Transcriptome profiling data reveals ubiquitin-specific peptidase 9 knockdown effects

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ABSTRACT

Ubiquitin specific peptidase 9 (USP9) is a deubiquitinase encoded by a sex-linked gene with a Y-chromosomal form (USP9Y) and an X-chromosomal form (USP9X) that escapes X-inactivation. Since USP9 is a key regulatory gene with sex-linked expression in the human brain, the gene may be of interest for researchers studying molecular gender differences and ubiquitin signaling in the brain. To assess the downstream effects of knocking down USP9X and USP9Y on a transcriptome-wide scale, we have conducted microarray profiling experiments using the human DU145 prostate cancer cell culture model, after confirming the robust expression of both USP9X and USP9Y in this model. By designing shRNA constructs for the specific knockdown of USP9X and the joint knockdown of USP9X and USP9Y, we have compared gene expression changes in both knockdowns to control conditions to infer potential shared and X- or Y-form specific alterations. Here, we provide details of the corresponding microarray profiling data, which has been deposited in the Gene Expression Omnibus database (GEO series accession number GSE79376). A biological interpretation of the data in the context of a potential involvement of USP9 in Alzheimer’s disease has previously been presented in Köglsberger et al. (2016). To facilitate the re-use and re-analysis of the data for other applications, e.g. the study of ubiquitin signaling and protein turnover control, and the regulation of molecular gender differences in the human brain and brain-related disorders,

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we provide a more in-depth discussion of the data properties, specifications and possible use cases.
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1. Data

Transcriptome-wide gene expression profiles of human DU145 prostate cancer cells were obtained for two different shRNA-knockdowns, one specific for the X-form of the gene USP9 (USP9X) and one combined knockdown of both the X- and Y-form of the gene (USP9X/Y, used to infer potential USP9Y-specific gene expression changes indirectly by comparing both knockdowns). The data additionally covers transcriptome measurements for three control conditions to assess the significance and specificity of the knockdown effects: A scrambled RNA control, a transfection reagent control, and an unperturbed control. For each of these target and control conditions, three replicates are included to account for biological and technical variation in subsequent statistical analyses. Apart from the raw microarray measurements in Affymetrix CEL-file format, we also provide the pre-processed data obtained from applying the GC-RMA approach for background
correction, normalization and probe replicate summarization [6]. To provide an overview of the significance and effect sizes of differentially expressed genes between USP9X and USP9Y combined knockdowns and control samples in this dataset, a volcano plot was generated (Fig. 1) and the top 50 differentially expressed unique genes with known gene symbols were visualized in a heat map (Fig. 2). Finally, to compare the expression changes in the two knockdowns against each other, a correlation plot of transcript log fold changes in the USP9X knockdown samples compared to controls vs. the transcript log fold changes in the USP9X/Y knockdown samples was created (Fig. 3).

**Fig. 1.** Volcano plot to visualize differential gene expression between USP9X and USP9Y combined knockdowns and control samples. For each transcript, the negative logarithm to base 10 for the adjusted p-value is plotted against the log fold change. To highlight transcripts with high effect size and significance, data points are colored orange if the absolute value of the log fold change is above 1, and green if additionally, the adjusted p-value is below 0.05 (all other data points are shown in black). For the data points that pass both these effect size and significance filters, the HGNC gene symbols and corresponding genetic probe set IDs (in brackets) are also displayed (multiple genetic probes map to the gene USP9X).
Fig. 2. Heat map for the top 50 shared differentially expressed unique genes with known gene symbols in the USP9XY and USP9X knockdowns. On the right, the standard HGNC symbols for these genes are provided, as well as the IDs of the corresponding microarray genetic probe sets (shown in brackets). The heat map color codes represent row-wise Z-scores computed for the gene expression levels on log-scale (see the color key at the top left).
2. Experimental design, materials and methods

2.1. Cell culture knockdown experiments

DU145 cells (ATCC #HTB-81) were cultured in Dulbecco's modified eagle medium (Gibco #41966–029), supplemented with 10% v/v heat-inactivated fetal bovine serum (Gibco #10500–064). Knock-down plasmids were designed to either jointly target USP9X and USP9Y or only target USP9X. Since USP9X and USP9Y have a high sequence homology, no potentially unspecific USP9Y shRNA plasmids were generated. Hairpin sequences and U6 promoters were subcloned into FastBac plasmids [7] (see [4] for details on the primers tested and the agarose gel electrophoresis results). Only one shRNA construct for each target was used, and since off-target effects can occur and the gene silencing...
efficiency may vary for different constructs, limitations may arise when interpreting data derived from a single construct. However, since the expression analysis only showed a highly significant inhibition of the target genes and no matched or similar effects for any other gene, we can exclude strong off-target changes and that larger amounts of target transcripts escape the inhibition to recover normal function. Cells were transfected via lipofectamine 2000 (ThermoFisher/Invitrogen #11668019) and incubated for 48 h.

2.2. Cell sorting, RNA isolation and microarray experiments

Before extracting RNA, the perturbed cells were enriched via fluorescence-activated cell sorting. The Qiagen RNeasy Mini Kit (Qiagen #74106) and DNasel (Qiagen #79254) were used to extract and treat the RNA, respectively. Reverse transcription was done as described previously [8]. RNA extracts were prepared for the microarray experiments using the GeneChip WT PLUS Reagent Kit (Affymetrix, Manual P/N 703174 Rev. 2 and User Manual GeneChip Expression Wash, Stain and Scan for Cartridge Arrays P/N 702731 Rev. 4). A NanoDrop ND-100 (Thermo Scientific) and the 2100 Agilent Bioanalyzer were used to check RNA quality and integrity, respectively. The purified, sense-strand cDNA was fragmented by uracil-DNA glycosylase (UDG) and apurinic/apyrimidinic endonuclease 1 (APE 1) at the unnatural dUTP residues. The fragmented cDNA was labelled by terminal deoxynucleotidyl transferase (TdT) using the Affymetrix proprietary DNA Labelling Reagent covalently linked to biotin. 5.5 μg of single-stranded cDNA was used for fragmentation and labelling, and the GeneChip Hybridization, Wash and Stain Kit was used to hybridize and wash the cartridges. Control Oligonucleotide B2 and 20X Eukaryotic Hybridization Controls were added to the hybridization cocktail containing the labelled sample and injected into the cartridge. Incubation lasted 16 hrs. at 45 °C with rotation at 60 rpm. Next, the Fluidics Station 450/250 was used to wash and stain the Affymetrix GeneChip Human Gene 2.0 ST probe arrays. Using the Affymetrix GeneChip Scanner 3000, the probe arrays were then scanned after completion of the wash protocols.

2.3. Microarray data processing and visualization

The raw microarray dataset was pre-processed and normalized using the GC-RMA approach [6], and the raw and processed data have both been made available in the Gene Expression Omnibus (GEO) database (series accession number GSE79376). Differentially expressed genes between different knockdown conditions (USP9X only and USP9X/Y combined knockdown) and control conditions were assessed using the empirical Bayes moderated t-statistic [9]. To adjust the resulting p-values for multiple hypothesis testing the Benjamini–Hochberg procedure was applied [10]. Gene expression patterns were visualized using a volcano plot to highlight altered patterns of gene expression between USP9X and USP9Y combined knockdowns and control samples (Fig. 1). The volcano plot compares the negative logarithm to base 10 of the adjusted p-values against the log fold change, revealing that numerous genes display alterations with high effect size and high significance. As a complementary visualization, a heat map and cluster dendrogram was generated for the top 50 shared differentially expressed unique genes with known gene symbols in the USP9XY and USP9X knockdowns compared to control samples, using hierarchical average linkage agglomerative clustering with a Pearson correlation distance metric (Fig. 2). These top 50 genes include USP9X as the gene with the most significant decrease in expression levels. To further confirm the successful knockdowns for USP9X and USP9Y/X, we determined the median log fold changes and p-values for all genetic probes mapping to these genes. For the USP9X/Y knockdown, a strong under-expression was observed for both the USP9X and the USP9Y gene (median log. fold change (logFC) USP9X: -2.8, median p-value USP9X: 1.1E-05; median logFC USP9Y: -1.4, median p-value USP9Y: 6.1E-03). For the USP9X knockdown, lower average expression levels for USP9X were also observed, but with a weaker effect (median logFC: -0.46, median p-value: 2.6E-01). To assess the similarity between the expression alterations for the two knockdowns, a correlation plot was generated, comparing the transcript log fold changes for the USP9X knockdown against those in the USP9X/Y knockdown (Fig. 3).
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Conflict of interest

None.

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