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Accessibility
Feather development genes and associated regulatory innovation predate the origin of Dinosauria

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The evolution of avian feathers have recently been illuminated by fossils and the identification of genes involved in feather patterning and morphogenesis. However, molecular studies have focused mainly on protein-coding genes. Using comparative genomics and more than 600,000 conserved regulatory elements, we show that patterns of genome evolution in the vicinity of feather genes are consistent with a major role for regulatory innovation in the evolution of feathers. Rates of innovation at feather regulatory elements exhibit an extended period of innovation with peaks in the ancestors of amniotes and archosaurs. We estimate that 86% of such regulatory elements were present prior to the origin of Dinosauria. On the branch leading to modern birds, we detect a strong signal of regulatory innovation near IGFBP2 and IGFBP5, which have roles in body size reduction, and may represent a genomic signature for the miniaturization of dinosaurian body size preceding the origin of flight.
Feathers constitute complex branched structures that arise through interactions between the dermis and epidermis (Widelitz et al. 2003; Mou et al. 2011; Ng et al. 2012; Li et al. 2013; Lin et al. 2013). Although feathers were long thought to be a key innovation associated with the origin of avian flight, paleontological discoveries over the past fifteen years indicate a more ancient origin; filamentous feather precursors are now known to be present in many lineages of non-avian dinosaurs, and pennaceous feathers clearly arose prior to the origin of flight (Xu et al. 2001; Norell and Xu 2005; Zheng et al. 2009; Kellner et al. 2010; Godefroit et al. 2014). At the same time, the molecular processes underlying feather development and deployment throughout the integument are becoming better known through studies of gene expression patterns (Antin et al. 2014) and natural mutants (Mou et al. 2011; Ng et al. 2012). Comparative genomics can offer insights into the evolutionary history of functional elements in the genome; however, aside from the β-keratins, which are known to have diversified extensively on the lineage leading to birds (Li et al. 2013), we know little about evolutionarily novel genes or noncoding regions associated with feather development. Recent studies have shown that regulatory changes underlie many key phenotypes in vertebrates (Karlsson et al. 2007; Chan et al. 2010; McLean et al. 2011; reviewed in Wray 2013), but regulatory innovations associated with the origins of feathers have not been systematically explored. In particular, conserved non-exonic elements (CNEEs) have emerged as important regulators of gene expression (Visel et al. 2008) and have revealed the evolutionary dynamics of genomic regions associated with novel phenotypes such as mammalian hair (Lowe et al. 2011).
Results and Discussion

Conserved non-exonic elements and constraint in the avian genome. We identified a set of 193 genes that have been associated with feather development through mutant phenotypes or spatiotemporally restricted expression patterns (Supplementary Materials and Supplementary Table 1). To investigate the evolutionary history of these genes and their potential regulatory elements, we constructed a 19-way whole-genome alignment referenced on the chicken genome (Hillier et al. 2004) containing four birds, two crocodilians, two turtles, a lizard, four mammals, a frog, and five actinopterygian (ray-finned) fish. Regions of the genome showing evolutionary constraint were identified using a phylogenetic hidden Markov model to detect regions of the alignment evolving more slowly than synonymous sites in coding regions. Overall, 957,409 conserved elements totaling ~71Mbp and spanning ~7.2% of the chicken genome were identified, a higher percentage than the 5% often reported for the human genome. This result is consistent with the small (1.2 Gb) size of the chicken genome relative to the human genome, making the total amount of sequence annotated as constrained about half of what is currently reported for human (Siepel et al. 2005; Lindblad-Toh et al. 2011). To identify putative regulatory elements we removed any regions overlapping an exon annotated in chicken, or another species, resulting in 602,539 CNEEs covering 4.4% of the chicken genome. We identified the gene that each CNEE is likely to regulate by assigning each CNEE to the gene with the closest transcription start site, and found that 13,307 of the CNEEs were associated with the 193 feather-related genes in the data set. Although regulatory elements can act over long genomic distances that include genes not regulated by the elements (Kleinjan and
van Heyningen 2005), experimentally identified enhancers tend to be closest to genes with expression in the same tissues and at the same times in development (Visel et al. 2009). Additionally, many regulatory regions undergo rapid evolution and turnover (Wray 2007; Wray 2013), and these will be missed by our analysis. Due to their different functions, we split the list of 193 feather related genes and their associated CNEEs into a structural set of 67 keratin genes and a patterning set of 126 non-keratin genes and analyzed these groups separately.

**An ancient genic toolkit and extended regulatory evolution are associated with feather origins.** The genic and regulatory components of the keratin and non-keratin sets show very different patterns across the 500My backbone of our tree, on the lineage leading from the common ancestor of vertebrates to the chicken in our tree (Figs. 1 and 2, Supplementary Fig. 1). The most ancient branch in our analysis, leading to the common ancestor of ray-finned fishes and other vertebrates, shows the strongest enrichment for the non-keratin feather genes (1.7 times expected), with smaller numbers of non-keratin feather genes arising on branches leading to tetrapods and less inclusive clades (Fig. 1A, Fig. 2). No members of this non-keratin feather gene set are reconstructed to have arisen after the ancestor of birds and turtles. Although ancient genes are more likely to be studied during chick development, the non-keratin genes in our study were even more ancient than we would expect taking into account this bias (Mann-Whitney U test; p < 0.022; Supplementary Figure 2). The inferred first appearance of non-keratin protein-coding regions that are involved, for example, in placode patterning and feather ontogeny in birds is consistent with these genes being part of an ancient developmental toolkit (Figs. 1 and 2).
Surprisingly, the CNEEs associated with non-keratin feather related genes show the highest rate of origin not on the internode between the ancestral archosaur and birds, where they exhibit a 25% higher-than-expected rate of origination, but instead on the branch leading to amniotes, where they exhibit a rate of origination 60% higher than expected (Figs. 1 and 2, Supplementary Fig. 1). The rate of origination for these CNEEs is greater than what would be expected from CNEEs uniformly distributed throughout the genome for 6 of the 8 branches along the lineage leading to chicken, suggesting a large amount of regulatory innovation over an extended time period (Figs. 1A and 2). Thus, the non-keratin genic component of feather development arose deep in vertebrates and the greatest signal of regulatory innovation was coincident with the burst of phenotypic change associated with the transition to land. Although information on the integument of the ancestral amniote remains exceptionally limited (Alibardi et al. 2009; Alibardi 2012), the accumulation of CNEEs inferred to have occurred at this time indicates a key role for regulatory change during this transition and in the subsequent evolution of vertebrate integumentary diversity. Consistent with this hypothesis, 32 genes in our feather gene set are here identified as shared with those involved in the development of mammalian hair (Lowe et al. 2011) (hypergeometric distribution, p < 1e-80; Supplementary Table 3) and as present in the amniote ancestor. Genes driving hair development have been previously shown to exhibit an increase in regulatory innovation on the branch leading to amniotes, followed by a peak on the branch leading to mammals and a decline more recently (Lowe et al. 2011).

Our analysis suggests that non-avian dinosaurs, as part of Archosauria, possessed the entirety of the known non-keratin protein-coding toolkit for making
feathers. Moreover, assuming a constant rate of genome-wide accumulation of CNEEs throughout vertebrates, we estimate that 86% of non-keratin feather gene CNEEs were also present in the archosaur ancestor. The CNEEs present in this ancestor may have less to do with feather origins but instead could be linked to the earlier amniote transition to land, with later, bird-specific CNEEs having feather-specific functions. These results are also consistent with new data on integumentary innovation and diversity in Archosauria: filamentous or bristle structures either originated once early in the clade or three or more times (Clarke 2013) in pterosaurs (Kellner et al. 2010), ornithischian (Zheng et al. 2009; Godefroit et al. 2014) and theropod dinosaurs (Norell and Xu 2005). Thus, the genic and regulatory complement identified in the ancestral archosaur was either a flexible toolkit coopted in multiple origins of new structures including feathers, or indicates an ancient origin in that clade for filamentous integumentary structures, often called feather precursors, on some part of the body or stage in development more than 100 million years before the origin of pinnate feathers in dinosaurs.

**Limited role of protein evolution in feather origins.** Our analysis detects the well-known burst of duplication in β-keratin genes within Archosauria (Greenwold and Sawyer 2010; Li et al. 2013) on the branch leading to birds (Figs. 1B, 2). The larger peak for keratin innovation is comprised of 57 β-keratins arising as an expansion of a gene cluster on chicken chromosome 27 and 5 β-keratins from duplications on chromosome 2. The small peak in the turtle-bird ancestor is due to the expansion of a β-keratin gene cluster on chromosome 25. Both of these results are consistent with
previous studies of β-keratin evolution (Greenwold and Sawyer 2010; Li et al. 2013).

However, this keratin burst constitutes the only, albeit substantial, signal of innovation at the protein level in pinnate feather origins. Notably, there is little evidence for regulatory innovation in the vicinity of β-keratin genes. We detected little additional cross-species constraint outside of the exonic regions in the keratin clusters than we would expect if CNEEs were randomly distributed in the genome. We only detected 15 CNEEs neighboring feather-related keratins in the lineage leading to birds, suggesting that regulatory evolution near β-keratins is not exceptional. Although the signature of CNEEs is likely complicated by a history of duplication and gene conversion in this multigene family, either the regulatory landscape around β-keratins does not appear noteworthy or their regulatory elements are under less severe constraint. These data are consistent with the idea that the keratin component of feathers arose primarily as a result of genic innovations.

Aside from β-keratin evolution, protein evolution appears to play a limited role in pinnate feather origins. We searched for signals of positive selection with respect to amino acid substitutions. After Bonferroni correction, only 3 of the 126 non-keratin feather genes showed signatures of positive selection on the archosaurian branch leading to birds (Supplementary Table 2). These results indicate that most non-keratin genes related to feather development exhibit regulatory, not protein-coding, innovations in the avian stem lineage, including living birds and non-avian dinosaurs, consistent with the hypothesis that regulatory innovations underlie adaptations in skin patterning and feather morphology.
Body size genes exhibit exceptional regulatory innovation in Dinosauria: Genes with an anomalously large number of regulatory elements arising in birds after their divergence from extant crocodilians may contribute to the origin of avian phenotypes. A genome-wide survey of 1 Mb genomic windows revealed 23 segments of the chicken genome possessing anomalously high numbers of CNEEs arising on the branch leading to birds (Fig. 3a; corrected p < 0.01; Supplementary Table 4). Although gene ontology analysis does not reveal significant enrichment for any functions for the set of genes near these innovation-rich segments, a number of these segments flank genes involved in body size, limb development, and integument (Fig. 3a). The region showing the greatest enrichment for bird-specific CNEEs in the entire chicken genome, over 500 percent more than expected (p < 1^{-53}), is centered in a 400-kb gene desert with insulin-like growth factor binding protein (IGFBP) 2 and 5 being the two closest genes (Fig 3b and c). IGFBP2 is expressed in the chick apical ectodermal ridge and at the tips of the growth plates in the wing bud, contains single nucleotide polymorphisms linked to phenotypic variation in the limbs of chickens (McQueeney and Dealy 2001; Li et al. 2006), and lies in the signaling pathway of both body size and limb length in mammals and birds (Fisher et al. 2005; Sutter et al. 2007). IGFBP5 also plays important roles in limb development (McQueeney and Dealy 2001) and the reduction of body size (Salih et al. 2004). Its widespread expression during chick development (Antin et al. 2014) is consistent with a role for IGFBP5-associated regulatory elements in body size reduction. Body size and limb length are known to vary extensively across Dinosauria and have been proposed to play a key role in dinosaur evolutionary dynamics (Benson et al.)
2014), with miniaturization indicated by the fossil record to have preceded the origin of flight in Paraves (Turner et al. 2007; Lee et al. 2014), and changes in limb scaling within Maniraptora and continuing into birds associated with the origin of flight (Xu et al. 2001). Thus, analysis of patterns of regulatory innovation offer the potential to link genome evolution to key shifts in shape and form occurring in deep time.

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**Author Contributions** DH, CBL and SVE conceived the study; CBL and SVE collected and analyzed data; CBL, SVE and JC wrote the paper, with comments from all other authors.

**Author Information:** All alignments and genomic coordinates of genes and CNEEs used in the analysis can be found at [http://hgdev-lowec.cse.ucsc.edu/](http://hgdev-lowec.cse.ucsc.edu/)
Competing financial interests statement. The authors declare no competing financial interests. Correspondence and requests for materials should be addressed to sedwards@fas.harvard.edu, lowec@stanford.edu, or Julia_Clareke@jsg.utexas.edu.

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Figures (main text)

Figure 1. Feather development genes are ancient whereas associated CNEEs peak in the amniote ancestor. Evolutionary dynamics of a) non-keratin feather development genes and associated CNEEs (n = 126 genes) and b) keratin genes and associated CNEEs (n = 67 genes). The black horizontal line indicates the null expectation of the number of new genes (comparison to all genes in the genome) or CNEEs (a uniform distribution throughout the genome). Points above this line indicate lineages on which a higher-than-expected number of genes or CNEEs have arisen. Points on the X-axis correspond to the ancestors depicted in Fig. 2, with spacing proportional to divergence times as recorded in timetree.org (Hedges et al. 2006). In b, the larger peak is comprised of β-keratins arising from expansions of gene clusters on chicken chromosomes 27 and 2. The small peak in the turtle-bird ancestor is due to the expansion of a β-keratin gene cluster on chromosome 25. Both of these results are consistent with previous studies of β-keratin evolution (Greenwold and Sawyer 2010; Li et al. 2013).

Figure 2. Major genomic events underlying the origin of feathers. The colored backbone of the tree is comprised of three tracks: CNEEs, non-keratin feather genes (n=126), and keratin genes (n=67). Rates of origination of these three genomic classes are indicated by the colors for each stem internode and track in the tree, with blue colors indicating low origination rates and red colors indicating high origination rates. Key events at the level of coding regions (genes) and regulatory elements are indicated.
The colors of the silhouettes at right indicate the percent of the feather regulatory component present in the chicken genome inferred to have arisen in the ancestor of each indicated taxon. For example, the fish are inferred to possess about 28% of the CNEEs associated with feather genes in chicken, whereas 86% of the observed chicken CNEEs are inferred to have arisen by the ancestral archosaur, including non-avian dinosaurs.

Figure 3. Identification of regions of the avian genome with signatures for exceptional regulatory innovation on the archosaur lineage that includes birds and other dinosaurs. a) A genome-wide plot of the density of conserved nonexonic elements (CNEEs) arising on the archosaurian branch leading to the avian ancestor. Red regions indicate those areas enriched compared to the distribution of CNEEs on other branches (gray line in ‘b’) and green squares indicate the 23 significant peaks of enrichment for bird-specific CNEEs relative to a uniform distribution throughout the genome. We examined the closest upstream and closest downstream genes and for select peaks a flanking gene is indicated along with a proposed role in avian morphological evolution (key at top); regulatory innovation may also have played a role in earlier dinosaur-lineage evolutionary dynamics. b) The densest region for bird-specific CNEEs in the chicken genome is in a gene desert on chromosome 7 with IGFBP2 being the closest well-annotated refseq gene and IGFBP5 being the closest gene prediction. CNEE density on all branches other than the one leading to birds is indicated in grey. c) UCSC Genome Browser shot of a CNEE-rich region in the vicinity
of IGFBP2 and IGFBP5, which function in limb development and body size regulation (see main text, Supplementary Table 4), showing CNEEs found only in birds (red boxes) or arising on deeper branches in the vertebrate tree (gray boxes). Regions of aligning sequence for representatives of the 19 included taxa are in green.
Extended accumulation of regulatory CNEEs with ~86% present in the archosaur ancestor

Burst of *keratin gene duplication* within archosauria

Evolution of full complement of *feather patterning genes*

Bony vertebrates

CNEEs

pattern genes

keratin genes

0.3 subst. / site

Galliformes

Chickens

Turkey

Pigeon

Zebra finch

Non-avian dinosaurs

American alligator

Saltwater crocodile

Soft-shell turtle

Painted turtle

Anolis lizard

Human

Mouse

Dog

Opposum

Xenopus frog

Tetraodon

Fugu

Stickleback

Medaka

Zebrafish

Reptilia

Amniota

Archosauria

Dinosauria

Aves

Bony vertebrates
Figure 3

(a) CNEEs on the avian branch
- Enriched compared to other branches
- Enrichment peaks compared to uniform distribution
- Genes associated with feathers
- Genes associated with limb development
- Genes associated with body size

(b) CNEEs per Mb

(c) Conserved non-exonic elements
- Alignment and conservation
Supplementary Material

Lowe et al. “Feather development genes and associated regulatory innovation predate the origin of Dinosauria”

Supplementary Materials and Methods:

**Genome-wide Multiple Alignment.** Our genome-wide alignment referenced on the chicken genome contains the following species and build numbers: Chicken (*Gallus gallus*; galGal3), Turkey (*Meleagris pavo*; melGal1), Pigeon (*Columba livia*; colLiv1), Zebra Finch (*Taeniopygia guttata*; taeGut1), American Alligator (*Alligator mississippiensis*; allMis1), American Crocodile (*Crocodylus acutus*; croPor0), Soft-shelled Turtle (*Pelodiscus sinensis*; pelSin1), Painted Turtle (*Chrysemys picta belli*; chrPic1), Green Anole (*Anolis carolinensis*; anoCar2), Human (*Homo sapiens*; hg18), House Mouse (*Mus domesticus*; mm9), Dog (*Canis lupus familiaris*; canFam2), Opossum (*Monodelphis domestica*; monDom4), Xenopus (*Xenopus tropicalis*; xenTro3), Tetraodon (*Tetraodon nigriviridis*; tetNig1), Fugu (*Takifugu rubripes*; fr2), Stickleback (*Gasterosteus aculeatus*; gasAcu1), Medaka (*Oryzias latipes*; oryLat2), and Zebrafish (*Danio rerio*; danRer5). Initial pairwise alignments were done between the chicken genome and all other genomes using Blastz(Schwartz et al. 2003). We then chained these local alignments together to construct whole-genome alignments(Kent et al. 2003). We applied a syntenic filter that requires a consistent order and orientation between the species, which helped distinguish orthologs from paralogs(Kent et al. 2003). We then used Multiz(Blanchette et al. 2004) to create the final multi-species genome-wide
alignment from the filtered pairwise alignments. We expect that additional avian genomes (Ellegren et al. 2012; Huang et al. 2013; Qu et al. 2013; Rands et al. 2013; Zhan et al. 2013) will add resolution to this analysis, but we suspect that the fraction of the genome estimated as constrained in our study will be similar to those employing additional genomes.

**Identification of Conserved Non-Exonic Elements.** We used a phylogenetic hidden Markov model (HMM) to define regions of cross-species conservation in our genome-wide alignments (Siepel, Haussler 2004). The HMM had both a neutral state and a conserved state that resisted substitutions. We defined the neutral state by fitting a time-reversible model of nucleotide substitution to four-fold degenerate sites in codons (Siepel, Haussler 2004). The conserved state has the same tree topology, but with the branch lengths reduced to three tenths of the neutral rate. We set the transition probabilities of the HMM by expecting a length of 45 base pairs and a coverage of 0.3. We then filtered this set of conserved regions by removing any elements overlapping exons according to spliced expressed sequence tags, Ensembl genes (Hubbard et al. 2002), RefSeq genes (Pruitt, Tatusova, Maglott 2005) or Transmap (Zhu et al. 2007).

**Enrichment calculations for feather-related genes.** There is variable statistical power to identify conserved elements depending on the evolutionary age of the element. For this reason we report the fraction of all CNEEs that arose on a branch that are associated with a given set of genes. We report enrichment as the fraction of CNEEs
arising on a branch near a set of genes divided by the fraction that would be expected near this set if CNEEs were uniformly distributed across the genome (Lowe, Bejerano, Haussler 2007). P-values for CNEE enrichment are calculated using the binomial distribution where the probability of success is the number of bases closest to a transcription start site for the set of genes being analyzed, divided by the number of bases in the genome. The number of trials is the number of CNEEs and the number of successes is the number of CNEEs closest to a transcription start site of a gene in the set being analyzed. For protein coding genes we report enrichment as the fraction of genes arising on a branch that are members of the gene set divided by the fraction of all genes that are in the gene set.

**Identifying the Branch of Origin for Genes and CNEEs.** For each gene or CNEE we used the branch leading to the most recent common ancestor of chicken and the most divergent species with an ortholog present in the alignment as the time of origin. For CNEEs, an ortholog was annotated as present if the species had orthologous bases for at least one-third of the bases in the CNEE. For genes, the species had to have orthologous bases present for at least 50 percent of the isoform with the most exons.

**Genomic locations experiencing anomalous regulatory evolution leading to Aves.**

We compared the genomic locations of CNEEs appearing on the branch leading to Aves against a uniform distribution along the genome. For each base in the genome we
calculated the number of CNEEs arising on the avian branch within 0.5Mb in either
direction (roughly 1Mb windows). To identify the genomic location of peaks in this data
set we fit a cubic spline with a smoothing parameter of 0.35. We calculated the
significance of each peak based on the binomial distribution with the number of avian
CNEEs divided by the number of bases being the probability of success, 1,000,001
being the number of trials, and Bonferroni correcting based on the number of bases in
the genome. This resulted in a significance threshold of 76 CNEEs/Mb \( (p < 0.01) \). We
also tested for significance against the distribution of CNEEs originating outside the
avian branch being the null hypothesis instead of a uniform distribution across the
genome. In this case we performed a fishers exact test for every base in the genome,
comparing the number of avian CNEEs within the window to the number of non-avian
CNEEs within the same range. We again used a p-value threshold of 0.01 after
correcting for the number of tests being the number of bases in the genome.

**Estimating the percentage of CNEEs near feather genes present in archosaurs.**

To estimate the percentage of CNEEs present in Aves near non-keratin feather genes
that were already present in archosaurs we needed to adjust the number, but not
percentage, of feather-related CNEEs detected on each branch for the power to detect
conserved elements that originated at that time. This is because for very recently
originated sequence it is difficult to notice a resistance to substitutions, and for very
ancient elements it is difficult to align over long distances, even when the sequence is
resisting many substitutions. To do this we assume that CNEEs in general, but not those near feather-related genes, have accumulated at a constant rate through time relative to the speed of substitutions in neutral DNA. For each branch in our analysis we maintain the fraction of CNEEs near feather-related genes, but adjust the counts so that the total number of CNEEs arising on each branch, divided by the length of the branch in substitutions per site, is a constant for all branches in our analysis.

\[ j = \{1, 2, 3, 4, 5, 6, 7\} \text{ with } 1 \text{ being the branch leading to Aves and higher numbers representing progressively more ancient branches.} \]

\[ b_j = \text{length in substitutions per site of branch } #j \]

\[ f_j = \text{number of feather-related CNEEs detect as originating on branch } #j \]

\[ a_j = \text{number of CNEEs detected as originating on branch } #j \]

Fraction present in archosaurs =

\[ \frac{\sum_{j=2}^{7} f_j a_j b_j}{\sum_{j=1}^{7} a_j b_j} \]

**Analysis of protein evolution.** Alignments of the protein-coding exons for the 126 non-keratin feather genes were extracted from the 19-way genomic alignment. To test the hypothesis that adaptive evolution of amino acid substitutions occurred on the branch leading to extant birds, we ran two branch-site models under Model A in the
package Phylogenetic Analysis using Maximum Likelihood 4 (PAML4, version 4.4). The null (neutral or purifying selection) model allows two values of $\omega$, the ratio of nonsynonymous to synonymous substitutions, to vary between 0 and 1 ($\omega_0$, site classes 0 and 2a) or to be constrained to equal 1 ($\omega_1$, site classes 1 and 2b). The alternate model (allowing for positive selection) enables $\omega$ to be equal to or exceed 1 for some sites (classes 2a and 2b) for the avian branch. For each gene we recorded the likelihood of both models, conducted a likelihood ratio test, and used Bonferroni correction with the number of tests being equal to the number of genes. The results suggest that 3 out of 126 genes rejected the null model at $p < 0.01$. 
Supplementary Figure 1 - Dynamics of feather-related gene and CNEE innovation across vertebrates. The key indicates symbols used to designate gene and CNEE trajectories. In both panels, the black horizontal line indicates the null expectation of the number of genes or CNEEs and the blue area indicates the 99% confidence limits on the fold-enrichment for CNEEs on a given branch. a, Dynamics of non-keratin feather-related genes and associated CNEEs (n = 126 genes). b, Dynamics of keratin genes and associated CNEEs (n = 67 genes). The green spike at the right consists of β-keratin genes inferred to have duplicated along the lineage leading to birds.
Supplementary Figure 2 - Feather related genes are more ancient than expected for the chicken genome. The distribution of origin for the 126 feather patterning genes (red) is more ancient when compared to both the entire set of Ensembl genes (blue; Mann-Whitney U test; p < 1e-9; see Hubbard et al. 2002) as well as the set of genes with Ensembl identifiers that have been deposited in a database of in situ hybridizations during chick development (green; Mann-Whitney U test; p < 0.022; (Antin et al. 2014)).
Supplementary Table 1 - Keratin and non-keratin feather gene set.

| Keratin or non-Keratin | Gene ID          | Gene Symbol | PMID     |
|------------------------|------------------|-------------|----------|
| Non-keratin            | ENSGALG000000004212 | Adam10      | 21780243 |
| Non-keratin            | ENSGALG000000012740 | Adam12      | 21158755 |
| Non-keratin            | ENSGALG000000016427 | Adam17      | 21780243 |
| Non-keratin            | ENSGALG000000008976 | Adam22      | 21780243 |
| Non-keratin            | ENSGALG000000008582 | Adam23      | 21780243 |
| Non-keratin            | ENSGALG000000003398 | Adam9       | 21780243 |
| Non-keratin            | ENSGALG000000004270 | Aldh1a2     | 21423653 |
| Non-keratin            | ENSGALG000000007129 | Aldh1a3     | 21423653 |
| Non-keratin            | ENSGALG000000023542 | Alpl        | 8563025  |
| Non-keratin            | ENSGALG000000006995 | ApoD        | 15580625 |
| Non-keratin            | ENSGALG000000021455 | Asip        | 18287407 |
| Non-keratin            | ENSGALG000000008830 | Bmp2        | 17948257 |
| Non-keratin            | ENSGALG000000012429 | Bmp4        | 8823374  |
| Non-keratin            | ENSGALG000000007668 | Bmp7        | 15272377 |
| Non-keratin            | ENSGALG000000015132 | Cdh2        | 19557684 |
| Non-keratin            | ENSGALG00000013034  | Ccr2        | 20336606 |
| Non-keratin            | ENSGALG00000007280  | Cldn1       | 15749086 |
| Non-keratin            | ENSGALG000000009641 | Col1a2      | 15272377 |
| Non-keratin            | ENSGALG00000002552  | Col3a1      | 15272377 |
| Non-keratin            | ENSGALG00000008494  | Creb        | 7556946  |
| Non-keratin            | ENSGALG00000010257  | Crp2        | 12128214 |
| Non-keratin            | ENSGALG00000011905  | Ctnnb1      | 12949772 |
| Non-keratin            | ENSGALG00000011182  | Delta1      | 9435296  |
| Non-keratin            | ENSGALG00000004256  | Dhrs3       | 21423653 |
| Non-keratin            | ENSGALG00000004481  | Eda         | 17948257 |
| Non-keratin            | ENSGALG00000004599  | Eda2r       | 17362907 |
| Non-keratin            | ENSGALG00000016809  | Edar        | 17948257 |
| Non-keratin            | ENSGALG00000014369  | Edaradd     | 15673574 |
| Non-keratin            | ENSGALG00000007466  | Ednrnb2     | 18625062 |
| Non-keratin            | ENSGALG00000012155  | Egf         | 12586066 |
| Non-keratin            | ENSGALG00000012363  | Egfr        | 12586066 |
| Non-keratin            | ENSGALG00000012120  | En1         | 9012506  |
| Non-keratin            | ENSGALG00000016428  | Enpp2       | 17366625 |
| Non-keratin            | ENSGALG00000005256  | Epha4       | 12949772 |
| Non-keratin            | ENSGALG00000001143  | Ets1        | 9074942  |
| Non-keratin            | ENSGALG00000014872  | Fgf10       | 15201222 |
| Non-keratin            | ENSGALG00000002203  | Fgf18       | 11669372 |
| Non-keratin            | ENSGALG00000011835  | Fgf2        | 15201222 |
| Non-keratin            | ENSGALG00000013663  | Fgf20       | 22712610 |
| Non-keratin            | ENSGALG00000007563  | Fgf3        | 15201222 |
| Non-keratin            | ENSGALG00000007562  | Fgf4        | 15201222 |
| Non-keratin            | ENSGALG00000007706  | Fgf8        | 9012506  |
| Non-keratin            | ENSGALG00000003311  | Fgfr1       | 15201222 |
| Non-keratin            | ENSGALG00000009495  | Fgfr2       | 21302260 |
| Protein Type | Gene ID | Gene Symbol |
|--------------|---------|-------------|
| Keratin      | ENSGALG000000024142 | Fker        |
| Non-keratin  | ENSGALG000000003578 | Fn1         |
| Non-keratin  | ENSGALG000000023293 | Foxe1       |
| Non-keratin  | ENSGALG00000003654 | Foxn1       |
| Non-keratin  | ENSGALG00000014908 | Fst         |
| Non-keratin  | ENSGALG00000006902 | Fz1         |
| Non-keratin  | ENSGALG00000023647 | Gbx1        |
| Non-keratin  | ENSGALG00000013342 | Gbx2        |
| Non-keratin  | ENSGALG00000011059 | Hoxa6       |
| Non-keratin  | ENSGALG00000009273 | Hoxd11      |
| Non-keratin  | ENSGALG00000009270 | Hoxd8       |
| Non-keratin  | ENSGALG00000009546 | Htra1       |
| Non-keratin  | ENSGALG00000005499 | If17rd      |
| Non-keratin  | ENSGALG00000001405 | Irf6        |
| Non-keratin  | ENSGALG00000007145 | Itgb1       |
| Non-keratin  | ENSGALG00000009020 | Jag1        |
| Non-keratin  | ENSGALG00000011696 | Jag2        |
| Non-keratin  | ENSGALG00000013925 | Kit         |
| Non-keratin  | ENSGALG00000011206 | Kitlg       |
| Keratin      | ENSGALG00000019719 | Krt19       |
| Keratin      | ENSGALG00000019716 | Krt20       |
| Keratin      | ENSGALG00000002349 | Krt75       |
| Non-keratin  | ENSGALG00000004284 | Lfg         |
| Non-keratin  | ENSGALG0000000936  | Lmx1        |
| Keratin      | ENSGALG00000024227 | LOC425854   |
| Keratin      | ENSGALG00000024220 | LOC427060   |
| Keratin      | ENSGALG00000024185 | LOC770926   |
| Non-keratin  | ENSGALG00000023459 | Mc1r        |
| Non-keratin  | ENSGALG00000007679 | Mitf        |
| Non-keratin  | ENSGALG00000003580 | Mmp2        |
| Non-keratin  | ENSGALG00000006438 | Mnx1        |
| Non-keratin  | ENSGALG00000015013 | Msl1        |
| Non-keratin  | ENSGALG0000002947  | Msx2        |
| Non-keratin  | ENSGALG0000005911  | Mtf2        |
| Non-keratin  | ENSGALG00000013956 | Myb         |
| Non-keratin  | ENSGALG00000016308 | Myc         |
| Non-keratin  | ENSGALG00000007839 | Ncam1       |
| Non-keratin  | ENSGALG00000003114 | Nog         |
| Non-keratin  | ENSGALG00000002375 | Notch1      |
| Non-keratin  | ENSGALG00000002922 | Notch2      |
| Non-keratin  | ENSGALG0000001024  | P63         |
| Non-keratin  | ENSGALG0000002553  | Pcdh1       |
| Non-keratin  | ENSGALG00000016941 | Pcdh17      |
| Non-keratin  | ENSGALG00000009732 | Pcdh18      |
| Non-keratin | ENSGALG000000006822 | Pcdh19 | 23318466 |
| Non-keratin | ENSGALG000000016944 | Pcdh8  | 23318466 |
| Non-keratin | ENSGALG000000016908 | Pcdh9  | 23318466 |
| Non-keratin | ENSGALG000000014634 | Pmel   | 17106652 |
| Non-keratin | ENSGALG000000016600 | Pomc   | 21187100 |
| Non-keratin | ENSGALG00000003446  | Prlr   | 22297689 |
| Non-keratin | ENSGALG00000012620  | Ptc1   | 16330160 |
| Non-keratin | ENSGALG000000010133 | Ptch2  | 11784016 |
| Non-keratin | ENSGALG000000007130 | Rac1   | 15136143 |
| Non-keratin | ENSGALG00000002859  | Rac3   | 15136143 |
| Non-keratin | ENSGALG000000011298 | Rarb   | 8619957 |
| Non-keratin | ENSGALG00000002841  | Rfng   | 10704863 |
| Non-keratin | ENSGALG00000016485  | Rhob   | 15136143 |
| Non-keratin | ENSGALG00000000569  | Sdc3   | 7729580 |
| Non-keratin | ENSGALG00000009241  | Sfrp2  | 15272377 |
| Non-keratin | ENSGALG00000006379  | Shh    | 18079472 |
| Non-keratin | ENSGALG000000016698 | Shox   | 16904661 |
| Non-keratin | ENSGALG00000004885  | Slc24a5| 18287407 |
| Non-keratin | ENSGALG00000003310  | Slc45a2| 18625062 |
| Non-keratin | ENSGALG00000007870  | Smad3  | 16330160 |
| Non-keratin | ENSGALG00000018639  | Smad7  | 16330160 |
| Non-keratin | ENSGALG00000015310  | Sodp   | 22802190 |
| Non-keratin | ENSGALG00000010797  | Sostdc1| 21423653 |
| Non-keratin | ENSGALG00000005984  | Sox18  | 11418236 |
| Non-keratin | ENSGALG00000005285  | Tbx4   | 16330160 |
| Non-keratin | ENSGALG00000009612  | Tgf2   | 8903351 |
| Non-keratin | ENSGALG00000011442  | Tgfr2  | 8903351 |
| Non-keratin | ENSGALG00000011294  | Thrb   | 10828844 |
| Non-keratin | ENSGALG00000008120  | Tle3   | 12666196 |
| Non-keratin | ENSGALG00000007113  | Tnc    | 12949772 |
| Non-keratin | ENSGALG00000017119  | Tnfrsf19| 17362907 |
| Non-keratin | ENSGALG00000016971  | TSC-22 | 11803572 |
| Non-keratin | ENSGALG00000004274  | Twist2 | 11744368 |
| Non-keratin | ENSGALG00000017237  | Tyr    | 8612714 |
| Non-keratin | ENSGALG00000015205  | Typr1  | 18625062 |
| Non-keratin | ENSGALG0000000839   | Wnt11  | 7779076 |
| Non-keratin | ENSGALG00000010797  | Wnt3   | 18394491 |
| Non-keratin | ENSGALG00000011358  | Wnt6   | 17948257 |
| Non-keratin | ENSGALG00000005123  | Wnt7a  | 12949772 |
| Non-keratin | ENSGALG00000005668  | Wsb1   | 10354473 |
| Keratin    | ENSGALG00000014956  | Wnt12  | 19512870 |
| Keratin    | ENSGALG00000010797  | Wnt3   | 18394491 |
| Keratin    | ENSGALG00000011358  | Wnt6   | 17948257 |
| Keratin    | ENSGALG00000005123  | Wnt7a  | 12949772 |
| Keratin    | ENSGALG00000005668  | Wsb1   | 10354473 |
| Keratin | ENSGALG00000022757 | 20482795 |
|---------|--------------------|---------|
| Keratin | ENSGALG00000024140 | 20482795 |
| Keratin | ENSGALG00000024143 | 20482795 |
| Keratin | ENSGALG00000024144 | 20482795 |
| Keratin | ENSGALG00000024145 | 20482795 |
| Keratin | ENSGALG00000024147 | 20482795 |
| Keratin | ENSGALG00000024154 | 20482795 |
| Keratin | ENSGALG00000024174 | 20482795 |
| Keratin | ENSGALG00000024179 | 20482795 |
| Keratin | ENSGALG00000024180 | 20482795 |
| Keratin | ENSGALG00000024181 | 20482795 |
| Keratin | ENSGALG00000024184 | 20482795 |
| Keratin | ENSGALG00000024186 | 20482795 |
| Keratin | ENSGALG00000024187 | 20482795 |
| Keratin | ENSGALG00000024188 | 20482795 |
| Keratin | ENSGALG00000024189 | 20482795 |
| Keratin | ENSGALG00000024190 | 20482795 |
| Keratin | ENSGALG00000024191 | 20482795 |
| Keratin | ENSGALG00000024192 | 20482795 |
| Keratin | ENSGALG00000024194 | 20482795 |
| Keratin | ENSGALG00000024197 | 20482795 |
| Keratin | ENSGALG00000024198 | 20482795 |
| Keratin | ENSGALG00000024200 | 20482795 |
| Keratin | ENSGALG00000024201 | 20482795 |
| Keratin | ENSGALG00000024202 | 20482795 |
| Keratin | ENSGALG00000024205 | 20482795 |
| Keratin | ENSGALG00000024206 | 20482795 |
| Keratin | ENSGALG00000024208 | 20482795 |
| Keratin | ENSGALG00000024209 | 20482795 |
| Keratin | ENSGALG00000024212 | 20482795 |
| Keratin | ENSGALG00000024215 | 20482795 |
| Keratin | ENSGALG00000024216 | 20482795 |
| Keratin | ENSGALG00000024218 | 20482795 |
| Keratin | ENSGALG00000024219 | 20482795 |
| Keratin | ENSGALG00000024221 | 20482795 |
| Keratin | ENSGALG00000024222 | 20482795 |
| Keratin | ENSGALG00000024225 | 20482795 |
| Keratin | ENSGALG00000024226 | 20482795 |
| Keratin | ENSGALG00000024228 | 20482795 |
| Keratin | ENSGALG00000024229 | 20482795 |
| Keratin | ENSGALG00000024230 | 20482795 |
| Keratin | ENSGALG00000024233 | 20482795 |
| Keratin | ENSGALG00000024236 | 20482795 |
| Keratin | ENSGALG00000024237 | 20482795 |
| Keratin | ENSGALG00000024238 | 20482795 |
| Keratin | ENSGALG00000024239 | 20482795 |
| Keratin | ENSGALG00000024240 | 20482795 |
| Keratin | ENSGALG00000024241 | 20482795 |
Gene ID provides Ensemble ID numbers for chicken. PMID, Pubmed ID, for source publication. To construct our candidate gene set for feather-related genes, we conducted a literature search in Web of Science through the search engine in Endnote X5, using “feather AND gene*” as keywords, with no constraints on year of publication. We conducted our last such systematic search on February 18, 2013, yielding approximately 400 abstracts, but added additional abstracts as recently as November 2013. We recorded genes for which a mutation is known to cause a feather phenotype. We also included genes where the pattern of expression was spatially restricted to placodes, feather buds, or feathers during development.

| Keratin | ENSGALG00000024242 | 20482795 |
|---------|--------------------|---------|
| Keratin | ENSGALG00000024245 | 20482795 |
| Keratin | ENSGALG00000024246 | 20482795 |
| Keratin | ENSGALG00000024247 | 20482795 |
**Supplementary Table 2 – Feather-related genes showing evidence of positive selection on the branch leading to birds.**

| Gene ID                 | Gene Symbol | omega class 2 (dn/ds) | ln(null model) | ln(allowing positive selection) | Test statistic (D) | P-value    |
|-------------------------|-------------|-----------------------|----------------|---------------------------------|--------------------|------------|
| ENSGALG00000002375      | NOTCH1      | 17.7                  | -60953.0571    | -60942.8249                    | 20.4645            | <8E-4      |
| ENSGALG00000002552      | COL3A1      | ds~ = 0               | -30962.3467    | -30953.8837                    | 16.9259            | <5E-3      |
| ENSGALG000000016941     | PCDH17      | 7                     | -17124.3081    | -17052.9813                    | 142.6535           | <1E-30     |

The test statistic D is twice the difference in log likelihood of the two models tested. P-value is adjusted for Bonferroni correction. Omega (\(\omega\)) indicates the signal for adaptive evolution on the branch leading to birds relative to the null model \(\omega = 1\).
Supplementary Table 3 - 32 Genes shared between feather and hair gene sets.

| Gene ID               | Gene Symbol |
|-----------------------|-------------|
| ENSGALG00000002375    | Notch1      |
| ENSGALG00000002922    | Notch2      |
| ENSGALG00000002947    | Msx2        |
| ENSGALG00000003310    | Slc45a2     |
| ENSGALG00000003446    | Prlr        |
| ENSGALG00000003654    | Foxn1       |
| ENSGALG00000004274    | Twist2      |
| ENSGALG00000004481    | Eda         |
| ENSGALG00000004885    | Slc24a5     |
| ENSGALG00000006379    | Shh         |
| ENSGALG00000007130    | Rac1        |
| ENSGALG00000007145    | Itgb1       |
| ENSGALG00000007280    | Cldn1       |
| ENSGALG00000007679    | Mitf        |
| ENSGALG00000007870    | Smad3       |
| ENSGALG00000011206    | Kitlg       |
| ENSGALG00000011905    | Ctnnb1      |
| ENSGALG00000012155    | Egf         |
| ENSGALG00000012363    | Egfr        |
| ENSGALG00000012620    | Ptc1        |
| ENSGALG00000013925    | Kit         |
| ENSGALG00000014369    | Edaradd     |
| ENSGALG00000014634    | Pmel        |
| ENSGALG00000015205    | Tyrp1       |
| ENSGALG00000016308    | Myc         |
| ENSGALG00000016427    | Adam17      |
| ENSGALG00000016600    | Pomc        |
| ENSGALG00000016809    | Edar        |
| ENSGALG00000017119    | Tnfrsf19    |
| ENSGALG00000017237    | Tyr         |
| ENSGALG00000021455    | Asip        |
| ENSGALG00000023293    | Foxe1       |

Hair gene set from Lowe et al. (2011).
## Supplementary Table 4 - Details of 23 regions of the chicken genome enriched for CNEEs arising on the branch leading to birds.

| chromosome | position | CNEEs/Mb | Gene upstream | Gene downstream | Gene prediction upstream (if closer) | Gene prediction downstream (if closer) |
|------------|----------|----------|---------------|-----------------|--------------------------------------|----------------------------------------|
| chr7       | 24496029 | 160.68   | TNS1          | IGFBP2          | _                                    | IGFBP5                                 |
| chr13      | 2187175  | 143.48   | DNAJC18       | PAIP2           | _                                    | SLC23A1                                 |
| chr13      | 1725780  | 138.48   | HDAC3         | PCDH1           | STARD10                              | _                                      |
| chr15      | 10772514 | 122.29   | DGCR6         | SF3A1           | RTN4R                                | OSP2                                   |
| chr15      | 10596727 | 119.21   | DGCR6         | SF3A1           | PRODH                                | RTN4R                                 |
| chr9       | 15448741 | 110.90   | TP63          | LPP             | _                                    | _                                      |
| chr9       | 14989167 | 103.35   | OSTN          | TMEM207         | IL1RAP                               | _                                      |
| chr6       | 23553695 | 96.69    | ATP6V0E2      | GOT1            | SLC25A28                             | _                                      |
| chr14      | 12900949 | 94.14    | AXIN1         | CLCN7           | _                                    | LUC7L                                  |
| chr8       | 20835190 | 90.99    | HY1           | ST3GAL3         | PTPRF                                | _                                      |
| chrZ       | 7694631  | 88.22    | CNTFR         | IL1RA           | _                                    | _                                      |
| chr19      | 4220049  | 86.95    | YWHAG         | HSPB1           | _                                    | _                                      |
| chr4       | 9222023  | 84.44    | AVP           | HTR7            | DDRGK1                               | _                                      |
| chr9       | 14259981 | 84.09    | ATP13A4       | FGF12           | HRASLS                               | MB21D2                                 |
| chr5       | 29829705 | 82.61    | COX16         | SLC8A3          | _                                    | _                                      |
| chr17      | 3783301  | 79.43    | TNC           | TLR4            | TRIM32                               | ASTN2                                  |
| chr17      | 3932933  | 79.38    | TNC           | TLR4            | TRIM32                               | ASTN2                                  |
| chr12      | 2017381  | 79.16    | CISH          | MST1            | DOCK3                                | MANF                                   |
| chr17      | 4269608  | 78.57    | TLR4          | BRIPN1          | _                                    | _                                      |
| chr7       | 27109460 | 77.65    | MRAS          | GLI2            | RALB                                 | _                                      |
| chr18      | 1148963  | 77.32    | MYOCD         | NDEL1           | DNAH9                                | SHISA6                                 |
| chr19      | 3930149  | 76.73    | CUX1          | PRKRI1P1        | _                                    | SH2B2                                  |
| chr8       | 5404210  | 76.17    | LMX1A         | PBX1            | _                                    | _                                      |

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