Phylogenetic Analysis of Local-Scale Tree Soil Associations in a Lowland Moist Tropical Forest

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Abstract

Background: Local plant-soil associations are commonly studied at the species-level, while associations at the level of nodes within a phylogeny have been less well explored. Understanding associations within a phylogenetic context, however, can improve our ability to make predictions across systems and can advance our understanding of the role of evolutionary history in structuring communities.

Methodology/Principal Findings: Here we quantified evolutionary signal in plant-soil associations using a DNA sequence-based community phylogeny and several soil variables (e.g., extractable phosphorus, aluminum, and manganese, pH, and slope as a proxy for soil water). We used published plant distributional data from the 50-ha plot on Barro Colorado Island (BCI), Republic of Panamá. Our results suggest some groups of closely related species do share similar soil associations. Most notably, the node shared by Myrtaceae and Vochysiaceae was associated with high levels of aluminum, a potentially toxic element. The node shared by Apocynaceae was associated with high extractable phosphorus, a nutrient that could be limiting on a taxon specific level. The node shared by the large group of Laurales and Magnoliidae was associated with both low extractable phosphorus and with steeper slope. Despite significant node-specific associations, this study detected little to no phylogeny-wide signal. We consider the majority of the ‘traits’ (i.e., soil variables) evaluated to fall within the category of ecological traits. We suggest that, given this category of traits, phylogeny-wide signal might not be expected while node-specific signals can still indicate phylogenetic structure with respect to the variable of interest.

Conclusions: Within the BCI forest dynamics plot, distributions of some plant taxa are associated with local-scale differences in soil variables when evaluated at individual nodes within the phylogenetic tree, but they are not detectable by phylogeny-wide signal. Trends highlighted in this analysis suggest how plant-soil associations may drive plant distributions and diversity at the local-scale.

Introduction

An important goal in tropical ecology is to identify the strength of plant associations with their physical environments in order to understand the factors underlying the distribution and abundance of species. Identifying plant-soil associations is not a new goal (for example, see studies reviewed in [1]). Many advances have been made at the levels of landscape and mesoscale (10^2–10^3 km²) and 1–100 km² respectively, as defined by [2], e.g. [3,4,5,6,7,8,9]), but progress in understanding local-scale (<1 km²) associations in the tropics has been slower for two reasons. First, intensive inventories are required to quantify the spatial distributions and abundances of tropical tree communities. Second, quantitative measurements of the abiotic environment are required at a high spatial resolution within the tree communities being inventoried. Work in Malaysia has succeeded in confronting both of these obstacles by mapping fine scale variation in soil texture [10,11], which can correlate with soil resources, within a 52-ha plot. Similarly, recent work on Barro Colorado Island (BCI) in the Republic of Panama has mapped the spatial distribution of soil nutrients and elements within the 50-hectare forest dynamics plot on the island [2]. The results of this work have shown that indeed some tree species in the BCI plot had significant associations with soil nutrients and elements.

Here we expand on previous work on local-scale plant-soil associations by moving the analyses beyond the species-level associations and investigating the degree to which closely related...
species share similar associations for soil variables on a local-scale. Such an approach may be useful for understanding local-scale patterns that exist across highly diverse tropical forests that may share few species but have ‘deeper’ phylogenetic overlap (e.g., shared genera and families). For example, physiological traits which may influence plant distributions are known to exist at phylogenetic nodes for aluminum accumulation (e.g., Vochysiaceae and Melastomataceae). This trait can serve as a mechanism of aluminum tolerance [12] and it is reasonable to hypothesize that, in some systems, it could translate into local-scale plant-soil associations for those groups. Similarly, node-specific plant associations could exist for other elements, including nutrients that may be limiting for plant productivity. In highly weathered soils in Hawaii [13] phosphorus has been shown to be a limiting nutrient and could be limiting to different degrees, perhaps on a taxon-specific basis, in other tropical forests.

The degree to which soil nutrient and element associations are similar amongst closely related species is also of practical importance for phylogenetic investigations into the community structure [14,15,16,17,18,19,20]. Recent work has shown that within some topographically defined habitats of the BCI forest plot, species assemblages tend to be more similar in their phylogenetic composition than expected [16,19]. Knowledge regarding the degree to which some closely related species occur on similar positions on soil resource gradients may help uncover mechanisms underlying empirical patterns of community phylogenetic structure.

Although it is not possible to conclusively determine that associations are the result of soil influencing plant distribution, as opposed to plants influencing soil variables, there are a few reasons why the sampling design of the BCI soil nutrient and element data set is more likely to detect plant responses to soil variation. As outlined by others [2], the high tree diversity homogenizes inputs, decreasing feedbacks of litter quality on soil properties. In addition, soil nutrients and elements are correlated with topography suggesting geological processes underlie nutrient and element variation in the plot. Furthermore, soil sampling was intensive but, due to the size of the plot area, data analysis is dependent on kriging between the 50 m×50 m (plus 3 random samples at alternate grid points) sampling locations to create estimates at a scale of 20 m×20 m. Considering this spatial scale, along with the high number of small individuals in the data set (~90 percent of the individuals in the BCI plot are less than 10 cm in diameter), strengthens the argument that associations are more likely to be due to soil influences on plant distribution than the reciprocal effect. However, an important caveat to this reasoning is when species are highly aggregated, in which case many individuals could have a cumulative effect on soil properties. Within the BCI 50-ha plot, it has been shown that most species are spatially aggregated but there is large variation in the degree of aggregation [21].

The present study used a phylogenetic hypothesis of the relationships among the taxa in the BCI forest plot to quantify the evolutionary signal in three soil nutrients and elements (extractable phosphorus, aluminum and manganese), pH, slope and a general index of soil fertility determined by using Principal Component Analyses (PCA) axes which combine 15 soil variables. Our main hypothesis was: if the evolution of soil associations has been relatively conserved for a given soil variable or group of variables (as evaluated through PCA axes), there will be node-specific signal in soil variables, suggesting that the association between edaphic variation and local-scale plant distribution within the BCI plot exist beyond the species-level. The null hypotheses being: an absence of signal in the soil variables. This would suggest that local-scale variation of edaphic factors are not important in determining distributions of clades of species; however, the associations between plant distributions and soil factors can still exist for individual species (as shown in [22]).

In contrast to our expectations for node-specific signal, we did not expect to find phylogeny-wide signal for extractable aluminum, manganese, phosphorus, or slope (to the extent that it is a proxy for water), due to the nature of these metrics. We are interested in whether groups of species show association with soil variables, which serve as the ‘trait’ values for the phylogenetic analyses. These are not trait values in the traditional sense, as they are not directly measured on individual organisms. However, the soil variables have the potential to represent the evolutionary history of an organism and we consider them to be ecological ‘traits’, which is one category of traits highlighted in Blomberg et al. [22], along with morphological, life history, physiological, and behavioral. For example, Blomberg et al. [22] consider mean annual temperature and seasonality as ecological traits which, similar to soil variables, are abiotic factors that could influence an organism’s range of potential environments and competitive ability.

Here we are investigating if taxa show tolerance, or intolerance, within the ranges of soil variables. It follows that, within a group, species exhibiting tolerance or intolerance would have average values of the soil metric of interest clustered toward the extreme of the gradient compared to those expected by chance. For tolerance, the shifted average would reflect either 1) the ability to occupy the more extreme sites while also existing on less extreme sites or 2) the limited distribution on the tail of the tolerance gradient due to a trade-off decreasing their presence in the rest of the gradient. In contrast, data for less tolerant taxa would extend over a range of less extreme values. Therefore, phylogeny-wide signal based on soil variables would be unlikely to be significant because well constrained trait values throughout the phylogeny, which might be expected for morphological or physiological traits, are not expected for this ecological trait. However, the phylogeny could show node-specific associations in the trait value (as hypothesized above).

**Methods**

Barro Colorado Island (BCI) is a semideciduous moist forest with average rainfall of 2,600 mm [23]. In the 50-ha plot, red light clays are the most abundant soil category but brown fine loams are also present [24] (for more information on BCI soils see [25]). Census data for the 50-ha plot was collected by Smithsonian Tropical Research Institute (STRI) Forest Dynamics project, lead by R. Condit, S. Hubbell and R. Foster [26,27]; data can be requested at http://ctfs.si.edu/datasets/bci/ and we used data from the 2005 census. All trees ≥1cm diameter are included in the census and the sampling protocol is described in [28].

For soil traits, we focus on previously published data on extractable phosphorus, aluminum, manganese, pH, and slope. We also use a general index of soil fertility using PCA axes created from analysis of 13 soil variables. Soil collection and analysis was conducted by J. Dalling, K. Harms, R. John, R. Stallard, and J. Yavitt [2,29]. Methods are described in [30]. Briefly, 300 samples were collected to 10 cm depth following a 50 m×50 m grid (plus 3 random samples at alternate grid points) and soil analyses included extractable aluminum, boron, calcium, copper, iron, potassium, magnesium, manganese, inorganic nitrogen, phosphorus and zinc, in addition to net nitrogen mineralization rates, and pH. These data were then used to create estimates at a scale of 20 m×20 m. Nitrogen was extracted using potassium chloride and estimated colorimetrically using an autoanalyzer, while all other nutrients and elements were extracted with a Mehlich-3 solution [31] and
analyzed using inductively coupled plasma-atomic emission spectrometry (ICP-AES) at Cornell University Laboratories [2]. Net nitrogen mineralization rates represent nitrogen mineralized during a 28 day in situ incubation. Soil pH was measured using a 1:3 ratio of field moist soil to distilled water [30]. In addition, although slope and elevation are topographic and not soil characteristics, we included them as soil variables to guide inference about the importance of soil water. For example, on BCI steep slopes have been shown to have higher water potentials than plateau areas [32,33]. The slope and elevation for each 400 m² subplot inside the BCI forest plot were downloaded with the original 2005 census data.

**Supertree and trait files**

The present study uses a previously published molecular phylogeny for the BCI forest plot published by [19]. The phylogeny was generated using a three-locus (rbcL, trnH-psbA, and matK) DNA barcode dataset generated for the plot. The sequencing, alignment and phylogenetic inference methods are discussed in detail elsewhere [19]. The phylogeny resolves the majority of the nodes with the phylogeny and provides a marked improvement over phylogenetic supertrees that paste species onto the phylogeny backbone using the taxonomic hierarchy [34].

To generate the species by soil variable association data set we first made geographic information system (GIS) map layers of each of the soil variables. Next, we performed a principal component analysis (PCA) to investigate suites of correlated soil variables. Data input into the PCA analysis were log transformed when necessary (i.e. extractable B, Ca, Cu, Fe, K, Mg, P and Zn, and elevation) and scaled. The resulting PCA scores were used to generate GIS map layers for the first three PCA axes. Next using the x-y coordinates in the original BCI census file we plotted the BCI tree data in the GIS on top of the soil variable maps and extracted the soil variable value for each individual stem. Then for each species we calculated the median value on which each species was found for each soil variable. Phylogenies and trait files for medians were created for all individuals ≥1 cm in diameter.

**Phylogenetic signal**

Phylogenetic signal is defined as the tendency for related species to resemble each other more than they resemble species drawn at random from the tree [22] and phylogenetic signal is often calculated using the mean contrast value for all nodes descendant of a node [22,35,36,37]. In this analysis we use three metrics (phylogeny-wide signal, node-specific signal and Blomberg’s K statistic) to measure the tendency of related species to resemble each other more than they resemble species drawn at random. Phylogeny-wide signal indicates if a pattern was detected across the entire phylogeny, while node-specific signal indicates if significant associations were detected for nodes within the phylogeny. Blomberg’s K statistic was used to quantify phylogenetic signal in the soil data and to compare the magnitude of effect sizes among traits as measured from the root node [38]. For consistency, we only refer to the metrics which use contrast values as phylogenetic signal (i.e. the phylogeny-wide signal and Blomberg’s K statistic; see below). We use the term evolutionary signal to refer to all three metrics, including node-specific signal.

The analysis of traits (AOT) module in the software Phylocom v4.0 [39] was used to test for phylogeny-wide signal and node-specific signal in soil variables. In the AOT procedure, phylogeny-wide signal was evaluated at the root node, while node-specific signal was tested for each node in the phylogeny. Phylogeny-wide signal here is based on the mean contrast value for all nodes descendant of a node, while node-specific signal is based on means of all taxa subtended by the node of interest. The analysis for Blomberg’s K statistic was performed using the R package Picante [40].

To be conservative with interpreting significant node-specific results from AOT module, in this paper we focus on nodes that were significant for both tip and ancestral averaging. Detailed information and references for all AOT can be found in [37]. The observed mean value of each node is compared to a null distribution of mean values that is generated by permuting trait values across the tips of the phylogeny 999 times. If the observed value for a node lands in the p = 0.05 tails of the null distribution, the trait has significant signal at that node and all descendant taxa have average trait values more similar than expected.

High and low tail tests were conducted for node-specific associations of all traits. However, for PCA axes, only associations with the high tail tests are considered. Significant associations with low values of a PCA axis are not biologically meaningful and therefore are not included in the results. Considering the number of species and the number of individuals per species for a given node is important for identifying significant node-trait associations that may be skewed by poor representation of one or several taxa. To address this, we list the number of individuals per species for each node of interest. Significant nodes that are dependent on a species that has less than five individuals are not included in the results and nodes representing large numbers of individuals are given emphasis during interpretation of the results.

**Results**

**Principal components analysis of soil traits**

The first three PCA axes were found to represent 74% of the total variance among the 15 soil variables used in this analysis, with the first axis representing more than three times more variance than the second (Table 1). The fourth axis explained only 8%, while the fifth explained only 3% of the variance. Therefore, only the first three axes are used as phylogenetic analysis ‘traits’ to represent combinations of soil variables that were well represented within the plot. Our analysis used additional variables compared to the PCA results reported in [2] (Table 1), leading to different results, most notably of axes 2 and 3.

**Signal in plant-soil associations**

Of the eight soil variables evaluated (medians of four soil variables, slope, plus 3 PCA axes), none were found to have a significant phylogeny-wide phylogenetic signal (p<0.05; data not shown). Only two traits (PCA axis 2 and 3, Table 2) were found to have a K statistic greater than unity. The lack of significance for most variables suggests that, at the phylogeny-wide level, closely related species do not have more similarity than expected by Brownian motion model of trait evolution. Thus, across the entire phylogeny there was little to no phylogenetic signal in species soil associations in the BCI forest plot.

Although the analysis lacked phylogenetic signal across the entire phylogeny, a number of significant signals at individual nodes (i.e., node-specific signals) were detected. Rate of significance was 3.8%, which is less than that expected given the number of tests conducted. Despite this, we do report on those instances where groups of closely related species share similar soil associations. Most notably, the node shared by the Laurales and Magnoliidales was significantly associated with low extractable phosphorus (Table 3, Figure S1), high slope (Table 4) and PCA axis 3 (Table 5). *Protium* (in Burseraceae) was also associated with low extractable phosphorus (Table 3). In contrast, Apocynaceae and nodes uniting groups within Apocynaceae were found to be associated with higher levels of extractable phosphorus (Table 4).
Table 1. Loadings for the first three axes of principal components analysis (PCA) and the mean and range of the soil variables in 20 m × 20 m data set.

| Soil variable       | Axis 1  | Axis 2  | Axis 3  | Trait mean and range             |
|---------------------|---------|---------|---------|----------------------------------|
| Aluminum            | 0.142   | 0.271   | 0.45    | 1012.1 (283.2–1563.8)            |
| Boron               | −0.3    | 0.169   | 0.169   | 0.93 (0.04–3.33)                 |
| Calcium             | −0.310  | 0.193   | 0.271   | 1697.3 (343.5–4416.5)            |
| Copper              | −0.234  | 0.255   | 0.155   | 7.1 (1.3–15.1)                   |
| Iron                | −0.219  | 0.207   | −0.226  | 178.0 (51.1–350.0)               |
| Potassium           | −0.367  | 0.143   | −0.469  | 167.4 (58.2–391.1)               |
| Magnesium           | −0.274  | 0.211   | 0.389   | 296.3 (43.9–861.3)               |
| Manganese           | −0.285  | 0.522   | 0.436   | 368.2 (9.4–781.5)                |
| Phosphorus          | 0.782   | −0.113  | −0.248  | 2.87 (0.30–8.30)                 |
| Zinc                | −0.355  | −0.132  | 0.115   | 5.1 (0.9–17.9)                   |
| Nitrogen            | −0.103  | 0.291   | 0.257   | 7.9 (12.5–47.1)                  |
| Nitrogen mineralization | −0.459 | −0.692  | −0.224  | 17.9 (−11.7–47.6)               |
| pH                  | −0.144  | −0.109  | −0.116  | 4.67 (3.64–5.35)                |
| Elevation           | 0.122   | −0.188  | 0.211   | 144.4 (121.3–159.3) m            |
| Slope               | −0.202  | −0.409  | 0.211   | 4.33 (0.02–16.66) degrees       |
| % variance          | 51%     | 13%     | 10%     |                                  |

Soil variables include Mehlich-3 extractable elements, potassium chloride extractable inorganic nitrogen, nitrogen mineralization rates, pH, slope and elevation. All elements are expressed as mg element kg⁻¹ dry soil. Nitrogen mineralization rates are expressed as ng N mineralized kg⁻¹ soil per 28 days. Percent variance for each axis is included. doi:10.1371/journal.pone.0013685.t001

Table 2. Bloomberg’s K statistic for soil traits.

| Trait     | K   | P value |
|-----------|-----|---------|
| Al        | 0.80| 0.091   |
| Mn        | 0.77| 0.102   |
| P         | 0.92| 0.010   |
| pH        | 0.64| 0.694   |
| elevation | 0.88| 0.091   |
| slope     | 0.83| 0.146   |
| PCA axis 1| 0.69| 0.556   |
| PCA axis 2| 1.45| 0.003   |
| PCA axis 3| 1.06| 0.044   |

Aluminum, manganese and phosphorus were extracted with Mehlich-3. See Table 1 for information on PCA axes. Significant results of a two tailed test are shown in bold. doi:10.1371/journal.pone.0013685.t002

Node-specific results for association with extractable aluminum and manganese, both potentially toxic elements, were intriguing. The most recent common ancestor node for Myrtaceae and Vochysiaceae, in addition to the node shared by Eugenia and Chamguava (in Myrtaceae) and a number of nodes shared by species of Ficus (in Moraceae) showed association with high extractable aluminum (Table 4, Figure S2). While the Ficus results are based on a small number of individuals, the node shared by Myrtaceae and Vochysiaceae and the node hosting Eugenia and Chamguava are based on robust sample sizes (Table 4). The most recent common ancestor node for Psidium guajava, Osmoxylon excelsa, and Maquira costaricana (in Moraceae) was associated with high manganese (Table 4). In addition, Protium (in Burseraceae) was associated with high manganese (Table 4). However, as summarized in the footnotes of Table 5, a number of groups showed association with more than one trait, as is the case for Protium, which was also significantly associated with low extractable phosphorus as mentioned above. A number of nodes also showed association with suites of traits as evaluated through the PCA axes (Table 5) and pH. The range of pH in this study is unlikely to directly influence plants but pH can be considered as an important proxy for nutrient and element availability. The node shared by Salicaceae and Clusiaceae was associated with low soil pH, as was the node uniting only Salicaceae and a number of nodes within Salicaceae (Table 3). Meliaceae, on the other hand, showed a significant association with high pH (Table 4).

Discussion

The present study asked whether closely related tropical tree species tend to share similar soil associations in a tropical forest. Overall there was little to no phylogeny-wide evolutionary signal in soil nutrient/element associations, however the results from individual nodes within the phylogeny show that medians for a number of soil metrics had significant signal, and these instances may be important for understanding plant distributions at the local scale in highly diverse tropical forest. This suggests that previous phylogenetic analyses of BCI assemblages that found significant phylogenetic structuring in different habitats [16,19] may be partially explained upon the basis of phylogenetically conserved soil nutrient and element associations. However, because many soil variables within the dataset are correlated (Table 7 in [2]), and we could not account for species aggregation due to dispersal limitation, relationships between soil variables and taxa may not be causal. Below we discuss the results with respect to the association between individual soil variables and particular clades to highlight how plant-soil associations may drive the maintenance of diversity and the distributions of tropical genera and families.

Aluminum

Aluminum is nonessential for plant growth and can be toxic, especially at pH less than 5.5 [41]. Lowland tropical forests supported by highly weathered soils often have high levels of extractable aluminum and relatively low soil pH, and floras of many forests on acidic soil have been suggested to confer tolerance to this element [41,42]. The range of soil pH values, which are below 5.5, indicate that aluminum toxicity could be a concern in the BCI 50-ha plot soil. Moreover, the range of Mehlich-3 extractable aluminum was large (283–1563 mg kg⁻¹, based on the 20m × 20m scale data), suggesting that the potential of toxicity could vary.

This analysis detected an association between high extractable aluminum and the node shared by Myrtaceae and Vochysiaceae, Eugenia and Chamguava (in Myrtaceae), nodes within Ficus (in Moraceae) and a node shared by two species of Psychotria (Rubiacae) (Table 4). To different degrees, all of these taxa with the exception of Ficus have been shown to be aluminum accumulators, which is one known mechanism for tolerating this potentially toxic element [43]. Vochysiaceae is well known to be a family characterized by ability to accumulate aluminum while species within Myrtaceae, and species within Eugenia, are noted as sometimes being aluminum accumulators [12]. Rubiiodaeae, which includes Psychotria, is also noted as a group of aluminum accumulators [44,45]. However, out of the 12 species of Psychotria present in the plot, only one node shared by several species of Psychotria was found to be significantly associated with high extractable aluminum and this node did not include P. horizontalis, which has over 3,000 individuals in the dataset.
It was interesting that Melastomataceae, which is a family well established as being aluminum accumulating [46], was not detected as having significant association with aluminum in this analysis. We note, however, that across the plot, the group could have a lower median of extractable aluminum yet still show preferential success in high aluminum areas compared to other taxa. In such a case an association would not be detected, although aluminum tolerance could still be an important strategy for taxa within this family. Although associations between Melastomataceae (and groups within this family) and aluminum were not detected in the current work, results do suggest that associations between aluminum and groups highlighted in this analysis, such as Myrtaceae and Vochysiaceae, Eugenia and Chamaedorea, and some groups of Ficus and Psychotria, could be important in determining local-scale plant distribution in BCI and other tropical forests that also show both high concentrations and large ranges of extractable aluminum.

Manganese

Manganese is an essential plant nutrient but can be toxic or induce other deficiencies when present in high concentrations [47]. Comparing the extractable manganese levels associated with the nodes showing significant association with high manganese in this analysis (which include the node hosting Poulsea armata, Olmeda apera and Maquia costaricana (in Moraceae), the node shared by three Zanthoxylum species (in Rutaceae), and Protium (in Burseraceae) (Table 4)) shows that the high manganese levels in this study are near levels that have been considered toxic in a tropical agricultural system which also used a Mehlich-3 extract [48]. However, clearly the value of direct comparisons of extractable manganese among systems is limited because plant ability to tolerate high levels of manganese is known to differ by species [49,50] and the effects of excess manganese can be dependent on concentrations of other ions [50]. Another line of evidence for the potential of excess manganese to influence plant distributions comes from temperate zones where high concentrations of manganese have been shown to negatively affect forest tree seedling performance [51,52,53,54].

Out of the three forests investigated in John et al. [2], the BCI average of extractable manganese was more than twice that for Yasuni (a lowland forest in Ecuador), and nearly two magnitudes greater than that of La Planada (a montane forest in Columbia; Table 2 in [2]). Although John et al. [2] did not find manganese to have a relatively strong effect on a species analysis of niche structure compared to other nutrients/elements for BCI, the effect of manganese tended to be greater on BCI compared to the other two sites (Figure 3 in [2]). Because some tropical forest soils are characterized by high levels extractable manganese, and negative effects of excess manganese on plant performance have been shown to vary on at least a species level, the potential role of manganese in structuring lowland tropical forests that show large ranges in extractable manganese concentrations is worthy of future consideration.

Phosphorus

Phosphorus is an important nutrient that is considered to have relatively low availability in highly weathered soils [55] and is thought to limit ecosystem processes in some tropical forests

Table 3. Significant node-specific evolutionary signals for low tail tests of trait (i.e. soil variable) medians.

| Trait | Node shared by: | Family | Individuals per species | T mean | T p-value |
|-------|----------------|--------|-------------------------|--------|-----------|
| P     | Protium*       | Burseraceae | 2829, 698, 2853, 9    | 1.87   | 0.05      |
| P     | Laurales & Magnoliiales* | 367, 27, 179, 112, 131, 2, 237, 162, 70, 59, 2115, 183, 1394, 44, 621, 472, 11327, 896, 123, 485, 794 | 2.19   | 0.02      |
| pH    | Salicaceae & Clusiaceae | 6, 24, 124, 34, 410, 26, 131, 14, 434, 67, 484, 23, 4, 41, 152, 393, 4602, 391, 1427 | 4.39   | 0.03      |
| pH    | Salicaceae*    | 6, 24, 124, 34, 410, 26, 131, 14, 434 | 4.02   | 0.002     |
| pH    | Casearia commersoniana, C. arborea, Zuelania guidonia, Laetia thamnia, L. procera, C. sylvestris, C. guianensis, C. aculeata | Salicaceae | 24, 124, 34, 410, 26, 131, 14, 434 | 3.99   | 0.004     |
| pH    | Casearia arborea, Zuelania guidonia, Laetia thamnia, L. procera, C. sylvestris, C. guianensis, C. aculeata | Salicaceae | 124, 34, 410, 26, 131, 14, 434 | 3.94   | 0.006     |
| pH    | Zuelania guidonia, Laetia thamnia, L. procera, Casearia sylvestris, C. guianensis, C. aculeata | Salicaceae | 34, 410, 26, 131, 14, 434 | 3.84   | 0.008     |
| pH    | Zuelania guidonia, Laetia thamnia | Salicaceae | 34, 410 | 2.24   | 0.006     |
| pH    | Psychotria marginata & P. gracilliflora* | Rubiaceae | 581, 53 | 4.37   | 0.044     |
| slope | Ficus obtusifolia & F. costaricana | Moraceae | 6, 8 | 2.13   | 0.046     |
| slope | Salicaceae* | See above |           |        |           |
| slope | Amauoa corymbosa, Borajoa panamensis, Albertia edulis | Rubiaceae | 22, 1, 370 | 2.17 | 0.008     |
| slope | Psychotria horizontalis, P. marginata, P. gracilliflora | Rubiaceae | 3119, 581, 53 | 2.38 | 0.024     |
| slope | Thevetia ahouna, Tabernaemontana arborea, Stemmadenia grandiflora, Raoulilla littoralis, Lacmellea panamensis* | Apocynaceae | 91, 1593, 1, 1, 100 | 2.40 | 0.008     |
| slope | Tabernaemontana arborea, Stemmadenia grandiflora, Raoulilla littoralis, Lacmellea panamensis* | Apocynaceae | 1593, 1, 1, 100 | 2.54 | 0.024     |
| slope | Elaeis oleifera, Chamaedorea topilote, Attalea butyrcacea* | Arecales | 21, 7, 34 | 2.28 | 0.024     |

All nodes listed were significant for both ancestral weighted trait and tip averaging analyses using data for all stems ≥1 cm. Tip (T) averaging means for Mehlich-3 extractable soil elements are shown in units of mg kg⁻¹ soil and slope is in degrees. Significant nodes which are dependent on a species that has less than five individuals are not shown. Nodes which were significant for more than one of the variables investigated are indicated with an asterisk. See footnotes of Table 5 for a summary of nodes with multiple associations.

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could be due to lower requirements or more effective acquisition may be relatively less sensitive to low levels of this nutrient, which level, even if a community as a whole is not limited. Analyses also that if taxa such as Apocynaceae are sensitive to low phosphorus example, see total soil phosphorus data in [58]), our results suggest [13,56,57]. Although soil phosphorus is relatively high on BCI (for groups, such as the Laurales and Magnoliales compared to [68], investigating differences in arbuscular associations among plant species [64,65] and plant species have been documented as having nonrandom associations with hosts [66,67]. Although a great deal of this diversity and trends among host taxa remains unknown [68], investigating differences in arbuscular associations among mechanisms linking taxa with phosphorus availability [59]. The could be related to root fungal associations, may be one mechanism linking taxa with phosphorus availability and/or correlations with nutrients/

| Trait | Node shared by: | Family | Individuals per species | T mean | T p-value |
|-------|----------------|--------|-------------------------|--------|-----------|
| Al    | *Ficus trigonata, F. papenoei, F. obtusifolia, F. costaricana, F. citrifolia, F. bullenei* | Moraceae | 5, 3, 6, 8, 1, 5 | 1172.53 | 0.002 |
| Al    | *Ficus papenoei, F. obtusifolia, F. costaricana, F. citrifolia, F. bullenei* | Moraceae | 3, 6, 8, 1, 5 | 1192.98 | 0.002 |
| Al    | Lythraceae, Vochysiaceae, Myrtaceae | Moraceae | 5, 27, 60, 46, 1816, 482, 1751, 611, 449 | 1130.94 | 0.002 |
| Al    | Vochysiaceae & Myrtaceae | Moraceae | 27, 60, 46, 1816, 482, 1751, 611, 449 | 1110.38 | 0.024 |
| Al    | *Eugenia & Champaguan* | Myrtaceae | 1816, 482, 1751, 611, 449 | 1116.23 | 0.044 |
| Al    | *Psychotria marginata & P. graciliflora* | Rubiaceae | See Table 3 | 1196.71 | 0.034 |
| Mn    | *Poulseia armata, Olmedia aspera, Maquira costaricana* | Moraceae | 1162, 149, 1396 | 454.64 | 0.022 |
| Mn    | *Zanthoxylum setulosum, Z. panamense, Z. ekmanii* | Rutaceae | 1, 178, 194 | 449.53 | 0.028 |
| Mn    | *Protium* | Bursaraceae | See Table 3 | 433.05 | 0.046 |
| Mn    | *Psychotria limonensis, P. granadensis, P. psychotrisfolia, P. chagrensis* | Rubiaceae | 65, 3, 1, 13 | 445.75 | 0.014 |
| P     | *Ficus papenoei, F. obtusifolia, F. costaricana, F. citrifolia, F. bullenei* | Moraceae | See above | 3.52 | 0.012 |
| P     | *Miconia impetiolaris, M. elata, M. dorsiloba, Clidemia septuplinervia* | Melastomataceae | 14, 12, 2, 3 | 3.45 | 0.048 |
| P     | *Apocynaceae* | Apocynaceae | See Table 3 | 3.90 | 0.006 |
| P     | *Thevetia ahouai, Tabernaemontana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmellea panamensis* | Apocynaceae | See Table 3 | 3.78 | 0.02 |
| P     | Tabernaemontana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmellea panamensis* | Apocynaceae | See Table 3 | 3.73 | 0.008 |
| P     | *Bactris major, B. barronis, Elaeis oleifera, Chamaedorea tepejilote, Attalea butryacea* | Arecaceae | 80, 5, 21, 7, 34 | 3.73 | 0.004 |
| P     | Elaeis oleifera, Chamaedorea tepejilote, Attalea butryacea* | Arecaceae | See Table 3 | 3.86 | 0.024 |
| pH    | *Sapium aucuparium, S. 'spnov', Hura crepitans* | Euphorbiaceae | 52, 3, 103 | 4.89 | 0.02 |
| pH    | *Meliaceae* | Moraceae | See above | 6.07 | 0.04 |
| slope | *Poulseia armata, Olmedia aspera, Maquira costaricana* | Moraceae | See above | 4.63 | 0.032 |
| slope | *Laurales & Magnoliales* | Moraceae | See Table 3 | 5.47 | 0.012 |
| slope | *Piper perlasense, P. colonense, P. reticulatum, P. imperialis, P. cordulatum, P. aequale* | Piperaceae | 17, 5, 131, 16, 50, 19, 27 | 5.45 | 0.02 |

See Table 3 for column details.
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[13,56,57]. Although soil phosphorus is relatively high on BCI (for example, see total soil phosphorus data in [58]), our results suggest that if taxa such as Apocynaceae are sensitive to low phosphorus availability, phosphorus limitation may exist on a taxon specific level, even if a community as a whole is not limited. Analyses also detected that the large clade of Laurales and Magnoliales was associated with low extractable phosphorus, suggesting this clade may be relatively less sensitive to low levels of this nutrient, which could be due to lower requirements or more effective acquisition strategies.

Taxon specific access to different forms of phosphorus, which could be related to root fungal associations, may be one mechanism linking taxa with phosphorus availability [59]. The large majority of tropical trees associate with arbuscular mycorrhizae [60], which is a wide spread plant-fungal association that is thought to be ancestral [61] and can improve phosphorus acquisition [62,63]. However, different arbuscular mycorrhizae have been shown to vary in their levels of benefit to different host species [64,65] and plant species have been documented as having nonrandom associations with hosts [66,67]. Although a great deal about the diversity of arbuscular fungi, the functional significance of this diversity and trends among host taxa remains unknown [68], investigating differences in arbuscular associations among groups, such as the Laurales and Magnoliales compared to Apocynaceae, could be especially interesting given the results of this study.

We do note, however, that the node shared by Laurales and Magnoliales was also significantly associated with slope (see below for discussion on slope) and the suite of traits influencing PCA axis 3, which includes low phosphorus and slope in addition to high aluminum and magnesium (Table 1). Similarly, Protium, another well represented group in this dataset (in Burseraceae), was also found to be significantly associated with low extractable phosphorus, suggesting this group too may have strategies for dealing with lower phosphorus availability. However, Protium was also significantly associated with high manganese, as discussed in the previous section. These results highlight the fact that this study is not based on experimental manipulations and we cannot conclude if associations are the result of directly related mechanisms, random correlations, or interactions between a number of traits. Our results suggest that a trade-off between an ability to tolerate lower phosphorus levels and compete in more favorable conditions may exist within the BCI plot, but this remains to be experimentally tested.

**Slope**

Mechanisms underpinning plant association with slope may be related to water availability and/or correlations with nutrients/
Sloped areas have been shown to be related to increased water potential during the dry season compared to plateau areas [32,33], and species densities on drier plateaus compared to wetter water potential during the dry season compared to plateau areas [32,33], and species densities on drier plateaus compared to wetter elements. Sloped areas have been shown to be related to increased water potential during the dry season compared to plateau areas [32,33], and species densities on drier plateaus compared to wetter

Slopes have been shown to be negatively correlated with drought [32,33], and species densities on drier plateaus compared to wetter

**Table 5. Significant node-specific evolutionary signals for high tail tests of PCA axes.**

| Trait          | Node shared by:                                                                 | Family                                                                 | Individuals per species | T mean | T p-value |
|----------------|------------------------------------------------------------------------------|------------------------------------------------------------------------|-------------------------|--------|-----------|
| PCA axis 1     | Solanaceae, Bignoniaceae, Acanthaceae, Verbenaceae                          |                                                                       | 69, 5, 4, 7, 230, 71, 5, 8, 280, 44 | 0.26   | 0.012     |
| PCA axis 1     | Bignoniaceae, Acanthaceae, Verbenaceae                                       |                                                                       | 230, 71, 5, 8, 280, 44  | 0.27   | 0.034     |
| PCA axis 1     | Psychotria marginata & P. graciliflora*                                      | Rubiaceae                                                             |                         |        |           |
| PCA axis 2     | Ficus papenoei, F. obtusifolia, F. costaricana, F. citrifolia, F. bullenei*  | Moraceae                                                              |                         | 0.11   | 0.004     |
| PCA axis 2     | Thevetia ahouai, Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis* | Apocynaceae                                                           |                         |        |           |
| PCA axis 2     | Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis* | Apocynaceae                                                           |                         |        |           |
| PCA axis 3     | Pseudobambus septentum & Ochroma pyramidal                                   | Malvaceae                                                             |                         |        |           |
| PCA axis 3     | Laurales & Magnoliales*                                                      |                                                                       |                         |        |           |
| PCA axis 3     | Laurales                                                                     |                                                                       |                         | 0.05   | 0.026     |
| PCA axis 3     | Siparuna                                                                     | Siparunaceae                                                          |                         |        |           |
| PCA axis 3     | Piper colonense, P. reticulatum, P. imperialis, P. cordulatum, P. aequale    | Piperaceae                                                            |                         |        |           |
| PCA axis 3     | Piper reticulatum, P. imperialis, P. cordulatum, P. aequale                  | Piperaceae                                                            |                         |        |           |

See Table 3 for column details.

Summary of nodes with multiple associations:

- Ficus papenoei, F. obtusifolia, F. costaricana, F. citrifolia, F. bullenei: high Al, P, and PCA axis 2.
- Poulsenia armata, Olmedia aspera, Maquira costaricana: high Mn and slope.
- Salicaceae: low pH and slope.
- Proton: low P, high Mn.
- Psychotria marginata & P. graciliflora: low pH, high Al and PCA axis 1.
- Thevetia ahouai, Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
- Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
- Laurales & Magnoliales: low P, high slope and PCA axis 3.
- Elaeis oleifera, Chamaedorea tepejilote, Attalea butyricca: low slope, high P.

See Table 3 for column details.

Summary of nodes with multiple associations:

- Ficus papenoei, F. obtusifolia, F. costaricana, F. citrifolia, F. bullenei: high Al, P, and PCA axis 2.
- Poulsenia armata, Olmedia aspera, Maquira costaricana: high Mn and slope.
- Salicaceae: low pH and slope.
- Proton: low P, high Mn.
- Psychotria marginata & P. graciliflora: low pH, high Al and PCA axis 1.
- Thevetia ahouai, Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
- Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
- Laurales & Magnoliales: low P, high slope and PCA axis 3.
- Elaeis oleifera, Chamaedorea tepejilote, Attalea butyricca: low slope, high P.

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- Salicaceae: low pH and slope.
- Proton: low P, high Mn.
- Psychotria marginata & P. graciliflora: low pH, high Al and PCA axis 1.
- Thevetia ahouai, Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
- Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
- Laurales & Magnoliales: low P, high slope and PCA axis 3.
- Elaeis oleifera, Chamaedorea tepejilote, Attalea butyricca: low slope, high P.

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- Thevetia ahouai, Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
- Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
- Laurales & Magnoliales: low P, high slope and PCA axis 3.
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- Laurales & Magnoliales: low P, high slope and PCA axis 3.
- Elaeis oleifera, Chamaedorea tepejilote, Attalea butyricca: low slope, high P.

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- Proton: low P, high Mn.
- Psychotria marginata & P. graciliflora: low pH, high Al and PCA axis 1.
- Thevetia ahouai, Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
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- Laurales & Magnoliales: low P, high slope and PCA axis 3.
- Elaeis oleifera, Chamaedorea tepejilote, Attalea butyricca: low slope, high P.

Summary of nodes with multiple associations:

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- Proton: low P, high Mn.
- Psychotria marginata & P. graciliflora: low pH, high Al and PCA axis 1.
- Thevetia ahouai, Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
- Taberna montana arborea, Stemmadenia grandiflora, Rauvolfia littoralis, Lacmella panamensis: low slope, high P and PCA axis 2.
- Laurales & Magnoliales: low P, high slope and PCA axis 3.
- Elaeis oleifera, Chamaedorea tepejilote, Attalea butyricca: low slope, high P.
ranges. For example, consider a system where potentially toxic concentrations are limited to high end of the gradient. Species within a group displaying tolerance would have either 1) similar average soil trait values on the extreme end of the tolerance gradient, reflecting both their ability to tolerate such conditions and an associated trade-off related to performance under more favorable conditions or 2) a larger range of soil trait values which is biased toward but is not limited to values at the extreme end if a trade-off did not exist. In both cases, and especially the former case, the tolerant species would have averages that are more similar to the extreme than expected by chance. On the other hand, trait values for non-tolerant taxa would span a larger range of less extreme values. In turn, the phylogeny-wide K statistic would be less than unity yet the phylogeny could show node-specific associations in the trait value. If on the other hand, the measured trait were the mechanism conferring the tolerance (i.e., a physiological trait), the species found on the large range of less extreme soil values could show a constrained trait value and a higher K statistic could be expected.

We suggest this may be the case for the trait of extractable soil aluminum in the current study, as it had a K statistic of less than unity (K = 0.80 with marginal significance of p = 0.09; Table 2) and node-specific results from the AOT showed a number of significant nodes associated with high aluminum. Again, it is important to note that the details of the tolerance related trait metric are key in determining expectations for the category of ecological tolerance. Studies that use a ranking of tolerance as the trait metric [80], as opposed to using the value of the environmental variable to which the organism is responding, would not follow the same expectations. Similarly, studies investigating tolerance and focusing on physiological adaptations to environmental stress as the trait variables would fall in the category of physiological traits, altering expectations for trait lability and K statistics.

Conclusion

The present work demonstrates that distributions of some plant taxa are associated with local-scale differences in soil variables when evaluated at individual nodes within the phylogenetic tree, but they are not detectable by phylogeny-wide signal. Some of the most intriguing results suggest aluminum and manganese, both potentially toxic elements, may structure plant distribution at the local-scale, as suggested by the significant associations between high extractable aluminum and the node shared by Vochysiaceae and Myrtaceae, in addition to the node shared by Eugenia and Changuana, and associations between high manganese and Protium armata, Omedula apera and Maquira costaricana (in Moraceae), the node shared by three Zanthoxylum species (in Rutaceae), and Protium (in Burseraceae). In addition, this study suggests slope may be an important variable underlying the distribution of Laurales and Magnoliidae, a large clade of plants, and Salicaceae, a well represented family.

The BCI 50-ha plot site was chosen to limit abiotic variation [70], which suggests that similar analyses of local-scale plant-soil associations in plots that are randomly laid out could detect more local-scale associations than found here. The results presented in this study highlight associations that are most promising for future work aimed at understanding the role of edaphic factors and evolutionary history in determining community assemblages, and the maintenance of diversity, at the local-scale. The degree to which associations are mechanistically underpinned by the soil variables highlighted in this analysis, versus being the result of correlations between multiple edaphic variables and other environmental factors, remains to be determined. In addition, understanding the extent to which patterns uncovered in this analysis hold among tropical forests is a valuable research direction.

Supporting Information

Figure S1 Extractable soil phosphorus. Mehlich-3 extractable soil phosphorus values (natural log transformed) are shown mapped on the molecular phylogeny for the BCI 50-ha plot. Found at: doi:10.1371/journal.pone.0013685.s001 (42.76 MB EPS)

Figure S2 Extractable soil aluminum. Mehlich-3 extractable soil aluminum values are shown mapped on the molecular phylogeny for the BCI 50-ha plot. Found at: doi:10.1371/journal.pone.0013685.s002 (10.72 MB EPS)

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Author Contributions

Conceived and designed the experiments: LAS NGS. Performed the experiments: LAS NGS. Analyzed the data: LAS NGS. Contributed reagents/materials/analysis tools: WJK DLE. Wrote the paper: LAS WJK DLE. NGS.

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