Role of endothelial nitric oxide synthase gene polymorphisms in predicting aneurysmal subarachnoid hemorrhage in South Indian patients

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Abstract. Endothelial nitric oxide synthase (eNOS) gene polymorphisms have been implicated as predisposing genetic factors that can predict aneurysmal subarachnoid hemorrhage (aSAH), but with controversial results from different populations. Using a case-control study design, we tested the hypothesis whether variants in eNOS gene can increase risk of aSAH among South Indian patients, either independently, or by interacting with other risk factors of the disease. We enrolled 122 patients, along with 224 ethnically matched controls. We screened the intron-4 27-bp VNTR, the promoter T-786C and the exon-7 G894T SNPs in the eNOS gene. We found marked interethnic differences in the genotype distribution of eNOS variants when comparing the South Indian population with the reported frequencies from Caucasian and Japanese populations. Genotype distributions in control and patient populations were found to be in Hardy-Weinberg equilibrium. In patients, the allele, genotype and estimated haplotype frequencies did not differ significantly from the controls. Multiple logistic regression indicated hypertension and smoking as risk factors for the disease, however the risk alleles did not have any interaction with these risk factors. Although the eNOS polymorphisms were not found to be a likely risk factor for aSAH, the role of factors such as ethnicity, gender, smoking and hypertension should be evaluated cautiously to understand the genotype to phenotype conversion.

Keywords: eNOS, SNP, aneurysm, subarachnoid hemorrhage, India

1. Introduction

Intracranial aneurysm is a fairly common condition that is often asymptomatic until the time of rupture, resulting in aneurysmal subarachnoid hemorrhage (aSAH), ensuing significant morbidity and mortality. It is the cause for an estimated five to 15 percent of all strokes, with an overall incidence of 16 to 22 per 100,000 population [2]. The reported prevalence rate of intracranial aneurysms in India varies from 0.75% to 10.3% as determined in angiographic and autopsy studies [9,18]. Current evidence supports the concept that the pathogenesis of intracranial aneurysms has a multifactorial origin, where the essential defect of the arterial wall may, at least in part be genetically determined [19].

Endothelial nitric oxide synthase (eNOS) expressed on endothelial cells plays a major role in shear-stress response related endothelial dysfunction, which has been recognized as the underlying pathological process of
intracranial aneurysms [22]. Endothelium derived nitric oxide (NO) synthesized by nitric oxide synthase has a powerful vasodilatory action and studies on the response of NO levels to shear-stress investigated in human cerebral endothelial cells has indicated a linear relationship [15]. In addition, it has been demonstrated that as blood vessels age, shear-stress induced eNOS becomes impaired leading to decreased NO bioavailability and endothelial dysfunction, which is likely to contribute to age-associated vascular stiffness [21]. A rat model of SAH used to demonstrate that cerebral microvessels have endothelial dysfunction, has revealed predisposition to vasoconstriction and spasm, suggesting impaired translation of eNOS [17]. Interestingly, aSAH is a late onset disorder and therefore reduced NO bioavailability due to eNOS expression variants could account for the etiology of aSAH.

Located on chromosome 7q35-36, eNOS has three well-defined polymorphisms; the T-786C SNP (rs2070744) in the promoter region, the 27-bp VNTR in intron 4 and the G894T SNP (rs1799983) in exon 7. These gene mutations have been shown to increase susceptibility either singly, or in combination with conditions that includes hypertension, stroke, cerebral small-vessel disease, and abdominal aortic aneurysms [1,6,8,16]. It has been demonstrated that the eNOS gene is functionally polymorphic, where some of the allelic variants may constitutively express altered phenotypic expression. These allelic variants of the gene, can result in decreased NO production, which may be explained either in terms of functional or quantitative deficiency (abnormalities of transcription, mRNA stability, formation of catalytically defective protein) or by enhanced degradation of the enzyme [4]. The eNOS gene variants with altered eNOS regulation might render the blood vessels susceptible to demographic factors.

Recently, a study done in a North American population had analyzed the role of these three eNOS gene polymorphisms as potential risk factors that can predict susceptibility to aSAH and cerebral vasospasm [10]. They had reported a positive association with regard to the 4a allele of the eNOS 27-bp VNTR polymorphism ($P = 0.007$), where they found an odds of approximately 3.95 for being an aSAH case for those persons with at least one 4a allele. In the present study, we have explored the hypothesis of whether polymorphisms in the eNOS gene are associated with aSAH in South Indian population, in relation to its other phenotypic variables and ethnicity.

2. Materials and methods

2.1. Study population

We studied 122 unrelated patients who had been consecutively admitted to the Department of Neurosurgery at Sree Chitra Tirunal Institute for Medical Science and Technology (SCTIMST), Kerala. The patient group consisted of 70 males and 52 females. Patients with aSAH were defined by symptoms suggestive of SAH combined with subarachnoid blood on computed tomography and a proven aneurysm. All patients had angiographically documented saccular aneurysms with SAH confirmed by cranial computed tomography (CT), magnetic resonance angiography (MRA) and digital subtraction angiography (DSA). The neurological grade was classified by the World Federation of Neurological Surgeons (WFNS) scale and all grades of SAH were eligible for inclusion. All preexisting medical conditions were documented. Exclusion criteria included nonsaccular aneurysm, arteriovenous malformations and haematological disorders. Demographic variables of aSAH patients in the South Indian population are shown in Table 1. The control group consisted of 224 unrelated, age and ethnically-matched volunteers recruited from individuals admitted to the hospital for any reason other than neurological diseases. This group comprised of 110 males and 114 females. All subjects were of the same ethnic origin and all participants gave informed consent for the study. The outline of the present study was approved by the Institutional Ethical Committee for Biomedical Subjects.

2.2. Genotyping of the eNOS polymorphisms

Peripheral venous blood from each participant was drawn and used for the isolation of genomic DNA using the standard organic extraction protocol. Presence of the T-786C and G894T SNPs were determined by polymerase chain reaction followed by restriction enzyme (RE) digestion. The 27-bp VNTR was genotyped as described previously [25]. The PCR primers and annealing temperatures are summarized in Table 2. PCR assays was carried out in a 20 µl volume with 100ng of genomic DNA, 10 pM of each primer, 2.5 mM dNTP (Amersham), 1.5 mM MgCl$_2$ and 10x PCR buffer [50 mM KCl, 500 mM Tris buffer, 160 mM (NH$_4$)$_2$SO$_4$, pH 8.8, and 0.1% Tween 20], 0.1% Triton X-100 and 0.5 U Taq polymerase (Sigma). After a hot start at 95°C for 4 minutes, amplification was achieved by 35 cycles of denaturation at 94°C for 30 seconds,
Table 1
Demographic and clinical characteristics of study subjects

| Characteristic(s)             | Patient Group | Control Group |
|------------------------------|---------------|---------------|
| Total no. of subjects        | 122           | 224           |
| Mean age (range), years      | 51.49 ± 11.4 years | 46.71 ± 14.1 years |
| Male, n                      | 70            | 110           |
| Mean age (range), years      | 48.9 (18–71) years | 46.7 (32–87) years |
| Female, n                    | 52            | 114           |
| Mean age (range), years      | 54.98 (20–75) years | 46.5 (31–81) years |
| Hypertension                 | 42            | 18            |
| Cigarette smoking            | 56            | 23            |
| Alcohol consumption          | 23            | 21            |
| Diabetes mellitus            | 7             | 29            |
| Admission WFNS grade, n      | n/a           | n/a           |
| I                            | 90            | n/a           |
| II                           | 17            | n/a           |
| III                          | 7             | n/a           |
| IV                           | 7             | n/a           |
| V                            | 1             | n/a           |
| Location, n                  | n/a           | n/a           |
| Internal carotid artery      | 28            | n/a           |
| Middle cerebral artery       | 26            | n/a           |
| Anterior communicating artery| 56            | n/a           |
| Anterior cerebral artery     | 11            | n/a           |
| Posterior communicating artery| 12        | n/a           |
| Basilar artery               | 5             | n/a           |
| Multiple, n                  | 17            | n/a           |
| Two aneurysms                | 14            | n/a           |
| Three aneurysms              | 3             | n/a           |

Table 2
Primer sequences

| Polymorphism | Primer pairs Sense/Antisense | Ta, (°C) | RE Digestion products, bp |
|--------------|------------------------------|---------|--------------------------|
| G894T        | 5'-AAGGCAGGAGACAGTGGATGGA-3' | 60      | BanII 248, 163, 85 |
|              | 5'-CCCAGTGCAATCCCCCTTGTGCTCA-3' |         |                         |
| T-786C       | 5'-GCATGCACTCTGGCCTGAAGT-3' | 60      | MspI 162, 116, 61, 46 |
|              | 5'-CAGGAAGCTGCCCTTCCAGTGC-3' |         |                         |
| 27-bp VNTR   | 5'-AGGCCCTATGGTGAGGTTTT-3' | 56      | ... 420, 393 |
|              | 5'-TCCTTTATCGTGTTGTTGTCAC-3' |         |                         |

annealing at specific temperatures, see Table 2, for 30 seconds, and extension at 72°C for 30 seconds, by using a thermal cycler (Eppendorff). PCR products were restriction digested with MspI and BanII restriction endonucleases (New England Biolabs), respectively. Restriction digested products were resolved using 3% NuSieve agarose gel and visualized under UV light after ethidium bromide staining. All three genotypes of the T-786C and G894T SNPs were further verified by direct sequencing using ABI PRISM BigDye Terminators v3.1 Cycle Sequencing Kit (Applied Biosystems), and analyzed using ABI PRISM 3730 Genetic Analyzer (Applied Biosystems).

2.3. Statistical analysis and haplotype construction

Deviations from the Hardy-Weinberg equilibrium were tested for all polymorphisms in cases and controls by comparing observed and expected genotype frequencies with an exact goodness of fit test. Odds ratios, as the estimates of relative risk of disease, with 95% confidence intervals were calculated to compare allele and genotype frequencies using chi-square analysis. Multiple logistic regression analysis was performed with the diagnosis as the dependent variable, and co-variates such as age at onset, age above 50 years, female gender, smoking, hypertension, diabetes mellitus, alcohol use, and the eNOS genotypes as the independent variables. For analyzing dominant inheritance model (e.g., eNOS 786CC + TC vs TT), genotypes were coded as 1 (heterozygous or homozygous for the minor allele) or 0 (the major allele homozygote). Regression analysis was conducted using the program SPSS 13.0 for Windows (SPSS Inc, Chicago, IL, USA) statistical analysis software. Additionally, the interaction of genotypes with the various non-genetic risk factors was
studied using the Crosstab option of the statistical analysis software program SPSS 13.0 (SPSS Inc, Chicago, IL). Haplotype and pairwise linkage disequilibrium analyses (LD) were performed by COCAPHASE program of the UNPHASED software suite, in which the expectation-maximization (EM) algorithm is used to estimate frequencies [5]. The extent of LD was expressed in terms of the maximum likelihood estimate of disequilibrium, \( D' \) and viewed in HAPLOVIEW. The power of the present study was calculated using the allele frequencies reported earlier for the 27-bp VNTR polymorphism, at a 5% test level (alpha = 0.05, two tailed) [10]. The statistical power of the study was calculated as described by Cohen [3]. Probability (\( P \)) values of less than 0.05 were considered statistically significant.

### 3. Results

The observed allele and genotype frequencies for T-786C and G894T SNPs, and intron 27-bp VNTR polymorphism is presented in Table 3. The observed genotype frequencies did not deviate from the Hardy-Weinberg equilibrium in both patients and control subjects. No significant differences in the distribution of allele and genotype frequencies in patient and control groups were observed (Table 3). We tested the LD between all the pairwise polymorphisms and observed that the T-786C and 27-bp VNTR loci in both patient and control groups showed linkage, \( D' = 0.6033 \) in controls and \( D' = 0.6247 \) in patients). However, we found a negative \( D' \) value (−1.00 in controls and −0.4117 in patients) for the association between the variants in the G894T and in 27-bp VNTR loci, indicating that the rarer variant 4a in intron 4 is associated with the wild-type variant Glu298 in exon 7 or vice versa. Comparison of haplotypes frequencies revealed no single haplotype overrepresented in aSAH patients (Table 4). T-4a-G was observed to be the most frequent haplotype and was observed in more than 60% of both the case and control populations.

The previously reported risk genotype, 4a/4b of 27-bp VNTR polymorphism was not significantly associated with an increased risk of developing aSAH. Likewise, the T-786C SNP, which was in strong linkage disequilibrium with the 27-bp VNTR, was again not significantly associated with aSAH. After controlling for age and sex multiple logistic regression analysis showed a lack of association of genotypes of the eNOS polymorphisms with aSAH. Non-genetic risk factors such as hypertension and smoking were identified as independent predictors of aSAH (\( P = 0.0001 \) for the entire model), as shown in Table 5. However, no significant interactions were observed between the risk alleles and the non-genetic risk factors.

The power calculation for the 27-bp VNTR 4a/4b allele was done as follows. The power required in this study to detect the specified difference in the frequen-
Caucasian patients, and severity of post-SAH cerebral vasospasm in
shown to significantly differentiate between the pres-

had been reported that heterozygosity
aSAH, and therefore, suggested its use as a diagnostic

Three times more prevalent in cases (49%) than controls

In the present study we were unable to observe any
significant variation in the eNOS polymorphisms at the
allele, genotype or haplotype levels, in South Indian
patients and controls. Although a strong rationale for

gene contributing to reduced NO bioavailability in cerebral blood vessels had
been proposed for the increased risk for aSAH, our re-
sults could not substantiate this. Initial studies done in
a North American population of Caucasian origin had

The impact of ethnicity on the distribution of these
clinically relevant eNOS gene polymorphisms had been
effectively studied in Caucasian, African-American and
Asian populations [23]. Likewise, we also observed sig-
ificant allelic and genotypic variability when compared to
different study populations (Table 6). The 4a risk allele
frequency of eNOS 27-bp VNTR in our study was comparable to the European population while
it was least frequent allele in Japanese population.

Though the risk genotype 4a/4b was significantly high-
er in South Indian patients when compared to North
American patients, it did not manifest its debilitating
phenotype. The eNOS -786C risk allele had a lower
frequency in the South Indian population (21.4%) in
comparison to the Caucasians of North America and
Europe, while in the heterozygous genotypic combina-
tion the -786C allele was comparable to these popula-
tions. The differences in allele frequencies among the
study populations were also reflected subsequently in
the haplotype frequencies. Khurana et al. reported no
significant LD among the three polymorphisms [11],
whereas, we found the T-786C SNP, in modest LD with
the 27-bp repeat in our population. This was in agree-
ment with the Japanese study that reported a similar
linkage for these two polymorphisms [13]. Moreover,
we found no significant difference in the LD pattern of
cases and controls (Fig. 1), ascertaining that the eNOS
gene polymorphisms can be ruled out as risk factors for
the aSAH phenotype. However, the arterial walls of in-
dividuals with risk genotypes are likely to be more sus-
ceptible to environmental insults which are influenced
by ethnicity and social factors.

Table 5

| Variables         | OR  | 95% C.I     | P-value |
|-------------------|-----|-------------|---------|
| Age at onset      | 1.007 | 0.96–1.05   | 0.754   |
| Age above 50 yrs  | 1.846 | 0.55–6.25   | 0.324   |
| Female sex        | 1.373 | 0.60–3.16   | 0.455   |
| Diabetes mellitus | 0.271 | 0.08–0.95   | 0.042   |
| Hypertension      | 7.327 | 2.54–21.17  | < 0.001 |
| Smoking habit     | 12.864 | 3.90–42.41  | < 0.001 |
| Alcohol use       | 0.377 | 0.11–1.32   | 0.128   |
| eNOS 786TC+CC     | 1.055 | 0.42–2.63   | 0.909   |
| eNOS 4ab+4aa      | 0.621 | 0.24–1.76   | 0.333   |
| eNOS 894GT+TT     | 0.909 | 0.42–1.98   | 0.811   |

In a complex disease, apart from the disease per se,
factors and circumstances that associate with the dis-
ease have as much importance. In India, in addition
to multicultural ethnicity the prevalence of risk factors
such as hypertension (54.5%), and smoking (18.4%),
that associate with aSAH are also reported to be high-
er than the global average burden of these individual
risk factors [7,24,26]. The difference of our results and
those of the earlier groups can be explained in terms of
the different allele frequencies, shown explicitly by the
varying LD patterns as well as the prevalence of risk
factors in the patient groups taken from different ethnic
backgrounds. The putative functions of the eNOS poly-
morphisms or its linked variants at other sites may be
dependent on the presence of environmental risk fac-
tors. However, our observations indicate hypertension
and smoking as significant independent risk factors,
which do not associate with the risk genotypes. It has
been reported that when the risk of disease conferred

4. Discussion

The 27-bp VNTR 4a/4b allele was calculated as

\[ p_{SAH} = 0.265 \]

and

\[ p_C = 0.133 \]

two frequencies of the VNTR 4a allele in patients with aSAH and controls, respectively, reported in a previous study [10]. Using the formula as described by Cohen, we obtained

\[ Z_{1-\beta} = 2.54. \]

This gives the value of \( t \)-statistic to be referred to the table of normal distribution to obtain the power, \( 1 - \beta = 0.994. \)
Table 6

Comparison of genotype and allele frequencies of eNOS polymorphisms in South Indian and other study populations

| Polymorphism         | South India (present study as reference) | N. America (Khurana et al. 2004) | Europe (Krex et al. 2006) | Japan (Krischek et al. 2006) |
|----------------------|------------------------------------------|----------------------------------|---------------------------|-------------------------------|
|                      | n-224                                    | n-90                             | n-192                      | n-176                         |
| T-786C SNP           |                                          |                                  |                            |                               |
| T/T                  | 0.603                                    | 0.310                            | 0.386                      | 0.820                         |
| T/C                  | 0.366                                    | 0.510                            | 0.467                      | 0.130                         |
| C/C                  | 0.031                                    | 0.180                            | 0.147                      | 0.050                         |
| allele T             | P < 0.0001                               | P < 0.0001                       | P < 0.0001                 |                               |
| allele C             | 0.786                                    | 0.570                            | 0.619                      | 0.850                         |
| allele T             | P < 0.0001                               | P < 0.0001                       | P < 0.0001                 | P = 0.016                     |
| allele C             | 0.214                                    | 0.430                            | 0.381                      | 0.150                         |
| 27-bp VNTR           |                                          |                                  |                            |                               |
| 4b/4b                | 0.638                                    | 0.780                            | 0.667                      | 0.810                         |
| 4b/4a                | 0.344                                    | 0.180                            | 0.291                      | 0.170                         |
| 4a/4a                | 0.018                                    | 0.040                            | 0.042                      | 0.020                         |
| allele 4b            | P = 0.005                                | P = 0.21                         | P = 0.0005                 |                               |
| allele 4a            | 0.810                                    | 0.870                            | 0.812                      | 0.900                         |
| allele 4b            | P = 0.09                                 | P = 0.94                         | P = 0.0006                 |                               |
| G894T SNP            |                                          |                                  |                            |                               |
| G/G                  | 0.710                                    | 0.220                            | 0.505                      | 0.820                         |
| G/T                  | 0.272                                    | 0.610                            | 0.400                      | 0.130                         |
| T/T                  | 0.018                                    | 0.170                            | 0.095                      | 0.050                         |
| allele G             | P < 0.0001                               | P < 0.0001                       | P = 0.001                  |                               |
| allele T             | 0.846                                    | 0.530                            | 0.705                      | 0.890                         |
| allele T             | P < 0.0001                               | P < 0.0001                       | P = 0.076                  |                               |
|                      |                                          |                                  |                            |                               |

*P value denotes individual difference between the present study and other study populations.

by the polymorphic variant of a haemostatic gene is reduced, it could be possible that environmental risk factors are so dominant that genetic polymorphisms provide a negligible and rather insignificant contribution to the overall risk [14].

In conclusion, we could not observe any role of haploinsufficiency or homozygosity of the risk alleles of eNOS gene in causing intracranial aneurysm or aSAH, suggesting that genetic variants in the eNOS gene responsible for the regulation of haemodynamic stress in the arterial endothelium are unlikely to be susceptibility factors for the disease. It is however possible, that rare variants in other regions in the eNOS gene may have a role in aSAH. It is also conceivable that causative polymorphisms could be lying in genes involved in the upstream events of NO formation. Our study contradicts the seminal work on the role of eNOS in aSAH in North American population but is in agreement with majority of the reports on lack of association in European and Asian populations. The study also supports that ethnic background coupled with different environmental factors might associate to aggravate the pathogenesis of the disease. However, the possibility of a multigenic combination in the NO pathway genes coupled with a genotypic tag for associated risk phenotypes such as hypertension and cigarette smoking as a causative event in the pathogenesis of aSAH cannot be ruled out.

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