**Toxocara Seroprevalence and Risk Factor Analysis in Four Communities of the Wiwa, an Indigenous Tribe in Colombia**

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**Abstract:** The life of the indigenous Wiwa tribe in northeast Colombia is characterized by lacking access to clean drinking water and sanitary installations. Furthermore, free-roaming domestic animals and use of yucca and/or manioc as a primary food source favor the transmission of soil-transmitted helminths, e.g., *Toxocara canis* and *Toxocara cati*, the roundworms of dogs and cats. Infection may result in the clinical picture of toxocarosis, one of the most common zoonotic helminthoses worldwide. To estimate the toxocara seroprevalence in four different villages of the Wiwa community, serum samples from 483 inhabitants were analyzed for anti-Toxocara-antibodies. Overall, 79.3% (383/483) of analyzed samples were seropositive. Statistically significant differences were observed between the four villages, as well as age groups (adults > adolescents > children), while sex had no effect. The high seropositivity rate demonstrates the risk of zoonotic roundworm infections and potential clinical disease in vulnerable indigenous inhabitants.

**Keywords:** toxocarosis; toxocariosias; soil-transmitted helminths; zoonosis; roundworms; IgG; ELISA

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1. Introduction

The Sierra Nevada de Santa Marta in the northeast of Colombia presents a great variety of climates and tropical ecosystems within a relatively small area. The ethnohistorical habitation of the region dates back at least to the 1st century AD with the pre-Columbian culture of the Tairona, of which the Kankuamo, Kogi, and Arhuacos, as well as the Wiwa, are the believed direct descendants [1]. The indigenous tribe of the Wiwa have suffered constant violations of their human rights and have historically been victims of, for example the Spanish colonial system, or illegal armed actors for the illicit cultivation of coca, poppy, and marijuana, but also of legal investors with great interest in the indigenous territory [2]. To avoid increasing confrontations, parts of the population moved to higher areas of the Sierra Nevada de Santa Marta, where the Wiwas live a secluded life, with a deep-rooted cultural and spiritual identity with the nature and wildlife surrounding them. Despite growing modernization and tourism in the territory, the Wiwa community strive to keep their ancient culture alive [2]. They live in small communities comprising 200–700 inhabitants per village, in which domestic animals have unrestricted access, even to the people’s living areas in simple clay huts. This, together with the use of natural resources such as yucca and/or manioc and home-grown vegetables as primary food sources, favors the transmission of soil transmitted helminths (STHs) such as *Toxocara canis* and *Toxocara cati*, the zoonotic roundworms of dogs and cats. Furthermore, a lack of access to clean water and sanitary installations affects environmental hygiene as well as personal hygienic standards, providing ideal transmission possibilities for STHs [3,4]. Consequently, high infection rates with *Ancylostoma* spp., *Necator americanus*, *Strongyloides*...
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stercoralis, Ascaris lumbricoides, Trichuris trichiura, and Hymenolepis nana were detected in members of the Wiwa community [5].

Additionally, environmental conditions, such as annual mean temperatures of 25–30 °C, high humidity, and soil moisture, are favourable for the development and tenacity of a broad range of (zoonotic) helminth eggs, such as Toxocara spp. [6–8]. Consequently, a particularly high seroprevalence of anti-Toxocara antibodies in humans is reported for some tropical and subtropical regions or countries [9]. Toxocarosis is one of the most common zoonotic helminthoses worldwide [10], and human infections are primarily acquired after accidental ingestion of embryonated eggs containing infective Toxocara spp. third-stage larvae (L3) in contaminated food, water, and soil, or via consumption of L3 in undercooked meat of paratenic hosts such as chicken [11]. By excreting eggs into the human environment, domestic dogs and cats play a pivotal role in the epidemiology of human toxocarosis [12].

Infections may remain asymptomatic; however, the migration and persistence of larvae in the tissues can also result in clinical disease, ranging from mild to severe, and rarely fatal organ injuries [13]. Overall, the disease is classified into four different syndromes. The visceral larva migrans (VLM) is a systemic disease caused by larval migration through major organs, while the covert toxocarosis represents a less severe form of VLM [14]. Larval migration into the eye or the optic nerve manifests in the ocular larva migrans (OLM), while neurotoxocarosis (NT) results from invasion of Toxocara larvae into the brain [15,16]. OLM and NT in particular may have severe consequences such as impaired vision and even blindness, or neurological deficits and neuropsychological disorders, exhibiting a lifelong burden, particularly in children [17,18]. The mostly rather unspecific symptoms, along with a lack of knowledge of the disease, are probably the main reasons for underestimating and underdiagnosing toxocarosis, which has been targeted by the US American Centers for Disease Control and Prevention (CDC) as one of the five neglected parasitic infections with priority for public health action [19]. Thus far, to our knowledge, nothing is known about the inhabitants’ health status with regard to Toxocara spp. As clinically apparent toxocarosis and its consequences affect the life of the individuals, the presented study aimed to estimate the seroprevalence in the indigenous tribe of the Wiwa, and to analyse potentially related risk factors. The results are necessary to gain knowledge about the risk of exposure, and to raise awareness for this neglected disease in indigenous communities.

2. Materials and Methods
2.1. Ethical Statement

Sample material was obtained in accordance with the international guidelines of the Declaration of Helsinki, approved by the local ethics committee of Valledupar, Cesar, Colombia (Acta no. 0022013). Written informed consent was obtained from each participant or the parent/legal guardian of a child prior to participation.

2.2. Sampling

In July and November 2014, blood samples of 483 indigenous volunteers from the Wiwa villages of Tezhumake (n = 173), Ashintukwa (n = 106), Cherua (n = 94), and Siminke (n = 110) were collected. Participants were categorized by their age (children: <6 years, n = 75; adolescents: 6–17 years, n = 217; adults: >17 years, n = 191) and sex (males: n = 247; females: n = 232). Furthermore, the villages were categorized regarding altitude, being categorized as below 1000 m above sea level (MASL) (Tezhumake: 700 MASL; Ashintukwa: 750 MASL; n = 279 inhabitants) and above 1000 MASL (Cherua: 1300 MASL; Siminke: 1100 MASL; n = 204 inhabitants). Collected blood samples were cooled in a specific cooling box at 4 °C and transported immediately to the laboratory for serum extraction. Sera were stored thereafter at −20 °C and transferred to Germany with a permanent cooling chain. After arrival, sera were stored at −20 °C until further analysis.
2.3. ELISA

To determine anti-Toxocara antibodies in the serum samples, the Toxocara canis IgG ELISA (product code: DE0450; Demeditec Diagnostics GmbH, Kiel, Germany), based on Toxocara excretory-secretory products (TES) as diagnostic antigen, was conducted in accordance with the manufacturer’s instructions. The described diagnostic specificity and sensitivity is given as 98.63% (95%-CI (96.52–99.62%)) and 96.92% (95%-CI (89.32–99.63%)). All runs included a positive and negative control, as well as a calibrator for semi-quantitative analysis. Briefly, all samples were diluted 1:101 with supplied IgG sample diluent, and a volume of 100 µl standards/controls as well as diluted sera were incubated as duplicates for 1 h at 37 °C. After incubation and washing steps consisting of three times 300 µl washing buffer, 100 µl horseradish peroxidase-labelled conjugate was added and again incubated for 30 min at 37 °C. After washing as described, colorimetric detection was initiated by applying 100 µl tetramethylbenzidine substrate solution for 30 min at room temperature and stopped with 100 µl stop solution. The optical density (OD) was measured using a Biowave 340 photometer (BioTek Instruments, Inc., Winooski, VT, USA) at a wavelength of 450 nm as well as a reference wavelength of 620 nm to exclude background signals. Result in units (U) were calculated, and samples were tested negative if U was <9, equivocal if U was ≥9 to <11, or positive if U was ≥11.

2.4. Statistical Analyses

Statistical analyses were conducted in R version 4.0.2 [20]. To determine Toxocara seroprevalence differences in the Wiwa, a general linear model (GLM) with binomial error structure and logit link function was conducted. Village, age group, and sex were included as fixed factors, and multiple comparisons between these factors were carried out using Tukey HSD contrasts with single-step p-value adjustment (package “multcomp” [21]). Final models were compared to null models containing only an intercept term (GLM) in a likelihood ratio test. As comparison of meters above sea level (MASL) could not be fitted in the multiple comparison analyses, villages were categorized according to their location in low altitude below 1000 MASL and high altitude above 1000 MASL, and comparison was repeated with MASL instead of village as a factor.

3. Results

3.1. Toxocara Seroprevalence in the Wiwa

Anti-Toxocara antibodies were detected in 79.3% of the 483 analyzed samples, and in 3.3%, the results were equivocal. In case of equivocal results, the manufacturer recommends repetition with freshly obtained serum after 2 to 4 weeks. As repetition of equivocal results was not possible, these samples were judged as negative to fit in the GLM.

The highest seroprevalence was detected in the village of Siminke (90.0%), the lowest in Tezhumake (70.5%, Table 1). Subgroup analyses showed a seroprevalence of 66.7% and 76.0% in children and adolescents compared to 88.0% in adults. Regarding sex, 75.7% of participating males and 83.2% of females tested positive. Furthermore, inhabitants of villages below 1000 MASL (Tezhumake and Ashintukwa) exhibited a lower seroprevalence than those living above 1000 MASL (Siminke and Cherua).

3.2. General Linear Modelling

Toxocara seroprevalences differed significantly between the communities. Overall, the seroprevalence in villages below 1000 MASL (Tezhumake and Ashintukwa) was statistically significantly lower compared to villages above 1000 MASL. Statistically significant differences were also determined between the age classes: the seroprevalence in adults was significantly higher than in adolescents and children, while sex had no effect on seropositivity. Detailed results are presented in Table 2.
Table 1. *Toxocara* seroprevalences including 95% confidence intervals (CI) in the Wiwa community regarding different risk factors.

| Group      | Sample Size | Seropositive | Seroprevalence | 95% CI         |
|------------|-------------|--------------|----------------|---------------|
| Total      | 483         | 383          | 79.3%          | 75.5–82.7%    |
| Villages   |             |              |                |               |
| Tezhumake  | 173         | 122          | 70.5%          | 63.3–76.8%    |
| Ashintukwa | 106         | 79           | 74.5%          | 65.4–75.4%    |
| Cherua     | 94          | 83           | 88.3%          | 80.3–93.3%    |
| Siminke    | 110         | 99           | 90.0%          | 83.0–94.3%    |
| Age        |             |              |                |               |
| children   | 75          | 50           | 66.7%          | 55.4–76.3%    |
| adolescents| 217         | 165          | 76.0%          | 77.9–87.5%    |
| adults     | 191         | 168          | 88.0%          | 82.6–91.8%    |
| Sex        |             |              |                |               |
| males      | 247         | 187          | 75.7%          | 70.0–80.6%    |
| females    | 232         | 193          | 83.2%          | 77.9–87.5%    |
| MASL       |             |              |                |               |
| below 1000 | 279         | 201          | 72.0%          | 66.5–77.0%    |
| above 1000 | 204         | 182          | 89.2%          | 84.2–92.8%    |

Abbreviation: MASL, meters above sea level.

Table 2. Results from the general linear model (GLM) testing the effect of different variables on the *Toxocara* seroprevalence in the North Colombian indigenous tribe of the Wiwa (n = 483). The final model was significantly different from a null model containing only an intercept term (likelihood ratio test, df = 6, \( \chi^2 = 58.643, p < 0.001 \)). Significant \( p \)-values are printed in bold.

| Variable                           | Estimate | Standard Error | z-Value | \( p \)-Value |
|------------------------------------|----------|----------------|---------|---------------|
| Intercept                          | 2.587    | 0.3921         | 6.598   | <0.001        |
| Village                            |          |                |         |               |
| Ashintukwa vs. Cherua              | 0.903    | 0.441          | 2.049   | 0.165         |
| Ashintukwa vs. Siminke             | 1.398    | 0.457          | 3.061   | 0.012         |
| Ashintukwa vs. Tezhumake           | -0.500   | 0.317          | -1.576  | 0.384         |
| Cherua vs. Siminke                 | 0.495    | 0.523          | 0.946   | 0.775         |
| Cherua vs. Tezhumake               | -1.403   | 0.410          | -3.423  | 0.003         |
| Siminke vs. Tezhumake              | -1.898   | 0.433          | -4.388  | <0.001        |
| Age                                |          |                |         |               |
| children vs. adolescents           | -0.852   | 0.326          | -2.617  | 0.024         |
| children vs. adults                | -2.099   | 0.400          | -5.248  | <0.001        |
| adolescents vs. adults             | -1.246   | 0.333          | -3.748  | <0.001        |
| Sex                                |          |                |         |               |
| male vs. female                    | -0.325   | 0.261          | -1.246  | 0.213         |
| MASL                               |          |                |         |               |
| below vs. above 1000 MASL          | -1.458   | 0.306          | -4.773  | <0.001        |

Multiple comparisons between the levels of the factors “village”, “age”, and “sex” were calculated using Tukey contrasts with single-step \( p \)-value adjustment. The comparison of MASL was calculated separately (null model containing only an intercept term with likelihood ratio test, df = 4, \( \chi^2 = 55.219, p < 0.001 \)). Abbreviation: MASL, meters above sea level.

4. Discussion

The worldwide distribution of *Toxocara* spp., their high reproduction rate, and the high tenacity of the eggs result globally in soil contamination of human environments [22,23], making toxocarosis one of the most common zoonotic helminthoses worldwide [10]. As tropical and subtropical conditions with year-round temperatures of >18 °C, high precipitation, and consequently high humidity favor the survival of *Toxocara* eggs, soil contamination rates are elevated at lower latitudes (0–20°) compared to higher ones (41–60°) [22]. Consequently, southeast Asia and South America are salient areas of *Toxocara* prevalence with,
estimated on the population size, high burdens of human toxocarosis [24]. Colombia is, with 47.5%, regarded as the country with the highest human seroprevalence in Latin America, even though only one municipality of Bogota was considered in the conducted meta-analysis [24]. However, the result is strengthened by Mendoza Meza [25], as 42.1% of examined schoolchildren in the city of Santa Marta at the Caribbean coast of north Colombia exhibited anti-Toxocara antibodies. Notably, the seroprevalence of 79.3% in the northeast Colombian indigenous Wiwa tribe investigated in the present study is remarkably higher than previously described for Colombia.

Besides tropical climate conditions, various further potential risk factors have been associated with human Toxocara infections, such as living in rural areas, having contact with dogs, cats, or soil, and the drinking of untreated water [24]. As the Wiwas live retracted in the mid-highlands of the Sierra Nevada de Santa Marta, their life is defined by a lack of access to clean drinking water (rivers and unprotected wells are used as a water source) or sanitary installations [1]. Their lifestyle, together with different cultural aspects such as top dressing, affects the environmental hygiene and hygienic standards of the residents [3,4]. Furthermore, companion as well as stray dogs and cats have unrestricted access to the villages and living areas, and represent a continuous source of environmental egg contamination [26]. Such high soil contamination may lead to contamination of vegetables grown close to the ground. Accordingly, Toxocara egg contamination of different vegetables such as lettuce, carrots, potatoes, zucchini, spinach, or cress have been described [27–29]. The Wiwa use yucca and manioc in combination with home-grown vegetables as a primary food source, and inadequately washed foods or foods washed with contaminated water can be a source for Toxocara infections.

The seroprevalence in the Wiwa population varied between the different villages. The seroprevalences of 88% and 90% in inhabitants of Cherua and Siminke were significantly higher compared to those of Tezhumake and Ashintukwa (70% and 75%). Although all communities have difficult living conditions, regional climatic conditions differ between the four villages. The daily average temperature in all four communities is above 18 °C, but Tezhumake and Ashintukwa are located at 700 and 750 MASL, and the climate at this altitude is characterized by a prolonged dry season of up to six months a year. In contrast, Cherua and Siminke are located at 1300 and 1100 MASL, where higher precipitation and higher average humidity and a subtropical or premontane humid forest result in an increased soil moisture. This favors the development and survival of embryonated Toxocara eggs, which may survive for up to four years in warm and humid environments [30–32]. Thus, climatic and associated abiotic factors may explain the observed seroprevalence differences between the villages.

Furthermore, Toxocara seropositivity in the Wiwas was associated with increasing age. However, age as a risk factor for human seropositivity remains debatable. Although children are regarded as being at high infection risk due to a less developed hygienic awareness compared to adults [9,33], elevated levels of anti-Toxocara antibodies in elderly people may result from a cumulative effect of Toxocara exposure during their lifespan. The constant rechallenge of the immune response due to less developed hygienic standards, leading to frequent and/or high Toxocara exposure of all inhabitants regardless of age, may result in the accumulation of detectable antibodies. On average, IgG antibodies against Toxocara can be detected for 2.7 years with TES-ELISA [34] as used in the present study, and in many patients requiring chemotherapeutic treatment, IgG persistence lasts over four years [34–36]. Nevertheless, about 67% seroprevalence in the Wiwa children is alarming, as toxocarosis can exhibit detrimental long-term consequences. Especially in children, neuroinvasion of Toxocara larvae has been associated with mental confusion and cognitive impairment, as well as deficits in the development of speech and poor reading achievements, a higher distractibility, and lower intelligence in kindergarten children [17,18,37,38].

Similar to age as an influencing factor, the impact of sex is controversially discussed. The majority of studies have identified males as being at higher risk for Toxocara expo-
sure [24,39,40], but females are at risk, too [41]. For males, the increased infection risk has been explained by a higher exposure to *Toxocara* eggs due to increased agricultural activities, and thus increased contact with *Toxocara* egg-contaminated soil and stray animals [42,43].

In the indigenous Wiwas, the seroprevalence did not differ statistically significant between sexes, as all inhabitants are presumed to be frequently exposed to *Toxocara* eggs.

Overall, the high seroprevalence of 79.3% in indigenous Wiwa indicates a high health risk of exposed people [19]. As infections might be accompanied by severe and detrimental symptoms, interventions and preventive measurements are necessary to eliminate the existing infection cycles. Although the installation of sanitary facilities remains difficult in these secluded villages, improved access to clean drinking water, and education in and implementation of adequate hygiene measures, such as washing of hands after soil contact or thorough washing of vegetables, are essential to prevent transmission [44,45]. Furthermore, the substantial reduction in infective *Toxocara* eggs in community environments would be an important measure. This comprises the avoidance of pet defecation in the villages and farmland and/or disposal of feces in these places [46,47]. However, avoiding defecation in crucial areas seems unrealistic, as animals are free-roaming and not housed and walked. Similarly, appropriate removal of feces, which is an effective measure to reduce environmental contamination, is not feasible to implement, as the contact with animal excrements is regarded as impure in the indigenous culture of the Wiwa. Frequent deworming of dogs and cats could be another approach to reduce environmental contamination, but this requires the provision of anthelmintics in sufficient quantities by governments or aid agencies.

5. Conclusions

The secluded life of the indigenous tribe of the Wiwas in north Colombia is characterized by, for example, a lack of access to clean drinking water and adequate hygiene standards. These well-known risk factors for zoonotic *Toxocara* infections are potentiated by nutrition habits and free-roaming dogs and cats in the Wiwa communities, resulting in a high seroprevalence of about 80% in the tested inhabitants. Deviating climatic conditions in the included villages are most likely responsible for the higher seroprevalences in communities above 1000 MASL with a subtropical or premontane humid forest compared to the lower altitudes with up to six months of dry season. Nevertheless, frequent *Toxocara* exposure is a common feature in all Wiwa communities, reflected by a cumulative effect with highest seroprevalences in adult inhabitants, and lacking sex differences. As toxocarosis can result in detrimental effects and long-term consequences, raising awareness thereof, and intervention measures, such as improvement in hygienic standards and reduction in soil contamination, are urgently needed in indigenous communities such as the Wiwa tribe.

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References

1. Ministerio Del Interior República de Colombia. Diagnóstico y Líneas de Acción para las Comunidades Wiwa de la Sierra Nevada de Santa Marta (Departamentos Cesar, Magdalena y Guajira) en el Marvo del Cumplimiento del Auto 004 de 2009. Available online: https://siic.mininterior.gov.co/sites/default/files/pueblo_wiwa_-_diagnostico_comunitario_0.pdf (accessed on 16 August 2021).

2. Ministerio de Cultura República de Colombia. Wiwa, la Gente que da Origen al Calor. Available online: https://www.mincultura.gov.co/prensa/noticias/Documents/Poblaciones/PUEBLO%20WIWA.pdf (accessed on 16 August 2021).

3. McMichael, A.J. The urban environment and health in a world of increasing globalization: Issues for developing countries. Bull. World Health Organ. 2000, 78, 1117–1126.

4. Montgomery, M.A.; Elimelech, M. Water and sanitation in developing countries: Including health in the equation. Environ. Sci. Technol. 2007, 41, 17–24. [CrossRef]

5. Kann, S.; Bruennert, D.; Hansen, J.; Mendoza, G.A.C.; Gonzalez, J.J.C.; Hanke, M.; Hagen, R.M.; Backhaus, J.; Frickmann, H. High Prevalence of Intestinal Pathogens in Indigenous in Colombia. J. Clin. Med. 2020, 9, 2786. [CrossRef] [PubMed]

6. Azam, D.; Ukpai, O.M.; Said, A.; Abd-Allah, G.A.; Morgan, E.R. Temperature and the development and survival of infective Toxocara canis larvae. Parasitol. Res. 2012, 110, 649–656. [CrossRef] [PubMed]

7. Gamboa, M.I. Effects of temperature and humidity on the development of eggs of Toxocara canis under laboratory conditions. J. Helminthol. 2005, 79, 327–331. [CrossRef]

8. Keegan, J.D.; Holland, C.V. A comparison of Toxocara canis embryonation under controlled conditions in soil and hair. J. Helminthol. 2013, 87, 78–84. [CrossRef] [PubMed]

9. Ma, G.; Holland, C.V.; Wang, T.; Hofmann, A.; Fan, C.K.; Maizels, R.M.; Hotez, P.J.; Gasser, R.B. Human toxocariasis. Lancet Infect. Dis. 2018, 18, e14–e24. [CrossRef]

10. Rubinsky-Elefant, G.; Hirata, C.E.; Yamamoto, J.C.; Quintero, C.L.A.; Hanke, M.; Hagen, R.M.; Backhaus, J.; Frickmann, H. High Prevalence of Intestinal Pathogens in Indigenous in Colombia. J. Clin. Med. 2020, 9, 2786. [CrossRef] [PubMed]

11. Morimatsu, Y.; Akao, N.; Akiyoshi, H.; Kawazu, T.; Okabe, Y.; Aizawa, H. A familial case of visceral larva migrans after ingestion of raw chicken livers: Appearance of specific antibody in bronchoalveolar lavage fluid of the patients. Am. J. Trop. Med. Hyg. 2006, 75, 303–306. [CrossRef]

12. Overgaauw, P.A.; van Knappen, F. Veterinary and public health aspects of Toxocara spp. Vet. Parasitol. 2013, 193, 398–403. [CrossRef] [PubMed]

13. Gillespie, S.H. Human toxocariasis. J. Appl. Bacteriol. 1987, 63, 473–479. [CrossRef]

14. Taylor, M.R.; Keane, C.T.; O’Connor, P.; Mulvihill, E.; Holland, C. The expanded spectrum of Toxocara disease. Lancet 1988, 1, 692–695. [CrossRef]

15. Fan, C.K.; Holland, C.V.; Loxton, K.; Barghouth, U. Cerebral Toxocariasis: Silent Progression to Neurodegenerative Disorders? Clin. Microbiol. Rev. 2015, 28, 663–686. [CrossRef] [PubMed]

16. Finsterer, J.; Auer, H. Neurotoxocarcisis. Rev. Inst. Med. Trop. Sao Paulo 2007, 49, 279–287. [CrossRef]

17. Jarosz, W.; Mizgajska-Wiktor, H.; Kirwan, P.; Konarski, J.; Rychlicki, W.; Wawrzyniak, G. Developmental age, physical fitness and Toxocara seroprevalence amongst lower-secondary students living in rural areas contaminated with Toxocara eggs. Parasitology 2010, 137, 53–63. [CrossRef]

18. Walsh, M.G.; Haseeb, M.A. Reduced cognitive function in children with toxocariasis in a nationally representative sample of the United States. Int. J. Parasitol. 2012, 42, 1159–1163. [CrossRef]

19. Woodhall, D.M.; Eberhard, M.L.; Parise, M.E. Neglected parasitic infections in the United States: Toxocariasis. Am. J. Trop. Med. Hyg. 2014, 90, 810–813. [CrossRef] [PubMed]

20. R Core Team, R. A Language and Environment for Statistical Computing; R Foundation for Statistical Computing: Vienna, Austria, 2020.

21. Hothorn, T.; Bretz, F.; Westfall, P. Simultaneous inference in general parametric models. Biom. J. 2008, 50, 346–363. [CrossRef]

22. Fakhri, Y.; Gasser, R.B.; Rostami, A.; Fan, C.K.; Ghasemi, S.M.; Javanian, M.; Bayani, M.; Armoon, B.; Moradi, B. Toxocara eggs in public places worldwide—a systematic review and meta-analysis. Environ. Pollut. 2018, 242, 1467–1475. [CrossRef]

23. Traversa, D.; Frangipane di Regalbuto, A.; Di Cesare, A.; La Torre, F.; Drake, J.; Pietrobelli, M. Environmental contamination by canine geohelminths. Parasit. Vectors 2014, 7, 67. [CrossRef]

24. Rostami, A.; Rahi, S.M.; Holland, C.V.; Taghipour, A.; Khalili-Fomeshi, M.; Fakhri, Y.; Omrani, V.F.; Hotez, P.J.; Gasser, R.B. Seroprevalence estimates for toxocariasis in people worldwide: A systematic review and meta-analysis. PLoS Negl. Trop. Dis. 2019, 13, e0007809. [CrossRef]

25. Mendoza Meza, D.L. Exposición al parasito Toxocara canis en una población escolar de la comuna 7 del distrito de Santa Marta, Colombia. Duazary 2013, 7, 183–190. [CrossRef]

26. Morgan, E.R.; Azam, D.; Fegler, K. Quantifying sources of environmental contamination with Toxocara spp. eggs. Vet. Parasitol. 2013, 4, 390–397. [CrossRef]
27. Abougrain, A.K.; Nahaisi, M.H.; Madi, N.S.; Saied, M.M.; Ghenghesh, K.S. Parasitological contamination in salad vegetables in Tripoli-Libya. Food Control 2010, 21, 760–762. [CrossRef]

28. Fallah, A.A.; Pirali-Kheirabadi, K.; Shirvani, F.; Saei-Dehkordi, S.S. Prevalence of parasitic contamination in vegetables used for raw consumption in Shahrekord, Iran: Influence of season and washing procedure. Food Control 2012, 25, 617–620. [CrossRef]

29. Klapec, T.; Borecka, A. Contamination of vegetables, fruits and soil with geohelminths eggs on organic farms in Poland. Ann. Agric. Environ. Med. 2012, 19, 421–425. [PubMed]

30. Deutz, A.; Fuchs, K.; Auer, H.; Kerbl, U.; Aspock, H.; Kofer, J. Toxocara-infestations in Austria: A study on the risk of infection of farmers, slaughterhouse staff, hunters and veterinarians. Parasitol. Res. 2005, 97, 390–394. [CrossRef] [PubMed]

31. Etewa, S.E.; Abdel-Rahman, S.A.; Abd El-Aal, N.F.; Fathy, G.M.; El-Shafey, M.A.; Ewis, A.M. Geohelminths distribution as affected by soil properties, physicochemical factors and climate in Sharky governorate Egypt. J. Parasit. Dis. 2016, 40, 496–504. [CrossRef] [PubMed]

32. Rocha, S.; Pinto, R.M.; Floriano, A.P.; Teixeira, L.H.; Bassili, B.; Martinez, A.; Costa, S.O.; Caseiro, M.M. Environmental analyses of the parasitic profile found in the sandy soil from the Santos municipality beaches, SP, Brazil. Rev. Inst. Med. Trop. Sao Paulo 2011, 53, 277–281. [CrossRef]

33. Hotez, P.J.; Wilkins, P.P. Toxocariasis: America’s most common neglected infection of poverty and a helminthiasis of global importance? PLoS Negl. Trop. Dis. 2009, 3, e400. [CrossRef] [PubMed]

34. Fillaux, J.; Magnaval, J.F. Laboratory diagnosis of human toxocariasis. Vet. Parasitol. 2013, 193, 327–336. [CrossRef]

35. Cypess, R.H. Visceral larva migrans. Cornell Vet. 1978, 68, 283–296.

36. Elefant, G.R.; Shimizu, S.H.; Sanchez, M.C.; Jacob, C.M.; Ferreira, A.W. A serological follow-up of toxocariasis patients after chemotherapy based on the detection of IgG, IgA, and IgE antibodies by enzyme-linked immunosorbent assay. J. Clin. Lab. Anal. 2006, 20, 164–172. [CrossRef]

37. Richartz, E.; Buchkremer, G. Cerebral toxocariasis: A rare cause of cognitive disorders. A contribution to differential dementia diagnosis. Der Nervenarzt 2002, 73, 458–462. [CrossRef]

38. Salvador, S.; Ribeiro, R.; Winckler, M.I.; Ohlweiler, L.; Riesgo, R. Pediatric neurotoxocariasis with concomitant cerebral, cerebellar, and peripheral nervous system involvement: Case report and review of the literature. J. Pediatr. 2010, 86, 531–534. [CrossRef]

39. Holland, C.V.; O’Lorcain, P.; Taylor, M.R.; Kelly, A. Sero-epidemiology of toxocariasis in school children. Parasitology 1995, 110 Pt 5, 533–545. [CrossRef]

40. Mughini-Gras, L.; Harms, M.; van Pelt, W.; Pinelli, E.; Kortbeek, T. Seroepidemiology of human Toxocara and Ascaris infections in the Netherlands. Parasitol. Res. 2016, 115, 3779–3794. [CrossRef] [PubMed]

41. Havasiova, K.; Dubinsky, P.; Stefanikova, A. A seroepidemiological study of human Toxocara infection in school children in the Netherlands. Parasitol. Res. 2016, 115, 3779–3794. [CrossRef] [PubMed]

42. Won, K.Y.; Kruszon-Moran, D.; Schantz, P.M.; Jones, J.L. National seroprevalence and risk factors for zoonotic Toxocara spp. infection. Am. J. Trop. Med. Hyg. 2008, 79, 552–557. [CrossRef] [PubMed]

43. Rubinsky-Elefant, G.; da Silva-Nunes, M.; Malafronte, R.S.; Muniz, P.T.; Ferreira, M.U. Human toxocariasis in rural Brazilian Amazonia: Seroprevalence, risk factors, and spatial distribution. Am. J. Trop. Med. Hyg. 2008, 79, 93–98. [CrossRef] [PubMed]

44. Fan, C.K.; Liao, C.W.; Cheng, Y.C. Factors affecting disease manifestation of toxocariasis in humans: Genetics and environment. Vet. Parasitol. 2013, 193, 342–352. [CrossRef] [PubMed]

45. Moreira, G.M.; Telmo Pde, L.; Mendonca, M.; Moreira, A.N.; McBride, A.J.; Scaini, C.J.; Conceicao, F.R. Human toxocariasis: Current advances in diagnostics, treatments, and interventions. Trends Parasitol. 2014, 30, 456–464. [CrossRef] [PubMed]

46. Woodhall, D.M.; Fiore, A.E. Toxocariasis: A Review for Pediatricians. J. Pediatr. Infect. Dis. Soc. 2014, 3, 154–159. [CrossRef] [PubMed]

47. Nijssse, R.; Mughini-Gras, L.; Wagenaar, J.A.; Franssen, F.; Ploeger, H.W. Environmental contamination with Toxocara eggs: A quantitative approach to estimate the relative contributions of dogs, cats and foxes, and to assess the efficacy of advised interventions in dogs. Parasit. Vectors 2015, 8, 397. [CrossRef] [PubMed]