CONCISE REVIEW

The emerging antioxidant paradigm of mesenchymal stem cell therapy

Rhian Stavely1,2 | Kulmira Nurgali1,3,4

1Institute for Health and Sport, Victoria University, Western Centre for Health, Research and Education, Sunshine Hospital, Melbourne, Victoria, Australia
2Department of Pediatric Surgery, Massachusetts General Hospital, Harvard Medical School, Boston, Massachusetts
3Department of Medicine Western Health, Faculty of Medicine, Dentistry and Health Sciences, The University of Melbourne, Melbourne, Victoria, Australia
4Regenerative Medicine and Stem Cells Program, Australian Institute of Musculoskeletal Science (AIMSS), Melbourne, Victoria, Australia

Correspondence
Kulmira Nurgali, MBBS, MSc, PhD, Western Centre for Health Research & Education, Sunshine Hospital, 176 Furlong Road, St Albans 3021, VIC, Australia.
Email: kulmira.nurgali@vu.edu.au

Abstract
Mesenchymal stem cells (multipotent stromal cells; MSCs) have been under investigation for the treatment of diverse diseases, with many promising outcomes achieved in animal models and clinical trials. The biological activity of MSC therapies has not been fully resolved which is critical to rationalizing their use and developing strategies to enhance treatment efficacy. Different paradigms have been constructed to explain their mechanism of action, including tissue regeneration, trophic/anti-inflammatory secretion, and immunomodulation. MSCs rarely engraft and differentiate into other cell types after in vivo administration. Furthermore, it is equivocal whether MSCs function via the secretion of many peptide/protein ligands as their therapeutic properties are observed across xenogeneic barriers, which is suggestive of mechanisms involving mediators conserved between species. Oxidative stress is concomitant with cellular injury, inflammation, and dysregulated metabolism which are involved in many pathologies. Growing evidence supports that MSCs exert antioxidant properties in a variety of animal models of disease, which may explain their cytoprotective and anti-inflammatory properties. In this review, evidence of the antioxidant effects of MSCs in in vivo and in vitro models is explored and potential mechanisms of these effects are discussed. These include direct scavenging of free radicals, promoting endogenous antioxidant defenses, immunomodulation via reactive oxygen species suppression, altering mitochondrial bioenergetics, and donating functional mitochondria to damaged cells. Modulation of the redox environment and oxidative stress by MSCs can mediate their anti-inflammatory and cytoprotective properties and may offer an explanation to the diversity in disease models treatable by MSCs and how these mechanisms may be conserved between species.

KEYWORDS
antioxidant, mesenchymal stem cell, mitochondria, multipotent stromal cell, oxidative stress, reactive oxygen species

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INTRODUCTION

Mesenchymal stem cells (multipotent stromal cells; MSCs) have been used as tools to treat a broad range of diseases in animal models due to their unique characteristics such as host immune evasion, rapid expansion, and their enfluenza in adult bone marrow and adipose tissue. The positive outcomes of these studies have driven hundreds of clinical trials into their application for diabetes, inflammatory disorders, and various liver, kidney, lung, cardiovascular, musculoskeletal, neurological, and gastrointestinal diseases. While several trials have demonstrated the therapeutic potential of MSCs, the failure to incorporate MSCs into current treatment regimens can be, in part, attributed to the lack of understanding pertaining to their biological mechanisms of action.

Initially, MSCs were explored as tools of regenerative medicine to replace damaged tissue. However, administered MSCs were rarely observed to differentiate and effectively engraft into host tissues despite demonstrating favorable effects in many disease models. Furthermore, the secretome of MSCs was identified to be therapeutic in many disease models in vitro and in vivo. Together, this resulted in a paradigm shift in recognition of the trophic actions of MSCs. Despite extensive research investigating the anti-inflammatory and trophic constituents of the MSC-derivered secretome, the therapeutic mechanisms of MSCs remain incompletely resolved. MSCs demonstrate therapeutic attributes across xenogeneic barriers and, therefore, the therapeutic mechanisms of MSCs may be similar between species. There is strong evidence that the effects of MSCs are mediated via the secretion of protein/peptide ligands; however, it is equivocal whether these ligands are effective across xenogeneic barriers.

Recently, the role of MSCs in ameliorating oxidative and nitrosative injury has received considerable attention. The reduction-oxidation (redox) environment regulates many physiological and pathophysiological mechanisms in cellular biology. Antioxidant effects of MSCs have been observed in various disease models such as diabetic injuries to the kidney, retina, sensory neurons, brain, and bone formation; chemotherapy- or radiation-induced injury to the lungs, gonads, aorta, and brain; ischemic injury of the brain, heart, kidney, and liver; and traumatic injury to the spine and testis, cognitive disorders, gastrointestinal inflammation, septic injuries, and aging (Figure 1; Table 1). MSCs can directly reduce oxidative stress-related injury in vitro in glial cells, neurons, cardiomyocytes, renal cells, endothelial cells, immune cells, hepatocytes, islet cells, fibroblasts, skeletal muscle, and other cells (Table 2). Oxidative stress is concomitant with cellular injury, inflammation, and dysregulated metabolism and, therefore, is key pathophysiological mechanism of many diseases. Oxidative stress and redox imbalance are mediated by molecular constituents that are present in all living cells and share similar functions. Thus, the ability of MSCs to regulate these processes may offer an explanation to the diversity of disease models treatable by MSCs and to the effects of MSCs conserved between species.

Oxidative stress refers to a deviation from the physiological redox state and an increase in pro-oxidants, or free radicals, that structurally change lipids, proteins, and DNA in a way that causes pathology or damage to a cell or tissue. The most widely studied free radicals are reactive oxygen species (ROS), which can also include reactive molecules that have a stable charge. The three major endogenous ROS include the superoxide anion (O$_2^-$), hydroxyl radical (•OH), and hydrogen peroxide (H$_2$O$_2$). O$_2^-$ is predominantly generated by nicotinamide adenine dinucleotide phosphate reduced (NADPH)-oxidase (NOX) family enzymes or, by the mitochondria, as a by-product of oxidative phosphorylation. The level of mitochondria-derived O$_2^-$ depends on metabolic substrates, cytosolic Ca$^{2+}$ levels, pH, and oxygen tension. O$_2^-$ generated from complexes of the electron transport chain (ETC) are highly reactive and can damage the mitochondrion. The detoxification of O$_2^-$ into H$_2$O$_2$ is mediated by superoxide dismutase (SOD). However, H$_2$O$_2$ can also be generated in various metabolic processes and by dual oxidases (DUOX). While H$_2$O$_2$ is more stable than O$_2^-$, its detoxification is crucial as it possesses a weak peroxide bond that makes it susceptible to reacting with metals, such as Fe$^{2+}$, to generate reactive •OH through the Fenton reaction. Both, H$_2$O$_2$ and O$_2^-$, are diffusible across cell membranes and can promote cell death and inflammatory signaling. Several studies have demonstrated that MSCs can reduce ROS and biomarkers of oxidative stress. In this review, evidence of direct and indirect antioxidant mechanisms of MSC therapies is explored.

Significance statement

The role of mesenchymal stem cells (MSCs) in ameliorating oxidative and nitrosative injury has received considerable attention in recent years. The reduction-oxidation (redox) environment regulates many physiological and pathophysiological mechanisms in cellular biology. Oxidative stress and redox imbalance are mediated by molecular constituents that are present in all living cells and share similar functions. The ability of MSCs to regulate these processes may offer an explanation to the diversity of disease models treatable by MSCs and to the effects of MSCs conserved between species. In this review, evidence of direct and indirect antioxidant mechanisms of MSC therapies is explored.

MSCs ARE RESISTANT AND RESPOND TO OXIDATIVE STRESS

The therapeutic properties of MSCs have been explored in many models of disease associated with high levels of ROS and biomarkers of oxidative injury. MSCs must survive these volatile environments to exert their therapeutic effects, which can present as a challenge for their engraftment after administration. Nonetheless, several studies
have demonstrated that MSCs are highly resistant to oxidative insult. The oxidative effects of ionizing radiation are limited on MSCs which have been attributed to their ability to directly scavenge free radicals.\textsuperscript{16} It has been demonstrated that MSCs are resistant to oxidative and nitrosative stimuli in vitro which is associated with constitutively expressed antioxidant enzymes SOD1, SOD2, catalase (CAT), and glutathione peroxidase (GPx), in addition to high levels of the antioxidant glutathione (GSH).\textsuperscript{17} Depletion of GSH results in a loss of tolerance to oxidative stress. MSCs also constitutively express heat-shock protein 70 (HSP70) and sirtuin (SIRT)\textsuperscript{3,18} which may also play a role in the resistance of MSCs to oxidative/nitrosative injury. SIRT1 is also required for MSC survival against H$_2$O$_2$ and its overexpression has a protective effect.\textsuperscript{19} Likewise, SIRT6 has been suggested to confer resistance to oxidative insult and basal ROS production in MSCs via downstream production of antioxidants including heme oxygenase-1 (HO-1).\textsuperscript{20} Overexpression of HO-1 ameliorates elevations in ROS and cellular senescence in SIRT6-null MSCs and, therefore, appears to be a critical component of the survival mechanism of MSCs in oxidative environment.\textsuperscript{20}

In addition to wielding constitutive antioxidants, MSCs are also capable of significant adaptations in response to redox stress. MSCs exposed to lipopolysaccharide (LPS) produce oxidative and nitrosative free radicals.\textsuperscript{18} In parallel, several adaptive processes are observed including the upregulation and/or nuclear translocation of redox-sensitive factors (nuclear factor kappa-B [NFkB], thioredoxin [TRX1], apurinic/apyrimidinic endonuclease redox effector factor-1 [APE1/Ref-1], nuclear factor erythroid 2-related factor 2 [NRF2], forkhead box O3 [FOXO3], and HO-1), as well as mitochondrial remodeling and autophagy. Similarly, MSCs exposed to hypoxic conditions (1.5%-2% O$_2$) exhibit increased intracellular ROS and cells respond by upregulating the expression of hypoxia-inducible factor 1 alpha (HIF-1x), erythropoietin receptor, CAT,
| Application                      | Model                                                                 | MSCs used                                                                 | Effects of MSC treatment                                                                 | Antioxidant mechanisms                                                                 | References |
|---------------------------------|-----------------------------------------------------------------------|--------------------------------------------------------------------------|------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------|------------|
| Aging                            | Aging-related erectile dysfunction (rat)                              | Rat AT-MSCs                                                              | Erectile response                                                                        | Lipid peroxidation, SOD activity                                                      | 62         |
| Premature aging (Bmi−/−) (mouse) | Mouse amniotic membrane MSCs                                          | AT-MSCs                                                                  | Survival time, Apoptosis in thymus and kidney, Proliferation in thymus and kidney, Mature immune cells, Skeletal muscle growth, Osteoporosis, Bmi-1 in liver, kidney, thymus, muscle, spleen, lung, and bone marrow | H2O2, CAT, SOD in the heart, liver, spleen, lung, kidney, BM and thymus, ROS in all except heart, DNA damage in cells of BM, spleen, lung and thymus, MSCs secrete SOD (total) and CAT | 33         |
| Chemotherapy and radiation       | Bleomycin-induced pulmonary fibrosis (rat)                           | Rat mesenchymal stem cells (H4320-1)                                      | Fibrosis                                                                                | Nrf2, αNQO1, HO-1, γGCS, Lipid peroxidation, SOD activity                             | 63         |
|                                 | Bleomycin-induced pulmonary fibrosis (mouse)                         | Human BM-MSCs cell line U6E7T-2                                           | Collagen                                                                                | DNA oxidation, ER stress marker BIP, Effects negated by silencing STC-1 and enhanced by STC-1 over expression | 127        |
|                                 | Cisplatin-induced acute kidney injury (rat)                          | Human UC-MSC exosomes                                                    | Blood urea nitrogen (MSC-CM and fibroblast exosomes had no effect), Creatinine (MSC-CM and fibroblast exosomes had no effect), Gross morphological damage, Apoptosis (TUNEL), PCNA, Bax, Bcl-2, pS6AMPK | DNA oxidation, GSH, Lipid peroxidation                                               | 27         |
|                                 | Cisplatin-induced cognitive impairment (mice)                         | Mouse BM-MSCs Intranasal delivery                                         | Cognitive function                                                                       | Maximal respiratory capacity and spare respiratory capacity of mitochondria, Morphologically atypical mitochondria | 112        |
|                                 | Cisplatin-induced gonadotoxicity (rat)                               | Rat BM-MSCs                                                              | Testis weight and testosterone levels, TNF                      | Lipid peroxidation, SOD activity, GSH, INOS                                     | 64         |
|                                 | Cisplatin-induced renal injury (mouse)                              | Mouse BM-MSC-CM                                                          | Weight loss, Serum creatinine levels, c-caspase 3 expression, Gross morphological damage | HO-1−/− MSCs did not demonstrate therapeutic value                                    | 94         |
| Lung radiation injury (mouse)    | Mouse aorta-derived and BM-MSCs                                      | Mouse BM-MSCs                                                            | Lung fibrosis                                                                            | Aorta and BM-MSCs secrete SOD1, SOD1 expression in irradiated lung, SOD1 mimetic replicated effect of MSCs   | 59         |
|                                 | Paclitaxel-induced neuropathy (rat)                                  | Rat BM-MSCs                                                              | Responses to thermal hyperalgesia and cold allodynia, Sciatic nerve: NGF, Pro-inflammatory cytokines, c-caspase 3 | Lipid peroxidation, GSH, INOS                                                      | 55         |
|                                 | Radiation-induced aortic injury (mouse)                              | Human BM-MSCs                                                            | Aorta thickness, Collagen, TGF, TNF, ICAM, Apoptosis, HO-1, CAT | Nitrotyrosine, Lipid peroxidation, CAT                                               | 82         |
|                                 | Radiation-induced neurological complication (mouse)                  | Human AT-MSCs                                                            | Cognitive function, Neuron loss, Caspase 3                                             | Lipid peroxidation in hippocampus and brain lateral ventricle, INOS                  | 41         |
| Hyperglycemic injuries           | Alloxan-induced diabetes (rat)                                       | Rat BM-MSCs                                                              | Insulin, Glucose levels, Total cholesterol, Triglycerides, Vitamin E                     | GSH, GST, SOD, NO, Lipid peroxidation                                               | 88         |
|                                 | Db/db mouse model of type 2 diabetes                                 | Mouse amniotic fluid MSCs                                                | Improved kidney function, Weight gain, Pro-inflammatory cytokines, Apoptosis, Overexpression of Sirtuin3 in MSCs improved all effects | Lipid peroxidation, 8-Isoprostane, GSH, GSSG                                         | 85         |
|                                 | Diabetes-induced cognitive impairment (mouse)                        | Rat BM-MSC and exosomes                                                  | Cognitive function Hippocampus (CA1): No change in neuronal numbers, Exosomes colocalized with astrocytes and can be detected in microglia and neurons | Lipid peroxidation                                                                 | 26         |
|                                 | Diabetic retinopathy (mouse)                                         | Mouse AT-MSCs Intravitreal injection                                     | Retinal ganglion cell loss, NFG, bFGF and GDNF, TSP1                                     | ROS and lipid peroxidation                                                           | 28         |
|                                 | STZ-induced diabetic osteoarthritis (mouse)                          | Mouse AT-MSCs                                                            | Chondrocytes, TNF, INF-β                                                              | Lipid peroxidation                                                                  | 128        |
|                                 | STZ-induced diabetic nephropathy (rat)                               | Rat BM-MSCs                                                              | Urinary albumin excretion and ameliorated glomerulosclerosis                            | Lipid peroxidation, ROS, SOD activity, GLUT1                                        | 29         |
TABLE 1 (Continued)

| Application | Model | MSCs used | Effects of MSC treatment | Antioxidant mechanisms | References |
|-------------|-------|-----------|--------------------------|------------------------|------------|
| STZ-induced sensorial diabetic Neuropathy (mouse) | Mouse BM-MSCs | Improved pain-like behaviors | Lipid peroxidation | 40 |
| STZ-nicotinamide (diabetes)-induced cardiac damage (rat) | Rat BM-MSCs | Normalization of gene expression associated with cardiac glucose and fatty acid uptake ([RGS-1, GLUT4, PPARα, PGC-1, CPT1a and SREBP-1c]) | Total antioxidant content in serum | 56 |
| Caecal ligation-puncture induced sepsis (rat) | Rat AT-MSCs | Apoptosis in serum-starved MSCs; TNFα, NFκB and Bax in lungs and kidney | Protein oxidation in kidney | 92 |
| DSS-induced colitis (mouse) | Mouse BM-MSCs | Mucosal permeability: D-lactic acid and Diamine oxidase | Lipid peroxidation | 65 |
| E. coli-induced Acute lung injury (mouse) | Mouse BM-MSCs | Edema | MPO activity | 37 |
| Endotoxin-induced inflammation in plasma (mouse) | Mouse BM-MSCs | Cys but ND to lung fibroblast | GSH, ND to glutathione disulfide (GSSG) or cysteine (CySS) | 51 |
| Freund’s adjuvant-induced arthritis (rat) | Rat BM-MSCs | Antinuclear antibodies (TNFα, IL-9 and IL-4) | Total antioxidant capacity | 66 |
| HOCl-induced systemic sclerosis (mouse) | Mouse BM-MSCs | Serum: Systemic sclerosis biomarker (SCL-70) Skin and lung: Collagen, α SMA, TGFβ1 | Advanced oxidation protein products | 57 |
| IL-10 –/− model of colitis (mouse) | Human BM-MSCs | TNFα, IFNγ, IL-4 and p-NFκB | Total antioxidant capacity | 34 |
| Immune complex-mediated dermal vasculitis (mouse) | Human AT-MSCs | Neutrophil accumulation | Effects dependent on SOD3 expression by MSCs | 79 |
| LPS-induced lung injury (rat) | Rat BM-MSCs | Lung edema Bronchoalveolar lavage protein | MPO, Lipid peroxidation, FASL | 107 |
| Sepsis-induced brain injury (rat) | Rat AT-MSCs | Apoptosis | Protein oxidation | 25 |
| Severe acute pancreatitis (rat) | Human BM-MSCs | Serum amylase and lipase Pancreatic damage | MSCs migrated to tissue stimulated: Lipid peroxidation | 67 |
| Severe acute pancreatitis (rat) | Rat BM-MSCs | Pancreatitis score | HO-1 | 30 |
| Kidney and bladder disease ischemia (rat) | Human amniotic fluid-derived MSCs | Bladder overactivity | DNA oxidation | 129 |
| Metabolic renovascular disease in swine | Swine AT-MSC extracellular vesicles | Capillary density | Lipid peroxidation | 31 |
TABLE 1 (Continued)

| Application                        | Model                  | MSCs used                | Effects of MSC treatment                                      | Antioxidant mechanisms                           | References |
|------------------------------------|------------------------|--------------------------|----------------------------------------------------------------|-------------------------------------------------|------------|
| Unilateral ureteral obstruction (rat) | Human UC-MSC-CM        | Renal tubular damage     | ROS                                                            | Lipid peroxidation                              | 24         |
|                                    |                        | Fibrosis                 |                                                                | GSH                                             |            |
|                                    |                        | Apoptosis                |                                                                |                                                 |            |
|                                    |                        | Cell proliferation       |                                                                |                                                 |            |
| Liver disease                      | Acetaminophen-induced acute liver failure (mouse) | Human UC-MSCs           | • MSC pretreatment and post-treatment of induced liver injury | GSH                                             | 38         |
|                                    |                        |                          | • Survival and liver weight                                   | SOD activity                                    |            |
|                                    |                        |                          | • Biomarkers of liver failure                                  | Lipid peroxidation only observed with pretreatment |            |
|                                    |                        |                          | • Apoptotic cells and necrotic tissue                          |                                                 |            |
|                                    |                        |                          | • IL-6 only observed with pretreatment                         |                                                 |            |
| Liver disease                      | CCl4-induced liver fibrosis (mouse) | Human BM-MSCs           | • Serum albumin                                                | Lipid peroxidation                              | 68         |
| Liver disease                      | CCl4-induced liver injury (mouse) | Allogeneic BM-MSC        | • Serum ALT and AST                                            | SOD activity, CAT and GSH                       |            |
| Liver disease                      | N-diethylthiolsalmine-induced hepatocarcinoma (rat) | Rat BM-MSCs             | • Expression of TNFα, IL-6, type 1 collagen and αSMA           | MPO                                             | 50         |
| Liver disease                      | CC14-induced rat liver injury (mouse) | Human BM-MSCs (cells and exosomes) | • MSCs outperformed hematopoietic stem cells in all assays | Lipid peroxidation                              |            |
| Placental insufficiency            | Human BM-MSC           | Serum ALT and AST        | GSH                                                            |                                                 | 89         |
|                                    |                        | Liver fibrosis           | Lipid peroxidation                                              |                                                 |            |
| Liver disease                      | Human BM-MSCs (cells and exosomes) | Human BM-MSCs (cells and exosomes) | • Fibrosis                                                     | Lipid peroxidation                              | 130        |
|                                    |                        |                          | • Pro-inflammatory cytokines                                   |                                                 |            |
|                                    |                        |                          | • Wnt signaling                                                 |                                                 |            |
| Lung diseases                      | Cigarette smoke-induced chronic obstructive pulmonary disease (guinea pig) | Guinea pig AT-MSCs IV and intratracheal delivery | • No effect on emphysema score                                | Thiol after IV administration                   | 131        |
|                                    | Mustard lung (human case study) | Human AT-MSCs           | • Functional respiratory improvement                            | Lipid peroxidation after IV and intratracheal delivery |            |
|                                    | Ovaebumin and aluminum hydroxide-induced asthma (mouse) | Human BM-MSC            | • Functional recovery                                          | GSH in sputum                                   | 49         |
|                                    |                        |                          | • Mucin                                                         | Nitrotyrosine                                   | 132        |
|                                    |                        |                          | • Collagen                                                      |                                                 |            |
|                                    |                        |                          |                                                                |                                                 |            |
| Lung diseases                      | APP/PS1 transgenic model of Alzheimer's disease (mouse) | Rat AT-MSCs             | Recognition in behavioral test                                 | Hippocampal GSSG/GSH                            | 32         |
|                                    | Chronic ethanol intake (rats) | Human AT-MSCs           | • AT-MSCs activated by TNFα and IFNγ                           |                                                 | 86         |
|                                    |                          |                          | • ETOH intake                                                  | Hippocampal GSSG/GSH                            |            |
|                                    |                          |                          | • Relapse after ETOH deprivation                               |                                                 |            |
|                                    | Collagenase induced-intracerebral hemorrhage (rat) | Rat BM-MSCs             | Apoptosis                                                      | INOS                                            | 39         |
|                                    |                          |                          | Edema                                                          |                                                 |            |
|                                    |                          |                          | Blood-brain barrier permeability                               | ODO-1                                           |            |
|                                    |                          |                          | Pro-inflammatory cytokines                                     | MPO                                             |            |
|                                    | Pilocarpine induction of temporal lobe epilepsy (rat) | Rat BM-MSCs             | • Caspase 3                                                    | GSH                                             | 90         |
|                                    |                          |                          | Glutamate [GABA] TNFα                                          | Lipid peroxidation                              |            |
|                                    |                          |                          | IL-1β                                                          | Paraoxonase-1                                   |            |
|                                    | Spontaneous stroke (rat) | Rat BM-MSCs             | • Bel-2 expression                                            | O2−                                             | 35         |
|                                    |                          |                          | Prevented hippocampal lesions                                  | Lipid peroxidation                              |            |
|                                    |                          |                          | • Apoptosis                                                    |                                                 |            |
|                                    | Tg2576 mice (Alzheimer’s disease) | Human UC-MSCs           | Improved cognitive function                                   | Lipid peroxidation                              | 43         |
|                                    |                          |                          | Effects on hippocampus                                         | eNOS [total NO]                                 |            |
|                                    |                          |                          | • No change in β-amyloid levels                                |                                                 |            |
|                                    |                          |                          | • Neurogenesis                                                 |                                                 |            |
|                                    | YG8 transgenic model of Friedreich’s ataxia (mouse) | Mouse BM-MSCs           | Improved performance on behavioral tests                      | SOD2 and SOD3 [CAT and Gpx1]                  | 81         |
|                                    |                          |                          | • BDNF, NT3, and NT4 in dorsal root ganglia (DRG)              |                                                 |            |
|                                    |                          |                          | • GFAP, TuJ1, and MAP2 in DRG                                  |                                                 |            |
|                                    |                          |                          | • Bcl-2                                                        |                                                 |            |
| Oxygen tension injuries            | Acute ischemic stroke (rat) | Swine AT-MSCs            | Infarct area                                                   | NOX1 and NOX2                                   | 102        |
|                                    |                          |                          | Inflammatory cytokines                                         | Protein oxidation                               |            |
|                                    |                          |                          | • c-caspase 3                                                  |                                                 |            |
|                                    |                          |                          | • c-PARP                                                       |                                                 |            |
|                                    |                          |                          | • γ-H2AX                                                       |                                                 |            |
|                                    |                          |                          | • cytosolic cytochrome c                                       |                                                 |            |
| Oxygen tension injuries            | Acute myocardial infarction (swine) | Swine BM-MSCs Autologous | • Bax, c-caspase 3 and c-PARP                                  | Oxidized protein                                | 103        |
|                                    |                          |                          | Inflammation                                                  | NOX1 and NOX2                                   |            |
|                                    |                          |                          | Infarct area                                                  | Protein oxidation                               |            |
|                                    |                          |                          | • Improved echocardiography parameters                       |                                                 |            |
Exposure of MSCs to hypoxia during in vitro culture can enhance their anti-inflammatory, antioxidative, and cytoprotective properties. This suggests that the ability of MSCs to tolerate and respond to oxidative environment may be critical to their engraftment and therapeutic efficacy at sites of tissue injury.
TABLE 1 (Continued)

| Application          | Model            | MSCs used                  | Effects of MSC treatment                                                                 | Antioxidant mechanisms                     | References |
|----------------------|------------------|----------------------------|------------------------------------------------------------------------------------------|--------------------------------------------|------------|
| Traumatic injuries   | Spinal cord injury (canine) | Canine AT-MSCs            | Motor function                                                                           | Lipid peroxidation and protein oxidation   | 42         |
|                      |                  |                            | [Hemorrhagic area]                                                                       |                                             |            |
|                      |                  |                            | [Microglia]                                                                              |                                             |            |
|                      |                  |                            | [TNFα, IL-6 and COX2]                                                                    |                                             |            |
| Testicular torsion   | Testicular torsion (rat) | Rat AT-MSCs               | Apoptosis                                                                                | Lipid peroxidation                         | 135        |
| injuries             |                  |                            |                                                                                         |                                             |            |

Abbreviations: ALT, alanine aminotransferase; APAF-1, apoptotic protease activating factor 1; AST, aspartate aminotransferase; AT-MSC, adipose tissue-derived MSC; BAX, Bcl-2-associated X protein; Bcl-2, B-cell lymphoma 2; BDNF, brain-derived neurotrophic factor; bFGF, basic fibroblast growth factor; BM-MSCs, bone marrow-derived MSC; CAT, catalase; c-caspase 3, cleaved-caspase 3; CM, conditioned medium; CO, carbon monoxide; COX2, cyclooxygenase-2; c-PARP, cleaved poly (ADP-ribose) polymerase; Cys, cysteine; DRG, dorsal root ganglion; DSS, dextran sulfate sodium; EGF, epidermal growth factor; ER stress, endoplasmic reticulum stress; GABA, gamma-aminobutyric acid; GDNF, glial cell-derived neurotrophic factor; GFAP, glial fibrillary acidic protein; GLUT1, glucose transporter 1; GPx, glutathione peroxidase; GSH, glutathione; H2O2, hydrogen peroxide; HO-1, heme oxygenase-1; I/R, ischemia / reperfusion; ICAM, Intercellular adhesion molecule; IFN, interferon gamma; IL, interleukin; iNOS, inducible nitric oxide synthase; IV, intravenous; LS, lipopolyaccharide; MAP2, microtubule associated protein-2; MPO, myeloperoxidase; NAD(P)H, nicotinamide adenine dinucleotide phosphate; ND, No difference; NFκB, Nuclear factor κB; NGF, Nerve growth factor; nNOS, neuronal nitric oxide synthase; NO, nitric oxide; NOX, NAD(P)H oxidase; NQO1, NAD(P)H quinone dehydrogenase 1; NT3 and 4, neurotrophin 3 and 4; O2−, superoxide; ONOO−, peroxynitrite; p38MAPK, p38 mitogen-activated protein kinases; pAkt, phosphorylated protein kinase B; PCNA, proliferating cell nuclear antigen; pGSK3, phosphorylated glycogen synthase kinase 3 beta; pJnk, phosphorylated c-Jun N-terminal kinases; pERK1/2, phosphorylated extracellular signal-regulated kinases 1/2; p38 MAPK, p38 mitogen-activated protein kinases; pAkt, phosphorylated protein kinase B; PCNA, proliferating cell nuclear antigen; pGSK3, phosphorylated glycogen synthase kinase 3 beta; pJnk, phosphorylated c-Jun N-terminal kinases; pERK1/2, phosphorylated extracellular signal-regulated kinases 1/2; RAGE, Receptor for Advanced Glycation End Products; ROS, reactive oxygen species; SOD, superoxide dismutase; TCA cycle, tricarboxylic acid cycle; TGFβ, transforming growth factor beta; TNFα, tumor necrosis factor alpha; TSG6, TNFα-stimulated gene-6; TSP1, thrombospondin 1; Tuj1, neuron-specific class III beta-tubulin; UC-MSC, umbilical cord-derived MSC; αSMA, alpha-smooth muscle actin; γGCS, gamma-glutamylcysteine synthetase; γH2AX, gamma-H2A histone family member X; Δψm, mitochondrial membrane potential; CySS, cystine (disulfide form of cysteine); GSGS, glutathione disulfide.

3 EFFECT OF MSCs ON OXIDATIVE STRESS BIOMARKERS AND ROS

In disease models, oxidative stress is typically quantified via biomarkers of oxidation to DNA and proteins or lipid peroxidation. Administration of MSCs has been demonstrated to reduce levels of one or more of these markers in a variety of animal models associated with oxidative stress (Table 1). Injection of MSCs themselves may not be critical to their antioxidant effects as administration of their conditioned medium (CM) also reduced lipid peroxidation in a model of ureteral obstruction-induced kidney injury.24 Administration of MSC-derived exosomes was also effective to rescue protein oxidation and lipid peroxidation in animal models of septic and hyperglycemic brain injury and cognitive impairment.25,26 Likewise, DNA oxidation and lipid peroxidation caused by cisplatin-induced kidney damage are alleviated by exosomes from human umbilical cord-derived MSCs (UC-MSCs); these results were confirmed in vitro with renal proximal tubular cells.27 Treatments with MSCs were also demonstrated to reduce levels of ROS in animal models of diabetic retinopathy and nephropathy, severe acute pancreatitis, ureteral obstruction-induced kidney damage, Alzheimer’s disease, and metabolic renovascular disease.24,28-32 Specifically, MSC treatments have been shown to reduce levels of H2O2 in intestinal inflammation and several organs in a model of premature aging.32,34 MSCs also reduced levels of O2− in colitis and spontaneous stroke.34,35

Typically, the reduction of oxidative stress markers by MSC treatments is associated with functional recovery and positive outcomes in animal models. The exception to this is the diversity of responses at various stages of hepatocarcinoma whereby antioxidant effects of MSCs reduce tumor burden at the early stages of disease by protecting the integrity of DNA but increase tumor progression at the late stages of the disease possibly by reducing ROS-associated cell death.36 The timing of treatments may also affect MSCs ability to attenuate oxidative stress as pretreatment with MSCs is more effective to prevent oxidative stress in septic lung injury and acute liver failure.37,38

Several studies have also investigated the therapeutic properties of MSCs on nitrosative stress which is particularly of interest in neural diseases. MSCs reduced the volatile peroxynitrite (ONOO−) in a model of intracerebral hemorrhage and nitrite levels in diabetic sensory neuropathy.39,40 In a model of radiation-induced neurological complications, intranasal delivery of MSCs reduced inducible nitric oxide synthase (iNOS) expression and oxidative stress biomarkers, which are associated with improved cognitive performance and neuronal survival.41 In a model of spinal cord injury, adipose tissue-derived MSCs (AT-MSCs) also demonstrated antioxidant activity with a reduction in lipid peroxidation and protein oxidation; however, no significant effects were observed for nitrosylation.42 Furthermore, MSCs have been associated with increased nitric oxide (NO) in a model of Alzheimer’s disease which may have been driven by preventing the loss of neuronal nitric oxide synthase (nNOS).43 Alternatively, several studies observed a reduction in inflammation-induced iNOS and nNOS expression after MSC treatment.37,44-47 Overall, while the majority of studies support antioxidative effects of MSCs, their antinitrative effects are unclear and likely to be disease and tissue specific. Furthermore, NO can be produced by nonhuman MSCs which is thought to be critical to their immunomodulatory function which may also explain these inconsistent effects on nitrosative stress.42,48
Although studies in cells and animal models unequivocally demonstrate that MSC treatments reduce levels of oxidative stress, albeit limited data exist from human studies. Nonetheless, favorable outcomes in a case study utilizing MSCs to treat the lungs of a subject previously exposed to sulfur mustard gas were attributed to the antioxidant properties of MSCs as evidenced by reduced lipid peroxidation levels in the sputum.\(^4\)

The antioxidant effects of MSC treatments are likely to be a specific property of these cells as they are more efficacious than hematopoietic stem cells and fibroblast at reducing oxidative stress in carbon tetrachloride-induced-liver injury and sepsis, respectively.\(^50,51\) Likewise, fibroblast exosomes have no effect on kidney injury-induced by ischemia and chemotherapy.\(^27,52\)

The alleviation of oxidative stress in animal models is associated with decreased pro-inflammatory cytokines and markers of cellular death highlighting the close association between these processes. The precise mechanisms of in vivo MSC treatments are difficult to determine as cell death, inflammation, and oxidative stress occur concomitantly and perpetuate each other. However, a growing body of evidence suggests that MSCs have a direct role on suppressing oxidative stress and ROS production which may mediate their antiapoptotic and anti-inflammatory effects.

### 4 ANTI-OXIDATIVE MECHANISMS OF MSCS

The potential for MSCs to attenuate oxidative injury is unequivocally demonstrated by the reduction in ROS and biomarkers of oxidative stress in many disease models. Evidence from in vitro models suggests that MSCs directly protect cells from oxidative stimuli (Table 2). This is often associated with a reduction in ROS suggesting that MSCs avert the negative effects of oxidative stress by reducing the oxidative stimuli. The antioxidative effects of MSCs often occur in a paracrine manner in vitro and the administration of MSC-conditioned medium (CM) can also reduce oxidative stress in vivo suggesting a paracrine component to their mechanism.\(^24\) Nevertheless, others report that the antioxidant effects of MSCs can be cell contact dependent\(^53;\) albeit, these mechanisms could be disease and tissue dependent. Currently, MSCs have been proposed to reduce oxidative injury via scavenging free radicals, enhancing host antioxidant defenses, modulating the inflammatory response, augmenting cellular respiration and mitochondrial functions, or donating their mitochondria to protect damaged cells (Figure 1).\(^37,50,51,53,54\)

#### 4.1 Antioxidant defense and scavenging

To maintain redox homeostasis and prevent excessive production of free radicals, cells rely on a complement of enzymatic antioxidants, including SODs, CAT, GPx, and small nonenzymatic antioxidants, such as GSH. After MSC treatments, the total antioxidant capacity of tissues are enhanced as observed in models of chemotherapy-induced neuropathy, hyperglycemia-induced cardiac damage, systemic sclerosis, hepatocarcinoma, and acute liver injury.\(^36,55-58\) MSCs are receptive to oxidative stimuli and exhibit all necessary machinery to efficiently process ROS.\(^18,59\) Furthermore, media conditioned by MSCs have potent antioxidant capacity indicating that MSCs actively secrete antioxidants.\(^60\) MSC-CM has more effective antioxidant properties than CM from lung fibroblasts.\(^61\) In several models of disease, MSC treatments upregulate the expression of antioxidant defense enzymes in vivo (Table 1). Therefore, the antioxidant effects of MSCs may be explained by their ability to directly scavenge free radicals and by enhancing antioxidant defenses in host tissues through upregulation of antioxidant enzymes.

Volatile O\(_2^-\) is produced during cellular respiration by the mitochondrial and NOX enzymes during tissue inflammation; O\(_2^-\) is eliminated by SOD which catalyzes its conversion to H\(_2\)O\(_2\). Antioxidant effects of MSC treatments have been associated with enhanced SOD activity or the expression of SODs in models of aging, age-related erectile dysfunction, chemotherapy-induced pulmonary fibrosis or gonadotoxicity colitis, pancreatitis, septic lung injury, arthritis, hepatic toxicity and hepatic ischemia reperfusion injury, Alzheimer’s disease, and ovarian autografts.\(^33,34,37,38,43,50,62-70\) MSCs secrete all isozymes of SOD including SOD1 and SOD2, which are archetypically not released extracellularly.\(^73,59,71,72\) Thus, it is difficult to interpret whether enhanced SOD activity and/or SOD expression is due to MSCs or host-tissue-derived SOD. In vitro studies, MSC-CM or MSCs in transwell cocultures promote SOD activity in tert-Butyl hydroperoxide or UV-exposed fibroblasts, H\(_2\)O\(_2\)-treated neural stem cells or retinal ganglion cells, and dexamethasone-induced muscle atrophy model.\(^60,73-76\) These studies suggest that MSCs enhance SOD activity in cells exposed to oxidative stimuli. Likewise, MSC-CM increases SOD1 expression in islet cells exposed to pro-inflammatory cytokines and SOD2 in tert-Butyl hydroperoxide-treated umbilical endothelial cells.\(^77,78\) In endothelial cells, increased SOD2 expression was regulated by signal transducer and activator of transcription (STAT3) signaling and knockdown of either SOD2 or STAT3 decreased the antiapoptotic effects of the MSC-CM.\(^77\) These findings suggest that MSC upregulation of SOD in host tissues may be critical to their antioxidant effects. Alternatively, MSCs stimulated with TNF-α and IFN-γ were found to secrete high levels of SOD3, which was a major contributor to the antioxidant properties of MSCs in the amelioration of NO-induced neuronal death in vitro.\(^71\) Likewise, SOD3 expression in MSCs was necessary to suppress neutrophil respiratory burst and the accumulation in immune complex-mediated dermal vasculitis.\(^79\) Similarly, silencing of SOD2 in MSCs inhibits the antioxidant properties and therapeutic efficacy of their exosomes in hepatic I/R injury in vivo and H\(_2\)O\(_2\)-treated human fetal hepatocytes which can be recovered by the addition of a SOD2 mimetic.\(^80\)

CAT and GPx are responsible for detoxifying H\(_2\)O\(_2\) by its conversion to oxygen and water. MSCs secrete CAT and upregulation of CAT expression is associated with the therapeutic properties of MSCS in models of Friedreich’s ataxia, radiation-induced aortic injury, septic...
the antioxidant effects of MSCs in several other models associate with upregulation of HO-1 such as radiation-induced aortic injury, septic lung injury, pancreatitis, and renal injuries caused by altered oxygen tensions and cisplatin.\textsuperscript{30,62,92-94} HO-1 was determined to partially contribute to the effects of MSCs in pancreatitis as its inhibition with zinc protoporphyrin negated some of the effects of MSC treatments, including upregulation of CAT and increased SOD activity, which may indicate that these processes are downstream of HO-1 activity.\textsuperscript{30} After MSC treatment of small bowel I/R injury, a larger number of HO-1 expressing cells were observed which did not appear to completely colocalize with engrafted MSCs suggesting that treatments may increase HO-1 expression in cells of the host.\textsuperscript{84} It has been demonstrated that overexpression of HO-1 in MSCs enhances their therapeutic activity in septic lung injury which was attributed to its pro-survival properties.\textsuperscript{95} Others have reported that MSCs still respond efficiently to oxidative insult with silenced HO-1 by upregulating GSH pathway enzymes.\textsuperscript{96} Nevertheless, the CM of MSCs derived from HO-1\textsuperscript{−/−} mice are unable to attenuate cisplatin-induced renal injury and therefore HO-1 appears to have an important role in the antioxidant properties of the MSC secretome.\textsuperscript{94}

Together, these studies demonstrate that antioxidants secreted by MSCs and their ability to upregulate host antioxidant defenses contribute to the suppression of oxidative stress. The exosomes derived from MSCs appear to be particularly rich in machinery to process ROS and can include, but not limited to, GPx, GSTs, SOD1-3, peroxiredoxin 1-6, CAT, cytoglobin, prostaglandin-endoperoxide synthase 1, peroxidioxidin, albumin, apolipoprotein E, glutathione-disulphide reductase, and thioredoxin reductase 1-2.\textsuperscript{31,80} Notwithstanding, recombinant application of factors secreted by MSCs, such as hepatocyte growth factor (HGF) and basic fibroblast growth factors (bFGF), has been demonstrated to upregulate Gpx1, CAT, and SOD activity via SIRT1 and FOXO1 during age-related loss of ovarian function.\textsuperscript{97} Mechanisms of antioxidant defense mediated by scavenging of ROS by MSCs and the host could occur independently or simultaneously and are likely to be disease-specific.

### 4.2 Antioxidant effects on inflammation

Immune function is regulated by free radicals and the redox system; leukocytes and pro-inflammatory mediators enhance the formation of free radicals and perturb the redox environment creating a positive feedback cycle.\textsuperscript{98} The immunomodulatory action of MSCs is a well-documented phenomenon; however, their role in the interactions between the immune system and oxidative stress is not fully understood. Oxidative and/or nitrosative free radicals unequivocally play a role in all grades of acute and chronic inflammation. At physiological levels, they act as cellular signals modifying function and initiating necessary cell death programs. However, excessive generation of free radicals and/or inadequate scavenging results in protein oxidation, lipid peroxidation, and DNA damage that can be detrimental both intrinsically to the cell and the surrounding microenvironment. The ROS and reactive nitrogen species involved can take many forms and...
| Cell types                              | Model                                                                 | MSCs used                              | Antioxidant and other effects of MSCs                                                                 | References |
|----------------------------------------|-----------------------------------------------------------------------|----------------------------------------|------------------------------------------------------------------------------------------------------|------------|
| Cardiomyocytes and endothelial cells   | Glucose-deprived hypoxia-reoxygenated H9c2 cardiomyocytes (rat)       | Rat BM-MSCs                            | ▪ Apoptosis                                                                                           | 121        |
|                                        |                                                                       | Direct coculture with GFP+ MSCs        | ▪ Bax                                                                                                 |            |
|                                        |                                                                       |                                        | ▪ Bcl-2                                                                                               |            |
|                                        |                                                                       |                                        | ▪ Caspase 3                                                                                           |            |
|                                        |                                                                       |                                        | ▪ Δψm                                                                                                 |            |
|                                        |                                                                       |                                        | ▪ MSCs transferred mitochondria to H9c2 via TNT structures                                            |            |
|                                        |                                                                       |                                        | ▪ Inhibition of TNT formation partially reversed these effects                                         |            |
|                                        | H2O2-treated RL14 cardiomyocytes and human umbilical vein endothelial cells (HUVEC) | Human AT-MSCs                          | ▪ MSCs engulf mitochondria from H2O2-treated cells                                                  | 53         |
|                                        |                                                                       |                                        | ▪ MSC coculture prevented cell death—no paracrine effect                                             |            |
|                                        |                                                                       |                                        | ▪ MSCs donate functional mitochondria to somatic cells exposed to H2O2                               |            |
|                                        |                                                                       |                                        | ▪ MSCs degrade engulfed mitochondria via autophagosomes                                              |            |
|                                        |                                                                       |                                        | ▪ MSCs do not prevent somatic cell death when mitophagy is inhibited                                 |            |
|                                        |                                                                       |                                        | ▪ Mitochondria sensing by MSCs HO-1 in MSC                                                          |            |
|                                        |                                                                       |                                        | ▪ HO-1 stimulated mitochondrial biogenesis in MSC which was necessary to prevent somatic cell death  |            |
|                                        |                                                                       |                                        | ▪ Doxorubicin caused increased mitochondrial O2•− production and MSCs protected cells via similar mechanism dependent on ROS generation and transfer of mitochondria from somatic cells |            |
|                                        | I/R of ventricular myocytes (mouse) in vitro                         | Mouse BM-MSC-CM                        | ▪ Cell loss                                                                                          | 54         |
|                                        |                                                                       |                                        | ▪ Early afterdepolarization of myocytes                                                             |            |
|                                        |                                                                       |                                        | ▪ Excessive depolarization of Δψm after reperfusion                                                |            |
|                                        |                                                                       |                                        | ▪ Exaggerated hyperpolarization of Δψm after acute reperfusion—effect prevented by PI3K, Akt, and I<sub>iATP</sub> inhibition |            |
|                                        |                                                                       |                                        | ▪ I<sub>iATP</sub> opener mimicked effects:                                                         |            |
|                                        |                                                                       |                                        | ▪ Δψm hyperpolarization                                                                             |            |
|                                        |                                                                       |                                        | ▪ Mitochondrial O2•−                                                                                 |            |
|                                        |                                                                       |                                        | ▪ ROS scavenger mimicked effects: ▪ cell loss, ▪ early after depolarizations, ▪ Δψm hyperpolarization, ▪ O2•− |            |
|                                        | Oxygen glucose deprivation and reoxygenation of human umbilical vein endothelial cells (HUVEC) | Human BM-MSCs                          | ▪ MSCs and HUVEC cells form tunneling nanotubes during oxygen glucose deprivation and reoxygenation | 136        |
|                                        |                                                                       |                                        | ▪ Exchange of mitochondria in HUVECs and MSCs confirmed by mtDNA and fluorescent dye                |            |
|                                        |                                                                       |                                        | ▪ Cell death                                                                                         |            |
|                                        |                                                                       |                                        | ▪ Oxygen consumption rate and                                                                       |            |
|                                        |                                                                       |                                        | ▪ extracellular acidification rate                                                                  |            |
|                                        |                                                                       |                                        | ▪ No effect by mitochondria-depleted MSCs                                                            |            |
|                                        | Cytarabine-treated human umbilical cord vein endothelial cells (HUVEC) | Human BM-MSCs                          | ▪ Tunneling nanotubes facilitate bidirectional mitochondria transfer between MSCs and endothelial cells | 122        |
|                                        |                                                                       |                                        | ▪ Unidirectional mitochondria donation to endothelial cells pretreated with cytarabine              |            |
|                                        |                                                                       |                                        | ▪ Apoptosis                                                                                          |            |
|                                        |                                                                       |                                        | ▪ Capillary formation                                                                               |            |
|                                        | tert-Butyl hydroperoxide-treated umbilical endothelial cells (human)  | Human placental MSC-CM                 | ▪ ROS                                                                                               | 77         |
|                                        |                                                                       |                                        | ▪ Apoptosis                                                                                          |            |
|                                        |                                                                       |                                        | ▪ No effect on SOD1, CAT and GPx1 mRNA                                                               |            |
|                                        |                                                                       |                                        | ▪ SOD2 mRNA and protein                                                                             |            |
|                                        |                                                                       |                                        | ▪ SOD2 expression correlated with IL-6-ST (gp130)-STAT3 signaling                                     |            |
|                                        |                                                                       |                                        | ▪ SOD2 and STAT3 siRNA in endothelial cells reduced protective effects of MSC-CM                      |            |

(Continues)
| Cell types                            | Model                                                                 | MSCs used                    | Antioxidant and other effects of MSCs                                                                 | References |
|--------------------------------------|-----------------------------------------------------------------------|------------------------------|-------------------------------------------------------------------------------------------------------|------------|
| Fibroblasts                          | tert-Butyl hydroperoxide-treated human dermal fibroblasts             | Human AT-MSC-CM              | ↑ Antioxidant capacity over normal culture media<br>↑ Morphological damage<br>↑ SOD activity in human dermal fibroblasts<br>↑ GPx activity in human dermal fibroblasts | 60         |
| UV-exposed fibroblasts (human)       |                                                                       | Human UC-MSC-CM              | ↑ Cell viability<br>↑ SOD activity<br>↑ ROS activity<br>↑ SOD activity<br>↑ ROS activity<br>↑ Antioxidant capacity | 73         |
| Gial cells and neurons               | Activated microglia and NO-induced neuronal death (rat)               | Human BM-MSCs                | ↑ Neuronal loss from activated microglia<br>↑ Neuronal loss from NO<br>↑ ROS<br>↑ SOD activity<br>↑ FIR activity<br>↑ BDNF activity | 71         |
|                                     | Amyloid-β oligomer-induced damage to hippocampal neurons (rat)        | Rat BM-MSCs and Transwell coculture and exosomes | ↑ ROS<br>↑ Neuronal loss from NO<br>↑ ROS activity<br>↑ BDNF activity<br>↑ CNTF activity<br>↑ SOD activity<br>↑ FIR activity<br>↑ BDNF activity<br>↑ CNTF activity | 83         |
| Glucose-deprived hypoxia-reoxygenated primary astrocytes (human) | Human dental pulp-derived and BM-MSCs and Transwell and CM astrocytes | ↑ Viability of astrocytes<br>↑ ROS<br>↑ SOD activity<br>↑ FIR activity<br>↑ BDNF activity<br>↑ CNTF activity | 137        |
| Glucose-deprived scratch injured T98G glioblastoma cells (human) | Human AT-MSC-CM                                                      | ↑ Viability<br>↑ ROS<br>↑ SOD activity<br>↑ FIR activity<br>↑ BDNF activity<br>↑ CNTF activity | 138        |
| H$_2$O$_2$-treated cortex-derived neural stem cells (rat) | Rat BM-MSC-CM                                                       | ↑ Apoptosis<br>↑ ROS<br>↑ SOD activity<br>↑ FIR activity<br>↑ BDNF activity<br>↑ CNTF activity | 74         |
| H$_2$O$_2$-treated motor neurons (NSC-34) expressing human mutant SOD1 (ALS) | Mouse AT-MSC exosomes                                           | ↑ Cell viability of naïve cells and SOD1 mutant cells<br>↑ Viability<br>↑ ROS<br>↑ SOD activity<br>↑ FIR activity<br>↑ BDNF activity<br>↑ CNTF activity | 139        |
| H$_2$O$_2$-treated retinal ganglion cells (RGCs) (rat) | Rat BM-MSCs and Transwell                                        | ↑ Apoptosis<br>↑ ROS<br>↑ SOD activity<br>↑ FIR activity<br>↑ BDNF activity<br>↑ CNTF activity | 75         |
| H$_2$O$_2$-treated SH-SY5Y neuroblastoma cells (human) | Human AT-MSC CM                                                    | ↑ Viability<br>↑ Antioxidant capacity<br>↑ ROS<br>↑ SOD activity<br>↑ FIR activity<br>↑ BDNF activity<br>↑ CNTF activity | 140        |
| Sevoflurane-induced apoptosis in human neuroglioma H4 cells | Rat BM-MSCs                                                        | ↑ Cell viability<br>↑ ROS<br>↑ c-caspase 3 and Bax<br>↑ ATP<br>↑ SOD activity<br>↑ FIR activity<br>↑ BDNF activity<br>↑ CNTF activity | 141        |
| Cell types          | Model                                                                 | MSCs used                          | Antioxidant and other effects of MSCs                                                                 | References |
|---------------------|-----------------------------------------------------------------------|------------------------------------|--------------------------------------------------------------------------------------------------------|------------|
| Hepatocytes         | Acetaminophen and H2O2-treated human hepatocytes (HepG2)              | Rat BM-MSC-CM                      | • Exosome-rich fractioned conditioned medium  
  |                     |                                                                       |                                    | Cell viability  
  |                     |                                                                       |                                    | ROS          | 142        |
|                     | H2O2-treated AML12 hepatocytes (murine)                               | Mouse BM-MSC extracellular vesicles| • ROS  
  |                     |                                                                       |                                    | Pro-inflammatory cytokines | 143        |
|                     | H2O2-treated human fetal hepatocytes (LO2 cells)                      | Human UC-MSC extracellular vesicles| • ROS  
  |                     |                                                                       |                                    | Mitochondrial O2⁻  
  |                     |                                                                       |                                    | Apoptosis    | 80         |
|                     |                                                                       |                                    | • Exosomes contain PRDX1-6, SOD1-2, CAT, TXN GSTO and GSTP1  
  |                     |                                                                       |                                    | Silencing of SOD2 in MSCs inhibits therapeutic effect of exosomes |           |
| Immune cells        | Cytarabine or methotrexate-treated immortalized human T lymphocytes (Jurkat cells) | Human BM-MSCs | • Jurkat cells transfer mitochondria to MSCs after exposure to chemotherapeutics. Few mitochondria transferred from MSCs to Jurkat cells  
  |                     |                                                                       |                                    | MSC direct coculture  
  |                     |                                                                       |                                    | Apoptosis    | 120        |
|                     |                                                                       |                                    | • Effects blocked by inhibition of mitochondrial transfer using cytochalasin D and anti-ICAM1 |           |
|                     | LPS-stimulated blood-derived monocytes (human)                        | Human AT-MSCs                      | • TNFα  
  |                     |                                                                       |                                    | NOx  
  |                     |                                                                       |                                    | COX2         | 110        |
|                     |                                                                       |                                    | • MPO  
  |                     |                                                                       |                                    | ROS          | 114        |
|                     | LPS-treated human monocyte-derived macrophages                        | Human BM-MSC CM and extracellular vesicles | • oxygen consumption rate  
  |                     |                                                                       |                                    | phagocytic phenotype  
  |                     |                                                                       |                                    | • This effect was partially reversed by Ab blocking extracellular vesicles (anti-CD44) |           |
|                     |                                                                       |                                    | • Extracellular vesicles from MSCs transfer mitochondria to macrophages |           |
|                     |                                                                       |                                    | • MSC-CM M2 phenotype (anti-inflammatory) |           |
|                     |                                                                       |                                    | • Effects abolished by damaging mitochondria in MSCs |           |
|                     | LPS-treated neutrophils (human)                                       | Human UC-MSCs Transwell and extracellular vesicles | • Lipid peroxidation  
  |                     |                                                                       |                                    | ROS          | 80         |
|                     |                                                                       |                                    | • No effect on cell numbers in vitro |           |
|                     | Macrophages in vitro (human and mouse)                               | Human BM-MSCs                      | • ROS-associated with NRLP3 inflammasome activation  
  |                     |                                                                       |                                    | NRLP3 associated caspase 1 activation  
  |                     |                                                                       |                                    | NRLP3 associated IL-1β and IL-18 secretion | 111        |
|                     |                                                                       |                                    | • TNFα and IL-6 transcription  
  |                     |                                                                       |                                    | • Effects inhibited by STC-1 siRNA |           |
|                     | PMA-activated neutrophils (mouse and human)                           | Human AT-MSCs                      | • Respiratory burst (ROS) dependent on SOD3 expression by MSCs  
  |                     |                                                                       |                                    | Apoptosis    | 79         |
|                     |                                                                       |                                    | • MPO protein and activity |           |

(Continues)
| Cell types | Model | MSCs used | Antioxidant and other effects of MSCs | References |
|------------|-------|-----------|--------------------------------------|------------|
| Islet cells | Cytokine cocktail-exposed islet cells (rat) | Human BM-MSCs | IL-1, TNFα and IFNγ cocktail, Insulin secretion, SOD1, NQO1, HO-1, Ferritin H | 78 |
| Hypoxia (1% O₂) exposed porcine islet cells | Human UC-MSC CM and exosomes | Apoptosis, ROS, Mitochondrial O₂⁻, GSH, GPx activity, Inhibition of ERK pathway reversed effects, MSCs secreted high levels of IL-6, MSC exosomes and recombinant IL-6 | 144 |
| Hypoxia-exposed neonatal porcine islet cell clusters (porcine) | Human UC-MSC CM and exosomes | Apoptosis, Oxygen consumption rate, Effects reduced after clearance of exosomes in conditioned media | 113 |
| Normoxia- and hypoxia-exposed WJ-MSC engineered islet-like cells (human) | Human WJ-MSCs | Normoxia (21% O₂) and hypoxia (2% O₂) WJ-MSCS formed monolayer while islet-like cells were free floating, Proliferation | 145 |
| Primary islet cells (mouse) | Mouse BM-MSCs Transwell coculture | | | 91 |
| Keratinocytes | High glucose and LPS-treated primary keratinocytes (rat) | Rat BM-MSC-CM | Viability, Wound assay closure, ROS, Dependent on ERK signaling | 146 |
| Lung epithelial cells | H₂O₂-treated human alveolar basal epithelial adenocarcinoma cells (A549) | Human BM-MSCs | Cell viability, Transcription and protein expression of STC-1 in H₂O₂-treated MSCs, Cell viability with Anti-STC-1, Cell viability with recombinant STC-1, Similar results in H1299 and PC9, Cell viability with STC-1 siRNA MSCs, ROS with STC-1 siRNA MSCs, mRNA expression of uncoupling protein 2 in A549, mRNA expression of uncoupling protein 2 with anti-STC-1 | 100 |
| Osteocytes | Mitochondrial DNA (mtDNA)-depleted 143B osteosarcoma cells (human) | Human WJ-MSCs | MSCs in direct co culture donated mitochondria, MSCs and mitochondria-depleted cells removed via auxotrophic restriction, Recovered cellular respiration (oxidative phosphorylation), Restoration of cellular proliferation and motility, Effects of mitochondria donation sustained for 45 passages | 124 |
| Renal cells | Cisplatin-treated renal proximal tubular cells (rat) | Human UC-MSC exosomes | ΔΨm, PCNA, Oxidized DNA, Lipid peroxidation, GSH, Bax, Bcl-2 | 27 |
| Cell types | Model | MSCs used | Antioxidant and other effects of MSCs | References |
|------------|-------|-----------|--------------------------------------|------------|
| H₂O₂-treated renal tubular epithelial cells (rat) in vitro | Rat BM-MSCs | Apoptosis | 93 |
| | | Cell loss | |
| | | Mitosis | |
| | | Mitox | |
| | | Bax expression | |
| | | p-ERK1/2 | |
| High glucose-treated glomerular mesangial cells (rat) | Rat BM-MSC-CM | ROS | 29 |
| | | GLUT1 | |
| | | Inhibition of HGF via antibody blocking inhibited antioxidant effect | |
| Hypoxia reoxygenation of rat kidney epithelial cells (NRK-52E) | Human WJ-MSC extracellular vesicles | ROS | 52 |
| | | Activated NRF2 | |
| | | ARE activity | |
| | | HO-1 | |
| Oxalate and calcium oxalate monohydrate-treated human proximal tubular epithelial (HK-2) | Human UC-MSC exosomes | Apoptosis | 147 |
| Skeletal muscle cells | Dexamethasone-induced muscle atrophy in L6 rat skeletal muscle cells | Human UC-MSC-CM | Muscle related gene expression (myogenin, desmin) | 76 |
| | | SOD activity | |
| | | ROS generation | |
| | | CAT, SOD1, GPx-1 in L6 cells | |
| Dexamethasone-induced muscle atrophy in L6 rat skeletal muscle cells | Human UC-MSC (isolated mitochondria) Centrifugal delivery of exogenous mitochondria | Cell proliferation | 125 |
| | | ATP content | |
| | | Mitochondrial O₂⁻ | |
| Trophoblasts | Hypoxia (1% O₂) trophoblast cells (mouse) | Mouse BM-MSCs Transwell | Mitofusin-2 | 148 |
| | | β-HCG and progesterone | |
| | | ATP levels | |
| | | Caspase 3 and 9 | |
| | | Bax, Bcl-2 | |
| | | Apoptosis | |

Abbreviations: ARE, antioxidant response element; AT-MSC, adipose tissue-derived MSC; BAX, Bcl-2-associated X protein; BDNF, brain-derived neurotrophic factor; BM-MSCs, bone marrow-derived MSC; c-caspase 3, cleaved-caspase 3; CM, conditioned medium; CNTF, ciliary neurotrophic factor; COX2, cyclooxygenase-2; ERK, extracellular signal-regulated kinases; GLUT1, glucose transporter 1; GPx, glutathione peroxidase; GST, glutathione S-transferase; H₂O₂, hydrogen peroxide; HGF, hepatocyte growth factor; HO-1, heme oxygenase-1; I/R, ischemia/reperfusion; ICAM, intercellular adhesion molecule; IFNγ, interferon gamma; IL, interleukin; LDH, lactate dehydrogenase; LPS, lipopolysaccharide; M2, type-2 macrophages; MPO, myeloperoxidase; NAC, N-acetylcysteine; NAD(P)H, nicotinamide adenine dinucleotide phosphate hydrogen; NO, Nitric oxide; NQO1, NAD(P)H quinone dehydrogenase 1; NRF2, nuclear factor erythroid 2-related factor 2; NLRP3, nod-like receptor protein-3; O₂⁻, superoxide; pERK1/2, phosphorylated extracellular signal-regulated kinases 1/2; PRDX1-6, peroxiredoxin; ROS, reactive oxygen species; SOD, superoxide dismutase; STAT3, signal transducer and activator of transcription 3; STC-1, stanniocalcin-1; TNFα, tumor necrosis factor alpha; TNT, tunneling nanotube; TXN, thioredoxin; UC-MSC, umbilical cord-derived MSC; β-HCG, β-human chorionic gonadotropin; Δψm, mitochondrial membrane potential.
be generated from a variety of sources. Large amounts of the highly reactive $\text{O}_2^-$ anion are generated from NOX expressed by innate leukocytes.\textsuperscript{99} SOD catalyzes the conversion of $\text{O}_2^-$ to $\text{H}_2\text{O}_2$ which phagocytes and neutrophils use to generate hypochlorous acid (HOCl) via myeloperoxidase (MPO).\textsuperscript{98} Collectively, this is referred to as respiratory burst, a crucial element of the bactericidal response and inflammatory signaling. Nonetheless, MPO activity and $\text{O}_2^-$ are also associated with various inflammatory diseases. In inflammatory bouts, there is often a parallel increase in the expression of iNOS in leukocytes and, thus, subsequent generation of the free radical NO. Nonimmune cells such as epithelial cells are also capable of expressing iNOS and NOX to generate NO and $\text{O}_2^-$.

MSC treatments reduce inflammation and oxidative stress in colitis, pancreatitis, arthritis, sepsis, vasculitis, stroke, myocardial infarction, hyperoxic lung injury, and I/R injury of the kidneys and bowel (Table 1). These effects have included a reduction in inflammatory cytokines TNF$\alpha$, IFN$\gamma$, interleukin (IL)-1$\beta$, IL-6, IL-9, and IL-4; decreased expression of ROS producing enzymes NOX, MPO, and iNOS; as well as a net reduction in the infiltration of immune cells such as neutrophils (Table 1). The anti-inflammatory effects of MSCs in pancreatitis were partially dependent on their expression of the antioxidant pathway enzyme HO-1.\textsuperscript{30} Previously, it was demonstrated in a model of sepsis that MSC treatments can reduce pro-inflammatory cytokines in the serum and normalize thiol/disulfide redox pairings responsible for free radical scavenging.\textsuperscript{51} Decreased levels of IL-1$\beta$ and TNF$\alpha$ superseded restoration of redox homeostasis. This suggests that the aversion of oxidative injury was secondary to the immunomodulation of pro-inflammatory signaling, at least in acute septic inflammation. Conversely, MSCs can directly reduce oxidative injury in many cell types in vitro; thus, it is likely that MSCs may also reduce oxidative stress in tissues by mechanism other than suppressing the immune system.\textsuperscript{27,54,60,75,77,100} This is highlighted in in vivo and organotypic ex vivo models of myocardial I/R injury where MSC-derived exosomes ameliorated infarction injury without altering leukocyte recruitment.\textsuperscript{101} In in vivo experiments, protein oxidation was reduced by MSC-derived exosomes after 1 hour; neutrophils were yet to infiltrate into the tissue. After 24 hours, MSCs reduced peripheral blood leukocyte numbers and neutrophil infiltration into the myocardium; thus, the antioxidative activity of MSCs preceded signals recruiting leukocytes.\textsuperscript{101} This suggests that MSCs can attenuate oxidative stress-induced tissue injury first, which can limit the recruitment of immune cells and subsequent inflammation in this model. This may be mediated by their ability to suppress NOX1 and 2 on resident cells which are downregulated by MSC treatments in acute myocardial infarction, sepsis-induced brain injury, acute ischemic stroke, I/R injury to kidneys, and small bowel which were all associated with reduced inflammation.\textsuperscript{25,84,102-104}

Neutrophils appear to be key mediators of oxidative stress in inflammation. These cells harbor an abundance of MPO, a major catalyst for hypochlorite and NO-derived oxidants.\textsuperscript{105,106} MSCs attenuate the infiltration of neutrophils and reduce MPO levels in several disease models.\textsuperscript{50,107,108} MSCs can also directly dampen the respiratory burst in neutrophils and suppress MPO activity required to produce free radical required for their pro-inflammatory function which was dependent on SOD3 and occurs in a paracrine manner.\textsuperscript{79,80,109} Likewise, MSCs can also directly decrease ROS and MPO in stimulated monocytes and macrophages which suppress their pro-inflammatory phenotype.\textsuperscript{110,111} These data suggest that MSCs not only suppress the immune system to prevent oxidative injury, but also that their mechanism of immunosuppression is reliant on their antioxidant properties.

### 4.3 Cellular bioenergetics

Free radicals are produced by several metabolic processes and the mitochondria during cellular respiration. Dysfunction in mitochondria can cause cellular injury which is mediated through the generation of $\text{O}_2^-$ and proteins that initiate cellular apoptosis. Depolarization of the mitochondrial membrane potential ($\Delta\psi_m$) is a hallmark of mitochondrial dysfunction leading to cell death. Hyperglycemia can also cause oxidative stress via several mechanisms including the formation of free radical as by-products of glucose auto-oxidation that deplete antioxidant defense and advanced glycation end products that induce cellular stress. In models of hyperglycemia, MSC treatments can reduce the expression of glucose and fatty acid transports in kidney and cardiac tissue cells, which prevents glucose transport and ROS generation.\textsuperscript{29,56} Therefore, the antioxidant effects of MSC treatments in models of diabetes may be downstream of glycemic control. Conversely, regulation of mitochondrial function and oxidative phosphorylation by MSCs has been implicated in several disease models. MSC treatments improve chemotherapy-induced cognitive impairment, which associates with enhanced respiratory capacity of the mitochondria.\textsuperscript{112} The effects of MSCs on the mitochondria appear to occur in a paracrine manner as MSC-CM increases the oxygen consumption rate of hypoxia-exposed neonatal porcine islet cells and LPS-treated macrophages.\textsuperscript{113,114} Similarly, MSC-derived extracellular vesicles suppress mitochondrial $\text{O}_2^-$ levels in $\text{H}_2\text{O}_2$-treated human fetal hepatocytes, which is associated with a reduction in apoptosis.\textsuperscript{90}

The potential for MSCs to directly attenuate mitochondrial dysfunction has been demonstrated in an in vitro model of I/R injury in mouse ventricular myocytes.\textsuperscript{54} Within 5 minutes of reperfusion, cells exhibited an exaggerated $\Delta\psi_m$ hyperpolarization, which was reduced by conditioning the reperfusion solution with MSCs. The exaggerated hyperpolarization was followed by a continuous depolarization in controls after 15 minutes which was also attenuated by the paracrine secretion of MSCs. Decay of the $\Delta\psi_m$ was likely a result of the mitochondrial permeability transition pore opening. The exaggerated hyperpolarization of the $\Delta\psi_m$ was also averted by a mitochondrial ROS scavenger which simultaneously decreased mitochondrial $\text{O}_2^-$ generation demonstrating the close relationship between these events. Similarly, MSC secretion decreased mitochondrial $\text{O}_2^-$, which led to the suggestion that MSCs may also attenuate $\Delta\psi_m$ dysfunction via scavenging of $\text{O}_2^-$. Depolarization of the $\Delta\psi_m$ in cisplatin-treated renal proximal tubular cells has also been reportedly attenuated using exosomes.
derived from UC-MSCs. In vitro, BM-MSCs were demonstrated to upregulate uncoupling protein 2 (UCP2) transcription in H2O2-treated alveolar basal epithelial adenocarcinoma cells which reduces the formation of mitochondria-derived O2− by lowering the proton-motive force across the mitochondrial membrane and provides another potential mechanism for the alleviation of mitochondrial dysfunction. This was regulated by the paracrine secretion of stanniocalcin-1 by MSCs, which enhanced UCP2, correlating with cell survival and decreased ROS generation. MSCs secreted stanniocalcin-1 may also attenuate inflammation as it decreases mitochondrial ROS and subsequent activation of the nucleotide-binding oligomerization domain (NOD)-like receptor protein 3 (NLRP3) inflammasome. BM-MSCs inhibited the activity of the NLRP3 inflammasome in primed macrophages which is responsible for recognizing damage-associated molecular patterns and initiating the inflammatory cascade through activation and secretion of IL-1. MSCs have also been demonstrated to secrete the redox-sensitive protein DJ-1, which has established roles in maintaining mitochondrial biogenesis and respiratory chain efficiency and could potentially mediate the neuroprotective effects of the MSC secretome as shown in Parkinson's disease models. Collectively, these studies demonstrate that MSCs can ameliorate mitochondrial dysfunction in a paracrine manner with diverse therapeutic outcomes.

4.4 Mitochondrial donation

Recently, a concept has emerged that MSCs may be able to alter oxidative phosphorylation and ROS generation in cells through donation of mitochondria themselves. Islam et al. observed mitochondrial transfer from human BM-MSCs to alveolar epithelium in a mouse model of LPS-induced lung injury. BM-MSC administration attenuated decreased intracellular ATP in the alveoli caused by lung injury; notably, ATP (visualized by a molecular probe) was predominantly restored at the site of mitochondrial transfer and immediately surrounding alveoli. MSCs with a mutation in connexin 43, a protein involved in the formation of gap junctions, were unable to transfer mitochondria despite being functionally competent and subsequently did not restore ATP, surfactant secretion, or reduce leukocyte infiltration. This phenomenon only occurred in LPS-exposed lungs indicating that mitochondrial transfer is dependent on stimulants from damaged tissues.

MSCs cultured in hyperoxic (21% O2; normoxic atmosphere) conditions produce high levels of mitochondrial O2−, depolarize Δψm, and induce mitophagy. Mitochondria are loaded into phagosomes and shuttled to the plasma membrane. These effects were reduced by culturing MSCs closer to a normoxic oxygen concentration (5% O2; hypoxic atmosphere). Macrophages have been observed to phagocytose these vesicles containing the partially depolarized mitochondria, which can fuse with endogenous mitochondria in macrophages. This protects silica-exposed macrophages by increasing their oxygen consumption rate and decreasing mitochondrial O2− production. These effects could not be elicited when MSCs were substituted by human fibroblasts. This suggests that mitochondrial transfer may be stimulated by oxidative stress in MSCs. Conversely, MSCs have also been demonstrated to engulf mitochondria from other somatic cells exposed to H2O2. MSCs degraded the engulfed mitochondria which stimulated HO-1 expression, mitochondrial biogenesis in MSCs, and the transfer of functional MSCs to damaged cells. Inhibition of mitophagy negated the cytoprotective effects of MSCs in other somatic cells, which suggests that MSC sensing of damaged mitochondria may mediate their therapeutic responses. Supporting this, the cell contact-dependent transfer of mitochondria from chemotherapy-treated T lymphocytes to MSCs was also determined to be critical to their ability to decrease mitochondrial O2− production and cell death in T lymphocytes. Nonetheless, in this study, mitochondrial transfer appeared to be predominately unidirectional and very few MSC-derived mitochondria were observed in T cells. This suggests that MSC sensing of mitochondria can promote therapeutic mechanisms other than mitochondrial donation.

Although extracellular vesicles can contain whole mitochondria, several studies suggest that the donation of mitochondria may be contact-dependent. MSCs have been found to transfer mitochondria via tunneling nanotubes (TNT) to glucose-deprived and hypoxia-reoxygenated cardiomyocytes which prevented Δψm depolarization and cell apoptosis. Albeit inhibition of TNT formation only partially reversed the effects of MSCs indicating other cytoprotective mechanisms were still active. Similarly, MSCs transfer mitochondria to chemotherapy-treated endothelial cells, which appears to occur in a unidirectional manner, unlike in T lymphocytes. Miro1 is important to TNT formation and its overexpression in MSCs can enhance mitochondrial transfer. The effects of MSC mitochondrial donation are sufficient to rescue cellular respiration, proliferation, and motility in mitochondria-depleted osteosarcoma cells. These effects can be maintained for 45 passages, which highlights the therapeutic potential of MSC-derived mitochondria. Exogenous application of mitochondria isolated from MSCs may also offer therapeutic benefit and are able to protect dexamethasone-treated muscle cells form oxidative stress in vitro. Albeit, delivery of MSC-derived mitochondria in vivo poses a challenge. While several studies have demonstrated contact-dependent transfer of mitochondria between MSCs and other cells, MSCs have also been reported to donate mitochondria to LPS-treated macrophages via secreted extracellular vesicles. Exposure of macrophages to MSC-derived exosomes promoted their indication to the type 2 phenotype, which exerted anti-inflammatory effects after adoptive transfer in septic lung injury. The effects of MSCs were dependent on enhancing mitochondrial function in macrophages and were inhibited by damaging mitochondria in MSCs and blocking extracellular vesicles. The therapeutic use of MSCs to deliver functional mitochondria to damaged tissue is an intriguing concept and warrants further study; however, another recent advancement reported by Panfoli et al. suggests that the exosomes of MSCs are capable of oxidative phosphorylation independent of the mitochondria. Subsets of MSC-derived exosomes isolated from the umbilical cord of term newborns were discovered to contain complexes of the ETC embedded in the membrane. These exosomes possessed an electrochemical membrane potential, consumed O2, and produced ATP.
The therapeutic application of these exosomes is yet to be investigated; nonetheless, this may present a viable tool to restore dysfunctional oxidative phosphorylation and ATP synthesis in damaged cells.

5 | CONCLUSION

The presented studies evidently demonstrate that MSCs exhibit antioxidant potential either directly via scavenging of ROS and donating mitochondria or indirectly by upregulation antioxidant defenses in other cells and altering cellular bioenergetics. These effects can occur in combination with the previously recognized trophic and vesicular components of the MSC secretome acting directly on regenerative pathways. Likewise, antioxidant and trophic pathways appear to mediate the cytoprotective effect of MSC treatments which are ROS dependent. MSCs have frequently been utilized in inflammatory diseases to modulate the immune response. In this context, immunosuppression can avert ROS generation which is generated by MPO and NOX enzymes as a part of the inflammatory response. However, MSCs have now been shown to exert immunosuppressive effects by dampening ROS production and enhancing mitochondrial function in macrophages and neutrophils. Therefore, the antioxidant properties confer a role in the trophic and anti-inflammatory mechanisms of MSC therapy. Considering that oxidative stress is implicated in almost every disease, these antioxidant properties, along with regenerative capacity of MSC secretome, may explain why MSC treatments are useful for such a spectrum of seemingly unlinked pathologies (Figure 1). Future studies should seek to clarify disease-specific nuances of the antioxidative mechanisms of MSCs and MSC-derived products. Likewise, improving the antioxidant effects of MSCs by enhancing the expression of antioxidant enzymes or promoting mitochondrial donation may be useful to optimize MSC-based therapies and improve outcomes.

CONFLICT OF INTEREST

The authors declared no potential conflicts of interest.

AUTHOR CONTRIBUTIONS

R.S.: conception and design, manuscript writing, final approval of manuscript. K.N.: conception and design, manuscript writing, financial support, final approval of manuscript.

DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no new data were created or analyzed in this study.

ORCID

Kulmira Nurgali https://orcid.org/0000-0002-2597-6929

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Author/s:
Stavely, R; Nurgali, K

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