Using Blue Mini-LEDs as a Light Source Designed a Miniaturized Optomechanical Device for the Detection of Direct Bilirubin

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Abstract
This study developed a miniaturized optomechanical device (MOD) for the feasibility study of direct bilirubin in urine using high-collimation blue mini-light-emitting diodes (Mini-LEDs) as the light source. The constructed MOD used optical spectroscopy to analyze different concentrations of direct bilirubin using the absorbance spectrum to achieve a noninvasive method for detection. The experimental results showed that between the absorbance and different concentrations of direct bilirubin at the blue Mini-LEDs central wavelength (462 nm) was the optimum fitting wavelength; in the direct bilirubin concentration range from 0.855 to 17.1 μmol/L, the coefficient of determination ($R^2$) was 0.9999, the limit of detection (LOD) of 0.171 μmol/L, and the limit of quantitation (LOQ) of 0.570 μmol/L. Therefore, we propose using blue Mini-LEDs as a light source to design a MOD to replace the invasive blood sampling method with a spectroscopic detection of direct bilirubin concentration corresponding to absorbance.

Keywords: Blue Mini-LEDs, Miniaturized optomechanical, Noninvasive, Direct bilirubin, Quantitative analysis, Coefficient of determination

Introduction
Bilirubin is a yellow pigment produced by the catabolism of hemoglobin in red blood cells and excreted from the body through urine and feces. Bilirubin is generally divided into three categories: direct bilirubin, indirect bilirubin, and total bilirubin (indirect plus direct bilirubin). Excess bilirubin causes the skin, eyes, and other tissues to turn yellowish-brown, a condition called jaundice [1–3]. Typically, full-term and premature babies have a 60% and 80% chance, respectively, of developing jaundice. As a result, bilirubin testing is a regular procedure performed on newborns [4, 5].

Jaundice occurring in newborns is generally classified as physiological or pathological jaundice. The majority of neonatal cases are physiological jaundice and do not have serious consequences. However, abnormally high bilirubin values can lead to neurotoxicity, and neurodevelopmental abnormalities, such as hearing loss, athetosis, and mental deficiencies [6–9]. Physiological jaundice in newborns typically appears on the second day of life, peaking on the fourth to fifth day, and then subsiding over a period of one to two weeks. The two main factors that account for pathological jaundice (cholestatic jaundice) are biliary atresia and neonatal hepatitis. In biliary atresia, early diagnosis and surgery are necessary to avoid the eventual need for a liver transplant. Neonatal jaundice lasting greater than two weeks, called delayed jaundice, should be investigated and further tests conducted to rule out pathological jaundice. When delayed jaundice is present in a newborn, the current recommendation of the American Academy of Pediatrics (AAP) is to draw blood for total and direct bilirubin testing to rule out the possibility of biliary atresia. The importance of indirect bilirubin
detection in diagnosing jaundice is minimal; only direct bilirubin is filtered by the kidneys and excreted into the urine. In addition, indirect bilirubin is insoluble in water. Therefore, the values obtained by urinary assessment of bilirubin are dominated by analysis of direct bilirubin [10–12]. In Taiwan, there is a stool color card in the child health handbook for parents to compare the color of their infant’s stool for early screening of biliary atresia. However, parents and health care workers have concerns about making errors when comparing infants’ stools with the stool color cards. Therefore, it is hoped that developing a noninvasive urine bilirubin test will provide them with accurate and immediate results.

Most bilirubin assays used in clinical chemistry laboratories are based on diazotization and oxidase methods [13, 14]. In a typical automated diazo colorimetric method, bilirubin and diazo ions are reacted under strongly acidic conditions of pH 1.7–2.0 to form a reddish colored azo compound. The fluorescence intensity of the azo compound is measured at 600 nm using a photometer, and the bilirubin concentration is determined [15]. Saifee et al. proposed a method that mixes normal and ultra-filtered plasma samples with O-desmethyl naproxen (ODMN) to detect total bilirubin. Their experimental results showed that total bilirubin’s absorption was in the wavelengths between 450 and 560 nm when combined with ODMN. After combining with the jaundice samples, the best absorption wavelength was found to be 520 nm [16]. Ponhong et al. proposed a hybrid analytical system for the determination of direct bilirubin in urine samples, which uses n-octyl-β-d-thioglucoside (OTG) as the solubilizer for bilirubin; when it reacts with diazotized sulfanilic acid, OTG-azobilirubin is formed. Analysis of the linear relationship between the direct bilirubin concentration and the OTG-azobilirubin found the coefficient of determination (R^2) was 0.994 in the concentration range of 0–1.0 mg/L, the detection limit was 4.7 μg/L, and the relative standard deviation value (RSD) was 1.9% [17]. Lano et al. proposed the analysis of total bilirubin concentration using a co-oxygenation saturation assay and Radiometer® ABL90; samples were centrifuged and analyzed for total bilirubin in plasma using diazo reagents. The mean deviation of the total bilirubin levels examined was −1.0 μmol/L, which is more precise than spectrophotometric methods with a mean deviation of −4.4 μmol/L [18].

Although the diazo assay for bilirubin is inexpensive and easy to automate, it is susceptible to hemolysis and lipid or albumin interference, causing a shift in the pH of the diazo reagent and unpredictable deviations in the determination of the bilirubin concentration. Therefore, the current diazo detection method for bilirubin can only be considered a qualitative analysis, as its reliability has not been able to reach the high accuracy standards required for quantitative analysis [19–21].

Bilirubin oxidase (BOD) is a multicopper oxidase (MCO), which uses metal ions to catalyze the oxidation of bilirubin to biliverdin, and is used in the determination of bilirubin concentration. BOD exhibits high activity and stability at neutral pH and is widely used as a test reagent [22–25]. Rachna et al. developed a biosensor by immobilization of BOD on graphene oxide nanoparticles. The biosensor showed a linear dependence for bilirubin concentration from 0.01 to 600 μmol/L and retained about 80% of its initial activity when repeatedly utilized, about 100 times, over 180 days. Compared with the standard colorimetric technique, the biosensor’s correlation of coefficient (r) was 0.99 [26]. Zheng et al. fabricated a novel electrochemical sensor based on Au nanoparticles/tetrathiafulvalene-carboxylate functionalized reduced graphene oxide 0D-2D heterojunction for the detection of bilirubin in human blood. The electrochemical sensor had a linear response range of 2.66 to 83 μmol/L for bilirubin, with a detection limit of 0.74 μmol/L [27]. Rahman et al. used Mn(II) ions and conductive polymers for the development of a current-based bilirubin biosensor, which was coated with a polyethyleneimine film of ascorbic acid oxidase; experimental data showed that the optimal linear range of bilirubin ranged from 0.1 to 50 μmol/L, with a detection limit of 40±3.8 nmol/L, with 3.8 nmol/L showing good stability and a fast response less than 5 s [28].

Although the oxidase method for the detection of bilirubin is widely used, the threshold of the technique is high, there is interference by substances, such as albumin or hemoglobin, and the separation of enzymes must be restricted to aqueous solvents. Therefore, there is much room for improvement to achieve quantitative detection [29–32].

Quantum dots are semiconductor nanostructure materials that modulate the radiation wavelength by changing the particle size and have wide absorption and narrow emission. Because quantum dots obtain the same full width at half maximum (FWHM) at different and consistent emission wavelengths, they are increasingly used in fluorescence detection of biological samples [33–35]. Jayasree et al. proposed a new type of biosensor using quantum dots with an emitted wavelength of 405 nm that chemically reacts with Fe^{3+} to quench blue fluorescence. The study showed that fluorescence was regained by adding bilirubin. Linear regression analysis was done using the ratio of fluorescence quenching and bilirubin concentration. The R^2 between the ratio of fluorescence quenching and bilirubin concentration was 0.99, and the limit of detection was 2.5 μmol/L. The data showed that the developed biosensor had good linearity of detection [36].
Zhao et al. reported a strategy to detect bilirubin using quantum dots modified with human serum albumin by cationic polymer polyethylenimine-induced aggregation, and then measuring the radioactive light intensity. This method was successfully applied to the quantitative determination of indirect bilirubin [37].

The fluorescence method detected purification bilirubin has a high linear correlation, but the detection method is dependent on chemical reactions. In addition, for biological samples with a complex composition, such as urine and blood, the anti-interference and linear correlation of fluorescence-based methods do not reach the expected level of accuracy [38–40]. Besides the methods above, the excess bilirubin in the blood will lead to the skin becoming yellow, and then, the method using diffuse reflectance spectroscopy to quantify bilirubin from skin reflectance spectra has been proposed [41]. There is also a noninvasive method by using a light beam incident on the nail and analyzing the reflected light through optical fibers based on diffuse reflection [42].

Compare LEDs with conventional light sources employed in analytical instruments, due to the narrow width of the LEDs emission spectrum, it is usually 30 nm which can improve the light utilization efficiency. The analytical application of LEDs is used in chromatography, absorption detection in capillaries, fluorescence spectrometry, micro-fluidic devices, photoacoustic spectroscopy [43].

The current noninvasive methods of bilirubin detection aim to quantify the total and direct bilirubin concentration. Because the blue wavelength of 462 nm is very close to the maximum absorbance wavelength of direct bilirubin. Therefore, our study used blue Mini-LEDs as a light source to design a miniaturized optomechanical device (MOD) to replace invasive blood sampling methods. The device used the spectroscopic detection of the direct bilirubin concentration corresponding to the absorbance. The device offers the benefits of being noninvasive, fast, and providing a digitally quantitative analysis.

**Materials and Methods**

**Absorbance Defined**

When a parallel light beam is directed at an absorbing substance, after passing through a certain thickness of the substance it absorbs some of the light energy, and the intensity of the transmitted light decreases. The derivation process of the Beer–Lambert law is shown in Eqs. 1–2:

\[
\text{Transmittance} = T = \frac{I_1}{I_0} \quad (1)
\]

\[
\text{Absorbance} = -\log_{10} T \quad (2)
\]

where \(T\) is the transmittance, \(A\) is the absorbance value, \(I_0\) is the intensity of incident light, and \(I_1\) is the intensity of transmitted light.

According to the Beer–Lambert law, the absorbance value is proportional to the concentration and thickness of the substance, meaning that when the absorbing material is more concentrated or thicker, because of the greater absorbance of the material, the reduction in light intensity is more pronounced, as shown in Eq. 3:

\[
A = \log_{10} \left( \frac{I_0}{I_1} \right) = \varepsilon \cdot \ell \cdot c \quad (3)
\]

where \(A\) is the absorbance value, \(\varepsilon\) is the absorbance coefficient of the substance, \(\ell\) is the thickness of the absorbing medium (cm), and \(c\) is the concentration of the absorbing substance (mg/dL).

Statistics commonly uses the \(R^2\) value to explain the correlation between the known data \(X\) (concentration) and the \(Y\) (absorbance) influenced by \(X\). The \(R^2\) value is expressed as a value between 0 and 1; the larger the value or closer to 1, the better the relationship for the linear regression. The \(R^2\) value can be calculated using Eq. 4 [44]:

\[
R^2 = \frac{\sum(Y_i - \bar{Y})^2}{\sum(Y_i - \bar{Y})^2} \quad (4)
\]

where \(\bar{Y}_i\) is the predicted value on the regression line, \(\bar{Y}\) is the average of all \(Y_i\), \((\bar{Y}_i - \bar{Y})^2\) is called the sum of squares for regression (SSReg), and \((Y_i - \bar{Y})^2\) is the sum of squares for total (SSTotal).

**Direct Bilirubin Sample**

Direct bilirubin powder (bilirubin conjugate, ditaurate, disodium salt – Calbiochem\textsuperscript{®}, Merck Millipore, Inc., USA, Catalog number 201102) is the standard currently used by medical institutions to simulate direct bilirubin. The product’s certificate of analysis was according to Regulation (EC) No. 1907/2006. The molecular formula and 3D structure of the direct bilirubin sample are shown in Fig. 1.

Direct bilirubin powder was mixed with deionized water to form 14 different standard solutions in the concentration range of 0.855–17.1 \(\mu\)mol/L. The weight of direct bilirubin and deionized water was measured using a precision scale and mixed uniformly by stirring at 500 rpm for 5 min with a magnetic stirrer.

**Design for a Mini-LEDs MOD**

The Hitachi U-3900 spectrometer has the disadvantage of being bulky and heavy to carry. This study used mini-LEDs as a light source to design a MOD, which consisted of mini-LEDs, a collimating lens, optical fiber, and a miniature spectrometer, as shown in Fig. 2.
A single mini-LED was fixed and driven on a breadboard. Use tape as a holder1 to hold the LEDs and optical fiber when they are centered. When light incident to an optical fiber with a numerical aperture of 0.22, fiber distribution panel of 200 µm, length of 1 m, a diameter of 3.2 mm and focused to a quartz cell that is filled with direct bilirubin through a collimating lens with a back focal length of 10 mm and a numerical aperture of 0.22. The light absorbed through the quartz cell was converted into parallel light by a reverse collimating lens and the incident light on the optical fiber. Finally, different concentrations of direct bilirubin were analyzed by a microspectrometer SE1030-025-FUVN (OtO Photonics, Inc., Hsinchu City, Taiwan). The quartz cell is fixed on the holder2, and the collimating lens is fixed on the holder2 by threads.

**Experimental Process**

The experimental procedure for the detection of direct bilirubin by the MOD developed in this study is shown in Fig. 3. Initially, 14 different concentrations of direct bilirubin standard solutions (0.855 to 17.1 μmol/L) were prepared. The absorbance of air was used as the baseline to zero-calibrate the spectrometer (Hitachi, Ltd., Tokyo, Japan, Model U-3900). After the calibration was complete, the direct bilirubin absorbance values of the standard solutions were measured, and the linear relationship between the absorbance and a standard solution’s concentration was analyzed to find the optimum fitting region. The measured band was then set to the identified optimum fitting region to analyze the relationship between direct bilirubin concentration and absorbance, and the data were analyzed using a binary linear regression; the $R^2$ value represented the linear relationship between direct bilirubin concentration and the absorption values. To conduct the reproducibility experiments, the developed MOD was rebuilt five times, and the average error of reproducibility was measured for each. Finally, the results of the constructed blue Mini-LEDs MODs and the Hitachi U-3900 spectrometer were compared to verify the developed MOD’s ability to detect direct bilirubin. The MOD experiments will fix the integration time of the microspectrometer and test in the dark room with a room temperature of 25 °C and humidity of 60%.

**Results and Discussion**

**Direct Bilirubin Absorbance Spectrum Analysis**

In this study, direct bilirubin was measured by the Hitachi U-3900 spectrometer first; the absorbance value was measured at a scan rate of 2 nm/s. Fourteen different concentrations of direct bilirubin were prepared, and their
absorbance values measured in the concentration range of 0.855–17.1 μmol/L. Figure 4 shows the absorbance spectra of the 14 different concentrations.

The absorbance spectra of direct bilirubin determined that the maximum absorbance wavelength was 448 nm, the linear regression is $y = 0.0276x + 0.0288$, and the $R^2$ reached 0.9978. The optimum fitting wavelength is 460 nm, and the linear regression is $y = 0.0265x + 0.0279$ with $R^2$ value of 0.998 as shown in Fig. 5.

**Detecting Direct Bilirubin Using Blue Mini-LEDs as a Light Source for a MOD**

This study used Blue Mini-LEDs (Harvatek corporation, Inc., Hsinchu City, Taiwan) as a light source; the chip size length, width, and height were 101.6, 152.4, and 150 μm, respectively. The package size was 800, 800, and 300 μm in length, width, and height respectively. The blue Mini-LEDs package fabrication process is shown in Fig. 6 and includes chip die bonding, molding with white glue, cutting the white glue, and separating it from the glass.

The emission spectrum of the blue Mini-LEDs is shown in Fig. 7. They have a maximum peak at wavelength 462 nm, an optimum fitting range between 390 and 490 nm band light source, and the FWHM of the spectrum is 36 nm.

The blue Mini-LEDs were driven at 2.8 V and 40 mA. The direct bilirubin was measured in the wavelength range of 430–500 nm as this was the optimum fitting
range after analysis. Figure 8 shows the absorbance spectrum of different concentrations (0.855 to 17.1 μmol/L) of direct bilirubin measured by the blue Mini-LEDs MOD.

The blue Mini-LEDs MOD measurement of direct bilirubin transmission spectrum from concentration 0.855 to 17.1 μmol/L has the best linearity relationship in the band 430 to 500 nm, with the \( R^2 \) value reaching above 0.99. Figure 9 shows the optimum fitting wavelength was 462 nm, the linear regression is \( y = 0.0385x + 0.0253 \), the \( R^2 \) value is 0.9999, LOD of 0.171 μmol/L, and LOQ of 0.570 μmol/L.

To conduct a reproducibility test, the blue Mini-LEDs MOD was rebuilt five times and each was used to measure three different concentrations of direct bilirubin at 462 nm; the highest concentration was 17.1 μmol/L, the medium concentration was 8.55 μmol/L, and the low concentration was 1.71 μmol/L. The experimental results are shown in Table 1. The standard deviation error \( (\sigma/\sqrt{n}) \), where \( \sigma \) is the standard deviation, and \( n \) is the number of samples) of the light penetration intensity at wavelength 462 nm of the 17.1, 8.55, and 1.71 μmol/L direct bilirubin was 0.073, 0.068, and 0.140%, respectively, and the average standard deviation error was 0.093%. According to the experimental results, the developed blue Mini-LEDs MOD has excellent reproducibility in the repeated setup and measurement of direct bilirubin.

The long-term stability test for MOD to detect direct bilirubin of 8.55 μmol/L was tested in a dark room with a room temperature of 25 °C and humidity of 60% as shown in Fig. 10. Besides, we also fix the integration time of the microspectrometer to 1 ms. After driven the LEDs for 1, 10, 30, 60 min, the absorbance for the direct bilirubin concentration of 8.55 μmol/L was 0.35615, 0.35364, 0.35479, 0.35278, respectively, and the standard deviation error was 0.07%.
Comparing Detection Fitting Between the Blue Mini-LEDs MOD and a Commercial Spectrometer

Figure 11 shows the $R^2$ measured by blue Mini-LEDs MOD and Hitachi U-3900 spectrometer in direct bilirubin (0.855 to 17.1 μmol/L) at the wavelength of 462 nm and 460 nm. The blue Mini-LEDs MOD developed in this study was compared with the Hitachi U-3900 spectrometer measured at a wavelength of 462 nm and 460 nm with different concentrations and absorbance, and the linear regression and $R^2$ values are $y = 0.0385x + 0.0253$, 0.9999 and $y = 0.0265x + 0.0279$, 0.998, respectively.

Conclusions

The experimental data showed that the blue Mini-LEDs MOD developed in this research had a good linear relationship in detecting direct bilirubin when compared with the Hitachi U-3900 spectrometer's analysis results. Therefore, the developed MOD could simplify the noninvasive quantitative detection of direct bilirubin.

In this study, a noninvasive spectroscopic detection method was proposed to detect direct bilirubin. Blue Mini-LEDs MODs and a Hitachi U-3900 spectrometer were used to analyze the relationship between direct bilirubin at different concentrations and the light absorbance change in the spectrum. The direct bilirubin concentration corresponds to absorbance and absorbance values, and linear regression analysis can provide a tool to achieve a noninvasive quantitative analysis. The experimental results show that the blue Mini-LEDs MOD's optimal fitting band of direct bilirubin at 14 different concentrations is 462 nm, in the direct bilirubin concentration range from 0.855 to 17.1 μmol/L, the linear regression was $y = 0.0385x + 0.0253$, the $R^2$ is 0.9999, LOD of 0.171 μmol/L, LOQ of 0.570 μmol/L and the average standard deviation error of measurement reproducibility is 0.093%. Comparing the blue Mini-LEDs MOD at 462 nm and the Hitachi U-3900 spectrometer at 460 nm, detect different concentrations of direct bilirubin, the linear regression and $R^2$ values are $y = 0.0385x + 0.0253$, 0.9999 and $y = 0.0265x + 0.0279$, 0.998, respectively. Therefore, the blue Mini-LEDs MODs have the advantage of small size and portability which can be used in the future as a noninvasive quantitative detection device for direct bilirubin measurement.

Table 1 The reproducibility measurement of the blue Mini-LEDs MOD

| Direct bilirubin concentration (μmol/L) | The absorbance for the first test (arb. u) | The absorbance for a second test (arb. u) | The absorbance for the third test (arb. u) | The absorbance for the fourth test (arb. u) | The absorbance for the fifth test (arb. u) | Standard deviation error (%) |
|----------------------------------------|--------------------------------------------|--------------------------------------------|--------------------------------------------|--------------------------------------------|--------------------------------------------|-----------------------------|
| 1.71                                   | 0.680                                      | 0.674                                      | 0.674                                      | 0.673                                      | 0.672                                      | 0.140                        |
| 8.55                                   | 0.354                                      | 0.352                                      | 0.351                                      | 0.351                                      | 0.350                                      | 0.068                        |
| 17.1                                   | 0.090                                      | 0.089                                      | 0.089                                      | 0.087                                      | 0.086                                      | 0.073                        |

Fig. 10 The absorbance for the direct bilirubin after driving the LEDs for 1, 10, 30, and 60 min

Fig. 11 The $R^2$ value of direct bilirubin absorbance at 462 nm and 460 nm by the blue Mini-LEDs MOD and the Hitachi U-3900 spectrometer
Abbreviations
MOD: Miniaturized optomechanical device; Mini-LEDs: Mini-light-emitting diodes; LOD: Limit of detection; LOQ: Limit of quantitation; FWHM: Full width at half maximum; AAP: American Academy of Pediatrics; ODMN: O-desmethyl-naproxen; RSD: Relative standard deviation value; BOD: Bilirubin oxidase; MCO: Multicopper oxidase.

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Author contributions
Z.T.Ye and H.C. Ku designed the experiments; Z.T.Ye, S.F. Tseng, and S.X. Tsou analyzed the data; Z.T.Ye, S.R. Chung and H.C. Ku discussed the results; and Z.T.Ye and S. F. Tseng contributed to the writing of the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials
The data presented in this study are available on request from the all authors.

Declarations

Ethics approval and consent to participate
Not applicable.

Consent for publication
All authors consent to the publication of this manuscript.

Competing interests
The authors declare that they have no competing interests.

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References
1. Huizebos CV, Vitek I, Zabeta CD, Dvořák A, Schenk P, Eline AEH, Cobbaert C, Tinbelli C (2021) Diagnostic methods for neonatal hyperbilirubinemia: benefits, limitations, requirements, and novel developments. Pediatr Res 90(2):277–283. https://doi.org/10.1038/s41390-021-01546-y
2. Harpavat S, Garcia-Prats JA, Anaya C, Brandt ML, Lupo PJ, Finegold MJ, Obuobi A, ElHennawy AA, Janriel WS, Shneider BL (2020) Diagnostic yield of newborn screening for biliary atresia using direct or conjugated bilirubin measurements. JAMA 323(12):1141–1150. https://doi.org/10.1001/jama.2020.0837
3. Liu Y, Li M, Song Y, Liu X, Zhao J, Deng B, Peng A, Qin L (2018) Association of serum bilirubin with renal outcomes in Han Chinese patients with chronic kidney disease. Clin Chim Acta 480:9–16. https://doi.org/10.1016/j.cca.2018.01.041
4. Afltheros CE, Wennberg RP, Ostrow JD, Tinbelli C (2009) Unbound (free) bilirubin: improving the paradigm for evaluating neonatal jaundice. Clin Chem 55(7):1288–1299. https://doi.org/10.1373/clinchem.2008.121269
5. Bell KG, Moussa M, Abd El-Rahman MK, Tan EW, Homer-VanSicklen SM, Whitesides GM (2019) Paper-based potentiometric sensing of free bilirubin in blood serum. Biosens Bioelectron 126:115–121. https://doi.org/10.1016/j.bios.2018.10.055
6. Petta S, Sebastiani G, Viganò M, Ampuero J, Wong VWS, Bournier J, Berzigotti A, Bugianesi E, Fracanzani AL, Camma C, Enea M, Grottes MD, Marocco VD, Younes R, Keyrouz A, Mazzola S, Mendoza Y, Pennisi G, Romerogomez M, Cravi A, Victor de Ledinghen (2021) Monitoring occurrence of liver-related events and survival by transient elastography in patients with nonalcoholic fatty liver disease and compensated advanced chronic liver disease. Clin Gastroenterol Hepatol. https://doi.org/10.1016/j.cgh.2020.06.045
7. Boshchaart N, Kok JH, Newsum AM, Ouweneel DM, Mentink R, Leeuwen TG, Alders MCG (2012) Limitations and opportunities of transcutaneous bilirubin measurement. Pediatrics 129(4):689–694. https://doi.org/10.1542/peds.2011-2586
8. Deneny PA, Seidman DS, Stevenson DK (2001) Drug therapy: neonatal hyperbilirubinemia. N Engl J Med 344(8):581–590. https://doi.org/10.1056/NEJM200102223440807
9. Oluwasanya BO, Kaplan M, Hansen TWR (2018) Neonatal hyperbilirubinemia: a global perspective. Lancet Child Adolesc Health 2(9):610–620. https://doi.org/10.1016/S2352-4642(18)30139-1
10. Hou JJ, Liu J, Fan YF, Zheng HJ, Zhao HY, Yang JM, Yan JM, Ma Y, Liu X, Li J, Jia XN, Chen PS (2020) High prevalence of breastmilk-acquired cytomegalovirus infection in jaundiced infants. J Clin Lab Anal 34(2):e23199. https://doi.org/10.1002/jcla.23199
11. Reismi PE, Raveendran J, Suneehs PV, Ramanchandran T, Nair BG, Satheesh Babu TG (2021) Screen-printed carbon electrode for the electrochemical detection of conjugated bilirubin. Elsevier Mater Lett. https://doi.org/10.1016/j.matlet.2021.130574
12. Moyer V, Freese DK, Whittington PF, Olson AD, Brewer F, Colletti RB, Heyman MB (2004) Guideline for the evaluation of cholestatic jaundice in infants: recommendations of the North American Society for Pediatric Gastroenterology, Hepatology and Nutrition. J Pediatr Gastroenterol Nutr 39(115–128. https://doi.org/10.1097/00005768-200408000-00001
13. Ellairaaja S, Shenbagavalli S, Ponnnariyappan S, Vasantha VS (2017) A green and facile approach for synthesizing imine to develop optical biosensor for wide range detection of bilirubin in human biofluids. Biosens Bioelectron 91:82–88. https://doi.org/10.1016/j.bios.2016.12.026
14. Singha SS, Mondal S, Bhattacharya TS, Das L, Sen K, Satpark B, Dase K, Singhaha A (2018) Au nanoparticles functionalized 3D-MoS2 nanoflower: an efficient SERS matrix for biomolecule sensing. Biosens Bioelectron 119:10–17. https://doi.org/10.1016/j.bios.2018.07.061
15. Dhungana N, Morris C, Krassowski MD (2017) Operational impact of using a vanadate oxidase method for direct bilirubin measurements at an academic medical center clinical laboratory. Pract Lab Med 8:77–85. https://doi.org/10.1016/j.plabm.2017.05.004
16. Saifee NH, Ranjitkar P, Greene DN (2016) Factors influencing naproxen metabolism interference in total bilirubin assays. Clin Biochem 49(6):514–517. https://doi.org/10.1016/j.clinbiochem.2015.11.010
17. Ponhong K, Teshima N, Grudpan K, Motomizu S, Sakai T (2011) Simultaneous injection effective mixing analysis system for the determination of direct bilirubin in urinary samples. Talanta 87:113–117. https://doi.org/10.1016/j.talanta.2010.09.048
18. Lano IM, Lyon AW, Wang L, Ruskin R, Lyon ME (2018) Comparative evaluation of neonatal bilirubin using radiometer whole blood co-oximetry and plasma bilirubin methods from Roche diagnostics and ortho clinical diagnostics. Clin Biochem 53:88–92. https://doi.org/10.1016/j.clinbiochem.2017.12.009
19. Devgun MS, Richardson C (2016) Direct bilirubin in clinical practice—interpretation and haemolysis interference guidance reassessed. Clin Biochem 49(18):1351–1353. https://doi.org/10.1016/j.clinbiochem.2016.09.005
20. Fevity J (2008) Bilirubin in clinical practice: a review. Liver Int 28(S):S92–603. https://doi.org/10.1111/j.1478-3231.2008.01716.x
21. Kazmierczak SC, Robertson AF, Catrous PG, Blirey KP, Keamer BL, Gourley GR (2002) Direct spectrophotometric method for measurement of bilirubin in newborns: comparison with HPLC and an automated diazo method. Clin Chem 48(7):1096–1097. https://doi.org/10.1093/clinchem/hvf709
22. Mano N (2012) Features and applications of bilirubin oxidases. Appl Microbiol Biotechnol 96(2):301–307. https://doi.org/10.1007/s00253-012-4312-9
23. Rawal R, Kharangarh PR, Dawra S, Bhardwaj P (2021) Synthesis, characterization and immobilization of bilirubin oxidase nanoparticles (BOxNPs).
with enhanced activity. Application for serum bilirubin determination in jaundice patients. Enzyme Microb Technol 143:109716. https://doi.org/10.1016/j.enzmctec.2020.109716

24. Mano N, Edembé L (2013) Bilirubin oxidases in bioelectrochemistry: features and recent findings. Biosens Bioelectron 50:478–485. https://doi.org/10.1016/j.bios.2013.07.014

25. Thangavel B, Lingagounder J, Berchmans S, Ganesh V (2021) Electrochemical grafting of 4-amino benzoic acid onto toray carbon-interfacial features and recent findings. Biosens Bioelectron 50:478–485. https://doi.org/10.1016/j.bios.2021.130292

26. Rachna R, Nidhi C, Monika T, Vinay G (2017) A contrivance based on electrochemical integration of graphene oxide nanoparticles/nickel nanoparticles for bilirubin biosensing. Biochem Eng J. https://doi.org/10.1016/j.bej.2017.06.006

27. Zheng Z, Feng Q, Zhu M, Shang J, Li M, Li C, Kou L, Zheng J, Wang C (2019) Electrochemical sensor for the discrimination of bilirubin in real human blood based on Au nanoparticles/tetrathiafulvalene-carboxylate functionalized reduced graphene oxide 0D–2D heterojunction. Anal Chim Acta 1072:46–53. https://doi.org/10.1016/j.aca.2019.04.040

28. Rahman MA, Lee KS, Park DS, Won MS, Shim YB (2008) An amperometric bilirubin biosensor based on a conductive poly-terthiophene–Mn(II) complex. Biosens Bioelectron 23(6):857–864. https://doi.org/10.1016/j.bios.2007.09.005

29. Okada H, Kawada K, Itoh S, Ozaki M, Kakutani I, Arai T, Koyano K, Yasuda S, Iwase T, Murao K (2018) Effects of bilirubin photoisomers on the measurement of direct bilirubin by the bilirubin oxidase method. Ann Clin Biochem 55(2):276–280. https://doi.org/10.1177/0004563217716474

30. Ouyang L, Yao L, Tang R, Yang X, Zhu L (2021) Biomimetic point-of-care testing of trace free bilirubin in serum by using glucose selective capture and surface-enhanced Raman spectroscopy. Sens Actuators B Chem. https://doi.org/10.1016/j.snb.2021.129941

31. Gu Y, Li X, Lv X, Jia Q (2017) High-Sensitive and selective sensing of free bilirubin using metal-organic frameworks-based energy transfer process. ACS Appl Mater Interfaces 9(36):30925–30932. https://doi.org/10.1021/acsami.7b09091

32. Pan X, Li LH, Lin HD, Tan JY, Wang HT, Liao ML, Chen CJ, Shan BB, Chen YF, Li M (2019) A graphene oxide–gold nanostar hybrid based-paper biosensor for label-free SERS detection of serum bilirubin for diagnosis of jaundice. Biosens Bioelectron. https://doi.org/10.1016/j.bios.2019.111713

33. Lin KB, Xing J, Quan L, de Arquer FPG, Gong XW, Lu JX, Xie LQ, Zhao W, Zhang D, Yan CZ, Li WQ, Lu Y, Liu Y, Kimman J, Sargent EH, Xiong QH, Wei ZH (2018) Perovskite light-emitting diodes with external quantum efficiency exceeding 20 percent. Nature 562(7726):245–248. https://doi.org/10.1038/s41586-018-0575-3

34. Liu JJ, Geng YJ, Li DW, Yao H, Huo ZP, Li YF, Zhang K, Zhu SJ, Wei HT, Xu WQ, Jiang JI, Yang B (2020) Deep red emissive carbonized polymer dots with unprecedented narrow full width at half maximum. Adv Mater 32(17):e1906641. https://doi.org/10.1002/adma.201906641

35. Yuan FL, Yuan T, Sui LZ, Wang ZB, Xi ZF, Li XJ, Zhan QQ, Gai HW (2018) Engineering triangular carbon quantum dots with unprecedented narrow bandwidth emission for multicolored LEDs. Nat Commun 9(1):2249. https://doi.org/10.1038/s41467-018-04635-5

36. Jayasree M, Aparna RS, Anjana RR, Devi JSA, John N, Abha K, Manikandan A, George S (2018) Fluorescence turn on detection of bilirubin using Fe(III) modulated BSA stabilized copper nanocluster: a mechanistic perception. Anal Chim Acta 1031:152–160. https://doi.org/10.1016/j.aca.2018.05.026

37. Zhao WF, Zong CH, Lei T, Tian WT, Su MW, Li XJ, Zhan QQ, Gai HW (2018) Ultrasensitive free bilirubin detection in whole blood via counting quantum dots aggregates at single nanoparticle level. Sens Actuators B Chem 275:95–100. https://doi.org/10.1016/j.snb.2018.08.032

38. Ahmad E, Mondal A, Sarkar A, Chakraborty S, Lohar S, Saha NC, Dhara K, Chattopadhyay P (2020) Bilirubin quantification in human blood serum by deoxygenation reaction switch-triggered fluorescent probe. ACS Appl Bio Mater 3(7):4074–4080. https://doi.org/10.1021/acsabm.0c00170

39. Yeh JT, Nam K, Yeh JT, Perminov N (2017) eUnaG: a new ligand-inducible fluorescent reporter to detect drug transporter activity in live cells. Sci Rep 7:41619. https://doi.org/10.1038/srep41619

40. Athar H, Ahmad N, Tayyab S (1999) Use of fluorescence enhancement technique to study bilirubin–albumin interaction. Int J Biol Macromol 25:353–358. https://doi.org/10.1016/S0141-8130(99)00056-2

41. Alla SK, Hudddle A, Butler JD, Bowman PS, Clark JF, Beyette FR (2011) Point-of-care device for quantification of bilirubin in skin tissue. IEEE Trans Biomed Eng. https://doi.org/10.1109/tbme.2010.2093132

42. Halder A, Banerjee M, Singh S, Adhikari A, Sarkar PK, Bhattacharya AM, Chakraborti P, Bhattacharyya D, Mallick AK (2019) Pal SK (2019) A novel whole spectrum-based non-invasive screening device for neonatal hyperbilirubinemia. IEEE J Biomed Health Inform 23:2347–2353. https://doi.org/10.1109/jbhi.2019.2892946

43. Bui DA, Hauser PC (2015) Analytical devices based on light-emitting diodes—a review of the state-of-the-art. Anal Chim Acta 853:46–58. https://doi.org/10.1016/j.aca.2014.09.044

44. Lee SH, Goddard ME, Wray NR, Visscher PM (2012) A better coefficient of determination for genetic profile analysis. Genet Epidemiol. https://doi.org/10.1002/gepi.21614

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