A single dose ChAdOx1 nCoV-19 vaccine elicits high antibody responses in individuals with prior SARS-CoV-2 infection comparable to that of double dose vaccinated SARS-CoV-2 infection naïve individuals

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Abstract

Background

A single dose COVID-19 vaccines, mostly mRNA-based vaccines, are shown to induce robust antibody responses in individuals who were previously infected with SARS-CoV-2, suggesting the sufficiency of a single dose to those individuals. However, these important data are limited to developed nations and lacking in resource-limited countries, like Ethiopia.

Methods

We compared receptor-binding domain (RBD)-specific IgG antibodies in 40 SARS-CoV-2 naïve participants and 25 participants previously infected with SARS-CoV-2, who received two doses of ChAdOx1 nCoV-19 vaccine. We measured the antibody response in post-vaccination blood samples from both groups of participants collected at four different post-vaccination time points: 8- and 12-weeks after each dose of the vaccine administration using an in-house developed ELISA.

Results

We observed a high level of anti-RBD IgG antibodies titers 8-weeks after a single dose administration (16/27; 59.3%) among naïve participants, albeit dropped significantly (p<0.05) two months later, suggesting the protective immunity elicited by the first dose ChAdOx1 nCoV-19 vaccine will likely last for a minimum of three months. However, as expected, a significant (p<0.001) increase in the level of anti-RBD IgG antibodies titers was observed after the second dose administration in all naïve participants. By contrast, the ChAdOx1 nCoV-19 vaccine-induced anti-RBD IgG antibody titers produced by the P.I participants at 8- to 12-weeks post-single dose vaccination were found to be similar to the antibody titers seen after a two-dose vaccination course among infection- naïve participants and showed no significant (p>0.05) increment following the second dose administration.

Conclusion

Taken together, our findings show that a single ChAdOx1 nCoV-19 dose in previously SARS-CoV-2 infected individuals elicits similar antibody responses to that of double dose vaccinated naïve individuals. Age and sex were not associated with the level of vaccine-elicited immune responses in both individuals with and without prior SARS-CoV-2 infection. Further studies are required to assess the need for a booster dose to extend the duration and amplitude of the specific protective immune response in Ethiopia settings, especially following the Omicron pandemic.

1. Background
COVID-19 continues to be a major public health concern, causing severe illness and deaths in Ethiopia and the rest of the world alike. Mass vaccination against SARS-CoV-2 is the most effective public health intervention to protect against morbidity and mortality related to SARS-CoV-2 infection [1]. Safe, efficacious, and licensed COVID-19 vaccines, including ChAdOx1 nCoV-19 (AZD1222; Oxford–AstraZeneca) are available [2–5], despite being challenged by the recurrent emergence of new SARS-CoV-2 variants. Real-world vaccine efficacy studies from developed countries have shown that the current vaccines are able to generate effective humoral and cellular immunity, albeit differential responses are observed between vaccine-induced immunity and hybrid (vaccine-induced immunity combined with natural infection) immunity [6, 7]. Several correlates of protection studies have demonstrated that higher antibody titers are associated with decreased risk of subsequent symptomatic SARS-CoV-2 infection [8–12], and several studies from developed countries have revealed the rapid waning of antibody levels among SARS-COV-2 infection naïve vaccine recipients compared to those individuals with hybrid immunity [7, 13–16]. Despite the importance of immune durability data for guiding national vaccination strategies, there is a dearth of studies from Ethiopia and other African countries looking at more locally relevant populations.

The ChAdOx1 nCoV-19 vaccine utilizes a replication-deficient adenoviral vector that induces expression of SARS-CoV-2 spike (S) protein in host cells, particularly in the skeletal muscle [17]. Vaccinated individuals subsequently generate antibodies against the spike protein, including those that target the receptor-binding domain (RBD), which contains many neutralizing epitopes. However, they do not generate antibodies against other SARS-CoV-2 structural and non-structural proteins, such as nucleocapsid (N) [18]. Studies have shown a strong correlation between anti-RBD IgG titers and SARS-CoV-2 neutralizing titers [15]. Therefore, in resource-limited countries, it is advantageous to use anti-RBD IgG testing as a proxy for virus neutralization to assess the protection offered by the ChAdOx1 nCoV-19 vaccine.

As part of the strategy to evaluate the Ethiopian national COVID-19 response through vaccination, we established a longitudinal cohort of healthcare professionals working at the Armauer Hansen Research Institute (AHRI), with and without evidence of prior SARS-CoV-2 infection and determined their levels of ChAdOx1 nCoV-19 vaccine-induced anti-RBD IgG titers across four-time points. The present study generated evidence of the duration of ChAdOx1 nCoV-19 vaccine-induced humoral responses and the long-term effect of prior SARS-CoV-2 infection on subsequent vaccine-induced responses.

2. Methods

2.1 Study Design and Participants

We conducted a longitudinal prospective study constituting healthcare professionals from AHRI, who were also one of the priority target recipients of the ChAdOx1 nCoV-19 vaccine. Vaccination was offered through the Ethiopian Ministry of Health national COVID-19 vaccination campaign. In this analysis, only participants who were vaccinated with the ChAdOx1 nCoV-19 vaccine were included. The study protocol
was reviewed and approved by the AHRI/ALERT Ethics Review Committee (PO/32/20) and only voluntary participants and who were able to give written informed consent were included in the study. About 5ml of venous blood was collected from a total of 65 (n=26 female and n=39 male) participants, aged 24-59 years (mean of 38.1 ± standard deviation of 8.36) before vaccination and four subsequent post-vaccination follow-ups (Figure 1) to monitor humoral responses to the vaccine. The first (F1) and second follow-ups (F2) were conducted at 8-and 12-weeks after the first dose, respectively, whereas the third (F3) and fourth (F4) follow-ups were done at 4 and 8 weeks after the second dose, respectively (Figure 1). All participants received their first dose between March 23, 2021 and March 31, 2021. The interval between the first and the second doses was 12 weeks (range 83-97 days). Participants were then stratified into two groups based on previous exposure to SARS-CoV-2 before vaccination and hereafter denoted as naïve and P.I. (Figure 1). The two groups: naïve (n=40) and P.I. (n=25) contained similar distributions of age and sex (Figure 1 and Table 1). Table 1 summarizes the demographics of each group of study participants. In addition, participants completed a questionnaire at each visit regarding their history of RT-PCR confirmed SARS-CoV-2 infection.
Table 1
Baseline characteristics and serostatus of study participants (n = 65) and longitudinal humoral immune response

| Character | n^a | Mean (SD^b) | % | GMTs^c |
|-----------|-----|-------------|---|---------|
| Age in years | 38.1 (8.4) | | | |
| <40 | | 58.5 | | |
| >=40 | | 41.5 | | |
| Sex | | | | |
| Male | 29 | 60.0 | | |
| Female | 39 | 40.0 | | |
| Naïve^d | 25 | 61.5 | | |
| BL^e | | 200.00 | | |
| F1^f | 27 | 2267.68 | | |
| F2^g | 25 | 699.73 | | |
| F3^h | 7 | 7186.10 | | |
| F4^i | 14 | 5966.97 | | |
| P.I.^j | 40 | 38.5 | | |
| BL | 25 | 1671.88 | | |
| F1 | 15 | 6783.12 | | |
| F2 | 18 | 4219.28 | | |
| F3 | 11 | 7972.68 | | |
| F4 | 10 | 5542.56 | | |

^a n=number participants; ^b SD=standard deviation; ^c GMTs=geometric mean titers; ^e BL=baseline or prevaccination; ^f F1= 8-weeks after the first vaccination; ^g F2=12-weeks after the first dose; ^h F3 = 8-weeks after the second vaccination; ^i F4 =8-weeks after the second vaccination; ^d naïve= participants without previous exposure to SARS-CoV-2 infection; ^j P.I.= participants with previously infected with SARS-CoV-2 infection.

2.2 ELISA Methods:
Prior to analyzing the presence of anti-RBD IgG antibodies, each serum sample was treated with Triton X-100 at the final concentrations of 1.0% and incubated at room temperature (RT) for 30 minutes. This procedure was performed aiming to reduce risk from any potential virus in serum [19]. Detection of anti-RBD IgG antibodies in the sera was done using a validated in-house ELISA as described previously [20]. To determine the end titer in seropositive serum samples, a two-fold serial dilution starting at 1:200 in a 96-well ELISA plate was done. The end titer was defined as a serum dilution at which the observed optical density (OD) at 450 nm reads matched to the OD\textsubscript{450} readout for pre-COVID-19 serum sample diluted at 1:200 and included in each ELISA run as a negative control. Inter-assay variability was normalized by including a convalescent serum in each run of titration ELISA.

Since the magnitude and durability of SARS-CoV-2 antibody response has been shown to positively correlate with the severity level of COVID-19 [14, 21] and yet we did not have clinical records of our P. I. participants. Thus, a convalescent sera panel (CSP; n=15) was collected from COVID-19 recovered individuals, three-months after being discharged from the Saint Paulo's Millennium College Hospital in 2020 (before the rollout of the vaccines) were included in the anti-RBD IgG antibodies titers analysis. Based on the level of anti-RBD IgG titers in these CSP as a reference, we inferred whether our P.I. participants have had symptomatic or asymptomatic SARS-CoV-2 infection.

2.3 Statistical analysis

GraphPad Prism version 8.0 for Windows (GraphPad Software, La Jolla California USA) was used for statistical analyses. We measured anti-RBD IgG end titers ranging from 1:200 to 1:51200. Graphs were plotted using log10-transformed anti-RBD IgG end titers values. Table 1 comprised the calculated geometric means (GM) titers. To compare the mean differences of anti-RBD IgG-antibodies titers across each time point of serum collection, we used the unpaired non-parametrical t-test with p <0.05 =*; p< 0.01=**; p<0.001=***; p<0.0001****). Because some participants dropped out and others joined, the number of blood samples provided after the first dose varied among participants enrolled at baseline (Figure 1 and Table 1). Participants who did not provide any post-vaccine samples were excluded from the analysis.

3. Results

Post-vaccination seropositivity

Figure 1b shows the number of participants who provided a blood sample for antibody test at each post-vaccination follow-up. While detectable anti-RBD IgG antibodies 8-weeks post-first dose were measured in 16 (59.3%) naïve and 14 P.I. (87.5%) participants, we detected in 15 (60.0%) naïve and 16 (88.9%) P.I. participants at 12-weeks post-first dose (Figure 2b). At 8-weeks post-second dose, anti-RBD IgG antibodies were detected in serum samples from all the 17 (7 naïve and 10 P.I.) participants that provided blood samples. At 12-weeks post-second dose, all 24 (14 naïve and 10 P.I) participants had detectable anti-RBD IgG antibodies. Notably, four naïve (2 males with the age of 36 and 59 and 2 females aged 32 and 45 years) participants had remained seronegative at 12-weeks post-first dose. On the other hand, 2
Comparison of anti-RBD IgG antibody titers

We first compared the pre-vaccination anti-RBD IgG titers level in P.I. participants with that of the titers in CSP. A significantly (p-value <0.0001) higher levels of anti-RBD IgG titers were observed among recovered COVID-19 patients 3-months after natural infection than in those P.I. participants (Fig. 2a).

We also compared the levels of anti-RBD IgG titers between naïve and P.I. participants across the four post-vaccination follow-ups. P.I. had a four-fold increase at 8-weeks post-first dose compared to their pre-vaccination anti-RBD IgG antibodies levels (p=0.0004; Table 1). We also observed higher anti-RBD IgG titer levels in naïve participants at 8-weeks post-first dose, yet rapidly (30%) declined at 12-weeks post-first dose (p<0.05; Figure 2c). By contrast, high and sustainable post-first dose antibody levels were observed in P.I. participants and were found to be comparable to those naïve participants post-second dose. This trend remained consistent when results were stratified by sex and age (Figure 3).

As shown in Figure 2c, we observed a significant (p-value <0.01) increase in the level of anti-RBD IgG antibodies in naïve participants 8-weeks post-second dose compared to the level of the titer that was observed 12-weeks post-first dose. On contrary, we did not observe a statistically significant (p-value >0.5) difference in the level of anti-RBD IgG antibodies titers in P.I. participants from those observed post-single dose (Figure 2d). Unlike the decline shown 12-weeks post-first dose, we did not see a significant (p-value >0.5) decline in mean anti-RBD IgG antibodies titers 12-weeks post-second dose in both naïve and P.I. participants (Table 1 and Figure 2b).

Figure 3 compares the levels of anti-RBD IgG antibody titers for different age groups and sex across the four post-vaccination time points. We observed, no statistically significant difference (p-value >0.5) in the level of anti-RBD IgG titers between age 21-40 years and age 40-59 years (Figure 3a and 3b) and between males and females (Figure 3c and 3d) irrespective of participants being naïve or P.I.

4. Discussion

Immune protection following either vaccination or natural infection with SARS-CoV-2 decreases overtime [22]. Although the minimum antibody titer that correlates with protection has not yet been established, a decreased antibody titer is shown to be associated with an increased risk of subsequent symptomatic SARS-CoV-2 infection [8, 10–12]. In the present study, immunologically naïve participants had relatively comparable anti-SARS-CoV-2 RBD IgG responses at 8-weeks post-first dose ChAdOx1 nCoV-19 with those of participants with previous exposure to SARS-CoV-2. This finding is consistent with other studies on mRNA-based vaccines [7, 13]. Notably, at three months post-single-dose, the level of anti-RBD IgG antibodies elicited by a single dose ChAdOx1 nCoV-19 vaccine dropped significantly in individuals who were not previously infected with SARS-CoV-2. This is consistent with the suggestion made by the UK Joint Committee on Vaccination and Immunization (JCVI) -that is protective immunity elicited by the first
dose ChAdOx1 nCoV-19 vaccine will likely last for a duration of 12 weeks [23]. As reported elsewhere [24], we also noted that four of our SARS-CoV-2 infection naïve participants experienced a delay in generating anti-RBD Ig antibodies. Unexpectedly, we also noted seroconversion in 2 males PI participants aged 30 and 42 years who had higher anti-RBD IgG antibody titers 8-weeks post-first dose, indicating evidence of a rare event of rapid waning of humoral response in single-dose vaccinated individuals with prior SARS-CoV-2 infection. This could be due to critical medical conditions such as immunosuppression, though in our study, such medical conditions were not systematically recorded. Similarly, seroreversion was observed after receiving two hepatitis A vaccination in HIV-positive patients [25].

As expected, individuals who naturally contracted SARS-CoV-2 prior to vaccination developed a more rapid and sustained response to the ChAdOx1 vaccine than immunologically naïve individuals. The vaccine-induced anti-RBD IgG antibody titers produced by participants with prior SARS-CoV-2 infection at three months after a single dose vaccination appeared to be comparable to the antibody titers levels seen after a two-doses for infection-naïve participants. Similar findings have been previously reported for the ChAdOx1 nCoV-19 [26] and BNT162b2 vaccines [7, 14, 15]. Similarly, SARS-CoV-2 immunologically naïve individuals developed relatively higher and durable anti-RBB IgG antibody titers after the second dose vaccination. However, we did not observe a statistically significant difference in antibody response after second dose between participants with and without evidence of previous SARS-CoV-2 infection. Our findings, along with other studies [6, 15, 27, 28] suggest a single vaccine dose in previously infected individuals elicits similar antibody responses to that of double dose vaccination.

Participants with SARS-CoV-2 antibodies at baseline before the first vaccine injection, regardless of their sex and age (ranged 21-59), have developed strong anti-RBD IgG antibodies to the COVID-19 vaccine with no statistically significant variability between the first and the second dose. Given the age range (21-59 years with an average of 38.1 years) of our participants, this finding not surprising; however, an age-dependent decreasing pattern of anti-RBD IgG antibodies titers was reported across similar age groups to our study [29, 30]. On the other hand, the absence of statistically significant antibody titers difference between male and female participants is surprising and inconsistent with previous reports [27, 30] yet agrees with the finding reported by Wheeler et al. [31] and Lee et al.[32].

Our findings are the first evidence from Ethiopia supporting the superiority of “hybrid immunity” in eliciting a strong immunity against SARS-CoV-2 infection, comparable to that of two doses of the ChAdOx1 nCoV-19 vaccine in infection naïve individuals. Recent studies have demonstrated that sera from mRNA-vaccinated individuals with prior infection provide broader cross-neutralizing antibodies against several SARS-COV-2 variants, including Delta variants [33]. Consistent with this, another recent study revealed the mechanism by which hybrid immunity improves B cells and antibodies against SARS-CoV-2 variants[34]. However, a study revealed that the anti-spike antibody titers of BNT162b2 recipients were remarkably higher than those of ChAdOx1 nCoV-19 recipients [13]. Additionally, recent studies showed increased risk of SARS-CoV-2 Omicron infection in both vaccinated and previously infected individuals through evasion of vaccine- or infection-induced immune response and suggested the need of the rapidly developing new, Omicron variant-specific vaccine (refs). It also remains to be determined if the
hybrid immunity elicited by the ChAdOx1 nCoV-19 vaccine will effectively protect vaccinees from the subsequent infections with different SARS-CoV-2 spike variants. Thus, further studies that aim to find a correlation between the level of ChAdOx1 nCoV-19 or any vaccine-induced antibody titer and clinical outcome are required.

5. Conclusions

Our findings demonstrate that administrating only one dose vaccine to individuals previously infected with SARS-CoV-2 infection could elicit antibody response comparable to that of two doses of ChAdOx1 nCoV-19 vaccinated SARS-CoV-2 naïve infection and an effective strategy to stretch the supply of vaccines. Age and sex were not associated with the level of vaccine and hybrid immune responses. However, our findings should not be translated to the Omicron variant as our study was not designed to determine the effectiveness of vaccine-elicited antibodies against Omicron. Thus further studies that compare neutralization of Omicron by sera of 1-dose vaccinated individuals with prior infection versus 2-dose vaccinated, 3-dose vaccinated naive individuals and vaccinated individuals with Omicron breakthrough infection are required to assess the need of third booster dose over Omicron-specific booster dose in Ethiopia settings especially following the Omicron pandemic.

This study is limited the by number of participant dropouts, lack of neutralization assays, lack of T-cell data and by our limitation to employ N-protein-based antibody test verification for indirect evidence of breakthrough infections.

Declarations

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Authors’ contributions

TG designed experiments, supervised laboratory testing, analyzed data, and interpreted results, discussed data, and wrote the original manuscript – critically revised and edited. TG, AM, AMi, and AA conceived the study. TG, AM, MA, LW, TAB, FG, AMi, and AA contributed reagents. TG, AM, MA, MT, FT, AMi, AA
contributed to the recruitment of participants. TAB, LW, and FGT critically revised the manuscript. DF, AyA, GT were responsible for laboratory analysis of blood samples under the supervision of TG. FT collected blood samples and FT and MT contributed to clinical data acquisition. TG, AM, AMi and AA prepared the study protocol. TAB, FGT, and AA contributed to funding acquisition. AM. MA, TAB, LW, AGB, FGT, AMi, and AA revised and edited the manuscript. AMi and AA are senior authors. All authors read and approved the final version of the manuscript.

**Competing Interests**

The authors declare no competing interests.

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Figures
Study design with the timeline for vaccination and sample collection. (b) Schematic representation of number participants at baseline and at four postvaccination time points. Light red color character (Naïve) represents participants with SARS-CoV-2 immunologically naïve and blue color (P.I.) represents participants with likely previous SARS-CoV-2 infection; BL = baseline or prevaccination; F1 = 8-weeks after the first dose; F2 = 12-weeks after the first dose; F3 = 8-weeks after the second dose; and F4 = 8-weeks after the second dose. The first dose and second dose were given to participants at BL and F3, respectively prior to blood sample collection.
Figure 2

Analysis of ChAdOx1 nCoV-19 vaccine-induced antibody response in naïve and previously infected (P.I.) participants profile at different time points. Comparison of anti-RBD IgG titers profile (a) between P.I. participants (red dots) before they received vaccination and convalescent serum panel (CSP: blue dots); (b) between naïve (blue dots) and P.I. (red dots) participants across different time points before and after vaccination; among naïve participants (c) and (d) P. I. participants across different points.

CSP = convalescent sera panel included as a reference to indirectly assess whether P.I. participants had
previously either asymptomatic or symptomatic SARS-CoV-2 infection. Each colored dot corresponds to an individual participant. Horizontal bars represent mean with 95% CI of anti-RBD IgG titer levels (transformed to Log_{10} value) within the indicated groups. The broken line denotes the assay detection limit. Unpaired non-parametrical t-test with p <0.05 =*; p < 0.01 =**; p < 0.001 =***; p < 0.0001 =****, ns=non-significant) was used to compare the mean differences of anti-RBD IgG-antibodies titers across each time point of serum collection. BL = baseline or prevaccination; F1 = 8-weeks after the first dose; F2 = 12-weeks after the first dose; F3 = 8-weeks after the second dose; and F4 = 8-weeks after the second dose. The first dose and second dose were given to participants at BL and F3, respectively prior to blood sample collection.

Figure 3
Antibody response by age and sex of naïve and previously infected (P.I.) participants following vaccination. Antibody response comparison of (a) naïve and (b) P.I. participants by age: 40-59 (blue dots) versus 21-39 years (red dots); Antibody response comparison of (c) naïve and (d) P.I. participants by sex: male (blue dots) versus female (red dots). For panel C male vs female comparison was not done at F3 since there was infection naïve female participants that provided blood sample at F3.