Comparative Morphology of the Papillae Linguales and their Connective Tissue Cores in the Tongue of the Greater Japanese Shrew-mole, Urotrichus talpoides

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Summary

The external morphology of the papillae linguales (papillae filiformes, papillae fungiformes and papillae vallatae) and their connective tissue cores (CTCs) of the greater Japanese shrew-mole (Urotrichus talpoides) were analysed by optical and scanning electron microscopy. Papillae filiformes were distributed over the dorsal surface of the apex linguae, and on the rostral and caudal regions of the corpus linguae but were less numerous in the mid-region. They were absent from the radix linguae. A pair of oval papillae vallatae was situated at the border between the corpus linguae and the radix linguae. Papillae foliatae were absent. The epithelial surface of each papilla filiformis consisted of a circular concavity, a ring-like wall and either a single thumb-like process or 2–3 slender pointed processes, depending on their location. The morphology of the CTCs of the papillae linguales also varied regionally. The papillae linguales of the Japanese shrew-mole were morphologically similar to those of other Talpidae and Soricidae, including the common shrew, particularly with respect to the papillae filiformes in the mid- and caudal regions of the corpus linguae.

Introduction

The mammalian family Talpidae (moles and shrew-moles), within the order Soricomorpha (shrews), contains well-known living representatives. They were formerly placed within the ‘Insectivora’ but the systematics of this grouping is problematic, particularly with respect to the position of the poorly known Asiatic (Japanese) shrew-moles (Symonds, 2005). Molecular phylogenetic studies (Tsuchiya et al., 2000; Nikaido et al., 2003; Shinohara et al., 2003) have shown that the Japanese shrew-mole (Urotrichus) was phylogenetically distant from European and Japanese moles (Talpa and Mogera, respectively). Consequently, the systematics of the subfamilies of the Talpidae remains controversial (Wilson and Reeder, 2005).

Many studies have demonstrated morphological diversity among the oral tissues of mammals (Nickel et al., 1979), particularly with respect to the three-dimensional structure of the papillae linguales and their connective tissue cores (CTCs) in terrestrial species (Kobayashi et al., 1989, 2005; Kobayashi, 1992; Adnyane et al., 2011; Watanabe et al., 2011). Following the early macroscopic studies of insectivores by Sonntag (1923), there have been several detailed morphological investigations into the papillae linguales of the Soricomorpha, for example, Talpidae such as Mogera (Kobayashi et al., 1983; Miyata et al., 1990), Talpa (Jackowiak, 2006) and Dymecodon (Kobayashi et al., 1983); and Soricidae including Sorex, (Kobayashi and Iwasaki, 1989; Jackowiak et al., 2004), Suncus (Kobayashi et al., 1983; Kobayashi and Iwasaki, 1989), Chimarrogale (Kobayashi and Iwasaki, 1989) and Crocidura (Kobayashi (1992), Kobayashi et al. (1989, 2005). However, information is lacking for the papillae linguales and for their underlining CTCs of the Japanese shrew-moles (Urotrichus).
The aim of this study was to analyse in detail the surface morphology of the papillae linguales on the dorsal surface of the tongue of the Japanese shrew-mole and, after exfoliation of the epithelium of their CTCs, to compare them with earlier descriptions of Talpidae and other Soricomorpha.

Materials and Methods

Animals and tissue preparation

Eight Japanese shrew-moles Urotrichus talpoides (three males, body length 10.3–10.8 cm, 12.5–15.0 g; five females, 10.2–10.8 cm, 12.3–15.5 g) that had died in road accidents were used for this study. They were fixed with 10% formalin for post-mortem autopsy and, shortly afterwards, tissue blocks were excised from various regions of the tongue.

Light microscopy

Tissue samples were dehydrated with a graded ethanol series, embedded in paraffin wax and sectioned at 4 μm. Sections were stained with haematoxylin–eosin and observed by bright-field microscopy (BH-2; Olympus, Tokyo, Japan).

Scanning electron microscopy

Tissue samples were immersed in 3.5 N HCl for 5 days at room temperature (25–28°C). The epithelium was then exfoliated from the underlying CTCs. Specimens were washed with tap water and treated with a 0.5% tannic acid solution. Post-fixation was accomplished by immersion for 10 min in 1% OsO4. The tissue was then washed and dehydrated with a graded ethanol series. After dehydration, specimens were freeze-dried using t-butyl alcohol (Inoue and Osatake, 1988), coated with Pt-Pd and observed with a scanning electron microscope (SEM) (S-800; Hitachi-Hi-Technologies, Tokyo, Japan).

Results

Macroscopic overview

Macroscopically, the tongue of the greater Japanese shrew-mole (Fig. 1) was elongated in rostro-caudal direction, with a rounded apex linguæ. No torus linguæ were present. Numerous papillae filiformes (Fig. 1; Fi) were distributed over the entire dorsal surface of the tongue except on the radix linguæ. Papillae fungiformes (Fig. 1; Fu) were scattered over the apex linguæ (Fig.1b; A) and on the rostral (Fig.1b; B) and caudal regions (Fig.1b; D) of the corpus linguæ but they were fewer in the mid-region (Fig.1b; C). Neither a sulcus medianus linguæ nor a sulcus terminalis linguæ were present. A pair of oval or rostrolaterally obliquely elongate papillae vallatae (Fig. 1b; PV) was situated at the boundary between the caudal section (Fig.1b; D) of the corpus linguæ and the radix linguæ (Fig.1b; E). Papillae foliatae and lateral organ-like structures (Sonntag, 1925) were absent. At the caudal end of the radix linguæ (Fig.1b; E), the papillae linguales were attenuated, forming only weak folds.

Microscopic observation

Papillae filiformes

Light microscopy revealed that the epithelium of papillae filiformes (Fig. 2d) was keratinized, especially on the caudal aspect. A concavity was present rostral to each papilla (Fig. 2d, 4a; arrows). Keratohyalin granules were observed in the apex linguæ and in the caudal parts of the papillae (Fig. 2d, 4a). The papillae filiformes on the apex linguæ were more erect than those in the caudal regions of the corpus linguæ, where the stratum...
granulosum containing keratohyalin granules exhibited a compressed appearance. A number of nuclei were present in superficial cells of the interpapillary epithelium throughout the dorsal surface of the tongue. In sagittal sections, the lamina propria of the CTCs of the papillae filiformes had a different appearance in different regions of the tongue. On the apex linguae, the CTCs were rounded and extended only into the lower third of the epithelium. In contrast, those in the caudal section of the corpus linguae had pointed tips and extended almost into the superficial epithelial layer.

Visualization of the external surface by scanning electron microscopy revealed that each papilla filiformis was associated with a round concavity (Fig. 2d and e, Fig. 3a and c, Fig. 4b) and was surrounded by a ring-like wall. On the apex linguae, a thumb-like conical process (Fig. 2e) was situated on the caudal part of the rim of each papilla filiformis and was inclined caudally. The appearance of the papillae filiformes varied depending on the area. In the rostral section of the corpus linguae (Fig. 1b), the processes of the papillae filiformes were somewhat elongated vertically, with a sharp tip (Fig. 3a).
Fig. 3. A set of papillae filiformes distributed on the rostral (see Fig. 1b, B) or middle (see Fig. 1b, C) part of the corpus linguae. (a) SEM micrograph of the epithelial surface of papillae filiformes in the rostral region of the corpus. The papillae filiformes in this area exhibit sharp main processes. Scale bar: 20 μm. (b) SEM micrograph of the CTCs of the papillae filiformes of the rostral part of the corpus after the removal of the epithelium. The CTCs of papillae filiformes in this area possess a notch at the centre of the CTC. Scale bar: 20 μm. (c) SEM micrograph of the epithelial surface of papillae filiformes situated in the middle region of the corpus. The papillae filiformes in this area have two sharp main processes. Scale bar: 20 μm. (d) SEM micrograph of the CTCs of the papillae filiformes in the middle region observed after epithelial exfoliation. The edges of the CTCs of papillae filiformes are thin and possess a deep cleft in the midline. Scale bar: 20 μm.

Fig. 4. A set of papillae filiformes distributed on the caudal section of the corpus linguae (see Fig. 1b, D). (a) Sagittal histological section of papillae filiformes on the caudal area of the corpus. Goblet-like concavities (arrows) are situated in front of each papilla. Numerous keratohyalin granules (asterisk) are present in the stratum corneum and appear to be located beneath the concavities. Scale bar: 60 μm. (b) SEM micrograph of the epithelial surface of papillae filiformes situated on the caudal area of the corpus. Papillae filiformes distributed in this area have three sharp main processes. Scale bar: 30 μm. (c) SEM micrograph of the CTC of the papillae filiformes at the caudal area of the corpus linguae after the removal of the epithelium. CTCs of papillae filiformes in this area possess two notches. A cradle-like concavity is situated in front of the CTC of papillae filiformes. Scale bar: 30 μm.
**Papillae filiformes** distributed over the middle of the corpus (Fig. 1b) possessed two sharp processes (Fig. 3c), whereas in the caudal region (Fig. 1b), there were three slender sharp conical processes (Fig. 4b). After exfoliation of the epithelium, the CTCs of the **papillae filiformes** reflected the morphological differences of the epithelial surface. Each CTC of the **papillae filiformes** on the apex linguae (Fig. 1b) consisted of a single, smooth, dome-like protuberance with a hemispherical depression on the rostral surface. CTCs that were distributed on the rostral section of the corpus (Fig. 1b) had a cleft at the top (Fig. 3b). In the middle section (Fig. 1b), the CTCs were sharp-edged with a V-shaped cleft (Fig. 3d). Furthermore, CTCs in the caudal region of the corpus (Fig. 1b) possessed two clefts (Fig. 4c), presumably reflecting their overlying epithelial processes. The **papillae filiformes** were approximately 50–170 μm long and 35–86 μm wide.

**Papillae fungiformes**

Under the light microscope, the apices of the **papillae fungiformes** (Fig. 2a) were dome-like. Keratinization of these papillae was weak and some of the more superficial epithelial cells retained nuclei. A few taste buds were observed in the epithelium at the tips of the **papillae fungiformes**.

In SEM observations, the external surface of **papillae fungiformes** distributed on the rostral part of the corpus linguae (Fig. 2b) was smooth and dome-like, although their size varied. After epithelial exfoliation, the CTCs of the **papillae fungiformes** on the apex linguae (Fig. 2c) were columnar with several vertically aligned, narrow ridges. **Papillae fungiformes** were approximately 78–148 μm in diameter.

**Papillae vallatae**

A pair of **papillae vallatae** (Fig. 1b) was situated at the boundary between the caudal section of the corpus (Fig. 1b) and the radix linguae (Fig. 1b). In the light microscope, numerous taste buds were observed in the inner epithelial wall of the circumferential furrow. Glandulae linguales were seen in the lamina propria (Fig. 5a) with their orifices opening into the circumferential groove. A concavity was present at the top of each **papilla vallata**.

In SEM preparations, the external surfaces of the **papillae vallatae** (Fig. 5b) were oval, or obliquely elongated in a rostrolateral direction. The dome-like papilla was surrounded by a circumferential ridge and furrow. Cleft-like grooves were seen at the top of the elongated papillae. After exfoliation of the epithelium, CTCs of **papillae vallatae** were concave and surrounded by a wall-like core in the circumferential furrow (Fig. 5c). **Papillae vallatae** were approximately 337–703 μm in diameter.

**Radix linguae**

The radix linguae exhibited a weakly folded appearance (Fig. 1b). Under the light microscope, large mucus-rich mixed glands of glandulae linguales were seen in the lamina propria. After the removal of the epithelium, the surface of the CTCs of the radix linguae exhibited numerous processes. Orifices of glandulae linguales were observed among the CTC processes.

**Discussion**

Our observations indicate that the tongue of the Japanese shrew-mole possesses an assortment of morphological characteristics, representing features of both the Talpidae (moles and shrew-moles) and other Soricomorpha (shrews). Table 1 compares the morphological characteristics that we observed in the Japanese shrew-mole with those reported in previous investigations on other species.

First, it may be noted that the Japanese shrew-mole and the furry-snouted mole (Kobayashi et al., 1983) possess bifid and/or trifid **papillae filiformes**, whereas other talpids, including the European mole (Jackowiak, 2006), the large Japanese mole (Kobayashi et al., 1983) and the small Japanese mole (Miyata et al., 1990), possess only simple conical **papillae filiformes**. In contrast, bifid and trifid **papillae filiformes** are widely distributed on the tongue of the shrew.

The distribution patterns of **papillae fungiformes** are variable, especially among the Talpidae. In the Japanese shrew-mole, **papillae fungiformes** were sparsely distributed marginally and/or medially in the apex, which is similar to the pattern in the large Japanese mole (Kobayashi et al., 1983). In the rostral section of the tongue of the Japanese shrew-mole, **papillae fungiformes** were located marginally and medially, as they are in the furry-snouted mole (Kobayashi et al., 1983). Furthermore, few **papillae fungiformes** were present in the middle section of the tongue in the Japanese shrew-mole, which differs from the arrangement in other Talpidae and in Soricidae. In addition, the radix linguae of the Japanese shrew-mole exhibited weak folds, a feature not reported in other species.

Three types of **papillae vallatae** are present among the Soricomorpha (shrews): a circular type and two discontinuous types. The **papillae vallatae** of the Talpidae (mole and shrew-moles), including the present species, appeared to be only of the circular type.

Morphological information concerning the CTCs beneath the epithelium of the **papillae linguales** of Soricomorpha is limited and exists only for the Japanese mole (Miyata et al., 1990) and the house musk-shrew (Kobayashi et al., 1989, 2005; Kobayashi, 1992). However, both species exhibit different combinations of morphological traits with respect to the lingual papillae. In Japanese
shrew-moles, we observed that the CTC of the *papillae filiformes* on the apex linguæ consisted of a single dome, each with a hemispherical indentation on the rostral aspect. Another talpid, the Japanese mole (Miyata et al., 1990) has rather similar CTCs, described as ‘wooden spoon-like’, but the top edge of these CTC was somewhat thinner. In contrast, the CTCs of *papillae filiformes* (Kobayashi et al., 1989, 2005; Kobayashi, 1992) on the apex of the tongue of the house musk-shrew are spherical processes upon which a shallow groove runs in the rostro-caudal direction. Furthermore, the CTCs of *papillae filiformes* on the caudal region of the corpus of the Japanese shrew-mole possessed three clefts (trifids), whereas in the Japanese mole, the CTCs of the *papillae filiformes in this region* are vertically elongated and more conical (Miyata et al., 1990). This, therefore, represents a morphological trait that is rather different from that of the Japanese shrew-mole, despite the similarity of the epithelial surfaces of their *papillae filiformes*. Unfortunately, most previous reports of the morphology of the *papillae linguæ* in Soricomorpha only provide information on their epithelial surface so that elucidation of their mor-

**Fig. 5.** A set of *papillae linguæ* distributed on the radix linguæ (see Fig. 1b, E). (a) Transverse histological section of a *papilla vallata*. Numerous taste buds (arrows) are present in the inner wall of the epithelium of the peri-papillary furrow. Glandulae linguæ (G) are located in the lamina propria. A concavity is present in the centre of the papilla. Scale bar: 100 µm. (b) SEM micrograph of the epithelial surface of a *papilla vallata*. The epithelium is obliquely elongated, and a cleft-like concavity is present at the tip of the papilla. Scale bar: 100 µm. (c) SEM micrograph of the *papillae vallatae* after exfoliation of their epithelium. A wall-like CTC of the furrow of a *papilla vallata* is recognizable. Numerous CTCs of *papillae filiformes* are also distributed in this area. However, each CTC of the *papillae filiformes* has many notches with a flame-like appearance. Scale bar: 100 µm. (d) Sagittal histological section of the radix linguæ. Mucus-rich mixed glands of *glandulae linguæ* (G) are situated deep in the lamina propria. Scale bar: 100 µm. (e) SEM micrograph of the epithelial surface of the radix linguæ. The surface is smooth and hilly. Scale bar: 50 µm. (f) SEM micrograph of the caudal section of the radix linguæ after the removal of the epithelium. Numerous thumb-like or ridge-like CTCs are distributed on the surface. Some appear to be arranged in rows. An orifice of the glandulae linguæ (o) can be seen. Scale bar: 50 µm.
Table 1. Comparison of morphological features of Japanese shrew-mole and other talpid and soricomorphan tongues

| Species                      | Macroscopic view | Papillae Linguae | Papillae Filiformes | Papillae Fungiformes | Papillae Folialae | Papillae Vallatae | Radix linguae |
|------------------------------|------------------|------------------|---------------------|----------------------|------------------|------------------|---------------|
| Taipidae                     |                  |                  |                     |                      |                  |                  |               |
| Japanese shrew-mole          | Ro               | Not tapered      | One thumb-like      | One sharp conical    | Bifid            | Trifid           | Sp            |
| (Present study)              |                  |                  |                     |                      | Ma/Med           | Few              | Ma/Med        |
| Furry-snouted mole           | Ro               | Not tapered      | One conical         | One conical          | Bifid            | Trifid           | Few           |
| (Kobayashi et al., 1983)     |                  |                  |                     |                      | Ma/Med           | Ma/Med           | Ma/Med        |
| European mole                | Ro               | Not tapered      | One conical         | One conical          | One conical      | One conical      | Ma            |
| (Jackowiak, 2006)            |                  |                  |                     |                      | Ma               | Ma Row7–10       |               |
| Large Japanese mole          | Ro               | Not tapered      | One conical         | One conical          | One conical      | One conical      | Ma            |
| (Kobayashi et al., 1983)     |                  |                  |                     |                      | Ma               | Ma Row7–10       |               |
| Small Japanese mole          | Ro               | Not tapered      | One conical         | One conical          | One conical      | One conical      | Ma            |
| (Miyata et al., 1990)        |                  |                  |                     |                      | Sp               | Ma               | Ma            |
| Soricidae                    |                  |                  |                     |                      | Ma/Med           | Ma               | Ma/Med         |
| Long-clawed shrew            | Ro               | Not tapered      | One conical         | One conical          | One conical      | One conical      | Ma            |
| (Kobayashi et al., 1989, 2005; Kobayashi, 1992) | | | | | | | |
| Shinto shrew                 | Ro               | Tapered          | One conical         | Bifid                | Bifid            | Trifid           | Ma Row        |
| (Kobayashi et al., 1989, 2005; Kobayashi, 1992) | | | | | | | |
| Common shrew                 | Ro               | Tapered          | One conical         | Bifid                | Bifid            | Trifid           | Ma Row        |
| (Jackowiak et al., 2004)     |                  |                  |                     |                      | Ma               | Ma               | Ma Row        |
| Japanese water shrew         | Ro               | Not tapered      | One conical         | Bifid                | Bifid            | Ma               | Sp            |
| (Kobayashi et al., 1989, 2005; Kobayashi, 1992) | | | | | | | |
| House musk shrew             |                  |                  |                     |                      | Ma               | Ma1/4           | Ma1/4         |

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phological traits is limited. As it is clear that CTCs also exhibit morphological differences, particularly those of the *papillae filiformes*, a more detailed analysis of the CTCs of other insectivores is greatly needed.

The above-mentioned morphological differences are presumably influenced by the animals’ dietary habits. Most shrews are carnivorous but may also take carrion. They are primarily insectivorous, but some also eat seeds, nuts and other plant material. The diet of moles consists largely of earthworms, beetles and fly larvae and, when available, slugs (Rudge, 1968; Macdonald, 1984). In both groups of animals, therefore, the principal food consists of insects but shrews also ingest plant materials. The diet of the Japanese shrew-mole comprises mostly insects, spiders, centipedes and earthworms (Komiya, 2002; Abe et al., 2005). It is noteworthy, therefore, that the bifid or trifid morphology of the *papillae filiformes* in Japanese shrew-moles was similar to that of shrews, despite their diet being identical with other moles. This implies that the morphological trait exhibited by the Japanese shrew-moles may have been influenced more by their evolutionary history than by the environment. Thus, the exceptional morphological traits shown by the *papillae filiformes* of the Japanese shrew-mole reflect their transitional taxonomic status.

Further studies, including both morphological- and molecular-based investigations, are required to more fully elucidate the basis of the morphological specializations of the *papillae linguales* of the Japanese shrew-mole.

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References

Abe, H., N. Ishii, T. Ito, Y. Kaneko, K. Maeda, S. Miura, and M. Yoneda, 2005: A guide to the mammals in Japan. Hadano: Tokai University Press.

Adnyane, I. K. M., A. B. Zuki, M. M. Noordin, and S. Agungpriyono, 2011: Morphological study of the lingual papillae in the Barking deer, *Muntiacus muntjak*. Anat. Histol. Embryol. 40, 73–77.

Inoue, T., and H. Osatake, 1988: A new drying method of biological specimens for scanning electron microscopy: The t-butyl alcohol freeze-drying method. Arch. Histol. Cytol. 51, 53–59.

Jackowiak, H., 2006: Scanning electron microscopy study of the lingual papillae in the European mole (*Talpa europaea, L.*, *Talpidae*). Anat. Histol. Embryol. 35, 190–195.

Table 1 (continued)

| Species                  | Macroscopic view | Papillae Linguales | Papillae Filiformes | Papillae Fungiformes | Papillae Foliatae | Papillae Vallatae |
|--------------------------|-------------------|--------------------|---------------------|---------------------|-------------------|------------------|
| **Apex**                 |                   |                    |                     |                     |                   |                  |
| Po                       | Not               | tapered            | Ma1/4               | Ma1/4               | Ma1/4             | Ma/Med           |
| Fo                       | Weak folds        |                    |                     |                     |                   |                  |
| Wa                       | Wart-like processes|                   |                     |                     |                   |                  |
| **Corpus**               |                   |                    |                     |                     |                   |                  |
| Po                       | Not               | tapered            | Ma1/4               | Ma1/4               | Ma1/4             | Ma/Med           |
| Fo                       | Weak folds        |                    |                     |                     |                   |                  |
| Wa                       | Wart-like processes|                   |                     |                     |                   |                  |
| **Radix linguae**        |                   |                    |                     |                     |                   |                  |
| Po                       | Not               | tapered            | Ma1/4               | Ma1/4               | Ma1/4             | Ma/Med           |
| Fo                       | Weak folds        |                    |                     |                     |                   |                  |
| Wa                       | Wart-like processes|                   |                     |                     |                   |                  |
| **Papillae Filiformes**  |                   |                    |                     |                     |                   |                  |
| 1983, Kobayashi et al.   | Not               | tapered            | Ma1/4               | Ma1/4               | Ma1/4             | Ma/Med           |
| 1989, Kobayashi, 1992    | Not               | tapered            | Ma1/4               | Ma1/4               | Ma1/4             | Ma/Med           |

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Jackowiak, H., S. Godynicki, M. Jaroszewska, and B. Wilczynska, 2004: Scanning electron microscopy of lingual papillae in the common shrew *Sorex araneus*, L. Anat. Histol. Cytol. 33, 290–293.

Kobayashi, K., 1992: Comparative anatomical studies on the tongues with special reference to the connective tissue cores of the lingual papillae. Odontology 80, 661–678.

Kobayashi, K., and S. Iwasaki, 1989: Comparative studies on the stereo architecture of the connective tissue papillae in some mammalian tongues. Prog. Clin. Biol. Res. 295, 303–308.

Kobayashi, S., T. Shimoda, and A. Shimamura, 1983: Comparative anatomical observation on the tongue of the *insectivora*. Okajimas Folia Anat. Jpn. 60, 211–217.

Kobayashi, S., S. Arai, S. Tomo, T. Shimoda, A. Shimamura, and H. Yamada, 1989: Scanning electron microscopic study on the lingual papillae of the Japanese *insectivora*. Okajimas Folia Anat. Jpn. 65, 413–427.

Kobayashi, K., H. Jackowiak, H. Frackowiak, K. Yoshimura, M. Kumakura, and K. Kobayashi, 2005: Comparative morphological study on the tongue and lingual papillae of horses (*Perissodactyla*) and selected ruminantia (*Artiodactyla*). Ital. J. Anat. Embryol. 110, 55–63.

Komiya, T., 2002: Mammalian in Japan. Field Best Encyclopaedia. Tokyo: Gakushu-Kenkyu-sha press.

Macdonald, D. W., 1984: Insectivores. The Encyclopedia of Mammals. New York: Facts on File Publications.

Miyata, K., S. Iwasaki, and K. Kobayashi, 1990: Fine morphological studies in the connective tissue and the epithelial cell of the lingual papillae in *Mogera wogura wogura*. Odontology. 78, 553–574.

Nickel, R., R. Schummer, and E. Seiferle, 1979: Digestive system. The Viscera of the Domestic mammals. Berlin: Verlag Paul Parey.

Nikaido, M., Y. Cao, M. Harada, N. Okada, and M. Hasegawa, 2003: Mitochondrial phylogeny of hedgehogs and monophyly of Eulipotyphla. Mol. Phylogenet. Evol. 28, 276–284.

Rudge, M. R., 1968: The Food of the Common Shrew *Sorex araneus* L. (Insectivora: Soricidae) in Britain. J. Anim. Ecol. 37, 565–581.

Shinohara, A., K. L. Campbell, and H. Suzuki, 2003: Molecular phylogenetic relationship of moles, shrew moles, and desmans from the new and old worlds. Mol. Phylogenet. Evol. 27, 247–258.

Sonntag, C. F., 1923: The Comparative Anatomy of the Tongues of the Mammalia.—IX. Edentata, Dermoptera, and *Insectivora*. Proc. Zool. Soc. Lond. 93, 315–529.

Sonntag, C. F., 1925: The Comparative Anatomy of the Tongues of the Mammalia.—XII. Summary. Proc. Zool. Soc. Lond. 21, 701–762.

Symonds, M. R. E., 2005: Phylogeny and life histories of the ‘*Insectivora*’: controversies and consequences. Biol. Rev. 80, 93–128.

Tsuchiya, K., H. Suzuki, A. Shinohara, M. Harada, S. Wakana, M. Sakaizumi, S. H. Han, L. Lin, K., and A. P. Kryukov, 2000: Molecular phylogeny of east Asian moles inferred from the sequence variation of the mitochondrial cytochrome b gene. Genes. Genet. Syst. 75, 17–24.

Watanabe, I., J. P. Guimaraes, M. O. Maia, T. C. Santos, J. R. Kfoury Jr, S. A. Boleta, S. R. Almeida, M. M. Righeti, and M. A. Miglino, 2011: SEM and neurohistological observations of nerve endings in the middle region of the tongue of the collared peccary (*Tayassu tajacu*): a silver impregnation method. Anat. Histol. Embryol., 40, 128–133.

Wilson, D. E., and D. A. M. Reeder, 2005: Mammal species of the world. A taxonomic and geographic reference. Baltimore: Johns Hopkins University Press.