Sea stars of the genus *Henricia* Gray, 1840 (Echinodermata, Asteroidea) from Vostok Bay, Sea of Japan

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**ABSTRACT**

We report seven species of the genus *Henricia* Gray, 1840 that were found in Vostok Bay, and two species from adjacent area, known from museum collection or seen in underwater footage. While existing literature reported no confirmed species from this area. Most of these species: *H. djakonovi*, *H. alexeyi*, *H. densispina*, *H. hayashii*, *H. granulifera*, *H. pacifica*, *H. asiatica*, and *H. oculata robusta* were reported from the Sea of Japan previously. *H. nipponica*, known from Japan, is reported from Russian seas for the first time. All studied taxa are re-described here using a range of morphological characters and partial 16S rRNA nucleotide sequences, life colorations of several species are reported for the first time, and an identification key is provided. Lectotype designations are fixed for studied series of species described by AM Djakonov.

**Subjects** Biodiversity, Marine Biology, Zoology

**Keywords** Echinasteridae, Local fauna, Western Pacific

**INTRODUCTION**

Sea stars of the genus *Henricia* Gray, 1840, belonging to the family Echinasteridae (Asteroidea, Spinulosida), have been a challenge for systematists to sort out because of their high species diversity, similar appearance, confusing taxonomic history, and often broad geographic distribution, especially in the northern Pacific (*Verrill, 1909; Verrill, 1914; Fisher, 1911; Fisher, 1928; Fisher, 1930; Hayashi, 1940; Shin & Rho, 1996; Djakonov, 1961; Jewett et al., 2015*). *Smirnov (2013)* has reported 30 nominal species in Russian Pacific seas belonging to either *Henricia* or the allied genus, *Aleutihenricia Clark & Jewett, 2010*. This number is based mainly on Djakonov’s publications (1948; 1950; 1958; 1961), Hayashi (1940), and the work by *Clark & Jewett (2010)*. Hence, species abundance in Russian Pacific is still in need of study.

Comprehensive accounts of the Vostok Bay marine biota began following the construction of the Vostok Marine Biological Station in 1970. The station was established by the Soviet Academy of Sciences and was a significant research site used by many Russian and USSR biologists. The echinasterids of this region have been overlooked. In recent literature, the reviews on echinoderm fauna of Vostok Bay (*Dautov & Tyurin,
2014) and of adjacent Far Eastern National Marine Reserve (Lebedev, 2015), this family was represented by “Henricia sp.” only. Earlier literature review of Peter the Great Bay biota (with relevant section based on Djakonov (1961) reported seven Henricia species (Adrianov & Kussakin, 1998). Thereby, the fauna of Henricia of Vostok Bay remains totally unknown except for a single Henricia sp. of 21 species potentially inhabiting the Sea of Japan (Smirnov, 2013). Recent studies reported detailed morphology of 13 species from the Sea of Japan (Shin & Ubagan, 2015a; Shin & Ubagan, 2015b; Ubagan & Shin, 2016; Chichvarkhin, 2017a; Chichvarkhin, 2017b; Chichvarkhin & Chichvarkhina, 2017a; Chichvarkhin & Chichvarkhina, 2017b) and adjacent waters of Yellow Sea (Xiao, Liao & Liu, 2011).

Many sea stars including the species of the genus Henricia are a popular objects in biochemical and biomedical research because of perspective drugs found in these animals (e.g., Fedorov et al., 2008; Utkina, 2009). Taxonomic confusions have prevented confident identifications for Henricia, which has limited potential applications for this research field. Recent studies of coastal marine benthic fauna of Russian Pacific shores revealed numerous previously overlooked species belonging to different taxonomic groups (e.g., Chichvarkhin et al., 2016; Ekimova et al., 2016; Temereva & Chichvarkhin, 2017), hence a discovery of new species in a poorly studied and confused group like the Echinasteridae in Vostok Bay remains possible. Therefore, this study aims to shed light to Henricia fauna occurring in Vostok Bay by delineating putative species by the means of DNA barcoding, then finding corresponding morphological characters that are diagnostic for identification.

**MATERIAL AND METHODS**

Observations and sample collections were made by SCUBA-diving in 2014 through 2017 in Vostok Bay of the Sea of Japan (Fig. 1). Most of the individuals belonging to abundant species were returned and released where each was collected. The images were taken with a Nikon D810 or D7000 cameras and a Nikkor 60/2.8 lens. Also, remotely controlled submersible apparatus Obzor-150 was used in 2015 at the depth of 45 m at the silty outer part of the bay. The specimens were preserved in 96% ethanol or dry with prominent portion of a ray preserved in separate vials in 96% ethanol and deposited in the Museum of National Science Center of Marine Biology, Russian Academy of Sciences (Vladivostok, Russia). Skeletal plates and spines were denuded using 5–15% sodium hypochlorite solution at room temperature. Scanning electron images of the spines were obtained using Zeiss Sigma and Zeiss Evo electron microscopes after carbon coating. Studied specimens are preserved in the collections of Zoological Institute of Russian Academy of Sciences, St. Petersburg, Russia (voucher code ZIN), National Science Center of Marine Biology, Russian Academy of Sciences (voucher code MIMB) and Hokkaido University, Sapporo, Japan (voucher code ZIHU). Sequenced specimens are listed in the Table 1.

DNA was extracted using the Diatom™ DNA Prep 100 kit (Isogene Lab, Moscow, Russia) according to the manufacturer’s protocol. Partial sequence for mitochondrial 16S rRNA gene (16S) was used in this study. This fragment was used as a good species-specific marker in several Henricia and echinoderm studies (Chichvarkhin & Chichvarkhina, 2017b).
The primers 16Sar and 16Sbr (Palumbi, 1996) were used to amplify the region of interest. Polymerase chain reaction amplifications were carried out in a 20-µL reaction volume, which included 4 µL of 5× Screen Mix by Eurogen Lab. We also attempted unsuccessfully to amplify the portion of the mitochondrial gene for cytochrome c oxidase subunit I (COI) that is commonly used in metazoan DNA barcoding. The primers normally employed by Folmer et al. (1994) do not work well for echinasterids. We also attempted unsuccessfully to amplify the portion of the mitochondrial gene for cytochrome c oxidase subunit I (COI) that is commonly used in metazoan DNA barcoding. The master mix for each sample was prepared using 34.75 µL H2O, 5.00 µL PCR Buffer (Evrogen, Moscow, Russia), 5.00 µL 25 mM MgCl₂, 1.00 µL 40 mM dNTPs, 1.00 µL 10 mM primer 1, 1.00 µL primer 2, 0.25 µL 5 mg/ml (10×) Taq (Evrogen, Moscow, Russia), and 1.00 µL extracted DNA. Amplification began with an initial denaturation for 1 min at 95 °C followed by 40 cycles of 15 s at 95 °C, 15 s at 52 °C, 30 s at 72 °C with a final extension of 7 min at 72 °C. Sequencing for both strands proceeded with the Big Dye v3.1 sequencing kit by Applied Biosystems. Sequencing reactions were analyzed using ABI 3130 and 3500 Genetic Analyzer (Applied Biosystems, Foster City, CA, USA) in National Scientific Center of Marine Biology (Vladivostok, Russia). The sequences were aligned with ClustalW software (Larkin et al., 2007). Tree-based methods for species delineation and identification were used, including the calculation of pairwise p-distances (i.e., the proportion of variable positions) and neighbor-joining (NJ) clustering using MEGA7 software (Kumar, Stecher & Tamura, 2016).
| Species                        | Voucher #      | GenBank #, 16S |
|-------------------------------|----------------|---------------|
| *Henricia alexeyi*, paratype  | MIMB-33244     | KY464043*a    |
| *Henricia alexeyi*            | MIMB-34803     | KY464042*a    |
| *H. alexyi*                   | MIMB-33243     | MG976220*a    |
| *H. pacifica*                 | MIMB-33247     | KY464039*a    |
| *H. pacifica*                 | MIMB-33246     | KY464040*a    |
| *H. pacifica*                 | MIMB-33248     | KY464041*a    |
| *H. ohshimai*                 | Japan          | DQ297093      |
| *H. ohshimai*                 | Japan          | AB084571      |
| *H. densispina*               | MIMB-34800     | MG976223*a    |
| *H. densispina*               | MIMB-34801     | MG976224*a    |
| *H. densispina*               | MIMB-34801     | MG976225*a    |
| *H. djakonovi*                | MIMB-33878     | KY464037*a    |
| *H. djakonovi*                | MIMB-33128     | MG976221      |
| *H. djakonovi*                | MIMB-33831     | MG976222      |
| *H. djakonovi*, holotype      | MIMB-33129     | KY464038      |
| *H. hayashii*                 | MIMB-33544     | KY744471      |
| *H. hayashii*                 | MIMB-33250     | KY744470      |
| *H. granulifera*              | MIMB-33251     | KY744469      |
| *H. oculata*                  | Atlantic Ocean, UK | AY652500    |
| *H. oculata*                  | Commander Is.  | KX986916*a    |
| *H. oculata*                  | Commander Is.  | KX986917*a    |
| *H. nipponica*                | MIMB-33252     | KX610476*a    |
| *H. nipponica*                | MIMB-33253     | KX610479*a    |
| *H. nipponica*                | Vostok Bay     | KX610478*a    |
| *H. nipponica*                | Vostok Bay     | KX610477*a    |
| *H. nipponica*                | Japan          | D63737        |
| *H. cf. nipponica*            | Japan          | D63739        |
| *H. tumida*                   | NE Pacific     | AF290033      |
| *H. sanguinolenta*            | Norway         | KT268115      |
| *H. obesa*                    | Australia      | KT268113      |
| *H. compacta*                 | Australia      | KT268112      |
| *H. lineata*                  | Avacha Bay     | MF133325      |
| *H. uluudax*                  | Avacha Bay     | KY934075      |
| *H. olga*, holotype           | MIMB-33539     | KY934079      |
| *H. sp.*                      | -              | HM473897      |
| *H. sp.*                      | -              | HM473896      |
| *H. sp.*                      | -              | HM473895      |
| *H. sp.*                      |                | HM543156      |
| *H. sp.*                      |                | DQ297093      |

**Notes.**
- *Data obtained in this study.*
The primers normally employed by Folmer et al. (1994) do not work well for echinasterids. Recent works (Laakmann et al., 2016; Layton, Corstorphine & Hebert, 2016) have demonstrated that the COI can distinguish some entities in the Echinasteridae. Knott, Ringvold & Blicher (2017) used Henricia-specific primers more successfully: they delimit almost all north Atlantic species, although we failed to amplify any product using these primers. Often, the COI can be poorly or not amplified with ‘Folmer’s’ primers (Folmer et al., 1994) commonly used in metazoan DNA barcoding, hence we are considering the 16S is a more robust and preferable marker for routine DNA barcoding of Asteroidea (Chichvarkhin, 2017b; Chichvarkhin, 2017b; Knott, Ringvold & Blicher, 2017). Recently, Knott, Ringvold & Blicher (2017) have developed Henricia-specific primers to amplify the COI. Amplified fragment was same informative as the 16S, hence we see no reason to use it as a marker in our study.

In his descriptions of Henricia species, Alexander M. Djakonov (Djakonov, 1948; Djakonov, 1958) never designated type materials; the specimens that we identified in ZIN collection by the labels written by him and his co-worker Zoya I. Baranova, and collection dates preceding a publication. Certain of these syntypes series contain or may contain specimens belonging to different species, so we have fixed a particular syntype as the lectotype in such cases, with all other specimens in each series becoming paralectotypes.

For the taxa below, the author (Djakonov, 1958; Djakonov, 1961) did not designate unique specimens as the holotypes according to International Code of Zoological Nomenclature (ICZN) Article 73.1.3, i.e., the series of the syntypes were used for description. For all these species, we identified the specimens marked as the “holotypes” in ZIN collection. But this method for a holotype fixation applied by Z.I. Baranova is incorrect according to ICZN Article 72.4.7, since these specimens unequivocally belong to the type series according to ICZN Article 72.4.1.1. To avoid confusion, here we are fixing unique specimens as the lectotypes of several of Djakonov’s taxa based on the specimens used in this study. Appropriate lectotypes designations are given in species sections below. We disregarded ”holotype” labels later added to particular syntype specimens in the ZIN collection in the hand-writing of Djakonov’s assistant, Z.I. Baranova, because such holotype designations are invalid according to the International Code of Zoological Nomenclature (ICZN) articles 72.4.1.1, 72.4.7, and 73.1.3.

RESULTS

Molecular analysis

We used 16S sequences from this study, or added from GenBank, were partial 16S rRNA nucleotide sequences ranging from 541–606 bp long. We produced an alignment of 632 bp including gaps; 162 sites within the ingroup were variable, and 129 of these were parsimony-informative. Neighbor-Joining tree (Fig. 2) shows well supported clusters for all were partial 16S rRNA nucleotide sequences ranging from 541–606 bp long. We produced an alignment of 632 bp. Uncorrected interspecific distances ranged from 5% (H. lineata –H. uluudax) to 11.5% (H. ohshimai –H. compacta).
Figure 2  NJ tree (p-distance) based on 16S rRNA partial nucleotide sequences. Bootstrap 1,000 replicates. The taxa marked with red were collected in Vostok Bay.

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Order Spinulosida *Perrier, 1884*
Family Echinasteridae *Verrill, 1870*
Genus *Henricia Gray, 1840*

*Spinohenricia Heding, 1935*
*Magdalenaster Koehler, 1907*
*Cyllaster A.H. Clark, 1916*

Subgenus *Henricia Gray, 1840*

Type species: *Asterias oculata* Pennant, 1777

1. *Henricia (H.) alexeyi* Chichvarkhin & Chichvarkhina, 2017a; Chichvarkhin & Chichvarkhina, 2017b (Fig. 3)

*Henricia alexeyi* Chichvarkhin & Chichvarkhina, 2017a: 203–207, figs. 1–3, 4G; Chichvarkhin & Chichvarkhina, 2017b: 25–27, fig B, C.

Type locality: Rudnaya Bay, Sea of Japan, Russia.

Examined material: holotype: MIMB–33243, Senkina Shapka pinnacle, Rudnaya Bay, 10–15 m depth, 8 October 2015, leg. A. Chichvarkhin. Paratypes: MIMB–33244, 1 spm, Senkina Shapka pinnacle, Rudnaya Bay, 10–15 m depth, 8 October 2015, leg. A. Chichvarkhin; MIMB–34803, 1 spm, Vostok Bay, 07 Jul 2017, leg. K. Dudka; MIMB–33241, 1 spm, Vostok Bay, Sea of Japan, Russia, Aug 2015, leg. K. Dudka; MIMB–33242, 1 spm, Vtoraya Priboika Inlet, Vostok Bay, Sea of Japan, Russia, 2 m depth, 25 Aug 2016, leg. I. Ekimova. ZIHU–2405, 3 syntypes of *H. ohshimai* (Hayashi, 1940), Onagawa shore, 1934.

External anatomy. Small size sea stars, R (long radius, mouth to ray tip) = 19–47 mm, r (short radius, mouth to interradius) = 4–9 mm, mean R: r = 4.7 (R:r of holotype 4.56, R = 20.5, r = 4.5). Rays five, cylindrical, slim, not swollen at base (Fig. 3). Abactinal skeleton open meshed. Abactinal plates oval, rod-shaped with low ridges or tubercles. Papular areas with 1–3 papulae. Abactinal pseudopaxillae (Fig. 3B) with 6–10 spines in two rows covered by thin skin. Abactinal spines short, stout, 300 µm long, 80–100 µm wide, most tapering distally (Fig. 3), bear 1–6 short thick subapical thorns faced upwards.

Superomarginal row well discernible consisting of plates larger than abactinal, with rather high ridges (Figs. 3A, 3C), consists of longitudinally elongated plates. Intermarginal row extended to 2/3 of ray length, second intermarginal row short, consists of eight plates. Intermarginal plates small, round. Inferomarginals plates triangular or sub-quadrate. Superomarginals bear 7–9 spines in two rows. Intermarginal pseudopaxillae with 4–6 spines in two rows. Inferomarginal pseudopaxillae bear 10–12 spines in 2 transversal rows.

Ventrolateral row consists of densely set oval plates. Second ventrolateral row extends to a half of ray length, consists of tiny round plates. Ventrolateral pseudopaxillae bear 3–5 spines in one row. Adambulacrals with 3–4 larger near-furrow spines and three small spines in one
irregular row (Fig. 3C, 3D) joined by skin membrane as a crest. Proximally, adambulacral, ventrolateral and inferomarginal spines arranged in crests with skin membrane (Fig. 3D). Deep furrow spine small, single. Oral plates each bear single apical, three marginal, and two suboral spines.
Color in life varies greatly from red to orange. Some specimens with broad darker areas on rays or almost white on disc and proximally on rays (Fig. 3). Anal area darker or same color as background; madreporite lighter or same color as background. Actinal side uniformly orange.

Ecology: inhabits open bays exposed to wave activity at 2–15 m depths. In Rudnaya Bay, specimens were found on sponges apparently damaged by these sea stars (Fig. 3F).

2. *Henricia (H.) pacifica* Hayashi, 1940 (Fig. 4)

*Henricia pacifica* Hayashi, 1940: 152–154, Pl. 9 figs. 7–10; *Hayashi, 1952*: 114; *Djakonov, 1961*: 16–17. (part.); *Chichvarkhin & Chichvarkhina, 2017a*: fig. 4H; *Chichvarkhin & Chichvarkhina, 2017b*: 27, fig D.

*Henricia sanguinolenta*—*Yavnov, 2010*: 125–126 (non *Müller, 1776*).

Type locality: Tsugaru Strait, Japan.

Examined material: MIMB–33245, (R = 46 mm) Vostok Bay, 10–15 m depth, Aug 2015, leg. I. Krasilnikov. 3 spms, MIMB–33246–33248, (R = 38, 48, 51 mm), Vostok Bay, 10–15 m depth, Aug 2015, leg. I. Krasilnikov; ZIHU–2404, *H. pacifica* holotype, Tsugaru Strait, Japan.

External anatomy. Medium size sea stars, mean R: r = 4.1 (R:r of holotype 4.2); R = 48–53 mm, r = 11–14 mm, R of holotype 50 mm, r = 12 mm. Rays five, slender, slim, cylindrical (Figs. 4A–4D). Abactinal skeleton open meshed. Abactinal plates oval, rod-shaped with well discernible smooth ridges (Fig. 4G). Abactinal spines set along ridges in groups of 4–10 spines in one row covered by skin membrane, so in dried specimens the spines become poorly distinguishable. Papular areas with 1–3 papulae. Abactinal spines long, tapering, 400 µm long, 100 wide at base, tapering to 30–50 µm distally (Figs. 4H–4K), bear 9–11 tiny thorns on apex and spine shaft and faced upwards or at angle to spine axis.

Superomarginal row discernible consisting of cross-shaped plates similar in size to neighboring plates with no ridges (Figs. 4E–4F). Intermarginal row extended to 2/3 of ray length. Intermarginal plates rod- and crescent-shaped. Inferomarginals plates sub-quadratce. Superomarginals bear 4–5 spines in one row. Intermarginal pseudopaxillae with 4–5 spines in two rows. Inferomarginal pseudopaxillae bear five spines in one transversal rows.

Ventrolateral plates sub-quadratce, smaller than inferomarginals. Pseudopaxillae bear 4–5 spines in one row. Adambulacrals with three larger thick near-furrow spines and four smaller spines in one row. Deep furrow spine thick, single. Oral plates each bear single apical, three large marginal, and three suboral spines.

Color, in life, bloody-red with light whitish tint proximally on rays and on disc, also on ray tips in some specimens (Figs. 4A–4D). Anal area not discernible from background; madreporite yellow. Actinal side uniformly orange.

Ecology: inhabits open bays exposed to wave activity at 15 m depths.
Figure 4  *Henricia pacifica* (Hayashi, 1940). (A) holotype life coloration; (B–D) paratypes; (E, F) actinal skeleton; (G) abactinal skeleton; (H–K) abactinal spines. i, inferomarginal; im, intermarginal; s, superomarginal; v, ventrolateral; v2, additional ventrolateral. Scale bars: A–D - 10 mm, E–G - 5 mm, H–K - 50 µm. Photos by A Chichvarkhin.
3. *Henricia (H.) nipponica* Uchida, 1928 (Fig. 5)

*Henricia leviuscula* var. *nipponica* Uchida, 1928: 794, pl. 32, figs. 6, 7.

*Henricia nipponica* Hayashi, 1940: 146–149, fig. 32–35.

*Henricia pachyderma*, Chichvarkhin & Chichvarkhina, 2017a: fig 4I; Chichvarkhin & Chichvarkhina, 2017b: 27, fig. E. (non Hayashi, 1940)

**Type locality:** Japan.

**Examined material:** 6 spms, MIMB–33252–33257, Vostok Bay, July 2015, leg. I. Krasilnikov; 1 spm, MIMB–33814, 4 Oct 2015, A. Chichvarkhin; 5 spms, Vostok Bay, 27 Aug 2016, leg. A. Goloseyev; 1 spm, Vostok Bay, 10 Jul 2017, leg. K. Dudka; 4 spms, Rudnaya Bay, 10 Oct 2015, leg. A. Chichvarkhin; 1 spm Sagami Bay, Misaki Marine Biological Station, Kanagawa, Japan, 3 Jun 2018, leg. A. Chichvarkhin.

**External anatomy.** R: \( r = 4.4–5.7, R = 35–50 \) mm. Rays five, abnormally six, cylindrical (Fig. 5A). Aboral skeleton wide meshed. Most abactinal plates rod-shaped, some branched with well discernible tubercle. Secondary plates of irregular shape in some meshes (Figs. 5B, 5D). Abactinal pseudopaxillae with 4–8 spines in one row. Papular areas with 1–3 papulae (most often three); papulae in live individuals large, 4–5 fold longer than wide (Fig. 5E). Abactinal spines long, 400 \( \mu \)m long, 60–100 \( \mu \)m wide at base, gradually tapering proximally (Figs. 5F–5I), with 8–12 tiny apical and subapical thorns, covered by skin membrane.

Superomarginal row poorly discernible, consists of plates similar to adjacent aborals, bearing 3–5 spines, same as abactinal spines, arranged in one row. Interomarginal row extended to a half of ray length; plates with 4–5 spines in one row. Plates of intermarginal and adjacent marginal rows connected by additional transversal plates. Inferomarginal plates large, transversally elongated, pseudopaxillae bear 8–10 spines in one transversal row. Ventrolateral plates pillow-shaped, with proximal outgrowth overlaying next proximal plate; pseudopaxillae bear 4–7 spines in one transversal row. Second ventrolateral row consists of smaller round plates bearing 2–3 spines; this row extends to a half of ray length. Adambulacrals with 3–4 larger near-furrow spines and 6–8 smaller spines in two irregular rows (Fig. 5C). Deep furrow spine small, single. Oral plates not fused, each bear single apical, four marginal, and four suboral blunt spines.

Color in life. Abactinal side dirty-orange to brown, actinal side orange, lighter than abactinal side (Fig. 5A). Anal area darker than background; madreporite light creamy.

Ecology: The species was found on rocky substrates at the depths of 1–20 m at water temperature of 2–22 °C.

**Distribution.** Abundant in the northern part of Japan (*Hayashi, 1940*). Reported here from Russian waters for the first time.
4. *Henricia (H.) oculata robusta* Djakonov, 1958

*Henricia oculata*: Jewett et al., 2015: 76, 77.

*Henricia aspera*—Djakonov, 1948: 30; 1950: 99–100; 1961: 24–25 (non Fisher, 1911).

*Henricia aspera* robusta: Djakonov, 1948: 99, fig. 193; 1950: 99; 1958: 310–311; 1961: 25;

Yavnov, 2010: 177, 179 (part.); ? Xiao, Liao, Liu, 2010: 5, figs. 4A–F, 12G.
Type locality: southern Kurile Islands, Pacific shore.
Examined material: 1 spm (*H. oculata* neotype) NHMD-71387 (AMUC-AST-223), Treadur Bay, Ravensport, 15 May 1968, leg. B. Brunn; 2 spms, NHMD-204701, Bay Fire, Isle of Man, Irish Sea, 24 Apr 1968, leg. B. Brunn; 2 spms, NHMD-76378 (ZMUC-AST-214), Plymouth, ca. 50 m, Engelska Kanal, Sep 1931, leg. Th. Horbensen; 1 spm (NHMD), Foró, >1,867 m, 8 Apr 1984, leg. Rostrup; *ZIN–15/15340*, 2 spms, Korea-Sakhalin Expedition st. 2, 1 Mar 1900, Peter the Great Bay, Bosphorus Eastern Str., 18 m, silt; *ZIN–8/15043*, 1 spm, Rossinant st. 1555, 3 Sep 1930, 83–86 m, Vladimir Bay, leg. Derjavin; *ZIN–26/15052*, 2 spms, Toporok st. 2, 16 Aug 1948, Musasi Bank, Sea of Japan, 93–96 m, leg. Ushakov; *ZIN–10/15156*, 2 spms, Nagasaki, 1901, leg. P.Yu. Schmidt; *ZIN–18/15343*, 2 spms, Nagasaki Apr 3, 1897, D-r Bunge; *ZIN–17/15342*, 2 spms, Nagasaki, Dec 12–14 1897, D-r Bunge; *ZIN–16/15341*, 1 spm, 23 Jul 1898, Nagasaki, D-r Bunge; *ZIN–14/15339*, 1 spm, 10—15 Dec 1897, Nagasaki, D-r Bunge.

Diagnosis. Aboral pseudopaxillae comprised of up to 10–20 spines in double row. Spines covered with thick skin membrane. Spines stout, crowned with 15–20 tiny thorns, lateral ribs poorly developed. Abactinal plates arched, suboval, with no ribs, arranged in relatively wide mesh with 1–3 papulae a mesh. Superomarginals larger than aboral plates, T-shaped, with central proximal outgrowth. Inferomarginals large, cross-shaped, elongated transversally. Adambulacral spines 5–6, largest one near furrow spatulate or slightly flattened.

External anatomy: R:r 3.7–4.1 R 45–57 mm, r 11–15 mm. The rays slender not very slim, slightly swollen at base. Whole body covered by thick skin membrane, which looks rather thin in dried specimens. Abactinal plates suboval, rod-shaped, arched, cover adjacent proximal plates (Fig. 1A). Abactinal pseudopaxillae (Fig. 1B) crowned with 10–20 spines arranged in two rows. Papular areas nearly same width/length as average abactinal pseudopaxilla, includes one to three papulae. Abactinal spines covered with thin skin membrane, vary in length, stout, 350–400 µm long, 100 µm wide at base; apex with 15–20 short thorns, lateral ribs poorly developed (Fig. 2); in some individuals, spines short and widened distally, in the others, spines slender and tapering at distal end (Figs. 2A–2C).

Superomarginal row bent dorsally at ray base (Fig. 1C), consists of T-shaped plates with central dorsal outgrowth overlapping adjacent abactinal plates; bear 8–10 spines arranged in 2 rows. In large specimen, two complete or qincomplete intermarginal rows consisting of rod-shaped plates that connected to adjacent marginal rows with similar rod-shaped transversal plates crowned with 6–8 spines in two rows; one of intermarginal row extended to or beyond half ray. In this way, supero- and inferomarginal rows in some specimens (e.g., *MIMB–33214*, Fig. 1C) are widely separated with two intermarginal rows and two series of additional transversal plates. But in most specimens (Fig. 3D) intermarginal rows and connecting plates are less developed. Inferomarginal row extended over entire ray length, its pseudopaxillae bear 14–20 spines in two transversal rows. Ventrolateral quadratic plates bear 10–13 relatively small spines in 2–3 rows. Second ventrolateral row consists of about 18 transversally elongated rod-shaped plates with low tubercle. Arched adambulacrals with slightly curved small furrow spine, one large slightly flattened to spatulate spine (Figs. 1C, 1D) faced into furrow (but in some individuals, more than one spine is spatulate,
Fig. 3F); the other 3–5 gradually smaller spines faced ventrally in one staggered row bearing numerous thorns on apices. Spatulate shape varies because it is constituted by soft tissue, not spine itself. Oral plates not fused, bear 4 marginal and 1–4 suboral spines.

Life coloration varies. Rays orange-red, pink, yellow, some individuals with purple to violet stripes (Figs. 2E, 2F). Madreporite slightly lighter than background.

Ecology. Living sea stars were found off Medny Island on rock at depths 5–10 m at water temperature of 4 °C. Probably, they fed on yellow colored sponge, which was detected nearby (Figs. 2E, 2F). In other areas, found at depths >100 m feeding on sponges, detritus, hydrozoans, and bryozoans (Jewett et al., 2015); brooding (Jewett et al., 2015).

Distribution. Amphiboreal species. NE Atlantic: British Islands, Irish Sea, English Channel, North Sea, Faroe Islands, France, Portugal, NW Atlantic: New Brunswick, northern Pacific: British Columbia, Alaska, Aleutian Islands, Commander Islands, Kurile Islands, Sea of Okhotsk, Sea of Japan, Pacific shore of Japan (Hayashi, 1940; Djakonov, 1961; Madsen, 1987; Clark & Jewett, 2010; Jewett et al., 2015). Unknown from Japan: the only specimen found in R. Hayashi’s collection preserved in Hokkaido University belongs to H. aspera Fisher, 1906.

Subgenus Aleutihenricia (Clark & Jewett, 2010)

Type species Henricia rudis Verrill, 1911 (= H. derjugini (Djakonov, 1950)

5. Henricia (A.) granulifera Djakonov, 1958 (Fig. 6; Supplemental Information 1, second sea star, 5–9th seconds)

We identified the specimen in ZIN collection (Fig. 6) identified as H. granulifera and labeled as the ‘holotype’ by Z.I. Baranova. We are designating this specimen as lectotype of Henricia granulifera (Djakonov, 1958). R = 80 mm, r = 19 mm; ZIN–4/14824 transect off Moibesi [Belinskoye, W Sakhalin, Sea of Japan], 30–37 m, 30 Aug 1949.

Henricia granulifera Djakonov, 1958: 311–312, figs. 21, 22; Djakonov, 1961: 27, Pl. 2, fig. 20, Pl. 17, figs. 68, 69; (Yavnov, 2010): 121–122; (Chichvarkhin & Chichvarkhina, 2017a): fig 4K; Chichvarkhin & Chichvarkhina, 2017b: 27, fig. F.

Type locality: Sea of Japan, Tatar Strait near Belinskoye, Sakhalin.

Examined material: 1 spm, MIMB–33251, Vostok Bay, July 2015, leg. I. Krasilnikov; 1 spm. Vostok Bay, 13 July 2017, leg. K. Dudka; 1 spm, ZIN–4/14824 (lectotype of H. granulifera) transect off Moibesi [Belinskoye, W Sakhalin, Sea of Japan], 30–37 m, 30 Aug 1949, Toporok stations #65–66; ZIHU–2411 (holotype of H. reticulata), Onagawa Iigohama, Japan; ZIHU–2412 (holotype of H. irregularis), Yezo Strait, Albatross 1906 station #5031, 86 fathoms depth, Japan.

External anatomy. Relatively large sea star, R 72 mm, r = 9, R: r = 8. Rays five, abnormally four, cylindrical. Disc small, aboral skeleton wide meshed. Abactinal plates rod-shaped, with 1–2 well discernible tubercles on each, form wide mesh. Small secondary plates of irregular shape in some meshes. Abactinal pseudopaxillae in groups of 3–5 spines in paxilla. Papular areas with 2–4 papulae. Abactinal spines long, 550 μm long, 100 μm wide at base, gradually tapering proximally (Figs. 6H–6P), with about 15 tiny and subapical thorns, some thorns located laterally on spine shaft; lateral thorns faced at an angle to spine shaft; spines not covered by skin membrane.
Figure 6  *Henricia granulifera*, Vostok Bay. (A, B) life coloration of abactinal and actinal sides, Vostok Bay; (C) denuded abactinal skeleton, Vostok Bay; (D) denuded abactinal skeleton, holotype; (E) actinal side, Vostok Bay; (F) actinal side, lectotype; (G–K) abactinal spines, Vostok Bay; (L–O) abactinal spines, *H. granulifera* lectotype. i, inferomarginal; im, intermarginal; s, superomarginal; v, ventrolateral. Scale bars: A, B - 10 mm, C–F - 5 mm, G–O - 50 μm. Photos by A. Chichvarkhin.

Full-size DOI: 10.7717/peerj.6585/fig-6
Marginal rows poorly discernible, consists of similar plates, bearing 3–5 spines; plates with well discernible tubercles. Superomarginal plates higher than adjacent plates, form more discernible bent row. Intermarginal row extended to full ray length. Inferomarginal plates large but not as high as superomarginals.

Ventrolateral plates small, round, with tubercle; bear 5–6 spines in a circle. Adambulacrals with 2–3 larger near-furrow spines and 5–6 smaller spines in two rows (Figs. 6C–6D). Deep furrow spine small, single. Oral plates not fused, each bear single apical, four marginal, and three suboral blunt spines.

Color in life. Abactinal side brown with spine apices orange (Fig. 6A); marginal rows brown as aboral side; ventrolateral, adambulacral, oral and furrow areas orange (Fig. 6B). Anal area small and lighter than background; madreporite bright orange.

Ecology. A single specimen of this species was found on rocky substrate at the depth of 15 m at water temperature of 16 °C.

Distribution. Known for the northwestern part of the Sea of Japan (Chichvarkhin & Chhichvarkhina, 2017), Korea (Shin & Rho, 1996) and Japan (Hayashi, 1940).

Subgenus Setihenricia Chichvarkhin & Chichvarkhina, 2017
Type species Henricia hayashi Djakonov, 1961

6. Henricia (S.) djakonovi Chichvarkhin, 2017 (Fig. 7)

Henricia djakonovi Chichvarkhin, 2017a: e2863; Chichvarkhin & Chichvarkhina, 2017a: Fig. 4E; Chichvarkhin & Chichvarkhina, 2017b: 27, fig. G.

Henricia pseudoleviuscula—Chichvarkhin, 2017a: e2863 (non Djakonov, 1958).

Henricia hayashi—Yavnov, 2010: 113–114 (non Djakonov, 1961).

Henricia leviuscula leviuscula—Xiao, Liao & Liu, 2011: 9–11, fig. 7 (non Stimpson, 1857).

Type locality: Rudnaya Bay, Sea of Japan, Russia

We identified H. pseudoleviuscula specimen in ZIN collection labeled as the “holotype” presumably by Z.I. Baranova. We are designating this specimen as lectotype of Henricia pseudoleviuscula (Djakonov, 1958). R = 41 mm, r = 8 mm; ZIN—1/12815, Toporok station 31, Moneron Is., 14 Aug 1947, 84 m depth.

Examined material: MIMB–331294 4 Jun 2016, Senkina Shapka pinnacle, Rudnaya Bay, 44.36°N 135.83°E, 16 m, leg. A. Chichvarkhin (holotype); MIMB–333104 4 Jun 2016, Senkina Shapka pinnacle, Rudnaya Bay, 44.36°N 135.83°E, 16 m, leg. A. Chichvarkhin (paratype); 11spms, MIMB–33128, MIMB–33131, 23–25 Aug 2015, Vostok Bay, leg. I. Krasilnikov; 1 spm, MIMB—, Vostok Bay, 10 Jul 2017, leg. K. Dudka; ZIN—1/12815, Toporok station 31, Moneron Is., 14 Aug 1947, 84 m depth (lectotype of H. pseudoleviuscula); 2 spms, ZIHU—2397, Bomasiri Shima [N. off Rebun Is., Japan], 1906 (H. reniossa holotype and paratype).

Chichvarkhin & Chichvarkhina (2017a) and Chichvarkhin & Chichvarkhina (2017b) described this species as one of two ‘tiger-striped’ sympatric morphs that co-occur in the northwestern Sea of Japan, while the other form was incorrectly referred to as H. pseudoleviuscula. This study found that both morphotypes had identical sequences, so they...
Figure 7  *Henricia djakonovi*, Vostok Bay. (A, B) life coloration; (C) denuded abactinal skeleton; (D) abactinal side of ethanol-preserved specimen; (E) actinal side of ethanol-preserved specimen; (F–H) abactinal spines; (I–J) abactinal spines, holotype; (K–L) abactinal spines of *H. pseudoleviuscula*, holotype; (M) aboral spines of *H. reniossa*, holotype. Scale bars: A, B - 10 mm, C - 5 mm, D - 3 mm, E - 1 mm, F–M - 50 µm. Photos by A. Chichvarkhin.

Full-size DOI: 10.7717/peerj.6585/fig-7
more likely represent discontinuous phenotypes of the same species (Fig. 2); both forms cannot be referred as *H. pseudoleviuscula* (*Djakonov, 1961*) because the lectotype of the latter possesses very different spines (Figs. 7K, 7L), similar to those of *H. reniossa* Hayashi, 1941 (Fig. 7M). Thus, our amended description of *H. djakonovi* here is expanded to include *H. pseudoleviuscula sensu* (*Chichvarkhin, 2017a*).

External anatomy. R of holotype = 52 mm, r = 11 mm, R: r = 4.7–5; R = 38. To avoid a future confusion, here we are re-describing *H. djakonovi* by combining the characters of both forms (i.e., *djakonovi* and *pseudoleviuscula sensu* *Chichvarkhin, 2017a*). mm, r = 7.5 mm in paratype. Rays slender not very slim, slightly swollen at base, tapering to blunt tips. Most abactinal plates (>50%) cross-shaped with two proximal outgrowths covering adjacent plates, their elevated proximal sides form crescent-shaped pseudopaxillae. The other plates triangular or irregular shaped, lacking proximal outgrowths, form round and oval pseudopaxillae. Plates convex without ridges or tubercles. Abactinal surface (Figs. 7A, 7C, 7D) thick, rigid; abactinal plates on disc close-set forming tight reticulation. Abactinal pseudopaxillae bear 20–30 spines, but some smaller pseudopaxillae bear 8–10 spines. Papular areas nearly twice smaller than average abactinal pseudopaxillae, include one or two papulae. Abactinal spines short, stout, 200–240 μm long, 60–100 μm wide, barrel-shaped (Figs. 7F–7J). Apex of most spines massive with few short thorns; apex in some specimens smooth, resembles glass droplet (Fig. 7J).

Superomarginal row bent dorsally at ray base, consists of square pillow-shaped plates. Inferomarginals and ventrolaterals are similarly pillow-shaped. Intermarginal rows consisting of usually one, but up to six, irregular shaped plates, constituting a proximal intermarginal area that provides an inflation of ray base. Superomarginals bear 20–35 spines, same as abactinal spines, arranged in four rows. Inferomarginal row extended over entire ray length (Fig. 7B), its pseudopaxillae bear 30–45 spines in 4–6 transversal rows.

Ventrolateral pseudopaxillae bear 25–30 thorny spines in 4–5 rows. Adambulacrals with 10–11 spines in two transversal rows bearing numerous small thorns on apices; 3–4 larger adambulacrals near furrow spines; size of the other spines facing ventrally gradually decreases outwards from the furrow (Fig. 7E). Deep furrow spine small, single. Oral plates not fused, each bear single apical, four marginal, and four suboral blunt spines.

Color in life. Rays brownish-orange, abactinal side of the disc dark (Fig. 7C). Anal area almost black; madreporite partially creamy, partially brownish. In ethanol-preserved specimens, the madreporite is the same color as abactinal side of disc. Abactinal side of rays marked with dark brown to black spots of irregular shape, some of them look like broad transversal lines. Actinal side is uniformly colored orange-red, same as abactinal background color of rays.

Ecology. The species was found on rocky substrates at the depths of 1–20 m at water temperature 2–22 °C. In aquarium conditions, some were (one was?) kept over 10 months in an aquarium without any macroscopic food supply, which could indicate that they survived by grazing microscopic food, such as encrusting protists or diatoms.

Distribution. Known for the northwestern part of the Sea of Japan from Peter the Great Bay to Oprichnik Bay. Also, reported from Yellow Sea as *H. l. leviuscula* (*Xiao, Liao & Liu, 2011*). A species, *H. pseudoleviuscula* *Djakonov, 1958*, with similar skeleton pattern lacking
the ridges on the plates had been described from Moneron Island in the northwestern Sea of Japan. Life color of this species remains unknown but aboral spines in *H. pseudoleviuscula* is similar to the spines of type specimens of *H. reniossa* Hayashi, 1940, that are rather different from *H. djakonovi* (Figs. 7K–7M).

7. *Henricia (S.) hayashii* Djakonov, 1961 (Figs. 8A–8N)

*Henricia hayashii* Djakonov, 1961: 22–23, Pl. 2, fig. 8, Pl. 13, figs. 54–55); Chichvarkhin & Chichvarkhina, 2017a: fig4A; Chichvarkhin & Chichvarkhina, 2017b: 27, fig. H.

? *Henricia pseudoleviuscula* –Yavnov, 2010: 139–140 (non Djakonov, 1958).

Type locality: Sea of Japan, S off Povorotnyi Bay, Russia.

Examined material: ZIHU–2938 (holotype of *H. reniossa* f. *tohokuensis* Hayashi, 1940, Soyomaru 1930 station 630, off Tobisima, 200 m depth, 10 Aug 1930; ZIHU–2401 (holotype and paratypes of *H. saghaliensis* Hayashi, 1940, 3 broken spms, Albatross 1906, station 5023 Cape Patience, Sakhalin; 15 spms, MIMB–33250, Vostok Bay, Jul 2015, leg. I. Krasilnikov; 9 spms, Vostok Bay, 21 Aug 2016, leg. I. Krasilnikov; 5 spms Rudnyaya Bay, 6 Jun 2016, leg. A. Chichvarkhin; *H. hayashii* det. A.M. Djakonov: 1 spm, ZIN–2/14793, KSE, Toporok st. #117, 22 Sep 1949, Nemoro Sea, 57 m depth, sand, rocks; 1 spm, ZIN–5/15938, Taimyr [vessel], 17 Oct 1812, La Perouse Str, Redun-to Isl., 45°20′N 141°31′E, 40 m depth; 1 spm, ZIN–3/8863, Sea of Japan, st. 219, 1912, leg. Pavlenko; 1 spm, ZIN–4/6038 (marked as paratype of *H. hayashii*), TIRKh #1521, trawl #1, 21 Aug 1930, Sea of Japan, S off Povorotnyi Bay, 44 m depth, leg. Shurin; 1 spm, ZIN–8/8629, Far-Eastern expedition of Zool. Inst. Ac. Sci., st. 37, 8 Sep 1934, Sea of Japan, Sioukhu [Xiaohu?] Gulf; 1 spm, ZIN–6/15939, s/s Storozh st. 3, 20 Jun 1908, Sea of Japan, Askold Is., 25 m depth, on rocks, leg. Brazhnikova; 1 spm, ZIN–7/15940, s/s Storozh st. 6, 6 Jul 1908, Sea of Japan, Peter the Great Bay, 40–48 ftns depth, on rocks, leg. Brazhnikova; 1 spm, ZIN–6/15939, s/s Storozh st. 3, 20 Jun 1908, Sea of Japan, Askold Is; 1 spm, ZIN, Toporok st. #102, 6 Oct 1948, Moneron Is., 45 m depth, on gravel –more similar to *H. djakonovi* because of the absence of the ridges on intermarginal pates; 1 spm, ZIN–4/6038 (lectotype of *H. hayashii*), south off Povorotnyi Cape, Sea of Japan, 21 Aug 1930, 44 m, leg. Shurin.

External anatomy. The specimens, found in Vostok Bay possesses

| R | r | R:r |
|---|---|-----|
| 48 | 9 | 5.3-5.3 |

Rays five, cylindrical, slender. Aboral skeleton closely meshed. A bactinal plates (Fig. 8C) rod-shaped and triangular, tri-lobed, with low but discernable ridge. Abactinal pseudopaxillae reniform with 15–25 spines in two rows (Fig. 8B). Papular areas with 1–3 papulae. Abactinal spines (Figs. 8G–8M) long, 420 μm long, 100–120 μm wide at base, gradually extending proximally, with 10–15 apical and subapical thorns, not covered by skin membrane.

Superomarginal plates similar to adjacent aborals but slightly larger, bear 25–30 spines, arranged in 3–4 irregular rows. Intermarginal row extended to a third of ray length; plates round, with about 10 spines in one row. Inferomarginal plates large, with well discernible ridges transversally elongated, pseudopaxillae bear 15–21 spines in 2–3 irregular rows (Fig. 8C).

Ventrolateral plates small, oval; pseudopaxillae bear 10 spines in two irregular rows. Adambulacrals with four larger near-furrow spines and eight smaller spines in two rows.
Figure 8  *Henricia hayashii*. (A) life coloration, Vostok Bay; (B) abactinal side, lectotype; (C) denuded abactinal side, Vostok Bay; (D) actinal side, lectotype, (E) denuded actinal side, Vostok Bay; (F) actinal side of ethanol-preserved specimen; (G–J) abactinal spines, Vostok Bay; (K–L) abactinal spines, holotype; (M) single deep adambulacral spines in the specimen from Peter the Great Bay, ZIN–7/15940; (N) double deep adambulacral spines in the specimen from Moneron Is., Scale bars: A - 10 mm, B - 1 mm, C–F - 5 mm, G–L - 50 µm, M, N - 500 µm. Photos by A. Chichvarkhin.

Full-size DOI: 10.7717/peerj.6585/fig-8
(Fig. 8F). Deep furrow spine small, single, double at distal quarter of ray in some specimens. Oral plates each bear single apical, three marginal, and 1–2 suboral spines.

Color in life. Abactinal side bright orange, actinal side orange, lighter than abactinal (Fig. 8A). Anal area darker than background; madreporite light orange.

Ecology: The species was found on rocky substrate at the depth of 5–20 m at water temperature 16 °C.

Distribution. Known for the northwestern part of the Sea of Japan (Djakonov, 1961). Morphologically similar H. reniosa asiatica (Djakonov, 1958) was described from the northwestern shore of the Sea of Japan (Dzhygit Bay). Known specimens of this H. reniosa asiatica possess similar tightly meshed skeleton with distinct ridges on the inferomarginal plates as in H. hayashii. But it is significantly larger than any H. hayashii specimen that we ever studied (R = 142 mm), and possesses very unusually shaped aboral spines with blunt apex and a number of tiny denticles on it (Fig. 8N).

We identified the specimen imaged on fig. 54 of Djakonov (1961) in Henricia hayashii description in ZIN collection, it was labeled as the “holotype” presumably by ZI Baranova (Fig. 8). We found that this specimen does not possess double deep ambulacral spines (Fig. 8M) as described by Djakonov (1961). Also, we found a specimen identified by Djakonov as H. hayashii with two deep adambulacral spines (Fig. 8N) conforming the original diagnosis. It was the only one specimen with broken ray tip from Moneron Island, i.e., this is the only specimen in which deep spines might be counted earlier by Djakonov and/or Baranova: it is nearly impossible to observe these spines in an undamaged specimen. Therefore, we suppose that double deep adambulacral spine is not a distinctive character in this species, e.g., it is also specific for H. densispina (Sladen, 1878). All studied specimens from continental shore of the Sea of Japan possess single deep spine. Here, we are designating one of these specimens as the lectotype of Henricia hayashii (Djakonov, 1961): ZIN–4/6038, South off Povorotny Cape, Sea of Japan, Russia, 21 Aug 1930, 44 m, leg. Shurin.

8. Henricia asiatica Djakonov, 1958 (Fig. 9; Supplemental Information 1, first sea star: 1–5th s)

 Henricia reniosa asiatica Djakonov, 1958: 303–305, fig. 14; 1961: 22, pl. 2, fig. 11, pl. 12, figs. 52–53.

 Henricia asiatica – Chichvarkhin & Chichvarkhina, 2017a: 208, Fig. 4B; Chichvarkhin & Chichvarkhina, 2017b: 25.

Type locality: Dzhygit Bay, Tatar Str., Sea of Japan, Russia.

We found a specimen (Fig. 9) in ZIN collection identified as H. reniosa asiatica and labeled as the “holotype”. We are designating this specimen as the lectotype of Henricia reniosa asiatica (Djakonov, 1958). R = 142 mm, r = 16 mm; ZIN–2/12820, Opposite Dzhygit Bay, Tatar Strait, 14 Sep 1933, 54 m depth, leg. Kobyakova.

Examined material: lectotype, ZIN–2/12820, Opposite Dzhygit Bay, Tatar Strait, 14 Sep 1933, 54 m depth, leg. Kobyakova; 1 spm, ZIN–7/14192, Kurile Islands, KSE, 22 Sep 1949, Shikotan Is., opposite Anama Bay, 50 m depth; 1 spm, ZIN–8/15941, Sea of Japan, near
Figure 9  *Henricia asiatica*, lectotype. (A) abactinal view; (B) actinal side; (C) denuded actinal side; (D) abactinal side; (E) abactinal spines; (F) original labels. Scale bars: A - 20 mm, B–D - 10 mm, E - 50 µm. Photos by A. Chichvarkhin.
Kekurnyi Cape [Olga Bay], st. 39/451, 27 Oct 1931, 75–83 m depth, gravel, leg. Tarasov; 1 spm, ZIN, Toporok st. 93, 1949, E off Shikotan Is., 286 m depth leg. Kobyakova & Savelieva; 1 spm, ZIN, Dzhygit Bay, Tatar Str., o/tr [Otter-trawl] 73, 14 Sep 1933, leg. Kobyakova.

The specimen of this large sea star, which superficially resembles *H. (S.) asiatica* (= *H. reniossa asiatica*), had been recorded with the use of remotely controlled submersible apparatus south-east off the outer part of Vostok Bay (opposite Passek Cape, 42°44.6′N 132°43.8′E) at the depth of 45 m (appendix 1). R of the largest individual is about 90–100 mm, aboral coloration dark orange/mahogany. One of these specimens was associated with a large mussel, *Crenomytilus grayanus* (Dunker, 1853), probably filtering plankton from water currents generated by the mollusk.

Brun (1976) reported observations interpreted as suspension feeding in *Henricia* species, hypothesizing that some *Henricia* species may engage in rheophilic feeding behavior, i.e., collecting suspended phytoplankton in water currents generated by the sponges. If further supported, this could explain why *Henricia* individuals are often encountered sitting on the sponges, which has often led to an assumed sponge diet. However, most sponges feed on microbial cells, so it remains to be demonstrated that any *Henricia* could capture such tiny food items. However, we never observed suspension feeding behavior of any *Henricia as Rasmunsen (1965) reported for H. sanguinolenta* (O.F. (Müller, 1776). Suspension feeding on phytoplankton is a possible feeding strategy at shallow water habitats but is poorly effective at the depths >100 m, hence this species is likely polyphagous even if they feed on phytoplankton at shallower depths.

External anatomy. large in size, R (long radius: mouth to ray tip) of lectotype 142 mm, r (short radius: mouth to interradius) 16 mm, R:r 8.9; disc small, rays long, slender, subcylindrical, tapering to sharp tips (Fig. 9A). Abactinal plates with no tubercles relatively small forming tight reticulation; plates very close set, appearing almost fused; plates crowned with about 10 blunt spines, each tipped with 5–10 very small thorns (Fig. 9E); papular areas rather small, with 1 papula (Fig. 9D); madreporite small, circular, located about 2/3 way between anus and edge of disc. Superomarginals 1/3 larger than abactinal plates bear 40–45 spines, inferomarginals heigh about twice length; twice as large as superomarginal plates and bearing 45 spines; double series of intermarginals extends about 1/3 of R, each plate bearing 15–25 spines; ventrolateral series extending full length of R. Adambulacrals (Figs. 3D, 3E) each with a single deep furrow spine and 30–35 actinal spines, 5 longer spines at furrow edge, and 10–20 smaller distally grading spines behind (Figs. 9B, 9C). Oral plates with 5 marginal and 4–5 suboral spines; in addition, there are 4–5 thick, blunt spines near the distal edge of plate. Color in life unknown, probably orange.

Distribution. It was reported by Djakonov (1961) from southern Kurile Islands and from the northwestern Sea of Japan including type locality. He also reported this species from Peter the Great Bay because of a confusion of Dzhygit Bay (Tatar Strait, actual type location) vs. Noviy Dzhygit Bay (Russkiy Island, Peter the Great Bay).
Figure 10  *Henricia densispina*, Vostok Bay. (A, B) life coloration; (C) denuded abactinal skeleton; (D) abactinal side of live specimen; (E) denuded actinal side; (F) actinal side of ethanol-preserved specimen; (G–K) abactinal spines; (L–M) abactinal spines of ZIN –61/15229 H. sp., Tatar strait, Sovetskaya Gavan; (N) distal fragment of ambulacral furrow with double deep ambulacral spines; (O) proximal fragment of ambulacral furrow with single deep ambulacral spine. Scale bars: A, B - 10 mm, C, E - 5 mm, D - 3 mm, F - 1 mm, G–M - 50 µm. Photos by A. Chichvarkhin.

Full-size DOI: 10.7717/peerj.6585/fig-10
9. *Henricia (S.) densispina* (*Sladen, 1878*) (Fig. 10)

*Cribrella densispina* *Sladen, 1878*: 432, Pl. 8, figs. 5–9.

*Henricia densispina*—*Fisher, 1919*: 436–437, Pl. 118, figs. 2, 3, Pl. 119, figs. 1–3 (part.?)

*Hayashi, 1940*: 140, Pl. 10, fig. 7, 8; *Djakonov, 1961*: 20–21, Pl. 11, figs. 48–49; *Xiao, Liao & Liu, 2011*: 5–7, figs. 5A–5E, 12D; non *Djakonov, 1958*: 310, fig. 20; non *Yavnov, 2010*: 111–112; *Chichvarkhin & Chichvarkhina, 2017b*: 27, fig. I.

Type locality: Korean Strait, West coast of Japan.

Examined material: 1 spm, *MIMB–33249*, Vostok Bay, 20 Jul 2016, leg. A. Goloseyev; 1 spm; *Henricia spiculifera* as identified by *Djakonov*: 1 spm, *ZIN–61/15229*, Lesseis(?)-Data (Datta) Cape, mouth of Tumin Riv. near Imperatorskaya Gavan [Sovetskaya Gavan], 28 May 1909, leg. V. K. Arseniev; 3 spms, *ZIN–1/15155*, Okhotsk Sea, 50 m, 13 Aug 1918, leg. Meder; 3 spm, *MIMB–34800–34802*, Vostok Bay, 06–26 Jul 2017, leg. K. Dudka; *H. tacita* det. Z.I. Baranova, ex *H. leviuscula dyscrita* det. A.M. *Djakonov*: 1 spm, *ZIN–20/8457* (ex *8/8457*), GGI, 3 Jun 1932, tr. 1, #14, Okhotsk Sea, 118–60 m depth, leg. Polutov; 1 spm, *ZIN–23/8455*, GGI, s/s Ara, st. 25/1, 4 Sep 1939, Okhotsk Sea, leg. Schmidt; 1 spm, *ZIN–6/8454*, GGI, s/s Ara, st. 25/8, 6 Sep 1939, Okhotsk Sea, 56°24′N 142°12′E, 80 m depth, leg. Schmidt; *H. tacita* lectotype: 1 spm, *ZIN–14/12800*, KSE st. 57, Mordvinova Bay, 2 Sep 1947, 52 m depth, on clay, sand, gravel.

External anatomy. The specimens, which we found in Vostok Bay possesses $R = 32.5–55$ mm, $r = 7–11$ mm, $R: r = 4.6–5$. Rays five, broad at base, tapering. Disc broad. Aboral skeleton widely meshed. Abactinal plates tri-lobed with well discernible tubercles. Abactinal pseudopaxillae with 18–25 spines in 3–5 rows. Papular areas with 1–2 papulae. Abactinal spines slender, 450 $\mu$m long, 80–90 $\mu$m wide, with 3–4 long fine apical thorns (Figs. 10G–10K), not covered by skin membrane.

Marginal rows irregular proximally, after a half of ray length well discernible. Superomarginal row, consists of plates similar but slightly larger than adjacent aborals, bearing about 25 spines, same as abactinal spines, arranged in four irregular rows. Intermarginal row extended to full ray length; plates with about 20 spines in one row. Plates of intermarginal row oval, smaller than superomarginals. Inferomarginal plates relatively large, transversally elongated, with ridges. Pseudopaxillae bear 30–35 spines in 4–5 irregular rows.

Ventrolateral plates round or oval; pseudopaxillae bear about 20 spines in four irregular rows. Adambulacrals with three larger near-furrow spines and 8–9 smaller spines in three rows (Fig. 9F). Deep furrow spine small, single but double at ray tip (Figs. 10N, 10O). Oral plates each bear single apical, four marginal, and three suboral spines.

Color in life. Abactinal and actinal sides bright orange (Figs. 10A, 10B). Anal area darker than background; madreporite light creamy. Light yellow to orange at near-furrow area.

Ecology. The species was found on rocky substrate at the depth of 10 m at water temperature 20 °C.

Distribution. Known from the northwestern part of the Sea of Japan and South China Sea (*Xiao, Liao & Liu, 2011*) and Japan (*Hayashi, 1940*). Similar species previously reported...
Table 2 Number and distribution of recorded specimens in this study.

| Species   | Inner part of the bay (2015) | Outer part of the bay (2015/2016/2017) | Bottom                      |
|-----------|------------------------------|----------------------------------------|-----------------------------|
| alexeyi   | 0                            | 2/1/2                                  | nearshore boulders          |
| pacifica  | 0                            | 5/0/0                                  | rocks, pinnacles            |
| nipponica | 8                            | 29/15/1                                | various rocky               |
| granulifera| 0                           | 1/0/1                                  | rocks, pinnacles            |
| asiatica  | 0                            | 1/0/0                                  | silty                       |
| djakonovi | 52                           | 324/21/8                               | various rocky               |
| hayashii  | 1                            | 14/2/0                                 | rocks, pinnacles            |
| densispina| 0                            | 0/1/3                                  | rocks, boulders             |

by Djakonov (1961) from Tatar Strait as H. spiculifera and H. densispina likely belongs to a
different species possessing short and more stout spines with usually four apical thorns. A
species inhabiting the Sea of Japan referred to as H. spiculifera or H. leviuscula spiculifera
(Hayashi, 1940; Djakonov, 1961) and probably from South China Sea (Xiao, Liao & Liu,
2011) belong to a distinct species possessing very fine slender spines crowned with 3 long
thorns, while in H. densispina the spines are stouter (Xiao, Liao & Liu, 2011).

The specimens from Vostok Bay also resemble H. leviuscula dyscrita (Fisher, 1911)
described from California. This name was reported by Djakonov (1948) from the western
Pacific, Okhotsk Sea. Later, he did not mention this name in his publications. We found
three specimens in ZIN collection identified as H. leviuscula dyscrita, but later they were
re-identified as H. tacita Djakonov, 1958. We compared these specimens with the specimen
ZIN–14/12800 marked as “H. tacita holotype” presumably by Z.I. Baranova after A.M.
Djakonov’s death (although the holotype was not designated for this name by the author):
this specimen from Aniva Bay, off Sakhalin Island possess less discernible superomarginal
plates, the ridges on proximal marginal plates, and wider rays than re-identified exemplars
from Okhotsk Sea, i.e., this specimen strikingly resembles H. reniossa Hayashi, 1940
described from northern Japan, although these two species possess very distinctive abactinal
spines. There are four extra “H. leviuscula dyscrita det. Djakonov” specimens from Okhotsk
are listed in ZIN catalogue (ZIN–4/3453 and ZIN–10/8450) collected by Ushakov in 1932,
but we did not succeed in finding them. All these specimens are likely the syntypes of
H. tacita Djakonov, 1958. To avoid future confusion, we are designating the lectotype for
H. tacita Djakonov, 1958 with the specimen ZIN–14/12800 because it fully conforms to
Djakonov’s original diagnosis and photographed specimen (Djakonov, 1961: Pl. 11, fig.
44).

The specimens from Vostok Bay are different from all specimens mentioned above.
They do not possess crescent-shaped pseudopaxillae and smaller papular areas, i.e., do not
belong to H. tacita or H. leviuscula dyscrita.

Abundance and spatial distribution of Henricia in Vostok Bay
During 2010, 2014–2017, we recorded 537 individuals of Henricia spp. in Vostok Bay
(Table 2). Two of these individuals of Henricia sp. were recorded by a submersible
apparatus, although trawling in this area did not allow us to collect any Henricia species. Just three species were found in periodically desalted (32‰ to 2‰) inner part of the bay, while the others were strictly associated with always highly mineralized (appr. 32‰) outer part, and occurred on mainly rocky bottoms but sometimes on a silty substrate.

## Key to the genus Henricia of Vostok Bay

1a Spines not covered by thick skin membrane; spines in abactinal and inferomarginal pseudopaxillae in 3 or more rows or in small pseudopaxillae with 3–5 spines, spines with no lateral thorns ................................................................. subgenus Setihenricia Chichvarkhin & Chichvarkhina, 2017a; Chichvarkhin & Chichvarkhina, 2017b (2)

1b Abactinal spines covered by thick skin membrane, spines in double or single rows along plate ridges, spines with lateral thorns ................................................................. 5

2a At least a half of abactinal pseudopaxillae are crescent-shaped (reniform).................................3

2b The pseudopaxillae are not reniform. Abactinal pseudopaxillae circular, closely set leaving space for one or two papulae, with 10–15 long spines crowned with 3–4 sharp thorns; disc broad; rays tapering; color bright orange to pink .............................................. H. (S.) densispina (Sladen, 1878)

3a Inferomarginal plates lack ridges or tubercles, color orange to brown with wide almost back stripes. .............................................. H. (S.) djakonovi Chichvarkhin & Chichvarkhina, 2017a; Chichvarkhin & Chichvarkhina, 2017b 3

3b Inferomarginal plates with transversal ridges, well developed proximally, color bright orange .................................................................................................................................4

4a Abactinal spines with a crown of relatively large thorns, R <60 mm ................................................................. H. (S.) hayashii Djakonov, 1961

4b Abactinal spines large with blunt tip with tiny thorns, R may exceed 100 mm ......................... H. (S.) asiatica Djakonov, 1958

5a Marginal rows bent, poorly discernible or rudimental; abactinal and inferomarginal spines in pseudopaxillae with 3–5 spines; spines with lateral thorns; color brown, spine apices, oral, adambulacral and ventrolateral areas bright orange .................................................................................................................................

........................................................................ (H. Aleutihenricia) granulifera Hayashi, 1940

5b Marginal rows well discernible ........................................................................................................ subgenus Henricia Gray, 1840 (7)

6a Inferomarginal plates transversally elongated, 3–4-fold larger then ventrolaterals; Abactinal spines in one row; 2–3 papulae in an area; intermarginal row extends to full ray length; additional plates between intermarginal, inferomarginal, and superomarginal rows present; papulae very long; color brown, dirty orange ................................................................. H. (H.) nipponica Uchida, 1928.

6b Inferomarginal plates not larger then ventrolaterals, transversally not elongated ...............7

7a Abactinal spines in one row; superomarginal plates similar to abactinal; color bloody red, proximally, on ray tips, and on disc may be with white tint. Abactinal spines covered by very thick skin; mainly one papula in an area; .............................................. H. (H.) pacifica Hayashi, 1940

7b Abactinal spines in two rows, some pseudopaxillae with one row ......................................8

8a Rays thick; superomarginal plates small, similar to adjacent plates. One or few intermarginal rows extended to less than half of ray length. Wide-meshed abactinal skeleton with 2–3 papulae in an area. Coloration varies greatly: yellow-orange, pink, red, also in combinations of these colors ................................................................. H. (H.) oculata robusta Djakonov, 1958

8b Rays slim; proximal adambulacral, ventrolateral and inferomarginal spines joined in crests by skin membrane; tightly-meshed abs; coloration varies: yellow and orange on disk and tips or red and white on disk or uniformly orange, ventral side lighter colored ..........................................

H. (H.) alexyi Chichvarkhin & Chichvarkhina, 2017a; Chichvarkhin & Chichvarkhina, 2017b
DISCUSSION

Smirnov’s (2013) review of the invertebrates of Russian Pacific waters estimates that there are 21 echinasterid species reported from the Sea of Japan. Some of them were confirmed to the eastern part of that sea (H. aniva Djakonov, 1958, H. pseudoleviuscula Djakonov, 1958, H. sachalinica Djakonov, 1958, H. tacita Djakonov, 1958, H. orientalis Djakonov, 1950). Some of the listed species are dubious reports, including some that were likely misidentified (H. spiculifera Clark, 1901, H. aspera robusta Djakonov, 1948). Occurrence of deep-water H. reniossa asiatica Djakonov, 1961, which very likely is a distinct species related to a shallow-water H. reniossa Hayashi, 1940, is possible in Peter the Great Bay but we did not find it in Vostok Bay, probably because we focused mostly on shallow depths. Therefore, the total number of species that we have confirmed for Vostok Bay or its vicinity is estimated to be less than or about 15 species. Several Henricia species are known from adjacent waters of Korea and Japan, e.g. H. ohshimai Hayashi, 1940, H. reniossa Hayashi, 1940, H. nipponica Uchida, 1928, so they could also be found here.

In general, Henricia sea stars are common in Vostok Bay. However, H. djakonovi dominates among the others making about 80% of found individuals (Chichvarkhin, 2017a). The second abundant species is H. nipponica (about 15% of found individuals), while the others are rather rare (Table 2). The H. densispina, H. hayashii, and H. granulifera were encountered within this study just once despite the fact that the two latter species were found abundant in Rudnaya Bay, where water salinity is high and remains stable throughout the year. We hypothesize that most Henricia species are intolerant of decreased salinity in the inner part of Vostok Bay, and are restricted to the outer bay where the salinity was closer to oceanic. All species treated here inhabit near-shore rocky areas, with none found on the shallow silty bottom that completely occupies the central part of Vostok Bay and its inner part. The only exceptions were two remotely-recorded exemplars from deeper water on a silty bottom.

CONCLUSIONS

This study revealed seven Henricia species that inhabit Vostok Bay, with two additional species suspected as present but not yet sampled. All these species can be clearly identified with DNA barcoding approach using 16S rRNA marker. Morphological characters are also very valuable for identification of most species. Vostok Bay does not constitute a good habitat for high Henricia species diversity because of relatively low salinity water, especially in summer time, less than 30 m depths, and silty bottoms. However, a discovery of a few overlooked species remains possible along the outer shore of the bay.

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**Author Contributions**
- Anton Chichvarkhin conceived and designed the experiments, performed the experiments, analyzed the data, contributed reagents/materials/analysis tools, prepared figures and/or tables, authored or reviewed drafts of the paper.
- Olga Chichvarkhina conceived and designed the experiments, performed the experiments, contributed reagents/materials/analysis tools, authored or reviewed drafts of the paper.
- Daiki Wakita performed the experiments, prepared figures and/or tables, authored or reviewed drafts of the paper.

**Data Availability**
The following information was supplied regarding data availability:
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