Management of gestational trophoblastic neoplasia

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Introduction

Gestational trophoblastic neoplasia is a success story in medical oncology as it is associated with highest cure rates even in patients with high tumour loads and perhaps would be an excitement to the practicing oncologist treating such cases.

It is just 100 years since Marchand [1] identified choriocarcinoma as a tumour arising from placental villous trophoblast. Earlier description of similar tumours failed to identify their tissue of origin. Gestational Trophoblastic Disease (GTD) is the terminological umbrella now used to span the spectrum of cellular proliferations ranging from villous forms of hydatiform mole through invasive mole and choriocarcinoma to placental site tumours [2]. Each form of GTD presents its own particular set of problems ranging from social to therapeutic. These proliferations are unique in several aspects. In the first place there is no known homolog of hydatiform mole in any other species. The rare identification of choriocarcinoma like tumours remains restricted to a rhesus monkey [3] and an armadillo [4]. In the human, histologically characterized choriocarcinoma behaves in a broadly similar manner whether it is genetically identical with the host as in germ cell origin, or whether it arises from a normal conception or an androgenetic hydatiform mole.

It is now more than 35 years since the sensitivity of invasive mole and choriocarcinoma to cytotoxic drugs was first recognized [5]. They remain the most sensitive and most curable of all human cancers.

GTD is still an important reproductive health problem worldwide. The problem is that much information of GTD has come from less developed countries, where proper diagnostic tools and up to date treatment cannot be employed. Maternal age, previous hydatiform mole, race and geographical region have been identified as clear risk factors for GTD. Etiological factors of GTD have long been studied but no definite causes have yet been found.

However, it can be speculated that during gametogenesis and fertilization, the Risk factors may act synergistically. Genetic sub classification may be helpful in this context. Geographic variations of incidence exist but are inextricably linked with above mentioned risk factors. Important prerequisites for accurate evaluation are common denominators, standard classification and definition of index cases. The developments and improvements in suction curettage, termination of pregnancy, contraceptive techniques, diagnostic imaging and biochemical testing have been associated with not only fall in birth rate but also with a reduction in trophoblastic disease [6-14].

Gestational Trophoblastic Disease (GTD) can be benign or malignant. Histologically, it is classified into hydatidiform mole, invasive mole (chorioadenoma destruens), choriocarcinoma, and Placental Site Trophoblastic Tumor (PSTT). Those that invade locally or metastasize are collectively known as Gestational Trophoblastic Neoplasia (GTN). Hydatidiform mole is the most common form of GTN. While invasive mole and choriocarcinoma are malignant, a hydatidiform mole can behave in a malignant or benign fashion.

In histologic section of a complete hydatidiform mole stained with hematoxylin and eosin, Villi of different cell types are present. The large villous in the center exhibits edema with a fluid-filled central cavity known as cisterna. Marked proliferation of the trophoblast is observed. The syncytiotrophoblasts stain purple, while the cytotrophoblasts have a clear cytoplasm and bizarre nuclei. No fetal blood vessels are in the mesenchyme of the villi. CHMs are usually diploid and androgenetic in origin, ~80% resulting from duplication of the haploid genome of a single sperm while 20% arise by dispermic fertilisation of an ovum. In either case maternal chromosomes are lost before, or shortly after, fertilisation. However, while nuclear DNA is entirely paternal in CHM, mitochondrial DNA remains maternal in origin [15]. No methods exist to accurately predict the clinical behavior of a hydatidiform mole by histopathology. The clinical course is defined by the patient’s serum human chorionic gonadotropin (hCG) curve after evacuation of the mole. In 80% of patients with a benign hydatidiform mole, serum hCG levels steadily drop to normal within 8-12 weeks after evacuation of the molar pregnancy. In the other 20% of patients with a malignant hydatidiform mole, serum hCG levels rise or plateau [16,17].

Hydatidiform mole is considered malignant when the serum hCG levels plateau or rise during the follow-up period and an intervening pregnancy is excluded. This occurs in 15-20% of hydatidiform moles [16,17].

A hydatidiform mole with a fetus or fetal tissue and a triploid karyotype is known as a partial or incomplete mole. Partial moles also have malignant potential, but only 2-3% become malignant [18-20]. An invasive mole has the same histopathologic characteristics of a hydatidiform mole, but invasion of the myometrium with necrosis and hemorrhage occurs or pulmonary metastases are present.

Histologically, choriocarcinomas have no villi, but they have sheets of trophoblasts and hemorrhage. Choriocarcinomas are aneuploid and can be heterozygous depending on the type of pregnancy from which the choriocarcinoma arose. If a hydatidiform mole preceded the choriocarcinoma, the chromosomes are of paternal origin. Maternal and paternal chromosomes are present if a term pregnancy precedes the choriocarcinoma. Of choriocarcinomas, 50% are preceded by a hydatidiform mole, 25% by an abortion, 3% by ectopic pregnancy, and the other 22% by a full-term pregnancy [16].

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Placental site trophoblastic tumor is a rare form of gestational trophoblastic neoplasia, with slightly more than 200 cases reported in the literature [21,22]. In patients with PSTT, intermediate trophoblasts are found infiltrating the myometrium without causing tissue destruction. The intermediate trophoblasts contain human placental lactogen (hPL) [23]. These patients have persistent low levels of serum hCG (100-1000 mIU/mL). However, serum hCG levels as high as 108,000 mIU/mL have been reported in patients with PSTT [24]. The most frequent sites of metastases of malignant gestational trophoblastic neoplasia are the lungs, lower genital tract, brain, liver, kidney, and gastrointestinal tract.

Epidemiology

The incidence is estimated at 1-3: 1000 pregnancies for CHM and 3: 1000 pregnancies for PHM, respectively [15]. GTD appears to be more frequent in Asia than in North America or Europe.

An increased risk of molar pregnancy is seen in the very young (<16 years), but is most associated with advanced maternal age (>45 years). Following a molar pregnancy, the risk of a further CHM or PHM increases to ~1%. After two molar gestations, the risk of a third mole is 15%–20%. The frequency of CC and PSTT is less clear, since these can arise after any type of pregnancy. CC develops after around 1:50 000 deliveries, while recent data suggest that PSTT represents 0.2% of UK GTD cases [25]. GTN risk may also relate to hormonal factors since women with menarche after 12 years of age, light menstrual flow and prior use of oral contraceptives are at increased risk.

Diagnosis

CHMs and PHMs most commonly present with vaginal bleeding in the first trimester of pregnancy. Previously reported features such as anaemia, uterine enlargement, pre-ecclampsia, hyperemesis, hyperthyroidism and respiratory distress are now rare [26] reflecting the introduction of routine ultrasonography in early pregnancy.

Characteristic sonographic findings for CHM in the second trimester, of a heterogeneous mass (‘snowstorm’), without foetal development and with theca lutein ovarian cysts, are not seen in the first trimester, and ultrasonography is not diagnostic reliably [27]. Indeed, false positive and negative rates are high with ultrasound, especially for PHM, and histological examination is essential to achieve a correct diagnosis [27]. All products of conception from nonviable pregnancies must undergo histological examination regardless of ultrasound findings [28]. The safest method of evacuation is suction dilatation and curettage (D&C) under ultrasound control to ensure adequate emptying of uterine contents and to avoid uterine perforation [15]. A proportion of women who miscarry or who undergo medical terminations will have unsuspected molar pregnancies. As histological examination is not routinely requested, the diagnosis of GTN can be delayed resulting in significantly greater morbidity [29]. Histological examination of every termination is impractical, and perhaps a simple measurement of the urine or serum hCG level 3–4 weeks post-treatment to ensure return to normal is indicated [29].

The other malignant forms of GTD, CC and PSTT/ETT can be much trickier to diagnose as the disease can develop months or many years after a prior pregnancy with protein presentations possible. Although change in menstruation is frequent, it does not always occur. It is therefore essential to measure the hCG in any woman of childbearing age who has unexplained metastatic disease. Biopsy of lesions without the ability to control bleeding is highly risky in this very vascular disease and is not essential before commencing chemotherapy. However, where complete excision is possible this can provide useful histological confirmation of the diagnosis and material for genetic analysis.

Indications for treatment

1. Plateaued or rising hCG after evacuation
2. Heavy vaginal bleeding or evidence of gastrointestinal or intraperitoneal haemorrhage
3. Histological evidence of choriocarcinoma
4. Evidence of metastases in the brain, liver or gastrointestinal tract, or radiological opacities of >2 cm on chest X-ray
5. Serum hCG of ≥20 000 IU/l >4 weeks after evacuation, because of the risk of uterine perforation.

Staging investigations and treatment stratification after a molar pregnancy

Most patients developing GTN post-HM are detected early via hCG monitoring and so extensive investigation is rarely required. Information to determine therapy can be obtained from the clinical history, examination, measurement of serum hCG and a Doppler pelvic ultrasound to confirm the absence of a pregnancy, to measure the uterine size/volume, spread of disease within the pelvis and its vascularity. The latter assessed by the Doppler pulsatility index is an independent prognostic factor for resistance to single-agent methotrexate MTX) therapy [30] and is now being evaluated in a prospective trial. Pulmonary metastases are most common, so a chest radiograph is essential [31].

Computed Tomography (CT) of the chest is not required if the chest X-ray (CXR) findings are normal, since discovery of micro metastases, which may be seen in ~40% of patients, does not influence outcome [32]. However, if lesions are noted on CXR, magnetic resonance imaging (MRI) of the brain and CT body are indicated to exclude more widespread disease involving, for example, the brain or liver, which would significantly alter management. FIGO reports data on GTN using prognostic scoring and anatomic staging systems [33].

The total score for a patient is obtained by adding the individual scores for each prognostic factor. Low risk, 0–6; high risk, ≥7. PSTT should not be scored and instead requires staging. Stage I, disease confined to the uterus; stage II, disease extending into the pelvis; stage III, disease spread to lungs and/or vagina; stage IV, all other metastatic sites including liver, kidney, spleen and brain.

Staging investigations for CC and PSTT/ETT

Women who present with an elevated hCG and suspected GTN (CC or PSTT/ETT) following a prior pregnancy require much more extensive staging investigations, which include a contrast enhanced CT of the chest and abdomen, MRI of the brain and pelvis, a Doppler ultrasound of the pelvis and may benefit from a lumbar puncture to assess the cerebrospinal fluid to serum hCG ratio. The latter if more than 1:60 suggests occult central nervous system disease. For CC, the FIGO scoring/staging system is the same as described above. However, PSTT/ETT has a discrete biological behavior with less hCG production, slower growth, late metastasis and slightly less chemo sensitivity. Consequently, the scoring system is not valid for PSTT/ETT, but FIGO staging is used to help adapt treatment intensity. Some investigators have recently started using Positron Emission Tomography (PET)/CT imaging, but experience is still quite limited. It appears that this
imaging modality is more helpful in relapsed disease to identify sites for resection and, as with other cancers, is prone to both false-positive and false-negative results [15].

Management of low-risk disease

About 95% of patients with HM who develop GTN are low risk (score 0–6). In women with stage I disease apparently confined to the uterine cavity, the role of second D&C in reducing the need for chemotherapy remains controversial. UK results indicate that this procedure is only valuable if the hCG is <5000 IU/l with disease in the cavity rather than myometrium. Indeed, the low efficacy of a second D&C, small risks of introducing infection, causing haemorrhage and uterine perforation should be balanced against the almost 100% cure rate and relative safety of chemotherapy [15].

There is no consensus on the best chemotherapy regimen for initial management of low-risk Gestational Trophoblastic Neoplasia (GTN), and first-line regimens vary by geography and institutional preference. Most regimens have not been compared head-to-head, and the level of evidence for efficacy is often limited to except as noted below. Even if there are differences in initial remission rate among the regimens, salvage with alternate regimens is very effective, and the ultimate cure rates are generally 99% or more. The initial regimen is generally given until a normal beta human chorionic gonadotropin (beta-hCG) (for the institution) is achieved and sustained for 3 consecutive weeks (or at least for two treatment cycles beyond normalization of the beta-hCG). A salvage regimen is instituted if any of the following occur:

- A plateau of the beta-hCG for 3 weeks (defined as a beta-hCG decrease of 10% or less for 3 consecutive weeks).
- A rise in beta-hCG of greater than 20% for 2 consecutive weeks.
- Appearance of metastases.

The use of chemotherapy in the first-line management of low-risk GTN has been assessed in a Cochrane Collaboration systematic review [34]. In that systematic review, four randomized controlled trials were identified [35-38].

Three of the randomized trials [36-38] compared the same two commonly used regimens:

- Biweekly (pulsed) dactinomycin (1.25 mg/m² intravenously [IV]).
- Weekly intramuscular methotrexate (30 mg/m²).

These three trials included a total of 392 patients. All three trials showed better primary Complete Response (CR) rates without the need for additional salvage therapy associated with pulsed dactinomycin (relative risk [RR] of cure, 3.00; 95% Confidence Interval [CI], 1.10–8.17), even though the magnitude of benefit showed substantial heterogeneity (I² statistic = 79%) [36-38]. Fewer courses of therapy were needed to achieve CR and cure with dactinomycin treatment. As expected, salvage chemotherapy was nearly uniformly successful, because almost all low-risk GTN patients are ultimately cured, irrespective of the initial chemotherapeutic regimen. There were no statistically significant differences in most toxicities.

The Cochrane systematic review also summarized the evidence from four nonrandomized trials, but comparisons across studies are difficult. The regimens evaluated in those studies are included in the lists below [34].

Commonly used treatment regimens include the following:

1. The 8-day Charing Cross regimen. Methotrexate (50 mg Intramuscularly [IM] on days 1, 3, 5, and 7) and folinic acid (7.5 mg orally on days 2, 4, 6, and 8). This may be the most common regimen worldwide [34,39], but it has not been directly compared with other regimens.
2. Biweekly pulsed dactinomycin (1.25 mg/m² IV).
3. Weekly methotrexate (30 mg/m² IM). Efficacy of this regimen appears to be low for choriocarcinoma and for patients with Fédération Internationale de Gynécologie et d’Obstétrique (FIGO) risk scores of 5 to 6.

Other regimens in less-common use include the following [34].

- An 8-day regimen of methotrexate (1 mg/kg IM days 1, 3, 5, and 7) and folinic acid (0.1 mg/kg IM days 2, 4, 6, and 8).
- Methotrexate 20 mg/m² IM days 1 to 5, repeated every 14 days.
- Dactinomycin 12 μg/kg/day IV days 1 to 5, repeated every 2 to 3 weeks. This regimen has fallen out of favour because of substantial alopecia and nausea.
- Methotrexate 20 mg IM daily, days 1 to 5; and dactinomycin 500 μgIV daily, days 1 to 5, repeated every 14 days.
- Dactinomycin 10 μg/kg/day, days 1 to 5, repeated every 2 weeks.
- Methotrexate 0.4 mg/kg/day IM daily on days 1 to 5, repeated after 7 days.
- Etoposide 100 mg/m²/day IV on days 1 to 5, or 250 mg/m² IV on days 1 and 3, at 10-day intervals [40].

As gestational trophoblastic neoplasia is a highly curable disease, the aim of treatment should be to minimize the drug toxicity but not at the cost of treatment efficacy.

Management of high-risk GTN

Multigent chemotherapy is standard for the initial management of high-risk Gestational Trophoblastic Neoplasia (GTN). A systematic literature review revealed only one randomized controlled trial (and no high-quality trials)—conducted in the 1980s—comparing multigent chemotherapy regimens for high-risk GTN [41]. In the trial, only 42 women were randomly assigned to either a CHAMOMA regimen (i.e., methotrexate, folinic acid, hydroxyurea, dactinomycin, vincristine, melphalan, and doxorubicin) or MAC (i.e., methotrexate, dactinomycin, and vincristine). The study showed better primary Complete Response (CR) rates without additional salvage therapy associated with pulsed dactinomycin (relative risk [RR] of cure, 3.00; 95% Confidence Interval [CI], 1.10–8.17), even though the magnitude of benefit showed substantial heterogeneity (I² statistic = 79%) [36-38]. Fewer courses of therapy were needed to achieve CR and cure with dactinomycin treatment. As expected, salvage chemotherapy was nearly uniformly successful, because almost all low-risk GTN patients are ultimately cured, irrespective of the initial chemotherapeutic regimen. There were no statistically significant differences in most toxicities. There was a statistically significant increase in dermatologic toxicity, including alopecia, associated with dactinomycin. However, in the largest study [38], there was statistically significantly more low-grade gastrointestinal toxicity, grade 2 nausea, grade 1 to 2 vomiting, and grades 1 to 3 neutropenia in the dactinomycin group. In that study, choriocarcinoma patients and patients with a risk score of 5 to 6 had a worse CR rate to initial treatment with single-agent therapy, and methotrexate was virtually ineffective [38].

The fourth randomized trial was a very small study of 45 patients and compared a 5-day regimen of dactinomycin (10 μg/kg) with an 8-day regimen of methotrexate (1 mg/kg) and folinic acid (0.1 mg/kg) on alternate days. There was a statistically significant decrease in risk of failure to achieve primary cure without the need for salvage therapy in the dactinomycin arm (RR, 0.57; 95% CI, 0.40–0.81) [35]. There was less alopecia associated with methotrexate but more hepatic toxicity.

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Other regimens in less-common use include the following [34].

- An 8-day regimen of methotrexate (1 mg/kg IM days 1, 3, 5, and 7) and folinic acid (0.1 mg/kg IM days 2, 4, 6, and 8).
- Methotrexate 20 mg/m² IM days 1 to 5, repeated every 14 days.
- Dactinomycin 12 μg/kg/day IV days 1 to 5, repeated every 2 to 3 weeks. This regimen has fallen out of favour because of substantial alopecia and nausea.
- Methotrexate 20 mg IM daily, days 1 to 5; and dactinomycin 500 μgIV daily, days 1 to 5, repeated every 14 days.
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- Etoposide 100 mg/m²/day IV on days 1 to 5, or 250 mg/m² IV on days 1 and 3, at 10-day intervals [40].

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and chlorambucil) [42]. There was substantially more life-threatening toxicity in the CHAMOMA arm and no evidence of higher efficacy. However, there were serious methodologic problems with this trial. It was reportedly designed as an equivalency trial, but owing to the small sample size, the trial was inadequately powered to assess equivalence. In addition, the characteristics of the patients randomly assigned to the two study arms were not reported (although the authors stated that there were no major differences in the patient populations assigned to each arm), nor was the method of randomization or allocation concealment described.

There are no randomized trials comparing regimens in common use to establish the superiority of one over another. Therefore, the literature does not permit firm conclusions about the best chemotherapeutic regimen [41]. However, since EMA/CO (i.e., etoposide, methotrexate, and dactinomycin/cyclophosphamide and vincristine) is the most commonly used regimen, the specifics are provided in Table below [43–45].

Cycles are repeated every 2 weeks (on days 15, 16, and 22) until any metastases present at diagnosis disappear and serum beta-human chorionic gonadotropin (beta-hCG) has normalized, then the treatment is usually continued for an additional three to four cycles.

Results of a large, consecutive case series of 272 patients with up to 16 years of follow-up showed a complete remission rate of 78% using this regimen, and these results are consistent with other case series in the literature that employed EMA/CO [43]. More than two-thirds of the women who did not have a complete response or subsequently had disease recurrence could be salvaged with cisplatin-containing regimens (with or without resection of metastases), yielding a long-term cure rate of 86.2% (95% CI, 81.9%–90.5%) [39]. Moreover, routinely when the addition of cisplatin plus etoposide was added to EMA/CO, a 9% improvement was reported in the survival results of these high-risk patients [46]. Among the women who had an intact uterus, about 50% of them retained their fertility. Patients with documented brain metastases received higher doses of systemic methotrexate as part of the EMA component (i.e., etoposide, methotrexate, folinic acid, and dactinomycin) of EMA/CO (1 g/m² intravenously [IV] for 24 hours, followed by folinic-acid rescue, 15 mg orally every 6 hours for 12 doses starting 32 hours after methotrexate). Patients with brain metastases received an increased dose of systemic methotrexate of 1 g/m² for 24 hours followed by folinic-acid rescue, 15 mg orally every 6 hours for 12 doses starting 32 hours after methotrexate. Patients with lung metastases received cranial prophylaxis with irradiation and intrathecal methotrexate 12.5 mg every 2 weeks with the CO (i.e., cyclophosphamide and vincristine) cycles.

Table 1. FIGO 2000 scoring system for GTN.

| Prognostic factor | Score |
|-------------------|-------|
| Age (years) <40   | 1     |
| ≥40               | 1     |
| Antecedent pregnancy (AP) Mole | 0     |
| Abortion | Term |
| Interval end of AP to chemotherapy in months | 4     |
| 4–6               | 7–12  |
| >12               | –     |
| hCG (IU/l) <10⁶   | 1     |
| 10⁶–10¹⁰          | 10¹⁰–10¹⁵ |
| >10¹⁵             | –     |
| Number of metastases | 0     |
| 1–4               | 5–8   |
| >8                | –     |
| Site of metastases Lung Spleen and kidney G1 tract Brain and liver | –     |
| Largest tumour mass | 3–5 cm |
| >5 cm             | –     |
| Prior chemotherapy | –     |
| Single drug       | >2 drugs |

Table 2. Specifics of the EMA/CO Regimen a,b,c.

| Day | Drug | Dose |
|-----|------|------|
| 1   | Etoposide | 100 mg/m² IV for 30 min |
|     | Dactinomycin | 0.5 mg IV push |
|     | Methotrexate | 300 mg/m² IV for 12 h |
| 2   | Etoposide | 100 mg/m² IV for 30 min |
|     | Dactinomycin | 0.5 mg IV push |
|     | Folinic Acid | 15 mg or PO every 12 h × 4 doses, beginning 24 h after the start of methotrexate |
| 8   | Cyclophosphamide | 600 mg/m² IV infusion |
|     | Vincristine | 0.8–1.0 mg/m² IV push (maximum dose 2 mg) |

IV=intravenously; PO=orally.

a Adapted from Bower et al. [43]
b Adapted from Escobar et al. [44]
c Adapted from Lurain et al. [45]

Table 3. TP/TE schedule for relapsed GTN.

| Regimen | Schedule |
|---------|----------|
|         | Day 1    |
| Dexamethasone | 20 mg oral (12 h pre-paclitaxel) |
| Dexamethasone | 20 mg oral (6 h pre-paclitaxel) |
| Cimetidine | 30 mg in 100 ml NS over 30 min i.v. |
| Chlorophenamine | 10 mg bolus i.v. |
| Paclitaxel | 135 mg/m² in 250 ml NS over 3 h i.v. |
| Mannitol | 10% in 500 ml over 1 h i.v. |
| Cisplatin | 60 mg/m² in 1 L NS over 3 h i.v. |
| Post-hydration | 1 L NS + KCl 20 mmol + 1 g MgSO₄ over 2 h i.v. |
| Day 15   |         |
| Dexamethasone | 20 mg oral (12 h pre-paclitaxel) |
| Dexamethasone | 20 mg oral (6 h pre-paclitaxel) |
| Cimetidine | 30 mg in 100 ml NS over 30 min i.v. |
| Chlorophenamine | 10 mg bolus i.v. |
| Paclitaxel | 135 mg/m² in 250 ml NS over 3 h i.v. |
| Etoposide | 150 mg/m² in 1 L NS over 1 h i.v. |

Table 4. Low-/high-risk post-chemotherapy patients, hCG concentration sampling.

| Year | Urine | Blood |
|------|-------|-------|
| 1    | Weekly | Weekly |
| Week 1–6 after chemotherapy | Weekly |
| Month 2–6 | Two weekly |
| Month 7–12 | Two weekly |
| Year 2 | Four weekly |
| Year 3 | Eight weekly |
| Year 4 | Three monthly |
| Year 5 | Four monthly |
| After Year 5 | Six monthly |

Examples of other regimens that have been used include the following [41]

- MAC: Methotrexate, folinic acid, dactinomycin, and cyclophosphamide.
- Another MAC: Methotrexate, dactinomycin, and chlorambucil.
- EMA: Etoposide, methotrexate, folinic acid, and dactinomycin (EMA/CO without the CO).
- CHAMOCA: Methotrexate, dactinomycin, cyclophosphamide, doxorubicin, melphalan, hydroxyurea, and vincristine.
CHOMOMA: Methotrexate, folic acid, hydroxyurea, dactinomycin, vincristine, melphalan, and doxorubicin.

Brain metastases are associated with poor prognosis, particularly when liver metastases are also present [47-49]. However, even patients with brain metastases may achieve long-term remission in 50% to 80% of cases [43,44,49]. Patients with Central Nervous System (CNS) metastases receive additional therapy simultaneously with the initiation of systemic chemotherapy. Some centers utilize whole-brain irradiation (30 Gy in 2 Gy fractions) with or without intrathecal methotrexate [47]. However, some investigators omit the cranial radiation, relying on replacement of the standard dose of methotrexate in the EMA/CO regimen with the higher dose of 1,000 mg/m² IV for 24 hours on the first day, as noted above, to achieve therapeutic CNS levels [49-52].

Management of drug-resistant disease

About 20% of high-risk GTN patients will progress on or after primary chemotherapy, but these individuals still have an excellent outcome with ~75%–80% still being salvaged. This is partly because relapse is detected early due to hCG monitoring so disease volume is small. Moreover, hCG monitoring enables the early detection of resistance during therapy. In relapsed patients, fluorine-18 fluoro-2-deoxy-D-glucose-PET (FDG-PET) scanning may help identify the site of active disease to facilitate surgical resection and cure. However, if surgery is not possible, several salvage regimens have been either created or adopted from the germ cell tumour setting. At Charing Cross Hospital, a regimen has been developed combining etoposide with cisplatin (EP) alternating weekly with EMA that omitted the second day of etoposide and ActD. Survival rates are >80% but toxicity is significant, and less toxic salvage therapies are required. Several cases of drug-resistant GTN have been reported to respond and/or be cured by paclitaxel-based single-agent or combination therapy, gemcitabine and capcitabine.

Of these, an alternating two weekly doublet of paclitaxel/cisplatin and paclitaxel/etoposide (TP/TE) appears from non-randomised data to be much better tolerated than EP/EMA and is effective in patients with relapsed and/or refractory GTN. In view of these results, the International Society of the Study of Trophoblastic Diseases (ISSTD) has recently proposed a randomised trial of TE/TP versus EP/EMA to determine the optimal therapy for patients relapsing after non-cisplatin/paclitaxel-based combination therapies such as EMA/CO.

Another approach in patients with refractory disease involves high-dose chemotherapy with peripheral stem-cell transplantation. However, cure are not common, so improved patient selection may be required to achieve better outcomes from this approach. PSTT differs from CC, growing more slowly, metastasising later, involving lymph nodes more commonly and producing less hCG [15]. However, like CC, it can arise after any type of pregnancy, including PPHM, and usually presents with abnormal vaginal bleeding [16]. PSTT may be suspected if the hCG level is low for the volume of disease present on imaging combined with an elevated free beta form of hCG, but none of these features are diagnostic.

Consequently histological confirmation is essential. A recent large population-based series of PSTT comprised 62 cases over 30 years, representing 0.2% of UK GTD cases, and examined prognostic features. On univariate analysis, stage, hCG, mitotic index and a duration of >4 years from the preceding pregnancy were prognostic, but the FIGO score was unhelpful. Only the duration from the prior pregnancy remained predictive of survival on multivariate analysis with 100% (13 of 13) dying and 98% (48 of 49) surviving for those ≥48 and <48 months, respectively. This effect was not explained by differences in disease stage or hCG levels, but may reflect a biological switch in the tumours after this time. In the absence of sufficient data regarding adjuvant therapy, currently 8 weeks of EP/EMA or TE/TP are advocated when there are poor risk factors such as disease presenting beyond 4 years of the antecedent pregnancy. While uterine-sparing surgery is possible [15], multifocal microscopic uterine disease can occur, which could compromise survival and careful counselling is required. Currently, it is thought that ETT behaves very similarly to PSTT but in reality, little data are available to be sure of this. PSTT and ETT are so rare that it is unlikely that their treatment will ever be fully optimised. Very rarely, multi-drug resistant disease develops that is not amenable to surgical resection or any other existing treatment, so it is unclear whether anything can be done in this case. Since GTN is very vascular it is plausible that vascular targeting agents such as bevacizumab might be active. The tumours can also overexpress epidermal growth factor receptor, leading to the question whether erlotinib or gefitinib could demonstrate efficacy. The potential for an anti-hCG targeted therapy has not been explored and could be of interest in women who have completed their families or have run out of other options.

Follow-up and long-term implications

The risk of relapse after chemotherapy is ~3% and most occur in the first year of follow-up. Therefore, careful hCG monitoring is required and pregnancy should ideally be delayed until beyond this period. Any method of contraception can be used including the oral contraceptive pill, as long as there are no other contraindications to their use.

Fortunately, apart from EMA/CO bringing forward the menopause date by 3 years, fertility is not otherwise affected with 83% of women becoming pregnant after either MTX/FA or EMA/CO chemotherapy [15]. Moreover, there is no obvious increase in the incidence of congenital malformations. When a patient does become pregnant, it is important to confirm by ultrasound and other appropriate means that the pregnancy is normal. Follow-up is then discontinued, but the hCG should be rechecked at 6 and 10 weeks after the pregnancy to ensure no recurrence or new disease.

Late sequelae from chemotherapy have been remarkably rare. In 279 patient on 15 years of follow-up, there was no significant increase in the incidence of second tumours following MTX therapy. In contrast, 26 patients receiving combination chemotherapy for GTN developed another cancer when the expected rate was only 16.45, a significant difference. Most of this risk appears to occur if combination chemotherapy is continued beyond 6 months.

References

1. Marchand F (1985) Über die sogenannten ‘decidualen’ Geschwülste im Anschluss an normale Geburt. Abort. Blasenmccle, undextraintra Schwangerschaft. Monatschr. Geburtshilfe Gyänikut 1: 419-438.
2. (1983) Gestational trophoblastic diseases. Report of a WHO Scientific Group. World Health Organ Tech Rep Ser 692: 7-81. [Crossref]
3. Lindsey JR, Wharton LR Jr, Woodruff JD, Baker HJ (1969) Intrauterine choriocarcinoma in a rhesus monkey. Pathol Vet 6: 378-384. [Crossref]
4. Marin-Padilla M, Benirschke K (1963) Thalidomide induced alterations in the blastocyst and placenta of the armadillo, dasypus novemcinctus mexicanus, including a choriocarcinoma. Am J Pathol 43: 999-1016. [Crossref]
5. Hertz R, LJ MC, Spencer DB (1956) Effect of methotrexate therapy upon choriocarcinoma and chorioadenoma. Proc Soc Exp Biol Med 93: 361-366. [Crossref]
6. Berkowitz RS, Goldstein DP (1996) Chorionic tumors. N Engl J Med 335: 1740-1748. [Crossref]
13. Wake N, Takagi N, Sasaki M (1978) Androgenesis as a cause of hydatidiform mole.
12. Kajii T, Ohama K (1977) Androgenetic origin of hydatidiform mole.
11. Lage JM, Wolf NG (1995) Gestational trophoblastic disease. New approaches to
9. 
8. Gestational Trophoblastic Tumor (GTT) (1997). In SGO Handbook, Staging of
Gynecologic Malignancies, 2nd Edition.
7. Suzuki T, Goto S, Nawa A, Kuruczhi O, Saito M, et al. (1993) Identification of the
pregnancy responsible for gestational trophoblastic disease by DNA analysis. Obstet
Gynecol 82: 629-634. [Crossref]
6. 
5. \[Crossref\]
4. Baergen RN, Rutgers JL, Young RH, Osann K, Scully RE (2006) Placental site
trophoblastic tumour: a study of 55 cases and review of the literature emphasizing
markers and long-term outcome of placental-site trophoblastic tumours: a retrospective
observational study. Lancet 374: 48-55. [Crossref]
3. Schmid P, Nagai Y, Agarwal R, Hancock B, Savage PM, et al. (2009) Prognostic
markers and long-term outcome of placental-site trophoblastic tumours: a retrospective
4. 
3. \[Crossref\]
25. \[Crossref\]
23. \[Crossref\]
24. Nigam S, Singhal N, Kumar Gupta S, Chhabra D, Manaktala U (2004) Placental site
trophoblastic tumour in a postmenopausal female--a case report. Gynecol Oncol 93: 550-
553. [Crossref]
22. \[Crossref\]
21. \[Crossref\]
20. Ginsberg A, Gillespie A, Tidy J, Everard R G N J, Wells M, et al. (2005) Placental site
trophoblastic tumour: clinical features and management. Gynecol Oncol 99: 603-607. [Crossref]
19. \[Crossref\]
18. Fowler DJ, Lindsay I, Seckl MJ, Sebire NJ (2007) Histomorphometric features of
hydatidiform moles in early pregnancy: relationship to detectability by ultrasound
examination. Ultrasound Obstet Gynecol 29: 76-80. [Crossref]
17. Hitchins RN, Holden L, Newlands ES, Begent RH, Rustin GJ, et al. (1988) Single agent
etoposide in gestational trophoblastic tumours. Experience at Charing Cross Hospital
16. \[Crossref\]
15. Hitchins RN, Holden L, Newlands ES, Begent RH, Rustin GJ, et al. (1988) Single agent
etoposide in “poor prognosis” metastatic gestational trophoblastic disease: a randomised controlled trial. Eur J Cancer 24: 1041-1046. [Crossref]
14. Engel L, Yan X, Zhang J, Wu T (2009) Combination chemotherapy for high-risk
gestational trophoblastic tumour. Cochrane Database Syst Rev 15: CD005196. [Crossref]
13. Alazzam M, Tidy J, Hancock BW, Osborne R (2009) First line chemotherapy in low-
risk gestational trophoblastic neoplasia: a gynecologic oncology group study. J Clin Oncol 27: 825-
831. [Crossref]
12. Khan F, Everard J, Ahmed S, Coleman RE, Aitken M, et al. (2003) Low-risk persistent
gestational trophoblastic disease treated with low-dose methotrexate: efficacy, acute and long-term effects. Br J Cancer 89: 2197-2201. [Crossref]
11. Lurain JR1, Singh DK, Bozorgi K, Fishman DA (2003) Treatment of high-risk
gestational trophoblastic neoplasia. J Reprod Med 48: 165-170. [Crossref]
10. Hancock BW, Tidy JA (2002) Current management of molar pregnancy. J Reprod Med 47: 347-354. [Crossref]
9. Watson EJ, Hernandez E, Miyazawa K (1987) Partial hydatidiform mole: a review. Obstet Gynecol Surv 42: 540-544. [Crossref]
8. Agarwal R, Teoh S, Short D (2012) Chemotherapy and human chorionic gonadotropin
concentrations 6 months after uterine evacuation of molar pregnancy: a retrospective
cohort study. Lancet 379: 130-135. [Crossref]
7. Yuen BH, Cannon W (1981) Molar pregnancy in British Columbia: estimated incidence and postevacuation regression patterns of the beta subunit of human chorionic gonadotropin. Am J Obstet Gynecol 139:216-219.
6. Goto S, Yamada A, Ishizuka T, Tomoda Y (1993) Development of postmolar
trophoblastic disease after partial molar pregnancy. Gynecol Oncol 48: 165-170. [Crossref]
5. Smith HO, Kohorn E, Cole LA (2005) Choriocarcinoma and gestational trophoblastic
disease. Obstet Gynecol Clin North Am 32: 661-684. [Crossref]
4. Garrett LA, Garner EI, Feltmate CM, Goldstein DP, Berkowitz RS (2008) Current management of gestational trophoblastic diseases. Gynecol Oncol 112: 654-662. [Crossref]
3. Darby S, Jolley I, Pennington S, Hancock BW (2009) Does chest CT matter in the staging of GTN? Gynecol Oncol 112: 155-160. [Crossref]
2. FIGO Oncology Committee (2002) FIGO staging for gestational trophoblastic neoplasia
2000. FIGO Oncology Committee. Int J Gynecol Obstet 77: 285-287. [Crossref]
1. FIGO Committee on Gynecologic Oncology. \[Crossref\]
50. Small W Jr, Lurain JR, Shetty RM, Huang CF, Applegate GL, et al. (1996) Gestational trophoblastic disease metastatic to the brain. *Radiology* 200: 277-280. [Crossref]

51. Crawford RA, Newlands E, Rustin GJ, Holden L, A’Hern R, et al. (1997) Gestational trophoblastic disease with liver metastases: the Charing Cross experience. *Br J Obstet Gynaecol* 104: 105-109. [Crossref]

52. Newlands ES, Holden L, Seckl MJ, McNeish I, Strickland S, et al. (2002) Management of brain metastases in patients with high-risk gestational trophoblastic tumors. *J Reprod Med* 47: 465-471. [Crossref]