Antispasmodic activities of the methanolic extract from aerial parts of *Origanum* species on excised rat ileum

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**ABSTRACT**

**Background and Aims:** *Origanum hypericifolium*, *Origanum minutiflorum*, *Origanum saccatum*, and *Origanum haussknechtii* are some of the endemic *Origanum* species in Turkey. In the literature, besides the known effects of these *Origanum* species, no antispasmodic activity was determined. Therefore, we investigated *Origanum* species for possible antispasmodic effects to rationalize its traditional medicinal use. The present study aimed to investigate the antispasmodic activity of *Origanum* species with the organ bath.

**Methods:** Rats were divided into 5 groups. While no treatment was given to the control group, plant extracts were given to other groups. Ileal tissues taken by rats were used for evaluating the contraction responses elicited with potassium chloride (KCl) and acetylcholine (ACh). Thereafter, the extracts of *Origanum* species were applied and cumulative concentration-response curves of these species were constructed in each experiment.

**Results:** All treatments with *Origanum* species have a relaxation effect on rat ileum as compared with the control group. According to the results of the isolated organ bath study, KCl and ACh-induced ileum contractions were significantly decreased by all *Origanum* species.

**Conclusion:** *Origanum* species that were used in the present study can prevent smooth muscle contraction of ileum, and therefore they can be used to reduce intestinal spasms.

**Keywords:** *Origanum hypericifolium*, *Origanum minutiflorum*, *Origanum saccatum*, *Origanum haussknechtii*, Organ bath, Antispasmodic effect

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INTRODUCTION

Turkey, an important gene center for the Lamiaceae family, is represented by 45 genera, 546 species, and 730 taxa (Celik, Herken, Arslan, Ozel, & Mercan, 2010). The endemic rate of the Lamiaceae family in Turkey is 53% (Cam et al., 2019). Lamiaceae members are mainly common in the mountainous areas of the Mediterranean region of Turkey (Taskin, Cam, Taskin, & Rayaman, 2019).

Oregano is a term that refers to members of Lamiaceae with many common characteristics, all of which contain carvacrol as the major component in their essential oils. Oregano includes Origanum, Thymus, Thymbra, Satureja, and Cordothymus of the Lamiaceae family. Origanum, one of these, is among important aromatic plants common in the world and comprises the largest portion of oregano used in trade (Baser, 2008). Hippocrates (500 BC) was aware of the antiseptic properties of oregano, and it is used for respiratory diseases and stomach pain. Dioscorides (1st century AD), the author of the famous for Materia Medica, has proposed oregano tea for snakebites. Paracelsus (16th century AD) suggested using oregano for the treatment of diarrhea, psoriasis, and fungal diseases (Baser, 2008).

Some of the endemic Origanum species in Turkey are Origanum hypericifolium Schwarz et Davis, Origanum minutiflorum Schwarz et Davis, Origanum saccatum Davis, and Origanum haussknechtii Boiss. The first of these is O. hypericifolium, which is used in traditional medicine for the treatment of some diseases, especially diabetes (Ocak, Celik, Ozel, Kocan, & Konuk, 2012). It mainly contains p-cymene, γ-terpinene, thymol, and carvacrol (Koseoglu, Taskin, Sadikoglu, & Bitis, 2016). The second Origanum species is O. minutiflorum that has α-pinene, camphene, α-terpinene, p-cymene, carvacrol-methyl-ether, and carvacrol (Ozcan & Chalchat, 2009). In addition to this, the fourth plant O. haussknechtii was collected from Burdur, Isparta-Çandır, Antalya-Alanya, and Erzincan-Kemaliye provinces, respectively. The plant materials were identified by Dr. Narin Sadikoglu. Voucher specimens are deposited in the herbarium of the Faculty of Pharmacy, Inonu University, and herbarium voucher numbers: Origanum 2009/016, Origanum 2009/013, Origanum 2009/033, Origanum 2009/002, respectively.

Materials and Methods

Materials

All chemicals used in the organ bath were purchased from Sigma (St. Louis, Missouri, USA).

Collection of plant material

O. hypericifolium, O. minutiflorum, O. saccatum, and O. haussknechtii were collected from Burdur, Isparta-Çandır, Antalya-Alanya, and Erzincan-Kemaliye provinces, respectively. The plant materials were identified by Dr. Narin Sadikoglu. Voucher specimens are deposited in the herbarium of the Faculty of Pharmacy, Inonu University, and herbarium voucher numbers: Origanum 2009/016, Origanum 2009/013, Origanum 2009/033, Origanum 2009/002, respectively.

Methods

Preparation of the extracts

The aerial parts of four dried Origanum species (20 g) were extracted with 100 ml portions of methanol by maceration at room temperature (25 °C) for 7 days stirring several times until the extracts remained colorless. The extracts were filtered and evaporated to dryness under reduced pressure at 45 °C in a rotary evaporator. The crude extracts were then transferred to vials and kept at +4 °C. The crude extract was administered by diluting with distilled water at different concentrations before being used in the organ bath.

Animals

All animal experiments were carried out with the approval of the Marmara University Animal Experiments Local Ethics Committee (permission number: 51.2013.mar). Adult female Sprague-Dawley rats (250-350 g) (n=8 in each experiment) were obtained from Marmara University The Experimental Animal Implementation and Research Center (Istanbul, Turkey). The rats were housed under controlled temperature (20-23 °C), in humidity (40-60%), and light (12 h light/dark regime)-regulated rooms. The animals were kept on a standard rodent pellet diet, with tap water available ad libitum. Before starting to the experiments, all necessary precautions were taken, and the factors that would adversely affect the parameters during the study were minimized.
Organ bath studies
Healthy animals were divided into 5 groups. In the organ bath system, the control group was given methanol as a vehicle, while the other groups were given plant extracts. Ileum was removed from the rats after the rats were sacrificed by guillotine. 2 cm-long part of ileum were sliced from all ileum; subsequently, the surrounding mesentery and fat tissues were carefully removed from ileum, and ileum was mounted vertically in an organ bath containing 20 ml Tyrode buffer (composition (mM): NaCl, 139.9; KCl, 2.68; CaCl₂, 1.8; MgCl₂, 1.05; NaHCO₃, 11.9; NaH₂PO₄, 0.42; and glucose 5.55) at pH 7.40. The temperature of the organ bath was maintained at 37 °C, and carbogen (95% O₂ and 5% CO₂ gas mixture) was perfused through the bath. After mounting, ileum was equilibrated for approximately 60 min before the experiments were started with a buffer change every 15 min. The initial tension of the preparation was set to about 1 g. The tension of ileum was measured with an isometric force transducer (PowerLab, ADInstruments, Australia) and recorded on-line on a computer via a four-channel transducer data acquisition system using appropriate software (LabChart, ADInstruments, Australia).

Contractility studies
In the isolated ileum, rhythmic and tonic contractions were elicited with 80 mM KCl and 10⁻⁶ M acetylcholine (ACh). Firstly, contraction with KCl was measured, this measurement was used for % of KCl maximum and then KCl was removed by changing the water bath three times. The contraction responses of ACh were determined 15 minutes after removing KCl and Origanum species were cumulatively (0.025, 0.05, 0.1, 0.25, 0.5, 1 mg/ml) administrated at the highest point of contraction with ACh. The relaxation response of ileal tissues were evaluated by adding cumulative concentrations of Origanum species on ACh induced precontractions. ACh induced contraction values were calculated according to KCl. Following the addition of each concentration of Origanum species, the recording was performed for 120 s. KCl induced contractions before the administration of Origanum species on rat ileum was obtained to investigate the viability of the tissues. Similar responses to KCl were observed for tissues among the groups. Concentration-response curves were fitted and areas under curves (AUC) were evaluated and analyzed statistically with Prism 6.05 (GraphPad Software Inc. San Diego, CA, USA). The AUC values and the maximal inhibitory effect of Origanum species (Eₘₐₓ) values were calculated according to % relaxation response of tissues to ACh contraction.

Statistical analysis
All values are presented as mean ± standard error of the mean (SEM). Differences in the contractile effect of KCl and ACh were analyzed using analysis of variance (ANOVA) tests with the Tukey multiple comparison test. Concentration-response curves were fitted, AUC determined, and Eₘₐₓ (%) calculated with the Prism 6.05 software. The value of p<0.05 was considered statistically significant.

RESULTS
The main focus of our study is to clarify and compare the effects of some endemic species of Origanum in Turkey on rat ileum relaxations and contractions in the organ bath experiments. Therefore, the effect of these plants used in folk medicine was verified, and hence, the relaxation effect of Origanum species in various diseases can be utilized. The results obtained from the in vitro organ bath are given below according to responses to KCl and ACh contractions on rat ileum.

Contraction responses of KCl on rat ileum
The results of isolated organ bath experiments studied on rat ileum demonstrated that there are no significant differences between groups on contraction responses of the ileum tissues to 80 mM KCl which acts via voltage-gated calcium channels on all groups. It is clearly seen that the contraction responses of the ileum tissues to 80 mM KCl was similar before the administration of Origanum species.

Relaxation responses of cumulative concentrations of Origanum species on rat ileum
The relaxation response of Origanum species at the concentration range of 0.025 to 1 mg/ml on ACh induced contraction on rat ileum was demonstrated in Figure 1. In the previous studies, some other Origanum species (O. acutidens, O. compactum, O. vulgare, and O. majorana) was found to display a relaxation response. Similar to the results of the previous studies, all treatments with Origanum species at the concentration range of 0.025-1 mg/ml have a relaxation response on rat ileum as compared with the control group. In comparison with the control group, the highest ileum relaxing effect of Origanum species belong to O. saccatum (p<0.05), O. haussknechtii (p<0.05), O. hypericifolium (p<0.01), and O. minutiflorum (p<0.001), respectively. Relaxation responses was augmented with the increase in concentration of Origanum species in all groups. Thus, concentration-dependent relaxations were observed and these relaxing effects were completely recovered during washout period.

Figure 1. Changes in the relaxation responses of the cumulative Origanum species after contraction with ACh. Concentration of Origanum species: 0.025, 0.05, 0.1, 0.25, 0.5, and 1 mg/ml. Each group (n=8) represents Mean ± SEM. The statistical analyses were carried out with ANOVA tests with the Tukey multiple comparison test. *p<0.05, **p<0.01, ***p<0.001 vs control group.
Besides, $E_{\text{max}}$ and AUC values for *Origanum* species were listed in Table 1. *O. hypericifolium* ($p<0.05$), *O. saccatum* ($p<0.01$), and *O. haussknecthii* ($p<0.05$) treatments increased the $E_{\text{max}}$ values compared to the control group.

### Table 1: Changes ($E_{\text{max}}$ and AUC values) in the relaxation response of *Origanum* species on ACh induced contractions in the rat ileum.

|                  | $E_{\text{max}}$ (%) | AUC (%) |
|------------------|----------------------|---------|
| Control          | 21.470±3.561         | 16.77   |
| *O. hypericifolium* | 38.570±4.301*        | 30.99   |
| *O. minutiflorum* | 26.870±2.704         | 22.46   |
| *O. saccatum*    | 49.950±5.405**       | 39.72   |
| *O. haussknechtii* | 43.750±5.211**      | 38.75   |

* $p<0.05$, ** $p<0.01$ vs control (vehicle) group (Mean ± SEM). AUC: Area under the curve. $E_{\text{max}}$ (%): the maximum relaxing effect of *Origanum* species.

### Discussion

In literature, the antispasmodic activity of *O. acutidens* essential oil in rat ileum was investigated by organ bath study. In that study, the essential oil of *O. acutidens* was applied in cumulative doses (0.1, 0.5, and 1 mg) and decreased ileal tissue contraction. According to their results, the antispasmodic effect of that essential oil is strong, concentration-independently and fully reversible (Goze et al., 2010). In another study, which investigated the antispasmodic activity of *O. compactum*, an organ bath study with the guinea-pig ileum and duodenum, and the rat fundus strip and duodenum were performed. According to their results, *O. compactum* showed antispasmodic activity on smooth muscles. In that study, it was suggested that the effects of *O. compactum* on muscle relaxation were caused by a decrease in the presence of the calcium for contraction of muscle due to inhibition of intracellular Ca$^{2+}$ release and extracellular Ca$^{2+}$ influx in the smooth muscle cells (Van Den Broucke & Lemli, 1980). In another study, the antispasmodic activity of *O. vulgare* was investigated with an organ bath study in the strips of rabbit urinary bladder. *O. vulgare* inhibited carbachol and K$^+$-induced contractions in concentration-dependent manner (Khan et al., 2011). The antispasmodic activity of the essential oil from the aerial part of *O. majorana* on muscle relaxation was investigated on rabbit and rat intestinal smooth muscle. According to these results, *O. majorana* essential oil inhibited the carbachol and KCl-induced contraction of tissue (Makrane et al., 2019). We investigated the antispasmodic effect of *Origanum* species and examined their relaxation response in ACh (10$^{-6}$M) induced contractions on rat ileum. As obtained from our results, *Origanum* species reduced contractions on rat ileum.

### Table 2: Changes ($E_{\text{max}}$ and AUC values) in the ACh induced contractions in the presence of *Origanum* species on the rat ileum.

|                  | $E_{\text{max}}$ (%) | AUC (%) |
|------------------|----------------------|---------|
| Control          | 90.110±3.310         | 72.10   |
| *O. hypericifolium* | 87.780±2.528         | 64.64   |
| *O. minutiflorum* | 87.980±5.301         | 52.96   |
| *O. saccatum*    | 65.220±13.980**      | 57.88   |
| *O. haussknechtii* | 74.550±5.172*        | 45.17   |

* $p<0.05$, ** $p<0.01$ vs control group (Mean ± SEM). AUC: Area under the curve. $E_{\text{max}}$ (%): the maximum contraction of ACh.
contraction of smooth muscle by the opening of the voltage-induced L-type calcium channels, thus resulting in the contraction of the smooth muscles because of the influx of extracellular Ca^{2+} (Khan et al., 2011).

ACh, due to its effect on muscarinic M_{4} receptors, leads to contraction by decreasing cyclic adenosine monophosphate (cAMP) levels, increasing the amount of intracellular calcium or affecting nicotinic receptor and opening ion channels, and thus produce depolarization (Nausch, Heppner, & Nelson, 2010). It is also well known that contraction induced by K^{+} in smooth muscles is caused by an increase in Ca^{2+} flow through voltage-gated Ca^{2+} channels, and therefore, any substance that inhibits high K^{+} mediated contractions of smooth muscles is claimed to be an inhibitor of Ca^{2+} influx (Chiworo & Ojewole, 2009). KC1 induced contraction on ileum smooth muscle is mainly caused by calcium influx through voltage-sensitive calcium channels (Ponce-Monter et al., 2006). The present findings show that the Origanum species inhibit ACh induced ileum contractions acting on the stress-sensitive calcium channels. These Origanum species led to concentration-dependent and reversible inhibition of the isolated rat ileum contractions. The contraction caused by ACh was inhibited by Origanum species in a concentration-dependent manner.

Additionally, according to our results, the inhibitory effects of Origanum species induced by ACh is not reversible. These effects are only reduced and eliminated by washing tissues. The relative decrease in the effects of the extract after tissue washing can be related to the removal of the extract from the surface of receptors. Moreover, prolonged ileum contractions without a reduction in contractile force with ACh in control tissues showed that the decrease in contractile strength was due to the performance of the extract rather than muscle fatigue.

Phytochemical studies reveal the presence of various substances in plants. The activities observed in the extracts of the Origanum species that we tested are probably due to the presence of these phytochemicals. For example, it is known that flavonoids exhibit antispasmodic activity. However, other phytochemicals in the extracts also contribute to the effects found (Tumen et al., 1995). Considering the phytochemical studies on these Origanum species, these plants contain tannins, flavonoids, sitosterols, phenolic glycosides, phenolic terpenoids, arbutin, and numerous essential oil molecules.

Carvacrol (C_{10}H_{14}O, a monoterpenoid phenol), one of the major components of Origanum species, is investigated in the Origanum species extract and it is known to have antispasmodic activity. Thus, the antispasmodic activity observed in the present study can be attributed to the carvacrol of Origanum (Baser, 2008). It is reported that carvacrol content of the O. hypericifolium, O. minutiflorum, O. saccatum, and O. haussknechtii are 42-82% (Baser, Ermin, Kurucuoglu, & Tumen, 1994), 34-64% (Baser, Tumen, & Sezik, 1991), 6.36-7.18% (Tumen et al., 1995), and 4.34% (Koseoglu et al., 2016), respectively. In addition to carvacrol, it is reported that thymol, another component of Origanum species, has antispasmodic effect; however, p-cymene has no antispasmodic effect. According to the study, which is a continuation of the same study, the basic fact emerging from the results is that pharmacological inhibition of smooth muscle activity plays a major role in the non-specific and non-competitive antagonism of thymol and carvacrol. Changes in the degree of effect of the Origanum species whose effects were investigated in our study are likely to vary according to the presence of carvacrol, thymol, and other components at different rates. However, further investigations are needed on that subject. Additionally, an edible plant with a long history of medical use is considered to be relatively safe, but detailed studies of the safety profile are needed before being recommended for clinical use.

Any report of antispasmodic properties of these Origanum species on the literature was not mentioned before. According to our results, the extracts of Origanum species reduced smooth muscle contraction on rat ileum. The effects of the extract were concentration-dependent and completely reversed during washing. The exact mechanism of Origanum species may be related to Ca^{2+} level changes in the ileum smooth muscle in the presence of extract. The antispasmodic effect of the extract observed in this study supports the clinical efficacy and use of the extracts of Origanum species in the treatment of spasmodic disorders. Further studies are needed to explain the underlying mechanism of action of Origanum species and evaluate its use in folk medicine. Due to the antispasmodic activity of Origanum species, these plants can be considered as a natural source and freely used in the food and pharmaceutical industry.

**CONCLUSION**

The antispasmodic effects of some Origanum species, such as Origanum hypericifolium, Origanum minutiflorum, Origanum saccatum, and Origanum haussknechtii, were investigated. The Origanum species have shown significant antispasmodic activity by acting on various pharmacological pathways. The contraction caused by ACh was inhibited by Origanum species in a concentration-dependent manner. Consequently, Origanum species can cause concentration-dependent and reversible inhibition of spontaneous contractions on isolated rat ileum, and they can inhibit contraction of the smooth muscle of ileum on rats. Thus, it can be used to decrease intestinal spasms. Further studies are needed to determine the mechanism of action and major effective compounds.

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**Ethics Committee Approval:** All animal experiments were carried out with the approval of the Marmara University Animal Experiments Ethics Committee (number 51.2013.mar).

**Author Contributions:** Conception/Design of Study- H.K.E., M.EÇ., A.N.H.Y., L.K.; Data Acquisition- M.EÇ., A.N.H.Y.; Data Analysis/Interpretation- M.EÇ., L.K.; Drafting Manuscript- M.EÇ., A.N.H.Y.; Critical Revision of Manuscript- L.K., H.K.E.; Final Approval and Accountability- M.EÇ., A.N.H.Y., L.K., TT, LB, H.K.E.; Technical or Material Support-T.T; Supervision- L.B.

**Conflict of Interest:** The authors have no conflict of interest to declare.

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