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Rickettsia massiliae in the Canary Islands

To the Editor: Rickettsia massiliae was recently recognized as a human tick-borne spotted fever group rickettsia (1). We report the finding of R. massiliae in Rhipicephalus pusillus ticks from Gran Canaria, Canary Islands, Spain. Introduction of this pathogen into the Canary Islands is thought to have resulted from translocation of the European wild rabbit Oryctolagus cuniculus (Linnaeus), a preferred host of R. pusillus ticks (www.kolonin.org/16_4.html), from the Iberian Peninsula 600 years ago (2).

We collected questing adult ticks in 2008 in Gran Canaria and identified 2 tick species, Hyalomma lusitanicum (n = 82 [46 females]) and R. pusillus (n = 8 [5 females]). Whole ticks were preserved in 70% ethanol and used for DNA extraction by using TriReagent (Sigma, St. Louis, MO, USA) according to the manufacturer’s instructions. We identified rickettsial sequences by using PCR primers that amplify fragments of 16S rRNA,ompB, atpA, dnaA, dnaK, and recA genes (Table). Ambicones were cloned into pGEM-T (Promega, Madison, WI, USA), and 3 independent clones were sequenced from both ends for each gene marker. Sequence similarity search was performed by using BLAST (www.ncbi.nlm.nih.gov). Rickettsial DNA was detected in 2 R. pusillus males only; sequences were identical in both ticks. Fragments of 16S rRNA were 99% identical to the R. massiliae strain Mtu5 (CP000683) isolated from R. sanguineus ticks in southern France (3), and fragments of ompB, atpA, dnaA, dnaK, and recA genes were 100% identical to the R. massiliae strain Bar29 (AF123710, AY124739, DQ821798, DQ821828, and AY124750, respectively), previously isolated from R. sanguineus ticks in Catalonia, Spain (4) (Table).

R. massiliae was first isolated in 1992 from R. sanguineus ticks collected near Marseille, France (5). Since then, the pathogen has been identified in different Rhipicephalus species in France, Greece, Portugal, Switzerland, Spain, North and Central Africa, Argentina, and the United States (6,7). R. massiliae has been identified in southern Spain (8) but not in the Canary Islands. R. pusillus ticks are commonly found in southern Europe (Portugal, Spain, and France) and northern Africa (Tunisia and Morocco). All stages of these ticks inhabit burrows of wild rabbits and feed on them (www.kolonin.org/16_4.html).

Wild rabbits were introduced into the Canary Islands at the end of 14th century during colonization by the kingdom of Castilla. Colonists were asked to bring rabbit couples with them to provide food in the islands (2), a practice continued by new colonists because of their interest in hunting this rabbit species. Introduction of wild rabbits by colonists led to establishment of parasites, such as helminths, coccidia, and viruses in the Canary Islands (9). R. pusillus, a common ectoparasite (tick) that feeds on wild rabbits on the Iberian Peninsula, was also introduced this way. R. massiliae could have been introduced in the islands by infected R. pusillus ticks or by infected wild rabbits if this species serves as a natural reservoir host for the pathogen.

To find evidence for this hypothesis, we tested blood and liver samples of 150 wild rabbits from both Canary Islands and Andalucia (southern Spain) by using Rickettsia-specific PCR primers (Table). No R. massiliae DNA was detected in the rabbit samples tested,
suggesting that the pathogen probably was introduced in the Canary Islands with infected *R. pusillus* ticks feeding on rabbits. Alternatively, *R. massiliae* infection levels in wild rabbits may be below the PCR detection limit and were not detected.

The Canary Islands are a popular tourist destination. The presence of *R. massiliae* in the islands constitutes a risk for human infection and should be considered in hospital diagnostic and wildlife management strategies. As with other *Rhipicephalus* spp., *R. pusillus* ticks could feed on humans under certain circumstances (10). Our results emphasize the risks associated with unsupervised animal translocations, a factor that probably plays a role in the introduction of ticks and tick-borne pathogens in different parts of the world.

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| Gene       | Primer sequence (5’ → 3’)   | Amplicon size, bp | PCR annealing conditions |
|------------|----------------------------|-------------------|--------------------------|
| 16S rRNA   | F: AGAGTTTGATCCTGCGGCTCAAG GGGTGCTGCTACACAGCAGAA AAGCTGAAGGCTCC | 416 | 1.  50°C/30 s |
| ompB       | R: AACGCTGATCTTCTTGCGGCTAC CCGTCACCGATATTAATTGCC | 618 | 2.  53°C/30 s |
| dnaK       | F: AGCGTCAAGCAACAGAAAGAT AAGCTGAAGGCTCC | 323 | 3.  50°C/30 s |
| dnaA       | R: CAAACGTTGAAGTGCTAAAGG AAGCTGAAGGCTCC | 241 | 4.  56°C/30 s |
| recA       | F: CCTACTAACTTTGTATAGAGTT AAGCTGAAGGCTCC | 428 | 5.  52°C/30 s |
| atpA       | R: CGCGTACCCCGACATATTTCC AAGCTGAAGGCTCC | 731 | 6.  48°C/30 s |

*GenBank accession numbers correspond to *R. massiliae* sequences identified in this study. PCRs were completed by employing the Access RT-PCR system (Promega, Madison, WI, USA) with 1 ng DNA, the oligonucleotide primers, and annealing conditions and with extension for 1 min at 68°C. F, forward; R, reverse.