.1. Bacterial Strains

The strains used in this study were kept in the Culture Collection of Food Microorganisms (CCFM) of Jiangnan University (Wuxi, Jiangsu, China).

*Lactobacillus* strains were cultured at 37 °C for 16 h under aerobic conditions with de Man, Rogosa, and Sharpe (MRS) broth (Sinopharm Chemical Reagent, Shanghai, China). *Bifidobacterium* strains were cultured at 37 °C for 30 h under anaerobic conditions with modified MRS broth containing 0.05% (w/v) L-cysteine-HCl (Sinopharm Chemical Reagent). Detailed information on the probiotics is shown in Table 1. The strains were cultured for three generations and centrifuged at 8000 rpm for 10 min (Eppendorf, Hamburg, Germany) to remove the supernatant. The strains were washed with phosphate buffered saline (PBS, Sinopharm Chemical Reagent) three times and then resuspended in normal saline at a concentration of $5 \times 10^9$ colony-forming units (CFU)/mL.

Table 1. Information of strains.

| Strain Number | Strain Original Number | Genus/Species          | Origin  |
|---------------|------------------------|------------------------|---------|
| CCFM667       | CCFM667                | *Bifidobacterium adolescens* |         |
| CCFM1078      | JSWX17M1               | *Bifidobacterium breve*  |         |
| CCFM1148      | JSWX23M8               | *Bifidobacterium animalis* |         |
| CCFM1074      | FJSWX1M3               | *Lactobacillus paracasei* | CCFM   |
| CCFM1147      | VCQQJ4174M3            |                        |         |
| CCFM1032      | FZJZ20M3               | *Limosilactobacillus reuteri* |         |
| CCFM1040      | FYNDL13                |                        |         |

CCFM: The Culture Collection of Food Microorganisms of Jiangnan University (Wuxi, Jiangsu, China).

2.2. Animal Experiments

Female BALB/c mice (6–8 weeks, 18–20 g, Beijing Vital River Laboratory Animal Technology, Beijing, China) were housed in a barrier condition. The temperature was 23 °C and the humidity was 55%. All animal procedures were performed according to the European Community guidelines (Directive 2010/63/EU) and approved by the Ethics Committee of Jiangnan University (JN. No20200515b1040650(066)).

After 1 week of adaptation, mice were separated into 10 groups: control group, IMQ (Aldara, 3M Pharmaceuticals, St. Paul, MN, USA) group, methotrexate (MTX, SPH Sine Pharmaceutical Laboratories, Shanghai, China) positive control group, and probiotic groups (seven groups). Six mice were included in each group. The probiotic groups were intra-gastrically administered 200 µL of strain suspension daily for 2 weeks, while the other mice received intragastric administration of the same volume of sterile saline. Two weeks later, shaving the dorsal skin and applying 62.5 mg IMQ cream to the dorsal skin and 20 mg to the right ear daily for seven consecutive days was performed [30], while mice in the control group received the same volume of Vaseline (Lircon Medical Technology, Dezhou, Shandong, China). During that time, probiotic groups received strain suspensions, the control and IMQ groups received sterile saline, while the MTX group received methotrexate dissolved in normal saline at 1 mg/kg/day [31,32]. Mice were sacrificed on the 8th day (Figure 1A).
pensions, the control and IMQ groups received sterile saline, while the MTX group received methotrexate dissolved in normal saline at 1 mg/kg/day [31,32]. Mice were sacrificed on the 8th day (Figure 1).

**Figure 1.** Effects of probiotics on psoriasis-like pathological characteristics. (A) Animal experiment design. (B) Ear thickness of mice. (C) Spleen weight of mice at D7. **p < 0.01 and ****p < 0.0001.

2.3. Ear Thickness Determination and Dorsal Skin Score

The ear thickness of the mice was measured with a digital vernier calliper (Guanglu Measuring Instrument, Guilin, Guangxi, China) daily during the IMQ receiving period. The lesion skin of mice was scored in terms of the clinical psoriasis area and severity index (PASI). Severity of thickening, scaling, and erythema were scored separately on a scale from 0–4: 0, none; 1, slight; 2, moderate; 3, marked; 4, very marked. Cumulative Score was the sum of the scores of the three.

2.4. Skin Histopathology

After mice were sacrificed, the skin was removed, sliced, embedded in paraffin wax (Sigma Aldrich, St. Louis, MO, USA), and stained with haematoxylin and eosin (Sigma Aldrich) for further microscopic examination. The digital scanner (Pannoramic MIDI, 3DHistech, Budapest, Hungary) was used to scan photomicrographs (20×).

2.5. Skin Cytokine Analysis

Crushing dorsal skin samples (100 mg) in 900 µL radioimmunoprecipitation assay lysis buffer (Beyotime Biotechnology, Shanghai, China) containing 2% (v/v) protease inhibitor mixture (Beyotime Biotechnology) and 2% (v/v) phosphatase inhibitor mixture (Beyotime Biotechnology) with the grinder (Scientz48, Scientz Biotechnology, Ningbo, Zhejiang, China) at 60 Hz for 30 s, five times. According to the manufacturer’s instructions, IL-22, IL-23, and IL-17 levels were measured using enzyme-linked immunosorbent assay (ELISA) kits (Enzyme-linked Biotechnology, Shanghai, China), and total protein levels were measured using an enhanced bicinchoninic acid protein assay kit (Beyotime Biotechnology).

2.6. SCFA Analysis

SCFA levels in the caecum (50 mg) were measured by gas chromatography-mass spectrometer (GC-MS, Shimadzu Corp., Kyoto, Japan) using a previously published method [33].
2.7. 16S rRNA Amplification Sequencing of Faecal Samples

A swab (Huangchenyang Technology, Shenzhen, Guangzhou, China) was used to collect skin bacteria. Faecal samples and swab samples were collected the day before euthanasia and frozen at −80 °C. According to the manufacturer’s instructions, DNA from faecal samples was extracted using a Fast DNA spin kit for faeces (MP Biomedicals, Santa Ana, CA, USA). The extracted DNA was then amplified by polymerase chain reaction (PCR). The V4 region of faecal samples of the 16S rRNA gene were amplified using the primers 341F and 806R. The detailed protocol has been described previously [34]. PCR products were purified using a TIANgel mini purification kit (Tiangen Biotech, Beijing, China) and then sequenced using the Illumina sequencing platform (Miseq, Illumina, Santiago Canyon, CA, USA).

2.8. Bioinformatics Analysis

All sequences were analysed using Quantitative Insights into Microbial Ecology (QIIME2). Alpha and beta diversity analyses were also performed using QIIME2. Heatmap analysis was performed using the MetaboAnalyst website (https://www.metaboanalyst.ca/MetaboAnalyst/ModuleView.xhtml, accessed on November 2020). The data were normalised by the median, transformed by log transformation, and scaled by the centred mean. Heat tree analysis and linear discriminant analysis (LDA) effect size were performed using the MicrobiomeAnalyst website (https://www.microbiomeanalyst.ca/MicrobiomeAnalyst/upload/OtuUploadView.xhtml, accessed on December 2020). The low count filter was a 20% prevalence filter, whereas the low-variance filter was based on the inter-quantile range. The data were normalised using total sum scaling. Differences were regarded significant at \( p < 0.05 \).

2.9. Statistical Analysis

Major statistical analyses were performed using GraphPad Prism (version 8, GraphPad, La Jolla, CA, USA) and the Statistical Package for the Social Sciences (version 22, SPSS, Chicago, IL, USA). Data are the mean ± standard error of the mean. The differences between multiple groups were calculated using one-way analysis of variance and Duncan’s multiple range test while \( p < 0.05 \) was considered to indicate statistical significance. \( * p < 0.05, ** p < 0.01, *** p < 0.001 \) and \( **** p < 0.0001 \). Differences between multiple groups were calculated using the Tukey–Kramer method, and differences were considered significant at \( p < 0.05 \).

3. Results

3.1. CCFM667, CCFM1078, CCFM1074, and CCFM1132 Ameliorated Psoriasis-Like Pathological Characteristics

In previous experiments, it was found that \( B. \) adolescentis CCFM667 reduced IL-17 levels in mice did ameliorate psoriasis, while \( B. \) animalis CCFM1148 and \( L. \) paracasei CCFM1147 failed to reduce IL-17 levels and did not. Therefore, in the selection of strains in this experiment, we selected strains that could or could not reduce the IL-17 level in other animal pathological models, including mice with asthma [29], constipation, and rheumatoid arthritis (as measured in other experiments, some data are not shown).

Continuous application of IMQ resulted in thickening of the ears (Figure 1B), swelling of spleens (Figure 1C), and erythema, scaling, and thickening of the dorsal skin of mice (Figure 2). These results are consistent with literature reports [30,35,36]. Except for \( L. \) paracasei CCFM1147, all probiotics effectively ameliorated the swelling of ears. In the spleen, only MTX and \( B. \) adolescentis CCFM667 effectively ameliorated spleen swelling.
With respect to the pathological characteristics of dorsal skin, MTX, B. adolescentis CCFM667, B. breve CCFM1078, L. paracasei CCFM1074, and L. reuteri CCFM1132 effectively ameliorated erythema, scaling, and thickening, while B. animalis CCFM1148, L. paracasei CCFM1147, and L. reuteri CCFM1040 had almost no effect. Although the skin thickening in the B. animalis CCFM1148 group was not serious, this group had very serious erythema. Pathological sections showed that the epidermal structure of the control group consisted of only one or two layers of cells, while the epidermal structure of the IMQ group was clearly thickened (Figure 3). Consistent with the pathological characteristics, MTX, B. adolescentis CCFM667, B. breve CCFM1078, L. paracasei CCFM1074, and L. reuteri CCFM1132 effectively reduced epidermal thickness. Therefore, B. adolescentis CCFM667, B. breve CCFM1078, L. paracasei CCFM1074, and L. reuteri CCFM1132 were considered effective in relieving psoriasis-like pathological characteristics, while B. animalis CCFM1148, L. paracasei CCFM1147, and L. reuteri CCFM1040 were considered to be ineffective strains.

Figure 2. Effects of probiotics on dorsal skin. (A) Representative photographs of dorsal skin of mice from D3 to D7. (B) PASI of mice in different groups. Cumulative Score is the sum of the scores of the severity of thickening, scaling, and erythema.

Figure 3. Representative photomicrographs of haematoxylin and eosin sections of dorsal skin tissue.
3.2. CCFM667, CCFM1078, CCFM1074, and CCFM1132 Suppressed the Psoriasis-Like Immune Response

Psoriasis is an immune-mediated skin disease, and the IL-23/Th17 axis takes part in its progression [5,6]. IL-23 drives the differentiation of Th17 cells, and Th17 cells produce IL-22 and IL-17 [10]. Therefore, levels of IL-23, IL-22, and IL-17 in skin lesions of mice were determined. Continuous application of IMQ resulted in increased levels of IL-23, IL-22, and IL-17, while MTX significantly suppressed this trend (Figure 4). In the probiotic groups, B. breve CCFM1078 significantly suppressed expression of all three cytokines, while B. adolescentis CCFM667, L. paracasei CCFM1074, and L. reuteri CCFM1132 significantly suppressed expression of two cytokines. B. animalis CCFM1148, L. paracasei CCFM1147, and L. reuteri CCFM1040 had no effect on ameliorating psoriasis-like symptoms and did not suppress the psoriasis-like immune response (Figure 2). Therefore, it was concluded that B. adolescentis CCFM667, B. breve CCFM1078, L. paracasei CCFM1074, and L. reuteri CCFM1132 were effective in ameliorating psoriasis through the IL-23/Th17 axis.

![Figure 4](image-url) Effects of probiotics on levels of IL-23/Th17 axis-associated inflammatory cytokines. (A) IL-23 levels (n = 6). (B) IL-22 levels (n = 6). (C) IL-17 levels (n = 6). * p < 0.05, ** p < 0.01, *** p < 0.001 and **** p < 0.0001.

3.3. Probiotics Exerted Different Effects on SCFA Metabolism

The main products of intestinal microbial fermentation are SCFAs, especially acetate, propionate, and butyrate [37]. Acetate, propionate, and butyrate levels were not significantly different in the control and IMQ groups (Figure 5A). Compared to the IMQ groups, acetate level in the B. breve CCFM1078, L. paracasei CCFM1074, and L. reuteri CCFM1132 groups significantly increased, while propionate levels in the B. adolescentis CCFM667 and L. paracasei CCFM1074 groups significantly increased, but butyrate level in all groups showed no significant difference. In short, the strains that effectively ameliorated psoriasis increased the levels of acetate or propionate.

Further correlation analysis was performed between the levels of SCFAs in the colon content and the levels of inflammatory cytokines in the skin tissue; the result showed that the level of acetate was significantly negatively correlated with the levels of IL-17 and IL-23, and the level of propionate was significantly negatively correlated with the levels of IL-23 (Figure 5B). It was concluded that probiotics that effectively alleviated psoriasis could inhibit the release of IL-23/Th17 axis inflammatory cytokines by promoting the production of SCFAs.
3.4. Probiotics Exerted Different Effects on Gut Microbial Composition

Changes in gut microbial diversity in patients with psoriasis have been reported in a large number of studies [18,19,21]. The α diversity is shown in Figure 6A, based on different indicators, including the Shannon, Evenness, and Faith indices, and observed operational taxonomic units. Considering species and abundance, the Shannon index showed that compared to the control group, the α diversity of the IMQ group significantly decreased, while application of *B. breve* CCFM1078, *L. paracasei* CCFM1074, and *L. paracasei* CCFM1147 significantly suppressed this trend. However, this trend was not observed when the evolutionary relationship was taken into consideration, as shown by the Faith index. The β diversity is shown in Figure 6B, based on the Bray–Curtis distance, which reveals the decline of common species between the control and IMQ groups. Only MTX and *L. paracasei* CCFM1074, which both relieved psoriasis-like pathological characteristics, suppressed this trend. The relative abundance at the phylum level is shown in Figure 6C. Bacteroidetes and Firmicutes had the highest abundance, and the ratio of Firmicutes to Bacteroidetes (F/B) in the control and IMQ groups was not significantly different, while MTX and *L. paracasei* CCFM1074, which both relieved psoriasis-like pathological characteristics, increased the F/B ratio (Figure 6D). These results showed that probiotics exerted different effects on the regulation of gut microbial composition.
3.5. Key Differences between Effective and Ineffective Probiotics in Ameliorating Psoriasis

To better explore the effect of probiotics on gut microbial composition, the gut microbiota at the genus level in different groups of mice was analysed using heatmap analysis (Figure 7). Although the control and IMQ groups were clustered into the same category, the abundance of some gut microbiota in the IMQ group was significantly reduced. In addition, *L. reuteri* CCFM1040 and CCFM1132, *L. paracasei* CCFM1074 and CCFM1137, and *Bifidobacterium* CCFM1078, CCFM1148, and CCFM667 were clustered into the same category. Therefore, it was speculated that the effects of probiotics of the same genus or species were similar in gut microbial composition in general, but there should be some key differences that made certain probiotics effective in ameliorating psoriasis, while others were ineffective.

To explore the key differences that made certain probiotics effective in ameliorating psoriasis, heat tree analysis [38] was performed. Differential gut microbiota between the control and IMQ groups, as well as *L. reuteri* CCFM1040 and CCFM1132 groups, *L. paracasei* CCFM1074, and CCFM1137 groups, *Bifidobacterium* CCFM1148, and CCFM667 groups, and *Bifidobacterium* CCFM1148, and CCFM1078 groups are shown in Figure 8 (p < 0.05).
To explore the key differences that made certain probiotics effective in ameliorating psoriasis, heat tree analysis [38] was performed. Differential gut microbiota between the control and IMQ groups, as well as L. reuteri CCFM1040 and CCFM1132 groups, L. paracasei CCFM1074, and CCFM1137 groups, Bifidobacterium CCFM1148, and CCFM667 groups, and Bifidobacterium CCFM1148, and CCFM1078 groups are shown in Figure 8 (p < 0.05).

Figure 7. Clustering analysis of gut microbiota at the genus level (n = 5/6).

Figure 8. Heat tree analysis of gut microbiota at the genus level (n = 5/6). Nodes marked with text represent a significance difference between the two groups, p < 0.05.
Based on heat tree analysis, the abundance of significantly differentiated gut microbiota was analysed further (Figure 9). Figure 9A shows the changes in the abundance of the family Rikenellaceae and that the genus Dorea in the IMQ group was suppressed by the effective strains, while the ineffective strains did not have such function. Figure 9B shows the increase in the abundance of the phylum Proteobacteria and the genus Bilophila in the effective Bifidobacterium CCFM1078 and CCFM667 groups compared to the ineffective Bifidobacterium CCFM1148 group. Figure 9C shows the increase in the abundance of the species ASF356 and Lachnospiraceae FCS020 group in the effective L. paracasei CCFM1074 group compared to the ineffective L. paracasei CCFM1147 group. This phenomenon was not found in the L. reuteri CCFM1040 and CCFM1132 groups. It was suggested that the above changes in the abundance of gut microbiota may be responsible for the different effects of the strains on psoriasis.

**Figure 9.** Difference analysis of gut microbiota between groups (n = 5/6). The abundance of key differential gut microbiota between (A) the control and IMQ groups, (B) the Bifidobacterium CCFM1078, CCFM667, CCFM1148 groups, (C) the L. paracasei CCFM1074 and CCFM1147 groups. *p < 0.05, **p < 0.01, ***p < 0.001 and ****p < 0.0001.

To further explore the role of the above differential gut microbiota, a correlation analysis was performed (Figure 10). The family Rikenellaceae, which was different between the control and IMQ groups, was positively correlated with spleen weight, ear thickness, cumulative scores, and IL-23/Th17 axis-associated inflammatory factor levels, while it was negatively correlated with the propionate level. This suggested that the development of psoriasis may lead to an increase in the abundance of the family Rikenellaceae, while B. adolescentis CCFM667, L. paracasei CCFM1074, and L. reuteri CCFM1132 reduced its abundance (Figure 9A). The Lachnospiraceae FCS020 group, which was different between the CCFM1074 and CCFM1147 groups (Figure 9C), was negatively correlated with IL-23 level. This suggested that the development of psoriasis may lead to a decrease in the abundance of the Lachnospiraceae FCS020 group, while CCFM1074 promoted its abundance. In addition, the genera Dorea and Bilophila, and the species ASF356 were significantly correlated with SCFA levels, but they did not correlate with psoriasis-like pathological characteristics.
Figure 10. Spearman correlation analysis between the key gut microbiota and psoriasis-like pathological characteristics.

In summary, the increase in the abundance of the family Rikenellaceae may be related to the development of psoriasis, while B. adolescentis CCFM667, L. paracasei CCFM1074, and L. reuteri CCFM1132 had the effect of restraining this trend.

4. Discussion

Psoriasis is an immune-mediated skin disease, and the IL-23/Th17 axis plays an important part in its progression. In the IL-23/Th17 axis, IL-23 induces the differentiation of Th17 cells and then promotes the secretion of IL-22 and IL-17, which eventually leads to the aggravation of psoriasis [39]. IL-23 inhibitors, such as guselkumab, risankizumab and tildrakizumab, are used in the management of psoriasis and other Th17 mediated inflammatory diseases [40,41]. Besides this, IL-17 inhibitors, including secukinumab, ixekizumab, and brodalumab [11], are also widely used to relieve psoriasis. Probiotics can affect the differentiation of T-lymphocyte through their metabolites on T-lymphocytes, thereby affecting the number of Th17 cells in the host, and thus affecting IL-17 levels. A study has reported that obese people have lower levels of IL-17 in their blood after ingesting yogurt containing probiotics [42]. Based on this, probiotic strains that could or could not decrease IL-17 levels in other animal pathological models, including mice with asthma [29], constipation, and rheumatoid arthritis (as measured in other experiments, some data are not shown) were selected and applied to IMQ-induced psoriasis-like mice to compare the ameliorating effects of the strain on psoriasis. ELISA results showed that B. adolescentis CCFM667, B. breve CCFM1078, L. paracasei CCFM1074, and L. reuteri CCFM1132 decreased levels of IL-23/Th17 axis-related inflammatory cytokines, including IL-23, IL-22, and IL-17, while B. animalis CCFM1148, L. paracasei CCFM1147, and L. reuteri CCFM1040 did not. Similarly, B. adolescentis CCFM667, B. breve CCFM1078, L. paracasei CCFM1074, and L. reuteri CCFM1132 ameliorated psoriasis-like pathological characteristics, including skin lesion PASI scores, ear thickness, and spleen weight, while B. animalis CCFM1148, L. paracasei CCFM1147, and L. reuteri CCFM1040 did not. Based on the above results, it is preliminarily believed that probiotics have the potential to ameliorate psoriasis by decreasing the levels of IL-23/Th17 axis-related inflammatory cytokines. This is consistent with that reported by...
Zuzana et al., who declared that gut microbiota directly regulates imiquimod-induced skin inflammation [43]. Other studies have also reported the function of probiotics in decreasing levels of IL-23/Th17 axis-related inflammatory cytokines [44,45].

In recent years, increasing attention has been paid to the relationship between intestinal microecology and human health. SCFAs, especially acetate, propionate, and butyrate [37], are the main products of gut microbial fermentation [37]. It has been reported that SCFAs could promote the differentiation of Treg cells and then affect the balance of Th17/Treg, so as to alleviate the occurrence of psoriasis [17,46]. Furthermore, Khyshtuk-tuev et al. have found decreased levels of acetate and propionate in the serum of patients with psoriasis [47]. In this study, B. breve CCFM1078, L. paracasei CCFM1074, and L. reuteri CCFM1132 increased acetate levels, and B. adolescentis CCFM667 and L. paracasei CCFM1074 increased propionate levels, while the ineffective strains B. animalis CCFM1148, L. paracasei CCFM1147, and L. reuteri CCFM1040 did neither. This suggested that acetate and propionate played a role in the alleviation of psoriasis. Besides this, the levels of acetate and propionate were significantly negatively correlated with the levels of IL-23/Th17 axis-related inflammatory cytokines. It was concluded that probiotics that effectively alleviated psoriasis could inhibit the release of IL-23/Th17 axis inflammatory cytokines by promoting the production of SCFAs.

Human health is closely related to the changes of gut microbiota, and a large number of studies have reported the changes of intestinal flora diversity in patients with psoriasis [19,20,48]. Many studies have also reported that there is a significant decrease in gut microbial diversity [17,18]. In this study, the α and β diversity of the gut microbiota in psoriasis-like mice decreased, and the relative abundance changed. Some studies report an increased ratio of F/B in patients with psoriasis, which is associated with altered carbohydrate metabolism of medium-chain fatty acids and SCFAs [19,49], while Masallat et al. have found that the F/B ratio is positively correlated with the PASI score [20]. However, in this study, the F/B ratio in the control and IMQ groups showed no significant difference, but was increased in the MTX and CCFM1074 groups, which both relieved psoriasis-like pathological characteristics. However, Scher et al. report a decreased abundance of Bacteroidetes in patients with psoriasis [18]. Therefore, the role of F/B ratio in the development of psoriasis needs further study.

Some studies have reported strain-specific effects of probiotics in ameliorating disease [50,51]. In this study, heatmap analysis showed that probiotics of the same genus or species had similar effects in modulating gut microbiota, although the effects of these probiotics in ameliorating psoriasis were different. Based on this, the abundance of gut microbiota was compared between the effective and ineffective probiotic groups, and some key differential gut microbiota were identified. The abundance of the family Rikenellaceae in the IMQ group significantly increased, while the effective strains, B. adolescentis CCFM667, L. paracasei CCFM1074, and L. reuteri CCFM1132, decreased its abundance. Meanwhile, the abundance of the family Rikenellaceae was positively correlated with psoriasis-like pathological characteristics and negatively correlated with propionate level. This suggested that the development of psoriasis may have correlation with the increase in the abundance of the family Rikenellaceae. Similarly, Sun et al. have found that the Th1 response in the colon of mice is enhanced after receiving antibiotics, while the abundance of the family Rikenellaceae increased [52], suggesting that Rikenellaceae abundance is related to immunity, but its specific mechanism remains unknown. Compared to the L. paracasei CCFM1147 group, L. paracasei CCFM1074 increased the abundance of the Lachnospiraceae FCS020 group, which was positively correlated with IL-23 levels. This suggested that the development of psoriasis may lead to a decrease in the abundance of the Lachnospiraceae FCS020 group, which was related to inflammatory responses. The key differential gut microbiota in the B. breve CCFM1078 group was not identified, but the B. breve CCFM1078 strain largely ameliorated psoriasis, and was most successful in suppressing inflammatory responses of all the probiotic groups; therefore, it was speculated that B. breve CCFM1078 ameliorated psoriasis directly through the IL-23/IL-17 axis rather than exerting effects on
gut microbiota. Due to the strain-specificity, this study believes that the effective strains can be developed into products to enable patients to increase the intake of the effective strains, thereby ameliorating psoriasis. In addition, probiotics could only alleviate psoriasis to a certain extent, and could not replace drugs. Probiotic intake should be used as a supplement to medication.

In this study, we found that probiotics could ameliorate the pathological characteristics of psoriasis in mice by inhibiting the IL-23/Th17 axis-related inflammatory response and regulating gut microbiota, providing a new possibility for the treatment of psoriasis. However, there are still shortcomings in this study, such as the lack of repeated experiments to verify the results. In addition, tests of immune cells were lacking to strengthen the link between gut microbial composition and immunological changes in the skin. These will be the direction of future research. In the future, this study will further confirm whether the effective strains have the same phenomenon and effects in the human body, and further explore the mechanisms.

5. Conclusions

In summary, the effects of seven probiotic strains in ameliorating psoriasis were explored in IMQ-induced psoriasis-like mice. B. adolescentis CCFM667, B. breve CCFM1078, L. paracasei CCFM1074, and L. reuteri CCFM1132 ameliorated psoriasis via suppressing the release of IL-23/Th17 axis-related inflammatory factors by promoting the content of SCFAs, while B. animalis CCFM1148, L. paracasei CCFM1147, and L. reuteri CCFM1040 did not have such effects. Among these, B. breve CCFM1078 exhibited the best anti-inflammatory effects. The development of psoriasis may lead to an increase in the abundance of the family Rikenellaceae while the effective strains B. adolescentis CCFM667, L. paracasei CCFM1074, and L. reuteri CCFM1132 decreased its abundance. These results revealed differences in the regulation of immune response and gut microbiota by strains, so it was confirmed that effects were strain-specificity. This study demonstrated the potential of probiotics in the treatment of psoriasis.

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# Abbreviations

| Abbreviation | Description |
|--------------|-------------|
| CCFM         | Culture Collection of Food Microorganisms |
| EILISA       | Enzyme-linked Immunosorbent Assay |
| IL           | Interleukin |
| IMQ          | Imiquimod |
| LDA          | Linear Discriminant Analysis |
| MRS          | Man, Rogosa and Sharpe |
| MTX          | Methotrexate |
| OD           | Optical density |
| PASI         | Psoriasis Area and Severity Index |
| PCA          | Principal Components Analysis |
| PCR          | Polymerase Chain Reaction |
| SCFAs        | Short-Chain Fatty Acids |
| Th           | Helper T cells |
| Tregs        | Regulatory cells |

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