Nitrous oxide, methane emissions and grain yield in rainfed wheat grown under nitrogen enriched biochar and straw in a semiarid environment

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ABSTRACT

Background. Soil application of biochar and straw alone or their combinations with nitrogen (N) fertilizer are becoming increasingly common, but little is known about their agronomic and environmental performance in semiarid environments. This study was conducted to investigate the effect(s) of these amendments on soil properties, nitrous oxide (N₂O) and methane (CH₄) emissions and grain and biomass yield of spring wheat (Triticum aestivum L.), and to produce background dataset that may be used to inform nutrient management guidelines for semiarid environments.

Methods. The experiment involved the application of biochar, straw or urea (46% nitrogen [N]) alone or their combinations. The treatments were: CN₀ –control (zero-amendment), CN5₀ –50 kg ha⁻¹ N, CN1₀₀ –100 kg ha⁻¹ N, BN₀ –15 t ha⁻¹ biochar, BN5₀ –15 t ha⁻¹ biochar + 50 kg ha⁻¹ N, BN1₀₀ –15 t ha⁻¹ biochar + 100 kg ha⁻¹ N, SN₀ –4.5 t ha⁻¹ straw, SN5₀ –4.5 t ha⁻¹ straw + 50 kg ha⁻¹ N and SN1₀₀ –4.5 t ha⁻¹ straw + 100 kg ha⁻¹ N. Fluxes of N₂O, CH₄ and grain yield were monitored over three consecutive cropping seasons between 2014 and 2016 using the static chamber-gas chromatography method.

Results. On average, BN1₀₀ reported the highest grain yield (2054 kg ha⁻¹), which was between 25.04% and 38.34% higher than all other treatments. In addition, biomass yield was much higher under biochar treated plots relative to the other treatments. These findings are supported by the increased in soil organic C by 17.14% and 21.65% in biochar amended soils (at 0–10 cm) compared to straw treated soils and soils without carbon respectively. The BN₁₀₀ treatment also improved bulk density and hydraulic properties (P < 0.05), which supported the above results. The greatest N₂O emissions and CH₄ sink were recorded under the highest rate of N fertilization (100 kg N ha⁻¹). Cumulative N₂O emissions were 39.02% and 48.23% lower in BN₁₀₀ compared with CN₀ and CN₁₀₀ respectively. There was also a ≈37.53% reduction in CH₄ uptake under BN₁₀₀ compared with CN₀–control and CN₅₀. The mean cumulative N₂O emission from biochar treated soils had a significant decrease of 10.93% and 38.61% compared to straw treated soils and soils without carbon treatment, respectively. However, differences between mean cumulative N₂O emission between straw treated soils and
soils without carbon were not significant. These results indicate the dependency of crop yield, N₂O and CH₄ emissions on soil quality and imply that crop productivity could be increased without compromising on environmental quality when biochar is applied in combination with N-fertilizer. The practice of applying biochar with N fertilizer at 100 kg ha⁻¹ N resulted in increases in crop productivity and reduced N₂O and CH₄ soil emissions under dryland cropping systems.

**Subjects** Agricultural Science, Ecosystem Science, Soil Science, Biogeochemistry

**Keywords** Semi-arid, Greenhouse gas, Rain-fed, Loess plateau, Grain yield, Carbon amendments, Fertilization

**INTRODUCTION**

Atmospheric methane (CH₄) and nitrous oxide (N₂O) are persistent greenhouse gases (GHG) influencing global warming (IPCC, 2014). Agriculture contributes significant amounts of N₂O and CH₄ to the atmosphere, however net GHG emissions as CO₂ from farming-related activities can be potentially reduced by increasing carbon (C) sequestration in soil and crop biomass (Wang et al., 2021). This may be achieved by implementing improved crop and fertilizer management practices that maximize biomass production and C returned to soil (Norton, 2014). There are no significant terrestrial sinks of N₂O hence reduction in its emission may only be achieved by managing nitrogen (N) inputs, and improving soil conditions and efficiency of applied fertilizer-N (Grace, 2016). However, in semi-arid regions of China, in an attempt to increase yields, farmers are compelled to apply more fertilizer, leading to an over-application (Xu & Yang, 2017). There is heavy dependence on mineral fertilizers to ensure adequate N supply for crops, and in most cases more fertilizer is applied than needed by the plant (Liu et al., 2016). This is a common practice in most farming communities in semi-arid regions of China (Wang et al., 2021). The situation has led to negative impact on the environment, and threatens the long-term sustainability of Chinese agriculture (Liu et al., 2016; Wang et al., 2021). Therefore, it is key to identify suitable agricultural practices that could help maximize crop production without compromising on environmental quality.

Current increases in atmospheric GHG levels require that novel approaches are undertaken to mitigate impacts of climate change, such as management practices capable of improving soil C sequestration (Woolf et al., 2010). Soil carbon sequestration through application of recalcitrant C-rich biochar is mentioned as a suitable means to mitigate climate change, and improve soil fertility (Laird et al., 2010) and crop productivity (Steiner et al., 2007). According to Saggar (2010) N₂O emissions are driven by the applications of fertilizer nitrogen (N), soil tillage and crop type, with their effects dependent on soil and weather conditions. Biochar application as a soil amendment, could therefore be an effective strategy for mitigating emissions and increasing crop yield. However, the effect of biochar on soil properties, GHG emissions and crop yield have been diverse. Several mechanisms have also been proposed in literature to explain the diverse effects, with limited amounts of evidence to support them. Yanai, Toyota & Okazaki (2007) reported
decreased N₂O and CH₄ soil emissions in response to biochar application. In contrast, Clough et al. (2013) observed no suppression of N₂O and CH₄ soil emissions, whilst similar effect was observed by Zhang et al. (2010). Zhang et al. (2011) also reported that biochar application in dryland significantly reduces soil CH₄ emission by 33% compared to soil without biochar. Zimmerman, Gao & Ahn (2011) attributed the positive effect of biochar application on soil CH₄ emissions to the inhibition of soil methanotrophs while Zhu et al. (2018) associated reduced soil CH₄ emissions to the change in the ratio of methanogenic to methanotrophic archaea. In general, most studies have found biochar amendments to either decrease or not significantly affect soil N₂O emissions; however, some few reports have found increased N₂O emissions following biochar amendments (Yeboah et al., 2018). Explanations for continued long-term suppression of N₂O emissions in biochar-amended soils include alterations in microbial communities due to physical habitat changes, physical and/or chemical protection of organic C and/or N by biochar and alteration of micro-scale soil redox status due to electrochemical properties of biochars (Rivka, David & Timothy, 2019). It is thus clear that, these effects have been shown to vary significantly depending upon the type of biochar used and the environmental and soil conditions under which the material is applied.

The Loess Plateau is an important agricultural area in China and is widely used for grain production (He et al., 2014). The area is one of the most severely eroded regions in China, which coupled with limited precipitation and high evaporation rates, often results in poor crop productivity (He et al., 2014). Many studies have indicated that human activities, such as land use is responsible for the degradation and loss of soil fertility in semi-arid regions of China (Xu & Yang, 2017; Zhang et al., 2017; Huang et al., 2019). Traditional methods of soil cultivation often accelerates the decline of soil fertility, and loss of soil organic C (Lamptey, Li & Xie, 2018). Given the fact that the population of semi-arid regions in China mainly relies on rainfed agriculture for their livelihood; developing environmentally friendly and sustainable nutrient management strategies is crucial. There is limited information on the specific impact of widely-used agronomic practices involving biochar, straw and nitrogen fertilizer used alone or combined on greenhouse gas emission and crop yield in drier lossiah soils (Solomon et al., 2007). Moreover, little is known about the effect of biochar application to soil under arid conditions (Arfaoui, Ibrahimi & Trabelsi, 2019). This study hypothesized that increased C inputs would raise the soils potential to reduce N₂O and CH₄ soil emissions whilst increasing grain yield. Therefore, the objectives of this study were to: 1) determine the effect of biochar, straw and nitrogen fertilizer applied alone or combined with fertilizer-N on soil properties, (2) assess the effect of biochar, straw and nitrogen fertilizer applied alone or combined with fertilizer-N on biomass and grain yield of spring wheat, and (3) determine the effects of biochar, straw and nitrogen fertilizer used alone or combined with fertilizer-N on N₂O and CH₄ emissions.

**MATERIALS & METHODS**

**Study site**

The study was conducted during the 2014, 2015 and 2016 growing seasons at the Dingxi Experimental Station (35°28’N, 104°44’E, elevation 1971-m above-sea-level) of the Gansu
Agricultural University in Northwestern China. The research station is located in the semiarid Western Loess Plateau, which is characterized by step hills and deeply eroded gullies (Feng et al., 2013). This area has Aeolian soils, locally known as Huangmian (Chinese Soil Taxonomy Cooperative Research Group, 1995), which equate to Calcaric Cambisols based on the FAO (1990) description. The soil type in the study area is sandy-loam with low fertility. The soil has a pH of ≈8.3, soil organic carbon (SOC) ≤8.13 g kg⁻¹, and Olsen-P ≤13 mg kg⁻¹ as described in Yeboah et al. (2018). The type of soil in the study area is the principal soil for cultivation of crops in the agro-ecological zone. Long term average rainfall, evaporation and aridity in the study area is 391.9 mm per annum; 1531 mm per annum and 2.53 respectively. The aridity index (AI) is the degree of dryness of the climate at the study area. In July, the daily maximum temperature can increase to 38 °C. Similarly, in January daily minimum temperature can drop to −22 °C. Annual cumulative temperatures >10 °C are 2240 °C and annual radiation is 5930 MJ m⁻², with 2477 h of sunshine as described in Yeboah et al. (2018). The agro-climatic conditions are similar to semiarid environments. The research site is characterized by continuous cultivation of the same field using conventional tillage practices. The preceding crop cultivated at the research site was potatoes (Solanum tuberosum L.). Seasonal rainfall recorded in 2014, 2015 and 2016 during the research was 174.6, 252.5 and 239.4 mm respectively (Fig. 1).

**Experimental design and description of treatment**

The experiment involved addition of different carbon (C) sources; namely: biochar and straw, and N fertilizer in the form of urea (46% N) arranged in a randomized block design with nine treatments and three replications (Yeboah et al., 2018). The treatments were: CN₀ – control (zero-amendment), CN₅₀ –50 kg ha⁻¹ N applied each year, CN₁₀₀ –100 kg ha⁻¹ N applied each year, BN₀ –15 t ha⁻¹ biochar applied in a single dressing in 2014, BN₅₀ –15 t ha⁻¹ biochar applied in a single dressing in 2014 + 50 kg ha⁻¹ N applied each year, BN₁₀₀ –15 t ha⁻¹ biochar applied in single dressing in 2014 + 100 kg ha⁻¹ N applied each year, SN₀ –4.5 t ha⁻¹ straw applied each year, SN₅₀ –4.5 t ha⁻¹ straw applied each year + 50 kg ha⁻¹ N applied each year and SN₁₀₀ –4.5 t ha⁻¹ straw applied each year + 100 kg ha⁻¹ N applied each year as described in Yeboah et al. (2018). The two C sources (biochar and straw) were applied at the same quantity based on the straw returned to the soil every year and straw C mineralization. Biochar was spread evenly on the soil surface in March 2014 and incorporated into the soil using a rotary tillage implement to a depth of ≈10 cm. The biochar was obtained from Golden Future Agriculture Technology Company Limited, Liaoning in China. Biochar was produced from maize straw using pyrolysis process at a temperature of 350–550 °C. This process converted about 35% of the maize straw to biochar. The biochar in the form of granules was milled to a size of <5 mm to allow for even mixing with the soil. The wheat crop of the previous season from the research station was used as a source of straw for the study. In the plots that received straw treatment, the straw from the previous wheat crop was weighed and returned to the original plots. This was done after threshing. Biochar analysis was conducted using the procedure as describe in Lu (2000). Total C and N and soil pH were determined using a CN Analyzer (analytikjena; multi N/C, 2100S, Germany) and Kjeldahl digestion and distillation (Bremner & Mulvaney,
Figure 1  Precipitation (mm) in (A) 2014, (B) 2015 and (C) 2016 cropping season at the experimental site.

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Table 1 Characterization of biochar and straw used in the study.

| Parameter | pH  | BD (g cm\(^{-3}\)) | SA (m\(^2\) g\(^{-1}\)) | Ca  | Mg  | K   | C   | N   | P   | Ash content (%) |
|-----------|-----|--------------------|-----------------|-----|-----|-----|-----|-----|-----|-----------------|
| Biochar   | 9.2 | 0.68               | 8.75            | 0.8 | 0.47| 0.51| 53.28| 1.04| 0.26| 25.5            |
| Straw     | 6.5 | /                  | /               | 0.53| 0.04| 0.47| 45.05| 0.94| 0.08| 8.9             |

Notes.
Values are means for \(n=2\).

1982) and pH meter (model: Sartorius PB–10, Germany). The soil pH was determined using soil to water ratio of 1:2.5. Similar protocol was used to determined total C and N, ash content and pH of the straw. Table 1 shows the chemical characterization of biochar and straw used in the experiment. All the treatments received a blanket application of Phosphorus (P) fertilizer which was applied equally at a rate of 46 kg ha\(^{-1}\) P in the form of ammonium dihydrogen phosphate (12% N, 52% P\(_2\)O\(_5\)). No–tillage seeder was used to incorporate the fertilizer to about 20 cm soil depth at planting. Based on the protocol described in Yeboah et al. (2016) spring wheat (Triticum aestivum L. cv. Dingxi 35) was sown in Mid-March at a rate of 188 kg ha\(^{-1}\) seeds at 20-cm row spacing. The crop was harvested either at the end of July or early August. The individual plot’s measured 3 m by 6 m and the plots were separated by 0.5 m width protection rows.

Soil sampling, measurements and analyses
Based on the protocol described in Yeboah et al. (2016), soil bulk density (BD) was determined by taking small cores and relating the oven–dried mass of soil to the volume of the core. Soil saturated hydraulic conductivity (Ksat) was determined at two points per plot using the disc permeameter method according to Carter (1993). Soil samples were collected from 0–10 and 10–30 cm depth and bulked for analysis. The samples were processed for analysis using the protocol described in Yeboah et al. (2018). Soil organic carbon (SOC) in the fine ground samples was determined by the modified Walkley & Black (1934) wet oxidation method (Nelson & Sommers, 1982).

Gas sampling and analysis
Collection of N\(_2\)O and CH\(_4\) gases were performed using the static chamber technique based on the procedure described by Zou et al. (2005). For each sampling event, gas collection was consistently performed between 08:00–12:00 h, based on the guidelines of Yeboah et al. (2016). Collection of samples for N\(_2\)O and CH\(_4\) analyses was conducted at 0, 10, and 20 min after chamber closure. Samples were collected between March and September and detailed sampling procedure could be found in Yeboah et al. (2016); Yeboah et al. (2018). Based on earlier studies conducted in low rainfall areas (e.g., Wang et al., 2010) emissions occurring during the dry season were expected to be low and therefore did not justify measurements over that period. Gas fluxes were measured over 14 sampling events per year. Whilst acknowledging that accurate estimates of total emissions cannot be determined from relatively few sampling events, the main purpose of this work was to quantify relative differences between-treatments, which therefore justifies the approach used in this study. A similar approach was also employed by Tullberg et al. (2018) to quantify soil emissions.
of GHG from tillage and traffic treatments in conservation agriculture areas with seasonal rainfall. The N\textsubscript{2}O and CH\textsubscript{4} concentration in samples were analyzed within 2 to 3 days after collection using gas chromatograph (GC). The GC system (Agilent 7890A, USA) equipped with flame ionization detector (FID) was used for CH\textsubscript{4} analysis and an electron capture detector (ECD) was used for N\textsubscript{2}O analysis. Rates of CH\textsubscript{4} and N\textsubscript{2}O fluxes were calculated by linear increment of the gas concentration at 0, 10 and 20 min. The calculation was only accepted when the R\textsuperscript{2} of the linear correlation was higher than 0.90 (\(p < 0.05\)). The average GHG fluxes were a mean of three replicates of each treatment over the sampling dates. Further procedure for the analysis and conditions of the column could be found in Yeboah et al. (2018) and Zou et al. (2005).

Estimations of nitrous oxide and methane emissions

The N\textsubscript{2}O (mg m\textsuperscript{-2} h\textsuperscript{-1}) and CH\textsubscript{4} (mg m\textsuperscript{-2} h\textsuperscript{-1}) emissions were calculated using Eq. (1) based on the protocol described in Yeboah et al. (2016):

\[
F = \frac{C_2 \times V \times M_0 \times 273/T_2 - C_1 \times V \times M_0 \times 273/T_1}{A \times (t_2 - t_1) \times 22.4}
\]

(1)

where: F are fluxes of N\textsubscript{2}O or CH\textsubscript{4} (mg m\textsuperscript{-2} h\textsuperscript{-1}), V is volume (m\textsuperscript{3}), M\textsubscript{0} is the molecular weight of the gas, C\textsubscript{1} and C\textsubscript{2} are the concentration of previous (0 mins) and current (20 mins) gas concentrations inside the chamber (mol mol\textsuperscript{-1}), T\textsubscript{1} and T\textsubscript{2} are temperature (Kelvin) recorded inside the chamber during current and previous samplings, and t\textsubscript{1} and t\textsubscript{2} are previous and current sampling times (h).

The cumulative emission of N\textsubscript{2}O and CH\textsubscript{4} in kg ha\textsuperscript{-1} was estimated using the equation as follows (Yeboah et al., 2016):

\[
M = \sum_{i=1}^{N} (F_{N+1} + F_N) \times 0.5 \times (t_{N+1} - t_N) \times 24 \times 10^{-2}
\]

(2)

where M is the N\textsubscript{2}O and CH\textsubscript{4} cumulative emissions during the period of measurement (kg ha\textsuperscript{-1}), F is N\textsubscript{2}O and CH\textsubscript{4} emission (in mg m\textsuperscript{-2} h\textsuperscript{-1}); and previous and current sampling emissions were N+1 and N respectively. The number of days from first sampling is represented by t.

Biomass and grain yield

Biomass and grain yield was determined by cutting the plants using hand sickles to five cm height aboveground. The outer edges of about 0.5 m was discarded from each plot. Both yields were determined on a dry-weight basis by oven–drying the plant material at 105 °C for 45 min and then to constant weight at 85 °C (Yeboah et al., 2016).

Statistical analyses

Statistical analyses were undertaken with the SPSS 22 (IBM Corporation, Chicago, IL, USA) with the treatment as the fixed effect and year as random effect. Tukey’s honestly significant was used to determine the differences between-treatments means. Significance differences were declared at probability level of 5%.
Table 2 Analysis of variance for carbon, nitrogen and year effects and their interaction.

| Sources | Soil bulk density | Ksat | Soil organic carbon | N₂O | CH₄ | Biomass yield | Grain yield |
|---------|------------------|------|---------------------|-----|-----|--------------|-------------|
|         | 0–5  | 5–10 |         | 0–5  | 5–10 | 10–30 |       |      |
| Carbon (C) |       |       |       |       |       | n.s.  |       |      |
| Nitrogen (N) |       | n.s.  |       |       |       | n.s.  |       |      |
| Year (Y)   | n.s.  |       |       | n.s.  |       | n.s.  |       |      |
| C × N      | n.s.  | n.s.  | n.s.  | n.s.  | n.s.  | n.s.  |       | n.s |
| C × Y      | n.s.  | n.s.  | n.s.  | n.s.  | n.s.  | n.s.  |       | n.s |
| N × Y      | n.s.  | n.s.  | n.s.  | n.s.  | n.s.  | n.s.  | n.s.  | n.s |

Notes.

* and ** indicate significant difference at *P* < 0.05 and *P* < 0.01, respectively. n.s. indicate no significance difference at *P* < 0.05.

RESULTS

Soil bulk density, saturated hydraulic conductivity and soil organic carbon

Soil samples taken during the study period showed significant differences in the bulk density depending on the type of treatment and the depth of sampling (Table 2). Bulk density increased with soil depth in many cases irrespective of treatment over the experimental period. Significant differences between treatments were minor in the upper layer in 2014, but significant treatment effect was recorded in the 5–10 cm soil depth as the experimental period progressed from 2014 to 2016 (Table 3). On average, the lowest bulk density (1.14 g cm⁻³) was recorded under biochar–amended soils, and the highest was observed under soils with carbon (1.21 g cm⁻³). The results obtained with the straw–amended soils showed a similar trend, except that differences were not significant at *P* < 0.05 in most cases.

Saturated hydraulic conductivity (Ksat) was significantly (*P* < 0.05) affected by carbon, N fertilizer and year but there was no significant interaction between treatment factors (Table 2). Application of BN₁₀₀ treatment enhanced mean saturated hydraulic conductivity by 23.7%, 24.3% and 20.4% relative to CN₀, CN₅₀ and SN₀, respectively (Table 4). Carbon and year had significant interaction (*P* < 0.05) on soil organic carbon, except at the 10–30 cm soil depth (Table 2). Similarly, carbon and fertilizer-N also interactively affected soil organic C in all the soil depth evaluated. Application of fertilizer-N at the 50 and 100 kg ha⁻¹ rate influenced SOC significantly (*P* < 0.05) under biochar treated soils, particularly in the depth of 0–5 cm (Table 5). However, N₁₀₀ had greater effect compared to N₅₀.

Nitrous oxide emissions

All the treatments were sources of nitrous oxide (N₂O) emission throughout the sampling period and the maximum observed N₂O emissions occurred in early July in each year of this study (Fig. 2). These responses were consistent with recorded soil moisture and temperature data. Significant differences (*P* < 0.05) were found among treatments at certain periods of measurement (Fig. 3). For example, in 2014, the maximal N₂O emission of BN₁₀₀ was 79.5 µg m⁻² h⁻¹ and the minimal was 36.5 µg m⁻² h⁻¹; they were significantly lower than those for CN₅₀ (100.7 µg m⁻² h⁻¹ for maximum and 55.8 µg m⁻² h⁻¹ for minimum) and CN₀ (98.1 µg m⁻² h⁻¹ for maximum and 50.2 µg m⁻² h⁻¹ for minimum). At a lesser
Table 3  Soil bulk density as affected by carbon addition sources.

| Treatment | C source | Mineral N | 0–5  | 5–10  | 10–30  | Mean | Mean |
|-----------|----------|-----------|------|-------|--------|------|------|
|           |          |           | 2014 | 2015  | 2016   |      |      |
| No carbon | N0       | 1.24a     | 1.27a | 1.29a  | 1.27a  | 1.29a|
|           | N50      | 1.17bc    | 1.25abc | 1.24ab| 1.25abc | 1.27a|
|           | N100     | 1.20ab    | 1.17cd | 1.12c  | 1.16bc | 1.27a|
| Biochar   | N0       | 1.17bc    | 1.25abc | 1.24ab| 1.25abc | 1.27a|
|           | N50      | 1.13cd    | 1.16cd | 1.14c  | 1.18bc | 1.24a|
|           | N100     | 1.15bcd   | 1.18bcd | 1.17bc| 1.19bc  | 1.27a|
| Straw     | N0       | 1.11d     | 1.08d  | 1.14c  | 1.16c  | 1.21a|
|           | N50      | 1.14cd    | 1.18cd | 1.16bc | 1.21abc | 1.25a|
|           | N100     | 1.17a     | 1.22a  | 1.23ab | 1.25ab | 1.29a|

Notes.
Values with different letters within a column are significantly different at \( P < 0.05 \).

Table 4  Saturated hydraulic conductivity as affected by carbon addition sources.

| Treatment | C source | Mineral N | 2014  | 2015  | 2016  | mean |
|-----------|----------|-----------|-------|-------|-------|------|
|           |          |           | 2014  | 2015  | 2016  |      |
| No carbon | N0       | 62.64b    | 68.86b | 62.95c | 64.82c |      |
|           | N50      | 65.77ab   | 64.06b | 63.71c | 64.51c | 67.89ab |
|           | N100     | 67.63ab   | 60.58b | 75.45abc | 67.63ab |      |
| Biochar   | N0       | 71.93ab   | 62.88b | 68.07bc | 67.63ab |      |
|           | N50      | 80.49a    | 70.94ab | 78.98ab | 76.80ab | 80.19a |
|           | N100     | 78.78ab   | 78.99a | 82.79a  | 80.19a  |      |
| Straw     | N0       | 68.66ab   | 65.65b | 65.42c  | 66.58bc | 66.82bc |
|           | N50      | 76.07ab   | 66.02b | 74.75abc | 72.28ab |      |
|           | N100     | 75.65ab   | 72.44ab | 71.24abc | 73.11ab |      |

Notes.
Values with different letters within a column are significantly different at \( P < 0.05 \). \( n = 3 \).

magnitude, SN0 and SN50 also produced significantly lower \( \text{N}_2\text{O} \) emission compared to CN0 and CN50. During this period the lowest seasonal \( \text{N}_2\text{O} \) emission was mostly recorded in the biochar treated soils and at a lesser magnitude in the straw treated soils.

There were no significant treatment interactions \((p < 0.05)\) effect on cumulative \( \text{N}_2\text{O} \) emission (Table 6), but treatment factors independently influenced cumulative \( \text{N}_2\text{O} \) emission. The highest cumulative \( \text{N}_2\text{O} \) emissions were consistently observed in the fertilized soils compared to the unfertilized soils, but differences were not always significant (Table 6). Application of BN0, BN50 and BN100 significantly decreased cumulative \( \text{N}_2\text{O} \) emission by 48.42%, 37.12% and 35.80% on average compared to CN100, respectively (Table 4). The mean cumulative \( \text{N}_2\text{O} \) emission of biochar was averaged at 1.83 kg ha\(^{-1}\) representing significant decrease of 10.93% and 38.61% compared to straw treated soils (2.03 kg ha\(^{-1}\))
### Table 5  Soil organic carbon as affected by different treatments.

| Treatment | C source | N rate | 0–10  | 10–30  | 0–10  | 10–30  | 0–10  | 10–30  | 0–10  | 10–30  | 0–10  | 10–30  |
|-----------|----------|--------|-------|--------|-------|--------|-------|--------|-------|--------|-------|--------|
|           |          |        | 2014  | 2015  | 2016  | Mean  | 2014  | 2015  | 2016  | Mean  | 2014  | 2015  | 2016  | Mean  |
| No carbon | N₀       |        | 9.64c | 9.86c | 10.43e| 9.98d | 9.29b | 9.58b | 9.48e | 9.45c |
|           | N₁₀      |        | 10.18bc| 9.92bc| 11.54d| 10.55cd| 10.34ab| 9.73b | 10.71d| 10.26bc|
|           | N₁₀₀     |        | 10.32bc| 10.90bc| 11.70d| 10.97bcd| 9.76ab| 10.10b| 11.05cd| 10.30bc|
| Biochar   | N₀       |        | 11.82ab| 10.28bc| 14.91b| 12.34b| 10.45ab| 10.27b| 12.47b| 11.06b |
|           | N₁₀      |        | 12.21ab| 14.04a | 16.01a| 14.09a| 11.59a | 12.66a| 14.41a| 12.89a |
|           | N₁₀₀     |        | 12.42a | 14.09a | 16.26a| 14.26a| 11.04ab| 13.75a| 15.41a| 13.40a |
| Straw     | N₀       |        | 9.71c | 10.14bc| 11.41d| 10.42cd| 9.58ab| 10.12b| 10.69d| 10.13bc|
|           | N₁₀      |        | 10.70abc| 10.64bc| 13.77c| 11.70bc| 10.70ab| 10.41b| 11.54bcd| 10.88b |
|           | N₁₀₀     |        | 11.08abc| 11.19b | 14.19bc| 12.15b| 10.92ab| 10.99b| 12.10bc| 11.34b |

**Notes.**

Values with different letters within a column are significantly different at *P* < 0.05. *n* = 3.

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**Figure 2** Seasonal N$_2$O fluxes for spring wheat in 2014 (A), 2015 (B) and 2016 (C) as affected by carbon addition sources. The vertical bars represent the least significant difference (LSD) at *P* < 0.05 among treatments within a measurement date.

Full-size [DOI: 10.7717/peerj.11937/fig-2](https://doi.org/10.7717/peerj.11937/fig-2)

**Figure 3** Seasonal CH$_4$ fluxes for spring wheat in 2014 (A), 2015 (B) and 2016 (C) as affected by carbon addition sources. The vertical bars represent the least significant difference (LSD) at *P* < 0.05 among treatments within a measurement date.

Full-size [DOI: 10.7717/peerj.11937/fig-3](https://doi.org/10.7717/peerj.11937/fig-3)
Table 6  Cumulative N₂O emissions of spring wheat as affected by carbon addition sources.

| Treatment | N rates | 2014     | 2015     | 2016     | Mean     |
|-----------|---------|----------|----------|----------|----------|
| No carbon | N₀      | 3.10a    | 2.09ab   | 2.00ab   | 2.40ab   |
|           | N₅₀     | 3.17a    | 2.00ab   | 1.75bc   | 2.31abc  |
|           | N₁₀₀    | 3.21a    | 2.37a    | 2.11a    | 2.56a    |
| Biochar   | N₀      | 2.37b    | 1.50c    | 1.32c    | 1.73c    |
|           | N₅₀     | 2.47b    | 1.73bc   | 1.41c    | 1.87bc   |
|           | N₁₀₀    | 2.48b    | 1.77bc   | 1.42bc   | 1.89bc   |
| Straw     | N₀      | 2.43b    | 1.87bc   | 1.24c    | 1.85bc   |
|           | N₅₀     | 2.83ab   | 1.78bc   | 1.54abc  | 2.05abc  |
|           | N₁₀₀    | 2.99a    | 1.98ab   | 1.61abc  | 2.19abc  |

Notes.
Values with different letters within a column are significantly different at p < 0.05.

and soils without carbon treatment (2.42 kg ha⁻¹). Straw treated soils had non–significant cumulative N₂O decrease of 0.39 kg ha⁻¹, or 19.40% less compared to no carbon soils.

Methane emissions

All the treatments had similar trends of seasonal CH₄ dynamics and were net carbon sinks over the three study years (Fig. 3). The minimum CH₄ consumption was recorded in April 2014 and 2015, and in September 2016. In the present study, a single peak was observed in June 2014, whiles double peaks were observed in May and July 2015 and 2016. During this period, the greatest seasonal CH₄ consumption of −79.94, −81.07 and −111.59 µg m⁻² h⁻¹ in 2014, 2015 and 2016 respectively were observed in BN₁₀₀ soils; it was 38.14%, 47.37%, 43.05% more compared to CN₀ (−57.87, −55.01 and −78.01 µg m⁻² h⁻¹). At a lesser extent, the maximum seasonal CH₄ consumption in SN₅₀ and SN₁₀₀ soils were significantly higher (p < 0.05) compared to the CN₀ and CN₅₀ soils. The results were clear that, the greater seasonal CH₄ consumption occurred with the higher N fertilizer soils and the greatest CH₄ uptake generally occurred in the biochar treated soils, followed by the straw treated soils and the least were observed in the no carbon soils.

Year individually had a significant effect (p < 0.05) on cumulative CH₄ emission (Table 7), and interaction between carbon and year significantly affected cumulative CH₄ emission. The results of cumulative CH₄ emission showed that increasing N fertilizer rates generally enhanced CH₄ consumption in all treatments. The use of BN₁₀₀ boosted cumulative CH₄ uptake in 2014 (by 21.9% and 18.2%), 2015 (by 83.6% and 59.1%) and 2016 (by 30.5% and 18.4%) compared to CN₀ and CN₅₀, respectively. Increasing the fertilizer rate from N₅₀ to N₁₀₀ resulted in significantly higher cumulative CH₄ consumption (p < 0.05) on straw treated soils in 2014 relative to N₀ on soils without carbon; the increase was 16.8%. In 2015, application of SN₁₀₀ increased cumulative CH₄ sink by 41.0%, 73.0%, 22.8% and 26.8% compared with CN₀, CN₅₀ and CN₁₀₀, respectively. The mean cumulative CH₄ consumption was greatest in biochar treated plots (−2.8 kg ha⁻¹), followed by straw treated soils (−2.6 kg ha⁻¹) and the least in no carbon soils (−2.3 kg ha⁻¹).
Table 7  Cumulative CH$_4$ emissions of spring wheat as affected by different treatment.

| Treatment | C source | N rates | 2014 | 2015 | 2016 | Mean |
|-----------|----------|---------|------|------|------|------|
| No carbon | $N_0$    |         | –1.80a | –1.79a | –2.83a | –2.14a |
|           | $N_{50}$ |         | –1.85ab | –2.07a | –3.12ab | –2.35ab |
|           | $N_{100}$ |         | –2.08bc | –2.00a | –3.13abc | –2.40abc |
| Biochar   | $N_0$    |         | –2.09bc | –3.14c | –3.31abc | –2.85bc |
|           | $N_{50}$ |         | –2.13bc | –2.23ab | –3.38abc | –2.58abc |
|           | $N_{100}$ |       | –2.19c | –3.29c | –3.70c | –3.06c |
| Straw     | $N_0$    |         | –1.91abc | –2.16ab | –3.19abc | –2.42abc |
|           | $N_{50}$ |         | –1.96abc | –2.21ab | –3.32abc | –2.50abc |
|           | $N_{100}$ |         | –2.10bc | –2.54b | –3.61bc | –2.75abc |

Notes. Values with different letters within a column are significantly different at $P < 0.05$.

### Biomass and grain yield

There was significant interaction effects between carbon and nitrogen, and nitrogen and year on biomass yield at $p < 0.05$ (Table 2). In addition, carbon, nitrogen and year individually had significant effect on biomass yield. Application of $N_{100}$ treatments on biochar treated soils (BN$_{100}$) increased biomass yield by 39.05% in 2014, 37.31% in 2015 and 30.02% in 2016 on average compared to soils without carbon (Table 8). Similarly, BN$_{100}$ significantly increased biomass yield in 2014 (by 35.06% and 26.43%), 2015 (by 40.04% and 23.11%) and 2016 (by 21.86% and 13.45%) compared to SN$_0$ and SN$_{50}$ sites, respectively. Application of SN$_{100}$ also caused significant increases in biomass yield compared to no carbon soils, an average increase of 32.09%, 29.32% and 32.56% were recorded in 2014, 2015 and 2016 respectively. The grain yield under $N_{100}$ fertilization was significantly increased ($p < 0.05$) by 35.87%, 29.45% and 13.34% under no carbon soils; 33.64%, 37.02% and 39.16% under biochar soils, and 31.89%, 32.35% and 24.08% under biomass treated soils in 2014, 2015 and 2016, respectively, compared to their corresponding $N_0$ soils (Table 9).

### DISCUSSION

The lowest cumulative N$_2$O emission was recorded in the biochar treated soils and at a lesser magnitude in the straw treated soils, whereas the highest N$_2$O emission was observed in the no carbon treated soils. In both cases, the highest rate of N fertilizer recorded the greatest N$_2$O emission. In contrast, Chatskikh (2007) and Kammann et al. (2012) reported that N$_2$O fluxes were significantly increased by addition of biochar, particularly when added with mineral N-fertilizer. It has been shown that the type and rate of fertilizer have an important impact on N$_2$O emissions (Bouwman, Boumans & Batjes, 2002). Some studies have reported that use of crop straw combined with mineral nitrogen fertilizer enhances soil quality while reducing N$_2$O emissions (Xu, Han & Ru, 2019; Sainju, 2016). Crop straw return commonly aims at improving soil carbon and nitrogen cycling (Xu, Han & Ru, 2019; Meng et al., 2017), thought it can also be a source of trace gas emissions (Cha...
Table 8  Biomass yield of spring wheat as affected by different treatment.

| Treatment | C source | N rates | 2014   | 2015   | 2016   | Mean   |
|-----------|----------|---------|--------|--------|--------|--------|
| No carbon | N_0      |         | 2776d  | 3030d  | 2455d  | 2754c  |
|           | N_50     |         | 3102c  | 3358bcd | 3022c  | 3161bc |
|           | N_100    |         | 3399bc | 3739b  | 3267bc | 3468b  |
| Biochar   | N_0      |         | 3295bc | 3530bc | 3147bc | 3324b  |
|           | N_50     |         | 3489b  | 3767b  | 3331bc | 3529b  |
|           | N_100    |         | 4291a  | 4630a  | 3788a  | 4236a  |
| Straw     | N_0      |         | 3170bc | 3312cd | 3118bc | 3200bc |
|           | N_50     |         | 3403bc | 3765b  | 3365b  | 3511b  |
|           | N_100    |         | 4082a  | 4345a  | 3633a  | 4020b  |

Notes. Values with different letters within a column are significantly different at $P < 0.05$.

Table 9  Grain yield of spring wheat as affected by different treatments.

| Treatment | C source | N rates | 2014   | 2015   | 2016   | Mean   |
|-----------|----------|---------|--------|--------|--------|--------|
| No carbon | N_0      |         | 1305d  | 1500d  | 1009d  | 1271d  |
|           | N_50     |         | 1538cd | 1896bc | 1043cd | 1492bcd |
|           | N_100    |         | 1770abc| 1927bc | 1144cd | 1614cd |
| Biochar   | N_0      |         | 1603bcd| 1789cd | 1124cd | 1505bcd |
|           | N_50     |         | 1905abc| 2133b  | 1233bc | 1757abc |
|           | N_100    |         | 2139a  | 2456a  | 1567a  | 2054a  |
| Straw     | N_0      |         | 1502cd | 1658cd | 1111cd | 1424cd |
|           | N_50     |         | 1852abc| 1944bc | 1182cd | 1659bc |
|           | N_100    |         | 1975ab | 2180ab | 1380ab | 1845ab |

Notes. Values with different letters within a column are significantly different at $P < 0.05$.

et al., 2016). Nitrogen fertilization has the greatest potential to increase N₂O emissions because mineral N controls both nitrification and denitrification. Other studies (Zhang et al., 2011) have shown that biochar combined with N-fertilizer can significantly reduce N₂O emissions. One mechanism that may explain lower (cumulative) N₂O fluxes from biochar + N-fertilizer-amended soils is the fact that relatively low C soils treated with N-fertilizer and biochar may retain relatively higher amounts of mineral N than soils untreated with N-fertilizer (Zhang et al., 2011). Nitrogen thereby retained provides a source of available N for plant uptake, which reduces N availability for microbes involved in denitrification processes. Since biochar has significant impact on soil environment and affects many soil parameters such as the availability of substrates (Van Zwieten et al., 2009), it is very likely that biochar will have significant effects on the production of N₂O. Their results is confirmed by the increased plant N uptake in this study (Table S4). Singh et al. (2010) reported that biochar can also reduce the N availability to microorganisms by absorption. In this study, improved soil porosity could also explain the decreased N₂O emission recorded.
when biochar was applied with N fertilizer. Soil aeration and improved porosity inhibit denitrification. Nitrogen dynamics are affected by changes in soil aeration, pH and the C/N ratio of the material incorporated into the soil. Biochar may suppress N₂O production from denitrification by increasing the air content of the soil or by absorbing water from the soil, thus improving aeration of the soil (Yanai, Toyota & Okazaki, 2007). Karhu et al. (2011) shared similar view and observed that biochar amendment modifies soil physical properties such as reducing soil bulk density or increasing water holding capacity (Karhu et al., 2011), thereby increasing soil aeration. This may lead to lower soil N₂O emissions, as soil aeration influences both nitrifier and denitrifier activity. Soils, which are not affected by compaction often exhibit adequate porosity and therefore the risk of denitrification is lower compared with soils that have impaired infiltration or internal drainage (Antille, 2018). In this study, lower N₂O emissions were also observed on the straw treated plots, although the effects were lesser relative to the biochar treated soils. The lower N₂O emission under straw treated soils could be attributed to the accumulation of organic matter on the soil surface that led to reduced bulk density and thus improved soil aeration.

Reductions in CH₄ emission were observed in biochar–amended soils and to a lesser extent on straw amended soils compared to their controls. Literature evidence indicated that biochar input to soil can potentially reduce CH₄ emissions (Yeboah et al., 2018). In contrast, Xie et al. (2021) showed that charcoal input into soil may increase soil methane fluxes. The mechanisms underlying changes in soil CH₄ emissions following biochar amendment are unclear (Lehmann et al., 2011). The greater uptake of CH₄ may be attributed to the protected environment created for the CH₄ oxidizers and improved soil porosity. In this study, the greater uptake of methane in the soils with carbon amendment, particularly biochar amended soils with N fertilizer may be attributed to the favorable environment created for the CH₄ oxidizers. The aerobic, well drained soils can be a sink for CH₄ due to the possible high rate of CH₄ diffusion and ensuing oxidation by methanotrophs. Combined application of biochar and inorganic N-fertilizer in this study improved soil physical properties (reduction in soil bulk density and increased soil saturated hydraulic conductivity). Such improved soil structural conditions are known to protect the ecological niche for methanotrophic bacteria, influence the gaseous diffusivity, and affect the rate of supply of atmospheric CH₄ (Hütsch, 1998; Serrano-Silva et al., 2014; Ma et al., 2016). Aerobic, well–drained soils behave as a sink for CH₄ due to the high rates of CH₄ diffusion and subsequent oxidation by methanotrophs (Serrano-Silva et al., 2014). However, these results do not appear to support the conclusions of Laird (2008) on the reduction observed in methane emissions from field plots, which was deduced as an increased CH₄ oxidation activity. Other studies have reported significant increase in CH₄ emissions following biochar or biomass application (e.g., Wang et al., 2012). The authors explained that, the increased availability of labile C substrates following biochar or biomass addition stimulates the activities of methanogenic bacteria which may account for increased CH₄ emissions. However, this could be a short-term effect since labile carbon fraction in the materials could be mineralized rapidly (Wang et al., 2012).

The results of this study indicate that when biochar was applied together with fertilizer N, both biomass and grain yield of spring wheat increased. This finding shows the potential
of biochar applied together with fertilizer N to improve nutrient use efficiency in spring wheat in semiarid environment (Solaiman et al., 2010). Diverse reasons have been given to the positive effect of biochar applied in combination with fertilizer N on crop yield. Bruun et al. (2011) reported that combined application of biochar and N fertilizer has the potential to improve soil properties and could therefore be responsible for the effect observed. Similarly, both Borchard et al. (2012) and Tammeorg et al. (2014) attribute increased crop productivity when biochar is applied together with N fertilizer to improve nutrient availability. In the current study, increased yield may be attributed to increased nutrient availability and improved soil physical and chemical properties (soil bulk density, saturated hydraulic conductivity and soil organic carbon), as reported in earlier work (Zhang et al., 2010). These results imply that, when biochar and inorganic fertilizers are applied together, an increased nutrient supply to plants may be the most important factor in increasing crop yields. The higher biomass and grain yield obtained on the carbon amended soils compared to the soils without carbon in this study is attributed to the fact that in drier soils, crop residues provide a better soil environment by reducing temperature, conserving water, and improving soil quality resulting in better yield (Zou et al., 2016). Positive effects of biochar combined with N fertilizer on increasing SOC and hydraulic conductivity as well as decreasing soil bulk density was observed in this study. Therefore, this study evidenced a positive effect of biochar amendment on soil quality and spring wheat yield consistent over three consecutive years. Furthermore, the lowest yield recorded on the no carbon soils throughout this study may be related to the removal of all the aboveground biomass at the end of the cropping season. Zhang, Yang & Wu (2008) showed that field practices with low carbon inputs to arable soils as crop biomass removal and manure abandonment deplete soil organic carbon and reduce crop productivity. Therefore, when biochar was applied and crop residues retained, it had immediate effect and the beneficial influence on biomass and grain yields were obtained.

CONCLUSIONS

Application of crop residue amendments combined with nitrogen fertilizer has been increasingly recommended as an effective management practice for mitigating greenhouse gas emissions while enhancing soil fertility, thereby increasing crop production. In this paper, we have shown that application of carbon amendment, especially biochar combined with N fertilizer in wheat grown under rain fed conditions in a semi-arid environment reduced nitrous oxide and methane emissions whilst increasing biomass and grain yield. This study confirmed our hypothesis that increased C inputs would increase the soils ability to reduce N₂O and CH₄ soil emissions whilsts increasing biomass and grain yield. The main conclusions derived from this work are: application of biochar + N-fertilizer (BN₁₀₀) or straw + N-fertilizer (SN₁₀₀) increased saturated hydraulic conductivity to significantly greater extent than the other treatments tested. This translated into higher biomass production and therefore grain yield in those treatments. These results indicate the dependency of crop yield on soil quality and imply that crop productivity could be increased without resource degradation when biochar is applied combined with N-fertilizer.
Application of biochar + N-fertilizer showed relatively lower N$_2$O emissions, including increased uptake of CH$_4$, but the effect of BN$_{100}$ was consistently greater. The findings of this study suggest that biochar applied together with N-fertilizer can concurrently improve soil physical and chemical properties as well as biomass and grain yield while reducing the effect of agricultural activities on the environment. Based on this results, the potential exist for developing crop and soil management interventions around biochar applied together with fertilizer N in semiarid environments. Further studies that focus on N$_2$O and CH$_4$ measurements after every rainfall, tillage and fertilization events are required for better recommendations.

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**ADDITIONAL INFORMATION AND DECLARATIONS**

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**Competing Interests**

The authors declare there are no competing interests.

**Author Contributions**

• Stephen Yeboah conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
• Wu Jun performed the experiments, analyzed the data, prepared figures and/or tables, and approved the final draft.
• Cai Liqun conceived and designed the experiments, authored or reviewed drafts of the paper, and approved the final draft.
• Patricia Oteng-Darko and Erasmus Narteh Tetteh analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.
Zhang Renzhi conceived and designed the experiments, performed the experiments, analyzed the data, authored or reviewed drafts of the paper, and approved the final draft.

Data Availability
The following information was supplied regarding data availability:

The raw data for N<sub>2</sub>O and CH<sub>4</sub>, grain and biomass yield, soil data and Plant N are available in the Supplemental Files.

Supplemental Information
Supplemental information for this article can be found online at http://dx.doi.org/10.7717/peerj.11937#supplemental-information.

REFERENCES

Antille DL. 2018. Evaluation of fertigation applied to furrow and overhead irrigated cotton grown in a Black Vertosol in Southern Queensland, Australia. Applied Engineering in Agriculture 34(1):197–211 DOI 10.13031/aea.12519.

Arfaoui A, Ibrahimi K, Trabelsi F. 2019. Biochar application to soil under arid conditions: a bibliometric study of research status and trends. Arabian Journal of Geosciences 12:45 DOI 10.1007/s12517-018-4166-2.

Borchard N, Wolf A, Laabs V, Ackersberg R, Scherer HW, Moeller A, Amelung W. 2012. Physical activation of biochar and its meaning for soil fertility and nutrient leaching—a greenhouse experiment. Soil Use Management 28:177–184 DOI 10.1111/j.1475-2743.2012.00407.x.

Bouwman AF, Boumans LJM, Batjes NH. 2002. Emissions of N<sub>2</sub>O and NO<sub>2</sub> from fertilized fields: summary of available measurement data. Global Biogeochemical Cycles 16:6-1–6-13 DOI 10.1029/2001GB001811.

Bremner JM, Mulvaney CS. 1982. Nitrogen-Total. In: Page AL, Miller RH, Keeney DR, eds. Methods of soil analysis. Part 2. Chemical and microbiological properties. Madison, Wisconsin: American Society of Agronomy, Soil Science Society of America, 595–624.

Bruun EW, Muller-Stover D, Ambus P, Hauggaard-Nielsen H. 2011. Application of biochar to soil and N<sub>2</sub>O emissions: potential effects of blending fast–pyrolysis biochar with anaerobically digested slurry. European Journal of Soil Science 62:581–589 DOI 10.1111/j.1365-2389.2011.01377.x.

Carter MR. 1993. Soil sampling and methods of analysis, Canadian Society of Soil Science. Boca Raton: CRC Press, 63–72.

Cha JS, Park SH, Jung SC, Ryu C, Jeon JK, Shin MC, Park YK. 2016. Production and utilization of biochar: a review. Journal of Industrial and Engineering Chemistry 40:1–15 DOI 10.1016/j.jiec.2016.06.002.

Chatskikh D. 2007. Soil tillage enhanced CO<sub>2</sub> and N<sub>2</sub>O emissions from loamy sand soil under spring barley. Soil and Tillage Research 97:5–18 DOI 10.1016/j.still.2007.08.004.
Chinese Soil Taxonomy Cooperative Research Group. 1995. *Chinese Soil Taxonomy (Revised Proposal).* Academic Sinica/Beijing: Institute of Soil Science/Chinese Agricultural Science and Technology Press.

Clough TJ, Condon LM, Kammann C, Müller C. 2013. A review of biochar and soil nitrogen dynamics. *Agrono* 3:275–293 DOI 10.3390/agronomy30.

FAO. 1990. *Soil map of the world: revised legend.* World soil resources report 60. Rome: Food and Agriculture Organization of the United Nations.

Feng X, Fu B, Lu N, Zeng Y, Wu B. 2013. How ecological restoration alters ecosystem services: an analysis of carbon sequestration in China’s Loess Plateau. *Scientific Reports* 3:2846.

Grace P. 2016. Soil Research. *Foreword* 54(5):i–ii DOI 10.1071/SRv54n5_FO.

He L, Cleverly J, Chen C, Yang X, Li J, Liu W, Yu Q. 2014. Diverse responses of winter wheat yield and water use to climate change and variability on the semiarid Loess Plateau in China. 2013. *Agronomy Journal* 106:1169–1178 DOI 10.2134/agronj13.0321.

Huang JP, Ma JR, Guan XD, Li Y, He YL. 2019. Progress in semi-arid climate change studies in China. *Advances in Atmospheric Sciences* 36(9):922–937 DOI 10.1007/s00376-018-8200-9.

Hütsch BW. 1998. Tillage and land use effects on methane oxidation rates and their vertical profiles in soil. *Biology and Fertility of Soils* 27:284–292 DOI 10.1007/s003740050435.

IPCC. 2014. Climate Change: mitigation of Climate Change. Intergovernmental Panel on Climate Change. *Available at http://www.ipcc.ch/report/ar5/wg3/*.

Kammann C, Ratering S, Eckhard C, Müller C. 2012. Biochar and hydrochar effects on greenhouse gas (carbon dioxide, nitrous oxide, and methane) fluxes from soils. *Journal of Environmental Quality* 41:1052–1066 DOI 10.2134/jeq2011.0132.

Karhu K, Mattila T, Mattila T, Bergström I, Regina K. 2011. Biochar addition to agricultural soil increased CH₄ uptake and water holding capacity — results from a short-term pilot field study. *Agriculture Ecosystems & Environment* 140(1):309–313 DOI 10.1016/j.agee.2010.12.005.

Laird DA. 2008. The charcoal vision: a win-win-win scenario for simultaneously producing bioenergy, permanently sequestering carbon, while improving soil and water quality. *Agronomy Journal* 100:178–181.

Laird D, Fleming P, Wang B, Horton R, Karlen D. 2010. Biochar impact on nutrient leaching from a Midwestern agricultural soil. *Geoderma* 158:436–444 DOI 10.1016/j.geoderma.2010.05.012.

Lamptey S, Li L, Xie J. 2018. Impact of nitrogen fertilization on soil respiration and net ecosystem production in maize. *Plant, Soil and Environment* 64:353–360 DOI 10.17221/217/2018-PSE.

Lehmann J, Rillig MC, Thies J, Masiello CA, Hockaday WC, Crowley D. 2011. Biochar effects on soil biota –a review. *Soil Biology and Biochemistry* 43:1812–1836 DOI 10.1016/j.soilbio.2011.04.022.
Liu QF, Chen Y, Li WW, Liu Y, Han J, Wen XX, Liao YC. 2016. Plastic-film mulching and urea types affect soil CO$_2$ emissions and grain yield in spring maize on the Loess Plateau. *China. Sci. Rep* 6:1–10 DOI 10.1038/s41598-016-0001-8.

Lu RK. 2000. *Analysis method of soil agricultural chemistry*. Beijing: China Agricultural Science and Technology Press.

Ma N, Zhang L, Zhang Y, Yang L, Yu C, Yin G, Doane ATimothy, Wu Z, Zhu P, Ma X. 2016. Biochar improves soil aggregate stability and water availability in a mollisol after three years of field application. *PLOS ONE* 11(5):e0154091 DOI 10.1371/journal.pone.0154091.

Meng F, Dungait JA, Xu X, Bol R, Zhang X, Wu W. 2017. Coupled incorporation of maize (*Zea mays* L.) straw with nitrogen fertilizer increased soil organic carbon in Fluvic Cambisol. *Geoderma* 304:19–27 DOI 10.1016/j.geoderma.2016.09.010.

Nelson DW, Sommers LW. 1982. Total carbon, organic carbon and organic matter. In: Page AL, Miller RH, Keeney DR, eds. *Methods of soil analysis Part 3: Chemical and Microbiological properties*. Agronomy monographs, Madison: Soil Science Society of America, Inc., 961–1010 DOI 10.2134/agronmonogr9.2.2ed.c29.

Norton R. 2014. Combating climate change through improved agronomic practices and input-use efficiency. *Journal of Crop Improvement* 28(5):575–618 DOI 10.1080/15427528.2014.924331.

Rivka BF, David AL, Timothy BP. 2019. Effect of biochar on soil greenhouse gas emissions at the laboratory and field scales. *Soil Systems* 3(8):2–18 DOI 10.3390/soilsystems3010008.

Saggar S. 2010. Estimation of nitrous oxide emission from ecosystems and its mitigation technologies. *Agriculture Ecosystem Environment* 136:189–191 DOI 10.1016/j.agee.2010.01.007.

Sainju UMA. 2016. Global meta-analysis on the impact of management practices on net global warming potential and greenhouse gas intensity from cropland soils. *PLOS ONE* 1(2):e0148527 DOI 10.1371/journal.pone.0148527.

Serrano-Silva N, Sarria-Guzman Y, Dendooven L, Luna-Guido M. 2014. Methanogenesis and methanotrophy in soil: a review. *Pedosphere* 24(3):291–307 DOI 10.1016/S1002-0160(14)60016-3.

Singh BP, Hatton BJ, Singh B, Cowie AL, Kathuria A. 2010. Influence of biochars on nitrous oxide. *Journal of Environmental Quality* 39(4):1224–1235 DOI 10.2134/jeq2009.0138.

Solaiman ZM, Blackwell P, Abbott IK, Storer P. 2010. Direct and residual effect of biochar application on mycorrhizal root colonisation, growth and nutrition of wheat. *Soil Research* 48:546–554 DOI 10.1071/SR10002.

Solomon S, Qin DM, Manning Z, Chen M, Marquis KB, Averyt M, Tignor, Miller HL. 2007. Technical Summary. in Climate Change 2007: the physical science basis, contribution of working group I to the fourth assessment report of the intergovernmental panel on climate change. Cambridge University Press, Cambridge, United Kingdom and New York, NY. Available at http://www.ipcc.ch.
Steiner C, Teixeira WG, Lehmann J, Nehls T, Macêdo JLV, Blum WEH, Zech W. 2007. Long term effects of manure, charcoal and mineral fertilization on crop production and fertility on a highly weathered central Amazonian upland soil. *Plant and Soil* 291:275–290 DOI 10.1007/s11104-007-9193-9.

Tammeorg P, Asko S, Pirjo M, Stoddard LF, Laura A, Juha H. 2014. Biochar application to a fertile sandy clay loam in boreal conditions: effects on soil properties and yield formation of wheat, turnip rape and fava bean. *Plant Soil* 374:89–107 DOI 10.1007/s11104–013–1851–5.

Tullberg J, Antille DL, Bluetta C, Eberhard J, Scheer C. 2018. Controlled traffic farming effects on soil emissions of nitrous oxide and methane. *Soil and Tillage Research* 176(1):18–25.

Van Zwieten L, Bhupinderpal-Singh, Joseph S, Kimber S, Cowie A, Chan Y. 2009. Biochar reduces emissions of non–CO$_2$ GHG from soil. In: Lehmann J, Joseph S, eds. *Biochar for environmental management*. London: Earthscan Publications Ltd, 227–249.

Walkley AJ, Black IA. 1934. Estimation of soil organic carbon by the chromic acid titration method. *Soil Science* 37:29–38 DOI 10.1097/00010694-193401000-00003.

Wang J, Xie J, Li L, Luo Z, Zhang R, Wang L, Jiang Y. 2021. The Impact of Fertilizer Amendments on Soil Autotrophic Bacteria and Carbon Emissions in Maize Field on the Semi-arid Loess Plateau. *Frontiers in Microbiology* 12:664120 DOI 10.3389/fmicb.2021.664120.

Wang W, Peng SS, Wang T, Fang JY. 2010. Winter soil CO$_2$ efflux and its contribution to annual soil respiration in different ecosystems of a forest–steppe ecotone, North China. *Soil Biology and Biochemistry* 42(3):451–458 DOI 10.1016/j.soilbio.2009.11.028.

Wang L, Tian H, Song C, Xu X, Chen G, Ren W, Lu C. 2012. Net exchanges of CO$_2$, CH$_4$ and N$_2$O between marshland and the atmosphere in Northeast China as influenced by multiple global environmental changes. *Atmospheric Environment* 63:77–85.

Wang L, Tian H, Song C, Xu X, Chen G, Ren W, Lu C. 2012. Biochar stability in soil: meta-analysis of decomposition and priming effects. *GCB Bioenergy* DOI 10.1111/gcbb.12266.

Woolf D, Amonette JE, Street-Perrott FA, Lehmann J, Joseph S. 2010. Sustainable biochar to mitigate global climate change. *Nature Communications* 1:1–9 DOI 10.1038/ncomms1053.

Xie Z, Xu Y, Liu G, Liu Q, Zhu J, Tu C, Amonette JE, Cadisch JWY, Hu S. 2021. Impact of biochar application on nitrogen nutrition of rice, greenhouse-gas emissions and soil organic carbon dynamics in two paddy soils of China. *Plant Soil* 370:527–540.

Xu C, Han X, Ru S. 2019. Cardenas, L.; Rees, R.M.; Wu, D.; Wu, W.; Meng, F. Crop straw incorporation interacts with N fertilizer on N$_2$O emissions in an intensively cropped farmland. *Geoderma* 341:129–137 DOI 10.1016/j.geoderma.2019.01.014.

Xu ZF, Yang ZL. 2017. Relative impacts of increased greenhouse gas concentrations and land cover change on the surface climate in arid and semi-arid regions of China. *Climatic Change* 144:491–503 DOI 10.1007/s10584-017-2025-x.
Yanai Y, Toyota K, Okazaki M. 2007. Effect of charcoal addition on N\textsubscript{2}O emissions from soil resulting from rewetting air–dried soil in short–term laboratory experiments. 
Soil Science and Plant Nutrition 53:181–188 DOI 10.1111/j.1747-0765.2007.00123.x.

Yeboah S, Lamptey S, Cai L, Song M. 2018. Short-term effects of biochar amendment on greenhouse gas emissions from rainfed agricultural soils of the semi–arid loess plateau region. Agronomy 8:74 DOI 10.3390/agronomy8050074.

Yeboah S, Zhang R, Cai L, Song M, Li L, Xie Z, Luo Z, Wu J, Zhang J. 2016. Greenhouse gas emissions in a spring wheat–field pea sequence under different tillage practices in semi-arid Northwest China. Nutrient Cycling in Agroecosystems 106:77–91 DOI 10.1007/s10705-016-9790.

Zhang A, Cui L, Pan G, Li L, Hussain Q, Zhang X, Zheng J,Crowley D. 2010. Effect of biochar amendment on yield and methane and nitrous oxide emissions from a rice paddy from Tai Lake plain, China. Agriculture, Ecosystems & Environment 139:469–475 DOI 10.1016/j.agee.2010.09.003.

Zhang A, Liu Y, Pan G, Hussain Q, Li L, Zheng J, Zhang X. 2011. Effect of biochar amendment on maize yield and greenhouse gas emissions from a soil organic carbon poor calcareous loamy soil from Central China Plain. Plant Soil 139:469–475 DOI 10.1007/s11104-011-0957.

Zhang YT, Guan XD, Yu PY, Xie KY, Jin CH. 2017. Contributions of radiative factors to enhanced dryland warming over East Asia. Journal of Geophysical Research 122:7723–7736 DOI 10.1002/2017JD026506.

Zhang QZ, Yang ZL, Wu WL. 2008. Role of crop residue management in sustainable agricultural development in the North China Plain. Journal of Sustainable Agriculture 32:137–148.

Zhu Z, Ge T, Luo Y, Liu S, Xu X, Tong C, Shibistova O, Guggenberger G, Wu J. 2018. Microbial stoichiometric flexibility regulates rice straw mineralization and its priming effect in paddy soil. Soil Biology & Biochemistry 121:67–76 DOI 10.1016/j.soilbio.2018.03.003.

Zimmerman AR, Gao B, Ahn MY. 2011. Positive and negative carbon mineralization priming effects among a variety of biochar-amended soils. Soil Biology & Biochemistry 43:1169–1179 DOI 10.1016/j.soilbio.2011.02.005.

Zou H, Ye X, Li J, Lu J, Fan Q, Yu N. 2016. Effects of straw return in deep soils with urea addition on the soil organic carbon fractions in a semi–arid temperate cornfield. PLOS ONE 11(4):e0153214 DOI 10.1371/journal.pone.0153214.

Zou J, Huang Y, Jiang J, Zheng X, Sass RL. 2005. A 3-year field measurement of methane and nitrous oxide emissions from rice paddies in China: effects of water regime, crop residue, and fertilizer application. Global Biogeochemical Cycles 19:19–31.