Severe community-acquired pneumonia caused by *Leptospira interrogans*: A case report and review of literature

Qiu-Hong Bao, Li Yu, Jian-Jun Ding, Ying-Jun Chen, Jun-Wei Wang, Jian-Ming Pang, Qi Jin

**Abstract**

**BACKGROUND**

*Leptospira* is an uncommon pathogen for adult severe community-acquired pneumonia and its nonspecific manifestations and limited diagnostic tests make it difficult to identify. Although conventional penicillin remains efficacious to treat *leptospirosis*, failure in early diagnosis and treatment can lead to progression into a deadly syndrome with multiple organ dysfunction. Next generation sequencing is of great value to understand cases with infection of unknown cause, which could help in the diagnosis of uncertain *Leptospira* infection.

**CASE SUMMARY**

We recently managed a patient with fever, cough and dyspnea on admission that progressed into persistent adult respiratory distress syndrome, hemoptysis and hematuria after admission. In this case, the rare *Leptospira* infection was clouded by the positive influenza tests at admission, delaying early *Leptospira*-targeted antibiotics administration. Next generation sequencing, a novel molecular diagnostic tool, provided a key hint to uncover the crucial pathogen, *Leptospira interrogans*, further supported by the possible occupational exposure history. Subsequent conventional penicillin and mechanical respiratory support were administered to cure the patient successfully without any sequela.

**CONCLUSION**

Clinicians must pay attention to possible exposure history and keep uncommon
A 54-year-old man presented with a 4 d history of cough with white phlegm, chills and fever as well as dyspnea for 1 d. No hemoptysis, chest pain, edema or rash were found.

History of present illness
Before this visit, he had been given 1.5 g of intravenous cefuroxime but received little improvement. According to the above examinations, the initial diagnosis was considered severe pneumonia caused by influenza virus. Thus, the patient was treated with methylprednisolone (40 mg, q12 h), oseltamivir (75 mg, bid), tienam (0.5 g, q8 h) and moxifloxacin (0.4 g qd). Unexpectedly the next morning, the patient’s condition worsened into cough with bloody sputum and shortness of breath with decreased oxygen saturation. A chest X-ray showed a right lower lobe infiltrate.

MANAGEMENT
Before this visit, the patient had been given 1.5 g of intravenous cefuroxime. However, little effectiveness was observed. There was a possibility of resistant bacteria, such as methicillin-resistant Staphylococcus aureus, which was considered for antibacterial therapy. However, his conditions worsened and oxygen saturation decreased.

Core Tip: Pneumonia caused by Leptospira interrogans is rare but life-threatening. In the early phase, the nonspecific symptoms of Leptospira pneumonia often lead clinicians to misdiagnosis as influenza or other illness. We herein report a 54-year-old man who presented with cough, fever and dyspnea and was misdiagnosed with influenza but was confirmed with Leptospira infection with the help of next generation sequencing. Possible exposure history and next generation sequencing during management of infections with unknown causes are important.
SpO₂ to 60%. Hence, the patient was managed with intubation and continuous mechanical ventilation of positive end-expiratory pressure 8 cm H₂O, FiO₂ 40% and SpO₂ 99%. Caspofungin was added to the initial therapeutic plan.

Further examinations were also performed to find the essential causes. Sputum smear test showed no positive findings. Bronchoscopy showed sporadic mucosal petechiae and bloody sputum in the bilateral bronchi (Figure 1), and bronchoalveolar lavage fluid (BALF) was collected. Echocardiography revealed left ventricular diastolic dysfunction and moderate tricuspid regurgitation with pulmonary arterial hypertension. The patient was negative for acid-fast bacilli stain, fungi test of BALF, tumor makers and a panel of respiratory viruses and autoantibodies. Under the current comprehensive treatment covering anti-infections of virus, bacteria and fungi, the patient did not improve and had persistent hemoptysis and hematuria under assisted ventilation. Inflammation and myocardial impairment-related laboratory values were elevated, and hemoglobin and platelet counts remained below normal levels (Figure 2).

The NGS report of the BALF on day 6 after admission indicated the cause. To our surprise, NGS indicated an infection of *Leptospira interrogans*. Subsequently, it was retrospectively interviewed that the patient worked as a rubbish cleaner of rivers with high exposure risk of *Leptospira* infection.

**History of past illness**
The patient reported a 30 yr history of smoking and drinking. He denied any history of hypertension, diabetes or other chronic disease.

**Personal and family history**
The patient and his families denied any history of hypertension, diabetes or other chronic disease.

**Physical examination**
On examination, his temperature was 37.5 °C, pulse was 118/min, respiratory rate was 19/min, and blood pressure was 81/54 mmHg at admission. He was in a conscious state but in compulsive position with cyanotic face. His superficial lymph nodes were not swollen. Lung examination showed bilateral rales. Other physical examinations were normal.

**Laboratory examinations**
Arterial blood gas analysis showed lower PaO₂ of 64 mmHg and SpO₂ of 94.3% with 2 L/min oxygen support. Laboratory investigations revealed decreased white blood cell count of 1.7 × 10⁹/L and platelet count of 56 × 10⁹/L, elevated C-reactive protein of 190.81 mg/L, creatinine phosphokinase of 372 U/L, creatinine of 117 μmol/L and activated partial thromboplastin ratio of 1.30 and a positive urine occult blood test. The throat swab for influenza virus antigen was positive.

**Imaging examinations**
Chest computed tomography (CT) indicated infectious lesions of both lungs and emphysema with pulmonary bullae (Figure 3A).

**FINAL DIAGNOSIS**
Severe pneumonia with *Leptospira interrogans* infection.

**TREATMENT**
After identification of the *Leptospira* pathogen, the patients finally received penicillin (400000 U, q12 h, im) for 1 wk as well as respiratory support, while previous administration of oseltamivir and caspofungin was stopped. After 1 wk, with advice of the infection department, the usage of penicillin was doubled for 2 d and then doubled again for another 1 wk.
Bao QH et al. Misdiagnosed pneumonia caused by Leptospira

Figure 1 Bronchoscopy findings. A-C: Bronchoscopy images of main trachea (A), left main bronchus (B) and opening of the right basal segmental bronchus (C) showed that the bronchi were patent, and sporadic mucosal petechiae and bloody sputum were seen in the bilateral bronchi.

Figure 2 Changes in laboratory parameters after admission. In the first 5 d of antiviral treatment after admission: inflammatory values were elevated; platelets and hemoglobin were decreased consistent with hemorrhagic complications; and elevated brain natriuretic peptide indicated the existence of myocardial impairment. After 5 d, penicillin was administered and the above vital parameters improved. WBC: White blood cell; CRP: C-reactive protein; PCT: Procalcitonin; PLT: Platelets; Hb: Hemoglobin; BNP: Brain natriuretic peptide.

OUTCOME AND FOLLOW-UP
After about 1 wk of penicillin treatment, the patient obviously got better with significantly relieved cough and hemoptysis as well as improved oxygenation status without mechanical ventilation (PaO₂ 78 mmHg, SpO₂ 95.3 under 5 L/min oxygen flow). Additionally, echocardiography of mild mitral and tricuspid regurgitation and decreased brain natriuretic peptide suggested the improvement of myocardial damage. Chest CT also showed the bilateral lung inflammation was greatly resolved (Figure 3C). After a total of 16 d of penicillin, the patient was discharged home without any complaint. Most laboratory values were returning to normal (Figure 2). Follow-up chest CT after discharge was almost completely cleared with no recurrence (Figure 3D).
DISCUSSION

The main pathogens of CAP in China are Streptococcus pneumoniae and Mycoplasma pneumoniae. Other common related-pathogens include Hemophilus influenzae, Chlamydia, Klebsiella pneumoniae and Staphylococcus aureus[1]. Respiratory virus, especially influenza, rhinovirus and coronavirus, is reported to be responsible for a third of adult pneumonia cases[4]. By comparison, pneumonia caused by Leptospira is rare and seldom reported. Leptospira interrogans, detected in this case, is one type of pathogenic spirochetes and could cause zoonotic leptospirosis. Leptospirosis is estimated to affect over one million people and accounts for about 60000 deaths annually worldwide[3]. Clinical manifestations of leptospirosis vary, ranging from subclinical status, mild influenza-like illness to severe multiple organ dysfunction, including kidney, liver, heart and lung. The typical severe syndrome, named “Weil’s disease”, is characterized by jaundice and renal failure with hemorrhagic complications[10].

Pulmonary involvement of leptospirosis occurs in 20%-70% of patients[11] and is mostly secondary to renal and hepatic dysfunction as an immunity-mediated status[11,12], including mild to severe pneumonia, deadly alveolar hemorrhage, adult respiratory distress syndrome, etc. [11] and manifesting as cough, dyspnea, hemoptyisis, etc. As observed in this case lung involvement of severe pneumonia was primary and main manifestations were relatively few. However, mortality from severe lung involvement like adult respiratory distress syndrome and pulmonary hemorrhage greatly exceeds that from Weil’s disease (> 50% vs > 10%), although optimal treatment is given[14]. It is the nonspecificity and diversity of clinical symptoms, particularly atypical manifestations, that lead to lack of clinical suspicion, which delays targeted detection and treatment of leptospirosis and increases mortality.

Common laboratory biochemical tests of leptospirosis usually show no specific abnormalities compared with most infectious diseases. For Leptospira-related pneumonia, chest CT may show ground-glass opacities, consolidation and small pleural effusion, which could also be observed in radiographs of influenza pneumonia[12,13]. Thus, it is possible to misdiagnose this case as influenza pneumonia, especially based on evidence from the initial throat swab test. During hospitalization, bronchoscopy with BALF collection was required because it could help to identify hemorrhage even if it remained occult and to obtain samples for etiological tests. Overall, for differential diagnosis, persistent hemorrhagic complications and related
abnormal values, which are relatively few in influenza-associated pneumonia under corresponding antiviral treatment, it may be the only hint to raise doubt of the initial diagnosis and treatment plan.

For the final etiological diagnosis, serological tests and isolation of Leptospira are usually considered as the prior standard diagnostic methods for leptospirosis, including conventional microscopic agglutination test and enzyme-linked immunosorben assay[19-20], whereas patient autoimmune status, previous medications with immunosuppressor and antibiotics and timing of specimen collection lead to an inevitable bias in etiological tests[21]. Polymerase chain reaction, as the representative of current molecular tests, is another widely-used and reliable diagnostic tool[1]. However, these methods share several common drawbacks, such as failing to discriminate among widely-diverse Leptospira serotypes and only targeting some specific pathogens based on clinical prejudgments[19-20].

NGS, applied in our case, is a rapid and high-throughput assay that determines the entire nucleotide sequences of a sample in a single run without any target primers or probes[19]. NGS could identify a variety of microorganisms at one detection and provide data about typing, virulence and antimicrobial resistance simultaneously[20]. Many previous reports confirmed that NGS offers great advantages in diagnosis and management of infectious diseases of undetermined, uncommon or emerging origins[20-22], and were superior to traditional detecting techniques. It should be noted that not all microorganisms detected by NGS, like internal colonizing bacteria, bear responsibility for the diseases. To identify the crucial pathogens, clinicians need to combine NGS results with contact history, clinical symptoms and other laboratory tests to make a comprehensive interpretation.

Penicillin remains the first choice for treatment of leptospirosis recommended by the World Health Organization because no penicillin-resistant strains have been found so far. Other antibiotics like doxycycline, tetracycline and amoxicillin are also preferred. Early and suitable administration of antibiotics could shorten disease course, decrease mortality and prevent deterioration[23]. For patients with severe pulmonary illness of leptospirosis, as observed in, hemodynamic and respiratory support are urgent. Mechanical ventilation should be a priority to deal with severe pulmonary hemorrhage[23]. Recently, extracorporeal membrane oxygenation, as a salvage intervention, is also confirmed to benefit the management of severe pulmonary leptospirosis[24]. Immunomodulation therapy is expected to promote recovery from leptospirosis. Rodrigo et al[25] reviewed that steroids may only improve the outcomes of pulmonary leptospirosis but not benefit other clinical subtypes. Several reports and trials supported utilization of glucocorticoids, and Shenoy et al[26] concluded use of corticosteroids within the first 24 h of onset of pulmonary illness could reduce the need for assisted ventilation[17,26-27]. Additionally, activated factor VII, desmopressin cyclophosphamide and hemofiltration have been used in several refractory cases[22,28].

Prevention measures play an important role in management of leptospirosis. It is well-known that pathogenic Leptospira invades bodies via skin and mucosa abrasion. Indirect infection by contact with contaminated water or soil is more common than direct contact with infected mammals. Avoiding exposure to pathogens, such as wearing boots and goggles, is the most important protective measures for people with potential occupational risk, like the river-trash cleaner in this case. Although vaccines against Leptospira have been available since 1886, its general application still faces great challenges. Current qualified vaccines are restricted to partial serotypes and provide short-term immunity and are only available in several specific epidemic areas[24]. In addition, oral doxycycline can be used as chemoprophylaxis for high-risk people such as rescuers in floods[25].

CONCLUSION

In summary, severe CAP caused by Leptospira is uncommon, but it poses a considerable threat to public health. Although the lack of specific clinical symptoms and high-efficient diagnostic tools make it hard to discriminate from other infectious diseases, many measures could be taken to correctly diagnose the disease. First, high clinical suspicion of Leptospira infection is vital to handle severe pneumonia patients, especially those with hemorrhage complications. Second, recognition of high-risk people in contact with possible infected mammals or contaminated water or soil will give strong evidence to diagnosis. Third, NGS has great advantages in identification of undetermined or uncommon pathogens, which could provide hints for diagnosis and
REFERENCES

1 Prina E, Ranzani OT, Torres A. Community-acquired pneumonia. Lancet 2015; 386: 1097-1108 [PMID: 26277247 DOI: 10.1016/S0140-6736(15)60733-4]

2 Bharti AR, Nally JE, Ricaldi JN, Matthias MA, Diaz MM, Lovett MA, Levett PN, Gilman RH, Willig MR, Gotuzzo E, Vinetz JM. Peru-United States Leptospirosis Consortium. Leptospirosis: a zoonotic disease of global importance. Lancet Infect Dis 2003; 3: 757-771 [PMID: 14632202 DOI: 10.1016/S1473-3099(03)00839-2]

3 Costa F, Hagan JE, Calcegno J, Kane M, Torgerson P, Martinez-Silveira MS, Stein C, Abela-Riddler B, Ko AI. Global Morbidity and Mortality of Leptospirosis: A Systematic Review. PLoS Negl Trop Dis 2015; 9: e0003898 [PMID: 26379143 DOI: 10.1371/journal.pntd.0003898]

4 Reis RB, Ribeiro GS, Felzemburgh RD, Santana FS, Mohr S, Melendez AX, Queiroz A, Santos AC, Ravines RR, Tassinari WS, Carvalho MS, Reis MG, Ko AI. Impact of environment and social gradient on Leptospira infection in urban slums. PLoS Negl Trop Dis 2008; 2: e228 [PMID: 18431445 DOI: 10.1371/journal.pntd.0000228]

5 Karpagam KB, Ganesh B. Leptospirosis: a neglected tropical zoonotic infection of public health importance—an updated review. Eur J Clin Microbiol Infect Dis 2020; 39: 835-846 [PMID: 31898795 DOI: 10.1007/s10096-019-03797-4]

6 Soo ZMP, Khan NA, Siddiqui R. Leptospirosis: Increasing importance in developing countries. Acta Trop 2020; 201: 105183 [PMID: 31542372 DOI: 10.1016/j.actatropica.2019.105183]

7 Marquez A, Djelouadji Z, Luttard V, Kodjo A. Overview of laboratory methods to diagnose Leptospirosis and to identify and to type leptospires. Int Microbiol 2017; 20: 184-193 [PMID: 29529330 DOI: 10.2436/20.1501.01.302]

8 Gwinn M, MacCannell D, Armstrong GL. Next-Generation Sequencing of Infectious Pathogens. JAMA 2019; 321: 893-894 [PMID: 30763343 DOI: 10.1001/jama.2018.21669]

9 Ruuskamen O, Labhi E, Jennings LC, Murdoch DR. Viral pneumonia. Lancet 2011; 377: 1264-1275 [PMID: 21435708 DOI: 10.1016/s0140-6736(10)6459-4]

10 AREAN VM. The pathologic anatomy and pathogenesis of fatal human leptospirosis (Weil's disease). Am J Pathol 1962; 40: 393-423 [PMID: 13862141]

11 Carvalho CR, Bethlem EP. Pulmonary complications of leptospirosis. Clin Chest Med 2002; 23: 469-478 [PMID: 12090240 DOI: 10.1016/s0272-5231(01)00010-7]

12 Marchiori E, Lourenço S, Setúbal S, Zanetti G, Gasparetto TD, Hochhegger B. Clinical and imaging manifestations of hemorrhagic pulmonary leptospirosis: a state-of-the-art review. Lung 2011; 189: 1-9 [PMID: 21152929 DOI: 10.1007/s00115-010-9273-0]

13 De Brito T, Silva AMGD, Abreu PAE. Pathology and pathogenesis of human leptospirosis: a commented review. Rev Inst Med Trop Sao Paulo 2018; 60: e23 [PMID: 29846473 DOI: 10.1590/s1679-9964201800023]

14 McBride AJ, Athanazio DA, Reis MG, Ko AI. Leptospirosis. Curr Opin Infect Dis 2005; 18: 376-386 [PMID: 16148523 DOI: 10.1097/01.qco.0000178824.05715.2c]

15 Koo HJ, Lim S, Choe J, Choi SH, Sung H, Do KH. Radiographic and CT Features of Viral Pneumonia. Radiographics 2018; 38: 719-739 [PMID: 29757717 DOI: 10.1148/radiographics.2018170048]

16 Agrawal VK, Bansal A, Pujani M. A rare case of leptospirosis with isolated lung involvement. Indian J Crit Care Med 2015; 19: 174-176 [PMID: 25806515 DOI: 10.4103/0972-5229.152765]

17 Haahe DA, Levett PN. Leptospirosis in humans. Curr Top Microbiol Immunol 2015; 387: 65-97 [PMID: 25388113 DOI: 10.1007/978-3-662-45059-8_5]

18 Héry G, Lefèvre J, Flècher E, Quentin C, Piau C, Le Tulzo Y, Tattevin P. Massive intra-alveolar hemorrhage caused by Leptospira serovar Djasiman in a traveler returning from Laos. J Travel Med 2015; 22: 212-214 [PMID: 25728613 DOI: 10.1111/jtm.12189]

19 Deukenberg RH, Bathoorn E, Chlebowicz MA, Couto N, Ferdous M, Garcia-Cobos S, Kooistra-Smid AM, Raangs EC, Rosena S, Veloo AC, Zhou K, Friedrich AW, Rossen JW. Application of next generation sequencing in clinical microbiology and infection prevention. J Biotechnol 2017; 243: 16-24 [PMID: 28042011 DOI: 10.1016/j.jbiotec.2016.12.022]
Bao QH et al. Misdiagnosed pneumonia caused by Leptospira

Wilson MR, Naccache SN, Samayoa E, Biagtan M, Bashir H, Yu G, Salamat SM, Somasekar S, Federman S, Miller S, Sokolic R, Garabedian E, Candotti F, Buckley RH, Reed KD, Meyer TL, Seroogy CM, Galloway R, Henderson SL, Gern JE, DeRisi JL, Chiu CY. Actionable diagnosis of neuroleptospirosis by next-generation sequencing. *N Engl J Med* 2014; 370: 2408-2417 [PMID: 24896819] DOI: 10.1056/NEJMoa1401268

Briese T, Pawska JT, McMullan LK, Hutchison SK, Street C, Palacios G, Khrstova ML, Weyer J, Swansepol R, Egholm M, Nichol ST, Lipkin WI. Genetic detection and characterization of Lujo virus, a new hemorrhagic fever-associated arenavirus from southern Africa. *PLoS Pathog* 2009; 5: e1000455 [PMID: 19478873] DOI: 10.1371/journal.ppat.1000455

Palacios G, Druce J, Du L, Tran T, Birch C, Briese T, Conlan S, Quan PL, Hui J, Marshall J, Simons JF, Egholm M, Paddock CD, Shieh WJ, Goldsmith CS, Zaki SR, Catton M, Lipkin WI. A new arenavirus in a cluster of fatal transplant-associated diseases. *N Engl J Med* 2008; 358: 991-998 [PMID: 18256387] DOI: 10.1056/NEJMoa073785

Goarant C. Leptospirosis: risk factors and management challenges in developing countries. *Res Rep Trop Med* 2016; 7: 49-62 [PMID: 30050339] DOI: 10.2147/RRTM.S102543

Rodrigo C, Lakshitha de Silva N, Goonaratne R, Samarasekara K, Wijesinghe I, Parththipan B, Rajapakse S. High dose corticosteroids in severe leptospirosis: a systematic review. *Trans R Soc Trop Med Hyg* 2014; 108: 743-750 [PMID: 25266477] DOI: 10.1093/trstmh/tru148

Shenoy VV, Nagar VS, Chowdhury AA, Bhalgat PS, Juvale NJ. Pulmonary leptospirosis: an excellent response to bolus methylprednisolone. *Postgrad Med J* 2006; 82: 602-606 [PMID: 16954459] DOI: 10.1136/pgmj.2005.044255

Niwattayakul K, Kaewtsi S, Chueasuwanchai S, Hoontrakul S, Chareonwat S, Suttinont C, Phimda K, Chierakul W, Silpasakorn S, Suputtamongkol Y. An open randomized controlled trial of desmopressin and pulse dexamethasone as adjunct therapy in patients with pulmonary involvement associated with severe leptospirosis. *Clin Microbiol Infect* 2010; 16: 1207-1212 [PMID: 19732091] DOI: 10.1111/j.1469-0691.2009.03037.x

Trivedi SV, Chavda RK, Wadia PZ, Sheth V, Bhagade PN, Trivedi SP, Clerk AM, Mevawala DM. The role of glucocorticoid pulse therapy in pulmonary involvement in leptospirosis. *J Assoc Physicians India* 2001; 49: 901-903 [PMID: 11837758]

Trivedi SV, Vasava AH, Patel TC, Bhatia LC. Cyclophosphamide in pulmonary alveolar hemorrhage due to leptospirosis. *Indian J Crit Care Med* 2009; 13: 79-84 [PMID: 19881188] DOI: 10.4103/0972-5229.56053

Tatopoulos A, Herbain D, Kazmierczak C, Bollaert PE, Gibot S. Parenteral use of recombinant activated factor VII during diffuse alveolar hemorrhage secondary to leptospirosis. *Intensive Care Med* 2010; 36: 555-556 [PMID: 19924396] DOI: 10.1007/s00134-009-1718-7

Xu Y, Ye Q. Human leptospirosis vaccines in China. *Hum Vaccin Immunother* 2018; 14: 984-993 [PMID: 29148958] DOI: 10.1080/21645515.2017.1405884

Schneider MC, Velasco-Hernandez J, Min KD, Leonel DG, Baca-Carrasco D, Gompper ME, Hartskeerl R, Munoz-Zanzi C. The Use of Chemoprophylaxis after Floods to Reduce the Occurrence and Impact of Leptospirosis Outbreaks. *Int J Environ Res Public Health* 2017; 14 [PMID: 28587195] DOI: 10.3390/ijerph14060594
