Respiratory MUC5B disproportion is involved in severe community-acquired pneumonia

Lu Fan1,2†, Yi Lu1,3†, Yan Wang4, Xiaomin Zhang1, Yuxuan Wu1, Hao Sun1* and Jinsong Zhang1*

Abstract
Background: Mucus production is a process involved in the pathogenesis of Community-acquired pneumonia (CAP). The study is to determine Mucin 5B (MUC5B) protein concentration and its proportion in the bronchoalveolar lavage fluid (BALF) of CAP patients and evaluate its value to help assess disease severity.

Methods: A total of 118 patients were enrolled in this cross-sectional study, including 45 with severe CAP (SCAP) and 73 with non-severe CAP (NSCAP). MUC5B concentration in BALF were determined by immunoblotting analysis. Total protein concentration of BALF was detected by Pierce BCA kit. Cytokines IL6, IL10, IFNγ, IL13, and IL17 in BALF were measured using commercial enzyme-linked immunosorbent assay (ELISA). Spearman's correlation analysis was applied to evaluate the relationships between MUC5B concentration or MUC5B/total protein ratio and the CURB-65 score, as well as cytokines. Logistic regression analysis was used to identify the independent factors associated with severe CAP. Receiver operating characteristic (ROC) curve was used to evaluate the assessment value of MUC5B/total protein ratio and other indexes for CAP severity.

Results: MUC5B concentration in the BALF of NSCAP group was higher than that in SCAP group [NSCAP 13.56 µg/ml (IQR 5.92–25.79) vs. SCAP 8.20 µg/ml (IQR 4.97–14.03), \(p=0.011\)]. The total protein concentration in the BALF of NSCAP group was lower than that in SCAP group [NSCAP 0.38 mg/ml (IQR 0.15–1.10) vs. SCAP 0.68 mg/ml (IQR 0.46–1.69), \(p=0.002\)]. The MUC5B/total protein ratio was remarkably higher in NSCAP group than that in SCAP groups [NSCAP 3.66% (IQR 1.50–5.56%) vs. SCAP 1.38% (IQR 0.73–1.76%), \(p<0.001\)]. MUC5B/total protein ratio was negatively correlated with total protein concentration (\(r_s = -0.576, p<0.001\)), IL6 (\(r_s = -0.312, p=0.001\)), IL10 (\(r_s = -0.228, p=0.013\)), IL13 (\(r_s = -0.183, p=0.048\)), IL17 (\(r_s = -0.282, p=0.002\)) and CURB-65 score (\(r_s = -0.239, p=0.009\)). Logistic regression identified that MUC5B/total protein ratio, IL6 level and CURB-65 score as independent variables related to CAP severity. ROC curve demonstrated best assessment value of MUC5B/total protein ratio for SCAP (AUC 0.803, \(p<0.001\)), with a sensitivity of 88.9% and a specificity of 64.4%.

Conclusions: Respiratory MUC5B disproportion is related to CAP severity. MUC5B/total protein ratio may serve as an assessment marker and a potential therapeutic target for severe CAP.

Keywords: Community-acquired pneumonia, MUC5B, Severity, Disproportion

Introduction
Community-acquired pneumonia (CAP) is one of major lethal infectious disease [1] with the incidence reaches 30–50% in adults [2]. Approximately 10% patients with CAP will develop severe CAP (SCAP) and require ICU treatment in which the mortality rate ranges from 19 to 50% [3]. Common symptoms of CAP include fever, cough and increased sputum production [4]. However, severe...
cases are prone to develop serious complications due to complexity and heterogeneity of variant risk factors [5], such as virulence and serotypes of pathogens, the age, immune state and comorbidities of patients, as well as genetic variants [6]. Given the inadequacy of the condition assessment of SCAP, a profound understanding of SCAP pathogenesis and a sensitive molecular marker are expected to improve patients’ diagnosis and treatment.

One major feature of CAP is mucus production and mucus is mainly composed of mucins [7, 8]. Previous animal experiments have proved that airway secreted mucin MUC5B but not Mucin 5AC (MUC5AC) plays a critical role in immune defense against bacterial infections [9]. MUC5B helps form gel in the airway as a defensive barrier and regulates the rheology of airway mucus [10]. Electron microscopy has shown that multiple MUC5B filaments always appear as strands lining in the airway. It traps and sweeps away pathogens from the lung by working with other types of mucins [11]. Previous studies showed that MUC5B participates in the development of pulmonary diseases, such as pulmonary fibrosis, chronic obstructive pulmonary disease (COPD) and bronchiectasis [12–14]. So far, whether MUC5B expression is involved in the development of SCAP has not been studied.

In our study, we compared the MUC5B concentration and its proportion in bronchoalveolar lavage fluid (BALF) of patients with NSCAP and SCAP. We analyzed the correlation between MUC5B expression and CAP severity to determine its value as a marker to help assess disease severity.

Material and methods

Study design

This cross-sectional study recruited patients from three hospitals including Jiangsu Province Hospital, Nanjing Chest Hospital and Qixia Branch of Jiangsu Province Hospital. The consecutive adult CAP patients treated in the respiratory and critical care general ward or intensive care unit (ICU) from Jan 2021 to Aug 2021 were screened for participants. The study was approved by the Institutional Review Board of coordinating center Jiangsu Province Hospital (No. 2021-SR-028). Written informed consent of the bronchoalveolar lavage (BAL) procedure and BALF sample utilization was obtained from each participant or from a relative or main care.

The inclusion criteria were as follows: 1. Men and women aged over 18 years old; 2. Diagnosed of CAP according the ATS guidelines; 3. Untreated or received treatment less than 24 h before admission; 4. Patients underwent BAL as standard of diagnosis [15, 16]; 5. Signed informed consent form. The exclusion criteria were as follows: 1. Patients with active pulmonary tuberculosis; 2. Patients with chronic pulmonary disease including chronic bronchitis, bronchiectasis, asthma, lung cancer, pulmonary embolism or interstitial lung diseases; 3. Patients with malignancy or severe immunosuppression; 4. Patients with pregnancy.

According to the ATS Guidelines [17] for the management of adults with CAP: CAP was diagnosed if the patient presented with at least one newly acquired respiratory symptoms such as cough, sputum and dyspnea setting on at communities, accompanied with fever, abnormal breath sounds and crackles. Each patient underwent a computerized tomography (CT) scan to identify patchy infiltrate shadows, lobar or segmental consolidation shadows, ground glass shadows or interstitial changes.

Severe CAP (SCAP) was defined during 24 h after admission by the criteria in ATS consensus guidelines 2007 [18]. The major criteria are as follows: 1. Patients needing invasive mechanical ventilation; 2. Patients showing septic shock needing vasopressors. The minor criteria included: 1. Respiratory rate ≥ 30 breaths/min; 2. PaO2/FiO2 ratio ≤ 250; 3. Multi-lobar infiltrates; 4. Confusion/disorientation; 5. Uraemia (BUN level ≥ 20 mg/dl); 6. Leucopenia (WBC < 4,000/mm³); 7. Thrombocytopenia (platelet count < 100,000 /mm³); 8. Hypothermia (core temperature < 36 °C); 9. Hypotension requiring aggressive fluid resuscitation. CAP patients presenting at least one major criterion or at least three minor criteria were diagnosed as SCAP.

Baseline clinical parameters of patients were collected from electronic hospital records (EHR). All participants received BAL procedure during 24 h after hospitalization. Through standardized bronchoscopy and performed the protocol of 2012 ATS guideline for BAL [19], the BAL was proceeded using a 120 ml lavage of 0.9% saline [20]. The BAL fluid (BALF) was then collected through a sputum box attaching to the suction canal of the bronchoscope with qualified at least 10% returned rate [21]. The BALF sample was filtered by gauze and immediately frozen at −80 °C for further experiment. CT scan and blood test were carried out before lavage. Blood indexes included white blood cells count (WBC), leukocyte classification, lactate dehydrogenase (LDH), C-reaction protein (CRP), erythrocyte sedimentation rate (ESR), fibrinogen (Fib), and d-dimer (D-D).

Inflammatory markers in BALF tested by ELISA

Concentration of cytokines including interleukin-6 (IL6), interleukin-10 (IL10), interferon-γ (IFNy), interleukin-13 (IL13), and interleukin-17 (IL17) in BALF was determined using commercial enzyme-linked immunosorbent assay (ELISA) kits specific for human following protocols (IL6, Cat. #: EHC007; IL10, Cat. #: EHC009; IFNy, Cat. #...
MUC5B expression measurement

The BALF was thawed and then centrifuged at 1200r for 10 min at 4 °C. MUC5B in the supernatant was measured by immunoblotting analysis using S&S MINIFOLD® and has been described in our previous study [22]. The MUC5B standard concentration driving from human saliva ranged from 0.87 to 0.01 (µg/ml). Samples were diluted with dilution buffer (3 M urea) with a ratio of 1:50. Standards and samples were run in duplicate (100 ul per sample). The protein blots binding to PVDF membrane were incubated with MUC5B primary antibodies (Santa Cruz Bio.), then with secondary goat anti-rabbit IgG Biotin conjugate and strep-HRP (Life Technologies). The blots were visualized using an enhanced chemiluminescence system. Immunoreactive dots were quantified using ImageJ software. Total protein concentration in BALF supernatant was determined by Pierce BCA protein assay kit (Cat. #: 23227; Thermo Scientific, Rockford, USA).

Statistical analysis

The current sample size achieved the study objectives with a desired power 0.80. Statistical analyses were performed with SPSS 17.0 (SPSS Inc., Chicago, IL). Continuous variables with normal distribution were presented as mean ± SD. Variables with abnormal distribution were presented as median (Interquartile range, IQR). Categorical variables were displayed as percentages. Student t-test was applied for comparison between normal distributed variables. Mann–Whitney U test was used to compare abnormal distributed variables. The Fisher's exact test or chi-square test was used to compare categorical variables. Spearman’s correlation coefficients ($r_s$) were measured to evaluate the relationships between MUC5B concentration or MUC5B/Total protein ratio and CURB-65 score, total protein concentration, as well as inflammatory markers in BALF. Binary logistic regression analysis was used to identify independent factors associated with severe CAP. The assessment value of MUC5B/Total protein ratio or other indexes for CAP severity was evaluated using receiver operating characteristic (ROC) curve. All statistics were two-sided, only a $p$ value less than 0.05 was considered statistically significant.

Results

Demographic characteristics and clinical parameters of participants

A total of 118 patients confirmed with CAP were included in the research and divided into NSCAP group ($n=73$) and SCAP group ($n=45$) (Fig. 1). Demographic characteristics and clinical parameters are described in Table 1. The frequency of difficulty breathing was significantly higher in SCAP group than in NSCAP group ($p=0.001$). Other symptoms including cough, sputum and fever showed no significant differences between the two groups. CURB-65 scores were significantly higher in SCAP group than that in NSCAP group ($p<0.001$). In laboratory parameters, percentage of neutrophils (NE%), serum levels of LDH, CRP, ESR, plasma level of Fib and D-D were remarkably higher in SCAP patients than those in NSCAP patients ($p<0.05$, respectively). Percentage of lymphocytes (LYM%) in SCAP group was lower than that in NSCAP group ($p=0.028$).

![Flowchart](image_url)  
Fig. 1 Study flowchart of patient enrollment. CAP community-acquired pneumonia, NSCAP non-severe CAP, SCAP severe CAP
Table 1 Demographic and clinical characteristics of the subjects enrolled in this study

| Characteristic                     | NSCAP   | SCAP     | p value |
|------------------------------------|---------|----------|---------|
| Demographics                       |         |          |         |
| N                                  | 73      | 45       |         |
| Age (years)                        | 50.9 ± 14.1 | 56.2 ± 16.7 | 0.068   |
| Gender, male (%)                   | 42 (57.5%) | 18 (40.0%) | 0.064   |
| Symptoms and CURB-65 score         |         |          |         |
| Cough                              | 62 (84.9%) | 40 (88.9%) | 0.542   |
| Sputum                             | 54 (74%) | 36 (80%) | 0.455   |
| Difficulty breathing               | 14 (19.2%) | 22 (48.9%) | 0.001   |
| Fever                              | 35 (47.9%) | 24 (53.3%) | 0.570   |
| CURB-65 score                      | 0 57 (78.1%) | 20 (44.4%) | <0.001  |
|                                   | 1 15 (20.5%) | 14 (31.1%) |         |
|                                   | 2 1 (1.4%) | 8 (17.8%) |         |
|                                   | 3 0 (0%) | 2 (4.4%) |         |
|                                   | 4 0 (0%) | 1 (2.2%) |         |
|                                   | 5 0 (0%) | 0 (0%) |         |
| Comorbid conditions                |         |          |         |
| Current or ex-smoker               | 21 (28.8%) | 6 (13.3%) | 0.053   |
| Hypertension                       | 15 (20.5%) | 15 (33.3%) | 0.121   |
| Diabetes mellitus                  | 9 (12.3%) | 6 (13.3%) | 0.874   |
| Chronic cardiac disease            | 1 (1.4%) | 1 (2.2%) | 0.728   |
| Prior malignancy                   | 5 (6.8%) | 0 (0%) | 0.073   |
| Immune disorder                    | 0 (0%) | 1 (2.2%) | 0.201   |
| Inflammatory biomarkers in Peripheral blood |         |          |         |
| WBC (*10^9/l)                      | 6.1 (4.9–8.0) | 6.5 (5.2–8.1) | 0.814   |
| NEUT (%)                           | 64.6 (54.6–73.4) | 71.0 (61.3–78.0) | 0.028   |
| LYM (%)                            | 26.7 (18.3–35.2) | 19.6 (16.4–30.0) | 0.028   |
| MO (%)                             | 6.0 (5.0–7.3) | 5.7 (4.5–7.9) | 0.459   |
| EO (%)                             | 1.5 (0.7–2.6) | 1.6 (1.7–2.4) | 0.934   |
| PLT (*10^9/l)                      | 230 (178.5–269.5) | 229 (168.5–249.5) | 0.458   |
| LDH (U/l)                          | 108.0 (20.8–177.0) | 203.0 (177.5–231.0) | <0.001  |
| CRP (mg/l)                         | 3.8 (0.6–29.0) | 16.0 (2.4–56.7) | 0.024   |
| ESR (mm/h)                         | 210 (80.0–320.0) | 565 (378–873) | <0.001  |
| Fib (g/l)                          | 3.3 (2.4–5.1) | 4.4 (3.2–5.9) | 0.012   |
| D-D (mg/l)                         | 0.3 (0.1–0.7) | 0.6 (0.4–1.2) | 0.002   |

Data presented as number (percentage) or mean ± SD or median (interquartile range)

CAP, community-acquired pneumonia; NSCAP, non-severe CAP; SCAP, severe CAP; CURB-65, confusion, urea level, respiratory rate, blood pressure, and age ≥ 65 years; WBC, white blood cell; NEUT%, percentage of neutrophils; LYM%, percentage of lymphocytes; MO%, percentage of monocytes; EO%, percentage of eosinophilic granulocytes; PLT, blood platelets; LDH, lactate dehydrogenase; CRP, C-reactive protein; ESR, erythrocyte sedimentation rate; Fib, fibrinogen; D-D, d-dimer

MUC5B condition and its correlation with related indexes

MUC5B concentration in the BALF of NSCAP group was higher than that in SCAP group [NSCAP 13.56 µg/ml (IQR 5.92–25.79) vs. SCAP 8.20 µg/ml (IQR 4.97–14.03), p = 0.011]. The total protein concentration in the BALF of NSCAP group was lower than that in SCAP group [NSCAP 0.38 mg/ml (IQR 0.15–1.10) vs. SCAP 0.68 mg/ml (IQR 0.46–1.69), p = 0.002]. The MUC5B/total protein ratio was remarkably higher in NSCAP group than that in SCAP groups [NSCAP 3.66% (IQR 1.50–5.56%) vs. SCAP 1.38% (IQR 0.73–1.76%), p < 0.001]. For inflammatory markers in BALF, the levels of IL6, IL13 and IL17 were lower in NSCAP group than those in SCAP group (p < 0.05). The levels of IL10 and IFNγ showed no significant difference between the two groups (Table 2).

The correlations among MUC5B level, total protein concentration, MUC5B/total protein ratio and the inflammatory parameters in BALF as well as CURB-65 score were investigated by Spearman’s correlation analysis (Table 3). MUC5B level had positive correlation with total protein concentration (rs = 0.423, p < 0.001) but no correlation with inflammatory cytokines. MUC5B/total protein ratio was negatively correlated with total protein concentration (rs = −0.576, p < 0.001), IL6 (rs = −0.312, p = 0.001), IL10 (rs = −0.228, p = 0.013), IL13 (rs = −0.183, p = 0.048) and IL17 (rs = −0.282, p = 0.002). MUC5B level and MUC5B/total protein ratio both had negative correlations with CURB-65 score (rs = −0.218, p = 0.018 for MUC5B, rs = −0.239, p = 0.009 for MUC5B/total protein ratio, respectively).

Assessment performance of MUC5B/total protein ratio for CAP severity

Though MUC5B concentration and total protein level showed significant differences between NSCAP and SCAP groups, the binary logistic regression analysis revealed that MUC5B concentration and total protein level were not independent factors associated with CAP severity. Logistic regression identified that MUC5B/total protein ratio, IL6 level and CURB-65 score remained as variables significantly related to CAP severity (Table 4).

ROC analysis was applied to evaluate whether MUC5B/total protein ratio could be used as a sensitive biomarker for CAP severity. Table 5 and Fig. 2 showed MUC5B/total protein ratio (AUC 0.803, p < 0.001) presented a better performance to assess the CAP severity with a sensitivity of 88.9% and a specificity of 64.4% comparing to CURB-65 score (AUC 0.692, p < 0.05). The total protein concentration in the BALF (AUC 0.791, p = 0.013) and the cytokine IL6 (AUC = 0.791, p < 0.001). The optimal cut-off point of MUC5B/total protein ratio to distinguish SCAP from NSCAP was 2.117%, with a positive predictive value of 60.6% and a negative predictive value of 90.4%.
In the present study, we showed that the MUC5B concentration and MUC5B/total protein ratio in SCAP group were obviously lower than those in NSCAP group. The MUC5B/total protein ratio was an independent factor associated with CAP severity. We speculate that the disproportion of respiratory MUC5B plays an important role in regulating pulmonary inflammation in severe CAP.

MUC5B is the main expressed mucin in normal human and mouse airway [23, 24]. It plays a constitutive role in mucus barrier which traps and eliminates particulates and pathogens via mucociliary clearance (MCC) [25, 26]. In vivo and vitro experiments showed MUC5B expression could be induced by multiple pathogens, such as Rhinovirus (RV) [27], Mengovirus [28], Pseudomonas aeruginosa (PA) [29], Mycoplasma pneumoniae [30], A. pleuropneumoniae [31] and Pneumocystis [32]. The mechanism mainly involves STAT3-STAT6/EGFR-FOXA2 signaling [29, 30, 33]. Overexpressed MUC5B can bind to pathogens to prevent their attachment to the epithelium and initiate the clearance of airway pathogens [34, 35]. Furthermore, Muc5b defect increases the accumulation of pathogens in mice respiratory tract, which leads to chronic bacterial infection and hardly dissolved inflammation [9, 36]. By contrast, Muc5AC is the other secreted mucin expressed in airway, but it is not required for MCC or for diminishing infections in the airway [9]. It was concluded that MUC5B is the only secreted mucin that regulates airway homeostasis and mucosal immunity in humans [9]. In our observational study, the result showed MUC5B/total protein ratio was significantly related with CAP severity. However, whether lower concentration of MUC5B was involved in the develop of CAP severity was uncertain. We assume that MUC5B disproportion may diminish the barrier function of respiratory tract during CAP pathological process, but the hypothesis needs to be proved by further animal experiments.

Furthermore, we assessed the relationship between MUC5B and inflammatory factors. The results showed

**Table 2** MUC5B concentration and inflammatory biomarkers in BALF of NSCAP patients and SCAP patients

| Related indexes | NSCAP | SCAP | p value |
|-----------------|-------|------|---------|
| MUC5B concentration (µg/ml) | 13.56 (5.92–25.79) | 8.20 (4.97–14.03) | 0.011 |
| Total protein concentration (mg/ml) | 0.38 (0.15–1.10) | 0.68 (0.46–1.69) | 0.002 |
| MUC5B/Total protein percent (%) | 3.66 (1.50–5.56) | 1.38 (0.73–1.76) | <0.001 |
| IL6 (pg/ml) | 1.81 (1.22–3.08) | 5.83 (2.50–15.73) | <0.001 |
| IL10 (pg/ml) | 0.18 (0.11–0.32) | 0.15 (0.12–0.27) | 0.632 |
| IFNγ (pg/ml) | 2.98 (1.90–4.04) | 3.27 (2.11–4.69) | 0.195 |
| IL13 (pg/ml) | 30.00 (28.78–31.66) | 31.08 (30.20–33.35) | 0.004 |
| IL17 (pg/ml) | 9.53 (6.52–11.23) | 12.04 (9.47–17.54) | <0.001 |

Data presented as median (interquartile range)

**Table 3** Correlation between MUC5B level, total protein concentration, MUC5B/total protein ratio and related indexes

| Related indexes | MUC5B | MUC5B/total protein ratio |
|-----------------|-------|--------------------------|
| rs, p value     | rs, p value |
| Total protein   | 0.423 <0.001 | −0.576 <0.001 |
| IL6             | −0.021 0.820 | −0.312 0.001 |
| IL10            | −0.095 0.307 | −0.228 0.013 |
| IFNγ            | −0.145 0.118 | −0.044 0.636 |
| IL13            | −0.101 0.276 | −0.183 0.048 |
| IL17            | 0.085 0.360 | −0.282 0.002 |
| CURB-65 score   | −0.218 0.018 | −0.239 0.009 |

rₜ, spearman rho correlation coefficients; CURB-65, confusion, urea level, respiratory rate, blood pressure, and age > 65 years

**Table 4** Logistic regression of variable parameters for determining the severity of CAP

| Variable        | OR    | 95% CI        | p value |
|-----------------|-------|---------------|---------|
| MUC5B           | 0.980 | 0.929–1.035   | 0.475   |
| Total protein   | 0.902 | 0.693–1.175   | 0.446   |
| MUC5B/total protein ratio | 0.417 | 0.230–0.756   | 0.004   |
| IL6             | 1.117 | 1.036–1.205   | 0.004   |
| IL13            | 0.944 | 0.742–1.201   | 0.638   |
| IL17            | 1.072 | 0.964–1.192   | 0.201   |
| CURB-65 score   | 3.494 | 1.450–8.415   | 0.005   |
| Constant        | 4.236 | 0.688         | 0.688   |

OR, Odds ratio; CI, confidence interval; CURB-65, confusion, urea level, respiratory rate, blood pressure, and age > 65 years

**Discussion**

In the present study, we showed that the MUC5B concentration and MUC5B/total protein ratio in SCAP group were obviously lower than those in NSCAP group.
that the SCAP patients exhibited an increase in a broad scope of cytokines in BALF, especially IL6, IL13 and IL17. MUC5B/total protein ratio was negatively correlated with levels of IL6, IL10, IL13 and IL17, especially IL6 (p = 0.001) and IL17 (p = 0.002). IL6 is a pro-inflammatory cytokine involved in the pathogenesis of airway inflammatory diseases [37]. It decides CD4+ T cell fate and promotes preferential Th2 differentiation [38, 39]. In vitro study has demonstrated that IL-6 regulates Muc5b expression via the ERK signaling pathway [40]. IL-17 is also a strong pro-inflammatory factor that can induce excess inflammation through cytokine cascade [41]. It has been reported that IL-17 mediates Muc5b expression by the ERK signaling pathway and NF-kB-based transcriptional mechanism [40, 42]. MUC5B disproportion was correlated with high IL6 and IL17, indicating its contribution to lung inflammatory augmentation in severe CAP. We suggest that multiple inflammatory pathways coordinate with MUC5B disproportion regulate CAP pathogenesis.

MUC5B disproportion could also be detected in other pulmonary diseases and smoke exposure. The SPIROMICS data showed that MUC5B concentration in sputum increased with COPD severity [13]. Ever-smokers (current and former smokers) without evidence of COPD also had a higher concentration of MUC5B in sputum than no-smoke controls [43]. Our studies excluded patients with history of COPD. But the percentage of ever-smokers in NSCAP group was slightly higher than the SCAP group. It seems that smoke exposure might be partially accountable to the higher MUC5B level in NSCAP group. But when comparing the MUC5B level between ever-smokers and no-smokers, no significant difference was detected neither in the NSCAP group nor in the SCAP group. We assumed there may be limited impact of smoke exposure on the level of MUC5B. In the future we would like to conduct a randomized controlled trial to clarify this issue. MUC5B disproportion could also be found in other pulmonary diseases. In Non-CF Bronchiectasis (NCFB) study, the ratio of MUC5AC to MUC5B was about 4 times higher in NCFB patients than in healthy controls [14]. In asthma, MUC5B levels decreased or remained the same, while other types of mucin levels were increased significantly [44]. The latest reports showed abnormal MUC5B expression in COVID-19 patients [45]. Further study found MUC5B playing a protective role against COVID-19 [46]. We suggested that regulating MUC5B proportion might be a therapy target for SCAP patients in future. A recent research has proven that restoring Muc5b level can improve lung function and alleviate inflammatory responses in a rodent model [47], but more studies are needed to confirm the role of MUC5B in SCAP development.

Table 5  

| Parameter                        | Cut-off value | AUC   | Sensitivity | Specificity | p value | 95% CI Lower limit | 95% CI Higher limit |
|----------------------------------|---------------|-------|-------------|-------------|---------|--------------------|--------------------|
| MUC5B/total protein ratio        | < 2.117       | 0.803 | 0.889       | 0.644       | < 0.001 | 0.726              | 0.880              |
| IL6 (pg/ml)                      | > 4.349       | 0.791 | 0.667       | 0.849       | < 0.001 | 0.708              | 0.874              |
| CRUB-65 score                    | –             | 0.692 | 0.556       | 0.781       | < 0.001 | 0.588              | 0.795              |

AUC, area under the curve; CI, confidence interval; CURB-65, confusion, urea level, respiratory rate, blood pressure, and age > 65 years

**Limitations**

First, this result was based on a small population sized cross-sectional study. The patients included in the study underwent BAL as standard of diagnosis. It may lead to a selection bias. In the further, we would like to conduct a randomized controlled trial to get results nearer to true circumstance. Second, the possible difference in MUC5B concentration between BALF and spontaneous sputum should be considered. Another issue is that we used...
immunoblotting to detect the concentration of MUC5B by antibody binding reaction, the accuracy of which might be affected by complex patterns of MUC5B glycosylation. These limitations will be overcome in future studies.

Conclusions
Respiratory MUC5B disproportion is related to CAP severity. MUC5B/total protein ratio is inversely correlated with the levels of inflammatory cytokines and may serve as an assessment marker and a potential therapeutic target for severe CAP.

Abbreviations
CAP: Community-acquired pneumonia; SCAP: Severe CAP; NSCAP: Non-severe CAP; COPD: Chronic obstructive pulmonary disease; MUC5B: Mucin 5B; BAL: Bronchoalveolar lavage fluid; ELISA: Enzyme-linked immunosorbent assay; ROC: Receiver operating characteristic; AUC: Area under the curve; ICU: Intensive care unit; CT: Computed tomography; EHR: Electronic hospital records; CURB-65: Confusion, urea level, respiratory rate, blood pressure, and age > 65 years; WBC: White blood cells; NE%: Percentage of neutrophils; LYM%: Percentage of lymphocytes; MO%: Percentage of monocytes; EOC%: Percentage of eosinophilic granulocytes; PLT: Blood platelets; LDH: Lactate dehydrogenase; CRP: C-reactive protein; ESR: Erythrocyte sedimentation rate; Fib: Fibrinogen; D-D: D-dimer; IL-6: Interleukin-6; IL10: Interleukin-10; IFN-γ: Interferon-γ; IL13: Interleukin-13; IQR: Interquartile range; CI: Confidence interval; OR: Odds ratio; r: Spearman rho correlation coefficients.

Acknowledgements
Not applicable.

Authors’ contributions
LF analysed the data, prepared the original draft. YL and YW recruited the subjects and did the BAL procedure. XZ and YW were responsible for data collection and validation. HS was responsible for conceptualization, methodology, formal analysis and validation. HS, LF and YW were responsible for study supervision and funding acquisition. All authors read and approved the final manuscript.

Funding
This work was supported by the National Natural Science Foundation of China (82072158), Jiangsu Province’s key provincial talents program (WSN-003).

Availability of data and materials
The datasets used and analysed during the current study are available from the corresponding author on reasonable request.

Declarations
Ethics approval and consent to participate
The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by Institutional Review Board of coordinating center Jiangsu Province Hospital (No. 2021-SR-028) and informed consent was taken from all individual participants.

Consent for publication
Informed consent was taken from all individual participants.

Competing interests
The authors declare that they have no competing interests.

Author details
1Department of Emergency, Jiangsu Province Hospital, The First Affiliated Hospital of Nanjing Medical University, 300 Guangzhou Rd, Nanjing 210029, People’s Republic of China. 2Department of Emergency, Northern Jiangsu People’s Hospital Affiliated to Yangzhou University, 98 Nantong West Rd, Yangzhou 225001, People’s Republic of China. 3Department of Respiratory Medicine, Qixia Branch of Jiangsu Province Hospital, 28 Yaoja Rd, Nanjing 210033, People’s Republic of China. 4Intensive Care Unit, Nanjing Chest Hospital, 215 Guangzhou Rd, Nanjing 210029, People’s Republic of China.

Received: 24 November 2021 Accepted: 23 February 2022

References
1. Welle T, Torres A, Nathwani D. Clinical and economic burden of community-acquired pneumonia among adults in Europe. Thorax. 2012;67(1):71–9.
2. Weiss AJ, Wier LM, Stocks C, Blanchard J. Overview of emergency department visits in the United States. Statistical Brief #174 (Healthcare Cost and Utilization Project [HCUP] Statistical Briefs [Internet]). 2011.
3. Woodhead M, Welch CA, Hamson DA, Bellingan G, Ayres JG. Community-acquired pneumonia on the intensive care unit: secondary analysis of 17,869 cases in the ICNARC Case Mix Programme Database. Crit Care. 2006;10(Suppl 1):S1.
4. Lanks CW, Musani AI, Hsia DW. Community-acquired pneumonia and hospital-acquired pneumonia. Med Clin North Am. 2019;103(3):487–501.
5. Christ-Crain M, Opal SM. Clinical review: the role of biomarkers in the diagnosis and management of community-acquired pneumonia. Crit Care. 2010;14(1):203.
6. Rautanen A, Mills TC, Gordon AC, Hutton P, Steffens M, Nau M, Zeman A, Chiche JD, Parks T, Chapman SJ, Davenport EE, Elliott KS, Bion J, Lichtner P, Meitinger T, Winken TF, Caulfield MJ, Mein C, Bloos F, Bobek I, Cotogni P, Stamm V, Saxapau S, Kobialy M, Ranieri VM, Rello J, Sogu K, Weiss YG, Rusuwum S, Schneider EM, Reinhart K, Holloway PA, Knight JC, Garraud CS, Russell JA, Wallay KR, Stuber F, Hill AV, Hinds CJ. Genome-wide association study of survival from sepsis due to pneumonia: an observational cohort study. Lancet Respir Med. 2015;3(1):S1–60.
7. Dekker J, Rossen JW, Buller HA, Einerich AW. The MUC family: an obituary. Trends Biochem Sci. 2002;27(3):126–31.
8. Rose MC, Veynow JA. Respiratory tract mucin genes and mucin glycoproteins in health and disease. Physiol Rev. 2006;86(1):245–78.
9. Roy MG, Livraghi-Butrico A, Fletcher AA, McElwee MM, Evans SE, Boerner RM, Alexander SN, Bellinghausen LK, Song AS, Petrova YM, Tuvim MJ, Adachi R, Romo I, Bordt AS, Bowden MG, Sisson JH, Woodruff PG, Thorn-Don J, Rousseau K, De la Garza MW, Moghaddam SJ, Karmouty-Quintana H, Blackburn MR, Drouin SM, Davis CW, Terrell KA, Grubb BR, O’Neal WK, Flores SC, Costa-Gomez A, Lozupone CA, Donnelly JM, Watson AM, Hennessey CE, Keith RC, Yang N, Barthel BM, Henson PM, Janssen WJ, Schwartz DA, Boucher RC, Dickey BF, Evans CM. MUC5B is required for airway defence. Nature. 2014;505(7483):412–6.
10. Sepper R, Prikk K, Metsis M, Sergejeva S, Pugatsjova N, Bragina O, Marran S, Fehniger TE. MUC5B expression by lung alveolar macrophages is increased in long-term smokers. J Leukoc Biol. 2012;92(2):319–24.
11. Ostedgaard LS, Möringer TQ, McMenimen JD, Savin NM, Parker CP, Thornell IM, Powers LS, Gasemier ND, Bouzek DC, Cook DP, Meyerholz DK, Abou AM, Stolz DA, Welsh MJ. Gel-forming mucins form distinct morphologic structures in airways. Proc Natl Acad Sci USA. 2017;114(26):E5842–7.
12. Seibold MA, Wise AL, Speer MC, Steele MP, Brown KK, Loyd JE, Fingerlin TE, Zhang W, Gudmundsson G, Groshong SD, Evans CM, Garantziotis S, Adler KB, Dickey BF, du Bois RM, Yang Y, Herron A, Kortvitsky D, Talbert JL, Markin C, Park J, Crevis AL, Slifer SH, Auerbach S, Roy MG, Lin J, Hennessy CE, Schwartz MI, Schwartz DA. A common MUC5B promoter polymorphism and pulmonary fibrosis. N Engl J Med. 2011;364(16):1503–12.
13. Kesimer M, Ford AA, Ceppe A, Radicioni G, Cao R, Davis CW, Doerschuk CM, Alexis NE, Anderson WH, Henderson AG, Bari RC, Bleecker ER, Christenson SA, Cooper CB, Han MK, Hanseil NN, Hastei AT, Hoffman EA, Kanner RE, Martinez F, Paine RR, Woodruff PG, O’Neal WK, Boucher RC. Airway
mucin concentration as a marker of chronic bronchitis. N Engl J Med. 2017;377(10):911–22.

14. Ramsey KA, Chen A, Radiccion I, Louie R, Martin M, Broomfield A, Sheng YH, Hasnain SZ, Radford-Smith G, Simms LA, Burni L, Thornton DJ, Bowler SD, Livengood S, Coppe A, Knowles MW, Noone PG, Donaldson SH, Hill DB, Ehr C, Button B, Alexis NE, Kesimer M, Boucher RC, McGuckin MA. Airway mucus hyperconcentration in non-cystic fibrosis bronchiectasis. Am J Respir Crit Care Med. 2020;201(6):661–70.

15. Wu X, Li Y, Zhang M, Li M, Zhang R, Lu X, Gao W, Li Q, Xia Y, Pan P, Li Q. Etiology of severe community-acquired pneumonia in adults based on metagenomic next-generation sequencing: a prospective multicenter study. Infect Dis Ther. 2019;8(4):1033–67.

16. Metlay JP, Waterer GW, Long AC, Anzueto A, Brozek J, Cohrs K, Cooley LA, Dean NC, Fine MJ, Flanders SA, Griffin MR, Metersky ML, Mush M, Restrepo MI, Whitney CG. Diagnosis and Treatment of Adults with Community-acquired Pneumonia. An official clinical practice guideline of the American Thoracic Society and Infectious Diseases Society of America. Am J Respir Crit Care Med. 2019;200(7):e45–67.

17. Niederman MS, Bass JJ, Campbell GD, Fein AI, Grossman RF, Mandell LA, Marinje T, Sarris GA, Torres A, Yu VL. Guidelines for the initial management of adults with community-acquired pneumonia: diagnosis, assessment of severity, and initial antimicrobial therapy. American Thoracic Society. Medical Section of the American Lung Association. Am Rev Respir Dis. 1992;145(5):1381–96.

18. Mandell LA, Wunderink RG, Anzueto A, Bartlett JG, Campbell GD, Dean NC, Dowell SF, File TM, Mush M, Niederman MS, Torres A, Whitney CG. Infectious Diseases Society of America/American Thoracic Society consensus guidelines on the management of community-acquired pneumonia in adults. Clin Infect Dis. 2007;44(suppl 2):S27–72.

19. Meyer KC, Raghu G, Baughman RP, Brown KK, Costabel U, du Bois RM, Drent M, Haslam PL, Kim DS, Nagai S, Rottoli P, Saltini C, Selman M, Strange C, Wood B. An official American Thoracic Society clinical practice guideline: the clinical utility of bronchoalveolar lavage cellular analysis in interstitial lung disease. Am J Respir Crit Care Med. 2012;185(9):1004–14.

20. Hellinger TP, Morris AC, McAuley DF, Walsh TS, Anderson NH, Singh S, Dark P, Roy AI, Baudouin SV, Wright SE, Perkins GE, Kefala K, Jeffers M, McMullan R, O’Kane CM, Spencer C, Laha S, Robin N, Gossain S, Gould K, Ruchaud-Sparaga MH, Scott J, Brownie EM, MacFarlane JG, Wiscombe S, Widdrington CE, Dimmick I, Laurensen IF, Nauwelaers F, Simpson AJ. Diagnostic accuracy of pulmonary host inflammatory mediators in the exclusion of ventilator-acquired pneumonia. Thorax. 2015;70(4):1–7.

21. Sengal S, Dhooon S, Choudhary H, Aggarwal AN, Garg M, Chakrabarti A, Agarwal R. Utility of serum and bronchoalveolar lavage fluid galactomannan in diagnosis of chronic pulmonary aspergillosis. J Clin Microbiol. 2019;57(3):e01821-e1918.

22. Monte AA, Sun H, Rapp-Olsson AM, Mohamed F, Gwarammana I, Buckley NA, Evans CM, Yang IV, Schwartz DA. The plasma concentration of MUC5B mannans in diagnosis of chronic pulmonary aspergillosis. J Clin Microbiol. 2019;57(3):e01821-e1918.

23. Inoue D, Yamaya M, Kubo H, Sasaki T, Hosoda M, Numasaki M, Tomioka Y, Rosenthal LA, Szakaly RJ, Amineva SP, Xing Y, Hill DB, Ehr C, Button B, Kesimer M, Hill DB, Sheehan JK, Boucher RC, McGuckin MA. Airway mucus hyperconcentration in non-cystic fibrosis bronchiectasis. Am J Respir Crit Care Med. 2020;201(6):661–70.

24. Hao Y, Kuang Z, Xu Y, Walling BE, Lau GW. Pyocyanin-induced mucin production is associated with redox modulation of FOXA2. Respir Res. 2013;14:82.

25. Hau Y, Kuang Z, Ju Y, Walling BE, Lau GW. Pyocyanin-induced mucin production is associated with redox modulation of FOXA2. Respir Res. 2013;14:82.

26. Mendez A, Rojas DA, Ponce CA, Basturmane R, Beltran CJ, Toledo J, Garcia-Angulo VA, Henriquez M, Vargas SL. Primary infection by Pneumocystis induces Notch-independent Clara cell mucin production in rat distal airways. PLoS ONE. 2019;14(6):e217684.

27. Rojas DA, Iturra PA, Mendez A, Ponce CA, Basturmane R, Gallo M, Borquez P, Vargas SL. Increase in secreted airway mucins and partial Muc5b STAT6/FoxA2 regulation during Pneumocystis primary infection. Sci Rep. 2019;9(1):2078.

28. Ermund A, Meiss LN, Rodriguez-Pineiro AM, Bahy A, Nilsson HE, Trillo-Muyo S, Ridley C, Thornton DJ, Wine JJ, Hebert H, Klymiuk N, Hansson GC. The normal trachea is cleaned by MUC5B mucin bundles from the submucosal glands coated with the Muc5b mucin. Biochim Biophys Acta. 2017;19(3):331–7.

29. White MR, Helmerhorst EJ, Ligtenberg A, Karpel M, Tickle T, Squire SA, Oppeheim FG, Hartshorn KL. Multiple components contribute to ability of saliva to inhibit influenza viruses. Oral Microbiol Immunol. 2009;24(1):18–24.

30. Janssen WJ, Stefanski AL, Bochner BS, Evans CM. Control of lung defense by mucins and macrophages: ancient defense mechanisms with modern functions. Eur Respir J. 2016;48(4):1201–14.

31. Kwak S, Choi YS, Na HG, Bae CH, Song SY, Kim YD. Glyoxal and methylglyoxal as E-cigarette vapor ingredient-induced pro-inflammatory cytokine and mucins expression in human nasal epithelial cells. Am J Rhinol Allergy. 2020;35(2):213–20.

32. Balkrishna A, Solleti SK, Singh H, Verma S, Sharma N, Nain P, Varshney A. Herbal decoction Divya–Swasari–Kwath attenuates airway inflammation and remodeling through Nrf-2 mediated antioxidant lung defense in mouse model of allergic asthma. Phytomedicine. 2020;78:153295.

33. Dienes Q, Rincon M.T. The effects of IL-6 on CD4 T cell responses. Clin Immunol. 2009;107(3):27–1.

34. Chen Y, Thi P, Zhao YH, Ho YS, DeSouza MM, Wu R. Stimulation of airway mucin gene expression by interleukin (IL)-17 through IL-6 paracrine/autoimmune loop. J Biol Chem. 2003;278(19):17036–43.

35. Iwakura Y, Ishigame H, Saijo S, Nakae S. Functional specialization of interleukin-17 family members. Immunol. 2011;142(2):149–62.

36. White MR, Helmerhorst EJ, Ligtenberg A, Karpel M, Tickle T, Squire SA, Oppeheim FG, Hartshorn KL. Multiple components contribute to ability of salivary glycoproteins during respiratory infections-relevance for SARS-CoV-2. MBio. 2020;11(6):e02374-e2420.

37. Lachowicz-Scroggins ME, Yuan S, Kerr SC, Dunican EM, Yu M, Carrington SD, Alcorn A, Olih Y, Khan H, Sea HW, Kim D, Kang J, Park C, Jang KY, Kim SH, Chae C. Expression of secreted mucins (MUC2, MUC5AC, MUC5B, and MUC6) and membrane-bound mucin (MUC4) in the lungs of pigs experimentally infected with Actinobacillus pleuropneumoniae. Res Vet Sci. 2021;129(3):486–91.

38. Mendez A, Rojas DA, Ponce CA, Basturmane R, Beltran CJ, Toledo J, Garcia-Angulo VA, Henriquez M, Vargas SL. Primary infection by Pneumocystis induces Notch-independent Clara cell mucin production in rat distal airways. PLoS ONE. 2019;14(6):e217684.

39. Rojas DA, Iturra PA, Mendez A, Ponce CA, Basturmane R, Gallo M, Borquez P, Vargas SL. Increase in secreted airway mucins and partial Muc5b STAT6/FoxA2 regulation during Pneumocystis primary infection. Sci Rep. 2019;9(1):2078.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.