The therapeutic landscape in rheumatoid arthritis: why are antigen-specific strategies needed?

Rheumatoid arthritis (RA) is a systemic inflammatory disease resulting from an autoimmune response to self-antigens, leading to inflammation of synovial tissue of joints and subsequent cartilage and bone erosion. Current disease-modifying anti-rheumatic drugs and biologic inhibitors of TNF, IL-6, T cells and B cells block inflammation nonspecifically, which may lead to adverse effects, including infection. They do not generally induce long-term drug-free remission or restoration of immune tolerance to self-antigens, and lifelong treatment is usual. The development of antigen-specific strategies in RA has so far been limited by insufficient knowledge of autoantigens, of the autoimmune pathogenesis of RA and of the mechanisms of immune tolerance in man. Effective tolerance-inducing antigen-specific immunotherapeutic strategies hold promise of greater specificity, of lower toxicity and of a longer-term solution for controlling or even preventing RA. This paper reviews current understanding of autoantigens and their relationship to immunopathogenesis of RA, and emerging therapeutics that aim to leverage normal tolerance mechanisms for implementation of antigen-specific therapy in RA.

Abstract

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Current drugs do not generally induce long-term drug-free remission or restoration of immune tolerance to self-antigens, and lifelong treatment is normally required. Moreover, approximately 30% of RA patients do not reach the desired treatment outcome – due to drug failure rates and/or toxicity [3]. For those patients experiencing serious adverse effects or treatment failure, disease progression may lead to deformity, disability and loss of productivity [4]. Furthermore, uncontrolled inflammation in RA is associated with a higher risk of atherosclerotic disease and cardiac complications with increased mortality [5]. Many of the current effective biologic drugs are very costly to produce in the quantities required to meet demand for the drug, placing a heavy burden on health authorities, patients and the community [6]. There is thus a recognised need for better treatments in RA. Ideally these treatments would target underlying disease pathogenesis with greater specificity, producing fewer side effects.

No therapeutic has yet achieved antigen specificity for controlling symptoms in RA. The development of such strategies has so far been limited by insufficient knowledge of RA autoantigens, of the autoimmune pathogenesis of RA and of the mechanisms of immune tolerance in man. On the contrary, excellent underpinning knowledge of inflammatory mediators of joint pathology, and the potent disease-modifying effect and the excellent revenues achieved by passive transfer of fully or partly humanised antibodies or soluble receptors targeting inflammatory pathways, have stimulated the development of many approaches over the past decades to reduce...
the inflammatory burden of RA. This development has improved the therapeutic options available to doctors and patients, and has captured a huge market in RA, predicted to be $12 billion annually by 2017. Effective tolerance-inducing antigen-specific immunotherapeutic strategies hold promise of greater specificity, of lower toxicity and of a longer-term solution to controlling or even preventing RA. After decades of disappointing results in this area, we are at last moving towards new therapeutic possibilities. This paper reviews current understanding of autoantigens and their relationship to immunopathogenesis of RA, and evolution of therapeutics developed to leverage normal tolerance mechanisms for implementation of antigen-specific therapy in RA.

Self-reactivity to post-translationally modified antigens in RA and its implications for immunotherapy

Autoreactivity towards a variety of self-proteins has been associated with the development and progression of RA. Some of the antigens described are joint-derived proteins, such as type II collagen and human cartilage-derived glycoprotein HCgp39 [7]. Other antigens are stress-associated proteins, including grp78/BiP, which is an intracellular chaperone involved in endoplasmic reticulum stress and angiogenesis in proliferative RA synovial tissue [8,9]. Endoplasmic reticulum stress may be provoked by many factors, including proinflammatory cytokines, cell death, misfolding of proteins in the endoplasmic reticulum, and reactive oxygen species [10]. As a result of intracellular Ca2+ flux in these settings, deamination or citrullination of arginine is enhanced through activation of the peptidyl arginine deiminase enzymes [11]. Owing to the reduction in positive charge, citrullination substantially affects the protein structure and function, for example, of enzymes. These changes can affect the quality and stiffness of the extracellular membrane at inflamed sites [12].

Citrullination also plays a physiological role in apoptosis and epigenetics [13]. Citrullinated self-antigens have emerged as a major group of post-translationally modified autoantigens in RA. Approximately 70% of RA patient sera contain autoantibodies reactive to a variety of citrullinated peptide antigens (ACPA) [14]. These autoantigens include vimentin, fibrinogen, collagen type II, α-enolase, clusterin, histones and peptidyl arginine deiminase-4 itself [11,15,16]. Citrullinated self-proteins are found in inflamed RA joints, but are not specific to RA. Post-translationally modified peptides probably represent an important means to diversify the repertoire of antigens (the immunopeptidome), and thus responding T cells, in response to inflammatory or stressful priming events such as infections. Smoking is an inducer of peptidyl arginine deiminase in the lung, and is likely to be a major trigger of post-translational modification of self-antigens in RA, including citrullination [17,18]. Stress-inducing environmental agents such as smoking, obesity, and exposure to toxins such as silica and mineral oils promote expression of post-translationally modified neo-antigens, which may trigger priming of autoreactive T cells in the peripheral repertoire of individuals genetically susceptible to RA.

The major histocompatibility complex (MHC) contributes about one-third of the genetic susceptibility to RA. Specific RA-associated human leukocyte antigen (HLA)-DR alleles encode a conserved amino acid sequence in the HLA-DR antigen-binding groove, known as the shared epitope (SE) [19]. ACPA – reflecting autoreactivity to citrullinated self-antigens – are much more likely to occur in patients with the HLA-DR SE [20]. Polymorphisms at amino acid positions 11, 71 and 74 of the DRβ chain and single amino acid associations in the peptide binding groove of HLA-B (position 9) and HLA-DPβ1 (position 9) were found almost completely to explain the MHC association with seropositive RA [21]. Amino acids at positions 71 and 74 form part of the fourth anchoring pocket (P4 of HLA-DR). The encoded amino acid at position 71 is positively charged, and therefore preferentially binds nonpolar amino acids, including citrulline, but not the unmodified positively-charged arginine [21].

It has been proposed that tightly-bound post-translationally modified peptide antigens are available for presentation to the T-cell antigenic receptor in the periphery, but not for negative selection in the thymus [22]. Deletion of autoreactive T cells in the thymus depends on strong T-cell receptor (TCR) recognition and signalling by peptide-MHC complexes, which would not occur with nonbinding native self-peptides. Peptides modified by a variety of post-translational modifications, including citrullination, transglutamination and oxidation, are implicated in a number of autoimmune diseases, including multiple sclerosis, type 1 diabetes and coeliac disease [23,24]. Post-translational modifications are likely to represent a general mechanism for immune priming to neoself-antigens for which T cells potentially reactive with the native self-antigen were poorly deleted in the thymus, if at all, due to insufficient binding strength of the native peptide to the susceptibility HLA allele.

Following the initial elucidation of citrullinated T-cell epitopes that bound strongly to HLA-SE molecules, and which stimulated T-cell responses and inflammatory arthritis in HLA-DR4 transgenic mice [25,26], we and others demonstrated citrullinated autoantigen-specific T-cell autoimmunity in RA patients carrying HLA susceptibility alleles [27-29]. We found that proinflammatory cytokines were secreted by peripheral blood (PB) CD4+ T cells of RA patients and healthy controls, in response to citrullinated but not unmodified peptides in
the context of the HLA-SE sequence. RA patient T cells secreted a broader range of cytokines than healthy control T cells. Of the peptides tested, citrullinated aggrecan was most immunogenic [27]. Together, these studies indicate that citrulline does indeed confer the capacity of self-peptides to bind the SE, and that PB T cells from HLA-SE-positive individuals respond to these peptides in vitro.

**Antigen-specific tolerising immunotherapy and dendritic cells**

Loss of tolerance to self-antigens is a critical component in the pathogenesis of autoimmunity. Both central (thymic) and peripheral tolerance mechanisms exist for the control of self-reactive T cells. Active mechanisms of peripheral tolerance include deletion of self-reactive cells after antigen recognition and regulation of self-reactive effector responses by specialised populations of regulatory T cells (Tregs). These mechanisms maintain the tolerant state in order to avoid autoimmune disease long term. Several T-cell and natural killer T-cell populations with the ability to inhibit the response of effector T cells have been described.

Autoreactivity of T cells and B cells towards self-antigens associated with the development and progression of RA presents an opportunity for antigen-specific tolerising immunotherapy to induce antigen-specific regulatory cells, including Tregs. Attempts to harness antigen-specific tolerance to modify the autoimmune response in RA have been made since the early 1990s. These include oral tolerance, TCR peptide vaccination and HLA-DR peptide vaccination. Antigens used in oral tolerance trials include type II bovine or chicken collagen, HCGp39, dnaJp1, and lyophilised *Escherichia coli* extract. Oral tolerance relies on presentation of the relevant peptides by constitutive mucosal dendritic cells (DCs), with the capacity to regulate pathogenic T cells by deletion, immune deviation or through Treg-mediated suppression (Figure 1a). While some evidence for induction of regulatory populations or immune deviation to less pathogenic T-cell cytokines was produced, efficacy of oral autoantigens was limited [30-32].

Tolerogenic antigen presentation after mucosal delivery by mucosal DCs, however, may be unreliable for several reasons. First, there may be a small tolerising antigen dose window, and disease flares may occur due to antigen sensitisation of the immune system [33]. This is a well-described side effect of specific immunotherapy for allergic desensitisation, which relies on immune deviation and induction of Tregs to crude allergen preparations delivered to the skin or sublingually. Second, the capacity of Tregs to suppress pathologic cytokine production by effector T cells is dysfunctional and DCs are activated at inflammatory sites in RA, and therefore unpredictable effects may occur without control over the antigen presenting cells at mucosal sites [34-37]. Third, the gastrointestinal tract of RA patients may be colonised with distinct microflora, including oral *Porphyromonas gingivalis*, which may themselves promote inflammatory arthritis [38]. Finally, effector-memory autoreactive T cells – present in RA synovium – are known to be more resistant to tolerance mechanisms than naïve T cells [39,40].

Alternatively, DCs presenting autoantigen in vitro or in vivo could be modified to promote their capacity to delete antigen-specific T cells or to induce antigen-specific Tregs in *vivo*: so-called tolerising DCs (Figure 1b). Such DCs, when delivered with appropriate autoantigens, should afford greater control over the outcome of antigen-specific immunotherapy through the induction of Tregs. If this strategy were effective, antigen-specific suppression of pathogenic T cells in RA patients would avoid generalised immune suppression, focusing tolerance mechanisms on relevant autoantigens. This effect might be anticipated to restore tolerance towards RA self-antigens with low toxicity, suitable for suppression of early disease or for prevention in individuals at risk.

**Translation of tolerising immunotherapeutic approaches in RA and other autoimmune and inflammatory diseases**

**Oral tolerance**

In animal models, orally administered type II collagen suppressed collagen-induced arthritis as well as adjuvant arthritis – the latter presumably through secretion of anti-inflammatory cytokines by bystander T cells, or linked suppression through induction of collagen type II-specific regulatory cells and their migration to the joint [41-43]. Oral mycobacterial heat shock protein 65 or human gp39 also attenuated collagen-induced arthritis and adjuvant arthritis [44,45]. Oral chicken or bovine type II collagen was safe when administered to patients with RA. While some clinical improvement was noted in open-labelled studies, placebo-controlled trials found no statistically significant improvement in collagen-fed patients, including among patients with early RA [30,46-48]. Oral human gp39 was also trialled by two companies, but with little evidence of efficacy [49]. Oral lyophilised *E. coli* extract contains several bacterial heat shock proteins with immunomodulatory properties. The extract’s clinical efficacy was equivalent to d-penicillamine in a placebo-controlled trial [50]. However, this extract is more likely to be a nonspecific immunomodulator than to induce antigen-specific tolerance.

Oral dnaJp1 was trialled in phase I and phase II clinical trials in RA. This bacterial heat shock protein sequence has been proposed to be cross-reactive with corresponding self-peptides in RA because it is homologous with the...
HLA-DR SE sequence. Oral dnaJp1 had an excellent safety profile and demonstrated immune modulatory effects. Whereas patients generally made Th1-type T-cell cytokine and proliferative responses to dnaJp1 at baseline, dnaJ-specific proliferation and IFNy decreased and IL-4 and IL-10 increased after the treatment [51]. In a placebo-controlled phase II clinical trial, the dnaJp1-treated group showed statistically significant improvement in American College of Rheumatology scores after 6 months of daily oral dnaJp1 peptide, associated with increased expression of regulatory molecules and decreased secretion of TNF in PB [52].

**T-cell receptors in RA and T-cell receptor peptide vaccination**

Diversity is generated through gene rearrangement within the Variable (V) domains of the TCR; namely, the

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**Figure 1. Principles underlying the induction of antigen-specific regulation.**

(a) Mucosal tolerance. Rheumatoid arthritis (RA) self-antigen is delivered to the mucosal immune system by the oral or nasal route. If taken up and presented by the appropriate CD103+ dendritic cells (DCs), which reside in the transforming growth factor-beta (TGFβ)-rich gut lamina propria and draining lymph nodes, antigen-specific regulatory T cells (Tregs) can be induced in the presence of all-trans retinoic acid (ATRA). These Tregs should suppress RA self-antigen-specific immune responses in the joint.

(b) Tolerising DC immunotherapy. DCs with tolerising capacity are generated from peripheral blood monocytes in vitro in the presence of inhibitory drugs to prevent DC activation after contact with Toll-like receptor ligands or cytokines. The DCs are then exposed to RA self-antigen and washed. The antigen-presenting DCs are injected and should induce antigen-specific Tregs in draining lymph nodes. These Tregs should suppress RA self-antigen-specific immune responses in the joint. Tolerizing DC + antigen may also be injected into joints, venous blood or lymphatics. Ag, antigen.
V and Junction gene segments of the Vα chain, and the V, Diversity and Junction gene segments of the Vβ chain. Junctional diversity also occurs through random insertions and deletions at these V-Diversity-Junction regions. This diversity is manifested in the complementarity determining regions (CDRs) of the TCR, which directly interact with the peptides presented by MHC class II (pMHC-II). Despite the potential breadth of the TCR repertoire, there are many examples in anti-viral immunity of biased or restricted Vα or Vβ usage and CDR3 sequences [53]. This effect is known as a public immune response, in which individuals share a similar T-cell repertoire to a given HLA-restricted peptide antigen. Although early studies suggested bias in the T-cell repertoire in RA, their interpretation is limited by the techniques used [54-56]. More recently, using real-time PCR, biased Vβ usage and CDR3 sequences were shown in the synovial fluid T-cell repertoire of RA patients, particularly in patients with HLA-DRB1*0405 [57]. Moreover, evidence of T-cell clonal expansion in synovial tissue of early RA patients was shown by Vβ TCR sequencing. While the most expanded clones were shared between different joints, CDR3 sequences were not shared between patients [58]. This study was insufficiently sensitive to address the question of public TCR sequences, however, because HLA-DRB1*04 alleles were not subtyped to allow comparison of sequences associated with a single MHC allele.

An alternative approach to antigen-specific vaccination is based on evidence from animal models, including arthritis and experimental allergic encephalomyelitis, that antigen-specific T-cell lines or clones prevent priming of autoimmune diseases [59]. The mechanism of this phenomenon was in part related to induction of Tregs reactive with the CDR3 regions of the autoreactive TCRs [60]. The approach has been pursued in a number of trials in multiple sclerosis patients. Although modest benefits have been reported, these are all open-labelled studies (reviewed in [61]). Based on some early work suggesting that Vβ3, Vβ14 and Vβ17 determinants were dominant in RA, a trial was undertaken of Vβ3, Vβ14 and Vβ17 peptides emulsified in incomplete Freund's adjuvant [62]. Vaccination with a limited number of peptides on a population basis assumes the presence of public TCR determinants. Given that patients were not recruited on the basis of a defined HLA-DR allele (no HLA typing was reported), the clinical effects of the peptide vaccination were predictably small [62].

**CD4+CD25+ natural regulatory T cells**

Mice thymectomised on day 3 after birth develop a syndrome of organ-specific autoimmune disease, including oophoritis, gastritis, and/or thyroiditis. The mice can be rescued from illness by transfer of CD4+CD25+ T cells from a syngeneic adult spleen, and depletion of this population from nonthymectomised mice leads to a similar spectrum of autoimmune disease [63-66]. Since these discoveries were made in the 1960s and 1970s, CD4+CD25+ Tregs (known as natural Tregs), which express the transcription factor FoxP3, have been well characterised as a thymic-derived suppressor or regulatory population that regulates effector T-cell proliferative and cytokine responses to self-antigens and foreign antigens, innate inflammatory cells and DCs [67-69]. Owing to their high expression of the high affinity IL-2 receptor CD25, they survive and expand in response to IL-2 and anti-CD3/CD28 activation in vitro and in vivo [70]. Bulk Tregs expanded ex vivo or in vivo (using immature DCs), when transferred to mice with collagen-induced arthritis, slowed the progression of disease in an antigen nonspecific manner, through local effects on the joints [71,72]. Autoantigen-specific TCR-transgenic Tregs expanded using DCs, peptide and IL-2 similarly restored normoglycemia to mice with type 1 diabetes [73]. Transfer of bulk IL-2-expanded and CD3/CD28-expanded Tregs [74] has been translated to patients with type 1 diabetes in a clinical trial, currently underway.

**Induced CD4+ regulatory T cells including T regulatory type 1 cells**

T regulatory type 1 (Tr1) cells can be induced in vitro and in vivo in the periphery by DCs expressing high levels of IL-10 and low IL-12 [75,76]. Tr1 cells secrete high levels of IL-10 and moderate levels of transforming growth factor-beta, granulocyte–macrophage colony-stimulating factor, IL-5 and IFNγ, but little IL-4 [77]. A review article describing a phase I clinical trial of IL-10-induced donor lymphocytes in patients with high-risk hematopoietic malignancies transplanted with CD34+ hemopoietic stem cells from haploidentical donors reported mild acute and no chronic graft-versus-host disease in patients receiving Tr1 cells [78].

Extra-thymic antigen-specific FoxP3+ Treg induction or conversion from CD4+FoxP3-CD25+ naïve T cells has been demonstrated in gut-associated lymphoid tissue and mesenteric draining lymph nodes. All-trans retinoic acid is a key regulator promoting this conversion, in the presence of gut-derived DCs, transforming growth factor-beta and antigen [79,80] (Figure 1a). The CD103+ DC subset, found in gut and mesenteric lymph node, plays a specialised role in Treg conversion, as expression of aldhl1a2 – a retinal dehydrogenase involved in the conversion of retinal (vitamin A) into all-trans retinoic acid – is increased in this subset [81,82]. All-trans retinoic acid promotes the generation of FoxP3+ Treg in the presence of anti-CD3 and in the absence of antigen-presenting cells, indicating that all-trans retinoic acid signals CD25+ Treg precursors through a retinoic acid
receptor, promoting FoxP3 expression in the presence of transforming growth factor-beta [83-85].

**Tolerising dendritic cell immunotherapy**

Tr1 Tregs can be induced in vivo by NF-κB or CD40-deficient DCs. NF-κB is a transcription factor family comprising five subunits involved in two pathways: the classical pathway (including p65 transcriptional activity) plays an important role in innate immunity, and the alternate pathway (including RelB activity) plays a role in immune tolerance and adaptive immune activation [86,87]. RelB activity is required for myeloid DC differentiation and function [88-90]. We showed that RelB regulates antigen-presenting cell function through regulation of CD40 and MHC molecule expression [91,92]. Antigen-exposed myeloid DCs, in which RelB function is inhibited, prevent priming of immunity and suppress a previously-primed immune response. RelB activity thus determines the consequences of presentation of antigen by myeloid DCs. Induction of suppression was specific for the antigen to which DCs had been exposed. This suppression resulted from induction in draining lymph node and spleen of antigen-specific Tregs resembling Tr1 cells, which were CD4+IL-10+. CD4+ splenic T cells from tolerant animals transferred antigen-specific tolerance to primed recipients. The DCs therefore induced an active infectious process of antigen-specific regulation [93].

We and others have developed other means to block production or nuclear translocation of RelB and other NF-κB family members in DCs, including anti-sense and soluble inhibitors, and the resulting modified DCs were consistently tolerogenic through induction of Tregs [91,94,95]. Induction of tolerance in response to one antigen does not simultaneously produce tolerance to antigens expressed elsewhere in the body. This property allows for exquisite specificity in tailoring therapeutic interventions to specific pathogenic antigens without global immune suppression. Given the potential for development of antigen-specific autoimmune immunotherapy using DCs treated with soluble inhibitors of NF-κB, such as Bay11-7082 (Bay-DCs) [94], we translated this concept to a phase 1 clinical trial of autologous Bay-DCs exposed to citrullinated peptides in HLA-DR-SE*ACPA* RA patients. In vitro studies of human blood Bay-DCs demonstrated similar phenotype and in vitro functional capacity to murine Bay-DCs [96]. Preclinical experiments in the mBSA antigen-induced arthritis model demonstrated suppression of established arthritis, equivalent to treatment with soluble TNF receptor, in mice treated with a single dose of Bay-DCs exposed to mBSA but not to control antigen. Skin-test reactivity to mBSA was simultaneously reduced and anti-mBSA antibodies were switched from Th1 to Th2 and regulatory isotypes, including IgG1 and IgA but not IgE [97]. Preliminary clinical data demonstrate the feasibility, safety, systemic and clinical effects of a single intradermal administration of Bay-DCs exposed to citrullinated peptides in RA patients [98].

A number of other approaches have been used to generate tolerising human or murine DCs from PB or bone marrow precursors, based on suppression of DC activation in response to inflammatory stimuli [99]. DCs are commonly generated from precursors, and then incubated with pathogen-associated molecular pattern activators, such as lipopolysaccharide or monophosphoryl lipid A, in the presence of NF-κB inhibitors, such as vitamin D3, glucocorticoids or IL-10, or the mammalian target inhibitor rapamycin. Alternatively, DCs are incubated with TNF, IL-6 or low-dose endotoxin to produce so-called semi-mature or tolerant DCs [100-104]. A phase 0 clinical trial of dexamethasone, vitamin D3 and monophosphoryl lipid A-modulated autologous DCs in RA patients is in progress [104], and interesting phase 1 safety, immunological and clinical data from a clinical trial of modified autologous DCs in long-standing type 1 diabetes patients were recently published [105].

**Conclusions and challenges for the field**

Restoration of tolerance to self-antigens is currently an exciting field. After many years in the doldrums, considerable progress has been made to understand how T-cell regulation occurs and how to control antigen-presenting cells for induction of tolerance. Moreover, persistent efforts to translate cellular therapeutic approaches from mouse models have seen the commencement and early reporting of data from proof-of-concept clinical trials of both tolerising DCs and Treg transfer in autoimmune disease and graft-versus-host disease.

There are several key challenges. First, personalised autologous cell therapy is extremely challenging to develop for widespread clinical use. A major challenge pertains to the regulatory requirements for standardisation, sterility and quality control of cell therapies. If used to obtain PB cells, leukapheresis is associated with a degree of morbidity, and is logistically difficult in many centres. There are also difficulties designing protocols with adequate control groups, and as trials continue in this area there will be difficulties comparing results from individual small trials in which varying cell culture protocols or antigen preparations have been used. The cost of carrying out autologous cell therapy trials is also a major impediment to the scale-up required for later-phase trials. Furthermore, this cost may be justifiable in cancer settings where therapeutic options are few, but this is more difficult in RA.

Clinical trials of various forms of antigen-loaded tolerising DCs or antigen-specific Tregs will therefore be useful to demonstrate proof-of-concept for RA.
immunotherapy. However, the future challenge is to translate such a concept to a therapeutic that is more broadly applicable than adoptive transfer of autologous cell products. To achieve this challenge, we formulated liposomes to target antigen and NF-κB inhibitor to DCs in situ. Curcumin, a natural lipophilic NF-κB inhibitor and antioxidant, was incorporated into 40 nm egg phosphatidyl choline liposome particles, which were also loaded with antigenic proteins or peptides. The liposomes encapsulated antigens and lipophilic NF-κB inhibitors stably and efficiently. We injected loaded liposomes into mice with antigen-induced inflammatory arthritis. When delivered intravenously or subcutaneously, the liposomes targeted antigen-presenting cells including DCs, macrophages and B cells in situ, suppressing responsiveness to NF-κB and inducing antigen-specific Tregs. This regulatory mechanism suppressed effector T-cell responses and the clinical signs of full-blown antigen-induced arthritis [106]. These results demonstrate that DC function can be effectively altered by NF-κB inhibitors using in vivo targeting.

The second major challenge is that the diversity of autoantigens in RA raises questions about which one(s) to target, and whether targeting a single autoantigen would fail due to subsequent epitope-spreading in autoreactive recipients. The mechanism of tolerance induction is relevant in this regard. Regulatory or suppressive T-cell responses targeted to specific antigens (antigen-specific Tregs) can simultaneously suppress bystander responses in the same location [107]. One might therefore anticipate that joint inflammatory responses might be suppressed by arthritogenic antigen-specific Tregs of a single specificity even if autoreactivity to multiple autoantigens were present. On the contrary, tolerance elicited by deletion or anergy of antigen-specific T cells would be particularly prone to failure, when autoreactive T cells of new autoantigenic specificities are induced by epitope spreading [108]. This issue brings perhaps the key challenge to the fore: the need for effective reagents to detect and measure the responses of autoantigen-specific CD8+ T cells in RA.

The development of multimeric pMHC complexes has revolutionised the detection of antigen-specific T cells in mice and humans. Tetramer or multimer reagents consist of four or more soluble MHC molecules each loaded with a single peptide bound to streptavidin and coupled to a fluorescent marker. By mimicking pMHC complexes normally expressed by antigen-presenting cells, tetramers bind to the receptors of antigen-specific T cells and allow them to be enumerated and functionally characterised or expanded. pMHC-I tetramers have greatly enhanced understanding of the CD8 T-cell response in viral infections and cancer. However, stable and specific pMHC-II tetramer production is technically more challenging. Nevertheless this technology has recently improved for the production of pMHC-II (human HLA-DR) tetramers, facilitating the understanding of autoimmune responses [109]. Low-frequency antigen-specific T-cell populations have been identified in the blood of patients with coeliac disease and type 1 diabetes, and analysis of the TCR repertoire can greatly enhance understanding of the way that post-translationally modified peptides may be driving autoimmune disease processes [110]. Although it is clear that ACPA are highly specific for RA, the understanding of T-cell responses towards such autoantigens in RA is still rudimentary. Ongoing development of a toolkit of more sensitive pMHC-II tetramer biomarkers, so that autoantigen-specific T-cell responses can be monitored during clinical trials of antigen-specific therapy, is clearly essential for ongoing development in this field.

ACPA may predate the onset of clinical RA by up to 15 years [111]. Addition of assays for rheumatoid factor isotypes and for multiple serum cytokines and chemokines has been shown to enhance the specificity for prediction of RA disease onset [16,112]. These advances in biomarkers for prediction of future disease, as well as evidence that treatment of patients with undifferentiated arthritis may delay the diagnosis of RA, suggest that preventive approaches might eventually be feasible in genetically at-risk individuals or families [113,114]. Clearly, demonstration of safety and efficacy using antigen-specific immunotherapeutic strategies in patients with early established RA would be an important first step along that road.

Abbreviations
ACPA, anti-citrullinated protein antibodies; CDR, complementarity determining region; DC, dendritic cell; HLA, human leukocyte antigen; IFN, interferon; IL, interleukin; MHC, major histocompatibility complex; NF, nuclear factor; PB, peripheral blood; PCR, polymerase chain reaction; pMHC, peptides presented by MHC; RA, rheumatoid arthritis; SE, shared epitope; TCR, T-cell receptor; Th, T-helper type; TNF, tumour necrosis factor; Tr1, T regulatory type 1; Treg, regulatory T cell; V, variable gene segment.

Competing interests
RT has filed a provisional patent surrounding technology for targeting DCs for antigen-specific tolerance, and is a director of a spin-off company that is commercialising vaccines which target DCs to suppress autoimmune diseases.

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