The Anti-Leukemia Effect of Ascorbic Acid: From the Pro-Oxidant Potential to the Epigenetic Role in Acute Myeloid Leukemia

S. Travaglini1, C. Gurnari1,2, S. Antonelli1, G. Silvestrini1, N. I. Noguera1,3, T. Ottone1,3 and M. T. Voso1,3*

1Department of Biomedicine and Prevention, University of Rome Tor Vergata, Rome, Italy, 2Department of Translational Hematology and Oncology Research, Taussig Cancer Institute, Cleveland Clinic, Cleveland, OH, United States, 3Neuro-Oncohematology Unit, IRCCS Fondazione Santa Lucia, Rome, Italy

Data derived from high-throughput sequencing technologies have allowed a deeper understanding of the molecular landscape of Acute Myeloid Leukemia (AML), paving the way for the development of novel therapeutic options, with a higher efficacy and a lower toxicity than conventional chemotherapy. In the antileukemia drug development scenario, ascorbic acid, a natural compound also known as Vitamin C, has emerged for its potential anti-proliferative and pro-apoptotic activities on leukemic cells. However, the role of ascorbic acid (vitamin C) in the treatment of AML has been debated for decades. Mechanistic insight into its role in many biological processes and, especially, in epigenetic regulation has provided the rationale for the use of this agent as a novel anti-leukemia therapy in AML. Acting as a co-factor for 2-oxoglutarate-dependent dioxygenases (2-OGDDs), ascorbic acid is involved in the epigenetic regulations through the control of TET (ten-eleven translocation) enzymes, epigenetic master regulators with a critical role in aberrant hematopoiesis and leukemogenesis. In line with this discovery, great interest has been emerging for the clinical testing of this drug targeting leukemia epigenome. Besides its role in epigenetics, ascorbic acid is also a pivotal regulator of many physiological processes in human, particularly in the antioxidant cellular response, being able to scavenge reactive oxygen species (ROS) to prevent DNA damage and other effects involved in cancer transformation. Thus, for this wide spectrum of biological activities, ascorbic acid possesses some pharmacologic properties attractive for anti-leukemia therapy. The present review outlines the evidence and mechanism of ascorbic acid in leukemogenesis and its therapeutic potential in AML. With the growing evidence derived from the literature on situations in which the use of ascorbate may be beneficial in vitro and in vivo, we will finally discuss how these insights could be included into the rational design of future clinical trials.

Keywords: acute myeloid leukemia, ascorbic acid, epigenetic regulation, oxidative stress, vitamin C
INTRODUCTION

Acute myeloid leukemia (AML) is a heterogeneous clonal disorder characterized by the uncontrolled proliferation of undifferentiated myeloid progenitor cells in the bone marrow and peripheral blood. Advances in DNA sequencing technologies have provided a detailed knowledge of the molecular landscape of AML, with a better understanding of the disease pathogenesis and prognosis (The Cancer Genome Atlas Research Network, 2013; Papaemmanuil et al., 2016; Awada et al., 2021). Despite high heterogeneity, the spectrum of genetic alterations have highlighted the presence of recurrent mutations in genes encoding epigenetic regulators (Gallipoli et al., 2015), including DNA methyltransferase 3A (DNMT3A), Ten-eleven-translocation 2 (TET2), Wilms’ tumor 1 (WT1), and isocitrate dehydrogenase 1 and 2 (IDH1/2) (Kunimoto and Nakajima, 2017). The study of the molecular architecture of such cases unveiled that alterations of these genes often represent founding events in the clonal hierarchy, suggesting their essential role in early phase of leukemia ontogenesis (Papaemmanuil et al., 2013; Hirsch et al., 2018; Yoshimi et al., 2019).

Epigenetic mechanisms play a major role in normal and, particularly, in malignant hematopoiesis, where the presence of recurrent alterations in transcription factors and chromatin regulators are able to drive hematopoietic malignancies (Hu and Shilatifard, 2016; Fennell et al., 2019). Epigenetic modifiers are a large and varied group of proteins involved in the modification of DNA at cytosine residues and post-translational acetylation and methylation of histones, whose aberrations lead to dysregulation of critical genes that control cell growth, differentiation and apoptosis, all important mechanisms in the pathogenesis of AML (Hou and Tien, 2016; Pastore and Levine, 2016). In contrast to genetic changes, epigenetic modifications refer to changes in gene expression, and are frequently reversible, providing the opportunities for targeted treatment (Wouters and Delwel, 2016).

A wide range of therapeutic strategies that target epigenetic alterations in AML have been successfully tested in preclinical studies and several drugs have already gained approval for clinical use, with the final goal to reverse epigenetic dysfunctions (Bewersdorf et al., 2019). Since these discoveries, a number of selective inhibitors of mutant FLT3, IDH1 and IDH2 have been developed and are now approved by the Food and Drug Administration, and drugs that target the potential epigenetic writers (DOT1L, PRMT5), readers (BRD2/3/4) and erasers (HDAC, LSD1) are currently under investigation (Das et al., 2021). However, the therapeutic efficacy of epigenetic drugs as single agents seems to be limited (Bewersdorf et al., 2019).

Ascorbic acid, also known as vitamin C or ascorbate, at physiological levels potentiates the effects of hypomethylating agents (HMA) by both causing DNA demethylation through active and passive mechanisms and selectively killing tumor cells (Ruiz et al., 2015; Liu et al., 2016; Gillberg et al., 2018). Indeed, ascorbic acid is an essential factor for epigenetic regulation and many studies reported that it also displays a pro-oxidant activity when used at high concentrations, in particular on various type of cancer cells, suggesting it as an emerging epigenetic therapy (Chen et al., 2005; Chen et al., 2008).

Besides its role in epigenetics, ascorbic acid is a pivotal regulator of many physiological processes in humans (Giansanti et al., 2021), including cellular immune responses (Ströhle et al., 2011), and represents an antioxidant molecule involved in the reactive oxygen species (ROS) scavenging that prevent oxidative DNA damage and other effects, which may lead to cancer transformation.

In this review, we provide an overview of the role of ascorbate as an essential factor for epigenetic regulation, highlighting its anti-leukemic mechanisms of action, currently under investigation in the treatment of AML.

ASCORBIC ACID: STRUCTURE, BIOSYNTHESIS AND UPTAKE

Ascorbic acid represents a wide spectrum antioxidant and an essential nutrient for humans. This vitamin exists in two redox states, ascorbate, the reduced active form, and dehydroascorbic acid (DHA), its oxidized form. At physiological conditions, ascorbic acid loses a proton to form the ascorbate anion (AscH⁻), the predominant ionic form of vitamin C, that can be oxidized by losing two protons, giving rise first to ascorbate radical (Asc·⁻) and then to a fully oxidized DHA (Ferrada et al., 2021).

Unlike most mammals, humans are unable to synthesize ascorbic acid from oxidated glucose, missing the gulonolactone (L-) oxidase (GULO), a key enzyme involved in the catalysis of the last enzymatic step in ascorbate synthesis, and are thereby dependent on vitamin C intake from the diet (Du et al., 2012). The dietary daily requirement of vitamin C is 75–90 mg/day, which is usually taken under the form of both ascorbate and DHA (Valdés, 2006; Testa et al., 2021). Indeed, even if DHA levels are extremely low under physiological conditions (Du et al., 2012), it is rapidly recycled back to ascorbate by DHA reductase, using glutathione (GSH) as a reducing agent (Mandl et al., 2009). This recycling process occurs intracellularly, where vitamin C is involved in several biological processes. Thus, its exogenous uptake is attained through the presence of specific membrane transporters, which determine the distribution of this molecule between extra- and intra- cellular compartments. Both ascorbic acid and DHA can be transported across the plasma membrane, although by distinct carriers (Savini et al., 2008). Under physiologic conditions, vitamin C is mainly present in plasma as ascorbate and is actively absorbed in the gastrointestinal tract, particularly at the level of the enterocytes of the small intestine (May, 2011; Du et al., 2012).

Ascorbate is transported into mammalians cells in an energy-dependent fashion, requiring two types of sodium-dependent transporters, SVCT1 and SVCT2, encoded by SLC23A1 and SLC23A2 genes, respectively, which show distinct tissue distributions (Tsukaguchi et al., 1999; Sotiriou et al., 2002; May, 2011). Conversely, DHA can be internalized by facilitative hexose transporters, particularly via GLUT1 and GLUT3 (Rumsey et al., 1997; Rumsey et al., 1999), encoded by
Liu et al. (2020) showed that including lymphoma, breast, head and neck, and gastric cancers in the tumor growth and spread (Wohlrab et al., 2017). Indeed, the deficiency due to a variety of factors such as decreased oral intake, inflammation, infection, and disease phases, particularly following chemotherapy and hematopoietic stem cells (HSCs) transplantation, resulting in shorter survival (Caraballoso et al., 2003; Mayland et al., 2005; Nannya et al., 2014; Wilson et al., 2014; Rasheed et al., 2019; White et al., 2020). Furthermore, it has also been indicated a potential role of vitamin C transporters in human cancer (Caprile et al., 2009; Hong et al., 2014; Wilson et al., 2014; Rasheed et al., 2019; White et al., 2020). By increasing the extracellular matrix components, the most abundant extracellular protein, promoting the proper folding of the stable collagen triple-helix conformation (Phillips et al., 1994). Under normoxic conditions, the hydroxylation of these proline residues induces proteasomal degradation of HIF-1α. In case of hypoxia, a scenarios which is very common in cancer, or in absence of ascorbate, hydroxylation is inhibited and HIF-1α is stabilized thereby initiating its downstream effects (Flashman et al., 2010). Furthermore, ascorbic acid, as a cofactor, is also involved in other important hydroxylation reactions, essential for catecholamines, L-carnitine, cholesterol and amino acids synthesis (Chatterjee et al., 1975).

Consistent with its role as a cofactor, ascorbic acid regulates DNA and histone methylation thanks to its ability to modulate 2-OGDD enzymes, which encompass demethylases involved in epigenetic regulation and in the maintenance of genomic stability (Gimmino et al., 2018). Particularly, the TET proteins are responsible for the active DNA demethylation and regulate gene transcription by converting 5-methylcytosine (5mC) to 5-hydroxymethylcytosine (5hmC), further to 5-formylcytosine (5fC) and 5-carboxylcytosine (5caC), to produce an unmethylated cytosine, thus completing the process of DNA active demethylation (Tahiliani et al., 2009; Ito et al., 2011). Ascorbic acid sustains and promotes the catalytic activity of the SLC2A1 and SLC2A3 genes, and then converted back to ascorbic acid (Savini et al., 2008) (Figure 1). As mentioned above, DHA is present at extremely low concentration in the blood of healthy subjects and at much higher levels in the intestine, and its uptake is inhibited by excess glucose (Vera et al., 1993; Malo and Wilson, 2000; Corpe et al., 2013; Lykkesfeldt and Tveden-Nyborg, 2019). However, the intracellular transport of DHA by GLUT transporters is not usually considered the principal route of uptake, even if the impact of ascorbate and/or DHA to cellular transport in vivo is not fully elucidated (Rumsey and Levine, 1998).

Vitamin C uptake and its whole body distribution is essential for many biochemical processes, some of them also influencing tumor growth and spread (Wohlrab et al., 2017). Indeed, the ascorbate concentration in plasma and tissues represent one of the main risk factor for cancer incidence (Linowiecka et al., 2020). Several studies reported also that cancer patients are more likely to experience vitamin C deficiency due to a variety of factors such as decreased oral intake, inflammation, infection, and disease phases, particularly following chemotherapy and hematopoietic stem cells (HSCs) transplantation, resulting in shorter survival (Caraballoso et al., 2003; Mayland et al., 2005; Nannya et al., 2014; Wilson et al., 2014; Rasheed et al., 2019; White et al., 2020). Furthermore, the TET proteins are responsible for the active DNA demethylation and regulate gene transcription by converting 5-methylcytosine (5mC) to 5-hydroxymethylcytosine (5hmC), further to 5-formylcytosine (5fC) and 5-carboxylcytosine (5caC), to produce an unmethylated cytosine, thus completing the process of DNA active demethylation (Tahiliani et al., 2009; Ito et al., 2011). Ascorbic acid sustains and promotes the catalytic activity of the

Ascorbic acid takes part in many biochemical processes in humans, acting as an essential enzymatic cofactor, a reducing and antioxidant agent and a scavenger of ROS in biological systems. The wide spectrum of its biological functions relies on its role as a specific cofactor for the catalytic activity of the 2-oxoglutarate-dependent dioxygenases (2-OGDDs), which catalyze the addition of hydroxyl group to various substrates (Kuiper and Vissers, 2014). For instance, under physiological conditions, ascorbate is essential in the biosynthesis of collagen, the most abundant extracellular protein, promoting the proper folding of the stable collagen triple-helix conformation (Phillips et al., 1994). By increasing the extracellular matrix components, these mechanisms are supposed to prevent cancer spread, thus walling in tumors (Cameron and Rotman, 1972; Cameron et al., 1979). In addition, ascorbate may also act as a cofactor in the hydroxylation of the hypoxia inducible factor-1α (HIF-1α), a transcription factor that regulates the expression of specific genes involved in the cellular response to hypoxia (Ivan et al., 2001; Jaakkola et al., 2001; Schofield and Ratcliffe, 2004; Ratcliffe, 2013; Simon, 2016). Under normoxic conditions, the hydroxylation of these proline residues induces proteasomal degradation of HIF-1α. In case of hypoxia, a scenarios which is very common in cancer, or in absence of ascorbate, hydroxylation is inhibited and HIF-1α is stabilized thereby initiating its downstream effects (Flashman et al., 2010). Furthermore, ascorbic acid, as a cofactor, is also involved in other important hydroxylation reactions, essential for catecholamines, L-carnitine, cholesterol and amino acids synthesis (Chatterjee et al., 1975).

Consistent with its role as a cofactor, ascorbic acid regulates DNA and histone methylation thanks to its ability to modulate 2-OGDD enzymes, which encompass demethylases involved in epigenetic regulation and in the maintenance of genomic stability (Gimmino et al., 2018). Particularly, the TET proteins are responsible for the active DNA demethylation and regulate gene transcription by converting 5-methylcytosine (5mC) to 5-hydroxymethylcytosine (5hmC), further to 5-formylcytosine (5fC) and 5-carboxylcytosine (5caC), to produce an unmethylated cytosine, thus completing the process of DNA active demethylation (Tahiliani et al., 2009; Ito et al., 2011). Ascorbic acid sustains and promotes the catalytic activity of

FIGURE 1 | Ascorbic acid uptake. Ascorbic acid, also known as Vitamin C, enters the cell either in its reduced form (ascorbate, ASC) by sodium-dependent vitamin C transporters (SVCTs) or in its oxidized form (dehydroascorbate, DHA) via facilitative glucose transporters (GLUTs). In the cytosol, DHA is rapidly reduced back to ASC in the presence of glutathione (GSH). Created with Biorender.com.
TET enzymes, most likely reducing Fe$^{3+}$ to Fe$^{2+}$ (Kuiper and Vissers, 2014). Recent studies have demonstrated that ascorbic acid, as a direct regulator of TET activity, may enhance 5hmC generation, in a time- and dose-dependent fashion (Yin et al., 2013; Young et al., 2015). Two mechanisms have been proposed to explain the stimulatory effect of ascorbate on TET enzymatic activity. The first is related to its role as a cofactor, capable to directly bind the catalytic domain of TET proteins by enhancing their enzymatic activity (Hore et al., 2016); this finding is also supported by the evidence that other antioxidant compounds do not show effects on TET activity in in vitro models (Tahiliani et al., 2009; Ito et al., 2011; Chen et al., 2013; Dickson et al., 2013; Minor et al., 2013; Yin et al., 2013). The second hypothesis posits a different mechanism, suggesting that the stimulatory role of ascorbate on TET activity is associated to its ability to promote the reduction of Fe$^{3+}$ to Fe$^{2+}$ (Hore et al., 2016).

In addition to its function as a cofactor of several enzymes, ascorbic acid is involved in many biological processes acting as an electron donor. At physiological concentrations, ascorbate is a potent free radical scavenger, with a protective effect against oxidative damage caused by ROS, commonly produced during normal cellular metabolism (Gaziano et al., 2012). The antioxidant mechanisms of ascorbic acid are based on its capacity to reduce potentially damaging ROS and produce chemically inert resonance-stabilized ascorbate radicals (AFR). The AFR is reduced back to ascorbate intracellularly by the activity of NADH$^-$ and NADPH dependent reductases, which display high affinity for the generated radicals (Duarte and Lunec, 2005; Santos-Sánchez et al., 2019). In case of accumulation of AFR in areas not accessible to these enzymes, two AFR molecules may either react with each other or dismutate to form one molecule each of ascorbate and DHA (Bielski et al., 1981). These may be then converted back into ascorbic acid for reuse or may be metabolized with further release of additional electrons (May et al., 2001; Wilson, 2002; Duarte and Lunec, 2005). This mechanism underlines the cytoprotective functions of Vitamin C, involved in the first line of antioxidant defense, including...
prevention of DNA mutation induced by oxidation (Noroozi et al., 1998; Pflaum et al., 1998; Lutsenko et al., 2002), protection of lipids against peroxidative damage (Barja et al., 1994; Sweetman et al., 1997), and repair of oxidized amino acid residues to maintain protein integrity (Kimura et al., 1992; Sweetman et al., 1997; Cadenas et al., 1998). The multifactorial roles of ascorbate discussed above are summarized in Figure 2.

ASCORBATE AS A KEY EPIGENETIC REGULATOR INVOLVED IN HEMATOPOIETIC STEM CELL FUNCTION AND LEUKEMOGENESIS

As previously mentioned, among the various biological effects induced by ascorbic acid, emerging evidences suggest its key role in the epigenetic reprogramming, an effect related to the ability of ascorbic acid to act as an electron donor and, particularly, a cofactor for 2-OGDDs. These enzymes require 2-oxoglutarate (2-OG) and molecular oxygen, as substrates, and non-heme iron (Fe2+) and ascorbic acid, as cofactors; therefore, the reduction of both substrates or cofactors result in a decreased activity of these enzymes (McDonough et al., 2016; Vissers et al., 2014; Martinez-Reyes and Chandel, 2020; Crake et al., 2021). This superfamily of enzymes include key epigenetic regulators of histone demethylation and DNA hydroxymethylation, which have been shown to have crucial roles in the epigenetic regulation of stem cells and cancer (Cimmino et al., 2018). These evidences are of particular interest for AML because many of these demethylase enzymes, which require ascorbic acid as a cofactor, are deregulated in the process of leukemogenesis (Das et al., 2021), including lysin-specific demethylases (Castelli et al., 2018), the Jumonji C (JmjC) domain containing histone demethylases (JHDMs) (Wang et al., 2011a; Staehele et al., 2021), the ALKB homolog (ALKBH) family of nucleic acid demethylases (Gerken et al., 2007; Yi et al., 2010) and TET enzymes (Castelli et al., 2018; Das et al., 2021). Of the context discussed above, it has been recently demonstrated that ascorbic acid, acting as an epigenetic modulator could promote DNA demethylation in embryonic stem cells (ESCs), stimulate the reprogramming of fibroblast to induced pluripotent stem cells (iPSCs) and humper the aberrant self-renewal of HSCs, through the stimulation of JHDM activity (Chung et al., 2010; Esteban et al., 2010; Cimmino et al., 2018). Recent studies on JHDMs have also clearly demonstrated the involvement of these enzymes in disease initiation and progression, suggesting new attractive targets for myeloid malignancies.

Particularly, several studies have investigated the specific relationship between the activity of TET enzymes, ascorbate and the development of hematologic malignancies. It has been recently reported that ascorbic acid can protect HSCs from epigenetic alterations driving leukemia progression, stimulating the catalytic activity of TET enzymes, which are known as bona fide tumour suppressors of the hematopoietic lineage. Among TET genes, alterations in TET2 are frequently reported in myeloid disorders, occurring in 10% of de novo AML, 30% of myelodysplastic syndrome (MDS) and almost 50% of chronic myelomonocytic leukemia (CMML) cases (Delhommeau et al., 2009; Papaemmanuil et al., 2016). TET2 is one of the principal epigenetic regulator of normal and malignant hematopoiesis, being able to regulate the differentiation and self-renewal of HSCs (Nakajima and Kunimoto, 2014), and is one of the most commonly mutated genes reported at high allele frequency in CD34+ hematopoietic stem and progenitor cells (HSPCs). These evidences suggested that TET2 mutations are early clonal events of the leukemic transformation in cells with multi-lineage potential.

Acting as a co-factor for the 2-OGDDs, ascorbate sustains and promotes the activity of TET enzymes, as already demonstrated by in vitro and in vivo evidences (Agathocleous et al., 2017; Cimmino et al., 2017; Vissers and Das, 2018). Two recent studies provide novel insights on how ascorbic acid regulates HSC functions and leukemogenesis by the enhancement and restoration of TET2 function, respectively.

In the first, Agathocleous et al. reported that vitamin C deficiency might play a role in leukemia progression. Interestingly, the authors showed that the more immature population of stem and progenitor cells display higher levels of ascorbate than more differentiated cells. Accordingly, they found also an overexpression of the Slek23a2 gene, which provides instructions for making a protein involved in vitamin C uptake in HSCs/MPPs (multipotent progenitors), determining an accumulation of ascorbate levels, which decreased with differentiation (Agathocleous et al., 2017). Using Gulo-depleted mice, which are unable to synthesize ascorbate from glucose, they showed that ascorbate depletion increased HSC pool, in part as a consequence of Tet2 reduced activity, a scenario which was reversed by dietary vitamin C supplementation. These findings suggest the beneficial role of ascorbate treatment in the setting of TET2 mutated leukemia. Furthermore, the presence of additional internal tandem duplications (ITDs) in the juxtamembrane domain of FLT3 cooperated with ascorbate deprivation in acceleration of leukemia development (Agathocleous et al., 2017), phenocopying TET2 loss.

In a second, independent study Cimmino et al. established a reversible mouse model with Tet2 knockout. Tet2-deficient mice showed defective self-renewal and differentiation capacity of HSC/HPC (hematopoietic progenitor cell), and defective genomic hydroxymethylation and DNA hypermethylation. Treatment with ascorbic acid pharmacologically mimicked TET2 restoration, inducing a reversal of defective DNA methylation and cell differentiation. As TET2 mutations are almost exclusively heterozygous, ascorbic acid was found capable to stimulate the activity of the non-mutated TET2 allele, leading to genome-wide DNA demethylation, differentiation and cell death. These evidences also suggest that the persistence of TET2 deficiency is needed to maintain leukemic self-renewal (Cimmino et al., 2017). Furthermore, the authors showed the ability of ascorbate to enhance the activity of poly(ADP ribose) polymerase (PARP) inhibitor to induce cell
death, providing a safe and effective combination strategy to selectively target TET deficiency in cancer (Cimmino et al., 2017).

Altogether, these data suggest that the restoration of TET2 activity via ascorbate supplementation could provide an opportunity to reverse disease progression in AML cases linked to heterozygous loss-of-function mutations in TET2, pointing out to its role as a potentially non-toxic therapy for TET-associated malignancies (Agathocleous et al., 2017; Cimmino et al., 2017).

**THE ANTI-CANCER EFFECTS OF ASCORBIC ACID AT PHARMACOLOGIC DOSES**

Among its many effects on cellular functions and metabolism, ascorbic acid has also shown a powerful anti-cancer effect against a number of human cell lines. Indeed, while generally regarded as an antioxidant, ascorbic acid may also have prooxidant activities at high pharmacological concentrations (Noguera et al., 2017), obtained by intravenous administration, and in the presence of free transition metal ions, especially iron. Particularly, ascorbate catalyzes the reduction of Fe$^{3+}$ to Fe$^{2+}$, leading to the formation of H$_2$O$_2$, through the so-called Fenton reaction, and exerting a cytostatic and cytotoxic effect against tumor cells, without harming normal cells. This pro-oxidant potential has been investigated in many studies in the prevention and treatment of cancers and is proposed to be dose-dependent (Putchala et al., 2013). Among the possible mechanisms, four main biological pathways have been proposed to explain how high-dose ascorbate targets several vulnerabilities of tumor cells: intracellular iron metabolism, DHA uptake via GLUT1, hypoxia pathways and epigenetic regulators (Figure 3).

One of the major determinants of ascorbate-mediated tumor cytotoxicity is represented by the amount and the availability of intracellular iron. The labile iron pool (LIP) represents a source of intracellular iron available for exchange between various cellular

---

**FIGURE 3** | The anti-cancer effects of ascorbic acid. (A) A high concentration of ascorbic acid (ASC) increases the labile iron pool (LIP) of tumor cells, induces the production of increased level of ROS directly damaging mitochondria and DNA, and ultimately stimulates apoptotic pathways. (B) Tumor cells can uptake DHA at higher rates and then internally reduce it to ASC. This reduction triggers scavenging of glutathione (GSH), inducing oxidative stress, the inactivation of glycolytic flux, an energy crisis and cell death. (C) In the hypoxic conditions of tumor microenvironment, there is a repression of HIF-1α hydroxilation. As a result, HIF-1α accumulates in the cytoplasm and translocates into the nucleus, promoting the transcription of its targets involved in processes such as angiogenesis, glycolysis and anaerobic metabolism, metastasis and resistance to therapy. Under ascorbic acid treatment, HIF-1α is hydroxylated by the prolyl hydroxylases, ultimately leading to polyubiquitination and proteosomal degradation of HIF-1α. (D) Ascorbic acid binds to the catalytic domain of TET and facilitates DNA demethylation and re-expression of important tumor suppressor genes. Created with Biorender.com.
compartments (Kakhlon and Cabantchik, 2002). The intracellular iron concentration has a key role in the ascorbate-mediated cytotoxicity, related to the formation of H$_2$O$_2$ and .OH, which directly damages mitochondria and DNA, stimulating apoptotic pathways. Particularly, tumor cells displayed low levels of antioxidant enzymes and an impaired redox balance in respect of their normal counterpart, rendering these cells more vulnerable to oxidative stress and identifying pro-oxidant stimulations as a strategy to induce their death (Moloney and Cotter, 2018). According to these findings, Schoenfeld et al. have explored in detail the sensitivity of glioblastoma and non-small cell lung cancer (NSCLC) cell lines to ascorbate and have demonstrated that an altered iron metabolism, associated with disturbances in oxidative stress, induces an increased level of mitochondrial ROS, ultimately leading to an increase in the level of LIP and sensitivity of cancer cells to ascorbic acid (Schoenfeld et al., 2017).

Interestingly, a dual relationship exists between LIP and ascorbate: pharmacological doses of ascorbate increase LIP in tumor cells, whereas an increased LIP in tumor cells enhances the toxicity of pharmacological doses of ascorbate (Du et al., 2015). Moreover, elevated LIP levels have been found in several tumor cell types as opposed to their normal counterpart, such as breast cancer cells, T-cell lymphoma and RAS-transformed cells (Yang and Stockwell, 2008; Kiessling et al., 2009; Pinnix et al., 2010). Accordingly, one could speculate that the intravenous iron injection may represent a powerful strategy to sensitize tumor cells to ascorbic acid via increased LIP levels (McCarty and Contreras, 2014).

The second mechanism involves DHA, the reversible oxidized form of ascorbic acid, based on the findings that tumor cells have the peculiar ability to uptake DHA at much higher rates and then internally reduce it to ascorbic acid (Nauman et al., 2018). As reported by Yun et al. (2015), in KRAS and Braf-mutated colorectal cancer, this reduction triggers scavenging of glutathione (GSH), induces oxidative stress, inactivates glyceraldehyde 3-phosphate dehydrogenase (GAPDH), inhibits glycolytic flux and leads to an energy crisis with subsequent cell death (van der Reest and Gottlieb, 2016). Acting as a glycolytic inhibitor, high-dose ascorbate could represent an effective strategy against tumor cells, frequently characterized by high glycolytic activity. The third anti-tumor mechanism is based on the activity of ascorbate as a cofactor for 2-OGDDs, including HIF-hydroxylases (Ozer and Bruick, 2007). As reported above, in the hypoxic condition of tumor microenvironment, there is a repression of HIF-1α hydroxylation, associated with HIF activity. HIF-1α promotes the transition of tumor cells from aerobic to anaerobic metabolism, increasing glycolysis to maintain energy production (Masoud and Li, 2015). Thus, it is conceivable that the previously discussed DHA mechanism works in conjunction with the oxygen pathways alterations, leading to a global disruption of metabolic functioning in the tumor cell, that trigger cell death (Nauman et al., 2018). Furthermore, several studies have reported elevated levels of HIF-1α in AML, where it mediates the capacity of leukemia cells to migrate and invade extramedullary sites, suggesting that hypoxia and HIF-mediated signaling may play a crucial role in leukemia. Thus, targeting HIF with ascorbic acid could represent a potentially useful approach in AML treatment (Wang et al., 2011b; Kawada et al., 2013; Forristal et al., 2015; Gao et al., 2015).

Lastly, ascorbate modulates DNA demethylation and epigenetically reprograms cancer cells through the interaction with TET enzymes family (Blaschke et al., 2013; Ito et al., 2013). Binding to the catalytic domain, it facilitates TET-mediated DNA demethylation and re-expression of important tumor suppressor genes, with subsequent increase in chemosensitivity (Blaschke et al., 2013; Shenoy et al., 2017).

TARGETING AML DRIVER MUTATIONS USING ASCORBIC ACID: FROM IN VITRO EXPERIENCES TO CLINICAL TRIALS

Despite a growing body of evidence suggesting the anti-cancer properties of ascorbic acid, few data are available on its role for the treatment of AML.

In an in vitro study, Parker and coworkers demonstrated that ascorbate, at concentrations of 0.25–1 mM, was able to induce a dose- and time-dependent inhibition of proliferation in various AML cell lines (HL-60, acute promyelocytic leukemia cell line NB4, retinoic acid-resistant APL cell line NB4-R1, K562 chronic myelogenous leukemia cell line, and KG1), and primary blasts (Park et al., 2004). The induction of apoptosis in these cells was due, at least in part, to the excessive increase of H$_2$O$_2$ levels (Park, 2013).

Another study performed by Kawada et al. (2013) attempted to determine whether high ascorbate may exert significant cytotoxic effects against human leukemic cells, K562, HL60, Jurkat (T-lymphoblastic leukemia) and Raji (B-lymphoblastic leukemia), and normal hematopoietic cells, confirming that the specific cytotoxic effects on leukemic cells were caused by the production of H$_2$O$_2$ with an effect directly proportional to the dose employed. These data were confirmed in vivo, whereby the intravenous administration of high ascorbate repressed the proliferation of leukemic cells injected in NSG (NOD scid gamma) mice. Moreover, the authors also showed that high dose ascorbate markedly inhibited the expression of HIF-1α in leukemic cells by blocking the transcriptional activation of NF-kB, constitutively upregulated in many types of leukemia and associated with leukemic progression (Braun et al., 2006; Packham, 2008; Reikvam et al., 2009).

Accordingly, Mastrangelo et al. (2015) tested the effects of high concentrations (0.5–7 mM) of ASC on a variety of human myeloid cell lines, HL60, K562, U937, NB4, NB4-R4 and arsenic trioxide (As$_2$O$_3$, ATO)-resistant NB4 (NB4/As), showing the high sensitivity of myeloid leukemia cell lines to the pro-oxidant effects of high doses ascorbate, with an average cytotoxic concentration of 3 mM. Surprisingly, ASC was found significantly more effective than ATO as a single agent in inducing apoptotic cell death in HL60 human cell lines in vitro. Since ATO also functions as a pro-oxidant factor, in a second study, Noguera et al. (2017) tested the effects of ascorbate in combination with ATO on AML and APL primary blasts and cell lines, including NB4, NB4-R4, NB4-ATO-R, and MV4-11.
AML-M5 derived cell line with t(4;11) and FLT3-ITD mutations], showing a synergistic effect. Particularly, the combination treatment was highly effective in APL samples, displaying a cytotoxic mechanism linked to ROS overproduction within leukemic cells and consequent induction of apoptosis. Moreover, high concentrations of ascorbate were able to downregulate the phosphorylation of FLT3 and its downstream target proteins STAT5a/b, suggesting the potential activity of the drug also in the subset of FLT3-ITD positive AML. In an attempt to further characterize the mechanistic underpinnings underlying the efficacy of ascorbic acid in the APL setting, the same authors found that the fusion protein PML/RARA inhibits NRF2 (NF-E2 p45-related factor 2) functions. This is a transcription factor that orchestrates cellular adaptive responses to stress, and whose nuclear transfer is prevented by ascorbate treatment, thereby enhancing its degradation into the cytoplasm. As a result, the inhibition of the NRF2 oxidative stress pathway clarifies the peculiar sensitivity of APL cells to the pro-oxidant activity of high-dose ascorbate and suggest its potential use in APL patients, especially in those resistant to ATO/retinoic acid treatment (Banella et al., 2019). Moreover, ascorbate also shows the ability to induce cell death by targeting glycolytic metabolism in primary AML blasts, through the inhibition of hexokinase 1/2 (HK1/2) and GLUT1 in hematopoietic cells, and, in combination with the metabolic inhibitor buformin, also decreases mitochondrial respiration and ATP production, sparing healthy CD34+ cells. Overall, these data clearly depict an effect of ascorbate on glycolysis and contribute to elucidate the targets and mechanisms through which this therapeutic agent exerts its anti-cancer effects (Banella et al., 2022).

Another study investigated instead the combinatorial effect in AML cells of low-dose of ascorbate with decitabine, an HMA widely used in the treatment of AML in elderly patients, not suitable for conventional intensive chemotherapy. Potentially, one of the mechanisms for the efficacy of decitabine treatment is the upregulation of TET2 proteins, among others. Acting as a direct regulator of TET activity, ascorbate increases 5hmC levels and potentially sensitizes patients to decitabine. Indeed, in vitro studies showed that this combination in NB4 and HL60 cells resulted in the most significant upregulation of the activity of TET2 enzyme. Furthermore, in vivo results displayed a synergistic effect of this two agents, which are able to improve the complete remission (CR) rate in elderly AML patients (Zhao et al., 2018). Overall, these observations are of potential interest particularly for MDS and AML patients, who could have an additional benefit from adding ascorbic acid to HMA therapy.

Interestingly, ascorbate specifically interacts with the C-terminus catalytic domain of TET2, inducing effects not shown with other strong reducing chemicals (Yin et al., 2013). As already mentioned, ascorbate can act as an epigenetic therapeutic in the presence of heterozygous TET2 mutations by restoring TET2 activity and providing an opportunity for reversing disease progression in AML cases linked to heterozygous loss-of-function mutations in TET2. Thus, the proposed mechanism of action requires the presence of residual functional enzyme (Das et al., 2021). However, TET2 mutations frequently occur in combination with other lesions, but little is known about how the genomic makeup may impact the up-regulation of TET2 activity induced by ascorbic acid as well as its subsequent effects on cell differentiation and survival (Papaemmanuil et al., 2016; Das et al., 2021). To further muddy the waters of the intertwining relation between TET enzymes and ascorbate in myeloid disorders, a reduced TET2 activity may also result from mutations in IDH1, IDH2 and WT1, frequently reported in AML (Figure 4). In the presence of such mutations, IDH enzymes produce 2-hydroxyglutarate (2-HG), an oncometabolite able to acts as a competitor for TET2, instead of its physiological substrate, 2-oxoglutarate (2-OG) (Figueroa et al., 2010), causing functional inactivation of TET2 enzyme. Moreover, the presence of mutated WT1, prevent the recruitment of TET2 to DNA and the activation of the WT1-target genes expression. These findings provide the evidence of decreased TET2 activity in both context of IDH and WT1 mutations, and are in line with a recent study reporting TET2 deficiency in up to 74% of patients with myeloid disorders, a result that goes beyond the mere presence of TET2 mutations. Indeed, loss of TET2 functions by mutations or down-modulation due to various mechanisms have been identified as a common lynchpin of myeloid malignancies, as also indicated by the meta-analysis performed in the above-mentioned study (Gurnari et al., 2022). The implications of this are obvious, and suggest the potential application of ascorbate in settings beyond the disruption of TET2-IDH-WT1 pathway, which is the scenario where we would expect its maximal therapeutic efficacy, as also demonstrated by anecdotal case reports (Das et al., 2019). Furthermore, TET3 upregulation has also been invoked as a potential mechanism compensating the general TET2opathy of myeloid disorders, and has been linked to better survival outcomes in MDS with TET2 deficiency. Perhaps, ascorbate treatment may rescue the fraction of patients not able to contra-balance TET2 loss, thereby improving their dismal clinical outcomes (Gurnari et al., 2021, 2022).

As said, IDH1/2 mutations alter the epigenome of AML cells. Mingay and coworkers explored the effect of ascorbic acid in the setting of a murine leukemic model expressing IDH1*R132H mutation. Ascorbate treatment induced a reduction in cell proliferation and an increased expression of genes involved in leukocyte differentiation in IDH1*R132H mice, not observed in IDH1*WT counterparts. These marked effects on cell differentiation were related to the induction of demethylation at the level of DNA binding sites of the hematopoietic transcription factors CEBPβ, HIF-1α, RUNX1 and PU.1 (Mingay et al., 2018). The previously mentioned study by Cimmino et al. (2017) provided the evidence that ascorbic acid treatment is able to induce the restoration of TET2 function in various leukemia models, by blocking aberrant self-renewal and leukemia progression. Indeed, mimicking TET2 restoration ascorbate treatment suppresses leukemic colony formation and leukemic progression of primary human leukemia patient-derived xenografts (PDX). Finally, a recent study investigated the effects of ascorbate on cell growth and differentiation of SKM-1 AML cell line, harbouring both TET2 and TP53 mutations, showing a
beneﬁcial anti-proliferative effect also in this subgroup of adverse-risk AML (Smith-Díaz et al., 2021).

Taken together these results identify ascorbic acid as a novel metabolic tumor suppressor involved in epigenetic remodeling and highlighted that supra-physiological doses could prevent myeloid disease progression, pointing out to its role as a potentially non-toxic therapy, especially for TET-deﬁcient malignancies (Agathocleous et al., 2017; Cimmino et al., 2017; Guan et al., 2020). These observations suggest the incorporation of high-dose ascorbate as an adjuvant to standard chemotherapy or HMA therapy in clinical trials. Indeed, many clinical trials (NCT03682029, NCT03999723) are currently investigating the effects of ascorbate alone or as an add-on to classic therapeutic schemes of AML and myeloid disorders. Beyond HMA, speciﬁc lines of research are exploring the possibility of combining ascorbate with class I/II histone deacetylases (HDAC) (Zhang et al., 2017) or sirtuin activators (Sun et al., 2018), with the rationale of regulating TET dioxygenase-dependent effects of vitamin C (Guan et al., 2021). A peculiar application currently under investigation is the scenario of clonal hematopoiesis of indeterminate potential (CHIP) or its age-related counterpart ARCH (age-related clonal hematopoiesis) (Miller and Steensma, 2020). To this end, a recent study showed that elderly individuals with inadequate vitamin C dietary intakes and plasma concentrations had higher odds of ARCH, typically TET2-related (Chen et al., 2021).

By this virtue, ascorbate could provide therapeutic opportunities able to overcome the TET2 impairments typical of myeloid neoplasms by re-establishing the net, residual TET-dioxygenase activity. Indeed, it is known that when TET2 expression is downregulated, two other dioxygenases (TET1, and especially TET3) maintain a minimal enzymatic activity critical for cell survival (Gurnari et al., 2022). Given the current evidence, newer approaches of personalized medicine should take into account not only cytogenetic and mutational characteristics but also both transcriptomic changes (e.g., RNA-seq) and assessment of vitamin C levels in patients at AML onset. This information would enable the identiﬁcation of therapeutic vulnerabilities in individual patients (e.g., ascorbate in TET2-IDH-WT1 impairment and/or vitamin C-deﬁcient patients). Ideally, with the use of sophisticated methods of artiﬁcial intelligence, one could speculate that the in silico creation of “digital twins” would allow testing of multiple, combinatorial therapeutic strategies (including ascorbate, conventional cytotoxic, HMA or new targeted agents), ultimately providing the best drugs to combine with this agent in a “synthetic lethally” fashion (Björnsson et al., 2019).

That said, the exact position of ascorbate in the therapeutic arsenal of AML is yet to be clearly deﬁned. Future data derived from ongoing clinical trials will shed light on its role within the treatment algorithm of AML, a disease where still only less than 30% of patients become long-term survivors and for which new treatment options are urgently needed.

CONCLUSION

The therapeutic potential of ascorbic acid in leukemia have been known for several decades. In particular, the observation that leukemic patients display low vitamin C plasma levels, due to the increased uptake by the actively proliferating leukocytes (Stephens and Hawley, 1936; Kyhos et al., 1945; Barkhan and Howard, 1958) suggested the rationale for the use of high-doses of ascorbic acid, not only as a prophylactic measure, but also to treat

**FIGURE 4** | Activity of pharmacologic doses of ascorbic acid in AML with TET2, IDH1/2 or WT1 altered pathways. (A) TET2 mutations result in a nonfunctional enzyme with hypermethylation of gene promoters, and prevention of oxidation of 5-methylcytosine (SmC) to 5-hydroxymethylcytosine (ShmC). (B) The presence of gain-of-function mutations in IDH1/2 genes results in the overproduction of the oncometabolite 2-hydroxyglutarate (2-HG) with the inhibition of TET2 activity. (C) WT1 mutations hamper the ability of TET2 to bind and activate WT1, inhibiting the expression of WT1-target genes. (D) Ascorbic acid treatment mimics TET2 restoration, inducing a reversal of defective DNA methylation and cell differentiation, ultimately inhibiting tumor progression. Created with Biorender.com.
a number of pathologic conditions, including cancer (Pauling, 1980). Its role as an anti-cancer agent has long been debated and the identification of potential mechanisms through which ascorbate exerts its biologic and pharmacologic activities, lead to hypothesize a new window of therapeutic opportunities.

Recent investigations highlighted the role of ascorbic acid as a critical regulator of cellular epigenetic processes, and a potential drug in the therapeutic armamentarium of acute myeloid leukemia through its stimulatory effect on TET2 function. Being epigenetic dysregulation a hallmark of AML and playing such a central role in the initiation and maintenance of the disease, the possibility to overcome the dysregulated gene-expression programs through an ascorbate based-epigenetic therapy represents a promising and cost-effective anti-leukemia approach.

Overall, these findings suggest the clinical benefit that could derive from the use of ascorbic acid both as a dietary supplement or as a therapeutic agent. Despite the wide number of in vitro experiments demonstrates the anti-leukemia activity of ascorbic acid, the lack of robust evidences about the precise mechanism of action, the tolerability and timing of pharmacological doses of ascorbic acid in vivo did not allow to design appropriate clinical trials. Future clinical trials are warranted to identify patients and specific AML subgroups who may benefit the most from this therapeutic strategy.

**AUTHOR CONTRIBUTIONS**

ST, CG, and MV composed, edited and finalized the review. SA, GS, and TO designed the figure and reviewed the text. All authors contributed to the article and approved the submitted version.

**FUNDING**

This work was supported by AIRC 5x1000 call “Metastatic disease: the key unmet need in oncology” to MYNERVA project, #21267 (Myeloid Neoplasms Research Venture AIRC. A detailed description of the MYNERVA project is available at http://www.progettoagimm.it), PRIN grant No. 2017WXR7ZT to MV, and Ministero della Salute, Rome, Italy (Finalizzata 2018, NET-2018-12365935, Personalized medicine program on myeloid neoplasms: characterization of the patient’s genome for clinical decision making and systematic collection of real world data to improve quality of health care).

**REFERENCES**

Agathocleous, M., Meacham, C. E., Burgess, R. J., Piskounova, E., Zhao, Z., Crane, G. M., et al. (2017). Ascorbate Regulates Haematopoietic Stem Cell Function and Leukaemogenesis. *Nature* 549, 476–481. doi:10.1038/nature23876

Aguilera, O., Muñoz-Sagastibelza, M., Torrejón, B., Borrero-Palacios, A., Del Puerto-Navedo, L., Martínez-Users, J., et al. (2016). Vitamin C Uncouples the Warburg Metabolic Switch in KRAS Mutant Colon Cancer. *Oncotarget* 7, 47954–47965. doi:10.18632/oncotarget.10087

Aldoss, I., Mark, L., Vrona, J., Ramezani, L., Weitz, I., Mohrlander, A. M., et al. (2014). Adding Ascorbic Acid to Arsenic Trioxide Produces Limited Benefit in Patients with Acute Myeloid Leukemia Excluding Acute Promyelocytic Leukemia. *Ann. Hematol. 93*, 1839–1843. doi:10.1007/s00277-014-2124-y

Awada, H., Durmaz, A., Gumrak, C., Kishitogari, A., Meggendorfer, M., Kerr, C. M., et al. (2021). Machine Learning Integrates Genomic Signatures for Subclassification beyond Primary and Secondary Acute Myeloid Leukemia. *Blood* 138, 188–195. doi:10.1182/blood.2020010603

Banella, C., Catalano, G., Travaglini, S., Divona, M., Masciarelli, S., Guerrera, G., et al. (2019). PML/RARA Interferes with NR2F Transcriptional Activity Increasing the Sensitivity to Ascorbate of Acute Promyelocytic Leukemia Cells. *Cancers* 12, 95. doi:10.3390/cancers12010095

Banella, C., Catalano, G., Travaglini, S., Pelosi, E., Ottone, T., Zaza, A., et al. (2022). Ascorbate Plus Buformin in AML: A Metabolic Targeted Treatment. *Cancers* 14, 2565. doi:10.3390/cancers14102565

Barja, G., López-Torres, M., Pérez-Campo, R., Rojas, C., Cadenas, S., Prat, J., et al. (1994). Dietary Vitamin C Decreases Endogenous Protein Oxidative Damage, Malondialdehyde, and Lipid Peroxidation and Maintains Fatty Acid Unsaturation in the guinea Pig Liver. *Free Radic. Biol. Med.* 17, 105–115. doi:10.1016/0891-5849(94)90108-2

Barkhan, P., and Howard, A. N. (1958). Distribution of Ascorbic Acid in Normal and Leukaemic Human Blood. *Biochem. J.* 70, 163–168. doi:10.1042/bj0700163

Bewersdorf, J. P., Shallis, R., Stahl, M., and Zeidan, A. M. (2019). Epigenetic Therapy Combinations in Acute Myeloid Leukemia: what Are the Options? *Ther. Adv. Hemotol.* 10, 2040627118816697. doi:10.1177/2040627118816698

Biewald, T. W., Shih, R., Stahl, M., and Zeitlin, A. M. (2015). Mechanism of the Disproportionation of Ascorbate Radicals. *J. Am. Chem. Soc.* 103, 3516–3518. doi:10.1021/ja50402a042

Björnsson, B., Borrebaeck, C., Borrebaeck, C., Elander, N., Gaslander, T., Gavel, D. R., et al. (2019). Digital Twins to Personalize Medicine. *Genome Med.* 12, 4. doi:10.1186/s13073-019-0701-3

Blaschke, K., Ebata, K. T., Karimi, M. M., Zepeda-Martinez, J. A., Goyal, P., Mahapatra, S., et al. (2013). Vitamin C Induces Tet-dependent DNA Demethylation and a Blastocyst-like State in ES Cells. *Nature* 500, 222–226. doi:10.1038/nature12362

Braun, T., Carvalho, G., Fabre, C., Grosjean, J., Fenaux, P., and Kroemer, G. (2006). Targeting NF-κb in Hematologic Malignancies. *Cell. Death Differ.* 13, 748–758. doi:10.1038/sj.cdd.4401874

Cadenas, S., Rojas, C., and Barja, G. (1998). Endotoxin Increases Oxidative Injury to Proteins in guinea Pig Liver: Protection by Dietary Vitamin C. *Pharmacol. Toxicol.* 82, 11–18. doi:10.1111/j.1749-6632.1998.tb01391.x

Cameron, E., Pauling, L., and Leibovitz, B. (1979). Ascorbic Acid and Cancer: a Review. *Cancer Res.* 39, 663–681

Cameron, E., and Rotman, D. (1972). Ascorbic Acid, Cell Proliferation, and Cancer. *Lancet* 299, 542. doi:10.1016/s0140-6736(72)90215-2

Caprile, T., Salazar, K., Astuya, A., Cisternas, P., Silva-Alvarez, C., Montecinos, H., et al. (2009). The Nar-dependent L-Ascorbic Acid Transporter SVCT2 Expressed in Brainstem Cells, Neurons, and Neuroblastoma Cells Is Inhibited by Flavonoids. *J. Neurochem.* 108, 563–577. doi:10.1111/j.1471-4159.2008.05788.x

Caraballoso, M., Sacristan, M., Serra, C., and Bonfill Cosp, X. (2003). Drugs for Preventing Lung Cancer in Healthy People. *Cochrane database Syst. Rev.*, CD002141. doi:10.1002/14651858.CD002141

Castelli, G., Pelosi, E., and Testa, U. (2018). Targeting Histone Methyltransferase and Demethylase in Acute Myeloid Leukemia Therapy. *Ott Vol.* 11, 131–155. doi:10.2147/OTT.S145971

Chatterjee, I. B., Majumder, A. K., Nandi, B. K., and Subramanian, N. (1975). Synthesis and Some Major Functions of Vitamin C in Animals. *Ann. N. Y. Acad. Sci.* 258, 24–47. doi:10.1111/j.1749-6632.1975.tb29266.x

Chen, J., Guo, L., Zhang, L., Wu, H., Yang, J., Liu, H., et al. (2013). Vitamin C Modulates TET1 Function during Somatic Cell Reprogramming. *Nat. Genet.* 45, 1504–1509. doi:10.1038/ng.2807

Chen, J., Nie, D., Wang, X., Wang, L., Wang, F., Zhang, Y., et al. (2021). Enriched Clonal Hematopoiesis in Seniors with Dietary Vitamin C Inadequacy. *Clin. Nutr. ESPEN* 46, 179–184. doi:10.1016/j.clnesp.2021.10.014
Chen, Q., Espey, M. G., Krishna, M. C., Mitchell, J. B., Corpe, C. P., Buettner, G. R., et al. (2005). Pharmacologic Ascorbic Acid Concentrations Selectively Kill Cancer Cells: Action as a Pro-drug to Deliver Hydrogen Peroxide to Tissues. Proc. Natl. Acad. Sci. U.S.A. 102, 13604–13609. doi:10.1073/pnas.050390102

Chen, Q., Espey, M. G., Sun, A. Y., Pooput, C., Kirk, K. L., Krishna, M. C., et al. (2008). Pharmacologic Doses of Ascorbate Act as a Prooxidant and Decrease Growth of Aggressive Tumor Xenografts in Mice. Proc. Natl. Acad. Sci. U.S.A. 105, 11105–11109. doi:10.1073/pnas.0804226105

Chung, T.-L., Brena, R. M., Kolle, G., Grimmond, S. M., Berman, B. P., Laird, P. W., Flashman, E., Davies, S. L., Yeoh, K. K., and Scher, J., et al. (2015). Vitamin C Promotes Widespread yet Specific DNA Demethylation of the Epigenome in Human Embryonic Stem Cells. Stem Cells 28, 1848–1855. doi:10.1002/stem.2493

Cimmino, L., Dolgalev, I., Wang, Y., Yoshimi, A., Martin, G. H., Wang, J., et al. (2017). Restoration of TET2 Function Blocks Abrupt Self-Renewal and Leukemia Progression. Cell. 170, 1079–1095. doi:10.1016/j.cell.2017.07.032

Cimmino, L., Neel, B. G., and Aifantis, I. (2018). Vitamin C in Stem Cell Reprogramming and Cancer. Trends Cell. Biol. 28, 698–708. doi:10.1016/j.tch.2018.04.001

Corpe, C. P., Eck, P., Wang, J., Al-Hasani, H., and Levine, M. (2013). Intestinal Dehydroascorbic Acid (DHA) Transport Mediated by the Facilitative Sugar transporter, GLUT2 and GLUT8. J. Biol. Chem. 288, 9092–9101. doi:10.1074/jbc.M112.436790

Cronin, E. R., Burgess, R. E., Roys, J. A., Phillips, E., Vissers, M. C. M., and Dachs, G. U. (2021). The Role of 2-Oxoglutarate Dependent Dioxigenases in Gliomas and Glioblastomas: A Review of Epigenetic Reprogramming and Hypoxic Response. Front. Oncol. 11, 619300. doi:10.3389/fonc.2021.619300

Das, A. B., Kakadia, P. M., Wojcik, D., Pemberton, L., Brottet, P. J., Bohlander, S. K., et al. (2019). Clinical Remission Following Ascorbate Treatment in a Case of Acute Myeloid Leukemia with Mutations in TET2 and WT1. Blood Cancer J. 9, 82. doi:10.1038/s41408-019-0242-4

Das, A. B., Smith-Diaz, C. C., and Vissers, M. C. M. (2021). Emerging Epigenetic Therapeutics for Myeloid Leukemia: Modulating Demethylase Activity with Ascorbate. haematol. 106, 14–25. doi:10.3324/haematol.2020.259283

Delhommeau, F., Dupont, S., Vallez, V. D., James, C., Trannoy, S., Massé, A., et al. (2009). Mutation in TET2 in Myeloid Cancers. N. Engl. J. Med. 360, 2289–2301. doi:10.1056/NEJMoai0810069

Dickson, K. M., Gustafson, C. B., Young, J. I., Züchner, S., and Wang, G. (2013). Ascorbate-induced Generation of 5-hydroxymethylcytosine Is Unaffected by Varying Levels of Iron and 2-oxoglutarate. Biochem. Biophysical Res. Commun. 439, 522–527. doi:10.1016/j.bbrc.2013.09.010

Du, J., Cullen, J. J., and Buettner, G. R. (2012). Ascorbic Acid: Chemistry, Biology and the Treatment of Cancer. Biochimica Biophysica Acta (BBA) - Rev. Cancer 1826, 443–457. doi:10.1016/j.ccr.2010.11.015

Du, J., Wagner, B. A., Buettner, G. R., and Cullen, J. J. (2015). Role of Labile Iron in the Toxicity of Pharmacological Ascorbate. Free Radic. Biol. Med. 84, 289–295. doi:10.1016/j.freeradbiomed.2015.03.033

Duarte, T. L., and Lunej, J. (2005). ReviewPart of the Series: From Dietary Antioxidants to Regulators in Cellular Signalling and Gene ExpressionReview: When Is an Antioxidant Not an Antioxidant? A Review of Novel Actions and Reactions of Vitamin C. Free Radic. Res. 39, 671–686. doi:10.1080/10715760501040025

Esteban, M. A., Wang, T., Qin, B., Yang, J., Qin, D., Cai, J., et al. (2010). Vitamin C Enhances the Generation of Mouse and Human Induced Pluripotent Stem Cells. Cell Stem. Cell. 6, 71–79. doi:10.1016/j.stem.2009.12.001

Fennell, K. A., Bell, C. C., and Dawson, M. A. (2019). Epigenetic Therapies in Acute Myeloid Leukemia: where to from Here? Blood 134, 1891–1901. doi:10.1182/blood.2019003262

Ferrada, L., Magdalena, R., Barahona, M. I., Ramirez, E., Sanzana, C., Gutierrez, J., et al. (2021). Two Distinct Faces of Vitamin C: AA vs. DHA. Antioxidants 10, 215. doi:10.3390/antiox10020215

Figuerola, M. E., Abdel-Wahab, O., Lu, C., Ward, P. S., Patel, J., Shih, A., et al. (2010). Leukemic IDH1 and IDH2 Mutations Result in a Hypermethylation Phenotype, Disrupt TET2 Function, and Impair Hematopoietic Differentiation. Cancer Cell. 18, 553–567. doi:10.1016/j.cccr.2011.01.015

Flashman, E., Davies, S. L., Yeoh, K. K., and Schofield, C. J. (2010). Investigating the Dependence of the Hypoxia-Inducible Factor Hydroxylases (Factor Inhibiting HIF and Prolyl Hydroxylase Domain 2) on Ascorbate and Other Reducing Agents. Biochem. J. 427, 135–142. doi:10.1042/BJ20091609
Implications for O2 Sensing. Science 292, 464–468. doi:10.1126/science.1039847

Jakkola, P., Mole, D. R., Tian, Y.-M., Wilson, M. L., Gielbert, J., Gaskell, S. J., et al. (2001). Targeting of HIF-a to the von Hippel-Lindau Ubiquitlation Complex by O2-Regulated Prolyl Hydroxylation. Science 292, 468–472. doi:10.1126/science.1059796

Kakhlon, O., and Cabantchik, Z. I. (2002). The Labile Iron Pool: Characterization, Measurement, and Participation in Cellular Processes. I. This Article Is Part of a Series of Reviews on ‘Iron and Cellular Redox Status.’ The Full List of Papers May Be Found on the Homepage of the Journal. Free Radic. Biol. Med. 33, 1037–1046. doi:10.1016/S0891-5849(02)00106-7

Kawada, H., Kaneko, M., Sawanobori, M., Uno, T., Matsuzawa, H., Nakamura, Y., et al. (2013). High Concentrations of L-Ascorbic Acid Specifically Inhibit the Growth of Human Leukemic Cells via Downregulation of HIF-1α Transcription. PLoS One 8, e62717. doi:10.1371/journal.pone.0062717

Kisieling, M. K., Klemke, C. D., Kaminiski, M. M., Galani, I. E., Krammer, P. H., and Guelow, K. (2009). Inhibition of Constitutively Activated Nuclear Factor-KB Induces Reactive Oxygen Species- and Iron-dependent Cell Death in Cutaneous T-Cell Lymphoma. Cancer Res. 69, 2365–2374. doi:10.1158/0008-5472.CAN-08-3221

Kimura, H., Yamada, Y., Morita, Y., Ikeda, H., and Matsu, T. (1992). Dietary Ascorbic Acid Depresses Plasma and Low Density Lipoprotein Lipid Peroxidation in Genetically Scorbatic Rats. J. Nutr. 122, 1904–1909. doi:10.1093/1945.002103000530006

Kuiper, C., and Vissers, M. C. M. (2014). Ascorbate as a Co-factor for Fe- and 2- Oxidative Stress. Curr. Opin. Struct. Biol. 20, 659–672. doi:10.1016/j.sbi.2010.08.006

Miller, P. G., and Steensma, D. P. (2010). Implications of Clonal Hematopoiesis for Precision Oncology. JCO Precis. Oncol. 4, 639–646. doi:10.1202/PO.2001144

Mingay, M., Chaturvedi, A., Bilenisky, M., Cao, Q., Jackson, L., Hui, T., et al. (2018). Vitamin C-Induced Epigenomic Remodelling in IDH1 Mutant Acute Myeloid Leukemia. Leukemia 32, 11–20. doi:10.1038/leu.2017.171

Minor, E. A., Court, B. L., Young, J. I., and Wang, G. (2013). Ascorbate Induces Ten-Eleven Translocation (Tet) Methylcytosine Dioxygenase-Mediated Generation of 5-hydroxymethylcytosine. J. Biol. Chem. 288, 13669–13674. doi:10.1074/jbc.C113.468000

Mooney, J. N., and Cotter, T. G. (2018). ROS Signalling in the Biology of Cancer. Seminars Cell. & Dev. Biol. 80, 50–64. doi:10.1016/j.semcdb.2017.05.023

Nakajima, H., and Kunimoto, H. (2017). TET2 as an Epigenetic Master Regulator for Normal and Malignant Hematopoiesis. Cancer Sci. 105, 1093–1099. doi:10.1111/cas.12484

Nannya, Y., Shinohara, A., Ichikawa, M., and Kurokawa, M. (2014). Serial Profile of Vitamins and Trace Elements during the Acute Phase of Allogeneic Stem Cell Transplantation. Biol. Blood Marrow Transplant. 20, 430–434. doi:10.1016/j.bbmt.2013.12.554

Nauman, G., Gray, J., Parkinson, R., Levine, M., and Paller, C. (2018). Systematic Review of Intravenous Ascorbate in Cancer Clinical Trials. Antioxidants 7, 89. doi:10.3390/antiox7030086

Ngo, B., Van Riper, J. M., Cantley, L. C., and Yun, J. (2019). Targeting Cancer Vulnerabilities with High-Dose Vitamin C. Nat. Rev. Cancer 19, 271–282. doi:10.1038/s41568-019-0135-7

Noguera, N. I., Pelosi, E., Angelini, D. F., Piredda, M. L., Guerrera, G., Piras, E., et al. (2017). High-dose Ascorbate and Arsenic Trioxide Selectively Kill Acute Myeloid Leukemia and Acute Promyelocytic Leukemia Blasts In Vitro. Oncotarget 8, 32550–32565. doi:10.18632/oncotarget.15925

Noroozi, M., Angerson, W. J., and Lean, M. E. (1998). Effects of Flavonoids and Vitamin C on Oxidative DNA Damage in Cancer Patients. Br. J. Nutr. 80, 32565. doi:10.1079/20016899.646.

Ozer, A., and Bruick, R. K. (2007). Non-herne Dioxygenases: Cellular Sensors and Regulators Jelly Rolled into One? Nat. Chem. Biol. 3, 144–153. doi:10.1038/ncbembio863

Packham, G. (2008). The Role of NF-KB in Lymphoid Malignancies. Br. J. Haematol. 143, 3–15. doi:10.1111/j.1365-2457.2008.07284.x

Papaemmanuil, E., Gerstung, M., Bullinger, L., Gaidzik, V. I., Paschka, P., Roberts, N. D., et al. (2016). Genomic Classification and Prognosis in Acute Myeloid Leukemia. N. Engl. J. Med. 374, 2209–2221. doi:10.1056/NEJMoa1516192

Papaemmanuil, E., Gerstung, M., Malcovati, L., Tauro, S., Gundem, G., Van Loo, P., et al. (2013). Clinical and Biological Implications of Driver Mutations in Myelodysplastic Syndromes. Blood 122, 3616–3627. quiz 3699. doi:10.1182/blood-2013-08-51886.

Park, S., Han, S.-S., Park, C. H., Hahn, E.-R., Lee, S. J., Park, H. K., et al. (2004). L-Ascorbic Acid Induces Apoptosis in Acute Myeloid Leukemia Cells via Hydrogen Peroxide-Mediated Mechanisms. Int. J. Biochem. Cell. Biol. 36, 2180–2195. doi:10.1016/j.biocel.2004.04.005

Park, S. (2013). The Effects of High Concentrations of Vitamin C on Cancer Cells. Nutrients 5, 3496–3505. doi:10.3390/nu5093496

Pastore, F., and Levine, R. I. (2016). Epigenetic Regulators and Their Impact on Therapy in Acute Myeloid Leukemia. Haematologica 101, 269–278. doi:10.3324/haematol.2015.140822

Pauling, L. (1980). Vitamin C Therapy of Advanced Cancer. N. Engl. J. Med. 302, 694–695. doi:10.1056/NEJM198003203021219

Plaum, M., Kielbassa, C., Garmyn, M., and Epe, B. (1998). Oxidative DNA Damage Induced by Visible Light in Mammalian Cells: Extent, Inhibition
Ascorbic Acid Treatment for AML

Staelhe, H. F., Pahl, H. L., and Jurti, J. S. (2021). The Cross Marks the Spot: The Emerging Role of ImC Domain-Containing Proteins in Myeloid Malignancies. Biomolecules 11, 1911. doi:10.3390/biom11121911

Stephens, D. J., and Hawley, E. E. (1936). The Partition of Reduced Ascorbic Acid in Blood. J. Biol. Chem. 115, 653–658. doi:10.1016/S0021-9258(17)47042-2

Strohle, A., Wolters, M., and Hahn, A. (2011). Micronutrients at the Interface between Inflammation and Infection Ascorbic Acid and Calciferol. Part 2: Calciferol and the Significance of Nutrient Supplements. IJAD 10, 64–74. doi:10.2174/18752811794352097

Sun, J., He, X., Zhu, Y., Ding, Z., Dong, H., Feng, Y., et al. (2018). SIRT1 Activation Disrupts Maintenance of Myelodysplastic Syndrome Stem and Progenitor Cells by Restoring TET2 Function. Cell. Stem Cell. 23, 355–369. e9. doi:10.1016/j.stem.2018.07.018

Sweetman, S. F., Strain, J. J., and McKelvey-Martin, V. J. (1997). Effect of Antioxidant Vitamin Supplementation on DNA Damage and Repair in Human Lymphoblastoid Cells. Nutr. Cancer 27, 122–130. doi:10.1080/01635589709514153

Tahiliani, M., Koh, K. P., Shen, Y., Pastor, W. A., Bandukwala, H., Brudno, Y., et al. (2009). Conversion of 5-methylcytosine to 5-hydroxymethylcytosine in Mammalian DNA by MLN Partner TET1. Science 324, 930–935. doi:10.1126/science.1170116

Testa, U., Pelosi, E., and Castelli, G. (2021). New Promising Developments for Personalized Therapeutic Applications of High-Dose Ascorbate as an Anticancer Drug. Hematol. Oncol. Stem Cell Ther. 14, 179–191. doi:10.1016/j.hemos.2020.11.002

The Cancer Genome Atlas Research Network (2013). Genomic and Epigenomic Landscapes of Adult De Novo Acute Myeloid Leukemia. N. Engl. J. Med. 368, 2059–2074. doi:10.1056/NEJMoa1301689

Tsukaguchi, H., Tokui, T., Mackenzie, B., Berger, U. V., Chen, X.-Z., Wang, Y., et al. (1999). A Family of Mammalian Nas-dependent L-Ascorbic Acid Transporters. Nature 399, 70–75. doi:10.1038/19986

Valdés, F. (2006). Vitamin C. Actas Dermosifiliogr. 97, 557–568. doi:10.1016/j.ad.2005.05.001

van der Reest, J., and Gottlieb, E. (2016). Anti-cancer Effects of Vitamin C Revisited. Cell. Res. 26, 269–270. doi:10.1038/cr.2016.7

Vera, J. C., Rivas, C. I., Fischbarg, J., and Golde, D. W. (1993). Mammalian Facilitative Hexose Transporters Mediate the Transport of Dehydroascorbic Acid. Nature 364, 79–82. doi:10.1038/364790a0

Vissers, M. C. M., and Das, A. B. (2018). Potential Mechanisms of Action for Vitamin C in Cancer: Reviewing the Evidence. Front. Physiol. 9, 809. doi:10.3389/fphys.2018.00809

Vissers, M. C. M., Kuiper, C., and Dachs, G. U. (2014). Regulation of the 2-oxoglutarate-dependent Dioxygenases and Implications for Cancer. Biochem. Soc. Trans. 42, 945–951. doi:10.1042/BST20140118

Wang, T., Chen, K., Zeng, X., Yang, J., Wu, Y., Shi, X., et al. (2011a). The Histone Demethylases Hdm1a/1b Enhance Somatic Cell Reprogramming in a Vitamin-C-dependent Manner. Cell. Stem Cell. 9, 575–587. doi:10.1016/j.stem.2011.10.005

Wang, Y., Liu, Y., Malek, S. N., Zheng, P., and Liu, Y. (2011b). Targeting HIF1A Eliminates Cancer Stem Cells in Hematological Malignancies. Cell. Stem Cell. 8, 399–411. doi:10.1016/j.stem.2011.02.008

White, R., Nonis, M., Pearson, J. F., Burgess, E., Morrin, R. H., Pullar, J. M., et al. (2020). Low Vitamin C Status in Patients with Cancer Is Associated with Patient and Tumor Characteristics. Nutrients 12, 2338. doi:10.3390/nut12082338

Wilson, J. K. (2002). The Physiological Role of Dehydroascorbic Acid. FEBS Lett. 527, 5–9. doi:10.1016/S0014-5793(02)03167-8

Wilson, M. K., Baguley, B. C., Wall, C., Jameson, B. M., and Findlay, M. P. (2014). Review of High-Dose Intravenous Vitamin C as an Anticancer Agent. Asia-Pac. J. Clin. Oncol. 10, 22–37. doi:10.1111/ajco.12173

Wohlrab, C., Phillips, E., and Dachs, G. U. (2017). Epigenetics and Approaches to Targeted Epigenetic Therapy in Acute Myeloid Leukemia. Blood 127, 42–52. doi:10.1182/blood-2015-07-604512

Yang, W. S., and Stockwell, B. R. (2008). Synthetic Lethal Screening Identifies Compounds Activating Iron-dependent, Nonapoptotic Cell Death in...
Oncogenic-RAS-Harboring Cancer Cells. *Chem. Biol.* 15, 234–245. doi:10.1016/j.chembiol.2008.02.010

Yi, C., Jia, G., Hou, G., Dai, Q., Zhang, W., Zheng, G., et al. (2010). Iron-catalysed Oxidation Intermediates Captured in a DNA Repair Dioxygenase. *Nature* 468, 330–333. doi:10.1038/nature09497

Yin, R., Mao, S.-Q., Zhao, B., Chong, Z., Yang, Y., Zhao, C., et al. (2013). Ascorbic Acid Enhances Tet-Mediated 5-methylcytosine Oxidation and Promotes DNA Demethylation in Mammals. *J. Am. Chem. Soc.* 135, 10396–10403. doi:10.1021/ja4028346

Yoshimi, A., Lin, K.-T., Wiseman, D. H., Rahman, M. A., Pastore, A., Wang, B., et al. (2019). Coordinated Alterations in RNA Splicing and Epigenetic Regulation Drive Leukaemogenesis. *Nature* 574, 273–277. doi:10.1038/s41586-019-1618-0

Young, J. I., Züchner, S., and Wang, G. (2015). Regulation of the Epigenome by Vitamin C. *Annu. Rev. Nutr.* 35, 545–564. doi:10.1146/annurev-nutr-071714-034228

Yun, J., Mullarky, E., Lu, C., Bosch, K. N., Kavalier, A., Rivera, K., et al. (2015). Vitamin C Selectively Kills KRAS and BRAF Mutant Colorectal Cancer Cells by Targeting GAPDH. *Science* 350, 1391–1396. doi:10.1126/science.aaa5004

Zhang, Y. W., Wang, Z., Xie, W., Cai, Y., Xia, L., Easwaran, H., et al. (2017). Acetylation Enhances TET2 Function in Protecting against Abnormal DNA Methylation during Oxidative Stress. *Mol. Cell.* 65, 323–335. doi:10.1016/j.molcel.2016.12.013

Zhao, H., Zhu, H., Huang, J., Zhu, Y., Hong, M., Zhu, H., et al. (2018). The Synergy of Vitamin C with Decitabine Activates TET2 in Leukemic Cells and Significantly Improves Overall Survival in Elderly Patients with Acute Myeloid Leukemia. *Leukemia Res.* 66, 1–7. doi:10.1016/j.leukres.2017.12.009

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher’s Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Travaglini, Gurnari, Antonelli, Silvestrini, Noguera, Ottone and Voso. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.