INTRODUCTION

Arthroplasty can fail because of aseptic loosening, instability, periprosthetic fracture or infection of the prosthesis. In general, the incidence of periprosthetic joint infection (PJI) is lower than aseptic loosening, but it is more serious and complex complication following arthroplasty. The haematogenous seeding could happen at any time during the rest of patient’s life after surgery. For doctor, the treatment and diagnose PJI are still challenging in the modern world. The implant as a foreign body increases the pathogenicity of bacteria and the presence of biofilm makes the diagnosis and treatment complex and difficult. Therefore, appropriate management protocol of PJI should be established, in a timely manner to take preventive measure and diagnosis, and accordant strictly with the results of culture and antimicrobial susceptibility test, to avoid the increase in the rate of bacterial resistance and select reasonably antibiotics combined with adequate operation procedure. Eventually, it achieves eradicating of infection, preserving joint function without any pain.

EPIDEMIOLOGY

The joint arthroplasty is a safe and cost-effective surgical technique. It can alleviate symptoms, recover functions and enhance quality of life, especially in the elderly population. With the increment of the joint prosthetic replacements, the number of postoperative complications has also increased. The infection rate after hip or shoulder replacement is usually less than 1%, after knee replacement less than 2% and after elbow replacement is between 1.9% and 10.3%. The reason for higher incidence in elbow region may be related to the more frequent rheumatic disorder, trauma or multiple reconstructive procedures compared to hip and knee surgery.

PJI also increases the medical costs, which are up to 24 times higher than without PJI, the major cost of PJI is generated by prolonged hospitalization, multiple surgeries and prostheses, and medical supplies.

Coagulase-negative staphylococci and Staphylococcus...
 aureus are the most common microorganisms in hip and knee PJI. Furthermore some pathogenic bacteria depend on different body regions, such as Propionibacterium acnes after shoulder replacement and gram-negative bacteria after hip arthroplasty.

**PATHOGENESIS AND CLASSIFICATION**

Development stages of the biofilm formation are divided into 4 phases; namely adhesion, proliferation, biofilm maturation and cellular detachment. The bacterium adheres to the foreign body material that is the first step which can cause biofilm-related infections. The materials of orthopedic implant (such as titanium, ceramics, hydroxyapatite, and polyethylene) is easy for the bacteria to colonize on. Biofilms is a complex community of microorganisms embedded in an extracellular matrix that forms on surfaces of prosthesis.

The prosthesis can get infected by three pathways: first, perioperative period, most commonly through intraoperative inoculation; second, haemogenous could happen at any time after implantation, pathogen from different parts, e.g. respiratory or urinary tract infection, skin infection and pneumonia; and third, direct contact with a nearby infected, e.g. infected soft tissue, septic arthritis or osteomyelitis. Table 1 shows the classification of PJI into acute and chronic infection. The early (immature) biofilm can be eradicated without remove prosthesis. If symptoms persist less than 4 weeks (hematogenous) or infection manifests less than 4 weeks after surgery (perioperative). Other procedures is defined as chronic PJI, the implant must be changed due to mature biofilm.

**DIAGNOSIS**

Early diagnosis is a positive factor to save the prosthesis and the joint function. No single indicator of a test using in clinic or in laboratory can hand out ideal sensitivity and specificity aiming to the diagnosis of PJI, so a mixture of multiple tests can reasonably increase the diagnostic accuracy. Some criteria from the Musculoskeletal Infection Society (MSIS), European Bone and Joint Infection Society (EBJIS) and Infectious Diseases Society of America (IDSA) have been recently published. The EBJIS criteria are more sensitive for the diagnosis of PJI than other criteria (Table 2).

**1. Laboratory Studies**

If PJI is suspected, serum C-reactive protein (CRP) is usually performed. This marker is inexpensive, rapid and has a better performance than erythrocyte sedimentation rate (ESR). However, CRP is not specific for infection and usually increases due to healing process after intervention. Pérez-Prieto et al. found that with normal CRP about one-third of chronic, low-grade infections would be missed. So the combination of CRP and ESR is used for diagnose the infection. It was reported that the sensitivity of combined ESR (>30 mm/hr) and CRP (>10 mg/L) was 96%, but low specificity 56%.

Synovial fluid aspiration and culture are the most accurate preoperative examination for the diagnosis of PJI. Synovial fluid leukocyte count and granulocyte percentage can accurately distinguish PJI from aseptic failure, and has a sensitivity of 86% compared with synovial fluid culture (52%). The patient with rheumatoid arthritis,

| Table 1. Classification of Periprosthetic Joint Infection (PJI) into Acute and Chronic Infection |
|----------------------------------|------------------|------------------|
| **Type of PJI**                  | **Acute PJI**    | **Chronic PJI**  |
| Pathogenesis:                    |                  |                  |
| - Perioperative origin           | Early postoperative | Delayed postoperative (low-grade) |
| - Hematogenous origin            | <4 weeks after surgery | ≥4 weeks after surgery |
| Biofilm age (maturity)           | <3 weeks of symptoms | ≥3 weeks of symptoms |
| Clinical features                | Immature | Mature |
| Causative microorganism          | Acute joint pain, fever, red/swollen joint | Chronic pain, loosening of the prosthesis, sinus tract (fistula) |
| Surgical treatment               | Débridement & retention of prosthesis (change of mobile parts) | Low-virulent: Coagulase-negative staphylococci (e.g. Staphylococcus epidermidis), Propionibacterium acnes |

[1] 1. Pérez-Prieto et al.
[2] 2. Coagulase-negative staphylococci (e.g. Staphylococcus epidermidis), Propionibacterium acnes
[3] 3. Change in one-, two-, or multiple stages

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periprosthetic fracture or dislocation and in the early postoperative period should be excluded, because the cell count shows falsely high\(^{11,16,21,22,24}\).

### 2. Histopathological Studies

Histopathological examination has a high sensitivity (95%) and specificity (92%) for the diagnosis of PJI. Based on histomorphological criteria, four types of periprosthetic membranes have been defined, wear particle-induced, infectious, combined and indeterminate type\(^{25}\). Nevertheless, the nature and degree of infiltration with inflammatory cells may vary markedly among specimens and even within individual tissue sections from the same patient. There is no comprehensively accepted definition about acute inflammation; normally an acute inflammation has been defined as \( \geq 2 \) granulocytes per high-power field.\(^{12}\)

Acute inflammation is variably defined as from \( \geq 1 \) to \( \geq 10 \) neutrophils per high-power field.\(^{12}\)

Leukocyte cutoffs are not interpretable within 6 weeks of surgery, in rheumatic joint disease, periprosthetic fracture or luxation. Leukocyte count should be determined within 24 hours; clotted specimens are treated with 10 μL hyaluronidase.\(^{12}\)

For highly virulent organisms (e.g., *Staphylococcus aureus, Escherichia coli*), already one positive sample confirms infection.\(^{12}\)

Under antibiotics and for anaerobes, <50 colony-forming unit (CFU)/mL can be significant.

### 3. Microbiological Studies

1) **Preoperative aspiration**

The synovial fluid culture has a sensitivity ranging from 50% to 70% and should be performed before revision surgeries (together with the determination of leukocyte count in the synovial fluid).

2) **Intraoperative specimens**

Intraoperative tissue samples provide accurate specimens for detecting the infecting microorganism(s), sensitivity ranging from 45% to 78%, and specificity from 91% to 96%\(^{23,25,27}\). At least three to five intraoperative tissue samples from different anatomical sites should be sampled for culture. Samples should always be collected from a zone in which the tissue structure is visibly inflamed, because it is informative. Prior to collecting microbiological samples, any antibiotic regimen should be discontinued for 2 weeks to progress the disease\(^{28}\).

3) **Sonication for removed implant**

Sonication is used for dislodging adherent microorganisms from the surface of prosthetic joint. The sonication fluid culture proves the higher sensitivity and specificity than periprosthetic tissue culture which is also valid at the patient who has received antibiotic treatment before surgery. Discontinued antibiotic therapy within 14 days will have higher sensitivity\(^{29}\). A study\(^{30}\) about inoculation of sonication fluid in blood culture bottles (BCB) greatly improves the result, even when the patients received antibiotics. The sonication fluid in BCB has 100% sensitivity and specificity, even the half of patients received antibiotics within 14 days. But the sensitivity is 87% for conventional synovial fluid culture and 59% in tissue culture. The sonication fluid in BCB also reduces the culture time; it detects all bacteria in only 5 days. Another study demonstrated that the sonication fluid in BCB has better sensitivity than agar plate culture and also reduces the culture time than agar plate\(^{30}\). But why sonication fluid in BCB increases the sensitivity is still unclear. The method of sonication fluid in BCB still need more clinic practice to find some details about this research.

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**Table 2. Definition of Periprosthetic Joint Infection, if at least one of the following 4 criteria is fulfilled**

| Diagnostic test                     | Criteria                                                                 | Sensitivity (%) | Specificity (%) |
|-------------------------------------|--------------------------------------------------------------------------|-----------------|-----------------|
| Clinical features                   | Sinus tract or visible purulence*                                        | 20-30           | 100             |
| Histology in periprosthetic tissue  | Acute inflammation in periprosthetic tissue\(^{1}\)                      | 95-98           | 95-98           |
| Leukocyte count in synovial fluid\(^{1}\) | \( >2,000/\mu L \) leukocytes or \( \geq 70\% \) granulocytes          | 93-96           | 93-96           |
| Microbiology (culture)              | Synovial fluid or Tissue samples\(^{1}\) or Sonication fluid (\( \geq 50 \) CFU/mL)\(^{1}\) | 60-80           | 97              |
|                                     |                                                                          | 70-85           | 92              |
|                                     |                                                                          | 85-95           | 95              |

* Metal-on-metal bearing components can simulate pus, but leukocyte count is usually normal, but metal debris visible.

\(^{1}\) Acute inflammation defined as \( \geq 2 \) granulocytes per high-power field.

\(^{1}\) Leukocyte cutoffs are not interpretable within 6 weeks of surgery, in rheumatic joint disease, periprosthetic fracture or luxation. Leukocyte count should be determined within 24 hours; clotted specimens are treated with 10 μL hyaluronidase.

\(^{1}\) For highly virulent organisms (e.g., *Staphylococcus aureus, Escherichia coli*), already one positive sample confirms infection.

\(^{1}\) Under antibiotics and for anaerobes, <50 colony-forming unit (CFU)/mL can be significant.
Conventional radiography is the most used in the first-step imaging diagnosis of PJI\(^5\). However, the sensitivity and specificity of X-ray plain film in the diagnosis of infection are low, and it is difficult to distinguish between aseptic loosening and PJI\(^3\). Computed tomography (CT) imaging occupies good contrast resolution of bone and surrounding soft tissue. Magnetic resonance imaging (MRI) can be used safely on patients with non-ferrimagnetic implants. MRI displays greater resolution for soft tissue abnormalities than CT and radiography and does not involve radiation. However, the patients must remain in an enclosed machine, which may be extremely problematic for claustrophobic patients. The main disadvantage of CT and MRI is imaging interference in the vicinity of metallic orthopedic implants. Fluorine 18-fluorodeoxyglucose (FDG) positron emission tomography (PET) is a fast, safety, high-quality imaging for detection of PJI\(^2\). One meta-analysis report\(^3\) pooled sensitivity and specificity of FDG-PET for the diagnosis of prosthetic hip or knee joint infection were 82.1% and 86.6%.

Polymerase chain reaction (PCR) techniques can identify an isolated bacterium and some difficult-to-cultured bacteria. When the patient receives antibiotics, the sensitivity and specificity are still high by multiplex PCR for diagnosis PJI and to distinguish aseptic loosening\(^4\). According to a meta-analysis of 14 studies\(^5\), the sensitivity and specificity of PCR in synovial fluid samples were 84% and 89% and the PCR in sonication fluid culture were 81% and 96% for the detection of PJI. The sensitivity in fresh samples was better than using frozen samples.

The alpha defensin lateral flow (ADLF) is a rapid biomarker for test PJI, but the sensitivity and specificity are a controversial idea\(^8\)-\(^3\). In a recent study\(^1\), the ADLF test was detected PJI with sensitivity and specificity in the criteria of MSIS 84.4% and 96.4%, IDSA 67.3% and 95.5%, and EBJIS 54.4% and 99.3%. The EBJIS criteria used for ADLF test are not a good screening diagnose to rule out the PJI, while it could be a good method to confirm PJI.

Microcalorimetry is able to be used to rapidly detect the existence of microorganisms through measuring microbial heat produced by microbial growth and metabolism. A study reports that in microcalorimetry of sonication fluid

**Fig. 1. Treatment algorithm of periprosthetic joint infection (PJI).**
TREATMENT

The goals of PJI treatment are to alleviate suffering, restore the normal joint function and eliminate the infection. Treatment decisions should be individualized, and involve a cooperation of a multi-disciplinary team in order to tender the best approach for each patient based on a critical review of the current information. An appropriate operation combining with antimicrobial concept is required for successful treatment. The existing recommendations for treatment of the PJI have been refined further by new scientific evidences and clinical experiences, as optimized and summarized in a surgical and antibiotic treatment algorithm in Fig. 1.

1. Surgical Therapy

Major surgical strategy for the treatment of PJI includes; débridement and implant retention, one-stage or two-stage implant replacement (Fig. 2).

1) Débridement with retention

Early studies of débridement combining with retention strategies to treat prosthetic joint infection have high failure rates. However, success rates can be greater than 80% when the following conditions are met: (1) prosthesis is stable; (2) a pathogen with susceptibility to antimicrobial agents is active against surface-adhering microorganisms; (3) there is no sinus tract or compromised soft tissue; (4) symptom duration of infection is less than 3 weeks. Based on a recent report, 90% of orthopedic device-related infections are successfully cured by surgical débridement and implant-retention plus antimicrobial therapy according to a predefined treatment algorithm, if patients fulfill the above selection criteria and the pathogen is susceptible to rifampin (for gram-positive pathogens) or ciprofloxacin (for gram-negative pathogens).

2) One-stage implant replacement

One-stage exchange is a single operation, which includes the removal of the old and reimplantation of a new prosthesis. The operation is mostly used in Europe, whereas two-stage replacement is often used in United States. One-stage exchange is suitable for patients who have good bone conditions and soft tissue without sinus tract, as well as known bacteria with no difficult-to-treat (DTT) infections caused by pathogens resistant to biofilm-active antimicrobials. If based on the indication, the success rate of one-stage exchange could be reach 100%. In a single center report, the success rate of one-stage replacement is from 85% to 90% over 35 years. The one-stage exchange is an effective surgery with high success rate, earlier mobility, shorter period of hospitalization and less cost than two-stage exchange.

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Fig. 2. Surgical treatment of periprosthetic joint infection (PJI).
wk: week; i.v.: intravenously; p.o.: per oral.
### Table 3. Antimicrobial Treatment of Periprosthetic Joint Infection

| Microorganism                        | Antibiotic (dose), check pathogen susceptibility                  | Duration  | Followed by                                                                 |
|--------------------------------------|------------------------------------------------------------------|-----------|-----------------------------------------------------------------------------|
| **Staphylococcus spp.**              |                                                                  |           |                                                                             |
| Oxacillin-/methicillin-susceptible   | Cefazolin (3 × 2 g, i.v.)*+                                       | 2 wk      | According to susceptibility,                                                |
|                                      | Rifampin (2 × 450 mg, p.o.)                                       |           | Levofloxacin (2 × 500 mg, p.o.) or                                         |
|                                      |                                                                  |           | Cotrimoxazol (3 × 960 mg, p.o.) or                                         |
|                                      |                                                                  |           | Doxycyclin (2 × 100 mg, p.o.) +Rifampin (2 × 450 mg, p.o.)                  |
| Oxacillin-/methicillin-resistant     | Vancomycin (2 × 1 g, i.v.)*+                                      | 2 wk      | Same combination as above for oxacillin-/methicillin-susceptible staphylococci |
|                                      | Rifampin (2 × 450 mg, p.o.)                                       |           |                                                                             |
| Rifampicin-resistant†                | Vancomycin (2 × 1 g, i.v.)*†                                      | 2 wk      | Long-term suppression for ≥1 yr, depending on susceptibility (e.g., with cotrimoxazol, doxycycline or clindamycin) |
| **Streptococcus spp.**               |                                                                  |           |                                                                             |
|                                      | Penicillin G (4 × 5 million U, i.v.)* or                          | 2 wk      | Amoxicillin (3 × 1,000 mg, p.o.) or                                         |
|                                      | Ceftriaxon (1 × 2 g, i.v.)                                        |           | Levofloxacin (2 × 500 mg, p.o.)                                            |
|                                      |                                                                  |           | (consider suppression for 1 year)                                          |
| **Enterococcus spp.**                |                                                                  |           |                                                                             |
| Penicillin-susceptible               | Ampicillin (4 × 2 g, i.v.)*+                                      | 2-3 wk    | Amoxicillin (3 × 1,000 mg, p.o.)                                           |
|                                      | Gentamicin (2 × 60-80 mg, i.v.)*                                  |           |                                                                             |
| Penicillin-resistant†                | Vancomycin (2 × 1 g, i.v.)*†                                      | 2-4 wk    | Linezolid (2 × 600 mg, p.o.), maximum 4 wk                                  |
| Vancomycin-resistant                 | Individual; removal of the implant or life-long suppression necessary |           |                                                                             |
| Gram-negative bacteria               |                                                                  |           |                                                                             |
| Enterobacteriaceae (Escherichia coli, Klebsiella, Enterobacter, etc.) | Ciprofloxacin (2 × 750 mg, p.o.)                                 |           |                                                                             |
| Nonfermenters (Pseudomonas aeruginosa, Acinetobacter spp.) | Piperacillin/tazobactam (3 × 4.5 g, i.v.) or Meropenem (3 × 1 g, i.v.) or Ceftazidim (3 × 2 g, i.v.) +Gentamicin (1 × 240 mg, i.v.) | 2-3 wk | Ciprofloxacin (2 × 750 mg, p.o.)                                            |
| Ciprofloxacin-resistant†             | Depending on susceptibility: meropenem (3 × 1 g), colistin (3 × 3 million U) and/or fosfomycin (3 × 5 g, i.v.) |           | Oral suppression                                                            |
| Anaerobes                            |                                                                  |           |                                                                             |
| Gram-positive anaerobes              | Penicillin G (4 × 5 million U, i.v.)* or                          | 2 wk      | Levofloxacin (2 × 500 mg, p.o.) or                                         |
| (Propionibacterium, Peptostreptococcus, Finegoldia magna) | Ceftriaxon (1 × 2 g, i.v.)                                      |           | Amoxicillin (3 × 1,000 mg, p.o.)                                           |
|                                      | +Rifampin (2 × 450 mg, p.o.)                                      |           | +Rifampin (2 × 450 mg, p.o.)                                                |
| Gram-negative anaerobes              | Clindamycin (3 × 600 mg, i.v.)                                    | 2 wk      | Metronidazol (3 × 500 mg, p.o.)                                             |
3) Two-stage implant replacement

It includes removal of the prosthesis and subsequently delayed reimplantation of a second prosthesis. The approach of short interval (2-4 weeks) is suitable for patients who have known and easily treatable organism, compromised soft tissue or sinus tract. The approach of long interval (8 weeks) is suitable for the organism which is unknown or DTT and strongly compromised soft tissue. Two-stage exchange is identified as a golden standard to treat the patients\(^{15}\), especially in DTT microorganisms such as enterococci or fungi, etc. The success rate of two stage usually >90\(\%\)^4, but the reinfection is important and easy to be ignored question, and the incidence of reinfection in one and two stage according to a meta-analysis shows 8.2\% versus 7.9\% (95% confidence intervals)^{46}. If more than three morbidities and a high ESR or CRP is present before reimplantation, the risk of reinfection is high\(^{16}\).

2. Antimicrobial Therapy

For all surgical procedures, a total duration of antibiotic treatment of 12 weeks is recommended (Fig. 2). Antibiotic treatment without surgery is not recommended and should be only performed, if the patient refuses surgery or the surgical procedure is associated with high risk for patient life. In this case, antibiotic suppression might be considered.

Rifampin is effective to the implant-associated infections causes by staphylococci and Propionibacterium spp., whereas ciprofloxacin has biofilm activity against gram-negative bacteria. In Table 3 the recommended antibiotic therapies targeting different microorganisms are summarized\(^4\).

| Microorganism          | Antibiotic (dose), check pathogen susceptibility | Duration | Followed by |
|------------------------|-------------------------------------------------|----------|-------------|
| Candida spp.           | Fluconazole-susceptible| 2 wk. | Fluconazole (1×400 mg, suppression for ≥1 year; p.o.) |
|                        | Fluconazole-resistant| 2 wk. | Caspofungin (1×100 mg, 1st day: 70 mg; i.v.) |
|                        | Individual (e.g., with voriconazole 2×200 mg, p.o.); removal of the implant or long-term suppression | | |
|                        | Caspofungin (1×100 mg, 1st day: 70 mg; i.v.) | 2 wk. | |

Total duration of therapy: 12 weeks, usually 2 weeks intravenously, followed by oral route. Laboratory testing 2 times/weekly: leukocytes, C-reactive protein, creatinine, estimated glomerular filtration rate, liver enzymes (AST/SGOT and ALT/SGPT).

Dose-adjustment according to renal function and body weight (<40 kg or >100 kg); the dosages needed renal adjustment are in bold.

Rifampin is administered only after the new prosthesis is implanted, wounds are dry and drains are removed; in patients aged >75 years, the rifampin dose should be reduced to 2×300 mg, p.o.

In case of anaphylaxis (such as Quincke’s edema, bronchospasm, anaphylactic shock) or cephalosporin allergy: vancomycin (21 g, i.v.).

* In case of cephalosporin allergy: vancomycin (21 g, i.v.).

Difficult-to-treat.

OUTLOOK

Diagnosis and treatment of PJI are still difficult and have a lack of an universal definition. In order to successfully prevent and treat PJI as well as preserve implant functions in the future, PJI management must contain the effective, timely and individualized diagnosis and treatment with interdisciplinary collaboration. On the other hand, research and development of new diagnostic methods with more accuracy, simplicity, and convenience are required.

CONFLICT OF INTEREST

The authors declare that there is no potential conflict of interest relevant to this article.
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