Ultrafast Excited-State Decay Mechanisms of 6-Thioguanine Followed by Sub-20 fs UV Transient Absorption Spectroscopy

Danielle C. Teles-Ferreira 1, Cristian Manzoni 2, Lara Martínez-Fernández 3, Giulio Cerullo 2,4,*, Ana Maria de Paula 5 and Rocío Borrego-Varillas 2,4,*

1 Instituto Federal de Minas Gerais, Campus Ouro Preto, Ouro Preto 35400-000, MG, Brazil; danielle.teles@ifmg.edu.br
2 Istituto di Fotonica e Nanotecnologie-Consiglio Nazionale delle Ricerche (IFN-CNR), Piazza Leonardo da Vinci 32, I-20133 Milano, Italy; cristian.manzoni@polimi.it (C.M.); giulio.cerullo@polimi.it (G.C.)
3 Departamento de Química, Facultad de Ciencias and Institute for Advanced Research in Chemistry (IADCHEM), Campus de Excelencia UAM-CSIC, Universidad Autónoma de Madrid, Cantoblanco, 28049 Madrid, Spain; lara.martinez@uam.es
4 Departamento de Física, Politecnico di Milano, Piazza Leonardo da Vinci 32, I-20133 Milano, Italy
5 Departamento de Física, Universidade Federal de Minas Gerais, Belo Horizonte 31270-901, MG, Brazil; ana@fisica.ufmg.br
* Correspondence: rocio.borrego@polimi.it

Abstract: Understanding the primary steps following UV photoexcitation in sulphur-substituted DNA bases (thiobases) is fundamental for developing new phototherapeutic drugs. However, the investigation of the excited-state dynamics in sub-100 fs time scales has been elusive until now due to technical challenges. Here, we track the ultrafast decay mechanisms that lead to the electron trapping in the triplet manifold for 6-thioguanine in an aqueous solution, using broadband transient absorption spectroscopy with a sub-20 fs temporal resolution. We obtain experimental evidence of the fast internal conversion from the S2(ππ*) state to the S1(ππ*) states, which takes place in about 80 fs and demonstrates that the S1(ππ*) state acts as a doorway to the triplet population in 522 fs. Our results are supported by MS-CASPT2 calculations, predicting a planar S2(ππ*) pseudo-minimum in agreement with the stimulated emission signal observed in the experiment.

Keywords: ultrafast spectroscopy; thiobases; global analysis

1. Introduction

In 1950, George Hitchings and Gertrude Elion [1] obtained, by exchanging oxygen with sulphur in nucleobases, a potent drug for the treatment of numerous diseases [2–5]. This discovery led them to be honoured with the Nobel Prize in 1988 in the category of Physiology and Medicine. Such nucleobase derivatives are known as thiobases, and there has been a great effort to unravel their energy deactivation pathways [6,7]. The sulphur substitution in the nucleobases causes noticeable changes in the photophysics of these molecules [8–19] in comparison with the canonical nucleobases. There is a redshift of the absorption spectrum and a loss of photostability when excited by ultraviolet (UV) light. Unlike the canonical bases, which quickly repopulate the ground state after UV photoexcitation via an internal conversion (IC) process mediated by conical intersections (CIs) [20,21], the thiobases population mostly decays to a long-lived triplet state with a high quantum yield via an ultrafast intersystem crossing (ISC) process [12,22–24].

Due to the high transfer efficiency to the triplet states and, consequently, the high rate of singlet oxygen formation, several applications of the thiobases have been demonstrated. They are used as chemotherapeutic agents for the treatment of some types of skin cancer, as they enhance the polymerase chain reaction for DNA/RNA replication, they are used as chromophores for investigating specific interactions between nucleic acids and...
proteins [25–33]. In addition to their practical applications, they provide an excellent model system for understanding how a single atom substitution can modify the energy relaxation processes of canonical DNA/RNA nucleobases.

One of the open questions in the ultrafast dynamics of thiobases concerns the mechanism behind the efficient triplet manifold formation by ISC, which occurs shortly after the bright, excited singlet state is populated. In the literature, at least two hypotheses have been proposed: (i) the population of the triplet states mediated by a dark singlet state [12,18,22,34–37] and (ii) the direct transition from the bright singlet state to the triplet states [12,18,22–24,34–36,38]. We have recently observed a third possibility: for water-solvated thiouracils, the triplet manifold is accessed by parallel channels. While part of the population decays directly from the photoexcited bright state, another part decays passing through a dark singlet state [36,39].

In the last few years, the process of triplet state formation by ISC in 6-thioguanine (6TG) has received considerable attention in the literature [6,18,23,40–45]. Time-resolved transient absorption and photoelectron spectroscopies have been applied both in solution and gas phase for the investigation of the ultrafast dynamics in time scales, ranging from hundreds of femtoseconds up to nanoseconds. However, the primary steps after UV photoexcitation are not experimentally documented, since technical challenges have long prevented access to the sub-100 fs time scale, a highly desired benchmark as these mechanisms determine the system response on longer time scales.

In this work, we investigate the ultrafast dynamics of buffered, solvated 6TG by means of broadband transient absorption spectroscopy (TAS) with sub-20 fs UV pump pulses. Thanks to our high temporal resolution, which exceeds by a one-order magnitude that of previous studies on this molecule, and the broad spectral probing window which extends from the UV to the visible range, we are able to obtain mechanistic insight regarding the ultrafast primary processes that lead to the formation of the triplet manifold and the time constants associated with it. In particular, we show that the internal conversion from $S_2(\pi\pi^*)$ to the $S_1(n\pi^*)$ state takes place in about 80 fs, with the $S_1(n\pi^*)$ acting as a doorway to the triplet population in approximately 500 fs. Therefore, our results demonstrate that the decay channel $S_2(\pi\pi^*) \rightarrow S_1(n\pi^*) \rightarrow T_1(\pi\pi^*)$, experimentally observed so far only in thiouracils [36,39], is more general and can also be found in purine thiobases.

2. Results and Discussion

Figure 1a shows the 6TG steady-state linear absorption spectrum (black) and the pump pulse spectrum used for the TAS measurements (blue), while Figure 1b displays the emission spectrum. The lowest energy absorption band of 6TG presents a maximum at 340 nm (Figure 1a, black curve). Several studies [43,46,47] have assigned this band to the $S_0(\pi\pi^*) \rightarrow S_2(\pi\pi^*)$ transition, which is the optically active state populated by the sub-20 fs pump pulse in TAS experiments.

Figure 1b shows the steady-state photoluminescence (PL) spectra of buffered, solvated, and $N_2$-pumped 6TG excited by a lamp at 340 nm. We observe a broad band that extends from 350 nm to 550 nm. Our PL spectrum is in line with the observations of Zou et al. [16]. In contrast, Reichardt et al., did not identify emission from 6TG within the sensitivity of their spectrometer [23]. Ashwood et al., observed two bands for an emission measurement at 77 K in a tris buffer 7.4 pH matrix: a weak one from 350 nm to 420 nm and another intense one from 430 nm to 550 nm [43]. The authors assigned the first band (350–420 nm, peaking at 385 nm) to the optically active state populated by the sub-20 fs pump pulse in TAS experiments.

Figure 1b shows the steady-state photoluminescence (PL) spectra of buffered, solvated, and $N_2$-pumped 6TG excited by a lamp at 340 nm. We observe a broad band that extends from 350 nm to 550 nm. Our PL spectrum is in line with the observations of Zou et al. [16]. In contrast, Reichardt et al., did not identify emission from 6TG within the sensitivity of their spectrometer [23]. Ashwood et al., observed two bands for an emission measurement at 77 K in a tris buffer 7.4 pH matrix: a weak one from 350 nm to 420 nm and another intense one from 430 nm to 550 nm [43]. The authors assigned the first band (350–420 nm, peaking at 385 nm) to the fluorescence emission. The intense band from 430 nm to 550 nm was attributed to the phosphorescence emission, which peaks at 480 nm, in reasonable agreement with the vertical excitation energy calculated for the $T_1(3\pi\pi^*)$ state at 454 nm [45] in water. As we show data at room temperature and the sample was pumped with $N_2$ (in order to avoid the formation of singlet oxygen and photoluminescence quenching), we expect an overlap of the contributions from phosphorescence and fluorescence. A decomposition of the PL spectrum reveals two Gaussians centred at 398 nm (red curve in Figure 1b) and 454 nm (green curve in Figure 1b). It should be noted that there is a
gap in the data from 374 to 393 nm due to a subtraction of a scattering peak from the excitation lamp. Table 1 reports the energies and oscillator strengths of 6TG in vacuum calculated at the MS-CASPT2//CASSCF(14,12)/ANO-L level of theory [48–52]. P1, P2, and P3 (coordinates reported in the Supplementary Materials, Table S1) are three single points along the minimum energy path relaxing the S\(_2\)(\(\pi\pi^*\)) state from the Franck–Condon region (Supplementary Materials, Figure S1). Their close adiabatic energies suggest a planar S\(_2\)(\(\pi\pi^*\)) pseudo-minimum. Since the maximum of the red curve in Figure 1b matches the emission wavelength calculated for this pseudo-minimum, we assign it to the fluorescence emission from the S\(_2\)(\(\pi\pi^*\)) state. The peak of the green curve is instead attributed to the phosphorescence from the T\(_1\)(\(3\pi\pi^*\)) state, as also predicted by previous works [7,43,53].

We performed broadband TAS experiments for 6TG in PBS solution by pumping it with sub-20 fs UV pulses centred at 330 nm and probing in a broad spectral window from 275 to 630 nm. The acquired TAS map and the transient spectra at selected time delays are displayed in Figure 2a,b, respectively, while Panel c shows the dynamics at selected wavelengths. An intense negative band peaking at 340 nm is partially superimposed with the lowest-energy band observed in the steady-state linear absorption spectra (Figure 1a), and is therefore assigned to ground state bleaching (GSB). At early times (<100 fs), we observed a negative band at about 375 nm, which is assigned to stimulated emission (SE) from the S\(_2\)(\(\pi\pi^*\)) state. The peak of the SE band at 375 nm is in satisfactory agreement with the vertical emission energy calculated from single points at S\(_2\)(\(\pi\pi^*\)) (P\(_1\), P\(_2\), and P\(_3\) in Figure 3a and Table 1) in vacuum. The estimated SE wavelength is also in line with the broadband fluorescence emission shown in Figure 1b, taking into account the experimental
uncertainties of determining the position of the peaks due to the overlapping signals. The SE band completely decays in less than 80 fs, leaving two photoinduced absorption (PA) bands, PA$_1$ peaking at 300 nm and the broader PA$_3$ peaking about 520 nm.

Figure 2. (a) TAS map of solvated 6TG as a function of the pump–probe delay when excited by a 330-nm pulse; (b) transient spectra at the time delays indicated in the legend; (c) dynamics at selected wavelengths during the first 500 fs. (d) Evolution associated spectra (EAS) obtained after the analysis of the TAS map shown in Panel (a).

Figure 3. Possible 6TG triplet formation mechanism upon UV photoexcitation. (a) Scheme of the potential energy curves in the vicinity of the S$_2$(ππ*)/S$_1$(nπ*) CI: a UV photon (purple arrow) excites the S$_2$(ππ*) state (yellow). Relaxation of the wave packet from S$_2$(ππ*) to S$_1$(nπ*) (blue) is mediated by a CI. (b) IC from S$_2$(ππ*) to S$_1$(nπ*) occurs in tens of femtoseconds as predicted in [18]. From here, several decay pathways are open: population of S$_1$(ππ*), which decays by IC to the ground state in tens of picoseconds [43], and ISC to the triplets in 520 fs from S$_1$(nπ*).
At later times (>200 fs), we observed the rise of another PA band peaking at ≈375 nm (PA2), while the PA3 band remained substantially unchanged. Previous TAS measurements with 200 fs of temporal resolution have also shown long-lived PA bands centred at about 390 nm and 520 nm, which were identified as signatures of the triplet state based on their theoretical calculations [23,43].

To interpret our results, we review the previous computational studies on 6TG. Photoexcitation by the UV pump pulse creates a wave packet in the bright state, S2(ππ*). Reichardt et al., by combining quantum-chemical calculations and TAS with 200 fs pulses, suggested that for 6TG, most of the population accesses the triplet manifold directly from the excited state, S2(ππ*), while a minor fraction decays by IC to the S1(ππ*), which, in-turn relaxes non-radiatively to the ground state in tens of picoseconds [23]. Theoretical studies based on MS-CASPT2/CASSCF [46] and TD-DFT [43,45] calculations have found two S1 minima: one of ππ* nature and another of ππ character. They proposed that the population from the S2(ππ*) state decays via IC in a barrierless fashion in tens of femtoseconds to the S1(ππ*), which acts as a doorway to the triplet manifold. The S1(ππ*) minimum (see Figure 3a) enables a CI to the ground state of about 25% of the population [43] in tens of picoseconds [18]. From the S1(ππ*) minimum, there is an efficient population of the T1(ππ*) and the T2(ππ*) triplet states, which were found to be isoenergetic [18,46]. Then, the population is trapped in the bright triplet state for hundreds of nanoseconds [18,43].

The evolution-associated spectra (EAS) for each species were obtained by a global fit of the TAS data using a sequential kinetic model (Figure 2d). The best fit was obtained with two time constants (τ1 = 81 ± 30 fs and τ2 = 522 ± 48 fs) and an offset. The offset in the global analysis refers to time constants that are beyond our 2 ps temporal observation window (blue curve in Figure 2d). This EAS (blue curve in Figure 2d) peaks at 375 nm, with a broad band extending from 400 to 650 nm. These spectral signatures are typical of the long-lived triplet state, which has a lifetime estimated to be 830 ns [43]. The 81 ± 30 fs time constant is reported for the first time in this study (black curve in Figure 2d). Four signatures are distinguishable in the short-lived EAS. The main negative peak at 340 nm is assigned to the GSB, which is overlapped with the red-shifted negative signal at around 375 nm. The latter is assigned to the SE from the photoexcited state, S2(ππ*), for the following reasons: (i) it is in satisfactory agreement with the vertical emission energy calculated for the S2(ππ*) state (Table 1: P1, P2, and P3 single points); (ii) it matches with the broad fluorescence emission maximum at 385 nm [43], and it lies within the broadband PL (Figure 1b). Therefore, we estimate that the IC from S2(ππ*) to S1(ππ*) occurs on a time scale of 81 ± 30 fs, which is in line with theoretical calculations that predicted IC in about 50 fs [18]. The two PA bands at 300 nm and from 380 to 650 nm could be excited-state absorption signals from S2(ππ*) since they evolve on the same time scale of the SE, but we might not exclude a minor direct channel to the triplet manifold [18]. In addition, due to the planarity of the S2(ππ*) potential energy surface, there might be some part of the population trapped in the S2(ππ*), which decays at later times. Higher-level computations including spectroscopic signals of the corresponding electronic states are needed for an unambiguous assignment. The mismatch between τ1 and τ2 and the observation of SE from S2(ππ*) delivers the experimental confirmation about the involvement of the dark S1(ππ*) mediating the ISC process, as suggested by previous theoretical works for 6TG [18,43]. This shows that the S1(ππ*) state acts as a doorway to the triplet manifold, as also observed experimentally in some thiopyrimidines [36,39]. The τ2 time constant is assigned to the lifetime of the S1(ππ*) state (red curve in Figure 2d). It is the same value (within the error margin) reported by Ashwood et al., [43] (0.56 ± 0.06 ps), which was assigned to the formation of the triplet manifold.

Figure 3 summarises the proposed photoexcitation scenario for 6TG. Excitation at 340 nm generates a wave packet in the S2(ππ*) state, the signature for which is identified as an SE band with a time constant as fast as 81 fs in the TAS experiments. The CASPT2/CASSCF(14,12)/ANO-L calculations of the potential energy surface reveal a planar character of the S2(ππ*) state before reaching the CI to the S1 state, which acts as a
doorway to access the triplet manifold in 522 fs. Our findings therefore provide experimental evidence of the involvement of the intermediate dark singlet state and highlight $S_2(\pi\pi^*)\rightarrow S_1(n\pi^*)\rightarrow T_1(\pi\pi^*)$ as the major decay pathway in 6TG.

3. Materials and Methods

3.1. Sample Preparation

6TG was purchased from Sigma-Aldrich (Sigma-Aldrich Chemie GmbH, Taufkirchen, Germany) with 98% purity. PBS solution was prepared by dissolving 0.15 g of monosodium phosphate and 0.27 g of sodium disodium phosphate in 200 mL of ultrapure water to obtain a pH of 7.4 and a concentration of 16 mM. The 6TG in PBS solution was prepared to obtain a maximum absorbance around 1 OD at the central wavelength of the pump beam (1.4 mM).

3.2. Linear Absorption

The linear absorption (LA) spectrum of each sample in PBS solution was measured using a JASCO Corp. V-570 (Jacso Inc., Easton, MD, USA) spectrometer at room temperature. The samples were placed in a 1 mm internal optical-path quartz cuvette. The spectra (Figure 1a) were background-corrected by subtracting the solvent spectrum measured under the same experimental conditions. The LA spectra were collected before and after each TAS experiment to assure that the sample had not degraded during the experiments.

3.3. Steady-State Photoluminescence

We acquired the steady-state emission spectra at room temperature using a Cary Eclipse spectrophotometer (Varian Inc., Palo Alto, CA, USA) with a lamp excitation. The spectra were measured at 1000 V of photomultiplier voltage with excitation and emission slit widths of 5 nm and averaging times of 0.9 s. The sample concentrations used were about 0.02 mM, and the contribution from the solvent solution was subtracted. After preparation, the sample was pumped with N$_2$ to avoid O$_2$ dissolved in the solution that could lead to the formation of singlet oxygen and photoluminescence quenching.

3.4. Transient Absorption Spectroscopy

TAS measurements were carried out with a home-made setup [54] fed by an 800 nm Ti:Sapphire laser system (Libra, Coherent, Santa Clara, CA, USA), which delivers 100 fs pulses at a 1 kHz repetition rate. A fraction of the beam seeds a broadband, visible non-collinear optical parametric amplifier (NOPA), which was mixed with the fundamental wave in a 50 µm-thick Type II β-barium borate crystal to generate broadband UV pump pulses at 330 nm [55]. Probe pulses covering the 275–700 nm spectral range were obtained by white light generation in a 2 mm-thick CaF$_2$ plate. The data were collected separated in two ranges: UV (275–340 nm) with thick CaF$_2$ plate pumped at 400 nm and visible (330–650 nm) with thick CaF$_2$ plate pumped at 800 nm. The two datasets were merged to obtain the data shown in Figure 2. The setup’s instrumental response function is estimated to be 20–30 fs (depending on the probe wavelength), and ultimately limits the precision of the determination of the faster time constant. For the measurements presented in this study, the pump energy was limited to 27 nJ (leading to a fluence of 340 µJ·cm$^{-2}$) to avoid sample photodamage and solvated electrons generated by two-photon absorption from the solvent. A flowing jet configuration was employed for the sample that was flown at a typical rate of 40 rpm. The experiments were performed using pump and probe beams with parallel polarisations.

3.5. Global Analysis and Data Processing

The TAS datasets were subjected to a global analysis using the free software Glotaran [56,57]. A singular-value decomposition was first performed in order to find the number of linearly independent vectors (decay constants) significantly different from the noise. We found three dominant components and three residual components associated
with the coherent artefact. A nonlinear, least-squares fit was then performed on the TAS map using a sequential kinetic model with three exponential functions. The time-dependent spectrum is thus represented by the function \( \psi(\lambda, t) \), and depends on the wavelengths of the pump and the probe:

\[
\psi(\lambda, t) \propto \sum_{l=1}^{n_{\text{comp}}} (e^{k_i t} * \text{IRF}(t)) \text{EAS}_l(\lambda)
\]

where \( k_i \) is the decay rate for each component, \( \text{IRF}(t) \) is the instrumental response function, and \( \text{EAS}_l(\lambda) \) are the amplitudes of the exponential decays, known as evolution-associated spectra (EAS).

The analysis also takes into account the coherent artefact, the oscillatory time profile of which was modelled by a sequence of exponentials and the dispersion due to the chirp of the probe (modelled by a third-order polynomial).

At least three independent datasets (i.e., recorded on three different days) were used in the analysis, and all uncertainties are reported as twice the standard deviation of the average lifetime. The larger error in the determination of the faster time constant stems from the presence of the coherent artefact, which affects the dynamics in the first 100 fs.

4. Conclusions

From experimental data obtained by TAS with sub-20 fs pump pulses and broadband probing in a joint UV and visible window, combined with global analysis, we demonstrate the involvement of a dark singlet state \( S_1(n\pi^*) \) in the decay mechanism of 6TG, which acts as a doorway to the triplet manifold. The ultrafast data are supported by theoretical studies and establish that the major decay pathway in 6TG to the triplet manifold is as follows: \( S_2(\pi\pi^*) \rightarrow S_1(n\pi^*) \rightarrow T_1(\pi\pi^*) \).

Supplementary Materials: The following supporting information can be downloaded online, Figure S1: Minimum energy path from which the single points calculations were extracted. CASSCF (14,12)/ANO-L level of theory.; Table S1: XYZ coordinates.

Author Contributions: Conceptualization, G.C., C.M. and A.M.d.P.; methodology, D.C.T.-F., C.M. and R.B.-V.; software, L.M.-F.; formal analysis, D.C.T.-F., L.M.-F., A.M.d.P. and R.B.-V.; investigation, D.C.T.-F., G.C., A.M.d.P. and R.B.-V.; writing—original draft preparation, D.C.T.-F. and R.B.-V.; writing—review and editing, G.C., A.M.d.P. and R.B.-V.; visualization, D.C.T.-F.; supervision, C.M., G.C., A.M.d.P. and R.B.-V.; project administration, G.C. and A.M.d.P.; funding acquisition, G.C. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Ministerio de Ciencia e Investigación (grant n. PID2019-110091GB-I00) and the European Commission (grant n. 765266, LightDyNAmics).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data are available from the authors upon reasonable request.

Acknowledgments: D.C.T.F. and A.M.d.P. acknowledge financial support from the funding agencies Fapemig, CNPq, and CAPES. L.M.F. thanks the PID2019-110091GB-I00 funded by the MCIN/AEI/10.13039/501100011033 project for financial support and the Centro de Computación Científica UAM (CCC-UAM) for computing time. G.C. acknowledges support from the H2020 Grant Agreement number 765266 (LightDyNAmics).

Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Not available.
References

1. Hitchings, G.H.; Elion, G.B.; Falco, E.A.; Russell, P.B.; Sherwood, M.B.; VanderWerff, H. Antagonists of nucleic acid derivatives. *J. Biol. Chem.* **1950**, *183*, 1–9. [CrossRef]

2. Pollum, M.; Jockusch, S.; Crespo-Hernández, C.E. Increase in the photoactivity of uracil derivatives by doubling thionation. *Phys. Chem. Chem. Phys.* **2015**, *17*, 27851–27861. [CrossRef] [PubMed]

3. Jordheim, L.P.; Durante, D.; Zoulim, F.; Dumontet, C. Advances in the development of nucleoside and nucleotide analogues for cancer and viral diseases. *Nat. Rev. Drug Discov.* **2013**, *12*, 447–464. [CrossRef] [PubMed]

4. Attard, N.R.; Karran, P. UVA photosensitization of thiopurines and skin cancer in organ transplant recipients. *Photochem. Photobiol. Sci.* **2012**, *11*, 62–68. [CrossRef]

5. Pridgeon, S.W.; Heer, R.; Taylor, G.A.; Newell, D.R.; O’Toole, K.; Robinson, M.; Xu, Y.Z.; Karran, P.; Boddy, A.V. Thiothymidine combined with UVA as a potential novel therapy for bladder cancer. *Br. J. Cancer* **2011**, *104*, 1869–1876. [CrossRef]

6. Arslancan, S.; Martínez-Fernández, L.; Corral, I. Photophysics and photochemistry of canonical nucleobases’ thioanalogs: From quantum mechanical studies to time resolved experiments. *Molecules* **2017**, *22*, 998. [CrossRef]

7. Ashwood, B.; Pollum, M.; Crespo-Hernández, C.E. Photochemical and Photodynamical Properties of Sulphur-Substituted Nucleic Acid Bases. *Photochem. Photobiol.* **2019**, *95*, 33–58. [CrossRef]

8. Nenov, A.; Conti, I.; Borrego-Varillas, R.; Cerullo, G.; Garavelli, M. Linear absorption spectra of solvated thiouracils resolved at the hybrid RASPT2/MM level. *Chem. Phys.* **2018**, *515*, 643–653. [CrossRef]

9. Middleton, C.T.; de La Harpe, K.; Su, C.; Law, Y.K.; Crespo-Hernández, C.E.; Kohler, B. DNA Excited-State Dynamics: From Single Bases to the Double Helix. *Annu. Rev. Phys. Chem.* **2009**, *60*, 217–239. [CrossRef]

10. Pecourt, J.M.L.; Peon, J.; Kohler, B. Ultrafast internal conversion of electronically excited RNA and DNA nucleosides in water. *J. Am. Chem. Soc.* **2000**, *122*, 9348–9349. [CrossRef]

11. Harada, Y.; Okabe, C.; Kobayashi, T.; Suzuki, T.; Ichimura, T.; Xu, Y.Z. Ultrafast intersystem crossing of 4-thiothymidine in aqueous solution. *J. Phys. Chem. Lett.* **2010**, *1*, 480–484. [CrossRef]

12. Sánchez-Rodríguez, J.A.; Mohamadzade, A.; Mai, S.; Ashwood, B.; Pollum, M.; Marquetand, P.; González, L.; Crespo-Hernández, C.E.; Ulrich, S. 2-Thiouracil intersystem crossing photodynamics studied by wavelength-dependent photoelectron and transient absorption spectroscopies. *Phys. Chem. Chem. Phys.* **2017**, *19*, 19756–19766. [CrossRef]

13. Jiang, J.; Zhang, T.-S.; Xue, J.-D.; Zheng, X.; Cui, G.; Fang, W.-H. Short-time dynamics of 2-thiouracil in the light absorbing S2(ππ*) state. *J. Chem. Phys.* **2015**, *143*, 175103. [CrossRef]

14. Koyama, D.; Milner, M.J.; Orr-Ewing, A.J. Evidence for a Double Well in the First Triplet Excited State of 2-Thiouracil. *J. Phys. Chem. B* **2017**, *121*, 9274–9280. [CrossRef]

15. Bai, S.; Barbati, M. Why Replacing Different Oxygens of Thymine with Sulfur Causes Distinct Absorption and Intersystem Crossing. *J. Phys. Chem. B* **2015**, *119*, 27851–27861. [CrossRef]

16. Martínez-Fernández, L.; Corral, I.; Granucci, G.; Persico, M. Competing ultrafast intersystem crossing and conversion involving: A time resolved picture for the deactivation of 6-thioguanine. *Chem. Sci.* **2014**, *5*, 1336. [CrossRef]

17. Ruckenbauer, M.; Mai, S.; Marquetand, P.; González, L. Photoelectron spectra of 2-thiouracil, 4-thiouracil, and 2,4-dithiouracil. *J. Chem. Phys.* **2016**, *144*, 074303. [CrossRef]

18. Cerullo, G.; Garavelli, M. A novel spectroscopic window on conical intersections in biomolecules. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 26553–26555. [CrossRef]

19. Borrego-Varillas, R.; Nenov, A.; Kabacinski, P.; Conti, I.; Ganzer, L.; Oriana, A.; Delno, I.; Weingart, O.; Manzoni, C.; Rivalta, I.; et al. Tracking excited state decay mechanisms of pyrimidine nucleosides in real time. *Nat. Commun.* **2021**, *12*, 7285. [CrossRef] [PubMed]

20. Xie, B.-B.; Wang, Q.; Guo, W.-W.; Cui, G. The excited-state decay mechanism of 2,4-dithiothymine in the gas phase, microsolvated surroundings, and aqueous solution. *Phys. Chem. Chem. Phys.* **2017**, *19*, 7689–7698. [CrossRef] [PubMed]

21. Reichhardt, C.; Guo, C.; Crespo-Hernández, C.E. Excited-state dynamics in 6-thioguanosine from the femtosecond to microsecond time scale. *J. Phys. Chem. B* **2011**, *115*, 3263–3270. [CrossRef] [PubMed]

22. Reichhardt, C.; Crespo-Hernández, C.E. Room-temperature phosphorescence of the DNA monomer analogue 4-thiouracil in aqueous solutions after UVA excitation. *J. Phys. Chem. Lett.* **2010**, *1*, 2239–2243. [CrossRef]

23. Pollum, M.; Jockusch, S.; Crespo-hernández, C.E. 2,4-Dithiouracil as a Potent UVA Chemotherapeutic Agent. *J. Am. Chem. Soc.* **2014**, *136*, 17930–17933. [CrossRef] [PubMed]

24. Sismour, A.M. The use of thymidine analogs to improve the replication of an extra DNA base pair: A synthetic biological system. *Nucleic Acids Res.* **2005**, *33*, 5640–5646. [CrossRef]

25. Heuberger, B.D.; Pal, A.; Del Frate, F.; Topkar, V.V.; Szostak, J.W. Replacing Uridine with 2-Thiouracil Enhances the Rate and Fidelity of Nonenzymatic RNA Primer Extension. *J. Am. Chem. Soc.* **2015**, *137*, 2769–2775. [CrossRef]

26. Favre, A.; Moreno, G.; Blondel, M.O.; Klíber, J.; Vinzenz, F.; Salet, C. 4-thiouridine photosensitized RNA-protein crosslinking in mammalian cells. *Biochem. Biophys. Res. Commun.* **1996**, *184*, 847–854. [CrossRef]
29. Pollum, M.; Lam, M.; Jockusch, S.; Crespo-Hernández, C.E. Dithionated Nucleobases as Effective Photodynamic Agents against Human Epidermoid Carcinoma Cells. *ChemMedChem* **2018**, *13*, 1044–1050. [CrossRef]

30. Harris, M.; Cote, H.; Oehoa, C.; Allavena, C.; Negredo, E.; Cahn, P.; Zala, C.; Raffi, F. A Randomized, Open-Label Study of a Nucleoside Analogue Reverse Transcriptase Inhibitor—Sparing Regimen in Antiretroviral-Naive HIV-Infected Patients. *J. Acquir. Immune Defic. Syndr.* **2009**, *50*, 339–340. [CrossRef]

31. Meisenheimer, K.M.; Koch, T.H. Photocross-Linking of Nucleic Acids to Associated Proteins. *Crit. Rev. Biochem. Mol. Biol.* **1997**, *32*, 101–140. [CrossRef]

32. Reelfs, O.; Karran, P.; Young, A.R. 4-thiouridylate sensitization of DNA to UVA offers potential for a novel photochemotherapy. *Photochem. Photobiol. Sci.* **2012**, *11*, 148–154. [CrossRef]

33. Massey, A.; Xu, Y.Z.; Karran, P. Photoactivation of DNA thiobases as a potential novel therapeutic option. *Curr. Biol.* **2001**, *11*, 1142–1146. [CrossRef]

34. Martínez-Fernández, L.; Granucci, G.; Pollum, M.; Crespo-Hernández, C.E.; Persico, M.; Corral, I. Decoding the Molecular Basis for the Population Mechanism of the Triplet Phototoxic Precursors in UVA Light-Activated Pyrimidine Anticancer Drugs. *Chem.—A Eur. J.* **2017**, *23*, 2619–2627. [CrossRef]

35. Cui, G.; Fang, W.-H. State-specific heavy-atom effect on intersystem crossing processes in 2-thiouracil: A potential photodynamic therapy photosensitizer. *J. Chem. Phys.* **2013**, *138*, 044315. [CrossRef]

36. Ashwood, B.; Jockusch, S.; Crespo-Hernández, C.E. Excited-state dynamics of the thiopurine prodrug 6-thioguanine: Can N9-glycosylation affect its phototoxic activity? *Molecules* **2017**, *22*, 579. [CrossRef] [PubMed]

37. Martinez-Fernandez, L.; Faleson, T.; Norman, P.; Santoro, F.; Coriani, S.; Improta, R. Optical absorption and magnetic circular dichroism spectra of thiouracils: A quantum mechanical study in solution. *Photochem. Photobiol. Sci.* **2017**, *16*, 1415–1423. [CrossRef] [PubMed]

38. Pollum, M.; Ortiz-Rodriguez, L.A.; Jockusch, S.; Crespo-Hernández, C.E. The Triplet State of 6-thio-2’-deoxyguanosine: Intrinsic Properties and Reactivity toward Molecular Oxygen. *Photochem. Photobiol. Biol.* **2016**, *92*, 286–292. [CrossRef] [PubMed]

39. Woodrow, B.; Jockusch, S.; Crespo-Hernández, C.E. Excited-state dynamics of the thiopurine prodrug 6-thioguanine: Can N9-glycosylation affect its phototoxic activity? *Molecules* **2017**, *22*, 579. [CrossRef] [PubMed]

40. Farrell, K.M.; McMillan, M.M.; Pettitkow, M.; Selby, T. Heavy-Atom-Substituted Nucleobases in Photodynamic Applications: Substitution of Sulfur with Selenium in 6-Thioguanine Induces a Remarkable Increase in the Rate of Triplet Decay in 6-Selenoguanine. *Photochem. Photobiol. Sci.* **2016**, *15*, 286–292. [CrossRef] [PubMed]

41. Martínez-Fernández, L.; González, L.; Corral, I. An ab initio mechanism for efficient population of triplet states in cytotoxic sulfur substituted DNA bases: The case of 6-thioguanine. *Chem. Soc. Trans.* **2012**, *48*, 2134–2136. [CrossRef]

42. Santhosh, C.; Mishra, P.C. Electronic structures and spectra of 6-mercaptoguanine and 6-thioguanine. *Spectrochim. Acta Part A Mol. Specros. Sci.* **1993**, *49*, 985–993. [CrossRef]

43. Andersson, K.; Malmqvist, P.A.; Roos, B.O. Second-order perturbation theory with a complete active space self-consistent field reference function. *J. Chem. Phys.* **1992**, *97*, 1218–1226. [CrossRef]

44. Roos, B.O.; Andersson, K. Multiconfigurational perturbation theory with level shift—the Cr2 potential revisited. *Chem. Phys. Lett.* **1995**, *245*, 215–223. [CrossRef]

45. Finley, J.; Malmqvist, P.A.; Roos, B.O.; Serrano-Andrés, L. The multi-state CASPT2 method. *Chem. Phys. Lett.* **1998**, *288*, 299–306. [CrossRef]

46. Pou-Amérgio, R.; Merchán, M.; Nebot-Gil, I.; Widmark, P.O.; Roos, B.O. Density matrix averaged atomic natural orbital (ANO) basis sets for correlated molecular wave functions—III. First row transition metal atoms. *Theor. Chim. Acta* **1995**, *92*, 149–181. [CrossRef]

47. Fernández Galván, I.; Vacher, M.; Alavi, A.; Angeli, C.; Aquilante, F.; Autschbach, J.; Bao, J.J.; Bokarev, S.I.; Bogdanov, N.A.; Carlson, R.K.; et al. OpenMolcas: From Source Code to Insight. *J. Chem. Theory Comput.* **2019**, *15*, 5925–5964. [CrossRef]

48. Zou, X.; Zhao, H.; Yu, Y.; Su, H. Formation of guanine-6-sulfonate from 6-thioguanine and singlet oxygen: A combined theoretical and experimental study. *J. Am. Chem. Soc.* **2013**, *135*, 4509–4515. [CrossRef]

49. Bologa, A.; Bologa, C.; Cerullo, G.; Manzoni, C. Ultraviolet Transient Absorption Spectrometer with Sub-20-fs Time Resolution. *Appl. Sci.* **2018**, *8*, 989. [CrossRef]
55. Varillas, R.B.; Candeo, A.; Viola, D.; Garavelli, M.; De Silvestri, S.; Cerullo, G.; Manzoni, C.; De Silvestri, S.; Cerullo, G.; Manzoni, C.; et al. Microjoule-level, tunable sub-10 fs UV pulses by broadband sum-frequency generation. Opt. Lett. 2014, 39, 3849–3852. [CrossRef]

56. Snellenburg, J.J.; Laptenok, S.P.; Seger, R.; Mullen, K.M.; van Stokkum, I.H.M. Glotaran: A Java-Based Graphical User Interface for the R Package TIMP. J. Stat. Softw. 2012, 49, 1–22. [CrossRef]

57. van Stokkum, I.H.M.; Larsen, D.S.; Van Grondelle, R. Global and target analysis of time-resolved spectra. Biochim. Biophys. Acta—Bioenerg. 2004, 1657, 82–104. [CrossRef]