Mitochondrial genome of captive Alpine musk deer, Moschus chrysogaster (Moschidae), and phylogenetic analyses with its coordinal species

Chao Yanga, d, Wei-Feng Wangb, Xiao-Juan Duc, Xiao Tanb, Li-Juan Suoa, Kun Bianb, Fei-Ran Lib, Jie Tanga, Ben-Mo Jiangc, Xue-Juan Lid and Yan Wanga

aShaanxi Institute of Zoology, Xi’an, PR China; bShaanxi Nature Reserve and Wildlife Management Station, Xi’an, PR China; cSichuan Fengchun Pharmaceutical Co. Ltd, Chengdu, PR China; dSchool of Life Sciences, Shaanxi Normal University, Xi’an, PR China

ABSTRACT

Alpine musk deer, Moschus chrysogaster, a solitary, primitive ungulate inhabiting high elevation areas (3000–4500 m) is an endangered species facing threat of extinction globally due to excessive hunting for its musk. In this study, we determined the complete mitochondrial genome of M. chrysogaster, which was 16,354 bp in length, and revealed the same gene order and genomic organization as typical Moschidae mitochondrial DNA. Start codons in 13 protein-coding genes (PCGs) were all typical ATGs except ATA for ND2 and ND3 and ATT for ND5. Stop codons were all typical types except an incomplete stop codon T for COX3, ND2, ND3, and ND4. Secondary structures in 22 transfer RNA genes all showed typical cloverleaf except tRNA-Ser(AGY), in which the dihydrouridine arm formed a simple loop. No repeat units were found in the control region. The topology structure indicated that M. cupreus was primitive and located at the root of the Moschidae clade. Phylogenetic reconstruction placed M. chrysogaster as a distinct lineage, closely related to the branch of M. leucogaster, M. berezovskii (wild) and predicted a sister relationship with M. moschiferus, M. anhuiensis, and M. berezovskii (captive). However, we suggested that the genetic resources of M. chrysogaster_JQ608470 should be further investigated.

As an endangered species, Alpine musk deer, Moschus chrysogaster, was listed on the IUCN Red List (Harris 2016) and categorized as a first-degree national protected species in China. This species occurs from the highlands of central China (the Helan Mountains from the northern edge of its distribution), to areas south and west of the Himalayas, extending to eastern Nepal, Bhutan, and northeastern India (Wemmer 1998). It is widely but discontinuously distributed across the mountainous parts of the Himalayas (Aryal et al. 2010). Owing to excessive hunting, as well as habitat loss and degradation, wild populations have been declining for decades (Meng et al. 2003). Only one legal institution exists in China to breed M. chrysogaster in captivity, and efforts are also under way to do so in India (Mithileshwari et al. 2016). To domesticate, genetically improve, and systematically conserve wild populations, it is necessary to define the genetic characteristics of M. chrysogaster (Yang et al. 2013).

The muscle tissue used for DNA extraction and analysis was sampled from a captive male Alpine musk deer, M. chrysogaster, that died of natural causes in Feng County, Shaanxi Province (N: 34.231295, E: 106.903539), and the specimen (voucher number: MS01) was deposited in the animal museum of the Shaanxi Institute of Zoology, Xi’an, China (contacts: Chao Yang, chaoy819@xab.ac.cn).

Total genomic DNA was extracted to construct paired-end libraries, tagged, and subjected to the high-throughput Illumina Xten platform sequencing with a 150 bp paired-end strategy. Clean reads were trimmed by removing regions with a Phred score of <10 and 7,259,638 paired-end raw reads were obtained. Then, MITObim version 1.9 (Hahn et al. 2013) was used to assemble the clean reads with the complete mitochondrial genome (mitogenome) of M. berezovskii (GenBank: MH047347) as a reference. A total of 79,140 individual mitochondrial reads were mapped to the reference mitogenome, giving an average coverage of 709.8X. Geneious version 2020.0.4 (Karse et al. 2012) and the MITOchondrial genome annotation server (MITOS; http://mitos.bioinf.uni-leipzig.de/index.py) were used for annotation of protein-coding genes (PCGs) in the mitogenome, and data were manually inspected to predict transfer RNA (tRNA) and ribosomal RNA (rRNA) genes.

Finally, the whole mitogenome sequence of M. chrysogaster was obtained and submitted to GenBank (accession number MW284875); the genome was 16,354 base pairs (bp) in length, including 13 PCGs, 2 rRNA genes, 22 tRNA genes, and one control region. Gene content and arrangement were identical to those of other typical mammalian mitogenomes (Pan et al. 2015). The base composition of the M. chrysogaster mitogenome was as follows: A = 34%, T = 28%, C = 25.1%,
G = 12.9%, and A + T = 62%. With the exception of ND2 and ND3, which start with ATA, and ND5, which starts with ATT, all PCGs had typical ATG start codons, and all of them ended with a complete triplet codon (TAA and AGA), except for COX3, ND2, ND3, and ND4, which ended with an incomplete T.

The length of 12S rRNA was 956 bp, and that of 16S rRNA was 1569 bp, located between tRNA-Phe and tRNA-Leu(UUR), separated by tRNA-Val. The length of the 22 tRNA genes ranged from 60 bp (tRNA-Ser(AGY)) to 75 bp (tRNA-Leu(UUR)), and all tRNA genes had typical cloverleaf secondary structures, with the exception of tRNA-Ser(AGY), in which the dihydouridine arm formed a simple loop. The control region was 924 bp in length, located between tRNA-Pro and tRNA-Phe, and the AT content was 63.6%. Similar to other musk deer mitogenomes, a tandem repeat was not found in this region (Kim et al. 2017).

To confirm the phylogenetic position of M. chrysogaster, RAxML (Stamatakis 2006), and MrBayes version 3.2.2 (Ronquist et al. 2012) were used on the concatenated datasets of 13 PCGs to reconstruct maximum likelihood (ML) tree and Bayesian inference (BI) tree, by referencing the best-partitioned scheme and optimal model analyzed in Partitionfinder version 1.1.1 (Lanfear et al. 2012). Hippopotamus amphibius (GenBank: AP003425) and Camelus bactrianus (GenBank: EF507798) were selected as outgroups. The ML tree reflected the same structure as the BI tree. The phylogenetic topology of the families obtained from them was congruent with a previous study in which Moschidae was a sister group to Bovidae (Hassanin and Douzery 2003; Yang et al. 2018). In Moschidae, as the last identified species, M. cupreus was at the base and formed a separate clade (Singh et al. 2020).

M. chrysogaster is more primitive and is a sister group to M. moschiferus, M. anhuiensis, and M. berezovskii (captive) (Su et al. 1999). From wild populations to those in captivity, more molecular data should be examined on the genetic diversity and geographical distribution of this species to increase musk reserves. The newly sequenced species obtained from Feng County, China, clustered with M. chrysogaster (GenBank: KP684123, KC425457) and were closely related to the M. leucogaster and M. berezovskii (wild) branch, with 94.60% sequence similarity to M. chrysogaster_JQ608470 (Miyaluo Nature Reserve, Sichuan, China) and with 99.80% and 99.96% sequence similarities to M. chrysogaster_KC425457 (Xinglong Mountain, Gansu, China) and M. chrysogaster_KP684123 (Qinghai Lake, Qinghai, China), respectively (Yang et al. 2013; Pan et al. 2015). Extraordinarily, the status of M. chrysogaster_JQ608470 was indicated either a misidentification or an unidentified hybrid individual (Figure 1).

Disclosure statement
No potential conflicts of interest were reported by the authors.
Funding
The study was supported by the Science and Technology Projects of Shaanxi Academy of Science, China under Grant [No. 2018K-01, 2019K-03] and Projects for the Department of Science and Technology of Shaanxi Province, China under Grant [No. 2018ZDXM-NY-071, 2019NY-089].

Data availability statement
The genome sequence data that support the findings of this study are openly available in NCBI GenBank (https://www.ncbi.nlm.nih.gov/) under accession no. MW284875. The associated BioProject, SRA, and Bio-Sample numbers are PRJNA685635, SRR13258650, and SAMN17088691, respectively.

References
Aryal A, Raubenheimer D, Subedi S, Kattel B. 2010. Spatial habitat overlap and habitat preferences of Himalayan Musk Deer (Moschus chrysogaster) in Sagarmatha (Mt. Everest) National Park, Nepal. Curr Res J Biol Sci. 2:217–225.
Hahn C, Bachmann L, Chevreux B. 2013. Reconstructing mitochondrial genomes directly from genomic next-generation sequencing reads—a baiting and iterative mapping approach. Nucl Acids Res. 41(13):e129–e129.
Harris R. 2016. Moschus chrysogaster. The IUCN red list of threatened species. [accessed 2020 Nov 24]. https://dx.doi.org/10.2305/IUCN.UK.2016-1.RLTS.T13895A61977139.en
Hassanin A, Douzery EJ. 2003. Molecular and morphological phylogenies of Ruminantia and the alternative position of the Moschidae. Syst Biol. 52(2):206–228.
Keare M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, et al. 2012. Geneious basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. Bioinformatics. 28(12):1647–1649.
Kim SI, Lee MY, Jeon HS, Han SH, An J. 2017. Complete mitochondrial genome of Siberian musk deer Moschus moschiferus (Artiodactyla: Moschidae) and phylogenetic relationship with other moschus species. Mitochondrial DNA Part B. 2(2):860–861.
Lanfear R, Calcott B, Ho SY, Guindon S. 2012. Partitionfinder: combined selection of partitioning schemes and substitution models for phylogenetic analyses. Mol Biol Evol. 29(6):1695–1701.
Meng XX, Yang QS, Xia L, Feng ZJ, Jiang YW, Wang PM. 2003. The temporal estrous patterns of female alpine musk deer in captivity. Appl Anim Behav Sci. 82(1):75–85.
Mithileshwari C, Srivastava T, Kumar V, Kumar A, Umamathy G. 2016. Non-invasive assessment of fecal prostagstags and pregnancy detection in Himalayan musk deer (Moschus chrysogaster). Theriogenology. 85(2):216–223.
Pan T, Wang H, Hu CC, Sun ZL, Zhu XX, Meng T, Meng XX, Zhang BW. 2015. Species delimitation in the genus Moschus (Ruminantia: Moschidae) and its high-plateau origin. PLOS One. 10(8):e0134183.
Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Syst Biol. 61(3):539–542.
Singh B, Gautam KB, Sahoo S, Kumar A, Gupta SK. 2020. Complete mitogenome of Kashmir musk deer (Moschus cupreus) and its comparative phylogenetic relationships. Mol Biol Rep. 47(12):9757–9763.
Stamatakis A. 2006. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics. 22(1):2688–2690.
Su B, Wang YX, Lan H, Wang W, Zhang YP. 1999. Phylogenetic study of complete cytochrome b genes in musk deer (genus Moschus) using museum samples. Mol Phylogenet Evol. 12(3):241–249.
Wemmer C. 1998. Deer status survey and conservation action plan. Gland, Switzerland and Cambridge: IUCN.
Yang C, Tang J, Bian K, Suo LJ, Yuan H, Wang Y, Huang Y. 2018. Next generation sequencing yields the complete mitogenome of captive forest musk deer, Moschus berezovskii (Ruminantia: Moschidae). Mitochondrial DNA Part B. 3(2):472–473.
Yang CZ, Xiang CK, Zhang XY, Yue BS. 2013. The complete mitochondrial genome of the Alpine musk deer (Moschus chrysogaster). Mitochondrial DNA Part A. 24(5):501–503.