Chapter from the book *Thyroid and Parathyroid Diseases - New Insights into Some Old and Some New Issues*

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Algorithm 1. Flowchart of the proposed method

1. Initialize the network with random weights.
2. For each input pattern:
   a. Forward propagate the input through the network.
   b. Compute the error between the output and the target.
   c. Backpropagate the error through the network.
   d. Update the weights using an appropriate optimization algorithm.
3. Repeat steps 2 until convergence is achieved.

Result: A trained feedforward neural network.
proliferation, differentiation and also stimulating the development of neuronal processes, axons and dendrites. As well TH mediate effects on CNS occur throughout life (Bauer et al., 2002a; Smith et al., 2002). Different studies have demonstrated the presence of thyroid receptors in rat CNS with a particular distribution during development and adulthood (Bradley et al., 1992; Bradley et al., 1989). Thus, TH regulate the expression of genes implicated in myelination, neuronal and glial cell differentiation (Bernal, 2005; Bernal & Nunez, 1995) and neuronal viability and function (Smith et al., 2002). These hormones are able to modify cell morphology by acting on cytoskeleton machinery required for neuronal migration and outgrowth (Aniello et al., 1991; Morte et al., 2010). Additionally, TH are present in noradrenergic nuclei of CNS (Rozanov & Dratman, 1996) probably acting as neuromodulator or co-neurotransmitter (Dratman & Gordon, 1996). In line with this, TH increase β-adrenergic receptors levels (Ghosh & Das, 2007; Whybrow & Prange, 1981) and improve both cholinergic (Smith et al., 2002) and serotonin neurotransmission in animals (Bauer et al., 2002a). The effect of TH on serotonin (5 HT) has been explained by a desensitization of 5HT1A autoreceptor in the raphe nuclei which probably results in enhancement of firing and release of serotonin from raphe neurons (Heal & Smith, 1988). Furthermore, these hormones can stimulate the expression of neuronal growth factor (NGF) suggesting certain trophic actions on CNS (Walker et al., 1979; Walker et al., 1981).

In mice models, maternal hypo and hyperthyroidism cause some malformation and developmental defects in the cerebellar and cerebral cortex of their newborns. Concomitantly, there is some degeneration, deformation and severe growth retardation in neurons of these regions in both groups (El-Bakry et al., 2010). Therefore, TH play an important role in brain development, neuronal migration and axonal projection to target cells. In vitro and in vivo studies have shown that TH exert a non genomic action over the actin citoskeleton development in astrocytes and neurons. The lack of TH impaire cell growth, granule cells migration and explain those defects in the hypothyroid brain (Farwell et al., 2006; Leonard&Farwell, 1997; Farwell&Dubord, 1996).

Moreover, both acute and chronic Thyroxine treatment in rats increases the cognitive function, probably through an enhancement in cholinergic neurotransmission (Smith et al., 2002).

In humans, TH deficiency during the fetal and postnatal periods may cause irreversible mental retardation, neurological and behavioral deficits, and long lasting, irreversible motor dysfunctions. In adulthood, hypothyroidism may also determine profound behavioral consequences such as depressive symptoms, impaired memory, impairment in learning, verbal fluency, and spatial tasks (Miller et al., 2007; Samuels et al., 2007). Probably these alterations are due to neurotransmission impairment in brain areas related to learning and memory, such as hippocampus. Thus, the reduction of TH levels in CNS, can promote an altered neurotransmission activity contributing to some mood disorders like major depression.

The biologically active thyroid hormone T3 exerts its effects by interacting with their specific nuclear thyroid receptors (TRs) that are positively regulated by its own ligand, acting as transcription factors. TRs are encoded for two different genes: TRβ located on chromosome 3, encodes three isoforms: β1, β2 and β3, and TRα located on chromosome 17, encodes the isoforms α1, α2 and α3. TRα1 is expressed early in the embryonic development and TRβ is expressed at later stages of development. By the other hand, the expression of these isoforms
is tissue dependent; in the brain, the main isoforms of TRs are: TR\(\alpha_1\), TR\(\alpha_2\), TR\(\beta_1\) and TR\(\beta_2\). TR\(\alpha_1\) and \(\alpha_2\) accounting for most of TRs in the organ, whereas TR\(\beta_1\) and \(\beta_2\) are detected in only a few areas as retina, cochlea, anterior pituitary and hypothalamus. In mice in which TR\(\alpha\) or TR\(\beta\) were inactivated, different phenotypes are observed indicating that TRs isoforms mediate specific functions but also, they can substitute each other to mediate some actions of T3 (Jones et al., 2007; Forrest et al., 1996a; Forrest et al., 1996b; Wikström et al., 1998; Fraichard et al., 1997; Göthe et al., 1999).

Some studies have reported, in propylthiouracyl-induced hypothyroidal adult rats, a decreased expression of TR\(\alpha_1\) and TR\(\beta\) in the hippocampus, associated with an increase in \(\beta\)-amyloid peptides in the same area. Hypoactivity of the thyroid signaling in the hippocampus could induce modifications in the amyloidogenic pathway and this could be related with a greater vulnerability of developing Alzheimer disease in hypothyroidal subjects (Ghenimi et al., 2010).

2.1 T3 generation in the central nervous system: The importance of deiodinases

Although both forms of TH (T4, T3) are present in the circulating blood, some studies have demonstrated that T4 is transported into the brain much more efficiently than T3 (Hagen & Solberg, 1974). In contrast to peripheral tissue, in the brain T4 and T3 are in equimolar range indicating mechanisms for an efficient transformation into biological active hormone.

TH production is regulated by the HPT axis, while its biological activity is mainly regulated by three selenodeiodinasas coded by different genes (D1, D2, D3). Deiodinases act at prereceptor level influencing both, extracellular and intracellular TH levels and its action. Whether it activates or inactivates it, will depend on the level where deiodination occurs (5 or 5\(^{\prime}\) position on the iodothyronine molecule). In the periphery, in the kidney and liver, D1 isoform is responsible for the production of most of the circulating T3 (Bianco et al., 2002).

In the CNS, the most important isoforms are D2 and D3. In the brain, T3 is produced locally by the action of D2 which is also expressed in pituitary, thyroid, brown adipose tissue, skeletal muscle, and aortic smooth muscle cell, in humans. D2 activity varies extensively in different brain regions, with the highest levels found in cortical areas and lesser activity in the midbrain, pons, hypothalamus and brainstem (Bianco et al., 2002; Gouveia et al., 2005; Zavacki et al., 2005). It has been described in adult rats, that approximately 80% of T3 bound to nuclear receptors is produced locally by D2 activity (Crantz et al., 1982). Moreover, inactivation of TH is mainly carried out by D3 as well as glucuronosyltransferase and sulfotransferases. D3 is highly expressed within the CNS, with low peripheral expression. D3 degrades T\(4\) to rT\(3\) and T\(3\) to 3,3\(^{\prime}\)-diiodothyronine (T2) therefore preventing or finishing actions of T3. Thus, combined actions of D2 and D3 can locally increase or decrease thyroid hormone signaling in a tissue -and a temporal- fashion, and more importantly in a way independent of thyroid hormone plasma levels. In addition, increasing evidences pointed out that deiodinase expression can be modulated by a wide variety of endogenous signaling molecules, suggesting a local modulation of T3 production in the brain (Gereben et al., 2008a, Gereben et al., 2008b). D2 enzymatic activity is increased also in hypothyroidism and decreased in hyperthyroidism (Kirkegaard & Faber, 1998).
2.2 Association between deionidase polymorphisms and thyroid hormone metabolism

Genetic variations in deionidase genes may impact significantly thyroid function and TH levels in euthyroid subjects (Hansen et al., 2007; Peeters et al., 2007; Peeters et al., 2006; Peeters et al., 2003). The effect of two polymorphisms in D1 gene, D1-rs11206244 (D1-C785T) and D1-rs12095080 (D1-A1814G) on thyroid hormone metabolism has been evaluated in randomly selected subjects (Peeters et al., 2003). The allele T of D1-rs11206244 was associated with high levels of rT3 and high rT3/T4 ratio and a low T3/rT3 in plasma; whereas the G allele of D1-A1814G was associated with a high T3/rT3 (de Jong et al., 2007; Peeters et al., 2003). These results suggest a lower activity in T carriers of rs11206244 than G carriers (Peeters et al., 2003).

Of special interest is the common polymorphism in humans: D2 rs225014 (D2-Thr92Ala), characterized by a threonine (Thr) change to alanine (Ala) at codon 92 (D2 Thr92Ala). It is associated with insulin resistance in different populations, suggesting that D2-generated T3 in skeletal muscle plays a role in insulin sensitivity (Mentuccia et al., 2002, Canani et al., 2005). The minor allele (G) is associated with a low D2 activity in thyroid samples obtained from patients (Canani et al., 2005). In accordance, G allele seems to predict the need for higher T4 intake in thyroidectomized patients (Torlontano et al., 2008). Nonetheless, it has been observed that GG subjects show a delayed serum T3 rise in response to TRH-mediated TSH secretion consistent with decreased D2 activity (Butler et al., 2010). Some studies have described a naturally occurring polymorphism located in 5'-untranslated region of the D2 gene (Coppotelli et al., 2006). In healthy blood donors, the minor allele of this polymorphism (D2-ORFa-Asp variant, rs12885300) is associated with an increase in circulating T3/T4 ratio but not with plasma T3 and TSH levels, suggesting an increased D2 gene expression (Peeters et al., 2005). In agreement, in vitro studies suggested that D2-rs1288530 polymorphism leads to higher activity of D2 at the pituitary level (Coppotelli et al., 2006). In a long case-control Chinese study, the haplotypes ORFa-3Asp-92Ala and ORFa-3Gly-92Ala indicated higher susceptibility for bipolar disorders, while ORFa-3Asp-92Thr probably played a protective role (He et al., 2009). According to this evidence, it is feasible that variants of D2 gene can produce “brain hypothyroidism” limiting T3 action on CNS affecting brain neurotransmission.

3. Thyroid and depression

The similarity and overlapping between symptoms of depression and thyroid disorders has been the theoretical base for the hypothesis regarding a possible relationship between both entities. As we mention above, hypothyroidism could induce cognitive dysfunction and depressive symptoms besides psychological distress in a very similar way to primary depression (Constant et al., 2005; Bould et al., 2011; Mowla et al., 2011). Likewise, TH effect as augmentation therapy in refractory depression, and thyroid disorders as risk factors for rapid-cycling in bipolar disorder sustain a possible association between both types of diseases.

The involvement of HPT axis in the pathogenesis of depression is supported by multiple data. There are few studies that show normal range TH levels during a depressive episode; however most of them demonstrate diverse changes in different hormones associated with this axis. Concerning TSH levels, data are contradictory, some authors have reported a decrease in basal TSH values as well as in those observed in response to exogenous TRH.
(Forman-Hoffman & Philibert, 2006; Stipcević et al., 2008) and other studies showed TSH elevation in bipolar depression (Brouwer et al., 2005; Saxena et al., 2000).

In reference to T3 levels, results are more conclusive, showing a trend to decrease in the presence of depression, as well as an association with high risk of long term relapse. In addition there seems to be a more pronounced T3 decrease in direct relation with the severity of depression (Stipcević et al., 2008; Saxena et al., 2000). Reported T4 levels in depression are also contradictory, since there is evidence showing a rise as well as a decrease of T4 during depressive episodes. (Saxena et al., 2000; Kirkegaard&Faber, 1998). In a study, with more than 6,000 subjects, it was shown that a low TSH and a high T4 levels were associated with depression specially in young men but, in women only a higher T4 levels correlated with current depression syndrome (Forman-Hoffman&Philibert, 2006). It is possible that these findings could be explained by a diversity of factors, such as differences in phenotypes of depressive patients, severity and duration of the disease, difficulties in isolating drugs effects in TH levels (antidepressants and mood stabilizers) and probably, gender and other differences.

Overt thyroid disease is infrequent among depressive patients. Nevertheless, many authors have seen that a subgroup of depressive patients manifest a subclinical hypothyroidism and this might be a negative prognostic factor (Fountoulakis et al., 2006). On the other side, some antidepressants as lithium inhibits TH secretion and could increase antithyroid antibodies, promoting hypothyroidism in susceptible subjects (Emerson et al., 1972; Myers et al., 1985).

There is still no hypothesis that can satisfactorily integrate these data. Interactions between TH and neurotransmitters, gene expression and neurohormonal receptors are not clear yet. For instance, 5 HT seems to inhibit TRH secretion and somatostatin TSH secretion (Kirkegaard&Faber, 1998); both of them are reduced in cerebro spinal fluid (CSF) in patients with psychiatric illness and affective disorders (Gerner&Yamada, 1982, Roy-Birne et al., 1983; Rubinow et al., 1983). Otherwise, T3 influx to intracellular level in the brain is determined by many factors, including T3 and T4 circulating levels, protein transporters, and deiodinase activity.

About a 25% of major depressed patients show a reduction in TSH release under TRH stimulation (Loosen 1985, Risco et al, 2003). It has been proposed that in them exist a blunted response due to the raise of circulating cortisol, associated to hypothalamic-pituitary-adrenal axis hyperactivity. This response has also been observed in bipolar disorders (Linkowsky et al., 1981). On the other hand, in rapid cycling depressives, TSH hypersecretion is observed in response to TRH (20% of basal TSH levels above the normal range) (Szabadi, 1991, Larsen et al., 2004).

Nevertheless, as we mentioned before, the mechanism by which TH affect the adult brain is not completely clear, because the complex interactions between neurotransmitters and thyroid. One hypothesis is that TH modulate the number of post-synaptic β-adrenergic receptors in the cerebral cortex and cerebellum. This could be relevant considering the influence of catecholamines deficit, mainly norepinephrine as a cause of depression (Atterwill et al., 1984). Another possible mechanism is the modulation of 5 HT and its receptors. It has been suggested that TH inhibit the impulse rate of neurons present at the raphe and reducing the release of 5HT. T3 administration to mice attenuates the function of 5HT1A and 5HT1B receptors, increasing the cortical and hippocampus synthesis and
Turnover of 5-HT. Administration of T3 plus electroconvulsive shock markedly potentiated its actions on 5-HT2-mediated responses. (Heal&Smith, 1988). These findings provide evidences for possible antidepressant effects of T3 and/or potentiating therapy by TH. This issue is relevant in patients suffering depressive disorders, related with reduction in monoamine neurotransmission such as serotonin (reviewed in Belmaker&Agam 2008).

A positive correlation between serotonin levels and circulating T3 has been described in humans. Indirect evidences showed that brain serotonin is increased in hyperthyroidism and decreased in hypothyroidism (Singhal et al., 1975). In the last situation, this is reversed with TH replacement (Bauer et al., 2002b, Strawn et al., 2004). In depressed subjects, the decrease in serotoninergic tone could be related to lower brain T3 levels, perhaps due to a reduction of deiodinases activity. Furthermore, an imbalance in T3 conversion could account for depressive disorder and/or clinical outcome to antidepressants therapy. It has been suggested that in depression, T3 may favor the release of cortical 5-HT and thus synergize the response to antidepressants. Administration of desipramine a selective serotonergic reuptake inhibitor (SSRI) in rats, induces an increase of D2 activity and T3 concentration in cortical tissue. Interestingly, T4 concentrations were significantly lowered after administration of the antidepressant but, serum T3 levels were significantly reduced only after toxic dosis of desipramine. Other commonly used SSRI, fluoxetine also decreases D3 activity (Eravci et al., 2000). Based on these data, one might suggest that depression occurs by the inhibition of D2, determining decreased T3 levels and secondarily, reduced levels of brain 5HT.

The efficacy of T3 as a supplement of sertraline therapy, another SSRI, was studied recently in relation D1 polymorphism (Cooper-Kazaz et al., 2009). Patients carrying the T allele of D1-rs11206244 showed a significant response to 8 week of antidepressant treatment in comparison with non-carriers of the allele. Additionally, there was no effect of T allele on sertraline response, suggesting that the polymorphism is not associated to antidepressant effect (Cooper-Kazaz et al., 2009). As we mentioned, the T allele of D1-rs11206244 showed lower T3 and higher rT3 than non-T carriers (de Jong et al., 2007; Peeters et al., 2003). Thus, it seems that patients genetically characterized by poor conversion of T4 to T3, are better responders to T3-antidepressant co-treatment (Cooper-Kazaz et al., 2007; 2008). Another study evaluated whether baseline thyroid function and D2 rs225014 (D2-Thr92Ala) predict response to paroxetine. It showed that high TSH levels predict the response, and heterozygous patients showed lower TSH levels than the wild-type allele (A) (Brouwer et al., 2006). However, up to date there is no study evaluating the influence of T3 and D2 polymorphisms on antidepressant response.

Based on these observations, we evaluated the presence of D2 polymorphism related with a lower activity of the enzyme: D2-Thr92Ala (T/C). The polymorphism was analyzed in 61 euthyroid patients with depression and 48 subjects of a population sample using the PCR-RFLP method. Clinical response to fluoxetine was evaluated before and after 8 weeks of treatment, using Hamilton Scale for Depression (HAM-D). We found that the CC genotype of Thr92Ala polymorphism was more frequent in depressed subjects and in non-responders patients (unpublished data). We concluded that Thr92Ala polymorphism of D2 gene could be considered a predictive marker of clinical response to fluoxetine, and hence of pharmacological therapy, but more studies are needed to confirm this preliminary results.

The presence of these polymorphisms could influence basal activity of type 2 deiodinase, and therefore of T3 bioavailability in the brain.
4. Use of thyroid hormone in depression

Several studies using thyroid hormones in the management of patients with mood disorders have been reported since the early seventies. TH have been used in euthyroid depressed patients to enhance the effects of antidepressants. In patients receiving electroconvulsive therapy, those treated with T3 required less sessions and presented less memory loss compared with placebo treated group (Stern et al., 1991). T3 has been employed in initial combination therapy, and T3 or T4 in refractory depression or non responder patients. T3 in doses of 20 to 50 µg is able to enhance the effect of tricyclic antidepressants and shorten the depression period but, many studies have not demonstrated differences in the number of patients recovered (Prange et al., 1969; Wilson et al., 1970; Coppen et al., 1972; Wheatley, 1972). A meta-analysis showed that when T3 was used in refractory depression in addition to tricyclic antidepressant therapy, patients treated with it were twice as likely to respond as controls, decreasing depression severity scores (Aronson et al., 1996). However, samples size were small and deserve more evidence. Other studies, using T3 augmentation to SSRI-resistant depression, observed an improvement in mood scores (Agid&Lerer, 2003, Iosifescu et al., 2005, Abraham et al., 2006). Some authors found that patients who responded to T3 had higher serum TSH levels than non-responders and T3 appears to be less effective in men than in women (Agid&Lerer, 2003). Other authors reported that patients with atypical depression experienced significantly greater clinical improvement in final HAM-D with higher rates of treatment response and remission compared to subjects with non-atypical major depressive disorder (Iosifescu et al., 2005). All those cases were treated mainly with fluoxetine in a daily dose of 20 to 40 mg/ and 25-50 µg of T3, with few side effects.

L-thyroxine (T4) added to antidepressants has been used less frequently than T3. Some authors have suggested that T4 augmentation is less effective than T3 (Joffe&Singer, 1990) and that supra physiological doses (250-600 ug/day) are needed, as has been demonstrated in patients with resistant major depression or refractory uni and bipolar disorders (Baumgartner et al., 1994, Bauer et al., 1998, 2002). These results support the theory of a reduced deiodination of T4 compatible with an inhibition of the D2 or a stimulation of the D3 in brain tissues resulting in reduced local T3 concentration.

Nevertheless, the addition of T4 (100 ug /day during 4 weeks) to serotonergic antidepressants obtained remission in 11 of 12 female patients with a resistant depressive episode but, these results did not show association with T3, T4 or TSH levels (Łojko & Rybakowski, 2007).

To date, the use of TH in mood disorders is controversial and the rationale for this therapy is still not completely clear. Main limitations of the studies are: small number of cases, lack of a placebo group, heterogeneity in diagnosis criteria, differences in observational period and in antidepressant therapy. For example, lithium has a known inhibitory effect on TH secretion; fluoxetine has a stimulatory effect over D2 as well as desipramine and both of them could induce deficit of T4.

In this line, we evaluated a group of euthyroidal adult female patients with major depression according to DSM IV-R criteria. All of them were free of antidepressants for at least for 6 month. We studied the effect of adding T3 in a dose of 50 ug per day (n=11) or placebo (n=10), to the standard antidepressant therapy with fluoxetine during 8 weeks. At the end of the observational period final HAM-D scores were similar in both groups. (See Table 1). Patients in T3 group showed significant T4, T3 and TSH changes; but they remain...
clinically euthyroid during the whole treatment period. Their body mass index, heart rate and other clinical parameters did not change. The placebo group showed a non significant increase of THS at the end of the observation time (See Figure 1, unpublished results).

|                | T3 Group | Placebo | p   |
|----------------|----------|---------|-----|
| **Age (y.o.)** | 40±12    | 36±10   | ns  |
| Initial        | 24±4     | 26±6    | <0.0001 |
| 2m             | 8±4      | 7±4     | ns  |
| Initial vs 2m  |          |         |     |

Table 1. Age and Hamilton score (HAM-D) with 21 items, in the groups with T3 addition or placebo. Both initial and 2 months means±SD were similar (using non paired t student test). The difference between initial and 2 month was highly significant in both groups (using paired t tests).

Fig. 1. TSH changes after addition of T3 or placebo in both groups. Measurements of TSH are shown at baseline, 1 month and 2 months using similar SSRI therapy. T3 hormone induced significant decreased TSH levels. No significant change was observed in placebo group.

Summarizing, our results suggest that TH addition to SSRI therapy in euthyroid depressed patients is safe and has not deleterious clinical effects in spite of TSH changes during treatment. Although, we could not demonstrated in this particular group, a significant antidepressant effect.
5. Hypothyroidism, depression and brain imaging

Single-photon tomography (SPECT), positron emission tomography (PET) and functional magnetic resonance imaging (fMRI) are able to capture physiological events linked to underlying neuronal activity. They have been employed to image and quantify brain perfusion, flow and metabolism in several conditions as well as the radionuclide techniques have been used to map neurotransmitters, receptors, drug actions and many metabolic pathways. Functional imaging in mood disorders may show abnormalities at different brain levels that could normalize with therapy. Several serotonin and adrenergic markers have also been employed to study negative emotional stimuli response in mood disorders. For instance: thalamic activity was increased by reboxetine, whereas citalopram primarily affected ventrolateral prefrontal regions. It would be interesting to have a method able to predict therapy responses to either noradrenergic or serotonergic antidepressants (Carey et al., 2004; Navarro et al., 2004; Zobel et al., 2005; Kohn et al., 2007; MacQueen, 2009; Brühl et al., 2011).

It is also known that even mild hypothyroidism may produce changes in brain regions modulating attention, motor speed, memory and visual-spatial processing. In severe hypothyroidism induced by thyroidectomy in cancer patients, it have been reported a clear parietal and partial occipital lobe hypoperfusion, measured with SPECT; the abnormalities improved after reaching normal thyroid function, in some subjects. However, fluorodeoxyglucose (FDG) and oxygen-15-labeled water studies, in similar patients, showed lower global brain glucose metabolism and flow. Hypothyroidal patients were also significantly more depressed, anxious and psychomotor slowed than euthyroidal subjects (Nagamachi et al., 2004; Constant et al., 2001).

Brain metabolism and flow are usually decreased in major depression and bipolar disease being metabolism inversely associated with the severity of depression. Changes are variable and as we mentioned earlier, could reverse with adequate therapy. Subgenual prefrontal cortex presents abnormal blood flow and metabolism in the depressed state. Prefrontal cortex and limbic structures are involved in emotion regulation and amygdala is involved in emotional memory formation (Buchsbaum et al., 1997; Kennedy et al., 2007; Chen et al., 2011). In major depression patients, glucose metabolism in orbitofrontal and inferior frontal cortex correlates with therapy response; responders have a significant decrease in the orbitofrontal and ventrolateral regions compared with non-responders, implicating ventral prefrontal subcortical circuits in response to specific therapy with SSRI. In major depression and bipolar patients, FDG has shown an inverse correlation between brain metabolism and circulating TSH (Brody et al., 1999; Marangell et al., 1997; Milak et al., 2005).

Cerebral fMRI has been reported to be helpful in major depression intending to predict therapy response using brain activation. Morphometric studies have evaluated hippocampus volume association with response to treatment. Patients who remit have larger pretreatment hippocampus volumes bilaterally compared with those who do not remit. There are similar preliminary findings for the anterior cingulate cortex. A recent work demonstrated a significantly thinner posterior cingulate cortex in non-remitters than in remitters, and also significant decrease in perfusion in frontal lobes and anterior cingulate cortex in non-remitters compared with healthy controls, at baseline (MacQueen, 2009; Järnum et al., 2011).

There are reports with increased perfusion in anterior cingulate and prefrontal medial cortex when using SSRI or amesergide. Responders and non-responders to cognitive behavior therapy versus antidepressive pharmacotherapy and deep brain stimulation could also be
differentiated using brain perfusion SPECT or glucose metabolism with PET (Vlassenko et al., 2004; Kennedy et al., 2007; Richieri et al., 2011).

Another work with fMRI demonstrated also that successful paroxetine treatment decreases amygdala activation, presumably by improved frontolimbic control, in line with SSRI, induced increased functional connectivity between pregenual anterior cingulated cortex, prefrontal cortex, and amygdala. Changes in amygdala activation when processing negative faces expressions might serve as an indicator for improved frontolimbic control required for clinical response (Ruhé, 2011).

We recently studied a group of major depression middle age patients using brain perfusion SPECT, all in their first episode of major depression and /or without any specific therapy for at least six months. Their initial HAM-D scores corresponded to 24±4.8; all of them received standard SSRI therapy. Ninety-three percent were responders at 2 months (HAM-D decrease >50%) and 59% were remitters (HAM-D score ≤5). There was association of decreased perfusion in diverse brain areas with HAM-D changes in the whole group using Statistical Parametric Analysis (SPM) as covariate (See Figure 2). We did not observe significant neocortical perfusion change after 2 months of standard dose of fluoxetine therapy. However, there was a bilateral decrease in parahippocampal gyrus, thalamus and striatum as well as in anterior cingulate gyrus (Brodmann 32 area) after SSRI therapy. No significant difference was observed between remitters and non-remitters.

Fig. 2. In the whole group, SPM8 analysis demonstrated association between decreased perfusion and HAM-D scores considered as a covariate, at baseline and after 2 months of therapy (non corrected p <0.001): -at left: in amygdala, anterior cingulate, globus pallidum, putamen and Brodmann area 9 (mid frontal gyrus) -bilaterally: both hyppocampal gyrus, mid and superior temporal and insulas and cerebellar hemispheres -at right: in central and supramarginal gyrus
As we mentioned before, a half of the women in our group received T3 in addition of SSRI and the other half a placebo instead of T3. Our results showed no evidence that adding T3 to SSRI therapy in unipolar major depression females produces significant change in regional cerebral blood flow at neocortical level (See Figure 3). Only a small difference was found at deep structure level that could imply diverse brain mechanism involved [data not published].

These findings are in agreement with other reports showing relative normalization of perfusion and metabolism that were abnormally increased at baseline in patients with mood disorders. Some of these regional metabolism changes are correlated with emotional behavior. The amygdala and limbic structures have been associated with face recognition and emotional processing. It is well known that there is increased perfusion and metabolism in specific brain areas, reflecting molecular abnormalities in neurotransmitter systems. The development of new molecular imaging methods could help in the individualization of antidepressant therapies (Chen et al., 2011).

![Image](https://www.intechopen.com)

**Fig. 3.** Absence of regional cerebral blood flow change after SSRI therapy in T3 group, using Statistical Parametric Analysis (SPM8) with significant level <0.001; uncorrected p value.

### 6. Conclusions

Depressive and thyroid disorders are important public health problems. There is strong experimental evidence showing thyroid involvement on early stages of CNS development and on metabolic function of the mature brain. It is also accepted that overt hyper or hypothyroidism are not found frequently among mood disorders patients except in those with bipolar disorders, indicating that in most cases the underlying abnormality is at cellular or molecular levels. Although there is a prolific literature on the relationship between thyroid function and depressive disorders, clear results in humans on the role of TH in antidepressant therapy are still lacking. There are no randomized controlled trials, and the number of patients included in existing studies is too small. On the other hand,
more research is needed in order to define the importance of genetic variants in deiodinases and the role of neuroimaging into the complex interactions between HPT function and mood disorders and in clinical response to treatments.

Therefore, considering the available evidence and our own experience, we can recommend this strategy only as an alternative treatment in major depression patients who have failed to respond to other measures.

7. Acknowledgments

The work described in this chapter was partially financed by the grant “Líneas de Investigación Prioritarias” #240/07 HCUCH from the Clinical Hospital of the University of Chile, Santiago, Chile.

The authors thank Dr. Rodrigo Jaimovich (Department of Radiology, School of Medicine, Pontificia Universidad Católica de Chile) who processed and analyzed all SPECT data; Dr. Tamara Galleguillos (Psychiatric Clinic of the Clinical Hospital of the University of Chile) by her collaboration in patient’s recruitment, selection and follow up; Mr. Egardo Caamaño for his technical laboratory support and Ms. Clara Menares who performed the molecular biology studies.

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