Quantifying male and female pheromone-based mate choice in *Caenorhabditis* nematodes using a novel microfluidic technique

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Abstract

Pheromone cues are an important component of intersexual communication, particularly in regards to mate choice. *Caenorhabditis* nematodes predominant rely on pheromone production for mate finding and mate choice. Here we describe a new microfluidic paradigm for studying mate choice in nematodes. Specifically, the Pheromone Arena allows for a constant flow of small molecule signals to be passed in real time from signaling worms to those making a choice without any physical contact. We validated this microfluidic paradigm by corroborating previous studies in showing that virgin *C. remanei* and *C. elegans* males have a strong preference for virgin females over mated ones. Moreover, our results suggest that the strength of attraction is an additive effect of male receptivity and female signal production. We go on to explicitly examine female choice and find that females are more attracted to virgin males. However, a female’s mate choice is strongly dependent on her mating status.

Introduction

A critical component of sexual reproduction is the ability to find and recognize the appropriate individual with which to mate. Individuals must be able to distinguish members of their own species – namely, conspecifics – from those of other species – heterospecifics. Perhaps equally important, is the ability to choose high quality individuals that are receptive to mating. The process of mate choice is shaped by sexual selection and relies on communication between the sexes [1-3]. In particular, sex pheromones – small chemicals produced by a signaler to induce a sexual response in a receiver – are a major means of intersex communication across a wide variety of both invertebrate and vertebrate taxa [reviewed in 4]. Some of the best studied sex
pheromones are the cuticular hydrocarbon family found in many insect species [5,6]. These pheromones have both species-specific and sex-specific effects [6]. For example, in *Drosophila* female hydrocarbons attract males, while male hydrocarbons have an anti-aphrodisiac effect on other males and increase female receptivity [7-9]. While these studies highlight the importance of pheromones in mate choice, many of the taxa studied have additional behaviors and traits that contribute to the mate choice process, potentially confounding the relative reliance on pheromone-based cues. *Caenorhabditis* nematodes rely almost exclusively on pheromone signals for mate choice [10,11]. Pheromone signals in *Caenorhabditis* have traditionally been studied using plate-based chemotaxis assays, where male attraction is quantified based on his ability to discriminate between a control medium and a female-conditioned medium [12,13]. In particular, these studies have identified ascaroside pheromones as important in mate choice signaling due to their sex-specific production and effects on sexually-associated behaviors [14,15]. Female pheromones act as a male attractant in both hermaphroditic *C. elegans* [12] and gonochoristic *C. remanei* [16]. This female-based signaling appears to be related to the amount of sperm stored [16-18] and targets male-specific neurons [13,14,16,19,20]. Recent work has turned to male-produced ascarosides, showing that hermaphrodites exposed to male pheromones alone have a decrease in lifespan [21-24]. However, such male-produced pheromones do not appear to elicit a female mate choice response [12,16,22]. While ascarosides play a predominant role in mate choice, they also influence other population behaviors, thus necessitating a precise combination and concentration of small molecules for signaling specific to male-female interactions [11,14]. Since pheromone cues depend on such a precise mixture, accurate intersex communication
assays necessitate a well-controlled environment where the concentration and diffusion of molecules is clearly defined and external signals are limited.

Microfluidic technology has proved to be an excellent method in which to study behavioral responses under precise environmental control [25-27]. Microfluidic devices scale on the nano- to micro-size and thus the small size of Caenorhabditis makes them suitable for manipulation within a microfluidic environment. Microfluidics offers many advantages over traditional plate-based assays, including better control of the concentration of molecules and their diffusion due to the laminar properties of microfluidics [28]. Several microfluidic devices have been designed to study behavioral responses such as chemotaxis, thermotaxis, and electrotaxis [25-27,29,30]. With respect to reproductive behavior, Chung et al. [29] found a behavioral response of individually isolated males when exposed to a uniform concentration of hermaphrodite-conditioned medium. In particular, they showed that males exposed to pre-conditioned medium spent more time performing sexually-associated behaviors than those exposed to a control medium. While their microfluidic device overcomes the issues of pheromone diffusion on agar plates, it cannot be used to study mate-choice searching patterns [as in 20,31] or direct choice comparisons between different attractants. Additionally, the pheromone signal contained in the pre-conditioned media is likely to decrease over the time required to study such locomotion patterns. Moreover, conditioned medium may be difficult to accurately reproduce as the concentration of small molecules will depend on the density of worms used to produce pheromones as well as the time spent in liquid culture. Given the limitations of exposure to a single pre-conditioned medium, the relationship between a pheromone signal and the elicited sexual response should be studied in real time. Given the
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We describe a new microfluidic paradigm, using the Pheromone Arena microfluidic
device, that overcomes the current limitations of traditional plate-based assays and existing
microfluidic technology to study sex-specific mate choice with constant exposure to pheromones
produced in real time. We show that the Pheromone Arena allows for small-molecule
communication alone without a decay of signal over time, thus validating its use for mate choice
assays. Using the Pheromone Arena, we show that males are more attracted to virgin females
than mated ones in both *C. remanei* and *C. elegans* with the degree of attraction being species-
dependent. Additionally, we show that females rely on pheromone cues, though their preference
is dependent on mating status.

**Materials and Methods**

**Worm culture and strains**

Wildtype *Caenorhabditis remanei* (strain EM464) and feminized *C. elegans* (strain
JK574: fog-2 mutation on the standard N2 laboratory background) were used in this study.
Feminization of *C. elegans* is achieved by blocking self-sperm production in hermaphrodites,
making them functionally female, and they will be referred to thusly. Use of a feminized *C.
elegans* hermaphrodites allowed for direct comparisons between species as well as preventing
any potential mate cue effects due to self-sperm [see 17,18]. Both strains were grown at 20°C on
NGM-agar plates seeded with OP50 *Escherichia coli* bacteria following Brenner [32].
Synchronized cultures of stage 1 larvae were prepared by hypochlorite treatment of gravid
females [33]. Larvae were matured to young adulthood in population densities of approximately
1,000 individuals. To maintain virgins, males and females were separated onto sex-specific
plates of 40-50 individuals 40-45 hours post-larval stage 1. Mating plates of 25 females and 25
males were created at the same time. At the start of all the choice assays, day 1 adult virgins
were 48 hours post-larval stage 1 and day 2 adults were 72 hours post-larval stage 1 (24 hours
after separation to virgin or mating plates).

**Microfluidic device manufacturing**

The Pheromone Arena (final design: v2.1; S1 File) was designed using CAD software
(Vectorworks 2013 SP5, Nemetschek Vectorworks, Inc). Single layer devices were fabricated
out of polydimethylsiloxane (PDMS) following soft lithography methods [34] and bonded to a
glass microscopy slide following exposure to air plasma. Holes for connecting tubing were
punched using a 1.25mm biopsy punch.

**Microfluidic set-up and experimental protocol**

To avoid blocking flow, air bubbles were evacuated from each device using a vacuum
chamber and replaced with M9 buffer. Worms were loaded into three chambers: ten worms each
were loaded into the two upstream “signaling” chambers of the device and 20 to 25 worms were
loaded into the single downstream “choice” chamber. To control for environmental and
observational biases, worm combinations were alternatively loaded into the left and right
upstream chambers.

Liquid was flowed through all inlets continuously, with the start of flow corresponding to
the beginning of an experiment. Flow was maintained at a constant rate using a pressurized air
system (S1 Fig.). Specifically, air exiting a pressurized one gallon tank was regulated to 1.5 PSI.
The air-line running from the tank was bifurcated to two tubing lines, each pressurizing a sealed
500mL bottle of M9. The bottle caps were modified to hold seven pieces of tubing: one for the
air-line (terminating at the cap) and six liquid supply lines. The liquid tubing lines extended below the M9 surface, allowing liquid to flow out of the bottle and into the microfluidic device once air pressure was applied from the air-line. Tubing lengths were equal for each partition of the set-up to maintain equal flow through all lines. Pressure could be maintained for three hours off a single air tank.

The head position of each worm in the downstream chamber was counted every 30 minutes and recorded as being located under the left or right upstream chamber. Worms located in the filter separators were not counted. If worms were flushed out of the downstream chamber (and therefore the device), the assay continued without counting these worms and thus some replicates had a decrease in sample size over time. If worms climbed or were flushed into a different chamber than the one into which they were loaded, the experiment was terminated.

Each choice combination was replicated multiple times \( (n \geq 3) \) over multiple days \( (n \geq 2) \). Day 2 adult worms were used for the male choice assays. Day 1 adult worms were used for female choice assays, except when comparing virgin versus mated female choice, which required using day 2 adult females. All data have been made available (S2 File).

**Statistical analyses**

Data were analyzed in R v3.2.1 (R Core Development Team 2015). Replicates were pooled for each choice assay by time point. An equality of proportions test was performed for each assay individually to determine if: i) males were more attracted to virgin females over mated females or ii) females were more attracted to virgin males over virgin females. The null hypothesis was that of no choice (using a probability of success = 0.5). A chi-square test for homogeneity was performed to determine if the proportion of males or females choosing virgin females or males, respectively, was equal across species combinations.
Results

Microfluidic design

The Pheromone Arena has three sequential components: three inlets with each with a distribution loading network, a main arena, and a single high resistance outlet (Fig. 1A). The arena area is further divided into three physically distinct chambers. The chambers all have a pillar-array to facilitate the natural, sinusoidal movement of worms [35,36]. The two upstream chambers hold the pheromone signalers. Two versions of the loading distribution networks were created to accommodate the size differences between males and females at day 2 of adulthood. Specifically, female signalers have a wider distribution network, while male signalers have a narrower distribution network coupled with the removal of the first two rows of pillars to prevent males from climbing out of the chamber (Fig. 1B). The upstream chambers are separated from the single, large downstream chamber by a very fine filter (Fig. 1C). This filter allows for small molecules to pass, but rarely can worms pass through. Worms making a choice were loaded into the downstream chamber by a third inlet. Due to laminar flow, two pheromone environments are created in the downstream chamber that mirror the signaling pair in the upstream chambers (Fig. 1D). All the chambers were greater than one-by-one worm length, allowing for free movement of individuals without density effects (Fig 1E).

No inherent biases were measured in the movement of worms in the downstream chamber. In particular, when no pheromone cue was present, males were equally likely to move to the left of right sides of the chamber in a random fashion (p = 0.45, n = 24 replicates).

Fig 1. Microfluidic Pheromone Arena for mate choice assays.
(A) Blueprint for the Pheromone Arena (v2.1; S1 File). The three inlets correspond to the three worm chambers: inlets 1 and 2 connect to the upstream chambers and inlet 3 connects to the downstream chamber. All the chambers have a pillar-array (shown as circles) spaced 100um apart to allow for natural worm movement. An 18um filter separates the downstream chamber from the upper two chambers and from the outlet. Extra resistance was added to the outlet distribution to decrease the overall flow rate. (B) Close-up of the loading distribution networks for the upstream chambers. The distribution network used depended on whether males or females were the pheromone signalers. (C) Close-up of the filter separating chambers. (D) Visualization of how the laminar flow dynamics create two distinct environments in the downstream chamber using red and blue dyes. (E) Visualization of worms in the device. After being loaded, worms move freely throughout their chamber without passing into another chamber.

**Males are attracted to virgin females**

Virgin *C. remanei* and *C. elegans* males were assayed for their ability to discriminate between conspecific virgin and mated females. Male choice was measured approximately every 30 minutes for 3 hours. Across all replicates, male choice varied little after 60 minutes (S2 Fig.). Therefore, we used data from this time point as a single comparative measure of male choice across assays.

Males were more attracted to virgin females than mated females in both *C. remanei* (proportions test: $\chi^2 = 46.2$, d.f. = 1, $p < 0.0001$, 95% C.I. of virgin attraction = 72.6-87.1%) and *C. elegans* (proportions test: $\chi^2 = 4.15$, d.f. = 1, $p = 0.04$, 95% C.I. of virgin attraction = 50.3-64.8%), though the ability to discriminate was much weaker in *C. elegans* (Fig. 2). To better understand this marked difference in male sensitivity, male choice was assayed for the ability to
discriminate between virgin and mated heterospecific females. In both heterospecific crosses males were more attracted to virgin females than mated females (C. remanei male proportions test: $\chi^2 = 42.0$, d.f. = 1, $p < 0.0001$, 95% C.I. of virgin attraction = 66.1-78.7%; C. elegans male proportions test: $\chi^2 = 24.2$, d.f. = 1, $p < 0.0001$, 95% C.I. of virgin attraction = 64.1-80.9%). For example, C. remanei males chose virgin C. elegans females more successfully than in the conspecific C. elegans assay. Similarly, C. elegans males had a much high ability to discriminate virgins when they were presented with C. remanei females. The strength of attraction to virgins for both heterospecific assays was between that of the conspecific assays, suggesting species-dependent pheromone effects in both males and females ($\chi^2 = 22.0$, d.f. = 3, $p < 0.0001$).

**Fig 2. Males are more attracted to virgin females than mated females within and between species.**

Virgin, day 2 adult C. remanei males (blue) and C. elegans males (orange) were given a choice between virgin and mated female pheromone (C. remanei females shown as circles and C. elegans females shown as triangles). Each replicate is represented as an individual point and the mean attraction to virgin females is given by the horizontal bar. Conspecific assays are shown as solid point and heterospecific assays as open points. The null hypothesis of no choice is given by the dashed line. In each assay males were more attracted to virgin females than mated ones. However, the strength of male attraction was dependent on the species of both the chooser and the signaler (Test of homogeneity across assays: $\chi^2 = 22.0$, d.f. = 3, $p < 0.0001$). Asterisks denote a significant proportions test.

**Females choose male pheromone over those of from females**
Virgin *C. remanei* and *C. elegans* females were assayed for their ability to discriminate between conspecific virgin males and females. Female choice increased slightly over time, but was consistent by 60 minutes, again making this time point a reflective measure of overall female choice (S3 Fig.). Interestingly, *C. remanei* and *C. elegans* followed a similar trend in choice of males over time, though at very different magnitudes. Specifically, female *C. remanei* chose conspecific male pheromones over female pheromones (proportions test: $\chi^2 = 14.8$, d.f. = 1, $p < 0.001$, 95% C.I. of male attraction = 58.0-74.0%) (Fig. 3). When females were given a choice between male pheromone and no pheromone, they still chose the male pheromone more than expected by chance ($\chi^2 = 7.19$, d.f. = 1, $p < 0.01$), suggesting that females are in fact attracted to males and not simply repulsed by other females. However, *C. elegans* females showed no clear differentiation between conspecific males and females (proportions test: $\chi^2 = 0.563$, d.f. = 1, $p = 0.45$, 95% C.I. of male attraction = 45.0-61.8%). Moreover, the heterospecific assays also showed a lack of discrimination between male and female pheromones, suggesting that female choice may be species-specific, or at least very weak at best within *C. elegans*.

**Fig 3. Female choice of virgin males over virgin females is species-specific.**

Virgin, day 1 adult *C. remanei* females (blue) and *C. elegans* females (orange) were given a choice between virgin male and female pheromone (*C. remanei* females shown as circles and *C. elegans* females shown as triangles). Each replicate is represented as an individual point and the mean attraction to males is given by the horizontal bar. Conspecific assays are shown as solid point and heterospecific assays as open points. The null hypothesis of no choice is given by the dashed line. Only when *C. remanei* females were given a choice of conspecifics were they more attracted to male pheromone than
female pheromone \( (\chi^2 = 14.8, \text{ d.f.} = 1, p < 0.001) \). All other comparisons failed to reject the null hypothesis of no female choice. Asterisks denote a significant proportions test.

Additionally, we examined if female choice was dependent on a female mating status. The female choice assay was replicated using mated and virgin \textit{C. remanei} females. Virgin females showed a strong preference for virgin male over female pheromones (proportions test: \( \chi^2 = 17.5, \text{ d.f.} = 1, p < 0.0001, 95\% \text{ C.I. of male attraction} = 60.3\text{-}69.4\% \)). However, mated females showed no obvious preference (proportions test: \( \chi^2 = 0.150, \text{ d.f.} = 1, p = 0.70, 95\% \text{ C.I. of male attraction} = 38.0\text{-}57.5\% \)).

\textbf{Fig 4. Female attraction depends on mating status.}

Virgin (light purple) and mated (dark purple) day 2 adult \textit{C. remanei} females were given a choice between virgin day 1 adult \textit{C. remanei} male and female pheromones. Each replicate is represented as an individual point and the mean attraction to males is given by the horizontal bar. The null hypothesis of no choice is given by the dashed line. Virgin females showed a strong preference for males over females \( (\chi^2 = 17.5, \text{ d.f.} = 1, p < 0.0001) \), while mated females displayed no choice \( (p = 0.70) \). Asterisks denote a significant proportions test.

\section*{Discussion}

Mate choice is ubiquitous across metazoans with sexual reproduction. Understanding the signal-receiver dynamics of mate choice beyond phenotypic traits – such as pheromone signals – provides valuable information on how individuals discern high quality mates with a propensity to mate. Here we proposed a new microfluidic paradigm to quantify pheromone communication in
nematodes. Our design allows for real time isolation of small molecules in a controlled environment as well as allowing for the natural searching and choice behaviors of receivers over time. While this design is not the first to use pillared-arenas [35-37], to the best of our knowledge no other worm-specific microfluidic devices have such a physically separated, sequential arena design. Moreover, the Pheromone Arena expands on previous worm choice devices [29,30] by allowing for natural searching behaviors in addition to measuring overall choice. Specifically, we examined male and female mate choice in *C. remanei* and *C. elegans* using a combination of conspecific and heterospecific assays to determine how mating status affects attraction. This study is the first use this combinatorial design coupled with real time pheromone signaling and spatial complexity.

The conspecific male choice results support previous studies [16,18] in showing that virgin males are strongly attracted to virgin females over mated females in both *C. elegans* and *C. remanei*. Interestingly, males from these species did not discriminate between female mating types with the same intensity [16]. In particular, *C. elegans* males had a reduced ability to discern virgin females from mated ones. However, when *C. elegans* males were presented with pheromones from *C. remanei* females, male attraction to virgins increased. Similarly, *C. remanei* males could distinguish between virgin and mated *C. elegans* females better than *C. elegans* males. Therefore, there appears to be a decrease in both female signal intensity as well as male receptor capability in *C. elegans*, leading to an overall decrease in mate choice ability. This diminution of choice is likely a result of the independent lineage transition to self-fertilizing hermaphroditism in *C. elegans*. Since fertilization is predominantly by selfing and males are rare within populations, sexual selection – apparently including mate recognition dynamics – is greatly reduced.
This hypothesis could be further tested by altering the number of signalers in the upstream chambers. The pheromone arena allows for exact control over the number of worms producing pheromone and, given the constant flow dynamics, altering the number of worms would in effect alter the concentration of pheromone signal in the downstream chamber.

Previous studies have used various concentrations of hermaphrodite-conditioned media and found different results in male attraction [16,18]. Our assays used the same number of females in both upstream chambers, however, modulating the number of females in each upstream chamber is promising future work.

Previous work has shown that females are attracted to isolated, male-produced ascaroside cues [15,38], however, this result has not been replicated when the signal is produced in vivo. We took a novel approach comparing female discrimination between male and female produced pheromones to determine if females are truly attracted to males or are simply repulsed by the presence of a high number of other females. A discernable choice of males over females was measured in the C. remanei conspecific assays. Additionally, females were attracted to male pheromone over a no pheromone control, suggesting this choice measured is true attraction to males. Despite making a choice, the intensity with which females choose males was much weaker than seen for the male choice assays. This weak attraction could potentially explain why previous plate-based assays – where male signals can be lost by diffusion or mixed with other signals from the environment – could not measure any female choice of males [13,16]. However, C. elegans females made no choice in the conspecific assays as was also seen for both heterospecific assays, suggesting species-specific effects. Together these results suggest that when sexual selection is strong, as in C. remanei, males take a more active searching approach, such that females are predominantly signalers, while males are active receivers and searchers.
This signal-receiver dynamic is in somewhat of a sex-role reversal from traditional sexual selection models of male signaling and female receiving, though still consistent with anisogamy. We further examine female choice based on a female’s mating status. Mating status is known to influence remating behavior in many species, such that mated females – or females with sperm – typically have a lower propensity to remate [39,40]. Moreover, mated females will run away from males to avoid remating [41]. These observations are consistent with our results as virgin females strongly preferred male pheromones while mated females made no choice between male and female pheromones.

While the Pheromone Arena is both an innovative and effective tool, there are several limitations to its use. Namely, the difference in size between older adult males and females poses a problem for intersex comparisons. Additionally, the design could be improved by limiting worms from reaching other chambers through the filter separators. This cross-chamber movement was particularly an issue with males as they have a small diameter and a highly-developed searching behavior that leads them to attempt to crawl through the filters to reach the virgin females. However, the filter between the upstream and downstream chambers is currently at the lower limit of what can accurately be manufactured using PDMS-based microfluidics and thus a significant design change would be required to prevent this tenacious behavior.

Despite these limitations, the Pheromone Arena was able to reproduce previously seen sexual behavior responses and go further into the study of species-specific sexual attraction in both males and females. In the future, this type of device could be used to study the effect of density or sex ratio on sexual attraction. Moreover, it would be possible to modify these devices to allow food delivery and perform longer term assays or study the influence of food availability on sexual attraction. Such assays would benefit from being coupled with an automated system.
[see 42] to obtain a more accurate counting via recordings and to facilitate high-throughput experiments.

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References

1. Andersson M, Simmons LW. Sexual selection and mate choice. Trends in Ecology & Evolution. 2006;21: 296–302. doi:10.1016/j.tree.2006.03.015

2. Jones AG, Ratterman NL. Mate choice and sexual selection: what have we learned since Darwin? Proc Natl Acad Sci USA. National Acad Sciences; 2009;106 Suppl 1: 10001–10008. doi:10.1073/pnas.0901129106

3. Andersson MB. Sexual selection. Princeton University Press; 1994.

4. Wyatt TD. Pheromones and animal behavior: communication by taste and smell. Cambridge University Press; 2003.

5. Howard RW, Blomquist GJ. Chemical ecology and biochemistry of insect hydrocarbons. Annu Rev Entomol. 1982.

6. Howard RW, Blomquist GJ. Ecological, behavioral, and biochemical aspects of insect hydrocarbons. Annu Rev Entomol. 2005;50: 371–393. doi:10.1146/annurev.ento.50.071803.130359

7. Grillet M, Dartevelle L, Ferveur J-F. A Drosophila male pheromone affects female sexual receptivity. Proc Royal Soc B. 2006;273: 315–323. doi:10.1098/rspb.2005.3332

8. Ferveur J-F. Cuticular hydrocarbons: their evolution and roles in Drosophila pheromonal communication. Behav Genet. 2005;35: 279–295. doi:10.1007/s10519-005-3220-5

9. Brent CS, Byers JA, Levi-Zada A. An insect anti-aphrodisiac. eLife. eLife Sciences Publications Limited; 2017;6: 1765. doi:10.7554/eLife.24063

10. Barr M. Male mating behavior. WormBook. 2006;: 1–11. doi:10.1895/wormbook.1.78.1
11. Chute CD, Srinivasan J. Chemical mating cues in C. elegans. Seminars in Cell and Developmental Biology. Elsevier Ltd; 2014;33: 18–24. doi:10.1016/j.semcdb.2014.06.002

12. Simon JM, Sternberg PW. Evidence of a mate-finding cue in the hermaphrodite nematode Caenorhabditis elegans. Proceedings of the National Academy of Sciences. 2002;99: 1598–1603. doi:10.1073/pnas.032225799

13. White JQ, Nicholas TJ, Gritton J, Truong L, Davidson ER, Jorgensen EM. The sensory circuitry for sexual attraction in C. elegans males. Current Biology. 2007;17: 1847–1857. doi:10.1016/j.cub.2007.09.011

14. Srinivasan J, Kaplan F, Ajredini R, Zachariah C, Alborn HT, Teal PEA, et al. A blend of small molecules regulates both mating and development in Caenorhabditis elegans. Nature. 2008;454: 1115–1118. doi:10.1038/nature07168

15. Izrayelit Y, Srinivasan J, Campbell SL, Jo Y, Reuss von SH, Genoff MC, et al. Targeted metabolomics reveals a male pheromone and sex-specific ascaroside biosynthesis in Caenorhabditis elegans. ACS Chem Biol. 2012;7: 1321–1325. doi:10.1021/cb300169c

16. Chasnov JR, So WK, Chan CM, Chow KL. The species, sex, and stage specificity of a Caenorhabditis sex pheromone. Proceedings of the National Academy of Sciences. 2007;104: 6730–6735.

17. Morsci NS, Haas LA, Barr MM. Sperm status regulates sexual attraction in Caenorhabditis elegans. Genetics. 2011;189: 1341–1346. doi:10.1534/genetics.111.133603

18. Leighton DHW, Choe A, Wu SY, Sternberg PW. Communication between oocytes and somatic cells regulates volatile pheromone production in Caenorhabditis elegans. Proc Natl Acad Sci USA. 2014;111: 17905–17910. doi:10.1073/pnas.1420439111

19. Narayan A, Venkatachalam V, Durak O, Reilly DK, Bose N, Schroeder FC, et al. Contrasting responses within a single neuron class enable sex-specific attraction in Caenorhabditis elegans. Proc Natl Acad Sci USA. 2016;113: E1392–401. doi:10.1073/pnas.1600786113

20. Barrios A, Nurrish S, Emmons SW. Sensory regulation of C. elegans male mate-searching behavior. Curr Biol. 2008;18: 1865–1871. doi:10.1016/j.cub.2008.10.050

21. Aprison EZ, Ruvinsky I. Sexually Antagonistic Male Signals Manipulate Germline and Soma of C. elegans Hermaphrodites. Curr Biol. 2016. doi:10.1016/j.cub.2016.08.024

22. Shi C, Runnels AM, Murphy CT. Mating and male pheromone kill Caenorhabditis males through distinct mechanisms. eLife. 2017;6: S62. doi:10.7554/eLife.23493

23. Dong L, Cornaglia M, Lehner T, Gijs MAM. On-chip microfluidic biocommunication assay for studying male-induced demise in C. elegans hermaphrodites. Lab Chip. Royal Society of Chemistry; 2016;16: 4534–4545. doi:10.1039/C6LC01005A
24. Maures TJ, Booth LN, Benayoun BA, Izrayelit Y, Schroeder FC, Brunet A. Males Shorten the Life Span of C. elegans Hermaphrodites via Secreted Compounds. Science. 2014;343:541–544. doi:10.1126/science.1244160

25. Gupta BP, Rezai P. Microfluidic approaches for manipulating, imaging, and screening C. elegans. Micromachines. 2016;7: 1–26. doi:10.3390/mi7070123

26. Bakhtina NA, Korvink JG. Microfluidic laboratories for C. elegans enhance fundamental studies in biology. Rsc Advances. 2014. doi:10.1039/C3RA43758B

27. Ben-Yakar A, Chronis N, Lu H. Microfluidics for the analysis of behavior, nerve regeneration, and neural cell biology in C. elegans. Current Opinion in Neurobiology. 2009;19: 561–567. doi:10.1016/j.conb.2009.10.010

28. Squires TM, Quake SR. Microfluidics: Fluid physics at the nanoliter scale. Rev Mod Phys. American Physical Society; 2005;77: 977–1026. doi:10.1103/RevModPhys.77.977

29. Chung K, Zhan M, Srinivasan J, Sternberg PW, Gong E, Schroeder FC, et al. Microfluidic chamber arrays for whole-organism behavior-based chemical screening. Lab Chip. 2011;11: 3689–3697. doi:10.1039/c1lc20400a

30. McCormick KE, Gaertner BE, Sottile M, Phillips PC, Lockery SR. Microfluidic Devices for Analysis of Spatial Orientation Behaviors in Semi-Restrained Caenorhabditis elegans. Samuel A, editor. PLoS ONE. 2011;6: e25710–8. doi:10.1371/journal.pone.0025710

31. Lipton J, Kleemann G, Ghosh R, Lints R, Emmons SW. Mate searching in Caenorhabditis elegans: a genetic model for sex drive in a simple invertebrate. J Neurosci. Society for Neuroscience; 2004;24: 7427–7434. doi:10.1523/JNEUROSCI.1746-04.2004

32. Brenner S. The genetics of Caenorhabditis elegans. Genetics. 1974;77: 71–94.

33. Kenyon C. The nematode Caenorhabditis elegans. Science. 1988;240: 1448–1453.

34. Qin D, Xia Y, Whitesides GM. Soft lithography for micro- and nanoscale patterning. Nature Protocols. 2010;5: 491–502. doi:10.1038/nprot.2009.234

35. Lockery SR, Lawton KJ, Doll JC, Faumont S, Coulthard SM, Thiele TR, et al. Artificial Dirt: Microfluidic Substrates for Nematode Neurobiology and Behavior. Journal of Neurophysiology. 2008;99: 3136–3143. doi:10.1152/jn.91327.2007

36. Park S, Hwang H, Nam S-W, Martinez F, Austin RH, Ryu WS. Enhanced Caenorhabditis elegans Locomotion in a Structured Microfluidic Environment. Brembs B, editor. PLoS ONE. 2008;3: e2550–5. doi:10.1371/journal.pone.0002550

37. Albrecht DR, Bargmann CI. High-content behavioral analysis of Caenorhabditis elegans in precise spatiotemporal chemical environments. Nat Meth. 2011;8: 599–605. doi:10.1038/nmeth.1630
38. Srinivasan J, Reuss von SH, Bose N, Zaslaver A, Mahanti P, Ho MC, et al. A Modular Library of Small Molecule Signals Regulates Social Behaviors in Caenorhabditis elegans. Sengupta P, editor. PLoS Biol. 2012;10: e1001237–14. doi:10.1371/journal.pbio.1001237

39. Chapman T. Seminal fluid-mediated fitness traits in Drosophila. Heredity. 2011;87: 511–521.

40. Yamane T, Goenaga J, Rönn JL, Arnqvist G. Male Seminal Fluid Substances Affect Sperm Competition Success and Female Reproductive Behavior in a Seed Beetle. Wicker-Thomas C, editor. PLoS ONE. 2015;10: e0123770–14. doi:10.1371/journal.pone.0123770

41. Garcia LR, LeBoeuf B, Koo P. Diversity in Mating Behavior of Hermaphroditic and Male-Female Caenorhabditis Nematodes. Genetics. 2007;175: 1761–1771. doi:10.1534/genetics.106.068304

42. Stroustrup N, Ulmschneider BE, Nash ZM, López-Moyado IF, Apfeld J, Fontana W. The Caenorhabditis elegans Lifespan Machine. Nat Meth. 2013;10: 665–670. doi:10.1038/nmeth.2475

Supporting Information

S1 File. Pheromone Arena microfluidic design. The Pheromone Arena has two versions to account for the difference in size between day 2 adult males and females: male choice of female signalers and female choice of male signalers. In the male choice version, the upstream loading distribution network is sized at 60um to account for the larger diameter of females, while the downstream chamber distribution channel has a final constriction size of 35um to prevent males from climbing back out of the downstream chamber. The female choice version has a smaller upstream distribution network (40um) and a larger downstream chamber distribution (60um).

Additionally, in the female choice version the two upstream chambers have the first two rows of pillars removed, again to prevent males from climbing out of the device. Both versions have increased resistance added to the outflow to decreased the overall flow rate through the device. These blueprints are accessible using CAD software. A master height of 65um is recommended.

S2 File. Data. Raw data for all male choice, female choice, and control experiments.
S1 Fig. Pressurized air flow set-up. A pressurized air system was used to maintain a constant flow rate through the microfluidic devices. A one gallon air tank was regulated to 1.5 PSI was sufficient to run experiments for up to 3 hours. The air-line running from the tank was bifurcated to pressurize two sealed 500mL bottles of M9 buffer. The bottle caps were modified to supply six liquid lines, which connected to two Pheromone Arenas, as well as hold the air-line, which terminated at the cap. The tubing lengths were equal for each partition of the set-up to maintain equal flow through all lines. The Pheromone Arena was kept on a confocal microscope at 20°C for the duration of each experiment. This figure was modified with permission from Stephen Banse.

S2 Fig. Virgin male choice over time. Virgin, day 2 adult C. remanei males were given a choice between virgin and mated conspecific female pheromone (blue) and C. elegans males were given a choice between virgin and mated conspecific female pheromone (orange). The weighted means and standard error are plotted over time. By 60 minutes into the experiment males had made a consistent choice (down triangle). The null hypothesis of no choice is given by the dashed line. In each assay males were more attracted to virgin females than mated ones.

S3 Fig. Virgin female choice over time. Virgin, day 1 adult C. remanei females were given a choice between virgin conspecific male and female pheromones (blue) and C. elegans males were given a choice between virgin conspecific male and female pheromones (orange). The weighted means and standard error are plotted over time. The null hypothesis of no choice is given by the dashed line. Only C. remanei females made a measurable choice of male pheromone over female pheromone.
Male Chooser

Attraction to Virgin Females (%)

Female Signal:
- C. remanei
- C. elegans

Conspecific Choice

Heterospecific Choice

C. remanei
C. elegans

C. remanei
C. elegans

Male Chooser

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