Research article

**Resistance of Leishmania (Leishmania) amazonensis and Leishmania (Viannia) braziliensis to nitric oxide correlates with disease severity in Tegumentary Leishmaniasis**

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**Abstract**

**Background:** Nitric oxide (NO•) plays a pivotal role as a leishmanicidal agent in mouse macrophages. NO• resistant *Escherichia coli* and *Mycobacterium tuberculosis* have been associated with a severe outcome of these diseases.

**Methods:** In this study we evaluated the *in vitro* toxicity of nitric oxide for the promastigote stages of *Leishmania (Viannia) braziliensis* and *Leishmania (Leishmania) amazonensis* parasites, and the infectivity of the amastigote stage for human macrophages. Parasites were isolated from patients with cutaneous, mucosal or disseminated leishmaniasis, and NO• resistance was correlated with clinical presentation.

**Results:** Seventeen isolates of *L. (L.) amazonensis* or *L. (V.) braziliensis* promastigotes were killed by up to 8 mM of more of NaNO₂ (pH 5.0) and therefore were defined as nitric oxide-susceptible. In contrast, eleven isolates that survived exposure to 16 mM NaNO₂ were defined as nitric oxide-resistant. Patients infected with nitric oxide-resistant *Leishmania* had significantly larger lesions than patients infected with nitric oxide-susceptible isolates. Furthermore, nitric oxide-resistant *L. (L.) amazonensis* and *L. (V.) braziliensis* multiplied significantly better in human macrophages than nitric oxide-susceptible isolates.

**Conclusion:** These data suggest that nitric oxide-resistance of *Leishmania* isolates confers a survival benefit for the parasites inside the macrophage, and possibly exacerbates the clinical course of human leishmaniasis.
Background
Leishmaniasis is a parasitic disease caused by the *Leishmania* spp. protozoa, transmitted to the skin of a mammalian host during the bite of an infected female sand fly vector. Infections range in severity from asymptomatic to disfiguring forms of tegumentary and potentially fatal visceral leishmaniasis [1,2]. American tegumentary leishmaniasis (ATL) presents a spectrum of clinical manifestations characterized by cutaneous (CL), mucosal (ML), disseminated (DL) and diffuse cutaneous leishmaniasis (DCL). The major *Leishmania* species that cause ATL in the New World are *L. (V.) braziliensis*, *L. (V.) guyanensis*, *L. (L.) amazonensis* and *L. (L.) mexicana*. The typical clinical manifestation of American CL is a single ulcerated lesion with elevated borders, frequently located on the inferior limbs [3]. Mucosal leishmaniasis (ML) is a destructive disease that predominantly affects the nasopharyngeal mucosa. The disease is most common in areas of *L. (V.) braziliensis* transmission and usually occurs months or years after cutaneous leishmaniasis [4]. Disseminated leishmaniasis (DL) has been reported almost exclusively in northern and northeastern Brazil. DL is characterized by the appearance of multiple pleomorphic lesions in two or more noncontiguous areas of the body [5].

*Leishmania* is a digenetic protozoan with an extracellular flagellated promastigote form which replicates and matures to the infectious metacyclic form in the gut of the sand fly vector. The promastigotes is transmitted to a mammalian host during the bite of an infected sand fly. Promastigotes undergo facilitated phagocytosis by a macrophage and convert to the obligate intracellular amastigote life stage [6,7]. Amastigotes survive in macrophage phagolysosomes, a hostile environment for many microbes. *Leishmania* spp. must undergo profound biochemical and morphological adaptations to survive successfully and multiply within macrophages [7]. The mechanisms through which the parasite resists killing within the toxic environment of the phagolysosome remain incompletely defined.

Leishmaniasis is controlled through cell-mediated immune defenses [8]. Murine models have illustrated that macrophages produce IL-12, which induces CD4+ T cells and NK cells to release interferon gamma (IFN-γ), polarizing the immune response toward a type 1 (Th1 type) phenotype [9,10]. In murine systems, IFN-γ has been shown to synergize with another macrophage derived cytokine, tumor necrosis factor alpha (TNF-α), activating nitric oxide synthase (iNOS or NOS2) to produce nitric oxide (NO•) with resultant in killing of intracellular parasites [11-13]. NO• is generated from the oxidation of the terminal guanidine nitrogen atoms of the L-arginine by NADPH dependent enzyme nitric oxide synthase (NOS) [14]. In murine models of leishmaniasis, NO•-dependent mechanisms have been shown to be critical for control of *Leishmania* infection[15,16]. The role of NO• in leishmanicidal activity of human macrophages, has been debated [17]. However, recent data suggest that NO• plays a role in the response of human macrophages to intracellular infections, but the nature of this role is not yet clear [18,19].

Putative NO•-mediated leishmanicidal actions in eukaryotic cells include inhibition of mitochondrial respiration, inactivation of peroxidases, increasing susceptibility to oxidant damage, inhibition of glycolysis, mutation of DNA, inhibition of DNA repair and synthesis, S-nitrosylation, ADP-ribosylation, tyrosine nitration of proteins, disruption of Fe-S clusters, zinc fingers or heme groups, and peroxidation of membrane lipids [20,21]. The *Leishmania* spp. possesses unique antioxidant mechanisms and enzymes. Notably, they convert their abundant GSH stores to trypanothione (TSH) and use TSH reductase/oxidase systems for redox cycling [22]. The protozoa express an iron superoxide dismutase (SOD) but not a manganese SOD, and they have peroxidoxins for handling oxidative stress [23-26]. Nonetheless, oxidant resistance in these parasites are inducible [27], and one expects these systems are susceptible to inactivation by oxidant species similar to other eukaryotes.

Resistance to nitric oxide has been described in *E. coli* and *M. tuberculosis*. Resistant isolates have been associated with a more severe outcome of disease than that caused by non-resistant strains [28]. However, natural NO• resistance in *Leishmania* spp. isolates has not previously been described. In the present study, we evaluated the effect of NO• generated from NaNO2 (pH 5.0) on the viability of *L. (V.) braziliensis* and *L. (L.) amazonensis* promastigotes. NO• resistant *Leishmania* amastigotes multiplied significantly better than nitric oxide-susceptible parasites during infection of human macrophages. Furthermore, NO• resistance was directly associated with lesion size, a clinical measure of disease severity.

Methods
Parasites
*L. (L.) amazonensis* and *L. (V.) braziliensis* parasites were obtained by needle aspiration of lesions from patients with CL, DL or ML. Parasites for study were randomly selected from frozen nitrogen *Leishmania* stocks by investigators blinded to the *Leishmania* species or clinical form of leishmaniasis. Parasites were speciated by isoenzyme electrophoresis and monoclonal antibodies as described. This analysis was performed by Departamento de Bioquimica e Biologia Molecular, Instituto Oswaldo Cruz, FIOCRUZ, Rio de Janeiro, Brazil [29].
Isolation and cultivation of L. (V.) braziliensis and L. (L.) amazonensis

Parasite isolates L. (V.) braziliensis (n = 17) and L. (L.) amazonensis (n = 11) were initially cultivated from patient specimens in tubes with biphasic medium (NNN) consisting of rabbit blood agar overlaid with liver infusion tryptose (LIT), supplemented with 10% heat inactivated fetal bovine serum medium (Sigma Chemical Co., St. Louis, MO). Following isolation, parasite isolates were cryopreserved. The time of storage of the selected strains was similar (p > 0.05). The parasites selected for study had not been previously passaged in liquid culture medium before the beginning of the present study. After selection, parasite isolates were expanded in Schneider’s insect medium (Sigma) pH 7.2 supplemented with 10% fetal bovine serum (FBS) and 2% male human urine at 25°C (complete Schneider medium).

Promastigote NO* susceptibility assays

Thymidine incorporation

L. (L.) amazonensis (n = 10) and L. (V.) braziliensis (n = 6) promastigotes in log phase growth were adjusted to 2 × 10^7 parasites/ml in Hanks' balanced solution (HBBS Sigma, pH 5.0). Twenty µl aliquots containing 4 × 10^5 parasites were placed in 96-well U shaped plates containing 180 µl of 0 to 16 mM NaNO2 (freshly prepared NO* donor) in Hanks Balanced Solution, pH 5.0. After 4 hours incubation at 25°C, plates were centrifuged (700 × g for 10 minutes). The viability of the remaining parasites was assessed by incubation for 20 hr in 200 µl of complete Schneider’s medium with 1 µCi of Thymidine (³H-TdR; ICN Immunochemicals, Costa Mesa, CA, USA) to allow them to enter logarithmic growth. Thymidine incorporation was assessed on a β counter.

MTT assay

L. (L.) amazonensis (n = 5) and L. (V.) braziliensis promastigotes (n = 14) in log phase growth were adjusted to 5 × 10^7 parasites/ml in Hanks’ balanced solution (HBBS Sigma, pH 5.0). Twenty µl aliquots were incubated in 180 µl of 0 to 16 mM NaNO2 (freshly prepared NO* donor) in Hanks Balanced Solution, pH 5.0 in 96-well U shaped plates. After 4 hrs incubation at 25°C, plates were centrifuged (700 × g for 10 minutes) and parasites were resuspended with 200 µl of complete Schneider medium. After an additional 20 hrs at 25°C and centrifugation, parasite viability was measured by incubation in 0.5 mg/ml of MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazo- lium bromide] in Hanks solution, pH 7.0 at 25°C for 4 hrs, followed by dilution in an equal volume 0.04 N HCl in isopropanol. Living mitochondria convert MTT to dark blue formazan that is soluble in acid-isopropanol and detectable on a microplate reader at 540 nm. The percentage of viability was calculated from the OD ratio of untreated versus NO*-treated parasites × 100 [27]. For each parasite isolate 3 experiments at least were performed to test for NO* susceptibility. The thymidine incorporation and MTT assays were done with 28 Leishmania isolates of both species. Seven isolates were tested with both methods.

The virulence of Leishmania spp. is highest in stationary phase, or metacyclic organisms. Nonetheless the MTT and [³H]-TdR uptake assays are most sensitive for log phase organisms. We previously reported that these assays of virulence and oxidant sensitivity in log phase correlate with oxidant sensitivity and virulence in stationary phase organisms [27,30]. As such, viability assays were performed using log phase promastigotes, whereas studies of interactions with mammalian cells utilized stationary phase organisms.

Macrophage cultures

Peripheral blood mononuclear cells (PBMCs) were isolated from the peripheral blood of three different healthy human donors. Briefly, heparinized blood was diluted 1:2 with 0.15 M NaCl and overlaid on Ficoll Hypaque (LSM; Organon Teknika corporation, Durham, NC, USA). After centrifugation, mononuclear cells were collected at the plasma – Ficoll interface, washed three times and resuspended in RPMI 1640 with 10% heat inactivated human AB serum, 100 U penicillin/ml and 100 µg streptomycin/ml (complete medium) (GIBCO BRL, Grand Island, NY). Monocytes were separated from 1 × 10^6 PBMCs by adherence to 8 well Lab Tek plates for 2 h at 37°C, 5% CO2, non-adherent cells were removed by washing, and complete medium was added. Adherent monocytes differentiated to macrophages over six days incubation at 37°C in 5% CO2.

Macrophage infection

One NO*-resistant and one NO*-susceptible isolate each of L. (V.) braziliensis and L. (L.) amazonensis (total 4 isolates) were selected for the macrophage infection assays. Three to 4 replicate assays were performed for each isolate. Promastigotes were maintained at 25°C in Schneider’s insect medium (Sigma) pH 7.2 supplemented with 10% fetal bovine serum (FBS) and 2% human male urine at 25°C (complete Schneider’s medium). Promastigotes in stationary- phase of growth were used in all experiments. All experiments were performed in 3 assays for L. (V.) braziliensis and 4 assays for L. (L.) amazonensis using PBMC/macrophages from 3 different healthy volunteers. The same donors were used for the different species so that the results are directly comparable. After Six-day monocyte-derived macrophages were infected with a 10:1 ratio of stationary - phase promastigotes to macrophages for 2 hours at 35°C, 5% CO2. Extracellular parasites were removed by gentle washing and infected macrophages were maintained for up to 96 h. Cells were stained with
Giemsa and the infection levels were enumerated microscopically by counting the infected cells and parasites per 100 macrophages by three independent observers, blinded to the experimental conditions.

**Epidemiological and clinical evaluations**

Clinical characteristics of the patients such as age, lesion size, Montenegro skin reaction, duration of disease and clinical manifestation of leishmaniasis were determined from clinical records after characterizing the NO* susceptibility of isolates. Adequate data were available for fourteen patients. Most patients were identified and diagnosed at the Corte de Pedra Health Post, located in an endemic area for cutaneous leishmaniasis situated in the southeast of the state of Bahia, Brazil. The remainder of patients was referred to the University Hospital Prof. Edgard Santos of the Federal University of Bahia, Brazil. This study was approved by the Ethical Committee of the Hospital Universitário Prof. Edgard Santos and an informed consent was obtained from all participants or their parents or guardians if patients were less than 18 years old.

**Statistical analysis**

Student's t-test was used to compare the age, lesion appearance, and human macrophage infection studies. Lesion size and Montenegro diameter were analyzed by Mann-Whitney nonparametric test. Fischer's Exact test was used to compare NO*-resistant versus NO*-susceptible *L. (L.) amazonensis* and *L. (V.) braziliensis*. An alpha of 5% (p ≤ 0.05) was considered for statistical significance (two tailed). Statistical analysis was performed using GraphPad Prism 3.0 (GraphPad software, San Diego, CA, USA).

**Results**

**Evaluation of Leishmania spp. promastigotes resistance to NO* and correlation with clinical disease**

The susceptibility of *L. (L.) amazonensis* (*n* = 11) and *L. (V.) braziliensis* (*n* = 17) promastigotes to NO* toxicity was evaluated using two measures of parasite viability: first, the rate of [3H]-thymidine incorporation into parasite DNA, and second, a colorimetric measure of mitochondrial activity according to MTT metabolism to formazan.

Evaluation of these records indicated that patients infected with NO-resistant *L. (L.) amazonensis* and *L. (V.) braziliensis* had significantly higher numbers of macrophages infected with NO*– resistant compared to NO*-susceptible parasites (mean ± SD = 315 ± 56 versus 47 ± 10, *p* = 0.008), and higher numbers of macrophages infected with NO*– resistant versus susceptible *L. (V.) braziliensis* (61 ± 4 versus 32 ± 5, *p* = 0.002). This suggests that parasites spread to new cells in the macrophage monolayer in vitro (Figure 2C and 2D). Although we did not evaluate the mechanisms, these data suggest that NO*-resistant amastigotes survive and multiply in resting human macrophages better than susceptible isolates.

**Epidemiological and clinical evaluations**

Clinical records were available for full analysis for 14 isolates from patients with CL. Evaluation of these records indicated that patients infected with NO*-resistant *L. (L.) amaz...
**Discussion**

In the current study we demonstrated for the first time that some isolates of both *L. (V.) braziliensis* and *L. (L.) amazonensis* promastigotes are resistant to killing by nitric oxide. Additionally, we showed that the amastigotes from two resistant isolates survived and multiplied better than susceptible isolates in resting human macrophages in vitro. Macrophages play a pivotal role in *Leishmania* infection. After phagocytosis, *Leishmania* promastigotes enter a parasitophorous vacuole within which the macrophage can provide a safe haven for the parasite to transform into amastigotes and proliferate in a naïve host [6,31]. In an immune host, macrophages can be activated by inflam-
Table 2: Thymidine incorporation assay of resistant L. (L.) amazonensis and L. (V.) braziliensis promastigotes to NO (NaNO2) donor in vitro.

| L (L.) amazonensis | NaNO2 Concentration (mM) |
|---------------------|--------------------------|
|                     | 0 | 0.25 | 0.5 | 1 | 2 | 4 | 8 | 16 |
| 10184               | 1804 | 2205 | 3077 | 2373 | 1284 | 1426 | 1648 | 1435 |
| 436                 | 1683 | 1928 | 2189 | 2544 | 2704 | 3755 | 4366 | 4047 |
| 10432               | 3874 | 4240 | 4285 | 3914 | 3453 | 3312 | 2880 | 2450 |
| 10048               | 826 | 1008 | 1167 | 1150 | 1243 | 1159 | 1114 | 1549 |
| 10076               | 1644 | 2094 | 2870 | 2090 | 1652 | 1860 | 3874 | 3817 |
| 484077              | 788 | 988 | 928 | 961 | 1009 | 1158 | 873 | 1674 |
| 10047               | 576 | 7551 | 5789 | 4416 | 9139 | 7024 | 19181 | 17839 |
| AC                 | 5995 | 10034 | 9731 | 7218 | 6162 | 16305 | 10955 | 11195 |

| L. (V.) braziliensis | NaNO2 Concentration (mM) |
|----------------------|--------------------------|
|                      | 0 | 0.25 | 0.5 | 1 | 2 | 4 | 8 | 16 |
| 13314               | 1626 | 2407 | 1829 | 1923 | 2340 | 3902 | 2950 | 2677 |
| H.R                 | 1958 | 1865 | 2367 | 2549 | 1993 | 2442 | 2909 | 3511 |

Table 3: Thymidine incorporation assay of susceptible L (L.) amazonensis

| L (L.) amazonensis | NaNO2 Concentration (mM) |
|---------------------|--------------------------|
|                     | 0 | 0.25 | 0.5 | 1 | 2 | 4 | 8 | 16 |
| 8653                | 1227 | 1244 | 513 | 262 | 174 | 102 | 81 | 61 |
| 9667                | 1364 | 1595 | 1255 | 1078 | 200 | 65 | 75 | 54 |

| L. (V.) braziliensis | NaNO2 Concentration (mM) |
|----------------------|--------------------------|
|                      | 0 | 0.25 | 0.5 | 1 | 2 | 4 | 8 | 16 |
| 13323               | 2339 | 3217 | 1636 | 215 | 92 | 88 | 76 | 83 |
| Lb-001              | 2213 | 2368 | 1978 | 1319 | 830 | 482 | 268 | 242 |
| 13352               | 1879 | 1227 | 1318 | 870 | 868 | 474 | 326 | 196 |
| 13468               | 1626 | 1796 | 169 | 78 | 73 | 85 | 89 | 97 |

4 × 10⁴ L. (L.) amazonensis and L. (V.) braziliensis promastigotes in HBSS pH 5.0 were exposed to increased NaNO2 concentrations (0.25–16 mM) for 4 hours. The Leishmania were then washed, distributed in parasite growth medium in the presence of 1 µCi/ml of ³H-TdR. After 20 hours of incubation, the incorporation of Thymidine was measured in Beta scintillation counter (CPM). Data are mean ± SD of NO•-resistant (Table 2) or NO•-susceptible (Table 3) isolates of L. (L.) amazonensis and L. (V.) braziliensis.
Table 4: MTT colorimetric assay of resistant *L. (L.) amazonensis* and *L. (V.) braziliensis* promastigotes to NO (NaNO₂) donor in vitro.

| *L. (L.) amazonensis* | NaNO₂ Concentration (mM) |
|------------------------|---------------------------|
|                        | 0  | 0.25 | 0.5 | 1   | 2   | 4   | 8   | 16  |
| 10184                  | 0.558 | 0.678 | 0.729 | 0.693 | 0.726 | 0.989 | 0.824 | 0.518 |
| 436                    | 0.172 | 0.231 | 0.197 | 0.257 | 0.207 | 0.197 | 0.216 | 0.265 |
| 10432                  | 0.098 | 0.081 | 0.072 | 0.158 | 0.131 | 0.153 | 0.154 | 0.112 |

| *L. (V.) braziliensis* | NaNO₂ Concentration (mM) |
|------------------------|---------------------------|
|                        | 0  | 0.25 | 0.5 | 1   | 2   | 4   | 8   | 16  |
| 13314                  | 0.379 | 0.455 | 0.404 | 0.421 | 0.438 | 0.462 | 0.506 | 0.47  |
| 14214                  | 0.242 | 0.248 | 0.285 | 0.261 | 0.268 | 0.294 | 0.295 | 0.279 |

Table 5: MTT colorimetric assay of susceptible *L. (L.) amazonensis*.

| *L. (L.) amazonensis* | NaNO₂ Concentration (mM) |
|------------------------|---------------------------|
|                        | 0  | 0.25 | 0.5 | 1   | 2   | 4   | 8   | 16  |
| 9667                   | 0.488 | 0.682 | 0.463 | 0.373 | 0.335 | 0.25 | 0.027 | 0.003 |
| 9986                   | 0.194 | 0.278 | 0.263 | 0.171 | 0.197 | 0.049 | 0.01  | 0.002 |

| *L. (V.) braziliensis* | NaNO₂ Concentration (mM) |
|------------------------|---------------------------|
|                        | 0  | 0.25 | 0.5 | 1   | 2   | 4   | 8   | 16  |
| 11155                  | 0.389 | 0.478 | 0.488 | 0.432 | 0.113 | 0.025 | 0.007 | 0.004 |
| 13396                  | 0.292 | 0.269 | 0.28  | 0.325 | 0.224 | 0.038 | 0.012 | 0.009 |
| 13223                  | 0.407 | 0.265 | 0.382 | 0.684 | 0.228 | 0.157 | 0.033 | 0.014 |
| Lb 001                | 0.249 | 0.178 | 0.191 | 0.167 | 0.169 | 0.125 | 0.091 | 0.03  |
| 9291                  | 0.628 | 0.645 | 0.437 | 0.253 | 0.101 | 0.033 | 0.051 | 0.02  |
| 13690                  | 0.558 | 0.646 | 0.448 | 0.326 | 0.13  | 0.055 | 0.023 | 0.073 |
| 14183                  | 0.178 | 0.166 | 0.168 | 0.096 | 0.048 | 0.035 | 0.017 | 0.009 |
| 9139                   | 0.446 | 0.277 | 0.221 | 0.305 | 0.182 | 0.093 | 0.047 | 0.012 |
| 13183                  | 0.598 | 0.39  | 0.38  | 0.373 | 0.02  | 0.036 | 0.014 | 0.007 |
| 13548                  | 0.547 | 0.58  | 0.894 | 0.584 | 0.382 | 0.246 | 0.053 | 0.032 |
| 14349                  | 0.812 | 0.556 | 0.426 | 0.404 | 0.208 | 0.108 | 0.085 | 0.086 |
| 14808                  | 0.261 | 0.231 | 0.251 | 0.160 | 0.096 | 0.074 | 0.024 | 0.08  |

\(1 \times 10^6\) *L. (L.) amazonensis* and *L. (V.) braziliensis* promastigotes contained were exposed for 4 hours to increasing concentrations of the NO⁻ donor NaNO₂ (0.25–16 mM) in HBSS pH 5.0. The cells were then washed and distributed in parasite growth medium for 20 hours. After this time the supernatants were removed and the parasites were incubated in HBSS, pH 7.0 plus 10 \(\mu\)l of MTT for 4 hours. Viability was measured by the conversion of MTT to formazan and is expressed as the OD at 540 nm. Data are mean ± SD of NO⁻- resistant *Leishmania* isolates (Table 4) or NO⁻ susceptible *Leishmania* isolates (Table 5).
Infection of human macrophages with resistant or susceptible L. (V.) braziliensis or L. (L.) amazonensis isolates. Human Mφ from 3 healthy donors were infected with NO•-susceptible or NO•-resistant L. (L.) amazonensis or L. (V.) braziliensis and evaluated at the designated time points for the level of intracellular infection. After monolayers were stained with Giemsa, the level of infection was expressed as number of amastigotes per 100 Mφ (A, B), and as the percentage of infected Mφ (C, D) for L. (L.) amazonensis (A, C) or L. (V.) braziliensis isolates (B, D). The data are expressed as the mean ± SEM from 3 separate experiments for L. (V.) braziliensis and the mean ± SEM from 4 experiments for L. (L.) amazonensis. Parasites were used in stationary phase of growth. Statistical analysis was performed using the paired t-test.

Figure 2
matory cytokines to produce toxic metabolites that result in intracellular killing of *Leishmania* [32], or their microbicidal capacity can be dampened or abrogated by suppressive cytokines, leading to disease symptoms [33]. Specifically, TNF-α and IFN-γ elaborated by macrophages or T cells synergize to up-regulate iNOS and the NADH oxidase, with the resultant production of reactive nitrogen intermediates (RNI) and reactive oxygen intermediates (ROI), respectively, that mediate killing of intracellular *Leishmania* [32-36]. Macrophages can alternatively produce IL-10 and TGF-β that inhibit leishmanicidal activity. Both of these cytokines enable the parasite to grow locally and disseminate to distant sites [37-39].

Many prior studies focus on the host immune response during leishmaniasis and the ability of host cells/cytokines to influence the outcome of *Leishmania* infection. During the current study we focused instead on the innate susceptibility of the parasite to leishmanicidal molecules, and their ability to resist to a host microbicidal response. We found that human *L. (V.) braziliensis* and *L. (L.) amazonensis* isolates differ in their innate susceptibility to killing by RNI in vitro, falling into two groups based on their resistance to nitric oxide. Susceptible isolates of both *Leishmania* species were nearly totally killed by 8 mM of acidified NaNO₂, whereas NO*-resistant isolates remained viable even in 16 mM NaNO₂. These divisions were biologically relevant, in that patients with NO*-resistant cutaneous isolates produced significantly larger cutaneous ulcers than NO*-susceptible *Leishmania* spp. isolates. Other clinical parameters were not different between NO*-resistant and NO*-susceptible *Leishmania*, arguing against a spurious association. These clinical data suggest that NO*-resistance may lead to more aggressive forms of clinical disease. Differences in the time of storage in liquid nitrogen or in the length of in vitro promastigote cultivation could not explain our observations, since both NO*-resistant and NO*-susceptible parasites had similar time of storage and were expanded in growth medium only after selection to the present study.

Interestingly, a higher proportion of *L. (L.) amazonensis* isolates than *L. (V.) braziliensis* isolates were NO*-resistant (73% versus 18%, respectively). A comparable published study showed that promastigotes and amastigotes of *L. (L.) enriettii* were more sensitive to NO* than *L. (L.) major* [21]. In conjunction with our data, this suggests that there are inter- and intra-species variations in susceptibility to toxic nitrogen products.

Isolate-specific differences in NO* susceptibility are consistent with the observed high degree of DNA polymorphism between isolates of *L. (V.) braziliensis* from several endemic areas of Brazil, documented in literature reports. Techniques used to discern these polymorphic sequences include multilocus enzyme electrophoresis (MLEE) and internal transcribed spacers (ITS) between the small and large subunits of the tRNA gene and polymorphic DNA amplification (RAPD) [40-42]. Utilizing polymorphic DNA amplification (RAPD) we reported DNA polymorphisms in *L. (V.) braziliensis* isolates from Corte de Pedra (CP), Bahia, the same location from which the current patient isolates were derived. In addition to finding polymorphism among the *L. (V.) braziliensis* isolates, we published that certain genotypes are associated with specific forms of leishmaniasis [42]. The current study extends these observations to suggest that there are biological differences between *L. (V.) braziliensis* and *L. (L.) amazonensis* isolates that correlate with the clinical course of disease. Murine resistance to *Leishmania* infections depends at least in part on NO*-mediated intracellular killing of parasites through the action of iNOS [type 2 NO*-synthase (NOS2)]. However, the contribution of iNOS to parasite killing in human macrophages remains debated. Some reports claim a role for nitric oxide in killing of intracellular *M. tuberculosis* by human alveolar macrophages [18,43]. Our group and another has published evidence for a role of nitric oxide in macrophage microbicidal activity toward *L. (L.) chagasi/infantum* [19,44]. Nonetheless prior studies have reported difficulty in demonstrating NO* production by human macrophages [17].

Figure 3
Association between NO*-susceptibility of the *Leishmania* isolate and size of the initial cutaneous lesion in CL patients. Patients (n = 14) with cutaneous leishmaniasis were assessed for lesion size at the time of clinical presentation. This is graphed with the correlating 4 NO*-resistant *Leishmania* (3 L. (L.) amazonensis and 1 L. (V.) braziliensis) or 10 NO*-susceptible *Leishmania* (2 L. (L.) amazonensis and 8 L. (V.) braziliensis), (p = 0.01, Mann-Whitney nonparametric test).
In addition to NO* derived from the macrophage, the Leishmania spp. themselves are able to produce NO* [45-47]. It is likely that the parasite additionally has innate mechanisms for NO* resistance in order to avoid toxicity from endogenous NO*. As such, the toxic effects of exogenous NO* generated by macrophages or added experimentally would be expected to represent the sum of NO* generated by the parasite plus exogenous NO*, minus the amount of NO* scavenged or inactivated by innate parasite defense mechanisms. We hypothesize that such anti-NO* defenses may be utilized by the parasite for anti-oxidant defense in human infections. This hypothesis is supported by our observation that the degree of NO* resistance correlates with the severity of lesion.

We have previously reported that L. chagasi isolates from Brazilians with visceral leishmaniasis are susceptible to killing by NO* [19]. We showed in the current study that Leishmania isolates obtained from of humans with CL differ in their susceptibility to NO*. NO*-resistant Leishmania isolates were found to enter macrophages at a similar rate as susceptible strains, but they resisted intracellular killing by 72 to 96 h after infection. The timing of intracellular killing is consistent with the kinetics of iNOS induction, which acts 48–72 hours after infection [19]. We hypothesize that NO* resistance is one of the mechanisms enhancing parasite survival. Alternatively or additionally, NO*-resistant parasite isolates could inhibit NO* production by macrophages, or other killing mechanisms such as ROI. Importantly, it has been shown that M. bovis inhibits NO*-mediated killing by murine macrophages [48], as do Cryptococcus neoformans [49], Trypanosoma cruzi [50], as well as L. (L.) amazonensis infection [51]. Other studies have reported that amastigote surface enzymes can inhibit NO* production and thereby reduce leishmanicidal activity [52,53]. Furthermore, the LPG-associated kinetoplastid membrane protein 11 has been reported to suppress iNOS activity, because it contains monomethylarginine residues, a structural analog of L-arginine, a known inhibitor of iNOS [31,54].

The meaning of differences in NO* susceptibility amongst different Leishmania spp. isolates is not entirely straightforward. The finding that NO*-resistant Leishmania exhibit improved survival within human macrophages may indicate evasion of iNOS catalyzed toxicity as in murine macrophages and a role for iNOS in control of cutaneous leishmaniasis. Alternatively, since NO* can also play a role in signaling within the infected cell, it is possible that NO* resistant isolates are changing the intracellular signaling, or resistant to alternate microbicidal effector molecules not tested here. It seems likely that NO* resistance may contribute to the apparent increased virulence of these parasites in a human host, based on the differences in severity of the clinical parameters evaluated in the present study (significantly larger lesion size, and trend toward more resistant isolates from ML compared to CL patients). Most certainly, factors other than NO* resistance determine in part the differences in lesion size. Such factors could include the magnitude of parasite inoculum, the host immune response, and the effect of saliva. Nonetheless, it is quite interesting that the NO* resistance correlates with disease severity in our small study. Future studies are needed to better determine the clinical effects of NO* resistance on human infection and response to therapy.

Conclusion
These data suggest that nitric oxide-resistance of Leishmania isolates confers a survival benefit for the parasites inside the macrophage, and possibly exacerbates the clinical course of human leishmaniasis.

Competing interests
The author(s) declare that they have no competing interests.

Authors’ contributions
AG, EMC and RPA participated equally in the study design, and AG and RPA performed all the parasites experiments. AG, ARJ, MEW, JLH, LWR and RPA drafted the manuscript. PTGL, JMBP and IC participated in the experiments of human macrophage infection.

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