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SCREENING OF CONTENT AND DYNAMIC OF ACCUMULATION OF POLYPHENOLS IN SOME BASIDIOMYCETES SPECIES

O. V. FEDOTOV, A. K. VELIGODSKA

Donetsk National University
Universitetska str., 24, Donetsk, 83000, Ukraine,
e-mail: bio.graff@yandex.ua

The aim of the study was to investigate the total content of polyphenolic substances in Basidiomycetes carpophores from 50 species, of which 27 belong to the order Polyporales and 23 to the order Agaricales. Introduced 23 strains of 8 species of Basidiomycetes. Methods. Gathered wild carpophores dried and crushed to a particle size of 0.1 till 0.01 mm and searching strains were cultured in Erlenmeyers flasks by surface method on standard glucose-peptone culture medium. Determination of total content of polyphenolic compounds was carried out in ethanol extracts of mycelial and mycelial complex, carotenoids, flavonoids, melanins, hyde derivatives, substances of polyphenol nols. It was determined the content of polyphenols ranging from more than 60 mg / g completely dry biomass. For introduced strains established dynamics of growth and accumulation of polyphenolic compounds in the mycelium and culture filtrate during fermentation on glucose-peptone medium. All cultures reach a maximum accumulation of biomass on the 12th day of growth. Shizophyllum commune Sc-1101 and 10 and F. velutipes F-202 have been identified as the most productive strains. The lowest accumulation of absolutely dry biomass was recorded for strain P. ostreatus P-192 and strain F. fomentarius Ff-09. Cultures have investigated individual value growth such as biomass accumulation in the applied cultivation conditions, which probably reflects the suitability of the medium for their growth and genotypic characteristics. Strains are overwhelmingly able to accumulate polyphenolic compounds in both mycelium and culture fluid during the whole period of cultivation. Maximum content of polyphenols in the mycelium to 96%, and in the culture fluid - for 91% of strains coincided with the end of their term cultivation. Calculated correlation coefficient between the content of polyphenols in the mycelium and culture fluid showed that there is a very high positive correlation of 73.2%, a positive high at 17.4% and the average 4.5% of experiment data. Conclusion. The strains of species Shizophyllum commune, Pleurotus ostreatus, Fistulina hepatica and Laetiporus sulphureus were selected for further research in order to obtain polyphenols mycelial and extracellular origin.

Key words: polyphenols, Basidiomycetes, carpophores, mycelium, cultural filtrate

Introduction. In the last decades an actual problem is searching for new biologically active substances (BAS’s) and their producers for the purpose of development and manufacturing application of modern drug and therapeutic products (Запрометов, 1993; Никитина, 2007).

In particular, polyphenol compounds, which are natural antioxidants preventing development of different pathogenic effects in a cell and as a result of different diseases, are the desired substances in different branches of industry and medicine (Запрометов, 1993; Федотов ат. ал., 2012 Asatiani ат. ал., 2010). These include phenolic acids and aldehyde derivatives, substances of polyphenoloxycarbon complex, carotenoids, flavonoids, melanins, tannins, etc. (Asatiani ат. ал., 2010; Wasser, 2010).

It is established that these substances are synthesised by almost all plant and fungi organisms (Никитина, 2007; Li Fu ат. ал., 2011). The traditional sources of polyphenols are plant raw materials - Camellia sinensis and Humulus lupulus, as well as the fruits of Vitis vinifera (Li Fu ат. ал., 2011, Halvorsen B.L. ат. ал., 2011). A number of studies are devoted to investigation of polyphenols in fungi, especially to investigation of total polyphenols in the fruit bodies of 49 species of edible fungi which belong to genera Boletus, Suillus, Volvariella, Pleurotus et al. (Guthalu Puttaraju Nethravathi ат. ал., 2006; Guo Ya–Jun ат. ал., 2012). However, these studies give a vague idea about the qualitative and quantitative content of polyphenols in higher basidi fungal and mycological material when cultivating them that makes it necessary to carry out further screening operations in this field.

The interest for basidiomycetes, including wood-destroying ones, is firstly associated with their ability to synthesise numerous BAS’s. Especially while destructing lignin-cellulose complex, they produce antioxidant substances - oxidoreductases, vitamins, polyphenols, free radical blockers et al., which provide adaptive mechanisms for antioxidant protection of xylotrophs (Peyrat–Maillard ат. ал., 2000; Fedotov,
The aim of this study was to determine total polyphenols in the carpophores and mycelium and in the culture filtrate of some basidiomycetes species.

**Materials and methods.** The carpophores, mycelium and culture filtrate of 50 macromycetes species, 27 of which belong to order Polyporales and 23 to order Agaricales, division Basidiomycetes were used as the materials for this investigation. The general information about the species of basidal fungi investigated is set forth in the published work (Fe-доров ат. al., 2012) and relevant sections of the article. Also 23 strains from a piliated fungi culture collection of the Physiology of Plants Department, Donetsk National University: *Fomes fomentarius* (L. ex Fr.) Gill. – T-10, Ff-09, Ff-1201; *Laetiporus sulphureus* (Bull.) Murrill. – Ls-08, Ls-09; *Fistulina hepatica* Schaff. ex Fr. – Fh-08, Fh-18; *Flammulina velutipes* (Curt.: Fr.) Sing. – F-03, F-06, F-1, F-202; *Pleurotus ostreatus* (Jacq.: Fr.) P. Kumm. – Hk-35, P-004, P-01, P-039, P-107, P-192, P-208; *Schizophyllum commune* Fr.: Fr. – Sc-10, Sc-1101, Sc-1102; *Trametes hirsuta* (Wulf.:Fr.) Pil. – Th-11 and *Trichaptum biforme* (Fr.) Ryv. – Tb-11. Most of the introduced strains are isolated into a pure culture from the wild fruit bodies (FBs) of basidiomycetes collected in different areas of Donetsk oblast, the taxonomic position of which was established according to the modern literature (Kirk аt al., 2001).

To determine total polyphenols, the collected FBs were dried and ground to the particle size of 0,1±0,01 mm, and the strains under investigation were cultivated superficially in Erlenmeyer flasks of 250 ml in a glucose-peptone growth medium (GPM, pH0 6,5±0,2) of 50 ml with the following composition (g/l): glucose – 10,0; peptone – 3,0; KH₂PO₄ – 0,6; K₂HPO₄ – 0,4; MgSO₄·7H₂O – 0,5; CaCl₂ – 0,05; ZnSO₄·7H₂O – 0,001. 10-day mycelial strain cultures on wort agar were used as inoculums. The incubation temperature was 27,5°C. The cultivation period was 6, 9 and 12 days. Upon completion of the cultivation period, a mycelium was separated from the culture broth by filtration at 5±1°C. The mycelium obtained was additionally dried a little on filtration paper and cooled down to ±0,5°C. The prepared mycelium was homogenized by grinding in a chilled mortar. The ground carpophores (GCs), homogenized mycelium (HM) and culture filtrate (CF) were used for further tests.

Absolutely dry biomass (ADB) of the GCs and mycelium was determined by gravimetric method (Государственная фармакопея, 1987).

The determination of total (W) polyphenols (PPs) was carried out in the alcoholic extracts of the mycological material using modified Folin–Ciocalteu method (Мусиенко ат. al., 2001) and calculated by the standart formula.

The tests were performed in three replications. Statistical manipulation was done using the programs for statistical manipulation of biological test results. A correlation analysis was performed to determine correlation level between PP concentration in the mycelium and CF of one-year cultures. A difference was considered as significant at confidence level P>0,95 (Присецкий, 1999).

**Results and their discussion.** Total polyphenols in 225 carpophores of 27 species of Polyporales fungi and in 220 ones of 23 species of Agaricales fungi were evaluated at the first step of investigation (Table 1).

The analysis of polyphenols in the carpophores of Polyporales fungi showed the following. Most of their FBs (85%) contain little polyphenols being within the range of 9 mg/g (*D. quercina*) to 39 mg/g (*F. pinicola*). The other group comprises 3 species of Polyporales fungi (*G. lucidum, L. sulphureus* and *G. applanatum*) containing polyphenols in their FBs in the amount of 89 mg/g to 161 mg/g (ADB). The fruit bodies of tinder fungus *F. fomentarius* have the highest concentration of polyphenols - more than 248 mg/g (ADB). To compare the results obtained, it should be noted that the method of reactant extraction from the carpophores of shelf fungus *Inonotus obliquus* (Ach. ex Pers.) Pil. with the maximum PP concentration of 140 mg/g (ADB) by water extraction has been patented (Сысоева ат. al., 2005; Патент 2448721).

The analysis of polyphenols in the carpophores of Agaricales fungi showed that most of them (74%) contain little polyphenols in their FBs but it is less compared to Polyporales fungi. Here they are within the range of 12 (*P. squarrosa*) to 37 mg/g (*P. citrinopileatus*). 5 species of Agaricales fungi (*P. ostreatus*, *S. rugosoanulata*, *T. flavovirens*, *F. velutipes* and *A. cylindracea*) can be put into a group with the medium polyphenol concentration of 53 to 101 mg/g in the fruit bodies. The highest polyphenol concentration of about 172 mg/g was registered in the wild fruit bodies of *F. hepatica*. However, this parameter is more than 1,5 times lower than the concentration of phenolic substances in the fruit bodies of tinder fungus *F. fomentarius*. In contrast, it should be noted that the mean concentration of polyphenols amounting to 450 mg/g in the plant raw material of *Camellia sinensis*, and 70 mg/g in the mycological material (fruit bodies of *P. ostreatus*) Li Fu аt al., 2011; Halvorsen B.L. аt al., 2011).

The analysis of polyphenols in the alcohol extracts from the fruit bodies of 50 basidiomycetes...
species made it possible to isolate some species of tinder fungi: *G. lucidum*, *L. sulphureus*, *G. applanatum* and *F. fomentarius*, and some species of Agaricales fungi: *S. rugosoannulata*, *A. cylindracea*, *T. flavovirens*, *F. velutipes*, *P. ostreatus* and *F. hepatica* having the high concentration of these substances of more than 60 mg/g (ADB).

The next step of the investigation was to isolate pure cultures from the carpophores as well as to study the growth and synthesis rate of polyphenols of some of them when cultivating in the GPM.

### Table 1

Total content of polyphenols in fruit bodies of some species of Basidiomycetes

| Species                              | Number of the samples | Polyphenols, mg/g |
|--------------------------------------|-----------------------|-------------------|
| **Order Polyporales**                |                       |                   |
| *Auricularia auricula-judae* *       | 12                    | 32.53 ± 3.52      |
| *Laetiporus roseum* *                | 3                     | 20.02 ± 0.58      |
| *Chaetoporus ambigus* *              | 6                     | 20.66 ± 0.95      |
| *Sparassis crispa* *                 | 9                     | 10.54 ± 0.19      |
| *Fibulopsis mollusca* *              | 6                     | 10.54 ± 0.35      |
| *Tyromyces lacteus* *               | 9                     | 16.07 ± 0.76      |
| *Tyromyces revolutus* *              | 3                     | 12.09 ± 0.16      |
| *Tyromyces undosus* *               | 6                     | 10.33 ± 0.13      |
| *Irpex lacteus* *                   | 9                     | 26.75 ± 0.43      |
| *Amylopora lenis* *                 | 3                     | 15.05 ± 0.21      |
| *Hydnellum aurantiacum* *           | 12                    | 13.33 ± 0.64      |
| *Fomitopsis pinicola* *             | 6                     | 39.19 ± 0.58      |
| *Daedalea quercina* *               | 6                     | 9.02 ± 0.13       |
| *Piptoporus betulinus* *            | 12                    | 15.10 ± 0.10      |
| *Polyporus squamosus* *             | 9                     | 23.20 ± 0.37      |
| *Laetiporus sulphureus* *           | 9                     | 117.04 ± 0.56     |
| *Ganoderma applanatum* *            | 9                     | 161.08 ± 0.19     |
| *Ganoderma lucidum* *               | 15                    | 89.06 ± 1.5       |
| *Inonotus obliquus* *               | 12                    | 20.55 ± 0.31      |
| *Phellinus igniarius* *             | 9                     | 34.53 ± 0.55      |
| *Phellinus pomaceus* *              | 9                     | 19.04 ± 0.59      |
| **Order Agaricales**                |                       |                   |
| *Agaricus arvensis* *               | 5                     | 24.57 ± 4.07      |
| *Agaricus bisporus** *              | 9                     | 35.44± 0.63       |
| *Agaricus campestris* *             | 5                     | 23.46 ± 0.10      |
| *Agrocybe cylindracea**             | 9                     | 75.85 ± 1.22      |
| *Coprinus comatus* *                | 15                    | 25.04 ± 0.58      |
| *Coprinus micaceus* *               | 15                    | 25.03± 0.15       |
| *Fistulina hepatica* *              | 9                     | 172.25 ± 0.20     |
| *Flammulina velutipes* *            | 27                    | 81.25 ± 7.75      |
| *Flammulina velutipes**             | 3                     | 65.06 ± 0.92      |
| *Lentinus edodes** *                | 9                     | 35.47 ± 0.42      |
| *Marasmius oreades* *               | 3                     | 37.08 ± 0.65      |
| *Pleurotus citrinopileatus**        | 3                     | 37.55 ± 0.11      |
| *Pleurotus eryngii**                | 6                     | 15.03 ± 0.42      |
| *Pleurotus ostreatus* *             | 34                    | 100.56 ± 3.15     |
| *Pleurotus ostreatus var. Florida** | 3                     | 53.07 ± 2.01      |
| *Kuehneromyces mutabilis* *         | 9                     | 32.28 ± 0.83      |
| *Pholiota aurivella* *              | 3                     | 18.02 ± 0.35      |
| *Pholiota squarrosa* *              | 3                     | 12.04 ± 0.65      |
| *Schizophyllum commune* *           | 21                    | 19.29 ± 0.27      |
| *Stropharia aeruginosa* *           | 3                     | 32.53 ± 0.54      |
The results of ADB accumulation by the strains while their growing (on the 6th, 9th and 12th day of cultivation) are shown on Figure 1. As can be seen, all cultures reach the maximum of this parameter on the 12th day of their growth. The most productive strains here are those of S. commune Sc-1101 and Sc-10 and that of F. velutipes F-202. The lowest values of ADB accumulation were registered for P. ostreatus strain P-192 and F. fomentarius strain Ff-09. Thus, the investigated cultures have their individual growth values – biomass accumulation in the cultivation conditions used that probably reflects suitability of these conditions for their growth.

It was established that most of strains were able to accumulate polyphenols both in the mycelium and CF during the whole cultivation period. The maximum of PP concentration in the mycelium for 96% and in the CF for 91% of the total strains coincided with the termination of their cultivation period. The investigation results of total polyphenols in the mycelium and culture filtrate while growing some basidiomycetes strains are shown on Figure 2 and 3.

| Strain                         | 6   | 59.56 ± 1.85 |
|--------------------------------|-----|--------------|
| Stropharia rugosoannulata **   | 5   | 21.37 ± 0.63 |
| Lyophyllum loricatum *         | 5   | 20.42 ± 0.12 |
| Lyophyllum connatum *         | 5   | 79.08 ± 0.20 |
| Tricholoma flavovirens *      | 5   | 31.48 ± 0.52 |
| Tricholoma sejunctum *        |     |              |

Note: "**" – wild in nature CF, "***" – commercial CF

**Fig. 1. Accumulation of absolute dry biomass of strains of Basidiomycetes on the 6th, 9th and 12th days of cultivation**

**Fig. 2. The total content of polyphenols of mycelium of strains of Basidiomycetes on 6th, 9th and 12th days of cultivation**

Note: DC – day of cultivation.
The trend of polyphenols in the mycelium of the strains under investigation has the following characteristics. The highest concentration of these substances within the range of 107.9 to 129.4 mg/g was registered for *P. ostreatus* strains P-039 and P-208, and *S. commune* strain Sc-1102 on the 12th day of their growth. The lowest values of PP concentration of 29.9 to 50.1 mg/g were registered for *F. velutipes* strains F-1, F-202 and F-03, and *P. ostreatus* strain P-192 at the end of their cultivation period.

The trend analysis of polyphenols in the culture filtrate of the strains under investigation showed the following. The highest PP concentration was registered on the 12th day of their growth within the range of 5.4 to 6.8 mg/ml for the strains of *F. hepatica* FH-18, *L. sulphureus* LS-08 and *P. ostreatus* P-01, and the lowest (0.9 to 1.7 mg/ml) for the strains of *F. hepatica* FH-08, *P. ostreatus* P-192 and *F. velutipes* F-1.

In all cases the concentration of polyphenols in the mycelium was much higher than the concentration of these substances in the culture filtrate and varied on the 12th day of cultivation from 11.4 times for *L. sulphureus* strain LS-08 to 52.9 times for *F. hepatica* strain FH-08. A considerable difference between ability of strains to synthesize and accumulate PPs in the mycelium and CF may be most likely explained by their genotype realization under test conditions. The calculation of a correlation coefficient between PP concentration in the mycelium and CF of one-year cultures showed the following. A very high positive correlation is observed in 73.2%, high positive one – in 17.4% and medium one – in 4.5% of tests.

Thus, the test results of total polyphenols in some basidiomycetes species enable us to make the following conclusions. The species of tinder fungi - *Ganoderma applanatum*, *Ganoderma lucidum*, *Laetiporus sulphureus* and *Fomes fomentarius*, and the species of Agaricales fungi - *Stropharia rugosoannulata*, *Agrocybe cylindracea*, *Tricholoma flavovirens*, *Flammulina velutipes*, *Pleurotus ostreatus* and *Fistulina hepatica* are characterized by the highest concentration of polyphenols in their carpophores. Most of introduced strains can accumulate polyphenols both in the mycelium and CF during the whole period of their cultivation. The strains of *P. ostreatus* P-01, *F. hepatica* FH-18 and *L. sulphureus* LS-08 are perspective for further investigations aimed to obtain polyphenols of extracellular origin and the strains of *S. commune* Sc-1102, and *P. ostreatus* P-039 and P-208 – those of mycelial one.

**References:**

1. Зареметов М.Н. Фенольные соединения / М.Н. Зареметов. – М.: Наука, 1993. – 271 с.
2. Никитина В.С. Антибактериальная активность полифенольных соединений, выделенных из растений семейств Geranieaeae и Rosaceae / В.С. Никитина; Л.Ю. Кузьмина, А.И. Мелентьев, Г.В. Шендель // Прикладная биохимия и микробиология. – 2007. – Т. 43, № 6. – С. 705–712.
3. Присядский Ю.Г. Статистическая обработка результатов биологических экспериментов / Ю.Г. Присядский. – Донецк: Кассиопея, 1999. – 210 с.
4. Сысоева М.А. Структурная организация и свойства полифенолов чаги / М.А. Сысоева, О.Ю. Кузнецов, В.С. Гамаюрова // Вестник Казанского технологического университета (КГТУ). 2005. – №1. – С. 244–250.
5. Федотов О.В. Колекция культур шапинковых грибов – основа микологических досліджень та стратегії збереження біорізноманіття базидіоміцетів / О.В. Федотов О.В. Колекція культур шапинкових грибов – основа микологічних досліджень та стратегії збереження біорізноманіття базидіоміцетів / О.В. Федотов О.В. Колекція культур шапинкових грибов – основа микологічних досліджень та стратегії збереження біорізноманіття базидіоміцетів / О.В.
Федотов О.В. Чайка Т.С. Волошко А.К. Велигодська / Вісник Донецького університету Сер. А: Природничі науки. вип. 1. – Донецьк: ДонНУ, 2012. – С. 209–213.
6. Asatiani M.D. Higher basidiomycetes mushrooms as a source of antioxidants / M.D. Asatiani, G. Elisashvili, A.Z. Songulashvili, V. Reznick, S.P. Wasser // Progress in Mycology. – 2010. – р. 311–327.
7. Kirk P.M. Ainsworth & Bisby’s Dictionary of the fungi. 9th ed. / P.M. Kirk, P.F. Cannon, J.C. David, J.A. Stalpers – Wallingford, CAB International, 2001. – 655 p.
8. Li Fu Total phenolic contents and antioxidant capacities of herbal and tea infusions / Li Fu, Bo–Tao Xu, Ren–You Gan, Yuan Zhang, Xiang–Rong Xu, En–Qin Xia, Hua–Bin Li // International Journal of Molecular Sciences. – 2011. – Vol. 12 (4). – Р. 773–779.
9. Guthalu Puttaraju Nethravathi Antioxidant Activity of Indigenous Edible Mushrooms / Nethravathi Guthalu Puttaraju, Sathisha Uparrahali Venkateshiah, Shylaja Mallaiha Dharmesh, Shashirekha Mysore Nanjaraju Urs and Rajarathnam Somasundaram // J. Agric. Food Chem. – 2006. – 54 (26). – Р. 9764–9772.
10. Shivashankara K.S. Bioavailability of Dietary Polyphenols and the Cardiovascular Diseases / K.S. Shivashankara, S.N. Acharya // The Open Agric. Journal. – 2010. – Vol. 3. – Р. 227–241.
11. Peyrat–Maillard M.N. Determination of the antioxidant activity of phenolic compounds by coulometric detection / M.N. Peyrat–Maillard, S. Bonnely, C. Bersot // Talanta. – 2000. – V. 51. – Р. 709–716.
12. Halvorsen B.L. A systematic screening of Total Antioxidants in dietary plants / B.L Halvorsen., K. Holte M.C., W.Myrstad // J. Nutr. – 2002. – V. 132. – Р. 461–471.
13. Fedotov O.V. Wood–destroying fungi as bio–sources of ferments for medicinal and nutritional purposes / O.V. Fedotov – Plant and Microbial Enzymes: isolation, characterization and biotechnology applications – Tbilisi: Myza, – 2007. – Р. 125–126.
14. Wasser S.P. Medicinal mushroom Science: History, Current Status, Future Trends, and Unsolved problems / S.P. Wasser // Int. J. Med. Chem. – 2010. – 12 (1). – Р. 1–16.
15. Guo Ya–Jun Antioxidant capacities, phenolic compounds and polysaccharide contents of 49 edible macro–fungi / Ya–Jun Guo, Gui–Fang Deng, Xiang–Rong Xu, Shan Wu, Sha Li, En–Qin Xia, Fang Li, Feng Chen, Wen–Hua Ling and Hua–Bin Li // Food & Function. – 2012. – Vol. 8. – Р. 709–716.
16. Мусиенко М.М. Спектрофотометрические методы в практике физиологии, биохимии и экологии растений / М.М. Мусиенко, Т.В. Паршикова, П.С. Славный. – К.: Фитосоциоцентр, 2001. – 200 с.
17. Государственная Фармакопея СССР. – XI изд. – Вып. 1. – М.: Медицина, 1987. – 336 с.
18. Патент 2448721 Россия. Способ получения экстракта чаги / Кузнецов О.Ю., Сысоева М.А. Заявка № 2010124076/15 від 11.06.2010, МПК A61K36/06, B01D11/02 (2006.01), Бюл. № 12 від 27.04.2012.

**СКРИНІНГ ВМІСТУ ТА ДИНАМІКА НАКОПИЧЕННЯ ПОЛІФЕНОЛЬНИХ РЕЧОВИН У ДЕЯКИХ ВИДІВ БАЗИДІОМІЦЕТІВ**

**О. В. Федотов, А. К. Велигодська**

**Метою** роботи було вивчення загального вмісту поліфенольних речовин у карпофорах 50 видів базидіоміцетів з яких 27 вважаємо до порядку Polyporales та 23 – порядку Agaricales. Інтродуковано 23 штами 8 видів базидіальної грибів. **Методи.** Зібрані дикорослі карпофори висушували та подрібнювали до розміру часток 0,1±0,01 мм, а дослідні штами культивували поверхнево в колбах Ерленмейєра на стандартному глюкозо–пептонному живильному середовищі. Визначення загального вмісту поліфенольних речовин проводили у спицях з яких 27 належать до порядку Polyporales та 23 до порядку Agaricales. Інтродуковано 23 штами 8 видів біомаси карпофорів та міцелію визначали ваговим методом. Відомо, що поліфеноли є силинг провокаторами патогенних процесів. Методикою обліку росту і накопичення поліфенолів в міцелії та культуральних рідин проконту – чога, або в міцелії та культуральних рідинних видів біомаси карпофорів та міцелію визначали ваговим методом.

Відомо, що поліфеноли є силинг провокаторами патогенних процесів. Методикою обліку росту і накопичення поліфенолів в міцелії та культуральних рідинних видів біомаси карпофорів та міцелію визначали ваговим методом.

**Висновок.** Одержано редколегією визначення надзвичайної природи загального вмісту поліфенольних речовин у карпофорах 50 видів базидіоміцетів з яких 27 вважаємо до порядку Polyporales та 23 – порядку Agaricales. Інтродуковано 23 штами 8 видів базидіальної грибів.

**Ключові слова:** поліфеноли, базидіоміцети, карпофори, міцелій, культуральний фільтрат

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