Liver cirrhosis is the terminal stage of most chronic liver conditions, with a high risk of mortality. Careful evaluation of the prognosis of cirrhotic patients and providing precise management are crucial to reduce the risk of mortality. Although the liver biopsy and hepatic venous pressure gradient (HVPG) can efficiently evaluate the prognosis of cirrhotic patients, their application is limited due to the invasive procedures. Child-Pugh score and the model for end-stage liver disease (MELD) score had been widely used in the assessment of cirrhotic prognosis, but the defects of subjective variable application in Child-Pugh score and unsuitability to all phases of liver cirrhosis in MELD score limit their prognostic values. In recent years, continuous efforts have been made to investigate the prognostic value of body fluid biomarkers for cirrhotic patients, and promising results have been reported. Since the collection of fluid specimens is easy, noninvasive, and repeatable, fluid biomarkers can be ideal indicators to predict the prognosis of cirrhosis. Here, we reviewed noninvasive fluid biomarkers in different prognostic functions, including the prediction of survival and complication development.

### 1. Introduction

Advanced liver cirrhosis (LC) is a life-threatening disorder with limited treatment options [1]. Annually, LC causes 1.03 million deaths worldwide, ranking as the 14th cause of disease death [2]. The patients with decompensated LC present an appropriate 10-fold risk of death compared with the general population [3]. The 6-week mortality of cirrhotic patients with variceal bleeding is about 10%–20% [4]. The mortality of cirrhotic patients with ascites is 14% within 1 year and 44% within 5 years [5]. The development of hepatocellular carcinoma (HCC) may accelerate the death of the disease at any stage. Precise assessment of prognosis followed by efficient therapies, such as endoscopic or pharmaceutical intervention, should be offered to patients with a high risk of death.

Liver biopsy and hepatic venous pressure gradient (HVPG) are able to well reflect the prognosis of LC patients [6–8], but they are limited in clinical applications due to the invasive procedures. Child-Pugh and the model for end-stage liver disease (MELD) scores are currently noninvasive methods to predict the prognosis of LC. However, Child-Pugh score has the defect of subjective variables used. Although the MELD score is more reproducible and accurate than Child-Pugh score in terms of the prediction of prognosis of LC, it is established in candidates for transjugular intrahepatic portosystemic shunt (TIPS); moreover, it is more suitable for end-stage patients [9]. Therefore, simple, reproducible, and noninvasive indicators are required to predict the prognosis of different types of LC patients.

Fluid biomarkers are ideal for predicting LC prognosis since they can be obtained by a simple, noninvasive, and reproducible way. Recently, lots of efforts have been made to explore the prognostic value of fluid biomarkers in different stages of LC. In the present paper, we summarized and discussed these biomarkers based on their own different prognostic value.

### 2. Prognostic Biomarkers for Prognosis of LC

#### 2.1. Prognostic Biomarkers of Survival Rate

There were several fluid biomarkers and panels of biomarkers developed to predict the overall survival of cirrhosis.

Red blood cell distribution width (RDW) is a common indicator of routine blood tests and it has a certain prognostic value. Turcato et al. [10] carried out a retrospective study that...
included 542 patients with acute decompensate LC and showed that the median RDW in this patient cohort was 15.7%; the mortality rate at 1 month was significantly lower in the patients with RDW less than 15.7% than in those with RDW more than 15.7% (8.2% vs. 21.8%). After adjusting for the age, hepatocellular carcinoma, renal function, hemoglobin value, and CLIF-C AD (Chronic Liver Failure Consortium Acute Decompensation) score, the dichotomous and continuous RDW was the independently predictive factor for predicting 1-month mortality (OR: 2.319 and 1.216, respectively). Besides, the model combined with RDW and CLIF-C AD score had a better ability of discrimination than CLIF-C AD (AUROC: 0.769 vs. 0.720; \( p = 0.006 \)).

C-reactive protein (CRP) is a nonspecific biomarker of inflammation [11, 12]. Recently, a number of studies showed that CRP had favorable prognostic value in LC. A retrospective study in 336 cirrhotic patients with spontaneous bacterial peritonitis (SBP) showed that the patients with serum levels of CRP \( \geq 5 \) mg/dL had significantly higher in-hospital mortality (45.5% vs. 28.6%) and lower rate of antibiotic responses (63.5% vs. 83.5%) than those with CRP < 5 mg/dL [13]. Another retrospectively observational study enrolled 202 LC patients with sepsis and 87 of them with repeated CRP measurements at day 4 and/or 5; the results showed that the ratio of the follow-up CRP level to the initial CRP level might be a useful prognostic factor to predict the mortality of LC patients (OR: 19.12), but not initial CRP level [14].

Blood leukocyte and neutrophil counts are conventional inflammatory markers. Recently, it is found that neutrophil to lymphocyte ratio (NLR) is a diverse functional prognosis predictor in some diseases, such as myocardial infarction, adult polymyositis and dermatomyositis, and spontaneous intracerebral hemorrhage [15–17]. NLR is related to immune dysregulation in patients with cirrhosis and is inexpensive to measure. Some studies showed that NLR is an emerging prognostic predictor in LC. Kalra et al. [18] evaluated the prognostic value of NLR in liver-related death in low MELD score patients listed for liver transplantation and found that high NLR was associated with liver-related death and independent of MELD score and cirrhosis stage. In another retrospective study, Rice et al. [19] found that increasing NLR was associated with death of LC patients within 1 year after nonelective hospitalization (HR: 2.17–2.84) and remained significant after adjusting for age, MELD score, hepatocellular carcinoma, and severity of acute-on-chronic liver failure (ACLF).

Aspartate transaminase (AST)/PLT (APRI) is proved to be a simple noninvasive index able to predict fibrosis and cirrhosis in patients with chronic hepatitis C [20]. Additionally, APRI is an independent prognostic index in primary biliary cirrhosis (PBC). Trivedi et al. [21] evaluated 1015 patients with PBC and found that in both derivation and validation cohorts, the patients with higher baseline or one-year APRI values had poorer transplant-free survival.

Serum albumin levels are able to predict the prognosis of LC. In compensated LC patients, serum albumin levels \(< 4 \) g/dL indicated a worse prognosis at 5 years compared with those with serum albumin levels \( \geq 4 \) g/dL [22]. In patients with decompensated LC, albumin is changed in both quantity and quality [23]. There are three homodimeric albumin isoforms, hdHA-DA, hdHA-L, and hdHA-native, in patients with cirrhosis and healthy controls. During 1 year follow-up, it was found that patients with a lower relative amount of the hdHA-DA or a higher relative amount of hdHA-native seemed to have a significantly longer survival (10.22 ± 0.44 vs. 8.28 ± 0.56 months; 9.94 ± 0.42 vs. 8.33 ± 0.64 months) [24].

Patients with LC are often complicated with electrolyte disorders. Hyponatremia is a common complication in LC. Several researches had proved that serum sodium levels are well prognostic in LC. Jenq et al. [25] observed 126 patients with LC who admitted to ICU and found that the patients with lower serum sodium concentration (\( \leq 135 \) mmol/L) at the first day of admission had a significantly higher in-hospital mortality (73% vs. 56%) and a poor 6-month survival rate after discharge, and the similar conclusion about 6-month survival was also achieved in the subset of patients with gastrointestinal hemorrhage.

Urokinase plasminogen activator receptor (uPAR) is a membrane glycoprotein. When a body suffers from a tumor, inflammation, and other pathological conditions, uPAR is upregulated and released into the blood circulation by hydrolysis of phospholipase or protease. suPAR is a soluble form of uPAR and a promising prognostic biomarker [26–28]. A study indicated that suPAR could predict the hospital and long-term mortalities in patients with critical illness and sepsis [29]. Zimmermann et al. [30] detected serum suPAR levels in 159 patients with chronic liver disease (98 cirrhosis) and demonstrated that serum suPAR levels were significantly higher in LC patients than those in non-LC patients, and the patients with suPAR levels > 9 ng/ml had significantly higher mortality during a median follow-up period of 478 days (RR: 8.5); they also found that suPAR levels were valuable for predicting 28-day mortality of decompensated cirrhosis and independent of MELD score and infection, with hazard ratio 4.83, sensitivity 71%, and specificity 71%, and remained valuable for 90-day mortality (HR: 2.93) [31].

Neutrophil gelatinase-associated lipocalin (NGAL) is a glycoprotein composed of a polypeptide chain with 178 amino acid residues. Due to its small molecular size, it can be filtered to the urine where it can be measured from. It shows diverse biological functions, such as embryonic development, cell differentiation, inflammation and immune response, apoptosis, signal transduction, carbohydrate lipid metabolism, tumor formation, invasion, and metastasis [32]. Recently, some studies reported that NGAL, in both blood and urine, is a novel sensitive biomarker with prognostic value for renal injury in LC patients. Gungor et al. [33] reported that plasma NGAL (pNGAL) had potential ability to predict the outcome of liver cirrhosis in patients with hepatorenal syndrome (HRS): the patients with pNGAL > 289.6 \( \mu g \) /L had a shorter survival time during 6-month follow-up, with AUROC 0.819 higher than CTP score (0.795), MELD-Na score (0.807), urine NGAL (0.686), and sensitivity 83.7% and specificity 72.2% in predicting mortality. Ariza et al. [34] investigated serum and urine NGAL in 716 patients in which 148 patients were with ACLF and
demonstrated that urinary NGAL (uNGAL) was markedly increased in ACLF compared with non-ACLF patients and was also an independent predictive factor for 28-day transplant-free mortality when combined with MELD score and leukocyte count (AUROC: 0.880).

Patients with LC are often associated with decreased body fat mass and insulin resistance. Resistin, a secreted protein produced by adipocytes and detectable in plasma, plays an important role in insulin resistance [35]. Yagmur et al. [36] detected resistin in 82 patients with chronic liver diseases (including 67 cirrhosis) and followed up for 6 years; they found that resistin levels were significantly elevated in cirrhosis patients compared with healthy controls and positively correlated with the stage of cirrhosis. However, da Silva et al. [37] did not observe a significant relationship between resistin levels and transplant-free survival: they detected plasma resistin and adiponectin levels and found that adiponectin, but not resistin, levels were associated with lower survival (HR: 1.034).

Copeptin is a fragment derived from the vasopressin precursor. Vasopressin is increased and plays a role in the development of complications in patients with cirrhosis. The measurement of vasopressin is difficult, but copeptin is more stable and easier to be detected in the blood. Moreno et al. [38] reported that plasma copeptin levels were increased along with the severity of liver disease in cirrhotic patients, with a 2.3-fold risk of death/liver transplantation in high-level patients compared with low-level ones, and independently predicted the 1-year mortality or liver transplantation (AUROC: 0.7). Kerbert et al. [39] investigated 61 LC patients at the waiting list for liver transplantation and found that patients with low levels of serum copeptin had a significantly higher transplant-free survival rate than those with high levels of copeptin. Subsequently, in 184 hospitalized cirrhotic patients, they demonstrated that patients with lower copeptin levels had a better transplant-free survival rate during 6 and 12 months of follow-up [40].

M30, M65, and M65EpiDeath belong to cytokerin 18-(CK-18-) based serum markers reflecting apoptotic or overall epithelial cell death. It was found that the increased serum levels of a fragment of CK-18 were associated with liver fibrosis and acute or chronic liver diseases [41–43]. Sekiguchi et al. [44] detected M30, M65, and M65EpiDeath in 130 PBC patients and found that they were all significantly higher in PBC than in healthy controls and were significantly correlated with fibrosis stage: the mortality was significantly higher in patients with high M65EpiDeath levels (HR: 6.13). Waidmann et al. [45] prospectively investigated 211 LC patients and found that these three CK-18-based cell death markers also could be potential biomarkers of severity of LC induced by other etiologies (mainly alcohol abuse and HCV infection), but only M65EpiDeath was associated with the patients prognosis, and interestingly, M65EpiDeath level was a factor for mortality in patients with compensated LC (HR: 11.483), but not decompensated LC.

Insulin-like growth factor-binding protein (IGFBP) is a group of polypeptides that are homologous with insulin. IGFBP-3, the most abundant type of IGFBPs in blood circulation, is mainly synthesized in Kupffer cells of the liver [46]. It is an indicator that can reflect the liver capacity. A study showed that serum IGFBP-3 was lower in cirrhotic patients than healthy controls, which indicated that low IGFBP-3 was associated with hepatocellular dysfunction [47]. Correa et al. [48] found that lower IGFBP-3 levels were associated with worse outcomes in patients with cirrhosis: the low IGFBP-3 group had lower survival rates than high IGFBP-3 group in outpatients followed up for a median of 20 months (72.1% vs. 88.6%) and in hospitalized patients at 90-day transplant-free survival (56.1% vs. 80.4%).

Pentraxin-3 (PTX-3) is a 40 kDa protein that contained 381 amino acids. Many cells are able to produce PTX-3 under inflammatory stimulation [49]. Narciso-Schiavon et al. [50] analyzed the circulating PTX3 levels in cirrhotic patients and found that higher PTX-3 levels indicated a significantly lower 90-day survival rate (54% vs. 77%). Fan et al. [51] also found that acute decompensated patients with high PTX-3 levels had a higher probability of in-hospital mortality (HR: 3.94) and 3-month mortality (HR: 3.52).

Myostatin is a cytokine able to strongly suppress skeletal muscle growth [52]. The serum concentration of myostatin in LC patients was significantly higher than the healthy [52, 53]. Nishikawa et al. [54] measured serum myostatin levels in 198 LC patients, and the results showed that the overall survival rates of patients with high myostatin were significantly lower than those with low myostatin: 1-, 3-, 5-, and 7-year cumulative survival rates were 96.0%, 77.9%, 53.0%, and 39.1%, respectively, in the high myostatin group and 96.4%, 87.6%, 77.6%, and 73.2%, respectively, in the low myostatin group after excluding hepatocellular carcinoma patients at baseline.

The levels of Wisteria floribunda agglutinin-1-colony-stimulating factor 1 receptor (WFA+-CSF1R) could reflect the progression of liver fibrosis and possibly evaluate the severity of LC [55]. Lio et al. [56] detected the levels of WFA+-CSF1R and total-CSF1R in 214 patients with liver diseases caused by HCV infection and found that patients with LC (n = 115) had significantly higher plasma levels of WFA+-CSF1R and total-CSF1R than the patients (n = 99) with chronic hepatitis (208.9 ng/ml vs. 82.3 ng/ml; 845.3 ng/ml vs. 536.4 ng/ml, respectively) and that the mortality was significantly higher in patients with WFA+-CSF1R ≥ 310 ng/ml (HR: 1.8), with an AUROC of 0.691. Wisteria floribunda agglutinin positive Mac-2 binding protein (WFA*-M2BP) is a novel and noninvasive biomarker for the assessment of liver fibrosis [57, 58]. Yamasaki et al. [59] enrolled 707 hepatitis C patients including 120 cases of LC and followed up for 20 years, and the results showed that the patients with the serum levels of WFA*-M2BP ≥ 1-4 and ≥ 4 at admission presented higher risk to develop HCC compared with WFA*-M2BP < 1 (HR = 5.115 and 8.318, respectively). In PBC patients, ones with serum WFA*-M2BP levels < 2.0 had a significantly higher survival rate of 10 years compared with the patients whose WFA*-M2BP value ≥ 2.0 (98.8% vs. 59.1%) [60].

Interleukin-22 (IL-22) is a crucial parameter of pathology in experimental liver damage and associated with the prognosis of LC. Kronenberger et al. [61] detected the
systemic levels of IL-22 in 120 LC patients and 40 healthy donors and found that IL-22 was more detectable in LC patients compared with healthy donors (74.1% vs. 10%), and the patients with elevated IL-22 levels had a reduced survival time (321 days vs. 526 days).

Lysophosphatidic acid (LPA), a secreted glycoprotein formed by lysophosphatidylcholine by autotaxin (ATX) which possesses both phosphodiesterase and lysophospholipase D activity, activates hepatic stellate cells and inhibits their apoptosis. A prospective cohort study included 270 cirrhotic patients and found that low levels of serum autotaxin indicated a longer survival time (HR: 0.575) [62].

Vitamin D is a necessary fat-soluble vitamin in the body, and its deficiency is associated with various chronic liver diseases [63]. A study prospectively recruited 324 patients with alcoholic liver disease in which there were 254 LC patients and were followed up for one year, and the low levels of vitamin D were associated with low overall survival time (HR: 5.95) [64].

Several biomarkers detected in urine also could predict the survival rate of patients with LC. Interleukin-18 (IL-18) is a proinflammatory protein able to regulate immune responses. Urinary IL-18 is associated with AKI and 90-day mortality in critically ill patients [65]. Recently, Tsai et al. [66] detected the concentration of the urinary IL-18 in 168 consecutive cirrhotic patients with severe sepsis and found that it was significantly higher in hospital survivors than in nonsurvivors and was an independent factor to predict mortality of the patients in the hospital. Monocyte chemoattractant protein-1 (MCP-1) is one of the key chemokines in regulating migration and infiltration of monocytes/macrophages and involved in various diseases. Graupera et al. [67] prospectively collected 218 patients with acute decompensated cirrhosis with a follow-up for at least 3 months and detected the levels of MCP-1, osteopontin (OPN), and trefoil-factor 3; the results showed that the levels of urinary MCP-1 were associated with 3-month readmission and death in univariate analysis, and in multivariate analysis, MCP-1 was an independent biomarker to predict 3-month hospital readmission (HR: 2.1) and the event combined with readmission or death (HR: 2.4).

Saliva samples can be collected easily, repetitively, and noninvasively. It is found that the saliva contains biomarkers associated with LC prognosis. Salivary caffeine clearance (CCl) could be used to evaluate the liver function and liver metabolic capacity [68, 69]. Jover et al. [70] calculated the CCl in 34 cirrhotic patients and followed up those patients for average 33.8 months and found that the survivors had a significantly higher CCl than the nonsurvivors (0.43 vs. 0.14) and CCl was an independent factor for predicting survival. However, the value of CCl is susceptible to other factors, such as treating with drugs that alter the hepatic metabolism of caffeine; thus, the application of CCl might be restricted in the clinic. Dysbiosis or altered gut microbiota in cirrhotic patients' stool and colonic mucosa would be likely associated with disease severity and systemic inflammation [71, 72]. Interestingly, salivary dysbiosis is related to the prognosis of LC patients. Bajai et al. [73] enrolled 102 cirrhotic patients and found that the patients who were hospitalized within 90 days had a lower salivary dysbiosis ratio compared with the patients' free hospitalization (0.15 ± 0.24 vs. 0.52 ± 1.2) and salivary dysbiosis ratio was an independent predictor of 90-day hospitalization (OR: 0.5).

2.2. Prognostic Biomarkers of Complication of Cirrhosis

2.2.1. Predicting the Renal Injury. It is not uncommon that cirrhotic patients combine with acute renal injury (AKI) that is associated with mortality [74]. Cirrhotic patients with renal failure had over 7-fold risk of death within 1 year compared with patients without renal failure [75]. Therefore, it is necessary to diagnose kidney injury early and implement effective strategies timely. Actually, serum creatinine (Cr) and its derived formulae in estimating glomerular filtration rate (GFR) are inaccurate to assess renal function [76].

Cystatin C (CysC) has been proposed as a specific marker for detecting a reduced GFR and an early indicator of impaired renal function. It is superior to Cr because its expression level is independent of age, sex, muscle mass, and inflammatory or malignant diseases [77]. HRS is a functional renal failure caused by intrarenal vasoconstriction [78]. Sharawy et al. [79] evaluated 80 cirrhotic patients with ascites and normal serum creatinine levels (<1.2 mg/dL) and found that serum CysC was a predictor of HRS development (OR: 2.1); when the cut-off value of serum CysC was set at 1.819 mg/L, the area under receiver operating characteristics curve (AUROC) for predicting the development of HRS within 12 months was 0.86, and the corresponding sensitivity and specificity were 88.8% and 67.7%, respectively. CysC was also a predictor for predicting the development of ACLF. Recently, Markwardt et al. [80] measured the plasma levels of CysC in 429 patients hospitalized for acutely decompensated cirrhosis and results showed that the baseline CysC level was not only a predictor of renal dysfunction (OR: 9.4) but also a predictor of HRS, ACLF, and 90-day mortality (OR: 4.2, 5.9, and 3.1, respectively). Maiwall et al. [81] developed AKI-Score including bilirubin, Cystatin C, and prior AKI to not only a predictor of renal dysfunction (OR: 2.1); when the cut-off value of serum CysC was set at 1.819 mg/L, the area under receiver operating characteristics curve (AUROC) for predicting the development of HRS within 12 months was 0.86, and the corresponding sensitivity and specificity were 88.8% and 67.7%, respectively. CysC was also a predictor for predicting the development of ACLF. Recently, Markwardt et al. [80] measured the plasma levels of CysC in 429 patients hospitalized for acutely decompensated cirrhosis and results showed that the baseline CysC level was not only a predictor of renal dysfunction (OR: 9.4) but also a predictor of HRS, ACLF, and 90-day mortality (OR: 4.2, 5.9, and 3.1, respectively). Maiwall et al. [81] developed AKI-Score including bilirubin, Cystatin C, and prior AKI to predict the new AKI in patients with LC. The C-index of the AKI-Score was 0.78 and 0.80 in training and validation groups, respectively, which was higher than MELD (0.71 and 0.73 in training and validation groups, respectively) and CTP (0.69 and 0.75 in training and validation groups, respectively). Besides, the AUROC of the AKI-Score (0.81 and 0.85 in training and validation groups, respectively) also was higher than the MELD score (0.70 and 0.74 in training and validation groups, respectively) and CTP (0.67 and 0.76 in training and validation groups, respectively).

NGAL is expressed in a variety of normal human tissues, such as bronchial epithelial cells, gastric wall cells, and proximal renal tubular epithelial cells. Nevertheless, NGAL shows a heterogeneous expression in the kidney, positive in proximal renal tubule epithelial cells, while negative in the distal renal tubule, glomerulus, and renal units [82]. Barreto et al. [83] found that levels of uNGAL were markedly higher in patients with AKI compared with ones without, with predictive value for persistent AKI (OR: 1.48) and 3-month survival rate (HR = 1.10); interestingly, patients with higher
uNGAL levels had a more probability to develop a second infection during hospitalization. Verna et al. [84] measured the levels of uNGAL in 118 patients with cirrhosis and also found that uNGAL could be a potential biomarker for predicting the mortality of inpatient with cirrhosis (OR: 6.05).

Aquaporin-2 (AQP2) is a water channel sensitive to vasopressin and located on the luminal side of collecting ducts of the kidney [85]. A portion of AQP2 can be excreted into urine and be detected. Busk et al. [86] analyzed baseline concentration of urine AQP2 in cirrhotic patients, and the results showed that it was able to significantly distinguish patients with the progression of renal insufficiency from the patients without and could be a predictor for 28-day mortality but not for 90-day mortality.

Kidney injury molecule-1 (KIM-1) is a type I transmembrane glycoprotein with a potential for the detection of tubular injury in renal diseases. Yap et al. [87] prospectively observed cirrhotic patients with normal serum creatinine and Child grade B or C for 12 weeks and found that urinary KIM-1 levels could be a potential biomarker for predicting the development of HRS in progressive cirrhotic patients (RR: 1.973), with AUROC of 0.78.

### 2.2.2. Predicting the Infection
LC patients are frequently complicated with a bacterial infection. A bacterial infection could promote the process of cirrhosis and eventually result in patient death [88, 89]. Preventive effective measures of cirrhotic patients with a high risk of infection might potentially decrease mortality. Several biomarkers had been found to have a potential ability to predict the infection in LC.

Lipopolysaccharide-binding protein (LBP), an acute-phase protein produced by hepatocytes, is found to be a biomarker able to predict the development of severe infection in LC patients. High serum LBP levels in the cirrhotic patients with ascites developing severe bacterial infection were significantly higher than those in patients with normal LBP levels (32.4% vs. 8.0%; RR: 4.49) [90]. LBP is a predictor of LC prognosis. The authors of [91] detected LBP levels in 88 patients with decompensated cirrhosis including 18 patients with infection and the others without infection and found that, in noninfective patients, those with lower level of LBP had a lower 90-day mortality than the high level (48.0% vs. 24.4%), and only high LBP (HR: 8.1) and MELD (HR: 1.1) were predictors of mortality in multivariate analysis, but this association was not observed in infective patients. In 58 critically ill cirrhotic patients with severe sepsis, Chen et al. [92] found that the cumulative survival rate at 28 days was higher in the high serum LBP group (>46 ng/mL) than that in the low serum LBP group (<46 ng/mL) (72.7% vs. 16.7%), with the AUROC of 0.809, sensitivity of 72.7%, and specificity of 83.3%. In addition, Papp et al. [93] did not observe an association between LBP levels and the probability of developing infections during a 3-month follow-up period in LC patients. Therefore, the value of LBP in the evaluation of infection and prognosis in LC patients needs more studies.

High blood levels of free cortisol indicate the severity of cirrhosis. Thevenot et al. [94] prospectively detected serum free cortisol (SFC) in 95 hemodynamically stable cirrhotic patients and found that the patients with levels of SFC ≥79 nmol/L had a significantly higher one-year mortality (26.2% vs. 3.4%). Another study including 143 LC patients with acute decompensation showed that patients with relative adrenal insufficiency had a higher probability of infection (41% vs. 21%), sepsis (27% vs. 9%), and death (22% vs. 7%) during follow-up of 3 months [95].

### 2.2.3. Predicting Gastroesophageal Varices
Acute variceal bleeding is a serious complication of LC. Despite the progress in the standardization of the supportive care and new therapeutic approach, the 6-week mortality of AVB was around 20% [96]. In the clinic, lacking fluid biomarkers could predict the occurrence of gastroesophageal varices and AVB.

Theoretically, lower platelet (PLT) counts indicate a higher risk of bleeding and poor prognosis in LC patients, but it is not supported by reports. Qamar et al. [97] found that, in patients with LC and portal hypertension, PLT counts could not predict the presence of gastroesophageal varices at admission and during follow-up. Other studies showed that PLT number was not a risk factor of major bleeding during a four-year observation period [98] and not a predictor of survival rate in patients with LC [99]. Kim et al. [100] found that the index named P2/MS, (PLT)²/[monocyte fraction (%)×segmented neutrophil fraction (%)], was negatively associated with the presence of esophageal varices in hepatitis B viral cirrhosis, and the patients with P2/MS ≥25 had lower risk of bleeding esophageal varices [101].

Patients with LC are often complicated with a disorder nutritional status, like protein-energy malnutrition (PEM) and sarcopenia, which is associated with the prognosis of LC [102]. Branched-chain amino acids (BCAAs) are energy substrates in muscles. Studies proved that cirrhotic patients had lower levels of BCAAs, but higher levels of aromatic amino acids (AAAs) [103, 104]. Ishikawa et al. [105] calculated the BCAA-to-tyrosine ratio (BTR) in 530 cirrhotic patients and found that patients with BTR <4 have more events including the worsening of esophageal and/or gastric varices, death, HCC, and liver failure (total events 53.3% vs. 11.8%, HR: 6.34).

### 2.3. Prognostic Biomarkers of Cirrhosis with ACLF
Acute-on-chronic liver failure (ACLF), characterized by multorgan failure, is the major cause of death in patients with acute decompensation of LC, and the mortality rate of patients with ACLF at 90 days is around 50% [106]. It is necessary to seek indicators for early diagnosis of ACLF.

Piano et al. [107] followed up 466 outpatients with cirrhosis and found that the baseline hemoglobin (HR = 0.07) is an independent prognostic biomarker of predicting the development of ACLF at one year.

uNGAL is a multifunctional biomarker to predict the prognosis of LC. Ariza et al. [34] found that uNGAL was not
only a biomarker that is independently associated with the ACLF (OR: 1.34) but also could predict the development of ACLF. After adjusting for the INR, Cr, and leukocyte count, the uNGAL still was an independent prognostic factor of the development of ACLF during hospitalization (OR: 1.31).

As previously described, the elevated copeptin levels indicated the higher short-term mortality, and the baseline serum copeptin was also a predictive biomarker of ACLF development. Kerbert et al. [108] followed up the patients without ACLF and identified that the copeptin was the independent prognostic factor for predicting the development of ACLF (OR:1.40).

2.4. Prognostic Biomarkers of Cirrhosis with Cardiovascular Dysfunction. Cirrhosis is associated with cardiovascular dysfunction. Several studies had proved that patients with cirrhosis were complicated with impaired cardiac contractility and performance [109, 110]. Brain natriuretic peptide (BNP) and its precursor NT-proBNP and C-type natriuretic peptide (CNP) are traditional biomarkers for evaluating cardiac dysfunction. Actually, those biomarkers had prognostic value in patients with LC. An earlier study found that increased BNP and NT-proBNP were associated with the severity of liver disease [111]. Pimenta et al. [112] found that BNP was an independent predictor of medium-term survival in decompensated cirrhosis. Higher serum BNP levels at preoperative liver transplantation or first postoperative day indicated a higher mortality rate [113, 114]. NT-proCNP is also associated with adverse prognosis of cirrhosis. Koch et al. [41] found that serum NT-proCNP was elevated in advanced liver diseases and indicated unfavorable prognosis when NT-proCNP levels >2 pmol/L (sensitivity 66.7%, specificity 72.8%, RR: 5.4).

2.5. Fluid Biomarkers to Improve the MELD Score and CTP. CTP and MELD are the most common score system to evaluate the prognosis of LC and have been widely used in clinical practice. Some studies attempted to incorporate prognostic biomarkers into CTP and MELD scores for improving the performance of the prediction of the prognosis of LC. MELD-Na score, combined with serum sodium and MELD score, was a presentative improvement of the MELD score. Heuman et al. [115] demonstrated that, in patients with a low MELD score (<21), the MELD-NA score had a better performance than the MELD score in predicting the 180-day mortality (c-statistic: MELD-Na vs. MELD: 0.768 vs. 0.638).

Combination of CRP and procalcitonin (PCT) with MELD score could improve the prediction of 30-day mortality (c-statistic: MELD-CRP, MELD-PCT, MELD-CRP-PCT vs. MELD: 0.79, 0.80, 0.81 vs. 0.76) and 90-day mortality (c-statistic: MELD-CRP, MELD-PCT, MELD-CRP-PCT vs. MELD: 0.83, 0.84, 0.85 vs. 0.81), and the similar results were obtained from the Mayo Clinic external validation cohort in the prediction of 30-day mortality (c-statistic: MELD-CRP vs. MELD:0.71 vs. 0.67) and prediction of 60-day mortality (c-statistic: MELD-CRP vs. MELD:0.69 vs. 0.65).

Kim et al. [116] established a model that combined serum sodium concentration with MELD score and found the MELD-Na score might have better performance in the assignment of priority of transplantation than MELD.

MELD-Cystatin Score showed a better performance than the MELD score and CTP score in predicting the probability of survival during a year follow-up (C-index: 0.84 vs. 0.83 and 0.74) [81]. But, Finkenstedt et al. [117] established a CysC-based MELD score (replace creatinine with CysC) and it was not superior to MELD score in predicting the 90-day mortality among LC patients. More studies are needed to clarify the prognostic value of incorporating CysC in the MELD model.

As mentioned, uNGAL is a biomarker that could predict the survival and occurrence of complications in patients with LC and also enhance the prognostic value of the MELD score. A model combined with uNGAL and MELD had a better performance in predicting the 28-day transplant-free mortality of LC patients (AUROC: model Vs. MELD: 0.86 vs. 0.81, p = 0.017) [34].

3. Conclusions

Liver biopsy and the measurement of HVPG can efficiently predict the prognosis of LC, but the invasive feature limits their applications in clinical practice. Although Child-Pugh score and MELD score are widely applied to evaluate the prognosis of LC, their drawbacks are obvious, such as subjectivity and incomprehensive scope of application. Accumulating evidences have shown that fluid biomarkers provide prognostic information of liver cirrhosis. Fluid biomarkers are objective measures and relative to different prognostic aspects of cirrhotic patients, including short- and long-term survival, complication development, and disease progression, and some biomarkers are “multifunctional”; for example, CysC predicts not only the mortality of LC but also the development of HRS. Importantly, fluid biomarkers have been found in blood, urine, and saliva, and all these fluids can be collected easily, repetitively, and noninvasively, which is favorable for clinical application. Some studies also attempted to incorporate those prognostic biomarkers into Child-Pugh and MELD score and got a better performance.

However, before these fluid biomarkers are applied to evaluate LC prognosis in clinical practice, several challenges are needed to be overcome, including clinical value to be confirmed by more studies, standardization of detection methods, and comparison with current methods in large samples. The overcoming of these obstacles will facilitate the utilization of fluid biomarkers in the prediction of LC prognosis and promote the precise management of LC patients by a combination of different biomarkers.

Disclosure

Si-Hai Chen and Qin-Si Wan are joint co-first authors.
Conflicts of Interest

The authors report no conflicts of interest in this work.

Authors’ Contributions

Si-Hai Chen and Kun-He Zhang were responsible for idea and design. Si-Hai Chen, Qin-Si Wan, and Kun-He Zhang contributed to literature search and data extraction. Si-Hai Chen, Qin-Si Wan, Ting Wang, and Kun-He Zhang wrote and revised the manuscript.

References

[1] P. Byass, “The global burden of liver disease: a challenge for methods and for public health,” BMC Medicine, vol. 12, no. 1, p. 159, 2014.

[2] R. Lozano, M. Naghavi, K. Foreman et al., “Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: a systematic analysis for the Global Burden of Disease Study 2010,” The Lancet, vol. 380, no. 9859, pp. 2095–2128, 2012.

[3] K. M. Fleming, G. P. Aithal, T. R. Card, and J. West, “All-cause mortality in people with cirrhosis compared with the general population: a population-based cohort study,” Liver International, vol. 32, no. 1, pp. 79–84, 2012.

[4] R. de Franchis and V. F. Baveno, “Revising consensus in portal hypertension: report of the Baveno V consensus workshop on methodology of diagnosis and therapy in portal hypertension,” Journal of Hepatology, vol. 53, no. 4, pp. 762–768, 2010.

[5] R. Planas, S. Montoliu, B. Ballesté et al., “Natural history of patients hospitalized for management of cirrhotic ascites,” Clinical Gastroenterology and Hepatology, vol. 4, no. 11, pp. 1385–1394, 2006.

[6] A. Berzigotti, M. Reig, J. G. Abraldes, J. Bosch, and J. Bruix, “Portal hypertension and the outcome of surgery for hepatocellular carcinoma in compensated cirrhosis: a systematic review and meta-analysis,” Hepatology, vol. 61, no. 2, pp. 526–536, 2015.

[7] C. Ripoll, R. Groszmann, G. Garcia-Tsao et al., “Hepatic venous pressure gradient predicts clinical decompenation in patients with compensated cirrhosis,” Gastroenterology, vol. 133, no. 2, pp. 481–488, 2007.

[8] C. Ripoll, R. J. Groszmann, G. Garcia-Tsao et al., “Hepatic venous pressure gradient predicts development of hepatocellular carcinoma independently of severity of cirrhosis,” Journal of Hepatology, vol. 50, no. 5, pp. 923–928, 2009.

[9] F. Durand and D. Valla, “Assessment of the prognosis of cirrhosis: Child-Pugh versus MELD,” Journal of Hepatology, vol. 42, no. 1, pp. S100–S107, 2005.

[10] G. Türcato, T. Campagnaro, A. Bonora et al., “Red blood cell distribution width independently predicts 1-month mortality in acute decompenation of cirrhotic patients admitted to emergency department,” European Journal of Gastroenterology & Hepatology, vol. 30, no. 1, pp. 33–38, 2018.

[11] K. M. Ho and J. Lipman, “An update on C-reactive protein for intensivists,” Anesthesia and Intensive Care, vol. 37, no. 2, pp. 234–241, 2009.

[12] K. Reinhart, W. Karzai, and M. Meisner, “Procalcitonin as a marker of the systemic inflammatory response to infection,” Intensive Care Medicine, vol. 26, no. 9, pp. 1193–1200, 2000.

[13] Y. Cho, S. Y. Park, J.-H. Lee et al., “High-sensitivity C-reactive protein level is an independent predictor of poor prognosis in cirrhotic patients with spontaneous bacterial peritonitis,” Journal of Clinical Gastroenterology, vol. 48, no. 5, pp. 444–449, 2014.

[14] Y. E. Ha, C.-I. Kang, E.-J. Joo et al., “Usefulness of C-reactive protein for evaluating clinical outcomes in cirrhotic patients with bacteremia,” The Korean Journal of Internal Medicine, vol. 26, no. 2, pp. 195–200, 2011.

[15] B. Azab, M. Zaher, K. F. Weiserbs et al., “Usefulness of neutrophil to lymphocyte ratio in predicting short- and long-term mortality after non-ST-elevation myocardial infarction,” The American Journal of Cardiology, vol. 106, no. 4, pp. 470–476, 2010.

[16] Y. J. Ha, J. Hur, D. J. Go et al., “Baseline peripheral blood neutrophil-to-lymphocyte ratio could predict survival in patients with adult polymyositis and dermatomyositis: a retrospective observational study,” PLoS One, vol. 13, no. 1, Article ID e0190411, 2018.

[17] Z. Ye, X. Ai, F. Fang et al., “The use of neutrophil to lymphocyte ratio as a predictor for clinical outcomes in spontaneous intracerebral hemorrhage,” Oncotarget, vol. 8, no. 52, pp. 90380–90389, 2017.

[18] A. M. Chavakis, S. M. Kanse, B. Yutzy, H. R. Lijnen, and K. T. Preissner, “Vitronectin concentrates proteolytic activity of the molecule,” Journal of Hepatology, vol. 60, no. 6, pp. 1249–1258, 2014.

[19] C. Ripoll, K. Bari, and G. Garcia-Tsao, “Serum albumin can identify patients with compensated cirrhosis with a good prognosis,” Journal of Clinical Gastroenterology, vol. 49, no. 7, pp. 613–619, 2015.

[20] F. Durand and D. Valla, “Assessment of the prognosis of cirrhosis: Child-Pugh versus MELD,” Journal of Hepatology, vol. 42, no. 1, pp. S100–S107, 2005.

[21] G. Türcato, T. Campagnaro, A. Bonora et al., “Red blood cell distribution width independently predicts 1-month mortality in acute decompenation of cirrhotic patients admitted to emergency department,” European Journal of Gastroenterology & Hepatology, vol. 30, no. 1, pp. 33–38, 2018.

[22] C. Ripoll, K. Bari, and G. Garcia-Tsao, “Serum albumin can identify patients with compensated cirrhosis with a good prognosis,” Journal of Clinical Gastroenterology, vol. 49, no. 7, pp. 613–619, 2015.

[23] F. Durand and D. Valla, “Assessment of the prognosis of cirrhosis: Child-Pugh versus MELD,” Journal of Hepatology, vol. 42, no. 1, pp. S100–S107, 2005.

[24] G. Türcato, T. Campagnaro, A. Bonora et al., “Red blood cell distribution width independently predicts 1-month mortality in acute decompenation of cirrhotic patients admitted to emergency department,” European Journal of Gastroenterology & Hepatology, vol. 30, no. 1, pp. 33–38, 2018.

[25] K. M. Ho and J. Lipman, “An update on C-reactive protein for intensivists,” Anesthesia and Intensive Care, vol. 37, no. 2, pp. 234–241, 2009.

[26] K. Reinhart, W. Karzai, and M. Meisner, “Procalcitonin as a marker of the systemic inflammatory response to infection,” Intensive Care Medicine, vol. 26, no. 9, pp. 1193–1200, 2000.

[27] Y. Cho, S. Y. Park, J.-H. Lee et al., “High-sensitivity C-reactive protein level is an independent predictor of poor
[28] M. Thune, B. Macho, and J. Eugen-Olsen, “suPAR: the molecular crystal ball,” Disease Markers, vol. 27, no. 3-4, pp. 157–172, 2009.

[29] A. Koch, S. Voigt, E. Sanson et al., “Circulating soluble urokinase plasminogen activator receptor is stably elevated during the first week of treatment in the intensive care unit and predicts mortality in critically ill patients,” Critical Care, vol. 15, no. 1, p. R63, 2011.

[30] H. W. Zimmermann, A. Koch, S. Seidler, C. Trautwein, and F. Tacke, “Circulating soluble urokinase plasminogen activator is elevated in patients with chronic liver disease, discriminates stage and aetiology of cirrhosis and predicts prognosis,” Liver International, vol. 32, no. 3, pp. 500–509, 2012.

[31] H. W. Zimmermann, P. A. Reuken, A. Koch et al., “Soluble urokinase plasminogen activator receptor is compartmentally regulated in decompensated cirrhosis and indicates immune activation and short-term mortality,” Journal of Internal Medicine, vol. 274, no. 1, pp. 86–100, 2013.

[32] J. B. Cowland and N. Borregaard, “Molecular characterization and pattern of tissue expression of the gene for neutrophil gelatinase-associated lipocalin from humans,” Genomics, vol. 45, no. 1, pp. 17–23, 1997.

[33] G. Gungor, H. Ataseven, A. Demir et al., “Neutrophil gelatinase-associated lipocalin in prediction of mortality in patients with hepatorenal syndrome: a prospective observational study,” Liver International, vol. 34, no. 1, pp. 49–57, 2014.

[34] X. Ariza, I. Graupera, M. Coll et al., “Neutrophil gelatinase-associated lipocalin is a biomarker of acute-on-chronic liver failure and prognosis in cirrhosis,” Journal of Hepatology, vol. 65, no. 1, pp. 57–65, 2016.

[35] C. M. Steppan, S. T. Bailey, S. Bhat et al., “The hormone resistin links obesity to diabetes,” Nature, vol. 409, no. 6818, pp. 307–312, 2001.

[36] E. Yagmur, C. Trautwein, A. M. Gressner, and F. Tacke, “Resistin serum levels are associated with insulin resistance, disease severity, clinical complications, and prognosis in patients with chronic liver diseases,” The American Journal of Gastroenterology, vol. 101, no. 6, pp. 1244–1256, 2006.

[37] T. Erotides da Silva, M. Costa-Silva, C. G. Correa et al., “Clinical significance of serum adiponectin and resistin levels in liver cirrhosis,” Annals of Hepatology, vol. 17, no. 2, pp. 286–299, 2018.

[38] J.-P. Moreno, E. Grandclement, E. Monnet et al., “Plasma copeptin, a possible prognostic marker in cirrhosis,” Liver International, vol. 33, no. 6, pp. 843–851, 2013.

[39] J. C. K. Annarein, L. Verbeke, F. W. T. Chiang et al., “Copeptin as an indicator of hemodynamic derangement and prognosis in liver cirrhosis,” PLoS One, vol. 10, no. 9, Article ID e0138264.

[40] J. C. K. Annarein, D. Weil, H. W. Verspaget et al., “Copeptin is an independent prognostic factor for transplant-free survival in cirrhosis,” Liver International, vol. 36, no. 4, pp. 530–537, 2016.

[41] A. Koch, H. W. Zimmermann, C. Baeck et al., “Serum NT-proCNP concentrations are elevated in patients with chronic liver diseases and associated with complications and unfavorable prognosis of cirrhosis,” Clinical Biochemistry, vol. 45, no. 6, pp. 429–435, 2012.

[42] G. V. Papanastoridou, E. Hadziyannis, E. Tsochatzis et al., “Serum apoptotic caspase activity as a marker of severity in HBeAg-negative chronic hepatitis B virus infection,” Gut, vol. 57, no. 4, pp. 500–506, 2008.

[43] X. Volkkmann, M. Anstaett, J. Hadem et al., “Caspase activation is associated with spontaneous recovery from acute liver failure,” Hepatology, vol. 47, no. 5, pp. 1624–1633, 2008.

[44] T. Sekiguchi, T. Umemura, N. Fujimori et al., “Serum cell death biomarkers for prediction of liver fibrosis and poor prognosis in primary biliary cirrhosis,” PLoS One, vol. 10, no. 6, Article ID e0131658, 2015.

[45] O. Waidmann, F. Brunner, E. Herrmann, S. Zeuzem, A. Piiper, and B. Kronenberger, “Cytokeratin 18-based cell death markers indicate severity of liver disease and prognosis of cirrhotic patients,” Liver International, vol. 36, no. 10, pp. 1464–1472, 2016.

[46] E. Arany, S. Afford, A. J. Strain, P. J. Winwood, M. J. Arthur, and D. J. Hill, “Differential cellular synthesis of insulin-like growth factor binding protein-1 (IGFBP-1) and IGFBP-3 within human liver,” The Journal of Clinical Endocrinology & Metabolism, vol. 79, no. 6, pp. 1871–1876, 1994.

[47] O. Colakoglu, B. Taskiran, G. Colakoglu, S. Kizildag, F. Ali Ozcan, and B. Unsal, “Serum insulin like growth factor-1 (IGF-1) and insulin like growth factor binding protein-3 (IGFBP-3) levels in liver cirrhosis,” Turkish Journal of Gastroenterology, vol. 18, no. 4, pp. 245–249, 2007.

[48] C. G. Correa, B. d. S. Colombo, M. F. Ronsoni et al., “Circulating insulin-like growth factor-binding protein 3 as prognostic biomarker in liver cirrhosis,” World Journal of Hepatology, vol. 8, no. 17, pp. 739–748, 2016.

[49] B. Bottazzi, C. Garlanda, G. Salvatori, P. Jeannin, A. Manfredi, and A. Mantovani, “Pentraxins as a key component of innate immunity,” Current Opinion in Immunology, vol. 18, no. 1, pp. 10–15, 2006.

[50] J. L. Narciso-Schiavon, J. G. Pereira, T. E. Silva et al., “Circulating levels of pentraxin-3 (PTX3) in patients with liver cirrhosis,” Annals of Hepatology, vol. 16, no. 5, pp. 780–787, 2017.

[51] W.-C. Fan, C.-C. Huang, Y.-Y. Yang et al., “Serum pentraxin-3 and tumor necrosis factor-like weak inducer of apoptosis (TWEAK) predict severity of infections in acute decompensated cirrhotic patients,” Journal of Microbiology, Immunology and Infection, vol. 50, no. 6, pp. 905–914, 2017.

[52] Y. Ellkina, S. von Haehling, S. D. Anker, and J. Springer, “The role of myostatin in muscle wasting: an overview,” Journal of Cachexia, Sarcopenia and Muscle, vol. 2, no. 3, pp. 143–151, 2011.

[53] P. S. Garcia, A. Cabbabe, R. Kambadur, G. Nicholas, and M. Caele, “Elevated myostatin levels in patients with liver disease,” Anesthesia & Analgesia, vol. 111, no. 3, pp. 707–709, 2010.

[54] H. Nishikawa, H. Enomoto, A. Ishii et al., “Elevated serum myostatin level is associated with worse survival in patients with liver cirrhosis,” Journal of Cachexia, Sarcopenia and Muscle, vol. 8, no. 6, pp. 915–925, 2017.

[55] M. Ocho, A. Togayachi, E. Iio et al., “A novel glycobiomarker, a serum ‘sweet-doughnut’ protein facilitates fibrosis evaluation and therapy of liver cirrhosis,” International Journal of Cancer, vol. 136, no. 6, pp. 1428–1437, 2014.

[56] E. Iio, M. Ocho, A. Togayachi et al., “Application of a glycoproteomics-based biomarker development method: alteration in glycan structure on colony stimulating factor 1 receptor as a possible glycoinhibitor candidate for evaluation of liver cirrhosis,” Journal of Proteome Research, vol. 13, no. 3, pp. 1428–1437, 2014.

[57] E. Iio, M. Ocho, A. Togayachi et al., “A novel glycobiomarker, Wisteria floribundaagglutinin macrophage colony-stimulating factor receptor, for predicting carcinogenesis of liver cirrhosis,” Critical Care Medicine, vol. 13, no. 4, pp. 1462–1471, 2016.

[58] A. Kuno, Y. Ikehara, Y. Tanaka et al., “A serum ‘sweet-doughnut’ protein facilitates fibrosis evaluation and therapy...
assessmnet in patients with viral hepatitis,” *Scientific Reports*, vol. 3, p. 1065, 2013.

[58] A. Kuno, T. Sato, H. Shimazaki et al., “Reconstruction of a robust glycodiagnostic agent supported by multiple lectin-assisted glycan profiling,” *Proteomics—Clinical Applications*, vol. 7, no. 9-10, pp. 642–647, 2013.

[59] K. Yamasaki, M. Tateyama, S. Abiru et al., “Elevated serum levels of Wisteria floribunda agglutinin-positive human Mac-2 binding protein predict the development of hepatocellular carcinoma in hepatitis C patients,” *Hepatology*, vol. 60, no. 5, pp. 1563–1570, 2014.

[60] T. Umemura, S. Joshita, T. Sekiguchi et al., “Serum Wisteria floribunda agglutinin-positive mac-2-binding protein level predicts liver fibrosis and prognosis in primary biliary cirrhosis,” *American Journal of Gastroenterology*, vol. 110, no. 6, pp. 857–864, 2015.

[61] B. Kronenberger, I. Rudloff, M. Bachmann et al., “Interleukin-22 predicts severity and death in advanced liver cirrhosis: a prospective cohort study,” *BMCMedicine*, vol. 10, p. 102, 2012.

[62] T. Pleli, D. Martin, B. Kronenberger et al., “Serum autotaxin is a parameter for the severity of liver cirrhosis and overall survival in patients with liver cirrhosis—a prospective cohort study,” *PloS One*, vol. 9, no. 7. Article ID e103532, 2014.

[63] M. T. Kitson and S. K. Roberts, “D-livering the message: the importance of vitamin D status in chronic liver disease,” *Journal of Hepatology*, vol. 57, no. 4, pp. 897–909, 2012.

[64] E. Trépo, R. Ouziel, P. Pradat et al., “Marked 25-hydroxyvitamin D deficiency is associated with poor prognosis in patients with alcoholic liver disease,” *Journal of Hepatology*, vol. 59, no. 2, pp. 344–350, 2013.

[65] S. Nisula, R. Yang, M. Poukkonen et al., “Predictive value of urine interleukin-18 in the evolution and outcome of acute kidney injury in critically ill adult patients,” *Survey of Anesthesiology*, vol. 59, no. 6, pp. 460–468, 2015.

[66] M.-H. Tsai, Y.-C. Chen, C.-W. Yang et al., “Acute renal failure in cirrhotic patients with severe sepsis: value of urinary interleukin-18,” *Journal of Gastroenterology and Hepatology*, vol. 28, no. 1, pp. 135–141, 2013.

[67] I. Graupera, E. Sola, N. Fabrellas et al., “Urinary monocyte chemotactic protein-1 is an independent predictive factor of hospital readmission and survival in cirrhosis,” *PloS One*, vol. 11, no. 6, Article ID e0157371, 2016.

[68] G. Jost, A. Wahlländer, U. V. Mandach, and R. Preisig, “Overnight salivary caffeine clearance: a liver function test suitable for routine use,” *Hepatology*, vol. 7, no. 2, pp. 338–344, 1987.

[69] F. W. Lewis and W. G. Rector Jr., “Caffeine clearance in cirrhosis. The value of simplified determinations of liver metabolic capacity,” *Journal of Hepatology*, vol. 14, no. 2-3, pp. 157–162, 1992.

[70] R. Jover, F. Carncier, J. Sanchez-Paya et al., “Salivary caffeine clearance predicts survival in patients with liver cirrhosis,” *The American Journal of Gastroenterology*, vol. 92, no. 10, pp. 1905–1908, 1997.

[71] J. S. Bajaj, D. M. Heuman, P. B. Hylemon et al., “Altered profile of human gut microbiome is associated with cirrhosis and its complications,” *Journal of Hepatology*, vol. 60, no. 5, pp. 940–947, 2014.

[72] J. S. Bajaj, P. B. Hylemon, J. M. Ridlon et al., “Colonic mucosal microbiome differs from stool microbiome in cirrhosis and hepatic encephalopathy and is linked to cognition and inflammation,” *American Journal of Physiology-Gastrointestinal and Liver Physiology*, vol. 303, no. 6, pp. G675–G685, 2012.

[73] J. S. Bajaj, N. S. Betrapally, P. B. Hylemon et al., “Salivary microbiota reflects changes in gut microbiota in cirrhosis with hepatic encephalopathy,” *Hepatology*, vol. 62, no. 4, pp. 1260–1271, 2015.

[74] J. M. Belcher, G. Garcia-Tsao, A. J. Sanyal et al., “Association of AKI with mortality and complications in hospitalized patients with cirrhosis,” *Hepatology*, vol. 57, no. 2, pp. 753–762, 2013.

[75] G. Fede, G. D’Amico, V. Arvaniti et al., “Renal failure and cirrhosis: a systematic review of mortality and prognosis,” *Journal of Hepatology*, vol. 56, no. 4, pp. 810–818, 2012.

[76] E. Cholongitas, V. Shusang, L. Marelli et al., “Review article: renal function assessment in cirrhosis—difficulties and alternative measurements,” *Alimentary Pharmacology & Therapeutics*, vol. 26, no. 7, pp. 969–978, 2007.

[77] Z.-H. Wan, J. J. Wang, S. L. You et al., “Cystatin C is a biomarker for predicting acute kidney injury in patients with acute-on-chronic liver failure,” *World Journal of Gastroenterology*, vol. 19, no. 48, pp. 9432–9438, 2013.

[78] F. Salerno, C. Camma, M. Enea, M. Rössle, and F. Womb, “Transjugular intrahepatic portosystemic shunt for refractory ascites: a meta-analysis of individual patient data,” *Gastroenterology*, vol. 133, no. 3, pp. 825–834, 2007.

[79] M. A. Sharawey, E. M. Shawky, L. H. Ali, A. A. Mohammed, H. A. Hassan, and Y. M. Fouad, “Cystatin C: a predictor of hepatorenal syndrome in patients with liver cirrhosis,” *Hepatology International*, vol. 5, no. 4, pp. 927–933, 2011.

[80] D. Markwardt, L. Holdt, C. Steib et al., “Plasma cystatin C is a predictor of renal dysfunction, acute-on-chronic liver failure, and mortality in patients with acutely decompensated liver cirrhosis,” *Hepatology*, vol. 66, no. 4, pp. 1232–1241, 2017.

[81] R. Maiwall, A. Kumar, A. Bhardwaj, G. Kumar, A. S. Bhadoria, and S. K. Sarin, “Cystatin C predicts acute kidney injury and mortality in cirrhotics: a prospective cohort study,” *Liver International*, vol. 38, no. 4, pp. 654–664, 2018.

[82] A. Friedl, S. P. Stoesz, P. Buckley, and M. N. Gould, “Neutrophil gelatinase-associated lipocalin in normal and neoplastic human tissues. Cell type-specific pattern of expression,” *Histochemical Journal*, vol. 31, no. 7, pp. 433–441, 1999.

[83] R. Barreto, C. Elia, E. Solà et al., “Urinary neutrophil gelatinase-associated lipocalin predicts kidney outcome and mortality in cirrhosis: a meta-analysis,” *Scientific Reports*, vol. 5, no. 4, pp. 5738–5755, 2015.

[84] E. C. Verna, R. S. Brown, E. Farrand et al., “Urinary neutrophil gelatinase-associated lipocalin predicts mortality and identifies acute kidney injury in cirrhosis,” *Digestive Diseases and Sciences*, vol. 57, no. 9, pp. 2362–2370, 2012.

[85] S. Nielsen, C. L. Chou, D. Marples, E. I. Christensen, B. K. Kishore, and M. A. Knepper, “Vasopressin increases water permeability of kidney collecting duct by inducing translocation of aquaporin-CD water channels to plasma membrane,” *Proceedings of the National Academy of Sciences*, vol. 92, no. 4, pp. 1013–1017, 1995.

[86] T. M. Busk, S. Möller, E. R. Pedersen et al., “Aqp0 in liver transplant recipients is a novel biomarker for the prognosis of de novo primary biliary cirrhosis,” *Digestive and Liver Disease*, vol. 49, no. 2, pp. 202–206, 2017.
M. Borzio, F. Salerno, L. Piantoni et al., “Bacterial infection in patients with advanced cirrhosis: a multicentre prospective study,” *Digestive and Liver Disease*, vol. 33, no. 1, pp. 41–48, 2001.

P. Tandon and G. Garcia-Tsao, “Bacterial infections, sepsis, and multiorgan failure in cirrhosis,” *Seminars in Liver Disease*, vol. 28, no. 1, pp. 026–042, 2008.

A. Albillos, A. de-la-Hera, and M. Alvarez-Mon, “Serum lipopolysaccharide-binding protein prediction of severe bacterial infection in cirrhotic patients with ascites,” *The Lancet*, vol. 363, no. 9421, pp. 1608–1610, 2004.

D. Agiasotelli, A. Alexopoulou, L. Vassileva et al., “High serum lipopolysaccharide binding protein is associated with increased mortality in patients with decompensated cirrhosis,” *Liver International*, vol. 37, no. 4, pp. 576–582, 2017.

Y.-Y. Chen, J.-M. Lien, Y.-S. Peng et al., “Lipopolysaccharide binding protein in cirrhotic patients with severe sepsis,” *Journal of the Chinese Medical Association*, vol. 77, no. 2, pp. 68–74, 2014.

M. Papp, Z. Vitalis, I. Altorjay et al., “Acutephaseproteinsincirrhosisaffectingmortalityinpatientswithcirrhosis,”*Liver International*, vol. 32, no. 4, pp. 603–611, 2012.

T. Thevenot, R. Dorin, E. Monnet et al., “High serum levels of free cortisol indicate severity of cirrhosis in hemodynamically stable patients,” *Journal of Gastroenterology and Hepatology*, vol. 27, no. 10, pp. 1596–1601, 2012.

J. Acevedo, J. Fernández, V. Prado et al., “Relative adrenal insufficiency in decompensated cirrhosis: relationship to short-term risk of severe sepsis, hepatorenal syndrome, and death,” *Hepatology*, vol. 58, no. 5, pp. 1757–1765, 2013.

G. García-Tsao, J. G. Abraldes, A. Berzigotti, and J. Bosch, “Portal hypertensive bleeding in cirrhosis: risk stratification, diagnosis, and management: 2016 practice guidance by the American Association for the study of liver diseases,” *Hepatology*, vol. 65, no. 1, pp. 310–335, 2017.

A. A. Qamar, N. D. Grace, R. J. Groszmann et al., “Platelet count is not a predictor of the presence or development of gastroesophageal varices in cirrhosis,” *Hepatology*, vol. 47, no. 1, pp. 153–159, 2008.

S. Basioli, V. Raparelli, L. Napoleone et al., “Platelet count does not predict bleeding in cirrhotic patients: results from the pro-liver study,” *American Journal of Gastroenterology*, vol. 113, no. 3, pp. 368–375, 2018.

E. G. Giannini, A. Moscatelli, M. Brunacci, P. Zentlin, and V. Savarino, “Prognostic role of mean platelet volume in patients with cirrhosis,” *Digestive and Liver Disease*, vol. 48, no. 4, pp. 409–413, 2016.

B. K. Kim, K.-H. Han, J. Y. Park et al., “Prospective validation of P2/MS noninvasive index using complete blood counts for detecting esophageal varices in B–viral cirrhosis,” *Liver International*, vol. 30, no. 6, pp. 860–866, 2010.

B. K. Kim, S. H. Ahn, K.-H. Han et al., “Prediction of esophageal variceal bleeding in B–viral liver cirrhosis using the P2/MS noninvasive index based on complete blood counts,” *Digestion*, vol. 86, no. 3, pp. 264–272, 2012.

N. Toshikuni, T. Arisawa, and M. Tsutsumi, “Nutrition and exercise in the management of liver cirrhosis,” *World Journal of Gastroenterology*, vol. 20, no. 23, pp. 7286–7297, 2014.

M. Holecek, “Ammonia and amino acid profiles in liver cirrhosis: effects of variables leading to hepatic encephalopathy,” *Nutrition*, vol. 31, no. 1, pp. 14–20, 2015.

H. Moriwaki, Y. Miwa, M. Tajika, M. Kato, H. Fukushima, and M. Shiraki, “Branched-chain amino acids as a protein- and energy-source in liver cirrhosis,” *Biochemical and Biophysical Research Communications*, vol. 313, no. 2, pp. 405–409, 2004.

T. Ishikawa, M. Imai, M. Ko et al., “Evaluation of the branched-chain amino acid-to-tyrosine ratio prior to treatment as a prognostic predictor in patients with liver cirrhosis,” *Hepatologyleasing*, vol. 8, no. 45, pp. 79480–79490, 2017.

R. Moreau, R. Jalan, P. Gines et al., “Acute-on-chronic liver failure is a distinct syndrome that develops in patients with acute decompensation of cirrhosis,” *Gastroenterology*, vol. 144, no. 7, pp. 1426–1437, 2013.

S. Piano, M. Tonon, E. Vettore et al., “Incidence, predictors and outcomes of acute-on-chronic liver failure in outpatients with cirrhosis,” *Journal of Hepatology*, vol. 67, no. 6, pp. 1177–1184, 2017.

A. J. C. Kerbert, H. W. Verspaget, A. A. Navarro et al., “Copeptin in acute decompensation of liver cirrhosis: relationship with acute-on-chronic liver failure and short-term survival,” *Critical Care*, vol. 21, no. 1, p. 321, 2017.

R. Estruch, J. Fernandez-Sola, E. Sacanella et al., “Relationship between cardiomyopathy and liver disease in chronic alcoholism+1,” *Hepatology*, vol. 22, no. 2, pp. 532–538, 1995.

M. Pozzi, S. Carugo, G. Boari et al., “Evidence of functional and structural cardiac abnormalities in cirrhotic patients with and without ascites,” *Hepatology*, vol. 26, no. 5, pp. 1131–1137, 1997.

J. H. Henriksen, J. P. Gotze, S. Fuglsang et al., “Increased circulating pro-brain natriuretic peptide (proBNP) and brain natriuretic peptide (BNP) in patients with cirrhosis: relation to cardiovascular dysfunction and severity of disease,” *Gut*, vol. 52, no. 10, pp. 1511–1517, 2003.

J. Pimenta, C. Paulo, A. Gomes et al., “B-type natriuretic peptide is related to cardiac function and prognosis in hospitalized patients with decompensated cirrhosis,” *Liver International*, vol. 30, no. 7, pp. 1059–1066, 2010.

F. H. Saner, T. Neumann, A. Canbay et al., “High brain-natriuretic peptide level predicts cirrhotic cardiomyopathy in liver transplant patients,” *Transplant International*, vol. 24, no. 5, pp. 425–432, 2011.

A. Toussaint, E. Weiss, L. Khoy-Ear et al., “Prognostic Value of Preoperative Brain Natriuretic Peptide Serum Levels in Liver Transplantation,” *Transplantation*, vol. 100, no. 4, pp. 819–824, 2016.

D. M. Heuman, S. G. Abou-Assi, A. Habib et al., “Persistent ascites and low serum sodium identify patients with cirrhosis and low MELD scores who are at high risk for early death,” *Hepatology*, vol. 40, no. 4, pp. 802–810, 2004.

W. R. Kim, S. W. Biggins, W. K. Kremers et al., “Hypotension and mortality among patients on the liver-transplant waiting list,” *New England Journal of Medicine*, vol. 359, no. 10, pp. 1018–1026, 2008.

A. Finkenstedt, L. Dorn, M. Edlinger et al., “Cystatin C is a strong predictor of survival in patients with cirrhosis: is a cystatin C-based MELD better?” *Liver International*, vol. 32, no. 8, pp. 1211–1216, 2012.