Double-Matched Matrix Decomposition for Multi-View Data

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ABSTRACT
We consider the problem of extracting joint and individual signals from multi-view data, that is, data collected from different sources on matched samples. While existing methods for multi-view data decomposition explore single matching of data by samples, we focus on double-matched multi-view data (matched by both samples and source features). Our motivating example is the miRNA data collected from both primary tumor and normal tissues of the same subjects; the measurements from two tissues are thus matched both by subjects and by miRNAs. Our proposed double-matched matrix decomposition allows us to simultaneously extract joint and individual signals across subjects, as well as joint and individual signals across miRNAs. Our estimation approach takes advantage of double-matching by formulating a new type of optimization problem with explicit row space and column space constraints, for which we develop an efficient iterative algorithm. Numerical studies indicate that taking advantage of double-matching leads to superior signal estimation performance compared to existing multi-view decomposition based on single-matching. We apply our method to miRNA data as well as data from the English Premier League soccer matches and find joint and individual multi-view signals that align with domain-specific knowledge. Supplementary materials for this article are available online.

1. Introduction

Multi-view data (collected on the same samples from multiple views or data sources) are increasingly common with advances in multi-omics and other data collection technologies. In matrix form, each view \( d \) corresponds to a matrix \( X_d \) with \( n \) rows for the matched samples, and \( p_d \) columns for corresponding measurements. While typically the distinct views are only matched by samples, in some cases the views are double-matched: matched by both samples (matched rows) and view features (matched columns). A motivating example is the miRNA data from The Cancer Genome Atlas (TCGA) project collected from both primary tumor and normal tissues of the same subjects; the measurements from two tissues represent two views \( X_1, X_2 \in \mathbb{R}^{n \times p} \) that are matched both by \( n \) subjects and \( p \) miRNAs. Our goal is to extract common (across tissues) as well as individual (tissue-specific) signals from each view, where common/individual signals have two meanings: common/individual signals across subjects, and common/individual signals across miRNAs.

Several methods have been proposed that allow to extract common (joint) structure from the multi-view data. Canonical correlation analysis (Hotelling 1992) seeks linear combinations of features from each view that have maximal correlation. Similarly, partial least squares (PLS) (Rosipal and Krämer 2005) maximizes the covariance, with OnPLS (Löfstedt and Trygg 2011), multiple coindertia analysis (Meng et al. 2014) and interbattery analysis (González Rojas 2016) considering extensions to more than two views. JIVE (Lock et al. 2013) decomposes each view into joint and individual signals, where joint signals are due to matched samples. CIFE (Zhou et al. 2016) and AJIVE (Feng et al. 2018) consider the same joint and individual decomposition as JIVE, however, use a different estimation procedure. Multi-Omics Factor Analysis (MOFA) (Argelaguet et al. 2018, 2020) disentangles common and individual information using group factor analysis with the sparsity structure. DISCO-CCA (Schouteden et al. 2014) uses simultaneous components model (Van Deun and Smilde 2009) with subsequent rotation. iNMF (Yang and Michailidis 2016) is a nonnegative matrix factorization extension of JIVE. SLIDE (Gaynanova and Li 2019) allows for partially-common structures when the number of views is larger than two.

Despite the considerable developments in multi-view data decompositions that extract joint and individual signals, these methods (JIVE, CIFE, AJIVE, MOFA, DISCO-CCA, SLIDE, iNMF) are designed for single-matched multi-view data (matched by samples) rather than double-matched in our motivating example. Applying these methods to double-matched data will lead to extraction of joint signals only in one direction. Let \( X_1, X_2 \in \mathbb{R}^{n \times p} \) be data matrices corresponding to double-matched views, and let \( \hat{A}_1, \hat{A}_2 \in \mathbb{R}^{n \times p} \) be estimated signal matrices obtained by applying one of the existing approaches (e.g., JIVE, CIFE, AJIVE, etc.). Then \( \hat{A}_1 \) and \( \hat{A}_2 \) will have joint signal in their column spaces (corresponding to matched rows), but no joint signal in their row spaces (corresponding to matched columns). A naive approach to estimate joint signal in the row space is to apply the same method to transposed \( X_1^T, X_2^T \in \mathbb{R}^{p \times n} \) leading to \( \hat{A}_1, \hat{A}_2 \in \mathbb{R}^{p \times n} \) with the joint signal corresponding to matched \( p \) features.
However, there is no guarantee that the estimated signals agree with each other, that is in general $\hat{A}_d \neq \tilde{A}_d$, which we confirm in our simulation studies (Section 3.2). Furthermore, some signal rank estimations methods, for example, permutation approach in Lock et al. (2013) or bi-cross-validation approach in Gaynanova and Li (2019), can lead to different estimated ranks for the same $X_1, X_2$ depending on whether the matching by rows or the matching by columns is used (Section 3.1).

Several methods consider the problem of extracting signal from double-matched multi-view data. Population value decomposition (Crainiceanu et al. 2011) is an extension of singular value decomposition to double-matched data, however, it only allows to extract joint signal, and does not extract individual signal. Similarly, 3-way PCA (Wold et al. 1987) and tensor decompositions (Zhou et al. 2016) extract joint signals, but not individual. Linked matrix factorization (O’Connell and Lock 2019) and bidimensional integrative factorization (Park and Michailidis 2016; Feng et al. 2018) are based on separating the signal matrix into joint and individual parts with respect to the matched dimension, that is $A_k = I_k + I_k$. For example, in JIVE (Lock et al. 2013; Zhou et al. 2016; Feng et al. 2018), the joint matrices $J_1$ and $J_2$ share the same column space, that is, $C(J_1) = C(J_2) = C(J)$. The individual matrices $I_1$ and $I_2$ are orthogonal to the joint space and have zero intersection of their respective column spaces, that is, $C(I_j) \perp C(I_k) \cap \{0\}$, and identify parts of the signal that are joint/individual across row dimension $n$ (samples) as well as parts of the signal that are joint/individual across column dimension $p$ (features).

Existing methods for estimation of $A_k$ in single-matched multi-view data (Lock et al. 2013; Zhou et al. 2016; Yang and Michailidis 2016; Feng et al. 2018; Gaynanova and Li 2019) are designed for the case where each pair of views is either matched by rows or by columns, but not simultaneously by both as in our motivating example.

In this work, we propose the double-matched matrix decomposition (DMMD) for multi-view data that allows to extract joint and individual signals in both row and column directions simultaneously, in contrast to existing approaches. First, we prove that DMMD decomposition exists, and characterize conditions for its uniqueness. Second, we propose an estimation approach that takes advantage of the fact that the signal matrices must coincide whether the joint and individual signals are considered in the row direction, or in the column direction. We pose this estimation as a new type of optimization problem with explicit row space and column space constraints, for which we develop an efficient iterative algorithm. Third, we show that DMMD has superior signal estimation performance compared to existing methods for single-matched data even when underlying true signal ranks are known (Section 3.2), thus, confirming the advantage of taking into account double-matched structure in estimation.

The rest of the article is organized as follows. In Section 2, we formulate the proposed double-matched matrix decomposition, and derive an algorithm for its estimation. In Section 3, we compare DMMD to existing methods on simulated data. In Section 4, we illustrate DMMD on the double-matched miRNA data from TCGA, and double-matched English Premier league soccer match data. In Section 5 we conclude with discussion.

2. Method

2.1. Notation

For a matrix $A \in \mathbb{R}^{n \times p}$, we let $A^T$ be its transpose, $C(A)$ be its column space and $R(A)$ be its row space. We use $\|A\|_F = \sqrt{\sum_{i=1}^n \sum_{j=1}^p a_{ij}^2}$ to denote its Frobenius norm. For two matrices $A_1 \in \mathbb{R}^{n \times p_1}$ and $A_2 \in \mathbb{R}^{n \times p_2}$, we write $[A_1, A_2] \in \mathbb{R}^{n \times (p_1 + p_2)}$ to denote the column-wise concatenation. We say $C(A_1)$ is orthogonal to $C(A_2)$ if for any vector $x_1 \in C(A_1)$ and any vector $x_2 \in C(A_2)$, it holds that $x_1 \perp x_2$. We use $\text{Id}$ to denote an identity matrix. We use $e_i = (0, \ldots, 0, 1, 0, \ldots, 0)^T$ with only the $i$-th element being one to denote the standard basis vector.

We use script-style letter $\mathcal{U}$ to denote a vector space formed by the matrix $U$ with columns corresponding to orthonormal basis vectors, $C(U) = \mathcal{U}$.

2.2. Model

We consider two double-matched data matrices $X_1 \in \mathbb{R}^{n \times p}$ and $X_2 \in \mathbb{R}^{n \times p}$. We assume additive decomposition $X_k = A_k + E_k$, $k = 1, 2$, where $A_k$ is the signal matrix and $E_k$ is the noise matrix. We further assume each signal matrix $A_k$ is low-rank, which is common in the literature (Udell and Townsend 2019). Our goal is to estimate $A_k$ from $X_k$, and identify parts of the signal that are joint/individual across row dimension $n$ (samples) as well as parts of the signal that are joint/individual across column dimension $p$ (features).

Existing methods for estimation of $A_k$ in single-matched multi-view data (Lock et al. 2013; Zhou et al. 2016; Yang and Michailidis 2016; Feng et al. 2018; Gaynanova and Li 2019) are based on separating the signal matrix into joint and individual parts with respect to the matched dimension, that is $A_k = I_k + I_k$. For example, in JIVE (Lock et al. 2013; Zhou et al. 2016; Feng et al. 2018), the joint matrices $J_1$ and $J_2$ share the same column space, that is, $C(J_1) = C(J_2) = C(J)$. The individual matrices $I_1$ and $I_2$ are orthogonal to the joint space and have zero intersection of their respective column spaces, that is, $C(I_j) \perp C(I_k) \cap \{0\}$, and identify parts of the signal that are joint/individual across row dimension $n$ (samples) as well as parts of the signal that are joint/individual across column dimension $p$ (features).

Our proposal is based on the observation that for double-matched signal matrices $A_k$, the JIVE decomposition must hold with respect to both dimensions (row and column) simultaneously. We formalize this observation in the following lemma, which is a generalization of Lemma 1 from Feng et al. (2018).

**Lemma 1.** Given two signal matrices $A_1, A_2 \in \mathbb{R}^{n \times p}$, there are unique sets of matrices $[I_{1\times1}, I_{2\times2}], [I_{1\times1}, I_{2\times2}], [I_{1\times1}, I_{2\times2}]$ and $[I_{1\times1}, I_{2\times2}]$ such that

1. $A_k = I_k + I_k = I_k + I_k$, $k = 1, 2$
2. $C(I_k) = M \subset C(A_k), \quad k = 1, 2$
3. $R(I_k) = N \subset R(A_k), \quad k = 1, 2$
4. $M \perp C(I_k), N \perp C(I_k), \quad k = 1, 2$
5. $C(I_{1\times1}) \cap C(I_{2\times2}) = \{0\}, R(I_{1\times1}) \cap R(I_{2\times2}) = \{0\}$

Here $M$ represents the joint column structure (common signal for $n$ samples) and $N$ represents joint row structure (common signal for $p$ features) of the signal matrices $[A_1, A_2]$. Similarly, $I_{1\times1}$ and $I_{2\times2}$ represent the individual column signals and individual row signals, respectively. Lemma 1 applies to double-matched matrices $[A_1, \ldots, A_k]$ from more than two views ($K \geq 2$); we only present case $K = 2$ as it is sufficient for motivating datasets.

In light of Lemma 1, we consider the following double-matched matrix decomposition (DMMD) for observed $X_1 \in \mathbb{R}^{n \times p}$ and $X_2 \in \mathbb{R}^{n \times p}$

$$X_k = J_k + I_k + E_k = J_k + I_k + E_k, \quad k = 1, 2;$$

(1)
Two double-matched matrices are decomposed into joint structure, individual structures and noise in both row and column directions according to DMMD model (1), here $n = 80$, $p = 40$, $\text{rank}(A_1) = 15$, $\text{rank}(A_2) = 12$, $\text{rank}(M) = 7$ and $\text{rank}(N) = 5$.

Figure 1. Summary of the proposed estimation approach for fitting DMMD model (1).

Step 1: Estimate proxy signals. Estimate the total ranks of $A_1$ and $A_2$, and construct proxy signal matrices $Z_1$ and $Z_2$ from $X_1$ and $X_2$ given those ranks.

Step 2: Estimate joint structure. Use proxy signals $Z_1$ and $Z_2$ to estimate basis vectors of $M$ (joint column structure) and $N$ (joint row structure).

Step 3: Estimate signals with given joint structure. Fit model (1) conditionally on the estimated $M$, $N$ from step 2 and estimated total ranks from step 1.

Figure 2 shows the flow chart summarizing DMMD estimation steps. In Section S5 of the supplementary materials, we describe a variation of Step 3 that allows for iterative updates of initial $M$, $N$ from Step 2 leading to iterative DMMD (DMMD-i). Numerically, the two approaches are very similar.

2.3. Estimation

To fit model (1), we propose the following estimation approach:
but DMMD-i has a significantly higher computational cost (Section 3).

2.3.1. Estimation of Proxy Signals
We estimate proxy low-rank signal matrices $Z_k$ from observed $X_k$ using low-rank singular value decomposition of $X_k$ (Jha and Yadava 2010). We propose to use the profile likelihood approach (Zhu and Ghodsi 2006) to estimate the total rank of the signal.

Let $d_1 \geq d_2 \geq \ldots \geq d_m$ be the ordered singular values of matrix $X_k$, where $m = \min(n, p)$. Given a fixed $q$ with $1 \leq q \leq m$, define sets $D_1 = \{d_1, d_2, \ldots, d_q\}$ and $D_2 = \{d_{q+1}, \ldots, d_m\}$. Zhu and Ghodsi (2006) assume that the elements of $D_1$ and $D_2$ come from the normal distributions $N(\mu_1, \sigma^2_1)$ and $N(\mu_2, \sigma^2_2)$, respectively. Let $f(\cdot; \mu, \sigma^2)$ be the probability density function of $N(\mu, \sigma^2)$. Then the log-likelihood is

$$l(q, \mu_1, \mu_2, \sigma^2) = \sum_{i=1}^{q} \log f(d_i; \mu_1, \sigma^2_1) + \sum_{j=q+1}^{m} \log f(d_j; \mu_2, \sigma^2_2).$$

Given $q$, the MLEs are $\hat{\mu}_1 = \sum_{i=1}^{q} d_i/q, \hat{\mu}_2 = \sum_{j=q+1}^{m} d_j/(m-q)$ and $\hat{\sigma}^2 = [(q-1)s^2_1 + (m-q-1)s^2_2]/m$, where $s^2_1$ and $s^2_2$ are the sample variances of elements in $D_1$ and $D_2$, respectively. We estimate the rank of signal $A_i$ by maximizing the profile likelihood over $q$ and set $r_1 = \hat{q}$, where $\hat{q}$ is the maximizer. The same approach is used for $X_2$. Given $r_k$, we obtain proxy signal matrix $Z_k$ by corresponding rank-$r_k$ SVD of observed $X_k$.

**Remark 1.** We use profile likelihood approach for rank estimation as it is fast and performs well in our simulations, however, an alternative rank estimation approach can be used in this step. Some examples are permutation method (Lock et al. 2013), edge distribution method (Onatski 2010) and Bi-Cross-Validation method (Owen and Perry 2009). We compare these approaches in simulations in Section 3.1.

2.3.2. Estimation of Joint Structure
In this section we estimate the joint column structure $M$ and the joint row structure $N$ based on the proxy signals $Z_1$ and $Z_2$.

From the proof of Lemma 1, the joint column structure $M = C(A_1) \cap C(A_2)$ and the joint row structure $N = R(A_1) \cap R(A_2)$. Thus, a naive way to estimate $M$ is to consider the intersection of column spaces of proxy signals $Z_1$ and $Z_2$, $C(Z_1) \cap C(Z_2)$, however, $Z_k$ is only an estimate of $A_k$. Thus, in practice $C(Z_1) \cap C(Z_2) = \{0\}$ due to the corruption of true joint structure by noise. To circumvent this difficulty, we propose to use principal angles to measure the similarity between $C(Z_1)$ and $C(Z_2)$. Both CCA (Avron et al. 2013) and inter-battery factor analysis (González Rojas 2016) also use the cosines of principal angles to measure similarity. We propose to separate the principal angles into two groups: small angles indicating common signals (albeit not exactly equal) and large angles indicating individual signals. Similar idea is used in Feng et al. (2018), however, our approach for determining the angle cutoff is different.

We first review principal angles. Let $\mathcal{U}$ and $\mathcal{V}$ be subspaces with $\dim(\mathcal{U}) = h_1, \dim(\mathcal{V}) = h_2$ in $\mathbb{R}^n$. Let $h = \min(h_1, h_2)$, then the principal angles $\Theta(\mathcal{U}, \mathcal{V}) = \{\theta_k \in [0, \frac{\pi}{2}] | k = 1, 2, \ldots, h\}$ between $\mathcal{U}$ and $\mathcal{V}$ are recursively defined by

$$\cos \theta_k = \max_{x \in \mathcal{U}, y \in \mathcal{V}} |x^T y| = |x_k^T y_k|$$

subject to $\|x\| = \|y\| = 1, x^T x = 0, y^T y = 0, i = 1, 2, \ldots, k - 1$.

The vectors $\{x_1, \ldots, x_h\}$ and $\{y_1, \ldots, y_h\}$ are called principal vectors. Principal angles can be calculated using singular value decomposition (Knyazev and Argentati 2002). Let $X \in \mathbb{R}^{n \times h_1}$ and $Y \in \mathbb{R}^{n \times h_2}$ be the orthogonal matrices formed by concatenating orthonormal basis vectors of $\mathcal{U}$ and $\mathcal{V}$ column-wise, respectively. Let SVD of $X^T Y$ be $USV^T$ where $U \in \mathbb{R}^{h_1 \times h_1}$ and $V \in \mathbb{R}^{h_2 \times h_2}$ be the orthogonal matrices. Using principal angles, we estimate joint column structure $M$ from $Z_1$ (with rank $r_1$) and $Z_2$ (with rank $r_2$) as follows.

First we calculate the principal angles $\theta_1, \ldots, \theta_l, l = \min(r_1, r_2)$, and principal vectors $\{u_1, \ldots, u_l\}$ and $\{v_1, \ldots, v_l\}$ between two column spaces $C(Z_1)$ and $C(Z_2)$. Since the joint rank could be 0 or $l$, we add artificial $\theta_0 = 0$ and $\theta_{l+1} = \pi/2$ angles into the principal angle vector. We then separate the angles into two groups by using profile likelihood as described in Section 2.3.1 to estimate the optimal cutoff $\hat{q}$. The estimated joint column rank is then $r_c = \hat{q} - 1$. We estimate the basis for joint column structure $M$ by calculating the element-wise average of principal vectors, for example, $w_i = \frac{1}{l} (u_i + v_i), i = 1, 2, \ldots, r_c$, corresponding to the smallest $r_c$ principal angles. The basis of joint row structure $N$ together with its rank $r_c$ is determined similarly from the row spaces $R(Z_1)$ and $R(Z_2)$. Let $M$ and $N$ be the matrices containing averaged principal vectors corresponding to $M$ and $N$, respectively. To form basis vectors, we orthogonalize $M$ and $N$ using Gram-Schmidt process. The full procedure is summarized in Figure 3.

**Remark 2.** An alternative rank estimation approach can be used in this step. Some examples are permutation method (Lock et al. 2013), Bi-Cross-Validation method (Owen and Perry 2009) and Wedin bound method (Feng et al. 2018). See Section 3.1 for comparison.

2.3.3. Estimation of Signals with Given Joint Structure
We consider the estimation of the signals $A_k$, $k = 1, 2$, in (1) given the joint structures $M$ and $N$. Our goal is to find the closest matrix to $X_k$ that simultaneously contains both given joint column structure $M$ and given joint row structure $N$, that is to solve

$$\min_{A_k \in \mathbb{R}^{n \times h}} \|X_k - A_k\|_F^2$$

subject to $C(M) \subset C(A_k), C(N) \subset R(A_k), \text{rank}(A_k) = r_k, k = 1, 2$.

where $r_k$ is the total rank for signal $A_k$ as in Section 2.3.1, $M$ is the joint column space with $r_c$ basis in $\mathbb{R}^{n \times r}$ and $N$ is the joint row space with $r_c$ basis in $\mathbb{R}^n$ estimated as in Section 2.3.2. In Section S5 of the supplementary materials, we describe a variation of (2) with additional minimization over joint structures that allows
Figure 3. Procedure of calculating joint structure in DMMD.

for iterative updates of initial M, N, leading to iterative DMMD (DMMD-i). We compare DMMD and DMMD-i in Section 3.

To solve (2), we first consider a simplified problem by removing the row-space constraint, for which we derive a closed-form solution.

Lemma 2. Given X ∈ ℝⁿˣᵖ, M ∈ ℝⁿˣᵣᵣ with orthonormal columns with rank(MMᵀX) = rₓ, and rank r with rₓ ≤ r ≤ min(n, p), consider

$$\text{minimize}_{A \in \mathbb{R}^{n \times p}} \|X - A\|_F^2$$

such that $C(M) \subseteq C(A), \text{ rank}(A) = r$. Let $A_M^* = MMᵀX + RRᵀX$, where the columns of R are the first $r - rₓ$ left singular vectors of (Id - MMᵀ)X. Then $A_M^*$ is the global minimizer of (3). Furthermore, if matrix (Id - MMᵀ)X has distinct $(r - rₓ)$ th and $(r - rₓ + 1)$ th singular values, then $A_M^*$ is the unique minimizer of (3).

From Lemma 2, the columns of matrix [M, R] are the basis vectors of the column-space of the solution $A_M^*$ to (3). Similarly, consider a simplified problem (2) with row-space constraint, but no column-space constraint.

Lemma 3. Given $X \in \mathbb{R}^{n \times p}$, $N \in \mathbb{R}^{p \times r}$ with orthonormal columns with rank(XNᵀN) = rₓ, and rank $r$ with $rₓ ≤ r ≤ \text{min}(n, p)$, consider

$$\text{minimize}_{A \in \mathbb{R}^{n \times p}} \|X - A\|_F^2$$

such that $C(N) \subseteq C(A), \text{ rank}(A) = r$. Let $A_N^* = XNᵀX + XSᵀS$, where the columns of S are the first $r - rₓ$ right singular vectors of X(Id - NNᵀ). Then $A_N^*$ is the global minimizer of (4). Furthermore, if matrix X(Id - NNᵀ) has distinct $(r - rₓ)$ th and $(r - rₓ + 1)$ th singular values, then $A_N^*$ is the unique minimizer of (4).

Algorithm 1 Iterative algorithm for (2)

1: Given: $X_k \in \mathbb{R}^{n \times p}$, $r_k$, $k = 1, 2$, $M \in \mathbb{R}^{n \times r}$, $N \in \mathbb{R}^{p \times r}$, $t_{max}, \epsilon > 0$
2: for $k = 1, 2$ do
3: SVD: $(\mathbf{Id} - MMᵀ)X_k = U_kD_kV_kᵀ$
4: $R_k(0) \leftarrow$ first $r_k - r_x$ columns of $U_k$
5: $\tilde{M}_k(0) \leftarrow [M, R_k(0)]$
6: $t \leftarrow 0$
7: while $t \neq t_{max}$ and $|X_k^{(t)} - L_k^{(t-1)}| > \epsilon$ do
8: SVD: $\tilde{M}_k^{(t)}(\tilde{M}_k^{(t)})ᵀX_k(\mathbf{Id} - NNᵀ) = U_k^{(t)}D_k^{(t)}V_k^{(t)}ᵀ$
9: $\tilde{S}_k^{(t+1)} \leftarrow$ first $r_k - r_x$ columns of $V_k^{(t)}$
10: $S_k^{(t+1)} \leftarrow [N, S_k^{(t+1)}]$  
11: SVD: $(\mathbf{Id} - MMᵀ)X_k\tilde{N}_k^{(t+1)}\tilde{N}_k^{(t+1)ᵀ} = U_k^{(t)}D_k^{(t)}V_k^{(t)}ᵀ$
12: $R_k^{(t+1)} \leftarrow$ first $r_k - r_x$ columns of $U_k^{(t+1)}$
13: $\tilde{M}_k^{(t+1)} \leftarrow [M, R_k^{(t+1)}]$
14: $t \leftarrow t + 1$
15: $L_k^{(t)} = \|X_k - \tilde{M}_k^{(t)}\tilde{M}_k^{(t)ᵀ}X_k\tilde{N}_k^{(t)}\tilde{N}_k^{(t)ᵀ}\|_F^2$
16: end while
17: return $A_k^* = \tilde{M}_k^{(t)}\tilde{M}_k^{(t)ᵀ}X_k\tilde{N}_k^{(t)}\tilde{N}_k^{(t)ᵀ}$
18: end for

The columns of matrix [N, S] are the basis vectors of the row-space of the solution $A_N^*$ to (4). Given the closed form solutions to (3) and (4), we propose an iterative algorithm for the full problem (2), where we alternate the update of column space with the update of the row space. The full algorithm is summarized in Algorithm 1. In words, we first initialize the full column space $M_0$ of each signal matrix, and update the row space that is not captured by $N$. Given the updated full row space of each signal matrix $\tilde{N}_k$, we then update the column space that is not captured by $M$. At each step, the current estimated signal matrix is $A_k^{(t)} = \tilde{M}_k^{(t)}\tilde{M}_k^{(t)ᵀ}X_k\tilde{N}_k^{(t)}\tilde{N}_k^{(t)ᵀ}$, which is feasible as long as it has full rank $r_k$ (this is always satisfied in our numerical studies because of noisy $X_k$). Once the estimated signal matrices $A_k$ are obtained, by construction it holds that for $k = 1, 2$: $J_{ck} = MMᵀA_k$, $I_{ck} = (\mathbf{Id} - MMᵀ)A_k$. Similarly, $J_k = A_kNNᵀ$, $I_k = A_k(\mathbf{Id} - NNᵀ)$.
Let $L_k^{(t)}$ be the objective function of (2) at iteration step $t$, 
$$L_k^{(t)} = L_k(R_k^{(t)}, S_k^{(t)}) = \|X_k - A_k^{(t)}\|_F^2 = \|X_k - (MM^T + R_k^{(t)}R_k^{(t)T})X_k(NN^T + S_k^{(t)}S_k^{(t)T})\|_F^2.$$ We show that Algorithm 1 is guaranteed to converge as it leads to nonincreasing sequence of $L_k^{(t)}$.

**Proposition 1.** If at each iteration step $t$ in Algorithm 1, $(MM^T + R_k^{(t)}R_k^{(t)T})X_k(\text{Id} - NN^T)$ is of rank at least $r_k - r_r$ and $(\text{Id} - MM^T)X_k(NN^T + S_k^{(t)}S_k^{(t)T})$ is of rank at least $r_k - r_e$, then the sequence of objective values $L_k^{(t)}$ is nonincreasing.

Proposition 1 only guarantees the convergence of objective values, but in practice we found that the sequences of $R_k^{(t)}$ and $S_k^{(t)}$ also converge. The convergence to the global minimizer is not guaranteed since problem (2) is nonconvex, thus, the output may depend on initial $R_k^{(0)}$. The proposed $R_k^{(0)}$ corresponds to global solution when $r_r = 0$ due to Lemma 2. Empirically this choice leads to a smaller objective value at convergence compared to a random $R_k^{(0)}$, and excellent signal estimation performance.

### 3. Simulation Studies

We generate the signal matrices $A_k \in \mathbb{R}^{n \times p}$ given the sample size $n$, the number of features $p$, the total signal ranks $r_k \leq \min(n, p)$, $k = 1, 2$, the rank of joint column structure $r_c \leq \min(r_1, r_2)$ and the rank of joint row structure $r_r \leq \min(r_1, r_2)$ in accordance with Lemma 1 (Section S3 of the supplementary materials). We then set $X_k = A_k + E_k$, where $E_k$ has independent entries $e_{kij} \sim \mathcal{N}(0, \sigma_k^2)$, $i \in [1, \ldots, n]$, $j \in [1, \ldots, p]$. We define the signal to noise ratio as
$$\text{SNR} = \frac{\|A_k\|_F^2}{E_k(\|E_k\|_F^2)} = \frac{r_k}{n \sigma_k^2},$$ and choose $\sigma_k$ to control the SNR at prespecified levels.

#### 3.1. Rank Estimation

We investigate the performance of profile likelihood (PL) method from Sections 2.3.1–2.3.2 on estimating the total signal ranks $r_k$, the joint column rank $r_c$ and the joint row rank $r_r$. We compare with permutation method used in rjive package (O’Connell and Lock 2016) and Bi-Cross-Validation (BCV) method (Owen and Perry 2009) implemented in SLIDE package (Gaynanova and Yuan 2021). JIVE and SLIDE rank selection methods are applied in two ways: (i) column space decomposition based on matched rows (samples); (ii) row space decomposition based on matched columns (features). When we perform JIVE on matched rows, we denote it as JIVE (Row), similarly for SLIDE. For total rank estimation, we also consider edge distribution (ED) method (Onatski 2010). We implement ED method in R ourselves by translating Python code from Shu, Wang, and Zhu (2020). For joint rank estimation, we also consider the Wedin threshold method (Feng et al. 2018) as implemented in ajive package (Carmichael 2021). The Wedin method is applied with the given true total ranks rather than estimated total ranks as the latter is not implemented in ajive.

We consider three settings, with 140 replications for each.

**Setting 1** $n = 240$, $p = 200$, $\text{SNR} = 1$, $r_1, r_2$ sampled from $\{2, 3, \ldots, 20\}$ with replacement and $r_c, r_r$ sampled from $\{1, 2, \ldots, \min(r_1, r_2), 5\}$ with replacement.

**Setting 2** Same as Setting 1 with $\text{SNR} = 0.5$.

**Setting 3** $n = 240$, $p = 200$, $\text{SNR} = 1$, $r_1, r_2$ sampled from $\{2, 3, \ldots, 20\}$ with replacement. In the first 35 replications, $r_c, r_r = 0$. In the next 35 replications, $r_c = 0, r_r = \min(r_1, r_2)$. In the third 35 replications, $r_c = \min(r_1, r_2), r_r = 0$. For the last 35 replications, $r_c = r_r = \min(r_1, r_2)$. This setting is used to demonstrate cases where there is either no joint structure or no individual structure.

Figure 4(a) displays the difference between the estimated total rank and the true rank $r_k$ for each method in Setting 1. ED works the best, followed by the proposed PL. The permutation approach in rjive works poorly in this setting, and is also not consistent (different ranks are estimated depending on whether the matching is done by rows or by columns). SLIDE rank estimation based on BCV also works poorly, however, this is likely due to automatic centering implemented in the package which will perturb the column-space of the true noncentered signal. Figure 4(b) displays the difference between the estimated...
joint rank and the true joint rank (either \( r_c \) or \( r_r \)). The Wedin bound method works perfectly, however, it uses the knowledge of true total ranks. The proposed PL works as well as Wedin bound without such knowledge with the exception of two cases. Both methods are significantly more accurate compared to other approaches. The results in Setting 2 are qualitatively similar to results in Setting 1 (Section S3 of the supplementary materials), however, the performance tends to be worse due to lower SNR. On total rank estimation, the median performance of ED and PL is still superior to the permutation method used in \( \cdot \cdot \cdot \) (Ahn and Horenstein 2013), and this assumption is satisfied in all of our settings. Since in practice this assumption may be violated, we use PL method as default. On joint rank estimation, Wedin bound method works perfectly in all of the settings, however, it does so by using true total ranks. Since in practice the true ranks are unknown, and PL works similarly, we use PL as default. We reach the same conclusions in a high-dimensional setting that mimics TCGA dataset in Section 4.1 (Section S3 of the supplementary materials).

3.2. Signal Identification

We investigate the performance of DMMD on estimating signals \( A_k \) in model (1) if the true ranks are known. We also consider DMMD-i (Section S5 of the supplementary materials) which allows iterative adjustment of initial \( M \) and \( N \) from Step 2. We measure the performance as

\[
\text{Relative Error}(\hat{A}_k, A_k) = \frac{||\hat{A}_k - A_k||_F^2}{||A_k||_F^2}.
\]

We also measure the relative error separately on joint \( I_{jk}, J_{jk} \) and individual \( I_{jk}, I_{j} \) in model (1). We compare with JIVE (Lock et al. 2013), SLIDE (Gaynanova and Li 2019) and AJIVE (Feng et al. 2018) using the same implementation as in Section 3.1. AJIVE, JIVE and SLIDE are fitted in two ways: (i) column space decomposition based on matched rows (samples); (ii) row space decomposition based on matched columns (features). We use JIVE (Row) to indicate model based on matched rows, and similarly JIVE (Column).
We consider three settings with 140 replications for each.

Setting 4  \[ n = 240, p = 200, r_1 = 20, r_2 = 18, r_c = 4, r_r = 3, \] 
SNR = 1.

Setting 5  Same as Setting 4 with SNR = 0.5.

Setting 6  \[ n = 240, p = 200, \] 
SNR = 1, \( r_1 = 20, r_2 = 18. \) In the first 35 replications, \( r_c, r_r = 0. \) In the next 35 replications, \( r_c = 0, r_r = \min(r_1, r_2) = r_2. \) In the third 35 replications, \( r_c = \min(r_1, r_2) = r_2, r_r = 0. \) For the last 35 replications, \( r_c = r_r = \min(r_1, r_2) = r_2. \) Like Setting 3 in Section 3.1, this setting is used to demonstrate cases where there is either no joint structure, or no individual structure.

All DMMD, JIVE, AJIVE, and SLIDE use true total ranks \( r_k, \) true joint rank \( r_c \) (for column decomposition) and true joint rank \( r_r \) (for row decomposition), \( k = 1,2, \) as input. Ranks misspecifications are investigated in Section S3 of the supplementary materials, where DMMD performs best in total signal estimation.

Figure 6(a) and (b) show relative errors of all methods in Setting 4 for estimated signals based on matched rows (\( J_{rk}, I_{rk} \)) and matched columns (\( J_{rk}, I_{rk} \)), respectively. The errors for total signal are the same for DMMD as it enforces model (1). In contrast, the errors for JIVE, AJIVE, and SLIDE depend on matching (by rows or by columns) as it affects the estimated signal. For joint signals, DMMD and SLIDE perform similar, and are both more accurate than JIVE and AJIVE. DMMD has the smallest errors on full signals and individual signals in all scenarios, confirming that taking into account double matching leads to more accurate signal estimation. The same conclusion holds in Setting 5 with smaller SNR (see Section S2 of the supplementary materials).

In Setting 6, either joint signal matrix or individual signal matrix is exactly equal to zero, thus, we use the absolute error, \( \| \text{Estimated Signal} - \text{True Signal} \|^2_F, \) rather than the relative error to measure the performance. Figure 7(a) and (b) show absolute errors of the four methods based on column space decomposition due to matched rows (\( J_{rk}, I_{rk} \)), or row space decomposition due to matched columns (\( J_{rk}, I_{rk} \)), respectively. When both joint column and row structures are absent (\( r_c = r_r = 0 \)), all methods perform the same, which is expected as the estimation is completely separate across two views. When \( r_c = r_r = r_2, \) DMMD gives smallest errors as it takes advantage of double matching. When \( r_c = r_r = 0, \) JIVE and SLIDE work better than DMMD on estimating joint row structure, whereas when \( r_c = r_r = r_2, \) they work better than DMMD on estimating joint column structure. A possible explanation for this is a different approach for estimating the joint structures used by the methods. DMMD uses element-wise averaging of pairs of basis vectors from each view with smallest principal angles as in AJIVE (Feng et al. 2018), whereas JIVE and SLIDE extract basis vectors from concatenated matrix of view-specific residuals after subtracting individual structures.

Overall, we find that DMMD has the smallest signal estimation error, with DMMD-i being slightly better than DMMD. This remains true in a high-dimensional setting that mimics TCGA dataset in Section 4.1 (Section S3 of the supplementary materials). While in Setting 6 JIVE and SLIDE sometimes lead to better performance, these cases correspond to absent individual structures in either row or column directions and absent joint structures in the other directions, which is rarely the case for real data. When individual structures are present, DMMD always leads to improved errors as it enforces equality in estimated total signals from row and column decomposition of double-matched data, which subsequently leads to more accurate estimation of individual structures, and consequently, of the total signal.

Computationally, DMMD is significantly faster than the competitors; its total run time (rank estimation plus signal identification) on data with \( n = 100, p = 800 \) is 24 sec on Intel(R) Core(TM) i5-7300U CPU @ 2.60GHz. As expected, DMMD-i comes with a significantly higher computational cost taking 618 sec on the same data. More details are in Section S3 of the supplementary materials. Given the relatively small improvement of DMMD-i over DMMD, in practice we recommend to use DMMD with large datasets for computational efficiency.
4. Application

4.1. Application to TCGA Data

We consider data from The Cancer Genome Atlas (TCGA) repository corresponding to the Breast Invasive Carcinoma (BRCA) cancer type. We use TCGA-Assembler 2 software pipeline (Wei et al. 2018) to obtain miRNA read counts corresponding to the primary tumor tissue (view 1) and to the normal tissue (view 2) of the same subjects. We log-transform the counts to account for skewness, and remove the samples and features with zero variance for both views. We then apply double-standardization as in Efron (2012) so that all rows and columns in each matrix have mean zero and sample variance one; such processing of multi-view data is also used in Risk and Gaynanova (2021). The double-standardization makes the estimated decomposition mean and scale-invariant. The final double-matched $X_1$ (primary tumor tissue) and $X_2$ (normal tissue) each contain $p = 734$ matched miRNAs from $n = 87$ matched samples. To evaluate possible biological relevance of obtained decomposition, we match each sample with one of the cancer subtypes: Luminal A (LumA), Luminal B (LumB), Basal-like (Basal), HER2-enriched (H) obtained from https://www.cbioportal.org/study/clinicalData?id=brca_tcga_pub. For 46 out of 87 subjects there are missing clinical records which are denoted as unknown.

Our goal is to extract common (across tissues) as well as individual (tissue-specific) signals from each view, where common/individual signals have two meanings: (i) across subjects, and (ii) across miRNAs. Here we consider two methods: JIVE (rank selection via permutation test with subsequent fitting of the JIVE model) and the proposed DMMDD (rank selection via profile likelihood with subsequent fitting of model (1)). For JIVE, we consider both matching by subjects (column space decomposition), and matching by miRNAs (row space decomposition).

First, we compare the estimated ranks. Permutation approach used by JIVE leads to inconsistent total ranks as the estimates depend on the type of matching: matching by samples leads to $\hat{r}_1 = 11, \hat{r}_2 = 9$, whereas matching by miRNAs leads to $\hat{r}_1 = 14, \hat{r}_2 = 11$. The PL method gives smaller estimated ranks $\hat{r}_1 = 8, \hat{r}_2 = 6$. Despite the discrepancy in total ranks between JIVE and DMMDD, both lead to the same estimated joint ranks with $\hat{r}_c = 0$ and $\hat{r}_r = 2$.

Next, we compare the variance explained by each method, together with the variance explained separately by joint/individual parts of the estimated signal. Figure 8 shows the percent variance explained by each part of the estimated decomposition separately for tumor and normal tissues. The total variance explained by estimated signal (joint plus individual) is the same for DMMDD regardless of the type of matching considered, whereas it changes for JIVE due to discrepancy in estimated signals. The overall variance explained is higher for JIVE as it estimates higher total ranks compared to DMMDD. Both DMMDD and JIVE show that the variance explained by joint structure is higher for normal tissue compared to primary tumor tissue. We believe that this is in agreement with what would be expected from biological knowledge since tumor tissue evolves from originally normal tissue, and becomes more heterogeneous as cancer develops.

We next display the found joint row structure ($\hat{r}_r = 2$) corresponding to matched miRNAs in Figure 9(a) and (b). An alternative vertical alignment of these heatmaps is in Figure S3, supplementary materials. The order of samples and miRNAs is the same in both tissues, and are determined based on the hierarchical clustering of the joint structure of the primary tumor tissue. Visually, the joint row structure captures the division of miRNAs in three clusters. While the displayed cluster partition is based on tumor tissue, the heatmap of normal tissue in Figure 9(b) has block structure based on the same partition.
To provide further interpretation of estimated decomposition, we next consider individual structures from DMMD with respect to matched subjects. Figure 10(a) displays the heatmaps of estimated $I_{ck} \in \mathbb{R}^{p \times n}$, $k = 1, 2$, from model (1) with samples sorted according to subtype information. Here rank($I_{c1}$) = 8 and rank($I_{c2}$) = 6. To better visualize the corresponding individual column spaces, Figure 10(b) shows the leading left singular vectors of $I_{c1}$ and $I_{c2}$, respectively, which are the leading basis vectors for corresponding individual column spaces. For the primary tumor tissue, the basis vector displays a strong contrast between the Basal and LumA subtypes, expresses 27.3% of the whole variation in individual structure. The effect of this basis vector can be seen in the whole signal heatmap in Figure 10(a), where the contrast in the same direction is observed roughly in the top half of the miRNAs in primary tumor tissue, and is observed in the opposite direction in the bottom half of the miRNAs. In contrast, the individual structure for normal tissue does not display this contrast. Furthermore, the leading basis vector for the individual structure of normal tissues does not appear to separate any of the cancer subtypes, which is in agreement with what would be expected from biological knowledge since it captures individual subjects structure in normal tissues that is not present in primary tumor tissue.

4.2. Application to Soccer Data

We consider data from soccer matches in the English Premier League obtained from https://www.kaggle.com/kenzeng24/premier-league-matches. Each row of data represents a soccer match played in the English Premier League, and each column represents a unique feature recorded for that match (e.g., Date, Home Team, Full Time Goals for each team, etc.). First, we filter the data by removing the matches with data quality issues (decimal values recorded for the number of yellow cards or red cards), and removing the matches corresponding to draw games. Second, we determine the winning and losing team for each of the matches, and extract 10 match statistics recorded for each team representing numbers of full-time goals, half-time goals, shots, shots on target, hit woodwork, corners, fouls, offsides, yellow cards, and red cards. To aid interpretation of extracted signals in terms of original untransformed match statistics, we do not apply double standardization from Section 4.1 to soccer.
Individual column (matched subjects) structures extracted by DMMD for primary tumor and normal tissues from double-matched TCGA-BRCA miRNA data. The samples are ordered according to the cancer subtype. The top row shows full individual signals, whereas the bottom row shows the leading individual basis vectors along with the percentage of variance explained (relative to the full individual signal).

**Table 1.** Joint row basis for winning and losing teams in English Premier League when $r_1 = r_2 = r_r = 1$.

| Signal         | Full time goals | Half time goals | Shots | Shots on target | Hit woodwork | Corners | Foul committed | Offsides | Yellow cards | Red cards |
|----------------|-----------------|-----------------|-------|-----------------|--------------|---------|----------------|----------|---------------|-----------|
| Joint          | 1.00            | 0.45            | 8.08  | 3.90            | 0.23         | 4.03    | 9.80           | 2.50     | 1.10          | 0.07      |

**Table 2.** Joint row basis and individual row basis for winning teams in English Premier League when $r_1 = 2, r_2 = r_r = 1$.

| Signal         | Full time goals | Half time goals | Shots | Shots on target | Hit woodwork | Corners | Foul committed | Offsides | Yellow cards | Red cards |
|----------------|-----------------|-----------------|-------|-----------------|--------------|---------|----------------|----------|---------------|-----------|
| Joint          | 1.00            | 0.47            | 7.85  | 3.78            | 0.21         | 4.01    | 11.35          | 2.76     | 1.30          | 0.08      |
| Win            | 1.00            | 0.33            | 5.09  | 2.93            | 0.21         | 1.62    | -4.98          | -0.39    | -0.88         | -0.08     |

In the end, we obtain two double-matched 558 × 10 matrices, one for the winning team and one for the losing team, where each row corresponds to a match, and each column corresponds to a team statistic from the match. Our goal is to investigate the relationship between match statistics that are (i) common across teams, and (ii) individual to the winning team.

First, we estimate the ranks of underlying signals. Both PL and ED automatically select $\hat{r}_1 = \hat{r}_2 = 1$, and PL determines $\hat{r}_c = \hat{r}_r = 1$, which is one of the special cases considered in simulation setting 6. DMMD works best in that setting, so we only report DMMD results. Table 1 displays the coefficient of the extracted row basis vector normalized to have coefficient 1 for Full time goals to assist interpretation. As this basis vector corresponds to joint structure across both winning and losing teams, we conclude that in English Premier League there is on average 1 goal for every 8.08 shots in the game. Also, on average, there are more goals in the second half game compared to the first half game.

The total ranks estimated by PL and ED are quite low due to low $p = 10$. ED inherently restricts the signal rank to be at most $0.1p$, and thus, cannot possibly estimate larger ranks for these data. PL relies on clustering singular values in two groups, which we suspect is less reliable when the number of singular values is small. Thus, here we consider an alternative approach to rank estimation, where for each team we pick the rank to explain 90% of the variation in the respective dataset. This approach leads to $\hat{r}_1 = 2, \hat{r}_2 = 1, \hat{r}_r = 1$. Thus, there is a rank 1 individual structure in winning team not present in the losing team. Table 2 displays the coefficients for the joint row basis vector, and the individual row basis vector for winning team as estimated by DMMD. Both vectors are normalized to have coefficient 1 corresponding to full time goals to assist interpretation. As in the previous analyses, there are approximately eight shots for every goal, and approximately four corners for every goal. However, the winning team tends to have a higher number of goals per shots while simultaneously having fewer fouls, offside, yellow cards and red cards. The coefficient for offside may appear
counter-intuitive, however, it can be interpreted as the attack of
winning team being less interrupted. Somewhat surprisingly,
the number of hitting woodwork does not seem to affect the
winning or losing conditions.

5. Discussion

We propose a new decomposition for multi-view data with
matched rows and columns, which we call DMMD. The main
novelty of our approach is in taking advantage of double-
matching property via explicit column and row space con-
straints in signal estimation, and in deriving the corresponding
optimization algorithm. The algorithm relies on estimated joint
column and row spaces; the proposed estimation works well
numerically but can be improved using iterative DMMD at the
expense of a significantly higher computational cost. While our
exposition has been limited to the case of two views, DMMD
can be applied to more views as Lemma 1, model (1), estimation
of proxy signals in Section 2.3.1 and the Algorithm 1 still
hold. Estimation of joint structures in Section 2.3.2 requires
modification for more than two views, which we outline in
Section S6 of the supplementary materials.

The method requires estimation of ranks corresponding to
different parts of the decomposition, and our empirical studies
indicate that the chosen profile likelihood (PL) approach for
rank estimation is competitive compared to alternative rank
estimation methods. However, we also found that PL can occa-
sionally severely overestimate the ranks, while other methods
tend to underestimate the ranks. To help identify whether severe
rank overestimation is an issue in practice, we recommend to
simultaneously consider several rank estimation approaches
(like we did in Section 4) to verify that the ranks estimated by
PL are not considerably higher than the ranks estimated using
other methods.

Several extensions of DMMD are of interest. First, general-
ization to more than two views described in Section S6 of the
supplementary materials allows to apply DMMD to longitudinal
data, albeit the method will treat all the time points inter-
changeably. An alternative approach for longitudinal data is to treat the
view at starting time as base view, and apply DMMD for each
consecutive view paired with the base view. This would lead to
a sequence of joint and individual structures ordered by time,
which could provide additional insights on the time/treatment
effects. It would be of interest to pursue such analyses in the
future. Second, DMMD is designed for fully observed data, thus,
imputation is required if some elements of $X_j$ are missing. As
the computations rely only on the leading singular vectors and
singular values, one possibility is to adapt soft-impute algorithm
(Mazumder, Hastie, and Tibshirani 2010) for DMMD. Third,
DMMD decomposition is not sparse, however, sparse exten-
sions can be pursued by replacing standard SVD in Algorithm 1
with its sparse analogs, for example, sparse PCA method of
Shen and Huang (2008) or penalized orthogonal iteration of
Jung, Ahn, and Jeon (2019). This, however, would increase
computational time, and create additional challenge of choosing
appropriate sparsity tuning parameters. Finally, while DMMD is
based on the matrix models, an alternative approach is to view
double-matched data as a three-way tensor and consider tensor
decompositions (Zhou et al. 2016). It is unclear, however, how to
extract individual information from the latter. Furthermore, we
find that existing tensor decompositions may have difficulties in
capturing joint row and column structures simultaneously
(Section S7 of the supplementary materials). It would be of
interest to investigate extensions of tensor decompositions that
will provide more flexibility as well as preserve the matrix inter-
pretation of joint and individual structures.

Supplementary Materials

Supplementary: Proofs of all statements, and additional numerical results
(.pdf file). The R code is available at https://github.com/justicesuker/
DMMD_Code.

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References

Ahn, S. C., and Horeinstein, A. R. (2013), “Eigenvalue Ratio Test for the
Number of Factors,” Econometrica, 81, 1203–1227. [1120]
Argelaguet, R., Arnol, D., Bredikhin, D., Deloro, Y., Velten, B., Marioni, J. C.,
and Stegle, O. (2020), “Mofa+: A Statistical Framework for Comprehen-
sive Integration of Multi-Modal Single-Cell Data,” Genome Biology, 21,
1–17. [1114]
Argelaguet, R., Velten, B., Arnol, D., Dietrich, S., Zenz, T., Marioni, J. C.,
Buettner, F., Huber, W., and Stegle, O. (2018), “Multi-Omics Factor Analy-
sis: Framework for Unsupervised Integration of Multi-Omics Data Sets,”
Molecular Systems Biology, 14, e8124. [1114]
Avron, H., Boutsidis, C., Toledo, S., and Zouzias, A. (2013), “Efficient
Dimensionality Reduction for Canonical Correlation Analysis,” in Inter-
national Conference on Machine Learning, pp. 347–355. PMLR. [1117]
Carmichael, I. (2021), “ajive: Angle Based Joint and Individual Variation
Explained,” R package version 0.1.0. Available at https://github.com/idc9/
r_jive. [1119]
Crainiceanu, C. M., Caffo, B. S., Luo, S., Zipunnikov, V. M., and Punjabi,
N. M. (2011), “Population Value Decomposition, a Framework for the
Analysis of Image Populations,” Journal of the American Statistical
Association, 106, 775–790. [1115]
Efron, B. (2012), Large-Scale Inference: Empirical Bayes Methods for Estima-
tion, Testing, and Prediction (Vol. 1). Cambridge: Cambridge University
Press. [1122]
Feng, Q., Jiang, M., Hannig, J., and Marron, J. (2018), “Angle-Based Joint
and Individual Variation Explained,” Journal of Multivariate Analysis,
166, 241–265, [1114,1115,1117,1119,1120,1121]
Gaynanova, I., and Li, G. (2019), “Structural Learning and Integra-
tive Decomposition of Multi-View Data,” Biometrics, 75, 1121–1132.
[1114,1115,1120,1121]
Gaynanova, I., and Yuan, D. (2021), “SLIDE: Structural Learning and
Integrative Decomposition of Multi-View Data,” R package version 1.0.
Available at https://github.com/irinagain/SLIDE. [1119]
Gonzalez Rojas, V. (2016), “Inter-Battery Factor Analysis via pls: The
Missing Data Case,” Revista Colombiana de Estadistica, 39, 247–266.
[1114,1117]
Hotelling, H. (1992), "Relations Between Two Sets of Variates," in Breakthroughs in Statistics, eds. S. Kotz, N. L. Johnson, pp. 162–190, New York: Springer. [1114]

Jha, S. K., and Yadava, R. (2010), "Denoising by Singular Value Decomposition and its Application to Electronic Nose Data Processing," IEEE Sensors Journal, 11, 35–44. [1117]

Jung, S., Ahn, J., and Jeon, Y. (2019), "Penalized Orthogonal Iteration for Sparse Estimation of Generalized Eigenvalue Problem," Journal of Computational and Graphical Statistics, 28, 710–721. [1125]

Knyazev, A. V., and Argentati, M. E. (2002), "Principal Angles Between Subspaces in an a-based Scalar Product: Algorithms and Perturbation Estimates," SIAM Journal on Scientific Computing, 23, 2008–2040. [1117]

Lock, E. F., Hoadley, K. A., Marron, J. S., and Nobel, A. B. (2013), "Joint and Individual Variation Explained (jive) for Integrated Analysis of Multiple Data Types," The Annals of Applied Statistics, 7, 523–542. [1114,1115,1117,1120]

Onatski, A. (2010), "Determining the Number of Factors from Empirical Distribution of Eigenvalues," The Review of Economics and Statistics, 92, 1004–1016. [1117,1119]

Owen, A. B., and Perry, P. O. (2009), "Bi-Cross-Validation of the SVD and Nonnegative Matrix Factorization," The Annals of Applied Statistics, 3, 564–594. [1117,1119]

Risk, B. B., and Gaynanova, I. (2021), "Simultaneous Non-Gaussian Component Analysis (Sing) for Data Integration in Neuroimaging," The Annals of Applied Statistics, 15, 1431–1454. [1122]

Rosipal, R., and Krämer, N. (2005), "Overview and Recent Advances in Partial Least Squares," in International Statistical and Optimization Perspectives Workshop’ Subspace, Latent Structure and Feature Selection’, pp. 34–51, Springer. [1114]

Zhu, M., and Ghodsi, A. (2006), "Automatic Dimensionality Selection from the Scree Plot via the Use of Profile Likelihood," Computational Statistics & Data Analysis, 51, 918–930. [1117]