Microbiome-aware ecotoxicology of organisms: relevance, pitfalls and challenges

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Abstract
Over the last 15 years, the advent of high-throughput ‘omics’ techniques has revealed the multiple roles and interactions occurring among hosts, their microbial partners and their environment. This microbiome revolution has radically changed our views of biology, evolution and individuality. Sitting at the interface between a host and its environment, the microbiome is a relevant yet understudied compartment for ecotoxicology research. Various recent works confirm that the microbiome reacts to and interacts with contaminants, with consequences for hosts and ecosystems. In this paper, we thus advocate for the development of a “microbiome-aware ecotoxicology” of organisms. We emphasize its relevance and discuss important conceptual and technical pitfalls associated with study design and interpretation. We identify topics such as functionality, quantification, temporality, resilience, interactions and prediction as major challenges and promising venues for microbiome research applied to ecotoxicology.

1. Introduction: the microbiome is relevant to ecotoxicology

The significance of microbes to multicellular organisms is long documented. Because only a fraction of microorganisms can be isolated in culture, it is the advent of high-throughput sequencing technologies which ultimately revealed how diverse and numerically abundant they were. Microorganisms form complex symbiotic communities of eukaryotes, bacteria, archaea, and viruses referred to as the microbiome (HMP, 2012; McFall-Ngai et al., 2013; Tipton et al., 2019). Over the last 15 years, the microbiome has been a new frontier in Life Sciences (Alivisatos et al., 2015), and microorganisms were shown to be involved and even necessary in many host functions including nutrition, defense, immunity, development and behavior (Rakoff-Nahoum et al., 2004; Archie and Theis, 2011; McFall-Ngai et al., 2013). A current paradigm assumes that most animals and plants harbor a microbiome (Rosenberg and Zilber-Rosenberg, 2018). However, some species of comb jellies and nematomorpha, as well as certain life stages of insects including honeybee larvae, are
apparently devoid of a microbiome, suggesting that this may be an over-simplification (Martinson et al., 2012; Hammer et al., 2019). In humans, the microbiome may represent as many cells as the hosts and up to 1,000 times more genes, questioning the concept of individuality and the limits of self (Gilbert et al., 2012; Rees et al., 2018). The holobiont concept, referring to the entity formed by a host organism and its various microbial associates, arise to encompass the complexity of hosts and their microbiome (Mindell, 1992; Bordenstein and Theis, 2015). All these discoveries have fueled a “microbiome revolution” that increasingly spreads through all fields of life sciences, with extensions to behavioral and human sciences (Blaser, 2014; Cryan and Dinan, 2019).

Recent research focuses mostly on the links between hosts and their microbiome, and the reciprocal influence they exert on each other, revealing its significance to host physiology, homeostasis, disease, health and fitness (Selber-Hnatiw et al., 2017). Interestingly, most members of the microbiome are located on epithelia (mucosa, skin...), *i.e.* animal or plant polarized tissues that separate the inside from the outside of the organism. Sitting at the interface between a host and its environment, epithelia and their associated microbiome are the hosts buffer and first line of defense against contaminants and environmental stressors (Barr et al., 2013). Deep-sea hydrothermal vent mussels are an example of this. They harbor bacteria located in the gill epithelium that oxidize hydrogen sulfide, a compound toxic to their hosts, and fix carbon that contributes hosts nutrition (Halary et al., 2008). Toxicology studies investigating the effects of chemical compounds on organisms examine the accumulation, bio-transformation, elimination and effects in tissues, and are thus beginning to account for associated microbiome. This paper aims to emphasize the relevance, pitfalls and promises of host-associated microbiome research for ecotoxicology and advocates for the emergence of a “microbiome-aware ecotoxicology” of multicellular organisms, *i.e.* an approach that fully incorporates the microbiome compartment as a dynamic interface interacting with host and environment (Figure 1).

### 2. The microbiome responds to and interacts with contaminants

Data on effects of environmental contaminants on microbiomes has been published over the last years, with a focus on animal gut-associated bacteria (Claus et al., 2016; Jin et al., 2017; Rosenfeld, 2017; Adamovsky et al., 2018). Published studies include controlled exposure of model organisms to various types of contaminants including pesticides, antibiotics, heavy metals, xenobiotics, or nanoparticles (Licht and Bahl, 2019; Monroy-Torres et al., 2019). Bacterial community composition is typically assessed using the 16S rRNA sequence as a taxonomic marker that identifies Operational Taxonomic Units (OTUs), a commonly used proxy for species. The effect of exposures on OTU richness and diversity is then evaluated using multivariate statistics to test whether contaminants interfere with microbiome composition (Ramette, 2007; Evariste et al., 2019). Many studies include complementary analyses on host parameters of toxicological relevance such as markers of the immune system, tissue histology and developmental markers.

Besides descriptive studies that correlate exposures and microbiome variations, functional studies directly investigate the interactions between the gut microbiome and contaminants (Figure 2; Schmidt et al., 2019). It has been shown that microbiomes of mammals guts are able to metabolize a wide range of xenobiotics (*e.g.* polycyclic aromatic hydrocarbons, polychlorobiphenyls, nitrotoluenes) and could protect animals from deleterious effects (Claus et al., 2016). But the microbiome may also activate some compounds and mediate toxicity to hosts. For instance, the human colon microbiome was shown to convert polycyclic aromatic hydrocarbons into estrogenic metabolites with consequences on hormonal equilibrium (Van de Wiele et al., 2005); and the nephrotoxicity of melamine in rats results from its conversion into toxic cyanuric acid mediated by
the bacterium *Klebsiella terrigena* (Zheng et al., 2013). Many pharmaceutical drugs such as lovastatin or loperamide were also proved to be activated in the small intestine by bacteria-mediated biotransformations (Lavrijsen et al., 1995; Yoo et al., 2014). Alternatively, xenobiotics can also alter activities of the gut microbiome (Licht and Bahl, 2019). Besides the obvious example of antibiotics, many molecules such as epoxiconazole or glyphosate, both pesticides, are known to induce shifts in microbiome compositions (Xu et al., 2014). Because the microbiome substantially responds and interacts with contaminants, it must now be considered a key player in toxicology.

3. Producing and interpreting microbiome data relevant to ecotoxicology

3.1. Conceptual pitfalls: identifying a “good” microbiome and a “good” model species

Ecotoxicology studies that address the microbiome rely on a microbial ecology background and need to consider the caveats associated with this discipline. Experimental investigations to date have focused mostly on bacteria. However, numerous studies have demonstrated the significance of Archaea and microbial Eukaryotes, including fungi, to host physiology (Hoffmann et al., 2013; Richard and Sokol, 2019), and the key role of phages in regulating bacterial populations (Mirzaei and Maurice, 2017; Bettarel et al., 2018). A comprehensive description of microbiome functioning requires all components to be accounted for (Rowan-Nash et al., 2019). However, the lack of universal, easy-to-obtain markers for some groups, notably for viruses, still precludes the development of systematic analyses that require deep metagenomics and specific expert analysis pipelines.

Besides, OTUs composition provides only a partial description of the real microbial diversity. Indeed, a single 16S rRNA-based OTU can encompass a diversity of distinct bacterial genotypes, potentially quite different in term of their respective functional phenotypes and responses to a stimulus (Andam, 2019; Hanafiah and Lopes, 2020). The genus *Vibrio* for example includes strains with very different lifestyles, including commensals, light-producing mutualists of the squid *Euprymna scolopes*, and pathogens of numerous metazoans, that all display almost identical 16S rRNA sequences (Sawabe et al., 2007). The relative abundances and dynamics of these different phenotypes can thus not be monitored using 16S rRNA.

Another major difficulty is the general lack of baseline knowledge regarding microbiomes of toxicology model species, for which very little-to-no data is available regarding wild populations (Uenishi et al., 2007; Hird, 2017; Shinohara et al., 2019). Besides, organisms used in tests are often sourced from rearing facilities, and have thus experienced domestication, a process documented to lead to massive changes in bacterial microbiome compositions. In vertebrates, changes include overall bacterial species richness decrease and shift in taxa abundances due to dietary, social and environmental conditions of captivity (reviewed in Hird, 2017; Alessandri et al., 2019). Effects in other taxa are less documented and less clear-cut. In the silkworm for example, domestication is associated with higher bacterial diversity (Chen et al., 2018). The representativity of model species in ecotoxicology versus their wild relatives thus remains to be evaluated in the light of their domestication history. Interestingly, humans are no exception to this trend, and gut microbiomes in industrialized societies greatly differ from the recent ancestral microbiome and from that of contemporary traditional populations (e.g. hunters-gatherers). Changes in diets, sanitation and medical practices have led to a functional shift from fiber to mucus degraders, high frequency of antibiotic resistance, loss of particular taxa (e.g. Spirochaetes) and overall diversity decrease (Blaser, 2016; Sonnenburg and Sonnenburg, 2019). This is assumed to result in non-optimal microbiomes associated with increased risk of chronic diseases. Lack of knowledge, along with inter-individual
variability (discussed below, 3.2), undermines the identification of the ‘normal’, balanced microbiome composition, i.e. the eubiotic state. This compromises the proper diagnosis of a dysbiosis (an ‘abnormal’, unbalanced) state upon exposure to contaminants (Figure 2). Indeed, although many factors can cause dysbiosis that may lead to health issues, it is not easy to establish what a healthy/eubiotic microbiome is (Iebba et al., 2016). Recently, authors insisted that dysbiosis due to stressors is first of all the destabilization of the stable eubiotic state. Changes in abundances of certain beneficial taxa are evident signs of dysbiosis, but interestingly, increased inter-individual variability in microbiome composition could be another signature of dysbiosis (Zaneveld et al., 2017).

In between the relative simplicity of most invertebrate-associated microbiomes in which a few OTUs are usually dominant (e.g. Raymann et al., 2017) and the extreme complexity of mammal-associated microbiomes (with hundreds to thousands of OTUs), teleost fish and their tens to a few hundred bacterial OTUs offer an interesting intermediate, besides their relevance to the monitoring of aquatic ecosystems (Llewellyn et al., 2014). Choosing a model thus involves addressing different levels of microbiome complexity, functionality and domestication history. Whether current models in toxicology are relevant to microbiome-aware ecotoxicology studies needs to be evaluated.

### 3.2. Technical pitfalls: performing the right experiment to detect effects

Most studies using controlled microcosms are monitoring various compartments that are potential sources of microbial diversity (e.g. food for animals, water for aquatic organisms). When scaling up to more holistic approaches such as mesocosms or the natural environment, potential sources of microorganisms dramatically increase, requiring the investigation of multiple compartments (e.g. food, prays, parasites, water, particles, sediments).

Fifteen years of human gut microbiome research revealed the high level of intra- (between body regions or life stages) and inter-individual heterogeneity in community compositions (HMP, 2012; Rothschild et al., 2018). Although less documented, high levels of intra- and inter-individual variation are reported in other taxa including fish (e.g. Atlantic cod, salmon, rainbow trout, zebrafish) (Star et al., 2013; Llewellyn et al., 2014; Lowrey et al., 2015; Duperron et al., 2019; Evariste et al., 2019). In the zebrafish for example, gut-associated communities become increasingly different from those in the environment, and inter-individual variation increases across development (Stephens et al., 2016). Skin-associated communities are different on different body regions in the rainbow trout (Lowrey et al., 2015). These examples emphasize the importance of replication levels, and of addressing the exact same life stage and tissue region in all individuals.

Sex-differentiated responses to compounds are commonly reported in ecotoxicology studies, for example in medaka fish exposed to cyanotoxins (Le Manach et al., 2016). Sex also influences microbiome composition in various vertebrates, probably due to hormones and sex-specific immunity responses (Haro et al., 2016). It also affects microbiome responses. In lab-reared stickleback fed different diets, diet induced changes in some bacterial taxa abundances, but effects on bacteria in males were uncorrelated with effects observed in females, supporting that diet effects were clearly sex-specific (Bolnick et al., 2014). Authors measured similar sex-specific diet effects in mice and humans. Sex-specific effects on microbiome responses to contaminants are also documented.

Exposure to silver nanoparticles was for example shown to modify the gut microbiome structure in male zebrafish, but not in females (Ma et al., 2018). Experimental design should thus carefully consider confounding factors of which sex is an important one.
4. The roads less traveled: challenges in microbiome-aware ecotoxicology

4.1. Functionality and integration

One major finding of the Human Microbiome Project was that despite high levels of intra- and inter-individual variation in the taxonomic compositions of bacterial communities, the functions they performed, as encoded by the metagenome, were highly conserved (HMP, 2012). Similar functions are thus performed by taxonomically distinct microorganisms. This concept known as functional redundancy is now recognized as key to the resistance and resilience of microbial communities (Allison and Martiny, 2008; Moya and Ferrer, 2016). Because of this, and the fact that closely related bacteria can display markedly different functionalities, community composition is not a reliable predictor of functions. Predictive tools for functional profiling based on composition (e.g. PICRUSt (Langille et al., 2013)) thus suffer limitations, and identity and functions should ideally be investigated in tandem. Functional capabilities can be evaluated through metagenomic sequencing, but genes and functions that are expressed at specific time points are better evaluated by metatranscriptomic or metaproteomic approaches. Metabolomics, which map metabolites, are another important tool that profiles ongoing metabolisms, and thus informs functions (Bundy et al., 2009; Gao et al., 2017) although, as for all of the above, the improvement of databases supporting metabolite identifications will be critical (Gertsman and Barshop, 2018). The integration of these approaches in multi-omics appears challenging, yet particularly promising for revealing the causal role of the microbiome and mechanisms involved in contaminants metabolism and toxic effects (Rohart et al., 2017; Koh and Bäckhed, 2020). This will be key in integrating the microbiome in adverse outcome pathways (AOPs) and risk assessment (figure 2 and Adamovsky et al., 2018).

4.2. Quantification

Microbiome-aware ecotoxicology should identify contaminant threshold values relevant to microbiomes. Indeed, microbial communities may shift rapidly and non-linearly between contrasting alternative, more or less stable states provided some parameters reach threshold values. The existence of yet-undescribed tipping points is for example hypothesized to explain the existence of bimodal distributions of abundances of certain bacteria in the human gut (Lahti et al., 2014; van Nes et al., 2016), and may be a trigger of dysbiosis. To identify tipping points in ecotoxicology, studies should examine dose-dependent responses and chronic exposure to low doses as done for toxicological effect on host traits, for example the determination of non-observable adverse effect limit (NOAEL).

Microbiome composition assessments also need to become more quantitative. Indeed, metabarcoding datasets produce taxa relative abundances tables, and their variations. In these, an increase in one group thus cannot be properly interpreted, as it may as well represent a lower decrease relative to other groups in a globally shrinking population. Bacterial densities in guts of distinct lineages of rainforest ants were for example shown to vary by orders of magnitude based on qPCR quantifications; interestingly, absolute abundance variations were better correlated with habitat (arboreal or terrestrial) and trophic position than actual community compositions (Sanders et al., 2017). Antibiotics, which are reported to affect relative abundances in bacterial communities, act first by affecting the total number of bacteria present, as was clearly demonstrated for streptomycin and sancomycin, this being their major influence on the microbiome (Willing et al., 2011; Vieira-Silva et al., 2019). Absolute abundances are relevant to our understanding of the environment-host-microbiome continuum, and should thus be informed whenever possible, for example using quantitative PCR (Tkacz et al., 2018). Quantifying bacteria within organisms is however challenging,
as demonstrated by the very different estimations of bacteria-to-human cell ratios found in the
literature (Sender et al., 2016).

4.3. Temporality and resilience

The nature and amplitude of variations are important aspects of microbiome response to
contaminants. Composition of communities and diversity indices are still the main endpoints of most
studies. However, the dynamics of these variations during and after exposure are certainly as
important. In humans, microbiome dynamics are individual-dependent (Flores et al., 2014).
Dynamics inform resilience, evaluating whether variations have long-term effects on the microbiome,
or whether it fully recovers and returns to a naive, pre-exposure stable state (Figure 2). Antibiotic
exposure was for example shown to affect human gut bacterial communities for several months post-
exposure, and similar effects can be expected with many contaminants (Dethlefsen and Relman,
2011; Francino, 2015). Whether iterative exposure to some contaminants may lead to habituation,
and thus become less influential to microbiomes over time, is also an important issue. Finally, how
microbiome resilience itself may be affected by environmental factors (e.g. temperature, pH,
interactions, seasonality) remains to be investigated.

4.4. Interactions and prediction

The holobiont is more than just the sum of its parts (Bordenstein and Theis, 2015). With
dozens-to-thousands distinct coexisting microbial taxa, and many more if phages are considered, an
animal’s gut or skin is a whole ecosystem in which multiple interactions among members and with
the host influence its functioning. These as well as interactions with the environment, including the
contaminants and microorganisms occurring there, need to be accounted for. For example, a new
method coupling spatial imaging of metabolites and bacterial genotypes, MetaFISH, was developed
to characterize interactions occurring between symbiotic bacteria and the gill epithelial cells of
hydrothermal vent mussels. It allowed the identification of metabolites located at the host-symbiont
interface on tissue sections at the micrometer scale (Geier et al., 2020). Such a method is promising
to monitor small-scale interactions between contaminants, the microbiome and the host and further
explore causality (Koh and Bäckhed, 2020). Co-occurrence networks that are based on positive or
negative correlations between the occurrence of microorganisms, functions, and environmental
parameters also help in exploring interactions and formulating hypotheses (reviewed in Faust and
Raes, 2012). A strong relationship between the presence of a contaminant and that of certain bacterial
taxa can suggest an ability to metabolize the former, which can then be tested (Claus et al., 2016).
Changes in the network structure itself can indicate microbial successions in time series experiments
or dysbiosis, and may support modelling approaches (Hoffmann et al., 2007; Zhou et al., 2010).

5. Conclusion: what can the microbiome do for ecotoxicology and vice versa?

By analogy with the famous essay written by Dobzhansky (1973), it is tempting these days to
suggest that “Nothing in Biology makes sense except in the light of the microbiome”. Ecotoxicology
is no exception to this trend, and must not lag behind other disciplines that have embraced the
microbiome revolution. However, the microbiome is not just another ecotoxicological endpoint, but a
peculiar and complex biological compartment that exhibits its own ecological, metabolic, functional
and thus ecotoxicological rules (Evariste et al., 2019). Instead, a microbiome-aware ecotoxicology of
organisms needs to develop (Figure 1). This involves questioning, and not only transferring, classical
ecotoxicology protocols and model organisms’ relevance to microbiome studies. Close cooperation
between microbial ecologists and ecotoxicologists is needed. They have a lot in common: the
 Complexity of microbiomes and their response mirrors that of contaminants and their interactions; and both domains start with reductionist approaches, and strive to scale up to holistic approaches that encompass ecosystems full complexity and produce real-life-relevant data.

A major challenge is to move on from observing correlations to addressing causality, and ultimately explain processes, e.g. demonstrate mitigating effects of the microbiome at the population level in a given ecosystem. Repeatability is a key point, which involves inter-studies comparisons and meta-analyses for which tools are becoming available (e.g. Amplicon Sequence Variants for OTU clustering (Callahan et al., 2017)). Microbiome features including taxa or functions may become bioindicators of contamination, as recently proposed in stream ecosystems (Simonin et al., 2019). Modelling interactions between environment, contaminants, microbiomes and hosts will become tractable, with a certain level of predictive power (Gould et al., 2018). No doubt the dialogue between disciplines will result in mutual enrichment, and will allow to make the most of the microbiome revolution applied to ecotoxicology.

Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Author Contributions

All authors (SD, SH, AG and BM) have contributed to the writing of the manuscript and the production of figures. All authors have read and approved the final version.

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References

Adamovsky, O., Buerger, A. N., Wormington, A. M., Ector, N., Griffitt, R. J., Bisesi, J. H., et al. (2018). The gut microbiome and aquatic toxicology: An emerging concept for environmental health. *Environ. Toxicol. Chem.* 37, 2758–2775. doi:10.1002/etc.4249.

Alessandri, G., Milani, C., Mancabelli, L., Mangifesta, M., Lugli, G. A., Viappiani, A., et al. (2019). The impact of human-facilitated selection on the gut microbiota of domesticated mammals. *FEMS Microbiol. Ecol.* 95. doi:10.1093/femsec/fiz121.

Alivisatos, A. P., Blaser, M. J., Brodie, E. L., Chun, M., Dangl, J. L., Donohue, T. J., et al. (2015). MICROBIOME. A unified initiative to harness Earth’s microbiomes. *Science* 350, 507–508. doi:10.1126/science.aac8480.
Allison, S. D., and Martiny, J. B. H. (2008). Colloquium paper: resistance, resilience, and redundancy in microbial communities. *Proc. Natl. Acad. Sci. U. S. A.* 105 Suppl 1, 11512–11519. doi:10.1073/pnas.0801925105.

Andam, C. P. (2019). Clonal yet Different: Understanding the Causes of Genomic Heterogeneity in Microbial Species and Impacts on Public Health. *mSystems* 4. doi:10.1128/mSystems.00097-19.

Archie, E. A., and Theis, K. R. (2011). Animal behaviour meets microbial ecology. *Anim. Behav.* 82, 425–436. doi:10.1016/j.anbehav.2011.05.029.

Barr, J. J., Auro, R., Furlan, M., Whiteson, K. L., Erb, M. L., Pogliano, J., et al. (2013). Bacteriophage adhering to mucus provide a non-host-derived immunity. *Proc. Natl. Acad. Sci. U. S. A.* 110, 10771–10776. doi:10.1073/pnas.1305923110.

Bettarel, Y., Halary, S., Auguet, J.-C., Mai, T. C., Van Bui, N., Bouvier, T., et al. (2018). Corallivory and the microbial debacle in two branching scleractinians. *ISME J.* 12, 1109–1126. doi:10.1038/s41396-017-0033-5.

Blaser, M. J. (2014). The microbiome revolution. *J. Clin. Invest.* 124, 4162–4165. doi:10.1172/JCI78366.

Blaser, M. J. (2016). Antibiotic use and its consequences for the normal microbiome. *Science* 352, 544–545. doi:10.1126/science.aad9358.

Bolnick, D. I., Snowberg, L. K., Hirsch, P. E., Lauber, C. L., Org, E., Parks, B., et al. (2014). Individual diet has sex-dependent effects on vertebrate gut microbiota. *Nat. Commun.* 5, 4500. doi:10.1038/ncomms5500.

Bordenstein, S. R., and Theis, K. R. (2015). Host Biology in Light of the Microbiome: Ten Principles of Holobionts and Hologenomes. *PLOS Biol* 13, e1002226. doi:10.1371/journal.pbio.1002226.

Bundy, J. G., Davey, M. P., and Viant, M. R. (2009). Environmental metabolomics: a critical review and future perspectives. *Metabolomics*. Available at: http://agris.fao.org/agris-search/search.do?recordID=US201301600087 [Accessed March 27, 2020].

Callahan, B. J., McMurdie, P. J., and Holmes, S. P. (2017). Exact sequence variants should replace operational taxonomic units in marker-gene data analysis. *ISME J.* 11, 2639. doi:10.1038/ismej.2017.119.

Chen, B., Du, K., Sun, C., Vimalanathan, A., Liang, X., Li, Y., et al. (2018). Gut bacterial and fungal communities of the domesticated silkworm (Bombyx mori) and wild mulberry-feeding relatives. *Isme J.* 12, 2252–2262. doi:10.1038/s41396-018-0174-1.

Claus, S. P., Guillou, H., and Ellero-Simatos, S. (2016). The gut microbiota: a major player in the toxicity of environmental pollutants? *Npj Biofilms Microbiomes* 2, 16003. doi:10.1038/npjbiofilms.2016.3.
Consortium, T. H. M. P. (2012). Structure, function and diversity of the healthy human microbiome. *Nature* 486, 207–214. doi:10.1038/nature11234.

Cryan, J. F., and Dinan, T. G. (2019). Talking about a microbiome revolution. *Nat. Microbiol.* 4, 552–553. doi:10.1038/s41564-019-0422-9.

Dethlefsen, L., and Relman, D. A. (2011). Incomplete recovery and individualized responses of the human distal gut microbiota to repeated antibiotic perturbation. *Proc. Natl. Acad. Sci. U. S. A.* 108 Suppl 1, 4554–4561. doi:10.1073/pnas.1000087107.

Duperron, S., Halary, S., Habiballah, M., Gallet, A., Huet, H., Duval, C., et al. (2019). Response of Fish Gut Microbiota to Toxin-Containing Cyanobacterial Extracts: A Microcosm Study on the Medaka (*Oryzias latipes*). *Environ. Sci. Technol. Lett.* 6, 341–347. doi:10.1021/acs.estlett.9b00297.

Evariste, L., Barret, M., Mottier, A., Mouchet, F., Gauthier, L., and Pinelli, E. (2019). Gut microbiota of aquatic organisms: A key endpoint for ecotoxicological studies. *Environ. Pollut. Barking Essex 1987* 248, 989–999. doi:10.1016/j.envpol.2019.02.101.

Francino, M. P. (2015). Antibiotics and the Human Gut Microbiome: Dysbioses and Accumulation of Resistances. *Front. Microbiol.* 6, 1543. doi:10.3389/fmicb.2015.01543.

Gao, B., Chi, L., Mahbub, R., Bian, X., Tu, P., Ru, H., et al. (2017). Multi-Omics Reveals that Lead Exposure Disturbs Gut Microbiome Development, Key Metabolites, and Metabolic Pathways. *Chem. Res. Toxicol.* 30, 996–1005. doi:10.1021/acs.chemrestox.6b00401.

Geier, B., Sogin, E. M., Michellod, D., Janda, M., Kompauer, M., Spengler, B., et al. (2020). Spatial metabolomics of in situ host-microbe interactions at the micrometre scale. *Nat. Microbiol.* 5, 498–+. doi:10.1038/s41564-019-0664-6.

Gertsman, I., and Barshop, B. A. (2018). Promises and Pitfalls of Untargeted Metabolomics. *J. Inherit. Metab. Dis.* 41, 355–366. doi:10.1007/s10545-017-0130-7.

Gilbert, S. F., Sapp, J., and Tauber, A. I. (2012). A symbiotic view of life: we have never been individuals. *Q. Rev. Biol.* 87, 325–341.

Gould, A. L., Zhang, V., Lamberti, L., Jones, E. W., Obadia, B., Korasidis, N., et al. (2018). Microbiome interactions shape host fitness. *Proc. Natl. Acad. Sci. U. S. A.* 115, E11951–E11960. doi:10.1073/pnas.1809349115.

Halary, S., Riou, V., Gaill, F., Boudier, T., and Duperron, S. (2008). 3D FISH for the quantification of methane- and sulphur-oxidising endosymbionts in bacteriocytes of the hydrothermal vent mussel Bathymodiolus azoricus. *ISME J* 2, 284–292.
Hammer, T. J., Sanders, J. G., and Fierer, N. (2019). Not all animals need a microbiome. *FEMS Microbiol. Lett.* 366. doi:10.1093/femsle/fnz117.

Hanafiah, A., and Lopes, B. S. (2020). Genetic diversity and virulence characteristics of *Helicobacter pylori* isolates in different human ethnic groups. *Infect. Genet. Evol. J. Mol. Epidemiol. Evol. Genet. Infect. Dis.* 78, 104135. doi:10.1016/j.meegid.2019.104135.

Haro, C., Rangel-Zúñiga, O. A., Alcalá-Díaz, J. F., Gómez-Delgado, F., Pérez-Martínez, P., Delgado-Listar, J., et al. (2016). Intestinal Microbiota Is Influenced by Gender and Body Mass Index. *PloS One* 11, e0154090. doi:10.1371/journal.pone.0154090.

Hird, S. M. (2017). Evolutionary Biology Needs Wild Microbiomes. *Front. Microbiol.* 8. doi:10.3389/fmicb.2017.00725.

Hoffmann, C., Dollive, S., Grunberg, S., Chen, J., Li, H., Wu, G. D., et al. (2013). Archaea and fungi of the human gut microbiome: correlations with diet and bacterial residents. *PloS One* 8, e66019. doi:10.1371/journal.pone.0066019.

Hoffmann, K. H., Rodriguez-Brito, B., Breitbart, M., Bangor, D., Angly, F., Felts, B., et al. (2007). Power law rank-abundance models for marine phage communities. *FEMS Microbiol. Lett.* 273, 224–228. doi:10.1111/j.1574-6968.2007.00790.x.

Iebba, V., Totino, V., Gagliardi, A., Santangelo, F., Cacciotti, F., Trancassini, M., et al. (2016). Eubiosis and dysbiosis: the two sides of the microbiota. *New Microbiol.* 39, 1–12.

Jin, Y., Wu, S., Zeng, Z., and Fu, Z. (2017). Effects of environmental pollutants on gut microbiota. *Environ. Pollut.* 222, 1–9. doi:10.1016/j.envpol.2016.11.045.

Koh, A., and Bäckhed, F. (2020). From Association to Causality: the Role of the Gut Microbiota and Its Functional Products on Host Metabolism. *Mol. Cell* 78, 584–596. doi:10.1016/j.molcel.2020.03.005.

Lahti, L., Salojärvi, J., Salonen, A., Scheffer, M., and Vos, W. M. de (2014). Tipping elements in the human intestinal ecosystem. *Nat. Commun.* 5, 1–10. doi:10.1038/ncomms5344.

Langille, M. G. I., Zaneveld, J., Caporaso, J. G., McDonald, D., Knights, D., Reyes, J. A., et al. (2013). Predictive functional profiling of microbial communities using 16S rRNA marker gene sequences. *Nat. Biotechnol.* 31, 814–821. doi:10.1038/nbt.2676.

Lavrijsen, K., van Dyck, D., van Houdt, J., Hendrickx, J., Monbaliu, J., Woestenborghs, R., et al. (1995). Reduction of the prodrug loperamide oxide to its active drug loperamide in the gut of rats, dogs, and humans. *Drug Metab. Dispos. Biol. Fate Chem.* 23, 354–362.

Le Manach, S., Khenfech, N., Huet, H., Qiao, Q., Duval, C., Marie, A., et al. (2016). Gender-Specific Toxicological Effects of Chronic Exposure to Pure Microcystin-LR or Complex Microcystis aeruginosa Extracts on Adult Medaka Fish. *Environ. Sci. Technol.* 50, 8324–8334. doi:10.1021/acs.est.6b01903.

Licht, T. R., and Bahl, M. I. (2019). Impact of the gut microbiota on chemical risk assessment. *Curr. Opin. Toxicol.* 15, 109–113. doi:10.1016/j.cotox.2018.09.004.
Llewellyn, M. S., Boutin, S., Hoseinifar, S. H., and Derome, N. (2014). Teleost microbiomes: the
state of the art in their characterization, manipulation and importance in aquaculture and
fisheries. *Front. Microbiol.* 5, 207. doi:10.3389/fmicb.2014.00207.

Lowrey, L., Woodhams, D. C., Tacchi, L., and Salinas, I. (2015). Topographical Mapping of the
Rainbow Trout (Oncorhynchus mykiss) Microbiome Reveals a Diverse Bacterial Community
with Antifungal Properties in the Skin. *Appl. Environ. Microbiol.* 81, 6915–6925.
doi:10.1128/AEM.01826-15.

Ma, Y., Song, L., Lei, Y., Jia, P., Lu, C., Wu, J., et al. (2018). Sex dependent effects of silver
nanoparticles on the zebrafish gut microbiota. *Environ. Sci. Nano* 5, 740–751.
doi:10.1039/C7EN00740J.

Martinson, V. G., Moy, J., and Moran, N. A. (2012). Establishment of characteristic gut bacteria
during development of the honeybee worker. *Appl. Environ. Microbiol.* 78, 2830–2840.
doi:10.1128/AEM.07810-11.

McFall-Ngai, M., Hadfield, M. G., Bosch, T. C. G., Carey, H. V., Domazet-Loso, T., Douglas, A. E.,
et al. (2013). Animals in a bacterial world, a new imperative for the life sciences. *Proc. Natl.
Acad. Sci. U. S. A.* 110, 3229–3236. doi:10.1073/pnas.1218525110.

Mindell, D. P. (1992). Phylogenetic consequences of symbioses: Eukarya and Eubacteria are not
monophyletic taxa. *Biosystems* 27, 53–62. doi:10.1016/0303-2647(92)90046-2.

Mirzaei, M. K., and Maurice, C. F. (2017). Ménage à trois in the human gut: interactions between
host, bacteria and phages. *Nat. Rev. Microbiol.* 15, 397–408. doi:10.1038/nrmicro.2017.30.

Monroy-Torres, R., Hernández-Luna, M. A., Ramírez-Gómez, X. S., and López-Briones, S. (2019).
Role of the Microbiome as the First Metal Detoxification Mechanism. *Prebiotics Probiotics -
Potential Benefits Nutr. Health.* doi:10.5772/intechopen.89232.

Moya, A., and Ferrer, M. (2016). Functional Redundancy-Induced Stability of Gut Microbiota
Subjected to Disturbance. *Trends Microbiol.* 24, 402–413. doi:10.1016/j.tim.2016.02.002.

Rakoff-Nahoum, S., Paglino, J., Eslami-Varzaneh, F., Edberg, S., and Medzhitov, R. (2004).
Recognition of commensal microflora by toll-like receptors is required for intestinal
homeostasis. *Cell* 118, 229–241. doi:10.1016/j.cell.2004.07.002.

Ramette, A. (2007). Multivariate analyses in microbial ecology. *FEMS Microbiol. Ecol.* 62, 142–160.
doi:10.1111/j.1574-6941.2007.00375.x.

Raymann, K., Shaffer, Z., and Moran, N. A. (2017). Antibiotic exposure perturbs the gut microbiota
and elevates mortality in honeybees. *Plos Biol.* 15, e2001861. doi:10.1371/journal.pbio.2001861.

Rees, T., Bosch, T., and Douglas, A. E. (2018). How the microbiome challenges our concept of self.
*PLOS Biol.* 16, e2005358. doi:10.1371/journal.pbio.2005358.
Richard, M. L., and Sokol, H. (2019). The gut mycobiota: insights into analysis, environmental interactions and role in gastrointestinal diseases. *Nat. Rev. Gastroenterol. Hepatol.* 16, 331–345. doi:10.1038/s41575-019-0121-2.

Rohart, F., Gautier, B., Singh, A., and Lê Cao, K.-A. (2017). mixOmics: An R package for ’omics feature selection and multiple data integration. *PLoS Comput. Biol.* 13, e1005752. doi:10.1371/journal.pcbi.1005752.

Rosenberg, E., and Zilber-Rosenberg, I. (2018). The hologenome concept of evolution after 10 years. *Microbiome* 6, 78. doi:10.1186/s40168-018-0457-9.

Rosenfeld, C. S. (2017). Gut Dysbiosis in Animals Due to Environmental Chemical Exposures. *Front. Cell. Infect. Microbiol.* 7, 396. doi:10.3389/fcimb.2017.00396.

Rothschild, D., Weissbrod, O., Barkan, E., Kurilshikov, A., Korem, T., Zeevi, D., et al. (2018). Environment dominates over host genetics in shaping human gut microbiota. *Nature* 555, 210–215. doi:10.1038/nature25973.

Rowan-Nash, A. D., Korry, B. J., Mylonakis, E., and Belenky, P. (2019). Cross-Domain and Viral Interactions in the Microbiome. *Microbiol. Mol. Biol. Rev. MMBR* 83. doi:10.1128/MMBR.00044-18.

Sanders, J. G., Lukasik, P., Frederickson, M. E., Russell, J. A., Koga, R., Knight, R., et al. (2017). Dramatic Differences in Gut Bacterial Densities Correlate with Diet and Habitat in Rainforest Ants. *Integr. Comp. Biol.* 57, 705–722. doi:10.1093/icb/icx088.

Sawabe, T., Kita-Tsukamoto, K., and Thompson, F. L. (2007). Inferring the evolutionary history of vibrios by means of multilocus sequence analysis. *J. Bacteriol.* 189, 7932–7936. doi:10.1128/JB.00693-07.

Schmidt, R., Ulanova, D., Wick, L. Y., Bode, H. B., and Garbeva, P. (2019). Microbe-driven chemical ecology: past, present and future. *ISME J.* 13, 2656–2663. doi:10.1038/s41396-019-0469-x.

Selber-Hnatiw, S., Rukundo, B., Ahmadi, M., Akoubi, H., Al-Bizri, H., Aliu, A. F., et al. (2017). Human Gut Microbiota: Toward an Ecology of Disease. *Front. Microbiol.* 8, 1265. doi:10.3389/fmicb.2017.01265.

Sender, R., Fuchs, S., and Milo, R. (2016). Are We Really Vastly Outnumbered? Revisiting the Ratio of Bacterial to Host Cells in Humans. *Cell* 164, 337–340. doi:10.1016/j.cell.2016.01.013.

Shinohara, A., Nohara, M., Kondo, Y., Jogahara, T., Nagura-Kato, G. A., Izawa, M., et al. (2019). Comparison of the gut microbiotas of laboratory and wild Asian house shrews (Suncus murinus) based on cloned 16S rRNA sequences. *Exp. Anim.* doi:10.1538/expianim.19-0021.

Simonin, M., Voss, K. A., Hassett, B. A., Rocca, J. D., Wang, S.-Y., Bier, R. L., et al. (2019). In search of microbial indicator taxa: shifts in stream bacterial communities along an urbanization gradient. *Environ. Microbiol.* 21, 3653–3668. doi:10.1111/1462-2920.14694.
Sonnenburg, E. D., and Sonnenburg, J. L. (2019). The ancestral and industrialized gut microbiota and implications for human health. *Nat. Rev. Microbiol.* 17, 383–390. doi:10.1038/s41579-019-0191-8.

Star, B., Haeverkamp, T. H. A., Jentoft, S., and Jakobsen, K. S. (2013). Next generation sequencing shows high variation of the intestinal microbial species composition in Atlantic cod caught at a single location. *BMC Microbiol.* 13, 248. doi:10.1186/1471-2180-13-248.

Stephens, W. Z., Burns, A. R., Stagaman, K., Wong, S., Rawls, J. F., Guillemin, K., et al. (2016). The composition of the zebrafish intestinal microbial community varies across development. *Isme J.* 10, 644–654. doi:10.1038/ismej.2015.140.

Tipton, L., Darcy, J. L., and Hynson, N. A. (2019). A Developing Symbiosis: Enabling Cross-Talk Between Ecologists and Microbiome Scientists. *Front. Microbiol.* 10. doi:10.3389/fmicb.2019.00292.

Tkacz, A., Hortala, M., and Poole, P. S. (2018). Absolute quantitation of microbiota abundance in environmental samples. *Microbiome* 6, 110. doi:10.1186/s40168-018-0491-7.

Uenishi, G., Fujita, S., Ohashi, G., Kato, A., Yamauchi, S., Matsuzawa, T., et al. (2007). Molecular analyses of the intestinal microbiota of chimpanzees in the wild and in captivity. *Am. J. Primatol.* 69, 367–376. doi:10.1002/ajp.20351.

Van de Wiele, T., Vanhaecke, L., Boeckaert, C., Peru, K., Headley, J., Verstraete, W., et al. (2005). Human colon microbiota transform polycyclic aromatic hydrocarbons to estrogenic metabolites. *Environ. Health Perspect.* 113, 6–10. doi:10.1289/ehp.7259.

van Nes, E. H., Arani, B. M. S., Staal, A., van der Bolt, B., Flores, B. M., Bathiany, S., et al. (2016). What Do You Mean, “Tipping Point”? *Trends Ecol. Evol.* 31, 902–904. doi:10.1016/j.tree.2016.09.011.

Vieira-Silva, S., Sabino, J., Valles-Colomer, M., Falony, G., Kathagen, G., Caenepeel, C., et al. (2019). Quantitative microbiome profiling disentangles inflammation- and bile duct obstruction-associated microbiota alterations across PSC/IBD diagnoses. *Nat. Microbiol.* 4, 1826–1831. doi:10.1038/s41564-019-0483-9.

Willing, B. P., Russell, S. L., and Finlay, B. B. (2011). Shifting the balance: antibiotic effects on host-microbiota mutualism. *Nat. Rev. Microbiol.* 9, 233–243. doi:10.1038/nrmicro2536.

Xu, C., Liu, Q., Huan, F., Qu, J., Liu, W., Gu, A., et al. (2014). Changes in Gut Microbiota May Be Early Signs of Liver Toxicity Induced by Epoxiconazole in Rats. *Chemotherapy* 60, 135–142. doi:10.1159/000371837.

Yoo, D.-H., Kim, I. S., Van Le, T. K., Jung, I.-H., Yoo, H. H., and Kim, D.-H. (2014). Gut microbiota-mediated drug interactions between lovastatin and antibiotics. *Drug Metab. Dispos. Biol. Fate Chem.* 42, 1508–1513. doi:10.1124/dmd.114.058354.

Zaneveld, J. R., McMinds, R., and Thurber, R. V. (2017). Stress and stability: applying the Anna Karenina principle to animal microbiomes. *Nat. Microbiol.* 2, 1–8. doi:10.1038/nmicrobiol.2017.121.
Zheng, X., Zhao, A., Xie, G., Chi, Y., Zhao, L., Li, H., et al. (2013). Melamine-induced renal toxicity is mediated by the gut microbiota. *Sci. Transl. Med.* 5, 172ra22. doi:10.1126/scitranslmed.3005114.

Zhou, J., Deng, Y., Luo, F., He, Z., Tu, Q., and Zhi, X. (2010). Functional molecular ecological networks. *mBio* 1. doi:10.1128/mBio.00169-10.

**Figures**

**Figure 1:** Ecotoxicology studies the effect of chemicals on organisms at the population level. A microbiome-aware ecotoxicology perspective acknowledges the importance of associated microorganisms in their hosts biology, at the level of individuals as well as populations. Indeed, the microbiome is interacting with hosts, environment and contaminants and may be linked with health status (here, light versus dark grey). The microbiome thus needs to be integrated as an element of the system, and protocols to investigate ecotoxicological effects at each level need to be adapted.
Figure 2: Sitting at the interface between environment and host, the microbiome may interact with contaminants. Sequestration, inactivation and degradation mitigate potential effects on host health, while activation or potentialisation reinforce the effect of contaminants. Microbiome composition, abundance and functions respond to exposure, and dysbiosis can occur. Post-exposure recovery leads to a new stable state, identical or altered compared to the pre-exposure state. Future lines of research are emphasized. Although contaminants may alter the host health through adverse outcome pathways, dysbiosis itself may also induce pathology and fitness loss, difficult to disentangle from each other.