Cytokine Responses in the Blood and Pleural Fluid of Pulmonary Tuberculosis Patients with and without HIV-1 Co-infection

Mahendra K Bhopale1*, Harshada Shah1, Babu Lal Bamboria2, Arti Julka3, Imran Patel4, Vijay K Mahadik1, Manju Raj Purohit5,6

1Department of Microbiology, R.D. Gardi Medical College, Agar Road, MP, India
2Department of Medicine, R.D. Gardi Medical College, Agar Road, MP, India
3Department of Pulmonary Medicine, R.D. Gardi Medical College, Agar Road, MP, India
4Central Laboratory, R.D. Gardi Medical College, Agar Road, Ujjain 456006 MP, India
5Department of Pathology, R.D. Gardi Medical College, Agar Road, MP, India
6Public Health Sciences, Karolinska Institute, Stockholm, Sweden

*Corresponding Author: Dr. Mahendra K Bhopale, Department of Microbiology, R.D. Gardi Medical College, Agar Road, Ujjain 456006 MP, India, E-mail: mbhopale@yahoo.com

Received: 19 July 2019; Accepted: 07 August 2019; Published: 14 August 2019

Abstract

Tuberculosis (TB) is an opportunistic infectious disease with more severe forms in HIV-1 infected patients. The aim of the present study is to investigate the role of IFN-γ in the other cytokines belonging to Th1, Th17, Th22, and Th3 groups and CD4+ cell counts in the HIV-1 infection in patients co-infected with Mycobacterium tuberculosis. Clinically diagnosed patients of HIV-1, TB and HIV-1 co-infected with TB (HIV-1+TB) groups were selected to test IFN-γ, IL-17, IL-22, TGF-β, and also CD4 counts in the blood for the study. Our results showed that IFN-γ and IL-17 cytokines were higher in HIV-1 and TB patients in serum when compared with healthy normal subjects, but there was an insignificant difference when compared with those in HIV-1+TB with TB patients’ samples. Pleural fluid samples of the HIV-1+TB patients showed significantly higher in IL-22 and IFN-γ cytokines than in TB patients, whereas IL-17 showed insignificant differences. CD4+ cells were counted in the blood of HIV-1, TB, and HIV-1+TB patients, however, the results showed that the counts were significantly lower than in the healthy normal group. There was a significantly lower CD4+ count in HIV-1+TB co-infected patients compared to TB patients, but not with HIV-1 patients. The present study suggests that IFN-γ and IL-17 play a significant role individually in HIV-1 and tuberculosis infected patients and IL-22 in pleural fluid in tuberculosis, which differs from those in HIV-1 co-infected tuberculosis patients due to the severely affected immune system.
Introduction

Human immunodeficiency virus (HIV) infection remains the most common risk factor for the development of active tuberculosis (TB). Co-infection with Mycobacterium tuberculosis in HIV-1 patients is a leading cause of death due to deterioration of immunological functions [1, 2]. HIV-1 activity increased in patients infected with pleural TB demonstrated in pleural fluid and pleural mononuclear cells [3]. IFN-γ as biomarker has been used in the clinical assessment in TB patients. Clinical manifestations of TB co-infected with HIV-1 patients are usually severe, with diffuse pulmonary involvement in the pleural fluid. IL-17, IL-22 and TGF-β cytokines play immunological roles, particularly in mycobacterial active tuberculosis and in granuloma formation, whereas the regulation of IL-17A and IL-22 requires TGF-β and IL-23 and IL-1β [4-5]. Between TB and HIV-1, both infections have profound effects on the immune system and the ability to disarm the host’s immune responses. HIV-1 co-infection with M. tuberculosis has the risk of reactivation of the latent stage of TB and exacerbates HIV-1 infection. CD4+ T cell counts of HIV-1 infected with pulmonary TB (PTB) are higher than other opportunistic infections due to T cell depletion and dysfunction [6], due to diminishing M. tuberculosis antigen-specific IL-17 and IL-22 production [7].

A laboratory study was conducted on clinical-microbiologically diagnosed patient samples of HIV-1, TB and HIV-1+TB patients to investigate cytokine responses of IFN-γ, IL-17, IL-22, TGF-β in serum and pleural fluid of pulmonary tuberculosis patients with and without HIV-1 co-infection.

Materials and Methods

The clinical study was conducted at the Microbiology Department, R.D. Gardi Medical College, Ujjain, (MP) India from 2014-2016. A Government approved guidelines adopted by an ethical committee established in R. D. Gardi Medical College, and the Institutional Review Board-approved written consent was obtained from all the participants. Patients on anti-tuberculous, anti retroviral therapy or corticosteroid treatment were not included in this study. A clinical study of patients was applied for the statistical analysis using an Inter Quartile range (IQR) for the difference between 3rd and 1st quartile, and the median for IQR measures. The diagnosis of HIV status of the patient was confirmed by Microlisa HIV kit and tuberculosis was confirmed by sputum or bronchoalveolar lavage positive by Ziehl Neelsen stain and mycobacterial culture. Serum and pleural fluid samples of HIV-1+TB and TB patients were collected and performed enzyme-linked immunosorbant assay (ELISA) kits, purchased from Ray Biotech, USA. The ELISA plate was read on ELISA reader at 450 nm by LisaScan EM, Erba Mannheim.
Pleural fluid samples of HIV-1+ and TB co-infected patients were tested for ADA (Adenosine Deaminase Activity). The titer in aspirated pleural fluid was measured by colorimetric method using ADA-MTB kit (Tulip diagnostics (P) Ltd. India). The total ADA activity was measured by spectrophotometer at 570-630 nm. CD4+ cells in the blood samples were counted in a BD FACSCount System of (Becton and Dickinson) in the Central Laboratory of R.D. Gardi Medical College. CD4 reagents were used specifically for enumerating the absolute cell counts of CD4+ T lymphocytes. The statistical analysis was applied to compare the cytokine levels between the two groups of the two-tailed p value of <0.05 for a nonparametric t-test test by GraphPad software (La Jolla, CA). The statistical t’ test (unpaired samples) was applied for the Linear regression (R2) analysis on p<0.05 of straight-line relationship between the individual Log2 value of an each cytokine and between two variable y against that of the independent variable x was determined by GraphPad Software and QuickCalcs Linear regression calculator.

3. Results

All clinically examined patients suffering from HIV-1, with and without TB, exhibited signs and symptoms like cough, breathlessness, chest pain and fever. Both groups showed pleural effusion in radiological examination (Table 1). We further conducted a study on cytokine assay and CD4+ counts on the blood samples from four groups: HIV-1 infected patients (n=5), TB patients (n=8), HIV-1+TB co-infected patients (n=7), and Healthy normal subjects (n=10). CD4+ cells count in the blood of HIV-1 (p=0.0001), TB (p=0.0021), HIV-1+TB (p=0.0001) show a significant difference when compared to the healthy normal group; and HIV-1+TB co-infected and TB patients (p=0.0001), but insignificant differences between HIV-1+TB and HIV-1 (p=0.8147) (Table 1, Figure 1).

Table 1: Study participant characteristics and clinical and laboratory findings.

| Patients’ characteristics | HIV-1+TB | TB |
|---------------------------|---------|----|
| Sex                       | Male    | Female | Male | Female |
| Participant (n)           | 7       | 0      | 10   | 5      |
| Median age (QIR) in years | 32 (22-43) | -     | 37 (30-45) | 35 (30-55) |
| Clinical symptoms         | Cough, Breathlessness, Chest pain, Weight Loss | Cough, Breathlessness, Chest pain, Weight Loss |

Radiological Studies

| Number of PIEF shown in Lung side | Left | Right | Left | Right | Left | Right | Left | Right |
|-----------------------------------|------|-------|------|-------|------|-------|------|-------|
|                                   | 3    | 4     | -    | -     | 3    | 8     | 2    | 3     |

Laboratory findings

| Pleural fluid appearance and sample in numbers | Turbid (1), Clear (2) Hazy (1) Yellowish (1) Turbid (5) | Turbid (5), Clear (4), Hazy (6) Yellowish (1) |
|-----------------------------------------------|--------------------------------------------------------|-----------------------------------------------|
| Protein (range) mg/ml                         | 4.5-7.00                                                | 3.9-7.00                                      |
| ADA (IQR)                                     | 14 (79-93)                                              | 32.5 (62-94.5)                               |
| CD4+ (IQR) in cells/mm³                      | 151 (54-205)                                            | 151.5 (340-459.5)                            |
FACS CD4⁺ cells in study groups

| No. of Samples | Patients   | Mean ± SE | Control       | No. of Samples | Mean ± SE | Statistical p value |
|----------------|------------|-----------|---------------|----------------|-----------|---------------------|
| 5              | HIV-1      | 116.00 ± 50.58 | Normal Healthy | 10             | 1016.40 ± 143.70 | p=0.0001**          |
| 8              | TB         | 412.00 ± 35.41 | Normal Healthy | 10             | 1016.40 ± 143.70 | p=0.0021**          |
| 7              | HIV-1+TB   | 130.71 ± 37.09 | Normal Healthy | 10             | 1016.40 ± 143.70 | p=0.0001**          |
| 7              | HIV-1+TB   | 130.71 ± 37.09 | HIV-1         | 5              | 116.00 ± 50.58  | p=0.8147            |
| 7              | HIV-1+TB   | 130.71 ± 37.09 | TB            | 8              | 412.00 ± 35.41  | p=0.0001**          |

Abbreviations: ADA-Adenosine Deaminase test for tuberculosis optical cut-off points was determined at 50 U/L measured by spectrophotometer at 570-630 nm; CD4⁺ counts was determined in cells/mm³ volume of blood sample; IQR-Inter Quartile range; FACS-Fluorescence-activated cell sorting; PIEF-Pleural effusion (fluid in the lungs); Statistical significance =**p=0.0001 (Highly significant).

Figure 1: CD4⁺ cells count in HIV-1, TB, HIV-1+TB patients shows a significant difference (*) as compared to healthy normal group, whereas insignificant differences between HIV-1+TB co-infected and HIV-1 patients.

The cytokine profiles of IL-17 (p=0.0032), IL-22 (p=0.0267) and IFN-γ (p=0.0001) in the HIV-1 infected patients in the serum showed significantly higher, whereas IL-17 (p=0.0079) and IFN-γ (p=0.0007) were significantly higher in TB patients when compared with healthy, normal subjects (Table 2, Figure 2). However, the serum levels of IL-17 (p=0.9253), IL-22 (p=0.8253), and TGF-β (p=0.7810) and IFN-γ (p=0.5066) were not significantly different in HIV+TB co-infected patients compared to TB; and IL-17 (p=0.5870), IL-22 (p=0.7872), and TGF-β (p=0.7360) and IFN-γ (p=0.8102) compared to HIV-1 patients. However, both IL-22 (p=0.0290) and IFN-γ (p=0.0484) were significantly higher in pleural fluid of TB than HIV-1+TB co-infected patients (Table 2, Figure 3). Regression (r) analysis and Linear regression (R²) analysis of the pleural fluid samples value in pg/ml or ng/ml were converted into Log² and calculated R² value to measure how close sample data to the regression line when compared statistically.
two groups. A comparative regression (r) analysis and Linear regression (R2) analysis in the pleural and serum samples of their mean values of TB and HIV-1+TB is illustrated (Figure 4).

Table 2: Comparison of cytokine levels among HIV-1+TB patients and normal healthy participants.

| ELISA test on serum samples | HIV-1 | Normal Healthy |
|-----------------------------|-------|----------------|
| No. of Sample | Cytokine | Mean ± SE | No. of Sample | Mean ± SE | p value |
| 5 | IL-17 | 387.00 ± 111.55 pg/ml | 10 | 95.30 ± 19.95 pg/ml | p=0.0032* |
| 5 | IL-22 | 55.58 ± 29.96 pg/ml | 10 | 3.38 ± 3.29 pg/ml | p=0.0267* |
| 5 | TGF-β | 1.620 ± 0.28 ng/ml | 10 | 1.625 ± 0.28 ng/ml | p=0.9914 |
| 5 | IFN-γ | 238.00 ± 44.62 pg/ml | 10 | 5.35 ± 1.20 pg/ml | p=0.0001** |

TB

| HIV-1+TB | Normal Healthy |
|---------|----------------|
| 8 | IL-17 | 317.13 ± 78.39 pg/ml | 10 | 95.30 ± 19.95 pg/ml | p=0.0079* |
| 8 | IL-22 | 35.082 ± 30.77 pg/ml | 10 | 3.385 ± 3.29 pg/ml | p=0.2670 |
| 8 | TGF-β | 1.580 ± 0.230 ng/ml | 10 | 1.625 ± 0.28 ng/ml | p=0.9074 |
| 8 | IFN-γ | 283.625 ± 75.124 pg/ml | 10 | 5.35 ± 1.20 pg/ml | p=0.0007** |

HIV-1+TB

| HIV-1 | Normal Healthy |
|-------|----------------|
| 7 | IL-17 | 305.57 ± 93.24 pg/ml | 5 | 387.0 ± 111.55 pg/ml | p=0.5870 |
| 7 | IL-22 | 44.380 ± 26.56 pg/ml | 5 | 55.58 ± 29.96 pg/ml | p=0.7872 |
| 7 | TGF-β | 1.475 ± 0.29 ng/ml | 5 | 1.620 ± 0.28 ng/ml | p=0.7360 |
| 7 | IFN-γ | 224.29 ± 34.62 pg/ml | 5 | 238.00 ± 44.62 pg/ml | p=0.8102 |

ELISA test on pleural fluid samples

| HIV-1+TB | TB |
|---------|-----|
| 7 | IL-17 | 174.685 ± 46.40 pg/ml | 8 | 187.87 ± 16.54 pg/ml | p=0.7820 |
| 7 | IL-22 | 128.491 ± 82.90 pg/ml | 8 | 1740.375 ± 607.42 pg/ml | p=0.0290* |
| 7 | TGF-β | 2.917± 0.878 ng/ml | 8 | 4.625 ± 0.679 ng/ml | p=0.1432 |
| 7 | IFN-γ | 37.43 ± 7.55 pg/ml | 8 | 371.0 ± 142.42 pg/ml | p=0.8102 |

Abbreviations: IL-17-Interleukin 17; IL-22-Interleukin 22; TGF-β-Transforming growth factor beta; IFN-γ-Interferon gamma; Statistical significance-* p=0.05 (significant); **p=0.0001 (Highly significant).
Figure 2: Cytokine values in the serum of HIV-1, TB, HIV-1+TB and healthy normal subjects. IL-17 and IFN-γ levels are significantly (*) high in HIV-1 and TB groups when compared with the healthy normal subjects. No significant difference in IL-17 and IFN-γ in HIV-1+TB and TB groups.

Figure 3: HIV-1+TB and TB patient’s lungs showing pleural effusion. IL-22 and IFN-γ cytokine values are significantly (*) higher in the pleural fluid samples of TB compared to HIV-1+TB patients.
Figure 4: Linear regression (R^2) analysis applied between HIV-1+TB and TB groups show a significant difference in IL-17 (r=0.7345; R^2=0.8051; p=0.0061) in pleural fluid whereas insignificant differences in IFN-γ (r=0.6392, R^2=0.3327, p=0.1752). In serum samples of HIV1+TB and TB patients, there was an insignificant difference in IL-17 (r=0.9107; R^2=0.0464, p=0.64270 whereas significant in IFN-γ (r=0.949; R^2=0.6481, p=0.0289) levels.

4. Discussion

Our present study indicates the role of IFN-γ and IL-17 with other cytokines like IL-22, TGF-β, and CD4 cells in HIV-1, TB and HIV-1 co-infected with TB patients. We measured the serum levels of pro-inflammatory cytokines (IFN-γ and IL-17) and anti-inflammatory cytokines (IL-22 and TGF-β). The main findings of our study showed: 1) the serum IFN-γ level was significantly higher in HIV-1 and TB patients, 2) patients with HIV-1+TB had lower IFN-γ levels and IL-22 and CD4 cell counts compared with the TB patients, 3) the level of IL-22 and IFN-γ were significantly lower in the pleural fluid of HIV-1 co-infected with TB than the TB patients, 4) CD4 counts were lower in HIV-1+TB patients than the TB patients and the normal healthy subjects, and 5) HIV-1+TB co-infected did not seem to affect serum level of IFN-γ and IL-17 and IL-22 and TGF-β compared with HIV-1 patients. CD4 counts were not statistically difference between TB positive with HIV-1 and TB negative HIV-1 patients.

Our study showed that IFN-γ, IL-17 and IL-22 cytokines were increased significantly in serum from HIV patients. There was a significant increase in IL-22 and IFN-γ in the pleural fluid of TB patients when compared with HIV-1+TB co-infected patients. This suggests that imbalances in Th17 in TB and HIV-1 co-infection were probably due to the interaction of Mycobacterium antigens with previous sensitized T-lymphocytes to release other different cytokines [8].
The importance of CD4 T lymphocyte function is seen in patients with HIV, where the risk of TB increases with the decrease of cell counts. Clinical assessment of IFN-γ level in the serum of TB has been used to determine as biomarker compared to the pathogenesis of HIV-1+TB. However, probably IFN-γ exhibits polyfunctional effects on immune activation, proinflammatory responses and immunomodulation. The present study revealed a high level of IFN-γ in TB than the HIV-1 patients with primary TB infection. In HIV-1 chronic disease, IFN-γ levels decline to a steady state that is often equivalent to healthy controls [9, 10]. The primary TB infection in HIV-1 patients has induced IFN-γ production that has reduced the adaptive immune responses to the development of HIV-1. Such inflammatory activities can promote HIV-1 infection and may cause a higher viral set point before T cell immunity can control the HIV-1 load.

HIV-1 exposure with pulmonary tuberculosis infection is associated IL17 and IL22 responses, both systemically and at the site of mucosal exposure impairs Th1 and Th17 [11]. IFN-γ-producing CD4 T act as co-stimulators to activate CD8 T lymphocytes is essential for CD8⁺ T cell function in Mycobacterium tuberculosis infection [12]. Interferon-γ and IL-17 play a pivotal role in the protective immune response against M. tuberculosis associated with viral replication [13, 14]. The importance of CD4 T lymphocyte function is seen in patients with HIV, where the risk of TB increases with the decrease of cell counts.

The immune responses to M. tuberculosis showed that B cell and humoral immunity interacts with T cells and other effector cells [15, 16]. A variety of effector lymphocyte subsets are induced by infection, but not all responding cells make IFN-γ [17]. On the other hand, IL-22 cytokine is produced by innate lymphoid cells and natural killer cells. Th1 and Th17 and Th22 cells bind to IL-22R1 receptor which is expressed on non-hematopoietic lung epithelial cells [18]. Th17 cells have a potential impact on tuberculosis (TB) infection when compared with healthy persons. IFN-γ and Th17 cells in tuberculous pleural effusion indicate that Th17 cells contribute to the immunopathogenesis of tuberculous pleural effusion [19]. In HIV infection, TGF-β+NK and IL-10+NK cells play negative regulatory roles. TGF-β is a cytokine release immunosuppressive factor in HIV disease and promotes virus replication in infected monocytes and peripheral blood mononuclear cells and increases susceptibility to opportunistic infections [20].

Our study concludes the findings that the serum IFN-γ level was significantly higher in HIV-1 patients and also in primary TB patients than the normal healthy subjects. CD4 counts were lower in the HIV-1+TB group than the TB and normal healthy subjects. Patients with primary HIV-1+TB had insignificant differences in IFN-γ, IL-17, IL-22 levels in serum and CD4 cell counts, compared with the HIV-1 control group. The level of IL-22 and IFN-γ were significantly lower in the pleural fluid of HIV-1 co-infected with TB than the TB patients. TGF-β did not seem to affect serum level in HIV-1+TB co-infection as there was no statistically significant difference between TB positive and TB negative HIV-1 patients. The role of IFN-γ and IL-17 and IL-22 did not appear significant high in HIV-1 co-infected tuberculosis (HIV-1+TB) patients due to severely affected immune system.
References

1. Bell LCK, Noursadeghi M. Pathogenesis of HIV-1 and Mycobacterium tuberculosis co-infection. Nature Reviews Microbiology 16 (2018): 80-90.

2. Pawlowski A, Jansson M, Skold M, et al. Tuberculosis and HIV co-infection. Plos Pathogens 8 (2012): e1002464.

3. Toossi Z, Johnson JL, Kanost RA, et al. Increased replication of HIV-1 at sites of Mycobacterium tuberculosis infection: potential mechanisms of viral activation. Journal of Acquired Immune Deficiency Syndromes 28 (2001): 1-8.

4. Ness-Schwickerath KJ, Morita CT. Regulation and function of IL-17A- and IL-22-producing γδ T cells. Cellular and Molecular Life Sciences 68 (2011): 2371-90.

5. Wu J, Wang S, Lu C, et al. Multiple cytokine responses in discriminating between active tuberculosis and latent tuberculosis infection. Tuberculosis (Edinb) 102 (2017): 68-75.

6. Canaday DH, Sridaran S, Van Epps P, et al. CD4+ T cell polyfunctional profile in HIV-TB coinfection are similar between individuals with latent and active TB infection. Tuberculosis (Edinb) 95 (2015): 470-75.

7. Zhang M, Wang Z, Graner MW, et al. B cell infiltration is associated with the increased IL-17 and IL-22 expression in the lungs of patients with tuberculosis. Cellular Immunology 270 (2011): 217-23.

8. Korb VC, Phulukdaree A, Laloo UG, et al. TB/HIV pleurisy reduces Th17 lymphocyte proportion independent of the cytokine microenvironment. Tuberculosis (Edinb) 99 (2016): 92-9.

9. Spellberg B, Edwards JE Jr. Type 1/Type 2 immunity in infectious diseases. Clinical Infectious Diseases 32 (2001): 76-102.

10. Roff SR, Noon-Song EN, Yamamoto JK. The Significance of Interferon-γ in HIV-1 Pathogenesis, Therapy, and Prophylaxis. Frontiers in Immunology 4 (2014): 498-509.

11. Murray LW, Satti I, Meyerowitz J, et al. Human Immunodeficiency Virus Infection Impairs Th1 and Th17 Mycobacterium tuberculosis-Specific T-Cell Responses. Journal of Infectious Diseases 217 (2018): 1782-1792.

12. Green AM, Difazio R, Flynn JL. IFN-γ from CD4 T cells is essential for host survival and enhances CD8 T cell function during Mycobacterium tuberculosis infection. Journal of Immunology 190 (2013): 270-277.

13. Wang F, Mao L, Hou H, et al. The source of Mycobacterium tuberculosis-specific IFN-γ production in peripheral blood mononuclear cells of TB patients. International Immunopharmacology (2016) 32: 39-45.

14. Sodhi A, Gong J, Silva C, et al. Clinical correlates of interferon γ production in patients with tuberculosis. Clinical Infectious Diseases 25 (1997): 617-620.

15. Kozakiewicz L, Phuah J, Flynn J, et al. The role of B cells and humoral immunity in Mycobacterium tuberculosis infection. Advances in Experimental Medicine and Biology 783 (2013): 225-250.

16. Chan J, Mehta S, Bharrhan S, et al. The role of B cells and humoral immunity in Mycobacterium tuberculosis infection. Seminars in Immunology 26 (2014): 588-600.

17. Cooper AM. Cell-mediated immune responses in tuberculosis. Annual Review of Immunology 27 (2009): 393-422. Seminars in Immunology 26 (2014): 588-600.
18. Ronacher K, Sinha R, Cestari M. IL-22: An Underestimated Player in Natural Resistance to Tuberculosis? Frontiers in Immunology 9 (2018): 2209-2216.
19. Wang T, Lv M, Qian Q, et al. Increased frequencies of T helper type 17 cells in tuberculous pleural effusion. Tuberculosis (Edinb) 91 (2011): 231-237.
20. Jiang Y, Yang M, Sun X, et al. IL-10+ NK and TGF-β+ NK cells play negative regulatory roles in HIV infection. BMC Infectious Diseases 18 (2018): 80-90.