Herbivory Drives the Spread of Salt Marsh Die-Off

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Abstract

Salt marsh die-off is a Western Atlantic conservation problem that has recently spread into Narragansett Bay, Rhode Island, USA. It has been hypothesized to be driven by: 1) eutrophication decreasing plant investment into belowground biomass causing plant collapse, 2) boat wakes eroding creek banks, 3) pollution or disease affecting plant health, 4) substrate hardness controlling herbivorous crab distributions and 5) trophic dysfunction releasing herbivorous crabs from predator control. To distinguish between these hypotheses we quantified these variables at 14 Narragansett Bay salt marshes where die-off intensity ranged from <5% to nearly 98%. Nitrogen availability, wave intensity and plant growth did not explain any variation in die-off. Herbivory explained 73% of inter-site variation in die-off and predator control of herbivores and substrate hardness also varied significantly with die-off. This suggests that salt marsh die-off is being largely driven by intense herbivory via the release of herbivorous crabs from predator control. Our results and those from other marsh systems suggest that consumer control may not simply be a factor to consider in marsh conservation, but with widespread predator depletion impacting near shore habitats globally, trophic dysfunction and runaway consumption may be the largest and most urgent management challenge for salt marsh conservation.

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Introduction

Salt marsh die-off is a growing phenomenon in the western Atlantic [1] and a serious conservation problem as salt marshes provide valuable ecosystem services to coastal habitats [2]. Landscape wide die-offs were first described over 35 years ago in Hudson Bay, Canada, where increasing goose populations, fueled by artificial nitrogen fertilizer use, led to runaway grazing and marsh collapse [3]. Nearly 15 years later, salt marsh die-offs were described on the Southeastern and Gulf coasts of the U.S. driven by snail herbivory and exacerbated by drought, which increased the vulnerability of marsh cordgrass to consumers and fungal infections [4]. Most recently, salt marsh creek bank die-offs of the cordgrass Spartina alterniflora are occurring in New England, driven by predator depletion and release of the herbivorous marsh crab Sesarma reticulatum (hereafter Sesarma) from consumer control [5]. Historical reconstructions have revealed that Cape Cod die-offs have been ongoing, unnoticed for over two decades as a consequence of human population growth and recreational fishing pressure depleting predator populations [6, 7].

While it has been hypothesized that human impacts causing trophic dysfunction are driving these die-offs, salt marshes are widely considered to be controlled exclusively by physical forces [8, 9]. Consequently, many recent studies and reviews of salt marsh die-off fail to even acknowledge the impact of predator depletion and herbivory on die-off [10–13]. For example, a recent study on Cape Cod proposed from studies at a single site that eutrophication was causing salt marsh die-offs worldwide [10]. Other hypothesized causes of marsh die-off include harsh physical conditions e.g. hypersaline and anoxic soils [13], fungal infections [14], boat wakes [15], pollution [15], substrate type [16] and positive interactions between eutrophication and increased herbivory [17].

Recently, creek bank salt marsh die-off has spread into Narragansett Bay, where it had been reported absent as recently as 2009 [5]. We use this spread of salt marsh die-off to examine the relative importance of proposed drivers of die-off. We studied 14 Narragansett Bay salt marshes with a broad range of die-off conditions. We used archived aerial photographs to quantify the temporal pattern of die-off and then tested the competing hypotheses listed above with field measurements. To distinguish between hypotheses we quantified die-off, nitrogen availability (eutrophication), common source transplant performance (pollution and disease), wave exposure (boat wakes), herbivory, substrate hardness and predation on herbivorous crabs at each site (trophic dysfunction). We then used multiple regression to quantify the contribution of each of these factors to the spread of die-off.

Ethics Statement

All necessary permits for field sampling were obtained from the Cape Cod National Seashore.

Materials and Methods

Extent and Time Course of Die-Off

Fourteen accessible salt marshes with a range of die-off conditions were identified in Narragansett Bay. The extent of die-off was measured at each site in July 2013 by walking two randomly placed 100 m creek bank transects and every 10 m
measuring the total width of the cordgrass zone and the width of the cordgrass zone denuded by die-off.

We quantified the history of die-off at each site using archived aerial photographs from 1997, 2003, 2008, and 2012. Marsh habitat was defined as the total area covered in visible salt marsh vegetation and die-off area was defined as the total area converted from marsh vegetation to barren peat over each time interval. All areas were defined and quantified using ArcGIS. Marsh linear extent was defined as the total length of marsh edge bordering water (length of marsh border exposed to die-off conditions). To test the hypothesis that salt marsh die-off in Narragansett Bay is recent, percent linear extent affected by die-off was calculated for each site and time point. We tested the hypothesis that the intensity of die-off is increasing over time by measuring the mean width of die-off and the mean rate of die-off expansion for each site and time period. Mean die-off width was calculated as total area of die-off divided by length of border affected. Rate of die-off expansion was calculated as mean die-off width per year. An ANOVA with a univariate split plot approach was used to analyze both percent linear extent and the width of die-off over time.

Herbivory

Sesarma herbivory was measured at each site the second week of July 2013. Herbivory was estimated by walking a 20 m transect along the creek bank of each site and every 2 m scoring 100 cordgrass stems for crab herbivore damage in a 50×50 cm quadrat. Sesarma grazing leaves characteristic rasped edges and clipped blades on transplanted cordgrass [5].

Substrate Hardness

Substrate hardness, an accurate proxy for peat density [20], was measured with a penetrometer in vegetated areas along the grazing border at all sites. The penetrometer was a top loading spring scale with a 2 cm diameter, 7 cm long rod mounted vertically on the weighing platform. The force required to push the rod vertically into the substrate (n = 10/site) was recorded avoiding burrows along the same 20 m transect line used to assess crab herbivory.

Predation Pressure on Sesarma

To quantify trophic dysfunction in Narragansett Bay, operationally defined as how severely predator depletion has reduced predation pressure on Sesarma leading to increased Sesarma densities and herbivory, we quantified Sesarma densities, predation pressure on tethered Sesarma, and top predator biomass at each study site. Sesarma abundance was estimated with nocturnal pitfall traps in late July when the crabs showed maximum activity (n = 8 traps/site) [6]. Sesarma were counted and measured the following morning. Sesarma tethering assessed inter-site variation in predation pressure. Sesarma were tethered to braided monofilament fishing line, secured with cyanoacrylic glue, and fastened using metal pins in Sesarma habitat at each site (n = 15 Sesarma/site). Caged crabs to assess tethering artifacts were not deployed because previous studies on Cape Cod and Long Island Sound found no tethering artifacts [5,6]. Tethered Sesarma behave normally on tethers, digging shallow burrows and foraging in the tethered area. Crabs were deployed overnight in August 2013 and mortality was scored the following morning. Predation mortality left broken carapace fragments attached to tethers, mortality from desiccation left dead intact crabs, while surviving tethered crabs were found in newly excavated shallow burrows.

Inter-site variation in predator populations was quantified with baited, commercial 30×60×90 cm traps with funnel openings (n = 3 traps/site) in August 2013. Traps were constructed of 2.5 cm PVC-coated steel mesh and deployed overnight in creeks >100 m apart. Traps have been shown to catch dominant Sesarma predators in the past and thus provide reliable estimates of predatory populations [6]. All individuals caught were identified to species, measured (carapace width for crabs, snout-vent length for fish) and pooled for each site. Multiple regression with leaf tissue percent nitrogen, cordgrass transplant biomass, chalk block dissolution and herbivore intensity was used to examine alternative hypotheses explaining inter-site variation die-off among sites. To examine the hypotheses that site differences in herbivory were being driven by inter-site differences in top predator abundance and predation on Sesarma [6,7] and/or that substrate hardness limits Sesarma burrowing and herbivory, we also performed linear regressions between predator biomass, predation rates on Sesarma, substrate hardness and herbivory. All variables were transformed as needed when necessary. Data was analyzed in JMP 10.
Results

Inter-site variation in die-off intensity ranged from <5% to nearly 98% from ground surveys in 2013. GIS analysis revealed that creek bank die-off (percent linear extent of die-off of available habitat) significantly increased over time ($F_{2,12} = 394.02$, $P < 0.0001$). Die-off was absent in Narragansett Bay in 1997, minimal (7%) in 2003, but by 2012 affected >85% of creek bank habitats (Fig 1). Paralleling the linear spread of die-off, the amount of marsh area also damaged by die-off increased over time ($F_{2,39} = 50.13$, $P < 0.0001$) nearly 3-fold between 2008 and 2012 (Fig 1).

Multiple regression between leaf tissue percent nitrogen, aboveground cordgrass transplant biomass, percent chalk block dissolution, herbivore intensity and extent of die-off revealed that inter-site variation in herbivory explained 73% of the inter-site variation in salt marsh creek bank die-off ($F_{1,8} = 18.49$, $P < 0.01$, Fig 2A), while nitrogen availability, growth potential and wave exposure did not significantly explain any residual inter-site variation in salt marsh die-off ($F_{1,8} = 0.07$, $P > 0.50$, Fig 2B; $F_{1,8} = 0.20$, $P > 0.50$, Fig 2C; $F_{1,9} = 0.17$, $P > 0.50$, Fig 2D, respectively).

We performed a separate polynomial regression between substrate hardness (N) and herbivory to test the hypothesis that substrate hardness dictates crab herbivory. Variation in substrate hardness in vegetated areas along the grazing border explained 46% of variation in salt marsh creek bank die-off ($F_{2,11} = 4.63$, $P < 0.05$, Fig 3).

To test the hypothesis that creek bank die-off is caused by trophic dysfunction due to overfishing, we performed regressions across trophic levels (Fig 4) to test if the previously established trophic cascade in Cape Cod and Long Island Sound marshes had spread to Narragansett Bay [6,7]. We found a positive relationship between predator abundance and predation pressure; predator abundance explained 31% of the inter-site variation in predation pressure ($F_{1,12} = 12.55$, $P < 0.01$, Fig 4A). Predation pressure was negatively associated with Sesarma abundance, explaining 39% of inter-site variation in Sesarma abundance ($F_{1,12} = 7.81$, $P < 0.05$, Fig 4B). Sesarma abundance was positively correlated with herbivory, explaining 30% of the inter-site variation in herbivory across sites ($F_{1,12} = 5.30$, $P < 0.05$, Fig 4C).

Discussion

Our results reveal that the spread of creek bank die-off into Narragansett Bay salt marshes is being driven by predator depletion, which releases herbivorous crabs from consumer control. Additionally, salt marshes with medium substrate hardness are the most vulnerable to die-off. Our results reject
the alternate hypotheses that eutrophication, boat wakes, plant disease and/or pollution are associated with the spread of salt marsh die-off into Narragansett Bay and suggest that they are not drivers of the widespread die-off of western Atlantic salt marshes.

Causation Based on Correlations

While many explanations have been proposed for salt marsh die-off, few have been rigorously tested by well-replicated field experiments. We found no evidence that eutrophication (Fig 2B), pollution or disease affecting plant growing conditions (Fig 2C), or wave exposure (Fig 2D) are directly influencing the spread of salt marsh die-off. None of these proposed die-off mechanisms have been tested experimentally at multiple sites. Based on studies at a single site, Deegan et al. (2012) proposed that eutrophication leads to decreased belowground cordgrass investment, decreasing cordgrass biomass and triggering global salt marsh creek bank collapse. In an identical experiment was independently replicated in Long Island Sound, no shift in biomass allocation or creek bank collapse was found in a 5-year time span [24]. In addition, at Sippiwissett Marsh on Cape Cod, >30 years of nitrogen additions have not led to creek bank collapse [25] and at our Cape Cod study sites we have never seen creek bank die-off without evidence of extensive Sesarma burrowing which increases vulnerability to calving and marsh loss [6,7].

Of all of the hypotheses proposed to cause die-off, herbivory remains the only one that has been widely examined experimentally in well-replicated studies across the Canadian subarctic [3], New England [6,7], Southeastern and Gulf coasts of the United States [4], and the east coast of South America [26]. Our study adds further support to the hypothesis that herbivory is the central driving mechanism of salt-marsh die off. Additionally, our study reveals that herbivory may be mediated by substrate hardness. Substrate hardness determines where runaway herbivory can occur because it limits crab burrowing in hard and soft substrates, leading to peak herbivory in medium hardness substrate creek banks, where burrows can be easily constructed and maintained [16].

Historically, salt marsh ecosystems have been thought to be controlled by physical forces based on correlations between marsh productivity and physical factors [8,9]. Globally, however, near shore food webs have been severely altered by overfishing and predator depletion has heavily impacted seagrass beds [27], kelp forests [28] and coral reefs [29]. Here we have shown that salt marsh ecosystems are also vulnerable to consumer control and consumer mediated salt marsh die-offs are an emerging phenomenon that needs to be addressed.

Management Implications

Our results predict a dire future for New England salt marshes. Salt marshes are one of the most valuable ecosystems on earth per unit area because they provide coastal storm buffering, biochemical processing, carbon sequestration, and nursery habitat ecosystem services [5]. With increasing human population growth in coastal areas, commercial and recreational fishing pressure and predator depletion will inevitably increase [30]. Globally, all salt

Figure 3. Nonlinear polynomial regression relationship between substrate hardness and die-off. Substrate hardness explained 46% of inter-site variation in die-off. Intermediate substrate hardness experiences the highest level of die-off. Soft substrate cannot support burrows while hard substrate is too hard to burrow in. doi:10.1371/journal.pone.0092916.g003

Figure 4. Trophic dysfunction and salt marsh die-off. As predator biomass decreases, predation mortality decreases (A), increasing Sesarma abundance (B), and leading to increased herbivory (C). doi:10.1371/journal.pone.0092916.g004
marsh conservation and management is based on the premise that physical factors exclusively control salt marshes. Our results and those from other marsh systems suggest that consumer control may not simply be a factor to consider in marsh conservation, but with widespread predator depletion impacting near shore habitats globally, trophic dysfunction and run away consumption may be the largest and most urgent management challenge for salt marsh conservation. Rebuilding near shore predatory fish stocks [31] and limiting recreational fishing to catch and release may be the most critical salt marsh conservation measures in saving salt marshes from the impending threat of sea level rise.

References

1. Bertness MD, Silliman BR (2000) Consumer control of salt marshes driven by human disturbance. Conserv Biol 22:618–623.
2. Barbier EB, Hacker SD, Kennedy C, Koch EW, Sier AC et al (2011) The value of estuarine and coastal ecosystem services. Ecol Monogr 81:169–193.
3. Jeffries RL, Jiao AP, Abraham KF (2006) A biotic agent promotes large-scale catastrophic change in the coastal marshes of Hudson Bay. J Eco 94:234–242.
4. Silliman BR, van de Koppel J, Bertness MD, Stanton LE, Mendelssohn IA (2005) Drought, snails, and large-scale die-off of southern US salt marshes. Science 310: 1803–1806.
5. Holdredge C, Bertness MD, Altiere AH (2009) Role of crab herbivory in die-off of New England salt marshes. Conserv Biol 23:672–679.
6. Altiere AH, Bertness MD, Coverdale TC, Herrmann NC, Altiere AH (2012) A trophic cascade triggers collapse of a salt marsh ecosystem with intensive recreational fishing. Ecology 93:1402–1410.
7. Coverdale TC, Herrmann NC, Altiere AH, Bertness MD (2013) Latent impacts: the role of historical human activity in coastal habitat loss. Front Ecol Environ 11:69–74.
8. Odum EP (1971) Fundamentals of Ecology (Saunders, Philadelphia).
9. Mendelssohn IA, Morris JT (2000) Eco-physiological constraints on the primary productivity of Spartina alterniflora. Concepts and Controversies of Tidal Marsh Ecology (Kluwer Academic Publishers, Dordrecht, The Netherlands), pp 59–80.
10. Deegan LA, Johnson DS, Warren RS, Peterson BJ, Fleeger JW et al. (2012) Coastal eutrophication as a driver of salt marsh loss. Nature 490:388–392.
11. Ogburn MB, Alber M (2006) An investigation of salt marsh dieback in Georgia using field transplants. Estuar Coast 29:54–62.
12. Kuch WM, Morris JT (2009) Relationships between Spartina alterniflora and Littoraria irrorata in a South Carolina salt marsh. Wetlands 29:818–825.
13. Alber M, Swenson EM, Adamowicz SC, Mendelssohn IA (2008) Salt marsh dieback: An overview of recent events in the US. Est Coast Shelf Sci 80:1–11.
14. Elmer WH, Usman S, Schneider RW, Marra RE, LaMondia JA et al. (2013) Sudden Vegetation Dieback in Atlantic and Gulf Coast Salt Marshes. Plant Dis 97:436–443.
15. Smith JP, Carullo M (2007) Survey of Potential Marsh Dieback Sites in Coastal Massachusetts (Massachusetts Office of Coastal Zone Management Special report).
16. Bertness MD, Holdredge C, Altiere AH (2009) Substrate mediates consumer control of salt marsh cordgrass on Cape Cod, New England. Ecology 90:2108–2117.

17. Bertness MD, Crain CM, Holdredge C, Sala N (2009) Eutrophication and Consumer Control of New England Salt Marsh Primary Productivity. Conserv Biol 22:131–139.
18. Bertness MD, Esuanchoke PI, Silliman BR (2002), Anthropogenic modification of New England salt marsh landscapes. Proc Natl Acad Sci USA 99:1395–1398.
19. Yund F, Gaines SD, Bertness MD (1994) Cylindrical tube traps for larval sampling. Limnol Oceanogr 36:1167–1177.
20. Bertness MD (1988) Peat accumulation and the success of marsh plants. Ecology 69:703–713.
21. Hines AH (1982) Allometric constraints and variables of reproductive effort in Brachyuran crabs. Mar Biol 69:309–320.
22. Olini EJ, Bishop JM (1983) Variations in total width–weight relationships of blue crabs, Callinectes sapidus, in relation to sex, maturity, molt stage, and carapace form. J Crustac Biol 3:575–581.
23. Wigley SE, McBride HM, McHugh NJ (2003) Length-weight relationships for 74 fish species collected during NEFSC research vessel bottom trawl surveys, 1992–99. NOAA Technical Memorandum NMFS-NE-171 (National Marine Fisheries Service, NOAA, Woods Hole, Massachusetts, USA).
24. Anisfeld SC, Hill TD (2012) Fertilization effects on elevation change and belowground carbon balance in a Long Island Sound tidal marsh. Estuar Coast 35:201–211.
25. Valiela I (1995) Marine Ecological Processes, 2nd Ed. Springer, Verlag. pp 686.
26. Alberti J, Escapa M, Iribarne O, Silliman BR, Bertness MD (2008) Crab herbivory regulates plant facilitative and competitive processes in Argentinean marshes. Ecology 89:155–164.
27. Jackson JBC, Kirby MX, Berger VH, Bjorndal KA, Bonfod LF et al. (2001) Historical overfishing and the recent collapse of coastal ecosystems. Science 293:609–637.
28. Estes JA, Palmisano JF (1974) Sea otters: Their role in structuring nearshore communities. Science 185:1058–1060.
29. Hughes TP (1994) Catastrophes, phase shifts, and large-scale degradation of a Caribbean coral reef. Science 265:1547–1551.
30. Lotze HK, Lenihan HS, Bourque BJ, Bradbury RH, Cooke RG et al. (2006) Depletion, degradation, and recovery potential of estuaries and coastal seas. Science 312:1806–1809.
31. Worm BR, Hilborn R, Baum JK, Branch TA, Collie JS et al. (2009) Rebuilding Global Fisheries. Science 325:378–385.

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Author Contributions

Conceived and designed the experiments: MDB. Performed the experiments: MDB CPB MCB SMC. Analyzed the data: CPB MCB SMC. Contributed reagents/materials/analysis tools: CPB MCB SMC. Wrote the paper: MDB CPB. Edited the manuscript: MDB CPB MCB SMC.