Identification of the novel localization of tenascinX in the monkey choroid plexus and comparison with the mouse

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Tenascin-X (Tn-X) belongs to the tenascin family of glycoproteins and has been reported to be significantly associated with schizophrenia in a single nucleotide polymorphism analysis in humans. This finding indicates an important role of Tn-X in the central nervous system (CNS). However, details of Tn-X localization are not clear in the primate CNS. Using immunohistochemical techniques, we found novel localizations of Tn-X in the interstitial connective tissue and around blood vessels in the choroid plexus (CP) in macaque monkeys. To verify the reliability of Tn-X localization, we compared the Tn-X localization with the tenascin-C (Tn-C) localization in corresponding regions using neighbouring sections. Localization of Tn-C was not observed in CP. This result indicated consistently restricted localization of Tn-X in CP. Comparative investigations using mouse tissues showed equivalent results. Our observations provide possible insight into specific roles of Tn-X in CP for mammalian CNS function.

Key words: tenascin-X, choroid plexus, monkey, mouse, Ehlers–Danlos syndrome, schizophrenia.

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ous system (CNS), Tn-X mRNA expression has only been shown in the rat meninges of the olfactory bulb (Deckner et al., 2000). Recently, we found novel Tn-X localizations in the adult mouse leptomeninges trabecula in the cerebral cortex and in the connective tissue in the lateral ventricle choroid plexus (CP; Imura and Sato, 2008). Our finding of Tn-X localization in CP, which produces cerebrospinal fluid (CSF), might be a key factor in the investigation of the association between CSF metabolism and enlarged ventricles in schizophrenia. Enlarged ventricles are typical structural abnormalities associated with schizophrenia (Staal et al., 1999). Furthermore, CP secretes biologically active molecules into the CSF for brain development, activity and protection (Strazielle and Gherzi-Egea, 2000; Brown et al., 2004; Thouvenot et al., 2006; Johanson et al., 2008). In these molecules, for instance, there is a brain-derived neurotrophic factor (BDNF), the gene expression level and polymorphism of which have been analysed in relation to the pathogenesis of schizophrenia (Buckley et al., 2007). One study reported that BDNF is able to stimulate Tn-X expression in vitro (Takeda et al., 2005).

The validity and limitations of animal models (rodents and monkeys) for use in the study of schizophrenia have been discussed (Tordjman et al., 2007). The authors concluded that monkeys appear to be an interesting social interaction model, more so than rodents, because of their complex well-organized social structure. In addition to differences in social structure, the dopaminergic system of rats and monkeys is quite different (García-Cabezas et al., 2009), and dysfunction of the dopaminergic system is related to schizophrenia (Wang et al., 2008).

The CSF outflow system has been studied in some animal models (Kapoor et al., 2008). An anatomical difference in arachnoid granulations has been shown between rodents and monkeys (Krisch, 1988). Arachnoid granulations in monkeys are structurally similar to those in humans (Cooper, 1958; Krisch, 1988). In contrast, arachnoid granulations in rodents are similar to those of cats and dogs (Krisch, 1988). It is possible that Tn-X localization in CP is different between rodents and monkeys.

Therefore, details concerning Tn-X localization in monkey CP need to be clarified. In the present study, we compared the immunohistochemistry of Tn-X in monkey CP with that in mouse CP. Subsequently, to verify the reliability of Tn-X localization, we compared it with Tn-C localization in corresponding regions using neighbouring sections.

**Materials and Methods**

Two male monkeys (Macaca mulatta) weighing 3.3 and 8.0 kg and five male ICR mice (11 weeks of age) were used as experimental animals in this study. The monkey tissues were a gift of Dr Kathleen Rockland (RIKEN Brain Science Institute). This study was conducted in accordance with the official Japanese regulations for research on animals (according to institutionally approved protocols; RIKEN Institute and the Nippon Dental University, approval no. 07-04) and in accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals (NIH Publication no. 80-23, revised 1996). The animals were perfused with 4% paraformaldehyde in 0.1 M phosphate buffered saline after being deeply anaesthetized with nembutal as previously reported (Imura and Rockland, 2007; Imura and Sato, 2008). The monkey CP was carefully dissected from the lateral and fourth ventricles. Mouse brains were removed from the skull. The tissues were rapidly frozen by immersing them in a cold acetone solution with dry ice and were cut transversely or horizontally at 30 µm with a cryostat. We used a previously described immunohistochemical to analyse Tn-X and Tn-C (Imura and Sato, 2008). Briefly, the sections were processed for either Tn-X or Tn-C single immunoperoxidase staining (goat polyclonal anti-Tn-X antibody, catalogue no. sc-5498 or anti-Tn-C antibody, catalogue no. sc-9871; Santa Cruz Biotechnology, Santa Cruz, CA). The sections were incubated for 2 days at 4°C. Finally, the sections were incubated with biotinylated anti-goat secondary rabbit antibody (Vector Laboratories, Burlingame, CA) and visualized using avidin-biotin complex conjugated horseradish peroxidase (ABC Elite kits, Vector Laboratories) with diaminobenzidine. Routine controls for immunostaining were used during these experiments, by omission of the two primary antibodies. Photographs were taken as digital images with a digital camera (VB7010; Keyence,
Figure 1 Immunohistochemistry (IHC) of tenascin-X (Tn-X) and tenascin-C (Tn-C) in monkey choroid plexus (CP) in fourth ventricle. (A) Low magnification of IHC of Tn-X in the lateral recess of fourth ventricle. Two boxed regions are re-photographed at higher magnification in B and D. (B, D) Tn-X positive fibrous structures are clearly observed within connective tissue of CP (arrows). (C, E) IHC of the Tn-C in neighboring sections to the B and D. Tn-C positive fibrous structures are not detected within the connective tissue of CP. CC: cerebellar cortex, EC: epithelial cell, MO: medulla oblongata. Scale bars=500 µm in A, 50 µm in B-E.
Japan) equipped with a microscope (DM-RE; Leica Microsystems, Germany). The images were adjusted for contrast and/or brightness to match the real image using standard image software (Adobe Photoshop 5.5, Adobe, San Jose, CA).

Results

We observed Tn-X and Tn-C localizations in the fourth and lateral ventricles of monkey CP (Figures 1 and 2A, B). Tn-X- positive (Tn-X+) fibrous structures were clearly observed in CP (Figures 1 and 2A, B) of both ventricles. The Tn-X+ fibrous structures were restricted to the interstitial connective tissue in CP (Figures 1 and 2A, B). In neighbouring sections, Tn-X+ fibrous structures were clearly confined to the connective tissue and did not colocalize with Tn-C in CP (Figure 1B, E). Tn-X was notably distributed around the blood vessels in CP. Dense Tn-X+ fibrils surrounded blood vessels in CP (Figure 2A, B). The presence of Tn-X+ and Tn-C+ was confirmed in some blood vessels in the parenchyma of the cerebellum and medulla oblongata (Figure 2C, E). These results were consistent with previous reports of vascular localization of Tn-X and Tn-C (Matsumoto et al., 1994; Hasegawa et al., 1997; Ballard et al., 2006). In addition, a leptomeningeal Tn-X+ trabecular structure was also observed in the cerebellum (Figure 2C). In contrast, no leptomeningeal localization of Tn-C was detected (Figure 2D). The Tn-X+ trabecula might correspond to the leptomeninges trabecula in the subarachnoid space (Haines, 1991; Imura and Sato, 2008). These observations in monkeys were similar to those of our previous findings of Tn-X localization in mouse CP of lateral ventricles and

Figure 2. Tenascin-X (Tn-X) positive vascular and leptomeningeal structures in monkey choroid plexus (CP), cerebellar cortex (CC), and medulla oblongata (MO). (A, B) Tn-X positive fibrous structures are clearly observed around blood vessels (asterisks) and within the connective tissue of CP in lateral ventricle (arrowheads). (C) Leptomeninges on the CC is strongly Tn-X positive (arrowheads). An arrow indicates Tn-X positive blood vessel. (D) Immunohistochemistry of tenasin-C in a neighboring section to the C. Tn-C positive leptomeninges is not observed in the CC (arrowheads). An arrow indicates Tn-C positive blood vessel, which is corresponding to the Tn-X positive blood vessel in C (arrow). E: Tn-X positive blood vessels are also observed in MO (arrows). EC: epithelial cell. Scale bars=50 µm.
leptomeninges (Imura and Sato, 2008).

In the fourth ventricle of mouse CP, Tn-X+ fibrils were restricted to the interstitial connective tissue and leptomeninges in the cerebellum, and no localization of Tn-C was observed (Figure 3). These findings in the fourth ventricle were homologous in the monkeys described above. Tn-X localization in CP was consistently confirmed using immunohistochemical techniques in the lateral and fourth ventricles of monkeys and mice.

**Discussion**

We observed Tn-X localization in monkey CP in the present study. These findings were not previously reported in primates.

In mouse CP, we confirmed our previous reports of Tn-X localization in the lateral and fourth ventricles (Imura and Sato, 2008; present study). Tn-X localization in both species was confined to the interstitial connective tissue in CP. Tn-X directly binds to type I collagen and regulates type VI collagen for collagen fibrillogenesis (Mao et al., 2002; Minamitani et al., 2004). Collagen fibril in the CP connective tissue has been clearly observed in monkeys (Ling 1981). Furthermore, the connective tissue of CP consists of type I collagen in mice (Masuda et al. 1998) and of type VI collagen in humans (Roggendorf et al., 1988). Because of the presence of collagen fibres and the role of Tn-X in collagen fibrillogenesis, Tn-X localization in CP has a functional role, although

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**Figure 3.** Immunohistochemistry (IHC) of the tenascin-X (Tn-X) and tenascin-C (Tn-C) in mouse choroid plexus (CP) in fourth ventricle. (A) Low magnification of IHC of Tn-X in a coronal section at fourth ventricle level. One boxed region is re-photographed at higher magnification in B. (B, D) Tn-X positive fibrous structures are clearly observed within connective tissue of CP. Three arrowheads in B indicate Tn-X positive leptomeninges on PFl (paraflocculus). The panel D is photographed from another section. (C, E) IHC of the Tn C in neighboring sections to the B and D. Tn C positive fibrous structures are not detected within the connective tissue of CP and leptomeninges. 4V: fourth ventricle, CC: cerebellar cortex, EC: epithelial cell, LR4V: lateral recess of 4V, MO: medulla oblongata. Scale bars=500 μm in A, 50 μm in B-E.
the type of collagen in monkey CP is unknown.

Tn-X deficiency is consistent with structural defects in human collagen. Patients with the connective tissue disorder EDS have been found to lack Tn-X (Burch et al., 1997); incomplete collagen fiber bundles are responsible for the symptoms. In addition to skin symptoms, vascular fragility and an abnormal perivascular structure have also been observed in EDS patients (Burch et al. 1997). In mouse and human blood vessels, Tn-X was confirmed by immunohistochemistry (Matsumoto et al. 1994; Hasegawa et al. 1997). Type VI collagen was observed in CP blood vessels in humans (Roggendorf et al., 1988). These previous reports might support our finding of dense Tn-X localization around blood vessels in CP (Figure 2).

Recently, Tn-X was shown to be significantly associated with schizophrenia in a single nucleotide polymorphism analysis in humans (Wei and Hemmings, 2004; Tochigi et al., 2007). Previous reports and our present findings possibly indicate a novel important role of Tn-X in the CNS. Tn-X interacts with vascular endothelial growth factor B (VEGF-B) and BDNF in endothelial cells and enhances the function of VEGF-B in endothelial cell proliferation (Ikuta et al., 2000), and Tn-X expression is stimulated by BDNF (Takeda et al., 2005). In the nervous system, VEGF-B stimulates neurogenesis in the adult brain (Sun et al., 2006), and BDNF signalling is impaired in patients with schizophrenia (Buckley et al., 2007). The secretion of BDNF in CP (Borlongan et al., 2004) and the strong expression of VEGF-B in CP of mouse embryo (Aase et al., 1999) suggest the possibility of an interaction between Tn-X and these two molecules in CP. To clarify this hypothesized interaction of Tn-X in CP, further investigation is needed in future studies.

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