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CASE REPORT

Description of ovine model for testing ventilator prototypes in the COVID-19 pandemic

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Abstract The COVID-19 pandemic has revealed a ventilator deficit in the global health system for this scenario. For this reason, several national and international projects have been developed to get done prototypes of ventilators which could be easy and fast to manufacture. One of the requirements of the AEMPS for conducting clinical studies with new prototypes is through the validation of these new prototypes in an animal model. Therefore, it is important to achieve an animal model which allows us to easily reproduce different clinical scenarios. In this article, we describe the use of a sheep as a research model to assess a prototype ventilator. The animal was anesthetized for 10h in which the prototype was tested in up to 6 different scenarios. This model is effective and easy to reproduce, making it an excellent choice for this kind of research.

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PALABRAS CLAVE
Experimento;
Oveja;
Ventilador;
Síndrome de distrés respiratorio agudo

Descripción de un modelo ovino para la prueba de ventiladores de urgencia en la pandemia de COVID-19

Resumen La pandemia COVID-19 ha puesto de manifiesto un déficit de ventiladores en el sistema sanitario para estas situaciones. Por ello, varios proyectos nacionales e internacionales se han desarrollado en pocas semanas para producir prototipos de ventiladores de fácil y rápida...
Introduction

Translational research was developed as a bridge between basic and clinical research1. This step in the development of new drugs, medical equipment or procedures is both a necessity and a legal requirement prior to performing a clinical trial.

Many patients required ventilatory support during the COVID-19 pandemic. According to various sources, up to 15.2%2 of these required the use of mechanical ventilation, and equipment such as intensive care ventilators became a fundamental factor in the treatment of these patients. Given the limited stock and manufacturing capacity of intensive care ventilators, hospitals were faced with the risk of ventilator shortages. Several national and international ventilator design and manufacturing projects have been launched to increase supplies. Most of these projects are aimed at developing open-source designs for low-cost ventilators that can be easily and rapidly manufactured in any country.

In this emergency situation, the Spanish Agency of Medicines and Medical Devices (AEMPS), the competent authority in Spain, published a resolution setting out the requirements for approving a ventilator prototype3. In addition to a technical description of the prototype and tests on a simulation lung, the agency required that the prototype also be tested on an animal model. However, the resolution only defines the animal model (porcine), but does not specify the characteristics of the experiment. This meant that potentially valid animal models were initially ruled out, and the minimum monitoring parameters to ensure the safety of the prototype were not defined.

In this article, we describe the animal model we used to test an open source ventilator developed during the COVID-19 pandemic. We hope that our experience will serve as a reference for other research groups.

Case report

The Animal Ethics and Experimentation Committee of the CEU Cardenal Herrera University obtained the approval of the Department of Agriculture, Rural Development, Climate Emergency and Ecological Transition of the Gener-
obtain a tidal volume (VT) of 6–7 ml/kg, respiratory rate 14 rpm, inspiration: expiration ratio 1:2 and FiO2 0.5.

2 Baseline ventilation with neuromuscular blockade. In this phase, atracurium (0.5 mg kg) was administered, and the stage 1 ventilatory parameters were maintained. Both stages lasted 150 min.

3 Respiratory distress model (ARDS). The ARDS model was induced by washing the alveolar surfactant with 60 ml/kg of physiological saline at 37 °C. To do this, 20 ml/kg of tempered saline was administered via a catheter inserted through the endotracheal tube. Following this, coupage was performed and the liquid content was recovered with a surgical aspirator. The off-white liquid collected is compatible with a mixture of saline solution and surfactant. After completing this procedure, the animal was re-connected to the ventilator and monitored to see if the ARDS model had been established. Three lavage procedures were required to achieve the ARDS model. ARDS was confirmed when peripheral arterial oxygen saturation had fallen to below 90%, following which a chest X-ray was performed and arterial blood gas was measured confirm a PaO2/FiO2 of less than 100.

4 Alveolar recruitment and ventilatory strategy. Once the ARDS model had been established, and after waiting 20 min, the following recruitment strategy was performed:

- Alveolar recruitment manoeuvre. Recruitment was performed in CMV-PC mode with a driving pressure (DP) of 15 cmH2O, a respiratory rate of 15 rpm, an inspiration: expiration ratio of 1:1, FiO2 of 0.8 and initial PEEP of 5 cmH2O. PEEP was increased in 5 cmH2O increments every 10 respiratory cycles, and then every 15 cycles when peak PEEP (25 cmH2O) had been reached and an airway opening pressure of 40 cmH2O had been achieved (duration of the manoeuvre: 160 s) (Fig. 2).
- Calculation of optimal PEEP. The working pressure was adjusted to obtain a VT of 6–7 ml/kg, a respiratory rate of 15 rpm, an inhalation:expiration ratio of 1:2, and best Cdyn PEEP, maintaining ventilation for 90 min.

5 Bronchoconstriction model: CMV-PC in a bronchoconstriction model after recovery from respiratory distress. When the animal had been stable for several hours and lung function had been restored, 0.03 mg/kg of iv neostigmine was administered to produce bronchoconstriction, which was confirmed by increased airway resistance on spirometry.

6 Spontaneous ventilation with different continuous positive airway pressure (CPAP) settings. Finally, ketamine, fentanyl, and propofol infusions were down-dosed by 50% to reduce depth of anaesthesia and regain spontaneous activity. At that time, different levels of CPAP were administered and their effects on the monitored parameters were assessed.

Fig. 3 shows the main oxygenation parameters obtained at each stage of the experiment.

Discussion

The choice of model in animal experimentation should be based on criteria of replacement, reduction and refinement. In our case, we were looking for a docile species with characteristics similar to humans that had previously been used in this type of experiment based on an easily reproducible protocol that tested the ventilator at different clinical settings. The model used had to be simple to execute with the usual drugs and techniques used in animal experimentation centres. The AEMPS indicates that the experiment should be performed in a porcine model. However, by the time the standard was published our project was well advanced, and we had already obtained permission to use a sheep model, which, a priori, we considered more suitable. After contacting the AEMPS and justifying the suitability of the sheep model, they authorized it use.

Sheep are gregarious, docile animals that have been domesticated for thousands of years. One of their main advantages over the porcine model is that sheep can be used to explore and administer drugs by any route, including intravenous, without the need for prior sedation.

The sheep model is widely used in translational medicine in different fields of research. Specifically, it has been used
to study different models of respiratory disease and different ventilatory strategies.

Although we might believe, at first glance, that a herbivorous and ruminant species such as the sheep is further removed from humans than other animals widely used in animal experimentation, such as the pig, the anatomy and physiology of the respiratory system of the sheep is more akin to the human system than other species. Sheep models are also very useful in experiments to study respiratory parameters and obtain measurements of respiratory mechanics.

Although the animal in our experiment was anaesthetized for 10h while we performed all the foregoing tests, the only complications observed were those associated with the respiratory distress model, which were resolved with a recruitment manoeuvre using the ventilator prototype. The use of advanced monitoring gave us detailed information on the cardiovascular and respiratory status of the animal at all times. This allowed us to confirm, on the one hand, that our manoeuvres had achieved the desired effect at all times, and on the other, that they did not cause any undesired side effects. Although the degree of monitoring was not defined in the AEMPS resolution, we believe that the foregoing are the minimum parameters that should be monitored when testing a ventilator prototype.

The sheep model has been widely used in the study of acute respiratory distress syndrome. This ARDS model has been induced using various techniques, such as, administering cotton smoke directly into the lung, administering lipopolysaccharides of Escherichia coli, or performing alveolar lavage with saline solution to remove alveolar surfactant phospholipids. The difference between these models depends on the product used to produce the tissue damage and the severity and pathophysiology of the
condition. The latter technique, alveolar lavage with saline, is the one used in our model. This ARDS model has been used in various studies to evaluate alveolar recruitment manoeuvres \textsuperscript{1,2}. The model used in our case is a simple, fast, safe and easily reproducible method to assess the effect of ventilatory manoeuvres. It also allows the syndrome to be resolved, so the animal can potentially be used for other manoeuvres or scenarios, as in this case. This allowed us to optimise the "reduction" principle, one of the 3 Rs formulated by Russel and Burch in England in 1959 that has since become central to animal experimentation\textsuperscript{1}.

The model used here is easy to induce with the usual drugs and techniques used in animal experimentation centres.

Finally, the blood gas measurements shown in the graph show good oxygenation in phases 1 and 2. The PaO\textsubscript{2}/FiO\textsubscript{2} <100 achieved after surfactant alveolar lavage reproduced a pattern of severe acute respiratory distress, and alveolar recruitment manoeuvres significantly improved oxygenation. Finally, oxygenation was also adequate in both phases 5 and 6.

## Conclusion

To sum up, the sheep model used in this experiment fulfilled all the criteria required to assess the ventilator prototype in different clinical settings. Therefore, we believe that the animal model used to test this prototype is easy to use, reproducible, and a single animal can be used to evaluate different clinical scenarios in the current COVID-19 health emergency.

## Conflict of interests

The authors have no conflict of interest to declare.

## References

1. Tannenbaum J, Bennett B. Russell and Burch’s 3Rs then and now: the need for clarity in definition and purpose. J Am Assoc Lab Anim Sci. 2015;54:120–32. PMCID: PMC4382615.
2. Villalonga R, Martín MC, Avilés-Jurado FX, Alvarez J, Aldecoa C, de Haro C, et al. Documento de consenso de la Sociedad Española de Medicina Intensiva, Crítica, y Unidades Coronarias (SEMICYUC), la Sociedad Española de Otorrinolaringología y Cirugía de Cabeza y Cuello (SEORL-CCC) y la Sociedad Española de Anestesiología y Reanimación (SEDAR) sobre la traqueotomía en pacientes con COVID-19. Rev Esp Anestesiol Reanim. 2020, http://dx.doi.org/10.1016/j.redar.2020.05.001.
3. Agencia española de medicamentos y productos sanitarios. Información sobre prototipos de respiradores, pruebas de seguridad y requisitos de investigación clínica. 27 de marzo 2020, https://www.aemps.gob.es/informa/notaslavitas/productosSanitarios/2020/NI-P511-2020-respiradores-pruebas-investigaciones.pdf.
4. Lachmann B, Robertson B, Vogel J. In vivo lung lavage as an experimental model of the respiratory distress syndrome. Acta Anaesthesiol Scand. 1980;24:231–6.
5. Ranieri VM, Rubenfeld GD, et al. Acute respiratory distress syndrome: the Berlin definition. JAMA. 2012;307:2526–33, http://dx.doi.org/10.1001/jama.2012.5669.
6. Ferrando C, Soro M, Canet J, Unzueta MC, Suárez F, Librero J, et al. Rationale and study design for an individualized perioperative open lung ventilatory strategy (iPROVE): study protocol for a randomized controlled trial. Trials. 2015;27:16, http://dx.doi.org/10.1186/s13063-015-0694-1, 193.
7. Meeusen E, Snibson K, Hirst S, Bischof R. Sheep as a model species for the study and treatment of human asthma and other respiratory diseases. Drug Discov Today Dis Models. 2009;6:e101–6, http://dx.doi.org/10.1016/j.ddmod.2009.12.002.
8. Ballard-Croft C, Wang D, Sumpter R, Zhou X, Zwischenberger J, et al. Large-animal models of acute respiratory distress syndrome. Ann Thorac Surg. 2012;93:1331–9, http://dx.doi.org/10.1016/j.athoracsur.2011.06.107.
9. Asmussen S, Ito H, Traber DL, Lee J, Cox R, Hawkins H, et al. Human mesenchymal stem cells reduce the severity of acute lung injury in a sheep model of bacterial pneumonia. Thorax. 2014;69:819–25, http://dx.doi.org/10.1136/thoraxjnl-2013-204980.
10. Cardenes N, Aranda-Valderrama P, Carney JP, Sellares Torres J, Alvarez D, Kocydirim E, et al. Cell therapy for ARDS: efficacy of endobronchial versus intravenous administration and biodistribution of MAPCs in a large animal model. Open Resp Res. 2019:1–10, http://dx.doi.org/10.1136/bmjrresp-2018-000308.
11. Yehya N. Lessons learned in acute respiratory distress syndrome from the animal laboratory. Ann Transl Med. 2019;7:503–15, http://dx.doi.org/10.21037/atm.2019.09.33.
12. Engel M, Nowacki RM, Reiss LK, Uhlig S, Willems CH, Kloosterboer N, et al. Comparison of recruitment manoeuvres in ventilated sheep with acute respiratory distress syndrome. Lung. 2013;191:77–86, http://dx.doi.org/10.1007/s00408-012-9428-2.