Functional Interaction between Human Histocompatibility Leukocyte Antigen (HLA) Class II and Mouse CD4 Molecule in Antigen Recognition by T Cells in HLA-DR and DQ Transgenic Mice

By Ken Yamamoto, Yoshinori Fukui, Yukio Esaki, Takeshi Inamitsu,* Tohm Sudo, Kazuaki Yamane, Nobuhiro Kamikawaji, Akinori Kimura, and Takehiko Sasazuki

From the Department of Genetics, Medical Institute of Bioregulation, Kyushu University, Higashi-ku, Fukuoka 812; and the *Department of Pediatrics, Hamanomachi Hospital, Chuo-ku, Fukuoka 810, Japan

Summary

Studies in vitro have suggested that a species barrier exists in functional interaction between human histocompatibility leukocyte antigen (HLA) class II and mouse CD4 molecules. However, whether mouse CD4+ T cells restricted by HLA class II molecules are generated in HLA class II transgenic mice and respond to peptide antigens across this barrier has remained unclear. In an analysis of T cell responses to synthetic peptides in mice transgenic for HLA-DR51 and -DQ6, we found that DR51 and DQ6 transgenic mice acquired significant T cell response to influenza hemagglutinin-derived peptide 307-319 (HA 307) and Streptococcus pyogenes M12 protein-derived peptide 347-397 (M6C2), respectively. Inhibition studies with several monoclonal antibodies showed that transgenic HLA class II molecules presented these peptides to mouse CD4+ T cells. Furthermore, T cell lines specific for HA 307 or M6C2 obtained from the transgenic mice could respond to the peptide in the context of relevant HLA class II molecules expressed on mouse L cell transfectants that lack the expression of mouse MHC class II. These findings indicate that interaction between HLA class II and mouse CD4 molecules is sufficient for provoking peptide-specific HLA class II-restricted T cell responses in HLA class II transgenic mice.
Materials and Methods

Mice. DRA-B6 and DQ6-B6 were generated and characterized as previously described (12, 13). DR51B-B6 were produced by injection of B6 oocytes with a 5.6-kb ApaI/EcoRI fragment containing the HLA-DR51B (DRB*0102) cDNA (14). This transgene includes a 4.0-kb ApaI/SacI fragment containing the 5' upstream region of DR51B gene and connected to the SV40 poly A signal sequence at the 3' side. Mice carrying the transgene were identified by Southern blot analysis using DR51B cDNA as a probe. DR51-B6 were obtained by crossing DRA-B6 with DR51-B6.

Reverse Transcriptase–PCR Analysis. Total cellular RNA was extracted from homogenized tissues or single cell suspensions. cDNA were made from RNA extracts digested by DNase I to avoid contaminating transgenic DNA and were then subjected to PCR to amplify the DR51B for $\alpha^b$ gene. The primers used were (5'-CACCTGATGTGCTGACCTCCAC-3') and (5'-CCAGGAAAGAGACAGGCACGCAC-3') for DR51B, and (5'-CGGAAGGCATTCGTGTTAGCAGGACG-3') and (5'-ATGCTAGACTCTGAGTAAAGAGCAGGCCCAGCAC-3') for $\alpha^b$. PCR products were electrophoresed in a 1.2% agarose gel, transferred onto a nylon membrane, and hybridized with end-labeled oligonucleotide probes for DR51B (5'-GAGGGCTGTCATTTCTTCAACG-3') and (5'-CGGCGGCTTGAACTCGAAGCAGGCCCAGCACAGC-3') as described (15).

Flow Cytometry. Cell surface expressions of HLA-DR51 molecules on B cells, dendritic cells, and macrophages were examined by flow cytometry (FCM) as follows. Spleen cells from each strain of mice were cultured with IL-4 (50 U/ml) for 48 h and then doubly stained with biotinylated goat anti-mouse IgG plus IgM (Tago, Inc., Burlingame, CA) followed by streptavidin-conjugated PE, and TAL16.1, specific for HLA-DR51 (16), followed by FITC-conjugated goat anti-mouse IgG (Jackson ImmunoResearch, West Grove, PA). IgG plus IgM positive cells were gated to be analyzed as B cells. Splenic dendritic cells were stained as described (17) and stained with TAL16.1. About 70–80% of cells prepared were positive for splenic dendritic cell marker 33D1 (18). Dish adherent spleen cells were prepared and cultured with IFN-γ (250 U/ml) for 48 h, and then doubly stained with M1/70HL, specific for MAC-1 antigen expressed on macrophages (Hybritech, Inc., San Diego, CA) and TAL16.1. MAC-1 positive cells were gated to be analyzed as splenic macrophages. All the cells expressed a comparable level of the I-A molecule. For analysis of the expression of $\alpha\beta\epsilon\beta$ molecules in DRA-B6 and DR51-B6, spleen cells were stained with biotinylated 17-3-3 (50 U/ml) for 48 h, and then doubly stained with M1/70HL, specific for MAC-1 antigen expressed on macrophages (Hybritech, Inc., San Diego, CA) and TAL16.1. MAC-1 positive cells were gated to be analyzed as splenic macrophages. All the cells expressed a comparable level of the I-A$^b$ molecule. For analysis of the expression of $\alpha\epsilon\beta$ molecules in DRA-B6 and DR51-B6, spleen cells were stained with biotinylated 17-3-3, mAb specific for $\beta^b$ (19), followed by streptavidin-conjugated PE. FCM was done by FACScan® (Becton Dickinson & Co., Mountain View, CA).

T Cell Proliferation Assays and Blocking Experiments. Mice were immunized by giving 0.2 ml of soluble HA 307 peptide (20 μg, PKYVKQNTLKLAT) or M6C2 peptide (20 μg, AKKQVEKDAKKA) emulsified with CFA (Difco, Detroit, MI) into the hind footpads and at the base of the tail. These peptides were synthesized by a peptide synthesizer (model 9050 Plus; Millipore Corp., Bedford, MA) and purified by reversed-phase HPLC. Popliteal and inguinal lymph nodes were removed 9–12 d later and single cell suspensions were prepared in RPMI 1640 medium (Gibco BRL, Gaithersburg, MD) supplemented with 10% heat-inactivated FCS, 100 U/ml penicillin, 100 μg/ml streptomycin, 2 mM L-glutamine, 50 μM 2-ME, and 20 mM Hepes (complete medium). To purify lymph node cells to T cells, I-A$^b$ positive cells were eliminated by using sheep anti-mouse IgG coated magnetic beads (Dynal, Oslo, Norway) and anti-I-A$^b$ mAb (Meiji, Odawara, Japan) as follows. Magnetic beads were added to the cell suspension and the preparation was centrifuged for 10 min at 1,500 rpm followed by resuspension of the cells and removal of the beads/IgG positive B cells. This step was repeated twice and cell suspensions were subsequently incubated with anti-I-A$^b$ mAb at a saturating concentration for 30 min at 4°C. The cells were then washed three times with complete medium and further enriched to T cells using magnetic beads. The enriched T cells (4 × 10$^6$) were cultured in 0.2 ml of complete medium in triplicate in 96-well flat-bottom tissue culture plates, containing 8 × 10$^4$ irradiated (3,000 rad) syngeneic spleen cells as APCs in the presence or absence of antigen peptide at 37°C in a humidified atmosphere of 5% CO₂ for 48 h. 1 μCi of [³H]thymidine was added during the final 12 h and then incorporated radioactivity was measured. In blocking experiments on the T cell responses with mAbs, a 200-fold diluted ascites form of anti-Lyt2.2 (anti-CD8; Meiji), 3JP (anti-Aœ), M6C2 (anti-DQ) (21), or a fourfold diluted culture supernatant GKL5 (anti-CD4) (22), or 1.3 μg/ml purified L243 (anti-DR) (23), or 10 μg/ml purified 17-3-3 (anti-β$^b$), 17/227 (anti-A$^b$) (24), 20-8-4S (anti-α$^b$) (25) was added to the culture.

Antigen-specific T Cell Line. T cells were purified from lymph node cells of DR51-B6 or DQ6-B6 preimmunized with 20 μg of peptide by eliminating I-A$^b$ positive cells as described above. Purified T cells (6 × 10$^5$) were cultured with 25 × 10$^5$ irradiated (3,000 rad) syngeneic spleen cells with HA 307 or M6C2 peptide at 10 μg/ml in complete medium for 5 d. Viable cells were collected by density gradient centrifugation with Lympholyte-M (Cedarlane Laboratories, Hornby, ON, Canada) and further cultured without stimulator cells and antigen for 5 d. The stimulation and rest cycle was repeated twice in medium supplemented with rat Con A supernatant (Becton Dickinson & Co.). A T cell line (5 × 10$^5$) was cultured with 3 × 10$^5$ indicated mouse L cell transfectants pretreated with mitomycin C in the presence or absence of peptide for 36 h. 1 μCi of [³H]thymidine was added during the final 6 h and then incorporated radioactivity was measured.

Results and Discussion

We previously reported that HLA-DQ6 molecules were expressed in the thymus, lungs, lymph nodes, and spleen in a tissue-specific manner as was MHC class II in DQ6-B6 (12). In DR51-B6 obtained by crossing DR51B-B6 with DRA-B6, the DR51B transgene was expressed in the thymus, spleen, and lung (Fig. 1 A). This suggested that HLA-DR51 molecules were expressed on various immunocompetent cells such as thymic epithelial cells, B cells, macrophages, and dendritic cells in DR51-B6 because DRA-B6 expressed DRα chain as DRαεβ$^b$ molecule on these cells (13). Cell surface expressions of DR51 molecules on these immunocompetent cells were confirmed by FCM analysis (Fig. 1 B). It should be noted that DR51-B6 also expresses a xenogeneic mixed isotype molecule, DRαεβ$^b$, as in DRA-B6 (Fig. 2). Expression of the DRαεβ$^b$ molecule in DRα single transgenic mice was not demonstrated by immunoprecipitation (26) nor was the AœDR51β molecule in DR51B-B6 demonstrated by FCM using anti-DRβ polyclonal antibody (data not shown). Thus, DR51-B6 apparently expresses three different MHC class II molecules, DR51, DRαεβ$^b$, and I-A$^b$. In DQ6-B6, mixed
isotype molecules were not detected by immunoprecipitation (12).

In a previous study (12), it was shown that DQ6-B6 did not produce antibodies specific for the DQ6 molecule, and that loss of MLR against the DQ6 molecule was observed in DQ6-B6, suggesting that DQ6-B6 acquired a tolerance to HLA-DQ6 molecule at the B and T cell levels. To investigate whether DR51-B6 acquired a tolerance to the HLA-DR51 molecule, mice were immunized with the EBV-transformed human B cell line, TOK, which expresses HLA-DR51 molecules, and the production of antibodies specific for DR51 molecule was examined. As expected, DR51-B6 did not produce antibodies against DR51 molecule, whereas B6, DRA-B6, and DR51B-B6 produced the antibodies (data not shown). In addition, a primary MLR using DR51-B6 spleen cells as stimulator showed that the transgenic DR51 molecule could stimulate T cells from B6, DRA-B6, and DR51B-B6, but not DR51-B6 (data not shown). These findings indicate that DR51-B6 acquired a tolerance to transgene products as had DQ6-B6, suggesting that transgenic HLA class II molecules contributed to eliminate self-reactive T cells as had MHC class II molecules.

Next, in order to examine whether HLA class II molecules expressed in mice would provoke T cell responses to foreign peptide antigens, mice were immunized with influenza A hemagglutinin-derived peptide 307-319 (HA 307), known to be a satisfactory DR binding peptide (27), or with Streptococcus pyogenes M12 protein-derived peptide 347-367 (M6C2),
Figure 3. Immune responses to peptide antigens in HLA transgenic mice. DR51-B6 and DQ6-B6 acquired the T cell response to HA 307 peptide (A) and M6C2 peptide (B), respectively. Anti-CD4 but not anti-CD8 mAb inhibited these T cell responses (C and D). Data presented are representative of three experiments and are expressed as the mean ± SD of triplicate wells.

which is a DQ6-restricted antigenic peptide in humans (28), and secondary proliferative T cell responses in vitro were measured. Fig. 3 A shows that DR51-B6 acquired the T cell response to HA 307, whereas no proliferative response was observed in B6, DRA-B6, or DR51B-B6. On the other hand, the immune response to M6C2 was observed only in DQ6-B6 (Fig. 3 B). DR51-B6 and DQ6-B6 did not acquire the T cell response to M6C2 and HA 307, respectively (data not shown). Anti-CD4 but not anti-CD8 mAb inhibited these responses (Fig. 3, C and D). These results suggest that the DR51 molecule, but not DRαEβ or I-Aβ, presented HA 307 to mouse CD4+ T cells in DR51-B6, whereas the DQ6 molecule presented M6C2 to CD4+ T cells in DQ6-B6. However, it remained to be determined whether functional interaction of mouse CD4 with HLA class II molecules was involved in these responses, because mouse CD4 might interact with the β chain of DRαEβ and/or I-Aβ expressed in addition to DR51 or DQ6 molecules on the same APC (9, 12).

To clarify this point, inhibitory effects by several mAbs specific for mouse MHC class II molecules on the T cell responses were examined. As shown in Fig. 4, mAbs specific for Eβ (17-3-3), which are known to react with DRαEβ (13, 29) or those for Aβ (17/227), failed to block the responses as well as had anti-Aα (3JP). By contrast, the response was completely inhibited by anti-DR (L243) in DR51-B6 and anti-DQ (HU11) in DQ6-B6. These results suggest that the DRαEβ or I-Aβ molecules did not play a major role in these responses.

To examine more closely the interaction of mouse CD4 with HLA class II, CD4+ T cell lines specific for HA 307 or M6C2 were obtained from DR51-B6 or DQ6-B6, respectively, and their restriction elements were investigated using mouse L cells transfected with various HLA class II as APCs that do not express endogenous mouse MHC class II molecules. Consistent with the blocking experiments, each T cell line specific for either HA 307 or M6C2 responded to the peptide antigen in the context of the relevant HLA class II molecules in the absence of mouse class II II (Fig. 5, A and B). These responses were restricted to DR51 or DQ6 molecules because the L cells expressing each of the DR1, DR2, or DR4 molecules, which are known to present HA 307 to
human T cells (27), and those expressing DQ4, which binds M6C2 (data not shown), could not present the antigen peptides to these T cell lines.

Spleen cells from bone marrow chimera-B6 and -DRA-B6, which were reconstituted with DR51-B6 BM, could stimulate the HA 307–specific DR51-restricted T cell line, but these bone marrow chimera mice did not acquire the T cell response to HA 307 (data not shown). These observations indicate that the responding T cells observed in the DR51-B6 express TCR antigens that were selected positively in the thymus by the transgene product, but not by I-A\(^d\) or DR\(\alpha \)EB\(\beta\) molecules. Previous study (2) has shown that interaction of CD4 with MHC class II molecules is critical for the generation of MHC class II–restricted mature T cells by analyzing T cell differentiation in CD4-deficient mice. Taken together, we conclude that HLA class II–restricted mouse CD4\(^+\) T cells were generated by significant interaction between HLA class II and mouse CD4 molecules in the thymus of the transgenic mice, and that these cells could recognize antigen peptides in the context of HLA class II molecules.

Our results would contradict the previous observations showing that a species barrier exists in functional interaction between HLA class II and mouse CD4 molecules (9–11). However, it has been also suggested that this barrier is partial and that CD4 can bind to the MHC class II \(\alpha\) chain or a conserved site that itself is not species specific (10, 30). We cannot assess in this study whether T cell responses in the HLA class II transgenic mice will be augmented by introducing human CD4, or whether a HLA class II transgenic mouse expressing chimeric human/mouse MHC class II molecules that contain mouse class II–CD4 interacting region will acquire stronger T cell response than the transgenic mice used in this study. However, one prominent feature of this study was that we established two strains of HLA class II transgenic mice...
expressing HLA-DR or -DQ molecules in a tissue-specific manner as MHC class II and T cell responses to peptide antigens were analyzed in an in vivo system. Therefore, our observations indicate that interaction between HLA class II and mouse CD4 molecules is physiologically sufficient for generating HLA class II–restricted mouse T cell repertoire and for provoking peptide-specific mouse T cell responses. Thus, we propose that HLA class II transgenic mice can serve to investigate the role of HLA class II molecules in regulating immune responses and disease susceptibility in humans.

Figure 5. Recognition of HA 307 and M6C2 peptide in the context of the relevant HLA class II molecule expressed on the mouse L cell transfectant by T cell lines established from DR51-B6 (A) and DQ6-B6 (B), respectively. Peptides were used at a concentration of 5 μg/ml. 98% of T lymphoblasts were positive for CD4. Data presented are representative of three experiments and are expressed as the mean ± SD of triplicate wells.

We thank M. Kimoto (Saga Medical School, Japan) and J. G. Bodmer (Imperial Cancer Research Fund, London, UK) for generous gifts of mAb, T. Ide for technical support, and M. Ohara for helpful comments.

This work was supported in part by grants from the Ministry of Education, Science and Culture, from the Science and Technology Agency, and from the Ministry of Health and Welfare, Japan.

Address correspondence to Takehiko Sasazuki, Department of Genetics, Medical Institute of Bioregulation, Kyushu University, 3-1-1 Maidashi, Higashi-ku, Fukuoka 812, Japan.

Received for publication 14 February 1994 and in revised form 25 March 1994.

References
1. Kruisbeek, A.M., J.J. Mond, B.J. Fowlkes, J.A. Carmen, S. Bridges, and D.L. Longo. 1985. Absence of the Lyt-2-, L3T4+ lineage of T cells in mice treated neonatally with anti-I-A correlates with absence of intrathymic I-A-bearing antigen-presenting cell function. J. Exp. Med. 161:1029.
2. Rahemtulla, A., W.P. Fung-Leung, M.W. Schilham, T.M. Kündig, S.R. Sambataro, A. Narendran, A. Arabian, A. Wakeham, C.J. Paige, R.M. Zinkernagel, et al. 1991. Normal
development and function of CD8+ cells but markedly decreased helper cell activity in mice lacking CD4. *Nature (Lond.)* 353:180.

3. Janeway, C.A. 1989. The role of CD4 in T-cell activation: accessory molecule or co-receptor? *Immunol. Today.* 10:234.

4. König, R., L.Y. Huang, and R.N. Germain. 1992. MHC class II interaction with CD4 mediated by a region analogous to the MHC class II binding site for CD8. *Nature (Lond.)* 356:796.

5. Cammarota, G., A. Scheirle, B. Takacs, D.M. Doran, R. Knorr, W. Bannwarth, J. Guadilla, and F. Sinigaglia. 1992. Identification of a CD4 binding site on the b2 domain of HLA-DR molecules. *Nature (Lond.)* 356:799.

6. Gay, M., P. Maddon, R. Sekaly, M.A. Talle, M. Godfrey, E. Long, G. Goldstein, L. Chess, R. Axel, J. Kappler, and P. Marrack. 1987. Functional interaction between human T-cell protein CD4 and the major histocompatibility complex HLA-DR antigen. *Nature (Lond.)* 328:626.

7. Rosoff, P.M., S.J. Burakoff, and J.L. Greenstein. 1987. The role of the L3T4 molecule in mitogen and antigen-activated signal transduction. *Cell.* 49:845.

8. Klein, J., and N. Takahata. 1990. The major histocompatibility complex and the quest for origins. *Immunol. Rev.* 113:5.

9. Lamarré, D., A. Ashkenazi, S. Fleury, D.H. Smith, R.-P. Sekaly, and D.J. Capon. 1989. The MHC-binding and gp120-binding functions of CD4 are separable. *Science (Wash. DC).* 245:743.

10. Vignali, D.A., J. Moreno, D. Schiller, and G.J. Hämmerling. 1992. Species-specific binding of CD4 to the b2 domain of major histocompatibility complex class II molecules. *J. Exp. Med.* 175:925.

11. Barzaga-Gilbert, E., D. Grass, S.K. Lawrance, P.A. Peterson, E. Lacy, and V.H. Engelhard. 1992. Species specificity and augmentation of responses to class II major histocompatibility complex molecules in human CD4 transgenic mice. *J. Exp. Med.* 175:1707.

12. Nishimura, Y., T. Iwanaga, T. Imamitsu, Y. Yanagawa, M. Yasunami, A. Kimura, K. Hirokawa, and T. Sasazuki. 1990. Expression of the human MHC, HLA-DQw6 genes alters the immune response in C57BL/6 mice. *J. Immunol.* 145:353.

13. Fukui, Y., Y. Esaki, A. Kimura, K. Hirokawa, Y. Nishimura, and T. Sasazuki. 1993. T-cell repertoire in a strain of transgenic C57BL/6 mice with the HLA-DRA gene on the X-chromosome. *Immunogenetics.* 37:204.

14. Kamikawaji, N., K. Fujisawa, H. Yoshizumi, M. Fukunaga, M. Yasunami, A. Kimura, Y. Nishimura, and T. Sasazuki. 1991. HLA-DR2-restricted CD4+ T cells specific to streptococcal antigen present in low but not in high responders. *J. Immunol.* 146:2560.

15. Senju, S., A. Kimura, M. Yasunami, N. Kamikawaji, H. Yoshizumi, Y. Nishimura, and T. Sasazuki. 1992. Allele-specific expression of the cytoplasmic exon of HLA-DQB1 gene. *Immunogenetics.* 36:319.

16. Sadler, A.M., J.M. Heyes, S.G.E. Marsh, P. Krausa, G.E. Reynolds, and J.G. Bodmer. 1993. The monoclonal antibody TAL16.1 recognizes the aspartic acid residue at position 70 in DRB gene products. *Tissue Antigens.* 41:42.

17. Crowley, M., K. Inaba, M. Witmer-Pack, and R.M. Steinman. 1989. The cell surface of mouse dendritic cells: FACS analysis of dendritic cells from different tissues including thymus. *Cell. Immunol.* 118:108.

18. Nussenzweig, M.C., R.M. Steinman, M.D. Witmer, and B. Gutchinov. 1982. A monoclonal antibody specific for mouse dendritic cells. *Proc. Natl. Acad. Sci. USA.* 79:161.

19. Ozato, K., N. Mayer, and D.H. Sachs. 1980. Hybridoma cell lines secreting monoclonal antibodies to mouse H-2 and Ia antigens. *J. Immunol.* 124:533.

20. Janeway, C.A., P.J. Conrad, E.A. Lerner, J. Babich, P. Wettstein, and D.B. Murphy. 1984. Monoclonal antibodies specific for Ia glycoproteins raised by immunization with activated T cells: possible role of T cell bound Ia antigens as targets of immunoregulatory T cells. *J. Immunol.* 132:662.

21. Kasahara, M., T. Takenouchi, H. Ikeda, K. Ogasawara, T. Okuyama, N. Ishikawa, A. Wakisaka, Y. Kikuchi, and M. Aizawa. 1983. Serologic dissection of HLA-D specificities by the use of monoclonal antibodies. *Immunogenetics.* 18:525.

22. Dialynas, D.P., D.B. Wilde, P. Marrack, A. Pierres, K.A. Wall, W. Havran, G. Otten, M.R. Loken, M. Pierres, J. Kappler, and F.W. Fitch. 1983. Characterization of the murine antigenic determinant, designated L3T4a, recognized by monoclonal antibody GK1.5: expression of L3T4a by functional T cell clones appears to correlate primarily with class II MHC antigen reactivity. *Immunol. Rev.* 74:29.

23. Lampson, L.A., and R. Levy. 1980. Two populations of Ia-like molecules on a human B cell line. *J. Immunol.* 125:293.

24. Lemke, H., G.J. Hämmerling, and U. Hämmerling. 1979. Fine specificity analysis with monoclonal antibodies of antigens controlled by the major histocompatibility complex and by the Qa/TL region in mice. *Immunol. Rev.* 47:175.

25. Ozato, K., and D.H. Sachs. 1981. Monoclonal antibodies to mouse MHC antigens. III. Hybridoma antibodies reacting to antigens of the H-2k haplotype reveal genetic control of isoform expression. *J. Immunol.* 126:317.

26. Lawrance, S.K., L. Karlsson, J. Price, V. Quaranta, Y. Ron, J. Sprent, and P.A. Peterson. 1989. Transgenic HLA-DRα faithfully reconstitutes IE-controlled immune functions and induces cross-tolerance to Eta in Eta mutant mice. *Cell.* 58:583.

27. O'Sullivan, D., J. Sidney, E. Appella, L. Walker, L. Phillips, S.M. Colén, C. Miles, R.W. Chestnut, and A. Sette. 1990. Characterization of the specificity of peptide binding to four DR haplotypes. *J. Immunol.* 145:1799.

28. Kamikawaji, N., K. Morii, Y. Fujita, H. Yoshizumi, V.A. Fichetti, A. Kimura, Y. Nishimura, and T. Sasazuki. 1992. Identification of T cell epitopes in M protein from type 12 streptococcus. In *HLA 1991,* Vol. II. K. Tsujii, M. Aizawa, T. Sasazuki, editors. Oxford University Press, Oxford, 604–606.

29. Fukui, Y., K. Yamamoto, N. Yokoyama, T. Iwanaga, C. Kurashima, Y. Esaki, A. Kimura, T. Akashi, K. Hirokawa, and T. Sasazuki. 1993. Restricted expression of transgenic HLA-DRA gene in thymic epithelial cells and its role in acquisition of T cell tolerance to self-superantigens and processed DRα-restricted peptide. *Eur. J. Immunol.* 23:1678.

30. Zhou, P., G.D. Anderson, S. Savarirayan, H. Inoko, and C.S. David. 1991. Thymic deletion of Vβ11+, Vβ5+ T cells in H-2E negative, HLA-DQ8+ single transgenic mice. *J. Immunol.* 146:854.

171 Yamamoto et al.