Spectroscopic Evaluation of Chalcone Derivatives and their Zinc Metal Complexes: A Combined Experimental and Computational Approach on the Binding of the Complexes with the Serum Albumin

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Research Article

Keywords: Chalcone, Zinc, Spectroscopy, Docking, Molecular dynamics, Complexes, DFT, BSA

DOI: https://doi.org/10.21203/rs.3.rs-136887/v1

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Abstract

Three chalcone derivatives (L₁, L₂, L₃) were synthesized using Claisen-Schmidt condensation reaction. Their molecular structures and spectroscopic properties (IR, UV-vis, ¹H NMR), were calculated at B3LYP level. Electrostatic interactions and HOMO-LUMO properties were calculated using TD-DFT method. Molecular docking was used to compare the HSA (human serum albumin) interactions with the ligands and their Zn complexes (C₁, C₂, C₃) which were synthesized by interaction between the ligands and the Zn (II) ion in a 2:1 M ratio. Elemental analysis, FT-IR, and UV–Vis spectroscopy studies investigated the structure of the synthesized complexes. UV–Vis, molecular docking and molecular dynamics were used to study the interactions of the Zn complexes with the BSA (bovine serum albumin). The biological activity of the Zn-Chalcone complexes was generally higher than the chalcones when evaluated spectroscopically and theoretically.

1. Introduction

Chalcones, are belonging to the flavonoid family, having a medicinal importance because of the CO-CH=CH- (ketoethylenic group). There are intermediates of flavonoids and they exhibit anti-inflammatory, antifungal, antimicrobial, antioxidant and antitumor activity [1,2]. This biological activity is because of the α, β-unsaturated keto function. Chalcone contains two aromatic rings linked by a three carbon α, β-unsaturated carbonic system and they are selected because of their low toxicity and possible chemical modification [3,4]. In nature, chalcone (1,3-diphenyl-2-propen-1-one) is one of the open chain flavonoids containing 15-carbon arranged in a C6C3-C6 configuration [5]. The bioactivity of chalcones have been studied and several biological activities have been found such as, antioxidant, cytotoxic, antiviral, tyrosinase inhibitory, antimalarial, antibacterial, and anti-inflammatory [6]. Additionally, the synthesis of Pd (II) and Pt (II) complexes and Co (II), Cu (II), Mn (III) of a chalcone compound has been proposed elsewhere [7,8] and this is because chalcones are effective metal ion chelators and they can easily create metal-coordinated compounds [9]. Chalcones are synthesized by Claisen–Schmidt condensation. This reaction involves cross aldol condensation of appropriate aldehydes and ketones by base catalysed or acid catalysed reactions followed by dehydration [10].

It is known that Zinc-containing compounds play a key role in carbonic anhydrases, peptidases, proteases, and deaminases [11]. Interestingly, the coordination chemistry of zinc in proteins and peptides involves N, O, and S donors of the side chains of histidine, glutamate/aspartate, and/or cysteine with any permutation of these ligands and with the number of protein ligands ranging from three to six [12]. On the other hand, Zn plays a vital role in a variety of biological processes but, excessive exposure of Zn²⁺ to human beings can cause toxicity, inducing a series of overt poisoning symptoms and neurodegenerative disorders [13].
HSA is responsible to maintaining stable osmotic pressure and to carrying endogenous or exogenous compounds. Additionally, most lipophilic drugs are substrates of HSA and are transferred to target tissues via the bloodstream. The chemical structure of HSA includes three homologous α-helical domains (I, II and III), each of which possesses two subdomains (A and B) [14]. Various binding and denaturation studies have shown to be a rapid and effective tool for the characterization of albumin binding sites and their enantioselectivity, and for the study of the changes in the binding properties of the protein caused by interaction between different ligands [15-18].

The last years more and more scientists are using computational chemistry and theoretical tools to evaluate their molecular structures [19-21]. Moreover, some researchers are using computational tools to evaluate the biological activity of the molecules of interest [22-26]. This is because of the quick results that the computational tool is giving to the researcher, and the added scientific value to the findings. All that, at minimum cost and resources. As the technological improvements are running fast, more and more theoretical tools are going to save time to the researcher and give a different perspective, using the predictive character of the computerized models.

Herein, we propose the synthesis of some chalcone analogues, and their complexation with Zn (II) metal ions. In total, 6 molecules were prepared and evaluated spectroscopically, both in situ and theoretically. More specifically, we used TD-DFT studies [26-31], to evaluate our chalcone derivatives in terms of structure and activity, molecular docking [32-35], to evaluate their interaction with human serum albumin (HSA) and spectroscopy to evaluate their binding interactions with bovine serum albumin (BSA). The biological activity of the chalcone derivatives, was compared with the biological activity of their counter Zn complexes.

2. Experimental

2.1. Materials

2-nitrobenzaldehyde, 2-(methylamino) benzaldehyde, 2-hydroxy-5-methylphenyl ethanone, 1-(2-hydroxy-5-methylphenyl) ethanone, 4-hydroxy-3-methoxyphenyl ethanone, were purchased from Merck. Solvents were used without any further purification. Other reagents were of analytical grade. BSA (bovine serum albumin) (purity ≥ 98%) was purchased from Merck.

2.2 Instruments

All the chalcone analogues and their zinc complexes were characterized by $^1$HNMR, recorded on Bruker 300 MHz spectrometer using DMSO-d6 as solvent and TMS as an internal standard. The chemical shifts were expressed in $\delta$ ppm. The absorption spectrum of all the reaction mixtures was then taken in a range of 200-400 nm in a JASCO (Tokyo, Japan) UV-visible spectrophotometer using a 1cm path length quartz cuvette.

2.3 Synthesis of chalcones analogues
2.3.1 Synthesis of (E)-1-(2-hydroxy-5-methylphenyl)-3-(2-nitrophenyl) prop-2-en-1-one

In a round bottom flask (100 ml), to a methanolic solution (20 ml) of 2-hydroxy-5-methylphenyl ethanone (0.003 mol) a 40% aqueous NaOH solution (15 mL) was gradually added. 2-nitrobenzaldehyde (0.003 mol) was added and the reaction took place for 20 h at 25°C. Any precipitate formed was removed by filtration and the filtrate was acidified with dilute HCl and then extracted with chloroform. The concentrate of the chloroform was chromatographed over silica gel to obtain the desired product. The compound then crystallized from chloroform-petroleum ether (50:50). Olive green solid, M. P: 55-57 °C; \( ^1 \)H NMR (300 MHz, DMSO-d6): \( \delta \) 2.36 (s, -CH\(_3\)), 5.37 (s, -OH), 7.04 (m, -CH aromatic), 7.36 (m, -CH aromatic), 7.44 (m, -CH aromatic), 7.65 (m, -H), 7.89 (m, -CH), 8.23 (m, -CH), 8.64 (m, -H) ppm.

2.3.2 Synthesis of (E)-3-(4-dimethylamino) phenyl)-1-(2-hydroxy-5-methylphenyl) prop-2-en-1-one

In a round bottom flask (100 ml), to a methanolic solution (20 ml) of 2-hydroxy-5-methylphenyl ethanone (0.003 mol) a 40% aqueous NaOH solution (15 mL) was gradually added. 4-dimethylamino benzaldehyde (0.003 mol) was added and the reaction took place for 20 h at 25°C. Any precipitate formed was removed by filtration and the filtrate was acidified with dilute HCl and then extracted with chloroform. Orange solid, M. P: 54-55 °C; \( ^1 \)H NMR (300 MHz, DMSO-d6): \( \delta \) 2.34 (s, -CH\(_3\)), 3.06 (s, -CH\(_3\)), 5.35 (s, -OH), 7.02 (m, -CH aromatic), 7.34 (m, -CH aromatic), 8.33 (d, -H) ppm.

2.3.3 Synthesis of (E)-1-(2-hydroxy-5methoxyphenyl)-3-(2-nitrophenyl) prop-2-en-1-one

In a round bottom flask (100 ml), to an ethanolic solution (20 ml) of 1-(4-hydroxy-3methoxyphenyl) ethanone 0.003 mol) a 40% aqueous NaOH solution (20 mL) was gradually added. 2-nitrobenzaldehyde (0.003 mol) was added and the reaction took place for 24 h at 25°C. Any precipitate formed was removed by filtration and the filtrate was acidified with dilute HCl. Yellow-green solid, M. P: 57-59 °C; \( ^1 \)H NMR (300 MHz, DMSO-d6): \( \delta \) 3.84 (s, -CH\(_3\)), 5.33 (s, -OH), 7.10 (m, -CH aromatic), 7.27 (m, -CH aromatic), 7.79 (m, -CH aromatic), 7.89 (m, -CH aromatic), 8.21 (m, -CH aromatic), 8.60 (d, -H).

2.4 Synthesis of Zinc (II) Complexes

Zn (II) complexes with ligands L\(_1\)-L\(_3\)

An ethanolic solution (30 ml) of Zn (II) chloride (0.01 mol) was added to a refluxing solution of appropriate chalcone analogue L\(_1\)-L\(_3\) (0.02 mol) in ethanol (30 ml). The reaction mixture was refluxed for 6 h. The coloured complexes were obtained, filtered off, washed with ethanol and dried under vacuum. Elemental Analysis: L1: C, 54.84; H, 3.45; Cl, 10.12; N, 4.00; O, 18.26; Zn, 9.33. L2: C, 62.04; H, 5.21; Cl, 10.17; N, 4.02; O, 9.18; Zn, 9.38. L3: C, 52.45; H, 3.30; Cl, 9.68; N, 3.82; O, 21.83; Zn, 8.92.

2.5 Theoretical Studies

2.5.1 Optimization and vibration frequency calculations were made at B3LYP level with ORCA version 4.0.1 program. B3LYP is a hybrid density functional theory method. Among the ever-increasing number of
DFT methods, the hybrid functional B3LYP, as a good compromise between computational cost, coverage, and accuracy of results [36]. It has become a standard method used to study organic chemistry in the gas phase. UV-vis spectra were calculated at the same level calculated by time-dependent density functional theory (TD-DFT) method. ORCA input files were created by AVOGADRO version 1.2.0 software. HOMO energy ($E_{HOMO}$) and LUMO energy ($E_{LUMO}$) were taken from the output file. Chalcone analogues and their zinc complexes were docked against human serum albumin. Molecular docking studies were carried out by using iGEMDOCK 2.1 software [11]. The HSA coded crystal structure was selected from the Protein Data Bank (www.rcsb.org). The population size was $= 200$, generations $= 70$, number of solutions $= 3$.

2.6 Effect of the ligand and the complex on the absorption spectrum of BSA

The effect of the ligands and their corresponding complexes, on the absorption spectrum of BSA, was studied using UV–visible spectrophotometry. Briefly, BSA (5 μM) were incubated in the absence and presence of 2-9 μM of L$_1$, L$_2$, L$_3$, C$_1$, C$_2$ and C$_3$ for 30 min at RT.

3. Results And Discussion

3.1 Synthesis of Chalcone Derivatives and their Zinc Complexes

Chalcone derivatives were synthesized based on Claisen-Schmidt condensation reaction. Acetophenones analogues reacted in (1:1) ratio with benzaldehydes analogues in methanolic solutions resulting the chalcone derivatives (L$_1$, L$_2$ and L$_3$). The synthetic routes and conditions are depicted in Fig. 1. Reactions took place at room temperature resulting colourful precipitates after 20 h mixing. Any precipitate formed was removed by filtration before acidification. The formation of carbanion or enolate ions is considered to be the first stage of the condensation reaction. Aromatic ketones having a hydrogen if treated with alkaline solutions (in this case NaOH), the hydroxide ion from the base will attack hydrogen $\alpha$ from the ketone so that a carbanion is formed which can be stabilized by resonance and release the H$_2$O molecule. Followed by the second stage which is a nucleophilic addition reaction. Here, the enolate or carbanion ion formed at stage one acts as a nucleophile that attacks the carbonyl group of benzaldehyde. An alkoxide ion is formed which has an excess of electron charge in the O atom. Next, is the formation of an aldol. Aldol is a compound formed from aldehydes and ketones where aldol takes protons from solvent molecules, H$_2$O. Alkoxide ions take hydrogen protons from H$_2$O molecules to form $\beta$-hydroxyketone (aldol). Then the hydroxide ion from H$_2$O binds to the sodium ion and returns to form a NaOH base catalyst. Finally, the dehydration reaction of the aldol compound takes place in Claisen-Schmidt reaction. Dehydration reaction is a reaction of the release of water molecules. Carbonyl $\beta$-hydroxy like aldol is easily dehydrated, because the double bonds in the compound conjugate with the carbonyl group. The disappearance of the chemical shift at 2.5 ppm attributed to -CH$_3$ hydrogens of acetophenone, it is a good evidence of the resulted chalcone derivatives. Moreover, the increase of the melting points from 47 °C to 57 °C is another indicator of the successful synthesis.
Zn (II) complexes ($C_1$, $C_2$ and $C_3$) resulted after mixing ethanolic chalcone derivative solution ($L_1$, $L_2$ and $L_3$) with ZnCl$_2$ at (1:2) ratio, using reflux for 6 h. Coloured complexes obtained after filtration. The three complexes are soluble to DMSO and DMF. Elemental analyser indicated that the complexes have 1:2 metal to ligands stoichiometry of the types [ZnL$_2$(Cl)$_2$], whereas L are the chalcone derivatives resulted after the condensation of the correct acetophenone analogue with the correct benzaldehyde analogue. In addition, low molar conductance values indicate that the complexes are not electrolytes.

3.2 Theoretical Studies on Chalcone Derivatives

3.2.1 Molecular Geometry

The ground state optimization structures of the synthesized chalcone molecules were obtained in the aqueous phase at B3LYP def2-TZVP Grid5 level and are given in Fig. 2.

The geometrical parameters have been procured from optimized molecular structure and summarized in Table 1, giving the main theoretically calculated bond lengths and bond angles of the molecules.

Table 1 Selected bond lengths (Å) and bond angles (°) of the synthesized chalcones.

|   | Atoms       | Angle  | Atoms       | Length   |
|---|-------------|--------|-------------|----------|
| L1 | C5-C1-O1    | 119.279| H4-C8       | 1.08865  |
|   | C2-C1-C5    | 118.142| C8-C9       | 1.40054  |
|   | C1-C7-H1    | 118.563| H13-O2      | 0.972961 |
|   | C14-C15-N   | 122.475| C10-O2      | 1.36579  |
|   | H10-C16-H12 | 107.425| O4-N        | 1.23645  |
|   | C10-O2-H13  | 109.185| N-C15       | 1.47324  |
|   | O3-N-O4     | 123.738|             |          |
|   | C15-N-O4    | 119.113|             |          |

|   | Atoms       | Angle  | Atoms       | Length   |
|---|-------------|--------|-------------|----------|
| L2 | C2-C1-C6    | 120.008| H15-C17     | 1.11298  |
|   | C1-C2-H1    | 120.004| C17-N       | 1.47004  |
|   | C5-C4-O2    | 120.001| C12-C11     | 1.3949   |
|   | C12-C13-N   | 119.995| C7-O1       | 1.20804  |
|   | H10-C16-H12 | 109.462| O4-N        | 1.23645  |
|   | C4-O2-H13   | 108.000|             |          |
|   | C17-N-C18   | 119.996|             |          |
|   | N-C17-H15   | 109.445|             |          |

|   | Atoms       | Angle  | Atoms       | Length   |
|---|-------------|--------|-------------|----------|
| L3 | C2-C1-C6    | 119.998| H13-C16     | 1.11302  |
|   | C2-C1-O3    | 120.001| C16-O3      | 1.40206  |
|   | C3-C2-H1    | 120.033| C13-C14     | 1.39477  |
|   | C14-C15-N   | 119.959| O5-N        | 1.31001  |
|   | C4-O2-H10   | 107.999| C15-N       | 1.24809  |
|   | C1-O3-C16   | 110.801| H10-O2      | 0.971976 |
|   | H12-C16-H13 | 109.525|             |          |
|   | O3-C16-H11  | 109.497|             |          |
|   | C15-N-O4    | 120.004|             |          |
3.2.2 Spectroscopy

The spectroscopic UV-vis spectrum of the three ligand molecules, and their electronic transitions can be computerized and analyzed by time-depended density functional theory or TD-DFT. Additional information in the molecular structure prediction can be taken by electronic spectroscopy. In Fig. 3 we can see the calculated UV-vis spectrum of the molecules taken in the aqueous phase using B3LYP def2-TZVP Grid5 level algorithm.

As seen in Fig. 3, for \( L_1 \) and \( L_3 \) are observed similar spectrums giving two bands for \( \pi\pi^* \) and \( \eta\pi^* \) transitions respectively. Most absorption spectroscopy of organic compounds is based on transitions of \( n \) or \( \pi \) electrons to the \( \pi^* \) excited state. This is because the absorption peaks for these transitions fall in an experimentally convenient region of the spectrum (200 - 700 nm). These transitions need an unsaturated group in the molecule to provide the \( \pi \) electrons. For \( L_2 \) molecule only one transition is observed the \( \pi\pi^* \). The similarity of the bands shapes and close wavelengths show that the structures of the studied molecules are quite similar as they are belonging to the same family of the chalcones. \(^1\)HNMR spectrum calculated data also suggest the similarity of the structures.

For structural characterization \(^1\)HNMR spectra of chalcone molecules were calculated and peaks were attributed to the correct chemical groups. Chemical shifts were given according to the TMS reference and calculated from \( \delta=\Sigma_{\text{TMS}}-\Sigma_{\text{relation}} \), whereas \( \Sigma_{\text{TMS}} \) corresponds to the shielding of proton to the TMS and \( \Sigma \), the shielding of proton in the sample. The \(^1\)HNMR spectrum calculated at B3LYP DEF2-TZVPP DEF2 level in aqueous phase and are given in Fig. 4.

Chemical shifts of \(^1\)HNMR, are given in Table 2.

**Table 2.** \(^1\)HNMR chemical shifts of chalcone molecules calculated at aqueous phase.
| Group | Shift (ppm) | Group | Shift (ppm) | Group | Shift (ppm) |
|-------|-------------|-------|-------------|-------|-------------|
| OH    | 5.35        | OH    | 5.35        | OH    | 5.35        |
| CH    | 7.02        | CH    | 7.02        | CH    | 7.1         |
| CH    | 8.21        | CH    | 6.71        | CH    | 7.33        |
| CH    | 7.43        | CH    | 7.43        | CH    | 8.21        |
| CH    | 8           | CH    | 7.2         | CH    | 7.27        |
| CH    | 7.34        | CH    | 7.34        | CH    | 8           |
| CH    | 7.89        | CH    | 7.15        | CH    | 7.89        |
| CH    | 7.79        | CH    | 6.74        | CH    | 7.89        |
| CH<sub>3</sub> | 2.34 | CH<sub>3</sub> | 3.06 | CH<sub>3</sub> | 3.83 |
| H     | 8.02        | CH<sub>3</sub> | 3.06 | H     | 8.62        |
| H     | 7.66        | CH<sub>3</sub> | 2.34 | H     | 7.66        |
| H     |             | H     | 8.33        |       |             |
| H     |             | H     | 7.42        |       |             |

As can be seen in Fig. 4, the spectra are similar for molecules L<sub>1</sub>, L<sub>3</sub> same as the UV-vis spectra. The only difference is the shift of the first chemical shift from 3.83 to 2.34 ppm due to the presence of the extra methyl group on L<sub>3</sub>. The presence of the 3.06 ppm chemical shift that it is characteristic for the L<sub>2</sub> and corresponds to the -CH<sub>3</sub> group of the molecule, dominates the anomeric region. Because they are attached to carbon atoms with low s-character hybridization (sp<sup>3</sup>) the found in a low ppm chemical shift. The aromatic region of the spectra of the three chalcone derivatives are quite similar. As can be seen from Table 2, the -OH group is present at three molecules as well, at the same chemical shift 5.35 ppm. The hydrogens of the phenol group of the chalcones can be found at 8.02 and 7.66 ppm for L<sub>1</sub>, 8.33 and 7.42 ppm for L<sub>2</sub>, and 8.62 and 7.66 ppm for L<sub>3</sub>.

An additional way to verify the structures of the chalcone derivatives, is the vibrational spectrum. It has been calculated by BP86 DEF2-SVP FREQ algorithm. The high intensity peaks with the harmonic vibration frequencies are given in Fig. 5.

The characteristic high frequency peak at 3500 cm<sup>-1</sup>, of s(O-H) is common for the three analogues, while the peak at 1200 cm<sup>-1</sup> for L<sub>1</sub>, indicating the s(N-H) vibration. The methoxy group of L<sub>3</sub> is responsible for the t(H-O-C-C) vibration at 800cm<sup>-1</sup> which is absent for L<sub>1</sub> and L<sub>2</sub>. The 1700 cm<sup>-1</sup> peak is responsible for the s(N-C) of the molecules, while the 1500 cm<sup>-1</sup> peak is responsible for s(N-O) that is why it is absent from L<sub>2</sub>.

### 3.2.3 Molecular Orbital Studies

The molecular orbital studies revealed the energy gap between the highest molecular orbital (HOMO) and the lowest molecular orbital (LUMO). The value of the energy difference between HOMO and LUMO as well as the highest occupied molecular orbital (EHOMO) and lowest unoccupied molecular orbital (ELUMO) energies plays a very important role in stability and reactivity of molecules. The EHOMO
energies of molecules show the molecule's ability to give electrons. On the other hand, ELUMO characterizes the ability of the compound to accept electrons. Thus, the one is nucleophile and the other electrophile. Molecules with small energy gaps are considering to have a higher chemical reactivity and softer structures, while molecules having larger energy gaps are considering to be more stable and chemically harder. The computed values of the three chalcone derivatives can be found in Fig. 6.

It seems that $L_3$, has the highest reactivity ($\Delta E = 0.38 \text{ eV}$) followed by $L_1$ ($\Delta E = 2.24 \text{ eV}$) and $L_2$ ($\Delta E = 2.34 \text{ eV}$). In Table 3, we can see all the calculated energy features of the three molecules including, enthalpy, entropy and Gibbs energy. Any process in which the number of particles in the system increases consequently results in an increase in disorder. This is why we can observe an increase in the entropy of the molecules.

Table 3. Calculated energy values of the molecules.

| Molecule                      | L1       | L2       | L3       |
|-------------------------------|----------|----------|----------|
| Potential Energy (Kcal/mol)   | -2.015   | -1.871   | -2.175   |
| Kinetic Energy (Kcal/mol)     | 1.003    | 9.317    | 1.089    |
| Magnitude (a.u)               | 2.202    | 3.991    | 2.326    |
| Zero Point Energy (Kcal/mol)  | 157.90   | 215.90   | 237.07   |
| Total Thermal Energy (Kcal/mol)| -1.013   | -9.385   | -1.085   |
| Total Enthalpy (Kcal/mol)     | -1.013   | -9.385   | -1.085   |
| Final Entropy (Kcal/mol)      | 37.09    | 33.74    | 33.11    |
| Gibbs Free Energy (Kcal/mol)  | -1.013   | -9.386   | -1.085   |
| $E_{\text{Lumo}}$ (eV)        | -1.817   | 0.58     | -4.259   |
| $E_{\text{Homo}}$ (eV)        | -4.057   | -1.757   | -4.646   |
| $\Delta E$ (eV)               | 2.24     | 2.34     | 0.38     |

The individual charge on each atom on the molecule, it is another factor used to characterised molecular structures and it is presented by the Mulliken population study. The Mulliken atomic charges have been calculated by DFT method and presented in Table 4.

Table 4. Calculated Mulliken atomic structures of the synthesised structures.
The O20 atom has the highest negative charge for \( L_1 \), while for \( L_2 \) the highest negative charge belongs to C2 atom. For \( L_3 \) the highest negative charge is that of C2 atom as well. On the other hand, N18 and H33 atoms are having the highest electropositive charge for \( L_1 \), and C7 and H33 atoms the highest electropositive charge for \( L_2 \). Finally, for \( L_3 \), C3, N19 and H31 are the most electropositive atoms. The negative charges are due to the electron withdrawing groups that the atoms are attached with and the positive charges are because of the negative charges of the adjacent groups.

### 3.3 Biological Evaluation of Chalcone Derivatives and their Zinc Complexes

#### 3.3.1 Molecular Docking

The examination of the biological activity of the three ligand molecules \((L_1, L_2, L_3)\) and their corresponding complexes \((C_1, C_2, C_3)\), calculated theoretically by molecular docking studies. Using this technique, we can predict the best drug candidate in terms of protein inhibition, on a specific targeted protein. By molecular docking, we can predict binding energies, types of interactions and the amino acid profile residue of the protein that interacts with the drug molecule. In this study, we investigated the binding affinity of our studied molecules with HSA (human serum albumin). HSA, is the main transport protein in human organisms, were drugs bind and transported throughout the blood transportation. The
interaction types between the chalcone molecules and their zinc complexes are given in Table 5. The energy function can be dissected into the following terms:

$$E_{\text{tot}} = E_{\text{bind}} + E_{\text{pharma}} + E_{\text{ligpre}} \quad (1)$$

where $E_{\text{bind}}$ is the empirical binding energy used during the molecular docking; $E_{\text{pharma}}$ is the energy of binding-site pharmacophores; and $E_{\text{ligpre}}$ is a penalty value if the ligand unsatisfied the ligand preferences. $E_{\text{pharma}}$ and $E_{\text{ligpre}}$ were used to improve the number of true positives by discriminating active compounds from hundreds of thousands of non-active compounds.

**Table 5.** Interaction types of the chalcone molecules and their zinc complexes.

| Molecule | Energy (Kcal/mol) | Van der Waals (Kcal/mol) | Hydrogen Bonding (Kcal/mol) | Electrostatic Forces (Kcal/mol) |
|----------|-------------------|--------------------------|-----------------------------|--------------------------------|
| L1       | -78.63            | -65.08                   | -13.55                      | 0                              |
| L2       | -76.66            | -59.16                   | -17.50                      | 0                              |
| L3       | -84.53            | -65.31                   | -19.22                      | 0                              |
| C1       | -82.64            | -62.09                   | -20.55                      | 0                              |
| C2       | -118.05           | -102.97                  | -15.08                      | 0                              |
| C3       | -94.29            | -73.61                   | -20.67                      | 0                              |

It can be seen that $C_2$, exhibit the highest binding affinity on the transport protein, followed by $C_3$ and $L_3$. The highest binding affinity exhibited by $C_2$ is because of the $\text{CH}_3$-$N$-$\text{CH}_3$ group which seems to interact better with the protein with van der Waals forces. The lowest binding affinity exhibited by $L_2$, which means that in general, the complexed zinc (II) molecules are more active molecules with only exception the $C_1$. In Table 6, the amino acid residue of the protein that interact with the studied molecules can be seen.

**Table 6.** Interactions formed between the studied molecules with the amino acids of the transport protein.
| Molecule | Amino Acid Residue |
|----------|--------------------|
| L1       | Hydrophilic: GLN 33 (-2.5), THR 83 (-8.5) TYR 84 (-2.5)  
Hydrophobic: LEU 31 (-4.8), ARG 81 (-4.1) GLU 82 (-9.6) THR 83 (-5.4) |
| L2       | Hydrophilic: TYR 140 (-4.1) GLU 141 (-3.5) ARG 144 (-9.9)  
Hydrophobic: PRO 35 (-6.9) PHE 36 (-5) GLU 37 (-5) TYR 140 (-9.3) |
| L3       | Hydrophilic: ASP 38 (-2.5) HIS 39 (-5.6) ARG 81 (-3.4) THR 83 (-6)  
Hydrophobic: LEU 31 (-2.5) HIS 39 (-5.6) ARG 81 (-3.4) THR 83 (-6) LEU 31 (-4.3) GLN 33 (-6.1) PRO 35 (-6.4) ASP 38 (-4.4) VAL 77 (-4.5) ARG 81 (-7) TYR 84 (-4) |
| C1       | Hydrophilic: ASP 38 (-11) THR 83 (-6.7) ARG 144 (-2.8)  
Hydrophobic: GLN 33 (-9.3) ASP 38 (-5.8) ARG 81 (6.3) LAU 112 (-4.9) ARG 144 (-5.1) |
| C2       | Hydrophilic: GLN 33 (-3.2) PHE 36 (-3.1) TYR 140 (-5) ARG 144 (-3.9)  
Hydrophobic: GLN 33 (-7.4) PRO 35 (-9) PHE 36 (-6.1) GLU 37 (-7.7) LEU 112 (-4) TYR 140 (-13.6) |
| C3       | Hydrophilic: ASP 38 (-10.7) ARG 81 (-8.6)  
Hydrophobic: GLN 33 (-11.8) PRO 35 (-7.7) ARG 81 (-11.9) THR 83 (-12.8) |

The best drug candidate of the six studied molecules with molecular docking (C$_2$), interacts both with hydrophilic and hydrophobic interactions. More specifically, C$_2$ exhibits hydrogen bonds with GLN 33, PHE 36, TYR 140 and ARG 144 amino acids. Additionally, C$_2$, exhibits van der Waals interactions with GLN 33, PRO 35, PHE 36, GLU 37, LEU 112 and TYR 140 amino acids. Docking poses are depicted in Fig. 7.

### 3.3.2 BSA Binding

The structural changes of the protein and the complexation with the studied molecules has been done using UV-vis absorption measurements. The compounds interacted on the site I of the protein. We performed binding studies on the BSA protein because it has a similar shape with the HSA. The UV-vis absorption spectrums in Fig. 8 shows the effect of L$_1$, L$_2$, L$_3$, C$_1$, C$_2$ and C$_3$ molecules on the BSA spectrum. Strong absorption peaks at 250 nm and 350 nm can be seen for L$_1$, L$_2$, L$_3$ which are increased in intensities as the concentration of the molecules increases. The red shift observed at 250-255 nm indicates the complex formation of the protein with the ligands. When the L$_1$, L$_2$ and L$_3$ complexed with Zn (II), we observe in the spectrum of the protein only one strong absorption peak at 210-220 nm depending on the complex molecule. Again, the red shift of 1-5 nm corresponds to the ligation of the molecules in the protein structure. The Zn (II) ions are responsible for the loosening and unfolding of the protein backbone with an increase of the hydrophobicity of its environment, more drastically than the of L$_1$, L$_2$, L$_3$ molecules which Zn is absent. Thus, the drastic change in the spectrum was happened by the complexed C$_1$, C$_2$, C$_3$ molecules. Additionally, the results indicated that the interaction of the Zn (II) complexed chalcones with BSA molecule has caused some conformational changes in the microenvironment around chromophore of BSA, that is why we cannot observe any peak at the 300 nm.

### 4. Conclusions
In this work, the synthesis of three chalcone derivatives and their corresponding Zn (II) molecules was presented and their structures evaluated spectroscopically and theoretically. Their spectroscopic and theoretical evaluation indicated that the Zn-chalcone molecules exhibited higher binding activity than their corresponding chalcone ligands. The binding activity was predicted with molecular docking studies and confirmed by spectroscopic BSA interactions of $L_1$, $L_2$, $L_3$, $C_1$, $C_2$ and $C_3$. From the highest to lowest activity the molecules are $C_2 > C_3 > L_3 > C_1 > L_1 > L_2$. Chalcones are biological active molecules and their interactions with Zn metal ion increases their binding activity on transport proteins. Additionally, chalcones could be used as biological chelators to reduce the toxic zinc concentrations in biological systems.

**Declarations**

**CRediT authorship contribution statement**

**Manos C. Vlasiou:** Conceptualization, Methodology, Software, Experimentation, Writing - original draft, writing – review & editing, validation.

**Declaration of competing interest**

The author declares that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

**Acknowledgements**

The author wants to acknowledge the Pharmacy Programme of the University of Nicosia and Dr. Demetris Apostolides of the Department of Chemistry, of the University of Cyprus for the NMR experiments.

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**Figures**

Figure 4
Calculated 1HNMR spectra of L1, L2 and L3 molecules.

**LUMO PLOT**

- **L1**: $E_{\text{LUMO}} = -1.817 \text{ ev}$
- **L2**: $E_{\text{LUMO}} = 0.58 \text{ ev}$
- **L3**: $E_{\text{LUMO}} = -4.259 \text{ ev}$

**HOMO PLOT**

- **L1**: $E_{\text{HOMO}} = -4.057 \text{ ev}$
- **L2**: $E_{\text{HOMO}} = -1.757 \text{ ev}$
- **L3**: $E_{\text{HOMO}} = -4.646 \text{ ev}$

$\Delta E_1 = 2.24 \text{ ev}$
$\Delta E_2 = 2.34 \text{ ev}$
$\Delta E_3 = 0.38 \text{ ev}$

Figure 6

HOMO-LUMO orbitals of L1, L2 and L3 molecules.