Drug Discovery in Spinal Cord Injury With Ankylosing Spondylitis Identified by Text Mining and Biomedical Databases

Chenfeng Wang, Hongdao Ma, Weiqing Wu and Xuhua Lu*

Department of Orthopaedics, Shanghai Changzheng Hospital, Naval Medical University, Shanghai, China

Spinal cord injury (SCI) and ankylosing spondylitis (AS) are common inflammatory diseases in spine surgery. However, it is a project where the relationship between the two diseases is ambiguous and the efficiency of drug discovery is limited. Therefore, the study aimed to investigate new drug therapies for SCI and AS. First, text mining was used to obtain the interacting genes related to SCI and AS, and then, the functional analysis was conducted. Protein–protein interaction (PPI) networks were constructed by STRING online and Cytoscape software to identify hub genes. Last, hub genes and potential drugs were performed after undergoing drug–gene interaction analysis, and MicroRNA and transcription factors regulatory networks were also analyzed. Two hundred five genes common to “SCI” and “AS” identified by text mining were enriched in inflammatory responses. PPI network analysis showed that 30 genes constructed two significant modules. Ultimately, nine (SST, VWF, IL1B, IL6, CXCR4, VEGFA, SERPINE1, FN1, and PROS1) out of 30 genes could be targetable by a total of 13 drugs. In conclusion, the novel core genes contribute to a novel insight for latent functional mechanisms and present potential prognostic indicators and therapeutic targets in SCI and AS.

Keywords: spinal cord injury, ankylosing spondylitis, text mining, drug discovery, bioinformatic analysis

INTRODUCTION

Spinal cord injury (SCI) is a serious complication of traumatic diseases such as spinal fracture or dislocation, burdening families, economics, and society (GBD 2017 US Neurological Disorders Collaborators et al., 2021). On the other hand, ankylosing spondylitis (AS) is a chronic disease that mainly affects the spine and accumulates the sacroiliac and surrounding joints (Braun and Sieper, 2007). However, the relationship between the two diseases is unclear and the efficiency of drug discovery is limited. In addition, if patients with AS happen to have SCI, then the question whether AS aggregates the injury degree of the spinal cord needs to be answered. Therefore, mining the underlying pathomechanism is a necessary way to further understand the relationship between the two diseases and discover potential therapies for destroying their interaction.

There is possibly an inevitable connection between the SCI and AS. The incidence of SCI appears to be higher in patients with AS than in the general population, which, of course, is directly related to the increasing incidence of vertebral column fracture in patients with AS (Jacobs and Fehlings, 2008). Under the pathologic mechanism of the disease, combined with an unstable spine, the AS population is predisposed to highly distracting injuries and spinal epidural hematoma
formation further increases the severity of SCI. Hence, they likely share similar pathogenesis, genes, and antigens.

In recent years, with the development of bioinformatics methods and computational technology, the genetic mechanism behind diseases can be interpreted to investigate deleterious variants associated with target drugs, such as cancers, degenerative diseases, and COVID-19 (Cheng et al., 2016; Woolston et al., 2019; DiNardo et al., 2020; Li et al., 2021). Having been applied in exploring biomarkers, searching for drug targets, and studying molecular regulation, bioinformatics analysis acts as a catalyst to clarify the potential pathogenic agents (Li et al., 2019a; Tolios et al., 2020). Text mining of biomedical literature has been understood as a valid way to reveal the relationship between genes and pathologies while combining with other bioinformatical tools is an effective protocol to verify the existing therapy.

Despite extensive research on SCI and AS, there still is a gap in understanding intersecting genes and latent targets for the treatment of SCI and AS. This study aimed to mine the correlations between the SCI and AS, further obtaining new drugs to control the progression of the two, including text mining, genetic functional analysis, protein–protein interaction (PPI) analysis, drug–gene interaction, and related regulatory molecular analysis. First, common genes were obtained from the SCI and AS preliminary gene lists. Second, interacting genes conducted functional analysis for clarifying related functions and
pathways. Third, PPI analysis is to further screen out core genes and clusters, further purposes drug selection. Last, the hub genes obtained from drug–gene interaction were undergone MicroRNA analysis and transcriptional factors (TFs) regulatory network to explore the potential mechanisms (Figure 1).

**METHODS AND MATERIALS**

**Text Mining**
Pubmed2ensemble, an available website, could be utilized to conduct text mining (http://pubmed2ensemble.ls.manchester.ac.uk/). Relevant kinds of literature and databases are mined in pubmed2ensemble to obtain significant genes. In this study, two queries were performed with “spinal cord injury” and “ankylosing spondylitis”. Two gene lists were generated, and each gene hits were extracted to acquire the common genes for the next analysis.

**Gene Ontology and Signal Pathways Analysis**
The DAVID (https://david.ncifcrf.gov/) was used to conduct gene ontology (GO) and signal pathways annotations for the genes corresponding to the SCI and AS intersection. Biological process, cellular component, and molecular function genes participated in explained functional information while performing pathways analysis with the Kyoto Encyclopedia of Genes and Genomes (KEGG). False discovery rate (FDR) < 0.05 was set as the cutoff.

**Protein–Protein Interaction Analysis and Gene Module Analysis**
First, the STRING database (https://www.string-db.org/), a useful tool to complete functional protein association networks, was selected to analyze the PPI. Second, we uploaded the genes to the site, and the maximum interaction score >0.9 (high confidence) was regarded as an important screening indicator. Third, the TSV file downloaded from the STRING was imported into the Cytoscape software for the Molecular Complex Detection (MCODE), finally building up the gene modules (cluster) to own the hub genes, which were applied to the subsequent steps of drug–gene interactions.

**Drug–Gene Interaction and Functional Analysis**
It was realizable for us to explore the interaction between drugs and genes, which would help us to search for the potential therapy target of SCI and AS. The drug–gene interaction database (https://www.dgidb.org/) is the same as the abovementioned website, open to everyone eager to find effective medicines to treat SCI and AS. The module genes would match with existing drugs whose criteria were obvious interaction type and approved by countries. Besides, final genes after interacting were continued to take part in the process of functional analysis, and FDR of consequences <0.05 was recognized as statistically significant.

**MicroRNA Analysis and TFs Regulatory Network of Hub Genes**
Targetscan (http://www.targetscan.org/) could be utilized to search for the microRNA of hub genes. In addition, TFs of core genes were relying on another online database named NetworkAnalyst (https://www.networkanalyst.ca/), a comprehensive network visual analytics platform for gene expression analysis.

**RESULT**

**Identification of Common Genes**
After eliminating the duplication of gene symbols, 605 genes were related to SCI, 218 genes were related to AS, and 205 common genes were found from text mining with conceptions of “spinal cord injury” and “ankylosing spondylitis”.

**Gene Ontology and Signal Pathways Analysis**
Common 205 genes were participating in the process of GO and KEGG analysis. Consequently, the number of GO terms for the biological process is 1,750, that of the cellular component is 37, and that of molecular function is 31. The five most enriched annotations of the biological process were 1) “response to organic substance” (FDR = 2.85E-35), 2) “response to external stimulus” (FDR = 7.78E-32), 3) “positive regulation of multicellular organismal process” (FDR = 1.62E-31), 4) “inflammatory response” (FDR = 1.81E-31), and 5) “positive regulation of response to stimulus” (FDR = 1.70E-30), respectively, containing 115, 96, 51, 58, and 93 genes from the query set. As for cellular component, the five most enriched annotation were 1) “extracellular space” (FDR = 1.71E-32), 2) “extracellular region part” (FDR = 2.19E-25), 3) “extracellular region” (FDR = 1.07E-24), 4) “cell surface” (FDR = 1.25E-22), and 5) “external side of plasma membrane” (FDR = 6.27E-19), containing 85, 123, 133, 54, and 31 genes. There are also five most enriched annotation of molecular function, 1) receptor binding (FDR = 4.41E-28), 2) cytokine receptor binding (FDR = 1.27E-18), 3) cytokine activity (FDR = 5.97E-17), 4) growth factor activity (FDR = 1.98E-12), and 5) identical protein binding (FDR = 3.12E-8).

In addition, the most enriched annotations of signaling pathways were that 1) Cytokine-cytokine receptor interaction (FDR = 3.17E-20), 2) HIF-1 signaling pathway (FDR = 2.22E-12), 3) NF-kappa B signaling pathway (FDR = 2.46E-11), 4) TNF signaling pathway (FDR = 5.86E-11), 5) Jak-STAT signaling pathway (FDR = 7.66E-8), respectively containing 39, 16, 15, 15, and 17 genes from the query set (Figure 2).

**Module Screening of Protein–Protein Interaction Network**
Meeting the limitation of high confidence and hiding non-interacting genes, candidate genes were selected to construct a PPI network via multiple protein functions of the STRING database. A “tsv” file was exported from the site and then imported into the Cytoscape software for the clustering analysis,
illustrating that there are 142 nodes and 507 edges (Figure 3A). MCODE app, a necessary tool, was used to complete cluster analysis to further screening for the target genes, eventually acquiring various clusters. Cluster 1 constructed by 12 nodes and 86 edges and cluster 2 built up by 18 nodes and 58 edges were chosen to collect important genes (Figures 3B,C).

Potential Therapeutics and Functional Analysis

Drug–gene interaction is the last step to search for the potential signatures and target medicines, investigating existing drugs to alleviate the damaging degree of SCI and AS. Among the PPI and MCODE analysis, a hub gene list that had been eliminated the duplicating part was a channel to the DGIdb (http://www.dgidb.org/), which was consist of 27 various clinical databases and literature, such as DrugBank and NCBI PubMed. With the limitations of presence or absence of approvals and definitive drug–gene interacting type, a sum of 13 drugs was shown in Table 1 and Figure 4, and the interacting score of Cysteamine was highest. Depending on the target drugs analysis result, nine genes (SST, VWF, IL1B, IL6, CXCR4, VEGFA, SERPINE1, FN1, and PROS1), which acted as the core to take part in the occurrence and development of SCI and AS underwent functional analysis (Table 2), which clarified HIF-1 signaling pathway and PI3K-Akt signaling pathway played an important role in the progression of the two diseases.

MicroRNA and TF Regulatory Network

Nine hub genes were corresponding to one or two conserved MircoRNA shown in Figure 4. The TFs regulatory network consisted of nine genes and 122 TFs (Figure 5). CXCR4 was regulated by 47 TFs, FN1 was regulated by 46 TFs, VEGFA was found to be regulated by 39 TFs, and SERPINE1 was regulated by 36 TFs. The numbers of VWF, IL6, PROS1, SST, and IL1B were 29, 25, 25, 18, and 15, respectively. In addition, 11 TFs were found with an interacting degree >4 in the network, implying they may play a key role in the progress of diseases (Table 3).

DISCUSSION

Leading to lifelong disability, SCI, a devastating disease, is damage to the spinal cord. As for AS, the autoimmune disease is characterized by inflammation of the sacroiliac joints and the attachment points of the spine. According to the study outcomes, patients with AS are more likely to fracture and develop SCI. However, there has been little or no research into drug discovery for SCI and AS diseases treatment. Therefore, we designed this study to identify the potential drugs of therapeutic value, further providing references for clinical diagnosis and treatment. These 13 drugs target the following nine genes: VEGFA (four drugs), IL1B (two drugs), SST, VWF, IL6, CXCR4, SERPINE1, FN1, and PROS1 (one drug each).

CXCR4 (C-X-C motif chemokine receptor 4), a member of the GPCR family, and CXCL12 constitute an axis to modulate the nerve system. The expression of the CXCL12/CXCR4 axis was increasing in the patients with SCI and AS (Knerlich-Lukoschus et al., 2011; Tysseling et al., 2011). Besides, a study found that CXCR4 could accelerate the ossification of fibroblasts in patients with AS (He et al., 2019). FN1 (fibronectin 1) is involved in the process of cell adhesion and migration (Cai et al., 2018) and could rely on the TGF-β/PI3K/Akt pathway to promote chondrocyte differentiation and collagen production in fractured bones (Zhang et al., 2021). As for PROS1 (protein s), one of its roles is to inhibit tumor metastasis by mediating inflammation and immunization (Maimon et al., 2021). Downregulated SERPINE1 (serpin family E member 1) could accelerate the progression of inflammation (Yang et al., 2017).
When it comes to the VWF (von Willebrand factor) and VEGFA (vascular endothelial growth factor), some reports have revealed the relationship between the two genes and the two diseases (Widenfalk et al., 2003; Divecha et al., 2005; Nesic et al., 2010; Taylan et al., 2012). Experiencing compressive injury, the levels of VWF and VEGFA expression in the rat spinal cord were decreased. Interestingly, the levels of two gene expressions were dramatically increased after decompression (Cheng et al., 2021), which was consensus with another study (Shen et al., 2020). Furthermore, VWF production in men with AS was more than...
TABLE 1 | Potential drugs targeting genes with SCI and AS association.

| Number | Drug        | Gene   | Type                  | Score | Approved? | PMID      |
|--------|-------------|--------|-----------------------|-------|-----------|----------|
| 1      | Afibercept  | VEGFA  | Inhibitor, Binder     | 2.35  | Yes       | 22813448 |
| 2      | Bevacizumab | VEGFA  | Inhibitor             | 1.02  | Yes       | 18182667 |
| 3      | Canakinumab | IL1B   | Inhibitor, Binder     | 10.34 | Yes       | 19169963 |
| 4      | Caplacizumab| VWF    | Inhibitor             | 13.67 | Yes       | None     |
| 5      | Cysteamine  | SST    | Binder                | 18.23 | Yes       | 2853942  |
| 6      | Menadione   | PROS1  | Activator             | 1.14  | Yes       | 12033454 |
| 7      | Ocrilasin   | FN1    | Inhibitor             | 1.29  | Yes       | 23193358 |
| 8      | Pegaptanib sodium | VEGFA | Antagonist          | 3.36  | Yes       | 23953100 |
| 9      | Plerixafor  | CXCR4  | Antagonist, Agonist   | 8.93  | Yes       | 17715128 |
| 10     | Ranibizumab | VEGFA  | Inhibitor             | 8.81  | Yes       | 18046235 |
| 11     | Rilonacept  | IL1B   | Inhibitor, Binder     | 3.45  | Yes       | 23319019 |
| 12     | Siltuximab  | IL6    | Antagonist            | 10.21 | Yes       | 8823310 |
| 13     | Urokinase   | SERPINE1 | Inducer, Substrate  | 3.19  | Yes       | 12709915 |

Each drug–gene interaction ensured that the hypothetical drug had an expected effect on the condition. The link to the source was tracked to confirm the report and evaluate related metadata such as approval status and available route drug use. Drugs that targeted the candidate genes through appropriate interactions were collected in the final list.

*aThe score is the combined number of database sources and PubMed references.*

FIGURE 4 | The interactions of hub genes with drugs and MicroRNA. The interaction of nine hub genes with 13 drugs and MicroRNA.
normal people, and it is likely to be recognized as a biomarker of AS (Taylan et al., 2012). In addition, a study found that early VEGF delivery for SCI treatment attributed to its power of protecting vessels and inhibiting cell apoptosis was advantageous (Widenfalk et al., 2003), and it even serves as an important criterion for assessing SCI (Nesic et al., 2010).

### TABLE 2 | Summary of gene set enrichment analysis.

| Category                     | Term                          | Count | FDR         | Genes                                           |
|------------------------------|-------------------------------|-------|-------------|-------------------------------------------------|
| Biological process           | Leukocyte migration           | 7     | 4.83E-06    | IL6, IL1B, PROS1, SERPINE1, FN1, CXCR4, VEGFA   |
| Biological process           | Platelet degranulation        | 5     | 5.39E-05    | VWF, PROS1, SERPINE1, FN1, VEGFA               |
| Biological process           | Cell motility                 | 8     | 5.35E-05    | IL6, SST, IL1B, PROS1, SERPINE1, FN1, CXCR4, VEGFA |
| Biological process           | Localization of cell          | 8     | 5.35E-05    | IL6, SST, IL1B, PROS1, SERPINE1, FN1, CXCR4, VEGFA |
| Biological process           | Locomotion                    | 8     | 1.73E-04    | IL6, SST, IL1B, PROS1, SERPINE1, FN1, CXCR4, VEGFA |
| Cellular component           | Platelet alpha granule lumen  | 5     | 5.73E-07    | VWF, PROS1, SERPINE1, FN1, VEGFA               |
| Cellular component           | Platelet alpha granule        | 5     | 1.02E-06    | VWF, PROS1, SERPINE1, FN1, VEGFA               |
| Cellular component           | Secretory granule lumen       | 5     | 1.18E-06    | VWF, PROS1, SERPINE1, FN1, VEGFA               |
| Cellular component           | Cytoplasmic membrane-bound vesicle lumen | 5     | 1.65E-06    | VWF, PROS1, SERPINE1, FN1, VEGFA               |
| Cellular component           | Vesicle lumen                 | 5     | 1.65E-06    | VWF, PROS1, SERPINE1, FN1, VEGFA               |
| Molecular function           | Receptor binding              | 7     | 0.001232431 | IL6, VWF, SST, IL1B, SERPINE1, FN1, VEGFA      |
| Molecular function           | Protease binding              | 3     | 0.045365023 | VWF, SERPINE1, FN1                              |
| Molecular function           | Growth factor receptor binding| 3     | 0.045365023 | IL6, IL1B, VEGFA                               |
| KEGG pathway                 | Complement and coagulation cascades | 3     | 0.046494339 | VWF, PROS1, SERPINE1                           |
| KEGG pathway                 | HIF-1 signaling pathway       | 3     | 0.046494339 | IL6, SERPINE1, VEGFA                           |
| KEGG pathway                 | PI3K-Akt signaling pathway    | 4     | 0.046494339 | IL6, VWF, FN1, VEGFA                           |

With a strict level, a p-value cutoff was set. Among the most importantly enriched biological process, cellular component, molecular function, and KEGG pathways above the cutoff, those most relevant to SCI and AS pathology were chosen from the researches and literature.

*FDR correction was performed to control for the false positive.

![FIGURE 5](image-url) The interactions of hub genes with TFs. The TFs regulatory network of nine hub genes. Red nodes represented nine hub genes and blue squares represented TFs.
GO analysis and KEGG enrichment analysis of DEGs showed that most DEGs were closely related to inflammatory response, response to external stimulus and response to organic substance. PI3K/Akt and HIF-1 are the major signaling pathways that have been identified to be significant in inflammatory diseases. PI3K/Akt signaling pathway is inhibited by endoplasmic reticulum stress after SCI, which is activated to realize motor recovery and nerve protection (Li et al., 2019b). In addition, owning the function of regulating spinal cord regeneration, PI3K/Akt is regarded as a potential target for SCI treatment (Lu et al., 2018). Concerning the HIF-1 signaling pathway, if combined with VEGF, then it is activated, secondary SCI will be inhibited, and local hypoxic ischemia environment will be improving, meanwhile promoting neurological recovery (Chen et al., 2017). Furthermore, the HIF-1 signaling pathway that can also be explored to prevent SCI involves the apoptosis of neuronal cells (Wang et al., 2016). On the basis of the previous studies, HIF-1 activity relies on the HIF-1α that is particularly controlled by the PI3K/Akt signaling pathway at the post-transcriptional protein level (DeBerardinis et al., 2008; Zhang et al., 2018).

The top five on the drug–gene interacting scoring system were Cysteamine (SST binder), Caplacizumab (VWF inhibitor), Canakinumab (IL1B inhibitor), Siltuximab (IL6 antagonist), and Plerixafor (CXCR4 inhibitor). Owning the highest interacting score in these drugs, Cysteamine deserves to be more explored for its therapeutic value for SCI patients with AS, which is mainly applied to cure Cystinosis (Hollywood et al., 2020), atherosclerosis (Wen et al., 2019), and Parkinson’s disease (Cicchetti et al., 2019). Caplacizumab is only used for acquired thrombotic thrombocytopenic purpura (Scully and Spero, 2019), whereas Siltuximab is mostly utilized for Castleman’s disease (van Rhee et al., 2014). Besides, Canakinumab has entered the clinical stage of lung cancer treatment, perhaps becoming the candidate for SCI or AS (Wong et al., 2020). On the basis of the existing researches, inhibition of CXCR4 could alleviate inflammatory pain induced by ischemia–reperfusion (Li et al., 2016).

More than a quarter of identified drugs target VEGFA, including Afibercept, Bevacizumab, Pegaptanib sodium, and Ranibizumab. Bevacizumab approved by the US Food and Drug Administration (FDA) has been used in the management of neurosurgery diseases such as radiation-induced myelopathy and gliomas (Psimaras et al., 2016; Levenbaum et al., 2019). Surprisingly, some drugs (Afibercept, Pegaptanib sodium, and Ranibizumab) for the therapy of age-related macular degeneration may also be a benefit for SCI patients with AS. As mentioned before, VEGFA owns the function of inducing angiogenesis, indicating that it plays a key role in the SCI and AS. Although the above drugs are not currently used for AS and SCI, they are approved by the FDA and may be possibly approved in the therapy of the two diseases in the future.

Interestingly, it is worth noting that STAT3 (signal transducer and activator of transcription 3), as the most connected TF, and MicroRNAs, respectively, corresponding to final genes play a significant role in the development of AS and SCI. The expression of STAT3 is high in patients with AS as well as SCI patients, so AS can further aggregate the degree of SCI and induce more severe inflammatory storms, finally influencing the prognosis of patients (Jo et al., 2020). Meantime, TFs and MicorRNA can also regulate the hub genes to mediate inflammations (Qiu et al., 2021). For example, MicroRNA-149 could suppress hepatic inflammatory response through antagonizing STAT3 signaling pathway (Zhang et al., 2017). Besides, MicroRNA-183 regulates LPS-induced oxidative stress by targeting FNI (Xie et al., 2020) and MicroRNA-140-3p can ameliorate the progression of osteoarthritis via targeting CXCR4 (Ren et al., 2020). Therefore, MicorRNA and TFs may act as important targets for the treatment of SCI patients with AS, providing direction for later researches.

All consequences suggested that common genes are not only responsible for the SCI and AS but also cover a wide range of digestive system neoplasm and blood system via cytokine–cytokine interaction, PI3K/Akt signaling pathway, and HIF-1 signaling pathway. What calls for special attention is that those alternating genes can aggravate the severity of the SCI and AS, thus building up a vicious circle, and there is also a potential risk of tumor formation. Therefore, it is important to give early interventions for SCI patients with AS through potential drugs therapy. On the one hand, the progression of SCI and AS can be postponed by early treatment, finally breaking the vicious circle. On the other hand, tumors will not be given a chance to develop if intervention is used in the early phase, ultimately maintaining body physical activity to promote recovery.

Regrettably, the limitation of this study is that, with the update of the database we used, our study needs to catch up with the development to repeat the study. In addition, the criteria of screening out core genes are subjective, implicating objective criteria need to be constructed to identify the candidate.

| TFs | Genes | Count |
|-----|-------|-------|
| STAT3 | SST, VWF, IL1B, IL6, CXCR4, VEGFA, SERPINE1, FN1, PROS1 | 9 |
| SOX2 | IL6, CXCR4, VEGFA, SERPINE1, FN1, PROS1 | 6 |
| MYC | VWF, IL6, CXCR4, VEGFA, FN1 | 5 |
| SMAD4 | IL1B, CXCR4, VEGFA, SERPINE1, FN1 | 5 |
| TRIM28 | IL6, VEGFA, SERPINE1, FN1, PROS1 | 5 |
| SALL4 | IL1B, IL6, CXCR4, FN1, PROS1 | 5 |
| GATA2 | VWF, IL1B, IL6, SERPINE1, FN1 | 5 |
| SP1 | SST, IL1B, IL6, SERPINE1, PROS1 | 5 |
| ESR1 | VWF, VEGFA, SERPINE1, FN1, PROS1 | 5 |
| HNF4A | VWF, IL1B, VEGFA, SERPINE1, FN1 | 5 |
| TP53 | VWF, IL1B, CXCR4, VEGFA, PROS1 | 5 |
CONCLUSION
In conclusion, our study obtained 205 genes common in both SCI and AS. GO and KEGG analyses of intersecting genes might reveal a novel prospective relationship between SCI and AS. Consequently, the final nine genes (SST, VWF, IL1B, IL6, CXCR4, VEGFA, SERPINE1, FN1, and PROS1) and 13 drugs may provide therapeutic value for SCI patients with AS.

DATA AVAILABILITY STATEMENT
The original contributions presented in the study are included in the article/Supplementary Material. Further inquiries can be directed to the corresponding author.

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AUTHOR CONTRIBUTIONS

CW designed the experiments, analyze the data, and wrote the manuscript. HM revised the manuscript. WW designed the experiments and analyze the data. XL devised the concept, designed the research, supervised the research, and wrote the manuscript. All authors read and approved the final manuscript.

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