Evaluation of biorelevant media to investigate the dissolution properties on flurbiprofen and to assess cytotoxicity effects on Caco-2 cell line

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ABSTRACT
Biorelevant media are used to simulate the physiological conditions in terms of components, pH, osmolality and buffer capacity of the human stomach and intestine in both fasted and fed states. In this study, we aim to apply the biorelevant media to Caco-2 cell lines to investigate the cytotoxicity effects via the cell viability ratio and to compare the solubilizing effects of various dissolution media on a poorly soluble model drug. Flurbiprofen (Biopharmaceutics Classification System, BCS Class 2) was selected as a model drug.

In dissolution studies the pH effects were predominant at higher pH values, while bile salt effects were dominant at lower pH values. The preparation method, bile salts and the phospholipids did not show any additional effect on Caco-2 cell viability. In the cytotoxicity test, fed state media caused an additional 10-15% decrease in cell viability, compared to the fasted state. Similar results were obtained when using the blank of these media which did not include the bile salt or phospholipid. From this, it is evident that this decrease resulted from the pH values, not the components. In conclusion, the cytotoxicity assessment showed that all the biorelevant media were compatible with 70-90% of cell viability for at least 24 h, and this ratio might be increased by modifying the pH.

Keywords: Biorelevant media, flurbiprofen, cytotoxicity

INTRODUCTION

Many studies have been conducted to develop a dissolution media which better reflects the contents of the human gastrointestinal system. In those studies, researchers added some enzymes, surfactants, bile salts, phospholipids, lipolysis products etc. to the media (Dressman et al. 1998; Galia et al. 1998; Tang et al. 2001; Wiedman et al. 2002; Jantraid et al. 2008). Additionally, pH, surface tension, and the osmolality of these media were taken into consideration. Regarding the effect of food on the absorption of BCS Class 2 drugs, the dissolution media for the fed state were also developed separately (Dressman et al. 1998; Galia et al. 1998; Jantraid et al. 2008).

Health authorities have not yet approved any biorelevant media other than compendial media. However, Dressman developed two basic media for reflecting intestine in the fasted and fed states (Dressman et al. 1998). These media are known as Fasted State Simulated Intestinal Fluid (FaSSIF) and Fed State Simulated Intestinal Fluid (FeSSIF). These media have attracted the attention of a many scientists and much research has been conducted in their use (Galia et al. 1998; Nicolaides et al. 1999; Kostewicz et al. 2002; Fagerberg et al. 2010). Moreover, several other media were developed based on these media (Marquez 2004; Jantraid et al. 2008; Fatouros et al. 2009; Kleberg et al. 2010; Klein 2010; Fuchs et al. 2013; Zhou et al. 2017). When the dissolution tests that were performed using these media provided a better correlation in vivo (Mathias et al. 2015; Xu et al. 2017), researchers started to use them in permeability studies (Patel et al. 2006; Birch et al. 2018). However, certain components in these media such as bile salts and phospholipids raised suspicions regarding the possible toxic effects on cell lines (Ingels
and Augustijns 2003; Birch et al. 2018). Apart from the components of these media, the preparation process is also important. Due to the physicochemical properties of bile salts and lecithin and the use of organic solvents in the process, the preparation of these media is not simple. To standardize the media preparation, an instant powder mix of these components was obtained (SIF Powder®) by Biorelevant.com (Boni et al. 2009).

Flurbiprofen is a rapidly absorbed, non-steroidal anti-inflammatory drug, with 96% oral bioavailability following oral administration. Absorption is increased with food (Pargal et al. 1996) and it eliminates 75-80% as metabolites and 20-25% as the unchanged drug in the urine. Flurbiprofen is a weak acid with a pKₐ of 4.22. It is a BCS Class 2 drug and practically insoluble in water. It has a solubility of 0.0080 mg/mL at pH 1.2 and its solubility increases with pH (Li and Zhao 2003). Yazdanian et al. calculated the permeability (P_app) of flurbiprofen as higher than verapamil used as the reference drug with a P_app of 20.1x10⁻⁶±2.7 x 10⁻⁶ cm/s (Yazdanian et al. 2004).

In this study, we aim to evaluate the possible cytotoxic effects of a high permeable model drug when prepared conventionally and when prepared from the instant powder. For this reason, we selected a BCS Class 2 drug - flurbiprofen. Moreover, we performed solubility and dissolution tests with the commercial product containing the model drug to compare the media against themselves. In addition to the biorelevant media, pharmacopoeial media and blank media not including the bile salts and phospholipids were also used to assess the pH effects.

**MATERIALS AND METHODS**

Sodium taurocholate 97% pure (high quality: HQ), egg-phosphatidylcholine, MTT and trypan blue were purchased from Sigma-Aldrich® (USA). All the chemicals and reagents were purchased from Merck® (Germany). SIF Powder® (Biorelevant.com, United Kingdom) is a patented formulation of sodium taurocholate and lecithin with the molar ratio of 4:1 which corresponds with Dressman’s formulation (Dressman et al. 1998).

The biorelevant media prepared using instant powder are defined as SIF-FaSSIF and SIF-FeSSIF in this study.

Dulbecco’s Modified Eagle’s Medium (DMEM), EDTA, Fetal bovine serum (FBS) and RPMI 1640 were purchased from Biochrom, Germany and Caco-2 cells were obtained from Cell Culture Collection, Turkey. Flurbiprofen (Sun Pharmaceuticals, India) was supplied from Drogsan Pharmaceuticals (Turkey). Ansaid® (Pfizer, Turkey) 100 mg film-coated tablets were purchased from a local drug market.

**Preparation of dissolution media**

The classical dissolution media of pH 1.2, pH 4.5, and pH 6.8 were prepared according to USP. FaSSIF and FeSSIF were prepared as previously reported (Marquez 2004). SIF-FaSSIF and SIF-FeSSIF; Blank FaSSIF, Blank FeSSIF were prepared in accordance with the protocols of Biorelevant.com. The compositions of the biorelevant media and blank media are shown in Table 1.

**Solubility**

Solubility measurements of the samples were performed using the shake-flask method and all the solubility experiments were conducted in triplicate. According to the method, the excess of the drug powder was added to 50 mL of different dissolution media and stirred at 37.0°C±0.1°C in a shaking incubator water bath. The equilibrium time was set to 24 h. The final solution was then filtered through a 0.45 μm (Millipore Millex-HV, USA) membrane filter and analyzed using a UV spectrophotometer (Shimadzu, UV-170, Japan). To evaluate the solubility results of the drug, dose number (D₀), which is defined as the ratio of drug concentration in the administered volume to the saturation solubility of the drug (Oh et al. 1993) was used. It was calculated using Equation 1.

\[
D₀ = \frac{M₀}{V₀ C₅}
\]

where M₀ is the dose of drug administered, V₀ is the administered volume, and C₅ is the saturation solubility. Fluid volume used with the drug was set as 250 mL - the volume of a glass

| Compositions | FaSSIF* | FeSSIF* | SIF-FaSSIF | SIF-FeSSIF | Blank FaSSIF | Blank FeSSIF |
|--------------|---------|---------|------------|------------|--------------|--------------|
| Sodium taurocholate | 3 mM | 15 mM | 3 mM | 15 mM | - | - |
| Lecithin | 0.75 mM | 3.75 mM | 0.75 mM | 3.75 mM | - | - |
| NaH₂PO₄·H₂O | 1.977 g | - | 1.977 g | - | 1.977 g | - |
| Glacial acetic acid | - | 8.65 g | - | 8.65 g | - | 8.65 g |
| NaCl | 3.093 g | 11.874 g | 3.093 g | 11.874 g | 3.093 g | 11.874 g |
| NaOH (pellets) | 0.174 g | 4.04 g | 0.174 g | 4.04 g | 0.174 g | 4.04 g |
| Deionized water (qs) | 500 mL | 1000 mL | 500 mL | 1000 mL | 500 mL | 1000 mL |
| pH | 6.5 | 5.0 | 6.5 | 5.0 | 6.5 | 5.0 |

*Lecithin was dissolved in dichloromethane, after emulification dichloroethane was evaporated.

FaSSIF: Fasted State Simulated Intestinal Fluid, FeSSIF: Fed State Simulated Intestinal Fluid; SIF-FaSSIF: Fasted State Simulated Intestinal Fluid prepared using instant powder; SIF-FeSSIF: Fed State Simulated Intestinal Fluid prepared using instant powder; Blank FaSSIF: Fasted State Simulated Intestinal Fluid media not including the bile salts and phospholipids; Blank FeSSIF: Fed State Simulated Intestinal Fluid media not including the bile salts and phospholipids.
of water. While a $D_n$ which is equal to or lower than 1 means a high-solubility, a $D_n$ higher than 1 indicates a low-solubility.

**In vitro dissolution studies**

USP apparatus 2 (PharmaTest, Germany) was used for all dissolution tests. The dissolution studies were carried out at 37±0.5°C in 900 mL of dissolution media and with a rotational speed of 50 rpm. At each predetermined sample time intervals (5, 10, 15, 20, 30, 45 and 60 minutes), 5 mL of the sample was taken, and 5 mL of blank medium was replaced. All samples were filtered using 0.45 μm (Millipore Millex-HV, USA) membrane filter and after diluting, they were analyzed by the validated UV spectrophotometric method (Shimadzu, UV-170, Japan). All experiments were performed in triplicate. The dissolution test results were evaluated using the similarity factor ($f_2$) to compare the mediums.

$$f_2 = 50.10\log\left(1 + \frac{1}{n}\sum_{i=1}^{n}(R_i - T_i)^2\right)^{0.5}\times 100$$  

Eq 2

Where $n$ is the number of time points, $R_i$ is the dissolved amount of the reference at time $t$, and $T_i$ is the dissolved amount of the test at time $t$.

**Spectrofotometric analysis**

A Shimadzu UV-170 Spectrophotometer (Japan) was used for UV analysis. All assay and dissolution studies were analyzed using a UV-spectrometer. The maximum absorbance values of the drugs in various media were different. The wavelengths ($\lambda_{max}$) that are being used were 246 nm (pH 4.5, pH 6.8, Blank FaSSIF, FaSSIF and Blank FeSSIF), 248 nm (SIF-FeSSIF) and 250 nm (FeSSIF and SIF-FeSSIF).

**Cytotoxicity assessments**

Caco-2 cells were grown at 37°C in an atmosphere of 5% $CO_2$ in Dulbecco’s Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum. A confluent cell line was washed with trypsin/EDTA solution (0.05%/0.02%) and kept in an incubator for 5-10 minutes and then centrifuged. The cell line was homogenized by DMEM with 10% of serum and 1% of antibiotic. Three passages were performed. For the cell count 0.1 mL of trypan blue was added to 0.9 mL of cell suspension and the cell count was conducted by a hemacytometer. The cell viability was then checked with an optical microscope. The cells were homogenized in DMEM containing 10% FBS and 1% antibiotic and then a 4x10^6 cell/mL cell suspension was prepared and transferred to 96-well cell plates (100 μL/well) and incubated in 5% $CO_2$ for 24 hours.

**MTT viability test**

The MTT [3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide] test was a colorimetric assay that can be used to determine cell viability (mitochondrial activity) measuring the extent of formazan formation after the lysis of the living material and the solubilization of formazan crystals (Berridge et al. 2005).

The MTT [3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide] test was performed to evaluate the cytotoxicity of sodium taurocholate, lecithin and flurbiprofen in biorelevant media with incubation times of 1 and 24 hours and concentrations of 100 μM and 500 μM. These concentrations were selected with reference to previous transport studies (Laitinen et al. 2007). The cell viability values were calculated as a percentage in control groups according to the following equation:

$$Cell\,\,viability\,\,(%) = \frac{A_{test}}{A_{control}}\times 100$$  

Eq 3

where $A_{test}$ is the absorbance of test and $A_{control}$ is the absorbance of DMEM.

**RESULTS AND DISCUSSION**

**Solubility**

The solubility studies were assessed following two methods: 1) the effect of pH and 2) the effect of concentrations of sodium taurocholate and lecithin. In addition, the preparation method of the biorelevant media was also considered. The solubility test results of flurbiprofen and estimated dose figures are summarized in Table 2. Flurbiprofen is a weak acid, thereby it is soluble when the pH value is greater than its $pK_a$ value of 4.22. For this reason, flurbiprofen did not practically dissolve at pH 1.2 and the solubility increased with pH. When the solubilization effects of sodium taurocholate and lecithin were not considered, flurbiprofen showed the maximum solubility at pH 6.8, the highest pH. The solubility at pH 6.8 was four times higher than at pH 4.5. The solubility values of flurbiprofen at different pH levels (Table 2) were in a strong correlation with the literature data (Li and Zhao 2003). To evaluate the effect of sodium taurocholate and lecithin in the fasted state (Table 3), Blank FaSSIF was compared with FaSSIF and SIF-FaSSIF. The solubility values of flurbiprofen in Blank FaSSIF, FaSSIF, and SIF-FaSSIF, while similar, were lower than the other media not including the bile salts and phospholipids.

**Table 2. Solubility and dose number values of flurbiprofen in different dissolution media.**

| Dissolution media  | Solubility±SD (mg/mL) | $D_n$±SD | Dissolved % in 30 min±SD |
|--------------------|-----------------------|----------|-------------------------|
| pH 4.5             | 0.055±0.0003          | 7.21±0.04| 39.6±0.4                |
| pH 6.8             | 2.53±0.05             | 0.323±0.006| 90.9±0.9               |
| Blank FaSSIF       | 1.78±0.03             | 0.224±0.003| 93.2±1.7               |
| FaSSIF             | 1.25±0.02             | 0.158±0.003| 95.7±3.0               |
| SIF-FaSSIF         | 1.70±0.01             | 0.237±0.001| 98.6±0.9               |
| Blank FeSSIF       | 0.075±0.0014          | 5.29±0.09 | 58.3±1.0               |
| FeSSIF             | 0.52±0.007            | 0.756±0.011| 76.1±2.4               |
| SIF-FeSSIF         | 4.20±0.02             | 0.0946±0.0005| 104.5±0.1             |

$D_n$: Dose number SD: Standard deviation, FaSSIF: Fasted State Simulated Intestinal Fluid, FeSSIF: Fed State Simulated Intestinal Fluid, Blank FaSSIF: Fasted State Simulated Intestinal Fluid prepared using instant powder, Blank FeSSIF: Fed State Simulated Intestinal Fluid prepared using instant powder, FaSSIF: Fasted State Simulated Intestinal Fluid media not including the bile salts and phospholipids, Blank FeSSIF: Fed State Simulated Intestinal Fluid media not including the bile salts and phospholipids.
the solubility at pH 6.8 media which has a higher pH value than biorelevant media. Interestingly, neither FaSSIF nor SIF-FaSSIF provided a more significant increase than Blank-FaSSIF. To investigate the food effect on flurbiprofen solubility, we compared the Blank FeSSIF, FeSSIF and SIF-FeSSIF. The solubility of flurbiprofen was increased seven-fold in FeSSIF compared with Blank FaSSIF. However, in SIF-FeSSIF flurbiprofen was eight times more soluble than FeSSIF and 56 times more soluble than Blank-FaSSIF. This dramatic variation between FeSSIF and SIF-FeSSIF is thought to result from the preparation processes. FaSSIF and FeSSIF were prepared using dichloromethane as a solvent and after the emulsification process, the organic solvent should be evaporated as reported in the literature (Dressman et al. 1998). Therefore, it is difficult to standardize the preparation method. It may be affected by the capacity of the equipment in the emulsification and evaporation processes. However, the SIF-Powder was simply dissolved in the blank mediums as described in Biorelevant media. Therefore, probable variations arising from using different analysts, equipment or time can be minimized.

Based on the D₀ values given in Table 3, flurbiprofen was found to have low solubility in pH 4.5 and blank FaSSIF (D₀>1), while it was highly soluble (D₀<1) in all the other media (Table 3).

| Table 3. Comparing the flurbiprofen dissolution curves in different media |
|---------------------------------------------------------------|
| Test media          | Reference media | f₂ value* | Similarity |
|---------------------|-----------------|-----------|------------|
| Blank FaSSIF        | FaSSIF          | 64.2      | Similar    |
| Blank FaSSIF        | SIF-FaSSIF      | 63.3      | Similar    |
| FaSSIF              | SIF-FaSSIF      | 54.0      | Similar    |
| Blank FeSSIF        | FeSSIF          | 36.2      | Not similar|
| Blank FeSSIF        | SIF-FeSSIF      | 16.3      | Not similar|
| FeSSIF              | SIF-FeSSIF      | 27.2      | Not similar|

*When similarity factor, f₂>50 dissolution curves are similar

FaSSIF: Fed State Simulated Intestinal Fluid, FeSSIF: Fed State Simulated Intestinal Fluid, SIF-FaSSIF: Fasted State Simulated Intestinal Fluid prepared using instant powder, SIF-FeSSIF: Fed State Simulated Intestinal Fluid prepared using instant powder, Blank FaSSIF: Fasted State Simulated Intestinal Fluid media not including the bile salts and phospholipids, Blank FeSSIF: Fed State Simulated Intestinal Fluid media not including the bile salts and phospholipids

In vitro dissolution studies
Dissolution studies were performed to witness the media effects on BCS Class 2 drug, Ansaid® (Pfizer, Turkey) 100 mg was used for the dissolution studies as a reference product for flurbiprofen. The mean dissolution profiles of flurbiprofen are shown in Figure 1. Since flurbiprofen is a weak acid with a pKₐ of 4.22, the ionized form of flurbiprofen increases as the pH of the dissolution media increases. After 1 hour, the dissolved ratio of flurbiprofen from tablets was 47% at pH 4.5 buffer, whereas the dissolved amount was 68% in Blank FeSSIF (pH 5.0). Although, the dissolution of flurbiprofen increased with a pH higher than pKₐ of 4.22, a complete dissolution was not observed at pH 5.0. Given the use of SIF-FeSSIF containing bile salt/lecithin media with the same pH, a great enhancement resulting in complete dissolution was observed. Nevertheless, in FeSSIF, media which includes the same amount of bile salt and lecithin, a complete dissolution was not achieved. When comparing FeSSIF and SIF-FeSSIF, the bile salt and lecithin containing media, this significant difference is thought to result from the preparation conditions. SIF-FeSSIF is an easy-to-prepare instant powder, whereas FeSSIF involves a complicated preparation process including an organic solvent evaporation step. The similarity factors of the biorelevant media and their blanks are given in Table 3. More than 85% of the drug was released in 30 minutes at pH 6.8, Blank FaSSIF, FaSSIF and SIF-FaSSIF with pH 6.5. While the pH effect was predominant at the higher pH values, bile salt effect was dominant at the lower pH levels. Therefore, thanks to the pH of FaSSIF 6.5, it may not be necessary to use the bile salts in media. However, the dissolution results of FeSSIF, Blank FeSSIF and SIF-FeSSIF were not found to be similar. SIF-FeSSIF significantly increased the release of flurbiprofen. This means that the preparation method is equally as important as the composition of the fed state biorelevant media. In line with these results, we decided to assess the preparation method of the biorelevant media when evaluating the cytotoxicity on the Caco-2 cell line.

Cytotoxicity results
The effects of sodium taurocholate and lecithin, DMSO, and flurbiprofen on the mitochondrial dehydrogenase activity were studied in the Caco-2 cell monolayers. Conventionally prepared biorelevant media (Dressman et al. 1998), and the media prepared using SIF-Powder” (the instant powder), were evaluated separately. To eliminate the pH effect, the media without bile salt and phospholipid (Blank media) were also investigated and DMEM without serum was used as a control. Additionally, a model drug (flurbiprofen) at two different concentrations (100 μM and 500 μM) was evaluated in these biorelevant media. Cell viability was scored according to the following classification (Dahi et al. 2006): More than 90 percent of cell viability was defined as "non-cytotoxic", 60-90 percent of cell viability was defined as "slightly cytotoxic", 30-59 percent of cell viability was defined as "moderately cytotoxic" and less than 30 percent of cell viability was defined as "severely cytotoxic".

Since the model drug possesses a high permeability, the initial measurement was carried out at the first hour and
and another measurement was performed, at the 24th hour. The cell viability percents of cells in Blank FaSSIF, FaSSIF and SIF-FaSSIF were 88.4%±7.2, 93.7%±6.7 and 91.3%±2.7 after 1 hour and 73.2%±1.5, 78.5%±1.6 and 77.5%±8.0 after 24 hours, respectively (Table 4). According to the cytotoxicity classification, it was accepted as “slightly cytotoxic”. When Blank FaSSIF without bile salts was compared with FaSSIF and SIF-FaSSIF which included 3 mM sodium taurocholate and 0.75 mM lecithin, no difference was observed. Moreover, the viability results at 24 hours were nearly 15% lower than the results at 1 hour. The viability percents of cells in Blank FeSSIF, FeSSIF and SIF-FeSSIF were 86.1%±2.4, 84.3%±4.2 and 82.4%±4.8 after 1 hour and 70.8%±8.2, 73.2%±3.0 and 69.1%±4.4 after 24 hours, respectively. According to the cytotoxicity classification, they were accepted as “slightly cytotoxic”. When Blank FeSSIF without bile salts were compared with FeSSIF and SIF-FeSSIF which included 15 mM sodium taurocholate and 3.75 mM lecithin, no difference was observed. However, the viability results at 24 hours were lower by between 10-15% when compare with the results at 1 hour (Figure 2). This decrease may result from the different pH of DMEM (pH 7.4) and the biorelevant media (pH 6.5 for FaSSIF and pH 5.0 for FeSSIF). Antoine et al. (2015) found similar results with 83%±24 and 69%±17 of cell viability after 2 hours using FaSSIF and FeSSIF, respectively. In another study (Patel et al. 2006), while cell viability results were close to our results for FaSSIF (nearly 70%), FeSSIF was found very cytotoxic on cells with about 10% of viability after 2 h. For this reason, Patel et al. (2006) recommended modifications on FeSSIF when using in permeability studies. Ingels and Augustijns (2003) found 96.9% ± 20.1 cell viability for FaSSIF, but very low cell viability (5.4%±0.3) for FeSSIF. These results are conflicting and none of them were comparable with the blank media. Therefore, our blank-controlled study has given us more reliable results. The cell viability results are given in Table 4. The cytotoxicity results of flurbiprofen in concentrations of 100 μM and 500 μM were also similar to the results of the biorelevant media without the drug. The results were between 80-94% (mean value of 86.3±6.3%) after 1 hour and 72-87% (mean value of 79.6±6.0%) after 24 hours. These results have shown us that no additional toxic effect associated with the drug was obtained considering the viability results of cells (Figure 3).

As flurbiprofen is a weak acid, its solubility was affected by both pH and the bile salt content of the dissolution media. The effect of the bile salt content was found more significant in the media at pH values lower than the pKa of flurbiprofen, while the pH effect was more distinctive at higher pH values. Likewise, biorelevant media which was prepared using SIF-Powder®, gave more notable results in solubility and dissolution rate. As for the results of MTT tests on Caco-2 cells, there was no significant difference between the biorelevant media prepared conventionally and media prepared using the instant powder. Additionally, no meaningful difference was observed between the biorelevant media and their blanks in the Caco-2 cell viability. This means that, contrary to expectations, the main effect on cell viability was pH, with regard to bile salt and lecithin effects. Although, the cell...
viability results for the media which reflect the fasted state were higher than the fed state media, all media were categorized as "slightly cytotoxic" with about 70-90% of cell viability. Therefore, all media might be used in Caco-2 permeability studies. The viability results of cells decreased in the ratio of 10-15% after 24 hours compared to the results at 1 hour. DMEM, a routinely used media, has a pH of 7.4 whereas the studied pH values for fasting and fed states were 6.5 and 5.0, respectively. This decrease in the viability results was related to the lower pH values. The cytotoxicity results using a model drug -flurbiprofen (BCS Class 2) - in two different concentrations were similar to the viability results of the media without any drug. This finding suggests that model drugs had no effect on cell viability.

In conclusion, biorelevant media is a solid method of approach in evaluating in vitro drug performance and development of new drug pharmaceuticals. However, the preparation of the method of the media is also vital for determining the efficiency of the tests. In that respect, dissolution media prepared from instant powder (such as SIF Powder®) are more useful for low soluble drugs. We also saw a dramatic difference in dissolution and solubility results in the fed state of flurbiprofen - a Class 2 drug. Furthermore, these media can be used for safely performing permeability studies since the bile salts and lecithin did not cause any cytotoxic effect on the Caco 2 cells. However, minor changes in pH of these media can affect the results. Therefore, pH adjustment should be considered in further investigations.

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**REFERENCES**

- Amidon G, Lennernäs H, Shah VP, Crison JR (1995). A theoretical basis for a pharmacokinetic drug classification: The correlation of in vitro drug product dissolution and in vivo bioavailability. *Pharm Res* 12(8): 413-420. [CrossRef]
- Antoine D, Pellequer Y, Tempesta C, Lorscheidt C, Kettel B, Tamaddon L, Jannin V, DeMarne F, Lamprecht A, Béduneau A (2015). Biorelevant media resistant co-culture model mimicking permeability of human intestine. *Int. J. Pharm.* 481: 27 – 36. [CrossRef]
- Berridge MV, Herst PM, Tan AS (2005). Tetrazolium dyes as tools in cell biology: new insights into their cellular reduction. *Biotechnol Annu Rev* 11: 127-152. [CrossRef]
- Birch D, Diedrichsen RG, Christophersen PC, Mu H, Nielsen HM (2018). Evaluation of drug permeation under fed state conditions using mucus-covered Caco-2 cell epithelium. *Eur J Pharm Sci* 118: 144-153. [CrossRef]
- Boni JE, Brickl RS, Dressman J, Pfefferle MR (2009). Instant FaSSIF and FeSSIF – biorelevance meets practicality. *Disp Tech* 16: 41-45. [CrossRef]
- Dahl JE, Frangou-Polyzois MJ, Polyzois GL (2006). In vitro biocompatibility of denture relining materials. *Gerodontology* 23: 17-22. [CrossRef]
- Dressman JB, Amidon GL, Reppas C, Shah VP (1998). Dissolution testing as a prognostic tool for oral drug absorption: Immediate release dosage forms. *Pharm Res* 15: 11-22. [CrossRef]
- Fagerberg JH, Tsinman O, Sun N, Tsinman K, Avdeef A, Bergström CAS (2010). Dissolution rate and apparent solubility of poorly soluble drugs in biorelevant dissolution media. *Mol Pharm* 7: 1419-1430. [CrossRef]
- Fatouros DG, Walrand I, Bergenstahl B, Müllerertz A (2009). Colloidal structures in media simulating intestinal fed state conditions with and without lipolysis products. *Pharm Res* 26: 361-374. [CrossRef]
- Fuchs A, Leigh M, Kloefker B, Dressman JB (2015). Advances in the design of fasted state simulating intestinal fluids: FaSSIF-V3. *Eur J Pharm Biopharm* 94: 229-240. [CrossRef]
- Galia E, Nicolaides E, Hörter D, Löbenberg R, Reppas C, Dressman JB (1998). Evaluation of various dissolution media for predicting in vivo performance of class I and II drugs. *Pharm Res* 15: 698-705. [CrossRef]
- Ingels F, Deferme S, Destexhe E, Oth M, Mooter G, Augustijns P (2002). Simulated intestinal fluid as transport medium in the Caco-2 cell-culture model. *Int J Pharm* 232: 183-192. [CrossRef]
- Ingels FM, Augustijns PF (2003). Biological, pharmaceutical and analytical consideration with respect to the transport media used in the absorption screening system Caco-2. *J Pharm Sci* 92: 1545-1558. [CrossRef]
- Jantraid E, Janssen N, Reppas C, Dressman JB (2008). Dissolution media simulating conditions in the proximal human gastrointestinal tract: an update. *Pharm Res* 25: 1663-1676. [CrossRef]
- Kleborg K, Jacobsen F, Fatouros DG, Müllerertz A (2010). Biorelevant media simulating fed state intestinal fluids: Colloid phase characterization and impact on solubilization capacity. *J Pharm Sci* 99: 3522-3532. [CrossRef]
- Klein S (2010). The use of biorelevant dissolution media to forecast the in vivo performance of a drug. *AAPS J* 12: 397-406. [CrossRef]
- Kostewicz ES, Draus U, Becker R, Dressman JB (2002). Forecasting the oral absorption behavior of poorly soluble weak bases using solubility and dissolution studies in biorelevant media. *Pharm Res* 19: 345-349. [CrossRef]
- Laitinen L, Takala E, Vuorela H, Vuorela P, Kaukonen AM, Marvola M (2007). Anthranoid laxatives influence the absorption of poorly permeable drugs in human intestinal cell culture model (Caco-2). *Eur J Pharm Biopharm* 66: 135-145. [CrossRef]
- Li P, Zhao L (2003). Solubilization of flurbiprofen in pH-surfactant solutions. *J Pharm Sci* 92: 951-956. [CrossRef]
- Marques M (2004). Dissolution media simulating fasted and fed states. *Disp Tech* 11: 16. [CrossRef]
- Mathias N, Xu Y, Vig B, Kestur U, Saari A, Crison J, Desai D, Vanarase A, Hussain M (2015). Food effect in humans: predicting the risk through in vitro dissolution and in vivo pharmacokinetic models. *AAPS J* 17: 988-998. [CrossRef]
- Nicolaides E, Galia E, Efthymiopoulos C, Dressman JB, Reppas C (1999). Forecasting the in vivo performance of four low solubility drugs from their in vitro dissolution data. *Pharm Res* 12: 1876-1882. [CrossRef]
- Oh DM, Curl RL, Amidon GL (1993). Estimating the fraction dose absorbed from suspensions of poorly soluble compounds in humans: a mathematical model. *Pharm Res* 10: 264-270. [CrossRef]
• Pargal A, Kelkar MG, Nayak PJ (1996). The effect of food on the bioavailability of ibuprofen and flurbiprofen from sustained release formulations. *Biopharm Drug Disp* 17: 511-519. [CrossRef]

• Patel N, Forbes B, Eskola S, Murray J (2006). Use of Simulated Intestinal Fluids with Caco-2 Cells and Rat Ileum. *Drug Dev Ind Pharm* 32: 151-161. [CrossRef]

• Tang L, Khan SU, Muhammad NA (2001). Evaluation and selection of bio-relevant dissolution media for a poorly water-soluble new chemical entity. *Pharm Dev Technol* 6: 531-540. [CrossRef]

• Wiedman TS, Liang W, Kamel L (2002). Solubilization of drugs by physiological mixtures of bile salts. *Pharm Res* 19: 1203-1208. [CrossRef]

• Xu H, Vela S, Shi Y, Marroum P, Gao P (2017). In vitro characterization of ritonavir drug products and correlation to human in vivo performance. *Mol Pharm* 14: 3801-3814. [CrossRef]

• Yazdanian M, Briggs K, Jankovsky C, Hawi A (2004). The high solubility definition of the current FDA guidance on biopharmaceutical classification system may be too strict for acidic drugs. *Pharm Res* 21: 293-299. [CrossRef]

• Zhou Z, Dunn C, Khadra I, Wilson CG, Halbert GW (2017). Statistical investigation of simulated fed intestinal media composition on the equilibrium solubility of oral drugs. *Eur J Pharm Sci* 99: 95-104. [CrossRef]