Alginate based antimicrobial hydrogels formed by integrating Diels–Alder “click chemistry” and the thiol–ene reaction

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In recent years medical devices manufacturers have been looking for antimicrobial coatings which are biocompatible and non-toxic for a wide range of medical devices. The demand for these antimicrobial coatings has increased significantly, owing to the increased incidence of hospital-associated infections (HAIs). Hydrogels have been widely used in biomedical applications due to their hydrophilicity, biodegradability, non-toxicity and biocompatibility. In this work, sodium alginate (SA) based antibacterial hydrogels SA/PEG–HH10 were designed and prepared by combining Diels–Alder (DA) click chemistry and the thiol–ene reaction. The hydrogels were first prepared using DA click chemistry with good mechanical strength, then the cysteine-terminated antimicrobial peptide HH10 was grafted into the hydrogel by the thiol–ene reaction between the oxy-norbornene group and the thiol group. The results showed that the antimicrobial hydrogels had a strong antibacterial property and good biocompatibility. Therefore, the antimicrobial hydrogels have significant potential application as coatings for implantable medical devices.

Introduction

Bacteria can easily adhere and gather on the surface of implantable medical devices and cause bacterial infection.1 Bacterial infection may induce inflammation and complications, and in the worst case, even cause implant failure.2–5 Once the failure has occurred, the implant needs to be removed and replaced with a new one, significantly increasing the patients’ pain and the costs of hospitalization.6–9 However, conventional antibiotic treatment often fails due to the low level of antibiotic at the site of infection. Furthermore, over prescription and misuse of conventional antibiotics have led to escalating drug-resistance associated with various bacteria. Therefore, there is a pressing need to develop new antimicrobials and relevant formulations to address these issues.

Hydrogels are three-dimensional networks which are formed by vastly hydrophilic macromolecules.10 Due to the unique properties of hydrogels, such as extracellular matrix-like water content, adequate viscoelastic, and desirable biocompatibility, they make excellent candidates for implantable medical device coatings.11 As coatings for implantable medical devices, the hydrogels not only need the above mentioned performance characteristics, but more importantly, good antimicrobial properties to ensure the success rate of implanted devices.11–16

For this purpose, the common strategies are blending or grafting antibacterial particles or groups into hydrogels.15 Employing hydrogels as supports to load the antibacterial particles (typically gold or silver), antibiotics or antimicrobial agents was the first methodology used to obtain antimicrobial hydrogels. However, the application of this approach is limited due to the cytotoxicity of nanoparticles and the short-term antimicrobial activity.17–19 The second methodology is chemical grafting of antimicrobial agents onto hydrogels. This approach can enhance the antimicrobial efficacy and specificity, reduce their cytotoxicity, and prolong their biostability and biocompatibility.20

Due to widespread resistance to conventional antibiotics, there is increasing interest in developing novel antimicrobials as an alternative antibacterial therapy. The broad-spectrum bactericidal activity of antimicrobial peptides (AMPs) makes them a promising candidate for therapeutic use. AMPs are small molecules that form an important part of the innate immune system. These small molecules are composed of 10–50 amino-acid residues and they show activities against a wide variety of bacterial, viral, fungal and protozoan infections.21,22 So far, several combinations of surfaces modified with AMPs as the active antibiotic have been reported.21–23 However, it is still
a challenge to graft AMPs into biomedical hydrogels with good mechanical strength, strong antibacterial properties and good biocompatibility.

Diels–Alder (DA) click chemistry, a selective [4 + 2] cycloaddition between a diene and a dienophile, can be used as a chemical cross-linking reaction for hydrogels, in particular due to its moderate reaction conditions, thermal reversibility, high efficiency and not requiring a catalyst. Consequently, the hydrogels formed by the DA reaction have played a significant role in biomedical fields. In our previous studies, we developed multifunctional hydrogels with elasticity, injectability, spatiotemporal patterning and tissue adhesive abilities based on DA click chemistry for cartilage tissue repair. In addition, the cyclohexene derivative (oxy-norbornene group) formed by DA click chemistry between furyl and maleimide can go on to efficiently react with the thiol group by the thiol–ene reaction, which can be used to further functionalize hydrogels. For instance, through integrating DA click chemistry and the thiol–ene reaction, we successfully introduced a RGDSC molecule. Then, the thiol-functionalized antimicrobial peptide HHC10 (HHC10–CYS) with a high activity against multidrug-resistant pathogens was selected to react in conjunction with the oxy-norbornene group of the hydrogel. This kind of antimicrobial hydrogel, integrated by DA click chemistry and the thiol–ene reaction, with good mechanical, strong antibacterial properties and desirable biocompatibility has significant potential application for use as a coating for implantable medical devices.

Materials and methods

Materials

2-Hydroxy-4-(2-hydroxyethoxy)-2-methylpropionophenone (I2959) and sodium alginate (SA, CP) were purchased from Aladdin. 2-Morpholinoethane sulfonic acid (MES), 4-{4,6-dimethoxy-1,3,5-triazin-2-yl}-4-methylmorpholiniumchloride (DMTMM, CP) and furylamine (furan) were purchased from Sigma-Aldrich (Guangzhou, China). Dimaleimide poly(ethylene glycol) (mal–PEG–mal) \( (M_w = 2 \times 10^4) \) was purchased from Shanghai Sunway Pharmaceutical Technology Co., Ltd (Shanghai, China). HHC10–CYS antimicrobial peptide (H-crkwwkwirw-NH2, 95%, water soluble) was purchased from Hefei peptide Biotechnology Co., Ltd. All other reagents and solvents were of analytical grade and used as received.

Methods

Synthesis of SA-furan

SA-furan was obtained by amidation between the carboxyl groups of SA and the amine groups of furan, as shown in Scheme 1. Briefly, SA (1 g, 5.05 mmol carboxyl groups) was dissolved in 250 ml of MES buffer (100 mM), then DMTMM (0.990 g, 2.5 mmol; 1.397 g, 5 mmol; 2.235 g, 7.5 mmol) was added to activate the polysaccharide carboxyl groups. After stirring for 1 h, furan (343 \( \mu l \), 2.5 mmol (SA-furan-1); 515 \( \mu l \), 5 mmol (SA-furan-2); 772 \( \mu l \), 7.5 mmol (SA-furan-3)) was added dropwise and the mixture was stirred at room temperature for 24 h. The reaction products with different degrees of substitution (DS) were dialyzed against distilled water for 5 days \( (M_w \text{ cut-off } 3500 \text{ Da}) \). Finally, the water was removed by lyophilization to obtain SA-furan as a white floccule.

The formation of SA/PEG hydrogels

Three different degrees of substitution of SA-furan (SA-furan-1, SA-furan-2 and SA-furan-3) were separately dissolved in distilled water with a concentration of 1.5% w/v. The mal–PEG–mal (the molar ratio of furly to maleimide was controlled at 1:1) was added into the solutions for 15 min and injected into the mold to form SA/PEG hydrogels \( \text{(SA/PEG-DS1, SA/PEG-DS2 and SA/PEG-DS3) at } 37 \degree \text{C, separately. Shown in Scheme 2.} \)

The morphology and gelation time of SA/PEG hydrogels

The morphology of the freeze-dried SA/PEG hydrogels \( \text{(SA/PEG-DS1, SA/PEG-DS2 and SA/PEG-DS3)} \) was observed using a scanning electron microscope (SEM) (Quanta 200, Netherland FEI). The gelation time of the hydrogels was obtained at 37 \degree \text{C and 50 \degree \text{C using a test tube inverting method as described in our previous work.} \)
**Mechanical properties**

A dynamic mechanical analyzer (DMA Q800 USA) with unconstrained compression mode was used to measure the compressive modulus of the SA/PEG hydrogels (SA/PEG-DS1, SA/PEG-DS2 and SA/PEG-DS3) (cut into cylinders of height 5 mm and diameter 10 mm). The linear ramp force of 1 N min$^{-1}$ up to 18 N was designed to obtain the breaking strength of the hydrogel. The strain from 5 to 10% was used to determine the value of the compressive modulus.

**Synthesis of the antimicrobial hydrogels SA/PEG–HHC10**

The synthesis the antimicrobial hydrogels SA/PEG–HHC10 is shown in Scheme 3. Different concentrations of HHC10–CYS antimicrobial peptides were dissolved in MES buffer to prepare a series of concentration solutions (1 mg ml$^{-1}$ (HHC10-I), 2 mg ml$^{-1}$ (HHC10-II), 3 mg ml$^{-1}$ (HHC10-III), 4 mg ml$^{-1}$ (HHC10-IV)). Then, the SA/PEG-DS2 hydrogels with the best mechanical properties in the group were immersed in the HHC10–CYS solution (each sample in 1 ml HHC10–CYS solution) for 24 h, then a photoinitiator I2959 was added and the hydrogels were exposed to 365 nm UV light for 10 min (5 min for each side). These were then washed with PBS buffer to remove the unreacted HHC-10 peptide to achieve SA/PEG–HHC10 antimicrobial hydrogel (SA/PEG–HHC-I, SA/PEG–HHC-II, SA/PEG–HHC-III and SA/PEG–HHC-IV). All the experiments were carried out under aseptic conditions.

**Organic elemental quantitative analysis of SA/PEG–HHC10**

SA/PEG–HHC10 hydrogels (SA/PEG–HHC-I, SA/PEG–HHC-II, SA/PEG–HHC-III and SA/PEG–HHC-IV) were freeze-dried and

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**Scheme 2** Formation of SA/PEG hydrogels.

**Scheme 3** Synthesis of SA/PEG–HHC10 antimicrobial hydrogels.
organic elemental quantitative analysis was conducted using an elemental analyzer (vario EL cube, Elementar), the N, S, C elements were measured and the extent of conjugation of HHC-10 antimicrobial peptide to the hydrogel was calculated. The mass of HHC10 in SA/PEG–HHC10 ($m_1$) can be calculated using the element conservation formula as follows:

$$m_1 = \left( m_{\text{Hgel}} \times S_{\text{Hgel}} \right) / S_{\text{H}}$$

where $m_{\text{Hgel}}$ is the mass of the SA/PEG–HHC10 hydrogels, $S_{\text{Hgel}}$ and $S_{\text{H}}$ is the S content of the SA/PEG–HHC10 hydrogels and HHC10–CYS, respectively.

Antibacterial assay of the SA/PEG–HHC10 hydrogel

The SA/PEG–HHC10 hydrogel samples (SA/PEG–HHC-I, SA/PEG–HHC-II, SA/PEG–HHC-III and SA/PEG–HHC-IV) group and blank group (without hydrogel sample) were cultured with 1 ml of the 1 × 10⁷ CFU ml⁻¹ *Escherichia coli* bacterial stock solution in a 48-well plate for 24 h at 37 °C. After culturing for 24 h, the cultured solutions were diluted 10⁴ times, 10 µl of 1 ml diluted solution was coated on the agarose culture plate. All agarose culture plates were cultured at 37 °C, with 5% CO₂ for 10 h to achieve a bacterial colony. The number of CFU in the blank group ($N_0$) and SA/PEG–HHC10 hydrogel samples groups ($N_x$) was obtained by the flat colony counting method. The sterilization rate ($ST_x$) was calculated using the following equation:

$$ST_x = \left( N_0 - N_x \right) / N_0 \times 100\%$$

Biocompatibility assay of SA/PEG–HHC10

Freeze-dried SA/PEG–HHC10 hydrogels (SA/PEG–HHC-I, SA/PEG–HHC-II, SA/PEG–HHC-III and SA/PEG–HHC-IV) were put in a 48-pore plate after radiation sterilization, and the sterile PBS (500 µl) was added, then the extra PBS was removed before adding Dulbecco’s modified Eagle’s medium (Gibco) (DMEM) with 10% fetal bovine serum, the hydrogels were immersed for 24 h. The human skin fibroblasts (HSF) were seeded onto the surface of the hydrogels with an inoculum density of 2 × 10⁴ CFU cm⁻². Cell viability was revealed by using the live–dead assay (live–dead assay Kit (Invitrogen)) after culture for 1 and 3 days.

Results and discussion

Characterization of SA-furan

SA-furan was obtained by coupling furfurylamine to SA using DMTMM reagent (Scheme 1). The chemical structure of SA-furan was confirmed by ¹H-NMR (Fig. 1). The peaks at 6.26, 6.46 and 7.51 ppm refer to the protons of furan, these peaks show that the furan group is successfully grafted to SA. The DS of SA-furan was calculated from the C and N content according to the elemental analysis, the calculation of DS was deduced as follows:

$$\frac{(M_{1C} + DS \times M_{2C})/(DS \times M_{2N})}{C\% / N\%} = \frac{(M_{1C} + DS \times M_{2C})}{(DS \times M_{2N})} = C\% / N\%$$

where $M_{2N}$ is the N mass of furan, $M_{1C}$ and $M_{2C}$ are the C mass of the SA and furan, respectively.

The DS values for SA-furan were 24.71 ± 1.21%, 35.52 ± 1.21%, and 44.63 ± 1.32% (Table 1) respectively.

Gelation time and morphology of the SA/PEG hydrogels

SA/PEG hydrogels with three different cross-linking densities were prepared (SA/PEG–DS1, SA/PEG–DS2, SA/PEG–DS3). The gelation time of the SA/PEG hydrogels at 37 and 50 °C is shown in Table 2. From Table 2 we can see that with the same DS, the gelation time of 50 °C was shorter than that of 37 °C, which can be explained by the fact that the DA reaction is thermally induced, the reaction rate is faster when the temperature is higher. Compared with 50 °C, the gelation time more sharply
decreased with increasing DS at 37 °C, this indicated that the DS was less influenced by the gelation time when the temperature is higher.

In order to reveal the relationship between the DS and network structure of the SA/PEG hydrogels, the microstructure of the freeze-dried hydrogel was observed using SEM micrographs. As can be seen from A, B and C of Fig. 2, the freeze-dried scaffolds exhibited a uniform pore size and good connectivity. The average pore size of SA/PEG-DS1, SA/PEG-DS2 and SA/PEG-DS3 were 165, 109 and 85 μm, respectively. It can be concluded that the pore size of the hydrogels decreased with an increase of DS, which was caused by the increasing of the crosslinking density of the hydrogels.

**Swelling properties of the SA/PEG hydrogels**

The ESR is an important parameter for the use of hydrogels in biomaterials. It was obtained by recording the water absorption of the hydrogels over time until it reached equilibrium. As shown in Fig. 3, the ESR of SA/PEG-DS1 was 45 g g⁻¹, SA/PEG-DS2 was 40 g g⁻¹ and SA/PEG-DS1 was 33 g g⁻¹. From these results we can see that an increase in the cross-linker concentration led to a decrease in the swelling capacity of the hydrogels. This behavior could have contributed to the increase in the concentration of the hydrophilic cross-linkers within the hydrogel matrix and a tough cross-linked network was obtained.

**Mechanical properties of the SA/PEG hydrogels**

Hydrogel samples were tested for their mechanical properties under compression. Typical compressive stress–strain curves of SA/PEG hydrogels with different DS are shown in Fig. 4. With the increasing of DS, the mechanical property increased and then decreased. The SA/PEG-DS2 had the highest value of stress-at-failure (33.01 ± 4.50 kPa) and compress modulus (52.53 ± 3.16 kPa). This is because the increasing of the crosslinking density of the SA/PEG hydrogels play a major role in the changing of the mechanical properties. When the proportion of flexible molecular chain (PEG) is increased, the increase of flexible segments of the SA/PEG hydrogels, instead of the crosslinking density, plays the main role in the changing of the mechanical properties.

**Mass of HHC10 in SA/PEG–HHC10 hydrogels**

The sulfur element (S) content of HHC10–CYS and SA/PEG–HHC10 antibacterial hydrogels were proceeded by organic elemental quantitative analysis. As shown in Table 4, with the increase of soaking concentration, the mass of HHC10 increased firstly and then decreased. The reasons might be the change of hydrogel network size. In the experiment we found that the equilibrium volume increased firstly and then decreased, which is consistent with the mass of HHC10 in the SA/PEG–HHC10 hydrogels (Table 4). The diffusion of peptides depends on the size of the hydrogel network, when the soaking
concentration increased to 4 mg ml⁻¹, the hydrogels became shrunken, the size of the hydrogel network became smaller than other groups, which caused a decrease in the mass of HHC10 in the SA/PEG–HHC10-IV hydrogel.

Antibacterial activity of the SA/PEG–HHC10 hydrogels
An ideal implantable medical devices coating should possess antimicrobial properties against infection. The antibacterial activity of the SA/PEG–HHC10 hydrogels against E. coli is evaluated in Fig. 5. After culture for 24 h, the sterilization rate of SA/PEG–HHC10-I was 98.5%, SA/PEG–HHC10-IV was 99.5% and both SA/PEG–HHC10-II and SA/PEG–HHC10-III were 100%. These results were in accordance with the mass of HHC10 in the SA/PEG–HHC10 hydrogels (Table 4) and show the strong antibacterial activity of the SA/PEG–HHC10 hydrogels.

Cytocompatibility of the SA/PEG–HHC10 hydrogel
Cytocompatibility of the SA/PEG–HHC10 hydrogel was assessed using a live-dead assay that stains live cells green (calcein AM) and dead cells red (ethidium homodimer). As shown in Fig. 6(A and B), with the extension of the incubation time, the number of HSF cells on the SA/PEG–HHC10 became more and the morphology of the cells became exclusively a fusiform shape. The results showed that the SA/PEG–HHC10 hydrogel had good cytocompatibility.

**Conflicts of interest**

There are no conflicts to declare.

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