Therapeutic function of a novel rat induced pluripotent stem cell line in a 6-OHDA-induced rat model of Parkinson's disease

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Abstract. Parkinson's disease (PD) is a progressive neurodegenerative movement disorder of the central nervous system that results from the loss of dopaminergic (DA) nigral neurons. Induced pluripotent stem cells (iPSCs) have shown potential for cell transplantation treatment of neurodegenerative disorders. In the present study, the small molecules CHIR99021 and RepSox (CR) significantly facilitated reprogramming and enhanced the efficiency of GFP/iPS-like colonies [rat iPSCs induced by OCT3/4, Sox2, Klf4, c-Myc, Nanog and Lin28 + CR (RiPSCs-6F/CR)] generation by ~4.0-fold during lentivirus-mediated reprogramming of somatic cells. The generation of iPSCs was confirmed by reverse transcription-PCR, immunofluorescence and western blot analysis. Subsequently, RiPSCs-6F/CR were stereotactically transplanted into the right medial forebrain bundle (MFB) of 6-hydroxydopamine-lesioned rats with PD. The transplanted RiPSCs-6F/CR survived and functioned in the MFB of rats with PD for ≥20 weeks, and significantly improved functional restoration from their Pd-related behavioral defects. Furthermore, the grafted RiPSCs-6F/CR could migrate and differentiate into various neurocytes in vivo, including γ aminobutyric acid-ergic, DA neurons and glial cells. In conclusion, the present study confirmed that RiPSCs-6F/CR induced by small molecules could be used as potential donor material for neural grafting to remodel basal ganglia circuitry in neurodegenerative diseases.

Introduction

Parkinson's disease (PD) is the second most common neurodegenerative disease after Alzheimer's disease, affecting 1-3% of the population >60 years old worldwide (1). The main pathology of PD is the progressive loss of dopaminergic (DA) neurons in the substantia nigra and formation of α-synuclein-containing Lewy bodies (2,3). Early PD responds well to DA drugs, such as L-3,4-dihydroxyphenylalanine, dopamine agonists, and monoamine oxidase and catechol-O-methyl-transferase inhibitors (4). However, over time, these drugs begin to lose their effectiveness and cause side effects, including movement disorders and neuropsychiatric complications (5,6). At present, there are two main surgical methods that can be used as an effective adjuvant therapy to drug treatment: Nucleus lesioning surgery and deep brain stimulation (DBS) (3). Nucleus lesioning is irreversible damage to the brain nuclei, such as the globus pallidus internus and the subthalamic nucleus (STN), accompanied by serious complications including hemiparesis, visual disturbances and permanent speech deficits, whereas DBS targets the STN and medial globus pallidus nucleus (7,8). However, DBS exhibits complications, such as tolerance and electrode displacement, and is associated with a high cost; therefore, it is not a popular approach (9-11).

In previous years, stem cell transplantation therapies have been considered a potential method for the treatment of PD (12,13). In 2006, induced pluripotent stem cells (iPSCs) reprogrammed from somatic cells became a hot topic in research (14,15). Notably, iPSC technology can efficiently generate patient- and disease-specific PSC lines, which can then differentiate into any desired cell type, potentially solving a major problem in stem cell therapy (14,16-19). Given the inherent self-renewal capacity, pluripotency and relatively low immunogenicity of iPSCs (20), they provide a promising patient-derived cell resource for human genetic disease modeling and toxicity studies, thereby reducing the overall cost and associated risks of drug development and clinical trials (21-23). Moreover, functional midbrain DA progenitors and neural progenitor cells (NPCs) derived from iPSCs could survive and restore motor function in the treatment of neurodegenerative diseases (24-26). Furthermore,
long-term survival and function of midbrain-like DA neurons derived from autologous human iPSCs have been reported in a non-human primate model of PD (27,28).

The present study used a lentivirus encoding six reprogramming factors to reprogram rat embryonic fibroblasts (REFs) to generate pluripotent cells in vitro, and the small molecules CHIR99021 and RepSox (CR) significantly enhanced the generation of iPSC colonies. Subsequently, the novel rat iPSCs were directly transplanted into the medial forebrain bundle (MFB) of rats with PD to investigate the functional effects of the transplanted cells in vivo, which provided experimental evidence for studying the pathogenesis of PD and identifying the potential of iPSCs for neural transplantation.

Materials and methods

Animals. A total of 48 specific-pathogen-free, 8-week-old, healthy male Sprague-Dawley (SD) rats (weight ~200 g) and two pregnant SD rats (weight ~350 g) were provided by the Experimental Animal Center of Anhui Medical University (Heifei, China). The rats were maintained in groups of five per cage under a 12-h light/dark cycle, the relative humidity was controlled at 40-70% and the temperature was controlled at 23±2˚C, with ad libitum access to food and water. During the experimental period, if any rat started to show signs of immobility, a huddled posture, inability to eat, ruffled fur or self-mutilation, they were immediately sacrificed. In addition, animals were euthanized to prevent further suffering if they were unable to stand or displayed agonal breathing, severe muscular atrophy, severe ulcers or uncontrolled bleeding. The animals were anesthetized with an intraperitoneal injection of 3% sodium pentobarbital (50 mg/kg; Merck KgaA) before 6-OHDA injection, cell transplantation and perfusion. Subsequently, the rats with unsuccessful modeling were euthanized by cervical dislocation under anesthesia or were injected with an overdose of sodium pentobarbital (150 mg/kg) to prevent further suffering if they were unable to stand or displayed agonal breathing, severe muscular atrophy, severe ulcers or uncontrolled bleeding. The animals were anesthetized with an intraperitoneal injection of 3% sodium pentobarbital (50 mg/kg; Merck KgaA) before 6-OHDA injection, cell transplantation and perfusion. Subsequently, the rats with unsuccessful modeling were euthanized by cervical dislocation under anesthesia or were injected with an overdose of sodium pentobarbital (150 mg/kg) for euthanasia. In addition, complete cardiac and respiratory arrest were observed to verify animal death. All of the experimental animal procedures were approved by the Institutional Animal Care and Use Committee of Bengbu Medical College (Bengbu, China; approval no. 2020-025).

Culture of REFs. Primary REFs were mechanically isolated and cultured from 16 embryonic day (E)16-18 rat embryos as previously described (29). Briefly, uteri were isolated from two E16-18 pregnant SD rats following deep anesthesia via a subcutaneous injection of 3% sodium pentobarbital (50 mg/kg), as aforementioned. The head, limbs, visceral tissues and gonads were removed from the isolated embryos. The remaining body parts were minced and trypsinized using 0.1 mM trypsin/1 mM EDTA solution for 20 min at 37˚C. Cells in the supernatant were collected by centrifugation (200 x g for 5 min at 4˚C), resuspended in fresh high-glucose Dulbecco’s modified Eagle’s medium (H-DMEM; Gibco; Thermo Fisher Scientific, Inc.) supplemented with 10% FBS (Gibco; Thermo Fisher Scientific, Inc.), 1% GlutaMax (Thermo Fisher Scientific, Inc.), 1% non-essential amino acid (NEAA), 100 U/ml penicillin and 0.1 mg/ml streptomycin, and cultured at 37˚C in a 5% CO2 incubator. In the present study, REFs within five passages were used to avoid replicative senescence and genetic characteristic instability. The expression of fibroblast-specific marker genes (CD34, COL1A1 and S100a4) and epithelial cell-specific marker genes (CDH1 and MUC1) were analyzed by reverse transcription-PCR (RT-PCR) as described previously (30-32). The expression of fibroblast-specific markers, including CD34 (1:500; Affinity Biosciences) and Vimentin (1:500; BIOUS) were also detected by immunofluorescence (IF) as described previously (31-35).

Plasmid construction and lentivirus production using second-generation lentivirus system. The cDNA of six reprogramming factors, namely OCT3/4, Sox2, Klf4, c-Myc, Nanog and Lin28 (OSKMNL), were inserted downstream of the EF-1α promoter and upstream of IRES-EF1GFP in the lentiviral vector to construct six Lenti-EF-1α-X-IRES-EF1GFP lentivirus plasmids, (SiDan Sai Biotechnology Co., Ltd.). The day before transfection, 293T cells (gifted by Dr Liang Meng, Bengbu Medical College) were seeded at 10⁷ cells per gelatin-coated T75 flask. The following day, six lentiviral plasmids (10 µg) were co-transfected into 293T cells with pSVSVG (5 µg) and pCMV-dr8.91 (7.5 µg) (both from Addgene, Inc.) using 56 µl FuGENE® 6 Transfection Reagent (Roche Diagnostics) according to the manufacturer’s protocols. Cells were then incubated overnight at 37˚C in an atmosphere containing 5% CO2. At 24 h post-transfection, the medium was replaced. After 48 and 96 h, the virus-containing supernatants were collected and filtered through a 0.45-µm polyvinylidene fluoride (PVDF) filter. Titers of six lentiviruses were then determined according to the proportion of green fluorescence-emitting cells to total 293T transfected cells.

Reprogramming rat fibroblasts to iPSCs. For reprogramming, the initial REFs from a male rat embryo at passage 3 (P3) were co-transduced with six lentiviruses carrying six reprogramming factors (OSKMNL) and GFP-tagged protein at day 0 with a multiplicity of infection of 10 for each lentivirus (10 viral particles/cell) and supplemented with 10 µg/ml polybrene. Cells were incubated in the virus/polybrene-containing supernatants for 24 h at 37˚C, and then the medium was changed to fresh complete medium (H-DMEM containing 10% FBS). At day 2 post-transduction, REFs were re-plated on an irradiated OriCell® ICR mouse embryonic fibroblast feeder layer (Cyagen Biosciences, Inc.). The following day, the culture medium was replaced with DMEM/F-12 supplemented with 1% KnockOut Serum Replacement (Gibco; Thermo Fisher Scientific, Inc.), 0.1 mM β-mercaptoethanol, 1% NEAA, 2 mM L-glutamine, 100 U/ml penicillin and 0.1 mg/ml streptomycin, which was further supplemented with 3 µM CHIR99021 (Sigma-Aldrich; Merck KGaA), 1 µM RepSox (Selleck Chemicals) and 15 ng/ml fibroblast growth factor 2 (R&D Systems, Inc.) on day 4 (36,37). The GFP+/iPS-like colonies [rat iPSCs induced by OCT3/4, Sox2, Klf4, c-Myc, Nanog and Lin28 + CR (RiPSCs-6F/CR)] were mechanically picked 20-30 days after viral transduction and re-cultured on feeder layers. In addition, REFs treated with empty lentiviral particles and CR were used as a negative control and original REFs were used as blank control. The RiPSCs-6F/CR were analyzed for chromosomal alterations by G-bband karyotype analysis at P6. The cells (~1x10⁶ cells) were treated with 1.0 g/l colchicine solution and were then treated with 0.025 M KCl hypotonic
solution for 30 min in a 37°C water bath and with carnoy fixative (methanol:glacial acetic acid, 3:1) in a 37°C water bath for 5 min. Giemsa staining was performed following standard method (38). Subsequently, the well-spread chromosome metaphases were observed under an oil immersion objective (inverted fluorescence microscope; magnification, x1,000) and analyzed with VideoTest-Karyo 3.1 software (NatureGene Corp.).

Alkaline phosphatase (AP) staining and IF. To detect AP activity, the compact cell colonies formed from REFs were washed with PBS three times and stained with an AP kit (cat. no. C3206; Beyotime Institute of Biotechnology) according to the manufacturer's protocol. The full and bright GFP-positive colonies were selected under a fluorescence microscope. To biologically characterize RiPSCs-6F/CR, cells seeded on coverslips were fixed with 4% (w/v) paraformaldehyde (PFA) for 18 min, permeabilized with 0.2% Triton X-100 for 8 min, and blocked with 10% goat serum and 10% donkey serum (both from Jackson ImmunoResearch Laboratories, Inc.) at room temperature for 1 h. Subsequently, the cells were incubated for 12 h at 4°C with the following primary antibodies against pluripotency markers: Anti-OCT4 (1:200; Abcam), anti-Nanog (1:300) and anti-Sox2 (1:400; both from Cell Signaling Technology, Inc.). The samples were then incubated with a cyanine 3 (Cy3) dye-conjugated secondary antibody (1:1,000; cat. no. 711-165-152; Jackson ImmunoResearch Laboratories, Inc.) for 1 h at room temperature. The cell nuclei were counterstained with DAPI (Thermo Fisher Scientific, Inc.) for 15 min at room temperature. Finally, the coverslips were mounted with ProLong™ Gold Antifade Mountant (Invitrogen; Thermo Fisher Scientific, Inc.) and observed under an inverted fluorescence microscope (Guangzhou Micro-shot Technology Co., Ltd.) (1.36).

Analyses of pluripotency markers for RiPSCs-6F/CR

Flow cytometry. Following digestion of RiPSCs-6F/CR with 0.25% trypsin to prepare a single-cell suspension and fixing with 4% (w/v) PFA for 30 min, the cells were permeabilized with 0.2% Triton X-100 for 10 min and blocked with 1% BSA (Sigma-Aldrich; Merck KGaA) + 10% donkey serum at room temperature for 1 h. Subsequently, the cells were incubated overnight at 4°C with the following primary antibodies against pluripotency markers: Anti-OCT4 (1:200; Abcam) and anti-Sox2 (1:400; Cell Signaling Technology, Inc.), and then with a Cy3 dye-conjugated secondary antibody (1:1,000; cat. no. 711-165-152, Jackson ImmunoResearch Laboratories, Inc.) for 1 h at room temperature. Finally, the cells were analyzed immediately by flow cytometry (DxP Athena 1L-3L; Cytek Biosciences) using FlowJo CE V10.1 software (FlowJo LLC).

RT-PCR. Total RNA was extracted from the cells using TRIzol® reagent (Invitrogen; Thermo Fisher Scientific, Inc.) and reverse transcribed into cDNA using PrimeScript™ RT Reagent Kit (Perfect Real time) (Takara Biotechnology Co., Ltd.) with the following parameters: 37°C for 15 min, 55°C for 5 sec and finally 4°C for 5 min. RT-PCR was carried out using TB Green Premix Ex Taq II (Takara Biotechnology Co., Ltd.) with a QuantStudio™ 6 Flex thermocycler (Applied Biosystems; Thermo Fisher Scientific, Inc.). The thermocycling conditions were as follows: Initial denaturation at 95°C for 30 sec, followed by 40 cycles of denaturation at 95°C for 10 sec and at 60°C for 20 sec. The amplified fragments were visualized by 1% agarose gel electrophoresis and stained with Gel-Red (cat. no. D0140; Beyotime Institute of Biotechnology, Inc.), and GAPDH was used as the internal control. The primers were synthesized by Sangon Biotech Co., Ltd., and their sequences are shown in Table SI.

Western blotting. Total proteins were extracted from the REFs and RiPSCs-6F/CR using the total protein extraction kit (cat. no. P0028; Beyotime Institute of Biotechnology) and quantified using a BCA protein assay kit (cat. no. P0010; Beyotime Institute of Biotechnology). Equivalent quantities of protein (40 µg/sample) were separated by SDS-PAGE on 10% gels and electroblotted onto PVDF membranes (0.45 µM; MilliporeSigma). The membranes were blocked with 5% skimmed milk powder diluted in TBS-0.05% Tween at room temperature for 1 h and immunoblotted overnight at 4°C with the following primary antibodies: Rabbit anti-OCT4 (1:200; Abcam), rabbit anti-Nanog (1:300), rabbit anti-Sox2 (1:500; both from Cell Signaling Technology, Inc.) and mouse anti-β-actin (1:1,000; Affinity Biosciences). Subsequently, the membranes were incubated with a HRP-labeled secondary antibody (1:5,000; cat. no. 111-035-003; Jackson ImmunoResearch Laboratories, Inc.) for 1 h at room temperature. The protein bands were then visualized using Clarity Western ECL Substrate (Bio-Rad Laboratories, Inc.) and were semi-quantified using Quantity One software version 1-D (Bio-Rad Laboratories, Inc.).

6-Hydroxydopamine (6-OHDA)-lesioned rat model of PD

Behavioral detection of rat models of PD

Sniff test. A total of 2 weeks after injection of 6-OHDA, the rats underwent a sniff test. Rats with PD usually sniff an unfamiliar environment standing on the ground, with movements of vibrissae and the head tilted upwards (39).

Apomorphine (APO)-induced rotation experiment. At week 2 after 6-OHDA unilateral injection, an intraperitoneal injection of APO (0.5 mg/kg; Sigma-Aldrich; Merck KGaA) was used to induce rotational behavior contralateral to the lesion side. A rotational speed of ≥210 revolutions/30 min was considered as the criterion for successful modeling of rats with PD. The experiment was performed at 4, 8, 12, 20 and 24 weeks after cell transplantation.

Rotarod test. A total of three rats were placed on the three channels of a rotarod test apparatus (SANS, Biotechnology Co., Ltd.). The rats placed on the rod synchronously were...
detected simultaneously to assess motor coordination. The test was processed at a constant speed of 300 rpm for 1,800 sec, and each animal underwent three trials. Each trial was automatically paused, and the time it took for the rat to fall off the rod or run for 1,800 sec was recorded. If a rat fell within 10 sec, three repetitions of the experiment would be performed. The experiment was performed at 4, 8, 12, 20 and 24 weeks after cell transplantation.

**Open-field assay.** Each rat was placed into the central grid of the open-field instrument (Noldus Information Tech, Inc.), and the surrounding curtains were quickly drawn. The EthoVision XT 10.1S system of open-field instrument (Noldus Information Technology BV) automatically captured the track of the rat within 5 min and analyzed its stay in the central grid and the total distance of movement. Zone heatmaps were obtained by tracing the path of the rat in the open field. The experiment was performed at 4, 8, 12, 20 and 24 weeks after cell transplantation.

**Transplantation of RIPSCs-6F/CR into the right MFB of rats with PD.** A total of 2 weeks after 6-OHDA injection, RIPSCs-6F/CR and RIPSCs-6F (rat iPSCs induced by OSMKML only) were resuspended in serum-free H-DMEM at a cell density of 1.25x10^7 cells/ml and stereotactically transplanted into the right MFB of model rats with PD according to the same two stereotaxic coordinates as the PD model. At each site, an aliquot (8 µl) of cell suspension including 1.0x10^5 cells was injected into each rat with PD using a microsyringe at a rate of 0.5 µl/min. The rats were divided into the following four groups: i) Control group containing 12 healthy rats, which were injected with 8 µl saline. A total of 30 model rats with PD (out of the initial 36 rats used for PD modelling, 83.3% success rate) were divided randomly into three groups, including a ii) vehicle group containing 12 PD model rats, which were injected with 8 µl H-DMEM; iii) a RIPSCs-6F/CR group containing 12 PD model rats injected with 1.0x10^5 RIPSCs-6F/CR; and iv) a RIPSCs-6F group containing six PD model rats injected with 1.0x10^4 RIPSCs-6F.

Behavioral analysis was performed on each group at 4, 8, 12, 20 and 24 weeks after cell transplantation. In total, two rats from each group were perfused, and coronal sections of the perfused brains were used for IF detection at 8 and 20 weeks after cell transplantation. On week 12, two rats from each group were perfused for hematoxylin and eosin (H&E) staining and tyrosine hydroxylase (TH)-3,3’-diaminobenzidine (DAB) detection by immunohistochemical analysis. Each test requiring sacrifice of experimental animals was repeated only once, i.e. two rats were sacrificed per test. And, the remaining six rats from each of the three groups were used for long-term monitoring of behavioral changes.

**Histology and immunohistochemistry (IHC).** Rats were deeply anesthetized with intraperitoneal injections of 3% pentobarbital sodium (50 mg/kg), and then transcardially perfused with 0.9% NaCl followed by 4% PFA. During PFA perfusion, the limbs of the rats twitched continuously and became rigid, and the liver and brain became white, thus confirming successful perfusion and rat euthanasia. The brain was collected and fixed in 4% paraformaldehyde at 4°C for 6 h, after which the brain tissue was transferred into 25% sucrose solution until it sank to the bottom at 4°C. The brain tissue was then incubated at -80°C overnight. Subsequently, the perfused brains were embedded using OCT embedding medium (Sakura Finetek USA, Inc.) and serial coronal sections (12 µm) were cut using a cryostat (CM-1850; Leica Microsystems, Inc.), mounted on gelatin-coated glass slides and frozen at -20°C. Subsequently, the sections were permeabilized with 0.2% Triton X-100 for 8 min at room temperature, and blocked with 10% goat serum and 10% donkey serum at room temperature for 1 h. The sections were then subjected to double I F staining using an anti-GFP antibody (1:200) and the following nerve-specific labeling antibodies: Anti-BIII tubulin (TUJ1; 1:500), anti-Nestin (1:200), anti-TH (1:300), anti-paired box protein 6 (PAX6; 1:200), anti-synaptophysin (SYN; 1:1,000; all from Abcam), anti-γ aminobutyric acid (GABA; 1:300; Sigma-Aldrich; Merck KGaA), anti-Sox2 (1:500), anti-postsynaptic density protein 95 (PSD95; 1:300; both from Cell Signaling Technology, Inc.) and anti-glial fibrillary acidic protein (GFAP; 1:300; NeuroMics). Details of all the primary antibodies used in the current study are listed in Table SII. Subsequently, the samples were incubated with appropriate Alexa Fluor 488 (1:500; cat. no. A-21202; Invitrogen; Thermo Fisher Scientific, Inc.) and Cy3-conjugated (1:1,000; cat. no. 711-165-152; Jackson ImmunoResearch Laboratories, Inc.) secondary antibodies, followed by incubation with DAPI for nuclear staining. Images were obtained using a multiphoton laser scanning confocal microscope (FV-1200MPE SHARE; Olympus Corporation).

Perfused brains collected at 12 weeks after cell transplantation were paraffin embedded and cut into 30-µm sections. Antigen retrieval was performed using citric acid antigen retrieval buffer (cat. no. G1202; pH 6.0; Wuhan Servicebio Technology Co., Ltd.) in a microwave oven. To block endogenous peroxidase activity, the sections were incubated in 3% hydrogen peroxide (Disinfection Technology Co., Ltd.) at room temperature in the dark for 25 min. Subsequently, sections were blocked with 3% BSA (G5001; Wuhan Servicebio Technology Co., Ltd.) for 30 min at room temperature. The sections were then incubated with an anti-TH (1:300; cat. no. ab112; Abcam) antibody for 1 h at room temperature. Subsequently, the sections were washed three times with Dulbecco's PBS, and were incubated with a HRP-conjugated goat anti-rabbit secondary antibody (1:1,000; cat. no. 111-035-003; Jackson ImmunoResearch Laboratories, Inc.) for 1 h at room temperature. The sections were then stained with 3,3’-diaminobenzidine tetrahydrochloride solution (cat. no. G1211; Wuhan Servicebio Technology Co., Ltd.) at room temperature; the color developing time (1-10 min) was controlled under take microscope.

Additionally, histopathological examination of 3-µm paraffin-embedded sections was routinely performed using a H&E staining kit (Wuhan Servicebio Technology Co., Ltd.). The paraffin-embedded sections were first dehydrated with xylene, then dehydrated with increasing ethanol concentrations, washed with PBS and stained with H&E staining solution at room temperature for 5 min. The survival and number of TH⁺ cells obtained from TH-DAB and H&E staining were calculated by whole brain scanning using a Nikon Imaging System (DS-U3; Nikon Corporation) and CaseViewer 2.0.
software (3DHISTECH, Ltd.). The number of TH⁺ cells in three randomly selected fields was counted using ImageJ software (1.51r, National Institutes of Health).

Statistical analysis. All quantitative data are presented as the mean ± standard error of the mean from at least three independent experiments. Statistical comparisons between two...
groups were performed using independent Student's t-test. For multiple comparisons, one-way ANOVA followed by Tukey's post hoc test was used to analyze the data. GraphPad Prism software 7.0 (GraphPad Software, Inc.) was used for statistical analyses and to produce the graphs. P<0.05 was considered to indicate a statistically significant difference.

Results

Morphological characteristics and identification of pluripotency of RiPSCs-6F/CR. The reprogramming procedure of rat fibroblasts transduced by the six reprogramming factors (OSKMNL) as well as cR is shown in (Fig. 1A). REFs were isolated, and a substantially homogeneous population of fibroblast-like cells was obtained after subculture for 3-4 passages (Fig. 1B). All REFs could express fibroblast-specific markers CD34 and vimentin, as determined by immunofluorescence (Fig. 1C). Moreover, the fibroblast-specific genes S100a4, COL1A1 and CD34 were also highly expressed in REFs, as determined by RT-PCR analysis (Fig. 1D). However, the REFs at P3 generation did not express epithelial cell marker genes CDH1 and MUC1 (Fig. 1D). Subsequently, REFs were transduced with lentiviral particles expressing OSKMNL, and the RT-PCR results confirmed that all OSKMNL genes were overexpressed after 3 days (Fig. 1E). The REFs were then cultured in the presence of the small molecules cR at day 4 and at day 8 after the initial transduction, small GFP+/iPS-like colonies could be observed. After 20 days, these GFP+/iPS-like colonies (referred to as RiPSCs-6F/cR) were picked and cultured on feeder layers for expansion and further characterization. The cell morphological changes throughout the induction process are shown in (Fig. 1F). In addition, after 20 days of treatment with empty lentiviral particles and cR, REFs showed a tendency to aggregate, but no obvious GFP+/iPS-like colonies could be observed. After 20 days, these GFP+/iPS-like colonies were picked and cultured on feeder layers for expansion and further characterization. The cell morphological changes throughout the induction process are shown in (Fig. 1F).
microscope, it could be observed that the stereoscopic effect was strong and the cells were closely arranged (Fig. 2A).

The majority of RiPSCs‑6F/cR colonies (90%) expressed high levels of AP and maintained iPS‑like morphology with GFP for >25 passages (Fig. 2A). Chromosome G‑banding analysis confirmed that the percentage of RiPSCs‑6F/cR with normal karyotypes 2n=42 was 96.6%, indicating that RiPSCs‑6F/cR were reproducible diploids, and there was no cross‑contamination of cells from other species (Fig. 2B). Specifically, RiPSCs‑6F/cR highly expressed pluripotency‑specific ESC markers, including OCT4, Sox2 and Nanog, as demonstrated by IF and flow cytometric analysis (Fig. 2C and D). RT‑PCR indicated that RiPSCs‑6F/cR highly expressed the six transcription factor genes (OSKMNL; Fig. 2E), and western blotting showed that RiPSCs‑6F/cR also expressed numerous ESC markers at the protein level, including Sox2, OCT4 and Nanog (Fig. 2F).

Figure 3. RiPSCs‑6F/CR significantly ameliorate the behavioral deficits of a rat model of Parkinson's disease. (A) 6‑Hydroxydopamine was stereotactically transplanted into the right medial forebrain bundle of each model rat at the labeled two coordinates. (B) Behavioral evaluation by sniff test, ipsilateral rotation and rotarod test. (C) Experimental rats were fixed on a stereotactic apparatus. (D) Transplantation of RiPSCs‑6F/CR did not lead to tumor formation at 8 weeks after transplantation. (E) Heat maps of open‑field assay. (F‑H) Motor behavior evaluation in three groups at 8 and 24 weeks after transplantation. *P<0.05, **P<0.01 and ***P<0.001 vs. RiPSCs‑6F/cR. RiPSCs‑6F, rat induced pluripotent stem cells induced by OSKMNL; RiPSCs‑6F/cR, rat induced pluripotent stem cells induced by OSKMNL + CR; OSKMNL, OCT3/4, Sox2, Klf4, c‑Myc, Nanog and Lin28; CR, CHIR99021 and RepSox.
Injection of RiPSCs-6F/CR ameliorates the motor deficits of 6-OHDA-lesioned model rats with PD. The PD rat models were prepared by stereotaxic injection of 6-OHDA into the right MFB of SD rats at two coordinates (Fig. 3A). A flow chart of the experimental procedures and animal groups described in the present study is shown in Fig. S1. At week 2 post-injection of 6-OHDA, SD rats exhibited PD-like symptoms, such as tail-pressing, back arching, sniffing and motor coordination disorder (Fig. 3B). In addition, the behavior of continuously turning >210 ipsilateral rotations/30 min to the contralateral side of the injury in APO-induced rotation was considered a main criterion for model rats with PD. A total of 30 rats were successfully modeled (83.3% success rate), as determined by a behavioral test. RiPSCs-6F/CR (1.0x10^5 cells/graft) were stereotactically transplanted into the right MFB of model rats with PD (n=12; Fig. 3C); all of the transplanted rats survived. In addition, transplantation of RiPSCs-6F/CR induced by OSKMN-L-CR did not lead to rejection or tumor formation (Fig. 3D). However, two out of six graft recipients of RiPSCs-6F developed tumors in the brain at 8 weeks after transplantation; therefore, all six rats in the RiPSCs-6F group were euthanized at 8 weeks due to tumorigenicity detection and no further behavioral testing was performed on the RiPSCs-6F group.

A total of 8 weeks after RiPSCs-6F/CR transplantation, the rats were more excited and active than those in the vehicle group, and the total distance of movement was significantly increased in the open-field test (P<0.01; Fig. 3E and F). In addition, the APO-induced rotational behavior was significantly reduced to 225.0±64.2 after 8 weeks of transplantation (Fig. 3D). Therefore, the results suggest that RiPSCs-6F/CR transplantation can ameliorate motor deficits associated with PD, particularly in the early stages of the disease.
8 weeks of transplantation with RiPSCs-6F/CR (P<0.01; Fig. 3G). Furthermore, the motor coordination ability of the RiPSCs-6F/CR group was also effectively improved according to the results of the rotarod test (P<0.05; Fig. 3H). Moreover, the motor deficits of rats with PD in the RiPSCs-6F/cR group were further improved 24 weeks after cell transplantation (Fig. 3F-H), which indicated that transplanted cells required a period to induce functional recovery in vivo. Notably, transplantation of RiPSCs-6F/CR into the MFB may significantly improve the dyskinesia of rats with PD after 8 weeks.

RiPSCs-6F/CR differentiate into targeted TH+ dopamine neurons in the MFB of model rats with PD. Whole-brain H&E staining analysis showed that the number of cells in the 6-OHDA-lesioned area of rats with PD was markedly lower than that of the healthy control group, and the cells were disorderly arranged (Fig. 4A). However, numerous viable rat RiPSCs-6F/CR grafts were observed in the injured area 12 weeks after transplantation and the cells were relatively neatly arranged (Fig. 4A).

In addition, TH-DAB staining of the whole brain showed that, compared with that in the healthy control group, the number of TH+ cells in the injured area of rats with PD (vehicle group) was significantly reduced, and the expression level of TH was also markedly reduced. However, numerous TH+ cells were present in the grafted area of RiPSCs-6F/CR and the surrounding MFB 12 weeks after transplantation. Microscopic imaging showed markedly increased TH labeling in the transplanted MFB compared with that in the rats of the vehicle group, indicating robust recovery of the transplanted MFB from the engrafted RiPSCs-6F/CR. Moreover, stereological...
cell counts of TH⁺ dopamine neurons showed that the number of viable DA neurons in the MFB of the RiPSCs-6F/cR group was significantly higher than that of the vehicle group (Fig. 4B and C). These results were in agreement with behavioral evaluations, and suggested that RiPSCs-6F/cR differentiated into targeted TH⁺ dopamine neurons in the microenvironment of the host brain.

RiPSCs-6F/cR differentiate into various types of functional neurons in the host MFB of rats with PD. The results of IF detection of frozen sections of rat brain tissue showed that GFP-positive cells formed a distinct graft area 2 weeks after transplantation of RiPSCs-6F/cR. Furthermore, a large number of the transplanted cells had migrated 2.3 mm from the graft area into the surrounding brain tissue 20 weeks after transplantation (Fig. 5A). In addition, numerous GFP⁺ cells also stained positive for TH, and certain TH⁺ cell clusters were dispersed throughout the engraftment area and integrated into the host brain (Fig. 5B). Since TH is a specific marker of DA neurons, this suggested that RiPSCs-6F/cR could differentiate into DA neurons in vivo. Moreover, the engrafted RiPSCs-6F/cR gave rise to various functional neurons around and within the graft area, which could express the neuronal marker TUJ1, the GABAergic neuron marker GABA (Fig. 5B), the glutamatergic neuronal marker PSD95 (Fig. 5C) and the glial marker GFAP (Fig. 5D).

Certain GFP-positive cells exhibited features of neural stem cells or neural precursor cells, and expressed the neural stem markers Sox2 and Nestin 20 weeks after transplantation (Fig. 6B-D). This indicated that the transplanted RiPSCs-6F/cR differentiated into neural precursor cells first and then into mature neurons in the brain microenvironment. In addition, the expression of SYN was markedly increased after RiPSCs-6F/cR transplantation, and 52% of GFP⁺ cells were also positive for SYN staining (Fig. 6E). Moreover, numerous SYN⁺, GFP⁻ patches were adjacent to the transplanted RiPSCs-6F/cR, indicating that host brain-derived presynaptic terminals connected with RiPSCs-6F/cR-derived neurons to form mature synapses (Fig. 6E). These data suggested that RiPSCs-6F/cR could differentiate into neural precursor cells, various types of specific functional astrocytes and neurons in the microenvironment of the host brain. Notably, no tumor formation was found in the 12 grafted rats 20 weeks after RiPSCs-6F/cR transplantation.

Discussion

The identification of a method capable of obtaining functional cell types is currently the most basic scientific issue in regenerative medicine research. Direct reprogramming has emerged as a promising approach to induce cell fate transition by introducing a combination of specific transcription factors (40). Moreover, previous reports have demonstrated that reprogramming efficiency could be significantly improved, and different functional cell types could be generated by the presence of certain small molecules, such as valproic acid [VPA, a histone deacetylase (HDAC) inhibitor], CHIR99021 [a glycogen synthase kinase-3β (GSK3-β) inhibitor], butyrate (an HDAC inhibitor), AZA (a DNA
RiPScs‑6F/cR grafts could differentiate into multiple types of engraftment area in the MFB of rats with PD. In addition, of transplantation, the RiPS cs‑6F/cR could form a distinct transplanted into the MFB of rats with PD. After 20 weeks functional active neurocytes and glial cells to promote behavioral recovery of motor dysfunction and neurological function associated with these cells (50‑56). Immune rejection, lower tumorigenicity and better stability of neural cells in vivo, as well the risk of genomic integration of exogenous transcriptional factors (43). In the present study, the small molecules RepSox (TGFβ receptor-1 inhibitor) and CHIR99021 promoted reprogramming and greatly improved the efficiency of RiPSCs‑6F/CR colony generation by ~4.0-fold; notably, 40‑50 iPSC colonies were generated from 1x10⁴ REFs within 25 days of infection.

Numerous studies have shown that inhibition of GSK3‑β by CHIR99021 or inhibition of TGF‑β signaling by RepSox can effectively replace Sox2 and c‑Myc for reprogramming by inducing Nanog (41,43). In addition, GSK3β is a master regulator of Myc threonine 58 phosphorylation and leads to ubiquitin‑dependent degradation of c‑Myc. Therefore, inhibition of GSK3‑β by Wnt signaling can promote self‑renewal and cell reprogramming by regulating the stability of c‑Myc (40,44). Inhibition of TGF‑β could induce mesenchymal to epithelial transition (MET) and increase Nanog expression (36). Together with the present findings, it may be concluded that CR could promote the reprogramming process by simultaneously inhibiting GSK3‑β and TGF‑β signaling. Furthermore, the resulting RiPSCs‑6F/CR had typical ESCs and iPSCs can improve the motor behavioral defects of 6‑OHDA‑lesioned rats by re‑innervating the striatum and restoring DA neurotransmission, but are also associated with the risk of tumor formation and teratomas in vivo, as well as the possibility of undifferentiated cells or proliferating non‑neural cells being present in the cell population (46‑49). To address these issues, the use of various functional midbrain dopamine neurons and NPCs derived from iPSCs has the potential for the treatment of neurodegenerative diseases due to the lower immune rejection, lower tumorigenicity and better stability associated with these cells (50‑56).

In the present study, induced RiPSCs were directly transplanted into the MFB of rats with PD. After 20 weeks of transplantation, the RiPSCs‑6F/CR could form a distinct engraftment area in the MFB of rats with PD. In addition, RiPSCs‑6F/CR grafts could differentiate into multiple types of functional active neurocytes and glial cells to promote behavioral recovery of motor dysfunction and neurological function of rats with PD, such as GFA+, PSD95+, PAX6+, GABA+ and TH+ cells. TH serves a key role in the regulation of DA biosynthesis in DA neurons (3). A sufficient number of surviving TH+ cells (DA neurons) derived from RiPSCs‑6F/CR was indicated to serve an important role in behavioral improvement in the current study. In addition, GABA+ cells (GABAergic neurons) derived from the RiPSCs‑GFP may be responsible for regulating the balance of excitatory and inhibitory signals in the dopamine pathway. In addition, synapse formation between donor- and host‑derived neurons could promote functional recovery and behavioral improvement. Notably, no tumor formation was observed in any of the transplanted rats within 20 weeks of RiPSCs‑6F/CR transplantation; however, two out of six graft recipients of RiPSCs‑6F developed tumors at 8 weeks after transplantation. This result may suggest that not all iPSCs will result in tumorigenesis after transplantation in vivo. It may also be due to the fact that the small molecules CR facilitate reprogramming, and reduce the tumorigenicity of iPSCs in vivo; however, the specific mechanism needs to be further investigated (57,58). It is known that inhibition of GSK3‑β by CHIR99021 can result in activation of β‑catenin/c‑Jun signaling and downregulation of NF‑κB activity to promote apoptosis and inhibit proliferation (59). In addition, inhibition of TGF‑β signaling by RepSox can induce MET and inhibit epithelial to mesenchymal transition, thereby inhibiting cell cycle progression and tumorigenesis (36). Small molecules are non‑integrative to the genome, and are thus much safer and more advantageous than the gene editing method in modulating cell function and cell fate changes (60).

Since the transplanted cells migrated in the microenvironment of the host brain, the percentage of survival and the potential mitotic activity of RiPSCs‑6F/CR after transplantation needs to be further investigated.

In conclusion, in the present study, the small molecules CR significantly facilitated reprogramming and promoted RiPSCs‑6F/CR colony generation during lentivirus‑mediated reprogramming of six transcription factors in REFs. Furthermore, the transplanted RiPSCs‑6F/CR could survive for ≥20 weeks in the MFB and could differentiate into multiple functional neurocytes to ameliorate neurological deficits in 6‑OHDA‑injured rats with PD.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

JX, YL and HZ constructed the reprogramming system and drafted the manuscript, and made an equal contribution to this
work. WW and YL established the Parkinson's disease model. WG and YG participated in the statistical analysis. CM and WW established the Parkinson's disease model. Patient consent for publication Not applicable.

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Competing interests The authors declare that they have no competing interests.

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