Plasma prolactin changes in anoestrous ewes after infusion of genistein into the third ventricle*

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ABSTRACT

Genistein, one of isoflavones, is structurally similar to mammalian endogenous oestrogens, which inhibit activity of tyrosine kinase, DNA topoisomerases, and possess antioxidant activity and oestrogenic activity. The aim of the present work was to study effects of the central administration of genistein on prolactin secretion in ewes during the seasonal anoestrus. The experiment was carried out on twelve Polish Lowland ewes. Infusions of genistein into the third ventricle of ewes were performed with calibrated 1.0-ml gas-tight syringes and a microinjection pump. Control infusions (n=4) from 12.01 to 16.01 h were done with a Ringer-Locke solution at flow rate 100 µl/h. The dose of genistein was 1 µg/100 µl/h (n = 6) and 10 µg/100 µl/h (n = 5) during 4-h infusions. Before the infusions, differences in prolactin concentration between control and experimental animals were insignificant. Prolactin concentrations were significantly (P<0.05) elevated after 2 h of genistein infusion (1 µg/h) as compared with the control group. The higher dose of genistein (10 µg/h) evoked an immediate increase of prolactin concentrations, which remained elevated (P<0.05) until the end of samples collection at 20.00 h. There was also significant (P<0.05) difference in prolactin concentrations, during the last 2 h of the blood collection, between ewes with different genistein doses. In conclusion, centrally administered genistein, stimulate the secretion of prolactin in dose-dependent manner in ewes during the seasonal anoestrus.

KEY WORDS: prolactin, genistein, intracerebroventricular, ewe

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INTRODUCTION

Isoflavones compounds, a class of phytoestrogens, which include formononetin, biochanin A, daidzein and genistein, occur in red clover (Trifolium pratense), and might be causing infertility in female sheep (Adams, 1995). In the earlier study Mathieson and Kitts (1980) found that genistein and coumestrol could bind to oestrogen receptors in pituitary and hypothalamic cytosol of the ewe. In general, phytoestrogens mimic the actions of oestradiol. Their affinity to oestrogen receptor alfa (ERα) as well as the oestrogen receptor beta (ERβ) is substantial (Kuiper et al., 1998). In a previous study we have shown that feeding red clover silage to ovariectomized ewes caused an increase in teat length, circumference and even stimulated a milky fluid secretion (Nwannenna et al., 1995). These changes were similar to that during treatment with oestradiol-17β implant. Ingestion of phytoestrogens stimulated also secretion of thyroid hormones and tended to increase follicle size and ERα immunoreactivity of thyroid glands of ovariectomized ewes (Madej et al., 2002). Treatment of intact and ovariectomized ewes with different doses of oestradiol-17β resulted in a stimulation of prolactin’s secretion (Baird et al., 1981; Elsasser et al., 1983; Rozell and Keisler, 1990). The aim of the present work was to study effects of the central administration of genistein on prolactin changes in intact ewes during the seasonal anoestrus.

MATERIAL AND METHODS

Animals, management and experimental procedure

Twelve Polish Lowland ewes were used in this experiment during the period corresponding to anoestrus. A steel guide cannulae (1.6 mm o.d.) was implanted under stereotaxic control in the third brain ventricle through a drill hole in the skull under general anaesthesia (Traczyk and Przekop, 1963; Welento et al., 1969; Misztal et al., 1997). General anaesthesia was induced with Vetbutal plus ketamine (Biowet, Poland). The guide cannula was fixed to the skull with stainless steel screws and dental cement. The external opening to the canal was closed with a stainless steel cup. After surgery, usually lasting 45 to 70 min, animals were injected with pain-relieving drugs and antibiotics (Betamox - ScanVet, Poland). The post-operative period was a minimum 10 days, under veterinary surveillance. Sixteen days after surgery, adequately recovered animals were used in the experimental studies.

Infusions of genistein into the third ventricle of ewes were performed with calibrated 1.0-ml gas-tight syringes and a microinjection pump CMA/100 (CMA/Microdialysis AB, Stockholm, Sweden). Control infusions (C-group, n=4) were
done with a Ringer-Locke solution from 12.01 to 16.01 h at flow rate 100 μl/h. The

dose of genistein (Sigma-Aldrich Corp. St. Louis, MO, USA) of concentration

1 μg/100 μl/h (GEN1-group, n=6) and 10 μg/100 μl/h (GEN10-group, n=5) was

infused during 4 h. Before infusion, an appropriate genistein solution was prepared

from the stock solution in ethanol (20 mg/ml), kept in -20°C. During the experiments,

animals were kept in cages of comfortable sizes, in which they could lie down and

have free access to hay and water. Blood samples (3 ml) were collected from the

jugular vein, through a catheter, at 10-min intervals from 08.00 to 20.00 h. Plasma

samples were stored at -20°C until assayed for prolactin by radioimmunoassay.

The Ethics Committee of the Kielanowski Institute of Animal Physiology and

Nutrition approved all described procedures at Jablonna, according to the Polish

Guide of Care and Use of Animals (August 2, 1997).

Prolactin determination

Plasma prolactin concentrations were assayed by a radioimmunoassay double-

antibody method, using antiovine-PRL and antirabbit-gammaglobulin antiserum

according to Wolinska et al. (1977). The prolactin standard was produced by

Professor Kochman from the Kielanowski Institute of Animal Physiology and

Nutrition (Kochman and Kochman, 1977) and is routinely applied in the analysis.

The intra- and interassay coefficients of variation were 9.0 and 12.0%, respectively.

The lowest detectable amount of prolactin was 2 μg/l.

Statistical analysis

The hormonal observations were first log transformed and then the repeated

measurement analysis of variance was performed using the MIXED procedure

on the generated averages according to the Statistical Analysis System program

package (Release 6.12, 1996, SAS Institute Inc., Cary, NC, USA). The statistical

model included dose (3 groups), time (twelve 1-h periods), the interaction between

dose and time, and the random effect of ewe within dose. From the analysis of

variance, the results of prolactin concentrations, presented as least squares means

(LS means) ± SEM, were back transformed.

RESULTS

No significant difference in the plasma concentration of prolactin was seen during

four h prior infusions between the C-group and the GEN1-group (Figure 1). During

that time, prolactin concentrations varied between 44.7 and 165.3 μg/l with maximum

values before 10.00 h. Infusion of Ringer-Locke solution did not changed significantly
prolactin secretion; its concentration varied between 39.0 and 77.0 μg/l (Figure 1). On the other hand, infusion of 1 μg/h of genistein into the third ventricle did result in the significant increase in prolactin from the initial concentration of 67.7±25.1 μg/l at 12.00 h to 142.0±25.1 μg/l at 16.00 h (P<0.05). Afterwards, prolactin concentration gradually decreased, reaching a level comparable to that of control ewes at 18.00 h (Figure 1). In the GEN1-group, two h after genistein infusion was stopped, prolactin concentrations were, overall, significantly (P<0.05) elevated, compared with that of the C-group.

![Figure 1](image_url)

Figure 1. Concentrations of prolactin (LS means ± SEM) in 10-min plasma samples over 12-h sampling in ewes during an infusion either of Ringer-Locke solution (from 12.01 to 16.01 h) (▼—▼) or genistein (1 μg/h from 12.01 to 16.01 h) (○—○). Arrows point to beginning and termination of the infusions.

No significant difference in the plasma concentration of prolactin was seen during four h prior infusions between the C-group and the GEN10-group (Figure 2). Infusion of 10 μg/h of genistein into the third ventricle resulted in the immediate increase of prolactin concentration, from the initial 62.0±43.2 μg/l at 12.00 h to 150.0±43.2 μg/l at 13.00 h (P<0.05). Before the infusion was stopped at 16.00 h, the concentration of prolactin had reached 220.0±43.2 μg/l (P<0.05). After 16.00 h, prolactin concentrations were still elevated (140.0-210.0 μg/l) until 20.00 h, when the last samples were collected. Overall, in the GEN10-group, beginning two h after start of infusion, prolactin concentrations were significantly (P<0.05) elevated, compared with that of the C-group. During the last 2 h of
Figure 2. Concentrations of prolactin (LS means ± SEM) in 10-min plasma samples over 12-h sampling in ewes during an infusion either of Ringer-Locke solution (from 12.01 to 16.01 h) (■—■) or genistein (10 μg/h from 12.01 to 16.01 h) (○—○). Arrows point to beginning and termination of the infusions experimental day (18.00 h to 20.00 h), prolactin concentrations in the GEN10-group were significantly (P<0.05) higher than that in GEN-1 (Figure 1 vs Figure 2).

DISCUSSION

The results from the present study show that the centrally administered genistein stimulates the secretion of prolactin in ewes during seasonal anoestrus. Several hypotheses can be advanced to explain this stimulatory effect of genistein. In the case of ovariectomized rats, ingestion of either genistein or oestradiol resulted in the stimulation of prolactin secretion (Santell et al., 1997). Santell's et al. (1997) concluded that genistein's action in the hypothalamus and pituitary is similar to that of oestradiol, leading to the synthesis and release of prolactin from the adenohypophysis in rats. Their conclusion was based on the findings of Jones and Naftolin (1990), who demonstrated that oestradiol acts on the synthesis and release of prolactin by decreasing the activity of tyrosine hydroxylase, which in turn decreases the concentration of dopamine in the hypothalamus. Thus, genistein's ability to inhibit mammary gland regression postovariectomy might be mediated through prolactin, which exerts its mitogenic effects on the mammary
gland (Santell et al., 1997). This suggestion is in agreement with our previous findings in the ovariectomized ewes (Nwannenna et al., 1995) which showed that the ingestion of phytoestrogens resulted in mammary gland development and milky fluid secretion.

Moreover, Stahl et al. (1998) reported that genistein induced prolactin secretion from the immortalized pituitary cell line PR1, stimulating the growth of these cells, and did so in an oestrogen receptor-dependent manner. Thus, genistein might also be active within lactotroph cells of the pituitary. Messina and Loprinzi (2001) described genistein as having oestrogen-like effects at lower concentrations (which may be considered physiological concentrations); at higher concentrations, as having other non-oestrogen receptor mediated effects, such as inhibition of the activity of one or more cellular molecules that control cell signaling, growth and death.

Daidzein, another isoflavone, given orally as a supplement to feed stimulated the secretion of LH and prolactin in pigs (Liu et al., 1999). Recently, Ren et al. (2001) reported that daidzein down-regulates ERβ gene expression in the hypothalamus of newborn pigs, possible indicating central effects of this isoflavone.

In earlier findings of Wolinska et al. (1977) it was suggested that in sheep the inhibitory system controlling the secretion and production of prolactin exist in the caudal medial basal hypothalamus. Indeed, the suppression of prolactin concentrations following infusion of D2-like (quinpirole) agonist into MBH (Anderson et al., 1997) supports findings of Wolinska et al. (1977) and Curlewis et al. (1991). Dopamine might also act through dopamine D2 receptors located on lactotrophs in the anterior pituitary in the ewe (Anderson et al., 1997; Bertrand et al., 1999). On the other hand a stimulatory system controlling the secretion and production of prolactin in sheep was suggested to operate via dopamine D1 receptors in the ventromedial hypothalamus (VMH) (Curlewis et al., 1995).

Intraventricular administration of oestradiol for 3 days resulted in an increase of serum prolactin in rats, consistent with the change in the prolactin mRNA level (Maeda et al., 1996). However, Blum et al. (1987) showed that a decrease in tyrosine hydroxylase gene transcription in rat hypothalamus occurred 20 min after oestrogen treatment and reached 5% of control level after an hour. In ovariectomized ewes, oestradiol treatment during long days resulted in reduction of pulsatile LH secretion, without affecting prolactin, due to stimulation of tyrosine hydroxylase in the lateral retroschiasmatic area (Gayrard et al., 1994).

Interestingly, melatonin infused into the third ventricle in anoestrous ewes stimulated the prolactin secretion within 30 min, and maintained high concentrations for an additional 2 to 3 h (Misztal et al., 1997). In the present study, the response of prolactin to the low-dose infusion of genistein (1 μg/h) resembled changes found during melatonin infusion at the dose of 100 μg/h (Misztal et al.,
1997), whereas the high-dose infusion of genistein (10 μg/h) resulted in a very high elevation of prolactin, which lasted even 4 h after the infusion of genistein was terminated. Misztal et al. (1997, 2001) found that the stimulation of prolactin secretion by melatonin was not mediated by the changes in dopamine release. Endogenous opioid peptides were not a major component of this melatonin action either (Misztal et al., 2001). It might be possible that TRH (Robinson et al., 1996) mediated this melatonin action.

Recently, Romanowicz et al. (2004) reported that in ovariectomized ewes during seasonal anoestrus, infused with 10 μg of genistein, i.e. the same dose as we used in the present study, the plasma prolactin concentrations were increased during and after infusion of genistein. It was suggested that the short-term stimulatory effect of genistein on prolactin secretion might be mediated through an inhibition of dopamine neurons.

The results from the present study lead us to conclusion that the centrally administered genistein may stimulate the secretion of prolactin in intact ewes during the seasonal anoestrus. To explain, however, the neuroendocrine effects of genistein in sheep, further studies need to be conducted.

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STRESZCZENIE

Zmiany stężenia prolaktyny w krwi u anestralnych owiec po infuzji genisteiny do trzeciej komory mózgowej

Genisteina, należąca do grupy związków izoflawnowych, jest strukturalnie podobna do endogennych estrogenów, hamuje aktywność kinazy tyrozynowej, topoizomerazy DNA oraz ma właściwości antyoksydacyjne. Celem pracy było zbadanie wpływu genisteiny, podawanej bezpośrednio do ośrodkowego układu nerwowego, na sekrecję prolaktyny u owiec w okresie anestralnym. Doświadczenie przeprowadzono na 12 owcach rasy polska nizinna. Infuzje genisteiny lub płynu Ringera-Locke’a (kontrola, n=4) do trzeciej komory mózgowej wykonywano przy pomocy pompy mikroiniekcyjnej od godziny 12.01 do 16.01, w tempie 100 μl/godz. Podano dwie dawki genisteiny, 1 μg/100 μl/godz. (4 μg, n=6) lub 10 μg/100 μl/godz. (40 μg, n=5). Próbki krwi pobierano z żyły szyjnej od godz. 8.00 do 20.00, poprzez kateter założony dzień przed doświadczeniem. Nie stwierdzono istotnych różnic w stężeniu prolaktyny przed infuzją między grupami doświadczalną i kontrolną. U owiec, które otrzymały mniejszą dawkę genisteiny, stwierdzono istotne (P<0,05) wzrost stężenia prolaktyny w ciągu dwóch godzin po zakończeniu infuzji, w porównaniu z owcami kontrolnymi. Większa dawka genisteiny spowodowała wyraźny wzrost stężenia prolaktyny bezpośrednio po rozpoczęciu infuzji. Stężenie prolaktyny utrzymywało się następnie na poziomie istotnie (P<0,05) wyższym niż u owiec kontrolnych, do zakończenia doświadczenia (godz. 20.00). Istotne różnice (P<0,05) w stężeniu prolaktyny stwierdzono również między owcami otrzymującymi różne dawki genisteiny w ciągu ostatnich dwóch godzin kolekcji krwi. Uzyskane wyniki wskazują, że genisteina infundowana do ośrodkowego układu nerwowego anestralnych owiec stymuluje sekrecję prolaktyny w sposób zależny od dawki.