Bio-sourced phosphoprotein-based synthesis of silver-doped macroporous zinc phosphates and their antibacterial properties†

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The usual sources of phosphorus for metal phosphates are obtained from phosphate rocks, of which resources are depleted. As a substitute for these mineral sources, an original method of synthesis has been developed to prepare macroporous zinc phosphates using casein phosphoprotein. This bio-sourced reactant plays during the synthesis the roles of both a phosphorus source and a reducing agent for silver nanoparticles. Thus, zinc phosphates loaded with different Ag contents (up to 6.4 wt%) are prepared via hydrothermal treatment at 100 °C. Silver nanoparticles co-crystallized with hopeite, Zn3(PO4)2 and/or Zn2P2O7. In addition, casein induces porosity within the zinc phosphate framework and provides macropores (diameter of >50 nm) during calcination. The antibacterial properties against Escherichia coli K12 bacteria of Ag-containing and Ag-free porous zinc phosphates (calcined at 750 °C) were also tested for the first time.

Phosphorus sources for the synthesis of these materials are derived from phosphate rock resources, principally sedimentary marine phosphorites.15 Approximately 82% of these phosphate rocks are exploited for the preparation of fertilizer, a few per cent for animal feed, and the rest for industrial purposes.16,17 Because of the importance of phosphorus and the constant demand for it, the level of exploitation of phosphate rocks would reach a peak in 2033 and 2034.17 Total resource exhaustion at the current rate of world consumption is expected to occur in a few hundred years.15,17 Orthophosphoric acid is the main product of the treatment of phosphate rocks. It is also the main phosphorus source used for the synthesis of metal phosphates.18 Hence, the depletion of stocks of phosphate rocks will have a direct impact on the supply of metal phosphates. Therefore, the development of bio-sourced syntheses of metal phosphates is particularly relevant.

The development of silver-supported solids, including metal phosphates, is of constant interest for biomedical and environmental applications owing to the antimicrobial activity of Ag+ ions against different Gram-positive and Gram-negative strains of bacteria.9,19 The bactericidal activity of Ag+ ions is attributed to a combination of different biocidal actions that lead to the death of bacteria, which include degradation of the bacterial cell wall by the interaction of Ag+ ions with the membrane and inactivation of protein synthesis and DNA replication.9,20,21 For this purpose, the incorporation of Ag nanoparticles into materials is an efficient method of providing antibacterial properties in an aqueous medium by allowing the release of Ag+ ions from oxidized Ag nanoparticles. When applied to porous zinc phosphates, doping with Ag
nanoparticles may significantly improve the antibacterial properties of the final material. In addition, porosity may bring added value by enhancing the retention of bacteria for their further degradation.

Furthermore, the use of inorganic matrices as supports for Ag nanoparticles has attracted interest because of their advantages in terms of stability, heat resistance and safety in comparison with organic compounds. Studies have shown that inorganic structures such as zeolites, zinc oxide, titanium dioxide, phosphates, ceramics and glass are suitable carriers for the release of Ag⁺ ions. In this context, several studies of the synthesis of metal phosphate materials loaded with Ag nanoparticles, which mainly focused on hydroxyapatite (calcium phosphate), have been reported. The incorporation of Ag nanoparticles into hydroxyapatite coatings has shown good results in the prevention of implant-associated bacterial infections, which is attractive in medical applications owing to the biocompatible properties of hydroxyapatite. Environmental applications of hydroxyapatite have also been reported. An example is the development of a filter material composed of porous hydroxyapatite loaded with Ag nanoparticles as a substitute for the typical zeolite/Ag, sand/Ag, fiberglass/Ag and resin/Ag filters used in water disinfection.

Porosity in metal phosphate materials is usually generated by organic molecules such as quaternary ammonium compounds or amines, of which the supply depends on oil resources. They are usually highly toxic and environmentally unfriendly. The use of new templates obtained from new raw materials represents one of the major challenges in the synthesis of porous solids. Few studies of the synthesis of bio-sourced porous and nonporous metal phosphates are available in the literature, except for the synthesis of calcium phosphate. Bio-sourced zinc phosphates were reported for the first time in a recent work published by our group. The present paper describes the first synthesis of bio-sourced porous zinc phosphates loaded with Ag nanoparticles with a study of their antibacterial properties. Casein phosphoprotein (CN) from bovine milk is used. It is composed of four principal types of caseins (i.e., αs1-, αs2-, β- and κ-casein), in which phosphorus is present as colloidal calcium phosphate (CCP) (inorganic phosphorus) and phosphoserine residues (SerP) (organic phosphorus). These are in the form of dispersed spherical colloidal particles known as micelles (diameter: 50–500 nm, average: 150 nm). Casein plays the role of a phosphorus source, generates porosity and permits the in situ synthesis of Ag nanoparticles without the use of any additional reducing agents such as commonly used hydrazine and sodium borohydride.

2. Experimental section

2.1. Preparation of zinc phosphates

A casein dispersion was prepared by adding 9.50 g (0.71 wt% of P) of casein powder from bovine milk (Sigma-Aldrich, technical grade) to 190 mL of a 3.0 mmol L⁻¹ aqueous solution of sodium hydroxide (pH 11.5) (NaOH pellets, Sigma-Aldrich, ≥97%) under magnetic stirring for 30 min. A 0.12 mol L⁻¹ aqueous solution of silver nitrate (AgNO₃, Sigma-Aldrich, ≥99%) was then added to the casein dispersion under stirring for 20 min at room temperature. Then, 32 mL of a 0.35 mol L⁻¹ aqueous solution of zinc nitrate (Zn(NO₃)₂·6H₂O, Alfa Aesar, ≥99%) was added to the prepared casein/silver ion dispersion, and finally the entire reaction mixture was maintained under magnetic stirring for 20 min at room temperature. The resulting solution was transferred to a 500 mL polypropylene flask and stored at 100 °C. After heating for 6 days, the product was filtered off, washed with distilled water and dried at 70 °C overnight. All chemicals and reagents were used as received without further purification.

2.1.1. Calcination conditions. The as-synthesized samples were calcined in a muffle furnace at different temperatures ranging from 250 °C to 750 °C using a heating rate of 5 °C min⁻¹. They were cooled to room temperature (20 ± 5 °C) before analysis. The samples were named as ZnPOs/Ag-250, ZnPOs/Ag-500, ZnPOs/Ag-600 and ZnPOs/Ag-750 for the calcined samples that contained silver and ZnPOs-250, ZnPOs-500, ZnPOs-600 and ZnPOs-750 for the calcined samples without silver (reference samples).

2.1.2. Structural and textural characterization of zinc phosphates. The crystallinity of the samples was characterized by powder X-ray diffraction (XRD). The XRD patterns of the as-synthesized samples and calcined samples were recorded using PANalytical X’Pert and STOE STADI P diffractometers, respectively. These were operated with Cu Kα radiation (λ = 1.5406 Å) for 2θ values of 3–70° with steps of 0.02 or 0.28° s⁻¹ at room temperature. X’Pert HighScore software was used to identify the crystalline phases present in the samples by comparison of the experimental diffraction patterns with reference patterns provided by the International Center for Diffraction Data (ICDD) (i.e., PDF-4+ powder diffraction files). The composition of the samples was determined by inductively coupled plasma atomic emission spectrometry (ICP-AES). The morphology of the samples was determined by scanning electron microscopy (SEM) using a Philips XL30 microscope under a vacuum of 10⁻⁵ mbar. Samples were dispersed on a support
covered with a graphite conductive tab and then coated with a thin film of gold (10 nm to 20 nm, deposited using a Bal-Tec 
SCD 004 sputter coater). Micrographs were obtained with an 
accelerating voltage of 7 kV at several magnifications. The sizes 
of silver nanoparticles were determined by transmission elec-
tron microscopy (TEM) with a JEOL ARM200 microscope oper-
ating at 200 kV. TEM images were analyzed using ImageJ® 1.50b 
software to calculate the average size of the silver nano-
particles and the standard deviation from 10 images taken of 
each sample. EDX mapping was performed using a Centurio 
JED-2300 1sr detector coupled to a TEM microscope. Ther-
mogravimetric (TGA) and differential thermal (DTA) analyses 
were performed with a Setaram Labsys thermoanalyzer. A 
sample of about 10 mg contained in an alumina crucible was 
placed in a heating chamber. Measurements were recorded 
under an air stream with a heating rate of 5 °C min⁻¹ up to 
900 °C. The presence of micro- and mesopores in the calcined 
samples was determined by nitrogen adsorption analysis at the 
boiling point of liquid nitrogen using a Micromeritics ASAP 
2020 analyzer. The samples were previously degassed at 150 
°C for 15 h in a vacuum. The specific surface area (S_{BET}) was 
determined by the BET (Brunauer, Emmett and Teller) multi-
point method in the relative pressure (p/p₀) range of 0.05 to 0.30. 
To determine the macroporosity of the samples, the mercury 
intrusion porosimetry technique (Autopore IV, Micromeritics, 
USA) was selected. This is based on the progressive intrusion 
of mercury into the pores of a material under controlled pres-
sure. About 30 mg of powder was introduced into the penetrometer.

2.2. Characterization of antibacterial properties induced by 
the release of silver

2.2.1. Bacterial strain and culture medium. Experiments 
were performed with the E. coli K12 SCC1 strain, which emits 
fluorescence at 508 nm under excitation at an appropriate 
wavelength (396 nm) thanks to the chromosomal insertion of 
GFP (green fluorescent protein). Bacteria were stored in a 
freezer at −80 °C. M63G nutrient medium (0.1 M KH₂PO₄, 
20 wt% (NH₄)₂SO₄, 0.1 wt% FeSO₄, 20 wt% MgSO₄, 6 M KOH, 
0.05 wt% vitamin B1 and 10 wt% glucose) adjusted to a pH of 
6.8 was sterilized by autoclaving for 20 min at 121 °C and 100 
kPa before being used for the culture of bacterial cells.

2.2.2. Microbial cell culture. Bacteria from frozen stock were 
inoculated onto a lysogeny broth (LB) agar plate (15 g L⁻¹ in 
water) and incubated overnight at 30 °C. A first pre-culture was 
prepared from one colony of the cultivated bacteria in 10 mL of 
fresh M63G medium and incubated at 30 °C for 14 h. A second 
pre-culture was prepared from the first pre-culture (10 vol% 
dilution) in 10 mL of fresh M63G medium and cultivated at 30 °C 
for 4 h. The bacterial suspension was centrifuged for 30 min 
at 30 000 rpm, and the pellet was re-suspended in 4 mL of fresh 
M63G medium. The absorbance of a bacterial suspension con-
taining about 5 × 10⁶ bacteria per mL measured at 600 nm 
(Abs₆₀₀ nm) was adjusted to 0.05. This final bacterial suspension 
was used to perform bacterial colonization experiments on the 
zinc phosphate materials.

2.2.3. Zinc phosphate materials used for microbiological 
assays. The antibacterial activity of the zinc phosphate mate-
rials was investigated after calcination at 750 °C (ZnP0s-750 and 
ZnP0s/Ag-750). A polished silicon wafer (SW) disk cut into 
squares (about 10 mm × 10 mm) was used as a control in the 
bacterial colonization assays. Zinc phosphates in powder form 
(m = 80–100 mg) were introduced into small sterile glass 
containers (diameter of 10 mm and height of 3 mm) until they 
were almost completely filled. The glass containers with zinc 
phosphates were then placed in Petri dishes (diameter of 
35 mm and height of 10 mm). The calcined material samples 
were sterilized by the thermal treatment at 750 °C for 4 h. 
Silicon wafer samples were sterilized by dry heating at 180 °C 
for 1 h. All further manipulations of material samples in the 
microbiological assays were performed under sterile conditions.

2.2.4. Bacterial colonization of zinc phosphates. Material 
samples were inoculated with 4 mL of a bacterial suspension 
with an Abs₆₀₀ nm value of 0.05. They were incubated for 
periods of 2 h and 3 h at 30 °C. The samples were then carefully 
rinsed with a fresh solution of NaCl (9 g L⁻¹ in water) to 
eliminate non-adherent bacteria from the material surface. 
For this purpose, 1 mL of liquid was removed by putting a 
pipette tip onto the wall of the Petri dish without touching 
the material surface and was replaced by 1 mL of a fresh 
solution of NaCl. This rinsing process was repeated 8 times. 
The material samples were kept immersed in the liquid during 
rinsing to avoid any detachment of adherent bacteria by the 
creation of a material–air interface.

Colonization by bacteria of the material samples was analyzed in situ in the last rinsing medium under a confocal 
laser scanning microscope (Carl Zeiss LSM700) equipped with a 
63× water immersion objective (working distance of 2.4 
mm). Fluorescence and reflection images were taken at the 
same time at 6 different locations on each material sample. 
Excitation sources at 488 nm and 405 nm, respectively, and 
a 490–555 nm bandpass emission filter were used for fluo-
rescence data capture. At each location, stacks of 2D x,y images 
at intervals of 1 μm in the z-direction) were acquired. 
In each experiment, one sample of each type of material was 
used. The experiment was repeated 6 times.

2.2.5. Effect of release of Ag⁺ ions on bacterial viability. 
The effect of the release of silver ions from Ag nanoparticles on 
the viability of adherent bacteria was investigated by staining 
with red-fluorescent propidium iodide (PI) (1 g L⁻¹, ≥94%, 
Sigma-Aldrich), which penetrates bacteria with a damaged 
membrane. As the last rinsing step, the material samples 
were rinsed with a solution of PI in NaCl with a concentration 
of 4 mg L⁻¹. They were then incubated for 15 min at 30 °C. 
The material samples were analyzed by confocal laser scanning 
microscopy according to the procedure described above, with 
emission fluorescence recorded in the range of 600–700 nm 
and excitation at 555 nm. Quantification of Ag⁺ ions released 
into the liquid was performed by ICP-AES in specific experi-
ments. Samples were exposed to 4 mL of M63G medium free 
from bacteria for 3 h at 30 °C.
2.2.6. Image processing and statistical treatment. Adherent bacteria on material surfaces were quantified from fluorescence micrographs using ImageJ® 1.50b software and plugins. A total of 36 images were analyzed for each sample, which enabled the calculation of the mean value and standard deviation associated with each type of material. The significance of the results was determined using the statistical Student’s t-test with a 95% confidence interval.

2.2.7. Bacterial growth in zinc phosphate environment. The antibacterial effect of the zinc phosphate materials and casein powder on bacteria growing without direct contact with the materials was investigated by a plate diffusion assay in their as-synthesized forms (ZnPO/CN and ZnPO/CN/Ag) and after calcination at 750 °C (ZnPOs-750 and ZnPOs/Ag-750). Because the as-synthesized material samples were prepared at high temperatures for a sufficiently long period of synthesis (100 °C and 6 days), the material samples were used without any additional sterilization treatments to prevent degradation of casein. The calcined material samples were sterilized by the thermal treatment at 750 °C for 4 h. Agar plates of LB medium containing 5 spaced wells (diameter of 5 mm and height of 4 mm) were fabricated using previously sterilized cylindrical templates of Teflon®. A 100 µL sample of a fresh bacterial suspension (Abs600 nm of 0.50, prepared from one colony of bacteria in 10 mL of M63G medium and cultivated for 14 h at 30 °C) was carefully spread on an LB agar plate. The wells were filled with samples of the zinc phosphates and casein reagent, as used for the preparation of zinc phosphate, and incubated overnight at 30 °C. Bacterial growth around the samples was examined to identify the potential presence of a growth inhibition zone (expressed in cm²). The experiment was conducted in triplicate.

3. Results and discussion

3.1. Preparation and characterization of zinc phosphates loaded with Ag nanoparticles

As-synthesized zinc phosphates named as ZnPO/CN/Ag (ZnPO = zinc phosphate, CN = casein, Ag = silver) and ZnPO/CN (reference) were prepared via the same procedure, and the molar compositions of the starting mixtures are reported in Table S1.

Table 1 Molar compositions of the starting mixtures heated at 100 °C for 6 days

| Sample          | ZnO | P2O5 | Na2O  | H2O   | Ag⁺ | pH |
|-----------------|-----|------|-------|-------|-----|----|
| ZnPO/CN/Ag      | 1   | 0.093| 0.027 | 1.085 | 0.021| 4.0|
| ZnPO/CN         | 0   | 0.093| 0.027 | 1.085 | 0.021| 4.5|

P from casein protein (0.71 wt% of P).

ZnPO/CN/Ag sample contained an additional Ag phase characterized by three peaks observed at 38.1°, 44.2° and 64.4° (2θ) (Fig. 1a), which correspond to the (111), (200) and (220) diffraction planes of face-centered cubic (fcc) silver, respectively (PDF 00-004-0783). No diffraction peaks of silver oxide or other impurities were detected. The XRD results reveal that the crystallization of Ag was not hampered by the co-crystallization of hopeite. Using the XRD Scherrer method, an average Ag crystal size of 27 nm was determined.

TEM images and the particle size distribution of the ZnPO/CN/Ag sample are shown in Fig. 2. The TEM observations revealed the presence of spherical Ag nanoparticles that are fairly well distributed in the solid. The size distribution has an average value of 26.6 ± 10.4 nm, with some aggregates with sizes ranging from 40 nm to 60 nm. These results are in agreement with the value obtained by the Scherrer method.

The formation of Ag nanoparticles was initially a result of the incorporation of the silver nitrate solution to disperse casein micelles in the acidic medium (pH ~5). This involved the formation of casein–Ag⁺ ion complexes thanks to electrostatic interactions between Ag⁺ ions and functional groups of amino acids present in casein molecules such as amine (–NH₂), thiol (–SH) and carboxyl (–COOH) groups. At this stage, no evident difference in color was observed between the casein/Ag⁺ ions dispersion and the casein dispersion (reference), which suggested that Ag particles were not immediately formed, at least at room temperature. The concentration of NaOH (i.e., 3.0 mmol L⁻¹) may have been too low to promote the reducing capacity of casein protein, which led to incomplete reduction of Ag ions. The final incorporation of the zinc nitrate solution in the casein/Ag⁺ ions dispersion resulted in the formation of Ag particles. Moreover, the dark brown color of the product obtained via synthesis at 100 °C suggests the formation of Ag nanoparticles. This suggests that previously formed casein–Ag⁺ ion complexes reacted in situ to provide Ag.
particles, which were probably stabilized by casein as previously observed by Ashraf et al.39

3.2. Calcination conditions

As-synthesized ZnPO/CN/Ag samples were calcined at different temperatures ranging from 250 °C to 750 °C to investigate thermal influences on the crystallization process (Fig. 3, Table 2). The XRD results for the sample heated at 250 °C show a broad band centered at about 20° (2θ) (casein) and a few small unidentified peaks (Fig. 3d). This indicates the transformation of hopeite via thermal treatment. The absence of a signal for crystalline silver suggests possible thermal instability of the Ag nanoparticles in the presence of casein associated with the low sensitivity of the diffractometer used for the calcined samples. The amorphous solid recrystallized at 500 °C (Fig. 3c), mainly into zinc orthophosphate (α-Zn3(PO4)2) (PDF 00-029-1390). At higher calcination temperatures ranging from 600 °C to 750 °C (Fig. 3a and b), crystallization of α-Zn2P2O7 (PDF 00-008-038) occurred, which was probably induced by phosphate groups of phosphoserine residues released during the combustion of casein. In addition, better crystallization of Ag0 was observed at 600 °C, as indicated by the sharpening and higher intensity of the Ag peaks at 38.1°, 44.2° and 64.4° (2θ). Another compound, which was probably silver zinc orthophosphate of the Ag3Zn6(PO4)5 type (PDF 00-038-0302), crystallized at and above 600 °C. This may result from either Ag nanoparticles that were oxidized by PO43− species present in the sample,53,54 as indicated by the decrease in the intensity of Ag peaks at 750 °C, or silver atoms

| Sample              | Temperature (°C) | Result (XRD)                                                                 |
|---------------------|------------------|-----------------------------------------------------------------------------|
| ZnPO/CN/Ag          | RT               | Caseina + hopeiteb + Ag                                                     |
| ZnPOs/Ag-250        | 250              | α.p.c + nid (traces)                                                        |
| ZnPOs/Ag-500        | 500              | α-Zn3(PO4)3 + γ-Zn2P2O7 (traces) + a.p.c + Ag (traces)                       |
| ZnPOs/Ag-600        | 600              | Ag + α-Zn3(PO4)3 + α-Zn2P2O7 + γ-Zn2P2O7 (traces) + Ag3Zn6(PO4)5 (traces)    |
| ZnPOs/Ag-750        | 750              | α-Zn3(PO4)3 + α-Zn2P2O7 + Ag + γ-Zn2P2O7 (traces) + Ag3Zn6(PO4)5 (traces)    |

a Major phase. b Zn3(PO4)3·4H2O. c a.p.: amorphous phase. RT: room temperature (sample before calcination). nid: not identified.
not involved in the crystallization of Ag at temperatures of less than 600 °C that participated in the crystallization of this new compound. Furthermore, in the absence of Ag in the starting mixture, the same zinc phosphate phases crystallized (i.e., α-$\text{Zn}_3(\text{PO}_4)_2$ and γ- and α-$\text{Zn}_3\text{P}_2\text{O}_7$, hopeite).\textsuperscript{17}

According to chemical analyses (Table 3), the weight percentages of Ag, Zn and P in ZnPO/CN/Ag were lower than those in ZnPOs/Ag-750, as was expected owing to the predominance of casein in the sample before calcination. Because calcium cations were undetectable in the as-synthesized samples but measurable in casein (i.e., 0.15 wt%), colloidal calcium may have been almost completely solubilized and Ca\textsuperscript{2+} ions eliminated in the filtrate of the as-synthesized sample, while phosphate ions may have been used for the synthesis of hopeite. The presence of calcium in the calcined samples suggests that (i) Ca\textsuperscript{2+} ions present in the as-synthesized samples were involved in electrostatic interactions with phosphate groups of phosphoserine and (ii) Ca\textsuperscript{2+} ions compensated for some negative charges that formed in the sample calcined at 750 °C.

Simultaneous TG and DT analyses were performed on the as-synthesized ZnPO/CN/Ag and ZnPO/CN samples. As was expected because of the large amount of casein in the samples, the TGA and DT A curves shown in Fig. 4 have similar profiles to those of casein. The first loss of weight in the TGA curves between room temperature and 200 °C is related to physisorbed water. It is associated with the small endothermic peak centered at about 100 °C in the DT A curves. Three main weight losses that are ascribed to the decomposition of casein are observed between 200 °C and 780 °C, which are associated with the broad exothermic band with a maximum at about 520 °C. The higher values of exothermic enthalpy observed for ZnPO/CN/Ag in comparison with the ZnPO/CN sample suggest that the presence of Ag nanoparticles protected by casein led to an increase in the energy required for the decomposition of casein. The weight losses determined from the TGA curves of the as-synthesized ZnPO/CN/Ag and ZnPO/CN samples are summarized in Table 4. They indicate that the samples had similar contents of casein.

Furthermore, the influence of the calcination temperature on the particle size of silver nanoparticles was investigated by TEM (Fig. 5). Elemental mapping of Ag showed highlighted silver nanoparticles and aggregates (larger pink spots). The nanoparticle size was 9.9 \textpm\textbf{2.7} nm, 25.4 \textpm\textbf{7.5} nm and 6.4 \textpm\textbf{3.0} nm for the samples calcined at 500, 600 and 750 °C, respectively. The formation of aggregates of Ag nanoparticles with sizes of around 80 nm was most prominent at 600 and 750 °C. Such large aggregates were not observed in the as-synthesized sample, in which casein stabilized silver nanoparticles and prevented them from approaching each other. In contrast, they may have formed after the combustion of casein, which led to the decomposition of casein and, therefore, possible close contact between nanoparticles.

SEM analyses of the as-synthesized ZnPO/CN/Ag and ZnPOs/Ag-500, -600 and -750 samples, as shown in Fig. 6, show that the surfaces of the as-synthesized samples were composed of large irregular particles with undefined shapes and rough surfaces. At 500 °C, pores were formed (diameter: 0.05–0.2 μm). Macroporosity increased at 600 °C with a heterogeneous pore size distribution (i.e., a range of 0.10–1.70 μm) and an ill-defined pore shape. This macroporosity disappeared at 750 °C while larger quasi-spherical pores with a heterogeneous pore diameter (17 μm to 100 μm) were formed (Fig. 6d). In the case of the sample calcined at 600 °C, silver nanoparticles and aggregates are seen in the SEM images as white particles (Fig. 6c1 and c2).

| Sample         | Ag (%)     | Ca (%)   | P (%)    | Zn (%)   |
|----------------|------------|----------|----------|----------|
| ZnPO/CN        | —          | <0.003   | 0.84 (±0.03) | 2.41 (±0.06) |
| ZnPO/CN/Ag     | 0.57 ± 0.01| <0.003   | 1.04 (±0.41) | 3.09 (±0.06) |
| ZnPOs-750      | —          | 0.063 (±0.002) | 15.60 (±0.54) | 45.64 (±1.40) |
| ZnPOs/Ag-750   | 6.39 ± 0.26| 0.105 (±0.003) | 14.07 (±0.48) | 46.75 (±1.89) |
| Casein         | —          | 0.145 (±0.004) | 0.77 (±0.03)   | —         |
In addition, nitrogen adsorption measurements revealed that the calcined samples had very small specific surface areas ($S_{BET} < 3 \text{ m}^2 \text{ g}^{-1}$) and did not contain any micro- or mesopores. Complementary analyses performed by Hg porosimetry on the ZnPOs/Ag-750 sample revealed a broad pore size distribution (10 to 200 nm) with an average diameter of 70 nm (Fig. 7). This is mainly attributed to the intra-particle porosity visible in SEM images. The foamy aspect of the material observed after calcination was preserved during the porosity experiments; this proves that the porosity was stable up to a pressure of 200 MPa. The Hg cumulative pore volume was $1.28 \text{ cm}^3 \text{ g}^{-1}$.

### 3.3. Antibacterial activity of zinc phosphates

Bacterial growth in close proximity to the zinc phosphate materials was investigated by depositing samples onto agar nutrient plates inoculated with a bacterial suspension, which allowed diffusion of compounds from the material. Zones of bacterial growth inhibition were more clearly defined around the ZnPOs/Ag-750 sample ($0.79 \pm 0.23 \text{ cm}^2$) in comparison with the other samples (Fig. 8). This is ascribed to the antibacterial activity of Ag$^+$ ions that were released by the oxidation of Ag nanoparticles present in the material in contact with the liquid[8,9,55] and diffused through the agar gel. In contrast, an
inhibition zone was not observed for the ZnPO/CN/Ag samples, which was probably due to the lower silver content (Table 3 and ESI† Table 1). In addition, the results indicate that the Ag-free ZnPO/CN and ZnPOs-750 samples did not release a sufficient quantity of any substance able to inhibit the growth of *E. coli*, such as zinc or casein. Pure casein powder, which was used as a positive control, was shown to provide a weak antibacterial effect (0.25 ± 0.16 cm²) after overnight incubation (Fig. 8), as already reported in the literature.⁵⁶

Bacterial colonization on the ZnPOs-750 and ZnPOs/Ag-750 materials was investigated using the *E. coli* K12 SCC1 strain (see ESI† for ZnPO/CN and ZnPO/CN/Ag). Fig. 9a(1–3) shows typical examples of micrographs acquired by confocal fluorescence microscopy after incubation for 3 h. Bacteria were uniformly distributed on all material surfaces and were preferentially located in surface cavities of the ZnPOs-750 and ZnPOs/Ag-750 materials, which were probably formed from macropores (pore diameter between 350 and 1000 nm).⁵⁷ A quantitative view of colonization after incubation for 2 h and 3 h is given in Fig. 9b, which reveals that the number of bacteria was significantly lower on zinc phosphate surfaces in comparison with the SW control and thus shows that the zinc phosphate surfaces were unfavorable for bacterial colonization.

**Fig. 6** SEM images of (a) ZnPO/CN/Ag, (b) ZnPOs/Ag-500, (c, c1 and c2) ZnPOs/Ag-600 and (d) ZnPOs/Ag-750. The inset in (c1) displays a magnified image of the enclosed area showing Ag nanoparticles with particle sizes ranging from 14 nm to 27 nm, which are also observed in (c2).

**Fig. 7** Pore size distribution of ZnPOs/Ag-750 determined by mercury porosimetry.
The capacity of zinc phosphates to affect bacterial proliferation was also investigated on the basis of a comparison between zinc phosphates in terms of colonization after the materials were exposed to bacteria for 2 h and 3 h (see ESI† for ZnPO/CN and ZnPO/CN/Ag). As shown in Fig. 9b, the number of adherent bacteria on the ZnPOs-750 material significantly increased between incubation durations of 2 h and 3 h (about +20%) (Table 5), as was expected owing to the normal growth of bacterial populations on surfaces. Nevertheless, it was significantly less than that measured on the SW control (about +87%), which is attributed to the action of ions such as Zn\(^{2+}\) that were released from the zinc phosphate materials (Table 5) and are known to exhibit inhibitory effects on bacteria.\(^{57,58}\)

Regarding potential antibacterial activity related to the Ag content of the materials, the results revealed a significant reduction in the bacterial population on an Ag-based (ZnPOs/Ag-750) in comparison with an Ag-free (ZnPOs-750) material after incubation for 2 h (Fig. 9b). A significant reduction in the number of adherent bacteria was also observed on ZnPOs/Ag-750 between incubation durations of 2 h and 3 h (~19%, Fig. 9b and Table 5). These results are consistent with the antibacterial activity expected for the amount of Ag\(^{+}\) measured after release in the M63G culture medium (10\(^{-5}\) M, Table 5), i.e.,

**Fig. 8** Antibacterial activity against *E. coli* tested by measuring the zone of growth inhibition after overnight incubation (inset: a magnified image of zone 5): (1) ZnPO/CN, (2) ZnPOs-750, (3) ZnPO/CN/Ag, (4) ZnPOs/Ag-750 and (5) casein samples on *E. coli* K12.

**Fig. 9** (a) 2D z-projection of confocal 3D images of *E. coli* K12 bacteria adherent on (a1) SW, (a2) ZnPOs-750 and (a3) ZnPOs/Ag-750 materials after incubation for 3 h. (b) Number of bacteria on SW control, ZnPOs-750 and ZnPOs/Ag-750 materials after incubation for 2 h and 3 h (average of 6 experimental replicates). The * and § symbols indicate significant differences in comparison with SW and ZnPOs-750, respectively (p-value < 0.05). Inset: a magnified image of the histogram showing the number of bacteria on the ZnPOs-750 and ZnPOs/Ag-750 materials.
large enough to induce the death of *E. coli* bacteria and a subsequent reduction in the bacterial population (minimum inhibitory concentration of 0.6 × 10⁻⁶ M, 0.4–2.0 × 10⁻⁶ M for Ag⁺, i.e., AgNO₃).³⁸⁻⁴⁴ However, bacterial staining by PI used as a marker of damaged *E. coli* K12 bacteria showed that bacteria that adhered to the ZnPOs/Ag-750 material displayed an intact membrane after incubation for 3 h, which indicates that the antibacterial activity was lower than that expected for such a released quantity of Ag⁺ ions (10⁻⁵ M). This is ascribed to the specific culture conditions in comparison with studies reported in the literature. In particular, the bacterial strain, culture medium and temperature are parameters known to affect bacterial sensitivity to antibacterial agents.⁶²,⁶³ Thus, reported studies⁴⁴,⁶⁵ and results obtained by our group using M63G culture medium⁶⁶ have shown that the interactions of Ag⁺ ions with compounds in different media reduce the amount of Ag⁺ available for antibacterial action. Despite these interactions, however, the results reported here demonstrate that Ag⁺ ions were released from the ZnPOs/Ag-750 material in sufficient amounts to affect both bacterial colonization and proliferation and therefore to endow it with antibacterial properties.

### 4. Conclusion

In this study, bio-sourced macroporous zinc phosphates with Ag nanoparticles (ZnPOs/Ag) were successfully obtained via an original method of synthesis. This new route for the synthesis of silver-based metal phosphates is advantageous because casein (CN) acts as a bio-source of phosphorus, a stabilizer and a reducing agent for the synthesis of Ag nanoparticles. It also induces intra-particle macroporosity. During calcination of the as-synthesized compounds (ZnPO/CN/Ag), the crystallization of Ag-supported zinc phosphates was improved up to 600 °C. A study of the antibacterial activity of Ag-containing and Ag-free zinc phosphates against *E. coli* K12 bacteria demonstrated that porous zinc phosphates inhibited bacterial adhesion over a short incubation period. The antibacterial activity of ZnPOs/Ag-750, which was related to the release of Ag⁺ ions from the material, indicated significant bactericidal properties and even led to reductions in populations of already adhered *E. coli* K12. Growth-inhibiting and bactericidal effects were also observed against bacteria present in the environment of the ZnPOs/Ag-750 material. To the best of our knowledge, antibacterial activity of zinc phosphates against *E. coli* is reported here for the first time. These materials could be used, for example, in the treatment of cooling water, heating systems and wastewater.

On the other hand, this work confirms the feasibility of the synthesis of (porous) metal phosphates without the use of reactants derived from phosphate rocks. In addition, such a method of synthesis could be extended to the synthesis of other metal-doped phosphates.

### Conflicts of interest

There are no conflicts to declare.

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