Clinical Efficacy of Electroneurography in Acute Facial Paralysis

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The estimated incidence of acute facial paralysis is approximately 30 patients per 100000 populations annually. Facial paralysis is an extremely frightening situation and gives extreme stress to patients because obvious disfiguring face may cause significant functional, aesthetic, and psychological disturbances. For stressful patients with acute facial paralysis, it is very important for clinicians to answer the questions like whether or not their facial function will return to normal, how much of their facial function will be recovered, and how long this is going to take. It is also important for clinicians to treat the psychological aspects by adequately explaining the prognosis, in addition to providing the appropriate medical treatment. For decades, clinicians have used various electrophysiologic tests, including the nerve excitability test, the maximal stimulation test, electroneurography, and electromyography. In particular, electroneurography is the only objective measure that is useful in early stage of acute facial paralysis. In this review article, we first discuss the pathophysiology of injured peripheral nerve. And then, we describe about various electrophysiologic tests and discuss the electroneurography extensively.

KEY WORDS: Acute facial paralysis · Electrophysiological test · Electroneurography · Electromyography.

Introduction

Acute facial paralysis is an acute peripheral facial weakness of various etiologies and its diagnosis can be established without difficulty in patients with unexplained unilateral isolated facial weakness. However, bilateral facial paralysis is more difficult to be notified than unilateral involvement because bilateral facial paralysis makes symmetric weakness. The onset is sudden and symptoms typically peak within a few hours to days. The most common cause of acute onset unilateral peripheral facial weakness is Bell’s palsy. Other etiologies include viral infection (herpes zoster virus, human immunodeficiency virus), Guillain-Barre syndrome, autoimmune disease, Lyme disease, Kawasaki disease, head or ear trauma, temporal bone fracture, barotrauma, acute or chronic otitis media, cholesteatoma, sarcoidosis, Melkerson-Rosenthal syndrome, and cerebrovascular accident [1].

Most facial weakness is apparent to a clinician as well as a patient. Its clinical diagnosis is based on both static and dynamic facial analysis during physical examination, and attempts have been recently made to standardize an objective measurement of facial function, for example, digital photographic and videographic interactive computer systems [2-5]. Several systems of clinical measurement of facial nerve function have been devised, but since the mid-1980s, the House-Brackmann grading system has been most widely accepted and endorsed by the American Academy of Otolaryngology-Head and Neck Surgery. However, in this grading system, regional descriptions of facial paralysis within each grade can overlap and lead to confusion in determining the appropriate grade. Therefore, modified House-Brackmann grading system [1] is made, focusing major functional criteria of the House-Brackmann system (absolute movement, synkinesis, eye closure, asymmetry at rest, and absolute paralysis). One of the limitations of both grading systems is the inadequacy...
to stratify the degree of degeneration or reflect the prognosis of facial paralysis.

For decades, clinicians have searched the prognostic tests of sufficient accuracy for acute facial paralysis. Since Esslen [6] introduced the use of electroneurography (ENoG) in the early 1970s, the prognosis of facial paralysis has been predicted based mainly on various electrophysiologic tests, including the nerve excitability test, the maximal stimulation test, ENoG, and electromyography (EMG). In particular, ENoG can determine the percentage of degenerating nerve fibers in early phase of acute facial paralysis. Various studies [7-11] have shown that ENoG can be used to predict the prognosis of acute facial paralysis and May and Shambaugh [12] reported that degeneration ≤ 25% within the first 2 weeks of onset indicated a satisfactory recovery of the facial function in 98% of Bell’s palsy cases.

This review article stated the clinical efficacy, advantages and disadvantages of ENoG as the prognostic test of acute facial paralysis. It also described the comparison with other electrophysiologic test, including nerve excitability test (NET), maximal stimulation test (MST), and EMG.

Review

Why is the electrophysiologiical test important?

Facial paralysis can leave aesthetic and functional sequelae to patients and it is very important to patients as well as clinicians to choose the best treatment options and determine its prognosis. However, subjective judgment such as the House-Brackmann grading system cannot give enough or objective information on the paralysis status, especially with regard to the treatment and prognosis. Therefore, objective recording and measuring the degree of neural degeneration and resultant myopathy have been used to determine the physiologic degree of nerve injury and predict the prognosis.

Electrophysiologic test is one of supportive tools for the diagnosis of neural-muscular system but not a method to diagnose the disease or confirm the etiology. Before the electrophysiologic tests, the history of the facial paralysis and physical examination are preceded and considered to interpret the results of electrophysiologic tests. The purposes of electrophysiologic tests are to localize the lesion site along the nerve, determine the severity of the injury, and differentiate whether an injured nerve is still degenerating or regenerating.

The pathophysiology of peripheral nerve after the injury

To understand the concept of peripheral neural injury, we should understand what happens after the injury of peripheral nerve. After the injury of peripheral nerve, pathophysiologic changes depend on the severity of the injury as well as the proximity of the injured segment to the cell body. After mild injury like neuropraxia, focal demyelination and remyelination occur. However, retrograde degeneration and regeneration of the axon occur in the case of severer injury. In mild injury, the regenerative and repair processes begin almost immediately, but nerve regeneration begins only after Wallerian degeneration has run its course in severer injury.

Degeneration of the injured nerve

Before regeneration of nerve fibers can occur, a series of degenerative processes must take place. Transection of the axon divided the nerve into proximal segment connecting to a cell body and distal one. Two segments retract in the direction opposite to the transection site each other and two axonal stumps become to be swollen because of accumulation of the cytoskeleton elements along fast axonal transport and slow axoplasmic flow. Within 24 hours post-injury, the nucleus migrates to the periphery of the cell and Nissl granules, rough endoplasmic reticulum, breaks up and disperses (chromatolysis). This chromatolysis ends in 10–21 days after the injury. Within 3 days post-injury, axonal sprout starts to grow from proximal stump and the rate of axonal regeneration is generally estimated to be 1 mm per day. Within 24 hours post-injury, Wallerian degeneration of axon and myelin sheath occur and Schwann cells proliferate. In Wallerian degeneration, the primary change is physical fragmentation of both axons and myelin sheath and both neurotubules and neurofilaments become disarrayed. By 48 to 96 hours post-injury, axonal continuity is lost and nerve conduction is lost. Myelin disintegration lags slightly behind that of axons but is well advanced by 36 to 48 hours. Disintegrated debris of axon and myelin sheath is removed by phagocytosis of macrophages within 12–14 days post-injury. Schwann cell plays a key role in Wallerian degeneration. It becomes active within 24 hours post-injury, divides rapidly into differentiated daughter cells, and up-regulates the gene expression for proteins to assist in the degeneration and repair process. Initial role of Schwann cell is to help removal of degenerated debris of axon and myelin sheath. Schwann cell and macrophage work together for phagocytosis and clear the site of injury in a process that requires 2 week to 3 months [13-16]

Regeneration of the nerve

Regenerative and repair processes begin almost immediately, although regeneration of the severer injured nerve begins only after Wallerian degeneration has run along the nerve. For more severe nerve injuries more than 3rd degree of Sunder-
land classification (neurotmesis of Seddon classification), endoneurial tube is disrupted and axon cannot regenerate into its original sheath. Proliferated Schwann cells form cytoskeleton framework (Bungner band) connecting both ends of transected nerve. The earliest signs of nerve regeneration are visible changes in the cell body that mark the reversal of chromatolysis. The metabolic machinery of the cell body is reprogrammed to produce proteins and lipid needed for axonal regrowth during the regeneration process. Both fast and slow axoplasmic transports supply the cytoskeletal materials from the cell body to the sites of axonal regeneration but this process results in swelling of both stumps of transected nerve within several hours post-injury. Axonal regrowth begins as early as 24 hours post-injury. During regeneration, axonal regrowth may be impeded by fibrous tissues and regenerated nerve with myelin sheath can make scar neuroma. Multiple axon sprouts may enter into each endoneurial sheath, even in milder injuries, that do not involve destruction of the sheath itself. However, only one axon sprout becomes to be myelinated. Sometimes, axon sprout may enter into endoneurial tube other than its own. If one axon sprout enters into endoneurial sheath other than its own, we called it as “simple misdirection”. If multiple branches of one axon sprout enter into endoneurial sheath other than its own, we called it as “complex misdirection”. Clinical examples of complex misdirection are synkinesis and mass movement. Axon sprout which does not enter into any endoneurial sheath becomes atrophic and breaks down [17,18].

The pathophysiologic concept of peripheral neural injury

To interpret the results of electrophysiological tests, we should understand the pathophysiologic concept on the degree of neural injury. In 1943, Seddon [19] described three basic types of peripheral nerve injury that include neuropraxia, axonotmesis, and neurotmesis (Fig. 1). In 1951, Sunderland [20] expanded Seddon’s classification to five degrees of peripheral nerve injury (Fig. 1). The 1st degree is essentially the same as neuropraxia of Seddon classification. The 2nd degree is same as axonotmesis of Seddon classification. The 3rd degree is axonotmesis as well as the disruption of endoneurium (intact epi- and perineurium). The 4th degree is axonotmesis, as well as the disruption of endo- and perineurium (intact epineurium). The 5th degree is same as neurotmesis of Seddon classification (complete transection). Sunderland classification is more suitable for the acute traumatic facial paralysis and Seddon classification is for acute inflammatory facial paralysis, such as Bell’s palsy. Neuropraxia means local injury of myelin with the axon still intact and functional and is considered as a temporary paralysis of the nerve fiber. So, the nerve action potential can propagate along the nerve in the case of neuropraxia (Seddon classification) or 1st degree injury (Sunderland classification). Major weakness of electrophysiological tests is that they cannot differentiate between axonotmesis and neurotmesis (Seddon classification) or between 2nd to 4th degree injuries (Sunderland classification).
Which electrophysiological tests are used to evaluate the facial nerve?

NET, MST, ENoG and EMG are representative electrophysiological tests, which have been widely used clinically. Among these, NET and MST involve the examiner’s visual evaluation of electrically-elicited facial movement. Therefore, these two tests have possible subjectivity. NET is the oldest and best known electrophysiological test with well-established clinical efficacy, introduced by Laumans and Jongkees in 1963 [21]. During this test, the lowest current elicits a facial twitch is defined as the threshold of excitation and the difference in thresholds between the two sides is calculated. During MST, facial movement of the paralyzed side is compared with that of the normal side at the level of maximal stimuli (current level at which the greatest amplitude of facial movement is seen at the normal side). Contrary to NET, MST and ENoG, EMG is the sole electrophysiological test which is very useful after loss of nerve excitability and completeness of degeneration. After 2 to 3 weeks after acute facial paralysis, tests of electrical stimulation (NET, MST, and ENoG) are no longer useful. After 10 to 14 days post-onset, fibrillation potentials or positive sharp waves seen on EMG can confirm the degeneration of facial nerve. In addition, polyphasic reinnervation potentials more useful, which may be seen as early as 4 to 6 weeks after the onset of paralysis.

What is the ENoG?

The ENoG records compound muscle action potential (CMAP) of facial muscle, which is elicited electrically. The facial nerve is stimulated transcutaneously at the stylomastoid foramen using a bipolar stimulating electrode. Responses to maximal electrical stimulation of the two sides recorded electrically by the second bipolar electrode pair placed in several sensory regions of facial nerve branches and compared between both sides. Contrary to NET or MST, ENoG calculates electrically-evoked responses objectively for the amplitude of electrically-evoked response is measured in mV. Typically, ENoG is delayed until 72 hours post-onset of paralysis because it takes some 72 hours for Wallerian degeneration to propagate from intratemporal portion (injured site) to portion distal to the stylomastoid foramen (electrically-stimulated site during ENoG). Other end of the timing window is the 21 day post-onset of paralysis. After this time, the nerve excitability is lost and the nerve degeneration is completed. In addition, ENoG may be interfered by possible collateral nerves which are regenerated after 2 weeks post-onset. Clinicians should keep in mind that ENoG should be performed for the first time at about 72 hours post-onset and again at 3 to 5 day intervals until a trend and confirmation can be determined. It is good to test the patient serially until a plateau can be determined.

The factors influencing the test results are 1) electrical impedance between electrodes and skin, 2) conduction velocity of the facial nerve, 3) transmission velocity at the junction of neuromuscular junction, 4) propagation velocity along the facial muscle, 5) the degree of synchrony of the facial muscle fibers, and 6) the population of facial nerve fibers still intact. Other factors include sweat and oil status of the skin, diameter of the electrodes, distance between electrodes, location of the electrodes, pressure on the electrodes, skin impedance, and response of the masseter muscles [22,23]. When proper ENoG response is not elicited, clinicians should consider 1) whether or not the electrodes are detached, 2) whether or not the stimulating electrode is malfunctioning, 3) whether or not the facial nerve is totally degenerated, or 4) whether or not the trigeminal nerve is stimulated.

During analysis of ENoG results, latency is not important. Of primary importance is the amplitude of CMAP of the facial nerve. The percentage response of ENoG is defined as a percentage of the amplitude of the paralyzed side divided by the amplitude of the normal side. Alternatively, percentage degeneration is calculated by 1 minus percentage response. The test-retest variability of ENoG amplitude is 6–20% [22,24]. ENoG test-retest variability between sides is known to be 3–20% in normal healthy subjects and asymmetry more than 30% is considered as a clinically significant difference [25].

The major advantage of ENoG is its usefulness to predict the prognosis in early stage of acute facial paralysis. Because conduction block and axonal degeneration progress together in Bell’s palsy, ENoG has the advantage of evaluating the portion of the axons which are not degenerated. Esslen [26] and Fisch [27] demonstrated that in the cases with Bell’s palsy of 95% degeneration, the prognosis for return of facial nerve function was greatly reduced, with a 50% chance of unfavorable recovery. May, et al. [8] reported the results of ENoG performed within the first 10 days after onset in cases with complete paralysis, which showed that percentage response of less than 10% was highly correlated with incomplete recovery, whereas percentage degeneration of less than 25% has a 98% chance of a satisfactory recovery. Fisch [28] stated that percentage degeneration of 95% within 2 weeks gave a 50% chance of a poor recovery and more gradual decrease in ENoG amplitude was related with a much better prognosis.

As mentioned above, it is important to repeat ENoG until a trend and confirmation can be determined. For example, in the case with percentage degeneration of 50% but complete palsy, we can draw that this facial paralysis may result from conduction block and the prognosis may be excellent. ENoG
is useful to predict the prognosis only during the first 2 weeks after the onset. After then, accurate prediction using ENoG is difficult because the regeneration process already begins at the proximal end or the collateral neural circuit can develop from a healthy nerve around a degenerated nerve.

**Conclusion**

ENoG is considered the most valuable test to predict the prognosis and its main indication is acute complete facial paralysis. Both the percentage degeneration and rate of degeneration are prognostic indicators. In Bell’s palsy, percentage degeneration of more than 90% within 14 days post-onset of complete paralysis indicates poor prognosis in more than 50% of cases.

Although electrophysiological tests provide useful neurophysiological information and serve as a guide to the management of acute facial paralysis, they have some limitations. They cannot distinguish between axonotmesis and neurotmesis and are not useful in cases of incomplete paralysis. In addition, they cannot give any useful information within first 72 hours after the onset.

**Conflicts of interest**

The author has no financial conflicts of interest.

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