ANTIOXIDANT POTENTIAL OF POLYSACCHARIDE ISOLATED FROM METHANOLIC EXTRACT OF TINOSPORA CORDIFOLIA STEM BARK

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ABSTRACT

Objective: Investigation of the antioxidant efficacy of a novel polysaccharide isolated from the methanolic extract of Tinospora cordifolia stem bark with reference to its action as free radical scavengers using in vivo and in vitro approaches was performed in the study.

Methods: Lipid peroxidation (TBARS), superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase (GPx) levels were determined with and without polysaccharide treatment in the cell lines and breast and liver tissues. 7, 12-Dimethylbenz (α) anthracene-induced albino Wistar rats were used as animal models.

Results: The IC50 concentration of polysaccharide was 100 µM for both the cell lines. The lipid peroxidation levels were decreased in both breast cancer cell lines and liver and breast tissues. Polysaccharide treatment showed remarkably low levels of TBARS in breast cancer cells, and a significant reduction in the activity of SOD, GPx, and CAT levels in cancer groups was found to be increased with polysaccharide treatment. The alteration was found to be induced in MCF-7 and MDA-MB-231 cell lines and in breast and liver tissues of in vivo models.

Conclusion: The results showed that the polysaccharide from T. cordifolia has antioxidant potential on human breast cancer cell lines and tissues with induction of apoptosis.

Keywords: Phytotherapy, Apoptosis, Antioxidant, Free radicals.

INTRODUCTION

Breast cancer is one of the most frequent cancers in women of the developed and developing countries. Oxygen free radicals are generated during various metabolic processes, and for the excess production of free radicals, an antioxidant defense mechanism is generated in the body [1]. Oxidative damage to cells and macromolecules is considered to be the cause of several diseases such as coronary heart disease, arthritis, and various neurodegenerative diseases including cancer [2]. Reactive oxygen species are involved in a variety of important pathophysiological conditions including mutagenesis and carcinogenesis.

In addition, there are emerging evidences, suggesting that 7, 12-dimethylbenz (α) anthracene (DMBA) induces the production of reactive oxygen species (ROS) that result in lipid peroxidation, DNA damage, and depletion of cell’s antioxidant defense systems [3]. Such alterations or imbalance in ROS and antioxidants can be balanced with the help of plants having antioxidant properties [4]. In recent days, plant-based therapies have emerged as a potential remedy for all human ailments including cancer [5]. Oxidative damage to biomolecules is caused due to the release of oxygen free radicals and oxidants which tend to generate oxidative stress [6,7]. The role of oxidative stress and lipid peroxidation in tumor development was studied in vivo [8]. There are certain detoxifying enzymes and small scavenger molecules at cellular levels acting as antioxidant defenses. The intracellular ROS-scavenging system includes superoxide dismutases (SOD), catalases (CAT), and glutathione peroxidase (GPx), which are potential free radical scavengers and are signified as cancer inhibitors at different stages of carcinogenesis [9]. Tinospora cordifolia is a widely used medicinal herb for its potential biological activity against numerous human disorders [10,11]. Various compounds explored from this plant have been recognized to possess several pharmacological activities [12], most importantly anti-diabetic [13] and anti-clastogenic [14] effects, and in such a way, the polysaccharide isolated from T. cordifolia has been experimentally reported to possess antidiabetic efficacy on type 1 diabetic animal models [15].

However, there are no proven evidences on the antioxidant potential of polysaccharide isolated from T. cordifolia, and hence, the present study was focused on identifying the antioxidant property of polysaccharide isolated from methanolic extract of T. cordifolia stem bark on MCF-7 and MDA-MB-231 cancer cell lines, and its in vivo activities on albino Wistar rat breast and liver tissues were also examined.
METHODS

Isolation of polysaccharide

Polysaccharide was isolated from the methanolic extract of T. cordifolia stem bark, by the protocol of Rajalakshmi et al. [15].

Cell line

Human breast cancer cell lines (MCF-7 and MDA-MB-231) were procured from ATCC and were cultured in DMEM culture medium with 10% FBS at 5% CO₂, and 37°C. Cells werePassaged using trypsin EDTA at 70–80% confluence.

Compound preparation

1 mM stock solutions of the polysaccharide were prepared with dimethyl sulfoxide (DMSO). From the stock, the compounds were prepared at different micromolar concentrations (10, 20, 30, 40, 80, and 100 µM) with serum-free medium for the test. The concentration of DMSO was aimed to not exceed 0.01%.

In vitro studies

Treatment protocol

Cells were seeded in 6-well plate (1.8 × 10⁶ cells/well) in medium containing 10% FBS and incubated for 24 h under 5% CO₂, at 37°C for attachment. The cells were then washed with 1× PBS, and polysaccharide (100 µM) was added to the wells containing MCF-7 cells and incubated for 24 h. The medium was then removed, and the cells were washed with PBS. Finally, the cells were collected by adding trypsin EDTA, pelleted, and the supernantant was used for further assays.

Determination of TBARS

A lipid peroxide level was estimated by the method reported by Ohkawa et al. [16]. The assay mixture containing 0.1 mL of the cell and tissue lysate, 0.2 mL of 0.1% sodium dodecyl sulfate, 1.5M of 20% acetic acid (adjusted to pH 3.5 with NaOH), and 1.5 mL of 0.6% aqueous solution of thiobarbituric acid was heated for 60 min at 95°C. Thereafter, the mixture was cooled and extracted with 5 mL of a mixture of n-butanol and pyridine (15:1 v/v). After centrifugation at 4000 rpm for 10 min, the organic phase was assayed spectrophotometrically at 532 nm and the lipid peroxide levels of TBARS were determined (nmol/mL).

Determination of SOD

SOD level was estimated by the method described by Kakkar et al. [17]. The assay mixture contained 0.1 ml of the cell and tissue lysate, 1.2 mL of sodium pyrophosphate buffer (pH 8.3, 0.025 M), 0.1 mL phenazine methosulfate (186 µM), 0.3 mL of 300 µM nitroblue tetrazolium, and 0.2 mL of NADH (780 µM). The reaction was started by the addition of NADH. After incubation at 30°C for 90 s, the reaction was stopped by the addition of 1 mL of glacial acetic acid. The reaction mixture was stirred vigorously with 4 mL of n-butanol, and the mixture was allowed to stand for 10 min. After centrifuging the mixture, the n-butanol layer was separated. Color intensity of the chromogen in n-butanol was measured at 520 nm spectrophotometrically, and the concentration of SOD was expressed as unit/min/mg of protein.

Determination of CAT

CAT level was measured by the method described by Aebi et al. [18]. A volume of 0.1 mL of cell and tissue lysate was added to a cuvette containing 1.9 mL of 50 mM phosphate buffer (pH 7.0). A reaction was started by the addition of 1 mL of freshly prepared 30 mM H₂O₂. The rate of decomposition of H₂O₂ was measured spectrophotometrically from changes in absorbance at 240 nm. The activity of CAT was expressed as µ mol H₂O₂ consumed/min/mg protein.

Assay of GPs

GPs level was measured by the method described by Rotruck et al. [19]. To 0.2 mL of the cell and tissue lysate, 0.2 mL of 0.4 mM EDTA, 0.1 mL of sodium azide, 0.2 mL of 2 mM GSH, and 0.1 mL Zn₂⁺ solution, 0.4 mL of 0.4 M phosphate buffer (pH 7.0) was added. The mixture was incubated at 37°C for 10 min, and 0.5 mL of 10% TCA was added and centrifuged at 2000 rpm for 10 min. The supernatant was collected and 0.1 mL of 0.04% DTNB solution was added to it. Optical density was read at 420 nm against blank, and the results were obtained. The activity of GPs was expressed as GSH utilized/min/mg of protein.

In vivo studies

Animals

Female albino Wistar rats aged between 50 and 55 days were procured from Tamil Nadu Veterinary Sciences, Madhavaram. The animals were maintained under controlled environmental conditions on alternative 12 h dark/light cycle. Commercial pelleted feed supplied by Sai Enterprises Ltd., Chennai, and water ad libitum were given to animals. This research work on albino Wistar rats was sanctioned and approved by the Institutional Animal Ethical Committee (IAEC No. 07/2013).

Experimental setup

The animals were divided into five groups of five animals each. Group I animals served as normal control, Group II was normal animals supplemented with polysaccharide (20 mg/kg body weight [bw]), Group III was animals treated with 20 mg of DMB in 1 mL corn oil to induce breast cancer, Group IV animals were treated with DMB and simultaneously supplemented with polysaccharide (20 mg/kg bw), and Group V animals were treated with DMB and simultaneously supplemented with paclitaxel (1 mg/kg bw). The overall induction and treatment period was 3 months for all groups. After the experimental period, the animals were sacrificed by decapitation, breast and liver tissues were dissected out, and tissue homogenates were prepared in 0.1 M Tris-HCl buffer pH 7.4 which was stored at 80°C, until its use for further analysis.

Antioxidant enzyme analysis

The breast and liver tissue homogenates were used for the estimation of antioxidant levels such as SOD [17], CAT [19], GPs [19], and lipid peroxidation (LPO) [16].

Statistical analysis

The data were analyzed using the SPSS Windows Students version software. For all the measurement, one-way ANOVA followed by Student–Newman–Keuls (SNK) test was used to assess the statistical significance of the difference between control and treated groups. A statistically significant difference was considered at the level of p < 0.05.

RESULTS

Effect of polysaccharide on TBARS level

Analysis of TBARS is the widely used method for the evaluation of lipid peroxidation. TBARS level in breast cancer cell lines was found to be altered, and a significant decrease in the levels was examined on treatment with 100 µM polysaccharide in both MCF-7 and MDA-MB-231 cell lines (Fig. 1). In vivo experiments on breast and liver tissues (Fig. 2) observed a significant increase in lipid peroxidation level in DMBA-induced rats (Group III) when compared to normal (Group I) animals. No such significant increase in TBARS level was observed in polysaccharide-supplemented DMBA-induced animals (Group IV).

Group I and Group V (standard drug-paclitaxel supplemented DMBA-induced animals) showed normal level of lipid peroxidation. Hence, the results suggested that the polysaccharide regulates a normal lipid peroxidation level in both in vitro and in vivo conditions.

Effect of polysaccharide on SOD, CAT, and GPs levels

Variation in the activities of antioxidant enzymes such as SOD, CAT, and GPs that are usually altered at lower levels in cancer conditions was examined. A significant increase in the activity of antioxidant enzymes was noted on polysaccharide-treated MCF-7 and MDA-MB-231 cell lines, when compared with the cancer control cells (Fig. 3). It is also proved through the in vivo studies on the breast (Fig. 4) and liver tissues (Fig. 5). Similarly, increase in the activity of these antioxidant enzymes was identified in polysaccharide-treated DMBA-induced animals and it was maintained near to normal, whereas DMBA-induced animals had a
reduction in the level of these enzymes. Therefore, it is suggested that normal levels of SOD, CAT, and GPx levels were maintained in both in vitro and in vivo conditions with polysaccharide treatment.

DISCUSSION

Natural antioxidants are characterized by their ability to scavenge free radicals. Medicines from herbal resources are preferred over the synthetic drugs for their better bioavailability and minimal side effects [20]. Plants that possess anticancer activity are also found to have a potential antioxidant activity, which is correlated with the cytotoxic efficacy of the molecules [21]. Many studies have found that the antioxidant property of a plant has a potential role in inhibiting the growth of cancer cells. In this study, the antioxidant potential of the polysaccharide isolated from the methanolic extract of T. cordifolia stem bark was studied on MCF-7 and MDA-MB-231 cell lines and on female albino Wistar rat breast and liver tissues.

Oxidative stress, which causes the development of cancer, is created by various underlying factors which include the excessive production of ROS that is responsible for the high level of lipid peroxidation and a deficit in antioxidant balance [22,23]. Antioxidant-rich plants are capable to inhibit the growth of several cancer cell lines, including breast cancer [24]. From the present study, it is found that, both in the in-vitro and in-vivo conditions, the level of lipid peroxidation was decreased and the level of antioxidant enzymes was increased in the polysaccharide-treated group of animals.

The SOD activity was very low in almost all types of cancer which indicates the reduction in the ROS defending mechanisms, and this low level of SOD leads to the increase of superoxide anions which in turn promotes cancer cell proliferation by acting as a second messenger. Inhibition of the development of cancer can be brought about by the increased level of SOD. The results of this study proved that the polysaccharide treatment increased the level of SOD in the cancer cell lines.
An anticancer agent with antioxidant activities balances the ROS levels, inhibits further proliferation of cancer cells, and allows apoptosis to occur. Besides, polysaccharide-treated MCF-7 and MDA-MB-231 cells enhanced the levels of the antioxidant enzymes and thereby decreased the oxidative stress. Increase in SOD and CAT is reported to inhibit tumor progression with less proliferation and migration of the cancer cells [25,26]. In the present study, there was a significant increase in the CAT levels in the polysaccharide-treated cancer cell lines when compared to DMBA-induced cancer groups.

GPx inhibits cancer initiation and metastasis in many of the cancers [27]. This may be due to the reduction of ROS-mediated DNA damage or mutations that induce carcinogenesis. Thus, the increase in the antioxidant enzymes activity is attributed to the induction of cell apoptosis. In this study, GPx levels in the polysaccharide-treated cancer groups were increased which were initially low.

Thus, the polysaccharide-treated groups exhibited normal range of lipid peroxidation and antioxidant levels. These alterations maintained a normal balance between the antioxidant defense system and the ROS generation, thereby inhibiting the cancer cell proliferation of MCF-7 and MDA-MB-231 cell lines.

CONCLUSION
Thus, the present study proved that the polysaccharide isolated from the methanolic extract of T. cordifolia stem bark has an efficient antioxidant property on both in vitro cell lines and in vivo animal models. Elevated levels of lipid peroxidation and decreased activities of antioxidant enzymes in both MCF-7 and MDA-MB-231 cell lines and in breast and liver tissues of DMBA-induced breast cancer female albino Wistar rats were prevented on treatment with the compound. Thus, it is evident that the abnormality with ROS and antioxidant enzymes was normalized by the polysaccharide which subsequently resulted in the inhibition of cancer. Hence, the novel polysaccharide can be used as an antioxidant agent with further explorations.

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Research guidance - Manikkam Rajalakshmi. Laboratory and experimental work and writing the article - Antony Ludas. Manuscript writing - Sabapathy Indu. Laboratory work - Sekar Hinduja. Laboratory work - Anthonisamy Kumari Nirmala.

CONFLICTS OF INTEREST
All authors have none to declare.
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