Two novel species and two new records of *Distoseptispora* from freshwater habitats in China and Thailand

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Abstract

During investigations into freshwater fungi from the Great Mekong Subregion, four *Distoseptispora* taxa were collected from China and Thailand. Based on morphological characteristics, and phylogenetic analyses of combined LSU, ITS, SSU, TEF1-\(\alpha\), and RPB2 sequence data, two new species *Distoseptispora bangkokensis* and *D. lancangjiangensis* are introduced, and two known species *D. clematidis* and *D. thyssanolaenae* were first reported in freshwater habitat. Illustrations and descriptions of these taxa are provided, along with comparisons with extant taxa in the genus.

Keywords

2 new taxa, Distoseptisporales, freshwater fungi, morphology, phylogeny, taxonomy
Introduction Distoseptisporaceae was introduced by Su et al. (2016) based on morphological and phylogenetic analyses, with *Distoseptispora* as type genus. Distoseptisporaceae is placed in Distoseptisporales, which was introduced by Luo et al. (2019), and currently comprises two families, Aquapteridosporaceae and Distoseptisporaceae (Luo et al. 2019; Wijayawardene et al. 2020; Hyde et al. 2021). Species of both families are commonly reported from freshwater habitats (Yang et al. 2015, 2018; u et al. 2016; Li et al. 2021; Hyde et al. 2016a, 2019, 2020; Luo et al. 2018, 2019; Song et al. 2020; Dong et al. 2021).

*Distoseptispora* as a single genus in Distoseptisporaceae was introduced by Su et al. (2016) with *D. fluminicola* as the type species. The genus is characterized by monoblastic, cylindrical, conidiogenous cells, with percurrent proliferation, acrogenous, solitary, brown or yellowish/reddish brown, olivaceous, distoseptate or euseptate, cylindrical, obclavate, rostrate conidia, truncate base, with rounded apices, basal cell with a cross wall and basal scar. This genus is not known for its sexual morph (Su et al. 2016; Yang et al. 2018; Hyde et al. 2019, 2020; Luo et al. 2019; Sun et al. 2020). Currently, 32 species are accepted in the genus of which 13 from terrestrial habitats and 19 were reported from freshwater environments (Su et al. 2016; Hyde et al. 2016a, 2019, 2020; Xia et al. 2017; Yang et al. 2018; Luo et al. 2018, 2019; Monkai et al. 2020; Song et al. 2020; Sun et al. 2020; Li et al. 2021; Index Fungorum 2021).

During our ongoing study of freshwater fungi along the north-south gradient in the Asian/Australian region (Hyde et al. 2016b), we collected four species in the genus. Two new species, *Distoseptispora bangkokensis* and *D. lancangjiangensis*, are introduced in this study, *D. clematidis* and *D. thysanolaenae* are newly recorded from freshwater habitats for the first time in China. Morphological descriptions and illustrations of the species and an updated multi-gene phylogenetic tree are provided to reveal their taxonomic position among the species in the Distoseptisporales, and also provided the comparison of morphological characteristics, habitats and hosts information of species newly added to *Distoseptispora* after Monkai et al. (2020) (Table 2).

Materials and methods

Isolation and morphology

Specimens of submerged decaying wood were collected from Dulongjiang, Nanpanjiang, Lancangjiang and Chao Phraya River in China and Thailand respectively. Multiple samples will be collected at each collection site at different times, allowing more strains to be obtained for each species. Methods of morphological observation and isolation follow Luo et al. (2018) and Senanayake et al. (2020). IFW (Tarosoft(R) Image Frame Work) was used for measurement of photomicrograph, and Adobe Photoshop CS5 software was used to process images for making photo-plates (Adobe Systems
Inc., USA). Single spore isolation was performed according to the following steps: The conidia suspension from specimens, absorbed with a sterilized pipette, was placed on potato dextrose agar (PDA) and incubated at room temperature overnight. Germinated conidia were transferred to new PDA/MEA (Beijing land bridge technology CO., LTD., China) plates and incubated in an incubator at room temperature (25 °C). Specimens were deposited in the Kunming Institute of Botany, Academia Sinica herbarium (KUN-HKAS), and Mae Fah Luang University herbarium (MFLU). Cultures were deposited in the Dali University Culture Collection (DLUCC), China General Microbiological Culture Collection Center (CGMCC), and Mae Fah Luang University Culture Collection (MFLUCC). Facesoffungi number was obtained as described in Jayasiri et al. (2015) and Index Fungorum number was also registered (http://www.indexfungorum.org/Names/Names.asp). In this study, multiple samples were collected for each sample site and related environment, but unfortunately, there were still no more strains for the two new species in the paper.

**DNA extraction, PCR amplification, and sequencing**

DNA extraction, PCR amplification, sequencing and phylogenetic analysis follow Dissayanake et al. (2020) with the following modifications. Fungal mycelia (200–500 mg) were scraped from grown on PDA/MEA plates using sterile scalpel, transferred to microcentrifuge tube with sterilized needles, and then grind with liquid nitrogen or quartz sand to break the cells. DNA was extracted using the T relief™ Plant Genomic DNA Kit (TSP101) according to the manufacturer’s instructions.

Five gene regions, LSU, ITS, SSU, TEF1-α, and RPB2 were amplified using LR0R/LR5, ITS5/ITS4, NS1/NS4, 983F/EF1-2218R, and RPB2-5F/RPB2-7cR (Vilgalys and Hester 1990; White et al. 1990; Liu et al. 1999) primer pairs respectively. Primer sequences are available at the WASABI database at the AFTOL website (aftol.org). The PCR mixture contained 12.5 μL of 2 × Power Taq PCR Master Mix (a premix and ready to use solution, including 0.1 Units/μL Taq DNA Polymerase, 500μm dNTP Mixture each (dATP, dCTP, dGTP, dTTP), 20 mm Tris-HCl pH 8.3, 100 Mm KCl, 3 mM MgCl₂, stabilizer and enhancer), 1 μL of each primer including forwarding primer and reverse primer (10 μm), 1 μL template DNA extract and 9.5 μL deionized water (Luo et al. 2018). The PCR cycling conditions of LSU, ITS, SSU and TEF1-α were as follows: 94 °C for 3 min, followed by 35 cycles of denaturation at 94 °C for 30s, annealing at 55 °C for 50s, elongation at 72 °C for 1 min, and a final extension at 72 °C for 10 min. The PCR thermal cycle of RPB2 has a total of 40 cycles, and the conditions are as follows: initially denature at 95 °C for 5 min, and then enter 40 cycles: denaturation at 95 °C for 1 min, annealing at 52 °C for 2 min, extension at 72 °C for 90s, and finally at 72 °C for 10 min. PCR products were then purified using minicolumns, purification resin, and buffer according to the manufacturer’s protocols (Amersham product code: 27–9602–01). The sequences were carried out at Beijing Tsingke Biotechnology Co., Ltd. (Beijing, P.R. China).
Table 1. Strains used for phylogenetic analysis and their corresponding GenBank numbers. The type strain are in bold font.

| Species                      | Source          | GenBank accession number       | Reference                      |
|------------------------------|-----------------|--------------------------------|--------------------------------|
| Aquapteridospora fusiformis  | MFLUCC 18-1606  | MK849798 MK828652 MN194056 – – | Luo et al. (2019)              |
| A. lignicola                 | MFLUCC 15-0377  | KU221018 – – – –              | Yang et al. (2015)             |
| Distoseptispora adscendens   | HKUCC 10820     | DQ408561 – – DQ435092 –      | Shenoy et al. (2006)           |
| D. appendiculata             | MFLUCC 18-0259  | MN163023 MN163009 MN174866 – – | Luo et al. (2019)              |
| D. aquatica                  | MFLUCC 15-0374  | KU376268 MF077552 – – – –    | Su et al. (2016)               |
| D. bambusae                  | MFLUCC 20-0091  | MT232718 MT232713 MT232880 MT232881 MT232716 | Sun et al. (2020) |
| D. bambusae                  | MFLUCC 14-0583  | MT232717 MT232712 – MT232882 – | Sun et al. (2020) |
| D. bangkokokensis            | MFLUCC 18-0262  | MZ518206 MZ518205 – – MZ518208 | This study                    |
| D. cangshanensis             | MFLUCC 16-0970  | MG979761 MG979754 MG988419 – – | Luo et al. (2018)              |
| D. caricis                   | CBS 146041      | MN567632 MN562124 – MN556805 – | Crous et al. (2019)            |
| D. clematidis                | MFLUCC 17-2145  | MT214617 MT310661 – MT394721 MT226728 | Phukhamsakda et al. (2020) |
| D. clematidis                | KUN-HKAS 112708 | MW879523 MW723056 MW729784 – MW774580 | This study                    |
| D. dehongensis               | KUMCC 18-0090   | MK079662 MK085061 MK087659 – – | Hyde et al. (2019)             |
| D. euseptata                 | MFLUCC 20-0154  | MW081544 MW081539 – MW151860 – | Li et al. (2021)               |
| D. euseptata                 | DLUCC 52024     | MW081545 MW081540 MW084994 MW084996 – – | Li et al. (2021) |
| D. fasciculata               | KUMCC 19-0081   | MW287775 MW286501 MW396656 – – | Dong et al. (2021)             |
| D. fluminicola               | MFLUCC 15-0417  | KU376270 MF077553 – – – –    | Su et al. (2016)               |
| D. guttulata                 | MFLUCC 16-0183  | MF077554 MF077543 MF135651 – MF077532 | Yang et al. (2018) |
| D. hydei                     | MFLUCC 20-0115  | MT742830 MT734661 – MT767128 – | Monkai et al. (2020)           |
| D. lancangjiangensis         | KUN-HKAS 112712 | MW879522 MW723055 MW882260 – | This study                    |
| D. leonensis                 | HKUCC 10822     | DQ408566 – – DQ435089 –      | Sheno et al. (2006)            |
| D. lignicola                 | MFLUCC 18-0198  | MK849797 MK828651 – – – MK828318 | Luo et al. (2019)             |
| D. longispora                | HFJAU 0705      | MH555357 MH555359 – – – MH555431 | Song et al. (2020)            |
| D. martinii                  | CGMCC 3.18651   | KX033566 KU999975 – – – KX033537 | Xia et al. (2017)             |
| D. multisepata               | MFLUCC 16-1044  | MF077555 MF077544 MF135652 MF135644 MF077533 | Yang et al. (2018) |
| D. multisepata               | MFLUCC 15-0609  | KX710140 KX710145 MF135659 – NG_065693 | Hyde et al. (2016) |
| D. neostrata                 | MFLUCC 18-0376  | MN163017 MN163008 – – –      | Luo et al. (2019)             |
| D. obclavata                 | MFLUCC 18-0329  | MN163010 MN163012 – – –      | Luo et al. (2019)             |
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| Species                  | Source          | GenBank accession number   | Reference       |
|--------------------------|-----------------|---------------------------|-----------------|
| **Species**              | **Source**      | **LSU** | **ITS** | **TEF1-α** | **RPB2** | **SSU** | **Reference** |
| *D. obpyriformis*        | MFLUCC 17-01694 | MG979764     | –      | MG988422   | MG988415 | –      | Luo et al. (2018) |
| *D. obpyriformis*        | DLUCC 0867      | MG979765     | MG979757 | MG988423   | MG988416 | –      | Luo et al. (2018) |
| *D. palmarum*            | MFLUCC 18-1446  | MK079663     | MK085062 | MK087660   | MK079661 | –      | Hyde et al. (2019) |
| *D. phangngaensis*       | MFLUCC 16-0857  | MF077556     | MF077545 | MF135653   | –      | MF077534 | Yang et al. (2018) |
| *D. rayongensis*         | MFLUCC 18-0415  | MH457137     | MH457172 | MH463253   | MH463255 | MH457169 | Hyde et al. (2012) |
| *D. rostrata*            | MFLUCC 16-0969  | MG979766     | MG979758 | MG988424   | MG988417 | –      | Luo et al. (2018) |
| *D. saprophytica*        | MFLUCC 18-1238  | MW287780     | MW286506 | MW396651   | MW504069 | –      | Dong et al. (2021) |
| *D. songkhaensis*        | MFLUCC 18-1234  | MW287755     | MW286482 | MW396642   | –      | –      | Dong et al. (2021) |
| *D. suoluoensis*         | MFLUCC 17-0224  | MF077557     | MF077546 | MF135654   | –      | MF077535 | Yang et al. (2018) |
| *D. suoluoensis*         | MFLUCC 17-1305  | MF077558     | MF077547 | –          | –      | MF077536 | Yang et al. (2018) |
| *D. tectonae*            | MFLUCC 12-0291  | KX751713     | KX751711 | KX751710   | KX751708 | –      | Hyde et al. (2016) |
| *D. tectonae*            | MFLUCC 16-0946  | MG979768     | MG979760 | MG988426   | MG988418 | –      | Luo et al. (2018) |
| *D. tectonigena*         | MFLUCC 12-0292  | KX751714     | KX751712 | –          | KX751709 | –      | Hyde et al. (2016) |
| *D. thailandica*         | MFLUCC 16-0270  | MH260292     | MH275060 | MH412767   | –      | MH260334 | Tilpromma et al. (2018) |
| *D. thysonolaenae*       | KUN-HKAS 102247 | MK064091     | MK045851 | MK086031   | –      | –      | Phookamsak et al. (2019) |
| *D. thysonolaenae*       | KUN-HKAS 112710 | MW879524     | MW723057 | MW729783   | –      | –      | This study |
| *D. xishuangbannaensis*  | KUMCC 17-0290   | MH260293     | MH275061 | MH412768   | MH412754 | MH260335 | Tilpromma et al. (2018) |
| *D. yunnanensis*         | MFLUCC 20-0153  | MW081546     | MW081541 | MW084995   | MW151861 | –      | Li et al. (2021) |
| *Myrmecridium aquaticum* | MFLUCC 15-0366  | MK849804     | –      | –          | –      | MK828323 | Luo et al. (2019) |
| *M. aquaticum*           | S-1158          | MK849803     | MK828656 | MN194061   | MN124540 | MK828322 | Luo et al. (2019) |
| *M. banksiae*            | CBS 132536      | JX069855     | JX069871 | –          | –      | –      | Crous et al. (2012) |
| *Pseudostanjehughesia*   | MFLUCC 16-0569  | MF077559     | MF077548 | MF135655   | –      | MF077537 | Yang et al. (2018) |
| *P. lignicola*           | MFLUCC 16-0532  | MK849787     | MK828643 | MN194047   | MN124534 | –      | Luo et al. (2019) |
| *Sporidesmium dulongense*| MFLUCC 17-0116  | MH795817     | MH795812 | MH801191   | MH801190 | –      | Luo et al. (2019) |
| *S. lagensiforme*        | DLUCC 0880      | MK849782     | MK828640 | MN194044   | MN124533 | –      | Luo et al. (2019) |
| *S. pyriformatum*        | MFLUCC 15-0620  | KX710141     | KX710146 | MF135662   | MF135649 | –      | Hyde et al. (2016) |
| *S. thailandense*        | MFLUCC 15-0617  | MF077561     | MF077550 | MF135657   | –      | –      | Yang et al. (2018) |
| *S. thailandense*        | MFLUCC 15-0964  | MF374370     | MF374361 | MF370957   | MF370955 | –      | Zhang et al. (2017) |

*1 Ex-type strain of *Distoseptispora submersa*. 
Phylogenetic analysis

Preliminary identification of genes obtained from fresh strains by GenBank database. The LSU, ITS, SSU, TEF1-α, and RPB2 used for phylogenetic analysis are selected based on the preliminary identification results and the related publications (Yang et al. 2018; Monkai et al. 2020). The sequences were aligned using MAFFT online service: Multiple alignment program for amino acid or nucleotide sequences MAFFT version 7 (Katoh and Standley 2013: http://mafft.cbrc.jp/alignment/server/index.html), and edited manually in BioEdit v. 7.0 (Hall 1999). The sequence dataset was combined using SequenceMatrix v.1.7.8 (Vaidya et al. 2011). The alignment formats were change to PHYLIP and NEXUS formats by ALigment Transformation EnviRonment (ALTER) website (http://sing.ei.uvigo.es/ALTER/).

Maximum likelihood (ML) analysis was carried out using the RAxML-HPC2 on XSEDE (8.2.12) (Stamatakis 2006; Stamatakis et al. 2008) of CIPRES Science Gateway website (Miller et al. 2010: http://www.phylo.org/portal2) and the estimated proportion of invariant sites is (GTRGAMMA+I) model.

Bayesian analyses were performed in MrBayes 3.2.6 (Ronquist et al. 2012) and the best-fit model (LSU, ITS, SSU, TEF1-α, and RPB2 are all GTR+I+G) of sequences evolution was estimated via MrModeltest 2.2 (Guindon and Gascuel 2003; Nylander 2004; Darriba et al. 2012). The Markov Chain Monte Carlo (MCMC) sampling approach was used to calculate posterior probabilities (PP) (Rannala and Yang 1996). Bayesian analyses of six simultaneous Markov chains were run for 10000000 generations with trees sampled every 1000 generations.

Phylogenetic trees were visualized using FigTree v1.4.0 (Rambaut 2012: http://tree.bio.ed.ac.uk/software/figtree/), editing and typesetting using Adobe Illustrator (AI) (Adobe Systems Inc., the United States). The new sequences were submitted in GenBank and the strain information used in this paper is provided in Table 1. The alignments and phylogenetic trees were deposited in TreeBASE (http://www.treebase.org/, accession number: 28758).

Results

Phylogenetic analysis

The dataset composed of LSU (1–744 bp), ITS (745–1310 bp), TEF1-α (1311–2161 bp), RPB2 (2162–3178 bp), and SSU (3179–4199 bp) gene, comprising a total of 4199 characters (including gaps), including 56 taxa with Pseudostanjehughesia aquitropica (MFLUCC 16-0569) and P. lignicla (MFLUCC 15-0352) as the outgroup taxa (Figure 1). The ML and BI phylogenetic analyses produced similar topology. The combined dataset analysis of RAxML generates a best-scoring tree (Figure 1), with the final ML optimization likelihood value of -30393.557997. The aligned matrix had 1624 distinct alignment patterns, with 36.44% completely undetermined characters or gaps. The
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| Species                    | Conidiophore (μm) | Conidia (μm) | Conidia septation | Conidia characteristic                           | Habitat     | Host                  | Reference   |
|----------------------------|-------------------|--------------|-------------------|-------------------------------------------------|-------------|-----------------------|-------------|
| *Distoseptispora bangkokensis* | 37–55 × 3–4       | 400–568 × 13–16 | Multi-distoseptate | Elongate, obclavate, rostrate, dark olivaceous to dark brown | Freshwater | Unidentified submerged wood | This study |
| *D. lancangjiangensis*    | 30–41 × 5–6       | 83–220 × 12–14 | 16–41-distoseptate | Oblclavate, cylindrical, elongated, straight or curved, brown to greenish-brown | Freshwater | Unidentified submerged wood | This study |
| *D. euseptata*            | 19–28 × 4–5       | 37–54 × 8–9   | 4–7-euseptate     | Oblpyriform to obclavate, straight or curved, olivaceous | Freshwater | Unidentified submerged wood | Li et al. 2021 |
| *D. fasciculata*          | 12–16 × 5–6       | 46–200 × 10–16.5 | 10–40-distoseptate | Subclavatril to obclavate, mostly curved, olivaceous when young, dark brown when mature | Freshwater | Unidentified submerged wood | Dong et al. 2021 |
| *D. longispora*           | 17–37 × 6–10      | 189–297 × 16–23 | 31–56-distoseptate | Oblavate, elongated, straight or slightly curved, to yellowish brown | Freshwater | Unidentified submerged wood | Song et al. 2020 |
| *D. suprophytica*         | 50–140 × 3.2–4.2  | 14.5–30 × 4.5–7.5 | 2–6-distoseptate | Subclavatril to obclavate, straight or curved, olivaceous to brown | Freshwater | Unidentified submerged wood | Dong et al. 2021 |
| *D. songkhlaensis*        | 70–90 × 4–5.5     | 44–125 × 9–14.5 | 9–16-distoseptate | Oblavate, straight or curved, olivaceous to brown | Freshwater | Unidentified submerged wood | Dong et al. 2021 |
| *D. yunnanensis*          | 131–175 × 6–7     | 58–108 × 8–10 | 6–10-euseptate    | Oblavate, rostrate, straight or slightly curved, mid olivaceous to brown | Freshwater | Unidentified submerged wood | Li et al. 2021 |

Table 2. Comparison of morphological characteristic, habitats and hosts’ information of species added to *Distoseptispora* after Monkai et al. (2020) (for other species see Monkai et al. 2020).

Base frequency and rate are as follows: $A = 0.243915$, $C = 0.259360$, $G = 0.279029$, $T = 0.217696$; rate $AC = 1.166355$, $AG = 2.813539$, $AT = 1.110401$, $CG = 0.796371$, $CT = 5.621229$, $GT = 1.000000$; gamma distribution shape: $\alpha = 0.221933$. Bootstrap support values with a maximum likelihood (ML) greater than 70%, and Bayesian posterior probabilities (PP) greater than 0.97 are given above the nodes.

The phylogenetic tree shows that the new species *Distoseptispora bangkokensis* (MFLUCC 18-0262) was placed as a sister taxon to *D. bambusae* (MFLUCC 14-0583 and MFLUCC 20-0091), *D. dehongensis* (KUMCC 18-0090), *D. euseptata* (MFUCC 20-0154 and DLUCC S2024), *D. lancangiangensis* (KUN-HKAS 112712), *D. suoluoensis* (MFLUCC 17-0224 and MFLUCC 17-1305), *D. thysanolaenae* (KUN-HKAS 102247 and KUN-HKAS 112710), and *D. yunnanensis* (MFLUCC 20-0153) with low bootstrap support with low bootstrap support (Figure 1), whereas *D. lancangiangensis* clustered with *D. suoluoensis* with 97%ML/0.98PP support. *Distoseptispora thysanolaenae* (KUN-HKAS 112710) and *D. clemtidis* (KUN-HKAS 112708) clustered with the ex-type strain of *D. thysanolaenae* (KUN-HKAS 102247) and *D. clermatidis* (MFLUCC 17-2145), respectively, with 100%ML/1.00PP and 97%ML/0.99PP bootstrap support.
**Taxonomy**

*Distoseptispora bangkokensis* H.W. Shen, D.F. Bao, K.D. Hyde & Z.L. Luo, sp. nov.

Index Fungorum Number No: IF558556

Facesoffungi Number No: FoF09993

Figure 2

Etymology. Referring to the collecting location, Bangkok, Thailand.

**Holotype.** MFLU 21-0110

**Description.** Saprobic on submerged wood in freshwater stream. **Sexual morph:** Undetermined. **Asexual morph:** Colonies effuse, glistening, hairy, brown to dark brown. **Mycelium** partly superficial in the substratum, composed of hyaline to pale brown, sep-
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Figure 2. Distoseptispora bangkokensis (MFLU 21-0110, holotype) A colonies on the substratum B conidiophores C conidiophores with conidia D conidiogenous cell E-G conidia H germinating conidium

Scale bars: 20 μm (B, C, H); 10 μm (D); 50 μm (E-G).
tate, branched hyphae. Conidiophores 37–55 × 3–4 μm (x = 46 × 3 μm, n = 15) macronematous, mononematous, solitary or in a small group of 2–4, cylindrical, straight or slightly flexuous, 3–8-septate, dark brown, paler at the apical part, rounded at the apex. Conidiogenous cells 6–8 × 3–4 μm (x = 7 × 3 μm, n = 15), integrated, terminal, monoblastic, cylindrical, brown. Conidia 400–568 × 13–16 μm (x = 484 × 15 μm, n = 20), 6–7 μm at the narrowest apical region, acrogenous, solitary, elongate, obclavate, rostrate, multi-distoseptate, tapering towards the apex, truncate at the base, rounded at apex, dark olivaceous to dark brown, straight or slightly curved, guttulate, thick-walled, smooth, conidia percurrent proliferation which forms another conidium at the apex.

**Culture characteristics.** Conidia cultivated on PDA within 12h and germ tubes produced at the ends. Colonies on PDA, reaching 6 cm in 1 month at room temperature (25 °C). Mycelium loose, flocculent, smooth edge, brown to dark brown, dark brown on the reverse.

**Material examined.** THAILAND, Bangkok Province, Khwaeng Phra Khanong Nuea, 13°42’41”N; 100°36’03”E, on submerged decaying wood, 1 October 2017, Zonglong Luo, S–3083 (MFLU 21-0110, holotype), ex-type living culture (MFLUCC 18-0262).

**Notes.** Distoseptispora bangkokensis is comparable to *D. cangshanensis* and *D. multiseptata* in having elongate, obclavate, or rostrate conidia (Su et al. 2016; Hyde et al. 2016a; Yang et al. 2018). However, *D. bangkokensis* has shorter and narrower conidiophores than those of *D. cangshanensis* (37–55 × 3–4 μm vs. 44–68 × 4–8 μm), but has longer conidia (400–568 μm vs. 58–166 μm); *D. multiseptata* (MFLU 17-0856) is similar to *D. bangkokensis* in conidial morphology, with conidia mostly 300–600 μm long (up to 700 μm) and significantly longer than those of the holotype (up to 380 μm long). However, Yang et al. (2018) did not give a detailed description of *D. multiseptata* (MFLU 17-0856). Phylogenetic analyses showed that *D. bangkokensis* clustered with *D. bambusae, D. dehongensis, D. euseptata, D. lancangjiangensis, D. suoluoensis, D. thysanolaenae,* and *D. yunnanensis* with low bootstrap support (26%ML/0.53PP, Figure 1). *Distoseptispora bangkokensis* is distoseptate conidia, and it is easily distinguished from *D. bambusae, D. euseptata, D. lancangjiangensis, D. suoluoensis,* and *D. yunnanensis,* which are euseptate. *Distoseptispora bangkokensis* is resemble to *D. dehongensis* and *D. thysanolaenae* in having obclavate, distoseptae conidia, but are distinguished by conidia characteristics, *D. bangkokensis* has elongate, obclavate, rostrate, multi-distoseptat, and longer conidia than *D. dehongensis* (400–568 × 13–16 μm vs. 17–30 × 7.5–10 μm) and *D. thysanolaenae* (400–568 × 13–16 μm vs. 30–70 × 5–8 μm), respectively.

**Distoseptispora lancangjiangensis** H.W. Shen, H.Y. Su, K.D. Hyde & Z.L. Luo, sp. nov.

Index Fungorum Number No: IF558555
Facesoffungi Number No: FoF09994
Figure 3

**Etymology.** Referring to the collecting location, Lancangjiang River in China.
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Figure 3. *Distoseptispora lancangjiangensis* (KUN-HKAS 112712, **holotype**) A colonies on the substrate B conidiophore and conidium C-E conidiophores F, G conidiogenous cells H conidiogenous cell with conidium I-Q conidia R germinating conidium S, T culture on PDA. Scale bars: 50 μm (B-E); 20 μm (F-R).
Holotype. KUN-HKAS 112712

Description. Saprobic on submerged wood in freshwater River. Sexual morph: Undetermined. Asexual morph: Colonies effuse, hairy, glistening, brown to dark. Mycelium partly immersed in the substratum, composed of hyaline to pale brown, septate, branched hyphae. Conidiophores 144–204 × 5–6 μm (\(\bar{x} = 175 \times 6 \mu m\), n = 20) macronematous, mononematous, solitary, inflate at the base, cylindrical, straight or slightly flexuous, 6–11-septate, dark brown, hyaline and rounded at apex. Conidiogenous cells 12–24 × 4–5 μm (\(\bar{x} = 18 \times 5 \mu m\), n = 20) integrated, terminal, monoblastic, cylindrical, brown. Conidia 64–84 × 9–10 μm (\(\bar{x} = 74 \times 10 \mu m\), n = 20), acrogenous, solitary, narrowly obclavate or obspathulate, tracted at base, tapering towards apex, 3–10-euseptate, brown to dark brown, thin-walled, becoming paler or hyaline towards apex, guttulate, with a darkened scar at base, smooth-walled.

Culture characteristics. Conidia cultivated on PDA within 12h and germ tubes produced at the apex. Colonies on PDA, reaching 4.5 cm in 1 month at room temperature (25 °C). Mycelium loose, flocculent, smooth edges, convex middle, pale brown to dark brown on the surface of PDA. Smooth, black on the reverse.

Material examined. China, Yunnan Province, Dali City, Lancangjiang River, 22°36’36”N; 100°37’59”E, on submerged decaying wood, 20 July 2017, Qishan Zhou and Qingxiong Ruan S–1864 (KUN-HKAS 112712, holotype; MFLU 21-0111, isotype), ex-type living culture (DLUCC 1864 = CGMCC 3.20265).

Notes. Phylogenetic analysis showed that Distoseptispora lancangjiangensis clustered as a sister taxon to D. suoluoensis with 97%ML/0.98PP support. Distoseptispora lancangjiangensis is similar to D. suoluoensis in having long conidiophores, monoblastic conidiogenous cells, and obclavate to rostrate, euseptate conidia. However, D. suoluoensis has yellowish-brown or dark olivaceous, verrucose conidia, while in D. lancangjiangensis conidia are brown to dark brown and smooth-walled. Moreover, D. lancangjiangensis has smaller conidia than those of D. suoluoensis (64–84 × 9–10 μm vs. 80–125 × 8–13 μm) (Yang et al. 2018). Distoseptispora lancangjiangensis and D. bambusae have similar conidial shapes, but D. lancangjiangensis is having longer conidia (64–84 × 9–10 μm vs. 45–74 × 5.5–10 μm) and longer conidiophores (144–204 × 5–6 μm vs. 40–96 × 4–5.5 μm). Furthermore, D. bambusae has polyblastic or monoblastic conidiogenous cells and olivaceous or brown conidia, while D. lancangjiangensis only has monoblastic conidiogenous cells and brown to dark brown conidia (Sun et al. 2020).

Distoseptispora clematidis Phukhams., M.V. de Bult & K.D. Hyde, in Phukham-sakda et al., Fungal Diversity 102: 168 (2020)
Index Fungorum Number No: IF557301
Facesoffungi Number No: FoF07261
Figure 4

Description. Saprobic on submerged wood in freshwater River. Sexual morph: Undetermined. Asexual morph: Colonies on the substratum superficial, effuse, scattered, hairy, dark brown. Mycelium partly immersed in substrate, composed of branched, smooth,
New species and records of *Distoseptispora* from freshwater habitats

Figure 4. *Distoseptispora clematidis* (KUN-HKAS 112708) **A** colonies on the substratum **B-C** conidiophores with conidia **D** conidiogenous cells **E-H** conidia **I** germinating conidium **J** culture on PDA Scale bars: 30 μm (**B, C, E-I**); 20 μm (**D**).
septate, brown to dark brown hyphae. *Conidiophores* 30–41 × 5–6 μm ($\bar{x} = 36 \times 6$ μm, n = 15), macronematous, mononematous, single or in a small group, straight or slightly flexuous, unbranched, septate, erect, 2–4-septate, cylindrical, smooth, dark brown to brown. *Conidiogenous cells* 7–9 × 5–6 μm ($\bar{x} = 8 \times 5$ μm, n = 15), monoblastic, integrated, determinate, terminal, cylindrical, pale brown to brown. *Conidia* 83–220 × 12–14 μm ($\bar{x} = 151 \times 13$ μm, n = 20), acrogenous, solitary, obclavate, cylindrical, elongated, straight or curved, truncate at base, rounded at apex, 16–41-distoseptate, slightly constricted at some septa, smooth, brown to greenish-brown, thick-walled.

**Culture characters.** Conidia cultivated on PDA within 12h and germ tubes produced at the ends. Colonies on PDA, attaining 4 cm after 1 month at room temperature (25 °C), gray at first, later becoming dark gray, loose, flocculent, smooth edge, dark brown on the reverse.

**Material examined.** **China**, Yunnan Province, Kunming City, Yiliang County, Nanpanjiang River, 24°38′28″N; 103°09′38″E, on submerged decaying wood, 12 June 2018; Hongwei Shen and Xiu He, S–1797 (KUN-HKAS 112708), living culture (DLUCC 1797).

**Notes.** Our new isolate clustered with the ex-type strain of *Distoseptispora clematidis* (MFLU 17-1501) (Phukhamsakda et al. 2020) with 97%ML/0.99PP bootstrap support (Figure 1). *Distoseptispora clematidis* (MFLU 17-1501) was collected on dead culms of *Thysanolaena maxima* (Roxb. ex Hornem.) Honda in Yunnan Province, China. Based on morphological analysis, the size and shape of the conidia and conidiophores of our new isolate are similar to *D. clematidis*. Therefore, we identified our new isolate as *D. clematidis* and it is a new record from freshwater habitats in China.

**Distoseptispora thysanolaenae** Goonas., Dayarathne, Phookamsak & K.D.Hyde, in Phookamsak et al., Fungal Diversity 95: 126 (2019)

Index Fungorum Number No: IF555408
FACESoffungi Number No: FoF05011

**Figure 5**

**Description.** Saprobic on submerged wood in freshwater River. **Sexual morph:** Undetermined. **Asexual morph:** Colonies on the substratum superficial, effuse, scattered, hairy, dark brown. Mycelium partly immersed, composed of branched, septate, smooth, brown to dark brown hyphae. *Conidiophores* 41–59 × 4–5 μm ($\bar{x} = 50 \times 5$ μm, n = 20) macronematous, mononematous, unbranched, single, erect, straight or slightly curved, smooth, 3–6-septate, pale brown to brown. *Conidiogenous cells* monoblastic, integrated, determinate, terminal, dark brown, cylindrical. *Conidia* 46–87 × 9–12 μm ($\bar{x} = 67 \times 10$ μm, n = 25) acrogenous, solitary, dry, smooth, obclavate, elongated, straight or slightly curved, truncate at base, tapering towards apex, 6–19-septate, dark grayish-brown to light yellow-green, thick-walled.
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Figure 5. Distoseptispora thysanolaenae (KUN-HKAS 112710) A colonies on the substratum B-D conidiophores with conidia E, F conidiogenous cells G-N conidia O germinating conidium P, Q culture on PDA Scale bars: 30 μm (B-D); 10 μm (E, F); 20 μm (G-O).
**Culture characteristics.** Conidia cultivated on PDA within 12 h and germ tubes produced at the apex. Colonies on PDA, reaching 6 cm after 6 weeks at room temperature (25 °C). Mycelium loose, flocculent, neat edges, convex in middle, pale brown to dark brown. Black, smooth on the back.

**Material examined.** CHINA, Yunnan Province, Lushui City, Nujiang River, 26°23’12”N; 98°53’94”E, on submerged decaying wood, 3 May 2016, Zonglong Luo and Songming Tang, S-876 (KUN-HKAS 112710), living culture (DLUCC 876 = KUNCC 21-10710)

**Notes.** Our new collection is identical to *Distoseptispora thysanolaenae* in characters of the conidiophores, conidiogenous cell, and conidia (Phookamsak et al. 2019). Furthermore, our new isolate phylogenetically clusters with the ex-type strain of *D. thysanolaenae* (KUN-HKAS 102247) with 100%ML/1.00PP support (Figure 1). *Distoseptispora thysanolaenae* was collected from terrestrial habitats in China, while, our new isolate was collected from freshwater habitat in China. Therefore, we identified our new collection as *D. thysanolaenae*, and it is new to freshwater habitats in China.

**Discussion**

*Distoseptispora* has been reported from both freshwater and terrestrial habitats. Of these, species have been collected from freshwater environments (Su et al. 2016; Hyde et al. 2016a, 2019, 2020; Luo et al. 2018; Xia et al. 2017, 2019; Yang et al. 2018; Tibpromma et al. 2018; Crous et al. 2019; Phookamsak et al. 2019; Monkai et al. 2020; Phukhamsakda et al. 2020; Song et al. 2020; Sun et al. 2020; Li et al. 2021). To date, 18 species of *Distoseptispora* have been reported from Thailand, 14 species from China. In this study, we collected four *distoseptispora*-like taxa from rivers and streams in China and Thailand. Phylogenetic analysis showed that all four species were well-placed in *Distoseptispora* (Figure 1). Two new species and records are introduced based on morphological and phylogenetic analysis.

Species of *Distoseptispora* are highly diverse in morphology, especially the conidial shape. Conidia of most species are obclavate to cylindrical or rostrate (e.g. *D. aquatica*, *D. tectonae*, and *D. suoluoensis*), but a few are ellipsoid to subglobose (e.g. *D. martini*), lanceolate (e.g. *D. guttulata* and *D. multiseptata*), and some species have conidia with a sheath at the apex (e.g. *D. appendiculata*) (Hyde et al. 2016a; Su et al. 2016; Xia et al. 2017; Yang et al. 2018; Luo et al. 2018, 2019). Some species also differ in the conidiogenous cells (*D. palmarum*, *D. dehongensis*, and *D. bambusae* are monoblastic or polyblastic, while the others are monoblastic) and conidial septate (*D. bambusae*, *D. euseptatensis*, *D. guttulata*, *D. lignicola*, *D. rayongensis*, *D. suoluoensis*, and *D. yunnanensis* are euseptate, while other species are distoseptate) (Yang et al. 2018; Hyde et al. 2019; Luo et al. 2019; Sun et al. 2020; Dong et al. 2021; Li et al. 2021).

Based on the key morphological characteristics, *viz.*, conidiophores, conidiogenous cells, and conidia, Subramanian (1992) redistributed seven genera, *viz.*, *Sporidesmium*, *Polydesmus*, *Sporidesmiella*, *Stanjehughesia*, *Repetophragma*, *Penzigomyces*, and *Ellisem-
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*bia* to accommodate several *Sporidesmium*-like taxa. Based on multi-gene phylogenetic analysis and morphology, Su et al. (2016) introduced a new *Sporidesmium*-like genus *Distoseptispora*. Some *Sporidesmium*-like taxa were introduced in different lineages and synonymized *Ellisembia* under *Sporidesmium*. Although *Distoseptispora* was only introduced from submerged wood in freshwater habitat in 2016 (Su et al. 2016), the genus has previously been reported from both freshwater and terrestrial habitats as species in other genera. For example, Cai et al. (2002), Ho et al. (2001, 2002) and Luo et al. (2004) reported *Distoseptispora* as other species (*Ellisembia*, *Sporidesmiella*, and *Sporidesmium*) from submerged wood in freshwater habitats, and Kodsueb et al. (2016), Mena-Portales et al. (2016) and Zhou et al. (2001) reported from terrestrial habitats. However, none of these records had molecular data and it is impossible to consider the placement of these isolates. In these species *distoseptispora/sporidesmium*-like genera, it is therefore better to describe taxa based on molecular data.

Based on phylogenetic analysis, Xia et al. (2017) transferred *Acrodictys martinii* to *Distoseptispora* as *Distoseptispora martinii*. The species is characterized by solitary erect, unbranched conidiophores, monoblastic conidiogenous cells with percurrent extensions and subhyaline to pale brown and solitary, transversal ellipsoid, oblate or subglobose, muriform conidia, separated by septa, sometimes with pores in the septa and pale brown to brown. However, the current understanding of *Distoseptisporaceae*, *D. martinii* is significantly different from other *Distoseptispora* taxa; thus, needs to be verified in the future (Luo et al. 2018; Sun et al. 2020).

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