Genetic and environmental factors influencing the contents of essential oil compounds in *Atractylodes lancea*

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Abstract

Rhizomes of *Atractylodes lancea* are used in traditional Japanese medicine (Kampo) and Chinese medicine to treat numerous diseases and disorders because they contain many pharmacologically active compounds. The major active compounds in *A. lancea* are essential oil compounds such as β-eudesmol, hinesol, atractylon, and atractylodin. The contents of the compounds in *A. lancea* exhibit high variability depending on their habitat. We cultivated clonal lines of *A. lancea* in different years (2016, 2017) and different locations (Hokkaido, Ibaraki) to investigate the influence of genetic and environmental factors on the contents of major compounds, namely, β-eudesmol, hinesol, atractylon, and atractylodin. Broad sense heritability of β-eudesmol, hinesol, atractylon, and atractylodin contents were 0.84, 0.77, 0.86, and 0.87, respectively. The effects of interannual variability on the contents of the compounds were lower than those of genotype. In addition, the cultivated environmental factors were assessed by different locations, and the correlations between Hokkaido and Ibaraki grown plants based on β-eudesmol, hinesol, atractylon, and atractylodin contents were 0.94, 0.94, 1.00, and 0.83, respectively. The results suggest that the contents of β-eudesmol, hinesol, atractylon, and atractylodin in *A. lancea* are largely influenced by genetic factors, and clonal propagation could be an effective strategy for obtaining populations with high contents of essential oil compounds. Furthermore, the contents of β-eudesmol, hinesol, atractylon, and atractylodin in *A. lancea* exhibited few correlations with rhizome yields. *A. lancea* cultivars with not only high contents of essential oil compounds but also high rhizome yield could be developed through selective breeding.

Introduction

*Atractylodes lancea* De Candolle (*Compositae*) is a medicinal plant that is distributed in East Asia, mainly in central China [1]. Dried rhizomes of *A. lancea* are prescribed in Japanese and Chinese herbal medicines as the crude drugs [2, 3]. In Japan, the crude drug is called "So-
articulated in the “author contributions” section. Tsumura & Co. was responsible for the study design, data collection and analysis, decision to publish, and preparation of the manuscript.

**Competing interests:** I have read the journal’s policy and the authors of this manuscript have the following competing interests: TT and BM are employees of Tsumura & Co. (https://www.tsumura.co.jp/english/). HE and RO are supervisors at the University of Tsukuba to which TT belongs. This does not alter our adherence to PLOS ONE policies on sharing data and materials.

**Materials and methods**

**Plant materials**

In the present study, seeds of *A. lancea* obtained in a previous study were used [21]. The seeds were originally obtained from the People’s Republic of China and were from different genotypes. In other words, the seeds were harvested in natural mating. The plants were identified by the first author himself as *A. lancea* due to morphological characteristics. A clonal line of *A. lancea* was propagated from a rhizome of a single plant. Twenty-five clonal lines (line1–line25) with different genotypes were used in the present study.
Cultivation of *A. lancea*

The 25 clonal lines were cultivated in an experimental field located in Ami-machi, Inashiki-gun, Ibaraki prefecture (35˚.99'N, 140˚.20'E), Japan, in 2017. Rhizomes of the 25 clonal lines were divided into about 50 g. The rhizomes were planted in November 25, 2016 and harvested on November 23, 2017. The cultivation was performed with 5–20 biological replicates for each clonal line. The replicates for each clonal line were as follows; line1–line17 (n = 20), line18–line24 (n = 10), line25 (n = 5).

To assess the effects of cultivation year on the contents of essential oil compounds in *A. lancea*, six clones (line1–line6) were also grown in the fields located in Ibaraki prefecture in 2016. The plants were cultivated from November 25, 2015 to November 23, 2016 in Ibaraki prefecture. The cultivation was performed with 20 biological replicates for each clonal line. The six clones were also cultivated in an experimental field located in Kyowa-town, Hokkaido prefecture (43˚.01'N, 140˚.53'E), Japan, in 2017, to evaluate the effects of cultivation location on the contents of essential oil compounds in *A. lancea*. Rhizomes of the plants were planted on October 21, 2016 and harvested on October 19, 2017. The plants were cultivated with 8–20 biological replicates for each clonal line. The replicates for each clonal lines were as follows; line1 (n = 20), line2 (n = 12), line3 (n = 8), line4 (n = 20), line5 (n = 18), line6 (n = 20).

**GC-MS analysis of essential oil compounds**

The harvested rhizomes of *A. lancea* were dried in a convection oven at 50˚C for 7 days and their dry weights were determined. The dried rhizomes were pulverized using a vibrating mill (Cosmic Mechanical Technology, TI-200). The contents of four essential oil compounds, namely, β-eudesmol, hinesol, atractylon, and atractylodin, were determined by gas chromatography-mass spectrometry (GC-MS) analysis as follows. Powdered samples of *A. lancea* rhizomes (0.5 g) were extracted with n-hexane (25 mL) for 15 min using a recipro shaker (Taitec, model SR-1) and centrifuged at 1660 × g for 10 min. The supernatant was separated, and the residue was re-extracted with n-hexane (20 mL) in a similar manner as above. The supernatant was combined, and phenanthrene (1.5 mg in 1 mL n-hexane) was added as an internal standard. A solution with a total volume of 50 mL was made by adding of n-hexane. An aliquot (1 μL) of the solution was injected into GC-MS. GC-MS analysis was carried out using an Agilent 7890 gas chromatograph equipped with a 5975 mass spectrum detector (MSD) (Agilent Technologies). Chromatography was performed with a DB-WAX GC column (polyethylene glycol, 30 m×250 μm i.d., 0.25 μm film; Agilent J&W Scientific). The column was used with the following temperature program: column held at 160˚C for 2 min after injection, increased by 5˚C/min to 200˚C, then increased by 8˚C/min to 240˚C, and held for 5min. Injection temperature was set to 160˚C. Helium was used as the carrier gas and the flow rate was 1.0 mL/min. The contents were calculated based on the dry weights of the powdered samples.

**Statistical analysis**

Statistical analyses, including one-way ANOVA, two-way ANOVA, and correlation analysis (Pearson’s correlation coefficient) were performed for β-eudesmol, hinesol, atractylon, and atractylodin contents using R (version 3.5.0). Broad sense heritability ($h_B^2$) was estimated from variance components in ANOVA as follows: $h_B^2 = \sigma_G^2 / (\sigma_G^2 + \sigma_E^2)$, where $\sigma_G^2$ = genotypic variance, $\sigma_E^2$ = environmental variance [11, 20]. Effective number of replicates ($r$) for the estimation of $\sigma_G^2$ was calculated using the following formula: $r = (\sum_{i=1}^a r_i - \sum_{i=1}^a r_i^2 / \sum_{i=1}^a r_i) / (a - 1)$, where $a$ = the number of clonal lines [22].
Results

Estimation of broad sense heritability for the contents of essential oil compounds

To evaluate the heritability of the contents of essential oil compounds in *A. lancea* under a microenvironment, 25 clonal lines were grown in Ibaraki prefecture in 2017 and the contents of the compounds in individual plantlets were determined. Figure 1 illustrates the distribution of each compound in different clonal lines. In most clonal lines, the ranges of variation in the contents of β-eudesmol, hinesol, atractylon, and atractylodin were lower than varietal differences, although some lines exhibited relatively high variations among individual plantlets in the same line, for example, line20 and line23 (Fig 1). One-way ANOVA and the estimation of broad sense heritability of those compound contents were performed using the above results. Table 1 shows that the differences in varietal compound contents were significant (P > 0.01) and the heritability for the contents of β-eudesmol, hinesol, atractylon, and atractylodin were high at 0.84, 0.77, 0.86, and 0.87, respectively (Table 1).

Correlation analyses among various compound contents and rhizome yield were performed, since there is potential linkage among different traits. According to the results, there was no significant correlation between rhizome yields and contents of atractylon or

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**Fig 1.** Range of variations for contents of the essential oil compounds in *A. lancea*. The boxes represent the values from the 25th to 75th percentile. The middle lines represent the median. The vertical lines extend from the minimum to the maximum values. Biological replicates within each clonal lines were as follows: line1–line17 (n = 20), line18–line24 (n = 10), line25 (n = 5). A; β-eudesmol contents, B; hinesol contents, C; atractylon contents, D; atractylodin contents.

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atractylodin, and the correlation coefficient between rhizome yields and contents of β-eudesmol and hinesol were relatively low, 0.20 and 0.28, respectively (Fig 2). In contrast, the correlation between β-eudesmol and hinesol contents was relatively high, although the correlations among the other compounds were comparatively low (Table 2).

### Environmental conditions of cultivation years and locations

Climatic datas for each cultivation years and locations were used the data issued by the Japan Meteorological Agency (JMA) [23]. Climatic conditions for the period of cultivation in Ibaraki prefecture were slightly different between 2016 and 2017. Mean temperatures in 2016 and 2017 were 14.9˚C and 15.4˚C, and accumulated rainfall in 2016 and 2017 were 1243 mm and 1196 mm, respectively. Environmental conditions of cultivation locations were more differed than that of cultivation years. Hokkaido is located in north of Japan, and mean temperature in Hokkaido is lower than mean temperature in Ibaraki prefecture. Mean temperatures for the period of cultivation in Hokkaido and Ibaraki prefecture were 15.4˚C and 8.3˚C, respectively. In addition, accumulated rainfall was differed between Hokkaido and Ibaraki prefecture. The accumulated rainfall for the period of cultivation in Hokkaido and Ibaraki prefecture was 1915 mm and 1487 mm, respectively. Further, there were difference in soil type and soil texture between 2 locations. The soil type and soil texture in Ibaraki prefecture were andosol and loam, and that in Hokkaido were alluvial soil and sandy loam.

### Effects of cultivation year on contents of the essential oil compounds

To examine the effect of cultivation year on contents of the essential oil compounds, six clones of *A. lancea* were grown in the same experimental field in 2016 and 2017, and contents of the compounds analyzed using two-way ANOVA. The effects of genotype (G), cultivation year (Y), and G × Y interaction were significant for contents of β-eudesmol, hinesol, and atractylon. However, mean squares of genotypes for each compound’s contents were higher than those of cultivation year and G × Y interaction (Table 3). In addition, interaction plots between genotype and cultivation year for β-eudesmol, hinesol, and atractylon showed that there were low qualitative interactions (Fig 3). Atractylodin contents were not significantly influenced by cultivation year, although there were significant differences for genotypes and G × Y interactions (Table 3). Fig 2D illustrates that qualitative interaction between genotype and cultivation year based on atractylodin contents was low, although in line5, the content levels of atractylodin in six clonal lines varied depending on cultivation year. In addition, the correlation coefficients

### Table 1. Broad-sense heritability for contents of the essential oil compounds in *A. lancea*.

| Essential oil compounds | Factors     | Df  | MeanSq | P-value | Effective replication | Genotypic variance | Environmental variance | Broad-sense heritability |
|-------------------------|-------------|-----|--------|---------|-----------------------|--------------------|------------------------|--------------------------|
| β-eudesmol              | Clonal lines | 24  | 539.1  | <2e-16** | 16.5                  | 32.2               | 6.3                    | 0.84                     |
|                         | Residuals   | 390 | 6.3    |         |                       |                    |                        |                          |
| hinesol                 | Clonal lines | 24  | 698.2  | <2e-16** | 16.5                  | 41.5               | 12.7                   | 0.77                     |
|                         | Residuals   | 390 | 12.7   |         |                       |                    |                        |                          |
| atractylon              | Clonal lines | 24  | 23.5   | <2e-16** | 16.5                  | 1.4                | 0.2                    | 0.87                     |
|                         | Residuals   | 390 | 0.2    |         |                       |                    |                        |                          |
| atractylodin            | Clonal lines | 24  | 6.1    | <2e-16** | 16.5                  | 0.4                | 0.1                    | 0.86                     |
|                         | Residuals   | 390 | 0.1    |         |                       |                    |                        |                          |

Df: degree of freedom, Mean Sq: Mean square
***: Probability value for test of significance < 0.01.
Fig 2. Relationships between contents of the essential oil compounds and rhizome weight in *A. lancea*. A; β-eudesmol contents, B; hinesol contents, C; atractylon contents, D; atractylodin contents.

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Table 2. Correlation matrix of β-eudesmol, hinesol, atractylon, and atractylodin contents.

|              | β-eudesmol | Hinesol | atractylon | atractylodin |
|--------------|------------|---------|------------|--------------|
| β-eudesmol   | -          | 0.59**  | 0.29**     | 0.09         |
| hinesol      | -          | -       | -0.13**    | 0.39**       |
| atractylon   | -          | -       | -          | -0.04        |
| atractylodin | -          | -       | -          | -            |

**: Probability value for test of significance < 0.01.

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between the two cultivation years for β-eudesmol, hinesol, atractylon, and atractylodin contents were 0.94, 0.94, 1.00, and 0.83, respectively (Fig 4).

Effects of cultivation location on the contents of essential oil compounds

To determine the effects of cultivation location on contents of the essential oil compounds, six clonal lines of \( A. \) lancea were cultivated in different locations. Two-way ANOVA results for β-eudesmol and hinesol contents revealed that there were significant differences in genotypes (G), locations (L), and G \( \times \) L interactions. In addition, mean squares of genotype were lower than those of location (Table 4). However, low qualitative interactions between genotype and location for contents of β-eudesmol and hinesol were observed (Fig 5). In addition, correlation coefficients for contents of β-eudesmol and hinesol between the two cultivation locations were 1.00 and 0.94, respectively (Fig 6).

Two-way ANOVA analyses of atractylon and atractylodin contents revealed that there were significant differences in genotypes, locations, and G \( \times \) L interaction. However, the mean squares of genotypes were higher than those of locations and G \( \times \) L interactions (Table 4). Fig 5C and 5D show that there were low qualitative interactions between genotype and location for contents of atractylon and atractylodin. In addition, correlation analyses of the contents of atractylon and atractylodin between the two cultivation locations revealed high correlations coefficients, 1.00 and 0.94, respectively (Fig 6).

Discussion

In the present study, broad sense heritability for β-eudesmol, hinesol, atractylon, and atractylodin contents in \( A. \) lancea was high (Table 1), suggesting that the contents of the essential oil compounds in \( A. \) lancea are highly influenced by genetic factors. In addition, correlations between each compound contents and rhizome yield were low (Fig 2), indicating that \( A. \) lancea strains, which have high contents of essential oil compounds could be selectively bred.
irrespective of rhizome yields. Therefore, cultivars of *A. lancea* with not only high contents of essential oil compounds but also high yields could be developed through selective breeding.

Correlation coefficients among different compound contents indicated low correlations, except in the case of the correlation between β-eudesmol and hinesol contents (Table 2). The biosynthetic pathways of β-eudesmol and hinesol are considered potentially closely associate since the compounds have similar chemical structures with the same molecular mass [24]. In addition, Takeda et al. [21] observed that the content ratio of β-eudesmol to hinesol keep constant in *A. lancea* clones. The above results could be the reason why the correlation coefficient between β-eudesmol and hinesol was high. When selectively breeding for the contents of the compounds, that the contents of β-eudesmol and hinesol could be linked. Conversely, the contents of the other compounds could be selected independently in the course of cultivar development.

We used the six clonal lines to investigate the influence of cultivation year on the contents of the essential oil compounds. Based on the results of the two-way ANOVA analyses, mean squares of genotype for contents of the essential oil compounds were higher than the interannual variability (Table 3). Additionally, there were low G × Y qualitative interactions (Fig 3), while the correlation coefficients between cultivation years and compound contents were relatively high (Fig 4). The results imply that, regardless of cultivation year, the contents of essential oil compounds in *A. lancea* clonal lines were stable, and genetic factors had greater influence than cultivation year did.
We cultivated six clones in two locations and evaluated the effects of cultivation location on the contents of essential oil compounds. The effect of cultivation location on the contents of essential oil compounds in *Atractylodes lancea* was larger than that of cultivation year, especially, the mean squares of cultivation location for $\beta$-eudesmol and hinesol contents were higher than those of genotype (Table 4). In general, secondary metabolites such as sesquiterpenes are induced by biological and abiotic stress via phytohormone signaling [25]. In *A. lancea*, the contents of essential oil compounds are induced by phytohormones such as jasmonic acid and ABA, via symbiosis with endophytes [14, 15]. In addition, increased soil acidity triggers an increase in $\beta$-eudesmol contents [16]. Therefore, the influence of cultivation location on the...
accumulation of β-eudesmol and hinesol in the present study could be induced by plant hormones due to biological or abiotic stress. It has been reported that mean temperature in their natural habitats are 11.1–16.0˚C, and accumulated rainfall are 850–1560 mm [13]. In the present study, there were differences in climatic and soil conditions between the 2 cultivation locations. Mean temperatures in Ibaraki were within the range of mean temperature for the natural habitats, whereas the average temperature of Hokkaido was lower than that of the natural habitats. It is possible that the contents of essential oil compounds in *A. lancea* could be influenced by these climatic differences. Furthermore, it has been reported that soil type and soil clay content are ecological factors having the high contribution rate against atractyloclin contents in *A. lancea* [26]. In the present study, soil type and soil texture were different between the 2 cultivation locations, and the contents of the essential oil compounds in *A. lancea* might be varied by these soil conditions. More detailed analyses on the influence of environmental factors are required, particularly on what are the major factors influencing the contents of the essential oil compounds.

Conversely, low qualitative interactions between genotype and location in the contents of β-eudesmol, hinesol, atractylon, and atractylodin were observed (Fig 5), and correlation coefficients between 2 locations for these compounds’ contents were high (Fig 6). The results suggest that genetic potential of *A. lancea* regarding the contents of β-eudesmol, hinesol, atractylon and atractylodin are stable regardless of environmental conditions, although the contents of the compounds are influenced by environmental factors as absolute values. It has been reported that artemisinin contents in *Artemisia annua* clones grown under different cultivation conditions were highly correlated [27]. The contents of β-eudesmol, hinesol, atractylon, and atractylodin in *A. lancea* exhibited similar results in the present study. The results indicate that the sesquiterpenoids and polyacetylene content levels among genotypes are not influenced by environmental conditions, although their absolute values varied depending on environments. Therefore, selective breeding could be effective in controlling the contents of essential oil compounds in *A. lancea*. In addition, cultivars exhibiting wide adaptation in the contents

### Table 4. Two-way ANOVA for β-eudesmol, hinesol, atractylon, and atractylodin contents in six *A. lancea* clones grown in Hokkaido and Ibaraki prefecture.

| Essential oil compounds | Factors    | Df   | MeanSq | P-value |
|-------------------------|------------|------|--------|---------|
| β-eudesmol              | Genotype (G) | 5    | 7.80   | 2.0E-16** |
|                         | Location (L) | 1    | 16.66  | 2.0E-16** |
|                         | G × L       | 5    | 0.24   | 2.2E-04** |
|                         | Residuals   | 206  | 0.05   |          |
| hinesol                 | Genotype (G) | 5    | 9.61   | 2.0E-16** |
|                         | Location (L) | 1    | 52.76  | 2.0E-16** |
|                         | G × L       | 5    | 1.58   | 2.3E-16** |
|                         | Residuals   | 206  | 0.08   |          |
| atractylon              | Genotype (G) | 5    | 0.83   | 2.0E-16** |
|                         | Location (L) | 1    | 0.23   | 2.0E-16** |
|                         | G × L       | 5    | 0.01   | 1.9E-08** |
|                         | Residuals   | 206  | 0.001  |          |
| atractylodin            | Genotype (G) | 5    | 0.16   | 2.0E-16** |
|                         | Location (L) | 1    | 0.01   | 4.6E-08** |
|                         | G × L       | 5    | 0.002  | 3.9E-05** |
|                         | Residuals   | 206  | 0.0004 |          |

Df: degree of freedom, Mean Sq: Mean square

**": Probability value for test of significance < 0.01.

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of essential oil compounds could be developed. Recent studies have proposed a putative sesquiterpenoid biosynthetic pathway based on transcriptome analyses of different *A. lancea* tissues, and several candidate genes related to sesquiterpenoid biosynthesis have been reported [28, 29]. As for sesquiterpenoid biosynthesis, β-eudesmol synthase mainly expressed in rhizomes has been identified in other plants such as *Zingiber zerumbet* [30]. Furthermore, high levels of SNPs (simple nucleotide polymorphisms) have been reported from leaf, stem, and root cDNA libraries of *A. lancea* [31]. The results suggest the potential of generating genetic markers for the contents of essential oil compounds in *A. lancea* using genetic association analysis.

In conclusion, the present study demonstrates that the contents of β-eudesmol, hinesol, atracytylon, and atractylodin in *A. lancea* are influenced mainly by genetic factors, and selective breeding could be an effective strategy for developing *A. lancea* populations that yield high contents of essential oil compounds. In addition, β-eudesmol, hinesol, atracytylon, and atractylodin contents could be selected regardless of rhizome yields in the course of *A. lancea* cultivar development. Consequently, *A. lancea* cultivars with high rhizome yields and high contents of each the essential oil compounds could be developed. However, since the contents of the essential oil compounds are influenced by environmental condition, further investigations on the effects of environment factors on the compound contents in *A. lancea* are required.

Fig 5. Interaction plots for contents of the essential oil compounds in six *A. lancea* clones grown in Hokkaido and Ibaraki prefectures. Data represent mean values of each compounds’ contents in *A. lancea* (n = 20 in line1, n = 12 in line2, n = 8 in line3, n = 20 in line4, n = 18 in line5, n = 20 in line6). A; β-eudesmol contents, B; hinesol contents, C; atracytylon contents, D; atractylodin contents.

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Supporting information

S1 Table. The data for Figs 1 and 2 and Tables 1 and 2. The contents of essential oil compounds in Atractylodes lancea grown in Ibaraki prefecture on 2017.
(PDF)

S2 Table. The data for Figs 3 and 4 and Table 3. The contents of essential oil compounds in A. lancea grown in each cultivation year.
(PDF)
S3 Table. The data for Figs 5 and 6 and Table 4. The contents of essential oil compounds in *A. lancea* grown in each cultivation location.

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**References**

1. Shiba M, Kondo K, Miki E, Yamaji H, Morota T, Terabayashi S, et al. Identification of medicinal Atractyloides based on ITS sequences of nrDNA. Biol Pharm Bull. 2006; 29: 315–320. PMID: 16462038
2. Japanese Pharmacopoeia, Society of Japanese Pharmacopoeia, 17th ed. Tokyo; 2016: 1803
3. Chinese Pharmacopoeia Commission. Chinese Pharmacopoeia (the first part). China Medical Science Press; 2015: 161.
4. Mizukami H, Okabe Y, Kohda H, Hiraoka N. Identification of the crude drug Atractyloides rhizome (Byaku-jutsu) and Atractyloides lancea rhizome (So-jutsu) using chloroplast TrnK sequence as a molecular marker. Biol Pharm Bull. 2000; 23: 589–594. PMID: 10823670
5. Kubo M, Nogami M, Nishimura M, Moriura T, Aichi S. Studies of crude drugs about its origin, process and quality. I. The preventive effects of chinese crude drug" Zhu" on experimental stomach ulcer and its pharmacological evaluation (1). Yakugaku Zasshi. 1983; 103: 442–448. PMID: 6631674
6. Wang HX, Liu CM, Liu Q, Gao K. Three types of sesquiterpenes from rhizomes of Atractyloides lancea. Phytochemistry. 2008; 69: 2088–2094. https://doi.org/10.1016/j.phytochem.2008.04.008 PMID: 18511090
7. Koonrungsesomboon N, Na-Bangchang K, Karbwang J. Therapeutic potential and pharmacological activities of Atractyloides lancea (Thunb.) DC. Asian Pac J Trop Med. 2014; 7: 421–428. https://doi.org/10.1016/S1995-7645(14)60089-9 PMID: 25066389
8. Nogami M, Moriura T, Kubo M, Tani T. Studies on the origin, processing and quality of crude drugs. II.: Pharmacological evaluation of the chinese crude drug "Zhu" in experimental stomach ulcer. (2). Inhibitory effect of extract of Atractyloides lancea on gastric secretion. Chem Pharm Bull. 1986; 34: 3854–3860. PMID: 3815607
9. Nakai Y, Kido T, Hashimoto K, Kase Y, Sakakibara I, Higuchi M, et al. Effect of the rhizomes of Atractyloides lancea and its constituents on the delay of gastric emptying. J Ethnopharmacol. 2003; 84: 51–55. PMID: 12499077
10. Nisikawa Y, Watanabe Y, Seto T. Studies on the evaluation of crude drugs (i) Comparative studies on the components of Atractyloides rhizomes. Syoyakugaku Zasshi. 1975; 29: 139–146.
11. Ukai Y. Genetic analysis of quantitative traits. Original ed. Tokyo: Igaku-Shuppan Inc.; 2002
12. Zhen O, Zhang L, Zhao M, Wang P, Wei Y, Fang J. Identification and quantification of sesquiterpenes and polyacetylenes in Atractylodes lancea from various geographical origins using GC-MS analysis. Braz J Pharmacogn. 2012; 22: 957–963.
13. Takeda O, Miki E, Terabayashi S, Okada M, Ye Lu, Hui-shen He, et al. Variation of essential oil components of Atractylodes lancea growing in China. Nat Med. 1995; 49: 18–23.
14. Ren CG and Dai CC. Jasmonic acid is involved in the signaling pathway for fungal endophyte-induced volatile oil accumulation of Atractylodes lancea plantlets. BMC Plant Biol. 2012; 12: 128. https://doi.org/10.1186/1471-2229-12-128 PMID: 22856333
15. Wang XM, Yang B, Ren CG, Wang HW, Wang JY, Dai CC. Involvement of abscisic acid and salicylic acid in signal cascade regulating bacterial endophyte-induced volatile oil biosynthesis in plantlets of Atractylodes lancea. Physiol Plant. 2015; 153: 30–42. https://doi.org/10.1111/ppl.12236 PMID: 24862990
16. Yuan Y, Liu YJ, Huang LQ, Cui GH, Fu GF. Soil acidity elevates some phytohormone and β-eudesmol contents in roots of Atractylodes lancea. Russ J Plant Physiol. 2009; 56: 133–137.
17. Takeda O, Miki E, Terabayashi S, Okada M, Lu Y, He HS, et al. A comparative study on essential oil components of wild and cultivated Atractylodes lancea and A. chinensis. Planta Med. 1996; 62: 444–449. https://doi.org/10.1055/s-2006-957936 PMID: 17252479
18. Kalton RR, Smit AG, Leffel RC. Parent-inbred progeny relationships of selected orchardgrass clones. Agron J. 1952; 44: 481–486.
19. McDonald ED, Kalton RR, Weiss MG. Interrelationships and relative variability among S1 and open-pollination progenies of selected bromegrass clones. Agron J. 1952; 44: 20–25.
20. Burton GW, Devane EH. Estimating heritability in tall fescue (Festuca arundinacea) from replicated clonal material. Agron J. 1953; 45: 478–481.
21. Takeda O, Miki E, Terabayashi S, Okada M, Shan-an HE YS. Seasonal variation of essential oil components in Atractylodes lancea (THUNB.) DC. propagated by division of their rhizomes. Chem Pharm Bull. 1997; 45: 1527–1529.
22. Yoshida M. Design of experiments for animal husbandry. 8th ed. Tokyo: Yokendo co.ltd.; 1998
23. Japan Meteorological Agency. Past meteorological data download; 2019. Database: figshare [Internet]. Available from: https://www.data.jma.go.jp/gmd/risks/obsdl/index.php
24. Yosioka I, Takahashi S, Hikino H, Sasaki Y. Studies on the constituents of Atractylodes. III. Separation of atractylol into eudesmol and hinesol. Chem Pharm Bull. 1958; 7(3): 319–323.
25. Sudha G, Ravishankar GA. Involvement and interaction of various signaling compounds on the plant metabolic events during defense response, resistance to stress factors, formation of secondary metabolites and their molecular aspects. Plant Cell Tissue Organ Cult. 2002; 71: 181–212.
26. Shoudong Z, Huasheng P, Lanping G, Tongren X, Yan Z, Meilan C, et al. Regionalization of Chinese material medical quality based on maximum entropy model: A case study of Atractylodes lancea. Sci Rep. 2017; 7: 42417 https://doi.org/10.1038/srep42417 PMID: 28205539
27. Jorge F. S. Ferreira J. E. Simon Janick J. Relationship of artemisinin content of tissue-cultured, house-grown, and field-grown plants of Artemisia annua. Planta Med. 1995; 61: 351–355. https://doi.org/10.1055/s-2006-958098 PMID: 17238088
28. Chen F, Wei YX, Zhang JM, Sang XM, Dai CC. Transcriptomics analysis investigates sesquiterpenoids accumulation pattern in different tissues of Atractylodes lancea (Thunb.) DC. plantlet. Plant Cell Tissue Organ Cult. 2017; 130: 73–90.
29. Huang Q, Huang X, Deng J, Liu H, Liu Y, Yu K, et al. Differential gene expression between leaf and rhizome in Atractylodes lancea: A comparative transcriptome analysis. Front Plant Sci. 2016; 7: 1–13. https://doi.org/10.3389/fpls.2016.00001
30. Yu F, Harada H, Yamasaki K, Okamoto S, Hirase S, Tanaka Y, et al. Isolation and functional characterization of a β-eudesmol synthase, a new sesquiterpene synthase from Zingiber zerumbet Smith. FEBS Lett. 2008; 582: 565–572. https://doi.org/10.1016/j.febslet.2008.01.020 PMID: 18242187
31. Ahmed S, Zhan C, Yang Y, Wang X, Yang T, Zhao Z, et al. The transcript profile of a traditional Chinese medicine, atractylodes lancea, revealing its sesquiterpenoid biosynthesis of the major active components. PLoS One. 2016; 11: 1–19.