Mast cells in vulnerable atherosclerotic plaques - a view to a kill

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Abstract

The aim of the present review is to discuss the participation of mast cells in the pathogenesis of erosion and rupture of atherosclerotic plaques, the major causes behind acute coronary syndromes and myocardial infarction. We present ex vivo observations describing mast cells and their activation in human atherosclerotic plaques and discuss in vitro and in vivo data showing that mast cells are potential regulators of inflammation, immunity and adverse remodeling, including matrix remodeling and cell death. Furthermore, we focus on studies that have been performed with human tissues and human mast cells, but when appropriate, we also discuss observations made in animal models. Finally, we present potential pharmacological means to modulate mast cell responses in the arterial vessel walls.

Keywords: apoptosis • atherosclerosis • endothelium • erosion • mast cell • pericellular matrix • plaque rupture • proteases • smooth muscle cell

Mast cells - an introduction

The origin of mast cells

Mature mast cells are progeny of multipotent haematopoietic stem cells that commit to the mast cell lineage already in the bone marrow, being positive for CD34 and c-kit, but negative for FcεRI[1–3]. The mast cell-committed progenitors leave the bone marrow, circulate in the blood as CD34+ and c-kit+ precursor cells [4], and adhere to activated endothelial cells via α4β1 integrins, vascular cell adhesion molecule-1
In the presence of specific chemotactic signals, such as stem cell factor (SCF) and eotaxin [6], the mast cell precursors migrate into tissues using as sensors of chemotaxis different chemokine receptors, such as CXCR2, CCR3, CXCR4 and CCR5 [7]. Recently, the expression of transcription factor T-bet by dendritic cells has been shown to regulate the homing of mast cell precursors [8]. In the tissues, the proliferation and differentiation of mast cell precursors depend on the presence of local growth factors and cytokines, notably SCF, IL-3, IL-4, IL-6, IL-9 and NGF [9–12], which are secreted by various tissue cells. The crucial requirement of a functional SCF and c-kit signaling system for mast cell growth and development is underscored by the fact that lack of c-kit signaling in mice results in mast cell deficiency [13–15], whereas an elevated expression of c-kit in patients induces mastocytosis [16, 17]. Similarly, injection of SCF into the skin of human beings results in local accumulation of mast cells [18]. Depending on the site of tissue infiltration, the precursor cells proliferate and differentiate either into connective tissue-type or mucosal-type mast cells, two well-described subsets of mast cells that differ in the quality and quantity of their stored mediators and also in their physiological functions [19].

**Mast cell subtypes**

The connective tissue-type and mucosal mast cells can be morphologically distinguished using histochemical, electron microscopic, biochemical or immunological criteria that determine their tissue distribution, specific structural features and mediator content [19]. Using a more simplistic approach, human mast cells have been divided into subtypes depending on their variable content of two neutral serine proteases, tryptase and chymase. Thus, mast cells containing only tryptase (MC\textsubscript{T}) represent the mucosal mast cells and are typically present in the lungs and in the intestinal mucosa. Mast cells containing both tryptase and chymase (MC\textsubscript{TC}) represent the connective tissue-type mast cells and are typically found in the skin, synovium and perivascular tissue. However, the two tissue subtypes can inter-change, their ultimate phenotype (MC\textsubscript{T} or MC\textsubscript{TC}) being determined by their microenvironment. Indeed, by varying the culture conditions in vitro human mast cells of different phenotypes may be obtained [20]. The presence of IL-4 [21] or IL-6 [10] has been shown to induce the differentiation of CD34\textsuperscript{+} precursor cells into chymase-containing human MC\textsubscript{TC}.

**Mast cell localization and physiological function**

Mast cells are generally found at the boundaries between the outside world and the internal milieu of the body, such as in the skin and in the mucosa of the pulmonary, gastrointestinal and genitourinary system. Moreover, they are prevalent in the conjunctivae and the mucosa of the nose. At these sites, they act as surveillance antennae of the local microenvironment and direct immune responses by regulating innate and adaptive immune mechanisms [22, 23]. In addition, mast cells are present in most vascularized tissues, where they reside in the vicinity of blood vessels and lymphatic vessels [24]. Although mast cells are best known for their ability to release histamine and to induce IgE-mediated type I hypersensitivity reactions [25, 26], they also initiate and regulate inflammatory responses, defend the host against bacterial and parasitic pathogens, regulate vascular functions, participate in wound healing and neovascularization and recruit and activate other types of inflammatory cells, and stromal cells, as well [27–30].

Based on the above findings, the mast cells may be considered ‘sentinels’ or ‘friends’ that play an important role in the normal homeostasis of the body. Indeed, several studies in mice have indicated that mast cell deficiency may be harmful and even lethal if the mast cell-deficient host is pre-disposed to exogenous insults, such as bacterial infections, acute septic peritonitis, ET-1 or snake and honeybee venom [31–34]. However, chronic local activation of mast cells in diseases, such as atherosclerosis [35], rheumatoid arthritis [36] and congestive heart failure [37], may result in a ‘foe’ response that is unregulated, and if not properly terminated, may turn out to be harmful and even lethal to the host.

**Mast cell – a potent effector cell**

Mast cells contain a wide variety of preformed mediators that are secreted acutely upon mast cell activation...
with ensuing degranulation and that participate in the
mast cell-mediated ‘friend or foe’ responses. The pre-
formed mediators can roughly be divided into five
classes of effector molecules, notably histamine, pro-
teoglycans, proteases, growth factors and cytokines,
all of which may have an impact on the vulnerability
of an atherosclerotic plaque. In addition to the preformed
mediators, the activated mast cells also produce newly
formed lipid mediators of which prostaglandins and
leukotrienes are the major ones, as well as cytokines
and chemokines. Below, we shortly discuss the mast
cell-derived mediators and their possible relation to
the pathogenesis of plaque erosion and rupture.

Histamine

Histamine is a biogenic amine that exerts its functions
via four different histamine receptors (H1, H2, H3 and
H4 receptors) differently expressed in various cells
and tissues. The physiological and pathophysiological
effects of histamine include blood vessel dilatation,
increased vasopermeability and translocation of P-
selectin to the endothelial cell surface with subse-
quent induction of leukocyte rolling in vivo [38]. Thus,
the histamine-mediated effects lead to local edema
(swelling) and attraction of inflammatory cells, which
may then increase the susceptibility of plaque erosion
and rupture. Furthermore, histamine has been shown
to induce the expression of Toll-like receptors 2 and 4
in endothelial cells, which may be of relevance in local
innate immune responses in the vulnerable plaque
[39]. In addition, histamine induces the expression of
tissue factor by smooth muscle and endothelial cells
[40], which may promote thrombosis in atherosclerotic
arteries. Since atherosclerotic coronary arteries con-
tain more histamine than healthy coronary arter-
ies and also are hyperreactive to it, histamine has a
potential of inducing vasospasm in human athero-
sclerotic coronaries [41].

Mast cell-derived proteoglycans

Human mast cell granules contain heparin and chon-
droitin sulphate proteoglycans that form the macro-
complexes in which proteases and growth factors are
embedded. Upon degranulation, the exocytosed
mast cell proteoglycans exert antithrombotic effects
[42] and a variety of functions on neighboring cells, in
particular the smooth muscle cells [43–45]. Heparin
may also bind lipoproteins, and thus play a role in
foam cell formation, i.e. in the accumulation of low-
density lipoprotein (LDL) cholesterol in macrophages
typically seen in the early stages of atherogenesis
[46]. Heparin is also important for the activity of mast
cell serine proteases, as it stabilizes the active
tetrameric form of tryptase [47] and protects chy-
mase and cathepsin G from their natural inhibitors
present in the interstitial fluid [48–50]. Thus, in contrast
to many proteases released by other inflammatory
cells, the mast cell-derived proteolytic enzymes are
secreted bound to proteoglycans that partially protect
them from inactivation in the presence of natural pro-
tease inhibitors. Mast cell heparin also binds several
growth factors, attenuates their inactivation and allows
their prolonged presence in the extracellular space.

Mast cell-derived proteases

Tryptase is a mast cell-specific neutral serine
protease, which is found in two isoforms, α- and
β-tryptase, the former being released from mast cells
constitutively and the latter upon degranulation [51].
The tetrameric form of β-tryptase is proteolytically
active when bound to heparin, but an active
monomeric form has also been described [52]. When
released by the mast cells, tryptase is capable of
activating neighboring cells by cleaving and activat-
ing protease-activated receptor (PAR)-2 [53], and
thrombin receptors [54]. In addition, tryptase has
been shown to degrade neuropeptides, such as
vasoactive intestinal peptide (VIP) and calcitonin
gene related peptide (CGRP) [55], and the pericellu-
lar matrix components fibronectin and vitronectin [56,
57], and to activate pro-matrix metalloproteinases
(MMP)-1, -2 and -3 [58, 59]. Interestingly, there are
no known natural inhibitors of the tetrameric form of
β-tryptase [60]; however, the disruption of
tryptase–proteoglycan interactions by lactoferrin can
inactivate tryptase by inducing its monomerization
[61]. Also, the reassembly and activity of monomers
can be inhibited by a variety of endogenous inhibitors,
such as antithrombin III and α-2-macroglobulin [52].
Furthermore, tryptases isolated from different tissues
are very heterogeneous and show variation in size,
charge and proteolytic activity [62, 63].

Chymase is a neutral serine protease secreted
exclusively by MC\textsubscript{TC} and it appears that chymase
plays a major role in the proteolytic activation of MMP-1, -2 and -9 [59, 64, 65] as well as in the physiologic degradation of fibronectin and thrombin in tissues [66]. Chymase may also act as an angiotensin-converting enzyme [67–69], release latent TGF-ß1 from the extracellular matrix [70], inhibit smooth muscle growth [71] and induce apoptosis of arterial smooth muscle cells [72] and endothelial cells [73]. In addition, mast cell proteases and endotoxin synergistically activate human coronary artery endothelial cells to generate interleukin-6 and interleukin-8, which may enhance the inflammatory response in atherosclerotic lesions [74].

Cathepsin G is an elastolytic, angiotensin II-forming serine protease that is present in the granules of human MCTC [75]. Our recent observations show that mast cells are a major local source of cathepsin G in the coronary arteries and that cathepsin G can degrade both fibronectin and VE-cadherin [76], which are necessary for adhesion of endothelial cells to their basement membrane and to each other.

Renin, an angiotensin I-forming enzyme has recently been shown to be present in cardiac mast cells of rats and guinea pigs, as well as in the human mastocytoma cell line, HMC-1 [77, 78]. If present also in mast cells of the arterial wall, renin may participate in the local production of angiotensin II, and participate in the adverse vascular effects mediated by the renin-angiotensin system (RAS). Angiotensin II has been shown to induce atherosclerosis through various processes, such as by inducing endothelial dysfunction, cellular proliferation and inflammation [79]. Mast cells are also known to produce MMPs, notably MMP-1 [80] and MMP-9 [81, 82], that may actively participate in plaque remodeling and increase the susceptibility to plaque erosion and rupture. Although mast cells have been shown to contain tissue inhibitor metalloproteinase 4 (TIMP-4) [83], the mast cell-derived proteases may promote the activity of MMPs by degrading the TIMPs [84]. The important role of various MMPs in plaque destabilization has been demonstrated in mouse models of atherosclerosis [85].

Growth factors and preformed cytokines

Human mast cells have also been shown to contain several different growth factors, such as vascular endothelial growth factor (VEGF), basic fibroblast growth factor (bFGF) and transforming growth factor (TGF)-ß, that upon secretion may recruit effector cells and enhance angiogenesis [86], and thus, participate in the pathogenesis of plaque erosion and rupture. Tumour necrosis factor (TNF)-α is a preformed cytokine that is stored in the granule compartment and released with the granules during mast cell activation and degranulation, and also secreted after mast cell activation as a newly formed cytokine [87]. TNF-α is known to induce either directly or indirectly endothelial dysfunction and expression of the endothelial cell adhesion molecules P-selectin, E-selectin and VCAM [88], which are responsible for leukocyte infiltration. TNF-α is also known to increase endothelial permeability at very low concentrations [89, 90], an effect that is mediated via TNF receptor 1 [91]. Interestingly, the TNF-α-mediated endothelial effects may be potentiated by interferon (IFN)-γ [92], a cytokine also found in atherosclerotic lesions.

Newly formed cytokines and chemokines

Upon activation, mast cells produce a plethora of different cytokines and chemokines that may actively participate in the pathogenesis of the atherosclerotic plaques. These include at least the following: TNF-α, IFN-α, IFN-β, IL-1α IL-1β, IL-6, IL-18, granulocyte macrophage-colony stimulating factor (GM-CSF), and leukaemia inhibitory factor (LIF), all of which are involved in the induction of inflammation [86]. Furthermore, mast cells also secrete cytokines that are capable of inducing either T helper 1-type (Th1) polarization (IL-12 and IFN-γ) or T helper 2-type (Th2) polarization (IL-3, IL-4, IL-5, IL-9, IL-13, IL-15 and IL-16) [93]. The activated mast cells also secrete IL-10 and TGF-β, which are known to attenuate the inflammatory response in the atherosclerotic lesions [94]. In addition to releasing cytokines, the activated mast cells are able to secrete both α-chemokines (CXCL1, CXCL2, CXCL8, CXCL9, CXCL10, CXCL11) and β-chemokines (CCL2, CCL3, CCL4, CCL5, CCL11, CCL20) that recruit additional effector cells and regulate immune responses [29, 86, 95].

Newly formed lipid mediators

Activated mast cells also mobilize arachidonic acid through the activation of cytosolic phospholipase A2,
with ensuing rapid generation and secretion of both prostaglandin D2 and leukotriene C4 [96]. The mast cell-derived eicosanoids interact with their respective cellular receptors and are known to serve diverse functions in vasoconstriction, cell trafficking, antigen presentation, immune cell activation, matrix deposition and fibrosis [96]. Interestingly, atherosclerotic arteries are hyperresponsive to the constricting effects of leukotrienes [97], whereas prostanoids, by being important mediators of inflammation and endothelial dysfunction are also likely to contribute to the hyperresponsiveness of the endothelium in atherosclerosis lesions [98].

Mast cells in the vessel wall

In the arterial vessel wall, mast cells localize preferentially to the subendothelial space of the intimal layer, i.e. to the close vicinity of the endothelial cells (ECs), which likely reflects the fact that ECs are a major source of SCF [99, 100], a vital growth factor for mast cells in the vessel wall. In the normal human aortic intima, approximately 60% of all mast cells (15 mast cells/mm²) are of the MC₇ subtype, and approximately 40% are of the MC₇C subtype [101]. It is evident that the microenvironment in the arterial wall is effectively influencing the mast cell phenotype, since both subtypes can be found in the same atherosclerotic plaques [101]. In healthy human arteries, there are no mast cells present in the medial layer, i.e. the layer that constitutes of concentric layers of contractile smooth muscle cells, and is responsible for the regulation of the arterial tone. However, the outer adventitial layer of a normal coronary artery contains high amounts of mast cells (19 mast cells/mm²). Even larger numbers of mast cells were found to be present in the adventitia of ruptured plaques (98 mast cells/mm²) than in the adventitia of non-ruptured plaques (41 mast cells/mm²) [102]. The majority of the adventitial mast cells contained both tryptase and chymase, and were the only cells in the coronary adventitia that contained histamine. Interestingly, the amounts of adventitial macrophages and CD4⁺ T-lymphocytes were also found to be increased in the segments with plaque rupture, indicating that the inflammatory process is not only restricted to the intimal layer of the lesion. Long-term cocaine abusers have increased level of atherosclerosis and increased numbers of adventitial mast cells in their coronary arteries, suggesting that adventitial mast cells may potentiate atherosclerosis, vasospasm, thrombosis and even premature sudden death [103]. Interestingly, while the normal coronary intima has only low numbers of mast cells, the intima of healthy aortas contains significant numbers of mast cells [101]. In sharp contrast to human beings, in the normal mouse no mast cells are present in the aortic intima or media. However, since the distance between the intima and adventitia is small in mice, the adventitial mast cells may well be capable of influencing the atherogenic processes that occur within the intima.

Mast cells and atherosclerosis

A short history

Originally, mast cells were suggested to participate in the pathogenesis of atherosclerosis by Constantinides in 1953 [104, 105]. Three years later McGovern suggested that, by residing in the immediate vicinity of endothelial surfaces, human mast cells may regulate arterial thrombus formation by releasing the antithrombotic mediator, heparin [106]. Shortly thereafter, the increased numbers of adventitial mast cells were found to correlate with the progression of atherosclerosis, their numbers being particularly high in areas of arterial thrombosis [107]. Importantly, the highest numbers of intimal mast cells have been found at sites of rupture and/or erosion in infract-related coronary arteries [35]. Similar to the coronary arteries, the numbers of mast cells in the carotid arteries increased as the atherosclerotic lesions became more advanced [108, 109]. However, in contrast to coronary and carotid lesions, the number of intimal mast cells in the aorta seemed to decline as the atherosclerotic disease progressed [101]. Interestingly, a general role for mast cells in atherosclerosis has been suggested in a study with Brown Norway rats, in which the authors showed that, after induction of atherosclerosis by immunization with ovalbumin, the intimal thickness correlated positively with the level of chymase [110]. In addition, preliminary data in our laboratory suggest that mast cell-deficient KitW-sh/W-sh mice on a LDLR⁻/⁻ background develop less atherosclerosis when fed an atherogenic western-type diet (Fig. 1). Thus, increasing in
vitro and in vivo evidence suggests that mast cells contribute to the early pathogenesis of atherosclerosis by promoting intimal lipid accumulation and foam cell formation by multiple mechanisms [46] (Fig. 2). Mast cells have also been shown to associate with neovessels formed within the atherosclerotic plaques [111–113]. Although neovessels are a common feature of advanced human atherosclerotic plaques, it is unclear whether angiogenesis, by causing intraplaque hemorrhage and growth of lipid core, is responsible for plaque instability [114]. Interestingly, data from clinical cancer trials have shown that prevention of angiogenesis with antibodies against VEGF, bevacizumab (Avastin®), leads to an increased risk of thromboembolic complications, including cerebrovascular events, myocardial infarction and deep vein thrombosis [115]. Furthermore, current evidence from clinical trials of both proangiogenic and antiangiogenic therapies suggests that inhibition of angiogenesis is not a viable therapeutic strategy for cardiovascular diseases, and that VEGF-induced angiogenesis in human arteries may rather play a protective role [116].

**Activated mast cells in human atherosclerotic plaques**

When compared with normal intima, the number of activated mast cells in atherosclerotic plaques is especially high in the shoulder regions prone to plaque rupture [117, 118]. Upon activation, mast cells release their preformed mediators in an active exocytotic process called degranulation, and most importantly, patients who died of acute myocardial infarction had an increased number of degranulated cell-deficient mice. To test the role of mast cells in the pathogenesis of atherosclerosis in vivo, mast cell-deficient mice (Kit^{-/-}/LDLR^{-/-}Kit^{W-sh/W-sh}) were crossed with LDLR-deficient mice (LDLR^{-/-}/Kit^{W-sh/W-sh}) double knockout mouse model. As shown in Figure 1, the extent of aortic atherosclerosis that is induced in an LDLR^{-/-}/Kit^{+/+} mouse (A) by a Western-type diet for 9 weeks is slightly attenuated in an LDLR^{-/-}/Kit^{W-sh/W-sh} mouse (B).

**Fig. 1** Diet-induced atherosclerosis is attenuated in mast cell-deficient mice. To test the role of mast cells in the pathogenesis of atherosclerosis in vivo, mast cell-deficient mice (Kit^{W-sh/W-sh}) were crossed with LDLR-deficient mice (LDLR^{+/+}) to obtain a LDLR^{-/-}/Kit^{W-sh/W-sh} double knockout mouse model. As shown in Figure 1, the extent of aortic atherosclerosis that is induced in an LDLR^{-/-}/Kit^{+/+} mouse (A) by a Western-type diet for 9 weeks is slightly attenuated in an LDLR^{-/-}/Kit^{W-sh/W-sh} mouse (B).
Fig. 2 A role for mast cells in the intra- and extracellular accumulation of low density lipoprotein (LDL) cholesterol in atherogenesis. Upon activation, the mast cell secretes its preformed granules by exocytosis into the subendothelial space of the arterial intima. Histamine, a mast cell-derived soluble mediator increases the transendothelial transport of plasma LDL into the subendothelial space, where it is bound by the heparin component of the insoluble granule remnants and degraded by chymase, a granule remnant neutral serine protease. The proteolytically modified LDL particles become unstable and fuse on the remnant surface, after which the macrocomplex is phagocytozed and degraded by the macrophage with the subsequent formation of a foam cell, a hallmark of an early atherosclerotic lesion. High density lipoprotein (HDL), being responsible for the efflux of LDL-derived cholesterol from the macrophage foam cell, is also proteolyzed by mast cell chymase and the high-affinity component of the HDL-dependent cholesterol efflux is so impaired. As a result, the balance between cholesterol influx and efflux is disturbed and a cholesterol-filled foam cell is formed. The foam cell may eventually die and so contribute to the formation of an extracellular lipid core, which is a hallmark of an advanced atherosclerotic lesion.
mast cells at the actual site of plaque erosion or rupture [35]. In general, mast cells can be activated by a wide variety of physiological and non-physiological mediators [86]. In atherosclerotic plaques, the accumulating modified lipids, other inflammatory components or the neighboring cells may be responsible for mast cell activation. Interestingly, the level of mast cell degranulation was elevated in intimal areas that contained an increased number of macrophages and T-lymphocytes, suggesting that factors responsible for mast cell degranulation in vivo may be derived from neighboring inflammatory cells. Since the complement anaphylatoxins, C5a and C3a are capable of inducing mast cell activation [119], and since the complement cascade is a functional component of the inflammatory process in advanced human coronary plaques [120], it is possible that they trigger mast cell-mediated responses in the coronary plaques. Recently, oxidized LDL that are present in high amounts in atherosclerotic lesions, have been shown to induce mast cell activation and secretion of IL-8 [121]. The mechanisms involved in this activation are still unclear, but may involve Toll-like receptors and some type of molecular mimicry. Smoking is also a risk factor for atherosclerosis, and nicotine present in smoke extract has been shown to activate human mast cells [122], and moreover chronic exposure to cigarette smoke induces the expression of mast cell proteases [123].

The activation of adventitial mast cells may occur by similar mechanisms. In addition, sensory nerves that are able to secrete substance P and CGRPs, are potential activators of the adventitial mast cells. Indeed, the number of mast cells were higher in the adventitia of advanced atherosclerotic coronary lesions (104 mast cells/mm²) compared to normal intima (31 mast cells/mm²), and the number of mast cell–nerve contacts (30 nerve contacts/mm²) were significantly greater than in normal intima (four nerve contacts/mm²) [124]. In atherosclerotic lesions, neurogenic activation of adventitial mast cells may involve the release of vasoactive compounds, such as histamine and leukotrienes, which can induce coronary vasoconstriction [124].

**Mast cell activation without degranulation**

Previously, activation of mast cells was thought to exclusively involve the process of degranulation. However, an evolving concept suggests that activation of mast cells in tissues during disease progression may occur without signs of degranulation [125–127]. Recently, both IL-1 and TNF-α have been shown to induce mast cell-mediated secretion of cytokines in the absence of degranulation, i.e. without secretion of histamine and other granule components. Since the physiological and pathophysiological compounds capable of activating mast cells are so diverse [86], it is also likely that different mechanisms of mast cell activation exist. These would then range from selective secretion of individual cytokines or chemokines to anaphylaxis with exhaustive degranulation and acute secretion of a multitude of cytokines and chemokines.

**Mast cell – an inflammatory cell**

Due to their strategic localization in the close vicinity of endothelial cells, mast cells are involved in the regulation of innate and acquired immune responses, such as the infiltration of T-cells, macrophages and neutrophils [23, 86]. In human atherosclerotic plaques, T-cells co-localize with mast cells, suggesting a close interaction between these cell types. The direct contact between the two cell types may not only influence the functions of the individual cells, but may exert a more general immunoregulatory function that affects their microenvironment [128–130]. For instance, inflammatory areas of ruptured human atherosclerotic plaques have been shown to contain CD30 [131], and CD30–CD30L interaction is known to induce degranulation-independent secretion of chemokines by mast cells [127]. In addition, mast cells may activate T-cells by releasing TNF-α [132] and by expressing the costimulatory OX40L, which induces a direct cell–cell interaction between mast cells and the OX40 receptor on T-cells [133].

How could the Th1/Th2 polarization in atherosclerotic lesions affect the local development and activation of mast cells? Since human mast cells obey SCF-dependent, cytokine-driven (IL-3, IL-5, IL-6, IL-9 and granulocyte/macrophage colony-stimulating factor) mitogenic responses, and since they have a unique profile of chemokine receptors (e.g. CCR3), an increased infiltration and differentiation of mast cells reflects a Th2-type polarization [7]. In contrast, a Th1-type polarization, involving high expression of IFN-γ is inhibitory for human mast cell growth and
differentiation [134]. Since atherosclerosis is generally considered to be a Th1-driven disease, which at the early stages of fatty streak formation may be counteracted by Th2-mechanisms [135], the role of mast cells may change with the progression of the disease. Indeed, in advanced stages of atherosclerosis, the Th2-driven processes might also be proatherosclerotic [136, 137]. Indeed, at least in the mouse severe hypercholesterolaemia is associated with advanced atherosclerotic plaques and with a switch from Th1 to Th2-driven processes. Since Th1 and Th2 cells counteract each other, the appearance of Th2 cytokines may have important consequences for the inflammatory/immune process in atherosclerosis [138], and especially in terms of mast cell proliferation and differentiation.

Mast cells also induce infiltration of monocytes and neutrophils [139] by secreting various chemokines, such as MCP-1 and IL-8. The participation of monocyte/macrophages in plaque remodeling that predispose to rupture is well established [140]. In addition, the mast cell-dependent recruitment of neutrophils may be of special importance in the infarct-related coronary plaques, which, in contrast to the more innocent stable plaques, often contain neutrophils [141]. Indeed, mast cells participate in the induction of antigen- and Th17 cell-dependent neutrophil-rich inflammatory responses [142], and IL-17, the main mediator of Th17-driven inflammation, is induced in unstable angina and acute myocardial infarction [143].

Mast cells are also known to cause endothelial dysfunction and activation of endothelial adhesion molecules [30], effects that also may predispose to plaque erosion and rupture. Mast cell-derived histamine increases the permeability of EC by binding to histamine receptors (H1 receptors) on the endothelial cell surface so increasing phosphorylation of adherent junction molecules and loosening of vascular endothelial (VE)-cadherin-mediated endothelial cell–cell adhesions [147–149], PAR-2 cleavage and activation by mast cell-derived tryptase, chymase and cathepsin G may also induce loosening of VE-cadherin-mediated cell–cell adhesions [150–152].

Activation of adventitial mast cells by nervous stimuli may regulate vessel tone and also various functions of vasa vasorum [124], and so participate in events leading to plaque erosion and rupture. Indeed, histamine released from activated adventitial mast cells may reach the media, where it could act on smooth muscle cells, and by provoking a local spasm, histamine could contribute to the onset of acute cardiac events [146, 153]. Interestingly, the proportion of activated and degranulated adventitial mast cells was found to be highest in the segments with ruptured plaques [102]. However, it is presently not known whether activation and degranulation of the adventitial mast cells precede the rupture or occur as a consequence of plaque rupture.

Animal models of plaque erosion and rupture

Studies with experimental animals have shown that most species spontaneously develop fatty streaks and some even more advanced atherosclerotic lesions with time, but that plaque rupture with subsequent thrombosis is exceedingly rare. In most animal models, dietary addition of fat and cholesterol have induced an accelerated development of advanced lesions, but again, without an increased frequency of plaque rupture [154]. Although some studies have shown that artificial induction of neointima by ligation of the common carotid artery [155], by non-constrictive cuffing of the femoral artery [156], or by a combination of both [157, 158] may lead to endothelial damage and thrombus formation, they are not optimal models to study mechanisms involved in the rupture of...
vulnerable atherosclerotic plaques in man. So far, only one group has showed that fat-fed apoE knock-out mice develop advanced atherosclerotic plaques in their brachiocephalic arteries with frequent rupture and formation of luminal thrombi [159]. According to these data, the ruptured plaques show many of the characteristics of vulnerable plaques in human beings [159], and the ruptures can be prevented by statin treatment [160]. Thus, by crossing the above apoE-deficient model with a mast cell-deficient mouse model (Kit$^{W-sh/W-sh}$), one would potentially create an animal model that would clarify the in vivo role of mast cells in plaque erosion and rupture. However, one should keep in mind that rodent mast cells are different from human mast cells in terms of both mediator contents and functions [161, 162], and that observations made in such animal models may not truly reflect the human disease. Indeed, one of
the major obstacles in the use of animal models for studying the role of mast cells in atherosclerosis is the lack of intimal mast cells in many commonly used laboratory animals.

Mice do have mast cells in the aortic adventitial layer, and since human adventitial mast cells are activated in the areas of atherosclerotic plaques [102], the adventitial mouse mast cells may be able to affect pathological processes in the arterial intima. Interestingly, another experimental approach to induce accelerated atherosclerosis in animal models has been the use of arterial cuffs [163, 164], that may activate the adventitial mast cells. In addition, recent data show that activation of adventitial mast cells promote atherogenesis and induce plaque destabilization in apoE-deficient mice [165]. A targeted activation of adventitial mast cells in advanced carotid atherosclerotic plaques sharply increases the incidence of intraplaque haemorrhage, macrophage apoptosis, vascular leakage and CXCR2/VLA-4-mediated recruitment of leukocytes to the plaque [165].

Clinical approaches to stabilize mast cells in the atherosclerotic plaque

Pathological vasoconstriction or spasm of inflamed atherosclerotic coronary segments plays an important role in acute coronary syndromes and has also been suggested to pre-dispose to plaque rupture and erosion [41, 166, 167]. Since mast cell-derived histamine and leukotrienes have been suggested to trigger coronary spasm [124, 146] and since both antihistamines and leukotriene receptor antagonists are available for human use [168], clinical studies regarding their use in the treatment of atherosclerosis may be feasible in the future [169]. Interestingly, an H1-antihistamine (desloratadine) has been shown to inhibit mast cell activation in vitro [170] and simultaneous inhibition of H1 and H2 receptors may be even more efficient in mast cell stabilization than inhibition of either receptor alone [171]. Taken together, antihistamines and leukotriene receptor antagonists might provide means to inhibit progression of atherosclerosis by reducing plaque inflammation. These drugs may potentially also prevent coronary spasms.

A major mechanism pre-disposing to plaque rupture is the increased degradation of extracellular and pericellular matrix components in the fibrous cap. The activated mast cells secrete proteases that either directly or indirectly can induce degradation of collagen, elastin and proteoglycans [172]. Degradation of the extracellular and pericellular matrices induces apoptosis and weakens the fibrous cap rendering it more susceptible to rupture and precipitation of an acute coronary syndrome. Although pharmacological inhibition of MMPs and cathepsins has been proposed in the treatment of atherosclerosis [173, 174], many of these proteases are crucial for normal vessel homeostasis [85, 175]. In addition, since the mast cells present in the plaque are filled with the neutral proteases, tryptase, chymase and cathepsin G, which they upon activation avidly secrete into the extracellular space, a therapeutic need to inhibit their activity in vivo may exist. However, although tryptase inhibitors have been used in the treatment of asthma and ulcerative colitis [176], they have not yet been tested in atherosclerosis. Heparin antagonists, such as protamine and Polybrene® also inhibit tryptase activity by causing dissociation of active tetrameric tryptase into inactive monomers [61]. Several orally active inhibitors of chymase are currently also available [177], and animal data support their benefits in a variety of cardiovascular diseases [178, 179]. However, human clinical data regarding the effects of chymase inhibitors in the pathogenesis of atherosclerosis are still lacking. Taken together, the key enzymes responsible for the pathological matrix degradation should be identified before specific antiprotease therapy can be envisioned, and even then the redundancy of proteases may render this approach unsuccessful.

For the time being, anti-inflammatory therapies aiming at lowering the numbers of mast cells or stabilizing mast cells may be the best way of lessening their proteolytic burden in the vulnerable cap. Although sodium cromolyn has successfully been used to stabilize human mast cells locally in the eye, it is presently not suitable for systemic use in human beings. Interestingly, intravenously and intraperitoneally given cromolyn treatment during dinitrophenyl-albumin (DNP) challenge of collar-induced carotid artery lesions in apoE-deficient mice normalized the extent of mast cell degranulation in the adventitia, while preventing intraplaque hemorrhage [165]. An orally absorbable phosphodiesterase (PDE5) inhibitor
zaprinast has also been shown to stabilize mast cells and to possess a moderate bronchodilator effect [180]. PDE5 inhibitors were tested also for treatment of angina pectoris, but proved not to have any beneficial effect over nitrates [181]. Since SCF is necessary for mast cell development, proliferation and survival, and participates in homing and adhesion of mast cells, it may also be a target of therapy [182]. Indeed, drugs targeting SCF and/or Kit, including anti-SCF antibodies, antisense oligonucleotides and Kit inhibitors have been studied for their anti-allergic properties [182], and thus, may be useful in stabilizing mast cells present in atherosclerotic plaques.

Furthermore, statins have been shown, at least in vitro, to inhibit the secretion of MMPs by macrophages [183], but no data on the effects of statins on the release of mast cell proteases appear to be available. However, previous studies in a rat mast cell line (RBL-2H3) have shown that statin-treatment (lovastatin) resulted in impaired tyrosine phosphorylation and in inhibition of the IgE-mediated degranulation [184]. In addition, recent results have shown that cerivastatin and atorvastatin, and partially also lovastatin act as inhibitors of growth and function of human mast cells [185]. However, simvastatin and pravastatin did not affect mediator release or growth of human mast cells, revealing that the inhibitory effects were not a class effect, but peculiarly specific for particular statins [185].

Conclusions

Accumulating evidence clearly suggests a role for mast cells in the pathogenesis of plaque erosion and rupture [144] (Fig. 3). Their strategic tissue location, vast content of powerful mediators ranging from vasoactive substances and proinflammatory cytokines to proteolytic enzymes, ability to rapidly become activated upon contact with specific triggers, and ability to regulate innate and acquired immune responses, are attributes which set the scene for their view to a kill. However, we eagerly await more animal models that would produce proof of concept and give us the confidence to develop means to treat against mast cell infiltration and activation in the pathogenesis of plaque erosion and rupture. Although we presently think that chronic activation of mast cells in the atherosclerotic lesions pre-disposes to plaque rupture, it may well be that the initial activation of the intimal and adventitial mast cells are necessary for the onset of immunological defense mechanisms in the arterial wall [135, 186]. Accordingly, mast cells have been shown important for the healing responses of injured arterial walls, in that they participate in the initiation of thrombus formation [187], as well as in the resolution of thrombus formation and in the process of neovascularization [29, 188]. Thus, mast cells, by being powerful regulators of immune responses and inflammation, may represent a double-edged sword in the pathogenesis of plaque erosion and rupture [135].

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