Widespread subcortical grey matter degeneration in primary lateral sclerosis: a multimodal imaging study with genetic profiling

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\textbf{ABSTRACT}

\textit{Background:} Primary lateral sclerosis (PLS) is a low incidence motor neuron disease which carries a markedly better prognosis than amyotrophic lateral sclerosis (ALS). Despite sporadic reports of extra-motor symptoms, PLS is widely regarded as a pure upper motor neuron disorder. The post mortem literature of PLS is strikingly sparse and very little is known of subcortical grey matter pathology in this condition.

\textit{Methods:} A prospective imaging study was undertaken with 33 PLS patients, 117 healthy controls and 100 ALS patients to specifically assess the integrity of subcortical grey matter structures and determine whether PLS and ALS have divergent thalamic, hippocampal and basal ganglia signatures. Volumetric, morphometric, segmentation and vertex-wise analyses were carried out in the three study groups to evaluate the integrity of thalamus, hippocampus, caudate, amygdala, pallidum, putamen and accumbens nucleus in each hemisphere. The hippocampus was further parcellated to characterise the involvement of specific subfields.

\textit{Results:} Considerable thalamic, caudate, and hippocampal atrophy was detected in PLS based on both volumetric and vertex analyses. Significant volume reductions were also detected in the accumbens nuclei. Hippocampal atrophy in PLS was dominated by dentate gyrus, hippocampal tail and CA4 subfield volume reductions. The morphometric comparison of ALS and PLS cohorts revealed preferential medial bi-thalamic pathology in PLS compared to the predominant putaminal degeneration detected in ALS. Another distinguishing feature between ALS and PLS was the preferential atrophy of the amygdala in ALS.

\textit{Conclusions:} PLS is associated with considerable subcortical grey matter degeneration and due to the extensive extra-motor involvement, it should no longer be regarded a pure upper motor neuron disorder. Given its unique pathological features and a clinical course which differs considerably from ALS, dedicated research studies and disease-specific therapeutic strategies are urgently required in PLS.

\textbf{Glossary}

ALS: Amyotrophic Lateral sclerosis, ANCOVA: Analysis of covariance, BG: Basal ganglia, C9orf72: chromosome 9 open reading frame 72, CA: Cornu ammonis, FOV: Field of view, FWE: Familywise error, GC-ML-GD: Molecular and granule cell layer of the dentate gyrus, GM: Grey matter, HATA: Hippocampal-amygdala transition area, HC: Healthy control, HSP: Hereditary Spastic Paraplegia, IR-SPGR: Inversion Recovery prepared Spoiled Gradient Recalled echo, LMN: Lower motor neuron, M: Mean, MNI152: Montreal Neurological Institute 152 standard space, PBA: Pseudobulbar affect, PCL: Pathological crying and laughing, PLS: Primary lateral sclerosis, pTDP-43: Phosphorylated 43kDa TAR DNA-binding protein, SD: Standard deviation, T1 W: T1-weighted imaging, TE: Echo time, TFCE – Threshold-free cluster enhancement, TI: Inversion time, TIV: Total intracranial volume, TR: Repetition time, UMN: Upper motor neuron

\textbf{Introduction}

Primary lateral sclerosis is an upper motor neuron predominant...
motor neuron disease, with markedly slower progression rates and longer survival than ALS. (Floeter and Mills, 2009, Chipika et al., 2019) It has a number of distinguishing and overlapping features with ALS, but owing to its low incidence and paucity of post-mortem studies, relatively little is known of its patterns of cerebral disease burden. PLS is often regarded as a 'pure' upper motor neuron disorder and many of its cardinal clinical features; spasticity, gait impairment, dysarthria and pseudobulbar affect are solely attributed to upper motor neuron dysfunction. The dominance of widespread pyramidal signs renders the clinical detection of subtle cerebellar and extrapyramidal signs challenging. Existing imaging studies of PLS primarily focus on corticospinal tract, (Iwata et al., 2011, Müller et al., 2018) corpus callosum, (Unrath et al., 2010, Agosta et al., 2014) brainstem (Bede et al., 2019) and precentral gyrus (Butman and Floeter, 2007, Schuster et al., 2013) pathology and few studies have endeavoured to specifically characterise extra-motor involvement. With very few exceptions, (Clark et al., 2018, Finegan et al., 2019) most PLS studies describe similar imaging patterns to ALS. (Müller et al., 2018, Van Weehaeghe et al., 2016) The notion that PLS selectively affects the UMN system has led to the assumption that extra-motor manifestations, such as neuropsychological, cerebellar and extrapyramidal deficits are rare. However, recent reports of cognitive deficits, (de Vries et al., 2019) descriptions of widespread cerebellar degeneration (Finegan et al., 2019, Clark et al., 2017, Floeter et al., 2014) and the observation of considerable subcortical TDP-43 burden (Kosaka et al., 2012) have gradually challenged this perspective. Accordingly, the objective of this study is the comprehensive characterisation of subcortical grey matter involvement in PLS. An additional objective of this study is the identification of subcortical signatures that may distinguish PLS from ALS. Our hypothesis is that basal ganglia, hippocampal and thalamic pathology can be detected in PLS, and the subcortical signature of PLS is different from ALS.

Methods

Participants and ethics

Thirty-three PLS patients, 100 patients with ALS and 117 healthy controls (HC) were included in a prospective neuroimaging study. The clinical and demographic profiles of the study participants are summarised in Table 1. The study was approved by the Ethics (Medical Research) Committee – Beaumont Hospital, Dublin, Ireland, and all participants provided informed consent prior to inclusion. Participating ALS patients had ‘probable’ or ‘definite’ ALS according to the revised El Escorial research criteria (Brooks et al., 2000) and PLS patients were diagnosed according to the Gordon criteria. (Gordon et al., 2006) Inclusion criteria included the ability to lie supine in the scanner for the duration of data acquisition. Exclusion criteria included implanted medical devices, such as baclofen pumps or pacemakers which would have precluded MR imaging. ALS patients with comorbid frontotemporal dementia were not included. (Racovsky et al., 2011) The healthy control cohort had no known neurological or psychiatric conditions, previous head injuries or established vascular risk factors.

Table 1. The demographic and clinical profile of study participants.

|                      | PLS n = 33 | ALS n = 100 | HC n = 117 | p value |
|----------------------|------------|-------------|------------|---------|
| Gender (male)        | 19 (62)    | 56          |            | 0.11    |
| Age-years (mean ± SD)| 60.5 ± 10.5| 59.8 ± 11.2 | 57.4 ± 11.9| 0.19    |
| Education-years (mean ± SD) | 12.9 ± 3.4 | 13.5 ± 3.2 | 14.3 ± 3.3 | 0.04*  |
| Handedness (right)   | 29         | 90          | 109        | 0.55    |
| ALSFRS-r (mean ± SD) | 34.4 ± 5.3 | 36.6 ± 7.5  | N/A        | 0.11    |

Genetics

Whole exome sequencing was performed in 29 of the 33 PLS patients, as previously described. (Finegan et al., 2019) Briefly, sequence data were assessed for quality, aligned to the GRCh37 reference genome, annotated and analysed using cutadapt V.1.9.1 (Martin, 2011), SAMtools V1.7 (Li et al., 2009), Picard V.2.15.0, Plink V.1.9 (Purcell et al., 2007), R V.3.2.3, SnpEff V.4.3 (Cingolani et al., 2012) and Gemini V.0.20.1 (Paila et al., 2013). Samples were compared to 135 Irish controls sequenced as described previously. (Project Min, 2018, McLaughlin et al., 2015) Putative variants were defined as protein altering variants in the exons and splice sites of 33 genes linked to ALS on the ALS online database (Abel et al., 2013) and 70 genes linked to HSP in the literature. (Klebe et al., 2015) The presence of the C9orf72 hexanucleotide repeat expansion was determined using repeat-primed polymerase chain reaction (PCR) as described previously (Byrne et al., 2012). Of the 100 ALS patients enrolled in neuroimaging, whole-genome sequence data was available for 44 patients (Project Min, 2018) and targeted DNA sequence data for a further 27. (Kenna et al., 2013) Patients were screened for previously reported ALS variants. C9orf72 repeat expansion status was determined in 97 patients. (Byrne et al., 2012)

Magnetic resonance imaging

Magnetic Resonance Imaging was performed on a 3 Tesla Philips Achieva system using an 8-channel receive-only head coil. T1-weighted images were acquired using a 3 D Inversion Recovery Prepared Spoiled Gradient Recalled echo (IR-SPGR) sequence; FOV: 256 × 256 × 160 mm, spatial resolution: 1 × 1 × 1 mm, TR/TE = 8/3.9 ms, TI = 1060 ms, flip angle = 8°, SENSE factor = 1.5 acquisition time = 7 min 30 s. A multimodal approach was implemented to comprehensively characterise the subcortical grey matter profile of the three study groups. First, total intracranial volumes (TIV) were calculated and the volumes of individual subcortical grey matter structures estimated. Subsequently, hippocampal segmentation was performed to estimate the volumes of specific subfields. Finally, morphometric analyses were carried out to identify focal pathological changes and vertex analyses were undertaken to characterise patterns of shape deformations.

Volumetric analyses

Total intracranial volume (TIV) was estimated for each participant to be used as a covariate for volumetric analyses. The FSL imaging suite was utilised for TIV estimations which were performed by linearly aligning each subject’s skull-stripped brain to the standard MNI152 brain image in MNI space, calculating the inverse of the determinant of the registration matrix and multiplying it by the size of the template. Registration to template was undertaken using FSL-FLIRT, (Jenkinson and Smith, 2001) and FSL-FAST was used for tissue type segmentation. (Zhang et al., 2001) Subsequent to brain extraction with FSL-BET, the subcortical segmentation and registration tool FIRST (Patenaude et al., 2011) of the FMRIB’s Software Library (FSL) was used to estimate the volumes of subcortical grey matter structures in the left and right hemispheres separately; the hippocampus, amygdala, thalamus, nucleus accumbens, caudate nucleus, putamen, and pallidum. Pipelines for subcortical segmentation and volume estimations were previously described. (Bede et al., 2013, Bede et al., 2018) Briefly, FSL-FIRST uses a two-stage affine registration procedure to register input T1 data sets to the Montreal Neurological Institute 152 (MNI152) standard space and a model-based approach is then implemented for the segmentation of subcortical structures. The accuracy of subcortical segmentation was individually verified for all participants. Subcortical mesh and volumetric outputs are generated following automatic boundary corrections. Analyses of covariance (ANCOVA) were
conducted to compare volumes of subcortical structures between study groups. Assumptions of normality, linearity and homogeneity of variances were verified. Volumes of subcortical grey matter structures were included as dependent variables, and study group allocation as the categorical independent variable. Age, education, gender and TIV were used as covariates and the ALS versus PLS contrasts were additionally corrected for symptom duration. A p-value ≤ 0.05 was considered significant. A table was generated with the estimated marginal means of volumes for each anatomical structure, standard error, between-group ANCOVA significance and p-values for Bonferroni-corrected post hoc testing. For illustrative purposes, estimated marginal means of volumes were plotted with confidence intervals to highlight group-specific volumetric traits for each structure. To illustrate the comparative subcortical profiles of PLS and ALS, percentage volume reductions were also calculated based on estimated marginal means for each structure with reference to healthy controls and plotted on a radar (spider) chart.

**Hippocampal subfields**

Version 6.0 of the FreeSurfer image analysis suite was used for the segmentation of the hippocampus into cytologically-defined subfields. (Fischl, 2012) T1-weighted data pre-processing included the removal of non-brain tissues, segmentation of the subcortical white matter and deep grey matter structures, intensity normalization, tessellation of the grey matter-white matter boundaries, and automated topology correction. The hippocampus was subsequently segmented into the following subfields using the hippocampal stream of the FreeSurfer package: CA1, CA2/3, CA4, fimbria, subiculum, hippocampal tail, molecular layer, molecular and granule cell layer of the dentate gyrus (GC-ML-DG), and hippocampal-amygdala transition area (HATA) (Iglesias et al., 2015, Christidi et al., 2019). Figure 1. To illustrate the comparative hippocampal profiles of PLS and ALS, percentage volume reductions were calculated based on estimated marginal means for each subfield with reference to controls and plotted on a radar (spider) chart.

**Vertex analyses**

As volumetric analyses only capture overall atrophy considering the entire structure, additional shape (vertex) and morphometric analyses were carried out to characterise deformation patterns and focal density reductions within the subcortical grey matter structures. Vertex-wise statistics were performed using FMRIB’s subcortical segmentation and registration tool FIRST. Vertex locations of individual study participants were projected on the surface of an average template shape as scalar values, positive values representing vertex locations outside the surface and negative values inside. Intergroup differences were explored using permutation-based non-parametric testing as implemented in FSL Randomise. (Winkler et al., 2014) Design matrices for the comparison of PLS patient with healthy controls included age, gender, TIV and education. The design matrix comparing the subcortical deformation profile of PLS and ALS patients included age, gender, TIV, education and symptom duration. The threshold-free cluster enhancement (TFCE) method (Smith and Nichols, 2009) was used and results were corrected for multiple comparisons across space (FWE < 0.05).

**Morphometry**

Following brain extraction and tissue-type segmentation, grey-matter partial volume images were aligned to MNI152 standard space using affine registration. Brain removal and tissue-type segmentation outcomes were individually verified. A study specific template was created, to which the grey matter images from each subject were non-linearly coregistered. A voxelwise generalized linear model and permutation-based non-parametric testing was used to highlight density alterations in a merged basal ganglia grey matter mask. (Winkler et al., 2014, Nichols and Holmes, 2002) The Harvard-Oxford subcortical probabilistic structural atlas was used to generate a merged basal ganglia mask incorporating the bilateral caudate, thalamus, accumbens, hippocampus, amygdala, putamen, and pallidum. (Frazier et al., 2005, Desikan et al., 2006)

**Results**

**Subcortical grey matter atrophy**

Subcortical segmentation confirmed statistically significant volume reductions in PLS compared to healthy controls in the following structures: left thalamus, right thalamus, left hippocampus and a trend.
of right accumbens atrophy (p = 0.056) accounting for age, gender, TIV and education. These structures also exhibited atrophy in the ALS group compared to controls, and significant additional volume reductions were noted in the left amygdala, left caudate and right accumbens.

Table 2. The comparison of the two patient cohorts confirmed disproportionate left pallidum and left accumbens atrophy in ALS. The volumes of other subcortical structures were not significantly different between the patient groups. Based on percentage volume reductions, the most affected structure in PLS was the accumbens nucleus followed by the thalamus, hippocampus and caudate. Figure 2. This was different from ALS where the second most affected structure was the amygdala.

Hippocampal subfield profiles

The evaluation of hippocampal subfields in PLS revealed focal atrophy in the dentate (GC-ML-DG), molecular layer, and CA4 subfields compared to controls as well as a statistical trend for volume reductions in CA1 and CA2/3. Figure 3. This pattern of selective subfield vulnerability was similar to that observed in ALS, with the exception that HATA volumes were significantly reduced in ALS which was not the case in PLS. Table 3. No significant volume reductions were identified in the hippocampal tail, subiculum, or fimbria in either patient group compared to controls. Moreover, no hippocampal segments were significantly different between ALS and PLS.

Vertex analyses

Shape analyses revealed statistically significant vertex deformations in PLS compared to controls in the bilateral thalamus, hippocampi and caudate nuclei. Figure 4. Patterns of surface-projected atrophy were strikingly symmetric in the two hemispheres. Preferential lateral hippocampal shape deformation and predominantly medial thalamic atrophy was observed. Most of the medial aspect of the body of the caudate nucleus, a small region of the lateral body of the caudate and the lateral- inferior aspect of the head of the left caudate was also affected in PLS. Vertex changes in the pallidum, and putamen did not reach significance compared to controls. The vertex-wise comparison of the ALS and PLS cohorts only reached significance in the left pallidum, left putamen and left caudate.

Focal morphometric alterations

The morphometric analyses restricted to the subcortical grey matter mask revealed left putamen and left pallidum pathology in PLS compared to controls at p < 0.05 TFCE FWE. Figure 6. This pattern is
Figure. 2. Left: The volumetric profile of subcortical grey matter structures in PLS, ALS and healthy controls (HC) based on estimated marginal means adjusted for age, gender, total intracranial volume (TIV) and education. Error bars represent 95% confidence intervals. Right: the comparative volumetric profile of subfields based on percentage reduction with reference to controls.

Figure. 3. Left: The volumetric profile of hippocampal subfield in PLS, ALS and healthy controls (HC) based on estimated marginal means adjusted for age, gender, total intracranial volumes (TIV) and education. Error bars represent 95% confidence intervals. Right: the comparative volumetric profile of subfields based on percentage change with reference to controls. CA: cornu ammonis, HATA: hippocampal-amygdala transition area, GC-ML-GD: molecular and granule cell layer of the dentate gyrus.
distinctly different from the right hippocampal and amygdala atrophy observed in ALS compared to controls. The direct comparison of ALS and PLS cohorts, revealed medial bi-thalamic atrophy in PLS compared to ALS and right putaminal changes in ALS compared to PLS. Figure 7.

Clinical and genetic profiling

Thirty-two of the 33 PLS patients had lower limb symptom onset. Ninety-one percent (n = 30) of the patients used a walking aid for safe ambulation and 79% (n = 26) experienced at least one fall during the 12-months preceding their MRI scan. The functional profile of the group based on ALSFRS-r demonstrated lower limb symptom predominance in the entire cohort: total ALSFRS-r M = 34.4, SD 5.3; bulbar sub-score M = 9.2, SD = 2.1; upper-limb sub-score M = 8.6, SD 2.1; lower limb sub-score ALSFRS-r sub-score M = 5.4, SD 1.6; respiratory sub-score M = 11.1, SD = 1.4. The Penn UMN score (Woo

Table 3

Hippocampal subfield volumes (mm3) in healthy controls (HC), ALS patients (ALS) and PLS patients (PLS) Estimated marginal means and standard error are adjusted for age, gender, education and total intracranial volume (TIV) Age = 58.77, Gender = 1.45, Edu = 13.78, TIV = 1429699.38. *Significant intergroup differences are flagged with asterisks. The ALS vs PLS post-hoc comparisons are adjusted for age, gender, education, TIV and symptom duration (age = 59.99, Gender = 1.39, Edu = 13.31, TIV = 1440387.86, Symptom Duration(m) = 45.71). ^ indicates statistical trends CA: cornu ammonis, HATA: hippocampal-amygdala transition area, GC-ML-GD: molecular and granule cell layer of the dentate gyrus

| Structure                                      | Study group | Estimated marginal mean- mm3 | Standard error | ANCOVA p value | ALS vs HC | PLS vs HC | ALS vs PLS |
|------------------------------------------------|-------------|-----------------------------|----------------|----------------|-----------|-----------|------------|
| Hippocampal Tail                               | HC          | 545.5                       | 6.2            | .107           | .310      | .206      | 1.0        |
|                                               | ALS         | 530.6                       | 6.6            | 1.5            |           |           |            |
|                                               | PLS         | 521.5                       | 11.5           |                |           |           |            |
| Subiculum                                      | HC          | 438.7                       | 4.0            | .077           | .289      | .133      | 1.0        |
|                                               | ALS         | 428.9                       | 4.3            |                |           |           |            |
|                                               | PLS         | 421.6                       | 7.4            |                |           |           |            |
| CA1                                            | HC          | 660.5                       | 5.8            | .029*          | .099      | .077      | 1.0        |
|                                               | ALS         | 642.2                       | 6.2            |                |           |           |            |
|                                               | PLS         | 632.9                       | 10.8           |                |           |           |            |
| Molecular Layer                                | HC          | 589.4                       | 5.0            | <.001*         | .002*     | .017*     | 1.0        |
|                                               | ALS         | 563.5                       | 5.4            |                |           |           |            |
|                                               | PLS         | 559.5                       | 9.3            |                |           |           |            |
| GC-ML-GD                                       | HC          | 311.8                       | 2.9            | <.001*         | .001*     | .006*     | .755       |
|                                               | ALS         | 296.5                       | 3.1            |                |           |           |            |
|                                               | PLS         | 292.3                       | 5.4            |                |           |           |            |
| CA2/3                                          | HC          | 227.6                       | 2.7            | <.001*         | <.001*    | .056      | 1.0        |
|                                               | ALS         | 212.6                       | 2.9            |                |           |           |            |
|                                               | PLS         | 214.1                       | 5.0            |                |           |           |            |
| CA4                                            | HC          | 270.0                       | 2.5            | <.001*         | .001*     | .011*     | .772       |
|                                               | ALS         | 257.0                       | 2.6            |                |           |           |            |
|                                               | PLS         | 254.6                       | 4.6            |                |           |           |            |
| Fimbria                                        | HC          | 77.1                        | 1.5            | .720           | 1.0       | 1.0       | 1.0        |
|                                               | ALS         | 75.6                        | 1.6            |                |           |           |            |
|                                               | PLS         | 74.9                        | 2.9            |                |           |           |            |
| HATA                                           | HC          | 65.1                        | 0.8            | .045*          | .047*     | .494      | 1.0        |
|                                               | ALS         | 62.2                        | 0.9            |                |           |           |            |
|                                               | PLS         | 62.7                        | 1.5            |                |           |           |            |

Figure 4. Subcortical shape alterations in PLS compared to controls. Surface projected patterns of atrophy are shown in orange over the 3-dimensional mesh representation of the thalami, caudate, and hippocampi shown in blue. p < 0.05 FWE.
profile was also consistent with selective functional impairment (higher scores indicate greater burden): total score (max = 32) M = 20.3, SD = 6.3; bulbar score (max = 4) M = 1.8, SD = 1.4; upper limb score (max = 14) M = 8.6, SD = 3.4; lower limb score (max = 14) M = 9.8, SD = 2.5. Finger-tapping rates were: right hand M = 2.59/s, SD = 1.32; left hand M = 2.34/s, SD = 1.12; right foot M = 1.41/s, SD = 0.69; left foot M = 1.36/s, SD = 0.78. Genetic testing data were available on 29 PLS patients. No PLS patient carried the

Figure. 5. Subcortical shape alterations in PLS compared to ALS. Surface projected patterns of atrophy are shown in red over the 3-dimensional mesh representation of the left pallidum shown in green. p < 0.05 FWE.

Figure. 6. Morphometric changes in PLS (top) and ALS (bottom) with reference to healthy controls in a subcortical grey matter mask (light blue). Radiological convention is used and MNI coordinates are provided. p < 0.05 TFCE FWE corrected for age, gender, total intracranial volume and education. Radiological convention is used; Rt - Right, Lt - Left.
c9orf72 hexanucleotide expansion and no PLS patients carried mutations previously implicated in ALS or HSP. Eleven of 100 ALS patients carried the C9orf72 repeat expansion.

Discussion

Our finding of considerable subcortical grey matter degeneration in PLS is consistent with emerging reports of subcortical pTDP-43 burden, and extra-motor clinical deficits. The importance of characterising extra-motor pathology in PLS is twofold; academic and clinical. From an academic perspective it provides compelling evidence that PLS is not a “pure” UMN disorder, and that the underlying pathology is not confined to the motor cortices and corticospinal tracts. While mechanisms of propagation in PLS remain elusive, it seems that progressive pathological changes take place in PLS, both spatially and chronologically. In ALS, a number of spreading mechanisms have been proposed; including prion-like propagation, progressive changes mediated by inflammation, impaired inhibitory function, hypermetabolic changes, none of which has been specifically studied in PLS. (Chipika et al., 2019, Schuster et al., 2015) In ALS, pathological staging systems have been developed based on TDP-43 disease burden, (Brettschneider et al., 2013) and attempted in vivo validations published based on imaging data, (Kassubek et al., 2014) but these have not been applied to PLS to date. (Brettschneider et al., 2013, Kassubek et al., 2014, Geser et al., 2011, Bede, 2019)

From a clinical perspective, the literature on cognitive and behavioural impairment in PLS is sparse and the suggestion that cognitive impairment might be present in a high proportion of PLS patients is relatively novel. (de Vries et al., 2019, de Vries et al., 2017) However, there is now a growing recognition that similarly to ALS, (Phukan et al., 2012, Elamin et al., 2011, Burke et al., 2016, Elamin et al., 2017) widespread deficits in social cognition, verbal fluency, executive function and memory impairment can also be detected in PLS. (de Vries et al., 2019) In our cohort of patients language deficits were present in 24%, verbal fluency was abnormal in 18% and 12% had abnormal memory performance on ECAS. (Abrahams et al., 2014) Given the high prevalence of verbal fluency deficits in the PLS group, it is noteworthy that significant bilateral thalamic morphological changes have previously been described in non-fluent variant primary progressive aphasia and significant hippocampal atrophy has been associated with behavioural-variant FTD; (Bede et al., 2018) observations which are consistent with our current findings. As in ALS, (Elamin et al., 2017) behavioural manifestations are not uncommon in PLS, which typically include loss of sympathy, apathy, as well as disinhibition. There has been also been reports of personality changes in PLS, such as late-onset obsessive compulsive disorder. (Bersano et al., 2018) Studies from other motor neuron diseases suggest that the basal ganglia may play a role in compensatory processes (Abidi et al., 2019) and their degeneration may contribute bulbar dysfunction. (Yunusova et al., 2019)

Similar to ALS, (Feron et al., 2018) extrapyramidal dysfunction has been previously reported in PLS, including freezing and postural instability (Mabuchi et al., 2004) which is sometimes referred to as “PLS-plus.” (Rowland, 2005) PLS cases were reported which initially presented with frank parkinsonism. (Gordon et al., 2006) However, the detection of extrapyramidal signs in the presence of widespread UMN degeneration is challenging based on clinical assessment alone, and may require computational gait analyses. (Feron et al., 2018) It is conceivable that extrapyramidal deficits contribute to gait impairment and fall risk in many patients with PLS. Based on anecdotal functional improvement on levodopa, a small clinical trial is currently underway to evaluate its use in PLS and ALS. (Clinicaltrials.gov 2019)
In addition to the neuropsychological and extrapyramidal manifestations of subcortical degeneration in PLS, it is likely that basal ganglia pathology also contributes to pseudobulbar affect (PBA). (Christidi et al., 2018, Bede and Finegan, 2018, Finegan et al., 2019) PBA is a very common symptom of PLS. (Thakore and Pioro, 2014) and in our cohort with 15 of 33 (45%) of PLS patients were affected. While PBA is classically linked to corticobulbar tract degeneration, it is increasingly clear that cerebellar and basal ganglia pathology also contribute to the aetiology of PBA. (Floeter et al., 2014, Bede and Finegan, 2018, Finegan et al., 2019) PBA has been previously linked to imaging changes in the putamen in ALS (Christidi et al., 2017), and basal ganglia pathology in stroke and MS. (Ghaffar et al., 2008) The clinical relevance of extra-motor changes in PLS is particularly important as this motor neuron disease phenotype carries a relatively good prognosis compared to ALS (Finegan et al., 2019) and extrapyramidal and neuropsychological impairment may impact on financial decisions, participation in clinical trials, driving, compliance with assistive devices, fall risk and impact on caregiver burden. (Olney et al., 2005, Elamin et al., 2013, Christidi et al., 2018)

While hippocampal degeneration is a recognised feature of ALS, (Machts et al., 2015, Christidi et al., 2018) it has not been specifically studied in PLS to date. Our volumetric analyses have captured overall hippocampal atrophy, and the parcellation of the structure revealed selective subfield vulnerability. The dentate (GC-ML-DG), molecular layer, and CA1, CA2/3 and CA4 subfields were preferentially affected with the relative sparing of the hippocampal tail, subiculum, and fimbria. The characterisation of affected and unaffected regions in PLS may have implications for the development of classification algorithms. (Bede et al., 2016, Grollemund et al., 2019) Two subcortical regions were identified where significant degeneration occurs in ALS, but not in PLS; the left amygdala and HATA. The majority of imaging studies in PLS describe imaging features similar to ALS. (Finegan et al., 2019, Schuster et al., 2016, Bede et al., 2018) The identification of PLS-specific imaging signatures which are distinct from ALS are clinically relevant, as they may contribute to the development of diagnostic applications for suspected PLS cases which don’t meet the current 4-year symptom duration criterion for diagnosis. (Clark et al., 2018, Schuster et al., 2017) Disease-specific imaging features are increasingly utilised in complex classification algorithms to provide a diagnostic probability, (Grollemund et al., 2019, Bede et al., 2017, Querin et al., 2018, Schuster et al., 2016) or to model prognostic outcomes. (Schuster et al., 2017)

Our finding of extensive subcortical degeneration in PLS is consistent with previous pathology reports. A post-mortem study on an individual with PLS reported marked atrophy of the thalamus and striatum. (Sugihara et al., 1999) Another study detected considerable atrophy of the thalamus and hippocampus. (Kosaka et al., 2012) Other sporadic reports exist of TDP-43 burden in the dentate gyrus and amygdala. (Fu et al., 2010) A recent post-mortem study of PLS identified significant basal ganglia involvement keeping with the extra-pyramidal signs recorded ante-mortem. (Hirsch-Reinshagen et al., 2019) Consistent with our imaging findings, hippocampal TDP-43 inclusions were also evident, specifically involving the dentate gyrus and cornu ammonis (CA).

Extra-motor involvement in ALS, (Christidi et al., 2018) particularly extensive temporal lobe and anterior frontal pathology, is often associated with C9orf72 hexanucleotide expansions. (Bede et al., 2013, Christidi et al., 2018) It is noteworthy that in our cohort of PLS patients, none of the study participants carried the pathological hexanucleotide expansion demonstrating that hippocampal and accumbens pathology cannot be solely attributed to C9orf72 mutations.

This study is not without limitations. Owing to the low incidence of PLS the sample size of our cohort is limited despite population-based recruitment efforts. Our study has a cross-sectional design which precludes the longitudinal assessment of subcortical changes in this cohort. As with previous comparative studies of PLS and ALS, PLS subjects in our cohort had substantially longer symptom duration than the ALS group. Furthermore, the PLS group and the healthy controls were not matched for education. Accordingly, both symptom duration and years of education were included as covariates in the statistical models, in addition to demographic factors. (Bede et al., 2013) An additional limitation of our study is the lack of post mortem data which would be instrumental in validating our imaging findings. As current diagnostic criteria require a symptom duration of four years, it is unclear whether the subcortical changes described herein are a late or early features of PLS. Very few longitudinal studies exist in PLS. Innovative studies of suspected PLS patients, or ‘pre-PLS’ cohorts have been previously undertaken to describe early imaging features. (Clark et al., 2018) Longitudinal studies in ALS indicate early ceiling effects in white matter metrics, which capture considerable integrity changes at baseline with limited progression over time. Grey matter metrics in ALS however show continued decline in the later stages of the disease making them superior monitoring marker candidates. (Westeneng et al., 2015, Bede and Hardiman, 2018) These observations provide the rationale to undertake longitudinal PLS studies focusing on both cortical and subcortical grey matter metrics.

Conclusions

PLS should no longer be regarded as a pure upper motor neuron disorder, as it is associated with marked thalamic, hippocampal and basal ganglia atrophy. The severity of subcortical grey matter degeneration observed in PLS is comparable to ALS, but the anatomical patterns are different. These data demonstrate that PLS exhibits a number of unique features that underpin a clinical course which is markedly distinct from ALS. In the era of precision medicine, PLS urgently needs dedicated imaging studies, disease-specific clinical trials, and staging systems to develop effective disease modifying strategies.

Declaration of Competing Interest

Peter Bede is the patron of the Irish motor neuron disease association (IMNDA), the head of the computational neuroimaging group (CNG) in Trinity College Dublin, member of the steering committee of the Neuroimaging Society of ALS (NISALS) and member of the biomedical research advisory panel of the UK MND association (MNDDA). These affiliations had no impact on the opinions expressed herein.

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Supplementary materials

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