Plasma p217+tau versus NAV4694 amyloid and MK6240 tau PET across the Alzheimer’s continuum

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Abstract

Introduction: We evaluated a new Simoa plasma assay for phosphorylated tau (P-tau) at aa217 enhanced by additional p-tau sites (p217+tau).

Methods: Plasma p217+tau levels were compared to 18F-NAV4694 amyloid beta (Aβ) positron emission tomography (PET) and 18F-MK6240 tau PET in 174 cognitively impaired (CI) and 223 cognitively unimpaired (CU) participants.

Results: Compared to Aβ− CU, the plasma levels of p217+tau increased 2-fold in Aβ+ CU and 3.5-fold in Aβ+ CI. In Aβ− the p217+tau levels did not differ significantly between CU and CI. P217+tau correlated with Aβ centiloids $P = .67$ (CI, $P = .64$; CU, $P = .45$) and tau SUVRMT $P = .63$ (CI, $P = .69$; CU, $P = .34$). Area under curve (AUC) for Alzheimer’s disease (AD) dementia versus Aβ− CU was 0.94, for AD dementia versus other dementia was 0.93, for Aβ+ versus Aβ− PET was 0.89, and for tau+ versus tau− PET was 0.89.
1 | INTRODUCTION

Assessing brain levels of amyloid beta (Aβ) plaque and tau aggregation has a central role in the selection of individuals for clinical trials and assessing the efficacy of therapeutic compounds. It may also aid earlier and more accurate diagnosis of Alzheimer’s disease (AD).  

Despite the extremely low concentration of Aβ and tau in plasma, progress in detection methodologies has recently allowed reliable measurement of plasma Aβ42/Aβ40 and phosphorylated tau (P-tau). In a recent report, the accuracy of plasma Aβ42/Aβ40 to predict the presence of Aβ in the brain using immunoprecipitation–mass spectrometry (IMS) assays was equal to 90%. However, despite the promising potential of IMS to provide an Aβ biomarker, it is expensive and difficult to widely deploy. Consequently, measures of plasma P-tau have recently received much attention. Measurement of plasma tau phosphorylated at threonine 181 (P-tau181) was the first P-tau to demonstrate good accuracy. P-tau181 can differentiate AD from cognitively unimpaired (CU) subjects and from other dementias. P-tau181 was also a good predictor of elevated brain Aβ and tau as measured by positron emission tomography (PET). Subsequently, plasma P-tau217 and P-tau231 have also shown strong association with AD pathology with accuracy comparable to cerebrospinal fluid (CSF) and PET measures, with some evidence of superior performance to plasma P-tau181.

More recently a high-sensitivity Simoa assay has been developed using a capture antibody (pT3) that was raised against tau in paired helical filaments (PHFs) of AD brain. The core requirement for this antibody binding is phosphorylation at aa217 (p217), with enhanced binding when other nearby phosphorylated sites are present, predominantly at aa212. The recognized epitope is thus referred to as “p217+tau” and its measurement in plasma has demonstrated good concordance with CSF markers of AD in a validation cohort of 227 subjects with an area under the curve (AUC) of 0.90 versus CSF Aβ42/40 and 0.95 versus CSF P-tau181.

Plasma p217+tau has not been compared to PET measures. Aβ PET scans were spatially normalized using CapAIBL and the standard Centiloid (CL) method was applied using the standard whole cerebellum mask as reference region. A CL value of 25 was selected to determine a positive Aβ PET scan (Aβ+). For the main analysis but Aβ PET thresholds between 15 and 50 CL were also investigated. Aβ PET agents in participants of the Australian Imaging, Biomarkers and Lifestyle study of aging (AIBL) and the Australian Dementia Network (ADNeT) trial screening program. To investigate both the preclinical trial screening and clinical diagnostic performance, we examined CU and cognitively impaired (CI) subjects separately and varied the threshold for Aβ PET from 15 to 50 centiloids (CLs).

2 | METHODS

2.1 | Participants

Three hundred ninety-seven participants in the AIBL and ADNeT cohorts had both 18F-NAV4694 Aβ PET and 18F-MK6240 tau PET. The AIBL cohort recruitment and evaluation is detailed elsewhere. The ADNeT cohort all had mild cognitive impairment (MCI) or mild dementia and was evaluated as per AIBL. A multi-disciplinary panel, blind to all imaging and blood results, classified all participants as CU or CI with MCI, AD, or non-AD dementia. A diagnosis of CU required performance within 1.5 standard deviation (SD) of the published norms for their age group on neuropsychological assessment. A diagnosis of MCI or dementia was assigned according to the criteria of Winblad et al. and Petersen et al. For this study, only individuals with a clinical diagnosis of AD and a positive Aβ PET scan were classified as AD. Those with dementia but a negative Aβ scan were classified as non-AD dementia.

Institutional ethical review committees approved the AIBL and ADNeT studies, and written informed consent was obtained from all participants.

2.2 | Image acquisition

A 20-minute Aβ PET acquisition was conducted 50 minutes post-injection of 200 MBq of 18F-NAV4694. On a separate day, a 20-minute tau PET acquisition was conducted 90 minutes post-injection of 185 MBq of 18F-MK6240.

Aβ PET scans were spatially normalized using CapAIBL and the standard Centiloid (CL) method was applied using the standard whole cerebellum mask as reference region. A CL value of 25 was selected to determine a positive Aβ PET scan (Aβ+) for the main analysis but Aβ PET thresholds between 15 and 50 CL were also investigated. 18F-MK6240 tau PET scans were spatially normalized using...
a CapAIBL principal component analysis (PCA)-based approach and scaled using the cerebellar cortex as the reference region. 18F-MK6240 tau standardized uptake value ratio (SUVR) values were estimated in two composite regions of interest (ROIs); a mesial temporal ROI (Me) comprising entorhinal cortex, hippocampus, parahippocampus and amygdala; and a meta temporal ROI (MT) composed of the Me ROI plus inferior and middle temporal and fusiform gyri. We also estimated 18F-MK6240 SUVR in Braak stages IV, V, and VI with ROIs derived from the Freesurfer Desikan-Killiany Atlas (see Figures S4-S5). In the CU group, we additionally estimated 18F-MK6240 SUVR in the entorhinal cortex, amygdala, hippocampus, and inferior temporal cortex separately. These regions were selected as we aimed to assess the early spectrum of Braak stages in CU. The threshold for tau positivity was the 95th percentile of the Aβ– CU group in each composite ROI and individual mesial temporal regions.

2.3 Plasma analysis and p217+tau assay design

Fasted blood sampling was performed 3.3 ± 6 months from the time of Aβ PET scan and 1.7 ± 2 months from the tau scan. Plasma from K2-EDTA tubes (7.5 mL S-movette 01.1605.008, Sarstedt) containing pre-added prostaglandin E1 (33 ng/mL of whole blood, Sapphire Biosciences) to prevent platelet activation, a potential source of peripheral Aβ, was centrifuged at room temperature at 200g for 10 minutes to collect platelet-rich plasma, and then at 800g for 10 minutes to provide plasma that was snap frozen within 2 hours of collection and stored in vapor phase liquid nitrogen prior to shipping on dry ice from Australia to Janssen R&D (La Jolla, CA, USA). Plasma ptau217+ assay was performed on a Single MOlecular Array (Simoa) HD-X platform blinded to all subject data using the technique described previously. Samples were measured in duplicate, yielding average precision of 12% CV (87% of the samples measured with CV < 20%). Functional assay range (after accounting for sample dilution) was: lower limit of detection (LLOD) = 1 fg/mL, lower limit of quantification (LLOQ) = 10 fg/mL, upper limit of quantification (ULOQ) = 20,000 fg/mL. All samples were >LLOD. Data analysis was then performed by AIBL investigators.

2.4 Statistical analysis

Demographic and clinical characteristics were compared between Aβ status within clinical classifications using standard generalized linear modeling (GLM) for quantitative features and the chi-square test for categorical comparisons. ROI, vertex, and voxel-wise correlation between the plasma p217+tau and CL or tau SUVR was performed using Spearman’s rank-based correlation (Spearman’s Rho [ρ]). Receiver-operating characteristic (ROC) AUC analyses were used to explore the discriminative performance of p217+tau. Optimal p217+tau thresholds for detecting Aβ+ and tau+ PET were calculated both using Youden’s index and two SD above the mean p217+tau levels in Aβ– CU group. In addition, 95% CIs were shown in square brackets and positive and negative predictive values (PPV, NPV) were not disease prevalence adjusted. Model based performance of p217+tau accounting for age, gender, and APOE ε4 allele status was done using GLM. AUC values between models were compared using DeLong’s method. CIs were computed using bootstrap. False discovery rate (FDR) corrected P-values <.05 were considered statistically significant.

3 RESULTS

3.1 Demographic and clinical characteristics

Three participants had a low CL but high Braak stage V/VI tau on 18F-MK6240 PET, suggesting that, similar to some PiB case reports, the Aβ PET results were falsely negative. These three subjects all showed clearly elevated plasma p217+tau levels and were kept in the analysis but are displayed with a diamond shape in the scatter plots.

The demographic and clinical characteristic of the cohort are presented in Table 1. Of the 397 participants, 56% were CU, 22% had MCI, and 21% had dementia, of which 82% in the dementia group were Aβ+.

Overall, 170 (43%) had Aβ+ PET and 143 (36%) had high tau PET in the meta temporal region (TMT+). Participants with dementia were ≈4 years younger than those in the MCI and CU groups (P < .05).

Stratification by Aβ status within each clinical classification is provided in Table S1.
TABLE 1  Demographic and clinical characteristics

|                | CU (n = 223) | MCI (n = 91) | Dementia (n = 83) |
|----------------|--------------|--------------|------------------|
| Age            | 75.2 (5.70)  | 73.6 (7.99)  | 70.7 (7.91)*     |
| Gender, N male (%) | 101 (45.3%) | 51 (56.0%)  | 49 (59.0%)       |
| Education (years)  | 14 (3.0)    | 12.5 (3.1)* | 12 (3.0)*        |
| APOE ε4+ (%)    | 72 (32.3%)  | 46 (50.5%)* | 46 (55.4%)*      |
| MMSE, median (IQR) | 29 (2.00)   | 27 (4.00)*  | 23 (4.00)**-**   |
| CDR SoB, median (IQR) | 0.0 (0.00)  | 1.0 (1.50)* | 4.0 (1.00)**      |
| AIBL PACC, mean (SD) | −0.4 (0.84) | −2.4 (1.12)* | −4.3 (2.39)**     |
| Centiloid, mean (SD) | 19.5 (38.11) | 72.8 (66.16)* | 93.9 (55.58)**   |
| Aβ+, N (%)     | 46 (20.6%)  | 56 (61.4%)* | 68 (81.9%)*      |
| MK6240 SUVR_{MT}, mean (SD) | 1.02 (0.23) | 1.48 (0.61)* | 1.93 (0.77)**      |
| Tau_{MT}, N (%) | 31 (13.9%)  | 48 (52.7%)* | 64 (77.1%)*       |
| Adjusted HV (SD) | 2.98 (0.26) | 2.78 (0.40)* | 2.57 (0.42)**     |
| Plasma p217+tau, mean (SD) | 87.8 (67.8) | 183.7 (141.0)* | 229.9 (150.7)**   |

Abbreviations: CU, cognitively unimpaired; MCI, mild cognitive impairment. the total number of individuals (N), positive Aβ scan (Aβ+), SUVR in the Meta Temporal region (SUVR_{MT}). P-value < .05 compared to CU. *P-value < .05 compared to MCI.

3.2 | Plasma p217+tau levels in clinical groups by Aβ PET status

Figure 1A shows the concentration of plasma p217+tau in the different clinical groups, subdivided into Aβ− and Aβ+. The mean p217+tau concentration in CU Aβ+ participants (162.7 fg/mL [SD: 85.76]) was approximately twice that of the CU Aβ− participants (71.2 fg/mL [SD: 46.63]; P < 10−18). Mean p217+tau concentration in MCI Aβ+ (247.1 fg/mL [SD: 141.23]) was not significantly different from those with Aβ+ dementia (AD) (260.5 fg/mL [SD: 148.43]) but both groups showed significantly higher concentration than the CU Aβ− group and the CU Aβ−, MCI Aβ−, and dementia Aβ− groups. Levels of p217+tau in the MCI Aβ− (82.1 fg/mL [SD: 55.81]) and the Aβ− non-AD dementia (93.4 fg/mL [SD: 54.97]) participants were not significantly higher than CU Aβ− participants.

When subdividing the CL scale into five levels (Figure 1B), the ptau217+tau concentration was progressively higher as CL values rose from 25 CL. This progressive increase remained in those who were tau PET negative (see Figure S1). Figure 1C shows the CL level with increasing p217+tau level and Figure 1D shows the probability of having a positive Aβ scan versus p217+tau level.

3.3 | Correlations of plasma p217+tau with Aβ PET and tau PET

Vertex-based analysis (Figure 2 and Figures S7-S11) and scatter plots (Figures 3-5) demonstrate moderate to strong correlations between plasma p217+tau and Aβ and tau PET. The correlation in all subjects between p217+tau and Aβ PET CL gave a Spearman’s ρ of 0.67 (P < 10−53). The correlation was significantly stronger in CI than in CU (CI: ρ = 0.64, P < 10−21; CU:ρ = 0.45, P < 10−12; Z-score difference 2.6).

The correlation between p217+tau and tau SUVR was moderate to strong in the meta temporal ROI (Spearman ρ = 0.63, P < 10−46) and the mesial temporal ROI (Me) (Spearman ρ = 0.60, P < 10−40), but was progressively lower when the ROI was restricted to higher Braak stage regions. In CI subjects, the correlation was again strongest in the meta temporal ROI (ρ = 0.69). In CU the correlation was lower; meta temporal ROI (ρ = 0.34); mesial temporal ROI (ρ = 0.33), whereas Figure 2 shows that the regional correlation matches the early areas of tau accumulation as seen in the CU and the predominant regions of tau accumulation as shown in the CI. The correlations persisted between ptau217+ and tau SUVR_{MT} when analysis included only the Aβ+ participants (ρ = 0.52 in CI; ρ = 0.37 in CU). There was no correlation between p217+tau and tau PET in Aβ− individuals (P > .05). Surface projection of the Pearson correlation is also available in Figure S7.

3.4 | Performance of plasma p217+tau versus Aβ PET

For discriminating between the diagnostic groups, the AUC of p217+tau for Aβ+ AD versus Aβ− CU was 0.94 [0.90 to 0.97] and for Aβ+ AD versus Aβ+ CU was 0.67 [0.57 to 0.77]. For Aβ+ AD versus all CU was 0.88 [0.84 to 0.92] and for Aβ+ AD versus Aβ− dementia the AUC was 0.93 [0.90 to 0.96].
For discriminating Aβ+ PET from Aβ− PET in the entire cohort, the AUC was 0.89 [0.86 to 0.93], or 0.90 if the three Aβ−/tau+ PET individuals (all three had elevated plasma p217+tau) were removed. The Youden index provided a threshold concentration of 126.7 fg/mL [100.0 to 134.4 fg/mL] that yielded an accuracy of 0.85, sensitivity 0.79 [0.75 to 0.89], specificity 0.89 [0.80 to 0.92], PPV 0.84, and NPV 0.85. P217+tau accurately identified Aβ+ individuals among CI participants (AUC = 0.89 [0.84 to 0.93]) and among CU (AUC = 0.84 [0.76 to 0.91]). The Youden threshold was also 126.7 fg/mL [97.4 to 177.2] for the CI (Figure 3), giving a sensitivity of 0.82 [0.66 to 0.92] and a specificity of 0.82 [0.75 to 0.98], PPV 0.92, and NPV 0.65. Youden index provided an optimal threshold of 100.3 fg/mL [100.3 to 137.3] for CU participants (Figure 3), with a sensitivity of 0.80 [0.78 to 0.95], specificity of 0.83 [0.67 to 0.90], PPV 0.49, and NPV 0.95. Applying the whole cohort p217+tau threshold of 126.7 fg/mL to CU gave a sensitivity of 0.72 [0.6 to 0.83], specificity of 0.90 [0.86 to 0.94], PPV 0.66, and NPV 0.92.

The Z-score threshold for p217+tau derived from CU with <15 CL on Aβ PET was higher at 164 fg/mL, resulting in lower sensitivity of 0.60, higher specificity of 0.96, and PPV 0.91, and NPV 0.76 for all subjects to detect positive Aβ PET. In CI, the higher threshold gave sensitivity 0.66, specificity 0.94, PPV 0.96, and NPV 0.53. In CU it gave sensitivity 0.43, specificity 0.96, PPV 0.74, and NPV 0.87.

Altering the CL threshold for Aβ+ PET between 25 to 50 CL had no effect on AUC. However, at 15 and 20 CL thresholds, the AUC decreased due to more p217+tau false-negative results (Figure 3).

3.5 | Performance of plasma p217+tau versus 18F-MK6240 tau PET

Plasma p217+tau accurately discriminated individuals with elevated tau in all examined ROIs, with the highest AUC in the meta temporal ROI (AUC = 0.89 [0.86 to 0.92]). The scatter plots and ROC curves are shown in Figure 4A and C for the mesial temporal and meta temporal ROI, respectively. The Youden index thresholds were 126.7
fg/mL (Figure 4 B and D), the same as for Aβ PET, and provided an accuracy of 0.81, sensitivity of 0.80 [0.71 to 0.93], and specificity of 0.82 [0.71 to 0.93]).

In CU, the highest AUC (0.88 [0.83 to 0.92]) was observed in the meta temporal region with a Youden index threshold of 97.8 fg/mL (Figure 4F), giving a sensitivity of 0.9 [0.77 to 0.97] and specificity of 0.75 [0.71 to 0.88]. Applying the 126.7 fg/mL threshold to the CU group gave a sensitivity of 0.71 [0.51 to 0.84] and specificity of 0.85 [0.80 to 0.89]. Correlation between p217+tau and tau PET was only significant (P < .05) in the CU subjects who were Aβ+ and was strongest in the meta temporal ROI (P = .37) (Figure 4E Aβ+ [dark blue] and Aβ− CU [light blue]).

In CI, the highest AUC were observed in the meta temporal and the inferior temporal regions, with AUCs of 0.86 [0.81 to 0.91] (Figure S3). The mesial temporal AUC was lower at 0.81 [0.74 to 0.88]. The Youden index threshold for the meta temporal ROI provided a threshold of 148.4 fg/mL, with a sensitivity of 0.73 [0.65 to 0.83] and specificity of 0.90 [0.82 to 0.97]. Applying the threshold of 126.7 fg/mL to the CI cohort gave a sensitivity of 0.82 [0.77 to 0.89] and a specificity of 0.71 [0.6 to 0.79].

3.5.1 p217+tau versus tau PET in subregions of the mesial temporal in CU

The AUC for p217+tau to detect regional tau+ PET in the CU was higher for the meta temporal region at 0.88 [0.83 to 0.92] than the amygdala 0.81 [0.74 to 0.88] (P = .2), hippocampus 0.81 [0.74 to 0.88], parahippocampus 0.86 [0.77 to 0.93] (P = .2), or entorhinal cortex 0.78 [0.71 to 0.83] (P = .01). Plotting of the data points (see Figure S13) shows that discordance was due predominantly to p217+tau+/tau PET− subjects with a disproportionately high prevalence of Aβ+ in this category compared to p217+tau−/tau PET−.

3.6 p217+tau predicts Aβ and tau pathology independent of confounders

AUC values from base models incorporating age, sex, and APOE ε4 allele status to predict Aβ+ and meta temporal tau+ PET were 0.72 [0.67 to 0.77] and 0.69 [0.64 to 0.76], respectively. Adding p217+tau to the base model significantly improved prediction (P-value via DeLong's
FIGURE 3 Plasma p217+ tau versus Centiloid measures of amyloid beta (Aβ). Scatter plot and receiver-operating characteristic (ROC) curve (A & B), with ROC curves using different CL thresholds to define Aβ+ PET; full cohort (A & B), cognitively unimpaired sub-cohort (C & D) and cognitively impaired sub-cohort (E & F). Clinical groups are color-coded with red for dementia, green for mild cognitive impairment (MCI), and blue for cognitively unimpaired (CU). Solid circles are tau PET positive (TMT+). The black dashed horizontal line corresponds to the p217+ tau threshold derived from the Youden index. In (C) and (D) the CU specific threshold is shown. The cognitively impaired (CI) group threshold was the same as the whole cohort threshold. The diamond shapes are the three Aβ−/TMT+ subjects ROC test <.0001) compared to the base model alone, and produced an AUC of 0.91 [0.86 to 0.93] for Aβ+ PET and an AUC of 0.89 [0.86 to 0.92] for meta temporal tau+ PET. The combined model, however, did not perform significantly better than p217+ tau alone that gave an AUC of 0.89 [0.86 to 0.93] to predict Aβ+ and 0.89 [0.86 to 0.92] for meta temporal tau+ PET.

3.7 | Modeling of 18F-MK6240 SUV and p217+tau as a function of CL

To unveil the chronological order of the emergence of elevation in biomarkers, we normalized 18F-MK6240 SUV and p217+ tau concentration to the same scale, using two different methods and mapped
FIGURE 4  Plasma p217+tau versus PET SUVR measures of tau. Scatter plot and ROC curve for mesial temporal regions of interest (ROI) (A & B) and meta temporal ROI (C & D) and in cognitively unimpaired (CU) alone (E & F). Clinical groups are color-coded with red for dementia, green for mild cognitive impairment (MCI), and blue for CU. Solid circles are Aβ PET positive. The black dashed horizontal line corresponds to the p217+tau threshold derived from the Youden index. Linear correlation and Spearman co-efficient are shown for the Aβ+ (dark blue) and Aβ− CU (light blue) in part E.
FIGURE 5  Modeling of $^{18}$F-MK6240 quantification and p217+tau as a function of CL using polynomial curves; (A) normalized by linear transform of p217+tau levels and each tau PET, regions of interest (ROIs), SUVR to a scale of zero to 100 where zero is the mean of the results for each marker in Aβ− CU (<15 CL) and 100 is the mean of the results for each marker from the 30 individuals with the highest values for each marker. (B) Normalized by Z-score using the results from Aβ to cognitively unimpaired (CU) to define the normal range for p217+tau level and each tau PET, ROI, SUVR. The horizontal line is +2 standard deviations (SD) (A) suggests that plasma p217+tau increases early, similar to amygdala tau at low levels of Aβ, whereas (B) shows that the wide normal range for p217+tau delays reaching a two SD threshold for significance.

against CL. Aβ+ PET level were used as a surrogate for duration of disease development, since abnormal accumulation of Aβ as measured by PET is the first hallmark pathology of AD and continues at a relatively slow and constant rate over decades. First, we linearly normalized the range of biomarkers from zero to 100, with zero being the mean of the CU Aβ− individuals with <15 CL to increase certainty of Aβ negativity, and 100 being the mean of the 30 individuals with the highest biomarker value. When normalizing the range of values between 0 and 100, (Figure 5A), the fitted polynomial curves of $^{18}$F-MK6240 in amygdala and plasma p217+tau were nearly identical and rose very early when Aβ level was quite low and before other tau PET regions began to rise. Second, we normalized each biomarker to a Z-score, again using those CU with <15 CL to provide the normal range. Figure 5B shows that although we have demonstrated in Figure 5A that plasma p217+tau rises early, it takes until 70 CL to exceed 2 SD of the normal range.

4 | DISCUSSION

We determined the performance of a plasma P-tau assay that utilizes an antibody raised against the PHFs of AD. Human PHF tau is phosphorylated at multiple sites. Plasma p217+tau measures tau that is phosphorylated at aa217 with binding enhanced by the simultaneous presence of phosphorylation at aa212, so to may better reflect the tau of AD than single-binding site p-tau assays. To evaluate this assay, we employed PET tracers, which may give greater sensitivity than those previously used for comparison to plasma P-tau. Plasmata p217+tau displayed high accuracy in detecting Aβ+ individuals across the clinical spectrum. Compared to Aβ− PET CU, plasma p217+tau concentration was 2-fold higher in Aβ+ CU and 3.5-fold higher in Aβ+ MCI and dementia. The AUC for Aβ+ AD versus Aβ− CU was 0.94, for Aβ+AD versus Aβ− dementia was 0.93, while the AUCs for all Aβ+ versus Aβ− and for tau+ versus tau− were equal at 0.89. In the CU group, the AUC was 0.84 for Aβ+ versus Aβ− PET and 0.88 for tau+ versus tau− PET. Adding age, sex, and APOEε4 did not significantly improve the prediction of Aβ+ PET (AUC 0.89 vs 0.91) or tau+ PET (unchanged at 0.89). These results compare favorably to other plasma P-tau assays, although the use of different assay methods, different anti-tau antibodies, and cohort variation prevents direct comparison. The range of AUC reported for Aβ+ versus Aβ− PET for plasma P-tau measures has ranged from 0.76 to 0.92. Wide intersubject variability in plasma P-tau in persons without evidence of AD pathology on amyloid PET remains a challenge and warrants further investigation.

Our data (Figures 3–5) suggest that plasma p217+tau begins to rise (1) soon after brain Aβ levels begin to trend up as assessed by PET, (2) concordant with the rise in tau in the amygdala region on PET, and (3) prior to the rise in meta temporal tau PET. This is supported by the vertex-based analysis of correlation in CU, that p217+tau and Aβ are highly associated in brain regions where early Aβ deposition occurs and with tau in the anteromedial temporal lobe (Figure 2). However, the plasma p217+tau level does not reach +2 SD above the mean found in Aβ− CU until Aβ reaches moderate levels (≥70 CL). These findings are consistent with reports for plasma P-tau measures of p181, p217, and p231. Wide intersubject variability in plasma P-tau in persons...
of tau PET and plasma P-tau measures to detect PART is unclear and requires further investigation.

Only longitudinal studies will determine if what appear to be false positive p217+tau results in CU compared to PET are due to variation and limitation in the plasma measure or are detecting very early stage AD prior to PET becoming significantly positive, and this work is in progress under the AIBL study umbrella. In contrast to the CU, the CI show more “false-negative” p217+tau results when compared to both Aβ and tau PET. Many of these false-negative p217+tau results were A+/T+ on PET, so this requires further investigation.

Reflecting the impact of disease prevalence in our study population (71% Aβ+ in CI vs 21% in CU), the p217+tau positive and negative predictive values of p217+tau for Aβ+ PET were very different for the CI and CU cohorts. The PPVs were high in the CI group for predicting Aβ (0.94) and tau (0.93), suggesting that p217+tau may be a good test to confirm AD in patients with objective cognitive impairment. However, the NPVs were relatively low at 0.66 and 0.65, suggesting that a negative p217+tau do not exclude AD. Lowering the p217+tau threshold to 100 fg/mL in the CI group only improved the NPV to 0.73, whereas lowering the PPV to 0.88 and at 75 fg/mL, the NPV was 0.78 and PPV 0.84. P217+tau has potential as a diagnostic tool and in combination with other biomarkers, as discussed by Schindler et al., suggesting that a negative p217+tau can largely exclude AD pathology in persons with normal cognitive test results. However, the associated lower PPV (0.50) in CI indicates that positive results would need confirmation by PET or CSF analysis. These findings show that triage with p217+tau may substantially reduce the number of screening Aβ PET studies needed to identify Aβ+ PET participants for preclinical AD trials, resulting in considerable recruitment cost savings.

This study has several limitations. First the results need validation in other cohorts and more diverse populations. Second, this is a cross-sectional study, and longitudinal analysis is required to confirm the predictive value of p217+tau as a marker of disease progression and to clearly determine when abnormal levels of p217+tau can be measured along the disease trajectory.

In conclusion, an elevated level of plasma p217+tau is associated with both elevated Aβ and tau across the clinical spectrum of AD. Elevated p217+tau strongly supports a diagnosis of AD in persons with MCI or dementia, whereas a low level in CU persons is strong evidence against preclinical AD.

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ETHICS APPROVAL AND CONSENT TO PARTICIPATE/PUBLICATION
This study was approved by the Austin Health Human Research Ethics Committee (HREC/18/Austin/201). All participants gave written consent for publication of de-identified data.

CONFLICTS OF INTEREST
Christopher C. Rowe has received research grants from NHMRC, Enigma Australia, Biogen, Eisai, and Abbvie. He is on the scientific advisory board for Cereave Technologies and consulted for Prothena, Eisai, Roche, and Biogen Australia. Victor Villemagne is and has been a consultant or paid speaker at sponsored conference sessions for Eli Lilly, Life Molecular Imaging, GE Healthcare, Abbvie, Lundbeck, Shanghai Green Valley Pharmaceutical Co Ltd, and Hoffmann La Roche. Ashley Bush is a shareholder in Alteity Ltd, Cogstate Ltd, and Mesoblast Ltd. He is a paid consultant for, and has a profit share interest in, Collaborative Medicinal Development Pty Ltd. He has received lecture fees from Biogen and Merck Sharp & Dohme P/L. Ziad Saad, Galen Triana-Baltzer, Randy Slemon, and Hartmut Kolb are employees of Janssen R&D. The other authors did not report any conflict of interest.

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SUPPORTING INFORMATION
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