Manipulation of Auxin Response Factor 19 affects seed size in the woody perennial *Jatropha curcas*

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Seed size is a major determinant of seed yield but few is known about the genetics controlling of seed size in plants. Phytohormones cytokinin and brassinosteroid were known to be involved in the regulation of herbaceous plant seed development. Here we identified a homolog of *Auxin Response Factor 19* (*JcARF19*) from a woody plant *Jatropha curcas* and genetically demonstrated its functions in controlling seed size and seed yield. Through Virus Induced Gene Silencing (VIGS), we found that *JcARF19* was a positive upstream modulator in auxin signaling and may control plant organ size in *J. curcas*. Importantly, transgenic overexpression of *JcARF19* significantly increased seed size and seed yield in plants *Arabidopsis thaliana* and *J. curcas*, indicating the importance of auxin pathway in seed yield controlling in dicot plants. Transcripts analysis indicated that ectopic expression of *JcARF19* in *J. curcas* upregulated auxin responsive genes encoding essential regulators in cell differentiation and cytoskeletal dynamics of seed development. Our data suggested the potential of improving seed traits by precisely engineering auxin signaling in woody perennial plants.
improvements by direct engineering of genes for either seed size or final seed yield. Therefore, it is necessary to identify more genes from model or non-model plants to understand the genetic network that controls seed size.

Plant hormones play pivotal roles in the developmental processes of diverse traits related to yield. For example, the phytohormones cytokinin and brassinosteroid are involved in the regulation of herbaceous plant seed development. Auxin regulates cell division and is further involved in almost every process of plant growth and development, including seed size. Auxin signaling initiates from hormone perception by an F-box protein receptor Transport Inhibitor Response 1 (TIR1) followed by degradation of negative regulators AUX/IAAs. Auxin Response Factors (ARFs) are then released. In the current auxin signaling model, Aux/IAAs dimerize with and repress ARF transcription factors in the absence of auxin. In the presence of auxin, Aux/IAAs interact with TIR1 resulting in repressor degradation, freeing ARFs for auxin-responsive gene transcription. The interaction between ARFs and IAAs is a key aspect of auxin regulation and occurs through the highly conserved COOH-terminal (CT) Phox and Bem1p (PBI) domain present in both types of proteins. Genetically, auxin induces the formation of Arabidopsis pluripotent cells via a root development pathway by ARF7, ARF19 and other downstream transcription factors such as Lateral Organ Boundaries-domain (LBD) LBD16, LBD17, LBD18 and LBD29. They bind to auxin-responsive elements (AuxREs) in promoter regions to enhance or repress auxin-regulated genes and auxin-inducible genes. In Arabidopsis, ARF2 negatively regulates auxin signaling and seed size via downregulation of cell division in the integument region of endosperm. However, arf2 plants exhibit pleiotropic effects, e.g. reduced seed set, which make these plants undesirable for agriculture. Rice THOUSAND-GRAIN WEIGHT 6 (TGW6) encodes an IAA-glucosidase and plays an essential role in the regulation of auxin homeostasis during endosperm development. Downregulation of this negative regulator in the auxin pathway increases grain weight and yield in rice. Activation of the BIG GRAIN1 (BGI) gene, which encodes a regulator in auxin transporting in rice, leads to significant increases in seed size and plant biomass, weight and yield. The YUCCA1 (ZmYUC1) gene in maize (Zea mays) encodes a seed-specific flavin monooxygenase that is involved in tryptophan-dependent IAA biosynthesis; its mutant has 40% less dry mass than wild-type seeds. However, which ARFs and how the ARFs and AUX/IAAs proteins control seed development remains largely unknown.

Compared with conventional herbaceous vegetable oil, seed oil from perennial woody plants has notable nutritional advantages as well as potential raw materials for biodiesel production. Woody plants are usually perennial with strong secondary xylem that can efficiently transport water and nutrients from root to leaf. Jatropha curcas (Jatropha) is one promising new energy crop with high seed oil content, tolerance to drought and an ability to thrive in poor soil. Furthermore, the reduction of greenhouse gas emission for generating 1 gigajoule energy can be at least 40–107% with respect to fossil diesel. Jatropha curcas (J. curcas) is one promising new energy crop with high seed oil content, tolerance to drought and an ability to thrive in poor soil. Furthermore, the reduction of greenhouse gas emission for generating 1 gigajoule energy can be at least 40–107% with respect to fossil diesel. However, Jatropha has some undesirable traits, such as a high male-flower ratio, high sensitivity to viral diseases, high polyunsaturated fatty acid level and low seed yields. To improve the inadequacy, biotechnological progress had been made in the seed oil profile, flowering traits and virus resistance of Jatropha. Pretreated cuttings Jatropha with indole-3-butryc acid (IBA) and 1-naphthalenecarboxylic acid (NAA) increased rooting, sprouting, and seeds yield. Auxin pretreated cuttings produced much longer and healthier plants that flowered and produced fruits and seeds a year earlier than the control. However, the detailed molecular mechanism how exogenous application of auxin can increase seed yield is still unknown.

Seed size in J. curcas is greater than in the congener J. integerrima (Fig. 1a). Using the inter-species crossing population J. curcas × J. integerrima, we conducted a whole-genome scan for quantitative trait loci (QTLs) and expression QTLs that affect seed oil traits in Jatropha. We screened key genes in auxin pathway including ARFs, IAAs and downstream effectors to identify candidate genes controlling seed size. Importantly, we detected a major QTL qSL11-a controlling seed length with a high likelihood of odd score (LOD score) of 16.69 and percentage of variance explained (PVE) 29.6% on Linkage group (LG) 11, were harboring the JcARF19 gene. In this study, we functionally characterized a key genetic factor JcARF19 which regulates seed size and seed yield in Jatropha. We found that JcARF19 functions as a positive upstream modulator. Its ectopic expression leads to increase in cell size and cell number in Jatropha. Transgenic expression of JcARF19 can also increase seed size and seed weight both in Arabidopsis and Jatropha.

Results
Down regulation of JcARF19 affected auxin signalling transduction in Jatropha. According to the whole-genome scan results, we hypothesized that JcARF19 might participate in controlling seed size. To test the hypothesis, firstly, we investigated potential tissue-specific roles of JcARF19 in Jatropha by profiling its expression patterns in roots, stems, leaves, fruits and seeds using quantitative real-time PCR. JcARF19 showed highly similar expression profiles to ubiquitous expression in all organs and highest in endosperm (Fig. 1b), providing evidence that JcARF19 might be related to seed traits in Jatropha. To test the function of JcARF19 in auxin signalling transduction and therefore plant development in Jatropha, we used synthetic tobacco rattle virus (sTRV) based virus-induced gene silencing (VIGS) method, which we developed previously and allowed us to rapidly identify gene function in various plants. At 27 days after inoculation of agrobacterium containing VIGS vectors of JcARF19 and positive control Jatropha Chlorata 42 (JcCH42) or empty vector control (EV), we observed distinct smaller newly expanding leaves in JcARF19-silenced plants than EV-treated plants (Fig. 1c) and the difference between them was remarkably as shown in Fig. 1d. Smaller leaf size was also observed in the positive control treatment containing the silenced marker gene, JcCH42, which encodes a subunit of Magnesium (Mg) chelatase, involved in photosynthesis (Fig. 1c). Recent research showed that auxin treatment enhances ARF19 binding to its target gene promoters, which correlates with the enhancement of transcriptional activity of the ARF19 in Arabidopsis thaliana (Arabidopsis). We next tested the gene expression of downstream transcription factors esp. Lateral Organ Boundaries-domain (LBDs). Upon auxin treatment, two genes JcLBD18 and JcLBD29 expression QTLs that affect seed oil traits in Jatropha...
which encode putative LBDs of *Jatropha* were up-regulated differentially (Fig. 1e), especially *JcLBD18* which had 7-fold higher expression in EV-treated plants. But in *JcARF19*-silenced plants, there was no obvious induction of *JcLBD18* or *JcLBD29* upon auxin treatment, indicating that *JcARF19* was essential for proper auxin-mediated signalling transduction process in leaf cells of *Jatropha*.

Transgenic overexpression of *JcARF19* increased seed size and dry seed weight in *Arabidopsis*. To further investigate the function of *JcARF19*, at first, we got partial cDNA sequence from a database of sequenced cDNA library prepared from *Jatropha* seeds. By integrating known genomic sequence of *JcARF19* and cloned sequenced information from 5′ RACE and 3′ RACE, we finally got the full-length coding sequence of *JcARF19* (Genbank accession NO. KX988008, detailed sequence information could be found in Supplementary file). *JcARF19* protein encoded 1133 amino acids and consists of major functional domains, an amine-terminus B3 DNA binding domain and a carboxyl-terminus (CT) Phox and Bem1p (PB1) domain. Amino acid sequence alignment showed that ARF19s from various plants had high sequence similarity in the three conserved domains (Fig. S1), suggesting functional conservation of ARF19 family proteins. To further investigated the function of *JcARF19*, we generated thirty transgenic lines for CaMV35S:*JcARF19* in *Arabidopsis*. An obvious increase in plant size and seed size was observed in the 35S:*JcARF19* plants compared to wild-type (WT) Col-0 plants (Fig. 2a–d). Seed length and seed dry weight of 35S:*JcARF19* lines were increased remarkably compared with those of WT Col-0 plants as well (Fig. 2e,f). We further conducted oil traits analysis to check the effects of *JcARF19* on oil yield or oil composition. There was no significant change in either oil content per dry seed weight or oil composition in transgenic line of 35S:*JcARF19* compared with those of WT Col-0 plants (Fig. S2). Since seed size and weight were significantly increased in transgenic line of 35S:*JcARF19*, the lipid
Increased seed size and seed numbers by overexpression of JcARF19 in Jatropha. Jatropha was also transformed with 35S:JcARF19 vector \(^{11}\). The ectopic expression of JcARF19 affected calluses formation during transformation procedure (Fig. S3), producing bigger calluses than that of empty vector (EV). After transferring to soil, the ectopic expression of JcARF19 also affected flowering time (Fig. 3a) and other characteristics (Fig. 3c–f). Under normal growth conditions in a greenhouse, WT Jatropha Jc-MD required around 8 months to produce the first inflorescence whereas 10 primary independent JcARF19 overexpression lines formed their first inflorescences after only 5 months, with a 3 months reduction (Figs 3a and 4a–c). Quantitative reverse transcriptase PCR (qRT-PCR) analysis verified the presence of the transgene and ectopic expression of JcARF19 in transgenic Jatropha plants (Fig. 3b). JcARF19 overexpression lines also had greater branching compared with WT Jc-MD control plants (Figs 3c and 4a). A four-fold increase in seed set from JcARF19 ectopic expression plants was collected within one year of transplanting. Furthermore, a 17.2% increase of single seed weight and 17.1% increase in fruit size were found in JcARF19 ectopic expression plants (Figs 3d–f and 4d–f). The longer cell length (18.4%) and higher cell number (16.5%) in JcARF19 ectopic expression plants explained the increased seed weights and lengths (Figs 3g,h and 4g,h). These results indicated that overexpression of JcARF19 increased length, number and weight of seeds and also oil yield by increasing seed cell number and length in Jatropha.

We further germinated T1 JcARF19-overexpression Jatropha seeds and found that the germination percentage of T1 JcARF19-overexpression Jatropha seeds was higher than these of the wild type control (Fig. S4).

Expression of Auxin responsible genes upon the ectopic expression of JcARF19 in developing seeds. In VIGS assay, we have demonstrated that JcARF19 is essential for auxin signaling transduction on downstream transcription factors JcLBD18 and JcLBD29 in Jatropha leaves (Fig. 1e). To further understand its role in auxin pathway during seed development, we analyzed the expression of JcLBD18 and JcLBD29 in JcARF19 overexpression seeds and found that both of two LBD genes were higher expressed either at early stage or middle stage compared with those of WT control. We further checked the expression of other downstream genes in auxin pathway such as cell cycle and cell number controlling. We found that the expression level of Expansin1 (JcEXP1) and AUXIN-REGULATED GENE INVOLVED IN ORGAN SIZE (JcARGOS) \(^{45}\), together with cell cycle regulators JcCDKA1, JcCYCD2 and JcCYCD5, are induced in JcARF19-ectopic expression Jatropha seeds endosperm, at either early or middle stage of seed development (Fig. 5a), providing molecular explanations for the increased cell number in JcARF19 overexpression Jatropha seeds. We found that the expression of several genes encoding important regulators in cell differentiation and cytoskeletal dynamics have been enhanced including ARGOS \(^{46}\), small GTPases auxin-Rho of Plants (ROP), ROP-interactive protein RIC and receptor-like auxin-(Transmembrane Kinase) TMK \(^{47,48}\), either in early or middle stages of seed development of J. curcas (Fig. 5b), which also explain the cell size expansion in JcARF19 ectopic expression Jatropha seeds.

Characterization of interaction between JcARF19 and JcIAA9. Each ARF transcription factor forms dimer with corresponding Aux/IAAs. Stimulated by eQTL clue of the genetic interaction of JcARF19 and
we identified previously, we hypothesized that ARF19 may function via direct protein-protein interaction with IAA9. The secondary structure of JcARF19 and JcIAA9 were predicted to have 4 α-helix and 5 β-sheet folding module (Fig. 6a). We found that JcARF19 interacts with JcIAA9 physically in vitro pull-down assays (Fig. 6b). We used glutathione S-transferase (GST) fused JcIAA9 as bait and JcARF19 as prey. We found that the GST-fused JcIAA9 COOH-terminal protein (JcIAA9-CT) could interact strongly with 6*Histidine-tagged ARF19 COOH-terminal protein (JcARF19-CT), in contrast to much weaker interaction found on the protein pair of J. integerrima (JiARF19 and JcIAA9). This difference was surprising because the ARF19-PB1 and IAA9-PB1 polypeptides of the two species differ by only one amino acid. The S→G mutation which located in interaction interface β5 of ARF19 proteins affects ARF19-IAA9 binding ability. We also provided evidence to show the physical interaction between JcARF19 and JcIAA9 in vivo and the vital role of key amino acid in heterodimer formation by Bimolecular Fluorescence Complementation (BiFC) assays (Fig. 6c). However, although we identified a putative protein-protein interaction pair of IAA9-ARF19 here (Fig. 6), it is still unclear what the significance of this interaction is so far and how IAA9 affects the function of ARF19 in auxin signaling pathway.

Discussion

In herbaceous plants, species with small seeds sometimes have larger seed set than larger-seed species, assuming a limited total amount of energy. This energetic trade-off has been observed in genetic mutants such as APETALA29–31 and CURLY LEAF2852. In this study we showed that manipulation of the auxin pathway in J.curcas not only increased seed size but also enhanced total seed yield. Previous knowledge on the molecular mechanisms of seed size was mainly limited to model herbaceous plants, particularly Arabidopsis and rice53. Our work suggests that manipulation of auxin is an alternative approach to increase seed size in woody plants.

Several lines of evidence support the involvement of JcARF19 in seed size determination. First, JcARF19 was mapped in the major quantitative trait locus (QTL) region and was significantly associated with seed size11. Second, by using expression QTL (eQTL) analysis to link variants with functional candidate genes, we provided evidences that seed traits were affected by the genetic interaction of JcARF19 and JcIAA911. Third, the C-terminal

Figure 3. Agronomic traits of JcARF19-overexpressing Jatropha. (a) Flowering time in 10 independent T0 transgenic Jatropha plants overexpressing JcARF19 and wild-type plant Jc-MD. Values are mean ± SD (n = 10), and Student T-test statistic was used for analysis. **Indicates P < 0.01, the same for the below traits analysis. (b) Relative JcARF19 expression level in leaves of T0 generation 35S:JcARF19 transgenic Jatropha lines (JcARF19OE #1, #10 and #13) and wild-type plant Jc-MD. Values are mean ± SD (n = 3). (c) Comparison of branch number of WT plant Jc-MD and ten T0 JcARF19 overexpression lines. Values are mean ± SEM (n = 10). (d) Comparison of seed number per tree per year of WT plant Jc-MD and ten T0 JcARF19OE lines. Values are mean ± SEM (n = 10). (e) Comparison of single seed weight of WT plant Jc-MD and three T0 JcARF19 overexpression (JcARF19OE #1, #10 and #13) line. Values are mean ± SEM (n = 50). (f) Comparison of seed length of WT plant Jc-MD and three JcARF19OE #1, #10 and #13 lines. Values are mean ± SEM (n = 50). (g) Comparison of cell length of WT plant Jc-MD and JcARF19OE #1 line. Values are mean ± SEM (n = 10). (h) Comparison of seed cell number of WT plant Jc-MD and JcARF19OE #1 line. Values are mean ± SEM (n = 10).
Figure 4. Functional analysis of JcARF19 in Jatropha. (a) Comparison of plant architecture of JcARF19 overexpression transgenic T0 plant (#1, right) with WT Jatropha plant (Jc-MD, left). (b) No inflorescence was found in WT Jatropha plant Jc-MD in the same stage as JcARF19-overexpressing transgenic T0 plant (#1). (c) JcARF19-overexpressing transgenic T0 plant (#1) with early inflorescence. Bar: 10 cm for (a–c). (d,e) Comparison of fruit size of WT Jatropha (Jc-MD, left) and transgenic T1 Jatropha plant overexpressing JcARF19 (right). Bar: 1 cm. (f) Comparison of seed size of WT Jatropha (Jc-MD, left) and transgenic T1 Jatropha plant overexpressing JcARF19 (right). Dashed line indicates the position of the cross-section. Bar: 1 cm. Comparison of cell size of WT Jatropha Jc-MD (g) and transgenic T1 Jatropha plant overexpressing JcARF19 (h). Bar: 50 μm.

Figure 5. Relative expression in developing endosperm of the JcARF19 overexpression plants. Early stage: 3WAF, Middle stage: 6WAF. Values are mean ± SD (n = 3). (a) Relative expression folds of cell cycle and cell number related genes (JcARF19, JcIAA9, JcLBD18, JcLBD29, JcEXP1, JcARGOS, JcCDKA1, JcCDKD2 and JcCYCD5) in JcARF19 #1 overexpression plants normalized with the wild type. (b) Relative expression folds of cell differentiation and cytoskeletal dynamics related genes (JcRIC1, JcRIC4, JcROP2, JcROP6, JcTMK1, JcTMK2 and JcTMK3) in JcARF19 #1 overexpression plants normalized with the wild type.
of JcARF19, which is essential for protein-protein interaction among ARF proteins, was identical between ARF19 homologs of bigger seed size in *Jatropha*. Point mutations on the single-nucleotide polymorphisms (SNPs) of JcARF19 affected their direct physical interactions (Fig. 6). Fourth, overexpression of JcARF19 increased seed size in both *Arabidopsis* and *Jatropha* by increasing both cell numbers and cell length. Fifth, ectopic expression of JcARF19 upregulated auxin responsive genes encoding important regulators involving in cell differentiation and cytoskeletal dynamics.

Woody plants such as *Jatropha* had longer life cycle than herbaceous ones. It took as long as two years for *Jatropha* to get the first flower blooming under our current lab condition in Beijing North of China, in contrast of 4–5 months in tropical countries such as Singapore. We tried to plant *Jatropha* in South of China and so far they do not flower yet. For this reason we used a few independent T0 Jatropha plants and we also performed genetic analyses on laboratory model plant *Arabidopsis*. The increased seeds yield of T3 JcARF19 ectopic Arabidopsis was also consist with the results of overexpression in *Jatropha*, confirming our claims of improved seed traits with JcARF19 ectopic expression strategy. Furthermore, we found that the germination percentage of T1 JcARF19-overexpression Jatropha seeds was obviously higher than that of the wild type (Fig. S4). Early research had reported that the auxin has close relationship with the seeds germination rate54. It indicated that the traits we observed at T0 generation can be inherited into the next generation and our improved seed traits by JcARF19 ectopic expression are reliable as well55. Nevertheless, the observed impact on plant architecture, seed size and yield by manipulation of ARF19 need not only more observations on T1 and T2 generations plants under greenhouse condition, more researches under field condition are also necessary for the feasibility of a big scale commercialization of this strategy. Auxin is a multifunctional hormone that regulates pattern formation in plants56. The location and timing of auxin accumulation and signal transduction play critical roles in various aspects of plant development57,58. In the future, to avoid growth abnormalities in auxin signaling pathway transgenic plants, it is advisable to use a weaker or an organ and developmental specific promoter rather than a stronger promoter like CaMV 35S promoter because the amount of the hormone produced by the transgene and the response should be confined to the target tissue at an appropriate level as did in cotton fiber cell and other success reports59.

**Figure 6. The key amino acid residue affected the ability of JcARF19 binding with JcIAA9.** (a) Predicated secondary structure of JcIAA9, JcARF19 and other related proteins. (b) GST pull-down assay. Recombinant bait protein GST-JcIAA9 COOH-terminal (residues 229–368 shows JcIAA9-CT containing protein-protein interaction domain III and domain IV of Aux/IAA protein family) strongly binds to prey protein JcARF19 COOH-terminal (residues 1005–1276), but weakly binds to prey protein JiARF19-CT (residues 1008–1279). (c) BiFC assay. Very strong fluorescence signal was only found in the combination of C-terminal of JcIAA9 and JcARF19 (nYFP-JcIAA9+JcARF19-cYFP), but not in the combination of C-terminal of JcIAA9 and JiARF19 (nYFP-JcIAA9+JiARF19-cYFP). Bar: 50 μm.
We present a common sharing genetic framework for the control of cell division, differentiation and size for various plant organs, e.g. seed and root. Given the early working model of ARF19 in auxin signaling transduction, the seed size controlling results in this study can be best integrated as the working model presented in Fig. S5. In this seed size controlling model, auxin activates the transcription of JcARF19 via RETINOBLASTOMA-RELATED (RBR) protein and cytokinin-dependent transcription factor ARABIDOPSIS RESPONSE REGULATOR12 (ARR12). JcARF19 is involved in promoting cell differentiation and thus cell number increasing in early stage of seed development by regulating the transcription of LBD18 and LBD29. ARGOS gene family is auxin-induced and involved in the regulation of cell number for the duration of organ growth. Ectopic expression of ARGOS prolongs the expression of INTEGRUMENTA (ANT) and cell cycle regulator CycD3; as well as the neoplastic activity of leaf cells. Overexpression of ARGOS genes modifies plant sensitivity to Ethylene, leading to improved drought tolerance in both Arabidopsis and maize. The auxin-(Transmembrane Kinase) TMK sensing and auxin-Rho of Plants (ROP) signaling networks have been demonstrated to control auxin signaling pathway. JcARF19 might be also involved in enlarged cell size by TMK Auxin-Sensing and ROP GTPase signaling complex in middle stage of seed development. Considering that orthologs of JcARF19 exist in many other plant species, including castor bean, alfalfa (Medicago sativa), soybean and apples, the manipulation of ARF19 may provide a broad application to increase plant biomass and seed productivity in many other species. This study provides evidence that an auxin signaling integrator ARF19 plays vital roles in determining seed size. ARF19 is conserved in higher plants and involved in auxin pathway signal transduction. Nevertheless, it is necessary to test the ARF19 ectopic expression Jatropha lines under field conditions to get conclusive statements of its commercial viability. Meanwhile besides of plant genome, plant rhizospheric or leaf-residing microorganisms via plant endogenous auxin signalling pathway have been successfully to improve crop yield dramatically including Jatropha. ARF19 transactivation factors mediated auxin pathway is essential for growth and yield promoting effect by beneficial microbes. Manipulation of the auxin signaling pathway can result in larger seed sizes and improved seed yield in J. curcas, this ARF19-ectopic expression plant may become a more attractive commercial plant.

Materials and Methods

Plant materials and growth condition. Three species of plants were used in this study. For Jatropha, seeds were obtained from the Jatropha curcas (Jc-MD) elite plants which were pre-selected by Drs. Yan Hong and Chenxin Yi. The seeds were germinated on ½ Murashige and Skoog salt medium at 25 °C under a 16 h light/8 h dark photoperiod with a light intensity of 100 μmol·m⁻²·s⁻¹. When two or three true leaves were grown, the seedlings were transplanted into pots filled with soil and grown at 25 °C in greenhouse under natural light condition. Plant management, including pesticide spraying, watering and artificial fertilization, was carried out according to normal practice. For Arabidopsis, seeds of ecotype Col-0 were vernalized on ¼ Murashige and Skoog salt medium at 4 °C in darkness for about three days and transferred into a growth chamber at 22 °C under 16 h dark/8 h light photoperiod. The seedling with four true leaves were transplanted into soil and grown in the same condition. Arabidopsis transformation was performed according to the floral-dipping method previously described. For Nicotiana benthamiana, seeds were sown in soil to germinate. The seedlings were transplanted in separated pots and grown in a greenhouse at 25 °C with 14 h dark/10 h light photoperiod.

RNA extraction and analysis. RNA was isolated and analyzed according to previously described methods. 100 mg samples from different organs were harvested and extracted with plant RNA purification reagent (Invitrogen, Carlsbad, CA, USA). Nanodrop (Thermo Scientific, Wilmington, DE, USA) was used to quantify RNA concentration. M-MLV reverse transcriptase (Promega, Madison, USA) was used for reverse transcription. For quantitative PCR analysis, THUNDERBIRD SYBR qPCR Mix (TOYOBO) was used and run in Bio-Rad CFX96 qPCR machine. Each treatment was repeated with three biological replicates, and with three technical replicates for each biological sample. The primers for target genes are listed in Table S1. Standard deviation was calculated based on the three biological replicates.

Virus induced gene silencing. We used the sTRV method described by Ye et al., using psTRV1 and psTRV2. PCR-based cloning was used to clone partial cDNAs of JcARF19 to psTRV2 to generate psTRV2 derivatives. Another psTRV2 clone with insertion of Jatropha Chlorata 42 (JcCH42) was served as a positive control, using psTRV1, psTRV2 and psTRV2 derivatives were electroporated into Agrobacterium strain AGL1. Vacuum agroinfiltration was used to inoculate those Agrobacterium into Jatropha seedling with two or three true leaves. At least 5 Jatropha seedlings were agroinfiltrated with psTRV1 and psTRV2-JcARF19, psTRV2-JcCH42 or psTRV2 vector only accordingly. After infiltration, plants were grown in a growth chamber at 25 °C with a 16 h light/8 h dark photoperiod. Phenotypes of Jatropha plants at 27 days post-infiltration were recorded and leaves in same leaf position were picked and leaf width were measured. Values (n = 5) were shown as mean ± SD and statistic analysis with Student T-test. *Indicates P < 0.01, **indicates P < 0.05.

IAA sensitivity test. Detached silenced systemic Jatropha leaves were kept in liquid MS medium containing 10 nM IAA following treated with vacuum infiltration. Treated leaves were incubated at 25 °C in an incubator for 6 hrs. After the IAA treatment, the leaves were subjected to RNA isolation and following quantitative real-time PCR analysis.
Transgenic plant plasmid construction.  JcARF19 gene was identified from a database of sequenced cDNA library prepared from Jatropha seeds (detailed sequence information could be found in Supplementary file). The full-length cDNA fragment of JcARF19 was PCR-amplified with primers (Table S1). The PCR fragment was inserted in the sense orientation into suitable sites of pCABMIA1300-3HA vector41.

Scanning electronic microscopy (SEM) and light microscopy.  For observation of Arabidopsis seeds with the scanning electron microscope (SEM), collected seeds from WT Col-0 and 35S:JcARF19 overexpression plants respectively were fixed with a tape inside a sample chamber, following freezing in liquid N2. Images were collected using a SEM (JSM-6360LV, JEOL, USA) with an acceleration voltage of 20 kV. For observation of Jatropha seed endosperm with light microscopy, endosperm discs from WT Jc-MD and 35S:JcARF19 overexpression plants respectively were excised from mature Jatropha endosperm (7 WAF, weeks after fertilization) and fixed overnight in 2.5% glutaraldehyde in 0.1 M phosphate buffer, pH 7.2 as described previously40. Endosperm discs were rinsed three times in 0.1 M phosphate buffer for 15 min each, and were then post-fixed in 1% (w/v) aqueous OsO4 for 1 h. Tissues were dehydrated in an ethanol series and embedded in Spurr’s resin. Semi-thin sections with thickness of 500 nm were stained in 0.1% toluidine blue and photographed with a Zeiss Axioplan 2 microscope (Carl Zeiss, Germany). Cell size and cell number per disc were analyzed with ImageJ and calculated, followed with statistic analysis with Student T-test. **Indicates P < 0.01. Values are mean ± SD (n = 10).

Arabidopsis seed size and weight measurement.  Mature seeds were harvested from WT Col-0 and 35S:JcARF19 overexpression plants grown under the same conditions. 100 seeds from ten independent transgenic lines and WT Col-0 were weighted and recorded with three technical replicates. Values (n = 10) are given as mean ± SD. A DMS000B microscope (Leica) and ImageJ analysis software were used to measure seed sizes. Values (n = 10) are given as mean ± SD. Statistic analysis with Student T-test. **Indicates P < 0.01.

Fatty acid analysis.  Total lipid was extracted and transmethylated from 100 dry Arabidopsis seeds as described previously40. The resulting FAMEs were separated and detected by GCMS-QP2010 Ultra (Shimadzu, Kyoto, Japan). The GC analysis was performed under conditions described before31. The data were presented based on three biological replicates and each biological replicate had three technical replicates. Values (n = 3) are given as means ± standard deviation.

Explant material preparation and Jatropha transformation method.  Cotyledons were harvested from WT plants Jc-MD sterilized seedlings that were 7–9 days old and were cut into small pieces (5 mm × 5 mm) used as explants. After co-cultivation, shoot regeneration, shoot elongation and rooting, we got the JcARF19 overexpression line. Detailed protocol can be found in Qu et al.31.

JcARF19-overexpressing Jatropha agronomic traits measurement and statistical analysis.  Wild-type Jc-MD and JcARF19 transgenic overexpression Jatropha plants were grown in the same condition. Flowering time was scored by the number of days from transplantation to soil to the day of first florescence emergence. 10 independent T0 JcARF19 overexpression plants and WT plant Jc-MD should be calculated and values are mean ± SD (n = 10). Branch number of each plants after 1 year of transplanting was recorded for either Jc-MD (n = 10) or T0 JcARF19 overexpression plants (n = 10). Values are mean ± SD (n = 10). Single seed weight and seed length for each seeds were measured for three of WT plants Jc-MD and T1 JcARF19 overexpression seeds of three lines, JcARF19OE #1, #10 and #13. Values are mean ± SEM (n = 50). Germination percentage were measured for five of WT plants Jc-MD and T1 JcARF19 overexpression seeds of JcARF19 OE #10 and #13. Values are mean ± SEM (n = 5). Student T-test was used for statistical analyses for all agronomic traits. *Indicates P < 0.01; **Indicates P < 0.05.

In vitro GST pull-down assay.  The C-terminal sequence of JcIAA9 and JcARF19 were amplified by PCR using Phusion High-Fidelity DNA Polymerase (Thermo-Fisher, Finnzymes, Espoo, Finland) and subcloned into pGEX6P-1 or pET28-SUMO vectors to generate GST fusion or 6×His fusion constructs. Point mutations were performed to generate vector of JcARF19 fusion with 6×His tag by QuikChange Site-Directed Mutagenesis Kit (Stratagene, Agilent, Wilmington, DE, USA). In vitro pull-down assays were performed with 2 μg of GST fusion proteins and 2 μg of His-tagged proteins. GST fusion proteins were incubated in a binding buffer (50 mM Tris-HCl at pH 7.5, 100 mM NaCl, 0.25% Triton X-100, 35 mM β-mercaptoethanol) with 25 μL of glutathione sepharose 4B (GE Healthcare, Uppsala, Sweden) for 3 h at 4 °C and GST beads were washed six times with binding buffer. His-tagged JcARF19-CT and JcARF19-CT proteins were added into GST beads and the mixture was incubated overnight at 4 °C. After washing again with binding buffer six times, pulled-down proteins were separated on 12% SDS–polyacrylamide gel and detected by Western blotting using anti-His or anti-GST antibody as previously described67,68.

Bimolecular fluorescence complementation (BiFC).  BiFC was carried using previously described vectors and methods67,68. The C-terminal sequence of JcARF19 and JcIAA9 were cloned in corresponding restrict enzyme sites of BiFC vectors. Point mutations were performed to generate JcARF19 by QuikChange Site-Directed Mutagenesis Kit (Stratagene, Agilent, Wilmington, DE, USA). The resulting cassettes including fusion proteins and constitutive promoters were cloned into pGreen binary vector HY105 and transformed into Agrobacterium. For BiFC experiments, 3-week-old Nicotiana benthamiana leaves were co-infiltrated with Agrobacterium as previously described. Two days after incubation, fluorescence and DAPI staining were analyzed by confocal microscopy67,70. The confocal laser scanning microscope technique we used was referred to the Leica SP8 microscope instruction.
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Author Contributions

J.Y. and C.M.W. conceived of the study, and design the experiments and draft the manuscript. C.M.W. carried out the gene expression analysis and analysis of the study. Z.F.X. participated in data interpretation and revised the manuscript. Y.W.S. helped to design the experiments, analyzed the data, drafted the manuscript and performed molecular cloning and analysis, genotyping. N.W. participated in the design of the study, performed molecular analysis and statistical analysis and drafted the manuscript. X.Y.J. performed chemical analysis, field traits collection and analysis. X.H.W. and Z.J.L. participated in the molecular analysis. H.Z.M. did the Jatropha transformation. G.H.Y. provided research materials. C.X.Z. provided important reagents. F.J.W. provided important reagents and help to revise the manuscript. All authors read and approved the final manuscript.

Additional Information

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