Technologies for the treatment of source-separated urine in the eThekwini Municipality

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ABSTRACT

In recent years, a large number of urine-diverting dehydration toilets (UDDTs) have been installed in eThekwini to ensure access to adequate sanitation. The initial purpose of these toilets was to facilitate faeces drying, while the urine was diverted into a soak pit. This practice can lead to environmental pollution, since urine contains high amounts of nutrients. Instead of polluting the environment, these nutrients should be recovered and used as fertiliser. In 2010 the international and trans-disciplinary research project VUNA was initiated in order to explore technologies and management methods for better urine management in eThekwini. Three treatment technologies have been chosen for the VUNA project. The first is struvite precipitation, a technology which has already been tested in multiple projects on urine treatment. Struvite precipitation is a simple and fast process for phosphorous recovery. Other nutrients, such as nitrogen and potassium, remain in the effluent and pathogens are not completely inactivated. Therefore, struvite precipitation has to be combined with other treatment processes to prevent environmental pollution and hygiene risks. The second process is a combination of nitrification and distillation. This process combination is more complex than struvite precipitation, but it recovers all nutrients in one concentrated solution, ensures safe sanitisation and produces only distilled water and a small amount of sludge as by-products. The third process is electrolysis. This process could be used for very small on-site reactors, because conversion rates are high and the operation is simple, as long as appropriate electrodes and voltages are used. However, nitrogen is removed and not recovered and chlorinated by-products are formed, which can be hazardous for human health. While urine electrolysis requires further research in the laboratory, struvite precipitation and nitrification/distillation have already been operated at pilot scale.

Keywords: Sanitation, source separation, nutrient recovery, eutrophication, hygiene, human health

BACKGROUND

In 2001, the South African Government issued a White Paper on Basic Household Sanitation (DWAF, 2001) with the aim of clearing the sanitation backlog by 2014. In response, the eThekwini Municipality’s Water and Sanitation Unit (EWS) started an integrated development planning (IDP) process to provide basic water and sanitation to peri-urban and rural areas in the metropolitan area of Durban (Gounden et al., 2006). In addition to providing 200 ℓ of free potable water per household and day, EWS installed urine-diverting dry toilets (UDDTs) to achieve access to sanitation. Urine diversion was chosen for several reasons:

- Building waterborne sewage infrastructure was too costly and water supplies were limited
- Emptying of conventional ventilated improved pit latrines (VIPs) was costly and access for collection and transport trucks was often problematic
- After a sufficiently long dehydration time, faeces could be safely disposed of on-site
- No new pits would have to be periodically excavated
- The risk of environmental pollution would be limited
- Initially, urine was not collected but infiltrated into the ground via a soak pit. The implementation of the new sanitation system was successful given that about 75 000 households had received UDDTs by 2013. However, a survey conducted in December 2010 and January 2011 revealed that user satisfaction was low (Roma et al., 2013).

To increase the acceptance and sustainability of the UDDT-based sanitation approach, the research project VUNA was initiated by the Swiss Federal Institute of Aquatic Science and Technology (Eawag), EWS, the University of KwaZulu-Natal and the Swiss Federal Institute of Science and Technology in Zurich. The VUNA project is based on the vision that nutrient recovery from urine lowers the cost of sanitation, provides a valuable local fertiliser and reduces pollution of water resources. The project partners are guided by the belief that these advantages can lead to a higher acceptance of urine separation and sanitation in general.

In this paper, we will focus on the technical aspects of the VUNA project. After discussing challenges of urine treatment, we will present the processes which have been chosen for the VUNA project, and we will discuss some of the project results.
COMPOSITION OF STORED URINE

Urine which has been collected in UDDTs is very different from fresh urine (Udert et al., 2006). In the collection tanks, urea and other organic substances are degraded by bacteria. The results are high ammonia concentrations, a high pH value and a strong, pungent smell caused by organic degradation products (Udert et al., 2003a; Troccaz et al., 2013). Due to the high pH value, nearly half of the nitrogen can volatilise as free ammonia (Siegrist et al., 2013). The high pH value also triggers the precipitation of about 30% of the phosphate as calcium phosphate and struvite (Udert et al., 2006). Tables 1 and 2 provide overviews of the composition of stored urine. Urine contains the majority of the nutrients in household wastewater, with the main nutrients being nitrogen, phosphorus, potassium and sulphur (Larsen and Gujer, 1996). Due to cross-contamination from faeces, urine can contain pathogens (Höglund et al., 1998), so that sanitisation measures have to be considered when treating urine.

In recent years, pollution by trace organic contaminants, particularly pharmaceutical residues, has caught much attention among scientists, environmental protection agencies and the public. These trace organic compounds can have adverse effects on ecosystems and their organisms. Based on current studies, direct adverse health effects from pharmaceuticals occurring in drinking water are not expected (Bruce et al., 2010), however, possible chronic effects on humans may still be unknown. Our measurements have shown a high content of pharmaceutical residues in urine, especially of antibiotics, in the peri-urban areas of Durban (authors own unpublished data).

| TABLE 1 |
| --- |
| Compounds in stored urine, their reuse potential and possible negative effects |
| Compound | Beneficial reuses | Negative impacts |
| --- | --- | --- |
| Water | Recycling, for example for irrigation | Water volume requires large storage tanks |
| Nitrogen | Fertiliser | Water weight makes transport expensive |
| | Raw product for chemical industry | Smell and toxicity of gaseous ammonia |
| | | Environmental pollution: nitrate in ground-water, eutrophication of receiving waters and soils, fish toxicity, atmospheric particles |
| Phosphorus | Fertiliser | Environmental pollution (eutrophication) |
| | Raw product for chemical industry | Precipitates block pipes and valves |
| Bicarbonate | None | Precipitates block pipes and valves |
| Sulfur | Fertiliser | Environmental pollution |
| | Raw product for chemical industry | Smell and toxicity of hydrogen sulphide |
| Potassium | Fertiliser | Salinisation of agricultural soils and groundwater |
| | Raw product for chemical industry | |
| Sodium chloride | Raw product for chemical industry | Salinisation of agricultural soils and groundwater |
| Bulk organic substances | None | Pungent smell |
| | | Treatment problems such as foaming |
| | | Organic reduction of sulfate and nitrate: sulhide production or NO and N₂O production (pollutants, climate gases) |
| Trace organic compounds | None | Human health concerns |
| Pathogens | None | Human health concerns |

| TABLE 2 |
| --- |
| Concentrations in stored urine |
| Theoretical values are based on medical studies and include the morning urine (Udert et al., 2006). The values for the women's urine tank at Eawag were measured between 2011 and 2013 (between 5 and 106 samples; Etter et al., 2013). They are lower due to ammonia volatilisation, dilution and the absence of morning urine. |

| Variable | Unit | Theoretical value | Women’s tank at Eawag |
| --- | --- | --- | --- |
|  |  | Mean | Standard deviation |
| Total ammonia | mg N·ℓ⁻¹ | 8 100 | 1 790 ± 180 |
| Total phosphate | mg P·ℓ⁻¹ | 540 | 108 ± 14 |
| Potassium | mg·ℓ⁻¹ | 2 200 | 897 ± 103 |
| Sulphate | mg·ℓ⁻¹ | 1 500 | 316 ± 14 |
| Calcium | mg·ℓ⁻¹ | 0 | 10 ± 11 |
| Magnesium | mg·ℓ⁻¹ | 0 | < 5 |
| Sodium | mg·ℓ⁻¹ | 2 600 | 966 ± 314 |
| Chloride | mg·ℓ⁻¹ | 3 800 | 1 830 ± 320 |

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Urine treatment has to prevent possible negative effects of urine, such as environmental pollution, hygienic risks and malodour. Inadequate sanitation is reportedly the main source of high nitrate concentrations in South African groundwater (Tredoux, 2004) and a major cause of eutrophication in urban areas of sub-Saharan Africa (Nyenje et al., 2009). However, urine is also a source of valuable nutrients. By recovering these nutrients as a fertiliser, the nutrient cycle to agriculture is closed and the environmentally adverse production of synthetic fertilisers is reduced. Besides these beneficial effects for the environment, financial valorisation of human excreta could offset the costs of sanitation (Tilley and Günther, 2013).

Therefore, many justifications for the separate treatment of urine, but realistically, it will only be implemented on a large scale if the investment and operational costs are low and if the treatment technologies are reliable and easy to operate. Furthermore, nutrients have to be recovered in a form which allows direct application in agriculture or further treatment in the chemical industry.

**THE AIM OF URINE TREATMENT**

Urine treatment has to prevent possible negative effects of urine, such as environmental pollution, hygienic risks and malodour. Inadequate sanitation is reportedly the main source of high nitrate concentrations in South African groundwater (Tredoux, 2004) and a major cause of eutrophication in urban areas of sub-Saharan Africa (Nyenje et al., 2009). However, urine is also a source of valuable nutrients. By recovering these nutrients as a fertiliser, the nutrient cycle to agriculture is closed and the environmentally adverse production of synthetic fertilisers is reduced. Besides these beneficial effects for the environment, financial valorisation of human excreta could offset the costs of sanitation (Tilley and Günther, 2013).

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**TECHNOLOGIES FOR URINE TREATMENT**

A wide range of processes for urine treatment has been proposed (Maurer et al., 2006) but only a few have been tested in the laboratory or at pilot scale (Larsen et al., 2013). To our knowledge, only the following 4 processes have been operated at pilot scale prior to the VUNA project:

- A combination of electrodialysis, microfiltration and ozonation (Prokn et al., 2007)
- Phosphorus recovery by struvite precipitation (Etter et al., 2011; Antonini et al., 2011; Winkler et al., 2013)
- Ammonia stripping (Antonini et al., 2011; Winkler et al., 2013)
- Acidification and solar evaporation (Antonini et al., 2012)

In many studies, urine has also been applied without intensive treatment, but some measures have to be taken to prevent hygienic and environmental risks. Höglund et al. (2002) recommend storing the urine for 6 months to ensure that pathogens (especially viruses) are inactivated. Furthermore, gentle spreading techniques have to be used to prevent high ammonia losses (Johansson, 2001).

For the VUNA project, we decided to investigate the following 3 processes:

- Struvite precipitation, because it has already been tested extensively in pilot studies and is a simple process to be implemented in the field
- Nitrification/distillation, because it is currently the only process combination for complete recovery of all nutrients in a small volume without chemical addition
- Electrolysis, because it allows building very small reactors, which could be integrated into toilets. On-site treatment would cut the costs of transportation

**Struvite precipitation**

Struvite precipitation is probably the best understood process for nutrient recovery from source-separated urine, because it has been tested in multiple pilot projects (Etter et al., 2011; Antonini et al., 2011; Winkler et al., 2013). The mineral struvite (MgNH₄PO₄·6H₂O) can easily be precipitated from stored urine, because most of the requirements (high pH value, high ammonia and phosphate concentrations) are given; only a magnesium source has to be added to precipitate nearly all phosphate as struvite (Wilsenach et al., 2007). Struvite precipitation only takes a few minutes if a very soluble magnesium source is used (Etter et al., 2011). Typical magnesium sources are magnesium salts such as MgO, MgCl₂, and MgSO₄. The dissolution rate of MgO in urine is slightly slower than for MgCl₂ and MgSO₄, but MgO is substantially cheaper (Sakthivel et al., 2012). Some waste products are also suitable magnesium sources, for example bittern (waste brine left over after sea salt production; Ye et al., 2011) and wood ash (Sakthivel et al., 2012).

In the latter case, pollution with heavy metals and precipitation of unwanted minerals can be problematic. A low overdose of 10% (that is 1.1 mol Mg·mol⁻¹ PO₄) is already sufficient to precipitate 95% of the phosphate as struvite, but overall struvite recovery is often limited by the efficiency of crystal recovery, for example during filtration (Etter et al., 2011). In order to maximise phosphate recovery from urine, the spontaneously precipitated phosphate in the collection tank has to be recovered as well (see above). Overall, struvite precipitation is a process for phosphorus recovery; however, while nearly all the dissolved phosphate can be removed, more than 97% of the nitrogen, and practically all the potassium and sulphur remain in the effluent (Etter et al., 2011).

**Nitrification and distillation**

The goal of a nitrification/distillation reactor setup is to recover the maximum of the urine’s nutrient content (nitrification) and to concentrate the nutrients in a very low volume (distillation). By reducing the water volume, the costs for storage and transportation can be minimised. Nitrification is an aerobic biological process which involves 2 bacterial groups, i.e. ammonia-oxidising bacteria and nitrite-oxidising bacteria. The activities of both bacterial groups have to be well-balanced, otherwise, nitrite will accumulate and inhibit the nitrite-oxidising
alkalinity has to be added, for example by dosing 1 mol KHCO₃ in low pH value. For complete conversion of ammonia to nitrate, ammonia is oxidised to nitrate, the process stops due to the limited by the alkalinity in stored urine; after half of the losses occur anymore (Udert et al., 2003b). Nitrate production close to 6. Under such conditions, no considerable ammonia losses occur anymore (Udert et al., 2003b). Without preliminary stabilisation of urine by nitrification or acid dosage (Ek et al., 2006), high ammonia losses will occur during distillation of urine (Udert and Wächter, 2012).

Electrolysis of urine

High degradation rates per surface area and simple operation make electrolysis an interesting process for small reactors which could be directly integrated into a toilet. A complete treatment of urine at the household would cut the costs for transportation to a treatment plant. Possibly, only a small amount of by-products (e.g. electrochemically precipitated struvite) would have to be collected, while the remaining water could be infiltrated. Drawbacks of electrolysis are a high demand for electricity, high prices for some of the most efficient electrodes, and the production of chlorinated by-products (Anglada et al., 2009). Since ammonia is degraded and not recovered in most electrolysis processes, electrolysis could be applied at locations where preventing hygiene and environmental problems is much more important than nutrient recovery and reuse.

Frequently, electrolytic oxidation processes in wastewater treatment are based on indirect oxidation. In most cases of indirect oxidation, chlorine is oxidised anodically to active chlorine, which, in turn, oxidises compounds such as ammonia or organics. Active chlorine is also an efficient disinfectant, especially for viruses and bacteria (but to a lesser extent for protozoa; von Gunten, 2013). However, indirect oxidation is an unspecific process, which often produces considerable amounts of by-products, such as chlorinated organic substances, chloride and perchlorate. A more specific electrolytic process is direct oxidation. In this case, the target compound, in our case mostly ammonia, is oxidised directly at the anode. Ideally, the anode potential, at which direct oxidation occurs, is below the potentials necessary to produce significant amounts of active chlorine or oxygen (Zöllig et al., 2015).

RESULTS OF URINE TREATMENT IN VUNA

Struvite precipitation

Reactor setup

We investigated 2 reactor setups: the first (Grau et al., 2012) is similar to a manually operated reactor built in a previous study in Nepal (Etter et al., 2011), the second is a fully automated reactor equipped with a process control for magnesium dosage (Grau et al., 2013). Besides the level of automation, the 2 setups also use different filtration methods: the manual reactor is equipped with cotton filter bags, which are later dried in the air (for 7 to 14 d) and the automated version has a module with flat cotton filters, which can be used up to 7 times. For both setups, solutions of MgSO₄ or MgCl₂ were used as magnesium sources. When the magnesium solution was added manually, a magnesium to phosphate (Mg:P) ratio of 1:1:1 was used. The reactor volumes were 40 ℓ for the manual and 50 ℓ for the automated version. The total processing time for the manual reactor was 50 min, of which 45 min were needed for filtration and only 5 min to fill the reactor, mix its content, and let the precipitant react in the precipitation reactor.

Automated magnesium dosage

In most reactors for struvite precipitation from urine, magnesium is dosed at the beginning of a process cycle. In this approach, the phosphate concentration needs to be known exactly for exact magnesium dosage. In VUNA’s automated reactor, turbidity or electrical conductivity were measured online, while the magnesium solutions were dosed continuously (Grau et al., 2013). Based on the online measurement signals, the endpoint of struvite formation could be predicted, which allowed for a more exact magnesium dosage. Turbidity measurements were

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more sensitive than electrical conductivity measurements; while the turbidity signal doubled between the beginning and the end of magnesium dosage, the conductivity signal changed only by 1 to 2%, which is only slightly higher than typical measurement accuracies of 0.5%. However, the conductivity signals better reflected the course of precipitation and allowed a more exact determination of the endpoint of struvite formation. Intermittent instead of continuous dosage resulted in more informative signals but the algorithms for process control also became more complex. Nevertheless, the experiments showed that online sensors can be used to optimise magnesium dosage.

**Electro-precipitation of struvite**

Besides being dosed as a powder or a solution, magnesium can also be dissolved electrochemically. Hug and Udert (2013) used a magnesium anode and a steel cathode to dissolve metallic magnesium for electro-precipitation of struvite. In a long-term experiment at an anode potential of −0.6 V vs. NHE, the precipitation rate was 90 mg P·cm⁻²·d⁻¹. Electrochemical dissolution is more expensive than using MgO or waste products, but cost calculations show that it is in the same range or even less expensive than using MgSO₄ or MgCl₂. The energy demand is low (1.7 Wh per gram P precipitated), so that energy costs are negligible compared to the costs for the metallic magnesium. Combined with reliable process control, e.g. using turbidity measurements, electro-precipitation could be a promising approach for remote operation of on-site reactors.

**Fertiliser value**

In pot experiments with ryegrass, chemical fertiliser (KH₂PO₄) and struvite precipitated from synthetic stored urine performed equally (Bonvin, 2013). By using isotopic tracers, a phosphorus uptake of 26% of phosphorus applied with struvite was determined, which was similar to the 28% of phosphorus taken up from the water soluble reference fertiliser. These results confirm previous studies on the fertiliser quality of struvite (Johnston and Richards, 2003; Römer, 2006). These studies also showed that struvite is a considerably better fertiliser than iron or aluminium phosphate recovered from wastewater.

**Pathogens and trace organic contaminants**

The fate of pathogens during struvite precipitation has been investigated by Decrey et al. (2011) and Schoger (2011). Decrey et al. focused on the most resistant pathogen groups, i.e. viruses and helminth eggs, and used phage ΦX174 as indicator for human viruses and *Ascaris suum* as indicator for worm eggs. The study revealed that the majority of helminth eggs are retained in the struvite cake, which forms on the filter. The phage concentration in the struvite cake could be attributed to the phages in the pore water. Drying of struvite reduced the number of phages by more than 3 log units (drying at 36°C and 35% to 42% relative humidity). The inactivation of phages was directly correlated to the moisture content of the struvite. *Ascaris suum* eggs inactivated to a much lower degree. Even at the minimum moisture content, the deactivation was only 2 log units. Schoger (2011) measured total heterotrophic bacteria as an indicator of bacterial pathogens. He found that bacteria also accumulated in struvite but died off as a function of struvite moisture content. However, inactivation levelled off around 3 log loss.

**Nitrification and distillation**

**Reactor setup**

Within the VUNA project, 2 reactor setups have been operated, one in Eawag’s main building in Dübendorf/Switzerland (Fig. 2) and the other one in Durban, South Africa. The basic principle of both setups is the same: stored urine is pumped into a nitrification reactor, where Kaldnes® K1 biofilm carriers (60% of the reactor bulk volume) are used as substratum for the slow-growing nitrifying bacteria. Microbiological analyses with
polymerase chain reaction (authors' own unpublished data) have shown that the nitrifying bacteria grow mostly on the biofilm carriers, while heterotrophic bacteria also grow in suspension. The maximum nitrification rate was 420 mg N·m⁻³·d⁻¹ or 1.4 g N·m⁻²·d⁻¹. This is in the range of maximum nitrification rates reported by Rusten et al. (1995) for municipal wastewater treatment (1.4 to 1.6 g N·m⁻²·d⁻¹).

The distiller is a commercially available reactor produced by KMU-Loft Cleanwater (ProwaDest E20, KMU-Loft Cleanwater GmbH, Hausen, Germany) with a minimum throughput of 20 ℓ·h⁻¹. About 85% of the energy needed for evaporation is recovered by vapour compression and subsequent heat exchange. About 95–97% of the water in urine is removed and the remaining concentrate is still a liquid without precipitates. A solid product mainly consisting of ammonium nitrate can be recovered from this process by evaporating the remaining water. However, this final process step does not improve the product quality substantially and requires much more elaborate equipment. The solid ammonium nitrate product is also more sensitive to heat and can explode if heated. Wächter et al. (2013) showed that the solid ammonium nitrate product from urine should not be exposed to temperatures above 96°C for an extended period of time to prevent fast thermal decomposition. If all ammonia is oxidised to nitrate by adding a base to the influent urine, the sensitivity to high temperatures is no longer critical anymore within common temperature ranges for distillation and drying processes (Wächter, 2013). However, small amounts of ammonia make the product temperature sensitive. For example, the temperature limit for drying a 1:9 ammonium nitrate nitrified urine solution is 168°C, but this is still substantially above the common temperature for distillation and drying processes. For the liquid products, the temperature limits are not critical: for the 1:1 ammonium nitrate solution, the limit is 217°C. The temperature limit for the 1:9 ammonium nitrate solution was not determined, because the limit for the 1:1 ammonium nitrate solution is already far above the operating range of the distiller. In the current setup, the nitrified urine is heated to 80°C for several hours. These conditions are far from being critical for the thermal stability of ammonium nitrate, and they ensure disinfection. For comparison, pasteurisation of sewage sludge requires heating the sludge to 70°C for at least 30 min (US EPA, 2003).

**Nutrient recovery**

Nearly all nutrients are recovered in the final product except for a negligible amount of nutrients in the withdrawn excess sludge, and some ammonia, which volatisises during distillation (less than 3% of the total nitrogen in urine; Udert and Wächter, 2012). Since about 30% of the phosphate precipitates in the collection tanks and might not enter the nitrification reactor, the solids in the urine storage tanks should be collected as well in order to maximise nutrient recovery.

**Chemical requirements and energy demand**

The process combination of nitrification/distillation does not require any chemicals with the exception of a small amount of corrosion inhibitor for the distillation reactor. Adding a base will be necessary if all of the ammonia is supposed to be oxidised to nitrate. If the base KHCO₃ was used, 7.1 g KHCO₃ per gram of total ammonia would be needed. Based on experience with the reactor at Eawag, the energy demand is about 50 Wh·ℓ⁻¹ for partial nitrification and 100 Wh·ℓ⁻¹ for complete nitrification (for undiluted urine). Distillation requires about 80 Wh·ℓ⁻¹. Additionally, pumps and process control will require some energy, so that our current estimate for the total energy demand is about 140 Wh·ℓ⁻¹.

**Process stability**

Since nitrification is a biological process, it is more sensitive to process disturbances than a chemical process. The most critical problem is nitrite accumulation (Udert and Wächter, 2012; Etter et al., 2013). Small changes in the inflow rate or in the reactor pH can destabilise the sensitive balance between ammonium and nitrite-oxidising bacteria, so that the intermediate nitrite can accumulate. Nitrite concentrations above approximately 50 mg N·ℓ⁻¹ will require immediate action to prevent further accumulation, and inhibition of the nitriifying bacteria. Simulations have shown that an inflow increase of only 10% can lead to serious nitrite accumulation. (Etter et al., 2013). Nitrite detection will be a key feature of stable urine nitrification reactors.

**Fertiliser value**

Bonvin (2013) tested the availability of nitrogen from synthetic solutions representing nitrified urine in pot experiments with ryegrass (Bonvin, 2013). The plants took up about 75% of the nitrogen in nitrified urine, compared to 77% in the case of the chemical fertiliser (NH₄NO₃). The advantage of urine-derived fertilisers is the content of other nutrients, which are often lacking in synthetic fertilisers. Therefore, it might be particularly well suited for regions where fertilisers are scarce and knowledge about optimal fertiliser application is low.

**Pathogens**

Nitrified urine contains some compounds of concern for fertilisers including pathogens, pharmaceutical residues and sodium chloride. Bischel et al. (2015) used bacteriophages to investigate the inactivation of viruses in laboratory-scale reactors for urine nitrification. The bacteriophage Qbeta was inactivated by aeration, while 2 other bacteriophages (ΦX174 and MS2) were hardly affected. However, the number of pathogenic bacteria (indicator organisms: *Salmonella typhimurium*, *Enterococcus* spp.) was reduced by about 1 log unit due to microbial activity. Worm eggs are presumably not inactivated during nitrification, but could partially be removed by sedimentation in the settling tank or by filtration if a membrane reactor was used for nitrification. While nitrification might not be a safe barrier against pathogens, distillation ensures that all pathogens will be killed, so that the concentrated product will be hygienically safe.

**Trace organic contaminants**

To date, we have not investigated the degradation of pharmaceutical residues during distillation, but it is unlikely that a strong reduction will occur, because they are mostly stable at temperatures up to 80°C. However, some pharmaceutical residues are degraded during nitrification. In a laboratory study with a nitrification reactor, we investigated the degradation of several pharmaceuticals (antibiotics, antivirals, beta blockers, antibiotics, anti-inflammatory agents, etc.) to determine if any residual concentrations could be a threat to the environment. While some pharmaceuticals were partially degraded in the nitrification reactor, others were not affected. The results indicate that distillation is a suitable method to remove trace organic contaminants from urine.
analgesics, and diuretics), which were dosed to stored urine. The pharmaceuticals were chosen based on measurements in large urine collection tanks in eThekwini. Several antiviral and antibiotic compounds and the beta blocker Atenolol had half-life times of 9 ± 3 d (Özel, 2013), which was shorter than the hydraulic retention time in the lab-scale reactor. However, some compounds were still detectable in considerable amounts in the effluent. To minimise the pharmaceutical load in the final product, the effluent of the nitrification reactor could be treated with activated carbon. Experiments with different dosages of activated carbon revealed that a near complete removal of the investigated pharmaceuticals is possible (Özel, 2013). In addition to removing the pharmaceuticals, the brown colour of the effluent is degraded. In a practical setting, a granular activated carbon filtration could be easily established.

**Sodium chloride removal**

Sodium chloride is not desirable in fertilisers due to the possible salinisation of agricultural soils. It should be noted that the ratio of nutrients to sodium and chloride in urine is not higher than in manure, which is commonly used as fertiliser (see e.g. Nunez-Delgado et al., 2002). Nevertheless, removing sodium chloride would further increase the quality of the urine-derived fertiliser. Laboratory experiments have revealed that up to 50% of the sodium chloride could be removed by sequential distillation at well-controlled and set temperatures (Huber, 2011). This might be an option for producing a fertiliser for agricultural soils with low tolerance for sodium chloride. However, for many soils the impact of sodium chloride from urine-derived fertiliser is probably too low to justify the higher complexity of a distillation process with sequential salt removal.

**Electrolysis**

**Direct oxidation on graphite electrodes**

Graphite is a cheap electrode material, which has been used traditionally for chlorine production (Hamann and Vielstich, 2007). While it is inexpensive, it is not stable at anode potentials in the chlorine evolution range and cannot be used for indirect oxidation of urine in the long term. However, below this range (below an anode potential of 1.6 V vs. SHE), graphite can be used for direct oxidation of compounds such as ammonia (Zöllig et al., 2015). Our experiments (Zöllig et al., 2015) on direct ammonia oxidation on graphite electrodes showed that this process requires high pH values, preferably above the $pK_a$ value for ammonia (9.5 at 25°C and an ionic strength of 0.5 mol·l$^{-1}$) (Stumm and Morgan, 1996), because free ammonia ($\text{NH}_3$) is the actual reactant on the electrode surface. In stored urine, the pH is slightly below this value, which might explain why the degradation in the batch experiments was only 5.2 g N·m$^{-2}$·d$^{-1}$ (at 1.31 V vs. SHE). In comparison, indirect oxidation of ammonia on graphite proceeded at a rate of 150 g N·m$^{-2}$·d$^{-1}$ (at 1.94 V vs. SHE) and the biological nitrification rate in a moving bed biofilm reactor can be 1.6 g N·m$^{-2}$·d$^{-1}$, which is only slightly lower. The energy demand per mass of N was 44 Wh·g$^{-1}$ and 87 Wh·g$^{-1}$ for direct and indirect oxidation respectively, and therefore significantly higher than the energy demand for nitrification (about 6 Wh·g$^{-1}$). Direct ammonia oxidation with graphite is a suitable process, if the pH is kept high, large electrode surface areas are provided and the anode potential can be kept constant. All these requirements are hard to fulfil in small on-site reactors. However, a major advantage of direct oxidation on graphite is that the formation of chlorinated by-products can be prevented.

**Indirect electro-oxidation**

In our experiments (authors’ own unpublished data), we used 2 common anodes for indirect electro-oxidation, a noble metal oxide electrode TDIROF (thermally decomposed iridium oxide film) and BDD (boron-doped diamond). With indirect electro-oxidation it is not necessary to adjust a certain anode potential, and the simpler and more common operational mode of applying a constant current density was chosen. Both electrodes can also be used for direct ammonia oxidation (Kapalka et al., 2010; Kapalka et al., 2011), but in these experiments, indirect oxidation was dominant due to the high anode potentials that resulted from the high current densities we applied. Preliminary results showed that depending on the current density, very high removal rates can be achieved. At the highest current density (20 mA·cm$^{-2}$), the maximum removal rates for organic substances were higher than 500 g COD·m$^{-2}$·d$^{-1}$ for TDIROF and 1 000 g COD·m$^{-2}$·d$^{-1}$ for BDD. The maximum ammonia removal rates were above 400 g N·m$^{-2}$·d$^{-1}$ and 200 g N·m$^{-2}$·d$^{-1}$ for TDIROF and BDD, respectively. Unfortunately, the excellent removal rates came with a price: the energy consumption was high and considerable amounts of chlorinated by-products were produced. Even at ideal conditions (high ammonia concentrations and new electrodes) the energy consumption per mass of N removed was above 40 Wh·g$^{-1}$ and 200 Wh·g$^{-1}$ for TDIROF.

![Image](http://dx.doi.org/10.4314/wsa.v41i2.06)

**Figure 3**

Laboratory setup at Eawag to determine the production of chlorinated by-products during urine electrolysis

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and BDD, respectively. There were two reasons for the high energy consumption with BDD. Firstly, the current efficiency for ammonia oxidation with BDD was only 20% compared to 50% for TDIROF, which could be explained by a higher preference of BDD for oxidising organic substances. Secondly, BDD required a higher cell voltage, 6 V compared to about 4 V with TDIROF at 20 mA·cm⁻²).

**By-product formation**

Whereas no chlorinated organic or inorganic by-products were found for direct oxidation on graphite, considerable amounts were detected in the experiments on indirect oxidation with TDIROF and BDD (authors’ own unpublished data). The chlorinated organic by-products were extracted from the urine and were captured in the off-gas to determine the amount of volatileised compounds. Six compounds were quantified with coupled gas chromatography/mass spectrometry (GC/MS): 1,2 dichloroethane, 1,2 dichloropropane, chlorobenzene, dichloromethane, trichloromethane, tetrachloromethane. Our preliminary measurements showed that about 50–100% of the chlorinated organic by-products volatilised. While BDD was more prone to the production of chlorinated organic by-products, chloride formation was doubled on TDIROF compared to BDD. Our results suggest that in practical settings for urine electrolysis, chlorinated by-products have to be captured in the off-gas as well as in the effluent to prevent health and environmental hazards.

**SUMMARY**

Table 3 provides an overview of criteria which are relevant for the choice of urine treatment technologies. Electrolysis, especially indirect oxidation, is a promising process with respect to sanitisation and removal of nitrogen and organic compounds. However, the process does not allow the recovery of most nutrients and might even pose a health risk due to the production of chlorinated by-products. Further research and development and a combination with other processes will be necessary to make electrolysis a suitable process for on-site urine treatment.

Struvite precipitation and nitrification/distillation have been tested at pilot scale and are close to commercialisation. Both processes target different markets. Struvite precipitation does not require complex reactor setups and can be operated by non-skilled workers. It is an efficient process for phosphorus recovery, but most nutrients end up in the effluent. Furthermore, sanitisation of struvite is incomplete without long drying phases. A combination with other processes, for example nitrification/distillation to treat the effluent, or heating of struvite for pathogen deactivation (and struvite transformation to MgHPO₄) is advisable.

Nitrification/distillation fulfils all requirements with respect to nutrient recovery, environmental protection and sanitisation, but the process is more complex and technically challenging than struvite precipitation. Water removal can even become dangerous, if the temperatures are high (above 96°C) and solid ammonium nitrate is produced. This problem could be solved by reliable automation, or by using other water

| TABLE 3 | Comparison of struvite precipitation, nitrification/distillation and electrolysis according to the research results in VUNA |
|-----------------------------|-------------------------------------------------|-------------------------------------------------|
| **Struvite precipitation** | **Nitrification/Distillation** | **Electrolysis** |
| Main product | Phosphate mineral (struvite) | Concentrated, complete nutrient solution | Effluent without ammonia, organic substances and pathogens |
| Nutrient recovery | Medium, mainly phosphate | High, nearly all nutrients | None, if not combined with other processes |
| Nutrient loss | High, most nutrients in the effluent | Low, little ammonia volatilisation | High ammonia is oxidised to nitrogen |
| Sanitation | Medium, depends on drying procedure | High, complete disinfection during distillation | Medium chlorination |
| Malodour removal | No | Yes | Yes |
| Removal of trace organic compounds | No | Partial degradation during nitrification | Unknown |
| Energy demand(1) | Very low, especially for manual reactor | Medium approx. 50 Wh ℓ⁻¹ partial nitrification approx. 80 Wh ℓ⁻¹ distillation | High, approx. 320 Wh ℓ⁻¹ (TDIROF) approx. 1 600 Wh ℓ⁻¹ (BDD) |
| Volume reduction of main product | High, phosphorus in struvite 250 to 630 times concentrated(2) | High, 20 to 30 times concentrated | No volume reduction |
| Other outputs | Phosphate depleted urine | Distilled water, excess sludge | Off-gas with chlorinated by-products |
| Process complexity | Low (manual reactor) | High | Low |
| Development level | High | High | Low |

(1) Calculated for a total ammonia concentration of 8 g N ℓ⁻¹. The values for electrolysis are based on the experimental values given in the text. These values are most probably too high, because they are based on small-scale laboratory experiments, which have not been optimised for low energy demand.
(2) Assuming phosphate concentrations between 200 and 500 mg P ℓ⁻¹ in stored urine.
removal processes which do not require high temperatures, for example humidification or reverse osmosis.

Removal of trace organic compounds, specifically pharmaceutical residues, might be necessary to license any fertiliser product from urine. However, it is unclear whether these compounds actually pose a major risk to the environment and human health when the urine fertilisers are applied to agricultural soils. Based on our current state of knowledge, removal of pathogens is much more important for human health.

High energy demand can be prohibitive for nitrification/distillation (and electrolysis) if the reactors are supposed to be operated in remote locations, where no centralised or decentralised energy supply (e.g. solar cells) is available. However, the energy demand of all processes is so low, that it should not pose a serious problem in cities, where at least intermediate volumes of electricity supply can be guaranteed.

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