Research Article

The Beneficial Effect of *Equisetum giganteum* L. against *Candida* Biofilm Formation: New Approaches to Denture Stomatitis

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*Equisetum giganteum* L. (*E. giganteum*), Equisetaceae, commonly called “giant horsetail,” is an endemic plant of Central and South America and is used in traditional medicine as diuretic and hemostatic in urinary disorders and in inflammatory conditions among other applications. The chemical composition of the extract EtOH 70% of *E. giganteum* has shown a clear presence of phenolic compounds derived from caffeic and ferulic acids and flavonoid heterosides derived from quercitin and kaempferol, in addition to styrylpyrones. *E. giganteum*, mainly at the highest concentrations, showed antimicrobial activity against the relevant microorganisms tested: *Escherichia coli*, *Staphylococcus aureus*, and *Candida albicans*. It also demonstrated antiadherent activity on *C. albicans* biofilms in an experimental model that is similar to dentures. Moreover, all concentrations tested showed anti-inflammatory activity. The extract did not show cytotoxicity in contact with human cells. These properties might qualify *E. giganteum* extract to be a promising alternative for the topic treatment and prevention of oral candidiasis and denture stomatitis.

1. Introduction

Denture stomatitis (DS) is an inflammatory disease that affects the oral mucosa underlying a removable dental prosthesis, both partial and total. DS is considered a very common disease affecting denture wearers, especially those with maxillary complete dentures [1, 2]. Due to the strong relationship between DS and *Candida* fungus, the use of proper hygiene aids by the denture wearer is fundamental, by physical means, such as brushing, and, if necessary, chemical, and immersion of dentures in disinfectant solutions. Local or systemic antifungal therapy may still be an option in some cases of DS [3]; however, one should know the predisposing factors related to the user and eliminate them or change them whenever possible. There is no sense in using drug therapy, if it is not associated with awareness and change in the factors that contribute to DS.

Currently, treatments directed to DS include topical antifungal therapy, such as several formulations based on nystatin, systemic antifungal medication such as azoles, attention to oral hygiene, denture disinfection procedures, removal of the denture overnight, and replacement of older dentures [4].

Continued use of topical antifungal agents, nystatin suspensions [5], and disinfectants, such as sodium hypochlorite (NaOCl), glutaraldehyde, and chlorhexidine, can induce changes in the properties of the resin surface, such as roughness, hardness, and wettability, which can contribute to fungal adhesion [5–7]. The continued use of systemic antifungal therapy can also lead to serious adverse effects, such as...
2. Materials and Methods

2.1. Plant Material and Extract Preparation. The aerial parts of E. giganteum were collected in November 2011 at the Jardim Botânico Municipal de Bauru, SP, Brazil (22°20'30"S and 49°00'30"W). Voucher specimens were prepared, identified, and deposited at the Herbarium of the UNESP, São Paulo State University “Júlio de Mesquita Filho”, UNBA (Bauru, SP, Brazil) under number 5795. The fresh plants were dried at 40°C for 48 h and the powdered raw material (1.3 kg) was extracted with EtOH/H₂O (7:3 v/v) by percolation at room temperature. The filtrate was concentrated to dryness under reduced pressure at 40°C providing the hydroethanolic extract (70% EtOH) with a yield of 8.24% (364 g).

2.2. Chemical Analysis by UHPLC-PAD-ESI-MSⁿ. The chemical profile of the 70% EtOH extract of E. giganteum was obtained by means of Accela High Speed LC (Thermo Scientific, San Jose, CA, USA), Thermo Scientific column (50 × 2.1 mm, 1.9μm), with PAD and coupled to an Accela (Thermo Scientific) LCQ Fleet with Ion Trap (IT) 3D and ionization by electrospray (ESI). The mobile phase consisted of ultrapure water (eluente A) and methanol (eluente B), both mixed in 0.1% of formic acid, and we utilized a proportion of 5–100 of A in B during 20 min. The experimental protocol used the following parameters: injection volume: 10.0 μL; column temperature: 25°C; and flow ratio: 0.4 mL min⁻¹ and the chromatogram at 254 nm. The effluent from the UHPLC was directed into the ESI probe. The second scan event was an MS/MS experiment using a data dependent scan on deprotonated molecule [M-H]⁻ with collision energy to 10–25%. After being dissolved in MeOH-H₂O (85:15), the extract was infused in the ESI source by flow injection analysis (FIA) using a syringe pump at 5 μg/mL.

2.3. Microorganisms. For this study, we used Staphylococcus aureus (ATCC6536), Escherichia coli (O:124), and C. albicans (SC 5314). C. albicans were grown in YEPD broth (Difco, Sparks, MD, USA) and tested in Sabouraud broth (Difco, Sparks, MD, USA). Bacteria were grown and tested in brain-heart infusion broth (BHI) (Difco, Sparks, MD, USA).

2.4. Antimicrobial Assay. In vitro antimicrobial action was analyzed by the broth microdilution method with the objective of obtaining the minimum inhibitory concentration (MIC) of the E. giganteum extract. Bacterial and fungal suspensions (1 × 10⁷ cells/mL) were, respectively, inoculated into BHI and Sabouraud broth using 96-well plates. Wells containing each inoculum with the 1% NaOCl solution (CTRL/NaOCl) and culture medium (Sabouraud or BHI broth) (CTRL/Medium) served as the positive and negative control, respectively.

2.5. Antiadherent Assay. Ninety specimens of heat-polymerized acrylic resin (2.5 × 2.0 × 0.5 cm) were fabricated according to the manufacturers’ directions (Lucitone 550; Dentsply International Inc., York, USA). Each surface of specimens was abraded manually with high grade silicon carbide abrasive paper (P80; Norton Abrasivos, São Paulo, Brazil) [27], to promote a rough surface of approximately 2 microns. After this, the specimens were ultrasonically cleansed in sterile water (Ultrasonic Cleaner, Arotec, São Paulo, Brazil) [28] and sterilized with ethylene oxide (Acceil, Campinas, Brazil).
According to the different treatments, the resin specimens were divided into five groups: E50, E25, and E16 (50, 25, and 16 mg/mL) of *E. giganteum* extract; CTRL/PBS, sterile phosphate buffered saline (pH 7.2); or CTRL/NaOCl-1% NaOCl solution. The treatments were performed in 6-well plates, at room temperature for 10 min. Sequentially, the specimens were incubated for 90 min at 37°C in RPMI-1640 (GIBCO, Grand Island, NY) at 37°C for 12 h at 75 rpm. Three specimens for group were stained by LIVE/DEAD BacLight Bacterial Viability Kit solution (Molecular Probes, Eugene, OR, USA) [29] and incubated in the dark for 15 min at room temperature.

Subsequently, the specimens were conditioned in the confocal laser scanning microscopy (Leica Microsystems GmbH, Mannheim, Germany), from which digital images were obtained. The viable cells were visualized in green, while dead cells were visualized in red. With the use of BioImage Software program, the biofilm mass was quantified including both viable and nonviable cells and expressed as μm². The more the biofilm mass on the specimens decreased, the higher the antiadherent activity was.

2.6. Cell Culture. HPEC were cultivated as previously described by Klingbeil et al. [30] and human monocytes were obtained from thirty-milliliter volumes of peripheral blood, which were drawn from apparently healthy five volunteers. After obtaining peripheral blood mononuclear cells by cell separation using Histopaque 1077 gradients, according to the manufacturer’s instructions (Sigma-Aldrich Company), monocytes were counted using neutral red (0.02%) and incubated in 96-well culture plates by 2 h at 37°C in 5% CO₂. Following, nonadherent cells were removed by aspiration. Monocytes were cultivated in RPMI-1640 (GIBCO, Grand Island, NY), supplemented with 10% FBS (GIBCO) and 1% penicillin-streptomycin (GIBCO). The medium was replaced every other day until the cells grew to about 70% confluence. The experimental protocol used in this study was approved by the Committee on Ethics in Human Experimentation, of Bauru School of Dentistry, University of São Paulo (grant number 6706412.0.0000.5417, November 8, 2012), and written informed consent was obtained from all patients. This study was performed in accordance with the Declaration of Helsinki.

2.6.1. Cell Viability Assay. The different extracts were evaluated regarding cytotoxicity by using MTT (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide) colorimetric assay (Sigma Chemical Co., St. Louis, USA). Briefly, monocytes (2 x 10⁵ cells/well) suspended in 200 μL of medium were plated into 96-well tissue culture plates for 24 h. Thereafter, 200 μL of fresh medium containing various concentrations of *E. giganteum* extracts (50, 25, 16, 8, and 4 mg/mL) was added to each well and incubated for time intervals of 1, 12, and 24 hours and compared with control cells (untreated and water). After these periods, the plates were washed twice in PBS and 200 μL of fresh medium mixed with 10 μL of 5 mg/mL MTT dye in PBS was added to the wells for 4 h in a humidified 5% CO₂ atmosphere at 37°C. After the removal of MTT solution, 150 μL of dimethyl sulfoxide (DMSO-LGC Biotecnologia, São Paulo, Brazil) was added and mixed to dissolve the dye crystals. The absorbance was measured at 500 nm by means of a multiplate reader (Synergy Mx Monochromator-Based, Biotek, Washington, DC, USA). The same assay was performed with human palatal epithelial cells (HPEC) (10⁴ cells/well) for 12 and 24 hours.

2.7. Measurement of Intracellular Reactive Oxygen Species (ROS). Endogenous amounts of ROS in human monocytes were determined using the fluorescent probe 2',7'-dichlorofluorescin diacetate (Cell Rox Deep Red Reagent; Life Technologies, Grand Island, NY, USA). Monocytes (2 x 10⁵ cells/well) were in vitro incubated with *E. giganteum* extract (50, 25, 16, 8, and 4 mg/mL) for 1 hour at 37°C in 5% CO₂ atmosphere. After, they were stimulated with LPS at 5 μg/mL or *C. albicans* SC5314 (1 x 10⁶ cells/mL) for three hours and incubated with 5 μM of Cell Rox at 37°C in 5% CO₂ atmosphere for 30 min. The fluorescence intensities (FIs) of the cells were measured using the multiplate reader at 640/665 nm, at 37°C.

2.8. Statistical Analysis. The data are expressed as the mean ± standard deviation of at least three independent experiments, in triplicate. The data were analyzed, where appropriate, by means of the one-way ANOVA, two-way ANOVA, or Kruskal-Wallis Test followed by the Tukey and Dunnett Test using the STATISTICA software program (version 12.0; StatSoft Inc., Tulsa, OK, USA). We considered *p* < 0.05 as a level of statistically significant difference.

3. Results

3.1. Compounds Isolated from *Equisetum giganteum* L. The constituents present in the hydroethanolic extract were identified by *R* in RP-UHPLC, UV-vis, and MS/MS spectra analysis and comparison with data from the literature [11]. In total, 13 constituents were identified in the 70% EtOH extract (Table 1). Figure 1 shows the structures of styrylpyrones and flavonoids identified. The UHPLC-PAD-ESI-MS analysis of the hydroethanolic extract highlighted UV spectrum characteristic of styrylpyrones with bands at 253–258 and 355–373 nm [31] and flavonoids at 240–290 nm related to A-ring and at 300–390 nm related to B-ring [32]. Diagnostics precursor ions at *m/z* 178 and 192 were related to presence of caffeic and ferulic acid derivatives. The neutral losses of 162 mass units allowed the identification of hexoses (sophorose or glucoside).

In Figure 2, the UPLC-PAD chromatogram of the *E. giganteum* 70% EtOH extract is presented.

3.2. Antimicrobial Activity of *E. giganteum*. The *E. giganteum* extract (E50, E25, E16, E8, and E4) showed antimicrobial activity against all strains studied (Figure 3). The strain most sensitive to the *E. giganteum* extract was *S. aureus.*
Table 1: UHPLC-PAD-ESI-MS data (UV-vis spectra and detected ions) and MS^n (product ions) of compounds detected in EtOH 70% of the aerial parts of *E. giganteum*.

| Peak | $R_t$ (min) | UV ($\lambda_{max}$) | UHPLC-MS ions [M-H]^− | ESI-IT-MS^n | Compound |
|------|-------------|----------------------|------------------------|-------------|----------|
| 1    | 9.99        | 266, 350             | 787                    | 463, 301    | Quercetin-tri-O-hexoside |
| 2    | 10.56       | 265, 344             | 771                    | 609, 428, 285 | Kaempferol-3-O-sophoroside-7-O-glu |
| 3    | 11.22       | 254, 366             | 585                    | 422, 379, 259 | 3-Hydroxyhispidin-3,4’-di-O-glucoside |
| 4    | 11.61       | 218, 295sh, 327      | 625.07                 | 463, 301    | Quercetin-3,7-di-O-glucoside |
| 5    | 11.69       | 278, 305sh, 327      | 625.01                 | 463, 301, 255, 178 | Quercetin-3-O-(caffeoyl)-glucoside |
| 6    | 12.08       | 264, 341             | 609                    | 447, 327, 285, 255 | Kaempferol-3,7-di-O-glucoside |
| 7    | 13.09       | 270, 361             | 625                    | 462, 301    | Flavonol-di-O-hexoside |
| 8    | 13.44       | 254, 331, 369        | 423                    | 379, 287, 261, 217 | Equisetumpyrone |
| 9    | 13.53       | 216, 291, 328        | —                      | 215, 178, 160, 142 | Caffeic acid derivative |
| 10   | 14.04       | 265, 287sh, 341      | 609                    | 285         | Kaempferol-3-O-sophoroside |
| 11   | 14.55       | 270, 347             | 463                    | 301         | Quercetin-hexoside |
| 12   | 14.82       | 275, 297sh, 331      | 463                    | 309, 192, 177, 133 | Ferulic acid derivative |
| 13   | 15.68       | 265, 344             | 447                    | 285         | Kaempferol-3-O-glucoside |

Figure 1: Structure of the flavonoids and styrylpyrones identified by UHPLC-PAD-ESI-MS^n in the hydroethanolic extract of *E. giganteum*.

At the highest concentration of the extract (50 mg/mL), the results were similar to 1% NaOCl for all strains tested. Even at its lowest concentrations, the extract still showed a microbicidal action.

3.3. Antibiofilm Effect of *E. giganteum* against *C. albicans* SC5314. The specimens pretreated with sterile phosphate buffered saline (CTRL/PBS) showed an expressive presence of biofilm mass of *C. albicans* on the acrylic resin specimen surface, thereby revealing biofilms at the stage of proliferation and filamentation (Figure 4(d)). As expected, this biofilm was mainly formed of viable (green) cells (Table 2). In contrast with PBS, 1% sodium hypochlorite pretreatments removed the majority of biofilm (Figure 4(e)), leaving only a few viable cells; but most of the cells were nonviable (Table 2).

The surface of resin specimens pretreated with the different concentrations of *E. giganteum* extract (50, 25, and 16 mg/mL) revealed that increasing the concentration of the extract improved its antiadherent activity, because biofilm formation decreased with an increase in amount of herbal extract used (Figures 4(a), 4(b), and 4(c)). As shown for 1% NaOCl, E50 significantly reduced the biofilm mass.
were performed in triplicate ($n = 15$ compared with the CTRL/Medium. Five independent experiments still be noted (Figure 4(a)). All groups of the on the specimen surfaces, although scarce viable cells can

![Figure 2: UHPLC-PAD analytical chromatogram of the 70% EtOH extract of the aerial parts of *E. giganteum* at $\lambda = 254$ nm. Mobile phase was water ultrapure (eluent A) and methanol (eluent B), both containing 0.1% of formic acid. The ratio was 5–100 of A in B in 20 min. Injection volume: 10.0 $\mu$L; Thermo Scientific column ($50 \times 2.1$ mm, $1.9 \mu$m); column temperature: $25^\circ$C; flow ratio: 0.4 mL-min$^{-1}$.](image)

on the specimen surfaces, although scarce viable cells can still be noted (Figure 4(a)). All groups of the *E. giganteum* extract presented lower numbers of remaining fungal cells in comparison with control (PBS), showing marked anti-adherent activity of the *E. giganteum* extracts against *C. albicans* biofilms. At 25 and 16 mg/mL, the values of biofilm mass were similar to those obtained for PBS; however, they differed statistically from both control (NaOCl) and E50. No statistical differences were observed between E50 and CTRL/NaOCl with respect to the total number of remaining nonviable cells.

**3.4. ROS Production in Monocytes Treated with *E. giganteum Extract***. The results revealed that all the extract concentrations (50, 25, 16, 8, and 4 mg/mL) were able to decrease the levels of ROS production by cells stimulated with LPS or *C. albicans*, returning to baseline levels. Values for cells treated with *E. giganteum* extract, which were not stimulated with LPS or *C. albicans*, remained at baseline levels (Figure 5).

**3.5. Cytotoxicity Test.** Both monocytes and HPEC remained viable in the presence of *E. giganteum* extract; but, at the lowest concentration (E4) for 12 hours on monocytes, there were significant differences between the cells treated with extract and untreated cells. In addition, at highest concentrations of *E. giganteum*, the percentage of cell viability was higher than it was for untreated cells, suggesting an increased cell metabolism (Figures 6(a) and 6(b)).

**4. Discussion**

*C. albicans* is the predominant yeast species isolated from patients with DS, due to its greater capacity to form biofilms on surfaces [33–36], including denture acrylic surfaces. Therefore, decontamination of dentures is a fundamental aspect of effective oral hygiene, in order to prevent the denture stomatitis. Typically, research on *Candida* biofilms has evaluated the efficacy of different disinfectant solutions such as 1 and 2% NaOCl and 2% glutaraldehyde on heat-polymerized acrylic resin [37].

Considering that various disinfectants affect the physical properties of resin surface of denture [37–44], alternative therapies, such as phytotherapeutic agents, with antimicrobial potential and ability to eliminate or prevent the adhesion of *C. albicans* to the inner surface of the denture, without damaging the mucosa or dentures, may be of extreme interest to removable denture wearers, particularly those with maxillary complete dentures [45, 46].

In the present study, the crude extract of *E. giganteum* showed a potent antimicrobial effect on *C. albicans* and on fungus adherence to heat-polymerized acrylic resin. Furthermore, the plant extract demonstrated a potential *in vitro* antibacterial effect on monocytes, while it had no cytotoxic effect *in vitro* on monocytes and HPEC. Our study is the first to verify the beneficial effect of *E. giganteum* against *Candida* biofilm formation on surfaces, and its biocompatibility.

Although *E. giganteum* does not have bacteriostatic potential, the antimicrobial action was evident in our findings, even at the lowest concentration tested (4 mg/mL). This

| Groups | Biofilm mass ($\mu$m$^3$) | (Viable + nonviable) |
|--------|--------------------------|---------------------|
| E50    | 4.263,03 ± 4.037,27       | (4.231,74 + 31,29)  |
| E25    | 31.408,34 ± 5.552,14      | (28.825,29 + 2.583,05) |
| E16    | 35.448,57 ± 4.950,06      | (34.063,22 + 1.385,35) |
| CTRL/PBS | 95.488,84 ± 40.389,17     | (94.060,39 + 1.428,45) |
| CTRL/NaOCl | 489,36 ± 244,11        | (386,17 + 103,19)  |

**Table 2: Mean ± S.D. of biofilm cells (biofilm mass), including those that are viable and nonviable (mean), remaining on the surface of the specimens after different pretreatments with extract: 50 mg (E50), 25 mg (E25), or 16 mg (E16), and PBS (CTRL/PBS) or 1% sodium hypochlorite (CTRL/NaOCl), for 10 minutes. The data were obtained by using the confocal laser scanning microscopy and Bioimage software program. Six independent experiments were performed in triplicate ($n = 18$).**
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Figure 4: Confocal microscopy images of Candida albicans biofilms after each treatment with extract. (a) E50; (b) E25; (c) E16; (d) CTRL/PBS; and (e) CTRL/NaOCl. Six independent experiments were performed in triplicate.

Figure 5: Production of ROS by human monocyte cells cultured and treated with LPS and C. albicans in the presence or absence of extracts as described in Materials and Methods. Results are expressed as mean ± S.D. of at least four experiments. Symbols indicate significant difference (p < 0.05): * in comparison with LPS-treated cells and δ compared with C. albicans-treated cells, in the presence of extract in all concentrations tested.

was shown not only against C. albicans, but also against the S. aureus and E. coli bacteria used as parameters for comparison in our study. At a concentration of 50 mg/mL, by means of antimicrobial assay based on the broth microdilution method, the herbal extract showed antimicrobial action similar to 1% sodium hypochlorite, an agent widely applied for immersion of dentures during treatment of DS.

We emphasize the values of antimicrobial action obtained in our study, which were higher than those reported in a previous study [12] that assessed the same herbal extract against Gram-negative and Gram-positive bacteria and obtained no activity against Escherichia coli (E. coli). In our study, another aspect that highlights the significant antimicrobial activity of E. giganteum is that this phytotherapeutic agent was effective against a clinical strain of E. coli (O1124), which usually presents more resistance and higher pathogenicity than standard strains when they are isolated from patients [47]. However, even at low concentrations of the extract (8 and 4 mg/mL), we observed that its antimicrobial activity against this strain of E. coli ranges from 65 to 54% of death.

The chemical composition of the extract EtOH 70% of E. giganteum has shown a clear presence of phenolic compounds derived from caffeic and ferulic acids and flavonoid heterosides derived from quercitín and kaempferol, in addition to styrylpyriones (Table 1; Figure 1). This herbal extract produces bioactive compounds such as flavonoids that are responsible for its anti-inflammatory and antioxidant effects, in addition to having analgesic, antimicrobial, and immunomodulatory properties [48, 49]. Therefore, the flavonoids present in the composition of E. giganteum probably are the constituents responsible for the microbicidal effects observed
three independent experiments; under treatments with different concentrations of resin surface pretreated with extract of contaminated specimens made of denture base material. The conditions of the intraoral environment, we developed significant difference compared with the control (p < 0.05). (b) The effects of E. giganteum on cell viability of HEPCs. The viability of cells under treatments with different concentrations of E. giganteum was investigated by MTT assays. Data shown are the mean ± S.D. of at least three independent experiments; * P < 0.05 represents a significant change compared with the control.

Figure 6: (a) Effects of E. giganteum on cell viability of human monocytes. The viability of cells under treatments with different concentrations of E. giganteum was investigated by MTT assays. Data shown are the mean ± S.D. of at least four independent experiments. Asterisks indicate significant difference compared with the control (p < 0.05). (b) The effects of E. giganteum on cell viability of HEPCs. The viability of cells under treatments with different concentrations of E. giganteum was investigated by MTT assays. Data shown are the mean ± S.D. of at least three independent experiments; * P < 0.05 represents a significant change compared with the control.

here, because they have the ability to inactivate microbial adhesion proteins and transport proteins and to rupture the microbial membrane.

Furthermore, in order to simulate the resinous portion of removable dentures that have been contaminated by C. albicans, as frequently it occurs with dentures under the conditions of the intraoral environment, we developed contaminated specimens made of denture base material. The resin surface pretreated with extract of E. giganteum at all concentrations showed a significant reduction in the biofilm mass of viable C. albicans cells in comparison with the untreated resin. In other words, we can assert that a brief 10-minute treatment of the resin surface with the extracts was able to significantly reduce the capacity of C. albicans to adhere to the surface. We emphasize that, at 50 mg/mL, the treatment with E. giganteum resulted in the lowest biofilm mass found, when compared with any of the other concentrations. Interestingly, after these pretreatments, the resin surface showed dispersed fungus in both its filamentous and yeast forms, especially at the highest concentration of the extract. As observed in our control samples, confluent biofilms presenting predominantly filamentous forms are developed on the resin surface. Taken together, we believe that the herbal extract was able to interfere in the expression of fungal virulence factors, in addition to having an antiahesive effect on the microorganism.

One possible hypothesis for explaining the effect of biofilm removal by E. giganteum extract could be based on biofilm pH. Since the most of biofilms are formed at neutral pH, the conditions having very high or very low pH values can provide changes on microbial metabolism and surface properties, interfering in their adhesion process to surfaces [50]. Hypochlorite solutions, for example, having alkaline pH (pH > 11), can increase the electrostatic repulsion between the yeast cells and the surface of the resin material, dissolving the biofilm cells [51]. Thus, the acidic pH (pH = 5.48) of the hydroalcoholic extract of E. giganteum, associated with antimicrobial activity, especially in higher concentrations, could explain their important antiahesive effect on C. albicans biofilms; however, further studies are required about the mechanisms involved.

In addition to the beneficial results with respect to the antimicrobial and antiahesive activities, all concentrations of E. giganteum extract demonstrated suppression of reactive oxygen species in human monocytes stimulated in vitro by LPS or C. albicans. Previous studies have also shown reduced levels of ROS resulting from the action of E. arvense and have attributed this effect to the presence of flavonoids [52]. In fact, flavonoids are able to eliminate ROS and RNS by donation of electron and hydrogen (H+) from the hydroxyl groups present in their composition, stabilizing these free radicals [49, 53] or suppressing the formation of ROS by inhibiting the enzymes involved in generating them [49, 54]. The presence of phenolic acids was also detected in the E. giganteum extract. This group of substances is characterized by their antioxidant properties, since they were able to reduce the levels of reactive oxygen species due to trapping free radicals directly or scavenging them through a series of combined reactions with antioxidant enzymes [49, 52, 53, 55]. Other inflammatory mediators should be evaluated for a more accurate conclusion and to imply that E. giganteum extract could negatively modulate monocyte-mediated inflammatory responses.

Another interesting finding was that the extract did not alter the viability of monocytes and HPEC for up to 24 hours. This has important clinical repercussions, since these cells are closely related to the inner surface of the denture, in denture wearers. The oral epithelial cells are the first cells to interact with C. albicans during the establishment of DS.

In summary, our results suggest that the phytotherapeutic E. giganteum extract has the following properties: antimicrobial against C. albicans, S. aureus, and E. coli; antiaherence
of C. albicans to heat-cured acrylic resin specimens; and anti-inflammatory effects on human monocytes activated by C. albicans. In addition, the phytherapeutic extract did not compromise the viability of human monocytes or human palatal epithelial cells. These properties might qualify E. giganteum extract to be a promising alternative for the topic treatment and prevention of oral candidiasis and denture stomatitis.

Conflict of Interests

The authors declare that they have no conflict of interests regarding the publication of this paper.

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