CASE REPORT

Exacerbation of mycosis fungoides leading to the diagnosis of chronic myelomonocytic leukemia

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INTRODUCTION

Patients with mycosis fungoides (MF) are at a greater risk (relative risk, 1.73; range, 1.32-2.4) to have secondary malignancies than healthy subjects, and co-occurrence of cutaneous T-cell lymphoma with other hematoproliferative diseases has been reported.1,2 Chronic myelomonocytic leukemia (CMML) is a rare myeloproliferative disease with features of myelodysplastic syndrome and myeloproliferative disorders leading to monocytosis in the peripheral blood.3 Cutaneous manifestations of CMML have been reported and seem to be associated with a poor prognosis and more likely progression to acute myeloid leukemia.4-6 Specific infiltrates in myeloproliferative disorders may clinically and histologically be difficult to distinguish from neutrophilic dermatoses, and neoplastic cells might accompany inflammatory or infectious dermatoses, as reported for B-cell chronic lymphocytic leukemia.7-11 We describe a case of CMML coinciding with MF presenting with Sweet-like infiltrates and neoplastic cells in MF plaques.

CASE REPORT

A 65-year old, otherwise healthy patient was first seen in our department in 2005 with suberythroderma and histologically confirmed diagnosis of MF. Treatment with systemic psoralen ultraviolet A (PUVA) initially led to a complete remission of disease symptoms over several years. Initial staging did not show any signs of visceral or blood involvement. In 2014, the patient experienced a relapse with erythematous plaques and patches predominantly on the trunk. Systemic PUVA was reintroduced, however, the skin condition worsened with erythematous patches affecting 80% body surface area and severe pruritus. In addition, the patient subsequently had multiple abscess-like lesions positive for methicillin-resistant staphylococcus aureus (Fig 1, A).

A first biopsy taken from an abscess found a dense superficial and deep perivascular and interstitial infiltrate of neutrophils and poorly differentiated myelomonocytic cells (myeloperoxidase [MPO]1+, elastase1+, CD51+, CD681+; Fig 1, B). Biopsies from erythematous skin lesions showed features typical of MF with a band-like and partly epidermotropic infiltrate of atypical T lymphocytes (CD4+, CD3+, CD8−; Fig 2). T-cell receptor rearrangement was negative in skin specimens, peripheral blood, and bone marrow.

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Abbreviations used:

AML: acute myeloid leukemia
CMML: chronic myelomonocytic leukemia
MF: mycosis fungoides
MPO: myeloperoxidase
PUVA: psoralen ultraviolet A

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Differential blood count showed a leukocytosis (36,000/μL) with monocytosis (11,000/μL), a microcytic and hypochromic anemia, thrombopenia (88,000/μL), elevated lactate dehydrogenase level (793 U/L) and C-reactive protein (8.8 mg/dL).

Repeated bone marrow biopsies and smears found a myelomonocytic infiltrate expressing CD33, lysozyme, and myeloperoxidase compatible with CMML.

Further workup, including abdominal and lymph node ultrasound scan, showed splenomegaly of 13.8 cm and palpable axillary and inguinal lymphadenopathies. We initiated an antibiotic treatment and high-dose oral corticosteroids (1 mg/kg). This treatment led to a transient regression of erythematous lesions, relapsing on corticosteroid dose tapering to less than 0.5 mg/kg. Treatment with

**Fig 1.** A, example of an abscess-like lesion on the patient's arm. B, Histologic picture of an abscess-like lesion with abundant superficial and deep interstitial and perivascular infiltrate of neutrophilic granulocytes with signs of leucocytoclasia. Additionally, larger MPO-positive myelomonocytic cells can be found.

**Fig 2.** Subepidermal infiltrate of CD3+ cells with hyperchromatic, atypical nuclei, and abundant epidermotropism. Single myelomonocytic cells around the deep dermal vessels.
low-dose methotrexate (15 mg weekly) had to be stopped after 1 injection because of dyspnea, vertigo, and radiologic signs of pneumonitis.

Monocytosis persisted over more than 6 months, and repeated bone marrow biopsies found blast infiltrates of up to 20%. The diagnosis of CMML was made, and an association to the exacerbation of MF had to be assumed. Consequently, a treatment with hydroxicarbamide, 500 mg/d, was started. Owing to lack of response, the treatment was changed to azacitidine, 75 mg/m². The patient responded well to the treatment with improvement of skin manifestations and pruritus (Fig 5). However, pruriginous erythematous patches and plaques remained, and a parallel treatment with bexarotene, 300 mg/m², was initiated.

The administration of azacitidine was reduced to 5 instead of 7 days in a 28-day cycle after the first 6 treatment cycles because of thrombocytopenia (33,000/μL). Under this combined treatment, the skin condition improved dramatically, lymphadenopathy was regressing, and blood count improved with near normalization of thrombocyte count and differential blood count. Neither CMML nor MF worsened. Bexarotene was continued at a dose of 300 mg/m². One year after initiation of the combination therapy, the patient had an increasing leukocytosis with up to 80% of blasts and progression to acute myeloid leukemia (AML). Treatment was changed from azacitidine to cytarabine, 20 mg, days 1 to 10 every 4 weeks. Treatment with bexarotene was continued. The patient died of leukemic disease 4 months after diagnosis of AML.

**DISCUSSION**

Various hypotheses on the pathogenesis of secondary hemoproliferative diseases exist; identical stem cell clones, immunosuppression, mutagenic therapies, and specific cytokine milieus may be possible causes. With regard to ultraviolet light treatment, such as PUVA irradiation, for cutaneous T-cell lymphoma, no correlation with hemoproliferative diseases has been shown. Changes in microenvironment and increased immunosuppression may also be explanations for the worsening of MF in our case. Interestingly, we could observe neoplastic myeloid cell infiltrates in MF plaques. One may speculate on a direct interaction between CMML and MF neoplastic cells or a locally altered immune response as potential triggers of MF exacerbation. Also, a “pseudo” MF could be discussed as reported in B-cell chronic lymphocytic leukemia. It has been described as lesions clinically and histologically indistinguishable from MF, but without evidence of a positive T-cell receptor rearrangement. Yet, the long duration from diagnosis of MF to diagnosis of CMML and the typical histologic picture are in favor of a true cutaneous lymphoma in our case.

In general, differentiation between specific infiltrates and reactive infiltrates containing neoplastic cells may be impossible. We observed Sweet-like infiltrates and abscess formation probably caused by skin infection triggered by immunosuppression in the context of CMML and MF. Here too, a non-negligible part of the infiltrate may have consisted of neoplastic CMML cells.

Treating 2 different neoplasms at a time may be challenging; azacitidine is a hypomethylating agent approved for the treatment of CMML with a white blood count of less than 13 G/L. In highly replicative CMML it seems to be less efficient, which is why our patient first received a cytoreductive agent (hydroxicarbamide) and azacitidine as second-line treatment.

Adding bexarotene meant combining 2 drugs with each specific and dose-dependent side effects and potential unknown interactions. Fortunately,
except for thrombocytopenia, no other side effects led to interruption or tapering of medication dose. The clinical picture could be stabilized under combination therapy over 12 months before progression to AML.

This case shows how 2 hemoproliferative diseases may influence one another and lead to therapeutic challenges that call for individual solutions. It also shows that neoplastic cells in CMML may be found not only in inflammatory or infectious dermatoses but also in cutaneous lymphoma plaques.

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