A pregnant Japanese woman returning from Africa with recurrent fevers

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Abstract: Certain clinical aspects of vivax malaria are no longer defined as benign. We present a case of vivax malaria with three relapses in a pregnant Japanese woman who had returned to Japan from the Comoros Islands in East Africa. Data on the successful delivery, examination of Duffy-blood group antigen, and microscopic findings of growing stages of Plasmodium vivax are thought to be of considerable interest.

Keywords: vivax malaria, Duffy-blood group, pregnancy

Introduction

There is growing evidence that the impact of Plasmodium vivax has been underestimated in developing and poor countries.1,2 The difficulty of differential diagnosis and lack of proper treatment of both blood-stage and latent parasites are two of the reasons for the prevalence of vivax malaria.

Case report

A 28-year-old Japanese woman who had been married to an African man and lived in the Comoros Islands for 1 year was admitted to our hospital in Japan because of chills and severe headache. She was 30 weeks pregnant and had taken medication against malaria 2 months previously when she was in Africa.

Laboratory findings showed leukocyte (6,600/µL) and platelet (6.6 × 104/µL) counts, as well as hemoglobin (9.7 g/dL), C-reactive protein (CRP; 19.2 mg/dL), fibrin/fibrinogen degradation products (FDP) (35.1 µg/mL), and FDP D-dimer (38.5 µg/mL) levels. Giemsa staining of peripheral blood using pH 7.4 buffer showed that 1.8% of the erythrocytes were infected by Plasmodium. After the ring form, schizont, and gametocyte of this parasite were determined (Figure 1), the results of the subsequent rapid diagnostic test and DNA assay led to a diagnosis of vivax malaria.

Seven-day medication (quinine 1500 mg/day) against malaria, administered under strict observation and with the patient’s agreement, led to remission. However, vivax malaria relapsed in the 34th week of pregnancy. The presence of latent parasites was suspected to be the cause of the fevers. The next quinine administration resulted in another remission. To exterminate the latent parasites, we assessed mother and child blood glucose-6-phosphate dehydrogenase (G6PD) activities after 39th week spontaneous vaginal delivery, after which she was treated again with primaquine (15 mg/day; 14 days). The new born baby’s weight was 2850 g. There were no
abnormalities in either the baby or the placenta. There was no mother-to-child transmission. Mother and child are healthy after 6 months of anti-relapse therapy.

Discussion

Four distinct Plasmodium species are known to regularly infect humans: P. falciparum, P. vivax, P. malariae, and P. ovale. Certain clinical aspects of vivax malaria are not defined as benign anymore because relapse, severe clinical cases, and drug resistance have been reported for P. vivax.\(^1\)\(^-\)\(^3\)

Relapse of P. vivax and P. ovale is known to be triggered by dormant hypnozoites in the liver.\(^2\)

Plasmodium falciparum malaria is defined as malignant and is a major public health problem in Africa, including the Comoros Islands,\(^4\) while vivax malaria is the major cause of malaria outside Africa, mainly afflicting Asia and the Americas.\(^2\)

Why would vivax malaria be less prevalent in Africa? It is known that Africans with Duffy-blood group antigen negative erythrocytes cannot be infected by P. vivax,\(^5\) because it requires the Duffy-blood group antigen as an obligate receptor for invasion,\(^6\) and the Duffy-blood group antigen negative phenotype Fy(a\(^-\) b\(^-\)) frequently occurs in the native African.\(^7\)\(^,\)\(^8\)

On the other hand, treatment of symptomatic vivax malaria during pregnancy is very difficult. It may lead to preterm delivery and fetal loss, and neonates of non-immune mothers may be at a particular risk of congenital malaria resulting from transplacental passage of parasites.\(^9\)

Quinine as well as chloroquine are the drugs of choice for the treatment of vivax malaria. Primaquine, the only therapeutic option for these dormant tissue forms, is contraindicated during pregnancy because of the risk of fetal hemolysis.\(^10\)

Even more noteworthy, low-birth weight associated with vivax malaria has also been reported during second and subsequent pregnancies.\(^11\)

We reported here repeated recurrences of vivax malaria in a Japanese primigravida from the Comoros Islands. Since the Duffy-blood group antigen of our patient was Fy(a\(^+\) b\(^-\)), our findings support the notion that we should pay careful attention to the occurrence of P. vivax even in Africa,\(^12\) in addition to the numerous local Duffy-blood group antigen-negative populations.

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Disclosure

The authors report no conflicts of interest in this work.

References

1. Price RN, Tjitra E, Guerra CA, et al. Vivax malaria: neglected and not benign. Am J Trop Med Hyg. 2007;77(Suppl 6):79–87.
2. Carlton JM, Adams JH, Silva JC, et al. Comparative genomics of the neglected human malaria parasite Plasmodium vivax. Nature. 2008;455(7214):757–763.
3. Argawal R, Nath A, Gupta G. Noninvasive ventilation in Plasmodium vivax related ALI/ARDS. Intern Med. 2007;46:2007–2011.
4. Rebaudet S, Bogreau H, Silai R, et al. Genetic structure of Plasmodium falciparum and elimination of malaria, Comoros archipelago. Emerg Infect Dis. 2010;16(11):1686–1694.
5. Chaudhuri A, Polyakova J, Zbrzezna V, et al. Cloning of glycoprotein D cDNA, which encodes the major subunit of the Duffy blood group system and the receptor for the Plasmodium vivax malaria parasite. Proc Natl Acad Sci U S A. 1993;90(22):10793–10797.
6. Michon P, Fraser T, Adams JH. Naturally acquired and vaccine-elicited antibodies block erythrocyte cytoadherence of the Plasmodium vivax Duffy binding protein. Infect Immun. 2000;(6896):3164–3171.
7. Matusuoka H, Hasegawa M, Ishii A, et al. A survey of malaria, glucose-6-phosphate dehydrogenase deficiency and Duffy blood group in six localities in Guatemala. J J Trop Med Hyg. 1988;16;1–9.
8. Howes RE, Patil AP, Piel FB, et al. The global distribution of the Duffy blood group. Nat Commun. 2011;2:266.
9. ter Kuile FO, Rogerson SJ. Plasmodium vivax infection during pregnancy: an important problem in need of new solutions. Clin Infect Dis. 2008;46:1382–1384.
10. Baird JK. Neglect of Plasmodium vivax malaria. Trends Parasitol. 2007;23(11):533–539.
11. Nosten F, McGready R, Simpson JA, et al. Effects of Plasmodium vivax malaria in pregnancy. Lancet. 1999;354(9178):546–549.
12. Mizuno Y, Kudo K, Kano S. Difficulty of proper diagnosis for an imported vivax malaria patient from Africa. Kansenshogakai Zasshi. 2007;81(5):597–599. Japanese.
Recurrent fevers in a pregnant woman returning from Africa