Supporting Information

Oxidative regulation of the mechanical strength of a C-S bond

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I. General procedures

Materials
Lab general solvents (dichloromethane, ethyl acetate, acetone, toluene, tetrahydrofuran, methanol) were purchased from VWR or Sigma Aldrich. 4-Dimethylaminopyridine, \( p \)-toluenesulfonic acid, N,N'-diisopropylcarbodiimide, 3,3'-thiodipropionic acid, glutaric acid, hydrogen peroxide, acetic anhydride, \( meta \)-chloroperoxybenzoic acid (mCPBA) and silica dioxide were purchased from Sigma Aldrich, Alfa Aesar or TCI and used without further purification.

Characterizations
\( ^1 \)H NMR spectra were collected on a Bruker Advance Neo - 500 MHz multinuclear NMR spectrometer. Chemical shifts are provided in ppm (\( \delta \)) and referenced to the residual \( ^1 \)H peak at 7.26 ppm in CDCl\(_3\). \( ^1 \)H shifts are reported as chemical shift, multiplicity, coupling constant if applicable, and relative integral. Multiplicities are reported as: singlet (s), doublet (d), doublet of doublets (dd), doublet of triplets (dt), doublet of doublet of doublets (ddd), doublet of doublet of triplets (ddt), triplet (t), triplet of doublets (td), quartet (q), pentet (p), multiplet (m), or broad (br). Coupling constants (\( J \)) are reported in Hz. Gel permeation chromatography (GPC) was performed on two Agilent PLgel mixed-C columns (10\(^5\) Å, 7.5x300 mm, 5 \( \mu \)m, part number PL1110-6500) using THF (stabilized with 100 ppm BHT) as the eluent. Molecular weights were calculated using a Wyatt Dawn EOS multi-angle light scattering (MALS) detector and Wyatt Optilab DSP Interferometric Refractometer (RI). The refractive index increment (dn/dc) values were determined by online calculation based on injections of known concentration and mass.

II. Synthesis procedures
1. **Synthesis of polymer PE-S**

   To a 50 mL oven dried round bottom flask, added glutaric acid I (528.4 mg, 4.0 mmol, 0.8 eq), 3,3’-thiodipropionic acid 2 (178.2 mg, 1.0 mmol, 0.2 eq), gem-dichlorocyclopropane (gDCC) diol 3$^1$ (855 mg, 5.0 mmol, 1.0 eq) and 4-(dimethylamino)pyridinium-4-toluenesulfonate (DPTS)$^2$ (588 mg, 2.0 mmol, 0.4 eq) as catalyst. 5 mL dry DCM was then added to the flask. After solids are completely dissolved, N, N'-diisopropylcarbodiimide (DIC) (2.32 mL, 15 mmol, 3.0 eq) was added dropwise to the solution. White precipitate formed during the addition. The flask was then sealed and allowed to stir at room temperature for 3 days. The resulting viscous mixture was precipitated from methanol three times. Obtained white gummy polymer PE-S was collected and dried under high vacuum (1.132 g, 82%). Obtained polymer was analyzed from GPC: $M_n = 71.7$ kDa, $D_M = 1.445$, dn/dc= 0.087. $^1$H NMR (500 MHz, CDCl$_3$) $\delta$: 4.33-4.15 (m, 10H), 2.84 (t, 2H), 2.68 (t, 2H), 2.45 (t, 8H), 2.20-2.06 (m, 5H), 1.99 (p, 4H). Calculated sulfide content: 2/10×100% = 20 mol%.

   Another PE-S polymer with $M_n = 60.0$ kDa ($D_M = 1.437$) was prepared using the same procedure.

2. **Synthesis of polymer PE-SO**

   The oxidation process was adapted from previous literature.$^3$

   Polymer PE-S (78 mg, $M_n = 71.7$ kDa, 0.05 mmol C-S bond) was weighted in a 10 ml scintillation vial. 1 mL DCM was added to completely dissolve the polymer. Acetic anhydride (5.2 μL, 0.055 mmol, 1.1 eq) and silica gel (50 mg) was added to the solution. Then, 30% hydrogen peroxide (57 μL, 0.5 mmol, 10 eq) was added to the mixture. The reaction was further stirred at room temperature for 6 h. The resulting mixture was then diluted with 2 mL DCM and silica gel was filtered using a syringe filter. Obtained DCM solution was condensed and precipitated from methanol three times to give polymer PE-SO (74 mg, 93.9%). GPC: $M_n = 59.9$ kDa, $D_M = 1.478$, dn/dc = 0.090. $^1$H NMR (500 MHz, CDCl$_3$) $\delta$: 4.36-4.16 (m, 10H), 3.16-3.05 (m, 1H), 3.01-2.93 (m, 1H), 2.93-2.84 (m, 2H), 2.45 (t, 8H), 2.20-2.06 (m, 5H), 1.99 (p, 4H). Calculated sulfoxide content: 1.9/10×100% = 19 mol%.

3. **Synthesis of polymer PE-SO$_2$**

   To a solution of PE-SO (156 mg, $M_n = 71.7$ kDa, 0.1 mmol C-S bond) in 3 mL THF, mCPBA (74 mg, 70%~75%, 0.3 mmol) was added in portions. The reaction was allowed to stir at room temperature overnight. After the reaction completed, the solution was condensed and precipitated from methanol three times to give a white polymer PE-SO$_2$ (158 mg, 99.9%). GPC: $M_n = 54.1$ kDa, $D_M = 1.478$, dn/dc = 0.091. $^1$H NMR (500 MHz, CDCl$_3$) $\delta$: 4.37-4.16 (m, 10H), 3.40 (t, 2H), 2.94 (t, 2H), 2.45 (t, 8H), 2.20-2.06 (m, 5H), 1.99 (p, 4H). Calculated sulfone content: 2/10×100% = 20 mol%.

### III. Sonication experiment

1. **General sonication procedures**

   A solution of 36 mg polymer (PE-S, PE-SO or PE-SO$_2$) in 18 mL dry THF ($c = 2$ mg/mL) was transferred into a Suslick cell. The solution was purged with N$_2$ for 10 min while cooled with ice bath. Pulsed ultrasound was applied (1s on, 1s off) at 30% amplitude. An aliquot of 0.8 mL sample was taken out for GPC analysis at each sonication time points: 0, 3, 6, 10, 15, 20, 30, 45 min. Each of these samples were further condensed in a 10 ml scintillation vial to give a thin layer of polymer at the vial bottom. The polymer was washed with methanol and further dried under high vacuum. Obtained dry polymers were further subjected to $^1$H NMR analysis.
2. Sonication of PE-S polymer

1) Calculation of PE-S polymer chain scission cycle

![Sonication time vs Retention time](image1)

**Figure S1.** Evolution of GPC traces during sonication of PE-S polymer (left) and corresponding plot of molecular weight versus sonication time (right).

The chain scission cycle (SC) describes the average number of times a polymer has undergone chain scission. SC of PE-S polymer at each sonication time was obtained from the following equation:

\[ SC = \frac{\ln M_n(o) - \ln M_n(t)}{\ln 2} \]

Where \( M_n(o) \) is the initial molecular weight; \( M_n(t) \) is sonicated molecular weight.

![Sonication time vs Scission cycle](image2)

**Figure S2.** Scission cycle (SC) of PE-S polymer at various sonication times.

2) Determination of ring-opening percentage of gDCC in PE-S polymer

In the \(^1\)H NMR spectrum, the peak at 4.17–4.32 ppm (\( H_a \)) corresponds to methylene protons adjacent to the cyclopropane in the pristine polymer. The peak at 6.13 ppm (\( H_a \)), 4.68–4.85 ppm (\( H_b \& H_c \)), and 6.39 ppm (\( H_d \)) are protons in the ring opened gDCC product. The ring-opening percentage can be obtained:

\[ \text{Ring opening} \% = \frac{4 \times \int H_a}{\int H_b + \int H_d + \int H_e} \times 100\% \]
Figure S3. Stack of $^1$H-NMR (CDCl$_3$, 500 MHz) spectra of sonicated PE-S polymer at various sonication times.

Figure S4. Ring-opening percentage of gDCC in PE-S polymer versus sonication time.

3) Evaluation of $\Phi$ value

$\Phi$ value, the slope of ring opening percentage of gDCC versus scission cycle (SC) plot, is used to evaluate the relative strength of C-S weak bonds.
**Figure S5.** Percentage of gDCC ring opening versus scission cycle plot for PE-S polymer. The slope of linear fitting through origin gives $\Phi = 0.334$ for $M_n = 72$ kDa and $\Phi = 0.382$ for $M_n = 60$ kDa.

3. **Sonication of PE-SO polymer**

The analysis of PE-SO polymer is the same as PE-S polymer.

1) Calculation of PE-SO polymer chain scission cycle

**Figure S6.** Evolution of GPC traces during sonication of PE-SO polymer (left) and corresponding plot of molecular weight versus sonication (right).
Figure S7. Scission cycle (SC) of PE-SO polymer at various sonication time

2) Determination of ring-opening percentage of gDCC in PE-SO polymer

\[
\begin{align*}
\text{O} & \quad \text{Cl} & \quad \text{Cl} \\
\text{H} & \quad \text{H} & \quad \text{H} \\
\text{O} & \quad \text{O} & \quad \text{He} \\
\end{align*}
\]

\[
\begin{align*}
\text{O} & \quad \text{Cl} & \quad \text{Cl} \\
\text{H} & \quad \text{H} & \quad \text{H} \\
\text{O} & \quad \text{O} & \quad \text{He} \\
\end{align*}
\]

45 min \quad H_a \\
30 min \\
20 min \\
15 min \\
10 min \\
6 min \\
3 min \\
0 min

\( \delta \) (ppm)

6.5 6.4 6.3 6.2 6.1 6.0 5.9 5.8 5.7 5.6 5.5 5.4 5.3 5.2 5.1 5.0 4.9 4.8 4.7 4.6 4.5 4.4 4.3 4.2 4.1 4.0

Figure S8. Stack of \(^1\text{H-NMR (CDCl}_3, 500 \text{ MHz)}\) spectra of sonicated PE-SO polymer at various sonication times.

Figure S9. Ring-opening percentage of gDCC in PE-SO polymer versus sonication time.

3) Evaluation of \( \Phi \) value
Figure S10. Percentage of gDCC ring opening versus scission cycle plot for PE-SO polymer. The slope of linear fitting through origin gives $\Phi = 0.125$.

4. Sonication of PE-SO$_2$ polymer

The procedure of analyzing PE-SO$_2$ polymer is the same as PE-S polymer.

1) Calculation of PE-SO$_2$ polymer chain scission cycle

Figure S11. Evolution of GPC traces during sonication of PE-SO polymer (left) and corresponding plot of molecular weight versus sonication (right).

Figure S12. Scission cycle (SC) of PE-SO polymer at various sonication time
2) Determination of ring-opening percentage of gDCC in PE-SO$_2$ polymer

![Diagram of ring-opening transformation]

Figure S13. Stack of $^1$H-NMR (CDCl$_3$, 500 MHz) spectra of sonicated PE-SO$_2$ polymer at various sonication times.

Figure S14. Ring-opening percentage of gDCC in PE-SO$_2$ polymer versus sonication time.
3) Evaluation of Φ value

![Figure S15](image)

**Figure S15.** Percentage of gDCC ring opening versus scission cycle plot for PE-SO₂ polymer. The slope of linear fitting through origin gives Φ = 0.365.

**IV. CoGEF modeling**

CoGEF modeling was conducted using Spartan '18 V1.4.1 software. The methyl ester form of each sulfide, sulfoxide or sulfone containing repeating unit was subjected to CoGEF modeling using DFT method on the theory level of B3LYP/61G*. The ground state geometry was first optimized, and then its end-to-end distance was constrained with step increasement of 0.1 Å. The optimized energy was plotted as a function of end-to-end distance.

1. CoGEF modeling of sulfide repeating unit

![Figure S16](image)

**Figure S16.** CoGEF modeling of sulfide containing unit. Left: the evolution of energy over increasement in end-to-end distance. Right: the length of two C-S bonds at different constrained distance.

2. CoGEF modeling of sulfoxide repeating unit
Figure S17. CoGEF modeling of sulfoxide containing unit. Left: the evolution of energy over increase in end-to-end distance. Right: the length of two C-S bonds at different constrained distance.

3. CoGEF modeling of sulfone repeating unit

Figure S18. CoGEF modeling of sulfone containing unit. Left: the evolution of energy over increase in end-to-end distance. Right: the length of two C-S bonds at different constrained distance.

Figure S19. Overlay of length of breaking C-S bonds at various constrained distance.
Table S1. Summary of $F_{max}$ obtained from GoGEF modeling of C-S bond containing species

| C-S bond | Sulfide | Sulfoxide | Sulfone |
|----------|---------|-----------|---------|
| $F_{max}$ (nN) | 4.61    | 3.78      | 4.07    |

V. Calculation of bond dissociation energy

The methyl ester form of each sulfide, sulfoxide or sulfone containing repeating unit was applied to calculate the C-S bond dissociation energy (BDE). The BDE was calculated using equation: $\text{BDE} = E(\text{Frag1}) + E(\text{Frag2}) - E(\text{SM})$.

![Chemical structures of Sulfide, Sulfoxide, and Sulfone]

The calculation was performed in Spartan '18 V1.4.1 software. The energy of each starting molecule was optimized using DFT method on theory level of B3LYP/6-311+G** (in nonpolar solvent, $\varepsilon = 7.43$), and the two radical fragments were optimized using the same method but with one unpaired electron. The obtained optimized energy for each molecule/fragment was provided in the following table.

Table S2. Summary of calculated bond dissociation energy (BDE) for C-S bond containing species

|          | Optimal energy ($E_h$) | BDE  |
|----------|------------------------|------|
|          | SM | Fragment 1 | Fragment 2 | $E_h$ | kcal/mol |
| Sulfide  | -1012.61727 | -307.133215 | -705.375901 | 0.10815 | 67.9     |
| Sulfoxide| -1087.82834 | -307.133215 | -780.621622 | 0.07350 | 46.1     |
| Sulfone  | -1163.06402 | -307.133215 | -855.834720 | 0.09609 | 60.3     |
VI. $^1$H NMR spectrum

$^1$H NMR (CDCl$_3$, 500 MHz) spectrum of PE-S polymer

$^1$H NMR (CDCl$_3$, 500 MHz) spectrum of PE-SO polymer
**VII. Reference**

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$^1$H NMR (CDCl$_3$, 500 MHz) spectrum of **PE-S** polymer