Coenzyme Q10 and Melatonin for the Treatment of Male Infertility: A Narrative Review

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Abstract: Background: Lifestyle and environmental factors can negatively impact fertility by means of oxidative stress. In this context, antioxidant supplementation therapy has gained much interest in recent years, and different molecules, alone or in combination, have been studied. Objective: The purpose of the present review is to investigate the evidence regarding the efficacy of coenzyme Q10 (CoQ10) and melatonin on male infertility. Methods: A literature search using PUBMED database from 2000 to October 2022 was performed to explore the role of CoQ10 and melatonin on male reproductive function. Conclusions: The analysis involved a narrative synthesis. CoQ10, alone or in combination, appears to reduce testicular oxidative stress and sperm DNA fragmentation and to improve sperm parameters; particularly sperm motility. Moreover, CoQ10 treatment is associated with higher pregnancy rates, both naturally and through assisted reproductive technology (ART). Larger studies are needed to precisely determine its clinical efficacy. Melatonin is a known antioxidant and preclinical studies have shown its ability to modulate reproductive function through hormonal and immune system regulation and sperm cell proliferation. Regardless, clinical studies are necessary to assess its potential in male infertility.

Keywords: coenzyme Q10; melatonin; male reproduction; oxidative stress; supplementation

1. Introduction

Infertility, which is defined as the inability to conceive after at least 12 months of regular, unprotected sexual activity, is estimated to affect between 8 and 12 percent of reproductive-aged couples [1,2]. Male factor infertility (MFI) can be identified in around 50% of cases in this scenario [1,3]. Couple’s infertility is a frequent ailment, and the Global Burden of Disease survey found that, over the past three decades (1990–2017), the age-standardized prevalence of infertility has increased annually by 0.29% in males and 0.35% in women [4].

In the context of male infertility, causes can be classified as (I) congenital (II) acquired and (III) idiopathic, which comprises nearly 30% of infertile couples [5]. Idiopathic infertility is assumed to be associated with risk factors that have a detrimental impact on the reproductive capacity of the male population [6–9]. Oxidative stress, which is known to play a pathogenic role in a variety of diseases [10,11], may also have a relevant impact on spermatozoa’s activity. Hence, lifestyle factors such as smoking, alcohol use, obesity, varicocele, infections and psychological stress, which have been associated with infertility and poor sperm quality [12–14] may indeed exploit their effect through oxidative stress [15–17]. The process is thought to influence between 30 and 80% of subfertility cases, and, for this reason, the category of MOSI (male oxidative stress infertility) was created [18]. In this regard, it was suggested that increased...
oxidative activity may negatively impact the reproductive function by reducing sperm concentration and motility [19]. Accordingly, infertile men display imbalance of the blood oxidation status and the level of reactive oxygen species (ROS) in seminal fluid has been correlated with sperm motility, morphology, and count in astheno- and oligoasthenoteratospermic males [20]. Moreover, reports have linked these factors to sperm DNA fragmentation (SDF), a recurrent disruption observed in idiopathic male infertility [21,22]. As a proof of concept, several authors have investigated the efficacy of antioxidant supplementation (such as L-carnitine, selenium, Coenzyme Q10, ubiquinol, and vitamins C and E) in infertile men, with positive results on sperm quality [23]. However, the limited understanding of MOSI’s etiology has prevented the development of established guidelines regarding its treatment [24].

Coenzyme Q10 (CoQ10) is a fat-soluble ubiquinone with intracellular antioxidant activity, the lack of which has been observed in various sub sub-fertile conditions, (e.g., varicocele, oligozoospermia) [25]. Besides oxidative status balance, the substance is primarily involved in mitochondrial activity and energy-dependent processes, such as sperm motility. Melatonin is a hormone secreted by the pineal gland of the brain [26], whose production is enhanced by darkness, and it is the primary regulator of circadian rhythms. In addition, melatonin and its metabolites act as free radical scavengers, hence protecting cells from oxidative stress, which could have a significant impact on infertility [27].

The purpose of this narrative review is to summarize and analyze the role of CoQ10 and melatonin in the treatment of male infertility.

2. Materials and Methods

The PubMed database was employed for the research. Studies were identified using combinations of the search terms “male reproduction”, “sperm”, “testis”, “gonad”, “fertility”, “semen quality” and “sex hormones”, in combination with “coenzyme Q10”, “melatonin” and “coenzyme”. We decided to limit the search to articles published from January 2000 to October 2022, in order to guarantee the relevance and currency of the paper. Only publications in English were included. Selection criteria included all published randomized controlled trials (RCTs) and non-randomized studies (NRSs) (e.g., observational, prospective, retrospective cohort studies, case-control studies), focusing on CoQ10 and melatonin and male reproductive function. Only clinical studies were considered. Articles identified through references were also included and analyzed. Original and review articles were included. After removing duplicates and papers not relevant to the topic of this article, we identified 61 papers to be included in this narrative review.

The narrative review checklist is reported in the Supplementary Table S1.

3. Results

Results from the retrieved articles are presented in subsections based on the type of treatment.

3.1. Coenzyme Q10

3.1.1. CoQ10 Monotherapy

Based on previous pilot studies, Balercia et. al. [28] investigated the effect of daily 200 mg CoQ10 administration in a randomized, double-blind, placebo-controlled trial conducted on 28 infertile men and 27 control patients. After 6 months, seminal fluid analysis showed improvements in total and forward motility (33.14 ± 7.12% to 39.41 ± 6.80%, p < 0.0001, and 10.43 ± 3.52% to 15.11 ± 7.34%, p < 0.0003, respectively). Moreover, a cause–effect relationship was suggested by the finding of a significant increase in CoQ10 levels both in seminal fluid plasma and sperm cells [28].

In 2009, a larger RCT (106 subjects vs. 106 controls) investigated the efficacy of a daily 300 mg regimen of CoQ10 in infertile men with idiopathic oligoasthenoteratospermia (iOAT) [29]. After 26 weeks, higher total sperm counts (47.8 ± 11.2 × 10⁶ vs. 57.6 ± 14.4 × 10⁶, p = 0.01) and motility (23.1 ± 2.1% vs. 27.6 ± 2.2%, p = 0.01) were found in the CoQ10 group. Notably, the CoQ10 group showed a significant increase in inhibin B levels and a significant
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A decrease in follicle stimulating hormone (FSH) values. Blood and seminal plasma CoQ10 were raised after treatment, and the latter strongly correlated with sperm count ($r = 0.77$, $p = 0.01$), sperm motility ($r = 0.76$, $p = 0.01$) and sperm morphology ($r = 0.54$, $p = 0.02$).

The same group subsequently studied the effect of a 26-week course of 200 mg daily CoQ10 vs. placebo. Compared to placebo, sperm density ($16.8 \pm 4.4 \times 10^6$/mL vs. $28.7 \pm 4.6 \times 10^6$/mL, $p = 0.005$), motility ($25.4 \pm 2.1%$ vs. $35.8 \pm 2.7%$, $p = 0.008$) and morphology ($14.8 \pm 4.1%$ vs. $17.6 \pm 4.4%$, $p = 0.01$) were significantly higher in the treatment group, showing a positive correlation with treatment duration. Moreover, the increase in sperm density and motility remained evident even 3 months after treatment, although less markedly [30].

Another group later examined CoQ10's potential in varicocele-related male infertility. After 3 months of 100 mg CoQ10 daily supplementation, Festa et al. reported a significant improvement in sperm density ($35.5 \pm 3.4 \times 10^6$/mL vs. $42.6 \pm 4.5 \times 10^6$/mL, $p = 0.03$), forward motility ($20.1 \pm 4.5%$ vs. $28.4 \pm 4.9%$, $p = 0.03$) and seminal plasma total antioxidant capacity (TAC) ($106.6 \pm 8.7$ s vs. $148.4 \pm 12.6$ s, $p < 0.01$) [31].

Investigating a dose response effect of CoQ10 integration on semen parameters, an RCT from 2019 by Alahmar et al. observed a stronger effect of a 3-month 400 mg regimen, compared to the standard 200 mg, on progressive and total motility and TAC [33].

Table 1. Summary of studies evaluating CoQ10 monotherapy. TAC: Total antioxidant capacity; CAT: catalase; SOD: superoxide dismutase. FSH = Follicle stimulating hormone; LH = Luteinizing hormone.

| Group               | Number of Patients | Therapy Administered         | Main Results                                                                 |
|---------------------|--------------------|------------------------------|------------------------------------------------------------------------------|
| Balercia et al., [28]| 28                 | CoQ10 200 mg/day for 6 months| • Increase in total and forward motility
• Increased levels of CoQ10 in sperm cells and seminal plasma |
Table 1. Cont.

| Group                             | Number of Patients | Therapy Administered                                               | Main Results                                                                                                                                                                                                 |
|-----------------------------------|--------------------|--------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Safarinejad et al., [29]          | 106                | CoQ10 300 mg/day for 6 and half months                             | • Higher total sperm counts and motility  
• Increase in frequency of acrosome reaction  
• Increase in inhibin B levels and a decrease in FSH and LH levels                                                                                                                                       |
| Safarinejad et al., [30]          | 228                | 114 CoQ10 (ubiquinol) 200 mg/day for 6 and half months 114 placebo | • Increase in sperm density, motility and morphology  
• Increase in inhibin B levels and a decrease in FSH and LH serum levels                                                                                                                                   |
| Festa et al., [31]                | 38                 | CoQ10 100 mg/day for 3 months                                      | • Increase in sperm density and forward motility  
• Increase in total antioxidant capacity                                                                                                                                                                    |
| Nadjarzadeh et al., [32]         | 47                 | CoQ10 200 mg/day for 3 months                                      | • Increase in total antioxidant capacity of seminal plasma.  
• No difference in semen parameters between the CoQ10 and placebo groups                                                                                                                                |
| Alahmar et al., [33]              | 65                 | CoQ10 200 mg/day for 6 months CoQ10 400 mg/day for 6 months        | • In both groups, increase in sperm concentration and progressive and total motility  
• In both groups, increase in TAC, CAT and SOD activity  
• Stronger improvement with higher dosage                                                                                                                                                           |
| Alahmar et al., [34]              | 35                 | CoQ10 200 mg/day for 3 months                                      | • Increase in sperm concentration, progressive sperm motility and total sperm motility  
• Improvement in TAC, SOD and CAT                                                                                                                                                                          |
| Alahmar et al., [35]              | 50                 | CoQ10 (ubiquinol) 200 mg/day for 3 months                         | • Increase in sperm concentration, progressive sperm motility and total sperm motility                                                                                                                                 |
| Alahmar et al., [36]              | 78                 | CoQ10 (ubiquinol) 200 mg/day for 6 months                         | • Increase in semen volume, sperm concentration, progressive and total sperm motility and normal morphology                                                                                               |

3.1.2. CoQ10 Combination Therapy

In 2012, Busetto et al. were the first to prospectively evaluate a combination therapy including CoQ10. In total, 114 patients with iOAT were treated by a single daily dose of a formulation containing multiple antioxidant molecules (L-carnitine, acetyl-L-carnitine, fructose, citric acid, selenium, zinc, ascorbic acid, cyano-cobalamin, folic acid) and 20 mg CoQ10 for 4 months. Only sperm motility significantly increased after treatment (18.3 ± 3.8% to 42.1 ± 5.5%, p < 0.05) [37].

Abad et al. evaluated a commercial multivitamin comprising 20 mg CoQ10, L-carnitine, vitamin C, E, B9, B12, zinc and selenium. In a group of 20 asthenoteratozoospermic men,
3 months of therapy markedly improved type A motility, type A + B motility and vitality, whereas sperm density and normal morphology showed a minor rise \((p = 0.042\) and \(p = 0.04\), respectively). Total and type B motility were unchanged [38].

In 2014, Kobori and colleagues conducted a larger, single-arm study employing a compound of CoQ10, vitamin C and E on 169 iOAT patients. Sperm concentration and motility significantly improved at 3 and 6 months. Sperm motility significantly improved at 3 and 6 months [39]. Moreover, during follow-up, 16 spontaneous pregnancies were achieved in this cohort.

A subsequent study with 20 patients with idiopathic asthenozoospermia evaluated the antioxidant capacity of a combination of CoQ10 (200 mg) and aspartic acid (2660 mg). After a 3-month treatment period, there was a significant improvement in sperm kinetics, but not in sperm count or in the number of atypical sperm cells. Levels of nitric oxide and peroxynitrite in seminal plasma decreased, whereas SOD activity increased. Moreover, the percentage of damaged DNA (assessed by comet assay) decreased significantly [40].

On the other hand, Gvozdjaková et al. observed an increase in sperm density in iOAT patients after 3 (39.8%, \(p < 0.001\)) and 6 months (78.0%, \(p < 0.001\)) of treatment with a multivitamin complex including carnitine, vitamin E and vitamin C and CoQ10 [41].

The effect of a combination of micronutrients (including 15 mg CoQ10 and carnitine) vs. carnitine alone was compared in a prospective, open-label, non-randomized study, by Lipovac et al. in infertile patients with at least one abnormal semen analysis. All the studied sperm parameters significantly improved after 3 months of treatment; however, in the combined micronutrient treatment group, the change of sperm density and progressive motility was higher [42].

In a 2018 double-blind, placebo-controlled trial, 77 infertile men with a high DNA fragmentation index (≥25%) were randomized to receive a commercial fertility supplement containing vitamins and antioxidants (including 10 mg CoQ10) or placebo twice a day for 6 months. After 3 months, the antioxidant group, compared to pre-treatment values, had higher sperm density (median: 24.4 \(\times\) 10⁶/mL vs. 27.2 \(\times\) 10⁶/mL, \(p = 0.028\)) [43].

In 2020, Terai et al. randomized 31 patients with idiopathic male infertility to receive an antioxidant supplement containing CoQ10 (90.26 mg), L-carnitine, zinc, astaxanthin, vitamin C, vitamin B12 and vitamin E or hochu-ekki-to (a Chinese herbal medicine). Sperm analysis and LH, FSH and testosterone serum concentrations were performed before and after 3 months of treatment. Both groups did not show any endocrinological or semen parameters increase. Total motile count was the only semen parameter to show a significant increase after treatment in the supplement group \((p = 0.04)\), whereas no significant changes in hormonal parameters were observed [44].

Arafa et al. have recently evaluated the effect of antioxidant supplementation on conventional semen parameters and advanced sperm function tests in a population of infertile men [45]. A total of 148 subjects (119 in the idiopathic male infertility and 29 in the unexplained infertility group) received a 3-month treatment with three capsules twice a day of an antioxidant formula which provided, among others, 200 mg CoQ10 per day. Sperm analysis revealed an improvement in all parameters investigated, except for semen volume and sperm viability.

In 2020, Sadaghiani et al. enrolled, in their single-blinded trial, 50 oligospermic and asthenospermic patients, who were active smokers, to evaluate the effect of antioxidant supplementation (30 mg CoQ10, 8 mg zinc, 100 mg vitamin C, 12 mg vitamin E, 400 µg folic acid once a day and 200 mg selenium every other day) on semen parameters [46]. After 3 months of supplementation, mean sperm volume and total sperm count increased from 3.48 ± 1.44 to 3.71 ± 1.42 mL \((p = 0.032)\) and from 21.76 ± 23.02 \(\times\) 10⁶ to 23.22 ± 23.28 \(\times\) 10⁶ \((p = 0.001)\), respectively. Additionally, total sperm motility, progressive motility and normal sperm morphology increased from 27.22 ± 13.69% to 31.85 ± 5.82% \((p = 0.001)\), from 9.82 ± 9.10% to 11.57 ± 10.18% \((p = 0.001)\) and from 23.22 ± 23.28% to 33.60 ± 20.01% \((p = 0.003)\), respectively [46].
The effect of a dietary supplement containing a mix of antioxidants and vitamins (including 40 mg CoQ10) versus placebo (1:1) was studied in the randomized, double-blind, placebo-controlled study by Kopets et al. [47]. Eighty-three patients with idiopathic male infertility were enrolled and randomized to receive a 6-month treatment. Normalization of semen analysis at 0, 2 and 4 months was the primary outcome, while the pregnancy rate was the secondary outcome. At 4 months, 69.0% of the patients in the treatment group and 22.0% in the placebo group had normal semen analysis ($p < 0.001$), whereas the pregnancy rate was significantly higher in the treatment than the placebo group (23.8% and 4.9%, $p = 0.017$) [47].

In 2021, Nazari et al. conducted an open study on 180 iOAT patients who received daily treatment of a combination of antioxidants (including 40 mg CoQ10) for 12 weeks. Sperm motility was unchanged, though sperm density ($25 \text{ vs. } 36 \times 10^6$/mL, $p = 0.004$) and morphology ($p = 0.01$) improved [48].

Similarly, Gual-Frau et al. analyzed the impact of a multivitamin compound with 20 mg of CoQ10 in infertile men with grade I varicocele [49]. They found that only the total number of sperm increased after treatment.

More recently, Ma et al. [50] performed a single-blind RCT with patients randomly allocated to receive L-carnitine complex nutrient treatment (study group—15 g/bag, orally one bag at a time, twice a day, $n = 73$) or CoQ10 (control group—10 mg tablet orally, thrice daily, $n = 70$) with Vitamin E (100 mg tablet orally, thrice daily) for three months. They found that CoQ10 plus Vitamin E therapy resulted in improvement of sperm motility, morphology and testosterone levels. Conversely, L-carnitine significantly improves sperm motility, morphology and concentration, while also improving testosterone and LH levels.

A summary of the studies evaluating CoQ10 combination therapy is presented in Table 2.

3.1.3. Effect of CoQ10 on Sperm DNA Fragmentation

Sperm DNA fragmentation (SDF) has progressively gained clinical importance in terms of reproductive outcomes both under natural and assisted reproductive technology (ART) conditions [51]. Several treatments of male infertility aim to reduce SDF values in order to improve the couple’s reproductive chance. While antioxidant therapy emerged as an effective option to improve semen parameters, its impact on SDF is still a matter of debate [52]. Here, we analyzed the literature with the specific focus of the effect of CoQ10 on SDF.

Antioxidant treatment was found to be effective in reducing SDF values in a study by Nadjarzadeh and colleagues [32]. Furthermore, in the subgroup with increased pretreatment SDF levels, supplementation increased the success rate of intracytoplasmic sperm injection (ICSI) [32].

Similarly, two years later, Abad et al. showed that a multicomponent antioxidant treatment could significantly diminish the progression of sperm DNA fragmentation over increasing incubation time; moreover, the proportion of sperm cells bearing degraded DNA was significantly reduced ($7.32 \pm 4.12\% \text{ vs. } 5.66 \pm 3.21\%, p = 0.04$) [38].

In 2015, Gual-Frau et al. investigated the effect of a multivitamin compound containing CoQ10 in infertile patients affected by varicocele and with high SDF levels. Three months of treatment lead to a 22.1% decrease in SDF ($p = 0.02$) and a 31.3% reduction in DNA-degraded sperm cells ($p = 0.07$) [49].

In a subsequent study, Alahmar et al. administered a 3-month course of 200 mg CoQ10 to 65 patients affected by idiopathic OA related infertility and 40 fertile subjects. The results showed an improvement in semen parameters and a reduction in OS markers and SDF in the former group [53].
Table 2. Studies evaluating CoQ10 in combination with other supplements. ROS: reactive oxygen species; SOD: superoxide dismutase; SDF: Sperm DNA fragmentation; ORP: oxidation reduction potential.

| Group | Patients | Therapy Administered | Main Results |
|-------|----------|----------------------|--------------|
| Busetto et al., [37] | 114 | L-carnitine, acetyl-L-carnitine, fructose, citric acid, selenium, zinc, ascorbic acid, cyanocobalamin, folic acid and 20 mg CoQ10 daily for 4 months | • Increase in sperm motility |
| Abad et al., [38] | 20 | 1500 mg L-carnitine, 60 mg vitamin C, 10 mg vitamin E, 200 µg vitamin B9, 1 µg vitamin B12, 10 mg zinc, 50 mg selenium and 20 mg CoQ10 daily for 3 months | • Improvement in type A + B motility, vitality, sperm density and normal morphology<br>• Reduction in DNA degraded sperm |
| Kobori et al., [39] | 169 | 80 mg vitamin C and 40 mg vitamin E and 120 mg CoQ10 daily for 6 months | • Increase in sperm motility and sperm concentration |
| Tirabassi et al., [40] | 20 | Aspartic acid 2660 mg and 200 mg CoQ10 daily for 3 months | • Improvement in sperm kinetics and increase in SOD activity<br>• Decrease in ROS levels in seminal plasma<br>• Correlation of seminal fluid oxidation state and DNA damage index |
| Gvozdjaková et al., [41] | 40 | 440 mg L-carnitine fumarate, 75 IU vitamin E, 12 mg vitamin C and 30 mg CoQ10 daily for 6 months | • Increase in sperm density<br>• Increase in seminal fluid CoQ10 concentration<br>• Decreased oxidative stress parameters |
| Lipovac et al., [42] | 143 group 1 156 group 2 | 440 mg L-carnitine, 250 mg L-arginine, 40 mg zinc, 120 mg vitamin E, 80 mg glutathione, 60 µg selenium, 800 µg folic acid and 15 mg CoQ10 daily for 3 months VS 500 mg l-carnitine/twice a day alone | • Improved semen parameters in the multivitamin group |
| Stenqvist et al., [43] | 37 group 1 40 group 2 | Vitamin C 30 mg, vitamin E5 mg, vitamin B12 0.5 lg, L-carnitine 750 mg, and folic acid 100 lg, zinc 3 mg, selenium 25 lg, with maltodextrin, calcium carbonate, citric acid, steviol glycoside, flavors, beta-carotene, silicon dioxide and coenzyme Q10 10 mg twice/daily VS placebo for 6 months | • Increased sperm density in group 1<br>• No differences in semen parameters or SDF |
| Terai et al., [44] | 15 group 1 16 group 2 | L-Carnitine 750.1 mg, Zinc 30 mg, Astaxanthin 16.05 mg, Vitamin C 1000 mg, Vitamin B12 60.1 µg, Vitamin E 150 mg and CoQ10 90.26 mg vs. Chinese herbal medicine hochu-ekki-to (HE), 3 times/daily for 3 months | • No difference in endocrinological or semen parameters<br>• In group 1 increase of total motile count |
| Arafa et al., [45] | 148 group 1 idiopathic infertility 29 group 2 unexplained infertility | Vitamin C, 120 mg vitamin D3, 1200 IU vitamin E (as mixed tocopherols), 200 IU vitamin K, 80 µg thiamin, 3 mg, riboflavin, 3.4 mg niacin, 20 mg vitamin B6, 25 mg folic acid, 800 µg vitamin B12, 1000 µg biotin, 600 µg pantothenic acid, 20 mg iodine, 150 µg zinc, 30 mg selenium, 140 µg copper, 1 mg manganese, 2 mg chromium, 120 µg molybdenum, 75 µg L-carnitine tartrate, 2000 mg L-arginine, 350 mg CoQ10, 200 mg N-acetyl L-cysteine, 200 mg grapeseed extract, 20 mg lycopene, 10 mg and benfotiamine 1 mg daily for 3 months | Group 1: <br>• Decrease in SDF values and ORP.<br>• Improvement of sperm parameters<br>Group 2: Improvement in progressive motility only |
| Sadaghiani et al., [46] | 50 | 30 mg CoQ10, 8 mg zinc, 100 mg vitamin C, 12 mg vitamin E, 400 µg folic acid once a day and 200 mg selenium every other day for 3 months | • Increase in mean volume of sperm and total sperm count<br>• Increased total sperm motility, progressive motility and normal sperm morphology |
| Kopets et al., [47] | 42 group 1 supplementation 41 group 2 placebo | L-carnitine/L-acetyl-carnitine 1990 mg, L-arginine 250 mg, glutathione 100 mg, co-enzyme Q10 40 mg, zinc 7.5 mg (75% of recommended daily allowance, RDA), vitamin B9 234 µg (117% RDA), vitamin B12 2 µg (80% RDA), selenium 50 mcg (91% RDA), excipients sorbitol, maltodextrin, orange/beta-carotene colorant, saccharide, acesulfame potassium, and silicon dioxide vs. placebo daily for 6 months | • 69.0% vs. 22.0% normal semen analysis in study group vs. placebo<br>• 23.8% vs. 4.9% spontaneous pregnancies in study group vs. placebo |
| Group             | Patients | Therapy Administered                                                                 | Main Results                                           |
|-------------------|----------|--------------------------------------------------------------------------------------|--------------------------------------------------------|
| Nazari et al., [48] | 180      | 1500 mg of L-Carnitine, 60 mg of vitamin C, 20 mg of coenzyme Q10, 10 mg of vitamin E, 10 mg of zinc, 200 µg of vitamin B9, 50 µg of selenium, 1 µg of vitamin B12 twice/daily for 3 months | Increase in sperm density and morphology               |
| Gual-Frau et al., [49] | 20       | 1500 mg L-Carnitine, 60 mg vitamin C, 20 mg coenzyme Q10, 10 mg vitamin E, 200 µg vitamin B9, 1 µg vitamin B12, 10 mg zinc, 50 µg selenium | Increase in total number of sperm, but other semen parameters were unaffected |
| Ma et al., [50]     | 70       | CoQ10 10 mg tablet orally, thrice daily plus Vitamin E 100 mg tablet orally, thrice daily, for three months | Improvement in sperm motility, morphology and testosterone levels |
On the other hand, an RCT from 2018 by Stenqvis et al. failed to confirm these findings. Seventy-seven infertile men received a compound containing, among others, 10 mg CoQ10 for 6 months. At the end of the treatment phase SDF and other sperm parameters were comparable between groups, although sperm density did rise in the antioxidant group compared to pre-treatment [43].

More recently, Arafa et al. conducted a trial categorizing subjects with idiopathic (119 patients) and unexplained male infertility (29 patients). The 148 subjects were administered a course of a mixed supplement composed of 200 mg CoQ10 and other antioxidants. In the first group, 3 months of treatment lead to an improvement in SDF and all semen parameters (with the exception of sperm volume and viability). On the other hand, SDF was stable in the unexplained infertility group [45].

As previously reported, Alahmar et al. investigated 50 patients with iOAT treated with a daily dose of 200 mg of CoQ10 and 50 fertile men (controls) [35]. They found a significant decrease in SDF levels after treatment in infertile men (38.6% ± 7.9 vs. 34.5% ± 9.3, \( p < 0.001 \)).

3.1.4. CoQ10: Effect on Natural Pregnancy and ART

Different authors have described an increase in pregnancy rates after CoQ10 administration [54,55] and this has been attributed to its positive effect on sperm concentration and motility. For instance, both Abad and Kobori have reported natural pregnancies after a combined antioxidant treatment which improved semen parameters [38,39].

Similarly, Kopets and colleagues studied the effect of a dietary supplement containing a mix of antioxidants and vitamins (including 40 mg CoQ10) in a 1:1 RCT. Four-month normalization of semen parameters was significantly more frequent in the treatment group (69.0 vs. 22.0%, \( p < 0.001 \)), as it was for pregnancy rate (23.8% and 4.9%, \( p = 0.017 \)) [47].

In a prospective series published in 2012, Safarinejad et al. enrolled 287 iOAT patients to be administered a 12-month course of daily 300 mg CoQ10. Patients were then kept on follow up for an additional year. Mean treatment to pregnancy time was 8.4 ± 4.7, with a total pregnancy rate of 34.1%; semen parameters were linearly associated with the latter [56].

Again, focusing on combination therapy, Gvozdjakova and colleagues investigated the efficacy of a mixed compound of carnitine, CoQ10 and vitamin C and E on OA-related infertility. After treatment, a 45% overall pregnancy rate, 7.5% of which through ART, was reported [41].

As previously reported, Alahmar et al. [36] investigated the effect of 200 mg CoQ10 for 6 months in 78 iOA infertile men vs. controls. Treatment significantly improved semen parameters, antioxidant biomarkers and SDF. The pregnancy rate was 24.2%, and time to pregnancy (TTP) was 20.52 ± 6.72 months following 6 months of CoQ10 therapy and another 18 months of follow-up. CoQ10 level, male age, semen parameters and ROS levels were found to be predictors of pregnancy outcomes and TTP.

3.2. Melatonin

3.2.1. Melatonin: Role in the Testis

In addition to its well-known effect on daily and annual rhythmicity, melatonin has been found to regulate reproductive function via modulating the release of gonadotropin-releasing hormones (GnRH) [57]. Furthermore, melatonin is absorbed by the testis, where it affects testicular function directly [58,59] by acting on receptors present on Leydig [60], Sertoli [61,62] and intratesticular inflammatory [63] cells. Intriguingly, melatonin and testosterone share comparable circadian rhythms, and in animal species, this hormone influences various aspects of testicular function to facilitate seasonal reproduction [64,65]. Specifically, melatonin was found to inhibit GnRH-induced testosterone release [60,66,67] and to stimulate the testicular conversion of testosterone into dihydrotestosterone (DHT) [68]. Subsequent studies demonstrated that this may occur via a local corticotropin-releasing hormone (CRH) system, all components of which were then identified in the testes of infertile men [67]. This indolamine
also protects the testis from local inflammatory processes and ROS. Consequently, melatonin exerts anti-proliferative and anti-inflammatory effects on testicular macrophages [69] and mast cells [63], and its testicular concentrations demonstrated a negative correlation with the concentration of pro-apoptotic molecules [63] and the number of macrophages [70] in biopsies from infertile patients. In addition, melatonin functions as a free radical scavenger [71], preventing apoptosis and restoring testicular function [72–74]. Accordingly, melatonin treatment reduces the severity of induced testicular damage in animal models [75–81]. Lastly, the hormone acts on Sertoli cells by enhancing their responsiveness to follicle stimulating hormone during development [82] and by inducing the secretion of growth factors active on spermatogonial stem cells proliferation [83–85].

Few clinical studies have investigated the impact of melatonin treatment on semen parameters (Table 3). Awad and colleagues investigated a possible correlation in both fertile and infertile men, categorizing the latter group by semen analysis alterations. The authors observed lower melatonin levels in men affected by non-obstructive azoospermia, in those displaying impaired sperm motility and leucocytospermia (the levels of whom were lowest) [86]. In 2016, Kratz et al. measured seminal plasma concentrations of melatonin and oxidative stress parameters in fertile normozoospermic and infertile terato- or azoospermic infertile subjects. The authors report higher levels of seminal plasma melatonin in normozoospermic fertile men, with no difference between different subgroups of infertile men. Moreover, infertile subjects displayed increased concentrations of oxidation products in seminal fluid, with azoospermic patients displaying the highest levels [87]. The study once again suggests the possible relation between melatonin and oxidative stress in the infertile male.

Table 3. Studies evaluating the association between melatonin and semen parameters. OA = Oligoasthenozoospermia; NOA = Non-obstructive azoospermia.

| Group             | Number of Patients                                                                 | Main Results                                                                 |
|-------------------|------------------------------------------------------------------------------------|------------------------------------------------------------------------------|
| Awad et al., 2006 | • Fertile normozoospermic men (n = 20) <br>• OA (n = 20) <br>• OA with leucocytospermia (n = 20) <br>• OA with varicocele (n = 20) <br>• NOA with high FSH (n = 20) <br>• NOA with normal FSH (n = 20) | • Serum and seminal plasma melatonin levels in all infertile groups were reduced significantly compared with their levels in the fertile group  <br>• OA with leucocytospermia had the lowest levels of melatonin <br>• Melatonin was positively correlated with sperm motility |
| Kratz et al., 2016| • azoospermia (n = 37) <br>• theratozoospermia (n = 29) <br>• fertile controls (normozoospermia, n = 37) | • Melatonin was lower in azoospermic (p < 0.0001) and theratozoospermic (p < 0.0001) patients versus fertile men |
| Lu et al., 2018   | 27 infertile men receiving 400 mg of melatonin for 3 months and 27 receiving placebo for 3 months after varicocelectomy | • Sperm concentration, motility and proportions of normally formed spermatozoa significantly improved in melatonin group compared with placebo group |

The potential clinical benefit of melatonin supplementation on semen parameters was tested after varicocele treatment by Lu and colleagues [88]. Fifty-four infertile, mildly oligospermic men affected by varicocele were randomized to receive either 400 mg of melatonin or placebo after varicocele treatment. At 3 and 6 months, the patients treated by melatonin showed a stronger improvement in terms of semen parameters (sperm
concentration, motility and proportions of normally formed spermatozoa), peripheral blood inhibin B levels and total antioxidant compared with the placebo group.

3.2.2. Melatonin in ART Outcomes

The role of melatonin on female reproduction has been more extensively studied. For instance, in a systematic review and meta-analysis from 2020, Hu and colleagues described the role of melatonin in ART. The authors concluded that melatonin is indeed associated with higher embryo quality and clinical pregnancy rate, though not with a more significant live birth rate [27]. Besides these findings, which are beyond the scope of this review, in this context, melatonin was shown to act, among other mechanisms, on inflammation, apoptosis and oxidative stress modulation [89,90]. These observations, thus, strengthen the ones made on male reproductive system, thus confirming this hormone’s potential.

4. Discussion

Supplementation therapy has been widely employed in the context of infertility as a mean to improve the success rate in achieving pregnancy both spontaneously and by means of ART. In this context, the evidence strongly suggests that oxidative stress plays a central role in idiopathic infertility [15–17], thus raising interest towards antioxidant therapy in this field.

Coenzyme Q10 is involved in mitochondrial energy production and has a key role as an antioxidant for cell membranes and lipoproteins. For this reason, different authors have raised interest towards its use in the field of reproductive medicine. As discussed in this review, CoQ10 monotherapy was shown to enhance sperm motility and concentration as well as improving sperm DNA fragmentation in infertile men. Moreover, the results also suggest that CoQ10 may itself be effective in improving conception rates. The improvement in semen parameters reported by many authors was then followed by a higher frequency of spontaneous pregnancy and better outcomes for ART. Nonetheless, it must be noted that these were mostly assessed as secondary outcomes, and one should be careful in interpreting these results. In fact, despite single studies suggesting a positive effect of CoQ10 on pregnancy outcomes, a previous systematic review and meta-analysis revealed that there is no evidence that CoQ10 increases either live birth or pregnancy rates, but there is a global improvement in sperm parameters [91].

Similar to other molecules, CoQ10 use in male infertility showed premising results, but studies are of low evidence, mainly with small sample size and with a wide range of dosage and timing of administration, thus precluding its widespread use [92]. In particular, it must be noted that results from studies dealing with combination therapy are quite heterogeneous. First, the dose of CoQ10 in these studies is generally lower (range 10–200 mg) compared to that investigated in monotherapy (200–300 mg); second, the presence of multiple antioxidant agents makes hard the interpretation of which is the real responsible for the biological effect; finally, the duration of treatment was not standardized. As a whole, caution should be used when interpreting results from studies dealing with antioxidant cocktails.

Similarly, we analyzed the existing evidence concerning the role of melatonin in male infertility, although the data regarding this molecule is limited. Besides its role as a free radical scavenger, and thus, its antioxidant potential, which might itself justify its employment, melatonin might have a more complex role. Animal models have in fact shown that this molecule may contribute to gonadal physiology by modulating androgen production (both centrally and locally), by acting as an immunomodulatory compound and by influencing the progression of germ cells to spermatozoa. Clinical data in men are scarce, whereas there is wider evidence regarding its efficacy on female fertility from ovarian aging to clinical improvement on in vitro fertilization success rate. In summary, preclinical data regarding melatonin show potential in the context of male infertility, though clinical studies are needed in order to assess the concrete possibility of its employment.
As previously mentioned, in an attempt to counteract the deleterious effects of OS, urologists resorted to antioxidant supplementation. Considering that indiscriminate and empirical use of these compounds can exert detrimental effects through a reduction state, physicians need to be aware of the “antioxidant paradox” [93]. The latter was termed by Halliwell et al. to describe that the overuse of antioxidants has no preventative or therapeutic effect at all. In this context, Symeonidis et al. [94], in a recent review, highlighted the need for redox balance, thus safeguarding redox homeostasis. This article signifies that moderation is the key to optimal sperm regulation.

5. Conclusions

In recent years, a growing body of literature has shown that an altered redox balance in seminal fluid may display deleterious effects on sperm homeostasis, leading to male infertility [95,96]. The present narrative review shows the antioxidant properties of CoQ10 and its beneficial role on semen parameters and sperm DNA fragmentation. Moreover, CoQ10 administration in couples resulted in improved ART outcomes, such as increased fertilization rates in IVF/ICSI.

Further in-depth interventions are needed to reveal the exact mechanism of action of CoQ10 and to determine the appropriate standardized dose and duration of CoQ10 supplementation in the treatment of specific male infertility cases. Additional evidence is needed to further support the treatment of male infertility with melatonin supplementation.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/nu14214585/s1, Table S1: Narrative Review Checklist.

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References
1. Vander, M.; Borght, E.C. Wyns, Fertility and infertility: Definition and epidemiology. Clin. Biochem. 2018, 62, 2–10. [CrossRef] [PubMed]
2. Barratt, C.L.; Björndahl, L.; De Jonge, C.J.; Lamb, D.J.; Osorio Martini, F.; McLachlan, R.; Oates, R.D.; van der Poel, S.; St John, B.; Sigman, M.; et al. The diagnosis of male infertility: An analysis of the evidence to support the development of global WHO guidance—Challenges and future research opportunities. Hum. Reprod. Update 2017, 23, 660–680. [CrossRef] [PubMed]
3. Zegers-Hochschild, F.; Adamson, G.D.; Dyer, S.; Racowsky, C.; De Mouzon, J.; Sokol, R.; Rienzi, L.; Sunde, A.; Schmidt, L.; Cooke, I.D.; et al. The International Glossary on Infertility and Fertility Care, 2017. Fertil. Steril. 2017, 108, 393–406. [CrossRef] [PubMed]
4. Sun, H.; Gong, T.-T.; Jiang, Y.-T.; Zhang, S.; Zhao, Y.-H.; Wu, E.Q.-J. Global, regional, and national prevalence and disability-adjusted life-years for infertility in 195 countries and territories, 1990-2017: Results from a global burden of disease study, 2017. Aging 2019, 11, 10952–10991. [CrossRef] [PubMed]
5. Minhas, S.; Bettocchi, C.; Boeri, L.; Capogrosso, P.; Carvalho, J.; Cilesiz, N.C. European Association of Urology Guidelines on Male Sexual and Reproductive Health: 2021 Update on Male Infertility. Eur. Urol. 2021, 80, 603–620. [CrossRef]
6. Gunes, S.; Arslan, M.A.; Hekim, G.N.T.; Asci, E.R. The role of epigenetics in idiopathic male infertility. J. Assist. Reprod. Genet. 2016, 33, 553–569. [CrossRef]
7. Muneer, A.; Pozzi, E.; Cakir, O.O. The role of nitric oxide (NO) donors in the treatment of male infertility. Curr. Pharm. Des. 2020. [CrossRef]
8. Alfano, M.; Pedercizi, F.; Locatelli, I.; Ipollito, S.; Longhi, E.; Zerbi, P. Impaired testicular signaling of vitamin A and vitamin K contributes to the aberrant composition of the extracellular matrix in idiopathic germ cell aplasia. *Fertil. Steril.* 2019, 111, 687–698. [CrossRef]

9. Adamopoulos, D.A.; Pappa, A.; Billia, E.; Nicopoulou, S.; Koukkou, E.; Michopoulos, J. Effectiveness of combined tamoxifen citrate and testosterone undecanoate treatment in men with idiopathic oligozoospermia. *Fertil. Steril.* 2003, 80, 914–920. [CrossRef]

10. Burton, G.J.; Jauniaux, E. Oxidative stress. *Best Pract. Res. Clin. Obstet. Gynaecol.* 2020, 55, 287–299. [CrossRef]

11. Kruk, J.; Aboul-Enein, H.Y.; Kładna, A.; Bowser, J.E. Oxidative stress in biological systems and its relation with pathophysiological functions: The effect of physical activity on cellular redox homeostasis. *Free Radic. Res.* 2019, 53, 497–521. [CrossRef] [PubMed]

12. Boeri, L.; Capogrosso, P.; Ventimiglia, E.; Pedercizi, F.; Frego, N.; Cazzaniga, W.; Chierigo, F.; Alfano, M.; Piemonti, L.; Viganò, P.; et al. Undiagnosed prepubertal diabetes is highly prevalent in primary infertile men—results from a cross-sectional study. *BJU Int.* 2019, 123, 1070–1077. [CrossRef] [PubMed]

13. La Vignera, S.; Condorelli, R.; Vicari, E.; D’Agata, R.; Calogero, A.E. Diabetes mellitus and sperm parameters. *J. Androl.* 2012, 33, 145–153. [CrossRef] [PubMed]

14. Boeri, L.; Capogrosso, P.; Ventimiglia, E.; Pedercizi, F.; Cazzaniga, W.; Chierigo, F.; Dehò, F.; Montanari, E.; Montorsì, F.; Salonia, A. Heavy cigarette smoking and alcohol consumption are associated with impaired sperm parameters in primary infertile men. *Asian J. Androl.* 2019, 21, 478. [CrossRef]

15. Amiri, I.; Najafi, R.; SHEYKH, N. Nitric oxide level in seminal plasma and its relation with sperm DNA damages. *Iran. Biomed. J.* 2007, 11, 259–264.

16. Lopes, F.; Pinto-Pinho, P.; Gaivão, I.; Martins-Bessa, A.; Gomes, Z.; Moutinho, O.; Oliveira, M.M.; Peixoto, F.; Pinto-Leite, R. Sperm DNA damage and seminal antioxidant activity in subfertile men. *Andrologia* 2021, 53, e14027. [CrossRef]

17. Boeri, L.; Capogrosso, P.; Ventimiglia, E.; Pedercizi, F.; Cazzaniga, W.; Chierigo, F.; Pozzi, E.; Clementi, M.; Viganò, P.; Montanari, E.; et al. High-risk human papillomavirus in semen is associated with poor sperm progressive motility and a high sperm DNA fragmentation index in infertile men. *Hum. Reprod.* 2019, 34, 209–217. [CrossRef]

18. Showell, M.G.; Mackenzie-Proctor, R.; Brown, J.; Yazdani, A.; Stankiewicz, M.T.; Hart, R.J. Antioxidants for male subfertility. *Cochrane Database Syst. Rev.* 2014, 12, CD007411. [CrossRef]

19. Agarwal, A.; Virk, G.; Ong, C.; Du Plessis, S.S. Effect of oxidative stress on male reproduction. *World J. Mens Health* 2014, 32, 1–17. [CrossRef]

20. Colagar, A.H.; Karimi, F.; Jorsaraei, S.G.A. Correlation of sperm parameters with semen lipid peroxidation and total antioxidants levels in astheno- and oligoagsheno- teratospermic men. *Iran. Res Crescent Med. J.* 2013, 15, 780–785. [CrossRef]

21. Nadjarzadeh, A.; Shidfar, F.; Amirjannati, N.; Vafa, M.R.; Motievalian, S.A.; Gohari, M.R.; Nazeri Kakhki, S.A.; Akhondi, M.M.; Sadeghi, M.R. Effect of Coenzyme Q10 supplementation on antioxidant enzymes activity and oxidative stress of seminal plasma: A double-blind randomised clinical trial. *Andrologia* 2014, 46, 177–183. [CrossRef] [PubMed]

22. Panner Selvam, M.K.; Sengupta, P.; Agarwal, A. Sperm DNA Fragmentation and Male Infertility. In *Genetics of Male Infertility*; Arafa, M., Elbardisi, H., Majzoub, A., Agarwal, A., Eds.; Springer: Cham, Germany, 2020; pp. 155–172.

23. Gambera, L.; Stendardi, A.; Ghelardi, C.; Fineschi, B.; Aini, R. Effects of antioxidant treatment on seminal parameters in patients undergoing in vitro fertilization. *Arch. Ital. Urol. Androl. Organo Uff. Soc. Ital. Ecogr. Urol. E Nefrol.* 2009, 81, 5–13. [CrossRef]

24. Camarda, G.; Stendardi, A.; Ghelardi, C.; Fineschi, B.; Aini, R. Effects of antioxidant treatment on seminal parameters in patients undergoing in vitro fertilization. *J. Endocrinol. Invest.* 2009, 32, 626–632. [CrossRef] [PubMed]

25. Agarwal, A.; Parekh, N.; Selvam, M.K.P.; Henkel, R.; Shah, R.; Homa, S.T.; Ramasamy, R.; Ko, E.; Tremellen, K.; Esteves, S. Male Oxidative Stress Infertility (MOSI): Proposed Terminology and Clinical Practice Guidelines for Management of Idiopathic Male Infertility. *World J. Mens Health* 2019, 37, 296–312. [CrossRef] [PubMed]

26. Balercia, G.; Mancini, A.; Paggi, F.; Tiano, L.; Pontecorvi, A.; Boscaro, M.; Lenzi, A.; Littarru, G.P. Coenzyme Q10 and male infertility. *Endocrinol. Invest.* 2009, 32, 626–632. [CrossRef] [PubMed]

27. Pereira, N.; Naufel, M.F.; Ribeiro, E.B.; Tufik, S.; Hachul, H. Influence of Dietary Sources of Melatonin on Sleep Quality: A Review. *J. Food Sci.* 2020, 85, 5–13. [CrossRef]

28. Hu, K.L.; Ye, X.; Wang, S.; Zhang, D. Melatonin Application in Assisted Reproductive Technology: A Systematic Review and Meta-Analysis of Randomized Trials. *Front. Endocrinol.* 2020, 11, 160. [CrossRef] [PubMed]

29. Balercia, G.; Buldregghi, E.; Vignini, A.; Tiano, L.; Paggi, F.; Amoroso, S.; Ricciardi-Lamonica, G.; Boscaro, M.; Lenzi, A.; Littarru, G. Coenzyme Q10 treatment in infertile men with idiopathic asthenozoospermia: A placebo-controlled, double-blind randomized trial. *Fertil. Steril.* 2009, 91, 1785–1792. [CrossRef]

30. Safarinejad, M.R. Efficacy of coenzyme Q10 on semen parameters, sperm function and reproductive hormones in infertile men. *J. Urol.* 2009, 182, 237–248. [CrossRef]

31. Festa, R.; Giacchi, E.; Raimondo, S.; Tiano, L.; Zuccherelli, P.; Silvestrini, A.; Meucci, E.; Littarru, G.P.; Mancini, A. Coenzyme Q10 supplementation in infertile men with low-grade varicocele: An open, uncontrolled pilot study. *Andrologia* 2014, 46, 805–807. [CrossRef]
32. Nadjarzadeh, A.; Sadeghi, M.R.; Amirjannati, N.; Vafa, M.R.; Motievalian, S.A.; Gohari, M.R.; Akhondi, M.A.; Yavari, P.; Shidfar, F. Coenzyme Q10 improves seminal oxidative defense but does not affect on semen parameters in idiopathic oligoasthenoteratozoosperma: A randomized double-blind, placebo controlled trial. J. Endocrinol. Invest. 2011, 34, e222–e228. [CrossRef]

33. Alahmar, A.T.; Sengupta, P. Impact of Coenzyme Q10 and Selenium on Seminal Fluid Parameters and Antioxidant Status in Men with Idiopathic Infertility. Biol. Trace Elem. Res. 2021, 199, 1246–1252. [CrossRef] [PubMed]

34. Alahmar, A.T.; Sengupta, P.; Dutta, S.; Calogero, A.E. Coenzyme Q10, oxidative stress markers, and sperm DNA damage in men with idiopathic oligoasthenoteratozoosperma. Clin. Exp. Reprod. Med. 2021, 48, 150–155. [CrossRef] [PubMed]

35. Alahmar, A.T.; Sengupta, P.; Dutta, S.; Calogero, A.E. Coenzyme Q10, oxidative stress markers, and sperm DNA damage in men with idiopathic oligoasthenoteratozoosperma. Clin. Exp. Reprod. Med. 2021, 48, 150–155. [CrossRef] [PubMed]

36. Alahmar, A.T. Naemi, R. Predictors of pregnancy and time to pregnancy in infertile men with idiopathic oligoasthenospermia. Clin. Exp. Reprod. Med. 2020, 47, 112–118. [CrossRef] [PubMed]

37. Busetto, G.M.; Koverech, A.; Messano, M.; Antonini, G.; de Berardinis, E.; Gentile, V. Prospective open-label study on the efficacy and tolerability of a combination of nutritional supplements in primary infertile patients with idiopathic astenoteratozoosperma. Arch. Ital. Urol. Androl. Organo Uff. Soc. Ital. Ecogr. Urol. E Nefrol. 2012, 84, 137–140.

38. Abad, C.; Amengual, M.J.; Gosalbez, J.; Coward, K.; Hannauoi, N.; Benet, J.; Garcia-Peiró, A.; Prats, J. Effects of oral antioxidant treatment upon the dynamics of human sperm DNA fragmentation and subpopulations of sperm with highly degraded DNA. Andrology 2013, 45, 211–216. [CrossRef]

39. Kobori, Y.; Ota, S.; Sato, R.; Yagi, H.; Sob, S.; Arai, G.; Okada, H. Antioxidant cosupplementation therapy with vitamin C, vitamin E, and coenzyme Q10 in patients with oligospermia. Arch. Ital. Urol. Androl. Origo Uff. Soc. Ital. Ecogr. Urol. E Nefrol. 2014, 86, 1–4. [CrossRef]

40. Tirabassi, G.; Vignini, A.; Tiano, L.; Buldrehgini, E.; Bruge, F.; Silvestri, S.; Orlando, P.; D’Aniello, A.; Mazzanti, L.; Lenz, A.; et al. Protective effects of coenzyme Q10 and aspartic acid on oxidative stress and DNA damage in subjects affected by idiopathic asthenospermia. Endocrine 2015, 49, 549–552. [CrossRef]

41. Gvozdjaková, A.; Kucharská, J.; Dubravický, J.; Mojto, V.; Singh, R.B. Coenzyme Q10, α-tocopherol, and oxidative stress could be important metabolic biomarkers of male infertility. Dis. Markers 2015, 2015, 827941. [CrossRef]

42. Lipovac, M.; Bodner, F.; Imhof, M.; Chedraui, P. Comparison of the effect of a combination of eight micronutrients versus a standard mono preparation on sperm parameters. Reprod Biol Endocrinol. RBE. Dicembre 2016, 14, 84.

43. Senuqvist, A.; Oleszczuk, K.; Leijonhufvud, I.; Giwercman, A. Impact of antioxidant treatment on semen parameters and antioxidant status in men with idiopathic oligoasthenoteratozoosperma. Clin. Exp. Reprod. Med. 2019, 46, 112–118. [CrossRef] [PubMed]

44. Terai, K.; Horie, S.; Fukuhara, S.; Miyagawa, Y.; Kobayashi, K.; Tsujimura, A. Combination therapy with antioxidants improves total motile sperm counts: A Preliminary Study. Reprod. Med. Biol. 2020, 19, 89–94. [CrossRef] [PubMed]

45. Arafa, M.; Agarwal, A.; Mazjoub, A.; Fanner Selvam, M.K.; Baskaran, S.; Henkel, R.; Elbardisi, H. Efficacy of Antioxidant Supplementation on Conventional and Advanced Sperm Function Tests in Patients with Idiopathic Male Infertility. Antioxidants 2020, 9, 219. [CrossRef] [PubMed]

46. Sadaghiani, S.; Fallahi, S.; Heshmati, H.; Teshnizi, S.H.; Chajian, H.A.; Ebrahimi, F.F.A. Effect of antioxidant supplements on sperm parameters in infertile male smokers: A single-blinded clinical trial. AIMS Public Health 2020, 7, 92–99. [CrossRef]

47. Kopets, R.; Kubiida, I.; Chernyavska, I.; Cherepanov, V.; Mazo, R.; Fedeyych, V. Dietary supplementation with a novel l-carnitine multi-micronutrient in idiopathic male subfertility involving oligo-, astheno-, teratozoospermia: A randomized clinical study. Andrology 2020, 8, 1184–1193. [CrossRef]

48. Nazari, L.; Salehpour, S.; Hosseini, S.; Allameh, F.; Jahanmardi, F.; Azizi, E.; Ghodssi-Ghassemabadi, R.; Hashemi, T. Effect of antioxidant supplementation containing L-carnitine on seminal parameters: A prospective interventional study. J.BRA Assist. Reprod. 2021, 25, 76–80. [CrossRef]

49. Gual-Frau, J.; Abad, C.; Amengual, M.J.; Hannauoi, N.; Checa, M.A.; Ribas-Maynou, J.; Lozano, I.; Nikolau, A.; Benet, J.; Garcia-Peiró, A.; et al. Oral antioxidant treatment partly improves integrity of human sperm DNA in infertile grade I varicocele patients. Hum. Fertil. Camb. Engl. 2015, 18, 225–229. [CrossRef]

50. Ma, L.; Sun, Y. Comparison of L-Carnitine vs. Coq10 and Vitamin E for idiopathic male infertility: A randomized controlled trial. Eur. Rev. Med. Pharmacol. Sci. 2022, 26, 4697–4704. [CrossRef]

51. Tharakan, T.; Bettocchi, C.; Carvalho, J.; Corona, G.; Jones, T.H.; Kadioglu, A.; Salamanca, J.I.M.; Serefolgu, E.C.; Verze, P.; Salonia, A.; et al. European Association of Urology Guidelines Panel on Male Sexual and Reproductive Health: A Clinical Consultation Guide on the Indications for Performing Sperm DNA Fragmentation Testing in Men with Infertility and Testicular Sperm Extraction in Nonazoospermic Men. Eur. Urol. Focus 2022, 8, 339–350. [CrossRef]

52. Agarwal, A.; Cannarella, R.; Saleh, R.; Harraz, A.M.; Kandil, H.; Salvio, G.; Boitrelle, F.; Kuroda, S.; Farkouh, A.; Rambhatla, A. Impact of Antioxidant Therapy on Natural Pregnancy Outcomes and Sperm Parameters in Infertile Men: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. World J. Mens Health 2022. [CrossRef] [PubMed]

53. Alahmar, A.T.; Calogero, A.E.; Sengupta, P.; Dutta, S. Coenzyme Q10 Improves Sperm Parameters, Oxidative Stress Markers and Sperm DNA Fragmentation in Infertile Patients with Idiopathic Oligoasthenozoosperma. World J. Mens Health 2021, 39, 346–351. [CrossRef] [PubMed]

54. Arhin, S.K.; Zhao, Y.; Lu, X.; Chetry, M.; Lu, J. Effect of micronutrient supplementation on IVF outcomes: A systematic review of the literature. Reprod. Biomed. Online 2017, 35, 715–722. [CrossRef] [PubMed]
55. Majzoub, A.; Agarwal, A. Antioxidant therapy in idiopathic oligoasthenoteratozoospermia. Indian J. Urol. 1JJU J. Urol. Soc. India 2017, 33, 207–214. [CrossRef]
56. Safarinejad, M.R. The effect of coenzyme Q10 supplementation on partner pregnancy rate in infertile men with idiopathic oligoasthenoteratozoospermia: An open-label prospective study. Int. Urol. Nephrol. 2012, 44, 689–700. [CrossRef]
57. Harumi, T.; Pandi-Perumal, S.R.; Martinez, G.R.; Di Mascio, P.; Hayashi, Y. Chromatographic analyses of biogenic melatonin and its related indolamines. In Melatonin from Molecules to Therapy; Pandi-Perumal, S.R., Cardinali, D.P., Eds.; Nova Science Publishers Inc.: New York, NY, USA, 2007; pp. 3–22.
58. Cardinale, D.P.; Lynch, H.J.; Wurtman, R.J. Binding of melatonin to human and rat plasma proteins. Endocrinology 1972, 91, 1213–1218. [CrossRef] [PubMed]
59. Reiter, R.J. Pineal control of reproduction. Prog. Clin. Biol. Res. 1981, 59, 349–355.
60. Reiter, R.J. Pineal melatonin: Cell biology of its synthesis and of its physiological interactions. Endocr. Rev. 1991, 12, 151–180. [CrossRef]
61. Reiter, R.J. The melatonin rhythm: Both a clock and a calendar. J. Photochem. Photobiol. B 1993, 49, 654–664. [CrossRef]
62. Frungieri, M.B.; Mayerhofer, A.; Zitta, K.; Pignataro, O.P.; Calandra, R.S.; Gonzalez-Calvar, S.I. Direct effect of melatonin on Syrian hamster testes: Melatonin subtype 1a receptors, inhibition of androgen production, and interaction with the local corticotropic-releasing hormone system. Endocrinology 2005, 146, 1541–1552. [CrossRef]
63. Rocha, C.S.; Martins, A.D.; Rato, L.; Silva, B.M.; Oliveira, P.F.; Alves, M.G. Melatonin alters the glycolytic profile of Sertoli cells: Implications for male fertility. J. Cell. Physiol. 2014, 239, 1067–1076. [CrossRef] [PubMed]
64. Yang, W.-C.; Tang, K.-Q.; Fu, C.-Z.; Riaz, H.; Zhang, Q.; Zan, L.-S. Melatonin regulates the development and function of bovine Sertoli cells via its receptors MT1 and MT2. Anim. Reprod. Sci. 2014, 147, 10–16. [CrossRef] [PubMed]
65. Rossi, S.P.; Windschuett, S.; Matzkin, M.E.; Terradas, C.; Ponzio, R.; Puigdomenech, E. Melatonin in testes of fertile men: Evidence for anti-proliferative and anti-oxidant effects on local macrophage and mast cell populations. Andrology 2014, 2, 436–449. [CrossRef]
66. Bellastella, A.; Criscuolo, T.; Manga, A.; Perrone, L.; Sinisi, A.A.; Faggiano, M. Circannual rhythms of plasma luteinizing hormone, follicle-stimulating hormone, testosterone, prolactin and cortisol in prepuberty. Clin. Endocrinol. 1983, 19, 453–459. [CrossRef] [PubMed]
67. Dixit, V.D.; Singh, B.; Singh, P.; Georgie, G.C.; Galhotra, M.M.; Dixit, V.P. Circadian and pulsatile variations in plasma levels of inhibin, FSH, LH and testosterone in adult Murrah buffalo bulls. Theriogenology 1998, 50, 283–292. [CrossRef]
68. Valenti, S.; Thellung, S.; Florio, T.; Giusti, M.; Schettini, G.; Giordano, G. A novel mechanism for the melatonin inhibition of testosterone secretion by rat Leydig cells: Reduction of GnRH-induced increase in cytosolic Ca2+. J. Mol. Endocrinol. 1999, 23, 299–306. [CrossRef]
69. Rossi, S.P.; Matzkin, M.E.; Terradas, C.; Ponzio, R.; Puigdomenech, E.; Levalle, O. New insights into melatonin/CRH signaling in hamster Leydig cells. Gen. Comp. Endocrinol. 2012, 178, 153–163. [CrossRef] [PubMed]
70. Frungieri, M.B.; Calandra, R.S.; Lustig, L.; Meineke, V.; Köhn, F.M.; Mayerhofer, A. Number, distribution pattern, and identification of macrophages in the testes of infertile men. Fertil. Steril. 2002, 78, 296–306. [CrossRef]
71. Pawlak, J.; Singh, J.; Lea, R.W.; Skwarlo-Sonta, K. Effect of melatonin on phagocytic activity and intracellular free calcium concentration in testicular macrophages from normal and streptozotocin-induced diabetic rats. Mol. Cell. Biochem. 2006, 275, 207–213. [CrossRef]
72. Zhang, H.M.; Zhang, Y. Melatonin: A well-documented antioxidant with conditional pro-oxidant actions. J. Pineal Res. 2014, 57, 131–146. [CrossRef]
73. Chen, C.; Ling, M.-Y.; Lin, F.-H.; Xu, L.; Lv, Z.M. Melatonin appears to protect against steroidogenic collapse in both mice fed with high-fat diet and H2O2-treated TM3 cells. Andrologia 2019, 51, e13323. [CrossRef] [PubMed]
74. Deng, S.L.; Wang, Z.P.; Jin, C.; Kang, X.L.; Batool, A.; Zhang, Y.; Liu, Y.X. Melatonin promotes sheep Leydig cell testosterone secretion in a co-culture with Sertoli cells. Theriogenology 2018, 106, 170–177. [CrossRef] [PubMed]
75. Li, C.; Zhou, X. Melatonin and male reproduction. Clin. Chim. Acta Int. J. Clin. Chem. 2015, 446, 175–180. [CrossRef] [PubMed]
76. Atesşahin, A.; Şahna, E.; Türk, G.; Çeribaş, A.O.; Yılmaz, S.; Yüce, A.; Bulmuş, Ö. Chemoprotective effect of melatonin against cisplatin-induced testicular toxicity in rats. J. Pineal Res. 2006, 41, 21–27. [CrossRef]
77. Ilbey, Y.O.; Ozbek, E.; Simsek, A.; Otnuncetemur, A.; Cekmen, M.; Somay, A. Potential chemoprotective effect of melatonin in cyclophosphamide- and cisplatin-induced testicular damage in rats. Fertil. Steril. 2009, 92, 1124–1132. [CrossRef]
78. Ji, Y.L.; Wang, H.; Meng, C.; Zhao, X.F.; Zhang, C.; Zhang, Y.; Zhao, M.; Chen, Y.H.; Meng, X.H.; Xu, D.X. Melatonin alleviates cadmium-induced cellular stress and germ cell apoptosis in testes. J. Pineal Res. 2012, 52, 71–79. [CrossRef]
79. Kanter, M. Protective effects of melatonin on testicular torsion/detorsion-induced ischemia-reperfusion injury in rats. Exp. Mol. Pathol. 2010, 89, 314–320. [CrossRef]
80. Lee, K.M.; Lee, I.C.; Kim, S.H.; Moon, C.; Park, S.H.; Shin, D.H.; Kim, S.H.; Park, S.C.; Kim, H.C.; Kim, J.C. Melatonin attenuates doxorubicin-induced testicular toxicity in rats. Andrologia 2012, 44, 796–803. [CrossRef]
81. Semercioz, A.; Onur, R.; Ogras, S.; Orhan, I. Effects of melatonin on testicular tissue nitric oxide level and antioxidant enzyme activities in experimentally induced left varicocele. Neurol. Endocrinol. Lett. 2003, 24, 86–90.
82. Zhang, K.; Lv, Z.; Jia, X.; Huang, D. Melatonin prevents testicular damage in hyperlipidaemic mice. Andrologia 2012, 44, 230–236. [CrossRef]
83. Heindel, J.J.; Jackson, F.L.; Berkowitz, A.S. Role of the pineal in the alteration of hamster Sertoli cell responsiveness to FSH during testicular regression. *J. Androl.* 1984, 5, 211–215. [CrossRef] [PubMed]

84. Niu, B.; Li, B.; Wu, C.; Wu, J.; Yan, Y.; Shang, R.; Hua, J. Melatonin promotes goat spermatogonia stem cells (SSCs) proliferation by stimulating glial cell line-derived neurotrophic factor (GDNF) production in Sertoli cells. *Oncotarget* 2016, 7, 77532–77542. [CrossRef] [PubMed]

85. Espino, J.; Ortiz, Á.; Bejarano, I.; Lozano, G.M.; Monllor, F.; García, J.F. Melatonin protects human spermatozoa from apoptosis via melatonin receptor- and extracellular signal-regulated kinase-mediated pathways. *Fertil. Steril.* 2011, 95, 7. [CrossRef] [PubMed]

86. Awad, H.; Halawa, F.; Mostafa, T.; Atta, H. Melatonin hormone profile in infertile males. *Int. J. Androl.* 2006, 29, 409–413. [CrossRef]

87. Kratz, E.M.; Piwowar, A.; Zeman, M.; Stebelová, K.; Thalhammer, T. Decreased melatonin levels and increased levels of advanced oxidation protein products in the seminal plasma are related to male infertility. *Reprod. Fertil. Dev.* 2016, 28, 507–515. [CrossRef] [PubMed]

88. Lu, X.L.; Liu, J.J.; Li, J.T.; Yang, Q.A.; Zhang, J.M. Melatonin therapy adds extra benefit to varicecelectomy in terms of sperm parameters, hormonal profile and total antioxidant capacity: A placebo-controlled, double-blind trial. *Andrologia* 2018, 50, e13033. [CrossRef] [PubMed]

89. Smits, R.M.; Mackenzie-Proctor, R.; Yazdani, A.; Stankiewicz, M.T.; Jordan, V.; Showell, G.M. Antioxidants for male subfertility. *Cochrane Database Syst. Rev.* 2019, 3, CD007411. [CrossRef]

90. Mannucci, A.; Argento, F.R.; Fini, E.; Coccia, M.E.; Taddei, N.; Becatti, M.; Fiorillo, C. The Impact of Oxidative Stress in Male Infertility. *Front. Mol. Biosci.* 2021, 8, 799294. [CrossRef]

91. Hussain, B. The antioxidant paradox. *Lancet* 2000, 355, 1179–1180. [CrossRef]

92. Symeonidis, E.N.; Evgeni, E.; Palapelas, V.; Koumasi, D.; Pyrgidis, N.; Sokolakis, I.; Hatziychristodoulou, G.; Tsampali, C.; Mykonias, I.; Zachariou, A.; et al. Redox Balance in Male Infertility: Excellence through Moderation-“Μέτρων ἀρχήν”. *Antioxidants* 2021, 10, 1534. [CrossRef] [PubMed]

93. Aitken, R.J.; Drevet, J.R.; Moazamian, A.; Gharagozloo, P. Male Infertility and Oxidative Stress: A Focus on the Underlying Mechanisms. *Antioxidants* 2022, 11, 306. [CrossRef] [PubMed]

94. Green, B.N.; Johnson, C.D.; Adams, A. Writing narrative literature reviews for peer-reviewed journals: Secrets of the trade. *J. Chiropr. Med.* 2006, 5, 101–117. [CrossRef]