Enzyme-linked immunosorbent assay of epidermal growth factor receptor in lung cancer: comparisons with immunohistochemistry, clinicopathological features and prognosis

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Summary The prognostic role of epidermal growth factor receptor (EGFR) remains controversial in patients with lung cancer. Previous assays for EGFR have primarily been qualitative or, at best, semiquantitative. In the present study, using fresh-frozen tissue from 190 unselected lung cancer patients, quantification of EGFR (EGFR±IHC) using a recently developed enzyme-linked immunosorbent assay (ELISA) technique was compared with results (EGFR±IHC) obtained using immunohistochemistry (IHC). Correlation between results obtained by the two different techniques was highly significant (r = 0.63, P < 0.001, n = 190). This correlation improved even further (r = 0.76) when sections were estimated using an IHC score that took into account percentage staining, intensity and relative tumour area. Furthermore, the relationship between clinicopathological features and prognosis was identical for the two methods. The expression of EGFR was highest in squamous cell carcinomas, but it was not correlated with other characteristics such as age, sex, histological grading, stage or prognosis. We conclude that evaluation of EGFR content using IHC and ELISA produces comparable results.

Keywords: epidermal growth factor receptor; enzyme-linked immunosorbent assay; immunohistochemistry, non-small-cell lung cancer; prognosis

Epidermal growth factor receptor (EGFR) is the protein product of the proto-oncogene HER-I. Ligand binding to the extracellular region of EGFR causes receptor dimerization, resulting in autophosphorylation and activation of cytoplasmic signal protein, which ultimately triggers DNA synthesis associated with proliferation and differentiation (Prigent and Lemoine, 1992). However, whether aberrant expression of EGFR is causally or consequentially related to the development of cancer remains to be established. Some studies have reported EGFR to be of prognostic importance in lung cancer (Volm and Mattern, 1993; Volm et al. 1993); unfortunately, the relationship to survival has been inconsistent (Pfeiffer et al. 1996a).

Immunohistochemistry (IHC) is the most widely applied technique to assess expression of EGFR in patients with non-small-cell lung cancer (NSCLC) (Pfeiffer et al. 1996a). Immunohistochemical results depend on the primary antibody, the visualization system and whether fixed or frozen tissues are used. Furthermore, the evaluation of immunohistochemical slides is inherently qualitative or, at best, semiquantitative. The enzyme-linked immunosorbent assay (ELISA) can quantify EGFR in cytosol extracts. We used a recently developed ELISA technique (Christensen et al. 1995) to measure the level of EGFR in a large cohort of lung cancer patients and compared the results with immunohistochemical analysis on cryosections and with clinical and pathological data. To our knowledge, this is the first study to compare the detection of EGFR using IHC and ELISA.

MATERIAL AND METHODS

Patients and tumour samples

Frozen tumour tissue from 190 previously untreated lung cancer patients (median age 61 years (range 42–79 years); 131 men and 59 women) were available for ELISA and IHC. All the patients were surgically treated at Odense University Hospital. Denmark, between 1984 and 1991: median follow-up was 66 months (range 39–119 months). The study was approved by the local ethics committee.

We analysed the relationship between EGFR and histological subtype and the correlation between IHC and ELISA for all lung cancer patients; however, all other analyses were restricted to 180 patients with NSCLC. The TNM stage of NSCLC was determined retrospectively, according to the new International Staging System for lung cancer (Mountain, 1987). No patient received post-operative adjuvant cytotoxic or radiation therapy.

Tissue preparation

Lung resections were received unfixed in the pathology laboratory, immediately after surgical removal. One piece of tumour measuring approximately 1 cm³ was cut out and divided in two. One part was placed in cryoconservation tubes, snap-frozen at –80°C, and stored until further examinations. The other part was formalin fixed and embedded in paraffin for histological classification.
ELISA

Biopsies were stored at -80°C until further analysis. We used a recently developed ELISA technique to measure EGFR content (EGFR ELISA) (Christensen et al. 1995). Briefly, biopsies (median 44 mg; range 5–192 mg) from 190 patients were cut, ultrasonically homogenized in buffer and centrifuged at 20 000 g. The membrane pellet was resuspended in buffer and the receptor was solubilized by incubation with 2% Triton X-100, followed by centrifugation at 20 000 g. EGFR was measured in the supernatant. We used EGFR1 (RPN 513; Amersham, UK) as catching antibody and biotinylated Ab-1 (GR01; Oncogene Science, USA) as detecting antibody. EGFR from placenta membranes was used as the calibration standard. The detection limit was 0.08 nmol l⁻¹.

Immunohistochemical analysis

EGFR immunostaining was performed on 5-μm cryostat sections, using the peroxidase labelled Streptavidin–Biotin (LSAB) technique and EGFR1 (Amersham, UK) as primary antibody (Pfeiffer et al. 1996a, b).

Immunohistochemical assessment

The percentage of positively reacting tumour cells (EGFR TUM: 0–100%) was estimated by one author (PP) (Pfeiffer et al. 1996a).

An immunohistochemical score (IHC score), which took into account percentage staining, intensity and relative tumour area, was also calculated. The average intensity of staining (INT SCO) was given a score between 0 and 3. The number of positively stained tumour cells (CELL SCO) was also scored between 0 and 3: 0 (0–19%); 1 (1–49%); 2 (50–79%); 3 (80–100%). In addition, the relative tumour area was assessed by evaluating the proportion of malignant cells compared with the entire histological section (AREA SCO) and expressed as a proportion (0–1.0). The IHC score was then
Table 1 Relationship between EGFR<sub>ELISA</sub> content and histological subtype in 190 patients with lung cancer.

|        | EGFR<sub>ELISA</sub> content |
|--------|-----------------------------|
|        | Low (%) | Medium (%) | High (%) |
| SqC    | 100     | 37        | 27       | 36       |
| AdC    | 57      | 74        | 10       | 16       |
| LaC    | 23      | 61        | 22       | 17       |
| Carcinoid | 9       | 100       | 0        | 0        |
| SCLC   | 1       | 100       | 0        | 0        |
| Total  | 190     | 54        | 20       | 26       |

NSCLC > SCLC + carcinoid, P < 0.001; SqC > AdC, P < 0.00001; SqC > LaC, P = 0.01. EGFR<sub>ELISA</sub> EGFR content measured using ELISA: SqC, squamous cell carcinoma; AdC, adenocarcinoma; LaC, large cell carcinoma; T, primary tumour; N, lymph node.

![Survival curve](image)

Survival (%)

Follow-up time (years)

Figure 3 Survival of 190 patients with NSCLC according to measurement of EGFR using ELISA (low, medium or high) (P = 0.6). ... low n = 89; ... medium n = 42; ... high n = 49

assigned to each tumour by multiplying INT<sub>SCO</sub>, CELL<sub>SCO</sub> and AREA<sub>SCO</sub>, the IHC score thus ranged from 0 to 9.0.

Statistical evaluation

All P-values are from two-sided tests. Statistical significance was claimed for P-values less than or equal to 0.05. Relative risk (RR) is followed by 95% confidence intervals (CI). The association between two variables was quantified using Spearman’s rank correlation coefficient (r<sub>S</sub>) test.

Survival curves were estimated according to the Kaplan–Meier method (Kaplan and Meier, 1958) and compared using the log-rank test (Peto et al, 1977) or, if there were three or more ordered categories, the log-rank test for trend. BMDP/PC Release 7.01 (BMDP Statistical Software, 1993, Los Angeles, CA, USA) was used for all statistical analyses.

RESULTS

The median EGFR<sub>ELISA</sub> was 1.0 nmol EGFR g<sup>-1</sup> protein (range 0.1–26.9 nmol EGFR g<sup>-1</sup> protein). This value was selected as a cut-off point and a second cut-off point (2.0 nmol EGFR g<sup>-1</sup> protein) was arbitrarily chosen (Figure 1A). The patients were thus categorized as low (< 1.0 nmol EGFR g<sup>-1</sup> protein), medium (1.0–2.0 nmol EGFR g<sup>-1</sup> protein) or high (> 2.0 nmol EGFR g<sup>-1</sup> protein).

The distribution of EGFR<sub>BK</sub> is shown in Figure 1B; the median EGFR<sub>BK</sub> was 80%. EGFR<sub>ELISA</sub> and EGFR<sub>BK</sub> results of paired samples from 190 patients with lung cancer are shown in Figure 2A. There was a highly significant correlation between EGFR<sub>BK</sub> and EGFR<sub>ELISA</sub> (r<sub>S</sub> = 0.63, P < 0.001). The association between ELISA and IHC was improved even further (r<sub>S</sub> = 0.76) when the IHC score was used, i.e. when we also took into account the intensity of staining and relative tumour area (Figure 2B).

The association between EGFR expression and other variables was similar whether we used results obtained using ELISA or IHC. We found expression of EGFR in all subtypes of NSCLC, but most frequently in squamous cell carcinomas (Table 1).

EGFR<sub>ELISA</sub> or EGFR<sub>BK</sub> was not correlated with age, tumour size, T status, lymph node involvement, stage, histological grading or time of diagnosis. Furthermore, squamous cell carcinoma was the only variable that correlated with EGFR expression in a logistic regression model. The relative risk for high EGFR<sub>ELISA</sub> and EGFR<sub>BK</sub> content for squamous cell carcinoma was 4.1 (95% CI 2.2–7.7) and 4.4 (95% CI 2.3–8.3), respectively.

We did not find any correlation between EGFR expression and survival in the entire group of patients with NSCLC (Figure 3), nor in any subgroup analysis.

DISCUSSION

Most studies on NSCLC have focussed on the protein level. Berger et al (1987) found a close correlation between IHC and auto-phosphorylation activity. Ligand-binding studies have shown that EGFR-binding characteristics were comparable in tumour and normal lung tissue (Hwang et al, 1986; Veale et al, 1989; Dittadi et al, 1991). These studies point to the fact that, if expressed in NSCLC, a normal functional EGFR is operating. Thus, this receptor can be detected by antibodies that recognize EGFR, using either ELISA or IHC.

It is often stated that patients with overexpression of EGFR have a shorter survival, but in a prior study (Pfeiffer et al, 1996a) we were unable to find a correlation between EGFR and prognosis. To substantiate these findings, we measured EGFR content in frozen tumour samples using a recently developed ELISA technique (Christensen et al, 1995). We found a highly significant correlation between EGFR<sub>BK</sub> and EGFR<sub>ELISA</sub>, and also an identical correlation with other variables. A high EGFR<sub>ELISA</sub> value always corresponded with a high EGFR<sub>BK</sub> value, whereas samples with a high EGFR<sub>BK</sub> showed variable values for EGFR<sub>ELISA</sub>. To interpret this discrepancy, one must realize that even although EGFR<sub>ELISA</sub> is a quantitative measure, ELISA calculates an average EGFR content in a homogenized tissue sample consisting of tumour tissue intermixed with various amounts of non-tumour tissue, including normal lung tissue, connective tissue and necrotic tissue. Another inconvenience is that it requires handling of fresh or frozen tissue. The major advantages of ELISA are its quantitative nature and the use of a calibration curve (Christensen et al, 1995).

Quantitation of EGFR has been reported in a few prior studies. Veale et al (1993) measured EGFR using the ligand-binding assay in 19 selected patients with NSCLC. They found that EGFR quantitation may give prognostic information and proposed confirmation in a larger prospective study. Recent studies have shown that...
EGFR content can be quantitated on tumour sections (Stanton et al. 1994; Robertson et al. 1996), but whether quantitative analysis will add to the prognostic significance of EGFR is doubtful. Furthermore, it is still complex to interpret results in heterogeneous tumours, i.e. what is the biological importance of a large amount of EGFR in few tumour cells compared with a lower amount in most tumour cells.

IHC represents a small sectional view of a larger tissue area; the result relies very much on the immunohistochemical technique, and the results are subjective and qualitative. The major advantage of the immunohistochemical technique is the maintenance of tissue architecture and in situ localization of the antigen. By contrast with ELISA, in which cancer cells cannot be distinguished from non-malignant tissue, this distinction can easily be made using immunohistochemical analysis. Also, IHC is also able to identify tumour positivity, even in very small tumour samples.

When we took into account the percentage of stained tumour cells, intensity of staining and cellularity, the correlation between ELISA and IHC was further improved. At least some of the remaining discrepancy may be due to intratumoral heterogeneity.

To our knowledge, a detailed comparison of EGFR expression using ELISA and IHC has not been published previously, but some studies have determined the related growth factor receptor p185HER-2 using IHC and ELISA (Dawkins et al. 1993; Dittadi et al. 1993; Nugent et al. 1994; Piffanelli et al. 1996). In brief, the overall agreement was comparable with that of the present study.

In agreement with most other studies, we found expression of EGFR (EGFR_{ILISA} or EGFR_{IHC}) in all subtypes of NSCLC, but most frequently in squamous cell carcinomas, and no correlation between EGFR expression and the size of the primary tumour, lymph node status or stage. The expression of EGFR without correlation to stage suggests an important step during early tumour genesis. It might provide the potential tumour cell with the ability continually to proliferate when the supply of growth factors is restricted and/or escape terminal differentiation.

In conclusion, detection of EGFR using IHC and ELISA produces comparable results, particularly when IHC is estimated using an immunohistochemical score that evaluates percentage staining, intensity and relative tumour area, however further methodological standardization is needed. Expression of EGFR was found in all histological subtypes of NSCLC, but especially in squamous cell carcinoma. Quantitative or qualitative EGFR expression was not correlated with expansion of tumour tissue or histological grading and was without prognostic value in patients with NSCLC.

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