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Mobile-Genetic-Element-Encoded Hypertolerance to Copper Protects Staphylococcus aureus from Killing by Host Phagocytes

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ABSTRACT Pathogens are exposed to toxic levels of copper during infection, and copper tolerance may be a general virulence mechanism used by bacteria to resist host defenses. In support of this, inactivation of copper exporter genes has been found to reduce the virulence of bacterial pathogens in vivo. Here we investigate the role of copper hypertolerance in methicillin-resistant Staphylococcus aureus (MRSA). We show that a copper hypertolerance operon (copB-mco), carried on a mobile genetic element (MGE), is prevalent in a collection of invasive S. aureus strains and more widely among clonal complex 22, 30, and 398 strains. The copB and mco genes encode a copper efflux pump and a multicopper oxidase, respectively. Isogenic mutants lacking copB or mco had impaired growth in subinhibitory concentrations of copper. Transfer of a copB-mco-carrying plasmid to a naive clinical isolate resulted in a gain of copper hypertolerance and enhanced bacterial survival inside primed macrophages. The copB and mco genes were upregulated within infected macrophages, and their expression was dependent on the copper-sensitive operon repressor CsoR. Isogenic copB and mco mutants were impaired in their ability to persist intracellularly in macrophages and were less resistant to phagocytic killing in human blood than the parent strain. The importance of copper-regulated genes in resistance to phagocytic killing was further elaborated using mutants expressing a copper-insensitive variant of CsoR. Our findings suggest that the gain of mobile genetic elements carrying copper hypertolerance genes contributes to the evolution of virulent strains of S. aureus that are better equipped to resist killing by host immune cells.

IMPORTANCE Methicillin-resistant Staphylococcus aureus (MRSA) poses a substantial threat to human health worldwide and evolves rapidly by acquiring mobile genetic elements, such as plasmids. Here we investigate how the copB-mco copper hypertolerance operon carried on a mobile genetic element contributes to the virulence potential of clinical isolates of MRSA. Copper is a key component of innate immune bactericidal defenses. Here we show that copper hypertolerance genes enhance the survival of S. aureus inside primed macrophages and in whole human blood. The copB and mco genes are carried by clinical isolates responsible for invasive infections across Europe, and more broadly among three successful clonal lineages of S. au-
reus. Our findings show that a gain of copper hypertolerance genes increases the resistance of MRSA to phagocytic killing by host immune cells and imply that acquisition of this mobile genetic element can contribute to the success of MRSA.

**KEYWORDS** MRSA, P-type ATPase, *Staphylococcus aureus*, copper tolerance, macrophages, metals, mobile genetic elements, multicopper oxidase

*Methicillin-resistant Staphylococcus aureus* (MRSA) is a major problem for animal and human health and is considered a global high-priority pathogen by the World Health Organization (1). One reason why MRSA continues to be a problem is that it evolves rapidly by acquiring mobile genetic elements (MGEs) such as plasmids. Many successful contemporary clones of MRSA carry copper tolerance genes located on MGEs (2–6), but the contribution of copper hypertolerance to the fitness and virulence of *S. aureus* has not yet been studied.

Copper is a key component of innate immune bactericidal defenses, and macrophages use copper to kill intracellular bacteria by actively importing it into the phagosome (7–10). Eukaryotic copper transport is facilitated by CTR1-mediated import into the cell and ATP7a-dependent transport into the phagolysosome (7, 11). Under aerobic conditions, excess copper is proposed to catalyze the production of hydroxyl radicals via the Fenton and Haber-Weiss reactions, which may cause oxidative damage to macromolecules due to their high redox potential. Copper toxicity (under all conditions or perhaps only anoxic conditions) involves the formation of adventitious Cu(I)-thiolate bonds, thus damaging enzymes that functionally require free cysteines or disulfide bonds, such as iron sulfur cluster proteins (12, 13). The toxic properties of copper are harnessed by host phagocytes, such as macrophages (11, 14). Infection signaling, which involves elevated levels of interferon gamma (IFN-γ) and a release of copper into the plasma, may trigger activation of macrophages and increased import of copper, which enhances killing of phagocytosed bacteria (7, 10, 15).

Pathogens have evolved mechanisms to counteract copper toxicity, mainly by limiting the copper concentration in their cytoplasm through efflux or sequestration by copper metallochaperones, metallothioneins, or storage proteins (16). Almost all bacteria possess genes that confer copper tolerance, from environmental bacteria isolated from black shale in copper-rich exploration regions (17) to human pathogens. Inactivation of copper exporter genes has been shown in vivo to reduce the virulence of bacterial pathogens such as *Mycobacterium tuberculosis* (18), *Streptococcus pneumoniae* (19), *Salmonella enterica* (10), and *Pseudomonas aeruginosa* (20). In some cases, the virulence defect has been shown to be due to the inability of these pathogens to resist copper-mediated killing within the macrophage phagosome (10). Data accumulated so far suggest that copper tolerance may be a general mechanism of virulence in bacteria and that pathogens are exposed to toxic levels of copper during infection (10, 18, 19, 21).

All *S. aureus* strains possess a conserved chromosomal operon, encoding the archetypal P$_{1B-1}$-type ATPase copper transporter CopA and a copper metallochaperone CopZ, that confers low-level resistance to copper (Fig. 1A) (22). A copper hypertolerance operon (copB-mco) has been reported in some clinically relevant strains of *S. aureus*, carried either on a replicating plasmid or on a plasmid integrated into the chromosome (Fig. 1A) (2, 3, 5). The copB gene encodes a second copper-exporting P$_{1B-3}$-type ATPase (CopB), and mco encodes a multicopper oxidase implicated in copper homeostasis and the oxidative stress response (23). A chromosomally encoded homolog of the Cu-sensitive operon repressor (CsoR), first characterized in *M. tuberculosis* (24), was shown to control transcription of both operons in *S. aureus* (2).

Here we investigated the role of copper hypertolerance in *S. aureus*. We found that the copB and mco genes carried on a MGE improved bacterial growth under copper stress and enhanced bacterial survival within macrophages and in whole human blood. Expression of copB and mco was detected by intracellular bacteria isolated from macrophages, and CsoR was responsible for regulating expression of these genes in...
Finally, we determined the extent of carriage of copB and mco genes in a collection of invasive S. aureus isolates from European hospitals and in a more diverse collection of whole-genome-sequenced isolates from around the world.

**RESULTS**

The tolerance of S. aureus to copper is enhanced by the copB-mco operon. The copB-mco copper hypertolerance operon is carried either on a replicating plasmid or on a plasmid integrated into the chromosome (2, 3, 5). The role of copper tolerance genes carried on MGEs in MRSA was studied using the copB-mco operon-carrying plasmid P2-hm (3), here named pSCBU. Plasmid pSCBU was previously found to be carried by a population of MRSA clonal complex 22 (CC22) bloodstream isolates from the United Kingdom and Ireland (3). For the purposes of this study, pSCBU was introduced into S. aureus CC22 strain 14-2533T (see Table S1 in the supplemental material). 14-2533T is a clinical isolate that is representative of the lineage where pSCBU was detected, but it does not carry the plasmid. This strain was chosen as a clean and receptive host to study plasmid-conferring phenotypes.

The level of copper tolerance in strain 14-2533T carrying copB and mco genes on the replicating plasmid pSCBU was determined by measuring the minimum inhibitory
concentrations (MICs) to copper salts (Table 1). Copper tolerance was the highest in strain 14-2533T carrying the replicating plasmid pSCBU (11 mM CuCl₂), whereas the same strain without pSCBU had a lower MIC (6 mM). The individual contributions of the copB and mco genes to copper tolerance were investigated by generating isogenic mutants carrying deletions in the copper tolerance genes on the plasmid pSCBU (pSCBUΔmco and pSCBUΔcopB [Table S1 and Fig. S1]). Deletion of mco or copB resulted in a decrease in the MIC to 8 mM or 6 mM CuCl₂, respectively (Table 1), indicating that these genes are the main contributors to pSCBU-mediated copper tolerance. Inactivation of the copA gene in the 14-2533T or 14-2533T(pSCBU) background did not change the MIC compared to the wild-type strain, suggesting that the copB-mco operon is the main mediator of copper hypertolerance (Table 1). In support of a role for copB-mco in overcoming copper toxicity, 14-2533T(pSCBU) contained less intracellular copper (83.89 μM per OD₆₀₀ unit) than strain 14-2533T without the plasmid (174.53 μM per OD₆₀₀ unit) following culture in tryptic soy broth (TSB) supplemented with a subinhibitory concentration of CuCl₂ (4 mM). When cultured in TSB without added CuCl₂, strain 14-2533T and 14-2533T(pSCBU) had similar intracellular copper contents (9.42 and 8.94 μM per OD₆₀₀ unit of copper, respectively), as measured using inductively coupled plasma mass spectrometry. This suggests that copper efflux is an important mechanism of copper hypertolerance.

The pSCBU plasmid also encodes a cadmium efflux system (cadA), which is known to protect from intracellular accumulation of toxic Cd(II), Zn(II), and Co(II) (25). For a control, cadmium and zinc tolerance of the pSCBU variants was tested. We observed that pSCBU conferred resistance to cadmium and zinc (Table 1), which was unaffected by mutations in copB and mco, demonstrating that these genes do not influence tolerance to these metals.

**CsoR binds to copB promoter DNA in a copper-dependent manner.** The *S. aureus* copper-sensing transcriptional regulator (CsoR) was previously shown to negatively regulate both chromosomal and plasmid-carried copper tolerance genes (2). Dissociation of CsoR from the GC-rich palindromic promoter regions has been shown to occur at two copper-regulated operons (copA-copZ and copB-mco) in a copper-dependent manner (Fig. 1A) (24). The *S. aureus* CsoR protein shares 24% amino acid sequence identity with CsoR from *Mycobacterium tuberculosis* (2). In *S. aureus*, residues Cys⁴¹, His⁶⁶, and Cys⁷⁰ coordinate Cu(I) and CsoR with an alanine substitution at position 41 fails to dissociate from DNA in the presence of Cu(I) (26). Electrophoretic mobility shift assays (EMSAs) performed anaerobically with recombinant CsoR and an ~250-bp DNA fragment representing the copA promoter (P_copA) confirmed that the wild-type CsoR repressor bound specifically to the copA promoter, whereas anaerobic incubation with Cu(I) prevented association of CsoR with the promoter DNA (Fig. 1B). In contrast, a CsoR variant carrying C41A, H66A, and C70A substitutions (C41A/H66A/C70A substitutions

**TABLE 1** Tolerance of *S. aureus* strains to metals

| Strain          | copAZ   | copB   | mco   | MIC (mM) to Cu₂SO₄ | cadA | MIC (mM) to CdCl₂ | MIC (mM) to ZnCl₂ |
|-----------------|--------|-------|-------|--------------------|------|-----------------|-----------------|
| 14-2533T        | ch     | NE    | NE    | 6                  | NE   | 20 μM           | 20 μM           |
| 14-2533T copA::spc⁴| copA mutant | NE | NE | 6 | NE | 20 μM | 20 μM |
| 14-2533T(pSCBU) | ch     | p     | p     | 11                 | p    | 20 mM          | 20 mM           |
| 14-2533T(pSCBU) | copA mutant | p | p | 11 | p | 20 mM | 20 μM |
| 14-2533T(pSCBU) | ch     | Δ     | p     | 6                  | p    | 20 mM          | 20 mM           |
| 14-2533T(pSCBU) | ch     | Δ     | p     | 8                  | p    | 20 mM          | 20 mM           |
| 14-2533T(pSCBUΔcopB) | ch | NE | NE | 6 | NE | 20 μM | 20 μM |
| 14-2533T(pSCBUΔmco) | ch | p | p | 10 | p | 20 mM | 20 mM |
| MRSA252        | ch     | ch    | ch    | 8                  | NE   | NT             | NT              |
| MRSA252        | ch     | ch    | ch    | 5                  | NE   | NT             | NT              |

aMICs were determined by a microdilution method.
bThe presence or absence and location of the copAZ, copB, mco, and cadA genes are shown as follows: ch, gene is incorporated into the chromosome; NE, gene is not carried by the strain; p, gene is carried on a replicating plasmid; Δ, gene has been deleted by mutation.
cNT, not tested.
dspc, spectinomycin resistance (aad9).
[CHC variant herein]) remained bound to the copA promoter DNA despite the presence of copper (Fig. 1C), suggesting that it is unable to coordinate Cu(I) and therefore to undergo its copper-dependent allosteric conformational change. Thus, the CsoR CHC variant is insensitive to copper, and derepression of CsoR-regulated genes will not occur in cells expressing this variant.

Since copB and mco genes are responsible for hypertolerance to copper in *S. aureus* (Table 1), binding of CsoR to the copB and mco promoter regions was investigated. CsoR bound to DNA containing the sequence of the regions upstream of both copA and copB (P<sub>copB</sub>, but not that upstream of mco (P<sub>mco</sub>) (Fig. 1D), consistent with copB and mco being cotranscribed as part of an operon, under the regulatory control of CsoR through binding to P<sub>copB</sub>. There is no obvious CsoR binding sequence within the short intergenic region (14 bp) between copB and mco or in the 3′ sequence of copB (2).

**Copper hypertolerance enhances growth of *S. aureus* at subinhibitory concentrations of copper.** To study the role of the copper-sensitive operon repressor CsoR in copper tolerance, site-directed mutagenesis was conducted on the *csoR* genes on the chromosomes of strain 14-2533T (CC22) and the CC30 strain MRSA252 to introduce amino acid substitutions (C41A/H66A/C70A) that generate the copper-insensitive CsoR variant (CsoR CHC) (Table S1).

The MRSA252 strain carries a chromosomally integrated plasmid bearing the copB-mco operon and was more tolerant to copper (MIC of 8 mM) than its isogenic CsoR CHC mutant (5 mM) (Table 1), showing that CsoR represses the copper tolerance phenotype in strain MRSA252. In contrast, the CsoR CHC variant-expressing strain 14-2533T CHC(pSCBU) exhibited a MIC (10 mM) similar to the MIC of the parent strain 14-2553T(pSCBU) (11 mM) and an elevated MIC compared to the plasmid-negative 14-2533T host strain (6 mM). This may reflect the fact that CsoR does not fully repress copB (and by extension mco) expressed from multicopy plasmids, as shown previously by Baker et al. using a *csoR*-deficient mutant of strain ATCC 12600 (2).

To determine whether expression of the copB and mco genes had an impact on bacterial growth under copper stress, we monitored the growth of cultures in TSB containing a concentration of copper below the MIC for all strains and mutants (4 mM [Fig. 2]). Strain 14-2553T(pSCBU) grew faster and to a higher OD<sub>600</sub> in subinhibitory concentrations of copper than the same strain without the plasmid or the mutants deficient in copB or mco (Fig. 2A). The defect in growth was more pronounced for the copB mutant than for the mco mutant. In contrast, the growth profile of the 14-2533T CHC(pSCBU) mutant was identical to the growth profile of the wild-type strain carrying the plasmid (Fig. 2A), possibly due to the fact that CsoR does not fully repress copB and mco expression when they are carried on a replicating plasmid (2). There was no growth advantage observed for strains growing in TSB lacking copper (Fig. 2B) or when low (micromolar) concentrations of copper salts were added to the growth medium (data not shown). As a control, the growth of strain MRSA252 and the MRSA252 CsoR CHC mutant were compared in TSB containing a subinhibitory concentration of copper (4 mM [Fig. 2C]). Strain MRSA252 grew more quickly and reached a higher OD<sub>600</sub> than the non-copper-responsive regulatory mutant MRSA252 CHC, but not in media without copper (Fig. 2D). Transcription of copB and mco in strains 14-2553T(pSCBU) and MRSA252 was quantified by reverse transcription-quantitative PCR (RT-qPCR) (Fig. 2E). An increase in the abundance of transcript was measured for strains carrying mco and copB genes in TSB cultures of 14-2553T(pSCBU) and MRSA252 supplemented with subinhibitory concentrations of CuCl<sub>2</sub> (4 mM) compared to TSB without added copper, which confirmed that expression of copB-mco can be induced by copper. For strain MRSA252 CHC, no copB or mco expression was detected in bacteria grown in TSB supplemented with CuCl<sub>2</sub> (4 mM), showing that this strain is unable to express the copB-mco operon in response to CuCl<sub>2</sub>. The expression of copB and mco in the 14-2533T CHC(pSCBU) mutant was induced to a much lesser extent by copper than in the parent strain 14-2533T(pSCBU), and the small increase was not statistically significant. RNA transcripts of mco or copB were not detected in their respective deletion mutants (Fig. 2E and Fig. S1), as expected. Inducible copper-dependent expression of the other
gene was detected in each of the mutants, showing that the respective gene deletions had not obstructed transcription of the other gene in this operon from the \( P_{\text{copB}} \) promoter (Fig. 2E).

**Copper hypertolerance genes increase \( S. \text{aureus} \) survival inside IFN-\( \gamma \)-activated macrophages.** Copper has previously been shown to be critical for the killing of bacteria following phagocytosis (7). In the presence of copper, activated macrophages upregulate expression of the copper importer, CTR1, and commence trafficking of the

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**FIG 2** Enhanced growth in subinhibitory concentrations of copper chloride requires expression of copper tolerance genes. Growth of \( S. \text{aureus} \) 14-2533T and MRSA252 variants was measured in TSB supplemented with subinhibitory (4 mM) concentrations of copper chloride (A and C) or TSB broth alone (B and D). Growth curves representing data obtained from at least three independent experiments are presented. (E) Fold change in expression of \( \text{copB} \) and \( \text{mco} \) in \( S. \text{aureus} \) cultured in TSB versus TSB with copper chloride (4 mM). The \( \Delta \Delta CT \) method was used to determine the relative expression levels of the \( \text{copB} \) and \( \text{mco} \) genes normalized to \( \text{gyrB} \). Values are means plus standard deviations (SD) (error bars) from three independent experiments, with statistical significance determined by analysis of variance (ANOVA). Values that are statistically significantly different by ANOVA are indicated by bars and asterisks as follows: *, \( P < 0.05 \); **, \( P < 0.01 \); ***, \( P < 0.001 \). Values that are not significantly different are indicated by bars labeled ns.
P-type ATPase ATP7A to the phagolysosomal membrane, which leads to an enhanced killing of intracellular bacteria (7, 10).

To investigate whether bacterial tolerance to copper might influence the outcome for *S. aureus* following phagocytosis by macrophages, experiments were performed to quantify the survival of bacteria following phagocytosis. The murine macrophage cell line (RAW264.7) was activated with IFN-γ/H9253 and treated with CuSO4 to induce expression of the relevant copper transporters (ATP7A and CTR1), which was confirmed using RT-qPCR (Fig. S2) (7, 27). IFN-γ-activated phages internalized the wild-type strain and mutants at similar levels (data not shown). However, 3 h after phagocytosis, intracellular levels of bacteria were significantly different in the strains. The 14-2533T(pSCBU) strain survived inside the macrophages at significantly higher levels than strain 14-2533T without the plasmid (Fig. 3A). Importantly, the copper-susceptible *copB* and *mco* mutants had a survival defect compared to their parent strain 14-2533T, suggesting that copper tolerance in *S. aureus* prevents killing by macrophages (Fig. 3A). The CsoR CHC mutant of 14-2533T(pSCBU) did not show a significant survival defect in macrophages (Fig. 3A), probably reflective of the fact that CsoR-regulated genes carried on plasmids are not efficiently repressed in this strain (2) (Fig. 2), thus, it behaves like the wild type. In contrast, the MRSA252 CsoR CHC mutant had a defect in macrophage survival (Fig. 3B).

![Graph A](image1.png)  ![Graph B](image2.png)

**FIG 3** Hypertolerance to copper increases resistance of *S. aureus* to macrophage killing. Mouse macrophage cell line (RAW264.7) was suspended in DMEM supplemented with mouse IFN-γ (40 ng/ml) and CuSO₄ (40 μM) and seeded in the wells of 24-well plates at 2 × 10⁶ cells per ml for 18 h at 37°C in 5% CO₂ (A and B) *S. aureus* strain 14-2533T (A) or MRSA252 (B) and derivatives were grown overnight in RPMI 1640 and then inoculated into the wells at a multiplicity of infection (MOI) of 10 in DMEM allowing phagocytosis for 30 min followed by killing of extracellular bacteria with gentamicin/lysozyme for 30 min. Macrophages were then lysed at this time point (time zero [T0]) and after 3 h of incubation (T3), and viable bacteria were counted to determine the levels of bacterial survival. The mutants expressing CsoR C41A/H66A/C70A (CHC) are indicated. Values are means plus SD from three independent experiments. Statistical significance is indicated as follows: **, P < 0.005; *, P < 0.05; ns, not significant.

To determine whether the *copB* and *mco* genes are expressed by bacteria residing inside activated macrophages, RT-qPCR was performed using RNA obtained from intracellular bacteria at 3 h postinfection. The relative transcription levels were compared between the wild-type strains and their isogenic CsoR CHC mutants. The *copB* and *mco* genes were found to be 44- and 28-fold upregulated, respectively, in wild-type MRSA252 compared to MRSA252 CHC recovered from infected macrophages (Fig. 4A). This demonstrated that (i) *copB* and *mco* are expressed by *S. aureus* inside the macrophage and (ii) this expression is dependent on CsoR within immune cells (Fig. 4A). The same experiment was carried out with strain 14-2533T(pSCBU) and showed that *copB* and *mco* are expressed intracellularly in macrophages (Fig. 4B). However, the increase
in expression of copB and mco was much less for 14-2533T(pSCBU) than in the MRSA252 strain (Fig. 4B), which is consistent with susceptibility results (Table 1 and Fig. 2), indicating a weaker transcriptional control of CsoR over the plasmid-borne genes compared to the genes carried on the chromosome of MRSA252.

Copper hypertolerance genes increase survival of *S. aureus* in whole human blood. To determine whether the enhanced ability of copB-mco-carrying strains to survive inside activated macrophages *in vitro* may be of relevance to infection of the human host, *ex vivo* infection studies were performed with whole human blood. Consistent with results obtained for intracellular survival within activated macrophages, copper-hypertolerant *S. aureus* 14-2533T(pSCBU) had an increased ability to survive in whole human blood compared to the 14-2533T strain without the plasmid (Fig. 5A). This protection from killing in blood was due to copper resistance genes, since the mco and copB mutants had a survival defect, similar to that of the plasmid-deficient 14-2533T strain (Fig. 5A). Protection from killing in blood could be attributed to resistance to phagocytic killing, since incubation in the cell-free plasma fraction of the same blood under the same conditions yielded similar values for the wild type and mutants (Fig. 5C). There was no significant difference in the survival of the copA mutants in blood (Fig. 5A), suggesting that copB and mco, but not copA, confer protection against cellular killing in human blood.

The CsoR CHC mutant of strain MRSA252 had a significant defect in survival in whole blood compared to the wild type but did not show a defect in growth in plasma (Fig. 5B and D). This showed that failure to derepress CsoR-regulated genes (Fig. 1) impaired the ability of *S. aureus* to survive in blood. Together, these results show the importance of copper hypertolerance for *S. aureus* to resist cellular killing in human blood.

The *copB-mco* operon is carried by invasive *S. aureus* isolates and by strains belonging to CC22, CC30, and CC398. The prevalence of the *copB-mco* operon was investigated by interrogating the whole-genome sequences (WGS) of 308 invasive *S. aureus* isolates (28) from hospitals across Europe. Mapping the *copB-mco* sequences against the WGS showed that this operon was present in 55 of the invasive isolates (17.9% [Fig. S3]). The copB and mco genes were carried by isolates from two major clonal complexes (CCs) within the population, clonal complex 22 (CC22) and CC30, and also a single CC8 isolate. All CC30 strains carried the *copB-mco* operon. The most prevalent sequence type (ST) in the CC30 population carrying *copB-mco* was sequence type 30 (ST30), but ST2868, ST36 (EMRSA-16), ST2858, ST2864, ST2879, ST39, ST1829,
ST2862, ST2881, and ST34 isolates also carried the copB-mco operon. Among the CC22 strains, 50% were found to carry the operon, and all of them belonged to ST22 apart from one ST2877 isolate. In summary, copB-mco was found to be present in invasive S. aureus strains from across Europe but predominantly in isolates from two important clonal groups, CC22 and CC30 (28).

To further explore the presence of copB homologs as well as related copper tolerance genes, we interrogated all publicly available S. aureus genomes (GenBank; \( n = 8,037 \)). While a conserved copA was found universally in 99.9% of all genomes, copB homologs were the second most prevalent copper tolerance gene at ca. 34.4% of all the genomes. The copB and mco homologs were found mostly in CC22, CC30, and CC398 and only sporadically in other clonal complexes. To further characterize the distribution...
of genes in these three CCs, we constructed phylogenetic trees of each CC and mapped the presence and absence of each \textit{copB} gene to each tree. Interestingly, the distributions of genes within each clade are strikingly different. For instance, CC30 genomes show a strong conservation of \textit{copB} loci with very few predicted losses, whereas CC22 and CC398 have much more sporadic distributions that suggest multiple acquisitions and losses. This pattern could signal stronger, or more persistent, selection for \textit{copB} loci in CC30 genomes compared to CC22 and CC398, where selection may be weaker or intermittent. We also found evidence of a more diverse context to the copper hypertolerance genes than the original context in which they were found, i.e., the \textit{copB-mco} operon, with an additional putative lipoprotein-encoding gene \textit{copL} (4, 29) frequently associated with the \textit{copB-mco} operon in CC398 strains and less frequently in CC22 and CC30 strains. These data indicate that the \textit{copB-mco} copper hypertolerance genes are widely distributed in CC22, CC30, and CC398 and imply the presence of selection pressure for hypertolerance to copper.

**DISCUSSION**

The connection between gain of copper tolerance and increased virulence of several human pathogens has been reported over recent years. Here we demonstrate that \textit{S. aureus} employs copper hypertolerance genes to resist macrophage killing and to survive in whole human blood. Presumably, better survival in human blood is due to an increased resistance to killing by the cellular component, because control experiments indicated that growth in blood plasma was not affected by copper resistance genes (Fig. 5). The increased resistance to phagocytic killing conferred by the \textit{copB-mco} operon is likely to affect the virulence potential of the bacterium \textit{in vivo} and may provide a selective advantage to the pathogen. Importantly, \textit{copB} and \textit{mco} were expressed within infected macrophages, and the expression of these genes was, at least partially, dependent on expression of copper-responsive CsoR (Fig. 4). This provides indirect evidence that the \textit{copB-mco} operon is expressed intracellularly in macrophages, in response to copper. ATP7A-dependent copper transport into the macrophage phagosome is required for bactericidal activity (7). Since copper hypertolerance genes confer resistance to killing in a macrophage cell line, it is tempting to speculate that the protective effect of CopB and Mco is due to increased tolerance of \textit{S. aureus} to copper within the phagosome. However, further investigation is needed to fully understand how CopB and Mco exert their effect in phagocytic cells.

By studying the genome sequences of a collection of invasive isolates obtained from hospitals across Europe, we determined the prevalence of \textit{copB-mco} to be 17.9% of all isolates, emphasizing the clinical relevance of this locus (5, 28, 30). The \textit{copB} and \textit{mco} genes were carried by all isolates belonging to CC30, by 50% of isolates belonging to CC22, and by a single CC8 isolate. The plasmid carrying the \textit{copB-mco} operon was also recently reported to be carried by 43 to 70% of bloodstream infection isolates of \textit{S. aureus} (mostly CC22) from the United Kingdom and Ireland sampled between 2001 and 2010 (3). There is evidence of extensive loss and gain of the pSCBU (p2-hm) plasmid (3), highlighting the mobility of the \textit{copB-mco} operon within populations of \textit{S. aureus}. The global significance of copper resistance in \textit{S. aureus} was further highlighted by the widespread presence of \textit{copB} and \textit{mco} in CC22, CC30, and CC398 strains (Fig. 6). Interestingly, CC398 is the most common CC found in European livestock. Previous studies have reported that 24.3% of livestock-associated MRSA carried the \textit{copB} gene (31). The use of copper compounds as feed supplements in animal husbandry may be selecting for the carriage of copper resistance genes by MRSA (32). Copper hypertolerance in \textit{S. aureus} is likely to have broader implications for human health, since the dominant clone of community-associated (CA)-MRSA in North America (USA300) and a closely related CA-MRSA clone found in South America (USA300-LV) both independently acquired a copper resistance locus as part of the arginine catabolic mobile element and the copper and mercury resistance element, respectively (4). In both cases, the copper resistance loci are adjacent to the staphylococcal cassette chromosome \textit{mec} element (SCCmec).
Consistent with a previous report (2), our data show that the copB-mco operon mediates copper hypertolerance in *S. aureus*. Disruption of the copB or mco gene inhibited the growth of *S. aureus* in subinhibitory concentrations of copper, demonstrating that carriage of both of these genes provides a fitness advantage to *S. aureus* under copper stress. Therefore, it can be concluded that both CopB-mediated copper efflux and the activity of Mco play a role in protecting *S. aureus* from copper.

The CsoR repressor, which has been previously implicated in transcriptional regulation of copA-copZ and copB-mco (2), was shown here to control expression of copA and copB-mco in a copper-dependent manner by binding directly to the DNA sequence upstream of copA and copB but not of mco (Fig. 1). Inactivation of the Cu(I)-coordinating residues Cys41, His66, and Cys70 (CHC) disrupted copper-dependent derepression of CsoR-regulated genes. Although continued association of CsoR CHC with the copA and copB promoter DNA was confirmed by EMSA using recombinant proteins (Fig. 2), repression of the copper tolerance phenotype by the CsoR CHC variant was completely effective only in live bacteria with the chromosomally encoded copB-mco operon (strain MRSA252 CHC). In contrast, the CHC mutant of 14-2533T(pSCBU) did not completely lose the copper hypertolerance phenotype shown by the parent strain carrying a wild-type copy of the csoR gene (Table 1 and Fig. 2 and 4), due to the fact that CsoR does not fully repress copB expression from a plasmid (2). It may be that CsoR diffuses poorly to the pSCBU plasmid-located genes or that there are too few copies of the CsoR protein in the cell to fully repress copB-mco if the plasmid is present in more than one copy. Surprisingly, mutation of copA had no significant effect on copper tolerance phenotypes under the conditions used here to study copB-mco function. Our recent study conducted with *S. aureus* USA300 JE2 strain revealed that mutation of copA can influence expression of other copper-regulated genes elsewhere on the chromosome (29). A more extensive analysis of the transcriptome of *S. aureus* copper-hypertolerant strains will be needed to fully understand the phenotypes of copA mutants.

Horizontal gene transfer represents a major driving force in the evolution of *S. aureus* (33). This study provides important new insights into the contribution of MGE-carried copper hypertolerance genes to the resistance of *S. aureus* to innate immune defenses. Due to the potential for MGEs to transmit rapidly in populations of *S. aureus*, our study shows that the spread of copper hypertolerance genes could have important implications for the evolution of *S. aureus* as a pathogen.

**FIG 6** Maximum likelihood trees of CC22, CC30, and CC398 showing distribution of copA, copB, mco, and copL genes. Trees are rooted in the longest branch (CC22 to *S. aureus* 08 01492; CC398 to *S. aureus* SO1977; CC30 to *S. aureus* MRSA252). The four rings show the presence of copA, copB, mco, copL as blue, purple, yellow, and red, respectively.
MATERIALS AND METHODS

Bacterial strains and growth conditions. *S. aureus* strains used in the study are listed in Table S1 in the supplemental material. Bacteria were grown on tryptic soy agar (TSA) plates or in liquid cultures in either tryptic soy broth (TSB) or RPM 1640 at 37°C with shaking (200 rpm). To select for strains carrying pSCBU, TSA was supplemented with CdCl₂, at 1 mM. Growth curves were obtained using microtiter plates in TSB containing copper salts (either CuCl₂ or CuSO₄). For macrophage and whole-blood survival assays, bacterial strains were cultured in RPMI 1640 in aerated 50-ml Falcon tubes at 37°C with shaking (200 rpm).

Construction of mutations in plasmid-borne and chromosomally integrated *copB* and *mco* genes. Plasmid pSCBU was extracted from strain SASCBU26 (34) and used to transform strain 14-2533T (Table S1). Mutations in *S. aureus*, including deletions in the native plasmid pSCBU (Table S1 and Fig. S1), were introduced using pIMAY (35). Plasmids with deletions of the copper tolerance genes, pSCBUΔmco and pSCBUΔcopB, were isolated in the 14-2533T (clonal complex 22 [CC22]) background (Table S1 and Fig. S1). It was necessary to purify and reintroduce each validated mutated plasmid into a clean background in order to eliminate a mixed population containing a mutated and wild-type copy of this multicopy plasmid. Strain 14-2533T copA::spc was constructed by transduction of copA::spc (29) into strain 14-2533T using phage 85.

Susceptibility testing. MICs of soluble metal salts were determined by the standard broth microdilution method according to the guidelines by Clinical and Laboratory Standards Institute (CLSI). The lowest concentration of a compound showing no visible growth was recorded as the MIC.

Production and purification of recombinant CsoR. The wild-type *csoR* gene was amplified (Table S2) from *S. aureus* genomic DNA and cloned into pGEM-T (Promega). An internal Ndel site was mutated silently using QuickChange site-directed mutagenesis (Stratagene), and then *csoR* was subcloned into vector pET29a via Ndel/BamHI digestion and ligation. The *cso* CHC mutant gene was amplified from the respective pIMAY construct. Constructs were confirmed by sequencing (GATC Biotech).

Escherichia coli BL21(DE3) cells transformed with the resulting vector, pET29a-CsoR or pET29a-CsoR-CHC, were cultured in lysogeny broth (LB) at 37°C with orbital shaking at 180 rpm, and protein expression was induced at an optical density at 600 nm (OD₆₀₀) of ~0.6 by the addition of 1 mM isopropyl-β-D-thiogalactopyranoside (IPTG), followed by further incubation at 30°C for 5 h. Cells were harvested, washed, resuspended in 25 mM Tris (pH 7.5) and 15 mM dithiothreitol (DTT) containing protease inhibitor cocktail (Sigma), and lysed by sonication.

The supernatant was clarified by centrifugation and filtration and purified by anion-exchange chromatography on a 5-ml Hitrap Q HP column and an Akta purifier (GE Healthcare). Protein was eluted with 1 M NaCl. This fraction was subsequently concentrated on a 1-ml heparin column (GE Healthcare) eluted with 1 M NaCl without EDTA. Washed pellets were stored at 50°C.

EMSAs. *S. aureus* MRSA252 genomic DNA was used to PCR amplify the putative promoter regions (i.e., the ~200 bp upstream of the start codon) of *copA*, *copB*, and *mco* (Table S2), which were cloned into vector pGEM-T, confirmed by sequencing. The promoter fragments (plus ~100 bp of flanking sequence from pGEM-T) were produced by PCR amplification from these pGEM-T constructs, plus a negative-control fragment containing only the pGEM-T sequences. These PCR products were purified and used in electrophoretic mobility shift assays (EMSAs).

EMSAs were performed by incubating fully reduced (as determined with Ellman’s reagent) recombinant CsoR variants (0 to 100 μM) with the respective promoter DNA plus the negative-control DNA (both 0.1 μM) in 20 mM HEPES (pH 7.0), 100 mM NaCl, 100 ng/μl poly(dI-dC) (Sigma), 1 mM DTT, 0.4 mg/ml bovine serum albumin (BSA) at room temperature for 30 min. All incubations were performed anaerobically inside an N₂ atmosphere glove box ([O₂] < 5 ppm) (Belle Technology), and Cu(II)-CsoR was prepared by anaerobically incubating protein for 10 min with 1 mol equivalent of Cu(II) prepared as previously described (36). After incubation, samples were resolved on 6% acrylamide (wt/vol) native PAGE for 60 to 80 min at 82 V and stained with 10% SYBR Safe solution (Invitrogen) for 20 min.

Inductively coupled plasma mass spectrometry. *S. aureus* 14-2533T and 14-2533T(pSCBU) bacteria were grown overnight in TSB and then subcultured into TSB or TSB supplemented with CuCl₂ (4 mM) for 16 h. Samples were normalized to the same OD₆₀₀ harvest by centrifugation, and washed twice in 50 mM Tris (pH 7.5), 100 mM NaCl, and 10 mM EDTA, followed by two washes in 50 mM Tris (pH 7.5) and 100 mM NaCl without EDTA. Washed pellets were stored at −20°C until use, then thawed, and digested with 65% (wt/vol) nitric acid (Merck) for 48 h. Digests were centrifuged at 21,000 × g for 20 min at 4°C, and the supernatants were analyzed using inductively coupled plasma mass spectrometry (Thermo X-series2). Samples were diluted 10-fold in 2% nitric acid containing 20 μg/liter platinum and indium as internal standards and analyzed (100 reads, 30-ms dwell, five channels, 0.02 atomic mass unit separation, each in triplicate) for ⁵⁵Mn, ⁶⁵Cu, ⁶⁶Zn, ¹¹⁴In, and ¹⁹⁵Pt in collision cell mode (3 ml/min 8% H₂ in He collision gas), and metal concentrations were determined by comparison to matrix-matched elemental standard solutions (Merck).

RNA extraction. (i) RNA isolation from *S. aureus*. To isolate RNA from *S. aureus*, bacterial cultures were grown in 20 ml TSB with or without copper salts (as indicated) to an OD₆₀₀ of ~0.6. Cultures were suspended in phenol:ethanol (5:95) mixture and incubated on ice for 1 h before pelleting the cells by centrifugation. At this step, pellets were either stored at −70°C or subjected to total RNA extraction. To extract RNA, the pellet(s) was gently suspended in 1 ml of TRIzol following lysis using FastPrep lysing beads (three times, each time for 45 s, 2-min intervals on ice). Aqueous lysate was then mixed with
chloroform (2:1) in Phase Lock Gel to separate the RNA-containing aqueous upper layer from the high-density organic lower phase. The upper phase was precipitated with isopropanol (1:1) following ultracentrifugation at top speed for 30 min. The pellet was washed with 70% (vol/vol) ethanol and centrifuged. Supernatant was removed, and the RNA pellet was dried.

(ii) RNA isolation from macrophages. RNA isolation from macrophages was performed using a modified TRizol-based method. RAW264.7 cells were lysed directly in the culture dish by adding 12 ml of TRizol per T-175 cm² flask and scraping the cells. Chloroform was added to the suspension at 0.2 ml per 1 mM TRizol reagent. Samples were immediately vortexed and incubated at room temperature for 2 to 3 min. Following centrifugation at 12,000 × g for 15 min at 4°C, the mixture separated into layers, and the upper aqueous layer was collected, precipitated with 0.5 ml isopropanol per 1 ml of TRizol, incubated at room temperature for 10 min, and centrifuged at 12,000 × g for 10 min at 4°C. The RNA pellet obtained was washed once with 75% ethanol (adding at least 1 ml per 1 ml of TRizol).

(iii) RNA isolation from intracellular S. aureus. To isolate RNA from intracellular S. aureus, a combination of the above methods was used. First, cells were infected in T-175 cm² flasks following gentamicin/lysozymin killing of extracellular bacteria and monolayer washing. The cells were then lysed with TRizol as described above. Centrifugation at 4,000 × g for 20 min was performed to separate the bacteria into a pellet. RNA from the bacterium-containing pellet and macrophage RNA-containing suspension was extracted by the respective methods.

All air-dried pellets were dissolved in RNase-free molecular-grade water, and their stability and purity were checked by gel electrophoresis. The concentrations were determined using a Thermo Fisher Scientific NanoDrop spectrophotometer.

RT-qPCR. RNA was digested by DNase I treatment (Qiagen) according to the manufacturer’s instructions and quantified using a NanoDrop spectrophotometer, and the integrity of RNA was assessed by electrophoresis. RNA was reverse transcribed to cDNA using High Capacity RNA-to-cDNA kit (Applied Biosystems). Reverse transcription-quantitative PCR (RT-qPCR) was performed using the Power SYBR green PCR master mix (Applied Biosystems). The relative levels of gene expression in the treated cells and the nontreated controls were calculated by relative quantification using gyrB as the reference gene and using the primers in Table S2. All samples were amplified in triplicate, and data analysis was conducted using StepOne software (Applied Biosystems).

Genomic DNA was isolated from cultured macrophages as described previously (http://cancer.ucsf.edu/_docs/cores/array/protocols/dna_cell_culture.pdf). The isolated DNA was used as the template to generate a standard curve.

Macrophage survival assays. A murine macrophage cell line (RAW264.7) was cultured in Dulbecco modified Eagle medium (DMEM) containing 10% (vol/vol) fetal bovine serum (FBS). To generate monolayers, 2 × 10⁶ cells per ml were seeded in the wells of 24-well plates (500 µl per well) and incubated for 24 h in serum-free DMEM supplemented with CuSO₄ (40 µM) and mouse interferon gamma (IFN-γ) (50 µg/ml) for 18 h at 37°C and 5% CO₂. Immediately before the infection, RAW264.7 monolayers were washed with ice-cold DMEM alone. S. aureus strains were cultured in RPMI 1640. Immediately before the experiment, bacteria were washed twice with DMEM and adjusted to an OD₆₀₀ of 0.05 (ca. 2 × 10⁷ CFU per ml) in DMEM and inoculated into the monolayers for 30 min. The monolayers were subsequently washed, and extracellular bacteria were killed by treatment with gentamicin (200 µg/ml) and lysostaphin (100 µg/ml) for 30 min. Monolayers were then washed and lysed with ice-cold water at time zero (T0) and after additional 3 h of incubation (T3) to determine the survival rates (CFU per milliliter). Lysates were plated on agar, and CFU were counted to determine the numbers of viable bacteria.

Human blood survival assays. The quantification of S. aureus in human blood was performed by the method of O’Halloran et al. (37). Briefly, S. aureus variants were grown in RPMI 1640 to stationary phase and diluted in RPMI 1640, 25 µl (containing ca. 1 × 10⁶ CFU/ml) was added to 475-ml fresh blood samples obtained from healthy human volunteers, and blood samples were treated with 50 mg/ml of hirudin anticoagulant (Refludan; Pharmion). The tubes were incubated at 37°C with gentle rocking, and after 3 h, serial dilutions were plated to determine the CFU/milliliter of viable bacteria. In parallel, an equal inoculum was incubated with cell-free plasma derived from the same donor’s blood. Bacterial numbers in plasma were quantified (CFU/milliliter) at the 3-h time point, and percent survival of the original inoculum was determined. Ethical approval for the use of human blood was obtained from the Trinity College Dublin Faculty of Health Sciences ethics committee.

Phylogenetic matrix construction and gene presence or absence. All preassembled genomes from public databases for CC22, CC30, and CC398 (n = 1,075, 320, and 707, respectively) were used for whole-genome alignment with reference to the S. aureus N315 genome, using the NUCmer and show-snps utilities of MUMmer (http://mummer.sourceforge.net) (38). The S. aureus genomes were assigned sequence types (STs) and CCs by the S. aureus multilocus sequence type (MLST) typing scheme https://pubmlst.org/saureus/ site at the University of Oxford (39) using the MLST typing perl script v. 2.9 for contigs (https://github.com/teemann/mlst), and thereafter membership in each CC was determined by clade membership in a large (n = 8,037; unpublished data) S. aureus data set composed of all publicly available preassembled genomes. All regions from the reference genome annotated as mobile genetic elements were excluded. We also applied a mask that excluded repetitive sequences from the reference genome that were >80% identical over at least 100 nucleotides to other genomic loci, based on pairwise MegaBLAST-based analysis (40). For each CC, a maximum likelihood phylogeny was constructed with RAxML v8.2.11 (41) using an ascertainment bias correction and the general time-reversible (GTR) substitution model (42) accounting for among-site rate heterogeneity using the I° distribution and four rate categories (43) (ASC_GTR Gamma model) for 100 individual searches with maximum parsimony
random-addition starting trees. Node support was evaluated with 100 nonparametric bootstrap pseudoreplicates (44). We used the copA, copB, and copL genes from strain TCH1516 and mco from strain CA12 to search for closely related genes in the genus *Staphylococcus* in GenBank (wgs and nr databases, 9,222 genomes as of 16 August 2017) using BLAST (tblastx with a cutoff value of 1e−130 for copA, copB, and mco and 1e−90 for copL) (45). The four genes were mapped to the three trees as high-quality circular representations using GraPhlAn software tool (https://bitbucket.org/nsegata/graphlan/). The richness of the color shows the percentage similarity with the seed sequence used.

**Statistics.** The data presented in this study are means ± standard deviations (SD) from three experiments unless stated otherwise. Statistical significance was assessed by two-way analysis of variance (ANOVA) and indicated in the figure legends.

**SUPPLEMENTAL MATERIAL**

Supplemental material for this article may be found at https://doi.org/10.1128/mBio.00550-18.

**FIG S1**, TIF file, 0.2 MB.

**FIG S2**, TIF file, 0.1 MB.

**FIG S3**, PDF file, 0.2 MB.

**TABLE S1**, DOCX file, 0.02 MB.

**TABLE S2**, PDF file, 0.01 MB.

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**REFERENCES**

1. World Health Organization. 2017. Prioritization of pathogens to guide discovery, research and development of new antibiotics for drug-resistant bacterial infections, including tuberculosis. World Health Organization, Geneva, Switzerland.

2. Baker J, Sengupta M, Jayaswal RK, Morrissey JA. 2011. The *Staphylococcus aureus* CsoR regulates both chromosomal and plasmid-encoded copper resistance mechanisms. Environ Microbiol 13:2495–2507. https://doi.org/10.1111/j.1462-2920.2011.02522.x.

3. Jamroz D, Coll F, Mather AE, Harris SR, Harrison EM, MacGowan A, Karas A, Elston T, Esteve Torok M, Parkhill J, Peacock SJ. 2017. Evolution of mobile genetic element composition in an epidemic methicillin-resistant *Staphylococcus aureus* temporal changes correlated with frequent loss and gain events. BMC Genomics 18:684. https://doi.org/10.1186/s12864-017-4065-z.

4. Planet PJ, Diaz L, Kolokotronis SO, Narechania A, Reyes J, Xing G, Rincon S, Smith H, Paneso D, Ryan C, Smith DF, Guzman M, Zunta J, Sebra R, Deikus G, Nolan RL, Tenover FC, Weinstock GM, Robinson DA, Arias CA. 2015. Parallel epidemics of community-associated methicillin-resistant *Staphylococcus aureus* USA300 infection in North and South America. J Infect Dis 212:1874–1882. https://doi.org/10.1093/infdis/jiv320.

5. Holden MT, Feil EJ, Lindsay JA, Peacock SJ, Day NP, Enright MC, Foster TJ, Moore CE, Hurst L, Akins R, Barron A, Benton NL, Bellamy SD, Chillingworth T, Churcher C, Clark L, Corston C, Cronin A, Doggett J, Dowd L, Feltwell T, Hance Z, Harris B, Hauser H, Holroyd S, Jagels K, James KD, Lennard N, Line A, Mayes M, Moule S, Mungall K, Ormond D, Quail MA, Rabinowitsch E, Rutherford K, Sanders M, Sharp S, Simmonds M, Stevens K, Whitehead S, Barrell BG, Spratt BG, Parkhill J. 2004. Complete genomes of two clinical *Staphylococcus aureus* strains: evidence for the rapid evolution of virulence and drug resistance. Proc Natl Acad Sci USA 101:9786–9791. https://doi.org/10.1073/pnas.0402521101.

6. Gómez-Sanz E, Kadlec K, Fessler AT, Zarazaga M, Torres C, Schwarz S. 2013. Novel erm(T)-carrying multiresistance plasmids from porcine and human isolates of methicillin-resistant *Staphylococcus aureus* ST398 that also harbor cadmium and copper resistance determinants. Antimicrob Agents Chemother 57:3275–3282. https://doi.org/10.1128/AAC.00171-13.

7. White C, Lee J, Kambe T, Fritsche K, Petris MJ. 2009. A role for the ATP7A copper-transporting ATPase in macrophage bactericidal activity. J Biol Chem 284:33949–33956. https://doi.org/10.1074/jbc.M109.070201.

8. Hodgkinson V, Petris MJ. 2012. Copper homeostasis at the host-pathogen interface. J Biol Chem 287:13495–13555. https://doi.org/10.1074/jbc.R111.316406.

9. Djoko KY, Ong CL, Walker MJ, McEwan AG. 2015. The role of copper and zinc toxicity in innate immune defense against bacterial pathogens. J Biol Chem 290:18954–18961. https://doi.org/10.1074/jbc.R115.647099.

10. Ladomerys E, Khan A, Shanbhag V, Cavet JS, Chan J, Weissman GA, Petris MJ. 2017. Host and pathogen copper-transporting P-type ATPases function antagonistically during *Salmonella* infection. Infect Immun 85:https://doi.org/10.1128/IAI.00351-17. 10.1128/IAI.00351-17.

11. Fest RA, Thiele DJ. 2012. Copper at the front line of the host-pathogen battle. PLoS Pathog 8:1002887. https://doi.org/10.1371/journal.ppat.1002887.

12. Macomber L, Imlay JA. 2009. The iron-sulfur clusters of dehydratases are primary intracellular targets of copper toxicity. Proc Natl Acad Sci USA 106:8344–8349. https://doi.org/10.1073/pnas.0812808106.

13. Gaupp R, Ledala N, Somerville GA. 2012. Staphylococcal response to oxidative stress. Front Cell Infect Microbiol 2:33. https://doi.org/10.3389/fcimb.2012.00033.
MGE-Encoded Copper Hypertolerance in S. aureus

14. Achard ME, Stafford SL, Bokil NJ, Chartres J, Bernhardt PV, Schembri MA, Sweet MJ, McEwan AG. 2012. Copper redistribution in murine macrophages in response to Salmonella infection. Biochem J 444:51–57. https://doi.org/10.1042/BJ20112180.

15. Hordyjewska A, Popiolek L, Kocot J. 2014. The many “faces” of copper in medicine and treatment. Biomolats 27:611–621. https://doi.org/10.1007/s10534-014-9736-5.

16. Gold B, Deng H, Bryk R, Vargas D, Eliezer D, Roberts J, Jiang X, Nathan C. 2008. Identification of a copper-binding metallothionein in pathogenic mycobacteria. Nat Chem Biol 4:609–616. https://doi.org/10.1038/ncb108.

17. Dziwiet L, Pyzik A, Szuplewska M, Matlakowska R, Mińcki S, Wiberg D, Schluter A, Puhler A, Bartosik D. 2015. Diversity and role of plasmids in adaptation of bacteria inhabiting the Lubin copper mine in Poland, an environment rich in heavy metals. Front Microbiol 6:152. https://doi.org/10.3389/fmicb.2015.00152.

18. Ward SK, Abomoeuk B, Hoye EA, Steinberg H, Talaat AM. 2010. CtpV: a putative copper exporter required for full virulence of Mycobacterium tuberculosis. Mol Microbiol 77:1096–1110. https://doi.org/10.1111/j.1365-2958.2010.07273.x.

19. Johnson MD, Kehl-Fie TE, Klein R, Kelly J, Burnham C, Mann B, Rosch JW. 2015. Role of copper efflux in pneumococcal pathogenesis and resistance to macrophage-mediated immune clearance. Infect Immun 83:1684–1694. https://doi.org/10.1128/IAI.00315-14.

20. Schwan WR, Warrener P, Keunz E, Stover CK, Folger KR. 2005. Mutations in the cua gene encoding a copper homeostasis P-type ATPase reduce the pathogenicity of Pseudomonas aeruginosa in mice. Int J Med Microbiol 295:237–242. https://doi.org/10.1016/j.ijmm.2005.05.005.

21. Neyrolles O, Mintz E, Catty P. 2013. Zinc and copper toxicity in host defense against pathogens: Mycobacterium tuberculosis as a model example of an emerging paradigm. Front Cell Infect Microbiol 3:8. https://doi.org/10.3389/fcimb.2013.00089.

22. Sithisak S, Knutsos L, Webb JW, Jayaswal RK. 2007. Molecular ample of an emerging paradigm. Front Cell Infect Microbiol 3:89. https://doi.org/10.1128/mBio.00444-16.

23. Purves T, James T, Riboldi GP, Zapotoczna M, Tarrant E, Andrew PW, Londono A, Planet PJ, Geoghegan JA, Waldron KJ, Morrissey JA. 2018. A horizontally gene transferred copper resistance locus confers hyper-resistance to bacteriophage copper toxicity and enables survival of community acquired methicillin resistant Staphylococcus aureus USA300 in macrophages. Environ Microbiol 20:1576–1589. https://doi.org/10.1111/1462-2920.14088.

24. Grundmann H, Aanensen DM, van den Wijngaard CC, Spratt BG, Hansen D, Friedrich AW, European Staphylococcal Reference Laboratory Working Group. 2010. Geographic distribution of Staphylococcus aureus causing invasive infections in Europe: a molecular-epidemiological analysis. PLoS Med 7:e1000215. https://doi.org/10.1371/journal.pmed.1000215.

25. Argudín MA, Butaye P. 2016. Dissemination of metal resistance genes among animal methicillin-resistant coagulase-negative Staphylococci. Res Vet Sci 105:192–194. https://doi.org/10.1016/j.rvsc.2016.02.009.

26. Yazdankhah S, Rudi K, Bernhoft A. 2014. Zinc and copper in animal feed – development of resistance and co-resistance to antimicrobial agents in bacteria of animal origin. Microb Ecol Health Dis 25 https://doi.org/10.3402/mehd.v25.235862.

27. Lindsay JA. 2014. Staphylococcus aureus genomics and the impact of horizontal gene transfer. Int J Med Microbiol 304:103–109. https://doi.org/10.1016/j.ijmm.2013.11.010.

28. Harris SR, Cartwright EJ, Torok ME, Holden MT, Brown NM, Ogily-Stuart AL, Ellington MJ, Quail MA, Bentley SD, Parkhill J, Peacock SJ. 2013. Whole-genome sequencing for analysis of an outbreak of meticillin-resistant Staphylococcus aureus: a descriptive study. Lancet Infect Dis 13:130–136. https://doi.org/10.1016/S1473-3099(12)70268-2.

29. Monk IR, Shah IM, Xu M, Tan MW, Foster TJ. 2012. Transforming the untransformable: application of direct transformation to manipulate genetically Staphylococcus aureus and Staphylococcus epidermidis. mBio 3:e00277-11. https://doi.org/10.1128/mBio.00277-11.

30. Vita N, Platsaki S, Basle A, Allen SJ, Paterson NG, Crombie AT, Murrell JC, Waldron KJ, Denison C. 2015. A four-helix bundle stores copper for methanol oxidation. Nature 525:140–143. https://doi.org/10.1038/nature14854.

31. O’Halloran DP, Wynne K, Geoghegan JA. 2015. Protein A is released into the Staphylococcus aureus culture supernatant with an unprocessed sorting signal. Infect Immun 83:1598–1609. https://doi.org/10.1128/IAI.03122-14.

32. Kurtz S, Phillippy A, Delcher AL, Smoot M, Shunwany M, Antonescu C, Salzberg SL. 2004. Versatile and open software for comparing large genomes. Genome Biol 5:R12. https://doi.org/10.1186/gb-2004-5-2-r12.

33. Jolley KA, Maidan MC. 2010. BIGSdb: scalable analysis of bacterial genome variation at the population level. BMC Bioinformatics 11:595. https://doi.org/10.1471-2105-11-595.

34. Morguilis A, Coulouris G, Raytisylis T, Madden TL, Agarwala R, Schaffer AA. 2008. Database indexing for production MegaBLAST searches. Bioinformatics 24:1757–1764. https://doi.org/10.1093/bioinformatics/btn322.

35. Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics 30:1312–1313. https://doi.org/10.1093/bioinformatics/btu033.

36. Lanave C, Preparata G, Saccone C, Serio G. 1984. A new method for calculating evolutionary substitution rates. J Mol Evol 20:86–93. https://doi.org/10.1007/BF02101990.

37. Yang Z. 1994. Maximum likelihood phylogenetic estimation from DNA sequences with variable rates over sites: approximate methods. J Mol Evol 39:306–314. https://doi.org/10.1007/BF02106154.

38. Felsenstein J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. Evolution 39:783–791. https://doi.org/10.1111/j.1558-5646.1985.tb04200.x.

39. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/ S0022-2836(05)80360-2.

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