Asperisporium and Pantospora (Mycosphaerellaceae): epitypifications and phylogenetic placement

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Key words
Ascomycota
Capnodiales
Dothideomycetes
Ictectype
Pseudocercospora ulmilofiliae

Abstract The species-rich family Mycosphaerellaceae contains considerable morphological diversity and includes numerous anamorphic genera, many of which are economically important plant pathogens. Recent revisions and phylogenetic research have resulted in taxonomic instability. Ameliorating this problem requires phylogenetic placement of type species of key genera. We present an examination of the type species of the anamorphic Asperisporium and Pantospora. Cultures isolated from recent port interceptions were studied and described, and morphological studies were made of historical and new herbarium specimens. DNA sequence data from the ITS region and nLSU were generated from these type species, analysed phylogenetically, placed into an evolutionary context within Mycosphaerellaceae, and compared to existing phylogenies. Epitype specimens associated with living cultures and DNA sequence data are designated herein. Asperisporium caricae, the type of Asperisporium and cause of a leaf and fruit spot disease of papaya, is closely related to several species of Passalora including P. brachycarpa. The status of Asperisporium as a potential generic synonym of Passalora remains unclear. The monotypic genus Pantospora, typified by the synnematous Pantospora guazumae, is not included in Pseudocercospora sensu stritcto or sensu lato. Rather, it represents a distinct lineage in the Mycosphaerellaceae in an unresolved position near Mycosphaerella microsora.

Article info Received: 9 June 2011; Accepted: 1 August 2011; Published: 9 September 2011.

INTRODUCTION

Mycosphaerella and related fungi are classified in Capnodiales (Dothideomycetes, Ascomycota) and include thousands of species (Crous et al. 2007). These fungi have diverse ecological roles, especially as saprophytes and parasites, and numerous species are of agricultural significance (Crous 2009). As plant pathogens, these fungi are found on plant taxa across the embaphytes (Farr & Rossman 2011) with most species exhibiting host specificity (Crous 2009).

In terms of morphology, generic concepts for Mycosphaerella and its related anamorphs have been challenging as the number of variable and overlapping characters has led to different classifications that emphasize different characters (Baker et al. 2000, Crous & Braun 2003, Crous et al. 2007, 2009b). With progress towards phylogenetic hypotheses based on DNA sequence data, it has become apparent that older morphological classifications are riddled with non-monophyletic groups and unexpected bedfellows (Crous et al. 2007, 2009b, Crous 2009).

The backbone of our understanding of the phylogeny of this group is skewed in large part, but not entirely, towards sampling from fungi associated with hosts in two plant families, Myrtaceae and Proteaceae (Crous et al. 2007, 2009a, b, Crous 2009). Furthermore, the number of genetic loci that have been examined is relatively small. While the type species of Mycosphaerella, M. punctiformis, has been placed in a phylogenetic context (Verkley et al. 2004), type species of large, related genera have yet to be placed (Crous & Braun 2003, Crous et al. 2007, 2009b).

During the course of conducting work related to plant protection and quarantine, we obtained new collections via port interceptions that were identified as Asperisporium caricae and Pantospora guazumae. These collections represent the generic types of Asperisporium (Ellis 1971) and Pantospora (Deighton 1976), respectively. Asperisporium is a small genus that includes roughly 12 species (Kirk et al. 2008). It shares many morphological features with Passalora such as pigmented conidia and thickened and darkened conidiogenous loci while it is differentiated by verrucose conidia (Crous & Braun 2003, Schubert & Braun 2005), but this distinction has been considered doubtful (Crous & Braun 2003, Schubert & Braun 2005). Asperisporium caricae is responsible for an important leaf and fruit spot disease of Carica papaya (papaw or papaya) (Stevens 1939) that is commonly referred to as black spot, blight or ‘rust’ of pawpaw (Ellis & Holliday 1972). The synnematous Pantospora is a monotypic genus that causes a leaf spot disease of Guazuma ulmilofilia (Deighton 1976). Deighton (1976) and Crous & Braun (2003) both noted its similarity to Pseudocercospora, especially in regards to the dictyospores in the type species of Pseudocercospora, P. vitis, and the latter authors formerly classified it in that genus. Since neither Asperisporium nor Pantospora have been placed phylogenetically (Crous & Braun 2003, Schubert & Braun 2005), we generated DNA sequence data from the ITS and nLSU, conducted analyses, present phylogenetic placements of these genera, and discuss them in the context of existing phylogenetic studies (Crous et al. 2007, 2009b). To further stabilize the application of Asperisporium and Pantospora, we herein epitypify the type species of these genera with collections associated with living cultures and DNA sequence data. Morphological descriptions of the designated epitypes and associated ex-epitype cultures are presented, and

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historical collections and descriptions were studied to confirm conspecificity of the new collections.

MATERIALS AND METHODS

Morphology and herbarium material

Dried herbarium material was rehydrated and viewed in 3 % KOH (Larget et al. 1977), and microscopic observations of cultures were made of material mounted in 3 % KOH or buffered Shear’s mounting fluid (Graham 1959). Herbarium acronyms follow Thiers (2011). See Farr & Rossman (2011) for additional information about collections housed at BPI.

Cultures

Isolates in pure culture were grown in plastic Petri plates on 2 % Difco potato-dextrose agar (PDA) and BBL Sabouraud dextrose agar (SDA), which were both prepared according to the manufacturers’ instructions. Cultures were incubated at 24 °C with a 12 h light/dark regimen. A subset of these cultures was transferred to a 12 h black light/dark regimen also at 24 °C approximately 2–2.5 wk to promote sporulation. Sporulation was also promoted by exposing 1 mo old cultures that were otherwise incubated in the dark at ambient room temperature to 1 h of UV at an intensity setting of 60 with a Fisher Biotech Transilluminator FBTIV-816 at approximately 2–3 d intervals. Terminology for colour includes general terms from author notes as well as standard terminology with the sample reference code in parentheses from Kornerup & Wanscher (1967). Voucher cultures were deposited at the Centraalbureau voor Schimmelcultures (CBS).

DNA extraction, PCR amplification, and sequencing

DNA was extracted from fresh mycelium using the Qiagen DNeasy Plant Mini Kit (Gaithersburg, Maryland). DNA sequence data were generated from the nuclear encoded ribosomal ITS region (ITS1, 5.8S, ITS2), the nuclear encoded ribosomal large subunit (28S nLSU), and a portion between amino acid motifs 5–7 of the nuclear DNA-dependant RNA Polymerase II gene’s large subunit (RPB2) for Asperisporium caricae and Pantospora guazumae. PCR cocktails for all reactions contained 0.2 µM of each forward and reverse primer, GoTaq Flexi Buffer (Promega; Madison, Wisconsin), 0.2 mM dNTPs, 2.0 mM Mg2+, and 5µL Promega GoTaq. Primers used for PCR and sequencing were fRPB2-5F and fRPB2-7cR (Liu et al. 1999) for RPB2, IT5S and IT54 (White et al. 1990) for the ITS, and LROR (Monclavo et al. 2000) and LR7 (Vilgalys & Hester 1990) for the nLSU. Thermal cycling conditions for RPB2 and nLSU were those of Malkus et al. (2006) and Reeb et al. (2004), respectively. Thermal cycling conditions for the ITS were: 95 °C for 60 s; 35 cycles at 95 °C for 15 s, 55 °C for 20 s, and 72 °C for 1 min; and a final extension at 72 °C for 3 min. Cycle sequencing was conducted using BigDye v. 3.1 (Applied Biosystems; Foster City, California) sequencing chemistry. Resulting fluorescence-labelled fragments were sequenced on an ABI 3730 capillary sequencer. Electropherograms were edited in the program Geneious Pro v. 5 (Drummond et al. 2010). Sequences were submitted to GenBank (http://www.ncbi.nlm.nih.gov).

Data matrix and phylogenetic analysis

For the purpose of determining preliminary phylogenetic positions of A. caricae and P guazumae among members of Mycosphaerellaceae and related genera, their nLSU sequences were manually incorporated into the alignment of Crous et al. (2009b) and analyzed phylogenetically using Maximum Parsimony (MP) as the optimality criterion in the program TNT v. 1.1 (Goloboff et al. 2008). Based on these results (not presented), a selection of taxa was made to provide the appropriate phylogenetic context for a fine-scale placement of these taxa within Mycosphaerellaceae. Additional taxa were included based on similarity and determined using BLAST results of Genbank. In all cases, the ITS and the nLSU sequences retrieved from GenBank to represent a particular taxon and combined for analysis were generated from the same culture.

Multiple sequence alignment of the ITS and the nLSU was conducted within the program Geneious Pro v. 5 (Drummond et al. 2010) using MUSCLE v. 3.6 (Edgar 2004), and adjusted manually. Insertions and deletions (indels) within the concatenated ITS and nLSU matrix were coded using the ‘simple indel coding’ method of Simmons & Ochoterena (2000) as implemented in Gapcode.py (Rhee 2007). The resulting alignment was deposited into TreeBASE as accession number SN11747. Bayesian Inference (BI) phylogenetic analysis was conducted on the concatenated ITS/nLSU matrix, including indel characters, in MrBayes v. 3.1.2 (Huelsenbeck & Ronquist 2001, Ronquist & Huelsenbeck 2003). Indel data were analyzed as ‘restriction’ data type and DNA sequence data were analyzed as ‘DNA’ data type. The GTR+I+G model of DNA sequence evolution was determined as the best fit using the Akaikie Information Criterion (AIC; Posada & Buckley 2004) in MrModelTest v. 2.2 (Nylander 2004) and implemented in the BI analysis. The coding parameter for the restriction data type was set to variable. All other parameters were left as default. The posterior probability (pp) distribution of trees was estimated based on the results of two independent runs of 1 million generations each of the Markov Chain Monte Carlo (MCMC) simulation, which sampled trees every 100 generations until the standard deviation of split frequencies reached less than 0.01. The burn-in was determined using the program Tracer v. 1.5 (Rambaut & Drummond 2007). The remaining trees were combined and used to build a 50 % majority rule consensus within the program FigTree v. 1.3.1 (Rambaut 2009).

RESULTS

Data matrix and phylogenetic analysis

The combined ITS, nLSU, plus indel matrix contained 1 304 characters, 53 of which were indels. Of these 1 251 nucleotide characters, 202 were variable (16.1 %) and 126 were informative (10.1 %). Thirty-one indel characters were informative (58.5 %). The ITS alignment was 747 nucleotide positions, of which 72 of which were variable (9.6 %) and 45 informative (6.0 %). The nLSU alignment contributed seven informative (10.1 %). Thirty-one indel characters were informative (60.9 %). The nLSU alignment contributed three of which were informative (42.9 %).

Preliminary MP analysis of the nLSU suggested that A. caricae and P guazumae occupy phylogenetic positions within the clade with 95 % bootstrap support (Crous et al. 2009b) containing Dothiostroma (clade 7), which is sister to the clade comprised of Pseudocercospora-like fungi (clade 6), Phaeophleospora (clade 5), and Lecanosticta (clade 4). Our combined analysis of ITS, nLSU, and indels resulted in strong support (pp = 1.0) for this clade and an increased level of resolution among its members relative to nLSU alone (Fig. 1). Within this clade, P guazumae occupies an unresolved position with Mycosphaerella microsora sister to a well-supported clade (p = 0.94) containing Passalora brachycarpa, M. ellipsoidea, M. aurantia, M. buckinghamiae, M. africana, and A. caricae. The latter four species are members of a well-supported (pp = 1.0) polytomy sister to Passalora brachycarpa.
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**Asperisporium** Maubl., Lavoura 16: 207/212. 1913 ‘1912’, and Bull. Trimestriel Soc. Mycol. France 29: 357. 1913. Note: Maublanc published his article in two languages in Lavoura and separately in Bull. Trimestriel Soc. Mycol. France. We were unable to confirm which printed issuance was first.

Typus genericus. *Asperisporium caricae* (Speg.) Maubl.

*Asperisporium caricae* (Speg.) Maubl., Lavoura 16: 207/212. 1913 ‘1912’, and Bull. Trimestriel Soc. Mycol. France 29: 357. 1913. — Fig. 2

Basionym. *Cercospora caricae* Speg., Anales Soc. Ci. Argent. 22: 215. 1886. Note: This species was published as ‘Cercospora? caricae’ to indicate doubt as to the generic classification, but it is still valid according to ICBN Art. 34.1 (McNeill et al. 2006).

≡ *Fusicladium caricae* (Speg.) Sacc., Rend. Congr. Bot. Palermo: 58. 1902. = *Pucciniopsis caricae* Höhn., Centralbl. Bakteriol., 2. Abth.: 60: 5. 1923, nom. illeg., non Earle 1902. Isonym (Spec.) Seaver, Scientific Survey of Porto Rico and the Virgin Islands, Vol. VIII, Part 1: 104. 1926.) Note: This combination was published as ‘Pucciniopsis? caricae’ to indicate doubt as to the generic classification, but it is still valid according to ICBN Art. 34.1. = *Scolicotrichum caricae* Ellis & Everh., J. Mycol. 7: 134. 1892 as ‘Scolicotrichum’.

≡ *Epiclinium cumminsii* Massee, Bull. Misc. Inform. Kew 1898: 133. 1898. = *Pucciniopsis caricae* Earle, Bull. New York Bot. Gard. 2: 340. 1902.

Types and typifications

Lectotypus of *Cercospora caricae* designated by Chupp, A Monograph of the Fungus Genus Cercospora: 106. 1953: Paraguay, Guarapi, on leaves of Carica papaya, Feb. 1881, coll. B. Balansa, No. 2739 (LPS).

Epitypus of *Cercospora caricae* hic designatus: Brazil, Intercepted at USA, Washington, Seattle, entering from Brazil, on fruit of Carica papaya, 16 Apr. 2010, coll. C. Weight, isolated by J.F. Bischoff from BPI 880773, epitype is a dried culture on SDA (BPI 881135); ex-epitype CBS 130298; GenBank accession nos: ITS (JN190955), LSU (JN190953), RP2B (JN190951).
Fig. 2  Cultures and microscopic features of Asperisporium caricae. a–d. Ex-epitype (CBS 130298) at approximately 1 mo at 24 °C with a 12 h light/dark regimen: a. PDA; b. reverse on PDA; c. SDA; d. reverse on SDA. — e. Lectotype packet, No. 2739 (LPS). — f–m. Ex-epitype (CBS 130298) on SDA; f–h. conidiophores and conidia; i–l. conidia; m. spermatia. — Scale bars = 10 µm for all.
Notes — Spegazzini (1886) cited two collections, no. 2739 and no. 3855, in the protologue without indicating either as type. These are syntypes according to ICBN Art. 9.4. Chupp (1953) lectotypified the species via ICBN Art. 7.11 when he indicated the word type for no. 2739 at LPS.

**Description of the epitype (preserved culture) and collection from which it was isolated**

Fruticosulous with spots scattered, 3–4 mm diam. *Sporodocha* formed on stromata, immersed becoming erumpent, punctiform, blackish to black. In culture on SDA, sporodocha of loosely to densely arranged conidiophores produced on darkly pigmented stromata. *Conidiophores* macronematous, mononematous, simple or less commonly branched, more or less straight to slightly sinuous, smooth, brownish, septate, 58–168 × 5–10 µm. *Conidigenous cells* integrated, terminal, polyblastic, as accepted at USA, Washington, Seattle, entering from Brazil, on fruit of *Carica papaya*, 16 Apr. 2010, coll. C. Weight (BPI 880773); culture isolated by J.F. Bischoff from BPI 880773, dried culture on SDA (BPI881135, designated epitype). — **COLOMBIA**, Intercepted at New York, J.F.K.I.A., #24116, on fruit of *Carica papaya*, 4 Jan. 1969, coll. C. Smock (BPI 424779). — **CUBA**, Santiago de Las Vegas, Experiment Station, on leaves of *Carica papaya*, 8 May 1915, coll. R.A. Jelie (BPI 424780). — **DOMINICAN REPUBLIC**, Valle del Cibao, SantiagoProv, Santiago, Hato del Yaque, Gardens, on leaves of *Carica papaya*, 19 Jan. 1931, coll. R. Ciferr. Mycotylo-Domingensia Exsiccate 311 (BPI 424783). — **HONDURAS**, Jamastran, on leaves of *Carica papaya*, 10 Mar. 1964, coll. A.S. Muller (BPI 424789). — **MEXICO**, Intercepted at Texas, Laredo, #008382, on fruit of *Carica papaya*, 20 Feb. 1975, coll. S. Kendall (BPI 424778). — **PUERTO RICO**, Yauco, on leaves of *Carica papaya*, 30 Mar. 1916, coll. W.H. Whetzel and E.W. Olive (BPI 424777). — **VENUEZUELA**, Federal District, Valle de Puerto La Cruz, El Limón, on leaves of *Carica papaya*, 15 Jan. 1925, coll. H. Sydow (BPI 1112163).

Notes — The collection from which the epitype was isolated is a cut portion of fruit. Unfortunately, it could not be well preserved before it was overgrown with anamorphic fungi including *Penicillium*. Thus, we have designed a dried culture isolated from this collection as the epitype. Conidiophores in culture were longer than those observed on host tissues (Ellis 1971, Ellis & Holliday 1972).

Ellis & Holliday (1972) listed three taxonomic synonyms. Examination of the protologues of these names supports the synonymy. Maublanc (1913a, b) described a teleomorph that he suggested was associated with *Asperisporium caricae* as *Sphaerella caricae*. This morph was later classified as *Mycosphaerella caricae* (Maubl.) Hansf., which makes it an illegitimate later homonym of *Mycosphaerella caricae* Syd. & P. Syd. These teleomorphic names have been considered to be conspecific and associated with the anamorphic *Phoma caricae* (Sivanesan 1984). Others expressed doubt that *A. caricae* has a known teleomorph since the connection has never been proven (Crous & Braun 2003). Recently, *M. caricae* Syd. & P. Syd. and *M. caricae* (Maubl.) Hansf. were listed separately with each having different anamorphs, but the type of the latter was not studied (Aptroot 2006). *Mycosphaerella caricae* Syd. & P. Syd. was transferred to *Stagonosporopsis* with no mention of the later homonym in the synonym list (Aveskamp et al. 2010).

No additional information about the life cycle of *A. caricae* in regards to its ascal state and the synonymy of these historical names can be added by this study. However, a structure of unproven identity that is presumed to be a spermogonium was found in culture. Its presumptive spermatogenous cells and spermatia are reminiscent of those described by Crous (1998) for *Mycosphaerella caricae*. The presumptive spermia did not germinate on PDA. The presence of spermogonium suggests that *A. caricae* has a sexual stage in its life cycle.

**Pantospora** Cif., Ann. Mycol. 36: 242. 1938.

Typus genericus. *Pantospora guazumae* Cif.

≡ *Dictyoccephal A.G. Meioeres, Universidade do Recife, Instituto de Micologia, Publicação No. 372: 13. 1962.

Typus genericus. *Cercospora ulmifoliae* Obreg.-Bot., which was indirectly referenced via the invalid *Dictyoccephala ulmifoliae* (Obreg.-Bot.) A.G. Meioeres (ICBN Art. 10.3).

**Pantospora** guazumae Cif., Ann. Mycol. 36: 242. 1938. — Fig. 3

≡ *Cercospora ulmifoliae* Obreg.-Bot., Caldaisa 1: 51. 1941.

≡ *Dictyoccephala ulmifoliae* (Obreg.-Bot.) A.G. Meioeres, Universidade do Recife, Instituto de Micologia, Publicação No. 372: 13. 1962 as ‘ulmifolia’, nom. inval. via ICBN Art. 33.4.

≡ *Psuedocercospora ulmifoliae* (Obreg.-Bot.) U. Braun & Crous, CBS Biodiversity Series 1: 415. 2003. Note: *Pseudocercospora guazumae* (Syd.) Deighton prevented a legitimate combination based on *Pantospora guazumae* (Crous & Braun 2005).
Types and typifications

Lectotypus of *Pantospora guazumae* designated by Deighton, Mycol. Pap. 140: 159. 1976: Dominican Republic, Valle del Cibao, prov. Santiago, Hato del Yaque, on leaves of *Guazuma ulmifolia*, 20 Apr. 1930, coll. R. Ciferri & A.M. Borgna Ciferri, Batey no. 1, Mycoflora Domingensis Exsiccata 210 (IMI 59269, K(M) 169346).

Epitypus of *Pantospora guazumae* hic designatus: Mexico, Intercepted at USA, Arizona, Nogales, entering from Mexico, on leaf of *Guazuma ulmifolia*, 12 Feb. 2009, coll. J. Moore (BPI 880778); ex-epitype CBS 130299, GenBank accession nos: ITS (JN190956), LSU (JN190954), RPB2 (JN190952).

Notes — Ciferri (1938) distributed the original material as number 210 in exsiccatea sets. These are syntypes according to ICBN Art. 9.4. Deighton (1976) lectotypified the species when he indicated that the collection at IMI was the type. This specimen is now housed at K.

**Fig. 3** Cultures and microscopic features of *Pantospora guazumae*. a–d. Ex-epitype (CBS 130299) at approximately 1 mo at 24 °C with a 12 h light/dark regimen: a. PDA; b. reverse on PDA; c. SDA; d. reverse on SDA. — e. Leaf spot on abaxial surface of *Guazuma ulmifolia* (BPI 880778, designated epitype); f. synnema on *Guazuma ulmifolia* (BPI 880778, designated epitype); g–i. conidia from ex-epitype (CBS 130299) on PDA. — Scale bars = 1 mm for e, 10 μm for f–i.
**Description of the epitype**

*Leaf spots* scattered, 1.5–2 mm diam, visible on both adaxial and abaxial surfaces, typically circular, dark brown to blackish with a distinct, lighter, occasionally purplish, margin on adaxial surface, discoloured brown with a distinct, lighter, occasionally purplish, margin on abaxial surface. *Caespituli* hypophyllous, scattered within margin of leaf spots, conidiophores densely aggregated forming synnemata. *Synnemata* basistromatic with immersed stromata, erect, more or less even with hyphal tips spreading apart at apex, up to 290 µm long, up to 40 µm wide along non-apical portion. *Conidiophores* branched, more or less straight to somewhat sinuous with apices more or less obtuse, often interwoven, smooth, pale brown to brown, septate, 3–6 µm, widest towards synnematal apex. *Conidigenous cells* integrated, terminal, blastic, slightly verrucose and with annelations, conidigenous loci visible, not significantly thickened or darkened. *Conidia* up to 43 × 13 µm, solitary, versiform, ellipsoid with short beaks to obclavate, slightly verrucose, brown, with multiple transverse, longitudinal, and occasionally oblique septa, hila visible but not thickened or darkened.

**Culture characteristics** — Colonies on PDA 12–13 mm after 1 mo at 24 °C with a 12 h light/dark regimen; mycelium forming a raised mound composed of tiers of smaller mounds, surface lanose, near brownish orange (7C7) to light brown (7D7); margin lobed, more or less concolorous; reverse almost black or black; medium becoming discoloured with a greyish red (7B6) to orange (6B7) soluble pigment. Surface mycelium with hyphae branching, walls smooth, at times covered with orange crystals, hyaline to brownish orange in Shear’s mounting fluid, purple in 3 % KOH, septate, 5–6.5 µm diam. Sporulation not observed.

Colonies on SDA 11–13 mm after 1 mo at 24 °C with a 12 h light/dark regimen; mycelium forming a raised mound composed of tiers of smaller mounds, surface lanose, multicoloured with near brownish orange (7C7) to light brown (7D4) to orangean tan to greyish hues; margin lobed, more or less concolorous; reverse almost black or black; medium becoming discoloured with a greyish red (7B6) to orange (6B7) soluble pigment. Surface mycelium with hyphae branching, walls smooth, at times covered with orange, pigmentary crystals, hyaline, orangish to brownish orange in Shear’s mounting fluid, purple in 3 % KOH, septate, 5–6.5 µm diam. Sporulation not observed.

Colonies on both media showing increased growth rates on a 12 h black light/dark regimen. Sporulation not observed. Colonies exposed to UV via the transilluminator showed increased growth rate, a significant increase in submerged hyphae with dark pigmentation at margins with a corresponding lack of mycelium above, and production of a few scattered synnemata. Mature synnemata were observed approximately 1.5 wk after the UV regimen was initiated. Synnemata in culture were of greater width than on host leaves. Conidia 32–60 × 6–16 µm, versiform, ellipsoid with short beaks, obclavate, cylindrical-obclavate, or more or less cylindrical, slightly verrucose, brown, with multiple transverse, longitudinal, and occasionally oblique septa. Both dictyosporous and scolecosporous conidia present, additional longitudinal and/or oblique septa may develop over time. Otherwise, synnemata similar in culture and on host leaves.

**Habitat & Distribution — Leaves of Guazuma ulmifolia (Malvaceae).** According to Farr & Rossman (2011), this fungus is known from North America (Cuba, Dominican Republic) and South America (Brazil, Colombia). This is the first report of this species from Mexico.

**Specimens examined.** COLOMBIA, Rioclaro, near Cali, on Guazuma ulmifolia, June 1938, coll. C. Garces-Orejuela (BPI 445535). – DOMINICAN REPUBLIC, Valle del Cibao, prov. Santiago, Hato del Yaque, on leaves of Guazuma ulmifolia, 20 Apr. 1930, coll. R. Ciferri & A.M. Borgrna Ciferri, Batey no. 1, Mycophora Domingensis Exsiccata 210 (IMI 59269, K(M) 169346, lectotype); (BPI 445536, syntype); Feb. 1932, coll. R. Ciferri (BPI 445537); 01 Sept. 1931, coll. R. Ciferri (BPI 445538); 01 Sept. 1931, coll. R. Ciferri (BPI 445539). – MEXICO, Intercepted at USA, Arizona, Nogales, entering from Mexico, on leaf of Guazuma ulmifolia, 12 Feb. 2009, coll. J. Moore (BPI 980778, designated epitype).

**Notes** — This epitype description is based on sparse, dried herbarium material. The epitype specimen is not fully mature as it possesses small leaf spots and lacks a large number of mature conidia. In the interest of preserving the specimen, the sample size of measured structures is relatively small. Discrepancies with the observations of previous authors should not be considered significant. As with and loosely following Medeiros (1962), UV was found to stimulate the production of synnemata in culture.

Both Deighton (1976) and Crous & Braun (2003) considered *Cercospora ulmifoliae* as a synonym of *Pantospora guazumae* even though they did not study Obregón-Botero’s original material of *C. ulmifoliae*. Based on the descriptions provided by Obregón-Botero (1941) and Chupp (1953), there can be no doubt that the two names are synonymous. There is no known sexual stage for *Pantospora guazumae*.

**DISCUSSION**

*Asperisporium* has been thought of as a likely synonym of *Passalora* since the two genera were separated on the basis of conidial surface ornamentation (Crous & Braun 2003, Schubert & Braun 2005). These authors tentatively maintained them as distinct due to the absence of DNA sequence data. The phylogenetic analyses place *Asperisporium caricae*, the type of the genus, in a relatively close relationship with several species of *Passalora* including *P. brachycarpa*. Sequence data are available for only a small number of the approximately 580 species of *Passalora* (Kirk et al. 2008) and the type species of the genus, *Passalora bacilligera*, has not been sequenced and placed phylogenetically. As *Passalora* currently stands, it is a polyphyletic genus (Crous et al. 2009b). A final conclusion cannot be made on the status of *Asperisporium*. The collective works of Patil & Thirumalachar (1966), Ellis (1976), Barreto & Evans (1995), Braun (2000a, b) and Braun & Crous (2007) cover nearly all of the remaining species that are currently classified in *Asperisporium*.

Crous & Braun (2003) formally classified *Pantospora guazumae* in *Pseudocercospora*, noted the lack of molecular data for it and explained why the presence of dictyosporous may not be a distinguishing character at the rank of genus. Based on the phylogenetic analyses, *Pantospora* is not closely related to two clades presented by Crous et al. (2009b), namely, *Pseudocercospora* including the type species, *P. vitis* (clade 16) and *Pseudocercospora*-like (clade 14). *Pantospora* is in an unresolved position near *Mycosphaerella microsora*. *Pantospora* appears to be a distinct lineage.

In summary, the type species of *Asperisporium*, *A. caricae*, and *Pantospora*, *P. guazumae*, have been placed phylogenetically in the *Mycosphaerellaceae*. Studies of their culture characteristics were made and both species were epitypified with herbarium material associated with living cultures and DNA sequence data. The phylogenetic placement of these genera demonstrates that previous generic concepts and the perceived values of particular morphological features were not necessarily congruent with phylogeny. Since this is becoming a repeated result (Crous & Braun 2003, Crous et al. 2007, 2009b, Crous 2009) and that sampling in terms of taxa and numbers of genetic loci remains small for such a large family, we advocate
a conservative and patient approach towards the creation of new taxonomic schemes and nomenclatural novelties in this group of fungi.

Acknowledgements We thank Jorge A. Chayle and other staff at LPS for providing information about and images of the original material of Asperisporium caricae and permission to reproduce the image of the prototype plate. Curators at K are acknowledged for their loan of the lectotype of Pantospora. Pedro W. Crous and Uwe Braun are thanked for their comments about preliminary phylogenetic trees and generic concepts. Amy Y. Rossman provided helpful comments to improve the manuscript prior to its submission.

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