Relationship of ethnicity and CD4 Count with glucose metabolism among HIV patients on Highly-Active Antiretroviral Therapy (HAART)

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Abstract

Background: HIV patients on HAART are prone to metabolic abnormalities, including insulin resistance, lipodystrophy and diabetes. This study purports to investigate the relationship of ethnicity and CD4+ T cell count attained after stable highly-active antiretroviral treatment (HAART) with glucose metabolism in hypertriglyceridemic HIV patients without a history of diabetes.

Methods: Demographic, anthropometric, clinical, endocrinologic, energy expenditure and metabolic measures were obtained in 199 multiethnic, healthy but hypertriglyceridemic HIV-infected patients [46% Hispanic, 17% African-American, 37% Non-Hispanic White (NHW)] on stable HAART without a history of diabetes. The relationship of glucose and insulin responses to ethnicity, CD4 strata (low (<300/cc) or moderate-to-high (≥ 300/cc)), and their interaction was determined.

Results: African-Americans had significantly greater impairment of glucose tolerance (P < 0.05) and HbA1c levels (P < .001) than either Hispanics or NHWs. In multivariate models, after adjusting for confounders (age, sex, HIV/HAART duration, smoking, obesity, glucose, insulin and lipids), African-Americans and Hispanics had significantly higher HbA1c and 2-hour glucose levels than NHW's. Demonstrating a significant interaction between ethnicity and CD4 count (P = 0.023), African Americans with CD4 <300/cc and Hispanics with CD4 ≥300/cc had the most impaired glucose response following oral glucose challenge.

Conclusions: Among hypertriglyceridemic HIV patients on HAART, African-Americans and Hispanics are at increased risk of developing diabetes. Ethnicity also interacts with CD4+ T cell count attained on stable HAART to affect post-challenge glycemic response.

Keywords: African American, Hispanic, Impaired glucose tolerance, HbA1c, Dyslipidemia

Background

HIV/AIDS affects U.S. minorities disproportionately, reflecting a change in the distribution of the disease by racial/ethnic groups since the beginning of the epidemic [1,2]. The widespread use of highly-active antiretroviral therapy (HAART) has slowed the progression of HIV infection to AIDS and has reduced associated morbidity and mortality [3]. However, HIV patients on HAART are prone to metabolic abnormalities, including insulin resistance, lipodystrophy and diabetes (1). Lipodystrophy - characterized by peripheral fat loss, visceral fat increase, hypertriglyceridemia, and low HDL-C - is often associated with insulin resistance [4,5], a risk factor for developing diabetes. Although the frequencies of impaired glucose tolerance (IGT) and diabetes have not been systematically evaluated in HIV patients of different ethnicities and geographic areas, the rates are likely to be higher than in the general population. An early study of HIV patients with lipodystrophy and abdominal fat accumulation reported a 35% prevalence of IGT and 8% prevalence of diabetes, compared to 5% and 0.5% respectively.
for matched non-HIV controls [4]. The high frequency of obesity (an independent risk factor for metabolic abnormalities and CVD) among HIV/HAART patients has an inimical effect on their immune reconstitution and body composition [6].

Diabetes and cardiometabolic syndrome are highly prevalent among ethnic minorities in non-HIV infected populations [7]. The relationship between HbA1c and fasting serum glucose is not homogenous across racial/ethnic groups: e.g., African-Americans and Hispanics have higher HbA1c than non-Hispanic Whites (NHWs) after adjusting for fasting glucose concentration, glucose area under the curve after oral glucose, insulin response and insulin resistance [8,9]. Possible important interactions between ethnicity and specific aspects of glucose metabolism may be obscured in epidemiologic studies that utilize limited parameters (e.g., only fasting serum glucose, or HbA1c) to measure dysglycemia: e.g., significant differences have been noted in HbA1c levels between African-Americans and NHWs at elevated serum glucose levels [8]. Among HIV patients, elevated HbA1c has been associated with higher CD4 cell count, older age, and ethnicity [10]. Similar data from relatively small studies and heterogeneous HIV populations suggest that metabolic (including glycemic) abnormalities among HIV patients may also be affected by ethnicity [11,12]. However, in the context of diabetes risk, the relationship of ethnicity and its interactions with metabolic parameters and the degree of immune reconstitution following HAART have not been investigated systematically in HIV patients.

The purpose of this study was to assess the relationship of ethnicity with several simultaneously measured glycemic parameters - HbA1c, fasting serum glucose and insulin, and post-challenge serum glucose and insulin levels - among hypertriglyceridemic HIV patients on stable HAART who participated in the Heart Positive study. We hypothesized that ethnicity and CD4 level on stable HAART would exert an influence on glycemic regulation in these hypertriglyceridemic but otherwise healthy HIV patients.

Methods
Heart Positive (Clinicaltrials.gov ID NCT00246376) was a large multiethnic study of healthy patients with HIV infection on stable HAART [13]. The primary outcomes have been reported [13]. The protocol was approved by the Institutional Review Boards of Baylor College of Medicine and the two recruitment centers. Written informed consent was obtained (in Spanish or English) from all subjects prior to their participation.

Subjects
HIV patients with no history of diabetes were recruited from two HIV centers in Houston: Legacy Community Health Services and Thomas Street Clinic, Harris County Hospital District. The sample comprised 202 patients on stable HAART with hypertriglyceridemia who underwent complete screening for the Heart Positive study. Racial/ethnic distribution of the subjects included NHWs, African Americans, Hispanics, and Native Americans. Three Native American subjects were excluded from this analysis due to their small numbers. All subjects were hypertriglyceridemic (fasting plasma triglycerides $>150$ mg/dL), on no lipid-lowering or anti-diabetic medications, on stable HAART for at least 6 months, with stable CD4 count and viral load, normal kidney and thyroid functions, and transaminase (AST / ALT) levels $\leq 2$ times upper limit of normal.

Measures

Demographic and clinical variables
Age, gender, ethnicity, number of years since HIV diagnosis, protease inhibitor (PI) use, smoking, alcohol use, drug abuse, and duration of HAART.

Body composition
Was measured by bioelectrical impedance analysis (Quantum II Analyzer, RJL Systems, Clinton Township, MI). Body Mass Index and waist and hip circumference were measured.

Glucose and insulin
Subjects ingested 75 g glucose (SunDex, Fisher Health Care, Houston, TX) in the morning after a 10 h overnight fast, with blood sampling at 0 (fasting), 30, 60, 90 and 120 minutes. Plasma glucose was measured by the glucose oxidase method and insulin by radioimmunoassay (Linco, St. Louis, MO). HbA1c was measured by HPLC at The Methodist Hospital clinical laboratory, Houston, TX.

CVD risk markers
Total cholesterol, HDL-C and triglyceride profiles were determined by an automated enzymatic method (Olympus AU400e Analyzer, Center Valley, PA) at the Baylor Atherosclerosis Core Laboratory (CLIA-CAP certified). LDL-C was calculated using the Friedewald formula. Fasting plasma total cholesterol, HDL-C and triglycerides were measured in the Baylor Atherosclerosis Clinical Laboratory using an Olympus AU400e automated analyzer. The median interassay c.v.’s for these assays were $< 2$–5%, and the median intra-assay c.v.’s were $< 2$–8%.

Energy expenditure
Resting energy expenditure (REE) was measured at the Baylor General Clinical Research Center (GCRC) by indirect calorimetry (Deltratrac, Sensormedics, Fullerton, CA) and calculated as $\text{REE} = \text{VO}2 \times [4.686 + (\text{RQ}-0.707) \times \text{KcM} / \text{Kg}]$.
0.361/0.293 [14]. For subjects who could not visit the GCRC (n = 68), REE was measured using a MedGem device (Microlife USA, Dunedin, FL) that measures VO₂ and calculates REE using a modified Weir equation.

**CD4+ T cell count and HIV-1 viral load**

CD4 count was measured by flow cytometry. To assess the relationship of CD4 levels to glucose abnormalities, subjects were divided into two groups: “low” (<300/cc; n = 53), and “moderate-high” (≥300/cc; n = 147). The cutoff of 300/cc was based on the World Health Organization’s current recommendation to the increase the threshold for the use of antiretroviral therapy (ART) for HIV-infected individuals to a CD4 count of ≤350 cells/ cc (from the previous level of ≤200 cells/μL) [15]. In view of these changes that occurred during the period of the Heart Positive study, we used a cutoff slightly lower than that of the WHO to define the CD4 strata. HIV-1 viral load (VL) was measured by LabCorp or Quest Diagnostics (Madison, NJ) using either of two quantitative real-time PCR assays (routine, with a lower limit of 400 copies/cc, or ultrasensitive, with a lower limit of 50 copies/cc).

VL values measured as <400/cc by the first assay were assigned a value of 200/cc and all the VL values were log-transformed prior to analysis.

**Statistical analysis**

Descriptive measures, chi-square (χ²) analysis and ANOVA were used to examine associations and differences in metabolic, hormonal, anthropometric and clinical parameters by ethnicity and CD4 strata. When differences in groups were observed, Bonferroni Post-Hoc analysis was performed to determine which groups were different from each other. Repeated measures ANOVA was used to assess the relationship of ethnicity with glucose measures following the oral challenge. Co-variates were age, sex, BMI, lean mass, REE/lean mass, HAART duration, PI usage, HIV duration, fasting glucose level and fasting insulin level. Two-way interactions between ethnicity and glucose, ethnicity and gender, ethnicity and age, and ethnicity and CD4 count were included in the model and removed if non-significant (p > 0.05). Linear regression was performed with fasting serum glucose, 2-hour glucose, and HbA1c as outcome variables, and age, sex, BMI, waist, lean mass, REE/lean mass, HAART duration, PI usage, HIV duration, fasting glucose, HOMA-IR, systolic and diastolic blood pressure, family history of diabetes and interactions as independent variables. To avoid collinearity, BMI and lean mass (and not waist-hip-ratio and fat mass) were included in the final model. All statistical analyses were performed using the Statistical Package for Social Sciences (SPSS) (version 16.0, SPSS Inc, Chicago).

**Results**

Table 1 summarizes demographic, anthropometric and clinical parameters of the patients by ethnicity and CD4 strata. Mean age was 43.6 ± 8.3 years, duration since HIV diagnosis 9.5 ± 5.9 years, and duration of HAART 5.6 ± 4.4 years. One quarter of the subjects (26%) had CD4 <300 / ml. Ethnic distribution was 46% Hispanic, 17% African-American and 37% NHW. The majority of the patients were indigent and uninsured. Plasma HIV-1 viral load was undetectable in 74%. There was no difference in the frequency of patients with undetectable viral load with respect to ethnic group or CD4 stratum. Fourteen subjects had HIV/HCV co-infection. However, mean fasting glucose was similar between those with and without the co-infection [P = 0.204].

Hispanics were significantly younger, had shorter HIV infection duration, fewer years of HAART and lower systolic blood pressure, and reported a lower rate of smoking, drug use and family history of diabetes, than African-Americans or NHWs (based on Bonferroni Post-Hoc analysis). Furthermore, they had significantly lower weight, height and body cell mass than African-Americans or NHWs. African-Americans had significantly more fat mass than Hispanics or NHWs (P < 0.001). Both African-Americans and Hispanics had higher BMI and percent body fat than NHW’s. NHWs had higher resting VCO₂, VO₂ and REE than Hispanics or African-Americans (P < 0.001). There were no significant ethnic differences in frequency of PI use or in fasting plasma levels of lipids, adiponectin, hsCRP, leptin, and free fatty acids. or.

No significant differences were noted for frequency of use of first generation PI drugs (indinavir, nelfinavir, saquinavir) compared to more recent PI drugs (atazanavir, darunavir), or for use of thymidine analogue drugs, with respect to ethnic group or CD4 stratum. No differences were noted between the low and moderate-high CD4 groups in demographic / clinical / anthropometric or endocrine / hormonal / energy expenditure parameters except for a higher frequency of PI use and lower body cell mass among subjects with moderate-high CD4 count, higher testosterone level in the low CD4 group, and higher VO₂ with a trend towards higher VCO₂ in the low CD4 group.

Insulin sensitivity and glycemic parameters by ethnicity and CD4 cell count are presented in Table 1. The 2-hour glucose value in the OGTT differed significantly by ethnicity but not by CD4 strata. IGT (defined by 2-h glucose level >140 but <200 mg/dl) was observed in 32% of the subjects - these were significantly older than those with normal glucose tolerance (P < 0.001). Compared to either Hispanics or NHWs, African-Americans manifested greater impairment of glucose tolerance, as indicated by higher 2-hour glucose (P < 0.05) and HbA1c.
| Variables | Total (n = 199) | CD4 Count* | CD4 Count* | F-Value | P | Ethnicity | Ethnicity | Ethnicity | F-Value/χ² value | P |
|-----------|----------------|------------|------------|---------|---|-----------|-----------|-----------|-----------------|---|
| Low (<300/cc; n = 53) | Moderate-High (≥300; n = 146) | | | | | | | | | |
| Age | 43.6 ± 8.3 | 43.7 ± 7.6 | 43.5 ± 8.5 | .026 | .871 | 46.9 ± 8.1 | 45.9 ± 7.1 | 39.9 ± 7.4 | 19.449 | <.001 |
| HIV duration (y) | 9.8 ± 5.9 | 9.9 ± 6.2 | 9.4 ± 5.9 | .215 | .643 | 12.7 ± 5.9 | 7.8 ± 5.1 | 7.4 ± 5.1 | 21.803 | <.001 |
| HAART duration (y) | 5.6 ± 4.4 | 5.3 ± 5.1 | 5.7 ± 4.1 | .417 | .519 | 7.1 ± 5.2 | 5.1 ± 4.1 | 4.6 ± 3.5 | 6.955 | .001 |
| PI Drug Use | 67.8% | 61.0% | 86.5% | 11.46 | .001 | 73.0% | 66.7% | 64.1% | 1.49 | .474 |
| Family History of Diabetes | 42.7% | 43.1% | 42.5% | .007 | .933 | 28.4% | 36.4% | 57.1% | 14.48 | .001 |
| Smoking | 61.8% | 65.4% | 61.0% | .319 | .572 | 75.7% | 69.7% | 47.8% | 14.51 | .001 |
| Drug Use | 28.1% | 28.8% | 28.1% | .011 | .916 | 35.1% | 36.4% | 19.6% | 6.24 | .044 |
| SBP (mm Hg) | 128.3 ± 14.4 | 130.5 ± 15.7 | 127.5 ± 13.9 | 1.741 | .188 | 131.5 ± 15.3 | 130.8 ± 15.3 | 125.5 ± 13.4 | 4.050 | .019 |
| DBP (mm Hg) | 82.4 ± 10.3 | 83.4 ± 11.4 | 82 ± 9.9 | .730 | .394 | 84.3 ± 11.2 | 82.1 ± 10.9 | 81.2 ± 9.3 | 1.914 | .150 |
| Log Viral load | 2.41 ± .40 | 2.41 ± .48 | 2.41 ± .38 | .002 | .961 | 2.36 ± .40 | 2.48 ± .08 | 2.41 ± .39 | .920 | .400 |
| Total Chol. (mg/dL) | 211.4 ± 48.5 | 207.6 ± 43.5 | 212.8 ± 50.2 | .441 | .507 | 218.1 ± 58.1 | 201.9 ± 42.0 | 208.3 ± 41.9 | 1.520 | .221 |
| TG (mg/dL) | 302 ± 163 | 321 ± 145 | 295 ± 169 | 1.030 | .311 | 323 ± 163 | 247 ± 118 | 302 ± 172 | 2.567 | .079 |
| HDL-C (mg/dL) | 39.7 ± 8.9 | 38.1 ± 8.3 | 40.3 ± 9.2 | .084 | .920 | 39.6 ± 9.5 | 39.2 ± 10.1 | 39.9 ± 8.4 | 19.6% | .364 |
| LDL-C (mg/dL) | 111.9 ± 40.4 | 105.3 ± 37.7 | 114.3 ± 41.2 | .1927 | .167 | 113.9 ± 43.7 | 113.3 ± 33.4 | 108.7 ± 41.2 | .369 | .692 |
| Metabolic | | | | | | | | | | |
| FFA (mmol/L) | .32 ± .22 | .29 ± .17 | .33 ± .24 | | | | | | | |
| Leptin (ng/mL) | 6.8 ± 5.9 | 5.5 ± 4.8 | 7.2 ± 6.3 | | | | | | | |
| Adiponectin (μg/mL) | 6.67 ± 5.39 | 7.40 ± 5.81 | 6.31 ± 5.16 | | | | | | | |
| hsCRP (mg/L) | 3.73 ± .548 | 3.51 ± .586 | 3.80 ± .533 | | | | | | | |
| Percent body fat (%) | 22.1 ± 8.2 | 20.5 ± 6.3 | 22.7 ± 8.8 | .010 | .916 | 23.3 ± 8.8 | 23.3 ± 8.8 | 23.3 ± 8.8 | .367 | .044 |
| BMI (kg/m²) | 26.8 ± 3.5 | 26.3 ± 3.2 | 27 ± 3.6 | | | | | | | |
| Waist circ. (cm) | 91.5 ± 11.7 | 90.2 ± 14 | 91.9 ± 10.7 | | | | | | | |
| Waist-hip ratio | .9 ± .1 | .9 ± .1 | .9 ± .09 | | | | | | | |
| VCO₂ (mL/min) | 221 ± 42.4 | 231.3 ± 44.3 | 216.8 ± 41 | | | | | | | |
| VO₂ (mL/min) | 265.3 ± 48.3 | 279.7 ± 48.4 | 259.8 ± 47.4 | | | | | | | |
| RQ | .83 ± .06 | .83 ± .07 | .84 ± .06 | | | | | | | |
| REE (kCal/24 h) | 1819.1 ± 357 | 1897 ± 320 | 1789.8 ± 367 | | | | | | | |
| Glycemia, Insulin Sensitivity | | | | | | | | | | |
| Insulinogenic Index | .59 ± .4 | .62 ± .4 | .64 ± .4 | | | | | | | |
| HOMA-IR | 3.3 ± 3.9 | 3.5 ± 4.1 | 3.3 ± 3.9 | | | | | | | |
| Fasting glucose (mg/dL) | 94.7 ± 14.2 | 95 ± 14.03 | 94.6 ± 14.4 | | | | | | | |
| OGTT Glucose 120 min (mg/dL) | 127.9 ± 41.7 | 124.7 ± 47.7 | 129.1 ± 39.4 | | | | | | | |
| Fasting Insulin (μU/ml) | 15.4 ± 25.6 | 14.7 ± 18.6 | 15.6 ± 27.8 | | | | | | | |
| OGTT Insulin 120 min (μU/ml) | 83.7 ± 76 | 75.1 ± 74.3 | 86.8 ± 76.7 | | | | | | | |
| HbA1c (%) | 5.3 ± 6 | 5.3 ± 6 | 5.3 ± 6 | | | | | | | |

Data are mean ± SE or percent.
(P < 0.001) levels. Hispanics had higher insulin levels at 90 min and 120 min of the OGTT than NHW’s or African-Americans. No ethnic difference was noted in calculated measures of insulin resistance such as area under the curve (AUC) for insulin or the insulinogenic index.

A linear regression model was fitted to determine parameters associated with HbA1c, fasting glucose and 2-hour glucose (Table 2a). Fasting glucose, age, female sex and ethnicity were significant predictors of HbA1c (R² = 0.395; F < .001). After adjusting for known confounders, HbA1c was significantly higher among African-Americans and Hispanics than among NHWs. After adjusting for confounders, fasting glucose levels were predicted only weakly by the Homeostasis Model Assessment measure of insulin resistance (HOMA-IR) and by PI use (R² = 0.153; F = .012). However, 2-hour glucose level (adjusted) was strongly and significantly predicted by age, female sex, ethnicity and fasting glucose, (R² = 0.565; F < .001). Hispanics and African-Americans had significantly higher 2-hour glucose levels than NHW’s. In the regression model, for the 2-hour glucose level, the significant interaction effect of ethnicity with fasting glucose (P < 0.001) highlighted differences among the three ethnic groups (NHW’s < African Americans < Hispanics) in this glycemic parameter. Outcomes of the regression model for the glucose response curves in the OGTT are shown in Table 2b. Age, female sex, HOMA-IR and AUC insulin were significant predictors in the model (R² = 0.392; F < 0.001). There was a significant interaction of ethnicity with CD4 strata.

Multivariate analysis (repeated measures ANOVA) assessed the effect of ethnicity on the OGTT glucose response curves after controlling for confounders (age, sex, race, obesity, fasting glucose, HOMA-IR, family history of diabetes, blood pressure, duration of HIV, and PI drug use) and also measured interactions of ethnicity with age, sex and CD4 strata. Ethnicity alone did not distinguish the glucose response curves in a statistically significant manner. Interaction effects of ethnicity*sex, and ethnicity*age were also not significant, but an interaction between ethnicity and CD4 strata was significant for the glucose response curves (P = 0.023). Interaction effects of ethnicity with CD4 count imply that the effect of ethnicity is modified by the CD4 count of the subjects. To elucidate how ethnicity affects the relationship between glycemic excursions and CD4 levels, we plotted post-challenge glucose data against ethnic groups for each CD4 stratum (Figure 1). In the moderate-high CD4 group, Hispanic subjects had the highest mean post-challenge glycemic excursion (Figure 1a), while in the low CD4 group, African-Americans had the highest mean post-challenge glycemic excursion (Figure 1b).

### Table 2 Multivariate regression models: factors associated with (a) HbA1c, Fasting Glucose, 2-Hour Glucose, and (b) glucose response curves in the OGTT

| Variable                  | Standardized Regression Coefficient | P-value |
|---------------------------|-------------------------------------|---------|
| (a) HbA1c                 |                                     |         |
| Fasting Glucose           | .533                                | .011    |
| Female (versus male)      | .356                                | .005    |
| Age                       | .193                                | .003    |
| Ethnic Category           | 3.01                                | .038    |
| White Referent            |                                     |         |
| African American          | .322                                | .005    |
| Hispanic                  | .167                                | .039    |
| (b) Fasting Glucose       |                                     |         |
| HOMA-IR                   | .325                                | <.001   |
| PI Drug                   | -.154                               | .050    |
| (b) 2-Hour Glucose        |                                     |         |
| Fasting Glucose           | .325                                | .010    |
| Age                       | .292                                | .032    |
| Female (versus male)      | .360                                | .003    |
| HOMA-IR                   | .141                                | .005    |
| Ethnic Category           | .146                                | <.001   |
| White Referent group      |                                     |         |
| African American          | -.126                               | .014    |
| Hispanic                  | -.380                               | <.001   |
| Ethnicity*Fasting Glucose | .165                                | <.001   |
| (b) Glucose Response      |                                     |         |
| Curves in OGTT            |                                     |         |
| Age                       | .276                                | <.001   |
| Female (versus male)      | .106                                | .031    |
| HOMA-IR                   | .136                                | .062    |
| AUC Insulin               | .145                                | .080    |
| Ethnicity *CD4            | -.172                               | .023    |

### Discussion

This is the first study to assess ethnic differences in glycemic parameters among HIV patients on a stable HAART regimen, specifically those with hypertriglyceridemia. The selection criteria for the Heart Positive study permitted us to investigate metabolic factors that differentiate glycemic control between ethnic groups of HIV patients without a history of diabetes, and to analyze a large data set of both fasting and post-prandial glucose responses. In bivariate analyses, African-Americans, Hispanics and NHW’s had similar fasting glucose levels, but African-Americans had higher HbA1c and 2-hour OGTT glucose values than Hispanics and NHWs. Significant
ethnic differences noted in the multivariate models for 2-hour glucose and HbA1c suggested that ethnicity affects postprandial glycemic control in HIV patients on HAART. These effects persisted after adjusting for group differences in age, sex, HIV years, HAART years, smoking, obesity, fasting glucose, fasting insulin, and lipid levels. Overall, African-Americans and Hispanics had higher glucose values after oral glucose challenge as well as higher HbA1c levels than NHW's, and an interaction between ethnicity and CD4 count modulated the glycemic response.

A previous study demonstrated that HIV-infected African American women have higher HbA1c levels than NHW's [10]. While controlling for ethnicity, age and CD4 count provided a reasonably accurate reflection of glycemic control in that study, the present results indicate that the greater impairment in glycemic control among HIV infected African-Americans is not due to unmeasured or suboptimally measured risk factors. A robust model that included all possible risk factors (demographic, anthropometric, clinical, and metabolic parameters) improved our confidence in the existence of ethnic disparities in specific glycemic parameters in this cohort who are at high risk for metabolic complications. It is possible that the effect of ethnicity in part “pre-dated” HIV infection or HAART therapy, since African American and

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Figure 1 Glucose response curves for HIV-infected African-Americans, Hispanics and NHW's in the oral glucose tolerance test. Multivariate models revealed a significant interaction between ethnicity and CD4+ T cell stratum that affected the glucose response curves ($P = 0.023$). a. Glucose response curves of patients with CD4+ T cell counts ≥ 300/cc on stable HAART; b. Glucose response curves of patients with CD4+ T cell counts < 300/cc on stable HAART.
Hispanics the general US population is known to have increased risk for impaired glucose tolerance and diabetes compared to NHW’s. The cross-sectional design of this study and absence of a HIV-negative control group did not permit us to estimate the specific effects of HIV / HAART on the risk of dysglycemia. However the interaction between ethnicity and CD4 level suggests that specific effects related to the immune status resulting from chronic HIV infection and its treatment magnify the risk of dysglycemia in minority ethnic groups.

Previous studies have shown inconsistent associations of CD4 cell count with abnormal glucose levels in HIV patients. Low CD4 cell count was associated with IGT and diabetes in patients co-infected with HIV, hepatitis C and hepatitis B virus [16]. Estrada et al. reported that patients with HIV-associated lipodystrophy and IGT have lower CD4 counts than normoglycemic patients [17]. El-Sadr et al. found an inverse relationship between CD4 counts and insulin resistance in HIV patients [18], but no relationship between CD4 counts and fasting glucose concentrations. This finding is consistent with our data which showed CD4-related differences relate to post-challenge glucose levels, not to fasting glucose levels. Shikuma et al. reported no association between fasting glucose levels and CD4 levels but did not measure post-challenge glucose [19]. In contrast to our results demonstrating no differences in either fasting or post-challenge insulin levels, these investigators also noted that their small sample (7–8 subjects in the respective groups) of HIV patients with CD4 <200/cc had higher fasting insulin levels than those with CD4 >500/cc.

The design of this study makes it difficult to specify the reasons for the relationships between ethnicity, CD4 counts and glucose tolerance in these patients. However, the literature suggests potential mechanisms that should be investigated in future studies. Strictly speaking these results may be generalizable to only to HIV patients with hypertriglyceridemia, however such patients comprise a large proportion of those with HIV infection on various forms of antiretroviral therapy. Additional limitations include the fact that not all ART drug classes or single drugs that could possibly affect insulin sensitivity were included in the multivariate analyses. However, drugs of the protease inhibitor class (which are the most strongly associated with both insulin resistance and defects in glucose uptake) were taken into consideration (i.e., use of PI was controlled for in the analyses).

Abnormal glycemic responses following an oral glucose challenge could be due to insulin resistance or impaired insulin secretion. HOMA-IR, a marker of insulin resistance calculated from fasting values of glucose and insulin, did not differ between patients of the different CD4 strata, suggesting that an aspect of the glycemic defect might involve defective insulin response to an oral challenge. Consistent with this notion, AUC insulin following the oral challenge and the insulinogenic index (a measure of insulin response relative to the glucose surge) did not differ between the two CD4 strata, indicating that persons with both low and high CD4 cell counts may have inadequate insulin secretory response in the face of higher post-challenge glucose levels. It is possible that either low or high CD4 levels might reflect underlying immunologic mechanisms that could affect beta cell mass or function. These possibilities are supported by recent studies showing that, compared to healthy non-HIV persons, HIV patients have higher rates of IGT [20] without necessarily having greater degrees of insulin resistance [21].

While the CD4+ T cell level (low or moderate-high) achieved on a stable HAART regimen was not by itself associated with dysglycemia, ethnicity appeared to interact with low or moderate-high CD4 levels to affect glycemic response to an oral glucose challenge. Activated CD4 cells play a significant role in non-insulin mediated glucose disposal [22]. The association of dysglycemia with low CD4 levels could be due to impaired non-insulin mediated glucose disposal caused by a diminished “sink” of activated CD4 cells (normally equal in total mass to perhaps a third of the liver) or to a greater degree of chronic systemic inflammation. Association of defective glucose disposal with high CD4 counts might be due to post-HAART recovery of CD4 T cells that are unable to utilize glucose effectively because of low GLUT-1 or IL7-R expression or to the influence of genetic factors such as CCL3L1 or CCR5 polymorphisms known to be associated with the development of type 1 diabetes [23]. These findings will be important in formulating hypotheses for an immunologic or molecular basis of impaired glucose metabolism in HIV patients.

These findings have prominent implications for clinical practice. Ethnicity should be considered to affect risk for developing dysglycemia and Metabolic Syndrome among HIV patients. In the U.S., the disproportionately higher burden of HIV/AIDS coupled with the projected population growth among African Americans and Hispanics underscores the need for early screening and intervention as a cost-effective public health strategy [24,25]. Consistent with non-HIV population data [26], we found that African-American ethnicity is associated with IGT, both independently and through an interaction with low CD4 count. Earlier studies investigating the association of ethnicity with IGT or insulin resistance in HIV patients have shown inconsistent results. NHW ethnicity, Hispanic ethnicity, and non-Caucasian race each have been shown to increase risk for insulin resistance or IGT [11]. However, El-Sadr et al. [18] actually found an inverse relationship between African-American ethnicity and insulin resistance in a large HIV cohort, while the Nutrition For Healthy Living study...
found no relation between ethnicity and insulin resistance in HIV patients [21]. These studies involved predominantly NHW’s [18], whereas our cohort had a mixed ethnic distribution resembling that of the overall US population.

Development of ethnically focused and culturally appropriate treatment approaches is highly relevant to clinical management of the growing HIV epidemic among minorities. The rate of African Americans living with HIV infection (1,202.2) is 8 times higher than that of NHWs (156.7) and 3 times higher than that of Hispanics (487.3) per 100,000 population [24]. HIV’s economic burden is predicted to grow markedly, based on the projected dramatic increase in the minority population as well as in HIV infections and complications associated with HAART [27]. In this context, the present findings in a large, multiethnic cohort of hypertriglyceridemic but otherwise apparently healthy HIV patients on stable HAART have important clinical implications for targeted therapy of HIV/HAART-associated glycemic complications.

Conclusion
Among hypertriglyceridemic HIV patients on HAART, African-Americans and Hispanics are at increased risk of developing diabetes. Ethnicity also interacts with CD4+ T cell count attained on stable HAART to affect post-challenge glycemic response. Clinicians should understand ethnic-specific factors in developing treatment strategies for dysglycemia, and should be aware of the need for vigilant screening and possibly earlier treatment for HIV-infected African-Americans with low CD4 count and Hispanics with high CD4 count following HAART.

Abbreviations
(HAART): Highly-active antiretroviral therapy; (BMI): Body mass index; (IGT): Impaired glucose tolerance; (PI): Protease inhibitors.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
RM carried out the data analysis and drafted the manuscript; PC completed the literature review; SR carried out the data analysis and edited the manuscript; DL carried out data analysis and edited the results section; SS finalized the data entry and cleaning; HP completed the laboratory analysis and edited the draft; IC participated in subject recruitment and coordinated the project; DL participated in manuscript writing and edited the drafts; RS edited the manuscript draft; AB supervised the project and drafted/editied the manuscript. All authors read and approved the final manuscript.

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