Diatom modulation of select bacteria through use of two unique secondary metabolites

Ahmed A. Shibli, Ashley Isaac, Michael A. Ochsenkühn, Anny Cárdenas, Cong Fei, Gregory Behringer, Marc Arnoux, Nizar Drou, Miraflo P. Santos, Kristin C. Gunsalus, Christian R. Voolstra, and Shady A. Amin

Unicellular eukaryotic phytoplankton, such as diatoms, rely on microbial communities for survival despite lacking specialized compartments to house microbiomes (e.g., animal gut). Microbial communities have been widely shown to benefit from diatom excretions that accumulate within the microenvironment surrounding phytoplankton cells, known as the phycosphere. However, mechanisms that enable diatoms and other unicellular eukaryotes to nurture specific microbiomes by fostering beneficial bacteria and repelling harmful ones are mostly unknown. We hypothesized that diatom exudates may tune microbial communities and employed an integrated multomics approach using the ubiquitous diatom Asterionellopsis glacialis to reveal how it modulates its naturally associated bacteria. We show that A. glacialis reprograms its transcriptional and metabolic profiles in response to bacteria to secrete a suite of central metabolites and two unusual secondary metabolites, rosmarinic acid and azelaic acid. While central metabolites are utilized by potential bacterial symbionts and opportunists alike, rosmarinic acid promotes attachment of beneficial bacteria to the diatom and simultaneously suppresses the attachment of opportunists. Similarly, azelaic acid enhances growth of beneficial bacteria while simultaneously inhibiting growth of opportunistic ones. We further show that the bacterial response to azelaic acid is numerically rare but globally distributed in the world’s oceans and taxonomically restricted to a handful of bacterial genera. Our results demonstrate the innate ability of an important unicellular eukaryotic group to modulate select bacteria in their microbial consortia, similar to higher eukaryotes, using unique secondary metabolites that regulate bacterial growth and behavior inversely across different bacterial populations.

Significance

Phytoplankton are major primary producers in the marine environment that excrete a wide range of metabolites. These exudates support the growth of surrounding bacteria that in turn provide phytoplankton cells with resources and growth cofactors to proliferate. Unlike multicellular eukaryotes with dedicated structures housing microbiomes, mechanisms enabling unicellular eukaryotic phytoplankton hosts to modulate potential symbionts and opportunists within a natural microbial community are unknown. Exposure of a host phytoplankton cell to its natural microbial community triggers major transcriptional and metabolic reprogramming to release unique secondary metabolites that selectively enable growth and attachment of symbiotic taxa while simultaneously suppressing the colonization of nonsymbiont bacteria. These results suggest strong and highly selective microbiome-modulating strategies shared across the unicellular and multicellular eukaryotic lineages.

Author contributions: A.A.S., A.I., M.A.O., G.B., C.R.V., and S.A.A. designed research; A.A.S., A.I., M.A.O., A.C., C.F., G.B., M.A., N.D., and M.P.S. performed research; A.A.S., A.I., M.A.O., A.C., C.F., N.D., and S.A.A. analyzed data; and A.A.S., A.I., M.A.O., A.C., K.C.G., C.R.V., and S.A.A. wrote the paper.

The authors declare no competing interest.

This article is a PNAS Direct Submission. This open access article is distributed under Creative Commons Attribution-NonCommercial-NoDerivatives License 4.0 (CC BY-NC-ND).

First published October 16, 2020.

www.pnas.org/cgi/doi/10.1073/pnas.2012088117

PNAS | November 3, 2020 | vol. 117 | no. 44 | 27445–27455
natural system derived from the environment by using multiotics to show that DOM secretions by the globally widespread diatom Asterionellopsis glaciica (20) modulate select bacterial behavior and growth. We hypothesize that diatom cells must adopt specific mechanisms to promote association with potentially beneficial symbionts while repelling opportunists to offset the lack of specialized compartments to house microbiomes. To this end, A. glaciica strain A3 was cultivated from its natural environment, then freed of its associated bacteria and left to acclimate until the time of reseeding, marked by the reintroduction of its natural bacterial consortium to the diatom. Transcriptional and metabolomic changes in both the diatom and the bacterial consortium at different time points were assessed, and potential representative symbiotic and opportunistic bacteria were cultivated from the consortium to further confirm hypotheses generated from multiotics experiments.

Results

To examine the interactions between the diatom and its bacterial consortium, we isolated A. glaciica A3 along with its natural microbial community (xenic A. glaciica), then cured it of bacteria using a suite of antibiotics to make it axenic, as described previously (21) (SI Appendix, Supplementary Methods). After ~170 generations of acclimating the axenic A. glaciica A3 culture to the absence of bacteria, the true bacterial consortium composition was harvested by filtration from xenic cultures immediately before the reseeding experiment. At the time of reseeding, one portion of this natural bacterial community was added to the acclimated axenic A. glaciica A3 culture, generating a reseeded A. glaciica A3 treatment to investigate the response of the diatom to bacterial exposure and the response of bacteria to diatom exudates (SI Appendix, Fig. S1). Two additional portions of the bacterial consortium were collected and used for shotgun metagenomics and metatranscriptomics (bacterial consortium control at 0.5 h). Diatom transcriptomic samples (at 0.5 and 24 h) were collected from the control axenic A. glaciica cultures and reseeded A. glaciica treatments. In addition, samples for metabolomics at two early (0.5 and 4 h) and two late (24 and 48 h) time points were collected (SI Appendix, Supplementary Methods and Fig. S1).

The composition of the microbial consortium collected at the time of reseeding showed the dominance of six bacterial families, with Flavobacteriaceae comprising 41% of all database reads, followed by Rhodobacteraceae (16%), Erythrobacteraceae (16%), Alteromonadaceae (9.28%), Pseudomonadaceae (10%), and Oceanospirillaceae (10.3%: Fig. 1A). To uncover how these families responded to diatom exudates, we assembled 10 near-complete bacterial genomes from the microbial consortium metagenome. The metagenomically assembled genomes (MAGs) belonged to most major families in the consortium, including Flavobacteriaceae (MAG9), Rhodobacteraceae (MAG3, MAG5, MAG6, and MAG11), Erythrobacteraceae (MAG10), Alteromonadaceae (MAG4 and MAG12), Oceanospirillaceae (MAG8), and Halomonadaceae (MAG13; SI Appendix, Table S1). Mapping metatranscriptomic reads to all MAGs showed that the four Rhodobacteraceae MAGs recruited ~41% of mRNA reads and were responsible for the majority of differentially expressed genes (SI Appendix, Table S2) at both early and late time points of reseeded samples relative to controls, despite representing ~10% of the bacterial consortium metagenome (SI Appendix, Table S1).

We examined the consortium metatranscriptome and confirmed that the Rhodobacteraceae (hereafter Roseobacters) exhibited the most transcriptionally rapid and diverse response within 0.5 h of reintroduction to the diatom, as evidenced by the large number of Gene Ontology terms associated with Roseobacter genes expressed after reseeding. In stark contrast, other bacterial families either displayed no significant response to reseeding (Flavobacteriaceae), displayed a decreasing response from 0.5 to 24 h (Erythrobacteraceae), or were responsive only at 24 h after reseeding (Alteromonadaceae, Pseudomonadaceae, and Oceanospirillaceae) relative to the consortium control (Fig. 1A).

The A. glaciica A3 transcriptome showed a major reprogramming of its transcriptional profile to differentially express ~14% of its protein-coding genes relative to axenic controls, coupled with temporal shifts in expression patterns (Fig. 1B). In response to consortium reseeding, transcripts for amino acid biosynthesis and fatty acid degradation were consistently up-regulated, while nitrate assimilation, photosynthesis, and carbon fixation were down-regulated throughout the reseeding experiment. At 0.5 h only, differentially up-regulated A. glaciica A3 transcripts included those for spermidine biosynthesis and transport and the tricarboxylic acid (TCA) and urea cycles, while transcripts for methionine biosynthesis and uroase activity were differentially up-regulated at 24 h only. We also observed down-regulated transcripts involved in the Calvin cycle at both 0.5 h and 24 h and tryptophan biosynthesis-related transcripts down-regulated at 24 h only (Fig. 1B and SI Appendix, Table S3).

The diatom and Roseobacter transcriptomic responses were coupled to major changes in the exometabolome. Exometabolomes sampled at two early and two late time points after reseeding (SI Appendix, Fig. S1B) were analyzed using a quadrupole time-of-flight mass spectrometer (Dataset S1). The DOM loadings varied between axenic and reseeded samples (Fig. S2B). Interestingly, based on Mahalanobis distances (Md), the DOM composition at early time points was significantly more distinct from late time points in the reseeded samples (Md = 3.88) than in axenic controls (Md = 3.06; Fig. 2 B and C), suggesting that DOM is temporally highly dynamic in response to consortium reseeding, similar to the diatom transcriptome. Analysis of the DOM elemental composition of extracted metabolites in axenic and reseeded samples using Fourier-transform ion cyclotron resonance mass spectrometry (FT-ICR-MS; Datasets S2 and S3) showed ~50% decrease in abundance of dissolved organic nitrogen (DON) in reseeded samples relative to axenic controls (SI Appendix, Fig. S2).

The identity of 28 metabolites common in axenic and reseeded diatom samples was confirmed (Fig. 2 D and SI Appendix, Table S4) using an in-house chemical library of >600 molecules (SI Appendix, Supplementary Methods), indicating these metabolites were present at the time of colony transfer by the diatom. A complete list of all chemical compounds identified in axenic and reseeded samples as a function of time, but a markedly lower overall accumulation in reseeded samples relative to axenic controls (e.g., leucine, threonine, 3-phosphoglucose), suggesting either diatom down-regulation of the biosynthesis of these molecules in reseeded samples and/or bacterial uptake in reseeded samples. Bacterial uptake was corroborated by the transcriptional response of the diatom to reseeding, which showed up-regulation of metabolite-specific biosynthesis genes and a concomitant up-regulation of specific Roseobacter transporters that take up these metabolites (SI Appendix, Table S5). Seven metabolites showed increases in relative abundance in reseeded samples compared to axenic controls (e.g., rosmarinic acid), suggesting either a signaling role for these diatom metabolites or coproduction by bacteria (Fig. 2D).

Based on the rapid response of the Roseobacters to diatom exudates, we built a conceptual model of diatom–Roseobacter interactions using the differential gene expression of A. glaciica A3, three Roseobacter MAGs (MAG3, MAG5, and MAG6), and identified exometabolites (Fig. 3 and SI Appendix, Tables S3–S5). In response to reseeding, the diatom up-regulated genes involved in the biosynthesis of spermidine (log2 fold-change = 3.8, P = 0.09) and its transport (log2 fold-change = 2.7, P = 0.04) at 0.5 h. Concomitantly, transcripts for spermidine uptake were up-regulated in both MAG3 (log2 fold-change = 5.5, P = 0.09) and MAG5 (log2 fold-change = 6.2, P = 0.08) at 0.5 h. The diatom increased transcription of glutamate dehydrogenase at
0.5 h (log₂ fold-change = 2.2, P = 0.079) and 24 h (log₂ fold-change = 5.4, P = 0.006) to fuel the TCA cycle and/or the urea cycle, both of which were up-regulated, by generating α-ketoglutarate and ammonia, respectively. Citrulline, a urea cycle intermediate released into the media, showed a differential decrease in abundance in reseeded samples versus axenic samples (P = 0.04 at 24 h; Fig. 2D), suggesting bacterial uptake. The diatom down-regulated homologs of phosphoglycerate kinase (22) involved in the conversion of 3-phosphoglycerate (3-PGA) to glyceraldehyde 1,3-diphosphate in the plastid (log₂ fold-change = -1.6, P = 0.06) and cytoplasm (log₂ fold-change = -5.3, P = 0.06). The 3-PGA transporters localized in the plastid were also down-regulated at 0.5 h after reseeding (log₂ fold-change = -7.0 and -4.0, P = 0.002 and P = 0.007, respectively), suggesting reduced transport of 3-PGA across the plastid membrane and a buildup of 3-PGA in the cytoplasm. The 3-PGA was released into the media and was presumably taken up by bacteria. Transporters for 3-PGA were not differentially expressed in MAG3, while a 3-PGA response regulator was up-regulated in MAG5 at 0.5 h (log₂ fold-change = 4.8, P = 0.03) or not differentially expressed suggest the diatom may be secreting the amino acids threonine and/or leucine. The secretion of threonine and leucine (P = 0.003 at 4 h; Fig. 2D) into the media was concordant with an up-regulation of their transporters and subsequent assimilation of leucine into branched-chain fatty acid biosynthesis in the three Roseobacter MAGs (Fig. 3 and SI Appendix, Tables S3 and S5).

To confirm the ability of Roseobacters to utilize diatom metabolites, we isolated bacteria from the bacterial consortium and sequenced their genomes (SI Appendix, Supplementary Methods). Two isolates were identified as Roseobacter species (Sulfitobacter pseudonitzschiiae F5 and Phaeobacter sp. F10) and one isolate as an Alteromonadaceae species (Alteromonas macleodii F12). The average nucleotide identities (ANIs) of A. macleodii F12 were 76.8% with MAG12 and 89.9% with MAG4, while the ANI of Phaeobacter sp. F10 and MAG6 was 99.9%, suggesting MAG6 represents the same species as Phaeobacter sp. F10. The average amino acid identity between S. pseudonitzschiiae F5 and all Roseobacter MAGs ranged from 61% to 66%, suggesting that they group at the genus level (cutoff 60% [23]). Phylogenetic analysis of isolate genomes and MAGs further confirmed this finding, as it clustered Phaeobacter sp. F10 close to MAG6 (SI Appendix, Fig. S3 and Table S6), while A. macleodii F12 clustered within the A. macleodii clade (SI Appendix, Fig. S4 and Table S7). Metagenomic read recruitment analysis using the three bacterial genomes indicated that, while S. pseudonitzschiiae F5 represented only 0.16% of metagenomic reads, Phaeobacter sp. F10 and A. macleodii F12 represented 11.41% and 2.82% of all metagenomic reads, respectively. In addition, mapping bacterial mRNA reads to the isolates showed that S. pseudonitzschiiae F5 recruited 5.3% of mRNA reads, Phaeobacter sp. F10 34.1%, and A. macleodii F12 9.1%, suggesting these bacteria are major players in the microbial community from a transcriptional perspective. Subsequently, 16 diatom metabolites from Fig. 2D were used to test the ability of S. pseudonitzschiiae F5 and A. macleodii F12 (classified as a potential symbiont and an opportunist, respectively, as discussed below) to utilize these metabolites.
as growth substrates. Despite the more rapid transcriptional responses of Roseobacters to reseeding (Fig. 1), both bacterial isolates were able to use most of these central metabolites as growth substrates (SI Appendix, Fig. S5).

We sought to examine if diatom secondary metabolites can account for the advantage Roseobacters have over other bacterial families in the microbial consortium, like the Alteromonadaceae. Cell attachment is an important mechanism used by bacteria to remain in the phycosphere to enhance access to diatom exudates (24). The motility of S. pseudonitzschiae F5, Phaeobacter sp. F10, and A. macleodii F12 was examined in the presence of a secondary metabolite not detected in diatoms before, rosmarinic acid, a common constituent of some terrestrial plants (25). Surprisingly, 2 μM rosmarinic acid significantly inhibited the motility of the potential symbionts S. pseudonitzschiae F5 and Phaeobacter sp. F10 and increased the motility of the potential opportunist A.

Fig. 2. SPE-extracted DOM profile is highly influenced by reseeding. (A–C) Principal components analysis (PCA) plots of axenic and reseeded untargeted exometabolome samples. PCA was performed based on Mahalanobis distances (M_D) comparing 1,237 SPE-extracted molecules between (A) axenic vs. reseeded samples and (B and C) early (0.5 and 4 h) and late (24 and 48 h) time points for (B) axenic and (C) reseeded conditions. Circles represent technical replicates (n = 3) of three biological replicates. (D) Euclidean hierarchical clustering of 28 exometabolites (SI Appendix, Table S4) identified in axenic and reseeded samples and confirmed using a library of in-house chemical standards. Colors represent average normalized relative abundance of each metabolite. Time points marked with an asterisk indicates a significant change in relative abundance across reseeded and axenic samples as determined with a Student’s t test (Bonferroni-adjusted P < 0.05). (i) Prospective refractory diatom metabolites, (ii) diatom metabolites possibly taken up by the consortium, and (iii) diatom metabolites with a potential signaling role.

as growth substrates. Despite the more rapid transcriptional responses of Roseobacters to reseeding (Fig. 1), both bacterial isolates were able to use most of these central metabolites as growth substrates (SI Appendix, Fig. S5).

We sought to examine if diatom secondary metabolites can account for the advantage Roseobacters have over other bacterial families in the microbial consortium, like the Alteromonadaceae. Cell attachment is an important mechanism used by bacteria to remain in the phycosphere to enhance access to diatom exudates (24). The motility of S. pseudonitzschiae F5, Phaeobacter sp. F10, and A. macleodii F12 was examined in the presence of a secondary metabolite not detected in diatoms before, rosmarinic acid, a common constituent of some terrestrial plants (25). Surprisingly, 2 μM rosmarinic acid significantly inhibited the motility of the potential symbionts S. pseudonitzschiae F5 and Phaeobacter sp. F10 and increased the motility of the potential opportunist A.
macleodii F12 (Fig. 4). To confirm whether reduced motility enables the symbionts to attach to the diatom, A. glacialis A3 was cocultured with each bacterial isolate. Indeed, S. pseudonitzschiæ F5 and Phaeobacter sp. F10 exhibited strong attachment in the diatom phycosphere while A. macleodii F12 showed no apparent attachment (Fig. 4).

In addition to rosmarinic acid, 100 μM azelaic acid, a byproduct of oleic acid metabolism, significantly inhibited the growth of A. macleodii F12 over a 24-h period, while the same concentration promoted growth of potential symbionts over a 48-h period (Fig. 5 A–C). Bacterial response to azelaic acid was shown to be controlled by a transcriptional regulator, AzeR (26). To shed light on the prevalence of the bacterial response to azelaic acid throughout the oceans, a hidden Markov model (HMM) profile of AzeR homologs detected in all three bacterial isolates was used to search the Tara Oceans database. The average abundance of AzeR reads relative to the total reads in the database was 0.03%. AzeR homologs were consistently distributed at surface and deep chlorophyll maximum depths across the oceans, with most homologs belonging to Alteromonadales (19%) and Rhodobacterales (18%; Fig. 5D). Mining the Pfam database for AzeR homologs indicated that the response to azelaic acid in publicly available bacterial genomes is mostly limited to the Proteobacteria phylum and is mostly restricted to six orders, including Alteromonadales and Rhodobacterales, to which the Alteromonadaceae and Roseobacters belong, respectively (SI Appendix, Fig. S6).

**Discussion**

Remineralization of phytoplankton-derived organic matter by heterotrophic bacteria plays a major role in the carbon cycle and accounts for the transformation of ~20 gigatons of carbon per year in the ocean’s euphotic zone (27). Our current understanding...
of the global passive and active release of DOM by phytoplankton has been largely studied in the context of primary production, grazing events, and virus-mediated cell lysis (28, 29). Still, underlying reasons for the active excretion by phytoplankton of significant amounts of low molecular weight organic compounds into the phycosphere (6) are still being debated (27, 30). Because of their microscopic size, transport of molecules around phytoplankton cells is mostly governed by diffusion, which leads to the accumulation of phytoplankton-derived DOM within the phycosphere (13). Bacteria in the ocean expend significant energy to track and colonize these DOM-rich hotspots to fuel their growth (31), employing a variety of mechanisms to succeed in the phycosphere, including establishing symbiotic exchanges with phytoplankton cells or producing algicidal agents that harm or kill phytoplankton (7, 32, 33). Therefore, it is imperative for phytoplankton cells to control the types of bacteria that come in contact with the phycosphere, as the outcome ultimately leads to survival or death. However, the mechanisms that enable ocean-drifting phytoplankton cells to attract beneficial bacteria and repel harmful ones in the phycosphere, if any, are mostly unknown.

The microbial community composition surrounding A. glacialis A3 is typical of bacteria associated with diatom cultures and blooms (34–38). Flavobacteria, the dominant lineage in the natural bacterial community associated with the diatom, often assimilate complex organic matter (e.g., polysaccharides) that require exoenzyme activity (39), especially during phytoplankton blooms (40), partially explaining their inactivity over shorter times with A. glacialis (<24 h; Fig. 4). Within 0.5 h of reintroducing the natural consortium to the axenic diatom culture, Roseobacters rapidly dominated the bacterial transcriptional activity (Fig. 4). The Roseobacter group spans >70 genera (41) with a highly versatile genetic repertoire (42, 43) that often dominates microbial assemblages surrounding particulate organic matter (44–46). They have been consistently shown to establish specific symbiotic relationships with diatoms (17, 18, 47) and are especially adept at acquiring phytoplankton-derived DOM (38, 48, 49). Despite their rapid response to A. glacialis A3 exudates relative to all other families, members of the Roseobacter group only represented 16.6% of the microbial consortium of the diatom, which is in line with the average Roseobacter group abundance in phytoplankton blooms (38). This discrepancy is potentially due to competition and chemical warfare between different bacterial taxa in the consortium, manifested by the overexpression of antibiotic resistance genes in all Roseobacter MAGs (Fig. 3 and SI Appendix, Table S5), which mitigates proliferation of any one bacterial group in the phycosphere. Indeed, production of diverse antimicrobial agents in a complex microbial community has been shown to maintain bacterial diversity (50), which explains why, despite being the most active, Roseobacters cannot solely dominate the phycosphere of A. glacialis A3. Interestingly, the microbial community composition of A. glacialis A3 based on metagenomics is different from the original microbial composition recovered with A. glacialis shortly after cultivation from the field based on 16S rRNA gene amplicon sequencing (21). While dominant families

![Fig. 4. Diatom secondary metabolite rosmarinic acid reduces motility and promotes attachment of potential Roseobacter symbionts to A. glacialis A3.](image-url)
like Rhodobacteraceae were common, they displayed large variation among the two datasets, either due to changes in the microbial community during laboratory cultivation or biases in DNA isolation and PCR amplification associated with amplicon sequencing.

Isolation and genome sequencing of *S. pseudonitzschiae* F5, *Phaeobacter* sp. F10, and *A. macleodii* F12 from the natural diatom microbial consortium provides an ample prospect to better understand phytoplankton modulation of different bacterial taxa in the phycosphere. Sulfitobacter and *Phaeobacter* are considered beneficial bacteria to diatoms, dinoflagellates, and macroalgae as they provide a variety of essential nutrients and cofactors that confer metabolic advantage to their hosts (17, 18, 51–56). Remarkably, several *S. pseudonitzschiae* strains (16S rRNA gene sequence identity >97%) have been isolated from several diatom species originating from different oceanic regions (18, 57, 58). One such strain, *S. pseudonitzschiae* SA11 (clustered near *S. pseudonitzschiae* F5; SI Appendix, Fig. S3), is a known diatom symbiont that enhances cell division of another diatom, *Pseudo-nitzschia multiserias*, via the hormone indole-3-acetic acid (18).

Preliminary growth experiments between *S. pseudonitzschiae* F5 and *A. glacialis* A3 indicate that it also enhances *A. glacialis* cell division, similar to *S. pseudonitzschiae* SA11 and *P. multiserias* (59). Thus, the current body of literature on the beneficial effects *Sulfitobacter* and *Phaeobacter* have on diatoms suggest they may be symbionts. *Alteromonas* has been labeled as a genus with an opportunistic lifestyle (60) due to its proliferation in resource-rich waters and during phytoplankton blooms (61, 62). They are able to outgrow native bacterial communities in coastal areas (63) and degrade a variety of algal-derived molecules such as silica (64), nutrients released during decaying phytoplankton blooms (65), and a variety of diatom-derived polysaccharides (66, 67). More specifically, *A. macleodii* competes with diatoms for nitrate (68), while closely related *Alteromonas* species contribute to the lysis of dinoflagellates (69) and produce an array of algicidal compounds effective against diatoms and other phytoplankton lineages (70–72). The placement of *A. macleodii* F12

![Fig. 5. Azelaic acid inhibits growth of *A. macleodii* F12 and promotes growth of Roseobacters. Growth of (A) *S. pseudonitzschiae* F5, (B) *Phaeobacter* sp. F10, and (C) *A. macleodii* F12 on 10% marine broth supplemented with 100 μM azelaic acid (squares) compared to controls (circles). Error bars represent SD of three replicates. Significance was determined by Student’s *t* test: *P* < 0.05 and **P** < 0.001. (D) Bacterial response to azelaic acid is geographically widespread throughout the oceans. The relative abundance of reads of the azelaic acid transcriptional regulator, AzeR, in the Tara Oceans database is 0.03%. The total percentage abundance of AzeR homologs according to their taxonomic distribution is shown in the top right box. Rhizobiales makes up the majority of hits (39%) in the “other” group. The color-coded donut plots represent the percentage taxonomic abundance of AzeR homologs from all size fractions (0 to 3 μm) at the surface (inner circle) and deep chlorophyll maximum (outer circle) from the Tara Oceans Microbiome Reference Gene Catalog. Numbers refer to the Tara Oceans stations; single donut plots depict surface samples only.

---

Shibl et al. PNAS | November 3, 2020 | vol. 117 | no. 44 | 27451
within the *A. macleodii* clade (SI Appendix, Fig. S4) suggests it is a common copiotrophic opportunistic opportunist (60). The conceptual model presented here (Fig. 3) clearly identifies the transcriptional and metabolomic responses of the host diatom and the surrounding Roseobacters. The combination of multiomics, bacterial isolation, and examination of the effects of different metabolites on these bacterial isolates provides several lines of evidence to support our conclusions. For example, up-regulation of the biosynthesis of metabolites (Fig. 3) by the diatom in response to reseeding is corroborated by the detection of these metabolites in the exometabolome, bacterial transcriptional responses toward these metabolites supported by metatranscriptomics, and growth experiments of bacterial isolates representing the *Roseobacter* group in the presence of these metabolites. Although we were not able to detect polyamines (e.g., spermidine) presumably produced by the diatom in our metabolome, the up-regulation of genes involved in spermidine uptake by MAG3 and MAG5 suggests that these diatom N-rich molecules may be rapidly utilized by the Roseobacters. Consistent with this observation, genes related to polyamine transformation were shown to be expressed mostly by Roseobacters in coastal waters, where diatoms usually dominate phytoplankton composition (73). In addition to spermidine, the rapid depletion of DON relative to DOC in the reseeded exometabolome (SI Appendix, Fig. S2) is supported by previous findings showing that labile N-containing compounds are preferentially utilized by Roseobacters in estuarine waters (74). These observations suggest DON is more labile than dissolved organic carbon in the phycosphere. The significant decrease in abundance of another DON molecule, citrulline, after the reseeding of bacteria implies its potential uptake (Fig. 2D). Although citrulline has been shown to support bacterial growth as a sole carbon source, uptake mechanisms have not been yet identified (75), complicating our ability to confirm bacterial uptake. However, growth of *S. pseudonitzschiae* F5 on citrulline confirms the ability of some members of the *Roseobacter* group to use it as a carbon source (SI Appendix, Fig. S5).

Of 1,237 detected metabolites, we were able to confirm the presence of 28 using a custom-curated chemical library of >600 biomolecules (Fig. 2D). Many of these confirmed metabolites have never been shown to be produced by diatoms before, suggesting they may be a rich source of metabolites in the ocean. In addition to several central metabolites, we observe the release of obscure secondary metabolites such as quinolinecarboxylic acid, 3-methylglutaric acid, suberic acid, and carnosine (Fig. 2D), which may play a role in symbiotic interactions or defense with different marine bacteria. Interestingly, other confirmed metabolites that have not been shown to be produced by diatoms before, such as rosarinic acid, azelaic acid, salicylic acid, hippurate, and N-acetyl-galactosamine (Fig. 2D and SI Appendix, Table S4), are involved in plant defense and inter-kingdom signaling mechanisms (76, 77). Production and secretion of these metabolites by the diatom hints at a defense system response (78) akin to land plants. Rosmarinic acid is one of the most frequently occurring defense compounds used by plants (25) and seagrasses (79), with a well-characterized biosynthetic pathway in plants (80). Similarly, azelaic acid is known as a natural signaling compound that induces systemic changes in plant defense mechanisms (81) and is produced by a marine fungus (82) and marine angiosperms (83). Other identified metabolites that display a shift in concentration after reseeding (Fig. 2D) may have important biological functions, including supporting bacterial growth (SI Appendix, Fig. S5). The significant shift in metabolic activity over time as the diatom host came in contact with the microbial consortia (Fig. 2 A–C and SI Appendix, Fig. S2) raises the question of the presence of more specialized compounds that potentially aid in shaping the phytoplankton microbiome. We sought to validate our hypothesis by examining the bacterial response to two of these secondary metabolites, rosmarinic acid and azelaic acid, using the isolated strains.

Rosmarinic acid was one of seven molecules that showed an increase in relative abundance within 0.5 h of reseeding relative to axenic controls (*P* = 0.025 and *P* = 0.006 at 0.5 and 48 h, respectively; Fig. 2D). This increase in abundance is either due to up-regulation of its biosynthesis by the diatom in response to reseeding, suggesting an interkingdom signaling function, or due to bacterial coproduction. Bacterial coproduction can be ruled out given that rosmarinic acid is only known to be produced by some land plants and seagrasses and has never been shown to be produced by prokaryotes (84). We mined the diatom genome for rosmarinic acid biosynthesis genes using plant homologs but were unable to find any matches, suggesting that diatoms may use a unique biosynthesis pathway different from legumes. Interestingly, rosmarinic acid significantly suppressed motility and promoted attachment of potential symbionts but had the opposite effect on *A. macleodii* F12 (Fig. 4). Rosmarinic acid was recently reported to be produced by *Arabidopsis thaliana* as a mimic of pathogenic bacterial quorum sensing autoinducers (85). It is likely that rosmarinic acid is also interfering with bacterial quorum sensing to control bacterial motility and attachment in the phycosphere, a hypothesis that appears to be supported by recent findings (59).

Azelaic acid, a C9-dicarboxylic acid and a byproduct of oleic acid metabolism, is also produced by the diatom (Fig. 2D). Azelaic acid primes plant defenses (86) and leads to the production of another defense signal, salicylic acid (87), which is also released by the diatom (Fig. 2D). The decrease in abundance of azelaic acid in reseeded exometabolomes (*P* = 0.0002, *P* = 0.001, and *P* = 0.018 at 0.5, 4, and 48 h, respectively; Fig. 2D) and its influence on growth of bacterial isolates suggest that the compound was assimilated by Roseobacters and Alteromonadaceae. Suberic acid (C8-dicarboxylic acid), a closely related metabolite also produced by the diatom, promotes the growth of *S. pseudonitzschiae* F5 and *A. macleodii* F12 alike (SI Appendix, Fig. S5). The similar structure and activity of both congeners suggest that, while azelaic acid inhibits growth of Alteromonadaceae, suberic acid may inhibit the growth of other bacteria. While such a strategy may enable diatoms to modulate different bacterial groups, Roseobacters gain an apparent advantage by utilizing a unique biosynthesis pathway for substrates from diatoms. Analysis of transporters in the genomes of *S. pseudonitzschiae* F5, *Phaeobacter* sp. F10, and *A. macleodii* F12 indicate that the Roseobacters possess a significantly higher number of transporters normalized to genome size relative to *A. macleodii* F12 (59). The mechanism of growth inhibition and promotion by azelaic acid remains unknown, and further work is needed to reveal its mechanism of action.

Bacteria in the phycosphere observe orders-of-magnitude higher concentrations of metabolites than cells outside (31). In contrast, these metabolites in bulk seawater potentially have much lower concentrations that fall within the nanomolar to picomolar range. This discrepancy between effective metabolite concentrations in the phycosphere and their measured concentrations in the environment and laboratory cultures is a byproduct of our inability to measure concentrations directly in the phycosphere. Recent findings show that bacterial community assembly in synthetic phycospheres can be predicted from the linear combination of taxa supported by growth on single phytoplankton central metabolites (88). Our findings further expand on our understanding of the role of metabolites in the phycosphere by incorporating host response to presence of different bacterial groups, manifested in the secretion of two unique secondary metabolites. Secretion of secondary metabolites by multicellular eukaryotes to modulate their microbiomes has been broadly reported (89–91). The ability of diatoms (and presumably other unicellular eukaryotes) to exert control over select microbial associates, suggesting a capacity to nurture microbiomes, may have
evolved earlier than the rise of multicellularity in eukaryotes. More interestingly, the ability of diatom-derived metabolites to have opposite phenotypic and/or behavioral effects on two different bacterial populations has not been widely shown, to our knowledge. This ability hints at complex evolutionary trajectories of how diatoms evolved the use of these metabolites and the role of secondary metabolism in interkingdom signaling. Further work is needed to characterize the mechanisms of action of these unique molecules in bacteria and to further identify other diatom metabolites and their role in modulating bacterial populations. Shedding light on these mechanisms has the potential to expand our understanding of food web dynamics and the role of phospho-sphere bacteria in carbon cycling.

Summary. Multicellular eukaryotes use diverse strategies to recruit and modulate microbiomes in specialized developmental organelles, such as the mammalian gut (92). In contrast, unicellular eukaryotes such as diatoms lack specialized organelles to house microbiomes, and, despite numerous observations that they possess unique microbial communities (93–95), it is not clear how they can modulate transient microbes. We show that, in addition to phytoplankton-derived central metabolites accessible to bacteria, the diatom A. glacialis A3 employs two unique secondary metabolites to promote the proliferation of select bacteria and demote others. The functional roles of signaling of secondary metabolites in marine environments are an important piece of the puzzle linking symbiotic exchanges between phytoplankton and bacteria with carbon cycling in the euphotic zone. Although signaling molecules are believed to constitute a minor fraction of DOM in the euphotic zone, their regulation of microbial metabolism and growth means they can exert a major influence on carbon cycling. This study provides a glimpse into the potential evolution of molecules from the same algal source that have opposite effects on two different groups of bacteria but a favorable outcome for the host. Such an efficient strategy to achieve two outcomes on symbionts and nonsymbionts in the euphotic zone (6, 96) suggests that microalgae and other unicellular eukaryotes modulate microbiomes.

Methods

Full details of the reseeding experimental design, bacterial isolation, DNA/RNA sequencing, exometabolite extraction, motility/growth assays, phylogenetic analysis, microscopy, and computational analysis are described in SI Appendix, Supplementary Methods. Briefly, the diatom A. glacialis strain A3 was isolated along with its natural bacterial community as previously described (21). A. glacialis A3 cultures were made axenic using antibiotics and left to acclimate in the absence of bacteria. To reseed the acclimated axenic A. glacialis A3 cultures, we used a consortium stock harvested from xenic A. glacialis A3 cultures after removing diatom cells. Ten near-complete bacterial genomes were assembled from the consortium metagenome. Subsequently, metatranscriptomes from the consortium were used to uncover how different bacterial families responded to diatom exudates and further examine the interactions between A. glacialis A3 and its natural bacterial associates. The A. glacialis A3 transcriptome was analyzed to reveal its gene expression profile after exposure to its natural microbial community. Exometabolites were analyzed at four different time points with a Bruker Impact II HD quadrupole time-of-flight mass spectrometer (QToF-MS), and 28 molecules were confirmed against a library of standards. In addition, Fourier-transform ion cyclotron resonance mass spectrometry (FT-ICR-MS) was used to determine the molecular composition of dissolved organic matter components to predict A. glacialis A3’s metabolic reprogramming. To confirm the ability of different taxa to utilize diatom metabolites as growth substrates, we isolated bacteria from the bacterial consortium and monitored their growth in supplemented media. Further, rosamarinic acid (2 mM) and azelaic acid (100 μM) were used to examine if diatom secondary metabolites can give Roseobacters an advantage over Alteromonadaceae by cell attachment with a bacterial motility assay or by modulation of bacterial growth, respectively. The global distribution and homology analysis of an azelaic acid transcriptional regulator was examined by querying a hidden Markov model profile against the Tara Oceans bacterial metagenomes database.

Data Deposition and Materials Availability. The A. glacialis strain A3 is available from the National Center for Marine Algae and Microbiota (NCMA) collection under the accession CCMP3542. The A. glacialis A3 genome is deposited at DDBJ/ENA/GenBank under the accession WKK00000000–WKF00000000 in NCBI-BioProject PRJNAS88343. DNA-sequ reads of A. glacialis A3 are deposited in NCBI under the BioProject PRJNAS88343. Metagenomic reads and RNA-seq reads of the bacterial consortium are deposited in NCBI under the BioProject PRJNAS75878. Metagenomically assembled genomes are deposited at DDBJ/ENA/GenBank under the accession WKF000000000–WKF000000000 in NCBI-BioProject PRJNAS88964. Whole-genome assemblies of consortium-isolated strains S. pseudontizschieae F5, Phaeobacter sp. F10, and A. macroedi F12 are deposited at DDBJ/ENA/GenBank under the accession WKF000000000, WKF0000000000, and CP04614-CP046144, respectively, in NCBI-BioProject PRJNAS88972. Mass spectral datasets are available in the MassIVE database under accession MSV000084592. All software packages used in this study are free and open-source.

ACKNOWLEDGMENTS. We thank the NYU Abu Dhabi Core Technology Platforms for support in genomics sequencing and mass spectrometry. We also thank Brian McParland for collecting water samples and Bryndan P. Durham and Elodie Ghedin for helpful comments on the manuscript. This project was supported by a grant from the NYU Abu Dhabi Research institute to K.C.G. (ADHPG-CCSB) and grants from NYU Abu Dhabi (AD179) and NOA (NA19NOS4780183) to S.A.A.
86. H. W. Jung, T. J. Tschaplinski, L. Wang, J. Glazebrook, J. T. Greenberg, Priming in systemic plant immunity. Science 324, 89–91 (2009).
87. J. Malamy, J. P. Carr, D. F. Klessig, I. Raskin, Salicylic acid: A likely endogenous signal in the resistance response of tobacco to viral infection. Science 250, 1002–1004 (1990).
88. H. Fu, M. Uchimiya, J. Gore, M. A. Moran, Ecological drivers of bacterial community assembly in synthetic phycospheres. Proc. Natl. Acad. Sci. U.S.A. 117, 3656–3662 (2020).
89. M. A. Ochsenkühn, P. Schmitt-Kopplin, M. Harir, S. A. Amin, Coral metabolite gradients affect microbial community structures and act as a disease cue. Commun. Biol. 1, 184 (2018).
90. A. Pascale, S. Proietti, I. S. Pantelides, I. A. Stringlis, Modulation of the root microbiome by plant molecules: The basis for targeted disease suppression and plant growth promotion. Front Plant Sci 10, 1741 (2020).
91. L. Peters et al., Secondary metabolites of Flustra foliacea and their influence on bacteria. Appl. Environ. Microbiol. 69, 3469–3475 (2003).
92. M. McFall-Ngai et al., Animals in a bacterial world, a new imperative for the life sciences. Proc. Natl. Acad. Sci. U.S.A. 110, 3229–3236 (2013).
93. M. Sapp et al., Species-specific bacterial communities in the phycosphere of microalgae? Microb. Ecol. 53, 683–699 (2007).
94. M. P. Sison-Mangus, S. Jiang, K. N. Tran, R. M. Kudela, Host-specific adaptation governs the interaction of the marine diatom, Pseudo-nitzschia and their microbiota. ISME J. 8, 63–76 (2014).
95. P. A. Ajani et al., The microbiome of the cosmopolitan diatom Leptocylindrus reveals significant spatial and temporal variability. Front. Microbiol. 9, 2758 (2018).
96. F. Azam et al., The ecological role of water-column microbes in the sea. Mar. Ecol. Prog. Ser. 10, 257–263 (1983).