Annotated mitochondrial genome with Nanopore R9 signal for *Nippostrongylus brasiliensis* [version 1; referees: 1 approved, 2 approved with reservations]

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**Abstract**

*Nippostrongylus brasiliensis*, a nematode parasite of rodents, has a parasitic life cycle that is an extremely useful model for the study of human hookworm infection, particularly in regards to the induced immune response. The current reference genome for this parasite is highly fragmented with minimal annotation, but new advances in long-read sequencing suggest that a more complete and annotated assembly should be an achievable goal. We de-novo assembled a single contig mitochondrial genome from *N. brasiliensis* using MinION R9 nanopore data. The assembly was error-corrected using existing Illumina HiSeq reads, and annotated in full (i.e. gene boundary definitions without substantial gaps) by comparing with annotated genomes from similar parasite relatives. The mitochondrial genome has also been annotated with a preliminary electrical consensus sequence, using raw signal data generated from a Nanopore R9 flow cell.

This article is included in the Nanopore Analysis gateway.
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**Introduction**

_Nippostrongylus brasiliensis_ is a parasitic nematode that naturally infects rodents. Its life cycle and morphology is comparable to *Necator americanus* and _Ancylostoma duodenale_, and it is thus an excellent murine model of human hookworm infection, a disease that affects approximately 700 million people worldwide. Like its human counterparts, _N. brasiliensis_ L3 larvae infect the host through the skin and migrate to the lungs where they feed on red blood cells (unpublished study; Haem metabolism is a check-point in blood-feeding nematode development and resulting host anaemia; Bouchery T, Filbey K, Shepherd A, Chandler J, Patel D, Schmidt A, Camberis A, Peignier A, Smith AAT, Johnston K, Painter G, Pearson M, Giacomin P, Loukas A, Bottazzi M-E, Hotez P, Le Gros G), causing extensive haemorrhage and anaemia – both hallmarks of hookworm infections. The larvae are coughed up and swallowed to enter the gastrointestinal tract. The nematode matures into a sexually active adult in the small intestine where it secretes eggs that enter the environment via the hosts’ faeces. Larvae hatch, undergo two molts to become infective L3 larvae, which propagates the lifecycle. The immunology of _N. brasiliensis_ infection has been studied extensively, and the parasite has been utilised as an inducer of potent Th2 responses in the lung and intestine, yielding important discoveries into cellular and molecular immune responses. The _N. brasiliensis_ model allows delination of hookworm-induced immune profiles that could be targeted in drug or vaccine design, and provides a simple and well-characterised murine model in which to test these interventions for efficacy. To underpin these studies, a high-quality reference genome is needed.

**Current reference genome**

The most recent NCBI reference genome sequence for _N. brasiliensis_ is a draft generated from Illumina HiSeq reads as part of the Wellcome Trust Sanger Institute (WTSI) 50 Helminth Genomes initiative. It is 294.4 Mbp in total length, and highly fragmented (29,375 scaffolds with an N50 length of 33.5kb, and a longest scaffold of under 400kb). The _N. brasiliensis_ reference genome would benefit from improvement, a goal that may be readily achieved with the advent of affordable long-read sequencing technologies.

**MinION sequencing**

The Oxford Nanopore Technologies’ (ONT) MinION platform is improving at a rapid pace, with improvements in flow cell chemistry and base calling software announced frequently. In 2015, the median accuracy of double-stranded MinION reads, using R7.3 sequencing pores, was about 89% pores, sequenced at 60 bases per second with a yield of about 200 Mb. The quality and length of sequences generated from R7.3 pores was sufficient to create a single-contig assembly of the _Escherichia coli_ K-12 MG1655 chromosome using nanopore reads alone, with consensus accuracy of 99.5%. An equimolar sample of _Mus musculus_, _E. coli_ and _Enterobacteriophage lambda_ DNA was sequenced in September 2016 on the International Space Station using R7.3 flow cells, producing approximately equal read counts for the different samples with a median accuracy of 83–92% for 2D reads across four runs.

The recent introduction of R9 sequencing pores in June 2016, together with improved software for base-calling the generated signal trace at 250 bases per second, has improved the median accuracy of high-quality double-stranded reads to 95%, and yield to 800 Mb (personal communication, September 2016; MinION Analysis and Reference Consortium). Consensus accuracy for an _E. coli_ K12 assembly consequently also increased to 99.96%. A rapid single-stranded sequencing kit was introduced in August 2016, reducing post-extraction sample preparation time to less than 15 minutes.

The R9.4 flow cell was commercially released by ONT a few months later in October 2016. This release brought together software and chemistry improvements that increased run flow cell yield into the gigabase range, and increased sequencing speed to 450 bases per second. Additional use cases for the MinION are evident with this increased yield: the R9.4 flow cells have already been used for sequencing human genomes using multiple flow cells, with observed yields of about 1–4Gb for each individual sequencing run.

**Scientific justification**

The mitochondrial genome is useful for epidemiology and population genetic analysis in nematodes, as it is rapidly evolving. An average cell has 100–1000 mtDNA molecules, compared to two nuclear DNA molecules, and this stoichiometric excess facilitates analyses, especially where starting materials are limited. The strict maternal inheritance of the mitochondrial chromosome, coupled with a general lack of recombination in this haploid replicon permits inference of maternal lineages. The ONT MinION can be deployed in infectious disease outbreak scenarios, and a “read until” methodology promises to make rapid, specific identification of known infectious agents possible. The technology has obvious utility in other areas of epidemiology and infection surveillance, and to enhance these applications it will be useful to develop the “read until” methodology to be able to detect a wider range of infectious agents from metagenomic sequencing. To do this, electronic signatures representing the MinION nanopore event signals could be used as a reference library to pre-screen raw signals from the pores before base calling. Here we present a complete mitochondrial genome for _N. brasiliensis_, assess its quality by gene prediction and phylogenetic analyses, and provide a validated electronic signal trace for the sequence.

This annotation represents the first hurdle in generating a complete genomic sequence for this model organism and provides crucial information for evolutionary and immunological studies. The rapid advancement of molecular technologies, such as qPCR, RNAseq, nanostring and high-throughput sequencing, has given researchers the capacity to acquire an expansive array of new knowledge and insight into how genetic pathways function and interact at a molecular level. However, the lack of a complete annotated reference genome for _N. brasiliensis_ thus far has restricted the full exploration into this important helminth.

**Methods and results**

Genomic DNA was extracted from adult _N. brasiliensis_ and sequenced on a MinION R9 flow cell. Reads from this sequencing run were then assembled, and the highest-coverage contig (mitochondrial DNA) was error-corrected and circularised for further analysis.
DNA extraction and library preparation

*N. brasiliensis* was originally sourced from Lindsey Dent of the University of Adelaide, South Australia and has been maintained for 22 years by serial passage at the Malaghan Institute. Female Lewis rats were bred and used for the maintenance of the *N. brasiliensis* life cycle at 4 months of age (weight over 150g; housed in IVC caging and given *ad libitum* access to food and water). For the purposes of this study, one rat was infected with 4000 infective larvae. After 7 days, to allow the worms to mature to the adult stage in the small intestine, the rat was euthanized, and the small intestine dissected and flushed with PBS to harvest worms, as outlined in Camberis *et al.*. Ethics approval for the maintenance of the *N. brasiliensis* life cycle is overseen and approved by the Victoria University of Wellington Animal Ethics Committee.

The harvested *N. brasiliensis* were washed in PBS by centrifugation to remove cellular debris. The nematodes were frozen at -80°C bead-beaten, and DNA extracted using Qiagen DNeasy Blood and Tissue DNA extraction kit, yielding approximately 4µg of high molecular weight double-stranded DNA (detected by the Quantus QuantiFluor dsDNA System). This DNA was treated with RNase. Two sequencing libraries were made using the Oxford Nanopore 2D genomic DNA sequencing kit, yielding in total about 70ng of adapter-ligated sequencing library. No effort was made to specifically isolate mitochondrial DNA. The first preparation was loaded onto an R9 MinION flow cell and sequenced for 6 hours, and the second preparation was loaded onto the same flow cell and sequenced for an additional 36 hours. Pore occupancy at 30 minutes into the first run was about 25%, while pore occupancy at 30 minutes into the second run was about 80%.

Whole-genome assembly with Canu

All FASTQ sequences (i.e. both 1D and 2D reads) were extracted from the base-called FAST5 files. These sequences were fed into Canu v1.3 to generate assembled contigs. The contig with the highest coverage was a 19907 bp sequence with similarity to other nematode mitochondrial genomes (see Supplementary File 1). This sequence had 98% identity to an unannotated *N. brasiliensis* contig in the Wellcome Trust Sanger Institute (WTSI) *N. brasiliensis* assembly.

Error correction and circularisation

Reads generated by WTSI (SRA ID: ERR063640) were mapped as pairs to the MinION mitochondrial contig using Bowtie2 in local mode. At each location, one read was randomly sampled from those that mapped to that location, representing a reference-based digital normalisation to approximately 100X coverage (see Supplementary File 2). The differences between these normalised reads and the MinION contig were evaluated using a custom script, producing a corrected sequence based on the consensus read alignments. The mapping and correction process was repeated with BWA-MEM on the corrected sequence (see Supplementary File 3) to identify additional variants that were missed by Bowtie2, due to multiple matches to duplicated regions.

Repeated sections of the linear contig (representing duplicated regions of the circular sequence) were merged to generate a circular consensus sequence, and the resultant sequence adjusted (by shifting sequence from the end to the start of the circular genome) so that the first base in the genome was set to the beginning of the COX1 gene (following the convention of OGRe, see http://drake.physics.mcmaster.ca/ogre). A final round of error correction was carried out on the circularised genome using Bowtie2-aligned reads from ERR063640 (see Supplementary File 4), producing a final mitochondrial genome length of 13,355 bp. The original 19 kbp contig thus contained about 6 kbp of duplicated sequence. MinION reads were mapped to the assembled genome to identify variants not present in the WTSI reads.

Comparison of WTSI and MIMR *N. brasiliensis* strains

After remapping the original R9 MinION reads back to the assembled and corrected genome with GraphMap, four locations were found with variant calls that contributed to more than 50% of the read coverage. Three of these variants involved transition mutations: \( T \rightarrow C \) at 5742, \( G \rightarrow A \) at 6102, and \( T \rightarrow C \) at 11460. One additional complementarity mutation was found: \( T \rightarrow A \) at 2860 (see Figure 1).

Mitochondrial genome annotation

Approximate gene boundaries were determined by a local NCBI BLASTx search, mapping the contig to mitochondrial protein sequences from *Necator americanus* (see Table 1; Supplementary File 5 and Supplementary File 6). Regions between genes were
### Table 1. mtDNA gene regions. Predicted gene features from the *Nippostrongylus brasiliensis* mitochondrial genome. Stop codons that end in hyphens (−) are completed by the addition of polyA sequence.

| Start | End  | Name       | Start Codon | Stop Codon |
|-------|------|------------|-------------|------------|
| 1     | 1575 | COX1       | ATT         | TAG        |
| 1820  | 2514 | COX2       | TTG         | TA−       |
| 2571  | 3522 | l-rRNA     | —           | —         |
| 3523  | 3857 | ND3        | ATA         | TAA       |
| 3858  | 5438 | ND5        | ATT         | TTA       |
| 5498  | 5578 | AT-rich    | —           | —         |
| 5689  | 6123 | ND6        | ATA         | TAA       |
| 6124  | 6356 | ND4L       | ATT         | TA−       |
| 6474  | 7223 | s-rRNA     | —           | —         |
| 7339  | 8206 | ND1        | ATT         | T−−       |
| 8207  | 8806 | ATP6       | ATT         | TAA       |
| 9003  | 9842 | ND2        | ATA         | TAA       |
| 10076 | 11186| CYTB       | ATA         | T−−       |
| 11242 | 12007| COX3       | ATA         | T−−       |
| 12062 | 13291| ND4        | GTC         | TAA       |

### Table 2. mtDNA tRNA sites. Predicted tRNA sites in the *Nippostrongylus brasiliensis* mitochondrial genome. One truncated tRNA site between the ND4 and COX1 genes (detected by cmscan) could not be fully annotated.

| Start | End  | Amino Acid | Codon |
|-------|------|------------|-------|
| 1589  | 1638 | Cys        | GCA   |
| 1649  | 1705 | Met        | CAU   |
| 1706  | 1760 | Asp        | GUC   |
| 1764  | 1819 | Gly        | UCC   |
| 2517  | 2570 | His        | GUG   |
| 5439  | 5497 | Ala        | UGC   |
| 5579  | 5633 | Pro        | UGG   |
| 5634  | 5688 | Val        | UAC   |
| 6356  | 6411 | Trp        | UCA   |
| 6417  | 6473 | Glu        | UUC   |
| 7224  | 7278 | Asn        | GUU   |
| 7279  | 7338 | Tyr        | GUA   |
| 8818  | 8880 | Lys        | UUU   |
| 8890  | 8944 | Leu        | UAA   |
| 8944  | 8997 | Ser        | UCU   |
| 9843  | 9901 | Ile        | GAU   |
| 9902  | 9959 | Arg        | ACG   |
| 9959  | 10013| Gin        | UUG   |
| 10022 | 10075| Phe        | GAA   |
| 11187 | 11241| Leu        | UAG   |
| 12008 | 12057| Thr        | UGU   |
| 13322 | 13355| —          | AUU   |

then scanned using Infernal cmscan\(^{29}\) to identify exact tRNA gene boundaries and codon sequences (see Table 2). The amino acid associated with each tRNA was identified using BWA-MEM to map annotated tRNA sequences from *Oesophagostomum columbianum*, *N. americanus*, *Strongylus vulgaris*, and *A. duodenale*. One tRNA region found by cmscan (between the ND4 and COX1 genes) could not be matched to any existing tRNA sequences. When this sequence was fed into RNAstructure\(^{30}\), the predicted secondary structure had no T-loop or D-loop, and an anticodon loop of 8 bases (Figure 2). The anticodon for this structure pairs with one of the two most common gene start codons (i.e. ATT), and could potentially pair with the other most common start codon through a wobble A-A pairing on the third base (see 31).

Precise gene start boundaries were determined by mapping open reading frames (ORFs) between the tRNA genes (codon translation table 5: Invertebrate Mitochondria with NCBI SmartBLAST (https://blast.ncbi.nlm.nih.gov/smartblast/smartBlast.cgi?CMD=Web). Stop boundaries were determined by looking for plausible in-frame stop sequences surrounding the end region of matching SmartBLAST hits. The boundaries for the ribosomal RNA genes were determined by a BLAST search against the four previously compared parasite species. Finally, the AT-rich region was identified as the region between tRNA-Ala and tRNA-Pro.

### Phylogenetic analyses

We identified orthologues of cytochrome oxidase 1 (COX1), cytochrome B (CytB), and the large ribosomal RNA subunit (l-rRNA) in other rhabditid nematodes using BLAST, and collated a dataset

![Figure 2. Predicted truncated tRNA structure.](attachment:RNAstructure.png)
from 49 taxa. Nucleotide sequences were aligned using clustalo\textsuperscript{32}, trimmed with trimAL, and phylogenies estimated using RAxML using the GTRGAMMA model. Bootstrap values were calculated from 100 iterations. Figures were generated using FigTree v1.4.2 (http://tree.bio.ed.ac.uk/software/figtree/). The *Nippostrongylus brasiliensis* sequences were placed within Strongylomorpha, as expected, and *N. brasiliensis* was found to be sister to *Heligmososoides polygyrus*, a finding in keeping with morphological systematics. Many internal nodes have very low bootstrap values, suggesting either low or conflicting signal in the data. Some groups were well supported, but these tend to be within rather than between genera. Overall the tree conforms to the classical morphological and global molecular phylogenies of the suborder, but cannot stand as indicators of those relationships independently (Figure 3).

Park and colleagues\textsuperscript{33} used whole mitochondrial genomes (i.e. all 12 protein coding loci) to develop a phylogeny of Nematoda, with the goal of analysing the placement of some unusual mitochondria from Ascaridia species, but including many strongyles. Our analyses are largely congruent with theirs, albeit with lower support (as noted above).

![Figure 3. Phylogenetic tree for mtDNA.](image)
Read mapping
The template and complement raw signal from the MinION reads mapped by GraphMap\textsuperscript{15} were extracted from the FAST5 files, and sorted into four groups:

1. Template sequence, mapped to coding strand
2. Template sequence, mapped to non-coding strand
3. Complement sequence, mapped to coding strand
4. Complement sequence, mapped to non-coding strand

A summary of mapping counts can be found in Table 3. Reads where the template fragment mapped to the non-coding strand were about two-thirds that of coding strand-mapped reads, with a similar proportion of reads distributed between the template and complement read fragments.

Event mapping
Event information (generated by the ONT cloud base caller Metrichor dragonet, version 1.22.4) was extracted for these sorted reads, and per-group median event currents were calculated for each pentamer found in the reference mitochondrial genome. An ideal signal trace of the mitochondrial genome was generated using these statistics for the four different signal groups (see Figure 4; Supplementary File 7).

Median complement events mapped to coding strand pentamers had a slightly higher event current when compared to template events (median difference = 3.94 pA, 90% range: 1.2 ~ 6.7, MAD = 1.53), and were lower in events mapped to non-coding pentamers (median difference = −2.08 pA, 90% range: −5.7 ~ 1.6, MAD = 2.93).

The median signal level for pentamers found in the \textit{N. brasiliensis} mitochondrial genome has a very strong positive correlation between read direction for the coding strand ($r = 0.982$, 90% range: 0.980 ~ 0.984) and the non-coding strand ($r = 0.974$, 90% range: 0.972 ~ 0.978), whereas there is weaker negative correlation between strands for the template direction ($r = −0.67$, 90% range: −0.70 ~ −0.63) and the complement direction ($r = −0.66$, 90% range: −0.69 ~ −0.62).

| Direction | Strand   | Count | Mean Length |
|-----------|----------|-------|-------------|
| Template  | Coding   | 26    | 5.0 kbp     |
| Complement| Non-coding| 25    | 4.8 kbp     |
| Template  | Non-coding| 17    | 5.3 kbp     |
| Complement| Coding   | 16    | 5.1 kbp     |

Figure 4. Ideal event plot, CytB gene tail. Ideal event trace for 200 pentamers at the tail end of the Cytochrome B gene. The complement sequence has a slightly lower current than the template sequence for reads mapped to the coding strand, and also a slightly lower current for reads mapped to the non-coding strand.
Raw signal mapping
Raw signal traces from both template and complement strands were converted to pA using scaling metadata in the FAST5 files, mapped to the GraphMap-aligned reference base positions using event metadata, and linearly interpolated to 11 samples per base using the R `approx` function (R version 3.3.1). Median signal traces (at a sub-base resolution) were generated by summarising the mapped signal at each interpolated location (Figure 5; Supplementary File 8).

The event data signal for template sequence mapped to the coding strand was loosely correlated with median raw signals in the middle of the interpolated region \( (r = 0.52, 90\% \text{ range: } 0.51 \sim 0.53) \), with other read groups demonstrating lower correlations \( (r = 0.29 \sim 0.44) \). This correlation disappeared when shifting the compared signal by one base in either direction \( (r = 0.03 \sim 0.09) \).

Discussion
Using a long-read assembler, and three passes of error correction with publicly-available data, we have created a full-length, error-free, de novo assembly of the mitochondrial genome of *N. brasiliensis*. This genome has been annotated with gene and rRNA boundaries, and compared with other related parasite species. An additional preliminary “electrical” annotation was generated from mapped nanopore read sequences.

Mitochondrial genome assembly
Low-cost long-read sequencing has made possible full-length assemblies of a number of different megabase-length genomes from nanopore data alone (e.g. 11.33–35), so it is not surprising that a full-length mitochondrial assembly was also possible using nanopore reads. The vast wealth of publicly-available data allows fast and low cost assembly, correction, and annotation of genomes, producing high-quality reference sequences that are of great benefit to medical research.

We were able to assemble the *N. brasiliensis* mitochondrial genome from a whole-genome sequencing nanopore dataset, by identifying assembly contigs with high relative coverage. The assembly is of high quality, based on read coverage, mapping of Illumina short reads, and annotation. The gene order is identical to that of *Caenorhabditis elegans* and other strongylophorm nematodes (see 36). Despite this shared structure, there is sufficient variation in sequences between species to generate resolved phylogenies\(^3\).

WTXI assembly of mtDNA
During the final preparation of this paper for publication, the WTSI deposited an annotated mitochondrial genome for *N. brasiliensis* (accession id: AP017690.1). This complements the introduction of the WormBase ParaSite resource for helminth genomics\(^9\). While the associated reference for the WTSI *N. brasiliensis* mitochondrial genome is not yet published, it is expected that this mitochondrial genome was assembled using a similar method to the WTSI’s previous work\(^3\) (i.e. a reference-based iterative mapping procedure using MITObim\(^3\)).

The sequence of this assembly differs only in an additional T insertion into a 10 base poly-T tract in the rRNA gene. While such polynucleotide tracts are problematic for MinION, the polyT region appears to be polymorphic, with some support for both variants in the WTSI reads (ERR063640). In addition the WTSI annotation excludes the AT-rich region.

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**Figure 5. Raw signal plot, CytB gene head.** Raw signal plot for 100 bases at the start of the Cytochrome B gene for template read directions (top) and complement read directions (bottom). Median raw signal current is shown as a thick red line, with individual raw signal observations shown in grey. Ideal event current for the observed pentamers is shown as black circles.
MinION whole genome sequencing data from a metazoan can be used for taxon identification

At the time of sequencing, no mitochondrial genome for *N. brasiliensis* was available. We thus explored the utility of the MinION data in species identification. As the mitochondrial genome is at a higher molarity than the nuclear genome, low-coverage sequencing of a target genome can yield deep coverage of the mitochondrial. Assembly of this replicon, and then analysis in a phylogenetic context was successful in placing *N. brasiliensis* in the Strongylomorpha. We suggest that this approach would be a useful technology for identification of unknown specimens in clinical practice, biosurveillance or biodiversity research programmes. In addition the nanopore electronic signal of the mitochondrial sequence could be used in a “read until” approach
diagnosis, using live monitoring to identify reads that likely derive from this, or a very similar genome. Usually, identification through sequencing is applied to amplification of specific target loci in a specimen or sample, an approach known as DNA barcoding. Direct sequencing of the whole genome of a specimen on MinION would allow both barcoding and produce additional sequences that could be used for, for example, population genetic diversity analysis.

Nanopore read analyses

Nanopore reads were separated into four different read groups to provide information that could be used to establish whether or not there are different sequencing features associated with template and complement strands. In general, the coding and non-coding strands had similar electrical profiles, as demonstrated by the event data (e.g. see Figure 4).

As this investigation is the first attempt to categorise the electrical properties of a complete mitochondrial genome, errors in the data analysis (e.g. due to incorrect mapping, low read coverage, and incorrect scaling parameters) cannot be excluded as an explanation for the difference in current that were observed between event data and raw signal. A comparison of raw signal current to the ideal current suggests that the pentamer model is probably sufficient to fully describe variation in signal in the mitochondrial genome. Although correlation between the signal and the ideal pentamer model is low for all four sequencing groups (template coding, template non-coding, complement coding, complement non-coding), this variation could be explained by errors in the raw signal mapping process, and other alternative mapping techniques (e.g. nanoraw
d may give better performance for linking raw signal to sequenced bases.

It is possible that the observed difference between the raw and ideal event signal may be due to methylation and other epigenetic modification of the mitochondrial genome. Methylation is a known feature of mitochondrial DNA (see 41), and methylation patterns can be observed as changes in the nanopore electrical signal
due to the lack of information about epigenetic patterns from de novo nanopore sequencing, this dataset is provided without additional epigenetic analysis as a source of discovery for other researchers.

Conclusions

The data presented here have been created from a minimally-prepared whole-genome DNA from *N. brasiliensis*, combining nanopore reads with publicly-available datasets. Using non-targeted sequencing, we have been able to generate a fully-annotated (gap-free) mitochondrial genome, with an initial electrical signal annotation having a resolution that is finer than a single base. The analysis proves that the efficiently MinION-generated mitochondrial genome of *N. brasiliensis* is of high enough quality for phylogenetic use.

We hope that the procedures discussed here will be sufficient to guide other researchers in annotating mitochondrial genomes and generating consensus signal traces, and that these data will contribute more generally towards improving the sequence base calling algorithms in the future for devices that implement sequencing by observation.

Data availability

Sequences have been deposited into NCBI Genbank, with accession number KY347017. Reads used to produce this assembly are associated with BioProject PRJNA328296. The assembly was error corrected using Illumina reads from a Wellcome Trust Sanger Institute sequencing run (ERR063640).

The *mpileup2proportion.pl* custom script that was used for error-correcting nanopore reads using Bowtie2-mapped short reads, as well as for generating count data for the read coverage plot, is available from David Eccles’ github repository (DOI, 10.5281/zenodo.164193)
. Read mapping group statistics were generated using the *fastx-grep.pl* and *fastx-length.pl* scripts also from this repository. These scripts have also been included here as a supplementary file (Supplementary File 9).

Author contributions

JC Preparation & extraction of DNA, phylogenetic analyses, manuscript preparation
MC Maintenance & propagation of *N. brasiliensis* larvae
TB *N. brasiliensis* sequencing project conception
MB Interpretation of phylogenetic results
GLG Project design & oversight
DAE Project design, MinION sequencing, assembly, data analysis, manuscript preparation

All authors have read the paper and provided edit suggestions where appropriate.

Competing interests

The present project has been fully-funded, but we have in the past received complimentary deliveries of flow cells and sequencing reagents from Oxford Nanopore Technologies, as part of the MinION Access Program.

The authors declare that there are no other competing interests.

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Supplementary files

Supplementary File 1 Original single-contig assembly generated by Canu v1.3. Click here to access the data.

Supplementary File 2 ERR063640 reads mapped to Canu-assembled genome, digitally normalised to one read per base. Click here to access the data.

Supplementary File 3 ERR063640 reads mapped to the first error corrected-genome, digitally normalised to one read per base. Click here to access the data.

Supplementary File 4 ERR063640 reads mapped to the second error corrected-genome, digitally normalised to one read per base. Click here to access the data.

Supplementary File 5 BED-format file of discovered mtDNA features (prior to correction of boundaries following protein translation). Click here to access the data.

Supplementary File 6 FASTA file containing subsequences of the mitochondrial genome representing discovered features. Click here to access the data.

Supplementary File 7 Event-level data aggregated for each base in the mitochondrial genome, including ideal current derived from pentamer signals. Click here to access the data.

Supplementary File 8 Data file containing interpolated raw signal-level data. Click here to access the data.

Supplementary File 9 Compressed file containing all Perl and R scripts used for data processing and analysis. Click here to access the data.

References

1. Hotez PJ, Bethony JM, Diemert DJ, et al.: Developing vaccines to combat hookworm infection and intestinal schistosomiasis. *Nat Rev Microbiol.* 2010; 8(11): 814–826. [PubMed Abstract] [Publisher Full Text]

2. Camberis M, Le Gros G, Urban J Jr: Animal model of *Nippostrongylus brasiliensis* and *Heligmosomoides polygyrus*. *Curr Protoc Immunol.* 2003; Chapter 19: Unit 19.12. [PubMed Abstract] [Publisher Full Text]

3. Bouchery T, Kyle R, Camberis M, et al.: ILC2s and T cells cooperate to ensure maintenance of M2 macrophages for lung immunity against hookworms. *Nat Commun.* 2015; 6: 6970. [PubMed Abstract] [Publisher Full Text]

4. Ohnmacht C, Schwartz C, Panzer M, et al.: Basophils orchestrate chronic allergic dermatitis and protective immunity against helminths. *Immunity.* 2010; 33(3): 364–374. [PubMed Abstract] [Publisher Full Text]

5. Chen F, Wu W, Millman A, et al.: Neutrophils prime a long-lived effector macrophage phenotype that mediates accelerated helminth expulsion. *Nat Immuno.* 2014; 15(10): 938–946. [PubMed Abstract] [Publisher Full Text] [Free Full Text]

6. Nellig DR, Wong SH, Belfosi A, et al.: Nuocytes represent a new innate effector leukocyte that mediates type-2 immunity. *Nature.* 2010; 464(7293): 1367–1370. [PubMed Abstract] [Publisher Full Text] [Free Full Text]

7. Holroyd N, Sanchez-Flores A: Producing parasitic helminth reference and draft genomes at the wellcome trust sanger institute. *Parasite Immunol.* 2012; 34(2-3): 100–107. [PubMed Abstract] [Publisher Full Text]

8. Wellcome Trust Sanger Institute: *Nippostrongylus brasiliensis* genome sequencing. *NCBI BioProject.* 2014. [Reference Source]

9. Howe KL, Bolt BJ, Shafie M, et al.: WormBase ParaSite - a comprehensive resource for helminth genomics. *Mol Biochem Parasitol.* 2016; pii: S0166-6851(16)30160-8. [PubMed Abstract] [Publisher Full Text]

10. Ip CL, Loose M, Tyson JRL, et al.: MiniON Analysis and Reference Consortium: Phase 1 data release and analysis [version 1; referees: 2 approved]. *F1000Res.*
11. Loman NJ, Quick J, Simpson JT: A complete bacterial genome assembled de novo using only nanopore sequencing data. Nat Methods. 2015; 12(8): 733–5.
Published Abstract | Publisher Full Text | Free Full Text

12. Castro-Wallace SL, Chu CY, John KK, et al.: Nanopore dna sequencing and genome assembly on the international space station. bioRxiv. 2016.
Published Full Text

13. JSCNASA and nScience: NASA AMA: We just sequenced DNA in space for the first time...ask us anything! The Winnower. 2016.
Published Full Text

14. Brown C: Inside the skunkwork. In: London Calling. Oxford Nanopore Technologies. 2016.
Reference Source

15. Simpson J: Supporting R9 data in nanopolish. Simpson Lab Blog. 2016.
Reference Source

16. Edwards A, Debonnaire AR, Sattler B, et al.: Extreme metagenomics using nanopore dna sequencing: a field report from svalbard, 78 n.
Reference Source

17. Brown C: Cliveome onth1 data release. Github repository. 2016.
Reference Source

18. Akeson M, Beggs AD, Nieto T, et al.: Na12878: Data and analysis for na12878 genome on nanopore. Github repository. 2016.
Reference Source

19. Brown WM, George M Jr, Wilson AC: Rapid evolution of animal mitochondrial DNA. Proc Natl Acad Sci U S A. 1979; 76(4): 1967–1971.
Published Abstract | Publisher Full Text | Free Full Text

20. Martin SA: Mitochondrial DNA repair. In: DNA Repair – On the Pathways to Fixing DNA Damage and Error. InTech, 2011.
Published Full Text

21. Pakendorf B, Stoneking M: Mitochondrial DNA and human evolution. Annu Rev Genomics Hum Genet. 2005; 6: 105–183.
Published Abstract | Publisher Full Text | Free Full Text

22. Cann RL, Stoneking M, Wilson AC: Mitochondrial DNA and human evolution. Nature. 1987; 329(6099): 31–36.
Published Abstract | Publisher Full Text | Free Full Text

23. Harrison RG: Animal mitochondrial DNA as a genetic marker in population and evolutionary biology. Trends Ecol Evol. 1989; 4(1): 6–11.
Published Abstract | Publisher Full Text | Free Full Text

24. Berlin K, Koren S, Chin CS, et al.: Assembling large genomes with single-molecule sequencing and locality-sensitive hashing. Nat Biotechnol. 2015; 33(6): 623–630.
Published Abstract | Publisher Full Text | Free Full Text

25. Langmead B, Salzberg SL: Fast gapped-read alignment with Bowtie 2. Nat Methods. 2012; 9(4): 357–359.
Published Abstract | Publisher Full Text | Free Full Text

26. Li H: Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. arXiv. 1303.3997v2: 2013.
Reference Source

27. Jameson D, Gibson AP, Huddet C, et al.: Ogre: a relational database for comparative analysis of mitochondrial genomes. Nucleic Acids Res. 2003; 31(1): 202–206.
Published Abstract | Publisher Full Text | Free Full Text

28. Sović I, Šikić M, Wilm A, et al.: Fast and sensitive mapping of nanopore sequencing reads with GraphMap. Nat Commun. 2016; 7: 11307.
PubMed Abstract | Publisher Full Text | Free Full Text

29. Navrotsky EP, Kolbe DL, Eddy SR: Infernal 1.0: inference of RNA alignments. Bioinformatics. 2000; 16(10): 1236–1237.
Published Abstract | Publisher Full Text | Free Full Text

30. Reuter JS, Mathews DH: RNAstructure: software for RNA secondary structure prediction and analysis. BMC Bioinformatics. 2010; 11(1): 129.
Published Abstract | Publisher Full Text | Free Full Text

31. Murphy FV 4th, Ramakrishnan V: Structure of a purine-purine wobble base pair in the decoding center of the ribosome. Nat Struct Mol Biol. 2004; 11(12): 1251–1252.
Published Abstract | Publisher Full Text

32. Park JK, Sultana T, Lee SH, et al.: Monophyly of clade III nematodes is not supported by phylogenetic analysis of complete mitochondrial genome sequences. BMC Genomics. 2011; 12(1): 392.
Published Abstract | Publisher Full Text | Free Full Text

33. Rose J, Thomson M, Patrick S, et al.: A single chromosome assembly of Bacteroides fragilis strain BE1 from Illumina and MinION nanopore sequencing data. Gigascience. 2015; 4(1): 60.
Published Abstract | Publisher Full Text | Free Full Text

34. Istance B, Friedrich A, d’Agata L, et al.: de novo assembly and population genomic survey of natural yeast isolates with the oxford nanopore minion sequencer. bioRxiv. 2016; 066613.
Published Full Text

35. Davis AM, Ioinville M, James S, et al.: Using minion nanopore sequencing to generate a de novo eukaryotic draft genome: preliminary physiological and genomic description of the extremophilic red alga galderia sulphuraria strain sag 107.7b. bioRxiv. 2016.
Published Full Text

36. Hu M, Gasser RB: Mitochondrial genomes of parasitic nematodes–progress and perspectives. Trends Parasitol. 2006; 22(2): 78–84.
Published Abstract | Publisher Full Text | Free Full Text

37. Hunt VL, Tsai IJ, Coghan A, et al.: The genomic basis of parasitism in the Strongyloides clad of nematodes. Nat Genet. 2016; 48(3): 299–307.
Published Abstract | Publisher Full Text | Free Full Text

38. Hahn C, Bachmann L, Chevreux B: Reconstructing mitochondrial genomes directly from genomic next-generation sequencing reads—a baiting and iterative mapping approach. Nucleic Acids Res. 2013; 41(13): e129.
Published Abstract | Publisher Full Text | Free Full Text

39. Loose M, Mallia S, Stout M: Real-time selective sequencing using nanopore technology. Nat Methods. 2016; 13(9): 751–4.
Published Abstract | Publisher Full Text | Free Full Text

40. Stober MH, Quick J, Egan R, et al.: De novo identification of dna modifications enabled by genome-guided nanopore signal processing. bioRxiv. 2016.
Published Full Text

41. Ghosh S, Singh KK, Sengupta S, et al.: Mitoeugenetics: the different shades of grey. Mitochondrion. 2015; 25: 60–66.
PubMed Abstract | Publisher Full Text

42. Simpson JT, Workman R, Zuzarte PC, et al.: Detecting DNA methylation using the oxford nanopore technologies MinION sequencer. bioRxiv. 2016; 047142.
Published Full Text

43. Eccles D: Bioinformatics scripts: Initial citable release. Zenodo. 2016.
Data Source
Open Peer Review

Current Referee Status: ✔️ ✔️ ✔️

Version 1

Referee Report 12 April 2017
doi:10.5256/f1000research.11363.r21379

Jianbin Wang
Department of Biochemistry and Molecular Genetics, University of Colorado School of Medicine, Aurora, CO, USA

In this manuscript, Chandler et al described in detail how they used the Nanopore sequencing technology to assemble the mitochondrial genome of Nippostrongylus brasiliensis. They also annotated the mitochondrial genome and did phylogenetic analysis among a selected group of nematodes. In addition, they characterized the Nanopore sequencing features for this genome. Overall, the authors have demonstrated that they can produce the complete mitochondrial genome from their Nanopore sequencing dataset.

The authors were able to recover the mitochondrial genome from a genomic DNA library due to the much higher copy number (often hundreds or thousands of times) of the mitochondrial DNA when compared to the nuclear genome. This approach has also been extensively used to recover mitochondrial and chloroplast genomes in whole genome shotgun libraries. In principle, it should work for any type of sequencing technology. The Nanopore sequencing technology is relatively new and is still fast evolving. In this case the technology does not seem to me to have a clear advantage over the Illumina or other sequencing approaches on mitochondrial genome assembly. In additional, the authors eventually used the Illumina data to do the error correction to make the final assembly. Nevertheless, the authors have presented a complete genome assembled from a combination of Nanopore and Illumina data with a full description of how they did this.

Not considering the novelty or significance of the work, I think the mitochondrial genome is properly assembled and annotated. The results are clear and the manuscript is well written.

Competing Interests: No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 12 Apr 2017

David Eccles, Malaghan Institute of Medical Research, New Zealand

Thank you very much for your review of our paper on mitochondrial genome sequencing with the MinION sequencer. We are currently working on updating the paper as per the reports of Christian Rödelsperger and Matthias Bernt, and intend to deliver a full response to them at that time.
This paper was intended as a stepping stone for investigating techniques that could be used to assemble a parasite genome from unamplified genomic DNA using the MinION. We discovered that the run yield in this case was not sufficient for assembling the entire *N. brasiliensis* genome, but being able to assemble a mitochondrial genome as a single contig has given us confidence that the technology is capable of improving on the existing Illumina-derived whole genome assembly. We did not intend to wow the world with this paper, rather it was an attempt to demonstrate methods and show how easy and quick it can now be to assemble a genome. Thank you for understanding this aspect of our paper.

At the time of sequencing, the base-calling software was not sufficiently accurate to generate a reliable sequence at a single base level. Understanding this, we used MinION reads for scaffolding, and Illumina reads (from a different strain) to correct the abundant base call errors. This approach has allowed a relatively cheap and fast assembly of the mitochondrial genome, such that comprehensive phylogenetic analyses can be carried out on the mitochondrial genes.

As you have mentioned, the nanopore sequencing technology is evolving fast. It is likely the case that updated base-calling software has improved base calling accuracy sufficiently that this approach can be carried out using MinION reads alone. I would like to carry out additional investigations on these data to discover if that is indeed the case, but would rather hold off on that until after we have published our attempts at whole genome assembly. Regardless, the mitochondrial sequences (including raw signal) are available for anyone else to determine themselves whether or not a high-quality MinION-only assembly is possible using re-called (but otherwise identical) nanopore sequence data.

**Competing Interests:** No competing interests were disclosed.
The Introduction basically describes the lifecycle of N. brasiliensis and the mode of infection. The authors might consider writing a more general introduction about nematodes, parasites, .. that it is important to study these parasites to develop treatments. In addition, there are multiple related parasites that are later part of the phylogenetic analysis. It would be good to give some information about those ones as well. e.g. what are their hosts?

**Section: Current reference genome**
Please provide the Genbank entry for the NCBI reference genome or provide the assembly that has been used for this study as supplemental data. Otherwise, it will be hard to reproduce the results.

**Section: Scientific justification**
please explain what a "read until" methodology is and provide some reference for the use of ONT MinION in studies of infectious disease outbreak.

Is the N. brasiliensis isolate that was used for sequencing have a strain ID? If yes, please specify and at least register a biosample for it and give the accession number. Was it the same isolate that was used for the NCBI reference genome.

**Section: Whole-genome assembly with Canu**
How much sequencing data was obtained? Please provide some more details about the assembly results. How many Contigs, total size.

For readers, that would like to use Nanopore technology to sequence their genomes it would be interesting to compare the quality of of the mitochondrial genome with nuclear contigs. I guess, that the lower coverage of nuclear contigs should also result in higher number mismatches with regard to the reference genome. A major finding of the paper could be that based on current nanopore technology, it only makes sense to do the multicopy mitochondrial genome. Such a statement could help people to plan their projects.

**Section: Error correction and circularisation**
How many sites had to be corrected. Error correction only makes sense, if the WTSI data is from the same isolate. Please clarify if this is the case. If it is the same isolate, there does the 2% mismatches come from in the "Whole-genome assembly with Canu" section.

**Section: Mitochondrial genome annotation**
"The amino acid associated with each tRNA was identified using BWA-MEM to map annotated tRNA sequences from Oesophagostomum columbianum, N. americanus, Strongylus vulgaris, and A. duodenale." Using BWA-mem to annotate tRNAs from other species sounds unusual. Do you have a reference where the performance of this methodology has ever been evaluated?

**Section: Phylogenetic analyses**
Please provide more information about the alignment, how many sites? amount of missing data? Please provide references for what is called "the classical morphological and global molecular phylogenies"

**Section: Read mapping/ Event mapping /Raw signal mapping**  (Table 3, Fig 4 and 5)
These sections seem to examine very specific aspects of the Nanopore sequencing technology and do not add any additional insights for the presented mitochondrial genome. I also have problems in understanding what kind of questions are asked. It seems to me, as if the authors try to examine whether Nanopore data has a preference for the template or complement strand or whether there is a bias for
coding or non-coding sequences. How well the sequencing signal corresponds to the basecalls in the final assembly and what features correlate with variation in sequencing signals. The presented results are not conclusive (no statistical tests have been done to assess the significance of the results) and are not really related to the rest of the manuscript. I would recommend to use this and other comparable data for a separate more methodological paper. One additional feature that could be tested would be how differences raw and ideal event current, sequencing coverage depend on GC content.

Minor comments

Section: DNA extraction and library preparation
The first paragraph should probably labeled as "Worm culturing" or something else. It has nothing to do with DNA extraction or library preparation

Section: MinION sequencing
"sequenced at 60 bases per second with a yield of about 200 Mb" does that mean per sequencing run?

This section sounds a bit like a promotion of MinION sequencing. I would recommend to reduce it only to the parts that are relevant for the current paper.

high through-put sequencing -> high-throughput

I wonder why the title has to have the information that R9 signal has been used. Probably most readers have heard about Nanopore sequencing but do not have a clue what R9 signal is. I would recommend to put this detailed information into the methods section but remove it from the title.

Is the work clearly and accurately presented and does it cite the current literature?
Yes

Is the study design appropriate and is the work technically sound?
Yes

Are sufficient details of methods and analysis provided to allow replication by others?
No

If applicable, is the statistical analysis and its interpretation appropriate?
Partly

Are all the source data underlying the results available to ensure full reproducibility?
Partly

Are the conclusions drawn adequately supported by the results?
Yes

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.
Author Response 12 Apr 2017

David Eccles, Malaghan Institute of Medical Research, New Zealand

Thank you very much for your review of our paper on mitochondrial genome sequencing with the MinION sequencer. We are currently working on updating the paper as per your report (and the report of Matthias Bernt), and will deliver a full response once the next revision of the paper is ready.

Competing Interests: No competing interests were disclosed.

Referee Report 20 February 2017

doi:10.5256/f1000research.11363.r19519

Matthias Bernt
Department of Computer Science, Leipzig University, Leipzig, Germany

The paper describes the sequencing of the mitochondrial genome of the *Nippostrongylus brasiliensis* with the novel Nanopore sequencing technique. To the best of my knowledge this seems to be one of the first mitogenomes that have been sequenced with this technology. The annotation of the genome and its use for phylogeny and taxonomic identification have been discussed.

Another group has sequenced the genome (including the mitogenome) has been sequenced using another NGS strategy. While this seem unfortunate its actually good for this study otherwise no reference data would have been available for comparison and error correction. I'm missing an analysis of the error rates of the sequencing without the correction that has used the read data from the other study. I'm wondering if the combination of data from MiniON sequencing and short read sequencing strategies might be a good general strategy?

The paper is well written and needs only a few corrections and additions. Details are given below.

Abstract:
=========

The term "electrical consensus sequence" might be puzzling for uninformed readers.

Introduction:
=============

"L3" is also difficult to understand for non experts. Maybe add 'stage'?

"highquality" missing space

MiniON sequencing
================

"R7.3" Can you explain what this means?
“89% pores” is unclear to me.

What are “2D reads”?

**Scientific justification**

"strict maternal inheritance": nothing in biology is strict. Check for paternal leakage or doubly-uniparental inheritance.

The term "read until" methodology is unclear.

**DNA extraction and library preparation**

Explain the abbreviation PBS

**Error correction and circularisation**

It needs to be explained what the custom script is doing.

“Repeated sections of the linear contig were merged... “ What happens with true repeats?

Since not all readers might know the color chartreuse I would suggest to order the colors as in the legend.

**Mitochondrial genome annotation**

I’m wondering why automatic methods for genome annotation have been ignored. Not saying that the applied approach is wrong.

When you use cmscan you need to state the used model as well.

“tRNA... codon sequences” Do you mean anticodon?

How about non-canonical start codons? How do you define “plausible” in frame stop?

For the truncated tRNAs there are examples known for Enoplea: see http://dx.doi.org/10.4161/rna.21630 and http://dx.doi.org/10.1016/j.biochi.2013.07.034

**Phylogenetic Analyses**

References for RAxML and trimAL are missing.

**Event Mapping**
"Event information" Specify what an event is.

"per-group" and later on "signal groups" You should reformulate this. Currently its a bit confusing.

Why pentamer?

What is an ideal signal trace?

Raw Signal Mapping

Has Graph Map been referenced?

Discussion

Are you really sure that the sequence is "error free"? In the end of the paper you write that its of "high enough quality...".

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

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