Overview of telomeres and the use of telomerase to compensate for telomere loss

Human chromosomes contain repeated TTAGGG DNA sequences at their ends (telomeres) that provide genomic stability (protect the ends from being recognized as DNA breaks needing repair) and a source of expendable DNA (a solution to the inability of the replication machinery to copy the extreme ends of chromosomes: the end replication problem) (see [1] for a review). A significant amount of truncation can occur during DNA replication without causing deleterious effects. Human telomeres progressively shorten with ongoing cell division until they reach a critical length that induces replicative senescence [2].

Recent research has led to increased knowledge of the structure and function of telomeres. Telomeres end in G-rich single-stranded 3’ overhangs, and can form a lariat structure called a t-loop. Telomere binding proteins such as TRF1 and TRF2 are thought to protect against nucleases and prevent DNA recombination or end joining [1]. Recent research has shown an increasing connection between DNA damage repair proteins (eg Mre11/Rad50/ Nbs1, Ku, etc) and telomere biology in mammals. Double-strand break recognition and repair factors associated with the BRCA1-associated genome surveillance complex have, for instance, been linked to telomeres and telomere binding proteins [3]. BRCA1 or BRCA2 might also play a role in telomere structure and function.

Although other mechanisms to maintain telomere stability are possible, the mechanism for lengthening telomeres in humans is almost always by the reactivation or upregulation of telomerase [4]. Human telomerase is a protein complex consisting of a human telomerase reverse transcriptase catalytic subunit (hTERT) that uses the human telomerase RNA component (hTR) of the complex as a template for adding TTAGGG repeats to the end of the chromosome [5–7]. Telomerase is only expressed in a small number of proliferating cell types, such as germ line and somatic stem cells. Most normal human cells lack telomerase activity and their telomeres shorten with each cell division, until they enter replicative senescence. Cells that lose critical cell-cycle checkpoint functions escape this initial growth arrest and divide until they enter crisis

AZT = 3’-azido-3’-deoxythymidine; FNA = fine-needle aspirate; hTERT = human telomerase reverse transcriptase catalytic subunit; hTR = human telomerase RNA component; PCR = polymerase chain reaction; TRAP = telomeric repeat amplification protocol.
when chromosome end fusions and apoptosis occur. Cells remain in this crisis period unless a rare cell acquires a mechanism, such as telomerase expression, that can lengthen telomeres. As cells continue to proliferate, the maintenance of telomeres involves a collection of factors including telomere binding and associated proteins, the telomerase ribonucleoprotein complex and, potentially, DNA damage repair proteins. A rare cell that can maintain telomeres is then able to grow continuously (ie becomes immortal) and this is generally believed to be a critical step towards cancer progression [8].

**Telomerase as a diagnostic target: incidence of telomerase in breast cancer**

The development of the telomeric repeat amplification protocol (TRAP) assay led to an expansion in the ability to detect telomerase activity in human cancer cells [9]. This sensitive polymerase chain reaction (PCR)-based assay can detect as few as 1 to 10 positive cells or 0.01% in a mixed population. Although preliminary data showed 88% of all stages of breast carcinoma having positive TRAP [9], closer investigation and careful handling of initially negative samples revealed the value may be closer to 95% [10]. As reviewed by Shay and Bacchetti, 75% of breast carcinoma in situ lesions, 88% of ductal and lobular carcinomas, 5% of adjacent tissues, and none of the normal tissues were TRAP-positive [9]. Yashima et al detected a progressive increase in the mean telomerase levels with the severity of histopathological change: 14% in benign breast diseases, 92% in carcinoma in situ lesions, and 94% in invasive breast cancers [11]. Expression of the hTERT mRNA can also be detected using real-time quantitative reverse transcriptase-PCR, and this assay revealed a statistical link between hTERT mRNA levels and the aggressiveness of breast tumors [12]. Both this semi-automated assay and the TRAP assay provide suitable methods for breast cancer diagnosis, but should be used in conjunction with other diagnostic tools to rule out false results.

Detection of telomerase activity in preoperative specimens, such as in fine-needle aspirates (FNAs), may improve diagnostic accuracy [13,14]. FNA cytology is known to be accurate, cost effective and have minimal risk [14]; however, difficulties still occasionally occur using cytology alone. Two groups separately compared the diagnostic utility of telomerase assays of FNAs with cytology preparations [13,14]. Poremba et al showed that 92% of FNAs from breast cancer patients were telomerase-positive, 94% of FNAs from patients with benign breast lesions were telomerase-negative (the positive cases were all fibroadenomas), and there was a strong correlation between TRAP and histologic diagnosis of atypia [13]. Hiyama et al observed that all atypical or intermediate cases with detectable telomerase activity in the FNAs were found to be carcinomas after surgery [14]. Furthermore, six out of seven tumors without telomerase activity were diagnosed as benign, while one half of the cases with detectable telomerase activity, initially designated by cytology as benign, were subsequently diagnosed as cancer. Detecting telomerase activity in FNAs is thus equivalent, if not better, than detection by cytology [14], and can be used in conjunction with other diagnostic tests. Finally, tumor-derived telomerase RNA found in the serum of breast cancer patients may have implications in diagnosis and in follow-up monitoring studies [15].

**Telomerase activity and prognosis in breast cancer**

With the increasing number of breast cancers detected by screening procedures, a marker is needed to stratify the risk of subsequent invasive cancer. Hoos et al found a significant correlation between telomerase activity and tumor size, lymph node status, and stage [16]. A significant association between telomerase-positive infiltrating breast carcinomas and lymphovascular invasion, a fundamental step in breast cancer metastasis and a predictor of survival, has also been observed, making telomerase a useful prognostic marker [17]. Clark et al reported, in a prognostic study involving 398 patients with lymph node-positive breast cancer, that increased telomerase activity was associated with decreased disease-free survival [18]. High telomerase activity in breast cancer is more-over associated with genetic aberrations in 3q (gain), 8q (gain), and 17p (deletion) [19]. These aberrations are common in breast cancers and involve the hTR (on 3q), c-myc (on 8q), and p53 (on 17p) genes, all of which have been associated with telomerase regulation [19]. Understanding the link between telomerase activity and genetic changes associated with breast cancer remain an important area of research today.

**Telomerase inhibition as an anticancer approach**

The average telomere length in breast cancer cells is usually well below that of normal cells. This difference in telomere length coupled with the more rapid rate of cell division in cancer cells makes the inhibition of telomerase an attractive potential breast cancer therapeutic target. Treatment with telomerase inhibitors may not have the toxicity found with other chemotherapeutic agents since telomerase is absent in most somatic cells (Fig. 1). While normal, proliferating telomerase-positive stem cells may also initially be affected, their telomeres are well above the critically short length that induces a DNA damage/growth arrest mechanism. Furthermore, most stem cells are quiescent, and telomere shortening normally only occurs with cell division. Since most breast cancer cells have very short telomeres, treatment with telomerase inhibitors should lead to growth arrest and cell death.
Advances in development of telomerase inhibitors

The telomerase protein complex allows for multiple sites for inhibition. Recent research includes targeting the RNA component of the telomerase (antisense oligonucleotides, hammerhead ribozymes), inhibition of the catalytic subunit (dominant negative mutant hTERT, reverse transcriptase inhibitors), immunotherapy, and small molecule inhibitors (reviewed in [20]). It is important in most approaches to confirm that telomerase inhibitors are acting specifically in a telomere-dependent mechanism. First, telomerase inhibitors should almost always lack immediate toxicity to the cell; they should reduce telomerase activity without initially compromising cellular proliferation. Second, without telomerase activity, telomeres should progressively shorten with each cell division. Cells should ultimately die or undergo growth arrest and the time required should be related to initial telomere length. Over 100 manuscripts have been published on telomerase inhibitors but only a small number meet the presented criteria [20], and fewer still have been tested in breast cancer cells.

Antisense oligonucleotides, dominant negative mutant hTERT and reverse transcriptase inhibitors have been studied in breast cancer [20–22]. The most widely used reverse transcriptase inhibitor is 3’-azido-3’-deoxythymidine (AZT) (reviewed in [20]). Melana et al observed that AZT inhibited the growth of breast cancer cells and telomerase activity at lower doses than in normal breast cells [21]. Multani et al, however, showed that AZT reduced the telomeric signals, as detected by fluorescence in situ hybridization, in human MCF-7 breast carcinoma cells within 72 h [22], a time too short to be explained by the inhibition of telomerase and progressive telomere shortening. The effects of AZT, while effective in inhibiting breast cancer cell growth in these studies, may not be due to a specific telomere-dependent mechanism, and telomerase inhibition may just be a side effect [20]. Whereas further research is needed to prove the specificity of AZT, studies using both antisense oligonucleotides, such as 2’-O-methyl RNA directed against the hTR template region, and dominant negative mutant hTERT have been shown to be effective specific telomerase inhibitors in immortalized breast epithelial and breast carcinoma cells in vitro (reviewed in [20]).

Standard chemopreventive agents have also been investigated for a possible role in telomerase inhibition and control of breast cancer. Retinoic acid and tamoxifen have
already been shown to inhibit telomerase activity in breast cancer cells [23,24], probably as a secondary consequence of inhibition of proliferation. While these agents are known for their antiproliferative effects, not much is known on their effects on immortalization, a critical step towards breast cancer progression [8]. We have recently investigated the possibility of using these agents or specific telomerase inhibitors to prevent cellular immortalization as a breast cancer chemoprevention strategy. We found that long-term treatment of precrisis breast epithelial cells with nontoxic doses of either chemopreventive or antitelomerase agents significantly lowered the frequency of spontaneous immortalization [25]. Our studies provide a new model for the screening of novel chemopreventive agents that target cellular immortalization.

Conclusion
Telomerase is an attractive target for diagnosis and therapy since it is expressed in over 90% of breast cancer cells, while it is not expressed in most normal cells. Whereas most of the progress on the role of telomerase in breast cancer has been in diagnostics, research into telomerase inhibitors is increasing. Inhibition of telomerase in vitro leads to progressive telomere shortening, eventually resulting in growth arrest or cell death due to the critically short telomeres inducing a DNA damage response. Breast cancer cells with already short telomeres would be most affected by telomerase inhibitors, whereas normal stem cells with longer telomeres would be relatively resistant. The effects of telomerase inhibitors would depend on initial telomere length and rate of cell division, and it may take weeks to months to see changes in tumor size. Combining telomerase inhibitors with current therapies to reduce tumor burden may thus provide a better regimen to target breast cancer and prevent recurrence.

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