EPAS1 variants in high altitude Tibetan wolves were selectively introgressed into highland dogs

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ABSTRACT

Background. Admixture can facilitate adaptation. For example, black wolves have obtained the variant causing black coat color through past hybridization with domestic dogs and have higher fitness than gray colored wolves. Another recent example of the transfer of adaptive variation between the two species has been suggested by the similarity between high altitude Tibetan mastiffs and wolves at the EPAS1 gene, a transcription factor induced in low oxygen environments.

Methods. Here, we investigate the directionality of admixture in EPAS1 between 28 reference highland gray wolves, 15 reference domestic dogs, and 21 putatively admixed highland wolves. This experimental design represents an expanded sample of Asian dogs and wolves from previous studies. Admixture was inferred using 17,709 publicly available SNP genotypes on canine chromosome 10. We additionally conducted a scan for positive selection in the highland dog genome.

Results. We find an excess of highland gray wolf ancestry at the EPAS1 locus in highland domestic dogs, suggesting adaptive introgression from wolves to dogs. The signal of admixture is limited in genomic extent to a small region on chromosome 10, indicating that it is the focus of selection in an oxygen-limited environment.

Discussion. Our results suggest that an adaptive variant of EPAS1 in highland wolves was transferred to highland dogs, carrying linked variants that potentially function in hypoxia response at high elevation. The intertwined history of dogs and wolves ensures a unique evolutionary dynamic where variants that have appeared in the history of either species can be tested for their effects on fitness under natural and artificial selection. Such coupled evolutionary histories may be key to the persistence of wild canines and their domesticated kin given the increasing anthropogenic modifications that characterize the future of both species.

INTRODUCTION

A novel mechanism that can promote accelerated adaptation is the spread of beneficial alleles through admixture between closely related species. Admixed individuals often display
novel gene combinations that can be tested by natural selection in a distinct ecological context (Arnold, 2016) or facilitate expansion into new ecological niches enhancing evolutionary persistence (Becker et al., 2013; Hedrick, 2013). Though adaptation may occur more quickly from standing variation, admixture is the only process that can transfer entire co-adapted multi-gene complexes (Hedrick, 2013). Examples of adaptive introgression include rodenticide warfarin resistance in mice (Song et al., 2011), black coat color in wolves (Anderson et al., 2009), beak morphology in Darwin’s finches (Grant & Grant, 2010), immunity genes in humans (Mendez, Watkins & Hammer, 2012a; Mendez, Watkins & Hammer, 2012b; Abi-Rached et al., 2011) and collar color in manikins (MacDonald et al., 2001).

Recently, the genetic basis underlying adaptation to high altitude in Tibetans was elucidated. Although several genes seem to be involved in the response, EPAS1 showed the strongest signal of selection in Tibetans (Beall et al., 2010; Simonson et al., 2010; Peng et al., 2011; Xu et al., 2011), and was obtained through adaptive introgression from Denisovan-like ancestors (Huerta-Sánchez et al., 2014). EPAS1 (or HIF2α) is a transcription factor induced in oxygen-limited (hypoxic) environments which dampens the erythropoietic response and is under natural selection (Beall et al., 2010). Previously, we showed EPAS1 experienced strong positive selection in highland Chinese wolves as indicated by divergent allele frequencies at non-synonymous SNPs (Zhang et al., 2014). Likewise, a similar signal on EPAS1 in the Tibetan mastiff and other highland dog breeds was detected (Gou et al., 2014). Recently, Miao, Wang & Li (2016) identified that the Tibetan mastiff EPAS1 locus was adaptively introgressed using the D statistic (Green et al., 2010; Durand et al., 2011). However, inferring the D statistic is challenging for small genomic regions as it shows more extreme values if the effective population size is low and if the interrogated regions have low genetic diversity (Martin, Davey & Jiggins, 2015), such as those found in high altitude wolves (Fan et al., 2016). Further, a single D-statistic test cannot be used to directly infer the direction of the introgression event and the authors suggest it likely was transferred from wolves to dogs since wolves had a more ancient history on the Tibetan Plateau. Finally, this survey was limited to Tibetan mastiffs as examples of high altitude dogs and only included a single low elevation dog breed for comparison.

Here, we investigate the hypothesis that EPAS1 in domestic highland dogs was obtained through introgression from highland wolves using a larger panel of reference canids. As such, we focused on the chromosome where this candidate gene resides, canine chromosome 10. We use complete sequence data of chromosome 10 to specifically assess if dogs and wolves derived EPAS1 mutations through introgression, and to define which of the two species was the donor if there was an introgression event. Dogs have dispersed into new environments along with the expansion of human civilization. According to previous studies of the history of human settlement on the Tibetan Plateau, the initial colonization occurred during the Paleolithic era (Zhao et al., 2009; Qi et al., 2013; Lu et al., 2016). Therefore, given the recent arrival of dogs to the Tibetan Plateau, the transfer of hypoxia adaptations from dogs to wolves would be a striking second example of variants selected under domestication transforming a related wild population. The first example is the K locus (a canine beta-defensin gene), which is a mutation that causes to canine
melanism that originated in the dog genome and was transferred through introgression to North American gray wolves over 10,000 years ago (Anderson et al., 2009). Alternatively, if the mutations originated in wolves, which have a much longer history in the Tibetan Plateau (Brantingham, Rhode & Madsen, 2010; Chen et al., 2015), then this would be the first example of adaptive introgression from wolves to dogs. A third possibility is the independent evolution of high elevation adaptations in dogs and wolves, a result which would highlight the relative mutability of EPAS1 as a response to environmental challenges. To better distinguish among these hypotheses, we use an expanded representation of dogs and wolves from both high and low elevation populations, and an analytical method that specifically assigns genomic regions to either species. We conclude that the recent arrival of dogs and the high diversity of and the deep divergence among EPAS1 haplotypes in highland wolves suggest the high altitude adapted variant originated in the highland wolves and was transferred to dogs, rather than evolving independently in both highland dogs and wolves.

**MATERIALS & METHODS**

**Samples and genome sequence data**

To determine if highland dogs inherited a beneficial allele through admixture with wolves, we obtained publically available genome sequence and SNP genotype data from 21 highland (>3,300 m) dogs (n = 10 indigenous dogs from a putatively admixed population, n = 11 Tibetan mastiffs) in China to compare to 15 reference dogs (n = 13 indigenous Chinese dogs; n = 1 basenji, n = 1 dingo), 22 low elevation Old World (OW) wolves, and six highland wolves from China (Fig. 1, Table S1). For this study, we obtained the genomic sequence and SNP variants for two genomes: one low elevation wolf from Inner Mongolia (im5) and one highland wolf from Tibet (ti3) (Table S1). The whole genome sequencing was
performed using an Illumina Hiseq 2,000 at Novogene (China). Two paired-end libraries with insert sizes of 300~500 bp were generated for each sample. Library preparation and sequencing was performed according to manufacturer’s protocols. Our newly generated sequence data can be found on NCBI’s Sequence Read Archive (SRA) under the accession number SRP096612. The 150 bp pair-end reads of each sample were aligned to the dog genome (CanFam3.1) using Bowtie 2 (Langmead & Salzberg, 2012) under the local alignment algorithm with the very sensitive model. Default options were used for all other parameters. Then, we applied Picard and GATK toolsets (DePristo et al., 2011) to process the alignments to identify SNPs. The pipeline is the same as used in our previous studies (Freedman et al., 2014; Zhang et al., 2014; Fan et al., 2016).

To obtain a set of polymorphic SNP genotypes, we excluded monomorphic sites and applied a strict inclusion threshold of no missing data to obtain 17,709 SNPs on canine chromosome 10. Further, to increase SNP density around the candidate gene EPAS1, we allowed on average 14% missing data, which provided an additional 1,113 SNP genotypes (total number of SNPs = 18,792). To assess if our assumed reference dogs were indeed non-admixed, we used the Ancestry function in Galaxy’s Genomic Diversity toolkit for K = 2–4 (Alexander, Novembre & Lange, 2009; Afgan et al., 2016) and performed a complete population structure analysis for all samples with 17,709 SNP genotypes.

Admixed ancestry analysis
To better explore the hypothesized introgression of EPAS1 in to highland dogs, we utilized ancestry information from a larger reference panel of dogs and gray wolves, and used a different methodological approach than previous work. Specifically, following methods in vonHoldt et al. (2011), we estimated the degree of per-SNP genetic differentiation ($F_{ST}$) between the reference dog and Old World wolves and employed a 2 standard deviation (SD) threshold to identify SNPs with significantly high levels of divergence. We then identified SNPs with strongly divergent allele frequencies between the two reference groups of dogs and wolves. Namely, sites with the highest $F_{ST}$ values, as defined by the 2SD threshold, were classified as ancestry informative markers (AIMs).

We assigned either dog or wolf ancestry to chromosome 10 SNPs from 21 highland dogs that are putatively admixed with OW wolves. Samples were assessed for outlier patterns using principal component analysis (PCA) in Galaxy’s Genomic Diversity toolkit (Afgan et al., 2016) on both the set of 17,709 and 980 AIM SNPs. Further, we surveyed local ancestry proportions for highland dogs using the program Admixture in Galaxy’s Genomic Diversity toolkit from the estimated genotype, two reference populations of dogs and wolves, and a switch penalty value of 10 to estimate per-site ancestry of the putatively admixed highland dogs (Table S1). We then surveyed average wolf ancestry across chromosome 10 in 100-SNP windows with a 50-SNP step in the putatively admixed highland dogs. Outlier regions were identified by a simple outlier threshold of wolf ancestry proportion >2SD above the chromosomal level (average wolf proportion, SD = 0.032, 0.09). We further repeated the PCA and completed a population structure analysis by using the Ancestry function in Galaxy (Alexander, Novembre & Lange, 2009) for SNP genotypes within any putatively introgressed regions. To phase our SNP genotypes using vonHoldt et al. (2017), PeerJ, DOI 10.7717/peerj.3522
SHAPEIT (Delaneau, Marchini & Zagury, 2012), we constructed a genetic map file as the ratio of inter-SNP distances to the position of the last SNP queried on chromosome 10. Additionally, we phased SNP genotypes using the program PCAdmix and plotted the per-individual loadings for the first two coordinates (Henn et al., 2012). Lastly, we applied a haplotype-based test (Mendez, Watkins & Hammer, 2012a; Mendez, Watkins & Hammer, 2012b) to determine if the shared EPAS1 haplotype in highland dogs and wolves was due to an adaptive introgression event or the alternative hypothesis that selection acted on a haplotype present in the ancestral population. We utilized a recently inferred genetic map (Campbell et al., 2016). We assume a generation time of three years (Freedman et al., 2014; Fan et al., 2016) and that all recombination events are detectable.

Variant effect predictions were conducted with transcripts annotated from the reference dog genome (CanFam 3.1), with SIFT (Sorting Intolerant From Tolerant) scores on non-synonymous substitutions, which range from 0 to 1, used to predict if the non-synonymous variants may be damaging. Specifically, if the SIFT score is ≤ 0.05 it is considered damaging, or tolerated when the score is >0.05 (McLaren et al., 2010).

Scan for positive selection
We scanned the target genomic region for signals of positive selection in the highland dog genome compared to the reference dogs using cross population extended haplotype homozygosity (XP-EHH) tests (Sabeti et al., 2007). This analysis was based on the phased haplotypes from 836 SNPs that had no missing data within the putatively introgressed region containing PRKCE and EPAS1. Per-SNP $F_{ST}$ was calculated as described above. We normalized both the $F_{ST}$ and XP-EHH scores as z-scores with a mean of zero and standard deviation of 1. Their product represents a composite “bivariate percentile score”. We identified outlier loci as those in the 97.5th percentile or greater (z-score > 2) of the bivariate percentile score distribution.

RESULTS

Two regions are enriched for wolf ancestry in highland dogs
To determine if admixture mapping is useful in searching for adaptively introgressed alleles in Chinese highland dogs, we first conducted a survey of genetic structure for 64 Old World canids along chromosome 10 (SNPs $n = 17,709$) (Fig. 1; Table S1). We found that the reference wolves represent two genetic clusters, high and low elevation individuals, with little evidence for admixture with dogs (Figs. 2, 3A). We further identified the reference dogs to be largely non-admixed with wolves, with the only exception being the basenji (bas; Fig. 2). Although the highland and reference dogs were similar in the PCA analysis (Fig. 3A), these two populations were distinct at $K = 4$ and displayed varying degrees of admixture with each other (Fig. 2). Further, highland dogs contained low chromosome-wide proportions of wolf ancestry on chromosome 10 (max = 0.027) (Table 1).

We surveyed ancestry across multiple dog and wolf populations based on 18,794 SNP genotypes on canine chromosome 10 to provide a higher density survey across the candidate gene, EPAS1. Using the 2SD threshold, we classified 980 $F_{ST}$ outlier SNPs as AIMS that were
informative for dog and wolf ancestry (all SNPs: $F_{ST}$ average = 0.04, SD = 0.10, $+2SD = 0.23$; AIMS: $F_{ST}$ average = 0.38, SD = 0.13), with an average density of 1 AIM per 70Kb across chromosome 10 (median = 26 Kb). Though non-model based, a PCA of 980 AIMS clearly identified four clusters, partitioned by species and elevation (Fig. 3B; Fig. S1). A sliding window analysis of AIMS along chromosome 10 revealed an enrichment of wolf ancestry in two regions (chr10: 8147438–8288850 containing the MSRB3 gene, average wolf proportion = 0.278; chr10: 48151501–48761234: PRKCE and EPAS1 genes, average wolf proportion = 0.355), thus identifying them as candidate outliers (Fig. 4). Genetic analysis of each outlier region varies in the degree of structure (Figs. 3C and 3D; Fig. S1) and observed outliers are found in three putatively introgressed genes. Following previous work and with the objective to explore the introgressed history of EPAS1 in high elevation adaptation, we focused on the region containing PRKCE and EPAS1 in subsequent analyses.

We conducted a PCA of phased haplotypes from this genomic region that contains both an excess of wolf ancestry in dogs and two functionally-relevant candidate genes. Our objective was to identify the potential source population of this putatively introgressed region in highland dogs. Low elevation wolves are spatially discrete compared to those from the highland, where 10 of 12 haplotypes are nested within the putatively admixed highland dogs (Fig. 5). This result suggests that the putatively introgressed region in the highland dog
Figure 3  Principal component analysis of (A) 17,709 SNPs, (B) 980 AIMs, and across two ancestry outlier genes (C) MSRB3 (100 SNPs) and (D) PRKCE/EPAS1 (167 SNPs).

genome originated from highland wolves in Qinghai and Tibet, China. From the phased haplotypes of the introgressed region containing PRKCE and EPAS1, we defined a 396 Kb subregion \( (n_{SNPs} = 467; \text{chr}10: 48305073–48700082) \) that had had a high proportion of wolf ancestry (wolf prop. > 0.45) across all highland dogs implying the origin of this region derives from admixture with highland gray wolves.

To better understand functional variation in this introgressed region, we determined the presence of damaging variation. We restricted our assignment of functional changes to SNPs within this candidate subregion and found one missense damaging non-synonymous variants in each of the PRKCE and EPAS1 genes (Table 2). The PRKCE variant was only observed in a single highland wolf (qh4), and the EPAS1 variant found in a putatively admixed highland Tibetan mastiff (DQZA81) and one low elevation indigenous reference dog (dc5).

Using the recent canine genetic map, we inferred that the 60 Kb length block containing the EPAS1 gene has a genetic map length of 0.032 cM. Consequently, the time until one of the two identical-by-descent copies of a EPAS1 haplotype are broken by a recombination event is exponentially distributed with a parameter of \( (0.00032 \times 2) \) per generation. We found that one of the two identical-by-descent copies of the EPAS1 haplotype has a 50%,
Figure 4  Local average wolf ancestry in a sliding window for highland dogs. Genes are indicated by black boxes.

Table 1  Wolf ancestry proportions for the putatively admixed Old World highland dogs for 17,709 SNPs on chromosome 10. (Abbreviations: Prop., proportion)

| Sample ID | Wolf prop. |
|-----------|------------|
| DQ25      | 0.007      |
| DQ28      | 0.017      |
| DQ29      | 0.029      |
| DQ32      | 0.002      |
| DQ33      | 0.005      |
| DQ45      | 0.010      |
| DQ52      | 0.004      |
| DQ54      | 0.002      |
| DQ59      | 0.002      |
| DQ60      | 0.008      |
| DQZA01    | 0.015      |
| DQZA06    | 0.021      |
| DQZA12    | 0.006      |
| DQZA18    | 0.006      |
| DQZA19    | 0.041      |
| DQZA23    | 0.011      |
| DQZA33    | 0.012      |
| DQZA55    | 0.002      |
| DQZA80    | 0.006      |
| DQZA81    | 0.006      |
| dtm       | 0.016      |

95% and 99% probability of at least one recombination event in 3,261, 14,092 and 21,663 years, respectively. On the other hand, estimates of the divergence time of dogs and wolves range from 11,700 or 29,000 years ago depending on the mutation rate used (Freedman et al., 2014; Skoglund et al., 2015; Fan et al., 2016). These divergence time estimates imply that recombination would have broken one of the two identical by descent copies of the EPAS1.
Figure 5  PCA of phased haplotypes for the putatively introgressed region containing the PRKCE and EPAS1 genes for the (A) whole sample set and (B) those samples with PC1 > 0. Geographic origins of each haplotype are provided. Refer to Fig. 1 for the map and Table S1 for more sample details.
Table 2  Putative functional consequences of SNP variants within the introgressed subregion (chr10: 48305073–48700082) of admixed highland dogs that contains PRKCE and EPAS1.

| CFA10 position | Allele | Gene  | Consequence | Impact     | Predicted SIFT phenotype (score) | Individuals carrying the allele |
|----------------|--------|-------|-------------|------------|---------------------------------|--------------------------------|
| 48,284,953     | T      | PRKCE | Missense    | Moderate   | Deleterious (0)                 | One highland wolf (qh4)        |
| 48,631,974     | C      | EPAS1 | Missense    | Moderate   | Deleterious (0.02)              | Tibetan mastiff (DQZA81), low elevation indigenous dog (dc5) |

haplotype at least once (with a 95% probability) if the ancestral haplotype was present in the population ancestral to dogs and wolves. Contrary to this expectation, highland dogs and wolves share the full 60 Kb EPAS1 haplotype, which is evidence in favor of an adaptive introgression event rather than selection on an ancestral haplotype in common ancestor of dogs and wolves.

Signal of selection in EPAS1

To confirm the signal of selection on EPAS1 as detected in previous studies, we conducted a cross-population analysis of extended haplotype homozygosity and genetic differentiation between the highland and low elevation dog genomes. This analysis supports the previous finding that EPAS1 is under strong positive selection (chr10: 48565127–48625694). However, this signal is discrete and does not encompass the adjacent gene, PRKCE (Fig. 6A), further supporting the specific role of EPAS1 in high elevation adaptation in domestic dogs. Additionally, phasing the 363 SNPs in EPAS1 also confirmed a shared haplotype between the highland dogs and wolves from Tibet (samples ti1, ti2, ti3) and Qinghai (samples qh1, qh2, qh4), China (Fig. 6B).

DISCUSSION

Adaptive introgression is an effective and rapid mechanism for species to adapt to new environments through the transmission of beneficial variants from related locally adapted taxa (Hedrick, 2013). Here, we highlight the role of adaptive introgression from Tibetan wolves to a diverse representation of high altitude dogs in China. We identified an introgressed haplotype containing the genes EPAS1 and PRKCE. Though introgressed as part of the same haplotype, only EPAS1 is under strong positive selection in highland dogs. However, the role of PRKCE in wild populations has yet to be explored. Experimentally, PRKCE null alleles in mice reduce pulmonary vasoconstriction in a hypoxic treatment (Littler et al., 2003). Analysis of diagnostic SNPs and the phased chromosomes revealed that the most likely source of the introgressed haplotype is from wolves in Tibet and Qinghai, China.

An analogous adaptive introgression process has been described between dogs and North American gray wolves. Specifically, the K locus, (a canine beta-defensin gene) has a dominantly inherited 3-bp mutation that causes black coat color in wolves and when heterozygous, confers higher fitness (Anderson et al., 2009; Coulson et al., 2011). This mutation, though Old World in origin, was transferred to North American gray wolves from New World dogs >10,000 years ago, yet no trace of dog ancestry remains in these wolves outside of the introgressed region (Anderson et al., 2009; Schweizer et al., 2016a; vanHoldt et al., 2017).
Figure 6  (A) Positive selection scan of 835 SNPs in the Old World highland dogs. (B) Phased haplotypes for 363 phased SNPs in EPAS1 (as indicated by the black bar). Rows are individuals; columns are SNP positions. Red color indicates the canfam3.1 reference allele; green the alternate allele.
Our finding of adaptive admixture in dogs, parallels the history of high altitude Tibetans whose EPAS1 haplotype derives from admixture from Denisovans but whose genome is otherwise very similar to geographically proximate low altitude Han populations (average $F_{ST} \sim 0.02$; Huerta-Sánchez et al., 2014). Given the absence of the EPAS1 highland haplotype in the genome of dogs from lowland regions, parallel mutation in highland dogs and wolves followed by an independent selective sweep would be needed to explain our results. This scenario is less parsimonious than adaptive introgression. The length of the conserved highland haplotype in dogs and wolves is also consistent with adaptive introgression, instead of selection on a haplotype that has not been broken up by recombination since at least the divergence of dogs and wolves. Finally, both the high diversity of and divergence among EPAS1 haplotypes suggest they evolved well before the arrival of dogs to the highlands about 10,000 years ago.

We identified two moderately deleterious missense variants in two genes in the candidate region containing PRKCE and EPAS1. One variant in EPAS1 was restricted to the Tibetan mastiff, whereas the second variant was found in PRKCE in a single highland admixed dog and a single low elevation indigenous dog. EPAS1’s function is central to hypoxia adaptation, which can be mitigated through a physiological decrease in the oxygen demand, typically through an upregulation of ATP-producing pathways and reduced activity of ATP-demanding pathways (e.g., ion pumping and protein synthesis) (Storey & Storey, 1990; Guppy, Fuery & Flanigan, 1994; Gnaiger & Kuznetsov, 2002). Our results suggest that adaptive variation of EPAS1 from highland wolves and variants in linked genes potentially facilitate the hypoxia response in highland dogs through adaptive introgression and that the EPAS1 variant unique to Tibetan mastiffs is not likely the causative variant in highland dogs. Critically, we show that this adaptation in dogs did indeed derive through hybridization with highland wolves confirming that adaptive variation can be transferred in both directions from wolves to dogs and dogs to wolves. The intertwined history of dogs and wolves ensures a unique evolutionary dynamic where variants that have appeared in the history of either species can be tested for their effects on fitness under natural and artificial selection. Such coupled evolutionary histories may be key to the persistence of wild canines and their domesticated kin given the increasing anthropogenic modifications that characterize the past, present and future of both species.

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The authors declare there are no competing interests.
Author Contributions
- Bridgett vonHoldt conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials-analysis tools, wrote the paper, prepared figures and/or tables, reviewed drafts of the paper.
- Zhenxin Fan conceived and designed the experiments, performed the experiments, analyzed the data, reviewed drafts of the paper.
- Diego Ortega-Del Vecchyo analyzed the data, reviewed drafts of the paper.
- Robert K. Wayne conceived and designed the experiments, contributed reagents/materials-analysis tools, wrote the paper, reviewed drafts of the paper.

Data Availability
The following information was supplied regarding data availability:
The only new sequences collected for this manuscript can be found on SRA (NCBI, accession SRP096612).

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REFERENCES
Abi-Rached L, Jobin MJ, Kulkarni S, McWhinnie A, Dalva K, Gragert L, Babrzadeh F, Gharizadeh B, Luo M, Plummer FA, Kimani J, Carrington M, Middleton D, Rajalingam R, Bekscak M, Marsh SG, Maiers M, Guethlein LA, Tavoularis S, Little AM, Green RE, Norman PJ, Parham P. 2011. The shaping of modern human immune systems by multiregional admixture with archaic humans. Science 334:89–94 DOI 10.1126/science.1209202.
Afgan E, Baker D, Van den Beek M, Blankenberg D, Bouvier D, Cech M, Chilton J, Clements D, Coraor N, Eberhard C, Gruning B, Guerler A, Hillman-Jackson J, Von Kuster G, Rasche E, Soranzo N, Turaga N, Taylor J, Nekrutenko A, Goecks J. 2016. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2016 update. Nucleic Acids Research 44(W1):W3–W10 DOI 10.1093/nar/gkw343.
Alexander DH, Novembre J, Lange K. 2009. Fast model-based estimation of ancestry in unrelated individuals. Genome Research 19(9):1655–1664 DOI 10.1101/gr.094052.109.
Anderson TM, vonHoldt BM, Candille SI, Musiani M, Greco C, Stahler DR, Smith DW, Padhukasahasram B, Randi E, Leonard JA, Bustamante CD, Ostrander EA, Tang H, Wayne RK, Barsh GS. 2009. Molecular and evolutionary history of melanism in North American gray wolves. Science 323(5919):1339–1343 DOI 10.1126/science.1165448.
Arnold ML. 2016. Divergence with genetic exchange. Oxford: Oxford University Press.
Beall CM, Cavalleri GL, Deng L, Elston RC, Gao Y, Knight J, Li C, Li JC, McCormack M, Montgomery HE, Pan H, Robbins PA, Shianna KV, Tam SC, Tsering N,
Veeramah KR, Wang W, Wangdui P, Weale ME, Xu Y, Xu Z, Yang L, Zaman MJ, Zeng C, Zhang L, Zhang X, Zhaxi P, Zheng YT. 2010. Natural selection on EPAS1 (HIF2α) associated with low hemoglobin concentration in Tibetan highlanders. *Proceedings of the National Academy of Sciences of the United States of America* 107(25):11459–11464 DOI 10.1073/pnas.1002443107.

Becker M, Gruenheit N, Steel M, Voelckel C, Deusch O, Heenan PB, McLenachan PA, Kardailsky O, Leigh JW, Lockhart PJ. 2013. Hybridization may facilitate *in situ* survival of endemic species through periods of climate change. *Nature Climate Change* 3(12):1039–1043 DOI 10.1038/nclimate2027.

Brantingham JP, Rhode D, Madsen DB. 2010. Archaeology augments Tibet’s genetic history. *Science* 329(5998):1468–1468.

Campbell CL, Bhérer C, Morrow BE, Boyko AR, Auton A. 2016. A Pedigree-based map of recombination in the domestic dog genome. *G3* 6:3517–3524 DOI 10.1534/g3.116.034678.

Chen FG, Dong GF, Zhang DJ, Liu XY, Jia X, An CB, Ma MM, Xie YW, Barton L, Ren XY, Zhao ZJ, Wu XH, Jones MK. 2015. Agriculture facilitated permanent human occupation of the Tibetan Plateau after 3600 B.P. *Science* 347(6219):248–250 DOI 10.1126/science.1259172.

Coulson T, MacNulty DR, Stahler DR, vonHoldt B, Wayne RK, Smith DW. 2011. Modeling effects of environmental change on wolf population dynamics, trait evolution, and life history. *Science* 334(6060):1275–1278 DOI 10.1126/science.1209441.

Delaneau O, Marchini J, Zagury J-F. 2012. A linear complexity phasing method for thousands of genomes. *Nature Methods* 9:179–181 DOI 10.1038/nmeth.1785.

DePristo MA, Banks E, Poplin R, Garimella KV, Maguire JR, Hartl C, Philippakis AA, Del Angel G, Rivas MA, Hanna M, McKenna A, Fennell TJ, Kernytsky AM, Sivachenko AY, Cibulskis K, Gabriel SB, Altshuler D, Daly MJ. 2011. A framework for variation discovery and genotyping using next-generation DNA sequencing data. *Nature Genetics* 43:491–498 DOI 10.1038/ng.806.

Durand EY, Patterson N, Reich D, Slatkin M. 2011. Testing for ancient admixture between closely related populations. *Molecular Biology & Evolution* 28(8):2239–2252 DOI 10.1093/molbev/msr048.

Fan Z, Silva P, Gronau I, Wang S, Armero AS, Schweizer RM, Ramierz O, Pollinger J, Galaverni M, Ortega Del-Vecchyo D, Du L, Zhang W, Zhang Z, Xing J, Vila C, Marques-Bonet T, Godinho R, Yue B, Wayne RK. 2016. Worldwide patterns of genomic variation and admixture in gray wolves. *Genome Research* 26(2):163–173 DOI 10.1101/gr.197517.115.

Freedman AH, Gronau I, Schweizer RM, Ortega Del-Vecchyo D, Han E, Silva PM, Galaverni M, Fan Z, Marx P, Lorente-Galdos B, Beale H, Ramirez O, Hormoxdiari F, Alkan C, Vila C, Squire K, Geffen E, Kusak J, Boyko AR, Parker HG, Lee C, Tadigotla V, Siepel A, Bustamante CD, Harkins TT, Nelson SF, Ostrander EA, Marques-Bonet T, Wayne RK, Novembre J. 2014. Genome sequencing highlights the dynamic early history of dogs. *PLOS Genetics* 10(1):e1004016 DOI 10.1371/journal.pgen.1004016.
Gnaiger E, Kuznetsov AV. 2002. Mitochondrial respiration at low levels of oxygen and cytochrome c. Biochemical Society Transactions 30(2):252–257 DOI 10.1042/bst0300252.

Gou X, Wang Z, Li N, Qiu F, Xu Z, Yan D, Yang S, Jia J, Kong Z, Wei Z, Lu S, Lian L, Wu C, Wang X, Li G, Ma T, Jiang Q, Zhao X, Yang J, Liu B, Wei D, Li H, Yang J, Yan Y, Zhao G, Dong X, Li M, Deng W, Leng J, Wei C, Wang C, Mao H, Zhang H, Ding G, Li Y. 2014. Whole-genome sequencing of six dog breeds from continuous altitudes reveals adaptation to high-altitude hypoxia. Genome Research 24(8):1308–1315 DOI 10.1101/gr.171876.113.

Grant PR, Grant BR. 2010. Conspecific versus heterospecific gene exchange between populations of Darwin’s finches. Philosophical Transactions of the Royal Society B 365:1065–1076 DOI 10.1098/rstb.2009.0283.

Green RE, Krause J, Briggs AW, Maricic T, Stenzel U, Kircher M, Patterson N, Li H, Zhai W, Fritz MHY, Hansen NF, Durand EY, Malaspinas AS, Jensen JD, Marques-Bonet T, Alkan C, Pruefer K, Meyer M, Burbano JA, Good JM, Schultz R,Aximu-Petri A, Butthof A, Hofer B, Hoffner B, Siegemund M, Weihmann A, Nusbaum C, Lander ES, Russ C, Novod N, Affourtit J, Egholm M, Verna C, Rudan P, Brajovic D, Kucan Z, Gusic I, Doronichev VB, Golovanova LV, Laluzeca-Fox C, De la Rasilla M, Fortea J, Rosas A, Schmitz RW, Johnson PLF, Eichler EE, Falush D, Birney W, Mullikin JC, Slatkin M, Nielsen R, Kelso J, Lachmann M, Reich D, Paabo S. 2010. A draft sequence of the Neandertal genome. Science 328(5979):710–722 DOI 10.1126/science.1188021.

Guppy M, Fuery CJ, Flanigan JE. 1994. Biochemical principles of metabolic depression. Comparative Biochemistry and Physiology Part B 109(2–3):175–189 DOI 10.1016/0305-0491(94)90001-9.

Hedrick P. 2013. Adaptive introgression in animals: examples and comparison to new mutation and standing variation as sources of adaptive variation. Molecular Ecology 22(18):4606–4618 DOI 10.1111/mec.12415.

Henn BM, Botigué LR, Gravel S, Wang W, Brisbin A, Byrnes JK, Fadhlaoui-Zid K, Zalloua PA, Moreno-Estrada A, Bertranpetit J, Bustamante CD, Comas D. 2012. Genomic ancestry of North Africans supports back-to-Africa migrations. PLOS Genetics 8(1):e1002397 DOI 10.1371/journal.pgen.1002397.

Huerta-Sánchez E, Jin X, Asan, Bianba Z, Peter BM, Vinckenbosch N, Liang Y, Yi X, He M, Somal M, Ni P, Wang B, Ou X, Huasang I, Luosang J, Cuo ZXP, Li K, Gao G, Yin Y, Wang W, Zhang X, Xu X, Yang H, Li Y, Wang J, Wang J, Nielsen R. 2014. Altitude adaptation in Tibetans caused by introgression of Denisovan-like DNA. Nature 512:194–197 DOI 10.1038/nature13408.

Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. Nature Methods 9:357–359 DOI 10.1038/nmeth.1923.

Little CM, Morris Jr KG, Fagan KA, McMurtry IF, Messing RO, Dempsey EC. 2003. Protein kinase C-epsilon-null mice have decreased hypoxic pulmonary vasoconstriction. The American Journal of Physiology Heart and Circulatory Physiology 284:H1321–H1331 DOI 10.1152/ajpheart.00795.2002.
Lu D, Lou H, Yuan K, Wang X, Wang Y, Zhang C, Lu Y, Yang X, Deng L, Zhou Y. 2016. Ancestral origins and genetic history of Tibetan highlanders. *American Journal of Human Genetics* 99:580–594 DOI 10.1016/j.ajhg.2016.07.002.

MacDonald DB, Clay RP, Brumfield RT, Braun MJ. 2001. Sexual selection on plumage and behavior in an avian hybrid zone: experimental tests of male-male interactions. *Evolution* 55(7):1443–1451 DOI 10.1111/j.0014-3820.2001.tb00664.x.

Martin SH, Davey JW, Jiggins CD. 2015. Evaluating the use of ABBA—BABA statistics to locate introgressed loci. *Molecular Biology & Evolution* 32(1):244–257 DOI 10.1093/molbev/msu269.

McLaren W, Pritchard B, Rios D, Chen Y, Flicek P, Cunningham F. 2010. Deriving the consequences of genomic variants with the ensembl API and SNP effect predictor. *Bioinformatics* 26(16):2069–2070 DOI 10.1093/bioinformatics/btq330.

Mendez FL, Watkins JE, Hammer MF. 2012a. A haplotype at STAT2 introgressed from Neanderthals and serves as a candidate of positive selection in Papua New Guinea. *The American Journal of Human Genetics* 91:265–274 DOI 10.1016/j.ajhg.2012.06.015.

Mendez FL, Watkins JC, Hammer MF. 2012b. Global genetic variation at OAS1 provides evidence of archaic admixture in Melanesian populations. *Molecular Biology and Evolution* 29:1513–1520 DOI 10.1093/molbev/msr301.

Miao B, Wang Z, Li Y. 2016. Genomic analysis reveals hypoxia adaptation of Tibetan Mastiff by introgression of grey wolf from Tibetan plateau. *Molecular Biology & Evolution* 34(3):734–743 DOI 10.1093/molbev/msw274.

Peng Y, Yang Z, Zhang H, Cui C, Qi X, Luo X, Tao X, Wu T, Ouzhuluobu, Basang, Chiwangsangbu, Danzengduojie, Chen H, Shi H, Su B. 2011. Genetic variations in Tibetan populations and high-altitude adaptation at the Himalayas. *Molecular Biology & Evolution* 28(2):1075–1081 DOI 10.1093/molbev/msq290.

Qi X, Cui C, Peng Y, Zhang X, Yang Z, Zhong H, Zheng H, Xiang K, Cao X, Wang Y, Ouzhuluobu, Basang, Ciwangsangbu, Bianba, Gonggalanzi, Wu T, Chen H, Shi H, Su B. 2013. Genetic evidence of paleolithic colonization and neolithic expansion of modern humans on the Tibetan plateau. *Molecular Biology and Evolution* 30:1761–1778 DOI 10.1093/molbev/mst093.

Sabeti PC, Varilly P, Fry B, Lohmueller J, Hostetter E, Cotsapas C, Xie X, Byrne EH, McCarroll SA, Gaudet R, Schaffner SF, Lander ES. The International HapMap Consortium. 2007. Genome-wide detection and characterization of positive selection in human populations. *Nature* 449:913–918 DOI 10.1038/nature06250.

Schweizer RM, Robinson J, Harrigan R, Silva P, Galaverni M, Musiani M, Green RE, Novenbre J, Wayne RK. 2016b. Targeted capture and resequencing of 1,040 genes reveal environmentally driven functional variation in gray wolves. *Molecular Ecology* 25(1):357–379 DOI 10.1111/mec.13467.

Schweizer RM, vonHoldt BM, Harrigan R, Knowles JC, Musiani M, Coltman D, Novembre J, Wayne RK. 2016a. Genetic subdivision and candidate genes under selection in North American gray wolves. *Molecular Ecology* 25(1):380–402 DOI 10.1111/mec.13364.
Simonson TS, Yang Y, Huff CD, Yun H, Qin G, Witherspoon DJ, Bai Z, Lorenzo FR, Xing J, Jorde LB, Prchal JT, Ge R. 2010. Genetic evidence for high-altitude adaptation in Tibet. Science 329(5987):72–75 DOI 10.1126/science.1189406.

Skoglund P, Ersmark E, Palkopoulou E, Dalén L. 2015. Ancient wolf genome reveals an early divergence of domestic dog ancestors and admixture into high-latitude breeds. Current Biology 25(11):1515–1519 DOI 10.1016/j.cub.2015.04.019.

Song Y, Endeplos S, Klemann N, Richter D, Matuschka FR, Shih CH, Nachman MW, Kohn MH. 2011. Adaptive introgression of anticoagulant rodent poison resistance by hybridization between old world mice. Current Biology 21(15):1296–1301 DOI 10.1016/j.cub.2011.06.043.

Storey KB, Storey JM. 1990. Metabolic rate depression and biochemical adaptation in anaerobiosis, hibernation and estivation. Quarterly Review of Biology 65(2):145–174 DOI 10.1086/416717.

vonHoldt BM, Pollinger JP, Earl DA, Knowles JC, Boyko AR, Parker H, Geffen E, Pilot M, Jedrzejewski W, Jedrzejewska B, Sidorovich V, Greco C, Randi E, Musiani M, Kays R, Bustamante CD, Ostrander EA, Novembre J. 2011. A genome-wide perspective on the evolutionary history of enigmatic wolf-like canids. Genome Research 21(8):1294–1305 DOI 10.1101/gr.116301.110.

Xu S, Li S, Yang Y, Tan J, Lou H, Jin W, Yang L, Pan X, Wang J, Shen Y, Wu B, Wang H, Jin L. 2011. A genome-wide search for signals of high-altitude adaptation in Tibetans. Molecular Biology & Evolution 28:1003–1011 DOI 10.1093/molbev/msq277.

Zhang W, Fan Z, Han E, Hou R, Zhang L, Galaverni M, Huang J, Liu H, Silva P, Li P, Pollinger JP, Du L, Zhang XY, Yue B, Wayne RK, Zhang Z. 2014. Hypoxia adaptations in the grey wolf (Canis lupus chanco) from Qinghai-Tibet Plateau. PLOS Genetics 10:e1004466 DOI 10.1371/journal.pgen.1004466.

Zhao M, Kong Q, Wang H, Peng M, Xie X, Wang W, Duan J, Cai M, Zhao S, Tu Y. 2009. Mitochondrial genome evidence reveals successful Late Paleolithic settlement on the Tibetan Plateau. Proceedings of the National Academy of Sciences of the United States of America 106:21230–21235 DOI 10.1073/pnas.0907844106.