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FLT3-ITDs Instruct a Myeloid Differentiation and Transformation Bias in Lymphomyeloid Multipotent Progenitors

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SUMMARY

Whether signals mediated via growth factor receptors (GFRs) might influence lineage fate in multipotent progenitors (MPPs) is unclear. We explored this issue in a mouse knockin model of gain-of-function Flt3-ITD mutation because FLT3-ITDs are paradoxically restricted to acute myeloid leukemia even though Flt3 primarily promotes lymphoid development during normal hematopoiesis. When expressed in MPPs, Flt3-ITD collaborated with Runx1 mutation to induce high-penetrance aggressive leukemias that were exclusively of the myeloid phenotype. Flt3-ITDs preferentially expanded MPPs with reduced lymphoid and increased myeloid transcriptional priming while compromising early B and T lymphopoiesis. Flt3-ITD-induced myeloid lineage bias involved upregulation of the transcription factor Pu.1, which is a direct target gene of Stat3, an aberrantly activated target of Flt3-ITDs, further establishing how lineage bias can be inflicted on MPPs through aberrant GFR signaling. Collectively, these findings provide new insights into how oncogenic mutations might subvert the normal process of lineage commitment and dictate the phenotype of resulting malignancies.

INTRODUCTION

Whether signals mediated via growth factor receptors (GFRs) might influence lineage fate in normal multipotent progenitors (MPPs) and stem cells remains unclear and disputed. New insights into this process have recently been gained through studies in vitro (Rieger et al., 2009); however, whether GFR signaling instructs lineage specification in vivo remains a key unresolved issue (Enver and Jacobsen, 2009). Studying the impact of gain-of-function mutations is an alternative approach to determine whether and how GFRs might instruct lineage fate in vivo. Indeed, activating mutations of GFRs and downstream signaling pathways are common events in cancer, particularly in hematopoietic malignancies such as acute myeloid leukemia (AML), and often show striking associations with distinct clinical and cell-lineage phenotypes (Croce, 2008). In some cases, this might simply reflect the fact that such mutations primarily target a specific cell lineage. However, more intriguingly, if a mutation targets a primitive multipotent cell, it might instruct lineage-fate decisions. As recent investigations of human AML have suggested that the propagating cell might frequently represent the counterpart of normal MPPs (Goardon et al., 2011), it is possible but still unclear to what degree GFR signaling mutations might also influence lineage specification in multipotent cells.

A good example of aberrant GFR signaling associated with specific leukemia phenotypes relates to the FMS-like tyrosine kinase 3 receptor (FLT3), which is expressed in the majority of cases of AML and acute lymphoblastic leukemia (ALL) (Stirewalt and Radich, 2003). Constitutively activating internal tandem duplications (ITDs) within the juxtamembrane domain of FLT3 occur in ~25% of cases of AML, conferring an adverse prognosis (Stirewalt and Radich, 2003). However, despite the high frequency of FLT3 expression in ALL and the key role of Flt3 in early lymphoid development (Sitnicka et al., 2002), including high-level Flt3 expression in lymphoid-primed MPPs (LMPPs) with combined lymphoid and myeloid potential (Adolfsson et al., 2005), FLT3-ITDs are rare in cases of ALL, occurring in <1% in larger series (Leow et al., 2011).

As is the case for most leukemic mutations, it is unclear how often FLT3-ITDs are a true initiating event in myeloid
malignancies. Indeed, a number of lines of evidence support the notion that FLT3-ITDs are often acquired secondarily to an initiating clonogenic event (Gale et al., 2008; Jan et al., 2012), although when involved in chronic myelomonocytic leukemia (CMML), they may indeed be the initiating mutation (Lee et al., 2007). Nevertheless, in line with the normal expression pattern of Flt3 (Adolfsson et al., 2005), results from studies in patients are compatible with the notion that FLT3-ITDs occur within the human MPP compartment (Levis et al., 2005) and are an essential driver mutation within the founding leukemic clone (Ding et al., 2012; Smith et al., 2012). Thus, even though FLT3-ITDs might occur secondarily to an initiating clonogenic event in many cases, these mutations occur in multipotent cells (Levis et al., 2005) and FLT3-ITD GFR signaling appears to be an essential requirement for leukemia propagation (Smith et al., 2012). It is possible, therefore, that FLT3-ITDs may act to dictate the lineage fate and phenotype of the resulting leukemia. Compatible with a role of Flt3-ITD signal in lineage determination, two different knockin mouse models of Flt3-ITD have been reported to develop a myeloproliferative phenotype exclusively (Lee et al., 2007; Li et al., 2008). However, although it was recently suggested that Flt3-ITDs deplete hematopoietic stem cells (HSCs) (Chu et al., 2012), the key progenitor population that propagates FLT3-ITD-induced myeloid disease, as well as the cellular and molecular bases of their myeloid lineage bias, remains unclear.

Using a mouse knockin model of the Flt3-ITD mutation, we investigated the cellular and molecular mechanisms by which constitutive GFR signaling might subvert lineage specification in MPPs and alter the cell fate of early lymphoid progenitors, in order to explain the myeloid bias of the resulting leukemias.

RESULTS

**Flt3-ITD Collaborates with Runx1 Mutation to Induce Aggressive AML**

To definitively determine whether physiologically expressed Flt3-ITD impacts the establishment of myeloid versus lymphoid leukemia development, we crossed Flt3ITD/ITD with Runx1fl/fl (Growney et al., 2005) and Mx1-Cre mice to induce Runx1 deletion in MPPs. Importantly, Runx1 loss-of-function mutation is associated with both lymphoid and myeloid leukemia (Grossmann et al., 2011; Schnittger et al., 2011). Unexpectedly, even without poly I:C induction, Flt3ITD/ITDRunx1fl/flMx1-Creet mice developed a high-penetration, short-latency acute leukemia (Figure 1A) characterized by marked leukocytosis, anemia, and thrombocytopenia (Figures S1A–S1C) and hepatosplenomegaly. Peripheral blood (PB) and bone marrow (BM) morphology resembled AML in all diseased mice (Figures 1B–1D), and myeloid lineage was confirmed by flow cytometry (Figures 1E and S1D). Development of ALL was never observed. Leukemias showed deletion of the Runx1 DNA-binding domain (Figure S1E). The uniform myeloid-lineage leukemias in Flt3ITD/ITDRunx1fl/flMx1-Creet mice demonstrate a key role for Flt3-ITD signaling in introducing a myeloid-lineage bias during leukemogenesis.

**Flt3-ITDs Expand Myeloid-Biased LMPPs**

We next investigated the cellular and molecular bases for FLT3-ITD-induced myeloid bias. As shown previously (Lee et al., 2007), the multipotent Lin−Sca1+c-Kit+ (LSK) compartment was expanded in Flt3ITD/ITD mice (Figure 2A). We applied CD150 and CD48 to determine the nature of the expanded cells within the LSK compartment (Kiel et al., 2005), as Flt3-ITD is not detectable at the cell surface (Figure S2A). Notably, the expansion of LSKs was wholly attributable to a marked expansion of LSKCD150−CD48+ MPPs in 7- to 10-week-old Flt3ITD/ITD mice (Figures 2B and 2C), in line with the recent observation that Flt3-ITD may suppress the HSC compartment (Chu et al., 2012). Heterozygous (Flt3ITD+/) mice had an intermediate phenotype between Flt3ITD−/− and Flt3ITD/ITD mice (Figures S2B and S2C). High-level Flt3 expression in the normal LSK compartment defines LMPPs that lack self-renewal and megakaryocytic (Mk) and erythroid (E) potential but sustain lymphomyeloid capability (Adolfsson et al., 2005), a progenitor that is also implicated in human AML (Goardon et al., 2011). Because Flt3ITD+ LMPPs reside almost exclusively in the LSKCD150−48+ compartment (Figure S2D), we explored whether the expanded LSKCD150−48+ cells in Flt3-ITD mice had LMPP-like characteristics. As expected for LMPPs (Mansson et al., 2007), HSC- and MkE-affected gene expression was downregulated in Flt3ITD/ITD LSKCD150−48+ MPPs (Figures 2D, 2E, and S2E), and in line with the molecular data they possessed little or no Mk potential in vitro (Figure 2F). In contrast, myeloid-affected gene expression was upregulated in Flt3ITD/ITD LSKCD150−48+ cells (Figure 2G), paralleled by high granulocyte-macrophage (GM) potential in vitro (Figure 2H). Early lymphoid transcriptional programs were downregulated in both Flt3ITD/ITD and Flt3ITD+/– LSKCD150−48+ MPPs (Figures 2I and 2J), paralleled by severely reduced B cell potential in vitro (Figure 2K). Importantly, the lymphoid transcriptional program was already suppressed in Flt3ITD/ITD LSKCD150−48+ MPPs in embryonic day 15 (E15) fetal liver (FL) (Figure S2F), at which time the phenotype and number of LSKCD150−48+ MPPs were unaffected (Figure S2G). In keeping with this suppression of early lymphoid programs in fetal MPPs, the number of B220+CD19+ B cells was suppressed in Flt3-ITD E15 FL (Figure S2H), preceding the emergence of a myeloproliferative phenotype at this early stage of ontogeny (Figure S2I). This supports the notion that lymphoid suppression and myeloid bias in MPPs occurs as a cell-intrinsic and direct consequence of Flt3-ITD signaling.

Because FLT3-ITDs in humans often occur secondarily to an initiating clonogenic event (Jan et al., 2012), it is possible that in such cases it is the other genetic events and not the FLT3-ITD that influence the leukemic phenotype. In order to address this issue, we used Vav-Cre mediated recombination to examine the impact of Runx1 loss of function on lineage priming in MPPs in the absence of Flt3-ITD. The data obtained demonstrate a significant upregulation of Il7r and Rag1 expression in MPPs with no significant impact on slngf expression (Figure S2J). This finding is in keeping with the high incidence of lymphoid malignancies in mouse models of Runx1 mutation (Jacob et al., 2010; Kundu et al., 2005; Putz et al., 2006) and contrasts markedly with the suppression of lymphoid programs caused by Flt3-ITDs in MPPs. This supports the notion that it is GFR signaling through Flt3-ITD that instructs the uniform myeloid phenotype of leukemias resulting from the collaboration between Runx1 deletion and FLT3-ITD in LMPPs.
Figure 1. Mutation of Runx1 in Flt3ITD/ITD Mice Results in High-Penetration Aggressive Myeloid Leukemia

(A) Leukemia-free survival curves of Flt3ITD/ITDRunx1fl/fl mice stratified according to Mx1Cre genotype (n = 50–68 mice of each genotype).
(B and C) Low-power (20×; B) and high-power (100×; C) morphology of typical leukemic cells in Flt3ITD/ITDRunx1fl/flMx1Cre+ mice.
(D) Differential morphology results from BM of WT (+/+), Flt3ITD/ITDRunx1fl/flMx1Cre−, and leukemic Flt3ITD/ITDRunx1fl/flMx1Cre+ mice; percentage of total BM cells. Mean (SEM) values for 4–11 mice of each genotype.
(E) Fluorescence-activated cell sorting (FACS) analysis of PB and BM from an Flt3ITD/ITDRunx1fl/flMx1Cre+ mouse with myeloid leukemia (representative of 19 further analyzed cases). Error bars represent SEM. See also Figure S1.
Figure 2. Flt3-ITDs Expand Myeloid-Biased MPPs

(A) Expansion of LSK cells in 8- to 10-week-old Flt3<sup>ITD/ITD</sup> mice (+/+: n = 27; ITD/ITD: n = 23).
(B) Expression of CD48 and CD150 on BM LSK cells from Flt3<sup>+/+</sup> and Flt3<sup>ITD/ITD</sup> mice.
(C) Expansion of LSKCD48<sup>+</sup>CD150<sup>−</sup>C0 MPPs in 8- to 10-week-old Flt3<sup>ITD/ITD</sup> mice (+/+: n = 27; ITD/ITD: n = 23).
(D) GSEA demonstrating downregulation of E-affiliated genes in Flt3<sup>ITD/ITD</sup> versus Flt3<sup>+/+</sup>LSKCD48<sup>+</sup>CD150<sup>−</sup>C0 MPPs (n = 4 of each genotype). FDR, false discovery rate; NES, normalized enrichment score.

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Flt3-ITD Suppresses Early T- and B-Lymphoid Progenitors

Because Flt3-ITD compromised lymphoid-transcriptional priming in LMPPs, which have been implicated as critical thymus-seeding progenitors (Luc et al., 2012), we next investigated whether T lymphopoiesis might be suppressed in Flt3-ITD mice. Thymic cellularity was found to be progressively reduced, and the earliest thymic progenitors (double-negative 1 Kit+) were almost completely lost in Flt3ITD/ITD mice (Figures 3A–3C). Also, subsequent stages of DN2Kit+ and DN3 thymocyte progenitors were severely reduced (Figures S3A and S3B). Of relevance, gene and protein expression for the chemokine receptor Ccr9, which is critical for migration of thymus-seeding progenitors from the BM to the thymus (Schwarz et al., 2007), was suppressed in Flt3ITD/ITD LSKCD150+48+ MPPs (Figures 3D, S3C, and S3D). Moreover, DN1 cells showed large reductions in expression of Notch1 and its targets (Hes1 and Hes5; Figure S3E), which are critical for early T cell development. Although expression of some early lymphoid genes (IiL2ra, Ii7r, and Gata3) was maintained, myeloid genes (Cebpa, Sfpi1, Csf1r, Csf2r, and Csf3r) were highly upregulated in DN1 thymic progenitors (Figure S3E). In keeping with this myeloid bias, Flt3ITD/ITD DN1 progenitors showed low T cell potential in vitro (Figure 3E).

Previous studies demonstrated a reduction of mature B cells in Flt3-ITD mice (Lee et al., 2007; Li et al., 2008). We investigated whether this might reflect Flt3-ITD-induced perturbation at the earliest B cell commitment stages, and in support of this we found a notable 11-fold expansion of Lin−CD19−CD24−AA4.1−CD43−B220+ pre-pro-B cells in Flt3ITD/ITD mice (Figures 3F and 3G), whereas pro-B cells, the next stage in B cell development, were severely reduced (Figures 3H and S3F). Because pre-pro-B cells are known to sustain low-level myeloid potential (Rumfelt et al., 2006), we next investigated lymphoid and myeloid gene expression in Flt3ITD/ITD pre-pro-B cells. Key early lymphoid genes (Cd79a, Ii7r, slgh, Eb1f, Pax5, Rag1, and Rag2) were all suppressed (Figure 3I), whereas myeloid-affiliated genes (Cebpa, Cebpb, Sfpi1, Csf1r, and Csf2r) were upregulated in Flt3ITD/ITD pre-pro-B cells (Figure 3J). In agreement with this, Flt3ITD/ITD pre-pro-B cells had severely reduced B cell potential (Figure 3K). These findings demonstrate that Flt3-ITDs markedly impair the earliest stage of both T and B lymphopoiesis and upregulate myeloid gene expression in the earliest B and T lymphoid progenitors, in agreement with the notion that myeloid bias initiates in multipotent LMPPs.

Flt3-ITD-Induced Myeloid Bias Is Dependent on Upregulation of Pu1

To further explore the mechanistic basis of Flt3-ITD-induced myeloid bias of early lymphoid progenitors, we examined Pu1 protein expression using a Pu1-YFP reporter line. Whereas in wild-type (WT) mice Lin− cells showed a bimodal distribution between Pu1low− and Pu1high cells, almost all Lin− cells in Flt3ITD/ITD mice were Pu1high (Figure 4A). Flt3ITD/ITD mice showed a modest increase in Pu1 expression in LSKCD150+48+ MPPs (Figure 4B) and markedly enhanced levels of Pu1-YFP in pre-pro-B cells (Figure 4C). In keeping with the observed increased expression of Pu1, gene set enrichment analysis (GSEA) demonstrated upregulation of Pu1 target genes (Steidl et al., 2006) in Flt3-ITD LSKCD150+48+ MPPs (Figure 4D). Furthermore, GSEA also demonstrated a similar upregulation of Pu1 target genes in FLT3-ITD mutated “LMPP-like” leukemia stem cells in human AML (Goardon et al., 2011) in comparison with FLT3-WT counterparts (Figure S4A).

Because transition from pre-pro-B cells to subsequent stages of B cell development is associated with downregulation of Pu1 expression (Figure S4B), we attempted to rescue the suppressed B cell phenotype in Flt3-ITD mice through generation of Flt3ITD/+/ mice. Notably, whereas Pu1 haploinsufficiency did not increase the number of B cells or their progenitors on a WT background, it resulted in a 10.3-fold increase in B cells in Flt3ITD/+/ mice (Figure 4E). Furthermore, Pu1 haploinsufficiency led to a 2-fold reduction in the expanded pre-pro-B cell population in Flt3-ITD mice (Figure 4F) together with a 13.5-fold rescue of the suppression of pro-B cells (Figure 4G). Strikingly, in the presence of Pu1 haploinsufficiency, the expansion of MPPs was also restored to WT levels (Figure 4H) together with a significant amelioration of the myeloproliferative phenotype in Flt3ITD/ITD mice (Figure 4I).

Stat3 is aberrantly activated by Flt3-ITD signaling (Schmidt-Arras et al., 2009) and Pu1 is a direct target gene of Stat3 (Hegde et al., 2009). In line with this, Flt3ITD/ITD BM showed aberrant activation of STAT3 (Figure S4C), and MPPs and pre-pro-B cells showed significantly increased expression of Stat3 target genes (Figures 4J–4L). Collectively, these data suggest that Pu1 upregulation is a key mediator of the Flt3-ITD-induced myeloid bias and progenitor phenotype.

DISCUSSION

In the studies presented here, we explored whether and how GFR signaling might influence lineage specification in vivo. To...
that end, we used a knockin model of Flt3-ITD to investigate whether oncogenic mutations that result in constitutive GFR signaling influence the lineage fate of MPPs in vivo.

In order to first establish the physiological relevance of Flt3-ITD as an AML-inducing mutation, we developed a mouse model in which Flt3-ITD was coexpressed with an inducible deletion of the DNA-binding domain of Runx1 (GIEWS, 2005) in MPPs. Importantly, although Runx1 loss-of-function mutations are found in patients with both ALL and AML, only the latter is often found in association with FLT3-ITDs (GROSSMANN et al., 2011; SCHNITTGER et al., 2011). Deletion of the DNA-binding domain of Runx1 during adult murine hematopoiesis results in mild myeloproliferation (GIEWS, 2005) or lymphoblastic leukemias/lymphomas (JACOB et al., 2010; KUNDU et al., 2005; PUTZ et al., 2006). Here, we demonstrate that the combination of Runx1 mutation with Flt3-ITDs results in aggressive leukemia with 100% penetrance and a uniform myeloid phenotype. In addition to demonstrating the functional relevance for FLT3-ITD-induced AML, this AML model has a number of important features. First, in contrast to many other mutation collaboration models, both mutations are targeted to the physiologically relevant endogenous genetic loci. Second, in isolation each mutation results in only a modest phenotype, but in combination a minor clone of deleted cells rapidly expands and becomes clonally dominant, paralleling somatic mutations during leukemogenesis. Third, the evolution of leukemia is rapid, supporting the notion that it occurs without the requirement for additional genetic events. Finally, the model recapitulates a particularly poor prognostic form of AML with a major unmet need for novel therapeutic approaches and thus provides a powerful model for future studies of the cellular and molecular mechanistic bases for collaboration of these mutations in AML.

The most primitive progenitor population that was expanded by Flt3-ITDs consisted of LMPP-like cells, demonstrating upregulation of the myeloid program, whereas the transcriptional expression of lymphoid-affiliated genes was markedly reduced in both heterozygous and homozygous Flt3-ITD mice. This myeloid transcriptional bias and suppression of lymphoid transcripts was also present during early B and T cell development and, importantly, was already present in Flt3-ITD FL MPPs, preceding the expansion of MPPs and development of a myeloproliferative phenotype. It is important to note that metastasis are essential for Flt3-ITD signaling and is occurring at the earliest B and T lymphoid progenitors.

**Figure 3. Early T and B Cell Progenitors Are Suppressed and Myeloid Biased in Flt3ITD/ITD Mice**

(A) Thymic cellularity of Flt3+/+ and Flt3ITD/ITD mice at 2–3 and 8–10 weeks (n = 7–10 mice of each genotype at each age).

(B) Representative DN staining of Flt3+/+ and Flt3ITD/ITD thymocytes from 2-week-old mice; percentages are relative to total thymocytes.

(C) Progressive reduction of DN1Kit+ cells in Flt3ITD/ITD thymus.

(D) Expression (SEM) of Ccr9 in LSKCD48+ 48+ BM cells from 8- to 10-week-old Flt3+/+ and Flt3ITD/ITD mice (n = 3–4 mice per genotype, 2–3 separately sorted replicates per mouse).

(E) T cell potential on OP9DL1 cells of DN1 cells (10/cells per well) from 2- to 3-week-old Flt3+/+ and Flt3ITD/ITD mice (n = 2 mice per genotype, 12 replicate wells per mouse). ND, not detected.

(F) Representative staining of early B cell progenitors in Flt3+/+ and Flt3ITD/ITD 2- to 3-week-old mice. pre-pro-B cells are identified as Lin- B220+CD19+ CD24+ AA4.1+CD43+; percentage of total BM cells.

(G) Increased numbers of pre-pro-B cells in 2- to 3-week-old Flt3ITD/ITD mice (mean [SEM] values for 2- to 3-week-old mice, n = 8–10 of each genotype).

(H) Reduced numbers of pro-B cells (Lin- B220+CD43+CD19+CD24AAD4.1+) in Flt3ITD/ITD mice.

(I) Mean (SEM) expression of lymphoid-affiliated genes in pre-pro-B cells from 2- to 3-week-old mice (n = 2 mice per genotype, 3 separately sorted replicates per mouse).

(J) Mean (SEM) expression of myeloid-affiliated genes in pre-pro-B cells from Flt3ITD/ITD mice.

(K) In vitro B cell potential (B220+CD19+ cells) of pre-pro-B cells from 2- to 3-week-old mice (n = 10 cells/well, 2 mice per genotype, 10 replicate wells per mouse).

Error bars represent SEM. See also Figure S3.
(legend on next page)
this is consistent with a model in which LMPPs, which normally express high levels of FLT3 (Adolfsson et al., 2005), acquire FLT3-ITD mutations that confer both a strong clonal advantage and a marked myeloid bias. This results in myeloid expansion and suppression of early lymphoid development, strongly supporting a fundamental role for FLT3-ITDs in promoting myeloid lineage leukemia development at the MPP level. Our study also further highlights LMPPs as a key target population in AML, as also supported by recent findings in human AML (Gordon et al., 2011). Transformation to an aggressive leukemia exclusively of a myeloid phenotype by introduction of Runx1 mutation demonstrates the functional relevance of this FLT3-ITD-induced myeloid bias and clonal dominance, providing insights into the process by which oncogenic mutations might determine the lineage fate of the resulting leukemias at the precommitment stage. Our findings are also of considerable relevance for normal hematopoiesis, as it remains disputed whether key cytokine receptor signaling pathways mediate critical in vivo functions in blood lineage development through purely permissive rather than instructive actions (Enver and Jacobsen, 2009). Thus, although Flt3-ITD elicits aberrant signaling, our findings clearly provide support for the notion that cytokine receptors are also capable of eliciting lineage-instructive signaling in MPPs in vivo.

**EXPERIMENTAL PROCEDURES**

**Animals**

All animals used were bred and maintained in accordance with regulations of the UK Home Office. Details regarding the mouse lines are provided in the Extended Experimental Procedures.

**Patient Samples**

BM samples from AML patients were obtained with informed consent and the approval of Oxford Ethics Committee B (protocols 06/Q1606/110 and 05/MRE077/74).

**Fluorescence-Activated Cell Sorting**

Details of the antibody staining panels and protocols are provided in the Extended Experimental Procedures. Antigens and antibodies used for identification of specific cell populations are shown in Tables S1 and S2.

**Western Blotting**

Cells were lysed in 1% NP40 buffer and analyzed by western blot as previously described (Pecquet et al., 2010) with antibodies against pStat3 (Cell Signaling) and beta-actin (Sigma).

**In Vitro Evaluation of Lineage Potentials**

Evaluation of megakaryocytic, GM, and lymphoid potentials was carried out as previously described (Månsson et al., 2007) and detailed in the Extended Experimental Procedures.

**Gene Expression by Dynamic Arrays**

Multiplex quantitative PCR was performed as previously described (Kharazi et al., 2011) and detailed in the Extended Experimental Procedures. Assays used for dynamic arrays are shown in Table S3.

**Microarray Analysis**

Analysis by Affymetrix Mouse Genome 430 2.0 Arrays was carried out at the Stanford Protein and Nucleic Acid Facility as described previously (Månsson et al., 2007) and in the Extended Experimental Procedures.

**ACCESSION NUMBERS**

The GEO accession number for the microarray data reported in this paper is GSE35805.

**SUPPLEMENTAL INFORMATION**

Supplemental Information includes Extended Experimental Procedures, four figures, and three tables and can be found with this article online at http://dx.doi.org/10.1016/j.celrep.2013.04.031.

**LICENSING INFORMATION**

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