Interactions of WASp, myosin-I, and verprolin with Arp2/3 complex during actin patch assembly in fission yeast

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East actin patches are dynamic structures that form at the sites of cell growth and are thought to play a role in endocytosis. We used biochemical analysis and live cell imaging to investigate actin patch assembly in fission yeast Schizosaccharomyces pombe. Patch assembly proceeds via two parallel pathways: one dependent on WASp Wsp1p and verprolin Vrp1p converges with another dependent on class 1 myosin Myo1p to activate the actin-related protein 2/3 (Arp2/3) complex. Wsp1p activates Arp2/3 complex via a conventional mechanism, resulting in branched filaments. Myo1p is a weaker Arp2/3 complex activator that makes unstable branches and is enhanced by verprolin. During patch assembly in vivo, Wsp1p and Vrp1p arrive first independent of Myo1p. Arp2/3 complex associates with nascent activator patches over 6–9 s while remaining stationary. After reaching a maximum concentration, Arp2/3 complex patches move centripetally as activator proteins dissociate. Genetic dependencies of patch formation suggest that patch formation involves cross talk between Myo1p and Wsp1p/Vrp1p pathways.

Introduction

During interphase, yeast cells contain two cytoskeleton structures composed of actin filaments, cables that depend on formins and actin patches that depend on the actin-related protein 2/3 (Arp2/3) complex for their formation (Chang and Peter, 2002; Weaver et al., 2003). Actin patches are highly dynamic, mobile structures (Doyle and Botstein, 1996; Waddle et al., 1996; Pelham and Chang, 2001; Smith et al., 2001) that form at the sites of active cell growth and may play a role in endocytosis (Mulholland et al., 1994). Budding yeast form patches at sites of endocytosis where actin assembly may either assist in severing the endocytic membrane or propel the vesicle toward cell interior (Munn, 2001; Engqvist-Goldstein and Drubin, 2003; Kaksonen et al., 2003; Huckaba et al., 2004; Jonsdottir and Li, 2004; Merrifield, 2004). Actin assembly is proposed to play similar roles in endocytosis in cultured vertebrate cells and Xenopus eggs (Merrifield et al., 2002; Sokac et al., 2003).

Yeast actin patches share many components involved in extension of the leading edge in motile cells (Pollard et al., 2000; Higgs and Pollard, 2001). Initiation of patch assembly in budding yeast involves coordinated activity of class 1 myosins Myo3p and Myo5p (Goodson et al., 1996; Anderson et al., 1998) and Wiskott-Aldrich Syndrome protein (WASP) family Las17p/Bee1p (Li, 1997). Both Las17p/Bee1p and myosins Myo3p/Myo5p have COOH-terminal domains with a tryptophan and a cluster of acidic residues that bind Arp2/3 complex (Higgs and Pollard, 2001; Weaver et al., 2003). Las17p/Bee1p and myosins Myo3p/Myo5p are genetically and functionally redundant because a single acidic domain either on a myosin-I or Las17p/Bee1p suffices to induce actin patch assembly (Evangelista et al., 2000; Lechner et al., 2000). Actin patch assembly in budding yeast also involves a proline-rich protein, verprolin Vrp1p (Donnelly et al., 1993; Munn et al., 1995; Vaduva et al., 1997), which interacts with both Las17p/Bee1p and Myo3p/Myo5p (Anderson et al., 1998; Naqvi et al., 1998; Evangelista et al., 2000). Vrp1p binding to type I myosins is needed to induce actin assembly in cell extracts (Geli et al., 2000). Class I myosins can also bind Las17p/Bee1p directly (Evangelista et al., 2000; Lechner et al., 2000). Interactions among activators may potentially lead to the formation of a large protein complex (Evangelista et al., 2000; Lechner et al., 2000, 2001) and coordinate the recruitment and activation of the Arp2/3 complex.
Fission yeast actin patches also depend on myosin-I and WASp (Lee et al., 2000). Like other WASp family members, Schizosaccharomyces pombe Wsp1p has a COOH-terminal VCA region with a central-acidic (CA) domain expected to bind Arp2/3 complex and a WASp homology 2 (WH2) domain (V) expected to bind actin monomer (Fig. 1 A), both required to activate Arp2/3 complex in other cells (Anderson et al., 2001; Hertzog et al., 2002). Myo1p is a long-tailed class 1 myosin with a motor domain, 2 IQ motifs, and a tail that includes a tail homology 1 (TH1) domain expected to bind lipids, a TH2 domain of unknown function, an Src homology 3 (SH3) domain expected to bind proline-rich sequences, and a CA domain (Fig. 1 A). S. pombe Vrp1p is a proline-rich protein with an NH2-terminal WH2 domain. A GST fusion to Myo1p TH2-SH3-CA binds and activates bovin-I gene ARPC3 as the faster migrating subunit in the ~20-kD ARPC3/ARPC4 doublet.

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Fission yeast survive deletion of either the single myosin-I gene myo1+ or the single WASp family gene wsp1+ with severe cytoskeletal defects, but deletion of both genes is lethal (Lee et al., 2000). Therefore, Myo1p and Wsp1p may have independent but overlapping functions and likely operate in two parallel pathways of actin assembly. Normal patch assembly may rely on coordinated activity of pathways involving Myo1p and Wsp1p, perhaps by direct physical interactions among activators.

Here, we use live cell imaging and genetic analysis in combination with biochemical analysis of purified S. pombe Arp2/3 complex, Myo1p tail, Wsp1p VCA, and full-length Vrp1p to study the mechanism of actin patch assembly.

### Results

#### Purification and interactions of Arp2/3 complex, Myo1p, Wsp1p, and Vrp1p

We used affinity and ion exchange chromatography to purify native S. pombe Arp2/3 complex and the following recombinant fusion proteins: GST fused to the TH2-SH3-CA, TH2-SH3, and SH3-CA domains of the myosin-I (Myo1p) tail (GST-Myo1p23A, -23, -3A); GST fused to the VCA region of Wsp1p (GST-Wsp1p-VCA); and full-length Vrp1p with a COOH-terminal His-tag (Fig. 1). S. pombe Arp2/3 complex consists of seven subunits (Fig. 1 C) with slightly different electrophoretic mobilities than the subunits in other species. S. pombe Arp2p and ARPC1 have similar electrophoretic mobilities. Mass spectrometry identified ARPC3 as the faster migrating subunit in the ~20-kD ARPC3/ARPC4 doublet.

We measured the affinities of these proteins for each other and for actin monomers with quantitative pull-down assays (Fig. 2 and Table I). Interaction of the Myo1p tail with Arp2/3 complex depended on the A domain, but not the TH2 domain. Like human GST-WASP-VCA (Hufner et al., 2001), GST-Wsp1p-VCA bound Arp2/3 complex much stronger (Kd ~0.05 μM) than GST-Myo1p23A (Kd = 1–2 μM). GST-Myo1p23A bound Vrp1p with a higher affinity (Kd = 3–6 μM) than muscle actin monomers or filaments (Kd > 20 μM; Lee et al., 2000). Because both GST-Myo1p23 and GST-Myo1p3A bound verprolin, the SH3 domain mediates this interaction as in budding yeast (Anderson et al., 1998). The affinity of verprolin for actin monomers (Kd ~1 μM) was similar to other proteins with WH2 domains (Hufner et al., 2001; Marchand et al., 2001; Martinez-Quiles et al., 2001; Hertzog et al., 2002).

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### Table I. Equilibrium dissociation constants (Kd) for interactions among Myo1p tail, Wsp1p VCA, verprolin, Arp2/3 complex, and actin

| Receptor          | Ligand            | Binding | Kd (μM) |
|-------------------|-------------------|---------|---------|
| GST-Myo1p23A      | Arp2/3 complex    | +       | 1.2/1.8 |
| GST-Myo1p3A       | Arp2/3 complex    | +       | 2.0/2.1 |
| GST-Myo1p23       | Arp2/3 complex    | −       | —       |
| GST-Wsp1p-VCA     | Arp2/3 complex    | +       | 0.05    |
| GST-Myo1p23A      | Verprolin         | +       | 4.6/5.6* (3–6 μM) |
| GST-Myo1p3A       | Verprolin         | +       | N.D.    |
| GST-Myo1p23       | Verprolin         | +       | N.D.    |
| Vrp1p-His         | G-actin           | +       | 1.0/0.9/0.8 |

Equilibrium dissociation constants were measured using quantitative pull-down assays. slashes separate values from independent experiments. N.D., not determined. *Values represent the best fits, the range of values from all fit attempts is in parentheses.
Activation of Arp2/3 complex by Myo1p, Wsp1p, and Vrp1p

We compared the ability of COOH-terminal fragments of Myo1p and Wsp1p each fused to GST to activate Arp2/3 complex. Purified *S. pombe* Arp2/3 complex had little effect on the spontaneous assembly of purified actin monomers (Fig. 3 A). GST-Wsp1p-VCA and GST-Myo1p-23A each activated Arp2/3 complex, resulting in nucleation of new filaments and acceleration of polymerization. Neither GST-Myo1p-23A nor GST-Wsp1p-VCA stimulated actin polymerization in the absence of Arp2/3 complex. GST-Wsp1p-VCA produced three times more actin filament barbed ends than GST-Myo1p-23A (Fig. 3 B). GST-Myo1p-23A did not increase Arp2/3 complex activation by GST-Wsp1p-VCA, so the two activators do not act synergistically (Fig. S1; available at http://www.jcb.org/cgi/content/full/jcb.200502053/DC1). Verprolin did not enhance the ability of GST-Wsp1p-VCA to stimulate actin polymerization by Arp2/3 complex (Fig. 3 A).

Full-length verprolin enhanced the ability of GST-Myo1p-23A to stimulate actin polymerization by Arp2/3 complex (Fig. 3 A). At a concentration (50 nM) of GST-Myo1p-23A that barely activated Arp2/3 complex, Vrp1p produced barbed ends in proportion to its concentration up to 1 μM. Verprolin alone did not activate Arp2/3 complex or stimulate actin polymerization, except for slight increase of rate late in reaction. Verprolin and GST-Myo1p-23A did not stimulate actin assembly without Arp2/3 complex. With a high concentration (500 nM) of verprolin and nanomolar Arp2/3 complex, GST-Myo1p-23A induced formation of five times more barbed ends than in the absence of verprolin (Fig. 3 B), similar to GST-N-WASP-VVCA, the strongest activator described to date (Zalevsky et al., 2001). This corresponds to about one barbed end for every three Arp2/3 complexes. In both the presence and absence of verprolin, the concentration of Myo1p tail required to achieve half-maximal yield of barbed ends was 150 nM, 10-fold higher than GST-Wsp1p-VCA or GST-N-WASP-VVCA (Fig. 3 B). Pre-existing filaments reduced the lag in the actin nucleation by Arp2/3 complex activated by Myo1p tail both in the presence and the absence of verprolin (Fig. 3 C).

The ability of Myo1p tail to activate Arp2/3 complex depended upon both the TH2 and A domains, both in the presence and the absence of verprolin (Fig. S1). In fact, GST-Myo1p-23 lacking the A domain inhibited Arp2/3 complex activation by GST-Myo1p-23A, reducing the number of actin filaments nucleated by Arp2/3 complex by 97% (Fig. 3 D). Verprolin partially overcame this inhibition, resulting in only a 63% decrease in the number of filaments nucleated by Arp2/3 complex. GST-Myo1p-3A tail construct lacking the TH2 domain reduced activation of Arp2/3 complex by GST-Myo1p-23A by 50% without verprolin and 18% with verprolin (Fig. 3 D). Neither GST-Myo1p-23 nor GST-Myo1p-3A inhibited Arp2/3 complex activation by GST-Wsp1p-VCA or assembly of actin alone (Fig. S1).

Arp2/3 complex produced many more branches when activated by GST-Wsp1p-VCA or GST-N-WASP-VVCA than GST-Myo1p-23A with or without verprolin (Fig. 4). Activation of Arp2/3 complex by Myo1p-tail ± verprolin produced only 3–7% filaments with branches, and these rare filaments had but single branches (Fig. 4, B and C). In contrast, 25% of the filaments produced by GST-Wsp1p-VCA and 34% of the filaments produced by GST-N-WASP-VVCA were branched, often with two or more branches (Fig. 4, E and F). The mean lengths of actin filaments in these micrographs agreed with those expected from the concentrations of actin polymer and filament ends calculated from the time course of polymerization (Fig. 4 A).
Deletions and functional fluorescent protein tags of Arp2/3 complex activators

None of the three S. pombe genes for factors involved in Arp2/3 complex activation (wsp1<sup>+</sup>, myo1<sup>+</sup>, vrp1<sup>+</sup>) is essential for viability. New Δmyo1 and Δwsp1 strains with complete ORF deletions showed defects (salt and temperature sensitivity, slow growth, reduced mating, misshapen cells) similar to the original strains (Lee et al., 2000), except that new Δwsp1 strain, unlike the original strain, was temperature sensitive. A Δvrp1 strain grew normally at 25°C without morphological defects, but had growth defects at 18 and 36°C as reported by Carnahan and Gould (2003). We constructed a few Δwsp1 Δvrp1 strains but failed to generate Δmyo1 Δvrp1 strains, indicating that Δmyo1 is synthetically lethal with both Δvrp1 and Δwsp1 (Lee et al., 2000; Carnahan and Gould, 2003).

We tagged the three nucleation-promoting factors and Arp2/3 complex with monomeric (m) and nonmonomeric fluorescent proteins (GFP, YFP, or CFP) directly in the genome so that native promoters controlled expression and cells depended entirely on these fusion proteins. We checked the tagged strains for defects like those observed when these genes are deleted. Cells depending on ARPC5-monomeric GFP (ARPC5-mGFP) is synthetically lethal with both Δvrp1 and Δwsp1 (Lee et al., 2000; Carnahan and Gould, 2003).

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cally lethal with Δmyo1. In contrast, we were unable to generate a mGFP-wsp1 Δmyo1 strain by genetic crosses. Because the time course of patch assembly appears entirely normal in strains depending on mGFP-Wsp1p (see further), inability to generate a mGFP-wsp1 Δmyo1 strain by genetic crosses may be due to mGFP-Wsp1p failing to function during mating, sporulation, or spore germination.

Localization of Arp2/3 complex and activators in dynamic actin patches

We used spinning disk confocal microscopy to localize Arp2/3 complex and mGFP-Myo1p, mGFP-Wsp1p, and Vrp1p-GFP in cortical patches of live yeast cells (Fig. S2, available at http://www.jcb.org/cgi/content/full/jcb.200502053/DC1). The signal from ARPC5-mGFP was the strongest, followed by mGFP-Myo1p, mGFP-Wsp1p, and the Vrp1p-GFP. All three activators had similar lifetimes (Table S1, available at http://www.jcb.org/cgi/content/full/jcb.200502053/DC1) of 10 (±3) seconds (range 6 to 18 s). In 3D reconstructions, mGFP-Myo1p fluorescence also faintly outlined the entire cell cortex, and mGFP-Wsp1p also had a high level of cytoplasmic fluorescence. The mGFP-Myo1p patches were stationary in time-lapse movies of Z-series of optical sections through entire cells, whereas some mGFP-Wsp1p and Vrp1p-GFP patches moved centripetally during the last 3 s of their lifetimes. ARPC5-mGFP localized to motile patches with average lifetimes of 22 s. ARPC5-mGFP patches were stationary during the first half of their lifetimes, as they formed in the cell cortex. Then they moved centripetally 0.4–2 μm over an average of 10 s before fading (Fig. S2). Disruption of actin filaments with 100 μM latrunculin A eliminated dynamic activator patches and resulted in clumping of mGFP-Wsp1p, Vrp1p-GFP, and occasionally mGFP-Myo1p at the ends of the cells (Fig. S2).

Z-series through entire cells expressing all combinations of pairs of the three activators tagged with YFP/CFP showed all three proteins localized in the same patches (Fig. 5; Table S2, available at http://www.jcb.org/cgi/content/full/jcb.200502053/DC1). mCFP-Wsp1p and Vrp1p-YFP fluorescence closely overlapped in 87% of patches. 80% of patches contained both mCFP-Myo1p and mYFP-Wsp1p and 78% contained mCFP-Myo1p and Vrp1p-YFP. The remaining ~20% of patches contained only one of the two fluorescently labeled activators, indicating differences in timing of patch assembly among activators. Patches of Myo1p appeared to be more spread out along the membrane than Wsp1p and Vrp1p by 1–2 pixels (100–200 nm).

Z-series through entire cells expressing pairs of patch proteins revealed that 65–75% of patches of Arp2/3 complex labeled with mCFP included activator proteins labeled with mYFP or YFP (Fig. 5, Table S2). Remaining patches either had the activator alone or Arp2/3 complex alone. A higher fraction of patches appeared to have both an activator and Arp2/3 complex when activators were tagged with CFP and Arp2/3 complex with YFP, owing to the weaker signal from CFP than...
YFP, especially at the initial stages of patch assembly. In all cases, the Arp2/3 complex fluorescence overlapped only partially with the activator fluorescence, with Arp2/3 complex deeper inside the cell than the activators.

**Time course of patch assembly**

We determined the time course of patch formation, maturation, and dissolution by time-lapse microscopy of cells expressing an activator tagged with YFP and Arp2/3 complex tagged with mCFP (Fig. 6, Videos 1 and 2). A time series in a single confocal plane (Fig. 7, A–D) showed that mYFP-Myo1p concentrated in a stationary patch 3 s before a spatially coincident ARPC5-mCFP signal appeared and then increased in intensity over the next 6–9 s. At this point the mYFP-Myo1p fluorescence began to decline and the ARPC5-mCFP signal moved away from the membrane for another 9–12 s. Tracking patches in 3D movies (Fig. S2, Table S1) showed that ARPC5-mGFP behaved similarly to ARPC5-mCFP in 2D, indicating that patches remained visible in single confocal mid-sections throughout the span of their movements. Arp2/3 complex patches moved for 400–1,800 nm (average, 800 nm) with velocities ranging from 30 to 230 nm/s (average, 90 nm/s). The behavior of patches marked with mYFP-Wsp1p and ARPC5-mCFP was similar (Fig. 6, E–G), although ARPC5 arrived 6 s after Wsp1p. The same was true for patches marked with Vrp1p-YFP and ARPC5-mCFP (Fig. 6, H–J). Thus, Wsp1p/Vrp1p arrive in patches ~3 s before Myo1p.

The onset of the activator patch disassembly always coincided with initiation of Arp2/3 complex movement. During rapid disassembly of the activator patches, mYFP-Myo1p remained stationary in the cell cortex for ~6 s, whereas mYFP-Wsp1p and Vrp1p-YFP moved along with Arp2/3 complex for ~3 s and 100–400 nm and then disappeared (Fig. 6, G and J). Thus, Wsp1p/Vrp1p disappeared ~3 s before Myo1p. The difference in timing of Myo1p and Wsp1p/Vrp1p arrival in patches explains the differences in the number of activator-only patches observed by colocalization at a single time point (Fig. 5). When activators were labeled with YFP and Arp2/3 complex with CFP, the fraction of patches with Wsp1p alone (23%) or Vrp1p alone (15%) was higher than Myo1p alone (8%). Wsp1p and Vrp1p arrive earlier than Myo1p, and therefore spend a smaller fraction of their lifetime associated with Arp2/3 complex patches. The lifetimes of YFP-labeled activator patches observed in single confocal sections were 6–9 s longer than lifetimes of GFP-labeled patches in 3D movies (Table S1). This is likely due to the faint signals when short exposure times were used to minimize photobleaching of GFP during 4D data collection.

**Time course and genetic dependence of activator patch assembly**

To observe directly the timing of activator assembly/disassembly in patches, we made movies of cells expressing pairs of activators tagged with YFP and CFP. Wsp1p and Vrp1p arrived at the site of patch assembly slightly before Myo1p, and Myo1p persisted longer than Wsp1p. In time series taken at 3-s intervals, mYFP-Wsp1p appeared 3–6 s and disappeared 3 s before mCFP-Myo1p (Fig. 7, A and B). Activators remained together in patches for 9–12 s. After that, Wsp1p and Vrp1p moved together a short distance into the cytoplasm for ~3 s while Myo1p remained in the cortical patch, persisting on average for ~3 s after Wsp1p disappeared. The time courses of concentration of Myo1p, Wsp1p, and Vrp1p at patches explain the existence of patches with Myo1p or Wsp1p alone. The timings observed for activator pairs were consistent with movies of each activator paired with Arp2/3 complex (Fig. 6).

Strains lacking one of the activators showed that Myo1p and Wsp1p arrive at patches independently, and that Wsp1p, but not Myo1p, is required to bring Vrp1p to patches. We made five double strains by crossing mGFP-Myo1p, mGFP-Wsp1p, and Vrp1p-YFP strains with Δmyo1Δ, Δwsp1Δ, and Δvrp1Δ strains (Fig. 7, C–H; Table S1). Immunoblots showed that none of the gene deletions altered the abundance of other GFP-tagged activators more than 30%. The localization and dynamics of mGFP-Myo1p and mGFP-Wsp1p in cortical patches was indistinguishable in vrp1Δ+ and Δvrp1Δ cells, but no Vrp1p-GFP patches formed in Δwsp1Δ cells. mGFP-Myo1p concentrated in dynamic cortical patches in Δwsp1Δ cells, but the lifetimes of these patches were almost twice that in Wsp1pΔ+ cells. Vrp1p-GFP formed dim dynamic cortical patches (presumably containing Wsp1p) in Δmyo1Δ cells; however, these patches were abnormal: up to half formed and faded repeat-
Figure 6. **Dynamics of activator patches and Arp2/3 complex patches in live cells expressing pairs of fluorescent fusion proteins.** Activators (Myo1p, Wsp1p, Vrp1p) are red and Arp2/3 complex (ARPC5) is green in color images. (A) Time series of mYFP-Myo1p (red) and ARPC5-mCFP (green) in a single confocal plane at 3-s intervals. The “max” frame is a maximum projection image of the entire time series. Bar, 2 μm. (B–J) Quantitative analysis of patch dynamics. (B–D) mYFP-Myo1p and ARPC5-mCFP. (E–G) mYFP-Wsp1p and ARPC5-mCFP. (H–J) Vrp1p-YFP and ARPC5-mCFP. (B, E, and H) Time courses of individual patches at 3-s intervals in a single optical plane. Bar, 0.5 μm. (C, F, and I) Kymographs of individual patches. At 3-s intervals a 20 × 10-pixel box was maximally projected onto a 20 × 1-pixel vertical lane. Eleven such lanes were combined horizontally to generate kymographs. (D, G, and J) Time courses of average intensity of activators and Arp2/3 complex and total distance moved by activators and Arp2/3 complex in 20–60 individual patches tracked in a single optical plane at intervals of (D) 2 s or (G and J) 3 s. Fluorescence intensity values were normalized, data from individual patches were aligned to the peak of activator patch YFP intensity for calculation of average normalized intensity and position. In all panels arrows and arrowheads mark initiation of activator or Arp2/3 complex patch assembly.
edly at the same site; some persisted over 20 s; and others were short-lived, with lifetimes of only 6 s.

**Discussion**

**Two parallel pathways of yeast actin patch assembly**

Biochemical analysis, time-lapse imaging of fluorescent fusion proteins, and genetic dependencies gave a consistent picture of actin patch formation, consisting of three phases (Fig. 8). Phase 1 begins with Wsp1p recruiting Vrp1p to a small patch near the inner surface of the plasma membrane over 3–6 s. Then, Myo1p joins this patch over the next 3 s. Myo1p localization requires actin filaments, but not Wsp1p or Vrp1p. Phase 2 starts with the arrival of Arp2/3 complex 3 s after Myo1p. Over the next 6–9 s, the concentrations of three activators and Arp2/3 complex steadily increase in the stationary patch. At phase 3, as the concentrations of all four proteins peak, the Arp2/3 complex patch begins to move toward cell interior at 30–230 nm/s. Myo1p stays behind and disappears in 6 s. Wsp1p and Vrp1p move a short distance with Arp2/3 complex away from the membrane while Myo1p remains behind. Activator patch disassembles in 3–6 s and Arp2/3 complex patch moves for 9–12 s.
The key feature of the actin patch assembly is that it proceeds through parallel Wsp1p and Myo1p pathways that both converge on Arp2/3 complex (Fig. 8). In vitro Wsp1p and Myo1p activate Arp2/3 complex independently, and in vivo either pathway is sufficient for patch assembly and cell viability (Lee et al., 2000). Localization and genetic analysis place verprolin in the Wsp1p pathway. However, physical interactions among activators, including Vrp1 binding to Myo1p, may bridge the two pathways.

Budding yeast also has parallel pathways to Arp2/3 complex dependent on myosin-I and WASp (Evangelista et al., 2000; Lechler et al., 2000, 2001; Kaksonen et al., 2003; Jonsdottir and Li, 2004). Budding yeast WASp Las17p/Bee1p localizes to patches before myosin-I Myo5p and actin polymerization begins after Myo5p arrival (Kaksonen et al., 2003; Jonsdottir and Li, 2004). Upon full assembly of actin and dissociation of activators, actin patches move over 500 nm with velocities of 200–400 nm/s (Waddle et al., 1996; Smith et al., 2000; Lechler et al., 2000, 2001; Kaksonen et al., 2003), similar to Arp2/3 complex patches in fission yeast.

On the other hand, patch assembly differs both qualitatively and quantitatively in the two yeast. First, the lifetime of Las17p/Bee1p patches (40 s) (Kaksonen et al., 2003) is significantly longer than fission yeast Wsp1p patches (10–17 s). Class 1 myosin patches (Jonsdottir and Li, 2004) have similar lifetimes in both yeast (10–20 s). Second, actin patches move slowly (25 nm/s) over a short distance (200 nm) before initiating fast long-range motility in budding yeast (Kaksonen et al., 2003) but not fission yeast. Third, deletion of budding yeast VRP1 depolarizes actin patches, whereas patches remain polarized in fission yeast Δvrp1. As a result, budding yeast Δvrp1 cells have severe endocytic and actin cytoskeletal defects (Donnelly et al., 1993; Munn et al., 1995; Vaduva et al., 1997), like Δlas17 cells (Li, 1997), whereas fission yeast without Vrp1p have no apparent defects at 25°C (this paper).

**Wsp1p-dependent pathway of actin assembly**

Fission yeast WASp homologue Wsp1p is the first nucleation-promoting factor to assemble into a patch. Like other WASp/WAVE family members (Higgs and Pollard, 2001), S. pombe GST-Wsp1p-VCA activates S. pombe Arp2/3 complex to nucleate branched actin filaments. GST-Wsp1p-VCA is less active than GST-N-WASP-VVCA, keeping with the trend that strength of nucleation-promoting factors is proportional to the number of acidic residues in the A-domain (Zalevsky et al., 2001). Wsp1p has 10 acidic residues compared with 18 for neuronal WASp (N-WASP).

Recruitment of Vrp1p to patches depends on the presence of Wsp1p, and the two proteins remain together throughout the life of a patch. The close association of WASp and verprolin in budding yeast and vertebrates (Lechler et al., 2001; Ho et al., 2004) is mediated through interaction of a conserved Ena/VASP homology 1 domain of WASp with a proline-rich sequence in the COOH-terminal half of verprolin (Ramesh et al., 1997; Naqvi et al., 1998; Volkman et al., 2002). WASp-interacting protein has been suggested to inhibit N-WASP (Martinez-Quiles et al., 2001; Ho et al., 2004), but this seems unlikely in S. pombe because activator and actin patches are normal in Δvrp1 cells.

**Myo1p-dependent pathway of actin assembly**

Myo1p localizes in patches independently of Wsp1p or Vrp1p. The tail of Myo1p weakly activates actin nucleation of unstable branches by Arp2/3 complex. The low activity may be related to the 20-fold lower affinity of Myo1p tail for Arp2/3 complex than Wsp1p or VCA. Accordingly, 10-fold more Myo1p tail is required to make half the number of filaments as Wsp1p-VCA. Verprolin strongly stimulates the activity of Myo1p tail, but does not reduce the concentration of Myo1p tail required for half-maximal activation. The ability of preformed filaments to reduce the lag phase of actin nucleation by Arp2/3 complex stimulated by Myo1p tail is evidence that nucleation involves binding of Arp2/3 complex to preexisting filaments, even though most of these branches dissociate before growing long enough to be detected by microscopy.

**Myo1p and Wsp1p/Vrp1p cooperate in patch assembly**

Cellular experiments suggest that patch assembly depends on coordination of the Wsp1p/Vrp1p and Myo1p pathways to Arp2/3 complex. Cells lacking either Myo1p or Wsp1p have depolarized actin patches and severe defects in growth, morphology, septation, and mating (Lee et al., 2000). Myo1p patches in Δwsp1 and Vrp1p patches in Δmyo1 are abnormal. Thus, Myo1p and Wsp1p/Vrp1p are recruited separately but cooperate in patch assembly by direct physical interaction or providing complementary biochemical activities.

Vrp1p strongly enhances the ability of Myo1p tail to activate Arp2/3 complex. High concentrations are required, owing to the low affinity of Vrp1p for Myo1p tail (Kd 3–6 μM). Vrp1p may provide a WH2 domain to complement the A domain provided by Myo1p, forming a complete VCA nucleation-promoting factor (Evangelista et al., 2000; Lechler et al., 2000; Machesky, 2000). Budding yeast Vrp1p WH2 domain can substitute for native WH2 in GST-Las17p/VCA (Lechler et al., 2001).

On the other hand, the available assays have detected no impact of the absence of Vrp1p on fission yeast actin patches. Although this observation does not rule out the possibility that Vrp1p stimulates Myo1p when concentrated in the patch, it suggests that other factors may bridge the two pathways. Conversely, each pathway may contribute nonredundant biochemical activities. Activation of Arp2/3 complex by Myo1p and Wsp1p/Vrp1p in vitro is not cooperative. As a stronger activator, Wsp1p may be the primary activator, whereas affinity of the motor domain of full-length Myo1p for actin filaments may also contribute to assembling activated Arp2/3 complex (Lechler et al., 2000, 2001), possibly by capturing and tethering actin filaments and/or cables. Myo1p may also help to retain Vrp1p and Wsp1p in the patch, consistent with the observation that Vrp1p patches in Δmyo1 are dim and show abnormal dynamics, similar to actin patches in budding yeast lacking type 1 myosins (Smith et al.,
Mounting evidence in budding yeast indicates that actin patches form at the sites of endocytosis (Kaksonen et al., 2003; Huckaba et al., 2004; Jonsdottir and Li, 2004), so we presume that fission yeast actin patches do as well. The fission yeast genome has homologues of the genes for all the budding yeast endocytic/actin cytoskeletal adaptor proteins that assemble into a nascent actin patch before Arp2/3 complex (Kaksonen et al., 2003). For example, deletion or depletion of the homologues SLA2 in budding yeast (Kaksonen et al., 2003), Hip1R in cultured cells (Engqvist-Goldstein et al., 2004), or end4+ in *S. pombe* (Iwaki et al., 2004) cause endocytic defects.

Electron micrographs show budding yeast actin patches associated with membrane invaginations and vesicles (Mulhol-land et al., 1994; Jonsdottir and Li, 2004; Young et al., 2004; Rodal et al., 2005). Actin patches purified from budding yeast contain a network of short (50 nm) branched filaments, as expected for assembly driven by Arp2/3 complex (Young et al., 2004). Based on orientation of filaments in actin comet tails in budding yeast Δsla2, Kaksonen et al. (2003) proposed that filament barbed ends push against the plasma membrane. This is consistent with the localization of Arp2/3 complex and actin observed by Rodal et al. (2005).

Fission yeast have structures called filasomes, 300-nm clouds of actin filaments surrounding vesicles 35–70 nm in diameter (Takagi et al., 2003). Filasomes were proposed to be exocytic vesicles, but they may actually be actin patches associated with endocytosis. If so, then 300 nm is the maximum length filaments grow in 6 s of patch assembly. That corresponds to elongation at a rate of ~20 subunits per second and free G-actin concentration of 2 μM. This may be an underestimate, as each filament may grow for <0.5 s, owing to termination by capping protein.

Actin assembly at the neck of an endocytic vesicle is proposed to assist in either pinching off or propelling the vesicle away from plasma membrane (Munn, 2001; Engqvist-Goldstein and Drubin, 2003; Merrifield, 2004). Fission yeast patches are stationary as they assemble and initiate motility only when fully grown. This is consistent with the idea that patches move after actin assembly severs the vesicle from the plasma membrane (Kaksonen et al., 2003).

Actin polymerization may drive short local movements of actin patches, but long-range movements seem to depend on association with moving actin cables in both fission (Pelham and Chang, 2001) and budding yeast (Huckaba et al., 2004). The brief (~20 s) lifetime and slow (30–230 nm/s) movements of Arp2/3 complex patches are significantly different from lifetime of over 30 s and rates of movement of 300 nm/s of actin patches labeled with coronin Crn1p-GFP or App1p-GFP (Pelham and Chang, 2001). We confirmed (unpublished data) that Crn1p-GFP behaves differently from GFP-tagged Arp2/3 complex, capping protein, fimbrin, thinfilin, and actin in that it lacks a distinct stationary phase and has a prolonged mobile phase. Crn1p-GFP may join patches at the onset of mobile phase and remain in the patch longer than other markers.

**Materials and methods**

**Bacterial expression constructs**

For NH2-terminal GST tagging, DNA inserts encoding Myo1p TH2-SH3-CA, TH2-SH3, and SH3-CA tail fragments, as defined by Lee et al. (2000), or Wsp1p VCA (aa 497–574), bovine N-WASP VCA (aa 422–505), and VVCA (aa 402–505) were subcloned into BamH1 and EcoR1 sites of pGEX-2T and pGEX-6P-1 (GE Healthcare). To add COOH-terminal Histag, full-length Vrp1p cDNA was subcloned into Nde1 and Xho1 sites of pET21a (Novagen). Myo1p and N-WASP inserts were PCR amplified using Turbo Pfu (Stratagene) from pBS-myo1 (Lee et al., 2000) and N-WASP-VVCA in pGEX-2T, respectively. To avoid introns, inserts for Wsp1p and Vrp1p constructs were amplified by RT-PCR (GIBCO BRL) from total *S. pombe* RNA.

**Protein purification**

Native *S. pombe* Arp2/3 complex was purified from protease-deficient TM011 cells resuspended in buffer U (50 mM Hepes, pH 7.5, 100 mM KCl, 3 mM MgCl2, 1 mM EGTA, 0.1 mM ATP, and 1 mM DTT) containing Complete (Roche) protease inhibitors and ruptured using a Microfluidizer (model M-110S; Microfluidics). After lysates were spun at 100,000 g, proteins were precipitated by 50% ammonium sulfate, solubilized in buffer U, and dialyzed against buffer A (50 mM Tris-HCl, pH 7.5, 25 mM KCl, 1 mM MgCl2, 1 mM EGTA, 0.1 mM ATP, and 1 mM DTT) containing 0.5 mM PMSF. Arp2/3 complex was bound to GST-N-WASP-VCA immobilized on glutathione-Sepharose, eluted with 1 M NaCl in buffer A, dialyzed against buffer B (10 mM Pipes, pH 6.8, 0.25 mM MgCl2, 0.25 mM EGTA, and 1 mM DTT), and further purified by ion exchange chromatography on a Source 15Q column (AKTA FPLC; GE Healthcare).

Vr1p-His was expressed in Escherichia coli strain Rosetta (DE3) plysS (Novagen) at 22°C and purified using Ni-NTA resin (QIAGEN). Eluate from Ni-NTA column was dialyzed against buffer QA (10 mM Tris-HCl, pH 8.0, 1 mM EGTA, and 1 mM DTT), passed over a Source 15Q column, and fractionated on a Source 15Q column. Pure Vr1p-His eluted in 145 mM NaCl.

GST fusion proteins were purified as described for human WASp GST-VCA (Higgs et al., 1999). GST-tagged Myo1p fragments were stored in buffer QA containing 275 mM NaCl.

**Actin polymerization assays**

Actin polymerization assays were performed using an Alphascan spectrofluorimeter (Photon Technology International) and analyzed as described previously (Higgs et al., 1999). Products of actin polymerization were
stained with equimolar rhodamine-phalloidin (Fluka) added at the reaction onset as described by Blanchain et al. (2000). Images were collected on a microscope (model IX71; Olympus) equipped with a 60×, 1.4 NA Plan-Apo lens using an ORCA-ER CCD camera (Hamamatsu Corporation) controlled by MetaMorph (Universal Imaging Corp.).

Quantitative pull-down assays
Equilibrium dissociation constants (Kd) were measured by quantitative pull-down assays (Lee et al., 1999). GST- or His-tagged receptors at variable concentrations [R] were immobilized on beads and incubated with soluble ligand at constant concentration [L]. Concentrations of unbound ligand were measured by gel densitometry and fraction of ligand bound [LR]/[L] was fitted to binding isotherm in KaleidaGraph (Synergy Software): 

$$[LR]/[L] = ([R] + [L] - K_d) / ([R] + [L] + K_d) = 2^{-4}*([R]([L])^{0.5})^2/[L].$$

Construction of yeast strains
Genes at their chromosomal loci were either deleted or tagged with fluorescent protein (FP) sequences using the PCR-based gene tagging technique (Bahl et al., 1998; Wu et al., 2003). In ∆pvr1, ∆myo1, ∆wsp1 pFA6a-kanMX6 cassette replaced the entire ORFs. Vrp1p was tagged at the COOH terminus with nonmonomeric FPs by integrating pFA6a-GFP(S65T)-kanMX6 cassette and YFP and CFP derivatives in place of the COOH terminus with monomeric FPs containing A206K mutation (Zacharias et al., 2002) and separated from the ARPC5 by GGGRGG linker. Myo1p and Wsp1p were tagged at their NH2 termini with monomeric FPs and expressed under control of native promoters by replacing myo1·Δ106 to +47 and wsp1·Δ112 to +3 with monomeric FP derivatives of pFA6a-kanMX6·3pmt1·GFP in which pmt1·GFP was replaced at the COOH terminus with monomeric FPs containing A206K mutation. Strains combining tags and deletions of two genes were constructed by genetic crosses. To promote mating, of two genes were constructed by genetic crosses. To promote mating, and S. Forsburg for valuable advice. We thank C. Dillingham and K. Macmillan for help with patch tracking. This work was supported by National Institutes of Health grants 26132 and 26338 (to T.D. Pollard) and American Heart Association post-doctoral fellowship 0225759T (to V. Sirotkin).

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Submitted: 8 February 2005
Accepted: 13 July 2005

Online supplemental material
Video 1 shows a time-lapse movie of mYFP-Myo1p (red) and ARPC5-mCFP (green). Video 2 shows a time-lapse movie of mYFP-Wsp1p (red) and ARPC5-mCFP (green). Fig. S1 shows truncation analysis of Myo1p tail. Fig S2 depicts localization and dynamics of GFP-tagged proteins. Table S1 lists lifetimes of GFP-tagged proteins in patches. Table S3 lists yeast strains. Online supplemental material available at http://www.jcb.org/cgi/content/full/jcb.200502053/DC1.

We are grateful to Drs. T. Toda and M. Yanagida for protease-deficient yeast strain, R. Li for a protocol to purify Arp2/3 complex, W.V. Lee for Myo1p construct, H. Higges for N-WASP construct, J. Kuhn for help with image analysis, and S. Fording for valuable advice. We thank C. Dillingham and K. Macmillan for help with patch tracking.

This work was supported by National Institutes of Health grants 26132 and 26338 to T.D. Pollard and American Heart Association post-doctoral fellowship 0225759T to V. Sirotkin.
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