Antioxidant activity and anti-inflammatory activity of ethanol extract and fractions of Doenjang in LPS-stimulated RAW 264.7 macrophages

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BACKGROUND/OBJECTIVES: Fermentation can increase functional compounds in fermented soybean products, thereby improving antioxidant and/or anti-inflammatory activities. We investigated the changes in the contents of phenolics and isoflavones, antioxidant activity and anti-inflammatory activity of Doenjang during fermentation and aging.

MATERIALS/METHODS: Doenjiang was made by inoculating Aspergillus oryzae and Bacillus licheniformis in soybeans, fermenting and aging for 1, 3, 6, 8, and 12 months (D1, D3, D6, D8, and D12). Doenjang was extracted using ethanol, and sequentially fractioned by hexane, dichloromethane (DM), ethylacetate (EA), n-butanol, and water. The contents of total phenolics, flavonoids and isoflavones, 2,2-diphenyl-1 picryl hydrazyl (DPPH) radical scavenging activity, and ferric reducing antioxidant power (FRAP) were measured. Anti-inflammatory effects in terms of nitric oxide (NO), prostaglandin (PG) E2 and pro-inflammatory cytokine production and inducible nitric oxide synthase (iNOS) and cyclooxygenase (COX)-2 expressions were also measured using LPS-treated RAW 264.7 macrophages.

RESULTS: Total phenolic and flavonoid contents showed a gradual increase during fermentation and 6 months of aging and were sustained thereafter. DPPH radical scavenging activity and FRAP were increased by fermentation. FRAP was further increased by aging, but DPPH radical scavenging activity was not. Total isoflavone and glycoside contents decreased during fermentation and the aging process, while aglycone content and its proportion increased up to 3 or 6 months of aging and then showed a slow decrease. DM and EA fractions of Doenjang showed much higher total phenolic and flavonoid contents, and DPPH radical scavenging activity than the others. At 100 μg/mL, DM and EA fractions of D12 showed strongly suppressed NO production to 55.6% and 52.5% of control, respectively, and PGE2 production to 25.0% and 28.3% of control with inhibition of iNOS or COX-2 protein expression in macrophages.

CONCLUSIONS: Twelve month-aged Doenjang has potent antioxidant and anti-inflammatory activities with high levels of phenolics and isoflavone aglycones, and can be used as a beneficial food for human health.

INTRODUCTION

Soybean, an important plant protein source for human and animals, has been reported to have beneficial activities, including antiobesity, antdiabetic, anti-inflammatory, and anti-cardiovascular disease properties [1]. Fermentation is an excellent processing method for improving nutritional and biological properties of soybean. Previous studies have shown that fermented soybean foods contain higher level of bioactive compounds [2,3] and exhibit stronger anti-oxidative [4-7] and anticancer activities [8,9] than unfermented ones. These results have drawn attention to the potential preventive role of fermented soybean foods against many chronic diseases.

Doenjang, a Korean long-term fermented soybean paste, is usually consumed as an ingredient or seasoning in various sauces, soups, or stews. It is regarded as a good source of essential amino acids and fatty acids, particularly for people on a grain and vegetable based diet. Doenjang has been manufactured every year for centuries at homes in Korea using traditional methods, which employ Bacilli subtilis, Bacillus licheniformis, and molds such as Rhizopus, Mucor and Aspergillus oryzae from rice straw and local environments [10,11]. However, with rapid urbanization and industrialization of society, manufacture by inoculation of some major microorganisms is increasing with...
shorter fermentation time and aging under controlled conditions in a factory.

It was reported that that Doenjang intake resulted in significant reduction of body weight gain [12-14], visceral adipocyte content and circumference [12], serum oxidative stress and proinflammatory cytokine levels in obese rats fed a high fat diet [4,13]. Inflammation is a primary host defense reaction, and considered a beneficial and necessary attempt of the organism to eliminate the aggressive agent and to start the healing process against many pathological conditions. There is convincing evidence of a close association between proinflammatory stress and metabolic stress in obesity and aging-related chronic diseases such as atherosclerosis and insulin resistance [15]. When the inflammation control system does not function properly and chronic inflammation persists, development of diseases including cancer may occur [16,17]. Therefore, pharmacological control of the many aspects of inflammation continues to be a major issue in the development of new drugs [18].

Overproduction of reactive oxygen species (ROS) can cause oxidative damage to membrane lipids, DNA, proteins, and lipoproteins, eventually leading to development of chronic diseases including cancer, chronic inflammation, and cardiovascular diseases. ROS is a stimulator of the inflammatory response via activation of signaling pathways including nuclear factor kappa B (NF-κB) [19].

Most isoflavones in raw, cooked, or unfermented soybean exist in the glycoside form. However, cleavage of β-glucosyl bond of isoflavone glycoside by β-glucosidase released from rapidly growing microorganisms during the process of fermentation results in higher contents of isoflavone aglycones, including genistein and daidzein [20-23]. Genistein, a major isoflavone aglycone in fermented soybean products, is reported to have various biological functions such as preventive coronary heart disease and osteoporosis, as well as anti-oxidant, anti-neoplastic, and anti-inflammatory compounds [15,24]. In terms of anti-inflammatory activity, genistein significantly reduces the production of nitric oxide (NO) and prostaglandin (PG) E2 by suppression of inducible nitric oxide synthase (iNOS) and cyclooxygenase (COX)-2 expression at the transcriptional level in lipopolysaccharide (LPS)-treated RAW 264.7 macrophages [25]. Genistein also effectively inhibited the LPS-induced overproduction of proinflammatory cytokines such as tumor necrosis factor (TNF)-α and interleukin (IL)-6 in a dose dependent manner via inhibition of NF-κB activation following adenosine monophosphate-activated protein kinase (AMPK) stimulation [26].

However, few studies have reported on antioxidant and anti-inflammatory activities of Doenjang during different stages of aging. In this study, we investigated the changes in antioxidant and anti-inflammatory activities of Doenjang during aging.

MATERIALS AND METHODS

Sample preparation, extraction, and fraction

Doenjang was manufactured by SunchangJangryu Co., located in Sun Chang County in Jeonlabuk-do, Korea. The preparation method is shown in Fig. 1.

![Fig. 1. Manufacturing process of Doenjang](image)

For our experiment, parts of steamed soybean (SB), Meju (MJ), and Doenjang aged for 0, 1, 3, 6, 8, and 12 months (D0, D1, D3, D6, D8, and D12) were sampled. Samples were freeze-dried (Samwon, Sungnam, Korea), powdered, and stored at -20°C. A 100 g portion of each dried sample was extracted with ethanol (v/v) 10 times with stirring for 24 h. The supernatant was removed and filtered with Whatman paper (No. 2), and the residue was extracted once more with ethanol and filtered. Two filtrates were combined, and evaporated for removal of ethanol using a rotary vacuum evaporator (EYELA, Tokyo, Japan), then concentrated ethanol extract was dissolved in distilled water and sequentially fractionated with hexane (Hx), dichloromethane (DM), ethylacetate (EA), and n-butanol (BT), leaving an aqueous layer (DW). All fractions were also concentrated, freeze-dried, dissolved in DMSO or distilled water at 100 mg/mL, and stored at -20°C.

Determination of total phenolic and flavonoid contents

For determination of total phenolic and flavonoid contents, 1.0 g of each dried sample powder was stirred in 50 mL of 80% ethanol for 24 h at room temperature and filtered with Whatman paper (No.2). Total phenolic concentration in the filtrate was determined according to Singleton et al. [27]. Briefly, 100 μL of samples was mixed with 1 mL of 0.2 N Folin-Ciocalteau reagent and 200 μL of 20% sodium carbonate, and stood in the dark for 1 h. The absorbance was read at 760 nm by spectrophotometer using tannic acid as a standard. Total flavonoid content was determined according to the modified AOAC method [28]. Briefly, 100 μL of samples was mixed with 900 μL of 90% diethylene glycol and then 20 μL of 1 N NaOH, and incubated in a 37°C water bath for 1 h. The absorbance was read at 420 nm by spectrophotometer using rutin as a standard.

Isoflavone analysis

Each ethanol extract was dissolved in 70% ethanol, centrifuged at 10,000 × g for 5 min, and the supernatant was filtered with
a 0.45 μm membrane (PVDF syringe filter, Whatman, Germany). Reversed-phase HPLC system (JASCO, Japan) with an ODS A303 column (25 μm, 4.6 × 250 mm, YMC, USA) was used for isoflavone analysis. A linear HPLC gradient was composed of (A) 0.1% acetic acid in acetonitrile and (B) 0.1% acetic acid in water. The solvent flow rate was 1.0 mL/min. Following injection of 20 μL of sample, eluted isoflavones were detected by the absorbance at 254 nm on a UV detector. Quantitative data for daidzin, glycitin, and genistin and their malonyl, acetyl, and acetylglucoside forms were obtained by comparison with 12 known standards. Isoflavone standards were obtained as follows: genistin, daidzein, and glycitein from Sigma (St. Louis, MO, USA), genistin, daidzin, and glycitin from Indofine (Hillsborough, NJ, USA), and malonylgenistein, malonyldaidzin, malonylglucit, acetylglycitein, acetyldaidzin, and acetylglycitin from LC Labs (Woburn, MA, USA). All data are presented as mean of two measurements.

Measurement of 2,2-diphenyl-1-picryl-hydrazyl (DPPH) radical scavenging activity

DPPH radical scavenging activity was determined as previously described [29]. The various concentrations of each sample in 10 μL were mixed with 190 μL of 200 μM DPPH in ethanol, incubated at 37°C for 30 min, and their absorbance was read at 517 nm by spectrophotometer. The reduction in absorbance is proportional to the ability of the sample to eliminate DPPH radicals. Ascorbic acid was used as a positive control. DPPH radical scavenging effect was calculated as follows; DPPH radical scavenging activity (%) = [(Acontrol - Asample)/Acontrol] × 100. IC50 value, the sample concentration needed to reduce 50% of DPPH radicals, was calculated from the equation between sample concentration and DPPH scavenging activity.

Measurement of Ferric reducing antioxidant power (FRAP)

The method described by Yildirim et al. [30] was used for determination of FRAP. Various concentrations of sample in 0.2 M phosphate buffer (pH 6.6), 1% potassium ferricyanide were incubated in a water bath at 50°C for 30 min, followed by addition of 10% trichloroacetic acid. Following centrifugation of solution at 3000 rpm for 10 min, supernatant was collected, mixed with 0.1% FeCl3 and the absorbance read at 700 nm by spectrophotometer. The result was expressed as μg/mL of ascorbic acid, positive control.

Cell culture and sample preparation

RAW 264.7 murine macrophage cells were cultured in DMEM and supplemented with 10% FBS, 2 mM glutamine, 100 unit/mL penicillin, and 50 μg/mL streptomycin at 37°C in a cell incubator of 5% CO2. Fractionated samples were dissolved in DMSO (100 mg/mL), filtered by sterile 0.2 μm syringe filter, and stored at -20°C.

Cell viability

Cell viability was determined by MTT assay [31]. RAW 264.7 cells were seeded in 96-well plate at a density of 5 × 103 cells/well. Following 24 h incubation in 10% FBS containing DMEM in a cell incubator, culture medium was replaced with serum-free medium including various concentrations of sample. Following further 24 h incubation, medium was replaced with 180 μL of serum-free and 20 μL of 5 mg/mL MTT solution. Following 4 h incubation at 37°C, the supernatant was discarded and 200 μL of DMSO was added to dissolve the formazan crystals formed in the cells. The absorbance was read at 540 nm using a microplate reader (Bio-Tech Instrument, Winoski, VM, USA). The results were expressed as percentage of control.

Measurement of NO, PGE2, IL-6, and TNF-α production

RAW 264.7 macrophage cells were seeded in 12-well plate at a density of 5 × 105 cells/well in 2 mL of DMEM containing 10% FBS, and incubated in a cell incubator for 24 h. Medium was replaced with 1% FBS and phenol red-free medium including various concentrations of sample, incubated for 2 h, and then LPS (1 μg/mL) was applied except negative control. Following further 22 h incubation, NO, PGE2, IL-6, and TNF-α concentrations were measured in cultured medium. NO concentration was measured by Griess reaction [32]. Each 100 μL of cultured medium was mixed with 100 μL of Griess reagent and incubated at room temperature. After 20 min, the absorbance was read at 540 nm using an ELISA plate reader. NO concentration was calculated from a standard curve drawn with known concentrations of sodium nitrite. PGE2, IL-6, and TNF-α concentrations were determined using ELISA kits (R&D Systems, Minneapolis, MN, USA) according to the manufacturer’s instructions.

Western blot analysis

Cells were washed with PBS and lysed with RIPA buffer (Sigma, Saint Louis, MS, USA) and protease and phosphatase inhibitor cocktail (Roche, Germany) on ice for 20 min. The cell debris was pelleted by centrifugation at 10,000 g for 15 min. The protein level of the supernatant was determined using Bradford method [33]. Total cell lysates were resolved in sodium dodecyl sulfate (SDS) sample buffer (Sigma, Saint Louis, MS, USA), separated by electrophoresis on 10% SDS polyacrylamide gel in the Mini-PROTEIN system (Bio-Rad) and transferred onto Immobilon membranes (Millipore, Bedford, MA). The membranes were blocked with 5% nonfat dry milk in Tris-buffered saline containing 0.1% Tween-20 (TBST, pH 8.0) at room temperature for 1 h, and probed with rabbit polyclonal antibodies against iNOS, COX-2, or anti-β-actin monoclonal antibody (Cell signaling, Technology Inc, OR, USA) diluted in TBST with 5% nonfat milk with overnight shaking at 4°C. After washing with TBST, the membranes were treated with secondary antibody (horseradish peroxidase-labeled IgG, 1:1000-1:2000) for 1 h at room temperature and washed. Protein bands were visualized by ECL solution (Pierce, Rockford, IL) and quantified using Luminescent Image Analyzer (Fujifilm, Japan).

Statistical analysis

Data were expressed as mean ± SD and analyzed for significance by t-test, ANOVA test and Duncan multiple-comparison test, and Pearson correlation test using SAS (ver 9.4) program. Differences were considered significant when the P values were < 0.05.
RESULTS

Contents of total phenolics and flavonoids in dried Doenjang

Contents of total phenolics and flavonoids in freeze-dried SB, MJ, and Doenjang are shown in Table 1. Total phenolic content was significantly increased by fermentation, and further increased by aging (P < 0.05). Different from total polyphenol content, total flavonoid contents in SB were not significantly changed by dry fermentation, slightly dropped in D0 and D1, and then recovered after 6 months of aging.

Antioxidant activity of ethanol extract

DPPH radical is a stable free-radical donor widely used for estimation of free-radical scavenging activity. It is reduced in the presence of an antioxidant molecule, producing a color change from violet to yellow. DPPH radical scavenging activities (IC50) of the samples are shown in Table 1. DPPH radical scavenging activity was rapidly increased (P < 0.05) by the fermentation and 1 month of aging, however, it was not significantly increased after 1 month of aging. IC50 was decreased from 21.0 mg/mL in SB to 15.5 mg/mL in MJ, 4.5 mg/mL in D1 and 3.9 mg/mL in D12. Total phenolic content (r = 0.8772, P < 0.001), aglycone isoflavone content (r = 0.9195, P < 0.001), and glycoside isoflavone content (r = 0.9595, P < 0.001) showed significant correlation with IC50 value for DPPH radicals, while no significant correlation was observed between total flavonoid content and DPPH radical scavenging activity (data not shown).

FRAP was determined at various concentrations of ethanol extract, 500, 1000, and 2000 μg/mL (Table 2). At all concentrations, FRAP in SB and MJ did not differ from each other, indicating that FRAP was not increased by dry fermentation. However, FRAP was rapidly increased by wet fermentation and aging (P < 0.05). At 2000 μg/mL, FRAP in MJ, 10.2 μg AA eq/mL, increased up to 19.9 μg AA eq/mL in D0, 32.1 μg AA eq/mL in D1 and 53.5 μg AA eq/mL in D12.

Total phenolic and flavonoid contents and antioxidant activity in different fractions

Total phenolic contents in DM and EA fractions were 330.35 mg TA/mg and 269.24 mg TA/mg, respectively, which were significantly higher than those in Hx, BT, and DW fractions (P < 0.05). Total flavonoid contents in DM and EA were 32.69 mg RT/mg and 32.02 mg RT/mg, respectively, which were also significantly higher than those in the other fractions (P < 0.05) (Table 3).

DPPH radical scavenging activities of DM and EA fractions were significantly higher (P < 0.05) than those of the other fractions. At 100 and 200 μg/mL, DPPH radical scavenging rate was 59.3% and 88.1% in DM fraction, and 80.9% and 94.9% in EA fraction, respectively (Table 3).

Isoflavone content

Three major isoflavone contents in samples are shown in Table 4. Total isoflavone content of diazzein, glycitein, and genistein showed a gradual decrease during fermentation and the aging process. Overall, in examination of the isoflavone composition, aglycone content and its proportion were increased by fermentation and aging. In detail, however, aglycone content peaked in D3, and its proportion peaked in D6, and then both of them showed a slight decrease until 12 months of aging.

Table 1. Changes in the contents of total phenolics and flavonoids and DPPH radical scavenging activity of Doenjang during fermentation and aging

| Total phenolics (mg TA/1/g of dry wt) | Total flavonoids (mg RT/1/g of dry wt) | DPPH radical scavenging activity (IC50) |
|--------------------------------------|---------------------------------------|----------------------------------------|
| SB | MJ | D0 | D1 | D3 | D6 | D8 | D12 |
| 3.07 ± 0.05 | 6.04 ± 0.09 | 7.05 ± 0.09 | 7.95 ± 0.09 | 14.61 ± 0.19 | 13.34 ± 0.12 | 13.34 ± 0.13 | 13.34 ± 0.15 |
| 1.00 ± 0.14 | 0.84 ± 0.10 | 0.66 ± 0.14 | 0.64 ± 0.16 | 0.77 ± 0.05 | 0.91 ± 0.06 | 0.86 ± 0.10 | 0.90 ± 0.06 |
| 4,262.4 ± 478.2 | 3,248.9 ± 306.5 | 2,207.8 ± 258.9 | 1,339.4 ± 70.2 | 1,264.2 ± 118.2 | 1,230.6 ± 87.9 | 1,152.9 ± 13.9 | 1,009.9 ± 6.5 |
| 2.9 ± 0.3 |

Data are presented as the mean ± SD (n = 3). SB: Steamed soybeans, MJ: Meju, D0: Doenjang without aging, D1: 1 month-aged Doenjang, D3: 3 month-aged Doenjang, D6: 6 month-aged Doenjang, D8: 8 month-aged Doenjang, D12: 12 month-aged Doenjang, AA: ascorbic acid, positive control
1) Tannic acid; 2) Rutin; 3) Concentration to reduce the initial DPPH concentration by 50%; 4) Means sharing the same alphabet in superscripts are not significantly different within each column at P < 0.05 by ANOVA and Duncan’s multiple range test.

Table 2. Changes in ferric reducing antioxidant power of ethanol extract from Doenjang during fermentation and aging (μg AA eq/mL)

| Concentration of ethanol extract (μg/mL) | 500 | 1000 | 2000 |
|-----------------------------------------|----|-----|-----|
| SB | 0.8 ± 1.2 | 3.1 ± 1.6 | 8.5 ± 1.6 |
| MJ | 1.5 ± 1.6 | 4.8 ± 1.2 | 10.2 ± 0.9 |
| D0 | 5.0 ± 0.6 | 9.4 ± 1.0 | 19.9 ± 1.2 |
| D1 | 8.3 ± 1.0 | 15.8 ± 1.6 | 32.1 ± 2.1 |
| D3 | 9.1 ± 0.4 | 18.1 ± 0.6 | 37.4 ± 1.4 |
| D6 | 8.7 ± 1.3 | 16.9 ± 1.7 | 35.4 ± 2.7 |
| D8 | 11.0 ± 1.6 | 21.8 ± 1.6 | 44.1 ± 2.7 |
| D12 | 13.1 ± 1.5 | 26.5 ± 2.2 | 53.5 ± 4.3 |

Data are presented as the mean ± SD (n = 3). SB: Steamed soybeans, MJ: Meju, D0: Doenjang without aging, D1: 1 month-aged Doenjang, D3: 3 month-aged Doenjang, D6: 6 month-aged Doenjang, D8: 8 month-aged Doenjang, D12: 12 month-aged Doenjang, AA: ascorbic acid, positive control
1) Means sharing the same alphabet in superscripts are not significantly different within each column at P < 0.05 by ANOVA and Duncan’s multiple range test.

Table 3. Contents of total phenolics and flavonoids, and DPPH radical scavenging activity in fractions of ethanol extract from Doenjang aged for 12 months

| Fractions | Total phenolics (μg TA/1/mg) | Total flavonoids (μg RT/1/mg) | DPPH radical scavenging activity (%) |
|-----------|-----------------------------|-------------------------------|--------------------------------------|
| Hx4 | 36.23 ± 0.47 | 0.75 ± 0.03 | 4.5 ± 0.4 |
| DM5 | 330.35 ± 7.5 | 32.69 ± 1.26 | 59.3 ± 1.4 |
| EA6 | 269.24 ± 1.63 | 32.02 ± 0.25 | 80.9 ± 2.1 |
| BT7 | 57.86 ± 1.04 | 3.17 ± 0.03 | 15.3 ± 1.1 |
| DW8 | 685.73 ± 0.37 | 15.0 ± 0.03 | 6.3 ± 0.5 |

Data are presented as the mean ± SD (n = 3).
1) Tannic acid; 2) Rutin; 3) Hexone fraction; 4) Dichloromethane fraction; 5) Ethylacetate fraction; 6) Butanol fraction; 7) Aqueous fraction; 8) Means sharing the same alphabet in superscripts are not significantly different within each column at P < 0.05 by ANOVA and Duncan’s multiple range test.
Table 4. Changes in the contents of three major isoflavone glycosides and aglycones of Doenjang during fermentation and aging

|                | SB      | MJ      | D0      | D1      | D3      | D6      | D8      | D12     |
|----------------|---------|---------|---------|---------|---------|---------|---------|---------|
| Glucoside      | 2.88 (96.7) | 1.14 (78.1) | 0.47 (38.2) | 0.26 (22.6) | 0.05 (4.4) | 0.03 (2.8) | 0.04 (4.0) | 0.04 (5.4) |
| β-glucoside    | 1.14 (78.1) | 0.47 (38.2) | 0.26 (22.6) | 0.05 (4.4) | 0.03 (2.8) | 0.04 (4.0) | 0.04 (5.4) | 0.04 (5.4) |
| Daidzin        | 0.42     | 0.27     | 0.10     | 0.03     | ND       | ND       | ND       | ND       |
| Glycitin       | 0.20     | 0.11     | 0.06     | 0.03     | ND       | ND       | ND       | ND       |
| Genistin       | 0.52     | 0.36     | 0.2      | 0.07     | ND       | ND       | ND       | ND       |
| malonylgucoside| 1.60     | 0.34     | 0.02     | 0.08     | 0        | 0        | 0        | 0        |
| daidzin        | 0.53     | 0.10     | 0.02     | 0.01     | ND       | ND       | ND       | ND       |
| glycitin       | 0.17     | 0.02     | ND       | ND       | ND       | ND       | ND       | ND       |
| genistin       | 0.09     | 0.22     | ND       | ND       | ND       | ND       | ND       | ND       |
| acetylglucoside| 0.14     | 0.06     | 0.09     | 0.05     | 0.05     | 0.03     | 0.04     | 0.04     |
| daidzin        | 0.06     | 0.03     | 0.04     | 0.04     | 0.03     | 0.03     | 0.03     | 0.04     |
| glycitin       | 0.01     | ND       | 0.04     | ND       | 0.01     | ND       | 0.01     | ND       |
| genistin       | 0.07     | 0.03     | 0.01     | 0.01     | 0.01     | ND       | ND       | ND       |
| Aglycone       | 0.13 (4.3) | 0.32 (21.9) | 0.76 (61.8) | 0.89 (77.4) | 1.08 (95.6) | 1.06 (97.2) | 0.95 (96.0) | 0.70 (94.6) |
| Daidzein       | 0.05     | 0.13     | 0.31     | 0.34     | 0.37     | 0.37     | 0.33     | 0.27     |
| Glycitein      | 0.02     | 0.07     | 0.15     | 0.17     | 0.20     | 0.20     | 0.17     | 0.11     |
| Genistein      | 0.06     | 0.12     | 0.30     | 0.38     | 0.51     | 0.49     | 0.45     | 0.32     |

Data are presented as mean of results from two independent experiments.
SB: Steamed soybeans, MJ: Meju, D0: Doenjang without aging, D1: 1 month-aged Doenjang, D3: 3 month-aged Doenjang, D6: 6 month-aged Doenjang, D8: 8 month-aged Doenjang, D12: 12 month-aged Doenjang; # percentage (%) of total isoflavone content, ND: not detected.

while glycoside content and its proportion were gradually decreased until 6 months of aging and then showed a slight increase until 12 months of aging.

Effect of DM and EA fractions of Doenjang on NO and PGE2 production in RAW 264.7 macrophages

DM and EA fractions of SB, D0, D6, and D12 did not show cytotoxicity to RAW 264.7 cells at the treated concentrations (10, 25, 50, and 100 μg/mL) as shown in Fig. 2. To investigate potential anti-inflammatory activity of Doenjang, the RAW 264.7 macrophages, which produce NO and PGE2 upon stimulation with LPS, were treated with DM and EA fractions of steamed soybeans and Doenjang samples (D0, D6, and D12), and the accumulated NO and PGE2 concentrations in cultured medium were measured. LPS (1 μg/mL) treatment resulted in significantly increased production of NO and PGE2, as shown in Fig. 3.

DM fractions of SB and D0 did not suppress the NO concentration at all treated concentrations. However, DM fraction of D6 significantly suppressed (P < 0.05) the NO concentration to 88.8% of LPS-treated control at 100 μg/mL, and DM fraction of D12 significantly suppressed (P < 0.05) the NO concentration to 83.3% and 55.6% of LPS-treated control at 50 and 100 μg/mL, respectively. DM fraction of SB did not influence the PGE2 concentration, while DM fraction of D0, D6, and D12 significantly suppressed (P < 0.05) it at 50 and 100 μg/mL in a dose dependent manner. At the highest concentration (100 μg/mL), DM fractions of D0, D6, and D12 suppressed the PGE2 concentration to 71.0%, 62.9%, and 25.0% of LPS-treated control, respectively.

EA fractions of SB and D0 did not suppress the NO concentration. However, EA fraction of D6 significantly suppressed (P < 0.05) the NO concentration to 89.3% and 84.6% of LPS-treated control at 50 and 100 μg/mL, respectively, and EA fraction of D12 significantly suppressed (P < 0.001) it to 74.3%...
and 52.5% of LPS-treated control at 50 and 100 μg/mL, respectively. EA fraction of SB did not influence the PGE2 concentration, while EA fractions of D0 and D6 significantly suppressed ($P < 0.05$) it at 100 μg/mL and EA fraction of D12 significantly suppressed ($P < 0.001$) it at 50 and 100 μg/mL. At 100 μg/mL, EA fractions of D0, D6, or D12 suppressed the PGE2 concentration to 68.7%, 74.4%, and 28.3% of LPS-treated control, respectively. DM and EA fractions of D12 commonly

Fig. 3. Effect of dichloromethane (DM) and ethylacetate (EA) fractions of ethanol extract from steamed soybeans and Doenjang on NO and PGE2 synthesis in LPS-treated RAW 264.7 macrophages. Each bar represents the mean ± SD (n = 3). Cells were treated with different concentrations of DM or EA fraction of sample extract for 2 h, followed by addition of LPS (1 μg/mL) and incubation for 22 h. NO (A) and PGE2 (B) concentrations in cultured medium were measured. SB: Steamed soybeans, D0: Doenjang without aging, D6: 6 month-aged Doenjang, D12: 12 month-aged Doenjang, NS: not significant. Means sharing the same alphabet in superscript within each sample are not significantly different at $P < 0.05$ by ANOVA and Duncan’s multiple range test.

Fig. 4. Effect of dichloromethane (DM) and ethylacetate (EA) fractions of ethanol extract from steamed soybeans and Doenjang on IL-6 and TNF-α synthesis in LPS-treated RAW 264.7 macrophages. Each bar represents the mean ± SD (n = 3). Cells were treated with different concentrations of DM or EA fraction of sample extract for 2 h, followed by addition of LPS (1 μg/mL) and incubation for 22 h. IL-6 (A) and TNF-α (B) concentrations in cultured medium were measured. SB: Steamed soybeans, D0: Doenjang without aging, D6: 6 month-aged Doenjang, D12: 12 month-aged Doenjang, Means sharing the same alphabet in superscript within each sample are not significantly different at $P < 0.05$ by ANOVA and Duncan’s multiple range test.
Effects of dichloromethane (DM) and ethylacetate (EA) fractions of ethanol extract from steamed soybeans and Doenjang on iNOS and COX-2 synthesis in LPS-treated RAW 264.7 macrophage cells. (A) Cells were treated with 50 or 100 µg/mL of DM or EA fraction of sample extract for 2 h, followed by addition of LPS (1 µg/mL) and incubation for 22 h. Cells were lysed and equal amounts of cellular proteins (80 µg/well) were resolved by SDS-PAGE and subjected to Western blotting. The proteins were visualized using iNOS and COX-2 antibodies and ECL detection system. (B) The visualized bands were quantified by densitometry. Each bar represents the mean ± SD (n = 3). * significantly different from LPS-treated control at P < 0.05 by ANOVA and Duncan’s multiple range test. SB: Steamed soybeans, D12:12 month-aged Doenjang showed significantly higher (P < 0.05) suppression effects on NO and PGE2 production at 100 µg/mL than those of SB, D0, and D6.

Effect of DM and EA fractions of Doenjang on IL-6 and TNF-α production in RAW 264.7 macrophages

To investigate potential anti-inflammatory activity of Doenjang, the accumulated IL-6 and TNF-α concentration in cultured medium was also measured. LPS (1 µg/mL) treatment resulted in significantly increased production of IL-6 and TNF-α, as shown in Fig. 4.

DM fractions of SB, D0, and D6 did not suppress the IL-6 and TNF-α concentration at all treated concentrations, while DM fraction of D12 significantly suppressed (P < 0.05) the IL-6 and TNF-α concentrations to 70.6% and 86.9% of the LPS-treated control at 100 µg/mL.

Comparing the effects of EA fractions on IL-6 production at 100 µg/mL, D0, D6, and D12 significantly suppressed (P < 0.05) the IL-6 concentration to 80.9%, 89.3%, and 40.1% of the LPS-treated control, respectively, while SB did not. In addition, EA fraction of D12 significantly suppressed (P < 0.05) the IL-6 concentration in a dose-dependent manner to 81.5%, 77.3%, and 40.1% of the LPS-treated control at 10, 50, and 100 µg/mL, respectively. EA fractions of all samples including SB significantly suppressed (P < 0.05) the TNF-α concentration to 91.2%, 90.4%, 89.2%, and 86.8% of the LPS-treated control at a low concentration of 10 µg/mL, respectively, however, significant suppression was not observed at 50 and 100 µg/mL.

Effect of DM and EA fractions of Doenjang on iNOS and COX-2 protein expression in RAW 264.7 macrophages

Because DM and EA fractions of D12 suppressed NO and PGE2 production in LPS-treated RAW 264.7 cells, Western blot analysis was performed to examine whether those fractions of D12 inhibited iNOS and COX-2 protein expression.

As shown in Fig. 5, extremely low detectable levels of iNOS and COX-2 proteins were expressed by the cells under the basal condition, however, iNOS and COX-2 protein expression was remarkably increased by LPS treatment. Treatment with DM fractions of D12 resulted in reductions of iNOS protein expression to 44.2% and 33.9% of the LPS-treated control at 50 and 100 µg/mL, respectively, and COX-2 protein expression was reduced to 87.1% and 71.6%, respectively, while DM fraction of SB did not reduce iNOS and COX-2 expression. Treatment with EA fraction of D12 resulted in reduction of iNOS protein expression to 75.3% and 32.7% of the LPS-treated control at 50 and 100 µg/mL, but COX-2 expression was not reduced.

DISCUSSION

ROS are responsible for damage to phospholipids in biomembranes by lipid peroxidation, and are known to be one of the stimulators of the inflammatory response [19, 34, 35]. Strong association of the ability to scavenge the DPPH radical with lipid peroxidation has been reported [36] and relatively high positive correlation between anti-inflammatory and antioxidant activity of plant extracts was observed [37-39].

Several studies found that phenolic compounds were responsible for the antioxidant activity in fruits, vegetables, and grains [40-42]. Soybeans contain natural antioxidants. Tocopherol and phosphatides are found in soybean oil, while the non-oil component contains many isoflavones, a class of phytochemicals. Isoflavonoids from legumes exist mainly in glycoside conjugates that are less bioavailable [43,44]. The combined isoflavone aglycone and phenolic acids were reported to account for nearly all in vitro antioxidant activity of soybean and soy products [7, 45, 46]. Dsiedzic et al. [47] reported that phospholipids rich in soy product can synergistically promote antioxidant activity of isoflavone.

Chai et al. [3], who reported that IC₅₀ for DPPH radical scavenging activity was soybean > Meju > Doenjang > Ganjang and was negatively correlated with total polyphenol content, total flavonoid content, and total protein content, suggested that vanillic acid and protocatechuric acid are the effective compounds of higher antioxidant activity of soybean fermented foods. Melanoidin produced during fermentation of Doenjang also contributed to increased antioxidant activity. Browning of Doenjang is the result of enzymatic and nonenzymatic
processes. Enzymatic browning is known to be mainly due to melanin produced by tyrosinase released from Bacillus, and nonenzymatic browning is mainly due to Maillard reaction of amino acid, sugars and carbonyl substance in Doenjang [48]. Kim et al. [49] already reported that melanoidin content was high in chloroform extract of Meju and methanol extract of Doenjang. Jeong et al. [50] isolated Lactobacillus from traditional Meju showing high DPPH radical scavenging activity. In addition, 7,3',4-trihydroxyisoflavone, 7,8,4'-trihydroxyisoflavone, and 6,7,4'-trihydroxyisoflavone, isolated in long-term fermented and aged traditional Doenjang, was reported to show strong activity for scavenging DPPH radical and superoxide [51,52].

In this study, total isoflavone content showed a gradual decrease with increase of processing time from 3.01 mg/g in SB to 1.46 mg/g of Mj, 1.23 mg/g in D0 and 0.74 mg/g in D12. The reason could not be clearly explained, however, it could be due to hydrolysis and release out from the solid part of Meju into the liquid part during wet fermentation in saline and chemical biotransformation during the aging process. Wang and Murphy [53] reported that 61% and 44% of isoflavones in soybeans were lost in tempeh and tofu, respectively.

Many researchers reported that aglycone isoflavone content and its proportion of the total isoflavone content was increased by fermentation [21-23]. In this study, total isoflavone content showed a gradual decrease with increase of time during fermentation and aging, however, aglycone content and its proportion were increased until 3 or 6 months of aging, and then slowly decreased until 12 months of aging. High β-glucosidase activity of Aspergillus oryzae and Bacillus licheniformis, used in this study, was reported [54,55]. In this study, it was observed that not only β-glucoside isoflavone but also malonylg glucoside and acetylglucoside contents were decreased by fermentation, thereby, the accumulated aglycone content increased by fermentation and aging, however, it reached the maximum level at 3 months of aging and then decreased slowly.

It was questionable why aglycone content in Doenjang decreased after 3 months of aging, and the antioxidant activity was sustained or rather increased up to 12 months of aging despite a decrease in aglycone content after 3 months of aging. Probable reason might be due to biotransformation of aglycones into hydroxyisoflavones, such as 8-hydroxydaidzein, hydroxyisoflavone, and hydroxyglycitein [53]. Lee et al. [55] recently reported that fermentation of a solution of ground steamed soybeans with Aspergillus oryzae at 30°C decreased acetylglucoside and glycoside content and aglycone content increased after the first 12 hours of fermentation, however, aglycone content was decreased after the second or third 12 hours of fermentation due to the conversion to hydroxyisoflavone, and the content of this substance showed strong correlation with antioxidant activity.

It is needed to age the initial Doenjang for food. It is usually sold after 3 months of aging following the fermentation process, and consumed for a long time approximately several months to years. The limitations of our study are that it was not sampled at over 12 months of aging with only one sample at each time. In addition, the aglycone isoflavone content was not measured in the different fractions of Doenjang ethanol extract.

In this study, it was observed that DM and EA fractions from Doenjang ethanol extract showed higher antioxidant activity and polyphenol content than the other fractions, and especially these two fractions of D12 extract showed the highest activity among samples. Therefore, we performed further experiments on anti-inflammatory activity of Doenjang using DM and EA fractions of D12 ethanol extract in LPS-activated RAW 264.7 macrophages.

Macrophage plays an important role in the inflammatory response. When activated, macrophages release NO, cytokines, and lipid mediators such as prostaglandins, which promote inflammation by stimulating cellular migration to the target site [15]. LPS, the major component of the outer membrane of Gram-negative bacteria, consists of a lipid and a polysaccharide joined by a covalent bond. LPS activates a number of major cellular pathways that play critical roles in the pathogenesis of inflammatory response [56].

NO is an important physiological messenger in many biological systems including immunological, neuronal, and cardiovascular tissues [57,58]. PGE2, a key inflammatory mediator, is converted from free arachidonic acid by cyclooxygenase [57]. During the inflammatory process, significant quantities of the pro-inflammatory mediators of NO and PGE2 were generated by iNOS and COX-2, respectively. COX enzyme possesses both cyclooxygenase and peroxidase activities. Prostaglandins produced by COX impair immune surveillance and modulate proliferation in a variety of cell types. The peroxidase function contributes to activation of carcinogens. High production of NO has been shown to cause DNA damage as well as mutations during infection and inflammation. Over-expression of either COX-2 or iNOS may be intimately involved in the pathogenesis of many diseases, such as colon cancer, multiple sclerosis, neurodegenerative diseases, and heart infarction [25].

NF-κB, activator protein-1, CCAAT/enhancer-binding protein and cyclic-AMP-response element-binding protein are commonly or individually involved in regulation of inflammatory genes including iNOS and TNF-α [59]. In particular, activation of NF-κB induces expression of various inflammatory target genes involved in inflammation, inducing iNOS, COX-2, IL-6, and TNF-α. [60]. One study [56] reported that genistein blocked LPS-induced intracellular inflammatory signaling cascades by suppressing ROS production and NF-κB signaling pathway. In addition, genistein potently suppressed binding of LPS to toll-like receptor 4 on BV2 microglial cell surface.

In this study, DM fraction of D12 extract obviously suppressed COX-2 protein expression while EA fraction of D12 extract did not, even though these DM and EA fractions markedly suppressed PGE2 production to a similar extent in LPS-activated macrophages. It is considered that suppressed PGE2 production by EA fraction may be due to inhibition of COX-2 activity rather than expression of COX-2 at the protein level, and direct suppression of microsomal PGE2 synthase (mPGE2) activity or inhibition of 15-OH PG dehydrogenase activity, converting PGs into inactive forms. Koebeler et al. [61] reported that curcumin suppressed PGE2 production via inhibition of COX-2 expression and direct activation of mPGE2 activity.

Increased total polyphenol content and/or isoflavone aglycone during fermentation is known to be associated with various
bioactivities [7,15,24,27,45]. Results of this study indicated that effective substances of antioxidant and anti-inflammatory activities of Doenjang could be phenolics, aglycone isoflavone and some other substances dissolved rich in DM and EA fractions. Taken together, it is concluded that 12 month-aged Doenjang has higher anti-inflammatory and antioxidant activities than soybeans or Doenjang aged for a shorter time, and can be used as a beneficial food for human health.

Further study is needed to investigate the underlying biomolecular mechanisms for anti-inflammatory activity of Doenjang in various systems, and to find an effective candidate substance dissolved in DM and/or EA fractions.

REFERENCES

1. Lee TH, Do MH, Oh YL, Cho DW, Kim SH, Kim SY. Dietary fermented soybean suppresses UVB-induced skin inflammation in hairless mice via regulation of the MAPK signaling pathway. J Agric Food Chem 2014;62:8962-72.

2. Kwon DY, Daily JW 3rd, Kim HJ, Park S. Antidiabetic effects of fermented soybean products on type 2 diabetes. Nutr Res 2010;30:1-13.

3. Chai C, Ju HK, Kim SC, Park JH, Lim J, Kwon SW, Lee J. Determination of bioactive compounds in fermented soybean products using GC/MS and further investigation of correlation of their bioactivities. J Chromatogr B Analyt Technol Biomed Life Sci 2012;880:42-9.

4. Chung SI, Rico CW, Kang MY. Comparative study on the hypoglycemic and antioxidative effects of fermented paste (doenjang) prepared from soybean and brown rice mixed with rice bran or red gingsem marc in mice fed with high fat diet. Nutrients 2014;6:4610-24.

5. Hu CC, Hsiao CH, Huang SY, Fu SH, Lai CC, Hong TM, Chen HH, Lu FJ. Antioxidant activity of fermented soybean extract. J Agric Food Chem 2004;52:5735-9.

6. Chen YC, Sugiyama Y, Abe N, Kuruto-Niwa R, Nozawa R, Hirota A. DPPH radical-scavenging compounds from Dou-chi, a soybean fermented food. Biosci Biotechnol Biochem 2005;69:999-1006.

7. Kwak CS, Lee MS, Park SC. Higher antioxidant properties of Chungkookjang, a fermented soybean paste, may be due to increased aglycone and malonylglyceride isoflavone during fermentation. Nutr Res 2007;27:719-27.

8. Jung KO, Park SY, Park KY. Longer aging time increases the anticancer and antimitastatic properties of doenjang. Nutrition 2006;22:539-45.

9. Park KY, Jung KO, Rhee SH, Choi YH. Antimitagenic effects of doenjang (Korean fermented soypaste) and its active compounds. Mutat Res 2003;523-524:43-53.

10. Park KJ, Kim YM, Lee BH, Lee BK. Fungal microflora on Korean home-made Meju. Korean J Mycol 1977;5:7-12.

11. Lee SS. Meju fermentation for a raw material of Korean traditional soy products. Korean J Mycol 1995;23:161-75.

12. Kwak CS, Park SC, Song KY. Doenjang, a fermented soybean paste, decreased visceral fat accumulation and adipocyte size in rats fed with high fat diet more effectively than nonfermented soybeans. J Med Food 2012;15:1-9.

13. Park NY, Rico CW, Lee SC, Kang MY. Comparative effects of doenjang prepared from soybean and brown rice on the body weight and lipid metabolism in high fat-fed mice. J Clin Biochem Nutr 2012;51:235-40.

14. Cha YS, Yang JA, Back HI, Kim SR, Kim MG, Jung SJ, Song WO, Chae SW. Visceral fat and body weight are reduced in overweight adults by the supplementation of Doenjang, a fermented soybean paste. Nutr Res Pract 2012;6:520-6.

15. Blay M, Espinel AE, Delgado MA, Baiges I, Bladé C, Arola L, Salvadó J. Isoflavone effect on gene expression profile and biomarkers of inflammation. J Pharm Biomed Anal 2010;51:382-90.

16. Scott A, Khan KM, Cook JL, Duronio V. What is ‘inflammation’? Are we ready to move beyond Celsus? Br J Sports Med 2004;38:248-9.

17. Prasad S, Phrommoni K, Yadav VR, Chaturvedi MM, Aggarwal BB. Targeting inflammatory pathways by flavonoids for prevention and treatment of cancer. Planta Med 2010;76:1044-63.

18. Witkamp R, Monshouwer M. Signal transduction in inflammatory processes, current and future therapeutic targets: a mini review. Vet Q 2000;22:11-6.

19. Bak MU, Jeong JH, Kang HS, Jin KS, Ok S, Jeong WS. Cedrela sinensis leaves suppress oxidative stress and expressions of iNOS and COX-2 via MAPK signaling pathways in RAW 264.7 cells. J Food Sci Nutr 2009;14:269-76.

20. Kwon SH, Lee KB, Im KS, Kim SO, Park KY. Weight reduction and lipid lowering effects of Korean traditional soybean fermented products. J Korean Soc Food Sci Nutr 2006;35:1194-9.

21. Jang CH, Lim JK, Kim JH, Park CS, Kwon DY, Kim YS, Shin DH, Kim JS. Change of isoflavone content during manufacturing of Cheong-gukjang, a traditional Korean fermented soyfood. Food Sci Biotechnol 2006;15:643-6.

22. Nakajima N, Nozaki N, Ishihara K, Ishikawa A, Tsuji H. Analysis of isoflavone content in tempeh, a fermented soybean and preparation of a new isoflavone-enriched tempeh. J Biosci Bioeng 2005;100:685-7.

23. Shon MY, Seo KI, Lee SW, Choi SH, Sung NJ. Biological activities of Chungkukjang prepared with black bean and changes in phytoestrogen content during fermentation. Korean J Food Sci Technol 2000;32:936-41.

24. Danciu C, Soica C, Csanay E, Ambrus R, Feflea S, Peev C, Dehelean C. Changes in the anti-inflammatory activity of soy isoflavonoid genistein versus genistein incorporated in two types of cyclodextrin derivatives. Chem Cent J 2012;6:58.

25. Liang YC, Huang YT, Tsai SH, Lin-Shiau SY, Chen CF, Lin JK. Suppression of inducible cyclooxygenase and inducible nitric oxide synthase by apigenin and related flavonoids in macrophages. Carcinogenesis 1999;20:1945-62.

26. Ji G, Zhang Y, Yang Q, Cheng S, Hao J, Zhao X, Jiang Z. Genistein suppresses LPS-induced inflammatory response through inhibiting NF-κB following AMP kinase activation in RAW 264.7 macrophages. PLoS One 2012;7:e53101.

27. Singleton VL, Orthofer R, Lamuela-Raventós RM. Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. Methods Enzymol 1999;299:152-78.

28. Chae SK, Kang GS, Ma SJ, Bang KW, Oh MW, Oh SH. Standard Food Analysis. Seoul: Jigu Publishing Co.; 1997.

29. Senba Y, Nishishita T, Saito K, Yoshikawa H, Yoshikawa H. Stopped-flow and spectrophotometric study on radical scavenging by tea catechins and the model compounds. Chem Pharm Bull (Tokyo) 1999;47:1369-74.

30. Yildirim A, Mavi A, Kara AA. Determination of antioxidant and antimicrobial activities of Rumex crispus L. extracts. J Agric Food Chem 2001;49:4083-9.
31. Mosmann T. Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. J Immunol Methods 1983;65:55-63.
32. Green LC, Wagner DA, Glogowski J, Skipper PL, Wishnok JS, Tannenbaum SR. Analysis of nitrate, nitrite, and [15N]nitrate in biological fluids. Anal Biochem 1982;126:131-8.
33. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem 1976;72:248-54.
34. Zhang R, Brennan ML, Shen Z, MacPherson JC, Schmitt D, Molenda CE, Hazen SL. Myeloperoxidase functions as a major enzymatic catalyst for initiation of lipid peroxidation at sites of inflammation. J Biol Chem 2002;277:46116-22.
35. Coulibaly AY, Kiendrebeogo M, Kehoe PG, Sombie PA, Lamien CE, Millogo JP, Nacoulma OG. Antioxidant and anti-inflammatory activities of Scoparia dulcis L. J Med Food 2011;14:1576-82.
36. Chang ST, Wu JH, Wang SY, Kang PL, Yang NS, Shyur LF. Antioxidant activity of extracts from Acacia confusa bark and heartwood. J Agric Food Chem 2001;49:3420-4.
37. Michel P, Dobrowolska A, K集成电路al, Owczarek A, Bazylko A, Granica S, Piwowarski JP, Olzewska MA. Polyphenolic profile, antioxidant and anti-inflammatory activity of eastern teaberry (Gaultheria procumbens L.) leaf extracts. Molecules 2014;19:20498-520.
38. Noh KH, Jang JH, Min KH, Chinzorig R, Lee MO, Song YS. Suppressive effect of green tea seed coat ethyl acetate fraction on inflammation and its mechanism in RAW264.7 macrophage cell. J Korean Soc Food Sci Nutr 2011;40:1205-10.
39. Velioglu YS, Mazza G, Gao L, Oomah BD. Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. J Agric Food Chem 1999;46:1143-7.
40. Zainol MK, Abd-Hamid A, Yusof S, Muse R. Antioxidative activity of extracts from Acacia confusa bark and heartwood. J Agric Food Chem 1996;44:2377-83.
41. Velioglu YS, Mazza G, Gao L, Oomah BD. Antioxidant activity and total phenolics in selected fruits, vegetables, and grain products. J Agric Food Chem 1999;46:1143-7.
42. Green LC, Wagner DA, Glogowski J, Skipper PL, Wishnok JS, Tannenbaum SR. Analysis of nitrate, nitrite, and [15N]nitrate in biological fluids. Anal Biochem 1982;126:131-8.
43. Coulibaly AY, Kiendrebeogo M, Kehoe PG, Sombie PA, Lamien CE, Millogo JP, Nacoulma OG. Antioxidant and anti-inflammatory activities of Scoparia dulcis L. J Med Food 2011;14:1576-82.
44. Pratt DE. Natural antioxidants of soybeans and other oil-seeds. In: Simic MG, Karel M, editors. Autoxidation in Food and Biological Systems. New York (NY): Plenum Press; 1980. p.283-293.
45. Pratt DE. Natural antioxidants of soybeans and other oil-seeds. In: Simic MG, Karel M, editors. Autoxidation in Food and Biological Systems. New York (NY): Plenum Press; 1980. p.283-293.
46. Sheih IC, Fang TJ, Wu TK, Chen RY. Effects of fermentation on antioxidant properties and phytochemical composition of soy germ. J Sci Food Agric 2014;94:3163-70.
47. Dzdziecz S, Hudson BJ. Hydroxy isoflavones as antioxidants for edible oils. Food Chem 1983;11:161-6.
48. Kwok SH, Shon MY. Antioxidant and anticarcinogenic effects of traditional Doenjang during maturation periods. Korean J Food Preserv 2004;11:461-7.
49. Kim MH, Im SS, Kim SH, Kim GE, Lee JH. Antioxidative materials in domestic Meju and Doenjang 2. Separation of lipophilic brown pigment and their antioxidative activity. J Korean Soc Food Sci Nutr 1994;23:251-60.
50. Jeong JK, Zheng Y, Choi HS, Han GJ, Park KY. Catabolylom enzyme activities and physiological functionalities of lactic acid bacteria isolated from Korean traditional Meju. J Korean Soc Food Sci Nutr 2010;39:1854-9.
51. Roh C, Jung U, Jo SK. 6,7,4′-Trifhydroxyisoflavone from Doenjang inhibits lipid accumulation. Food Chem 2011;129:183-7.
52. Park JS, Park HY, Kim DH, Kim DH, Kim HK. Ortho-dihydroxyisoflavone derivatives from aged Doenjang (Korean fermented soypaste) and its radical scavenging activity. Bioorg Med Chem Lett 2008;18:5006-9.
53. Wang HJ, Murphy PA. Mass balance study of isoflavones during soybean processing. J Agric Food Chem 1996;44:2377-83.
54. Wu CH, Chou CC. Enhancement of aglycone, vitamin K2 and superoxide dismutase activity of black soybean through fermentation with Bacillus subtilis BCRC 14715 at different temperatures. J Agric Food Chem 2009;57:10695-700.
55. Lee S, Lee MK, Oh DK, Lee CH. Targeted metabolomics for Aspergillus oryzae-mediated biotransformation of soybean isoflavones, showing variations in primary metabolites. Biosci Biotechnol Biochem 2014;78:167-74.
56. Jeong JW, Lee HH, Han MH, Kim GY, Kim WJ, Choi YH. Anti-inflammatory effects of genistein via suppression of the toll-like receptor 4-mediated signaling pathway in lipopolysaccharide-stimulated BV2 microglia. Chem Biol Interact 2014;212:30-9.
57. Sunh YH, Chun HK, Kim SE, Kim YM, Seo JH, Shin MC, Shin MS, Yi JW, Shin DH, Kim H, Kim CJ. Anti-inflammatory and analgesic effects of the aqueous extract of Comi fructus in murine RAW 264.7 macrophage cells. J Med Food 2009;12:788-95.
58. Breed DS, Snyder SH. Nitric oxide: a physiologic messenger molecule. Annu Rev Biochem 1994;63:175-95.
59. Lee AK, Sung SH, Kim YC, Kim SG. Inhibition of lipopolysaccharide-inducible nitric oxide synthase, TNF-α and COX-2 expression by sauchinone effects on Ls:Bx phosphorylation, CEBP and AP-1 activation. Br J Pharmacol 2003;139(11):11-20.
60. Lee EJ, Kim C, Kim KY, Kim SM, Nam D, Jang HJ, Kim SH, Shim BS, Ahn KS, Choi SH, Jung SH, Ahn KS. Inhibition of LPS-induced inflammatory biomarkers by ethyl acetate fraction of Patrinia scabiosaefolia through suppression of NF-κB activation in RAW 264.7 cells. Immunopharmacol Immunotoxicol 2012;34:282-91.
61. Koebeler A, Northoff H, Werz O. Curcumin blocks prostaglandin E2 biosynthesis through direct inhibition of the microsomal prostaglandin E2 synthase-1. Mol Cancer Ther 2009;8:2348-55.