Ezrin/Radixin/Moesin (ERM) Proteins Bind to a Positively Charged Amino Acid Cluster in the Juxta-Membrane Cytoplasmic Domain of CD44, CD43, and ICAM-2

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Abstract. CD44 has been identified as a membrane-binding partner for ezrin/radixin/moesin (ERM) proteins, plasma membrane/actin filament cross-linkers. ERM proteins, however, are not necessarily colocalized with CD44 in tissues, but with CD43 and ICAM-2 in some types of cells. We found that glutathione-S-transferase fusion proteins with the cytoplasmic domain of CD43 and ICAM-2, as well as CD44, bound to moesin in vitro. The regions responsible for the in vitro binding of CD43 and CD44 to moesin were narrowed down to their juxta-membrane 20–30–amino acid sequences in the cytoplasmic domain. These sequences and the cytoplasmic domain of ICAM-2 (28 amino acids) were all characterized by the positively charged amino acid clusters. When E-cadherin chimeric molecules bearing these positively charged amino acid clusters of CD44, CD43, or ICAM-2 were expressed in mouse L fibroblasts, they were co-concentrated with ERM proteins at microvilli, whereas those lacking these clusters were diffusely distributed on the cell surface. The specific binding of ERM proteins to the juxta-membrane positively charged amino acid clusters of CD44, CD43, and ICAM-2 was confirmed by immunoprecipitation and site-directed mutagenesis. From these findings, we conclude that ERM proteins bind to integral membrane proteins bearing a positively charged amino acid cluster in their juxta-membrane cytoplasmic domain.

Ezrin/radixin/moesin (ERM) proteins are thought to function as general cross-linkers between plasma membranes and actin filaments (Bretscher, 1983; Pakkanen et al., 1987; Lankes et al., 1988; Tsukita et al., 1989; Algrain et al., 1993; Arpin et al., 1994; Tsukita et al., 1997a,b). In cultured cells, ERM proteins are mostly coexpressed and concentrated just beneath specialized domains of plasma membranes such as microvilli and cell–cell or cell–substrate adhesion sites, where actin filaments are densely associated. In differentiated tissues, however, their expression levels are specifically regulated (Bretscher, 1983; Pakkanen et al., 1987; Lankes et al., 1988; Tsukita et al., 1989, 1992; Sato et al., 1991, 1992; Berryman et al., 1993; Franck et al., 1993; Amieva et al., 1994; Takeuchi et al., 1994b; Henry et al., 1995). The suppression of ERM protein expression with antisense oligonucleotides in cultured cells destroys cell surface structures such as microvilli and cell adhesion sites (Takeuchi et al., 1994b).

Sequencing of cDNAs has revealed that the amino acid sequence identity among ERM proteins is 70–80% (Gould et al., 1989; Turunen et al., 1989; Funayama et al., 1991; Lankes and Furthmayr, 1991; Sato et al., 1992). The sequences of their amino-terminal halves are highly conserved (~85% identity) and homologous to the amino-terminal ends of some membrane-associated proteins, such as band 4.1 protein, talin, merlin–schwannomin (a tumor suppressor molecule for neurofibromatosis type II), indicating that the ERM family is included in the band 4.1 superfamily (Conboy et al., 1986; Rees et al., 1990; Rouleau et al., 1993; Trofatter et al., 1993; Takeuchi et al., 1994a; Arpin et al., 1994; Tsukita et al., 1997a,b). Because the amino-terminal domain in band 4.1 protein is responsible for its direct association with the integral membrane protein, glycoporphin C (Bennet, 1989), ERM proteins were thought to associate with integral membrane proteins through their
that ERM proteins bind to integral membrane proteins by immunoprecipitation. Based on these findings, we concluded that ERM binding was also confirmed by transfection and immunofluorescence microscopy. Rat anti-ERM mAb (TK89) (Takeuchi et al., 1994b). TK89 recognizes ezrin, radixin, and moesin both by immunoblotting and immunofluorescence microscopy. Rat anti-E-cadherin mAb (ECCD-2; Shirayoshi et al., 1986) was provided by M. Takeichi (Kyoto University, Kyoto, Japan).

Production and Purification of Glutathione-S-Transferase (GST) Fusion Proteins with Cytoplasmic Domains of Integral Membrane Proteins

Various cDNA fragments for mouse CD44 and mouse ICAM-2 were obtained by reverse transcriptase-polymerase chain reaction, using mouse lung total RNA as a template, and those for rat CD44 were generated by PCR, using LSP-1 (Yonemura et al., 1993) as a template. They were then subcloned into pBluescript SK− (Stratagene, La Jolla, CA) and confirmed by sequencing with an ABI PRISM cycle sequencing kit (PerkinElmer Corp., Foster City, CA).

The PCR products subcloned into pBluescript SK− were excised and subcloned into pGEX2T vector (Pharmacia Diagnostics AB, Uppsala, Sweden) to produce GST fusion proteins with full-length and various truncated cytoplasmic domains of CD44 and CD43. The cytoplasmic domain of mouse CD44 contains 70 amino acids (He et al., 1992), and the following GST–CD44 cytoplasmic domain fusion proteins were produced (Fig. 1): G-44 containing the whole cytoplasmic domain of amino acids (a.a.)1–70, G-44/1–19 containing a.a.1–19, G-44/1–31 containing a.a.1–31 plus INN at the carboxyl terminus as a result of construction, and G-44/19–70 containing a.a.19–70. The cytoplasmic domain of rat CD 43 contains 124 amino acids (Kilien et al., 1987), and the following GST–CD43 cytoplasmic domain fusion proteins were produced (see Fig. 1): G-43 containing the entire cytoplasmic domain of a.a.1–124, G-43/1–31 containing a.a.1–31 plus INN5 at the carboxyl terminus, G-43/1–39 containing a.a.1–39 plus EFIVTD, G-43/1–64 containing a.a.1–64 plus EFIVTD, G-43/38–124 containing a.a.38–124, G-43/62–124 containing a.a.62–124, and G-43/78–124 containing a.a.78–124. The cytoplasmic domain of mouse ICAM-2 contains 28 amino acids (Xu et al., 1992), so only G-ICAM-2 containing the whole cytoplasmic domain was produced. We produced GST fusion proteins with the whole cytoplasmic domain of mouse E-cadherin (G- Ecad; Nagafuchi et al., 1987) and occludin (G-Oc; Furuse et al., 1994) as controls. Site-directed mutagenesis was performed by PCR, using appropriate mutagenic primers in pBluescript SK− vectors containing cDNAs encoding CD44, CD43, or ICAM-2 (see Fig. 8).

The GST fusion proteins were produced and purified basically according to the method of Smith and Johnson (1988) in Escherichia coli IM109 on SF9 cells. Synthesis of the GST fusion proteins was induced by incubating bacteria with 0.2 mM isopropyl β-D-thiogalactopyranoside for 2–5 h at 37°C. The cells were sedimented by centrifugation and the cell pellet was solubilized in buffer A (20 mM Tris buffer, pH 8.0, 150 mM NaCl, 1 mM EDTA, 1 mM DTT, 1.5% Sarkosyl, 1 mM PMSF, 20 μg/ml leupeptin) at 4°C according to the method of Frangioni and Neel (1993). Sarkosyl effectively decreased degradation of the fusion proteins during purification, which had not been technically circumvented in our previous study using E. coli (Hirao et al., 1996). After sonication, the cell debris was removed by centrifugation (10,000 g, 10 min, at 4°C) and the supernatant was mixed with an equal volume of buffer B (the same as buffer A except that 5% Triton X-100 was used instead of sarkosyl). The supernatant was mixed with glutathione-Sepharose 4B beads (Pharmacia Diagnostics AB) that had been washed with buffer C (1:1 mixture of buffers A and B) and then gently shaken for 10–30 min at 4°C. The beads were washed with buffer C to remove unbound bacterial proteins and stored on ice. The amount of GST fusion protein bound to the beads was estimated by SDS-PAGE.

Materials and Methods

Cells and Antibodies

SF9 and High Five cells (Invitrogen, Carlsbad, CA) were cultured in TC-100 medium (GIBCO BRL, Gaithersburg, MD) supplemented with tryptophane phosphate broth (GIBCO BRL) and 10% FCS at 27°C. Mouse fibroblastic L cells (Earle, 1943) were cultured in DME with 10% FCS. Monolayers of transfected ERM proteins, using mouse anti-ERM mAb, CR22 (Sato et al., 1991), which has higher affinity for moesin than for ezrin and radixin, rat antiezrin mAb (M11), rat antiradixin mAb (R2-1), rat antimoesin mAb (M22), and rabbit anti-ERM pAb (TK89) (Takeuchi et al., 1994b) were used as reagents. Mouse anti-ERM mAb, CR22, was kindly provided by Dr. H. Nagafuchi (Kyoto University, Kyoto, Japan).

In Vitro Binding Assay between ERM Proteins and GST Fusion Proteins

Mouse ezrin, radixin, and moesin were produced by recombinant baculovirus infection and purified as described (Hirao et al., 1996). For each reaction, 15–60 μl of glutathione-Sepharose bead slurry containing a GST fusion protein was suspended in 1 ml of buffer D (10 mM Hepes buffer, pH 7.5, 40 or 150 mM KCl, 1 mM MgCl2, 1 mM EDTA, 1 mM DTT, 2 μg/ml 100 medium (GIBCO BRL, Gaithersburg, MD) supplemented with tryp-
ml leupeptin) in a 1.5-ml tube, and recovered as a pellet by centrifugation (10,000 g, 1 min). After removing the supernatant, the pellet was resuspended in 1 ml of buffer D and this wash was repeated three times. Ezrin, radixin, or moesin was added to make 100–200 μl of bead suspension in buffer D containing 0.5–1 μg of ERM proteins. The beads were incubated for 30 min at room temperature with occasional mixing, and washed five times with buffer D by centrifugation. GST fusion protein was eluted with its associated protein using 150 μl of 50 mM Tris buffer (pH 8.0) containing 30 mM glutathione. The amount of GST fusion protein in each eluate was determined by SDS-PAGE. An appropriate amount of each eluate was again subjected to SDS-PAGE to contain the same amount of GST fusion protein. The amount of bound ERM protein was determined by immunoblotting with specific mAbs followed by densitometric scanning using a software NIH Image V1.54 and then relative amount of moesin bound per GST fusion protein (mol) was calculated.

**SDS-PAGE and Immunoblotting**

SDS-PAGE (12.5 or 10%) was performed according to the conventional method, and gels were stained with Coomassie brilliant blue R-250. For immunoblotting, proteins were electrophoretically transferred from gels onto nitrocellulose membranes. After incubation with first antibody, bound antibodies were visualized using biotinylated secondary antibody followed by densitometric scanning using a software NIH Image V1.54 and then relative amount of moesin bound per GST fusion protein (mol) was calculated.

**Mammalian Expression Vectors and Transfection**

As shown in Fig. 1b, a series of E-cadherin chimeric proteins with full-length or truncated cytoplasmic domains of mouse CD44, rat CD43, and mouse ICAM-2 were expressed in mouse L cells as described (Yonemura et al., 1993). These constructs corresponded to those of various GST fusion proteins (see Fig. 1a). Appropriate restriction sites were introduced to the extracellular domain of mouse E-cadherin (from the amino terminus to BstPI site) and several amino acids of the extracellular domain/transmembrane domain/cytoplasmic domain of CD44, CD43, or ICAM-2.

All E-cadherin/CD44 chimeric molecules contained four amino acids of the extracellular domain of CD44. In E-44/20–70, N in the transmembrane domain located near the transmembrane–cytoplasmic junction was converted to T. All E-cadherin/CD43 chimeric molecules contained 23 amino acids of the extracellular domain of CD43. Both E-43/1–31 and E-43/1–47 contained additional FGIL at the carboxyl terminus, and E-43/1–9,49–124 contained RSA between amino acids 9 and 49. In our previous study, E-43, E-43/1–47, and E-43/1–9,49–124 were called CLS-1, CLS1-A, and CLS1-B, respectively (Yonemura et al., 1993). The E-cadherin/ICAM-2 chimera (E-ICAM-2) contained 3 amino acids of the extracellular domain of ICAM-2. Site-directed mutagenesis was performed by PCR using appropriate mutagenic primers in pBluescript SK+ vectors containing cDNAs encoding E-44, E-43, or E-ICAM-2 (see Fig. 8).

L cells were transfected with DNA using lipofectin or lipofectamine reagent (GIBCO BRL). Cells cultured on coverslips were washed twice with Opti-MEM (GIBCO BRL), and were incubated for 3–5 h with 1 ml Opti-MEM containing 1 μg of plasmid DNAs and 10 μl of the reagents, followed by the addition of 3 ml of normal medium containing FCS. Cells were then cultured for 2–3 d. L cells were also transfected by microinjection using a set of manipulators (MN-188 and MO-189; Narishige, Tokyo, Japan) connected to a microinjector 5242 (Eppendorf, Inc., Hamburg, Germany). Expression vectors in injection buffer (100 mM KCl, 10 mM Hepes buffer, pH 7.5) were injected into the nuclei of cells cultured on coverslips. Cells were examined 12–24 h after injection.

**Immunofluorescence Microscopy**

All procedures were performed at room temperature. Cells were fixed with 1–4% formaldehyde in 0.1 M Hepes buffer (pH 7.5) for 10–15 min. After three washes with PBS containing 30 mM glycine (G-PBS), cells were soaked in blocking solution (G-PBS containing 2% normal goat
serum) for 5 min and incubated with anti–E-cadherin mAb (ECCD-2) diluted with the blocking solution for 30 min. The cells were then washed three times with G-PBS, treated with 0.2% Triton X-100 in G-PBS for 10 min, and washed with G-PBS. The cells were soaked in blocking solution for 10 min, incubated with CR22 for 30 min, washed three times with G-PBS, and incubated with secondary antibodies. FITC-conjugated goat anti–rat Ig antibody (BioSource, Camarillo, CA) and rhodamine-conjugated goat anti–mouse IgG antibody (Chemicon International, Inc., Temecula, CA) were used as secondary antibodies. Cells were washed three times, and then mounted in 90% glycerol-PBS containing 0.1% para-phenylenediamine and 1% n-propylgalate. Specimens were observed using a Zeiss Axiohot photomicroscope (Carl Zeiss, Oberkochen, Germany). Images were taken on T-MAX 400 film (Eastman Kodak Co., Rochester, NY), or recorded with a cooled CCD camera (SenSys 0400, 768X512 pixels; Photometrics, Tucson, AZ) controlled by a Power Macintosh 7600/132 and the software package IPLab Spectrum V3.1 (Signal Analytics Corp., Vienna, VA).

**Immunoprecipitation**

Confluent monolayer cultures of stable L transfectants expressing E-43 or E-43/1–9,49–124 (Yonemura et al., 1993) on 10-cm dishes were used for immunoprecipitation. All procedures were carried out on ice. Cells were washed twice with a solution containing 150 mM NaCl, 1 mM CaCl₂, 1 mM MgCl₂, and 10 mM Hepes (pH 7.5), and then lysed in 1 ml of lysis buffer (150 mM NaCl, 1 mM CaCl₂, 1 mM MgCl₂, 10 mM Hepes (pH 7.5), 0.1% Nonident P-40, 1 mM DTT, 20 µg/ml leupeptin) for 30 min. The lysate was removed from the dish after fully dislodging any remaining cellular debris with a rubber policeman. After centrifugation at 12,000 g for 20 min, the supernatant was incubated for 1 h with 10 µl of protein G–Sepharose 4B beads (Zymed Labs, Inc., South San Francisco, CA) conjugated with anti–E-cadherin mAb, ECCD-2. The beads were collected and washed with lysis buffer five times by centrifugation at 1,250 g for 2 min. The immune complexes were eluted from the beads in 300 µl of 1 M CH₃COOH for 10 min. The supernatant was freeze-dried and separated by SDS-PAGE followed by immunoblotting with ECCD-2 or TK9.

**Results**

**In Vitro Binding of Moesin to GST Fusion Proteins with CD44, CD43, and ICAM-2**

We established an in vitro binding assay to evaluate the interaction between recombinant ERM proteins and GST fusion protein with the cytoplasmic domain of CD44 (Hirao et al., 1996). Using this assay, we first compared the binding abilities of CD44, CD43, and ICAM-2 to recombinant moesin as a representative ERM protein. GST fusion proteins with the whole cytoplasmic domain of CD44 (G-44), CD43 (G-43), or ICAM-2 (G-ICAM-2) were purified on glutathione-Sepharose beads. As controls, GST fusion proteins with the whole cytoplasmic domain of E-cadherin (G-E-cad) and occludin (G-Oc) were also purified. These fusion protein-bound glutathione-Sepharose beads were incubated with recombinant moesin, washed, and eluted with glutathione. The eluate contained GST fusion protein and its associated protein. Moesin association with GST fusion proteins was evaluated by immunoblotting with anti-moesin mAb followed by densitometry (Fig. 2). At low ionic strength (40 mM KCl), G-43 and G-44 bound to moesin with similar affinity. At physiological ionic strength (150 mM KCl), G-43 still bound to moesin, whereas the binding ability of G-44 to moesin was significantly decreased as previously reported (Hirao et al., 1996). G-ICAM-2 bound to moesin with affinity similar to G-43 at physiological ionic strength. In contrast, G-E-cad, G-Oc and GST showed no binding affinity to moesin. Considering that the dissociation constant between the cytoplasmic domain of CD44 and moesin at 40 mM KCl is ~10 nM (Hirao et al., 1996), these findings showed that moesin directly and specifically binds to the cytoplasmic domains of CD43 and ICAM-2 at physiological ionic strength. We checked here that ezrin also bound to CD43 (data not shown), but other binding combinations remain to be examined.

**Moesin-binding Sites in the Cytoplasmic Domain of CD44 and CD43**

Because there is no significant homology among the cytoplasmic domains of CD44, CD43, and ICAM-2, we attempted to define the region responsible for moesin binding of each protein. Mouse CD44, rat CD43, and mouse ICAM-2 have 70, 124, and 28 amino acids in their cytoplasmic domains, respectively. We then constructed various deletion series of GST–CD44 and GST–CD43 fusion proteins (Fig. 1a), and performed an in vitro binding assay
using recombinant moesin at physiological ionic strength. As shown in Fig. 3a, in the CD44 deletion series, G-44/1–19 and G-44/1–31 bound to moesin whereas G-44/19–70 did not. This indicates that the amino acid sequence of the juxta-membrane region (a.a.1–19) is responsible for moesin binding. The binding affinities of G-44/1–19 and G-44/1–31 to moesin at 150 mM KCl were similar to that of G-44 at 40 mM KCl ($K_d \approx 10$ nM), suggesting an intramolecular suppressive interaction between the amino- and carboxyl-terminal parts of the cytoplasmic domain of CD44 under physiological conditions. We also found that G-44/1–19 and G-44/1–31 bound to ezrin and radixin with affinity similar to moesin under physiological conditions (data not shown).

The CD43 deletion series yielded rather complex results (Fig. 3b). G43/1–31 and G43/62–124 bound to moesin with affinities similar to the full-length cytoplasmic domain of CD43 (G-43). However, a construct longer than G43/1–31 (G43/1–39) weakly bound to moesin, and that longer than G43/62–124 (G43/38–124) also showed weak binding. Furthermore, G43/1–64 and G43/78–124 hardly bound to moesin. These observations indicate that there are two moesin-binding regions in the cytoplasmic domain of CD43 at least in vitro. One is located in the juxta-membrane domain (a.a.1–31) and the other is in the middle part of the cytoplasmic domain (a.a.62–78). The region between these two domains (a.a.32–61) appeared to be inhibitory for moesin binding. Similar results were obtained using ezrin (data not shown).

Colocalization of E-cadherin Chimeric Molecules Containing Cytoplasmic Domains of CD44, CD43, and ICAM-2 with ERM Proteins in Transfected Cells and Responsible Domains

To assess the physiological relevance of the above in vitro binding results, we constructed and introduced E-cadherin chimeric molecules consisting of the extracellular domain of E-cadherin and whole or various deletion constructs of the transmembrane/cytoplasmic domain of CD44, CD43, or ICAM-2 (Fig. 1b) into mouse L fibroblasts that did not express any endogenous E-cadherin. Using an anti–E-cadherin mAb specific for the extracellular domain of E-cadherin, we then compared their subcellular distributions with those of ERM proteins by immunofluorescence microscopy. In parent L cells, ezrin, radixin, and moesin were precisely colocalized at microvilli, and the full-length E-cadherin introduced in L cells was not concentrated at microvilli (data not shown; see Yonemura et al., 1993). Because in L cells moesin was predominant among ERM proteins, the data obtained with antimoesin mAb (CR-22) are shown here.

As shown in Fig. 4a, E-44 containing the full-length cytoplasmic domain of CD44 was co-concentrated with moesin at microvilli. Considering that endogenous CD44 was reported to be colocalized with moesin in microvilli of cultured fibroblasts (Tsukita et al., 1994), this finding indicates that the cytoplasmic, but not the extracellular do-
main of CD44 is responsible for its co-concentration with moesin in microvilli. We constructed various cytoplasmic domain deletion mutants of this chimeric molecule and introduced them into L cells (Fig. 1b). E-44/1–19 (Fig. 4, c and d) as well as E-44/1–32 (data not shown) colocalized with moesin at microvilli, whereas E-44/20–70 was diffusely distributed over the cell surface and not concentrated at microvilli (Fig. 4, e and f). These observations indicated that the juxta-membrane region (a.a.1–19) of the cytoplasmic domain of CD44 is sufficient for E-cadherin–CD44 chimeric molecules to colocalize with moesin at microvilli (see Fig. 7).

In our previous study using the same transfection system, we found that E-43, which contains the full-length cytoplasmic domain of CD43, was co-concentrated with moesin at microvilli, and that E-43/1–47 was also colocalized with moesin, whereas E-43/1–9,49–124 was not concentrated at microvilli (Yonemura et al., 1993; see Fig. 7). In this study, to further define the responsible domain, E-43/1–31 was introduced into L cells (Fig. 5, a and b). E-43/1–31 was highly concentrated at microvilli together with moesin, indicating that the juxta-membrane region (a.a.1–31) of the cytoplasmic domain of CD43 is responsible for colocalization of these E-cadherin–CD43 chimeric molecules with moesin.

We also produced an E-cadherin chimera with the whole cytoplasmic domain of ICAM-2 (E-ICAM-2) and transfected it into L cells. As shown in Fig. 5, c and d, this molecule was again highly concentrated at microvilli together with moesin.

**Coimmunoprecipitation of ERM Proteins with E-43 from L Cell Transfectants**

The association of ERM proteins with CD44 inside cells was detected by immunoprecipitation from BHK cells, but the detection of ERM protein–integral membrane protein complex by immunoprecipitation was difficult, because the molecular complex resists the detergent extraction due to its tight association with actin-based cytoskeletal components (Tsukita et al., 1994). However, probably because relatively large amounts of E-cadherin–CD43 chimeric molecules appeared on the cell surface of L cell transfectants, and probably because anti–E-cadherin mAb was very potent for immunoprecipitation, we were able to compare the amounts of ERM proteins in E-cadherin immunoprecipitates from E-43 stable transfectants with that from E-43/1–9,49–124 stable transfectants (in the former transfectants E-43 was co-concentrated with moesin at microvilli, whereas in the latter E-43/1–9,49–124 was not [see Fig. 7]). As shown in Fig. 6, ERM proteins were coimmunoprecipitated with E-43, but not with E-43/1–9,49–124. This finding indicates not only that the juxta-membrane domain of CD43 is responsible for the CD43–ERM pro-
Positively Charged Amino Acid Clusters Responsible for ERM Protein–Integral Membrane Protein Binding

As summarized in Fig. 7, in vitro binding analysis identified a.a.1–19 of the cytoplasmic domain of CD44, a.a.1–31, and 62–78 of the cytoplasmic domain of CD43, and the whole cytoplasmic domain (a.a.1–28) of ICAM-2 as moesin-binding sites. On the other hand, transfection experiments with E-cadherin chimeric molecules indicated that a.a.1–19 of CD44-, a.a.1–31 of CD43-, and a.a.1–28 of ICAM-2 cytoplasmic domains are responsible for co-concentration of these integral membrane proteins with ERM proteins at microvilli. Together with the results obtained from immunoprecipitation analysis, we concluded that the in vitro moesin-binding ability of a.a.62–78 of the CD43-
cytoplasmic domain was not indispensable in the cells, and that the juxta-membrane 20–30-amino acid sequence of the cytoplasmic domains of CD43, CD44, and ICAM-2 can bind to ERM proteins and recruit these integral membrane proteins to microvilli together with ERM proteins. These three juxta-membrane domains showed no significant similarity in their amino acid sequences, but they were characterized by many positively charged amino acids, such as R and K (Fig. 8). To evaluate the importance of the positively charged amino acid clusters of CD44, CD43, and ICAM-2 in their binding to ERM proteins, we substituted the juxta-membrane KKK in G-44, KRR in G-43, and RRR in G-ICAM-2 with QIN, NGG, and GGA, respectively (Fig. 8). Using these site-directed mutants of GST fusion proteins (G-44/KKK:QIN, G-43/KRR:NGG, and G-ICAM-2/RRR:GGA, respectively) and recombinant moesin, we then performed an in vitro binding assay. As shown in Fig. 9, all of the mutant proteins lost their binding ability to moesin. Furthermore, when E-cadherin chimeric molecules with corresponding mutants (E-44/KKK:QIN, E-43/KRR:NGG, and E-ICAM-2/RRR:GGA) were constructed and introduced into L cells, all of these molecules were distributed diffusely on the cell surface (Fig. 10). They were not excluded from microvilli, but were not co-concentrated with moesin at microvilli. These findings indicated the importance of the juxta-membrane positively charged amino acid clusters in the ERM protein–integral membrane interaction.

Discussion

Binding of ERM Proteins to CD43 and ICAM-2 As Well As CD44 through Their Juxta-Membrane Positively Charged Amino Acid Clusters

In this study, we first compared CD44 and ICAM-2 with CD44 in terms of moesin association in vitro. Close analyses using various deletion mutants revealed that moesin bound in vitro to juxta-membrane positively charged amino acid clusters of these membrane proteins. Judging from the affinity of moesin–CD44 binding (Kd = ~10 nM; Hirao et al., 1996), moesin binding to the clusters of CD43 and ICAM-2 appeared to be physiologically significant. Next, by transfecting various mutants of E-cadherin/CD-44 (E-44), E-cadherin/CD-43 (E-43), and E-cadherin/ICAM-2 (E-ICAM-2) chimeric molecules into L cells, we narrowed down the domains that were required for their colocalization with moesin at microvilli, and found that their juxta-membrane positively charged amino acid clusters were again responsible. These observations, together with the results of immunoprecipitation and site-directed mutagenesis studies, led us to conclude that moesin bound not only to the juxta-membrane region of CD44 but also to those of CD43 and ICAM-2 in vivo.

Most of the in vitro data presented here were from experiments using recombinant moesin, but we also confirmed that ezrin and radixin behaved in the same manner as moesin in several experiments. CD44 was reported to be associated with not only moesin but also ezrin and radixin both in vitro and in vivo (Tsukita et al., 1994; Hirao et al., 1996). Furthermore, in this study, ezrin and radixin,
as well as moesin, were coimmunoprecipitated with E-43, and all ERM proteins were colocalized with E-44, E-43, and E-ICAM-2 in L cell transfectants. We therefore concluded that not only moesin, but also ezrin and radixin, bound to the juxta-membrane region of CD43, ICAM-2, as well as CD44.

**Specificity of the Binding of ERM Proteins to the Juxta-Membrane Positively Charged Amino Acid Clusters**

Most of the transmembrane proteins have positively charged amino acid residues in their juxta-membrane regions, and these residues are thought to form an anchor, arresting translocation across the bilayer during biosynthesis and assuring the correct topological orientation of membrane proteins (Boyde and Beckwith, 1990). For example, there are several positively charged amino acid residues in the juxta-transmembrane domains of E-cadherin and occludin, which did not bind to ERM proteins. As shown in Table I, we found that the balance of positively and negatively charged amino acid residues in the whole cytoplasmic domain was significantly different between ERM-binding proteins such as CD44, CD43, and ICAM-2, and ERM-nonbinding proteins such as E-cadherin and occludin. The calculated isoelectric points of the whole cytoplasmic domains of CD44, CD43, ICAM-2, E-cadherin, and occludin were 8.17, 9.24, 12.98, 3.89, and 5.85, respectively, indicating that, at neutral pH, CD44, CD43, and

| Integral membrane protein | Calculated isoelectric point | ERM binding |
|---------------------------|-----------------------------|-------------|
| CD44                      | 8.17                        | Weak        |
| CD43                      | 9.24                        | Strong      |
| ICAM-2                    | 12.98                       | Strong      |
| E-cadherin                | 3.89                        | Undetectable|
| Occludin                  | 5.85                        | Undetectable|

The isoelectric points were calculated using a program from GENETYX software package (Software Development Co., Tokyo, Japan) using the sequence data obtained from EMBL/GenBank/DBJ under accession numbers X66081 (mouse CD44), Y00090 (rat CD43), X6549 or S46669 (mouse ICAM-2), X06115 (mouse E-cadherin), and D21837 (chick occludin).
ICAM-2 have a net positive charge over their whole cytoplasmic domain and that E-cadherin and occludin have a net negative charge.

Several integral membrane proteins, L-selecin (Picker et al., 1991), ICAM-1 (Carpén et al., 1992), integrins α4β7 and α4β1 (Berlin et al., 1995), and P-selectin glycoprotein ligand-1 (Moore et al., 1995) were reported to be localized at microvilli. L-selecin and ICAM-1 have short cytoplasmic domains (~10 and ~30 amino acids, respectively) and their net charges are highly positive. It is postulated that α-actinin is a membrane–cytoskeleton linker for these membrane proteins. However, L-selecin lacking the α-actinin–binding site, which bears only a positively charged six–amino acid cytoplasmic domain (RRLKKG), is still localized at microvilli (Carpén et al., 1992; Pavalko et al., 1995), and ICAM-1 was reported to associate with ezrin (Heland er et al., 1996). Both α4 integrin and P-selectin glycoprotein ligand-1 have positively charged amino acid clusters of ~30 amino acids in their juxta-membrane region.

Taken together, for integral membrane proteins, the positively charged amino acid cluster in the juxta-membrane cytoplasmic domain might be a default signal for ERM binding and/or microvillar localization within cells. When the positively charged amino acid cluster is followed by a negatively charged amino acid cluster, i.e., the net charge of the whole cytoplasmic domain is negative or nearly neutral, the former cluster may be masked by the latter, losing its binding ability to ERM proteins. This hypothesis may explain the binding ability of integral membrane proteins to ERM proteins. However, the ERM-binding or ERM-colocalization ability of some of the truncated or site-directed mutants cannot be explained by this hypothesis. For example, although the isoelectric point of the cytoplasmic domain of E-43/1–64 was calculated as 10.73, this mutant does not bind to moesin (Fig. 3 b). The cytoplasmic domain of E-ICAM-2/RRR:GGA also has a net positive charge, but does not colocalize with moesin (Fig. 9). These findings suggest that the three-dimensional structure in or around the juxta-membrane positively charged amino acid clusters is also important for the specificity of ERM–protein–integral membrane protein interaction.

Physiological Relevance of the Occurrence of Multiple Membrane Binding Partners for ERM Proteins

At present, it is not clear how many types of integral membrane proteins function as binding partners for ERM proteins in situ, but judging from the expression and distribution of CD44, CD43, and ICAM-2 in tissues, the occurrence of many other ERM-binding partners can be expected. In cells within tissues, as yet undetermined regulatory mechanisms may determine the combination of ERM proteins and their membrane-binding partners, resulting in the specific expression and distribution of ERM proteins in a cell type-specific manner (Franck et al., 1993; Ami et al., 1994).

We found previously that at physiological ionic strength the association between CD44 and ERM proteins requires PIP2 in vitro, and that it is regulated by the Rho signaling pathway in vivo, which is thought to generally regulate actin-based cytoskeletal organization (Hirao et al., 1996). However, in marked contrast to CD44, even at physiological ionic strength, CD43 and ICAM-2 bound to ERM proteins in vitro with a relatively high affinity in the absence of PIP2. It is not clear at present whether the binding of ERM proteins to CD43 and ICAM-2 (and to as yet unidentified ERM membrane-binding partners) in vivo is also regulated by the Rho signaling pathway. Although this study was focused on the identification of membrane-binding partners other than CD44 and on their ERM protein-binding sites (positively charged amino acid clusters), the data obtained here would also provide some clues to understand the regulatory mechanism of ERM–membrane interaction in general. For example, the carboxyl terminus–truncated CD44 binds to ERM proteins in vitro even in the absence of PIP2 at physiological ionic strength. Studies are currently underway in our laboratory to clarify the regulatory mechanism of ERM–membrane interaction.

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