Acaricidal effect of an isolate from *Hoslundia opposita* vahl against *Amblyomma variegatum* (Acari: Ixodidae)

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**ABSTRACT**

**Background:** *Hoslundia opposita* Vahl. (Lamiaceae), a common local shrub in Ghana, is traditionally known not only for its pharmacological benefits but also for its insecticidal properties. Its acaricidal property, however, has not been investigated. **Objective:** To test the acaricidal effects of the crude extract and fractions of *H. opposita* leaves as well as to isolate and characterize the acaricidal principles. **Materials and Methods:** The crude methanolic extract, pet. ether, ethyl acetate and aqueous fractions of the leaves of *H. opposita* were tested against the larvae of the cattle tick, *Amblyomma variegatum*, using the Larval Packet Test. A bioassay-guided isolation was carried out to identify the acaricidal principle obtained from the ethyl acetate fraction. **Results:** The active principle was characterised as ursolic acid, a triterpene previously isolated from the leaves of the same plant. The extract and fractions were less potent than the control, malathion (LC₅₀ 1.14 × 10⁻⁴ mg/ml). Among the plant samples however the crude methanolic extract exhibited the highest effect against the larvae (LC₅₀ 5.74 × 10⁻³ mg/ml), followed by the ethyl acetate fraction (LC₅₀ 8.10 × 10⁻² mg/ml). Ursolic acid, pet. ether and aqueous fractions however showed weak acaricidal effects with LC₅₀ values of 1.13 mg/ml, 8.96 × 10⁻¹ mg/ml and 1.44 mg/ml, respectively. **Conclusion:** Ursolic acid was not as potent as the crude methanolic extract and the ethyl acetate fraction from which it was isolated. The overall acaricidal effect of *H. opposita* may have been due to synergy with other principles having acaricidal properties.

**Key words:** Acaricide, Larval Packet Test, ticks, ursolic acid

**INTRODUCTION**

Ghana has an estimated cattle population of more than 1.5 million.[¹] It is a source of livelihood for many in the rural areas.² Considered to be the greatest animal disease problem in Africa, ticks and tick-borne diseases (TDBs) are therefore a great threat and concern to cattle owners in Ghana.[³,⁴]

A myriad of problems are caused by ticks, notably disease transmission, reduction of animal growth rate, milk yield, livestock productivity and market opportunities.[⁵] Ticks and TDBs have substantial control costs, requiring considerable expenditures to treat, limit their spread and protect productivity.[⁵,³]

The use of chemical acaricides in tick control is associated with a number of problems, including environmental pollution, chemical residues in meat, wool and milk products, toxicity to farm animals, high cost of chemicals, pesticide poisoning and the rapid development of tick resistance.[³,⁶]

The search for safer and effective alternatives has soared and successes have been obtained through natural products, especially those from plant sources.[⁷,⁸] They are economically and environmentally friendly and less-toxic to humans and animals.

*Hoslundia opposita* Vahl. (fam. Lamiaceae) is a common perennial shrub that survives well in both wet and dry areas of Ghana. It has been shown to have several pharmacological benefits, both traditionally and scientifically. Essential oils from the leaves are extensively used as insect repellents. Although *H. opposita* has been widely studied for many pharmacological activities, its acaricidal potential, however, is yet to be sourced.
This study therefore seeks to investigate the acaricidal potential of *H. opposita* leaf extract against a common cattle tick in Ghana, *Amblyomma variegatum*.

**MATERIALS AND METHODS**

**Collection and authentication of plant material**

Fresh leaves of *H. opposita* were collected from a farmland at Juaben in the Ashanti region of Ghana. It was authenticated at the Department of Pharmacognosy, Faculty of Pharmacy and Pharmaceutical Sciences, Kwame Nkrumah of Science and Technology, Kumasi, Ghana, where a voucher specimen (HO1/2010) has been deposited at the department's Herbarium.

**Extraction and isolation of the active acaricial Principle**

Approximately 1.0 kg of the air-dried leaves was cold-macerated with 5.0 L of methanol for 7 days. With the aid of the rotary evaporator, the extract was concentrated under reduced pressure and further dried to give a solid mass of 90.9 g (9.09% w/w).

Subsequently, 50 g of the crude methanolic extract (CME) was partitioned between 5 × 100 ml aliquots of petroleum ether (40–60°C) and water [1:1]. The petroleum ether fraction was collected, concentrated into a solid friable mass of 23.7 g and designated F1. Then, 5 × 100 ml aliquots of ethyl acetate were shaken with the aqueous extract. The ethyl acetate fraction was also collected and concentrated into a solid friable mass of 5.5 g and designated F2. The aqueous fraction left (F3) was then concentrated and freeze-dried to yield 10.0 g of amorphous mass.

Five grams of the ethyl acetate fraction (active) was repeatedly column-chromatographed on silica gel 60 F254 (70–230 mesh ASTM), using pet. ether–ethyl acetate step gradient. Guided by their thin layer chromatography (TLC) profiles, fractions collected were bulked into four main fractions labeled A to D. Fraction C (active) was repeatedly column-chromatographed on silica gel 60 F254 (70–230 mesh ASTM), employing gradient elution with pet. ether–ethyl acetate mixture (increasing polarity). Eluates collected were bulked into three main fractions, C1 to C3. Fraction C2 (active) afforded an off-white amorphous powder, X (78 mg), after washing contaminants off with 5% ethyl acetate in pet. ether.

**Bioactivity studies**

**Biological material**

Engorged adult female ticks of *A. variegatum* were obtained from the Kumasi Abattoir and bred for eggs. The 5-day old, unfed larvae were used in this experiment.

**Bioactivity studies of extracts and fractions**

The crude extract and fractions were screened for acaricidal effect using the Larval Packet Test (LPT). Twenty 5-day old, unfed *A. variegatum* larvae were brought into contact with drug-impregnated filter papers (Watman No. 1) and observed for mortality after 24 h. A larva was recorded dead if it showed no sign of mobility. Concentrations used were serially diluted and ranged from 5.0 to 5.0 × 10⁻⁶% w/v. A broad-spectrum acaricide, malathion, was used as the positive control, while methanol was used as the negative control. All experiments were run in triplicate.

**Acaricidal activity of the isolate**

Concentrations ranging from 2.0 to 2.5 × 10⁻⁶% w/v were prepared by serial dilutions. These were tested against the 5-day-old, unfed larvae as described above, using methanol and malathion as negative and positive controls, respectively.

**Statistical analysis**

Results were expressed as LC₅₀ values compared with the control. One-way ANOVA was used for comparison of the means.

**RESULTS AND DISCUSSIONS**

A number of ethnomedicinally important plant species abound in Ghana. Essential oils from such plants are commonly used as repellents or pesticides. Traditional livestock farmers employ various herbs to protect their animals and themselves against ticks and TBDs. Although research into local acaricidal plants has been scanty, one study by Annan et al.⁹ that investigated the acaricial effects of a common shrub, *Plumbago zeylanica*, identified the naphthoquinone, plumbagin, as the main acaricial principle. Their finding therefore suggests the possibility of many more acaricial principles lying hidden in many local plant species used as acaricides.

Isolate X was isolated as an off-white amorphous powder with an Rₐ value of 0.38 in pet. ether:ethyl acetate (8:2) and 0.67 in chloroform:methanol (9:1). It recorded a melting point of 270–273°C (uncorrected). ESI-TOF-MS data of X showed a molecular ion peak [M⁺] at m/z 456.4, which corresponds to the molecular formula 

\[ C_{30}H_{48}O_3 \].

¹H-NMR spectral data showed that X has a vinylic proton triplet at δ = 5.14 and an alcoholic proton triplet at δ = 3.09. It also showed an allylic proton multiplet at δ = 1.87 and several saturated alkane protons with chemical shifts ranging from δ = 0.63 to 1.52.
The DEPT 135 spectrum showed seven methyl groups at δ 18.20, 62.31, 62.49, 62.76, 62.95, 63.05, 63.29, 63.67 and 63.87. Nine methylene groups were shown at δ 24.09, 26.70, 28.30, 29.55, 36.81, 39.09, 39.35, 39.80 and 47.70. Seven quaternary carbons were also shown at δ 78.70, 81.38, 81.80, 82.08, 82.37, 82.56, 82.79 and 82.98. The MS, 1H-NMR and 13C-NMR spectral data compared closely with that of ursolic acid as reported by the literature [10] [Table 1]. The structure of isolate X was therefore assigned as ursolic acid in CDCl3/CD3OD. The MS, 1H-NMR and 13C-NMR spectral data are presented in Table 1.

### Biological activity

The crude methanolic extract was the most potent among the test samples against *A. variegatum* larvae (LC50 5.74 × 10⁻² mg/ml). The ethyl acetate fraction also showed a relatively high potency (LC50 8.10 × 10⁻² mg/ml). The pet ether and aqueous fractions showed weak acaricidal effects, recording LC50 values of 8.96 × 10⁻¹ mg/ml and 1.44 mg/ml, respectively. Ursolic acid also exhibited a weak acaricidal effect against the larvae of *A. variegatum*. At 2.0% w/v, it showed 58.5% mortality, with an LC50 value of 1.13 mg/ml. This was significantly lower (P < 0.001) than the effect exhibited by the control, malathion (LC50 1.14 × 10⁻¹ mg/ml) [Table 2].

Malathion is a broad-spectrum acaricide commonly used in Ghana. It is one of several others used by livestock farmers. However, due to the high cost associated with its use, most resource-poor farmers have resorted to cheaper alternative means of arriving at similar outcomes. One such alternative is their traditional knowledge and practices of herbal acaricides. Oils from various parts of plants have been used to repel or kill pests, including seeds, fruit peels and flowers.

Ursolic acid is a triterpene, a known component of many essential oils. Literatures supporting the ethnomedicinal use of essential oils in pest control continue to increase. These effects have been attributed to the terpene components of these plants.

### Table 1: 1H-NMR and 13C-NMR spectral data for ursolic acid in CDCl3/CD3OD

| Position | 1H-NMR (ppm) | 13C-NMR (ppm) | Reference [10] |
|----------|--------------|---------------|----------------|
| C-1      | 3.09         | 38.57         | 38.90          |
| C-2      | 2.09         | 26.67         | 3.11           | 27.10          |
| C-3      | 1.21         | 55.15         | -              | 55.60          |
| C-6      | 0.63         | 32.93         | 5.16           | 33.40          |
| C-7      | 5.14         | 39.35         | -              | 39.80          |
| C-9      | 0.72         | 47.45         | -              | 47.90          |
| C-10     | 0.72         | 36.81         | -              | 37.20          |
| C-11     | 0.72         | 23.15         | 2.11           | 23.60          |
| C-12     | 2.10         | 125.40        | -              | 125.80         |
| C-13     | 1.52         | 138.08        | 0.90           | 138.50         |
| C-14     | 1.68         | 41.95         | 0.74           | 42.40          |
| C-15     | 1.00         | 29.55         | 0.70           | 29.90          |
| C-16     | 0.68         | 24.09         | 0.84           | 24.50          |
| C-17     | 0.68         | 47.70         | 1.01           | 48.10          |
| C-18     | 0.68         | 52.72         | -              | 53.20          |
| C-19     | 0.87         | 38.98         | 0.86           | 39.40          |
| C-20     | 0.77         | 38.80         | 0.78           | 39.20          |
| C-21     | 0.77         | 30.56         | 30.90          |
| C-22     | 0.77         | 36.71         | 37.10          |
| C-23     | 0.77         | 27.88         | 28.30          |
| C-24     | 0.77         | 15.25         | 15.60          |
| C-25     | 0.77         | 15.45         | 15.80          |
| C-26     | 0.77         | 16.73         | 17.20          |
| C-27     | 0.77         | 23.37         | 23.70          |
| C-28     | 0.77         | 180.61        | 180.80         |
| C-29     | 0.77         | 16.85         | 17.10          |
| C-30     | 0.77         | 20.98         | 21.30          |

ppm: parts per million

### Table 2: Acaridal effects of crude extract and fractions of *H. opposita* leaves against larvae of *A. variegatum* compared with control

| Material             | LC50 (mg/ml) | Range of % mortality (%) |
|----------------------|--------------|--------------------------|
| Crude methanolic extract | 5.74 × 10⁻² | 8.5–100.0                |
| Pet. ether fraction  | 8.96 × 10⁻¹ | 6.5–45.0                 |
| Ethyl acetate fraction | 8.10 × 10⁻² | 13.5–95.0                |
| Aqueous fraction     | 1.44         | 0.0–13.5                 |
| Ursolic acid         | 1.13         | 8.5–58.5                 |
| Malathion            | 1.14 × 10⁻⁴  | 38.5–100.0               |

LC50: concentration required to kill 50% of the test organisms, mg/ml: mass in milligrams per 1 ml; %: percent
Acaricidal effects may have been higher in the crude methanolic extract and the ethyl acetate fraction due to the fact that there was a combined effect by other components of the extract and fraction. Previous phytochemical screening of *H. opposita* showed the presence of other medicinally important terpenes like menthol, thymol, β-caryophyllene, oleanolic acid and germacrene-D. A study has even reported on the presence of benzyl benzoate, a known pesticide[15] identified from samples of essential oils obtained from the plant in West Africa. The combined effects of components such as these may have contributed to the high effects observed. Some structure–activity studies on terpenes suggest that molecules possessing free alcoholic, phenolic or potential olefinic modifications showed the most potent acaricidal activity.[16,17]

**CONCLUSION**

The results of this study suggest that the crude methanolic extract and ethyl acetate fraction of the leaves of *H. opposita* have acaricidal activity against the larvae of *A. variegatum*. Ursolic acid has been found to be a component of the ethyl acetate fraction, which contributes to the acaricidal activity of the plant. The finding therefore corroborates the traditional use of the plant as a repellent; however, its great potential would be limited by using it only as a repellent. The alcoholic extract of the leaves can serve the livestock industry better as a safer and cheaper alternative to chemical acaricides.

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