Effects of Ficus carica paste on constipation induced by a high-protein feed and movement restriction in beagles

Hong-Geun Oh, Hak-Yong Lee, Min-Young Seo, Young-Rye Kang, Jung-Hoon Kim, Jung-Woo Park, Ok-Jin Kim, Hyang-Im Back, Sun-Young Kim, Mi-Ra Oh, Soo-Hyun Park, Min-Gul Kim, Ji-Young Jeon, Min-Ho Hwang, Sook-Jeong Shin, Soo-Wan Chae

1Huvet Co., Ltd, Iksan, Korea
2Center for Animal Resources Development, Wonkwang University, Iksan, Korea
3Clinical Trial Center for Functional Foods, and Chonbuk National University Hospital, Jeonju, Korea
4Clinical Trial Center, Chonbuk National University Hospital, Jeonju, Korea
5Department of Pharmacology, Chonbuk National University Medical School, Jeonju, Korea

Constipation is one of the most common functional digestive complaints worldwide. We investigated the laxative effects of figs (Ficus carica L) in a beagle model of constipation induced by high protein diet and movement restriction. The experiments were consecutively conducted over 9 weeks divided into 3 periods of 3 weeks each. All 15 beagles were subjected to a non-treatment (control) period, a constipation induction period, and a fig paste treatment period. We administered fig paste (12 g/kg daily, by gavage) for 3 weeks following a 3-week period of constipation induction in dogs. Segmental colonic transit time (CTT) was measured by counting radiopaque markers (Kolomark) using a radiograph performed every 6 h after feeding Kolomark capsules, until capsules were no longer observed. Fig paste significantly increased fecal quantity in constipated dogs, and segmental CTT was also reduced following fig paste administration. There were no significant differences in feed intake, water intake, body weight, or blood test results, between the constipation and fig paste administration periods. Our results demonstrate that fig is an effective treatment for constipation in beagles. Specifically, stool weight increased and segmental CTT decreased. Fig pastes may be useful as a complementary medicine in humans suffering from chronic constipation.

Key words: Constipation, fig paste, segmental colonic transit time, Kolomark, beagle dog

Received 26 September 2011; Revised version received 22 October 2011; Accepted 26 October 2011

Constipation is prevalent in modern societies and is a common gastrointestinal symptom in clinical practice, affecting 5 to 20% of the general population [1]. Constipation is typically defined as stool frequency of less than 3 bowel movements per week [2]. Constipation is thought to be partially caused by psychosocial and behavioral factors, such as decreased mobility, inadequate caloric intake, and anorectal sensation changes. Furthermore, it has a multifactorial etiology, including co-morbid illness and medication side effects, including those induced by narcotic analgesics, anticonvulsants, antidepressants, and anticancer drugs [3-5]. In addition, repeated use of a purgative medicine can cause chronic constipation, diarrhea, enteritis, and colorectal dysfunction, and is a risk factor for colorectal neoplasm [6,7]. Although constipation is a symptom, rather than a disease, it may be the cause of severe secondary diseases resulting from enteral fermentation, inducing toxic gas, and therefore requires active prevention and proper treatment [8].

*Corresponding authors: Soo-Wan Chae, Department of Pharmacology, School of Medicine, Chonbuk National University, 567 Baekjaedaero, Jeonju, Jeonbuk 561-712, Korea
Tel: +82-63-250-2348; Fax: +82-63-250-2349; E-mail: soowan@jbnu.ac.kr
Okjin Kim, Center for Animal Resources Development, Wonkwang University, 460 Iksandaero, Iksan, Jeonbuk 570-749, Korea
Tel: +82-63-850-6668; Fax: +82-63-850-7308; E-mail: kimoj@wku.ac.kr

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

275
common functional food remedy for constipation is dietary fiber. There are various studies on the treatment of constipation using natural substances \[9,10\].

The common fig (\textit{Ficus carica} L) is a deciduous broadleaf shrub belonging to the Moraceae family and is widely known as one of the first edible fruits cultivated by humans in areas with subtropical climate \[11\]. The fig originates from Carica in Asia Minor and the primary fig producers now are America and the Mediterranean \[12\]. Figs are high in natural and simple sugars, minerals, water, and fiber. They contain substantial levels of potassium, calcium, magnesium, iron, copper, manganese, and sodium, while they are low in fat \[13,14\]. They are a good source of flavonoids and polyphenols as well as being rich in the phytosterols lanosterol and stigmasterol \[12\]. Several reports have shown that the leaf, stem, and woody tissue contain antioxidants and antibiotics \[15,16\].

The purpose of the present study was to evaluate the laxative effect of figs by measuring stool weight and colonic transit time (CTT) after fig paste administration for 3 weeks in beagles with constipation induced by a high protein diet and exercise restriction.

**Materials and Methods**

**Materials**

Figs were supplied by the Black Raspberry Research Institute (Gochang, Korea) and made into a paste at the Research Center for Industrial Development of BioFood Materials (Chonbuk University, Jeonju, Korea).

**Animals**

Fifteen healthy beagle dogs (4 years old; Marshall Beijing, Beijing, China) with a weight range of 10-13 kg were purchased from Orient Co (Seongnam, Korea). All animals were subjected to a physical examination and were quarantined. The animals were housed individually in clean cages (H100×W120×L120 cm) placed in a well-ventilated house with a controlled temperature (25°C), 12/12-h light/dark cycle, and illumination of 200 lux. All experiments complied with ethical standards and were approved by the Animal Ethics Committee at Wonkwang University (Iksan, Korea).

**Constipation model**

We created a canine model of constipation with 15 beagles who were individually housed in general cage (H100×W120×L120 cm). In order to induce a constipation condition, the dogs were restrained in a small cage (H38×W60×L50 cm) for 1 week and moved to indoor kennels.

The experiment was performed in the same 15 dogs and separated to three time sections which were control period, constipation period (constipation induction in the small cage), and +fig paste period (fig paste administration), for 3 weeks each. In control period, dogs were given feed containing 23% of proteins at the rate of 3% of weight and 100 mL/kg of water. Being changed to over 40% of high proteins (feline feed, Purina Korea, Seoul, Korea) in constipation and +fig paste periods, feed was cut down to 2% of weight to inhibit the movement of digestive duct. Water also reduced to 80 mL/kg. Dogs during constipation and +fig paste periods were individually housed in small cage to restrict movement. Dogs during +fig paste period were administered 12 g/kg of fig paste.

**Measurement of body weight, fecal weight, and feed and water intakes**

We measured body weights and stool weights each week during the experimental periods. Feed and water intakes were measured for each 24-h period.

**Blood biochemical analysis**

We analyzed serum chemical parameters for the assessment of fig paste’s safety in the experimental beagles. Venous blood was collected at the last day of each experimental period. The whole blood was centrifuged at 2,000 g for 20 min at 4°C to obtain serum. Blood lipids including total cholesterol (TC), high-density lipoproteins (HDL) and triglycerides (TG) were analyzed using commercial kits (Sigma Diagnostics, St. Louis, MO, USA). Serum aspartate aminotransferase (AST) and alanine aminotransferase (ALT) as hepatotoxicity parameters were measured using automated techniques at the Department of Diagnostic Analysis, Wonkwang University Hospital (Iksan, Korea).

**Measurement of segmental CTT**

Based on preliminary tests, in which normal CTT was less than 6 h, 1 capsule of Kolomark (20 Ring; Konsyl Pharmaceuticals, Easton, MD, USA) containing 20 radiopaque rings, was ingested at 09:00, and simple abdominal radiographs were taken in the ventro-dorsal position at 6-h intervals until the Kolomark was egested. The localization of markers on abdominal films relied on identifying bony structures as suggested by Arhan et al. \[17\]. Markers located to the right of the vertebral spinous processes above a line from the seventh lumbar vertebra to the right pelvic outlet were assigned to the right colon, markers to the left of the vertebral spinous processes and above an imaginary line from the seventh lumbar.
Effects of *Ficus carica* paste on constipation

**Table 1. Effects of fig paste in change of feed and water intakes (n=15)**

| Period         | Feed intake (g) | Water intake (mL) |
|----------------|-----------------|-------------------|
| Control        | 318±13          | 879±103           |
| Constipation   | 212±16          | 762±58            |
| +fig paste     | 235±12          | 894±43            |

Column values with different superscripts are significantly different (*P*<0.05).

**Figure 1. Effects of fig paste on change in body weight in each beagle experimental period (n=15).**

**Figure 2. Effects of fig paste on fecal weight in each experimental period (n=15). Bars with different letters from the control are significantly different (*P*<0.05).**

Serum concentrations of blood lipids and hepatotoxicity parameters such as TC, HDL, TG, AST, and ALT were not significantly different among the 3 experimental periods (Table 2). The results of all experimental period were within the normal range, and no abnormal symptoms were observed.

### Blood biochemistry

The movement of Kolomark rings was observed using time-dependant radiography (Figure 3). Kolomark rings moved from the right colon to the rectosigmoid colon over a period of time. The transit time of Kolomark rings in the gastrointestinal tract was shortest over constipation period and was shorter during the +fig paste period than during constipation period. There was no remarkable findings on the abdominal radiography. We counted Kolomark rings using radiography (Figure 4, *P*<0.05). The numbers of Kolomark rings in the right colon were higher during +fig paste period than during constipation period after 6 h, but the numbers of Kolomark rings in the right, left, rectosigmoid and total colons were fewer during +fig paste period than during constipation period after 18 h (Table 3, *P*<0.05). There were no Kolomark rings in the right colon during the control period after 12 h, and all rings had been

### Results

**Feed and water intakes and body weights**

Feed intake was 318±13, 212±16, and 235±12 g during the control, constipation, and +fig paste periods, respectively (Table 1). This showed a significant tendency to decrease feed intake between the constipation and +fig paste periods compared with the control period, although the difference in feed intake was not statistically significant between constipation and +fig paste periods. There were no changes in water intake or body weight among the 3 periods (Table 1 and Figure 1).
egested after 18 h. The CTT of +fig paste period was 18 h, which was shorter than 30 h during constipation period. The final Kolomark ring was egested after 48 h during constipation period (data not shown). The CTT in the right colon, left colon, rectosigmoid colon, and total colon during control period was significantly faster than during constipation and +fig paste periods. Whereas the CTT in the left colon and total colon during +fig paste period was significantly shorter than during constipation period, there were no statistically significant differences in the CTT in the right and rectosigmoid colons between constipation and +fig paste periods.

### Table 2. Effects of fig paste on blood biochemical parameters (n=15)

| Period         | TC (mg/dL)  | HDL (mg/dL) | TG (mg/dL) | AST (mg/dL) | ALT (mg/dL) |
|----------------|-------------|-------------|------------|-------------|-------------|
| Control        | 157.07±6.00 | 123.87±4.33 | 32.93±3.24 | 36.67±1.38  | 72.40±15.80 |
| Constipation   | 158.13±7.82 | 124.67±6.06 | 39.00±5.76 | 32.27±1.83  | 89.60±27.96 |
| +fig paste     | 164.60±7.63 | 128.60±5.89 | 53.07±5.45 | 36.07±3.81  | 82.27±25.48 |

TC: total cholesterol, HDL: high-density lipoproteins, TG: triglycerides, AST: aspartate aminotransferase, ALT: alanine aminotransaminase.

### Figure 3. Effects of fig paste on radiographic measurement of Kolomark transit in each experimental period (n=15). Rt.: right, Lt.: left, Rs.: rectosigmoid.

The Rome criteria define chronic constipation on the basis of the presence of 2 or more of a list of 5 symptoms related to the ability to defecate over a period of 3 months or more, such as a frequency of 2 or less bowel movements per week, stools of hard mass over 25%, uncomfortable feeling after evacuation, fecal weight below 35 g, and excessive abdominal press at defecation [18]. Chronic constipation can be caused by peripheral neuropathy (Chagas disease or Hirschsprung’s disease), obstructive disease (colon cancer, hernia, or intestinal stricture), endocrine disease (hypothyroidism, diabetes mellitus, or hypokalemia), drugs (morphine, anticholinergic compounds, or calcium) and idiopathic factors [19].

Recently, the use of herbal remedies as constipation treatments has become a common practice in many countries. This study clearly demonstrates that fig extract or paste has a laxative activity. Figs contain numerous components: vitamins, minerals, cellulose, and amino acids [20,21]. They are reported to have antioxidant effects, as well as beneficial effects on cardiovascular, respiratory, and inflammatory diseases [22,23].

Figs contain cellulose, and beneficial effects of cellulose have been reported for cardiac disease, hypertension, diabetes, obesity, gastrointestinal disease, hyperlipidemia, and immune function [24-30]. Lee and Hwang [31] reported that cellulose increases fecal excretion by increasing water content and bulk, and elevating viscosity, and that both water-soluble and insoluble cellulose increases fecal egestion [32]. Fecal water content and volume are increased by eating fiber that does not decompose and is not digested by coliform bacilli [33,34]. Fig paste administration significantly decreased the feed intake both during constipation and +fig paste periods compared with control period. Our results show a tendency to increase during +fig paste period compared with constipation period, but this difference was not statistically significant. There were no changes in water intake and body weight (Table 1 and Figure 1). All serum chemistry results were also in the normal range following fig administration, indicating that fig paste
is safe to use (Table 2). The weight of stools significantly decreased during constipation period compared with control period, which was increased by fig paste administration, although the water content of stools was not increased by fig paste administration, although the water content of stools was not increased by fig paste administration.

CTT is the basic test used to assess movement disorder in the diagnosis of chronic constipation and irritable bowel syndrome [35]. It involves the use of radiography to count the number of markers remaining after a certain time period following administration of a gelatin capsule that contains 20 radiopaque markers; segmental CTT is also reported by Arhan et al. [17] and Metcalf et al. [36].

In the present study, considering that CTT in dogs is faster than in humans, we took abdominal radiographs every 6 h until there was no remaining Kolomark in the body (Figure 3) and measured CTT by counting the number of Kolomark rings in the intestinal tract (Table 3 and Figure 4). CTT was shortest during the control period, of all 3 periods. CTT in the left colon was faster during +fig paste period than during constipation period, whereas in the right colon and rectosigmoid colon, there was no difference between constipation period and +fig paste period. In our study, we confirmed that total CTT was fastest during control period followed by +fig paste period, and this result indicates that CTT was decreased by fig administration. Soluble cellulose accelerates peristaltic activity by maintaining higher acidity, because fermented cellulose results in acid production by coliform bacilli in rats [37]. Furthermore, Gordon [38] reported that cellulose shortens large intestine transit time, so that fecal excretion time decreases accordingly. Similar to our results, Spiller et al. [39] reported that intestinal transit time was diminished following administration of food-mixed cellulose and 5% guar gum, compared with rats given normal food [39]. Moreover, Jenkins et al. [40] reported that cellulose increases fecal egestion and reduces intestinal transit time, as well as reducing fecal sojourn time within intestines [40-42]. The shortened CTT indicates that fig paste stimulates intestinal peristalsis and accelerates fecal movement. Recently, intestinal ileum peristalsis was reported to be enhanced in rats by treatment with a low concentration of fig extract, but reduced by a high concentration [43]. Similarly, Baldassano et al. [44] reported that intestinal ileum peristalsis declined in a dose-dependent manner in the isolated mouse intestinal ileum after treatment with 10-320 mg/mL fig extract. Although these results differ from ours, this might be due to different experimental conditions with regard to animal species, type of figs used,

### Table 3. Effects of fig paste on radiographic measurement of Kolomark transit for each experimental period (n=15)

| Time  | Treatment | Right colon | Left colon | Rectosigmoid colon | Total colon |
|-------|-----------|-------------|------------|--------------------|-------------|
| 6 h   | Control   | 2.00±1.03   | 5.50±1.80  | 3.20±1.55          | 10.70±2.32  |
|       | Constipation | 8.53±1.86  | 10.93±1.82 | 0.40±0.40          | 19.87±0.50  |
|       | +fig paste | 14.80±1.83  | 4.07±1.60  | 0.13±0.09          | 19.00±1.00  |
| 12 h  | Control   | 0.00±0.00   | 2.50±1.18  | 5.90±1.59          | 8.40±1.70   |
|       | Constipation | 3.87±1.78  | 14.00±1.74 | 1.67±0.87          | 19.53±0.35  |
|       | +fig paste | 2.27±1.18   | 10.60±2.07 | 4.47±1.78          | 17.33±0.67  |
| 18 h  | Control   | 0.00±0.00   | 0.00±0.00  | 0.00±0.00          | 0.00±0.00   |
|       | Constipation | 0.67±0.37  | 15.33±1.60 | 3.40±1.56          | 19.40±0.48  |
|       | +fig paste | 0.00±0.00   | 6.67±2.09  | 7.40±1.93          | 14.07±2.12  |
| 24 h  | Control   | 0.00±0.00   | 0.00±0.00  | 0.00±0.00          | 0.00±0.00   |
|       | Constipation | 0.53±0.32  | 11.40±2.25 | 4.47±1.88          | 16.40±1.95  |
|       | +fig paste | 0.00±0.00   | 2.47±1.48  | 9.40±2.36          | 11.87±2.46  |
| 30 h  | Control   | 0.00±0.00   | 0.00±0.00  | 0.00±0.00          | 0.00±0.00   |
|       | Constipation | 0.00±0.00  | 5.40±2.1   | 8.53±2.29          | 13.93±2.32  |
|       | +fig paste | 0.00±0.00   | 0.20±0.20  | 2.60±1.68          | 2.80±1.81   |

Data are numbers of markers present. Column values with different superscripts are significantly different (P<0.05).

**Figure 4.** Effects of fig paste on change in segmental CTT for each experimental period (n=15). Bars with different letters from the control are significantly different (P<0.05). Rt.: right, Lt.: left, Rs.: rectosigmoid.
and dosage.

In conclusion, feeding fig paste increased fecal weight, which had been decreased by diet-induced constipation, and shortened CTT. The results of serum chemistry showed that oral administration of fig paste is safe, and we did not observe any abnormal symptoms in experimental animals. Therefore, fig paste may be suitable for human patients suffering from constipation, particularly where it is due to diet. In addition, our findings on the use of beagles as a constipation model could be useful in other constipation studies.

Acknowledgments

This work was supported by the Yeong-Am Figs of Food Cluster project of the Ministry for Food, Agriculture, Forestry and Fisheries, Republic of Korea (Grant: 2008-001).

References

1. Drossman DA, Richter JE, Talley NS. The Functional Gastrointestinal Disorders, 1st ed, Brown and Company, Boston, 1994; pp. 217-263.
2. Read NW, Timms JM, Barfield LJ, Donnelly TC, Bannister JJ. Impairment of defecation in young women with severe constipation. Gastroenterology 1986; 90(1): 53-60.
3. Elliot DL, Watts WJ, Girard DE. Constipation. Mechanisms and management of a common clinical problem. Postgrad Med 1983; 74(2): 143-149.
4. Enck RE. Constipation: etiologies and management. Am J Gastroenterol 1998; 93(4): 861-964.
5. Arhan P, Devroede G, Jehannin B, Lanza M, Faverdin C, Dormic C, Persoz B, Tétreault L, Perey B, Pellerin D. Segmental colonic transit time. Dis Colon Rectum 1981; 24(8): 625-629.
6. Drossman DA, Thompson WG, Talley NJ, Funch-Jensen P, Jassens J, Whitehead WE. Identification of functional gastrointestinal disorders. Gastroenterol Int 1990; 3: 159-172.
7. Faubrich WS, Schaffner F, Berk JE. Bockus Gastroenterology, 5th ed, W.B. Saunders, Philadelphia, 1995; pp 102-112.
8. Solomom A, Golubowicz S, Yablówicz Z, Grossman S, Bergman M, Gottlieb HE, Altman A, Korem Z, Flisheam MA. Antioxidant activities and anthocyanin content of fresh fruits of common fig (Ficus carica L.). J Agric Food Chem 2006; 54(20): 7717-7723.
9. Vebere R, Colaric M, Stampar F. Phenolic acids and flavonoids of fig fruit Ficus carica in the northern Mediterranean region. Food Chem 2007; 106(1): 153-157.
10. Guarerra PM. Traditional phytotherapy in Central Italy (Marche, Abruzzo, and Latium). Fitoterapia 2005; 76(1): 1-25.
11. Jeong WS, Lachance PA. Phytotherapies and fatty acids in fig (Ficus carica, var. Mission) fruit and tree components. J Food Sci 2001; 66(2): 278-281.
12. Brown L, Rosner B, Willett WW, Sacks FM. Cholesterol-lowering effects of dietary fiber: a meta-analysis. Am J Clin Nutr 1999; 69(1): 30-42.
13. Montonen J, Knekt P, Jarvinen R, Aromaa A, Reunanen A. Whole-grain and fiber intake and the incidence of type 2 diabetes. Am J Clin Nutr 2003; 77(2): 622-629.
14. Laiton D, Arnaud N, Bertras S, Planells R, Clerc E, Herberg S, Boutron-Ruault MC. Dietary fiber intake and risk factors for cardiovascular disease in French adults. Am J Clin Nutr 2005; 82(6): 1185-1194.
15. Liu S, Stampfer MJ, Hu FB, Giovannucci E, Rimm E, Manson JE, Hennekens CH, Willett WC. Whole-grain consumption and risk of coronary heart disease: results from the Nurses' Health Study. Am J Clin Nutr 1999; 70(3): 412-419.
16. Petruzziello L, Iacopini F, Bulajic M, Shah S, Costamagna G. Review article: uncomplcated diverticulitis of the colon. Aliment Pharmacol Ther 2006; 23(10): 1379-1391.
17. Watzl B, Girmbach S, Roller M, Inulin, oligofructose and immunomodulation. Br J Nutr 2005; 93(Suppl 1): S49-S55.
18. Whelon SP, Hyre AD, Pedersen B, Yi Y, Whelton PK, He J. Effect of dietary fiber intake on blood pressure: a meta-analysis of randomized, controlled clinical trials. J Hypertens 2005; 23(3): 475-481.
19. Lee HJ, Hwang EH. Effects of alginic acid, cellulose and pectin level on bowel function in rats. J Nutr 1997; 30(5): 185-189.
20. Kue K, Uebenhoer A, Press AG. Influence of senna, fibre, and fibre+senna on colonic transit in loperamide-induced constipation. Pharmacology 1993; 47(Suppl 1): 242-248.
21. Wintola OA, Summono TO, Afolayan AJ. The effect of Aloe vera leaf juice in the treatment of loperamide-induced constipation in Wistar rats. BMC Gastroenterol 2010; 10: 95.
22. Guarrera PM. Traditional phytotherapy in Central Italy (Marche, Abruzzo, and Latium). Fitoterapia 2005; 76(1): 1-25.
23. Jeong WS, Lachance PA. Phytotherapies and fatty acids in fig (Ficus carica, var. Mission) fruit and tree components. J Food Sci 2001; 66(2): 278-281.
24. Brown L, Rosner B, Willett WW, Sacks FM. Cholesterol-lowering effects of dietary fiber: a meta-analysis. Am J Clin Nutr 1999; 69(1): 30-42.
25. Montonen J, Knekt P, Jarvinen R, Aromaa A, Reunanen A. Whole-grain and fiber intake and the incidence of type 2 diabetes. Am J Clin Nutr 2003; 77(2): 622-629.
26. Laiton D, Arnaud N, Bertras S, Planells R, Clerc E, Herberg S, Boutron-Ruault MC. Dietary fiber intake and risk factors for cardiovascular disease in French adults. Am J Clin Nutr 2005; 82(6): 1185-1194.
27. Liu S, Stampfer MJ, Hu FB, Giovannucci E, Rimm E, Manson JE, Hennekens CH, Willett WC. Whole-grain consumption and risk of coronary heart disease: results from the Nurses' Health Study. Am J Clin Nutr 1999; 70(3): 412-419.
28. Petruzziello L, Iacopini F, Bulajic M, Shah S, Costamagna G. Review article: uncomplicated diverticulitis of the colon. Aliment Pharmacol Ther 2006; 23(10): 1379-1391.
29. Watzl B, Girmbach S, Roller M, Inulin, oligofructose and immunomodulation. Br J Nutr 2005; 93(Suppl 1): S49-S55.
30. Whelton SP, Hyre AD, Pedersen B, Yi Y, Whelton PK, He J. Effect of dietary fiber intake on blood pressure: a meta-analysis of randomized, controlled clinical trials. J Hypertens 2005; 23(3): 475-481.
31. Lee HJ, Hwang EH. Effects of alginic acid, cellulose and pectin level on bowel function in rats. J Nutr 1997; 30(5): 185-189.
32. Lupton JR, Morin JL, Robinson MC. Barley bran flour accelerates gastrointestinal transit time. J Am Diet Assoc 1993; 93(8): 881-885.
33. Gordon DT. The importance of total dietary cellulose in human nutrition and health. J Nutr 1992; 25(1): 75-76.
34. Nyman M, Schweizer TF, Tyrén S, Reimann S, Asp NG. Effect of dietary fiber intake on blood pressure: a meta-analysis of randomized, controlled clinical trials. J Hypertens 2005; 23(3): 475-481.
35. Lee HJ, Hwang EH. Effects of alginic acid, cellulose and pectin level on bowel function in rats. J Nutr 1997; 30(5): 465-477.
36. Lupton JR, Morin JL, Robinson MC. Barley bran flour accelerates gastrointestinal transit time. J Am Diet Assoc 1993; 93(8): 881-885.
37. Gordon DT. The importance of total dietary cellulose in human nutrition and health. J Nutr 1992; 25(1): 75-76.
38. Nyman M, Schweizer TF, Tyrén S, Reimann S, Asp NG. Fermentation of vegetable fiber in the intestinal tract of rats and effects on fecal bulking and bile acid excretion. J Nutr 1990; 120(5): 459-466.
39. Wald A. Colonic transit and anorectal manometry in chronic idiopathic constipation. Arch Intern Med 1990; 251(5): 1713-1716.
40. Metcalf AM, Phillips SF, Zinsmeister AR, MacCarty RL, Beart RW, Wolf BG. Simplified assessment of segmental colonic transit. Gastroenterology 1987; 93(8): 881-885.
41. Lee HJ, Hwang EH. Effects of alginic acid, cellulose and pectin level on bowel function in rats. J Nutr 1997; 30(5): 465-477.
39. Spiller GA, Chernoff MC, Hill RA, Gates JE, Nassar JJ, Shipley EA. Effect of purified cellulose, pectin, and a low-residue diet on fecal volatile fatty acids, transit time, and fecal weight in humans. Am J Clin Nutr 1980; 33(4): 754-759.

40. Jenkins DJ, Reynolds D, Leeds AR, Waller AL, Cummings JH. Hypcholesterolemic action of dietary fiber unrelated to fecal bulking effect. Am J Clin Nutr 1979; 32(12): 2430-2435.

41. Park EY, Lee SS. Effect of dietary fiber on the serum lipid level and bowel function in aged rat. Kor J Nutr 1996; 29(9): 934-942.

42. Vahouny GV, Kritchesky D. Dietary Fiber in Health and Disease. Plenum Press, New York 1982; 263-415.

43. Amos S, Binda L, Chindo B, Aka B, Abdurahman M, Danamallam HU, Wambebe C, Camanlal K. Evaluation of methanolic extract of Ficus platyphylla on gastrointestinal activity. Indian J Exp Biol 2001; 39(1): 63-67.

44. Baldassano S, Tesoriere L, Rotondo A, Seio R, Livrea MA, Mule F. Inhibition of the mechanical activity of mouse ileum by cactus pear (Opuntia ficus indica, L, Mill.) fruit extract and its pigment indicaxanthin. J Agric Food Chem 2010; 58(13): 7565-7571.