Immune-Related IncRNAs to Construct Novel Signatures and Predict the Prognosis of Rectal Cancer

Xiao-Liang Xing¹,2, Chaoqun Xing¹,2, Zhi Huang¹,2, Zhi-Yong Yao¹,2* and Yuan-Wu Liu³*

¹ School of Public Health and Laboratory Medicine, Hunan University of Medicine, Huaihua, China, ² Department of Pediatrics, Xiangya Hospital, Central South University, Changsha, China, ³ Beijing Advanced Innovation Center for Food Nutrition and Human Health, China Agricultural University, Beijing, China

Colorectal cancer (CRC) is one of the most common cancers. Almost 1/3 of CRC are rectal cancer, and 95% of rectal cancers are rectal adenocarcinoma (READ). Increasing evidences indicated that long noncoding RNAs (lncRNAs) have important role in the genesis and development of cancers. The purpose of our present study was to identify the differential expression IncRNAs which potentially related with immune cells infiltration and establish a risk assessment model to predict the clinical outcome for READ patients. We obtained three immune-related differential expression IncRNAs (IR-DELs) (C17orf77, GATA2-AS1, and TPT1-AS1) by differential expression analysis following correlation analysis and Cox regression analysis. A risk assessment model were constructed by integrating these analysis results. We then plotted the 1-, 3-, and 5-year ROC curves depending on our risk assessment model, which suggested that all AUC values were over 0.7. In addition, we found that the risk assessment model was correlated with several immune cells and factors. This study suggested that those three signatures (C17orf77, GATA2-AS1, and TPT1-AS1) screened by pairing IR-DELs could be prognosis markers for READ patients and might benefit them from antitumor immunotherapy.

Keywords: READ, immune, LncRNAs, overall survival, risk model

INTRODUCTION

Colorectal cancer (CRC) is one of the most prevalent cancer which accounts for approximately 10% of cancer cases and deaths annually. There are almost 1.8 million new cases of CRC and 881,000 deaths occurred in the world globally (1). Rectal cancer as a subtype of CRC which accounts for one-third of newly diagnosed CRC cases every year (2); 95% of rectal cancer is the rectal adenocarcinoma (READ). The treatment guidelines state that preoperative neoadjuvant chemotherapy could improve the radical resection rate and reduce local recurrence. Thus, even though surgery is the primary treatment for READ, preoperative neoadjuvant chemoradiotherapy is still the key to the treatment of READ (3, 4). However, previous studies demonstrated that almost 30% of patients treated with a curative treatment will develop distant metastases finally (5–8). It is very important to find suitable biomarkers for the prognosis of READ patients.
Recently, increasing evidences indicated that immune cells and immune factors participated in antitumor effects, including tumor initiation and progression (9, 10). The immunotherapy has promising advantages for cancers in the treatment efficiency and long-term survival of patients (11). CRC is a type of tumor which is accompanied by high density of tumor-infiltrating lymphocytes. The CRC patients, for example, with higher CD8+ T lymphocytes always displayed a better prognosis (12–14). Actually, previous studies demonstrated that signatures focusing on the tumor immune infiltrations have shown a promising predictive and prognostic value in diagnosis, evaluation, and treatment of cancer (15).

Long noncoding RNAs (lncRNAs) are kind of RNA which not coding for any protein or peptides and account for approximately 80% of the human transcriptome. Most of lncRNAs are more than 200 nucleotides and regulate 70% of the gene expression. In recent years, increasing studies have shown that lncRNAs are deeply involved in both normal cell development and cancer genesis (16, 17). In the present study, we aimed to investigate the relationships of lncRNAs with gene expression profile and the tumor-immune microenvironment in READ, and construct a risk assessment model for prognosis prediction in READ patients and benefit them from antitumor immunotherapy.

MATERIAL AND METHOD

Data Source and Differentially Expression Analysis

The data used in this study was obtained from openly database TCGA (The Cancer Genome Atlas), which included 10 normal and 166 READ samples. The mRNA expression profile data were analyzed by DESeq2 package in R (3.6.1) software as cutoff with padj <0.05, |logFC| ≥0.5, and basemean >50. Gene transfer format (GTF) files were downloaded from Ensembl (http://asia.ensembl.org) for gene annotation. A list of recognized immune-related genes (IR-Genes) was downloaded from the ImmPort database (http://www. immport.org) and was used to screen IR-lncRNAs by a coexpression strategy. The extent of immune cell infiltration was obtained from Tumor Immune Estimation Resource (TIMER) (https://cistrome.shinyapps.io/timer/). Spearman’s correlation analysis was performed between IR-DEGs and their corresponding lncRNAs as cutoff with r ≥0.3 and p-value <0.05 to screen the immune-related differentially expressed lncRNAs (IR-DEGs).

Survival Analysis

According to the median expression value or median risk score, patients were divided into high-expression/risk group and low-expression/risk group. Survival, Survminer, and RegParallel package in R were used to carry out univariate and multivariate Cox regression analysis.

Construction of Specific Risk Assessment Model

The risk model was established based on the expression data and multiplied Cox regression coefficients. The formula was set as followed, risk score = expression level of C17orf77 * (−1.5905) + expression levels of GATA2-AS1 * (−1.3003) + expression level of TPT1-AS1 * (−1.4072) (18, 19). To further investigate the relevance of the risk model with the clinical pathological features of READ, we analyzed the relationship between the risk value and vital status, gender, pathologic NMT, and pathologic stage. We then constructed time-dependent receiver operating characteristic (ROC) curves within 1-year, 3-year, 5-year, and clinical features and estimated its utility as a prognostic model for predicting the survival status.

Statistical Analysis

A repeated-measure ANOVA followed by Bonferroni post hoc tests or unpaired two-tail Student’s t-test was used as indicated. All statistical analyses were performed using the Graphpad Prism 6.0.1.

RESULTS

Identification of Immune-Related Differentially Expressed lncRNAs

We downloaded the expression data of 176 samples (10 control vs. 166 READ) from TCGA database. Through differential expression analysis by DEseq2, we screened 7,283 differentially expression genes (DEGs) (4,120 DEGs were upregulated and 3,163 DEGs were downregulated) (Figure 1A). Of which, we obtained 514 immune-related DEGs (IR-DEGs) by cross-analysis with the recognized immune-related genes (Figure 1B) and 420 differential expression lncRNAs (DELs) by cross-analysis with GTF annotation data (Figure 1C). We then introduced the Spearman’s correlation analysis for those 514 IR-DEGs and 420 DELs and identified that there were 9,973 IR-DEGs-IR-DELs pairs involved 508 IR-DEGs and 409 IR-DELs.

The purpose of this study was to identify the IR-DELs which could use to construct a risk assessment model to predict the outcome of READ. Therefore, we performed univariate Cox regression analysis for those 409 IR-DELS and found 22 IR-DEL expression were associated with the overall survival (OS) of READ (Figure 1D). The expressions of those DELs are shown in Figure 1E. Subsequently, we also performed multivariate Cox regression analysis for those 22 IR DELs and found three IR-DELs (C17orf77, GATA2-AS1, and TPT1-AS1) were still associated with the OS of READ (Figure 1F).

Establishment of Specific Risk Assessment Model

The expressions of those three DELs (C17orf77, GATA2-AS1, and TPT1-AS1) were increased significantly in the cancer group (Figure 1E). We used those three DELs to construct a specific risk assessment model. The risk scores and survival outcome of each READ cases are displayed in Figures 2A, B. The expression of C17orf77 and GATA2-AS1 were decreased significantly while the expression of TPT1-AS1 was comparable between the low-risk and high-risk groups (Figure 2C).
After regrouping the READ patients by risk value, we analyzed the relationship of the risk model with the OS of READ and found that low-risk READ patients exhibited a better OS (Figure 2D). Meanwhile, we also calculated the areas under curve (AUCs) for each receiver operating characteristic (ROC) curve of those three IR-DELs and drew the curved line (Figure 2E). To validate the optimality, we also plotted the 1-, 3-, and 5-year ROC curves, which suggested that all AUC values were over 0.7 (Figures 2F–H).

Clinical Evaluation by Risk Assessment Model
The TNM staging system was widely used for cancer stage classification and determining the appropriate treatment strategy. Therefore, we analyzed the relationship of the risk score with the clinical characteristics. As shown in Figure 3, we found that the risk score in the death READ patients was significantly higher than that in the alive READ patients (Figure 3A). There was a comparable risk value between male and female (Figure 3B). The risk score showed several differences among pathologic N, pathologic M, and pathologic stage system except pathologic T system (Figures 3C–I). We also calculated the AUC for each ROC curve of those clinical characters drew the curved line; the result is shown in Figure 3J. The AUC value of the risk model in predicting the survival status was the highest (0.7126).

Estimation of Tumor-Infiltration With the Risk Assessment Model
Firstly, we analyzed the differences of immune infiltration between normal and READ patients and found that there are 50 immune cells and factors that were different between control and cancer. Because IncRNAs and immune-related genes were initially connected, we consequently investigated the relationship of the risk model with the tumor immune microenvironment (Supplementary Table 1). We then investigated whether those 64 immune cells and factors were different between low-risk group and high-risk group. After this analysis, we found five immune cells and immune factors were different (Figures 4A–E). A detailed Spearman’s correlation analysis was conducted; the result is shown in Figure 4F. READ patients in the high-risk group were more positively associated with Macrophage_TIMER.

DISCUSSIONS
CRC is one of the most common cancers which comprised colon cancer and rectal cancer. While colon cancer and rectal cancer are very similar in several ways, their treatments are quite different. Due to the improvement of the treatment over the last few decades, the OS rate of READ has greatly improved. However, the additional long-term survival for READ patients is hard to achieve even under extensive treatment compared with
other cancers. Previous studies indicated that almost 30% of patients treated with a curative treatment will eventually develop distant metastases (5–8). Therefore, the early diagnosis and prediction of prognosis in patients with READ are important for their long-term survival.

Increasing evidence indicated that lncRNAs deeply participated in the tumor genesis, development, and metastasis (20–22). In another aspect, lncRNAs were screened to be the prognosis biomarkers for cancers, including READ. Chao et al. (2019) found lncRNA D16366 could be a potential biomarker for diagnosis and prognosis of hepatocellular carcinoma (23). Zhao et al. (23) found five lncRNAs (AC079789.1, AC106900.2, AL121987.1, AP004609.1, and LINC02163) were independently associated with the prognosis of patients with rectal cancer through using univariate and multivariate Cox regression analyses (24). Those five lncRNAs could be the potential biomarkers for READ (24). Qi et al. (25) found that lncRNA KCNQ1OT1 and SNHG1 were unveiled as common diagnostic biomarkers for the initiation and metastasis of colon and rectal cancers (25).

Increasing evidences indicated that immune cells and factors play an important role in antitumor immunity and antitumor initiation and progression (9, 10). Immune checkpoint blockade therapy has become an important cancer treatment option and immunotherapy has promising advantages in terms of efficiency and long-term survival (11). Antibody drugs, such as anti-PD-1 and anti-PD-L1, demonstrate obvious advantages such as broad applicability across cancer types and durable clinical response when treatment is effective (26). In the present study, we aimed to investigate the relationships of IR-DELs with immune cells and immune factors of READ and construct a risk assessment model to predict the prognosis for patients with READ and distinguishing those who could benefit from antitumor immunotherapy. First, we found that 23 IR-DELs were associated with the OS of READ by differential expression analysis, correlation analysis, and univariate Cox regression analysis. Three IR-DELs (C17orf77, GATA2-AS1, and TPT1-AS1) were independently associated with the OS of READ. Previous studies indicated that OS-related genes could be used to construct the risk assessment model. According to previous studies, we also constructed a risk assessment model by using those three IR-DELs (C17orf77, GATA2-AS1, and TPT1-AS1) (18, 19).

The correlation analysis results showed that risk value was correlated with pathologic N, pathologic M, pathologic stage, several immune cells, and immune factors. The AUC value was over 0.7 which proved that the model had certain accuracy in predicting the prognosis. However, whether this model could be used for clinical prognosis, diagnosis still needs further validation studies.

Chromosome 17 Open Reading Frame 77 (C17orf77) is a gene in cancer that has not been well studied yet. In 2019, Shaikh et al. found that C17orf77 may hold a putative role in both pathogenesis of smoking and nonsmoking-related head and neck squamous cell carcinoma tumors and could be considered...
a potential biomarker for separating these tumors (27). GATA-binding protein 2 (GATA2) is a member of the GATA family of zinc-finger transcription factors. GATA2 exists as an acetylated protein in immature precursor cells. GATA2-AS1 is the antisense RNA of GATA2. In 2019, Zhang et al. found that GATA2-AS1 could repress nonsmall-cell lung cancer cell proliferation via regulating GATA2. These results indicated that GATA2-AS1 could be a potential target for lung cancer drugs (28). In 2020, Li et al. found that GATA2-AS1 was a tumor-associated lncRNAs in COAD by bioinformatics analysis (29). TPT1-AS1 is the antisense RNA of tumor protein, translationally controlled 1 (TPT1). Previous studies indicated that TPT1-AS1 could regulate the genesis and development of several cancers by different ways. Huang et al. and Gao et al. found that TPT1-AS1 could promote cell growth in cervical cancer and glioblastoma via acting as a sponge for miR-324-5p and miR-23a-5p, respectively (30, 31). Tumors grow and evolve through constant crosstalk with the surrounding microenvironment, and emerging evidences indicate that angiogenesis and immunosuppression frequently occur simultaneously in response to this crosstalk. Although VEGFR2 was downregulated in the prostate cancer, VEGFR2 was upregulated in the high-risk prostate cancer and predicted clinical progression. Krebs et al. found miR-221-3p upregulation as an escape mechanism from VEGFR2 inhibition in prostate cells (32). Zhang et al. (33) and Zhang et al. (34) found that TPT1-AS1 could promote angiogenesis, progression, and metastasis of CRC through TPT1-AS1/NF90/VEGFA signaling pathway and upregulating the TPT1-mediated FAK and JAK-STAT3 signaling pathways, respectively (33, 34). Our present study indicated that C17orf77, GATA2-AS1, and TPT1-AS1 could be the prognosis biomarkers for READ which reinforced the role of those lncRNAs in predicting the outcome of cancers as biomarkers.

In conclusion, we identified three IR-DELS correlated with OS and constructed a specific risk assessment model for prognostic prediction of READ. Those three IR-DEL classifiers also demonstrated considerable predictive accuracy for predicting the OS. In addition, the risk model was correlated with immunotherapy-related biomarkers, suggesting its application...
value for predicting the efficiency of treatment. However, we also recognized some shortcomings and limitations in this study, especially for the lack of clinical validation. Therefore, we will collect clinical samples and carry out the verification in the future work.

DATA AVAILABILITY STATEMENT

Publicly available datasets were analyzed in this study. This data can be found here: https://portal.gdc.cancer.gov/.

AUTHOR CONTRIBUTIONS

Z-YY and Y-WL conceived and designed the experiments. X-LX performed the analysis. CX and ZH helped to analyze the data. Y-WL and X-LX wrote the paper. All authors contributed to the article and approved the submitted version.

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SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fonc.2021.661846/full#supplementary-material

REFERENCES

1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global Cancer Statistics 2020: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. CA Cancer J Clin (2021) 71:209–49. doi: 10.3322/caac.21660
2. Mondaca S, Yaeger R. Genetics of Rectal Cancer and Novel Therapies: Primer for Radiologists. Abdom Radiol (NY) (2019) 44:3743–50. doi: 10.1007/s00261-019-02051-x
3. Braendengen M, Tveit KM, Berglund A, Birkemeyer E, Frykholm G, Pahlman L, et al. Randomized Phase III Study Comparing Preoperative Radiotherapy With Chemoradiotherapy in Nonresectable Rectal Cancer. J Clin Oncol (2008) 26:3687–94. doi: 10.1200/JCO.2007.15.3858
4. Colorectal Cancer Collaborative, G. Adjuvant Radiotherapy for Rectal Cancer: A Systematic Overview of 8,507 Patients From 22 Randomised Trials. Lancet (2001) 358:1291–304. doi: 10.1016/S0140-6736(01)06409-1
5. JI D, Yi H, Zhang D, Zhan T, Li Z, Li M, et al. Somatic Mutations and Immune Alternation in Rectal Cancer Following Neoadjuvant Chemoradiotherapy. Cancer Immunol Res (2018) 6:1401–16. doi: 10.1158/2326-6066.CIR-17-0630
6. Sauer R, Liersch T, Merkel S, Fietkau R, Hohenberger W, Hess C, et al. Preoperative Versus Postoperative Chemoradiotherapy for Locally Advanced Rectal Cancer: Results of the German CAO/ARO/AIO-94 Randomized Phase...
17. Zhao J, Lawless MW. Long Noncoding RNAs and Their Role in the Liver

16. Schmitt AM, Chang HY. Long Noncoding RNAs in Cancer Pathways.

10. Chen YP, Zhang Y, Lv JW, Li YQ, Wang YQ, He QM, et al. Genomic Analysis

12. Bindea G, Mlecnik B, Tosolini M, Kirilovsky A, Waldner M, Obenauf AC,

11. Yang L, Wang L, Zhang Y. Immunotherapy for Lung Cancer: Advances and

14. Galon J, Angell HK, Bedognetti D, Marincola FM. The Continuum of Cancer

7. Engelen SM, Maas M, Lahaye MJ, Leijten JW, van Berlo CL, Jansen RL, et al.

Modern Multidisciplinary Treatment of Rectal Cancer Based on Staging With
Magnetic Resonance Imaging Leads to Excellent Local Control, But Distant
Control Remains a Challenge. *Eur J Cancer* (2013) 49:2311–20. doi: 10.1016/
j.ejca.2013.03.006

8. van Gijn W, Marijnien CA, Nagtegaal EM, Putter H, Wiggers T, et al. Preoperative Radiotherapy Combined With Total Mesorectal Excision for Resectable Rectal Cancer: 12-Year Follow-Up of the Multicentre, Randomised Controlled TME Trial. *Lancet Oncol* (2011) 12:575–82. doi: 10.1016/S1470-2245(11)70097-3

9. Berraondo F, Minute L, Ajona D, Corrales M, Melero I, Pio R. Innate Immune
Mediators in Cancer: Between Defense and Resistance. *Immunol Rev* (2016)
274:290–306. doi: 10.1111/imr.12464

13. Mlecnik B, Tosolini M, Kirilovsky A, Berger A, Bindea G, Meatchi T, et al.
Histopathologic-Based Prognostic Factors of Colorectal Cancers Are Associated
With the State of the Local Immune Reaction. *J Clin Oncol* (2011) 29:610–8. doi: 10.1200/JCO.2010.30.5425

11. Yang L, Wang L, Zhang Y. Immunotherapy for Lung Cancer: Advances and
Prospects. *Am J Clin Exp Immunol (2016)* 5:1–20.

12. Bindea G, Mlecnik B, Tosolini M, Kirilovsky A, Waldner M, Obenauf AC,
et al. Spatiotemporal Dynamics of Intratumoral Immune Cells Reveal the
Immune Landscape in Human Cancer. *Immunity* (2013) 39:78–92. doi: 10.1016/j.immuni.2013.10.003

13. Mlecnik B, Tosolini M, Kirilovsky A, Berger A, Bindea G, Meatchi T, et al.

14. Galon J, Angell HK, Bedognetti D, Marincola FM. The Continuum of Cancer
Immunosurveillance: Prognostic, Predictive, and Mechanistic Signatures.
*Immunity* (2013) 39:11–26. doi: 10.1016/j.immuni.2013.07.008

15. Shaikh I, Ansari A, Ayachit G, Gandhi M, Sharma P, Bhairappanavar S, et al.
Differential Gene Expression Analysis of HNSCC Tumors Depicted by Tobacco
Dependent and Independent Molecular Signatures. *Oncotarget* (2019) 10:6168–83. doi: 10.18632/oncotarget.27249

16. Schmitt AM, Chang HY. Long Noncoding RNAs in Cancer Pathways.

17. Zhao J, Lawless MW. Long Noncoding RNAs and Their Role in the Liver

18. Huang M, Zhang T, Yao ZY, Xing C, Wu Q, Liu YW, et al. MicroRNA Related
Prognosis Biomarkers From High Throughput Sequencing Data of Kidney
Renal Clear Cell Carcinoma. *BMC Med Genomics* (2021) 14:7: doi: 10.1186/s12920-021-00932-z

19. Xing XL, Yao ZY, Xing C, Huang Z, Peng J, Liu YW. Gene Expression and
DNA Methylation Analyses Suggest That Two Related Immune Genes Are
Prognostic Factors of Colorectal Cancer. *BMC Med Genomics* (2021) 14:116:
doi: 10.1186/s12920-021-00966-3

20. Choudhari R, Sedano MJ, Harrison AL, Subramani R, Lin KY, Ramos EL, et al.
Long Noncoding RNAs in Cancer: From Discovery to Therapeutic Targets.
*Adv Clin Chem* (2020) 95:105–47. doi: 10.1016/bs.acc.2019.08.003

21. Luo X, Qu Y, Jiang Y, Chen F, Jiang L, Zhou Y, et al. Long non-Coding RNA
Implicated in the Invasion and Metastasis of Head and Neck Cancer: Possible
Function and Mechanisms. *Mol Cancer* (2018) 17:14. doi: 10.1186/s12943-018-0763-7

22. Chi Y, Wang D, Wang J, Yu W, Yang J. Long Non-Coding RNA in the
Pathogenesis of Pancreatic Cancer. *Cells* (2019) 8:1–44. doi: 10.3390/cells8091015

23. Chao Y, Zhou D. IncRNA-D16366 Is a Potential Biomarker for Diagnosis and
Prognosis of Hepatocellular Carcinoma. *Med Sci Monit* (2019) 25:6581–6.
doi: 10.12659/MSM.915100

24. Zhao K, Wang M, Kang H, Wu A. A Prognostic Five Long-Noncoding RNA
Signature for Patients With Rectal Cancer. *J Cell Biochem* (2019) 1–7. doi:
10.1002/jcb.29549

25. Qi X, Lin Y, Liu X, Chen J, Shen B. Biomarker Discovery for the Carcinogenic
Heterogeneity Between Colon and Rectal Cancers Based on IncRNA-
Associated ceRNA Network Analysis. *Front Oncol* (2020) 10:535985. doi:
10.3389/fonc.2020.535985

26. He X, Xu C. Immune Checkpoint Signaling and Cancer Immunotherapy. *Cell
Res* (2020) 30:660–9. doi: 10.1038/s41412-020-0343-4

27. Shakk A, Anvari A, Ayacit G, Gandhi M, Sharma P, Bhairappanvar S, et al.
Differential Gene Expression Analysis of HNSCC Tumors Depicted by Tobacco
Dependent and Independent Molecular Signatures. *Oncotarget* (2019) 10:6168–83. doi: 10.18632/oncotarget.27249

28. Zhang L, Gao L, Shao M, Sun GY. A MYC Target Long Non-Coding RNA
GATA2-AS1 Regulates Non-Small Cell Lung Cancer Growth. *Neoplasma*
(2019) 66:954–62. doi: 10.4149/ne_2019_190210N112

29. Li Z, Tan H, Yu H, Deng Z, Zhou X, Wang M. DNA Methylation and Gene
Expression Profiles Characterize Epigenetic Regulation of IncRNAs in Colon
Adenocarcinoma. *J Cell Biochem* (2016) 121:2406–15. doi: 10.1002/jcb.29463

30. Jiang H, Huang G, Zhao N, Zhang T, Jiang M, He Y, et al. Long non-Coding
RNA TPT1-AS1 Promotes Cell Growth and Metastasis in Cervical Cancer via
Acting As a Sponge for miR-324-3p. *J Exp Clin Cancer Res* (2018) 37:169:
doi: 10.1186/s13046-018-0846-8

31. Zhang Y, Sun J, Qi Y, Wang Y, Ding Y, Wang K, et al. Long Non-Coding RNA
TPT1-AS1 Promotes Cell Growth and Metastasis in Cervical Cancer by
Upregulating the TPT1-Mediated FAK and JAK-STAT3 Signalling Pathway. *Adv Clin Chem* (2020) 12:6191–205. doi: 10.18632/aging.103016

32. Zhang L, Ye F, Zuo Z, Cao D, Peng Y, Li Z, et al. Long Noncoding RNA Tpt1-
As1 Promotes the Progression and Metastasis of Colorectal Cancer by
Upregulating the Tpt1-Mediated Fak and Jak-Sta3 Signalling Pathways. *Aging (Albany NY)* (2021) 13:3779–97. doi: 10.18632/aging.202339

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