A combined cryo-EM and molecular dynamics approach reveals the mechanism of ErmBL-mediated translation arrest

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Nascent polypeptides can induce ribosome stalling, regulating downstream genes. Stalling of ErmBL peptide translation in the presence of the macrolide antibiotic erythromycin leads to resistance in Streptococcus sanguis. To reveal this stalling mechanism we obtained 3.6-Å-resolution cryo-EM structures of ErmBL-stalled ribosomes with erythromycin. The nascent peptide adopts an unusual conformation with the C-terminal Asp10 side chain in a previously unseen rotated position. Together with molecular dynamics simulations, the structures indicate that peptide-bond formation is inhibited by displacement of the peptidyl-tRNA A76 ribose from its canonical position, and by non-productive interactions of the A-tRNA Lys11 side chain with the A-site crevice. These two effects combine to perturb peptide-bond formation by increasing the distance between the attacking Lys11 amine and the Asp10 carbonyl carbon. The interplay between drug, peptide and ribosome uncovered here also provides insight into the fundamental mechanism of peptide-bond formation.

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Ribosomes are the protein-synthesizing machines of the cell. The active site for peptide-bond formation, the peptidyl-transferase centre (PTC), is located on the large ribosomal subunit. During translation elongation, peptide-bond formation occurs when a peptidyl-tRNA is located at the P-site and an aminoacyl-tRNA at the A-site of the PTC. The α-amino group of the aminoacyl-tRNA makes a nucleophilic attack on the carbonyl carbon of the peptidyl-tRNA, transferring the peptidyl moiety on the A-tRNA and prolonging the nascent polypeptide chain by one amino acid. As the nascent polypeptide chain is synthesized, it passes through a tunnel located on the large ribosomal subunit and emerges on the solvent side where protein folding occurs. A growing body of evidence has revealed that the ribosomal tunnel is not a passive conduit for the nascent polypeptide chain but rather plays an important role in protein folding, co-translational targeting and translation regulation.

Translation can be regulated by intrinsic properties of the nascent chain, such that the amino-acid sequence of the polypeptide is sufficient to modulate its rate of synthesis, and in some cases even induce translation arrest. Well-characterized bacterial examples include the MrfM and SecM leader peptides, which interact with the ribosomal tunnel to induce translational stalling and thereby regulate expression of a downstream gene in the operon. Nascent polypeptide-mediated translation regulation can also require the presence of an additional ligand or cofactor. Well-characterized examples include the TnaC leader peptide, which requires the presence of free tryptophan for stalling to occur, and the Erm leader peptides, which induce translation stalling in the presence of the macrolide antibiotics.

In the case of Erm peptides, translational arrest on the leader peptide leads to expression of the downstream macrolide resistance determinant, usually a methyltransferase that confers resistance by methylating A2058, a 23S ribosomal RNA (rRNA) nucleotide that comprises part of the macrolide-binding site within the ribosomal tunnel. For example, in Streptococcus sanguis the expression of the ErmB methyltransferase is controlled by programmed arrest during translation of the upstream ermB leader peptide. In the absence of erythromycin (ERY), ErmB expression is repressed because the ribosome-binding site (RBS) and AUG start codon of the ermB mRNA are sequestered in a stem–loop structure. However, in the presence of sub-inhibitory concentrations of ERY, ribosomes translating the ErmBL leader peptide become stalled, leading to an alternative stem–loop structure in the mRNA that exposes the RBS and start codon of the ermB gene and thus allows ribosome binding and induction of ErmB expression. The drug-dependent nature of the stalling ensures that expression only occurs when the drug is present, which is critical for survival because of the fitness cost associated with the methylation of A2058.

Recent studies demonstrated that polymerization of the ErmBL nascent chain halts because the ribosome is unable to catalyse peptide-bond formation between the 10 amino-acid-long ErmBL-tRNA<sub>Asp</sub> (codon 10) in the ribosomal P-site and Lys-tRNA<sub>Asp</sub> (codon 11) in the A-site (Fig. 1a). The ErmBL-dependent translational arrest occurs with both macrolides (for example, ERY) and ketolides (for example, telithromycin), which contrasts with other leader peptides, such as ErmCL, where stalling is specific for macrolides. Mutagenesis studies identified four residues within ErmBL (Arg<sub>7</sub> and Val<sub>9</sub>-Lys<sub>11</sub>) that are critical for stalling and two of which (Arg<sub>7</sub> and Asp<sub>10</sub>) play a role in determining the antibiotic specificity of ErmBL. Interestingly, the compromised stalling efficiency of the ErmBL-R7A mutant can be rescued by compensatory mutations of 23S nucleotide U2586, located within the ribosomal tunnel, but not by mutations in the neighbouring tunnel nucleotide A2062 that impair ErmCL stalling. The first structural insights into how the ErmBL leader peptide, the macrolide antibiotic and the ribosomal tunnel cooperate to inactivate the PTC of the ribosome were derived from a cryo-electron microscopy (cryo-EM) structure of the ErmBL-stalled ribosomal complex (ErmBL-SRC) at 4.5–6.6-Å resolution. On the basis of this structure, it was suggested that peptide-bond formation cannot occur because the ErmBL nascent chain stabilizes a non-productive state of the ribosome that prevents accommodation of Lys-tRNA at the A-site.

Here we present cryo-EM structures of the ErmBL-SRC in the presence and absence of A-tRNA at 3.6-Å resolution, which were used as a basis to perform extensive molecular dynamics (MD) simulations. Our analyses revealed a complex network of hydrogen-bonding interactions between the C-terminal region of ErmBL and nucleotides of the 23S rRNA, consistent with the importance of these residues for translational stalling. Unexpectedly, the C-terminal Asp<sub>10</sub> residue of ErmBL was observed in a rotated conformation, promoting a unique path of the nascent chain such that it bypasses the tunnel-bound macrolide. In the cryo-EM structure, the unusual conformation of ErmBL appears to distort the terminal A76 ribose of the P-tRNA, which is supported by MD simulations performed after removal of ERY, showing that the ribose distortion is alleviated as the ErmBL nascent chain moves into the volume previously occupied by the drug. In addition, MD simulations predict that the presence of ERY also influences the position of the aminoacyl moiety of the A-tRNA via a cascade of 23S rRNA rearrangements. Thus, our results illustrate how ErmBL and ERY cooperate to perturb both the A- and P-tRNA positions at the PTC to inhibit peptide-bond formation and induce translational arrest.

**Results**

**Cryo-EM structures of the ErmBL-SRC.** The ErmBL-SRCs were generated by translation of a dicistronic 2XermBL mRNA in the presence of 20 μM ERY using an Escherichia coli lysate-based in vitro translation system. The ErmBL-SRC disomes were then isolated by sucrose gradient purification, converted to monosomes and analysed using single-particle cryo-EM (see Methods). In *silico* sorting of cryo-EM images yielded two major homogeneous subpopulations of ribosomes bearing P-tRNA and E-tRNAs, but differing in the presence (termed ErmBL-AP-E-SRC; 85,393 particles, 33%) or absence (termed ErmBL-PE-SRC; 75,839 particles, 30%) of A-tRNA (Fig. 1b and Supplementary Fig. 1). Both subpopulations could be refined to an average resolution of 3.6 Å (using a Fourier shell correlation (FSC) cutoff of 0.143; Supplementary Fig. 2), with local resolution calculations indicating that the ribosomal core reaches towards 3.1 Å (Fig. 1c and Supplementary Fig. 2). At this resolution, base separation is clearly observed for rRNA nucleotides (Fig. 1d), as is the density for the majority of the amino-acid side chains in ribosomal proteins (Fig. 1e). Density is also observed for the lysyl moiety attached to the CCA end of the A-tRNA as well as for the ErmBL leader peptide attached to CCA end of the P-tRNA (Fig. 1f).

**A molecular model for the ErmBL nascent chain.** The ErmBL nascent chains of both ErmBL-SRCs were well resolved within the ribosomal tunnel, as indicated by local resolution (Supplementary Fig. 3). Moreover, the electron density for ErmBL in the ErmBL-PE-SRC was essentially identical to that in the ErmBL-AP-E-SRC (Supplementary Fig. 3), indicating that the presence of the A-tRNA does not significantly influence the conformation of the ErmBL nascent chain. With the exception of the side chain of Asn8 for which we observe no density, we are able to present a
complete model for the ErmBL nascent chain residues Val3–Asp10 (Supplementary Table 1). In contrast, the N-terminus of ErmBL appears to be flexible, consistent with the reduced local resolution of this region, thus preventing the N-terminal Met1 residue from being modelled and allowing only the backbone for Leu2 to be tentatively assigned (Fig. 2a and Supplementary Fig. 3). The overall backbone trace of the ErmBL nascent chain within the ribosomal tunnel reported here is similar to the path reported previously for ErmBL based on a 4.5–6.6 Å cryo-EM map9 (Supplementary Fig. 4). Specifically, ErmBL adopts a unique conformation that enables it to bypass the macrolide ERY bound within the tunnel without even contacting it (Fig. 2a). This finding is consistent with biochemical studies demonstrating that ErmBL stalling occurs with a variety of macrolides and thus appears insensitive to the chemical nature of the drug9,11. The ErmBL path is distinct from that reported for the drug-dependent ErmCL-stalling peptide13, which establishes intimate interactions with the drug (Fig. 2b), consistent with the findings that ErmCL stalling occurs with macrolide antibiotics containing a C3-cladinose sugar, such as ERY, but not with ketolides, such as telithromycin that lack the C3-cladinose5,10.

This structure contrasts with the available structures of other polypeptide chains within the ribosomal tunnel14–20, which revealed paths that would be sterically obstructed by the presence of the macrolide, as exemplified here by the TnaC nascent chain14 (Fig. 2c). As noted9, the path of ErmBL illustrates the principle as to how specific amino-acid sequences allow translation of a subset of proteins, even in the presence of a macrolide-obstructed tunnel21,22.

Interactions of ErmBL within the ribosomal tunnel. In contrast to the previous reconstruction of ErmBL29, the significantly higher resolution of the ErmBL-SRCs reported here permits a detailed analysis of the interactions of ErmBL with components of the ribosomal tunnel (Fig. 3a). The majority of the interactions are localized within the last four C-terminal amino acids of ErmBL, namely Arg7–Asp10, which collectively establish a total of nine potential hydrogen-bond interactions with nucleotides of the 23S rRNA: Asp10 can form three hydrogen bonds, one from the backbone carboxyl oxygen with the N3 of U2506 and two from the Asp10 side chain oxygen atoms with the N1 and/or N2 of G2061 as well as the N3 of U2506 (Fig. 3b). These interactions are likely to be important since most mutations of Asp10 abolish MD simulations of the ErmBL-SRC. To capture the structural behaviour and energetics of the ErmBL peptide in the ribosome and to understand how these are influenced by ERY, all-atom explicit-solvent MD simulations were carried out. The simulations were started from two initial structures: (i) the ErmBL-APE-SRC structure (referred to as + ERY) and (ii) the ErmBL-APE-SRC structure after computational removal of ERY (– ERY). From each of these structures, two independent simulations were started. All simulations of the fully solvated complexes were set up using a previously established protocol23,24 and carried out using GROMACS25, the amber99sb force field26 and the SPC/E water model27. The root mean square deviation (r.m.s.d.) of the simulations from the cryo-EM structure remained low (r.m.s.d. < 5.2 Å; Supplementary Fig. 5, total simulation time ∼ 12 μs) in comparison with other, shorter MD simulations of ribosomes started from high-resolution X-ray structures (r.m.s.d. 6–10 Å)23,28. This underscores the quality of our simulation set-up and force field as well as that of the starting structure. Removal of ERY leads to increased dynamics and flexibility of the ErmBL nascent chain, in particular the N-terminal residues 2–5 of the ErmBL peptide (Fig. 2d), as seen by the root mean square fluctuations (r.m.s.f.’s; Fig. 2e). The absence of the drug allows the ErmBL nascent chain to explore the lumen of the tunnel, and to move into the volume otherwise occupied by the drug (Fig. 2d–f), consistent with the idea that ERY promotes the unique conformation of the ErmBL nascent chain.

Figure 1 | Cryo-EM structure of the ErmBL-SRC. (a) Schematic for ermBL-dependent regulation of ermB translation in the presence of ERY. (b) Cryo-EM maps of the ErmBL-APE-SRC and ErmBL-PE-SRC, with 30S (yellow), 50S (grey), A-tRNA (orange), P-tRNA (green) and E-tRNA (magenta). (c) Transverse sections of the cryo-EM maps of the ErmBL-APE-SRC and ErmBL-PE-SRC, coloured according to local resolution. (d–f) Examples of electron density (grey mesh) of the ErmBL-APE-SRC map illustrating (d) base separation within an rRNA helix, (e) side chains of ribosomal protein α-helix and (f) aminoacylated-CCA end of the A-tRNA (orange) and ErmBL peptide attached to the CCA end of the P-tRNA (green).
Overlapping volume of ErmBL peptide and ERY (Å$^3$)

Figure 2 | Path of ErmBL within the ribosomal tunnel. (a,b) Path of (a) ErmBL (green) and (b) ErmCL (yellow) within the ribosomal tunnel with bound ERY (red mesh). (c) Path of TnaC (pink) compared with the path of ErmBL (green) within the ribosomal tunnel with superimposed ERY (red mesh). (d) Path of ErmBL after 700-ns simulation time in absence of ERY (green) compared with ErmBL as seen in the cryo-EM structure (grey). (e) Overlap between the ErmBL peptide and the volume occupied by ERY in the course of the simulations.

translational stalling$^{9,11}$. All these interactions were also observed throughout the MD simulations. In particular, the hydrogen bonds between Asp10 and G2061 were very stable in all the simulations (62% average occupancy; Supplementary Fig. 6), with the formation of additional hydrogen bonds also being seen between the amide group of Asp10 and 23S rRNA nucleotide C2063 (74% average occupancy; Supplementary Fig. 6). The backbone carboxyl of Asn8 is within hydrogen-bonding distance of the ribose 2$'$OH of A2062, whereas no density is observed for the side chain, consistent with the report that Asn8Ala mutations do not affect ErmBL stalling$^9$. In this respect, we note that the most favoured rotamers for the Asn side chain would sterically clash with the desosamine sugar of ERY. In addition, in the simulations with ERY, although the backbone dihedral angles of Asn8 remain almost constant (Supplementary Fig. 7), the side chain of Asn8 is more mobile than the neighbouring residues (Fig. 2e). Indeed, the Asn8 side chain adopts multiple conformations with transient hydrogen bonding to U2506, A2062 and C2063 (Supplementary Fig. 6), thus explaining the lack of density for this side chain (Fig. 3c).

The well-resolved density (Fig. 3d) and low mobility (Fig. 2e) for the Arg7 side chain can be explained by the multiple interactions that this positively charged residue establishes by protruding into a negatively charged rRNA pocket of the tunnel wall (Fig. 3d and Supplementary Fig. 6). Specifically, two hydrogen bonds are possible with the two terminal amino groups of Arg7, namely to the phosphate oxygen atoms of C2063 and U2441 (Fig. 3d), which were also observed in the MD simulations (with an average occupancy of 13% and 87%, respectively, Supplementary Fig. 6). Additional hydrogen bonding is possible to the bridging ribose oxygen of A2439 and the ribose 2$'$OH of U2586 as well as between the backbone of NH of Arg7 and the O2 of U2586, although these interactions were not seen in the MD simulations (Supplementary Fig. 6). While an Arg7Ala mutation abolishes stalling$^9$, compensatory mutations of U2586 were shown previously to restore stalling$^9$, supporting the intimate association between these two entities. What is unclear from the structure is why the Val9Ala mutation also abolishes ErmBL-stalling activity$^9$. In the ErmBL-SRC, the side chain of Val9 is well resolved, yet it does not appear to come into close proximity of any components of the ribosomal tunnel (Fig. 3e). We speculate that the Val9 side chain may, therefore, be important for indirectly promoting the correct conformation of the ErmBL nascent chain: one possibility is that Val9 provides some steric constraints that are necessary for the Arg7 side chain to be optimally oriented, since these two residues are close to each other within the ribosomal tunnel. Interestingly, in the absence of ERY, all the other peptide residues exhibit larger fluctuations than those seen in the simulations with ERY. The only exception is Arg7, the fluctuations of which are unchanged. This observation, together with the fact that the interaction enthalpies of Arg7 with the tunnel wall remain essentially unchanged upon removal of the antibiotic, suggests that the free energy barrier for detaching the Arg7 from the tunnel-bound conformation might be rate-limiting.
In every single structure, the side chain of the Phe thermo1. In contrast to the previous study, the higher resolution stabilization an unaccommodated Lys-tRNA at the A-site of the A-tRNA, since it was previously proposed that ErmBL stabilized an unaccommodated Lys-tRNA at the A-site of the PTC. In contrast to the previous study, the higher resolution achieved here enables us to more accurately place the CCA end of the A-tRNA and also to model the lysyl moiety (Fig. 5a), which was not visible before9. Comparison of the position of the CCA.

Figure 3 | Interaction of the ErmBL nascent chain within the ribosomal tunnel. (a–f) Interactions of ErmBL (green) with 23S rRNA nucleotides (blue). In c, different rotamers of the N8 side chain are illustrated and ERY is shown in red.

for full detachment of the peptide from the tunnel wall. Apparently, this barrier is too high to be overcome on the microsecond simulation timescale.

In contrast to the C terminus, the N-terminal half of ErmBL (residues Leu2–Met6) is not critical for stalling and each residue can be individually mutated to alanine without loss of activity9. This observation is consistent with both the decreased local resolution in this region (Supplementary Fig. 3) as well as with the large fluctuations of these residues observed in the MD simulations (Fig. 2e), indicating that the N terminus of ErmBL is flexible and does not adopt a single defined conformation. In addition, the sequence of the N-terminal MLVFQM is very hydrophobic and therefore provides few possibilities for hydrogen-bonding interactions. Instead, the Phe4 side chain appears to pack against the U2609–A752 and U2586–U2585 base pairs, respectively (Fig. 3f). This observation is consistent with both the decreased local resolution in this region (Supplementary Fig. 3) as well as with the large fluctuations of these residues observed in the MD simulations (Fig. 2e), indicating that the N terminus of ErmBL is flexible and does not adopt a single defined conformation. In addition, the sequence of the N-terminal MLVFQM is very hydrophobic and therefore provides few possibilities for hydrogen-bonding interactions. Instead, the Phe4 side chain appears to pack against the U2609–A752 and U2586–U2585 base pairs, respectively (Fig. 3f).

Rotation of Asp10 and an alternate path for ErmBL. To understand how interactions of ErmBL within the ribosomal tunnel lead to translational arrest by preventing peptide-bond formation with the incoming Lys-tRNA in the A-site, we compared the ErmBL-SRC with available X-ray crystallography structures of ribosomal states with aminoacylated-tRNAs in the P-site. To date, these comprise structures of the Haloarcula marismortui 50S subunits bearing CC-puromycin (CC-pnm) or CC-Phe-caproic-acid-biotin tRNA analogues29–31 and of Thermus thermophilus 70S ribosomes bearing fMet-tRNA32 or Phe-tRNA33. In every single structure, the side chain of the Phe or fMet has the same orientation relative to the P-tRNA (Fig. 4a,b), namely pointing towards the direction of A2062. Unexpectedly, the Asp10 side chain attached to the P-tRNA in the ErmBL-SRC is rotated by 180° around the ester bond that links it to the 3′OH of A76, relative to the Phe/fMet side chains in the crystal structures (Fig. 4a,b). This unusual orientation might of course simply be due to the different respective tRNAs—a peptidyl-tRNA for the ErmBL structure versus an aminoacyl-tRNA in the other crystal structures. To investigate this possibility, we also compared the ErmBL-SRC with structures of other ribosomal states bearing peptidyl-tRNAs in the P-site.

There are currently four such cryo-EM structures available, the resolutions of which allow the orientation of the amino acid attached to the P-tRNA to be determined: TnaC-SRC14, ErmCL-SRC13, Sec61beta20 and MifM-SRC18. In all four structures, the amino-acid side chain attached to the P-tRNA is oriented similar to the Phe/fMet side chain in the crystal structures (Fig. 4c–f). This finding suggests that the difference is not due to different peptides on the P-tRNA. In addition, differences in species are likely to be irrelevant here, since the ErmCL- and TnaC-SRC were also formed using Escherichia coli ribosomes13,14. Finally, the rotation is also unlikely to be due to the nature of the amino acid because the residue that is attached to the P-tRNA in the MifM-SRC is also an Asp18. These findings lead us to conclude that the 180° rotation of the Asp10 residue in the ErmBL-SRC is likely to be a consequence of its context within the ErmBL nascent chain. It would therefore seem logical to assume that the interactions that stabilize the ErmBL in a specific conformation cooperatively stabilize the rotated state of Asp10.
**Figure 4 | Rotation of the C-terminal amino acid and an alternate path of ErmBL.** (a,b) Orientation of ErmBL D10 compared with (a) CC-Phe-caproic-acid-biotin (CC-pcb) tRNA analogues (pink, PDB1VQ6; purple, PDB1VQN; and teal, PDB1Q86 (ref. 74)), and Phe-tRNA (blue, PDB2WDL) and with (b) *T. thermophilus* 70S ribosomes bearing fMet-tRNA (brown, PDB4QCM and yellow, PDB 4Z3R). (c-f) Alternate path of ErmBL (green) compared with (c) that of ErmCL (yellow, PDB 3JZ), (d) MifM (light blue, PDB3J9W), (e) TnaC (pink, PDB4UY8 (ref. 14)) and (f) Sec61β (dark blue, PDB 3J92 (ref. 20)) in relation to the position of ERY (red).

**Figure 5 | A-site tRNA accommodation at the PTC of the ErmBL-APE-SRC.** (a–d) Atomic model for the CCA end of the A-site tRNA (orange) and (a) electron density (grey mesh) in the ErmBL-APE-SRC compared with (b) unaccommodated (pink, PDB1VQ6 (ref. 30)) and (c) accommodated states (brown, PDB4QCM; purple, PDB1VQN) and (d) with both. (e) Positions of the α-amino group of the amino acid attached to the A-site tRNA in the ErmBL-APE-SRC compared with its position in the pre-peptide-bond formation state (ref. 32). (f–h) Position of 23S rRNA nucleotide U2506 (blue) in (f) ErmBL-PE-SRC and (g) ErmBL-APE-SRC including electron density (grey mesh) and (h) compared with the unaccommodated and accommodated states of the PTC. (i) Comparison of 23S rRNA nucleotide U2585 in ErmBL-APE-SRC (blue), compared with the unaccommodated and accommodated states of the PTC. The nascent chain and A-tRNA in ErmBL are shown in green and orange, respectively.
end of the Lys-tRNA in ErnBL-SRC with the unaccommodated C-ppm and accommodated CC-ppm on the H. marismortui 50S subunit, as well as with accommodated CCA end of the Phe-tRNA in the T. thermophiles pre-attack complexes, suggests that the Lys-tRNA in the ErnBL-SRC is in a fully or near-fully accommodated state (Fig. 5b–e). The A-tRNA is shifted by 1.3 Å between the unaccommodated and accommodated states, when measuring the displacement using the ribose O4' atom. In contrast, the Lys-tRNA in the ErnBL-SRC is shifted by only 0.9 and 0.6 Å relative to the unaccommodated and accommodated states, respectively.

Accommodation of the A-tRNA is accompanied by corresponding conformational changes within the 23S rRNA nucleotides of the PTC; specifically, shifts in U2506 and U2584-U2585 are thought to be diagnostic for A-tRNA binding. In the ErnBL-PE-SRC, there is no clear density for U2506 (Fig. 5). However, upon binding of Lys-tRNA to the A-site, as in the ErnBL-PE-SRC, U2506 adopts a well-defined conformation (Fig. 5g), which is similar to that observed in the accommodated state (Fig. 5h). In both the ErnBL-SRC structures, the density for U2585 is weaker than other PTC nucleotides, but appears to predominantly adopt an intermediate position between the accommodated and unaccommodated U2585 states (Fig. 5i), hinting that the Lys-tRNA is not fully accommodated as suggested previously.

We note however that regardless of whether the Lys-tRNA is fully or near-fully accommodated, in the refined model of the Lys-tRNA in the ErnBL-PE-SRC 3' amino group is shifted by 1.5 Å compared with the accommodated pre-attack A-tRNAs (arrowed in Fig. 5e). The presence of the lysyl moiety on the A-tRNA indicates that the nucleophilic attack has not occurred, consistent with the previous biochemical evidence indicating that the lysine amino acid does not get incorporated into the ErnBL nascent chain.

Perturbation of the P-tRNA prevents peptide-bond formation. Comparison of the CCA end of the P-tRNA in the ErnBL-SRC (Fig. 6a,b) with the accommodated position of CC-ppm, Phe-tRNA or fMet-tRNA reveals a significant shift by 1–2 Å of the ribose and nucleobase of A76 (Fig. 6c,d), which was also seen at lower resolution. We note that the distance for the nucleophilic attack of the 3' amino group of the Lys-tRNA on the carbonyl carbon of the P-tRNA is 4.1 Å in the ErnBL-SRC (Fig. 6e), whereas the equivalent distances in the pre-attack complexes are 3.2–3.3 Å (refs 32; Fig. 6f), which may contribute to the inhibition of peptide-bond formation observed in the ErnBL-SRC. Consistently, the conformation of the P-tRNA A76 ribose was most similar to that of the pre-attack state in the MD simulations where ERY was absent (green traces in Fig. 6g), as compared with when the antibiotic was present (red traces in Fig. 6g). This finding suggests that the conformation of ErnBL induced by the presence of ERY in the ribosomal tunnel is responsible for the perturbed position of the P-tRNA.

The most recent model for peptide-bond formation, based on high-resolution X-ray crystallography structures, suggests that for the nucleophilic attack to proceed, a proton needs to be transferred from the attacking nucleophile to a water molecule (W1) via a proton wire formed by the 2' OH of the P-site A76 ribose and the 2'OH of A2451 (ref. 32; Fig. 6h and Supplementary

Figure 6 | Perturbation of the P-tRNA by ErnBL prevents peptide-bond formation. (a–d) Atomic model for the CCA end of the P-site tRNA (green) and (a,c,d) electron density (grey mesh) in the ErnBL-PE-SRC compared with (b) P-tRNAs in the pre-peptide-bond formation state (brown; PDB4QCM; blue, PDB2WDA; green). with (c,d) zoom onto the ribose of A76. (e,f) Relative orientation of the A-site 3' amino group and the P-site carbonyl carbon in (e) ErnBL-PE-SRC and (f) the pre-catalysis state. (g) Deviation of the ribose of A76 from the pre-attack state as a function of simulation time. (h) Conformation of the PTC in the ErnBL-SRC overlayed with the network of hydrogen bonds, creating a proton wire that enables peptide-bond formation. Cryo-EM map (grey mesh) with models for 23S rRNA (blue), L27 (brown) and P-tRNA (green).
The pre-attack complexes, nucleotide A2602 and the N-terminal regions of L27 become ordered\textsuperscript{3,23}, which, together with the non-bridging phosphate-oxygen (OP1) of A76 of the A-tRNA and the 2′-OH of A2451, coordinate W1 (ref. 32; Fig. 6h and Supplementary Fig. 8). In the ErmBL-SRC, we observed no density for A2602 nor for the N terminus of L27, both of which undergo large fluctuations in the MD simulations, suggesting that the W1 is not firmly coordinated (Fig. 6h). Stabilization of the very N-terminus of L27 has only been observed in the pre-attack structures because of interaction of the side chain of His3 of L27 with the OP1 of C75 of the A-tRNA and the backbone carboxyl of Ala2 of L27 with the N6 of A76 of the P-tRNA\textsuperscript{3,23,25}. Such simultaneous interaction of L27 with the A- and P-tRNA would be prohibited in the ErmBL-SRC by the shifted A76 nucleobase of the P-tRNA (Fig. 6h and Supplementary Fig. 8). We note however that previous studies have demonstrated that ribosomes lacking L27 are not impaired for peptide-bond formation\textsuperscript{34} and that mutations of A2506 affect predominantly peptidyl-tRNA hydrolysis rather than peptide-bond formation\textsuperscript{35,36}. The positioning of the positively charged lysine side chain of the A-tRNA is important for the conformational change to inactivate the PTC and induce translational arrest (Fig. 8). In the absence of ERY, and following the reduction in stalling efficiency of a Lys11Ala\textsuperscript{34} mutation in ErmBL\textsuperscript{9} may be because of the loss of interaction between the A-tRNA and the A-site crevice. To test this idea, we extended the MD simulations of the ErmBL-SRC in the presence and absence of ERY by 1 μs following \textit{in silico} mutation of Lys11 to Ala (Supplementary Fig. 5). In the presence of ERY, the Ala11 moves away from the crevice and towards the P-site (Fig. 7e, lower panel in Fig. 7d and Supplementary Fig. 9), leading to d2 distances shifted towards those seen with Lys11 in the absence of antibiotic (green trace in Fig. 7c). Moreover, the nucleotides retain a similar conformation as in the Lys11 simulation (Fig. 7e, lower panels in Fig. 7a,b and Supplementary Fig. 9), indicating that the motions of Ala11 and the crevice nucleotides are largely uncoupled.

To investigate whether the coupling between the aminoacyl moiety of the A-tRNA and the nucleotides of the A-site crevice is related to the strength of their interaction with one another, we calculated the interaction enthalpies for both Lys11 and Ala11 (see Methods). As predicted, the strongest interactions (more negative interaction enthalpies) were observed for the simulations with Lys11, regardless of the presence or absence of ERY (red and green curves in Fig. 7f, respectively). In contrast, much weaker interaction enthalpies were observed when Lys11 was substituted with Ala11 (cyan and orange curves in Fig. 7f), consistent with the notion that the positively charged lysine side chain is responsible for mediating interaction with the negatively charged nucleotides of the A-site crevice. This finding also suggests that ErmBL-mediated stalling should be influenced by the nature of the A-site aminoacyl moiety and should be efficient when the A-site contains a long positively charged side chain such as arginine. To test this hypothesis, we performed \textit{in vitro} translation of wild-type ErmBL containing Lys11 in the A-site, or with Lys11 mutated to one of the other 19 proteinogenic amino acids, and used the toe-printing assay to monitor the ribosomes stalled in the presence of ERY (Fig. 7g). Translational arrest was performed in the absence of the amino acid isoleucine (Ile), enabling ribosomes that readthrough the ErmBL stall site to be captured on the subsequent Ile codon of the mRNA, as observed in the absence of ERY (Fig. 7g). Quantitation and adjustment for loading indicated that as expected the nature of the A-site aminoacyl moiety had a dramatic influence on the efficiency of stalling, with Arg11 being threefold more efficient than the wild-type Lys11. It is important to note that for ErmBL comprising A-site amino acids, such as Cys11, Met11 or Phe11, where little stalling was observed, there was also no readthrough to the Ile codon, suggesting that in these cases, ERY induces high levels of peptidyl-tRNA drop-off, as reported previously\textsuperscript{11,40,41}.

**Discussion**

On the basis of our cryo-EM structures and MD simulations of the ErmBL-SRC, we propose a model for how the ErmBL nascent chain, together with the macrolide ERY and components of the ribosome, interplay to inactivate the PTC and induce translational arrest (Fig. 8). In the absence of ERY, and following accommodation of an aminoacyl-tRNA at the A-site of the PTC,
the α-amino group of the A-tRNA is positioned to make a nucleophilic attack on the carbonyl carbon of the peptidyl-tRNA in the P-site (Fig. 8a). In all structures to date, the side chain of the C-terminal amino acid, which is attached to the P-tRNA, is oriented such that all nascent polypeptide chains follow a similar path through the upper region of the ribosomal tunnel. This canonical pathway, which we term Path1, is sterically occluded by the presence of macrolide antibiotics (binding site dashed in Fig. 8a), explaining why this class of antibiotics induces peptidyl-tRNA drop-off for most nascent polypeptide chains40,41. In the presence of ERY, we observed a non-canonical path for the ErmBL nascent chain in the cryo-EM structure, which we termed Path2 (Fig. 8b). In the MD simulations, we observe that it is the presence of ERY that restricts the ErmBL conformations towards Path2. These restricted conformations in turn promote interaction of critical residues, such as Arg7 (R7) of ErmBL, with...
In addition, it was possible to change for example, D10G/E or D10P/Y mutations restrict stalling to ErmBL stalling but can also alter the antibiotic specificity, that mutations of R7 and D10 not only have a dramatic effect on our findings are consistent with a recent study demonstrating 10 why ErmBL stalling occurs with both macrolides and ketolides9.

Our findings are consistent with a recent study demonstrating that mutations of R7 and D10 not only have a dramatic effect on the A76 ribose of the P-tRNA is also perturbed. The perturbed and conformation of the ErmBL nascent chain, we observe that the drug interacts and stabilizes a conformation of the ErmBL-SRC formation is abolished by a -amino group of the A-tRNA (Fig. 8b), providing a structural explanation for the reduced rate of peptide-bond formation, namely, by restricting the path of the ErmBL such that orientation of the A-site crevice nucleotides (blue), which through electrostatic interactions keeps lys11 away from the P-site. We propose that the increased distance between the attacking amide group of the Lys11 and the carbonyl carbon of Asp10 hinders peptide-bond formation.

23S rRNA nucleotides on one side of the ribosomal tunnel (Fig. 8b). In addition, we observe that Asp10 (D10) of ErmBL adopts an unusual rotated conformation, which together with the R7 interaction, stabilizes the ErmBL nascent chain in the alternative Path2 conformation, rather than the canonical Path1. Path2 is directed away from the tunnel-bound drug, thus preventing direct peptide–drug interactions and explaining why ErmBL stalling occurs with both macrolides and ketolides9. Our findings are consistent with a recent study demonstrating that mutations of R7 and D10 not only have a dramatic effect on ErmBL stalling but can also alter the antibiotic specificity, for example, D10G/E or D10P/Y mutations restrict stalling to ERY or Tel, respectively11. In addition, it was possible to change the ERY dependency of ErmCL to also include Tel by substituting the ile9/Ser10 with Asp/Lys combination as in ErmBL11.

Presumably as a consequence of the unusual rotation of D10 and conformation of the ErmBL nascent chain, we observe that the A76 ribose of the P-tRNA is also perturbed. The perturbed A76 of the P-tRNA has a suboptimal geometry for nucleophilic attack of the a-amino group of the A-tRNA (Fig. 8b), providing a structural explanation for the reduced rate of peptide-bond formation and increased ribosome stalling during translation of ErmBL9. In addition, we have observed biochemically and inferred from the MD simulations that the nature of the A-site amino acid also influences the efficiency of translational stalling. On the basis of the MD simulations, we propose that the movement of the A-site crevice nucleotides (G2505-U2506/C2452) is coupled with the movement of the A-tRNA towards the P-tRNA during peptide-bond formation, thus allowing the a-amino group of the A-tRNA to make a nucleophilic attack on the carbonyl carbon of the peptidyl-tRNA in the P-site (Fig. 8a). We propose that the coupled movement results because of the interaction between the positively charged Lys11 (K11) of the A-tRNA and the negatively charged environment of the A-site crevice (Fig. 8a). In the MD simulations in presence of ERY, we observe that the drug interacts and stabilizes a conformation of the A-site crevice that prevents the coupled movement of the A-tRNA towards the P-site, thus suggesting that ERY can influence peptide-bond formation by allosterically influencing nucleotides within the A-site of the PTC (Fig. 8b). Consistently, uncoupling of the A-tRNA and A-site crevice is observed in the MD simulations with Lys11Ala mutant, indicating that the reduced interaction between Ala11 of the A-tRNA with the A-site crevice allows the A-tRNA to move towards the P-tRNA and undergo peptide-bond formation. Biochemically, we also observed that stable ErmBL-SRC formation is abolished by a Lys11Ala mutation, consistent with previous findings9, but that also other positively charged amino acids, such as Trp, His, Gln and especially Arg, promoted ribosome stalling (Fig. 7g).

In conclusion, this study illustrates how ERY exerts a dual effect on the PTC to inhibit peptide-bond formation, namely, by (i) restricting the path of the ErmBL such that orientation of the P-tRNA is perturbed and (ii) stabilizing the A-site crevice to prevent movement of the A-tRNA towards the P-site. Inhibition by P-tRNA perturbation has been suggested previously for the SecM arrest peptide16. Recent cryo-EM structures of other arrest peptides, including the drug-dependent ErmCL-SRC13, reveal that PTC silencing is induced by distinct allosteric conformational rearrangements that differ to those reported here for ErmBL. Thus, further investigations will be necessary to ascertain the extent to which other drug-dependent stalling systems utilize similar mechanisms to induce translational arrest or whether further novel mechanisms will be uncovered.

Methods

Generation and purification of ErmBL-SRC. ErmBL-SRCs were generated following the same procedure as previously described9. The 2XermBL construct was synthesized (Eurofins, Martinsried, Germany) such that it contained a T7 promoter followed by a strong RBS spaced by seven nucleotides (nts) to the ATG start codon of the first ermBL cistron. A linker of 22 nts separated the stop codon of the first ermBL cistron and the start codon of the second ermBL cistron. The linker also comprised the strong RBS 7 nts upstream of the ATG start codon of the second ermBL cistron, enabling initiation of translation independent from the first ermBL cistron. Each ermBL cistron encoded amino acids 1–17 corresponding to ErmBL leader peptide (Genbank accession number K00531) present on macrolide resistance plasmid pAM77 from S. sanguis strain a1 (ref. 42). The complete sequence of 2XermBL construct is: 5’-TAAATACGACTCACTATAGGAGGTTTTATAAGGGAAAAATTG TTTGATATTCAAATGGGAAAGTAAAGGAGTAAGAAACCATGCAATTTTGGAAA TAAAGTTTATAGGAGGAAAAAATGTTGTAATTTCATAGGTGAAAAGGAAAAGGA AAAGGAGGAAAAGAACACCAACAAAATTGTTGTAATTTCATAGGTGAAAAGGAAAAGGA AAAGGAGGAAAAGAACACCAACAAAATTGTTGTAATTTCATAGGTGAAAAGGAAAAGGA AAAGGAGGAAAAGAACACCAACAAAATTGTTGTAATTTCATAGGTGAAAAGGAAAAGGA AAAGGAGGAAAAGAACACCAACAAAATTGTTGTAATTTCATAGGTGAAAAGGAAAAGGA AAAGGAGGAAAAGAACACCAACAAAATTGTTGTAATTTCATAGGTGAAAAGGAAAAGGA AAAGGAGGAAAAGAACACCAACAAAATTGTTGTAATTTCATAGGTGAAAAGGAAAAGGA AAAGGAGGAAAAGAACACCAACAAAATTGTTGTAATTTCATAGGTGAAAAGGAAAAGGA
In vitro translation of the T2ermBL construct was facilitated by using the Rapid Translation System RTS 100. Each hybrid kit (SPRIIME, Cat. No. 240110) for in vitro coupled transcription-translation system with the PUREExpress in vitro Protein Synthesis Kit (NEB) was used. Each translation reaction consisted of 1 μl of 0.5 μl Uso inactivation mixture, 0.5 μl Uso tRNA mixture, 1 μl of 0.7 pmol DNA template (ermBL variants) (5′-ataatacgacggccaggttggataggaattctatc-3′, Metabion) was annealed to the mixed RNA-cistrons of the disomes, generating a DNA-RNA hybrid that could be cleaved by RNase H (NEB) treatment in buffer A at 25 °C for 1 h. After digestion of the disomes, ErmBL-SRC monosomes were again purified and concentrated by centrifugation using Amicon Ultra-0.5 ml Centrifugal Filters (Merck-Millipore) according to the manufacturer’s protocol.

The position of the ribosome on the mRNA was monitored with a toe-printing assay based on an in vitro-coupled transcription–translation system with the PUREExpress in vitro Protein Synthesis Kit (NEB) at 37 °C for 15 min at 500 r.p.m. in 1.5 ml reaction tubes. Ile-tRNA aminoacylation was performed using the Rapid Translation System RTS 100. Concentrated by centrifugation through Amicon Ultra-0.5 ml Centrifugal Filters (Merck-Millipore) according to the manufacturer’s protocol.

The data were provided as a series of seven frames (dose per frame of 60 e⁻/Å²/frames) at 300 kV with a pixel size of 1.108 Å and defocus range of −17 to −15 Å. The data set of 255,369 particles could be sorted into two main subpopulations using an incremental K-means-like method of unsupervised three-dimensional sorting (Supplementary Fig. 1). A minor subpopulation (94,139 particles, 37%) was defined by the presence of stoichiometric densities for the P- and E-tRNAs, lacking A-tRNA (L1 stalk in the ‘in’ position) and differing by the presence of sub-stoichiometric density for the A-tRNA, which was not sorted further because of the low particle numbers. The major subpopulation (161,231 particles; 63%) was defined by the presence of stoichiometric densities for the P- and E-tRNAs and sub-stoichiometric density for the A-tRNA. This population was further sorted into two additional subpopulations, both containing stoichiometric densities for the P- and E-tRNAs (L1 stalk in the ‘on’ position) and differing by the absence of the ErmBL-SRC or absence of A-tRNA (ErmBL-PE-SRC). Both subpopulations could be refined to an average resolution of 3.6 Å (0.143 FSC) and a local resolution extending towards 3.0 Å for the core of the 30S and 30S subunits as computed using ResMap (Supplementary Fig. 2). The final maps were subjected to the program EM-BCFAC40 in order to apply an automatically determined negative B-factor for sharpening of the map.

**Molecular modelling and PDB alignments**. Initial models for the two ErmBL-SRCs were obtained by combining: (i) a model of the E. coli 70S ribosome derived from a crystal structure at 2.8 Å resolution (PDB4U27 ref. 54); (ii) (A)/P-E-tRNAs and mRNA extracted from the 2.55 Å crystal structure of a pre-attack complex of the T. thermophilus 70S ribosome (PDB4QCM32, mutated all E-site tRNAs to tRNAVal, nucleotides 16 and 17 were mutated to C and 5MU, respectively). Further, additional force field parameters for ERY were added as described above. Force field parameters for ERY were obtained using the ANTECHAMBER module in AMBER. Parameters were converted for use in GROMACS 5.0 using ACPYPE64. Esther bonds between amino acids and the A76 of the A- and P-site tRNAs were treated as described earlier. Point charges and atom types for the backbone atoms of the uncharged amino acids lysine and alanine attached to the A-site tRNA were derived as described above for ERY.

**System setup**. In contrast to the cryo-EM structure, for the A-site tRNA19, nucleotides 16–17 were not modelled. To add the missing nucleotides, first, coordinates of nucleotides 15–18 were extracted from the tRNA52 in a ribosome EF–Tu complex structure57. Next, the extracted nucleotides were rigid-body-fitted to the cryo-EM structure using backbone atoms (P, C, O3′) of nucleotides 15 and 18. The fitted nucleotide C16 was mutated to a H2U and then nucleotides 16–17 were included in the model. In the cryo-EM structure, the tRNA54 in the E-site also lacked coordinates for nucleotides 16–17. These were extracted from tRNA55 (ref. 65) and rigid-body-fitted in the same manner as described for the A-site tRNA. Here, to match the tRNAVal, nucleotides 16 and 17 were mutated to C and 5MU, respectively. Further, the E-site tRNA lacked coordinates for the CCA-tail nucleotides. To model these nucleotides, first, tRNAVal, nucleotides 16 and 17 were mutated to C and 5MU, respectively. Further, the E-site tRNA lacked coordinates for the CCA-tail nucleotides. To model these
The N-terminal end of protein L27 (amino acids 1–10) was not resolved in the cryo-EM structure. To test whether the conformation of L27 in the pre-attack complex is stable in the presence of ERY, the N terminus was modelled on the basis of the crystal structure of a pre-attack state (T. thermophilus, PDB ID 1YV4 (ref. 32)). To that aim, first, the phosphates of rRNA nucleotides within a distance of 5 Å to amino acids 1–10 were extracted from the cryo-EM structure. L27 amino acids 1–10 were aligned to the cryo-EM structure by rigid-body fitting to the corresponding phosphate atoms. Finally, to match the E. coli sequence63, amino acids G6 and L7 were mutated to A and G, respectively.

The L1 protein and 23S rRNA H38 nucleotides 886–891 were not modelled in the cryo-EM structure. To model the L1 protein, coordinates of the L1 stalk rRNA (nucleotides 2992–3005) and of the tip of the L1 stalk rRNA (nucleotides 1869–1883) were extracted from the cryo-EM structure and from all trajectories from previous MD simulations of translation intermediates that include the L1 protein24. The coordinates from each frame of the trajectories were compared with those extracted from the cryo-EM structure by calculating the r.m.s.d. From the frame with the lowest r.m.s.d. (0.38 nm), the coordinates of the L1 protein were extracted and added to our model. To model the missing H38 nucleotides, first, nucleotides 883–888 and 892–893 as well as all residues within a radius of 1.2 nm were extracted from the cryo-EM structure and all trajectories. As described above, from the trajectories, the frame with the lowest r.m.s.d. (0.12 nm) was selected and the coordinates of nucleotides 886–891 were added to our model.

SD simulations. Starting structures of three states were used for the MD simulations: the model of the ribosome in complex with tRNAs, ErmBL and ERY (+ ERY); the model of the same complex with an alternative conformation of ErmBL (+ ERY); and the model after removal of ERY (– ERY). The alternative conformation of ErmBL (+ ERY) was obtained by perturbing the ψ dihedral of ErmBL-Trp265 and acidic Asp10 from 170° to 90°, as described in the following subsection. For each of the three starting structures, two independent simulations were carried out.

The protonation states of amino acids were determined using WHATTF86. Each structure was first solvated in a dodecahedron box keeping a minimum distance of 1.5 nm between the model atoms and the box boundaries. Next, the simulation system was neutralized with K+ ions and explicit salt (7 mM MgCl2 and 150 mM KC1) was added using the programme GENION from the GROMACS suite25. All MD simulations were carried out with GROMACS 5 (ref. 25). Lennard–Jones and short-range electrostatic interactions were calculated within a distance of 1 nm. Long-range electrostatic interactions, beyond 1 nm, were calculated by particle-mesh Ewald summation67 with a 0.12 nm grid spacing. The bond lengths were constrained using the LINCS algorithm68. The temperature of solute and solvent was controlled independently at T = 300 K using velocity rescaling29 with a coupling time constant of τf = 0.1 ps. An integration time step of 4 fs was used, applying virtual site constraints56. The coordinates were recorded for analysis every 5 ps.

The system was then equilibrated in four steps:

- Energy minimization using steepest decent,
- 0–50 ns: Berendsen barostat71, (τf = 1 ps) and position restraints on all the heavy atoms included in the cryo-EM structure (k = 1,000 kJ mol−1 nm−2);
- 50–70 ns: Linearly decreasing the position restraint force constant k to zero,
- 70–2,070 ns: Parrinello–Rahman barostat22, (τf = 1 ps) and no position restraints.

To study the effects of the ErmBL Lys11Ala mutation, the final structure of one trajectory for each state (t = 2,070 ns) was extracted. In this structure, the Lys11 of ErmBL was then mutated to an alanine. To preserve the neutral charge of the system, one randomly chosen water molecule, which was at least 2 nm away from any ribosomal atoms, was replaced by a K+ ion. The simulation system was subsequently, first, energy-minimized and, second, equilibrated with Berendsen barostat22 (as described above) for 20 ns (2,070–2,090 ns) and, finally, with Parrinello–Rahman barostat for 1,980 ns (2,090–3,070 ns).

Structural deviations. To monitor the stability of the simulations, the r.m.s.d. of all ribosome–ErmBL–tRNA complex atoms from their initial coordinates (cryo-EM structure) was calculated for each frame after rigid-body fitting of all Cα atoms (Supplementary Fig. 5).

To investigate whether, upon removal of ERY, the P-site A76 ribose moves towards its unstalled pre-attack state conformation, for each frame and simulation, the coordinates of the ribose and phosphate group atom of nucleotides C74–C75 of the P-site tRNA were rigid-body-fitted to their coordinates in the pre-attack structure24. Finally, the r.m.s.d. of the A76 ribose atoms from their pre-attack conformation was calculated (Fig. 6g in the main text).

ErmBL peptide dihedral angles. To monitor the conformational dynamics of the ErmBL peptide, dihedral angles for each amino acid were calculated (Supplementary Fig. 1). Particular, the ψ dihedral of Asp10 was found to spontaneously switch between the two starting conformations (+ ERY and + ERY). Therefore, we consider the simulations with ERY as exploring the same state. The results from the + ERY simulations support all the conclusions drawn from the − ERY simulations described in the main text. For clarity, these results are only reported as Supplementary Information.

Overlap of ErmBL with the volume occupied by ERY. To answer the question whether ERY restricts the conformation of the ErmBL peptide for each simulation and all frames at intervals of 1 ns, the overlapping volume between the peptide coordinates and the coordinates of ERY from the cryo-EM structure were calculated after alignment of the surrounding 23S rRNA phosphates. To that aim, the vander-Waals volume of ERY alone in the starting structure V(0)(t), of the ErmBL peptide together and of ERY and ErmBL amino acids 2–11 in the cryo-EM structure. Second, all trajectories were superimposed by least-square fitting using these P atoms. Then, the coordinates of the backbone atoms of nucleotides 2504–2506 were extracted from the superimposed trajectories, concatenated and the atomic displacement covariance matrix was calculated. Finally, the trajectory of each superimposition was projected on the first eigenvector of this matrix (Fig. 7a and Supplementary Fig. 9).

Distance of the crevice nucleotides from the A-site. The positively charged side chain of the ErmBL amino acid Lys11, which is attached to the A-site tRNA in the stalled complex, was found to interact with 23S nucleotides U2504, U2506 and C2452. The side chain specifically interacts with certain parts of these nucleotides: the U2504 ribose atoms, U2506 phosphate atoms and the C2452 base atoms. To measure the width of the A-site crevice57, first, the centres of mass of these nucleotide parts were calculated for each frame in each of the superimposed trajectories (see section Conformation of 23S nucleotides 2504–2506). Next, the mean position of the carbon atom of the ErmBL amino acid Asp10, which is attached to the P-site tRNA, was calculated from all superimposed trajectories. Finally, to quantify the width of the A-site crevice, the distance between these mean positions of the carbon atom and centre of geometry of the three centres of mass (U2504, U2506 and C2452) was calculated (d1 in Fig. 7c and Supplementary Fig. 9).

Interaction enthalpies of Lys11 and Ala11. To estimate the relative strength of the interaction of the A-site tRNA amino acids Lys11 and Ala11 with 23S nucleotides U2504, U2506 and C2452, the interaction enthalpies, that is, the sum of electrostatic and vander-Waals interactions were calculated. The electrostatic and vander-Waals interactions were calculated with the programmes and Lennard–Jones parameters from the amber99sb forcefield26 using GROMACS55. The sum of these interaction enthalpies is shown in Fig. 7i and Supplementary Fig. 9. Note that the estimated interaction strength calculated here represents a qualitative estimate of the free energy differences and does not include entropic contributions.

Hydrogen bonds. Hydrogen bonds of the A76 of A-site and P-site tRNAs and ErmBL amino acids 6–11 were monitored for all simulations using the GROMACS programme gmx hbonds55. For all hydrogen bonds with an occupancy > 20% in at least one simulation, the hydrogen bond occupancy was averaged over windows of 20 ns (Supplementary Fig. 6).

Data availability. The cryo-EM maps and associated atomic coordinates have been deposited in the EMDB and PDB with the accession codes EMDB-8175 and PDB ID 5TJE (ErmBL–APE–SRC) and EMDB-8176 and PDB ID 5U8 (ErmBL–AP–SRC). The additional MD data that support the findings of this study are available from the corresponding author upon request.

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Author contributions

S.A. prepared ErmBL-SRC and performed single-particle cryo-EM analysis. S.A. and C.A.I modelled and refined the ErmBL-SRC. L.V.B., H.G. and A.C.V. performed MD simulations. M.G. performed toe-printing analysis. All authors analysed data, prepared figures and wrote the paper.

Additional information

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