Lipase catalyzed epoxidation of fatty acid methyl esters derived from unsaturated vegetable oils in absence of carboxylic acid

Alejandro Sustaita-Rodríguez†, Víctor H. Ramos-Sánchez†, Alejandro A. Camacho-Dávila†, Gerardo Zaragoza-Galán†, José C. Espinoza-Hicks† and David Chávez-Flores*†

Abstract

Nowadays the industrial chemistry reactions rely on green technologies. Enzymes as lipases are increasing its use in diverse chemical processes. Epoxidized fatty acid methyl esters obtained from transesterification of vegetable oils have recently found applications as polymer plasticizer, agrochemical, cosmetics, pharmaceuticals and food additives. In this research article, grapeseed, avocado and olive oils naturally containing high percents of mono and poly unsaturations were used as starting materials for the production of unsaturated fatty acid methyl esters. The effect of lauric acid as an active oxygen carrier was studied on epoxidation reactions where unsaturated fatty acid methyl esters were converted to epoxy fatty acid methyl esters using immobilized Candida antarctica Lipase type B as catalyst and hydrogen peroxide as oxygen donor at mild temperature and pressure conditions. After this study it was confirmed by 1H NMR, 13C NMR and GC–MS that the addition of lauric acid to the enzymatic reaction is unnecessary to transform the alkenes in to epoxides. It was found that quantitative conversions were possible in despite of a carboxylic acid absence.

Keywords: Epoxidation, FAME, Lauric acid, Lipase

Introduction

In response to a serious concern over the depleting fossil reserves and the negative environmental impact caused by their products and processes, there has been a growing trend towards utilization of renewable feedstocks as a source of chemicals and materials [1]. Vegetable oils and the product of their transformations (fatty acids and alkyl esters) are important raw materials for producing a variety of products such coatings, paints, lubricants, soaps [2], and inks [3]. For more complex molecules such polymers, copolymers and their composites, the oils have to be chemically modified. One of the most interesting functional group for this purpose is the epoxy ring.

Epoxides are major key raw materials for various industrial products such as polymer plasticizer, agrochemical, cosmetics, pharmaceuticals and food additives. Epoxidation consists on the formation of an oxirane (epoxy) group by the reaction of peroxyacids (peracids) and olefinic double bonds [4]. Epoxides derived from vegetable oils are used as PVC stabilizers, plasticizers and in polyurethane production. They can also be useful as reactive diluents for paints, production of surfactants, corrosion protections agents and additives to lubricants. Using epoxidized FAMEs from unsaturated vegetable oils it is possible to obtain polymers and composites with better mechanical, electric, thermal properties than those of the polymers obtained from petrochemical products and greater resistance to oxidation than the latter ones [5–7].

During industrial epoxidation, the required peroxyacids are commonly produced by the reaction of acetic or formic acids and hydrogen peroxide in presence of strong mineral acids, such as H₂SO₄ and H₃PO₄ or by the direct
addition of any peroxycid which can cause equipment corrosion and promote undesirable oxirane ring-opening reactions [8, 9].

Recently, enzymatic epoxidation reactions have been better accepted compared with chemical synthesis due to the milder reactions conditions as formation of stable hydroperoxides directly from fatty acids, high stereoselectivity, significant suppression of side reactions and high conversion [10–12].

**Materials and methods**

Lipase B from *Candida antarctica* (Novozym 435) immobilized on macroporous polycrylate resin beds, chloroform-*,d*, cyclohexane, isooctane, dodecanoic acid (lauric acid) and pentadecanoic acid were acquire from Sigma-Aldrich. Potassium hydroxide, hydrogen peroxide 30% was purchased from J.T. Baker, toluene was acquired from Fermont, sodium hydroxide, potassium sulphate and anhydrous ethanol and methanol 99% were purchased Mallinckrodt. Vegetable oils (grapeseed, avocado and olive oils) were gotten from a local market (Additional file 1).

**Synthesis procedure**

All experiments on this report were made at least by triplicate. Synthesis of epoxides involved two steps: in the first stride, different vegetable oils (grapeseed, avocado and olive) were converted into methyl esters by transesterification reaction using KOH as catalyst and methanol as acyl acceptor.

In a second step 1 g of methyl esters, 100 mg of Novozym 435, 1 mmol of lauric acid and 6 mL of toluene were added into a 50 mL round bottom flask, and then 5 mL of distillate water was added to the reaction mixture to create a biphasic system which was heated to 45 °C and homogenized with stirring. Once the temperature was reached and kept for 5 min, slowly 1 mL of hydrogen peroxide was added to the reaction system and the reactions were stirred at this temperature for 16 h. Upon completion of reaction time, samples were cooled to 4 °C and Novozym 435 was recovery by vacuum filtration, using filter paper number 41. The filtered reaction mixture was firstly washed several times with distilled water and the separated organic layer was dried with anhydrous sodium sulphate for 1 h to remove trace water and decompose traces of unreacted peroxides. Finally, product was separated from solvent using a vacuum rotary evaporator at 90 °C for 1 h. Experiments were repeated in absence of lauric acid under the same conditions of reactions.

**Characterization of FAMEs and its epoxides**

Fatty acid methyl esters (FAMEs) and its epoxidized products were analysed by gas chromatography (Agilent series 7890B) coupled with mass spectrometer (Agilent series 5975C) in a HP-INNOWAX column, 30 m × 0.25 mm, 0.25 μm and by nuclear magnetic resonance trough 1H-NMR and 13C-NMR experiments using a Bruker 400 MHz magnet and deuterated chloroform solvent for sample preparation.

**Results and discussion**

**FAMEs characterization**

FAMEs from different vegetable oils were characterized by GC–MS, 1H-NMR and 13C-NMR analytical techniques. A proton NMR spectrum is shown on Figs. 1a, c, d and 2, in which can be observed the characteristic peak of methoxy protons as a singlet at 3.67 ppm. Also, is possible observe a triplet at 2.30 ppm that indicates presence of carbonyl α-CH₂ protons. These two peaks confirmed the presence of methyl esters and its integration areas confirms the conversion on each sample. Other peaks were observed at 0.88 ppm for terminal or deshielded methyl protons, 1.27 ppm for methylene protons ay the alkyl chain and a signal at 1.62 ppm was observed owing to β-carbonyl methylene protons. Protons on the unsaturation or double bonds were identified at 5.34 ppm.

FAMEs conversion (Table 1) was determined choosing relevant signals for integration (methoxy protons at 3.67 ppm and α-CH₂ protons at 2.30 ppm). The equation used to quantify the conversion was [13]:

$$C = \frac{2A_{Me}}{3A_{CH_2}} \times 100\%$$  (1)

where: C = percentage conversion of triglycerides to the corresponding methyl esters; $A_{Me}$ = integration value of the methoxy protons of the methyl esters; $A_{CH_2}$ = integration value of α-methylene protons.

Saturated fatty acid methyl ester (SFAME), monounsaturated fatty acid methyl ester (MUFAME) and polyunsaturated fatty acid methyl ester (PUFAME) content (Table 1) was calculated using the integral for different protons [14] and the equations are given below:

$$\text{PUFAME} = \frac{Z}{\text{Y}}$$  (2)

$$\text{MUFAME} = \left[ \frac{X}{\text{Y}} - \text{PUFAME} \right]$$  (3)

$$\text{SFAME} = \left[ 1 - \frac{X}{\text{Y}} \right]$$  (4)

**Effect of lauric acid in enzymatic epoxidation of FAMEs from vegetable oils**

Initial experiments were performed in presence of carboxylic acid as lauric acid because has been reported that the use of linear chain fatty acids favours the reaction conversion into the corresponding epoxide due to the fatty acids acts as precursor in the epoxide formation [15–17]. Originally FAMEs from different vegetable
Fig. 1 $^1$H-NMR spectrum of a avocado epoxide, b FAME avocado, c grapeseed epoxide, d FAME grapeseed
oils did not contain epoxy groups but after epoxidation reactions their presence could be identified with chemical shifts due to double bonds (~5 ppm). Figures 1b, d and 2b shows the disappearance of this peaks which means epoxidation was completely carried out (conversion >99%). A new signals at 2.90 ppm confirmed the existence of an epoxy ring corresponding to methine CH proton and at 1.50 ppm corresponding to the new methylene group alpha to the new epoxy group. Finally, signal at 1.63 ppm indicates a methylene group between the two new epoxy groups.

In this case, the use of lauric acid in the reaction system and its effect can be explained according to the proposed reaction mechanism shown in Fig. 3 (methyl oleate is used as example because of is the mainly component in FAMEs derived from olive and avocado oils while methyl linoleate corresponds to FAME derived from grapeseed oil) in which the enzyme catalyzed the formation of peracid from lauric acid and H₂O₂, then perlauric acid transfers the oxygen to the double bond in FAMEs to yield epoxystearic and diepoxystearic acid methyl ester respectively.

Experiments were also performed in absence of lauric acid in order to evaluate its effect in the epoxide formation. For these assays characteristics signals on the NMR spectra corresponding to epoxides were identified at 2.90, 1.50, and 1.63 ppm as in the previous experiments. Also, it was noticed the disappearance of peak belonging to double bonds (~5 ppm) where it can be inferred that conversion was more than 99% such as in the previous experiments. All ¹H NMR signals from FAME and epoxides of FAME matched perfectly with previous reports [18].

| Oil       | C (%) | SFAME (%) | MUFAME (%) | PUFAME (%) |
|-----------|-------|-----------|------------|------------|
| Olive     | 95.19 | 13.46     | 78.84      | 7.69       |
| Avocado   | 97.43 | 8.17      | 77.40      | 14.42      |
| Grapeseed | 96.74 | 14.67     | 19.92      | 65.41      |

In Fig. 2 ¹H-NMR spectrum of a olive epoxide and b FAME olive
Several researches have reported that addition of fatty acids in reaction system is crucial to carry out successful epoxidations but according to the results obtained from this paper it is possible to establish that lauric acid or any fatty acid is not critical or crucial to get high epoxidation or conversions. As a likely explanation, in Fig. 4 is shown the proposed reaction mechanism to describe the pathway in which enzymatic epoxidation without lauric acid is accomplished.

Reaction mechanism involves two steps: Firstly, saturated and insaturated methyl esters present in the mixture of FAMEs from vegetable oils react with $\text{H}_2\text{O}_2$ to form the corresponding peracid, secondly peracid spontaneously donates the oxygen to a double bond of the unsaturated FAMEs (methyl oleate and methyl linoleate are used as example) to yield epoxystearic and diepoxystearic acid methyl ester respectively.

As an alternative to the mechanism given in Fig. 4, it is proposed a hydrolysis of the ester bond as a side reaction (Fig. 5). In this case Novozym 435 catalyses the hydrolysis of ester to produce its corresponding acid which is oxidized by the enzyme itself to form a peracid. Finally, peracid donates the oxygen to double bond and epoxides are brought about. Therefore, the final product will be a mixture of epoxidized methyl esters and fatty acids [1].

GC/MS analysis was used to evaluate the resulting epoxides structures on proposed mechanism. The epoxy stearate (9,10-epoxy octadecanoate) was identified at RT: 33.69 min and the mass spectrum allowed to define its molecular structure. The molecular ion ($M^+$) was detectable at 312, the major fragments m/z 155 (M-157; loss of ($\text{CH}_2)_7$-COOCH$_3$) and 199 (M-113; loss of ($\text{CH}_2)_7$-CH$_3$) represent the result of cleavage at the epoxy ring [19].

The corresponding peak to diepoxystearate (9,10-12,13-diepoxy octadecanoate) appeared at RT: 37.38 min and the molecular ion ($M^+$) was perceptible at 326. The spectrum was similar at MS spectrum previous reported for the same compound [18, ..]
Characteristic peaks were identified at m/z 295 (M−31), 277 (M−49), 255 [CH₂CH(O)CHCH₂CH(O)CH(CH₂)₇COOCH₃−14]⁺, 237 [CH₂CH(O)CHCH₂CH(O)CH(CH₂)₇COOCH₃−32]⁺, and 211 [O=CHCH₂CH(O)CH(CH₂)₇COOCH₃−31]⁺.

**Conclusion**

It is possible to obtain epoxides fatty acid methyl esters by an enzymatic reaction under mild temperature and pressure conditions (45 °C, 1 atm., 250 rpm) and high selectivity by the use of Novozym 435 as biocatalyst and hydrogen peroxide as oxygen donor. The reactions were carried out with conversions higher than 99%. This study also demonstrates that the addition of a carboxylic acid is not essential or crucial to perform epoxidation reactions successfully due to the fact that the unsaturated FAMEs present in the mixture generated by enzymatic hydrolysis act as precursors in the epoxide formation.

**Additional file**

Additional file 1. Additional figures.

**Abbreviations**

FAMEs: fatty acid methyl esters; PVC: polyvinylchloride; SFAME: saturated fatty acid methyl ester; MUFAME: monounsaturated fatty acid methyl ester; PUFAME: polyunsaturated fatty acid methyl ester.

**Authors’ contributions**

ASR carried out the experimental work. DCF is the corresponding author and wrote most of the manuscript. VRS contributed to data analysis and interpretation. JCEH and ACD carried out the spectroscopic characterization and GZG carried out the chromatographic analysis. All authors read and approved the final manuscript.

**Acknowledgements**

We gratefully acknowledge the financial support of the Facultad de Ciencias Químicas of the Universidad Autónoma de Chihuahua and the National Council of Science and Technology of Mexico (CONACYT).

**Competing interests**

The authors declare that they have no competing interests.

**Availability of data and materials**

Not applicable.

**Ethics approval and consent to participate**

Not applicable.

**Funding**

Not applicable.
Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Received: 1 November 2017   Accepted: 30 March 2018
Published online: 11 April 2018

References
1. Orellana-Coca C, Ulrika T, Dietlind A, Bo M, Ranji H (2005) Chemo-enzymatic epoxidation of oleic acid and methyl oleate in solvent-free medium. Biocatal Biotransform 23(6):431–437
2. Hill K (2000) Fats and oils as oleochemical raw materials. Pure Appl Chem 72(7):1255–1264
3. Blayo A, Gandini A, Le Nest JF (2001) Chemical and rheological characterization of some vegetable oils derivatives commonly used in printing inks. Ind Crops Prod 14:155
4. Okieimen FE, Bakare OI, Okieimen CO (2002) Characterization of linseed oil epoxidized at different percentages. Ind Crops Prod 15:139
5. Biswas A, Adhvaryu A, Gordon SH, Erhan SZ, Willet JL (2005) Synthesis of diethylamine-functionalized soybean oil. J Agric Food Chem 53:9485–9490
6. La Scala J, Wool RP (2005) Property analysis of triglyceride-based thermosets. Polymer 46:61–69
7. Amoah J, Quayson E, Hama S, Yoshida A, Hasunuma T, Ogino C, Kondo A (2007) Simultaneous conversion of free fatty acids and triglycerides to biodiesel by immobilized Aspergillus oryzae expressing Fusarium heterosporum lipase. Biotechnol J 12:1–4
8. Petrovic ZS, Zlatanic A, Lava CC, Sinadinovic-Fiser S (2002) Epoxidation of soybean oil in toluene with peroxyacetic and peroxyformic acids-kinetics and side reactions. Eur J Lipid Sci Technol 104:293–299
9. Gerbase AE, Gregorio JR, Martinelli M, Brasil MC, Mendes ANF (2002) Epoxidation of soybean oil by the methyltrioxorhenium-CH3O2/CH2O2 catalytic biphasic system. J Am Oil Chem Soc 79:179–181
10. Lu H, Shangde S, Yanlan B, Goulong Y, Rulan M, Huifang Y (2010) Enzymatic epoxidation of soybean methyl esters in the presence of free fatty acids. Eur J Lipid Sci Technol 112:1101–1105
11. Furuhashi T, Nakamura T, Fragner L, Roustan V, Schön V, Weckwerth W (2016) Biodiesel and poly-unsaturated fatty acids production from algae and crop plants a rapid and comprehensive workflow for lipid analysis. Biotechnol J 10:262–267
12. Lotti M, Pleiss J, Valero F, Ferrer P (2015) Effects of methanol on lipases: molecular, kinetic and process issues in the production of biodiesel. Biotechnol J 10:22–30
13. Naureen R, Tariq M, Yusuffi, Chowdhury A, Ashraf A (2015) Synthesis, spectroscopic and chromatographic studies sunflower oil biodiesel using optimized base catalysed methanalysis. J Biol Sci 22:332–339
14. Usman A, Thoss V, Darko G, Itoho AU (2016) Determination of triacylglycerol composition of Trichilia emetica seed oil using GC–MS and 1H NMR spectroscopy. Adv Anal Chem 6(1):10–16
15. Zhang X, Wan X, Cao H, Dewil R, Deng L, Wang F, Tan T, Nie K (2017) Chemo-enzymatic epoxidation of Sapindus mukurrossi fatty acids catalyzed with Candida sp. 99–125 lipase in a solvent-free system. Ind Crops Prod 98:10–18
16. Wang X, Tang Q, Maria-Popowicz Q, Yang B, Wang Y (2015) A mechanistic study into the epoxidation of carboxylic acid and alkene in a mono, diacylglycerol lipase. Biochem Biophys Res Commun 460:392–396
17. Campanella A, Baltanas AB (2006) Degradation of the oxirane ring of epoxidized vegetable oils in liquid–liquid heterogeneous reaction systems. Chem Eng J 118:141–152
18. Xia W, Budge SM, Lumsden MD (2016) 1H-NMR characterization of epoxides derived from polyunsaturated fatty acids. J Am Oil Chem Soc 93:467–478
19. Kaneshiro ES, Ellis J, Guo Z, Jayaishamul K, Maiorano JN, Kallam KA (1996) Characterizations of neutral lipid fatty acids and cis-9,10-epoxy octadecanoic acid in Pneumocystis carinii carinii. Infect Immun 64:4105–4114
20. Piazza GJ, Nuñez A, Foglia TA (2003) Epoxidation of fatty acids, fatty methyl esters, and alkenes by immobilized oat seed peroxygenase. J Mol Catal B Enzym 21(3):143–151