Electrocardiographic Abnormalities in Trypanosoma cruzi Seropositive and Seronegative Former Blood Donors

Antonio L. Ribeiro1,*, Ester C. Sabino2, Milena S. Marcolino1, Vera M. C. Salemi3, Barbara M. Ianni3, Fábio Fernandes3, Luciano Nastari3, André Antunes4, Márcia Menezes4, Cláudia Di Lorenzo Oliveira5, Vandana Sachdev6, Danielle M. Carrick7, Michael P. Busch8, Eduard L. Murphy8, for the NHLBI Retrovirus Epidemiology Donor Study-II (REDS-II), International Component

1 Hospital das Clínicas and Faculdade de Medicina, Universidade Federal de Minas Gerais, Belo Horizonte, Brazil, 2 Department of Infectious Disease and Institute of Tropical Medicine, University of São Paulo, São Paulo, Brazil, 3 Cardiomyopathy Unit of the Heart Institute (InCor) da Faculdade de Medicina da Universidade de São Paulo, São Paulo, Brazil, 4 Centro de Ciências Biológicas e da Saúde, Universidade Estadual de Montes Claros, Montes Claros, Brazil, 5 Campus Donna Lindu da Universidade Federal de São João del Rei, Divinópolis, Brazil, 6 National Heart, Lung and Blood Institute, Bethesda, Maryland, United States of America, 7 Westat, Rockville, Maryland, United States of America, 8 Blood Systems Research Institute (BSRI) and University of California at San Francisco, San Francisco, California, United States of America

Abstract

Background: Blood donor screening leads to large numbers of new diagnoses of Trypanosoma cruzi infection, with most donors in the asymptomatic chronic indeterminate form. Information on electrocardiogram (ECG) findings in infected blood donors is lacking and may help in counseling and recognizing those with more severe disease.

Objectives: To assess the frequency of ECG abnormalities in T. cruzi seropositive relative to seronegative blood donors, and to recognize ECG abnormalities associated with left ventricular dysfunction.

Methods: The study retrospectively enrolled 499 seropositive blood donors in São Paulo and Montes Claros, Brazil, and 483 seronegative control donors matched by site, gender, age, and year of blood donation. All subjects underwent a health clinical evaluation, ECG, and echocardiogram (Echo). ECG and Echo were reviewed blindly by centralized reading centers. Left ventricular (LV) dysfunction was defined as LV ejection fraction (EF)<50%.

Results: Right bundle branch block and left anterior fascicular block, isolated or in association, were more frequently found in seropositive cases (p<0.0001). Both QRS and QTc duration were associated with LVEF values (correlation coefficients -0.159, p<0.0003, and -0.142, p=0.002) and showed a moderate accuracy in the detection of reduced LVEF (area under the ROC curve: 0.778 and 0.790, both p<0.0001). Several ECG abnormalities were more commonly found in seropositive donors with depressed LVEF, including rhythm disorders (frequent supraventricular ectopic beats, atrial fibrillation or flutter and pacemaker), intraventricular blocks (right bundle branch block and left anterior fascicular block) and ischemic abnormalities (possible old myocardial infarction and major and minor ST abnormalities). ECG was sensitive (92%) for recognition of seropositive donors with depressed LVEF and had a high negative predictive value (99%) for ruling out LV dysfunction.

Conclusions: ECG abnormalities are more frequent in seropositive than in seronegative blood donors. Several ECG abnormalities may help the recognition of seropositive cases with reduced LVEF who warrant careful follow-up and treatment.

Introduction

Chagas disease (ChD), caused by a flagellate protozoan, Trypanosoma cruzi (T. cruzi), is a major health problem in Latin America, where more than 8 million persons are infected [1,2]. Chronic cardiopathy is the most important and severe manifestation of human Chagas disease, eventually affecting approximately 20% to 40% of those in the chronic phase of the disease [1,2]. Due to migratory movements, ChD is now a world-wide challenge, since hundreds of thousands of chronically infected persons are now living not only in T. cruzi endemic countries but also in developed countries, mainly in Europe and the United States and Canada.
Methods

blood-borne T. cruzi populations including the United States, Canada, Spain and Portugal. Disease transmission via blood transfusions, universal blood bank screening for ChD has been established in most endemic countries, as well as in non-endemic countries with large immigrant populations. Blood donor screening leads to large numbers of new diagnoses of chronic T. cruzi infection. Counseling these individuals should address the recognition of those with more severe disease that deserve to be rigorously evaluated by experienced cardiologists and treated more promptly. The electrocardiogram is an important exam that can help in the recognition of cardiac disease and the evaluation of prognosis in ChD patients, but its role in blood donors has not been studied. The authors describe some electrocardiographic abnormalities that are typical of the infected blood donors, as well ECG abnormalities that help in the identification of those with severe cardiac involvement. These results may guide the evaluations of patients with incidentally detected T. cruzi infection from blood bank testing or public health screening.

Since one of the mechanisms of transmission of the disease is via blood transfusions, universal blood bank screening for ChD has been established in most endemic countries, as part of South American regional initiatives of elimination of transmission of the disease. Non-endemic countries with large immigrant populations including the United States, Canada, Spain and Portugal, have also begun to institute interventions to prevent blood-borne T. cruzi transmission. Blood donor antibody screening results in large numbers of new diagnoses of chronic T. cruzi infection, most of them in the asymptomatic, indeterminate form of infection. Counseling these individuals should address the recognition of those with more severe disease that deserve to be rigorously evaluated by experienced cardiologists and treated more promptly.

Electrocardiogram (ECG), one of the most important tests in evaluation of ChD, is used to define the clinical stage of the disease with potential prognostic implications. Most ECG studies of newly diagnosed ChD patients were performed decades ago, generally in patients identified by community or hospital based sampling; information on ECG findings in seropositive blood donors is lacking as is data relative to matched seronegative controls evaluated in parallel. Additionally, most studies of ECG findings are not accompanied by systematic results that use core-lab reading centers. Results of ECG and Echo were reviewed blindly by centralized reading centers. T. cruzi antibody status was confirmed on plasma samples collected at the time of enrollment.

Ethics statement

The study follows the Declaration of Helsinki of the Ethical Principles for Medical Research Involving Human Subjects, was approved by the Brazilian National Ethical Committee (CONEP # 1312/2006) and all subjects gave written informed consent.

Blood bank screening and enrollment procedures

In 1998–2002, Fundação Pro-Sangue in São Paulo performed T. cruzi antibody screening using three serological methods: ELISA, hemaglutination and immunofluorescence for Chagas disease. For the purpose of this study, we included as seropositive cases donors who were positive by all three assays at the time of donation and on a separate sample obtained at the time of counseling, generally one to four months after the donation. In Montes Claros, Hemominas screened all donations with two serological assays: ELISA and hemaglutination. All donors reactive to both assays at the time of donation and counseling were considered eligible for this study. Follow-up samples from all enrolled subjects were retested for T. cruzi antibody under code at the REDS-II Central Laboratory in the US using an FDA-approved assay manufactured by Ortho Diagnostics. All 483 control donors tested negative on their follow-up samples collected at the time of enrollment and clinical assessments for this study. Of the 499 case donors who originally screened as seropositive at the time of donation in 1998–2002, serum collected in 2008–2010 from 498 tested repeat reactive, while the samples from one donor tested just below the assay cutoff consistent with slowly progressive seroreversion.

Measurements

A face to face T. cruzi risk factor and health history questionnaire was administered by a trained nurse in each site. The questionnaire collected detailed information regarding demographics, cities of residence, physical activity, medical history, exposure to T. cruzi, previous ChD diagnoses, cardiac and GI symptoms, past medical history and medication history. All subjects received a physical examination by a non-blinded physician with recording of height, weight, blood pressure, heart rate and physical exam findings.

Resting 12-lead ECG were recorded using the same model of machine at both sites (General Electric MAC 1200 electrocardiograph; GE Healthcare, Waukesha, WI) using standardized procedures. All ECGs were processed blindly by the central ECG laboratory (Epidemiological Cardiology Research Center, Wake Forest University, Winston-Salem, NC), where they were visually inspected for technical errors and inadequate quality and processed with the 2001 version of the GE Marquette 12-12SL program. ECGs were analyzed electronically, with manual over-reading by trained cardiologists to ensure quality control. ECGs were classified by Minnesota code criteria using variables that were derived from the median complex of the Marquette measurement matrix. In this study, major and minor ECG

Author Summary

Chagas disease (ChD), caused by the protozoa Trypanosoma cruzi, is endemic in most Latin American countries and may be transmitted via blood transfusions. Cardiac disease is a major feature of chronically infected patients and may be lethal. Universal blood bank screening for ChD has been established in most Latin American countries, as well as in non-endemic countries with large immigrant populations, including the United States, Canada, Spain and Portugal. Blood donor screening leads to large numbers of new diagnoses of chronic T. cruzi infection. Counseling these individuals should address the recognition of those with more severe disease that deserve to be rigorously evaluated by experienced cardiologists and treated more promptly. The electrocardiogram is an important exam that can help in the recognition of cardiac disease and the evaluation of prognosis in ChD patients, but its role in blood donors has not been studied. The authors describe some electrocardiographic abnormalities that are typical of the infected blood donors, as well ECG abnormalities that help in the identification of those with severe cardiac involvement. These results may guide the evaluations of patients with incidentally detected T. cruzi infection from blood bank testing or public health screening.
abnormalities were defined as previously established [22], modified to include ECG abnormalities typical of Chagas cardiomyopathy with prognostic significance, as frequent supraventricular or ventricular premature beats [10]. Old myocardial infarction (MI) on ECG was defined by the presence of major Q wave abnormalities (MC 1.1.x or 1.2.x) or minor Q waves abnormalities with ST segment or T-wave abnormalities (1.3.x and [4.1.x, 4.2, 5.1, or 5.2]) [18].

Echocardiographic studies were performed using Sequoia 512 ultrasound machine (Acuson, Mountain View, CA, USA) at São Paulo site and GE Vivid3 (GE Healthcare, Waukesha, WI) at Montes Claros site. Cardiac measurements were performed according to the guidelines of the American Society of Echocardiography [23]. Studies were recorded in digital format and all measurements were performed on digital loops using a Digisonics offline analysis station (version 3.2 software, Digisonics, Houston, Tex) at the Cardiovascular Branch, Echocardiography Laboratory, National Heart, Lung, and Blood Institute, Bethesda, Maryland. LV ejection fraction (LVEF) was calculated based on modified form of Simpson’s biplane method [23] and, if atrial fibrillation is present, LVEF is estimated by a visual method. For this study, LV systolic dysfunction was defined as LVEF<0.50.

Statistical analysis

Statistical analyses were conducted using SAS 9.2 and SPSS 18. Distributions of data were examined for normality by using Kolmogorov-Smirnov tests. Continuous variables were expressed as median [interquartile range (IQR)] and differences between seropositive and seronegative donors were compared using Wilcoxon-Mann-Whitney test, since these variables were not normally distributed. Categorical variables were summarized as counts and percentages and differences were compared using the Chi-square test or Fisher exact test. Association between quantitative ECG variables and LVEF was evaluated by Spearman correlation coefficient (rs). A p-value<0.05 was considered significant.

Receiver Operator Characteristic (ROC) curves were plotted in order to evaluate the accuracy of ECG measurements in detecting reduced LVEF and the area under the curve (AUC) was calculated. Sensibility, specificity and positive and negative predictive values of abnormal ECG, wide QRS duration ($\geq$120 ms) and long QTc interval ($\geq$440 ms) were calculated with 95% confidence intervals. These cut points were selected since they are well-established in cardiology practice.

Results

ECG abnormalities in *T. cruzi* seropositive and seronegative blood donors

The study sample consisted of 499 seropositive and 488 seronegative donors. Seropositive had a higher proportion of non-white skin color and lower weight, body mass index, total cholesterol levels and pulse heart rate, as well as higher NT-proBNP values (table 1). LV systolic dysfunction was more commonly found in seropositive cases. The groups were comparable with regard to age, gender or other major cardiovascular risk factors (table 1). Quantitative variables are shown in table 2. All ECG intervals were longer in seropositive donors, and both heart rate and HRV indexes had lower values. ECG abnormalities were similar in seropositive and seronegative donors, except for a higher frequency in seropositive subjects of right bundle branch block and left anterior fascicular block, isolated (16% vs. 2%, p<0.001 for RBBB and 15% vs. 2%, p<0.001 for LAFB) or in association (4% vs. 0, p<0.001); and a higher frequency of left ventricular hypertrophy in seronegative subjects (<1% vs. 1%, p = 0.004).

Seropositive cases showed more abnormal ECGs (51% vs. 32%, p<0.001) than seronegative donors due to a higher prevalence of major (20% vs. 9%) ECG abnormalities. They also presented a higher number of major ECG abnormalities per tracing when compared to seronegative donors: 20% vs. 7% had one and 6% vs. 2% had 2 or more ECG abnormalities, respectively (p<0.001).

ECG abnormalities in *T. cruzi* seropositive blood donors with and without LV dysfunction

LVEF was reduced in 36 out of 497 seropositive subjects with available data [prevalence of LV systolic dysfunction of 7.2%]; in two patients LVEF measurement was not obtained due to technical reasons. Most seropositive subjects had LV systolic dysfunction considered mild (LVEF ranging between 40 and 49%, n = 17, 3.4%) or moderate (LVEF from 30–39%, n = 11, 2.2% of total); only 8 (1.6%) showed markedly depresses LVEF (<30%). Seropositive blood donors with and without LV dysfunction had comparable demographic and medical characteristics, although NT-proBNP levels were higher in those with LVEF below 50% (45 [25–76] vs351 [109–789], p<0.001).Seropositive donors with LV dysfunction showed longer PR, QRS and corrected QT intervals/durations (Figure 1), although both the heart rate and HRV indexes were not different between groups (data not shown).

Both QRS and QTc duration were associated with LVEF values (rs = −0.159, p<0.0005, rs = −0.142, p = 0.002), and showed moderate accuracy in the detection of reduced LVEF (ROC: 0.778 and 0.790, both p<0.0001, Figure 2). None of the other quantitative ECG variables showed significant correlation with measured LVEF.

Several ECG abnormalities were more commonly found in seropositive donors with depressed LVEF (table 3), including rhythm disorders (frequent supraventricular ectopic beats, atrial fibrillation or flutter and pacemaker), intraventricular blocks (right bundle branch block and left anterior fascicular block) and ischemic abnormalities (old myocardial infarction and major and minor ST abnormalities).

Almost all seropositive donors (33/36) with depressed LVEF showed at least one major or minor ECG abnormality. Only three subjects presented with normal ECG and abnormal LVEF: 0.46, 0.45 and 0.40, respectively; this last subject is a hypertensive patient with typical features of hypertensive cardiomyopathy in the Echo study who was not classified as having Chagas cardiomyopathy by the expert adjudication process employed in the parent study [19]. Seropositive donors with depressed LVEF had a higher prevalence of ECG abnormalities (69% vs. 25%, p<0.001) and a higher number of major ECG abnormalities per tracing when compared to seropositive donors with normal LVEF (39% vs. 19% with one and 31% vs. 4% with 2 or more abnormalities, p<0.001). The diagnostic performance of selected ECG abnormalities was evaluated (table 4). An abnormal ECG (with minor or major abnormalities) is a sensitive marker for recognition of seropositive donors with depressed LVEF, with a negative predictive value of 99% (96–100%).

Discussion

In this analysis of ECG findings from a large, controlled study focused on former blood donors with and without *T. cruzi* seropositivity, the frequency of major ECG abnormalities was higher in seropositive donors than in seronegative subjects; right bundle branch block and/or left anterior were more commonly found in seropositive than in well-matched seronegative donors.
When considering only seropositive donors, those with LV dysfunction rarely presented with a normal ECG, having one or more major ECG abnormalities, including rhythm disturbances, intraventricular blocks and ischemic abnormalities. The presence of either major or minor ECG abnormalities is therefore a sensitive marker of the presence of LV dysfunction in ChD, and, equally important, the absence of ECG abnormalities has high negative predictive value. Consequently, we recommend that initial clinical evaluation of seropositive blood donors (and probably seropositive subjects identified through other population based screening programs) can be limited to ECG with more expensive echocardiography performed on patients with abnormal ECG and/or clinical findings suggesting of ChD.

Our findings suggest that seropositive blood donors have a similar profile to community ChD populations in terms of the prevalence and type of ECG abnormalities [12,14,15,24], albeit with significantly lower rates and severity of abnormalities than in hospital and clinic-based series [11,16,16,25–28]. However, the mean age of blood donors was higher than those studied in early community-based series and the prevalence of concurrent chronic conditions in blood donors was relatively high (24% hypertension, 5% diabetes), in contrast to some studies in which those conditions were excluded [16,28]. Considering this higher frequency of ECG abnormalities in seronegative donors, right bundle branch block and/or left anterior fascicular block was the only finding more

### Table 1. Demographic and medical characteristics of the study population, by T.cruzi antibody status.

|                          | Seropositive | Seronegative | p-value* |
|--------------------------|--------------|--------------|----------|
| **N = 499**              | **N = 488**  |              |          |
| **Site**                 |              |              |          |
| Sao Paulo                | 255 (51)     | 239 (49)     |          |
| Montes Claros            | 244 (49)     | 249 (51)     |          |
| **Male gender**          | 261 (52)     | 241 (49)     |          |
| **Age (years)**          | 48 (40–57)   | 49 (42–58)   |          |
| **Skin Color**           |              |              | 0.0005   |
| White                    | 155 (31)     | 203 (42)     |          |
| Black                    | 56 (11)      | 28 (6)       |          |
| Mixed                    | 274 (55)     | 249 (51)     |          |
| Other                    | 11 (2)       | 7 (1)        |          |
| **Refuse to answer/missing** | 3 (1)       | 1 (<1)       |          |
| **Height (cm)**          | 163 (157–170)| 165 (158–172)|          |
| **Weight (Kg)**          | 71 (64–79)   | 73.5 (66–84) | <0.0001  |
| **BMI**                  | 26 (24–29)   | 27 (25–30)   | 0.0009   |
| **Pulse (bpm)**          | 65 (60–72)   | 70 (60–75)   | 0.002    |
| **Systolic BP (mm Hg)**  | 125 (114–140)| 125 (115–140)|          |
| **Hypertension (self-reported)** | 113 (23)    | 119 (24)     |          |
| **Diabetes (self-reported)** | 27 (5)      | 24 (5)       |          |
| **Total Cholesterol (mg/dL)** | 195 (168–226)| 204.5 (175–230)| 0.025   |
| **Smoking (current or former smoker)** | 216 (43)    | 233 (48)     |          |
| **Glucose (mg/dL)**      | 87 (79–96)   | 87 (80–98)   |          |
| **LVEF<0.50**            | 36 (7)       | 11 (2)       | 0.0002   |
| **NT-pro BNP (pg/mL)**   | 48 (27–90)   | 37 (23–64)   | <0.0001  |

*Data is presented as median (interquartile range) or numbers (%).

**p-value**: p < 0.05.

Table 2. Quantitative ECG measurements in T.cruzi seropositive and seronegative subjects.

| ECG variable          | Seropositive | Seronegative | p-value* |
|-----------------------|--------------|--------------|----------|
| **N = 499**           | **N = 488**  |              |          |
| Heart Rate (bpm)      | 62 (57–69)   | 64 (58–72)   | 0.003    |
| PR duration (ms)      | 160 (144–178)| 156 (142–168)| 0.007    |
| QRS duration (ms)     | 90 (84–104)  | 88 (82–95,75)| <0.001   |
| QT duration (ms)      | 420 (402–439)| 412 (392–432)| <0.001   |
| QTc duration (ms)     | 429 (412–447)| 427 (411–442)| 0.061    |
| HRV sdnn (ms)**       | 16 (10–25)   | 22 (14–35)   | <0.001   |
| HRV rmssd (ms)**      | 18 (11–29)   | 24 (15–39)   | <0.001   |

*Data is presented as median (interquartile range).

**p-value**: p < 0.05.

**HRV data available in 497 seropositive and 461 seronegative subjects. QTc: corrected QT interval by Bazett’s formula; HRV: heart rate variability; rmssd: root mean square of successive differences in normal RR intervals; sdnn: standard deviation of all normal RR intervals.

doi:10.1371/journal.pntd.0002078.t001

doi:10.1371/journal.pntd.0002078.t002
Figure 1. Quantitative ECG measurements in *T. cruzi* seropositive blood donors by left ventricular ejection fraction. A. PR interval. B. QRS duration. C. QT interval corrected by Bazett’s formula. doi:10.1371/journal.pntd.0002078.g001

Figure 2. Association between left ventricular ejection fraction and ECG measurements. A – Left ventricular ejection fraction and QRS duration; B - Left ventricular ejection fraction and QT corrected interval; and C: Diagnostic accuracy (area under the ROC curve) of QRS duration and QT corrected interval in the detection of reduced left ventricular ejection fraction. doi:10.1371/journal.pntd.0002078.g002
Table 3. ECG findings in T. cruzi seropositive blood donors by LVEF.

| ECG variables | LVEF<0.50 | LVEF>0.50 | p-value* |
|---------------|-----------|-----------|----------|
| N=36          | N=461     |           |          |
| Rhythm        |           |           |          |
| Sinus tachycardia | 0         | 0         |          |
| Sinus bradycardia | 1 (3)     | 20 (4)    |          |
| Frequent VPB   | 2 (6)     | 10 (2)    |          |
| Frequent SPB   | 3 (8)     | 6 (1)     | 0.02     |
| Atrial fibrillation | 2 (6)     | 0         | 0.005    |
| or flutter     |           |           |          |
| Pacemaker      | 3 (8)     | 2 (<1)    | 0.003    |
| Intraventricular block |       |           |          |
| LBBB          | 1 (3)     | 0         |          |
| RBBB          | 10 (28)   | 70 (15)   | 0.04     |
| LAFB          | 13 (38)   | 61 (13)   | <0.0001  |
| LAFB+RBBB     | 9 (25)    | 9 (2)     | <0.0001  |
| Incomplete LBBB | 0         | 13 (3)    |          |
| Incomplete RBBB | 5 (14)    | 30 (7)    |          |
| Ativoventricular block |       |           |          |
| First degree  | 2 (6)     | 12 (3)    |          |
| Second degree | 0         | 1 (<1)    |          |
| Third degree  | 0         | 0         |          |
| Ischemic      |           |           |          |
| Old MI        | 4 (11)    | 8 (2)     | 0.008    |
| Major isolated ST-T abnormalities | 7 (19) | 16 (3) | <0.0001 |
| Minor isolated ST-T abnormalities | 12 (33) | 47 (10) | <0.0001 |
| ST segment elevation | 0 | 20 (4) |          |
| Other         |           |           |          |
| LVH           | 0         | 1 (<1)    |          |
| Short PR interval | 0       | 16 (3)    |          |
| Major QT prolongation | 1 (3)  | 2 (<1)   |          |
| Minor QT prolongation | 1 (3) | 7 (2)    |          |
| Low QRS amplitude | 1 (3) | 16 (3)   |          |

*Reported if <0.05. VPB: ventricular premature beats; SPB: supraventricular premature beats; LBBB: left bundle branch block; RBBB: right bundle branch block; LAFB: left anterior fascicular block; Old MI: Old myocardial infarction, defined as major Q wave or minor Q waves with ST-T abnormalities; LVH: Left ventricular hypertrophy; major QT prolongation: QTc>111%; minor QT prolongation: QTc>111%; minor QT prolongation index: (QT/656) * (HR/100). doi:10.1371/journal.pntd.0002078.t003

LV systolic dysfunction, defined as reduced LVEF (<0.50), is a major marker of higher risk of death in ChD [30]. LV dysfunction, generally mild or moderate, occurred in a minority of cases in this sample (7.2%), reflecting the low risk profile of the seropositive donor population studied (the donors had to be clinically asymptomatic to give blood donations in 1990–2002, 0–12 years prior to the rigorous assessment including ECG and Echo that was the basis of the current analysis). In contrast, Ribeiro et al. [31] found a prevalence of more severe LV dysfunction (defined as LVEF≤0.40) of 9.1% and 14.5% in two samples from a Brazilian Outpatient Clinic that is a regional reference center for blood banks and primary care units, and Salles et al. [28] observed LV dysfunction in 109 out of 738 patients (14.8%) at another Brazilian Reference Outpatient Clinic.

In this study, several ECG abnormalities typical of ChD were predictive for LV dysfunction among seropositive donors. Right bundle branch block, frequently combined with left anterior fascicular block, is the most characteristic ECG abnormality in ChD [10] and is associated with higher risk of death in longitudinal studies [13,32]. ChD patients with pacemakers have lower LVEF comparative to pacemaker patients without ChD [33]. In contrast to what we found in this study, frequent ventricular ectopic beats have been repeatedly related to LV dysfunction [26,34] and the observed association of supraventricular ectopic beats with LV dysfunction has not been reported before. Since frequent supraventricular ectopic beats can precede the development of atrial fibrillation, we hypothesize that higher left atrial pressure and volume secondary to LV systolic dysfunction may lead to frequent atrial ectopic beats and, after years, to atrial fibrillation. Atrial fibrillation is a late abnormality in the natural history of ChD, related to other ECG abnormalities, LV dysfunction and higher risk of death [12,35]. Pathological q waves, ischemic ST-T abnormalities and abnormal T waves have also been reported to be markers of risk in Chagas cardiopathy [27,36]. Prolonged QRS duration and QTc interval were both related to LV dysfunction [27,37] and to worse prognosis [27,38]. Since prolonged excitation time in ventricular conduction defects may induce secondary prolongation of the QT interval [39], the significance of prolonged QTc interval in seropositive donors should be interpreted with caution, considering that no correction for QRS duration was made in this study. Those with more than one ECG abnormality are at greater risk of having LV dysfunction, as reported in other studies [26,40].

Heart rate variability indexes SDNN and RMSSD, calculated from standard 10-seconds ECG tracing, were reduced in seropositive donors but these findings were not correlated to the presence of LV systolic dysfunction. Both SDNN and RMSSD indexes from a 10-second ECG are markers of parasympathetic modulation of the sinus node [41]. In some studies in other settings [42–44], 10-s HRV indexes were predictive of the risk of death. Impairment of cardiac vagal modulation has been consistently reported in ChD [45–47] and occurs early in the evolution of the disease, preceding LV systolic dysfunction [48]. However, the association of vagal dysautonomia with LVEF and with prognosis in ChD is still controversial [46,49].

Data from our study suggest that the ECG can be useful to guide the management of seropositive blood donors: an abnormal ECG is a sensitive marker of LV dysfunction, while a normal ECG carries a high negative predictive value. Indeed, a normal ECG is an established marker of excellent prognosis in medium-term follow-up of T. cruzi seropositive subjects [12,13,16]. The diagnostic performance of a single ECG interval measurement, such as a normal QRS duration or corrected QT interval, is not good, as previously reported [37].
The main strength of this analysis is that it is based on findings from a large controlled, rigorously conducted study, with central and blinded reading of both ECG and Echo results, and with classification using an internationally accepted ECG code, the Minnesota Code. This is, to the best of our knowledge, the only study with these features in a population of seropositive and matched seronegative blood donors. Because these blood donors were healthy at study baseline, the incidence of pathology will be lower than in prevalent patient cohorts. On the other hand, because patients with chronic conditions, including hypertension and diabetes, were not excluded, it allows the generalization of findings to other seropositive blood donor populations in endemic and non-endemic countries. The main limitations are related to the cross-sectional status of the current analysis, with no information on the prognostic role of the observed abnormalities. We also lacked baseline measurements on the cohort to assure the absence of cardiac pathology, although all were healthy enough to donate blood. Moreover, blood donors from non-endemic countries may have different epidemiological profiles due to country of origin, socio-economic status and time since acquisition of infection prior to detection as seropositive donors, as well as possible differences in disease progression depending on the strain of T. cruzi.

In conclusion, we identified several ECG abnormalities that are predictive of LV dysfunction in ChD. Due to the study setting involving previously healthy seropositive donors who developed incident ChD, these study findings may be extrapolated to other low-risk populations. In particular, the results may guide the evaluations of patients with incidentally detected T. cruzi seropositivity from blood bank testing in endemic and increasingly in non-endemic countries, and from public health screening in endemic countries.

Author Contributions
Conceived and designed the experiments: ALR ECS VMCS BMI FF LN MPB ELM. Performed the experiments: ALR ECS VMCS BMI FF LN AA MM CDLO VS MPB ELM. Contributed reagents/materials/analysis tools: ALR ECS VMCS BMI FF LN AA MM CDLO VS. Wrote the paper: ALR ECS VMCS BMI MPB ELM.

References
1. Ribeiro AL, Nunes MP, Teixeira MM, Rocha MO (2012) Diagnosis and management of Chagas disease and cardiomyopathy. Nat Rev Cardiol 9: 576–589. doi: 10.1038/nrcardio.2012.109.
2. World Health Organization (2010) First WHO report on neglected tropical diseases: working to overcome the global impact of neglected tropical diseases. pp. 1–172.
3. Bern C, Montgomery SP (2009) An estimate of the burden of chagas disease in the United States. Clin Infect Dis 49: e52–e54.
4. World Health Organization (2010) Control and prevention of Chagas disease in Europe. Report of a WHO Informal Consultation jointly organized by WHO headquarters and the WHO Regional Office for Europe; Geneva, Switzerland, 17–18 December 2009. WHO/HTM/NTD/IDM/2010.1.
5. Dias JC (2007) Southern Cone Initiative for the elimination of domestic populations of Triatoma infestans and the interruption of transfusion Chagas disease: historical aspects, present situation, and perspectives. Mem Inst Oswaldo Cruz 102: 111–118.
6. Salvatella R (2007) Andean subregional Chagas disease area and the Andean Initiative of Chagas Disease. Mem Inst Oswaldo Cruz 102: e29–e30.
7. Gascon J, Bern C, Pinazo MJ (2009) Chagas disease in Spain, the United States and other non-endemic countries. Acta Trop 115: 22–27.
8. Sabino EC, Salles NA, Carraro AL, Rocha MO, Klatt H, et al. (2010) Enhanced classification of Chagas serologic results and epidemiologic characteristics of seropositive donors at three large blood centers in Brazil. Transfusion 50: 2620–2637. doi: 10.1111/j.1537-2995.2010.02576.x [pii].
9. Zaniello BA, Kessler DA, Vine KM, Grima KM, Weisenberg SA (2012) Risk of seropositivity from blood bank testing in endemic and increasingly non-endemic countries. Acta Trop 115: 22–27.
10. Biolo A, Ribeiro AL, Clausell N (2010) Chagas cardiomyopathy–where do we stand after a hundred years? Prog Cardiovasc Dis 52: 300–316.
11. Porto CC (1964) O eletrocardiograma no prognóstico e evolução da Doença de Chagas. Arq Bras Cardiol 17: 313–346.
12. Dias JC, Klotzki E (1960) The prognostic value of the electrocardiographic features of chronic Chagas’ disease. Rev Inst Med Trop Sao Paulo 10: 158–162.
13. Maguire JH, Hoff R, Sherlock I, Guimaraes AC, Sleegh AC, et al. (1987) Cardiac morbidity and mortality due to Chagas’ disease: prospective electrocardiographic study of a Brazilian community. Circulation 75: 1140–1145.
14. Williams-Bangero S, Magalhaes T, Rainwater E, Blanqero J, Correa-Oliveira R, et al. (2007) Electrocardiographic characteristics in a population with high rates of seropositivity for Trypanosoma cruzi infection. Am J Trop Med Hyg 77: 495–499.
15. Gonçalves JG, Prata A, Dias JC, Macedo V (2011) [The electrocardiographic survey]. Rev Soc Bras Med Trop 44 Suppl 2: 40–46. S0037-8682(11)00008-0 [pii].
16. Nascimento BR, Araujo CG, Rocha MO, Domingues JD, Rodrigues AR, et al. (2012) The prognostic significance of electrocardiographic changes in Chagas disease. J Electrocardiol 45: 43–48. S0022-0736(11)00155-5 [pii];10.1016/j.jelectrocard.2011.04.011 [doi].
17. Valério L, Roure S, Sabria M, Balanzó X, Valles X, et al. (2011) Clinical, electrocardiographic and echocardiographic abnormalities in Latin American migrants with newly diagnosed Chagas disease 2005-2009, Barcelona, Spain. Euro Surveill 16: pii: 19971.
18. Princeas, R J., Crow, R S., and Blackburn H. (1982) The Minnesota Code Manual of Electrocardiographic Findings. Littleton, MA: John Wright-PSG.
19. Sabino EC, Ribeiro AL, Salles NA, Oliveira CD, Anneses AP, et al. (2013) Ten-year Incidence of Chagas cardiomyopathy among asymptomatic, T. cruzi seropositive former blood donors. Circulation. In Press.
20. Salles NA, Sabino EC, Chiquet MG, Haef-Neuj, J, Mayer A, et al. (1996) Risk of exposure to Chagas’ disease among seroreactive Brazilian blood donors. Transfusion 36: 969–973.
21. Sabino EC, Lee TH, Montalvo L, Nguyen ML, Leiby DA, et al. (2012) Antibody levels correlate with detection of Trypanosoma cruzi DNA by sensitive polymerase chain reaction assays in seropositive blood donor and possible resolution of infection over time. Transfusion E-pub. 2011.10.1111/j.1537-2995.2012.03902.x [doi].
22. Denes P, Larson JC, Lloyd-Jones DM, Princeas RJ, Greenland P (2007) Major and minor ECG abnormalities in asymptomatic women and risk of cardiovascular events and mortality. JAMA 297: 978–983. S0022-0737(07)00351-7 [pii];10.1001/jama.297.9.978 [doi].
23. Lang RM, Bierig M, Deverux RB, Fleckskamp FA, Foster E, et al. (2005) Recommendations for chamber quantification: a report from the American College of Cardiology/American Heart Association Task Force on Chamber Quantification. J Am Soc Echocardiogr 18: 1440–1511.
35. Espinosa RA, Pericchi LR, Carrasco HA, Escalante A, Martinez O, et al. (1991)

34. Guerrero L, Carrasco H, Parada H, Molina C, Chuecos R (1991) [Ventricular...

33. Rincon LG, Rocha MO, Nunes MC, Rocha MO (2009) Old age as an emerging issue: 10 year follow-up of the Bambui population-based cohort study (Brazil). Int J Cardiol 145: 302–303.

32. Rodriguez-Salas LA, Klein E, Acquatella H, Catalioti F, Davalos V, et al. (1998) Electrocardiographic ventricular repolarization parameters in chronic Chagas' disease as predictors of asymptomatic left ventricular systolic dysfunction. Pacing Clin Electrophysiol 26: 1326–1335.

31. Ribeiro AL, Moraes RS, Ribeiro JP, Ferlin EL, Torres RM, et al. (2001) Cardiovascular autonomic function in Chagas' disease. Sao Paulo Med J 119: 802–813.

30. Rocha MO, Nunes MC, Ribeiro AL (2009) Morbidity and prognostic factors in chronic chagasic cardiopathy. Int J Cardiol 130: 195–202.

29. Lima-Costa MF, Peixoto SV, Ribeiro AL (2010) Chagas disease and mortality in old age as an emerging issue: 10 year follow-up of the Bambui population-based cohort study (Brazil). Int J Cardiol 145: 302–303. S0167-5273(10)00103-8 [pii];10.1016/j.ijcard.2010.02.036 [doi].

28. Salles GF, Cardoso CR, Xavier SS, Sousa AS, Hasslocher-Moreno A, Cardoso C (2003) Prognostic value of QT interval parameters for mortality risk stratification in Chagas' disease: results of a long-term follow-up study. Circulation 108: 305–312.

27. Salles G, Xavier S, Sousa A, Hasslocher-Moreno A, Cardoso C (2003) Relationship of electrocardiographic abnormalities and seropositivity to Trypanosoma cruzi within a rural community in Northeast Brazil. American Heart Journal 103: 267–294. doi: 10.1016/S0002-0703(03)90529-X.

26. Garzon SA, Lorga AM, Nicolau JC (1995) Electrocardiography in Chagas' heart disease. Sao Paulo Med J 113: 802–813.

25. Arteaga-Fernandez E, Barreto AC, Mady C, Ianni BM, Bellotti G, et al. (1985) [The electrocardiogram in patients with positive serological reactions for Chagas’ disease. Study of 600 cases]. Arq Bras Cardiol 44: 333–337.

24. Maguire JH, Mott KE, Lehman JS, Hoff R, Muniz TM, et al. (1983) Relationship of electrocardiographic abnormalities and seropositivity to T. cruzi Seropositive Former Blood Donors