Structures Showing Negative Correlations of Signal Intensity with Postnatal Age on T1-weighted Imaging of the Brain of Newborns and Infants

Saeka Hori1*, Toshiaki Taoka2, Tomoko Ochi1, Toshiteru Miyasaka1, Masahiko Sakamoto1, Katsutoshi Takayama1, Takeshi Wada1, Kaoru Myochin1, Yukihiro Takahashi3, and Kimihiko Kichikawa1

Purpose: Although the neonatal and infantile brain typically shows sequential T1 shortening according to gestational age as a result of myelination, several structures do not follow this rule. We evaluated the relationship between the signal intensity of various structures in the neonatal and infantile brain on T1-weighted imaging (T1WI) and either postnatal or gestational age.

Materials and Methods: We examined magnetic resonance images from 120 newborns and infants without any abnormalities in the central nervous system. Written informed consent was obtained from all parents and the institutional review board approved the study. Gestational age at examination ranged from 35 weeks, 3 days to 46 weeks, 6 days, and postnatal age ranged from 7 days to 127 days. Signal intensity on T1WI was evaluated on a scale from Grade 1 (indistinguishable from surrounding structures) to Grade 4 (higher than cortex and close to fat). We evaluated relationships between the T1 signal grades of various structures in the neonatal brain and postnatal or gestational age using Spearman's correlation analysis.

Results: Significant positive correlations were identified between T1 signal grade and gestational age in the pyramidal tract (P < 0.001). Conversely, significant negative correlations were evident between T1 signal grade and postnatal age (P < 0.001), in structures including the stria medullaris thalami, fornix cerebellar vermis, dentate nucleus and anterior pituitary gland.

Conclusion: Significant negative correlations exist between signal intensity on T1WI and postnatal age in some structures of the neonatal and infantile brain. Some mechanisms other than myelination might play roles in the course of signal appearance.

Keywords: newborn, brain, magnetic resonance imaging, T1-weighted images, development

Introduction

Signal intensity on the T1-weighted image (T1WI) of the neonatal brain is widely recognized as developing from hypointensity to hyperintensity according to gestational age, in a process that is attributed to myelination.1 However, recent reports have demonstrated that some structures show hyperintensity on T1WI soon after birth and a subsequent decrease in signal intensity according to postnatal age. These structures include the anterior lobe of the pituitary gland, subthalamic nucleus, and globus pallidus.2–4 This signal intensity pattern, hyperintensity to hypointensity according to postnatal age, is speculated to be due to different mechanisms from myelination. For example, the hyperintensity of the pituitary gland just after birth is reported to be due to the hyperplasia of prolactin cells.2 In our previous report which pointed out T1 hyperintensity in the subthalamic nucleus and globus pallidus just after the birth, we made speculation about the mechanism that might be due to various processes other than myelination including the rapid proliferation of oligodendroglial cells, cerebral glial reaction by stressful environment at delivery or influence from maternal thyroid hormone.4 Above mentioned speculation made us consider that there would be other structures in the brain which shows a similar...
signal transition pattern, because the mechanism we speculated might occur in anywhere in the brain. And it would be important to be aware of the distribution of those structures.

We selected following structures to evaluate the time course. As the basis of our discussions, we evaluated the structures which show typical and early signal transition by myelination called as “early myelinators” such as pyramidal tract, and “intermediate myelinator” that is the corpus callosum. The structures of limbic system were selected because limbic system are formed earlier than the structures related to neocortex, and may show different tissue development compared to the early or intermediate myelinator structure. In addition, several structures in the cerebellum were also evaluated in order to myelination related to the cerebellum. In the current study, we examined the signal intensities of the above-mentioned structures in the neonatal and infantile brain on T1WI and evaluated correlations between signal intensities and postnatal or gestational age.

Materials and Methods

Subjects

Subjects were newborns and infants who had been admitted to the neonatal intensive care unit (NICU) of our institute and showed normal development throughout two years of follow-up. We retrospectively analyzed 170 consecutive newborns admitted to the NICU for perinatal troubles who had undergone screening magnetic resonance imaging (MRI) of the brain between January 1, 2007 and August 7, 2008. Written informed consent was obtained from all parents prior to the enrollment of subjects and all study protocols were approved by the institutional review board in our institute. Reasons for MRI were as follows: low birth weight or premature birth, 140 cases; neonatal asphyxia, 11 cases; respiratory distress syndrome, 2 cases; transient tachypnea of the newborn, 6 cases; intrauterine growth restriction, 2 cases; ABO incompatibility, 2 cases; congenital diaphragmatic hernia, 2 cases; congenital esophageal atresia, 3 cases; pneumothorax, 1 case; and meningoecele, 1 case. From this population, we excluded 18 cases with pathological status detected in the central nervous system: congenital cytomegalovirus infection, 1 case; hydrocephalus, 2 cases; intracranial hematoma, 1 case; subependymal hemorrhage, 1 case; periventricular leukomalacia, 2 cases; chromosomal abnormality, 3 cases; Langerhans cell histiocytosis, 1 case; myotonic dystrophy, 2 case; hyperinsulinemia, 1 case; the epilepsy of unknown cause, 3 case; and congenital ichthyosis, 1 case. We also excluded 11 cases with developmental abnormalities based on evaluation with the Enjoji Infantile Developmental Test. We also excluded 21 cases that were lost to follow-up two years after birth. In total, we excluded 50 patients over the two years of follow-up. Ultimately, 120 newborns and infants were identified as showing normal development with normal findings on both MRI and developmental testing throughout the two years of follow-up.

We defined the terms concerning the chronological age of the newborn in the current study as follows. Gestational age to refer to the age of the newborn at birth based on the number of weeks gestation, i.e. from the first day of the last menstrual period to the day of delivery, and postnatal age to refer to the period from birth to the day of the MRI examination. In the current study, the gestational age ranged from 248 to 328 days (mean: 272.0 days, median: 268 days), and the postnatal age ranged from 8 to 119 days (mean: 33.1 days, median: 26 days).

Imaging and data analysis

MRI was performed using a 1.5T clinical MR unit (Magnetom Sonata; Siemens AG, Erlangen, Germany) with a standard 8-channel head coil. All examinations included axial sections perpendicular to the brainstem with conventional spin-echo T1-weighted (repetition time, 500 ms; echo time, 12 ms; averaging, 1; acquisition time, 3 min 26 sec). Section thickness was 6 mm with a 2 mm gap. Images were obtained using a 256 × 256 displayed matrix and a 230-mm field of view. Images were acquired during natural sleep with some foam cushions to provide some restraint.

MR images were retrospectively and independently reviewed by two radiologists (S.H., T.T). The radiologists were blinded to the ages of newborns and infants. We selected the following structures bilaterally (Fig. 1): [Early myelinator and intermediate myelinator] pyramidal tract in the precentral gyrus, corona radiata and posterior limb of the internal capsule; corpus callosum, [Limbic system] fornix; stria medullaris thalami; [Cerebellar structure] cerebellar vermis; dentate nucleus of the cerebellum; the decussation of the superior cerebellar peduncles; flocculus cerebellum; superior cerebellar peduncle; inferior cerebellar peduncle; [Other] the ventrolateral nucleus of the thalamus; and anterior pituitary gland. We examined the signal intensity on T1WI of each structure and qualitatively classified signal intensity on T1WI into 4 grades (Fig. 2): Grade 1, indistinguishable from surrounding structures; Grade 2, intensity between the cortex and surrounding structures; Grade 3, higher intensity than cortex, but close to the cortex; Grade 4, higher intensity than the cortex and close to fat. We evaluated the relationship between T1 signal grade and gestational or postnatal age using Spearman’s rank correlation analysis and compared correlation coefficients (rs) between gestational age and postnatal age in each structure. The strength of a calculated absolute rs value was interpreted as follows: 0.00 to 0.19, very weak to negligible; 0.20 to 0.39, weak; 0.40 to 0.69, moderate; 0.70 to 0.89, strong; 0.90 to 1.00, very strong correlation. We evaluated rs values as meaningful when the strength was categorized as moderate, strong or very strong.

Results

Figure 3 shows signal grades and gestational/postnatal ages of the structures of interest. Table 1 shows the correlation coefficient for each structure calculated between the T1 signal
grade as mentioned above and the gestational or postnatal age. Structures showing positive correlations with gestational age were as follows. Pyramidal tract in the precentral gyrus, corona radiata and posterior limb of the internal capsule showed moderate positive correlations between $T_1$ signal grade and gestational age ($P < 0.001$), but no correlation between $T_1$ signal grade and postnatal age.

Structures showing negative correlations with postnatal age were as follows. The fornix, stria medullaris thalami, cerebellar vermis, and anterior pituitary gland showed moderate to strong negative correlations between $T_1$ signal grade and postnatal age. In these structures except the fornix, weak negative correlations were noted between $T_1$ signal grade and gestational age. In the fornix, no significant correlation was apparent between $T_1$ signal grade and gestational age.

The dentate nucleus of the cerebellum showed a strong negative correlation with both postnatal and gestational ages.
Table 1. The correlation coefficient for each structure calculated between the T$_1$ signal grade as mentioned above and the gestational/postnatal age

| Structure                                      | Correlation coefficients with postnatal age | Correlation coefficients with gestational age |
|------------------------------------------------|---------------------------------------------|----------------------------------------------|
| Precentral gyrus                               | NS                                          | 0.53 ($P < 0.001$)                           |
| Pyramidal tract in the corona radiata          | NS                                          | 0.61 ($P < 0.001$)                           |
| Posterior limb of the internal capsule         | NS                                          | 0.51 ($P < 0.001$)                           |
| Fornix                                         | $-0.43$ ($P < 0.001$)                       | NS                                           |
| Stria medullaris thalami                       | $-0.52$ ($P < 0.001$)                       | $-0.24$ ($P < 0.01$)                        |
| Cerebellar vermis                              | $-0.47$ ($P < 0.001$)                       | $-0.28$ ($P < 0.01$)                        |
| Dentate nucleus                                | $-0.82$ ($P < 0.001$)                       | $-0.8$ ($P < 0.001$)                        |
| Flocculus cerebellum                           | NS                                          | $-0.54$ ($P < 0.001$)                       |
| Ventrolateral nucleus of the thalamus          | NS                                          | NS                                           |
| Corpus callosum                                | NS                                          | NS                                           |
| Decussation of superior cerebellar peduncles   | NS                                          | NS                                           |
| Superior cerebellar peduncle                   | $-0.35$ ($P < 0.001$)                       | NS                                           |
| Inferior cerebellar peduncle                   | $-0.27$ ($P < 0.001$)                       | $-0.24$ ($P < 0.01$)                        |
| Anterior pituitary gland                       | $-0.64$ ($P < 0.001$)                       | $-0.33$ ($P < 0.001$)                       |

The flocculus cerebellum showed a moderate negative correlation between T$_1$ grade and gestational age and no correlation with postnatal age. The ventrolateral nucleus of the thalamus, corpus callosum, the decussation of the superior cerebellar peduncles, superior cerebellar peduncle and inferior cerebellar peduncle showed either no significant correlation or only weak correlations with postnatal age and gestational age.

Discussion

The motivation of this study is to figure out the structures which show significant negative correlations between their T$_1$ signal intensities and postnatal age, not gestational age. In our previous report, T$_1$ signal intensities of the subthalamic nucleus and globus pallidus showed significant negative correlations to the postnatal age, and there was a question that if there are some other structures showing a similar pattern. So, we selected following structures to evaluate the time course. As sites expected to show typical T$_1$ signal changes according to gestational ages, we evaluated white matter of precentral gyrus, pyramidal tract in the corona radiata and the posterior limb of the internal capsule. These areas are called as “early mielinators” in which myelination begins before the birth and becomes histologically mature by six postnatal months. Corpus callosum was selected as an “intermediate myelinator” in which myelination begins after birth and becomes histologically mature by six postnatal months. As limbic system structure which shows early formation compared to neocortes, fornix and stria medullaris thalami are selected. Stria medullaris thalami is reported to show the onset of myelination before birth and the myelination completely in rather early period, and fornix is reported to show onset of myelination after birth and the myelination complete in rather late period. We also selected cerebellar structures including cerebellar vermis, dentate nucleus of the cerebellum and flocculus cerebellum, the decussation of superior cerebellar peduncles, superior cerebellar peduncles and inferior cerebellar peduncles. Ventrolateral nucleus of the thalamus was selected to represent the central gray matter to compare with the result of our previous study for subthalamic nucleus and globus pallidus. We also evaluated anterior pituitary gland, in which transient T$_1$ hyperintensity has already reported and can be act as a control of the current study.

The present study observed moderate to strong positive correlations between the T$_1$ signal intensity of the pyramidal tract and gestational age, apparently corresponding to myelination. In contrast, moderate to strong negative correlations were identified between T$_1$ signal intensity and postnatal age in various structures, including the fornix, medullary stria of thalamus, cerebellar vermis, dentate nucleus of the cerebellum and anterior pituitary gland. Also in our previous study on subthalamic nucleus and globus pallidus, hyperintensities on T$_1$WI in these structures were observed just after birth and these hyperintensities diminished in older subjects and the disappearance of this hyperintensity was well correlated with postnatal age. One of the explanations for the negative correlation to the age which is shown in the signal intensity may be the relative signal contrast to the surrounding structure. The signal of a structure will be looking lower when the surrounding structure myelinates especially when the evaluation is undergone by comparison with surrounding structure like the current study. However, if so there
Fig 3. Correlation between signal grade and gestational/postnatal age. Signal grades are plotted vertically and ages are plotted horizontally. Graphs on the left side are plotted using gestational age, and graphs on the right side using postnatal ages. (A) Precentral gyrus, (B) Pyramidal tract in the corona radiata, (C) Posterior limb of the internal capsule, (D) Fornix, (E) Stria medullaris thalami, (F) Cerebellar vermis, (G) Dentate nucleus of the cerebellum, (H) Flocculus cerebellum, (I) Ventrolateral nucleus of the thalamus, (J) Corpus callosum, (K) Decussation of superior cerebellar peduncles, (L) Superior cerebellar peduncles, (M) Inferior cerebellar peduncles, (N) Anterior pituitary gland.
should be a negative correlation to the gestational age, not postnatal age. We hypothesized that mechanisms other than myelination may play roles in these signal changes.

At birth, newborns may be under various stresses and environmental challenges, including exposure to the hypoxic extraterine environment, and body temperature drop immediately after birth. In order to adapt to extraterine life, a number of changes take place in the newborn body. Maternal and fetal hormone levels also change dynamically before and after delivery. For example, a massive release of oxytocin is the trigger of parturition and crossing easily the placenta from the mother to the newborn. After birth, thyroid-stimulating hormone (TSH) and tri-iodothyronine (T3) rise sharply and concurrently in the newborn, peaking at around 2 h. The surge in TSH is believed to be due to extraterine cooling. Glucocorticoids also increase soon after birth. We speculated that these various stresses and hormonal changes during delivery might be associated with the signal course in the current study in the following hypotheses.

In general, T1 shortening is caused by various factors including lipids, paramagnetic effects, the immobilization of water molecules, and relative hyperintensity. Hyperintensity on T1WI can be observed in various conditions in the central nervous system. The perirolandic gyrus of infants at 41-44 weeks old reportedly show hyperintensity on T1WI despite the relatively scant myelination, which may reflect the accelerated neuronal development associated with the rapid proliferation of oligodendroglial cells, synapses and dendrites. The periventricular hyperintensity on T1WI in patients with Alexander disease is known to be due to the accumulation of Rosenthal fibers, which are swollen, protein-rich astrocytes. These conditions seem to be associated with glial reactions, and can be classified as “glia-related hyperintensity”. In addition, the putamen reportedly shows transient hyperintensity on T1WI after transitional middle cerebral artery occlusion, which is suggested to be caused by accumulation of manganese-binding proteins.

In the current study, the structures of the limbic systems including fornix and medullary stria of the thalamus as well as the dentate nucleus of the cerebellum and cerebellar vermis showed negative correlations between T1 signal intensity and postnatal age, which were similar tendency with subthalamic nucleus and globus pallidus in our previous study. There may be several cause for these phenomenon and following descriptions are our speculations. One of our hypotheses is that glial reaction may be associated with hyperintensity on T1WI. GFAP is known as a major intermediate protein of astrocytes and plays important roles in brain development. Modulators of GFAP expression reportedly include several hormones such as T3, glucocorticoids and several growth factors. Since T3 and glucocorticoids begin to increase soon after birth, as mentioned above, these hormones might increase GFAP levels and lead to hyperintensity on T1WI. Oligodendrocytes are also stimulated by several mediators, including neuregulins, and some hormones including thyroid hormone and progesterone, which might cause transient hypermyelination leading to hyperintensity on T1WI. These reaction might take place in the perinatal period in which various stresses and environmental challenges described above exists.

Our next hypothesis is that high concentrations of neuronal transmitters might contribute to hyperintensity on T1WI. For example, gamma-aminobutyric acid (GABA) is known as the primary excitatory neurotransmitter in immature neurons and switches to act as an inhibitory neurotransmitter during delivery, which has recently been reported to be triggered by massive releases of oxytocin. The subthalamic nucleus and external globus pallidus are connected to each other by both glutamatergic and GABAergic neurons. High activity might be seen in these structures in association with the switching of GABA activity, which might cause hyperintensity on T1WI soon after birth. Our observation on pituitary gland as a control of the current study agreed with the previous report which shows negative correlations were identified between T1 signal intensity and postnatal age. This transient hyperintensity in the perinatal pituitary gland is speculated to be caused by hyperplasia of prolactin cells, which may be another mechanism for T1 hyperintensity just after birth.

Several limitations to the current study must be considered. First, subjects were not completely “normal” because they had required admission to the NICU for observation following perinatal troubles. Second, this was a qualitative analysis. However, signal intensity is usually evaluated by comparison with surrounding structures at routine image interpretation in clinical practice, as shown in the current study. Third, no histological proof has been provided for these signal changes. Finally, the cause of the signal changes must be limited to speculation.

Conclusions

In addition to previously reported subthalamic nucleus and globus pallidus, significant negative correlations between signal intensity on T1WI and postnatal age exist in the limbic structures including stria medullaris thalami and fornix, as well as in the cerebellar vermis, dentate nucleus and anterior pituitary gland. We speculated that some mechanism other than myelination might play a role in these signal changes. We should be aware that some structures show hyperintensity on T1WI soon after birth, regardless of gestational age in physiological status. We would therefore take into account not only the gestational age, but also the postnatal age for interpretation of the neonatal and infantile brain.
Conflicts of Interest

The authors declare that they have no conflicts of interest.

References

1. Barkovich AJ. Concepts of myelin and myelination in neuroradiology. AJNR Am J Neuroradiol 2000; 21: 1099–1109.
2. Kitamura E, Miki Y, Kawai M, Itoh H, Yura S, Mori N, et al. T1 signal intensity and height of the anterior pituitary in neonates: correlation with postnatal time. AJNR Am J Neuroradiol 2008; 29:1257–1260.
3. Miki Y, Kataoka ML, Shibata T, Haque TL, Kanagaki M, Shimono T, et al. The pituitary gland: changes on MR images during the 1st year after delivery. Radiology 2005; 235:999–1004.
4. Taoka T, Aida N, Ochi T, Takahashi Y, Akashi T, Miyasaka T, et al. Transient hyperintensity in the subthalamic nucleus and globus pallidus of newborns on T1-weighted images. AJNR Am J Neuroradiol 2011; 32:1130–1137.
5. Kinney HC, Karthigasan J, Borenshteyn NI, Flax JD, Kirschner DA. Myelination in the developing human brain: biochemical correlates. Neurochem Res 1994; 19: 983–996.
6. Hokama T, Gushi Ken M, Nosoko N. Iron deficiency anaemia and child development. Asia Pac J Public Health 2005; 17:19–21.
7. Huang H, Zhang J, Wakana S, Zhang W, Ren T, Richards LJ, et al. White and gray matter development in human fetal, newborn and pediatric brains. Neuroimage 2006; 33:27–38.
8. Folkert R, Del Bigio M. Disorders of the perinatal period. (Greenfield’s Neuropathology). Love S, Perry A, Ironside J, Budka H, editors. London: CRC Press; 2015.
9. Perrone S, Salvi G, Bellieni CV, Buonocore G. Oxidative stress and nutrition in the preterm newborn. J Pediatr Gastroenterol Nutr 2007; 45 Suppl 3:S178–182.
10. Soll RF. Heat loss prevention in neonates. J Perinatol 2008; 28 Suppl 1:S57–59.
11. Henning SJ. Postnatal development: coordination of feeding, digestion, and metabolism. Am J Physiol 1981; 241:G199–214.
12. Tyzio R, Cossart R, Khalilov I, Minlebaev M, Hübner CA, Represa A, et al. Maternal oxytocin triggers a transient inhibitory switch in GABA signaling in the fetal brain during delivery. Science 2006; 314:1788–1792.
13. Simiša S, Koivisto M, Ranta T, Leppäluoto J, Reinilä M, Haapalahiti J. Serum tri-iodothyronine, thyroxine, and thyrotropin concentrations in newborns during the first 2 days of life. Arch Dis Child 1975; 50:565–567.
14. Fisher DA, Odell WD. Acute release of thyrotropin in the newborn. J Clin Invest 1969; 48:1670–1677.
15. Korogi Y, Takahashi M, Sumi M, Hirai T, Sakamoto Y, Ikushima I, et al. MR signal intensity of the perirolandic cortex in the neonate and infant. Neuroradiology 1996; 38:578–584.
16. van der Knaap MS, Naidu S, Breiter SN, Blaser S, Stroink H, Springer S, et al. Alexander disease: diagnosis with MR imaging. AJNR Am J Neuroradiol 2001; 22: 541–552.
17. Vázquez E, Macaya A, Mayolas N, Arévalo S, Poca MA, Enríquez G. Neonatal Alexander disease: MR imaging prenatal diagnosis. AJNR Am J Neuroradiol 2008; 29:1973–1975.
18. Shan DE, Ho DM, Chang C, Pan HC, Teng MM. Hemichorea-hemiballism: an explanation for MR signal changes. AJNR Am J Neuroradiol 1998; 19:863–870.
19. Fujioka M, Taoka T, Hiramatsu KI, Sakaguchi S, Sakaki T. Delayed ischemic hyperintensity on T1-weighted MRI in the caudoputamen and cerebral cortex of humans after spectacular shrinking deficit. Stroke 1999; 30: 1038–1042.
20. Gomes FC, Paulin D, Moura Neto V. Gial fibrillary acidic protein (GFAP): modulation by growth factors and its implication in astrocyte differentiation. Braz J Med Biol Res 1999; 32:619–31.
21. Wood TL, Bercury KK, Ciélli SE, Mursch LE, Min J, Dai J, et al. mTOR: a link from the extracellular milieu to transcriptional regulation of oligodendrocyte development. ASN Neuro 2013; 5:e00108.