MicroRNA-365 Inhibits Cell Growth and Promotes Apoptosis in Melanoma by Targeting BCL2 and Cyclin D1 (CCND1)

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Background: MicroRNA-365 (miR-365) is involved in the development of a variety of cancers. However, it remains largely unknown if and how miRNAs-365 plays a role in melanoma development.

Material/Methods: In this study, we overexpressed miR-365 in melanoma cell lines A375 and A2058, via transfection of miR-365 mimics oligos. We then investigated alterations in a series of cancer-related phenotypes, including cell viability, cell cycle, apoptosis, colony formation, and migration and invasion capacities. We also validated cyclin D1 (CCND1) and BCL2 apoptosis regulator (BCL2) as direct target genes of miR-365 by luciferase reporter assay and investigated their roles in miR-365 caused phenotypic changes. To get a more general view of miR-365’s biological functions, candidate target genes of miR-365 were retrieved via searching online databases, which were analyzed by Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analyses for potential biological functions. We then analyzed The Cancer Genome Atlas (TCGA) Skin Cutaneous Melanoma (SKCM) dataset for correlation between miR-365 level and clinicopathological features of patients, and for survival of patients with high and low miR-365 levels.

Results: We found that miR-365 was downregulated in melanoma cells. Overexpression of miR-365 remarkably suppressed cell proliferation, induced cell cycle arrest and apoptosis, and compromised the migration and invasion capacities in A375 and A2058 cell lines. We also found that the phenotypic alterations by miR-365 were partially due to downregulation of CCND1 and BCL2 oncoproteins. The bioinformatics analysis revealed that predicted targets of miR-365 were widely involved in transcriptional regulation and cancer-related signaling pathways. However, analysis of SKCM dataset failed to find differences in miR-365 level among melanoma patients at different clinicopathologic stages. The Kaplan-Meier analysis also failed to discover significant differences in overall survival and disease-free survival between patients with high and low miR-365 levels.

Conclusions: Our findings suggested that miR-365 might be an important novel regulator for melanoma formation and development, however, the in vivo roles in melanoma developments need further investigation.

MeSH Keywords: Apoptosis • bcl-Associated Death Protein • Cyclin D1 • Melanoma • MicroRNAs

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Background

Melanoma, a skin malignancy notorious for its aggressiveness and high metastatic potential, has had a fast increase in its incidence rate, and accounts for over 65% of skin cancer-related deaths [1,2]. Most localized melanomas are curable with surgical resection, while the prognosis of patients with distant metastases is usually poor with a 10-year survival rate of only 16%, mainly due to innate or acquired resistance to therapeutic regimens [3]. This situation highlights the significance of further understanding of the underlying molecular mechanisms that contribute to melanoma development and metastasis.

MicroRNAs (miRNAs) are small non-coding RNAs that negatively regulate gene expression post-transcriptionally by pairing to the 3'-untranslated region (3'-UTR) of target mRNAs [4,5]. MiRNAs are involved in regulating a series of cellular functions, including development, differentiation, apoptosis, and proliferation. Deregelated miRNA expression has been shown to contribute to the development of cancers, including breast cancer [6], digestive tract cancers [7], melanoma [5], and so forth. MiR-365 is located on chromosome 16p13.12, a region that has been involved in multiple oncogenic processes. The expression pattern and biological roles of miR-365 are cancer type-dependent. MiR-365 is highly expressed in cutaneous squamous cell carcinoma [8,9] and breast cancer [10], while it is downregulated in colon cancer [11], lung cancer [12], and melanoma [13]. MiR-365 may display either a pro-proliferative or pro-apoptotic role in a specific cancer type. The roles of miR-365 in melanoma development is very poorly understood. Bai et al. have shown that miR-365 was downregulated in melanoma tissues and ectopic expression of miR-365 suppressed cell cycle progression and promoted apoptosis by targeting NR1P1, which is an essential regulator of cell migration and invasion, suggesting that miR-365 exerted a tumor-suppressive effect in melanoma [13].

Cyclin D1 (CCND1) is a member of the cyclins, which are essential in regulating cell cycle [14]. CCND1 is a well-established human oncogene [14], which is commonly overexpressed in different types of cancers such as breast cancer [15], lung cancer [16], and melanoma [17]. CCND1 overexpression can result in a number of potentially oncogenic effects and have been shown associated with poor patient outcome [18]. BCL2 apoptosis regulator (BCL2) belongs to the BCL2 family proteins, which are important regulators of apoptosis [19]. Antiapoptotic BCL2 family members, including BCL2, BCLXL, MCL1, and BCLW, inhibit apoptosis by sequestering the activators from interacting with BAX and BAK [20]. Overexpression of anti-apoptotic BCL2 has been observed in many types of cancers, such as follicular lymphoma [21], breast cancer [22], prostate cancers [23] and melanoma [24]. Upregulated expression of BCL2 protein promotes tumorigenesis and tumor progression and is associated with poor patient prognosis [25]. CCND1 and BCL2 have been reported as targets genes of miR-365 in colon cancer [11]. Thus, in this study we investigated the functional relationship between miR-365 and these 2 genes.

In this study, to further explore the roles of miR-365 in melanoma development and reveal the underlying molecular mechanisms, we investigated the effects of miR-365 overexpression on cell cycle, apoptosis, cell migration and invasion in 2 melanoma cell lines, A375 and A2058. We also investigated the roles of CCND1 and BCL2 in the cellular effects of miR-365. To obtain a comprehensive understanding of the potential biological functions of miR-365, the Gene Ontology (GO) analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis of predicted targets of miR-365 were carried out. In addition, analysis of the The Cancer Genome Atlas (TCGA) datasets for melanoma patients was also performed to investigate the association between miR-365 level and the clinicopathologic features and outcomes of melanoma patients.

Material and Methods

Cell culture

NHEM (Normal Human Epidermal Melanocytes) cell line was obtained from Miao Tong Biological Technology (Shanghai, China) and cultured in M2 medium. The human melanoma cell lines A375, A2058, SK-MEL-2, and SK-MEL-28 were obtained from China Center for Type Culture Collection (Wuhan, China). These cell lines were cultured in Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10% fetal bovine serum. All cell lines were incubated at 37°C with 5% CO₂ in a humidified atmosphere.

Transfection of miR-365 mimics

To transiently overexpress miR-365, A375 and A2058 cells were transfected with miR-365 mimic oligos (Life Technologies, USA) at a final concentration of 100 nM by using lipofectamine 2000 (Thermo Fisher Scientific, USA) according to the manufactures instructions. The control cells were transfected with the non-targeting control oligo (NC oligo for short, Life Technologies, USA) at the same concentration.

Quantitative real-time-PCR

Total RNA, including miRNA, was extracted from cells using the miRNeasy mini kit (Qiagen, USA) according to manufacturer’s instructions. For measuring BCL2 and CCND1 mRNA levels, 1.5 µg total RNA was reversely transcribed into cDNA using the Omniscript RT kit (Qiagen, USA) according to manufacturer’s instructions. The qRT-PCR was then performed to detect the levels of BCL2 and CCND1 as described by literature [26]. Briefly, 2 µL of cDNA (diluted in 1: 10) was used as templates

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for qRT-PCR using the iQ SYBR Green Supermix (Bio-Rad, USA) on a CFX Real-Time PCR Detection System (Bio-Rad, USA). The amplification protocol is as following: 95°C for 3 min; followed by 35 cycles of 95°C for 15 sec and 60°C for 30 sec. GAPDH was used as an internal control. Primer sequences are as follows: GAPDH forward: 5’-AAGCCTGCGGTGACTAC-3’ and reverse: 5’-GGGCGCAGATCAGGCAAA-3’; BCL-2 forward: 5’-ATGTTGCTGGA GACGCTCA-3’ and reverse: 5’-GGGCGCTACAGTCTCCAAACAAA-3’; CCND1 forward: 5’-CAGTATCCGCAAACACGGC-3’ and reverse: 5’-AAGTTTGGGCTCCTCAG-3’. All reactions were performed in triplicate. Fold changes in mRNA expression levels were calculated using the 2-ΔΔCT method [27]. For measuring miR-365 levels, a miScript II RT kit (Qiagen, USA) was used to converted miRNAs to cDNA and to add a universal tag to each cDNA product. The levels of miR-365 was detected via performing qRT-PCR assay using the miScript SYBR Green PCR kit (Qiagen, USA) according to the following protocol: 95°C for 15 min, followed by 40 cycles of 94°C for 15 sec, 55°C for 30 sec, and 70°C for 30 sec [28]. The qRT-PCR primers for miR-365 was 5’-GCTAATGCCCCTAAAAATCC-3’. U6 primer was 5’-GAAGCATAACAGAGAATTTAGCA-3’. All reactions were performed in triplicate. The expression level of miR-365 was normalized to that of U6 snRNA. Fold changes in miR-365 levels were calculated using 2-ΔΔCT method [27].

**MTT assay**

Cell viability was measured by using a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay (Sigma, USA) as described in published study [29]. Briefly, cells were seeded in 96-well plates at 5000 cells/well in triplicates. To measure cell viability, we added 10 µL MTT (5 mg/mL) to each well and incubated the plates for 4 hours at 37°C. Then 100 µL solubilization buffer was added to each well without removing medium. The plates were incubated for overnight at 37°C and the absorbance at 595 nm was measured by using a spectrophotometer. Results shown are the mean ± standard error (SEM) of 3 independent experiments.

**Cell cycles analysis**

Cells were harvested by trypsinization and fixed in 70% ethanol at −20°C for at least overnight. The cells were then treated with 50 µL of 100 µg/mL boiled RNase A (Sigma, USA) for 30 min followed by staining with 200 µL propidium iodide (from 50 µg/mL stock solution). Cell cycle was acquired using a flow cytometry (LSR II, BD Biosciences, USA) and data was analyzed using the FlowJo software (FlowJo LLC, USA).

**Apoptosis by flow cytometry**

Cell apoptosis was detected using Annexin V-FITC apoptosis detection reagent (BD, USA) according to the manufacturer’s instructions. Briefly, cells were washed twice with cell staining buffer and resuspended in binding buffer at 1×10^5 cells/100 µL. To 100 µL of cell suspension, 5 µL of Annexin V-FITC was added and cells were incubated at room temperature for 15 min followed by addition of another 400 µL binding buffer. Before measuring samples, 8 µL of Hoechst 33258 was added to each sample. Cell apoptosis was then detected by using a flow cytometry (LSR II, BD Biosciences, USA) and data was analyzed using the FlowJo software (FlowJo LLC, USA).

**Western blotting**

Western blot analysis for BCL2 and CCND1 levels was performed as described by Li et al. [30], with minor modifications. Briefly, cells were lysed, and the protein concentration of total cell lysates was measured by using the BCA assay (Beyotime, China). About 25 µg of whole cell lysates were subjected to electrophoresis and then transferred to an Immobilon-P Membrane (Merck Millipore, USA). After blocked by 5% non-fat milk for 1 hour at room temperature, the membranes were incubated with rabbit anti-human BCL2 or rabbit anti-human CCND1 (both 1: 1000, Cell Signaling, USA) and mouse anti-human β-actin (1: 2000, Abcam, USA) for overnight at 4°C. After incubation with corresponding secondary antibodies for 1 hour, the membrane was washed 3 times with TBST, and then incubated with BeyoECL Plus chemical luminescence solution (Beyotime, China). The membrane was then imaged using a ChemiDoc XRS imaging system and analyzed using the QuantityOne software (Bio-Rad, USA).

**Colony formation assay**

For the colony formation assay, cells were seeded at 300 cells/well on 6 well plates in triplicate and maintained in complete culture medium for 12 days until obvious colonies were formed. Colonies were then fixed by 70% ethanol for 5 min and stained with Coomassie blue dye for 5 min. A colony with more than 50 cells were considered as a colony and counted.

**Transwell migration and invasion assay**

Cells were transfected with miR-365 mimics or control oligos. Then cells were resuspended in 200 µL serum free medium and seeded in the upper part of the Transwell-inserts (8 µm pore membranes) in the 24-well plate with 20% FBS supplemented DMEM medium placed in the lower compartment. For the Matrigel invasion assay, 50 µL Matrigel at 300 µg/mL was added to a 24-well Transwell insert and solidified in a 37°C incubator for 1 hour to form a thin gel layer before cells were loaded. After incubation for 24 hours, cells that passed through the membrane were fixed on the membrane using ethanol and then stained with crystal violet. The average number of cells that passed through the membrane from 15 representative fields (5 replicates per individual experiments) was counted under a phase contrast microscope.
Luciferase reporter assay

The region of 3'UTR of the human CCND1 and BCL2 mRNA containing the miR-365 targeting site were cloned in between the XhoI(5') and NotI(3') sites in a psi-CHECK2 vector (Promega, USA). A375 cells were co-transfected with 1 μg of p3'UTR-CCND1 or p3'UTR-BCL2 or psi-CHECK2 vector and 100 nM miR-365 mimic oligos using Lipofectamine 2000 (Invitrogen, USA). Cells were harvested 24 hours after transfection and assayed by using a Dual Luciferase Reporter Assay (Promega, USA) according to the manufacturer’s instructions. The assay was performed in triplicates. The expression level of CCND1 and BCL2 3’UTR was defined as Renilla luciferase activity normalized to firefly luciferase activity for each well.

Bioinformatics analysis

The targets genes of miR-365 were predicted using the mi-Randa, DIANA-microT and TargetScan database and the predicted targets that showed up in at least 2 databases were selected for further analysis. To further investigate the functional roles of miR-365, target genes were subjected to Gene Ontology (GO) analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway enrichment analysis using DAVID Bioinformatics Resources [31].

Analysis of TCGA melanoma dataset

Skin Cutaneous Melanoma (SKCM) dataset for 470 melanoma samples was obtained from The Cancer Genome Atlas (TCGA). Level 3 miRNA-Seq data generated from the miSeq Illumina platform and clinicopathological features of patients were retrieved from the TCGA Data Portal (released on 04/08/16). All samples had clinical and follow-up information. The dataset contained 448 patients with miRNA-Seq data, therefore only patients with miRNA data were used in this analysis. As the data were obtained from TCGA, further approval by an ethics committee was not required. This study meets the publication guidelines provided by TCGA. Expression of miR-365 in patients at different disease stages was compared by using one-way ANOVA. Overall survival (OS) and disease-free survival (DFS) were assessed by using the Kaplan-Meier method and curves were compared by univariate (log-rank) test.

Statistical analysis

Statistical analysis was performed using the GraphPad Prism 6 (GraphPad Software Inc., USA). The difference between 2 groups was analyzed by Student’s t-test; the means of more than 2 groups were compared by using one-way ANOVA followed by the Tukey-Kramer method. Each experiment was repeated at least 3 times and data was presented as mean ± SEM. A P value ≤0.05 was considered to be significant.

Results

MiR-365 inhibited cell proliferation and induced apoptosis in melanoma

Initially, we found out that miR-365 was downregulated in 4 melanoma cell lines, A375, A2058, SK-MEL-2, and SK-MEL-28 compared to a melanocyte cell line, NHEM (Figure 1A). We then investigated the biological functions of miR-365 in melanoma cell lines. MiR-365 was overexpressed in A375 and A2058 melanoma cell lines by transient transfection of a miR-365 mimic oligo, and the overexpression of miR-365 was validated by qRT-PCR (Figure 1B). The results of MTT assays revealed that ectopic expression of miR-365 significantly reduced cell viability in both A375 and A2058 cells (Figure 1C). We then explored the potential mechanisms for the decreased cell viability in miR-365 overexpressing cells. The colony formation assay indicated that melanoma cells overexpressing miR-365 has decreased proliferation ability than the control cells (Figure 1D). The cell cycle analysis showed a significant increase in population at the G0/G1 phase and a decrease in the population at S phase (Figure 1E), suggesting a G1/S blockage was caused by miR-365 overexpression. In addition, we also found that miR-365 led to a substantial increase in cell apoptosis (Figure 1F). Together, these results indicated that miR-365 played a tumor suppressive role in melanoma likely through inhibiting cell proliferation, blocking cell cycle progression and enhancing cell apoptosis.

miR-365 suppressed melanoma migration and invasion

Bai et al. reported that miR-365 levels were inversely correlated with melanoma metastasis [13]. We therefore investigated the effects of miR-365 overexpression on the migration and invasion abilities of A375 and A2058 melanoma cells. The Transwell migration assay and Matrigel invasion assay indicated that overexpression of miR-365 significantly suppressed melanoma cell migration (Figure 2A) and invasion (Figure 2B) capacities, respectively.

miR-365 targeted BCL2 and CCND1

Both BCL2 and CCND1 were reported as direct targets of miR-365 in colon cancer [11]. However, these regulatory associations have not been established in melanoma and the roles of BCL2 and CCND1 in miR-365-mediated phenotypes in melanoma have not been studied. Thus, we examined the effects of miR-365 overexpression on the endogenous mRNA and proteins levels of BCL2 and CCND1 in A375 and A2058 melanoma cells. As expected, mRNA (Figure 3A) and protein levels (Figure 3B) of BCL2 and CCND1 were significantly downregulated in cells overexpressing miR-365 compared to control cells. To investigate whether BCL2 and CCND1 were direct targets...
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of miR-365 in melanoma we constructed luciferase reporter plasmids containing parts of BCL2 and CCND1 3’UTR with either a wildtype or deleted miR-365 target sites (Figure 3C). As shown by the results of luciferase reporter assays, co-transfection of miR-365 suppressed the luciferase activity of the reporters containing wild-type miR-365 target sites of BCL2 and CCND1 but not that of the reporters containing deleted miR-365 target sites in A375 cells (Figure 3D), indicating that both BCL2 and CCND1 were directly regulated by miR-365 in melanoma cells.

Figure 1. MiR-365 inhibited cell proliferation and induced apoptosis in melanoma. (A) The expression of miR-365 was measured in melanocytes and 4 melanoma cell lines, A375, A2058, SK-MEL-2, and SK-MEL-28 by qRT-PCR. A375 and A2058 were transfected with 100 nM miR-365 mimics or control oligos for 48 hours. (B) The expression levels of miR-365 were measured by qRT-PCR. (C) The cell viability was measured by MTT assay. (D) The colony formation assay was performed to analyze the effects of miR-365 on cell proliferation. In each well of a 6 well plate, 300 cells were plated and incubated for 10–12 days until colonies were visible. Cell cluster with over 50 cells was counted as a colony. (E) Cell cycle was analyzed by using PI staining flow cytometry. (F) Cell apoptosis was measured by using Annexin V-FITC/Hoechst 33258 double staining flow cytometry. Data shown are the mean ±SEM of 3 independent experiments. * P<0.05, ** P<0.01, and ns, no significance.
The effects of miR-365 were partially mediated through targeting CCND1 and BCL2

To investigate the role of CCND1 in miR-365-induced cell cycle arrest, we co-transfected A375 and A2058 cells with miR-365 mimic oligos and a construct encoding CCND1 or an empty vector. We found out that restoration of CCND1 levels eliminated the effects of miR-365 overexpression on cell cycle arrest in both cell lines (Figure 4A). To investigate the role of BCL2 in miR-365-induced cell apoptosis, we co-transfected A375 and A2058 cells with miR-365 mimic oligos and a construct coding BCL2 or an empty vector. The results revealed that co-transfection of miR-365 mimics with BCL2 expressing plasmid led to significant reduction in apoptotic rate in both cell lines (Figure 4B). These data suggested that the anti-tumor effects of miR-365 were at least partially mediated by inhibition of its target genes, BCL2 and CCND1.
Figure 3. MiR-365 targeted BCL2 and CCND1. A375 and A2058 were transfected with 100 nM miR-365 mimics or control oligos for 48 hours. (A) mRNA levels of BCL2 and CCND1 were measured by qRT-PCR. (B) Protein levels of BCL2 and CCND1 were measured using western blot. (C) An illustration of the targeting sites of miR-365 on 3’UTR of BCL2 and CCND1 and the constructs of the luciferase reporter plasmids. (D) The dual-luciferase reporter assay was performed to show that BCL2 and CCND1 were directly targeted by miR-365. A375 cells were co-transfected with indicated oligos and constructs. The Renilla and firefly luciferase activity was then measured using the dual-luciferase kit as instructed by the manufacture’s protocol. Data shown are the mean ±SEM of three independent experiments. * P<0.05, ** P<0.01, and ns, no significance.
GO analysis and KEGG pathway enrichment analysis

To gain further insight into the cellular functions of miR-365, the predicted target genes of miR-365 were retrieved using the miRanda, DIANA-microT and TargetScan database (Figure 5) and subjected to GO analysis and KEGG pathway enrichment analysis. The GO function enrichment analysis explored the functional roles of miR-365 target genes in terms of biological processes, cellular components and molecular functions. The top enriched biological processes associated with "regulation of gene transcription" (Table 1); the top enriched cell components included "nucleus", "nucleoplasm", and "neuron projection" (Table 2); the top enriched molecular functions were involved in "sequence-specific DNA binding", "transcriptional activator activity" and "protein binding" (Table 3). KEGG pathway enrichment analysis was also performed to identify significant pathways enriched in miR-365 target genes. KEGG analysis indicated that these genes were mainly involved in
miR-365 was not correlated with clinicopathological stage and survival of melanoma patients

We then analyzed the SKCM datasets in TCGA for correlation between miR-365 level and clinicopathological stage and the survival of melanoma patients. The results revealed that miR-365 level was not obviously different among patients at different stage (P>0.05, Figure 6A), N stage (P>0.05, Figure 6B), M stage (P>0.05, Figure 6C) and overall clinicopathological stage (P>0.05, Figure 6D). Furthermore, Kaplan-Meier analysis failed to discover significant differences in OS and DFS between patients with high and low miR-365 levels (P>0.05, Figure 6E).

Table 1. The top 10 enriched biological processes in the GO functional analysis.

| GO category | GO term                                                  | Gene number | P-value     |
|-------------|----------------------------------------------------------|-------------|-------------|
| GOTERM_BP   | Positive regulation of transcription from RNA polymerase II promoter | 58          | 2.90E-09    |
| GOTERM_BP   | Transcription, DNA-templated                              | 84          | 7.50E-07    |
| GOTERM_BP   | Positive regulation of transcription, DNA-templated       | 32          | 6.50E-06    |
| GOTERM_BP   | Neuron migration                                          | 13          | 1.40E-05    |
| GOTERM_BP   | Positive regulation of sodium ion transport               | 6           | 1.10E-04    |
| GOTERM_BP   | Axon guidance                                             | 14          | 2.10E-04    |
| GOTERM_BP   | Transcription from RNA polymerase II promoter             | 28          | 2.70E-04    |
| GOTERM_BP   | Small GTPase mediated signal transduction                 | 17          | 5.30E-04    |
| GOTERM_BP   | Negative regulation of cell migration                     | 10          | 6.70E-04    |
| GOTERM_BP   | Regulation of transcription, DNA-templated                | 59          | 6.80E-04    |

Table 2. The top 10 enriched cell components in the GO functional analysis.

| GO category | GO term                                                  | Gene number | P-value     |
|-------------|----------------------------------------------------------|-------------|-------------|
| GOTERM_CC   | Nucleus                                                   | 175         | 3.50E-05    |
| GOTERM_CC   | Nucleoplasm                                               | 101         | 5.80E-05    |
| GOTERM_CC   | Neuron projection                                          | 17          | 5.50E-04    |
| GOTERM_CC   | Cytoplasmic, membrane-bounded vesicle                     | 11          | 2.50E-03    |
| GOTERM_CC   | Cell-cell junction                                        | 12          | 3.60E-03    |
| GOTERM_CC   | Intracellular                                             | 51          | 3.80E-03    |
| GOTERM_CC   | Transcription factor complex                               | 13          | 4.10E-03    |
| GOTERM_CC   | NuA4 histone acetyltransferase complex                    | 4           | 9.30E-03    |
| GOTERM_CC   | Cytosol                                                   | 105         | 1.00E-02    |
| GOTERM_CC   | Membrane                                                  | 72          | 1.20E-02    |

Discussion

Investigation into the molecular mechanisms responsible for melanoma carcinogenesis and progression, especially the exploration of dysregulated miRNAs in melanoma, represents a popular and promising field of study and is essential for developing novel therapeutics.

miR-365 is located in the chromosome region 16p13.12. The expression pattern of miR-365 is cancer type-dependent. MiR-365 was highly expressed in cutaneous squamous cell carcinoma [8,9], breast cancer [32], and invasive pancreatic ductal adenocarcinoma [10]. In contrast, miR-365 was downregulated in colon cancer [11] and lung cancer [12]. The function of miR-365 is complicated, it may function as a tumor suppressor or an oncogene depending on the cancer types. For example, miR-365 was reported to suppress cell cycle progression.
and promote apoptosis of colon cancer cells by probably targeting Cyclin D1 and Bcl-2 [11,33]; miR-365 expression levels were reduced in lung cancer tissues and ectopic miR-365 expression could inhibit cell proliferation of lung cancer cell lines by targeting thyroid transcription factor 1 (TTF-1), which is an essential factor in lung developmental and a prognostic marker for non-small cell lung cancer. However, miR-365 was rarely studied in melanoma. In the only literature studying miR-365 in melanoma, Bai et al. found that the expression of miR-365 was significantly downregulated compared with that in matched normal tissue [13] and overexpression of miR-365 inhibited growth, invasion and metastasis of malignant melanoma through targeting neuropilin1 (NRP1). Therefore, we carried out the present study to further explore the roles of miR-365 in melanoma. In this study, we initially found that miR-365 expression was lower in melanoma cells compared with non-transformed melanocyte (Figure 1A). And the levels of miR-365 was not relevant to the status of BRAF mutation, as it was significantly decreased in both BRAF wildtype cell lines, A375, A2058, and SK-MEL-28, and in BRAF mutant cell lines, SK-MEL-2. Next, we found that overexpression of miR-365 led to significant decrease in cell viability (Figure 1C). Investigation into the underlying mechanism showed that ectopic expression of miR-365 remarkably suppressed clonogenic capacity and induced cell cycle arrest and apoptosis in melanoma cell lines (Figure 1D–1F). In addition, miR-365 significantly compromised the migration and invasion capacities of melanoma cell in vitro (Figure 2A, 2B). These results were in consistence with Bai et al. study in melanoma and suggested that miR-365 functioned as a tumor suppressor in melanoma.
Regarding the target genes of miR-365, a variety of genes have been reported, including CCND1, BCL2, NRP1, TTF-1. Among these genes, CCND1 is considered as an important proliferation-promoting molecule; BCL2 is a fundamental anti-apoptotic gene with a recognized role in cancer development (Figure 4D). However, this regulatory association has not been reported in melanoma. We found that overexpression of miR-365 downregulated the mRNA and protein levels of these 2 genes in A375 and A2058 cell lines (Figure 4A, 4B).

To confirm that CCND1 and BCL2 are direct targets of miR-365, we performed a luciferase reporter assay, showing that CCND1 and BCL2 were directly targeted by miR-365 in melanoma cells (Figure 4E). Finally, to explore the roles of CCND1 and BCL2 in miR-365-mediated cell cycle arrest and apoptosis, we co-transfected melanoma cells with miR-365 mimic oligos and a construct encoding either CCND1 or BCL2. The results revealed that restoration of CCND1 or BCL2 attenuated the effects of miR-365 overexpression on cell cycle and apoptosis, respectively (Figure 5A, 5B). Our result was consistent with Nie et al. study which has shown that BCL2 and CCND1 are direct

Figure 6. MiR-365 was not correlated with clinicopathological stage and survival of melanoma patients. The miRNA-Seq and clinical data from the TCGA SKCM datasets were obtained. (A) The expression levels of miR-365 among different clinicopathological stage were compared using one-way ANOVA. (B) Overall survival and (C) Disease-free survival were assessed by the Kaplan-Meier method and curves were compared by using the log-rank test. Significance was defined as P<0.05.
targets of miR-365 in colon cancer and that knockdown of the endogenous CCND1 or BCL2 was able to mimic the effect of miR-365 [11]. Together, these results suggested that miR-365 probably is a novel tumor suppressor in melanoma through targeting CCND1 and BCL2, as well as other potential targets. We should notice that the regulatory network by miRNAs is very complicated. One gene is regulated by multiple miRNAs, and a specific miRNA can regulate multiple genes. Therefore, other target genes in addition to CCND1 and BCL2 may also participate in the functions of miR-365.

This notion highlighted the need for further studies to reveal the entire “targetome” of miR-365 in the development of melanoma. To gain further insight into the cellular functions of miR-365, we then performed the GO functional enrichment analysis and KEGG pathway enrichment analysis for the predicted target genes of miR-365 retrieved from the miranda, DIANA-microT and TargetScan database. GO function enrichment analysis revealed that the most prominent functions of miR-365 target genes were involved in regulation of gene transcription. KEGG pathway enrichment analysis indicated that miR-365 target genes were mainly involved in cancer-related pathways. Thus, analysis of the “targetome” of miR-365 further revealed the association of miR-365 with cancer and dysregulated gene transcription may contribute to the function of miR-365.

We then analyzed the TCGA SKCM dataset for correlation between miR-365 levels and clinicopathological stage or the prognosis of melanoma patients. The results revealed that miR-365 levels were not obviously different among patients with different T stage, N stage, M stage, and overall clinicopathological stage (all P>0.05, Figure 5A). Furthermore, Kaplan-Meier analysis failed to discover significant difference in OS and DFS between patients with high and low miR-365 levels (both P>0.05, Figure 5B, 5C). The prognosis of patients can be influenced by many factors, such as disease stage and treatment. MiR-365 is just one of the many factors, therefore, its effect on patient outcome may be covered or counteract by the effect of other factors, although miR-365 showed obvious biological effects in melanoma cell lines. The discrepancy between in vitro and in vivo data again highlighted the complexity of miRNA regulatory network and further studies are guaranteed to completely understand the roles miR-365 in melanoma development.

Conclusions

In sum, our data suggested that miR-365 was downregulated in melanoma cells and played a tumor suppressive role in melanoma development through regulating multiple target genes that are essential to key cellular functions, such as cell proliferation, apoptosis, migration, and invasion. However, the TCGA data failed to discover the association of miR-365 with progression and development of melanoma patients. Further studies in animal models and human samples are required for the full understanding of the roles miR-365 in melanoma development.

Conflict of Interest

None.
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