Anatomical, Chemical and Mechanical Characteristics of Beech Wood Degraded by Two Pleurotus Species

Anatomical, kemijska i mehanička svojstva bukova drva degradiranoga gljivama roda *Pleurotus*

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ABSTRACT • The aim of this study was to determine the destructive capabilities of the two white rot fungi *Pleurotus cornucopiae* (Pc) and *P. eryngii* (Pe) compared with the standard fungus *Trametes versicolor* (Tv) on beech wood samples after 60 days of incubation. Understanding of the white rot decay is important as it is necessary for the development of effective solutions for wood protection. Measurements of mass loss, chemical, mechanical properties and light microscopical investigations were conducted prior to and after incubation. Mass loss of samples was found to be 9-22 % depending on fungi species. Impact bending strength is not as sensitive as presumed in classical literature. Light microscopy analysis revealed that decay patterns were similar for both fungi. Wood cell wall thinning, fungal colonization hyphae were also the same for both fungi. Results indicated considerable wood attack by both *Pleurotus* species, Pc being more destructive than Pe.

Keywords: *Pleurotus cornucopiae; Pleurotus eryngii; Trametes versicolor; wood decay; chemical analysis; light microscopy*

SAŽETAK • Cilj rada bio je utvrditi učinak dviju gljiva bijele truleži – *Pleurotus cornucopiae* (Pc) i *P. eryngii* (Pe) – na uzorcima bukova drva nakon 60 dana inkubacije u usporedbi s učinkom standardne gljive *Trametes versicolor* (Tv). Razumijevanje degradacije drva zbog bijele truleži iznimno je važno za razvoj učinkovitih rješenja zaštite. U pokusu su prije i nakon inkubacije drva provedena mjerenja mjerjenja gubitaka mase, kemijskih i mehaničkih svojstava te je obavljeno ispitivanje svjetlosnim mikroskopom. Utvrđeno je da je gubitak mase uzorka iznosio 9 – 22 %, ovisno o vrsti gljive. Čvrstoća drva na savijanje nije toliko osjetljiva na utjecaj gljiva kao što se navodi u klasičnoj literaturi. Analiza slika dobivenih svjetlosnim mikroskopom pokazala je da su procesi propadanja drva pri zaživi objima gljivama slični. Stanjivanje stijenki drvnih stanica i hije kolonizacije obiju gljiva također su bile jednake. Rezultati su pokazali znatnu degradaciju drva napadnutoga gljivama roda *Pleurotus*, s tim da je gljiva Pc destruktivnija od gljive Pe.

Ključne riječi: *Pleurotus cornucopiae; Pleurotus eryngii; Trametes versicolor; propadanje drva; kemijska analiza; svjetlosna mikroskopija*
1 INTRODUCTION

Wood is one of the most important building materials. It has been used for various applications such as construction, furniture, poles, and sports equipment. However, non-durable and susceptible wood species are prone to fungal degradation. Degradation develops if the moisture content of wood exceeds certain limit, which is associated to fiber saturation point (Schmidt 2006). Wood-decaying fungi play a prominent number of ecological roles in forest ecosystems that affect the health, diversity, productivity, and development of their biotic communities such as mycorrhizal associations with vascular plants, pathogens of commercial tree species, decomposers of coarse organic material, and food resources for wildlife (Marcot, 2017).

There are various classifications of wood-degrading fungi, and the most important is based on the color of degraded wood; white-, brown-, and soft-rot, blue-stain and sap-stain fungi (Zink and Feng, 1989; Schmidt, 2006). The white-rot fungi predominantly associated with hardwood wood species, where two degradation patterns are described, namely, selective and non-selective white rot as described by Eriksson et al. (1990). The selective fungi degrade and consume predominately hemicellulose and lignin, while the non-selective white rot fungi, beside hemicellulose and lignin, degrades cellulose as well (Eriksson et al., 1990; Zabel and Morrell, 1992; Eaton and Hale, 1993; Schwarze et al. 2004; Schmidt, 2006; Bari et al., 2019).

In the nature, different fungal species colonize a variety of substrates. Some fungi are more specialized than the others (Bari et al., 2019). In this regards, Pleurotus species are reported as one of the most important and robust white rot fungi. For example, in the Northern forests of Iran, a colony of them can be found on beech, hornbeam, oak, and aspen wood, clearly proving their flexibility (Ershad, 2009).

Most of forests in Iran are located in the northern parts, bordering the coastal plain at the Caspian Sea and on the northern slopes of the Alborz Mountain range. These forests cover an area 850 km long and vary in width from 20 to 70 km. The forests of this region are known as Hyrcanian forests. These Hyrcanian forests comprise a little more than 1.9 million ha of almost 100 % hardwoods, mainly beech (Fagus orientalis) and hornbeam (Carpinus betulus) (Kiaei and Samariha, 2001). Pleurotus genus is one of the most important basidiomycetes from commercial perspective, due to their gastronomic, nutritional and medicinal properties. Another factor that contributes to their commercial importance is the fact that they can be easily cultivated on a wide range of substrates, from straw to wood (Solár et al., 2007; Aghajani et al., 2018; Humar, 2013). Preferential degradation of wheat straw lignin was studied by Martinez et al. (1994), who concluded that Pleurotus eryngii and P. ostreatus are the most promising fungi. They reported that P. eryngii was the most successful organism examined, exhibiting nearly 50 % reduction of Klason lignin during a solid-state fermentation (SSF) experiment.

Beech wood is an important wood species, but unfortunately it is very susceptible to fungal degradation; hence it was used in the respective study. The objectives of the present study were to screen the capabilities and decay patterns of the two Pleurotus species, P. cornucopiae and P. eryngii, by applying them on beech wood samples and determine the biological, chemical, and mechanical properties of decayed wood as well as compare their degradation capacities to standard white-rot fungus Tremetes versicolor. These data are important because of constructional and biotechnological reasons. Degraded wood can be used in various fermentation processes from biogas to bioethanol production (Taherzadeh et al., 2008).

2 MATERIALS AND METHODS

2.1 Fungi

2.1.1  Gljive

Fruiting bodies of Pleurotus cornucopiae (Paullet) Rolland (Pc) and Pleurotus eryngii (De Quél. (Pe) were collected from living beech trees (Fagus orientalis Lipsky.) at Hezarjarib forests, (located in Neka, Iran) during the spring 2017. Macro- and microscopic identification was carried out in accordance with the keys of Eriksson and Ryvarden (1975), Gilbertson and Ryvarden (1986), Ryvarden (1991), Ryvarden and Gilbertson (1993).

2.2 Wood samples

2.2.1  Uzorci drva

Wood blocks were obtained from (Fagus orientalis) trees at breast height and air-dried to reach 23±2 % moisture content. Specimens of (5×2.5×1.5) cm³ according to the EN113 standard (1997) were used for the determination of mass loss (ML), and (6×5×0.6) cm³ according to ASTM-D256-04 standard (ASTM 2004) for testing impact bending strength. The specimens used to evaluate impact bending strength were cut in cross section. Ten replicate specimens were prepared from different disks for each test. They were kept in a conditioning chamber (25 °C, and 40±3 % RH) for 4 weeks before testing.

2.3 Mass loss after biological test

2.3.1  Gubitak mase nakon biološkog testa

In order to evaluate the degradation capabilities of the Pleurotus species, beech wood samples were oven dried at 103±3 °C for 24 h and weighed prior to fungal exposure. Wood blocks were sterilized at 121°C for 20 min and exposed to fungi according to EN113. Fungi were incubated for 60 days at 22±2 °C and relative humidity of 65±5 %. Ten replicates were used for each treatment. After exposure, surface mycelium was scraped off and wood samples were weighed before drying at 103 °C for 24 h to determine the final moisture content (MC). After drying, the mass loss (ML) was obtained (Eq. 1 and 2).

\[
MC(\%) = \frac{M_w - M_d}{M_d} \times 100 \tag{1}
\]

\[
50 \%
\]
M ( ML/%) = \frac{M_i - M_a}{M_i} \times 100 \quad (2)

Where MC is moisture content (%), ML is mass loss (%), \(M_i\) dry mass before decay (g), \(M_w\) wet mass after decay (g), and \(M_a\) dry mass after decay (g).

2.4 Chemical analyses

Changes in the chemical constituents of the wood cell walls of sound wood controls and samples, following exposure to fungi, were evaluated according to TAPPI standards test methods. The Klason lignin was determined according to T-222 om-98 of TAPPI standard. Oven-dried, extractive-free sawdust (1g) was mixed with 15 ml of 72 % sulfuric acid for 2 h at room temperature. The mixture was diluted with 560 ml of distilled water, heated for 4 h, and the insoluble materials were filtered off. The residue was washed and dried at 103 °C. The lignin content was calculated using Eq. (3):

\[ KL(\%) = \frac{S_i - KL}{S_i} \times 100 \quad (3) \]

Where \(S_i\) is the dried weight of sawdust and \(KL\) is the dried weight of extracted Klason lignin.

Cellulose content was determined in accordance with T-17 wd-70 of TAPP; 2 g of sawdust (free from extractives) were mixed with 96 % EtOH (100 ml) and 65 % nitric acid (50 ml). The mixture was heated under reflux for 1 h, cooled and filtered. The residue was washed with distilled water and dried at 103 °C. Cellulose content was then calculated by Eq. (4):

\[ Cellulose(\%) = \frac{S_i - EC}{S_i} \times 100 \quad (4) \]

Where \(S_i\) is the dried weight of sawdust and \(EC\) is the dried weight of extracted cellulose.

2.5 Impact bending strength

Impact bending strength was performed according to ASTM-D256-04 and calculated using Equation 3. Before the Impact bending strength test, all samples were conditioned in a standard climate at 20 °C and 65 % relative humidity until constant mass was achieved.

\[ I = \frac{F_{max}}{A} \quad (5) \]

Where \(I\) is resistance to impact (J/m²), \(F_{max}\) is force (J) and \(A\) is cross section area (m²).

2.6 Light microscopy

In order to monitor wood degradation, a GSL-1 sliding microtome (WSL, Switzerland) was used to cut thin wood sections (10–15 μm) of the blocks (20×10×8, mm³). The sections were stained with safranin (0.5 % aqueous), Astra Blue (0.3 % aqueous) solution and mixed in a 1:1 ratio, washed in distilled water for 1–3 min and dehydrated by an alcohol series. After rinsing in xylol for 1–2 min, sections were mounted in Moutan glaze (Kimianovin, Tehran, Iran) on microscope slides. To avoid buckling of the sample, a 50 g weight was placed on the cover glass edges while the slide was drying at 60 °C for 12 h. Dried sections were examined and photographed with an Olympus E−210 microscope and with an Olympus E−450 camera.

2.7 Statistical analyses

Comparison between mass loss and changes in chemical components of the wood was carried out using a Student t-test for each exposure period (95 % level of confidence). Two-way ANOVA was conducted to examine the effect of decay condition on mass and chemical losses. All statistical analyses were performed using the SPSS software program, version 23.

3 RESULTS AND DISCUSSION

3.1 Mass loss

Wood density is one of the first and elementary information. The average dry density of beech wood was 0.63 g cm⁻³. The ring width was between 2.80 and 3.40 mm. Mass loss (ML) of beech wood samples exposed to the fungi after incubation is shown in Figure 1. Average mass losses were 17.40 %, 8.70 %, and 21.76 % after 60 days incubation for P. cornucopiae (Pe), P. eryngii (Pe), and T. versicolor (Tv), respectively. The results indicated that Pe were more effective than Pc. However, Tv caused most ML. The minimum ML of 20 % by Tv is necessary for beech wood after 16 weeks (112 days) of incubation in accordance with EN-113 (1997). On the other hand, the average ML was 20 % and 40 %, respectively, in size of 30×10×5 mm after 12 weeks of exposure (Bravery, 1978). Bari et al. (2019) showed that Pleurotus ostreatus and Tv produced the same ML in beech wood after 120 days of incubation.

3.2 Moisture content after decay

The moisture content (MC) of wood blocks after fungal incubation is shown in Figure 2. Generally, the MC was 77.09 %, 65.80 %, and 108.51 % after 60 days of incubation for Pe, Pc, and Tv, respectively. Since fungi need moisture for their enzymes to cleave the cell wall components (Baldrian, 2008), the water is necessary for their function. According to Figure 2, the results demonstrated that the MC of the decayed wood blocks increased with the mass losses caused by both decay fungi. The increase of the mass loss could increase the moisture content in wood blocks. Similar works (Bami and Mohheby, 2011) showed that white-rot fungi caused higher water content in decayed wood samples.

3.3 Cell wall components analysis

Average lignin and cellulose contents of sound and decayed beech wood samples, after 60 days of degradation by fungi, is shown in Figure 3. The graph indicates that the three white-rot fungi severely degraded cellulose and lignin. With regard to lignin, average degradation by fungi was 16.73 %, 16.63 %, and 13.67 % after 60 days of incubation for Pe, Pc, and Tv, re-
respectively, while for the sound wood it was 23.63 %. According to Koshijima and Watanabe (2003) and Schmidt (2006), white-rot fungi are the most efficient lignin degraders in nature and they play a key role in carbon recycling on Earth. They break-down the lignin units by secretion of different enzymes to reach the necessary carbon. Average degradation of cellulose by fungi was 32.77 %, 34.53 %, and 28.64 % for Pe, Pc and Tv, respectively. Cellulose is the main carbon source for fungi, especially basidiomycetes (Schmidt, 2006). White rot fungi are divided in selective and simultaneous white-rot species (e.g. Eriksson et al. 1990; Schmidt, 2006). Figure 3 shows that the three fungi caused simultaneous white-rot in beech wood samples. Karim et al. (2016) showed that *Pleurotus ostreatus* decomposed beech and oak wood samples in natural and controlled conditions also follow a similar lignin degradation pattern. However, indications of selective digestion were also found in some wood cells. However, several researchers (e.g. Martinez et al. 2001, 2005) reported that many *Pleurotus* species caused the selective rot pattern. Cellulose, lignin content of decayed wood samples in the present study, is much lower than reported by Chen et al. (2017) and Hosseiniihashemi et al. (2017). Olfat (2014) indicated that the mass loss of beech wood was 47.5 % after 16 weeks and 13.2 % after 10 weeks. Mass loss values of this study are comparable with the values given by Witomski et al. (2012).

### 3.4 Monitoring beech wood degraded by white-rot fungus

Wood density is one of the first and basic information. The average density of beech wood is 0.63 g cm⁻³. Growth rings are distinct because of the unusually light color of latewood. Two forms of degradation for white rot were described in this study. It is known that *P. eryngii* (Pe) and *P. cornucopiae* (Pc) caused selective lignin degradation. In the selective delignification type, lignin is degraded earlier than cellulose or hemicellulose in the process of decay. During the initial stages of decay, the cellulose is left unchanged during delignification. In some cases, hyphae in the cell lumen grow, so that lignin is separated from the adjacent cell wall (Anagnost, 1998; Schwarze, 2007). In

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**Figure 1** Average percent mass loss of beech wood samples decayed by white rot fungi after 60 days of incubation

**Slika 1.** Prosječni postotni gubitak mase uzoraka bukovine zbog djelovanja gljiva bijele truleži nakon 60 dana inkubacije

**Figure 2** Moisture content of beech wood samples due to fungal metabolism after 60 days of incubation

**Slika 2.** Sadržaj vode u uzorcima bukovine zbog metaboližma gljiva nakon 60 dana inkubacije

**Figure 3** Average percentage of chemical composition in decayed beech wood samples after 60 days of incubation

**Slika 3.** Prosječni postotak promjene kemijskog sastava u uzorcima bukovine djelovanjem gljiva nakon 60 dana inkubacije
the tangent section, the beech wood incubated with *P. eryngii*, as seen in Figure 4a. The hyphae, growing in the cell lumen of the fiber-tracheids, are seen in the early stages of delignification in the secondary walls. As seen in the radial section, the beech wood incubated with *P. cornucopiae*, the hyphae penetrate into the cell walls, and then first separate the middle lamella, so that the cells tend to separate from each other (Tuar et al. 1995; Seshikala and Charya1, 2012). Cellulose is relatively unchanged during selective delignation, at least in the early stages of decay (Figure 4b). *T. versicolor* cause simultaneous white rot in angiosperms, but only rarely in gymnospermous wood. In many studies, it was reported that this type of white decay degrades the adjacent cell wall for hyphae growing in contact with the lumen surface (Anagnost 1998; Schwarze 2007; Karim et al. 2017; Silva-Castro et al. 2018). The enzymes of *T. versicolor* cause the degradation in all the components of the lignified cell wall. The decomposition of cellulose, hemicellulose and lignin occurs at almost the same rate. As erosion proceeds on the lumen surface, the cell wall becomes thin evenly, as opposed to forming channels (Anagnost, 1998; Schwarze, 2007). This degradation form of *T. versicolor* in beech wood is characterized in Figure 4c. In the transverse sections, advanced thinning resulted in the localized removal of the cell wall and middle lamella.

### 3.5 Mechanical evaluation

**Ocjena mehaničkih svojstava**

Figure 5 shows the effects of the cell wall degradation on the impact bending strength after exposure to the white-rot fungi. The average decrease of impact bending strength by the fungi was 3.32 %, 3.68 %, and 3.17 for *Pe*, *Pc*, and *Tv*, respectively, while it was 4.59 % for the control sample. Overall, both fungi showed a similar effect on the reduction of impact strength. Toughness or impact strength is the ability of wood to absorb the force of impact bending and characterizes the ability of material to withstand impact loads. Impact strength is expressed as the energy consumed while breaking wood with defined dimensions. This mechanical property is most sensitive to decay and, unlike other strength properties that decrease gradually as decay progresses, impact strength declines rapidly during incipient wood decay (Rowell, 2005).

![Figure 4](image_url)

**Figure 4** Light micrographs of beech wood degradation after 60 days of exposure to white-rot fungi; (a) Transverse section of beech wood incubated with *P. eryngii* (*Pe*). (b) Radial longitudinal section of beech wood incubated with *P. cornucopiae* (*Pc*). (c) Transverse section of beech wood incubated with *T. versicolor* (*Tv*)

![Figure 5](image_url)

**Figure 5** Average percentage of impact bending strength in decayed beech wood samples after 60 days of incubation

**Slika 4.** Svjetlosne mikrografije degradacije uzoraka bukovine nakon 60 dana izlaganja gljivama bijele truleži: a) poprečni presjek uzorka bukovine inkubiranoga gljivom *P. eryngii* (*Pe*), b) radijalni uzdužni presjek uzorka bukovine inkubiranoga gljivom *P. cornucopiae* (*Pc*), c) poprečni presjek uzorka bukovine inkubiranoga gljivom *T. versicolor* (*Tv*)

**Slika 5.** Prosječni postotak smanjenja savojne žilavosti degradiranih uzoraka bukovine nakon 60 dana inkubacije
Figures 6 shows the correlation between mass loss and impact bending strength data. As can be seen in these figures, the correlation is not very tight.

4 CONCLUSION

Anatomical, chemical and mechanical properties were investigated of beech wood exposed to the two white-rot fungi for 60 days of incubation. The fungus clearly caused simultaneous decay pattern of cell wall polymers in the wood. Results indicated that both Pleurotus species created a considerable mass loss, which was accompanied by losses in chemical and mechanical properties. Altogether, under the conditions of the present research, it was concluded that the decay capacity of \textit{P. eryngii} was more aggressive than that of \textit{P. cornucopiae} in some test cases. According to the obtained results of the present study, the capability of wood rotting fungi for biotechnological applications such as biopulping, bioremediation, biochelation and recycling of treated wood is indisputable. However, their advantages and disadvantages should be considered before attempting industrial-scale operations.

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