AMELIORATIVE ROLE OF SILYMARIN AGAINST ASPARTAME–INDUCED BIOCHEMICAL CHANGES, OXIDATIVE STRESS, INFLAMMATORY EFFECT AND GENOTOXICITY IN MALE ALBINO RATS

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ABSTRACT

Objective: Aspartame (ASP) is one of the most common artificial sweeteners. It has been recorded to be safe by World Health Organization. However, numerous publications have concluded that ASP is a genotoxic and carcinogenic sweetener.

Methods: The current study aims to examine the effect of ASP consumption (250 mg/kg body weight/day for 90 d) on some biochemical parameters, oxidative/antioxidative status in different tissues, Tumor Necrosis Factor-α (TNF-α), chromosomal aberration (CA) frequency and mitotic index (MI%) in addition to the possible ameliorative role of silymarin (50 mg/kg body weight/day for 90 d) against ASP-induced toxicity in male albino rats.

Results: The present results have confirmed that ASP is able to induce significant increase in the blood glucose level, liver, kidney and lipid function tests, Malondialdehyde (MDA) level, serum TNF-α level, frequency of CA and MI%. Meanwhile, Gluthathione reduced level (GSH), Gluthathione-S-transferase (GST) and catalase activity (CAT) were decreased by ASP administration. Recovery group showed slight enhancement in all parameters but remained significant as compared to the control group. Co-administration of ASP with silymarin showed greater improvement than the recovery group.

Conclusion: Silymarin have an ameliorative role against biochemical oxidative stress, inflammatory changes in blood and different tissues, chromosomal aberrations and MI% induced by ASP administration.

Keywords: Aspartame, Silymarin, Inflammatory marker, Oxidative/anti-oxidative status, Chromosomal aberrations

INTRODUCTION

Aspartame (L- aspartyl-L-phenylalanine methyl ester) is considered as non-nutritive and non-caloric artificial sweetener consumed by more than two hundred million people worldwide. It was used in nearly 6 thousands nourishment items, including dry refreshment, breakfast grains, biting gum, refrigerated and nonrefrigerated drinks, yogurt items and pharmaceuticals [1].

ASP is quickly metabolized into 3 segments, 50% phenylalanine, 10% methanol and 40% aspartate [2]. At the point when aspartame is absorbed by the body, aspartic acid is changed over into alanine and oxaloacetate [3]. Phenylalanine is changed into tyrosine, phenylethylamine and phenylpyruvate to a lesser degree [4]. Methanol is changed over into formaldehyde and afterward changed over to formate. These procedures are joined by the development of superoxide anion and hydrogen peroxide [5]. Formic acid is the real metabolite known for its destructive impacts of acute toxicity in humans and animals [6].

ASP is a multipotential cancer-causing operator and builds the danger of having various sorts of tumors [7]. Likewise, there was a relationship between aspartame dosing and the danger of having type 2 diabetes [8], preterm delivery [9], nephrotoxicity [10], liver toxicity [11] and histopathological effects in the parotid salivary glands [12]. ASP caused genotoxicity; it significantly induced DNA fragmentation in bone marrow cells and liver of albino rat’s mothers and their offspring compared with the control animals [13].

Silymarin is a polyphenolic compound obtained from the seeds and fruits of Silybum marianum. This plant is an ancient medicinal plant used for liver, gallbladder disorders and hepatoprotection management [14]. Silymarin, the unrefined business result of milkthistle[15], is a complex of 7 flavonolignans (the most significant being silybin, isosilybin, silydianin, and silychristin) and one flavonoid (taxifolin); half of 80% of the extract concentrate. Silymarin is utilized as a correlative prescription elective throughout the world [16]. It fills in as a cancer prevention agent by searching for free-radicals and expanding glutathione concentration [17]. Thus, the present study aimed to evaluate the toxicity of ASP in addition to the possible ameliorative role of silymarin using CA assay and measurements of MIP, blood glucose level, liver, kidney, cardiac function tests, MDA level and serum TNF-α level, GSH level, GST and catalase activity in male albino rats.

MATERIALS AND METHODS

Chemicals

Aspartame (ASP) was acquired from Al-Ameryapharma company, Egypt, as tablets; every tablet contains 20 mg of aspartame. Silymarin was purchased from the pharmacy (Legalon® 140 mg, each capsule contains 140 mg Silymarin). Doses of aspartame or silymarin were determined by dissolving estimated amounts in refined water, and they were managed at a volume of 10 ml/kg.

Experimental design

The present work was done using 50 adult male albino rats (190–200 gm weight). The animals were kept up under standard research facility conditions with water and nourishment. The investigation was approved as recommended in the guidelines given by the Ethical Review Committee of Zagazig University, Egypt. The rats were divided into five equal groups. Group 1 (Control): received ordinary diet and water. Group 2 (Silymarin): animals were orally administrated with a dose of silymarin (50 mg/kg/day) for 90 d. This dose was based on experiments assessing that LD50 was 385 mg/kg in rats [18]. Group 3 (Aspartame): received ASP by gastric tube (250 mg/kg/d) [19] for 90 d. This dose was compared to the satisfactory every day admission in people (defined by the WHO) of
40–50 mg/kg/d. Species correction required a five to six times higher dose in rats than in humans, as rats metabolize aspartame faster than humans [20]. Group 4 (Recovery Group) administrated aspartame as in Group 3, but animals were left for two weeks after stopping ASP to recover before they were killed. Group 5 (Aspartame+silymarin) rats co-administered silymarin with ASP daily for 90 d.

Collection and sampling of blood
At the end of the experimental period, 5 rats from each group were used for the chromosomal preparation and MI measurement from their bone marrow. The remaining 5 animals in each group had fasted for 12 hand were presented to mild anesthesia. Blood was gathered from their orbital sinus and blood serum was set up by accumulation of blood in an anticoagulant–free tube for separation of serum which was moved into another tube and kept frozen at-20 °C [21].

Tissue processing for oxidative parameters and histopathological examination
After blood collection, liver, kidney and heart tissues were removed, sectioned, and either homogenized with ice-cold phosphate-buffered saline (pH 7.4) or processed for general histology. The homogenates were centrifuged and the resulting supernatant was moved into another tube and protected until utilized. The other parts of tissues were fixed in 10% buffered formalin for histopathological study [22].

Biochemical measurements

- **Blood glucose, kidney, liver and lipid functions in blood serum**
  Fasting blood glucose was analyzed using colorimetric method [23], Urea [24] and creatinine concentration [25]. The biochemical parameters evaluated were liver biomarkers such as, bilirubin [26], ALT, AST [27] and ALP [28] using diagnostics kits. There were serum lipid profiles (Triglyceride and Total cholesterol) [29, 30]

- **Oxidative stress marker measurements**
  MDA [31], Glutathione reduced (GSH) [32], Glutathione–S-transferase (GST) [33] and Catalase (CAT) [34] were determined according to kits purchased from Biodiagnostic Company (Biodiagnostic, Egypt).

- **Inflammatory markers**
  Rat Tumor Necrosis Factor-α (TNF-α) was determined by ELISA according to Catalogue No. 201-11-0765 kits

Chromosomal aberration assay
Chromosomes were extracted from white blood cells in the bone marrow of rats from each group according to the method described by [35]. By Giemsa stain (1.5%), the slides were stained for about 20 min. The slides were carefully scanned for the metaphase spreads under the ×10 objective and then under ×40 objective of the light microscope. The well-spread metaphases were examined under ×100 (oil immersion) and the photomicrographs of the well-spread chromosomes were taken. A total of 50 well-spread metaphases from each rat were analyzed.

- **MI determination**
  The MI was examined for each rat in all groups according to the following formula: MI= (Number of dividing cells/Total examined cells) ×100

Statistical analysis
All outcomes were analyzed by SPSS programming (variant 14). Data were expressed as mean±SE. A comparison of mean values of factors among various groups was done using the ANOVA test. P<0.05 was considered to be significant [36].

RESULTS

Effect of aspartame administration on body weight of all studied groups
Data showed that there was a significant increase in the final body weight in group 3 (aspartame) (P< 0.001) and Group 4 (Recovery Group) (P< 0.05) which amounted to 27.1 % and 13.8 % respectively. In group 5 (Aspartame+silymarin) there was a statistically non-significant increase in the final body weight, which amounted to 7 % (table 1) and (fig. 1).

| Groups                  | Variable                  | Initial body weight (g) (mean±SEM) % change | Final weight (g) (mean±SEM) % change |
|-------------------------|---------------------------|--------------------------------------------|-------------------------------------|
| Group 1 (Control)       | 150.5±3.06                | 342.25±15.75                               |
| Group 2 (Silymarin)     | 146.4±2.37                | 350.8±7.94                                 |
| Group 3 (Aspartame)     | 154.8±1.52                | 435±8.94 ***                               |
| Group 4 (Recovery Group)| 148.6±3.67                | 389.4±7.56                                 |
| Group 5 (Aspartame+silymarin) | 146.6±3.25 | 366.40±10.27                               |

P > 0.05

Significant differences are indicated (One-way ANOVA followed by post hoc Duncan’s multiple range test). *P<0.05, **P<0.01, ***P<0.001 significantly different from control.

**Fig. 1: Initial body weight and final body weight of all studied groups**
Effect of aspartame administration on glucose, liver, kidney and lipid function tests

Data presented in (table 2) and (fig. 2, fig. 3) showed that the mean level of glucose was increased in groups 3, 4 and 5. This increase was statistically significant in group 3 (P<0.01) and group 4 (P<0.05). In group 5 silymarin administration decreased glucose level which was statistically non-significant compared to the control group (P>0.05).

Also, data showed a significant (P<0.05) increase in ALT activity in group 3. This value was decreased in group 4 and 5 (P>0.05). The mean value of AST activity increased in group 3, 4 and 5 but this increase was statistically non-significant (P>0.05) compared to control group. ASP administration caused slight decrease in the mean level of albumin (P>0.05) and a slight increase in total bilirubin (P>0.05).

Data confirmed that ASP administration caused a slight increase in urea concentration (P>0.05). ASP caused a significant (P<0.01) decrease in group 4. But this was still statistically significant. Group 5 had marked decrease which was statistically non-significant (P>0.05) as compared to the control group.

The present result declared that the dosing of ASP caused a significant increase in the mean level of Cholesterol (P<0.01) and triglyceride in group 3 (P<0.001). These values showed a slight decrease in group 4. But this was still statistically significant. Group 5 had marked decrease which was statistically non-significant (P>0.05) as compared to the control group.

Effect of aspartame administration on oxidative and antioxidative parameters in liver tissue

Data showed that the dosing of ASP caused a marked significant increase on hepatic MDA level (P<0.001) and a significant decrease on GSH levels (P<0.05), GST and catalase activities (P<0.001) as compared to the control group. The elevation on MDA level showed a slight decrease in group 4 (P<0.01) and a marked decrease in group 5 but it remained significant (P<0.05) at the statistical level. Also, the decrease on hepatic GSH level, GST and catalase activities were enhanced in group 4 but, it was statistically significant as group 3 and showed a marked elevation in group 5 which was statistically non-significant (P>0.05) as compared to the control group (table 3).

Table 2: Effect of aspartame administration on glucose, liver, kidney and lipid function tests

| Parameters          | Group 1 (Control) | Group 2 (Silymarin) | Group 3 (Aspartame) | Group 4 (Recovery Group) | Group 5 (Aspartame + silymarin) |
|---------------------|-------------------|---------------------|---------------------|--------------------------|---------------------------------|
| Glucose (mg/dl)     | 115.6±4.9         | 120.75±3.9         | 151±10.1            | 141±5.5                  | 125.7±4.14                     |
| ALT (U/l)           | 41.7±1.1          | 40.36±0.73         | 46.17±2.09          | 43.08±0.46               | 40.92±0.96                     |
| AST (U/l)           | 198.7±6.93        | 210.8±9.16         | 228.77±4.14         | 223±10.7                 | 209.8±5.65                     |
| ALB (g/dl)          | 3.22±0.07         | 3.14±0.06          | 3.17±0.06           | 3.20±1.16                | 3.14±0.09                      |
| T. bilirubin (mg/dl)| 0.43±0.06         | 0.45±0.02          | 0.52±0.02           | 0.50±0.03                | 0.41±0.02                      |
| Urea (mg/dl)        | 29.5±1.84         | 29±2.2             | 32±0.7              | 31.2±1.56                | 30.1±1.7                       |
| Creatinine (mg/dl)  | 0.56±0.03         | 0.57±0.02          | 0.80±0.04"         | 0.71±0.03"               | 0.58±0.04"                     |
| Cholesterol (mg/dl) | 111.5±2.0         | 109.8±1.8          | 127.2±2.4"         | 125.4±2.7"               | 117.7±2.3                      |
| Triglyceride (mg/dl)| 88.7±6.7          | 86.0±2.2           | 154±5.2"            | 135.2±5.08"              | 98.1±6.4                       |

Values are expressed as the means±SEM. Significant differences are indicated (One-way ANOVA followed by post hoc Duncan’s multiple range test). *P<0.05, **P<0.01, ***P<0.001 significantly different from control.

Fig. 2: Effect of aspartame administration on glucose, ALT, AST, urea, cholesterol and triglyceride

![Fig. 2: Effect of aspartame administration on glucose, ALT, AST, urea, cholesterol and triglyceride](image1)

Fig. 3: Effect of aspartame administration on albumin, total bilirubin and creatinine

![Fig. 3: Effect of aspartame administration on albumin, total bilirubin and creatinine](image2)
The elevation in MDA level showed slight decrease in elevation in kidney MDA level (P<0.01) and a significant decrease in GSH activities (P <0.001) as compared to control values. The elevation in MDA level showed a highly significant elevation in cardiac MDA level (P<0.001) and a marked decrease in group 5 but, it was still significant (P<0.05) at the statistical level. The decreased kidney GSH level, GST and catalase activities were enhanced in group 4, but the values were remained statistically significant as group 3 and showed marked elevation in group 5 which was statistically non-significant (P > 0.05) in case of GSH and catalase and significant (P<0.05) in MDA and GST as compared to control group (Table 4).

Table 3: Effect of aspartame administration on oxidative and anti-oxidative parameters in liver tissue

| Groups           | Liver tissue | MDA (nmol/g) | GST (U/g) | GSH (mmol/g) | CAT (U/g) |
|------------------|--------------|--------------|-----------|--------------|-----------|
| Group 1 (Control)|              | 31.4±0.79    | 427.7±29.8| 2.26±0.22    | 193.4±2.5 |
| Group 2 (Silymarin) |             | 36.4±2.3    | 412.5±43.3| 2.20±0.18    | 197.8±0.8 |
| Group 3 (Aspartame) |            | 156.4%      | -3.5%     | -2.7%        | 2.3%      |
| Group 4 (Recovery Group) |         | 84.4±1.4*** | 144.4±1.33*** | 0.80±0.06* | 183.1±0.94*** |
| Group 5 (Aspartame+silymarin) |       | 65.9±6.6**  | 157.8±0.8*** | 1.02±0.17* | 187.3±1.4**  |
|                  | 107.2%      | -63.1%      | -5.4%     | -3.2%        |
|                  | 40.0±1.9*   | 400.9±20.2  | 1.28±0.24 | 192.9±6.8   |
|                  | 26.1%       | -0.2%       | -4.3%     | -0.3%        |
| P < 0.001        | P < 0.001   | P < 0.001   | P < 0.001 | P < 0.001   |

Values are expressed as the means±SEM, % change. Significant differences are indicated (One-way ANOVA followed by post hoc Duncan’s multiple range tests) *P<0.05, **P<0.01, ***P<0.001 significantly different from control.

Table 4: Effect of aspartame administration on oxidative and anti-oxidative parameters in heart tissue

| Groups              | Kidney tissue | MDA (nmol/g) | GST (U/g) | GSH (mmol/g) | CAT (U/g) |
|---------------------|---------------|--------------|-----------|--------------|-----------|
| Group 1 (Control)   |               | 50.1±3.5     | 369.0±9.4 | 3.12±0.08    | 197.7±0.8 |
| Group 2 (Silymarin) |               | 53.5±0.97    | 370.4±2.0 | 3.35±0.12    | 198.5±0.4 |
| Group 3 (Aspartame) |               | 107.3±4.1**  | 305.5±1.4*** | 1.95±0.27** | 191.9±1.0*** |
| Group 4 (Recovery Group) |           | 84.6%        | -17.2%    | -37.5%      | -2.9%     |
| Group 5 (Aspartame+silymarin) |       | 93.6±3.1**  | 338.6±6.4** | 2.03±0.36** | 192.6±2.6*** |
|                  | 61.1%        | -8.3%        | -34.9%    | -2.6%        |
|                  | 78.2±1.08*   | 341.3±7.6*  | 2.61±0.04 | 197.2±0.71  |
|                  | 34.6%        | -7.5%        | -16.3%    | -0.3%        |
| P < 0.001         | P < 0.001    | P < 0.001    | P < 0.001 | P < 0.001  |

Values are expressed as the means±SEM, % change. Significant differences are indicated (One-way ANOVA followed by post hoc Duncan’s multiple range tests) *P<0.05, **P<0.01, ***P<0.001 significantly different from control.

Table 5: Effect of aspartame administration on oxidative and anti-oxidative parameters in heart tissue

| Groups              | Heart tissue | MDA (nmol/g) | GST (U/g) | GSH (mmol/g) | CAT (U/g) |
|---------------------|--------------|--------------|-----------|--------------|-----------|
| Group 1 (Control)   |              | 20.1±0.26    | 198.4±14.4| 2.6±0.05     | 188.8±5.3 |
| Group 2 (Silymarin) |              | 18.3±1.79    | 208.5±10.7| 2.8±0.03     | 190.7±4.4 |
| Group 3 (Aspartame) |              | 28.5±10.19***| 128.3±9.3*** | 2.2±0.15** | 154.3±2.4*** |
| Group 4 (Recovery Group) |          | 41.9%        | -35.3%    | -16.5%      | -18.2%    |
| Group 5 (Aspartame+silymarin) |        | 27.3±11.0*   | 162.1±4.7* | 2.3±0.05*  | 162.9±2.1** |
|                  | 35.8%        | -18.2%       | -10.2%    | -13.7%      |
|                  | 21.1±0.35    | 184.3±9.3    | 2.6±0.02  | 181.1±5.2   |
|                  | 5%           | -7.1%        | -1.1%     | -4.1%       |
| P < 0.001         | P < 0.001    | P < 0.001    | P < 0.001 | P < 0.001  |

Values are expressed as the means±SEM, % change. Significant differences are indicated (One-way ANOVA followed by post hoc Duncan’s multiple range test). *P<0.05, **P<0.01, ***P<0.001 significantly different from control.

Results confirmed that ASP administration caused a highly significant elevation in kidney MDA level (P<0.01) and a significant decrease in GSH levels (P<0.01) GST and catalase activities (P<0.001) as compared to the control group. The elevation in MDA level showed slight decrease in group 4 (P<0.01) and a marked decrease in group 5 but, it was still significant (P<0.05) at the statistical level. The decreased kidney GSH level, GST and catalase activities were enhanced in group 4, but the activities were remained statistically significant as group 3 and showed marked elevation in group 5 which was statistically non-significant (P > 0.05) in case of GSH and catalase and significant (P<0.05) in MDA and GST as compared to control group (Table 4).

Effect of aspartame administration on oxidative and antioxidative parameters in kidney tissue

Effect of aspartame administration on oxidative and antioxidative parameters in heart tissue

The present study recorded that ASP administration caused a highly significant elevation in cardiac MDA level (P<0.01) and a significant decrease in GSH levels (P<0.01), GST and catalase activities (P<0.001) as compared to control values. The elevation in MDA level showed a slight decrease in group 4 (P<0.05) and marked decrease in group 5 which was statistically non-significant (P > 0.05). The decreased cardiac GSH level, GST and catalase activities were enhanced in group 4 but still statistically significant as group 3 and showed a marked elevation in group 5 which was statistically non-significant (P 0.05) as compared to control group (Table 5). Data showed that ASP has an effect on the different tissues. Recovery period (14 d) was not enough to restore the oxidative and anti-oxidative parameters to its normal levels. Also, co-administration of ASP with silymarin showed great improvement than the recovery group (fig. 4, fig. 5, fig. 6, fig. 7).

Table 5: Effect of aspartame administration on oxidative and anti-oxidative parameters in heart tissue
Fig. 4: Effect of aspartame administration on percent change of MDA in aspartame, recovery and aspartame+silymarin groups in compared to the control group in different tissues.

Fig. 5: Effect of aspartame administration on percent change of GST in aspartame, recovery and aspartame+silymarin groups in compared to the control group in different tissues.

Fig. 6: Effect of aspartame administration on percent change of GSH in aspartame, recovery and aspartame+silymarin groups in compared to the control group in different tissues.

Fig. 7: Effect of aspartame administration on percent change of CAT in aspartame, recovery and aspartame+silymarin groups in compared to the control group in different tissues.
Table 6: Effect of aspartame administration on serum levels of TNF-α in all studied groups

| Groups                      | TNF-α (ng/l) | P value |
|-----------------------------|--------------|---------|
| Group I (Control)           | 96.8±2.3     |         |
| Group 2 (Silymarin)         | 96.6±1.1     | 0.21 %  |
| Group 3 (Aspartame)         | 129.6±2.8    | 0.01 %  |
| Group 4 (Recovery Group)    | 113.8±4.4    | 0.17 %  |
| Group 5 (Aspartame+silymarin)| 102.7±2.8    | 0.01 %  |

Values are expressed as the means±SEM, % change. Significant differences are indicated (One-way ANOVA followed by post hoc Duncan’s multiple range test). *P<0.05, **P<0.01, ***P<0.001 significantly different from control.

Effect of aspartame administration on serum levels of TNF-α in all studied groups

Results indicated that there was a highly significant increase in the mean level of serum TNF-α in group 3 (P<0.001) as compared to the control group. Group 4 showed a significant increase (P<0.05) but less than that of group 3. In group 5, there was a statistically non-significant increase (table 6) (fig. 8).

The effect on the chromosomal aberration frequency

Under the conditions of the present investigation, the control rats recorded chromosomal aberration (CA) frequency within the normal values while oral administration of ASP to male rats induced significant (p<0.001) increase in the percentage of total chromosomal aberration (TCA) reaching 63.00± 2.89 as compared with 15.60± 0.67 in control group. TCA included the total numerical aberration (TNA) and the total structural aberration TSA (table 7) (fig. 9).

The numerical aberrations were mainly the aneuploidy (hyperdiploid) and polyplody (table 7). (fig. 9, fig. 10). Meanwhile, the deletions, dicentric chromosomes, exchange fig. and breaks were the most abundant structural aberrations (table 7). (fig. 9, fig. 11); all of these aberrations significantly (p<0.001) increased as compared with control values.

In the present study, when rats were orally administered with ASP then left for two weeks post-treatment to recover (recovery group), the frequency of TCA (TNA and TSA) was clearly reduced as compared with ASP group. Hyperdiploid and polyplody were decreased but their values remained significant (p<0.01) as compared with control (table 7), (fig. 9). Except for exchange figures, all types of structural aberrations decreased and became non-significant at the statistical level. Thus, better CA improvement after stoppage of ASP may require an extension of the recovery period to more than two weeks.

According to the result of the present study, silymarin treated rats showed a significant (p<0.05) decrease in the frequency of TNA as compared with the control value. Herein, aneuploidy (hyperdiploid) was significantly (p<0.05) decreased and all other types of CA were non-significantly changed. This confirmed that silymarin itself does not induce CA and is able to repair the chromosomal damage (table 7), (fig. 9).

Moreover, the present data revealed that silymarin drug as an antioxidant alleviated the genotoxicity of ASP by clear reduction in the frequency of TCA (TNA and TSA) in bone marrow cells of rats co-administered both ASP plus silymarin. All types of CA (hyperdiploid, polydiploid, deletions, dicentric chromosomes and breaks) were highly reduced and became non-significant as compared with control values. However, the frequency of TCA (TNA and TSA mainly exchange figures) remained significant as compared with control values (table 7), (fig. 9). The ameliorative role of silymarin against ASP toxicity might be a result of its anti-oxidative activity; it acts as a scavenging agent against the free-radicals induced by ASP that were the main cause of ASP genotoxicity repairing the chromosomal damage in bone marrow cells of ASP treated rats.

The effect on the percentage of the mitotic index

According to the present work, the percentage of MI was increased in ASP group as compared with a control value (table 7), (fig. 12). Increased MI rate might result from a high progression rate of cells from the stage of DNA synthesis (S-phase) to mitosis (M-phase) of the cell cycle. This indicates the potential of ASP to induce more cells to divide and hence inducing cancer cells.

The present data recorded that the value of MI was relatively decreased in the recovery group as compared with the aspartame group (table 7), (fig. 12). Rats had been left for 14 d post-treatment with ASP to recover and this period was not enough to reach a complete recovery.

In addition, data showed that there was no significant change in MI% in silymarin treated rats. The animals administered with both ASP plus silymarin recorded an improvement in MI% as compared with rats treated with ASP alone, but the value didn’t reach the control level (table 7), (fig. 12).
Table 7: The frequencies of chromosomal aberrations (numerical and structural) and MI% in bone marrow cells of male albino rats after ASP and/or silymarin administration

| Groups                  | No. of rats | No. of scored cells 50/rat | Chromosomal aberrations | Mitotic index | TCA % | Metaphases /1000 cell |
|-------------------------|-------------|----------------------------|-------------------------|---------------|-------|-----------------------|
|                         |             |                            | Numerical aberrations   |               |       |                       |
|                         |             |                            | Aneuploidy               | Polyplody     | TNA   | Deletion              | Dicentric | Exchange | Break | TSA   |
| Control                 | 5           | 250                        | 9.00±0.00               | 1.00±0.1      | 12.2±0.1 | 0.40±0.1              | 1.20±0.0  | 0.40±0.0  | 0.20±0.0 | 3.40±0.0 | 15.60±1.0 | 1.10 |
| Silymarin               | 5           | 250                        | 4.60±1.4               | 2.40±0.7      | 9.2±0.6    | 1.00±0.1              | 0.20±0.0  | 1.40±0.0  | 0.60±0.0 | 4.00±0.0 | 13.20±1.0 | 1.12 |
| Aspartame               | 5           | 250                        | 14.00±1.4             | 14.00±1.4     | 35.40±1.4 | 5.40±0.6              | 6.00±0.6  | 3.00±0.0  | 27.60±1.4 | 6.00±0.4 | 2.89***   | 5.50 |
| Aspartame recovery      | 5           | 250                        | 13.20±1.4             | 4.20±1.4      | 18.40±1.4 | 0.20±0.0              | 1.20±0.2  | 0.40±0.0  | 0.40±0.0 | 6.80±0.0 | 25.20±1.4 | 4.44 |
| Aspartame+silymarin     | 5           | 250                        | 11.00±0.7             | 3.00±0.7      | 16.20±0.7 | 1.20±0.2              | 0.40±0.0  | 0.60±0.0  | 4.80±0.0 | 24.60±1.4 | 4.39***   | 3.09 |

Values are expressed as the mean±SEM. Significant differences are indicated *P<0.05, **P<0.01, ***P<0.001 significantly different from control.

Fig. 9: Histogram of mean values of different types of chromosomal aberrations in bone marrow cells of male albino rats after ASP and/or silymarin administration

Fig. 10: Several metaphases from bone marrow cells of albino rat after ASP administration showed; Deletion (De), Dicentric (Di), Exchange fig. (Ex) and Break (Br)
Fig. 11: Aberrant metaphases from bone marrow cells of albino rat after Asp administration showed: (A) Deletion (De); (B) Dicentric (Di); (C) Dicentric (Di) and Break (Br); (D) Exchange (Ex).

Fig. 12: Histogram showed MI% of male albino rats after ASP and/or silymarin administration.

Histopathological examination on liver, kidney and heart tissue

Findings of histopathological examination of liver tissue showed that the control group and Silymarin group showed a normal structure in the hepatic tissues which consisted of central vein and sheets of hepatocyte. Aspartame group showed diffuse hepatocytes vacuolations. Recovery group showed normal hepatic cords with the slightly congested hepatoporal blood vessel. Aspartame plus silymarin group showed mild picture of hepatocytes vacuolations (fig. 13).

According to the present study, the histopathological examination of kidney showed that the control group and silymarin group showed normal structure appearance of real tissue renal parenchyma, normal glomeruli and renal tubules. Aspartame group showed diffuse vacuolation of glomeruli, renal tubules together with congested interstitial blood vessel with the thickened wall. Recovery group showed normal renal parenchyma. Note the normal glomeruli and renal tubules. Aspartame plus silymarin group showed normal renal parenchyma. Note the normal glomeruli and renal tubules (fig. 14).

In the current study, the histopathological examination of cardiac tissue demonstrated that the control group and the silymarin group showed normal myocardial muscles. Aspartame group showed hyalinized myocardial muscle with dilated and thickened walled blood vessel. The recovery group showed normal myocardial muscle with a slightly dilated blood vessel. In Aspartame plus silymarin group, there were normal myocardial muscles (fig. 15).
Fig. 13: Histopathological examination of liver tissue. A1 and A2 refer to group 1 (normal control) and group 2 (Silymarin) showed normal structure all clearance of hepatic tissue which consists of central vein and sheets of the hepatocyte. A3 refer to group 3 (Aspartame) showed diffuse hepatocytes vacuolations (arrow). A4 refer to group 4 (Recovery) showed normal hepatic cords with slightly congested hepatoporal blood vessel (arrow). A5 (Aspartame+silymarin) showed mild picture of hepatocytes vacuolations, (H&E X 200 and 400)

Fig. 14: Histopathological examination of kidney tissue. B1 and B2 refer to group 1 (normal control) and group 2 (Silymarin) showed normal structure appearance of real tissue renal parenchyma, normal glomeruli and renal tubules. B3 refers to group 3 (Aspartame) showed diffuse vacuolation of glomeruli, renal tubules together with congested interstitial blood vessel with the thickened wall (arrow). B4 refer to group 4 (Recovery) showed normal renal parenchyma, note the normal glomeruli and renal tubules. B5 (Aspartame+silymarin) showed normal renal parenchyma, note the normal glomeruli and renal tubules, (H&E X 200 and 400)
DISCUSSION

The synthetic and some of the naturally present food additives have been reviewed and documented for toxicity. Limit values have been evaluated for dietary intake by humans on the basis of the conclusion of data attained in experimental animals.

In the current investigation, we studied the ASP induced toxic effects and biochemical variations in blood and different tissues in addition to the CA frequencies and MI% in experimental rats. The effect of co-administration of ASP and silymarin and the role of silymarin in the amelioration of these toxic effects were also examined.

The effect on body weight

Data showed that there was a significant increase in the final body weight in group 3 (aspartame) (P < 0.001) and Group 4 (Recovery Group) (P < 0.05) which amounted to 27.1 % and 13.8 % respectively. In group 5 (Aspartame+silymarin) there was a statistically non-significant increase in the final body weight which amounted to 7 % (table 1) and (fig. 1).

These results are in line with who reported that rats, when treated with aspartame, had an increase in food and water intake. ASP increases appetite in rats. Appetite is known as a highly regulated phenomenon characterized by hunger and satiety as crucial factors in controlling food intake [37, 38].

Sweetness without energy has impacts on appetite because of a natural sweetness-energy association. Due to this association, it has been proposed that sweetness could be an indication of the arrival of the associated energy and its physiological and psychological effects on appetite [39].

In the case of the absence of the associated energy, as in the case of artificial sweeteners, the body needs energy, leading to the stimulation of appetite [40]. ASP increases appetite through its metabolites by different mechanisms. An increased phenylalanine concentration may encourage admission by means of the hypothalamic adrenoreceptors implicated in the central appetite control mechanisms, stimulating appetite [41]. The aspartate, which is the metabolite of aspartame, reached the brain in the arcuate (ARC) nucleus. This nucleus is the main place where the synthesis of neuropeptide Y (NPY), stimulates carbohydrate intake. Also, chronic ASP administration significantly decreased leptin concentration in plasma [42]. Leptin inhibits food intake in the brain, but lower concentrations of leptin, could stimulate appetite [43].

Increased fluid intake can be associated with the intensive sweet taste of ASP and its hedonic impact. ASP is about 200 times sweeter than sucrose [44].

Data had revealed that there was a significant increase in the final body weight in group 4 amounting 13.8 %. This indicated improvement when rats were recovered for 14 d. These results are in accordance with who reported that improvement of the ASP produced changes but never returned to control ones in rats that received ASP and were then left for 4 w to recover [45].

In group 5, there was a statistically non-significant increase in the final body weight which amounted to 7 %. This indicates the role of silymarin in restoring the side effects of ASP.

Effect on glucose, liver, kidney and lipid function tests

Data presented in (table 2) and (fig. 2) showed that the mean level of glucose was increased in groups 3, 4 and 5. This increase was statistically significant in group 3 (P < 0.01) and group 4 (P < 0.05). In group 5 silymarin administration decreased glucose level which was statistically non-significant compared to the control group (P > 0.05).

These results are in line with who reported that the control group demonstrated a glucose concentration underneath those of aspartame groups (P < 0.05) [38]. Also, previous data declared that ASP consumption significantly increased blood glucose level (P < 0.05), in a dose and time-dependent fashion [46].
Artificial sweeteners, including ASP, bring about impedance in insulin sensitivity [47]. Moreover, phenylalanine, resulting from ASP metabolism, may increase insulin and glucagon values in serum of healthy subjects [48]. It may induce an elevation of hepatic glucose production and its level in the blood of rats [49].

In group 4, there was a statistically significant (P<0.05) increase in glucose level indicating an improvement but not returning to normal values. In group 5, silymarin administration decreased the elevation in glucose level which was statistically non-significant (P>0.05) compared to the control group. These results were in agreement with who reported that there was an amelioration of blood glucose concentration in nickel animals treated with silymarin extract [50]. It was suggested that the protective role of silymarin was due to its antioxidant properties, an expansion of plasma and pancreatic glutathione concentrations or both [51].

Also, data showed a significant (P<0.05) increase in ALT activity in group 3. This value was decreased in group 4 and 5 (P>0.05). The mean value of AST activity increased in group 3, 4 and 5 but this increase was statistically non-significant (P>0.05) compared to the control group. ASP administration caused slight decrease in the mean level of albumin (P>0.05) and slight increase in total bilirubin (P>0.05) (table 2) and (fig. 2).

Data supported who recorded that a highly significant increase in ALT, AST and ALP activity was recorded in Wistar albino rats treated with ASP compared to controls [52].

At this point, ASP was retained from the intestinal lumen and compasses to liver by means of entrance course likewise to other amino acids. [53] Methanol is a chemical compound that is harmful to liver cells. It is oxidized to formaldehyde resulting in formate production [53]. When liver cells are damaged, it releases variety of enzymes that normally found in cytoplasm into the bloodstream. Therefore, their levels in the serum are valuable as a demonstrative marker of the degree and kind of hepatic damage.

Previous studies indicated a critical diminishing of level of serum albumin in contrast with the untreated group. Oral dosing of ASP had no impact on total serum protein at 45 d, but it essentially diminished at 90 d of administration [54].

Other studies revealed that administration of silymarin concomitantly with dexamethasone-induced a significant reduction of liver function enzymes (ALT, AST and ALP) and the significant increase in total protein and globulin as compared with dexamethasone-treated group alone [55]. This may be attributed to the protection of liver cells directly through stabilizing the cell membrane by preventing liver glutathione depletion and inhibiting lipid peroxidation, regulating cell membrane permeability and integrity inhibiting leukotriene and scavenging reactive oxygen species [56].

Data confirmed that ASP administration caused a slight increase in urea concentration (P>0.05). ASP caused a significant (P<0.01) increase in the mean level of creatinine. These values showed a slight decrease in group 4. But this was still statistically significant. Group 5 had marked decrease which was statistically non-significant (P>0.05) as compared to control group (table 2) and (fig. 2).

These outcomes agreed with who announced that the administration of ASP had a fundamentally expanded level of cholesterol and triglycerides in rats [37, 54].

Cholesterol is a major component of cell membranes. It is important for tissue growth and the production of steroid hormones. The regulation of cholesterol metabolism is regulated by the liver. ASP causes changes in different biochemical parameters, lipid metabolism and may be the cause of hyperglycemia and hypercholesterolemia [59].

Other authors have shown that hypersensitivity and atherosclerosis resulted from long-term ASP administration. Oxidative stress and oxidative damage of tissue could be the initial markers of some chronic diseases, like diabetes [60]. In diabetes, high glucose causes increased production of ROS in all tissues resulting from glucose auto-oxidation and protein glycosylation [61].

**The effect on oxidative and antioxidative parameters in liver tissue**

Data showed that the dosing of ASP caused a marked significant increase on hepatic MDA level (P<0.001) and a significant decrease on GSH levels (P<0.05). GST and catalase activities (P<0.001) as compared to the control group. The elevation on MDA level showed slight decrease in group 4 (P>0.01) and a marked decrease in group 5 but it remained significant (P<0.05) at the statistical level. Also, the decrease on hepatic GSH level, GST and catalase activities were enhanced in group 4 but, it was statistically significant as group 3 and showed marked elevation in group 5 which was statistically non-significant (P>0.05) as compared to the control group (table 3).

Previous data concluded that ASP administration caused a significant decrease in the hepatic levels of antioxidant enzymes (SOD, CAT and GPx). It may be due to the destroying effect caused by methanol associated free radicals or formaldehyde obtained from methanol oxidation [62].

Free radicals, including superoxide radical, hydroxyl radical, hydrogen peroxide and lipid peroxide radicals, are delivered as an ordinary outcome of biochemical procedures in body cells and expanded introduction to lethal compounds [63]. Free radicals induce peroxidation of polyunsaturated fatty acid in the cell membrane. This causes a chain reaction of lipid peroxidation, harms the cellular membrane and causes further oxidation of membrane lipids and proteins, DNA, RNA and other components [64]. The protective role of silymarin on the cells is mainly due to its antioxidant activity. Overall, the protective role of silymarin on liver includes its activity against lipid peroxidation as a result of free radical scavenging and its capacity to build GSH content in addition to silymarin’s ability to regulate the cell membrane permeability and to increase membrane stability in the presence of harmful agents damage [65].

**The Effect on oxidative and antioxidative parameters in kidney tissue**

Results confirmed that ASP administration caused a highly significant elevation in kidney MDA level (P<0.01) and a significant decrease in GSH levels (P<0.01). GST and catalase activities (P<0.001) as compared to the control group. The elevation in MDA level showed slight decrease in group 4 (P<0.01) and a marked decrease in group 5 but, it was still significant (P<0.05) at the statistical level. The decreased kidney GSH level, GST and catalase activities were enhanced in group 4, but the values were remained statistically significant as group 3 and showed marked elevation in group 5 which was statistically non-significant (P>0.05) in case of GSH and catalase and significant (P<0.05) in MDA and GST as compared to control group (table 4).

Results were in line with who revealed that oral administration of ASP to rats caused a significant increase in the levels of oxidative stress markers as compared to the control group. This indicated that ASP caused renal toxicity by oxidative stress due to the production of a number of free radicals by its metabolites (methanol) [57].

Other studies showed that GSH, GST, GPx and SOD activities were significantly diminished (P<0.01) in the kidney, while MDA level
witnessed a highly significant expansion (P<0.01) in ASP treated rats as compared to controls.

The reduction in GSH action was brought about by methanol since methanol metabolism relied upon GSH. Maybe it was brought about by its quick interaction with formaldehyde, created during methanol metabolism and forming nucleophilic adducts and/or lipid peroxidation products [66]. The lessening in action of cancer prevention agent chemicals is connected with the activity formaldelhyde and free radicals. Formaldehyde rapidly interacts with the amino acids of soluble proteins causing hydroxymethyl derivatives and intra and intermolecular bridges in proteins. Also, free radicals produced through the methanol oxidation can cause the formation of protein peroxides. These changes may result in denaturation, aggregation and fragmentation of proteins, changing physicochemical properties and potentially losing enzymatic activities [67].

The effect on oxidative and antioxidative parameters in heart tissues

The present study recorded that ASP administration caused a highly significant elevation in cardiac MDA level (P<0.001) and a significant decrease in GSH levels (P<0.01), GST and catalase activities (P<0.001) as compared to control values. The elevation in MDA level showed a slight decrease in group 4 (P<0.05) and a marked decrease in group 5 which was statistically non-significant (P>0.05). The decreased cardiac GSH level, GST and catalase activities were enhanced in group 4 but still statistically significant as group 3 and showed marked elevation in group 5 which was statistically non-significant (P>0.05) as compared to control group (table 5).

ASP administration for 3 mo to Wistar albino rats caused a significant decrease in enzymatic (SOD, CAT and GPx) and non enzymatic (GSH, Vit-C and Vit-E) antioxidants level when compared to the control animals [68].

Data showed that ASP has an effect on the different tissues. Recovery period (14 d) was not enough to restore the oxidative and anti-oxidative parameters to its normal levels. Also, co-administration of ASP with silymarin showed great improvement than the recovery group (fig4, fig 5, fig 6, fig 7).

The effect on serum levels of TNF--α

Tumor necrosis factor-α (TNF-α) is a pro-inflammatory cytokine produced in blood. It initiates the immune response. TNF-α is secreted by different types of cells. It is produced by activated macrophages, endothelial cells and B lymphocytes. It induced apoptosis, increased blood coagulation, increased expression of adhesion molecules on WBCs and endothelial cells, as well as the release of different cytokines, chemokines, leukotrienes and ROS [67]. These ROS are known to regulate different important cellular events in response to TNF-α, including NF-kB, cellular proliferation, and apoptosis [69].

Results presented in (table 6) indicated that there was a highly significant increase in the mean level of serum TNF-α in group 3 (P<0.001) as compared to control group. Group 4 showed a significant increase (P<0.05) but less than that of group 3. In group 5, there was statistically non-significant increase.

The high level of TNF-α is generally found in persons with metabolic disorder diseases like obesity and diabetes. It was reported that ASP acted as a chemical stressor and increased oxidative stress leading to inflammation [70]. TNF-α was observed to straight-forwardly interfere with tissue insulin receptors and leads to blocking insulin’s biological actions on cell. This causes insulin resistance because of TNF-α exposure [71].

These findings were in line with who confirmed that ASP increased serum TNF-α in mice as compared to controls [72]. Silymarin was able to restrict the excessive inflammation caused by stress by acting on a variety of molecular targets. It inhibited the production of TNF-α in a dose-dependent manner in rats [73].

The effect on the chromosomal aberration frequency

Under the conditions of the present investigation, the control rats recorded chromosomal aberration (CA) frequency within the normal values while oral administration of ASP to male rats induced significant (p<0.001) increase in the percentage of total chromosomal aberration (TCA) reaching 63.00± 2.89 as compared with 15.60± 0.67 in control group. TCA included the total numerical aberration (TNA) and the total structural aberration TSA (table 7) (fig. 9).

The numerical aberrations were mainly the aneuploidy (hyperdiploid) and polyplody (table 7), (fig. 9, fig. 10). Meanwhile, the deletions, dicentric chromosomes, exchange fig. and breaks were the most abundant structural aberrations (table 7), (fig. 9, fig. 11); all of these aberrations significantly (p<0.001) increased as compared with control values.

These results supported other previous studies who confirmed that ASP induced a significant increase in the TCA (structural and numerical) in liver and bone marrow cells of albino rats’ mothers and their offspring [13]. In, in Swiss albino mice [74, 75] and in human lymphocytes as compared with control values. Chromosome breaks, Chromatid breaks, single chromatid union and sister union were the most common on CA [76].

On the contrary, the present results disagreed with who reported that ASP had a moderate genotoxic effect [77]. Also, ASP and acesulfame-K administration induced no significant increase in the frequency of CA in Swiss albino mice bone marrow cells [78].

Genotoxicity of ASP in bone marrow cells of rats might be caused mainly by the toxic methanol (a metabolite of ASP) where ASP can be hydrolyzed into phenylalanine, aspartic acid, a cyclized diketopiperazene and methanol. Methanol further hydrolyzed to formaldehyde, which is concluded to be one of the DNA damaging agents and known to induce CA in living cells [79]. Also, ASP could induce DNA fragmentation and CA by increasing methanol levels in plasma which, in turn, induced a significant increase in the free radical production causing an imbalance in the ratio of antioxidant/pro-oxidant in the brain tissue in male waster rats [67, 80, 81]. Methanol and phenylalanine are the metabolic components of ASP having a genotoxic effect for humans [76].

In the present study, when rats were orally administered with ASP then left for two weeks post-treatment to recover (recovery group), the frequency of TCA (TNA and TSA) was clearly reduced as compared with control (table 7) and (fig. 9). Except for exchange figures, all types of structural aberrations decreased and became non-significant at the statistical level. Thus, better CA improvement after the stoppage of ASP may require an extension of the recovery period to more than two weeks.

This result completely agreed with [82] who confirmed that the administration of ASP had harmful effects on the sciatic nerve and one-month discontinuation of ASP was not enough to obtain complete recovery.

It is known that anti-oxidizing agents are confirmed to be useful in preventing or treating damage induced by free radicals. So, silymarin was used in the present investigation to ascertain whether the co-administration of ASP and silymarin is effective in reducing genotoxicity induced by ASP in bone marrow of albino rats.

According to the result of the present study, silymarin treated rats showed significant (p<0.05) decrease in the frequency of TNA as compared with control value. Herein, aneuploidy (hyperdiploid) was significantly (p<0.05) decreased and all other types of CA were non-significantly changed. This confirmed that silymarin itself does not induce CA and is able to repair the chromosomal damage (table 7) and (fig. 9).

These results completely confirmed who explained that silymarin drug as antioxidant alleviated the genotoxicity of ASP by clear reduction in
the frequency of TCA (TNA and TSA) in bone marrow cells of rats co-
administered both ASP plus silymarin. All types of CA (hyperdiploidy,
polydiploidy, deletions, dicentric chromosomes and breaks) were
highly reduced and became non-significant as compared with control
values. However, the frequency of TCA (TNA and TSA) mainly
elevated in ASP exposed rats as compared with control values (table 7)
and (fig. 9). The ameliorative role of silymarin against ASP toxicity might be a result of its anti-oxidative activity; it
acts as a scavenging agent against the free-radicals induced by ASP
that were the main cause of ASP genotoxicity repairing the chromosomal damage in bone marrow cells of ASP treated rats.

These results were confirmed by who recorded that the combined
treatment with silymarin and cyclophosphamide revealed that the silymarin successfully decreased frequency of all types of CA in mice [88]. Silymarin is a powerful antioxidant agent; it increases the activity of both glutathione peroxidase and superoxide dismutase promoting the free radical scavenging activity of glutathione of glutathione peroxidase system [85, 86].

The effect on the percentage of the mitotic index

According to the present work, the percentage of MI in increased in ASP group as compared with control value (table 7) and (fig. 12). Increased MI rate might result from a high progression rate of cells from the stage of DNA synthesis (S-phase) to mitosis (M-phase) of the cell cycle. This indicates the potential of ASP to induce more cells to divide and hence inducing cancer cells.

Results fully agreed with who recorded that ASP induced a significant increase in the percentage of MI in Allium cepa L.
meristematic cells compared with control group [87]. MI is significant for measuring the cell division rate [88].

In contrast, it was reported that ASP decreased the MI at all treatment periods and all concentrations in dose-dependent manner [74, 76]. Also, ASP did not significantly affect MI% [75]; many in vivo and in vitro studies showed that ASP is not genotoxic agent [89].

The present data recorded that the value of MI was relatively decreased in the recovery group compared with the aspartame group (table 7) and (fig. 12). Rats had been left for 14 d post-treatment with ASP to recover and this period was not enough to reach a complete recovery.

This confirmed the data concluded by who reported that the improvement from ASP induced cytotoxic effect was gradual and incomplete [19].

In addition, data showed that there was no significant change in MI% in silymarin treated rats. The animals administered with both ASP plus silymarin recorded an improvement in MI% as compared with rats treated with ASP alone, but the value didn't reach the control level (table 7) and (fig. 12).

These data supported who recorded that silymarin is famous as a safe herbal product [90]. It has a powerful anticlastogenic activity on genome of mice in germ and somatic cells. It has the ability to reduce the induction of sister chromatid exchange, giving a confirmation that it plays a remarkable role in repairing DNA damage. This role may be due to the action of silymarin as a strong antioxidant, or a DNA stabilizing agent. Thus, the usage of silymarin for the treatment of many diseases is not only safe but also has an anti-mutagenic activity warranted [91].

Histopathological examination

Findings of histopathological examination of liver tissue showed that the control group and Silymarin group showed a normal structure in the hepatic tissues which consisted of central vein and sheets of hepatocyte. Aspartame group showed diffuse hepatocytes vacuolations. Recovery group showed normal hepatic cords with slightly congested hepatopetal blood vessel. Aspartame plus silymarin group showed mild picture of hepatocytes vacuolations (fig. 13).

These results were in line with [52] who recorded critical changes in the livers of ASP treated animals when compared to control. Significant amount of neutrophil infiltration was noticed in the liver cells. ASP treatment caused necrosis in liver histopathology with the spillage of blood cells. There were clear toxicological consequences for the hepatic tissue; significantly obtained in female rats treated with a dose of aspartame of 500 mg kg-1 daily in drinking water [92].

According to the present study, histopathological examination of kidney showed that control group and silymarin group showed normal structure appearance of real tissue renal parenchyma, normal glomeruli and renal tubules. Aspartame group showed diffuse vacuolation of glomeruli, renal tubules together with congested interstitial blood vessel with thickened wall. Recovery group showed normal renal parenchyma. Note the normal glomeruli and renal tubules. Aspartame plus silymarin group showed normal renal parenchyma. Note the normal glomeruli and renal tubules (fig. 14).

These observations agreed with who viewed that the histopathological profile of the rat kidney treated with ASP showed congestion in the sclerotic cortical blood vessels with swelling in the coating epithelial cells of the tubules [93].

In the current study, the histopathological examination of cardiac tissue demonstrated that control group and silymarin group showed normal myocardial muscles. Aspartame group showed hyalinized myocardial muscle with dilated and thickened walled blood vessel. Recovery group showed normal myocardial muscle with slightly dilated blood vessel. In Aspartame plus silymarin group there were normal myocardial muscles (fig. 15). Results disagreed with who reported that there was no morphological difference in heart of ASP treated Wistar albino rats when compared to control [68].

CONCLUSION

The results of the current study have proved that ASP has cytotoxic effect that may evidenced by inducing a significant increase in the liver, kidney and lipid function tests, blood glucose level, serum TNF-
α level, MDA level, CA frequency and MI%. This is in addition to decreasing GSH level, GST and catalase activity. Recovery period (14 d) after ASP stoppage was not enough to restore the oxidative and anti-oxidative parameters to normal levels. Silymarin, as an antioxidant, has the ability to ameliorate ASP toxicities. Therefore, it is important to be very careful when adding ASP in beverages and foods as a sweetener. It must be added together with antioxidants like silymarin.

ABBREVIATION

Total numerical aberrations, TNA; Total structural aberrations, TSA; Total chromosomal aberrations, TCA.

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AUTHORS CONTRIBUTIONS

All the authors have contributed equally.

CONFLICT OF INTERESTS

Declared none

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