Research Article

A Novel Prognostic Model Based on Seven Necroptosis-Related miRNAs for Predicting the Overall Survival of Patients with Lung Adenocarcinoma

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Received 11 November 2021; Accepted 11 February 2022; Published 24 March 2022

Academic Editor: Vitale Miceli

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Lung adenocarcinoma (LUAD) remains one of the leading causes of cancer-related deaths worldwide. This study is aimed at constructing a risk scoring model based on necroptosis-related miRNAs to predict prognosis of LUAD. Expression profile of miRNA in LUAD was downloaded from The Cancer Genome Atlas (TCGA) database. We screened the differentially expressed necroptosis-related miRNAs between LUAD patients and normal samples, thus constructed a seven miRNA-based risk stratification on the basis of the TGCA cohort. This risk stratification was prove to be effective in predicting the overall survival (OS) of patients with LUAD. Furthermore, we constructed a nomogram model based on the combination of risk characteristics and clinicopathological features, which was also prove to be accurate and efficient in predicting OS of LUAD patients. Functional enrichment analyses on the targeted genes of these miRNAs with prognostic value were carried out. Results indicated that these targeted genes were closely related to the development and metastasis of tumors. In summary, our research has developed a prognostic model based on the expression of miRNAs related to necroptosis. This model might be used to predict the prognosis of LUAD accurately, which might be helpful in improving treatment efficacy of LUAD.

1. Introduction

Lung cancer is a common but complex malignant disease, and most patients with lung cancer were diagnosed at advanced stage [1]. According to the latest global cancer statistics report by the International Agency for Research on Cancer, lung cancer remains the leading cause of cancer death worldwide, 2.2 million people were diagnosed with lung cancer in 2020 worldwide, and 1.79 million were died of lung cancer [2]. Unfortunately, the morbidity and mortality of lung cancer are still rising globally [3]. Lung adenocarcinoma (LUAD) represents the most frequent pathological type in lung cancer, which accounts for about 40% of the total lung cancers [4, 5]. Although advances have been made in current treatment strategies for lung cancer (surgical resection, chemotherapy, radiotherapy, molecular targeted therapy, and immunotherapy), the prognosis of lung cancer is still not satisfied. The incidence of 5-year survival of patients with lung cancer remains only 4%-17%, and the incidence of 5-year survival is less than 5% in patients with metastasized tumor [6–8]. Therefore, early detection with a comprehensive and accurate risk assessment is of great significance in diagnosing and monitoring of lung cancer. Current tools of risk assessment and monitoring of lung cancer are mostly based on clinical characteristics and pathological parameters, in which TNM stratification is most commonly used. However, the existing TNM model is often associated with a limited predictive confidence in predicting prognosis of lung cancer, which was comprised of huge
heterogeneity among individuals. Therefore, there is a unmet need for cooperating clinicopathological characteristics of the genome in assess individual survival prognosis.

Necroptosis is a newly identified caspase-independent function of programmed cell death, which is different from apoptosis [9]. Morphological manifestation of necroptosis is demonstrated by swelling and rounding cells, explosive plasma membrane rupture, mitochondrial dysfunction and loss of mitochondrial membrane potential, and cell membrane perforation [10]. When the expression of caspase-8 is inhibited or at a low level in cells, receptor-interacting protein 1 (RIP1) can recruit receptor-interacting protein 3 (RIP3) to form a RIP1-RIP3 complex, which induces a mixed lineage of pseudokinases. The kinase domain-like protein (MLKL) is phosphorylated to form necrosomes, which leads to necroptosis [11]. Necroptosis has been proved to have a double-edged sword effects on cancer, and the relationship between necroptosis and cancer is complicated. Although it is reported that necroptosis can inhibit tumor development and metastasis when apoptosis is blocked, its key regulators will also provide assistance for tumor metastasis and progression [12]. Emerging evidences have indicated that necroptosis can inhibit tumor development and metastasis, thus may be utilized as a potential method in treating cancers [13–15]. For example, necroptotic MiR-7-5p inhibits tumor metastasis by targeting NOVA2 in lung cancer [16]. Although previous studies have reported that necroptosis has antitumor effects in various types of cancer including LUAD, the knowledge of the prognostic value of necroptosis-related microribonucleic acid (miRNA, a set of short noncoding RNA with regulatory functions [17]) is still poor.

In this study, we constructed a risk scoring stratification based on miRNAs related to necroptosis. Meanwhile, to improve the prognosis confidence of this model, we have established a nomogram that integrates risk scores and clinical factors. This study is aimed at evaluating the prognostic value of necroptosis-related miRNAs in LUAD and develops a prognostic nomogram based on necroptosis-related miRNAs as a accurate prediction tool to effectively evaluate the clinical prognosis of patients with LUAD.

2. Materials and Methods

2.1. Data Collection. The schematic diagram was shown in Figure 1. Transcriptomic data sets of miR-seq were downloaded from the public lung adenocarcinoma (LUAD) database of TCGA (https://portal.gdc.cancer.gov/), which contain 521 LUAD patient samples and 46 normal control samples. Clinical data of corresponding patients was also downloaded. We excluded data from patients with no OS information. Thus, a total of 499 LUAD patients were included in the subsequent analyses. Detailed information on these 499 patients with LUAD is presented in Supplementary Table 1. This research complies with the TCGA data access policy. Meanwhile, this study was exempted from relevant ethical approval because the TCGA database is publicly available.

2.2. Differentially Expressed Necroptosis-Related miRNAs between LUAD Patients and Normal Control Samples. Thirteen miRNAs related to necroptosis were profiled according to the previous publication [12] (miR-495, miR-331-3p, miR-15a, miR-148a-3p, miR-7-5p, miR-141-3p, miR-425-5p, miR-200a-5p, miR-210, miR-223-3p, miR-500a-3p, miR-181-5p, miR-16-5p). By using the “tidyverse” R package, the expression of 13 necroptosis-related miRNAs was profiled with data downloaded from the TCGA database and was used for subsequent analysis. Then, the “limma” R package was used to identify the differentially expressed necroptosis-related miRNAs between LUAD patient samples and normal control samples, with false discovery rate (FDR) < 0.05 as statistically significant.

2.3. Establishment of a Prognostic Stratification Based on the TCGA Cohort. In order to assess the prognostic value of these necroptosis-related miRNAs, we used a Cox regression analysis to identify the miRNAs significantly related to the OS of LUAD. A total of 7 necroptosis-related miRNAs were identified to be significantly related to OS. Due to the limited number of identified miRNAs (less than 10), no further LASSO Cox regression analysis was performed. At last, 7 necroptosis-related miRNAs were retained to establish a risk scoring model. The risk score of each patient is calculated as the sum of the remaining scores of each miRNA, and each miRNA score is calculated by multiplying the miRNA coefficient by the miRNA expression level. The specific risk score formula is as follows: risk score = \sum_{i=1}^{n} coei \times expi (n represents the number of miRNAs, coei represents the regression coefficient of miRNAi, and expi represents the expression level of miRNAi) [18, 19]. According to the median value of the risk score, TCGA LUAD patients were divided into high- and low-risk groups. The “survival” R package, “timeROC” R package, and “ROCR” R package were used to compare the OS of the high- and low-risk groups, and 3-year and 5-year ROC curve analysis was performed. In addition, to investigate whether the risk score could be used as an independent factor of OS in LUAD, univariate and multivariate Cox regression analysis was performed. Meanwhile, the “pheatmap” package was utilized to plot a heat map of risk characteristics and related clinical factors.

2.4. Establishment of the Forecastive Nomogram. The “survival” R package and the “rms” R package were used to construct a predictive nomogram model, which includes risk scores and associated clinical factors (age, gender, smoking history (years), and TNM staging). A calibration curve was established to evaluate the consistency between the results of predictive model and the actual clinical results when nomogram model was applied.

2.5. Functional Enrichment Analysis. Potential targeted genes of these miRNAs were identified by taking the intersection of predicted genes by miRDB (http://www.mirdb.org/), miRTarBase (http://miurtherbase.mbc.nctu.edu.tw/php/index.php), and TargetScan (http://www.targetscan.org). Identified targeted genes were analyzed using gene ontology (GO) and Kyoto
3. Results

3.1. Differentially Expressed Necroptosis-Related miRNAs between Tumor and Normal Samples. A total of 7 differentially expressed miRNAs were identified between tumor and normal samples, among which miR-141-3p, miR-148a-3p, miR-200a-5p, miR-331-3p, miR-425-5p, and miR-500a-3p were upregulated in the cancer group, while miR-223-3p was downregulated (Figure 2(a)). The expression level of differentially expressed miRNAs was shown by a heat map. Rows represented miRNAs, and columns represented biological samples. The red color represented higher expression, and the green color represented lower expression (Figure 2(b)).

3.2. Establishment of a Prognostic Model Based on the TCGA Cohort. Cox regression analysis was used to identify those necroptosis-related miRNAs which were significantly related to OS of LUAD. A total of 7 necroptosis-related miRNAs were identified and were used to establish a risk scoring model. Formula of scoring for each individual was

\[ \text{Score} = -0.03043 \times \text{miR-141-3p} - 0.14387 \times \text{miR-148a-3p} - 0.02951 \times \text{miR-200a-5p} - (0.07482 \times \text{miR-223-3p}) + 0.01450 \times \text{miR-331-3p} + 0.18310 \times \text{miR-425-5p} - 0.11271 \times \text{miR-500a-3p}. \]

LUAD patients were assigned to the high-risk group, patients in the low-risk group were associated with lower incidence of deaths and longer OS compared to patients in the high-risk group (Figure 3(b), left side of the dotted line). The survival curve showed that OS of patients in the low-risk group was longer than patients in the high-risk group in the TCGA LUAD cohort \((P = 0.0053, \text{Figure 3(c)})\). Sensitivity and specificity of the prognostic model were calculated using a ROC method. Results revealed that the AUC of 3-year and 5-year OS was 0.631 and 0.605, respectively (Figure 3(d)).

3.3. Independent Prognostic Value of Risk Factors. To evaluate whether these miRNA-based risk scores could be used as independent prognostic factors in the TCGA cohort, univariate and multivariate Cox regression analysis was performed. Results of univariate Cox regression analysis showed that the 7 necroptosis-related miRNA risk scores were independent prognostic factors in the TCGA cohort (HR = 1.5299, 95% CI [1.1034-2.1213], \(P\) value = 0.0108 \((P < 0.05, \text{Figure 4(a)})\)). Meanwhile, results of multivariate Cox regression analysis also confirmed that the risk scores were prognostic factors for LUAD patients after integrating the other confounding factors (HR = 1.5520, 95% CI [1.1060-2.1779], \(P\) value = 0.0110 \((P < 0.05, \text{Figure 4(b)})\)).

3.4. Establishment of Risk Features and Clinicopathological Characteristic Combined Nomogram. Multiple factors and the risk value of each patient were integrated and calculated using a visualized nomogram. This nomogram combined risk characteristics and clinicopathological characteristics, which included five factors as risk characteristics, age, gender, smoking history (years), and disease stage and was used to predict the 3-year and 5-year OS of LUAD patients. According to the score value corresponding to each factor on the scale, a total score from the nomogram according to the individual situation was calculated for each patient, and the total score was used to predict the 3-year and 5-year OS (Figure 5(a)). Confidence of the nomogram was confirmed by the calibration chart, which indicated that the nomogram was associated with a ideal performance in predicting the 3-year and 5-year OS of LUAD patients (Figures 5(b) and 5(c)).

3.5. GO Enrichment and KEGG Pathway Analysis. A total of 302 targeted genes of these miRNAs with potential prognostic value were identified by Venn analysis using miRDB, miRTarBase, and TargetScan databases (Figure 6(a)).
Figure 2: Differentially expressed necroptosis-related miRNAs. (a) A volcano plot of expression change comparing the tumor and normal samples, with the seven downregulated and upregulated miRNAs. (b) The notably differentially expressed necroptosis-related miRNA in each biological specimen from TCGA LUAD cohort were displayed by heat map.
whole list of the targeted genes is in Supplementary Table 2. GO enrichment and KEGG pathway analysis of these targeted genes were further performed. The top 8 enriched biological processes, cell components, and molecular functions with the highest reliability based on the P value were shown in Figure 6(b).

PI3K–Akt signaling pathway, microRNAs in cancer, MAPK signaling pathway, FoxO signaling pathway, and proteoglycans in cancer were the most significantly enriched pathways. Illustrated by the KEGG analysis, these targeted genes of miRNAs were mainly related to cell proliferation, migration, invasion, and autophagy (Figure 6(c)).

4. Discussion

Necroptosis represents a form of cell death which is associated with the morphological features of necrotic cells, as well as intrinsic signal transductions similar to that of apoptotic cells. However, necroptosis differs from apoptosis, and it contributes to inhibition of tumor proliferation and metastasis [20–22]. Since the term necroptosis was proposed, a large number of studies have focusing on it. However, the prognostic value of necroptosis-related miRNAs on LUAD has not been fully established. In this present study, we investigated the prognostic value of 13 necroptosis-related miRNAs.
|   | P  | Hazard ratio     |   | P-value | Hazard ratio     |
|---|----|------------------|---|---------|------------------|
| Age | 0.2423 | 1.0099(0.9934–1.0267) | Age | 0.0175 | 1.0203(1.0035–1.0373) |
| Sex | 0.8004 | 1.0423(0.7559–1.4371) | Sex | 0.3884 | 0.8650(0.6222–1.2026) |
| Smoking | 0.9498 | 1.0048(0.8645–1.1680) | Smoking | 0.4362 | 1.0634(0.9109–1.2415) |
| Stage | <0.0001 | 1.6850(1.4545–1.9520) | Stage | <0.001 | 1.7313(1.4843–2.0194) |
| RiskScore | 0.0108 | 1.5299(1.1034–2.1213) | RiskScore | 0.0110 | 1.5520(1.1060–2.1779) |

**Figure 4:** Univariate and multivariate Cox regression analyses for the risk score. (a) Univariate analysis for the TCGA cohort. (b) Multivariate analysis for the TCGA cohort. (c) Heatmap of clinicopathological characteristics of the low- and high-risk patients.
and their potential regulatory mechanisms in LUAD. By profiling the expression levels of 13 known necroptosis-related miRNAs in LUAD patients and normal control tissues, we found that most miRNAs (7/13) were differentially expressed between LUAD patients and normal control tissues. To evaluate the prognostic value of these necroptosis-related miRNAs, a Cox regression analysis was used to construct a risk stratification of 7 necroptosis-related miRNAs. Results indicated that this miRNA-based stratification was associated with significant value in distinguishing patients with low or high risk. In addition, based on the 7 necroptosis-related miRNA risk stratification and clinicopathological characteristics, we established a nomogram in the LUAD cohort. This nomogram was shown to be associated with significant superiority in accurately predicting OS of LUAD comparing to conventional evaluation tools (such as TNM staging) and promoted the clinical application of the necroptosis-related miRNA risk stratification to a large extent.

By using the GO enrichment and KEGG pathway analysis, we further investigated the molecular functions
Figure 6: Continued.
underlying these necroptosis-related miRNAs. Our results indicated that targeted genes of these necroptosis-related miRNAs were mainly related to functions of cell proliferation, focal adhesion, cell matrix adhesion, and cell aggregation adhesion, which were reported to be closely associated with tumor cell migration and tumor metastasis. [23–25]. Therefore, targeted therapy by inhibiting these cell-cell interactions may represent an important therapeutic strategy to improve the outcome of LUAD treatment. According to our finding, the most enriched PI3K-Akt signaling pathway was considered to be the key signal of cell proliferation, migration, and even chemotherapy resistance in lung cancer cells [26, 27]. In addition, the MAPK signaling pathway and Ras signaling pathway were also reported to be closely related to the regulation of lung cancer cell proliferation, differentiation, and migration [28, 29]. Inhibition of these pathways may contribute to the inhibition of the growth and metastasis of lung cancer and may help improve the prognosis of LUAD.

Mechanistically, the risk hazard ratio of miR-223-3p was 1.07770 in this study, suggesting that the miR-223-3p over-expression was related to the poor prognosis of LUAD. Studies have shown that this miRNA plays a tumor-promoting effect in several types of cancers asides lung cancer. For example, upregulation of miR-223-3p in non-small-cell lung cancer enhances tumor cell viability, migration, and invasion. High expression of this miRNA is related to a significant low survival rate and inferior outcomes of patients [30]. Similar phenomena were also reported by Liu et al. and other [31]. In addition, the level of upregulation of miR-223-3p was positively correlated with the depth of tumor invasion and the degree of lymph node metastasis in gastric cancer [32]. All these observations have indicated that miR-223-3p was associated with an inferior prognosis. The risk characteristic hazard ratio of miR-425-5p in this study was also greater than 1, which was 1.20093, suggesting that the high expression of miR-425-5p may be associated with a cancer-promoting effect. A recent study found that miR-425-5p is overexpressed in lung cancer tissues, which enhances lung cancer cell proliferation and colony formation. It was also found that miR-425-5p promotes the development of lung cancer through the PTEN/PI3K/AKT signal axis [33]. In addition, the elevated expression of miR-425-5p significantly promoted the invasion and metastasis of gastric cancer and was an independent prognostic factor for a poor recurrence-free survival in patients with GC [34].

The risk characteristic hazard ratio of miR-141-3p in our study was 0.97003, indicating that miR-141-3p was a protective miRNA for LUAD. Studies have confirmed that the expression level of miR-141-3p in non-small-cell lung cancer tissues is downregulated, and its low expression is associated with poor prognosis, suggesting that miR-141-3p may be a tumor suppressor, and the high expression of this factor can help inhibit non-small-cell lung cancer cells. The development of [35].
In this study, the risk characteristic hazard ratios of miR-148a-3p and miR-500a-3p were also less than 1, which were 0.86600 and 0.89341, respectively, suggesting that miR-148a-3p and miR-500a-3p were also protective miRNAs. These observations are consistent with previous studies. Studies by Xie et al. and Bai et al. both confirmed that miR-148a-3p was significantly downregulated in non-small-cell lung cancer, and its overexpression can inhibit the proliferation and epithelial-mesenchymal transition of non-small-cell lung cancer and plays an inhibitory role on the tumor cells [36, 37]. Compared with normal tissues, the expression level of miR-500a-3p in lung cancer tissues was significantly reduced, and low expression level of miR-500a-3p was closely related to the adverse clinical outcome of lung cancer [38]. The risk characteristic hazard ratio of miR-200a-5p in our study was 0.97092, suggesting that miR-200a-5p may be a tumor suppressor factor.

There are no functional study identifying the role of miR-200a-5p in LUAD. A number of studies [39, 40] have shown that miR-331-3p plays an inhibitory role in tumor proliferation, migration, and invasion. However, the risk characteristic hazard ratios of miR-331-3p in our study was higher than 1, indicating that the miR-331-3p overexpression might be related to poor prognosis, which was in contrary to previous studies. However, owing to the fact that the modulatory effects of miRNAs are impacted by tumor microenvironment, the role of specific miRNA in different types of cancer may be inconsistent or even the on the opposite site [41]. It is necessary to continue to investigate the effect of miR-331-3p in LUAD. To overcome limitations of this present study, large-scale independent research is needed to verify the effectiveness of this risk stratification in the future.

5. Conclusion

We analyzed the prognostic value of necroptosis-related miRNAs in LUAD for the first time. Generally, 13 necroptosis-related miRNAs to were screened, and 7 miRNAs were identified to be associated with significant prognostic value. These miRNAs play an important role in the development and metastasis of LUAD and may be potential therapeutic targets for LUAD. In addition, the nomogram, which was established by the combination of risk characteristics and clinicopathological characteristics in the LUAD cohort, could effectively predict the clinical prognosis of LUAD and provide an accurate and individualized prediction tool for clinic use. In summary, this study helps to understand the biological behavior and potential treatment targets of LUAD from the perspective of necroptosis.

Data Availability

Transcriptional data sets of miR-seq were downloaded from the public lung adenocarcinoma (LUAD) database of TCGA (https://portal.gdc.cancer.gov/), which contain 521 LUAD patient samples and 46 normal control samples. Clinical data of corresponding patients was also downloaded. Potential targeted genes of these miRNAs were identified by taking the intersection of predicted genes by miRDB (http://www.mirdb.org/), miRTarBase (http://miRTarBase.mbc.nctu.edu.tw/php/index.php), and TargetScan (http://www.targetscan.org).

Conflicts of Interest

The authors have no conflicts of interest to disclose.

Authors’ Contributions

Xiaohua Hong and Guangyao Wang contributed equally to this work.

Acknowledgments

This work was supported by the National Natural Science Foundation of China (81760848) and Innovation Project of Guangxi Graduate Education (YCBZ2021077).

Supplementary Materials

Supplementary 1. Table S1: detailed information on these 499 patients with LUAD.

Supplementary 2. Table S2: the detailed results of the targeted genes.

References

[1] F. Nasim, B. F. Sabath, and G. A. Eapen, “Lung cancer,” The Medical Clinics of North America, vol. 103, no. 3, pp. 463–473, 2019.
[2] H. Sung, J. Ferlay, R. L. Siegel et al., “Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries,” CA: a Cancer Journal for Clinicians, vol. 71, no. 3, pp. 209–249, 2021.
[3] B. C. Bade and C. S. Dela Cruz, “Lung cancer 2020: epidemiology, etiology, and prevention,” Clinics in Chest Medicine, vol. 41, no. 1, pp. 1–24, 2020.
[4] H. M. Zhang, B. Yang, H. F. Chen et al., “Prognosis-related miRNA bioinformatics screening of lung adenocarcinoma and its clinical significance,” Chinese Journal of Applied Physiology, vol. 34, no. 6, pp. 530–535, 2018.
[5] K. Inamura and Y. Ishikawa, “MicroRNA in lung cancer: novel biomarkers and potential tools for treatment,” Journal of Clinical Medicine, vol. 5, no. 3, p. 36, 2016.
[6] F. R. Hirsch, G. V. Scagliotti, J. L. Mulshine et al., “Lung cancer: current therapies and new targeted treatments,” Lancet, vol. 389, no. 10066, pp. 299–311, 2017.
[7] K. C. Arbour and G. J. Riel, “Systemic therapy for locally advanced and metastatic non-small cell lung cancer,” Journal of the American Medical Association, vol. 322, no. 8, pp. 764–774, 2019.
[8] D. Anusewicz, M. Orzechowska, and A. K. Bednarek, “Lung squamous cell carcinoma and lung adenocarcinoma differential gene expression regulation through pathways of notch, hedgehog, Wnt, and ErbB signalling,” Scientific Reports, vol. 10, no. 1, p. 21128, 2020.
[9] N. Robinson, R. Ganesan, C. Hegedűs, K. Kovács, T. A. Kufer, and L. Virág, “Programmed necrotic cell death of
macrophages: focus on pyroptosis, necroptosis, and parthanatos," Redox Biology, vol. 26, p. 101239, 2019.

[10] V. Nikoletopoulou, M. Markaki, K. Palikaras, and N. Tavernarakis, "Crosstalk between apoptosis, necrosis and autophagy," Biochimica et Biophysica Acta, vol. 1833, no. 12, pp. 3448–3459, 2013.

[11] P. Vandenabeele, L. Galluzzi, T. Vanden Berghe, and G. Kroemer, "Molecular mechanisms of necroptosis: an ordered cellular explosion," Nature Reviews. Molecular Cell Biology, vol. 11, no. 10, pp. 709–714, 2010.

[12] Y. Liu, Q. Chen, Y. Zhu et al., "Non-coding RNAs in necroptosis, pyroptosis and ferroptosis in cancer metastasis," Cell Death Discovery, vol. 7, no. 1, p. 210, 2021.

[13] J. E. Park, J. H. Lee, S. Y. Lee et al., "Expression of key regulatory genes in necroptosis and its effect on the prognosis in non-small cell lung cancer," Journal of Cancer, vol. 11, no. 18, pp. 5503–5510, 2020.

[14] H. Y. Tan, N. Wang, Y. T. Chan et al., "ID1 overexpression increases gefitinib sensitivity in non-small cell lung cancer by activating RIP3/MLKL-dependent necroptosis," Cancer Letters, vol. 475, pp. 109–118, 2020.

[15] M. Li, M. Pan, J. Wang et al., "miR-7 reduces breast cancer stem cell metastasis via inhibiting RELA to decrease ESAM expression," Molecular Therapy Oncolytics, vol. 18, pp. 70–82, 2020.

[16] H. Xiao, "MiR-7-5p suppresses tumor metastasis of non-small cell lung cancer by targeting NOVA2," Cellular & Molecular Biology Letters, vol. 24, no. 1, p. 60, 2019.

[17] U. Vösa, T. Vooder, R. Kolde, J. Vilo, A. Metspalu, and T. Annilo, "Meta-analysis of microRNA expression in lung cancer," International Journal of Cancer, vol. 132, no. 12, pp. 2884–2893, 2013.

[18] M. Cao, J. Cai, Y. Yuan et al., "A four-gene signature-derived risk score for glioblastoma: prospects for prognostic and response predictive analyses," Cancer Biology & Medicine, vol. 16, no. 3, pp. 595–605, 2019.

[19] X. Zhao and L. Cui, "A robust six-miRNA prognostic signature for head and neck squamous cell carcinoma," Journal of Cellular Physiology, vol. 235, no. 11, pp. 8799–8811, 2020.

[20] Z. Fu, B. Deng, Y. Liao et al., "The anti-tumor effect of shikonin on osteosarcoma by inducing RIP1 and RIP3 dependent necroptosis," BMC Cancer, vol. 13, no. 1, p. 580, 2013.

[21] K. E. Lawlor, N. Khan, A. Mildenhall et al., "RIPK3 promotes cell death and NLRP3 inflammasome activation in the absence of MLKL," Nature Communications, vol. 6, no. 1, p. 6282, 2015.

[22] K. Newton, "RIPK1 and RIPK3: critical regulators of inflammation and cell death," Trends in Cell Biology, vol. 25, no. 6, pp. 347–353, 2015.

[23] T. Baronsky, D. Ruhlandt, B. R. Brückner et al., "Cell-substrate dynamics of the epithelial-to-mesenchymal transition," Nano Letters, vol. 17, no. 5, pp. 3320–3326, 2017.

[24] E. Bonastre, E. Brambilla, and M. Sanchez-Cespedes, "Cell adhesion and polarity in squamous cell carcinoma of the lung," The Journal of Pathology, vol. 238, no. 5, pp. 606–616, 2016.

[25] I. Skerpagnias, D. Anastasakis, K. Grafanaki et al., "Contribution of miRNAs, tRNAs and tRFs to aberrant signaling and translation deregulation in lung cancer," Cancers, vol. 12, no. 10, p. 3056, 2020.

[26] C. H. Wang, X. F. Li, L. F. Jin, Y. Zhao, G. J. Zhu, and W. Z. Shen, "Dieckol inhibits non–small–cell lung cancer cell proliferation and migration by regulating the PI3K/AKT signaling pathway," Journal of Biochemical and Molecular Toxicology, vol. 33, no. 8, article e22346, 2019.

[27] W. Yang, W. Xiao, Z. Cai, S. Jin, and T. Li, "miR-1269b drives cisplatin resistance of human non-small cell lung cancer via modulating the PTEN/PI3K/AKT signaling pathway," Oncotargets and Therapy, vol. Volume 13, pp. 109–118, 2020.

[28] C. Liu, H. Li, J. Jia, X. Ruan, Y. Liu, and X. Zhang, "High metastasis-associated lung adenocarcinoma transcript 1 (MALAT1) expression promotes proliferation, migration, and invasion of non-small cell lung cancer via ERK/mitogen-activated protein kinase (MAPK) signaling pathway," Medical Science Monitor, vol. 25, pp. 5143–5149, 2019.

[29] J. Fan, D. Ren, J. Wang et al., "Brucine D induces lung cancer cell apoptosis and autophagy via the ROS/MAPK signaling pathway in vitro and in vivo," Cell Death & Disease, vol. 11, no. 2, p. 126, 2020.

[30] S. Li, Y. Feng, Y. Huang et al., "MiR-223-3p regulates cell viability, migration, invasion, and apoptosis of non-small cell lung cancer cells by targeting RHOB," Open Life Sciences, vol. 15, no. 1, pp. 389–399, 2020.

[31] C. Liu, Z. Yang, Z. Deng et al., "Upregulated IncRNA ADAMTS9-AS2 suppresses progression of lung cancer through inhibition of miR-223-3p and promotion of TGFBR3," IUBMB Life, vol. 70, no. 6, pp. 536–546, 2018.

[32] Y. Zhu, K. Li, L. Yan, Y. He, L. Wang, and L. Sheng, "miR-223-3p promotes cell proliferation and invasion by targeting Arid1a in gastric cancer," Acta Biochimica et Biophysica Sinica Shanghai, vol. 52, no. 2, pp. 150–159, 2020.

[33] J. S. Zhou, Z. S. Yang, S. Y. Cheng, J. H. Yu, C. J. Huang, and Q. Feng, "miRNA-425-5p enhances lung cancer growth via the PTEN/PI3K/AKT signaling axis," BMC Pulmonary Medicine, vol. 20, no. 1, p. 223, 2020.

[34] Z. Zhang, M. Wen, J. Guo et al., "Clinical value of miR-425-5p detection and its association with cell proliferation and apoptosis of gastric cancer," Pathology, Research and Practice, vol. 213, no. 8, pp. 929–937, 2017.

[35] W. Li, Y. Cui, D. Wang, Y. Wang, and L. Wang, "MiR-141-3p functions as a tumor suppressor through directly targeting ZFR in non-small cell lung cancer," Biochemical and Biophysical Research Communications, vol. 509, no. 3, pp. 647–656, 2019.

[36] Q. Xie, Z. Yu, Y. Lu, J. Fan, Y. Ni, and L. Ma, "microRNA-148a-3p inhibited the proliferation and epithelial–mesenchymal transition progression of non-small–cell lung cancer via modulating Ras/MAPK/Erk signaling," Journal of Cellular Physiolo, vol. 234, no. 8, pp. 12786–12799, 2019.

[37] Y. Bai, L. Lang, W. Zhao, and R. Niu, "Long non-coding RNA HOXA11-AS promotes non-small cell lung cancer tumorigenesis through microRNA-148a-3p/DNMT1 regulatory axis," Oncotargets and Therapy, vol. Volume 12, pp. 11195–11206, 2019.

[38] X. H. Liao, Z. Xie, and C. N. Guan, "MiRNA-500a-3p inhibits cell proliferation and invasion by targeting lymphocyte antigen 6 complex locus K (LY6K) in human non-small cell lung cancer," Neoplasma, vol. 65, no. 5, pp. 673–682, 2018.

[39] H. Li, M. Wang, H. Zhou, S. Lu, and B. Zhang, "<em>eBLN3P</em> promotes the progression of liver cancer via alteration of microRNA-144-3p/Dock4"
signal,” *Cancer Management and Research*, vol. Volume 12, pp. 9339–9349, 2020.

[40] Q. Q. Tian, J. Xia, X. Zhang, B. Q. Gao, and W. Wang, “miR-331-3p inhibits tumor cell proliferation, metastasis, invasion by targeting MLLT10 in non-small cell lung cancer,” *Cancer Management and Research*, vol. 12, pp. 5749–5758, 2020.

[41] A. A. Svoronos, D. M. Engelman, and F. J. Slack, “OncomiR or tumor suppressor? The duplicity of microRNAs in cancer,” *Cancer Research*, vol. 76, no. 13, pp. 3666–3670, 2016.