Evaluation of *Giardia lamblia* thioredoxin reductase as drug activating enzyme and as drug target

David Leitsch a, b, *, Joachim Müller a, Norbert Müller a

a Institute of Parasitology, Vetsuisse Faculty, University of Bern, Länggassstrasse 122, CH-3012, Bern, Switzerland

b Institute of Specific Prophylaxis and Tropical Medicine, Medical University of Vienna, Kinderspitalgasse 15, A-1090, Vienna, Austria

**Abstract**

The antioxidative enzyme thioredoxin reductase (TrxR) has been suggested to be a drug target in several pathogens, including the protist parasite *Giardia lamblia*. TrxR is also believed to catalyse the reduction of nitro drugs, e.g. metronidazole and furazolidone, a reaction required to render these compounds toxic to *G. lamblia* and other microaerophiles/anaerobes. It was the objective of this study to assess the potential of TrxR as a drug target in *G. lamblia* and to find direct evidence for the role of this enzyme in the activation of metronidazole and other nitro drugs.

TrxR was overexpressed approximately 10-fold in *G. lamblia* WB C6 cells by placing the trxR gene behind the arginine deiminase (ADI) promoter on a plasmid. Likewise, a mutant TrxR with a defective disulphide reductase catalytic site was strongly expressed in another *G. lamblia* WB C6 cell line. Susceptibilities to five antigiardial drugs, i.e. metronidazole, furazolidone, nitazoxanide, albendazole and auranofin were determined in both transfectant cell lines and compared to wildtype. Further, the impact of all five drugs on TrxR activity in vivo was measured.

Overexpression of TrxR rendered *G. lamblia* WB C6 more susceptible to metronidazole and furazolidone but not to nitazoxanide, albendazole and auranofin. Of all five drugs tested, only auranofin had an appreciably negative effect on TrxR activity in vivo, albeit to a much smaller extent than expected. Overexpression of TrxR and mutant TrxR had hardly any impact on growth of *G. lamblia* WB C6, although the enzyme also exerts a strong NADPH oxidase activity which is a source of oxidative stress.

Our results constitute first direct evidence for the notion that TrxR is an activator of metronidazole and furazolidone but rather question that it is a relevant drug target of presently used antigiardial drugs.

© 2016 The Authors. Published by Elsevier Ltd on behalf of Australian Society for Parasitology. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

**1. Introduction**

*Giardia lamblia* (syn *duodenalis*, *intestinalis*) is a microaerophilic protist parasite that occurs in all parts of the world and infects hundreds of millions of people every year (Centers for Disease Control and Prevention, CDC). It colonizes the small intestine and causes gastrointestinal symptoms like nausea, diarrhea, bloating and malabsorption of nutrients. Although not life-threatening in most cases, *Giardia* infections can be persistent and cause growth retardation in children (Buret, 2008). Treatment mainly relies on 5-nitroimidazoles, such as metronidazole and tinidazole, or albendazole, a benzimidazole drug (Leitsch, 2015). 5-nitroimidazoles have been in use against practically all microaerophilic or anaerobic pathogens for more than 50 years due to the comparably low rate of resistance (Leitsch, 2015). However, metronidazole-resistant microaerophiles and anaerobes, including isolates of *G. lamblia*, do occur. Due to the importance of 5-nitroimidazoles, especially metronidazole which is listed among the “essential medicines” by the WHO (World Health Organisation, 2015), a large number of studies on 5-nitroimidazole action and resistance have been conducted throughout the last 30 years.

5-Nitroimidazoles are essentially prodrugs and not very reactive. Reduction at the nitro group, however, activates nitroimidazoles which react with numerous cell constituents - in *G. lamblia* e.g. DNA (Uzlikova and Nohynkova, 2014), proteins (Leitsch et al., 2012), and thiols (Leitsch et al., 2012). Due to the extremely low reduction potential of 5-nitroimidazoles reduction at the nitro group occurs quantitatively only in microaerophilic and anaerobic...
organisms (Müller, 1983). In the protist parasites G. lamblia, Entamoeba histolytica and Trichomonas vaginalis, several enzymatic pathways were identified that are likely to play a role in 5-nitroimidazole reduction, including the central metabolic enzyme pyruvate:ferredoxin oxidoreductase (PFOR) together with ferredoxin (Townson et al., 1996; Rasoloson et al., 2002; Leitsch et al., 2011) and thioredoxin reductase (TrxR) (Leitsch et al., 2007, 2009, 2011), a central redox regulator. Further, another 5-nitroimidazole reducing enzyme, nitroreductase 1 (NR1), was identified in G. lamblia (Müller et al., 2007). A correlation between expression levels of nitroreductase 1 and PFOR/ferredoxin and metronidazole sensitivity in G. lamblia is well documented, as PFOR and nitroreductase 1 are less strongly expressed in many metronidazole-resistant cell lines (Müller et al., 2008; Leitsch et al., 2011). Moreover, overexpression of NR1 from a plasmid renders G. lamblia more sensitive to metronidazole (Nillius et al., 2011). Direct evidence for a role of TrxR in 5-nitroimidazole reduction in vivo, however, has been missing so far.

Importantly, TrxR was not only found to reduce 5-nitroimidazoles but also to be targeted by reduced nitroimidazole intermediates (Leitsch et al., 2007, 2009, 2011), resulting, at least in E. histolytica (Leitsch et al., 2007) and T. vaginalis (Leitsch et al., 2009), in a diminished thioredoxin reducing activity of the enzyme (Leitsch et al., 2007, 2009). Thus, TrxR has an intriguing double role as an activator and target of 5-nitroimidazoles. It was hypothesised that inhibition of TrxR could be one of the major toxic effects brought about by 5-nitroimidazoles (Leitsch et al., 2007, 2009). The TrxR/thioredoxin (Trx) redox system has multiple roles in most organisms, including reduction of peroxiredoxins and maintaining the activity of enzymes like ribonucleotide reductase and methionine sulfoxide reductase. In G. lamblia, however, the role of the TrxR/Trx system is still poorly understood. Although G. lamblia TrxR displays marked disulphide reduction and NADPH oxidase activities (Tejman-Yarden et al., 2013), a functional thioredoxin has not been identified so far (Leitsch et al., 2011; manuscript in preparation). Further, several enzymes known to be dependent on thioredoxin-mediated reduction, such as ribonucleotide reductase, are absent from the parasite. However, despite the current lack of knowledge about the physiological role of TrxR it is generally believed to be an important target not only of metronidazole (Leitsch et al., 2012) but also of aurano-, a rheumatic drug that has been repro-}

### 2. Materials and methods

#### 2.1. Chemicals

Metronidazole, furazolidone, nitazoxanide, auranofin and albendazole were purchased from Sigma (St. Louis, Mo, USA). All drugs tested are depicted in Fig. 1. Growth medium constituents were purchased from Merck (peptone from casein, yeast extract, sodium chloride, glucose, ammonium iron (III) citrate). Fetal calf serum was purchased from Biochrom (Bioswisstec AG, Schaffhausen, Switzerland).

#### 2.2. Cell culture

G. lamblia WB C6 (ATCC 50803) trophozoites were axenically cultivated in Keister's modified Diamond's medium. Media were sterile-filtered. Subcultures were performed every third day.

#### 2.3. Construction of a TrxR overexpressing transfectant

The TrxR gene (GLS0803_9827; XM_001707116) was amplified from genomic DNA isolated from WB C6 (ATCC 50803) with primers bearing 50 bp of the upstream region and 50 bp of the downstream region, respectively, of the arginine deiminase gene (GLS0803_112103; XM_001705703), and PacI and XbaI restriction sites for cloning into the pPac-VInteg vector (Stefanić et al., 2009). The primer sequences were as follows: (forward) CATCTA-GAAACCTCTACACTGAGGT TGTAACACTCCGGAGAAAAAATCCCTATGCATCATGTCTGCTCAAGCATTCA, (reverse) CATAATTAAACGGGATCAGTGCAATCTCGAGAGTTAGTGTATGTGATGAT. Transfections of the new plasmid pTrxR into WB C6 trophozoites were performed in a BTX Electro cell manipulator 600 (Harvard Apparatus) with the settings 500 V, 800 μF, and 720 Ω. Transfected WB C6 cells were cultured in the presence of 100 μg/ml of the plasmid-encoded puromycin N-acetyl-transferase (pac) gene by adding puromycin to the growth medium (100 μg/ml). The resulting plasmid pTrxR-mut was transfection into WB C6 as described above.

#### 2.4. Construction of an episomal mutagenized TrxR gene

The second cysteine of the active site of TrxR on pTrxR was mutated to serine using the QuikChange II XL Site-Directed Mutagenesis kit (Agilent) according to the manufacturer's instructions. The mutagenesis primers introduced one single nucleotide exchange in order to alter a cysteine codon (TGC) to a serine codon (AGC). The sequences of the primers were as follows: (forward) GTGGCCTGGCGCTGACATGG; (reverse) GGGGATCCCAAGCCGAAAGCCGAGCC. The resulting plasmid pTrxR-mut was transfected into WB C6 as described above.

#### 2.5. Two-dimensional gel electrophoresis of G. lamblia protein extracts (2DE)

Two-dimensional gel electrophoresis (2DE) with G. lamblia cell extracts was performed as described previously (Leitsch et al., 2011, 2012). Gels were stained with Coomassie Blue R-250 and evaluated using Melanie™ 4 software (Genebio).

#### 2.6. mRNA quantification of expression by real-time RT–PCR

For quantification of TrxR mRNA expression, cells were harvested as described above and RNA was extracted using a Qiagen RNeasy Kit (Qiagen, Hilden, Germany). Synthesis of first-strand cDNA was performed using a Qiagen OmniscriptRT Kit (Qiagen). The primers used for the amplification of a 189 bp TrxR gene fragment: (forward) CGTTGGCCACGATCCCC; (reverse) GGGAATTCGTGGCCAAGCT. TrxR mRNA levels were calculated using actin mRNA as internal standard (primers: ACTQuanF, ACATATGAGCGCTTGACAGATG; ACTQuanRT TCGGGGAAGGCGCTGCAAC). Quantitative RT–PCR was performed on a LightCycler™ Instrument (Roche Diagnostics, Rotkreuz, Switzerland) as described previously (Nillius et al., 2011). Expression levels of TrxR mRNA were calculated as arbitrary units in relation to the quantity of actin mRNA. PCRs were performed in triplicate in three independent experiments.
2.7. Drug susceptibility and growth assays

Culture medium (10 mL) was inoculated with 10,000 trophozoites/mL and drugs were added in appropriate amounts. After incubation (37 °C, 48 h), cell numbers were determined in a Bürker-Türk chamber and IC50 values were calculated using Grafit 7 software (Erithacus software). The generation times of the cell lines containing 1 mM EDTA, 0.5 mM NADPH and 50 μM furazolidone or nitazoxanide were added to Tris pH 7.5 buffer, DTNB by thiols and proteins was calculated by adding 10 μM DTNB (Merck). Net TrxR activity without background reduction of Tris pH 7.5 buffer containing 0.5 mM NADPH, 1 mM EDTA and 1 mM lamblia extract was measured at 37 °C used in all measurements. Disulphide reductase activity in cell extracts of the respective cell lines was measured photometrically using cell extracts of the respective cell lines. Cells were harvested and lysed by resuspension of the pellet in 10 times the volume of ultrapure water. Cell debris was removed (20,000 g for 10 min) and protein concentrations of the supernatants determined by Bradford assay. 50 μM recombinant G. lamblia TrxR and 100 μM of either furazolidone or nitazoxanide were added to Tris pH 7.5 buffer, containing 1 mM EDTA, 0.5 mM NADPH and 50 μM cytochrome c (Leitsch et al., 2011). Reduction of cytochrome c (Sigma, St. Louis, Mo, USA) was measured in a Lambda 25 UV/Vis spectrometer (Perkin Elmer) at λ = 550 (Δε550 = 20 mM−1 cm−1). Recombinant G. lamblia TrxR was expressed and purified as described (Leitsch et al., 2011).

2.8. Nitroreductase assay

5 μg/mL recombinant G. lamblia TrxR and 100 μM of either furazolidone or nitazoxanide were added to Tris pH 7.5 buffer, containing 1 mM EDTA, 0.5 mM NADPH and 50 μM cytochrome c (Leitsch et al., 2011). Reduction of cytochrome c (Sigma, St. Louis, Mo, USA) was measured in a Lambda 25 UV/Vis spectrometer (Perkin Elmer) at λ = 550 (Δε550 = 20 mM−1 cm−1). Recombinant G. lamblia TrxR was expressed and purified as described (Leitsch et al., 2011).

2.9. Disulphide reductase and NADPH oxidase assays with G. lamblia cell extracts

Disulphide reductase and NADPH oxidase activities of TrxR were measured photometrically using cell extracts of the respective G. lamblia cell lines. Cells were harvested and lysed by resuspension of the pellet in 10 times the volume of ultrapure water. Cell debris was removed (20,000 g for 10 min) and protein concentrations of the supernatants determined by Bradford assay. 50 μg protein/mL were used in all measurements. Disulphide reductase activity in G. lamblia extracts was measured at 37 °C by quantifying reduction of DTNB (5,5’-dithiobis-(2-nitrobenzoic acid)) to TNB (2-nitro-5-thiobenzoate) at λ = 412 (Δε412 = 13.6 mM−1 cm−1) in 100 mM Tris pH 7.5 buffer containing 0.5 mM NADPH, 1 mM EDTA and 1 mM DTNB (Merck). Net TrxR activity without background reduction of DTNB by thiols and proteins was calculated by adding 10 μM of the non-competitive flavin inhibitor diphenyleneiodonium (DPI). NADPH oxidase activity was measured in 100 mM Tris pH 7.5 buffer containing 0.2 mM NADPH and 1 mM EDTA by determining oxidation of NADPH at λ = 340 (Δε340 = 6.2 mM−1 cm−1). Again, 10 μM DPI were added in order to determine the net NADPH oxidase activity of TrxR.

2.10. Western blotting

Polyclonal antibodies were raised in rabbit against G. lamblia TrxR (GenicBio Biotech, Shanghai, China). Sera were diluted 1:1000 for the detection of TrxR on western blots of G. lamblia cell extracts using anti-rabbit IgG (Sigma-Aldrich; 1:5000). Western blotting was performed according to standard protocols.

3. Results

3.1. Generation of TrxR overexpressing transfectant WB C6 lines

In order to assess the role of TrxR in the reduction of metronidazole and other drugs containing a nitro group, an expression construct based on pPAV-VInteg (Stefanic et al., 2009) was devised for the manipulation of TrxR expression levels in G. lamblia. Preliminary experiments harnessing 2DE and RT-qPCR had shown that episomal expression of the TrxR gene under its own promoter did not result in recognisable higher copy numbers of TrxR protein and mRNA, respectively (data not shown). The position of TrxR in 2DE gels had been identified earlier (Leitsch et al., 2011) and was hence known to us at the start of the study. Several promoters of very highly expressed proteins were tested as replacement of the TrxR gene, one of the most strongly expressed proteins in G. lamblia known to us at the start of the study. Several promoters of very highly expressed proteins were tested as replacement of the TrxR gene, one of the most strongly expressed proteins in G. lamblia as recognisable on 2DE gels (Supplementary Fig. 2). The resulting vector was named pTrxR and transfected into G. lamblia WB C6 trophozoites. Transfectants bearing pTrxR (WB TrxR) had an approximately 10-fold higher expression level of TrxR (Fig. 2) than wildtype WB C6, rendering TrxR one of the most prominent proteins in 2DE gels. The elevated expression level of TrxR in WB pTrxR was further confirmed in a western blot assay using an anti-TrxR antibody (Supplementary Fig. 3). The increase in protein levels was mirrored by strong increases of TrxR mRNA levels (Table 1). In order to check whether TrxR was functional in WB TrxR, disulphide

![Fig. 1. The five anti-Giardia drugs tested in this study. 1, metronidazole; 2, furazolidone; 3, nitazoxanide; 4, albendazole, and 5, auranofin.](image-url)
TrxR did hardly, if at all, affect growth of the transfected cell lines in a statistically significant manner. The very high expression levels of native TrxR and mutant TrxR in WB C6 and WB TrxR-mut, respectively, show that NADPH oxidase activity is independent of the catalytic site, as shown by the activity of DPI in reactions, both disulfide reductase and NADPH oxidase activities. IC50 values determined for wildtype WB C6, WB TrxR, and WB TrxR-mut for all three drugs were similar to those in WB TrxR but disulfide reductase activity was practically the same generation time (410 ± 10 min) as WB C6 pTrxR (435 ± 17 min) and WB C6 pTrxR-mut (430 ± 10 min). Further, no effect of altered TrxR levels on motility or shape could be observed under the light microscope. WB TrxR-mut was used as a control in the ensuing drug susceptibility assays in order to demonstrate that increased nitroreductase activity of the enzyme and not overexpression of the enzyme itself is responsible for altered drug susceptibilities.

### 3.2. Evaluation of G. lamblia TrxR as an activator of and target of anti-Giardia drugs

Five anti-Giardia drugs (Fig. 1) were selected for susceptibility testing in the transfected cell lines: the 5-nitroimidazole metronidazole, the 5-nitrofuran furazolidone, the nitrothiazolide nitazoxanide, the benzimidazole albendazole, and auranothin. The first three drugs have nitro groups which need to be reduced for activity (Müller et al., 2007; Leitsch, 2015) and are thus potential substrates for TrxR. Albendazole, however, lacks a nitro group and does not require reduction for toxicity. Also Auranothin, a novel anti-Giardia drug known to semicompetitively inhibit TrxR in vitro (Tejman-Yarden et al., 2013), lacks a nitro group. Thus, prior to the susceptibility assays we hypothesised that WB TrxR would be more susceptible to the three nitro drugs but less susceptible to auranothin, due to much higher concentrations of the target molecule, i.e., TrxR. Susceptibility to albendazole was expected to be unchanged in WB TrxR.

Indeed, overexpression of TrxR had no influence on the susceptibility to albendazole (Table 2). In contrast to our prediction, however, overexpression of TrxR did not confer more tolerance to auranothin but had no effect at all (Table 2). In accordance with TrxR’s capacity to function as a nitroreductase, however, overexpression of functional TrxR rendered WB TrxR more sensitive to metronidazole and furazolidone (Table 2) than wildtype or WB TrxR-mut. Surprisingly, the effect of TrxR overexpression was not

### Table 1

| TrxR expression | Relative expression level (protein) | Relative expression level (mRNA) | Disulphide reductase activity in cell extract (nmol min⁻¹ mg⁻¹) | NADPH oxidase activity in cell extract (nmol min⁻¹ mg⁻¹) |
|-----------------|-----------------------------------|---------------------------------|---------------------------------------------------------------|---------------------------------------------------------|
| WB C6 (wildtype)| 1×10                              | 1×1                             | 10.2 ± 1.5                                                    | 27 ± 7                                                   |
| WB TrxR         | 1×10                              | >2.5                            | 109 ± 14                                                      | 213 ± 17                                                 |
| WB TrxR-mut     | 1×6.5                             | 1×11                            | 12.1 ± 2.3                                                    | 278 ± 27                                                 |

### Table 2

| Drug            | IC50 Wildtype | IC50 WB TrxR | IC50 WB TrxR-mut |
|-----------------|---------------|--------------|------------------|
| Metronidazole (µM) | 3.44 ± 0.06  | 2.33 ± 0.03* | 3.71 ± 0.36      |
| Furazolidone (µM)  | 0.73 ± 0.00  | 0.52 ± 0.03** | 0.79 ± 0.02      |
| Nitazoxanide (µM)  | 1.86 ± 0.05  | 1.63 ± 0.04  | 1.76 ± 0.03      |
| Albendazole (nM)  | 59.3 ± 0.4   | 60.8 ± 0.4   | 59.9 ± 1.4       |
| Auranothin (µM)   | 16.5 ± 1     | 16.8 ± 0.1   | ND               |

*IC50 values determined for wildtype WB C6, WB TrxR, and WB TrxR-mut for all five drugs. Asterisks indicate statistically significant differences to wildtype (unpaired t-test, two sided; *P < 0.01; **P < 0.05). ND means not determined.

---

**Fig. 2.** Sections of 2D-gels from cell extracts of G. lamblia WB C6 without plasmid (wildtype), with a plasmid harbouring the trxR gene behind the ADI promoter (WB TrxR), and with a plasmid harbouring a mutated trxR gene behind the ADI promoter (WB TrxR-mut). TrxR is encircled. The degree of overexpression of TrxR (functional or mutated) in relation to wildtype is indicated beneath the gel sections. The gel shown is representative for three biological replicates in total.
significant in case of nitazoxanide although this drug has a nitro group which was shown to be essential for toxicity (Müller et al., 2007). In order to check whether nitazoxanide is a potential substrate for *G. lamblia* TrxR, we used purified recombinant *G. lamblia* TrxR for measuring reduction of nitazoxanide along the lines described for metronidazole and other 5-nitroimidazoles in an earlier study (Leitsch et al., 2011). Indeed, no nitroreductase activity of recombinant TrxR activity could be observed with nitazoxanide as substrate (Table 3). In contrast, furazolidone, was strongly reduced by recombinant *G. lamblia* TrxR (Table 3). This suggests that the elevated expression levels of TrxR in WB TrxR are responsible for a higher susceptibility to metronidazole and furazolidone, whereas the susceptibility to nitazoxanide remains unchanged because it is not a substrate of TrxR.

As a second step we delineated the role of TrxR as a drug target and assessed the effect of the five drugs on disulphide reductase activity of TrxR in WB TrxR (Fig. 3). Inhibition of TrxR by metronidazole and furazolidone was not significant, whereas nitazoxanide and albendazole did not inhibit TrxR at all (Fig. 3). Auranoﬁn was the only drug that substantially reduced disulphide reductase activity of TrxR in WB TrxR (Fig. 3). When applied at 20 μM auranoﬁn reduced TrxR activity to 60% as compared to control but even at a concentration of 50 μM, which is about the 300-fold dose of the IC_{50} as determined with recombinant *G. lamblia* TrxR in vitro (Tejman-Yarden et al., 2013), residual activity of TrxR in WB TrxR did not drop below 40% as compared to untreated WB TrxR. This indicates that the dose-response curve is rather flat and that even much higher concentrations of auranoﬁn would not reduce TrxR activity in WB TrxR below the rate as observed in wildtype (Fig. 3). Indeed, TrxR activity in WB TrxR treated with 50 μM auranoﬁn, was still 4–5-fold higher than in untreated wildtype (Fig. 3), at least after 2 h of incubation. Our results demonstrate that TrxR is a target of auranoﬁn in the living parasite. It is not likely, however, that TrxR is a critical target of auranoﬁn in *G. lamblia* since the susceptibility of WB TrxR to auranoﬁn is equal to that of wildtype (Table 2). In fact, the results of this experiment suggest that *G. lamblia* TrxR is not a relevant target of any drug presently used for the treatment of giardiasis.

### 4. Discussion

We devised a plasmid-based expression construct that coupled the trxR gene to the ADI promoter, resulting in approximately tenfold higher intracellular concentrations of the enzyme as compared to wildtype. Overexpression of TrxR rendered *G. lamblia* more susceptible to metronidazole and furazolidone (Table 2), two drugs which are reduced by recombinant TrxR *in vitro* (Leitsch et al., 2011) (Table 3). Although indirect evidence for the reduction of metronidazole *in vivo* by TrxR had already been presented for *E. histolytica* (Leitsch et al., 2007), *T. vaginalis* (Leitsch et al., 2009), and *G. lamblia* (Leitsch et al., 2007), this study constitutes first direct evidence that TrxR is an activator of nitro drugs in the living parasite. It can be ruled out that the higher susceptibility of WB TrxR to metronidazole and furazolidone was due to a decreased fitness caused by overexpression of TrxR as such because the susceptibilities to the other drugs tested were unaltered. Further, TrxR had to be functional in order to render WB C6 more susceptible to metronidazole and furazolidone (Table 2), as overexpression of mutated TrxR in WB TrxR-mut did not result in increased sensitivity to metronidazole and furazolidone (Table 2). Finally, since NADPH oxidase activity in WB TrxR-mut was even higher than in WB TrxR, it can be also ruled out that TrxR indirectly renders metronidazole susceptible to *G. lamblia* more susceptible to metronidazole and furazolidone by causing oxidative stress through generation of superoxide radicals. It is evident, however, that the effect of TrxR overexpression on metronidazole and furazolidone susceptibility was only moderate. This finding is in line with the notion that several factors contribute to nitro drug reduction in *G. lamblia* and other microaerophilic parasites. One well documented example is nitroreductase 1 from *G. lamblia* which, when overexpressed, increases susceptibility to metronidazole quite to the same extent as shown here for TrxR (Nillius et al., 2011; reviewed in Leitsch, 2015).

Surprisingly, the direct effect of four of the five drugs on TrxR activity was minute, if at all measurable. Only auranoﬁn, a compound known to inhibit *G. lamblia* TrxR effectively *in vitro* also

---

**Table 3**

| Drug                  | Reduction by *G. lamblia* TrxR |
|-----------------------|--------------------------------|
| Metronidazole (1 mM)  | 81 ± 30 nmol min^{-1} mg^{-1}   |
| Furazolidone (100 μM) | 1094 ± 25 nmol min^{-1} mg^{-1} |
| Nitazoxanide          | 0                              |

---

**Fig. 3.** Impact of the drugs tested on disulphide reductase activity of TrxR in cell extracts of WB TrxR after 2 h of treatment. Untreated WB TrxR represents 100% activity. TrxR activities from untreated wildtype and wildtype treated with auranoﬁn are given for comparison. Asterisks indicate statistically significant differences to TrxR activity in untreated WB TrxR (paired t-test, two sided; *P < 0.05*).
inhibited TrxR \textit{in vivo} to a relevant extent (Fig. 3). Still, even if auranofin was applied in very high concentrations (50 \(\mu\text{M}\)) the residual TrxR activity was still 4 to 5-times higher than in untreated wildtype (Fig. 3). Since wildtype and WB TrxR were equally susceptible to auranofin (Table 2), we conclude that TrxR, quite in contrast to the commonly held belief (Tejman-Yarden et al., 2013; Watkins and Eckmann, 2016), is not the primary target of this drug in \textit{G. lamblia}.

Acknowledgements

David Leitsch was supported by grant J3492 of the Austrian Science Fund (FWF). Norbert Müller [NM] and Joachim Müller [JM] were supported by the Swiss National Science Foundation (grants SNF 31003A_138353 [NM and JM] and 31003A_163230 [NM]).

Appendix A. Supplementary data

Supplementary data related to this article can be found at http://dx.doi.org/10.1016/j.ijpddr.2016.07.003.

References

Buret, A.G., 2008. Pathophysiology of enteric infections with \textit{Giardia duodenalis}. Parasite 15, 261–265.

Centers for Disease Control and Prevention, 2016. Parasites—\textit{Giardia}. https://www.cdc.gov/parasites/giardia/, Last time accessed: June 27th.

Debnath, A., Parsonage, D., Andrade, R.M., He, C., Cobo, E.R., Hirata, K., Chen, S., Garcia-Rivera, G., Orozco, E., Martinez, M.B., Gunatilleke, S.S., Barrios, A.M., Arkin, M.R., Poole, L.B., McKerrow, J.H., Reed, S.L., 2012. A high-throughput drug screen for \textit{Entamoeba histolytica} identifies a new lead and target. Nat. Med. 18, 956–960.

Leitsch, D., 2015. Drug Resistance in the microaerophilic parasite \textit{Giardia lamblia}. Curr. Trop. Med. Rep. 2015 (2), 128–135.

Leitsch, D., Kolarich, D., Wilson, I.B.H., Altmann, F., Duchêne, M., 2007. Nitroimidazole action in \textit{Entamoeba histolytica}: a central role for thioredoxin reductase. Plos. Biol. 5, 1820–1834.

Leitsch, D., Kolarich, D., Binder, M., Stadlmann, J., Altmann, F., Duchêne, M., 2009. \textit{Trichomonas vaginalis}: metronidazole and other nitroimidazole drugs are reduced by the flavin enzyme thioredoxin reductase and disrupt the cellular redox system. Implications for nitroimidazole toxicity and resistance. Mol. Microbiol. 72, 518–536.

Leitsch, D., Schlosser, S., Burgess, A., Duchêne, M., 2012. Nitroimidazole drugs vary in their mode of action in the human parasite \textit{Giardia lamblia}. Intern. J. Parasitol. Drugs Drug Resis 2, 166–170.

Müller, J., Ley, S., Felger, I., Hemphill, A., Müller, N., 2008. Identification of differentially expressed genes in a \textit{Giardia lamblia} WB GS clone resistant to nitozoxamide and metronidazole. J. Antimicrob. Chemother. 2008 (62), 72–82.

Müller, J., Wasting, J., Sanderson, S., Müller, N., Hemphill, A., 2007. A novel \textit{Giardia lamblia} nitroreductase, GlNR1, interacts with nitozoxamide and other thiazolides. Antimicrob. Agents Chemother. 51, 1979–1986.

Müller, M., 1983. Mode of action of metronidazole on anaerobic bacteria and protozoa. Surgery 93, 165–171.

Nillius, D., Müller, J., Müller, N., 2011. Nitroreductase (GlNR1) increases susceptibility of \textit{Giardia lamblia} and \textit{Escherichia coli} to nitro drugs. J. Antimicrob. Chemother. 66, 1029–1035.

Rasoloson, D., Vanacová, S., Tomková, E., Rázga, J., Hrdý, I., Tachezy, J., Kulda, J., 2002. Mechanisms of in vitro development of resistance to metronidazole in \textit{Trichomonas vaginalis}. Microbiology 48, 2467–2477.

Schlosser, S., Leitsch, D., Duchêne, M., 2013. \textit{Entamoeba histolytica}: identification of thioredoxin- targeted proteins and analysis of serine acetyltransferase-1 as a prototype example. Biochem. J. 451, 277–288.

Stefanić, S., Morf, L., Kulangara, C., Regós, A., Sonda, S., Schraner, S., Spycher, C., Wild, P., Hehl, A.B., 2009. Neogenesis and maturation of transient Golgi-like cisternae in a simple eukaryote. J. Cell Sci. 122, 2846–2856.

Tejman-Yarden, N., Miyamoto, Y., Leitsch, D., Santini, J., Debnath, A., Gut, J., McKerrow, J.H., Reed, S.L., Eckmann, L., 2013. Auranofin, a repurposed drug, is effective against metronidazole-resistant \textit{Giardia lamblia}. Antimicrob. Agents Chemother. 57, 2029–2035.

Townson, S.M., Upcroft, J.A., Upcroft, P., 1996. Characterisation and purification of pyruvate:ferredoxin oxidoreductase from \textit{Giardia duodenalis}. Mol. Biochem. Parasitol. 79, 183–189.

Uzlikova, M., Nohynkova, E., 2014. The effect of metronidazole on the cell cycle and DNA in \textit{metronidazole}-susceptible and -resistant \textit{Giardia} cell lines. Mol. Biochem. Parasitol. 198, 75–81.

Watkins, R.R., Eckmann, L., 2016. Treatment of giardiasis: current status and further directions. Curr. Infect. Dis. Rep. 18, 288.