Evaluation of Guanfacine as a Potential Medication for Alcohol Use Disorder in Long-Term Drinking Rats: Behavioral and Electrophysiological Findings

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One of the main treatment challenges in alcohol use disorder (AUD) is the high rate of craving in combination with decreased cognitive functioning including impaired decision making and impulse control that often lead to relapse. Recent studies show that guanfacine, an a2-adrenoceptor agonist and FDA-approved ADHD medication, attenuates stress-induced relapse of several drugs of abuse including alcohol. Here we evaluated guanfacine’s effects on voluntary alcohol intake, the alcohol deprivation effect (ADE), alcohol seeking behavior, and cue/priming-induced reinstatement in Wistar rats that had voluntarily consumed alcohol for at least 2 months before treatment. In addition, guanfacine’s ability to regulate glutamatergic neurotransmission was evaluated through electrophysiological recordings in medial prefrontal cortex (mPFC) slices prepared from long-term drinking rats (and alcohol-naive controls) that had received three daily guanfacine (0.6 mg/kg/day) or vehicle injections in vivo. Guanfacine decreased alcohol intake in high, but not low, alcohol-consuming rats and the effects were generally more long lasting than that of the AUD medication naltrexone. Repeated guanfacine treatment induced a long-lasting decrease in alcohol intake, persistent up to five drinking sessions after the last injection. In addition, guanfacine attenuated the ADE as well as alcohol seeking and cue/priming-induced reinstatement of alcohol seeking. Finally, subchronic guanfacine treatment normalized an alcohol-induced dysregulated glutamatergic neurotransmission in the mPFC. These results support previous studies showing that guanfacine has the ability to improve prefrontal connectivity through modulation of the glutamatergic system. Together with the fact that guanfacine appears to be clinically safe, these results merit evaluation of guanfacine’s clinical efficacy in AUD individuals.

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INTRODUCTION

Alcohol use disorder (AUD) is a chronic relapsing disorder significantly contributing to the global burden of disease (Rehm et al, 2009). Attenuation of alcohol’s reinforcing properties and prevention of relapse has historically been the major focus in medication development. Recently, the focus has shifted into understanding the role of neurobiological dysregulations in the prefrontal cortex (PFC) induced by chronic alcohol and drug use (eg, a heightened stress system and cognitive impairments) (Goldstein and Volkow, 2011) and evaluating these as potential treatment targets for AUD (Goddard et al, 2010; Gamo and Arnsten, 2011; Holmes et al, 2013). In particular, the noradrenergic and glutamatergic systems in the PFC have been identified as potential core substrates regulating the involved deficits as they might mediate, for example, compulsive drug use and/or cue- and stress-induced craving that in turn may trigger relapse (Goddard et al, 2010; Gamo and Arnsten, 2011; Holmes et al, 2013).

Guanfacine, an α2-adrenoceptor agonist, was recently FDA approved for the treatment of attention-deficit hyperactivity disorder (ADHD) in children and adolescents (Sallee et al, 2009a,b; Sallee and Eaton, 2010). The potential therapeutic efficacy of guanfacine in ADHD was based on the extensive preclinical work done by Arnsten and colleagues (Arnsten et al, 2010; Arnsten and Jin, 2014) showing that guanfacine has the ability to improve prefrontal connectivity, possibly through modulation of the glutamatergic system including N-Methyl-D-aspartate (NMDA) and α-Amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors. Guanfacine’s ability to also improve working memory deficits produced by the NMDA receptor antagonist phencyclidine (Marrs et al, 2005) further highlights the interconnection between the noradrenergic and glutamatergic systems in disorders characterized by cognitive deficits such as ADHD, but also possibly in dependence
disorders. Therefore, evaluation of guanfacine’s potential as a novel medication for dependence disorders has recently been initiated. Indeed, in a human brain imaging study, guanfacine attenuated cocaine craving, arousal, and anxiety while increasing PFC activity in cocaine-dependent individuals (Fox et al., 2012). Guanfacine also attenuated cocaine and alcohol craving in cocaine-dependent individuals co-abusing alcohol (Fox et al., 2014). Furthermore, guanfacine blunted alcohol-seeking behaviors during stress-induced reinstatement (Le et al., 2011) and comorbid depression (Riga et al., 2014) in rats. Collectively, these studies indicate that guanfacine might have the ability to reduce drug use via modulation of craving and dampening of a heightened stress system. However, there is, to our knowledge, no systematic evaluation of guanfacine’s ability to attenuate alcohol-related behaviors without the involvement of stress. In addition, guanfacine’s ability to affect the glutamatergic system in the PFC following voluntary chronic alcohol consumption needs to be elucidated.

The aim of this study was to evaluate the effect of guanfacine (alone and in comparison with the AUD medication naltrexone) on alcohol-mediated behaviors using a battery of animal models that have shown predictive validity for the clinical population, at least for varenicline (Steensland et al., 2012; McKee et al., 2012; Mitchell et al., 2012; Litten et al., 2013) and the monoamine stabilizer (−)-OSU6162 (Steensland et al., 2012, and unpublished data from our research group). Finally, the glutamatergic transmission in the medial PFC (mPFC) was evaluated using electrophysiological recordings in brain slices from long-term drinking rats (and alcohol-naive controls) treated with guanfacine in vivo.

**MATERIALS AND METHODS**

Detailed descriptions of the animals (male Rcc Wistar Han Rats (Harlan, The Netherlands)), housing conditions, chemicals, the intermittent-access 20% ethanol two-bottle-choice (IA20E) paradigm (Wise, 1973; Simms et al., 2008), the operant self-administration paradigms (cue/priming-induced reinstatement and progressive ratio (PR)) (Simms et al., 2010; Steensland et al., 2012), the electrophysiological recordings (Arvanov et al., 1997; Arvanov and Wang, 1998; Konradsson et al., 2006), and descriptions of statistical analyses are available in Supplementary Information.

**Treatment Schedules in the Two-Bottle-Choice Experiments**

For the acute guanfacine treatment, 18 rats that had voluntarily consumed alcohol (IA20E) (Wise, 1973; Simms et al., 2008) for ~5 months were divided into high (4.3 ± 0.2 g/kg per 24 h; n = 11) and low alcohol-consuming rats (1.9 ± 0.2 g/kg per 24 h; n = 7) based on their alcohol consumption during the four drinking sessions preceding the first treatment occasion. All rats received all treatments, guanfacine (0.6 and 0.3 mg/kg) and vehicle, 30 min before the bottles were presented. All injections were given once a week 7 days apart according to a Latin square design, and thus each rat served as its own control.

For the repeated guanfacine treatment, a second group of rats (n = 12) voluntarily consuming high amounts of alcohol (4.3 ± 0.2 g/kg per 24 h; IA20E) for ~7 months were divided into two groups with equal alcohol consumption. Thereafter, they received either guanfacine (0.6 mg/kg) or vehicle treatment 30 min before five consecutive alcohol-drinking sessions. After the fifth injection, all rats were subjected to a 3-week washout period under regular IA20E drinking. Thereafter, the administration schedule was repeated with treatments reversed between the groups. Thus, each animal served as its own control. Eight alcohol sessions after each treatment period were recorded for measurements of potential rebound increase in alcohol intake.

To compare the efficacy of guanfacine with an established AUD treatment, a third group of rats (n = 11) voluntarily consuming high amounts of alcohol (4.4 ± 0.2 g/kg per 24 h) for ~6 months (IA20E), were given guanfacine (0.6 mg/kg), naltrexone (2 mg/kg), or vehicle according to a Latin square design as described for the acute guanfacine experiments above. The naltrexone dose was chosen based on previous dose–response studies in the IA20E model (Simms et al., 2008).

To evaluate the ability of guanfacine to decrease voluntary alcohol intake after a period of protracted abstinence, we used the alcohol deprivation model that is based on the observation that voluntary alcohol intake will increase temporarily (compared with baseline) when alcohol is reintroduced after a period of forced abstinence in alcohol-experienced rats (ie, the alcohol deprivation effect (ADE)) (Spanagel and Holter, 2000). Two groups of rats were alcohol-deprived during 17 days after at least 8 weeks of voluntary consumption of high (4.7 ± 0.2 g/kg per 24 h) or moderate (2.6 ± 0.2 g/kg per 24 h) amounts of alcohol (IA20E), respectively. Each group was thereafter divided into three subgroups with equal alcohol intake during the last three drinking sessions, and subjected to vehicle (saline), guanfacine (0.6 mg/kg), or naltrexone (2 mg/kg) treatment (n = 6–9 per group) 60 min before alcohol was reintroduced. The alcohol intake was measured after 4 and 24 h of alcohol access.

The effect of guanfacine on NMDA- and AMPA-induced currents in pyramidal cells in layer V or VI of the prelimbic cortex in mPFC was evaluated in the long-term drinking rats from the ADE experiment. After the ADE experiment, each rat was given at least 1 week of alcohol consumption before receiving three daily consecutive injections (i.p.) of either guanfacine (0.6 mg/kg, n = 7) or vehicle (n = 5). Each rat received the same treatment (guanfacine or vehicle) as in the ADE experiment. The brains were collected 24 h after the last injection and slices were prepared as described previously (Arvanov et al., 1997; Konradsson et al., 2006). As a control, two groups of alcohol-naive age-matched rats underwent the same guanfacine or vehicle treatment as above (n = 5–6 per group). Details of the electrophysiological recordings are found in Supplementary Information.

The ability of guanfacine (0.6 mg/kg) to decrease alcohol intake without inducing sedative effects was evaluated in two experiments. (1) Acute guanfacine or vehicle treatment was given to rats (n = 8) that had consumed 0.175% NaCl solution for 7 months (intermittent-access two-bottle-choice schedule) as we have previously shown that compounds that specifically decrease the intake of reinforcing solutions, such as alcohol, saccharine, or sucrose, have no effect on the intake of a salty solution (Steensland et al., 2010; Steensland...
et al., 2012). All rats got both guanfacine and vehicle treatments using a within-subject design with 7 days between each injection. (2) Three daily guanfacine or vehicle injections were given to alcohol-naive rats (same rats as in the electrophysiological experiment) and the effect on the water intake was measured 23 h after each injection (n = 5–6 rats per treatment group).

RESULTS
Acute Guanfacine Treatment Decreased Alcohol Intake in Rats Voluntarily Consuming High Amounts of Alcohol
The effects of acute guanfacine treatment (0.3 and 0.6 mg/kg) on voluntary alcohol consumption was evaluated in a group of rats that had voluntarily emerged into high (4.3 ± 0.2 g/kg per 24 h; n = 11) or low (1.9 ± 0.2 g/kg per 24 h; n = 7) alcohol consumers following ~ 5 months of IA20E. Analysis of the alcohol intake showed an overall main effect of treatment (4 h (F(3, 48) = 18, P < 0.001) and 24 h (F(3, 48) = 18, P < 0.001)) and type (high or low drinkers) at both time points (4 h (F(1, 16) = 36, P < 0.001) and 24 h (F(1, 16) = 33, P < 0.001)). Furthermore, there was a significant interaction of treatment × type (4 h (F(3, 48) = 6.7, P < 0.01) and 24 h (F(3, 48) = 5.0, P < 0.01)). The post hoc analysis revealed that both doses of guanfacine significantly decreased voluntary alcohol intake compared with vehicle in the high-alcohol-consuming rats (Figure 1a). However, there was no significant effect on the alcohol intake following guanfacine treatment in the rats consuming low amounts of alcohol (Figure 1b). Furthermore, there was a significant decrease in alcohol preference in the rats consuming high but not low, amounts of alcohol (treatment: 4 h (F(3, 48) = 5.0, P < 0.01) and 24 h (F(3, 48) = 31, P < 0.001); type: 4 h (F(1, 16) = 21, P < 0.001) and 24 h (F(1, 16) = 21, P < 0.001); treatment × type: 4 h (F(3, 48) = 3.2, P < 0.05) and 24 h (F(3, 48) = 6.1, P < 0.001); Supplementary Table S1), whereas there was no significant effect on the water intake in either group (treatment: 4 h (F(3, 48) = 2.0, nonsignificant (n.s.)) and 24 h (F(3, 48) = 32, P < 0.001); type: 4 h (F(1, 16) = 9.1, P < 0.01) and 24 h (F(1, 16) = 5.3, P < 0.05); treatment × type: 4 h (F(3, 48) = 1.9, n.s.) and 24 h (F(3, 48) = 1.2, n.s.; Supplementary Table S1). When analyzing the alcohol intake during the first drinking session after the treatment in the rats consuming high amounts of alcohol, there was no overall main effect at any time point (4 h (F(2, 32) = 0.2, n.s.) and 24 h (F(2, 32) = 0.06, n.s.; Figure 1c). Thus, no post hoc analysis was performed. Finally, guanfacine treatment had no significant effect on voluntary intake of a salty solution compared with vehicle (Supplementary Table S2).

Repeated Guanfacine Treatment Induced Long-Lasting Decrease in Voluntary Alcohol Intake
The effects of repeated guanfacine treatment (0.6 mg/kg/day) during five consecutive drinking sessions were evaluated in rats that had voluntarily consumed high amounts of alcohol (4.3 ± 0.2 g/kg per 24 h) for ~ 7 months. When analyzing the effects on alcohol intake at the 4 and 24 h time points (Figure 2), there was an overall main effect on treatment (4 h (F(1, 11) = 144, P < 0.0001) and 24 h (F(1, 11) = 137, P < 0.0001)), day (4 h (F(12, 132) = 9.7, P < 0.0001) and 24 h (F(12, 132) = 13, P < 0.0001)), and a significant interaction of treatment × day (4 h (F(12, 132) = 11, P < 0.0001) and 24 h (F(12, 132) = 15, P < 0.0001)). Planned comparisons revealed that guanfacine significantly decreased voluntary alcohol intake compared with vehicle on all five treatment days and remained significantly lower compared with vehicle for up to five alcohol drinking sessions after the guanfacine treatment was terminated. In addition, there was an overall main effect at both time points for alcohol preference for treatment (4 h (F(1, 11) = 72, P < 0.0001) and 24 h (F(1, 11) = 99, P < 0.0001)), day (4 h (F(12, 132) = 7.0, P < 0.0001) and 24 h (F(12, 132) = 20, P < 0.0001)), and a significant interaction of treatment × day (4 h (F(12, 132) = 5.4, P < 0.0001) and 24 h (F(12, 132) = 17, P < 0.0001)) as well as for the water intake (treatment: 4 h (F(1, 11) = 21, P = 0.001) and 24 h (F(1, 11) = 90, P < 0.0001); day: 4 h (F(12, 132) = 2.4, P < 0.01) and 24 h (F(12, 132) = 20, P < 0.0001); treatment × day: 4 h (F(12, 132) = 1.9, P < 0.05) and 24 h (F(12, 132) = 15, P < 0.0001)). Planned comparisons showed that guanfacine significantly decreased alcohol preference and increased water intake compared with vehicle. Furthermore, there was no overall main effect of treatment (F(1, 9) = 0.02, n.s.) or day (F(3, 27) = 0.9, n.s.) on the water intake in the alcohol-naive rats that received three consecutive guanfacine or vehicle injections (Supplementary Table S3).

Guanfacine Was More Efficacious Than Naltrexone to Decrease Alcohol Intake in Rats Voluntarily Consuming High Amounts of Alcohol
The efficacy of guanfacine (0.6 mg/kg) to attenuate voluntary alcohol intake was compared with that of the AUD medication naltrexone (2 mg/kg) in rats that had voluntarily consumed high amounts of alcohol (4.4 ± 0.2 g/kg per 24 h; n = 11) for ~ 6 months before the treatment. There was an overall main effect on alcohol intake at both the 4 h (F(2, 20) = 40, P < 0.0001) and 24 h (F(2, 20) = 60, P < 0.0001) time points. Planned comparisons revealed that both guanfacine and naltrexone significantly decreased voluntary alcohol intake compared with vehicle at the 4 h time point (Figure 3a, left panel), whereas only guanfacine significantly decreased the intake compared with vehicle at the 24 h time point (Figure 3a, right panel). In addition, guanfacine significantly decreased alcohol intake compared with naltrexone at both time points.

In the ADE experiment, two groups of rats were subjected to a forced alcohol abstinence period after at least 8 weeks of voluntary intake of moderate or high amounts of alcohol, respectively. Following the reintroduction of alcohol, the vehicle-treated rats drinking moderate (Figure 3b), but not high (Figure 3c), amounts of alcohol significantly increased their alcohol intake at both time points compared with corresponding baseline before the abstinence period. Both guanfacine and naltrexone significantly decreased alcohol intake after the abstinence period compared with respective baseline in both moderate (Figure 3b) and high (Figure 3c) drinkers. The effect of guanfacine was more long lasting than that of naltrexone, especially in high drinkers.
Acute Guanfacine Treatment

**Figure 1** Acute guanfacine treatment (0.3 and 0.6 mg/kg) significantly decreased voluntary alcohol intake compared with vehicle in rats voluntarily consuming (a) high (4.3 ± 0.2 g/kg per 24 h; n = 11) but not (b) low (1.9 ± 0.2 g/kg per 24 h; n = 7) amounts of alcohol for ~5 months before the treatment. (c) There was no rebound increase in alcohol intake during the first drinking session after treatment in rats consuming high amounts of alcohol. All values are expressed as mean ± SEM; ***P < 0.001 compared with corresponding vehicle ((a, b) two-way repeated-measures ANOVA followed by planned comparisons; (c) one-way repeated-measures ANOVA followed by Newman–Keuls multiple comparison test).
Guanfacine Significantly Attenuated Alcohol Seeking and Cue/Priming-Induced Reinstatement of Alcohol-Seeking Behavior

The effect of guanfacine on alcohol-seeking behavior was evaluated using the PR test in rats that had been voluntarily exposed to alcohol for ~5 months using operant self-administration (see details in Supplementary Information). There was an overall main effect on active lever presses (F(2, 24) = 16, P < 0.0001), breakpoint (F(2, 24) = 28, P < 0.0001), and inactive lever presses (F(2, 24) = 6.5, P < 0.01). The post hoc analysis shows that both guanfacine doses significantly inhibited the number of active lever presses as well as the breakpoint compared with vehicle (Figure 4a). The higher guanfacine dose significantly (P = 0.01) decreased the number of inactive lever presses compared with vehicle (vehicle: 2.0 ± 0.5; guanfacine (0.3 mg/kg): 1.3 ± 0.4; guanfacine (0.6 mg/kg): 0.5 ± 0.2).

After the PR test and 2 weeks of baseline self-administration of alcohol, the alcohol-seeking behavior was extinguished during 16 extinction sessions (without the presence of olfactory, visual, and auditory cues) where pressing the active lever produced no delivery of alcohol. During the reinstatement session (where the cues together with a small amount of alcohol in the liquid dispenser (equivalent to one reward) were reintroduced), there was an overall main effect on the number of active lever presses (F(3, 30) = 11, P < 0.001). The post hoc analysis revealed that the vehicle pretreated group reinstated (ie, significantly increased the number of active lever presses compared with that during extinction) and that guanfacine pretreatment attenuated this response (Figure 4b, left panel). There was no overall main effect on the number of inactive lever presses (F(3, 30) = 0.4, n.s.) (Figure 4b, right panel).

Guanfacine Normalized Alcohol-Induced Dysregulated Glutamatergic Neurotransmission in Pyramidal Cells of the mPFC

Guanfacine (0.6 mg/kg/day, i.p.) or vehicle was given during three consecutive days to two groups of rats that had voluntarily been drinking high amounts of alcohol for at least 11 weeks. Alcohol-naïve age-matched controls underwent the same treatment. The electrophysiology recordings were performed in vitro (brains were collected 24 h after the last injection) on cells located in layers V and VI of the prelimbic cortex in the mPFC (located medial to the forceps minor and easily identified in a slice). The electrophysiological criteria used for distinguishing presumed pyramidal versus nonpyramidal cells are described elsewhere (Arvanov et al., 1997; Konradsson et al., 2006). In general, the pyramidal cells exhibited longer spike duration (>1 ms at

Figure 2 Repeated guanfacine treatment (0.6 mg/kg/day over 5 consecutive alcohol drinking sessions) significantly decreased voluntary alcohol intake ((a) 4 h time point and (d) 24 h time point) and preference for alcohol ((b) 4 h time point and (e) 24 h time point) compared with vehicle in rats that had been consuming high amounts of alcohol (4.3 ± 0.2 g/kg per 24 h) for ~7 months before the treatment. There was no tolerance development during treatment and the significant decrease persisted for up to five drinking sessions after the last injection. In addition, there was a significant increase in the water intake ((c) 4 h time point and (f) 24 h time point) compared with vehicle. All values are expressed as mean ± SEM; n = 12; *P < 0.05, **P < 0.01, ***P < 0.001 compared with corresponding vehicle (two-way repeated-measures ANOVA followed by planned comparisons).
Figure 3  Both guanfacine (0.6 mg/kg) and naltrexone (2 mg/kg) significantly decreased alcohol intake compared with vehicle at the 4 h time point (a, left panel) in rats that had voluntarily consumed high amounts of alcohol for ~6 months before the treatment (n = 11). At the 24 h time point (a, right panel) only guanfacine significantly decreased the alcohol intake compared with vehicle. At both time points, guanfacine significantly decreased the alcohol intake compared with naltrexone. All values are expressed as mean ± SEM; ***P < 0.001 compared with vehicle or as indicated (one-way repeated-measures ANOVA followed by planned comparisons). To evaluate the effect of guanfacine and naltrexone on the alcohol deprivation effect (ADE), two groups of rats voluntarily consuming (b) moderate (2.6 ± 0.2 g/kg per 24 h) or (c) high (4.7 ± 0.2 g/kg per 24 h) amounts of alcohol for at least 8 weeks were subjected to a period of 17 days with forced alcohol abstinence before alcohol was reintroduced. The rats were pretreated with guanfacine (0.6 mg/kg), naltrexone (2 mg/kg), or vehicle before renewed alcohol access (n = 6–9 per treatment group). An ADE was present in vehicle-treated rats consuming moderate (b) but not high (c) amounts of alcohol. Both guanfacine- and naltrexone-treated rats significantly decreased their alcohol intake in both high (c) and moderate (b) drinkers compared with corresponding baseline before the abstinence period, with the effects of guanfacine being more pronounced and long lasting than naltrexone. All values are expressed as mean ± SEM; *P < 0.05, **P < 0.01, ***P < 0.001 compared with corresponding values before the abstinence period (paired Student’s t-test within each treatment).
half-maximum spike amplitude) than interneurons, and showed pronounced spike-frequency adaptation in response to constant-current depolarizing pulses. In contrast, interneurons exhibited a brief duration of their action potentials and generally lacked pronounced spike-frequency adaptation. There was an overall main effect on both NMDA-induced ($F(3, 19) = 22, P = 0.0001$; Figure 5a) and AMPA-induced ($F(3, 19) = 30, P < 0.0001$; Figure 5b) inward currents in pyramidal cells of mPFC slices. The post hoc analysis revealed that long-term drinking rats had a significant increase in both NMDA and AMPA currents compared with the alcohol-naive rats. The in vivo guanfacine treatment significantly reversed this alcohol-induced dysregulation, whereas there was no significant effect of guanfacine in the alcohol-naive rats.

DISCUSSION

The main findings in this study were that guanfacine, a FDA-approved ADHD medication, attenuated voluntary alcohol consumption and relapse behavior in Wistar rats that had voluntarily consumed alcohol for at least 2 months before treatment. The effects of guanfacine were generally more pronounced and long lasting than that of the AUD medication naltrexone. In fact, repeated guanfacine...
treatment induced long-lasting reduction in voluntary alcohol intake even when the medication was no longer on board. In addition, we show, to our knowledge for the first time, that \textit{in vivo} guanfacine treatment normalized dysregulated glutamatergic neurotransmission in the mPFC, induced by long-term voluntary alcohol consumption.

In this study, guanfacine demonstrated several desirable effects of a potential AUD medication. First, repeated guanfacine treatment significantly decreased alcohol intake in rats voluntarily consuming high amounts of alcohol, without tolerance development to guanfacine’s effect. Second, the alcohol intake was significantly reduced compared with vehicle for up to five drinking sessions (equivalent to 12 days given the IA20E schedule) after termination of the repeated, but not acute, guanfacine treatment, indicating that sustained guanfacine treatment might induce long-lasting neurochemical alterations beneficial for maintenance of reduced alcohol consumption. Finally, guanfacine decreased voluntary alcohol intake after a long period of forced abstinence as well as alcohol seeking under PR schedule and cue/priming-induced reinstatement. The role of guanfacine as a relapse prevention agent is further supported by previous animal and human studies showing that the compound attenuates stress-induced relapse of alcohol and cocaine (Le \textit{et al}, 2011; Fox \textit{et al}, 2012, 2014) and the motivational drive to seek alcohol in depressed rats (Riga \textit{et al}, 2014).

The electrophysiological experiment in this study showed that guanfacine can normalize alcohol-induced dysregulation of the glutamatergic neurotransmission in the mPFC. These results reflect an \textit{in vivo} pharmacologically induced response in the mPFC as the brain slices were prepared from long-term drinking rats that were treated with three daily guanfacine injections (0.6 mg/kg/day, i.p.) or vehicle injections \textit{in vivo}. Brains were collected 24 h after the last injection. Alcohol-naive, age-matched rats were used as controls. The long-term alcohol consumption significantly increased both the NMDA- and AMPA-induced currents compared with alcohol-naive controls. Guanfacine treatment normalized this alcohol-evoked facilitation while having no significant effect in the alcohol-naive rats. The holding potential was $-60\text{ mV}$. Representative traces showing the NMDA- and AMPA-induced currents following guanfacine and vehicle treatment in alcohol-naive and alcohol-drinking rats are shown in (A) and (B), respectively. The summarized data (mean ± SEM) are found in (C); $n=6$ cells per treatment in the long-term-drinking rats and $n=4$ cells per treatment in the alcohol-naive rats; ***$p<0.0001$ as indicated in the figure (one-way ANOVA, Newman–Keuls multiple comparison test).

**Figure 5** The electrophysiology recordings were conducted \textit{in vitro} in pyramidal cells in layer V or VI in slices prepared from the prelimbic cortex of the mPFC of rats that had voluntarily been drinking high amounts of alcohol for at least 11 weeks. Before the experiment the rats received three daily guanfacine (0.6 mg/kg/day, i.p.) or vehicle injections \textit{in vivo}. Brains were collected 24 h after the last injection. Alcohol-naive, age-matched rats were used as controls. The long-term alcohol consumption significantly increased both the NMDA- (a) and AMPA-induced (b) currents compared with alcohol-naive controls. Guanfacine treatment normalized this alcohol-evoked facilitation while having no significant effect in the alcohol-naive rats. The holding potential was $-60\text{ mV}$. Representative traces showing the NMDA- and AMPA-induced currents following guanfacine and vehicle treatment in alcohol-naive and alcohol-drinking rats are shown in (A) and (B), respectively. The summarized data (mean ± SEM) are found in (C); $n=6$ cells per treatment in the long-term-drinking rats and $n=4$ cells per treatment in the alcohol-naive rats; ***$p<0.0001$ as indicated in the figure (one-way ANOVA, Newman–Keuls multiple comparison test).
expression of subunits of these receptors (see review (Holmes et al, 2013)). Such alcohol-induced dysregulation is supported by the present findings, showing increased NMDA- and AMPA-induced currents in pyramidal cells in the mPFC in rats that had consumed high amounts of alcohol for ~3 months. Interestingly, 3 days of guanfacine treatment normalized this alcohol-induced hyperglutamatergic transmission, providing support for a role of this x-2-adrenoceptor agonist as a modulator of the glutamatergic system in the mPFC. In fact, guanfacine has previously been shown to inhibit the production of CAMP, with related blockade of CAMP-dependent and hyperpolarization-activated cyclic nucleotide-gated channels leading to enhanced excitatory transmission and improved PFC connectivity (Wang et al, 2007). This mechanism together with previous studies showing that guanfacine improves impulsive behavior in rats (Fernando et al, 2012) as well as working memory (with corresponding increase in blood flow in the PFC) in monkeys (Avery et al, 2000) strengthen the case of guanfacine’s therapeutic profile as a cognitive enhancer.

In this study it was found that the effects of guanfacine were selective to high alcohol consumption as shown by: (1) a significant effect of guanfacine on the NMDA- and AMPA-induced currents in long-term drinking, but not alcohol-naive rats and (2) reduced alcohol intake in rats that had voluntarily been drinking higher, but not lower, amounts of alcohol for ~5 months before the treatment. This selective effectiveness is supported by previous clinical studies indicating that guanfacine’s therapeutic efficacy may be more specific in conditions characterized by dysfunction in the catecholamine systems (Arnsten et al, 1996) such as ADHD and impulse control disorders (Taylor and Russo, 2001). Indeed, alcohol and substance use disorders share common pathophysiology with ADHD (including PFC connectivity (Wang et al, 2007). This mechanism together with previous studies showing that guanfacine improves impulsive behavior in rats (Fernando et al, 2012) as well as working memory (with corresponding increase in blood flow in the PFC) in monkeys (Avery et al, 2000) strengthen the case of guanfacine’s therapeutic profile as a cognitive enhancer.

In rats, an increased demand for alcohol after protracted abstinence (ie, the ADE) has been suggested to be induced by cues (eg, the smell of alcohol) or a priming stimulus (eg, intake of small amount of alcohol) (Spanagel and Holter, 2000) and to be dissociated from normal eating or drinking behavior (Sanchis-Segura and Spanagel, 2006). Thus, the ADE has been suggested to resemble a typical relapse situation in AUD patients. In this study, guanfacine and naltrexone significantly attenuated ADE occurring after vehicle treatment in rats voluntarily consuming moderate amounts of alcohol (2.6 ± 0.2 g/kg per 24 h). In contrast, the vehicle-treated rats consuming high amounts of alcohol (4.7 ± 0.2 g/kg per 24 h) did not display an ADE. Nevertheless, both guanfacine and naltrexone decreased alcohol intake compared with corresponding baseline before the abstinence period also in the high-alcohol-consuming rats. The lack of ADE in rats voluntary drinking high amounts of alcohol might be because of a ceiling level in alcohol intake. This hypothesis is supported by a previous study showing that AA rat selectively bred for high alcohol preference (consuming similar levels of alcohol as the high-alcohol-consuming Wistar rats in the present study) do not display an ADE (Sinclair and Tiihonen, 1988). However, it should be noted that an ADE has been found in other rat strains selectively bred for high alcohol preference (McKinzie et al, 1998; Vengeliene et al, 2003), indicating that other factors, in addition to the level of baseline consumption, are important for the induction of an ADE. Regardless of the presence of an ADE or not, our results indicate that guanfacine might have the ability to prevent a slip from becoming a relapse after a period of prolonged abstinence.

It has previously been shown that a markedly higher guanfacine dose than used in the present study (1.5 vs 0.6 mg/kg) decreased voluntary alcohol intake in alcohol-preferring AA rats (Opitz, 1990). However, in a pilot study, the dose of 1.5 mg/kg was clearly sedating (unpublished data from our research group), and 1 mg/kg of guanfacine induced sedative effects in the novel open field test (Smith and Aston-Jones, 2011). Thus, it is possible that the decreased alcohol intake previously observed by Opitz (1990) could be a nonspecific effect related to the sedative effects of guanfacine at a high dose. In this study, with the highest dose of 0.6 mg/kg, the nonspecific effects are less likely as guanfacine significantly increased the water intake (in the presence of alcohol) and had no significant effect on water intake in alcohol-naive rats or voluntary intake of a salty solution. In addition, there was no significant effect on the inactive lever in the cue/priming-induced reinstatement experiment (although the highest guanfacine dose significantly decreased the number of inactive lever presses in the PR test (vehicle: 2.0 ± 0.5; guanfacine: 0.5 ± 0.2 lever presses). However, it should be noted that the general responding on the inactive lever was too low to be a reliable marker for possible locomotor deficits. Furthermore, previous studies showed that guanfacine (0.5 mg/kg) did not affect explorative behavior (Riga et al, 2014) or operant self-administration of sucrose (Le et al, 2011). Thus, collectively the present and previous results indicate that guanfacine’s ability to attenuate voluntary alcohol intake, alcohol-seeking behavior, and cue/priming-induced reinstatement is most likely not caused by sedation or a general effect on locomotor behavior.
Guanfacine’s effect on human alcohol consumption remains to be investigated; however, we hypothesize that the present results showing that guanfacine attenuates several voluntary alcohol-mediated behaviors in animal models with predictive validity are suggestive of the putative effect of guanfacine as a novel AUD medication. The present and previous promising clinical and animal studies with guanfacine in different dependency disorders (Le et al, 2011; Fox et al, 2012, 2014; Riga et al, 2014), together with the findings that guanfacine appears to be safe and well tolerated in humans (Sallee et al, 2009a, b; Sallee and Eaton, 2010; Fox et al, 2012, 2014), merits evaluation of guanfacine’s clinical efficacy in AUD patients.

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The authors declare no conflict of interest.

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