Introduction

RA is a common systemic autoimmune disease with a prevalence of about 1% worldwide. Although the etiology of the disease is still unknown, several risk factors have been identified. Like many autoimmune diseases, RA occurs more frequently in women than in men (3:1 ratio), suggesting a role for sex hormones. Genetic studies have demonstrated that a genetic predisposition resides in the HLA-DR locus. There is also evidence that environmental factors, such as infectious agents, oral contraceptives and smoking, may play a role [1].

RA is characterized by inflammation of the synovial membrane of diarthrodial joints. Early indications of RA are swelling and pain of the proximal interphalangeal and metacarpophalangeal joints. Later, the larger joints become affected, especially those of the knee, elbow and ankle. Large numbers of activated leukocytes infiltrate the synovial membrane, causing hyperplasia and inflammation, which in most cases leads to progressive destruction of cartilage and bone. Since RA is a systemic autoimmune disease, other parts/organs of the body may become affected at a later stage. An example of this is the formation of rheumatoid noduli. Peak onset typically occurs in the fourth and fifth decades of life.

With more sophisticated types of therapy becoming available, it is becoming more important to diagnose RA at an early stage of the disease, so that earlier treatment can be applied and major damage of joint tissue can be prevented.

Keywords: autoantibodies, autoimmunity, citrulline, rheumatoid arthritis
It is during the first period of the disease, when not all clinical parameters are manifest, that a specific and sensitive serological test is needed. Even more helpful would be the serological test being able to predict, in an early phase of RA, the erosive or nonerosive progression of the disease.

Over the past 25 years, many autoantibody activities in RA have been described. This review summarizes the most relevant, and attempts to discuss their specificity and possible diagnostic and prognostic value. We shall distinguish between RA-associated antibodies, which are present in RA but also in other diseases, and RA-specific autoantibodies, which appear to be exclusively present in RA. It is, of course, the category of RA-specific antibodies that is most relevant for the clinician.

Autoantibodies associated with RA
Rheumatoid factor
The so-called ‘rheumatoid factor’ (RF) was first described about 75 years ago; since then a vast amount of work has been performed on the incidence, nature and specificity of RF (reviewed in [2]). The RF antibody is present in about 75% of RA patients, but its specificity is limited since RF is also found in patients with other autoimmune diseases (e.g. Sjögren’s syndrome), infectious diseases (e.g. hepatitis, tuberculosis), and to a certain extent in the healthy population (3–5%) and healthy elderly individuals (10–30%). Despite its relatively low specificity, the presence of RF is widely used as a diagnostic marker for RA.

RFs are antibodies directed to the crystallizable fragment of IgG molecules. They are found in every immunoglobulin subclass (IgE, IgM, IgA and IgG). IgG RF has a self-binding capacity that can result in the formation of very large immune complexes, which are able to (further) activate the immune system (reviewed in [3]). It is not clear whether RF is directly related to the symptoms of RA, although RF is found significantly more often in cases of aggressive joint inflammation. In those cases, RF titers are linearly related with the severity of inflammation. The clinical relevance of RF has been reviewed frequently (see, for example, [3,4]). Since the presence of RF is one of the American College of Rheumatology criteria for RA, the test is performed on a routine basis in most clinical laboratories. Some reports mention raised IgA-RF levels as a parameter for disease activity [5,6]. A combined routine determination of IgM-RF, IgG-RF and IgA-RF is recommended for an improved sensitivity, for diagnostic specificity and for predictive value [7,8].

Anti-R33 antibodies
Hassfeld et al. [9] described a 33 kDa antigen recognized by sera from RA patients on immunoblots from soluble nuclear extracts of HeLa cells. The antigen, referred to as RA33, was recognized by 36% of the RA sera and by only 1% of the normal sera. Characterization of RA33 revealed that RA33 is identical to the A2 protein of the heterogeneous nuclear ribonucleoprotein complex [10]. Heterogeneous nuclear ribonucleoproteins are large nuclear structures present in every eukaryotic cell composed of long stretches of heterogeneous nuclear RNA associated with about 30 different proteins, with molecular weights varying between 34 and 120 kDa. The most abundant proteins in the heterogeneous nuclear ribonucleoprotein complexes, the so-called ‘core’ proteins, are named A1, A2, B1, B2, C1 and C2.

It has been suggested that RA33 is a marker for early arthritis [11]. However, the autoantibodies to RA33/A2 protein are not exclusively found in RA patients. They are also present in systemic lupus erythematosus (SLE) (25% of investigated sera) and mixed connective tissue disease sera [11,12].

Anticalpastatin
Calpains are calcium-ion-dependent neutral cysteine proteinases. Two forms of calpains exist: µ-calpains (calpain I, which needs micromolar concentrations of Ca²⁺ for activity) and m-calpains (which require millimolar amounts of Ca²⁺ for activity). Substrates of these enzymes are highly diverse and include cytoskeletal proteins, nuclear proteins, cytokines and extracellular matrix proteins including proteoglycans [13]. Elevated levels of extracellular calpain in the inflamed synovium have been reported, which suggests that calpains might be secreted by synovial cells and could play a role in cartilage degradation in RA [14,15].

Calpastatin is the natural inhibitor of calpains. Using immunoblots of recombinant protein, autoantibodies directed to calpastatin can be found in about 45% of the RA sera, but also sera from SLE, myositis and systemic sclerosis contain antibodies directed to calpastatin [16,17]. A recent study using recombinant fusion proteins with truncated forms of calpastatin showed that anticalpastatin antibodies can also be found in sera from healthy individuals, even in comparable numbers, relative to RA patients [18]. Autoantibodies directed to calpastatin could increase calpain activity, leading to enhanced cartilage damage, and could therefore contribute to the severity of the disease [14].

Other RA-associated autoantibodies
RA patients produce autoantibodies directed to many autoantigens, but most of them are not specific for RA. Among these nonspecific antibodies are those directed to a wide variety of cartilage proteins such as collagen and fibronectin, but also antiphospholipid antibodies and antineutrophil cytoplasmic antibodies (ANCA) can often be found.

Although autoantibodies against several types of collagens have been described, the autoimmune reaction
directed to collagen type II (anti-CII) seems predominant. Anticollagen antibodies are present in about 30% of RA patients and only in HLA-DR4-positive ones [19]. Although anti-CII autoantibodies can be found in serum of only a small proportion of the RA patients, these antibodies and anti-CII-producing B cells are present in the joints of the majority of these patients [20]. It is not clear whether the occurrence of anticollagen autoantibodies plays a role in the pathogenesis of RA or is merely a result of the disease [3,19]. Anticollagen autoantibodies show a low specificity and can also be found in other autoimmune disorders (e.g. SLE, 20% occurrence; systemic sclerosis, 15% occurrence; relapsing polychondritis, 50% occurrence) and infectious diseases (e.g. leprosy, 50% occurrence). Anti-CII IgG titers in serum and synovial fluid appear to directly correlate with levels of acute-phase reactants, and of cytokines such as tumor necrosis factor-α and interleukin-6 [21]. A possible explanation for these findings is cross-reactivity with the first complement component, C1q, which has a collagen-like tail [22].

Fibronectin is a dimeric glycoprotein that interacts with collagen(s), fibrin(ogen), heparin and various cell surface adhesion molecules. It is present in connective tissue matrices and on the surface of epithelial cells. The protein is also (in a soluble form) present in plasma and other body fluids. Autoantibodies to fibronectin are found in about 14% of RA patients and show a low specificity (SLE, 34%) [23].

ANCA were initially discovered in serum of RA patients by immunofluorescence on neutrophilic granulocytes and can be subdivided into two groups depending on their localization: cytoplasmic ANCA, and perinuclear ANCA (reviewed in [24]). ANCA can be found in up to one-third of the RA patients, but can also be detected in other autoimmune conditions and various infectious diseases [25–27].

Autoantibodies directed to the nonhistone chromosomal proteins HMG1 and HMG2 can be detected in about 25–40% of the RA patients, but similar frequencies have also been reported for SLE, Sjögren's syndrome and scleroderma patients (see references in [28]).

Antiphospholipid antibodies, like autoantibodies directed to cardiolipin, are not only found in autoimmune diseases such as RA (4–49% occurrence) and SLE (~40% occurrence), but are also present in many infectious diseases (e.g. syphilis, 75% occurrence; leprosy, 67% occurrence). The incidence in RA can vary strongly, depending on the methodology of detection and the selection of patients (reviewed in [29]).

**Autoantibodies specific for RA Sa protein**

The antigen Sa is recognized by autoantibodies in about 40% of the RA sera with a rather high specificity [30,31]. These antibodies can already be detected at an early stage of the disease, albeit with a rather low sensitivity (23%) [30,32]. The antigen is a 48–50 kDa protein, predominantly present in human placenta extracts [31]. It can also be detected in human spleen and RA pannus tissue, but not in cultured cells. As explained by Menard et al. [33], Sa possibly is a citrullinated form of the intermediate filament protein vimentin (in cultured cells, vimentin is normally not citrullinated). Other workers have postulated that part of the Sa activity could be identical to human α-enolase [34]. If the Sa protein indeed is citrullinated, the antigen can be categorized in the filaggrin/cyclic citrullinated peptide (CCP) family of citrullinated auto-antigens (see below).

**Heavy chain binding protein (p68)**

Autoantibodies directed to the p68 protein, recently identified as heavy chain binding protein (BiP), occur in about 64% of RA patients and appear to be highly specific for the disease [35–37]. The reported specificity of these antibodies still awaits further confirmation by clinical studies. The BiP antigen is also a target of RA-specific T-cell and B-cell immune responses [38], and is overexpressed in RA synovial tissue compared with control tissue. BiP is a ubiquitously expressed chaperone protein predominantly localized in the endoplasmic reticulum. The major B-cell epitope of this antigen appears to be an N-acetylgalactosamine carbohydrate group, which is possibly important for its cellular localization [39]. The BiP protein can be detected in the cytoplasm and endoplasmic reticulum of cultured cells, but also in low abundance at the cell surface. Under stress (especially heat shock) conditions, the antigen can relocate to the nucleus [39]. This nonphysiological localization of the antigen, accompanied by a shift in its glycosylation pattern, might trigger antigenicity of BiP during the pathogenesis of RA. Anti-BiP antibodies can also be detected in mice with established collagen-induced or pristane-induced arthritis [37].

**Glucose-6-phosphate isomerase**

The household enzyme glucose-6-phosphate isomerase (GPI) was recently described as a novel autoantigen in RA [40]. The antibodies could be detected in 64% of RA patients but not in controls. Interestingly, anti-GPI antibodies are also present in T-cell receptor-transgenic mice. In this mouse model system, GPI can serve as an autoantigen for both B and T cells, and passive transfer of anti-GPI to normal healthy animals induces arthritis. The anti-GPI antibodies in humans appear to be produced locally, since anti-GPI titers were higher in RA synovial fluids as compared with RA sera. Immunohistochemistry of synovia from RA patients showed high concentrations of GPI on the surface of the synovial lining and on the endothelial cell surface of arterioles [40]. It is intriguing and significant that the same autoantigen is targeted by autoantibodies in RA patients and in a mouse model of RA.
Antiperinuclear factor/antikeratin antibodies/antifilaggrin/anti-CCP

In 1964 a highly specific RA antibody system directed to a protein component present in the keratohyaline granules in the cytoplasm of differentiating buccal mucosa cells was described. The antigen was referred to as perinuclear factor, and the antibody activity became known as antiperinuclear factor (APF) [41]. APF antibodies combined a relative high sensitivity (depending on the cohort of patients being studied, they were present in 49–91% of RA patients) with a strong specificity (73–99%, reviewed in [42]). The APF test never became popular because of several practical inconveniences. Only a few percent of an average (laboratory) population can donate buccal mucosa cells differentiated far enough to contain the perinuclear factor. Another inconvenience is the fact that the antibody activity can only be measured by indirect immunofluorescence, making interpretation of test results sometimes very difficult [42].

A related group of RA-specific autoantibodies, the so-called antikeratin antibodies (AKA), was first described in 1979 [43]. These antibodies stain keratin-like structures in the cornified layer of esophagus cryostat sections but do not recognize cytokeratins, as is suggested by the name. AKA can be detected by indirect immunofluorescence in 36–59% of RA sera with a specificity of 88–99% (reviewed in [42]). Several studies have shown that the APF and AKA antibodies target the same antigen, the epithelial protein filaggrin (for references see [44]).

Filaggrin (filament aggregating protein) is involved in the organization of cytoskeletal structures in epithelial cells and is synthesized as a large, heavily phosphorylated precursor protein, profilaggrin. During differentiation of epithelial cells, profilaggrin is partly dephosphorylated and proteolytically cleaved into 10–12 filaggrin subunits. Finally, about 20% of the arginine residues are enzymatically deiminated to citrulline residues [45]. Conversion of the basic amino acid arginine into the neutral residue citrulline is catalyzed by the enzyme peptidylarginine deiminase (Fig. 1). It is this modification that has been shown to be essential for the autoantigenicity of filaggrin [46,47].

Immunoblotting assays and an enzyme-linked immunosorbent assay (ELISA) using filaggrin purified from human skin as an antigen have been developed for the detection of antifilaggrin antibodies (AFA). Vincent et al. [48] could detect AFA in 41% of RA sera with 99% specificity. When combining the AFA immunoblot assay with AKA testing, a much higher sensitivity (64%), without loss of specificity, could be achieved [48]. However, the sensitivity of the assay appears to be dependent on the method for purification of the filaggrin. Slack et al. calculated sensitivities of 12 and 16% for two different filaggrin preparations, while only one of five positive sera reacted with both preparations [49]. The AFA-ELISA is somewhat more sensitive (47–54%) than the immunoblot assay [50,51], but the results are in good agreement with APF data obtained with the same set of sera [52]. Using in vitro deiminated recombinant filaggrin, Nogueira et al. [53] could detect AFA in about 52% of RA sera. The differences in AFA, APF and AKA results and the influence of the antigen preparation imply that several (rather than one single) epitopes are recognized.

The AFA-ELISA test detected 45% of RF-negative erosive cases, in particular those with larger numbers of erosive joints. Testing for AFA could therefore supplement RF in the prediction of erosive disease. The test could also be helpful in the diagnosis of RA in those cases that do not completely fulfill the 1987 American College of Rheumatology criteria [52]. In a case–control study, pre-illness AFA titers were found to directly be related to the risk of developing RF-positive RA, but not RF-negative arthritis [54]. AFA are present early in the disease, as is the case with APF and anti-CCP antibodies (see below), and raised AFA levels correlate with the severity of the disease. However, there is discussion whether disease activity and radiological damage can be predicted by AFA levels [51,52,55].

One disadvantage of the AFA test is the difficulty of obtaining an antigen preparation that is pure and contains a reproducible citrulline content. These problems do not occur when citrullinated peptides are used as the antigen, as has been shown by Schellekens et al. [46]. RA sera, however, show different patterns of reactivity with the citrullinated peptide variants, which emphasizes the heterogeneous nature of the autoimmune response. With a combination of nine citrullinated peptide variants, a sensitivity of 76% could be reached with a specificity of 96% [46]. The test could be improved and simplified using a single cyclic peptide that has a three-dimensional structure optimally recognized by the heterogeneous population of RA autoantibodies [56]. Recent studies indicate that this anti-CCP test (manufactured by Euro-Diagnostica
BV, Arnheim, The Netherlands) is extremely specific (~98%) and has a reasonable sensitivity of 68–75% [57,58] (van Boekel and de Jong, unpublished observations, 2001). Anti-CCP antibodies can be detected very early in RA, although with a somewhat lower sensitivity (40–60%) [32,57–60]. Anti-CCP appears to be a good prognostic marker and has a high discriminating power between erosive and nonerosive RA [61]. RA patients positive for anti-CCP develop significantly more radiological damage than anti-CCP-negative patients [59,60], although anti-CCP combined with RF appears to be an even better prognostic marker [56,58,61].

The results described (supported by the fact that non-filaggrin-like citrulline-containing antigens are also recognized by RA sera [62]) suggest that, in principle, most citrullinated proteins/peptides will be recognized by autoantibodies in RA sera, all be it with different sensitivities and specificities. Since filaggrin is not present in the joint, it is reasonable to assume that other citrulline-containing proteins are present in the RA synovium. In RA pannus tissue, AFA are produced by local plasma cells and constitute a higher proportion of IgG than in serum [63]. Furthermore, B cells from the synovial fluid of anti-CCP-positive RA patients spontaneously produce anti-CCP antibodies, while peripheral blood B cells or B cells from anti-CCP-negative RA patients do not [64]. This suggests an antigen-driven maturation of CCP-specific B cells at the site of inflammation in RA. One of these possible autoantigens, citrullinated fibrin, has recently been identified [65]. The possible role of citrullinated fibrin and other citrullinated antigens in the pathophysiology of RA remains to be elucidated [66].

Conclusions and discussion

RA is accompanied by the occurrence of many autoantibodies in the serum of the patient. Most of these antibodies are not specific for RA because they also occur in other inflammatory conditions. Other antibodies appear to be more specific and are, in some cases, almost exclusively present in RA.

The possible pathogenic nature of autoantibodies in RA is still controversial (reviewed in [67]). The evidence for the pathogenic involvement of RF in RA can, at best, be considered indirect [67]. There is an association of RF with the severity of RA, and RF-positive patients can turn RF-negative following therapy [3,68]. Administration of human IgG or IgM monoclonal RF to mice (suffering from collagen-induced arthritis) results in an enhanced activity and severity of the disease and in an increase in anticollagen antibody levels [69]. RF has the capacity to activate the complement cascade, which could be an important factor in the disease activity and the chronic character of RA [3,70]. RF is also found in healthy individuals, however, and possibly even plays a physiological role in immune complex clearance [3].

Anticollagen antibodies possibly have pathogenic potential. Injection of serum IgG from a RF-negative patient with high levels of anti-CII antibodies induces a mild form of inflammatory arthritis in mice [71,72], but the pathogenic involvement of anti-CII antibodies is much less evident in human RA. Anticalpastatin antibodies could contribute to the pathology by inhibiting calpastatin functioning [13]. The participation of other autoantibody systems (such as anticitrullinated protein, anti-GPI and anti-Sa antibodies) in RA pathogenesis is still unclear, although their high specificity adds to the increasing body of evidence that they may be important in maintaining RA.

Some of the RA-specific antibodies might be very helpful in the early diagnosis and prognosis of the disease. These antibodies are presented in Table 1. Most promising for the future appears to be the growing family of antigens containing one or more citrulline residues. Some of these antigens (i.e. filaggrin and CCP) have recently been applied in simple and easy-to-use ELISA tests, and have been shown to be very specific for RA, to be able to diagnose RA in a very early phase of the disease, and to predict erosive disease. As such, this class of anticitrullinated protein autoantibodies has the potential to become the serological marker for RA of the future.
Although we know that the anticitrullinated protein antibodies are locally generated in the inflamed joint, we also know that both filaggrin and CCP are not present in this tissue. It was recently shown that fibrin in the pannus tissue is citrullinated. This intriguing finding underlines the idea that local citrullination of extravasal proteins might be one of the initiating event(s) leading to autoimmunity in RA.

The key question why only patients with RA develop antibodies to citrullinated (synovial) proteins remains to be answered. Further research will hopefully reveal the significance of protein citrullination in the immunopathology of RA.

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