Dual-Targeted Theranostic Delivery of miRs Arrests Abdominal Aortic Aneurysm Development

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Abdominal aortic aneurysm (AAA) is an often deadly disease without medical, non-invasive treatment options. The upregulation of vascular cell adhesion molecule-1 (VCAM-1) on aortic endothelium provides an early target epitope for a novel biotechnological theranostic approach. MicroRNA-126 was used as a therapeutic agent, based on its capability to downregulate VCAM-1 expression in endothelial cells and thereby reduces leukocyte adhesion and exerts anti-inflammatory effects. Ultrasound microbubbles were chosen as carriers, allowing both molecular imaging as well as targeted therapy of AAA. Microbubbles were coupled with a VCAM-1-targeted single-chain antibody (scFv) toward VCAM-1 (M126) as a therapeutic approach to downregulate VCAM-1 expression and ultimately to deliver anti-inflammatory therapy to the area of AAA development.

For AAA diagnosis/imaging, ultrasound is ideal as it is highly mobile, has no side effects, and provides real-time imaging and overall is highly cost-effective. MBs are US Food and Drug Administration (FDA) approved as ultrasound contrast agents and can be used as drug carriers, allowing localized delivery of drugs upon the selective delivery of an ultrasound burst. We have previously shown that MBs can be used for theranostic approaches, e.g., by conjugation with activated platelet-targeted antibodies and fibrinolytic drugs, allowing simultaneous diagnosis and treatment of acute thrombosis.

Undoubtedly, identifying targets for AAA treatment that non-invasively impede upon aneurysmal growth is of great clinical relevance and has inspired recent novel studies.12 Our approach specifically and non-invasively targets inflamed ECs via a single-chain antibody (scFv) toward VCAM-1. In addition to this scFv-based targeting, ultrasonic bursting provides additional selectivity for transfection of miR-126 mimetics. This unique, dual-targeted...
The delivery system is well suited to provide a high therapeutic payload directly to the site of inflamed endothelium at the AAA.

Overall, the presented combination of molecular imaging of AAA and targeted drug delivery approach holds immense potential for clinical translation in a disease with limited therapeutic options and often fatal outcome.

RESULTS AND DISCUSSION

To allow targeting of MBs, a single-chain antibody targeting mouse VCAM-1 (scFv mVCAM-1) was adapted for MB conjugation by subcloning into the pAC6 vector system, consisting of a C-terminal AviTag, which enables in vivo biotinylation during protein production (Figure 1A). DNA amplification and restriction digest were evaluated by electrophoresis (Figures 1B and 1C), and production of the adapted scFv mVCAM-1 was confirmed by western blotting (Figures 1D and 1E). The non-binding control (scFv Mut) was generated previously. Mouse VCAM-1-expressing Mus musculus axillary lymph node/vascular epithelium (SVEC4-10) cells were used to confirm the binding specificity of scFv mVCAM-1. Binding of commercially available anti-CD106 together with a goat-anti-rat-fluorescein-isothiocyanate (GAR-FITC) secondary antibody confirmed VCAM-1 expression on SVEC4-10 cells after transfection with A126 and decreased expression with M126 as compared to those with S126 (n = 3). Representative flow cytometry dot plots are shown below each bar graph. Representative images show successful transfection of miR-126 using Targ MB via microscopy using bright field and TRITC fluorescence channel; scale bar = 10 µm. Flow cytometry analysis detected Cy3 (on miR) after transfection into SVEC4-10 cells (n = 9). Flow cytometry assays show no change in VCAM-1 expression when non-Targ MB was used for transfection of miR-126. (k) Flow cytometry assays show decreased expression of VCAM-1 after transfection with Targ MB-M126 as compared to Targ MB A126 (n = 5); assays with two groups were analyzed using Student’s t tests and those with more than two groups with equal numbers using repeated-measures one-way ANOVA with Bonferroni post tests.
Three different cholesterol- (for attachment to MBs and transfer through the cell membrane) and fluorescence-tagged miR-126 oligos were used: (1) anti-miR-126 (A126), which induces VCAM-1 expression; (2) mimic-miR-126 (M126), which represses VCAM-1 expression; and (3) scrambled-miR-126 (S126) as control. Changes in VCAM-1 expression were assessed after the respective transfection into SVEC4-10 cells using qRT-PCR and flow cytometry. We observed increased VCAM-1 expression using A126 and decreased expression with M126 as compared to S126 (Figure S1C). Similarly, flow cytometry assays demonstrated significantly more VCAM-1 expression with A126-transfected SVEC4-10 cells and less with M126, as compared to those with S126 (Figure 1G).

MBs were first conjugated with either scFv\textsubscript{VCAM-1} (Targ\textsuperscript{MB}) or non-binding scFv\textsubscript{Mut} (non-Targ\textsuperscript{MB}), followed by incubation with the fluorescence-tagged miR therapeutics. The 3' cholesterol tag has been proven to increase the binding capacity of the miRs to MBs\textsuperscript{1}; however, it is yet to be investigated how long these bonds are stable in vivo in the presence of other hydrophilic blood components. For additional selectivity of our miR therapeutics, we employed an ultrasonic drug delivery approach using ultrasound bursts. After incubation of Targ\textsuperscript{MB} with SVEC4-10 cells and washing, we burst the Targ\textsuperscript{MB} to deliver the Cy3-tagged miR. Fluorescence was visible in the nuclei of cells incubated with Targ\textsuperscript{MB}; however, not with the non-Targ\textsuperscript{MB} (Figures 1H and S1B). Similar results were demonstrated on flow cytometry (Figure 1D).

Following the promising results from our dual-targeting and ultrasonic drug delivery, we investigated whether the expression of VCAM-1 on SVEC4-10 cells could be altered using this approach. Flow cytometry results showed no alteration in VCAM-1 expression of SVEC4-10 cells post-transfection with non-Targ\textsuperscript{MB}-A126, non-Targ\textsuperscript{MB}-M126, non-Targ\textsuperscript{MB}-S126, or non-Targ\textsuperscript{MB} alone as controls (Figure 1J). Similarly, no differences in VCAM-1 expression were observed post-transfection using just Targ\textsuperscript{MB} alone as control or Targ\textsuperscript{MB}-S126 (Figure 1K). However, a significant decrease in VCAM-1 expression was seen in cells transfected using Targ\textsuperscript{MB}-M126 as compared to Targ\textsuperscript{MB}-A126 (Figure 1K), demonstrating the specific targeting ability of our scFv\textsubscript{VCAM-1} and successful ultrasonic drug delivery, which resulted in successful modulation of VCAM-1 expression.

As a first in vivo proof of concept, we validated the efficiency and effectiveness of our Targ\textsuperscript{MB}-M126 using a murine model of acute inflammation intraperitoneal (i.p.) injection of lipopolysaccharide (LPS). Mice were then randomized before being treated with Targ\textsuperscript{MB}-M126, Targ\textsuperscript{MB}-A126 or Targ\textsuperscript{MB}-S126. After 5 min, ultrasound bursts were applied to the abdominal and thoracic aorta. Using qRT-PCR, we observed decreased VCAM-1 gene expression of the thoracic aorta in animals treated with Targ\textsuperscript{MB}-M126 and increased expression in those treated with Targ\textsuperscript{MB}-A126 compared to Targ\textsuperscript{MB}-S126 (Figure 2B). Immunohistochemistry using goat-anti-mouse VCAM-1 polyclonal (GAM-VCAM-1) antibody also showed decreased VCAM-1 expression in animals treated with Targ\textsuperscript{MB}-M126 as compared to those treated with Targ\textsuperscript{MB}-A126 or Targ\textsuperscript{MB}-S126 (Figure 2A).

Following the promising results from the in vivo acute inflammation model, we investigated our targeted theranostic approach for the prevention/treatment of aneurysm severity in an angiotensin (Ang)-II-induced AAA murine model (Figure 2C). Ultrasound monitoring of the abdominal aorta showed an increase in the suprarenal (pre-renal) diameter 1 week post-Ang-II induction for all treatment groups and a slight increase in the infrarenal (post-renal) diameter, as compared to baseline measurements (Figure S2). In order to determine the diagnostic capability of the Targ\textsuperscript{MB} on the abdominal aorta wall, we measured the contrast intensity both before and 20 min post-injection. A significant difference in contrast intensity was observed in the Ang-II-induced AAA animals, 20 min after the injection of Targ\textsuperscript{MB}, demonstrating that our unique VCAM-1-targeted MBs can bind directly at the site of inflamed ECs where AAA develops (Figure 2D). No increase in contrast intensity was seen in healthy PBS control mice (Figure 2E).

We then utilized our theranostic approach of Targ\textsuperscript{MB} loaded with miRs to investigate the impact of Targ\textsuperscript{MB}-M126 upon aneurysm severity. Weekly intravenous injection, ultrasound imaging, and ultrasonic bursting (abdominal aorta, both suprarenal and infrarenal) were performed. Ultrasound measurements of the abdominal aorta 4 weeks post-treatment showed a greater difference in vessel diameter (compared to baseline before disease induction) for animals treated with Targ\textsuperscript{MB}-A126 or Targ\textsuperscript{MB}-S126 as compared to those treated with Targ\textsuperscript{MB}-M126 (Figures 2F and 2G). At week 4, 3D ultrasound reconstruction of the abdominal aorta showed massive areas of plaque buildup and aneurysm in animals treated with Targ\textsuperscript{MB}-A126 or Targ\textsuperscript{MB}-S126, but not in those treated with Targ\textsuperscript{MB}-M126 (Figure 2H and 2I). These results were further confirmed macroscopically on vessel isolation (Figure 3B). Furthermore, immunohistochemistry using GAM-VCAM-1 antibody confirmed the decreased VCAM-1 expression for animals treated with Targ\textsuperscript{MB}-M126 as compared to those treated with Targ\textsuperscript{MB}-A126 or Targ\textsuperscript{MB}-S126 (Figure 3C). Martius Scarlet Blue (MSB) staining also showed plaque buildup and aneurysm in the abdominal aortas of animals treated with Targ\textsuperscript{MB}-A126 or Targ\textsuperscript{MB}-S126 whereas very little plaque buildup was observed with Targ\textsuperscript{MB}-M126 (Figure 3D).

Ultrasound imaging for the anatomical diagnosis and monitoring of AAA is well established in clinical practice. This study extends the use of ultrasound in AAA toward a unique theranostic application, which allows simultaneous diagnosis and therapy of AAA. We demonstrate that MBs can be used as functional ultrasonic contrast agents to assess the inflammatory state of the endothelium, as well as for targeted delivery of therapeutic anti-inflammatory drugs. The targeted delivery of miRs is achieved by conjugating a scFv, which specifically binds to mouse VCAM-1, onto MBs, thereby increasing the therapeutic payload and allowing targeted drug delivery specifically at the site of AAA. We cannot exclude the possibility that there might be free cholesterol-tagged miRs present in the blood stream, which could also be transferred into the cells within the area of disease, potentially facilitated by the ultrasound destruction of our targeted MBs. However, our study demonstrates that the anti-miR-126,
TargMB-A126 increases VCAM-1 expression and further aggravates AAA, whereas treatment with a mimic of miR-126, TargMB-M126, downregulates VCAM-1 expression on the intraluminal surface of the aneurysmal area and effectively prevents the development of AAA. To the best of our knowledge, this is the first study to present targeted theranostic MBs for the therapeutic delivery of miRs in an AAA model. Our targeting approach is specific for areas with VCAM-1 expression, thereby allowing the miR therapeutics to be delivered directly to the area of interest. The use of molecular ultrasound technology to destroy the MBs bound to the wall of the inflamed aorta and to release the miR therapeutics enhances the specificity of this novel therapeutic approach. These data provide proof of concept for a novel, non-invasive, and risk-free theranostic strategy (Figure S3) that allows monitoring and holds great potential to ultimately prevent aneurysmal growth of the abdominal aorta.

We used scFvs for targeting of our MBs because they have major advantages over traditional antibodies. ScFvs are small antibody fragments lacking the Fc region and therefore are minimally immunogenic. In addition, scFvs allow optimization via molecular biology methods, including fusion with drugs, as well as incorporation of specific tags and sequences for purification, biocjugation, and imaging. Our group has developed several highly specific scFvs for diagnostic imaging, targeted therapy, and theranostic approaches.

Pioneering studies have analyzed the circulating levels of vascular and inflammation-associated miRs in atherosclerosis, acute myocardial infarction, and AAA. In particular, one such miR that has been found to be upregulated in AAA is the endothelium-related...
miR-126. Therapeutic miR approaches have attracted major interest, with both pre-clinical and clinical reports describing their immense potential to regulate specific gene expression in a broad range of diseases. Several miR-based therapies have been assessed in AAA, such as targeting miR-21, miR-29, miR-181, miR-205, and miR-712. Within the setting of AAA, dysregulation of ECs...
augments localized inflammation and expression of integrins and adhesion molecules on the endothelial luminal surface, consequently increasing leukocyte attachment, rolling, and ultimately migration/infiltration into the intimal layer.\textsuperscript{5,23} miR-126 is understood to be exclusive to ECs and therefore has been identified as a crucial endogenous mediator of vascular integrity and angiogenesis and regulator of adhesion molecules.\textsuperscript{5–7} Our in vitro work also highlights the use of M\textsubscript{126} for the suppression of VCAM-1 expression, and the opposite has been observed using A\textsubscript{126}.

A recent study provided compelling evidence of miR-126’s anti-inflammatory effects via increased localized enrichment of CXCL12 following apoptotic microparticle blebbing from ECs, of which miR-126 is a major constituent.\textsuperscript{34} This study also showed that miR-126 therapy via delivery of endothelial apoptotic bodies decreased overall atherosclerotic lesion size in mouse models of atherosclerosis,\textsuperscript{34} further validating a role for miR-126 in disease models of endothelial dysfunction and inflammation. Our acute inflammatory model shows that targeted delivery of miR-126 can alter VCAM-1 expression on the thoracic and abdominal aorta of LPS-treated mice.

Based on the presented in vivo data, the clinical translation of our targeted theranostic approach appears highly promising for patients with AAA. The VCAM-1-targeting MBs enrich the miR therapeutics at the site of activated ECs, and together with the localized application of an ultrasound burst, non-specific uptake of miR therapeutics in other cell types can be minimized. Overall, the presented theranostic approach has the potential to provide a long-sought-after medical treatment option for patients with AAA.

MATERIALS AND METHODS

Generation of Anti-mVCAM-1 Single-Chain Antibody Containing an AviTag Motif at the C Terminus

The generation of the scF\textsubscript{v}m\textsubscript{VCAM-1} has been described previously and was a gift from Prof. Claudia Gottstein.\textsuperscript{13} The scF\textsubscript{v}m\textsubscript{VCAM-1} was subcloned into the pAC6 vector system (Avidity, Aurora, CO), which consists of the AviTag motif at the C-terminal. PCR was performed with a sense primer that anneals at the beginning of the scFv sequence and an antisense primer that anneals directly to the 6\textsuperscript{His} tag region at the end of the scFv. The following primers were used for PCR: sense strand: 5’–CAG CCG GCC ATG GCG CAG GTT CA–3’ and anti-sense strand: 5’–TCA GAT CCC GGG TAA TGG TGA TGG TGA TGA TG–3’. The antisense strand includes the Xmal restriction site. After amplification by PCR, the constructs were then digested with the restriction enzymes Notl and Xmal (both New England Biolabs, Ipswich, MA) and cloned into pAC6. Electrophoresis on a 0.8% agarose gel with SYBR Safe DNA gel stain (Invitrogen, Carlsbad, CA) was utilized to analyze amplifications of DNA by PCR and restriction digests. Ligation of the plasmids was performed with T4 ligase (NEB, Ipswich, MA) at 16°C overnight. The resulting plasmid constructs were then transformed into Turbo competent E. coli cells (NEB, Ipswich, MA). The transformed cells were grown in Luria-Bertani (LB) media containing 100 μg/mL ampicillin and 100 mM glucose at 37°C, and the plasmids were purified using a Plasmid Mini-Prep Kit (QIAGEN, Hilden, Germany). The respective plasmids were analyzed by electrophoresis on a 0.8% agarose gel with SYBR green, and constructs were confirmed by DNA sequencing. The DNA was then transformed into electrocompetent cell EVB101 (Avidity, Aurora, CO) by electroporation with Bio-Rad Gene Pulser. All agarose gels were visualized using a Bio-Rad Gel-Doc system with Quantity One software (Bio-Rad, Hercules, CA).

Expression, In Vivo Biotinylation, and Purification of scF\textsubscript{v}m\textsubscript{VCAM-1}-AviTag Construct

Bacterial strains of E. coli harboring the plasmid pAC6-scF\textsubscript{v}m\textsubscript{VCAM-1}-AviTag were used to inoculate 5 mL of tryptone-HEPES-yeast (THY) media containing 10 μg/mL chloramphenicol and 100 μg/mL ampicillin at 37°C.\textsuperscript{14} After culturing overnight, 5 mL of the overnight culture was transferred into 1 L of THY media containing 100 μg/mL ampicillin in a non-baffled Erlenmeyer flask. Glucose was added to the THY media to 0.5% final concentration. In vivo biotinylation was induced by 50 mM Biotin-D, 50 mM bicine, and 1.5 mM isopropyl β-D-thiogalactoside, when the optical density 600 (OD\textsubscript{600}) of the culture reached 0.7 units. After 3 hr induction, the cells were spun down and resuspended in BugBuster Master Mix (Novagen, Darmstadt, Germany). The cell debris was spun down, and the inclusion bodies were used for protein extraction via 8 M urea. All purified single-chain antibodies carry a 6\textsuperscript{His} Tag at the C-terminal end of their amino acid sequence for purification by immobilized metal affinity chromatography (IMAC) or for fluorescence-activated cell sorting (FACS) analysis. Proteins were purified with a nickel-based metal affinity chromatography column, Ni-NTA column (Invitrogen, Carlsbad, CA), according to the manufacturer’s instruction manual. Fractions of 1 mL were collected and dialyzed against 6 M urea, decreasing slowly in 2 M increments until PBS.

Evaluation of Purity and Efficiency of the In Vivo Biotinylation of the scF\textsubscript{v}m\textsubscript{VCAM-1}-AviTag Construct

The purity of the proteins and the efficiency of in vivo biotinylation were analyzed by western blotting. 30 μL of each purified protein and 6 μL of 5× reducing SDS loading buffer were added to 1.5-ML tubes and denatured at 96°C for 5 min. The samples were run on SDS-PAGE gel in SDS running buffer at 100 V for 2 hr. The gel was then used for western blotting at 100 V for 1 hr 30 min. The membrane was blocked with 1% BSA and hybridized with a specific horse-radish peroxidase (HRP). Anti-6\textsuperscript{His}-tag antibody HRP was used to detect the protein construct, and streptavidin-HRP (BD Biosciences, San Jose, CA) was used to determine the success of the in vivo biotinylation of the protein construct. Secondary hybridization was performed with SuperSignal West Pico chemiluminescent substrate (Thermo Scientific, Waltham, MA), an enhanced chemiluminescent (ECL) substrate for the HRP enzyme.

Cholesterol-Tagged miRNA

The sequences of the miR-126 are as follows: mimic-miR-126 (M\textsubscript{126}) 5’–UGG UAC CGU GAG UAA UAA UGC G–3’; anti-miR-126 (A\textsubscript{126}) 5’–CGT ATT ATT ACT CAC GGT ACG A–3’; and scramble-miR-126 (S\textsubscript{126}) 5’–CGC UCA UUC UGC CGG UUG
UUA UG-3', All miRs were modified with 2-O-methyl, 5'-Cy3, and 3'-cholesterol (VBC Biotech Services, Vienna, Austria).

**mVCAM-Expressing SVEC4-10 Cell Culture**

SVEC4-10 mouse endothelial cells (ATCC, Manassas, VA) were cultured in DMEM (Invitrogen, Carlsbad, CA), supplemented with 1% penicillin/streptomycin, 1% L-glutamine (200 mM), and 10% fetal calf serum to the media (Gibco, Waltham, MA). When the monolayer adherent SVEC4-10 cells reached confluency of 90%, a dissociation reagent containing 0.05% trypsin-EDTA (Gibco, Waltham, MA) was used to detach the cells into a suspension.

**Flow Cytometry**

SVEC4-10 cells were incubated with 10 µg/mL of purified scFvscCAM-1 for 10 min at 37°C followed by Penta-His AlexaFluor 488 conjugate (QIAGEN, Hilden, Germany) or R-phycocerythrin streptavidin (Jackson ImmunoResearch Laboratories, West Grove, PA) for 15 min. As a positive control and to determine the VCAM expression, the cells were incubated with CD106 (anti-VCAM) rat-anti-mouse antibody for 10 min at 37°C followed by a goat-anti-rat FITC (GAR-FITC) secondary antibody for 15 min. Samples were fixed with 1% Cellfix (BD Biosciences, San Jose, CA) and analyzed by FACS Calibur (BD Biosciences, San Jose, CA). The cell population was distinguished using the forward and sideward light scatter profile. A gate was set around the cell population, and 10,000 cells were analyzed. For determination of VCAM-1 expression, single staining using CD106 and GAR-FITC secondary antibody was used. For determination of miR-126 transfection, the Cy3 on the 5’ of the oligo was used.

**qPCR for Gene Expression**

For in vitro work, the total RNA was extracted using Trizol reagent (Invitrogen, Carlsbad, CA) under RNase-free conditions. A DNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA) was used to obtain cDNA from the total RNA samples, according to the manufacturer’s instructions. qRT-PCR of miR was carried out using a TaqMan Reverse Transcription Kit (Applied Biosystems, Foster City, CA), according to the manufacturer’s instructions. For in vivo work, total RNA was extracted using a fibrous tissue isolation kit (QIAGEN, Hilden, Germany) under RNase-free conditions as per the manufacturer’s instructions. Initially, two-step qRT-PCR was performed using identical reagents as mentioned previously; however, low yields of RNA on account of the small sample size of the thoracic aorta led to the use of a one-step QuantiFast SYBR green qRT-PCR kit (QIAGEN, Hilden, Germany) for all further assays. All samples were performed in triplicate. The expression levels of VCAM-1 were calculated using GAPDH as an internal control.

**Preparation of Microbubbles**

VisualSonics Target-Ready MBs (8.4 × 10⁸ MBs; VisualSonics, Toronto, Canada) were resuspended in 1 mL sterile saline (0.9% sodium chloride) according to the manufacturer’s instructions. 20 µg of scFvscCAM-1 or a non-binding scFvMut were added to each milliliter of MBs to generate TargMB or non-TargMB as control, respectively, and incubated at room temperature for 20 min. The average radius of a MB is around 1.5 µm, and MBs bear an average of 7,600 streptavidin molecules per µm² MB surface, resulting in approximately 6,000 biotinylated scFv molecules per µm² of the MB. TargMB or non-TargMB was incubated with 100 µg/mL of one of the three cholesterol-tagged miRs: M126, A126, or S126. This creates 1 molecule of cholesterol-miR to every 200 lipids on the MBs. The following terms are used for this manuscript: TargMB-M126 (VCAM-1-targeted MBs with mimic-miR-126); TargMB-A126 (VCAM-1-targeted MBs with anti-miR-126); and TargMB-S126 (VCAM-1-targeted MBs with scramble-miR-126). For in vitro experiments, 10 µL of MB, containing 0.2 µg of scFv and 100 ng of miR, were used. For in vivo experiments, 100 µL of MB, containing 2 µg of scFv and 10 µg of miR, were given to each animal per injection.

**Ultrasound Burst for Targeted Transfection of miR-126 Using VCAM-1-Targeted Microbubbles Loaded with miR-126**

SVEC4-10 cells were seeded into 35 × 10 mm Petri dishes (Sarstedt, Neumabrecht, Germany) and incubated to allow attachment overnight. Petri dishes were carefully filled with PBS with calcium and magnesium to the brim and taped shut using Parafilm M (Bemis, Neenah, WI). The TargMB-M126, TargMB-A126, or TargMB-S126 was injected using a 30 G needle into the Petri dishes through the Parafilm M. The Petri dishes were placed with the bottom facing up and were gently shaken for 5 min. A washing step was performed to remove unbound MBs before ultrasound bursting was employed to destroy the bound MBs and enable the miR to enter the cells. For washing, the Parafilm M was removed, the supernatant was discarded, and fresh media were added. Ultrasound was performed on the cells using color Doppler mode for 30 s per field of view (4 fields of view per Petri dish) to destroy the MBs. Cells were incubated for 48 hr before microscopy.

**In Vivo Mouse Experiments**

Male C57BL/6 mice or apolipoprotein E knockout (Apo E−/) mice were obtained from the Alfred Medical Research and Education Precinct Animal Services and assigned randomly to one of the three groups. Animals were injected with TargMB-M126, TargMB-A126, or TargMB-S126. All experiments involving animals were approved by the Alfred Medical Research and Education Precinct Animal Ethics Committee (E/1406/2013/B and E/1187/2012/B). The animals were anesthetized using 1.5%–2% isoflurane. Breathing of the animals was monitored to determine the amount of anesthetics used. For animals undergoing ultrasound, the fur was removed by shaving cream (Dove, Sydney, Australia). Mice were placed on a 37°C heated mat to prevent hypothermia.

**Acute Inflammatory Mouse Model**

Male C57BL/6 mice, 6 to 7 weeks of age (20–25 g), were used for this model. Animals were given an i.p. injection of 50 µg LPS diluted to 100 µL with PBS with calcium and magnesium. 18 hr post-injection, the animals were anesthetized using 1.5%–2% isoflurane and placed on the VisualSonics imaging station. The animals were assigned...
or TargMB-S126 was injected via the tail vein. After 5 min of circulation, we used the burst mode on the Vevo2100 high-frequency small-animal ultrasound scanner and burst the MBs that were attached on the thoracic and abdominal aorta. 48 hr post-ultrasound, the abdominal aorta and the thoracic aorta of the animals were removed for histology and qRT-PCR, respectively.

**AAA Mouse Model**

Male ApoE<sup>−/−</sup> mice, 24 weeks of age, were used for this model. The animals were anesthetized using 1.5%–2% isoflurane and placed on a heated mat. A small incision was made in the skin between the scapulae. The connective tissue was carefully spread to create a pocket for implantation, the animals were assigned randomly to one of three groups, and Targ<sup>MB-M126</sup>, Targ<sup>MB-A126</sup>, or Targ<sup>MB-S126</sup> was injected via the tail vein. 3 days post-LPS injection, embedded in optimal cutting temperature compound (OCT; Sakura Finetek, Torrance, CA) and snapped frozen, and finally, 6-µm sections were obtained using a cryostat (Microm HM525 NX, Thermo Fisher Scientific, Waltham, MA). In order to determine VCAM-1 expression, multiple sections per animal were selected at random and underwent immunostaining using goat-anti-mouse VCAM-1 polyclonal antibody (Santa Cruz Biotechnology, Dallas, TX) following acetone and formalin fixation steps. The sections were then incubated with the appropriate biotinylated secondary antibody (Vector Laboratories, Burlingame, CA), and expression was determined via DAB substrate (Vector Laboratories, Burlingame, CA). All relevant controls were performed. Sections of abdominal aorta were also stained for fibrin using MSB. Thawed sections were treated with Weigert’s iron hematoxylin (Sigma-Aldrich, MO), incubated with 96% ethanol, and stained with Picric Orange G solution (EMS, PA). Sections were then further stained with Brilliant Crystal Scarlet 6R (Sigma-Aldrich, St. Louis, MO), and fibrin was differentiated using 1% phosphotungstic acid (EMS, Hatfield, PA). Finally, sections were stained with Aniline Blue solution (EMS, Hatfield, PA), rinsed in 1% acetic acid, dehydrated in 100% ethanol, cleared in xylene, and mounted using Depex mounting medium (Labchem, Zelienople, PA). Sections were rinsed in dH<sub>2</sub>O between each step. For this MB trichrome stain, erythrocytes, fibrin, and collagen are represented by orange, bright red, and blue, respectively. Sections were imaged using microscopy.

**In Vivo Ultrasound Molecular Imaging**

Ultrasound imaging was performed using a Vevo2100 small-animal high-resolution imaging scanner (VisualSonics, Toronto, Canada) using a 22- to 55-MHz high-frequency transducer (lateral resolution, 100 µm; transverse resolution, 40 µm; focal length, 10 mm; low acoustic pressure or mechanical index, 0.14). The animals were anesthetized using 1.5%–2% isoflurane and placed on the imaging station. Fur was removed by shaving with cream (Dove, Sydney, Australia). For imaging of the abdominal aorta, the transducer was placed directly over the abdomen of the mouse, with the transducer marker facing 12 o’clock. All anatomical imaging was performed in fundamental brightness mode (B mode) using transmit power, 100%; dynamic range, 65 dB. We ensured that the region of interest (abdominal aorta and the branches of the renal arteries) was well visualized via imaging. The diameter of the abdominal aorta was measured both suprarenal (above the renal arteries) and infrarenal (below the renal arteries). The position of the renal arteries ensures that the measurements were repeatedly performed at the same area weekly and therefore ensures its repeatability. For contrast imaging and ultrasonic burst, we used a 13- to 24-MHz high-frequency non-linear contrast transducer. The animals were assigned randomly to one of the three groups, and Targ<sup>MB-M126</sup>, Targ<sup>MB-A126</sup>, or Targ<sup>MB-S126</sup> was injected via the tail vein. Upon the injection of MBs, we imaged using the nonlinear contrast mode (using transmit power, 10% and dynamic range, 40 dB). After 5 min of circulation, we used the burst mode on the Vevo2100 ultrasound scanner to burst the MBs that were attached on the thoracic and abdominal aorta. When we employed the burst mode to destroy the MBs, we used a continuous 10-MHz high-power destructive pulse (using transmit power, 100%; high mechanical index, ~0.63; average power, ~0.0676 W/cm<sup>2</sup>) applied for 2 s every 10 s, which destroys the MBs within the beam elevation. During the burst mode, the transducer is mechanically moved to cover the desired target area. 15 cycles of burst were performed. Videos and images were acquired throughout the imaging procedure. Analysis was performed using VisualSonics imaging software (VisualSonics, Toronto, Canada).

**Immunohistochemistry**

The abdominal aorta was harvested following 4 weeks of Ang-II treatment or 3 days post-LPS injection, embedded in optimal cutting temperature compound (OCT; Sakura Finetek, Torrance, CA) and stained with Brilliant Crystal Scarlet 6R (Sigma-Aldrich, MO), incubated with 96% ethanol, and stained with Picric Orange G solution (EMS, PA). Sections were then further stained using goat-anti-mouse VCAM-1 polyclonal antibody (Santa Cruz Biotechnology, Dallas, TX) following acetone and formalin fixation steps. The sections were then incubated with the appropriate biotinylated secondary antibody (Vector Laboratories, Burlingame, CA), and expression was determined via DAB substrate (Vector Laboratories, Burlingame, CA). All relevant controls were performed. Sections of abdominal aorta were also stained for fibrin using MSB. Thawed sections were treated with Weigert’s iron hematoxylin (Sigma-Aldrich, MO), incubated with 96% ethanol, and stained with Picric Orange G solution (EMS, PA). Sections were then further stained with Brilliant Crystal Scarlet 6R (Sigma-Aldrich, St. Louis, MO), and fibrin was differentiated using 1% phosphotungstic acid (EMS, Hatfield, PA). Finally, sections were stained with Aniline Blue solution (EMS, Hatfield, PA), rinsed in 1% acetic acid, dehydrated in 100% ethanol, cleared in xylene, and mounted using Depex mounting medium (Labchem, Zelienople, PA). Sections were rinsed in dH<sub>2</sub>O between each step. For this MB trichrome stain, erythrocytes, fibrin, and collagen are represented by orange, bright red, and blue, respectively. Sections were imaged using microscopy.

**Microscopy**

In vivo experiments were visualized with an IX81 Olympus microscope (Olympus, Tokyo, Japan) and Cell^P 1692 (ANALYsis Image Processing) software, using bright field with a 20× objective and the tetramethylrhodamine (TRITC) fluorescence channel. Fluorescence was quantified using ImageJ software. Histological sections were visualized with an Olympus BX5050F-3 microscope (Olympus, Tokyo, Japan) and CellSens software. All microscopy experiments were performed at ambient temperature.

**Statistical Analysis**

Unless otherwise specified, data are expressed as mean ± SD. Analyses containing 2 groups were analyzed with the Student’s t test. All analyses containing more than two groups were analyzed with one-way ANOVA with repeated measures (if equal numbers)
or ordinary (if unequal numbers), comparing all groups with one another, corrected by post hoc Bonferroni analysis, and the corrected p values are given. Statistical analyses were performed using Graphpad Prism 6.0.

SUPPLEMENTAL INFORMATION
Supplemental Information includes three figures and three movies and can be found with this article online at https://doi.org/10.1016/j.ymthe.2018.02.010.

AUTHOR CONTRIBUTIONS
X.W. and K.P. conceived and designed the research. X.W., A.K.S., J.D.H., A.L.L., M.-K.A., J.P., B.L., Y.Y., M.W., E.Y., and Y.-C.C. performed the experiments. X.W., A.K.S., J.D.H., J.P., and K.P. analyzed the data. X.W., A.K.S., and K.P. wrote the manuscript.

CONFLICTS OF INTEREST
The authors have declared that no competing interests exist.

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