Roles of interleukin-31 (IL-31) in the development and progression of human epithelial ovarian cancer are largely unknown. Studies report that the polymorphisms, rs7977932 C>G and rs4758680 C>A in IL-31, affect the expression level of IL-31. In the present study, we examined 412 patients with epithelial ovarian cancer and 428 healthy individuals to explore whether these polymorphisms are associated with the epithelial ovarian cancer in Chinese women. The genotype of the polymorphisms in each individual was identified. The associations of the polymorphisms with patients’ clinical characteristics and outcomes were evaluated. For rs7977932, the frequency of the CG/GG was significantly decreased in patients with epithelial ovarian cancer. However, the frequency of the rs4758680 CA/AA was significantly increased in those patients. Moreover, the frequency of rs7977932 CG/GG genotype was significantly higher in patients with less advanced FIGO stages. Kaplan-Meier curve showed that patients with CG/GG genotypes of rs7977932 had a decreased risk for recurrence compared to those with CC genotype. Our findings suggested that rs7977932 and rs4758680 of IL-31 may be associated with the development and progression of the epithelial ovarian cancer in the Chinese population. IL-31, therefore, may be a potential therapeutic target for the development of drugs to treat the disease.

1. Introduction

Ovarian cancer is the most lethal gynecological cancer, and the overall five-year survival rate is less than 40% [1]. The pathogenesis of ovarian cancer, especially the epithelial ovarian cancer, is still largely unknown. Data from several investigations indicate that mutations in genes, such as tumor protein 53, breast cancers 1 and 2, are associated with the development of epithelial ovarian cancer [2–4]. In addition, other factors, such as inflammatory cytokines, may also be implicated in the development of epithelial ovarian cancer at all stages [1].

Interleukin-31 (IL-31) is mainly produced by the activated T helper type 2 cells as well as other types of cells, including monocytes, macrophages, and immature and mature dendritic and mast cells [5–7]. It has been reported that IL-31 may be involved in the pathogenesis of cutaneous allergic diseases and atopic dermatitis [8]. Recent studies suggest that IL-31 may also contribute to hematopoietic malignancies and tumor growth in human lymphoma [5, 9–11]. Results from other investigations demonstrate that IL-31 can significantly increase expressions of epidermal growth factor and vascular endothelial growth factor [12, 13]. Thus, IL-31 has been considered an angiogenic factor to lung
epithelial cells, which may play a role in the development of lung cancer [12, 14]. In contrast to the aforementioned finds, the results from other independent investigations demonstrate that IL-31 is highly effective in suppressing the proliferation of lung epithelial cells and has the antiproliferative effect on the cells [14, 15]. IL-31 and its receptor, IL31RA, are highly expressed in tumor specimens from ovarian cancer patients and other human and mouse cancer cell lines [16]. IL-31 may have anticancer effects on certain kinds of tumors. For instance, murine colon carcinoma cells without IL-31 have greater invasive, migratory, and tumor growth properties, while these effects can be reversed by supplementing exogenous IL-31 to the cells. Moreover, the tumor growth in mice can also be inhibited by the administration of exogenous IL-31 [16]. The aforementioned information prompted us to investigate potential roles of IL-31 in epithelial ovarian cancer.

Single nucleotide polymorphisms (SNPs) in IL-31 provide us with an opportunity to investigate potential effects of the factor in the development and progression of epithelial ovarian cancer in humans. The SNP is the most common type of the genetic variation in human genome. A SNP can alter the expression of a gene, which leads to a specific phenotype of cells. For example, polymorphisms in IL-31 have been associated with the development of diseases and cancer [16–20]. In particular, systemic lupus erythematosus (SLE) patients carrying rs7977932 G allele in IL-31 have the high-level protein of IL-31 in serum [19]. Recent data show that the level of IL-31 mRNA in white blood cells is increased in dilated cardiomyopathy (DCM) patients, associated with a lower frequency of the CA/AA at rs4758680, in IL-31 [20]. However, there is no information of the polymorphisms, rs7977932 and rs4758680 of IL-31, in epithelial ovarian cancer patients.

In this present investigation, we retrospectively examined the genotypes of the rs7977932 and rs4758680 in 412 epithelial ovarian cancer patients. The rs7977932 and rs4758680 locate within IL-31 and are 2031 base pairs apart. The associations of the rs7977932 and rs4758680, with the epithelial ovarian cancer, were statistically calculated. The associations of the rs7977932 and rs4758680 with the cancer features, survival outcome, and recurrence were also evaluated.

### 2. Materials and Methods

#### 2.1. Study Subjects

Four hundred and twelve patients, aged 51.2 years (average) ± 9.6 years (SD), were diagnosed with epithelial ovarian cancer and treated at the West China Second University Hospital. Four hundred and twenty-eight healthy female individuals, aged 49.7 years (average) ± 13.5 years (SD), were enrolled in this hospital-based retrospective case-control study (Table 1). We excluded individuals with the borderline ovarian tumors, two or more different malignancies and autoimmune diseases from this study. The tumor stage was classified based on the International Federation of Gynecology and Obstetrics ovarian cancer staging criteria (FIGO, 2014). Healthy individuals were from a routine physical examination in the hospital. They had no personal or family history of cancer, autoimmune diseases, or other serious illnesses. The present investigation has been approved by the Ethics Committee of the West China Second Hospital, and the consent form was obtained from all participants.

### 2.2. Determination of Genotypes

The SNPs, rs7977932 (C > G) and rs4758680 (C > A), were genotyped by restriction fragment length polymorphism (RFLP) of PCR products. The three kinds of SNP genotypes were also confirmed by direct DNA sequence. For PCR, the genomic DNA of each patient was isolated from the paraffin section of the tumor tissue sample by the DNA isolation kit (BioTeke, Beijing, China). Genomic DNAs of the healthy individuals were extracted from 200 µL EDTA-anticoagulated peripheral blood sample by the whole blood DNA isolation kit (BioTeke, Beijing, China). For genotyping of the rs7977932, the primers for PCR were forward 5′-GGTCAGTGTTGGGTTTGCAAT and reverse 5′-TTGGTGATGGCACAGCCTCATAC-3′. For the rs4758680, the primers for PCR were forward 5′-GGTCTGTGGGTGGAGACAGCCTCATAC-3′ and reverse 5′-TTCCCGGAGTAAGATGAAGCCTCATAC-3′. The PCR reaction was 25 µl including 0.5 mmol/L each PCR primer, 100 ng of the genomic DNA, 0.15 mmol/L of dNTPs, 2.5 µl of 10× Taq Buffer, and 1 U of Taq DNA polymerase (BioTeke, Beijing, China). The PCR condition was 95°C for 4 minutes followed by 36 cycles at 95°C for 30 seconds, 60°C for 30 seconds, and 72°C for 30 seconds and a final extension by 72°C for 10 minutes. PCR products were then digested by the restriction enzyme ScrFI (for rs7977932) overnight and the enzyme MboII (for rs4758680) for 35 minutes at 37°C. The rs7977932 allele C was not digested by ScrFI, and the size of the PCR product was 131 base pairs. The rs7977932 allele G was digested by ScrFI to generate two DNA fragments, 74 and 57 base pairs. The rs4758680 allele A was not digested by

| Table 1: Descriptive characteristics of the epithelial ovarian cancer patients. |
|-------------------------------|------------------|
| Variable                      | Value            |
| Sample size                   | n = 412          |
| Mean age ± SD, range (years)  | 51.2 ± 9.6, 15–85|
| FIGO stage, number (%)        |                  |
| I                             | 43 (10.44%)      |
| II                            | 48 (11.65%)      |
| III                           | 297 (72.09%)     |
| IV                            | 24 (5.83%)       |
| Tumor grade, number (%)       |                  |
| G1                            | 7 (1.70%)        |
| G2                            | 25 (6.07%)       |
| G3                            | 380 (92.23%)     |
| Histological type, number (%) |                  |
| Serous                        | 333 (80.83%)     |
| Clear cell                    | 16 (3.88%)       |
| Endometrioid                  | 9 (2.18%)        |
| Mucinous                      | 9 (2.18%)        |
| Mixed and other               | 45 (10.92%)      |
2.4. Statistical Analysis. The differences of genotypes between the epithelial ovarian cancer patients and the controls were analyzed using SNPStats online software (https://www.snpstats.net/snpstats/start.htm), which provided the genotypic association including codominant, dominant, recessive, or overdominant genetic models. Odd ratios (OR) with a 95% confidence interval (95%CI) were obtained accordingly. Hardy-Weinberg equilibrium and the association of the genotypes and the alleles with patients’ clinical characteristics were assessed by chi-square analysis. Kaplan-Meier curve and Cox proportional hazard models were applied for the evaluation of the role of IL-31 SNPs and other factors on prognosis of epithelial ovarian cancer patients. The level of the statistical significance was set at $P < 0.05$.

3. Results

3.1. The SNPs of IL-31, rs7977932, and rs4758680, in Epithelial Ovarian Cancer Patients. The SNPs rs7977932 and rs4758680 were genotyped in the 412 patients with epithelial ovarian cancer and 428 control individuals. Genotypes of the SNPs, rs7977932 (CC, CG, and GG) and rs4758680 (CC, CA, and AA), were identified. The frequencies of all observed genotypes in both patients and controls were in agreement with the Hardy-Weinberg equilibrium ($P > 0.05$). Genotype distributions and allele frequencies of the SNPs of the IL-31 in patients and controls are shown in Table 2. For the rs7977932 C>G polymorphism, G allele carriers were associated with a significantly decreased epithelial ovarian cancer development ($P = 0.0024$, OR = 0.55, 95% CI = 0.37–0.81) in a dominant model (CG/GG versus CC). Similarly, significant associations were also observed in a codominant model (CG versus CC; $P = 0.01$, OR = 0.54, 95% CI = 0.36–0.82) and an overdominant model (CG versus CC/GG; $P = 0.0033$, OR = 0.55, 95% CI = 0.36–0.83). And the overall allele G is significantly lower in epithelial ovarian cancer patients than in the controls (G versus C; $P = 0.0033$, OR = 0.58, 95% CI = 0.40–0.84). For the rs4758680 C>A polymorphism, A allele carriers were associated with a significantly increased epithelial ovarian cancer development ($P = 0.012$, OR = 1.45, 95% CI = 1.09–1.92) in a dominant model (CA/AA versus CC). Similarly, significant associations were also observed in a codominant model (CA versus CC; $P = 0.042$, OR = 1.45, 95%CI = 1.09–1.96) and an overdominant model (CA versus CC/AA; $P = 0.017$, OR = 1.43, 95%CI = 1.06–1.92). And the overall allele A is significantly higher in epithelial ovarian cancer patients than the controls (A versus C; $P = 0.024$, OR = 1.32, 95% CI = 1.04–1.67).

3.2. IL-31 SNPs and Epithelial Ovarian Cancer Patients’ Characteristics. To further evaluate whether the genotype and allele polymorphism were associated with certain clinical features of epithelial ovarian cancer patients, we performed the stratification analyses for genotype and allele distribution in epithelial ovarian cancer patients with different FIGO stages (I + II and III + IV), tumor grades (G1 + G2 and grade G3), histological types (serous and others), and ages (<50 and ≥50 years old). No statistically significant differences were found for any subgroups of the two SNPs except for the FIGO stages. For rs7977932, the percentage of CG/GG genotype was significantly higher in patients at early stages (I and II) compared with those at advanced stages (III and IV) ($P = 0.042$, OR = 0.50, 95% CI = 0.26–0.99). The proportion of patients with G allele was also significantly higher at early stages than at advanced stages ($P = 0.017$, OR = 0.48, 95% CI = 0.26–0.88) (Table 3).

3.3. Survival and Recurrence Analysis. In the present study, no significant associations between the two SNPs (rs7977932 and rs4758680) and the overall survival of the epithelial ovarian cancer patients were observed by both the univariate and multivariate analysis (data not shown). However, the univariate analysis (Kaplan-Meier curve) showed that patients with CG/GG genotypes of rs7977932 had a decreased risk for recurrence compared to those with CC genotype ($P = 0.045$, Figure 1). No significant association was found between rs4758680 and the recurrence outcomes of the patients. Then, multivariate analysis using Cox proportional hazard models, adjusted by FIGO stage, histology type, and tumor grade, was carried out to examine the effects of IL-31 polymorphisms on patients’ outcomes. IL-31 polymorphisms rs7977932 yielded no independent prognostic effect ($P = 0.199$), while FIGO stage (HR = 0.249, 95% CI = 0.074–0.837, $P = 0.025$) and histological types (HR = 2.207, 95% CI = 1.021–4.774, $P = 0.044$) were independent prognostic factors for recurrence outcomes.

4. Discussion

In this investigation, the frequency of the CG/GG of the rs7977932 was significantly decreased in the patients than
Table 2: Distribution of SNPs in *IL-31* among the cases and controls and their association with the epithelial ovarian cancer.

| Model          | Genotype | Case, n (%) | Control, n (%) | OR (95% CI) | P value | Genotype | Case, n (%) | Control, n (%) | OR (95% CI) | P value |
|----------------|----------|-------------|----------------|-------------|---------|----------|-------------|----------------|-------------|---------|
|                |          | rs7977932   |                |             |         |          | rs4758680   |                |             |         |
| Codominant     | CC       | 368 (89.3%) | 351 (82%)      | 1.00        |         | CC       | 252 (61.2%) | 297 (69.4%)    | 1.00        |         |
|                | CG       | 41 (9.9%)   | 72 (16.8%)     | 0.54 (0.36–0.82) | 0.01*   | CA       | 141 (34.2%) | 114 (26.6%)    | 1.45 (1.09–1.96) | 0.042*   |
|                | GG       | 3 (0.7%)    | 5 (1.2%)       | 0.57 (0.14–2.44) |         | AA       | 19 (4.6%)   | 17 (4%)        | 1.32 (0.67–2.56) |         |
| Dominant       | CC       | 368 (89.3%) | 351 (82%)      | 1.00        |         | CC       | 252 (61.2%) | 297 (69.4%)    | 1.00        |         |
|                | CG/GG    | 44 (10.7%)  | 77 (18%)       | 0.55 (0.37–0.81) | 0.0024* | CA/AA    | 160 (38.8%) | 131 (30.6%)    | 1.45 (1.09–1.92) | 0.012*   |
| Recessive      | CC/CX    | 409 (99.3%) | 423 (98.8%)    | 1.00        |         | CC/CA    | 393 (95.4%) | 411 (96%)      | 1.00        |         |
|                | GG       | 3 (0.7%)    | 5 (1.2%)       | 0.62 (0.15–2.63) | 0.51    | AA       | 19 (4.6%)   | 17 (4%)        | 1.16 (0.60–2.27) | 0.65    |
| Overdominant   | CC/GG    | 371 (90%)   | 356 (83.2%)    | 1.00        |         | CC/AA    | 271 (65.8%) | 314 (73.4%)    | 1.00        |         |
| Allele         | CG       | 41 (9.9%)   | 72 (16.8%)     | 0.55 (0.36–0.83) | 0.0033* | CA       | 141 (34.2%) | 114 (26.6%)    | 1.43 (1.06–1.92) | 0.017*   |
|                | C        | 775 (94%)   | 774 (90%)      | 1.00        |         | C        | 645 (78%)   | 708 (83%)      | 1.00        |         |
|                | G        | 47 (6%)     | 82 (10%)       | 0.58 (0.40–0.84) | 0.0033* | A        | 179 (22%)   | 148 (17%)      | 1.32 (1.04–1.67) | 0.024*   |

n corresponds to the number of individuals; *P < 0.05.
Table 3: Association between SNPs, rs7977932, and rs4758680 and clinic features of the epithelial ovarian cancer.

| Clinical features          | rs7977932 Genotype frequency (%) | OR (95% CI) | P value | rs4758680 Genotype frequency (%) | OR (95% CI) | P value |
|---------------------------|----------------------------------|-------------|---------|----------------------------------|-------------|---------|
| FIGO stage I-II II-IV     | 76 (83.52%) 292 (90.97%)        | 1.00        |         | CC 55 (60.44%) 197 (61.37%)     | 1.00        |         |
| CC                        | 15 (16.48%) 29 (9.03%)          | 0.50 (0.26–0.99) | 0.042*  | CA/AA 36 (39.56%) 124 (38.63%)  | 0.96 (0.60–1.55) | 0.872  |
| CG/GG                     | 165 (90.66%) 612 (93.33%)      | 1.00        |         | C 141 (77.47%) 504 (78.50%)    | 1.00        |         |
| C                         | 17 (9.34%) 30 (4.67%)           | 0.48 (0.26–0.88) | 0.017*  | A 41 (22.53%) 138 (21.50%)    | 0.94 (0.63–1.40) | 0.766  |
| Tumor grade G1-G2 G3      | 28 (87.50%) 340 (89.47%)       | 1.00        |         | CC 21 (65.63%) 231 (60.79%)   | 1.00        |         |
| CC                        | 4 (12.50%) 40 (10.53%)         | 0.82 (0.28–2.47) | 0.961  | CA/AA 11 (34.38%) 149 (39.21%) | 1.23 (0.58–2.63) | 0.590  |
| C                         | 60 (93.75%) 717 (94.34%)       | 1.00        |         | C 53 (82.81%) 592 (77.89%)    | 1.00        |         |
| G                         | 4 (6.25%) 43 (5.66%)           | 0.90 (0.31–2.50) | 0.779  | A 11 (17.19%) 168 (22.11%)   | 1.37 (0.70–2.68) | 0.360  |
| Histological type Serous Others | 295 (88.59%) 73 (92.41%)   | 1.00        |         | CC 202 (60.66%) 50 (63.29%)  | 1.00        |         |
| CC                        | 38 (11.41%) 6 (7.59%)          | 0.64 (0.26–1.57) | 0.323  | CA/AA 131 (39.34%) 29 (36.71%) | 0.89 (0.54–1.49) | 0.666  |
| C                         | 625 (93.84%) 152 (96.20%)      | 1.00        |         | C 520 (78.08%) 125 (79.11%)   | 1.00        |         |
| G                         | 41 (6.16%) 6 (3.80%)           | 0.60 (0.25–1.44) | 0.250  | A 146 (21.92%) 33 (20.89%)   | 0.94 (0.62–1.44) | 0.777  |
| Age (years) <50 ≥50       | 164 (87.70%) 204 (90.67%)      | 1           |         | CC 112 (59.89%) 140 (62.22%)  | 1           |         |
| CC                        | 23 (12.30%) 21 (9.33%)         | 0.73 (0.40–1.37) | 0.332  | CA/AA 75 (40.11%) 85 (37.78%) | 0.91 (0.61–1.35) | 0.629  |
| C                         | 349 (93.32%) 428 (95.11%)      | 1.00        |         | C 288 (77.01%) 357 (79.33%)   | 1.00        |         |
| G                         | 25 (6.68%) 22 (4.89%)          | 0.72 (0.40–1.30) | 0.269  | A 86 (22.99%) 93 (20.67%)    | 0.87 (0.63–1.22) | 0.420  |

* P < 0.05.
in the healthy individuals. This suggested that the lower frequency of the CG/GG increased the risk of the epithelial ovarian cancer. It indicates that the CG/GG may be associated with the protective effect of IL-31 against the development of the epithelial ovarian cancer. Our result is in line with the previous observations that the CG polymorphism is associated with the elevated level of IL-31 in serum [16]. The increase in the expression of IL-31 may be able to reverse cancer invasive and migratory abilities and inhibit tumor growth [19]. For the rs4758680, the frequency of the CA/AA was significantly higher in the epithelial ovarian cancer patients than in the healthy individuals. This suggested that the CA/AA might be a risk factor for the development of the epithelial ovarian cancer. A recent study reports that the level of IL-31 mRNA expressed in white blood cells is increased in DCM compared with that in healthy individuals, while the frequency of the CA/AA genotypes is lower in DCM [20]. Although the genotypes of CA/AA may affect the expression of IL-31, there is no direct evidence to show whether the higher CA/AA is associated with the reduced level of IL-31. Taken all these findings together, it indicates that the CG/GG associated with the increased expression of IL-31 may have an antiepithelial ovarian cancer effect in a Chinese population.

We also found that the percentage of the CG/GG was significantly higher in patients with the epithelial ovarian cancer at the early stages I and II, compared with that in patients at the advanced stages III and IV. This suggests that the lower frequency of the CG/GG may lead to the development of more severe epithelial ovarian cancer. Thus, the increased level of IL-31 by the genotypes of the CG/GG may contribute to the reduction of the epithelial ovarian cancer severity. Our result is also in agreement with the previous finding by others that IL-31 is highly effective in suppressing the proliferation of lung epithelial cells [15].

In the follow-up investigation, we found that patients with CG/GG genotypes of rs7977932 had a decreased risk for recurrence compared to those with CC genotype. However, these genotypes were not independent risk factors for the prognosis of the patients with the epithelial ovarian cancer. Results from the Cox proportional hazard modeling showed that the independent, significant prognostic factors were FIGO stage and histological type. Variables, such as tumor grade, and the genotypes of the two SNPs (rs7977932 and rs4758680) were insignificant in predicting the epithelial ovarian cancer 5-year recurrence. These results partially support the finding by others that IL-31 might play an antiangiogenic role in tumor formation [16]. A lower recurrence risk of the epithelial ovarian cancer in patients with the CG/GG genotypes may represent the potential antiangiogenic effect of IL-31 since tumor growth is highly dependent on angiogenesis [21–23]. However, the FIGO stage and histological type play more dominant roles in the prognostic outcome of epithelial ovarian cancer, and these SNPs are not an independent prognostic factor for epithelial ovarian cancer. Interestingly, data from a study of patients with breast cancer demonstrate that there is a significant extended survival to breast cancer patients with a higher level expression of IL31RA [16, 24].

Lastly, in the present study, there were no associations between rs4758680 polymorphism and the patients’ clinical characteristics such as epithelial ovarian cancer stages by FIGO, or between the rs4758680 polymorphism and the recurrence and survival risks of epithelial ovarian cancer. One of the possible explanations is that the polymorphism rs4758680 alone may not be sufficient enough to contribute to the stages and outcomes of the cancer.

5. Conclusion

Results from our present investigation indicate the significant association of the SNPs rs7977932 and rs4758680 in IL-31 with the development of epithelial ovarian cancer in a Chinese population. Combined with the findings from other independent research groups, it suggests that IL-31 can protect ovarian epithelial cells from being malignant. Thus, IL-31 may potentially be used as an antiovarian cancer drug to treat the disease [21].

Conflicts of Interest

The authors have no conflict of interests to declare.

Authors’ Contributions

Chenlu Liu and Yanyun Wang contributed equally to this work.
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References

[1] M. P. Jammal, A. Martins-Filho, T. P. Silveira, E. F. Murta, and R. S. Nomelini, “Cytokines and prognostic factors in epithelial ovarian cancer,” Clinical Medicine Insights: Oncology, vol. 10, no. 10, pp. CMO.S38333–CMO.S38336, 2016.

[2] Y. Miki, J. Swensen, D. Shattuck-Eidens et al., “A strong candidate for the breast and ovarian cancer susceptibility gene BRCA1,” Science, vol. 266, no. 5182, pp. 66–71, 1994.

[3] K. B. Kuchenbaecker, S. J. Ramus, J. Tyrer et al., “Identification of six new susceptibility loci for invasive epithelial ovarian cancer,” Nature Genetics, vol. 47, no. 2, pp. 164–171, 2015.

[4] L. Havrilesky, K. M. Darcy, H. Hamdan et al., “Prognostic significance of P53 mutation and P53 overexpression in advanced epithelial ovarian cancer: a gynecologic oncology group study,” Journal of Clinical Oncology, vol. 21, no. 20, pp. 3814–3823, 2003.

[5] E. Ferretti, C. Tripodo, G. Pagnan et al., “The interleukin (IL)-31/IL-31r axis contributes to tumor growth in human follicular lymphoma,” Leukemia, vol. 29, no. 4, pp. 958–967, 2015.

[6] M. Lange, J. Glei, M. Zablotna et al., “Interleukin-31 polymorphisms and serum IL-31 level in patients with mastocytosis: correlation with clinical presentation and Pruritus,” Acta Dermato Venereologica, vol. 97, no. 1, pp. 47–53, 2017.

[7] S. R. Dillon, C. Sprecher, A. Hammond et al., “Interleukin 31, a cytokine produced by activated T cells, induces dermatitis in mice,” Nature Immunology, vol. 5, no. 7, pp. 752–760, 2004.

[8] H. M. Hermanns, “Oncostatin M and interleukin-31: cytokines, receptors, signal transduction and physiology,” Cytokine & Growth Factor Reviews, vol. 26, no. 5, pp. 545–558, 2015.

[9] T. Ishii, J. Wang, W. Zhang et al., “Pivotal role of mast cells in pruritogenesis in patients with myeloproliferative disorders,” Blood, vol. 113, no. 23, pp. 5942–5950, 2009.

[10] L. A. Nattkemper, M. E. Martinez-Escala, A. B. Gelman et al., “Cutaneous T-cell lymphoma and pruritus: the expression of IL-31 and its receptors in the skin,” Acta Dermato Venereol, vol. 96, no. 7, pp. 894–898, 2016.

[11] H. Ohmatsu, M. Sugaya, H. Sugah et al., “Serum IL-31 levels are increased in patients with cutaneous T-cell lymphoma,” Acta Dermato Venereologica, vol. 92, no. 3, pp. 282–3, 2012.

[12] W. Naumnik, B. Naumnik, K. Niewiarowska, M. Ossolinska, and E. Chyczewska, “Novel cytokines: II-27, II-29, II-31 and II-33. Can they be useful in clinical practice at the time diagnosis of lung cancer?,” Experimental Oncology, vol. 34, no. 4, pp. 348–353, 2012.

[13] W. K. Ip, C. K. Wong, M. L. Y. Li, P. W. Li, P. F. Y. Cheung, and C. W. K. Lam, “Interleukin-31 induces cytokine and chemokine production from human bronchial epithelial cells through activation of mitogen-activated protein kinase signaling pathways: implications for the allergic response,” Immunology, vol. 122, no. 4, pp. 532–541, 2007.

[14] X. Zeng, Z. Zhang, Q.-Q. Gao et al., “Clinical significance of serum interleukin-31 and interleukin-33 levels in patients of endometrial cancer: a case control study,” Disease Markers, vol. 2016, Article ID 9262919, 7 pages, 2016.

[15] S. Chattopadhyay, E. Tracy, P. Liang, O. Robledo, S. Rose John, and H. Baumann, “Interleukin-31 and oncostatin-M mediate distinct signaling reactions and response patterns in lung epithelial cells,” Journal of Biological Chemistry, vol. 282, no. 5, pp. 3014–3026, 2017.

[16] S. Davidi, E. Fremder, T. Kan et al., “The antiangiogenic role of the pro-inflammatory cytokine interleukin-31,” Oncotarget, vol. 8, no. 10, pp. 16430–16444, 2017.

[17] C. C. Lan, H. P. Tu, C. S. Wu et al., “Distinct Spink5 and IL-31 polymorphisms are associated with atopic eczema and non-atopic hand dermatitis in Taiwanese nursing population,” Experimental Dermatology, vol. 20, no. 12, pp. 975–979, 2011.

[18] A. Rabenhorst and K. Hartmann, “Interleukin-31: a novel diagnostic marker of allergic diseases,” Current Allergy and Asthma Reports, vol. 14, no. 4, p. 423, 2014.

[19] H. T. Huang, J. M. Chen, J. Guo, Y. Lan, and Y. S. Wei, “The association of interleukin-31 polymorphisms with interleukin-31 serum levels and risk of systemic lupus erythematosus,” Rheumatology International, vol. 36, no. 6, pp. 799–805, 2016.

[20] H. Song, Y. Peng, B. Zhou et al., “Associations between Interleukin-31 gene polymorphisms and dilated cardiomyopathy in a Chinese population,” Disease Markers, vol. 2017, Article ID 4191365, 10 pages, 2017.

[21] J. Folkman, “Anti-angiogenesis: new concept for therapy of solid tumors,” Annals of Surgery, vol. 175, no. 3, pp. 409–416, 1972.

[22] Y. Shaked, E. Henke, J. M. L. Roodhart et al., “Rapid chemotheraphy-induced acute endothelial progenitor cell mobilization: implications for antiangiogenic drugs as chemosensitizing agents,” Cancer Cell, vol. 14, no. 3, pp. 263–273, 2008.

[23] Y. Shaked and R. S. Kerbel, “Antiangiogenic strategies on defense: on the possibility of blocking rebounds by the tumor vasculature after chemotherapy,” Cancer Research, vol. 67, no. 15, pp. 7055–7058, 2007.

[24] G. Balazs, Kaplan-Meier Plotter, 2009, http://kmplot.com.