Stroke-prone salt-sensitive spontaneously hypertensive rats show higher susceptibility to spreading depolarization (SD) and altered hemodynamic responses to SD

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Abstract

Spreading depolarization (SD) occurs in a plethora of clinical conditions including migraine aura, delayed ischemia after subarachnoid hemorrhage and malignant hemispheric stroke. It describes waves of near-breakdown of ion homeostasis, particularly Na\(^+\) homeostasis in brain gray matter. SD induces tone alterations in resistance vessels, causing either hyperperfusion in healthy tissue; or hypoperfusion (inverse hemodynamic response = spreading ischemia) in tissue at risk. Observations from mice with genetic dysfunction of the ATP1A2-encoded \(\alpha_2\)-isoform of Na\(^+\)/K\(^+\)-ATPase (\(\alpha_2\)NaKA) suggest a mechanistic link between (1) SD, (2) vascular dysfunction, and (3) salt-sensitive hypertension via \(\alpha_2\)NaKA. Thus, \(\alpha_2\)NaKA-dysfunctional mice are more susceptible to SD and show a shift toward more inverse hemodynamic responses. \(\alpha_2\)NaKA-dysfunctional patients suffer from familial hemiplegic migraine type 2, a Mendelian model disease of SD. \(\alpha_2\)NaKA-dysfunctional mice are also a genetic model of salt-sensitive hypertension. To determine whether SD thresholds and hemodynamic responses are also altered in other genetic models of salt-sensitive hypertension, we examined these variables in stroke-prone spontaneously hypertensive rats (SHRsp). Compared with Wistar Kyoto control rats, we found in SHRsp that electrical SD threshold was significantly reduced, propagation speed was increased, and inverse hemodynamic responses were prolonged. These results may have relevance to both migraine with aura and stroke.

Keywords

Spreading depolarization, spreading depression, salt-sensitivity, hypertension, stroke, migraine

Received 28 June 2022; Revised 22 August 2022; Accepted 13 September 2022

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Introduction

The neurovascular unit (NVU) is comprised of vascular cells [endothelium, vascular smooth muscle cells, pericytes], glia (astrocytes, microglia, oligodendrocytes), and neurons. For a long time, there was a clear division of tasks between neurons on the one hand, which perform the core function of the brain, namely information processing, and the remaining cell types on the other hand, which feed the neurons, ensure the removal of metabolic waste products, for these purposes form a barrier between circulation and parenchyma (the blood-brain barrier (BBB)), match regional cerebral blood flow (rCBF) to neuronal energy demand and perform immunological surveillance. In recent decades, however, it has become increasingly clear that this view is too simplistic and that, for example, astrocytes serve important roles in information processing as they affect recruitment and function of neurons at local and network levels. Similarly, only the other way round, the dogma of stroke was for a long time that it is always a primarily vascular disorder in which neurons and astrocytes are only passively involved. However, in the past 25 years, even this view has fundamentally changed, as it has been discovered that primary neuronal dysfunctions of ion homeostasis, so-called spreading depolarizations (SD), which initiate and maintain neuronal cytotoxic edema, can secondarily trigger severe vasoconstriction and spreading ischemia, leaving neurons trapped in the disrupted state that eventually leads to ischemic necrosis. SD is observed as a large negative direct current (DC) shift and associated with a near-complete breakdown of transmembrane ion gradients and neuronal swelling. In alternating current (AC) electrocorticography (ECoG), SD triggers a rapidly evolving reduction in the amplitudes of spontaneous activity, termed spreading depression. SD-induced spreading ischemia has become the best characterized stroke mechanism in humans in terms of the pathophysiological relationships between DC/AC-ECoG, rCBF, and tissue partial pressure of oxygen (pO₂), as it typically occurs in patients during neurocritical care in whom neurosurgical intervention is indicated, allowing implantation of invasive probes for advanced neuro- and vascular monitoring before, during, and after the development of ischemic infarcts. In particular, SD-induced spreading ischemia is a key process in the pathogenesis of early and delayed infarcts in patients after aneurysmal subarachnoid hemorrhage (aSAH). Furthermore, spreading ischemia was found in patients with traumatic brain injury (TBI) and exacerbated preexisting ischemia in malignant hemispheric stroke (MHS) after middle cerebral artery (MCA) occlusion (MCAO). SD induces tone alterations in resistance vessels, causing either predominant hyperperfusion followed by a mild oligemia (physiological hemodynamic response) in healthy tissue, or severe and prolonged initial hypoperfusion (inverse hemodynamic response = spreading ischemia) when the NVU is severely disturbed. Spreading ischemia is distinguished from primary ischemia, such as occurs in the setting of embolic/thrombotic occlusion of a major cerebral artery. Whereas in the case of spreading ischemia, SD occurs first and is followed by ischemia with a latency of several seconds, and both SD and ischemia spread in the tissue, in the case of severe primary ischemia, ischemia occurs first, followed by SD with a substantial latency of ~1–5 minutes, and only SD, but not ischemia, spreads in the tissue. Nevertheless, the process of spreading ischemia can also build up on incomplete primary ischemia, as is the case, for example, in the context of photothermolysis in mice. In this model, the initial ischemia as a direct consequence of photothermolysis is relatively mild. Only when the first SD occurs between 3 and 35 minutes after the onset of photothermolysis does it result in severe spreading ischemia that produces an ischemic core and becomes progressively milder as it migrates out of the photothermolitic zone. In addition, even if the ischemic core is already permanently depolarized, severe, sporadic SD-induced spreading ischemia may still occur in the penumbra several hours after MCAO, for example, in cats, resulting in delayed growth of the core. Such severe, sporadic SD-induced spreading ischemia in penumbral tissue also occurs in rats, including spontaneously hypertensive rats (SHR), and in patients. In spreading ischemia, the rCBF decline leads to a repercussion on the negative DC shift which becomes longer-lasting compared to the one in normal tissue. This results from mismatch between energy demand and supply, which causes insufficiency of membrane pumps to repolarize the neurons. In the setting of short-duration SD and normal hemodynamic response, SD does not cause neuronal damage locally. However, spreading ischemia can lead to infarction even in tissue that was not yet ischemic at the onset of SD. Importantly, SD properties can change continuously and massively depending on local conditions as an SD moves from one site to another through the tissue. Overall, rather than the number of SDS, their local duration, measured as the local duration of the negative DC shift and depending in particular on both baseline rCBF and hemodynamic response, indicates the local risk of cell death development. In this concept, baseline rCBF and hemodynamic response are complex variables, through which the local duration of SD and neuronal cytotoxic edema, and thus the local...
risk of cell death development can be modulated, for example, by preconditioning or pharmacological treatment.6

Furthermore, a relevant aspect is that although SD begins in neurons and other cell types of the NVU are only secondarily involved,31–34 the dysfunction causative for the initiation of SD in neurons may well be of astrocytic origin.35 This has been particularly well evidenced, for example, for familial hemiplegic migraine type 2 (FHM2), a rare Mendelian model disease of SD due to loss-of-function mutations in ATP1A2, the gene encoding the α2 isoform of Na+/K+-ATPase (NaKA).36–41

Another disease that has long been considered primarily vascular and is now increasingly thought to involve neuronal and astrocytic brain dysfunction causally in its pathogenesis is salt-sensitive hypertension. In patients, salt-sensitivity is present when the blood pressure changes by 5% to 10% or at least 5 mmHg, in response to increased NaCl intake.42 Fifty-one percent of the hypertensive and 26% of the normotensive population are salt-sensitive.43 Animal studies suggest that genetic predisposition has a significant influence on whether or not an individual responds to NaCl intake with hypertension.42 Thus, polygenic animal models such as SHR,44 SHR-derived stroke-prone SHR (SHRsp)45 and Dahl salt-sensitive rats were bred, which are characterized by a particularly high salt-sensitivity.46–49 Dysfunctions in two organs in particular, namely kidney and brain, are held responsible for salt-sensitivity. At the level of the brain, the aldosterone-epithelial Na⁺ channel (ENaC)-endogenous ouabain-angiotensin II receptor type 1 (AT1R) cascade has been implicated as the causative factor in this common disorder.50 This cascade involves three hormone-receptor pairs: (1) aldosterone and mineralocorticoid receptors, (2) endogenous ouabain and α2NaKA, and (3) angiotensin II and AT1R.50 It is assumed that a set-point adjustment in this neural circuitry causes increased salt-sensitivity. Interestingly, at the center of this set-point adjustment is not a change in a neuronal hormone receptor but in an astrocytic/vascular hormone receptor, namely α2NaKA.50–52 According to this concept, salt-sensitive hypertension is a model disease in which hormonal astrocytic/vascular changes unbalance a neural circuitry, which in turn leads to a complex systemic vascular disorder.50,52 However, genetic and pharmacological reductions in α2NaKA-activity not only lead to salt-sensitivity,51,53–55 but also to a higher susceptibility to SD57–41,56 and a higher propensity to react to SD with inverse hemodynamic responses.46,56 This commonality suggests that considering all three conditions together may yield new insights.

The very first question that arises here is whether genetic predisposition to salt-sensitivity in general, i.e., in patients with salt-sensitive hypertension or in other experimental models than α2NaKA-dysfunctional mice, is associated with increased susceptibility to SD and increased propensity to react to SD with inverse hemodynamic responses. SHRsp, along with α2NaKA-dysfunctional mice, are another important genetic model for salt-sensitivity, thought to be due to a dysfunctional aldosterone-ENaC-endogenous ouabain-AT1R cascade in the brain.46–50,57 Although the ATP1A2 gene indeed shows characteristic differences in SHRsp compared with normotensive Wistar-Kyoto (WKY) or Sprague-Dawley rats and SHRsp exhibit greatly enhanced sensitivity to ouabain,57,59 the mechanisms underlying the dysfunctionality of the aldosterone-ENaC-endogenous ouabain-AT1R cascade may be more complicated in SHRsp than α2NaKA-dysfunctional mice. However, to explore this further was not the purpose of the present work; rather, we performed here a deep phenotyping of SHRsp with respect to SD and the inverse hemodynamic response because we considered this the first logical step to determine whether there might be a fundamental association between the genetic predispositions to salt-sensitivity, SD, and inverse response beyond α2NaKA-dysfunctional mice.

SHRsp develop progressive hypertension during young adulthood. The phenotype is polygenic and associated with over-activity of the renin-angiotensin-aldosterone system (RAAS). Histologic studies of small penetrating arteries showed several features of fibrinoid vasculopathy in common with human small vessel disease.50 SHRsp are regarded as a model of endothelial dysfunction.61 The incidence of ischemic strokes is rare when SHRsp are 12–14 weeks old, but exceeds 80% by 30 weeks. The onset of cerebrovascular lesions in SHRsp is dramatically accelerated by exposure of the rats to the so-called Japanese dietary regimen based on a high Na⁺ intake, high Na⁺/K⁺ ratio, and a modified protein composition.49,62,63 This enhancement of the SHRsp phenotype is more complex than pure salt-sensitivity, because the lower-protein diet accelerated stroke development for the same salt load, although blood pressure tended to be somewhat lower.63

To ensure that our results are not directly influenced by previous strokes, we studied 12–14-week-old SHRsp compared with WKY control rats. In in vivo experiments, we compared the electrical SD threshold, SD propagation speed, normal hemodynamic response to SD and post-SD edema formation between SHRsp and WKY on regular diet. In another series, spreading ischemia was induced by topical application of artificial cerebrospinal fluid (aCSF) on the brain containing...
the nitric oxide synthase (NOS) inhibitor NG-nitro-L-arginine (L-NNA) and increased K⁺ concentration ([K⁺]aCSF) as reported previously. Spreading ischemia was compared between four groups: SHRsp and WKY on Japanese diet and on regular diet.

Materials and methods

Animals

The reporting complies with the ARRIVE Guidelines. All animal experiments were authorized by the animal welfare authorities in Berlin, Germany: Berlin State Office for Health and Social Affairs (LAGeSo), G0203/13, and all experimental procedures were conducted in accordance with the Charité Animal Welfare Guidelines. The animals were housed in groups (two animals/cage) under a 12 h light/dark cycle with food and tap water available ad libitum.

In the first 9 weeks, all animals received a regular diet. In series 1A and 1B, both SHRsp and WKY rats continued to receive regular diet as shown in Figure 1(a). In contrast, in series 2, both SHRsp and WKY rats were divided into two subgroups each after the first 9 weeks. One received normal rat chow (containing 22% protein, 2.7 mg/g Na⁺, 7.4 mg/g K⁺, 0.05 mg/g methionine) and tap water, the other received a high-salt, mildly reduced protein Japanese diet [containing 17.5% protein, 3.7 mg/g K⁺, and 0.03 mg/g methionine (ssniff Spezialdiäten GmbH, Soest, Germany, Supplementary Table 1)] and 1% NaCl added to the drinking water. Series 3 only contained SHRsp rats that received Japanese diet after the first 9 weeks. All experiments were performed when the animals were 12–14 weeks old. Male SHRsp rats (n = 59) and male WKY rats (n = 46) were anesthetized with 100 mg/kg body weight (BW) thiopental sodium intraperitoneally (Trapanal®, BYK Pharmaceuticals, Konstanz, Germany), tracheotomized and artificially ventilated (Effenberger Rodent Respirator; Effenberger Med.-Techn. Gerätbauf, Pfaffing/Attel, Germany) to maintain an arterial partial pressure of CO₂ (pCO₂) between 35 and 45 mmHg, an arterial pO₂ between 90 and 130 mmHg and an arterial pH between 7.5 and 7.45. The right femoral artery and vein were cannulated and saline solution was continuously infused to keep the vessels open (1 ml/h). Systemic mean arterial pressure (MAP, Pressure Monitor BP-1, World Precision Instruments, Berlin, Germany) and expiratory pCO₂ (Heyer CO₂ Monitor EGM I, Bad Ems, Germany) were continuously monitored. Arterial pO₂, pCO₂ and pH were serially measured (AVL Medizintechnik GmbH, Bad Homburg, Germany). A physiological body temperature was maintained using a rectal probe connected to a heating pad (Homeothermic Blanket Control, Harvard Apparatus, Cambridge, MA, U.S.A.). Anesthesia was assessed by testing motor responses and changes in MAP to foot-pinchig. If necessary, additional doses of thiopental (25 mg/kg BW) were applied.

Cranial window preparation and experimental set-up

Figure 2 illustrates the experimental set-ups and paradigms. In series 1A, one rostral and one caudal burr hole (diameter: ~1.5 mm; 2–3 mm lateral to midline; ~1.5 mm and ~6.5 mm caudal to bregma) were placed over the parietal cortex for epidural recordings with two Ag/AgCl electrodes. A third burr hole for a laser-Doppler flow (LDF) probe (Periflux 4001, Perimed, Järfälla, Sweden) was implanted between the two electrodes to measure rCBF as reported previously (Figure 2(a)). A fourth burr hole was drilled over the frontal cortex at a distance of 4 mm to the rostral burr hole for placement of a bipolar stimulation electrode (NE-200, Rhodes Medical Instruments, Summerland, CA, USA) (Figure 2(b)).

In series 1B and 3 (Figure 2(c)), and series 2 (Figure 2(d)), a craniotomy (4x5mm) was performed over the parietal cortex, the dura was removed, and a closed cranial window was implanted as reported previously. ECoG was subdurally recorded at the closed window using an Ag/AgCl electrode inserted into the space between cortex and coverslip. Laser speckle contrast analysis (LASCA) imaging was employed to map cerebral perfusion levels in the window area as reported previously (PeriCam PSI HR, Perimed Instruments, Järfälla, Sweden, analysis with PIMSoft, Perimed Instruments). The cortical surface at the cranial window was continuously superfused with aCSF containing in mM: 127.5 NaCl, 24.5 NaHCO₃, 6.7 urea, 3.7 glucose, 1.5 CaCl₂, and 1.2 MgCl₂. In series 2, the Na⁺ concentration in aCSF ([Na⁺]aCSF) was simultaneously decreased from 152 to 120 mM to keep osmolarity constant when [K⁺]aCSF was increased from 3 to 35 mM to induce spreading ischemia. The aCSF was equilibrated with a gas mixture containing 6% O₂, 5.9% CO₂, and 87.5% N₂ to achieve physiological levels of pO₂, pCO₂ and pH. An additional Burr hole was implanted rostrally to either place a bipolar stimulation electrode (series 1B and 3) (Figure 2(c)) or to perform epidural recordings with an Ag/AgCl electrode (series 2) (Figure 2(d)).

Electrical stimulation of SD

See Supplementary Materials and Methods.
Figure 1. Experimental set-ups and protocols. (a) In the first 9 weeks, all animals received a regular diet. After that, part of the animals continued to receive regular diet and another part was fed with Japanese diet. The experiments were performed in 12 to 14 weeks old animals. (b) Original recording of a spreading depolarization (SD) in series 1A. Trace 1 from top to bottom shows the normal hyperemic regional cerebral blood flow (rCBF) response to SD. Traces 2 and 4 demonstrate the SD-induced spreading depression of activity in the alternating current (AC)-frequency band and traces 3 and 5 the negative direct current (DC) shift of SD at the two epidural recording sites. Vertical lines are electrical stimulation artifacts. The insets on the right give the DC/AC-electrocorticography (ECoG) changes of the SD on an expanded time scale and y axis. The SD propagates from the rostral to the caudal epidural recording site (Epi_DC/AC 1 to Epi_DC/AC 2). (c) Post-mortem magnetic resonance imaging (MRI) using a T2-weighted (T2w) fast spin echo (FSE) protocol of a naive stroke-prone spontaneously hypertensive rat (SHRsp) and an SHRsp after stimulation of one SD (right hemisphere) (series 1A). High signal intensity (bottom – overlaid in red) suggests either cerebrospinal fluid (CSF) or vasogenic edema. and (d) Voxel-based analysis of variability in T2w image intensity to identify edema. A Gaussian mixture model with three Gaussian probability density functions was applied to model the variety of intensities. Accordingly, voxels were clustered as a mixture of three Gaussians, namely: “low”, white matter; “medium”, mostly gray matter; and “high”, ventricles or abnormally hyperintense regions. Hyperintensities were quantified by including voxels above a segmentation threshold, defined as intersection between 2nd and 3rd Gaussian fit (arrow).
Figure 2. (a) Experimental set-up and protocol (series 1A). To determine the threshold of spreading depolarization (SD), two silver/silver chloride (Ag/AgCl) electrodes were inserted over the right parietal cortex which measured the epidural direct current (DC)/alternating current (AC)-electrocorticogram (ECoG). Another burr hole was implanted in-between to measure the corresponding regional cerebral blood flow (rCBF) changes using a laser-Doppler flowmetry (LDF) probe. SDs were elicited by current pulses of increasing intensity using a frontal bipolar stimulation electrode. The right panel shows the burr holes for epidural recordings and the lower panel the experimental protocol of series 1A. (b) Bipolar stimulation electrode (2 × 0.2 mm insulated stainless steel wires separated by a 0.5 mm shaft). (c) The left panel shows the experimental set-up of series 1B and 3. aCSF: artificial cerebrospinal fluid, LASCA: laser speckle contrast analysis imaging (sampling rate: 0.5 Hz, spatial resolution: 20 μm, averaged frames per image: 10). The right panel shows the closed cranial window used in series 1B and 3 and the lower panel the experimental protocol. (d) The left panel shows the experimental set-up, the right panel the closed cranial window and the lower panel the experimental protocol of series 2.
**Animal recording techniques**

Subdural and epidural DC/AC-ECoG was performed to monitor Sds and SD-induced activity depression as reported previously.\(^7,6,8\) Electrodes were connected to a differential amplifier (Jens Meyer, Munich, Germany). Analog-to-digital conversion was performed using a Power 1401 (Cambridge Electronic Design Limited, Cambridge, UK). Relative rCBF changes were calculated in relation to baseline after a stabilization period (\(=100\%\)). If possible, a zero level was established at the end of the experiment after cardiac arrest by air embolism. Regions of interest (ROI) in LASCA imaging covered an area of 0.2 mm\(^2\). MAP, expiratory pCO\(_2\), DC/AC-ECoG and rCBF (LDF) were continuously recorded using a personal computer and Spike 2 software (version 6, Cambridge Electronic Design Limited, Cambridge, UK).

**Experimental paradigms**

In series 1A, we first determined the electrical threshold of SD in SHRsp and WKY rats. SD propagation speed was calculated from the delay between the SD-initiating electrical stimulus and SD onset at the rostral window and the distance between stimulation site and rostral window. After recovery from SD, intracardial perfusion of phosphate-buffered saline (PBS) containing 4\% paraformaldehyde (PFA) was performed (Figure 2(a)). Brains were removed and fixed for 48 hours in 4\% PFA, and were then transferred to store-solution (0.25\% PFA in PBS) for ex vivo magnetic resonance imaging (MRI).

In series 1B and 3, normal aCSF was brain topically applied throughout the experiment (Figure 2(c)). We determined the electrical threshold of SD and recorded the normal hemodynamic response in the window area with LASCA imaging. In order to determine the SD propagation speed in these experiments, a rostral and a caudal ROI were defined. When SD was electrically triggered, this was detected as a hyperemic wavefront in the window area that propagated from rostral to caudal. SD propagation speed was calculated from the delay between the appearance of the hyperemic wavefront at two different ROIs.

**Ex-vivo MRI**

See Supplementary Materials and Methods.

**Experimental design and statistical analysis**

See Supplementary Materials and Methods.

**Results**

**BW before surgery, and body temperature and MAP after surgery**

Figure 3 summarizes the effects of rat strain and diet type on BW development before the experiments, as well as on body temperature and MAP measured immediately after surgery. In essence, the results show the expected differences between the four groups. In addition, when experiments were pooled, BW immediately before the experiments and body temperature after surgery significantly correlated both in SHRsp and WKY (Figure 3(b)).

**Electrical threshold and propagation speed of SD in SHRsp and WKY rats**

Thresholds to trigger SD were compared only between SHRsp and WKY receiving a regular diet. Experiments from series 1A (Figure 1(b)) and 1B were pooled. We found that the median SD threshold was significantly lower in SHRsp than WKY (Figure 4(a)). In addition, we found that the median propagation speed of SD was significantly faster in SHRsp than WKY (Figure 4(b)). In series 1A, the amplitude of the epidural negative DC shift was not different between SHRsp and WKY (Figure 4(c)). In series 1B, the amplitude of the subdural negative DC shift was not different between SHRsp and WKY (Figure 4(d)). In series 1A, the peak of the LDF-determined hyperemia was not significantly different between SHRsp and WKY (Figure 4(e)). However, in series 1B, we determined the
median of 5 ROIs in the window area for each animal and found that the LASCA-determined hyperemia was significantly higher in SHRsp than WKY (Figure 4(f)). Figure 4(g) to (j) examines possible relationships between SD propagation speed and SD threshold, on the one hand, and MAP and BW, on the other hand, for the pooled data of SHRsp and WKY.
Overall, post-mortem MRIs showed structural changes after the experiments. Thus, lateral ventricles enlarged in comparison to lateral ventricles of naïve rats (Figure 1(c)). In each hemisphere, we analyzed cortex, hippocampus, amygdala, diencephalon, and corpus callosum. For these structures, (i) volumetric differences of the respective structure and (ii) differences in the percentage of voxels with hyperintense (abnormal) T2w signal (ABT2w%) were compared for the between-subjects variable (SHRsp versus WKY) and the within-subjects variable (hemisphere that was electrically stimulated versus contralateral hemisphere) using two-way repeated-measures ANOVA on ranks with post-hoc Bonferroni t-tests. Significant volumetric differences were found for hippocampus, amygdala and diencephalon between SHRsp and WKY (Figure 5(a)). In contrast, significant differences in ABT2w% indicating edema were found for the whole hemisphere, isocortex, hippocampus, and amygdala between the hemisphere that was electrically stimulated and the contralateral hemisphere but not between SHRsp and WKY (Figure 5(b)). The side on which SD occurred showed more edema than the contralateral side.

**SD-induced spreading ischemia**

Brain topical application of L-NNA and elevated \([K^{+}]_{aCSF}\) led to spontaneously recurring SDs (Figure 6(a)) that induced spreading ischemias in all
groups (Figure 6(b) and (c)). Supplementary Video 1 shows the LASCA-recorded perfusion changes of the first SD-induced spreading ischemia of Figure 6(a) to (c). In SHRsp with Japanese diet, SHRsp with regular diet, and WKY with regular diet, we found that the negative DC shift was always significantly longer when SD in series 2 induced spreading ischemia than when SD in series 1B or 3 induced normal spreading hyperemia (Figure 6(e)). In the left panel of Figure 6(d), the resulting linear relationship between duration of initial hypoperfusion and duration of negative DC shift is shown for the pooled experiments from series 2. The same linear relationship has been demonstrated for the spreading ischemia continuum previously in other settings, including experimental models and patients with aSAH.9,69,70 Interestingly, systemic hypotension
alone even in the absence of concomitant NOS inhibition and elevated $[K^+]_{aCSF}$ led to initial hypoperfusion in response to SD in previous experiments in rats. However, when considering the mixed population of SHRsp and WKY in series 2, this effect appeared to be reversed, and higher blood pressure correlated with prolonged initial hypoperfusion (right panel in Figure 6(d)). In Figure 7, several variables of the first SD-induced spreading ischemia are compared between the four groups of animals. Figure 7(a) illustrates that the duration of SD-induced initial hypoperfusion was significantly longer in SHRsp on Japanese diet than WKY on either Japanese or regular diet. In addition, the duration of the negative DC shift was significantly longer in SHRsp on Japanese diet than WKY on Japanese diet (Figure 7(f)). The amplitude of the negative DC shift did not differ (Figure 7(g)). Perfusion minima during spreading ischemia or perfusion maxima during hyperemia subsequent to spreading ischemia did not show strong differences (Figure 7(b) and (e)). The propagation speed of SD-induced spreading ischemia showed a significant difference between WKY on Japanese diet and regular diet (Figure 7(c)). More importantly, the propagation speed of SD-induced spreading ischemia in series 2 was significantly higher than the propagation speed.
of SD-induced spreading hyperemia in series 1A, 1B or 3 in SHRsp with Japanese diet and normal diet and WKY with normal diet (Figure 7(h)).

**SHRsp receiving Japanese diet and subjected to physiological aCSF show a normal hemodynamic response to SD**

An example for this is given in Supplementary Video 2. The median amplitude of the hyperemic response to SD was 163 (interquartile range (IQR): 159–201) % in this group. The median electrical SD threshold was 2300 (830–2300) mA (n = 6), the median propagation speed 3.7 (3.2–4.0) mm/min (n = 7), and the median amplitude of the subdural negative DC shift 1.8 (1.2–2.5) mV (n = 6).

**Discussion**

We found that SHRsp have a significantly lower electrical threshold to trigger SD in vivo and show higher SD speed under otherwise physiological conditions. This was investigated under thiopental anesthesia, which is known to enhance the inhibitory action of GABAA receptors. GABAA receptor activation has been shown to limit SD velocity. Although this effect should affect both strains in a similar way, it cannot be excluded that it influenced the result. Edema on T2w-MRI of the SD-exposed hemisphere compared with the contralateral side, was most likely vasogenic and a consequence of BBB opening to SD, but did not differ between SHRsp and WKY. Consistent with vascular impairment in SHRsp on Japanese diet, SD-induced spreading hyperemia in series 1A, 1B or 3 in SHRsp with Japanese diet and normal diet and WKY with normal diet (Figure 7(h)).

**Figure 7.** Statistical comparisons of spreading ischemia variables between stroke-prone spontaneously hypertensive rats (SHRsp) on Japanese diet, SHRsp on regular diet, Wistar-Kyoto rats (WKY) on Japanese diet and WKY on regular diet. The analysis of these variables has previously been explained using original traces of a spreading ischemia in a Wistar rat in figure 2 in literature. For the one-way analysis of variance (ANOVA) with post-hoc Bonferroni t-tests, the duration of the initial hypoperfusion in response to spreading depolarization (SD) was submitted to logarithmic transformation to approach normal distribution [SHRsp on Japanese diet versus (1) WKY on Japanese diet: p = 0.003, and versus (2) WKY on regular diet: p = 0.006]. The perfusion minima during spreading ischemia showed a significant difference between WKY on Japanese diet and regular diet (one-way ANOVA with Bonferroni t-test: p = 0.013). When evaluating perfusion minima, it is important to remember that baseline blood flow in the brain is already reduced by approximately 40% compared with normal blood flow in awake animals because of barbiturate anesthesia. The propagation speed of SD-induced spreading ischemia showed a significant difference between WKY on Japanese diet and regular diet (one-way ANOVA with Bonferroni t-test: p = 0.040). The duration of the late hyperperfusion following the initial hypoperfusion during spreading ischemia differed significantly between SHRsp and WKY on Japanese diet [Kruskal-Wallis One Way Analysis of Variance on Ranks (KW-ANOVA) and post-hoc Dunn’s test (pD)]. The perfusion maxima of the late hyperperfusion following the initial hypoperfusion differed significantly between SHRsp and WKY on regular diet (ANOVA with Bonferroni t-test: p = 0.020). The propagation speed of SD-induced spreading ischemia in series 2 was significantly higher than the propagation speed of SD-induced spreading hyperemia in series 1B or 3 (Mann-Whitney Rank Sum Tests).
ischemia lasted significantly longer than in WKY on Japanese or regular diet using a standard model of spreading ischemia. Our results support the hypothesis that in the SHRsp rat model, similar to $\alpha_2$NaKA-dysfunctional mice, genetic predisposition to salt-sensitivity is associated with cerebrovascular dysfunction and increased susceptibility to SD. These findings may have implications for a variety of conditions in which SDs occur and hypertension is a vascular risk factor. For example, this applies to delayed cerebral ischemia after aSAH, in which the occurrence of SDs and spreading ischemia has been unequivocally demonstrated in patients and chronic hypertension is considered a risk factor. However, the findings are also interesting for the association between migraine with aura and stroke. The vascular risk factor hypertension, which is significantly more common in migraine patients, has been suggested as one of the possible mediators for this association. However, it may not be mediation in the strict sense, but the basic mechanisms underlying the development of (1) salt-sensitive hypertension, (2) cerebrovascular dysfunction, and (3) SD, respectively, may overlap. In this context, for example, clinical data are interesting, showing that migraine is primarily a disease of premenopausal women but is associated with increased rates of incident hypertension after menopause.

**Salt-sensitivity**

High salt intake is attributed to escalating cardio- and cerebrovascular morbidity and mortality. Salt-sensitivity is usually defined as a significant blood pressure response to increased NaCl intake, whereas KCl intake lowers blood pressure. The Na$^+$ sensor(s) that mediate this pressure response have not been fully elucidated. However, tight molecular links are thought to exist between Na$^+$ and the dynamic function of vessels, mediating the pressor and depressor effects of high- and low-salt diets. One link is the RAAS. When salt intake is reduced, RAAS activity enhances the expression of Na$^+$ channels and NaKA in the kidney as well as renal Na$^+$ reabsorption and vascular and central mechanisms that increase arterial tone and thirst. Nevertheless, the possibility that renal signals drive the blood pressure response to high-salt diet seems unlikely because the circulating RAAS is normally suppressed under high-salt diet.

In contrast, increasing evidence suggests that the Na$^+$ sensor and the key to salt-sensitivity are found in the brain. This research essentially began with papers by Takahashi and colleagues in 1988, who proposed that a hypothalamic NaKA inhibitor links brain RAAS activation to the hypertensinogenic effects of salt. Thus, a high-salt diet increases the Na$^+$ concentration in the plasma, and many patients with essential hypertension have a slightly elevated but significant increase in [Na$^+$/plasma]. Moreover, when controlled for age, patients with refractory hypertension had increased tissue Na$^+$ content, compared with normotensive controls as assessed using $^{23}$Na-MRI. Plasma Na$^+$ enters the CSF via the fenestrated capillaries of the circumventricular organs and choroid plexus, and elevated CSF Na$^+$ concentration ([Na$^+$/CSF]) then stimulates magnocellular neurosecretory cells of the hypothalamus. Salt uptake from plasma into CSF is physiological and, accordingly, [Na$^+$/CSF] increased significantly in response to a high-salt diet not only in salt-sensitive but also salt-resistant hypertensive patients. NaCl uptake from plasma into CSF occurs in a delayed fashion. Thus, intravenous infusion of a small volume of hypertonic saline in healthy dogs resulted in an increase in [Na$^+$/plasma] from ~149 to ~163 mM within 15 min, whereas [Na$^+$/CSF] reached the same elevated value as [Na$^+$/plasma] only after 90 min. Importantly, even if [Na$^+$/CSF] increases in isolation by intracerebroventricular application and [Na$^+$/plasma] remains unchanged, a short-term pressor response occurs that is associated with enhanced sympathathic activity. Prolonged administration of NaCl into the CSF led to sustained systemic hypertension. This pressor response to elevated [Na$^+$/CSF] is enhanced in various strains of salt-sensitive rodents, including SHRsp, and several lines of experimental evidence suggest that this enhancement involves the central release of endogenous ouabain. The specific relevance of the ouabain-binding site on $\alpha_2$NaKA in salt-sensitive hypertension is based in particular on studies in genetically modified mice. Thus, in a study of isoflurane-anesthetized $\alpha_2$NaKA-deficient knockout mice carrying a null mutation in one ATP1A2 allele, the haploinsufficiency resulted in an increase in baseline blood pressure. Whereas this effect on baseline blood pressure was not significant when the mice were conscious, the increased salt-sensitivity was also convincingly demonstrated in conscious $\alpha_2^{+/\text{WT}}$ mice. Thus, the pressor response to both an increase in [Na$^+$/CSF] and intracerebroventricular ouabain was significantly greater in adult $\alpha_2^{+/\text{WT}}$ mice than in their WT littermates. And, most importantly, neither increased [Na$^+$/CSF] nor intracerebroventricular ouabain induced any pressor response in knockout/knockin mice with an ouabain-resistant $\alpha_2$NaKA, in contrast to WT animals. Further indirect support that salt-sensitivity involves the $\alpha_2$NaKA comes from the finding that blood from volume-expanded rodents contained increased amounts of a NaKA inhibitor that was not present in animals with electrolytic lesions of the anteroventral third ventricular area (AV3V) of the hypothalamus.
the development of hypertension in the presence of AV3V lesions does not occur in many animal models of hypertension, and the humoral NaKA inhibitor cannot be detected.\textsuperscript{108} In addition to salt-sensitive hypertension, also adrenocorticotropic-induced hypertension, which is commonly employed as a model of stress-related hypertension, depended on \(\alpha_2\)NaKA.\textsuperscript{109}

Overall, elevated [Na\(^+\)]\(_{\text{CSF}}\) is now thought to cause sustained activation of central angiotensinergic pathways leading to sympathoexcitation and hypertension, which can be prevented by blockade of central AT1Rs.\textsuperscript{50} Sustained activation of these pathways depends on slow neuromodulation in which ENaCs of the ependyma lining the cerebroventricular compartment are activated by binding of aldosterone to central mineralocorticoid receptors. ENaC activation in turn stimulates the production of endogenous ouabain. Endogenous ouabain enhances protein expression of angiotensin converting enzyme, AT1Rs and NADPH oxidase subunits, decreases neuronal NOS and thereby persistently upregulates the central angiotensinergic pressure pathways. Of particular interest in the sustained activation of this neuronal network is that the effect of endogenous ouabain is not mediated by \(\alpha_2\)NaKA on neurons but by \(\alpha_2\)NaKA, which is not present on neurons but only on astrocytes and vascular cells in the adult brain.\textsuperscript{50,110,111}

\(\alpha_2\)NaKA plays a particular role in membrane microdomains

In contrast to \(\alpha_1\)NaKA, \(\alpha_2\)NaKA is not ubiquitously distributed in the plasma membrane but restricted to microdomains neighboring the junctional sarco(endo)plasmic reticulum (jSER).\textsuperscript{112,113} Plasmalemmal Na\(^+\)/Ca\(^{2+}\)-exchanger (NCX1) and jSER Ca\(^{2+}\)-ATPase (SERCA) are found in the same microdomains similar to different transient receptor protein canonical proteins as components of receptor- and store-operated cation-selective channels.\textsuperscript{112,114} Altogether, these structures form the plasmerosome as a functional unit in which plasma membrane and jSER are only 12–20 nm apart. As a consequence, Na\(^+\) and Ca\(^{2+}\) diffusion between plasmamembrane and cytosol is limited, resulting in [Na\(^+\)]\(_{\text{cyt}}\) and [Ca\(^{2+}\)]\(_{\text{cyt}}\) gradients between plasmamembrane and cytosol.\textsuperscript{110,113} Decline in \(\alpha_2\)NaKA activity, e.g., by ouabain, results in increased Ca\(^{2+}\) uptake by the jSER because of a decrease in Ca\(^{2+}\) efflux via plasmalemmal NCX1. Overall, these changes increase the potential for Ca\(^{2+}\) mobilization from the jSER and thus enhance cellular Ca\(^{2+}\) signaling.

In addition to these ionic mechanisms at the plasmerosome, \(\alpha_2\)NaKA seems to interact with neighboring membrane proteins and organized cytosolic cascades of signaling proteins to communicate with intracellular organelles.\textsuperscript{115} These signaling pathways are rapidly activated by the interaction of low concentrations of ouabain with \(\alpha_2\)NaKA and independent of changes in cytoplasmic Na\(^+\) ([Na\(^+\)]\(_{\text{cyt}}\)), K\(^+\) ([K\(^+\)]\(_{\text{cyt}}\)) and Ca\(^{2+}\) ([Ca\(^{2+}\)]\(_{\text{cyt}}\)) concentrations. They include activation of non-receptor cellular tyrosine protein kinase (cSrc), which leads to tyrosine phosphorylation of a number of cellular proteins and augmented generation of reactive oxygen species (ROS) by mitochondria. The increase in ROS is an essential second messenger for many, though not all, of the downstream events connected with \(\alpha_2\)NaKA. Through these signaling pathways, \(\alpha_2\)NaKA appears to be engaged in multiple regulatory processes affecting cell metabolism, growth, proliferation and migration as well as intercellular communication, i.e., processes typically involving protein phosphorylation/dephosphorylation, gene transcription and translation, as well as Ca\(^{2+}\) signaling.\textsuperscript{50} For example, the \(\alpha_2\)NaKA-cSrc kinase pathway was involved in enhanced responses to vasoconstrictors in isolated MCAs of mice carrying a G301R point mutation in one ATP1A2 allele, resulting in \(\alpha_2\)NaKA haploinsufficiency (ATP1A2\(^{+/−}\)-G301R), compared with WT mice.\textsuperscript{78} ATP1A2\(^{−/−}\)-G301R MCAs showed increased depolarization and stronger sensitization to [Ca\(^{2+}\)]\(_{\text{cyt}}\), although the increases of [Ca\(^{2+}\)]\(_{\text{cyt}}\) were surprisingly less than in WT. This was associated with increased cSrc activation and increased phosphorylation of the myosin targeting subunit (MYPT1) of myosin light chain phosphatase and abolished by cSrc inhibition.

\(\alpha_2\)NaKA and SD

In patients, loss-of-function mutations in the ATP1A2 gene cause FHM2, which is clinically characterized by complicated migraine aura,\textsuperscript{56,76,116} Direct clinical evidence from ECoG recordings\textsuperscript{117} and indirect evidence from measurements of rCBF or its surrogates\textsuperscript{18,118,119} suggest that migraine aura is one of the clinical manifestations of SD. Accordingly, ATP1A2\(^{+/−}\)-R887 mutant mice showed both reduced electrical threshold for SD and increased SD propagation speed \textit{in vivo}.\textsuperscript{37} It is assumed that this results from enhanced glutamatergic transmission. Thus, \(\alpha_2\)NaKA specifically colocalizes in astrocytes with the glutamate transporters GLAST and GLT1 (EAAT1, EAAT2).\textsuperscript{120–123} Analysis at the ultrastructural level demonstrated that this macromolecular complex occurs preferentially in astrocytic processes around asymmetric glutamatergic synaptic junctions, but not around GABAergic terminals.\textsuperscript{120} Under physiological conditions, glutamate is first transported into the cell together with three Na\(^+\) ions and one proton following the [Na\(^+\)] gradient, whereupon a K\(^+\) ion is transported out of the cell in return.\textsuperscript{124–126} Thus, the driving force for glutamate uptake is indirectly
provided by the work of \( z_2 \)NaKA as it maintains the ion gradients in the corresponding microdomains.

\( z_2 \)NaKA and impaired hemodynamic responses to SD

The normal hemodynamic response to SD in naive cortex is somewhat different in mice than in higher species such as rats, swine, and humans.\(^{31}\) The mouse response typically begins with marked initial hypoperfusion, followed by a short peak that barely reaches baseline and renewed, very prolonged rCBF reduction by \(~60\%\).\(^{127}\) Concurrent severe hemoglobin desaturation indicates that O\( \_2 \)metabolism becomes at least partially supply limited, and decrease in blood volume implies vasoconstriction as the mechanism.\(^{128}\) Thus, in the continuum of higher species between normal spreading hyperemia and spreading ischemia when NVU is impaired, the normal mouse hemodynamic response to SD occupies an intermediate position.\(^{31}\) Interestingly, this more vasoconstrictor than vasodilator response to SD was shifted even further toward vasoconstriction in heterozygous \( z_2 \)NaKA knockout mice compared with WT mice, an effect that was not observed in either \( z_1 \)NaKA or \( z_3 \)NaKA heterozygous knockout mice.\(^{40}\)

With regard to the induction of spreading ischemia with a NOS inhibitor and increased \([K^+]_{aCSF}\) in rats, it is relevant, that prolonged exposure of tissue to elevated \([K^+]_{aCSF}\) results in an ex vivo detectable decrease in the \( z_2/z_3 \)NaKA fraction and that elevated \([K^+]_{aCSF}\) can be replaced in the pharmacological protocol for induction of spreading ischemia by an ouabain concentration that inhibits \( z_2 \)NaKA but not \( z_1 \)NaKA.\(^{56}\)

In addition to \( z_2 \)NaKA-dysfunction, there may be further mechanisms specifically contributing to the impaired recovery from the inverse hemodynamic response to SD in SHRsp on Japanese diet. These include endothelial dysfunction triggered by salt loading because the mitochondrial Na\(^+\)/Ca\(^{2+}\) exchanger (NCLX) causes increased Na\(^+\) import into the mitochondrial matrix, which impedes oxidative phosphorylation, thereby limiting ATP production in endothelial cells.\(^{129,130}\) NOS inhibition by L-NNA used to generate spreading ischemia was 91% in a previous study with a similar design.\(^5\) Mechanisms that further reduce the low residual NOS activity might also contribute to prolonged spreading ischemia. These include impaired regulation of endothelial NOS phosphorylation and activation in SHRsp\(^{61,131}\) and salt-loading-triggered endothelial dysfunction as a result of a gut-initiated adaptive immune response mediated by Th17 lymphocytes.\(^{132–134}\)

Conclusion

The hypothesis that a mechanistic link exists between (1) salt-sensitive hypertension, (2) SD, and (3) inverse hemodynamic responses was derived mainly from genetically \( z_2 \)NaKA-dysfunctional mice. Here, we found these associations in SHRsp, a polygenic rat model of salt-sensitive hypertension and propensity to stroke characterized by augmented ouabain-induced vasoconstriction.\(^{57,59}\) Our work is the first step necessary to determine whether these associations apply not only to \( z_2 \)NaKA-dysfunctional mice but more generally. A limitation is that we performed deep phenotyping with respect to SD but no specific studies to determine whether the observed phenotype is directly associated with \( z_2 \)NaKA dysfunction. To our knowledge, \( z_2 \)NaKA functionality has not been sufficiently investigated in SHRsp. However, the ATP1A2 gene was previously found to have restriction fragment length polymorphisms between the genomes of normotensive Sprague-Dawley rats and WKY on the one hand and SHR and SHRsp on the other.\(^{58}\) salt-loading was found to decrease myocardial \( z_2 \)NaKA in SHRsp but not WKY.\(^{135}\) and ouabain-induced vasoconstriction is greatly enhanced in SHRsp compared to WKY, suggesting \( z_2 \)NaKA dysfunction.\(^{57,59}\) Overall, it needs further investigation whether the phenotype we observed indeed partly or completely results from \( z_2 \)NaKA dysfunction or has other causes. In addition, we suggest deep phenotyping related to SD in further genetic models of salt-sensitivity such as Dahl salt-sensitive rats to determine whether the dysfunctional aldosterone-ENaC-endogenous ouabain-AT1R cascade in the brain,\(^{50}\) is generally or only partially associated with an increased propensity for SD and inverse hemodynamic responses, as this may offer novel targets for the treatment of a number of important clinical conditions and stroke in particular.\(^{28,31,81}\)

List of abbreviations

AC, alternating current; ABT2w, abnormal T2-weighted signal; aCSF, artificial cerebrospinal fluid; Ag/AgCl electrode, silver/silver chloride electrode; \( z_2 \)NaKA, \( z_2 \)-isoform of Na\(^+\)/K\(^{-}\)-ATPase; ANOVA, analysis of variance; ARRIVE, Animal Research: Reporting of In Vivo Experiments; aSAH, aneurysmal subarachnoid hemorrhage; AT1R, Angiotensin II receptor type 1; ATP1A2, sodium/potassium-transporting ATPase subunit alpha 2; AV3V, anteroventral third ventricular area of the hypothalamus; BBB, blood-brain barrier; BW, body weight; CaCl\(_2\), calcium chloride; cSrc, non-receptor cellular tyrosine protein kinase; DC, direct current; EAAT1/EAAT2, excitatory amino acid transporter1/2; ECoG, electrocorticogram; ENaC, epithelial sodium channel; FHIM2, familial hemiplegic migraine type 2; FSE, fast spin echo; GABA, gamma-aminobutyric acid; GLAST, glutamate/aspartate transporter (EAAT1); GLT1,
glutamate transporter type 1 (EAAT2); jSER, junctional sarco(endo)plasmic reticulum; KCl, potassium chloride; LaGeSo, Landesamt für Gesundheit und Soziales Berlin; LASCA, Laser speckle contrast analysis; LDF, laser-Doppler flowmetry; L-NNA, N\textsuperscript{0}-nitro-L-arginine; MAP, mean arterial pressure; MCA, middle cerebral artery; MCAO, middle cerebral artery occlusion; MgCl\textsubscript{2}, magnesium chloride; MHS, malignant hemispheric stroke; MRI, magnetic resonance imaging; MWU, Mann-Whitney U test; MYPT1, myosin phosphatase target subunit 1; N\textsubscript{2}, nitrogen; NaCl, sodium chloride; NADPH, reduced nicotinamide adenine dinucleotide phosphate; NaHCO\textsubscript{3}, sodium bicarbonate; NaKA, Na\textsuperscript{+}/K\textsuperscript{+}-ATPase; NCLX, mitochondrial Na\textsuperscript{+}/Ca\textsuperscript{2+}-exchanger; NCX1, plasmalemmal Na\textsuperscript{+}/Ca\textsuperscript{2+}-exchanger; NOS, nitric oxide synthase; NVU, neurovascular unit; PBS, phosphate-buffered saline; pCO\textsubscript{2}, arterial partial pressure of carbon dioxide; PFA, paraformaldehyde; O\textsubscript{2}, arterial partial pressure of oxygen; p\textsubscript{o2}, tissue partial pressure of oxygen; RAAS, renin-angiotensin-aldosterone system; rCBF, regional cerebral blood flow; ROI, region of interest; ROS, reactive oxygen species; SD, spreading depolarization; SERCA, sarcoplasmic/endoplasmic reticulum calcium ATPase; SHRsp, stroke-prone spontaneously hypertensive rats; T2w, T2-weighted magnetic resonance imaging; TBI, traumatic brain injury; WKY, Wistar Kyoto rats; WT, wild type.

Funding

The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: Supported by Deutsche Forschungsgemeinschaft (DFG DR 323/5-1 and DFG DR 323/10-1), and Era-Net Neuron EBio2, with funds from BMBF (01EW2004) to Dr. Dreier.

Declaration of conflicting interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Author’s contributions

Eun-Jeung Kang: designed and performed the experiments, analyzed data, contributed to the figures and the draft, and approved the manuscript before submission. Ofer Prager: was involved in the scanning and analysis of the post-mortem MRIs, contributed to the figures and the intellectual content, and approved the manuscript before submission. Sebastian Major: contributed to the experimental design and intellectual content, supervised the data storage and documentation, and approved the manuscript before submission. Dominik N. Müller: contributed to the intellectual content, and approved the manuscript before submission. Alon Lublinsky: analyzed the post-mortem MRIs, contributed to the figures and the intellectual content, and approved the manuscript before submission. Ana I. Oliveira-Ferreira: contributed to the intellectual content, and approved the manuscript before submission. Jens P. Dreier: planned the study, designed experiments, analyzed data, contributed to the figures, drafted and finalised the manuscript, and approved the manuscript before submission.

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Supplemental material

Supplemental material for this article is available online.

References

1. Santello M, Toni N and Volterra A. Astrocyte function from information processing to cognition and cognitive impairment. Nat Neurosci 2019; 22: 154–166.
2. Dreier JP, Lemalle CL, Kola V, et al. Spreading depolarization is not an epiphenomenon but the principal mechanism of the cytotoxic edema in various gray matter structures of the brain during stroke. Neuropharmacology 2018; 134: 189–207.
3. Van Harrevelde A. Changes in the diameter of apical dendrites during spreading depression. Am J Physiol 1958; 192: 457–463.
4. Kirov SA, Fomitcheva IV and Sword J. Rapid neuronal ultrastructure disruption and recovery during spreading depolarization-induced cytotoxic edema. Cereb Cortex 2020; 30: 5517–5531.
5. Dreier JP, Körner K, Ebert N, et al. Nitric oxide scavenging by hemoglobin or nitric oxide synthase inhibition by N-nitro-L-arginine induces cortical spreading ischemia when K\textsuperscript{+} is increased in the subarachnoid space. J Cereb Blood Flow Metab 1998; 18: 978–990.
6. Dreier JP. The role of spreading depression, spreading depolarization and spreading ischemia in neurological disease. Nat Med 2011; 17: 439–447.
7. Dreier JP, Fabricius M, Ayata C, et al. Recording, analysis, and interpretation of spreading depolarizations in neurointensive care: review and recommendations of the COSBID research group. J Cereb Blood Flow Metab 2017; 37: 1595–1625.
8. Leão AAP. Spreading depression of activity in the cerebral cortex. J Neurophysiol 1944; 7: 359–390.
9. Dreier JP, Major S, Manning A, et al. Cortical spreading ischaemia is a novel process involved in ischaemic damage in patients with aneurysmal subarachnoid haemorrhage. Brain 2009; 132: 1866–1881.
10. Luckl J, Lemalle CL, Kola V, et al. The negative ultra-slow potential, electrophysiological correlate of infarction in the human cortex. Brain 2018; 141: 1734–1752.
11. Bosche B, Graf R, Ernestus RI, et al. Recurrent spreading depolarizations after subarachnoid hemorrhage decreases oxygen availability in human cerebral cortex. Ann Neurol 2010; 67: 607–617.
12. Sugimoto K, Nomura S, Shira S, et al. Cilostazol decreases duration of spreading depolarization and
spreading ischemia after aneurysmal subarachnoid hemorrhage. *Ann Neurol* 2018; 84: 873–885.

13. Dreier JP, Winkler MKL, Major S, et al. Spreading depolarizations in ischaemia after subarachnoid haemorrhage, a diagnostic phase III study. *Brain* 2022; 145: 1264–1284.

14. Hinzman JM, Andaluz N, Shutter LA, et al. Inverse neurovascular coupling to cortical spreading depolarizations in severe brain trauma. *Brain* 2014; 137: 2960–2972.

15. Woitiz J, Hecht N, Pinczolits A, et al. Propagation of cortical spreading depolarization in the human cortex after malignant stroke. *Neurology* 2013; 80: 1095–1102.

16. Van Harreveld A and Ochs S. Electrical and vascular concomitants of spreading depression. *Am J Physiol* 1957; 189: 159–166.

17. Lauritzen M. Pathophysiology of the migraine aura. *The spreading depression theory*. *Brain* 1994; 117: 199–210.

18. Hadjikhani N, Sanchez Del Rio M, Wu O, et al. Mechanisms of migraine aura revealed by functional MRI in human visual cortex. *Proc Natl Acad Sci U S A* 2001; 98: 4687–4692.

19. Leao AAP. Further observations on the spreading depression of activity in the cerebral cortex. *J Neurophysiol* 1947; 10: 409–414.

20. Dreier JP, Major S, Foreman B, et al. Terminal spreading depolarization and electrical silence in death of human cerebral cortex. *Ann Neurol* 2018; 83: 295–310.

21. Shin HK, Dunn AK, Jones PB, et al. Vasoconstrictive neurovascular coupling during focal ischemic depolarizations. *J Cereb Blood Flow Metab* 2006; 26: 1018–1030.

22. Strong AJ, Anderson PJ, Watts HR, et al. Peri-infarct depolarizations lead to loss of perfusion in ischaemic gyronecphalic cerebral cortex. *Brain* 2007; 130: 995–1008.

23. Bere Z, Obrenovich TP, Kozak G, et al. Imaging reveals the focal area of spreading depolarizations and a variety of hemodynamic responses in a rat microembolic stroke model. *J Cereb Blood Flow Metab* 2014; 34: 1695–1705.

24. Zhao HT, Tuohy MC, Chow D, et al. Neurovascular dynamics of repeated cortical spreading depolarizations after acute brain injury. *Cell Rep* 2021; 37: 109794.

25. Takeda Y, Zhao L, Jacewicz M, et al. Metabolic and perfusion responses to recurrent peri-infarct depolarization during focal ischemia in the spontaneously hypertensive rat: dominant contribution of sporadic CBF decrements to infarct expansion. *J Cereb Blood Flow Metab* 2011; 31: 1863–1873.

26. Nedergaard M and Hansen AJ. Spreading depression is not associated with neuronal injury in the normal brain. *Brain Res* 1988; 449: 395–398.

27. Dreier JP, Ebert N, Priller J, et al. Products of hemolysis in the subarachnoid space inducing spreading ischemia in the cortex and focal necrosis in rats: a model for delayed ischemic neurological deficits after subarachnoid hemorrhage? *J Neurosurg* 2000; 93: 658–666.

28. Hartings JA, Shuttleworth CW, Kirov SA, et al. The continuum of spreading depolarizations in acute cortical lesion development: Examining leao’s legacy. *J Cereb Blood Flow Metab* 2017; 37: 1571–1594.

29. Dijkhuizen RM, Beekwilder JP, van der Horp HB, et al. Correlation between tissue depolarizations and damage in focal ischemic rat brain. *Brain Res* 1999; 840: 194–205.

30. Zhao L and Nowak TS Jr. Preconditioning cortical lesions reduce the incidence of peri-infarct depolarizations during focal ischemia in the spontaneously hypertensive rat: interaction with prior anesthesia and the impact of hyperglycemia. *J Cereb Blood Flow Metab* 2015; 35: 1181–1190.

31. Dreier JP and Reiffurth C. The stroke-migraine depolarization continuum. *Neuron* 2015; 86: 902–922. Review.

32. Somjen GG. Mechanisms of spreading depression and hypoxic spreading depression-like depolarization. *Physiol Rev* 2001; 81: 1065–1096.

33. Chuquet J, Hollender L and Nimchinsky EA. High-resolution in vivo imaging of the neurovascular unit during spreading depression. *J Neurosci* 2007; 27: 4036–4044.

34. Peters O, Schipke CG, Hashimoto Y, et al. Different mechanisms promote astrocyte Ca2+ waves and spreading depression in the mouse neocortex. *J Neurosci* 2003; 23: 9888–9896.

35. Largo C, Cuevas P, Somjen GG, et al. The effect of depressing glial function in rat brain in situ on ion homeostasis, synaptic transmission, and neuron survival. *J Neurosci* 1996; 16: 1219–1229.

36. De Fusco M, Marconi R, Silvestri L, et al. Haploinsufficiency of ATP1A2 encoding the Na+/K+ pump alpha2 subunit associated with familial hemiplegic migraine type 2. *Nat Genet* 2003; 33: 192–196.

37. Leo L, Gherardini L, Barone V, et al. Increased susceptibility to cortical spreading depression in the mouse model of familial hemiplegic migraine type 2. *PLoS Genet* 2011; 7: e1002129.

38. Parker PD, Suryavanshi P, Melone M, et al. Non-canonical glutamate signaling in a genetic model of migraine with aura. *Neuron* 2021; 109: 611–628 e618.

39. Unekawa M, Ikeda K, Tomita Y, et al. Enhanced susceptibility to cortical spreading depression in two types of Na(+)K(+)-ATPase alpha2 subunit-deficient mice as a model of familial hemiplegic migraine 2. *Cephalalgia* 2018; 38: 1515–1524.

40. Reiffurth C, Alam M, Zahedi-Khorasani M, et al. K(+)-ATPase alpha isoform deficiency results in distinct spreading depression phenotypes. *J Cereb Blood Flow Metab* 2020; 40: 622–638.

41. Smith SE, Chen X, Brier LM, et al. Astrocyte deletion of alpha2-Na/K ATPase triggers episodic motor paralysis in mice via a metabolic pathway. *Nat Commun* 2020; 11: 6164.

42. Cuevas S, Asico LD, Jose PA, et al. Renal hydrogen peroxide production prevents salt-sensitive hypertension. *J Am Heart Assoc* 2020; 9: e013818.

43. Felder RA, White MJ, Williams SM, et al. Diagnostic tools for hypertension and salt sensitivity testing. *Curr Opin Nephrol Hypertens* 2013; 22: 65–76.

44. Okamoto K and Aoki K. Development of a strain of spontaneously hypertensive rats. *Jpn Cere J* 1963; 27: 282–293.
45. Yamori Y, Nagaoka A and Okamoto K. Importance of genetic factors in hypertensive cerebrovascular lesions; an evidence obtained by successive selective breeding of stroke-prone and -resistant SHR. *Jpn Circ J* 1974; 38: 1095–1100.

46. Maier B and Kubis N. Hypertension and its impact on stroke recovery: from a vascular to a parenchymal overview. *Neural Plast* 2019; 2019: 6843895.

47. Griffin KA, Churchill PC, Picken M, et al. Differential salt-sensitivity in the pathogenesis of renal damage in SHR and stroke prone SHR. *Am J Hypertens* 2001; 14: 311–320.

48. Nagaoka A, Iwatsuka H, Suzuoki Z, et al. Genetic predisposition to stroke in spontaneously hypertensive rats. *Am J Physiol* 1976; 230: 1354–1359.

49. Rubattu S, Stanzione R and Volpe M. Mitochondrial dysfunction contributes to hypertensive target organ damage: Lessons from an animal model of human disease. *Oxid Med Cell Longev* 2016; 2016: 1067801–1067809.

50. Van Huysse JW, Dostanic I, Lingrel JB, et al. Hypertension from chronic Central sodium chloride in mice is mediated by the ouabain-binding site on the Na, K-ATPase alpha(2)-isoform. *Am J Physiol Heart Circ Physiol* 2011; 301: H2147–2153.

51. Blaustein MP and Hamlyn JM. Ouabain, ouabain and aldosterone in the brain: a neuro-modulatory pathway underlying salt-sensitive hypertension and heart failure. *Cell Calcium* 2020; 86: 102151.

52. Zhang J, Lee MY, Cavalli M, et al. Sodium pump alpha2 subunits control myogenic tone and blood pressure in mice. *J Physiol* 2005; 569: 243–256.

53. Hou X, Theriault SF, Dostanic-Larson I, et al. Enhanced pressor response to increased CSF sodium concentration and to central ANG I in heterozygous alpha2 Na+-K+-ATPase knockout mice. *Am J Physiol Regul Integr Comp Physiol* 2009; 296: R1427–1438.

54. Van Huysse JW. Endogenous brain Na pumps, brain ouabain-like substance and the alpha2 isoform in salt-dependent hypertension. *Pathophysiology* 2007; 14: 213–220.

55. Major S, Petzold GC, Reiffurth C, et al. A role of the sodium pump in spreading ischemia in rats. *J Cereb Blood Flow Metab* 2017; 37: 1687–1705.

56. Imaizumi T, Takeshita A, Ashihara T, et al. Augmented vascular responses to ouabain in SHRSPr during salt loading. *Clin Exp Hypertens* A 1987; 9: 753–771.

57. Nojima H, Hayaga Y and Kawakami K. The Na,K-ATPase alpha 2 subunit gene displays restriction fragment length polymorphisms between the genomes of normotensive and hypertensive rats. *J Hypertens* 1989; 7: 937–940.

58. Bruner CA, Myers JH, Sing CF, et al. Genetic basis for altered vascular responses to ouabain and potassium-free solution in hypertension. *Am J Physiol* 1986; 251: H1276–1282.

59. Hainsworth AH and Markus HS. Do in vivo experimental models reflect human cerebral small vessel disease? A systematic review. *J Cereb Blood Flow Metab* 2008; 28: 1877–1891.

60. Daneshtalab N and Smeda JS. Alterations in the modulation of cerebrovascular tone and blood flow by nitric oxide synthases in SHRsp with stroke. *Cardiovasc Res* 2010; 86: 160–168.

61. Aiba I, De Paolis P, Porcellini A, et al. Defective suppression of the aldosterone biosynthesis during stroke permissive diet in the stroke-prone phenotype of the spontaneously hypertensive rat. *Basic Res Cardiol* 2000; 95: 84–92.

62. Yamori Y, Horie R, Tanase H, et al. Possible role of nutritional factors in the incidence of cerebral lesions in stroke-prone spontaneously hypertensive rats. *Hypertension* 1984; 6: 49–53.

63. Dreier JP, Petzold G, Tille K, et al. Ischaemia triggered by spreading neuronal activation is inhibited by vasodilators in rats. *J Physiol* 2001; 531: 515–526.

64. Blaustein MP and Hamlyn JM. Ouabain, endogenous ouabain and ouabain-like factors: the Na(+)/K+ pump/ouabain receptor, its linkage to NCX, and its myriad functions. *Cell Calcium* 2020; 86: 102159.

65. Van Huysse JW, Dostanic I, Lingrel JB, et al. Hypertension from chronic Central sodium chloride in mice is mediated by the ouabain-binding site on the Na, K-ATPase alpha(2)-isoform. *Am J Physiol Heart Circ Physiol* 2011; 301: H2147–2153.

66. Schoknecht K, Kikihia M, Lemule CL, et al. The role of spreading depolarizations and electrographic seizures in early injury progression of the rat photothrombosis stroke model. *J Cereb Blood Flow Metab* 2021; 41: 413–430.

67. Oliveira-Ferreira AI, Milakara D, Alam M, COSBID study group, et al. Experimental and preliminary clinical evidence of an ischemic zone with prolonged negative DC shifts surrounded by a normally perfused tissue belt with persistent electrocorticographic depression. *J Cereb Blood Flow Metab* 2010; 30: 1504–1519.

68. Kang EJ, Major S, Jorks D, et al. Blood-brain barrier opening to large molecules does not imply blood-brain barrier opening to small ions. *Neurobiol Dis* 2013; 52: 204–218.

69. Dreier JP, Windmuller O, Petzold G, et al. Ischemia caused by inverse coupling between neuronal activation and cerebral blood flow in rats. In: Tomita M, Kanno I and Hamel E (eds) *Brain activation and CBF control*. Amsterdam: Elsevier, 2002, pp.487–492.

70. Bere Z, Obrenovitch TP, Bari F, et al. Ischemia-induced depolarizations and associated hemodynamic responses in incomplete global forebrain ischemia in rats. *Neuroscience* 2014; 260: 217–226.

71. Sukhotinsky I, Dilekoz E, Moskowitz MA, et al. Hypoxia and hypotension transform the blood flow response to cortical spreading depression from hyperemia into hypoperfusion in the rat. *J Cereb Blood Flow Metab* 2008; 28: 1369–1376.

72. Aiba I, Carlson AP, Sheline CT, et al. Synaptic release and extracellular actions of Zn2+ limit propagation of spreading depression and related events in vitro and in vivo. *J Neurophysiol* 2012; 107: 1032–1041.
73. Guedes RC and Barreto JM. Effect of anesthesia on the propagation of cortical spreading depression in rats. *Braz J Med Biol Res* 1992; 25: 393–397.
74. Bar-Klein G, Lubinsky S, Kamistsky L, et al. Imaging blood-brain barrier dysfunction as a biomarker for epilepsy. *Brain* 2017; 140: 1692–1705.
75. Sadeghian H, Lacoste B, Qin T, et al. Spreading depolarizations trigger caveolin-1-dependent endothelial transcytosis. *Ann Neurol* 2018; 84: 409–423.
76. Dreier JP, Jurkat-Rott K, Petzold GC, et al. Opening of the blood-brain barrier preceding cortical edema in a severe attack of FHM type II. *Neurology* 2005; 64: 2145–2147.
77. Gursoy-Ozdemir Y, Qiu J, Matsuoka N, et al. Cortical spreading depression activates and upregulates MMP-9. *J Clin Invest* 2004; 113: 1447–1455.
78. Staelehr C, Hangaard L, Bouzinova EV, et al. Smooth muscle Ca(2+) sensitization causes hypercontractility of Middle cerebral arteries in mice bearing the familial hemiplegic migraine type 2 associated mutation. *J Cereb Blood Flow Metab* 2019; 39: 1570–1587.
79. de Rooij NK, Rinkel GJ, Dankbaar JW, et al. Delayed spreading depolarization of cortical spreading depression in rats. *Neurol Sci* 2020; 91: 152–407.
80. Ohman J, Servo A and Heiskanen O. Risks factors for cerebral infarction in good-grade patients after aneurysmal subarachnoid hemorrhage: a systematic review of clinical, laboratory, and radiological predictors. *Stroke* 2013; 44: 43–54.
81. Lemaire CL, Luckl J, Horst V, et al. Migraine aura, transient ischemic attacks, stroke, and dying of the brain share the same key pathophysiological process in neurons driven by Gibbs-Donnan forces, namely spreading depolarization. *Front Cell Neurosci* 2022; 16837650.
82. Oie LR, Kurth T, Gulati S, et al. Migraine and risk of stroke. *J Neurol Neurosurg Psychiatry* 2020; 91: 593–604.
83. Etminan M, Takkouche B, Isorna FC, et al. Risk of ischemic stroke in people with migraine: systematic review and meta-analysis of observational studies. *Bmj* 2005; 330: 63.
84. Schurks M, Rist PM, Bigal ME, et al. Migraine and cardiovascular disease: systematic review and Meta-analysis. *BMJ* 2009; 339: b3914.
85. Spector JT, Kahn SR, Jones MR, et al. Migraine headache and ischemic stroke risk: an updated meta-analysis. *Am J Med* 2010; 123: 612–624.
86. Hu X, Zhou Y, Zhao H, et al. Migraine and the risk of stroke: an updated meta-analysis of prospective cohort studies. *Neurul Sci* 2017; 38: 33–40.
87. Mahmoud AN, Mentias A, Elgendi AY, et al. Migraine and the risk of cardiovascular and cerebrovascular events: a meta-analysis of 16 cohort studies including 1 152 407 subjects. *BMJ Open* 2018; 8: e020498.
88. Gardener H, Monteith T, Rundek T, et al. Hypertension and migraine in the Northern Manhattan study. *Ehnh Dis* 2016; 26: 323–330.
89. MacDonald CJ, El Fatouhi D, Madika AL, et al. Association of migraine With incident hypertension after menopause: a longitudinal cohort study. *Neurology* 2021; 97: e34–e41.
90. Scher AI, Terwindt GM, Picavet HS, et al. Cardiovascular risk factors and migraine: the GEM population-based study. *Neurology* 2005; 64: 614–620.
91. Tietjen GE and Maly EF. Migraine and ischemic stroke in women. A narrative review. *Headache* 2020; 60: 843–863.
92. Neal B, Wu Y, Feng X, et al. Effect of salt substitution on cardiovascular events and death. *N Engl J Med* 2021; 385: 1067–1077.
93. Dietz R, Schonig A, Dart AM, et al. Modulation of sympathetic vasoconstriction by potassium. *J Cardiovasc Pharmacol* 1984; 6 Suppl 1: S230–235.
94. Hamlyn JM and Blaustein MP. Salt sensitivity, endogenous ouabain and hypertension. *Curr Opin Nephrol Hypertens* 2013; 22: 51–58.
95. Bonvalet JP. Regulation of sodium transport by steroid hormones. *Kidney Int Suppl* 1998; 65: S49–S56.
96. Horisberger JD and Rossier BC. Aldosterone regulation of gene transcription leading to control of ion transport. *Hypertension* 1992; 19: 221–227.
97. Takahashi H, Matsusawa M, Suga K, et al. Hypothalamic digitalis-like substance is released with sodium-loading in rats. *Am J Hypertens* 1988; 1: 146–151.
98. Takahashi H, Matsusawa M, Ikegaki I, et al. Brain renin-angiotensin system and the hypothalamic, digitalis-like Na+, K+-ATPase inhibitor in rats. *Clin Exp Hypertens A* 1988; 10: 1285–1287.
99. He FJ, Markandu ND, Sagnella GA, et al. Plasma sodium: ignored and underestimated. *Hypertension* 2005; 45: 98–102.
100. Suckling RJ, He FJ, Markandu ND, et al. Dietary salt influences postprandial plasma sodium concentration and systolic blood pressure. *Kidney Int* 2012; 81: 407–411.
101. de Wardener HE, He FJ and MacGregor GA. Plasma sodium and hypertension. *Kidney Int* 2004; 66: 2454–2466.
102. Kopp C, Linz P, Dahlmann A, et al. 23Na magnetic resonance imaging-determined tissue sodium in healthy subjects and hypertensive patients. *Hypertension* 2013; 61: 635–640.
103. Kawano Y, Yoshida K, Kawamura M, et al. Sodium and noradrenaline in cerebrospinal fluid and blood in salt-sensitive and non-salt-sensitive essential hypertension. *Clin Exp Pharmacol Physiol* 1992; 19: 235–241.
104. Ito N, Suzuki K, Koie H, et al. The effect of 7.2% hypertonic saline solution on the duration of sodium-loading in rats. *Am J Med* 1992; 13: 297–312.
106. James PF, Grupp IL, Grupp G, et al. Identification of a specific role for the Na,K-ATPase alpha 2 isoform as a regulator of calcium in the heart. *Mol Cell* 1999; 3: 555–563.

107. Bealer SL, Haywood JR, Gruber KA, et al. Preoptic-hypothalamic periventricular lesions reduce natriuresis to volume expansion. *Am J Physiol* 1983; 244: R51–57.

108. Brody MJ. Central nervous system and mechanisms of hypertension. *Clin Physiol Biochem* 1988; 6: 230–239.

109. Lorenz JN, Loreaux EL, Dostanic-Larson I, et al. ACTH-induced hypertension is dependent on the ouabain-binding site of the alpha2-Na+/-K+/ATPase subunit. *Am J Physiol Heart Circ Physiol* 2008; 295: H273–280.

110. Blaustein MP and Hamlyn JM. Signaling mechanisms that link salt retention to hypertension: endogenous ouabain, the Na(+)-pump, the Na(+)/Ca(2+)-exchanger and TRPC proteins. *Biochim Biophys Acta* 2010; 1802: 1219–1229.

111. Blanco G. Na,K-ATPase subunit heterogeneity as a mechanism for tissue-specific ion regulation. *Semin Nephrol* 2005; 25: 292–303.

112. Juhaszova M and Blaustein MP. Distinct distribution of different Na+ pump alpha subunit isoforms in plasma-lemma. Physiological implications. *Ann N Y Acad Sci* 1997; 834: 524–536.

113. Blaustein MP, Juhaszova M, Golovina VA, et al. Na/Ca exchanger and PMCA localization in neurons and astrocytes: functional implications. *Ann N Y Acad Sci* 2002; 976: 356–366.

114. Pulina MV, Zulian A, Berra-Romani R, et al. Upregulation of Na+ and Ca2+ transporters in arterial smooth muscle from ouabain-induced hypertensive rats. *Am J Physiol Heart Circ Physiol* 2010; 298: H263–274.

115. Xie Z and Askari A. Na(+)/K(+)/Ca(2+)-ATPase as a signal transducer. *Eur J Biochem* 2002; 269: 2434–2439.

116. Jurkat-Rott K, Freilinger T, Dreier JP, et al. Variability of familial hemiplegic migraine with novel A1A2 Na+/K+/Ca2+-ATPase variants. *Neurology* 2004; 62: 1857–1861.

117. Major S, Huo S, Lemale CL, et al. Direct electrophysiological evidence that spreading depolarization-induced spreading depression is the pathophysiological correlate of the migraine aura and a review of the spreading depolarization continuum of acute neuronal mass injury. *GeroScience* 2020; 42: 57–80.

118. Olesen J, Larsen B and Lauritzen M. Focal hyperemia followed by spreading oligemia and impaired activation of rCBF in classic migraine. *Ann Neurol* 1981; 9: 344–352.

119. Lauritzen M, S, Olsen T, Lassen NA, et al. Changes in regional cerebral blood flow during the course of classic migraine attacks. *Ann Neurol* 1983; 13: 633–641.

120. Cholet N, Pellerin L, Magistretti PJ, et al. Similar perisynaptic glial localization for the Na+,K+-ATPase alpha 2 subunit and the glutamate transporters GLAST and GLT-1 in the rat somatosensory cortex. *Cereb Cortex* 2002; 12: 515–525.

121. Rose EM, Koo JC, Antflick JE, et al. Glutamate transporter coupling to Na+,K+-ATPase. *J Neurosci* 2009; 29: 8143–8155.

122. Melone M, Ciriachi C, Pietrobon D, et al. Heterogeneity of astrocytic and neuronal GLT-1 at cortical excitatory synapses, as revealed by its colocalization With Na+/K+-ATPase alpha isoforms. *Cereb Cortex* 2019; 29: 3331–3350.

123. Capuani C, Melone M, Tottene A, et al. Defective glutamate and K+ clearance by cortical astrocytes in familial hemiplegic migraine type 2. *EMBO Mol Med* 2016; 8: 967–986.

124. Kanner BI. Structure and function of sodium-coupled GABA and glutamate transporters. *J Membr Biol* 2006; 213: 89–100.

125. Wadiche JJ, Amara SG and Kavanaugh MP. Ion fluxes associated with excitatory amino acid transport. *Neuron* 1995; 15: 721–728.

126. Bergles DE and Jahr CE. Synaptic activation of glutamate transporters in hippocampal astrocytes. *Neuron* 1997; 19: 1297–1308.

127. Ayata C, Shin HK, Salomone S, et al. Pronounced hypoperfusion during spreading depression in mouse cortex. *J Cereb Blood Flow Metab* 2004; 24: 1172–1182.

128. Yuzawa I, Sakadzic S, Srinivasan VI, et al. Cortical spreading depression impairs oxygen delivery and metabolism in mice. *J Cereb Blood Flow Metab* 2012; 32: 376–386.

129. Hernansanz-Agustin P, Choya-Foces C, Carregal-Romero S, et al. Na(+)-controls hypoxic signalling by the mitochondrial respiratory chain. *Nature* 2020; 586: 287–291.

130. Geisberger S, Bartolomaeus H, Neubert P, et al. Salt transiently inhibits mitochondrial energetics in mononuclear phagocytes. *Circulation* 2021; 144: 144–158.

131. Yoshitomi H, Xu Q, Gao M, et al. Phosphorylated endothelial NOS Ser1177 via the PI3K/akt pathway is depressed in the brain of stroke-prone spontaneously hypertensive rat. *J Stroke Cerebrovasc Dis* 2011; 20: 406–412.

132. Kleineveldt MF, Manzel A, Titzje J, et al. Sodium chloride drives autoimmune disease by the induction of pathogenic TH17 cells. *Nature* 2013; 496: 518–522.

133. Faraco G, Brea D, Garcia-Bonilla L, et al. Dietary salt transiently inhibits mitochondrial energetics in mononuclear phagocytes. *Circulation* 2021; 144: 144–158.

134. Quintas LE, Noel F and Wibo M. Na(+)/K(+) pump alpha isoforms expression in plasma-lemma. *Nature* 2016; 8: 1273–1283.

135. Hartings JA, Strong AJ, Fabricius M, et al. Spreading depolarizations and late secondary insults
after traumatic brain injury. J Neurotrauma 2009; 26: 1857–1866.

137. Rocha-de-Melo AP and Guedes RC. Spreading depression is facilitated in adult rats previously submitted to short episodes of malnutrition during the lactation period. Braz J Med Biol Res 1997; 30: 663–669.

138. Guedes RCA. Cortical spreading depression: a model for studying brain consequences of malnutrition. In: Preedy VR, Watson RR and Martin CR (eds) Handbook of behavior, food and nutrition. London: Springer, 2011, pp.2343–2355.

139. Schoknecht K, Prager O, Vazana U, et al. Monitoring stroke progression: in vivo imaging of cortical perfusion, blood-brain barrier permeability and cellular damage in the rat photothrombosis model. J Cereb Blood Flow Metab 2014; 34: 1791–1801.

140. Vinogradova LV, Koroleva VI and Bures J. Re-entry waves of leao’s spreading depression between neocortex and caudate nucleus. Brain Res 1991; 538: 161–164.

141. Cain SM, Bohnet B, LeDue J, et al. In vivo imaging reveals that pregabalin inhibits cortical spreading depression and propagation to subcortical brain structures. Proc Natl Acad Sci U S A 2017; 114: 2401–2406.

142. Linde R, Schmalbruch IK, Paulson OB, et al. The Kety-Schmidt technique for repeated measurements of global cerebral blood flow and metabolism in the conscious rat. Acta Physiol Scand 1999; 165: 395–401.

143. Todd MM and Weeks J. Comparative effects of propofol, pentobarbital, and isoflurane on cerebral blood flow and blood volume. J Neurosurg Anesthesiol 1996; 8: 296–303.