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Published in:
Cancer immunology and immunotherapy

DOI:
10.1007/s00262-010-0908-z

Citation for published version (APA):
van Luijn, M. M., Chamuleau, M. E. D., Ressing, M. E., Wiertz, E. J., Ostrand-Rosenberg, S., Souwer, Y., ... Ham, S. M. (2010). Alternative II-independent antigen-processing pathway in leukemic blasts involves TAP-dependent peptide loading of HLA class II complexes. Cancer immunology and immunotherapy, 59(12), 1825-1838. DOI: 10.1007/s00262-010-0908-z
Alternative Ii-independent antigen-processing pathway in leukemic blasts involves TAP-dependent peptide loading of HLA class II complexes

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Abstract During HLA class II synthesis in antigen-presenting cells, the invariant chain (Ii) not only stabilizes HLA class II complexes in the endoplasmic reticulum, but also mediates their transport to specialized lysosomal antigen-loading compartments termed Mfcs. This study explores an alternative HLA class II presentation pathway in leukemic blasts that involves proteasome and transporter associated with antigen processing (TAP)-dependent peptide loading. Although HLA-DR did associate with Ii, Ii silencing in the human class II-associated invariant chain peptide (CLIP)-negative KG-1 myeloid leukemic cell line did not affect total and plasma membrane expression levels of HLA-DR, as determined by western blotting and flow cytometry. Since HLA-DR expression does require peptide binding, we examined the role of endogenous antigen-processing machinery in HLA-DR presentation by CLIP- leukemic blasts. The suppression of proteasome and TAP function using various inhibitors resulted in decreased HLA-DR levels in both CLIP- KG-1 and ME-1 blasts. Simultaneous inhibition of TAP and Ii completely downmodulated the expression of HLA-DR, demonstrating that together these molecules form the key mediators of HLA class II antigen presentation in leukemic blasts. By the use of a proteasome- and TAP-dependent pathway for HLA class II antigen presentation, CLIP- leukemic blasts might be able to present a broad range of endogenous leukemia-associated peptides via HLA class II to activate leukemia-specific CD4+ T cells.

Keywords MHC class II · Tumor cells · Antigen presentation · Leukemia · Immune surveillance

Introduction

Professional antigen-presenting cells (APC), such as dendritic cells (DCs) and macrophages, can activate T cells by efficiently processing antigens from both endogenous and exogenous sources and presenting them at the plasma membrane.

HLA class I antigen-processing machinery is essential for the generation of endogenous peptides and loading onto HLA class I molecules in the endoplasmic reticulum (ER). One of its major components is the proteasome, a multicatalytic protease complex that specifically cleaves ubiquitin-conjugated cytoplasmic proteins into small peptides [1]. Another key player, the transporter associated...
with antigen processing (TAP), mediates translocation of these peptides into the ER [2], where further trimming to nonameric peptides may be necessary for HLA class I binding [3]. Upon complete folding in the ER lumen, the class I/peptide complex is exported via the default secretory pathway to the plasma membrane.

Exogenously derived peptides are generated in the endosomal/lysosomal pathway and loaded onto HLA class II molecules in specialized lysosomal antigen-loading compartments, termed MIIcs (or MHC class II-containing compartments) [4, 5]. Newly synthesized HLA class II αβ heterodimers associate with the invariant chain (Ii or CD74) in the ER lumen, which not only ensures stabilization and proper folding, but also transport to the MIIcs [6, 7]. During transport to the MIIcs, Ii is proteolytically degraded, leaving a small fragment bound to the peptide-binding groove [8]. In the MIIcs, this class II-associated invariant chain peptide (CLIP) is exchanged for an exogenous peptide by the peptide editor HLA-DM (DM) [9], and the class II/peptide complex is transported to the plasma membrane.

As extensively described in myeloid DCs, exogenous peptides are able to interact with HLA class I molecules via different processing pathways, a mechanism known as cross-presentation [10, 11]. Endogenous peptides, on the other hand, can be loaded onto HLA class II molecules as well, although it remains controversial which processing pathways underlie this type of peptide loading. In many professional APCs, endogenous antigens are transferred to the endosomal/lysosomal pathway for loading onto HLA class II molecules [12]. Autophagy has been reported to play a role in this process by delivering cytoplasmic material for degradation in the lysosomes [13–15], but also the proteasome could be involved by degrading cytoplasmic antigens before entering the endosomal/lysosomal pathway [16, 17].

Although HLA class II transport into this pathway is classically known to be dependent of Ii, the absence of Ii in tumor APCs was shown to clearly increase HLA class II-restricted presentation of endogenous peptides and their capacity to activate tumor-specific CD4+ T cells [18, 19]. Taking into account that the association of Ii with HLA class II interferes with endogenous peptide binding in the ER [20, 21], this suggests that in certain tumor APCs, loading of HLA class II molecules with endogenous peptides occurs via HLA class I antigen-processing machinery instead of the endosomal/lysosomal pathway. It has recently been shown that both the proteasome and TAP were involved in the processing of HLA class II-restricted viral epitopes [22], but less is known about this function of HLA class I antigen-processing machinery in HLA class II+ tumor APCs. A better understanding of endogenous antigen processing for HLA class II-restricted presentation in HLA class II+ tumor cells could be important for developing approaches to optimize presentation of, yet unidentified, tumor-associated antigens (TAAs), thereby circumventing tumor immune escape.

We previously demonstrated that a low amount of CLIP on HLA class II+ leukemic blasts predicts an improved clinical outcome for patients with acute myeloid leukemia (AML) and enhances recognition by allogeneic CD4+ T cells [23]. In the current study, we show that an Ii-independent, but proteasome- and TAP-dependent HLA class II presentation pathway is active in a human CLIP- myeloid leukemic cell line, which indicates that HLA class I antigen-processing machinery mediates HLA class II-restricted presentation of endogenous peptides in these tumor cells.

Materials and methods

Cell lines and culturing

The human myeloid leukemic cell lines KG-1 and Kasumi-1 were obtained from the American Type Culture Collection (ATCC) and frozen in aliquots at low cell passage. Culturing of the KG-1 cell line was performed at 3 × 10^5 cells/ml in IMDM (Gibco) supplemented with 20% FBS, 1% l-glutamine (Gibco), 25 mM Hepes (Sigma-Aldrich, St Louis, MO, USA) and 50 μM 2-ME (Gibco). The Kasumi-1 cell line was maintained in RPMI 1640 medium (Gibco, Paisley, UK) supplemented with 1% l-glutamine and 15% heat-inactivated FBS (Greiner, Alphen a/d Rijn, The Netherlands) with a cell density of 5 × 10^5 cells/ml. Cells were incubated at 37°C in a humidified atmosphere containing 5% CO2 and the culture medium was refreshed every 3 days.

Antibodies and immunofluorescence staining

The following mouse monoclonal antibodies (MoAbs) were used: PE-labeled anti-HLA-ABC (W6/32 clone; Dako, Glostrup, Denmark) and IgG2a isotype control (BD, San Jose, CA, USA); APC-labeled anti-HLA-DR (clone L243) and IgG2a isotype control (BD); and 7-amino-actinomycin D (7AAD; Via-Probe, BD). The PIN1.1 antibody (anti-Ii) was kindly provided by Peter Cresswell (Howard Hughes Medical Institute, Yale University School of Medicine, New Haven, CT, USA) [24]. For immunofluorescence stainings, 1 × 10^5 cells were pre-incubated with 10% human gamma-globulin (60 mg/ml; Sanquin) for 10 min. Mouse MoAbs were added during 15 min for extracellular and 30 min for intracellular staining. To analyze total protein expression, cells were fixed with PBS 1% paraformaldehyde, permeabilized using PBS 0.1% saponin (Sigma-Aldrich) and stained with the
MoAb of interest. We examined the total amount of Ii by staining with PIN1.1 followed by a second incubation step of 20 min with rabbit anti-mouse (RAM) IgG conjugated to PE (Dako). Stained cells were incubated at room temperature (RT), washed twice with PBS containing 0.1% HSA and 0.05% sodium azide, and measured on a FACSAria flow cytometer (BD). The same apparatus was used for flow cytometric sorting of cells. Sorted cells were washed three times with PBS before subsequent analysis or culture in medium. We performed flow cytometric analyses by using CellQuest software (BD) and determined extracellular expression levels on viable (7AAD−) cells. Mean fluorescence intensity (MFI) was defined as the median value of all gated cells relative to that of isotype control staining.

DNA constructs and retroviral vectors

Epstein-Barr virus (EBV)-encoded BNLF2a and bovine herpes virus 1 (BHV-1)-encoded UL49.5 were amplified from viral DNA with specifically selected primers and PCR products were cloned into pLZRS-IRES-EGFP, as previously reported [25, 26]. Information about this vector can be obtained at http://www.stanford.edu/group/nolan/retroviral_systems/retsys.html. Ii silencing was performed using a pSIREN-RetroQ retroviral vector (Clontech) that contains a specifically prepared Ii-siRNA insert (sequence 53) with a puromycin resistance gene [27].

Retroviral production, transductions and transfections

BNLF2a and UL49.5 retroviruses were produced using the Phoenix amphotropic packaging system, as described at http://www.stanford.edu/group/nolan/protocols/pro_helper_dep.html. For the production of UL49.5 retroviruses, we used a pLZRS vector that encodes a mutant form of UL49.5, containing two alanines instead of lysines within the cytoplasmic tail. This construct did not alter UL49.5-mediated degradation and inhibition of the TAP complex [28]. Retroviral transductions and transfections were performed according to standard protocols [23]. The possibility of off-target effects during these procedures were excluded by using an irrelevant GFP-encoding retrovirus (supplementary material, Fig. S2) and irrelevant siRNAs [27].

For transfection experiments, 2 μg of pSIREN-RetroQ-Ii-siRNA53 DNA was delivered into 2 × 10⁶ cells using the Amaxa nucleofection technology (Lonza, Basel, Switzerland). The optimized protocol for the KG-1 cell line is available at http://www.lonzabiomed.com. Before treated cells were resuspended in culture medium again, we first recovered them in IMDM (Gibco) without the addition of supplements for 30 min at 37°C. Two days after transfection, we added 1.0 μg/ml puromycin to select Ii-siRNA53-transfected cells.

Co-immunoprecipitations and western blotting

To immunoprecipitate Ii proteins, 10 × 10⁶ myeloid leukemia blasts were washed three times with sterile PBS (pH 7.4). Cell lysates were prepared by snap freezing the samples in liquid nitrogen and incubating them upon thawing for 45 min in 250 μl ice-cold lysis buffer, which consisted of sterile PBS supplemented with 1% Ipegal and 15% protease inhibitor cocktail (Complete; 1 tablet/7.5 ml H₂O₂; Boehringer Mannheim Biochemica, Mannheim, Germany). After removal of unlysed cell remnants and nuclei by centrifugation in an Eppendorf micro-centrifuge (5 min, 10,000 rpm), the protein content of the supernatant was determined by the Bio-Rad protein assay (Biorad Laboratories, Hercules, CA, USA). Then, 0.5 mg total cell lysates were diluted in 0.5 ml ice-cold lysis buffer and precleared with 1.0 μg of mouse IgG1 MoAb together with 20 μl Protein G PLUS-Agarose reagent (Santa Cruz Biotechnology, Santa Cruz, CA, USA) for 30 min. It was immunoprecipitated using the PIN1.1 antibody and Protein G PLUS-Agarose reagent (20 μl) in an overnight incubation. Ii immunoprecipitates were collected by centrifugation and washed four times with PBS before resuspending in reducing electrophoresis sample buffer.

For western blotting, samples were electrophoresed on 12.5% polyacrylamide gels that contained SDS and transferred onto methanol-activated PVDF membranes. Different protein amounts from total cell lysates were loaded on gel: 10 μg for HLA-DRz, 20 μg for Ii and 70 μg for TAP1. Mouse anti-DRz (clone 1B5 [29], 1:10,000), anti-Ii (clone PIN1.1 [24], 1:1,000) and anti-TAP1 (clone 148.3 [30], 1:100) were used as primary MoAbs. To lower the staining of interfering immunoprecipitated heavy and light chains, mouse Trueblot™ ULTRA reagent (eBioscience, San Diego, CA, USA) was added. Protein complexes were visualized by Amersham ECL Western Blotting Detection Reagents (GE Healthcare, Buckinghamshire, UK).

Proteasome inhibition assays

MG-132 and bortezomib (PS-341, Velcade) were kind gifts from Dr. Gerrit Jansen (VU University Medical Center, Amsterdam, The Netherlands). Each of these inhibitors was added to 5 × 10⁵ cells/ml culture medium per well in increasing concentrations using six-well plates and cells were incubated overnight at 37°C in a humidified atmosphere containing 5% CO₂. Following incubation, cells were washed twice with PBS containing 0.1% HSA and 0.05% sodium azide and centrifuged for immunofluorescence staining.
Results

HLA-DR expression and transport to the plasma membrane in CLIP⁻ KG-1 blasts can occur in the absence of the invariant chain.

To study the li-dependency of HLA-DR expression by CLIP⁻ leukemic blasts, we silenced li expression in the KG-1 myeloid leukemic cell line, which was found to lack CLIP at the plasma membrane [23].

Western blot analysis demonstrated that li expression was strongly inhibited in li-siRNA-treated KG-1 (KG-1/li-siRNA) blasts (Fig. 1a). Interestingly, the total amount of HLA-DR in KG-1/li-siRNA blasts was similar to that in wild-type blasts. These results were confirmed by flow cytometry, which showed that li was down-regulated for >98% and, despite a very small population of HLA-DR⁻ li⁻ blasts, total HLA-DR expression remained unaffected in the vast majority of permeabilized, li-negative KG-1/li-siRNA blasts (Fig. 1b, left panels).

Since the formation of HLA-DR molecules was not impaired in KG-1/li-siRNA blasts, we next questioned whether their transport to the plasma membrane was disturbed by FACS analysis of HLA-DR expression on intact cells (Fig. 1b, right panel). Again, no differences in expression were encountered, implicating that the absence of li does not interrupt HLA-DR transport to the plasma membrane.

To assess if this li-independent HLA-DR expression resulted from an inability of li to interact with HLA-DR molecules, we immunoprecipitated li from KG-1 blasts and determined its association with HLA-DR by western blotting. As a positive control, the CLIP⁺ Kasumi-1 myeloid leukemic cell line was used, in which HLA-DR expression did depend on the function of li [23]. Immunoprecipitation of li from these cell lines revealed a similar association with HLA-DR (Fig. 1c). Thus, the observed li-independent HLA-DR expression in KG-1 blasts is not a reflection of inefficient binding of li to HLA-DR.

These findings show that in CLIP⁺ KG-1 blasts, HLA-DR processing and transport to the plasma membrane can occur independently of li expression, indicating the involvement of other proteins in CLIP⁻ leukemic blasts that stabilize HLA class II molecules.

HLA-DR expression and transport to the plasma membrane in CLIP⁻ KG-1 blasts is dependent on the function of the proteasome.

Newly formed HLA-DR molecules are typically stabilized by li (and later by CLIP) prior to peptide binding in the MIICs [31, 32]. This raises the question of which alternative mechanism stabilizes these molecules in cells that do not express li. We studied if li-independent HLA-DR expression in KG-1 blasts may be accomplished by loading of cytoplasmic peptides that are processed via HLA class I antigen-processing machinery.

For classical loading of HLA class I molecules, cytoplasmic antigens need to be cleaved into small peptides by the proteasome. To investigate the role the proteasome in HLA-DR peptide loading, we blocked proteasomal activity in KG-1 blasts by two inhibitors: MG-132, which inhibits all catalytic β subunits, and bortezomib, which specifically suppresses the β5 and, at higher concentrations, the β1 subunit of the 26S proteasome [33].

After incubation with each inhibitor, HLA class I expression was decreased on the total KG-1 blast population, as determined by flow cytometry (Fig. 2a), indicating the effect of proteasome inhibition on endogenous peptide loading. Interestingly, HLA-DR expression was strongly down-regulated as well, in a dose-dependent manner (supplementary material, Fig. S1a). In line with its broad catalytic subunit specificity, MG-132 had a stronger effect on both HLA class I and HLA-DR expression than bortezomib (Fig. 2b). Similar effects of proteasome inhibition were found for the myeloid leukemic cell line ME-1 (supplementary material, Fig. S1b). ME-1 blasts also lack CLIP at the plasma membrane [23], indicating that our findings reflect a general mechanism for leukemic blasts that do not express CLIP.

As CLIP is abundantly expressed on Kasumi-1 blasts [23], in contrast to the KG-1 and ME-1 myeloid leukemic cell line, it is likely that cytoplasmic antigens in these blasts are not processed for presentation by HLA-DR molecules. Indeed, MG-132 and bortezomib did not reduce plasma membrane expression of HLA-DR on Kasumi-1 blasts (Fig. 2c), while HLA class I expression was affected. This demonstrates that HLA-DR expression on these blasts does not involve proteasome activity and confirms that the
effect of both inhibitors in KG-1 and ME-1 blasts does not result from aspecific protein down-regulation.

In conclusion, these data indicate that, in contrast to CLIP\(^{+}\) Kasumi-1 blasts, the transport of HLA-DR molecules to the plasma membrane in at least a subpopulation of KG-1 blasts and ME-1 blasts relied on proteasomal activity, which supports the notion that Ii-independent HLA-DR expression in CLIP\(^{+}\) leukemic blasts is related to loading with cytoplasmic peptides.

HLA-DR expression and transport to the plasma membrane in CLIP\(^{+}\) KG-1 blasts is partly dependent on the function of the TAP molecule

To further provide evidence that HLA class I antigen-processing machinery is involved in cytoplasmic peptide loading of HLA-DR molecules, we suppressed TAP-mediated peptide processing in KG-1 blasts using two different viral proteins: BNLF2a, which interferes with peptide and ATP binding to TAP [25], and UL49.5, which prevents conformational changes of TAP required for peptide translocation and targets TAP for proteasomal degradation [26].

After transduction with BNLF2a/GFP- and UL49.5/GFP-encoding retrovirus, GFP\(^{+}\) KG-1 blasts were flow cytometrically sorted and analyzed for TAP expression by western blotting (Fig. 3a). Although BNLF2a has been described to only functionally suppress the TAP complex, a lower amount of TAP1 was observed in BNLF2a-containing KG-1 (KG-1/BNLF2a) blasts, as compared to wild-type blasts. In UL49.5-containing KG-1 (KG-1/UL49.5) blasts, TAP1 expression was completely down-regulated, which agrees with the function of UL49.5 in targeting TAP for proteasomal degradation. TAP2 expression was not affected by these inhibitors, indicating that the inhibition of TAP function was mediated by down-modulation of TAP1.

To test the impact of TAP inhibition on Ii-independent HLA-DR expression, we analyzed the level of HLA-DR expression on TAP-inhibited KG-1 blasts by flow cytometry (Fig. 3b). HLA class I levels were determined as a positive control for the function of TAP in classical endogenous peptide loading. GFP\(^{+}\) KG-1/BNLF2a and KG-1/UL49.5 blasts not only showed decreased HLA class I expression, but also a blast subpopulation with clearly reduced HLA-DR levels (total down-regulation of 21 and 36%, respectively) at the plasma membrane. This effect was the strongest in GFP\(^{+}\) KG-1/UL49.5 blasts (53% of GFP\(^{+}\) blasts obtained a DR\(^{−}\) phenotype), probably as the cause of higher UL49.5 levels implicated by increased GFP expression compared to KG-1/BNLF2a blasts (Fig. 3b) and the complete degradation of TAP (Fig. 3a). In CLIP\(^{−}\) ME-1 blasts, TAP inhibition by using UL49.5 retrovirus resulted in down-regulation of HLA-DR on the total blast population (supplementary material, Fig. S3).

To investigate whether TAP is also involved in Ii-dependent HLA-DR expression, CLIP\(^{+}\) Kasumi-1 blasts were transduced with UL49.5/GFP-encoding retrovirus as well. After flow cytometric sorting and gating of GFP\(^{+}\) blasts, no differences were found in plasma membrane expression of HLA-DR compared to wild-type blasts (Fig. 3c). The strong reduction in TAP1 (Fig. 3a) and HLA class I (Fig. 3c) expression showed that the function of TAP was indeed inhibited in Kasumi-1/UL49.5 blasts.

By different ways of TAP inhibition, we demonstrate that in contrast to CLIP\(^{+}\) Kasumi-1 blasts, HLA-DR transport to the plasma membrane is dependent on the function of TAP in a subpopulation of KG-1 blasts. This suggests that Ii-independent HLA-DR expression in CLIP\(^{−}\) leukemic blasts results from loading with cytoplasmic peptides processed by HLA class I antigen-processing machinery.

Ii expression is inversely correlated to the TAP-dependency of HLA-DR expression and transport to the plasma membrane in CLIP\(^{−}\) KG-1 blasts

Although HLA-DR expression was totally absent in a subpopulation of TAP-inhibited KG-1 blasts, the other subpopulation was still able to transport HLA-DR molecules to the plasma membrane. To explain this difference in effect, we flow cytometrically sorted the DR\(^{−}\) (TAP-dependent) and DR\(^{+}\) (TAP-independent) subpopulation of GFP\(^{+}\) KG-1/UL49.5 blasts. Plasma membrane expression of HLA class I was equally reduced on both subpopulations compared to wild-type blasts, as determined by flow cytometry (Fig. 4a). This indicated that UL49.5 protein levels and the degree of TAP inhibition did not differ between these subpopulations.

We next studied whether the differences in TAP dependency of HLA-DR expression by KG-1 blasts involved the function of Ii. Flow cytometric analysis revealed a close association between total HLA-DR and Ii expression in GFP\(^{+}\) KG-1/BNLF2a and KG-1/UL49.5 blasts, as well as in the DR\(^{−}\)- and DR\(^{+}\)-sorted subpopulations of GFP\(^{+}\) KG-1/UL49.5 blasts (Fig. 4b). During cell culture, both HLA-DR and Ii expression were up-regulated again in the
Fig. 3 TAP inhibition in KG-1 and Kasumi-1 blasts and the effect on HLA-DR expression. 

a KG-1 and Kasumi-1 blasts were transduced with retrovirus encoding GFP together with either BNLF2a or UL49.5 proteins, two different inhibitors of TAP. After flow cytometric sorting of GFP⁺ blasts, TAP1 and TAP2 expression were determined by western blotting. All lanes were equally loaded with protein, as β-actin was expressed equally. Molecular weights: TAP1, 71 kD; TAP2, 75 kD; β-actin, 42 kD.

b, c Flow cytometrically sorted GFP⁺ KG-1 (b) and Kasumi-1 (c) blasts containing either UL49.5 or BNLF2a were analyzed for GFP as well as plasma membrane HLA class I (‘ABC’) and HLA-DR expression (unfilled histograms) by flow cytometry. GFP and plasma membrane HLA class I levels were evaluated to assess the transduction efficiency and the degree of TAP inhibition, respectively. The filled histograms represent stainings of the same cells with control IgG.
DR\(^-\)-sorted subpopulation, while the DR\(^+\)Ii\(^+\) phenotype remained stable in the DR\(^+\)-sorted subpopulation of GFP\(^+\) KG-1/UL49.5 blasts (data not shown).

Thus, in one KG-1 blast subpopulation, TAP suppression accompanied an unexpected, transient decline in Ii amount, probably resulting in the total absence of HLA-DR. In another subpopulation of KG-1 blasts, the expression of Ii was not affected by TAP inhibition, which indicated that HLA-DR expression in these blasts is rescued by association with Ii.

TAP and Ii together form the key mediators of HLA-DR expression and transport to the plasma membrane in CLIP\(^-\) KG-1 blasts.

To actually show the capacity of Ii in rescuing HLA-DR expression in TAP-inhibited KG-1 blasts, we retrovirally transfected the DR\(^+\)-sorted subpopulation of GFP\(^+\) KG-1/UL49.5 (KG-1/UL49.5/DR\(^+\)) blasts with Ii-siRNAs. In agreement with the flow cytometric data (Fig. 4b), total HLA-DR expression levels in GFP\(^+\) KG-1/UL49.5/DR\(^+\) blasts were slightly increased in comparison to those in wild-type blasts, as determined by western blotting (Fig. 5a). This indicates that in the absence of TAP, the association of HLA-DR molecules with Ii enhances their stability. The total amount of HLA-DR strongly declined after Ii silencing compared to non-silenced GFP\(^+\) KG-1/UL49.5/DR\(^+\) and wild-type blasts. We confirmed these results by flow cytometry (Fig. 5b; supplementary material, Fig. S4), showing similar reduction in HLA-DR expression after >90% Ii down-regulation in GFP\(^+\) KG-1/UL49.5/DR\(^+\) blasts. This was contrary to the observations in KG-1/Ii-siRNA blasts with the TAP complex still functional (Fig. 1b).

Thus, TAP-independent HLA-DR expression and transport to the plasma membrane in KG-1 blasts is confined to the function of Ii, consistent with the classical pathway of HLA class II antigen presentation.
To study if the function of TAP is also the determining factor in Ii-independent HLA-DR expression, we transduced KG-1/Ii-siRNA blasts with BNLF2a/GFP or UL49.5/GFP retrovirus and flow cytometrically sorted GFP\(^+\) blasts. The inhibition of TAP resulted in a strong down-regulation of total and plasma membrane expression of HLA-DR in both GFP\(^+\) KG-1/Ii-siRNA/BNLF2a and KG-1/Ii-siRNA/UL49.5 blasts (Fig. 6). The difference in effect of TAP inhibition on HLA-DR expression could be explained by the higher transduction efficiency of UL49.5/GFP compared to BNLF2a/GFP retrovirus (data not shown), as noticed earlier in wild-type blasts (Fig. 3b).

In accordance with the requirement of Ii in TAP-independent HLA-DR expression, the function of TAP is critical for Ii-independent HLA-DR expression and transport to the plasma membrane in KG-1 blasts. Overall, simultaneous inhibition of TAP- and Ii-dependent antigen-processing pathways in these blasts strongly down-modulates HLA-DR expression, showing that TAP and Ii together are the key regulators of HLA class II antigen presentation.

**Discussion**

For an effective anti-tumor immune response, CTLs and CD4\(^+\) T cells need to be stimulated with TAAs presented by tumor APCs [34]. Professional APCs present these
peptides after recognition, engulfment and processing of exogenously derived TAAs. Certain tumor cells, however, can also serve as potent APCs by not only expressing costimulatory molecules, but also presenting their own peptides to T cells. Since impaired TAA presentation is a known tumor escape mechanism [35], the components involved in processing and presentation of endogenous TAAs in tumor cells are interesting targets for immunomodulation. Previously, it has been reported that the CLIP− KG-1 myeloid leukemic cell line is able to stimulate T cell reactivity and presents HLA-DR molecules that are loaded with peptides from endogenous rather than exogenous sources [36, 37]. In this report, we show that in addition to classical Ii-dependent HLA-DR antigen presentation, KG-1 blasts use HLA class I antigen-processing machinery, including the proteasome and TAP complex, to process endogenous antigens for presentation via HLA-DR molecules.

The Ii-independency of HLA-DR expression in KG-1 blasts (Fig. 1a, b) is not unique to this myeloid leukemic cell line, as we recently found the same in the THP-1 myeloid leukemic cell line treated with Ii-siRNAs [23]. Although Ii plays an essential role in targeting HLA-DR into the endosomal/lysosomal pathway [7], the components involved in processing and presentation of endogenous TAAs in tumor cells are interesting targets for immunomodulation. Previously, it has been reported that the CLIP− KG-1 myeloid leukemic cell line is able to stimulate T cell reactivity and presents HLA-DR molecules that are loaded with peptides from endogenous rather than exogenous sources [36, 37]. In this report, we show that in addition to classical Ii-dependent HLA-DR antigen presentation, KG-1 blasts use HLA class I antigen-processing machinery, including the proteasome and TAP complex, to process endogenous antigens for presentation via HLA-DR molecules.

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loaded with proteasome-generated and TAP-translocated peptides.

The difference in TAP dependency of HLA-DR expression within the total KG-1 blast population might be explained by a differential expression of specific HLA-DR alleles with a high affinity for endogenous peptides. It was reported that two allelic subtypes, DRB1*1101 and DRB1*1401, on KG-1 blasts predominantly presented endogenous peptides [36]. In addition, HLA-DR4 and -DR7 molecules have been shown to associate with endogenous peptide presentation on tumor cells [42, 43]. Unexpectedly, BNLF2a or UL49.5 proteins abrogated the expression of Ii in the DR\(^{-}\) KG-1 blast population (Fig. 4b), but were recovered again during culture (data not shown). The suppression of TAP might therefore transiently elicit additional effects on the HLA class II-restricted presentation pathway in this subset of blasts, as we found reductions in HLA-DM expression levels as well (data not shown). This suggests that the master regulator of the HLA class II presentation pathway, CIITA, might be affected, which is currently under investigation.

By simultaneous inhibition of Ii and TAP, we were able to discriminate between the Ii-dependent and Ii-independent HLA class II presentation pathway of endogenous peptides in these tumor cells. We found that in the DR\(^{+}\) subpopulation of TAP-inhibited KG-1 blasts, only the Ii-dependent pathway was functional, as Ii silencing strongly reduced expression levels of HLA-DR (Fig. 5a). This seems to be a threshold effect as the amount of HLA-DR expressed is not directly proportional to the quantity of Ii (Fig. 5b). In KG-1/Ii-siRNA blasts, HLA-DR expression was clearly down-modulated after TAP suppression (Fig. 6), showing that the Ii-independent pathway depends on the function of TAP. The inability to express HLA-DR molecules in the absence of both Ii and TAP function agreed with the DR\(^{-}\) phenotype found for the TAP-deficient KG-1 subpopulation lacking Ii expression (Fig. 4b).

This study provides important insights into the intracellular mechanisms of tumor cells to process endogenous antigens for HLA class II-restricted presentation. Extensive interplay between HLA class I and II presentation pathways was shown in blasts of the CLIP\(^{-}\) KG-1 myeloid leukemic cell line, which constitutively express HLA class II molecules loaded with endogenous peptides. These findings indicate that CLIP\(^{-}\) leukemic blasts may be able to present tumor-associated peptides on both HLA class I and II molecules and underlines the hypothesis that abnormalities in the HLA class I antigen-processing machinery in certain tumor cells may result in tumor escape from both HLA class I- and HLA class II-restricted T cell recognition. Besides their potential as antigen presenters, KG-1 blasts can also be differentiated into functional DC-like cells [37, 44], which makes these tumor cells ideal candidates for the use as cellular vaccines. In general, the involvement of endogenous antigen processing in HLA class II presentation by tumor cells might introduce new immunomodulatory approaches to simultaneously induce HLA class I and II tumor antigen presentation.

Acknowledgments This work was financially supported by Stichting Vanderes (grant 125), the Dutch Cancer Foundation (RUL 2005-3259 and NKI 2001-2415), the Netherlands Scientific Organization (Vidi 917.76.330) and the Landsteiner Foundation for Blood Research (grant 0533).

Conflict of interest The authors declare that they have no conflict of interest.

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