Phosphorylation Modulates Survivin Function in Behcet’s Disease

Yasamin Pahlavan1,2,3,4, Nasser Samadi1,3,5, Khalil Ansarin1,6, Alireza Khabbazi1,2,6

1Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran.
2Connective Tissue Diseases Research Center, Tabriz University of Medical Sciences, Tabriz, Iran.
3Department of Molecular Medicine, Faculty of Advanced Medical Sciences, Tabriz University of Medical Sciences, Tabriz, Iran.
4Students Research Committee, University of Tabriz Medical Sciences, Tabriz, Iran.
5Department of Biochemistry, Faculty of Medicine, Tabriz University of Medical Sciences, Tabriz, Iran.
6Rahat Breath and Sleep Research Center, Tabriz University of Medical Sciences, Tabriz, Iran.

Introduction

Behcet’s disease (BD) is a vasculitis characterized by recurrent attacks and frequent relapses of manifestations including oral aphthous ulcers, genital sores, skin lesions, arthritis, uveitis, gut and central nervous system involvement with various occurrence in patients.1 BD is more distributed in ancient Silk Road countries.2 The etiopathology of BD is not completely clear. It is reported that environmental factors including health status of oral and dental, also genital system, stress, smoking, and nutrition have direct correlation with incidence of BD. A line of evidence indicate the familial hereditary basis for BD patients.3,5 Deregulated activation of innate immune response including NK cells, γδ T cells and neutrophils are involved in pathogenesis of BD.1,6 Defective regulation of apoptosis process has been introduced in the pathogenesis of BD.7 Survivin encoded by baculoviral inhibition of apoptosis proteins (IAPs) repeat containing 5 (BIRC5) gene. This gene is located on chromosome 17q25 and have 4 exons and 3 introns, a single BIR domain without C-terminus ring finger domain.4 There are several spliced variants encoded by BIRC5 gene including Survivin wild type (142 amino acid), ΔEx3 (137 amino acid), 2B (165 amino acids), 3B (120 amino acid), 2α (74 amino acids), 3α (78 amino acids). Phosphorylation at Thr 34 is necessary for antiapoptotic activity of survivin protein.7 High BIRC5 gene expression considered in fetal development and pathologic conditions including tumors and autoreactive cell survival.8,9,10 Recently, survivin dysregulation is taken into consideration strongly in prognosis and disease activity of autoimmune diseases.10,12 Defective apoptosis

Abstract

**Purpose:** Survivin is critical for proliferation, maturation, homeostasis and differentiation of effector and memory lymphocytes. In this study the baculoviral inhibitors of apoptosis proteins (IAPs) repeat containing 5 (BIRC5) mRNA, survivin, and phosphorylated survivin expression were evaluated in peripheral blood mononuclear cells (PBMCs), and plasma of patients with Behcet’s disease (BD).

**Methods:** In this study, 26 Iranian Azari patients diagnosed with BD and 30 healthy controls were recruited. Total RNA was extracted from PBMCs. The expression level of survivin was measured by quantitative real-time polymerase chain reaction (PCR). Survivin plasma levels were measured using survivin Enzyme-linked immunosorbent assays. Also, western blotting analysis was performed to measure phosphorylated-survivin and survivin levels in PBMCs and plasma of patients with BD.

**Results:** In a pilot study, we showed that BIRC5 gene expression increased in BD patients compared with healthy controls ($P<0.05$). Western blotting analysis indicated that there was an increase in phosphorylated survivin expression in PBMCs of BD patients. Our data from western blot analysis showed survivin level in plasma samples of BD patients was similar to healthy controls. No significant differences were observed between plasma survivin levels in the BD patients compared with control group ($P>0.05$). The expression of phosphorylated survivin at Thr34 in PBMCs of BD patients with active disease was increased. Plasma phosphorylated survivin levels in having BD patients were also downregulated compared to healthy individuals.

**Conclusion:** Analysis of PBMCs indicated increasing expression level of phosphorylated survivin in PBMCs of BD patients. There was also a downregulation in phosphorylated survivin levels in plasma of BD patients.
is the irrevocable problem in autoimmune diseases. Survivin is an anti-apoptotic protein expressed in fetal development, tumor cells and immune cells. Survivin is able to inhibit caspase 3 function in apoptosis process by interfering during G2/M phase of mitosis in deregulated cell cycle checkpoints to promote abnormal cell survival. Studies indicated that posttranslational modifications on survivin like phosphorylation and acetylation have a key role in regulation of cellular functions for survivin. Survivin can phosphorylate in several sites including Thr 34, Thr 53, Thr 117, and Serin 20 and etc. These modifications provide various molecular functions for survivin. Survivin phosphorylation on Thr34 is regulated by CDK1/P34cdc2 cyclin B1 supported by human cervical adenocarcinoma, oral sub-mucosal fibrosis and oral squamous cell carcinoma studies. Unlike other IAP family members, survivin has only one BIR domain. Threonine 34 amino acid residue is located in this BIR domain. In fact, recent studies confirmed that phosphorylation of survivin on Thr34 increase cyto-protective effects on cancer cells.

Considering to unknown etiopathogenesis of BD, it is required to identification of specific diagnostic and prognostic markers for disease activity and therapy. Survivin regulates this process under influence of phosphorylation at Thr34 by p34 cdk2 kinase. In this paper, we aimed to evaluate the role of phosphorylated survivin parallel with survivin expression in peripheral blood mononuclear cells (PBMCs) and circulation of BD patients.

Materials and Methods

Study design and sample collection
In this study, Iranian Azari patients with diagnosis of BD (n = 26) including 16 patients with active disease and 10 patients with inactive BD and 30 age, sex and ethnically matched healthy controls were recruited in the study. Whole blood sample was collected from both patients and healthy individuals. BD diagnosis was based on International Criteria of Behcet’s Disease. Cases were called from the BD clinic of Connective Tissue Diseases Research Center at Imam Reza Hospital in Tabriz University of Medical Sciences between February 2018 and December 2018. Excluding criteria were having renal diseases, liver diseases, malignancy, human immunodeficiency virus infection, diabetes mellitus, other inflammatory, and autoimmune diseases and smoking. BD activity was measured using the Behcet’s Disease Current Activity Form (BDCAF), Total Inflammatory Activity Index (TIAI) and Iranian Behcet’s Disease Dynamic Activity Measure (IBDDAM). According to Indexes, patients with BDCAF ≥ 1 classified as active.

Sampling
Five milliliters of peripheral blood was obtained from the cubital vein of participants and directly transferred into sodium citrate tubes. PBMCs isolated by Ficoll (Lymphodex, Inno-Train, Germany) density-gradient centrifugation (Sigma) according to manufacture instruction. Then PBMCs were stored at -70°C until next step.

Quantitative real-time PCR
The expression level of survivin was measured by real-time polymerase chain reaction (PCR) (Roche Diagnostics GmbH, Sandhofer Strasse 116, 68305 Mannheim, Germany). Total RNA was extracted from PBMCs by high pure RNA isolation kit (BioFACT™ Total RNA Prep Kit, Korea) according to the manufacturer’s instructions. Amplification of complementary DNA (cDNA) was performed by reverse transcription using the reverse transcription reagent kit (BioFACT, Korea). Glyceraldehyde-3-phosphate dehydrogenase (GAPDH) used as a reference gene. The specific primers and PCR program are survivin: Forward: ATTTGTATCGCCCTCCTCCC, survivin: Reverse: TCCAGAGTTTTCGCGGAAG, GapDH: Forward: AATGGGCGACGTTTAGAAAA, GapDH: Reverse: GCCCAAATCGCAGAACAG. The ΔΔCt formula was used for calculating the relative expression levels of survivin.

Enzyme-linked immunosorbent assays (ELISA)
Survivin plasma levels was measured in plasma by Human survivin ZellBio GmbH ELISA kit [Cat. No: ZB-13904C-H9648, Germany] according to the manufacturer’s recommendations.

Western immunoblotting analysis
Cells were harvested, and total protein lysates were resolved. Western blotting analysis performed on all samples from PBMCs and plasma, by anti-phosphorylated-survivin (Thr 34), (anti-p-survivin (Thr 34), Cat. No: sc-16320, SANTA CRUZ) and survivin antibodies (anti-survivin, Cat. No: sc-17779, SANTA CRUZ), and anti- β-Actin (Anti-β-Actin, Cat. No, sc-130657, SANTA CRUZ). Proteins were first separated by electrophoresis in an acrylamide gel containing sodium dodecyl sulfate (SDS). After separation, protein bands were transfected on the membrane, then emerged with a little luminescence by CDK1/P34cdc2 cyclin B1 supported by human cervical adenocarcinoma, oral sub-mucosal fibrosis and oral squamous cell carcinoma studies. Unlike other IAP family members, survivin has only one BIR domain. Threonine 34 amino acid residue is located in this BIR domain. In this paper, we aimed to evaluate the role of phosphorylated survivin parallel with survivin expression in peripheral blood mononuclear cells (PBMCs) and circulation of BD patients.

Statistical analysis
Statistical analysis was performed using SPSS software version 16.0 (SPSS, Chicago, IL, USA). The normal distribution of the data was tested with the Kolmogorov–Smirnov test with Lilliefors correction. The quantitative data were presented as mean ± standard deviation (SD). Differences in the expression levels of survivin in PBMCs and plasma between the groups were analyzed by the
Mann Whitney U-test. P-value < 0.05 was considered as statistically significant.

Results and Discussion

**BIRC5 gene is overexpressed in PBMCs in BD**

Up-regulation of BIRC5 in autoimmune diseases is reported in recent studies. High BIRC5 level is found in PBMCs of patients with rheumatoid arthritis (RA),12 myasthenia gravis21 and multiple sclerosis.22 We first analyzed BIRC5 gene expression levels in PBMCs by quantitative PCR analysis. Our data revealed that mRNA expression levels of survivin was significantly higher in BD patients compared to healthy controls (P<0.05, Figure 1).

Because the pattern of mRNA expression varies with protein content and researchers found that RNA may have function without translation to the protein,23 we also measured expression of survivin and phosphorylated survivin on Thr34 (p-survivin) in PBMCs by western blot analysis. Therefore, although, analyses showed similar expression patterns of survivin in PBMCs of BD patients and healthy controls (Figure 2A, Figure 2E. lower panel, Fold of control: 1.36), the expression of phosphorylated survivin at Thr34 was increased in PBMCs of Behcet's patients. However, these differences were insignificant in the point of statistic (P>0.05, Figure2B, Figure 2E. upper panel, Fold of control: 2.20).

We next classified patients into two groups: patients with active disease and patients with inactive disease. Then, statistical analysis was performed. Our data showed similar expression patterns of survivin in PBMCs between BD patients with active disease and inactive disease (Figure 2C, Figure 2E. lower panel, Fold of control: 1.20), the expression of phosphorylated survivin at Thr34 in PBMCs was tending to increase in Behcet's patients with active disease (Figure 2D, Figure 2E. upper panel, Fold of control: 2.04). Although, this difference did not reach a statistically significant level (P>0.05).

**Free circulating survivin protein and survivin phosphorylation on Thr34 in BD.**

Previous studies investigated the association between free circulating survivin levels and its impact on disease activity and prognosis in autoimmune diseases and cancers. There is evidence about presence of survivin in circulation. High serum survivin level is reported in other autoimmune diseases including RA,4 systemic sclerosis,4 and inflammatory arthritis which almost has similar clinical manifestation with BD.7,24 Therefore, in this study, we measured survivin and phosphorylated survivin on Thr34 in the plasma samples of BD patients and healthy individuals by western immunoblotting analysis. We next investigated the free circulating survivin in BD patients using enzyme linked immunosorbent assay. No significant differences were observed between plasma survivin levels in the BD patients compared with control group (P>0.05, Figure 3A). This level between BD patients with active and inactive forms of disease was also
Previous studies performed on autoimmunity diseases and cancers investigated the relationship between serum and plasma survivin level and prognosis. Also, our data from western blot analysis showed that the survivin level in plasma samples of Behcet’s patients was similar to healthy controls, (Figure 4A, Figure 4C, lower panel, Fold of control: 1.05). The phosphorylated survivin level was lower in patients with BD. However, these differences were insignificant in the point of statistic (P > 0.05, Figure 4B, Figure 4C, upper panel, Fold of control: 0.64). There was considerable down regulation in p-survivin level in BD patients compared with healthy individuals.

In this study, we had shown that survivin levels in PBMCs of Behcet’s patients critically modulated by phosphorylated survivin on Thr34. Other studies indicated that phosphorylation and acetylation, as post-translational modifications, have a key role in regulating different cellular and molecular function for survivin. As regards, survivin overexpression in PBMCs reported in RA, systemic sclerosis, myasthenia gravis.10 We also investigated levels of survivin and phosphorylated survivin on Thr34 in plasma samples of Behcet’s patients. The phosphorylated survivin was increased in PBMCs of Behcet’s patients which tended to increase in BD patients with active disease. Also, our data showed that phosphorylated survivin having a downward trend in the circulation of BD patients. Recent studies have identified the presence of survivin in normal and pathologic adult tissues including polymorphonuclear cells,25 T cells26 and serum of patients with cancer.27 These findings suggested that survivin could have different distribution inside and outside the cell which could have undesirable consequences.28 Survivin could be inhibited by ubiquitin proteosomal degradation, the X-linked inhibitor of apoptosis (XIAP)-XAF1 complex29 and some of medications,17 suggesting decreased level of phosphorylated form of survivin in plasma of this patients.

The expression of phosphorylated survivin at Thr34 was higher in PBMCs of BD patients with active disease compared with patients with inactive disease. Phosphosurvivin which is mainly localized in the nucleus, critically regulates survivin levels inside of malignant cells, suggesting why phosphorylated survivin showed higher level inside the cell and lower level in plasma, respectively. According to gene expression and proteomic data, researchers estimated the possible kinetic parameters about synthesis, stability and degradation of RNA and proteins beside transcriptional and post-transcriptional regulation. Also, depend on stage of disease progression, survivin expression has different distribution. In active status of the disease the expression of survivin may increase, as well as advanced stage of disease, comparable with silenced stage. Regulation of protein concentration is affected by miss folding stress which may show distinct expression patterns inside and outside of the cells.

Autoimmune diseases arise due to cellular and molecular dysregulation of immune system beside genetics and epigenetics dysregulation.30-31 The immunopathology and molecular bases of BD were not identified well. Genetic and environmental factors, lymphocytes activation, autoimmunity, and auto-inflammatory process and cell death and survival imbalance involved in BD initiation and progression.32,33 For instance, activation of regulatory T cells and decrease of NK cells may be observed in peripheral blood cells in Behcet’s patients which reflects homing of cytotoxic cells toward inflammation through cytokines belong to Th2 subset in Behcet’s patients that results in cytotoxicity during active phase of the disease.2,10 Studies reported that activation and proliferation of γδ T cells, which acts against various microbial infections, provide the establishment of BD.3 They are several evidence of inconsistency in apoptosis of immune cells in the pathogenesis of BD.17,34 Levels of apoptosis regulating genes such as Fas/APO1 and Bcl-2 is increased
in inflammatory sites in BD.\textsuperscript{16,17,35} Apoptosis associated regulator molecules including Fas antigen (CD95) is identified on lymphocytes in PBMCs of Behcet’s patients.\textsuperscript{7}

Extracellular survivin may play a significant role in inflammatory arthritis as previously reported in human leukocytes in autoimmune arthritis context.\textsuperscript{11} In this context, survivin regulates T cell stimulation and skew immune reactions toward Th2 responses,\textsuperscript{36,37} similar to observed phenotype in Behcet’s patients. Survivin is the inhibitor of the apoptosis process which may prevent caspase-dependent pathway in this process. The mechanism of action for survivin is binding and suppressing the caspase 9 in intrinsic pathway of apoptosis followed by inhibition of caspase 3 and 7, which are executive caspses.\textsuperscript{34}

Various functions of survivin are originated from different cellular localization, phosphorylation, and acetylation. Protein phosphorylation is involved in cytoprotection, transition from cell cycle,\textsuperscript{3} subcellular localization.\textsuperscript{31} Phosphorylated survivin has a critical role in controlling the cellular stability and expression of apoptosis regulators.\textsuperscript{5,35} Interestingly, evidence showed that survivin has several promitotic and antiapoptotic functions. Some studies mentioned that phosphorylation of survivin on Thr34 by mitotic kinase p34 cdk cyclin B1, is the key step for activation of survivin.\textsuperscript{4,37} Phosphorylated survivin on Thr34 bind to caspase 9 in Hela cells.\textsuperscript{30} Failure of survivin phosphorylation impairs complex formation with caspase 9. Phosphorylation on serine 20 reduces the antiapoptotic role of survivin. Phosphorylation on Thr34 stimulates binding to caspase 9 and regulates the intrinsic pathway of apoptosis. Phosphorylation of survivin on Thr34 by CDK1/P34\textsuperscript{40,42} cyclin B1 was found in human cancer cells.\textsuperscript{37}

In this study for a first time, the plasma level of survivin measured in BD patients. Although survivin is found and act in intracellular locations such as mitochondria, cytoplasm, and nucleus of cells, there is some evidence of presence of this molecule in extracellular space and circulation under pathologic conditions. Several studies considered survivin as a valuable serologic marker in RA which can predict the development and response to treatment in early RA and improve RA risk estimation even development of arthralgia in RA patients.\textsuperscript{7,24}

Generally, most of the functions of survivin in regulation of inflammation is not well understood. Although, further studies required for understanding the role of survivin in BD, our findings may open a new perspective in BD pathogenesis from the apoptosis and inflammation sight and crosstalk between BD and survival of autoreactive cells. Further studies should be performed on the function of phosphorylated survivin in cell organelles and circulation in autoimmune diseases.

**Ethical Issues**
The research protocol was approved by the Ethics Committee of Tabriz University of Medical Sciences (IR TBZMED. REC.1397.190) and the study was conducted according to the Declaration of Helsinki (2008). Written informed consents were taken from all study participants.

**Conflict of Interest**
Authors declare no conflict of interest in this study.

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