Introduction

The hypnotic agent propofol (2,6-diisopropylphenyl) is widely used during the induction and maintenance of anesthesia. It has the advantages of minimal side effects, a controllable anesthetic state, a quick onset, and rapid patient emergence from general anesthesia. Apart from these anesthetic properties, propofol has additional antiarrhythmic and proarrhythmic effects.

Antiarrhythmic property of propofol

Several case reports have documented the antiarrhythmic effects of propofol. Kannan and Sherwood\textsuperscript{[1]} reported that a 68-year-old man with a previous myocardial infarction experienced supraventricular tachycardia. In this patient, administration of adenosine or carotid sinus massage had no effect, but propofol converted the supraventricular arrhythmia to sinus rhythm before electrical cardioversion. Hermann and Vettermann reported another case of ectopic supraventricular tachycardia that was terminated by propofol\textsuperscript{[2]}. Propofol has also been shown to terminate ventricular tachycardia (VT) storm\textsuperscript{[3, 4]}. Burjorjee and Milne reported that propofol resolved the recurrent episodes of VT that were not terminated by maximal antiarrhythmic therapy in a 65-year-old man\textsuperscript{[3]}; similar results were observed in a second study of a patient with comparable disease\textsuperscript{[4]}. Additionally, propofol converted atrial fibrillation, which was not terminated by intravenous infusion of amiodarone, to sinus rhythm\textsuperscript{[5]}. Propofol can significantly decrease P-wave dispersion\textsuperscript{[6]}, which may represent the reason for termination of atrial fibrillation. Furthermore, propofol may improve cardiac conduction. After propofol injection, the delta wave of Wolff-Parkinson-White syndrome has been shown to disappear and the P-R interval to normalize, though the delta wave returned immediately after propofol discontinuation\textsuperscript{[7, 8]}. The Q-T interval of long Q-T syndrome was shortened when propofol was used; therefore, propofol may have the potential to prevent episodes of VT that are due to Q-T interval dispersion\textsuperscript{[9, 10]}. Reperfusion of the heart after short-term ischemia may lead to potentially lethal arrhythmias, but propofol can alleviate reperfusion-induced arrhythmia. In the isolated rat myocardium, propofol had antiarrhythmic effects against reperfusion-induced arrhythmia at concentrations of 1, 10, and 20 μmol/L, and the incidence and duration of sustained arrhythmias were decreased significantly\textsuperscript{[11]}. From the above studies, we conclude that propofol is likely to inhibit arrhythmias. How-
ever, clinical conditions are often complex, and the general classification of propofol as a unique agent for antiarrhythmic therapy cannot yet be made. Thus, more studies are needed to understand the detailed mechanism by which propofol exerts its antiarrhythmic effects.

**Proarrhythmic properties of propofol**

Some studies have demonstrated that propofol has the potential to block the conduction system of the heart and thereby induce arrhythmia. A study of 60 children with paroxysmal supraventricular tachycardia undergoing radiofrequency catheter ablation showed that atioventricular (AV) node (AVN) conduction was slowed with propofol\(^{[12]}\). Atrial wavelength and AVN are electrophysiologic factors critical to the pathogenesis of supraventricular tachydysrhythmias. A study focusing on the effects of anesthetics on such electrophysiologic factors in guinea pig hearts has indicated that propofol does not change the atrial wavelength. However, this study revealed that propofol prolonged the AVN effective refractory period (ERP) in concentration- and frequency-dependent manners, though no significant effect on atrial conduction velocity was detected\(^{[13]}\). Additionally, at concentrations greater than 25 μmol/L, propofol slowed the atrial rate and AVN conduction and prolonged the Wenckebach cycle length in a concentration-dependent manner. The frequency of propofol administration also correlated with stimulus-to-His bundle intervals, such that the higher the frequency, the longer the S-H interval\(^{[14]}\).

Propofol (3 μmol/L) significantly prolonged the rabbit AV conduction interval in a dose-dependent (3 to 100 μmol/L) manner. At higher concentrations (10 to 100 μmol/L), the AVN Wenckebach cycle length and its refractory period were also prolonged. In addition, conduction through the His-Purkinje system (HV interval) and the atrial tissue (SA interval), as well as the spontaneous cycle length, were lengthened in a dose-dependent manner (30 to 100 μmol/L). By contrast, propofol had no effect on the refractory period of the atrium, ventricle, or the His-Purkinje system\(^{[15]}\). Therefore, propofol inhibits AV conduction in a concentration-dependent fashion.

Conversely, some research suggests that propofol does not affect the conduction system. For example, propofol had no effect on the electrophysiologic properties of the AVN and pathway conduction in atioventricular nodal reentrant tachycardia patients\(^{[16]}\). Additionally, propofol did not cause sinoatrial node depression or a pathologic increase in atioventricular conduction\(^{[17]}\). Studies in normal pig hearts demonstrate that propofol promotes a dose-related depression of sinus node and His-Purkinje system functions but has no effect on the AVN function or the conduction properties of atrial and ventricular tissues\(^{[18]}\).

The effects of propofol on the conduction system may be impacted by age and drug concentration. At a low concentration (3 μmol/L), propofol induced a significant lengthening of the AV conduction interval in adult rabbit hearts but not neonatal hearts. At a higher concentration (10 μmol/L and above), propofol significantly prolonged the AV conduction interval in hearts from both neonates and adults. The AV Wenckebach cycle length was also lengthened, with a more significant change in the adult hearts. However, with concentrations of propofol up to 100 μmol/L, the neonatal hearts progressed to complete AV block, which did not occur in the adults. The spontaneous heart rate and conduction through the atrial tissue (SA interval) and the His-Purkinje system (HV interval) were all slowed by propofol at 30 μmol/L or above. Additionally, the lengthening of the SA interval was more pronounced in the neonatal hearts, where the atrial refractory period was prolonged by propofol at 10 μmol/L and above\(^{[19]}\).

Based on the above study, the effects of propofol on the conduction system remain controversial. One reasonable explanation is that different concentrations promote divergent effects: at clinically relevant or low concentrations, propofol does not significantly suppress conduction, but at high concentrations, it blocks conduction.

In a canine study, propofol reduced the arrhythmogenic plasma concentration of epinephrine in a concentration-dependent manner, which suggests that propofol enhances epinephrine-induced arrhythmias\(^{[20]}\). Furthermore, propofol can directly induce arrhythmia.

Rewari and Kaul\(^{[21]}\) have reported that one long Q-T interval syndrome patient undergoing surgery experienced ventricular premature beats immediately upon propofol administration and then progressed to polymorphic ventricular tachycardia that intermittently changed to ventricular fibrillation. Another report described a 78-year-old woman with normal cardiac function who suffered torsade de pointes and then progressed to ventricular fibrillation after propofol infusion\(^{[22]}\). Prolonged high-dose propofol infusion can induce some particular arrhythmias. An infant also developed dramatic cardiac conduction disturbances and tachyarrhythmias after prolonged high-dose propofol infusion; this patient’s electrocardiograph resembled that of a patient with Brugada syndrome\(^{[23]}\). A study of a chronic propofol abuser with a propofol blood concentration as high as 0.73 μg/g also revealed a Brugada-like electrocardiograph. This patient subsequently developed a long Q-T interval, idioventricular rhythm, and ventricular fibrillation\(^{[24]}\).

These cases demonstrate that propofol can block conduction and induce bradycardia. Polymorphic ventricular tachycardia and some particular arrhythmias similar to Brugada syndrome can also result from propofol treatment. It should be noted that the most common abnormality is bradyarrhythmia, whereas others, such as AV block, are relatively rare and only occur at high propofol concentrations. A systematic review showed that the incidence of bradycardia (<50 beats per minute) was 4.8% in the presence of propofol\(^{[25]}\). The reason for the conduction block could be the ability of propofol to modify the activity of human atrial muscarinic cholinergic receptors, and this effect may be related to the drug-induced bradycardia\(^{[26]}\). Further, an M-2-muscarinic receptor-mediated mechanism may be the reason for propofol-induced conduction block. In a study of isolated guinea pig hearts, the negative dromotrophic effect of propofol was attenuated by a muscarinic receptor activator.
Propofol and action potential duration

Action potential duration (APD) is determined by several inward and outward ion currents, including $I_{Kur}$, $I_{Cat}$, $I_{Kr}$, and $I_{Ca}$. Changes in any individual current may prolong or shorten the APD, and APD dispersion can produce ventricular arrhythmia. Propofol (25 and 50 μmol/L) shortened monophasic APD at 90% repolarization (MAPD 90) in Langendorff-perfused guinea pig hearts\[27\]. With single guinea pig cardiac myocytes, propofol (10 and 100 μmol/L) shortened the APD. A concentration-dependent shortening of the APD was produced by propofol (1–100 μmol/L) in these cells\[28\]. However, the concentration of propofol required to reduce the APD by 50% was at least 100 μmol/L, and neither resting membrane potential nor action potential amplitude was significantly affected\[29\]. High-dose propofol (1000 μmol/L) also shortened the APD of single dog ventricular cells\[30\]. Propofol reduced the APD 90 and the APD 90 (the time required for 20% or 90% repolarization, respectively) in guinea pig cardiomyocytes in a concentration-dependent manner, and the APD 90 was reduced by 21% at 100 μmol/L\[31\]. In seeming contrast, however, propofol lessened the ischemia-induced decrease in the APD 90 at 1 μmol/L and 10 μmol/L and attenuated the APD dispersion around the “border zone”, which is between the normal and ischemic zones of guinea pig right ventricular muscle strips. Specifically, the APD was shortened by 27%, 31% and 3% at doses of 1, 10, and 20 μmol/L propofol, respectively\[31\]. This effect is of particular importance because the decrease in APD dispersion between the normal and ischemic areas results in a reduction in the incidence of reentrant tachycardia. These paradoxical findings indicate that the propofol-induced shortening or prolongation of the APD is related to the cell surroundings. In normal conditions, propofol reduces the APD, whereas in ischemic conditions, it lessens the shortening trend.

Propofol and ion channels

Propofol can influence some ion currents, including the ATP-sensitive potassium current, delayed rectifier K+ current, transient outward rectifier K+ current, inward rectifier K+ current, sodium current, and L-type Ca2+ current. These effects cumulatively result in changes to the APD.

ATP-sensitive potassium channels (K$_{ATP}$), which are composed of the Kir6.x and SUR subunits, are regulated by cytosolic nucleotides and link cell metabolism to electrical activity and K+ fluxes. There are two types: sarcolemmal K$_{ATP}$ (sarcK$_{ATP}$) channels and mitochondrial K$_{ATP}$ (mitoK$_{ATP}$) channels. A number of studies have examined the effect of propofol on mitoK$_{ATP}$ channels.

In isolated guinea pig myocardial cells, propofol alone had no significant effects on mitoK$_{ATP}$ channels at a concentration of 50 μmol/L, but it dose-dependently inhibited isoflurane-induced mitochondrial K$_{ATP}$ channel opening\[32\]. In isolated ischemia-reperfused guinea pig hearts, propofol (35 μmol/L) had no effect on mitoK$_{ATP}$ channels\[33\]. A study of single rat ventricular myocytes showed that propofol inhibited mitoK$_{ATP}$ channel activity, but the concentration required was very high (>31 μmol/L)\[34\]. By contrast, propofol has differing effects on sarcoK$_{ATP}$ channels. For example, studies\[35, 36\] on recombinant cardiac sarcoK$_{ATP}$ channels showed that propofol inhibited the channel with half the maximal inhibitory concentration (IC$_{50}$) more than 70 μmol/L; in the presence of MgADP, the IC$_{50}$ was as high as 183 μmol/L. Because propofol protein binding exceeds 95%, free fractions of propofol are less than 2 μmol/L\[34\]. Hence, we can conclude that propofol inhibits K$_{ATP}$ channels only at high concentrations and has no significant effect on either sarcoK$_{ATP}$ or mitoK$_{ATP}$ channel activities at clinically relevant concentrations.

In the human heart, the delayed rectifier K+ current ($I_{K}$) can be separated into at least three different components: ultra-rapid ($I_{kur}$), rapid ($I_{kr}$) and slow ($I_{ks}$). The effects of low- and high-dose propofol on some of these components have been examined. Two independent studies assessed the effect of propofol treatment on $I_{K}$\[30, 37\]. In one, propofol (28 μmol/L) induced significant depression of the $I_{K}$ component in single dispersed guinea pig ventricular myocytes. Data from the second study indicates that propofol inhibits $I_{K}$ at a concentration of 50 μmol/L. In another study, propofol suppressed the $I_{K}$ amplitude in a concentration-dependent manner (IC$_{50}$=36 μmol/L) in differentiated H9c2 cells. The H9c2 cell line was established from an embryonic rat cardiac ventricle, and it has properties similar to neonatal and adult cardiomyocytes\[38\]. It is important to note that the concentrations required for $I_{K}$ suppression are higher than what is currently used in the clinic.

The slowly activating component of the $I_{KS}$ contributes to human atrium and ventricle repolarization, particularly during action potentials with a long duration, and is a dominant determinant of the physiological heart rate-dependent shortening of the APD\[39\]. A study of guinea pig ventricular myocytes indicated that propofol inhibited the $I_{KS}$ (IC$_{50}$=23 μmol/L)\[39\], whereas data from another study in these cells demonstrated that propofol (100 μmol/L and 300 μmol/L) selectively inhibited the $I_{KS}$ of isolated guinea pig ventricular myocytes. Therefore, propofol was preferred as the agent to separate $I_{K}$ into $I_{KR}$ and $I_{KS}$\[40\]. Propofol also inhibited the $I_{KS}$ (IC$_{50}$=250 μmol/L), which is the current induced by the mRNA that encodes the minK protein that has similar electrophysiological properties to $I_{KS}$. This particular study measured the channel activity after expression via injection into Xenopus oocytes\[41\].

The rapid activating component of the delayed rectifier potassium current ($I_{K,R}$) is characterized by rapid activation, rapid inactivation and strong inward rectification, which promotes phase 3 of repolarization. Thus, $I_{K,R}$ plays an important role in governing the cardiac APD and refractoriness. Pharmacologically, $I_{K,R}$ is the target of class III antiarrhythmic drugs; however, to the best of our knowledge, little research concerning the effects of propofol on the $I_{K,R}$ component has been published. Heath and Terrar\[42\] showed that propofol had no effect on the channel, and it was selected as an agent...
to differentiate $I_{K1}$ and $I_{Kf}$. Similarly, no group has reported how propofol modulates the ultrarapid activating component of the delayed rectifier potassium current ($I_{KDR}$), which exists only in human atria and not ventricular tissue. $I_{KDR}$ is the predominant delayed rectifier current responsible for human atrial repolarization. Hence, studies of the effects of propofol on this component will be of great clinical value.

The transient outward rectifier K$^+$ channel current ($I_o$) is a voltage-gated channel current that is responsible for early rapid repolarization (phase 1). $I_o$ also determines the height of the early plateau, thus influencing activation of other currents that control repolarization$^{[31]}$. Propofol (60 μmol/L) inhibited the $I_o$ of canine ventricular cells, and the inhibition was not voltage dependent$^{[42]}$. Propofol (25 and 50 μmol/L) also significantly decreased the $I_o$ in rat ventricular myocytes but did not affect the voltage-dependent manner and the outward rectifier character$^{[43]}$. In addition, the steady-state voltage-dependent inactivation curve of $I_o$ was shifted to a more negative membrane potential, and the $I_o$ of rabbit atrial myocytes was suppressed by propofol with a 50% effective dose (ED$_{50}$) of 5.7 μmol/L. At 3 μmol/L, propofol slightly inhibited $I_o$, suggesting that while propofol does inhibit $I_o$ at high concentrations, the suppression is minimal or nonexistent at clinically relevant concentrations$^{[35]}$.

The inward rectifier K$^+$ current ($I_{Ki}$) plays a significant role in cardiac myocytes where it maintains the resting membrane potential and shapes the late repolarization phase (phase 3) of the action potential. Whether propofol inhibits the $I_{Ki}$ is not clear. Propofol (28 μmol/L) had no effect on the $I_{Ki}$ of single dispersed guinea pig ventricular myocytes$^{[37]}$ nor was an effect observed using a higher concentration (60 μmol/L) on single dispersed canine ventricular cells$^{[42]}$. Further, propofol (2.5 μmol/L) did not alter $I_{Ki}$ conductance in rat ventricular myocytes$^{[44]}$. By contrast, another study indicated that propofol suppressed $I_{Ki}$, although the decrease in $I_{Ki}$ occurred to a much lesser extent: the currents were decreased by only 18% when cells were exposed to 3 μmol/L propofol, and the inward rectification was not affected$^{[35]}$.

Cardiac sodium channels transmit a large inward depolarizing current ($I_{Na}$) during phase 0 of the cardiac action potential, and it has been shown that propofol inhibits sodium currents. For example, in isolated rabbit ventricular myocytes, the Na$^+$ current was decreased in dose-dependent and frequency-dependent manners by propofol with an ED$_{50}$ for current inhibition of 6.9 μmol/L. In part, this suppression was due to a negative shift of the steady-state voltage-dependent inactivation and a decreased rate of recovery from inactivation$^{[35]}$. Propofol inhibited $I_{Na}$ in isolated rat myocardial cells, and the effect was enhanced at depolarized resting potentials$^{[45]}$. In another study, single channel conductance was not changed by propofol, but a dose-dependent suppression of rat whole cell sodium currents (ED$_{50}$=14.8 μmol/L) was detected. The most reasonable explanation for this apparent dichotomy is that a shorter mean channel open time accompanied by an increased channel re-opening could result in slowed macroscopic inactivation$^{[46]}$. Hyperpolarization-activated, cyclic nucleotide-gated (HCN) channels conduct a monovalent cationic current (I(f) that contributes to autorhythmicity in the heart. Propofol inhibited and slowed the activation of recombinant HCN1, HCN2, and HCN4 channels at clinically relevant concentrations, in which the HCN1 current was the most sensitive of the three. HCN4 is the predominant subtype in the sinoatrial node (SA node)$^{[47, 48]}$. In this study, HCN4 channel activation was decreased more significantly than other isoforms with 20 μmol/L propofol. In addition, propofol reduced the heart rate in an isolated guinea pig heart preparation over the same range of concentrations. These data indicate that propofol modulation of HCN channel gating is an important molecular mechanism that contributes to the bradycardiac effect of propofol$^{[49]}$.

The L-type Ca$^{2+}$ current ($I_{Ca}$) is important in heart function because it triggers excitation-contraction coupling, modulates action potential shape, and is involved in cardiac arrhythmia. The negative inotropic effect of propofol can therefore be best explained by inhibition of the L-type Ca$^{2+}$ current ($I_{Ca}$). Though propofol did not alter steady-state $I_{Ca}$, it reduced the isoproterenol-stimulated increase in $I_{Ca}$, in a dose-dependent manner (0.1–10 μmol/L)$^{[50]}$. Propofol caused a statistically significant decrease in the $I_{Ca}$ of guinea pig cardiac myocytes in a concentration-dependent manner (1–100 μmol/L), even at low concentrations (1 μmol/L)$^{[50]}$. In single dog ventricular cells, propofol decreased the $I_{Ca}$ at 100 μmol/L$^{[50]}$. Propofol (10 and 100 μmol/L) also inhibited the $I_{Ca}$ of H9c2 cells and guinea pig cardiac myocytes$^{[28, 37]}$. In rat ventricular myocytes, propofol depressed the $I_{Ca}$ by 28% and 57% at 25 and 50 μmol/L, respectively$^{[51]}$. These data from several different model systems definitively prove that propofol inhibits $I_{Ca}$ even at concentrations used in the clinic (1 μmol/L).

In conclusion, propofol can inhibit sodium, calcium and potassium currents, but we should note that the concentrations needed for inhibition are much higher than clinically achievable concentrations. At clinically relevant concentrations, propofol is likely to inhibit $I_{K1}$, $I_{Na}$, and $I_{Ca}$, but controversy still exist, most notably the fact that propofol shortened the APD but lessened the ischemia-induced decrease of the APD.

**Propofol and gap junctions**

Gap junctions comprise intercellular channels that couple cardiac myocytes electrically and metabolically by facilitating the intercellular exchange of ions, signaling molecules, and other molecular information between neighboring cells in the heart. The constituent proteins of gap junction channels, connexins, play a critical role in impulse propagation and electrical synchronization between myocytes. Multiple connexin types are expressed in the heart, among which connexin 43 (Cx43) is the principal gap junction protein in ventricular myocardium$^{[52, 53]}$. Ventricular arrhythmia following acute myocardial infarction is always lethal. Propofol preconditioning has the ability to prevent the ischemic heart from progressing to a lethal ventricular arrhythmia. Cx43 is a principal cardiac gap-junction channel protein that undergoes progressive dephosphoryla-
tion during acute myocardial infarction (MI). The dephosphorylation of Cx43 decreases gap junction conduction, which produces the substrate for reentrant arrhythmia. Propofol treatment preserved Cx43 phosphorylation during acute myocardial ischemia, and this might protect the heart from serious ventricular arrhythmias during acute coronary occlusion[58]. However, one study showed that the electrical uncoupling correlated with an intercalated disk occurred 10–15 min after ischemia[59]. In the study of Hirata et al, most of the severe arrhythmias occurred between 5 and 10 min in all groups[53]. As such, the importance of the role played by Cx43 in arrhythmogenesis is debatable.

Propofol and the autonomic nervous system

The Bezold-Jarish reflex sensitivity is an index of vagal nerve activity. Activation of the Bezold-Jarish reflex by injection of some agents produced a profound reduction in heart rate. It has been postulated that propofol-induced bradycardia may be related to the Bezold-Jarish reflex. Vincze[60] and Ebert[61] indicated that propofol lowered the Bezold-Jarish reflex sensitivity, but other researches indicated that the cause of acute bradycardia after propofol administration did not involve the Bezold-Jarish reflex in humans[58] or rabbits[59].

Whether the block of cardiac conduction by propofol is due to direct or indirect effects by the autonomic nervous system is controversial. Ikeno[62] reported that propofol did not affect the cardiac conduction system when the autonomic nervous system was completely blocked. Therefore, it was thought that the conductance change was an indirect effect of propofol. Propofol also reduced the cardiac parasympathetic tone, but the suppression of sympathetic tone was more than that of parasympathetic tone[62,63].

Conclusion

Though propofol can induce various types of arrhythmias, some of which are severe, the incidence of arrhythmia is relatively rare at clinically relevant concentrations of propofol. Numerous studies conclusively show that propofol has the potential to inhibit arrhythmia. The potential arrhythmogenic mechanisms of propofol include ion channel inhibition, uneven suppression of the autonomic nervous system, and protection of gap junctions during ischemia. The controversies in the field can be attributed in part to the use of different species and experimental conditions and the promiscuous effects of propofol at various concentrations. The currently available data regarding the effects of propofol on arrhythmogenesis are not sufficient. Previous studies mostly focused on normal conditions, but arrhythmias are always induced in abnormal conditions, such as ischemia and electrolyte disturbances. In addition, there is no pharmacogenetic data on the opposing propofol effects at clinically used doses. Available clinical data largely came from sporadic reports rather than large-sized, blinded and controlled trials. We believe future studies will expand to these areas to fully characterize the arrhythmic and antiarrhythmic properties of propofol.

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