Mosquitoes with their ability to transmit several pathogens of human disease pose a serious threat to healthcare worldwide. Although much has been done to prevent the disease transmission by mosquitoes. The rising rate of resistance in mosquitoes towards conventionally used control strategies necessitates developing of novel strategies to counter disease transmission. The mosquito brain plays a key role in host-seeking, finding mates and selection of oviposition sites. However, not much is know about the underlying physiological processes in mosquito brain. The data presented in this study describes the proteins that have been identified in the brain tissue of adult female Anopheles stephensi and their associated processes. Interpretation of the data can be related to the previously published article “Integrating transcriptomics and proteomics data for accurate assembly and annotation of genomes” [1].

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Specifications Table

| Subject                          | Biotechnology                          |
|---------------------------------|----------------------------------------|
| Specific subject area           | Proteomics and Vector Biology          |
| Type of data                    | Table                                   |
|                                 | Chart                                   |
|                                 | Graph                                   |
| How data were acquired          | LTQ-Orbitrap Velos and LTQ-Orbitrap-Elite mass spectrometers (Thermo Scientific, Bremen, Germany) |
|                                 | Proteome Discoverer 2.1, with SEQUEST and MASCOT search engine (Matrix Science, London, UK; version 2.2) |
|                                 | Anopheles stephensi (Indian strain) protein database (www.VectorBase.org, release February 25, 2014) |
| Data format                     | Raw, Analyzed                           |
| Parameters for data collection  | An. stephensi mosquitoes were maintained under laboratory conditions at 27 ± 2°C, 75% relative humidity and photoperiod of 12h of day-night cycle. The adult female mosquitoes were fed on 10% glucose solution soaked in cotton pad. Brain tissues were dissected under dissecting microscope using 0.65% normal saline and further stored at -80°C. |
| Description of data collection | Brains were homogenized in using sonication and extracted proteins were fractionated at peptide and protein level. Digested peptides were analysed using tandem mass spectrometry. Data obtained from tandem mass spectrometry was searched against the An. stephensi reference protein database of identification of proteins. Identified proteins were then analysed for their functions and interacting partners using bioinformatic tools. |
| Data source location            | Panaji, Goa, India                      |
|                                 | Bangalore, Karnataka, India             |
| Data accessibility              | Repository name: PRIDE (PProtein IDENTification Database) |
|                                 | Data identification number: PXD001128 |
|                                 | Direct URL to data:                     |
|                                 | https://www.ebi.ac.uk/pride/archive?keyword=PXD001128 |
|                                 | Analyzed data is provided along with this article as excel sheets. |
| Related research article        | Prasad et al. Integrating transcriptomic and proteomic data for accurate assembly and annotation of genomes. Genome Res. 2017;27(1):133–144. doi:10.1101/gr.201368.115 |

Value of the Data

- The data provides information on proteins expressed and provides insights into the underlying processes in the adult female An. stephensi brain.
- The data provides a platform for researchers working towards identification of key proteins associated with mosquito vector-pathogen interactions, host-seeking behavior and transmission blocking vaccines.
- The data provides a comprehensive list of genes that can be studied further in a variety of experiments to understand mosquito physiology, immunity, complexities of host attraction processes (host seeking behavior), avoidance of insecticide sprayed surfaces, resting, feeding and reproductive behavior and also oviposition site preferences of female An. stephensi mosquitoes in nature.

1. Data Description

The dataset provided in this article describes the proteins identified in female Anopheles stephensi brain and their associated processes and function Fig. 1. Tandem mass spectrometers-based acquisition of peptides derived from adult female An. stephensi brain samples led to the
identification of 4,662 proteins Supplementary Table S1, S2. The data provided in this article can be related to our previously published article “Integrating transcriptomics and proteomics data for accurate assembly and annotation of genomes” [1]. The identified proteins were further categorized based on their Gene Ontology term associations. Details of the Gene Ontology categorization for the identified proteins are provided in Fig. 2 and Supplementary Table S3. Fig. 3 illustrates the predicted interacting protein clusters along with their associated processes for the proteins identified in female *Anopheles stephensi* brain. The complete details of predicted protein-protein interaction have been provided in Supplementary Table S4. List of genes expressed in the female *An. stephensi* brain and with potential role in sporogony [2] have been provided in Supplementary Table S5. Processes and pathways for these 28 genes have been illustrated in Fig. 4 and the complete details are provided in Supplementary Table S6 and S7. The complete list of proteins identified in female *Anopheles stephensi* brain and mapping to Immunodb data is provided in Supplementary Table S8. Further analysis of these 54 genes for their associated pathways have been illustrated Fig. 5 and the complete details of the analysis result have been provided in Supplementary Table S9 and S10.
Fig. 2. Gene Ontology analysis of the identified protein in An. stephensi brain. A) Biological processes; B) Molecular function; C) Cellular components of the identified proteins. Only p value significant terms and groups were considered. Blue line represents term p value while red line represents group p value.

2. Experimental Design, Materials, and Methods

2.1. Maintenance of mosquito colony

An. stephensi mosquitoes required for the study were obtained from the insectary of ICMR- National Institute of Malaria Research, Field Station Goa. Mosquitoes were maintained under laboratory conditions and dissection of mosquito tissues were carried out as mentioned in our previous study [1].

2.2. Protein extraction

As previously discussed, proteins were extracted from adult female brains by homogenization using a probe sonicator in 200 μl of 4% SDS buffer. The extracted protein amount was quantified using Lowry’s method (Bio-Rad DC Protein assay) [3].
Fig. 3. Predicted protein-protein interaction map of sub-set of proteins identified in the An. stephensi brain along with the associated processes. The various processes involved is depicted in different color shades with the details provided in the box provided in the right-hand corner below.

2.3. Fractionation

Fractionation was carried out as discussed in our previously published study [1]. Briefly, for in-gel digestion, extracted proteins (300 μg) was fractionated on 10% SDS PAGE stained with Coomassie blue and bands were cut out based on intensity of protein bands. Excised bands were processed for overnight trypsin digestion using sequencing grade trypsin (Promega). For in-solution digestion, 500 μg of protein were trypsin digested and lyophilized prior to bRPLC fractionation. Fractionated peptides were concatenated into 14 fractions.

2.4. Mass spectrometry analysis

As discussed in our previously published study, fractions were reconstituted in 0.1% formic acid prior to mass spectrometry analysis [1]. Fractions were analysed on LTQ-Orbitrap Velos and
Fig. 4. Representation of various processes and pathways and their associated genes that are expressed in An. stephensi brain and have a role in vector-pathogen interactions. A) Biological processes associated with An. stephensi brain related genes that play key roles in vector-pathogen interactions. B) Associated pathways and their genes of the genes involved in vector-pathogen interactions and identified in An. stephensi brain.

LTQ-Orbitrap-Elite mass spectrometers (Thermo Scientific, Bremen, Germany) interfaced with Easy-nLCII (Thermo Scientific, Bremen, Germany). Data acquisition was carried out in a data dependent manner as described in our previous study [1].

2.5. Data analysis

Mass spectrometry data obtained from our previous study [1] was searched against the reference database containing 11,789 An. stephensi proteins along with common mass spectrometry contaminants. Data was analyzed using Proteome Discoverer, version 2.1 (Thermo Fischer Scientific, Bremen, Germany). Overall, 4,662 proteins were identified. Database search parameters included trypsin as digestion enzyme. Amino acid modification considered were Carbamidomethylation of cysteine as static and oxidation of methionine as variable. Results were retrieved using 1% false discovery rate (FDR).
2.6. Functional categorization and prediction of interaction map

The identified An. stephensi proteins were used to fetch corresponding ortholog genes of An. gambiae using Biomart online tool available through Vector Base [4-5]. The corresponding An. gambiae genes were then used to fetch Gene Ontology terms using Cluego plugging and the resulting figure was generated using Cluepedia plugging in Cytoscape [6-8]. A predicted protein-protein interaction map for the identified proteins were also generated using STRING (Search Tool for the Retrieval Interacting Genes/Proteins) online tool (version 10.5).plugin (version 1.1.0) [9]. Highest confidence parameter was used to retrieve interaction data from STRING database.
Possible immunological associations of the identified proteins were inferred from the information provided in Immunodb [10].

Ethics Statement

No human subjects or lab animals were used for this study. Mosquitoes used for the study were obtained after approval from National Institute of Malaria Research (NIMR) Scientific Research and Ethics Committee.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships which have, or could be perceived to have, influenced the work reported in this article.

Acknowledgments

We thank Institute of Bioinformatics and ICMR- National Institute of Malaria Research for institutional support and research facilities. We thank Science and Engineering Research Board (SERB), Government of India, for funding (EMR/2014/000444) to NIMR and IOB to characterize proteome and transcriptome of mosquito vectors. The NIMR publication committee has approved this manuscript. We also thank Karnataka Biotechnology and Information Technology Services (KBITS), Government of Karnataka for the support to the Center for Systems Biology and Molecular Medicine at Yenepoya (Deemed to be University) under the Biotechnology Skill Enhancement Programme in Multiomics Technology (BiSEP GO ITD 02 MDA 2017).

Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.dib.2020.106243.

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