MyD88-deficient bone marrow cells accelerate onset and reduce survival in a mouse model of amyotrophic lateral sclerosis

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Increasing evidence suggests that neurotoxicity of secreted superoxide dismutase 1 (SOD1) mutants is associated with amyotrophic lateral sclerosis (ALS). We show here that mutant SOD1 protein activates microglia via a myeloid differentiation factor 88 (MyD88)–dependent pathway. This inflammatory response is also associated with a marked recruitment of bone marrow–derived microglia (BMDM) in the central nervous system. We then generated chimeric SOD1G37R and SOD1G93A mice by transplantation of bone marrow (BM) cells from MyD88-deficient or green fluorescent protein (GFP)–expressing mice. SOD1G37R mice receiving MyD88−/− BM cells exhibit a significantly earlier disease onset and shorter lifespan compared with mice transplanted with control GFP cells. This compelling beneficial effect of MyD88–competent BMDM is a previously unrecognized natural innate immune mechanism of neuroprotection in a mouse model of late-onset motor neuron disease.

Introduction

Amyotrophic lateral sclerosis (ALS), also known as Lou Gehrig’s disease in the United States, is an adult-onset neurodegenerative disease affecting primarily motor neurons in the brain and spinal cord. The loss of motor neurons leads to the progressive atrophy of skeletal muscles and, ultimately, paralysis and death within 3–5 yr after symptom onset. About 90% of ALS cases are sporadic with no known genetic component and 10% are familial. Missense mutations in the gene encoding copper/zinc superoxide dismutase 1 (SOD1) are associated with 20% of familial ALS (Rosen et al., 1993). Transgenic mice overexpressing mutant SOD1 develop a motor neuron disease resembling ALS (Gurney et al., 1994). The inbred C57BL/6 SOD1G37R mice (line 29) have a life span of ~11–12 mo (late onset), whereas C57BL/6 SOD1G93A mice exhibit a life span of ~4–5 mo (early onset). Although these mice are two well-accepted animal models of ALS, the mechanisms leading to neurodegeneration may differ among them.

The mechanism by which SOD1 mutants cause motor neuron death is still unknown. To date, the most promising hypothesis is that the toxicity of mutant SOD1 results from the propensity of the misfolded protein mutants to aggregate rather than the aberrant copper-mediated catalysis (Johnston et al., 2000; Julien, 2001). Although SOD1 is traditionally regarded as a cytosolic protein, it is noteworthy that both wild-type (WT) and mutant SOD1 proteins can be secreted (Turner et al., 2005). A recent study proposed the selective secretion of mutant SOD1 mediated by chromogranins, which constitutes a potentially toxic pathway that can induce inflammation and neuronal death (Urushitani et al., 2006). Mutant SOD1-mediated toxicity is non–cell autonomous. The specific expression of mutant SOD1 in neurons, astrocytes, or microglia could not provoke motor neuron disease (Gong et al., 2000; Pramatarova et al., 2001; Beers et al., 2006). Over the years, increasing evidence has indicated the importance of a motor neuron milieu and the contribution of nonneuronal cells to neurodegeneration in ALS. Clement et al. (2003) demonstrated that WT nonneuronal cells delay degeneration and significantly extend survival of mutant-expressing motor neurons. More recently, it was reported that the reduced mutant SOD1 expression in microglia contributes to motor neuron protection (Boillee et al., 2006) and that WT microglia extend survival of SOD1G93A mice deficient in PU.1 (Beers et al., 2006).

Microglia are the main immune cells of the central nervous system (CNS). They produce numerous inflammatory mediators, which are detected in the CNS of both mouse models and ALS patients (Nguyen et al., 2002). Up-regulation of toll-like...
Figure 1. Exogenous recombinant G93A form of SOD1 protein induces expression of gene encoding proinflammatory molecules. (A) Representative photomicrographs of in situ hybridization signals showing the mRNA expression of TLR2, MCP-1, and IL-1β in the brain of adult C57BL/6J mice at 2 mo of age. Mice received a single intracerebral injection with 1 μl saline, 1 μg/μl of recombinant WT SOD1, or 1 μg/μl G93A mutant SOD1 proteins. Mice were killed 24 h after the injection. The coronal sections were taken from x-ray films. (B) Quantitative analysis of TLR2, MCP-1, and IL-1β mRNA expression. Results represent means ± the SEM of three or four mice per group. Asterisk indicates a significant difference (P < 0.05) from the other groups. (C) The same treatments were given to MyD88−/− mice. Representative in situ hybridization signals in the brain of MyD88−/− mice. (D) Quantitative analysis of mRNA expression in MyD88−/− mice. (E) Expression of the gene encoding PLP1 (a marker of oligodendrocytes) in the brains of adult C57BL/6J that received saline, recombinant WT SOD1, or G93A mutant SOD1 proteins. No sign of demyelination was found. (F) FJB staining was...
receptor 2 (TLR2), a reliable index of proinflammatory signaling in microglia/macrophages, also takes place in the spinal cord of SOD1G37R mice (Nguyen et al., 2004). Myeloid differentiation factor 88 (MyD88) is an adaptor protein that plays a critical role in mediating nuclear factor κB signaling and cytokine gene expression (Akira et al., 2006). The innate immune system is seriously compromised in MyD88-deficient mice (Adachi et al., 1998). Macrophages from MyD88 knockout mice did not produce interleukin (IL) 6 or TNF-α in response to lipopolysaccharide (Kawai et al., 2001). Mice deficient in MyD88 (MyD88−/−) have impaired IL-1– and IL-18–mediated functions and defects in T-cell proliferation (Adachi et al., 1998). We used MyD88−/− mice to study the role of this pathway in the pathogenesis of this motor neuron disease.

We show here that extracellular mutant SOD1 induces microglial activation in vivo via a MyD88-dependent pathway. Our data from chimeric mice suggest a critical neuroprotective role of MyD88 in bone marrow–derived microglia (BMDM) in SOD1 mutant mice.

**Results**

**Extracellular SOD1 mutant G93A induces inflammation without being neurotoxic**

Adult male C57BL/6J mice at 2 mo of age were given an intracerebral injection of saline, 1 μg of recombinant human SOD1 WT, or 1 μg SOD1 mutant G93A protein. Mice were killed 3 or 24 h after the infusion. In situ hybridization was then performed across the brains of the mice using the probes for TLR2, monocyte chemoattractant protein 1 (MCP-1), TNF-α, IL-1β, IκBα, and IL-12 genes. 3 h after the treatment, the hybridization signals were comparable between different treatments (unpublished data). However, 1 d after the injection, the signal for TLR2, MCP-1, and IL-1β mRNA was higher in the brain of mice that received mutant G93A compared with WT SOD1 and saline (Fig. 1, A and B). Such an increase in gene expression was significantly prevented in MyD88-deficient mice (Fig. 1, C and D). It is interesting to note that the IL-1β mRNA signal remained undetectable in the brain of MyD88−/− mice. The basal levels of TLR2 were significantly higher in MyD88−/− compared with WT mice. These data suggest that SOD1 mutant G93A induces proinflammatory signaling via the MyD88 pathway. WT SOD1 and G93A caused a more variable expression pattern of the other immune genes measured in the present experiments. They were therefore not assessed in the brain of MyD88-deficient mice.

We next tested whether such inflammatory reaction was dependent on the potential neurotoxicity properties of the mutant protein. The expression of proteolipid protein 1 (PLP1; a marker of altered oligodendrocytes) and fluoro-Jade B (FJB; neurodegeneration) was not different in the brains of mice administered either saline, WT SOD1, or mutant SOD1 (Fig. 1, E–G). Except for the damage caused by the cannula (Fig. 1, F and G), there was no anatomical evidence of demyelination or neurodegeneration after intracerebral administration of the mutant protein. These data indicate that activation of microglia is not a consequence of the neurotoxic effects of mutant SOD1 but a direct action of the protein on these immune cells.

**Infiltration of BMDM by mutant SOD1 and SOD1 mice**

To determine whether this immune reaction caused infiltration of BMDM, we generated chimeric mice by transplanting GFP bone marrow (BM) cells into irradiated WT mice. 3 mo later, these mice received a single intracerebral infusion of either 1 μl saline, 1 μg/μl of recombinant human SOD1 WT, or 1 μg/μl SOD1 mutant G93A protein. 7 d after the injection, a significantly greater number of GFP cells was found in the brains of mice that received G93A protein compared with those challenged with SOD1 WT protein or saline (Fig. 2, A and B). Colocalization of green autofluorescence from GFP BM cells with Iba-1 immunostaining (Fig. 2 C, red) provided clear anatomical evidence that these GFP cells differentiated into microglia and not other cell types (Fig. 2 C). The G93A mutant is therefore able to activate microglia and provoke infiltration of BMDM.

To determine whether these events also take place in the CNS of mouse models of ALS, we generated chimeric mice by transplanting BM cells expressing GFP into irradiated SOD1G37R and SOD1G93A transgenic mice. Mice were killed 3, 6, and 8 mo after transplantation (SOD1G37R; Fig. S1, available at http://www.jcb.org/cgi/content/full/jcb.200705046/DC1) or at the end stage (Fig. 3). A limited number of GFP cells were found in various regions of the CNS in both SOD1G37R and WT groups of mice that were killed 3 and 6 mo, respectively, after being transplanted with BM cells. Numerous GFP cells were detected in the spinal cord and brain stem of SOD1G37R animals at 8 mo of age (Fig. S1). At the end stage, BMDM massively infiltrated all affected regions of SOD1G37R and SOD1G93A chimeric mice (Fig. 3 A). All these GFP cells were immunoreactive for Iba-1 in the brain and spinal cord of SOD1G37R and SOD1G93A chimeric mouse models (Fig. 3 B). The data indicate that the infiltration process begins in the spinal cord and slowly progresses to more rostral degenerating regions and that all these cells differentiate into microglia in the CNS of mouse models of ALS.

**MyD88−/− BM-derived cells significantly affect the onset and lifespan of SOD1G37R mice**

Based on the critical role of the MyD88 pathway in the activation of microglia by mutant SOD1 and the marked recruitment of BMDM in SOD1G37R and SOD1G93A chimeric mice, we attempted to explore the role of MyD88-competent BMDM. SOD1G37R and SOD1G93A transgenic mice were irradiated at 2 mo of age and transplanted with MyD88−/− or GFP BM stem cells. C57BL/6J WT mice transplanted with MyD88−/− or GFP BM stem cells...
were used as controls. We defined disease onset by the age at which a 30% decrement in motor performance was measured using a rotarod device. There was no significant difference in either symptom onset or lifespan between GFP and SOD1 G93A chimera (GFP−SOD1 G93A) and MyD88−/−−SOD1G93A groups (unpublished data). Both groups of mice became paralyzed 1 mo after transplantation. However, SOD1 G37R mice receiving MyD88−/−BM cells exhibited a significantly earlier disease onset and shorter lifespan compared with mice receiving GFP cells. The disease onset occurred at the mean age of 283 ± 2.76 d in MyD88−/−−SOD1 G37R mice compared with 327 ± 4.13 d in GFP−SOD1 G37R mice (Fig. 4 A; P < 0.001). A remarkable difference was also found in lifespan, with a mean of 297 ± 2.4 d for MyD88−/−−SOD1 G37R mice and 348 ± 3.5 d for GFP−SOD1 G37R mice (Fig. 4 B; P < 0.001). However, the disease duration was not significantly different between these two groups (Fig. 4 C; P > 0.05). Consistent with their earlier disease onset, MyD88−/−−SOD1 G37R mice started to lose body weight earlier than GFP−SOD1 G37R mice (Fig. 4 D). The mean body weight of GFP−SOD1 G37R mice was not as high as nonirradiated SOD1 G37R mice, suggesting a potential side effect of irradiation and BM transplantation process but not on the lifespan. GFP−WT and MyD88−/−−WT mice did not exhibit any signs of paralysis (unpublished data).

Loss of motor neurons and axons in MyD88−/−−SOD1 G37R chimeric mice

To compare the number of motor axons and neurons between the different groups, MyD88−/−−SOD1 G37R chimera were killed at the end stage together with MyD88−/−−WT and GFP−SOD1 G37R chimeric mice. All the groups therefore had the same age but only MyD88−/−−SOD1 G37R animals were at the end stage.
L5 roots and lumbar spinal cords were taken from these mice and processed for the quantification of motor axons and neurons. As depicted by Fig. 5 (A and E), MyD88−/−–SOD1G37R had significantly fewer motor axons and neurons than MyD88−/−–WT and GFP–SOD1G37R chimeric mice. MyD88−/−–SOD1G37R mice had 230 ± 15 axons, whereas 787 ± 36 axons were counted in GFP–SOD1G37R mice (P < 0.05) and 1,001 ± 21 axons in MyD88−/−–WT (P < 0.001). Nissl-stained sections were used for the quantification of motor neurons in the ventral horn of lumbar spinal cords. Here again, the number of motor neurons in MyD88−/−–SOD1G37R mice (6.8 ± 0.1) was significantly lower compared with that of MyD88−/−–WT (17.4 ± 0.5) and GFP–SOD1G37R (14.7 ± 0.9) chimeric mice (Fig. 5, B and F).

It is interesting to note the strong microgliosis in the spinal cord of SOD1G37R, especially in MyD88−/−–SOD1G37R chimeric mice (Fig. 5, C and D). Both the number of microglial cells and

**Figure 3.** Widespread distribution of BMDM in SOD1 chimeric mice. SOD1G37R and SOD1G93A mice were transplanted with BM cells from GFP transgenic mice. Spinal cords and brains were taken at the end stage of the animals. (A) GFP-positive cells were detected in the lumbar spinal cord and throughout the affected regions of these two types of chimeric mice. (B) Confocal images show that BM-derived GFP-positive cells (green) in the brain and spinal cord are all microglial cells (Iba-1, red; Cy3) in both types of chimeric mice. DAPI was used to stain nuclei (blue). Bars: [A] 100 μm; [B] 20 μm.
the hybridization signal for the gene encoding the innate receptor TLR2 were significantly higher in mice that were transplanted with MyD88-deficient BM cells (Fig. 5, C, D, and G). Transcriptional activation of TLR2 is a reliable marker of the innate immune response by microglia (Laflamme et al., 2001; Nguyen et al., 2002). Whether such a microgliosis contributes or is a consequence of the rapid and marked neurodegeneration in MyD88−/−–SOD1G37R mice remains an open question.

Role of MyD88 in SOD1G93A and SOD1G37R mice

Because MyD88 plays a critical role in the BMDM of SOD1G37R animals and MyD88−/−–SOD1G37R chimeric mice exhibited a robust innate immune reaction in the spinal cord (Fig. 5, C and D), we created SOD1G93A and SOD1G37R mice in the context of a MyD88 gene knockout. However, we were not able to generate a single SOD1G93A;MyD88−/− mouse after >1 yr of crossbreeding. It is possible that MyD88 is critical for the development and survival of these mice because of the high copy number of mutant SOD1 in the SOD1G93A line, which is a rapid and severe mouse model of motor neuron degeneration. In contrast, we were able to generate SOD1G37R;MyD88−/− mice, but no significant difference was detected in the disease onset, duration, or survival between G37R+/−;MyD88−/− and G37R+/+;MyD88−/+; MyD88+/−; MyD88−/+ mice were much lower than the other groups of mice (Fig. 6 D).

Characteristics of the spinal cord from the crossed mice

Crossed mice were killed at 8 mo (presymptomatic stage), 11 mo (symptomatic stage), and the end stage. To examine whether the context of MyD88 deficiency affects motor neurons, Nissl staining was performed and motor neurons were counted in the ventral horn of lumbar spinal cords. No significant difference was found between G37R+/−;MyD88+/+ and G37R+/−;MyD88−/+ mice at the presymptomatic stage. Although not statistically significant, G37R+/−;MyD88−/+ mice appeared to have fewer motor neurons than G37R+/−;MyD88+/+ mice at 11 mo. However, G37R+/−;MyD88−/+ mice had significantly fewer motor neurons in the lumbar spinal cord compared with G37R+/−; MyD88−/+ mice at the end stage of the disease (P < 0.05; Fig. 7 A). Immunohistochemistry with the use of microglial cell marker anti–rabbit Iba-1 did not reveal any major differences in the number of microglia between these two groups at the presymptomatic stage, symptomatic stage, or end stage (Fig. 7 B). TLR2 mRNA expression was found to be similar at the presymptomatic and symptomatic stages between G37R+/−; MyD88−/+ and
G37R+/±:MyD88+/+ groups, but the signal was much higher in the lumbar spinal cord of G37R+/−:MyD88−/− compared with G37R+/−:MyD88+/+ at the end stage of the disease (P < 0.05; Fig. 7 C).

Discussion

Although ALS was first described in 1869, the mechanisms involved in the pathogenesis of this neurodegenerative disease still remain largely unknown. Several mechanisms that are not mutually exclusive have been proposed. These include oxidative stress, glutamate-induced excitotoxicity, cytoskeletal abnormalities, protein aggregation, mitochondrial dysfunction, and, more recently, inflammation. The role of inflammation and microglia in ALS and other CNS diseases is currently a matter of great debate and controversy. Although numerous cytokines are up-regulated in microglia of mice expressing mutant SOD1 transgenes, deletion in the gene encoding IL-1β and TNF does not change the outcomes of the diseases (Nguyen et al., 2001; Gowing et al., 2006). However, SOD1G37R mice that had MyD88-competent BMDM developed the disease later, survived longer, and had less neurodegeneration than those that were transplanted with MyD88-deficient BM cells. These data suggest a novel neuroprotective role of competent BMDM in a mouse model of ALS.

The discovery that ~20% of familial ALS cases are caused by mutations in SOD1 has enabled the development of animal
and cell culture models and led to much of our current understanding of the neurodegenerative mechanisms in ALS. SOD1 is a ubiquitously expressed protein, which protects cells from damage by free radicals. ALS often starts focally. It is still unclear how the toxicity of mutant SOD1 is propagated from one localized group of cells to another. Based on the recent evidence that extracellular mutant SOD1 proteins could be selectively secreted and trigger microgliosis and neuronal death in cultured cells (Urushitani et al., 2006), we injected WT SOD1 and mutant proteins into the mouse CNS. G93A mutant protein was used in our experiments because it was the only SOD1 mutant protein available to us. In agreement with this study, our data demonstrate that mutant SOD1 stimulates inflammation and recruitment of BMDM. WT SOD1 protein was used as a control to ascertain that these effects were specific to the mutant protein and not to potential traces of endotoxin. The inflammatory response caused by G93A is largely dependent on MyD88 signaling but it is still not known whether this protein binds to specific immune receptors in microglia. However, G93A is able to stimulate TNF production from the microglial cell line (Urushitani et al., 2006) and an acute intracerebral infusion of this protein is not neurotoxic. We therefore suggest that the inflammatory properties of mutant SOD1 are directly mediated by the MyD88 pathway in microglia and not via other products released by dying cells.

Previous studies have shown that in chimeric SOD1G93A mice, BM-derived cells differentiate into microglia in the spinal cord and brain. In addition, it has been suggested that the number of GFP-positive cells in the spinal cord is associated with disease progression (Corti et al., 2004; Solomon et al., 2006). In this paper, we generated chimeric SOD1G37R and SOD1G93A mice and confirmed the extensive distribution of GFP-positive cells throughout the brain and spinal cord in both mouse models of ALS. In accordance with the previous studies, we observed few GFP-positive cells in the CNS before the disease onset (Fig. S1), but the number of GFP cells greatly increased during disease progression (Table S1, available at http://www.jcb.org/cgi/content/full/jcb.200705046/DC1). More importantly, transplantation of BM from GFP-expressing mice does not affect disease progression of either SOD1G93A or SOD1G37R mice when compared with their respective nonirradiated SOD1 groups. In contrast, transplantation of MyD88-deficient BM cells dramatically changed the disease onset and progression only in mice that express human mutant G37R.

The rationale to investigate the role of this adaptor protein was based on our initial observation that the MyD88 pathway mediates microglial activation and infiltration induced by mutant SOD1. The results from the chimeric mice suggest that BMDM acts as a natural defense mechanism against secreted mutant SOD1.
Indeed, MyD88−/− BM transplantation led to the earlier disease onset and shorter lifespan of SOD1G37R mice compared with mice that received GFP cells. GFP–SOD1G37R and nonirradiated SOD1G37R mice were used as controls to exclude the possibility that these effects were attributable to irradiation. Histological analysis revealed a significant motor neuron and axon loss in MyD88−/−–SOD1G37R mice compared with the GFP–SOD1G37R and MyD88−/−–WT mice at the same age, which explains the intriguing earlier disease onset and death of MyD88−/−–SOD1G37R mice. We also found a more robust innate immune reaction in the spinal cord of these mice.

It has been shown that WT nonneuronal cells delayed disease onset by a mean of 1.2 mo for SOD1G37R chimeras and extended their survival by 1.1 mo (Clement et al., 2003). A recent study demonstrated that substitution of WT microglia for SOD1G93A-expressing microglia prolonged the survival and disease duration of SOD1G93A mice but had no effect on the onset (Beers et al., 2006). However, reduced levels of mutant SOD1 in microglia did not change onset and the early disease phase but clearly slowed later paralysis (Beers et al., 2006; Boilée et al., 2006). These data do not necessarily contradict the experiments using MyD88-deficient mice. Indeed, the disease progression may be influenced by the levels of extracellular mutant SOD1 that resident microglia contribute to this extracellular pool. This may be an explanation for the neuroprotective properties of SOD1-deficient microglia and such effects may not be associated with the immune functions of these cells.

We have previously reported the existence of different populations of microglia that may have somewhat opposite roles. The double-edged sword of these cells that has been intensively reviewed in the past few years may also depend on the origin of microglia in the adult CNS. BMDM are very efficient in restricting amyloid deposits in a mouse model of Alzheimer’s disease, whereas their resident counterparts seem unable to phagocyte this toxic protein (Simard et al., 2006). The results from this paper support this concept and imply that impairment of such natural function of BMDM accelerates the neurodegenerative properties of secreted mutant SOD1. It is interesting to note that transplantation of MyD88-deficient BM cells did not change onset or survival of mice expressing human SOD1G93A.

These mice reach paralytic end stage ~2 mo after BM transplantation. The hematopoietic system takes 7–9 wk to be fully restored after lethal irradiation and the percentage of GFP- or MyD88-deficient cells is low during the first 4 wk of the chimera. This may explain why these cells are unable to modulate the disease progression in such an early onset model of neurodegeneration. This is not the case in mice expressing SOD1G37R because restoration of BMDM is completed in these animals several months before the first symptomatic signs. The early onset may therefore explain why MyD88−/− BM cells failed to change the mean life expectancy of SOD1G93A.

Because of the potent beneficial role of WT BMDM in SOD1G37R mice, the lack of significant difference in the disease onset and lifespan between G37R+/−, MyD88+/+ and G37R+/−, MyD88−/− at the presymptomatic, symptomatic, or end stage (P > 0.05). [C] In situ hybridization was performed using antisense probe for TLR2 gene. Quantitative analysis of TLR2 mRNA expression showed a significant difference between the groups G37R+/−, MyD88+/+ and G37R+/−, MyD88−/− at the end-stage (P < 0.05). These data indicate that microglial cells were more activated in G37R+/−, MyD88−/− mice.

Figure 7. Number of motor neurons and microglia in the spinal cord of SOD1G37R mice in a MyD88 knockout context. Crossed mice were killed at 8 mo (presymptomatic stage), 11 mo (symptomatic stage), and the end stage. Results represent mean ± the SEM of four mice per group. (A) Motor neurons were counted in the lumbar spinal cord. Note a significant difference between the groups of G37R+/−, MyD88+/+ and G37R+/−, MyD88−/− at the end stage (P < 0.05). (B) Immunohistochemical staining of lumbar spinal cord with the use of microglial cell marker anti–rabbit Iba-1 followed by the incubation with biotinylated secondary antibody. The numbers of microglial cells were estimated and expressed as the number of cells per cubic millimeter. No significant difference was found between the groups of G37R+/−, MyD88+/+ and G37R+/−, MyD88−/− at the presymptomatic, symptomatic, or end stage (P > 0.05). (C) In situ hybridization was performed using antisense probe for TLR2 gene. Quantitative analysis of TLR2 mRNA expression showed a significant difference between the groups of mice at the presymptomatic stage (P < 0.05). These data indicate that microglial cells were more activated in G37R+/−, MyD88−/− mice.
models of ALS. A recent study has shown that sodium valproate, the histone deacetylase inhibitor, exerts neuroprotective effects both in vitro and in vivo but it does not improve the survival of SOD1G37R mice (Rouaux et al., 2007).

It is also important to mention that G37R+/−:MyD88−/− mice were much smaller than their littermates based on weekly body weight. In addition, the numbers of pups per carriage was lower in G37R+/−:MyD88−/− mice, with a mean of 4.5 compared with 7 pups from SOD1, 6 from MyD88−/−, and 8 from WT mice. Moreover, we were not able to generate a single G93A−/−:MyD88−/− mouse after >1 yr of crossbreeding between SOD1G37R and MyD88-deficient mice. We have not been able to generate homozygote MyD88−/− mice in using another mouse model of brain disease. Indeed, for >2 yr, we have attempted to breed amyloid precursor protein (APP)/presenelin 1 and APP/presenelin 1. Compared with SOD1G93A, the lower level viable in the presence of highly toxic proteins, such as G93A mouse. This indicates that these MyD88 homozygotes are not G93A mice were much smaller than their littermates based on weekly 06:00 and off at 20:00) with free access to rodent feed and water. All animal protocols were approved by the Laval University Animal Welfare Committee in accordance with the Canadian Council on Animal Care guidelines.

BM transplantation
Irradiation and BM transplantation were performed as described previously (Simard and Rivest, 2004; Simard et al., 2006). In brief, mice at 2 mo of age were exposed to 10 gray total-body irradiation using a cobalt-60 source (Theratron-780; MDS Analytical Technologies). A few hours later, the animals were injected via a tail vein with ∼14 × 10⁶ BM cells freshly collected and purified from donor mice. GFP or MyD88−/− mice at 3–5 mo of age were used as cell donors. Irradiated mice transplanted with BM cells were housed in autoclaved cages and treated with antibiotics (0.2 mg trimethorpine and 1 mg sulfamethoxazole per 1 ml of drinking water was given 7 d before and 2 wk after irradiation).

Intracerebral injection
Animals were anesthetized with isoflurane (Baxter Healthcare Corporation) and placed on a stereotaxic apparatus (David Kopf Instruments). The injection site was reached using a small cannula (28 gauge; Plastic One) at the coordinates −2.0 mm lateral and −3.0 mm dorsoventral from the bregma. The animals received an infusion of 1 μl of either sterile saline, 1 μg/μl of recombinant human WT SOD1, or 1 μg/μl of recombinant G93A mutant of human SOD1 protein (provided by M. Urrutia, Shiga University of Medical Science, Shiga, Japan) solution over a period of 2 min by a microinjection pump (A-99; Razel Scientific Instruments). The animals were killed at different time points (3, 24, or 168 h) after the injection.

Materials and methods

Animals
Adult male C57BL/6J mice were purchased from the Jackson Laboratory at 2 mo of age. Hemizygous transgenic mice expressing GFP under the control of the chicken β-actin promoter and cytomegalovirus enhancer were initially also obtained from the Jackson Laboratory. A colony was then established and maintained in a C57BL/6J background. Transgenic mice carrying G37R (line 29) and those harboring the G93A mutant of human SOD1 (B6SJLtGcNSOD1-G93A) were obtained from J.P. Julien (Laval University, Québec, Canada). MyD88−/− mice (in C57BL/6 background) were provided by S. Akira (Osaka University, Osaka, Japan). SOD1G37R and SOD1G93A mice were crossed with MyD88−/− mice to generate G93A−/−:MyD88−/− and G37R−/−:MyD88−/− mice by breeding G93A−/− and G37R−/− with MyD88−/− mice, yielding offspring that were heterozygous for the MyD88 gene. The heterozygous offspring were intercrossed to obtain breeding pairs of the following genotypes: G37R−/−:MyD88−/−, G37R+/−:MyD88−/−, MyD88−/−, and G37R−/−:MyD88+/−. With the breeding procedure, we were not able to generate G93A−/− in a context of MyD88 knockout (see Discussion). Animals were acclimated to standard laboratory conditions (14 h light and 10-h dark cycle; lights on at 06:00 and off at 20:00) with free access to rodent feed and water. Tissue collection
Mice were deeply anesthetized via an intraperitoneal injection of a mixture of ketamine hydrochloride and xylazine and rapidly perfused transcardially with 0.9% saline followed by ice-cold borax-buffered 4% PFA, pH 9.5. Brains and spinal cords were rapidly removed from the animals, postfixed for 2–3 d, and placed in a solution containing 10% sucrose diluted in 4% PFA-borax buffer overnight at 4°C. The frozen tissue were mounted on a microtome (Reichert-Jung; Cambridge Instruments Company) and cut into 25-μm coronal sections. The slices were collected in a cold cryoprotectant solution (0.05 M sodium phosphate buffer, pH 7.3, 30% ethylene glycol, and 20% glycerol) and stored at −20°C.

In situ hybridization, immunohistochemistry, and immunofluorescence
In situ hybridization, immunohistochemistry, and immunofluorescence were performed as described previously (Laframme et al., 1999; Millecamps et al., 2006; Simard et al., 2006). 35S-labeled complementary RNA probes for IL-1β, TNF-α, MCP-1, IL-6, and mouse PLP1 were used for in situ hybridization. For immunohistochemistry and immunofluorescence, rabbit polyclonal anti-ionized calcium binding adaptor molecule 1 (Iba-1, 1:2,000; Wako Chemicals USA) was used as the primary antibody. Microglia that were immunoreactive for Iba-1 were counted in the lumbar spinal cord using unbiased stereological techniques. The density of labeled cells was estimated by the optical fractionator method using Stereo Investigator software (MBF Bioscience). The lumbar spinal cord was traced with a C. The frozen tissues were mounted on a microtome (Reichert-Jung; Cambridge Instruments Company) and cut into 25-μm coronal sections. The slices were collected in a cold cryoprotectant solution (0.05 M sodium phosphate buffer, pH 7.3, 30% ethylene glycol, and 20% glycerol) and stored at −20°C.

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Mice were deeply anesthetized via an intraperitoneal injection of a mixture of ketamine hydrochloride and xylazine and rapidly perfused transcardially with 0.9% saline followed by ice-cold borax-buffered 4% PFA, pH 9.5. Brains and spinal cords were rapidly removed from the animals, postfixed for 2–3 d, and placed in a solution containing 10% sucrose diluted in 4% PFA-borax buffer overnight at 4°C. The frozen tissue were mounted on a microtome (Reichert-Jung; Cambridge Instruments Company) and cut into 25-μm coronal sections. The slices were collected in a cold cryoprotectant solution (0.05 M sodium phosphate buffer, pH 7.3, 30% ethylene glycol, and 20% glycerol) and stored at −20°C.

In situ hybridization, immunohistochemistry, and immunofluorescence
In situ hybridization, immunohistochemistry, and immunofluorescence were performed as described previously (Laframme et al., 1999; Millecamps et al., 2006; Simard et al., 2006). 35S-labeled complementary RNA probes for IL-1β, TNF-α, MCP-1, IL-6, and mouse PLP1 were used for in situ hybridization. For immunohistochemistry and immunofluorescence, rabbit polyclonal anti-ionized calcium binding adaptor molecule 1 (Iba-1, 1:2,000; Wako Chemicals USA) was used as the primary antibody. Microglia that were immunoreactive for Iba-1 were counted in the lumbar spinal cord using unbiased stereological techniques. The density of labeled cells was estimated by the optical fractionator method using Stereo Investigator software (MBF Bioscience). The lumbar spinal cord was traced with a 10× Plan Apochromat objective and sampled using a 40× Plan Apochromat objective (Nikon). The counting parameters were the distance between counting frames (300 μm), the counting frame size (100 × 100 μm), the dissector height (13 μm), and the guard zone thickness (1.5 μm).

Morphological and morphometric analysis
Mice were deeply anesthetized, perfused with 0.9% saline, and fixed with 3% glutaraldehyde in PBS buffer, pH 7.4. L5 roots and dorsal root ganglion tissue samples were immersed in fixative overnight, rinsed in PBS buffer, and postfixed in 1% osmium tetroxide. After three washes with PBS buffer, samples were dehydrated in a graded series of ethanol and embedded in Epon (Marivac). The thin sections of L5 ventral root were stained with toluidine blue and examined under a light microscope. Axons in the L5 ventral root were counted with Stereo Investigator software.
FJB staining
FJB method was performed as described previously [Turin and Rivest, 2006]. In brief, mounted brain sections were dried under a vacuum, dehydrated through graded concentrations of alcohol (50, 70, and 100% for 1 min), rehydrated through graded concentrations of alcohol (100, 70, and 50% for 1 min), and rinsed for 1 min in distilled water. They were then dipped and shaken in 0.06% potassium permanganate for 10 min, rinsed for 1 min in distilled water, dipped, and shaken in a solution containing 0.004% FJB (Histochem), 0.1% acetic acid, and 0.0002% DAPI (Invitrogen) for 20 min. The slides were thereafter rinsed three times in distilled water for 1 min each, dried, dipped in xylene three times for 2 min each, and coverslipped with distrene plasticizer xylene mounting media [Electron Microscopy Sciences]. The number of FJB-positive cells was counted and estimated with Stereo Investigator software.

Nissl staining and motor neuron counting
Mouse lumbar spinal cord was cut into 25-μm-thick transversal sections. The sections were washed in distilled water, dehydrated through graded concentrations of alcohol (50, 70, 95, and 100%) to xylene, rehydrated through graded concentrations of alcohol (100, 95, 70, and 50%), and then stained with a 0.25% thionin solution. Thereafter, the sections were dehydrated and coverslipped. After the staining, the large motor neurons in the ventral horn were counted with Stereo Investigator software. Only large multipolar motor neurons with a cross-sectional area ≥250 μm² were considered [Fischer et al., 2004].

Assessment of motor function
The motor function was tested using a rotarod device (10 rpm; Economex Controller; Columbus Instruments) weekly. The amount of time the mice remained on the rotarod was recorded for up to 180 s. The trial was conducted three times for each mouse and the best result of these trials is used as the riding time of each mouse.

Confocal laser scanning and regular microscopy
For cell phenotyping, tissue samples were analyzed for colocalization with cell type–specific markers using a confocal laser scanning microscope (BX-61) equipped with imaging software [Fluoview VS500 4.3; both from Olympus]. Confocal images were acquired with a 60× Plan Apochromat 1.4×/immersion objective [NA 1.35; Olympus] by sequential scanning using a two-frame Kalman filter, low speed scans, and a z separation of 0.40 μm. Regular 2D images were captured on a microscope [C-80; Nikon] fitted to a digital camera (Retiga EXI Fast, Qimaging) and a super-high-pressure mercury lamp [Nikon]. The images were then processed to enhance contrast and sharpness using Photoshop CS2 and figures were assembled using Illustrator CS [both from Adobe].

Quantitative analysis
Quantitative analyses of hybridization signals were performed as described previously [Glezer et al., 2003]. The intensity of mRNA signals were measured on x-ray film [Biomax RMF 72554]. The intensity of mRNA signals was estimated with Stereo Investigator software.

Statistical analysis
The data were analyzed by one-way analysis of variance followed by Bonferroni’s post hoc analysis. P < 0.05 was considered statistically significant.

Online supplemental material
Fig. S1 shows distribution of BMDM in WT and SOD1G37R chimeric mice. Table S1 shows recruitment of BMDM (GFP+) /Iba-1+) in the CNS of GFP–SOD1G37R mice at 3, 6, and 8 mo after transplantation. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.200705046/DC1.

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