Varadia, a new helicarionoidean semi-slug genus from India’s Western Ghats (Stylommatophora: Helicarionoidea)

Amrut R. BHOSALE 1,*, Ahmed J. SAADI 2, Christopher M. WADE 3, Tejas U. THACKERAY 4, Asif S. TAMBOLI 5, Suhas K. KADAM 6, Dipak V. MULEY 7 & Dinarzarde C. RAHEEM 8,**

1,7 Department of Zoology, Shivaji University, Kolhapur, 416004, India.
1 Department of Zoology, Sadguru Gadage Maharaj College, Karad, India.
1 Foundation for Biodiversity Conservation, A1903, Shubh Kalyan, Nanded City, Pune 411041, India.
2,3 School of Life Sciences, The University of Nottingham, University Park, Nottingham NG7 2RD, UK.
2 Unit for Integrative Zoology, Department of Evolutionary Biology, University of Vienna, UZA1 Althanstraße 14, 1090 Vienna, Austria.
4 Thackeray Wildlife Foundation, Vaibhav Chambers, BKC, Bandra, Mumbai, 400051, India.
5,6 Department of Biochemistry, Shivaji University, Kolhapur 416004, India.
3 Research Institute for Dok-do and Ulleung-do Island, Department of Biology, School of Life Sciences, Kyungpook National University, 80 Daehak-ro, Buk-gu, Daegu- 41566, Republic of Korea.
8 Department of Life Sciences, Natural History Museum, London, SW7 5BD, UK.

* Corresponding author: amrutb18@gmail.com
** Co-corresponding author: d.raheem@nhm.ac.uk
2 Email: ahmed.saadi@nottingham.ac.uk; ahmed.saadi@univie.ac.at
3 Email: chris.wade@nottingham.ac.uk
4 Email: tejasthackeray@gmail.com
5 Email: asift456@gmail.com
6 Email: suhas397@gmail.com
7 Email: drdvmuley@gmail.com

Abstract. We here describe a new Indian helicarionoidean genus, Varadia Bhosale & Raheem gen. nov., containing the single species Varadia amboliensis Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. This new semi-slug is endemic to the northern and central Western Ghats and is primarily a forest-
living species. We describe and figure the shell, reproductive system, radula, spermatophore and external morphology of this new species, and detail its known distribution. We explore its relationships to other helicarioideans using phylogenetic analyses of DNA sequence data for part of the ribosomal RNA gene cluster and discuss the morphology of the new genus in relation to other, primarily South Indian, helicarioidean taxa. Based on characters of the reproductive system, particularly the male genitalia and the gametolytic sac, we provisionally place *Varadia* gen. nov. in the Macroclymidae (Ariophantidae). This is consistent with the results of our molecular phylogenetic analyses. The combination of large size, broad, densely tuberculated shell lobes and a shell with ca 4 whorls and a disproportionately large body whorl makes *V. amboliensis* gen. et sp. nov. unique among the helicarioidean taxa of the Western Ghats. The new semi-slug is also highly distinctive in the morphology of its male genitalia.

**Keywords.** *Varadia amboliensis* gen. et sp. nov., Macroclymidae, taxonomy, phylogenetics, ribosomal RNA gene cluster.

Bhosale A.R., Saadi A.J., Wade C.M., Thackeray T.U., Tamboli A.S., Kadam S.K., Muley D.V. & Raheem D.C. 2021. *Varadia*, a new helicarioidean semi-slug genus from India’s Western Ghats (Stylommatophora: Helicarioidea). *European Journal of Taxonomy* 757: 50–79. https://doi.org/10.5852/ejt.2021.757.1413

**Introduction**

The stylommatophoran superfamly Helicarioidea Bourguignat, 1877 is particularly diverse in the tropics and subtropics of Asia, Africa and Australia (Blanford & Godwin-Austen 1908; Hausdorf 2000). South Asia is a major centre of diversity with at least 21 genera and more than 400 described species (Blanford & Godwin-Austen 1908), representing two of three currently recognized helicarioidean families (Helicarioidea Bourguignat, 1877 and Ariophantidae Godwin-Austen, 1888; Bouchet *et al.* 2017). A substantial part of this diversity is restricted to the Western Ghats–Sri Lanka (WGSL) hotspot, which has a taxonomically diverse helicarioidean fauna (123 species in 15 genera; Naggs & Raheem 2000; Raheem *et al.* 2014). This fauna is dominated by species belonging to the ariophantid subfamily Ariophantinae, a group that in its strictest sense is restricted to Peninsular India and Sri Lanka and is characterized by distinctive reproductive anatomy (Blanford & Godwin-Austen 1908; but see Zilch 1959 and Bank 2017). The WGSL fauna also includes taxa belonging to other helicarioidean genera, such as *Eurychlamys* Godwin-Austen, 1899 (Helicarioidea: Helicarioinidae), *Macrochlamys* sensu Godwin-Austen (1883) (Ariophantidae: Macroclymidae Godwin-Austen, 1888), *Pseudaustenia* Cockerell, 1891 (Helicarioidea: Helicarioinidae), *Satiella* Blanford & Godwin-Austen, 1908 (Helicarioidea: Durgellinae Godwin-Austen, 1888) and *Sitala* Adams, 1865 (Helicarioidea: Durgellinae). The helicarioideans of the WGSL hotspot are diverse in external morphology (Blanford & Godwin-Austen 1908). They range from snails, such as *Ariophanta* Desmoulins, 1829, *Euplecta* Semper, 1870, *Eurychlamys* and *Sitala*, with well-developed, robust or heavy shells into which the body can be fully withdrawn, to taxa with greatly reduced external or internal shells (semi-slugs and slugs, respectively, sensu Cameron 2016). Semi-slug taxa include *Indrella ampulla* (Benson, 1850), in which the shell is globose, fragile and not covered by the mantle; *Satiella* and *Ratnadvipia karui* Raheem & Naggs, 2006, both characterized by flexible and mostly proteinaceous shells that are largely covered by the mantle; and *Pseudaustenia*, which has an auriform shell that is almost entirely exposed.

Our current understanding of this taxonomically and morphologically diverse fauna has many gaps and is largely based on publications and collections from the British colonial period (see monographs by Blanford & Godwin-Austen 1908; Raheem *et al.* 2014). Many taxa were poorly described/defined and/or were not illustrated in the original literature, hence making their identification problematic. The genus-level placement of some taxa is uncertain and questionable (e.g., at least some of the western and southern Indian species assigned to the genus *Macrochlamys* may belong to *Eurychlamys*; Blanford &
Godwin-Austen 1908; Raheem et al. 2014). Data on the reproductive anatomy, external morphology and radula are scarce or lacking for most species. Knowledge of species’ distributions is poor; while many species are known only from single localities and have not been recorded for over a century, the snail fauna of substantial parts of the Western Ghats remains to be explored (Raheem et al. 2014). To date, although South Asian taxa have been included in molecular phylogenetic analyses of deep-level pulmonate relationships (e.g., Wade et al. 2001, 2006; Herbert & Mitchell 2009; Ramirez et al. 2012), studies focussing on the molecular systematics and diversification of South Asian helicarionoideans have yet to be carried out. With the exception of the substantial work done on Australian helicarionids (e.g., Hyman et al. 2007, 2017; Hyman & Ponder 2010; Hyman & Köhler 2019), this reflects the general scarcity of such studies globally (but see Schilthuizen et al. 2019; Pholyotha et al. 2020a).

The taxonomic study of the WGSL land-snail fauna has seen a renewed interest in recent years (e.g., Raheem & Naggs 2006; Raheem et al. 2014; Aravind & Páll-Gergely 2018; Bhosale et al. 2019, in press; Páll-Gergely et al. 2020). This includes the first modern surveys of the land-snail fauna of the northern Western Ghats by one of us (Bhosale et al. 2016, 2019, in press; Bhosale 2018). Begun as PhD fieldwork, these surveys focussed initially on Kolhapur District, Maharashtra, a 7685-km² area in the extreme south of the northern Western Ghats, which had not been previously explored by malacologists. It was during these surveys that a large, forest-living semi-slug with a *Macrochlamys*-like shell and broad, conspicuously tuberculate shell lobes was encountered. Based on comparative morphological study and molecular phylogenetic analyses of part of the ribosomal RNA (rRNA) gene cluster, we here describe this semi-slug as a new Indian helicarionoidean genus, *Varadia* Bhosale & Raheem gen. nov., containing the single species *V. amboliensis* Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. We discuss the morphology of this new genus in relation to other, primarily South Indian, taxa and explore its phylogenetic relationships to other helicarionoideans, provisionally placing it in the subfamily *Macrochlamydyinae* (Ariophantidae).

**Material and methods**

Living snails and shells of *Varadia amboliensis* gen. et sp. nov. were collected and/or observed by A. Bhosale at 5 localities in the state of Maharashtra, India in September 2017, November 2019 (during the autumn) and in September 2020, towards the end of the monsoon rains (Fig. 1). After sampling of foot tissue for DNA analysis, snails were euthanized following the guidelines of the American Veterinary Medical Association (2020) and preserved in 80% ethanol for anatomical study. The whole radulae were prepared following Bhosale et al. (2019) and SEM images were taken on a TESCAN VEGA3 SEM. All radula counts were carried out in the area of greatest width. Morphological studies of collected material were carried out at the Department of Zoology, Shivaji University, Kolhapur, India, with type and other material being deposited in the collections of the Bombay Natural History Society (BNHS), Mumbai and Zoological Survey of India (ZSI), Western Regional Centre (WRC), Pune, India. Unless stated otherwise, we follow the morphological terminology of Raheem & Naggs (2006) and Bhosale et al. (2019) for the mantle and reproductive system (the terms ‘proximate’ and ‘distal’ are used with reference to the genital orifice); Hyman & Köhler (2019) for the spermatophore; and Cox (1960) for the shell (including shell sculpture). Shell whorl counts (to the nearest quarter whorl) are based on the approach of Kerney & Cameron (1979). Shell height and width were measured as shown in Fig. 2. Unless stated otherwise, all photographic images are by A. Bhosale.

For the molecular phylogenetic study, our ingroup consisted of 30 taxa belonging to the order Limacoidea Batsch, 1789 (Table 1). The outgroup comprised three species of Arionoidea Gray, 1840; a recent study has shown that the orders Arionoidea and Limacoidea are sister groups (Saadi & Wade 2019). Unless stated otherwise, we have followed the higher-level classification of Bouchet et al. (2017). Phylogenetic analyses were based on the region of the rRNA gene cluster described by Wade & Mordan (2000). DNA sequence data for eight of the 33 taxa were generated during the course of the present study; data for
the remainder were generated in previous studies by Wade et al. (2001, 2006). For the newly sequenced samples, DNA was extracted from a 1–2-mm piece of foot tissue using a CTAB DNA extraction protocol (Goodacre & Wade 2001). PCR amplifications were done by adding 2.5 μl of DNA extract to a reaction mix containing 1× PCR reaction buffer (including 1.5 mm MgCl₂), 0.2 mm each dNTP (Sigma-Aldrich, USA), 0.2 μM each primer and 1 unit Taq DNA polymerase (Sigma-Aldrich, USA) in a final volume of 25 μl. The PCR conditions were as follows: 96°C for 1 min; 30 cycles of 94°C for 30 s, 50°C for 30 s

Fig. 1. Distribution of Varadia amboliensis Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. (coloured symbols) in the Western Ghats (green shading; sensu Irfan-Ullah & Davande 2008). The yellow square indicates the type locality of the new species and the red circles are the other localities.
and 72°C for 1 min; and a final extension step of 72°C for 5 min. Amplified products were purified using the HiPurA™ Quick Gel Purification Kit. Sequencing was carried out using a BigDye Terminator ver. 3.1 Cycle Sequencing Kit (Applied Biosystems, USA) with both DNA strands sequenced on a 3500 Genetic Analyzer (Applied Biosystems, USA); sequencing was done at the Department of Biochemistry, Shivaji University, Kolhapur. DNA sequences were assembled using STADEN ver. 1.5.3 (Staden et al. 2000). Subsequently, the sequences were manually aligned using the Genetic Data Environment ver. 2.2 package (Smith et al. 1994).

Phylogenetic analyses were based on a dataset of 874 unambiguously aligned nucleotide sites for 33 taxa. We used maximum likelihood (ML; Felsenstein 1981), neighbour joining (NJ; Saitou & Nei 1987) and Bayesian (Larget & Simon 1999) approaches, with the nucleotide substitution model being GTR + Γ (Lanave et al. 1984; Gu et al. 1995). ML analyses were carried out using RAxMLHPC2 (Stamatakis 2014). Each RAxML analysis involved a single programme run and consisted of a rapid bootstrap analysis with the extended majority rule bootstopping criterion, followed by a search for the best-scoring ML tree (Stamatakis 2014); analyses were run on the CIPRES Science Gateway ver. 3.3 (http://www.phylo.org/index.php, Miller et al. 2010) and each run was repeated at least once. NJ analysis was performed using PAUP* ver. 4.0b10 (Swofford 2002) with model parameters estimated following an iteration process; for each tree the parameters were estimated and used to build the next tree until there was no further improvement of the likelihood score. Bootstrap resampling (Felsenstein 1985) with 1000 replicates was undertaken for the NJ trees. Bayesian analyses were done using MrBayes ver. 3.1.2 (Ronquist & Huelsenbeck 2003). Each MrBayes analysis involved two independent MCMC runs (with four chains per run) for 5 million generations, sampling every 100 generations; each analysis was repeated at least once. To ensure adequate chain swapping, a range of heating parameters were tested with the optimal parameter used to construct the final trees. Only after the Bayesian MCMC searches had reached a stationary phase (indicating convergence of the chains onto the target distribution) was the run ended. A consensus tree was built using the last 75% of trees (burnin = 5001). Trees were rooted on a composite outgroup comprising three members of the Arionoidea, *Arion hortensis* Féreussac, 1819, *Meghimatium bilineatum* (Benson, 1842) and *Philomyrus carolinianus* (Bosc, 1802). Branches with bootstrap support (BS) values ≥ 70% and posterior probabilities (PP) ≥ 0.95 were considered to be well/strongly supported (Hillis & Bull 1993; Alfaro & Holder 2006). The frequency of all bipartitions in the bootstrap trees used for computing the NJ and ML consensus (50% majority-rule) trees were estimated using PAUP*.

![Fig. 2. Shell height and width, as measured for this study. The shell axis is parallel to the height and perpendicular to the width.](image-url)
BHOSALE A.R. et al., Varadia, a new semi-slug genus from India’s Western Ghats

Table 1 (continued on next two pages). Taxa used in the DNA analyses with locality data, GenBank accession numbers and source. Subfamily-level placements are shown only for helicarionoidean genera. Superfamily-level classification follows Bouchet et al. (2017). Genus- and species-level names follow Wade et al. (2006), Raheem et al. (2014) and Bank & Neubert (2017).

| Taxa | Sources for family- and subfamily-level placement | Locality | DNA sequence data GenBank acc. nos | Source |
|------|---------------------------------------------------|----------|-----------------------------------|--------|
| **INGROUP** | | | | |
| **LIMACOIDEA** | | | | |
| Agriolimacidae | | | | |
| 1. Deroceras reticulatum (Müller, 1774) | Bouchet et al. (2017) | Kirkdale, Derbyshire, UK | AY014118, AY014119 | Wade et al. (2001) |
| Vitrinidae | | | | |
| 2. Vitrina pellucida (Müller, 1774) | Bouchet et al. (2017) | Kirkdale, Derbyshire, UK | AY014113 | Wade et al. (2001) |
| 3. Azorivitrina laxata (Morelet, 1860) | Bank & Neubert (2017) | São Miguel, Azores | AY014112 | Wade et al. (2001) |
| **GASTRODONTOIDEA** | | | | |
| Oxychilidae | | | | |
| 4. Oxychilus alliarius (Miller, 1822) | Bouchet et al. (2017) | Deepdale, Derbyshire, UK | AY014114 | Wade et al. (2001) |
| 5. Oxychilus cellarius (Müller, 1774) | Bouchet et al. (2017) | Co. Kerry, Ireland | AY014116 | Wade et al. (2001) |
| 6. Oxychilus navarricus helveticus (Blum, 1881) | Bouchet et al. (2017) | Kirkdale, Derbyshire, UK | AY014115 | Wade et al. (2001) |
| Pristilomatidae | | | | |
| 7. Vitrea crystallina (Müller, 1774) | Bouchet et al. (2017) | New Forest, Hampshire, UK | AY014113 | Wade et al. (2001) |
| **PARMACELLOIDEA** | | | | |
| Milacidae | | | | |
| 8. Tandonia budapestensis (Hazay, 1881) | Bank & Neubert (2017) | Kirkdale, Derbyshire, UK | AY014117 | Wade et al. (2001) |
| **TROCHOMORPHOIDEA** | | | | |
| Trochomorphidae | | | | |
| 9. Trochomorpha pallens Pease, 1870 | Bouchet et al. (2017) | Faatoai Valley, Moorea | AY014109, AY014110 | Wade et al. (2001) |
| 10. Videna gouldiana (Pilsbry, 1901) | Hyman & Ponder (2010) | Amami Island, Ryukyu, Japan | AY841313, AY841314 | Wade et al. (2006) |
| Dyakiidae | | | | |
| 11. Asperitas inquinata (v.d. Busch, 1842) | Hausdorf (1995) | Java | AY014108 | Wade et al. (2001) |
Table 1 (continued). Taxa used in the DNA analyses with locality data, GenBank accession numbers and source. Subfamily-level placements are shown only for helicarionoidean genera. Superfamily-level classification follows Bouchet et al. (2017). Genus- and species-level names follow Wade et al. (2006), Raheem et al. (2014) and Bank & Neubert (2017).

| Taxa | Sources for family- and subfamily-level placement | Locality | DNA sequence data | Source |
|------|---------------------------------------------------|----------|-------------------|--------|
|      |                                                   | GenBank acc. nos |                   |        |
| 12. Craterodiscus pricei McMichael, 1959 | Wade et al. (2006) | Ravenshoe, NE Queensland, Australia | AY014123 | Wade et al. (2001) |
| 13. Euconulus fulvus (Müller, 1774) | Hyman & Ponder (2010); Wade et al. (2006) | New Forest, Hampshire, UK | AY014098 | Wade et al. (2001) |
| 14. Louisia barclayi (Benson, 1850) | Hyman & Ponder (2010); Wade et al. (2006) | Mauritius | AY014102 | Wade et al. (2001) |
| 15. Hiona sp. | Hyman & Ponder (2010); Wade et al. (2006) | Moorea | AY014104, AY014105 | Wade et al. (2001) |

HELICARIONOIDEA

Helicarionidae

| Helicarioninae | |
|----------------|-----------------------------|
| 16. Fastosarion brazieri (Cox, 1873) | Bouchet et al. (2017); Hyman & Ponder (2010); Zilch (1959) | Mossman, Queensland, Australia | AY014099 | Wade et al. (2001) |
| 17. Harmogenanina argentea (Reeve, 1852) | Bouchet et al. (2017); Wade et al. (2006); Zilch (1959) | Réunion | AY014101 | Wade et al. (2001) |
| 18. Plegma caelatura (Férussac, 1821) | Bouchet et al. (2017); Wade et al. (2006); Zilch (1959) | Réunion | AY014103 | Wade et al. (2001) |
| 19. Eurychlamys platychlamys (Blanford, 1880) | Blanford & Godwin-Austen (1908) | Sagar Upavan, Mumbai, Maharashtra, India | MW583029 | This study |
| 20. Mariaella dussumieri Gray, 1855 | Blanford & Godwin-Austen (1908) | Ramling Temple, Kolhapur, Maharashtra, India | MW583030 | This study |

Durgellinae

| 21. Satiella sp. | Bouchet et al. (2017) | Jawali, Kolhapur, Maharashtra, India | MW583028 | This study |
Table 1 (continued). Taxa used in the DNA analyses with locality data, GenBank accession numbers and source. Subfamily-level placements are shown only for helicarionoidean genera. Superfamily-level classification follows Bouchet et al. (2017). Genus- and species-level names follow Wade et al. (2006), Raheem et al. (2014) and Bank & Neubert (2017).

| Taxa                     | Sources for family- and subfamily-level placement | Locality                        | DNA sequence data | Source                      |
|-------------------------|---------------------------------------------------|----------------------------------|-------------------|-----------------------------|
| **Ariophantidae**       |                                                   |                                  |                   |                             |
| Ariophantinae           |                                                   |                                  |                   |                             |
| 22. *Ariophanta belangeri* (Deshayes, 1834) | Bouchet et al. (2017)                         | Kagal, Kolhapur, Maharashtra, India | MW583023         | This study                  |
| 23. *Ariophanta intumescens* (Blanford, 1866) | Bouchet et al. (2017)                         | Mhalunge, Kolhapur, Maharashtra, India | MW583024         | This study                  |
| 24. *Ariophanta bistrialis* (Beck, 1837) | Bouchet et al. (2017)                         | Sri Lanka                       | AY014106, AY014107 | Wade et al. (2001)          |
| 25. *Euplecta gardeneri* (Pfeiffer, 1846) | Bouchet et al. (2017); Blanford & Godwin-Austen (1908) | Sri Lanka                       | AY841310, AY841311 | Wade et al. (2006)          |
| 26. *Ratnadvipia* sp. | Bouchet et al. (2017); Blanford & Godwin-Austen (1908) | Sri Lanka                       | AY841312         | Wade et al. (2006)          |
| **Macrochlamydinae**    |                                                   |                                  |                   |                             |
| 27. *Macrochlamys indica* Benson in Godwin-Austen, 1883 | Bouchet et al. (2017)                         | Shivaji University, Kolhapur, Maharashtra, India | MW583025         | This study                  |
| 28. *Macrochlamys pedina* (Benson, 1865) | Bouchet et al. (2017)                         | Khandala, Maharashtra, India     | MW583026         | This study                  |
| 29. *Varadia amboliensis* gen. et sp. nov. | this study                                       | Amboli, Maharashtra, India      | MW583027         | This study                  |
| **Urocyclidae**         |                                                   |                                  |                   |                             |
| 30. *Rhysotina hepaztion* (Gould, 1848) | Bouchet et al. (2017)                         | São Tomé                         | AY014100         | Wade et al. (2001)          |
| **OUTGROUP**            |                                                   |                                  |                   |                             |
| **ARIONOIDEA**          |                                                   |                                  |                   |                             |
| **Arionidae**           |                                                   |                                  |                   |                             |
| 31. *Arion hortensis* Férussac, 1819 | Bouchet et al. (2017)                         | Kirkdale, Derbyshire, UK         | AY014143         | Wade et al. (2006)          |
| **Philomyidae**         |                                                   |                                  |                   |                             |
| 32. *Philomyrus carolinanus* (Bosc, 1802) | Bouchet et al. (2017)                         | Wake Co., USA                    | AY841349         | Wade et al. (2006)          |
| 33. *Meghimatium bilineatum* (Benson, 1842) | Tsai et al. (2011); Saadi & Wade (2019)       | Mauritius                        | AY841348         | Wade et al. (2006)          |
Results

Systematic descriptions

Phylum Mollusca Linnaeus, 1758
Class Gastropoda Cuvier, 1795
Subclass Heterobranchia sensu Bouchet et al., 2017
Order Stylommatophora sensu Bouchet et al., 2017
Superfamily Helicarionoidea Bourguignat, 1877
Family Ariophantidae Godwin-Austen, 1888
Subfamily Macrochlamydinae Godwin-Austen, 1888

Varadia Bhosale & Raheem gen. nov.
urn:lsid:zoobank.org:act:BB363A8F-6796-4874-BBF3-6CE1A207A3C3

Type species

Varadia amboliensis Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. (here designated).

Diagnosis

Only known large semi-slug species from the Western Ghats having broad shell lobes (covering nearly all of the dorsal shell surface when fully extended) and a depressed, discoid shell with a large body whorl. Shell thin, glossy golden brown, with 4–4½ rapidly increasing whorls and barely raised spire. Extensive, largely free penial sheath enclosing substantial part of male genitalia. When sheath is in situ and penis in relaxed state, male genitalia constitute a single elongate, irregularly-shaped mass, with all of penis and epiphallial caecum, most of epiphallus and part of flagellum enclosed by sheath; only vas deferens and most of flagellum is visible outside intact sheath. Penial sheath divisible into thick proximal part and thin distal part, with transition between these two parts occurring between distal penis and epiphallial caecum. With penial sheath removed, following evident: region extending from penis to epiphallial caecum long and cylindrical, with distinctive S-shaped bend in penis and associated band of muscle running along length of penis; proximal ¾ of epiphallus held in long, conspicuous loop; and penial retractor muscle with two branches, one inserting subterminally on epiphallial caecum and one inserting on apex of loop of epiphallus. Inner wall of penis divisible into three regions: proximal penis (one major and several minor longitudinal pilasters), mid-penis (several minor longitudinal pilasters) and distal penis (a few minor pilasters with associated regular transverse ridges in interspaces). Inner wall of epiphallial caecum shows one long major longitudinal pilaster, a large mass of reticulate ridges proximally and several minor longitudinal pilasters distally. Gametolytic gland elongated and long; gametolytic sac 3–3.5 times as long as gametolytic duct. Amatorial organ absent. Spermatophore consists of elongated, soft capsule and long tail-pipe, with U-shaped bend at capsule–tail-pipe junction and funnel-like opening at tip of tail-pipe; surface smooth apart from four ribs running obliquely along length of tail-pipe and short spines near end of tail-pipe.

Etymology

The new genus is named in honour of the herpetologist Dr Varad Giri, who has made a major contribution to the modern study and conservation of the Indian herpetofauna; masculine.

Varadia amboliensis Bhosale, Thackeray, Muley & Raheem gen. et sp. nov.
urn:lsid:zoobank.org:act:5C93F719-2DEF-4A9A-8973-06B7FE6E6528
Figs 3–12

Diagnosis

As genus-level diagnosis.
Fig. 3. Apertural, lateral, apical and umbilical views of the shell of *Varadia amboliensis* Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. A–D. Holotype (BNHS GAS 113). E–H. Paratype (BNHS GAS 116).
Etymology
Named after the type locality, Amboli, in the Sindhudurg District of southern Maharashtra, India. In recent years, Amboli has emerged as a hotspot for the discovery of new species (particularly reptiles and amphibians) in the northern Western Ghats.

Type material

Holotype
INDIA • Maharashtra State, Sindhudurg District, Amboli, Hiranyakeshi temple; 15°57′17.8″ N, 74°01′39.1″ E; 839 m a.s.l.; 2019; A. Bhosale leg.; BNHS GAS 113.

Paratypes
INDIA • 21 specimens (17 whole preserved specimens and 4 shells); same locality data as for holotype; 2019; A. Bhosale leg.; BNHS GAS 114–127, ZSI Moll/1820–1826 • 3 preserved specimens; same locality data as for holotype; 2020; A. Bhosale leg.; BNHS GAS 136–138.

Other material examined
INDIA – Maharashtra State • 1 specimen (sampled for DNA analysis); Sindhudurg District, Amboli Forest Park; 15°57′37.4″ N, 73°59′58.1″ E; 724 m a.s.l.; 2017; A. Bhosale leg.; BNHS GAS 129 • 9 preserved specimens; Sindhudurg District, near Amboli waterfall; 15°56′26.9″ N, 73°59′41.2″ E; 645 m a.s.l.; 2020; A. Bhosale leg.; BNHS GAS 130–135, BNHS GAS 139–141 • 1 shell; Kolhapur District, Kodali; 15°46′42.4″ N, 74°10′40.0″ E; 620 m a.s.l.; 2019; A. Bhosale leg.; BNHS GAS 128.

Description

Shell. Adult shell thin, depressed, glossy and appearing non-umbilicate, with ca 4–4.5 rapidly increasing whorls and colour ranging from golden-brown to reddish yellow (Fig. 3). Shell measurements (n = 35): width 20.2–26.3 mm; height 10.2–15.0 mm. Spire only slightly raised with flat apex and suture only slightly impressed. Body whorl disproportionately large, rounded at periphery, gently convex beneath. Aperture large, crescent-shaped, with width greater than height (Fig. 3A, E). Apertural margin simple, thin and delicate; in lateral view angled forward, with upper apertural margin noticeably anterior to the lower margin. When shell is viewed from below, basal margin curved (not straight) and expanded columellar margin reflected over, covering umbilical region. Shell surface smooth and glossy to naked eye, with irregular, faint collabral striae; under SEM, seen to be finely and closely sculptured with well-defined spiral lines on protoconch (first 1.5–2 whorls) and indistinct and irregular oblique lines on teleoconch (Fig. 4).

Body and mantle. Total adult body length, excluding extended tentacles, ranges from 4.8 to 6.9 cm (n = 5). Living snail glossy grey or greyish white with irregular dark mottling; head and tail dark grey or blackish with tentacles tending to be paler at their tips (Fig. 5–6). Surface of mantle densely and conspicuously covered by small, irregular tubercles that appear lighter on top. Sole tripartite with well-defined sole furrows dividing it into three distinct longitudinal tracts; central tract paler than lateral ones. Tail with large slit-like caudal pit (sensu Hausdorf 1998: 51); caudal horn prominent when extended (Fig. 6A) but when retracted gives tail truncated and blunt appearance (Fig. 6B). Mantle consists of two broad shell lobes (right and left) and two dorsal lobes (right and left) (Fig. 7). Shell lobes may cover nearly all of dorsal surface of shell (Fig. 5), but individuals have also been observed with shell lobes largely retracted and much of shell exposed (Fig. 6). Left dorsal lobe extends as far as base of tentacles when snail is resting (i.e., body not fully extended) and tentacles are retracted.

Reproductive system. Male genitalia consist of proximally penis and distally epiphallic caecum (= epiphallic retractor caecum of Hyman & Ponder 2010: 139) and epiphallus. These three regions are
**Fig. 4.** Shell sculpture of a paratype (BNHS GAS 119) of *Varadia amboliensis* Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. **A.** Protoconch showing irregular collabral lines; black arrow indicates the approximate point of transition from the protoconch to the teleoconch. **B–C.** Close longitudinal striae of protoconch (B) and teleoconch (C). The direction of the longitudinal striae and the faint collabral striae (barely evident) are shown, respectively, by the white and black arrows in C.

**Fig. 5.** Live individual of *Varadia amboliensis* Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. from the site near Amboli waterfall. Image: O. Yadav.
held together by a penial sheath (= penial tunica of Hausdorf 1998), which is largely independent of wall of penis, epiphallic caecum and epiphallus (Figs 8A, 9A–C). Penial sheath encloses all of penis, all of epiphallic caecum, much of epiphallus and part of flagellum; it holds proximal three quarters of epiphallus in a loop against epiphallic caecum, with remaining part of epiphallus (i.e., part closest to vas deferens and flagellum) lying outside sheath, along with a substantial part of flagellum (Fig. 9A). Proximal part of penial sheath is thick and covers penis; distal half of this sheath is thin and covers epiphallus and epiphallic caecum. Thick penial sheath attached to proximal end of penis, close to genital atrium. Thin penial sheath attached to distal end of epiphallic caecum and is open where penial retractor muscle inserts on epiphallic caecum (Fig. 9A–C); an extension of the thin penial sheath also encloses a sizeable section of flagellum (this section located about halfway along length of flagellum). Epiphallus passes through and is attached to penial sheath in region where thick penial sheath transitions into thin penial sheath.

With penial sheath dissected open, penis seen to have noticeable S-shaped bend midway; this bend is associated with a band of muscle that extends for some distance along penis, on either side of bend (Fig. 9D–E). Distally, penis branches into wider-lumened epiphallic caecum and narrower-lumened epiphallus. Epiphallus passes into much narrower-lumened vas deferens; junction between these two regions marked by long, bluntly pointed flagellum, which is similar in length to epiphallus. Penial retractor muscle, which originates on inner lung wall, inserts in two places (Fig. 9E): subterminally on epiphallic caecum, and on apex of loop of epiphallus (i.e., about three quarters of distance from vas deferens to penis). Junction between two branches of penial retractor muscle located near most distal part of epiphallic caecum. Irregular small holes/pores visible on inner surface of thin part of penial sheath (i.e., with sheath cut open and pinned out).

On the basis of the morphology of its inner wall, penis divisible into three morphologically distinct regions, proximal penis, mid-penis and distal penis, with S-shaped bend of penis including all of mid- and

Fig. 6. Live individuals of *Varadia amboliensis* Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. from Hiranyakeshi Temple, Amboli, showing shell lobes in retracted state. A. Right lateral view. B. Left lateral view. Note that the caudal horn is extended in A and fully retracted in B.
distal penial regions (Fig. 10). Proximal penis shows one major and several minor longitudinal pilasters; close study at low magnification (4×) of holotype and one paratype (BNHS GAS 114) showed that pilasters are interspersed by fine, obliquely longitudinal ridges that are close and irregular. Mid-penis ornamented by several thin longitudinal pilasters. Distal penis also with thin longitudinal pilasters, but here they are fewer in number and are contiguous with uniform, widely spaced transverse ridges that

---

**Fig. 7.** Arrangement of the mantle in *Varadia amboliensis* Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. A. Left lateral view. B. Dorsal view. C. Right lateral view. The shell is shaded in grey. Abbreviations: ldl = left dorsal lobe; rdl = right dorsal lobe; lsl = left shell lobe; rsl = right shell lobe.
extend outwards on either side of each pilaster. Opening of epiphallus into most proximal part of epiphallic caecum clearly visible (Fig. 10). Inner wall of epiphallic caecum (Fig. 10) has one major longitudinal pilaster (surface marked by irregular, fine longitudinal and/or transverse ridges; not shown in Fig. 10) running along its length; a large, reticulate mass of ridges proximally; and several short longitudinal pilasters distally. The short pilasters tend to be crenulated proximally and are smoother distally. Lumen of vas deferens widens with increasing distance from epiphallus, with part of vas deferens nearest to epiphallus being noticeably narrower-lumened than remaining two thirds (Fig. 8A). Right eye retractor

**Fig. 8.** Reproductive system of the holotype (BNHS GAS 113) of *Varadia amboliensis* Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. **A.** Gross morphology. **B.** Dissection of capsular gland, showing morphology of inner wall. Abbreviations: ag = albumen gland; cg = capsular gland; ec = epiphallic caecum; ep = epiphallus; esh = extension of penial sheath around epiphallus; fl = flagellum; ga = genital atrium; gd = gametolytic duct; gs = gametolytic sac; hd = hermaphrodite duct; ir = irregularly-marked papillate ridges; lw = lung wall; ot = ovate testes; ov = oviduct; pe = penis; pg = prostate gland; pr = penial retractor muscle; thsh = thick penial sheath; tsh = thin penial sheath; so = spermoviduct; va = vagina; vd = vas deferens.
muscle passes between male and female genitalia. Amatorial organ absent. Genital atrium cylindrical, well defined but short, with junction between male genitalia and vagina located at a short distance from genital orifice. Vagina cylindrical and shorter in length than genital atrium (Fig. 8A). Proximal part of oviduct, near junction with gametolytic gland, consists of pale yellowish, indistinctly-defined region, which is most likely the capsular gland (see Dasen 1933); inner wall of this gland irregularly marked by papillate ridges and papillae (Fig. 8). Gametolytic gland (Fig. 8A) comprises narrow gametolytic duct and long, voluminous sac that is 3–3.5 times length of duct; duct noticeably constricted at its junction with sac and has 1–3 longitudinal ridges on its inner wall.

**Fig. 9.** Male genitalia of the holotype (A, D–E; BNHS GAS 113) and one of the paratypes (B–C; BNHS GAS 116) of *Varadia amboliensis* Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. A. After dissecting out from the body and with penial sheath in situ. B–C. With extension of penial sheath around epiphallus dissected open; dorsal (B) and lateral (C) views are shown. D–E. With penial sheath cut open; dorsal (D) and lateral (E) views are shown. Thick part of penial sheath shown by bold outline and thin part by dotted outline. Abbreviations: bm = band of muscle; ec = epiphallic caecum; ep = epiphallus; esh = extension of penial sheath around epiphallus; fl = flagellum; ga = genital atrium; gd = gametolytic duct; pe = penis; pr = penial retractor muscle; thsh = thick penial sheath; tsh = thin penial sheath; vd = vas deferens.
Fig. 10. Inner surface of penis in the holotype (BNHS GAS 113) of *Varadia amboliensis* Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. Note that the short longitudinal pilasters of the epiphallic caecum are crenulated proximally. The close, fine longitudinal ridges between the longitudinal pilasters of the proximal penis are not illustrated.
Fig. 11. Spermatophore of *Varadia amboliensis* Bhosale, Thakeray, Muley & Raheem gen. et sp. nov.  
A. Fragments of two spermatophores (part of the tail-pipe is missing in both), as found in situ in the gametolytic sac of the paratype BNHS GAS 125.  
B–D. Morphology of the fully-intact spermatophore found in the gametolytic sac of the paratype ZSI Moll/1820.  
B. The upper arrow indicates the U-shaped bend at the junction of the capsule and tail-pipe, and the lower arrow shows the funnel-like opening at the tip of the tail-pipe. Also shown is the cross section of the spermatophore (indicated by the dotted line) close to the tip of the tail-pipe; note the hollow central area.  
C. Detail of a small section of the tail-pipe (indicated by rectangle in B), showing one of the four longitudinal ribs and the hair-like spines on the spermatophore surface.  
D. Detail of tip of tail-pipe, showing the funnel-like opening that leads into the hollow central area (shown by arrow). Note the hair-like spines.  
Abbreviations: cap = capsule; gd = gametolytic duct; gs = gametolytic sac; o = opening at tip of tail-pipe; sp = spermatophore; tp = tail-pipe.
One or two spermatophores (i.e., only one wholly intact; the rest damaged/partially digested) present in gametolytic gland of each of six specimens (Fig. 11A). Intact spermatophore consists of elongated, soft capsule with long tail-pipe. Sharply-angled, U-shaped bend at junction of capsule and tail-pipe; apex of bend noticeably hooked towards tail-pipe. Capsule wider-lumened than tail-pipe and twisted spirally (Fig. 11B). Tail-pipe flexible, internally hollow and externally sculptured obliquely along its length with four fine ribs. Tail-pipe in vicinity of tip hollow centrally and this passes into funnel-like opening (perforation) (Fig. 11D); surface of spermatophore near tip of tail-pipe has short, hair-like spines that point towards capsule (Fig. 11C).

**Radula and Jaw.** Central tooth tricuspid, with large mesocone, which is shorter than tooth base, and smaller, more basal ectocones (Fig. 12A–B). Inner laterals 17–21, uniformly tricuspid (Fig. 12A–B); mesocone large, equal in size to those of central tooth and shorter than tooth base, endocone barely defined and ectocone prominent but more basal than other cusps. Outer 2 lateral teeth grade into marginal teeth. Marginal teeth 45–53, uniformly bicuspid (endocone absent), with shorter, narrower and more basal ectocone (Fig. 12C–D). Formulae for the 8 specimens examined are as follows (the plus sign indicates that the outermost marginal teeth could not be counted):

- Holotype BNHS GAS 113  (+50.20.1.18.2.50+)
- Paratype BNHS GAS 114  (+49.21.1.19.2.49+)
- Paratype BNHS GAS 115  (53.19.1.17.2.53)
- Paratype BNHS GAS 116  (+45.21.1.19.2.45+)
- Paratype BNHS GAS 117  (+50.19.1.17.2.50+)
- Paratype BNHS GAS 118  (+48.23.1.21.2.48+)
- Paratype BNHS GAS 119  (+48.19.1.17.2.48+)
- Paratype BNHS GAS 120  (52.22.1.20.2.52)

Jaw oxygnath (smooth), having a concave cutting edge with well-defined or barely evident median projection (Fig. 12E).

**Distribution and ecology**

*Varadia amboliensis* gen. et sp. nov. is endemic to the northern and central Western Ghats of India and is currently known from only 5 localities. These are: Hiranyakeshi temple, Amboli, Sindhudurg District, Maharashtra State (15°57′17.8″ N, 74°01′39.1″ E; 839 m a.s.l.); Amboli Forest Park, Sindhudurg District, Maharashtra State (15°57′37.4″ N, 73°59′58.1″ E; 724 m a.s.l.); near Amboli waterfall, Sindhudurg District, Maharashtra State (15°56′26.9″ N, 73°59′41.2″ E; 645 m a.s.l.); Kodali, Kolhapur District, Maharashtra State (15°46′42.4″N, 74°10′40.0″E; 620 m a.s.l.); Yana Forest, Uttara Kannada District, Karnataka State (14°35′16.4″N 74°34′00.3″E; 272 m a.s.l.) (A. Bhosale, 2018, personal observation). The species occurs at elevations ranging from 272 to 839 m. Although it has been observed among human habitation on forest edges (Fig. 13), *V. amboliensis* gen. et sp. nov. appears to be primarily a species of tropical semi-evergreen and evergreen forest (sensu vegetation classification of Pascal 1991). The range of this species, as currently known, is restricted and disjunct. While 4 of the 5 known localities are in the extreme south of Maharashtra State (northern Western Ghats), the only other known locality, Yana Forest in northern Karnataka (central Western Ghats), is ca 160 km to the south. Further surveys are required to establish if this species occurs in the intervening area.

*Varadia amboliensis* gen. et sp. nov. is primarily a ground-living snail. It can be encountered at night in leaf litter or on rocks and the bases of trees; in rainy weather it can be seen on the exterior walls of buildings close to the forest edge (e.g., it was observed at the entrance of Amboli Forest Park in September 2017). The species can be seen throughout the monsoon (June to October) and as late as the end of November.
A few individuals have been seen in late February (late winter) on the banks of fast-flowing streams at Amboli.

This species appears to be omnivorous. It has been observed feeding on decaying plant matter (leaf litter, discarded banana peel) and on the remains of at least two different invertebrate taxa (a cricket and an earthworm) (A. Bhosale, personal observation) (Supp. file 1). Data on its predators are scarce,

![Fig. 12. Radula and jaw of a paratype (BNHS GAS 127) of Varadia amboliensis Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. A. Central (indicated by arrows), inner lateral and outer lateral teeth. B. Central and inner lateral teeth. C–D. Marginal teeth. E. Jaw. Note median projection.](image-url)
but a scorpion of the genus *Heterometrus* Ehrenberg, 1828 (Scorpionidae) was observed feeding on an individual of this species (Supp. file 1).

**DNA analysis**

The three phylogenetic analyses (NJ, ML and Bayesian) yielded broadly similar results (Fig. 14), with disagreements occurring only for internal branches lacking strong support in any of the analyses. Most deeper relationships within the Limacoidea were not strongly supported, with the optimal ML and NJ trees having fewer strongly supported branches than the Bayesian tree. All three trees included a sister-

---

**Fig. 13.** Habitat of *Varadia amboliensis* Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. A. Hiranyakeshi temple. B–C. Amboli Forest Park; snails were observed on the walls of the temple and other structures and in the surrounding forest. D–E. Amboli waterfall; snails were observed on the wall near the sunshade (D) and on the roadside safety barrier (E). F. Yana Forest, where snails were found on the forest floor, close to the footpath.
Fig. 14. Bayesian majority rule consensus tree of the rRNA gene cluster dataset (874 bp) for the Limacoidea. Ingroup taxa are colour-coded by their family-level placements, with subfamily-level placements being shown only for Ariophantidae (see Table 1). The tree is rooted with an outgroup consisting of three members of the Arionoidea (Arion hortensis Férussac, 1819, Meghimatium bilineatum (Benson, 1842) and Philomycus carolinianus (Bosc, 1802)). Clades A and B are indicated by arrows. Branch support values are shown in the following sequence: Bayesian posterior probabilities, ML bootstraps and NJ bootstraps. Symbols: ‘*’ = maximal support (Bayesian = 1; NJ, ML = 100%); ‘-’ = differences in branching pattern between Bayesian and NJ trees. Scale bar indicates substitutions per site.
group relationship between Varadia gen. nov. and the always maximally supported Macrochlamys clade (Macrochlamys indica Benson, 1883 + M. pedina (Benson, 1865)), but this was not strongly supported in any of the analyses (Bayesian: PP = 0.62; ML: BS = 60%; NJ: BS = 44%). All analyses also provided maximal support for the clade composed of five of the six helicarionid taxa, Fastosarion brazieri (Cox, 1873), Eurychlamys platychlamys (Blanford, 1880), Harmogenanina argentea (Reeve, 1852), Plegma caelatura (Férussac, 1821) and Satiella sp. (clade B in Fig. 14). Within clade B, support for the sister-group relationship between Fastosarion Iredale, 1933 and Eurychlamys was consistently strong (ML: BS = 90%; NJ: BS = 96%) or maximal (Bayesian). Two of the three analyses (ML: BS = 71%; Bayesian: PP = 0.98) provided strong support for clade A, comprising clade B, Rhysotina hepatzion (Gould, 1848), the Macrochlamys clade, Varadia gen. nov. and Mariaella dussumieri Gray, 1855. All analyses provided maximal support for the monophyly of the Trochomorphidae Möllendorff, 1890, of the Dyakiidae Gude & B.B. Woodward, 1921 and of the Euconulidae H.B. Baker, 1928 (i.e., clade comprising Euconulus fulvus (Müller, 1774) and Louisia barclayi (Benson, 1850)).

Examination of all bipartition frequencies for the ML bootstrap trees (n = 1008) showed that the best supported bipartition that is not compatible with Varadia gen. nov. forming a clade with the two species of Macrochlamys is one in which Macrochlamys forms a clade with the five helicarionids, Fastosarion brazieri, Eurychlamys platychlamys, Harmogenanina argentea, Plegma caelatura and Satiella sp. (BS = 12%). Similarly, for the NJ bootstrap trees (n = 1000), the best supported bipartition that is incompatible with the clade of Varadia gen. nov. + Macrochlamys is the clade uniting all the helicarionids and Rhysotina hepatzion with Macrochlamys (BS = 14%).

**Discussion**

Two of the three phylogenetic analyses (ML and Bayesian) showed strong support for clade A (consisting of clade B, Rhysotina hepatzion, the Macrochlamys clade, Varadia gen. nov. and Mariaella dussumieri). We note, however, that while this clade is strongly supported (BS = 71%) in the ML tree it is only very weakly supported in the NJ tree (BS = 30%), so its robustness requires substantial further investigation (e.g., by expanded sampling of taxa and/or gene regions). The five members of the Ariophantinae sensu stricto (Blanford & Godwin-Austen 1908; an almost exclusively Peninsular Indian–Sri Lankan group) lie outside clade A. Although this suggests that Varadia gen. nov. is not a member of the Ariophantinae sensu stricto, the position of Varadia gen. nov. within clade A was not compellingly resolved, and that includes uncertainty about the sister group of Varadia gen. nov.

On the basis of characters of the reproductive system, particularly the male genitalia and the gametolytic sac, we provisionally place Varadia gen. nov. in the Macrochlamydinae. This is consistent with the results of our DNA analyses and there are two key points here. First, although the sister-group relationship of Varadia gen. nov. to Macrochlamys was not strongly supported (Bayesian: PP = 0.62; ML: BS = 60%; NJ: BS = 44%), it was consistently recovered by all analyses, and is much better supported than the best supported incompatible bipartition in both the ML (BS = 12%) and NJ (BS = 14%) analyses, though, taxon sampling could be an issue here, and further investigations are required to address this. Second, in two of the three analyses, Varadia gen. nov. was strongly supported (ML: BS = 71; Bayesian: PP = 0.98) as a member of a clade uniting the helicarionids with the Macrochlamys clade and the urocyclid Rhysotina hepatzion. In V. amboliensis gen. et sp. nov., as in many Macrochlamydinae, the penial retractor muscle inserts not on the tip or distal-most end of the epiphallus caecum but on the side. Hausdorf (1998) considered the lateral insertion of the penial retractor muscle on the epiphallus caecum to be the only known autapomorphy of the Macrochlamydinae; we note, however, that in the Indian species
Macrochlamys pedina and in several Thai members of the Macrochlamydinae (e.g., Sarika Godwin-Austen, 1907 and some Taphrenalla Pholyotha & Panha, 2020), the penial retractor muscle inserts on the tip of the epiphallic caecum (Blanford & Godwin-Austen 1908; Pholyotha et al. 2020a, 2020b).

While the form of the epiphallic caecum in South Asian Macrochlamydinae is varied, the most widespread form is a distinctly coiled mass that is enclosed in a thin and translucent outer covering (e.g., Macrochlamys, Euaustenia Cockerell, 1891 and Bensonies H.B. Baker, 1938; Blanford & Godwin-Austen 1908: 77–141, 148–152, 172–177). The epiphallic caecum may also be a loose, open coil (M. richilensis Godwin-Austen, 1907, M. zemoensis Godwin-Austen, 1907; Blanford & Godwin-Austen 1908: 90–93), an irregular mass (M. castaneolabiata Godwin-Austen, 1883; Blanford & Godwin-Austen 1908: 101–102, 105) or straight and cylindrical (e.g., M. pedina; Blanford & Godwin-Austen 1908: 132–133). It is this last form of epiphallic caecum that occurs in V. amboliensis gen. et sp. nov. The new genus shares three other characters with some genera in the Macrochlamydinae (e.g., Macrochlamys and Bensonies): a well-developed epiphallus, a well-developed flagellum and a long, elongated gametolytic gland with a long stalk (cf. Blanford & Godwin-Austen 1908: 77–179).

In external morphology and in the form of the radula tooth V. amboliensis gen. et sp. nov. is similar to other Macrochlamydinae, as well as to some helicarionids. The broad shell lobes found in V. amboliensis gen. et sp. nov. also occur in other Indian Macrochlamydinae, such as Parvatella Blanford & Godwin-Austen, 1908 and Euaustenia from northern India, as well as in Indian helicarionids, such as Satiella (Blanford & Godwin-Austen 1908; Bhosale et al. 2019). We suggest that the expansion/retraction of the shell lobes in V. amboliensis gen. et sp. nov. and in other taxa (e.g., Eurychlamys platychlamys) may be related to air humidity, with the shell being largely covered by the mantle during rainy weather and the mantle lobes being largely retracted during drier conditions. In the new species the tripartite structure of the sole is clearly evident, the furrows demarcating the three sole fields being well defined. This condition is characteristic of both the Macrochlamydinae and the Helicarionidae; in the Ariophantinae sensu stricto, the tripartite structure is not obvious because the sole furrows are indistinct (Hausdorf 1998; Hyman & Ponder 2010; see also Blanford & Godwin-Austen 1908).

In the radula of V. amboliensis gen. et sp. nov., the central tooth has a large mesocone and two small ectocones; the lateral teeth have a large mesocone, a small/minute endocone and a larger, more basal ectocone; and the marginal teeth are bicuspud (endocone absent with no subdivision of the ectocone). Some Macrochlamydinae (e.g., Godwin-Austen 1908: M. indica; Hyman & Ponder 2010: M. petrosa (T. Hutton, 1834); Pholyotha et al. 2020a: Taphrenella) and helicarionids, such as Eurychlamys platychlamys (Bhosale et al. 2019) and the Australian genera Helicarion Féroussac, 1821 and Stanisicarion Hyman & Ponder, 2010 (Hyman & Ponder 2010) exhibit similar radular tooth form.

There are several clear differences between V. amboliensis gen. et sp. nov. and Macrochlamys. The amatorial organ (= stimulator of Hausdorf 1998), which is usually but not always present in the Macrochlamydinae (Blanford & Godwin-Austen 1908), is absent in the new genus. In comparison, the amatorial organ is nearly always present in the Ariophantinae sensu stricto, is usually absent in the Helicarionidae (Blanford & Godwin-Austen 1908)—all the Australian taxa lack it (Hyman & Ponder 2010)—and is sometimes absent in the Durgellinae (Blanford & Godwin-Austen 1908; e.g., present in Durgella W.T. Blanford, 1863 but absent in Satiella). We also note that V. amboliensis gen. et sp. nov. and Macrochlamys differ in the morphology of the capsular gland. In Macrochlamys and in the Ariophantinae (Dasen 1933) the capsular gland is a wide-lumened structure that is constricted at both ends and distinctly yellow in colour; in V. amboliensis gen. et sp. nov. it is slightly swollen but is otherwise not well defined. Another difference between the two genera is in the diameter of the vas deferens. This is more-or-less uniform in Macrochlamys; in V. amboliensis gen. et sp. nov., the section of the vas deferens (ca 60–70% of length) nearest to the spermoviduct/prostate gland is markedly wider (i.e., as wide as the epiphallus).
Also, on the basis of available data, the insertion of the penial retractor muscle at two separate points (near the tip of epiphallic caecum and on the epiphallus) and the presence of a thin but noticeable band of muscle on the outer wall of the penis are unique to *Varadia* gen. nov.

Published data on the morphology of the penial sheath and inner wall of the penis are scarce for South Asian helicarionoideans (but see Hyman & Ponder: *M. petrosa*; Raheem & Naggs 2006: *Ratnadvipia* Godwin-Austen, 1899; Bhosale et al. 2019: *Eurychlamys*). While detailed anatomical studies of Thai and Myanma *Macrochlamydinae* have been published recently (e.g., Pholyotha et al. 2018, 2020a, 2020b), these do not include details on the structure of the penial sheath. We compared *M. cf. indica* from Kolkata (formerly Calcutta), West Bengal, India (4 specimens dissected; reg no. BNHS GAS 142 to 145) with *V. amboliensis* gen. et sp. nov., and this revealed some clear differences between the two. In the Kolkata examples of *M. cf. indica*, the penial sheath extends from the most proximal part of the penis to the area where the penis gives way to the coiled epiphallic caecum; it is attached to the penial wall at these two ends but is otherwise free. In *V. amboliensis* gen. et sp. nov., in contrast, the penial sheath extends over the whole of the epiphallic caecum, part of the epiphallus and a small part of the flagellum. Interestingly, our dissections of *M. cf. indica* indicate that like in *V. amboliensis* gen. et sp. nov., the penial sheath is divisible into a proximal section that is thick and opaque, and a distal section that is thin and transparent (to our knowledge, this has not been reported before). These dissections also suggest that the section of the epiphallus nearest to the epiphallic caecum and penis is firmly attached to the penial sheath; again, this is similar to *V. amboliensis* gen. et sp. nov. Further studies of other species of *Macrochlamys* as well of other taxa in the subfamily *Macrochlamydinae* are required to investigate the generality of these observations. Internally, the penis of *M. cf. indica* is divisible into a proximal section that is thick and opaque, and a distal section with a well-developed penial verge (= penial papilla) demarcating the two; a similar arrangement has also been reported for Thai and Myanma *Macrochlamys* (Pholyotha et al. 2018, 2020b). In contrast, in *V. amboliensis* gen. et sp. nov., a penial verge is absent and the penis is divisible into three parts on the basis of the morphology of the inner surface.

The whole spermatophore and spermatophore fragments recovered from *V. amboliensis* gen. et sp. nov. were found in a loosely coiled state in the gametolytic sac. These spermatophores are broadly similar with those described and figured for *M. pedina* and *M. flemingi* (L. Pfeiffer, 1856) by Godwin-Austen (1899: 133, pl. 83: figs 5, 5a–c; pl. 87: figs 2, 2a–d), the capsule being relatively long and elongated in all three species. In *M. pedina* and *M. flemingi*, however, the entire length of the tail-pipe is covered with relatively large, branching spines that point towards the capsule; in *V. amboliensis* gen. et sp. nov., the spines are minute and confined to the free, terminal part of the tail-pipe (cf. the Australian helicarionid genus *Fastosarion*; Hyman & Köhler 2019). Godwin-Austen (1899: 133, 134; pl. 94: 1, 2, 2a, 4) also described and figured the spermatophores of *M. udus* Godwin-Austen, 1899 and *M. lecythis* (Benson, 1852). In these species, the capsule is short relative to the tail-pipe and spines (of varied complexity) are present only at the two ends of the tail-pipe. Spermatophores of broadly similar morphology have also been reported for some Thai *Macrochlamys* species (e.g., *M. aurantia* Pholyotha & Panha, 2018 and *M. coleus* Pholyotha & Panha, 2018; Pholyotha et al. 2018). The spermatophore of *V. amboliensis* gen. et sp. nov., unlike those described to date for *Macrochlamys*, has a pronounced U-shaped bend at the junction of the capsule and tail-pipe. We note that similarly bent spermatophores are characteristic of some urocyclid taxa (Van Goethem 1977: 34–35); these urocyclids, like *V. amboliensis* gen. et sp. nov., also have a perforated tip to the tail-pipe.

In external morphology (the combination of large size, broad, densely tuberculated shell lobes and a shell with ca 4 whorls and a disproportionately large body whorl), *V. amboliensis* gen. et sp. nov. is unique among the helicarionoidean taxa of the Western Ghats. *Eurychlamys platychlamys*, *M. indica* and *M. pedina* are superficially similar, but the first two are smaller (particularly *E. platychlamys*) and all have tightly-wound shells composed of ca 5 whorls (cf. Raheem et al. 2014: fig. 67a–b; Bhosale et al.
In addition, the shell lobes in the two species of *Macrochlamys* are narrow, ribbon-like and highly mobile (Bhosale et al. 2019: fig. 8a; A. Bhosale, unpublished data).

**Acknowledgements**

We are grateful to the Maharashtra Forest Department, Government of India (GoI) for providing the relevant collecting and fieldwork permits. Financial support was provided to AB by the IPLS programme, Department of Biotechnology, GoI (grant no. BT/PR4572/INF/22/147/2012); DST PURSE Phase I and Phase II of the Department of Science and Technology, GoI; and the Systematics Research Fund of the Systematics Association and the Linnean Society of London, UK. Lab facilities were provided by the Department of Zoology, Shivaji University, Kolhapur, India. We would like to thank the following people at Shivaji University: Shantakumar Manne (Head of Department of Zoology), for his help during the study; J.P. Jadhav (Head of Department of Biotechnology), for facilitating this work under a DBT-IPLS project; S.P. Govindwar (former Head of Department of Biochemistry), for giving AB permission to work in the molecular biology lab in his department; and Mansingh Nimbalkar (Department of Botany), for assisting AB with SEM work. Omkar Yadav, Tejas Patil, Rupesh Yadav, Gopal Raut, Pramod Lawand, Rupali Khandekar, Mayur Jadhav and Shriraj Patil assisted with fieldwork. Santosh Saste (DFO), Akshay Khandekar, Swapnil Pawar, Priyanka Patil, Yogesh Mane and Suraj Patil helped us at various stages. We would also like to acknowledge Fred Naggs for helping us in our initial efforts to identify the new genus, Rahul Khot for assisting us with the deposition of specimens at BNHS, P.S. Bhatnagar and S. Talmale for helping us with the deposition of specimens at WRC ZSI, David Gower for feedback on phylogenetic analysis, David Reid for advice on the formation of taxon names, Bernhard Hausdorf and Isabel Hyman for reviewing the manuscript; Thierry Backeljau for valuable editorial feedback; and František Kovářík for identifying the scorpion photographed preying on *Varadia* gen. nov.

**References**

Alfaro M.E. & Holder M.T. 2006. The posterior and the prior in Bayesian phylogenetics. *Annual Review of Ecology, Evolution, and Systematics* 37: 19–42. [https://doi.org/10.1146/annurev.ecolsys.37.091305.110021](https://doi.org/10.1146/annurev.ecolsys.37.091305.110021)

American Veterinary Medical Association. 2020. AVMA guidelines for the euthanasia of animals: 2020 edition. Available from [https://www.avma.org/resources-tools/avma-policies/avma-guidelines-euthanasia-animals](https://www.avma.org/resources-tools/avma-policies/avma-guidelines-euthanasia-animals) [accessed 29 Oct. 2020].

Aravind N.A. & Páll-Gergely B. 2018. *Dicharax (?) bawai* n. sp from southern India (Gastropoda: Cyclophoroidea: Alycaeidae). *Archiv für Molluskenkunde* 147: 55–62. [https://doi.org/10.1127/arch.moll/147/055-062](https://doi.org/10.1127/arch.moll/147/055-062)

Bank R. 2017. MolluscaBase classification of the recent terrestrial Gastropoda of the World. Last update: 16 July 2017. Available from [http://www.molluscabase.org](http://www.molluscabase.org) [accessed 21 Nov. 2018].

Bank R. & Neubert E. 2017. MolluscaBase checklist of the land and freshwater Gastropoda of Europe. Last update: 16 July 2017. Available from [http://www.molluscabase.org](http://www.molluscabase.org) [accessed 21 Nov. 2018].

Bhosale A.R. 2018. *DNA Barcoding and Phylogenetic Study of Freshwater and Land Molluscs from Kolhapur District*. PhD thesis, Shivaji University, Kolhapur, India.

Bhosale A.R., Patil T.S., Yadav R.B. & Muley D.V. 2016. On the occurrence of *Theobaldius(?) tristis* (Blanford, 1869) (Caenogastropoda: Cyclophoridae) in the northern Western Ghats, Maharashtra, India. *Journal of Threatened Taxa* 8: 8934–8937. [https://doi.org/10.11609/jott.2357.8.6.8934-8937](https://doi.org/10.11609/jott.2357.8.6.8934-8937)

Bhosale A.R., Muley D.V., Yadav O.V. & Raheem D.C. 2019. Re-description of the land snail *Eurychlamys platychlamys* (Blanford, 1880) (Pulmonata: Helicarionidae) from the Western Ghats of India. *Archiv für Molluskenkunde* 148: 117–128. [https://doi.org/10.1127/arch.moll/148/117-128](https://doi.org/10.1127/arch.moll/148/117-128)
Bhosale A., Thackeray T. & Rowson B. In press. Perrottetia rajeshgopali, a new species of Perrottetia Kobelt, 1905 (Stylommatophora: Streptaxidae) from the Western Ghats, India. Archiv für Molluskenkunde.

Blanford W.T. & Godwin-Austen H.H. 1908. The Fauna of British India Including Ceylon and Burma. Mollusca. Testacellidae and Zonitidae. Taylor and Francis, London.

Bouchet P., Rocroi J.P., Hausdorf B., Kaim A., Kano Y., Nützel A., Parkhaev P., Schrödl M. & Strong E.E. 2017. Revised classification, nomenclator and typification of gastropod and monoplacophoran families. Malacologia 61: 1–526. https://doi.org/10.4002/040.061.0201

Cameron R. 2016. Slugs and Snails. Collins New Naturalist Library, Book 133. William Collins, UK.

Cox L.R. 1960. General characteristics of Gastropoda. In: Moore C. (ed.) Treatise on Invertebrate Palaeontology Part I, Mollusca 1: 184–1169. Geological Society of America and University of Kansas Press, USA.

Dasen D.D. 1933. Structure and function of the reproductive system in Ariophanta ligulata. Proceedings of the Zoological Society of London 103: 97–118. https://doi.org/10.1111/j.1096-3642.1933.tb01579.x

Felsenstein J. 1981. Evolutionary trees from DNA sequences: a maximum likelihood approach. Journal of Molecular Evolution 17: 368–376. https://doi.org/10.1007/BF01734359

Felsenstein J. 1985. Confidence limits on phylogenies: an approach using the bootstrap. Evolution 39: 783–791. https://doi.org/10.1111/j.1558-5646.1985.tb00420.x

Godwin-Austen H.H. 1882–1888. Land and Freshwater Mollusca of India, Including South Arabia, Baluchistan, Afghanistan, Kashmir, Nepal, Burmah, Pegu, Tenasserim, Malay Peninsula, Ceylon, and Other Islands of the Indian Ocean. Supplementary to Messrs. Theobald and Hanley's Conchologia Indica. Vol. 1. Taylor & Francis, London.

Godwin-Austen H.H. 1897–1920. Land and Freshwater Mollusca of India, Including South Arabia, Baluchistan, Afghanistan, Kashmir, Nepal, Burmah, Pegu, Tenasserim, Malay Peninsula, Ceylon, and Other Islands of the Indian Ocean. Supplementary to Messrs. Theobald and Hanley's Conchologia Indica. Vol. 2. Taylor & Francis, London.

Goodacre S.L. & Wade C.M. 2001. Molecular evolutionary relationships between partulid land snails of the Pacific. Proceedings of the Royal Society B 268: 1–7. https://doi.org/10.1098/rspb.2000.1322

Gu X., Fu Y.X. & Li W.H. 1995. Maximum likelihood estimation of the heterogeneity of substitution rate among nucleotide sites. Molecular Biology and Evolution 12: 546–557. https://doi.org/10.1093/oxfordjournals.molbev.a040235

Hausdorf B. 1995. A preliminary phylogenetic and biogeographic analysis of the Dyakiidae (Gastropoda: Stylommatophora) and a biogeographic analysis of the other Sundaland biota. Cladistics 11: 359–376. https://doi.org/10.1016/0748-3007(95)90021-7

Hausdorf B. 1998. Phylogeny of the Limacoidea sensu lato (Gastropoda: Stylommatophora). Journal of Molluscan Studies 64: 35–66. https://doi.org/10.1093/mollus/64.1.35

Hausdorf B. 2000. Biogeography of the Limacoidea sensu lato (Gastropoda: Stylommatophora): vicariance events and long-distance dispersal. Journal of Biogeography 27: 379–390. https://doi.org/10.1046/j.1365-2699.2000.00403.x

Herbert D.G. & Mitchell A. 2009. Phylogenetic relationships of the enigmatic land snail genus Prestonella: the missing African element in the Gondwanan superfamily Orthalicoidea (Mollusca: Stylommatophora). Biological Journal of the Linnean Society 96: 203–221. https://doi.org/10.1111/j.1095-8312.2008.01109.x

Hillis D.M. & Bull J.J. 1993. An empirical test of bootstrapping as a method for assessing confidence in phylogenetic analysis. Systematic Biology 42: 182–192. https://doi.org/10.1093/sysbio/42.2.182
BHOSALE A.R. et al., Varadia, a new semi-slug genus from India’s Western Ghats

Hyman I.T. & Köhler F. 2019. Phylogeny and systematic revision of the helicarionid semislugs of eastern Queensland (Stylommatophora, Helicarionidae). Contributions to Zoology 88: 351–451. https://doi.org/10.1163/18759866-20191416

Hyman I.T. & Ponder W.F. 2010. A morphological phylogenetic analysis and generic revision of Australian Helicarionidae (Gastropoda: Pulmonata: Stylommatophora), and an assessment of the relationships of the family. Zootaxa 2462: 1–148. https://doi.org/10.1164/zootaxa.2462.1.1

Hyman I.T., Ho S.Y. & Jermin L.S. 2007. Molecular phylogeny of Australian Helicarionidae, Euconulidae and related groups (Gastropoda: Pulmonata: Stylommatophora) based on mitochondrial DNA. Molecular Phylogenetics and Evolution 45: 792–812. https://doi.org/10.1016/j.ympev.2007.08.018

Hyman I.T., de la Iglesia Lamborena I. & Köhler F. 2017. Molecular phylogenetics and systematic revision of the south-eastern Australian Helicarionidae (Gastropoda, Stylommatophora). Contributions to Zoology 86: 51–95. https://doi.org/10.1163/18759866-08601004

Irfan-Ullah M. & Davande S. 2008. Western Ghats Ecogeographic boundary as developed from physiographic and biogeographic data. ATREE. Available from http://indiabiodiversity.org/map?layers=lyr_7_wg_boundary&title=WGBoundary [accessed 21 Nov. 2018].

Kerney M.P. & Cameron R.A.D. 1979. Land Snails of Britain and North-west Europe. Collins, London.

Lanave C., Preparata G., Saccone C. & Serio G. 1984. A new method for calculating evolutionary substitution rates. Journal of Molecular Evolution 20: 86–93. https://doi.org/10.1007/BF02101990

Larget B. & Simon D.L. 1999. Markov chain Monte Carlo algorithms for the Bayesian analysis of phylogenetic trees. Molecular Biology and Evolution 16: 750–759. https://doi.org/10.1093/oxfordjournals.molbev.a026160

Miller M.A., Pfeiffer W. & Schwartz T. 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees. In: Proceedings of the Gateway Computing Environments Workshop (GCE), 14 Nov. 2010, New Orleans, LA: 1–8. https://doi.org/10.1109/GCE.2010.5676129

Naggs F. & Raheem D. 2000. Land snail diversity in Sri Lanka. Natural History Museum, London.

Páll-Gergely B., Hunyadi A., Grego J., Sajan S., Tripathy B. & Chen Z.Y. 2020. A review of the Diapheridae (Gastropoda: Eupulmonata: Streptaxoidea), with special emphasis on India and Myanmar. Raffles Bulletin of Zoology 68: 682–718.

Pascal J.-P. 1991. Floristic composition and distribution of evergreen forests in the Western Ghats, India. Palaeobotanist 39: 110–126.

Pholyotha A., Sutcharit C. & Panha S. 2018. The land snail genus Macrochlamys Gray, 1847 from Thailand, with descriptions of five new species (Pulmonata: Ariophantidae). Raffles Bulletin of Zoology 66: 753–781.

Pholyotha A., Sutcharit C., Tongkerd P., Jeraththitikul E. & Panha S. 2020a. Integrative systematics reveals the new land-snail genus Taphrenalla (Eupulmonata: Ariophantidae) with a description of nine new species from Thailand. Contributions to Zoology 1(aop): 1–49. https://doi.org/10.1163/18759866-bja10013

Pholyotha A., Sutcharit C., Tongkerd P., Lin A. & Panha S. 2020b. Taxonomic revision of the land snail genera Macrochlamys Gray, 1847 and Sarika Godwin-Austen, 1907 (Eupulmonata: Ariophantidae) from south-eastern Myanmar, with descriptions of three new species. Molluscan Research 40: 183–204. https://doi.org/10.1080/13235818.2020.1723041
Raheem D. & Naggs F. 2006. The Sri Lankan endemic semi-slug *Ratnadvipia* (Limacoidea: Ariophantidae) and a new species from southwestern Sri Lanka. *Systematics and Biodiversity* 4: 99–126. https://doi.org/10.1017/S1477200005001854

Raheem D.C., Taylor H., Ablett J., Preece R.C., Aravind N.A. & Naggs F. 2014. A systematic revision of the land snails of the Western Ghats of India. *Tropical Natural History Supplement* 4: 1–285. Available from https://ti01.tci-thaijo.org/index.php/tnh/article/view/103091 [accessed 21 Nov. 2018].

Ramírez R., Borda V., Romero P., Ramírez J., Congrains C., Chirinos J., Ramírez P., Velásquez L.E. & Mejia K. 2012. Biodiversity and endemism of the western Amazonia land snails *Megalobulimus* and *Systrophia*. *Revista Peruana de Biología* 19: 59–074. https://doi.org/10.15381/rpb.v19i1.798

Ronquist F. & Huelsenbeck J.P. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572–1574. https://doi.org/10.1093/bioinformatics/btg180

Saadi A. J. & Wade C. M. 2019. Resolving the basal divisions in the stylommatophoran land snails and slugs with special emphasis on the position of the Scolodontidae. *Molecular Phylogenetics and Evolution* 139: 106529. https://doi.org/10.1016/j.ympev.2019.106529

Saitou N. & Nei M. 1987. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution* 4: 406–425. https://doi.org/10.1093/oxfordjournals.molbev.a040454

Schilthuizen M., Clavera A.P., Khoo M.S., Bondar C.A., Elder C.H., Bouma A.M., Eddington T.D., Reisinger C., Cosentino E., Rossato M. & Delledonne M. 2019. Bringing the lab to the field: a new lowland *Microparmarion* semi-slug (Gastropoda: Ariophantidae) described and DNA-barcoded in the forest. *Journal of Molluscan Studies* 85: 35–40. https://doi.org/10.1093/mollus/eyy052

Smith S.W., Overbeek R., Woese C.R., Gilbert W. & Gillevet P.M. 1994. The genetic data environment, an expandable GUI for multiple sequence analysis. *Bioinformatics* 10: 671–675. https://doi.org/10.1093/bioinformatics/10.6.671

Staden R., Beal K.F. & Bonfield J.K. 2000. The Staden package, 1998. *Methods in Molecular Biology* 132: 115–130. https://doi.org/10.1385/1-59259-192-2:115

Stamatakis A. 2014. The RAxML v8.1.X Manual. Heidelberg Institute for Theoretical Studies. Available from http://www.exelixis-lab.org [accessed 21 Nov. 2018].

Swofford D.L. 2002. PAUP*. *Phylogenetic Analysis Using Parsimony (*and Other Methods), Version 4.0b10.* Sinauer Associates, Sunderland, MA.

Tsai C.L., Lu C.C. & Kao H.W. 2011. Morphology and molecular phylogeny of the east and southeast Asian *Meghimatium* slugs (Gastropoda: Pulmonata: Philomyidae) and description of a new species. *Zootaxa* 2890: 1–19. https://doi.org/10.11646/zootaxa.2890.1.1

Van Goethem J.L. 1977. Révision systématique des Urocyclinae (Mollusca, Pulmonata, Urocyclidae). *Musee royal de l’Afrique Centrale, Tervuren, Belgique, Annales, Sciences zoologiques* 218: 1–XI, 1–355.

Wade C.M. & Mordan P.B. 2000. Evolution within the gastropod molluscs; using the ribosomal RNA gene-cluster as an indicator of phylogenetic relationships. *Journal of Molluscan Studies* 66: 565–569. https://doi.org/10.1093/mollus/m004.4.565

Wade C.M., Mordan P.B. & Clarke B. 2001. A phylogeny of the land snails (Gastropoda: Pulmonata). *Proceedings of the Royal Society of London B* 268: 413–422. https://doi.org/10.1098/rspb.2000.1372

Wade C.M., Mordan P.B. & Naggs F. 2006. Evolutionary relationships among the pulmonate land snails and slugs (Pulmonata, Stylommatophora). *Biological Journal of the Linnean Society* 87: 593–610. https://doi.org/10.1111/j.1095-8312.2006.00596.x
Zilch A. 1959–1960. Gastropoda. Teil 2. Euthyneura. In: Schindewolf O.H. (ed.) Handbuch der Paläozoologie, 6 (2). Borntraeger, Berlin.

Printed versions of all papers are also deposited in the libraries of the institutes that are members of the EJT consortium: Muséum national d’histoire naturelle, Paris, France; Meise Botanic Garden, Belgium; Royal Museum for Central Africa, Tervuren, Belgium; Royal Belgian Institute of Natural Sciences, Brussels, Belgium; Natural History Museum of Denmark, Copenhagen, Denmark; Naturalis Biodiversity Center, Leiden, the Netherlands; Museo Nacional de Ciencias Naturales-CSIC, Madrid, Spain; Real Jardín Botánico de Madrid CSIC, Spain; Zoological Research Museum Alexander Koenig, Bonn, Germany; National Museum, Prague, Czech Republic.

Supplementary file
Supp. file 1: A–B. Varadia amboliensis Bhosale, Thackeray, Muley & Raheem gen. et sp. nov. feeding on a banana skin (A; photographed at Amboli) and on a dead cricket (B). C. A scorpion of the genus Heterometrus Ehrenberg, 1828 feeding on V. amboliensis gen. et sp. nov. (image: V. Giri).
https://doi.org/10.5852/ejt.2021.757.1413.4535