Abstract: In the present research study, a 2:1 type of smectite clay minerals, namely natural saponite (NSAP) and synthetic saponite (SSAP), was demonstrated for the first time to be controlled drug release host materials for the model drug quinine hydrochloride dihydrate (QU). The popular sol–gel hydrothermal technique was followed for the synthesis of saponite. The QU was ion exchanged and intercalated into an interlayered gallery of synthetic as well as natural saponite matrices. The developed QU-loaded hybrid composite materials along with the pristine materials were characterized by powder X-ray diffraction (PXRD), Fourier transformed infrared spectroscopy (FTIR), thermal gravimetric analysis (TGA), the Brunauer–Emmett–Teller method (BET) for surface area (SA), and scanning electron microscopy (SEM). The characterization of material results using DSC, FTIR and PXRD confirmed the presence of saponite clay mineral phases in the original and the synthesized saponite samples. Similarly, the drug-loaded composites confirmed the successful intercalation of QU drug on the natural and synthesized saponite matrices. The oral drug release performance of both nanocomposites along with pure quinine drug was monitored in sequential buffer environments at 37 ± 0.5 °C. These composite hybrid materials showed the superior controlled release of QU in gastric fluid (pH = 1.2) and intestinal fluid (pH = 7.4). QU release was best fitted in the Korsmeyer–Peppas kinetic model and demonstrated a diffusion-controlled release from nanocomposite layered materials. The observed controlled drug release results suggest that the applied natural/synthetic saponite matrices have the potential to provide critical design parameters for the development of bioengineered materials for controlled drug release.

Keywords: quinine hydrochloride dihydrate; controlled drug release; nanocomposite; saponite; oral drug delivery

1. Introduction

The term drug delivery is a pervasive one in pharmaceutical fields, referring to delivering the therapeutic agents in the patient body. Drug delivery systems are different types of routes manipulated for therapeutic effects in our body, such as oral, topical, transmucosal, parenteral, inhalation, etc. However, the oral drug delivery system is one of the most preferred routes, and a convenient option among all type of drugs with a broad range of formulation choices such as tablets, capsules, etc. Usually, conventional drug delivery systems produce unacceptable toxicity, reduced efficiency of drug concentration (more concentration or less concentration), and the therapeutic effect can have some side
effects on adsorption sites. In the oral controlled drug delivery system, the amount of drug release would be the same as the standard dosage and at uniform time intervals. By controlling the drug delivery manner, there are more advantages at the action site, such as delivering the drug to the target site, maintenance of drug levels within the desired range, reduction of side effects, and improved patient compliance. Nowadays, better bioavailability, biodegradation, and non-toxic compounds such as polymers, supramolecules, and dendrimers have been employed to improve carriers of drug delivery materials [1,2].

Natural and synthetic clay minerals are important potential materials for material scientists due to their distinctive structural and textural properties and features [3,4]. The clay materials and their composites have been reported over the years in numerous fields of research for several different technological applications [5–11]. In the biomedical field, the development of nanocomposites was previously extensively used for a wide array of biomedical applications with several bioengineered materials, and controlled drug release approaches in the pharmaceutical industry are considered one of the fastest growing research areas of the past few years [12–14]. In controlled drug release system applications, clay–drug nanocomposites are a novel class of composite materials in which clay with excellent nanometer-sized composites is applied as a solid matrix for ion exchanged intercalation of various water-soluble as well as water-insoluble drug molecules [15–17]. The nanocomposite should be designed to effectively load the various drug molecules as well as deliver them at the predetermined rate for a predesigned period at a physiological location in the human body. For prevention of the side effects of the toxic drugs, the drug can be delivered to the body in small dosages as well as in controlled release patterns to minimize the side effects and increase patient compliance [18,19].

Recently, our group published several research reports on the different clay-based nanomaterial for drug release applications, where we used different classes and types of drugs [20–26]. From our past experience, we observed that naturally available montmorillonite [27], hectorite [28], etc., are commonly used as pharmaceutical host materials as the carrier of active ingredients for controlled release of hydrophilic drugs. The abundant natural availability and low cost are major advantages in the application of natural clay minerals. However, the original rigid textural properties along with the presence of impurity phases are the strong limitations in the application of natural clay minerals [29]. Recently we proved that synthetic hectorite clay minerals could be synthesized with high purity phases as well as that the textural properties could be tuned according to the requirements of the application area [21,26,30–32]. Saponite clay belongs to the naturally occurring 2:1 triocahedral layered phyllosilicate family and is comprised of layers of Si(IV) tetrahedra and Al(III) or Mg(II) octahedra with definite interlayer spacing. The gallery cations can be readily replaced by a variety of functional cations for potential applications in catalysis and adsorption [33]. Considering scarce natural sources, as well the chemical composition sometimes being extremely variable due to the presence of impurities, synthetic analogs of saponite have attracted attention in recent years because of their high purity, adjustable compositions, and superior textural properties [34,35]. We believe that a saponite matrix has unique ion exchange properties that can be used for controlled release of drug molecules by strong electrostatic interactions or hydrogen bonding or dipole interactions with the interlayer gallery of minerals. As per our knowledge, there are still no reports available on the quinine drug release applications of natural and synthetic saponite clay as a host material. Here in this investigation, we are the first to report the interaction of the antimalarial drug quinine hydrochloride dihydride with natural and synthetic saponite clay.

QU is an antiprotozoal and an antimyotonic drug that is known for the treatment of malaria caused by *Plasmodium* species. Controlled release of QU provides a reduction of side effects and increases patient compliance compared to the immediate release of the dosage [26]. The primary objective of this present study was to synthesize saponite smectite clay mineral, its characterizations, and its incorporation of QU molecule. Secondly, the natural saponite–QU nanocomposite was also used to compare with synthetic saponite–QU nanocomposite for controlled release application. The release study of the QU–nanocomposite in physiological conditions demonstrated its controlled release in
intestinal pH. The overall scheme of QU–nanocomposite synthesis and QU control release is depicted in Figure 1.

Figure 1. Synthesis and quinine (QU) loaded natural saponite (QNSAP) and quinine (QU) loaded synthetic saponite (QSSAP) for controlled release of QU; (A) QU molecular structure, (B) QU intercalation by ion exchange in the interlayer gallery of clay by direct insertion or exchange reactions and (C) PXRD characterizations of QU from QNSAP and QSSAP nanocomposites and controlled release patterns.

2. Material and Methods

2.1. Materials

Natural saponite (NSAP) was obtained from the source of clay repository, Clay Minerals Society and used without further modification. Aluminum isopropoxide was purchased from Sigma Aldrich (St. Louis, MO, USA). Quinine hydrochloride dihydrate (QU, C\textsubscript{20}H\textsubscript{24}N\textsubscript{2}O\textsubscript{2}HCl 2H\textsubscript{2}O) obtained from Sigma Aldrich, USA. Sodium Silicate (Na\textsubscript{2}SiO\textsubscript{2}) Mg (CH\textsubscript{3}COO)\textsubscript{2}, NaOH, KH\textsubscript{2}PO\textsubscript{4} and HCl were obtained from S.D. fine chemicals (Mumbai, India) and used as received. The cation exchange capacity of the natural Saponite and synthetic Saponite is 76 and 59.7 mequiv/100 g clay, respectively [36,37].

2.2. Synthesis of Saponite

Synthetic Saponite clay was prepared by the sol–gel hydrothermal crystallization technique [37]. The gel was formed by mixing [Al(OCH(CH\textsubscript{3})\textsubscript{2})\textsubscript{3}], NaOH, Mg (CHCOO)\textsubscript{2} (with a gel composition of 1SiO\textsubscript{2}, 0.834 Mg (CH\textsubscript{3}COO\textsubscript{2}), 0.113 NaOH, 18.32 H\textsubscript{2}O) in water. The saponite clay with an octahedral sheet consisting of two metal ions incorporated clay was prepared by diluted 40 g of Na\textsubscript{2}SiO\textsubscript{2} by the
addition of 100 mL demineralized water; the 11.9 g of Al[OCH(CH\(_3\)]\(_2\)] solution was dissolved in 80 mL 2M NaOH solution. The aluminum-containing solution was gradually added to Si containing solution under continuous stirring. The gel was formed by allowing it to stand ideal for 1 h. simultaneously, 36 g of Mg(CH\(_3\)COO\(_2\)] dissolved 200 mL of de-ionized water was slowly added into the above Si/Al solution under stirring at 90 °C for 2 h. The gel so formed was aged for 4 h and transferred to the autoclave for hydrothermal treatment at 200 °C for 72 h. The obtained product was filtered and washed with deionized water to remove the unreacted ions. The synthesized saponite is designated as SSAP.

2.3. Synthesis of Clay Drug Nanocomposite

The drug-loaded nanocomposite was prepared by adsorption method. One gram of synthetic saponite (SSAP) or and natural saponite (NSAP) was added to the aqueous QU solution (1 g for QU in 200 mL of water) with continuous stirring (500 rpm) for 24 h at 35 °C. The composite was separated by centrifugation to obtain the hybrid solid residue, dried at 80 °C for 12 h and ground to obtain a fine powder. The obtained QU-loaded nanocomposite was designated as QSSAP and QNSAP for synthetic and natural clay respectively. The drug encapsulation efficiency of clay minerals was calculated spectrophotometrically at \(\lambda_{\text{max}} = 331\) nm using superannuated solution. The amount of QU adsorbed on clay was calculated by the difference in initial concentration and the equilibrium concentration of drug in solution. The QU incorporated clay minerals become hydrophobic in nature.

2.4. Characterization

Powder X-ray diffraction (PXRD) analysis was carried out on Rigaku, Miniflex II Desktop diffractometer (Tokyo, Japan) with a scanning rate 3°/min in the 20 range 2–70°. Fourier transform infrared spectra (FT-IR) were recorded using KBr pellets on Perkin-Elmer GX- FTIR spectrometer. Thermogravimetric analysis was carried out in the temperature range of 50–800 °C at the heating rate of 10 °C/min under a nitrogen atmosphere using Mettler- Toledo TGA/SDTA 851e. The surface area of all samples were determined by N\(_2\) sorption at 77 K (ASAP 2020, Micrometrics Inc, Norcross, GA, USA) after activating the sample at 423 K under high vacuum. UV-vis absorbance of QU solutions was measured at \(\lambda_{\text{max}} = 331\) nm using UV-visible spectrophotometer (UV-2550, Shimadzu, Japan) equipped with a quartz cell of path length = 1 cm. Separation of all the suspended samples was achieved by Kubota 6500 (Tokyo, Japan) centrifugation instrumentation. Drug release experiments were performed at 35 °C using a Julabo shaking water bath (SW23). FE-SEM analysis was carried out by a JEOL JSM 7100 F instrument.

2.5. In Vitro Drug Release Test

In vitro drug release study of native QU and drug-loaded QNSAP and QSSAP nanocomposite was successfully carried out using the dialysis bag technique in the physiological environment by the pH gradient method. The buffer solution of \(\text{pH} = 1.2\) (0.1 M HCl) and phosphate buffer solution \(\text{pH} = 7.4\) (4.4 g of Na\(_2\)HPO\(_4\), 0.38 g of KH\(_2\)PO\(_4\), and 16 g of NaCl) was used for the QU release study. The dialysis bags were equilibrated for a few hours with the release medium prepared in 0.1 M EDTA solution. The weighed quantities of nanocomposite (containing approximately 50 mg of QU) were placed in a dialysis bag containing 5 mL of the release medium. The drug release test was carried out using dialysis bag for nanocomposites (QU, QNSAP, and QSSAP) in a shaker water bath at preset temperature (37 ± 5 °C) with 150 mL of dissolution buffer media. The first two hours of the release was carried out in gastric (0.1 M HCl, \(\text{pH} = 1.2\)) environment and the second phase of the release study was completed in the intestinal pH. After every 30-min time interval, 2 mL of the dissolution medium was collected, which was compensated with the same volume of fresh buffer solution to maintain a constant volume. After 2 h, the release media was changed to phosphate buffer solution (\(\text{pH} = 7.4, 150\) mL) and the release study was continued for the next 8 h. Similarly, at 1-h time intervals, 2 mL of the solution was collected and refilled by the same volume of fresh buffer solution of pH 7.4. All the samples were analyzed for QU content by UV–visible spectrophotometer at \(\lambda_{\text{max}} = 331\) nm. These studies were
performed in triplicate, and the average values were used in data analysis. To understand the release mechanism of QU from nanocomposites materials, in vitro release data was plotted and fitted in first order, Higuchi, Korsmeyer–Peppas, and parabolic diffusion kinetic models [23].

3. Results and Discussion

3.1. Powder X-Ray Diffraction Analysis (PXRD)

The PXRD patterns of the QU-loaded natural/synthetic nanocomposites are depicted in Figure 2A–D, and Table 1. Generally, 2:1 smectite class of saponite clay minerals show the three characteristic reflection phases in the PXRD patterns at 2θ = 6.8–7.5° (d-spacing 1.25–1.51 nm), 19.4° (d spacing 0.47 nm) and 60.5° (d spacing 0.153 nm) due to (001), (110), and (060) planes typical for the saponite clay. The observed PXRD patterns of the natural/synthetic saponite before drug loading clearly showed three distinct peaks reflection confirming the 2:1 type of saponite clay phases in the original and synthetic saponite samples [34,35]. According to the previous report, first order 001 reflection, basal spacing around 1.515 nm, represent the presence of Ca-saponite, whereas at ~1.25 nm represent the Na-saponite [38]. The natural saponite showed the 001 peaks with a maximum at 2θ = 7.16° with d spacing 1.27 nm, close to the reported Na-saponite. However, for the drug-loaded nanocomposite, PXRD patterns showed the expansion of 001 nanolayer sheets by shifting the 2θ values to 4.77°, indicating the expansion of interlayer distance to 1.8 nm. Additionally, in QNSAP an increase of the intensity and a narrowing of the basal reflection was observed due to a more ordered organization of the layered sheets, occurring in the presence of organic quinine molecules inside the interlayer space [35]. Similarly, synthetic saponite demonstrated a characteristic peak corresponding to 001 planes at 2θ = 6.93°, with an interlayer spacing of 1.3 nm, close to the natural Na-based saponite. After drug loading, QSSAP sample showed complete exfoliation due to high drug loading.

![Figure 2. X-ray diffraction patterns of (A) NSAP and QNSAP with (B) lower angle peaks signature, (C) SSAP and QSSAP and (D) lower angle scanning plot.](image-url)
Table 1. The PXRD patterns 2θ valued along with (001) d-spacing of original clay and developed nanocomposites.

| Clay and Nano Composites | 2θ  | 001 Basal Spacing (nm) |
|--------------------------|-----|------------------------|
| NSAP                     | 7.06| 1.27                   |
| QNSAP                    | 4.89| 1.8                    |
| SSAP                     | 6.93| 1.3                    |
| QSSAP                    |     | Exfoliation            |

3.2. Fourier Transform Infrared Spectroscopy (FTIR)

Figure 3A,B depicts the FTIR spectrum of QU drug, NSAP, QU-loaded QNSAP nanocomposite and QU, SSAP, and QU-loaded QSSAP nanocomposite respectively. The natural saponite and synthetic saponite depict the absorption bands at 3689/3685 cm⁻¹ and 1640 cm⁻¹ due to –OH bands of Mg (OH) and octahedral smectite water molecules adsorbed onto the clay surface respectively [8]. The QU-loaded QNSAP, and QSSAP nanocomposite showed the absorption bands at 3679 and 3675 cm⁻¹. The absorption band at 3400 cm⁻¹ is due to the -OH stretching of both the clay and drug-loaded clay nanocomposite. For QU-loaded natural saponite and synthetic saponite aliphatic (C-H) band vibration are shown at 2340 cm⁻¹ and 2949 cm⁻¹. The –NH₂ stretching band of QU-loaded QNSAP and QSSAP are shown at 1639 cm⁻¹ and 1638 cm⁻¹ respectively. The QNSAP and QSSAP vibration stretching band are observed at 1513 cm⁻¹. The FTIR absorption spectra of the clay and drug-loaded clay are shown in Table 2. The functional group identification by the FTIR clearly demonstrates the intercalation of QU in the clay interlayer space. In the previous report, we already confirmed the intercalation of quinine organic moiety on synthetic hectorite clay matrix by solid-state \(^{13}\)C MAS-NMR studies [26].

Figure 3. FTIR spectra of (A) QU, NSAP, and QNSAP, and (B) QU, SSAP and QSSAP. Thermal Gravimetric Analysis (TGA) of (C) NSAP, QNSAP, (D) SSAP and QSSAP.
3.3. Thermal Gravimetric Analysis (TGA)

The thermal gravimetric analysis (TGA) of clay and drug-loaded clay nanocomposite are shown in Figure 2C,D. The first mass loss below 200 °C in NSAP, SSAP, and nanocomposite QNSAP and QSSAP corresponds to the loss of physisorbed water. The second weight loss of endothermic peak, observed only in the nanocomposite QNSAP and QSSAP at 200–400 °C, corresponds to the decomposition of quinine moiety [21].

3.4. Differential Scanning Calorimetry Analysis (DSC)

The DSC analysis of NSAP, SSAP, and nanocomposite QNSAP and QSSAP is shown in Figure 4A. In NSAP and SSAP, and QNSAP and QSSAP nanocomposite, the first dissociation peak was observed in the endothermic range at 50–180 °C, due to dissociation of the physisorbed water moiety. The second endothermic peak observed in QNSAP and QSSAP in the range of 180–250 °C in the DSC curve was due to the decomposition of the organic moiety.

3.5. BET Surface Area analysis

The N₂ sorption studies were carried out via analysis by Brunauer–Emmett–Teller (BET) method, to analyze the surface area, pore volume, and pore diameter of NSAP, SSAP, QNSAP, and QSSAP. N₂ adsorption–desorption data of all the materials are shown in Figure 3B,C and Table 3. The observed results demonstrated that N₂ isotherm of natural and synthetic saponite represented type IV isotherm

| Sample Type | Assignments | Absorption Bands of NSAP and SSAP (cm⁻¹) | QU (cm⁻¹) | QNSAP and QSSAP (cm⁻¹) |
|-------------|-------------|------------------------------------------|-----------|------------------------|
| Clay        | Mg(OH)₂ stretching -OH (for H₂O) | 3686, 3685 3433, 1640 | - | 3678, 3675 3432, 1640 |
|             | Si-O        | 1008, 1029 | - | 1023, 1014 |
|             | Si-O-Al     | 680, 669 | - | 667 |
|             | Si-O-Mg     | 528, 467 | - | 482, 456 |
| QU          | V(C-H) aliphatic | - | 2947 | 2949 |
|             | V(Qu-NH₂ group): | - | 1618 | 1611, 1629 |
|             | V(C=O), alkene | - | 1505 | 1511, 1513 |
|             | V(Q ring)   | - | 1468 | 1451, 1467 |
|             | δ (CH₂-methoxy) | - | 1235 | 1260, 1241 |

### Table 2. FTIR patterns wavenumber assignment for pristine clay, drug-loaded nanocomposite, and quinine drug.
nature, according to the International Union of Pure and Applied Chemistry (IUPAC) classification [39]. Additionally, these samples showed $H_3$ hysteresis loops. Such hysteresis is usually found in solids consisting of aggregates or agglomerates of particles forming slit-shaped pores, with the non-uniform size or shape, confirming layered clay material. The surface area of the synthetic saponite was 881.66 m$^2$/g, close enough to the previously published literature [40]. Further after drug incorporation, the surface area of both NSAP (70 m$^2$/g) and SSAP (881.66 m$^2$/g) decreased to 36.7 and 181 m$^2$/g, respectively. It clearly shows the successful loading of the QU molecule in the clay layer. Similarly, the pore diameter of NSAP (30 nm) and SSAP (15.59 nm) was increased to 33.3 and 18.05 nm, respectively. The increased pore diameter in the drug-loaded samples may be due to the incorporation of organic drug moiety in the clay. These types of pores can be explained as pseudo pores, generally observed in clay samples after organic modification [21]. The surface morphology of NSAP, SSAP with drug incorporated saponite composites QNSAP and QSSAP was studied by SEM (Figure 5A–D). This is additional evidence for quinine drug incorporation into the interlayer gallery of natural saponite and exfoliated structure of synthetic saponite clay hybrid.

### Table 3. Brunauer–Emmett–Teller (BET) surface area analysis results of original and developed composites samples.

| Sample | BET Surface Area, (m$^2$/g) | Pore Volume (cm$^3$/g) | Pore Size (nm) |
|--------|-----------------------------|------------------------|----------------|
| NSAP   | 70.0                        | 0.45                   | 30.0           |
| QNSAP  | 36.6                        | 0.21                   | 33.3           |
| SSAP   | 881.0                       | 3.44                   | 15.6           |
| QSSAP  | 181.0                       | 0.82                   | 18.05          |

### Figure 5. FE-SEM images of (A) NSAP, (B) QNSAP, (C) SSAP, and (D) QSSAP.
3.6. In Vitro Drug Release Study of Clay and Nanocomposite

In vitro drug release studies were carried out from the QU, QNSAP and QSSAP natural and synthetic smectite clay–drug nanocomposite at intestinal (pH = 7.4) gastric fluid (pH = 1.2) buffer environment. Total drug loading capacity of nanocomposite is given in Table 4. The controlled drug release of QU and QU-loaded nanocomposite is depicted in Figure 6. At pH = 1.2, 82% QU (without clay composite), 61% quinine from QNSAP nanocomposite, and 40% quinine from QSSAP clay composite were released in two hours. After replacing the release media with a buffer solution of pH = 7.4 (phosphate buffer), 88% QU (without clay composite), 70% QU was release from QNSAP clay drug nanocomposite and 55% from QSSAP over 10 h. Drug release from QSSAP is lower as compared to QNSSAP nanocomposite in both buffer medium. The high surface area, cation exchange capacity, pore volume, higher adsorption capacity and increase in pore size after drug loading in QSSAP may be the reason for controlled release of QU compared to natural saponite and native QU. The release of drug was in the following order QSSAP > QNSAP > QU.

![In vitro drug release study](image)

**Figure 6.** In vitro release of QU from QNSAP and QSSAP nanocomposites and comparison with QU control free drug.

| Composite | Drug Loading (mg/g) |
|-----------|---------------------|
| QNSAP     | 211                 |
| QSSAP     | 242                 |

**Table 4.** Quinine drug encapsulation efficiency: results of drug-loaded composite.

To the assessment of possible QU release mechanism from QNSAP and QSSAP nanocomposite, the release data were fit into an appropriate mathematical model (Figure 7 and Table 5). The release pattern correlated best to the Korsmeyer–Peppas model. According to Korsmeyer–Peppas, the value of n characterizes the release mechanism of the drug as described in Table 1 and the calculated value of n for QU release followed the diffusion controlled mechanism (fickian diffusion) pattern. [41].
Figure 7. The fitting of QU control release data to first order, Higuchi, Korsmeyer–Peppas and parabolic diffusion kinetic models.

Table 5. First order, Higuchi, Korsmeyer–Peppas and parabolic diffusion kinetic models fitting parameters were derived from Figure 6. Here linear correlation coefficient ($r^2$) and rate constant (K) of the diffusion kinetic models applied to QU release from QNSAP and QSSAP (data considered from first 6 h of release experiments).

| Kinetic Models          | Parameters | Formulations |
|------------------------|------------|--------------|
|                        | QU         | QNSAP        | QSSAP        |
| First order            | $r^2$      | 0.6642       | 0.8062       | 0.8261       |
|                        | K          | -0.16496     | -0.11478     | -0.08894     |
| Higuchi-equation       | $r^2$      | 0.6348       | 0.88         | 0.8912       |
|                        | $K_H$      | 15.23        | 21.41        | 19.88        |
| Korsmeyer–Peppas       | $r^2$      | 0.8842       | 0.8772       | 0.925        |
|                        | N          | 0.1483       | 0.3555       | 0.3399       |
|                        | $K_{HP}$   | 7.83         | 3.151        | 2.51         |
| Parabolic-diffusion    | $r^2$      | 0.611        | 0.8318       | 0.8661       |
|                        | $K_P$      | 0.15         | 0.2126       | 0.2017       |

4. Conclusions

Synthetic saponite was successfully synthesized using the sol–gel hydrothermal method. For the first time, the synthetic saponite was used as a host material for the drug release study with quinine as a guest molecule. We also compared the natural saponite clay with QU nanocomposite and pure...
QU for controlled release studies. The results confirmed that the synthetic saponite clay provided better-controlled release patterns compared to natural saponite clay and pure QU. The present study also concludes that the synthetic/natural Saponite clay minerals are better materials to serve facile and efficient drug release vehicles.

Author Contributions: H.C.B. was responsible for overall supervision of this work and experiments design, discussion, manuscript editing as well as overall guidance. K.S. and R.R.P. carried out the experiments and characterization and writing the manuscript under H.C.B., and B.D.K. helped to design release experiments and helped in writing the manuscript.

Funding: The authors acknowledge the support provided by CSIR, New Delhi, India, for funding through Network Project on CSC-0135. H. C. Bajaj acknowledges CSIR New Delhi for Emeritus scientist fellowship. Authors are also thankful to the Analytical Science Division and Centralized Instrumentation Facility of the Institute for the kind support.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Reinholz, J.; Landfester, K.; Mailänder, V. The challenges of oral drug delivery via nanocarriers. Drug Deliv. 2018, 25, 1694–1705. [CrossRef] [PubMed]
2. Webber, M.J.; Langer, R. Drug delivery by supramolecular design. Chem. Soc. Rev. 2017, 46, 6600–6620. [CrossRef] [PubMed]
3. Bergaya, F.; Lagaly, G. Chapter 1 General Introduction: Clays, Clay Minerals, and Clay Science. In Developments in Clay Science; Bergaya, F., Theng, B.K.G., Lagaly, G., Eds.; Elsevier: Amsterdam, The Netherlands, 2006; Volume 1, pp. 1–18.
4. Carrado, K.A.; Decarreau, A.; Petit, S.; Bergaya, F.; Lagaly, G. Chapter 4 Synthetic Clay Minerals and Purification of Natural Clays. In Developments in Clay Science; Bergaya, F., Theng, B.K.G., Lagaly, G., Eds.; Elsevier: Amsterdam, The Netherlands, 2006; Volume 1, pp. 115–139.
5. Murray, H.H. Overview — clay mineral applications. Appl. Clay Sci. 1991, 5, 379–395. [CrossRef]
6. Ingole, P.G.; Pawar, R.R.; Baig, M.I.; Jeon, J.D.; Lee, H.K. Thin film nanocomposite (TFN) hollow fiber membranes incorporated with functionalized acid-activated bentonite (ABn-NH) clay: towards enhancement of water vapor permeance and selectivity. J. Mater. Chem. A 2017, 5, 20947–20958. [CrossRef]
7. Chuaicham, C.; Pawar, R.; Sasaki, K. Dye-sensitized Photocatalyst of Sepiolite for Organic Dye Degradation. Catalysts 2019, 9, 235. [CrossRef]
8. Pawar, R.R.; Lalmumiasa; Bajaj, H.C.; Lee, S.-M. Activated bentonite as a low-cost adsorbent for the removal of Cu(II) and Pb(II) from aqueous solutions: Batch and column studies. J. Ind. Eng. Chem. 2016, 34, 213–223. [CrossRef]
9. Pawar, R.R.; Gupta, P.; Lalmumiasa; Bajaj, H.C.; Lee, S.-M. Al-intercalated acid activated bentonite beads for the removal of aqueous phosphate. Sci. Total Environ. 2016, 572, 1222–1230. [CrossRef] [PubMed]
10. Lee, C.; Jung, J.; Pawar, R.R.; Kim, M.; Lalmumiasa; Lee, S.-M. Arsenate and phosphate removal from water using Fe-sericite composite beads in batch and fixed-bed systems. J. Ind. Eng. Chem. 2017, 47, 375–383. [CrossRef]
11. Pawar, R.R.; Lalmumiasa; Kim, M.; Kim, J.-G.; Hong, S.-M.; Sawant, S.Y.; Lee, S.M. Efficient removal of hazardous lead, cadmium, and arsenic from aqueous environment by iron oxide modified clay-activated carbon composite beads. Appl. Clay Sci. 2018, 162, 339–350. [CrossRef]
12. Yang, P.; Gai, S.; Lin, J. Functionalized mesoporous silica materials for controlled drug delivery. Chem. Soc. Rev. 2012, 41, 3679–3698. [CrossRef]
13. Tibbitt, M.W.; Rodell, C.B.; Burdick, J.A.; Anseth, K.S. Progress in material design for biomedical applications. Proc. Natl. Acad. Sci. USA 2015, 112, 14444–14451. [CrossRef] [PubMed]
14. Bhat, S.; Kumar, A. Biomaterials and bioengineering tomorrow’s healthcare. Biomatter 2013, 3, e24717. [CrossRef] [PubMed]
15. Hun Kim, M.; Choi, G.; Elzataryah, A.; Vinu, A.; Bin Choy, Y.; Choy, J.-H. Review of Clay-drug Hybrid Materials for Biomedical Applications: Administration Routes. Clays Clay Miner. 2016, 64, 115–130. [CrossRef]
16. Khurana, I.S.; Kaur, S.; Kaur, H.; Khurana, R.K. Multifaceted role of clay minerals in pharmaceuticals. Future Sci. OA 2015, 1, FSO6. [CrossRef] [PubMed]
17. Massaro, M.; Colletti, G.C.; Lazzara, G.; Riela, S. The Use of Some Clay Minerals as Natural Resources for Drug Carrier Applications. J. Funct. Biomater. 2018, 9. [CrossRef]
18. Tiwari, G.; Tiwari, R.; Srivastawa, B.; Bhati, L.; Pandey, S.; Pandey, P.; Bannerjee, S.K. Drug delivery systems: An updated review. Int. J. Pharm. Investig. 2012, 2, 2–11. [CrossRef]
19. Senapati, S.; Mahanta, A.K.; Kumar, S.; Maiti, P. Controlled drug delivery vehicles for cancer treatment and their performance. Signal Trans. Target. Ther. 2018, 3, 7. [CrossRef]

20. Joshi, G.V.; Kevadiya, B.D.; Patel, H.A.; Bajaj, H.C.; Jasra, R.V. Montmorillonite as a drug delivery system: Intercalation and in vitro release of timolol maleate. Int. J. Pharm. 2009, 374, 53–57. [CrossRef]
21. Joshi, G.V.; Pawar, R.R.; Kevadiya, B.D.; Bajaj, H.C. Mesoporous synthetic hectorites: A versatile layered host with drug delivery application. Microporous Mesoporous Mater 2011, 142, 542–548. [CrossRef]
22. Kevadiya, B.D.; Chettiar, S.S.; Rajkumar, S.; Bajaj, H.C.; Gosai, K.A.; Brahmbhatt, H. Evaluation of clay/poly (l-lactide) microcomposites as anticancer drug, 6-mercaptopurine reservoir through in vitro cytotoxicity, oxidative stress markers and in vivo pharmacokinetics. Colloids Surf. B Biointerfaces 2013, 112, 400–407. [CrossRef]
23. Kevadiya, B.D.; Joshi, G.V.; Bajaj, H.C. Layered bionanocomposites as carrier for procainamide. Int. J. Pharm. 2010, 388, 280–286. [CrossRef] [PubMed]
24. Kevadiya, B.D.; Rajkumar, S.; Bajaj, H.C.; Chettiar, S.S.; Gosai, K.; Brahmbhatt, H.; Bhatt, A.S.; Barvaliya, Y.K.; Dave, G.S.; Kothari, R.K. Biodegradable gelatin–ciprofloxacin–montmorillonite composite hydrogels for controlled drug release and wound dressing application. Colloids Surf. B Biointerfaces 2014, 122, 175–183. [CrossRef] [PubMed]
25. Patel, H.A.; Somani, R.S.; Bajaj, H.C. Preparation and characterization of phosphonium montmorillonite with enhanced thermal stability. Appl. Clay Sci. 2007, 35, 194–200. [CrossRef]
26. Pawar, R.R.; Kevadiya, B.D.; Brahmbhatt, H.; Bajaj, H.C. Template free synthesis of mesoporous hectorites: Efficient host for pH responsive drug delivery. Int. J. Pharm. 2013, 446, 145–152. [CrossRef] [PubMed]
27. Kevadiya, B.D.; Joshi, G.V.; Patel, H.A.; Ingle, P.G.; Mody, H.M.; Bajaj, H.C. Montmorillonite-Alginate Nanocomposites as a Drug Delivery System: Intercalation and In Vitro Release of Vitamin B1 and Vitamin B6. J. Biomater. Appl. 2010, 25, 161–177. [CrossRef]
28. Kumaresan, S.; Pawar, R.R.; Lee, S.-M.; Bajaj, H. Quinine Intercalated Natural Hectorite Clay Layered Hybrid Nanocomposite for Controlled Drug Delivery Application. Sci. Technol. J. 2016, 1, 7–13.
29. Pawar, R.R.; Lakhmunsiaima; Gupta, P.; Sawant, S.Y.; Shahmoradi, B.; Lee, S.-M. Porous synthetic clay-alginate composite beads for effective adsorption of methylene blue dye from aqueous solution. Int. J. Biol. Macromol. 2018, 114, 1315–1324. [CrossRef]
30. Sawant, S.Y.; Pawar, R.R.; Somani, R.S.; Bajaj, H.C. Facile hard template approach for synthetic hectorite hollow microspheres. Mater. Lett. 2014, 128, 121–124. [CrossRef]
31. Sethia, G.; Patel, H.A.; Pawar, R.R.; Bajaj, H.C. Porous synthetic hectorites for selective adsorption of carbon dioxide over nitrogen, methane, carbon monoxide and oxygen. Appl. Clay Sci. 2014, 91–92, 63–69. [CrossRef]
32. Pawar, R.R.; Patel, H.A.; Sethia, G.; Bajaj, H.C. Selective adsorption of carbon dioxide over nitrogen on calcined synthetic hectorites with tailor-made porosity. Appl. Clay Sci. 2009, 46, 109–113. [CrossRef]
33. Vicente, M.A.; Suárez, M.; López-González, J.D.D.; Bañares-Muñoz, M.A. Characterization, Surface Area, and Porosity Analyses of the Solids Obtained by Acid Leaching of a Saponite. Langmuir 1996, 12, 566–572. [CrossRef]
34. Szabó, T.; Mitea, R.; Leeman, H.; Premchandra, G.S.; Johnston, C.T.; Szekeres, M.R.; Dékány, L.; Schoonheydt, R.A. Adsorption of Protamine and Papain Proteins on Saponite. Clays Clay Miner. 2008, 56, 494–504. [CrossRef]
35. Bisio, C.; Carniato, F.; Paul, G.; Gatti, G.; Boccaleri, E.; Marchese, L. One-Pot Synthesis and Physicochemical Properties of an Organoo-Modified Saponite Clay. Langmuir 2011, 27, 7250–7257. [CrossRef] [PubMed]
36. Okada, T.; Seki, Y.; Ogawa, M. Designed Nanostructures of Clay for Controlled Adsorption of Organic Compounds. J. Nanosci. Nanotechnol. 2014, 14, 2121–2134. [CrossRef] [PubMed]
37. Vogels, R.J.M.J.; Kloprogge, J.T.; Geus, J.W. Synthesis and characterization of saponite clays. Am. Mineralogist 2005, 90, 931–944. [CrossRef]
38. Post, J.L. Saponite from Near Ballarat, California. Clays Clay Miner. 1984, 32, 147–153. [CrossRef]
39. Sing, K.S.W.; Everett, D.H.; Haul, R.A.W.; Moscou, L.; Pierotti, R.A.; Rouquérol, J.; Siemieniewska, T. Reporting Physisorption Data for Gas/Solid Systems With Special Reference to the Determination of Surface Area and Porosity. *Pure Appl. Chem.* 1985, 57, 603-619. [CrossRef]

40. Shao, H.; Pinnavaia, T.J. Synthesis and Properties of Nanoparticle Forms Saponite Clay, Cancrinite Zeolite and Phase Mixtures Thereof. *Microporous Mesoporous Mater* 2010, 133, 10–17. [CrossRef]

41. Dash, S.; Murthy, P.N.; Nath, L.; Chowdhury, P. Kinetic modeling on drug release from controlled drug delivery systems. *Acta Poloniae Pharm.* 2010, 67, 217–223.

© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).