INTRODUCTION

Soil organic matter (SOM) from the forest soils is found at various stages of decomposition as well as processing and remains the main source of organic carbon fractions and acts as an important store and source of plant nutrients [Gałka and Łabaz 2014]. Humus is the stable fraction of SOM and the methods for the separation and analysis of these fractions are therefore required [Skjemstad et al. 2006]. The forest soils usually have higher SOM than the arable soils. The accumulation of SOM in low-productivity soils depends on the land use intensity and agrotechnical operations, fertilisation as well as the removal of biomass [Kazlauskaitė-Jadzevice et al. 2019]. Even limited agrotechnical treatments affect the soil properties. Soil tillage is one of the main agrotechnical operations because of its influence on the physical, chemical and biological properties of soil [Halpern et al. 2010, van Eerd et al. 2014, Stanek-Tarkowska et al. 2018]. Murty et al. [2002] pointed out that the conversion of forest to uncultivated grazing land did not, on average, lead to a decrease in the content of SOM, although individual areas may lose or gain soil organic carbon (SOC), depending on the fertilization applied, water retention or plant residues. The evaluation of those processes should be based not only on the current content of SOC but also on the forecast SOM transformations, especially in terms of the rate of its mineralization. An ability to detect the SOC change as a result of land-use or management change is important to allow making decisions to mitigate the fertility decline. Among the methods which give possibility for
assessment the susceptibility to chemical oxidation of SOC, the method with a solution of potassium permanganate VII was use [Loginow et al. 1987]. The modification and standardization of this method [Blair et al. 1995, Lefroy 1993] allows determining the content of labile carbon (C_L) and non-labile carbon (C_NL) of SOM [Skjemstad et al. 2006]. The content of C_L can provide a considerable indicator of SOC transformations in soil [Conteh et al. 1999]. Considering the interactions between C_organ C_L and C_NL, one can determine the following: Lability Index (LI), Carbon Pool Size Index (CPI) and Carbon Management Index (CMI) in soil. On the basis of those indices it is possible to evaluate the SOC management as a measure of relative permanence of various methods of soil use. In the studies on SOC in cultivated soils, the obtained results are referred to the reference objects. These are usually non-cultivated soils, forest soils, pastures [Blair et al. 1995, Blair et al. 1997, Szombathova 1999], as well as urban soils [Vaseneva et al. 2013]. Szombathova [1999] compared the forest soil with the soils under organic and integrated farming using CMI, CPI, LI. She also demonstrated that the total carbon content in the forest soil was almost threefold higher than in the arable soils and the values of LI in the soil under organic farming were higher than in the forest soil. Strzączyńska et al. [2009] showed that in the ectohumus under the plantings of black locust, the values of CPI and CMI are more favourable than under the trees of Scots pine. The soil studies of the static fertilisation experiment exposed to long-term agrotechnical treatments showed the applicability of these indices to the evaluation of the effect of the type of fertilisation on the organic carbon management in soil [Cieścińska 2007a,b].

The aim of the present research was to compare the content of SOC and its fraction susceptible to oxidation in the arable horizon of the cultivated (hunting) plots and the humus horizon of the soils under forest.

MATERIALS AND METHODS

The research material was made up of the samples of forest soils taken in the area of the Szubin Forest Division, in the Kujawsko-pomorskie Province, in Poland. The forests stand composition is dominated by pine which found perfect development conditions in those habitats. Other species include: spruce, beech, oak, ash tree, birch, alder, larch, aspen, and hornbeam tree. The humus horizon was only a few centimetres thick (from 7.0 to 10.0 cm). The cultivation of soil in the hunting plots located inside the forests aims at supplementing and enhancing the feeding conditions of forest animals. The selection of the plant species grown can considerably enrich the feeding base for the forest animals. Due to the dominant effect of woody plants on a relatively small area of the hunting plots, these soils can be also considered as forest soils. The cultivated plots area varied from 0.9 to 2.4 ha. The plots were exposed to basic agrotechnical treatments, including mineral fertilisation. The soil samples were taken from the arable horizon (Ap) of eight cultivated hunting plots and from the humus horizon (A) under four forests (references samples F1–4). The arable soil sampling sites were selected randomly in the middle part of the cultivated plots. The averaged samples of the soils of the plots and under forests were made from three individual samples. The collected soil samples were dried and sieved through a 2-mm sieve. The following parameters were determined in the soil samples:

a) grain size composition – using the areometric and sieve method,
b) pH w 1 M KCl and pH in H_2O dest. – potentiometric method,
c) hydrolytic acidity – according to the Kappen method (the air-dried soil samples were treated in the solution of sodium acetate),
d) exchangeable cations according to the method with 0.1 M BaCl_2 solution (the content of cations was measured by AAS apparatus PHILIPS PU 9100X),
e) C_organ and total nitrogen (N_T) Vario Max CN-Elementar Analysensystem GmbH analyzer,
f) content of the available forms of phosphorus (P_2O_5) and potassium (K_2O) with the Egner-Riehm method (pH 3.6, 0.02 mol/L hydrochloric acid and 0.04 mol/L calcium lactate, soil:solution ratio 1:50), as well as the content of Mg available to plants following the Schachtschabel method (0.0125 mol/L CaCl_2 with soil:solution ratio 1:10. Phosphorus was determined with the Genesis 6 spectrophotometer (Madison, USA), potassium and magnesium – using atomic absorption spectrometry (AAS, Philips 9100, Cambridge, UK).
The oxidation of organic carbon with potassium permanganate VII (333 mmol/dm$^3$) in neutral environment was prepared according to a method devised by Łoginow et al. [1987]. The calculations considered the following:

- L (lability of carbon) = having determined the Corg, the soils were exposed to chemical oxidation. In the soil samples of the hunting plots and the reference soil samples (from forests) the contents of labile fraction (CL) and non-labile fraction (CNL) of carbon were assayed. The fractions determined that way were used to determine Carbon Management Index (CMI), Carbon Pool Size Index (CPI) and Lability Index (LI) according to Blair et al. [1997]. Corg oxidised by KMnO$_4$/Corg remaining unoxidised by KMnO$_4$;
- L = CL/CNL;
- LI (Corg lability idicator) = L in cultivated soil/L in reference soil×100%;
- CPI = Corg in cultivated soil (g/kg)/Corg in reference soil (g/kg);
- CMI = CPI×LI.

Compliant with the concept by Blair et al. [1995], the reference soil should be the control in which the dynamics of changes in the content of carbon is conditioned by the natural processes or similar to the natural ones. The reference soils in this experiment was made up of the averaged sample from the forests (F1–4) surrounding the plots investigated.

The soil properties were treated with standard statistics and statistical tests (ANOVA). The statistical analyses were made using Statistica 10.0 (StatSoft Inc, Tulsa, USA). The significance of the differences between means was evaluated drawing on the Tukey test for uneven number. The results of the analyses were also verified statistically applying according to the WARD’s method.

RESULTS AND DISCUSSION

The soils investigated in the surface horizon showed the grain size composition of fine loamy sand, except for the samples E, S2 and W2 – the grain size composition of fine sand (Table 1). Bruonic Arenosols showed a strong acid reaction. The lowest values of exchangeable acidity were noted for the soils under forests (Table 2). The acidic pH of forest soils must have been due to the decay of pine needles which are the main component of the plant litter. Strong acidic pH of the needles is characteristic for Pinus species, which can affect the soil pH under the tree stand [Parzych et al. 2017]. In the arable horizon of cultivated plots, higher values of pH were found, which could have been an effect of the agricultural use of those soils. The exchangeable acidity ranged from pH 3.67 to 4.45. Significantly higher values of hydrolytic acidity (P=0.018) and insignificantly lower values of base saturation were noted in the A horizon of soils under forests in comparison with the Ap horizon (Table 3). In the arable soils, most probably more microbiologically active, the nutrient forms were more easily available to the plants and sorption and the buffer soil properties counteracted its excessive acidification [Sapek 2009]. A slight variation in the physical and physicochemical properties of cultivated soils (Fig. 1) was confirmed by the results of cluster analysis. The agrotechnical treatments also increased the content of the plant-available forms of P, K and

| Sample          | Depth (cm) | Grain size composition [%] | Textural group |
|-----------------|------------|-----------------------------|----------------|
|                 | L (lability of carbon) | Sand 2–0.05 | Silt 0.05–0.002 | Clay <0.002 |
| Forests F1–4   | 0–19       | 86.8                      | 7.3            | 5.9          | LFS |
| Field D        | 0–28       | 88.2                      | 5.5            | 6.3          | LFS |
| Field E        | 0–28       | 91.5                      | 5.4            | 3.1          | FS  |
| Field N        | 0–28       | 87.0                      | 6.1            | 6.9          | LFS |
| Field S1       | 0–30       | 87.8                      | 6.9            | 5.3          | LFS |
| Field S2       | 0–29       | 91.4                      | 5.9            | 2.7          | FS  |
| Field S3       | 0–27       | 87.6                      | 5.9            | 6.5          | LFS |
| Field W1       | 0–28       | 83.1                      | 9.5            | 7.4          | LFS |
| Field W2       | 0–27       | 94.6                      | 3.1            | 2.3          | FS  |

FS – fine sand; LFS – fine loamy sand
Mg in the soils of most cultivated plots (Table 4). Sienkiewicz et al. [2011] found that keeping soil as a bare field as well as leaving natural plants for several years resulted in the depletion of macronutrients in soil. During the decomposition of needle litter in forest soils, the chemical traits might change due to the release of chemical elements from or their immobilization. The amounts of phosphorus may increase during the initial stages of decomposition but K and Mg were released at the rates most similar to the organic matter weight loss [Staaf and Berg 1982]. Potassium and magnesium are essential nutrients for plants and are easily removed from decomposing litter in the forest ecosystems. The concentrations of P significantly increased during the decomposition of needle litter, while the concentrations of K decreased [Kainulainen and Holopainen 2002].

The decay rate of SOM in a natural forest is lower than in an agricultural field [Krishna and Mohan 2017]. Kramer et al. [2006] found that SOM increases the availability of nutrients and improves the fertilization efficiency due to its high cation exchange capacity which prevents the nutrient losses. Higher mean content of Corg (Table 5) was not ed in the humus horizon of the soil under forests (31.6 g/kg) compared with the arable horizon of hunting plots. Under the specific conditions found in the ectohumus of forest soils, an accumulation of organic remains occurred, which transformed into humus in the humification process [Krishna and Mohan 2017]. In general, the forest soils have most of SOM in upper soil horizons. After tillage organic matter is mixed in the topsoil. Different conditions occur in the arable horizon of cultivated soils in which mineral fertilisation or inadequate crop rotation can accelerate rate of the SOM mineralization [Murty et al. 2002]. In such case, the advantage of that process over humification regularly decreases the content of humus in soil [Haynes 2005]. The results confirm the earlier studies that soil cultivation on hunting plots reduced the content of Corg and Norg as well as the values of the C:N ratio. It also confirms that in the soil of arable plots, the mineralization process was more intense [Kondratowicz-Maciejewska and Kobierski 2012].

It was demonstrated that only in two hunting plots (S2 and W1) the CMI accounted for more than 50% of the value of the reference soil (Table 5) and in the others – the value was lower than 30%. The deteriorated carbon management in the soil of the Ap horizon of the cultivated plots is a consequence of the decrease in the total organic carbon content, as compared with its content in the A horizon of forest soils.

Table 2. Selected properties of soils

| Sample       | pH 1 M KCl | Hh [mmol/kg] | TEB [mmol/kg] | CEC [mmol/kg] | (V) [%] |
|--------------|-----------|-------------|---------------|---------------|--------|
| Forests F1–4 | 3.48 (±0.10) | 80.2 (±1.04) | 41.4 (±0.43) | 121.6 | 34.0 |
| Field D      | 3.98 (±0.03) | 45.7 (±0.55) | 50.6 (±0.51) | 96.3 | 52.5 |
| Field E      | 4.51 (±0.04) | 25.7 (±0.47) | 60.1 (±1.02) | 85.8 | 70.0 |
| Field N      | 4.11 (±0.08) | 40.5 (±0.47) | 58.4 (±1.01) | 98.9 | 59.0 |
| Field S1     | 4.22 (±0.08) | 46.5 (±0.55) | 49.0 (±0.43) | 95.5 | 51.3 |
| Field S2     | 4.28 (±0.07) | 38.7 (±0.38) | 54.4 (±0.66) | 93.1 | 63.0 |
| Field S3     | 4.27 (±0.14) | 34.5 (±0.78) | 50.0 (±0.81) | 89.5 | 58.4 |
| Field W1     | 4.32 (±0.06) | 47.5 (±0.50) | 49.1 (±0.58) | 96.6 | 50.8 |
| Field W2     | 4.37 (±0.03) | 34.5 (±0.77) | 37.0 (±0.61) | 71.5 | 51.7 |

(Hh) – hydrolytic acidity; (TEB) – total exchangeable bases; (CEC) – cation exchange capacity CEC= TEB+Hh; (V) – bases saturation V=(TEB/CEC)·100%

*Significant differences
The degradation process of the pool of total organic carbon in the soil of the plots is reflected by low CPI values (0.23–0.71) (Table 5). A decrease in the total content of organic carbon in the soils of cultivated plots even by 76.7% (W2) was observed, as compared with the forest soil. The lowest decrease of C_{org} was found for the plots S2 and W1, reaching 29.1% and 33.5%, respectively. The results were confirmed by Sapek [2009] who claims that the forest soils, due to the specificity of use, are richer in organic carbon and introducing agrotechnical treatments results in a decrease in the humus content. The losses, however, can be limited and one can even increase the content of C_{org} and N_{T} in soils by applying the adequate crop rotation and fertilisation [Cieścińska 2007 a, b]. The results were confirmed by Sapek [2009] who claims that the forest soils, due to the specificity of use, are richer in organic carbon and introducing agrotechnical treatments results in a decrease in the humus content. The losses, however, can be limited and one can even increase the content of C_{org} and N_{T} in soils by applying the adequate crop rotation and fertilisation [Cieścińska 2007 a, b]. The mean contents of C_{org} and N_{T} as well as the value of the C:N ratio (Table 5) were higher for the soil samples under forest than on the arable plots. One can thus claim that in the soils of cultivated plots, the process of mineralization was most intensive. On the surface of hunting plots, on the other hand, the uncollected plant residue provides fresh organic material which, when introduced to soil, intensifies the mineralization of SOM. The LI values in the soils of cultivated plots were relatively high. As for three plots (E, S2 and W1) the rate of the processes of mineralization of organic carbon (LI over 80) was similar to the soil material under forests (Table 5). Unlike the plots S2 and W1, where CMI assumed the highest value, plot E, despite a high LI value (84.2) showed a low CMI value (only 26.9). Such an unfavourable CMI value of the soil of that plot E was affected by a relatively low value of CPI (0.32). That soil must probably be characterised by favourable conditions for the mineralisation of organic carbon and reduction of the total organic carbon pool, which, as a result, definitely leads to the soil humus degradation. Besides, the soil samples from the Ap horizon of the hunting plots S2 and W1 showed similar properties of the content of C_{org}, N_{T} and C_{L} as compared with the averaged samples of A horizon under forests (Fig. 2).

**CONCLUSIONS**

Long-term tillage of soil in the hunting plots caused a decrease in the content of C_{org} as well as its labile and non-labile fractions. The values of Carbon Management Index indicate that
cultivation of plants, which enrich the feeding base of forest animals can impact on the dynamics of transformation and mineralisation rate of \( C_{\text{org}} \) in arable soils compared with forest soils. As a result low values of the CPI index indicate that the stock of organic carbon in hunting plots has decreased. Favourable conditions of the SOM decomposition in the arable Brunic Arenosols were observed from a definitely lower values of the C:N ratio and a higher values of base saturation than in the reference forest soils. Tillage, placement, and incorporation of residue and nutrients contributed to the degradation of the pool of organic carbon in the sandy soils.

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### Table 5. Parameters of organic matter and Carbon Management Index CMI of cultivated soils and uncultivated forest soils

| Sample       | \( C_{\text{org}} \) [g/kg] | \( C_{\text{L}} \) [g/kg] | \( C_{\text{NL}} \) [g/kg] | CPI | L | LI | CMI | \( N_T \) [g/kg] | C:N |
|--------------|-----------------------------|---------------------------|---------------------------|-----|---|----|-----|----------------|-----|
| Forests F1–4| 31.6 (±0.25)                | 7.48 (±0.05)              | 24.1                      | -   | 0.310 | - | - | 1.94 (±0.05) | 16.3 |
| Field D      | 12.2 (±0.52)                | 1.83 (±0.05)              | 10.4                      | 0.39 | 0.176 | 60.0 | 21.6 | 1.08 (±0.03) | 11.3 |
| Field E      | 10.3 (±0.25)                | 2.14 (±0.06)              | 8.2                       | 0.32 | 0.261 | 84.2 | 26.9 | 0.96 (±0.02) | 10.7 |
| Field N      | 11.4 (±0.21)                | 1.79 (±0.10)              | 9.6                       | 0.36 | 0.186 | 60.0 | 21.6 | 1.17 (±0.01) | 9.7  |
| Field S1     | 12.6 (±0.21)                | 2.23 (±0.03)              | 10.4                      | 0.40 | 0.214 | 69.0 | 27.6 | 1.18 (±0.04) | 10.7 |
| Field S2     | 22.4 (±0.42)                | 4.67 (±0.06)              | 17.7                      | 0.71 | 0.264 | 85.2 | 60.5 | 1.86 (±0.04) | 12.0 |
| Field S3     | 7.49 (±0.04)                | 1.38 (±0.04)              | 6.1                       | 0.24 | 0.226 | 72.9 | 17.5 | 0.84 (±0.06) | 8.9  |
| Field W1     | 21.0 (±0.40)                | 4.30 (±0.07)              | 16.7                      | 0.66 | 0.257 | 82.9 | 54.7 | 1.94 (±0.04) | 10.8 |
| Field W2     | 7.36 (±0.07)                | 1.26 (±0.02)              | 6.1                       | 0.23 | 0.207 | 66.8 | 15.4 | 0.54 (±0.03) | 13.7 |

\( C_{\text{org}} \) – total organic carbon; \( N_T \) – total nitrogen; \( C_{\text{L}} \) – labile fraction of carbon; \( C_{\text{NL}} \) – non-labile carbon; CPI – Carbon Pool Size Index; L – lability of carbon; LI – Lability Index

\[
C_{\text{NL}} = C_{\text{org}} - C_{\text{L}}; \quad \text{CMI} = \text{CPI} \times \text{LI}; \quad \text{CPI} = \frac{C_{\text{org}} \text{sample}}{C_{\text{org}} \text{reference}}; \quad \text{LI} = \left( \frac{L \text{sample}}{L \text{reference}} \right) \times 100; \quad L = \frac{C_{\text{L}}}{C_{\text{NL}}}
\]

**Fig. 2.** The cluster analysis based on: \( C_{\text{org}} \), \( N_T \), \( C_{\text{L}} \).
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