Research Progress of Exosome-Loaded miRNA in Osteosarcoma

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Abstract
Currently, although the improvement of surgical techniques and the development of chemotherapy drugs have brought a certain degree of development to the treatment of osteosarcoma, the treatment of osteosarcoma has many shortcomings, and its treatment is limited. MiRNAs and exosomes can be used as diagnostic tools, and they play an important role in the occurrence and chemotherapy resistance of osteosarcoma. Therefore, providing a new method for the treatment of osteosarcoma is the key to solving this problem. To systematically summarize the research status of exosome drug-loaded miRNA in osteosarcoma, we identified and evaluated 208 studies and found that exosome-carrying miRNA can be used as an index for the diagnosis and prognosis of osteosarcoma and share a certain relationship with chemosensitivity. In addition, exosomes can also be used as a carrier of genetic drugs able to regulate the progression of osteosarcoma. Based on the above findings, we propose suggestions for the future development of this field, aiming to bring new ideas for the early diagnosis and treatment of osteosarcoma.

Keywords
exosomes, miRNA, osteosarcoma, chemotherapy, targeted therapy

Introduction
Osteosarcoma (OS) is a primary malignant tumor that occurs frequently among teenagers. Its clinical manifestations are mainly local pain and swelling. Currently, the main clinical treatment for OS is surgery combined with chemoradiotherapy, and first-line chemotherapeutic drugs used in OS treatment mainly include platinum, doxorubicin, and methotrexate.1 In addition, cyclophosphamide, bleomycin, and pingyangmycin have also been applied in the clinic.2 Although the application of these drugs has improved the prognosis of OS, the clinical outcomes of patients with OS are not ideal due to the widespread emergence of drug resistance. Therefore, solving the problem of OS drug resistance represents a main breakthrough in promoting the development of OS treatment.3 Moreover, explaining the mechanism of OS resistance is helpful for the implementation of individualized precise treatment strategies in OS, which will bring more hope to OS patients. Exosomes are vesicles with diameters of 40-100 nm that are secreted to the outside of cells after fusion of the cell membrane and the vesicles. Since its discovery in 1983, mechanisms defining the production, composition, transport, and delivery of exosomes have been widely reported, and increasing attention has been paid to its role as a carrier of endogenous information.3 The ab initio synthesis of exosomes is related to the endosome maturation pathway. Early endosomes can be formed by plasma membrane invagination, and the fusion of early endosomes or endoplasmic reticulum, Golgi budding vesicles, and early endosomes can provide content for early endosomes. Early endosomes (Rab5 as a marker) form late endosomes (Rab7 as a marker) through acidification and material-exchange maturation. Late endosomes can eventually form multivesicular bodies

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(MVBs), which contain intralocular vesicles (ILVs), which are formed by inward depression of the MVB membrane. Multivesicular and lysosomal or autophagic lysosomal fusion results in material degradation, and fusion with the plasma membrane leads to the secretion of lumen vesicles to the outside of the cell, which are exosomes. Exosomes have been identified in different types of body fluids (blood, lymph, urine, saliva, semen, and milk). They are considered an intercellular information transmitter because they carry cell-derived DNA, mRNA, miRNA, proteins, and lipids. Numerous studies have confirmed that exosomes from different sources play different roles in the treatment of diseases (Figure 1). In OS, exosomes are used as diagnostic markers, and have also been proposed as potential therapeutic targets due to their effects on the occurrence and progression of tumors.

Noncoding RNAs are a type of RNA that lack a protein-coding function, and mainly comprises housekeeping and regulating noncoding RNAs. Noncoding RNAs are found in exosomes and play a role in cell proliferation, differentiation, migration, and apoptosis by regulating gene expression through different targets. As a widely used regulator of noncoding RNAs, miRNAs play a regulatory role in many diseases. The expression of miRNA in OS cell lines or tissues is significantly different from that in normal or organisms, and the change in miRNA content can regulate the progression of OS by regulating the expression of downstream targets and activating signaling pathways.

Based on the above characteristics, whether exosomes can be used as miRNA carriers to regulate the occurrence and development of OS and be further applied to the clinical diagnosis and treatment of OS is the key focus of this review. Herein, the role of miRNA, exosomes, and exosomes loaded with miRNA are reviewed and prospectively analyzed in terms of their potential application for the diagnosis and prognosis, progression, and chemotherapy resistance in OS.

**Application of miRNA in OS Therapy**

**Effects of miRNA on proliferation, migration, invasion, and apoptosis of OS cells**

By transfection or knockout of miRNA in animal models, many researchers have found that changes in miRNA expression are closely related to the proliferation, migration, invasion, and apoptosis of OS cells. This suggests that miRNAs can regulate the proliferation, migration, invasion, and apoptosis of OS cells through changes in their expression. Its mechanism is to regulate its own expression by biding to the 3'-UTR of the target protein, and further regulating the downstream signaling pathway. Zhao et al. identified 190 miRNAs in OS cell lines or tissues, of which 82 miRNAs were upregulated and 102 miRNAs were downregulated. Herein, we list several miRNAs (Tables 1 and 2) that have been the

![Figure 1. Production and transport of exosomal miRNAs.](image-url)

After the completion of transcription, translation, and other expression activities, miRNAs are enclosed into exosomes in the cytoplasm and excreted by the cells, and are then transported to the target cells through the liquid system in the body where they exert their physiological activity.
focus of extensive research to elucidated the respective mechanisms, and elaborate on the activity of specific miRNA on several common targets or signaling pathways. The Wnt/β-catenin signaling pathway is a Wnt pathway. Upon activation, β-catenin enters the nucleus and binds to T cells to promote the upregulation of cell proliferation-related genes (such as survivin, c-myc, and cyclin D1) in OS, while apoptosis-related genes (such as BIM, Bax, Mcl-1, and Bcl-xL) are downregulated in OS, thereby altering the expression of HOXA9 and downregulates Wnt/β-Catenin signal transmission, thereby inhibiting cell proliferation and promoting cell apoptosis.1,7 Yu et al.16 found that miRNA-107 can regulate the Wnt/β-catenin signaling pathway and inhibits the proliferation and migration of OS cells in vitro.

The PI3K/Akt signaling pathway, activated by its upstream or bypass signals, acts on the downstream signaling, to promote cell growth and inhibit apoptosis, which is closely related to the occurrence and development of tumors. miRNA-21 can inhibit PTEN expression, activate the PI3K/Akt signaling pathway, and promote OS cell proliferation.11 MiRNA-133b inhibits the activation of the PI3K/Akt signaling pathway by targeting FGFR1, thus inhibiting the development of OS.38 MiRNA-144 can further inhibit the activation of related signaling pathways and induces apoptosis of OS cells by inhibiting the synthesis of mTOR protein.44

### Effects of miRNA on Chemoresistance of OS

At present, the clinical treatment of OS is still a combination of surgery and adjuvant radiotherapy and chemotherapy, but for patients with OS in the middle and late stages, chemotherapy is the only choice to extend overall survival.45 Therefore, the drug resistance of commonly used chemotherapy drugs has gradually become one of the biggest obstacles in OS treatment. Meanwhile, cisplatin (CDDP) and doxorubicin (DOX) are commonly used to treat OS, but CDDP resistance, DOX resistance, and multidrug resistance are very common.1

### Tables

#### Table 1. Upregulated miRNA.

| miRNA   | Target gene(s) | Effect(s)                                                   | References                        |
|---------|----------------|------------------------------------------------------------|-----------------------------------|
| miRNA-21| Caspase 8, PTEN | Inhibition of OS cell apoptosis; Promotion of OS cell       | Lv C et al.;11 Xu B et al.12       |
|         | UQCR1, β-catenin| proliferation and invasion                                 |                                   |
| miRNA-214| CADM1, TRAF3, PTEN, | Inhibition of OS cell apoptosis; Promotion of OS cell     | Zhao X et al.;6 Zhu XH et al.;     |
|         | UQCR1, PTEN, | proliferation and invasion                                 | Cai H et al.13; Rehei AL et al.14;|
| miRNA-221| CDKN1B, p27 | Inhibition of OS cell apoptosis; Promotion of OS cell       | Liu CJ et al.15; Zhu J et al.;16;  |
| miRNA-155| MAP3K10, NF-κB, RIPK1, | Inhibition of OS cell apoptosis; Promotion of OS cell     | Hu XH et al.17; Sun X et al.;9;    |
| miRNA-199a-5p| HBPI | proliferation and invasion                                 | Wang C et al.;18; Lu S et al.;19;  |
|          | PIAS3, p27, CD44| Inhibition of OS cell apoptosis; Promotion of OS cell       | Bhattacharya S et al.20; Wang C et al.;21; Gao Y et al.22 |

Notes: Five upregulated miRNAs in OS lines or tissues with clear research mechanisms and extensive research and their target genes, effects, and references.

#### Table 2. Downregulated miRNA.

| miRNA   | Target gene(s) | Effect(s)                                                   | References                        |
|---------|----------------|------------------------------------------------------------|-----------------------------------|
| miRNA-34a| Dusp1, sox-2 | Promote cell cycle and apoptosis of OS cells and inhibit cell | Gang L et al.;22; Zou Y et al.24   |
| miRNA-143| Bcl-2, MAPK7, | Promotion of OS cell apoptosis; Inhibition of OS cell cell  | Li WH et al.;25; Dong X et al.;26;  |
|         |cox-2, FOSL2, PAI-1 | proliferation, migration, and invasion; Increased risk of lung | Fang Y et al.;27; Sun X et al.;28;  |
| miRNA-124| TGF-β, SPHK1, Snail2, TRAF6, B7-H3, ROR2 | Promote apoptosis of OS cells; Inhibition of cell proliferation, migration, and invasion | Yu B et al.;30; Zhou et al.;31; Huang et al.;32; Meng et al.33; Wang et al.34; Zhang et al.35 |
| miRNA-133b| Sirt1, FOXC1, FGFR1 | Inhibition of OS cell proliferation, migration, and invasion | Ying et al.;36; eng et al.;37; Gao et al.38 |
| miRNA-150| IGF2BP1, ROCK1, ZEB1, Ezrin | Inhibition of OS cell proliferation, migration, and invasion; Prevention of rapid growth, early metastasis, and poor prognosis of OS cells | Qu et al.;39; Wang et al.;40; Li et al.;41; Xu et al.;42; Zhan et al.43 |

Notes: Five downregulated miRNAs in OS lines or tissues with clear research mechanisms and extensive research and their target genes, effects, and references.
Different types of miRNAs play different roles in the process of OS resistance.66,47 They can play a positive role in promoting OS resistance; although, some miRNAs can also have a negative role in inhibiting or even reversing OS resistance. The above effects are mainly mediated by regulating the expression of target proteins responsible for regulating the resistance of OS.47

MiRNA-34a is a marker RNA that promotes multidrug resistance in OS in vivo by inhibiting the expression of Dll1, CD117, and AGTR1.48,50 MiRNA-199a-3p targets the downregulation of AK4 expression, reducing the multidrug resistance of OS.51 MiRNA-143 increases OS sensitivity to DOX by downregulating Atg2b and Bcl-2 expression.52

Another important mechanism of miRNAs in OS resistance is the regulation of autophagy, which affects the sensitivity of OS to drugs. MiRNA-140-5p can upregulate HMGN5 and mediates autophagy by interacting with nucleosomes, thus promoting chemotherapy resistance.53 At the same time, it can inhibit autophagy mediated by IP3K2, reverse the induction of chemotherapy drugs, and enhance the resistance of OS.54

MiRNA-155 targets PTEN expression, enhances PI3K/Akt/mTOR signaling pathway activation, inhibits DOX-induced apoptosis and autophagy, and reduces OS sensitivity to DOX.55 MiRNA-199a-5p can inhibit CDDP-induced autophagy and it enhances drug resistance by downregulating the expression of Bcl1.56 In addition, a study by Li et al.,57 the authors knocked out miRNA-214 in order to determine whether miRNA-214 plays a role in the response to radiation therapy, and found that both in vivo and in vitro, miRNA-214 can enhance the radiosensitivity of OS, which is achieved by targeting PHLDA2 to inhibit the PI3K/Akt pathway. These studies indicated that, irrespective of anti-chemotherapy or anti-radiotherapy, the regulation of miRNA content can play a therapeutic role in OS.

Application of Exosomes in OS Therapy

Exosomes Secreted by Different Cells Regulate the Progress of OS in Different Ways

As an important carrier of information interaction in the tumor microenvironment (TME), exosomes can be produced and secreted by multiple cells.68 At present, some studies have clarified the mechanisms involving OS cells-associated with exosome transport to shape their supportive TME and promote growth and vascularization. In specific exosomes, miRNA-148a and miRNA-21 are known to help shape the TME.61,62 The TME is conducive to the proliferation and metastasis of OS cells and provides a foundation for the development of OS. Therefore, by regulating the production, secretion, and delivery of exosomes, tumor progression can be regulated by affecting the TME.63,64 At the same time, exploiting the natural targeting of exosomes in gene therapy65 and drug delivery66 seems to be a feasible scheme, which will revolutionize the development of OS treatment. Exosomes secreted from different cells have different functions that are activated through different regulatory pathways. There are three main regulatory approaches (Figure 2).

(i) Exosomes can directly regulate the proliferation and apoptosis of OS cells by regulating signaling pathways. HuBMSCs transfer exosomes to cancer cells. Exosomes derived from hBMSCs activate the Hedgehog signaling pathway in OS cell lines, thereby increasing the proliferation of OS cells and promoting OS growth.57

(ii) By changing the tumor microenvironment, bone resorption and tumor angiogenesis can be promoted, which in turn promotes OS. The tumor microenvironment plays an important role in tumorigenesis and tumor development. The main cell components and activities include fibroblasts and osteoclasts, and immune cell mesenchymal cells and neovascularization. In the development of OS, osteoclast proliferation enhances osteolysis and enhances the destruction of OS to bone. Tumor blood vessels provide the demand for rapid growth of the OS tissue.68 Raimondi et al.69 found that OS-derived exosomes promote osteoclast gene expression, induce osteoclast maturation, and enhance absorption activity. It can also stimulate the release of angiogenic factors, induce tumor angiogenesis, and promote the progression of OS.
(iii) OS progression is regulated by immune regulation. Troyer et al.\textsuperscript{70} isolated exosomes from dog OS cells and healthy osteoblasts, determined their protein components, and evaluated the direct impact of these two sources of exosomes on the dog immune system. It was found that the exosomes derived from OS contain protein components that inhibit immune function, including TGF-β, α-fetoprotein, and heat shock protein. These exosomes can directly reduce the rate of T cell proliferation, promote the differentiation of CD4+ cells into a T-regulated phenotype (Foxp3+), and reduce the expression of CD25+ on the surface of CD8+ cells, thereby inducing immunosuppression and promoting the immune escape of OS cells. Osteoblast exosomes can also reduce T cell activity, but they are not as marked as OS-derived exosomes and do not promote the T-regulatory phenotype due to the lack of TGF-β. Dong et al.\textsuperscript{71} found that osteosarcoma-derived exosomes can induce T cell proliferation and cytotoxic T cell responses by loading tumor-derived exosomes with dendritic cells \textit{in vitro}, thus inhibiting the growth of OS cells.

**Application of Exosomes Loaded miRNA in OS Therapy**

\textbf{miRNAs in Exosomes Can Be Used as Biomarkers for OS Diagnosis and Prognosis}

There are several free miRNAs in the blood of healthy and tumor patients. Many researchers collected blood samples from OS patients and healthy individuals and detected the content of miRNA in serum by PCR. It was found that 21 miRNAs in the serum of OS patients were altered, including 13 upregulated miRNA (i.e., miRNA-17,\textsuperscript{72} and 8 were downregulated (i.e., miRNA-195\textsuperscript{73}). Compared with healthy individuals, both upregulated and downregulated serum miRNAs are associated with tumor volume, distant metastasis, advanced clinical stage, short overall survival period, and poor prognosis. This suggests that serum miRNA can be used as a diagnostic and prognostic indicator of OS. However, the expression of miRNA in the serum is not stable, which raises additional research questions.

Given the lipid or lipoprotein complexes present in the blood circulation, apoptotic bodies, microbubbles, and exosomes can resist RNA degradation enzymes, and circulate stably exist in the blood.\textsuperscript{74} Fujiwara et al.\textsuperscript{75} purified exosomes from a number of OS cell lines (U2OS, HOS, 143 B, and SAOS2) and MSCs, and analyzed the expression of miRNAs by qRT-PCR. Both miRNA-17-5p and miRNA-25-3p were found to be enriched in exosomes and both were upregulated in OS cells. This indicated that miRNA-17-5p and miRNA-25-3p could not only exist stably in serum but could also be used as diagnostic biomarkers. Li et al.\textsuperscript{5} determined that miRNA-744-5p expression in the exosomes of OS patients was higher than that in healthy individuals. In addition, exosome miRNA is clinically important in predicting OS in patients with poor response to chemotherapy.\textsuperscript{76} This confirms our conjecture that in the future, the detection of miRNA content in exosomes may be a new biomarker for the diagnosis and prognosis of OS.

**Relationship Between Changes of miRNA in Exosomes and the Chemosensitivity of OS**

Xu et al.\textsuperscript{76} selected 31 healthy individuals as controls, 25 patients with good response to OS, and 28 patients with adverse reactions to OS. The expression of 746 miRNAs was analyzed using TaqMan miRNA array. The authors found that 12 miRNAs were upregulated and 18 miRNAs were downregulated in exosomes of OS patients with poor chemotherapy response compared with those of OS patients with good chemotherapy response. Further analysis showed that the levels of miRNA-124, miRNA-133a, miRNA-199a-3p, and miRNA-385 in serum exosomes of patients with adverse reactions to chemotherapy were significantly decreased, while the contents of miRNA-135b, miRNA-148a, miRNA-27a, and miRNA-9 were significantly increased. This suggested that the differential expression of miRNAs in exosomes was closely related to the adverse effects of chemotherapy. In addition, we also found that the differentially expressed miRNAs in the exosomes were enriched in different biological pathways, among which the Hippo signaling pathway, PI3K-Akt signaling pathway, Ras signaling pathway, ubiquitin mediated proteolysis, choline metabolism, and other biological pathways were highly correlated with differentially expressed miRNA.
Exosomes as Carrier of Gene Therapy in OS Therapy

MiRNA-208a in exosomes derived from BMSCs can enhance the activity of OS cells, promote clone formation and migration ability of OS cells, inhibit the expression of 3'-UTR by targeting PDCD4, and activate the ERK1/2 pathway, thereby enhancing the invasion of OS. Wang et al. identified 13 types of miRNAs using miRNA chip analysis and found that these miRNAs were significantly increased in exosomes derived from cancer-related fibroblasts (CAFs) and corresponding para-cancerous fibroblasts (PAFs). In addition, CAF can transfer miRNA-1228 to human OS cells through exosomes, promoting the proliferation and migration of OS, which is achieved by targeting the inhibition of SCAI expression. Instead, miRNA-675 in exosomes from metastatic OS promotes the proliferation and migration of stromal cells by downregulating CALN1. Moreover, tumor-derived exosomes act as messengers between metastatic cancer cells and stromal cells, and remodel the tumor microenvironment (TME) in OS by transferring miRNAs. In addition, miRNA-135b can be isolated from OS-derived exosomes. Therefore, if OS cells can capture the exosomes secreted by immune cells, we can inhibit the proliferation of OS cells by exploiting natural exosomes.

All of these studies suggest that exosome-loaded miRNAs can regulate OS progression by regulating OS proliferation, invasion, or drug resistance. Therefore, we can speculate that artificially overexpressing or inhibiting some miRNAs and using exosomes as carriers to target the proliferation or invasion of OS cells, may represent a potential approach to prevent the progression of OS. Shimbo et al. proved that our conjecture is feasible. They introduced a synthesized double-stranded miRNA-143 into BMSCs and found that the secretion of exosomes containing miRNA-143 increased and miRNA-143 was transferred to the outside of exosomes, which was then captured by OS cells, inhibiting their migration. This suggests that miRNA-143 in exosomes can be effectively transferred to receptor cells and to exert their activity. Therefore, exosomes can be used as carriers for gene therapy to regulate OS progression.

Discussion

Based on the in-depth review of current research, we believe that exosome-loaded miRNA not only can be used as a noninvasive biological marker for the diagnosis, prognosis of OS and the evaluation of resistance to chemotherapy, but these exosomes can also express human miRNAs. Further, these exosomes can be used as an important carrier of targeted therapy to achieve precise regulation of OS progression compared with treatment with miRNA or exosomes alone. Although there are few studies in this area, a novel approach to OS therapy may involve loading exosomes with miRNAs that can inhibit the growth of OS and enhance the sensitivity to chemotherapy drugs in the future.

Nonetheless, the following concerns should be addressed prior to the application of exosomes loaded with miRNAs for the treatment of OS.

1. Removing bioactive substances from exosomes that do not play a therapeutic role,
2. Determining whether ectopic expression and load have side effects and whether there are ethical concerns.
3. Ensuring the integrity of the carrier and miRNAs in the transfer mechanism
4. The results of in vitro and animal experiments can be clinically converted.

If the above four problems can be resolved, then exosomes loaded with miRNA will play a fundamental role in the treatment of OS and will provide additional possibilities for clinical treatment.

Conclusions

Through the evaluation of existing studies, it is easy to demonstrate that exosomes loaded with miRNA have great potential for the diagnosis, prognosis evaluation, and pre-assessment of chemotherapy resistance of OS. However, there are still many problems to be resolved for the application of exosomes as a targeted therapy carrier for OS; which requires more research to demonstrate the feasibility of this approach.

Appendix

Abbreviations

CAF cancer-related fibroblasts
CDDP cisplatin
DOX doxorubicin
EMT epithelial–mesenchymal transition
HuBMSCs human bone marrow mesenchymal stem cells
ILVs intralocular vesicles
MVBs multivesicular bodies

MRI magnetic resonance imaging
MSCs mesenchymal stem cells
OS osteosarcoma
PAF para-cancerous fibroblasts
PCR polymerase chain reaction
RT radiation therapy
TME tumor microenvironment

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References
1. Tang Z, Lu Y, Chen Y, Zhang J, Chen Z, Wang Q. Research
progress of MicroRNA in chemotherapy resistance of osteosarcoma.
Technol Cancer Res Treat. 2021;20:15330338211034262.
doi:10.1177/15330338211034262. PMID: 34323141.
2. Smrke A, Anderson PM, Gulia A, Gennatas S, Huang PH, Jones
RL. Future directions in the treatment of osteosarcoma. Cells.
2021;10(1):172. doi:10.3390/cells10010172.
3. Wortzel I, Dror S, Keni
4. Doyle LM, Wang MZ. Overview of Extracellular Vesicles, Their
Origin, Composition, Purpose, and Methods for Exosome
Isolation and Analysis. Cells. 2019;8(7):727. doi:10.3390/
cells8070727. PMID: 31311206.
5. Yao L. Experimental Study on the Regulation of Exosome
miRNA-744-5p on Osteosarcoma Cell Metastasis. Anhui Medical
University; 2019.
6. Zhao X, Wang Q, Lin F, et al. RNA Sequencing of Osteosarcoma
Gene Expression Profile Revealed that miR-214-3p Facilitates
Osteosarcoma Cell Proliferation via Targeting Ubiquinol-
Cytochrome c Reductase Core Protein 1 (UQCRC1). Med Sci
Monit. 2019;25:4982-4991. doi:10.12659/MSM.917375. PMID:
31276465; PMCID: PMC6626500.
7. Zhang ZF, Wang YJ, Fan SH, et al. MicroRNA-182 down-
regulates Wnt/β-catenin signaling, inhibits proliferation, and
promotes apoptosis in human osteosarcoma cells by targeting
HOXA9. Oncotarget. 2017;8(60):101345-101361. doi:10.18632/
oncotarget.21167. PMID: 29254169.
8. Zhu XB, Zhang ZC, Han GS, Han JZ, Qiu DP. Overexpression of
miR-214 promotes the progression of human osteosarcoma by
regulating the Wnt/β-catenin signaling pathway. Mol Med Rep.
2017;15(4):1884-1892. doi:10.3892/mmr.2017.6203. PMID:
28260089.
9. Sun X, Geng X, Zhang J, Zhao H, Liu Y. miR-155 promotes the
growth of osteosarcoma in a HBP1-dependent mechanism. Mol Cell
Biochem. 2015;403(1-2):139-147. doi:10.1007/s11010-015-2344-z.
PMID: 25666090.
10. Yu M, Guo D, Cao Z, Xiao L, Wang G. Inhibitory Effect of
MicroRNA-107 on Osteosarcoma Malignancy Through Regulation
of Wnt/β-catenin Signaling in Vitro. Cancer Invest. 2018;
36(3):175-184. doi:10.1080/07357907.2018.1439055. PMID:
29567602.
11. Lv C, Hao Y, Tu G. MicroRNA-21 promotes proliferation, in-
vasion and suppresses apoptosis in human osteosarcoma line
MG63 through PTEN/Akt pathway. Tumour Biol. 2016;37(7):
9333-9342. doi:10.1007/s13277-016-4807-6. PMID: 26779632.
12. Xu B, Xia H, Cao J, Wang Z, Yang Y, Lin Y. MicroRNA-21
Inhibits the Apoptosis of Osteosarcoma Cell Line SAOS-2 via
Targeting Caspase 8. Oncol Res. 2017;25(7):1161-1168. doi:
10.3727/096504017X14841698396829. PMID: 28109080; PMCID:
PMC7841250.
13. Cai H, Miao M, Wang Z. miR-214-3p promotes the prolifera-
tion, migration and invasion of osteosarcoma cells by targeting
CADM1. Oncol Lett. 2018;16(2):2620-2628. doi:10.3892/ol.
2018.8897. PMID: 30013657.
14. Rehei AL, Zhang L, Fu YX, et al. MicroRNA-214 functions as
an oncogene in human osteosarcoma by targeting TRAF3. Eur
Rev Med Pharmacol Sci. 2018;22(16):5156-5164. doi:10.
26355/eurrev_201808_15711.
15. Liu CJ, Yu KL, Liu GL, Tian DH. MiR-214 promotes osteo-
sarcoma tumor growth and metastasis by decreasing the ex-
pression of PTEN. Mol Med Rep. 2015;12(4):6261-6266. doi:
10.3892/mmr.2015.4197. PMID: 26252022.
16. Zhu J, Liu F, Wu Q, Liu X. MiR-221 increases osteosarcoma cell
proliferation, invasion and migration partly through the
downregulation of PTEN. Int J Mol Med. 2015;36(5):
1377-1383. doi:10.3892/ijmm.2015.2352. PMID: 26397386.
17. Hu XH, Zhao ZX, Dai J, Geng DC, Xu YZ. MicroRNA-221
regulates osteosarcoma cell proliferation, apoptosis, migration,
and invasion by targeting CDKN1B/p27. J Cell Biochem.
2019;120(3):4665-4674. doi:10.1002/jcb.27755. PMID:
30582227.
18. Wang C, Zhang X, Zhang C, Zhai F, Li Y, Huang Z. MicroRNA-
155 targets MAP3K10 and regulates osteosarcoma cell growth.
Pathol Res Pract. 2017;213(4):389-393. doi:10.1016/j.prp.
2016.12.028. PMID: 28214207.
19. Lu S, Liao QS, Tang L. MiR-155 affects osteosarcoma cell
proliferation and invasion through regulating NF-κB signaling
pathway. Eur Rev Med Pharmacol Sci. 2018;22(22):7633-7639.
doi:10.26355/eurrev_201811_16380.
20. Bhattacharya S, Chalk AM, Ng AJ, et al. Increased miR-155-5p
and reduced miR-148a-3p contribute to the suppression of
osteosarcoma cell death. Oncogene. 2016;35(40):5282-5294.
doi:10.1038/onc.2016.68. PMID: 27041566.
21. Wang C, Ba X, Guo Y, et al. MicroRNA-199a-5p promotes
tumour growth by dual-targeting PIAS3 and p27 in human
osteosarcoma. Sci Rep. 2017;7:41456. doi:10.1038/
rep41456.
22. Gao Y, Feng Y, Shen JK, et al. CD44 is a direct target of miR-199a-3p and contributes to aggressive progression in osteosarcoma. Sci Rep. 2015;5:11365. doi:10.1038/srep11365.

23. Gang L, Qun L, Liu WD, Li YS, Xu YZ, Yuan DT. MicroRNA-34a promotes microRNA-34a upregulation in human osteosarcoma stem-like cells and promotes invasion, tumorigenic ability and self-renewal capacity. Mol Med Rep. 2017;15(4):1631-1637. doi:10.3892/mmr.2017.6187. PMID: 28260055.

24. Li WH, Wu HJ, Li YX, Pan HG, Meng T, Wang X. MicroRNA-124 promotes apoptosis of osteosarcoma cells by caspase-3 activation via targeting Bcl-2. Biomed Pharmacother. 2016;80:8-15. doi:10.1016/j.biopharm.2016.03.001. PMID: 27133034.

25. Zhou Y, Han Y, Zhang Z, et al. MicroRNA-124 upregulation in osteosarcoma and migration in osteosarcoma. Express Ther Med. 2015;9(6):2374-2378. doi:10.3892/etm.2015.2420. PMID: 26136990.

26. Sun X, Dai G, Yu L, Hu Q, Chen J, Guo W. miR-143, 145 upregulation and migration in osteosarcoma by targeting FOSL2. Sci Rep. 2018;8(1):606. doi:10.1038/s41598-017-18739-3. PMID: 29387244; PMCID: PMC5768133.

27. Fang Y, Zhang Z, Wang Q, Zhao J. Expression and clinical significance of cyclooxygenase-2 and microRNA-143 in osteosarcoma. Exp Ther Med. 2015;9(6):2374-2378. doi:10.3892/etm.2015.2420. PMID: 26136990.

28. Sun X, Dai G, Yu L, Hu Q, Chen J, Guo W. miR-143-3p inhibits the proliferation, migration and invasion in osteosarcoma by targeting MAPK7. Arch Biochem Biophys. 2017;630:47-53. doi:10.1016/j.abb.2017.07.011. PMID: 28734729.

29. Hirahata M, Osaki M, Kanda Y, et al. PAI-1, a target gene of miR-143, regulates expression of Snail2 in osteosarcoma. J Exp Clin Cancer Res. 2016;35:73. doi:10.1186/s13046-016-0533-2. PMID: 27592320; PMCID: PMC5119899.

30. Yu B, Jiang K, Zhang J. MicroRNA-124 suppresses growth and aggressiveness of osteosarcoma and inhibits TGF-β-mediated AKT/GSK-3β/SNAIL-1 signaling. Mol Med Rep. 2018;17(5):6736-6744. doi:10.3892/mmr.2018.8637. PMID: 29488603.

31. Zhou Y, Han Y, Zhang Z, et al. MicroRNA-124 upregulation inhibits proliferation and invasion of osteosarcoma cells by targeting sphenosine kinase 1. Hum Cell. 2017;30(1):30-40. doi:10.1007/s13577-016-0148-4. PMID: 27743351.

32. Huang J, Liang Y, Xu M, Xiong J, Wang D, Ding Q. MicroRNA-22 regulates a tumor-suppressive miRNA by inhibiting the expression of Snail2 in osteosarcoma. Oncol Lett. 2018;15(4):4979-4987. doi:10.3892/ol.2018.7994. PMID: 29552134; PMCID: PMC5840501.

33. Meng Q, Zhang W, Xu X, et al. The effects of TRAF6 on proliferation, apoptosis and invasion in osteosarcoma are regulated by miR-124. Int J Mol Med. 2018;41(5):2968-2976. doi:10.3892/ijmm.2018.3458. PMID: 29436576.

34. Wang L, Kang FB, Sun N, et al. The tumor suppressor miR-124 inhibits cell proliferation and invasion by targeting B7-H3 in osteosarcoma. Tumour Biol. 2016;37(11):14939-14947. doi:10.1007/s13277-016-5386-2. PMID: 27644254.
48. Pu Y, Zhao F, Wang H, Cai S. MiR-34a-5p promotes multi-chemoresistance of osteosarcoma through down-regulation of the DLL1 gene. Sci Rep. 2017;7:44218. doi:10.1038/srep44218.

49. Pu Y, Zhao F, Wang H, et al. MiR-34a-5p promotes the multi-drug resistance of osteosarcoma by targeting the CD117 gene. Oncotarget. 2016;78(1936):28420-28434. doi:10.18632/oncotarget.8546. Erratum in: Oncotarget. 2017;8(36):60723. PMID: 27056900.

50. Pu Y, Zhao F, Li Y, et al. The miR-34a-5p promotes the multi-chemoresistance of osteosarcoma via repression of the AGTR1 gene. BMC Cancer. 2017;17(1):45. doi:10.1186/s12885-016-3002-x. PMID: 28073349.

51. Lei W, Yan C, Ya J, Yong D, Yujun B, Kai L. MiR-199a-3p affects the multi-chemoresistance of osteosarcoma through targeting AK4. BMC Cancer. 2018;18(1):631. doi:10.1186/s12885-018-4460-0. PMID: 29866054; PMCID: PMC5987492.

52. Zhou J, Wu S, Chen Y, et al. microRNA-143 is associated with the survival of ALDH1+CD133+ osteosarcoma cells and the chemoresistance of osteosarcoma. Exp Biol Med. 2015;240(7):867-875. doi:10.1177/1535370214563893. PMID: 25576341; PMCID: PMC4935406.

53. Meng Y, Gao R, Ma J, et al. MicroRNA-140-5p regulates osteosarcoma chemoresistance by targeting HMGN5 and autophagy. Sci Rep. 2017;7(1):416. doi:10.1038/s41598-017-00405-3. PMID: 28341864; PMCID: PMC5428500.

54. Wei R, Cao G, Deng Z, Su J, Cai L. miR-140-5p attenuates chemotherapeutic drug-induced cell death by regulating autophagy through inositol 1,4,5-trisphosphate kinase 2 (IP3K2) in human osteosarcoma cells. Biosci Rep. 2016;36(5):e00392. doi:10.1042/BSR20160238. PMID: 27582507.

55. Wang L, Tang B, Han H, et al. miR-155 Affects Osteosarcoma Cell Autophagy Induced by Adriamycin Through Regulating PTEN-PI3K/AKT/mTOR Signaling Pathway. Cancer Biother Radiopharm. 2021;36(7):614. doi:10.1089/cbr.2020.00392. PMID: 28817816.

56. Li S, Wang X. The potential roles of exosomal noncoding RNAs in osteosarcoma. J Cell Physiol. 2021;236(5):3354-3365. doi:10.1002/jcp.31001. PMID: 33044018.

57. Qi J, Zhou Y, Jiao Z, et al. Exosomes derived from human bone marrow mesenchymal stem cells promote tumor growth through hedgehog signaling pathway. Cell Biol Int. 2021;45(4):858-868. doi:10.1002/cbin.11532. PMID: 33325136.

58. Yang L, Huang X, Guo H, et al. Exosomes as efficient nanocarriers in osteosarcoma: Biological functions and potential clinical applications. Front Cell Dev Biol. 2021;9:737314. doi:10.3389/fcell.2021.737314. PMID: 34712664.

59. Lu T, Jia P, Song X, et al. Exosomal circ_103801-dependent manner. Exp Biol Med. 2020;245(5):666-677. doi:10.1093/carcin/bgz130.

60. Kok VC, Yu CC. Cancer-derived exosomes: Their role in cancer biology and biomarker development. Int J Nanomedicine. 2020;15:8019-8036. 10.2147/ijn.272378. PMID: 33116515.

61. Raimondi L, De Luca A, Gallo A, et al. Osteosarcoma cell-derived exosomes affect tumor microenvironment by specific packaging of microRNAs. Carcinogenesis. 2020;41(5):666-677. doi:10.1039/carcin/bg2130.

62. Wang S, Ma F, Feng Y, Liu T, He S. Role of exosomal miR-21 in the tumor microenvironment and osteosarcoma tumorigenesis and progression (Review). Int J Oncol. 2020;56(5):1055-1063. doi:10.3892/ijo.2020.4992. PMID: 32319566.

63. Biru B, Durhik CP, Kacham S, et al. Stem cells in tumour microenvironment aid in prolonged survival rate of cancer cells and developed drug resistance: Major challenge in osteosarcoma treatment. Curr Drug Metab. 2020;21(1):44-52. doi:10.2174/138920021666200214120226.

64. Li S, Wang X. The potential roles of exosomal noncoding RNAs in osteosarcoma. J Cell Physiol. 2021;236(5):3354-3365. doi:10.1002/jcp.31001. PMID: 33044018.

65. Pan Y, Lin Y, Mi C. Cisplatin-resistant osteosarcoma cell-derived exosomes confer cisplatin resistance to recipient cells in an exosomal circ 103801-dependent manner. Cell Biol Int. 2021;45(4):858-868. doi:10.1002/cbin.11532. PMID: 33325136.

66. Raimondi L, De Luca A, Gallo A, et al. Osteosarcoma cell-derived exosomes affect tumor microenvironment by specific packaging of microRNAs. Carcinogenesis. 2020;41(5):666-677. doi:10.1039/carcin/bg2130.

67. Luy N, Kong Y, Mu L, et al. Hepatic arterial infusion of oxaliplatin plus fluorouracil/leucovorin vs. sorafenib for advanced hepatocellular carcinoma. J Hepatol. 2018;69(1):60-69. doi:10.1016/j.jhep.2018.02.008. PMID: 29471013.

68. Raimondi L, De Luca A, Gallo A, et al. Osteosarcoma cell-derived exosomes affect tumor microenvironment by specific packaging of microRNAs. Carcinogenesis. 2020;41(5):666-677. doi:10.1039/carcin/bg2130.

69. Brady JV, Troyer RM, Ramsey SA, et al. A preliminary proteomic investigation of circulating exosomes and discovery of biomarkers associated with the progression of osteosarcoma in a clinical model of spontaneous disease. Transl Oncol. 2018;11(5):1137-1146. doi:10.1016/j.tranon.2018.07.004. Epub 2018 Jul 24. PMID: 30053712.
Neurosci. 2019;12:240. 10.3389/fnmol.2019.00240. PMID: 31636538.

75. Fujiwara T, Uotani K, Yoshida A, et al. Clinical significance of circulating miR-25-3p as a novel diagnostic and prognostic biomarker in osteosarcoma. Oncotarget. 2017;8(20):33375-33392. doi:10.18632/oncotarget.16498. PMID: 28380419.

76. Xu JF, Wang YP, Zhang SJ, et al. Exosomes containing differential expression of microRNA and mRNA in osteosarcoma that can predict response to chemotherapy. Oncotarget. 2017;8(44):75968-75978. doi:10.18632/oncotarget.18373. PMID: 29100284.

77. Qin F, Tang H, Zhang Y, Zhang Z, Huang P, Zhu J. Bone marrow-derived mesenchymal stem cell-derived exosomal microRNA-208a promotes osteosarcoma cell proliferation, migration, and invasion. J Cell Physiol. 2020;235(5):4734-4745. doi:10.1002/jcp.29351. PMID: 31637737.

78. Wang JW, Wu XF, Gu XJ, Jiang XH. Exosomal miR-1228 from cancer-associated fibroblasts promotes cell migration and invasion of osteosarcoma by directly targeting SCAI. Oncol Res. 2019;27(9):979-986. doi:10.3727/096504018X15336368805108. PMID: 30180920; PMCID: PMC7848259.

79. Gong L, Bao Q, Hu C, et al. Exosomal miR-675 from metastatic osteosarcoma promotes cell migration and invasion by targeting CALN1. Biochem Biophys Res Commun. 2018;500(2):170-176. doi: 10.1016/j.bbrc.2018.04.016. PMID: 29626470.

80. Shimbo K, Miyaki S, Ishitobi H, et al. Exosome-formed synthetic microRNA-143 is transferred to osteosarcoma cells and inhibits their migration. Biochem Biophys Res Commun. 2014;445(2):381-387. doi:10.1016/j.bbrc.2014.02.007. PMID: 24525123.