Antidiabetic Phytochemicals From Medicinal Plants: Prospective Candidates for New Drug Discovery and Development

Safaet Alam¹, Md. Moklesur Rahman Sarker¹,²*, Taposhi Nahid Sultana³, Md. Nafees Rahman Chowdhury⁴, Mohammad A. Rashid⁵, Nusrat Islam Chaity¹, Chao Zhao⁶, Jianbo Xiao⁷, Elsaoy E. Hafez⁸, Shah Alam Khan⁹ and Isa Naina Mohamed¹⁰*

¹ Department of Pharmacy, State University of Bangladesh, Dhaka, Bangladesh, ² Pharmacology and Toxicology Research Division, Health Med Science Research Limited, Dhaka, Bangladesh, ³ Department of Pharmacy, University of Asia Pacific, Dhaka, Bangladesh, ⁴ Department of Pharmacy, University of Dhaka, Dhaka, Bangladesh, ⁵ Department of Pharmaceutical Chemistry, Faculty of Pharmacy, University of Dhaka, Dhaka, Bangladesh, ⁶ College of Food Science, Fujian Agriculture and Forestry University, Fuzhou, China, ⁷ Department of Analytical Chemistry and Food Science, Faculty of Food Science and Technology, University of Vigo, Vigo, Spain, ⁸ Plant Protection and Biomolecular Diagnosis Department, ALCRI (Arid Lands Cultivation Research Institute), City of Scientific Research and Technological Applications, Alexandria, Egypt, ⁹ College of Pharmacy, National University of Science & Technology, Muscat, Oman, ¹⁰ Pharmacology Department, Medicine Faculty, Universiti Kebangsaan Malaysia (The National University of Malaysia), Kuala Lumpur, Malaysia

Diabetes, a chronic physiological dysfunction affecting people of different age groups and severely impairs the harmony of peoples' normal life worldwide. Despite the availability of insulin preparations and several synthetic oral antidiabetic drugs, there is a crucial need for the discovery and development of novel antidiabetic drugs because of the development of resistance and side effects of those drugs in long-term use. On the contrary, plants or herbal sources are getting popular day by day to the scientists, researchers, and pharmaceutical companies all over the world to search for potential bioactive compound(s) for the discovery and development of targeted novel antidiabetic drugs that may control diabetes with the least unwanted effects of conventional antidiabetic drugs. In this review, we have presented the prospective candidates comprised of either isolated phytochemical(s) and/or extract(s) containing bioactive phytoconstituents which have been reported in several in vitro, in vivo, and clinical studies possessing noteworthy antidiabetic potential. The mode of actions, attributed to antidiabetic activities of the reported phytochemicals and/or plant extracts have also been described to focus on the prospective phytochemicals and phytosources for further studies in the discovery and development of novel antidiabetic therapeutics.

Keywords: diabetes mellitus, antidiabetic, antihyperglycemic, phytochemical, phytomedicine, bioactive compound, drug discovery, drug development
INTRODUCTION

Diabetes mellitus is a type of chronic metabolic disorder categorized by insufficiency in insulin activity and/or insulin secretion. Anomalies in proteins, carbohydrates and lipids metabolism can arise due to the lack of insulin, an anabolic hormone (1). These abnormalities in metabolism are caused by low levels of insulin, insulin resistance of target tissues, insulin receptor level, primarily skeletal muscles, and adipose tissue and to a lesser degree, liver, signal transduction system, and/or effector enzymes or genes and/or signal transduction pathway (2). Diabetes is one of the most abundant metabolic diseases across the world accounting for about 2.8% of the population worldwide and is projected to reach 4.4% by 2030 which has already risen to an unprecedented extent of the epidemic (3). Despite being a non-communicable disorder, diabetes is considered one of the five biggest morbidities worldwide (1). Diabetes category and frequency vary depending on the severity of the symptoms. Some patients with diabetes are asymptomatic, particularly patients with type 2 diabetes during the initial periods of illness whereas others have noticeable hyperglycemia. Uncontrolled and unmonitored diabetes can lead to stupor, coma and even death if kept untreated because of ketoacidosis or rare non-ketotic hyperosmolar disorders (4). The development of diabetes may include the interaction of genetic and non-genetic factors (5). Despite diabetes classification being crucial and having repercussions for treatment policies, this is somehow ambiguous and many diabetic individuals do not easily accommodate into one class, exclusively younger adults and 10% of the initially classified patients can need revision afterward (6). The standard classification of diabetes as type 1, type 2 and gestational diabetes mellitus (GDM) as introduced by the American Diabetes Association (ADA) in 1997 remains the best-accepted and adopted by ADA (4).

Currently, there are many antidiabetic drugs available in the market to treat hyperglycemia which notably works via improvement of insulin sensitivity, complementing insulin, upraising insulin secretion and stimulating glucose uptake. But metformin and sulfonylureas type antidiabetic drugs are compromised with several unwanted side effects such as diarrhea and lactic acidosis (demonstrated by metformin) and hepatic failure, weight gain, tachycardia and hypothyroidism (demonstrated by sulfonylureas) (7). Plant is always considered as one of the most reliable sources of curing agents of diseases and many of those synthetic drugs are either directly or indirectly derived from them. Plants and plant products can exert promising antidiabetic efficacy based on recent studies (Figure 1). Plant sources of antidiabetic agents are very much popular from the ancient era as they are relatively safer and much cheaper alternatives than synthetic drugs and are also mentioned in many folkloric medicines including the Indian, Korean and Chinese culture. Traditional herbal medicines and functional foods are believed to ameliorate diabetic syndromes via six notable mechanism of actions including enhanced insulin secretion and sensitivity, glucose uptake by muscle cells and adipose tissues and inhibition of glucose absorption from intestine and glucose production from hepatocytes along with demonstrating anti-inflammatory properties (7). As a result, functional foods and phytotherapies are becoming popular across the world day by day (8). In the current review, we have compiled most notable medicinal and dietary plants along with their isolated antidiabetic phytochemicals to give distinct insights into the establishment of novel functional foods and drug moieties against diabetes. The graphical abstract of the manuscript has been presented in Figure 2.

LATEST RESEARCHES ON THE MOLECULAR MECHANISMS AND PATHOGENIES OF DIABETES MELLITUS

In type 1 Diabetes, the pancreatic β-cells undergo autoimmune destruction by CD4+ and CD8+ T cells and macrophages which result in insulin deficiency (9, 10). Islet cell antibodies are found in nearly 85% of the patients, and most of them act against the glutamic acid decarboxylase (GAD) found inside the β-cells of the pancreas (9).

The metabolic disorders related to type 1 diabetes mellitus are a result of deficiency of insulin secretion caused by the immune destruction of islets of Langerhans of the pancreas. Besides, pancreatic α-cells start to function abnormally and secrete an excessively large amount of glucagon in patients with type 1 diabetes mellitus, which further aggravates the metabolic disorders already caused by insulin deficiency (11). A deficiency of insulin causes lipolysis to occur at an uncontrolled rate which causes the amount of free fatty acids in the blood to rise resulting in a reduction of glucose metabolism in the peripheral tissues (11). The deficit of insulin also causes a reduction of glucokinase enzyme in the liver and the GLUT-4 transporter protein in adipose tissue resulting in an inability of the target tissues to respond normally to insulin.

Impaired secretion of insulin through destruction of the insulin secreting β-cells, and diminished insulin activity through insulin resistance marks the underlying mechanisms of the pathogenesis of type 2 diabetes (9). The mitochondria-endoplasmic reticulum contacts are known as mitochondria-associated membranes (MAMs) play an important role in the regulation of lipid exchange, signaling of calcium, cell survival, and homeostasis in cellular metabolism. These MAM contacts are known to contain several insulin signaling proteins such as AKT kinase, mTORC2, PP2A, and PTEN and thus participate in insulin signaling. A growing number of studies have shown that these MAMs are involved in causing dysfunction of the insulin producing β cells, resistance to insulin in the peripheral tissues, leading to type 2 diabetes mellitus (12). miRNAs are small RNA consisting of 20–24 nucleotides that regulates early development, fat metabolism, cell proliferation, differentiation, apoptosis, and death. Recent studies have shown that these miRNAs contribute to the pathogenesis of type 2 diabetes mellitus and may be developed into new biomarkers (13).

As reactive oxygen species impact chemical changes in all cellular components and produce lipid peroxidation, oxidative...
stress also causes type 2 diabetes mellitus. As a result, lipid peroxidation is another important cause of type 2 diabetes mellitus (14). In excess amounts of hydrogen peroxide (H$_2$O$_2$), DNA, RNA, and lipids are severely damaged. Catalase (CAT) is the major H$_2$O$_2$ regulator, and it neutralizes H$2$O$_2$ by catalytically converting it to water and oxygen. When catalases (CAT) are deficient, pancreatic islet-cells are more susceptible to excessive formation of reactive oxygen species (ROS) and oxidative stress, which leads to pancreatic islet dysfunction and overt type 2 diabetes mellitus (15). In numerous illness states involving oxidative stress as a significant causal factor, such as diabetes and obesity, plasma levels of oxidized low-density lipoprotein (oxLDL) are elevated (16). Nuclear factor kappa B (NF-B), NH2-terminal Jun kinases, and p53 MAPK are transcriptionally regulated pathways that have been considered one of the most important stress-signaling pathways, and oxidative stress plays a role in the development of type 2 diabetes mellitus through their involvement (14).

Type 2 diabetes mellitus is linked by decreased physical activity and exercise training, as well as increased sedentary habits, which are linked to elevated indicators of chronic systemic inflammation (17). Proinflammatory molecules such as interleukin 6 (IL-6), C-Reactive Protein (CRP), tumor necrosis factor-alpha (TNF-) and interleukin 1 (IL-1) are released into the bloodstream and inside specific organs in this scenario, causing metabolic inflammation (18). IL-1 is involved in the pancreatic autoimmune response, decreasing -cell activity and activating the nuclear factor kappa-light-chain-enhancer of activated B cells (NF-B) transcription factor, inhibiting -cell function and inducing death.

The importance of the gut microbiota in the development of diabetes has been demonstrated, and new studies suggest that dysbiosis can increase type 2 diabetes mellitus (19). Experiments in animal models showed that a high-fat diet can increase the synthesis of lipopolysaccharide (from Gram-negative bacteria) by up to thrice, contributing to low-grade inflammation and insulin resistance (20, 21). Intestinal dysbiosis can also impair short-chain fatty acid production, which is important for gut barrier integrity, pancreatic cell proliferation, and insulin biosynthesis (22, 23). Dysbiosis can also affect the production of other metabolites such as branched amino acids and trimethylamine, causing glucose homeostasis to be disrupted and type 2 diabetes mellitus to develop (24, 25). The clinical consequences of the gut microbiome are still being researched, and more study is needed to better understand the link between gut bacteria and type 2 diabetes mellitus (26).

**ARTICLES SEARCH STRATEGY**

An extensive literature search was carried out through the following databases: Web of Science, Scopus, PubMed/Medline, ScienceDirect, ClinicalTrials.gov, Wiley Online Library and Google Scholar. The following keywords were used:
‘Antidiabetic’, ‘Diabetes’, ‘Phytochemical’, ‘Bioactive compound’, ‘Type 2 diabetes’, ‘Pharmacology’, and ‘Clinical trial’. Only peer-reviewed scientific journals were considered during the process. Plants with reported antidiabetic phytochemicals along with mechanism of actions have been considered only for the review work. Of the 598 identified papers and clinical trials records, 295 unique articles were included and reported in this comprehensive review following inclusion criteria.

**Anoectochilus roxburghii (WALL.) LINDL.**

Anoectochilus roxburghii (Wall.) Lindl. (family: Orchidaceae) is a perennial herb which mainly occurs in China, Taiwan, Japan, Sri Lanka, India, and Nepal. Polysaccharides from A. roxburghii lower blood glucose levels by enhancing the body’s antioxidant capacity, reducing blood lipid levels, modulating the activity of glucose-metabolizing enzymes, minimizing tissue damage such as pancreas, and promoting damaged tissue repair (27). Kinsenoside, extracted from A. roxburghii was found to demonstrate significant hypoglycemic activity. Previous studies have shown that kinsenoside could help in the restoration of damaged δ cells in pancreas and function against oxidative stress and NO factor and also regulates antioxidant enzymes, scavenging of free radicals (28).

**Bacopa monnieri (L.) WETTST.**

Bacopa monnieri (L.) Wettst. (family: Scrophulariaceae) is a creeping herb which occurs across India. Several compounds including tetracyclic triterpenoid saponins, Bacosides A and B, Hersaponin, alkaloids viz. Herpestine and Brahmine and flavonoids have been isolated from the plant (29). Bacosine, a triterpene isolated from the ethyl acetate fraction of the ethanolic extract of B. monnieri showed pronounced reduction in blood glucose levels in diabetic rats in a dose-dependent mode; however, no such effect has been observed on normal rats. Thus, bacosine is known to possess antihyperglycemic properties rather than hypoglycemic activity. It has been suggested that bacosine works in a way similar to insulin and that its antihyperglycemic activity might be attributed to the increase in the consumption of peripheral glucose as well as protect against oxidative damage in alloxan induced diabetes. Bacosine caused a pronounced increase (p < 0.001) of glycogen content in the liver of diabetic rats and consequently proved its insulin-like activity resulting in an increased uptake of glucose (29).

**Berberis aristata DC.**

Berberis aristata DC (family: Berberidaceae) is native to Northern Himalayan part and is known locally as ‘Daruhaldi’ or ‘Citra’. Root extract B. aristata regulates glucose homeostasis by reducing gluconeogenesis and oxidative stress and exhibits a strong anti-hyperglycemic activity (30, 31). The major antidiabetic compound extracted from this plant is berberine. Berberine is known to act through several mechanisms, including insulin-mimetic activity; improving the action of insulin by triggering AMPK (5’ adenosine monophosphate-activated protein kinase); reducing insulin resistance through protein kinase C-dependent up-regulation of insulin receptor expression; causing glycolysis; and by enhancing GLP-1 (Glucagon-like peptide 1) secretion and regulating its release, and by inhibiting DPP-IV (Dipeptidyl peptidase-4) (30).

According to 32, berberine extracted from B. aristata in a manner of 0.5 gm thrice a day on type 2 diabetic patients showed equal efficacy to metformin monotherapy in the reduction of fasting blood glucose, HbA1c, postprandial blood glucose, postprandial insulin and basal insulin (32). Another clinical trial also showed the efficacy of berberine as adjuvant therapy in poorly controlled type 2 diabetic patients (32). In other clinical trials, berberine also showed promising efficacy compared to rosiglitazone and metformin by reducing HbA1c, fasting blood glucose, postprandial blood glucose despite having some adverse effects on liver (33). In another 6 months long randomized-controlled trial on 85 type 1 diabetic patients (39 males and 46 females), twice daily intake of a tablet containing 588/105 mg combination of Berberis aristata/Silybum marianum decreased insulin use by the body during insulin therapy. Moreover, there was a decrease in Hb1Ac level in comparison to baseline and in fasting and postprandial plasma glucose levels in comparison to both baseline and placebo (34). This combination, which is made to improve the low bioavailability of berberine in oral route, has also been reported to show notable antidiabetic effects in 136 obese/overweight type 2 diabetic patients, as per another year-long placebo-controlled study (35).

**Bixa Orellana L.**

Bixa Orellana L. (family: Bixaceae) also known by the name ‘Achueté’, is a rapidly growing shrub that can grow up to 3 to 5 meters in height. The plant originated in Brazil but also grows in South and Central America. Besides, it also occurs in tropical countries such as Peru, Mexico, Ecuador, Indonesia, India, Kenya, and East Africa (36). B. Orellana lowered the blood glucose levels in dogs with streptozotocin-induced diabetes (36, 37). It has been suggested that B. Orellana causes a reduction in blood glucose level by increasing peripheral utilization of glucose (38, 39), increasing plasma insulin levels and increasing the binding of insulin to insulin receptors (40). Bixin, a high carotenoid content and a natural pigment found in B. orellana showed prominent hypoglycemic actions. Bixin is commercially named “Annatto” and a very promising wellspring of new medicines as well imperative nutraceuticals (36).

**Bumelia sartorum MART.**

Bumelia sartorum Mart. (family: Sapotaceae) is a large and tall tree commonly known as ‘Quixaba’, ‘Quixabeira’, ‘Tranceporeira’,...
‘Sacutiaba’ and ‘Rompe-gibao’ in northeastern Brazil. It naturally grows from north of Minas Gerais to Piauí (41). An unsaturated triterpene acid named basic acid, isolated from the ethanol extract of root bark of B. surtorum exhibited significant hypoglycemic activity in alloxan induced diabetic rat models. Moreover, basic acid was found to significantly increase the glucose uptake and glycogen synthesis process in isolated rat diaphragm. In alloxan-diabetic rats following basic acid treatment, a significant increase in plasma insulin levels was found. It has been suggested that basic acid increases insulin secretion from pancreatic beta-cells. This could be the underlying mechanism by which basic acid shows its hypoglycemic property which was found to be approximately equal to that of chlorpropamide (42).

**Callistemon rigidus R.Br.**

*Callistemon rigidus* R.Br. (family: Myrtaceae) is an evergreen plant which is native to Australia. Noteworthy antidiabetic compounds piceatannol and scirpusin B were isolated from the stem bark of the plant using 1H- and 13C-NMR technology (43). These compounds can suppress the activity of α-amylase in isolated mouse plasma Methanol extract of *C. rigidus* can also demonstrate prominent repressing activity on α-amylase. Besides, scirpusin B can regulate α-amylase in mouse GIT to demonstrate antidiabetic efficacy. These compounds are also expected to abate increment of postprandial glucose level and can offer a very good wellspring of antidiabetic drug development (44).

**Catharanthus roseus (L.) G.DON**

*Catharanthus roseus* (L.) G.Don (family: Apocynaceae) is a shrub-type plant that can grow up to 30–100 cm in height. The plant originated from Madagascar but is available around the world due to its high survival rate (45). Its leaf extracts exhibited significant dose-dependent blood sugar-lowering activity in normal and streptozotocin-induced diabetic model rats. The blood sugar-lowering potential of the leaf extract was equivalent to that of the commercially available drug Tolbutamide in the animal models (45, 46). Compared to normal animals, the enzymic activities of glycinogen synthase, glucose 6-phosphate-dehydrogenase, succinate dehydrogenase and malate dehydrogenase were found to decrease in the liver of diabetic animals and were significantly increased after treatment with dichloromethane–methanol (DCMM) extract of leaves and twigs of *C. roseus* at dose 500 mg/kg p.o. for 7 days. It has been suggested that *C. roseus* exhibits its anti-diabetic activity by increasing glucose metabolism in treated rats (47). Among isolated compounds, especially alkaloids including vindoline, vindolidine, vindolicine and vindolinine, isolated from *Catharanthus roseus* leaves induced increased glucose uptake in myoblast C2C12 or pancreatic β-TC6 cells where vindoline showed maximum efficacy. First three compounds did not exhibit any cytotoxicity towards pancreatic β-TC6 cells even when administered in the maximum dose of 25.0µg/mL. Vindoline, vindolidine and vindolinine also revealed improved protein tyrosine phosphatase-1B (PTP-1B) inhibition actions which can play a pivotal role in type 2 diabetes management (45).

**Chamaemelum nobile (L.) ALL.**

*Chamaemelum nobile* (L.) All. (family: Asteraceae) is a native South-eastern Moroccan shrub-type which is locally known as ‘Babounge’. It is very much popular throughout Europe, most notably in France and USA (48). Its aqueous extract mitigated blood glucose concentration in streptozotocin induced diabetic rat models except changing plasma insulin level which indicated an insulin secretion independent pathway (48). *C. nobile* may also exhibit its hypoglycaemic activity in the gastrointestinal tract by slowing down the digestion process and reducing the rate of carbohydrate absorption (48). Chamaemeloside, 3-hydroxy-3-methylglutaric acid (HMG) containing flavonoid glucoside is the most notable isolated antidiabetic compound from this plant which revealed hypoglycemic activity in Swiss-Webster mice models by reducing plasma glucose concentration. The reduction of fasting glucose level and improved glucose tolerance referred that it might act following more than one mechanism (49). The underlying mechanism of action may be attributed to the stimulation of the utilization of peripheral, especially in muscle and adipose tissue. In 8 week long clinical study on 26 pre-diabetic volunteers (21 were male and 5 were female; mean age: 50.5 ± 8.5 years), a mixed herbal extract supplementation was made by combining hot water extract of *Anthemis nobilis* (Roman chamomile), which is a synonym of *Chamaemelum nobile* and *Vitis vinifera* in the dose of 1200 mg reportedly reduced abnormal glucose values and thus the risk of developing diabetes (50).

**Chichorium intybus L.**

*Chichorium intybus* L. (family: Asteraceae) is an erect herb-type perennial plant that grows up to 1 m. It is abundantly found in Asia, Africa, Europe, and Southern America. It is well-known as ‘Chicory’ (51). Its ethanolic extract (CIE) was found to show a marked reduction in the hepatic glucose-6-phosphatase (Glc-6-Pase) activity when compared to the control group. The decrease in the activity of hepatic Glc-6-Pase could lead to a reduction in the production of hepatic glucose, which in turn reduces the blood glucose level in CIE-treated diabetic rats (52). Chlorogenic acid and chicoric acid are the two most notable antidiabetic phytoconstituents isolated from this plant which can increase glucose uptake in L6 muscular cells. Both phytochemicals can also upraise insulin secretion from the INS-1E insulin-secreting cell line and rat islets of Langerhans. Besides, chicoric acid can exert both insulin-secreting and sensitizing activities (53). A randomized, double-blind clinical trial on 100 type 2 diabetic
patients (55 were male and 45 were female) reported a reduction in HbA1c value from 8.6% at baseline to 7.42% after 12 weeks and thus indicated the potentiality of Cichorium intybus seed supplementation as an adjunct therapy in type 2 diabetes mellitus (54). A similar 4 weeks long randomized, double-blind, placebo-controlled study in 47 healthy subjects (8 were male and 39 were female; age range: 33-70 years) reported that the seed supplementation at 300 ml/day dose caused improvement in adiponectin level and fecal properties along with the antihyperglycemic activity, indicating that the supplement, which included inulin-type fructans, is effective in delaying diabetic mellitus onset and helps improving bowel movements (55).

**Cinnamomum verum J. PRESL**

*Cinnamomum verum* J. Presl (family: Lauraceae) is an evergreen plant with a height of 10-15 m which is local to Southern India and Sri Lanka. Apart from these places, it is widely available in other Asian, Australian, Caribbean and African countries but most notably in China, Indonesia, Madagascar, Vietnam and Burma. It is well-known as 'Ceylon cinnamon', 'True cinnamon', 'Darchini', 'Dalchini' and 'Mexican cinnamon'. The name Ceylon came after the former name of Sri Lanka, its native place. Bark (after drying) is the most important part of the plant with remarkable medicinal values (56). Methanolic extract from *C. verum* can suppress the activity of α- glucosidase and α-amylase (57). According to the study, cinnamon aqueous extract also showed notable antidiabetic activity in alloxan induced diabetic rat models by reducing fasting blood sugar, triglycerides and total cholesterol when tested for thirty days long (57). Interestingly, a lower dose of cinnamon extract i.e. 200 mg/kg showed maximum antidiabetic efficacy. In another research, it was revealed that hydro-alcoholic extract of Cinnamon can ameliorate postprandial glycemia more than its aqueous extract (58). Cinnamon can also increase the uptake of glucose by upraising the number of insulin receptors, glucose transporter 4 and activating glycogen synthase to diminish glucose levels (56, 59). Cinnamon extract was also co-administered with other herbs to evaluate the synergistic activity on diabetic complications. Again, a combination of methanolic cinnamon extract along with green tea can also decrease blood glucose concentration and body weight significantly in streptozotocin induced diabetic rat models by showing synergism (56, 60). Among isolated phytochemicals, it is believed that cinnamon polyphenols like eugenol and pyrogallol can demonstrate antidiabetic properties by renovating beta cells which leads to hypoglycemic and hypolipidemic actions (56). According to Tulini et al. solid lipid microparticles (SLM) of proanthocyanidin rich cinnamon extract can improve the antidiabetic efficacy of foods (61). Again, cinnamaldehyde can ameliorate the uptake of glucose by upraising the amount of AKT2 and aortic nitric oxide synthase 3 (eNOS), insulin receptor substratel (IRS1) and p-85 regulatory subunit of PI3K (PI3K-P85) while concurrently abating the expression of NADPH oxidase 4 (NOX4) which eventually balance the increased glucose concentration (56, 59). Cinnamon supplementation at the dose of 500 mg showed prominent antidiabetic action in a 3 month long randomized, triple-blind placebo-controlled, parallel clinical trial in 138 type 2 diabetic patients (63 were male and 75 were female; age range: 30-80 years) by causing a reduction in all glycemic parameters, namely FPG by -13.1 ± 1.7 mg/dl, HbA1C by -0.27 ± 0.04%, 2HPP by -16.9 ± 2.5 mg/dl, insulin resistance (HOMA-IR) by -1.01 ± 0.11 and fasting insulin by -1.77 ± 0.41 mlU/L) with no side effects while a better glycemic control was observed in patients whose BMI values were greater than 27 (59). Another phase 1 clinical study on 30 healthy adults (50% were male; mean age: 38.8 ± 10.4 years; age range: range 21–58 years) showed that cinnamon had no significant toxicity or side effects (62). In a review of 8 randomized-controlled trials, it has been found that sole therapy of powdered or aqueous form of cinnamon, at different doses starting from 0.5 g to 5 g per day, ameliorated glycemic control in type 2 diabetic and prediabetic patients (impaired fasting glycemia or impaired glucose tolerance) (63).

**Costus pictus D. DON**

*Costus pictus* D. Don (family: Zingiberaceae) is a rhizomatous medicinal herb which is popularly known as ‘Insulin plant’ for its strong antidiabetic efficacy. It demonstrated antidiabetic action by the inhibition of α-amylase and α-glucosidase activity (64). *C. pictus* can also improve the secretion of insulin in diabetic rat models along with improvement in glucose utilization (65). It has been found that upon the administration of aqueous extract of *C. pictus* to diabetic rats, *C. pictus* causes a marked reduction in blood glucose levels and an increase in plasma insulin level. Regulation of glucose homeostasis by improved peripheral glucose utilization, increased hepatic glycogen synthesis and/or decrease of glycogenolysis, inhibition of intestinal glucose absorption, and lowering of the glycaemic index of carbohydrates might be responsible for the antidiabetic effect of *C. pictus* (66). Earlier researchers also found that β-amyrin and methyl tetracosanate are the major bioactive phytoconstituents which exhibited ameliorated glucose uptake in 3T3-L1 adipocytes (67, 68). In another study conducted on *C. pictus*, β- L- Arabinopyranose methyl glycoside was reported responsible for antidiabetic property (69). Ingestion of the leaves of *C. pictus* by diabetic patients showed statistically significant reduction in their fasting and postprandial blood glucose levels, as per a cross-sectional clinical study (70).

**Curcuma longa L.**

*Curcuma longa* L. (family: Zingiberaceae) also known as ‘Turmeric’ is a moderately tall perennial plant containing underground rhizomes. It is grown in tropical regions like Pakistan, China, Peru and India. The curcuminoids bisdemethoxycurcumin, curcumin and demethoxycurcumin were isolated from *C. longa* and were found to exhibit
α-glucosidase inhibitory activity (71). Among the three curcuminois, bisdemethoxycurcumin showed the most potent α-glucosidase inhibition (72). Besides, volatile oils extracted from both fresh and dried turmeric rhizomes showed potent glucosidase inhibitory activity in a dose-dependent mode, and dried rhizomes increased the glucosidase inhibitory action significantly. Potent α-glucosidase and α -amylase inhibitory activity was exhibited by Aromatic-Turmerone, the main volatile component in turmeric rhizome (73). Turnerin, a water-soluble protein found in turmeric rhizomes inhibits α-amylase and α-glucosidase activities. Thus, turmeric rhizomes exert inhibitory action against enzymes related to type 2 diabetes (71). A combination of Curcuma longa and Allium sativum at 2.4 g total dose showed prominent antihyperglycemic action in type 2 diabetic patients by reducing fasting blood glucose, 2 h postprandial glucose, HbA1C and body mass index levels without showing any side effects (74). In addition, in six clinical trials, treating type 2 diabetic patients with curcuminois ranging from 0.25 g to 1 g per day also ameliorated glycemic control by decreasing fasting blood glucose, HbA1C, HOMA-IR (insulin resistance) levels and increasing adiponectin level without causing any major side effects. Improvement in diabetes-associated endothelial dysfunction and hyperlipidemia was observed as well (75).

**Cryptolepis sanguinolenta (LINDL.) SCHLTR.**

Cryptolepis sanguinolenta, (Lindl.) Schltr. (family: Apocynaceae) is a scrambling thin-stemmed shrub indigenous to West Africa which is commonly found in tropical rainforests, thickets, and mountainous ecologies (76). It was found to reduce the intestinal absorption of glucose and its transport from the gut significantly in a dose-dependent mode in the normoglycemic rats (76, 77). The study also revealed that treatment with C. sanguinolenta increased the size of β cells which might have improved the production and activity of insulin resulting in reduced blood glucose levels. Additionally, C. sanguinolenta also increases the uptake of glucose by 3T3-L1 cells, and improves insulin-mediated disposal of glucose. The hypoglycemic activity exhibited by the extract of C. sanguinolenta may be due to the presence of its alkaloid constituents. It has been reported that insulin resistance is reduced by alkaloids in mice and high fat-fed rats. They can activate AMP-activated protein kinase in 3T3-L1 adipocytes and L6 myotubes and promote the translocation of GLUT4 in L6 myotubes in a manner that is independent of phosphatidylinositol 3-kinase. Besides, they also improve the uptake of glucose in HepG2 and 3T3-L1 cells. In addition, they probably suppress the activity of the α-glucosidase enzyme and cause a reduction in the absorption of glucose. It has also been reported to significantly reduce the plasma levels of IL-6 with increased insulin sensitivity. This mechanism of action might also be responsible for the hypoglycemic activity of alkaloids-containing C. sanguinolenta stem extract (77). Cryptolepine, an indoloquinoline alkaloid purified from C. sanguinolenta, was found to reduce plasma glucose level significantly in a mouse model of diabetes, and in that model, it was approximately as effective as ciglitazone. It was suggested that cryptolepine works directly at the cellular level to enhance the glucose transport in 3T3-L1 cells and thus causes a reduction in blood glucose level (78).

**Euclea undulate THUNB. VAR. MYRTINA**

Euclea natalensis Thunb. var. myrtina (family: Ebenaceae) is a multi-stemmed, dioecious shrub or little tree growing up to about 6 m height. It is distributed in Botswana, Zimbabwe, Namibia, Swaziland, Mozambique and South Africa. Its crude acetone root bark extract was found to show antidiabetic activity in type 2 induced diabetic rat models (79). Besides, the plant extract of E. undulata was found to inhibit α-glucosidase and α-amylase activity (80). Past studies have revealed the presence of epicatechin and α-amyrin-3O-β-(5-hydroxy) ferulic acid in the crude acetone extract of the root bark of E. undulata. It has been reported that epicatechin may have the ability to lower blood glucose levels and α-amyrin-3O-β-(5-hydroxy) ferulic acid can inhibit α-glucosidase (81).

**Gymnema sylvestre R. BR.**

Gymnema sylvestre R. Br. (family: Asclepiadaceae) is an evergreen, woody climber and endogenous plant which is widely available in central and southern India and in the southern part of China, Sri Lanka, Malaysia and tropical Africa, Malaysia (82). G. sylvestre can improve average blood glucose levels in animal models and can stimulate insulin secretion from the MIN-6, HIT-T15 and RINm5F β-cells by upraising membrane permeability (83). Gymnema sylvestre is thought to act by several mechanisms including regeneration of islet cells, increase in the secretion of insulin and glucose utilization by insulin-dependent pathway, increase the phosphorylase enzyme activity, decrease in gluconeogenic enzymes and sorbitol dehydrogenase, reduction of glucose absorption from the gut wall (84). Antihyperglycemic compounds like gymnemagenin and gymnemic acids were discovered by LC/MS analysis from the ethanol extract of G. sylvestre leaves which demonstrated blood glucose level lowering activity in rat models (83). Gymnemic acid is a complex mixture of several (more than seventeen) saponins which are mainly dammarene and oleanane. 3-O-β-D-glucopyranosyl (1-6)-β-D-glucopyranosyloleanolic acid 28-O- β-D-glucopyranosyl ester, longispinogenin 3-O-β-D-glucuronopyranoside, oleanolic acid 3-O-β-D-xylopyranosyl(1-6)-β-D-glucopyranosyl(1-6)-β-D-glucopyranoside and 3-O-β-D-glucopyranosyl(1-6)-β-D-glucopyranosyl oleanolic acid 28-O-β-D-glucopyranosyl ester and 21-β-benzoylitsatiskogenin 3-O-β-D-glucuronopyranoside 3-O-β-D-xylopyranosyl (1-6)-β-D-glucopyranosyl (1-6)-β-D-glucopyranosyl oleanolic acid 28-O-β -D-glucopyranosyl ester are contributing to major oleanane-
triterpene glycosides. There are also seven novel dammarane saponins from the leaf extract of *G. sylvestre* known as gymnemasides I-VII. The introduction of gymnemic acid IV can decrease whose efficacy is comparable to the commercially available drug glibenclamide. A recent study also revealed that crystallographic investigation of gymnemagenin certainly indicated its good gelling with the target protein's crystallographic constitution (aldose reductase, dipeptidyl peptidases, fructose 1,6-bisphosphate, glucokinase, 1β-hydroxysteroid dehydrogenase, cytochrome 450, protein kinase B, tyrosine phosphatases, Insulin receptor substrate, cholesteryl ester transfer protein, glutamine fructose-6-phosphate amidotransferase, AMP-activated protein kinase and Glucose transporter) which contribute to its carbohydrate management property (83). In addition, the leaf extract of *Gymnema sylvestre* at 400 mg b.i.d dose lessened fasting blood glucose levels by 11%, postprandial blood glucose levels by 13% and HbA1c value by 0.6% in a 3 month long open-label trial consisting of 65 type 2 diabetic patients (85). Such lessening of the fasting blood glucose levels and post-prandial blood glucose levels were additionally consolidated by two other open-label trials on diabetic patients where the leaf extract of *G. sylvestre* were administered in the dose of 6–10 g for 15–21 days (86, 87). Moreover, in a 18-20 month-long controlled, open-label study on 22 subjects suffering from type 2 diabetes, 400 mg/day supplementation of *G. sylvestre* leaf extract exhibited a better reduction in blood glucose, glycosylated haemoglobin and glycosylated plasma protein levels compared to conventional therapy of tolbutamide or glibenclamide alone. Interestingly, of the total 22 patients participating in the trial, 5 patients could maintain blood glucose homeostasis with 400 mg/day dose of *G. sylvestre* alone even after ceasing their conventional drug therapy (88). A similar open-label study on 27 type 1 diabetic patients observed that the leaf extract supplementation reduced the levels of glycosylated plasma protein and serum amylase and increased serum C-peptide levels in contrast to the conventional therapy alone through the possible mechanism of regenerating the residual beta cells of the pancreas (89).

**Gynura divaricate (L.) DC.**

*Gynura divaricate* (L.) DC. (family: Asteraceae) is a traditional Chinese herbal plant locally known as ‘Bai Bei San Qi’. It also cultivates in the eastern and northern Taiwan coasts though is widely found in various part of Asia (90). *G. divaricate* can improve glucose metabolism in animal models including mice and rats. A study also revealed its low toxicity profile in both *in vivo* and *in vitro* testing along with significant reduction of fasting serum glucose and improved pancreatic damage (91). Among hypoglycemic phytoconstituents of the plant aerial part, a few major compounds are nystose, β-D-fructofuranose, 1-kestose, sucrose and 1F-β-fructofuranosylnystose which are fructooligosaccharides. The hexose transport assay showed that Nystose delivered the most powerful hypoglycemic activity among these five isolated phytocompounds (92). PKM1/2, PI3K, p-AKT, and GLUT4 play a vital role in the insulin signaling pathway in diabetes. High blood glucose-induced cell death and senescence in nucleus pulposus cells are inhibited by the activation of the PI3K/AKT signaling pathway. PKM1/2 is involved in glycolysis, and GLUT4 is a glucose transporter which is present on the cell membrane. Past studies also suggested that increasing GLUT4 expression promotes the glucose uptake and utilization. It has been found that *Gynura divaricate* increases the expression levels of PKM1/2, PI3Kp85, p-AKT, and GLUT4 resulting in the reduction of blood glucose levels (93).

**Hordeum vulgare L.**

*Hordeum vulgare* L. (family: Poaceae) is an annual herbaceous monocotyledonous grass commonly known as ‘Barley’ which originated in the Fertile Crescent including Israel, Jordan, Syria and southern Turkey to Zagros Mountains in Iran (94, 95). It contains high levels of dietary fiber such as β-glucans whose oral administration into type 2 diabetic and high-fat diet induced obese mice resulted in a significant lowering of blood glucose level (96). The underlying mechanism is thought to be the suppression of sodium-glucose transporter-1 expression in the intestinal mucosa. Besides, it also promotes glycogen synthesis and inhibits fat accumulation in the liver, and depresses macrophage infiltration and the production of pro-inflammatory cytokines (97). It was also found to promote glucose uptake and reduce gluconeogenesis by downregulating some genes responsible for gluconeogenesis. Barley is also rich in magnesium acting as a co-factor for more than 300 enzymes as well as for those which are involved in glucose metabolism and insulin secretion. It has been observed that regular consumption of whole grains can lower the risk of type II diabetes by 31%. This could be due to its high fiber content (98). β-glucans from barley also improved glycemic control in diabetic patients, mainly by abating postprandial blood glucose levels, according to multiple clinical studies. It has been suggested through clinical trials that high β-glucans containing foods have lower glycemic index (GI) and diabetic people should substitute the high-GI foods in their diet with low-GI foods in order to control the disease (99, 100).

**Larrea tridentata (SESSE & MOC. EX DC.) COVILLE**

*Larrea tridentata* (Sessé & Moc. ex DC.) is a highly branched and evergreen shrub from Zygophyllaceae which is widely available in North American warm deserts and Mexico (101). It contains Masoprocol, a lipoxygenase inhibitor as the major antihyperglycemic compound which decreased plasma glucose level in type-2 diabetic mice models without changing the concentration of plasma insulin. Additionally, it has been shown to improve oral glucose tolerance and enhance insulin activity in lowering the plasma glucose levels in masoprocol-treated db/db mice (102).
**Lobelia chinensis** LOUR.

*Lobelia chinensis* Lour. (family: Campanulaceae), known as 'Asian Lobelia' or 'Chinese Lobelia' is distributed throughout China, Taiwan, Korea, and Japan. It is a plant from which two new pyrrolidine-type alkaloids, radicamines A and radicamines B, were found to inhibit α-glucosidase activity and demonstrate antidiabetic efficacy (103). Besides, it has also been reported that the active ingredients of 5-hydroxymethylfural and acacetin in *L. chinensis* has been shown to promote the secretion of insulin, improve insulin resistance, and stimulate the utilization of glucose by acting on GSK3B, MAPK, INR, and dipeptidyl peptidase-4 (DPP4) (104).

**Lupinus perennis L.**

*Lupinus perennis* L. (family: Fabaceae) is a perennial herb which is abundantly found in Canada and USA (105). It is a plant from the leaves of which the compounds lupanine, 13-a-OH lupanine, and 17-oxo-lupanine were extracted and enhanced the secretion of insulin from isolated rat islets in a glucose-dependent manner. It was assumed that these quinolizidine alkaloids can increase insulin release by reducing K+ permeability in the β-cell plasma membrane. The fact that 13-a-OH lupanine and 17-oxo-lupanine stimulate insulin secretion only at high glucose concentrations indicates that it would reduce the risk of hypoglycemia which could be of additional value when considering their potential use in the treatment of type 2 diabetes (106).

**Matricaria chamomilla L.**

*Matricaria chamomilla* L. (family: Asteraceae), commonly known as 'German chamomile', 'Hungarian chamomile' or 'wild chamomile', is a herbaceous plant natively distributed in European and West Asian regions (107). Its aerial part’s ethanol extract dose-dependently lessened postprandial blood glucose levels and showed protective action on pancreatic β-cells of streptozotocin (STZ)-induced diabetic rats. The same study also reported a significant reduction in oxidative stress related to hyperglycemia (108). A similar study on STZ-induced diabetic rats showed that a 21 day long 200 mg kg⁻¹ body weight dose of *M. chamomilla* leaf extract significantly diminished the fasting blood glucose levels by 62.2% (109). HbA1C and blood glucose values were also reduced in STZ-induced female fertile diabetic rats after administering the plant’s aerial part-derived ethanolic extract (110). In addition, another animal study reported the effectiveness of *M. chamomilla* flower extract and the isolated compounds quercetin, esculetin, umbelliferone and luteolin in preventing the progression of hyperglycemia. It was observed that quercetin and esculetin moderately inhibited the enzymatic activity of sucrase in rats and all the compounds halted sorbitol from accumulating in the erythrocytes of humans. Quercetin and the hot water extract also suppressed blood glucose levels in a 21 days long feed test on STZ-induced diabetic rats. Furthermore, esculetin diminished hyperglycemia in a dicarboxylic loading test on mice. It was also reported that the extract exhibited good inhibitory activity against the aldose reductase enzyme (111). Apigenin, which is another compound isolated from the plant, showed antihyperglycemic action by causing increment in blood insulin and diminution in blood glucose levels in alloxan induced diabetic mice (112). In an 8 week long randomized controlled trial on 64 type 2 diabetic patients (12 were male and 52 were female; age range: 30-60 years), chamomile tea in the dose of 3 g/150 mL thrice a day showed a significant diminution in the HbA1C level by 5.01%, HOMA-IR level by 39.76%, and serum insulin level by 32.59% compared to baseline values and caused improvement in antioxidative activity (113). Another 4 week long randomized-controlled trial on 50 type 2 diabetic patients observed that twice-daily infusion of 10g/100 ml chamomile as a supplementation significantly improved glycemic control by lessening fasting blood glucose and 2h postprandial blood glucose levels. Moreover, lipid profile was also ameliorated in the patients (114).

**Momordica charantia L.**

*Momordica charantia* L. (family: Cucurbitaceae) is a flowering vine cultivated in Asia including Bangladesh, India and in other regions like East Africa and South America as well. The fruit has a distinct bitter taste for which this plant is known as bitter gourd which is also well known as ‘Korolla’, ‘Karela’, ‘Bitter melon’ or ‘Balsam pear’. The bitterness becomes more intensified when it ripens. The plant produced notable antidiabetic and hypoglycemic actions which ascertain the adjuvant use of the plant along with conventional commercialized drugs (115). The oral consumption of the juice of *M. charantia* seeds showed prominent hypoglycemic activity in streptozotocin induced type 1 diabetic rat models. There are many bioactive phytocompounds isolated from *M. charantia* producing remarkable antidiabetic activity. Among saponins, 3-hydroxycucurbita-5, 24-dien-19-al-7, 23-di-O-β-glucopyranoside and Momordicine- I were extracted from corolla exhibiting promising insulin-releasing properties in MIN6 β-cells. Charantin, a cucurbitane type triterpenoid extracted from the same plant has also showed tremendous antidiabetic activity which is even more potent than standard oral hypoglycemic drug tolbutamide (115). Polypeptide-p or p-insulin, insulin-like hypoglycemic protein type substance which demonstrated blood glucose-lowering activity in human upon subcutaneous administration was isolated from corolla. The insulin mimicking activity of Polypeptide-p can be considered as a plant based alternative of insulin in type 1 diabetic patients (115). Vicine, a glycol alkaloid is another isolated compound from *M. charantia* which can promote hypoglycemia in non-diabetic fasting rat models upon intraperitoneal administration (115). Among other isolated compounds, Momordicoside U showed moderate activity during in vitro insulin secretion property screening and 5β,19-epoxy-3 β,25-dihydroxycucurbita-6,23(E)-
diene and 3 β,7 β,25-trihydroxycucurbit-5,23(E)-dien-19-α both revealed hypoglycemic activity in diabetes induced male mice models (116, 117). A meta-analysis study of ten clinical trials conducted in 1045 type 2 diabetic patients observed that M. charantia possessed significant glycemic control improving ability since it lessened FPG, HbA1c and PPG levels without causing any side effects. In addition, prediabetic subjects also had a reduction in their FPG levels (118, 119). Such action is reported to occur because of increased insulin secretion, as per another randomized-controlled trial. Moreover, anthropometric parameters, namely weight, body mass index, waist circumference, fat percentage were also decreased (120). Another trial on maturity-onset diabetic patients reported the antihyperglycemic action of M. charantia since it decreased mean blood glucose levels in fasting conditions as well as at 1, 2 and 12 hours after oral intake of 50 g glucose (121).

**Moringa oleifera LAM.**

*Moringa oleifera* Lam. (family: Moringaceae) is a perennial angiosperm plant native to Asia and greatly found in Malaysia and other tropical countries. Local name of *M. oleifera* is ‘Sajna’, ‘Soanjna’ and ‘Sojanjna’ and the English name is ‘Drum stick tree’ (122). It is a plant whose leaf’s alcoholic extract along with its antidiabetic phytocompounds like flavonoids, alkaloids, tannins, steroids and glycosides is assumed to be effective to treat diabetic complications. Quercetin and kaempferol, two major phytoconstituents, isolated from *M. oleifera* notably reduced serum glucose (33.34%) along with augmentation in serum insulin level when introduced to diabetic rat models for four weeks (123). In another study, moringinine, quercetin and chlorogenic acid, notable phytochemicals extracted from this plant were introduced to diabetic rat models to evaluate antidiabetic efficacy. The outcome showed alleviated serum glucose, total cholesterol and triacylglycerol level at a dose of 150 mg/kg after 21 days of care. In addition, in diabetic rats, it also restored the normal histological structure of the pancreas (124). Past studies have indicated that the glucose uptake in the rat soleus muscle is stimulated by kaempferol via the PI3K and PKC mechanisms. When administered orally, kaempferol was found to reduce fasting blood glucose levels significantly and serum HbA1c levels besides improving insulin resistance. Additionally, Quercetin blocks the transport of fructose and glucose by GLUT2 in the brain and promotes the translocation and expression of GLUT4 in skeletal muscle (125). In addition, in a randomized control design trial, promising inhibition in the increment of serum glucose after 2h of 75g oral glucose intake was observed after taking capsules processed with *M. oleifera* leaf in a population pool of 18-55 years old (126). The efficacy of *M. oleifera* in twenty diabetic and ten healthy individuals of 35 to 60 years old was evaluated in another clinical trial. The concentration of glucose, triglycerides, glycosylated hemoglobin, total cholesterol and low-density lipoprotein cholesterol decreased significantly while high-density lipoprotein cholesterol was upraised. This hypoglycemic efficacy has been assumed to be attributed to phenols, tannins, flavonoids, alkaloids and carotenoids (122). In another study, an assessment was conducted to evaluate the ability of *M. oleifera* leaf powder to inhibit the activity of α-amylase in *vitro*. The study found that *M. oleifera* leaf powder decreased α-amylase enzyme activity by 68.2 ± 3.2% (127). The study also further evaluated in *vivo* activity of the leaf powder on postprandial blood glucose levels in the subjects of Saharawi refugee camps (17 diabetic and 10 healthy individuals). The study displayed that administration of 20g leaf powder improved postprandial glycemic index at 90, 120 and 150 min. as well as improved the mean glycemic index in diabetic patients compared to the control group which indicates the candidacy of *M. oleifera* as an antihyperglycemic herbal drug (127).

**Morus alba L.**

*Morus alba* L. (family: Moraceae), known as ‘Mulberry’ is a quickly growing tree growing as long as 20 m. It is native to China though cultivated sporadically in Japan and Korea (128). Three major compounds, namely Moracin M, steppogenin-4′-O-β-D-glucosiade and mulberroside A showing efficacy in alloxan induced mice models by demonstrating hypoglycemic efficacy and decreasing fasting blood glucose level were isolated from the plant (129). Moreover, the alkaloids extracted from the leaves of mulberry were found to exhibit hypoglycemic effects in streptozotocin- (STZ-) induced diabetic mice. It has been reported that 1-deoxynojirimycin (DNJ), a mulberry alkaloid, reduces the activity of α-glucosidase by competitive inhibition. Upon oral administration of starch and sucrose in Kunming mice, flavonoids from mulberry leaf reduced blood glucose level and inhibited α-glucosidase activity. In the laboratory experiment, two flavonoids (isoquercitrin and astragalin) were found to inhibit α-glucosidase activity. Polysaccharides isolated from the leaves of mulberry were reported to reduce plasma glucose level, improve glucose tolerance, increase the hepatic glycogen content, and inhibit α-glucosidase activity. The extracted polysaccharides α-arabinose, α-xylitol, α-glucose, α-rhamnose, and α-mannose were found to repair pancreatic β-cells, resulting in increased insulin secretion and reduced accumulation of liver fat in diabetic rats (130). A randomized, placebo-controlled study on 10 type 2 diabetic (age range: 59–75 years) and 10 healthy subjects (age range: 24–61 years) involving the ingestion of 1 g leaf extract of *Morus alba* showed remarkably reduced blood glucose levels after 2 hours in comparison to the placebo group (131). Another trial showed the effectiveness of the leaf extract in suppressing insulin and postprandial blood glucose levels (132). In addition, fasting blood glucose was better reduced by *M. alba* compared to glibenclamide in 24 type 2 diabetic patients (24 male; age range: 40–60 years), as per a 30 day long randomized controlled trial (133). Moreover, a 25% inhibition in carbohydrate absorption of healthy subjects was observed in another crossover trial, suggesting that *M. alba* could also be
used as a supplementation in the treatment of type 2 diabetes (134). 1-Deoxynojirimycin (DNJ) from the leaves of *M. alba* in 0.8 and 1.2 g doses also notably reduced the postprandial blood glucose levels and insulin secretion of 24 healthy subjects (mean age: 25.3 ± 0.7 years) in a randomized controlled trial. Such efficacy advocates for the use of 1-Deoxynojirimycin as a dietary supplement in the treatment of diabetes mellitus (135).

**Nelumbo nucifera GAERTN.**

*Nelumbo nucifera* Gaertn. (family: Nelumbonaceae), commonly known as ‘Chinese water lily’, ‘Indian lotus’ and ‘Sacred lotus’, is a large aquatic rhizomatous perennial plant (136). Nuciferine, an alkaloid was extracted from the plant through identifying by NMR spectroscopy and was found to increase insulin secretion in both isolated islets and INS-1E cells. It was found that nuciferine stimulates both the first phase and the second phase of insulin secretion. These results indicated that the nuciferine acts by closing K-ATP channels and also through stimulation of K-ATP channel-independent amplification pathways. Besides, it shows less cytotoxicity than Glibenclamide (137). In addition, *N. nucifera* seeds achieved a low glycemic index (GI) in a randomized crossover trial on healthy subjects (138).

**Nigella sativa L.**

*Nigella sativa* L. (family: Ranunculaceae) is a herbaceous plant which occurs in several southern Mediterranean and Middle Eastern countries. The seeds of *Nigella sativa* are known by the name ‘Black seed’ or ‘Black cumin’. Its seed’s ethanolic extract was found to enhance insulin secretion, stimulate proliferation of pancreatic β-cells, and enhance glucose uptake in muscle and fat cells (139). It has been suggested that *Nigella sativa* exhibits its hypoglycemic effect due to the presence of thymoquinone, dithyomoquinone, linoleic acid and oleic acid which might be responsible for stimulating pancreatic β-cells causing insulin secretion, reducing hepatic gluconeogenesis, and inducing insulin sensitivity in peripheral tissue. A placebo-controlled participant blinded clinical study on 114 type 2 diabetic patients found that *N. sativa* supplementation plays an important role in the amelioration of oxidative stress, the latter being responsible for diabetes mellitus pathogenesis. It has been found that the former does so by improving total antioxidant capacity, glutathione and superoxide dismutase values. The same study perceived that reduction in insulin resistance was also significant in the diabetic patients in the group taking *N. sativa* supplementation compared to the placebo group (140). Supplementation of *Nigella sativa* in type 2 diabetes patients was found to improve fasting blood glucose, HbA1c, total-cholesterol, and LDL level significantly (33). As such, a systematic review done to assess the effect of *N. sativa* on type 2 diabetes mellitus has also suggested that it could be adjunctively used with other oral antidiabetic medications to manage the disease (141).

**Panax ginseng C. A. MEYER**

*Panax ginseng* C. A. Meyer (family: Araliaceae) is also known as ‘Asian ginseng’. It is a plant that has been reported to modify blood glucose levels by increasing insulin sensitivity, ameliorating the function of pancreatic β-cells, and stimulating glucose uptake by elevating the production of glucose transporters (GLUT). The berry extract of *P. ginseng* stimulates the β-cell proliferation leading to increased insulin secretion to control the level of blood glucose in streptozotocin (STZ) -induced diabetic mice. Besides, it also results in improved sensitivity to insulin in C57BL/6 mice over 15 months old (142). Ginseng is known to contain ginsenosides, a group of steroidal saponins, including neutral ginsenosides and malonyl ginsenosides. Ginseng and neutral ginsenosides were found to lower blood glucose, increase insulin sensitivity, regulate lipid metabolism, and reduce body weight (143, 144). Administration of malonyl ginsenosides in high fat diet/streptozotocin diabetic rats was found to remarkably reduce fasting blood glucose levels, improve glucose tolerance and insulin sensitivity without affecting body weight (143, 144). The study results suggest that malonyl ginsenosides could be used to treat type-2 diabetes. In addition, a meta-analysis of sixteen clinical trials revealed that Ginseng caused a significant reduction in fasting blood glucose levels in both diabetic and non-diabetic patients, while the other glycemic parameters were contradictory in terms of outcomes (145). Besides, a randomized, placebo-controlled, double-blind study in 20 type 2 diabetic patients (mean age: 51.5 ± 1.9 years) observed about 45% reduction in HOMA-IR in the group which was given *P. ginseng* supplementation compared to 12% reduction for the placebo group (146). Moreover, combinatory administration of *Panax ginseng* and *Panax quinquefolius* in a 12 week long randomized controlled trial on 80 type 2 diabetic subjects (49 were male and 31 were female) reduced another glycemic control parameter, namely HbA1c by −0.35 ± 0.1% along with blood lipid parameters (total cholesterol, LDL-C, triglycerides) and 24-hour systolic blood pressure without causing any side effects (147).

**Pandanus amaryllifolius ROXB.**

*Pandanus amaryllifolius* Roxb. (family: Pandanaceae) is a shrub native to Thailand. This plant is also known as *Pandanus odorus*. Local name of the plant is ‘Tœi-hom’ (148). In recent studies, *P. amaryllifolius* exhibited very prominent hypoglycemic activities. The root extract of the plant evidently lowered blood glucose levels in streptozotocin induced mice models. The leaf extract could also show promising antihyperglycemic activity by stimulating insulin production and glucose uptake along with inhibition of α-glucosidase enzyme (148). 4-Hydroxybenzoic
acid is the major hypoglycemic phytoconstituents isolated from the aqueous extract of this plant root. Upon oral administration, this moiety upraised serum insulin and liver glycogen level in normal rat models (149). A clinical study performed on 30 healthy subjects (15 were male and 15 were female; age range: 15-25 years) showed that intake of 30 g of *P. amaryllifolius* containing tea caused a significant lessening of postprandial blood glucose (148).

**Punica granatum L.**

*Punica granatum* L. (family: Punicaceae) locally known as ‘Dalim plant’ is a plant native to some Asian countries including India, Bangladesh, Iran and Malaysia along with some other countries of America and European continents, such as United States of America, Peru, Turkey, etc. The fruit of the plant (commonly known as ‘Dalim’ in Asian countries) is popularly consumed as fresh form along with processed juice, jam, paste and wine (150). The fruit aqueous extract of *P. granatum* can notably decrease fasting glucose level along with promising increment in the expression levels of Glut-4, Glut-2, Akt and IRS-1 followed by improved glucose uptake and its storage in alloxan induced male Wistar rat models (151). It is blessed with several polyphenolic compounds like punicalagin, valoneic acid dilactone, anthocyanin, phenolic and non-phenolic Acids, Glutenins and Tannins (151). Among those, valoneic acid dilactone is the main antidiabetic principle which showed its antidiabetic efficacy by inhibiting the activity of aldose reductase enzyme in a dose dependent pattern. Protein tyrosine phosphatase 1B (PTP1B) was also inhibited by valoneic acid dilactone which can also ameliorate the level of blood glucose in alloxan induced diabetic rat models. Other possible mechanisms attributed to antidiabetic action of valoneic acid dilactone may be attributed to improved insulin secretion from rom pancreatic β-cells or its release from the bound form along with insulin-mimetic actions or amended glucose utilization technique (150). In addition, an 8-week long double blind randomized-controlled clinical study on 52 type 2 diabetic patients with obesity (26 were male and 26 were female; age range: 30-50 years) found that supplementation of *P. granatum* significantly reduced fasting blood glucose from 161.46 mg/dl to 143.50 mg/dl. An increase in GLUT-4 gene expression was also observed in the patients (152). Furthermore, another single-blind, randomized-controlled clinical study conducted on 44 type 2 diabetic patients (23 were male and 21 were female; age range 56 ± 6.8 years) found that the juice of *P. granatum* significantly ameliorated oxidative stress, suggesting that its consumption could retard the onset of oxidative stress associated diabetes mellitus (153).

**Pongamia pinnata (L) PIERRE**

*Pongamia pinnata* (L.) Pierre (family: Fabaceae) is a medium-sized glabrous tree, commonly known as ‘Karanja’ in Hindi, ‘Indian Beech’ in English, and ‘Pongam’ in Tamil. It occurs throughout India, and mainly found in tidal forests of India. The flowers of *P. pinnata* reportedly possessed anti-hyperglycemic and anti-lipidperoxidative properties (154). It has been found that oral administration of the aqueous (PPAE) and ethanolic (PPEE) extracts of the leaves of *P. pinnata* PPEE in alloxan diabetic rats resulted in a pronounced reduction in the plasma glucose level. This may be attributed to the enhancement of the effect of insulin by increasing the insulin release from the pancreatic β-cells or its release from the bound insulin. The significant glucose-lowering effect of PPAE and PPEE could also result from increased peripheral glucose utilization (155). Karanjin, one of the isolated compounds from this plant was reported to possess hypoglycemic activity in normal and in alloxan-induced diabetic rats. Pongamol and karanjin extracted from the chloroform-soluble fraction of the ethanolic extract of *P. pinnata* fruits exhibited significant glucose-lowering activity (156). The underlying mechanism of anti-hyperglycemic activity of the compounds may be attributed by the inhibition of PTPase-1B, a major mediator of insulin signaling and insulin resistance (156).

**Psacalium peltatum (KUNTH) CASS.**

*Psacalium peltatum* (Kunth) Cass. (family: Asteraceae) is a plant natively grown and known as ‘Matarique’ in Mexico. Petaltosa, an ulopyranose compound, has been obtained from the roots and rhizomes of *P. peltatum* which has shown anti-hyperglycemic activity on mice with mild diabetes, although the efficacy decreased on mice models with severe diabetes (157). The hypoglycemic effect of the compound was reported to be similar to tolbutamide, and the possible underlying mechanism of action could be attributed to enhanced secretion of insulin from the islets of Langerhans or an increased utilization of glucose by peripheral tissues (157).

**Silybum marianum (L.) GAERTN**

*Silybum marianum* (L.) Gaertn (family: Asteraceae), also known by the common name ‘Milk thistle’ is an annual or biennial herb native to the Mediterranean regions of Europe, North Africa and the Middle East and in some parts of USA (158, 159). Its major component is silymarin, a mixture of silybins (silybins A and B), isosilybin (isosilybins A and B), silychristin and silydianin (160). Administration of silymarin in patients with type 2 diabetes resulted in a significant reduction in HbA1c level, fasting plasma glucose (FPG), daily blood glucose average and glucosuria, daily insulin requirement, fasting insulin, as well as an increase in serum glutamic oxaloacetic transaminase (SGOT), serum glutamic pyruvic transaminase (SGPT) and HDL levels. When silymarin was supplemented with glibenclamide in type 2 diabetes patients, a reduction in postprandial hyperglycemia was also observed (33). A flavonolignans, silychristin A, extracted from *Silybum marianum* demonstrated a marked reduction of both postprandial and/or fasting hyperglycemia.
and improvement of the function of β-cells in STZ-induced T1DM. It has been suggested that silychristin A exerts its glucose-lowering effect by protecting the β-cells from oxidative stress-induced damage and blocking the activity of the α-glucosidase enzyme (159).

**Swertia chirayita BUCH HAM.**

Swertia chirayita Buch Ham., (family: Gentianaceae) locally named as ‘Chirayata’, ‘Chirayta’ and ‘Chireetta’ is a popular medicinal herb native to temperate Himalayan region (161). Promising antidiabetic efficacy of *S. chirayita* with improved insulin secretion was reported during cell line based evaluation technique using insulin secretion from monolayers of BRIN-BD11 clonal pancreatic cells (162). There are several antidiabetic compounds found in this species exerting prominent antidiabetic efficacy. According to 163, 1,5,8-trihydroxy-3-methoxyxanthone extracted from the aerial parts and roots of the *S. chirata* can demonstrate antidiabetic efficacy by lowering blood sugar levels (163). Gentianine, another antidiabetic compound of this plant is the active metabolite of swertiamarin and is believed to attribute to the efficacy of swertiamarin (164). Promising amelioration in adipogenesis associated expression of PPAR-γ, GLUT-4 and adiponectin by gentianine administration expressed that the compound is responsible for antidiabetic efficacy of swertiamarin. Magniferin, a potent phytoconstituent found in *S. chirata* can also exhibit antihyperglycemic potency by exhibiting glucosidase and 2,2-diphenyl-1-picrylhydrazyl radical inhibition action. Besides, as a co-therapy with metformin and gliclazide it cured renal injury symptoms due to diabetic neuropathy (165). Moreover, compounds, found in *S. chirata* like amarogentin is used in the preparation of different forms of commercially available drugs to treat diabetic complications (166). A 30 day long clinical study done on 12 type 2 diabetic patients found that ingestion of *S. chirayita* in grounded powder form caused 14.5% reduction in blood glucose level. A reduction was also noticed in lipid profile (total cholesterol by 8.6%, LDL-c by 14.4% and, tryglycerides by 10.5%) (167).

**Syzygium cumini (L.) SKEELS**

*Syzygium cumini* (L) Skeels (family: Myrtaceae) is known as ‘Jamun’, ‘Jambul’ and ‘Jambol’ in India and Malaya. The seeds of *S. cumini* are thought to lower blood sugar levels by increasing either insulin secretion from β-cells of the islets of Langerhans of Pancreas or its release from the bound form. Mycaminose, isolated from the seeds of *S. cumini* was found to produce a remarkable reduction in blood glucose level (168). It was suggested that the mode of action of mycaminose is similar to Glibenclamide, a commercially available anti-diabetic drug (168). In a double-blind randomized controlled trial involving 99 type 2 diabetic patients, 10 g daily *Syzygium cumini* supplementation significantly lessened fasting blood glucose by 30%, post-prandial blood glucose by 22% and HbA1c value from 8.99 ± 1.39% to 8.31 ± 1.40% after 90 days (169).

**Tinospora cordifolia (WILLD.) MIERS**

*Tinospora cordifolia* (Willd.) Miers (family: Menispermacae) is commonly known as ‘Guduchi’ or ‘Amrita’ and is found in the Indian subcontinent and China. It has exhibited blood and urinary glucose-lowering activity along with suppression in the increase of blood glucose level in animal models (170, 171). It is considered as an antidiabetic herbal drug in the Indian Ayurvedic Pharmacopoeia too due to its alkaloids, diterpenoids and glycosidic constituents. Among the alkaloids, magnoflorine was found to be the most potent α-glucosidase inhibitor (172). Besides, a norclerodane diterpenoid, tinosporaside, extracted from *T. cordifolia* possessed 28% antihyperglycemic activity when it was compared with Metformin 20.6% in diabetic rat models (173). It has been found that the isoquinoline alkaloid rich fraction (AFTC) isolated from the stem of *T. cordifolia* (AFTC) significantly reduced the synthesis of glucose in rat hepatocytes like insulin did and it also stimulated secretion of insulin in RINm5F cells like tolbutamide. The underlying mechanism may be attributed to the promotion of insulin release and insulin-mimicking activity (174). In addition, the powdered stem of *T. cordifolia* at the oral dose of 50 mg/kg of body weight significantly reduced the fasting blood glucose and HbA1c levels by 9% and 14%, respectively in type 2 diabetic patients (175).

**Trigonella foenum-graecum L.**

*Trigonella foenum-graecum* L. (family: Fabaceae) known as Fenugreek or Methi is a legume and a popular seasoning worldwide to improve the taste and flavor of food (176). Its seed water-soluble compound GII extract when administered for 15 days in the subdiabetic and moderately diabetic rabbits and for 30 days in the severely diabetic rabbits resulted in the elevation of hepatic and muscle glycogen content, stimulated hexokinase, glucokinase, pyruvate kinase, malic enzyme, glucose-6-phosphate dehydrogenase, superoxide dismutase, glutathione peroxidase, and reduced the activity of glucose-6-phosphatase, sorbitol dehydrogenase, aldose reductase. Partially damaged pancreatic cells were also regenerated following the administration of GII (177). Trigonelline, nicotinic acid and coumarin are antidiabetic phytochemicals that were isolated from Fenugreek seed. These three antidiabetic compounds extracted from fenugreek showed prominent efficacy in alloxan induced severe and moderate diabetic rabbit models (178). A meta-analysis on 10 clinical trials conducted on type 2 diabetic patients revealed that fenugreek can significantly improve glycemic control by altering parameters, namely fasting blood glucose level by -0.96 mmol/l, HbA1c value by -0.85% and 2 hour postprandial glucose level by -2.19 mmol/l (179). As per a 2 month long double blind placebo controlled trial on 25 subjects
with type 2 diabetes mellitus, hydroalcoholic extract of fenugreek seed also decreased insulin resistance compared to control which was apprehended by an increase in insulin sensitivity percentage (112.9 ± 67% vs 92.2 ± 57%, respectively) and beta-cell secretion percentage (86.3 ± 32% vs 70.1 ± 52%, respectively) through HOMA-IR test (180).

**Vitis vinifera L.**

*Vitis vinifera* L. (family: Vitaceae), known as ‘Grapevine’ or ‘Red grape’ is native to southern Europe and western Asia. However, it is cultivated worldwide which makes it the largest fruit crop in the world. It contains many active components in its seed and skin, including polyphenols, flavonoids, proanthocyanidins, anthocyanins, procyanidins, and resveratrol, a stilbene derivative (181). The *kir6.2* channel is encoded by the KCNJ11 gene. It has been shown that congenital hyperinsulinism is caused by a mutation in this gene and has a significant role in the development of type-1 diabetes. Pterostilbene has promising inhibitory efficacy on both normal and mutant models of *kir6.2* as an active component of *V. vinifera*. Again, quercetin, myricetin and resveratrol are three other most notable polyphenols found in red grapes to treat diabetic complications. Quercetin can demonstrate improved expression of adiponectin in white adipose tissue and blood concentration, despite inhibition of poly (ADP-ribose) polymerase γ expression followed by improved insulin sensitivity (182). Quercetin can also inhibit glucose uptake at the level of glucose transporters (GLUTs) (183). Besides, to treat hyperglycemia, Myricetin is also used as traditional medicine in northern Brazil. Myricetin can promote glucose uptake in the liver and soleus muscles as well as hepatic glycogen synthase (184). Myricetin can also improve insulin resistance in fructose chow-fed rat models (185). In addition, it can also halt advanced glycation end products in diabetic conditions (184). Furthermore, antihyperlipidemic and human pancreatic alpha-amylase inhibitions are a few other promising mechanisms by which myricetin can produce a significant antidiabetic effect (186). Resveratrol, a phytochemical from stilbene class of polyphenolic compounds isolated from red grape can also demonstrate strong antidiabetic activity. It can also protect against diabetic nephropathy while administration of resveratrol along with protective activities in renal dysfunction and oxidative stress (187). Resveratrol can effectively restore cellular homeostasis by activating the redox plasma membrane system, which functions as a compensatory mechanism in the cell to preserve redox status (184, 188). Besides, resveratrol administration to diabetic rats has resulted in decreased concentration of glycosylated hemoglobin (189). The antihyperglycemic efficacy of resveratrol, demonstrated in diabetic animals has been suggested to be due to the stimulatory activity on the transportation of intracellular glucose. The involvement of resveratrol can also promote glucose uptake in diabetic rat models (190). Improved expression of the insulin-dependent glucose transporter (GLUT4) was reported after ingestion of resveratrol in the study conducted on diabetic rat models (191, 192). Resveratrol was also reported to modulate the function of sirtuin-1, which ameliorates homeostasis of whole-body glucose and insulin sensitivity in diabetic rats (192). Moreover, in a 3 month long randomized controlled clinical trial on 62 subjects with type 2 diabetes mellitus (age range: 30-70 years), resveratrol supplementation exhibited antihyperglycemic action by lowering the value of hemoglobin A1c compared to the control (9.65 ± 1.54 vs 9.99 ± 1.50, respectively) (193). In another 45 days long randomized double-blinded placebo-controlled parallel clinical study on 66 subjects (age range: 20-65 years) with type 2 diabetes mellitus, 1 g daily resveratrol supplementation exhibited antihyperglycemic action by lowering the values of fasting blood glucose, hemoglobin A1c, insulin secretion, and insulin resistance compared to baseline (194). In another 30 days long randomized, double-blind, crossover trial, resveratrol supplementation lowered the response of postprandial glucagon in obese type 2 diabetic patients (195). Quercetin also reduced fasting blood glucose and insulin secretion, as per a meta-analysis of randomized controlled trials (196).

**Zingiber officinale ROSCOE**

*Zingiber officinale* Roscoe (family: Zingiberaceae) also known as ‘Ginger’ is a perennial plant with slender, brightly green, grassy leaves and yellowish green flowers which is grown in the tropics. Rhizome serves as the edible and medicinal part of the plant. Its administration to streptozotocin (STZ)-induced diabetic rats was found to reduce serum glucose, cholesterol and triacylglycerol levels significantly. Besides, raw ginger also exhibited effectiveness in reversing diabetic proteinuria in diabetic rats (197). Several active constituents have been isolated from Ginger including gingerols and their related dehydration products, the shogaols, as well as volatile oils including sesquiterpenes, such as β-bisabolene and monoterpenes, mainly geranial and neral (197). A previous study revealed that 6-shogaol and 6-gingerol can suppress the development of diabetic complications as well as advanced glycation end products (AGEs) by arresting methylglyoxal, the precursor of AGEs (198). 6-gingerol can also arrest Nε-carboxymethyl-lysine (CML), a marker of AGEs through activation of Nrf2 (199). 6-paradol and 6-shogaol facilitated glucose consumption by increasing AMPK phosphorylation in 3T3-L1 adipocytes and C2C12 myotubes. Furthermore, 6-paradol considerably decreased the concentration of blood glucose in high-fat diet-fed mouse models (200). Besides, 6-gingerol, in type 2 diabetic mice, aided glucose-stimulated insulin secretion and improved glucose tolerance by upraising glucagon-like peptide 1 (GLP-1). In addition, 6-gingerol therapy galvanized glycogen synthase 1 and increased glucose transporter type 4 (GLUT4) cell membrane presentations which amplified skeletal muscles’ glycogen storage (201). A meta-analysis study on 10 randomized-controlled clinical trials conducted on 490 type 2 diabetic subjects revealed that ginger improves glycemic control by lowering fasting blood glucose and HbA1c levels. Insulin sensitivity was also improved. Lipid profile improved as
mainly in the saliva and pancreatic juice. Inhibition of this antidiabetic actions are summerized in antidiabetic plants along with their phytochemicals and possible phytochemicals in a single study. However, a few more notable antidiabetic actions are summerized in Table 1 in addition to the aforementioned plants.

OTHER PROMISING PLANTS AND PHYTOCOMPOUNDS

Researchers suggested that there are almost 800 dietary and non-dietary plants with antidiabetic properties (77). So it is not possible to describe every plant and its isolated bioactive phytochemicals in a single study. However, a few more notable antidiabetic plants along with their phytochemicals and possible antidiabetic actions are summerized in Table 1 in addition to the aforementioned plants.

MOLECULAR MECHANISMS OF MEDICINAL PLANTS AND/OR EXTRACTED PHYTOCHEMICALS TO TREAT DIABETES MELLITUS

a. Inhibition of α Glucosidase Secreted From Brush Border of the Small Intestine

Mammalian α glucosidase, a membrane-bound hydrolytic enzyme found in the mucosal brush border of the epithelia of the small intestine plays a key role in carbohydrate digestion. Inhibitors of this α glucosidase enzymes delay the cleavage of carbohydrates resulting in reduced glucose absorption and an attenuated postprandial glycemic level. Thus, α glucosidase inhibitors could show a beneficial effect in the management of non-insulin-dependent diabetes mellitus (NIDDM) by causing a reduction in postprandial blood glucose levels (265, 266).

b. Inhibition of DPP-4 Enzyme

Glucagon like peptide-1 (GLP-1) and Glucose dependent insulinotrophic polypeptide (GIP) are incretin hormones which can initiate the differentiation of β-cells, stimulate the biosynthesis and secretion of insulin and inhibit gastric emptying. However, these hormones are rapidly broken down by a serine peptidase enzyme known as dipeptidyl peptidase-4 (DPP-4). Therefore, inhibitors of DPP-4 can be used in the treatment of type 2 diabetes. These DPP-4 inhibitors exhibit their antidiabetic activity via prolongation of GLP-1 and GIP activity, stimulation of insulin release and inhibition of glucagon secretion which ultimately leads to regulation of the blood glucose level (267, 268).

c. Inhibition of α Amylase Secreted From Salivary Gland

Hydrolysis of α-1,4-glucan polysaccharides, such as starch and glycogen is carried out by the enzyme, α-amylase which is found mainly in the saliva and pancreatic juice. Inhibition of this enzyme helps in the prevention of high postprandial blood glucose levels (269).

d. Increased Insulin Secretion

Past studies have shown that an increase in intracellular calcium ion [Ca2+]i was associated with insulin secretion. Generally, the release of insulin from the vesicles of the pancreatic β-cells is stimulated by a rise in [Ca2+]i. Membrane depolarization caused by the closure of the ATP-sensitive K+ channels of the insulin secreting β-cells leads to activation and opening of the voltage-dependent Ca2+ channels which increases the [Ca2+]i. These increased intracellular calcium levels stimulate the secretion of insulin from pancreatic β-cells. However, a few phytochemicals, for example, p-methoxycinnamic acid, has shown to increase insulin release by acting on the L-type Ca2+ channels rather than the ATP-sensitive K+ channels. This may also lead to a rise in cAMP via the inhibition of phosphodiesterase (270, 271).

e. Increased Insulin Sensitivity and Enhanced Glucose Uptake by Muscle Cells and Adipose Tissue

Some phytochemicals improve the sensitivity of non-pancreatic cells to insulin resulting in improved glycemic control. In skeletal muscle and adipose tissue, glucose uptake is enhanced via the activation of a series of events which take place following an increase in insulin levels. When insulin binds to the insulin receptors, it causes phosphorylation of protein substrates leading to the activation of phosphatidylinositol 3-kinase (PI3K) and downstream signaling through PKB/Akt and PKC-β/ζ. As a result, GLUT4, insulin-regulated glucose transporter protein is recruited to the cell membrane and an increase in the uptake of circulating glucose by the muscle cells and adipose tissue occurs via facilitated diffusion through GLUT4 transporter protein (272).

f. Nourishment of Pancreatic β-Cells

Survival, restoration and maintenance of the mass/function of pancreatic β-cells can hinder the pathogenesis of diabetes mellitus. β-cells from the pancreas secrete the hormone insulin which is crucially salient in maintaining homeostasis for glucose metabolism in the body. β-cells are impaired in type 1 diabetes due to macrophage, cytokine and T-cell mediated autoimmune reactions. In the case of type 2 diabetes, β-cells could possibly be debilitated or rendered dysfunctional due to factors like oxidative stress, enduringly elevated glucose or lipid levels, and the release of the inflammatory mediators. To inhibit such destruction, β-cells can be fortified against reactive oxygen species (ROS) accretion and lipid peroxidation mediated cell death by augmenting both non-enzymatic (e.g., reduced glutathione) and enzymatic (e.g., superoxide dismutase, glutathione S transferase, glutathione peroxidase, catalase) antioxidants which can enhance the antioxidant capacity of the cell. In addition, increment of the secretion of β-cell anti-apoptotic genes (e.g., Bcl-2 proteins) and minimizing the secretion of pro-apoptotic genes (e.g., caspases) hinders DNA and subsequent cell damage. Inhibition of the pro-inflammatory...
| Sl. No. | Name of the plant | Family | Compounds | In vitro/In vivo study with mechanism of action | Clinical Trials | References |
|--------|-------------------|--------|-----------|-----------------------------------------------|----------------|------------|
| 1      | Allium sativum L. | Amaryllidaceae | Allicin, Allin, Diallyl trisulfide, S-allyl cysteine, Allyl mercaptan, Ajoene | Inhibition of DPP-4 enzyme | Nine randomized-controlled trials on with a duration of 1 to 2 weeks | 0.05g to 1.5g dose of garlic | 768 type 2 diabetic patients | Reduction of fasting blood glucose level, with a further reduction in glycosylated hemoglobin and fructosamine in 12 to 24 weeks (204–206) |
| 2      | Aegle marmelos Correa. | Rutaceae | Aegeline | Elevation of blood insulin levels along with liver glycogen | Leaf juice supplementation | 20 g/100 ml dose for 4 weeks | 60 type 2 diabetic patient (25 were male and 35 were female; age range: 25-69 years old) | Reduction in fasting blood glucose, glycosylated hemoglobin (by 20%) and postprandial blood glucose (by 31%) (207) |
| 3      | Artocarpus heterophyllus Lam. | Moraceae | Gallic acid, Catechin, Caffeic acid, Rutin and Quercetin | Inhibits α-glucosidase activity in a dose-dependent manner, increase liver glycogen, increases glucose transporter 2 concentration, reduces blood glucose level | 6 month long randomized double blind placebo controlled cross over trial | Mixed herbal preparation (Artocarpus heterophyllus + Salacia reticulata + Cinnamomum zeylonicum + Pterocarpus marsupium) | 51 type 2 diabetic patients | HbA1C value and the dose of glibenclamide was significantly lowered. (208–210) |
| 4      | Bauhinia forficata Link | Fabaceae | Kaempferitrin | Exhibits hypoglycemic effect in normal and in alloxan-induced diabetic rats, reduces plasma and urinary glucose level in streptozotocin-diabetic rats | 3 month long quasi-experimental pilot study | Daily dose of 10% B. forficata tea in 200 mL water twice daily | 25 type 2 diabetic patients | 0.25% reduction in HbA1C; amelioration in lipid profile (211–213) |
| 5      | Beta vulgaris L. | Chenopodiaceae | Betavulgarosides (II, III, IV), Apigenin 8-C-β-D-glucopyranoside (vitexin), Acacetin 8-C-β-D-glucopyranoside, Acacetin 8-C-α-L-rhamnoside D-pinotol | Reduces blood glucose level, inhibits α-glucosidase activity | 75 day long clinical trial | Infusion of B. forficata leaves | 30 healthy volunteers | Significantly reduced glycemic profile (214) |
| 6      | Bougainvillea spectabilis Wild. | Nyctaginaceae | D-pinitol | Increase in the activity of glucose-6-phosphate dehydrogenase and hepatic, skeletal muscle glycogen content | Randomized-controlled clinical study | 1.2 g dose | 66 type 2 diabetic patients (20 were male and 46 were female; age range: 20-75 years) | Reduction in HbA1C, HOMA-IR and fasting blood glucose levels (215–217) |

(Continued)
| Sl. No. | Name of the plant | Family | Compounds | In vitro/in vivo study with mechanism of action | Clinical Trials | Outcomes | References |
|---------|-------------------|--------|-----------|-----------------------------------------------|-----------------|----------|------------|
|        |                   |        |           |                                               | Description     | Dosage   | Participants |                      |
| 7       | Cecropia obtusifolia Bertol. | Urticaceae | Isoorientin, Chlorogenic acid | Decreases HbA1c, reduces blood glucose level | Randomized parallel single-blind placebo and cross-over-controlled trial | 6.0 g dose | 30 healthy subjects (11 were male and 19 were female; age range: 18–65 years) | Reduction in blood glucose and insulin levels by 45 and 60 minutes, respectively | (222) |
| 8       | Centella asiatica (L.) Urb. | Apiaceae | Asiaticoside (triterpene saponin compound), Madecassic Acid, Asiatic acid, Brahmoside and Brahimnoside (glycosides) | Increase secretion of insulin from pancreatic β-cells, asiatic acid protects pancreatic β-cells from death via activation of Akt kinase and Bcl-xL, promotes proliferation of pancreatic β-cells, reduce blood glucose level and increase serum insulin level | Randomized, double-blind, placebo-controlled, crossover trial | 0.6g dose | 20 healthy subjects (12 were male and 8 were female; age range: 18-25 years) | Reduction in post-prandial blood glucose levels | (223) |
| 9       | Lagerstroemia speciosa (L.) Pers. | Lythraceae | Corosolic acid (2α-hydroxyursolic acid) | Glucose transport-stimulating activity, stimulate glucose uptake in 3T3-L1 cells, reduce blood glucose level | Randomized parallel single-blind placebo and cross-over-controlled trial | 6.0 g dose | 30 healthy subjects (11 were male and 19 were female; age range: 18–65 years) | Reduction in blood glucose and insulin levels by 45 and 60 minutes, respectively | (222) |

(Continued)
| Sl. No. | Name of the plant | Family | Compounds | In vitro/in vivo study with mechanism of action | Clinical Trials | References |
|--------|------------------|--------|-----------|-----------------------------------------------|-----------------|------------|
| 10     | Laminaria japonica Aresch. | Laminariaceae | Butyl-isobutyl-phthalate, polysaccharides | Inhibits the activity of α-glucosidase, reduces fasting blood glucose level, increases plasma insulin level | 1 year long open label trial | (235–238) |
|        |                  |        |           |                                               | 100 mg tablet daily containing water soluble L. speciosa extract | Fasting plasma glucose reduced by 16.6%, ameliorated glycated albumin and glucose tolerance |
|        |                  |        |           |                                               | 15 type 2 diabetic patients | |
| 11     | Mangifera indica L. | Anacardiaceae | Mangiferin, Kaempferol | Improves oral glucose tolerance, reduces fasting plasma glucose level, inhibits the activity of α-amylase and α-glucosidase | 12 week long double-blind randomized placebo-controlled trial | (239–244) |
|        |                  |        |           |                                               | 4 capsules daily containing 350 mg of L. japonica extract | Lessened blood glucose levels, reduced weight, symptomatic relief |
|        |                  |        |           |                                               | 31 subjects | |
|        |                  |        |           |                                               | 37 type 2 diabetic patients | |
| 12     | Salacia chinensis L. | Celastraceae | Salasones A, B, and C, Salaquinone A, Salasol A, 22-dihydroxyolean- | Inhibits α-glucosidase activity, inhibits rat lens aldose reductase | Randomized, double-blind, placebo- | (245–251) |
|        |                  |        |           |                                               | 1000 mg hydroalcoholic extract | Long term glucose hemostasis is achieved in obese subjects |
|        |                  |        |           |                                               | 21 healthy lean and obese subjects | Reduction in post-prandial blood glucose levels |
|        |                  |        |           |                                               | 30 healthy adult volunteers | |

(Continued)
| Sl. No. | Name of the plant | Family | Compounds | In vitro/in vivo study with mechanism of action | Clinical Trials | References |
|--------|-------------------|--------|-----------|-----------------------------------------------|----------------|-----------|
| 12     | 12-en-29-oic acid, Tingenone, Tingenine B, Reged A. Triptocalline A |        |           | controlled, crossover trial                    |                |           |
|        | S. chinensis      |        |           | Randomized double-blind, placebo controlled, crossover trial |                |           |
|        | S. chinensis      |        |           | Randomized double-blind, placebo controlled, crossover trial |                |           |
|        | Various doses of the extract of S. chinensis (200 mg, 300 mg, and 500 mg) |        |           | Double blind randomized controlled trial       |                |           |
|        | Either 300 mg or 500 mg dose of S. chinensis extract |        |           | Reduced glycemic indices supporting it’s αGI activity, gastrointestinal peptides were affected which might lead to appetite modification |                |           |
|        | 35 healthy subjects |        |           | Ameliorated postprandial glucose level and insulin response |                |           |
|        | 48 healthy overweight or obese participants |        |           | Glycemic indices supporting it’s αGI activity, gastrointestinal peptides were affected which might lead to appetite modification |                |           |
| 13     | Salacia reticulata Wight | Celastraceae | Salacinol, Kotalanol | Inhibits α-glucosidase enzyme |                | (210, 252–257) |
|        | 6 month long randomized double blind placebo controlled cross over trial |        |           | Mixed herbal preparation (Salacia reticulata + Artocarpus heterophyllus + Cinnamomum zeylionicum) |                |           |
|        | 51 type 2 diabetic patients |        |           | HbA1C value and the dose of glibenclamide was significantly lowered. |                |           |
| Sl. No. | Name of the plant | Family | Compounds | In vitro/in vivo study with mechanism of action | Clinical Trials | References |
|--------|-------------------|--------|------------|-----------------------------------------------|----------------|------------|
| 6      | Pterocarpus marsupium | Fabaceae | Pterocarpus | 6 week long placebo-controlled, cross-over trial | 20 type 2 diabetic subjects | Lessened fasting blood glucose, glycated hemoglobin levels and BMI value | (258, 259) |
| 14     | Scoparia dulcis L. | Scrophulariaceae | Scutellarein, Apigenin, Luteolin, Scopadulcic acid B, Betulinic acid, Scoparic acid A | Inhibition of α-glucosidase activity, stimulates the beta cell to secrete insulin, ameliorated glucose uptake activity, exhibits PPAR-γ agonistic activity and increase insulin sensitivity | 35 subjects with type 2 diabetes | Reduced fasting blood glucose and HbA1C level | (260–264) |
| 15     | Stevia rebaudiana Bertoni | Asteraceae | Stevioside | Antihyperglycaemic, insulinotropic and glucagonostatic actions | 20 type 2 diabetic patients | Reduced fasting and post-prandial blood glucose levels, ameliorated lipid profile | (260–264) |
transcription factor NF-κB reduces the inflammation stimulated production of inducible nitric oxide synthase (iNOS) and NO, hence reducing cell damage through increasing Ca²⁺ level in ER and mitigating ER stress, inactivating the JNK pathway and thus actuating the PI3K/Akt signaling which supports cell proliferation, survival and growth. Suppression of the deterioration of β-cell through these mechanisms halts reduced insulin secretion, thereby avoiding the state of hyperglycemia (273, 274).

g. Reduction of HbA1c and Glycated Plasma Protein Concentration

Since diabetes mellitus is a condition where blood carbohydrate concentration increases, the monosaccharides nonenzymatically react with the proteins in blood (mainly hemoglobin A and albumin) and adheres to form a modified protein complex (Schiff base) through a process called glycation. The produced glycated hemoglobin (HbA1c) and glycated plasma proteins are concerned with much significance in the research. HbA1c value is often examined as it is one of the major markers for the diagnosis of diabetes mellitus. These glycated products can further encounter intramolecular rearrangements followed by other irreversible reactions (condensation, cross-linking, glycoxidation, cyclization, dehydration, etc.) to become advanced glycation end products (AGEs) which can accumulate and cause deleterious effects on metabolic and vascular health, leading to added diabetic complications. Glycation inhibitors hinder this process through various mechanisms, namely competitively binding with the amino group of the protein, binding at the site of glycation, cutting the open chain structure of monosaccharides and, adhering to the intermediaries of the glycation reaction. Hence, the concentration of HbA1c and glycated plasma proteins is lessened and the aftermath of glycation and diabetic complications can be avoided (275–277).

h. Enhancement of GLP-1

Glucagon-like peptide-1 (GLP-1) is a hormone secreted by the L cells in the distal ileum and colon of the gastrointestinal system. Secreted upon nutrient intake, GLP-1 subsequently binds to GLP-1 receptor (a G-protein-coupled receptor) on pancreatic β-cell to exert its effects, namely, raising the glucose-dependent secretion of insulin and lessening the secretion of glucagon, decelerating gastric emptying, subduing of appetite with imparting a feeling of fullness. Circulatory GLP-1 faces immediate degradation by the enzyme dipeptidyl peptidase 4 (DPP-4) and hence has a short half life of about 2 min. As such, alternative compounds with the functionality of serving as agonists to GLP-1 receptor (GLP-1R) and being resistant to degradation by DPP-4 enzyme have been recognized feasible to employ the proper effect of GLP-1. The GLP-1 receptor (GLP-1R) agonists enhance insulin biosynthesis by increasing the transcription of the insulin genes through activating cAMP/ PKA-dependent and -independent signaling mechanisms, increasing Ca²⁺ levels intracellularly, and activating the insulin gene promoting transcription factor pancreas duodenum homeobox 1 (Pdx-1). The agonist binding also results in inhibition of ATP-sensitive K⁺ channels, leading to depolarization of the membrane and simultaneous influx of extracellular Ca²⁺. ATP synthesis is also enhanced in mitochondria. The combined effect of surged ATP and intracellular Ca²⁺ level is the exocytosis of insulin storage granule. As a result, insulin secretion capability and reserve of β-cell is maintained. Glucagon secretion is inhibited by GLP-1R agonists either directly by acting on α-cells of the pancreas, or less likely indirectly along with the stimulated secretion of insulin. This inhibition ameliorates glycemic control as reduced glucagon level diminishes glucose production from the liver, which in turn reduces the required insulin in the bloodstream. Enhancement of GLP-1 also facilitates the indirect suppression of gastric emptying through the vagus nerve and its involvement with the central nervous system (CNS) located GLP-1Rs, thus relaying a sensory message to the brainstem (278, 279).

i. Regulation of GLUT-4

Belonging to the group of sugar transporter proteins (GLUT1-GLUT12, and HMIT), glucose transporter type 4 (GLUT-4) is a 12-transmembrane domain containing transporter which allows insulin induced blood glucose influx into skeletal muscle and fat cells through facilitated diffusion process and hence maintains the homeostasis of glucose metabolism in the body. The transporter typically resides intracellularly but relocates to the cell membrane upon stimulation of insulin or during exercise through independent mechanisms. The receptor binding of insulin in target cells activates insulin receptor (IR) tyrosine kinase, thus beginning phosphorylation of tyrosine moiety of insulin receptor substrate proteins (IRS) followed by phosphoinositide 3-kinase (PI3K) recruitment. Afterward, PI3K catalyzed phosphorylation of phosphatidylinositol-4, 5-bisphosphate (PIP2) produces phosphatidylinositol-3, 4, 5-trisphosphate (PIP3), which in turn triggers the phosphorylation mediated activation of other protein kinases (Akt, αPKCa/δ) that eventually mobilize the effectors, namely Rab proteins. Rab proteins (Rab8 and Rab14) lead to GLUT-4 translocation into the cell membrane from intracellular GLUT4 storage vesicle (GSV) which increases glucose internalization up to 10-20 times (280, 281).

CHEMICAL CLASS WISE FEW MOST PROMINENT ANTIDIABETIC PHYTOCHEMICALS ALONG WITH THE REPORTED MECHANISM OF ACTIONS

Based on previous research studies, few phytochemicals have been already recognized as the most prominent antidiabetic lead compounds. Those are currently under exclusive assessment so that novel antidiabetic drugs can be introduced in the coming days. In Tables 2–5 the few most prominent phytochemicals with the reported mechanism of actions are represented briefly.
Alongside these phytochemicals, concerned researchers should also evaluate other aforementioned phytochemicals in this review work to establish absolute safety and toxicity profile as well as the mechanism of antidiabetic actions.

**FUTURE RESEARCH DIRECTIONS**

Diabetes is conventionally treated and managed by taking synthetic antidiabetic medications commercially available in the market. The major classes among these medicines are sulphonylureas (glibenclamide), biguanides (metformin), thiazolidinediones (pioglitazone), DPP-4 inhibitors (sitagliptin), alpha-glucosidase inhibitors (acarbose), glinides (repaglinide) and GLP-1 agonists (exenatide) (7, 284). Despite the mass prevalence and usage, the synthetic drugs accompany various side effects which include hypoglycemia (for sulphonylureas, glinides), weight gain (for sulphonylureas, thiazolidinediones), cardiovascular risk (for sulphonylureas, thiazolidinediones), pancreatitis (for DPP-4 inhibitors, GLP-1 agonists), hepatitis (for thiazolidinediones, DPP-4 inhibitors), cancer risk (for DPP-4 inhibitors, GLP-1 agonists), gastrointestinal effects (for biguanides, GLP-1 agonists), lactic acidosis (for biguanides) (285). These adversities and constraints associated with the prevailing synthetic medications entail the researchers to search for antidiabetic drugs from plant sources with a better safety and efficacy profile. Along the lines of many other diseases, diabetes mellitus has been treated with plant based medications for a long while owing to factors like marked efficacy, less toxicity and side effects, low cost, and availability (286, 287). An exclusive development of plant based drugs has seemingly occurred through evolutionary mechanisms, imparting the capability to interact with biomolecules (288, 289). Isolated phytochemicals are either used as drugs or availed as chemical leads or their analogs for synthesizing biologically active compounds. The prevalence of phytochemicals in the pharmaceutical scenario can be perceived by surveying all the authorized drugs registered globally within the time frame of 25 years before 2007, where roughly 50% of the drugs were majorly plant based natural products or their synthetic derivatives (285, 290). As an example, metformin, the extensively used drug in treating type 2 diabetes, is a drug derived from the plant *Galegine officinalis* (291). Antidiabetic action being one of the most popular fields of use of phytochemicals houses compounds from various chemical classes, namely flavonoids (quercetin), alkaloids (berberine), terpenes (thymoquinone), phenylpropanoids (chlorogenic acid) and others. The phytochemicals reported in this study revealed prominent antidiabetic action through various mechanisms like inhibiting α-glucosidase, α-amylase and DPP-4 enzyme, increasing insulin sensitivity and secretion, increasing glucose uptake by muscle cells and adipose tissue, nourishing pancreatic β-cells, etc. These various ways of phytocompounds to exert antidiabetic action illustrates the effective diversity they can offer. Thus, potential phytocompound(s), isolated from medicinal plants or dietary materials, with proven preclinical and clinical antidiabetic efficacy can be the prospective and potential candidates for the development of novel antidiabetic drugs. For example, Charantin and Polypeptide-p isolated from *Momordica charantia* L. have been reported to exhibit potential antidiabetic activities in preclinical and clinical studies which have been included in this manuscript. These two compounds can be prospective candidates for the development of novel antidiabetic medicaments after confirming their toxicity and further clinical trials. Traditional medicinal approaches like Ayurveda, Unani and so on also utilized plant based remedies to treat diabetic illness. However, further research is necessary to disclose the absolute and exact mechanism of action of these compounds which would facilitate their outset as drugs or chemical leads. Indeed, serving as drugs or drug templates is not the only purpose of plant derived compounds, rather, they also guide in the recognition and revelation of complex and novel molecular pathways and targets involving the health condition (292). Hence, further research on these phytochemicals could enable the discovery of several targets for therapeutic intervention against diabetes mellitus. In addition, elucidation of the feasibility and toxicity profile despite the mentioned predating advantage as plant based products is also a salient research concern.

**DISCUSSION**

The two most pronounced variations of diabetes are Type 1 diabetes mellitus and Type 2 diabetes mellitus both of which result in a hyperglycemiac state. Type 1 diabetes mellitus is an autoimmune illness typified by the demolition of pancreatic β-cells followed by dreadful insulin scarcity. On the other hand, Type 2 diabetes mellitus is more familiar and a major portion of diabetic patients (90 to 95%) are suffering from this dysfunction, which is categorized by peripheral insulin resistance and anomalies in the secretion of insulin (284). However, it is a non-communicable illness, experts are warning us about a figure of almost 438 million diabetic patients in 2030 (7) which considers the dreadfulness of this disease. In a broader sense, the causative factors of diabetes can reside in insulin resistance, abnormal insulin secretion and hepatic glucose synthesis along with impaired fat metabolism. Insulin resistance refers to the state where the efficacy of the insulin on target tissues becomes compromised particularly on adipocytes, hepatocytes and skeletal muscles (293) which leads to hyperglycemia by impairing utilization of glucose and increasing hepatic glucose output (294). Despite having a good number of commercially available antidiabetic drugs, the side effects delimit their unquestionable implications. In contrast, nutraceuticals and phytomedicines offer a low incidence of adverse effects that can be a fantastic alternative to regular drugs in combating diabetes and its related complications (7). Plant-derived medicaments have also been mentioned in various ethnic and traditional practices including the Indian, Koran, Chinese and Mexican cultures as well as in Western and Ayurvedic herbalism approaches which accredits their tremendous antidiabetic
TABLE 2 | Antidiabetic potential of alkaloids extracted from medicinal plants and their mechanism of actions.

| Sl. No | Compounds | Plant source | Study model | Mechanism of action | Reference |
|--------|------------|--------------|-------------|---------------------|-----------|
| 1      | Aegeline   | Aegle marmelos Correa aristata DC. | In vivo (Diabetic rat) | Lowering of blood glucose level due to similarity in structure and action with b3-AR agonists | (207) |
| 2      | Berberine  | Berberis | In vivo (Diabetic rat) | Improving the action of insulin by triggering AMPK, reducing insulin resistance through protein kinase C-dependent up-regulation of insulin receptor expression; causing glycolysis; enhancing GLP-1 secretion and regulating its release, inhibiting DPP-IV | (30) |
| 3      | Vindoline, Vindolol, Vindoline, Vindoline | Catharanthus roseus (L.) G.Don | In vivo | Vindoline, vindolol, vindoline and vindoline induced increased glucose uptake in myoblast C2C12 or pancreatic β-T1C6 cells, Vindoline, vindolol and vindoline also improved protein tyrosine phosphatase-1B (PTP-1B) inhibitory functions | (45) |
| 4      | Cryptopine | Cryptopis sanguinolenta (Lindl.) Schtr. Lobelia chinensis Lour. | In vivo (Diabetic mouse) | Enhanced glucose transport | (78) |
| 5      | Radicamines A, Radicamines B | Lupine, 13α-OH lupine, 17α-oxo-lupine | In vivo (Diabetic rat) | Enhanced the secretion of insulin in a glucose-dependent manner by reducing K+ permeability in the β-cell plasma membrane | (106) |
| 6      | Moringine | Moringa oleifera Lam. | In vivo (Diabetic rat) | Aiding the restoration of the normal histological structure of the pancreas | (122) |
| 7      | 1-deoxyojirimycin | 1-deoxyojirimycin | In vivo (Diabetic mouse) | Reduction in the activity of α-glucosidase by competitive inhibition | (130) |
| 8      | Nuciferine | Nelumbo nucifera Gaertn. | In vivo (Diabetic mouse) | Increase in insulin secretion in both isolated islets and INS-1E cells, stimulation of both the first phase and the second phase of insulin secretion by closing K-ATP channels and also through stimulation of K-ATP channel independent amplification pathways. | (137) |
| 9      | Gentianine | Swertia chirayita Buch Ham. | In vitro | Promising amelioration in adipogenesis associated expression of PPAR-γ, GLUT-4 and adiponectin | (164) |
| 10     | Magnoflorine | Tinospora cordifolia (Willd.) Miers | In vitro | Potent inhibition of α-glucosidase | (172) |

(Continued)

TABLE 3 | Antidiabetic potential of phenolics extracted from medicinal plants and their mechanism of action.

| Sl. No | Compounds | Subclass | Plant source | Study model | Mechanism of action | Reference |
|--------|------------|----------|--------------|-------------|---------------------|-----------|
| 1      | Piceatannol | Stilbenes | Callistemon rigidus R.Br. | In vivo (Diabetic mouse) | Suppression in the activity of α-amylase. | (44) |
| 2      | Scirpusin B | Stilbenes | Callistemon rigidus R.Br. | In vivo (Diabetic mouse) | Regulation of α-amylase in mouse GIT. Suppression in the activity of α-amylase. | (44) |
| 3      | Chamaemeloside | Flavonoids | Chamaemelum nobilis (L.) All. | In vivo (Diabetic mouse) | Potential suppression in the production of hepatic glucose, as such, reduced gluconeogenesis. Potential effects on intestinal absorption, hepatic or peripheral disposal of glucose as well Renovation of beta cells | (49) |
| 4      | Pyrogallol | Phenols | Cinnamomum verum J. Presl | In vivo (Diabetic mouse) | Inhibition of α-glucosidase activity | (56) |
| 5      | Bisdemethoxycurcumin, Curcumin, Demethoxycurcumin | Phenols | Curcuma longa L. | In vitro | (Continued) |
| 6      | Acacetin | Flavonoids | Lobelia chinensis Lour. | Network pharmacological model | Promotion of secretion of insulin, improvement of insulin resistance, and stimulation of the utilization of glucose by acting on GSK3B, MAPK, INR, and dipeptidyl peptidase-4 (DPP4) | (104) |
| 7      | Coumarins | Coumarins | Aegle marmelos Correa aristata DC. | In vivo (Diabetic rat) | Stimulation of insulin secretion from beta cells of the isles of Langerhans | (282) |
| 8      | Quercetin | Flavonoids | Maticaria chamomilla L. Moringa oleifera Lam. | In vivo (Diabetic rat) | Quercetin moderately inhibited the enzymatic activity of sucrase | (111) |
|        |            |          |              |              | Halted sorbitol from accumulating in erythrocytes Aiding the restoration of the normal histological structure of the pancreas | (122) |

(Continued)
| Sl. No | Compounds        | Subclass       | Plant source                        | Study model   | Mechanism of action                                                                 | Reference |
|-------|------------------|----------------|-------------------------------------|---------------|-------------------------------------------------------------------------------------|-----------|
| 9     | Luteolin         | Flavonoids     | Matricaria chamomilla L.            | In vivo (Diabetic rat) | Blocking the transport of fructose and glucose by GLUT2 in the brain and promoting the translocation and expression of GLUT4 in skeletal muscle | (125)     |
| 10    | Esculetin, Umbelliferone | Coumarins     | Matricaria chamomilla L.            | In vivo (Diabetic rat) | Improvement of the activity of adiponectin in white adipose tissue and blood concentration, in spite of an inhibition of poly (ADP-ribose) polymerase γ expression followed by improved insulin sensitivity. Inhibition of glucose uptake at glucose transporters (GLUTs) level | (182, 183) |
| 11    | Isoquercitrin, Astragalin | Flavonoids    | Morus alba L.                       | In vivo (Diabetic mouse) | Esculetin showed moderate inhibition in the enzymatic activity of sucrase. Esculetin and umbelliferone halted sorbitol from accumulating in erythrocytes | (111)     |
| 12    | Valoneic acid dilactone | Tannins       | Punica granatum L.                  | In vivo (Diabetic rat) | Inhibition of the activity of aldose reductase and protein tyrosine phosphatase 1B (PTP1B). Improvement in insulin secretion from pancreatic β cells or its release from the bound form along with insulin mimetic actions or amended glucose utilization technique | (150)     |
| 13    | Karanjin         | Flavonoids     | Pongamia pinnata (L.) Pierre         | In vitro      | Inhibition of PTPase-1B                                                                  | (156)     |
| 14    | Pongamol         | Phenols        | Pongamia pinnata (L.) Pierre         | In vitro      | Inhibition of PTPase-1B                                                                  | (156)     |
| 15    | Silychristin A   | Flavonolignans | Silybum marianum (L.) Gaertn         | In vivo (Diabetic rat) | Improvement of the function of β-cells along with glucose lowering effect by protecting the β-cells from oxidative stress-induced damage and blocking the activity of α-glucosidase enzyme | (159)     |
| 16    | Mangiferin       | Xanthonoid     | Swertia chirayita Buch Ham.          | In vivo (Diabetic rat) | inhibition of the activity of aldose reductase and protein tyrosine phosphatase 1B (PTP1B). Improvement in insulin secretion from pancreatic β cells or its release from the bound form along with insulin mimetic actions or amended glucose utilization technique | (165)     |
| 17    | Pterostilbene    | Stilbenoids    | Vitis vinifera L.                   | Wild-type and mutant Kir6.2 models | Promising inhibitory efficacy on both normal and mutant models of kir6.2 channel which is encoded by the KCNJ11 gene, whose mutation causes congenital hyperinsulinism | (181)     |
| 18    | Myricetin        | Flavonoids     | Vitis vinifera L.                   | In vivo (Diabetic rat) | Promotion of glucose uptake in liver and soleus muscles as well as hepatic glycogen synthase, halting advanced glycation end products in diabetic condition | (184)     |
| 19    | Resveratrol      | Stilbenoids    | Vitis vinifera L.                   | In vivo (Diabetic rat) | Improvement of insulin resistance                                                      | (185)     |
| 20    | 6-shogaol        | Phenols        | Zingiber officinalis Roscoe         | In vitro | Human pancreatic α-amylase inhibition                                                  | (186)     |
| 21    | 6-gingerol       | Phenols        | Zingiber officinalis Roscoe         | In vivo (Diabetic mouse) | Stimulation of the transportation activity of intracellular glucose and promotion of glucose uptake | (190)     |
| 22    | 6-parodol        | Phenols        | Zingiber officinalis Roscoe         | In vitro | Improvement in the expression of insulin-dependent glucose transporter (GLUT4)      | (191, 192) |

References:

1. Alam, Roscoe, of... Endocrinology | www.frontiersin.org February 2022 | Volume 13 | Article 800714
TABLE 4 | Antidiabetic potential of terpenes extracted from medicinal plants and their mechanism of actions.

| Sl No | Compounds | Subclass | Plant source | Study model | Mechanism of action | Reference |
|-------|------------|----------|--------------|-------------|---------------------|-----------|
| 1     | Bacosine   | Triterpenoids | Bacopa monnieri (L) Wettst. | In vivo (Diabetic rat) | Increase in the consumption of peripheral glucose and protection against oxidative damage. Increase in the level of liver glycogen as well improved glucose uptake in 3T3-L1 adipocytes. | (29) |
| 2     | Bassic acid | Triterpenoid | Bumelia santorum Mart. | In vivo (Diabetic rat) | Increase in glucose uptake and glycogen synthesis. Increase in insulin secretion from the pancreatic beta-cells. | (42) |
| 3     | β-amyrin   | Triterpenoids | Costus pictus D. Don | In vitro | Improved glucose uptake in 3T3-L1 adipocytes. | (67) |
| 4     | Turmerone   | Sesquiterpenoids | Curcuma longa L. | In vitro | Inhibition of α-glucosidase and α-amylase activity. | (73) |
| 5     | α-amyrin-3O-β-(5-hydroxy) ferulic acid | Triterpenes | Euclea undulate Thunb. var. myrtilloides | In vitro | Inhibition of α-glucosidase activity. | (81) |
| 6     | Gymnemagenin | Triterpenoids | Gymnema sylvestre R. Br. | Crystallographic investigation | Exhibition of good gelling property with various target protein’s crystallographic constitution which contribute to its carbohydrate management property. | (83) |
| 7     | Thymoquinone, Dithymoquinone | Monoterpene, Diterpene | Nigella sativa L. | | Potential stimulation in pancreatic β-cells causing insulin secretion, reduced hepatic gluconeogenesis, and induced insulin sensitivity in peripheral tissue. | (139) |

TABLE 5 | Antidiabetic potential of other notable phyto compounds extracted from medicinal plants and their mechanism of actions.

| Chemical Class | Compounds | Plant source | Study model | Mechanism of action | Reference |
|---------------|-----------|--------------|-------------|---------------------|-----------|
| Phenylpropanoids | Chlorogenic acid | Cichorium intybus L. | In vivo (Diabetic rat) | Increased glucose uptake in L6 muscular cells, elevated insulin secretion from the INS-1E insulin-secreting cell line and rat islets of Langerhans. Aiding the restoration of the normal histological structure of the pancreas. | (53) |
| Chovic acid | | Morinda oleifera Lam. | In vivo (Diabetic rat) | Increased glucose uptake in L6 muscular cells, elevated insulin secretion from the INS-1E insulin-secreting cell line and rat islets of Langerhans along with insulin secreting and sensitizing action. | (53) |
| Eugenol | | Cinnamonum verum J. Presl | In vivo (Diabetic mouse) | Renovation of beta cells. | (56) |
| Cinnamaldehyde | | Cinnamonum verum J. Presl | In vivo (Diabetic mouse) | Ameliorating the uptake of glucose by upraising the amount of AKT2 and aortic nitric oxide synthase 3 (eNOS), insulin receptor substrate1 (IRS1) and p-85 regulatory subunit of PI3K-P85 while concurrently abating the expression of NADPH oxidase 4 (NOX4). | (56) |
| Saponins | 3-hydroxycucurbita-5, 24-dien-19-al-7, 23-di-O-glucopyranoside, Momordica charantia L. | Momordica charantia L. | In vivo (Diabetic rat) | Promising insulin releasing property. | (115) |
| Lipid | Methyl tetracosanate | Costus pictus D. Don | In vivo (Diabetic rat) | Improved glucose uptake in 3T3-L1 adipocytes. | (68) |
| Fatty acid | Linoleic acid, Oleic acid | Nigella sativa L. | Human | Potential stimulation in pancreatic β-cells causing insulin secretion, reduced hepatic gluconeogenesis, and induced insulin sensitivity in peripheral tissue. | (139) |
| Protein | Turmerin | Curcuma longa L. | Human | Inhibition of α-glucosidase and α-amylase activity. | (73) |
| Polypeptide-p | Morinda oleifera Lam. | Momordica charantia L. | Human | Insulin mimicking activity. | (115) |
| Carbohydrate | α-arabinose, α-xylene, α-glucose, α-rhamnose, α-mannose | Morus alba L. | In vivo (Diabetic rat) | Repair of pancreatic β-cells. | (130) |
| Peitralosa | Psacaliaum peltatum (Kunth) Cass. | Human | | Potentially enhance secretion of insulin from the islets of Langerhans or increase utilization of glucose by peripheral tissues. | (157). |
| Kinsenoside | Anoectochilus roxburghii (Wall) Lindl. | In vivo (Diabetic rat) | | Restoration of damaged pancreatic β cells, functionality against oxidative stress and NO factor, regulation of antioxidant enzymes and scavenging of free radicals. | (28) |
| D-o-pitol | Bougainvillia spectabilis Wild. | In vivo (Diabetic mouse) | | Exhibition of an insulin-like impact by acting through a post-receptor insulin action pathway, affecting the uptake of glucose. | (283) |

(Continued)
potential (7, 295). Hence, the reported aforementioned phytochemicals in this extensive review can be considered as very promising wellsprings to develop novel antidiabetic therapeutics to heal diabetes and related complications.

**CONCLUSION**

A collection of anti-diabetic plants used in the treatment of diabetes mellitus has been reviewed in this article. Several shards of scientific evidence have proved that those phytochemicals possess antihyperglycemic potentials and can be effectively implicated in the management of diabetic and metabolic complications avoiding notable side effects exerted by conventional drugs. Although dietary and non-dietary plants are always considered as promising avenues of remedies to treat different types of disease states, together with diabetes and others, many plants and plant-derived bioactive phytoconstituents have not yet been researched well. In order to explore and validate proper mechanistic pathways of pharmacological activities demonstrated by the reported antidiabetic phytochemicals, further investigations are warranted. In spite of considering plants and/or dietary plant materials as safe for intake, yet the prospective antidiabetic phytochemicals should also be evaluated for toxicity studies for the establishment of therapeutically effective and safe phytomedicines.

**AUTHOR CONTRIBUTIONS**

MMRS and SA conceptualized the study. SA, MNRC and TNS searched in the databases and collected articles. SA, TNS and MNRC wrote the manuscript. MAR, NIC, CZ, JX, EH, SAK, and INM critically revised the manuscript. SA, TNS and NIC edited the final manuscript as per review comments. MMRS and INM critically evaluated, revised the manuscript and supervised the project. All authors contributed to the article and approved the submitted version.

**REFERENCES**

1. Kharroubi AT, Darwish HM. Diabetes Mellitus: The Epidemic of the Century. *World J Diabetes* (2015) 6:850–67. doi: 10.4239/wjd.v6.i6.850
2. Hunter SJ, Garvey WT. Insulin Action and Insulin Resistance: Diseases Involving Defects in Insulin Receptors, Signal Transduction, and the Glucose Transport Effector System 1. *Am J Med* (1998) 105(4):331–45. doi: 10.1016/S0002-9343(98)00300-3
3. Wild S, Roglic G, Green A, Sicree R, King H. Global Prevalence of Diabetes: Estimates for the Year 2000 and Projections for 2030. *Diabetes Care* (2004) 27(5):1047–53. doi: 10.2337/diacare.27.5.1047
4. American Diabetes Association. Diagnosis and Classification of Diabetes Mellitus. *Diabetes Care* (2014) 37 Suppl 1:S81–90. doi: 10.2337/dc14-S081
5. Patel DK, Prasad SK, Kumar R, Hemalatha S. An Overview on Antidiabetic Medicinal Plants Having Insulin Mimetic Property. *Asian Pac J Trop Biomed* (2012) 2:320–30. doi: 10.1016/S2221-1691(12)60032-X
6. Cakan N, Kizilbash S, Kamat D. Changing Spectrum of Diabetes Mellitus in Children: Challenges With Initial Classification. *Clin Pediatr (Philai)* (2012) 51:939–44. doi: 10.1177/0009922812441666
7. Li QQ, Kam A, Wong KH, Zhou X, Omar EA, Alqahtani A, et al. Herbal Medicines for the Management of Diabetes. In: Ahmad SI, editor. *Diabetes: An Old Disease, a New Insight, Advances in Experimental Medicine and Biology*. New York, NY: Springer (2013). p. 396–413. doi: 10.1007/978-1-4614-5441-0_28
8. Kazeem MI, Davies TC. Anti-Diabetic Functional Foods as Sources of Insulin Secretagog, Insulin Sensitizing and Insulin Mimetic Agents. *J Funct Foods* (2016) 20:122–38. doi: 10.1016/j.jff.2015.10.013
9. Baynes HW. Classification, Pathophysiology, Diagnosis and Management of Diabetes Mellitus. *J Diabetes Metab* (2015) 6(5):1–9. doi: 10.4172/2155-6156.1000583
10. Paschou SA, Papadopoulou-Marketou N, Chrousos GP, Kanaka-Gantenbein C. On Type 1 Diabetes Mellitus Pathogenesis. *Endocrine Connect* (2018) 7:R38–46. doi: 10.1530/EC-17-0347
11. Holt RI. Diagnosis, Epidemiology and Pathogenesis of Diabetes Mellitus: An Update for Psychiatrists. *Br J Psychiatry* (2004) 184(S47):s55–63. doi: 10.1192/bjp.184.47.s55
12. Yang S, Zhou R, Zhang C, He S, Su Z. Mitochondria-Associated Endoplasmic Reticulum Membranes in the Pathogenesis of Type 2 Diabetes Mellitus. *Front Cell Dev Biol* (2020) 8:571554. doi: 10.3389/fcell.2020.571554
13. Miao C, Zhang G, Xie Z, Chang J. MicroRNAs in the Pathogenesis of Type 2 Diabetes: New Research Progress and Future Direction. *Can J Physiol Pharmacol* (2018) 96(2):103–12. doi: 10.1139/cjpp-2017-0452
14. Rehman K, Akash MSH. Mechanism of Generation of Oxidative Stress and Pathophysiology of Type 2 Diabetes Mellitus: How Are They Interlinked? *J Cell Biochem* (2017) 118(11):3577–85. doi: 10.1002/jcb.26097
15. Asmat U, Abad K, Ismail K. Diabetes Mellitus and Oxidative Stress—A Concise Review. *Saudi Pharm J* (2016) 24(5):547–53. doi: 10.1016/j.jsps.2015.03.013
16. Tripkov A, Resanovic I, Stanimirovic J, Radak D, Mousa SA, Cenic-Milosevic D, et al. Oxidized Low-Density Lipoprotein as a Biomarker of Cardiovascular Diseases. *Crit Rev Clin Lab Sci* (2015) 52(2):70–85. doi: 10.3109/10408363.2014.992063
17. Esser N, Legrand-Poels S, Piette J, Scheen AJ, Paquot N. Inflammation as a Link Between Obesity, Metabolic Syndrome and Type 2 Diabetes. *Diabetes Res Clin Pract* (2014) 105(2):141–50. doi: 10.1016/j.diabres.2014.04.006
18. Venkatasamy VV, Pericherla S, Manthuruthil S, Mishra S, Hanno R. Effect of Physical Activity on Insulin Resistance, Inflammation and Oxidative Stress in Diabetes Mellitus. *J Clin Diag Res: JCDR* (2013) 7(8):1764. doi: 10.7860/jcdr/2013/6518.3306
19. Sircana A, Framarin L, Leone N, Ferrutti M, Castellino F, Parente R, et al. Altered Gut Microbiota in Type 2 Diabetes: Just a Coincidence? *Curr Diabetes Rep* (2018) 18(10):1–11. doi: 10.1007/s11892-018-1057-6

20. Li X, Watanabe K, Kimura I. Gut Microbiota Dysbiosis Drives and Implies Novel Therapeutic Strategies for Diabetes Mellitus and Related Metabolic Diseases. *Front Immunol* (2017) 8:1882. doi: 10.3389/fimmu.2017.01882

21. Cani PD, Amar J, Iglesias MA, Moggi P, Knauf C, Bastelica D, et al. Metabolic Endotoxemia Initiates Obesity and Insulin Resistance. *Diabetes* (2007) S67(1):1761–72. doi: 10.2337/db06-1491

22. Tan J, McKenzie C, Potamitis M, Thorburn AN, Mackay CR, Macia L. The Role of Short-Chain Fatty Acids in Health and Disease. *Adv Immunol* (2014) 121:91–119. doi: 10.1016/B978-0-12-800100-4.00003-9

23. Tang C, Ahmed K, Gille A, Lu S, Gröne HJ, Tunaru S, et al. Loss of FFA2 and FFA3 Increases Insulin Secretion and Improves Glucose Tolerance in Type 2 Diabetes. *Nat Med* (2015) 21(2):173–7. doi: 10.1038/nn.3779

24. Neis EP, Dejong CH, Rensen SS. The Role of Microbial Amino Acid Metabolism in Host Metabolism. *Nutrients* (2015) 7(4):2930–46. doi: 10.3390/nu7042930

25. Shan Z, Sun T, Huang H, Chen S, Chen L, Luo C, et al. Association Between Microbiota-Dependent Metabolite Trimethylamine-N-Oxide and Type 2 Diabetes. *Am J Clin Nutr* (2017) 106(3):888–94. doi: 10.3945/ajcn.117.157107

26. Galicia-Garcia U, Benito-Vicente A, Jefari S, Larrea-Sebal A, Siddiqi H, Uribe KR, et al. Pathophysiology of Type 2 Diabetes Mellitus. *Int J Mol Sci* (2020) 21(17):6275. doi: 10.3390/ijms21176275

27. Ye S, Shao Q, Zhang A. *Anoectochilus Roxburghii*: A Review of Its Traditional Uses, Chemical Constituents, and Biological Health Promoting Properties. *Front Immunol* (2020) 11:693. doi: 10.3389/fimmu.2020.00651

28. Potdar D, Hirwani RR, Dhulap S. Phyto-Chemical and Pharmacological Review of Cinnamomum Verum J. L. H. bark of Callistemon Rigidus Showing Inhibitory Effects on Mouse Alpha-Amlyase Activity. *Bioll Pharm Bull* (2006) 29:1275–7. doi: 10.1248/bpb.29.1275

29. Chandra K, Jain V, Jabin A, Dwivedi S, Joshi S, Ahmad S, et al. Effect of an Antidiabetic Extract of Bumelia Sartorum. *Phytotherapy* (2013) 2014:e857292. doi: 10.13040/IJPSR.0975-8232.1(1).29-46

30. Russell KRM, Omoruyi FO, Pascoe KO, Morrison EYSA. Hypoglycaemic Activity of Bixa Orellana Extract in the Dog. *Methods Find Exp Clin Pharmacol* (2008) 30:301–5. doi: 10.1055/s-2008.1148067

31. Mishra D. An Analytical Review of Plants for Anti-Diabetic Activity With Their Phytoconstituent & Mechanism of Action. *Int J Pharm Sci Res* (2009) 1:29–46. doi: 10.3344/ijbpr.0975-8232.1(1).29-46

32. Yin J, Xing H, Ye J. *Endiandra ritro* Bark of Callistemon Rigidus: A Comparative Evaluation of Some Blood Sugar Lowering Agents of Plant Origin. *J Ethnopharmacol* (1999) 67:367–72. doi: 10.1016/S0378-7349(99)00095-1

33. Singh SN, Vats P, Suri S, Shyam R, Kumria MML, Ranganathan S, et al. Effect of an Antidiabetic Extract of Catharanthus Roseus in Enzymic Activities in Streptozotocin Induced Diabetic Rats. *J Ethnopharmacol* (2001) 76:247–79. doi: 10.1016/S0378-7349(01)00254-9

34. Eddouks M, Lemhadri A, Zeggwagh NA, Michel J-B. Potent Hypoglycaemic Activity of the Aqueous Extract of Chamaemelus nobile in Normal and Streptozotocin-Induced Diabetic Rats. *Diabetes Res Clin Pract* (2005) 67:189–95. doi: 10.1016/j.diabres.2004.07.015

35. König GM, Wright AD, Keller WJ, Judd RL, Bates S, Day C. Hypoglycaemic Activity of an HMG-Containing Flavonoid Glucoside, Chamaemeloside, From Chamaemelus nobile. *Planta Med* (1998) 64:612–4. doi: 10.1055/s-2006-957532

36. Yonei Y, Miyazaki R, Takehashi Y, Takehashi H, Nomoto K, Yagi M, et al. Anti-Glycation Effect of Mixed Herbal Extract in Individuals With Pre-Diabetes Mellitus. *Anti-Aging Med* (2010) 7:26–35. doi: 10.3793/jaam.7.26.7.26

37. Street RA, Sidana J, Prinsloo G. Cichorium Intybus: Traditional Uses, Phytochemistry, Pharmacology, and Toxicology. *Evidence-Based Complement Altern Med* (2013) 2013:579319. doi: 10.1155/2013/579319

38. Pushparaj PN, Low HK, Manikandan J, Tan BKH, Tan CH. Anti-Diabetic Effects of Cichorium Intybus in Streptozotocin-Induced Diabetic Rats. *J Ethnopharmacol* (2007) 111:430–4. doi: 10.1016/j.jep.2006.11.028

39. Tousch D, Lajoix A-D, Hoby E, Azazy-Mihau J, Ferrare K, Jannahatt C, et al. Chioric Acid, a New Compound Able to Enhance Insulin Release and Glucose Uptake. *Biochem Biophys Res Commun* (2008) 377:131–5. doi: 10.1016/j.bbrc.2008.09.088

40. Chandra K, Jain V, Jabin A, Dwivedi S, Joshi S, Ahmad S, et al. Effect of Cichorium Intybus Seeds Supplementation on the Markers of Glycemic Control, Oxidative Stress, Inflammation, and Lipid Profile in Type 2 Diabetes Mellitus: A Randomized, Double-Blind Placebo Study. *Phytother Res* (2020) 34:1609–18. doi: 10.1002/ptr.6624

41. Nishimura M, Okhawara T, Kanayama T, Kitagawa K, Nishimura H, Nishihira J. Effects of the Extract From Roasted Chicory (Cichorium Intybus L.) Root Containing Inulin-Type Fructans on Blood Glucose, Lipid Metabolism, and Fecal Properties. *J Tradit Complement Med* (2015) 5:161–7. doi: 10.1016/j.jtcme.2014.11.016

42. Singh N, Rao AS, Nandala A, Kumar S, Yadav SS, Ganaie SA, et al. Phytochemical and Pharmacological Review of Cinnamomum Verum J. L. *Front Pharm Sci* (2021) 338:127773. doi: 10.1016/j.foodchem.2020.127773

43. El-Dessoky GE, Aboul-Soud MAM, Al-Nemair KS. Antidiabetic and Hypolipidemic Effects of Ceylon Cinnamon (Cinnamomum Verum) in
Alam et al. Antidiabetic Phytochemicals for Drug Discovery and Development

64. Tran N, Pham B, Le L. Bioactive Compounds in Anti-Diabetic Plants: From

59. Zare R, Nadjarzadeh A, Zarshenas MM, Shams M, Heydari M. Efficacy of Cinnamon in Patients With Type II Diabetes Mellitus: A Randomized Controlled Clinical Trial. Clin Nutr Edinb Scotl (2019) 38:549–56. doi: 10.1016/j.clnu.2018.03.003

63. Medagama AB. The Glycaemic Outcomes of Cinnamon, a Review of the Experimental and Clinical Trials. Nutr J (2015) 14:108. doi: 10.1186/s12937-015-0098-9

60. Joshi SC, Jain PK, Sharma P. Antiatherosclerotic and Lipid-Lowering Effects of Cinnamomum Zeylanicum (Ceylon Cinnamon) in Healthy Adults: A Phase I Clinical Trial. BMC Complement Altern Med (2017) 17:550. doi: 10.1186/s12906-017-2067-7

69. Correia J. The Effect of Cinnamon on Pancreatic Islet Function and the Isolation of a Potentially Useful New Antihyperglycaemic Agent. Diabetes Metab J Br Diabet Assoc (2017) 41:107–11. doi: 10.1016/j.dia.2016.10.001

73. Lekshmi PC, Arimboor R, Raghu KG, Menon AN. Turmerin, the Antioxidant and Safety of Cinnamomum Zeylanicum (Ceylon Cinnamon) in Healthy Subjects. Nutr J (2013) 12:51. doi: 10.1186/1475-2891-12-51

78. Sutrisno A, Rijal AS, Salehi B, et al. Hypolipidemic Activity of Cinnamomum Zeylanicum in Normal and Diabetic Persons. Biomed Res Int (2014) 2014:830285. doi: 10.1155/2014/830285

80. Devasagayam PA, Jaiswal SK, Maheshwari A, et al. Hypoglycemic Activity of Cinnamomum Zeylanicum: Potential for Increasing Antioxidant Content in Functional Foods for Diabetic Population. Food Res Int (2016) 85:10–8. doi: 10.1016/j.foodres.2016.04.006

87. Chaudhary S, Kalra SN, Juyal S, et al. The Antidiabetic Potential of the Ethanobotanical Approach to Drug Discovery. Indian J Diabetic Assoc (2019) 31:2914–24. doi: 10.4103/1939-8407.75707

88. Pivari F, Mingione A, Brasacchio C, Soldati L. Curcumin and Type 2 Diabetes Mellitus: Prevention and Treatment. Nutrients (2019) 11:1837. doi: 10.3390/nu11081837

89. Marojoy A. Eucaele Undulata Thumb: Review of its Botany, Ethnomedicinal Uses, Phytochemistry and Biological Activities. Asian Pac J Trop Med (2017) 10:1030–6. doi: 10.1016/j.ajptm.2017.10.005

90. Deutschlander MS. Isoflavonoids from Euclea Undulata Thumb. Harfield, Pretoria: University of Pretoria (2010). Doctoral dissertation.

91. Deutschlander MS, Lai N, Van de Venter M, Hussein AA. Hypoglycemic Evaluation of a New Triterpene and Other Compounds Isolated From Euclea Undulata Thumb. Var. Myrtina (Ebenaceae) Root Bark. J Ethnopharmacol (2011) 133:1091–5. doi: 10.1016/j.jep.2011.05.038

92. Tiwari P, Mishra BN, Sangwan NS. Phytochemical and Pharmacological Properties of Gymnema Sylvestre: An Important Medicinal Plant. BioMed Res Int (2014) 2014:830285. doi: 10.1155/2014/830285

93. Khare AK, Tondon RN, Tewari JP. Hypoglycaemic Activity of an Indigenous Plant Gymnema Sylvestre leaf Extract in the Control of Blood Glucose in Insulin-Dependent Diabetes Mellitus Patients. J Ethnopharmacol (2017) 189:1030–7. doi: 10.1016/j.jep.2017.02.026

94. Thakur G, Sharma R, Sanodiya BS, Pandey M, Bisen P. Gymnema Sylvestre: An Alternative Therapeutic Agent for Management of Diabetes. Pharm Sci Technol Today (2012) 2:2001–6. doi: 10.7324/JAPS.2012.21201

95. Leach MJ. Gymnema Sylvestre for Diabetes Mellitus: A Systematic Review. J Altern Complement Med N Y N (2017) 13:977–83. doi: 10.1089/acm.2006.6387

96. Khare AK, Tondon RN, Tewari JP. Hypoglycaemic Activity of an Indigenous Drug (Gymnema Sylvestre, ‘Gurmār’) in Normal and Diabetic Persons. Indian J Pharmacol (1983) 27:257–8.

97. Kumar SN, Mani UV, Mani I. An Open Label Study on the Supplementation of Gymnema Sylvestre: An Important Medicinal Plant. Alternat Ther Med (2010). Doctoral dissertation, University of Pretoria.

98. Baskaran K, Kizar Ahamath B, Radha Shanmugasundaram K, Shanmugasundaram ER. Antidiabetic Effect of a Leaf Extract From Gymnema Sylvestre in Non-Inulin-Dependent Diabetes Mellitus Patients. J Ethnopharmacol (1999) 30:295–300. doi: 10.1016/S0378-8741(98)00108-6

99. Shanmugasundaram ERB, Rajeswari G, Baskaran K, Kumar BRR, Shanmugasundaram KR, Ahmath BK. Use of Gymnema Sylvestre Leaf Extract in the Control of Blood Glucose in Insulin-Dependent Diabetes Mellitus. J Ethnopharmacol (1990) 30:281–94. doi: 10.1016/0378-8741(90)90107-5

100. Puranik R, Jadhav S, Patil V, et al. Antidiabetic Potential of the Ethanobotanical Approach to Drug Discovery. Indian J Diabetic Assoc (2019) 31:2914–24. doi: 10.4103/1939-8407.75707

101. Wang R, Xu Y, Zhou S, Tian S, Cao S. Isolation and Identification of Novel Anti-Diabetic Phytochemicals from Dracaena Fragrans. J Nat Prod (2019) 82:394. doi: 10.1021/acs.jnatprod.9b00513

102. Xu W, Lu Z, Wang X, Zheng MH, Lin M, Li C, et al. Gymnema Sylvestre Exerts Hypoglycemic Effects by Regulating the P38/ERK Signaling
Antidiabetic Phytochemicals for Drug Discovery and Development

101. Arteaga S, Andrade-Cetto A, Ca... 102. Luo J, Chuang T, Cheung J, Quan J, Tsai J, Sullivan C, et al. Masoprocol

107. McKay DL, Blumberg JB. A Review of the Bioactivity and Potential Health

106. Garc... 96. Cao Y, Sun Y, Zou S, Li M, Xu X. Orally Administered Baker

105. Kato A, Minoshima Y, Yamamoto J, Adachi I, Watson AA, Nash RJ. –56. doi: 10.2134/agronmono26.24

97. Cao Y, Zou S, Xu H, Li M, Tong Z, Xu M, et al. Hypoglycemic Activity of the

98. Re... 95. Von Bothmer R, Jacobsen N. Origin, Taxonomy, and Related Species. Barley

99. Reid DA. Morphology and Anatomy of the Barley Plant. Barley (1985) 26:73–101. doi: 10.2134/agronmono26.24

110. Darvishpadok A, Azemi M, Namjooan F, Khodayar M, Ahmadpour F, Panahi M. Effect of Matricaria Chamomilla L. on Blood Glucose and Glycosylated Hemoglobin in Femile Fertile Diabetic Rates. Res Pharm Sci (2012) 7:19

111. Kato A, Minoshima Y, Yamamoto J, Adachi I, Watson AA, Nash RJ. Protective Effects of Dietary Chamomile Tea on Diabetic Complications. J Agric Food Chem (2008) 56:8206–11. doi: 10.1021/jf8014365

112. Panda S, Kar A. Apigenin (4’,5,7-Trihydroxyflavone) Regulates Hyperglycaemia, Thyroid Dysfunction and Lipid Peroxidation in Alloxan-Induced Diabetic Mice. J Pharm Pharmacol (2007) 59:1543–8. doi: 10.1211/jpp.59.11.0012

113. Rafraf M, Zemestani M, Asghari-Jafarabadi M. Effectiveness of Chamomile Tea on Glycemic Control and Serum Lipid Profile in Patients With Type 2 Diabetes. J Endocr Invest (2015) 38:163–70. doi: 10.1007/s40618-014-0170-x

114. Kasef F, Yazdanpanah, Zeinab, Biergani AN, Yazdi NB, Yazdanpanah, et al. The Effect of Chamomile (Matricaria recutita L.) Infusion on Blood Glucose, Lipid Profile and Kidney Function in Type 2 Diabetic Patients: A Randomized Clinical Trial. Prog Nutr (2018) 20:110–8. doi: 10.23751/pn.v20i1-S.5884

115. Joseph B, Jini D. Antidiabetic Effects of Momordica Charantia (Bitter Melon) and its Medicinal Potency. Asian Pac J Trop Dis (2013) 3:93–102. doi: 10.1016/j.japt.2013(08)0052-3

116. Harinantenaina I, Tanaka M, Takaoka S, Oda M, Mogami O, Uchida M, et al. Momordica Charantia Constituents and Antidiabetic Screening of the Isolated Major Compounds. Chem Pharm Bull (Tokyo) (2006) 54:1017–21. doi: 10.1248/cpb.54.1017

117. Ma J, Whittaker P, Keller AC, Mazzola EP, Pawar RS, White KD, et al. Cucurbitane-Type Triterpenoids From Momordica Charantia. Planta Med (2010) 76:1758–61. doi: 10.1055/s-0030-1249807

118. Krawinkel MB, Ludwig C, Swai ME, Yang Ry, Chun KP, Habicht SD. Bitter Gourd Reduces Elevated Fasting Plasma Glucose Levels in an Intervention Study Among Prediabetics in Tanzania. J Ethnopharmacol (2018) 216:1–7. doi: 10.1016/j.jep.2018.01.016

119. Peter EK, Kasimi FM, Deyno S, Mtewa A, Nagendrappa PB, Tolo CU, et al. Momordica Charantia L. lowers Elevated Glycaemia in Type 2 Diabetes Mellitus Patients: Systematic Review and Meta-Analysis. J Ethnopharmacol (2019) 231:31–24. doi: 10.1016/j.jep.2018.10.033

120. Cortez-Navarrete M, Martinez-Abundis E, Perez-Rubio KG, Gonzalez-Ortiz M, Méndez-Del Villar M. Momordica Charantia Administration Improves Insulin Secretion in Type 2 Diabetes Mellitus. J Med Food (2018) 21:672–7. doi: 10.1089/jmf.2017.0114

121. Akhtar MS. Trial of Momordica Charantia Linn (Karela) Powder in Patients With Maturity-Onset Diabetes. JPMA J Pak Med Assoc (1982) 32:106–7.

122. Muhammad HI, Asmawi MZ, Khan NAK. A Review on Promising Phytochemical, Nutritional and Glycemic Control Studies on Moringa oleifera L. in Experimental Diabetes. Fitoterapia (2008) 80:475–80. doi: 10.1016/j.fitote.2009.06.009

123. Villarruel-Lopez A, Lopez-de la Mora DA, Vazquez-Paulino OD, Puebla-Mora AG, Torres-Vitela MR, Guerrero-Quiroz LA, et al. Effect of Moringa Oleifera Consumption on Diabetic Rats. BMC Complement Altern Med (2018) 18:127. doi: 10.1186/s12906-018-2180-2

124. Sandoval MAS, Jimeno CA. Effect of Malunggay (Moringa oleifera) Capsules on Lipid and Glucose Levels. Acta Med Philippina (2013) 47 (3):22–7. doi: 10.4795/amp.v47is3.1285

125. Sandoval MAS, Jimeno CA. Effect of Malunggay (Moringa oleifera) Extract on Blood Lipid Levels. Fitoterapia (2009) 80:475–7. doi: 10.1016/j.fitote.2009.06.009

126. Tian S, Tang M, Zhao B. Current Anti-Diabetes Mechanisms and Clinical Trials Using Morus Alba L. J Tradit Chin Med Sci (2016) 3:3–8. doi: 10.1016/j.jt cms.2016.04.001

Pathway and Fatty Acid Metabolism Signaling Pathway. Nutr Diabetes (2020) 10:31. doi: 10.1038/s41387-020-00134-z

94. Reid DA. Morphology and Anatomy of the Barley Plant. Barley (1985) 26:73–101. doi: 10.2134/agronmono26.24

95. Von Bothmer R, Jacobsen N. Origin, Taxonomy, and Related Species. Barley (1985) 26:19–56. doi: 10.2134/agronmono26.24
166. Takino Y, Koshioka M, Kagawuchi M, Miyahara T, Tanizawa H, Ishii Y, et al. Quantitative Determination of Bitter Components in Swertiae Herba. Plantia Med (1990) 38:351–5. doi: 10.1055/s-2008-1074888

167. Sharma S, Anjaneyulu M, Kulkarni SK, Chopra K. Resveratrol, a Polyphenolic Phytorexin, Attenuates Diabetic Nephropathy in Rats. Pharmacology (2006) 75:25–7. doi: 10.1159/000089720

168. Rizvi SI, Pandey KB. Activation of the Erythrocyte Plasma Membrane Redox System by Resveratrol: A Possible Mechanism for Antioxidant Properties. Pharmacol Rep (2010) 62:726–32. doi: 10.1177/1445614310070330-3

169. Palsamy P, Subramanian S. Ameliorative Potential of Resveratrol on Proinflammatory Cytokines, Hyperglycemia Mediated Oxidative Stress, and Pancreatic Beta-Cell Dysfunction in Streptozotocin-Nicotinamide-Induced Diabetic Rats. J Cell Physiol (2010) 224:423–32. doi: 10.1002/jcp.21138

170. Pandey KB, Rizvi SI. Resveratrol Up-Regulates the Erythrocyte Plasma Membrane Redox System and Mitigates Oxidation-Induced Alterations in Erythrocytes During Aging in Humans. Rejuvenation Res (2013) 16:322–40. doi: 10.1089/rej.2013.1419

171. Chi T-C, Chen W-P, Chi T-L, Kuo T-F, Lee S-S, Cheng J-T, et al. Phosphatidylinositol-3-Kinase Is Involved in the Antihyperglycemic Effect Induced by Resveratrol in Streptozotocin-Induced Diabetic Rats. Life Sci (2007) 80:1713–20. doi: 10.1016/j.lfs.2007.02.002

172. Singh D, Chaudhuri PK. Chemical and Pharmacology of Tinospora Cordifolia. J Ethnopharmacology, Phytochemistry, and Pharmacology. Evid Based Complement Alternat Med (2016) 2016:9232593. doi: 10.1155/2016/9232593

173. Kumar V. A Clinical Trial to Assess the Antidiabetic, Antislipidemic and Antioxidant Activities of Tinospora Cordifolia in Management of Type – 2 Diabetes Mellitus. J Pharm Sci Res (2016) 7:757–64. doi: 10.1002/psr.20140975-8232.7(2).757-64

174. Wani SA, Kumar P. Fenugreek: A Review on its Nutraceutical Properties and Utilization in Various Food Products. J Saudi Soc Agric Sci (2016) 17(2):97–106. doi: 10.1016/j.jssas.2016.01.007

175. Patil MR, Mishra S. Hypoglycemic Activity of Alkaloidal Fraction of Tinospora Cordifolia. Phytomed Int J Phytother Phytopharm (2011) 18:1045–52. doi: 10.1016/j.phymed.2011.05.006

176. Kumar V. A Clinical Trial to Assess the Antidiabetic, Antislipidemic and Antioxidant Activities of Tinospora Cordifolia in Management of Type – 2 Diabetes Mellitus. J Pharm Sci Res (2016) 7:757–64. doi: 10.1002/psr.20140975-8232.7(2).757-64

177. Sang S, Ahmedna M, El-Shazly M, Cheng Y-B, et al. 6-Paradol and 6-Shogaol, the Pungent Compounds of Ginger, Promote Glucose Utilization in Adipocytes and Myotubes, and 6-Paradol Reduces Blood Glucose in High-Fat Diet-Fed Mice. Int J Mol Sci (2017) 18:168. doi: 10.3390/ijms1810168

178. Samad MB, Mohsin S, Raza BA, Hussain MT, Mahzabeen S, Unnoor N, et al. [-Gingerol, From Zingiber Officinale, Potentiates G1P-1 Mediated Glucolysis in Ginger and Apple Alleviates Hyperglycemia in Mice With High Fat Diet-Induced Obesity via Nrf2 Mediated Pathway. Food Chem (2017) 226:79–88. doi: 10.1016/j.foodchem.2017.01.056

179. Wei C-K, Tsai Y-H, Korinek M, Hung P-H, El-Shazly M, Cheng Y-B, et al. 6-Paradol and 6-Shogaul, the Pungent Compounds of Ginger, Promote Glucose Utilization in Adipocytes and Myotubes, and 6-Paradol Reduces Blood Glucose in High-Fat Diet-Fed Mice. Int J Mol Sci (2017) 18:168. doi: 10.3390/ijms1810168

180. Palsamy P, Subramanian S. Ameliorative Potential of Resveratrol on Proinflammatory Cytokines, Hyperglycemia Mediated Oxidative Stress, and Pancreatic Beta-Cell Dysfunction in Streptozotocin-Nicotinamide-Induced Diabetic Rats. J Cell Physiol (2010) 224:423–32. doi: 10.1002/jcp.21138

181. Chi T-C, Chen W-P, Chi T-L, Kuo T-F, Lee S-S, Cheng J-T, et al. Phosphatidylinositol-3-Kinase Is Involved in the Antihyperglycemic Effect Induced by Resveratrol in Streptozotocin-Induced Diabetic Rats. Life Sci (2007) 80:1713–20. doi: 10.1016/j.lfs.2007.02.002

182. Wein S, Behm N, Petersen RK, Kristiansen K, Wolffram S. Quercetin from Vitis Vinifera (Grape) and Its Bioactive Compounds. Nutr J (2003) 2:189. doi: 10.1186/1475-2891-2-189

183. Sampath C, Rashid MR, Sang S, Ahmedna M. Specific Bioactive Compounds in Ginger and Apple Alleviate Hyperglycemia in Mice With High Fat Diet-Induced Obesity via Nrf2 Mediated Pathway. Food Chem (2017) 226:79–88. doi: 10.1016/j.foodchem.2017.01.056
Glycosidase Inhibitory Activities. *J Ocean Univ China* (2015) 14:651–62. doi: 10.1007/s11802-015-2684-3

238. Park MJ, Ryu HK, Han JS. Effects of Laminaria Japonica Extract Supplement on Blood Glucose, Serum Lipids and Antioxidant Systems in Type II Diabetic Patients. *J Korean Soc Food Sci Nutr* (2007) 36(11):1391–8. doi: 10.3746/jkfn.2007.36.11.1391

239. Muruganandan S, Srinivasan K, Gupta S, Gupta PK, Lal J. Effect of Mangiferin on Hyperglycemia and Atherogenicity in Streptozotocin Diabetic Rats. *J Ethnopharmacol* (2005) 97:497–501. doi: 10.1016/j.jep.2004.12.010

240. Aderibigbe AO, Emudianughe TS, Lawal BS. Evaluation of the Antidiabetic Action of Mangifera Indica in Mice. *Phytother Res* (2001) 15:456–8. doi: 10.1002/ptr.859

241. Villas Boas GR, Rodrigues Lemos JM, de Oliveira MW, Dos Santos RC, Stefanello da Silveira AP, Barbieri Bacha F, et al. Aqueous Extract From Mangifera Indica Linn. (Anacardiaceae) Leaves Exerts Long-Term Hypoglycemic Effect, Increases Insulin Sensitivity and Plasma Insulin Levels on Diabetic Wistar Rats. *PloS One* (2020) 15:e0227105. doi: 10.1371/journal.pone.0227105

242. Evans SF, Meister M, Mahmood M, Eldoumi H, Peterson S, Perkins-Vezzie P, et al. Mango Supplementation Improves Blood Glucose in Obese Individuals. *Nutr Metab Insights* (2014) 7:NLM17028. doi: 10.4137/NLM17028

243. Na L, Zhang Q, Jiang S, Du S, Zhang W, Li Y, et al. Mangiferin Levels on Diabetic Wistar Rats. Hypoglycemic Effect, Increases Insulin Sensitivity and Plasma Insulin Levels in Obese Mice. *J Ocean Univ China* 2007.36.11.1391

244. Kobayashi M, Akaki J, Ninomiya K, Yoshikawa M, Muraoka O, Morikawa T, Singh RG, Rathore SS, Kumar R, Usha, Agarwal A, Dubey GP. Jeykodi S, Deshpande J, Juturu V. Salacia Extract Improves Postprandial Glucose and Insulin Response: A Randomized Double-Blind, Placebo-Controlled, Placebo-Controlled, Cross-Over Trial. *Tetrahedron Lett* (1998) 46:1339–40. doi: 10.1016/cpb.46.1339

245. Kajimoto O, Kawamori S, Shimoda H, Kawahara Y, Hirata H, Takahashi T. Effects of a Diet Containing Salacia Reticulata on Mild Type 2 Diabetes in Humans. A Placebo-Controlled, Cross-Over Trial. Nippon Eiyo Shokuryo Gakkaishi J. *Ipn Soc Nutr Food Sci* (2003) 53:199–205. doi: 10.4327/jnfs.53.199

246. Radha R, Amrithiveni M. Role of Medicinal Plant Salacia Reticulata in the Management of Type II Diabetic Subjects. *Anc Sci Life* (2009) 29:14–6.

247. Shivasprasad H, Bhanumathy M, Sushma G, Midhun T, Ravendrak K, Sushma K, et al. Salacia Reticulata Improves Serum Lipid Profiles and Glycemic Control in Patients With Pre diabetes and Mild to Moderate Hyperlipidemia: A Double-Blind, Placebo-Controlled, Randomized Trial. *J Med Food* (2013) 16:564–8. doi: 10.1089/jmf.2013.2751

248. Pamunuwga V, Karunaratne DN, Waisundara VY. Antidiabetic Properties, Bioactive Constituents, and Other Therapeutic Effects of Scoparia Dulcis. *Evid Based Complement Alternat Med* (2016) 2016:8243215. doi: 10.1155/2016/8243215

249. Senadhira SPAS, Ekanayake S, Wanigatunge C. Anti-Hyperglycaemic Effects of Herbal Porridge Made of Scoparia Dulcis Leaf Extract in Diabetics – A Randomized Crossover Clinical Trial. *BMC Complement Altern Med* (2015) 15:410. doi: 10.1186/s12906-015-0935-6

250. Jeppesen PB, Gregersen S, Alstrup KK, Hermansen K. Stevioloside Induces Anti hyperglycaemic, Insulinotropic and Glucagonostatic Effects In Vivo: Studies in the Diabetic Goto-Kakizaki (GK) Rats. *Phytochem Int J Phytother Phytopharm* (2002) 9:9–14. doi: 10.1094/jpfh-7113-00081

251. Ritu M, Nandini J. Nutritional Composition of Stevia Rebaudiana, a Sweet Herb, and its Hypoglycaemic and Hypolipidaemic Effect on Patients With Non-Inulin Dependent Diabetes Mellitus. *J Sci Food Agric* (2016) 96:4231–4. doi: 10.1002//jsfa.7627

252. Mishra N. An Analysis of Antidiabetic Activity of Stevia Rebaudiana Extract on Diabetic Patient. *J Nat Sci Res* (2012) 1:1.

253. Gregersen S, Jeppesen PB, Holst JI, Hermansen K. Anti hyperglycaemic Effects of Stevioside in Type 2 Diabetic Subjects. *Metabolism* (2004) 53:73–6. doi: 10.1016/metabolism.2003.07.013

254. Antonsen V, Martin CK, Han H, Coulon S, Cefalu WT, Geiselman P, et al. Effects of Stevia, Aspartame, and Sucrose on Food Intake, Satiety, and Postprandial Glucose and Insulin Levels. *Appetite* (2010) 55:37–43. doi: 10.1016/j.appet.2010.03.009

255. Agu KC, Eluehike N, Ofeimun RO, Abile D, Ideho G, Ogedengbe MO, et al. Possible Anti-Diabetic Potentials of Annona Muricata (Soursop): Inhibition of α-Amylase and α-Glucosidase Activities. *Clin Pyscoph* (2019) 5(1):1–13. doi: 10.1186/s40816-019-0116-0

256. Youn JY, Park HY, Cho KH. Anti-Hyperglycaemic Activity of Commelina Communis L.: Inhibition of α-Glucosidase. *Diabetes Res Clin Pract* (2004) 66:149–55. doi: 10.1016/diabres.2003.08.015

257. Gao Y, Zhang Y, Zhu J, Li B, Li Z, Zhu W, et al. Recent Progress in Natural Products as DPP-4 Inhibitors. *Future Med Chem* (2015) 7(8):1079–89. doi: 10.4155/fmc.15.49

258. Elya B, Handayani R, Sauriasiri R, Hasyiyyat US, Permana IT, Permatasari YI. Antidiabetic Activity and Phytochemical Screening of Extracts From Indonesian Plants by Inhibition of Alpha Amylase, Alpha Glucosidase and Dipeptidyl Peptidase IV. *Pakistan J Biol Sci* (2015) 18(6):2789. doi: 10.2973/pjab.2015.2789

259. Mechchate H, Es-Safi I, Loub A, Alqahtani AS, Nasr FA, Noman OM, et al. *In Vitro Alpha-Amylase and Alpha-Glucosidase Inhibitory Activity and In Vivo Antidipabetic Activity of Withania Frutescens L. Foliar Extract. Molecules* (2021) 26(2):89. doi: 10.3390/molecules26020923

260. Adisakwattana S, Moonsan P, Tiyboch-Ammun S. Insulin-Releasing Properties of a Series of Cinnamic Acid Derivatives In Vitro and In Vivo. *J Agric Food Chem* (2008) 56(17):7838–44. doi: 10.1021/jf080120v

261. Zhao C, Yang C, Wai STC, Zhang Y, Portillo M, Paoli P, et al. Regulation of Glucose Metabolism by Bioactive Phytochemicals for the Management of
Type 2 Diabetes Mellitus. *Critt Rev Food Sci Nutr* (2019) 59(6):830–47. doi: 10.1080/10408398.2018.1501658

272. Schenk S, Sabeti M, Oldfjord JM. Insulin Sensitivity: Modulation by Nutrients and Inflammation. *J Clin Invest* (2008) 118(9):2992–3002. doi: 10.1177/0003522406290693

273. Ghorbani A, Rashidi R, Shafiee-Nick R. Flavonoids for Preserving Pancreatic Beta Cell Survival and Function: A Mechanistic Review. *Biomed Pharmacother* (2019) 111:947–57. doi: 10.1016/j.biopha.2018.12.127

274. Oh YS. Plant-Derived Compounds Targeting Pancreatic Beta Cells for the Treatment of Diabetes. *Evidence-Based Complement Altern Med* (2015) 2015:629863. doi: 10.1155/2015/629863

275. Ghazanfari-Sarabi S, Habihi-Rezaei M, Eshraghi-Naeeni R, Moosavi-Movahedi AA. Prevention of Haemoglobin Glycation by Acetylsalicylic Acid (ASA): A New View on Old Mechanism. *PloS One* (2019) 14(4): e0214725. doi: 10.1371/journal.pone.0214725

276. Welsh KJ, Kirkman MS, Sacks DB. Role of Glycated Proteins in the Diagnosis and Management of Diabetes: Research Gaps and Future Directions. *Diabetes Care* (2016) 39(8):1299–306. doi: 10.2337/dc15-2727

277. Harding JJ, Ganca E. Protection Against Glycation and Similar Post-Translational Modifications of Proteins. *Biochim Biophys Acta (BBA)-Proteins Proteomics* (2006) 1764(9):1436–46. doi: 10.1016/j.bbapap.2006.08.001

278. Baggio LL, Drucker DJ. Biology of Incretins: GLP-1 and GIP. *Gastroenterology* (2007) 132(6):2131–57. doi: 10.1016/j.gastro.2007.03.054

279. Saefer JCF, Kushner P, Aguilar R. User’s Guide to Mechanism of Action and Clinical Use of GLP-1 Receptor Agonists. *Postgrad Med* (2015) 127(8):818–26. doi: 10.1080/00325481.2015.1090295

280. Huang S, Czech MP. The GLUT4 Glucose Transporter. *Cell Metab* (2007) 5(4):237–52. doi: 10.1016/j.cmet.2007.03.006

281. Tunduguru R, Thurmond DC. Promoting Glucose Transporter-4 Vesicle Trafficking Along Cytoskeletal Tracks: PAK-Ing Them Out. *Front Endocrinol* (2017) 8:329. doi: 10.3389/fendo.2016.00435

282. Ruhil S, Balhara M, Dhanikhar S, Chhillar AK. Aegle marmelos (Linn.) Correa: A potential Source of Phytomedicine. *J Med Plants Res* (2011) 5(9):1497–507. doi: 10.5897/JMPR.2010048

283. Bates SH, Jones RB, Bailey CJ. Insulin-Like Effect of Pinitol. *British J Pharmacol* (2000) 130(8):1944–8. doi: 10.1038/sj.bjp.0703523

284. Unger J, Parkin CG. Type 2 Diabetes: An Expanded View of Pathophysiology and Therapy. *Postgrad Med* (2010) 122:145–57. doi: 10.3810/pgm.2010.05.2152

285. Osadebe PO, Odoh EU, Uzor PF. Natural Products as Potential Sources of Antidiabetic Drugs. *Br J Pharm Res* (2014) 4(17):2075–95. doi: 10.9734/BJPR/2014/8382

286. Verspoor EJ. Recommended Testing in Diabetes Research. *Planta Med* (2002) 68(07):581–90. doi: 10.1055/s-2002-32894

287. Chopra A, Saluja M, Tillo G. Ayurveda—modern Medicine Interface: A Critical Appraisal of Studies of Ayurvedic Medicines to Treat Osteoarthritis and Rheumatoid Arthritis. *J Ayurveda Integ Med* (2010) 1(3):190. doi: 10.4103/0975-9476.72620

288. Piggott AM, Karuso P. Quality, Not Quantity: The Role of Natural Products and Chemical Proteomics in Modern Drug Discovery. *Comb Chem High Throughput Screen* (2004) 7(7):su27–30. doi: 10.2174/1386207043328409

289. Ortholdan JY, Ganesan A. Natural Products and Combinatorial Chemistry: Back to the Future. *Curr Opin Chem Biol* (2004) 8(3):271–80. doi: 10.1016/j.cob.2004.04.011

290. Kennedy DO, Wightman EL. Herbal Extracts and Phytochemicals: Plant Secondary Metabolites and the Enhancement of Human Brain Function. *Adv Nutr* (2011) 2(1):32–50. doi: 10.3945/an.110.000117

291. Newman DJ, Cragg GM. Natural Products as Sources of New Drugs Over the 30 Years From 1981 to 2010. *J Natural Prod* (2012) 75(3):311–35. doi: 10.1021/np300906x

292. Hung DY, Jamison TF, Schreiber SL. Understanding and Controlling the Cell Cycle With Natural Products. *Chem Biol* (1996) 3(8):623–39. doi: 10.1016/S1074-5519(96)90129-5

293. Montecucco F, Steffens S, Mach F. Insulin Resistance: A Proinflammatory State Mediated by Lipid-Induced Signaling Dysfunction and Involved in Atherosclerotic PlaqueInstability. *Mediators Inflamm* (2008) 2008:767623. doi: 10.1155/2008/767623

294. Taton J, Czech A, Piatkiewicz P. Insulin as the Main Regulator of Cellular Glucose Utilization—Aetiological Aspects of Insulin Resistance. *Endokrynol Pol* (2010) 61:388–94.

295. Taheri Rouhi SZ, Sarker M, Rahmat A, Alkhatani SA, Othman F. The Effect of Pomegranate Fresh Juice Versus Pomegranate Seed Powder on Metabolic Indices, Lipid Profile, Inflammatory Biomarkers, and the Histopathology of Pancreatic Islets of Langerhans in Streptozotocin-Nicotinamide Induced Type 2 Diabetic Sprague–Dawley Rats. *BMC Complement Altern Med* (2017) 17:156. doi: 10.1186/s12906-017-1667-6

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## GLOSSARY

| Abbreviation | Description |
|--------------|-------------|
| ADA          | American Diabetes Association |
| AGEs         | Advanced glycation end-product |
| AMP          | Adenosine monophosphate |
| AMPK         | 5′-adenosine monophosphate activated protein kinase |
| b3-AR        | Beta-3 adrenergic receptor |
| BMI          | Body mass index |
| CIE          | Cichorium intybus |
| CML          | Carboxymethyl-lysine |
| DCMM         | Dichloromethane-methanol |
| DNJ          | 1-deoxynojirimycin |
| DPP-4        | Dipeptidyl peptidase-4 |
| eNOS         | Endothelial nitric oxide synthase |
| FBS          | Fetal bovine serum |
| FPG          | Fasting Plasma Glucose |
| GDM          | Gestational diabetes mellitus |
| GI           | Glycemic index |
| Glc–6-Pase   | Glucose-6-phosphatase |
| GLP-1        | Glucagon-like peptide-1 |
| GLUT-4       | Glucose transporter-4 |
| GSK3B        | Glycogen synthase kinase 3B |
| HbA1c        | Hemoglobin A1c |
| HDL-c        | High-density lipoprotein cholesterol |
| HepG2        | Hepatoma cell line |
| HMBA         | 2-hydroxy-4-methoxy benzoic acid |
| HMG          | 3-hydroxy-3-methylglutaric acid |
| HOMA-IR      | Homeostatic model assessment insulin resistance |
| IL-6         | Interleukin-6 |
| INR          | International normalized ratio |
| INS-1E       | Rat insulinoma cell line INS-1E |
| IRS1         | Insulin receptor substrate1 |
| K-ATP        | ATP-sensitive potassium channel |
| LC/MS        | Liquid chromatography-mass spectrometry |
| LDL-c        | Low-density lipoprotein cholesterol |
| MAPK         | Mitogen-activated protein kinase |
| NADPH        | Nicotinamide adenine dinucleotide phosphate |
| NMN          | Nuclear magnetic resonance |
| NO           | Nitric oxide |
| NOX4         | Nicotinamide adenine dinucleotide phosphate oxidase 4 |
| p-AKT        | Phosphorylated Akt |
| PKC          | Protein kinase C |
| PPAR         | Peroxisome proliferator-activated receptor |
| PTP1B        | Protein tyrosine phosphatase 1B |
| PX-407       | Poloxamer-407 |
| SOOT         | Serum glutamic oxaloacetic transaminase |
| SGPT         | Serum glutamic pyruvic transaminase |
| SLM          | Solid lipid microparticles |
| STZ          | Streptozotocin |
| TC           | Total cholesterol |
| TG           | Triglycerides |
| 2HPP         | 2 hour postprandial glucose |
| [Ca2+]i      | Calcium ion |
| ATP          | Adenosine triphosphate |
| PI3K         | Phosphatidylinositol 3-kinase |
| PKB          | Protein kinase B |
| GIP          | Glucose dependent insulinotropic polypeptide |
| MAMs         | mitochondria-associated membranes |
| mTORC2       | Mammalian target of rapamycin complex 2 |
| PKC-λ/ζ      | Protein kinase C zeta/lambda |
| PTEN         | Phosphatase and tensin homolog |
| PP2A         | Protein phosphatase 2 |
| GAD          | Glutamic acid decarboxylase |
| CD4          | Cluster of differentiation 4 |
| CD8          | Cluster of differentiation 8 |