Diagnostic and Prognostic Significance of Dysregulated Expression of Circular RNAs in Osteosarcoma

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Abstract

Background

CircRNAs have emerged as pivotal regulators in osteosarcoma tumorigenesis and progression, but their prognostic and diagnostic significance remain unclear. Herein, we aimed to perform an updated meta-analysis to explore the clinical, diagnostic and prognostic values of circRNAs in osteosarcoma.

Methods

Several databases, including PubMed, Web of Science, EMBASE, Scopus and Cochrane Library, were systematically searched up to April 10, 2020. Eligible studies regarding the relationship between circRNAs levels and clinicopathological, diagnostic and prognostic values in osteosarcoma patients were included in this study. Pooled odds ratios with corresponding 95% confidence intervals were used to measure clinical characteristics, while hazard ratios with 95% CIs were adopted to assess overall survival (OS) and disease-free survival (DFS).

Results

Overall, 26 relevant studies involving 1,652 patients with osteosarcoma were enrolled, with eighteen studies on clinicopathological parameters, ten on diagnosis and eighteen on prognosis. For clinical parameters, overexpression of oncogenic circRNAs was intimately correlated with larger tumor size ($P < 0.00001$), advanced Enneking stage ($P < 0.00001$), poor differentiation ($P = 0.0001$), and distant metastasis (DM) ($P < 0.00001$). In contrast, the downregulated circRNAs showed negative correlation with Enneking stage ($P = 0.002$) and DM ($P < 0.0001$). For the diagnostic values, the summary area under the curve of circRNA for the discriminative efficacy between osteosarcoma patients and non-cancer counterparts was estimated to be 0.86 (95% CI: 0.83–0.89), with a weighted sensitivity of 0.80 (95% CI: 0.74–0.84), specificity of 0.80 (95%: 0.75–0.84), and diagnostic odds ratio of 15.48 (10.85–22.10), respectively. For the prognostic significance, oncogenic circRNAs had poor OS (HR = 1.92, 95% CI: 1.68–2.19) and DFS (HR = 2.65, 95% CI: 2.02–3.49), while elevated expression of tumor-suppressor circRNAs were closely related to longer OS (HR = 0.44, 95% CI: 0.28–0.69).

Conclusions

Taken together, our study showed that aberrantly expressed circRNA signatures could serve as potential biomarkers in diagnosis and prognosis in patients with osteosarcoma.

Background
Osteosarcoma is a common primary bone malignancy that mainly affected children and adolescents [1]. In spite of substantial achievement in diagnosis and treatment, the 5-year survival of osteosarcoma remains unsatisfied [2], largely due to the insufficiency of accurate predictive biomarkers [3]. Therefore, it is urgent to identify more ideal biomarkers with both diagnostic and prognostic value in osteosarcoma.

Recent advances in transcriptomics have provided novel perspective for management of cancer [4]. Circular RNAs (circRNAs) are a cluster of endogenous non-coding RNAs (ncRNAs) with no or limited capacity for coding proteins [5–8]. It is formed from back-splicing of pre-RNAs without 3’-terminal poly A tail and 5’-terminal cap [9]. Accordingly, it is more stable than the linear mRNA due to the covalently continuous loop [10]. Mechanistically, circRNAs could function as nuclear transcription regulator, attaching elements of RNA-binding protein (RBP), or competing endogenous RNA (ceRNA) to sponge microRNA (miRNA), thus in turn to play a critical role in various biological process of human diseases including cancer [11]. Interestingly, several circRNAs have been found to encode target genes, and thereby serve as tumor promoter or suppressor in cancer progression [7, 8].

Nowadays, circRNAs have attracted the researches attention as potential biomarkers and therapeutic targets in carcinomas, which are attributed to their abundance in tissues, structural stability and tissue-specific expression profile [9]. More recently, emerging studies have highlighted the role of circRNAs in osteosarcoma [12–19]. CircRNAs are reported to modulate osteosarcoma malignant properties including cell proliferation, apoptosis, migration and resistance to chemotherapy [12, 20, 21]. CircRNAs could interact with cancer-related signaling pathway, such as Akt/GSK-3β, and wnt/β-catenin pathway, or act as a ceRNA to sponge miRNA and consequently regulate target genes in osteosarcoma progression [12, 22, 23]. Moreover, a growing number of evidences showed that dysregulated circRNAs are closely correlated with survival outcome and clinicopathological parameters including clinical stage, chemo-sensitivity, and metastasis [13, 23, 24]. However, the clinical prognostic and diagnostic significance of circRNAs may be jeopardized due to the controversial results as well as limited sample size in previous studies. The aim of this meta-analysis was to further clarify the pooled clinicopathological, diagnostic and prognostic significance of circRNAs in osteosarcoma.

**Methods**

**Search strategy**

This study was performed in accordance with the Preferred Reporting Items for Systematic Review and Meta-analysis (PRISMA) Checklist [25]. Several electronic databases, including PubMed, Web of Science, EMBASE, and the Cochrane Library database, were extensively searched from inception to April 10, 2020 for eligible studies that assessed the clinicopathological, diagnostic or prognostic significance of circRNAs in osteosarcoma. The following terms were used in databases for literature retrieval: (“circular RNA” or “circRNA” or “hsa_circ”) and (osteosarcoma). Patients with osteosarcoma were considered as the “case group”, while those with benign lesions or healthy were taken as “control group”.

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Study Selection Criteria And Data Extraction

Studies were eligible if met the following criteria: 1) cohort or case-control study; 2) diagnosis of osteosarcoma was histo-pathologically confirmed; and 3) correlation between circRNAs expression with clinicopathological features, diagnostic accuracy or prognosis utility were provided or extractable;

Besides, those studies were excluded if 1) not relevant with circRNAs or osteosarcoma; 2) in vivo studies, case reports, reviews or letters without original data; 3) duplicate studies;

Two independent investigators (CHZ and JYH) evaluated the enrolled studies and extracted data carefully, and a third researcher (LQ) would be consulted to reach a consensus if disagreement occurred. The following data were extracted in eligible studies: first author, publication year, circRNA type, sample type, sample size, detection assay, reference gene, regulation pattern, area under curve (AUC), circRNA expression levels, survival outcomes including overall survival (OS), disease-free survival (DFS) or progression-free survival (PFS), survival analysis method, follow-up duration (months), age, gender, tumor size, Enneking stage, differentiation based on WHO grade, and distant metastasis (DM).

Quality Assessment, Sensitivity Analysis And Publication Bias

The quality of included studies on diagnosis was assessed according to the Quality Assessment for Studies of Diagnostic Accuracy II (QUADAS II) checklist [26], while studies on prognosis were rated by the Newcastle-Ottawa Scale (NOS) as previously described [27, 28]. The studies were considered of high quality if the QUADAS II score was ≥ 4, or NOS score was ≥ 6.

In order to increase the credibility of this study, sensitivity analysis was performed to identify the potential source of heterogeneity. Publication bias was investigated by using both funnels plots and Begg’s as well as Egger’s test.

Statistical analysis

Statistical analyses were performed by using STATA software (13.0) and Meta-Disc (1.4). Pooled odds ratios (ORs) with 95% confidence intervals (CIs) were adopted for evaluation of clinical features, while hazard ratios (HRs) with 95% CIs to measure OS and DFS. The Chi-square test was utilized to assess the heterogeneity among studies. If \( P < 0.05 \) or \( I^2 > 50\% \), the random-effect model would be used due to the significant heterogeneity. Otherwise, the fixed-effect model would be adopted in the analyses. In addition, the publication bias was conducted by Deeks’ funnel plot asymmetry test, Begg’s and Egger’s test [29].

Results

Characteristics of the enrolled studies
The search procedure was presented in Fig. 1. Among the potential literature retrieved from databases, 170 studies were initially assessed. After removing 90 duplicate publications, the remaining 80 studies were evaluated for titles and abstracts. Of these, 32 irrelevant articles were further excluded after abstract review and only 48 studies remained for full-text verification. Moreover, 22 studies were further eliminated with a variety of reasons, including five studies not related to circRNAs or osteosarcoma, six studies did not report outcomes, two reviews and case reports, four animal studies, and five lack of extractable data. Finally, 26 studies comprising 1,652 osteosarcoma cases were enrolled in this quantitative study, with 18 on clinicopathological features, ten on diagnosis [13, 18, 19, 21, 24, 30–34] and 18 on prognosis [12–14, 16–20, 22–24, 30–32, 34–42]. Among the 1,652 eligible osteosarcoma cases, patients were separated by age, gender, or other clinicopathologic features. As for age, the included studies had divergent cut-off values, with six studies on upregulated circRNAs using ≥ 25 or < 25 years old [12, 14, 18, 19, 32–34, 36–40], whereas other two studies on downregulated circRNAs applying ≥ 18 or < 18 years old to separate enrolled patients [13, 30]. In consideration of the consistency, only these studies with common cut-off value were included in the following meta-analysis with regard to patients’ age. Besides, all cases in studies on clinicopathological feature analysis were divided into male and female. Other clinicopathologic parameters consist of tumor size (≥ 5 cm/<5 cm), Enneking stage (IIB-III/I-IIA), distant metastasis (positive/negative), and WHO grade (III/I-II) were also presented in this analysis.

The baseline characteristic features of enrolled studies were shown in detail in Table 1 and Table 2. All studies were conducted in China, and the publication year ranged from 2017–2020. All osteosarcoma cases were reliably diagnosed based on histopathology. The sample size was varied from 30 to 170. CircRNAs expression was measured by quantitative real-time polymerase chain reaction (qRT-PCR) and the reference gene was GAPDH. CircRNAs were regarded as oncogenes or tumor-suppressor in terms of their expression levels. Specifically, 21 circRNAs were identified as oncogenic circRNAs with upregulated expression pattern [12, 14, 16–23, 31–34, 36, 37, 39–43], while the other five were recognized as tumor suppressors [13, 24, 30, 35, 38] with downregulated expression, respectively. The duration for follow-up ranged from 40 to 125. For diagnostic assay, ten studies with data on sensitivity, specificity and AUC [13, 18, 19, 21, 24, 30–34].
Table 1
Main characteristics of studies for diagnosis analysis in osteosarcoma

| Study          | Publication year | CircRNA     | Sample type | Sample size | Method      | Reference gene | Regulation pattern | AUC   | Ref. |
|----------------|------------------|-------------|-------------|-------------|-------------|-----------------|--------------------|-------|------|
| Li, S et al    | 2020             | circ_000190| serum       | 60          | qRT-PCR     | GAPDH           | downregulated      | 0.889 | [30] |
| Hu, Y et al    | 2019             | circ-LARP4 | tissue      | 72          | qRT-PCR     | GAPDH           | downregulated      | 0.829 | [13] |
| Zhen, S et al  | 2019             | circLRP6   | tissue      | 50          | qRT-PCR     | GAPDH           | upregulated        | 0.874 | [31] |
| Zhu, K et al   | 2019             | circ_000885| serum       | 30          | qRT-PCR     | GAPDH           | downregulated      | 0.783 | [32] |
| Ma, X et al    | 2018             | circ_HIPK3 | serum       | 50          | qRT-PCR     | GAPDH           | upregulated        | 0.783 | [24] |
| Xu, B et al    | 2018             | CDR1as     | tissue      | 38          | qRT-PCR     | GAPDH           | upregulated        | 0.857 | [33] |
| Zhou, X et al  | 2018             | circ_0008717| tissue      | 45          | qRT-PCR     | GAPDH           | upregulated        | 0.782 | [34] |
| Zhu, K et al (a)| 2018             | circ_PVT1  | serum       | 50          | qRT-PCR     | GAPDH           | upregulated        | 0.871 | [19] |
| Zhu, K et al (b)| 2018             | circ_0081001| serum       | 50          | qRT-PCR     | GAPDH           | upregulated        | 0.898 | [18] |
| Liu, X et al   | 2017             | circ-NT5C2 | tissue      | 52          | qRT-PCR     | GAPDH           | upregulated        | 0.753 | [21] |

Abbreviations: AUC, area under curve; CircRNA, circular RNA; qRT-PCR, quantitative real time polymerase chain reaction; Ref, reference
### Table 2
Main characteristics of the meta-analysis for prognosis association of circRNAs in osteosarcoma

| Study | Publication Year | CircRNAs | CircRNA expression | Sample type | Detection assay | Regulation pattern | Survival indicators | Survival analysis | Follow-up months | Ref. |
|-------|-----------------|----------|--------------------|-------------|----------------|--------------------|--------------------|------------------|------------------|------|
| Wang, L et al | 2020 | circ_0001 658 | 21 | High | tissue | qRT-PCR | upregulated | N/A | N/A | [36] |
| Li, S et al | 2020 | circ_0000 190 | 30 | High | tissue | qRT-PCR | downregulated | N/A | N/A | [30] |
| Wang, Y et al | 2020 | circTCF25 | 26 | High | tissue | qRT-PCR | upregulated | N/A | N/A | [37] |
| Ji, X et al | 2020 | circ_0016 21 | 20 | High | tissue | qRT-PCR | upregulated | OS | Univariate | 60 | [12] |
| Zhu, K et al | 2019 | circ_0000 885 | 25 | High | tissue | qRT-PCR | upregulated | OS/DFS | Multivariate | 60 | [32] |
| Zeng, S et al | 2019 | circL RP6 | N/A | High | tissue | qRT-PCR | upregulated | OS/DFS | Multivariate | 125 | [31] |
| Wang, L et al | 2019 | circ_0021 347 | 35 | High | tissue | qRT-PCR | downregulated | OS | Univariate | 40 | [38] |
| Qi, H et al | 2019 | circ_0000 502 | 34 | High | tissue | qRT-PCR | upregulated | OS | Multivariate | 60 | [43] |
| Jin, J et al | 2019 | circ_1008 76 | 24 | High | tissue | qRT-PCR | upregulated | OS | Univariate | 60 | [16] |
| Pan, G et al | 2019 | circ MM P9 | 24 | High | tissue | qRT-PCR | upregulated | OS | Univariate | 60 | [39] |
| Li, L et al | 2019 | circ_0001 721 | 28 | High | tissue | qRT-PCR | upregulated | OS | Multivariate | 60 | [20] |
| Jin, Y et al | 2019 | circ_0102 049 | 38 | High | tissue | qRT-PCR | upregulated | OS | Multivariate | 60 | [17] |
| Study                  | Publication Year | CircRNA expression | Sample type | Detection assay | Regulation pattern | Survival indicators | Survival analysis | Follow-up months | Ref. |
|-----------------------|------------------|--------------------|-------------|-----------------|-------------------|-------------------|-----------------|------------------|------|
| Hu, Y et al           | 2019             | circ-LAR P4        | High: 36    | Low: 36         | qRT-PCR           | downregulated     | OS/DFS          | Univariat e      | 48   | [13] |
| Ma, X et al           | 2018             | circ_HIPK 3        | High: 37    | Low: 45         | qRT-PCR           | downregulated     | OS              | Univariat e      | 60   | [24] |
| Zhu, K et al          | 2018             | circVT1            | High: 30    | Low: 50         | qRT-PCR           | upregulated       | OS              | Univariat e      | 60   | [19] |
| Zhu, K et al          | 2018             | circ_0081001       | High: 27    | Low: 55         | qRT-PCR           | upregulated       | OS              | Univariate       | 60   | [18] |
| Zhu, K et al          | 2018             | circ_0004674       | High: 23    | Low: 37         | qRT-PCR           | upregulated       | OS              | Univariate       | 60   | [40] |
| Zhu, X et al          | 2018             | circ_0008717       | N/A         | N/A             | qRT-PCR           | upregulated       | OS/PFS          | Multivariate     | 80   | [34] |
| Huang, L et al        | 2018             | circNASP           | High: 19    | Low: 20         | qRT-PCR           | upregulated       | N/A             | N/A              | N/A  | [14] |
| Nie, W et al          | 2018             | circ_NT5 C2        | High: 86    | Low: 84         | qRT-PCR           | upregulated       | OS/DFS          | Multivariate     | 60   | [41] |
| Li, B et al           | 2018             | circ_0007534       | High: 31    | Low: 26         | qRT-PCR           | upregulated       | OS              | Multivariate     | 60   | [23] |
| Wu, Z et al           | 2018             | circ_0002052       | High: 54    | Low: 54         | qRT-PCR           | downregulated     | OS/PFS          | Univariat e      | 60   | [35] |
| Zhang, H et al        | 2017             | circUBAP2          | High: 42    | Low: 50         | qRT-PCR           | upregulated       | OS              | Univariat e      | 60   | [42] |
The quality of studies was evaluated by QUADAS II and NOS scores. For diagnostic studies, the rating scores of QUADAS II ranged from 4 to 6. While for prognostic studies, the NOS scores were from 6 to 7, suggesting high methodological quality in all selected studies.

Expression Of Circrna With Clinicopathological Parameters In Osteosarcoma

The correlation between circRNAs and clinicopathological parameters in patients with osteosarcoma were demonstrated in Table 3. Overexpression of oncogenic circRNAs were significantly correlated with poor clinical features (tumor size: OR = 4.27, 95% CI: 2.25–8.10; Enneking stage: OR = 5.52, 95% CI: 2.79–10.94; differentiation: OR = 3.06, 95% CI: 1.72–5.45; DM: OR = 4.55, 95% CI: 2.55–8.12). In contrast, upregulated expression of tumor-suppressor circRNAs were remarkably associated with improved clinicopathological characteristics (Enneking stage: OR = 0.33, 95% CI: 0.16–0.68; DM: OR = 0.18, 95% CI: 0.08–0.40). Besides, no obvious difference was noted in terms of age and gender.
Table 3
Clinicopathological characteristics of circRNAs in osteosarcoma

| Clinicopathological parameters | Upregulated circRNAs | Downregulated circRNAs |
|-------------------------------|----------------------|------------------------|
| Age                           | 1.02                 | 0.94                   |
| Gender (male/female)          | 1.24                 | 1.01                   |
| Tumor size (≥5 cm/<5 cm)      | 4.27                 | N/A                    |
| Enneking stage (IIB-III/I-IIA) | 5.52                 | 0.33                   |
| WHO grade (III/I-II)          | 3.06                 | N/A                    |
| Distant metastasis (P/N)      | 4.55                 | 0.18                   |

**Abbreviations:** CI, confidence interval; N, negative; N/A, not available; OR, odds ratio; P, positive

**Diagnosis Analysis**

As demonstrated in Fig. 2, the forest plots showed that the pooled diagnostic value of circRNA in separating osteosarcoma from controls were as follows: sensitivity (SENS) of 0.80 (95% CI: 0.74–0.84), specificity (SPEC) of 0.80 (95% CI: 0.75–0.84), positive likelihood ratio (PLR) of 3.95 (95% CI: 3.19–4.89), negative likelihood ratio (NLR) of 0.26 (95% CI: 0.20–0.33) and combined diagnostic odds ratio (DOR) of 15.48 (95% CI: 10.85–22.10) (Fig. 2A, 2B, and 2C). Moreover, a summary receiver operator characteristic (SROC) curve was presented in Fig. 2D, and the AUC was 0.86 (95% CI: 0.83–0.89). These results indicated that circRNAs could be ideal diagnostic biomarker for osteosarcoma.
Additionally, a small set of oncogenic circRNAs with different diagnosis efficiency were synthesized in order to explore the combination panel of certain circRNAs with even higher diagnostic potential, which may facilitate the clinical application of the circRNAs for diagnosis. Among them, the pooled analysis containing circPVT1, circ_0081001, and CDR1as, has the highest diagnostic efficiency [18, 19, 33]. As shown in Figure S1, the forest plot of the pooled DOR, SENS and SPEC of these three circRNAs panel were 32.21 (95%: 9.51–109.1), 0.85 (95% CI: 0.78–0.90), and 0.83 (95% CI: 0.71–0.91), respectively. Notably, the AUC under SROC curve was 0.9116, indicating the combination of these circRNAs may have good diagnostic performance as the full panel of included circRNAs in osteosarcoma. However, more caution should be taken in the interpretation of this result since the comparatively small study numbers and limited sample size may produce outlying outcome and thereby introduce potential bias.

**Expression Of Circrnas With Prognosis In Osteosarcoma**

Survival analysis showed that overexpression of oncogenic circRNAs was significantly correlated with worse OS (HR = 1.92, 95% CI: 1.68–2.19), and DFS (HR = 2.65, 95% CI: 2.02–3.49) as shown in Fig. 3A and 3B, respectively. We identified one outlier study performed by Zheng S et al. in the combined effect of oncogenic circRNAs by sensitivity analysis. Besides, elevated expression of tumor-suppressor circRNAs predicted favorable OS (HR = 0.44, 95% CI: 0.28–0.69), as depicted in Fig. 3C. The fixed-effect model was applied in these studies since no obvious heterogeneity was noted.

**Sensitivity Analysis And Publication Bias**

The sensitivity analysis was performed in the prognostic effect sizes by omitting the enrolled studies one by one. For pooled effects of upregulated circRNAs in osteosarcoma, one study conducted by Zheng S et al. [31] was identified as the outlier (Fig. 4A). Notably, the predictive significance of OS for upregulated circRNA did not alter after excluding the aforementioned outlier data (HR = 2.51, 95% CI: 2.08–3.02). No outliers were found in other pooled effects (Fig. 4B, 4C).

The Deeks’ funnel plot test (P = 0.27) demonstrated the absence of publication bias existed in diagnostic analysis (Fig. 5A). For prognosis analysis of upregulated circRNA on DFS and downregulated circRNA on OS, the Begg’s funnel plot also showed symmetry and revealed no evidence of publication bias among the eligible studies (Fig. 5C, 5D). Nevertheless, the funnel plot of upregulated circRNA profile on OS showed significant asymmetry, indicating a possible publication bias (Fig. 5B). Therefore, a “trim and fill” method was applied to trace the possible impacts from bias as previously described. However, the predictive value of upregulated circRNAs on OS was not altered after adjustment, suggesting a closely correlation between upregulated circRNAs and poor OS among osteosarcoma patients (Fig. 5C).

**Discussion**
Emerging studies have established an important role of circRNAs in cancer initiation and progression. In recent years, circRNAs have been recommended as novel diagnostic and prognostic biomarkers in several cancers [44], including colorectal cancer [29], hepatocellular cancer [45, 46], gastric cancer [47], esophageal cancer [48], and lung cancer [9, 49]. Our study implicated a marked correlation between abnormal circRNAs expression levels with clinical, diagnostic and prognostic significance in osteosarcoma. In particular, oncogenic circRNAs with higher expression pattern were strikingly associated with unfavorable clinical features, including larger tumor size, advanced clinical stage, poor differentiation and DM, while the tumor-suppressive circRNAs showed an opposite correlation. For diagnosis roles, our study suggested the AUC of 0.86, with 80% sensitivity and 80% specificity of circRNAs in osteosarcoma. Besides, for the prognostic value, abnormal expression of circRNAs was remarkably related with OS as well as DFS. Furthermore, there were two studies reported the correlation between circRNA levels with PFS. Specifically, one demonstrated that highly expressed circ_0008717 was associated with worse PDS in osteosarcoma patients [34], while another study conducted by Wu Z et al. [35] showed that downregulated circ_0002052 predicted unfavorable PFS. Given the discrepant expression pattern of these two circRNA, it was unable to perform a pooled analysis concerning the prognostic significance of circRNA on PFS. Therefore, more studies describing the relationship between circRNAs and PFS in osteosarcoma are still needed.

Previously, Huang X et al. performed a meta-analysis on predictive values of circRNAs in osteosarcoma in 2019 [15]. However, only 13 studies were enrolled in their study, with 9 about clinical features, 11 on prognosis and 5 about diagnosis. Therefore, the pooled resulted may be underpowered due to insufficient data. Since the studies on circRNAs in osteosarcoma are emerging, we updated the predictive value of circRNAs in osteosarcoma in the study, with 26 studies comprising 1,652 patients, which markedly increased the statistical power and made the pooled results more credible. Besides, Huang X and colleagues did not report the association between upregulated circRNA with DFS of osteosarcoma due to limited eligible studies, whereas our study report for the first time that oncogenic circRNA was significantly correlated with poor DFS in addition to OS.

The diagnostic value of circRNAs as potential biomarkers for osteosarcoma was extensively explored in our study. Aberrant expression of circRNAs was shown in a wide range of sample sources from osteosarcoma cell lines or patients, including tumor tissue and plasma. Given the fact that circRNA is widely expressed in human samples and stable in structure, it may be ideal biomarker candidate with diagnostic significance in osteosarcoma [50]. The SROC in our study showed that the pooled AUC of circRNA in osteosarcoma was 0.86, indicating that 86% of randomly chosen osteosarcoma patients would have abnormal expression levels of circRNAs when compared with controls. Compared with receiver operating characteristic (ROC), which only compares single test accuracy over divergent thresholds for positivity, the each data point in SROC originates from a different study rather than a different threshold [51, 52]. Thus, SROC analysis could provide better evaluation for overall accuracy across SENS and SPEC from multiple studies [53]. Of note, an AUC under SROC curve over 0.75 usually indicates good diagnostic accuracy. Accordingly, the AUC of 0.86 suggests that our analysis was accurate and credible [53].
Moreover, the respective PLR and NLR were 3.95 and 0.26, which indicated that the circRNA signature reached 3.95 between the true positive and false positive rates, and the probability of osteosarcoma patients that tested negative for circRNAs versus the case tested positive had a ratio of 0.26. In addition, the pooled DOR of 15.48 was obtained, implicating that dysregulated circRNAs were a powerful predictive biomarker for osteosarcoma diagnosis. Currently, serum alkaline phosphatase (ALP), with AUC of 0.673, is one of the well-known biomarkers for osteosarcoma diagnosis [19]. Moreover, the sensitivity of ALP in OS diagnosis is also comparatively low, since over 40% of osteosarcoma patients may have normal ALP expression [3]. Our results indicated that dysregulated circRNAs may be even better than ALP for separating osteosarcoma patients from normal subjects with both enhanced sensitivity and specificity, suggesting a potential application in clinical practice.

However, several obstacles also remain in the utility of circRNAs for diagnosis. For instance, circRNAs obtained from tissue may be invasive to the patients, and detection of circRNAs from exosome may be expensive or technically difficult, which could limit the widespread application of multiple circRNAs as biomarkers [6]. Therefore, we also explored the diagnosis accuracy of the combination of a small set of certain oncogenic circRNAs, which may be more available than the full panel of included circRNAs in clinical application. Our preliminary pooled result showed that combination of three oncogenic circRNA, including circPVT1, circ_0081001, and CDR1as, has an AUC of 0.9116, with SENS of 0.85 and SPEC of 0.83 in diagnosis. However, as we mentioned above, the interpretation of the small set of circRNAs in separating osteosarcoma patients from control should be more cautious due to the small study number as well as limited sample size.

Additionally, it worth noted that several other limitations remain to be addressed in this study. First, the included number of article on downregulated circRNAs was still comparatively small, making it difficult to conduct a stratified analysis in terms of certain clinical characteristics. Second, some HRs with 95% CIs were indirectly extracted from Kaplan-Meier curves, which may generate possible bias. Third, the detailed function and underlying mechanisms of circRNAs in osteosarcoma were not thoroughly annotated. Fourth, all of the studies were performed in China, which may introduce population bias. Thus, more studies with different ethnic groups are still warranted in future studies.

**Conclusion**

This study indicated a significant correlation between aberrant expression of circRNAs with the clinical, diagnostic and prognostic values in osteosarcoma patients. Therefore, circRNAs could serve as promising biomarkers and therapeutic targets for osteosarcoma.

**Abbreviations**

ALP: alkaline phosphatase; AUC: area under the curve; ceRNA: competing endogenous RNA; CIs: confidence intervals; CircRNAs: circular RNAs; DFS: disease-free survival; DM: distant metastasis; DOR: diagnostic odds ratio; HRs: hazard ratios; miRNA: microRNA; ncRNAs: non-coding RNAs; NLR: negative likelihood ratio;
ORs: odds ratios; OS: overall survival; PFS: progression-free survival; PLR: positive likelihood ratio; PRISMA: Preferred Reporting Items for Systematic Review and Meta-analysis; QUADAS: Quality assessment for studies of diagnostic accuracy; RBP: RNA-binding protein; ROC: receiver operating characteristic; SENS: sensitivity; SPEC: specificity; SROC: summary receiver operator characteristic

Declarations

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Not applicable.

Authors’ contributions

Conception and design: CT; Collection and assembly of data: CHZ and JYH; Data analysis and interpretation: All authors; Manuscript writing: CT and ZHL; Final approval of manuscript: All authors.

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Availability of data and materials

The data used and analyzed in the study is available from the corresponding author on reasonable request.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing Interest

Chao Tu and Zhihong Li are members of the editorial board of BMC Cancer. Other authors declare that they approve this article and have no competing interests.

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Figures
Figure 1

Publication bias judged by the funnel plots for diagnostic and prognostic analysis. Deek’s funnel plot for the diagnostic effect (A). Begg’s test for upregulated circRNA signature in predicting OS (B) and DFS (D), and downregulated circRNA in predicting OS (E) in osteosarcoma. The “trim and fill” method was applied to assess the possible effects of bias on the overall pooled effects of upregulated circRNA signature (C). The squares indicate the imputed studies. OS, overall survival; DFS, disease-free survival.
Figure 2

Sensitivity analysis of the upregulated circRNA profiles for OS (A) and DFS (B), and downregulated circRNA (C) signature for OS in osteosarcoma. OS, overall survival; DFS, disease-free survival.
Figure 3

Forest plots describing the OS (A) and DFS (B) for the upregulated circRNAs, and OS (C) for downregulated circRNAs in osteosarcoma. OS, overall survival; DFS, disease-free survival.
Figure 4

Forest plots of the combined sensitivity and specificity (A), PLR and NLR (B), DOR (C), and the SROC curve (D) in predicting diagnosis of osteosarcoma. DOR, diagnostic odds ratio; NLR, negative likelihood ratio; PLR, positive likelihood ratio; SROC, summary receiver operator characteristic.
Figure 5

The PRISMA flow diagram of study selection process.

Supplementary Files

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- Additionalfile1PRISMAChecklist.pdf