Effects of long noncoding RNA on prognosis of oral squamous cell carcinoma
A protocol for systematic review and meta analysis
Qingjie Lin, DDSa, Yong Zhang, DDSb, Yanguo Liu, DDS, Xin Xu, PhDa,∗

Abstract
Background: Long noncoding RNA (lncRNA) is reported to be upregulated in many tumors. Although the expression of lncRNA in oral squamous cell carcinoma has been assessed, the association between lncRNA expression and prognosis or clinicopathological feature still remains controversial. Therefore, we conducted a meta-analysis to verify whether lncRNA expression was related to prognosis or clinicopathological features in patients with oral squamous cell carcinoma.

Methods: We searched Embase, PubMed, Web of Science, Cochrane library, Chinese National Knowledge Infrastructure, and Wanfang databases from inception to February 2021. The language included Chinese and English. The published literature on lncRNA expression and prognosis or clinicopathological characteristics of patients with oral squamous cell carcinoma was statistically analyzed. The combination of hazard ratios (HRs), odds ratios (OR), and 95% confidence intervals (95% CIs) were applied to evaluate the effects of lncRNA on the prognosis and clinicopathological features of oral squamous cell carcinoma.

Results: This study could provide a comprehensive review of the available evidence of lncRNA on the prognosis and clinicopathological features of oral squamous cell carcinoma.

Conclusion: The conclusion of our study will provide the updated evidence to judge the lncRNA on the prognosis and clinicopathological features of oral squamous cell carcinoma.

Abbreviations: CIs = confidence intervals, DFS = disease-free survival, HRs = hazard ratios, lncRNAs = long noncoding RNAs, OS = overall survival, PRISMA-P = Preferred Reporting Items for Systematic Reviews and Meta Analysis Protocols, OR = odds ratio.

Keywords: long noncoding RNA, meta-analysis, oral squamous cell carcinoma, prognosis, protocol

1. Introduction
Oral cancer is one of the common oral malignant tumors, accounting for more than 90%, while oral squamous cell carcinoma occupies over 80% of head and neck malignant tumors, often occurring in tongue, gingiva, buccal mucosa, and other parts of body.[1–3] In recent years, the incidence of oral squamous cell carcinoma has increased year by year and the patients tended to be younger.[4] Clinically, surgical treatment, radiotherapy, and chemotherapy are the main treatment methods.[5] However, due to its high degree of malignancy and local invasion and lymph node metastasis, up to 60% of patients with oral squamous cell carcinoma are in the late stage of clinical progress. As a result, the survival rate of patients is low, and the life quality of patients is seriously affected.[6] In the past 30 years, the overall survival (OS) rate of patients with oral squamous cell carcinoma reached about 50%, without any significant improvement.[7] Early diagnosis and treatment of oral squamous cell carcinoma are the key steps to control the disease and improve the survival rate. Therefore, it is clinically important to find new markers for diagnosis, prognosis, and treatment of oral squamous cell carcinoma.

With the length of more than 200bp long-stranded noncoding RNA (lncRNA) lacks the potential of protein coding and is a RNA transcription product. It is involved in DNA replication, RNA transcription, protein translation, cell development, and differentiation, and is an important regulator of cell biology.[8–11] In recent years, it has been obvious that lncRNA plays an important role in the occurrence and development of tumors.[12]
lncRNA is also closely related to its occurrence and progression, and may be applied as a biomarker or therapeutic target for the diagnosis and prognosis of oral squamous cell carcinoma.[13–16]

Many studies have revealed that lncRNA is closely associated with the clinicopathological features, survival, and biological behavior of tumor cells in patients with oral squamous cell carcinoma.[17–23] However, there still exists the lack of effective evidence-based medicine to prove it. Therefore, this study conducted a systematic evaluation and meta-analysis of the included articles to further explore the relationship between lncRNA and clinicopathological features and the prognosis of patients with oral squamous cell carcinoma.

2. Materials and methods

2.1. Study registration

This protocol has been registered at Open Science Framework and its registration number is DOI 10.17605/OSF.IO/Z5Q3K. Any significant amendments of this protocol will be recorded in the Open Science Framework before the review is completed. According to the Preferred Reporting Items for Systematic Reviews and Meta-analysis Protocols (PRISMA-P) statement,[24] this protocol is drafted.

2.2. Inclusion and exclusion criteria

The study would be included in this meta-analysis if it meets the following criteria: (1) Patients were diagnosed with oral squamous cell carcinoma; (2) lncRNA expression level was detected; (3) Patients were divided into two groups based on the lncRNA expression level; (4) Efficient data were provided; (5) Full-text was available; (6) The research data were complete, and literatures on hazard ratio (HR), 95% confidence interval (CI), and observation indexes can be extracted. The following studies would be excluded from this meta-analysis: duplicated publications or patients, reviews, case reports, letters, comments, animal experiments, cell experiments, or studies without efficient data.

2.3. Search strategy

Embass, PubMed, Web of Science, Cochrane library, Chinese National Knowledge Infrastructure, and Wanfang databases were comprehensively searched up to February 2021. The language was Chinese and English. We also checked the references of retrieved articles to avoid missing relative studies. The combined method of MeSH Term and free words would be adopted for literature retrieval. A search strategy of PubMed is summarized in Table 1, which is created on the basis of the Cochrane handbook guidelines. The search strategies of other databases would be established similarly.

2.4. Data collection and analysis

2.4.1. Selection of studies. All reviewers received evidence-based training and adhered to the summarized process. The 2 reviewers independently screened the literature based on the title, abstract, and key words of literatures, and excluded the irrelevant literatures. The rest of literatures were further confirmed by 2 researchers after reading the full text. The excluded research and the reasons for the exclusion were recorded. The differences between the 2 reviewers were resolved through consensus or a third independent reviewers. The process of the selection is illustrated in Figure 1.

### Table 1

| Number  | Search terms                                      |
|---------|---------------------------------------------------|
| #1      | Mouth Neoplasms[MeSH]                             |
| #2      | Cancer of Mouth[Title/Abstract]                   |
| #3      | Mouth Cancer[Title/Abstract]                      |
| #4      | Oral Cancer[Title/Abstract]                       |
| #5      | Oral Neoplasms[Title/Abstract]                    |
| #6      | Cancer of the Mouth[Title/Abstract]               |
| #7      | Neoplasms, Mouth[Title/Abstract]                  |
| #8      | Neoplasms, Oral[Title/Abstract]                   |
| #9      | Cancer, Mouth[Title/Abstract]                     |
| #10     | Cancer, Oral[Title/Abstract]                      |
| #11     | Cancers, Mouth[Title/Abstract]                    |
| #12     | Cancers, Oral[Title/Abstract]                     |
| #13     | Mouth Cancers[Title/Abstract]                     |
| #14     | Mouth Neoplasms[Title/Abstract]                   |
| #15     | Neoplasms, Mouth[Title/Abstract]                  |
| #16     | Neoplasms, Oral[Title/Abstract]                   |
| #17     | Oral Cancers[Title/Abstract]                      |
| #18     | Oral Neoplasms[Title/Abstract]                    |
| #19     | Oral squamous cell carcinoma[Title/Abstract]      |
| #20     | or/1-19                                           |
| #21     | Long non-coding RNA[Title/Abstract]               |
| #22     | LncRNA[Title/Abstract]                            |
| #23     | or/21-22                                          |
| #24     | Prognos*                                          |
| #25     | Survival                                          |
| #26     | or/24-25                                          |
| #27     | #20 and #23 and #26                               |

2.4.2. Data extraction and management. Two authors extracted the data and assessed the quality of included studies independently. Any disagreement during this process was resolved through group discussion.

According to the inclusion and exclusion criteria, the literature was screened and the relevant data were extracted. The data extracted from the literature include the first author, publication year, number of patients and region, lncRNA name, cut-off value of lncRNA expression level, detection methods, prognostic indicators such as OS and DFS, and clinicopathological data, including age, sex, tumor differentiation, tumor diameter, depth of tumor invasion, lymph node metastasis, distant metastasis, TNM stage.

2.4.3. Assessment of risk of bias in included studies. The quality of all the included studies will be evaluated by 2 reviewers independently based on the Newcastle–Ottawa scale (NOS) that is used to evaluate the quality of observational studies.[25] Disagreement will be reported and resolved by a third reviewer. Meanwhile, score < 6 is considered low quality, and ≥6 is classified as high quality.

2.4.4. Measures of prognosis. OS and disease-free survival (DFS) would be taken as prognostic outcomes. The results would be expressed as HRs with 95% CIs.

2.4.5. Dealing with missing data. If there are insufficient or missing data in the literature, we would contact the author via email. If the data is not available, we would only analyze the currently available data and discuss its potential impacts.
2.4.6. Assessment of heterogeneity and data synthesis. All analysis was conducted with Review Manager 5.3 (The Cochrane Collaboration, Copenhagen, Denmark) and Stata 12.0 (Stata, College Station, TX) for Windows. For OS and DFS, HR and corresponding 95% CI were applied as the summary measures. For clinicopathological parameters, odds ratio (OR) and corresponding 95% CI were applied. Besides, inter-study heterogeneity was assessed by carrying out Chi-squared test and \( I^2 \) statistic. \( I^2 \leq 50\% \) or \( P \) value for heterogeneity >.10 proved that there was no obvious heterogeneity among studies. As a result, a fixed-effect model should be utilized. If not, a random-effect model should be applied. The association was considered to be significant when \( P < .05 \).

2.4.7. Assessment of reporting biases. If sufficient studies are included (at least 10), we will test the reporting bias in the meta-analysis by using an inverted funnel plot.\(^{26} \) Funnel plots, Begg test, and Egger test were performed to evaluate the publication bias.

2.4.8. Subgroup analysis. We will conduct a subgroup analysis based on the detection method of lncRNA expression, race, and the source of survival data.

2.4.9. Sensitivity analysis. If sufficient studies are available, we will perform sensitivity analyses to confirm the robustness of the primary results. The meta-analysis will be respectively processed by excluding studies with small sample size, and low methodological quality.

2.4.10. Ethics and dissemination. The content of this article does not involve moral approval or ethical review and would be presented in print or at relevant conferences.

3. Discussion

A large number of studies at home and abroad have confirmed that lncRNA is involved in the pathological process of the occurrence and development of a variety of tumors at different levels.\(^{27-29} \) lncRNA can be used as a regulator of gene
expression through gene modification, transcription, and post-transcriptional processing. Many studies have revealed that lncRNA can participate in the proliferation, apoptosis, angiogenesis, metastasis, and invasion of oral squamous cell carcinoma through various pathways and molecular mechanisms. In the past few years, a large number of studies have illustrated that the upregulation and downregulation of lncRNA are involved in the development and progression of oral squamous cell carcinoma. Therefore, the expression of lncRNA may be related to the prognosis of oral squamous cell carcinoma, while controversial results have been discovered. Here, we conducted this meta-analysis for the first time to further summarize the prognostic value of lncRNA expression in oral squamous cell carcinoma.

There are several key points in our research. First of all, as far as we know, this is the first meta-analysis to explore the prognostic value of lncRNA expression in oral squamous cell carcinoma. Second, this study comprehensively analyzed the prognostic and pathological parameters, which further confirmed the prognostic role of lncRNA expression in oral squamous cell carcinoma. Third, our research strictly follows the rules of PRISMA, so the method is normative. Nevertheless, our research has no limitations. There are few studies on the meta-analysis, and a relatively small sample size may reduce the reliability of the results. Furthermore, there is a great difference in the critical value among the included studies, which may limit the clinical application of this conclusion. Most importantly, although there are no restrictions on the country in the process of literature selection, most of the studies are carried out in China. Consequently, this conclusion may be difficult to extend to other countries. Therefore, more high-quality and extensive researches should be carried out to clarify this issue.

Nevertheless, our study will provide evidence to support the prognostic role of lncRNA expression in oral squamous cell carcinoma and provide relevant strategies for the accurate treatment of tumors.

**References**

[1] Fitzsimonds ZR, Rodriguez-Hernandez CJ, Bagatikar J, et al. From beyond the pale to the pale riders: the emerging association of bacteria with oral cancer. J Dent Res 2020;99:1–9.

[2] Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin 2020;70:7–30.

[3] Lell M, Detmar K, Schoen M, et al. Imaging of head and neck cancer. Der Nuklearmediziner 2020;43:103–14.

[4] Zhang N, Zeng L, Wang S, et al. LncRNA FER1L4 promotes oral squamous cell carcinoma progression via targeting miR-133a-5p/Prx1 axis. Oncotargets Ther 2021;14:793–806.

[5] Bilde A, von Buchwald C, Johansen J, et al. The Danish national guidelines for treatment of oral squamous cell carcinoma. Acta Oncol (Stockholm, Sweden) 2006;45:294–9.

[6] Hedberg ML, Goh G, Chiosea SI, et al. Genetic landscape of metastatic and recurrent head and neck squamous cell carcinoma. J Clin Invest 2015;126:169.

[7] Song Y, Qin X, Zhang J, et al. Expression of long noncoding RNA COL11A1-208 in oral squamous cell carcinoma and its clinical significance. J Shaanxi Jiao Tong Univ (Med Sci) 2020;40:1334–9.

[8] Heo JB, Lee YS, Sung S. Epigenetic regulation by long noncoding RNAs. Science (New York, NY) 2013;321:685–93.

[9] Ying Q, Ya-Ni K, Xiao-Dong Z. Unexpected roles of long non-coding RNAs in cancer biology. J Shanghai Jiaotong Univ Sci 2014;19:544–9.

[10] Niu X, Yang B, Liu F, et al. LncRNA HOXA11-AS promotes OSCC progression by sponging miR-98-5p to upregulate YBX2 expression. Biomed Pharmacother 2020;121:109623.

[11] Sur S, Nakanishi H, Steele R, et al. Long non-coding RNA ELDR enhances oral cancer growth by promoting ILF3-cyclin E1 signaling. EMBO Rep 2020;21:e51042.

[12] Schmitt AM, Chang HY. Long noncoding RNAs in cancer pathways. Cancer Cell 2016;29:452–63.

[13] Kasper G, Marta B, Tomasz K, et al. LncRNA in HNSCC: challenges and potential. Contemp Oncol 2017;21:259–66.

[14] A GF , B MR , C MJ , et al. Expression and function of long non-coding RNAs in head and neck squamous cell carcinoma. ScienceDirect. Exp Mol Pathol 1043;112:33.

[15] Yu L, Shao X, Huo L, et al. Long non-coding RNA [lncRNA] metastasis-associated long adenosine transcript 1 (MALAT1) promotes cell proliferation and migration by regulating miR-133a-5p/Prx1 axis. OncoTargets Ther 2021;14:793–806.

[16] Ghafouri-Fard S, Dashti S, Taheri M. The role of long non-coding RNA in oral squamous cell carcinoma. J Biosci Bioeng 2015;127:110202.

[17] Vishwakarma S, Pandey R, Singh R, et al. Expression of H19 long non-coding RNA is down-regulated in oral squamous cell carcinoma. J Biosci 2020;45:145.

[18] Zheng X, Tian X, Zhang Q, et al. Long non-coding RNA SAMMSON as a novel potential diagnostic and prognostic biomarker for oral squamous cell carcinoma. J Dent Sci 2020;15:329–35.

[19] Huang Z, Sang T, Zheng Y, et al. Long non-coding RNA PANDAR overexpression serves as a poor prognosis biomarker in oral squamous cell carcinoma. Int J Clin Exp Pathol 2018;11:2728–34.

[20] Geng YD, Wang SB, Lu TQ, et al. Expression and functions of long non-coding RNA actin filament-associated protein 1-antisense RNA1 in oral squamous cell carcinoma. Hua Xi Kou Qiang Yi Xue Za Zhi 2019;37:594–601.

[21] Yang ZG, Huang HY, Xu X. Long non-coding RNA PCAT-1 expression in oral squamous cell carcinoma and its clinical significance. Shanghai Kou Qiang Yi Xue 2019;28:67–70.

[22] Shao T, Huang J, Zheng Z, et al. SCCA, TSGF, and the long non-coding RNA COL11A1-208 in oral squamous cell carcinoma. Hua Xi Kou Qiang Yi Xue Za Zhi 2017;35:101.

[23] Shao T, Huang J, Zheng Z, et al. SCCA, TSGF, and the long non-coding RNA COL11A1-208 in oral squamous cell carcinoma. J Biosci 2020;45:145.

[24] Shamseer L, Moher D, Clarke M, et al. Preferred reporting items for systematic review and meta-analysis protocols (PRISMA-P) 2015: elaboration and explanation. BMJ (Clinical research ed) 2015;350:g7647.

[25] Stang A. Critical evaluation of the Newcastle-Ottawa scale for the assessment of the quality of nonrandomized studies in meta-analyses. Eur J Epidemiol 2010;25:603–5.
[26] Duval S, Tweedie R. Trim and fill: a simple funnel-plot-based method of testing and adjusting for publication bias in meta-analysis. Biometrics 2000;56:455–63.

[27] Huang Z, Zhuo W, Xu R, et al. The relationship between long noncoding RNA (lncRNA) Small Nucleolar RNA Host Gene 12 (SNHG12) expression in solid malignant tumors and prognosis of tumor patients: a systematic review and meta-analysis. Medicine 2020;99:e2224.

[28] Poursheikhani A, Nokhandani N, Yousefi H, et al. Clinicopathological significance of long non-coding RNA GHET1 in human cancers: a meta-analysis. Curr Pharm Biotechnol 2020;21:1422–32.

[29] Qian P, Xu Z, Chen H, et al. Abnormally expressed lncRNAs in the prognosis and clinicopathology of oesophageal cancer: a systematic review and meta-analysis. J Genet 2020;99:43.

[30] Feng H, Zhang X, Lai W, et al. Long non-coding RNA SLC16A1-AS1: its multiple tumorigenesis features and regulatory role in cell cycle in oral squamous cell carcinoma. Cell Cycle (Georgetown, Tex) 2020;19:1641–53.

[31] Wang F, Ji X, Wang J, et al. LncRNA PVT1 enhances proliferation and cisplatin resistance via regulating miR-194-5p/HIF1a axis in oral squamous cell carcinoma. OncoTargets Ther 2020;13:243–52.

[32] Sun M, Shen Z. Knockdown of long non-coding RNA (lncRNA) colon cancer-associated transcript-1 (CCAT1) suppresses oral squamous cell carcinoma proliferation invasion, and migration by inhibiting the Discoidin Domain Receptor 2 (DDR2)/ERK/AKT Axis. Med Sci Monit 2020;26:e920020.

[33] Zhao J, Bai X, Feng C, et al. Long non-coding RNA HCP5 facilitates cell invasion and epithelial-mesenchymal transition in oral squamous cell carcinoma by miR-140-5p/SOX4 axis. Cancer Manage Res 2019;11:10455–62.

[34] Huang GZ, Wu QQ, Zheng ZN, et al. Identification of candidate biomarkers and analysis of prognostic values in oral squamous cell carcinoma. Front Oncol 2019;9:1054.