The rice endophyte *Harpophora oryzae* genome reveals evolution from a pathogen to a mutualistic endophyte

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The fungus *Harpophora oryzae* is a close relative of the pathogen *Magnaporthe oryzae* and a beneficial endosymbiont of wild rice. Here, we show that *H. oryzae* evolved from a pathogenic ancestor. The overall genomic structures of *H.* and *M. oryzae* were found to be similar. However, during interactions with rice, the expression of 11.7% of all genes showed opposing trends in the two fungi, suggesting differences in gene regulation. Moreover, infection patterns, triggering of host defense responses, signal transduction and nutritional preferences exhibited remarkable differentiation between the two fungi. In addition, the *H. oryzae* genome was found to contain thousands of loci of transposon-like elements, which led to the disruption of 929 genes. Our results indicate that the gain or loss of orphan genes, DNA duplications, gene family expansions and the frequent translocation of transposon-like elements have been important factors in the evolution of this endosymbiont from a pathogenic ancestor.

Rice has been a major food source for people in Asia and Africa for centuries. However, a large proportion of the rice yield is annually lost to agricultural diseases and pests1. Rice blast, caused by the fungal pathogen *Magnaporthe oryzae*, is one of the most severe rice diseases and has been found almost everywhere rice is grown2,3. Beneficial relationships between plants and microorganisms often occur in the rhizosphere and play important roles in ecosystems by improving plant growth or by helping plants to overcome biotic or abiotic stress4. Whereas mycorrhizal symbioses have long attracted significant interest5, root endophytes have only recently been recognized to also be fundamental components of ecosystems6,7. The fungus *Harpophora oryzae* was first described among endophytes residing in domestic Chinese wild rice (*Oryza granulata*), where *H. oryzae* can strongly promote rice growth and biomass accumulation8. In addition, *H. oryzae* can protect rice roots from invasion by *M. oryzae* and can induce systemic resistance to rice blast, which makes it an attractive candidate for biocontrol9. Phylogenetic analyses have shown that *Harpophora* has a close relationship to other members of the Magnaporthaceae, such as *Gaeumannomyces* and *Magnaporthe*, most of which are plant pathogens10,11. *H. oryzae*, similar to *M. poae* and *G. graminis*, but unlike *M. oryzae*, only infects plant roots10. Some morphological aspects of penetration into the plant, such as the formation of appressoria or hyphopodia, are also similar between *M. oryzae* and *H. oryzae*. However, *H. oryzae* only infects roots from the epidermis to the cortex without penetrating the stele9, whereas *M. oryzae* can invade vascular tissue, from which it systemically spreads to the aerial parts of the plant12.

The morphologic and phylogenetic relationships between *H. oryzae* and its close plant pathogenic relatives render it an attractive model for examining the evolutionary mechanisms that lead to a beneficial endophyte in an ancestral neighborhood of pathogens. Here, we describe the genome sequence of *H. oryzae* and present a comparative analysis of the transcriptomes of *H. oryzae* and *M. oryzae* during rice root infection. We show that *H. oryzae* arose from a pathogenic ancestor and that this change was accompanied by a significant expansion of transposable elements, the development of a retrotransposon surveillance pathway, and the concomitant expansion of gene families related to a biotrophic lifestyle.
Results and Discussion

Genome sequencing and general features. The genome of *H. oryzae* R5-6-1 was sequenced using a combination of 454 pyrosequencing and the Illumina HiSeq 2000 sequencing platform. The total reads were 6562 Mb in length, representing an approximately 129-fold genome sequence coverage (Supplementary Table S1). A 50.78-Mb draft genome sequence of *H. oryzae* was assembled and contained 247 scaffolds (size: >200 bp; Table 1). The genome of *H. oryzae* was approximately 8% larger than those of *M. oryzae*, *M. poae* and *G. graminis* (Supplementary Table S2). A total of 14,575 protein-coding genes have been predicted for *H. oryzae*, 90.1% of which were validated using mRNA sequences. By the Core Eukaryotic Genes (CEGs) Mapping Approach pipeline (CEGMA v 2.0), the completeness of the *H. oryzae* genome was assessed to be 99.2%. The details of the sequence analysis are provided in the supplementary material (Supplementary Note 1).

Syntenic analysis and phylogenetic relationships. Pairwise sequence comparisons of the genome sequence of *H. oryzae* and those of its nearest phylogenetic neighbors revealed high degrees of macrosynteny between *H. oryzae* and *M. poae* or *G. graminis*. By contrast, only mesosynteny (i.e., chromosomes with similar gene contents but with different orders and orientations of genes) was observed between *H. oryzae* and *M. oryzae* (Supplementary Fig. S1). A phylogenetic tree derived from a combined analysis of 280 single-copy orthologs (i.e., genes in the same Markov cluster (MCL); see below) from *H. oryzae* and 16 other fungi was consistent with known species phylogeny (Fig. 1a). However, species sharing the same nutritional lifestyle (i.e., pathogens, symbionts or saprophytes) were located randomly on the branches rather than clustering together, indicating that these lifestyles have been gained and lost several times during evolution. Ancestral state reconstruction (ASR) further showed that *Harpophora, Magnaporthiopsis* and *Gaumannomyces* formed a monophyletic clade together with *Nakatakea* and *Magnaporthe*. Bayesian inference (BI), maximum likelihood (ML) and maximum parsimony (MP) analyses all suggested that the ancestral state of *H. oryzae* and related species in clade A was that of a plant pathogen (Fig. 1b). Inferring a time scale from the phylogenetic analysis further revealed that *M. oryzae* diverged from *G. graminis*, *H. oryzae* and *M. poae* approximately 67 million years (MY) ago (Fig. 1a), which corresponds well with the divergence times among the Triticeae (barley, wheat and oats, 13–25 MY) 

Transposable elements. One of the most striking features observed in the *H. oryzae* genome, compared with its closest phylogenetic neighbors, was its relatively high number of transposable elements (TEs) (Supplementary Table S3; S4), i.e., 15% in *H. oryzae* vs. 10.2%, 0.65% and 7.2% in *M. oryzae*, *M. poae* and *G. graminis*, respectively. LTR retrotransposons (i.e., Gypsy, Copia and LINE Tad1) were the most abundant class of TEs in the genomes of *H. oryzae*, *M. oryzae*, and *G. graminis* (50.7, 72 and 72.5% of all TEs, respectively). TcMarF01 was the most abundant class II TE in all three genomes. The higher number of TEs in *H. oryzae* was primarily due to the presence of larger numbers of unclassified TEs (38.3% of TEs, 2912 kb for *H. oryzae* vs. 8.2% of TEs, 341 kb for *M. oryzae*). Generally, TEs were more common in regions of the *H. oryzae* genome with lower gene density (Supplementary Fig. S2). Regions containing either *H. oryzae*-specific orphan genes (BLASTP: E-value < 1E-10) or duplicated genes (paralogs, see below) were located in the vicinity of areas with a high frequency of TEs (Supplementary Fig. S2). Consistent with this finding, most of the 929 TE-disrupted genes and 368 TE-containing genes were *H. oryzae* specific and had unknown functions (Supplementary Fig. S3). Among these genes, 83 and 15 encode proteins that contain a signal sequence and are therefore likely secreted. These results strongly suggest that the TEs have had an important influence on the dynamics of the evolution of the *H. oryzae* genome. We also found evidence of RIP activity in all *H. oryzae* TE families (Supplementary Fig. S4). Because RIP is active only under conditions of sexual recombination, we also searched for the presence of mating type genes. We found orthologs of MAT1-1-1 (HAOR_11805), MAT1-1-2 (HAOR_11806) and MAT1-1-3 (HAOR_11807), but not of a MAT1-2 gene, suggesting that *H. oryzae* is heterothallic and principally able to perform sexual reproduction. The fact that RIP has not protected *H. oryzae* against its significant transposon expansions is likely due to the existence of very rare sexual activity, which is similar to *M. oryzae*.

Comparative genomic analysis. An MCL analysis identified a total of 19,338 ortholog groups (157,857 genes) that were clustered between *H. oryzae* and other 16 fungal genomes [comprising phytopathogens (*M. oryzae*, *M. poae*, *G. graminis*, *Colletotrichum higginsianum*, *Fusarium graminearum*, *Botrytis cinerea*, *Sclerotinia sclerotiorum* and *Ustilago maydis*), symbionts (*Epichloë festucae*, *Tuber melanosporum*, *Laccaria bicolor* and *Piriformospora indica*), saprophytes (*Aspergillus nidulans*, *Neurospora crassa* and *Chaetomium globosum*) and the yeast *Saccharomyces cerevisiae*. On average, each group contained approximately 8.2 genes. Two ortholog groups were shared only by *H. oryzae* and other symbiotic fungi, and 13 were shared only by *H. oryzae* and saprophyte genomes; however, 1641 were shared exclusively by *H. oryzae* and pathogenic fungi, which is consistent with the origin of *H. oryzae* from pathogenic ancestors (vide supra). No MCL clusters were identified that were shared by all symbiotic fungi but were absent from the genomes of pathogens and saprophytes. Thus, we did not identify any common “symbiosis-determining genes”. This result is in accordance with other observations and with the observed “loss and gain” of the nutritional lifestyle (see above), thus suggesting that each lifestyle evolved independently of the others.

A total of 11,895 genes of *H. oryzae* clustered into 9315 MCL groups (Supplementary Fig. S1; Supplementary Table S2). Of these, 3834 genes (1254 groups) were present in one or more paralogs, considerably more than in *M. oryzae*, *M. poae* or *G. graminis* (1383 genes (570 groups), 1514 genes (663 groups) and 2200 genes (Fig. 1a). These divergence times among *G. graminis*, *M. poae* and *H. oryzae* correlate with the divergence times between the Triticeae (barley, wheat and oats, 13–25 MY). Therefore, these results suggest that differentiation among *M. oryzae*, *M. poae* and *G. graminis* and *H. oryzae* occurred in response to the divergence of their hosts.

Table 1 | Genome characteristics of *Harpophora oryzae* R5-6-1

| Genome features | Harpophora oryzae |
|-----------------|------------------|
| Assembly size (Mb) | 50.78 |
| Scaffold number | 247 |
| N50 scaffold (kb) | 755 |
| N90 scaffold (kb) | 220 |
| GC-content (%) | 56.6 |
| Protein coding genes | 14575 |
| Mean transcript length (bp) | 1651 |
| Number of exons | 4178 |
| Average exon length (bp) | 519 |
| Number of Introns | 26117 |
| Average intron length (bp) | 137 |
| Mean number of introns/gene | 1.7 |
| Mean number of exons/gene | 2.8 |
| Percentage coding (%) | 42 |
| Gene density (number of genes per Mb) | 287 |
| RNA genes | 243 |
| Repetitive DNA (%) | 14.97 |
| Secreted proteins (SP) | 2287 |
| Small secreted cysteine-rich proteins (SSCRPs) | 167 |
In addition, most of these genes occurred in duplicated DNA sequences (Supplementary Fig. S2), thus illustrating that the larger genome size of *H. oryzae* is the result of significant gene duplication. A total of 119 groups (192 genes) in *H. oryzae* were not found in *M. oryzae*, *M. poae* or *G. graminis*, whereas orthologs (generally fewer than 4) were found in the genomes of some of the 13 fungi (Supplementary Fig. S5). The highest number of orthologs was shared between *H. oryzae* and *C. globosum* (Supplementary Fig. S5), and many of them were present only in these two fungi (Supplementary Fig. S5). While it is theoretically possible that these genes were obtained by horizontal gene transfer (HGT), we consider this rather unlikely because of the high number of these genes; the rate of HGT within filamentous fungi is typically very low. Rather, the close similarity of the nutritional lifestyles of *H. oryzae* and *C. globosum* may have led to the maintenance of these genes in their genomes while they were lost in other species. These genes can thus be considered typical saprophytic growth genes.

The search for genes involved in the clusters of orthologous group (COG) classifications revealed many more genes in the ‘Lipid transport and metabolism’ cluster for the *M. oryzae* genome (1490 genes) than in the *H. oryzae* (395 genes), *M. poae* (274 genes) and *G. graminis* (342 genes) genomes (Fig. 2). Lipid transport and metabolism...
are necessary for M. oryzae to complete appressorium-mediated infection, especially the prepenetration of the leaf cuticle21. The loss of most genes in the ‘Lipid transport and metabolism’ cluster is consistent with the loss of the capability for appressorium-mediated leaf infection in root-infection-only species (see below).

A total of 573 gene families were shown by a CAFE22 analysis to have undergone expansion in H. oryzae (p < 0.01, at least 4 genes in total), whereas only 10 gene families have undergone contraction (Supplementary Table S5). The most expanded gene families were transposons, as noted above, and genes that encode proteins involved in DNA binding, such as centromere protein B (CENP-B). CENP-B proteins are transposase-derived centromeric proteins; in S. pombe, they localize to and recruit histone deacetylases to silence Tf1 retrotransposon surveillance pathway in H. oryzae reveals the activity of a retrotransposon surveillance pathway in H. oryzae, which may reflect an attempt to counteract the strong transposon activity.

Other genes for which expansion was demonstrated included those encoding chitinases, the transport and metabolism of carbohydrates (and other solutes), kinesin light chains, serine/threonine protein kinases, secondary metabolite biosynthesis and inorganic ion transport and metabolism. This expansion of genes involved in signaling and transport indicates that H. oryzae has developed a complex regulatory machinery to sense signals received from the external environment and couple them with intracellular signaling and transport pathways.

The root colonization strategies and transcriptome analyses during fungal infection. To identify which of these gene expansions might be related to the shift of H. oryzae from a plant pathogen to a plant endosymbiont, we performed genome-wide expression profiling using RNA-seq (Supplementary Note 3), and we verified selected genes using qPCR (Supplementary Note 4). To obtain detailed information about the root colonization strategy employed by H. oryzae and its differences from that of M. oryzae and thus to choose the most appropriate time points for RNA isolation, we first inoculated roots of in vitro-cultivated rice plants with conidia of DsRed2-tagged H. oryzae strain Ho19red and eGFP-tagged M. oryzae strain Ho31gfp6, and documented infection using fluorescence and confocal microscopy (Supplementary Fig. S6).

By 1–2 days after inoculation (DAI), numerous runner hyphae of H. oryzae were present along the longitudinal axis of the root surface, and the fungal penetration of the rhizodermis was observed. Melanized appressoria, which are typically associated with leaf infection, were not observed on the roots. Instead, hyphopodia were present on the surface and penetrated the epidermal cells. Upon penetration (4 DAI), fungal growth was visible in the epidermal cells of the root, but not in the cortical cell layers. Some invasive hyphae (IH) were strictly confined to certain epidermal cells, which contained large numbers of IH. At 6 DAI, mild colonization of cortical cell layers was observed. No IH were detected in vascular tissue, even at a late stage (20 DAI). At 20 DAI, the biomass of the aerial part of the rice colonized by H. oryzae was significantly increased compared with the control plants (Supplementary Fig. S7).

Similar to H. oryzae, the conidia of M. oryzae germinated within 2 DAI and produced fungal hyphae (Supplementary Fig. S6). Numerous runner hyphae were present and formed infectious structures (hyphopodia) and penetrated epidermal cells via penetration pegs12. However, no strictly confined IH were observed in M. oryzae at 4 DAI. The IH had rapidly colonized the vascular tissues by 6 DAI, when the systemic infection of leaves and stems commenced (Supplementary Fig. S6). Based on these results, we chose roots infected by H. oryzae at 2, 6 and 20 DAI and roots infected by M. oryzae at 2 and 6 DAI for the RNA-seq analysis.

The deficiency of the appressorium-mediated leaf infecting ability of H. oryzae. The genes controlling appressorium-mediated penetration in M. oryzae have been extensively studied22. These genes involve four key signaling pathways: the cyclic AMP-protein kinase A (cAMP-PKA) pathway, the MAP kinase (MAPK) pathway, the cell

![Figure 2](image-url)
wall integrity MAPK pathway and the osmoregulation pathway. H. oryzae contains orthologs of all these genes, in addition to genes involved in ROS23 and autophagy24, which are also important for the formation and function of appressoria (Supplementary Table S10). Furthermore, most of the experimentally verified virulence-associated genes of M. oryzae are also present in H. oryzae (Supplementary Table S11). However, all three of the fungi that only infect roots (H. oryzae, M. poae and G. graminis) lack 19 virulence genes, including mpg1, moac and mors7, which are important for interactions with the leaf surface or the penetration ability of appressoria25–27. Tucker29 proposed that the ability to develop appressorium-mediated infection was acquired after the ability to infect roots, and the former ability required the acquisition of new genes, such as mpg1, or gene functions. Because the ancestor of H. oryzae was a plant pathogen, it is likely that H. oryzae lost the ability to infect leaves owing to the loss of genes indispensable for appressorium-mediated infection. It has been shown that H. oryzae could restrict root infection by M. oryzae30, suggesting that competition between these relatives may have partly resulted in obligate root infection by H. oryzae.

The differences in transduction of the extracellular signal. A total of 73 G-protein-coupled receptors (GPCRs) were found in the H. oryzae genome, including 60 phl11-like GPCRs, whereas only 25, 26 and 27 GPCRs were found in the H. oryzae, M. poae and G. graminis genomes, with 16, 19 and 17 phl11-like GPCRs, respectively (Supplementary Table S12). phl11-like GPCRs were present in all the main clades in the phylogenetic tree of H. oryzae (Supplementary Fig. S11), suggesting that H. oryzae possesses orthologs of all of them. However, there were obvious significant differences in the expression of the abundantly expressed phl11-like GPCRs (Supplementary Fig. S11). Similar findings were obtained for cAMP receptor-like GPCRs (Supplementary Fig. S11). In addition, the orthologous gene (determined by bidirectional best hits) of the H. oryzae Gα subunit magC was expressed much more strongly in H. oryzae. Taken together, these results suggest that H. oryzae and H. oryzae differ in their responses to extracellular signals from the host, which may be part of the difference between endophytic and pathogenic interactions. These results also make it likely that signals produced by the same host may be received differently by M. oryzae and H. oryzae.

Different nutritional preferences of H. and M. oryzae. H. oryzae possesses the highest number of carbohydrate-active enzymes (CAZymes; 606 genes) compared with its relatives, which is due to the high numbers of glycoside hydrolases, carbohydrate-binding domains and polysaccharide lyases (Supplementary Table S15). A total of 109 plant cell wall-degrading enzymes (CWDEs) are shared by H. oryzae and M. oryzae (Supplementary Fig. S10; S12). Genes that were particularly enhanced in H. oryzae include the GH55 exo-β-1,3-glucanases, GH78 α-rhamnosidases, GH95 α-fucosidas, PL1 polygalacturonate lyases and the CBM67 rhamnose-binding module. Except for GH55, which may be involved in H. oryzae cell wall turnover or antagonism against competing fungi, the other genes all function in hydrolyzing the hemicellulose side chains and pectin of the plant and, thus, in penetrating the roots. These data are also in accordance with the findings that, compared with necrotrophic and hemibiotrophic fungi, plant pathogenic fungi tend to have fewer CWDEs, such as enzymes of GH61, GH78, PL1 and PL3 as well as enzymes containing CBM1, CBM18 and CBM50 domains31. Genome-wide expression profiling using RNA-seq revealed that most CWDEs were expressed in both H. oryzae and M. oryzae at all stages of their interactions with rice. However, there were remarkable differences in the expression patterns of these CWDEs: 44 genes clustered into six groups (Fig. 3a) and were regulated in essentially opposite directions in the two fungi. Most of them were downregulated in H. oryzae but upregulated in M. oryzae except genes in the group B and D (Fig. 3a). While the expressions of genes in the group B were not changed in M. oryzae, they were suppressed in H. oryzae. Similarly, the gene expressions in the group D kept stable in H. oryzae but upregulated in M. oryzae. These results show that the expression levels of CWDEs were tightly restricted at the onset of the biotrophic phase in H. oryzae. A similar pattern was recently reported in a comparison of the infection of living and dead roots by Piriformospora indica32. Those of the present study are consistent with the scenario that following successful colonization, M. oryzae is inclined to kill the root cells and commence necrotrophic nutrition, whereas H. oryzae prefers to keep the root cells alive and exhibits biotrophy. Quantification using GC/MS revealed that the glucose content of roots infected with H. oryzae was significantly higher than that of roots infected with M. oryzae and with sterile water (control samples) at all infection stages (Fig. 3b). Glucose could inhibit the expression of CWDEs by plant-colonizing fungi, which is known as glucose repression33,35. Differences in the allocation of carbon by the plant during endophytic and pathogenic interactions suggest that the host response may play an important role in the evolutionary process of the pathogen, which is consistent with the finding that differentiation among H. oryzae and related species was accompanied by the divergence of their hosts (vide supra).

Differences in triggering of host defense responses of H. and M. oryzae. Among the 173 orthologs of virulence-associated genes shared by M. oryzae and H. oryzae, 21 (12.1%) genes were significantly regulated in either M. oryzae or H. oryzae (Supplementary Fig. S15). However, most of these genes were downregulated in H. oryzae but upregulated in M. oryzae, suggesting that the expression of virulence-associated genes in H. oryzae was suppressed. The reduced expression of these genes was also important to decrease plant immune responses. 167 small secreted cysteine-rich proteins (SSCRPs) were identified in the H. oryzae genome, more than the numbers found in related species (Supplementary Table S13). These proteins are considered to facilitate infection by reprogramming host cells and modulating plant immune responses36. Interestingly, none of the significantly regulated SSCRPs in H. oryzae were significantly regulated in M. oryzae, and vice versa (Supplementary Fig. S15; Supplementary Table S14). Thus, the differential expression of these genes in H. oryzae and M. oryzae may be a mechanism involved in their different infection strategies.

The H. oryzae CAZyme also encodes an increased arsenal of enzymes and proteins that interact with chitin (82 genes vs. 61 in M. oryzae, Supplementary Fig. S12; Supplementary Table S14). Interestingly, only 40 of these proteins are orthologs (Supplementary Fig. S10). A phylogenetic analysis showed that the CBM50 (LysM) chitin-binding domains had undergone species-specific expansion, with tandem repeats arising from gene duplication (Supplementary Fig. S13). Most of the H. oryzae-specific genes (42 genes) were secreted extracellularly (Supplementary Table S16; Supplementary Table S17). Similar to the GH55 endo-β-1,3-glucanases, these genes could function in H. oryzae cell wall turnover or antagonism against competing fungi. However, LysM-bearing proteins function to scavenge the chitin of the plant host and therefore prevent the recognition of the fungus by the plant or other hosts37,38. In M. oryzae, LysM effectors also suppress chitin-triggered immunity39,40, although the mechanism by which fungal LysM effectors compete with plant receptors for chitin binding remains unknown.

Genes involved in the secondary metabolite. As is typical of fungi, H. oryzae and M. oryzae also encode genes (37 and 35, respectively) for secondary metabolite (SM) synthesis, numbers that are much higher than those of M. poae and G. graminis (Supplementary Table S18). A total of 60% of SMs were shared between H. oryzae and M. oryzae (Supplementary Fig. S10). A phylogenetic analysis revealed the specific expansion of polyketide synthase (PKS) genes in H. oryzae (Supplementary Fig. S13). In addition, cytochrome P450 (CYP)
monooxygenases were expanded in *H. oryzae* (211 genes) and *M. oryzae* (228 genes) compared with *G. graminis* and *M. poae* (Supplementary Table S19). Similar to SM genes, only few of the orthologous CYPs were shared between the *M. oryzae* and *H. oryzae* genomes when orthologs were considered (Supplementary Fig. S10). Interestingly, genes involved in abscisic acid signaling, which might play roles in the manipulation of plant hormone metabolism, exhibited lower (or downregulated) expression in *H. oryzae*, whereas genes involved in the synthesis of auxin and gibberellin exhibited higher (or upregulated) expression in *H. oryzae* (Supplementary Table S20). We note, however, that *H. oryzae* possesses no orthologs of the *cps/ks* genes, which are essential for gibberellin biosynthesis in *Fusarium fujikuroi* (Supplementary Table S20). Thus, our results suggest that *H. oryzae* may influence the biosynthesis of this plant hormone instead of producing it directly. The rice growth-promoting function of *H. oryzae* may therefore be related to auxins and/or gibberellins, which exhibit remarkable promotional effects on plants.

**Horizontal gene transfer (HGT) in the *H. oryzae* genome.**

*HAOR* 13605, which encodes a periplasmic phosphate ABC transporter, was highly expressed (Supplementary Table S24). Interestingly, this gene appears to have been obtained by horizontal gene transfer from a *Granulicella* sp. (a bacterium belonging to the phylum Acidobacteria) (Supplementary Fig. S14). However, since this protein lost its function as a phosphate-specific transporter (see Supplementary Note), *HAOR* 13605 is more likely to function as a sensor of nutrients that regulate carbon metabolism, which requires further investigation.

**Conclusion**

In this study, we generated a new model of a mutualistic endophytic fungus that originated from pathogenic ancestors, and this model provided us with the first chance to study this evolution by comparative genomic and transcriptomic analyses. Our research highlights the importance of both internal dynamics (such as transposable elements) and external pressures (such as competing relationships with relatives and different responses from hosts) in the evolutionary transformation of *H. oryzae* from a pathogen to a mutualistic endophyte. Furthermore, horizontal gene transfer may have also partly affected the *H. oryzae* genome. *H. oryzae* has also been shown to be useful for agricultural management, and the availability of its genome will

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### Figure 3 | Different expression patterns of fungal genes encoding plant cell wall-degrading enzymes and different plant carbon allocation in endophytic and pathogenetic interactions.

(a) Significantly regulated CAZymes in either *H. oryzae* or *M. oryzae* with enzymatic domains functioning in degrading cellulose, hemicellulose and pectin. Orthologous genes are aligned in the same lines, and the short line refers to orthologous genes that have been lost. DA12, DA16 and DA120 refer to transcripts expressed by *H. oryzae* (RDA1) or *M. oryzae* (GDA1) infecting rice roots at 2, 6 and 20 days after inoculation, respectively. Red, increase in transcript abundance; green, decrease in transcript abundance. (b) GC-MS quantification of glucose and fructose in rice roots infected with *H. oryzae* (HOR), *M. oryzae* (MOR) or sterile water (Control, CON). Blue and red asterisks indicate significant differences of HOR when compared with CON and MOR, respectively (p < 0.01). The y-axis indicates the contents (mg) of D-glucose/D-fructose in 1 mg dry weight of rice roots.
greatly promote the progress of research into the fundamental mechanisms of mutualistic symbiosis and will pave the way for future roles of *H. oryzae* in agricultural applications.

Methods

**Fungal strains and plant materials.** The endophytic *Harpospora oryzae* strain R5-6-1 and the blast fungus *Magnaporthe oryzae* strain Guy11 have been studied in our laboratory for several years8,9,43. The DsRed2-tagged *H. oryzae* and eGFP-tagged *M. oryzae* were used to monitor the infection process. The rice cultivar CO-39 (*Oryza sativa*), for which the genome sequence is available4, was used as a compatible host plant for inoculation experiments. All the fungal strains were cultured on complete medium (CMY) at 25 °C. The *H. oryzae* strains were cultured in the dark, whereas the *M. oryzae* strains were cultured under a 16-h-light/8-h-dark photoperiod. Conidia were harvested from 10-d-old cultures of *M. oryzae*. For *H. oryzae*, germinating phialidic conidia were harvested from 4-d-old potato dextrose broth (PDB, with 5 g glucose/L)5. Rice seeds were surface sterilized as previously described43 and planted in half-strength Murashige & Skoog (MS) solid medium at 30 °C in the dark for 4 days to promote root growth. The plants were then grown vertically under a 16-h light/8-h-dark photoperiod at 28°/24 °C for an additional 6 days. Inoculations were performed as previously described29. Infection assays were conducted using roots laid on sterile filter paper placed on 1/2 MS solid medium. Each plate of four plants received a total of 10 conidia of *M. oryzae* or 10 conidia of *H. oryzae*, which were distributed randomly on top of the root system. The inoculated plants were grown horizontally at 26 °C under 16-h-light/8-h-dark photoperiod, with the roots in darkness at all times. The root infection process was monitored using an LSM780 laser-scanning confocal microscope (Carl Zeiss Inc., Jena, Germany). Genomic DNA was extracted from 100 mg fungal material grown in PDB (with 5 g glucose/L) using a DNeasy Plant Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer’s instructions.

**Phylogenetic analysis and ancestral state reconstruction (ASR).** Six loci were used in this analysis (Supplementary Table S1). All sequences were aligned, concatenated and manually adjusted using Geneious Pro v8.8.3 (http://www.geneious.com/). GTR + G + 1 model was selected as the best-fit model for the datasets using jModelTest v2.1. The phylogenetic analyses were performed using the maximum-likelihood (ML) criterion implemented in RAxML48 through the RAxML-Http BlackBox web server at CIPRES (http://www.phyl.org/) with the best-fit model. Maximum parsimony (MP) analyses were performed using PAUP v4b10, Bayesian inference analyses (BI) were performed with the best-fit model using the Markov chain Monte Carlo method in MrBayes v3.2.1. Information about the studied characters in ASR were retrieved from Luo and Zhang25 and from Yuan et al., MP- and ML-based ASRs were conducted in Mesquite v2.75 (http://mesquiteproject.org/). To account for phylogenetic mapping uncertainty, we also employed a BI approach to analyze ancestral states using the ‘Multistate’ option in BayesTraits v2.0.1. More details are provided in the Supplementary Notes.

**Genome sequencing, assembly and analysis.** The genome of *H. oryzae* R5-6-1 was sequenced with the Roche/454 Pyrosequencing Platform and the Illumina HiSeq2000 sequencing platform. Low-quality data that had a QUAL value of less than 30 and consisted of short reads (length < 50 bp) were filtered from the raw data. The high-quality reads underwent primary assembly using the GS De Novo Assembler (Newbler 2.9.7, Roche) and ALLPATHS-LG (version:allpathslg-43894) and were then scaffolded using SSPACE v2.015. Finally, gaps were filled using SOAP GapCloser v1.0. The completeness of the *H. oryzae* genome was assessed using CEGMA v2.016. The *H. oryzae* R5-6-1 genome sequence has been deposited to GenBank under accession number JNVV00000000. De novo analysis was employed to examine the repetitive sequences. The repetitive elements were identified and classified using a de novo repetitive sequence search with RepeatModeler v1.07 (http://www.repeatmasker.org/RepeatModeler.html). For repeat annotation, the repeat library produced by RepeatModeler was used directly with RepeatMasker v4.0.3–4 (http://repeatmasker.org). The RIP indices were determined with the software RIPCAL by comparisons with the non-repetitive genome17,18.

**Gene prediction and annotation.** The annotation of the genomic sequences of *H. oryzae* was performed with Augustus v2.719 and trained with the assembled RNA-seq transcripts, and the annotated information from *M. oryzae* was incorporated as a reference. Genes were annotated using BLAST2 v2.2.27 against the NR database and InterProScan (http://www.ebi.ac.uk/interpro/) (E-value < 1E-4). InterProScan annotation IDs. Exon junctions were obtained from RNA-seq using TopHat v2.0.9.12. Transcripts with a significant P value (<0.05) and a greater than twofold change (log2) in transcript level were considered differentially expressed. All the P values were corrected for false discoveries resulting from multiple hypothesis testing using the Benjamini-Hochberg procedure. Heatmaps of gene expression profiles were generated using R (R-project.org) based on significant expression changes (log10 FPKM plus 1).

**Quantification of glucose and fructose in rice roots.** The contents of glucose and fructose in rice roots were quantified using gas chromatography-mass spectrometry (GC-MS). All the conditions were as described for the RNA-seq preparation. GC-MS was performed in parallel with cultures harvested for RNA-seq with three additional control samples. The control samples for 2 DAL 6 DAL and 20 DAL were prepared using sterile water instead of a conidia suspension for inoculation. Each mixture of roots from 12 independent rice plants was considered an experimental repetition. Total RNA was extracted using TRizol reagent (Invitrogen) according to the manufacturer’s protocol. The RNA integrity of all the samples was verified on an Agilent 2100 Bioanalyzer. Nine *H. oryzae* libraries (three developmental stages and three biological replicates) were sequenced using Illumina TruSeq RNA Sample Preparation Kit and were sequenced using the Illumina HiSeq 2000 based on 100 bp paired-end read sequencing. The insult sizes of all the libraries were 180 bp for both *H. oryzae* and *M. oryzae*. All the clean reads were mapped to the genome sequence using TopHat v 2.0.9.12, and an expression profile was created using Cufflinks v2.0.2. The abundances were reported as normalized fragments per kb transcript per million (FPKM). SSCRs shorter than 200 aa in length and containing at least 4 cysteine residues were identified among the predicted secreted proteins. Cytochrome P450s were classified based on de novo BLAST searches (E-value < 1E-5, coverage >55%) against protein sequences in the Cytochrome P450 Database (FCPD) (http://psort.soic.iastate.edu/). Phytohormone-related genes were identified by performing BLASTP against the Arabidopsis Hormone Database (AHDB) v2.0 (http://ahd.cb.pku.edu.cn/).

**Transcriptome analysis.** Roots infected with *H. oryzae* were harvested in liquid nitrogen 2, 6 and 20 days after inoculation (DAI); the same procedure was applied to *M. oryzae*. Except that the harvesting occurred 20 DAI. Roots from 12 independent rice plants were considered an experimental replicate. Total RNA was extracted using TRIzol reagent (Invitrogen) according to the manufacturer’s protocol. The RNA integrity of all the samples was verified on an Agilent 2100 Bioanalyzer. Nine *H. oryzae* libraries (three developmental stages and three biological replicates) and six *M. oryzae* libraries (two developmental stages and three biological replicates) were sequenced using the Illumina TruSeq RNA Sample Preparation Kit and were sequenced using the Illumina HiSeq 2000 based on 100 bp paired-end read sequencing. The insert sizes of all the libraries were 180 bp for both *H. oryzae* and *M. oryzae*. All the clean reads were mapped to the genome sequence using TopHat v 2.0.9.12, and an expression profile was created using Cufflinks v2.0.2. The abundances were reported as normalized fragments per kb transcript per million (FPKM). SSCRs shorter than 200 aa in length and containing at least 4 cysteine residues were identified among the predicted secreted proteins. Cytochrome P450s were classified based on de novo BLAST searches (E-value < 1E-5, coverage >55%) against protein sequences in the Cytochrome P450 Database (FCPD) (http://psort.soic.iastate.edu/). Phytohormone-related genes were identified by performing BLASTP against the Arabidopsis Hormone Database (AHDB) v2.0 (http://ahd.cb.pku.edu.cn/).

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