DeepInsight: A methodology to transform a non-image data to an image for convolution neural network architecture

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It is critical, but difficult, to catch the small variation in genomic or other kinds of data that differentiates phenotypes or categories. A plethora of data is available, but the information from its genes or elements is spread over arbitrarily, making it challenging to extract relevant details for identification. However, an arrangement of similar genes into clusters makes these differences more accessible and allows for robust identification of hidden mechanisms (e.g. pathways) than dealing with elements individually. Here we propose, DeepInsight, which converts non-image samples into a well-organized image-form. Thereby, the power of convolution neural network (CNN), including GPU utilization, can be realized for non-image samples. Furthermore, DeepInsight enables feature extraction through the application of CNN for non-image samples to seize imperative information and shown promising results. To our knowledge, this is the first work to apply CNN simultaneously on different kinds of non-image datasets: RNA-seq, vowels, text, and artificial.

In the post-genomic era, though an abundance of data is accessible, the information is indiscriminately spread over across high dimensional data space, making it challenging to differentiate phenotypes. The same problem of associating relevant features towards a class label lies for other kinds of data (e.g. vowels, text). It becomes critical to arrange elements in an appropriate manner which can enable extraction of relevant features for analyses. Accordingly, the arrangement of information turns to be an important phase via sorting and positioning of the elements in the right order for the subsequent step. We refer this phase as the element arrangement step. The identification or classification of phenotypes or class labels can conceivably be improved following the three steps: element arrangement, feature extraction and developing a suitable classifier.

The conventional machine learning (ML) techniques for classification or detection problem, requires a sample in the form of a feature vector (i.e., a column vector of size $p \times 1$). This feature vector obtained from a feature extraction technique is processed to be categorized into one of the defined groups. The features in this vector form are generally considered mutually independent (particularly in the order of appearance) by ML techniques. Consequently, changing the order of features bears no direct impact in classification or phenotype detection, which makes the element arrangement step redundant for many state-of-the-art ML classifiers like random forest\textsuperscript{1,2} and decision trees\textsuperscript{3}. However, the reliability of ML techniques is dependent on the feature extraction technique.

On the other hand, convolution neural network (CNN) architecture from deep neural networks accepts a sample as an image (i.e. a matrix of size $m \times n$) and performs feature extraction and classification via hidden
layers (such as convolutional layers, RELU layer, max-pooling layers). It does not require additional feature extraction techniques as it automatically derives features from the raw elements. The second advantage is that it finds higher-order statistics of image and nonlinear correlations. Third, convolutions neurons process data for its location. This method is perhaps the first one of converting gene expression into image samples and applying for RNA-seq data by first performing gene selection followed by constructing an image based on chromosome location. This method is perhaps the first one of converting gene expression into image samples and applying for RNA-seq data by first performing gene selection followed by constructing an image based on chromosome location.

The description of these datasets is as follows. The first is an RNA-seq or gene expression dataset which is 60483 genes or dimensions. This is a 10-class dataset, representing ten types of cancer. The second is a speech dataset from the TIMIT corpus\textsuperscript{24,25}. Here a set of 10 distinct monophthong vowels are extracted, then each vowel is subdivided into three segments, and each segment is used to generate mel-frequency cepstral coefficients.
with energy-delta-acceleration (MFCC_EDA) feature vectors. A total of 12579 samples with 39 dimensions are obtained. The third dataset is Relathe (text) which is derived from newsgroup documents and partitioned evenly across different newsgroups. It contains 1427 samples and 4322 dimensions. It is a two class problem. The next two are artificial datasets. One is Madelon which has 2600 samples and 500 dimensions. This is a two-class classification problem with continuous input variables. It is multivariate and highly non-linear. The other is ringnorm-DELVE, which is an implementation of Leo Breiman’s ringnorm example. It is 20 dimensional, 2 class classification with 7400 samples. Each class is drawn from a multivariate normal distribution, where class 1 has zero mean and covariance four times the identity, and class 2 has a mean $a = 2/\sqrt{20}$ with unit covariance.

These datasets are summarized in Table 1.

### Comparison and classification performance.

The existing state-of-the-art classifiers such as random forest, decision tree, and ada-boost were employed for comparison purposes. The hyperparameters of competing methods were optimized using grid search optimization. As discussed in Section 4.3 and Supplement File 1, the DeepInsight method employs two types of normalizations (norm-1 and norm-2), and the validation error is evaluated on both these norms. The norm which gives the lowest validation error is used for further processing. The pixel frame size is fixed at $120 \times 120$. However, for RNA-seq dataset, the analysis was done on $200 \times 200$ pixel size since the number of elements or features is very large (60483) resulting in lossy compression (as discussed in Supplement File 2) compared to other datasets studied in this work. The validation errors for both the norms are depicted in Supplement File 3 for all the datasets after executing DeepInsight. The best fit model on the validation set is used to evaluate performance on a separate test set.

The purpose of this comparison is to show that DeepInsight can also produce a competitive performance on different kinds of datasets. The performance regarding classification accuracy is depicted in Table 2 (see Supplementary File 4 for a brief discussion on codes).

| Datasets          | Decision Tree | Ada-Boost | Random Forest | DeepInsight |
|-------------------|---------------|-----------|---------------|-------------|
| RNA-seq            | 85%           | 84%       | 96%           | 99%         |
| Vowels             | 75%           | 45%       | 90%           | 97%         |
| Text               | 87%           | 85%       | 90%           | 92%         |
| Artificial (Madelon) | 65%      | 60%       | 62%           | 88%         |
| Artificial (Ringnorm-DELVE) | 90%   | 93%       | 94%           | 98%         |
| Average            | 80%           | 73%       | 86%           | 95%         |

Table 2. Classification accuracy on different kinds of datasets using various models.

As anticipated, the proposed DeepInsight method produced very promising results. The obtained results enable us to use CNN architecture for various kinds of non-image datasets. This increases the possibility of utilizing deep learning networks. One can envisage the immense possibility of applying this algorithm to a wide variety of applications.

In this work, we were able to integrate many properties of CNN for non-image samples through the inception of DeepInsight method. A non-image sample, in the form of vectors were transformed into meaningful images for the processing of CNN. This strategy does not solve all problems related to genomic data, however, it is a step forward in integrating the merits of CNN. Deep neural network architectures, encompass many advantages:
feature extraction, dimension reduction, finding hidden structure from sparse and hyper-dimensional data, data augmentation and up-sampling, semi-supervised learning with labeled/non-labeled samples, and optimum action selection with time-series data. Therefore, for a broader context, deep neural network architectures have the potential to offer a solution for genomic analysis for a variety of input samples ranging from DNA sequences to protein sequences (which may be considered as time-series data). The DeepInsight method increases the versatility of CNN architectures. The characteristics of CNN such as automatic feature extraction, reducing the need of neurons and consequently enabling to train a model much deeper, weight sharing capability to mitigate memory requirement, utilization of neighborhood information (i.e., processing subarea of pixel frame at a time), and, GPU utilization make CNN a potent tool for classification and analysis. These attributes of CNN are utilized for non-image cases by the proposed technique. Further, we have shown the effectiveness of DeepInsight on several kinds of datasets and obtained very promising results. For RNA-seq data the maximum classification accuracy achieved by DeepInsight was 99%. For vowels, text, Madelon and ringnorm the accuracies were 97%, 92%, 88% and 98%, respectively.

Further extensions of the current version of the algorithm can be considered. The present technique employs gray-scale or single layer (i.e., 2D matrix) for classification. This can be extended to incorporate multiple layers and therefore can be applied to solve problems related to multi-omics data (e.g., gene-expression, methylation, mutation) as well. Moreover, different kinds of data (e.g., clinical and non-clinical) can be normalized into a single layer (if multi-layer is prohibited due to computing resources) for analysis and classification. This technique can be useful for a number of applications where data is not in image form.

**Methods**

**DeepInsight method.** The concept of DeepInsight is to first transform a non-image sample to an image form and then supply it to the CNN architecture for the prediction or classification purpose. A simple illustration is given in Fig. 1a, where a feature vector $x$ consisting of gene expression values is transformed to a feature matrix $M$ by a transformation $T$. The location of features in the Cartesian coordinates depends on the similarity of features. For example, features $g_1$, $g_3$, $g_6$ and $g_d$ are closer to each other in Fig. 1a. Once the locations of each feature are determined in a feature matrix, then the expression values or feature values are mapped. This will generate a unique image for each sample (or feature vector). $N$ samples of $d$ features will provide $N$ samples of $m \times n$ feature.
matrices. This 2D matrix form will have all the \( d \) features. Thereafter, this set of \( N \) feature matrices are processed to the CNN architecture for learning the model and providing prediction.

If the data dimensionality is extremely large and difficult to handle due to hardware limitations, then the dimensionality reduction technique (DRT) may be considered before applying DeepInsight. The DRT can be either in the form of feature selection or feature extraction depending upon the nature of the problem. The application of DRT will provide a small feature set which will help in faster processing, however, can risk classification performance. On the other hand, if noisy or redundant features are removed then it could help to get higher processing speed as well as better accuracy. Since the application of DRT is case dependent, we have described DeepInsight without applying DRT.

**DeepInsight pipeline.** In this section, we briefly discuss the transformation of a non-image sample to its image form. A general overview of this transformation is depicted in Fig. 1b. The training set is used to find the location of features. If the training set consisting of \( n \) samples is defined as \( \chi = \{x_1, x_2, \ldots, x_n\} \) where a feature vector has \( d \) features or \( x \in \mathbb{R}^d \), then we can also define a gene or feature set \( G = \{g_1, g_2, \ldots, g_d\} \) where \( g \in \mathbb{R}^n \); i.e., a feature \( g_j \) has \( n \) training samples. Basically, \( G \) can be obtained by transposing \( \chi \). We used this feature set \( G \) and applied similarity measuring technique or dimensionality reduction technique like t-SNE or kernel principal component analysis (kPCA) to obtain a 2D plane (please see details about t-SNE and kernel PCA in Supplement File 5. These are non-linear dimensionality reduction techniques. A number of linear dimensionality reduction techniques also exist but not implemented in this work). The points in this Cartesian plane are the features or genes. These points only define the location of features, not the feature itself or expression values. Once the location of features is defined, the convex hull algorithm is used to find the smallest rectangle containing all the
points. Since the image should be framed in a horizontal or vertical form for the CNN architecture, a rotation is performed. Thereafter, the Cartesian coordinates are converted to pixels. The conversion from Cartesian coordinates to pixel frames is done by averaging some features as the image size has a pixel limitation. The pixel frame will, therefore, consist of the positions of features (or genes) for a sample \( x_i \) (for \( j = 1, 2, \ldots, n \)). Once the location is determined, the next step is to map the feature (or gene expression) values to these pixel locations. If more than one feature acquired the same location in the pixel frame, then, during mapping of the features, the respective features will be averaged and placed in the same location. Therefore, if the resolution of image or grid size is very small (compared to the number of features given), then many features overlap with each other and image representation may not be very accurate. An appropriate resolution should be selected given the hardware capacity and the number of features required to process. Alternatively, dimensionality reduction may be applied a priori. The details about the procedure is given in Supplementary File 6.

**Feature normalization.** The single layer of the image has 256 shades which are normalized in the range of [0, 1]. Therefore, feature values are to be normalized before applying the image transformation. In this work, we performed two types of normalizations: (1) each feature is assumed independent and therefore normalized by its minimum and maximum, and (2) the topology of mutual features are retained up to some extent by normalizing it with the one maximum value from the entire training set. These normalizations are explained in detail in the Supplementary File 1. DeepInsight evaluates validation set performance on both the types of normalizations and accepts the one with the lowest validation error.

**CNN architecture.** In this section, we describe the CNN architecture of the DeepInsight method. Once a feature vector is transformed into an image, it can then be further processed to the CNN architecture (an illustration of two types of cancer samples in image form is shown in Fig. 2a).

We developed a parallel CNN architecture so that different filter-sizes can be effectively used to train the model. Our CNN architecture is shown in Fig. 2b. In this architecture, we have four layers in parallel where each layer consists of a 2D convolution layer, a batch normalization layer, a ReLU activation layer, and a max pooling layer. Batch normalization is used to prevent overfitting during training, and the max pooling layer is used to down-samples the size of the image in each layer. The outputs of the fourth convolution layer (in the parallel architecture) are combined and fed to a fully connected layer. Finally, a SoftMax layer is used to give the output as class labels.

The CNN architecture of DeepInsight has various hyperparameters such as convolution layers, filter sizes, learning rate and so on. We tuned these hyperparameters by applying Bayesian optimization technique for all the trials. We obtained a set of hyperparameters that gave the best performance on the validation set. The parameter details and validation error during the training phase are discussed in Supplementary File 2 and Supplement File 3.

Once the CNN model is trained using the optimal hyperparameters, then any novel sample can be identified into one of the categories or classes.

For an illustration, two samples are derived from distinct types of cancers, texts and vowels to observe the difference between samples. The transformed samples by DeepInsight method are shown in Fig. 3. This method provides interesting localities by performing element arrangement, then feature dissimilarity is further captured by feature extraction and classification through the application of CNN. Moreover, these samples can now be visualized, and their relative difference in particular regions might lead to different class labels (or phenotypes).

**Data Availability**

RNA-seq data is available from TCGA (https://cancergenome.nih.gov). Vowels data can be extracted from TIMIT Acoustic-Phonetic Continuous Speech Corpus (https://catalog.ldc.upenn.edu/LDC93S1). Text data is available from http://featureselection.asu.edu/datasets.php. Madelon dataset is available from UCI repository http://archive.ics.uci.edu/ml/datasets/madelon, and ringnorm dataset is available from University of Toronto at https://www.cs.toronto.edu/~delve/data/ringnorm/desc.html.

**Code Availability**

All the source code, a sample dataset and documentaries are available at http://www.riken.jp/en/research/labs/ims/med_sci_math/ or http://www.alok-ai-lab.com.

**References**

1. Ho, T. K. In 3rd International Conference on Document Analysis and Recognition 278–282 (Montreal, QC, 1995).
2. Ho, T. K. The Random Subspace Method for Constructing Decision Forests. IEEE Transactions on Pattern Analysis and Machine Intelligence 20, 832–844 (1998).
3. Rokach, L. & Maimon, O. Data mining with decision trees: theory and applications. (World Scientific Pub Co Inc, 2008).
4. Habibi, A. H. & Jahani, H. E. Guide to Convolutional Neural Networks: A Practical Application to Traffic-Sign Detection and Classification. (Springer International Publishing, 2017).
5. Lecun, Y., Bottou, L., Bengio, Y. & Haffner, P. Gradient-based learning applied to document recognition. Proceedings of the IEEE 86, 2278–2324 (1998).
6. Krizhevsky, A., Sutskever, I. & Hinton, G. E. In Proceedings of the 25th International Conference on Neural Information Processing Systems - Volume 1 1097–1105 (Lake Tahoe, Nevada, 2012).
7. Simonyan, K. & Zisserman, A. Very Deep Convolutional Networks for Large-Scale Image Recognition. arXiv preprint arXiv 1409, 1556 (2014).
8. He, K., Zhang, X., Ren, S. & Sun, J. In 2016 IEEE Conference on Computer Vision and Pattern Recognition (CVPR) 770–778 (Las Vegas, NV, 2016).
9. Yan, M. L., Chen, Q. & Shuicheng. Network In Network. In 2015 IEEE Conference on Computer Vision and Pattern Recognition (CVPR) 1–9 (Boston, MA, 2015).
12. Sermanet, P. et al. In International Conference on Learning Representations (ICLR2014), CBLS (2014).
13. Redmon, J., Divvala, S. K., Girshick, R. B. & Farhadi, A. In IEEE Conference on Computer Vision and Pattern Recognition (CVPR) 779–788 (2016).
14. Girshick, R., Donahue, J., Darrell, T. & Malik, J. In IEEE Conference on Computer Vision and Pattern Recognition (CVPR) 779–788 (2016).
15. Girshick, R. In IEEE International Conference on Computer Vision (ICCV) 1440–1448 (Santiago, Chile, 2015).
16. Ren, S., He, K., Girshick, R. & Sun, J. In Advances in Neural Information Processing Systems 91–99 (2015).
17. Singh, R., Lanchantin, J., Robins, G. & Qi, Y. DeepChrome: deep-learning for predicting gene expression from histone modifications. Bioinformatics 32, 639–648 (2016).
18. Liu, J., Wang, X., Cheng, Y. & Zhang, L. Tumor gene expression data classification via sample expansion-based deep learning. OncoTarget 8, 109646–109660 (2017).
19. Zeng, T., Li, R., Mukamala, R., Ye, J. & Ji, S. Deep convolutional neural networks for annotating gene expression patterns in the mouse brain. BMC Bioinformatics 16 (2014).
20. Gao, X., Zhang, J., Wei, Z. & Hakonarson, H. DeepPolyA: A Convolutional Neural Network Approach for Polyadenylation Site Prediction. IEEE Access 6, 24340–24349 (2018).
21. Xu, J. et al. In IJCAI’15 Proceedings of the 24th International Conference on Artificial Intelligence 1369–1375 (Argentine, 2015).
22. Zhang, X., Zhao, J. & LeCun, Y. In NIPS’15 Proceedings of the 28th International Conference on Neural Information Processing Systems - Volume 1 (Montreal, Canada, 2015).
23. Lyu, B. & Haque, A. Deep Learning Based Tumor Type Classification Using Gene Expression Data, https://www.biorxiv.org/content/early/2018/07/11/364323 (2018).
24. Garofolo, J. S. et al. (Philadelphia: Linguistic Data Consortium, 1993).
25. Young, S. et al. The HTK Book (for HTK Version 3.2). (Cambridge University Engineering Department, 2002).
26. Mitchell, T. M. Machine Learning. (McGraw-Hill, Inc., 1997).
27. Guyon, I., Gunn, S., Nikravesh, M. & Zadeh, L. A. Feature Extraction: Foundations and Applications. (Springer-Verlag, 2006).
28. Breiman, L. Bias, Variance, And Arcing Classifiers. (Technical Report 460, Statistics Department, University of California, 2000).
29. Goodfellow, I., Bengio, Y., Courville, A. & Bach, F. Deep Learning. (The MIT Press, 2016).
30. Maaten, L. J. P. V. D. & Hinton, G. Visualizing High-Dimensional Data using t-SNE. Journal of Machine Learning Research 9, 2579–2605 (2008).
31. Sharma, A. & Paliwal, K. A new perspective to null linear discriminant analysis method and its fast implementation using random matrix multiplication with scatter matrices. Pattern Recognition 45(6), 2205–2213 (2012).
32. Sharma, A. & Paliwal, K. A gradient linear discriminant analysis for small sample sized problem. Neural Processing Letters 27(1), 17–24 (2008).
33. Sharma, A. & Paliwal, K. Linear discriminant analysis for the small sample size problem: an overview. International Journal of Machine Learning and Cybernetics 6(3), 443–454 (2015).
34. Sharma, A. & Paliwal, K. F. Principal component analysis using fixed-point algorithm. Pattern Recognition Letters 28(10), 1151–1155 (2007).

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A.S. perceived, developed and wrote the first draft of the manuscript, E.V. prepared figures and contributed in writing, D.S. and K.A.B. collected the samples, T.T. perceived and provided resources. All authors read and approved the final manuscript.

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