Comparative analysis of the expression patterns of metalloproteinases and their inhibitors in breast neoplasia, sporadic colorectal neoplasia, pulmonary carcinomas and malignant non-Hodgkin's lymphomas in humans

AE Kossakowska¹, SA Huchcroft², SJ Urbanski³ and DR Edwards¹

¹Department of Pathology, University of Calgary and Foothills Hospital, Calgary, Alberta T2N 2T9, Canada; ²Department of Epidemiology and Preventive Oncology, Alberta Cancer Board, Calgary, Alberta, Canada; ³Department of Pharmacology and Therapeutics, University of Calgary, Calgary, Alberta T2N 4N1, Canada.

Summary Matrix metalloproteinases (MMPs) and their inhibitors (tissue inhibitors of metalloproteinases, TIMPs) play essential roles in the remodelling of the extracellular matrix (ECM). Results of in vivo and in vitro studies suggest that the balance between MMPs and TIMPs is altered in neoplasia, contributing to the invasive and metastatic properties of malignant tumours. In this study we have analysed the expression of five MMP genes and TIMP-1 and TIMP-2 in 37 benign and malignant lesions of human breast using Northern blot analysis. MMP-9 (92 kDa gelatinase) and MMP-11 (stromelysin 3) were most consistently expressed by carcinomas. Based on detection of either MMP-9 or MMP-11 mRNAs, we were able to distinguish between malignant and benign lesions with a predictive and 85% specificity. Subsequently, these results were compared with results for carcinomas of colon and lung and malignant non-Hodgkin’s lymphomas (NHL). Elevated MMP-9 and TIMP-1 expression was observed in all four systems. MMP-11 characterised all carcinomas as well as carcinomas in situ but was not detectable in NHL. Our data therefore argue that there are remarkably similar patterns of specific functions involved in ECM remodelling that correlate with malignancy in different human tumours of different histogenesis. However, MMP-11 expression is a characteristic of tumours of epithelial origin that is not found in lymphoid neoplasia. Thus it suggests that MMP-11 may play a regulatory role in the invasion and metastasis of carcinomas.

Keywords: breast neoplasia; extracellular matrix; metalloproteinases; tissue inhibitors of metalloproteinases

The ability to breach tissue boundaries through active destruction of extracellular matrix (ECM) is a sine qua non of tumour invasion, dissemination and metastasis formation (Liotta et al., 1991). It is, in fact, the explicit definition of malignancy. It is therefore axiomatic that the events involved in tumour-mediated ECM dissolution can help to provide an understanding of tumour progression at the molecular level. Over the past two decades, there has been an explosion of information about the molecular processes involved in ECM breakdown (reviewed in Liotta et al., 1991; Mignatti et al., 1986; Mignatti and Rifkin, 1993; Stetler-Stevenson et al., 1993; Alexander and Werb, 1991). The picture of normal ECM remodelling that has emerged is of a highly regulated process involving: (1) multiple secreted proteases from different families; (2) the controlled local activation of these enzymes; and (3) the actions of specific protease inhibitors. The family of zinc-dependent matrix metalloproteinases (MMPs) that includes collagenases, stromelysins and gelatinases (also known as type IV collagenases) has achieved pre-eminence since in vivo work has shown that, in many situations, they are the principal players responsible for the actual attack on ECM components (Alexander and Werb, 1991; Mignatti et al., 1986). In particular, several family members, including the 72 kDa and 92 kDa type IV gelatinases A and B (MMP-2 and MMP-9) can degrade basement membrane components and have been shown to be instrumental in tumour invasion in vivo (Liotta et al., 1991; Mignatti and Rifkin, 1993; Stetler-Stevenson et al., 1993; Collier et al., 1988; Wilhelm et al., 1989; Nakajima et al., 1987; Watanabe et al., 1993; Jurgenson et al., 1993). In one of these studies we showed, using deregulation of the balance between MMPs and their inhibitors, the tissue inhibitors of metalloproteinases (TIMPs) have been linked to invasive and metastatic behaviour (Liotta et al., 1991; Albini et al., 1991; Khokha et al., 1989, 1992; DeClerck et al., 1991, 1992).

Several pieces of evidence suggest that MMP/TIMP imbalance may be a significant factor in the pathophysiology of mammary cancer. Work with rodent mammary carcinoma cell lines has shown that metastatic ability correlates with increased type IV collagenase activity (Nakajima et al., 1987) and decreased TIMP expression (Korcak et al., 1991). Most importantly, a novel member of the MMP family, designated stromelysin 3 (MMP-11), was recently isolated on the basis of its elevated expression in stromal cells surrounding invasive human breast carcinomas (Basset et al., 1990; Wolff et al., 1993). Moreover, Zucker et al. (1993) have found evidence for elevated levels of MMP-9 circulating in plasma of breast cancer patients. Other data from in situ hybridisation and immunohistochemical studies have implicated MMP-2 in the invasiveness of mammary carcinomas (Monteagudo et al., 1990; Poulsom et al., 1993; Clavel et al., 1992).

While powerful insights can be provided by studies of transformed or tumour-derived cells in vitro, it is necessary to examine fresh human tumours to understand the contribution of MMPs and TIMPs and other ECM remodelling proteins to malignant behaviour. We have therefore carried out systematic studies of the expression at the RNA level of MMPs and TIMPs in human sporadic colorectal neoplasia (HSCN) (Urbanski et al., 1993), lung carcinomas (Urbanski et al., 1992) and malignant non-Hodgkin's lymphomas (NHL) (Kossakowska et al., 1991, 1992). In one of these studies we showed, using statistical analysis, that the level of expression of MMP-9 transcripts correlated well with the clinical aggressiveness of immunoblastic NHL, strongly implicating this enzyme in dissemination of these tumours (Kossakowska et al., 1992).

In the present work, we turn our attention to human breast cancer, carrying out a systematic assessment of the expression at the RNA level of five members of the MMP family, namely interstitial collagenase (MMP-1), MMP-2, matrilysin (MMP-7), MMP-9 and MMP-11, along with...
TIMP-1 and TIMP-2. Expression patterns are assessed statistically for relationships with the morphological characteristics of 37 breast lesions.

Materials and methods

Tissue collection and preparation

All tissue was received fresh from the operating room. Analysed cases included 17 carcinomas (12 infiltrating duct, two lobular, two medullary and one mucinous), two intraductal carcinomas, one large intraductal papilloma, five fibroadenomas, three cases of benign non-proliferative breast disease, seven samples of breast tissue adjacent to either fibroadenoma (one case) or carcinoma (three showing benign proliferative breast disease without atypia and three showing non-proliferative breast disease) and two metastatic breast carcinomas (one to skin and one to axillary lymph node). The tissue submitted for RNA extraction was embedded in OCT. Cryostat sections taken adjacent to the analysed tissue and stained with haematoxylin and eosin precisely identified the analysed lesions.

Histological assessment

All breast pathology was assessed independently by two pathologists. Diagnoses and terminology were those in current use in the surgical pathology division of the Department of Pathology, University of Calgary. Sections taken adjacent to the tissue used for RNA extraction were assessed for degree of desmoplasia in epithelial neoplastic lesions and for stromal proliferation in fibroadenomas and non-neoplastic breast (none, –; mild, +; moderate, ++; severe, ++++) in order to assess stromal activity and its correlation with MMP-11 expression.

RNA extraction and Northern blot analysis

Total cellular RNA from tissue samples was extracted by the acid–guanidinium thiocyanate–phenol–chloroform (AGPC) method (Chomczynski and Sacchi, 1987). Frozen tissue was homogenised in guanidinium isothiocyanate solution and subsequently extracted with acid phenol–chloroform. The precipitated RNA was washed, ethanol-precipitated, resuspended in 0.5% sodium dodecyl sulphate (SDS) solution and stored at −70°C. Northern blot analysis was performed according to previously described methodology (Sambrook et al., 1989; Urbanski et al., 1992). Total cellular RNA (10 μg) was applied to each well and electrophoresed through formaldehyde-containing agarose gels. The products of electrophoresis were transferred onto Duralon membranes and fixed with a UV cross-linker. Six identical blots containing the same amount of each sample were prepared simultaneously and hybridised with 32P nick-translated DNA probes in standard 50% formamide-containing hybridisation buffer (Sambrook et al., 1989). The blots were washed, autoradiographed and subsequently stripped of the labelled probe for reuse by incubation with prehybridisation solution for 15 min at 80°C. Autoradiographic signals were quantified using a Pharmacia UltraScan XL laser densitometer, and all signals were within the linear response range of the X-ray film and the detection system. Each blot carried a control sample of RNA derived from Hs68 human foreskin fibroblasts that had been stimulated for 8 h with 10−7 M phorbol myristoyl-13 acetate. Signals were normalised between different blots based on the signals from the Hs68 RNA control. Signals were expressed as strong (+ + +), corresponding to normalised densitometric measurements greater than 3.0 absorbance units, moderate (+ +, densitometry measurements between 1.0 and 2.9) or weak (+, densitometry measurements less than 1.0). Samples were loaded on the gels in coded fashion and the blots were interpreted without knowledge of the tissue type.

DNA probes

The human TIMP-1, TIMP-2, MMP-1, MMP-2, MMP-7, MMP-9 and MMP-11 DNA probes have been described previously (Urbanski et al., 1992, 1993; Kossakowska et al., 1991, 1992, 1993).

Statistical analysis

The relationship between the malignant phenotype and MMP and TIMP mRNA values was assessed using the chi-square statistic (Norusis, 1988). The potential of MMP and TIMP mRNA measurements to discriminate between tumours with and without the malignant phenotype was assessed by calculating their sensitivity, specificity and overall predictive accuracy (Rimm et al., 1980). Sensitivity measures the proportion of those tumours with malignant phenotype that are identified by the expression of the mRNA transcripts. Specificity measures the proportion of tumours without the malignant phenotype which are correctly identified by their absence of mRNA transcripts. Predictive accuracy is the proportion of all tumours correctly identified as either malignant or benign by whether they express the mRNA transcripts.

The specimens were further subdivided into seven categories of neoplasia. Kendall’s tau was the statistic (Norusis, 1988; Rimm et al., 1980) used to test the association between mRNA expression and degree of neoplasia.

Results

The human breast tissue specimens ranged along the continuum of neoplasia from morphologically benign (n=16) and preinvasive epithelial lesions (n=2) through malignant (n=17) to metastatic (n=2). Figure 1 shows representative Northern blot results for 13 samples of human breast. Levels of expression of MMP and TIMP transcripts were assessed as strong (+ + +), moderate (+ +), weak (+) or absent (−) by visual comparison with signals obtained from human fibroblast RNA that was included as a control on each blot. The full results of this analysis are shown in Tables I and II.

It can be seen that the genes were expressed to varying

Figure 1 Representative results of the Northern blot analysis of non-neoplastic and neoplastic breast lesions probed for MMP-9, MMP-11, TIMP-1 and TIMP-2. F, fibroblasts; FA, fibroblast; I, infiltrating; ID, infiltrating duct; IC, intraductal; N, normal; N+, non-neoplastic breast adjacent to fibroadenoma; N−, non-neoplastic breast adjacent to carcinoma; MET, metastasis.
Malignant disease had a higher prevalence than benign, and only one-quarter had detectable MMP-1, MMP-2, MMP-7 and TIMP-9 transcripts. Secondly, mRNAs encoding MMP-1, MMP-2, MMP-7 and TIMP-2 appeared unrelated to the malignant phenotype, since they were expressed in both benign and malignant lesions.

Table I: Summary of expression of MMP and TIMP RNAs in malignant breast tissues

|         | MMP-1 | MMP-2 | MMP-7 | MMP-9 | MMP-11 Desmoplasia | TIMP-1 | TIMP-2 | L | S |
|---------|-------|-------|-------|-------|---------------------|--------|--------|---|---|
| Ductal  | 1     | NA    | NA    | NA    | –                   | NA     | NA     | ++| NA|
|         | 2     | –     | –     | –     | –                   | +      | +      | ++| + |
|         | 3     | +     | +     | –     | –                   | +      | +      | ++| + |
|         | 4     | –     | +     | +     | –                   | +      | +      | ++| + |
|         | 5     | +     | –     | +     | –                   | +      | +      | ++| + |
|         | 6     | –     | –     | –     | +                   | +      | +      | ++| + |
|         | 7     | –     | –     | –     | –                   | +      | +      | ++| + |
|         | 8     | NA    | NA    | NA    | NA                  | +      | +      | ++| + |
|         | 9     | –     | –     | –     | –                   | 0      | NA     | NA|
|         | 10    | –     | –     | –     | –                   | +      | +      | ++| + |
|         | 11    | –     | –     | –     | +                   | +      | +      | ++| + |
|         | 12    | NA    | NA    | NA    | +                   | +      | +      | ++| + |
| Lobular | 1     | NA    | NA    | NA    | NA                  | +      | +      | ++| + |
|         | 2     | NA    | NA    | –     | +                   | +      | +      | ++| + |
| In situ | 1     | –     | +     | +     | –                   | +      | +      | ++| + |
|         | 2     | –     | –     | –     | –                   | 0      | NA     | NA|
| Miscellaneous | 1 | –       | –   | +   | +                   | +      | +      | ++| + |
|         | 2     | –     | –     | +     | +                   | +      | +      | ++| + |
| Metastasis | 1 | +     | –     | +     | –                   | 0      | NA     | NA|
|         | 2     | +     | –     | –     | –                   | 0      | NA     | NA|

Table II: Summary of expression of MMP and TIMP RNAs in benign breast tissues

|         | MMP-1 | MMP-2 | MMP-7 | MMP-9 | MMP-11 Stromal proliferation | TIMP-1 | TIMP-2 | L | S |
|---------|-------|-------|-------|-------|------------------------------|--------|--------|---|---|
| Adjacent to FA | 1 | +     | –     | –     | –                            | –      | –      | + | + |
| Benign non-proliferative breast disease | 1 | –     | –     | NA    | NA                           | –      | –      | + | – |
|         | 2     | +     | +     | +     | –                            | –      | –      | + | + |
|         | 3     | +     | +     | –     | –                            | +      | +      | + | + |
| Adjacent to carcinoma | 1 | +     | –     | –     | –                            | +      | +      | + | + |
|         | 2     | +     | +     | –     | –                            | +      | +      | + | + |
|         | 3     | NA    | NA    | NA    | NA                           | –      | –      | + | + |
|         | 4     | –     | –     | –     | –                            | –      | –      | NA| NA|
|         | 5     | –     | –     | –     | –                            | –      | –      | + | + |
|         | 6     | –     | –     | –     | +                            | NA     | NA     | NA|
| Fibroadenomas | 1 | +     | +     | +     | –                            | +      | +      | + | + |
|         | 2     | –     | –     | –     | –                            | –      | –      | + | + |
|         | 3     | NA    | NA    | NA    | NA                           | +      | +      | + | + |
|         | 4     | –     | –     | –     | –                            | +      | +      | + | + |
|         | 5     | –     | –     | –     | –                            | +      | +      | + | + |
| Papilloma with moderate dysplasia | 1 | –     | –     | –     | –                            | –      | –      | + | NA|

NA, not available.

degrees: TIMP-1 and TIMP-2 transcripts were present in almost all specimens, MMP-11 and MMP-9 in about one-half, and only one-quarter had detectable MMP-1, MMP-2 and MMP-7 RNAs. Secondly, mRNAs encoding MMP-1, MMP-2, MMP-7 and TIMP-2 appeared unrelated to the malignant phenotype, since they were expressed in both benign and malignant lesions.

The third and most important point from this study comes from statistical analysis of the expression of MMP-11, MMP-9 and TIMP-1. In the 31 specimens for which results were available for all three, MMP-11 distinguished malignant disease with the greatest predictive accuracy (84% vs 77% and 71% respectively). Fourteen of the 18 malignant specimens and 12 of the 13 benign were correctly identified by the presence or absence, respectively, of transcripts for MMP-11. This yielded a predictive accuracy of 84% [(14+12)/(18+13)]. However, as shown in Figure 2, MMP-9 was the most strongly related to phenotype among the benign (n=13), malignant (n=16) and metastatic (n=2) categories (P=0.0004) and along a seven category continuum (P=0.0003). This was likely because MMP-11 transcripts were not expressed in the two specimens of metastatic disease (Tables I and II). Detection of either MMP-11 or MMP-9 increased the predictive accuracy to 90% with 94% sensitivity and 85% specificity. Eleven of the 13 benign specimens expressed neither MMP-11 nor MMP-9, while 17 of the 18 malignant specimens expressed either or both of MMP-11 and MMP-9 [(11+17)/(13+18)=90% predictive accuracy] (Table III). Specimens that deviated from the pattern of non-
expression of MMP-11 and MMP-9 in benign tissues and expression in malignant tissues probably represent transitional stages on the progression continuum. Thus, the one malignant primary tumour that expressed neither MMP-11 or MMP-9 was carcinoma in situ. Two of the 13 benign tumours expressed either MMP-11 or MMP-9; these were a fibroadenoma expressing MMP-9 and benign tissue adjacent to a carcinoma and expressing MMP-11.

We also observed that some MMP and TIMP genes showed differential expression at the periphery of the tumours compared with samples from the centre (Table IV). Transcripts for MMP-11 and TIMP-1 were elevated in the periphery of two of the five tumours analysed in this way, though only one case showed elevation of both transcripts. Also, in one case we found increased MMP-7 RNA levels in the periphery. Such patterns were not seen for MMP-1, MMP-2, MMP-9 or TIMP-2. Histological examination of sections adjacent to areas of tumour taken for RNA extraction were assessed for degree of desmoplasia as an indicator of stromal activity. As can be seen from Tables I and II, degree of desmoplasia was fairly well correlated with MMP-11 expression.

The comparison of the expression patterns of MMPs across four tumour systems is summarised in Figure 1. The most striking pattern is the association of MMP-11 expression with all types of carcinoma that we have analysed, and the absence of MMP-11 transcripts in NHL. Regardless of the type of malignancy, TIMPs 1 and 2 were expressed by over 80% of malignant lesions and MMP-9 by at least 50%. The expression of MMP-2, MMP-1 and MMP-7 showed the greatest variability across types of malignancy. But consistent features of all tumour types are the expression of MMP-9 or overexpression of TIMP-1.

Discussion

The natural history of malignant tumours involves multiple episodes of ECM breakdown mediated by secreted ECM-degrading proteinases (Liotta et al., 1991; Mignatti and Rifkin, 1993). A comprehensive picture of the molecular machinery that can participate in matrix turnover has emerged from in vitro studies of degradative enzymes and their natural inhibitors (Mignatti and Rifkin, 1993; Stetler-Stevenson et al., 1993; Alexander and Werb, 1991). However, most of the current literature that deals with the roles of proteinases in tumour invasion and metastasis has come from studies of transformed or tumour-derived cell lines in culture. To fully appreciate their biological significance, it is necessary to study factors that have been implicated in ECM destruction in the natural context of the complex tumour-host interactions in freshly resected human tumours. The members of the MMP and TIMP gene families are the focus of this work.

![Figure 2](image-url)  
**Figure 2** MMP mRNA expression (percentage of samples) in breast tissue by neoplastic phenotype. The breast tissue specimens were arranged in a seven category continuum of increasing malignancy (from left to right). The figure shows the percentage of samples in each category that expressed detectable transcripts of MMP-9 and MMP-11. MMP-9 was most strongly related to degree of neoplasia, whereas MMP-11 most clearly distinguished the neoplastic phenotype from the benign. □, MMP-9; ■, MMP-11.

| Tissue type (n)   | MMP-9 only | Both MMP-9 and MMP-11 | MMP-11 only | Total (MMP-11 and MMP-9) |
|------------------|------------|-----------------------|-------------|--------------------------|
| Benign (13)      | 1          | 0                     | 1           | 2                        |
| Malignant primary (16) | 1          | 9                      | 5           | 15                       |
| Metastatic (2)   | 2          | 0                     | 0           | 2                        |

Table III Frequency of mRNA transcript expression for MMP-9 and MMP-11 by tissue type (n = 31)

| Location | MMP-1 | MMP-2 | MMP-7 | MMP-9 | MMP-11 | TIMP-1 | L | S |
|----------|-------|-------|-------|-------|--------|--------|---|---|
| Centre   | —     | —     | +     | +     | +      | ++     | + | +|
| Periphery| —     | —     | NA    | NA    | +      | +      | + | +|
| Centre   | —     | —     | —     | —     | +      | +      | + | +|
| Periphery| NA    | NA    | NA    | NA    | +      | +      | + | +|
| Centre   | NA    | NA    | —     | +     | NA     | ++     | + | +|
| Periphery| —     | +     | +     | +     | +      | +      | + | +|
| Centre   | —     | —     | —     | —     | +      | +      | + | +|
| Periphery| —     | —     | —     | +     | +      | +      | + | +|

Table IV Expression patterns of mRNA transcripts of MMPs and TIMPs in the central and peripheral samples from breast carcinomas (n = 5)

NA, not available.
This study reports a thorough analysis of expression at the RNA level of MMP-1, -2, -7, -9 and -11 and TIMP-1 and -2 in a spectrum of breast tissues ranging from normal to overtly malignant. Comparative assessment of expression levels alongside pathological characteristics of the lesions allows us to identify which of these genes are likely important correlates of the malignant phenotype. Further, to explore the generality of the associations at which we arrive, we have contrasted the data obtained from human breast carcinomas with those from other tumours of both epithelial (lung and colon carcinomas) and non-epithelial origin (non-Hodgkin’s lymphomas; NHL).

A number of caveats to the present study must be acknowledged. First, we recognise that other enzymes of the aspartyl, cysteine and serine proteinase families and their specific inhibitors are also likely to be significant participants in ECM remodelling during cancer progression. Second, we have limited our analyses to mRNA detection since it provides the simplest and most reliable method of quantification of expression for the functions involved. Since MMPs and TIMPs are for the most part secreted freely diffusible proteins, both quantification at the protein level and immunohistochemical localisation present significant problems (Clavel et al., 1992; Hemby et al., 1986). Although it is possible that mRNA levels for MMPs and TIMPs may not always reflect amounts of protein products secreted, as has been reported for MMP-1 in a rat epithelial cell line (Whitelock et al., 1993), RNA analyses provide a generally reliable means of assessment of expression. Third, it is clear that the activities of MMPs are controlled at multiple levels including expression, activation and via interaction with TIMPs. Thus expression and secretion of protein product may not, in all cases, be the chief arbiters of functionality (Azzam et al., 1993). For instance, activation of pro-72 kDa gelatinase A (MMP-2) involves binding of the pro-MMP2/TIMP-2 complex to a cell-surface receptor followed by proteolytic cleavage by a transmembrane metallocproteinase (MMP-MMP-1), thereby restricting MMP-2 activity to the pericellular environment of cells carrying appropriate activation machinery (Emonard et al., 1992; Monsky et al., 1993; Kleiner and Steitler-Stevenson, 1993; Sato et al., 1994; Vassali and Pepper, 1994). Finally, there is the possibility that the expression of genes that are important in ECM destruction may be spatially restricted to the invasion front of the tumour (Stetler-Stevenson et al., 1993). This may complicate quantification owing to variable tissue sampling and because the expressing cells may be vastly outnumbered by non-expressing cells. Because of these concerns, it is important to emphasise that while our data identify two MMP genes by virtue of the association of their expression with malignancy, we cannot exclude the possibility that others may also play significant roles.

From our studies it is apparent that expression of MMP-11 and MMP-9 is likely associated with malignant phenotype in human breast carcinomas. Using detection of either MMP-11 or MMP-9 transcripts as a criterion, we were able to distinguish malignant from normal tissue with a predictive accuracy of 90% with 94% sensitivity and 85% specificity. Moreover, these genes are consistently found to be expressed in carcinomas in three different tissues of origin. As in HSCN (Urbanski et al., 1993), preinvasive lesions of human breast are intermediate in their expression patterns of MMPs and TIMPs between normal and frankly malignant tissue.

However, the presence of MMP-11 and MMP-9 transcripts is not simply related to epithelial proliferation, as one case of large papilloma (breast) with atypia showed neither MMP-11 nor MMP-9 expression. Similar observations were made previously in HSCN where many colonie adenomas did not show altered MMP expression (Urbanski et al., 1993). Thus, the processes that control cell proliferation and invasion are to some degree, independent. This conclusion differs from that of a recent in vitro study of human A431 epidermoid carcinoma cells, in which MMP-9 expression was observed in sparse, proliferating cultures but not in dense, contact-inhibited ones (Xie et al., 1994). The results with cultured A431 cells may reflect the behaviour of malignant cells in a highly advanced tumour in vivo, in which cells at the invasion front are more metabolically active than cells in the centre of the tumour, and thus also express MMP-9 to the highest level. However, our data clearly show that in benign proliferative breast lesions in vivo, MMP-9 expression and epithelial proliferation are uncoupled. We suggest that altered expression of both MMP-9 and MMP-11 characterises epithelial lesions committed to malignant transformation and may reflect underlying alterations in regulatory tumour-suppressor genes or proto-oncogenes that prepare the cells for imminent invasion.

Several reports localised MMP-11 to the invasive front in breast carcinomas where it is expressed not by tumour cells but by stromal cells (Basset et al., 1990; Wolf et al., 1993). Similar findings have been reported for basal cell carcinoma (Wagner et al., 1992) and head and neck squamous cell carcinomas (Muller et al., 1993). Support for the notion that tumour–stromal interactions are important for setting up MMP-11 expression comes from our observations that expression of MMP-11 transcripts was highest in samples taken from the tumour periphery in two out of five cases, and that MMP-11 expression was also detectable in tissue immediately adjacent to a carcinoma. Moreover, degree of desmoplasia correlated well with MMP-11 expression (Tables I and II). It is therefore of considerable interest that we find that MMP-11 expression is restricted to carcinomas and preinvasive lesions but is undetectable in 42 cases of NHL that we have also analysed. As progression of NHL is not associated with destruction of basement membrane it is likely that MMP-11 expression is intimately linked with events that lead to the invasion of tumours of epithelial origin through this biological barrier. However, MMP-11 does not itself display detectable proteolytic activity towards ECM components (G Murphy et al., 1993), though it can cleave and inactivate some of these proteinases (Pei et al., 1994). Thus, it is possible that it may perform a regulatory role by controlling the activities of other proteinases. Wolf et al. (1993) have noted similarities in the expression patterns of MMP-11 and urokinase plasminogen activator in breast carcinomas, suggesting that both enzymes may cooperate during cancer progression.

Although we found MMP-11 expression in 84% of primary breast carcinomas, we failed to detect it in the skin...
and nodal metastases. These findings do not agree with those of Wolf et al. (1993) who observed MMP-11 in 11 out of 13 (85%) lymph node metastases of breast carcinoma. We note however, that these authors report lower levels of MMP-11 detection in metastases to other locations (67% to bone and 50% to other organs). If MMP-11 is indeed linked to invasive spread of tumour cells away from primary carcinomas we think it reasonable, particularly in the light of our inability to detect MMP-11 in NHL, that the different stromal environment of nodal metastases will probably be reflected in the elaboration of different ECM degradative functions, which may also vary from site to site. Clearly, the role of MMP-11 in the growth of metastases is a topic for further investigation, using a larger series of tumour specimens. MMP-9 is the other consistent marker of malignancy that is expressed at elevated levels, not only in breast cancer but also in the other three types of malignancy that we have compared. Previously, we showed that MMP-9 expression in a subset of malignant high-grade NHL correlated with clinical aggressiveness as judged by poorer survival (Kossakowska et al., 1992). Our findings are supported by the demonstration of elevated levels of MMP-9 protein in the plasma of patients with colon and breast cancer (Zucker et al., 1993). In a zymographic analysis of bladder cancer specimens, Davies et al. (1993) have correlated increased amounts of MMP-9 and active MMP-2 with tumour grade and invasion. The links between MMP-9 and cellular invasion are now very strong and include work with oncogenically transformed rat fibroblasts (Bernhard et al., 1992), oral squamous cell carcinoma-derived cell lines (Juarez et al., 1993), U937 monoblastoid cells (Watanabe et al., 1993) as well as non-neoplastic cell types capable of invasion, such as preimplantation mouse and human blastocysts (Behrendsen et al., 1992, Librach et al., 1991). Unlike the predominantly stromal expression of MMP-11 and MMP-2 transcripts (Basset et al., 1990, 1994; Wolf et al., 1993; Polette et al., 1994; Soini et al., 1994) MMP-9 RNAs are localised primarily in epithelial cells of breast carcinoma (Soini et al., 1994). This indicates that the increased expression of MMP-9 and MMP-11 seen in malignant tumours involves distinct mechanisms.

Although our data point to an association of both MMP-9 and MMP-11 with the malignant phenotype in breast cancer, the situation with MMP-1, MMP-2 and MMP-7 is less clear. Transcripts for the last genes were not consistently found in malignant tumours and were frequently detected in benign lesions or normal tissues adjacent to cancer. However, as we pointed out earlier, the absence of association between gene expression and malignancy does not indicate non-involvement of the products of these genes in the malignant process. In fact, our data indicate an association of MMP-1 with the malignant phenotype in colon and lung carcinomas but not in breast carcinoma or NHL. Its involvement may thus depend on the anatomical location of the tumour. In colonic neoplasia, MMP-1 transcripts are localised to eosinophils, fibroblasts and endothelial cells (Gray et al., 1993), again demonstrating the importance of tumour–host interactions.

Our data on TIMP-1 and TIMP-2 reveal that transcripts for both are present in most tissues whether benign or malignant. Consistently we have seen an elevation of TIMP-1 expression in all types of malignant lesions studied, and we previously showed localisation of TIMP-1 transcripts to stromal cells in NHL (Kossakowska et al., 1991) and lung carcinomas (Urbanski et al., 1992). TIMP-1 transcription is highly inducible in vitro by diverse cytokines whereas TIMP-2 is expressed in a more constitutive fashion (Leco et al., 1992; Stelter-Stevenson et al., 1990). So tumour–host interactions would again appear to be important for the selective up-regulation of TIMP-1 expression. We do not yet know the full significance of increased TIMP-1 production on tumour pathophysiology. It could be interpreted as an attempt by the host to limit the ECM remodelling that is elicited by the presence of the tumour cells. However, as both TIMP-1 and TIMP-2 have growth modulatory actions, in addition to their roles as MMP inhibitors (Bertaux et al., 1991; Hayakawa et al., 1992; AN Murphy, et al., 1993; Nemeth and Goolsby, 1993), it is possible that increased TIMP-1 production at the tumour–host interface may in certain situations stimulate tumour growth rather than having the expected effect of limiting cellular invasion.

In summary, the observations presented here confirm a strong association between expression of MMP-11 mRNA and malignant phenotype in human breast epithelial neoplasms, and for the first time, indicate that MMP-9 is also consistently overexpressed in these carcinomas. This subset of MMPs is further implicated as a factor common to the malignant process in carcinomas of other organs, but we find no evidence for involvement of MMP-11 in NHL, which suggests that this proteinase characterises progression of carcinomas. The proteinases that appear to be necessary for invasion can be expressed in preinvasive states, indicating their potential applications as prognostic indices and as targets for therapeutic intervention. Further research into the molecular mechanisms underlying the expression of MMP-11 and MMP-9 may permit manipulation of the neoplastic process to inhibit or prevent acquisition of the invasive phenotype.

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