Anthocyanins inhibit high-glucose-induced cholesterol accumulation and inflammation by activating LXRα pathway in HK-2 cells

Chunyang Du1,2,*
Yonghong Shi1,2,*
Yunzhuo Ren1,2
Haijiang Wu1,2
Fang Yao1,2
Jinying Wei1,2
Ming Wu1,2
Yanjuan Hou1,2
Huijun Duan1,2

1Department of Pathology, Hebei Medical University, Shijiazhuang, People’s Republic of China; 2Hebei Key Laboratory of Kidney Diseases, Shijiazhuang, People’s Republic of China

*These authors contributed equally to this work

Abstract: The dysregulation of cholesterol metabolism and inflammation plays a significant role in the progression of diabetic nephropathy (DN). Anthocyanins are polyphenols widely distributed in food and exert various biological effects including antioxidative, anti-inflammatory, and antihyperlipidemic effects. However, it remains unclear whether anthocyanins are associated with DN, and the mechanisms involved in the reciprocal regulation of inflammation and cholesterol efflux are yet to be elucidated. In this study, we evaluated the regulation of cholesterol metabolism and the anti-inflammatory effects exerted by anthocyanins (cyanidin-3-O-β-glucoside chloride [C3G] or cyanidin chloride [Cy]) and investigated the underlying molecular mechanism of action using high-glucose (HG)-stimulated HK-2 cells. We found that anthocyanins enhanced cholesterol efflux and ABCA1 expression markedly in HK-2 cells. In addition, they increased peroxisome proliferator-activated receptor alpha (PPARα) and liver X receptor alpha (LXRα) expression and decreased the HG-induced expression of the proinflammatory cytokines intercellular adhesion molecule-1 (ICAM1), monocyte chemoattractant protein-1 (MCP1), and transforming growth factor-β1 (TGFβ1), as well as NFκB activation. Incubation with the PPARα-specific inhibitor GW6471 and LXRα shRNA attenuated the anthocyanin-mediated promotion of ABCA1 expression and cholesterol efflux, suggesting that anthocyanins activated PPARα-LXRα-ABCA1-dependent cholesterol efflux in HK-2 cells. Moreover, the knockout of LXRα abrogated the anti-inflammatory effect of anthocyanins, whereas the PPARα antagonist GW6471 does not have this effect. Further investigations revealed that LXRα might interfere with anthocyanin-induced decreased ICAM1, MCP1, and TGFβ1 expression by reducing the nuclear translocation of NFκB. Collectively, these findings suggest that blocking cholesterol deposition and inhibiting the LXRα pathway-induced inflammatory response might be one of the main mechanisms by which anthocyanins exert their protective effects in DN.

Keywords: 3-O-β-glucoside chloride, cyanidin chloride, diabetic nephropathy, inflammation, liver X receptor alpha

Introduction
Diabetes mellitus (DM), which is characterized by long-term hyperglycemia and various diabetes-related complications, is currently a major worldwide epidemic. Of all the important and serious complications, diabetic nephropathy (DN) has become the leading cause of end-stage renal disease, and contributes profoundly to patient morbidity and mortality.1 Although hyperglycemia and hypertension play key roles in the pathogenesis of DN, potential additional risk factors and the underlying mechanisms behind these effects remain unclear. Moreover, current therapies, including strict glycemic control, intensive insulin treatment, and reduction of hypertension, delay the progression of DN, but do not sufficiently prevent progression to end-stage renal disease.2 Therefore, clarifying novel pathogenic mechanisms and exploring additional effective therapeutic strategies is needed urgently.
Hyperglycemia is a primary risk factor for DN, and sustained hyperglycemia promotes lipid accumulation and inflammatory infiltration in the kidneys of patients with type 2 DM. Recently, the intrarenal accumulation of lipids and proinflammatory cytokines was observed in diabetic humans and experimental animal models. Lipotoxicity and inflammation might play an important role in the pathogenesis of DN. Accumulating evidence has suggested that monocyte chemoattractant protein-1 (MCP1), transforming growth factor-β1 (TGFβ1), and intercellular adhesion molecule-1 (ICAM1) are important proinflammatory cytokines involved in the development of DN. In addition, activated nuclear factor-κB (NFκB) translocates from the cytoplasm to the nucleus to trigger the expression of its target genes including MCP1, ICAM1, and TGFβ1 to induce persistent and enhanced inflammation, which leads to glomerulosclerosis and tubulointerstitial fibrosis.

Dysregulated cholesterol metabolism has also been linked to lipid accumulation and inflammation in diabetes. Ruan et al. showed that interleukin 1β (IL-1β) promotes intracellular lipid accumulation in mesangial cells by inhibiting cholesterol efflux. In addition, cholesterol accumulation and ABCA1 downregulation in podocytes plays a pathogenic role in DN. ABCA1, a member of the ATP-binding cassette transporter family, facilitates the efflux of cellular cholesterol to extracellular apoA1 or high-density lipoprotein (HDL). ABCA1 expression was reduced in the kidneys of diabetic NOD mice, which was accompanied by increased levels of renal cholesterol. Acute renal tubular injury can also trigger the downregulation of ABCA1 and cholesterol accumulation.

Numerous studies reveal that peroxisome proliferator-activated receptor alpha (PPARα) and liver X receptor alpha (LXRα), which are upstream of ABCA1, activate a transcriptional cascade to modulate the expression of ABCA1 and cholesterol efflux. High-glucose (HG) was sufficient to downregulate PPARα and LXRα expression, which was associated with severe dyslipidemia and inflammation in diabetic kidneys. The increase in PPARα expression reduces triglycerides, total cholesterol, and low-density lipoprotein (LDL) levels markedly, and elevates high-density lipoprotein (HDL) levels. PPARα deficiency accelerates dyslipidemia in animal models of DN. A study by Johnston and Waxman demonstrated that PPARα could increase the expression of LXRα in macrophages and thereby promote the expression of ABCA1, which subsequently mediates cholesterol efflux to apoA1. In addition to their important role in regulating lipid metabolism, PPARα and LXRα exhibit anti-inflammatory effects in many inflammatory and metabolic diseases. Recent studies found that the PPARα activator fenofibrate ameliorates inflammation by inhibiting NFκB activity and suppressing the production of the proinflammatory cytokines ICAM1, TGFβ1, and MCP1. The activation of LXRα also inhibits the production of the proinflammatory cytokines MCP1, ICAM1, and TGFβ1. Thus, PPARα and LXRα are physiological regulators that lie at the intersection of lipid metabolism and inflammation.

In recent years, the ingestion of naturally grown fresh vegetables, fruits, and crops was revealed to have preventive effect in diabetes and its complications. For example, anthocyanins are the largest group of water-soluble pigments, and belong to the family of phenolic flavonoid compounds that are responsible for the blue, red, purple, and pink colors of many plants in nature. Anthocyanins exhibit powerful antioxidative properties in diabetic animal models and also exert protective effects against hyperglycemia-induced kidney injury. In addition, many studies suggested that anthocyanins also have anti-inflammatory and anti-hyperlipidemic effects. Purple corn anthocyanins (PCA) prevent HG-induced glomerulosclerosis and renal fibrosis via NFκB-dependent mechanisms. Anthocyanin-rich foods and extracts increase ABCA1 levels significantly and suppress cholesterol accumulation in the liver and aorta. However, no studies have revealed the mechanisms involved in the reciprocal regulation of inflammation and cholesterol efflux by anthocyanins in DN.

In this study, we examined the effect of anthocyanins (cyanidin-3-O-β-glucoside chloride [C3G] and cyanidin chloride [Cy], an aglycon of C3G; Figure 1) on HG-mediated cholesterol accumulation and inflammation, as well as their possible molecular mechanism of action in HK-2 cells.

Materials and methods

All procedures were performed in compliance with the guidelines established by the Ethics Review Committee for Experimentation of Hebei Medical University.
Reagents
C3G (PubChem CID: 197081) and Cy (PubChem CID: 68247) were obtained from Sigma-Aldrich Co. (St Louis, MO, USA). GW6471, dimethyl sulfoxide (DMSO) and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazoliumbromide (MTT) were purchased from Sigma-Aldrich Co. The pLV-NR1H3-shRNA plasmid was constructed by Yingrun Biotechnologies (Changsha, People’s Republic of China; GenBank number NM_005693). FuGENE-HD transfection reagent was obtained from Promega (Madison, WI, USA). Antibodies against ICAM1, MCP1, PPARα and NFκB p65 were obtained from Abcam (Cambridge, UK). TGFβ1 antibody was purchased from Proteintech (Chicago, IL, USA). Antibodies against LXRα and ABCA1 were obtained from Sangon Biotech Co, Ltd. (Shanghai, People’s Republic of China). Histone H3 was purchased from Signalway Antibody (College Park, MD, USA). Human ICAM1 (H140114-19a), MCP1 (H140114-113a), and TGFβ1 (H140114-17a) enzyme-linked immunosorbent assay (ELISA) kits were purchased from Neobioscience (Shenzhen, People’s Republic of China). TRIzol reagent and a nuclear protein extraction kit were obtained from Invitrogen (Carlsbad, CA, USA). SYBR Premix Ex TaqTMII was from TaKaRa Bio Inc. (Shiga, Japan).

Cell culture
HK-2 cells were obtained from ATCC (American Type Culture Collection, Manassas, VA, USA). They were cultured in Dulbecco’s Modified Eagle’s Medium/Nutrient Mixture F-12 (DMEM-F12, 3:1) supplemented with 10% fetal bovine serum in a humidified 5% CO₂ atmosphere at 37°C. Cells were made quiescent by culturing in serum-free medium for 24 hours (h).

According to the different experiments, HK-2 cells were pretreated with or without C3G or Cy (50 μM) for 1 h, before exposure to HG (30 mM) for 24 or 48 h. Additional cells were stimulated with normal glucose (NG) (5.6 mM) as a normal control, and NG plus mannitol (24.4 mM) as an osmotic control. In the experiments intended to evaluate the specific involvement of PPARα in the activation of its target genes, cells were treated with 25 μM GW6471, a PPARα antagonist, 30 minutes (min) before the treatment with anthocyanins. All cell experiments were repeated at least three times.

Cell viability
MTT assays were used to determine the effects of C3G and Cy on cell viability. HK-2 cells in 96 well plates were exposed to HG (30 mM) alone for 24 h or preincubated with different concentrations of C3G and Cy (0–100 μM) for 1 h, followed by stimulation with or without HG (30 mM) for 24 h. Subsequently, 20 μL MTT (5 mg/mL) was added to each well, and the cells were further incubated for an additional 4 h. The supernatant was removed and the formazan crystals were dissolved with 150 μL/well of DMSO. The optical density was measured at 570 nm using a microplate reader (Synergy 2; BioTek, Winooski, VT, USA).

Flow cytometry
To assess the effect of anthocyanins on HG-induced intracellular reactive oxygen species (ROS) generation in HK-2 cells, flow cytometry with CM-DCHF-DA (CM-DCHF-DA is chloromethyl-2′,7′-dichlorodihydrofluorescein diacetate; Invitrogen) was used to evaluate the changes in hydroxyl radicals quantitatively. Briefly, cells were grown to 60%–70% confluence, and then incubated under the different experimental conditions in 6 well plates for 48 h. The media was then replaced with fresh serum-free media containing 10 μM DCHF-DA at 37°C for 30 min. Thereafter, cells were washed, trypsinized, and suspended in 500 μL phosphate-buffered saline (PBS), and the fluorescence intensity was measured immediately using a flow cytometer (BD ImmunoCytometry Systems, Franklin Lakes, NJ, USA).

Transient transfection
HK-2 cells were transfected transiently using FuGENE-HD transfection reagent according to the manufacturer’s instructions. Cells were seeded in 6 well plates at 80% confluence, and then transfected with 3.0 μg pLV-NR1H3-shRNA plasmid or scrambled control shRNA plasmid with 9 μL transfection reagent in 2 mL normal DMEM-F12 (3:1) medium. After 24 h, the cells were washed once with serum-free DMEM-F12 (3:1) medium, treated with HG or HG plus anthocyanins for 48 h, and then analyzed using Western blotting and filipin staining.

ELISA assay
The supernatants from different groups of cells were collected and the concentrations of the proinflammatory cytokines, ICAM1, MCP1, and TGFβ1, were quantified using commercially available ELISA kits (Neobioscience), according to the manufacturer’s instructions.

Western blotting
Nuclear and cytoplasmic proteins were extracted from HK-2 cells using a Nuclear and Cytoplasmic Protein Extraction Kit (Invitrogen). Proteins (50 μg/lane) were separated by sodium dodecyl sulfate–polyacrylamide gel electrophoresis (SDS-PAGE), and transferred to polyvinylidene difluoride membranes (Millipore, Billerica, MA, USA). The membranes were incubated overnight at 4°C with anti-ABCA1, -PPARα, -LXRα, -MCP1, -ICAM1, -TGFβ1, -β-actin, and -histone H3.
antibodies. Next, the membranes were incubated with goat anti-rabbit or mouse IgG horseradish peroxidase-conjugated secondary antibodies, and then scanned using an Odyssey Fc System (LI/-COR; Amersham, Piscataway, NJ, USA). The intensity of the bands was measured using LabWorks 4.5 software (UVP, Upland, CA, USA).

RNA isolation and quantitative real-time polymerase chain reaction (RT-qPCR)
Total RNA and cDNA were prepared from cultured cells using TRIzol reagent (Invitrogen) and a TaKaRa RNA PCR kit (AMV) version 3.0 (TaKaRa Bio Inc.), respectively. cDNA was amplified using PCR with specific primers for PPARα, LXRα, ABCA1, MCP1, ICAM1, TGFβ1, and 18s rRNA (Table 1). RT-qPCR was performed using SYBR Premix Ex TaqTMII and an Agilent Mx3000P QPCR System (Agilent, CA, USA). The relative changes in gene expression were calculated using the 2^ΔΔCT method, and all experiments were repeated at least thrice.

Oil Red O staining
HK-2 cells were allowed to adhere to polylysine-coated cover-slips and grown for 24 h. Cells were pretreated with or without C3G or Cy (50 µM) for 1 h, and then exposed to HG (30 mM) for 24 or 48 h. Subsequently, total cellular cholesterol was measured using Oil Red O staining. Cells were fixed in 4% paraformaldehyde in PBS for 30 min and then stained for 15 min in 1% Oil Red O dissolved in 60% isopropanol. Then, the sections were washed with 70% alcohol for 5 seconds (s) to remove background staining. Finally, the cells were rinsed in tap water and counterstained with Harris hematoxylin for 10 s. The stained sections were imaged with an Olympus microscope (Olympus Corporation, Tokyo, Japan).

Cholesterol efflux assays
Cholesterol levels were assayed using an Amplex Red Cholesterol Assay kit (Molecular Probes; Invitrogen) according to the manufacturer’s instructions. Briefly, HK-2 cells were treated as described above in 96 well plates, and then 50 µL Amplex Red reagent was added. After 45 min incubation at 37°C in the dark, sample fluorescence was measured using a microplate reader (Synergy 2; BioTek) with excitation at 530/525 nm and emission at 590/535 nm. Both the extracellular (in the growth medium) and intracellular cholesterol were analyzed. Cholesterol esterase was omitted from the assay. Each sample was analyzed in triplicate, and at least three independent experiments were performed.

Immunocytochemistry
Endogenous peroxidase activity was blocked using 3% hydrogen peroxide for 10 min, and the cells were incubated overnight at 4°C with anti-NFκB p65 (1:100) monoclonal antibody. Subsequently, the cells were incubated with biotin-labeled anti-rabbit IgG for 1 h, and then stained using a streptavidin-peroxidase detection system. Negative control samples were incubated with PBS instead of the primary antibody.

Statistical analysis
Values are expressed as means ± SDs. Significant differences between groups were determined using one-way analysis of variance using SAS software (release 8.01; SAS Institute, Inc., Cary, NC, USA); P<0.05 was considered to be statistically significant.

Results
Effect of anthocyanins on cell viability and radical scavenging
The potential cytotoxic effects of anthocyanins were evaluated first using MTT assays. The results showed that C3G and Cy did not affect the viability of HK-2 cells at the concentrations used (10, 20, and 50 µM; Figure 2A and B). Thus, C3G and Cy did not have cytotoxic effects in HK-2 cells. Because anthocyanins have powerful antioxidative properties, we next evaluated whether C3G and Cy reduced HG-induced ROS production in HK-2 cells, and the doses required for these effects. As shown in Figure 2C and D, HG increased intracellular ROS levels significantly, and this effect was reduced by C3G and Cy in a dose-dependent manner. These results indicate that C3G and Cy could reduce the intracellular ROS levels effectively.

Table 1 Primer sequences used for real-time polymerase chain reaction

| Gene   | Forward primer     | Reverse primer     | Product (base pair) |
|--------|--------------------|--------------------|--------------------|
| PPARα  | AGAAGCTGTCAACACAGTAGC | CGCGTGGACTCCGTAAGAT | 132                |
| LXRα   | GCTGAGACCTCTGCGATCG | CAAGGCAAAACTCGGATCA | 108                |
| ABCA1  | GCACCTGGAGAATGCTGAAAA | AGTTCCGGGAAGCTCTGTCCAC | 144             |
| ICAM1  | CGCTGCCGCACTCTGGTTC | CATGTCAACAGCGAGGACCC | 117                |
| MCP1   | GGCCCTTCTGTGCGCTCTGC | CGTGGATTCTTCTATGCTGC | 152                |
| TGFβ1  | GCCGACTAATCAGCGCAAGGAGG | CTCTCGGAAGCTGTAGGTGGT | 126                |
| 18S    | GCAAGCAAGGAGATGACGAGT | CTGCCGCAAGTTAGTTTGTCC | 92                  |
in a dose-dependent manner under HG conditions. Taken together with the results of MTT assays, we selected the dose of 50 µM of anthocyanins for use in subsequent experiments. In addition, mannitol had no effect on ROS levels.

**Anthocyanins enhance cholesterol efflux and ABCA1 expression in HG-stimulated HK-2 cells**

To determine whether anthocyanins weakened or prevented HG to decrease cholesterol efflux, we first analyzed cholesterol levels inside the cell using Oil Red O staining. Compared with the NG groups, HG enhanced cholesterol levels significantly inside the cell after treatment for 24 h, and this continued to increase until 48 h. In contrast, treatment with C3G and Cy markedly attenuated HG-induced cholesterol levels increasing inside the cell (Figure 3A). Next, we investigated the changes in cholesterol concentrations in the culture media and inside the cell using cholesterol efflux analysis. The results were consistent with Oil Red O staining (Figure 3B and C).

Because ABCA1 is important for cholesterol efflux, we next examined the effect of anthocyanins on ABCA1 expression. The mRNA and protein levels of ABCA1 significantly decreased after treatment with HG for 24 h, and remained at low levels until 48 h. Treatment with C3G and Cy reversed the HG-induced decrease in ABCA1 markedly (Figure 3D and E). These results suggest that anthocyanins enhanced cholesterol efflux from HK-2 cells by upregulating ABCA1.

**Anthocyanins increase the gene expression of PPARα and LXRα in HG-stimulated HK-2 cells**

To investigate the potential mechanism by which anthocyanins induce ABCA1 expression upregulation and cholesterol efflux, we analyzed the expression of PPARα and LXRα. As shown in Figure 4, HG treatment decreased the protein and mRNA expression of PPARα and LXRα at 24 and 48 h, respectively. Treatment with 50 µM C3G and Cy reversed HG-induced decreased protein (Figure 4A) and mRNA (Figure 4B) PPARα and LXRα levels markedly.

The upregulating of ABCA1 gene expression by anthocyanins is dependent on PPARα/LXRα in HG-stimulated HK-2 cells

We first treated HK-2 cells with both HG and C3G or Cy in the presence of 25 µM GW6471 (PPARα inhibitor) and

---

**Figure 2** Effects of C3G and Cy on the cell viability and ROS levels in HK-2 cells.

**Notes:** Cytotoxic effects of C3G (A) and Cy (B) on HK-2 cells was determined by MTT assay. Cells were cultured with different concentrations of C3G and Cy (0–100 µM) in the absence or presence of 30 mM HG for 24 h. The values presented are the means ± SD of three independent experiments. *P<0.05 versus control group. Effects of C3G (C) and Cy (D) on radical scavenging activity in HG-stimulate HK-2 cells. Cells were preincubated with or without different concentrations of C3G and Cy (0–100 µM) for 1 h and then treated with 30 mM HG for 24 h. Quantification of the ROS levels as detected by flow cytometry. The values presented are the means ± SD of three independent experiments and differences between mean values were assessed by an ANOVA. **P<0.01 versus control group, #P<0.01 versus HG group.

**Abbreviations:** ANOVA, analysis of variance; C3G, cyanidin-3-O-β-glucoside chloride; Cy, cyanidin chloride; DCHF-DA, dichlorodihydrofluorescein diacetate; h, hour/s; HG, high-glucose; M, mannitol; NG, normal glucose; ROS, reactive oxygen species.
Figure 3 Anthocyanins enhance cholesterol efflux and ABCA1 expression in HG-stimulated HK-2 cells.

Notes: Cells were preincubated with or without C3G and Cy (50 µM) for 1 h and then treated with 30 mM HG for 24 or 48 h. (A) Cholesterol mass inside the cell was measured by Oil red staining. (B and C) Effect of C3G and Cy on cholesterol concentration in the medium (B) and intracellularly (C) was assayed by an Amplex Red Cholesterol Assay kit. (D) The expression levels of ABCA1 mRNA were analyzed by RT-qPCR. (E) The expression levels of ABCA1 protein were detected by Western blot. The values presented are the means ± SD of three independent experiments. *P < 0.05, **P < 0.01 versus control group, #P < 0.05, ##P < 0.01 versus HG group.

Abbreviations: C3G, cyanidin-3-O-β-glucoside chloride; Cy, cyanidin chloride; h, hour/s; HG, high-glucose; NG, normal glucose; RT-qPCR, quantitative real-time polymerase chain reaction.

observed the effect of GW6471 on LXRα and ABCA1 gene expression and cholesterol efflux. The results showed that GW6471 reduced LXRα and ABCA1 expression compared with the NG group. The coincubation of anthocyanins and GW6471 blocked the anthocyanin-induced increase in LXRα and ABCA1 protein (Figure 5A) and mRNA (Figure 5B) expression and the cholesterol efflux (Figure 5C and D).

These results suggest that anthocyanins induce ABCA1 gene expression via the PPARα pathway.

Next, we used transfection to determine the effects of silencing LXRα on anthocyanin-induced ABCA1 gene expression and cholesterol efflux. Compared with scrambled control shRNA, LXRα shRNA suppressed LXRα expression by 76% (P < 0.01; Figure 6A). In addition, silencing LXRα
reversed the effects of anthocyanins on ABCA1 expression (Figure 6B and C) and cholesterol efflux (Figure 6D). These results indicate that anthocyanins increase ABCA1 expression and cholesterol efflux via an LXRα-dependent pathway.

Effects of anthocyanins on cytokine production in HG-stimulated HK-2 cells

To analyze the anti-inflammatory effects of anthocyanins in the progression of the DN, we measured the protein and mRNA levels of ICAM1, MCP1, and TGFβ1 using Western blotting, quantitative real-time (qRT)-PCR, and ELISA. Western blotting analysis revealed that HG increased the protein levels of ICAM1, MCP1, and TGFβ1 at 24 and 48 h significantly compared with NG group. In contrast, these HG-induced increases were decreased significantly by C3G and Cy (Figure 7A). The results of qRT-PCR (Figure 7B) and ELISA (Figure 7C) were consistent with these observations.

The anthocyanin-induced reduction in ICAM1, MCP1, and TGFβ1 expression is dependent on the activation of LXRα, but not PPARα

Recent reports22–26 demonstrated that PPARα and LXRα mediated the expression of some inflammatory cytokines. Therefore, we next investigated the possible effects of PPARα and LXRα on the inhibition of HG-mediated inflammatory signaling by anthocyanins. First, HK-2 cells were transfected with pLV-NR1H3-shRNA plasmid and then incubated with HG in the presence or absence of C3G or Cy for 48 h. Compared with the anthocyanin treatment groups, transfection with pLV -NR1H3-shRNA reversed the effects of anthocyanins on HG-induced ICAM1, MCP1, and TGFβ1 expression significantly (Figure 8A and B). Next, we used the specific PPARα inhibitor GW6471 to observe the role of PPARα in the anthocyanin-mediated inhibition of HG-induced ICAM1, MCP1, and TGFβ1 expression. Co-pretreatment with 25 µM GW6471 and C3G or Cy for 48 h slightly reversed the inhibitory effects of C3G or Cy on HG-induced ICAM1, MCP1, and TGFβ1 expression (Figure 8A and B). These results suggest that anthocyanins decreased the expression of ICAM1, MCP1, and TGFβ1 by upregulating LXRα transcriptional activity, but not PPARα.

Effects of anthocyanins on NFκB activation in HG-stimulated HK-2 cells

As shown in Figure 9, HG enhanced the nuclear and decreased the cytoplasmic localization of NFκB p65 (P<0.05). However, HG-induced NFκB p65 nuclear translocation was
Figure 5 Effects of GW6471 on anthocyanins-induced LXRα and ABCA1 gene expression and the cholesterol efflux in HK-2 cells.

Notes: Cells were preincubated with both GW6471 (25 µM) and C3G or Cy for 1 h, followed by treatment with 30 mM HG for 48 h. (A) The expression levels of LXRα and ABCA1 protein were detected by Western blot. (B) The expression levels of LXRα and ABCA1 mRNA were analyzed by RT-qPCR. (C) Cholesterol mass inside the cell was measured by Oil red staining. (D) Cholesterol concentration in cells was assayed by an amplex red cholesterol assay kit. The values presented are the mean ± SD of three independent experiments. *P < 0.01 versus control group, #P < 0.05 versus HG group. $P < 0.05 versus HG + C3G group, $P < 0.01 versus HG + Cy group.

Abbreviations: C3G, cyanidin-3-O-β-glucoside chloride; Cy, cyanidin chloride; h, hour/s; HG, high-glucose; RT-qPCR, quantitative real-time polymerase chain reaction.

Attenuation of the anti-inflammatory effects of anthocyanins is most likely mediated by NFκB signaling.

Anthocyanin-induced NFκB activity reduces cytokine production via the LXRα pathway

To test whether NFκB activation involved in the anti-inflammatory effects of anthocyanins is mediated by the LXRα pathway, we next detected the expression of NFκB p65 when LXRα was silenced in HK-2 cells using pLV-NR1H3-shRNA plasmid. As expected, the inhibitory effect of anthocyanins on NFκB activity was reversed markedly by transfection with pLV-NR1H3-shRNA (Figure 10A). We then used immunocytochemistry to investigate the nuclear translocation of NFκB p65. Data revealed that NFκB p65 was localized predominately in the cytoplasm in control cells, whereas HG treatment caused pronounced NFκB p65 nuclear staining. However, C3G and Cy decreased the nuclear distribution of NFκB p65 (Figure 10B). Moreover, transfection with pLV-NR1H3-shRNA led to predominantly nuclear NFκB p65 staining after HG treatment. Taken together, these data suggest that anthocyanins inhibit HG-induced inflammatory gene expression by activating LXRα and affecting the nuclear translocation of NFκB.

Discussion

The drugs used to treat diabetes and its complications often cause a variety of side effects. Therefore, increasing attention has focused on studying phytochemicals that are easy to obtain and lack numerous side effects. Anthocyanins are naturally occurring phytochemicals and are widely distributed in vegetables,
Recent work suggested that decreased cellular cholesterol efflux contributes to the progression of DN. Renal cholesterol accumulation was reported in podocytes, diabetic NOD mice, and an animal model of acute renal tubular injury. ABCA1 is a well-characterized cholesterol transporter that regulates cellular lipid homeostasis. Some reports showed that anthocyanins significantly ameliorated hypercholesterolemia and suppressed cholesterol accumulation by upregulating ABCA1 in the liver and aorta, suggesting the anthocyanins might remove cholesterol from tissues. Preclinical studies revealed that dietary anthocyanin-rich extracts ameliorated hyperglycemia and hyperlipidemia in high-fructose-fed rats and experimental type 2 diabetic mice. Here, we showed that HG inhibited cholesterol efflux and increased the intracellular cholesterol content in HK-2 cells significantly, which was accompanied by reduced ABCA1 expression. C3G and Cy treatment reversed these effects: increased ABCA1 mRNA and protein expression and intracellular cholesterol

Abbreviations: C3G, cyanidin-3-O-β-glucoside chloride; Cy, cyanidin chloride; h, hour/s; HG, high-glucose; RT-qPCR, quantitative real-time polymerase chain reaction.
efflux were observed in HK-2 cells cotreated with HG and anthocyanins.

The levels of proinflammatory cytokines such as ICAM1, MCP1, and TGFβ1 are elevated in diabetic kidneys, suggesting that renal inflammation might contribute to the pathogenesis of DN. Recent studies focused on the anti-inflammatory properties of anthocyanins for application in a large number of inflammatory diseases. C3G inhibited the expression of TNFα, IL1β, IL6, IL10, and IFNβ in LPS-stimulated alveolar

Figure 7 Effects of anthocyanins on HG-induced MCP1, ICAM1, and TGFβ1 expression in HK-2 cells.
Notes: Cells were preincubated with or without C3G and Cy (50 µM) for 1 h and then treated with 30 mM HG for 24 or 48 h. The expression levels of MCP1, ICAM1, and TGFβ1 protein were detected by Western blot (A) and mRNA was analyzed by RT-qPCR (B). Levels of MCP1, ICAM1, and TGFβ1 in culture supernatants were measured by ELISA (C). The values presented are the mean ± SD of three independent experiments. *P<0.05, **P<0.01 versus control group, #P<0.05, ##P<0.01 versus HG group.
Abbreviations: C3G, cyanidin-3-O-β-glucoside chloride; Cy, cyanidin chloride; h, hour/s; HG, high-glucose; NG, normal glucose; RT-qPCR, quantitative real-time polymerase chain reaction; ELISA, enzyme-linked immunosorbent assay.
The protective effects of anthocyanins in diabetic nephropathy

Figure 8 Effects of GW6471 and plV-nr1h3-shRNA plasmid on anthocyanins-mediated reduction of MCP1, ICAM1, and TGFβ1 in HK-2 cells.

Notes: (A) The expression levels of MCP1, ICAM1, and TGFβ1 protein were detected by Western blot. (B) The expression levels of MCP1, ICAM1, and TGFβ1 mRNA were analyzed by RT-qPCR. The values presented are the mean ± SD of three independent experiments. *P<0.01 versus control group, #P<0.01 versus HG group. $P<0.01 versus HG + C3G group, †P<0.01 versus HG + Cy group.

Abbreviations: C3G, cyanidin-3-O-β-glucoside chloride; Cy, cyanidin chloride; HG, high-glucose; RT-qPCR, quantitative real-time polymerase chain reaction.

The protective effects of anthocyanins in diabetic nephropathy

Macrophages in a model of acute lung injury. In addition, PCA inhibited MCP1 expression and attenuated the induction of ICAM1 to ameliorate the diabetic state in diabetic kidneys. PCA also retarded diabetes-associated renal fibrosis and mesangial inflammation by disturbing TGFβ1 signaling. This study revealed that C3G and Cy inhibited the production of MCP1, ICAM1, and TGFβ1 both intracellularly and extracellularly. NFκB is an important transcription factor that plays a critical role in inducing cytokine production; the anthocyanin-mediated inhibition of HG-induced cytokine production was likely mediated by the NFκB pathway.

Figure 9 Effects of anthocyanins on HG-induced NFκB expression in HK-2 cells.

Notes: Cells were preincubated with or without C3G and Cy (50 µM) for 1 h and then treated with 30 mM HG for 24 or 48 h. The expression levels of NFκB p65 cytoplasmic protein and nuclear protein were detected by Western blot. The values presented are the mean ± SD of three independent experiments. *P<0.05 versus control group, #P<0.05 versus HG group.

Abbreviations: C3G, cyanidin-3-O-β-glucoside chloride; Cy, cyanidin chloride; h, hour/s; HG, high-glucose; NG, normal glucose.
Figure 10 Knockdown of LXRα abrogated the anthocyanins-induced NFκB gene expression in HK-2 cells.

Notes: Cells were transfected with pLV-NR1H3-shRNA plasmid or scrambled control shRNA plasmid for 24 h and then treated with HG (30 mM) in the presence or absence of C3G and Cy (50 µM) for 48 h, respectively. (A) The expression levels of NFκB cytoplasm protein and nuclear protein were detected by Western blot. (B) Nuclear translocation of NFκB p65 was investigated by immunocytochemistry staining. The values presented are the mean ± SD of three independent experiments. *P<0.01 versus control group, #P<0.01 versus HG group. $P<0.01 versus HG + C3G group, ^P<0.01 versus HG + Cy group.

Abbreviations: C3G, cyanidin-3-O-β-glucoside chloride; Cy, cyanidin chloride; h, hour/s; HG, high-glucose; NG, normal glucose.
Nuclear LXR, and particularly LXRα, is an extensively studied and validated gene whose primary function is to maintain cellular cholesterol homeostasis.18,19 LXRα regulates intracellular cholesterol levels by mediating the expression of its target gene ABCA1 by binding to an LXR element in the ABCA1 promoter.42 In addition to LXRα, the nuclear receptor superfamily member PPARα plays a critical role in lipid metabolism and regulates the expression of ABCA1.18,19 The activation of PPARα reduces triglyceride, total cholesterol, and LDL levels markedly, and consequently elevates HDL levels.43 Moreover, previous studies demonstrated that crosstalk occurs between PPARα and LXRα during the regulation of cholesterol efflux and that PPARα induces LXRα expression directly via a response element upstream of the LXRα promoter region. This leads to the induction of ABCA1, followed by a transcriptional cascade that regulates cholesterol removal from macrophages.18,19 In addition, PPARα agonists induce ABCA1 expression and cholesterol efflux in macrophages in an LXRα-dependent manner.22,23

To investigate the molecular mechanisms by which C3G and Cy increased intracellular cholesterol efflux and ABCA1 expression in HK-2 cells, we measured their effects on PPARα and LXRα activation in the presence and absence of GW6471, a PPARα inhibitor, and pLV-NR1H3-shRNA, an LXRα silencing plasmid. Data revealed that C3G and Cy increased PPARα and LXRα expression. In addition, GW6471 and LXRα shRNA abolished the anthocyanin-mediated increase in cholesterol efflux and ABCA1 expression. Moreover, GW6471 also attenuated anthocyanin-mediated LXRα expression significantly. In contrast, LXRα shRNA did not affect anthocyanin-induced PPARα expression. Therefore, these combined findings illustrate a complex pathway involving PPARα, LXRα, and ABCA1 that participates in the anthocyanin-mediated increase in cholesterol efflux from HK-2 cells.

Accumulating evidence suggests that PPARα and LXRα also play an anti-inflammatory role in many inflammatory and metabolic diseases.22,35 Our previous experiments demonstrated that anthocyanin inhibited the production of proinflammatory cytokines. Therefore, we detected the effects of GW6471 and pLV-NR1H3-shRNA plasmid on the anthocyanin-mediated decrease in the proinflammatory cytokines MCP1, ICAM1, and TGFβ1. Taken together, these results suggest that the anti-inflammatory effects of the anthocyanins C3G and Cy in HK-2 cells are partly mediated by inhibiting the LXRα-stimulated nuclear translocation of NFκB, which further decreases the levels of the proinflammatory cytokines MCP1, ICAM1, and TGFβ1.

In conclusion, this study demonstrated that anthocyanins modulate cholesterol metabolism and the inflammatory response in HG-stimulated HK-2 cells, suggesting that anthocyanins might have novel therapeutic effects in DN. In addition, C3G and Cy had similar anti-hyperlipidemic and anti-inflammatory properties. Therefore, we hypothesize that the anti-hyperlipidemic and anti-inflammatory activities are due to their aglycone moiety. However, more work is needed to clarify the structure–function relationship and the effects of anthocyanins.

Acknowledgment

This study was supported by the National Natural Science Foundation of the People’s Republic of China (numbers 81270804 and 81370825) and the Hebei Science and Technology Department Program (12966118D).

Disclosure

All authors report no conflicts of interest in this work.

References

1. Kelly KJ, Dominguez JH. Rapid progression of diabetic nephropathy is linked to inflammation and episodes of acute renal failure. Am J Nephrol. 2010;32:469–475.
2. Abdel-Rahman EM, Saadulla L, Reeves WB, Awad AS. Therapeutic modalities in diabetic nephropathy: standard and emerging approaches. J Gen Intern Med. 2012;27:458–468.
3. Herman-Edelstein M, Scherzer P, Tobar A, Levi M, Gafter U. Altered renal lipid metabolism and renal lipid accumulation in human diabetic nephropathy. J Lipid Res. 2014;55:561–572.
4. Zhang MH, Feng L, Zhu MM, et al. The anti-inflammation effect of Moutan Cortex on advanced glycation end products-induced rat mesangial cells dysfunction and High-glucose-fat diet and streptozotocin-induced diabetic nephropathy rats. J Ethnopharmacol. 2014;151:591–600.

5. Kuhad A, Sachdeva AK, Chopra K. Attenuation of renoinflammatory cascade in experimental model of diabetic nephropathy by salomol. J Agric Food Chem. 2009;57:6123–6128.

6. Tesch GH, MCP-1-CCCL2, a new diagnostic marker and therapeutic target for progressive renal injury in diabetic nephropathy. Am J Physiol Renal Physiol. 2008;294:F697–F701.

7. Hills CE, Al-Rasheed N, Al-Rasheed N, Willars GB, Brunskill NJ. C-peptide reverses TGF-beta-induced changes in renal proximal tubular cells: implications for treatment of diabetic nephropathy. Am J Physiol Renal Physiol. 2009;296:F614–F621.

8. Li J, Lim SS, Lee JY, et al. Purple corn anthocyanins dampened high-glucose-induced mesangial fibrosis and inflammation: possible renoprotective role in diabetic nephropathy. J Nutr Biochem. 2012;23:320–331.

9. Korrapti MC, Howell LH, Shaner BE, Megyesi JK, Siskind LJ, Schnellmann RG. Suramin: a potential therapy for diabetic nephropathy. PLoS One. 2013;8:e73655.

10. Soetikno V, Sari FR, Veeraveedu PT, et al. Curcumin ameliorates macular degeneration by inhibiting NF-kappaB. J Nutr Biochem. 2006;17:1633–1643.

11. Zielonka SK, Zarkowski P, Zawadzki S, et al. Naturally occurring bioactive compounds: therapeutic potential for diabetic nephropathy. J Nutr Biochem. 2006;17:381–392.

12. Tang SC, Leung JC, Chan LY, Tsang AW, Lai KN. Activation of tubular epithelial cells in diabetic nephropathy and the role of the peroxisome proliferator-activated receptor-gamma agonist. J Am Soc Nephrol. 2006;17:1633–1643.

13. Xie X, Peng J, Chang X, et al. Activation of RhoA/ROCK regulates NF-kappa B signaling pathway in experimental diabetic nephropathy. Mol Cell Endocrinol. 2013;369:86–97.

14. Ruan XZ, Moorhead JF, Fernando R, Wheeler DC, Powis SH, et al. Cyanidin-3-glucoside-rich extract reduces the cholesterol exporter ABCA1 in mouse macrophages and reverses cholesterol efflux pathway. J Nutr Biochem. 2012;23:320–331.

15. Tang C, Kanter JE, Bornfeldt KE, Leboeuf RC, Oram JF. Diabetes reduces the cholesterol exporter ABCA1 in mouse macrophages and kidneys. J Lipid Res. 2010;51:1719–1728.

16. Zager RA, Johnson AC, Hanson SY, Shah VO. Acute tubular injury causes dysregulation of cellular cholesterol transport proteins. Am J Pathol. 2003;163:313–320.

17. Schmitz G, Langmann T. Transcriptional regulatory networks in lipid metabolism control ABCA1 expression. Biochim Biophys Acta. 2005;1715:1–19.

18. Nakaya K, Tohyama J, Naik SU, et al. Peroxisome proliferator-activated receptor-alpha activation promotes macrophage reverse cholesterol transport through a liver X receptor-dependent pathway. Arterioscler Thromb Vasc Biol. 2011;31:1276–1282.

19. Proctor G, Jiang T, Iwahashi M, Wang Z, Li J, Levi M. Regulation of retinal fatty acid and cholesterol metabolism, inflammation, and fibrosis in Akita and OVE26 mice with type 1 diabetes. Diabetes. 2006;55:2502–2509.

20. Park CW, Kim HW, Ko SH, et al. Accelerated diabetic nephropathy in mice lacking the peroxisome proliferator-activated receptor alpha. Diabetes. 2006;55:885–893.

21. Johnston TP, Waxman DJ. The induction of atherogenic dyslipidaemia in poloxamer 407-treated mice is not mediated through PPARalpha. J Pharm Pharmacol. 2008;60:753–759.

22. Balakumar P, Kadian S, Mahadevan N. Are PPAR alpha agonists a rational therapeutic strategy for preventing abnormalities of the diabetic kidney? Pharmacol Res. 2012;65:430–436.

23. Kouroumichakis I, Papanas N, Zarogoulidis P, Liakopoulos V, Maltezos E, Mikhalilidis DP. Fibrates: therapeutic potential for diabetic nephropathy? Eur J Intern Med. 2012;23:309–316.
