Isolation, Identification and Evaluation of Antimicrobial Activity of *Streptomycetes flavogriseus*, strain ACTK2 from Soil Sample of Kodagu, Karnataka State (India)

Nooshin Khandan Dezfully¹; Janardhana Gottravalli Ramanayaka ²,

¹Department of Studies in Microbiology, University of Mysore, Manasagangotri, Karnataka, India
²Mycology and Phytopathology Laboratory, Department of Studies in Botany, University of Mysore, Manasagangotri, Karnataka, India

Received: September 28, 2013; Revised: January 30, 2014; Accepted: February 2, 2014

Background: The search for novel antibiotics continues to be of immense importance in research programs around the world for pharmaceutical, industrial and agricultural applications. filamentous soil bacteria, belonging to the *Streptomycetes* genus, are widely used as an important biological tool for their ability to produce a wide range of novel secondary metabolites, such as "antibiotics".

Objectives: The aim of the present study was to isolate and identify a strain of *Streptomycetes* with high antibiotic production capability.

Materials and Methods: The soil sample was collected randomly from the agricultural land of Kushalnagar Taluk of Kodagu district, Karnataka, India. The ACTK2 strain was isolated by serial dilution method and identified based on cultural, morphological, microscopic, biochemical and sequence analysis of 16S rRNA gene parameters. The isolated ACTK2 was analyzed for antimicrobial activities by perpendicular streak and disc diffusion methods, against the Gram-positive bacteria *Staphylococcus aureus* (MTCC 96), *Bacillus subtilis* (MTCC 121), Gram-negative *Escherichia coli* (MTCC 729), *Enterococcus aerogenes* (MTCC 2829) and filamentous fungi (*Trichoderma harzianum* (MTCC 6046), *Fusarium proliferatum* (MTCC 9375)). Further, an antimicrobial metabolite from the ACTK2 strain was extracted by solvent extraction method, using n-butanol. The production of the antimicrobial compound by the ACTK2 strain was optimized by using different nutritional media and cultural conditions.

Results: The strain *Streptomycetes flavogriseus* designated as ACTK2 (Accession number KC990785) isolated from the soil sample of Kushalnagar Taluk, Kodagu, Karnataka, India, exhibited a broad spectrum of antimicrobial activity against test microorganisms. The optimum growth and antimicrobial compound production by strain ACTK2 was found to be a maximal pH 8, in the shaker incubator at 28ºC, for a period of 10 days.

Conclusions: The crude n-butanol extract of the ACTK2 strain of *S. flavogriseus* showed a broad spectrum of antimicrobial activities against the test organisms and this opened further research investigations on purification and structural characterization of the active compounds from the crude extract.

Keywords: Streptomycetaceae; India; Strain; Antibiotics

1. Background

The search for isolating novel antibiotics, effective against resistant pathogenic microorganisms from unexplored habitats around the world, continues to be an important sector of research. There is an emerging crisis secondary to the development of antibiotic resistant microbial pathogens and toxicity of several antibiotics in use (1, 2). As a result of the extensive screening, about 17000 antibiotics from different microbial sources have been isolated (3). The role of actinomycetes, as a source of bioactive molecule compounds, are well known, as these organisms, especially *Streptomycetes*, provides more than half (70%) of the naturally occurring antibiotics extensively used by the pharmaceutical industry (3, 4).

During the past two decades, there has been a decline in the discovery of new compounds from soil-derived actinomycetes (5). Hence, in the quest for exploring newer ecosystems for the isolation of actinomycetes and to discover high efficacy antibiotics, it is crucial to overcome the problem of antibiotic resistance. The Western Ghats region of India is being explored for its rich and varied heritage of microbial diversity. Several strains of actinomycetes with antimicrobial activity have been isolated and characterized from soil samples of Coimbatore, Kerala, Tamil Nadu, India (6-10). However, there is a lack of adequate data on isolated actinomycetes strains with potential antibiotic properties from the Kodagu region of Karnataka.

2. Objectives

The objective of the present study was to isolate, identify and evaluate the antimicrobial activity for optimiza-
tion of the strain Streptomyces obtained from a soil sample of Kodagu district of Karnataka, India.

3. Materials and Methods

3.1. Sample Collection

Soil samples were collected randomly from agricultural lands of Kushalnagar Taluk, Kodagu district, Karnataka, India during 2011 using an open-end soil borer (20 cm in depth and 25 cm in diameter) from a depth of 10 - 20 cm, which were then air dried for 24 to 48 hours (11). The soil sample was then treated with 1% CaCO$_3$ and kept at ambient temperature for 2 - 3 days (12) for the next step of analysis.

3.2. Isolation of Actinomycetes

Soil samples were serially diluted (13) and plated on Actinomycetes Isolation Agar medium procured from Difco (Hi media, Mumbai, India) and incubated at 28˚C for 6 days. The suspected colonies were picked up and purified on yeast extract-malt extract agar medium (ISP-2) supplemented with 50 µg/mL Nystatin, as antifungal agent (14). The pure culture of ACTK2 was maintained on ISP-2 slants at 4˚C. The spore mass and mycelium of the pure isolate were stored at -20˚C as glycerol suspension (20%, v/v) for further analysis (Difco, Hi media, Mumbai, India) (15).

3.3. Taxonomical Characterization of the Strain ACTK2

The strain of ACTK2 was identified based on cultural, morphological, physiological, biochemical and sequence analysis of 16S rRNA gene parameters.

3.3.1. Cultural and Morphological Characteristics

The cultural characteristic of pure isolate was studied after incubation of strain ACTK2 for 7-14 days at 28˚C using various media: Tryptone Yeast Extract Agar (ISP-1), Yeast Extract-Malt Extract Agar (ISP-2), Oat Meal Agar (ISP-3), Starch-Casein nitrate Agar medium (ISP-4), Glycerol-Asparagine Agar (ISP-5), Peptone Yeast Extract Iron Agar (ISP-6), Modified ISP-2 (MISP-2), Starch Agar, Potato Dextrose Agar, Czapek Dox Agar, Nutrient Agar, Modified Nutrient Glucose Agar (MNGA) and Mueller Hinton Agar, procured from Hi-Media, Mumbai, India. Morphological parameters such as aerial and substrate mycelium, arrangements of spores and Gram staining reaction of the ACTK2 strain (16) were carried out as described in Bergey’s Manual of Systematic Bacteriology and by Shirling and Gottlieb (17, 18).

3.3.2. Physiological and Biochemical Characteristics

Various tests, such as the catalase test, citrate utilization test, nitrate reduction test, starch hydrolysis (19), gelatin hydrolysis (20) and production of melanin pigmentation on ISP-6 (18), using standard media from Hi media, Mumbai, India, were carried out to analyze the physiological and biochemical characteristics of ACTK2. The analysis of 2-6 diaminopimelic acid (DAP) isomers, present in the whole cell wall composition of the strain ACTK2, was determined by thin layer chromatography (5 D Fine-Chem Ltd., Mumbai, India) (21).

3.3.3. Molecular Identification of ACTK2

The isolated ACTK2 was grown by agitation in 500 mL flasks containing 100 mL of ISP-2 medium for 4 days at 28˚C. After the incubation, biomass was harvested by centrifugation at 10000 rpm for 5 minutes and washed twice with double-distilled water. The genomic DNA of ACTK2 was isolated using the AMpurE Bacterial Genomic DNA Mini kit (Ammon’s Bioscience, Bangalore, India) (22). The formed DNA pellet was air dried and dissolved in 100 µL of TE buffer (100 mM NaCl, 1 mM EDTA, 100 mM Tris-HCl, pH 8.0) and used for polymerase chain reaction (PCR) studies. The 16S ribosomal RNA gene was amplified by using PCR with Taq DNA polymerase and universal primer pairs 27F (5’-AGAGTTTGATCCTGGCTCAG-3’) and 1492R (5’-GTTACCTTGTAGGACTC-3’) (23).

Amplification was performed using Thermal Cycler (Peqlab Biotechnologie GmbH, Erlangen, Germany), in a total volume of 50 µL containing 50 ng/mL DNA, 100 pmol of each primer, 10 mM dNTPs, 2 µL10X Taq Polymerase buffer and 0.25 U Taq DNA polymerase (Ammon’s PCR Master-Mix kit, Ammon’s Bioscience, Bangalore, India). The PCR conditions included, denaturation at 94°C for 5 minutes followed by 35 cycles of denaturation at 94°C for 30 seconds, primer annealing at 55°C for 30 seconds and primer extension at 72°C for 1.5 minutes and final extension at 72°C for 5 minutes. The PCR products were detected by 1.0% (w/v) agarose gel containing ethidium bromide (0.5 µg/mL) and were visualized by the ultraviolet (UV) fluorescence gel documentation system (UTP-Bio Doc, USA).

3.3.3.1. Cloning and Sequencing

The PCR products were cloned in the pGEM-T Easy Vector (Promega Corp., Fitchburg, Wisconsin, USA) and transformed to Escherichia coli cells for nucleotide sequence determination using an automated sequencer (Genetic Analyzer 3130, Applied Biosystems, Waltham, Massachusetts, USA). The nucleotide sequences analysis of the 16S rRNA gene of ACTK2 was compared with the reference species of bacteria contained in the GenBank database, using the NCBI BLAST available at http://blast.ncbi.nlm.nih.gov website.

3.3.3.2. Phylogenetic Analysis of Isolate ACTK2

The phylogenetic tree was derived from the distance matrices using the neighbor-joining method (24). All the
analyses were performed on a bootstrap dataset containing 100 replicates (generated by the program) (24).

3.4. Antimicrobial Activity

The test organisms employed included the following: Gram-positive *Staphylococcus aureus* (MTCC 96), *Bacillus subtilis* (MTCC 9375) and Gram-negative *E. coli* (MTCC 729), *Enterococcus aerogenes* (MTCC 2829). All the test organisms used in this study were obtained from the Microbial Type Culture Collection (MTCC), Institute of Microbial Technology (IMTECH), Chandigarh (India). The filamentous fungi *Trichoderma harzianum* (MTCC6046) and *Fusarium proliferatum* (MTCC 9375) were obtained from the Department of Studies in Microbiology and Department of Studies in Botany, University of Mysore, Mysore, Karnataka, India. Primary screening of antimicrobial activity of *Streptomyces flavogriseus* (ACTK2) was determined by using the perpendicular streak technique (25). Colonies were streaked on the center of ISP-2 plates and incubated at 28°C for 4 - 5 days. The test organism was inoculated perpendicular to the strain ACTK2 and the plate was incubated at 37°C for 24 hours for bacteria and at 28°C for 96 hours for fungi (8). The zone of inhibition was determined using a millimeter scale.

3.5. Optimization of Fermentation Parameters

To determine the optimal nutritional and cultural conditions for the growth and formation of antimicrobial compound by the ACTK2 strain using different media, such as ISP-2 Broth, Starch Casein Broth, MISP-2 Broth, ISP-4 Broth and MNG Broth were used. Culture conditions at different pH values (4, 6, 8 and 10), temperatures (20, 28, 37 and 45°C) and incubation periods (3, 4, 5, 7, 10 and 15 days) were studied for microbial growth and production of antimicrobial compound. Fermentation was carried out in an 250 mL Erlenmeyer flask, containing 50 mL of medium inoculated with 100 µL of the spore suspension of the strain ACTK2 (1.5 × 10^5 CFU/µL) and incubated in an orbital shaker at 100 rpm (26).

3.6. Determination of dry Weight of Mycelium

Growth of the isolate ACTK2 was measured as dry weight of the mycelium after 3, 4, 5, 7, 10 and 15 days of incubation. The biomass was separated after centrifugation at 4000 rpm for 10 minutes and transferred to a pre-weighted dry filter paper using a clean spatula and was kept in an oven at 60°C for 12 hours to reach a constant weight. The amount of growth was expressed as mg/50 mL culture medium (27).

3.7. Extraction of Antimicrobial Compound From ACTK2

The antimicrobial compound was extracted from the mycelium-free supernatant by the solvent-liquid extraction method (28). N-butanol was added to the supernatant in the ratio of 1:4(v/v) and shaken vigorously. The butanol phase having antimicrobial compound was separated and the extraction process was repeated three times. The n-butanol phase was concentrated by using a rotary vacuum flash evaporator at 60°C, 200 rpm. The obtained residue (crude extract) was dissolved in 1 mL of n-butanol solvent and stored at 4°C.

3.7.1. Determination of the Antimicrobial Activity

The antimicrobial activity of the n-butanol extract of ACTK2 was determined by the disc diffusion method (29) against the test organisms. The amount of 20 mL of sterilized molten Muller Hinton Agar was seeded with 50 µL of test organisms (1.5 × 10^5 CFU/µL), swirled gently and aseptically poured into Petri dishes and allowed to solidify. Sterile filter paper disc containing 50 µL of the crude solvent extract was placed at the center of the Petri dish. Control was maintained by loading 50 mL of n-butanol in place of crude extract. The plates were kept at 4°C for 2 hours for diffusion of the antibiotic and then incubated at 37°C for bacteria and at 28°C for 48 hours for fungi. The inhibition zone around the disk was measured with a millimeter scale and the results were tabulated and analyzed.

4. Results

The aerobic actinomycete designated as strain ACTK2 (accession number KC990785) isolated from the soil sample of Kushalnagar Taluk, Kodagu district, Karnataka, India, was a Gram-positive, *S. flavogriseus*. The culture was a slow growing while the filamentous aerial mycelium was grayish to whitish yellow in color producing straight to flexuous chains of spores. The sporulation on the agar medium was observed after 2 - 3 days of incubation (Table 1). No diffusible pigment was detected on the used media, except for ISP-3 medium. These morphological characteristics strongly suggested that the ACTK2 strain belonged to the genus *Streptomyces*. The physiological and biochemical characteristics of ACTK2 are presented in Table 2. The ACTK2 hydrolyzed Starch, reduced Nitrate, liquefied Gelatin, but it did not hydrolyze Citrate and produced Hydrogen Sulphide (H₂S). The ACTK2 strain was Catalase positive and Oxidase negative. Melanin production was not observed on ISP-6. The cell wall composition of the strain ACTK2 was found to be L-DAP (cell wall chemotype-1) which had characteristics of the genus *Streptomyces*.

The results of the PCR and sequencing of the strain ACTK2 are presented in Figures 1 and 2. The alignment of the nucleotide sequences (1489 bp) of strain ACTK2 in the gene bank using NCBI exhibited a similarity level of 100% with *S. flavogriseus*. The phylogenetic tree obtained by applying the neighbor joining method is shown in Figure 3. Primary screening of ACTK2 for its antimicrobial activity showed a broad spectrum antimicrobial
Table 1. Cultural Characteristics of the *Streptomyces flavogriseus* Isolate ACTK2 on Different Media After 14 Days of Incubation at 28 °C.\(^a\)

| Sl. No. | Medium   | Growth | Aerial Mycelium       | Substrate Mycelium | Spores   | Soluble Pigment |
|---------|----------|--------|-----------------------|--------------------|----------|-----------------|
| 1       | ISP-1    | Good   | Moderate, Ashes       | Dark brown         | Ashes    | None            |
| 2       | ISP-2    | Good   | Abundant, Ashes       | Dark brown         | Ashes    | None            |
| 3       | ISP-3    | Good   | Abundant, Gray        | Black              | Gray     | Brown           |
| 4       | ISP-4    | Good   | Abundant, Gray        | Black              | Gray     | Light brown     |
| 5       | ISP-5    | Good   | Moderate, Whitish gray| Whitish yellow     | Grayish white | Light brown     |
| 6       | ISP-6    | Poor   | None                  | Yellowish brown    | None     | Medium yellow brown |
| 7       | MNGA     | Moderate | Poor, Light gray    | Whitish gray       | Whitish gray | None            |
| 8       | NA       | Moderate | Poor, Light gray    | Whitish gray       | Gray     | None            |
| 9       | MHA      | Moderate | Moderate, Grayish white | Whitish gray       | Gray     | None            |
| 10      | PDA      | Moderate | Poor, Creamish yellow | Whitish yellow     | Whitish gray | None            |
| 11      | Starch Agar   | Moderate | Poor, Whitish yellow | Whitish yellow     | Whitish gray | None            |
| 12      | CZP      | Good   | Abundant, gray        | Dark brown         | gray     | Brownish black  |
| 13      | MISP-2   | Good   | Abundant, ashes       | Dark brown         | ashes    | Light brown     |

\(^a\) Abbreviations: ISP-1-6: International Streptomyces Project medium; MNGA: Yeast Extract-Malt Extract Dextrose Agar; NA: Nutrient Agar; MHA: Muller-Hinton Agar; PDA: Potato Dextrose Agar; CZP: Czapek Dox Agar; MISP-2: Modified Yeast Extract-Malt Extract Agar.

Table 2. Morphological, Biochemical and Chemotaxonomic Characteristics of *Streptomyces flavogriseus* ACTK2 Isolated From Soil Samples of Kodagu District, Karnataka (India).\(^a\)

| Sl. No, Properties | *Streptomyces flavogriseus* ACTK2 |
|--------------------|----------------------------------|
| **I. Morphological characteristics** | | |
| 1. Sporophore morphology | Flexible spiral |
| 2. Color of aerial mycelium | Ashes gray |
| 3. Color of substrate mycelium | Dark gray |
| 4. Spore mass | Ashes gray |
| **II. Biochemical characteristics** | | |
| 1. Catalase test | + |
| 2. Oxidase test | - |
| 3. Citrate utilization | - |
| 4. Gelatin hydrolysis | + |
| 5. Nitrate reduction | + |
| 6. Starch hydrolysis | + |
| 7. Urease test | + |
| 8. Melanin production | - |
| **III. Chemotaxonomic characters** | | |
| 1. Cell wall amino acid analysis | L-DAP |
| **IV. Grams staining** | + |

\(^a\) Abbreviation: L-DAP, L-diaminopimelic acid.

Figure 1. Agarose gel (1%) Showing PCR Products (1.5 Kb) of ACTK2 strain *S. flavogriseus*, Along with Marker DNA (2.0 kb).
activity against test bacteria and *F. proliferatum*. The strain ACTK2 did not show any activity against *T. harizianum* (Figure 4). The n-Butanol extraction of ACTK2 showed maximum inhibitory activity against *E. coli* followed by *B. subtilis*, *S. aureus*, *F. proliferatum* and *S. aureus*, respectively. Among the Gram-positive and Gram-negative bacteria, *E. coli* and *B. subtilis* were the most sensitive to antimicrobial compounds produced by ACTK2. The optimization studies revealed that ACTK2 was able to grow in all the tested culture media. However, the maximum growth (biomass) and production of antimicrobial compound was observed in MISP-2 Broth with an average dry weight of mycelium (0.8 mg/50 mL) followed by Inorganic salts-Starch Broth (ISP-4) (0.7 mg/50 mL), Starch Casein Broth (0.4 mg/50 mL), Yeast Extract-Malt Extract Glucose Broth (ISP-2) (0.3 mg/50 mL) and Modified Nutrient Glucose Broth (MNB) (0.2 mg/50 mL), respectively (Figure 5).

The ACTK2 could grow and produce the antimicrobial compound up to a pH 10.0. The maximum production was observed at a pH 8.0 (Figure 6). The results suggest that strain ACTK2 was slightly alkalophilic in nature. The ACTK2 showed a narrow range of incubation temperature for relatively good growth and antimicrobial production. Higher growth (0.8 mg/50 mL), as well as antimicrobial activity, were noticed against *B. subtilis* at 28˚C (Figure 7), indicating that ACTK2 was mesophilic in nature. An incubation period of up to 10 days was found to be optimal for maximum growth (0.8 mg/50 mL), as well as antimicrobial production (Figure 8). The results of optimization of nutritional and cultural conditions showed that n-butanol extract of strain ACTK2 exerted maximum activity against *E. coli* (16 mm), followed by *B. subtilis* (15 mm), *F. proliferatum* (13 mm), *Staph. aureus* (13 mm) and *Staph. aureus* (11 mm).

**Figure 2.** Partial Sequence (1499bp) of 16r RNA of *S. flavogriseus*, strain ACTK2.
Figure 3. 16S rRNA tree showing the phylogenetic relationship by neighbor-joining method between strain ACTK2 (Accession number KC990785) with other known Streptomyces species.

Figure 4. The Antimicrobial Activity of S. flavogriseus ACTK2. A) Plate showing ACTK2 inhibiting the growth of tested bacteria B) ACTK2 inhibited the growth of F. proliferatum by perpendicular streak method.

5. Discussion

The emerging crisis of antibiotic resistant pathogens indicates an increasing necessity for the survey of unexplored and underexplored niche habitats. The isolation of actinomycetes can contribute to the discovery of novel, safe, effective, broad-spectrum antimicrobial compounds, as part of the strategy to control the drug-resistant pathogens (30, 31). During the course of screening of bioactive compounds for the isolation of new antibiotics each year, thousands of actinomycetes, particularly Streptomyces strains, are screened by pharmaceutical research laboratories as sources for novel antimicrobial compounds (8, 27, 32-34). In the present study, a broad spectrum of antimicrobial producing actinomycetes, designated as S. flavogriseus (ACTK2) isolated from agricultural soil sample of Kushalnagar, Kodagu, Karnataka state, India were selected for optimization of their antimicrobial activities. The isolate ACTK2 was identified through conventional and molecular methods.
The isolate exhibited broad antimicrobial activities against Gram-positive, Gram-negative bacteria and fungus *F. proliferatum* in the primary and secondary screening process. In a study conducted in Iran (35) with different strains of *Streptomyces*, there was no significant correlation between the activity of intact bacteria in primary screening and their extract in secondary screening, which was similar to the present study on ACTK2.

The results of the optimal nutritional media showed that antibiotic production was higher in the medium containing glucose. The importance of glucose in the nutritional medium for the synthesis of a wide range of antibiotics by different *Streptomyces* species has been reported by many investigators (27, 36-38). Although the strain ACTK2 was able to grow in five different media tested, the highest growth (0.8 mg/50 mL) and antimicrobial activity were obtained in MISP-2 Broth medium supplemented with starch (as a carbon source) and CaCO₃ (as growth promoters). This result is quite comparable with the *S. fulvissimus* and *Pseudonocardia* species, for which MISP-2 was found to be a suitable medium for the antibiotic production (39, 40). Optimal cultural conditions at different pH levels and incubation temperatures and time for antimicrobial production by *S. flavogriseus* improved in the medium containing MISP-2 Broth.

The results revealed that the maximum growth (0.8 mg/50 mL) and in-vitro antimicrobial compound production by *S. flavogriseus* could be obtained in MISP-2 medium having a pH 8, for 10 days of incubation at 28°C. The growth profile, closely coupled with the metabolic capacities of the produced organism *S. flavogriseus*, which greatly influenced the biosynthesis of antibiotics. The findings confirm that the nature of the medium composition and cultural conditions strongly affect and enhance the antimicrobial compound of ACTK2, which is comparable to the surveys conducted on different *Streptomyces* spp. by other investigators (9, 27, 32-34, 36). In this study, the *S. flavogriseus* designated as ACTK2, isolated from agricultural soil sample of Kushlanagar Taluk of Kodagu, Karnataka, India, showed broad spectrum antimicrobial activity against Gram-positive, Gram-negative bacteria and the fungus *F. proliferatum*. Further studies on the purification and chemical characterization of the antimicrobial compound from the strain of *S. flavogriseus* will be useful for pharmaceutical applications.

References
1. Spellberg B, Powers JH, Brass EP, Miller LG, Edwards JJ. Trends in antimicrobial drug development: implications for the future. *Clin Infect Dis.* 2004;38(9):1279-86.
2. Bull AT, Ward AC, Goodfellow M. Search and discovery strategies for biotechnology: the paradigm shift. *Microbiol Mol Biol Rev.* 2000;64(3):573-606.
3. Berdy J. Bioactive microbial metabolites. *J Antimicrob Chemother.* (Tokyo). 2005;58(1):23-26.
4. Saadoun I, Gharailieh R, The Streptomyces flora of Badia region of Jordan and its potential as a source of antibiotics active against antibiotic-resistant bacteria. *J Arid Environ.* 2003;53(3):365-71.
5. Mincer TJ, Jensen PR, Kauffman CA, Fenical W. Widespread and persistent populations of a major new marine actinomycete taxon in ocean sediments. *Appl Environ Microbiol.* 2002;68(10):5005-11.
6. Sundaramoorthy C, Vengadesh PK, Gupta S, Kartich K, Tamilselvi N. Production and characterization of antibiotics from soil-isolated actinomycetes. *Int J Pharm.* 2011;42(4):314-8.
7. George M, George G, Hatha AAM. Diversity and antibacterial activity of actinomycetes from wetland soil. *South Pac J Nat Appl Sci.* 2010;28(1):52.
8. Arasu MV, Duraihandiyavan V, Agastian P, Ignacimuthu S. Antimicrobial activity of Streptomyces spp. *Curr Sci* recovered from Western Ghats of Tamil Nadu. *J Med Microbiol.* 2008;57(1):347-53.
9. Valas Arasu M, Duraihandiyavan V, Agastian P, Ignacimuthu S. In vitro antimicrobial activity of Streptomyces spp. *Curr Sci* isolated from Western Ghats rock soil (India). *J Med Microbiol.* 2009;57(1):22-8.
10. Santhanam P, Malisamani Selvam M. Antimicrobial activity of actinomycetes isolated from The Western Ghats of Tamilnadu. *Int J Pharm Sci* 2012;4(1).
11. Saadoun I, al-Momani F, Malkawi HI, Mohammed MJ. Isolation, identification and analysis of antibacterial activity of soil streptomycetes isolates from north Jordan. *Microbiol.* 1999;300(39):41-6.
12. Tsao PH, Leen C, Keitt GW. An enrichment method for isolating Actinomycetes that produce diffusible anti-fungal antibiotics. *Phytopathol.* 1960;50(8):388-9.
13. El-Nakeeb MA, Lechevalier HA. Selective isolation of aerobic Actinomycetes. *Appl Microbiol.* 1961;31(7):7-7.
14. Williams ST, Davies FL. Use of Antibiotics for Selective Isolation and Enumeration of Actinomycetes in Soil. *J Gen Microbiol.* 1965;38:253-63.
15. Welllington EMH, Williams ST. Preservation of actinomycete inoculum in frozen glycerol. *Microbiol Lett.* 1977;6(23):451-7.
16. Hucker GJ, Conn HJ. Methods of Gram staining. *Technical Bulletin of the New York State Agricultural Experimental Station.* 1923.
17. Williams ST, Sharpe ME, Holt JG. *Bergey’s Manual of Systematic Bacteriology* 2nd ed; Baltimore: Williams and Wilkins; 1989.
18. Shirling EB, Gottlieb D. Methods for characterization of *Streptomyces* species. *International Journal of Systematic Bacteriology.* 1966;36(1):313-40.
19. Gordon RE. The taxonomy of soil bacteria. In: Gray TRG, Parkison D editors. *The Ecology of soil bacteria*. Liverpool: Liverpool University Press; 1968.
20. Waksman SA. The actinomycetes – their nature, occurrence, activities, and importance. *J Am Med Assoc.* 1950;144(6):505-6.
21. Hasegawa T, Takizawa M, Tanida S. A rapid analysis for chemical grouping of aerobic actinomycetes. *J Gen Microbiol.* 1981;129(4):339-22.
22. Pospiech A, Neumann B. A versatile quick-prep of genomic DNA from gram-positive bacteria. *Trends Genet.* 1995;11(6):237-8.
23. Weisburg WG, Barns SM, Pelletier DA, Lane DJ. *16S rDNA amplification for phylogenetic study.* *J Bacteriol.* 1991;173(2):697-703.
24. Saitou N, Nei M. The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol.* 1987;4(4):406-25.
25. Egorov NS. *Antibiotics: A Scientific Approach.* Moscow: Mir Publishers; 1985.
26. Shuler ML, Kargi F. *Bioprocess engineering, basic concept.* New Jersey: Prentice-Hall Inc; 1992.
27. Singh LS, Mazumder S, Bora TC. Optimisation of process parameters for growth and bioactive metabolite produced by a salt-tolerant and alkaliophilic actinomycete, Streptomyces tanashiensis strain A2D. *J Med Microbiol.* 2009;58(4):225-33.
28. Liu CM, Westley JW, Herrman TE, Prosser BL, Palleroni N, Evans RH, et al. Novel polyether antibiotics X-14873A, G and H produced by a *Streptomyces*: taxonomy of the producing culture, fermentation, biological and ionophorous properties of the antibiotics. *J Antimicrob Chemo* (Tokyo). 1986;29(2):1271-2.
29. Bauer AW, Kirby YM, Sherris JC, Rusk M. Antibiotic susceptibility testing by a standardized single disk method. *Am J Clin Pathol.* 1966;45(4):493-9.
30. Demain AL, Sanchez S. Microbial drug discovery: 80 years of progress. *J Antimicrob Chemo* (Tokyo). 2009;62(1):3-16.
31. Livermore DM, British Society for Antimicrobial Chemotherapy Working Party on The Urgent Need: Regenerating Antibacterial Drug D, Development. Discovery research: the scientific challenge of finding new antibiotics. J Antimicrob Chemother. 2011;66(4):3941-4.
32. Laidi Rahah F, Elshafei A, Saker M, Cheikh B, Hocine H. Screening, Isolation and Characterization of a Novel Antimicrobial Producing Actinomycete, Strain RAf10. Biotechnology(Faisalabad). 2007;6(4):489-96.
33. Elleuch L, Shaaban M, Smaoui S, Mellouli I, Karray-Rebai I, Fouarati-Ben Fguira L, et al. Bioactive secondary metabolites from a new terrestrial Streptomyces sp. TN262. Appl Biochem Biotechnol. 2010;162(2):579–93.
34. Thumar JT, Dhulia K, Singh SP. Isolation and partial purification of an antimicrobial agent from halotolerant alkaliphilic Streptomyces aburaviensis strain Kut-8. World Journal of Microbiology and Biotechnology. 2000;26(11):2081–7.
35. Maleki H, Dehnad A, Hanifian S, Khani S. Isolation and Molecular Identification of Streptomyces spp. with Antibacterial Activity from Northwest of Iran. Bioimpacts. 2013;3(3):329–34.
36. Sujatha P, Bapi Raju KV, Ramana T. Studies on a new marine streptomycete BT-408 producing polyketide antibiotic SBR-22 effective against methicillin resistant Staphylococcus aureus. Microbiol Res. 2005;160(2):319–26.
37. Pandey A, Shukla A, Majumdar SK. Utilization of carbon and nitrogen sources by Streptomyces kanamyceticus M 27 for the production of an Anti bacterial antibiotic. Afr J Biotechnol. 2005;4(9):309-10.
38. Vasavada SH, Thumar JT, Singh SP. Secretion of a potent antibiotic by salt-tolerant and alkaliphilic actinomycete Streptomyces sannanensis strain RJF-3. Curr Sci. 2008;91(10):1393-7.
39. Malik H, Sur B, Singhal N, Bihari V. Antimicrobial protein from Streptomyces fulvissimus inhibitory to methicillin resistant Staphylococcus aureus. Indian J Exp Biol. 2008;46(4):254-7.
40. Usha Kiranmayi M, Sudhakar P, Sreenivasulu K, Vijayakrishna M. Optimization of Culturing Conditions for Improved Production of Bioactive Metabolites by Pseudonocardia sp. VUK-10. Mycobiology. 2011;39(3):174-81.