NMR-based plasma metabolic profiling in patients with unstable angina

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A B S T R A C T

Objective(s): Unstable angina (UA) is a form of the acute coronary syndrome (ACS) that affects more than a third of the population before age 70. Due to the limitations of diagnostic tests, appropriate identification of UA is difficult. In this study, we proceeded to investigate metabonome profiling in UA patients compared with controls to determine potential candidate biomarkers.

Materials and Methods: Ninety-four plasma samples from UA and 32 samples from controls were analyzed based on 1H NMR spectroscopy. The raw data were processed, analyzed, and subjected to partial least squares-discrimination analysis (PLS-DA), a supervised classification method with a good separation of control and UA patients was observed. The most important variables (VIP) ≥1 were selected and submitted to MetaboAnalyst pathway enrichment to identify the most important ones.

Results: We identified 17 disturbed metabolites in UA patients in comparison with the controls. These metabolites are involved in various biochemical pathways such as steroid hormone biosynthesis, aminoacyl-tRNA biosynthesis, and lysine degradation. Some of the metabolites were deoxycorticosterone, 17-hydroxyprogesterone, androstenedione, androstanedione, etiocholanolone, estradiol, 2-hydroxyestradiol, 2-hydroxyestrone, 2-methoxyestradiol, and 2-methoxyestrone. In order to determine test applicability in diagnosing UA, a diagnostic model was further created using the receiver operating characteristic (ROC) curve. The areas under the curve (AUC), sensitivity, specificity, and precision were 0.87, 90%, 65%, and 91%, respectively, for diagnosing of UA.

Conclusion: These metabolites could not only be useful for the diagnosis of UA patients but also provide more information for further deciphering of the biological processes of UA.

Introduction

Acute coronary syndrome (ACS) is a type of coronary artery disease (CAD) that refers to a range of conditions compatible with acute myocardial ischemia and/or infarction due to a sudden reduction in coronary blood flow (1). ACS is classified into myocardial infarction (MI) and unstable angina (UA). UA is a condition in which the heart does not get enough blood and oxygen supply. This condition may be dangerous and can lead to a heart attack. (2-5). The mortality rate in patients with UA was reported at 4.5% after 30 days of MI. Often, more tissue injury is required for the detection of myocardial injury because CK-MB is less sensitive than troponin (6). For MI, more tissue injury is required for the detection of myocardial injury because CK-MB is much less sensitive than troponin (10). Besides, exercise testing with myocardial perfusion imaging is relatively accurate, but it is costly (11), and slow in terms of speed for an emergency department (12). UA is not recognizable in the early stages, and symptoms alone are not adequate to distinguish between different ACS types (13). Due to the above limitations, finding new biomarkers using non-invasive techniques is an urgent
need. Advances in "omic" technologies have advantages such as sensitivity, speed, and robustness for identifying new biomarkers in cardiovascular diseases (CVD) (9).

One of the new "omic" approaches is the field of metabolomics. To understand the disease, the metabolism provides a "snapshot" of all metabolites in a biological sample such as blood, plasma, serum, urine, and many other samples that may be obtained from the patients or the experimental models (14). Metabolites are described as primitive reporters of disease because their increase in biological specimens are often directly associated with pathogenic mechanisms (15). One of the highly used techniques is the proton NMR (1H NMR), which has functions for analysis of urine and plasma specimens. Some of the benefits of NMR include no need for degradation and chemical manipulation of specimens and also providing accurate information from the molecular concentrations and structures of metabolites (16). Contrary to genes and proteins that are regulated epigenetically and by post-translational modifications, respectively, metabolites serve as direct signatures of biochemical activity and hence are more easily associated with phenotypes (17). Metabolic changes due to a particular disease can be detected in biological fluids before clinical symptoms develop and produce useful fingerprints (18). Metabolomics has been served for new biomarker discovery and consequently better diagnosing and characterizing of disease, including cancers (brain, lung, ovarian, and breast), neurological disorders (Alzheimer), inborn errors in metabolism (argininosuccinic aciduria, tyrosinemia type II, homocystinuria, and phenylketonuria), diabetes mellitus, tuberculosis, and others (19, 20).

Metabolomics will increase our understanding of the pathophysiological processes involved in UA and help us to identify potential novel biomarkers to develop new diagnostic strategies (16, 21). In the past, there have been few studies investigating the metabolomics approach for identifying novel biomarkers in UA patients in comparison with healthy controls (22-25). In this study, we have used 1H NMR-based spectrometry metabolic profiling of UA patients to find possible candidate metabolites and to provide useful information about important altered metabolic pathways. Our study was the first study carried out in Iran on UA using 1H NMR. Also, this is the first study in which obstruction has been measured in all three coronary heart arteries. More importantly, the first study is on which controls had autosomal dominant coronary arteries. The demographic data for the patients and controls and was only applied to the symptoms of the disease and the wave of S-T segment changes (24).

**Materials and Methods**

**Clinical characteristics of patients**

From September 2017 to May 2018, we recruited patients with symptoms of UA who referred to the emergency department at Mousavi Hospital and Rohani Hospital, Zanjan and Babol, Iran, respectively. All patients underwent coronary angiography and the percentage of obstruction was determined in the three main coronary arteries: Left Anterior Descending (LAD), Left Circumflex Artery (LCX), and Right Coronary Artery (RCA). From the patients with UA symptoms who were a candidate for coronary artery bypass grafting (CABG) and percutaneous coronary intervention (PCI), we selected 94 UA patients based on having an obstruction in any of the coronary arteries, and also 32 controls that did not have any obstruction in their coronary arteries. The demographic data for the patients with UA and the controls are shown in Table 1.

**Ethics, consent, and permissions**

Diagnosis criteria of UA refer to the ‘‘2014 AHA/ACC guideline for the management of patients with Non–ST-Elevation Acute Coronary Syndromes’’. All stages were carried out in accordance with the Helsinki Declaration. The study was ethically approved by the ethics committee of the Zanjan University of Medical Science (ethics approval code, ZUMS.REC.1396.98). Informed consent was taken from all patients before their inclusion in the study.

**Inclusion, exclusion, and rejection criteria**

All selected UA and the controls were diagnosed by physical examination, cardiac biomarkers, and confirmed by coronary angiography. Patients with obstruction over 75% in one of the three main coronary arteries (LAD, LCX, or RCA) were included in the UA group. Also, patients who were accepted as UA patients but in angiography did not have any obstruction in coronary arteries were included in the control group. The exclusion criteria include a history of heart, liver, or metabolic disease, diabetes, rheumatism, cancer, hematopoietic system, pulmonary embolism, autoimmune disorders, severe infectious diseases, trauma, and a recent surgical procedure. Also, patient samples whose files were not available or lacked clinical information were excluded from the study.

**Sample collection, preparation, and NMR spectroscopy**

Five-milliliter blood samples were taken from patients with UA symptoms and placed in tubes containing EDTA as an anticoagulant and immediately centrifuged (2-6E, Sigma, Germany) at 3500 rpm for 10 min to separate plasma. The plasma samples were stored at -80 °C until NMR analysis. The 1H NMR data were acquired at room temperature on a Bruker 400 NMR spectrometer using Carr–Purcell–Meiboom–Gill (CPMG) pulse program (27). In brief, 400 μl of plasma samples were mixed with 100 μl of D.O. For calibration of the chemical shift, Tetramethylsilane (TMS) was applied as the internal reference in NMR tubes.

**Pre-processing and Multivariate statistical analysis of 1H NMR spectra**

All NMR data in the FID time domain were imported into the MestReNova software package (version 6.0.2-5475) for data processing. The FID data were subjected to Fourier transform to obtain the spectra in the frequency domain. Then data were phased and baseline-corrected and exported as an ASCII file and imported into MATLAB.
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(R2008a; Mathworks, Natick, MA) for further analysis. To eliminate water resonance, the spectral regions from 4.7 to 5 ppm were removed. In the next step, each $^1$H NMR spectrum from plasma was segmented into equal widths (0.003 ppm), corresponding to regions 0–9 ppm. Figure 1 shows the final binned $^1$H NMR spectra of all samples. Then the spectral data were mean-centered and scaled according to the Pareto scaling method (28, 29).

A home-written program in MATLAB was used for multivariate statistical analysis. After pre-processing of data, principal component analysis (PCA) was applied to data as an unsupervised classification method. Since the separation of groups was not good (data not shown), PLS-DA as a supervised classification method was applied.

The corresponding loading plot from PLS-DA was obtained, and the important variables were selected according to VIP≥1 (Figure 2).

Figure 3 shows the score plot of data from PLS-DA. The proximity of patient and control profiles can be a good reason for some patients to overlap in the PCA analysis, which probably indicates that the metabolic profile of these patients is close to the metabolic profile of the controls.

The loading plot is shown in Figure 4, and the important variables are shown in this figure.

**Identification of metabolites and pathways**

Following maximum class separation, the Human

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**Table 1.** Demographic information and clinical characteristics of unstable angina (UA) patients and controls

| Variables                      | UA Patients (n=94) | Control (n=32) | P-value  |
|-------------------------------|-------------------|----------------|---------|
| Age (Mean±SD)                 | 64.07±10.5        | 54.13±11.7     | < 0.001 |
| Gender                        |                   |                |         |
| Male                          | 54 (57.4 %)       | 14 (43.8 %)    | 0.179   |
| Female                        | 40 (42.6 %)       | 18 (56.3 %)    |         |
| BMI                           | 26.44±3.9         | 28.56±4.9      | 0.023   |
| Smokers                       | 9 (12.3 %)        | 5 (15.6 %)     | 0.756   |
| Ex-smokers                    | 3 (4.1 %)         | 4 (12.5 %)     | 0.196   |
| Hypertension                  | 44 (60.3%)        | 15 (46.9%)     | 0.203   |
| Hyperlipidemia                | 47 (64.4%)        | 17 (53.1%)     | 0.276   |
| Overweight                    | 41 (60.3%)        | 24 (77.4%)     | 0.096   |
| Triglyceride (mg/dl)          | 136.67±58.1       | 113.13±41.5    | 0.337   |
| Total cholesterol (mg/dl)     | 182.25±59.5       | 215.88±99.6    | 0.355   |
| HDL (mg/dl)                   | 45.50±10.6        | 43.50±8.7      | 0.664   |
| LDL (mg/dl)                   | 101.91±47.1       | 133.86±83.8    | 0.313   |
| Ejection fraction (%)         | 48.20±8.3         | 49.29±11.0     | 0.635   |
| Mean LAD stenosis (%)         | 91.45±7.2         | N/A            |         |
| Mean LCX stenosis (%)         | 67.27±36.8        | N/A            |         |
| Mean RCA stenosis (%)         | 60.97±39.2        | N/A            |         |
| Single Stenosis               | 6 (8.2%)          | N/A            |         |
| Stenosis count                |                   |                |         |
| Double Stenosis               | 16 (21.9%)        | N/A            |         |
| Triple Stenosis               | 51 (69.9%)        | N/A            |         |

BMI: Body mass index; HDL: High density lipoprotein; LDL: Low density lipoprotein; N/A: Not applicable; LAD: Left anterior descending; LCX: Left circumflex artery; RCA: Right coronary artery.
Metabolome Database (HMDB) was used to obtain metabolites based on the chemical shifts of class separation (30). Selected chemical shifts that are shown in Figure 4 were entered into HMDB to identify altered metabolites. Then, we uploaded the selected metabolites into MetaboAnalyst 4.0 (31) in order to identify significant altered biochemical pathways based on P-values less than or equal to 0.05 (Table 3).

Results

Clinical characteristics and demographic information

Patients were selected based on UA symptoms and lack of cardiac biomarkers and coronary angiography, a golden standard method for detecting atherosclerotic plaques verified the percentage of stenosis in each of LAD, LCX, and RCA. Obstruction over 75% was considered as significant, and patients with at least one blockage were enrolled in the UA group. Patients without any significant obstruction were included in the control group. For patients with UA and control, a data collection form was completed that included clinical characterizations and demographic information of each patient and control. In the next stage, the informed consent form was completed for patients. In our study, NMR spectrometry was carried out for the 204 plasma samples of UA patients and 56 from controls. Some patients in case and control groups were rejected from our study because of incomplete information and having special diseases. Also, diabetic patients in cases and controls were rejected from the study. Therefore, we removed their NMR information from the study. Hence, 94 cases and 32 control samples remained in the study. The detailed demographics and clinical data of the study's subjects are indicated in Table 1.

Metabolite identification

We applied multivariate analysis to the obtained raw data from NMR spectroscopy to find best chemical shifts. The information about all analysis presented in Figure 2. After pre-processing and multivariate statistical analysis, selected chemical shifts obtained from NMR spectra were uploaded into the Human Metabolome Database (HMDB). We selected 17 important metabolites for examining their relationship with CVD, atherosclerosis and also finding their increased or decreased levels. 17 affected metabolites were identified in the 4 significant altered biochemical pathways (Table 2). We considered four significant pathways based on P-value less than or about 0.05.

The important altered biochemical pathways were steroid hormone biosynthesis, aminoacyl-tRNA biosynthesis, lysine degradation, phenylalanine, tyrosine, and tryptophan biosynthesis. The most important metabolites associated with the most important biochemical pathways were deoxycorticosterone, 17-hydroxyprogesterone, androstenedione, androstanedione, etiocholanolone, estradiol, 2-hydroxyestradiol, 2-hydroxyestrone, 2-methoxyestradiol, 2-methoxyestrone, L-arginine, L-methionine, L-tryptophan, L-tyrosine, aminoacidic acid, N6-acetyl-L-lysine, L-pipeolic acid. 10 metabolites were altered in the steroid hormone biosynthesis pathway included deoxycorticosterone, 17-hydroxyprogesterone, androstenedione, androstanediol, etiocholanolone, estradiol, 2-hydroxyestradiol, 2-hydroxyestrone, 2-methoxyestradiol, 2-methoxyestrone. 4 metabolites were altered in aminoacyl-tRNA biosynthesis pathway included L-arginine, L-methionine, L-tryptophan, L-tyrosine. 3 metabolites were altered in the lysine degradation pathway included aminoacidic acid, N6-acetyl-L-lysine, and L-pipeolic acid. Also, two
metabolites L-tyrosine and L-tryptophan were altered in another biochemical pathway called phenylalanine, tyrosine and tryptophan biosynthesis pathway.

Among 17 selected metabolites, 12 metabolites had decreased levels, and 5 metabolites had increased levels in UA patients in comparison to the control group (Table 2).

As shown in Table 2, compared with the controls, UA patients had higher levels of 5 metabolites; deoxycorticosterone, androstenedione, etiocholanolone, 2-hydroxyestradiol and L-methionine and lower levels of 12 metabolites: 17-hydroxyprogesterone, androstanedione, 2-hydroxyestrone, estradiol, 2-methoxyestradiol, 2-methoxyestrone, L-arginine, L-tryptophan, L-tyrosine, amino adipic acid, N6-acetyl-L-lysine, and L-pipecolic acid. The significant altered biochemical pathways are shown in Figure 5, an overview

Table 2. Selected important metabolites obtained from unstable angina (UA) patients

| No. | Metabolites                  | HMDB ID       | Pathway                                   | Change* |
|-----|------------------------------|---------------|-------------------------------------------|---------|
| 1   | Deoxycorticosterone         | HMDB0000016   | Steroid hormone biosynthesis              | ↑       |
| 2   | 17-Hydroxyprogesterone       | HMDB0000374   | Steroid hormone biosynthesis              | ↓       |
| 3   | Androstenedione              | HMDB000053    | Steroid hormone biosynthesis              | ↑       |
| 4   | Androstanedione              | HMDB0000899   | Steroid hormone biosynthesis              | ↓       |
| 5   | Etiocholanolone              | HMDB000490    | Steroid hormone biosynthesis              | ↑       |
| 6   | Estradiol                    | HMDB000151    | Steroid hormone biosynthesis              | ↓       |
| 7   | 2-Hydroxyestradiol           | HMDB000338    | Steroid hormone biosynthesis              | ↑       |
| 8   | 2-Hydroxyestrone             | HMDB000343    | Steroid hormone biosynthesis              | ↓       |
| 9   | 2-Methoxyestradiol           | HMDB000405    | Steroid hormone biosynthesis              | ↓       |
| 10  | 2-Methoxyestrone             | HMDB000010    | Steroid hormone biosynthesis              | ↓       |
| 11  | L-Arginine                   | HMDB000517    | Aminoacyl-tRNA biosynthesis               | ↓       |
| 12  | L-Methionine                 | HMDB0000696   | Aminoacyl-tRNA biosynthesis               | ↑       |
| 13  | L-Tryptophan                 | HMDB000929    | Aminoacyl-tRNA biosynthesis               | ↓       |
| 14  | L-Tyrosine                   | HMDB000886    | Aminoacyl-tRNA biosynthesis               | ↓       |
| 15  | Amino adipic acid            | HMDB0062715   | Lysine degradation                        | ↓       |
| 16  | N6-Acetyl-L-lysine           | HMDB0033891   | Lysine degradation                        | ↓       |
| 17  | L-Pipecolic acid             | HMDB0000716   | Lysine degradation                        | ↓       |

Figure 5. Graphical overview of metabolic pathway topology analysis of unstable angina (UA) metabotypes. The graph was generated using MetaboAnalyst 4.0 web-based software
of pathway analysis. As it is observed, the pathway of steroid hormone biosynthesis has high pathway impact value compared with the 3 other biochemical pathways in pathway topology analysis (Table 3).

The significant altered metabolic pathways are shown in Table 3. As noted, we found the important pathways based on P-values less than or equal to 0.05.

To validate the PLS-DA model and test and also its applicability in diagnosing UA, the ROC curve analysis was carried out to determine the clinical efficacy of these potential biomarkers. Accordingly, the sensitivity and specificity were determined for the test. The areas under the curve (AUC) for UA were 0.99. The sensitivity, specificity, and precision of the test were 0.87, 90%, 65%, and 91%, respectively, for diagnosing UA (Figure 6).

### Discussion

Because UA can lead to MI, it is important that the disease is diagnosed as soon as symptoms appear, and urgent proceedings are taken to prevent MI. H NMR metabolomics approach was applied to investigate the novel candidate metabolites in UA patients. We identified some of the disturbed important metabolic pathways in UA patients, which include steroid hormone biosynthesis, aminoacyl-tRNA biosynthesis, lysine degradation, and phenylalanine, tyrosine, and tryptophan biosynthesis pathways.

The first altered pathway in our study was steroid hormone biosynthesis. The affected metabolites in this pathway were in the group of mineralocorticoid and sex hormones. We identified 10 metabolites in the pathway of steroid hormone biosynthesis including 1 metabolite from mineralocorticoid group, a class of corticosteroids, and 9 metabolites from the sex hormone group. In mineralocorticoid group, deoxycorticosterone and in sex hormone group 17-hydroxyprogesterone, androstenedione, androstanedione, etiocholanolone, estradiol, 2-hydroxyestradiol, 2-hydroxyestron, 2-methoxyestradiol, and 2-methoxyestron were detected compared to the controls.

Mineralocorticoids are produced in the adrenal cortex and affect electrolyte and fluid balance, such as salt and water. Deoxycorticosterone is a precursor to aldosterone, which is the primary mineralocorticoid. Corticosterone, the main glucocorticoid, is produced from deoxycorticosterone involved in the regulation of energy, immune reactions, and stress responses.

Glucocorticoids are steroid hormones that have inflammatory and immunosuppressive effects on a wide variety of cells. One of the effects of glucocorticoids is to reduce the expression of pro-inflammatory genes (32). Therefore, this may be due to an increase in the inflammatory response, which is one of the predisposing factors for atherosclerotic plaque formation and rupture. Among these steroid hormones, aldosterone has some major deleterious effects on the cardiovascular system, including endothelial dysfunction and myocardial necrosis (33).

Aldosterone binds to mineralocorticoid receptors (MR) and regulates gene expression involved in sodium retention, potassium secretion, and water reabsorption, all of these actions may result in increased blood pressure. Moreover, MR is expressed in a variety of other tissues, and its activation could lead to tissue injury. Indeed, it has been shown that MR activation in the cardiovascular system promotes hypertension, fibrosis, and inflammation. Confirming this issue, the use of MR antagonists is helpful for patients with heart failure (HF) and prevents mortality and morbidity (34).

In addition, an increasing number of clinical and experimental evidence suggests that MR is involved in a wide range of diseases, especially CVDs. MR activation is involved under different pathophysiologic conditions of the cardiovascular system, such as high blood pressure,
HF, and MI (35). Several mechanisms including oxidative stress, inflammation, and fibrosis show the pathophysiological role of MR in the cardiovascular system (34). Furthermore, it has recently been shown that the proatherogenic effects of aldosterone are mediated by an increase in expression of ICAM-1 induced by endothelial MR activation in an atherosclerotic mouse model (36). Some studies done using animal models show the beneficial effects of MR antagonists on the endothelial dysfunction related to MI (37).

The other metabolite in the pathway of steroid hormone biosynthesis were sex hormones. Some of the studies mentioned sex steroids have immunomodulating actions, but their roles in inflammation are complex. Understanding the interplay of atherosclerosis and sex steroid hormones and their receptors on the vessel wall has great importance for clinical practice. Atherosclerosis is caused by a chronic inflammatory condition in the vessel wall, which leads to vasoconstriction or obstruction and is associated with vascular dysfunction (38). Other important considerations in the gender differences observed in CVD between men and women are related to a number of biological agents that affect the type of plaque build-up and vascular response to plaque (39). Compared to men or women after menopause, there is a relatively lower CVD in women before menopause (38). Supporting evidence is limited on low levels of testosterone and its metabolites such as dihydrotestosterone (DHT) and estradiol in the genesis of atherosclerosis and cardiovascular disease in older men. Observational studies link lower T levels with carotid atherosclerosis, aortic and peripheral vascular disease, and cardiovascular events and deaths (40).

One of the affected pathways in UA patients was the aminoacyl-tRNA biosynthesis pathway, which consists of four metabolites L-arginine, L-methionine, L-tryptophan, and L-tyrosine. Aminoacyl-tRNA synthetases include an old family of enzymes that harbor all cells and exist in three major kingdoms of life. Aminoacyl-tRNA synthetases are necessary and ubiquitous ‘house-keeping’ enzymes responsible for charging amino acids to their cognate tRNAs and giving the substrates to synthesize proteins. They perform the esterification reactions that connect amino acids with their tRNAs bearing the correct anticodon to protect the perfect transfer of information directed by the genetic code. Generally, the aminoacylation reaction includes a two-step process: first activation of amino acids by ATP and creating an intermediate aminoacyl adenylate, and then transfer to the 3′-end of tRNA to form the aminoacyl-tRNA end-product. Recent studies have shown several roles for aminoacyl-tRNA synthetases in diseases and their application as pharmacological targets and therapeutic reagents. In addition to the canonical functions of aminoacyl-tRNA synthetases, including aminoacylation and editing, they have the noncanonical activities that are unrelated to aminoacylation, consisting of translation control, transcription regulation, signal transduction, immune cell migration, endothelial cell detachment, stimulation of endothelial cell migration, angiogenesis, inflammation, and tumorigenesis (41, 42). Charcot–Marie–Tooth disease is an inheritable human disease caused by mutations in cytoplasmic aminoacyl-tRNA synthetases. Diseases related to the heart muscle such as cardiomyopathies caused by the deterioration of the myocardium function result in a mutation in aminoacyl-tRNA synthetases (42). In a study aminoacyl-tRNA biosynthesis was one of the altered pathways in UA patients (22). In this study, 3 altered metabolites L-arginine, L-tryptophan, and L-tyrosine decreased in UA patients, but L-methionine increased.

L-arginine is the only substrate of NO production that affects the cardiovascular system. Many studies have clearly shown a beneficial effect of L-arginine on endothelium hypofunction and thus reduced NO synthesis. Additionally, experiments on animals have been performed and in vitro data also indicate that L-arginine may have antiaggregatory, anticoagulant, and profibrinolytic effects (43). L-Arginine has antioxidiant and antiapoptotic effects and increases the relaxation in smooth muscle cells. It also inhibits the expression of adhesion molecules, chemotactic peptides, platelet aggregation, and decreases endothelin-1 expression (44). Production stimulation of endogenous nitric oxide could inhibit atherogenesis, and therefore may be beneficial in patients with risk factors for atherosclerosis (45). In our study, the L-Arginine level was lower compared with the controls. L-methionine as a substrate for cysteine, taurine, S-adenosyl methionine, and glutathione, plays a critical role in the metabolism and health of many species. Methionine is also an important part of angiogenesis. In our study, L-methionine level was higher compared with the controls. It has been shown that high level of methionine can be atherogenic in susceptible mice and causes atherosclerosis and CVD (46). Also, it was shown that a high intake of methionine could increase the risk of acute coronary events in middle-aged men (47). High doses of methionine can lead to an acute increase in plasma homocysteine levels, which can be a sign of the susceptibility to CVD (48). It is known that homocysteine, by harmful effects on cardiovascular endothelium and smooth muscle cells, can mediate cardiovascular problems with alterations in subclinical arterial structure and function (49).

L-tryptophan is an essential amino acid and a major component of protein synthesis in humans and animals. Research has displayed that the L-kynurenine pathway has a key role related to the pathological regulation in both innate and adaptive immune systems (50). The activity of indoleamine 2,3-dioxygenase, an enzyme in the kynurenine pathway, indicated a significant association with atherosclerosis risk factors such as age, LDL cholesterol, and BMI into the female population (51). The increased degradation of tryptophan due to the production of pro-inflammatory cytokines produced by monocytes leads to increased kynurenine and inflammation. The increase of kynurenine/tryptophan ratio increases risk of cardiovascular events (52, 53). Furthermore, increase in expression of indoleamine 2,3-dioxygenase was identified in the macrophages in the atherosclerotic plaques in human (54), and also a study illustrated that low levels of tryptophan in plasma and high levels of kynurenine to tryptophan ratio are features of individuals that suffer from coronary heart disease (55). L-tyrosine or 4-hydroxyphenylalanine is a non-essential amino acid with a polar side group. There is evidence that inflammation and immune activation also damage the conversion of phenylalanine.
to tyrosine. A relationship has been demonstrated in patients suffering from sepsis, cancer, or HIV-1 infection and also in the healthy elderly that have high levels of phenylalanine and as result, phenylalanine to tyrosine ratio (56). It has been described that phenylalanine and Phe/Tyr ratio are associated with some markers of immune activation such as neopterin (57, 58). In a study using a small group of patients, the same associations were seen in patients suffering from CAD (56). The pathway of lysine degradation was another pathway that changed in this study in which three metabolites, aminoadipic acid, N6-acetyl-L-lysine, and L-pipecolic acid were altered. The levels of these metabolites in UA were decreased compared with the controls.

Aminoadipic acid is an intermediate metabolite in the degradation of lysine and saccharopine. Lysine catabolism occurs through one of several pathways, the most common of which is the saccharopine pathway. Aminoadipic acid may be used as a precursor to the downstream enzyme for tryptophan metabolism. It has also been shown that aminoadipic acid inhibits the production of kynurenic acid (59). As a result, the level reduction of aminoadipic acid may be related to an increase in the level of kynurenine and in turn, with inflammation and atherosclerosis.

It is known that aminoadipic acid, as an atherogen, is produced by oxidation of lysine in proteins into the atherosclerotic plaques at high levels (60). The role of aminoadipic acid as a marker was verified in oxidative stress (61). Some studies have shown that aminoadipic acid is increased in prostate biopsies obtained from patients with prostate cancer (62). Therefore, in some of the circumstances, aminoadipic acid can have a role as an atherogen and a metabolotoxin. An atherogen is a compound that leads to atherosclerosis and CVD. A metabolotoxin is an endogenous metabolite that causes adverse effects on human health at chronically high levels. In our study, the reduction of aminoadipic acid can lead to reduction of lysine biosynthesis and consequently may decrease collagen synthesis within the plaque. This event may cause an unstable plaque. L-pipecolic acid is a normal metabolite present in human blood. This metabolite accumulates in body fluids of infants with some of the genetic peroxisomal disorders, such as Zellweger syndrome, neonatal adrenoleukodystrophy, and infantile Refsum disease. The levels of L-pipecolic acid are also raised in patients with chronic liver diseases. L-pipecolic acid is the substrate of the enzyme for tryptophan biosynthesis (56). As a result, the level reduction of aminoadipic acid may be related to an increase in the level of kynurenine and in turn, with inflammation and atherosclerosis.

Conclusion

We conducted the first study on the metabolome profile of UA patients in the Iranian population based on NMR-based spectroscopy. The patients with diabetes were excluded from our research in order to eliminate interferences. In our study, some of the altered pathways and metabolites in the UA patients were reported for the first time. We identified two new important disturbed pathways including steroid hormone biosynthesis, phenylalanine, tyrosine, and tryptophan biosynthesis in UA patients. Also, we have reported the aminocyt-tRNA biosynthesis pathway and lysine degradation that agree with a previous study about UA patients (22). The identified metabolites include 17 new metabolites that have been listed in Table 2. Approximately, most of the identified metabolites were new, and they have not been reported in articles about ACS metabolomics. These results need to be confirmed by other methods for use in clinics, but this finding may provide for a better diagnosis of UA in the future to predict heart attacks in patients.

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Conflicts of Interest

The authors declare that there are no conflicts of interest.

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References

1. Amsterdam EA, Wenger NK, Brindis RG, Casey DE Jr, Ganiats TG, Holmes DR Jr, et al. 2014 AHA/ACC guideline for the management of patients with non-ST-elevation acute coronary syndromes: executive summary: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines. Circulation 2014; 130:2354-2394.
2. Kibos A, Guerchicot A. Susceptibility genes for coronary heart disease and myocardial infarction. Acute Card Care 2011; 13:136-142.
3. Ambrose JA, Winters SL, Stern A, Eng A, Teichholz LE, Gorlin R, et al. Angiographic morphology and the pathogenesis of unstable angina pectoris. J Am Coll Cardiol 1985; 5:609-616.
4. Allender S, Scarborough P, O’Flaherty M, Capewell S. 20th century CHD morbidity in England and Wales: population trends in CHD risk factors and coronary death. BMC Public Health 2008; 8:148.
5. Unal B, Critchley JA, Capewell S. Explaining the decline in coronary heart disease mortality in England and Wales between 1981 and 2000. Circulation 2004; 109:1101-1107.
6. Das R, Kilcullen N, Morrell C, Robinson MB, Barth JH, Hall AS. The British Cardiac Society Working Group definition of myocardial infarction: implications for practice. Heart 2006; 92:21-26.
7. Ali SE, Farag MA, Holvoet P, Hanafi RS, Gad MZ. A comparative metabolomics approach reveals early biomarkers for metabolic response to acute myocardial infarction. Sci Rep 2016; 6:36359.
8. Libby P, Tabas I, Fredman G, Fisher EA. Inflammation and its resolution as determinants of acute coronary syndromes. Circ Res 2014; 114:1867-1879.
9. Barderas MG, Laborde CM, Posada M, de la Cuesta F, Zubiri I, Vivanco F, et al. Metabolomic profiling for identification of novel potential biomarkers in cardiovascular diseases. Biomed Res Int 2011; 2011.
10. Amsterdam EA, Wenger NK, Brindis RG, Casey DE Jr, Ganiats TG, Holmes DR Jr, et al. 2014 AHA/ACC Guideline for the Management of Patients with Non-ST-Elevation Acute Coronary Syndromes: a report of the American College of Cardiology/American Heart Association Task Force on
Practice Guidelines. J Am Coll Cardiol 2014; 64:e139-e228.
11. Ritchie JL, Bateman TM, Bowon RO, Crawford MH, Gibbons RJ, Hall RJ, et al. Guidelines for clinical use of cardiac radionuclide imaging: a report of the American College of Cardiology/American Heart Association Task Force on assessment of diagnostic and therapeutic cardiovascular procedures (Committee on Radionuclide Imaging)—developed in collaboration with the American Society of Nuclear Cardiology. J Nucl Cardiol 1995; 2:172-192.
12. Sabatine MS, Liu E, Morrow DA, Heller E, McCarroll R, Wiegand R, et al. Metabolomic identification of novel biomarkers of myocardial ischemia. Circulation 2005; 112:3868-3875.
13. Robert P, Giugliano CPC, and Eugene Braunwald. Non–ST Elevation Acute Coronary Syndromes. Braunwald's Heart Disease, 10th Edition 2015:1338.
14. Stringer KA, McKay RT, Karnovsky A, Quemerais B, Lacy P. Metabolomics and Its Application to Acute Lung Diseases. Front Immunol 2016; 7:44.
15. Gerszten RE, Wang TJ. The search for new cardiovascular biomarkers Nature 2008; 451:949-952.
16. Pouralijan Amiri M, Khoshkam M, Salek RM, Madadi R, Farzamzadeh Ganji G, Ramazani A. Metabolomics in early detection and prognosis of acute coronary syndrome. Clin Chim Acta 2019; 495:43-53.
17. Patti GJ, Yanes O, Siuzdak G. Innovation: Metabolomics: the apogee of the omics trilogy. Nat Rev Mol Cell Biol 2012; 13:263-269.
18. Capati A, Ijare OB, Bezabeh T. Diagnostic Applications of Nuclear Magnetic Resonance-Based Urinary Metabolomics. Magn Reson Insights 2017; 10:1178623x17694346.
19. Schoeman J, Loots D. Improved disease characterisation and diagnostics using metabolomics: A review. Cell Tissue Res 2011; 11:2673.
20. Emwas AH, Salek R, L. Griffin J, Merzaban J. NMR-based metabolomics in human disease diagnosis: Applications, limitations, and recommendations. Metabolomics 2013; 9:1049-1072.
21. Lewis GD, Asnani A, Gerszten RE. Application of metabolomics to cardiovascular biomarker and pathway discovery. J Am Coll Cardiol 2008; 52:17-123.
22. Li Z, Liu X, Wang J, Gao J, Guo S, Gao K, et al. Analysis of urinary metabolomic profiling for unstable angina pectoris disease based on nuclear magnetic resonance spectroscopy. Mol Biosyst 2015; 11:3387-3396.
23. Sun M, Gao X, Zhang D, Ke C, Hou Y, Fan L, et al. Identification of biomarkers for unstable angina by plasma metabolomic profiling. Mol Biosyst 2013; 9:3059-3067.
24. Shi C-H, Zhao H-H, Hou N, Chen J-X, Shi Q, Xu X-G, et al. Identifying metabolite and protein biomarkers in Unstable angina in-patients by feature selection based data mining method. Chem Res Chinese U 2011; 27:87-93.
25. Yao W, Gao Y, Wan Z. Serum metabolomics profiling to identify biomarkers for unstable angina. Biomed Res Int 2017; 2017.
26. Zhao H, Chen J, Shi Q, Ma X, Yang Y, Luo L, et al. Metabolomics-based study of clinical and animal plasma samples in coronary heart disease with blood stasis syndrome. Evid Based Complement Alternat Med 2012.
27. Loria JP, Rance M, Palmer AG. A relaxation-compensated Carr- Purcell- Melboom- Gill sequence for characterizing chemical exchange by NMR spectroscopy. J Am Chem Soc 1999; 121:2331-2332.
28. van den Berg RA, Hoefsloot HC, Westerhuis JA, Smilde AK, van der Werf MJ. Centering, scaling, and transformations: improving the biological information content of metabolomics data. BMC Genomics 2006; 7:142.
mechanisms, and therapeutic implications. Oxid Med Cell Longev 2017; 2017:1602074.
52. Mangge H, Summers KL, Meinitzer A, Zelzer S, Almer G, Prassl R, et al. Obesity-related dysregulation of the tryptophan-kynurenine metabolism: role of age and parameters of the metabolic syndrome. Obesity (Silver Spring) 2014; 22:195-201.
53. Mangge H, Stelzer I, Reininghaus EZ, Weghuber D, Portolache TT, Fuchs D. Disturbed tryptophan metabolism in cardiovascular disease. Curr Med Chem 2014; 21:1931-1937.
54. Niinisalo P, Oksala N, Levula M, Pelto-Huikko M, Jarvinen O, Salenius JP, et al. Activation of indoleamine 2,3-dioxygenase-induced tryptophan degradation in advanced atherosclerotic plaques: Tampere vascular study. Ann Med 2010; 42:55-63.
55. Wirleitner B, Rudzite V, Neurauter G, Murr C, Kalhins U, Erglis A, et al. Immune activation and degradation of tryptophan in coronary heart disease. Eur J Clin Invest 2003; 33:550-554.
56. Murr C, Grammer TB, Meinitzer A, Kleber ME, Marz W, Fuchs D. Immune activation and inflammation in patients with cardiovascular disease are associated with higher phenylalanine to tyrosine ratios: the ludwigshafen risk and cardiovascular health study. J Amino Acids 2014; 2014:783730.
57. Neurauter G, Schroeder-Kolp N, Scholl-Burgi S, Spener-Unterweger B, Schubert C, Ledochowski M, et al. Chronic immune stimulation correlates with reduced phenylalanine turnover. Curr Drug Metab 2008; 9:622-627.
58. Capuron L, Schroedl Nadel S, Feart C, Aubert A, Higuere D, Barberger-Gateau P, et al. Chronic low-grade inflammation in elderly persons is associated with altered tryptophan and tyrosine metabolism: role in neuropsychiatric symptoms. Biol Psychiatry 2011; 70:175-182.
59. Wu HQ, Ungerstedt U, Schwarcz R. L-alpha-aminoadipic acid as a regulator of kynurenic acid production in the hippocampus: a microdialysis study in freely moving rats. Eur J Pharmacol 1995; 281:55-61.
60. Lin H, Levison BS, Bufta JA, Huang Y, Fu X, Wang Z, et al. Myeloperoxidase-mediated protein lysine oxidation generates 2-aminoadipic acid and lysine nitrile in vivo. Free Radic Biol Med 2017; 104:20-31.
61. Zeitoun-Ghandour S, Leszczyszyn MI, Blindauer CA, Geier FM, Bundy JG, Sturzenbaum SR. C. elegans metallothioneins: response to and defence against ROS toxicity. Mol Biosyst 2011; 7:2397-2406.
62. Jung K, Reszka R, Kamarge B, Bethan B, Stephan C, Lein M, et al. Tissue metabolite profiling identifies differentiating and prognostic biomarkers for prostate carcinoma. Int J Cancer 2013; 133:2914-2924.
63. Mihalik SJ, Moser HW, Watkins PA, Danks DM, Poulos A, Rhead WJ. Peroxisomal L-pipecolic acid oxidation is deficient in liver from Zellweger syndrome patients. Pediatr Res 1989; 25:548-552.
64. Armstrong DW, Zukowski J, Ercal N, Gasper M. Stereochemistry of pipecolic acid found in the urine and plasma of subjects with peroxisomal deficiencies. J Pharm Biomed Anal 1993; 11:881-886.
65. Langeland R. [Construction of facilities in the health sector]. Sykepleien 1976; 63:782-784, 793.