Morpho-physiological acclimation to canopy coverage of *Araucaria angustifolia* during the establishment in the Atlantic Forest, Argentina

Aclimatación morfo-fisiológica durante el establecimiento de *Araucaria angustifolia* bajo cobertura de canopeo en el Bosque Atlántico, Argentina

Flavia Y Olguin a,b, Ana P Moretti a,b, Martín A Pinazo c, Corina Graciano a,b,d *

a INFIVE (CONICET- Universidad Nacional de La Plata), La Plata, Argentina.
b CCT CONICET La Plata, La Plata, Argentina.
c INTA EEA Montecarlo, Montecarlo, Misiones, Argentina.
d Corresponding author:

SUMMARY

*Araucaria angustifolia* is an ecologically and commercially important species of the Atlantic Forest. However, its ability to regenerate continuously under the shade of the canopy in the rainforest is uncertain. The aims were: to evaluate the morpho-physiological responses of araucaria to different intensities of canopy coverage to know its shade acclimation capacity; and to evaluate survival and growth under different shade intensities in gaps. First, seedlings were grown in pots at full sun or under canopy shade and morpho-physiological traits were measured after 6 and 12 months. Survival was 100% under full sun and 40% under shade. One year after planting, the ability to use high-intensity light was similar in plants under both conditions, although shade plants did not grow and showed little capacity to acclimate to shade. Plants under full sun duplicated their size in six months and had higher capacity to deliver water to the leaves. However, shade plants had higher specific hydraulic conductance. After that, in a field experiment, seedlings were planted in six forest-gaps to record survival and growth for 30 months. Survival was high in all positions within the gaps. In the center of the gaps, where incident radiation was higher, growth was higher. We conclude that araucaria has low ability to acclimate to low radiation, nevertheless, it can survive at least 30 months under the canopy. Consequently, araucaria should not be planted below dense canopy shade. The microenvironment in the center of 195-293 m² gaps is proper for araucaria growth.

Key words: abiotic stress, ecophysiology, Paraná pine, rainforest gaps, sun and shade tolerance.

INTRODUCTION

Tropical and subtropical forests have heterogeneous light environment. The forest canopy decreases the quantity and changes the quality of light that arrives to the understory and severely limits growth and survival of seedlings of shade-intolerant species (Niinemets 2010). The removal of trees of the upper stratum, either by anthropic interven-
tion or by natural causes, generates openings in the canopy (gaps) that increase the sunlight that arrive to the understory. However, more subtle changes in the magnitude and spatial distribution of light under closed canopies can also influence long-term survival and recruitment of seedlings (Zhang et al. 2012). Plants can adjust genotype expression in response to those changes in light availability, in order to change phenotype to tolerate or optimize light interception and use (Vieira et al. 2015). Thus, the survival and growth of plants in changing environments highly depend on the ability to acclimate to different micro-environmental conditions (Gianoli and Valladares, 2012) to catch the resources that limit growth.

_Araucaria angustifolia_ (Bertol.) Kuntze (araucaria) is a conifer that represents the predominant species of Araucaria Moist Forests in the Atlantic Forest Biome. The Atlantic Forest is a subtropical forest that extends from the Southeast of Brazil to the Northeast of Argentina. Araucaria Forests occupied 177,600 km² in Brazil and 2,100 km² in Argentina (Arnoni Costa et al. 2017). However, during the last two centuries, its distribution has shrunk, due to the extraction driven by the high quality of araucaria wood and expansion of agricultural areas. Despite its ecological and commercial importance, the capacity of araucaria to regenerate and develop continuously in the native forest is uncertain (Longhi et al. 2018). The study of the ecology and management of tropical and subtropical forests is based on the classification of the species according to the response to relevant disturbances and the need of certain levels of water or light (i.e. pioneer, heliophyte, shade tolerant). Araucaria is considered a long-lived pioneer species that depend on large openings of the canopy to have a successful regeneration (Souza et al. 2014, Longhi et al. 2018). This idea is consistent with the success of araucaria plantations in deforested areas. However, mature araucaria forests have distribution of diameter compatible with species that regenerate under the canopy (Paludo et al. 2011). Consistently, it has been reported that the low availability of light does not limit the natural regeneration of araucaria in the forest and that the species can tolerate and maintain different rates of growth under shade (Duarte and Dillenburg 2000, Duarte et al. 2002, Franco and Dillenburg 2007). Those reports indicate that araucaria can regenerate under native forest canopy. However, native araucaria forests have high mortality and low natural regeneration rates in the understory (Beckert et al. 2014). These results show that it is necessary to deepen the research on possible responses to canopy coverage to determine the causes of the low natural regeneration in the rainforest and to identify the best sites for planting araucaria in native forest restoration programs.

The aims are: 1) to evaluate the morpho-physiological responses of araucaria to different intensities of canopy coverage to know its shade acclimation capacity; 2) to evaluate survivor and growth under different shade intensities in gaps.

The hypothesis is that araucaria has low capacity to acclimate to shade under canopy and therefore, its survival and growth will be lower than in open areas. The originality of this work is that the capacity of araucaria to acclimate to shade is described in an integrative way and related to the causes of the death of araucaria regeneration in the rainforest.

**METHODS**

**Plant material, growth conditions and coverage treatments.** A pot experiment and a field experiment were performed in Campo Anexo Manuel Belgrano (CAMB) belonging to EEA INTA Montecarlo, placed in San Antonio, Misiones, Argentina (26°02’56.91” S; 53°46’14.15” W, 552 m a.s.l.). The mean annual temperature is 21 °C, with a mean monthly amplitude of 10 °C; mean annual rainfall is 2000 mm, distributed evenly throughout the year. Araucaria seeds were collected from the forest and germinated in trays under a semi-covered greenhouse with shade cloths (40 % of irradiance reduction) for both experiments. In the pot experiment, 15-month-old plants were transplanted into 7-L pots filled with 1:1 mixture of soil from the forest and pine bark compost. Twenty-five plants were randomly placed under canopy shade in a remnant native forest and fifteen plants were randomly placed under full sun (n = 40). Photon flux density of photosynthetic active radiation (PAR) and the red/ far-red radiation ratio were measured with a Red Tide USB 650 spectroradiometer (Ocean Optics, Dunedin, Florida, USA), at 670 nm and 730 nm respectively. PAR was significantly higher under the full sun (~1800 µmol photons m⁻² s⁻¹) than under the canopy (~300 µmol photons m⁻² s⁻¹). Red/ far red ratio was near 1 under the full sun and 0.5 under the canopy shade. In addition, air and soil temperatures and relative humidity were measured with thermo-hygrometers (TFA 30.5000.02, Wertheim, Germany), at midday in 15 plants per treatment to describe the environment. Air temperature was similar in both positions (around 28 °C), air relative humidity was higher under the canopy (39±1 %) contrasted with full sun (32±2 %). The mean ± standard error of the height and the collar diameter of the plants at the beginning of the experiment were 28±4 cm and 5±1 mm, respectively. Measurements and destructive sampling were performed in five plants per treatment after the summer (six months after planting) and after the winter (12 months after planting). In the field experiment, 4-month-old araucaria plants were planted in six forest gaps as a grid with a distance of 4 m x 4 m, 41 plants per gap (n= 246). Extended gap sizes varied between 195 to 293 m², considered medium gaps in tropical rain forests (Brokaw 1982, De Lima and De Moura 2008). Canopy openness and transmitted light were estimated with hemispherical photographs taken in the center of each gap (table 1). Pictures were taken before sunrise to avoid sunfleck distortions in the images. Photographs were taken with a 180° fish-eye Nikon FC-E9 lens with a 1m height system.
Table 1. Description of the gaps used to plant *A. angustifolia* in the field experiment.

| Gap | Area (m²) | Canopy openness (%) | Transmitted total light (%) |
|-----|-----------|---------------------|---------------------------|
| 1   | 293.0     | 20.9                | 24.3                      |
| 2   | 264.5     | 21.4                | 32.3                      |
| 3   | 194.7     | 27.9                | 42.1                      |
| 4   | 222.1     | 38.0                | 50.5                      |
| 5   | 214.9     | 25.1                | 46.3                      |
| 6   | 220.3     | 23.2                | 30.5                      |

Growth measurements. In both experiments, survival was registered by counting alive plants and height was measured with metal tape every six months. The pot experiment lasted one year, while the field experiment was measured for 30 months. Collar diameter was measured with digital caliper since the beginning in the pot experiment and since the first year after planting in the field experiment. In the pot experiment, the number of whorls with branches and the number of total branches per plant were recorded. Also, the total dry weight for taproots, fine roots, stems and leaves was measured after drying at 65 °C to constant weight. Root to shoot (stems + leaves) dry weight ratio was expressed in leaf area basis (µg cm⁻²) and leaf dry weight basis (mg g⁻¹), considering SLA of each plant.

Leaf traits and hydraulic structure (Pot experiment). Specific leaf area (SLA) was measured on five upper-latest expanded leaves per plant. Leaves were scanned and the area was expanded with the software Image Tool v. 1.28 CMEIAS Update. Leaf area of the whole plant (whole-plant LA) was calculated and the number of total branches per plant were recorded. Leaves were scanned and the area measured on five upper-latest expanded leaves. Leaf area ratio (LAR) was calculated as leaf area divided by total plant dry weight. Chlorophyll a, chlorophyll b, total chlorophyll and carotenoids were extracted during 48 h in dark from one intact leaf section of 6.25 mm² in 1-mL of N, N-dimethylformamide. Absorbance at 664, 647 and 480 nm wavelengths were read with a spectrophotometer (UV-160A, Shimadzu, Kyoto, Japan) (Wellburn, 1994) in one sample per plant. Chlorophyll a/b ratio was calculated and interpreted as acclimation changes of the proportion of the antenna complex regarding photosystem reaction centers. Chlorophyll concentration was expressed in leaf area basis (µg cm⁻²) and leaf dry weight basis (mg g⁻¹), considering SLA of each plant.

The response of photosynthesis to light was analyzed through the measurement of photosystem II electron transport rate (ETR) at different PAR levels using a chlorophyll fluorometer (FMS2, Hansatech, Norfolk, UK). Measurements were performed on the upper-latest expanded and not shaded leaves, under natural light conditions along the day. Potential functions (ETR = a PARᵇ) were selected to adjust data of each treatment and sampling date because that model had lower Bayesian information criterion (BIC) and Akaike Information Criterion (AIC) concerning linear, polynomial, exponential and logarithmic functions. Maximum quantum yield of photosystem II (Fv/Fm) was measured on the same plants with the saturating pulse method (FMS2, Hansatech, Norfolk, UK) after 30 minutes of dark acclimation to ensure the relax of non-photochemical dissipation mechanisms.

Figure 1. Photon flux density of photosynthetic active radiation (PAR) for each ring at noon along 30 months after planting: 6, 18 and 30 months are in autumn, while 12 and 24 months are in spring. Rings are concentric: ring 1 is the center of the gap, while ring 4 is the outer. Different letters indicate significant differences between means for each date (Kruskal Wallis test, *P* < 0.05).

Densidad de flujo de fotones de la radiación fotosintéticamente activa (PAR) para cada anillo al mediodía a lo largo de 30 meses después de la plantación: 6, 18 y 30 meses son en otoño, mientras 12 y 24 meses son en primavera. Los anillos son concéntricos: anillo 1 es el centro del gap, mientras que el anillo 4 es el borde. Letras diferentes indican diferencias significativas entre medias para cada fecha (Kruskal Wallis, *P* < 0.05).
To know the capacity of araucaria seedlings to conduct water to the photosynthetic tissues under different canopy coverage, the hydraulic conductivity of the main stem (kh$_{stem}$), the hydraulic conductance of the root system (K$_{root}$) and the shoot (stem + leaves) (K$_{shoot}$) were measured with the pressure-drop hydraulic flow meter (Melcher et al. 2012). The perfusion was done with de-gassed filtrated distilled water, using 4.5 kPa pressure. The main stem was cut at collar. Afterwards, another cut was performed 1-cm above the collar under water and immediately connected to the measuring system. Hydraulic conductance was measured when the water flow was stable. Subsequently, the portion with leaves and branches were cut and the hydraulic conductivity of the stem basal portion (10 cm long) was measured. To measure the hydraulic conductance of the root, the final part of the main root was cut to perfuse water from the tap-root to the collar of the plant. The root was always kept under water during the measurement. Hydraulic conductance of the whole plant (K$_{plant}$) was calculated as equation 1.

$$K_{plant} = 1/(1/K_{root} + 1/K_{shoot}) \quad [1]$$

The standardized conductance of the whole plant was obtained as K$_{plant}$ divided by dry weight of the whole plant (K$_{plant}$: total DW). Shoot conductance was expressed both divided by its dry weight (K$_{shoot}$: shoot DW) and on leaf area basis (K$_{shoot}$: LA). The conductivity of the root was divided by its dry weight (K$_{root}$: root DW). The specific hydraulic conductivity of the stem (ks$_{stem}$) and the leaf specific hydraulic conductivity of the stem (kl$_{stem}$) were calculated as the ratio between kh$_{stem}$ and the cross-section of the sapwood and leaf area, respectively.

Statistical analyses. PAR in the field experiment had no normal distribution; therefore, non-parametric analysis with Kruskal Wallis test, $P < 0.05$ was done, to compare means in each date.

Morphological and physiological modifications analyzed here will be interpreted as responses to changes in all the environmental variables that are modified when plants accclimate to full sun compared to plants acclimated to canopy shade. In the pot experiment, ANOVA ($P < 0.05$) was performed with coverage (full sun and canopy shade) and growth time (6 and 12 months) as factors; the interaction between factors was analyzed. As interactions were significant, means were compared with the post hoc analyses DGC test ($P < 0.05$) (Di Rienzo et al. 2002). In the field experiment, height and collar diameter were analyzed with General Mixed Linear Models ($P < 0.05$), with withingap position (rings) and date as fixed factors and gap as random factor. Interaction was analyzed. As interactions were significant, means were compared with DGC test ($P < 0.05$). Survival at 30 months after planting was analyzed by nonparametric ANOVA, with the Kruskal Wallis test, $P < 0.05$. All analyses were done with software Infostat (Di Rienzo et al. 2017).

RESULTS

Pot experiment. Survival was 100% under full sun and 40% under canopy shade, six and 12-months after planting. At the end of the pot experiment, height, collar diameter, number of branches and number of whorls with branches were higher in plants growing under full sun than in those growing under the canopy shade (figure 2). Collar diameter, number of branches and number of whorls increased along time in plants under full sun while these traits did not increase in plants under canopy shade (figure 2). Total and stem dry weight increased over time only in plants under full sun (table 2). Shoot to root ratio was similar in plants under full sun and under canopy shade and steady over time (table 2). Root dry weight was similar in plants under full sun and under canopy shade six months after planting. However, plants under full sun invested more dry weight in fine roots contrasted with plants under canopy shade (table 2). Leaf dry weight was higher in plants under full sun than in plants under canopy shade, six and 12 months after planting. The shoot dry weight (stems + leaves) doubled in 6 months in plants under full sun while it did not increase in plants under canopy shade (table 2).

Whole-plant leaf area (LA) was higher in plants under full sun than in plants under canopy shade in both sampling dates (table 2). The leaf area ratio (LAR) was higher in plants under canopy shade than in plants under full sun and it decreased in both treatments between six and 12 months after planting (table 2). The specific leaf area (SLA) was always higher in plants under canopy shade than in plants under full sun and SLA decreased over time in both treatments (table 2). Chlorophyll concentration per leaf area was higher in plants under full sun than in plants under canopy shade and it was similar over the time in both conditions (table 2). The chlorophyll concentration per leaf weight and chlorophyll a/b ratio were similar in both treatments and sampling dates (table 2). Carotenoid concentration in leaf area basis was higher in plants under full sun, while carotenoid concentration in leaf dry weight basis decreased in both treatments, 12 months and six months after planting.

The capacity to conduct water of whole plants (K$_{plant}$) as well as of organs (K$_{shoot}$, K$_{root}$, kh$_{stem}$) was similar in plants under full sun and under canopy shade 6 months after planting (table 3). K$_{shoot}$ and K$_{plant}$ decreased after planting in plants under full sun, but not in plants under canopy shade. Conversely, K$_{root}$: root DW increased one year after planting in plants under canopy shade, but not in plants under full sun (table 3). The conductance of the whole-plant standardized by total dry weight (K$_{plant}$: total DW) and the conductance of the shoot standardized by shoot dry weight (K$_{shoot}$: shoot DW) and by leaf area (K$_{shoot}$: LA) were higher in plants under canopy shade than in plants under full sun (table 3). Therefore, plants under full sun had higher conductance because they were bigger than plants under canopy shade. The capacity to driving water of the...
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Figure 2. Height (A), collar diameter (B), number of the total branches (C) and number of whorls with branches (D) during the first year in the pot experiment. As interaction between treatment x date was significant for all the variables, different letters show significant differences between means (DGC, \( P < 0.05 \)).

**DISCUSSION**

**Survival and growth.** In the pot experiment, survival was low below the canopy shade while in the field experiment survival was high in all the rings and mortality was no associated with within-gap position. Therefore, in the pots, plants could not sustain the cost of acclimation and some plants died, as radiation was very low to have positive carbon gain. This was evident because the survivors under the canopy shade made many morphological and physiological changes to acclimate to low radiation, in spite of their lower growth regarding plants under full sun. Lower growth was reflected in lower height, collar diameter and number of branches 30 months after planting, as both treatments started with the same plant size. Growth graphically can be seen as the slope of each variable along time: lower slope, lower growing rate and vice versa (figure 2). The results obtained in the pot experiment are related to the results in the field experiment, where the highest growth in height and collar diameter was recorded in plants in the positions of gaps with higher radiation (figures 1 and 4), which is straightforward related to coverage. Results suggest that the growth of araucaria under canopy shade during the first...
years is lower than in micro environments with higher level of radiation. Plants acclimated to low irradiances may have higher activity in the apical meristem, leading to reduced branches resulting in taller plants with thinner stems (Valladares and Niinemets 2008). Similar average heights, around 23 cm, were reported in young plants of the same species under controlled conditions with low radiation (200 μmoles m⁻² s⁻¹) (Einig et al. 1999). Contrary to our results, Duarte and Dillenburg (2000) reported higher araucaria height three months after planting at higher irradiances, however after that date, no differences between light conditions were observed. The contradiction may be due to the use of an artificial shade, whereas we used a native forest canopy shade. The proportion of red light (i.e. red: far-red ratio) or the intensity of blue light are essential in the morphogenetic response of plants to the environment and an artificial shade may not produce the same effect than a natural canopy. On the other hand, Franco and Dillenburg (2007) reported higher araucaria heights 3 months after planting at lower irradiance, where the growth in light-limiting conditions could have been favored by the mobilization of the storage compounds of the seeds (Einig et al. 1999). However, no increase in height was recorded after that period in shade plants, whereas plants at full sun increased in height and increased investment in branching. This last response coincides with the growing response described in our pot experiment (figure 2), possibly because the measurements were done six months after setting the coverage conditions. Although increase in height can be a helpful response to a short duration shade, conserving photoassimilates to sustain respiration may be more advantageous under continuous shading (Franco and Dillenburg, 2007).

Table 2. Dry weight partition and pigment concentration in the pot experiments in full sun and canopy shade treatments at six and 12 months; Shoot/Root: relationship between shoot dry weight and root dry weight; SLA: specific leaf area of plant; LA: leaf area of plant; LAR: leaf area ratio (LA/total dry weight); Chlorophyll a/b: relationship between chlorophyll a and chlorophyll b. Significant interaction or significant factors are indicate as: * = P < 0.05; ** = P < 0.01; *** = P < 0.001 and ns = no significant. If the interaction CxT is significant, different letters show significant differences between means (P < 0.05).

| Variable                        | 6 months | 12 months | P- value |
|---------------------------------|----------|-----------|----------|
|                                 | Full sun | Canopy shade | Full sun | Canopy shade | Coverage | Time | CxT |
| Total dry weight (g)            | 30 (b)   | 4 (c)     | 80 (a)   | 7 (c)       | ***      | ***  | **  |
| Taproot dry weight (g)          | 4 (b)    | 1 (b)     | 15 (a)   | 2 (b)       | ***      |     * |     |
| Fine root dry weight (g)        | 4 (b)    | 1 (c)     | 10 (a)   | 1 (c)       | ***      | **   | **  |
| Root dry weight (g)             | 8 (b)    | 1 (b)     | 25 (a)   | 2 (b)       | ***      | **   | **  |
| Leaf dry weight (g)             | 15 (b)   | 2 (c)     | 28 (a)   | 2 (c)       | ***      |     * |     |
| Stem dry weight (g)             | 8 (b)    | 1 (b)     | 27 (a)   | 2 (b)       | ***      | ***  | **  |
| Shoot dry weight (g)            | 23 (b)   | 3 (c)     | 55 (a)   | 4 (c)       | ***      | **   | **  |
| Shoot to Root ratio             | 2.98     | 2.67      | 2.30     | 2.35        | ns       | ns   | ns  |
| SLA (cm² g⁻¹)                   | 67       | 86        | 37       | 69          | **       | **   | ns  |
| Whole-plant LA (cm²)            | 989      | 187       | 1054     | 153         | ***      | ns   | ns  |
| LAR (cm² g⁻¹)                   | 33       | 43        | 13       | 24          | **       | ***  | ns  |
| Chlorophyll a/b                 | 2.06     | 2.31      | 2.01     | 2.17        | ns       | ns   | ns  |
| Total chlorophyll (µg cm⁻²)     | 105      | 60        | 115      | 70          | **       | ns   | ns  |
| Total chlorophyll (mg g⁻¹)      | 7        | 5         | 4        | 5           | ns       | ns   | ns  |
| Carotenoids (µg cm⁻²)           | 14       | 8         | 14       | 9           | **       | ns   | ns  |
| Carotenoids (mg g⁻¹)            | 0.90     | 0.65      | 0.51     | 0.61        | ns       | *    | ns  |

The maximum yield of photosystem 2 (Fv/Fm at dark) remained over 0.80 in both treatments and sampling dates (F = 1.68; P = 0.23), therefore no damage to the photosynthetic apparatus was registered. Electron transport rate (ETR) at high PAR was higher in plants under full sun than in plants under canopy shade six months after planting (figure 3), whereas the relationship between ETR and PAR was similar in both treatments 12 months after planting (figure 3).
Morphological acclimation to shade. One of the main mechanisms to adjust plant morphology to resource imbalances is assigning biomass to the organs that acquire the strongest limiting resource (Duarte and Dillenburg 2000). The accumulation of total dry weight was significantly lower in plants under canopy shade and there was no carbon gain between measurements (table 2). Shoot, leaf and fine roots dry weights were lower in plants under canopy shade than in plants under full sun (table 2). This response coincides with reports for the same species, with a shade that implied 90% reduction in sunlight (Duarte and Dillenburg 2000, Franco and Dillenburg 2007) and under full darkness (Dillenburg et al. 2010). A reduction of carbon allocation to the root system may be considered a strategy of shade-tolerant plants to prolong their survival (Valladares and Niinemets 2008), nonetheless in araucaria, under the shade, neither root nor shoot increased in six months. Therefore, there was no growth rather than acclimation. In another point of view, higher investment in roots in plants under high irradiances may be considered a functional response to higher evapotranspiratory demand, that may be related to higher soil temperature and lower relative humidity at full sun compared with the environment under canopy cover. However, in our experiment, shoot to root ratio was similar in plants under full sun and under canopy shade (table 2). Despite, in most of the species studied, an increase in the shoot to root ratio has been evident in response to low irradiances, even for araucaria (Duarte and Dillenburg 2000, Franco and Dillenburg 2007). Therefore, in our experiment, plants under full sun grew markedly and they did not change shoot to root ratio that seems optimal to tolerate full sun conditions; plants under the canopy shade did not grow, hence they could not adjust dry weight partitioning to better tolerate low radiation.

Although there were no changes along the first year after planting in dry weight in plants under canopy shade, there were adjustments of SLA, which was higher in plants under canopy shade in both dates (table 2) as was reported for araucaria (Duarte and Dillenburg 2000, Franco and Dillenburg 2007). It is expected that leaves developed in environments with higher radiation have additional layers of mesophyll, increasing the thickness of the leaf and decreasing its SLA. In contrast, plants developed in environments with lower radiation tend to make morphological modifications to maximize the interception of light with a low carbon investment (Niinemets 2010). Shade plants and shade-tolerant plants allocate more resources to the production of leaves, resulting in higher proportion of leaf area to total dry weight (LAR). For example, in six species

| Table 3. Conductance (K) and hydraulic conductivity (k) 6 and 12 months after planting, for full sun and canopy shade treatments in the pot experiment. K<sub>plant</sub>: conductance of whole plant; K<sub>root</sub>: conductance of root; K<sub>shoot</sub>: conductance of shoot (leaves+stem); kh<sub>stem</sub>: hydraulic conductivity of stem; ks<sub>stem</sub>: specific hydraulic conductivity of stem (kh<sub>stem</sub>xylem area); kl<sub>stem</sub>: specific leaf conductivity of stem (kh<sub>stem</sub>leaf area); K<sub>shoot</sub>LA: stem conductance standardized by the leaf area of the plant. LA means leaf area, DW means dry weight. Significant interaction or significant factors are indicated as: * = P < 0.05; ** = P < 0.01; *** = P < 0.001 and ns = no significant. If the interaction CxT is significant, different letters show significant differences between means (P < 0.05). |
|---|---|---|---|---|---|
| 6 months | 12 months | P-value |
| Full sun | Canopy shade | Full sun | Canopy shade | Coverage | Time | CxT |
| K<sub>plant</sub> (g MPa<sup>-1</sup> h<sup>-1</sup>) | 0.59<sup>(b)</sup> | 0.27<sup>(b)</sup> | 5.84<sup>(a)</sup> | 1.01<sup>(b)</sup> | * | * | * |
| K<sub>root</sub> (g MPa<sup>-1</sup> h<sup>-1</sup>) | 33<sup>(b)</sup> | 22<sup>(b)</sup> | 84<sup>(b)</sup> | 200<sup>(a)</sup> | *** | ** | ** |
| K<sub>shoot</sub> (g MPa<sup>-1</sup> h<sup>-1</sup>) | 0.62<sup>(b)</sup> | 0.24<sup>(b)</sup> | 6.69<sup>(a)</sup> | 1.08<sup>(b)</sup> | * | * | * |
| kh<sub>stem</sub> (g m MPa<sup>-1</sup> h<sup>-1</sup>) | 5 | 4 | 40 | 21 | Ns | ** | ns |
| ks<sub>stem</sub> (g m MPa<sup>-1</sup> h<sup>-1</sup> m<sup>-2</sup>)x10<sup>4</sup> | 23 | 78 | 58 | 274 | Ns | ns | ns |
| kl<sub>stem</sub> (g m MPa<sup>-1</sup> h<sup>-1</sup> m<sup>-2</sup>) | 56 | 236 | 408 | 1436 | Ns | * | ns |
| K<sub>plant</sub> total DW (g MPa<sup>-1</sup> h<sup>-1</sup> g<sup>-1</sup>) | 0.02 | 0.07 | 0.07 | 0.17 | ** | ** | ns |
| K<sub>root</sub> root DW (g MPa<sup>-1</sup> h<sup>-1</sup> g<sup>-1</sup>) | 7<sup>(b)</sup> | 21<sup>(b)</sup> | 4<sup>(b)</sup> | 107<sup>(a)</sup> | ** | * | * |
| K<sub>shoot</sub> shoot DW (g MPa<sup>-1</sup> h<sup>-1</sup> g<sup>-1</sup>) | 0.03 | 0.08 | 0.12 | 0.26 | ** | *** | ns |
| K<sub>shoot</sub> LA (g MPa<sup>-1</sup> h<sup>-1</sup> m<sup>2</sup>) | 6 | 14 | 62 | 95 | Ns | *** | ns |
LAR increased as light intensity decreased (Moraes Neto et al. 2000). Likewise, shade-tolerant temperate gymnosperms had higher LAR than intolerant gymnosperms (Valladares and Niinemets 2008). Consistently LAR was higher in araucaria plants under canopy shade than in plants under full sun; nevertheless, it is important to highlight that there was no increment in leaf area neither in shade plants nor in sun plants. However, unlike shade plants, sun plants increased total dry weight. That means that the higher LAR in araucaria under the canopy shade, contrasted with sun plants, is expressing that dry weight is more reduced than leaf area: i.e. araucaria under the canopy cannot fix carbon, although leaf area persists for six months. After that, LAR decreased over time (table 2). In the case of sun plants this change can be related to a phenological stage because bigger trees have less proportion of leaves compared with stems and roots. In the case of shade pants, this happened because leaf area started to reduce. Therefore, araucaria under canopy shade can increase SLA, although it cannot actively change LAR or shoot to root ratio, because no growth occurs.

**Physiological acclimation to canopy shade.** Concentration of chlorophyll per gram of leaves was higher in araucaria plants growing under the shade for two months, while chlorophyll a/b ratio was similar in plants under shade and full sun (Franco and Dillenburg 2007), as observed in many species. Contrary to expectations, the concentration of chlorophyll per unit area was lower in plants under canopy shade than in plants under full sun, while chlorophyll concentration in dry weight basis and chlorophyll a to b ratio were similar between treatments (table 2). The low concentration of chlorophyll we observed is possibly indicating that araucaria did not achieve a positive carbon balance under high coverage, as reflected in the null growth (figure 2). Similarly, in another study, there were no differences in chlorophyll concentration per leaf area or dry weight, or in chlorophyll a/b ratio in araucaria plants growing under different levels of irradiance during 3-months (Duarte et al. 2002). The difference in chlorophyll concentration may be given by the differences in the type of shade of each experiment since in our experiment the reduction of PAR was given by the canopy shade and in other experiments by polyethylene shade cloths. The concentration of photosynthetic pigments may vary in response to light levels and quality. Generally, the leaves acclimated to low irradiances have higher concentration of chlorophyll per unit area and leaf weight, in relation to leaves acclimated to high irradiances to optimize photophosphorylation and, consequently, energy production (Valladares and Niinemets, 2008). In most plants, the proportion of chlorophyll a to b tends to decrease as shade increases, because chlorophyll b is mainly present in the photosystem 2 antennae that increase their proportion under low irradiances. In light-demanding species, the concentration of chlorophyll increases only in response to a moderate shade (36 % incident light) while under denser shade (8 % incident light) the concentration does not change or even decreases, as in our experiment. However, in shade semitolerant species both moderate and severe shading lead to an increase in chlorophyll concentration (Strauss-Debenedetti and Bazzaz 1991). Therefore, chlorophyll concentration is another physiological trait that shows that araucaria cannot acclimate to the shade.

After six months of acclimation, sun plants were able to fix more carbon at high irradiances than did canopy plants (figure 3). This is consistent with the higher chlorophyll concentration and lower SLA, i.e. thicker leaves of sun plants compared with shade plants. However, after one year, plants acclimated to shade had the same response of...
physiological acclimation to open areas. A physiological trait that can test if plants under full sun are suffering stress by excess of light is Fv/Fm. This ratio can take values between 0 and 0.85 and connotes a direct relationship with the percentage of functional centers of photosystem 2 (PSII). In general, a decrease from 0.85 indicates damage and loss of function at photosynthetic apparatus level because the PSII is very susceptible to excess of light, thus the rate of damage is linearly dependent on the photon fluence rate. If plants are suffering by excess of light, Fv/Fm should decrease as well as the concentration of chlorophyll. None of these changes occurred in the plants exposed to full sun. The only protection to excess of light was an increment in carotenoids concentration in sun respect to shade plants (table 2). Therefore, according to our results, araucaria could be classified as a light-demanding species since the concentration of chlorophyll decreased under dense shade (12 % incident light) and there are no signals of stress by excess of light.

On the other hand, in open areas, the evaporative demand is higher than under the canopy due to the higher temperature and air movement. Consequently, the ability of the whole plant and different organs to carry water was higher in plants under full sun than in plants under the canopy shade. However, the capacity standardized by the size was higher in canopy shade plants (table 3). This indicates that plants under canopy shade can drive more water with less investment of dry weight. Possibly the highest specific conductance could be achieved with larger vessels and/or thinner walls (Hacke et al. 2017). Therefore, this higher efficiency of driving water can increase risk of hydraulic failure if the availability of water in the soil decreases or the saturation deficit of the atmosphere increases. Higher vulnerability to cavitation was reported for four conifers, with different shade tolerance, cultivated under canopy of deciduous forest compared with plants growing under full sun (Schoonmaker et al. 2010). Evapotranspiration and temperature are lower under the canopy than under full sun. However, a rapid change in environmental conditions or the occurrence of days with extreme heat and drought can expose plants to situations of high water deficit and produce generalized embolisms. In this situation, the death
of the plants can occur, as observed in shade plants in our pot experiment six months after planting. Although, a recent study has demonstrated that leaves of araucaria are capable of absorbing water from the atmosphere and that this water can be transported through the xylem to the soil close to the root, to improve plant water status (Cassana et al. 2016). This type of response demonstrates that the species has the ability to adjust its hydraulic architecture one year after planting, to ensure the adequate delivery of water to the leaves under high radiation and evaporative demands, without meaningful water stress.

Finally, we found no evidence that araucaria under full sun or in the open areas in the gaps suffer stress; nevertheless, we found that shade severely limits growth (table 2 and figure 4). Araucaria has nearly no capacity to acclimate to shade, as no active change in morphology or physiology was observed. However, it is very important to highlight that araucaria plants under the shade can survive at least for 30 months in medium gaps (figure 4).

CONCLUSIONS

Araucaria has low survival under very dense canopies with no dry mass gain. In forest medium gaps, with intermediate shade, the survival is higher than under the dense canopy and mortality is not related with the position of the plant in the gaps. However, growth rate is higher in positions with higher radiation. Consistently, araucaria acclimates properly to full sun and no water or excess of light stresses occur under this growing condition. We demonstrate that araucaria has low capacity to produce morphophysiological adjustments in response to the conditions provided by the forest canopy and these may explain its limited regeneration in the understory. Among the possible strategies to acclimate to shade observed in other species to enhance light use, only increase in specific leaf area is observed. The results suggest that the success of araucaria in restoration plans under canopy shade may not be guaranteed, because probably after 30 months, plants under the dense canopy would die by carbon starvation. However, as plant standardized hydraulic conductance is higher in shade plants than in sun plants hydraulics fails could compromise araucaria survival before carbon starvation, particularly if coverage is removed suddenly or under warmer climate. It would be advisable to carry out studies on the aptitude of the species to tolerate sudden variations in the canopy coverage and also over large gaps.

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