Review Article

Anisi Stellati Fructus, a Significant Traditional Chinese Medicine (TCM) Herb and Its Bioactivity against Gastric Cancer

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Anisi stellati fructus (ASF) is the fruit of Illicium verum Hook F. (Chinese star anise), which is native to many countries, and is a significant Chinese medicinal herb. Gastric cancer (GC) is one of the major fatal types of cancers with multiple stages and a poor prognosis. The present review aims to discuss the bioactive properties of ASF and its phytocompounds against GC, with a particular insight into the molecular mechanisms and signaling pathways involved in its anti-GC mechanism. Furthermore, it highlights the potential mechanism of action of major phytocompounds of ASF against GC. Clinical studies (in vitro and in vivo) regarding the action of ASF and its major bioactive compounds such as quercetin, luteolin, kaempferol, d-limonene, and honokiol against GC were reviewed. For this review, search of literature was performed in Science, PubMed, Google Scholar, Web of Science, and Scopus related to ASF and its phytocompounds, from which only relevant studies were chosen. Major bioactive compounds of ASF and their extracts have proven to be effective against GC due to the mechanistic action of these compounds involving signaling pathways that target cancer cell apoptosis, proliferation, and tumor metastasis in GC cells. Existing reports of these compounds and their combinatory effects with other modern anticancer agents have also been reviewed. From its traditional use to its role as an anticancer agent, ASF and its bioactive phytocompounds have been observed to be effective in modern research, specifically against GC. However, further studies are required for the identification of molecular targets and pharmacokinetic potential and for the formulation of anti-GC drugs.

1. Gastric Cancer (GC): Epidemiology and Etiology

Cancer is broadly defined as the uncontrolled proliferation of cells. It is classified into several stages before being termed fatal for a living system [1]. The World Health Organization (WHO) entails several factors to be involved in its progression, including lifestyle, diet, environment, gender, genetics, and overall health [2]. According to the Global Cancer Observatory (GCO), gastric cancer (GC) ranks the fifth most common cancer to be diagnosed globally and remains the third leading cause of cancer mortality after lung and colorectal cancer, respectively (5-year survival rate > 25%), with approximately 1 in 12 (< 8%) cancer-related deaths being attributable to GC [3]. The mortality risk of GC from birth till the age of 70 is more than 1% and 0.5% for males and females, respectively [4]. According to the GLOBOCON project, there were more than 100,000 new cases of GC and more than 700,000 GC-related deaths in 2018. The occurrence of GC is variable with respect to region and ethnicity and is reported to be more prevalent (more than 2 times) in males than females in developed regions. In more than five countries of the world, GC has the highest prevalence among all types of cancers in males [5]. In countries of Central, Eastern Asia, and Africa, the prevalence rate of GC is the highest, whereas the lowest rate of incidence is in Korea, with almost 4 cases per 100,000 cases for males, respectively [6].

Though GC is one of the most fatal types of cancers, it is also one of the most influential types on the basis of human behaviors and therefore is preventable [7]. Till the 1980s, it was the leading cause of cancer-related deaths until it was
overshadowed by lung cancer, as the rate of incidence of the latter was on the increase than the former, particularly in developed countries. Nevertheless, GC sustains a high mortality rate worldwide attributing to life-year-burden [8]. There are a variety of factors that affect its development and manifestation, which can either be genetic or environmental and triggered by the presence or abundance of pathogens in the surroundings [9]. However, drastic changes in lifestyle, profound awareness, and pathogenic eradication have caused the incidental rate of GC to substantially decline over the years [10–12]. Nevertheless, GC is a progressive form of cancer, which is comprised of multiple stages, commencing from chronic superficial gastritis, atrophic gastritis, intestinal metaplasia, and dysplasia and adenocarcinoma [13]. Apart from these stages, gastric precancerous lesions (GPLs), including intestinal metaplasia and dysplasia, are significant indicatives of GC occurrence [14]. Various reports demonstrate the yearly prevalence of GC-related intestinal metaplasia, mild and moderate dysplasia, and severe dysplasia to be 0.25, 0.6, and 6% after a 5-year period of GC diagnosis [15]. Hereditary diffuse gastric cancer (HDGC) is one of the most common forms of GC, which is associated with familial history, caused by mutations in the cadherin 1 (CDH1) gene [16]. Other changes leading to the development of cancer that occur due to long-term inflammation include lack of balance between epithelial cell differentiation and apoptosis, atrophy and achlorhydria, and gastric colonization by enteric microorganisms with nitrate reductase activity [17]. These EPIYA motifs are significant as their phosphorylation status, and the amount is an indicator for GC [28]. Tyrosine-phosphorylated CagA activates tyrosine phosphatase in the host cell, which in turn activates ERK1/2 and C-terminal Src kinase, while the interaction between the two results in cell elongation [30]. Even in its non-phosphorylated form, CagA has several pathogenic effects by targeting various cellular components like E-cadherin, c-Met, and Grb-2 to mediate proinflammatory responses, disrupting cell to cell apical junctions, activating β-catenin, and resulting in the loss of cell polarity [31, 32]. The highly pathogenic cytotoxin secreted by H. pylori, vacuolating cytotoxin A (VacA), is associated with various genotypes and vacuolating activities. Patients infected with H. pylori strains that express VacA with s1 or m1 genotype are reported to have an elevated risk of gastric cancer. Therefore, VacA expression has been designated as a significant biomarker for the development of gastric cancer [33]. For the evasion of the bacterium from the host’s immune system, genetic diversity is one of the major factors that H. pylori undertakes for the initiation of chronic inflammation and host colonization. Previous literature reported the involvement of CagA and VacA genes in the progression of neoplastic or nonneoplastic GC [34]. Moreover, H. pylori produces the enzyme urease, which releases ammonium and carbon dioxide for the neutralization of stomach acids, allowing the survival of the bacterium. Ammonia further causes alterations in the tissue structure, whereas carbon dioxide offers protection to H. pylori from host immune cells and induces angiogenesis, thus promoting GC [35]. Apart from GC, disturbance in the tissue microenvironment (TME) of the host gastric mucosa [36, 37] also involves H. Pylori infection. Furthermore, other bacterial species than H. pylori can also be attributable to carcinogenesis in the gastric mucosa [38]. Apart from bacterial pathogens, the occurrence of GC can be attributable to the infection caused by viruses, including Epstein-Barr virus (EBV), which has been reported to be associated with more than 9% of GC cases [39]. Its role in the manifestation and growth of GC has been regarded as a complex one, as many other factors are commonly associated with the incidence of this disease [40]. Like H. pylori, EBV’s role in GC is also linked with genetic mutation, characterized here by the posttranscriptional genetic regulation of EBV by miRNAs [41].

2. Association of Helicobacter pylori with GC

The major risk factor for GC is the bacterium Helicobacter pylori, which was discovered by Barry Marshall and Robin Warren in 1982, prior to which other factors such as lifestyle, diet, and stress were considered to be major risk factors for gastric disease [18]. H. pylori is a Gram-negative, spiral-shaped bacterium, which predominantly resides in the human stomach and is reported to colonize the human gut of immunocompromised individuals during infection [19]. It is a stomach pathogen that is responsible for stomach-related ulcers (duodenal and gastric) and cancer (gastric, mucosa-related lymphoid tissue lymphoma) in infected hosts [20]. The incidence of H. pylori has been reported to elevate the risk of stomach cancer to a fivefold ratio within a decade of infection. Moreover, more than 90% of noncardia subtypes are reported to be associated with the pathogen [21]. Polymorphisms of IL-10 and IL-17, which are associated with GC, are also involved in the interaction with H. pylori infection [22]. Histopathologically persistent gastritis, gastric decay, intestinal metaplasia, dysplasia, and malignancy are different phases which ultimately occur due to H. pylori infection, more than often leading to mortality [23]. The role of H. pylori in the initiation of GC involves direct infection and inflammation that occurs in the gastrointestinal mucosa of the host [24]. For instance, genes for the Type IV secretion system, which are reported to be essential for the transport of CagA proteins into human epithelial cells by H. pylori, were observed to be abundant in the intragastric microbiome of intestinal metaplasia patients [25]. Cag-Pathogenicity Island (cagPAI) consists of more than 30 genes that encode the Type IV secretion system and the CagA protein. Strains of H. pylori that express capPAI are associated with pathogenesis of gastric disease, peptic ulcers, and gastric cancer [26]. CagL, another protein, aids in translocating CagA and secreting IL-8 from H. pylori. In vitro studies have reported the role of CagA in inducing tumorigenesis in AGS cells [27]. Abl and Src kinases phosphorylate CagA on tyrosine residue at 4 distinctive EPIYA motifs inside the host cell, which results in several morphological cellular changes and increased cellular migration [28]. These EPIYA motifs are significant as their phosphorylation status, and the amount is an indicator for GC [29]. Tyrosine-phosphorylated CagA activates tyrosine phosphatase in the host cell, which in turn activates ERK1/2 and C-terminal Src kinase, while the interaction between the two results in cell elongation [30]. Even in its non-phosphorylated form, CagA has several pathogenic effects by targeting various cellular components like E-cadherin, c-Met, and Grb-2 to mediate proinflammatory responses, disrupting cell to cell apical junctions, activating β-catenin, and resulting in the loss of cell polarity [31, 32].
3. Treatment of GC: Conventional and Modern Methods

3.1. Conventional Methods and Chemotherapeutic Agents. Chemotherapy, radiotherapy, tumor resecting surgery, immunotherapy, and targeted therapy have all proven to be effective against GC and adenocarcinoma, which implies that a multidisciplinary approach is pertinent for the suitable selection of treatment. Chemotherapy for resectable GC is now acceptable, but classification of GC on the grounds of various molecular subtypes is significant for a more personalized, therapeutic approach. Random clinical trials prove evidence that perioperative and postoperative chemotherapy, chemoradiation, and immunotherapy are also considerable options for treatment [42]. Several cytotoxic agents are reported to be active in stages of advanced GC, including irinotecan, platinum, and taxanes. Oxaliplatin is the preferred platinum in most treatment regimens [43]. In the second-line of treatment for metastasized GC, monoclonal antibodies such as ramucirumab have proven a boost in survival of GC patients, with both its singular therapy and use in combination with paclitaxel being deemed effective. Kinase inhibitors such as lenvatinib and regorafenib have also been investigated in their use in immunotherapy in GC in East-Asian populations [44]. Nevertheless, the choice and options available for treatment are dependent on the prognosis of disease and the response of the patient to the preferred method, as these treatments come with their set of side effects.

3.2. Traditional Chinese Medicine (TCM) and GC. Over the past years, significant research in the field of cancer has resulted in scientific breakthroughs, including cancer immunotherapy, whose efficacy is dependent on the inflammation regulating the tumor microenvironment [45]. However, this approach is restricted to a slow rate of response, thus making it appropriate only for some patients [46]. This rate can be accelerated by the combination of immunotherapy with other agents, which can lead to an increase in the cure rate [47]. Traditional Chinese medicine (TCM) is a conjugative discipline, combining personalized medicine with therapeutics and cancer therapy. However, a greater part of the patients presently uses TCM as an alternate method of pain alleviation rather than the main method of treatment, despite the fact that it has been directly associated with treating major diseases such as cancer in recorded medical history [48].

Conventional treatments for diagnosed GC include tumor resection during early stages, but unfortunately, in patients with advanced stages of nonresectable tumor, the patients are advised noninvasive herbal therapy regimens meant to only alleviate their pain and increase their life expectancy [49]. Since time immemorial, plants have been used as therapeutic sources for the treatment of infections, diseases, and healthcare. Decades worth of studies have proven their efficacious potential and discovery of medicinal plant-derived drugs. The Chinese Pharmacopeia deciphers the involvement of Chinese herbs, plants, and their concoctions in addressing, managing, and curing clinical and terminal diseases. TCM includes medicinal plants and their associated constituents that can be used for therapeutic as well as theranostic purposes. They provide a wide spectrum of information about the mode of action of medicinal plants regarding managing a variety of diseases and simultaneous information about the human physiological systems [50]. Earliest reports of its use date back to 200 AD for the improvement of healthcare. Currently, traditional medicinal documentation of Ayurvedic and Chinese medicinal plants is available online in various databases. They contain comprehensive information about different plant parts, secondary metabolites, and bioactive compounds. In developing countries, people still employ traditional concoctions, which have been passed on from generations to cure diseases. This practice is now being opted by even those who reside in developed countries, where innovative research is being conducted to unlock the bioactive mechanisms effective against various diseases such as cancer [51]. Bioactive phytoconstituents that address cancer cells include alkaloids, saponins, terpenoids, tannins, and flavonoids [52]. In a similar manner, TCM has proven to be effective in the prevention of GPL progression [53].

Traditional medicine is comprised of the comprehension of beliefs, knowledge, and information used in amalgamation with disparate therapies and other practices, which gives way to the incorporation of herbal medicines prepared from animals, minerals, and/or plant and its constituents for diagnosis, treatment, and prevention of infection and disease [54]. Therefore, in context, every geographical locale inherits its own rich knowledge of traditional medicine, which may be passed onto generations and spanning decades. In developing and developed countries, this built-up indigenous knowledge lives on to serve fundamental roles in employing localized resources like plants and herbs [55]. Therefore, this review sheds light on one of these herbs, Anisi stellati fructus, for its mechanism of action and treatment of GC.

4. An Example of TCM: ASF and Its Activity against GC

Anisi stellati fructus (ASF) is the star-shaped fruit of Illicium verum Hook F. (Chinese star anise), which belongs to the Illiciaceae family [56], but the former is reported to be a member of the Schisandraceae family [57]. I. verum is a highly regarded Chinese medicinal herb which is also mentioned in the Chinese Pharmacopoeia [58]. There are various species in the genus, which demonstrate variance due to their distinctive morphology, composition, and growth habitat. I. verum is an aromatic, medium-sized tree which is grown in areas native to Jamaica, Laos, Japan, Indonesia, Philippines, and north-east and south-west Vietnam and China, respectively [56, 59], and is widely distributed in many regions of Asia and North America [60]. In various regions all over the world, it goes by different local names, including “Badiyan” (Persian), “Badiyaan” and “Badiyaan ka phool” (Urdu), “Phoolchakri” (Hindi), “Badiane” (French), and star anise (English). In TCM, it is commonly known as Ba Jiao Hui Xiang [58, 61].
4.1. ASF in Culinary Practice. ASF is characterized by 6–8 ridged follicles that are star-shaped, woody, and wrinkled in texture. It is generally regarded as safe and nontoxic for consumption [62] and has traditionally been used as a staple spice in various cuisines, including Oriental, Indian, and Pakistani cuisines. It is a major component of the widely used five-spice powder (locally known as garam masala in the Indian subcontinent) used in the preparation of stews and curries [63]. In European countries, it is used in the preparation of alcoholic drinks along with various teas, fruit jams, and condiments [64].

4.2. Medicinal Uses of ASF. The use of ASF in treating various infections and diseases is practiced in various regions, including Asia and North America [65]. It is reported to possess antimicrobial, antiviral, and antioxidant properties [66]. In Chinese, Ayurvedic, and Unani medicine, it is reported to improve digestion and alleviate symptoms of dysentery, dyspepsia, asthma, flatulence, menstruation irregularities, colic, inflammation, bronchitis, and rheumatic diseases. Using it in a concoction for herbal teas can relieve cough and flu and can reinvigorate various organs of the human body [61].

4.3. Anticancer Properties of ASF. Though the effects of ASF and its extracts and decoctions against cancer and tumor growth have not been thoroughly established, few studies have brought them to light, providing a brief yet informative insight into its anticancer mechanism. Extracts of ASF inhibited angiogenesis in Human umbilical vein endothelial cells (HUVECs) at concentrations of 10 μg/ml, suggesting its anticancer activity [67]. Another study reported that the oral administration of ASF resulted in the decrease of metastasis in lung cancer cells with little or no cytotoxic effects [68]. In chronic myeloid leukemia (CML) cells, ASF and its combinational treatment with imatinib yielded anti-angiogenic activity, indicating that ASF could be considered as a potential agent for CML therapy [69]. Therefore, these studies suggest that ASF possesses significant anticancer activity, which could be further elucidated by similar findings.

4.4. Chemical Constituents of ASF. The fruit of I. verum is reported to contain various alkaloids, essential oils, and tannins, with significant amounts of cis- and trans-anethole, limonene, safrole, α- and β-pinene, β-phellandrene, α-terpineol, and farnesol [70]. Flavonoids like quercetin and kaempferol and their glucosides, phenolic compounds like shikimic acid, and fatty acids such as linoleic, myristic, stearic, betulinic, and phenyl propionic acid are also reported to be active constituents of ASF [48, 71]. Furthermore, the essential oils of ASF are comprised of flavonoids, terpenes, sesquiterpenes, and lignans which are equally significant respective to their medicinal and therapeutic properties. Additional bioactive compounds which are found in ASF essential oils are myrcene, limone, linalool, luteolin, estragole, caryophyllene, γ-terpineol, and α-humulene [72]. Moreover, recent studies have reported the presence of other compounds such as β-sitosterol, α-phellandrene, β-myrcene, mairin, honokiol, cineol, and safrole [73–75].

5. Major Bioactive Compounds of ASF and Their anti-GC Effects

ASF and its compounds have been extensively reported for their anticancer activity [76, 77]. Kim et al. reported that the oral administration of ASF significantly decreased the metastasis in malignant cancer cells, which ultimately resulted in the reduction of MMP-9, MMP-13, MMP-14, uPA, and gelatinase activities by its treatment [68]. It also inhibited the activation and phosphorylation of NF-κB, AP-1, and p38 pathways, respectively, as well as suppressing tumor angiogenesis in cells. Another recent study examined the effect of ASF extract on CML cells, which demonstrated that the treatment induced cytotoxicity and proliferation inhibition in a dose-dependent manner. The combination of ASF extract with imatinib (IM) also leads to an apoptotic effect in the cells, which was not as pronounced as the singular treatment of IM on CML cells [69]. In a similar manner, many bioactive compounds of ASF (present in a major or minor amount) have been identified to be effective against GC, which are mentioned in detail in the next section.

5.1. Quercetin. Quercetin (3,3′,4′,5,7-pentahydroxyavone) is a flavonoid which is abundantly found in many foods and plants, with many properties attributed to the compound, including antioxidative, anti-inflammatory, antimicrobial, and anticancer activities, respectively [78, 79]. The treatment of quercetin in cancer cells has been reported to induce apoptotic, antilucre, and chemopreventive effects [80]. Furthermore, quercetin facilitates the prevention of mucosal damage in gastric ulcer formation, which is also said to be associated with its antibacterial action against H. pylori [81]. Various studies have reported the effectiveness of quercetin against cancer proliferation and angiogenesis. Its effect on gastric cancer apoptosis was revealed through Bax, BCL-2, and caspase analyses [82]. The regulation of P450 enzyme expression also resulted in the activation of procarcinogens by quercetin. Its treatment has also been affiliated with the enhancement of DNA repair and the elimination of carcinogens and actively proliferating cancer cells [83]. Quercetin also mediates apoptosis by the expression of various proteins such as mitogen-activated protein kinases (MAPK), phosphatidylinositol 3-kinases (PI3K), and protein kinase C (PKC) through regulating the expression of the BCL-2 family [84]. In two studies, it was demonstrated that quercetin and its combinative treatment with other cancer-inhibiting agents induce autophagy in GC cells via the negative regulation of the Akt-mTOR signaling pathway, which leads to the overall inhibition of cellular proliferation [85]. The administration of quercetin also leads to the reduction in the invasion and migration of GC cells through downregulating the expression of uPAR and uPA proteins, respectively [86]. In GC cell lines, the treatment of quercetin is demonstrated to reduce the progression of cellular growth in the cell cycle phases [87]. The combinative treatment of
quercetin with curcumin resulted in the decrease of Akt and ERK phosphorylation, which suggested cellular apoptosis via the mitochondrial pathway in GC cells [88]. In in vivo models, the treatment of quercetin stimulated the generation of nitric oxide synthase (nNOS), which reacted with ROS for the inhibition of cellular proliferation of cells of the gastric mucosa, which were previously treated with ethanol [89]. Other in vivo studies depicted the administration of oral quercetin to reduce COX-2, Twist1, and ITGβ6 levels [90]. Furthermore, the downregulation of angiogenesis-related factors (VEGFA and VEGFR-2) also suggested the effectiveness of quercetin against GC cells [91]. In the AGS cell line, quercetin leads to the reduction and increase in the expression of antiapoptosis (MCL-2, BCL-2, and BCL-x) and prosapoptosis (Bad, Bax, and Bid) related proteins, respectively [92]. The blocking of the phosphoinositide 3-kinase (PI3K-) Akt pathway is one of the mechanisms through which quercetin is reported to inhibit the mitochondrial pathways leading to the progression of GC, which was reported in many studies (Figure 1) [84, 93]. Recent in silico analyses have employed the network pharmacology approach for understanding the mechanism of quercetin and its involvement in molecular pathways against GC [94, 95].

5.2. Luteolin. Luteolin (3′,4′,5,7-tetrahydroxyflavone) is another well-known flavonoid that has been widely reported to be effective against the progression of several types of cancers, including GC [96]. In a study, luteolin was reported to inhibit the proliferation of GC cells through suppressing the Notch signaling pathway [97]. The administration of luteolin leads to the reduction in cell viability and induction of cell cycle arrest, as well as apoptosis in GC cell lines [98]. The treatment of GC cell lines with luteolin alone induced apoptosis, while its combinatorial treatment together with cisplatin resulted in the down and upregulation of CDC2, CDC25C, Cyclin-B1, and p21/cip1, respectively, leading to the effective inhibition of cell growth [96]. Lu et al. reported that luteolin was attributable to the downregulation of c-Met, MMP9, and Ki-67 in GC cells while promoting the induction of apoptosis via activating apoptotic proteins (CAS3 and PARP1), thus suggesting that luteolin can target Akt/ERK signaling pathway for its anti-GC effect [97].

BCL-2 is an apoptosis regulating protein that is characteristically found to be overexpressed in various cancers [98]. In a study, new findings demonstrated luteolin to downregulate the expression of the protein via the upregulation of miR-34a, thus aiding in inhibiting cellular growth in GC [99]. The suppression of phosphorylation of MAPK, AKT, and PI3K signaling pathway, as well as the induction of apoptosis in GC cells, was also observed through the regulation of CAS3, CAS9, and Bax/BCL-2 ratio by luteolin [100]. It is reported that treating GC cells with luteolin leads to the inhibition of STAT3 phosphorylation, reducing the growth of tumors in vivo [101]. Notch signaling pathway is reported to be associated with cellular angiogenesis and the regulation of AKT, MMP-9, and NF-κB signaling pathways [102], the latter of which in turn regulates VEGF expression in various human tissues [103]. In GC, luteolin impedes the expression of VEGF via the downregulation of the Notch1 pathway, the study findings of which have been proven previously by two studies [97, 104]. Additionally, treating H. pylori-infected GC cells with luteolin resulted in the induction of IL-8 and NF-κB at both protein and mRNA levels, respectively [105]. Furthermore, two recent studies have investigated the combinatory synergistic effect of luteolin with oxaliplatin in GC cells. The combined treatment demonstrated the positive effect of both agents on the inhibition of cellular proliferation, activation of CytC/caspase signaling, and the induction of cell cycle arrest (GC/M phase) and cellular apoptosis [106] (Figure 2).

5.3. Kaempferol. Kaempferol is a flavonoid compound that is abundantly found in various plants (edible and medicinal). Its various biological activities also include the anti-inflammatory property, which is apparently useful in inhibiting the expression of several proinflammatory cytokines, NF-κB, STAT1, and AP-1 [107, 108]. Therefore, kaempferol has been investigated and reported to be effective in many types of cancers, including GC [109, 110]. In a study conducted in Spain, kaempferol consumption reduced the risk of GC, while an in vivo study reported the inhibition of cancerous cell growth in GC xenografts, thereby suggesting the antiproliferative and metastasis-inhibiting ability of the flavonoid [111, 112]. Li et al. examined the effect of kaempferol against injury to the gastric mucosa, which was induced by ethanol [113]. The treatment with kaempferol demonstrated its protective effect by facilitating the inhibition of MPO and proinflammatory cytokine levels, as well as improving NO production in cells. In AGS cells, kaempferol leads to a decrease in the expression of IL8, IL-1β, and TNF-α. Moreover, its anti-inflammatory effect was observed by the suppression of the translocation of CagA and VacA proteins of H. pylori in GC cells [114]. Kaempferol is also reported to induce autophagy and apoptosis in GC cells via activating the IRE1-JNK-CHOP signaling and AMPK/aULK1 pathway and by positively regulating ER stress in GC cells [111, 115] (Figure 3). A recent study also shed light on the mode of action of the compound against GC through a network pharmacology approach [116]. Shrestha et al. reported the reduction in G9a expression, as well as inhibition of mTOR signaling and cellular proliferation in GC cells after kaempferol treatment [117].

5.4. Honokiol. Honokiol (3,5-di-(2-propenyl)-1,1-biphenyl-2,2-diol) is a small, biphenolic lignan. It has been reported to suppress Akt and NF-κB activation, which in turn results in the phosphorylation and degradation of IκBα, respectively [118]. It is also reported to mediate the suppression of STAT3 activity previously induced by IL-6, where the activated form of the former has been associated with many cancer cells [119]. The treatment with honokiol has been reported to be effective against GC, where it was attributable to the induction of apoptosis and downregulation of COX-2 and PPAR-γ in GC cells [120]. The significant anticancer activity was observed to be correlated with GRP94 levels, which were found to be reduced after administration of
**Figure 1:** Anticancer effect of quercetin against GC.

**Figure 2:** Anticancer mechanisms of luteolin against GC.
honokiol in mice in a dose-dependent manner [121]. Therefore, it can potentially serve as a promising anticancer therapeutic target for several pathways [122]. Liu et al. observed that honokiol increases SHP-1 activity that subsequently leads to the deactivation of the STAT3 pathway, thereby suggesting that honokiol actively inhibits cellular angiogenesis and proliferation of GC cells [123]. In a study, treatment with honokiol revoked the down- and upregulation of E-cadherin and TPL2, respectively, the latter of which was observed to be subsequently associated with decreased growth and vascular density in vivo [124]. This mechanism of action, combined with the inhibition of epithelial-to-mesenchymal transition as well as regulation of apoptosis (induced by ER stress), serves a key role in the therapeutic action of honokiol against GC. In a recent study, the anticancer properties of honokiol were attributable to its ability to downregulate PPAR-γ activity, as well as the expressions of CDC25C, CDC2, and Cyclin B1, which aid in inducing ER stress which in turn decreases vascular density [125].

5.5. D-Limonene. D-Limonene is a monoterpenic compound that has been reported to possess anticancer activities against different cancers [126, 127]. Lu et al. evaluated that d-limonene can inhibit the proliferation of GC cancer cells via the induction of apoptosis in cancer cells [128]. It has also been attributable to strong antioxidant activity resulting in the inhibition of H₂O₂-induced CAS3, CAS9, and p38/MAPK activation, as well as a decline in the BCL-2/Bax ratio, thereby indicating that it could offer protection against oxidative stress [129]. Another study evaluated that oral administration of limonene (≥400 mg/kg) in mice led to a reduction in tumor mass weight [128]. The combination of d-limonene with berberine and its singular treatment on GC cell line resulted in the increased expression of CAS3 and ROS, reduced expression of BCL-2, and cell cycle arrest, indicating that d-limonene causes apoptosis via the regulation of the mitochondrial pathway [130–132].

6. Bioactivity of ASF Compounds against *H. pylori*

*Helicobacter pylori* is often recognized to have a strong correlation with the occurrence of gastric diseases, including GC. Many bioactive compounds have been associated with the anti-*H. pylori* activity and other protective effects, which subsequently promote good gastric health and protection from several diseases. In a study, luteolin was observed to inhibit the activity of the arylamine N-acetyltransferase (NAT) enzyme, which is responsible for the N-acetylation of PABA and AF in *H. pylori* [133]. Another study demonstrated the protective effect of quercetin against *H. pylori* in the corpus mucosa [134]. Though the main mechanism of
Table 1: The anticancer effect of various bioactive compounds of ASF against gastric cancer in vitro.

| Compound     | Cell line                          | Concentration used                  | Effect on protein/pathway (s)                                                                 | References |
|--------------|------------------------------------|-------------------------------------|---------------------------------------------------------------------------------------------|------------|
| Quercetin    | AGS and MKN28                       | 10–160 μM                           | Inhibit Akt-mTOR pathway                                                                     | [85]       |
|              | AGS                                 | Quercetin alone (6.25, 12.5, 25, 50, and 100 μM) With SN-38 (5-25 nM) | Downregulate VEGFA and VEGFR-2 ↓ COX-2, Twist1, and ITGβ6 Inhibit EBNA-1 and LMP-2 proteins | [91]       |
|              | SNU719 and MKN74                     | N/A                                 | ↑ Cleaved CAS3, CAS9, and PARP Induce p53, Bax, and Puma Induce CAS3, Bcl-2, and Bax          | [90]       |
|              | BGC-823                             | 5, 30, 60, 90, and 120 μmol/L       | ↓ Bcl-2/Bax ratio ↑ CAS3 expression ↓ Cell migration and invasion                           | [82]       |
|              | BGC823 and AGS                       | 10 μM                               | ↓ MMP2 and MMP9 activity inhibit Pak1-Limk1-cofilin, NF-κB, PKC-δ, and ERK1/2 signaling, and AMPKα activation | [86]       |
|              | GCSC                                | 20–100 μM                           | Inhibit (PI3K)-Akt signaling                                                                | [84]       |
|              | HGC-27, NUGC-2, MKN-7, and MKN-28   | 70 μM (IC50=32–55 μM)               | Cell cycle arrest (Gi to S phase)                                                            | [87]       |
|              | AGS                                 | 50 μM (24h)                         | ↓ CDC2, cyclin B1, and CDC25C levels ↑ Apoptosis, CAS3, CAS6, CAS9, Bax, and p53 ↓ BCL-2    | [96]       |
|              | AGS                                 | 80 μM (48 and 72 h)                 | ↑ IL-8 expression ↑ NF-κB mRNA expression ↑ Cleaved CAS3 and PARP; induce apoptosis          | [105]      |
|              | CRL-1739                            | 30 μM                               | Downregulate MMP9 expression and c-Met/Akt/ERK signaling ↑ Apoptosis                          | [97]       |
|              | MKN45 and SGC7901                   | 20 μM (24h)                         | Inhibit GC cell proliferation, cyclin D1, cyclin E, BCL-2, MMP2, MMP9, N-cadherin, and vimentin Induce p21, Bax, E-cadherin expression, Notch1, PI3K, AKT, mTOR, ERK, STAT3, and p38 signaling pathway | [98]       |
|              | MKN45 and BGC823                    | 40 μM                               | ↑ Cleaved Bax and CAS3 Downregulate ERK1/2 phosphorylation and activation ↑ Apoptosis         | [100]      |
| Luteolin     | BGC-823                             | 0–60 μM (48 h)                      | ↑ Cleaved PARP and CAS3 Downregulate ERK1/2 phosphorylation and activation ↑ Apoptosis        | [106]      |
|              | MFC                                 | Luteolin alone (20 μM) and/or oxaliplatin (5 μM (24h)) | Combined treatment induced cell cycle arrest (G2/M phase) Induce apoptosis Combined treatment inhibited proliferation Induce ERK1/2 phosphorylation, JNK, and P38 MAPK signal transduction Inhibit PI3K/AKT and ERK1/2 MAPK intracellular signaling Induce apoptosis | [106]      |
|              | SGC-7901                            | 40 μM (24h)                         | ↓ Cyclin B1, CDK1 and CDC25C, COX-2, p-AKT, and p-ERK ↑ Cleaved CAS3, CAS9, PARP           | [101]      |
|              | MKN28, SGC7901, and GSE-1           | 60 μM                               | Activate IRE1-INK-CHOP signaling pathway Induce apoptosis LC3-I to LC3-II conversion Downregulate p62 | [112]      |
| Kaempferol   | AGS, SNU-216, NCI-N87, SNU-638, and MKN-74 | 25 μM, 50 μM, and 100 μM (24 h)     | Induce apoptosis Activate IRE1-INK-CHOP signaling pathway                                    | [117]      |
|              | Rh30                                | 25 or 50 μM kaempferol and quercetin (24 h) | Induce apoptotic markers (cleaved PARP and CAS3) Inhibit cell growth, survival, migration, and invasion by blocking mTOR signaling | [117]      |
action of quercetin is not clear, it is reported to be associated with the declining activity of urease, as well as its ability to chelate iron, which is a major cofactor imperative for \textit{H. pylori} growth. Treatment of quercetin in \textit{H. pylori}-infected animals is reported to reduce the process of inflammation and bacterial count in the gastric mucosa [135].

\textit{β}-Caryophyllene is a naturally occurring bicyclic sesquiterpene that is widely found in many medicinal plants and is reported to possess many protective abilities, including antibacterial and anti-inflammatory properties. In a study, \textit{β}-caryophyllene demonstrated significant gastroprotective activity in an ethanol-induced gastric ulcer model, reducing the lesions by more than 70% [136]. A recent study by Shim et al. revealed that the treatment of \textit{H. pylori}-caused gastrointestinal disease with \textit{β}-caryophyllene demonstrated remarkable improvement in physiological symptoms and subsequently resulted in IL-1\textit{β} decrease in serum [137]. Furthermore, the anti-\textit{H. pylori} action of similar compounds has also been well-reported in many studies [114, 138, 139].

7. Future Perspectives and Conclusions

Medicinal herbs such as ASF have been traditionally used for the treatment of several ailments. These herbs are deemed indispensable as therapeutic candidates for treating various diseases such as cancer. The protective effects of ASF and its compounds against GC are majorly related to modulating major hallmarks of cancer, such as the inhibition of cellular proliferation, angiogenesis, inducing apoptosis as well as the suppression of cell migration, and the promotion of immune cell secretion against GC cells (Table 1). Furthermore, the ability to use ASF and its compounds in combination with other anticancer agents and/or as adjuvants in cancer immunotherapy is also an exciting and encouraging field of study, as results have been promising in recent investigations. These properties could provoke great economic interest for their pharmaceutical applications when the compounds are extracted, as compared to their laboratory synthesis. In this regard, future studies and phytochemical analyses of ASF and its bioactive compounds may result in the discovery of novel anti-GC targets and may also lead to improvements in their chemical synthesis. This approach has been further elucidated in studies of single, isolated compounds, such as quercetin, where its standalone efficacy against GC is observed to be far greater than the herb on the whole. Therefore, these phytochemicals pose a superior anti-GC potential, which could be elucidated in further \textit{in vitro} and \textit{in vivo} studies and clinical trials.

Data Availability

No data were used to support this study.

Conflicts of Interest

The authors declare no conflicts of interest.

Authors’ Contributions

All authors contributed equally to this work.

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References

[1] B. E. Clurman and J. M. Roberts, “Cell cycle and cancer,” \textit{Journal of the National Cancer Institute}, vol. 87, no. 20, pp. 1499–1501, 1995.

[2] Y.-J. Surh, “Cancer chemoprevention with dietary phytochemicals,” \textit{Nature Reviews Cancer}, vol. 3, no. 10, pp. 768–780, 2003.

[3] Y. Yamaoka, “How to eliminate gastric cancer-related death worldwide?” \textit{Nature Reviews Clinical Oncology}, vol. 15, no. 7, pp. 407–408, 2018.

[4] F. Bray, J. Ferlay, I. Soerjomataram, R. L. Siegel, L. A. Torre, and A. Jemal, “Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries,” \textit{CA: A Cancer Journal for Clinicians}, vol. 68, no. 6, pp. 394–424, 2018.
[73] M. A. Rashid and R. H. Zuberi, "Pharmacognostical studies of standardisation for a medicinal spice, the fruit of Illicium verum Hook. F.," *PharmacTutor*, vol. 4, pp. 36–41, 2016.

[74] J. K. Patra, G. Das, S. Bose et al., "Star anise (Illicium verum): chemical compounds, antiviral properties, and clinical relevance," *Phytotherapy Research*, vol. 6614, pp. 1–20, 2020.

[75] M. Sharafan, K. Jafernik, H. Ekiert et al., "Illicium verum (Star anise) and trans-anethole as valuable raw materials for medicinal and cosmetic applications," *Molecules*, vol. 27, no. 3, pp. 650–665, 2022.

[76] J. A. Al Mollah, "Spices, herbal xenobiotics and the stomach: friends or foes?" *World Journal of Gastroenterology*, vol. 16, pp. 2710–2719, 2010.

[77] M. Asif, A. H. S. Yehya, M. A. Al-Mansoub et al., "Anticancer attributes of Illicium verum essential oils against colon cancer," *South African Journal of Botany*, vol. 103, pp. 156–161, 2016.

[78] C. Chen, J. Zhou, and C. Ji, "Quercetin: a potential drug to reverse multidrug resistance," *Life Sciences*, vol. 87, no. 11–12, pp. 333–338, 2010.

[79] M. Erboga, C. Aktas, Z. F. Erboga, Y. B. DONMEZ, and C. Chen, J. Zhou, and C. Ji, "Quercetin: a potential drug to reverse multidrug resistance," *Molecules*, vol. 10, no. 10, pp. 2866–2870, 2010.

[80] S. Ranganathan, D. Halagowder, and N. D. Sivasithambaram, "Quercetin suppresses twist to induce apoptosis in MCF-7 breast cancer cells," *PLoS One*, vol. 10, no. 10, Article ID e014370, 2015.

[81] A. Kahraman, N. Erkasp, T. Koken, M. Serteser, F. Aktepe, and S. Erkasp, "The antioxidative and antihistaminic properties of quercetin in ethanol-induced gastric lesions," *Toxicology*, vol. 183, no. 1–3, pp. 133–142, 2003.

[82] P. Wang, K. Zhang, Q. Zhang et al., "Effects of quercetin on the apoptosis of the human gastric carcinoma cells," *Toxicology in Vitro*, vol. 26, no. 2, pp. 221–228, 2012.

[83] A. M. Ekstrom, M. Serafini, O. Nyren, A. Wolk, C. Bosetti, and R. Bellocchio, "Dietary quercetin intake and risk of gastric cancer: results from a population-based study in Sweden," *Annals of Oncology: Official Journal of the European Society for Medical Oncology*, vol. 22, no. 2, pp. 438–443, 2011.

[84] X. Shen, Y. Si, Z. Wang, J. Wang, Y. Guo, and X. Zhang, "Quercetin inhibits the growth of human gastric cancer stem cells by inducing mitochondrial-dependent apoptosis through the inhibition of PI3K/Akt signaling," *International Journal of Molecular Medicine*, vol. 38, no. 2, pp. 619–626, 2016.

[85] K. Wang, R. Liu, J. Li et al., "Quercetin induces protective autophagy in gastric cancer cells: involvement of Akt-mTOR- and hypoxia-induced factor 1α-mediated signaling," *Autophagy*, vol. 7, no. 9, pp. 966–978, 2011.

[86] H. Li and C. Chen, "Quercetin has antimetastatic effects on gastric cancer cells via the interruption of uPA/uPAR function by modulating NF-xb, PKC-δ, ERK1/2, and AMPKα," *Integrative Cancer Therapies*, vol. 17, no. 2, pp. 511–523, 2017.

[87] M. Yoshida, T. Sakai, N. Hosokawa et al., "The effect of quercetin on cell cycle progression and growth of human gastric cancer cells," *FEBS Letters*, vol. 260, no. 1, pp. 10–13, 1990.

[88] J.-Y. Zhang, M.-T. Lin, M.-J. Zhou et al., "Combinational treatment of curcumin and quercetin against gastric cancer MGC-803 cells in vitro," *Molecules*, vol. 20, no. 6, pp. 11524–11534, 2015.
signaling pathway,” Molecular Oncology, vol. 10, no. 9, pp. 1473–1484, 2016.

[104] M. Imran, A. Rauf, T. Abu-Izneid et al., “Luteolin, a flavonoid, as an anticancer agent: a review,” Biomedicine and Pharmacotherapy, vol. 112, Article ID 108710, 2019.

[105] M. Borzym-Kluczyk and K. Leszczynska, “Luteolin alters MUC1 extracellular domain, sTantigen, ADAM-17, IL-8, IL-10 and NF-kB expression in Helicobacter pylori-infected gastric cancer CRL-1739 cells: a preliminary study,” Biomedical Reports, vol. 14, no. 2, pp. 19–27, 2021.

[106] J. Ma, X. Chen, X. Zha et al., “Luteolin potentiates low dose oxaliplatin induced inhibitory effects on cell proliferation in gastric cancer by inducing G2/M cell cycle arrest and apoptosis,” Oncology Letters, vol. 23, pp. 16–28, 2022.

[107] S.-H. Lee, Y.-J. Kim, S.-H. Kwon et al., “Inhibitory effects of flavonoids on TNF-alpha-induced IL-8 gene expression in HEK 293 cells,” BMB Reports, vol. 42, no. 5, pp. 265–270, 2009.

[108] S. Chen, J. Ma, L. Yang et al., “Anti-glioblastoma activity of kaempferol via programmed cell death induction: involvement of autophagy and pyroptosis,” Frontiers in Bioengineering and Biotechnology, vol. 8, pp. 1–10, 2020.

[109] W. Liao, L. Chen, X. Ma, R. Jiao, X. Li, and Y. Wang, “Protective effects of kaempferol against reactive oxygen species-induced hemolysis and its antiproliferative activity on human cancer cells,” European Journal of Medicinal Chemistry, vol. 114, pp. 24–32, 2016.

[110] M. Swiec, A. Herok, K. Piwowarczyk et al., “Potentially bioaccessible phenolics from mung bean and adzuki bean sprouts enriched with probiotic-antioxidant properties and effect on the motility and survival of AGS human gastric carcinoma cells,” Molecules, vol. 25, no. 13, pp. 2963–2975, 2020.

[111] R. Garcia-Closas, C. A. Gonzalez, A. Agudo, and E. Riboli, “Intake of specific carotenoids and flavonoids and the risk of gastric cancer in Spain,” Cancer Causes and Control, vol. 10, no. 1, pp. 71–75, 1999.

[112] T. W. Kim, S. Y. Lee, M. Kim, C. Cheon, and S. G. Ko, “Kaempferol induces autophagic cell death via IRE1-JNK-CHOP pathway and inhibition of G9a in gastric cancer cells,” Cell Death & Disease, vol. 9, no. 9, pp. 875–914, 2018.

[113] Q. Li, X. Hu, Y. Xuan et al., “Kaempferol protects ethanol-induced gastric ulcers in mice via pro-inflammatory cytokines and NO,” Acta Biochimica et Biophysica Sinica, vol. 50, no. 3, pp. 246–253, 2018.

[114] M. J. Yeon, M. H. Lee, D. H. Kim et al., “Anti-inflammatory effects of kaempferol on Helicobacter pylori-induced inflammation,” Bioscience Biotechnology and Biochemistry, vol. 83, no. 1, pp. 166–173, 2019.

[115] P. B. Bhosale, S. E. Ha, P. Vetrivel, H. H. Kim, S. M. Kim, and G. S. Kim, “Functions of polyphenols and its anticancer properties in biomedical research: a narrative review,” Translational Cancer Research, vol. 9, no. 12, pp. 7619–7631, 2020.

[116] L. Yang, H. Li, M. Yang et al., “Exploration in the mechanism of kaempferol for the treatment of gastric cancer based on network pharmacology,” BioMed Research International, vol. 2020, Article ID 5891016, 11 pages, 2020.

[117] R. Shrestha, K. Mohankumar, G. Martin et al., “Flavonoids kaempferol and quercetin are nuclear receptor 4A1 (NR4A1, Nur77) ligands and inhibit rhodanymosarcocoma cell and tumor growth,” Journal of Experimental Clinical Cancer Research, vol. 40, no. 392, pp. 1–17, 2021.

[118] S. I. Grivennikov and M. Karin, “Dangerous liaisons: STAT3 and NF-kappaB collaboration and crosstalk in cancer,” Cytokine Growth Factor Reviews, vol. 21, no. 1, pp. 11–19, 2010.

[119] P. Rajendran, F. Li, M. K. Shanmugam et al., “Honokiol inhibits signal transducer and activator of transcription-3 signaling, proliferation, and survival of hepatocellular carcinoma cells via the protein tyrosine phosphatase SHP-1,” Journal of Cellular Physiology, vol. 227, no. 5, pp. 2184–2195, 2012.

[120] S. H. Liu, C. C. Shen, Y. C. Yi et al., “Honokiol inhibits gastric tumourigenesis by activation of 15-lipoxigenase-1 and consequent inhibition of peroxisome proliferator-activated receptor-gamma and COX-2-dependent signals,” British Journal of Pharmacology, vol. 160, pp. 1963–1972, 2010.

[121] M. L. Sheu, S. H. Liu, and K. H. Lan, “Honokiol induces calpain-mediated glucose-regulated protein-94 cleavage and apoptosis in human gastric cancer cells and reduces tumor growth,” PLoS One, vol. 2, no. 10, Article ID e1096, 2007.

[122] S. Arora, S. Singh, G. A. Piazza, C. M. Contreras, J. Panyam, and A. P. Singh, “Honokiol: a novel natural agent for cancer prevention and therapy,” Current Molecular Medicine, vol. 12, no. 10, pp. 1244–1252, 2012.

[123] S. H. Liu, K. B. Wang, K. H. Lan et al., “Calpain/SHP-1 interaction by honokiol dampening peritoneal dissemination of gastric cancer in nu/nu mice,” PLoS One, vol. 7, no. 8, Article ID e43711, 2012.

[124] H.-C. Pan, D.-W. Lai, K.-H. Lan et al., “Honokiol thwarts gastric tumor growth and peritoneal dissemination by inhibiting Tpl2 in an orthotopic model,” Carcinogenesis, vol. 34, no. 11, pp. 2568–2579, 2013.

[125] A. Rauf, S. Patel, M. Imran et al., “Honokiol: an anticancer lignan,” Biomedicine and Pharmacotherapy, vol. 107, pp. 555–562, 2018.

[126] I. Kaji, M. Tatsuta, H. Iishi, M. Baba, A. Inoue, and H. Kasugai, “Inhibition by d-limonene of experimental hepatocarcinogenesis in sprague-dawley rats does not involve p21ras plasma membrane association,” International Journal of Cancer, vol. 93, no. 3, pp. 441–444, 2001.

[127] K. Z. Guyton and T. W. Kensler, “Prevention of liver cancer,” Current Oncology Reports, vol. 4, no. 6, pp. 464–470, 2002.

[128] X.-G. Lu, L. B. Zhan, B. A. Feng, M. Y. Qu, L. H. Yu, and H. Kasugai, “Inhibition by d-limonene of experimental hepatocarcinogenesis in sprague-dawley rats does not involve p21ras plasma membrane association,” International Journal of Cancer, vol. 93, no. 3, pp. 441–444, 2001.

[129] J. H.-Y. Zhang, “Synergistic inhibitory effect of berberine and honokiol,” Frontiers in Sustainable Food Systems, vol. 2, no. 10, Article ID e1096, 2007.

[130] X.-Z. Zhang, L. Wang, D.-W. Liu, G.-Y. Tang, and Z.-Q. Zou, “D-Limonene against oxidative stress-induced cell damage in human lens epithelial cells via the p38 pathway,” Oxidative Medicine and Cellular Longevity, vol. 2016, Article ID 5962832, 12 pages, 2016.

[131] X. Yu, H. Lin, Y. Wang et al., “D-limonene exhibits anti-tumor activity by inducing autophagy and apoptosis in lung cancer,” OncoTargets and Therapy, vol. 11, pp. 1833–1847, 2018.

[132] X.-Z. Zhang, L. Wang, D.-W. Liu, G.-Y. Tang, and H.-Y. Zhang, “Synergistic inhibitory effect of berberine and d-limonene on human gastric carcinoma cell line MGC803,” Journal of Medicinal Food, vol. 17, no. 9, pp. 955–962, 2014.

[133] J. Zhou, M. Azrad, and L. Kong, “Effect of limonene and berberine on human cancer cells,” Frontiers in Sustainable Food Systems, vol. 5, pp. 1–11, 2021.

[134] J. L. Zhou, S. S. Hsia, M. H. Kuo et al., “Inhibitory actions of luteolin on the growth and arylamine N-acetyltransferase
activity in strains of *Helicobacter pylori* from ulcer patients,” *Toxicology in Vitro*, vol. 15, no. 3, pp. 191–198, 2001.

[134] R. González-Segovia, J. L. Quintanar, E. Salinas, R. Ceballos-Salazar, F. Aviles-Jiménez, and J. Torres-López, “Effect of the flavonoid quercetin on inflammation and lipid peroxidation induced by *Helicobacter pylori* in gastric mucosa of Guinea pig,” *Journal of Gastroenterology*, vol. 43, no. 6, pp. 441–447, 2008.

[135] A. Haghi, H. Azimi, and R. Rahimi, “A comprehensive review on pharmacotherapeutics of three phytochemicals, curcumin, quercetin, and allicin, in the treatment of gastric cancer,” *Journal of Gastrointestinal Cancer*, vol. 48, no. 4, pp. 314–320, 2017.

[136] M. Lemos, J. R. Santin, C. S. Mizuno et al., “*Copaifera langsdorffii*: evaluation of potential gastroprotective of extract and isolated compounds obtained from leaves,” *Revista Brasileira de Farmacognosia*, vol. 25, no. 3, pp. 238–245, 2015.

[137] H. I. Shim, D. J. Song, C. M. Shin et al., “Inhibitory effects of β-caryophyllene on *Helicobacter pylori* infection: a randomized double-blind, placebo-controlled study,” *Korean Journal of Gastroenterology*, vol. 74, no. 4, pp. 199–204, 2019.

[138] A. González, S. Salillas, A. Velázquez-Campoy et al., “Identifying potential novel drugs against *Helicobacter pylori* by targeting the essential response regulator HsrA,” *Scientific Reports*, vol. 9, no. 1, pp. 11294–11313, 2019.

[139] H. T. Trung, H. T. T. Huynh, L. N. T. Thuy, H. N. Van Minh, M. N. T. Nguyen, and M. N. L. Thi, “Growth-inhibiting, bactericidal, antibiofilm, and urease inhibitory activities of *Hibiscus rosa-sinensis* L. flower constituents toward antibiotic sensitive-and resistant-strains of *Helicobacter pylori*,” *ACS Omega*, vol. 5, no. 32, pp. 20080–20089, 2020.