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Fluorogenic Substrates for In Situ Monitoring of Caspase-3 Activity in Live Cells

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Abstract

The in situ detection of caspase-3 activity has applications in the imaging and monitoring of multiple pathologies, notably cancer. A series of cell penetrating FRET-based fluorogenic substrates were designed and synthesised for the detection of caspase-3 in live cells. A variety of modifications of the classical caspase-3 and caspase-7 substrate sequence Asp-Glu-Val-Asp were carried out in order to increase caspase-3 affinity and eliminate caspase-7 cross-reactivity. To allow cellular uptake and good solubility, the substrates were conjugated to a cationic peptoid. The most selective fluorogenic substrate 27, FAM-Ahx-Asp-Leu-Pro-Asp-Lys(MR)-Ahx, conjugated to the cell penetrating peptoid at the C-terminus, was able to detect and quantify caspase-3 activity in apoptotic cells without cross-reactivity by caspase-7.

Introduction

Fluorogenic substrates and activity-based probes enable the study of protease function and have been used to elucidate the role of caspases in the progression of diseases such as cancer [1–6], neurodegenerative disorders [7–10], and sepsis [11,12]. Caspases are an important family of cysteine-dependent aspartate proteases that exist within cells as inactive zymogens with their cleavage giving active enzymes initiating cellular apoptosis [13–15]. Inappropriate control of this apoptotic machinery is implicated in many diseases [14,15], notably cancer [16,17]. As a part of the apoptotic cascade, executioner caspase-3 activates several important cellular substrates [14–20], such as PARP and ICAD, and its decreased activity is a prognostic indicator of chemoresistance in breast and ovarian cancer [21,22]. The ability to monitor caspase-3 activity in situ could provide a means, not only to elucidate its complex role in biological processes, but also to monitor the efficacy of anticancer drugs and to identify patients for whom discontinuation of ineffective toxic treatment is warranted, for example, due to acquired drug resistance [23].

Current methods are able to detect caspase-3 activity in vitro although they often display promiscuity and cannot be used to monitor caspase-3 within cells [24–26]. The majority of
fluorogenic caspase-3 substrates are based on a four-residue recognition sequence Asp-Glu-Val-Asp (DEVD) [23,27,28], established via combinatorial library methods [29,30]; however, this sequence is also efficiently cleaved by caspase-7, which shares very similar substrate specificities with caspase-3. In mouse macrophages, 46 out of the 55 identified protein cleavage sites (within 48 proteins) were cleaved by both enzymes with only 3 sites specifically cleaved by caspase-3 [31]. Incorporation of unnatural amino acids into the recognition sequence has yielded caspase-3 substrates with increased selectivity [32,33]. Recently, Wolan achieved live cell imaging of caspase-3 activity in apoptotic cells, with selectivity over caspase-7, with a near-infrared fluorogenic pentapeptide substrate (incorporating the unnatural amino acid β-homo-Leu) coupled to a cell penetrating peptide derived from the viral SV40 Large T-antigen nuclear localising signal [34].

Here, FRET-based fluorogenic substrates, incorporating a tetrapeptide recognition sequence Asp-X3-X2-Asp, were designed and synthesised for the selective, in situ monitoring of caspase-3 activity. To allow detection in live cells, the substrates were conjugated to a cationic peptoid-based cellular delivery vehicle.

**Results and Discussion**

**Substrate design and synthesis**

In order to improve selectivity towards caspase-3 over caspase-7, permutations of the classical tetrapeptide substrate Asp-Glu-Val-Asp (X1-X3-X2-X4) were explored. All known caspase-3 substrates contain Asp at position X1 and 80% contain an Asp at the X4 position of the sequence. The positions X3 and X2 are more varied, with no clear amino acid preference being reported for the X3 position (~20% of the substrates contain Glu and ~15% Phe or Val at this position). Approximately 40% of known caspase-3 substrates contain Val at the X2 position; however, Pro at the X2 position is known to increase specificity for caspase-3 over caspase-7 [35]. With the aim of improving caspase-3 selectivity, X3 and X2 modifications of the tetrapeptide sequence were carried out, retaining Asp at X1 and X4 positions. The X3 position was changed to Pro, Gly, Ala, Leu, Asn and Val, and the X2 position had Val (substrates 1–9) or Pro (substrates 10–14) (Table 1) [36,37]. For each substrate, the corresponding d-amino acid sequence was synthesised as a control (compounds 15–24, respectively, see ESI). 5(6)-Carboxyfluorescein was coupled to the N-terminus of the substrates via a 6-aminohexanoic acid (Ahx) spacer, and a quencher moiety was introduced next to the caspase cleavage site via Lys side chain modification (separated by an Ahx spacer from the Asp-X3-X2-Asp) (Fig 1). As the choice of the quencher can affect the rate of cleavage and level of background fluorescence, three different quenchers, methyl red (MR), Black Hole Quencher1 (BHQ1), and 5(6)-carboxytetraethylrhodamine (TAMRA) were evaluated. A cationic, “lysine-like” nonaresidue peptoid was incorporated onto the C-terminus to ensure cellular uptake of the substrates [38]. Unlike many common cell penetrating peptides [39–41] this peptoid is resistant to proteolysis, non-toxic in vivo, and has demonstrated a highly efficient cell entry profile [42–44]. In addition, peptoid-based delivery systems are not prone producing immunogenic responses associated with virus-derived sequences [45,46].

The peptides 1–14 and controls 15–24 were synthesised on a Rink amide-functionalised aminomethyl polystyrene resin (1% DVB, 100–200 mesh, loading 1.2 mmol/g) using an Fmoc/tBu-based strategy with microwave heating (S1 Fig) [47]. First, the nonapeptoid was synthesised using N-Fmoc-(6-Boc-aminohexyl)glycine [48] and DIC and Oxyma. Fmoc-Lys (Dde)-OH was coupled onto the peptoid, followed by Fmoc-Ahx-OH, and the substrate sequence (Asp-X3-X2-Asp), Fmoc-Ahx-OH, and 5(6)-carboxyfluorescein. The Lys side chain Dde protecting group was selectively removed with 2% hydrazine (v/v), followed by coupling
of the carboxy-functionalised quencher. After deprotection and cleavage from the resin with TFA–TIS–DCM, peptides 1–24 were purified by preparative HPLC and analysed by MALDI-TOF MS.

The effect of the quencher on caspase-3 cleavage

To optimise the fluorogenic substrates, MR (λ_{Abs} 480 nm), TAMRA (λ_{Abs} 555 nm), and BHQ1 (λ_{Abs} 534 nm) were evaluated as quenchers for 5(6)-carboxyfluorescein (λ_{Ex/Em} 488/528 nm) in compounds 1, 2, and 3, using the classical substrate Asp-Glu-Val-Asp. Only 1 (MR as quencher) showed notable time-dependent increase in fluorescence upon incubation with caspase-3 and 7 (2 and 3, incorporating TAMRA and BHQ1, did not demonstrate significant increase in fluorescence) (Fig 2).

Kinetic studies with Caspase-3 and 7

The ability of 1 and 5–24 (bearing MR as the quencher of choice) to act as a substrate for caspase-3, as well as caspase-7, was investigated by determining the catalytic efficiency (k_{cat}/K_M) for each substrate with both enzymes (Table 2). None of the d-amino acids containing sequences 15–24 or 4, which has a Pro residue at position X_3, showed any change in fluorescence intensity over time. With caspase-3, all the substrates with Val in X_2 position (substrates 5–9) showed similar catalytic efficiency (k_{cat}/K_M 0.7–2.0 μM^{-1}min^{-1}) as Asp-Glu-Val-Asp (substrate 1, 1.4 μM^{-1}min^{-1}). Substrate 5 (Asp-Gly-Val-Asp) showed 5-fold selectivity over caspase-7 (Table 2). Pro at the X_3 position increased specificity for caspase-3 over caspase-7 [49], particularly with substrates 11 (Asp-Ala-Pro-Asp), 12 (Asp-Leu-Pro-Asp) and 14 (Asp-Val-Pro-Asp), which exhibited 8–20-fold selectivity over caspase-7, along with increased caspase-3 affinity (K_M 0.2–0.4 μM) and catalytic efficacy (k_{cat}/K_M 3.4–8.1 μM^{-1}min^{-1}).

To confirm how specific 11, 12 and 14 were for caspase-3, these substrates were incubated with high enzyme concentrations (10–20 μM substrate, 0.4 μM enzyme) and the caspase-mediated cleavage analysed by MALDI-TOF MS. As expected, all the substrates were cleaved by

| substrate | X_4-X_3-X_2-X_1 | quencher (Q) |
|-----------|-----------------|--------------|
| 1         | Asp-Glu-Val-Asp | MR           |
| 2         | Asp-Glu-Val-Asp | TAMRA        |
| 3         | Asp-Glu-Val-Asp | BHQ1         |
| 4         | Asp-Pro-Val-Asp | MR           |
| 5         | Asp-Gly-Val-Asp | MR           |
| 6         | Asp-Ala-Val-Asp | MR           |
| 7         | Asp-Leu-Val-Asp | MR           |
| 8         | Asp-Asn-Val-Asp | MR           |
| 9         | Asp-Val-Val-Asp | MR           |
| 10        | Asp-Gly-Pro-Asp | MR           |
| 11        | Asp-Ala-Pro-Asp | MR           |
| 12        | Asp-Leu-Pro-Asp | MR           |
| 13        | Asp-Asn-Pro-Asp | MR           |
| 14        | Asp-Val-Pro-Asp | MR           |

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caspase-3 at the X₁ position (between Asp and the Ahx spacer), with the parent compound no longer detected after 2h; however, these substrates were also (partially) cleaved by caspase-7. Remarkably, MS analysis revealed that caspase-7 exhibited a different cleavage pattern cleaving the substrates between the Ahx spacer and the Lys(MR) (S2–S4 Figs).

Substrate optimisation

To eliminate the caspase-7 cross reactivity, three new substrates, all bearing Asp-Leu-Pro-Asp, were synthesised using d-Lys or N-Methyl-Lys as the quencher attachment point (25 and 26, respectively) and switching the position of the Ahx spacer (27) (Fig 3). Substrate 25 was cleaved by both caspase-3 and 7, whereas the N-methylated substrate 26 was not cleavage by either enzyme (S6 and S7 Figs). Substrate 27 showed good affinity for caspase-3 (K<sub>M</sub> 1.1 ± 0.3 μM) with k<sub>cat</sub> of 2.1 ± 0.8 min<sup>-1</sup> and a catalytic efficiency of 0.5 ± 0.08 μM<sup>-1</sup> min<sup>-1</sup>. Remarkably, caspase-7 (0.4 μM) did not show any cleavage of this substrate (S7 and S8 Figs).

Detecting caspase-3 activity in live cells

Caspase-3 activity in HEK293T cells was evaluated using the caspase-3 selective substrate 27 with apoptosis induced by staurosporine. Flow cytometry analysis of cells treated with 27
(10 μM) showed a 2.5-fold increase in the fluorescence intensity (λ<sub>Ex/Em</sub> 488/530 nm) of the cells after apoptotic stimulation by staurosporine (1 μM), with no increase observed in fluoresce without it (Fig 4). This increase in fluorescence with substrate 27 suggested that the concentration of caspase-3 in apoptotic cells was approximately 15.7 ± 0.5 nM per cell (28271 molecules per cell) in the execution phase of apoptosis (based on the V<sub>max</sub> and the k<sub>cat</sub> of 27, see supporting information) [50]. In live-cell confocal imaging of caspase-3 activation with 27 (10 μM), fluorescence “turn-on” was only detected in the cytoplasm of apoptotic HEK293T cells, with no increase in fluorescence observed in non-apoptotic cells (Fig 5A, 5B and S9 Fig). No fluorescence “turn-on” was observed in apoptotic MCF-7 cells (Fig 5C), which lack functional caspase-3 but express caspase-7 [34,51,52], confirming the isoform selectivity. Substrate 27 was nontoxic in an MTT assay at 10 μM concentration (S10 Fig).

Table 2. Kinetic analysis of the fluorogenic substrates. The kinetic parameters (n = 3) were determined for the fluorogenic substrates 1 and 5–14 (measured using substrate range of 0.1–8 μM) with caspase-3 and caspase-7. Substrate 4 or the d-amino acids containing 15–24 were not cleaved by either enzyme.

|   | K<sub>M</sub> (μM) | K<sub>cat</sub> (min<sup>-1</sup>) | k<sub>cat</sub>/K<sub>M</sub> (μM<sup>-1</sup>min<sup>-1</sup>) |   | K<sub>M</sub> (μM) | k<sub>cat</sub> (min<sup>-1</sup>) | k<sub>cat</sub>/K<sub>M</sub> (μM<sup>-1</sup>min<sup>-1</sup>) |
|---|------------------|-----------------|-----------------|---|------------------|-----------------|-----------------|
| 1 | 0.6 ± 0.1        | 0.9 ± 0.04      | 1.4             | 5 | 0.5 ± 0.1        | 1.1 ± 0.1       | 2.1             |
| 5 | 0.3 ± 0.1        | 0.5 ± 0.03      | 1.7             | 6 | 1.5 ± 0.4        | 1.1 ± 0.1       | 0.7             |
| 6 | 0.8 ± 0.2        | 0.9 ± 0.1       | 1.2             | 7 | 0.9 ± 0.2        | 1.0 ± 0.1       | 1.1             |
| 8 | 0.4 ± 0.1        | 0.8 ± 0.04      | 2.0             | 9 | 0.6 ± 0.1        | 0.8 ± 0.03      | 1.3             |
| 10| 0.7 ± 0.4        | 0.55 ± 0.2      | 0.8             | 10| 0.3 ± 0.05       | 0.2 ± 0.01      | 0.6             |
| 11| 0.4 ± 0.1        | 1.3 ± 0.1       | 3.4             | 11| 4.9 ± 2.1        | 1.6 ± 0.3       | 0.3             |
| 12| 0.2 ± 0.1        | 1.4 ± 0.2       | 5.8             | 12| 1.4 ± 0.3        | 0.9 ± 0.07      | 0.7             |
| 13| 0.8 ± 0.2        | 1.0 ± 0.2       | 1.2             | 13| 1.2 ± 0.2        | 1.0 ± 0.06      | 0.8             |
| 14| 0.2 ± 0.04       | 1.7 ± 0.1       | 8.1             | 14| 3.6 ± 0.9        | 1.4 ± 0.2       | 0.4             |

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Fig 3. Structural modifications to the fluorogenic substrates with the aim of eliminating caspase-7 cross-reactivity. Substrate 25 has a D-Lys residue, 26 an N-Methyl-Lys, and in substrate 27 the Ahx spacer has been moved between the Lys and the peptoid moiety. Caspase-3 selectivity was achieved with 27.

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Conclusions

Fluorogenic, FRET-based substrates for monitoring the enzymatic activity of caspase-3 \textit{in situ} were synthesised with permutations of the substrate sequence Asp-X$_2$-X$_3$-Asp with the aim of improving selectivity for caspase-3 over caspase-7. The fluorogenic substrates had 5(6)-carboxyfluorescein in the amino-terminus and an optimised quencher, methyl red, introduced via Lys.
side chain modification. These fluorogenic substrates were conjugated at the C-terminus to a cationic cell delivery vehicle to allow efficient cellular uptake. Substrates with Pro in the $X_2$ position (instead of Val), showed selectivity for caspase-3 over caspase-7, along with increased caspase-3 affinity, particularly when $X_3$ was Ala, Leu or Val. Mass spectrometry studies revealed unexpected cleavage pattern of the fluorogenic substrates with caspase-7, while optimisation of the substrate via spacer relocation yielded a caspase-3 selective substrate 27 (FAM-Ahx-Asp-Leu-Pro-Asp-Lys(MR)-Ahx-peptoid). In apoptotic cells, the optimised substrate 27 allowed imaging of caspase-3 activity in situ. Flow cytometry analysis gave approximate quantification of the concentration of caspase-3 in a cell to be 16 nM (28271 molecules per cell). Future work is aimed at the use of this caspase-3 selective sequence in in vivo near-infrared fluorescence imaging techniques, especially for cancer, which require stable, highly specific, and sensitive fluorogenic substrates.

Materials and Methods

Synthesis of fluorogenic substrates 1–27

A highly optimised microwave-based solid-phase strategy was used to synthesise the cell penetrating peptide–peptoids [38]. All solvents and reagents were obtained from commercial suppliers and used without purification. A Rink-amide functionalised aminomethyl polystyrene (1% DVB, 100–200 mesh, loading 1.2 mmol/g) resin was used for the synthesis of the peptides with an Fmoc-based strategy (S1 Fig). Coupling of the Fmoc-amino acids and fluorophores:
The resin (1 eq) was pre-swollen in DCM, washed with DMF, and added a pre-activated mixture (10 min) of the carboxylic acid (3 eq), DIC (3 eq) and Oxyma (3 eq) in DMF (0.1 M). This reaction mixture was stirred in the microwave (Biotage Initiator) for 20 minutes at 60°C after which the resin was washed with DMF, DCM and MeOH. Fmoc deprotection: The resin was shaken with 20% piperidine in DMF (2 × 10 min), and subsequently washed with DMF, DCM and MeOH. Dde deprotection: The resin was shaken with 2% hydrazine in DMF (v/v) (2 × 10 min), and subsequently washed with DMF, DCM and MeOH. Cleavage from the resin and deprotection: A solution of TFA/TIS/DCM (90:5:5) was added to the resin (20 μL of the cleavage cocktail per mg of resin) and left to shake for 5 hours. The resin was filtrated and washed with DCM, and the collected filtrate was evaporated under reduced pressure and the compound precipitated using cold diethyl ether. Peptides 1–24 were purified by preparative HPLC and analysed by MALDI-TOF MS. For the characterisation of the peptides, see S1 Table.

Kinetic assays with caspase-3 and 7

Caspase-3 or caspase-7 (R&D systems, USA) was added to 100 μL of caspase assay buffer with substrates 1–14 at concentrations from 0.1 μM to 8 μM in a 96-well plate ($n = 3$) to give final enzyme concentration of 20 nM (1–9) or 15 nM (10–14). Fluorescence ($\lambda_{Ex/Em}$ 485/528 nm) was recorded on a Biotek Synergy HT Multi-Mode Microplate Reader every 2 min. Control samples had the same composition but no enzyme. The rate (μM/min) was calibrated using a 5(6)-carboxyfluorescein conversion factor (0.0055 μM/RFU) and data plotted against time (min). For initial cleavage rate (0–5 min), plots were fitted using linear regression analysis and the Michaelis-Menten data generated using GraphPad Prism 5.

Caspase-3 detection in live cells

Cell culture was performed in a Heracell 150 incubator (Heraeus) and in a Herasafe KS 18 class II negative-flow cabinet (Heraeus). HEK293T and MCF-7 cells (cultured in high glucose (4.5 mg/mL) DMEM supplemented with 4 mM glutamine, 100 units/mL penicillin, 10 mg/mL
streptomycin and 25 mg/mL amphotericin B, and 10% FBS) were seeded onto a 48-well plate at a density of $10^4$ cells per well. After 12 hours, the media was removed and substrates 1–14 added at 10 μM in fresh media. Selected wells were also treated with staurosporine (1 μM). After 5 hours, the cells were washed twice with PBS, detached with trypsin/EDTA, harvested with 2% FBS in PBS supplemented with Trypan Blue (0.04%) for analysis on a BD FACSAnia flow cytometer. Fluorescence was evaluated as mean fluorescence intensity (MFI) and estimated <100 u.a. for untreated control cells (consistent values independently of staurosporine addition). Apoptotic cells treated with substrate 1 were used as a positive control to obtain the maximum of fluorescence signal. 50,000 events per sample were plotted in two-dimensional dot plots based on forward and side scattering. The cellular size and complexity (SSC-H vs. FSC-H) were used to gate two populations (alive/apoptotic cells) (debris excluded). The data were analysed using the software Flowjo 7.5.

For confocal microscopy, the cells were fixed with 4% paraformaldehyde in PBS and the nuclei stained with Hoechst-33342 (1% w/v in PBS). Cellular fluorescence of cells was analysed using an Inverted Leica DM IRB with filter I3 (450–490 nm) and a Leica SP5 Confocal (FITC and DAPI channel).

Supporting Information

S1 Fig. Solid phase synthesis of the fluorogenic substrates 1–24.
(PDF)

S2 Fig. MALDI-TOF MS spectra of substrate 11.
(PDF)

S3 Fig. MALDI-TOF MS spectra of substrate 12.
(PDF)

S4 Fig. MALDI-TOF MS spectra of substrate 14.
(PDF)

S5 Fig. MALDI-TOF MS spectra of substrate 25.
(PDF)

S6 Fig. MALDI-TOF MS spectra of 26.
(PDF)

S7 Fig. Analysis of substrate 27 with caspase-3 and caspase-7.
(PDF)

S8 Fig. MALDI-TOF MS spectra of 27.
(PDF)

S9 Fig. Substrate 27 selectively labels apoptotic cells.
(PDF)

S10 Fig. Cell viability.
(PDF)

S1 File. Quantification of caspase-3 in apoptotic cells by flow cytometry.
(PDF)

S1 Table. MALDI-TOF MS and HPLC analysis of substrates 1–27.
(PDF)
Author Contributions
Conceived and designed the experiments: AMPL MLSG AL MB. Performed the experiments: AMPL MLSG ERM. Analyzed the data: AMPL MLSG AL MB. Wrote the paper: AL AMPL MB.

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