Effects of different ingredients of zedoary on gene expression of HSC-T6 cells

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Abstract

Aim: To investigate the effects of four different ingredients of zedoary (Curcuma aromatica oil, Curcumol, β-elemence, and Curcumin) on the gene expressions of hepatic stellate cells (HSCs), and to explore the molecular mechanism of zedoary against hepatic fibrosis at gene network level.

Methods: We detected the mRNA sequences of 50 liver fibrosis-related genes in GenBank and designed oligonucleotide probes. We synthesized oligonucleotides with PE8909 DNA synthesizing instrument, and carried out oligonucleotide microarray with OGR-04 dropping platform to study the mechanism of herbal medicines as red sage root, zedoary, Chinese caterpillar fungus, hantianchiang A, extracts from peach seeds. Although many medicines can be used to treat liver fibrosis, effective medicines are still hard to find. Herbal medicines have the characteristics of multiple targets and poly-functioning routes, but genes in the organisms alone form a strict and complicated network. Therefore, study of herbal medicines should focus on the molecular mechanism at the level of genetic network based on the integral bio-system. Genetic chip technology is characterized by high communication, low consumption and miniaturization, thus providing a technological platform to study the mechanism of herbal medicines against liver fibrosis.

We found that zedoary could inhibit the proliferation of hepatic stellate cells (HSCs). Curcumin can be used to treat inflammation and tumors and Curcuma aromatica oil functions as an anti-inflammatory, anti-virus, anti-tumor and anti-thrombus agent. Now more than 20 chemical ingredients such as Curcumol, Epicurzerenone, β-elemence, Camphene, Isoborneol, Borneol, Cineole, and 4-methyl-pyrazine have been identified from Curcuma.
aromatic oil. It was reported that Curcumol and Elemence can function as anti-tumor and virus agents\(^{[5,6]}\). Xi et al\(^{[1]}\) found that zedoary could protect hepatic cells against necrosis and degeneration as well as proliferation of fibrous tissues.

To study the molecular mechanism of zedoary against liver fibrosis, we used the genetic expression spectrum chips to represent 50 genes related to liver fibrosis and substituted HSC-T6 for original HSCs, and investigated the effects of four different ingredients of zedoary (Curcuma aromatica oil, Curcumol, β-elemence, Curcumin) on the gene expression of activated HSCs.

### MATERIALS AND METHODS

**Materials**

Colchicine was purchased from American ALEXIS Co, Elemence injection and curcumin were obtained from Jingang Pharmacuetical Corporation Ltd, Dalian, China. Curcumol and Curcuma aromatica oil were from Pharmaceutical University, Shenyang, China. HSC-T6 was provided by the Institute of Hepatology, Shanghai University of Traditional Chinese Medicine and pharmacology. AXSys Probe Punctum-controlling software was purchased from Cartesian Technologies Co., and ImaGene 4.2 figure-analyzing softwares was from American Biodiscovery Co.

**Preparation of gene probes**

**Design of oligonucleotide probe** Oligonucleotide probes were designed by the design software of oligonucleotide probe. The coding region near the 3' end was selected for BLAST analysis. One or two probes related to liver fibrosis whose homology was less than 70% were used as spare probes.

**Synthesis of oligonucleotide** Oligonucleotides were synthesized by the chemical standard of standard subphosphorus imide using PE8909 DNA synthesizer. N-MMTr-6- ammonia-2-cyanogen-N and N-diisopropyl-subimide ammonia were modified by 5- or 3-amino-group.

**Preparation of probes** In brief, 0.5 µg/µL oligonucleotide probe was resolved into 3×SSC solution, glass chip was de-ion water for 10 min, and then dried for later use.

**Preparation of medical culture medium** Colchicine was dissolved in double-vaporing water to get the original solution (3.2 mg/mL). Curcumin was mixed with 950 mL/L alcohol to get the original solution (320 mg/L). Curcumol was mixed with 950 mL/L alcohol plus Tween-80 to get the original solution (3.2 mg/mL). Curcuma aromatica oil was mixed with Tween-80 (ratio, 1:10) plus 950 mL/L alcohol to get the original solution (2.5 mg/mL). Elemence injection (5 mg/mL) was used. All the medicines were filtrated through 0.45 micropores and stored at 4 °C.

**Culture of HSC-T6 cells**

The ampoule was taken out of the liquid nitrogen jar (wearing protective glasses and gloves) and put into a porcelain enamel vessel containing 36-37 °C water with shaking. The pocket was cut and the ampoule was taken out, sterilized with 700 mL/L alcohol. The cell suspension was aspirated and put into centrifuge tube, then 10 mL culture medium was added, centrifuged for 5 min at 500-1 000 r/min and rinsed. The culture medium was changed on the next day.

**Test of cytotoxicity (MTT assay)**

The 96-well plates were incubated at 37 °C until HSC-T6 cells were grown in a single layer. The culture medium was incubated for 48 h, and then 5 mg/mL MTT was added and incubated for 48 h. The A value of the solution was tested in enzymatic marking instrument (wave length on the light-filtrating slice is 492 nm). According to the experiment of cell toxicity, we took the appropriate concentrations of medicines that resulted in over 50% of cell survival. The formula of cell survival rate: cell survival rate (%) = (medicine group/control group)×100%.

**Incubation of HSC-T6 cells with medicines at different times**

HSC-T6 cells were incubated with different concentrations of Colchicine, Curcuma aromatica oil, Curcumol, β-elemence, and Curcumin, followed by collecting them at 1, 6, 12, and 24 h, respectively.

**Extraction and evaluation of cellular tRNA**

**Extraction of cellular tRNA** HSC-T6 cells were washed softly with germ-free PBS. One-milliliter of TRIzol reagent was used to blow the cells and to make them dissolve completely. Then 0.2 mL of methylene trichloride was added, followed by centrifugation at 12 000 g for 15 min at 4 °C. The supernatant was aspirated with 200 µL tip (dealt with DEPC) and moved to another EP tube. Then 0.5 mL of isopropylalcohol was added and put aside for 15 min, and centrifuged at 12 000 g for 10 min at 4 °C. The RNA was washed with 750 mL/L alcohol (dealt with DEPC), and then 1 mL of 750 mL/L alcohol (dealt with DEPC) was added, centrifuged at 7 500 r/min for 5 min at 4 °C, and stored at -20 °C.

**Evaluation of purity** DEPC water was added to 2 µL of RNA to make a total volume of 100 µL. Ultraviolet spectrophotometer was used to measure the value of \(A_{260}\) and \(A_{280}\) as well as \(A_{260}/A_{280}\). The formula of concentration of RNA: RNA (µg/µL) = \(A_{260}×40×\) dilution multiple/1 000.

**Evaluation of integrity** Five-microliter of RNA samples were put into 10 mL/L formaldehyde degeneration sepharose for cataphoresis, followed by painting with EB and examination under ultraviolet light.

**Reverse transcription system and the labeling and purification of cDNA**

**Reverse transcription system and conditions** Ten-microliter of TRNA (10 µg/µL), 0.3 µL of positive control, 4.0 µL of oligo (dT)16 (0.5 µg/µL) and 0.5 µL of
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RNA was put on 10 mL/L formaldehyde agarose gels for catalysis, and the results were recorded with gel photography. Three strips (28S, 18S, and 5S) can be seen in Figure 1A. After the analysis by software, the ratio of 28S and 18S was found to be between 1.5 and 2.0, showing that the RNA was integral and without degradation. Using ultraviolet spectrophotometer, the value of $\Delta_{260/280}$ was found to be between 1.7 and 2.0, showing that the RNA was pure and without protein pollution or phenol.

In addition, most of the cDNAs were observed between 0.5 and 2 kb (Figure 1B). Distribution of probes is shown in Table 1.

**Hybridization and wash**

Two microliters of probes labeled by fluorescence were diluted to 8 µL for hybridization, and the hybridization liquor was moved onto the cover glass chip by the capillarity between cover glass chip and carry sheet glass chip. Then the glass chips were put into a hybridization box and 5 µL of 3× SSC was added to keep the humidity. The probe washing temperatures varied according to the different probes, usually under room temperature. The order of washing liquor was lotion A–C.

**Analysis of data from fluorescence**

After hybridization, the genetic chips were scanned by Scanner Genepix 4 000B. ImaGene 4.2 was used to analyze the ratio of Cy3: Cy5 and the intensity of two kinds of fluorescence signals. Housekeeping gene and positive control were taken to balance the data of fluorescence of Cy3 and Cy5. Ratio of Cy3: Cy5 > 2 or < 0.5 was used to evaluate the differences of genetic expression.

### Table 1 Genetic probe matrix related to liver fibrosis

| Gene          | β-actin | β-actin | GAPDH | GAPDH |
|---------------|---------|---------|-------|-------|
| TIMP1         |         |         |       |       |
| TIMP1         |         |         |       |       |
| TIMP2         |         |         |       |       |
| TIMP2         |         |         |       |       |
| TIMP3         |         |         |       |       |
| TIMP3         |         |         |       |       |
| MMP2          |         |         |       |       |
| MMP2          |         |         |       |       |
| MMP2          |         |         |       |       |
| MMP8          |         |         |       |       |
| MMP8          |         |         |       |       |
| TGFβ1         |         |         |       |       |
| TGFβ1         |         |         |       |       |

### RESULTS

**Effects of different medicines on the growth of HSCs**

After treatment of HSC-T6 cells with different concentrations of Colechicine, Curcuma aromatica oil, Curcumin, β-elemence, and Curcumin for 48 h, along with deduction of the concentrations, the survival ratio of HSC-T6 increased.

**Infuence of four different ingredients of zedoary on the genetic expression of HSCs**

RNA was put on 10 mL/L formaldehyde agarose gels for catalysis, and the results were recorded with gel photography. Three strips (28S, 18S, and 5S) can be seen in Figure 1A. After the analysis by software, the ratio of 28S and 18S was found to be between 1.5 and 2.0, showing that the RNA was integral and without degradation. Using ultraviolet spectrophotometer, the value of $\Delta_{260/280}$ was found to be between 1.7 and 2.0, showing that the RNA was pure and without protein pollution or phenol.

In addition, most of the cDNAs were observed between 0.5 and 2 kb (Figure 1B). Distribution of probes is shown in Table 1.

**Hybridization**

In the hybridization scanning figures of probes (Figures 2A, 3A, and 4A), the green spots represent the locations of downregulated genetic expression. ImaGene 4.2 software was used to analyze the intensity and ratio of Cy3 and Cy5. After correction of housekeeping gene and positive
control, the two conditions were taken to evaluate the difference of genetic expression as follows: (1) the ratio of Cy3 and Cy5 >2 or <0.5; and (2) one of Cy3 and Cy5 >1 000. In the scattering diagram (Figures 2B, 3B, and 4B), the spots on the opposite angles show that the intensity of two groups (treatment group and control group) was the same. The farther they depart from the opposite angles, the bigger the difference of genetic expression would be. The different effects of medicines on genetic expression are shown in Tables 2-4. After culture of HSC-T6 cells in a medium containing 6.25 μg/mL of Colchicine for 12 h, the expression of TIMP-1 decreased 2.2-folds, which was in agreement with a previous report[9], suggesting that the genetic probe testing system in the experiment was dependable. Furthermore, Figures 3 and 4 stand for the scattering diagram and the scattering diagram of Curcuma aromatica oil group and Curcumol group, respectively, and the data of analysis are shown in Tables 3 and 4.
vascular endothelial growth factor B (VEGFB), vascular endothelial growth factor A (VEGFA), growth factor 1 (HGF1), hepatocyte growth factor 2 (HGF2), tumor necrosis factor α (TNFα), insulin-like growth factor 1 (IGF1), insulin-like growth factor 2 (IGF2), endothelin 1 (ET-1), intercellular adhesion molecule 1 (ICAM-1), vascular cell adhesion molecule 1 (VCAM-1), murine macrophage inflammatory protein 2 (MIP-2), monocyte chemotactic protein 1 (MCP-1), transforming growth factor β receptor I (TGFβRI), transforming growth factor β receptor II (TGFβRII), platelet-derived growth factor receptor α (PDGFRα), platelet-derived growth factor receptor β (PDGFRβ), tumor necrosis factor α receptor 1 (TNFRα1), platelet-activating factor receptor (PAF), and endothelin receptor A (ETRA). TGFβ1 is the strongest factor which promotes the synthesis of extracellular matrix (ECM). PDGF can accelerate the proliferation of HSCs; (2) expressing MMPs (matrix metalloproteinases) and TIMPs (tissue inhibitor of metalloproteinases), such as MMP2, MMP3, MMP8, TIMP-1, TIMP-2, and TIMP-3, which take part in the degradation of ECM; (3) gene expression related to the preliminary activation of HSCs, such as c-myc, Ets-1, STAT1, c-jun B, c-jun D; (4) involving in gene expression related to the biological oxidation[14], such as cytokrome P450d, cytokrome P450a, cytochrome P450e, cytochrome P450-LA, and cytochrome P450 (4A3), CYP1B1, CYP2D4; and (5) house keeping gene, such as β-actin and GAPDH.

How to choose the genes related to liver fibrosis is the key of the design of oligonucleotide probes. The mRNA sequences which contain 40 amino acids are selected and then oligonucleotide probes are designed using the design software of oligonucleotide probe. By and large, the length of oligonucleotide probe is 15-80 nt, the content of GC is 45-55%. Probes of the coding region approaching end 3 are chosen for BLAST analysis. One to two probes whose homology of sequences was less than 70% are chosen as gene distinctive oligonucleotide probes.

Because some sequences of mRNA related to collagen I, III, IV are so short, the probes designed through software are easy to cause cross reactions which may result in false positive results, and such genes, therefore, cannot be chosen. However, the aforementioned 50 genes, in general, can represent approximately the changes which take place in the process of preliminary and persistent activation of HSCs in liver fibrosis.

HSC-T6 cells are SV40 transfected HSCs of Sprague-Dawley rats. The cells can be steadily cultured and their phenotype is activated HSCs which can express high-level collagen I and TIMP-1 mRNA, etc. In our study, HSC-T6 was substituted for the original HSC. Yin et al.[15,16] took HSC-T6 as model cells to investigate the influence of compound 861 on the gene expression of MMP-3 and TIMP-1 and they found that 0.25, 0.5, and 1.0 mg/mL of compound 861 could increase the expression of MMP3 and inhibit the expression of TIMP1.

TIMPs can prevent MMPs from degrading ECM[17-21]. Using genetic chip technology, we found that after HSC-T6 cells were cultured in a medium containing 6.25 μg/mL...
Colehicine for 12 h, the expression of TIMP-1 decreased 2.2-folds. Expression of TIMP-2 decreased 2.3-folds after HSC-T6 cells were cultured in a medium containing 78.125 μg/mL Curcuma aromatica oil for 24 h. These results showed that Curcuma aromatica oil, like Colehcinics, can also inhibit the expression of TIMPs and reduce the inhibition of MMPs which, in turn, help MMPs to degrade ECM. This may be one of the mechanisms for zedoary against liver fibrosis.

IL-6 is also called hepatic cell stimulating factor which can directly stimulate hepatic cells proliferation, induce the expression of IL-6 receptors in Liver, and stimulate fibroblastic cells to synthesize collagens. In our study, 78.125 μg/mL Curcuma aromatica oil could decrease the expressions of IL-6, TIMP1 and other genes related to hepatic fibrosis, thereby enhancing MMPs to degrade ECM. It might be another mechanism of zedoary against hepatic fibrosis.

TGFβ1 plays an important role in hepatic fibrosis which can activate HSCs and, as a transcription factor of collagen, accelerate its expression. In our study, the expression of TGFβ1 decreased 2.3-folds after culture of HSC-T6 cells in a medium containing 1.5625 μg/L Curcumol for 12 h, suggesting that Curcumol can inhibit the synthesis of ECM through the inhibition of TGFβ1 which might be an important mechanism of zedoary against hepatic fibrosis.

Cytochrome P450 (or CYP450) involves in the synthesis of steroid hormone, bile acid and bile pigments as well as the process of the bio-transformation of medicine and poison. Yang et al. have shown that lipid peroxidation takes part in the activation of HSCs. Svegliati Baroni et al. have indicated that HCM/Fe might induce a significant increase in collagen type I accumulation in HSC culture media, and HSCs proliferation may be associated with changes in the Na+/H+ exchanger activity. Nicto et al. transfected CYP2E1 to HSC-T6 cells, and found that there was an increase in the level of reactive oxygen species and type I collagen mRNA. It has been reported that when HSCs were cultured with HepG2 cells which overexpress CYP2E1 together, the level of collagen markedly increased, suggesting that the solved oxidants can activate HSCs. In our study, we found that after culture of HSC-T6 cells in a medium containing 1.5625 μg/mL Curcumol for 12 h, the expression of P450α decreased 2.1-folds, suggesting that the metabolism of Curcumol in HSC-T6 cells might bring forth oxidation-conjugation reaction through the P450 enzyme system, induce a decline of oxidative stress and lipid peroxidation, and thus inhibit the activation of HSCs. This may be one of the mechanisms of zedoary against hepatic fibrosis.

In conclusion, two different ingredients of zedoary (Curcuma aromatica oil, Curcumol), when treated with HSC-T6 cells for 24 and 12 h, can decrease the expression of TIMP-2, IL-6, TGFβ1 and P450α by different degrees, indicating the molecular mechanisms of zedoary against hepatic fibrosis at gene network level. But the changes of genes and the expression of proteins might not be the same events. Along with the development of protein histology, further research needs to testify the proteins related to liver fibrosis, for example, to examine the content of type I collagen using ELISA or investigate the influence of Curcuma aromatica oil and Curcumol on protein expression of HSC-T6 cells by protein chip technology.

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