Cytogenetic studies of Brazilian pediatric myelodysplastic syndrome cases: challenges and difficulties in a large and emerging country

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Abstract

Myelodysplastic syndromes (MDS) and juvenile myelomonocytic leukemia (JMML) are rare hematopoietic stem cell diseases affecting children. Cytogenetics plays an important role in the diagnosis of these diseases. We report here the experience of the Cytogenetic Subcommittee of the Brazilian Cooperative Group on Pediatric Myelodysplastic Syndromes (BCG-MDS-PED). We analyzed 168 cytogenetic studies performed in 23 different cytogenetic centers; 84 of these studies were performed in patients with confirmed MDS (primary MDS, secondary MDS, JMML, and acute myeloid leukemia/MDS+Down syndrome). Clonal abnormalities were found in 36.9% of the MDS cases and cytogenetic studies were important for the detection of constitutional diseases and for differential diagnosis with other myeloid neoplasms. These data show the importance of the Cooperative Group for continuing education in order to avoid a late or wrong diagnosis.

Key words: Childhood myelodysplastic syndromes; Juvenile myelomonocytic leukemia; Cytogenetics; Karyotype; Diagnosis

Introduction

The Brazilian Cooperative Group on Pediatric Myelodysplastic Syndromes (BCG-MDS-PED) was founded in January 1997 as a working group of hematologists, pediatric hemato-oncologists, molecular biologists, and other related professionals with the following objectives: to provide support for diagnosis, to offer treatment advice and educational instructions, to perform studies and research, as well as to share experiences about cases of pediatric (age <18 years) myelodysplastic syndromes (MDS). The BCG-MDS-PED consists of eight distinct committees: morphology (cytology, histopathology, and immunophenotyping); genetics (cytogenetics, molecular biology, and clinical genetics); epidemiology; secondary MDS; Down syndrome (DS); bone marrow failures (congenital, acquired, and after infections); myeloproliferative disorders and therapeutics, in order to cover all aspects of MDS. We report here the experience of the Cytogenetic Subcommittee.

Material and Methods

Since January 1985, 350 cases have been referred to the BCG-MDS-PED, coming from 68 different public or private centers, specialized or general, and located in 19 of the 26 states of Brazil. Some cases were included retrospectively before the BCG-MDS-PED was founded. The morphology committee reviewed peripheral blood/ bone marrow smears and biopsies and classified
the cases according to the WHO-Pediatric-Hasle’s classification (1).

Karyotypes were obtained at local centers when available and reviewed by the Cytogenetic Committee (EDRP, MLC, SRCT) or at one of the four university centers in São Paulo. Karyotyping was performed in bone marrow samples using the Giemsa banding technique and chromosomal abnormalities were described according to the International System for Human Cytogenetic Nomenclature (ISCN 2009) (2). Chromosome fragility was induced with the diepoxybutane (DEB) test in suspected cases of Fanconi anemia. Some cytogenetic centers performed FISH studies for monosomy 7 in cases with less than 20 metaphases analyzed and BCR/ABL1 rearrangements as a differential diagnosis between juvenile myelomonocytic leukemia (JMML) and chronic myeloid leukemia (CML).

After the review, 156 cases were confirmed as childhood MDS (de novo or secondary), JMML, or acute myeloid leukemia/MDS with DS (AML/MDS-DS), 129 cases did not meet the diagnostic criteria and 65 cases had no definitive diagnosis (in revision, inconclusive or insufficient sample).

For the 82 MDS patients with cytogenetic studies and evolution, we calculated overall survival and survival according to the cytogenetic profile (normal, chromosome 7 abnormality and other abnormalities). The same evaluation was performed for the 65 patients with well-known therapy, censoring the time of allogeneic hematopoietic stem cell transplantation (HSCT) (performed in 13 of them). Statistical analysis was performed using the Kaplan-Meier product limit estimation (survival curves) and log rank test to compare groups. The data were analyzed using the SPSS software.

Results

Of the 350 cases included in the BCG-MDS-PED database, 168 had been submitted to conventional cytogenetic studies (bone marrow karyotype) and 76 were reviewed by the committee. At least 23 cytogenetic centers located in 7 different Brazilian States, most of them in São Paulo (capital and interior of the state) were responsible for the study. Figure 1 shows the origin of all 350 patients and the distribution of the cytogenetic centers.

Although the number of cases with karyotype studies was small, we observed an increase in the percentage of cases with cytogenetics according to the period of diagnosis: 8.3% (1985 to 1990), 29.4% (1991-1995), 49.3% (1996-2000), 56.6% (2001-2005), and 50% (2005-2010). The abnormal cytogenetic studies according to final diagnosis are shown in Table 1.

Childhood MDS was confirmed in 84 cases who had conclusive cytogenetic studies (5 cases showed no metaphases), including 11 with refractory cytopenia (RC), 48 with refractory anemia with excess of blasts (RAEB) or RAEB in transformation (RAEB/T), 4 with unclassified MDS, 11 with JMML, 7 with AML/MDS-DS, and 3 with secondary MDS. Results were abnormal in 45.2% of cases (38/84). Clonal abnormalities were found in 31 (36.9%) cases, 4 of them also with constitutional trisomy 21. Seven cases (8.3%) showed only constitutional abnormalities, including 4 cases with trisomy 21, 1 with balanced translocation t(8;9) and 2 with a positive DEB test.

Monosomy 7/deletion 7q was the most frequent clonal abnormality, found in 11 cases (13.9%), followed by trisomy 8 (4.7%) and 21; 2 or more abnormalities were seen in 5.9% of cases. Clonal abnormalities were more frequent in AML/MDS-DS (42.8%), RAEB/T (37.5%), JMML (36.4%), and secondary MDS (33.3%) than in RC (27.2%). In AML/MDS-DS, trisomy 8 and an extra copy of an isodisomy of chromosome 21 were found as clonal abnormalities in 2 cases and 1 case, respectively.

The median overall survival was 31 months for the MDS group (n = 82 patients), 122 months for the subgroup with chromosome 7 abnormality (n = 11 patients), 35 months for the subgroup with abnormal karyotype but no chromosome 7 abnormality (n = 19 patients), and 29 months for normal karyotype (n = 52 patients). No significant differences were detected in the survival of these 3 groups (P = 0.4; Figure 2A). The median survival obtained by censoring the time of autologous HSCT was 29 months for the MDS group (n = 65 patients), a time not reached by the subgroup with chromosome 7 abnormality (n = 8 patients), 36 months for the subgroup with abnormal karyotype but no chromosome 7 abnormality (n = 17 patients), and 28 months for patients with a normal karyotype (n = 40). Again, no significant differences were detected in the survival of these 3 groups (P = 0.4; Figure 2B).

Of 129 cases reviewed by the Morphology Committee who did not show childhood MDS, 53 had cytogenetic studies (but 4 cases showed no metaphases), 15 (30.6%) showed karyotype abnormalities, 1 of them characterized as constitutional in one boy with aplastic anemia who showed a balanced translocation involving the long arm of chromosome 3 and short arm of chromosome 5. Eight cases were diagnosed as AML, 3 with t(8;21), 2 with trisomy 8, one with del(11q23), and another with t(15;17). Karyotyping confirmed 3 cases of CML with Philadelphia chromosome, and 1 case of primary myelofibrosis with additional material on chromosome 22.

An interesting t(1;5)(q23;q33) translocation was detected in an 11-month-old girl with anemia, thrombocytopenia, leukocytosis with monocytosis, and marked eosinophilia. She was first diagnosed by our Committee as having unclassified MDS or MDS/myeloproliferative disorder. In cooperation with the Dana-Farber Cancer Institute and Harvard Medical School, the t(1;5)(q23;q33)
translocation was cloned, showing the fusion of the platelet-derived growth factor receptor beta (PDGFRB) gene to the myomegalin gene. After imatinib therapy, the patient achieved a long-term hematological remission, and is still alive, more than 10 years after the diagnosis (3). This disease is now called myeloid and lymphoid neoplasm with eosinophilia and abnormalities in the PDGFRB gene, according to the WHO 2008 classification (4).

Discussion

Childhood MDS and JMML are a group of clonal hematopoietic disorders occurring in 1.2 and 1.8 children per million per year, respectively (1). Cytogenetics plays an important role in the diagnosis of these rare diseases. The incidence of clonal cytogenetic abnormalities in childhood MDS ranges from 50 to 80% and monosomy 7/deletion 7q was the most frequent abnormality described in Europe (5,6) and in the USA (7). Trisomy 21 (inherited or acquired) was also common, but trisomy 8 and 5q deletions (often described in adult MDS) were detected in only a few cases (7). The European Working Group of Myelodysplastic Syndrome in Childhood (EOWG-MDS group) demonstrated that 36% of JMML cases showed clonal abnormalities, and monosomy 7 was detected in 26% of the cases (8). The role of cytogenetics in the prognosis of childhood MDS was published in few studies. In 1995, Passmore et al. (9) proposed a score that included platelet counts and fetal hemoglobin; but it was not validated by others (7). The French group

Figure 1. Map of Brazil showing the distribution of 350 patients included in the Brazilian Cooperative Group on Pediatric Myelodysplastic Syndromes (BCG-MDS-PED) database (1987-2007). Most of the cytogenetic centers are concentrated in the Brazilian states of the South and Southeast.
Table 1. Cytogenetic abnormalities in 52 patients with confirmed myelodysplastic syndromes and with other diagnoses.

| Diagnosis    | Karyotype studies                                      | Complementary cytogenetic tests |
|--------------|--------------------------------------------------------|---------------------------------|
| RC           | 40~46,XY,del(7)(q32)[6]/46,XY[7]                        |                                 |
| RC           | 45,XY,-7[20]                                           |                                 |
| RC           | 46,XX,del(p1.1)[6]/47,XX,-q(3)[4]                      |                                 |
| RAEB         | 45,XX,-7[7]/46,XY[7]                                   | FISH positive for monosomy 7    |
| RAEB         | 45,XX,-7[5]/46,XY[2]                                   | FISH positive for monosomy 7    |
| RAEB         | 45,XX,-7[10]/46,XX[5]                                  | FISH positive for monosomy 7    |
| RAEB         | Absence of metaphases                                  | FISH positive for monosomy 7    |
| RAEB         | 46,XX,del(12)(p?)[6]/47,XX,-q(3)[4]/46,XX[5]           |                                 |
| RAEB         | 46,XY,+8(20)/46,XY[20]                                 | positive DEB test               |
| RAEB/T       | 45,XX,del(7)[10]                                       |                                 |
| RAEB/T       | 47,XX,+8[10]/46,XY[10]                                 |                                 |
| JMML         | 45,XY,-7[20]                                           |                                 |
| JMML         | 45,XY,-7[17]                                           |                                 |
| JMML         | 46,XY,add(9)(p?)[4]/46,XY[8]                           |                                 |
| JMML         | 47,XY,+21[20]                                          |                                 |
| U-MDS        | 47,XY,+mar[2]/46,XY[8]                                 |                                 |
| Secondary MDS| 46,XX,del(21)(q?)[2]/46,XX[9]                         |                                 |
| MDS/AML+DS   | 48,XY,der(21;21)(q10;q10)c,+der(21;21)(q10;q10),+21[3] |                                 |
| MDS/AML+DS   | 47,XY,der(21;21)(q10;q10)c,+21[14]                     |                                 |
| MDS/AML+DS   | 48,XX,+8,+21c[6]/47,XX,+21c[2]                         |                                 |
| MDS/AML+DS   | 46,XY,add(7)(p22),-17,+21c[20]                         |                                 |
| MDS/AML+DS   | 48,XX,+8,+21c[4]/47,XX,+21c[2]                         |                                 |
| MDS/AML+DS   | 47,XX,+21c[20]                                         |                                 |
| MDS/AML+DS   | 47,XX+21c[20]                                          |                                 |
| MDS/AML+DS   | 47,XX+21c[20]                                          |                                 |
| MDS/AML+DS   | 47,XY+21c[20]                                          |                                 |
| AML          | 45,XY,-del(7)(q32?),t(8;21)[q22;q22][20]                |                                 |
| AML          | 46,XY,add(3)[q28],t(8;21)[q22;q22],del(9)(q?21)[12]/46,XY[1] |                                 |
| AML          | 46,XY, t(15;17)[q22;q21][20]                           |                                 |
| AML          | 46,XX,del(11)(q23)[5]/46,XX[21]                        |                                 |
| AML          | 45,XY,-7[15]                                           |                                 |
| AML          | 45,XY,-7[11]                                           |                                 |
| AML          | 46,XX,del(7)(q22)[12]/46,XX[2]                         |                                 |
| AML          | 47,XX,+8[30]                                           |                                 |
| Aplastic anemia| 46,XY,t(3:5)(q25;p15)c[20]                           |                                 |
| CML          | 46,XX, t(9;22)[q34.1;11.2][20]                         |                                 |
| CML          | 46,XX, t(9;22)[q34.1;11.2][20]                         |                                 |

Continued on next page
studying 44 children with MDS showed that monosomy 7 was associated with intermediate prognosis (Groupe Français de Cytogénétique Hématologique, 1997) (5). Monosomy 7 was not associated with unfavorable prognosis in JMML by the EOWG-MDS group (8). In 2010, the EOWG-MDS group studied prospectively 192 children with advanced MDS and identified that the presence of a structurally complex karyotype was the strongest independent prognostic marker predicting poor outcome, with only 14% of a 2-year survival in this group (10).

Before the founding of the BCG-MDS-PED in 1997, there were almost no studies describing the incidence, clinical evolution or transformation of childhood MDS in Brazil. At first, the idea of this cooperative group was to review cases of the entire country and give diagnostic support (morphology, genetics, etc.). In parallel, a teaching initiative was designed in order to disseminate the concepts and tools for a correct and quick MDS diagnosis in children. The results of these initiatives are shown here. More than 350 cases were reviewed, 156 of which were correctly classified as MDS according to the WHO pediatric approach. Morphology, histology, and cytogenetic analysis were reviewed by a central committee and the evaluation was confirmed. As expected, most of these cases (113/157) were de novo MDS and a few were secondary. Also, AML and JMML were well defined, as well as DS cases.

Table 1. Continued.

| Diagnosis       | Karyotype studies                                                                 | Complementary cytogenetic tests                          |
|-----------------|-----------------------------------------------------------------------------------|----------------------------------------------------------|
| CML             | 46,XX,del(4),t(9;22)(q34.1;q11.2)[20]                                              | FISH positive for BCR/ABL1 rearrangement                 |
| Myelofibrosis   | 46,XX,add(22)(q13)[4]/46,XX[7]                                                     |                                                          |
| MLN PDGFRB      | 46,XX,t(1;5)(q23;q33)[24]                                                         | FISH positive for PDGFRB rearrangement                    |

AML = acute myeloid leukemia; CML = chronic myeloid leukemia; DEB = positive diepoxybutane test; FISH = fluorescence in situ hybridization; JMML = juvenile myelomonocytic leukemia; MDS/AML+DS = myelodysplastic syndrome/AML associated with Down syndrome; MLN PDGFRB = myeloid and lymphoid neoplasia with eosinophilia and abnormalities in the PDGFRB gene; RC = refractory cytopenia; RAEB = refractory anemia with excess of blasts; RAEB/T = RAEB in transformation; U-MDS = unclassified MDS.

Figure 2. Kaplan-Meier overall survival curves for patients with myelodysplastic syndromes according to the cytogenetic profile and to chromosome 7 abnormalities vs other abnormalities vs normal. A, Overall survival. B, Survival with censoring of the time of allogeneic hematopoietic stem cell transplantation.

Brazil is a huge country, with a territorial extent of 8,511,996.3 km², with more than 5500 cities and 180 million inhabitants, most of them concentrated in the Southwest region, which is also the most economically
rich and with an estimated adequate number of physicians/inhabitants. As shown, most of the MDS cases came from São Paulo State (Southeast). In Northern or Central states of Brazil not even a single MDS case was detected, a fact that raises concern about the possibility that cases may still be misdiagnosed in these regions. This indicates the need for continuing education and awareness of pediatricians regarding the different aspects of disease manifestation in order to avoid a late or wrong diagnosis. Very interesting lessons can be drawn from this collaborative study: the joint effort brought benefits to patients, relatives and physicians as well as some social, scientific, and economical profits.

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