Cartilage shavings collected from equine metacarpophalangeal joints

Five 3 mm diameter explant discs cut and placed in 1 ml serum free DMEM to establish explant cultures.

- Untreated (DMEM alone)
- IL-1β (10 ng/ml)
- Carprofen (100 µg/ml)
- Carprofen + IL-1β

Trypsin protein digestion

DMMB assays to assess GAG release

Quantitative western blots of selected proteins

High throughput shotgun MS/MS

Protein identification via MASCOT database search

Carprofen inhibits the release of matrix metalloproteinases 1, 3, and 13 in the secretome of an explant model of articular cartilage stimulated with interleukin 1β

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Carprofen inhibits the release of matrix metalloproteinases 1, 3, and 13 in the secretome of an explant model of articular cartilage stimulated with interleukin 1β

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Abstract

Introduction: Arthritic diseases are characterized by the degradation of collagenous and noncollagenous extracellular matrix (ECM) components in articular cartilage. The increased expression and activity of matrix metalloproteinases (MMPs) is partly responsible for cartilage degradation. This study used proteomics to identify inflammatory proteins and catabolic enzymes released in a serum-free explant model of articular cartilage stimulated with the pro-inflammatory cytokine interleukin 1β (IL-1β). Western blotting was used to quantify the release of selected proteins in the presence or absence of the cyclooxygenase-2 specific nonsteroidal pro-inflammatory drug carprofen.

Methods: Cartilage explant cultures were established by using metacarpophalangeal joints from horses euthanized for purposes other than research. Samples were treated as follows: no treatment (control), IL-1β (10 ng/ml), carprofen (100 μg/ml), and carprofen (100 μg/ml) + IL-1β (10 ng/ml). Explants were incubated (37°C, 5% CO2) over twelve day time courses. High-throughput nano liquid chromatography/mass spectrometry/mass spectrometry uncovered candidate proteins for quantitative western blot analysis. Proteoglycan loss was assessed by using the dimethylmethylene blue (DMMB) assay, which measures the release of sulfated glycosaminoglycans (GAGs).

Results: Mass spectrometry identified MMP-1, -3, -13, and the ECM constituents thrombospondin-1 (TSP-1) and fibronectin-1 (FN1). IL-1β stimulation increased the release of all three MMPs. IL-1β also stimulated the fragmentation of FN1 and increased chondrocyte cell death (as assessed by β-actin release). Addition of carprofen significantly decreased MMP release and the appearance of a 60 kDa fragment of FN1 without causing any detectable cytotoxicity to chondrocytes. DMMB assays suggested that carprofen initially inhibited IL-1β-induced GAG release, but this effect was transient. Overall, during the two time courses, GAG release was 58.67% ± 10.91% (SD) for IL-1β versus 52.91% ± 9.35% (SD) with carprofen + IL-1β.

Conclusions: Carprofen exhibits beneficial anti-inflammatory and anti-catabolic effects in vitro without causing any detectable cytotoxicity. Combining proteomics with this explant model provides a sensitive screening system for anti-inflammatory compounds.
**Introduction**

Articular cartilage is a highly specialized load-bearing tissue that covers the ends of long bones in synovial joints and provides a strong and resilient surface for smooth and frictionless articulation as well as cushioning of the underlying bone [1,2]. The major biologic constituents of the extracellular matrix (ECM) of cartilage include collagens, proteoglycans, and noncollagenous proteins [1]. The chondrocyte is the main cell type found within the ECM of skeletally mature cartilage [3]. Chondrocytes synthesize all the ECM components in cartilage [3] and maintain this macromolecular framework in response to biochemical and biomechanical stimuli [4]. The ECM contains a specific combination of structural proteins and glycoproteins that are unique to cartilage. In addition, the ECM contains a number of other smaller noncollagenous proteins, including thrombospondin 1 (TSP-1) and fibronectin 1 (FN1). Fragments of FN1 are released in osteoarthritis (OA) and are thought to promote further cartilage degradation by upregulating catabolic signaling [5,6].

In diseases such as OA and rheumatoid arthritis (RA) chondrocytes are targeted, via specific cell-surface cytokine receptors, by pro-inflammatory cytokines such as interleukin-1β (IL-1β) and tumour necrosis factor-α (TNF-α). Although other pro-inflammatory cytokines (i.e. IL-6, IL-8 and IL-17) are involved, IL-1β and TNF-α are the predominant pro-inflammatory and catabolic cytokines involved in joint disease initiation and progression [7,8]. These pro-inflammatory cytokines suppress collagen and proteoglycan synthesis and drive inflammatory signaling, and protease expression/activation [9].

Previous studies from our research group have used an explant model of articular cartilage to study the major proteins released in response to IL-1β stimulation [10]. Subsequent high-throughput MS analysis of this model identified qualitative differences in MMP-1, -3, and -13 expression between untreated and IL-1β-stimulated explants in the spent culture media [11]. In this study, we used high-throughput proteomics and quantitative western blotting to evaluate the release of these MMPs in response to IL-1β stimulation in the presence and absence of carprofen (marketed as Rimadyl) [12], a non-steroidal anti-inflammatory drug (NSAID) developed by Pfizer Animal Health. Carprofen is a selective cyclooxygenase 2 (COX-2) inhibitor capable of blocking synthesis of the key inflammatory bioactive lipids like prostaglandin E2 (PGE2). It is used clinically to provide 24-hour relief of pain and inflammation in geriatric dogs and horses with joint pain, OA, hip dysplasia, and other forms of joint disease. The principal hypothesis of this study was that by combining proteomics with western blotting, we could determine and characterize effects of anti-inflammatory compounds (by using carprofen as an anti-inflammatory agent) in an in vitro model of cartilage (Figure 1). MMP-1, -3, and -13 were studied as surrogate in vitro biomarkers of inflammation to determine whether carprofen has the capacity to reduce the release of these catabolic enzymes. The effect of carprofen on

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**Figure 1** Schematic overview of the experimental design used in this study. Explant cultures were incubated for 6 days in designated treatments before conditioned supernatant was removed and treatment replenished for cultures up to 12 days.
cytokine-stimulated GAG release was also studied in cultures up to 6 and 12 days.

Methods
Animal tissues and statement of ethical approval
The animals used in this study were sourced from two UK-based abattoirs. Animals were euthanized for non-research purposes, having been stunned before slaughter in accordance with Welfare of Animals (Slaughter or Killing) Regulations 1995 [13]. Approval for the use of abattoir-derived animal tissues was obtained from the local Ethical Review Committee (Ethics Committee of the School of Veterinary Science and Medicine) with input from members of the University of Nottingham Animal Welfare and Ethical Review Body (AWERB).

Cartilage dissection and explant culture
Macroscopically normal articular cartilage samples were obtained from weight-bearing regions of the metacarpal-phalangeal joints of horses of mixed breed, age, and sex. Articular cartilage was harvested from the metacarpal-phalangeal joints. The joints were first shaved to remove excess hair and disinfected by washing with Hibiscrub and then Trigene, before being soaked in bleach for 1 hour. Afterward, the skin and flexor tendon was carefully removed from the joint without breaking into the synovial structure. A sterile scalpel was used to open the joint, which was then washed with sterile PBS (Gibco). Articular cartilage shavings of equal thickness were taken from the end surface of the third metacarpal and placed in 50 ml of serum-free DMEM (Thermo Scientific) + 4% Pen/Strep (Sigma) (Penicillin/Streptomycin solution: (5,000 units/ml penicillin, 5 mg/ml streptomycin)) (pre-warmed to 37°C). These cartilage shavings were transferred to a Falcon tube containing washing solution (sterile PBS + 10% Pen/Strep) and were twice washed with rotation for 20 minutes.

Cartilage-explant discs were prepared by using a 3-mm biopsy punch (Kai Medical), with discs placed into wells (five discs per 1 ml DMEM + 2% Pen/Strep) before incubation for 24 hours (37°C, 5% CO₂). After this equilibration period, media was removed and replaced with treatment media required for the purposes of the experiment. Once explants had been treated and incubated for the desired 6-day time courses, the supernatants from three animals (untreated control and IL-1β stimulated) and two from a pool of supernatants from three animals (untreated control and IL-1β stimulated). To prepare samples for MS, reduction of disulfide bonds was performed by addition of DTT (Bio-Rad) to a final concentration of 10 mM, followed by vortexing and incubation at 37°C for 30 minutes. Alkylation of thiol groups performed by addition of IAA (Bio-Rad) to a 55 mM concentration, vortexed and incubated for 45 minutes at 37°C (in the dark). Ice-cold acetone (1.2 ml) (Fisher) was added and incubated at −20°C for 1 hour, and then after centrifugation at 15,000 rcf for 5 minutes, the acetone was discarded. Proteins in the pellet were digested with 20 µg/µl trypsin gold (Promega) at 37°C overnight (16 hours). Trypsin digestion was terminated by addition of formic acid at a final concentration of 0.1%. Samples were zip-tipped with C18 resin (Millipore) by using 20 µl 50% methanol and 0.1% formic acid to elute. Excess solvent was evaporated off by heating at 70°C, to leave 10 µl of samples, which was transferred to glass vials ready to be loaded onto the nanoLC column. For each sample, 5 µl was loaded and analyzed by nanoLC-MS/MS on an amaZon ETD (Bruker). A flow rate of 250 nl/min was used to separate peptides. Solution A (100% H₂O + 0.1% formic acid) and solution B (100% ACN + 0.1% formic acid) were set to create a gradient of 10% solution B to 30% solution B over the course of an hour (0.3% per minute). From each MS scan, the five most abundant peptides were selected for fragmentation.

MS data processing
Mascot Daemon was used to search the Swiss-Prot database (all mammals) by submitting the .MGF files. The search parameters were as follows: Instrument: ESI-TRAP, peptide charge, 2+ and 3+ ions; peptide tolerance, 0.5 Da; 13C = 1; max missed cleavages = 1. Fixed modifications: carbamidomethyl (C) and variable modifications: oxidation (M). Individual ions Mascot scores above 42 indicated identity or extensive homology. Only protein identifications with probability-based protein family mascot MOWSE scores above the significant threshold of >42 (P < 0.05) were accepted and included if consistently identified in the specific treatments.
SDS-PAGE with Protean 3 mini-gels
For each sample, 10 μl collected culture supernatant was added to 10 μl of Laemmli sample buffer (62.5 mM Tris-HCl, pH 6.8, 2% SDS, 25% glycerol, 0.01% wt/vol bromophenol blue) (Bio-Rad) containing dithiothreitol (DTT) (Bio-Rad) 0.15 M. Samples were heated for 5 minutes at 95°C before loading into a 15-well 10% acrylamide gel. To reference protein molecular weights Precision Plus Protein Standards (Bio-Rad) were also loaded. A voltage of 120 V was applied until the dye front reached the bottom of the gel.

Western blotting
Western blots on days 0 to 6 of incubation were completed in three animals. Each sample of 50 μl was concentrated by lyophilization (Heraeus-Christ) before resuspension in 20 μl Laemmli buffer (Bio-Rad) before performing SDS-PAGE (see previous method). Positive controls were prepared by crushing articular cartilage with a mortar and pestle under liquid nitrogen, and suspended in Laemmli buffer. After SDS-PAGE, gels were electroblotted onto low-fluorescence PVDF membrane (GE Healthcare) at 80 V for 2 hours. Membranes were blocked for 1 hour with 5% BSA (Sigma) in PBS-Tween 20 (0.1%) (Fisher). Primary antibodies were diluted as required in blocking solution before overnight incubation at 4°C. Dilutions for primary antibodies were as follows: MMP-1 (1:800 dilution, Aviva), MMP-3 (1:1,000 dilution, Aviva), MMP-13 (1:1,000 dilution, Aviva), FN1 (1:1,000 dilution, Aviva), TSP1 (1:1,000, Abcam). The antibodies from Aviva systems biology were specifically designed to cross-react with multiple different species, including the horse.

After the overnight primary incubation, membranes were incubated with secondary antibodies for 1 hour. The secondary antibodies used in this study were all HRP conjugated and diluted to appropriate working concentrations by using 3% BSA blocking solution. Secondary antibodies were either anti-rabbit (1:100,000 dilution, Bio-Rad) for MMP-1, MMP-3, MMP-3, and FN1 primary antibodies or anti-rabbit/mouse (1:40,000 dilution, Dako) for the TSP-1 primary antibody.

Ampliflu Red Western Blot Kit (Sigma) was used to stain protein bands with red fluorescence, which was next detected on an FX-Imager (Bio-Rad) by using Quantity One software (Bio-Rad). Densitometry was performed on the images obtained by using ImageJ (NIH). Statistical analysis applying an unpaired t-test was completed, and the results graphically displayed using GraphPad Prism 6.

Chondrocyte cell death in the cartilage explants was monitored indirectly by using western blotting to detect β-actin release by explant cultures as an indicator of cytotoxicity and cell lysis. We compared the release of β-actin in untreated (control) cartilage explants and those that were stimulated with IL-1β, carprofen, and carprofen + IL-1β. For this purpose, we used a primary antibody to β-actin (1:7,000 dilution, Sigma) and a secondary anti-mouse HRP conjugate (1:100,000 dilution, Cell Signaling).

DMMB assays
For evaluation of proteoglycan release, the metachromatic dye 1,9 dimethylmethene blue (DMMB) was used to quantify the amount of sulfated glycosaminoglycans (GAGs) released into the medium. DMMB, papain, and chondroitin sulfate standards (Sigma) were prepared according to the method developed by Farndale et al. [14]. Supernatant samples were diluted to appropriate concentrations, to be within the accurate range (0 to 75 μg/ml) of the standard curve. On a 96-well plate, 40 μl of blank pain solution (duplicate), each standard (duplicate) and sample dilutions (triplicate) were added. Next, 200 μl of DMMB solution was added to each well, before reading the plate at 540 nm (spectrophotometer) within 10 minutes. To calculate the percentage of total GAG release, corresponding explant discs were digested overnight by incubating with papain solution at 60°C, before their GAG content was measured, also by DMMB assay. Total GAG release levels per well were calculated, before dividing the total GAG release by measurements for individual supernatant samples to provide the percentage GAG release in response to various treatments. Measurements from three animals (three replicates per treatment) were combined to provide final values for each incubation period.

GraphPad Prism 6 software was used produce graphic images and complete statistical analysis with one-way ANOVA by using the Tukey multiple comparison test.

Results and discussion
Carprofen decreases IL-1β-stimulated MMP-1 release
Mass spectrometric analysis of explant cultures incubated for 6 days identified MMP-1 in IL-1β-stimulated samples analyzed from all the animals (Table 1). MMP-1 was not identified by MS in untreated control samples. Untreated samples and carprofen-only treatments did not produce detected levels of MMP-1 by using western blot analysis. Western blots of day-6 explant cultures treated with IL-1β and IL-1β + carprofen contained protein bands at around 53 kDa, consistent with the predicted molecular mass of MMP-1 (Figure 2A). Densitometry and statistical analysis showed a significant decrease in IL-1β-stimulated MMP-1 release in response to carprofen treatment.

Carprofen decreases IL-1β-stimulated MMP-3 release, whereas TSP-1 is unaffected
MMP-3 was identified in both untreated and IL-1β-stimulated explant media (after 6 days of incubation) by qualitative MS analysis (Table 1). The mascot scores were considerably higher in IL-1β treatments compared...
| Protein                                      | Accession number | Mascot scores |
|----------------------------------------------|------------------|---------------|
| **ECM proteins**                             |                  | Individual animal | Pooled untreated | Individual animal | Pooled IL-1β stimulated |
| Cartilage oligomeric matrix protein          | COMP_BOVIN       | 4090           | 4763             | 7555             | 10590                  |
| Fibronectin                                  | FINC_BOVIN       | 4422           | 6222             | 3786             | 3492                   |
| Chondroadherin                               | CHAD_HUMAN       | 1022           | 3069             | 1978             | 3689                   |
| Thrombospondin 1                            | TSP1_BOVIN       | 741            | 1256             | 3456             | 2782                   |
| Aggrecan core protein                        | PGCA_HUMAN       | 1815           | 3389             | 2528             | 6412                   |
| Decorin                                      | PGS2_HORSE       | 665            | 802              | 513              | 1130                   |
| Biglycan                                     | PGS1_HORSE       | 1099           | 1062             | 2047             | 1720                   |
| Fibromodulin                                 | FMOD_HUMAN       | 1307           | 3200             | 1381             | 1930                   |
| Collagen alpha-1(III) chain                  | CO2A1_BOVIN      | 137            | 59               | 133              | 82                     |
| Collagen alpha-2(VI) chain                   | CO6A2_HUMAN      | 190            | 79               | 79               | 79                     |
| Collagen alpha-1(X) chain                    | COAA1_MOUSE      | 77             | 102              | 102              | 102                    |
| Prolargin                                    | PRELP_MOUSE      | 124            | 147              | 311              | 301                    |
| Cartilage intermediate-layer protein 1 (CILP-1) | CILP1_MOUSE     | 201            | 789              | 423              | 264                    |
| Proteoglycan 4 (Lubricin)                    | PRG4_HUMAN       | 139            | 49               | 49               | 49                     |
| SPARC (secreted protein acidic, cysteine rich) | SPRC_PIG        | 108            | 96               | 96               | 96                     |
| **Non-ECM secreted proteins**                |                  |                |                  |                  |                        |
| Clusterin                                    | CLUS_HORSE       | 2767           | 4723             | 1785             | 4876                   |
| Chitinase-3-like protein 1 (YKL-40)          | CH3L1_BOVIN      | 574            | 1178             | 414              | 2018                   |
| Serum amyloid A protein                      | SAA_CANFA        | 43             |                  |                  |                         |
| Metalloproteinase inhibitor 1                | TIMP1_HORSE      | 83             | 312              | 457              | 146                    |
| Alpha-1-antiproteinase 2                     | A1AT2_HORSE      | 99             | 342              | 50               |                         |
| MMP-1                                        | MMP1_HORSE       | 83             | 312              | 457              | 146                    |
| MMP-3                                        | MMP3_HORSE       | 653            | 956              | 5441             | 4580                   |
| MMP-13                                       | MMP13_HORSE      | 1700           | 1700             | 1700             | 1700                   |
| TNF-R superfamily, member 11b (Osteoprotegerin) | TR11B_RAT   | 252            | 319              | 388              | 638                    |
| Procollagen C-endopeptidase enhancer 2       | PCOC2_HUMAN      | 163            | 171              | 171              | 171                    |
| Lysozyme C                                   | LYSM_BOVIN       | 72             | 168              | 179              | 148                    |
| Secreted frizzled-related protein 3          | SFRP3_BOVIN      | 74             | 98               | 98               |                         |
| Lactadherin                                  | MFGM_PIG         | 83             | 63               | 63               |                         |
| Extracellular superoxide dismutase [Cu-Zn]   | SODE_HUMAN       | 67             | 72               | 72               |                         |
| Macrophage migration inhibitory factor       | MIF_HUMAN        |                |                  |                  |                         |
| Ribonuclease 4                               | RNAS4_HUMAN      | 48             | 46               | 46               | 46                     |
| **Intracellular proteins**                   |                  |                |                  |                  |                        |
| Vimentin                                     | VIME_PANTR       | 84             |                  |                  |                         |
| Alpha-enolase                                | ENOA_BOVIN       |                |                  |                  |                         |
| Phosphoglycerate mutase 1                    | PGAM1_MOUSE      | 149            | 104              | 104              |                         |
| Phosphatidylethanolamine-binding protein 1    | PEBP1_HUMAN      | 423            | 148              | 148              |                         |
| Phosphoglycerate kinase 1                    | PGK1_HORSE       | 792            | 651              | 651              |                         |
with untreated samples, giving an indication that IL-1β stimulation was having an impact on MMP-3 release. Because the MS analysis was qualitative, western blotting was used to provide accurate quantification of MMP-3 release.

Western blotting was used to quantify changes in MMP-3 release from explants in response to IL-1β stimulation and/or carprofen treatment. Upregulation of MMP-3 and its increased proteolytic activity is known to target several ECM substrates [15]. Western blotting for TSP-1 was therefore carried out by using the same membranes to determine whether the release of this ECM protein is affected by IL-1β stimulation and carprofen treatment. Bands for MMP-3 were visible at different intensities at 54 kDa in all treatment groups. An additional band a few kDa below 54 kDa appeared in all samples stimulated with IL-1β. This lower band corresponds to the active form of MMP-3 from which the propeptide domain has been cleaved.

Western blots of untreated and carprofen-alone samples revealed only a single band at 54 kDa corresponding to the nonactive zymogen form of MMP-3 [16,17].

Comparisons of MMP-3 release in explant cultures 6-day incubations (n = 3) (Figure 2) provided evidence to support the assertion that carprofen significantly decreases cytokine-stimulated MMP-3 release. MMP-mediated degradation of the ECM may be inhibited by carprofen, as treatment with this NSAID reduced the IL-1β-stimulated release and activation of MMP-3.

Carprofen decreases IL-1β-stimulated MMP-13 release
Release of MMP-13 was not detected by MS analysis of untreated explant culture media. However, MMP-13 was found in the media from all explant cultures stimulated with IL-1β (Table 1). This suggests that MMP-13 release was initiated in the explant model after IL-1β stimulation, but western blotting was required for quantification.

MMP-13 release was increased by IL-1β stimulation, and corresponding bands at 54 kDa were seen only in the presence of this cytokine (that is, not in untreated or carprofen-only explant cultures) (Figure 2). For IL-1β + carprofen treatments, faint bands were present after 6 days of incubation. Significantly less MMP-13 release occurred in the presence of carprofen when the IL-1β and IL-1β + carprofen samples were compared.

Carprofen treatment decreases the IL-1β-stimulated production of the 60 kDa fragment of FN1
Western blotting of FN revealed bands at 230 kDa representing the glycosylated monomer of this proteoglycan, and a further band at 60 kDa (Figure 3). Stimulation with IL-1β increased the density of the lower band (60 kDa), suggesting that this is a degradation product induced by this pro-inflammatory cytokine. A significant reduction was noted in the intensity of this band when carprofen was present along with the IL-1β.

Carprofen does not inhibit GAG release throughout the entire time course
A significant increase in GAG release in IL-1β-stimulated cultures was found compared with untreated controls (Figure 4). Carprofen treatment alone did not have any effect on the level of GAG release, as GAG levels were similar to untreated explant cultures throughout the entire time course.
time course. During 0 to 6 days of culture, comparing IL-1β with carprofen + IL-1β shows that carprofen caused a significant decrease in GAG release. The later time point of 6 to 12 days showed that carprofen + IL-1β had only delayed the GAG release compared with IL-1β (Figure 4).

The degradation of the ECM in articular cartilage is known to be facilitated by pro-inflammatory cytokines such as IL-1β in degenerative joint diseases such as OA [7,8]. Consequently, IL-1β is frequently used in culture models of arthritis to mimic the environment present within OA joints [18,19]. The use of physiologically relevant tissue-culture models (Figure 1) can provide mechanistic insights into the inflammatory and catabolic responses of cartilage by using tissues from euthanized cadavers, reducing the need to use animal models. In this study, we used IL-1β-stimulated cartilage explants in a static culture system. We previously used a targeted high-throughput proteomic approach to identify the major proteins present in the secretome of articular cartilage exposed to IL-1β [10,11].

Here we compared the effects of stimulation with IL-1β in the presence and absence of carprofen. One of the important issues addressed was the potential cytotoxic effects of carprofen on chondrocytes within the cartilage explants, especially because the culture system used was serum free and involved long incubation periods. The cytotoxicity of carprofen at 100 μg/ml was assessed by monitoring β-actin release from the explants by using western blotting. Western blot experiments with antibodies to β-actin were used to demonstrate that chondrocytes within the explants did not undergo cell death and lysis when treated with carprofen.

The data obtained indicated that IL-1β stimulation increases β-actin release, but addition of carprofen alone does not increase cytotoxicity compared with untreated controls (see Additional file 2: Figure S2).
Qualitative MS techniques identified a number of proteins including MMP-1, MMP-3, and MMP-13 in the secretome of IL-1β-stimulated samples (Table 1). These proteins were then selected as markers for studying the effects of carprofen. Treatment with carprofen resulted in a qualitative reduction in the levels of these proteins, even in the continued presence of IL-1β.

This qualitative approach also confirmed that MMP-1 and MMP-13 are found only above the identification threshold in IL-1β-stimulated samples, compared with untreated samples. Quantitative western blotting was used to demonstrate that the released levels of all three MMPs are significantly elevated after IL-1β stimulation. IL-1β signaling initiates the active form of NF-κB, causing transcription of pro-inflammatory and pro-apoptotic genes, initiating the release of various catalytic enzymes, including aggrecanases, cathepsins, ADAMTS, and MMPs (-1, -3, and -13) [20-22].

MMP-13 is a collagenase and has a major role in type II collagen degradation [23,24]. It also has the ability to degrade other collagens present in cartilage (types VI and IX), aggrecan, and various other ECM proteoglycans and constituents [25]. It has been identified in investigations into OA cartilage as a protein that is differentially upregulated in OA [26,27]. The progression of OA has been shown to be slowed by deletion of the MMP-13 gene in a MLI-induced OA mouse model [28]. MMP-1 is another member of the collagenase family that is effective at cleaving collagen I, II, and III [29]. The enzyme is able to unwind the triple helix of α chains before it cleaves collagen [29]. A quantitative analysis of chondrocytes has shown that IL-1β stimulation caused a significant increase in MMP-1 secretion [30]. Expression of the MMP-1 gene in equine articular cartilage has also been shown to be upregulated by IL-1β and LPS intra-articular injections [31]. MMP-3 (stromelysin-1) is known to activate both MMP-1 and MMP-13 by cleavage of their propeptide domains [32], which will lead to increased collagen degradation. In this study, we found that MMP-3 was detectable in the untreated secretome, but levels were significantly higher after IL-1β stimulation. OA cartilage secretome has been show to contain increasingly high levels of MMP-3 [33,34] and is shown to be induced in cartilage and chondrocytes by proinflammatory cytokines [30,35,36].

Measurements of MMP-1, MMP-3, and MMP-13 within in vitro models of cartilage inflammation can be used as screening systems for drugs and anti-inflammatory compounds. Interestingly, the reduced levels of these enzymes in the presence of carprofen suggests that this NSAID has the capacity to inhibit MMP release and activation and can be used as a reference drug in studies investigating the effects of compounds with potential anti-inflammatory properties. No previous proteomic studies report the use of carprofen as an inhibitor of MMP release and activity in explant cultures of cartilage stimulated with IL-1β. Therefore, this study expands on previous work in this area by testing the effects of carprofen in a novel context [10]. Carprofen is commonly used as a 24-hour treatment for arthritic dogs and is also available for use in large animals, including horses. The COX-2 enzyme is responsible for inducing metabolism of arachidonic acid, leading to the production of various prostaglandins, including PGE₂, which contributes to inflammatory signaling in synovial joint arthritis. It
would therefore be expected that inhibition of COX-2 by carprofen in this explant model would attenuate the inflammatory effects stimulated by IL-1β stimulation. Significant decreases in the IL-1β-stimulated release of active MMP-1, MMP-3, and MMP-13 were observed in the presence of carprofen.

These findings suggest that as well as alleviating pain, the use of this drug may help protect cartilage against the catabolic effects of proteases and therefore afford a certain level of protection against MMP activation and ECM loss.

The involvement of MMP action in the progression of OA has led to studies on the therapeutic effects of MMP inhibitors. Joint damage, osteochondral angiogenesis, and perceived pain were reduced by treatment with MMP inhibitors in a rat meniscal transection model of OA [37]. Whereas MMPs can contribute to collagen degradation, ADAMTS is believed to be the main enzyme responsible for aggrecan, GAG, and proteoglycan loss [38]. A DMBB assay measuring the GAG release in the cartilage explant culture time course indicated that, although inflammatory MMPs are significantly reduced by carprofen, equivalent GAG release occurs when comparing IL-1β and carprofen + IL-1β-treated explants cultured for up to 12 days.

**Conclusions**

In summary, the cartilage explant model used here has allowed us to use MMP-1, -3, and -13 as biomarkers to study the anti-inflammatory and anti-catabolic effects of carprofen. This may represent a useful approach for screening putative anti-inflammatory drugs with added anti-MMP activity in vitro and may be used for testing the effects of naturally occurring compounds on MMP expression in cartilage. This model also incorporates important elements of the replacement, refinement, and reduction of research by using animals (3Rs), thus providing an alternative to in vivo models of arthritis.

**Additional files**

**Additional file 1: Figure S1.** TSP-1 release stimulated by IL-1β is not significantly decreased after carprofen + IL-1β treatment. Western blots for TSP-1 produced a band at about 125 kDa that was detected only in the presence of IL-1β. Cartilage explants were cultured for 6 days with three treatment replicates from the same individual animal (n = 3). Explant treatments: untreated control, IL-1β (10 ng/ml), carprofen (100 μg/ml), or carprofen (100 μg/ml) + IL-1β (10 ng/ml).

**Additional file 2: Figure S2.** Cartilage explant cultures treated with IL-1β or carprofen + IL-1β did not show significant differences in β-actin release. (A) Representative western blot confirming the absence of β-actin in explant cultures that were untreated or treated with carprofen alone. IL-1β and carprofen + IL-1β treatments induced β-actin release, as evidenced by detection of an approximate 42 kDa band. (B) Graphical representation of the densitometric analysis of β-actin bands. An unpaired t-test was applied to assess statistical significance. ns, no significant difference; +, positive control for β-actin (equine chondrocyte lysate). Cartilage explants were cultured for 6 days with three treatment replicates from two animals (n = 6). Explant treatments: untreated control, IL-1β (10 ng/ml), carprofen (100 μg/ml), or carprofen (100 μg/ml) + IL-1β (10 ng/ml).
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MS; mass spectrometry; nanoLC-MS/MS; nano liquid chromatography–
mass spectrometry/mass spectrometry; NF-kB: nuclear factor kappa B;
NSAID: nonsteroidal anti-inflammatory drug; OA: osteoarthritis;
PBS: phosphate-buffered saline; PGE2: prostaglandin E2;
PvDF: polyvinylidene difluoride; RA: rheumatoid arthritis; SDS: sodium
dodecysulfate; SDS-PAGE: sodium dodecyl sulfate polyacrylamide gel
electrophoresis; TIMPs: tissue inhibitors of MMPs; TNF-α: tumor necrosis
factor alpha; TSP-1: thrombospondin 1.

Competing interests
This article was written by the authors within the scope of their academic
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Authors’ contributions
All authors have made substantial intellectual contributions to the
conception and design of the study, data acquisition, analysis, and
interpretation. AW carried out the experimental work and contributed to
data collection, interpretation, and analysis. JRS contributed to the qualitative
MS data. AM conceived the study design and supervised AW with the
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