ASCITIC INTERLEUKIN 6 AND SERUM PROCALCITONIN FOR EARLY DETECTION OF SPONTANEOUS BACTERIAL PERITONITIS IN ADVANCED LIVER CIRRHOSIS

Marwa Ibrahim Metawea (1), Doaa Ahmed Elwazzan (2), Reham Abdel Haleem Abo Elwafa (3), Sara Lotfy Asser (4)

1) Department Hepatology, Faculty of Medicine, Alexandria University.
2) Department Tropical Medicine, Faculty of Medicine, Alexandria University.
3) Department Clinical Pathology, Faculty of Medicine, Alexandria University.
4) Department Microbiology, Faculty of Medicine, Alexandria University.

ABSTRACT

Background: Liver cirrhosis is associated with frequent bacterial infections that increase mortality. Spontaneous bacterial peritonitis (SBP) is an important cause of mortality and morbidity in such patients with ascites. A polymorphonuclear (PMN) cell count >250/μl in the ascitic fluid is the current gold standard for diagnosing SBP which is considered a subjective test. Early diagnosis of SBP although vital is difficult in these patients. Interleukin 6 is a pro-inflammatory marker that increases earlier in bacterial infection than other inflammatory markers. This is crucial in cirrhotic patients to initiate treatment accordingly.

Objective: Evaluate the role of IL 6 together with other inflammatory markers in early diagnosis of SBP and as prognostic markers as well.

Patients and methods: The study was conducted on 60 cirrhotic patients with ascites divided into 2 groups. Group I included 30 cirrhotic patients with sterile ascites and group II, 30 patients with SBP. Liver profile, serum creatinine, serum sodium, ascitic IL6, C-reactive protein, serum procalcitonin, and ascitic fluid analysis were done for all patients in both groups. Exclusion criteria included: acute infection, diabetes mellitus, coronary vascular disease, collagen vascular disease, and any form of sepsis.

Results: Ascitic IL6 mean value was 2171.5(62.2 – 5000) and 342.5(146 – 2567) pg/ml in groups I and II respectively which was significantly higher in group I than among group II patients (P<0.001). Furthermore, the mean s. procalcitonin was 0.8(0.4-1.1) and 0.4(0.04-0.7) ng/ml among groups In and II respectively and it was significantly higher among the patients with SBP (P<0.001). Diagnosis of SBP among patients with liver cirrhosis could be suggested when ascitic IL6 and serum procalcitonin are assessed at a cut-off values of >780 pg/ml and >0.4ng/ml respectively.

Conclusion: Ascitic IL6 and serum procalcitonin can be used as a valuable surrogate serum marker for early diagnosis of SBP in cirrhotic patients.

Keywords: Spontaneous bacterial peritonitis (SBP), inflammatory markers, IL 6, highly sensitive C-reactive protein and procalcitonin

INTRODUCTION

Bacterial infections are one of the deleterious complications in patients with cirrhosis as they carry high morbidity and mortality. Among these bacterial infections, spontaneous bacterial peritonitis (SBP), is the most common with an incidence ranging between 10 to 30% and around 32% mortality rate(1,2).

Spontaneous bacterial peritonitis is defined as an ascitic fluid infection in the absence of detectable intra-abdominal surgically-treatable source. SBP is clinically suspected in patients with ascites due to advanced cirrhosis who develop symptoms such as fever, abdominal pain/tenderness or diarrhea. With progression of sepsis it can cause altered mental status, paralytic ileus, hypotension, hypothermia and septic shock. Laboratory abnormalities are those of sepsis and are non-specific, they may show peripheral leukocytosis, metabolic acidosis, and azotemia. About 13% of patients with SBP may lack signs of infection at time of diagnosis(2).

Spontaneous bacterial peritonitis is classified into: typical SBP with PMNL count >250 /mm3 and positive ascitic fluid culture without any evidence of external or intra-abdominal source of infection or malignancy, culture-negative SBP with PMNL >250/mm3 and a negative ascitic fluid culture, bacterascites with a positive ascitic fluid culture and an ascitic PMNL count <250 cells/mm3, sterile ascites with PMNL <250/ mm3 and a negative ascitic fluid culture. (2)
antibacterial polypeptides that end up in bacterial translocation in decompensated liver cirrhosis. ii During acute infections and inflammation as in the case of SBP, there is release of a number of pro-inflammatory and inflammatory mediators and cytokines which might have deleterious effect despite their role in host defense mechanisms.

Interleukin 6 is a pro-inflammatory cytokine produced by numerous cells, including leukocytes, adipocytes, endothelial cells, fibroblasts, and myocytes. It acts as a hepatocyte stimulatory factor to induce acute phase reactions. Acute phase reactants produced, whether positive such as C-reactive protein and fibrinogen or negative as albumin, indicate the presence of inflammation and correlate with its intensity. iii, iv, v

Another pro-inflammatory marker is procalcitonin (PCT) which is a peptide precursor of calcitonin. It is secreted mainly by parafolicular thyroid cells. It is also secreted by neuroendocrine cells of the lung and intestine in response to sepsis, which makes it a good diagnostic and monitoring tool in such patients. It was found also that PCT level correlates with the severity and extent of infection. vi, vii

**PATIENTS AND METHODS**

**Study design:**

This study was conducted on 60 patients admitted at the internal and tropical medicine departments, Alexandria University hospitals suffering from Hepatitis C virus induced liver cirrhosis and ascites. The patients were classified according to presence or absence of SBP at time of admission into two groups:

**Group 1:** Included 30 patients with SBP based on PMNL count >250/mm3 with/without positive culture.

**Group 2:** Included 30 patients with sterile cirrhotic ascites (PMNL <250 mm3 and a negative ascitic fluid culture). Patients with clinical evidence of infections other than SBP, malignancy, auto-immune diseases and on drugs affecting CRP level as statins and fibrates were excluded from the study. A written informed consent was obtained for all patients enrolled in the study.

About 6 ml of venous blood were withdrawn aseptically into two vacutainer tubes as following: Four ml were delivered into a plain tube, to separate serum for routine investigations. Two ml were delivered into EDTA vacutainer tube to separate plasma for procalcitonin. Samples for PCT were separated and analyzed within four hours of the blood draw. If not, samples were stored at 2-8 °C for up to 24 hours.

Twenty ml of ascitic fluid were tapped into 2 sterile containers, the first one was for the physical, chemical and microscopic examination of ascitic fluid and the second one was for culture and sensitivity.

All study subjects were investigated for fasting blood sugar, 2-hours post prandial blood sugar, liver profile, renal function tests, and highly sensitive CRP. All routine investigations were measured by the Biosystems A25 Automated chemistry analyzer (Biosystems, Spain).

Serum sodium and potassium were measured using AVL 9180 Series Electrolyte Analyzers (Roche Diagnostics, USA). Procalcitonin was measured by Chemiluminescent assay (CLIA) on the ADVIACentaur® (Siemens, Germany). viii

IL6 was measured in ascitic fluid by Enzyme linked immunosorbent assay using Human IL-6 Quantikine ELISA Kit (R&D, USA) according to the manufacturer instructions.ix

**STATISTICAL ANALYSIS OF THE DATA**

Data were fed to the computer and analyzed using IBM SPSS software package version 20.0. Comparisons between groups for categorical variables were assessed using Chi-square test. Mann Whitney test was used to compare two groups for abnormally distributed quantitative variables. While Student t-test was used for comparing the two studied groups for normally distributed quantitative variable. Significance of the obtained results was judged at the 5% level.

**RESULTS:**

Sixty patients with HCV induced liver cirrhosis and ascites were enrolled in this study. The cases of SBP (group I) were 30 of which 18 (60%) patients were men, mean age was 60 ± 9.9 years. Among the 30 other cases without SBP (group II): 18 (60%) were males, the mean age was 57.3 ± 10.2 years. (Table I)

| Test of Sig. | p |
|-------------|---|
| χ²= 0.000    | 1.000 |
| t= 1.042     | 0.302 |

*Statistically significant at p ≤ 0.05

| Group I (n = 30) | Group II (n = 30) |
|------------------|-------------------|
| Male 18(60%)     | 18(60%)           |
| Female 12(40%)   | 12(40%)           |
| Age (years) 60 ± 9.9 | 57.3 ± 10.2 |

χ², p; χ² and p values for **Chi square test** for comparing between the two groups, p: t and p values for **Student t-test** for comparing between the two groups
Among group I, the mean serum albumin was 2.6 ± 0.4 g/dl, serum bilirubin was 1.9(0.4 – 6) mg/dl, ALT ranged from 12 to 156 u/l with a mean 31.5 u/l, mean AST was 34.5(29 – 156) u/l, mean prothrombin time was 14.6 ± 1.1 sec with mean INR 1.3 ± 0.1. Regarding group II, the mean s. albumin was 2.7 ± 0.3 g/dl, mean s. bilirubin was 1.3 mg/dl ranging from 0.5 to 2.3, mean ALT and AST were 33.5 u/l for both, mean PT was 14.8 ± 2.1 seconds and mean INR 1.4 ± 0.3. Serum LDH was higher among group I, but with no significant difference. (Table 2)

Regarding the CBC findings, the mean HB was 11.9 ± 1.4 and 10.7 ± 1.2 g/dl among the patients in groups I and II respectively, it was significantly lower in group II than group I (p=0.001). The mean WBC was significantly higher among group I than group II (P<0.001), while platelets count was significantly higher in group I than group II (P=0.023). (Table 3)

Table (4) showed the results of renal function tests, serum Na and K in the studied groups. The mean blood urea and serum creatinine in group I were 65 (32-195) and 1.5 (0.7-1.3) mg/dl respectively, while in group II blood urea and creatinine were 45(24-190) and 1.1(0.7-1.3) mg/dl respectively.

Creatinine was significantly higher among group I than among group II (P= 0.001).

The mean serum Na was 133.7± 4.4 and 133.5 ± 3 mmol/L among groups I and II respectively, while mean serum K was 4.5(2.9 – 6) and 4.4(3 – 7) mmol/L in groups I and II respectively. No significant difference was found between both groups regarding serum Na and K (P=0.84).

Regarding the degree of ascites, 21 patients (70%) had moderate ascites and 9 patients (30%) had tense ascites among group I. In group II; 15 patients (50%) had mild ascites, 6 patients (20%) had moderate ascites and 9 patients (30%) had tense ascites. Significant difference was detected between the two studied groups regarding the degree of ascites (P<0.001). (Table 5)

Table 2: Distribution of the studied cases according to the liver functions

|                      | Group I (no=30) | Group II (no=30) | Test of sig. (t) | P     |
|----------------------|-----------------|------------------|-----------------|-------|
| Serum Albumin (g/dl) | 2.6 ± 0.4       | 2.7 ± 0.3        | 1.555           | 0.125 |
| Serum bilirubin (mg/dl) | 1.9(0.4 – 6)    | 1.3(0.5 – 2.3)   | 351.0           | 0.141 |
| ALT (u/l)            | 31.5(12 – 156)  | 33.5(10 – 67)    | 447.0           | 0.965 |
| AST (u/l)            | 34.5(29 – 156)  | 33.5(16 – 60)    | 370.5           | 0.239 |
| PT (sec)             | 14.6 ± 1.1      | 14.8 ± 2.1       | 0.570           | 0.572 |
| INR                  | 1.3 ± 0.1       | 1.4 ± 0.3        | 1.771           | 0.085 |
| Serum LDH (mmol/l)   | 381.5(213 – 480)| 320(120 – 394)   | U= 318.0        | 0.051 |

Table 3: Comparison between group I and group II according to CBC findings

|                      | Group I (no=30) | Group II (no=30) | Test of sig. | P     |
|----------------------|-----------------|------------------|--------------|-------|
| HB (g/dl)            | 11.9 ± 1.4      | 10.7 ± 1.2       | t=3.483      | *0.001* |
| WBCs (x10³/mm³)      | 10.4 ± 1.7      | 5.5 ± 1.7        | t= 11.435    | <0.001* |
| PLT (x10³/mm³)       | 134(87 – 353)   | 115(69 – 257)    | t= 297.0     | *0.023* |

Table 4: Renal function tests, serum Na and K in the studied groups.

|                      | Group I (no=30) | Group II (no=30) | Test of sig. | P     |
|----------------------|-----------------|------------------|--------------|-------|
| Blood urea (mg/dl)   | 65(32 – 195)    | 45(24 – 190)     | U= 360.0     | 0.182 |
| Serum Creatinine (mg/dl) | 1.5(0.7 – 4)    | 1.1(0.7 – 1.3)   | U= 234.0     | *0.001* |
| Na (mmol/L)          | 133.7 ± 4.4     | 133.5 ± 3        | t=0.206      | 0.838 |
| K (mmol/L)           | 4.5(2.9 – 6)    | 4.4(3 – 7)       | U= 436.5     | 0.841 |

Table 5: Distribution of the studied groups according to the degree of ascites

| Ascites (No (%))     | Group I (no=30) | Group II (no=30) | Test of sig. | P     |
|----------------------|-----------------|------------------|--------------|-------|
| Mild                 | 0(0%)           | 15(50%)          |              |       |
| Moderate             | 21(70%)         | 6(20%)           | Z=23.333     | <0.001* |
| Tense                | 9(30%)          | 9(30%)           |              |       |
Using Child-Pugh classification, 21 (70%) patients with SBP (group I) were Child B and 9 (30%) patients were Child C (mean of the score was 8.9 ± 1). Among group II, 27 (90%) patients were Child B and 3 (10%) patients were Child C (mean of the score was 8.2±1.1). Significant difference was detected between group I and group II regarding Child Pugh score (P=0.011). (Table 6)

**Ascitic fluid analysis in both groups revealed the following: (Table 7)**

The mean protein content in the ascitic fluid was (2.2 ± 0.6) and (3.6 ± 0.8) gm/dl in groups I and II respectively, it was significantly lower in group I than in group II (P<0.001).

The mean ascitic glucose was (112.2 ± 53) and (104 ± 27.9) mg/dl in groups I and II respectively. It was significantly higher in group I than in group II (P<0.001).

The culture of ascitic fluid was positive in 24 patients (80%) in group I, while it was positive in 3 patients (10%) in group II which was significantly different (P<0.001).

Among the inflammatory markers used to diagnose patients with SBP was IL6 in ascitic fluid; its mean value was 2171.5(62.2 – 5000) and 342.5(146 – 2567) pg/ml in groups I and II respectively which was significantly different among the two groups (U=153) (P<0.001). (Table 8)

Furthermore, the mean serum procalcitonin was 0.8(0.4-1.1) and 0.4(0.04-0.7) ng/ml among the patients in groups I and II respectively and it was significantly higher in patients of SBP (P<0.001). (Table 8 and figure 1) Serum CRP was higher among cases of SBP in group I, but with no significant difference. (Table 8)

Receiver operating characteristic curves (ROC curves) of ascitic IL6 as well as serum procalcitonin were done to estimate the cutoff points at which SBP can be diagnosed. The ROC curve for ascitic IL6 was significant (<0.001) (AUC=0.830) and showed that the cutoff point to diagnose SBP was >780 pg/ml, with sensitivity of 90%, specificity of 80%, positive predictive value of 81.8% and negative predictive value of 88.9%. (Figure 2, Table 9)

The ROC curve for serum procalcitonin was significant (<0.001) (AUC=0.875) and showed that the cutoff point to diagnose SBP was >0.4 ng/ml, with sensitivity of 90%, specificity of 70%, positive predictive value of 75% and negative predictive value of 87.5% ( Figure 3 , Table 10).

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**Table 6: Distribution of the studied groups according to Child Pugh score**

| Child score (No (%)) | Group I (no=30) | Group II (no=30) | Test of sig. | P  |
|----------------------|-----------------|-----------------|--------------|----|
| A                    | 0(0%)           | 0(0%)           |              |    |
| B                    | 21(70%)         | 27(90%)         | t=2.633      | 0.011* |
| C                    | 9(30%)          | 3(10%)          | X²=3.750     | 0.053 |

**Table 7: Different lab parameters and culture of ascitic fluid in the studied groups**

|                        | Group I (no=30) | Group II (no=30) | Test of sig. | P  |
|------------------------|-----------------|-----------------|--------------|----|
| **Ascitic Protein (g/dl)** | 2.2 ± 0.6       | 3.6 ± 0.8       | t=7.806      | <0.001* |
| **Ascitic Glucose (mg/dl)** | 112.2 ± 53      | 104 ± 27.9      | t=0.750      | 0.457 |
| **Ascitic LDH (U/L)**   | 272(50 – 550)   | 54.5(20 – 140)  | U= 126.0*    | <0.001* |
| **Ascitic PMNL (cells/mm<sup>3</sup>)** | 300(60 – 440)   | 52.5(18 – 210)  | U= 45.0*    | <0.001* |
| **Ascitic RBC (cells/mm<sup>3</sup>)** | 38.5(12 – 2250) | 40.5(10 – 140)  | U=355.5      | 0.162 |
| **Ascitic Lymphocytes (cells/mm<sup>3</sup>)** | 77.5(12 – 400)  | 34.5(10 – 160)  | U=297.0*    | 0.023* |
| **Ascitic Culture**     | Positive 24(80%) | 3(10%)          | X²=29.697*   | <0.001* |

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Table 8: Comparison between the studied groups according to some Inflammatory markers

|                  | Group I (n=30) | Group II (n=30) | Test of sig. | P     |
|------------------|----------------|-----------------|--------------|-------|
| Ascitic IL6 (pg/ml) | 2171.5(62.2 – 5000) | 342.5(146 – 2567) | U= 153.0* | <0.001* |
| Serum procalcitonin (ng/ml) | 0.8(0.4 – 1.1) | 0.4(0.04 – 0.7) | U=112.5 | <0.001* |
| Serum CRP (mg/l)  | 68(32 – 98) | 56.5(12 – 83) | U=337.5 | 0.096 |

Figure (1): Comparison between the two studied groups according to Serum procalcitonin (ng/ml)

Figure (2): ROC curve for ascitic IL6 to detect SBP patients group I from group II
Table 9: Accuracy of IL6 to detect SBP.

|               | AUC   | P        | 95% C.I   | Cut off     | Sensitivity | Specificity | PPV  | NPV  | LL    | UL    |
|---------------|-------|----------|-----------|-------------|-------------|-------------|-------|-------|-------|-------|
| a.IL6         | 0.830 | <0.001†  | 0.713     | >780pg/ml   | 90.0%       | 80.0%       | 81.8% | 88.9% |       |       |

AUC: Area Under a Curve  
p value: Probability value  
CI: Confidence Intervals  
LL: Lower limit  
UL: Upper Limit  
NPV: Negative predictive value  
PPV: Positive predictive value  
*: Statistically significant at p ≤ 0.05

Table 10: Accuracy of serum procalcitonin to detect SBP

|               | AUC   | P        | 95% C.I   | Cut off     | Sensitivity | Specificity | PPV  | NPV  | LL    | UL    |
|---------------|-------|----------|-----------|-------------|-------------|-------------|-------|-------|-------|-------|
| Serum procalcitonin | 0.875 | <0.001†  | 0.791     | >0.4ng/ml   | 90.0%       | 70.0%       | 75.0% | 87.5% |       |       |

DISCUSSION

Liver cirrhosis carries a high risk of sepsis because of impairment of both innate and humoral immunity together with bacterial translocation resulting from defective intestinal mucosal barriers. Sepsis in such patients carries very bad prognosis as a consequence of release of inflammatory mediators that may lead to multi-organ dysfunction. This necessitates rapid diagnosis and treatment of SBP. Serum inflammatory markers as PCT and CRP together can be used as rapid diagnostic tool in such patients.

PCT was first discovered in 1975 as a precursor for calcitonin hormone produced by parafollicular cells in thyroid gland. Calcitonin is involved in calcium and phosphorus homeostasis. In healthy individuals all PCT is converted into calcitonin and is not released into the circulation. During sepsis, inflammatory response results in production and release of PCT. It is either produced by a direct or indirect mechanism. The direct mechanism involves induction of hepatocytes and peripheral blood monocytes by toxic metabolites and bacteria lipopolysaccharides. The indirect one is activated by the release of inflammatory mediators such as IL6.

Interleukin 6, a pro-inflammatory cytokine secreted by monocytes and macrophages, requires signal transduction by STAT3/ Jak-kinase pathway to up-regulate acute-phase genes in the liver.

In our study we examined the role of ascitic IL 6 together with other acute phase reactants including Hs-CRP and procalcitonin in early diagnosis of SBP in sixty cirrhotic patients. Twenty percent of the patients (6/30) in group I had culture-negative SBP versus 90 % (27/30) patients of group II, and all of the culture-positive SBP patients (80%, 24/30) had E. colt positive cultures.
In our study, there was no statistical difference between both groups as regards liver enzymes (AST and ALT) and serum LDH. Serum LDH (381.5(213 – 480) vs 320(120 – 394) mmol/l) level was higher in patients in group I compared to patients with sterile cirrhotic ascites in group II, but with no statistical significance, p < 0.051.

We found that serum CRP (68(32 – 98) vs 56.5(12 – 83)) ng/ml level was higher in patients in group I than their levels in patients with sterile cirrhotic ascites in group II, but with no statistical significance, p < 0.096. On the other hand, serum PCT showed statistically significant difference between the two groups being higher in group I compared to patients in group II, p<0.001. The mean serum PCT was 0.8(0.4-1.1) and 0.4(0.04-0.7) ng/ml in groups I and II respectively.

As regards ascitic fluid parameters, significant difference was detected between the two studied groups regarding the degree of ascites. In group I, none of the cases had mild ascites while 70 % patients had moderate ascites and 30 % had tense ascites. In group II, half of the cases had mild ascites, 20 % patients had moderate and the remaining 30% had tense ascites.

Ascitic fluid PMNL was significantly higher in patients with SBP in group I than group II. The mean ascitic fluid protein was much lower in group I (2.2 ± 0.6) g/dl than group II (3.6 ± 0.8) g/dl. Also, we found that there was a statistically significant increase of the level of ascitic IL6 in group I, 2171.5 pg/dl (62.2 – 5000), compared to patients in group II, 342.5(146 – 2567) pg/dl, p<0.001.

The cutoff points for PCT and IL6 to diagnose SBP were >0.4 ng/ml and >780 pg/ml respectively, with sensitivity of 90% for each of them.

In literature there is controversy on the diagnostic value of PCT and CRP in SBP in end stage liver disease. Two meta-analyses, one included three studies by Su et al and the other included seven studies, both found moderate to high accuracy of PCT to rule in or rule out SBP. Our results are comparable to the results reported by Navasa et al and Suliman et al who reported that IL6 both in serum and ascitic fluid was higher in SBP compared to sterile ascites. The latter also concluded that serum and ascitic IL6 correlated with the development of renal impairment and Child-Pugh class in patients with SBP. Another finding reported by Suliman et al was the decline in ascitic IL6 following treatment of SBP.

One study conducted by Ahmed et al found similar findings as our study. It reported that there was significant increase in serum and ascitic IL6 in SBP compared to sterile ascites (553 [432–956] ng/mL vs 195 [95–347] ng/mL; p<0.001). The study also reported statistically significant increase in serum PCT in SBP versus non-SBP. A cutoff value of 0.94 ng/mL, serum PCT had a diagnostic sensitivity of 94.3% and specificity 91.8% for detecting SBP. In our study, the cutoff value of PCT was lower, >0.4 ng/ml, with 90% sensitivity and 70% specificity.

On the other hand, Magdalena et al found no difference in serum and ascitic PCT levels among SBP and non SBP patients which contradicts our results.

In conclusion, the combined use of serum PCT, CRP and ascitic IL6 can be used for early diagnosis of patients with SBP, although further studies with larger number of patients are needed to confirm these findings.

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