ABSTRACT

Background: Many factors are affecting intrauterine growth. The role of Wingless-type (Wnt) inducible signaling pathway protein-1 (WISP1), a novel adipokine and placental proteoglycans in intrauterine growth, is not known. We aimed to measure umbilical cord blood levels of glucose, insulin, leptin, WISP1, and placental proteoglycans [glypican-1 (GPC1), glypican-3 (GPC3), and syndecan-1 (SDC1)] which are thought to have an important role in fetal growth and investigate their relation with birth weight.

Methods: Full-term neonates were included in this prospective, cross-sectional study and classified as appropriate for gestational age (AGA), small for gestational age (SGA), and large for gestational age (LGA) according to their birth weight. Umbilical cord blood levels of glucose, insulin, leptin, WISP1, GPC1, GPC3, and SDC1 were measured.

Results: Leptin levels were higher in LGA newborns compared to AGA and SGA newborns, while WISP1, GPC1, GPC3, and SDC1 levels were not different between the three groups. Leptin and GPC1 levels were higher in infants of mothers with gestational diabetes mellitus compared to infants of non-diabetic mothers, while WISP1, GPC3, and SDC1 were not different between the groups. Leptin was positively correlated with insulin, birth weight, and maternal weight. While there was a strong correlation between the WISP1, GPC1, GPC3, and SDC1 levels; there was no correlation between the birth weight, maternal weight, glucose, insulin, and WISP1, GPC1, GPC3, and SDC1 levels.

Conclusion: Umbilical cord blood levels of GPC1, GPC3, SDC1, and WISP1 were not different between SGA, AGA, and LGA infants. The significance of serum levels of these adipokines and proteoglycans remains to be elucidated.

Keywords: Umbilical cord blood, WISP1, proteoglycan, glypican-1, glypican-3, syndecan-1

INTRODUCTION

Fetal growth is a process that depends on the interaction of fetal, placental, and maternal environment and is controlled by genetic, hormonal, nutritional, and maternal factors. The developmental origins of health and disease hypothesis, also known as Barker hypothesis, proposes that suboptimal fetal environment and impaired fetal growth may have significant consequences on an offspring’s long-term health, such as obesity, metabolic syndrome, and cardiovascular disease. Both large for gestational age (LGA) and small for gestational age infants.
There is growing evidence that adipose tissue plays an important role in fetal growth. The placenta affects fetal growth by regulating the transfer of various nutrients to the fetus. Adipokines secreted from adipose tissue can affect inflammation and the transfer of placental nutrients to the fetus. Leptin is a key adipokine mainly produced in white adipose tissue. It has a critical role in energy homeostasis and neuroendocrine regulation of body fat content. A systematic review and meta-analysis reported a positive and moderate correlation between umbilical cord leptin levels and birth weight in different population groups.

Wingless-type (Wnt) inducible signaling pathway protein-1 (WISP1) is a novel adipokine, whose serum levels are elevated in obesity and insulin resistance. Several studies suggest that WISP1 may have a role in the impaired glucose metabolism. Proteoglycans represent a special class of glycoproteins that are highly glycated. It consists of a core protein with a glycosaminoglycan chain attached by one or more covalent bonds. Glypicans are heparan sulfate proteoglycans and human genome comprises six glypican family members (glypican-1 to glypican-6). The placenta contains heparan sulfate and chondroitin sulfate or dermatan sulfate proteoglycans. Placental heparan sulfate proteoglycans enclose glypicans, syndecans, and perlecans and of the glypican family only glypican-1 (GPC1) and glypican-3 (GPC3) are expressed in the placenta. In a recent article, both placental expressions of GPC1 and GPC3 were reduced in SGA pregnancies when compared with appropriate for gestational age (AGA) pregnancies. Similarly, placental expression of syndecan-1 (SDC1) was significantly decreased in fetal growth restriction samples. Therefore, it is likely that fetal growth is closely related to the placental expression of glypicans.

In the present study, we aimed to investigate, for the first time, the umbilical cord levels of placental proteoglycans (GPC1, GPC3, SDC1), WISP1 and leptin levels in infants with SGA, AGA, and LGA.

**MATERIALS AND METHODS**

**Subjects**

This prospective, cross-sectional study was carried out in term neonates who were born at Department of Gynecology and Obstetrics of Aydın Adnan Menderes University between May 2019 and December 2019. The study was conducted according to the Helsinki Declarations, with ethical approvals obtained from the Aydın Adnan Menderes University, Faculty of Medicine (no. 2019/83). Informed consent was obtained from the parents of each infant.

The gestational age was determined according to the last menstrual date and/or ultrasonography of the first trimester. Body weight of infants was measured using a portable, electronic weighing scale sensitive to the 10 g (Seca, Hamburg, Germany). Fenton growth charts were used to evaluate auxologic parameters and infants with a birth weight below the 10th percentile were defined as SGA, those between the 10th and 90th percentiles as AGA, and those with a birth weight above the 90th percentile were defined as LGA. Maternal weight was measured just before the birth. We defined gestational weight gain during pregnancy as the difference between weight before pregnancy and weight just before birth (measured without heavy clothing).

**Homeostatic Model Assessment for Insulin Resistance (HOMA-IR)** was calculated using the formula: (fasting glucose x fasting insulin)/405. All pregnant women were screened using a 50 g glucose challenge test at 24-28 weeks of gestation. Women with post-load glucose ≥140 mg/dL underwent a standard 100 g, 3-hour oral glucose tolerance test. The diagnosis of gestational diabetes mellitus (GDM) was made if at least 2 of 4 diagnostic criteria were met: (i) fasting plasma glucose ≥95 mg/dL, (ii) for 1 hour ≥155 mg/dL, (iii) 2 hours ≥140 mg/dL, and (iv) 3 hours >140 mg/dL.

**Exclusion Criteria**

Infants with a major congenital anomaly or chromosomal abnormality, history or findings of any inherited metabolic disease, a history of congenital or perinatal infection, an Apgar score of <8 in the first and fifth minutes, antenatal maternal steroid prophylaxis, pregnancies complicated by preeclampsia or any chronic disease, a history of alcohol or smoking, and premature born infants (gestational age <37 weeks) were excluded from the study.

**Laboratory Measurements**

After birth, umbilical cord blood collected from patients was immediately centrifuged at 1500× g for 10 minutes and serum samples were separated. Serum samples were stored at −80 °C before performing assays and were thawed only once prior to use. Serum glucose and insulin levels were measured by a clinical chemistry analyzer (Abbott ARCHITECT C8000 and i2000, respectively). Serum levels of leptin, SDC1, GPC3, GPC1, and WISP1/CCN4 were measured by a commercial enzyme-linked immunosorbent assay kit according to the manufacturer’s instructions (FineTest, Wuhan). Samples were measured with a microplate reader at 450 nm wavelength (Epoch, Biotek). All assays coefficient of variation were determined with inter-assay cv: <10% and intra-assay cv: <8%, respectively.

**Statistical Analysis**

Statistical analyses were performed using the SPSS Statistics for Windows, Version 26.0 (IBM SPSS Statistics for Windows, Version 26.0. Armonk, NY: IBM Corp). Categorical data were presented with n and %, and numerical data with mean ± standard deviation if normally distributed, and median (25–75p) if non-normally distributed. Descriptive statistics (kurtosis and skewness), visual methods (histogram), and analytical tests (Shapiro–Wilk test) were used to determine the normal distribution of numerical variables. In the comparison of independent two groups, a student t-test was used if the data were normally distributed, and Mann–Whitney U test was used if the data were non-normally distributed. In the comparison of independent three groups, ANOVA was used if the data was normally distributed (when an overall significance was observed, Tukey’s and Games–Howell post hoc test were performed in case of homogeneity of variances, or not, respectively), and Kruskal–Wallis test was used if the data were non-normally
distributed (when an overall significance was observed, Dunn’s post hoc test was performed). Pearson correlation was used if the data were normally distributed, and a Spearman correlation test was used if the data were non-normally distributed. Type I error was determined as 5% and a \( P \) value was < .05 was considered statistically significant.

**RESULTS**

A total of 84 term neonates were included in the study. Seventeen infants were SGA (20%), 49 were AGA (58%), and 18 were LGA (22%). Median gestational age was 38 weeks and mean birth weights of the SGA, AGA, and LGA infants were 2388, 3321, and 4055 g, respectively. Demographic and clinical features of the subjects are given in Table 1. Mean serum glucose levels were similar between SGA, AGA, and LGA infants. Median serum insulin level was higher in LGA infants than those of SGA infants (\( P = .010 \)). Similarly, median leptin levels of LGA infants were found higher compared with the AGA and SGA infants (\( P = .002 \), Figure 1). Besides, serum levels of WISP1, GPC1, GPC3, and SDC1 levels were not different between SGA, AGA, and LGA infants (Table 1). Also, similar results were observed when infants of mothers with GDM were not included in the analysis (data were not given).

Thirty infants (36%) were born to mothers with GDM. Birth weight, birth weight SDS, maternal weight at birth, serum levels of leptin, and GPC1 were higher in infants of GDM mothers compared to infants of non-diabetic mothers (Table 2). When infants of GDM mothers were compared according to their birth weight; serum WISP1, GPC1, GPC3, and SDC1 levels were not different between SGA, AGA, and LGA infants. Maternal weight at birth and serum leptin levels were higher in LGA infants of GDM mothers than those of AGA infants of GDM mothers (\( P < .001 \) and \( P = .008 \), respectively).

A positive correlation was found between leptin and insulin levels (\( r = 0.291, P = .007 \)), birth weight (\( r = 0.422, P < .001 \)), birth weight SDS (\( r = 0.388, P < .001 \)), and maternal weight (\( r = 0.422, P < .001 \)). Similarly, a strong positive correlation was demonstrated between WISP1 and GPC1 (\( r = 0.736, P < .001 \)), GPC3 (\( r = 0.731, P < .001 \)), and SDC1 (\( r = 0.652, P < .001 \)). There was no correlation between the birth weight, maternal age, maternal weight, glucose, insulin levels, and serum levels of WISP1, GPC1, GPC3, and SDC1 (Figure 2).

![Figure 1. Comparison of umbilical cord leptin levels of infants. SGA, small for gestational age; AGA, appropriate for gestational age; LGA, large for gestational age. *Kruskal–Wallis test, \( P = .002 \).](image)

### Table 1. Demographic, Clinic and Laboratory Characteristics of the Subjects

|                  | SGA (n = 17) | AGA (n = 49) | LGA (n = 18) | \( P \) |
|------------------|-------------|-------------|-------------|--------|
| Gestational age (week) | 38 (37-39) | 38 (38-39) | 38 (37-38) | .328   |
| Gender           |             |             |             |        |
| Female (%)       | 10 (58.8)   | 22 (44.9)   | 7 (38.9)    | .471   |
| Male (%)         | 7 (41.2)    | 27 (55.1)   | 11 (61.1)   |        |
| Maternal age (year) | 31.6 ± 5.6 | 32.8 ± 4.4  | 31.9 ± 4.5  | .574   |
| Maternal weight at birth (kg) | 65.0 (60.8–76.0) | 79.5 (72.3–84.8) | 102.5 (92.0–112.0) | \(<.001^\ast\) |
| Maternal weight gain during pregnancy (kg) | 9.0 (7.0–12.8) | 13.0 (10.0–15.8) | 15.0 (10.3–18.8) | \(0.026^\ast\) |
| Glucose (mg/dL)  | 51.5 ± 11.1 | 59.7 ± 16.8 | 62.2 ± 15.7 | .101   |
| Insulin (µIU/mL) | 3.3 (2.7–3.8) | 4.7 (3.6–7.3) | 7.5 (1.8–16.6) | \(0.01^\ast\) |
| HOMA-IR          | 0.4(0.3–0.6) | 0.8 (0.5–1.3) | 1.4 (0.5–2.2) | \(0.01^\ast\) |
| Leptin (pg/mL)   | 21.5 (107.9–662.8) | 569.5 (202.2–911.3) | 1203.8 (699.5–1372.5) | \(0.002^\ast\) |
| WISP1 (pg/mL)    | 171.3 (129.6–230.8) | 152.2 (129.6–185.3) | 156.8 (132.3–189.0) | .169   |
| Glypican-1 (pg/mL) | 267.9 (267.2–299.2) | 267.0 (266.5–280.0) | 267.1 (266.8–296.4) | .099   |
| Glypican-3 (ng/mL) | 4.3 (2.0–6.5) | 2.7 (2.0–4.9) | 4.4 (1.9–8.8) | .283   |
| Syndecan-1 (ng/mL) | 7.2 ± 2.1 | 6.4 ± 2.6 | 6.7 ± 2.0 | .492   |

Bold values are statistically significant values and correct.

SGA, small for gestational age; AGA, appropriate for gestational age; LGA, large for gestational age; HOMA-IR, Homeostatic Model Assessment for Insulin Resistance; WISP1, WNT1-inducible-signaling pathway protein 1.

*Normally distributed data were given as mean ± SDS (one-way ANOVA test), non-normally distributed data were given as median [25p–75p] (Kruskal–Wallis test).

*Categorical variables were expressed as n (%) (chi-squared test).

*SGA versus AGA, AGA versus LGA, SGA versus LG; *SGA versus AGA, *AGA versus LGA, SGA versus LG.
DISCUSSION

This study evaluated umbilical cord blood levels of GPC1, GPC3, SDC1, and WISP1 in infants according to the birth weight (SGA, AGA, and LGA). The evaluation of both classical adipokine (leptin) and novel adipokines and proteoglycans may provide clues for intrauterine growth. Although we know that this study cannot establish causality between the adipokine levels and long-term metabolic risk due to its cross-sectional structure, it does not change the fact that we need novel markers to predict both intrauterine growth and long-term metabolic risk in that population.

Proteoglycans are structural molecules that affect the placenta’s cellular functions and the angiogenesis process. SDC1, GPC1, and GPC3 proteoglycans which are expressed in syncytiotrophoblast cells are essential members of the placental barrier. SDC1 may have an important role in modifying growth factor interaction and angiogenesis in the placenta. A few studies investigating the relation between maternal serum SDC1 levels and pregnancy outcomes reported similar plasma soluble SDC1 levels in those of infants of mothers with non-GDM, this difference was not statistically significant. Also, serum WISP1 levels were not different between SGA, AGA, and LGA infants.

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Table 2. Comparison of Demographic, Clinic and Laboratory Characteristics of the Infants of GDM Mothers With Infants of Non-diabetic Mothers

|                          | Infants of GDM Mothers (n = 30) | Infants of Non-diabetic Mothers (n = 54) | P* |
|--------------------------|---------------------------------|--------------------------------------|----|
| Gestational age (week)   | 38 (37-38)                      | 38 (37-39)                           | .238 |
| Gender                   |                                 |                                      |    |
| Female (%)               | 14 (46.7)                       | 25 (46.3)                            | .974 |
| Male (%)                 | 16 (53.3)                       | 29 (53.7)                            |    |
| Birth weight (g)         | 3526.6 ± 501.2                  | 3158.5 ± 621.0                       | .007 |
| Birth weight SDS         | 0.7 ± 1.1                       | −0.1 ± 1.2                           | .003 |
| Birth weight status      |                                 |                                      |    |
| SGA                      | 3 (10)                          | 14 (26)                              | .022* |
| AGA                      | 16 (53)                         | 33 (61)                              |    |
| LGA                      | 11 (37)                         | 7 (13)                               |    |
| Maternal age (year)      | 32.4 ± 4.1                      | 32.4 ± 4.9                           | .956 |
| Maternal weight at birth (kg) | 92.0 (83.5-103.5)               | 78.0 (70.0-90.5)                     | .001 |
| Maternal weight gain during pregnancy (kg) | 12.0 (9.0-17.3)               | 13.0 (10.0-15.0))                    | .913 |
| Glucose (mg/dL)          | 60.3 ± 14.4                     | 57.6 ± 16.6                          | .452 |
| Insulin (µIU/mL)         | 6.6 (3.1-10.7)                  | 4.7 (3.3-7.6)                        | .221 |
| HOMA-IR                  | 0.9 (0.3-1.7)                   | 0.6 (0.4-1.2)                        | .223 |
| Leptin (pg/mL)           | 848.6 (479.7-1240.5)            | 597.3 (190.9-895.9)                  | .033 |
| WISPI (pg/mL)            | 179.1 (144.3-197.4)             | 154.4 (126.8-192.8)                  | .139 |
| Glypican-1 (pg/mL)       | 270.1 (266.9-304.5)             | 267.1 (266.4-282.2)                  | .031 |
| Glypican-3 (ng/mL)       | 2.6 (2.0-4.7)                   | 2.9 (1.9-5.3)                        | .688 |
| Syndecan-1 (ng/mL)       | 6.7 ± 2.3                       | 6.5 ± 2.4                            | .815 |

Bold values are statistical significant values and correct.

GDM, gestational diabetes mellitus; HOMA-IR, Homeostatic Model Assessment for Insulin Resistance; WISPI, WNT1-inducible-signaling pathway protein 1. *Normally distributed data were given as mean ± SD (one-way ANOVA test); non-normally distributed data were given as median [25p-75p] (Kruskal–Wallis test); *Categorical variables were expressed as n (%) (chi-squared test).

The significance was lost after the post hoc analysis.
preeclamptic and normotensive pregnant women. On the contrary, some studies reported lower serum SDC1 levels in preeclamptic pregnant women. To the best of our knowledge, there is no study investigating the relation between umbilical cord blood SDC1 levels and birth weight of infants in mothers with GDM and non-GDM. A study investigating the role of endothelial glycocalyx constituents in predicting GDM reported that SDC1 have no role in prediction of GDM. In the present study, we find no difference between umbilical cord blood levels of GPC1 and GPC3 in SGA, AGA, and LGA infants, and no correlation between the levels of these proteoglycans and birthweight. To the best of our knowledge, there is no study investigating serum levels of GPC1 in patients with diabetes mellitus. In the present study, serum levels of GPC1 were higher in infants of mothers with GDM compared to infants of mothers with non-GDM.

There are some limitations in our study. The sample size was relatively small, and the design of the study was cross-sectional. Since there was no correlation between serum levels of
GPC1, GPC3, WISP1 and birth weight, we thought that the small sample size did not affect the results of the study.

In conclusion, umbilical cord blood levels of GPC1, GPC3, SDC1, and WISP1 were not different between SGA, AGA, and LGA infants. The significance of serum levels of these adipokines and proteoglycans, which are thought to have an important role on fetal growth, remains to be elucidated.

**Ethical Committee Approval:** Ethical approval was received from the Aydın Adnan Menderes University, Faculty of Medicine (2019/83).

**Informed Consent:** Informed consent was obtained from the parents of each infant.

**Peer Review:** Externally peer-reviewed.

**Author Contributions:** Concept – Ah.A., Ay.A.; Design – Ah.A., Ay.A.; Supervision – Ah.A., Ay.A., A.B.A., A.T., E.Z.; Resource – Ay.A.; Materials – Ay.A., A.T., Ö.C., E.Z., A.B.A.; Data Collection and/or Processing – Ay.A., A.T., Ö.C., E.Z., A.B.A.; Analysis and/or Interpretation – Ay.A., S.O., A.T., Ö.C., M.K.T., Ah.A.; Literature Search – Ah.A., Ay.A., Ö.C.; Writing – Ah.A., Ay.A., Ö.C.; Critical Reviews – Ah.A., M.K.T., Ö.C., S.O., A.B.A.

**Conflict of Interest:** The authors have no conflicts of interest to declare.

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