Anisakidae in fishing products sold in Sicily

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Abstract

One of the parasite diseases associated with the consumption of raw fish that occurs with some frequency is the anisakiasis, a human disease caused by the accidental ingestion of larval nematodes of the genus Anisakis, family Anisakidae. At the National Reference Centre for Anisakiasis (C.Re.N.A.) from October 2012 to February 2013, a number of 231 bony fish (Trichiluriidae, Engraulidae, Scombridae and Clupeidae) were received from different fishing sites in Sicily. Anisakis pegreffii is the main species detected in fish, as identified by molecular analysis based on polymerase chain reaction-restriction fragment length polymorphism, while Anisakis simplex sensu stricto was found only in Scomber scombrus caught in the Mediterranean Sea (Fishing Areas 37), in the Spanish coast (Fishing Areas 37) and in the Atlantic Ocean (Fishing Areas 34). Larvae of the genus Pseudoterranova were found only in fish caught in the Norwegian Sea.

Introduction

In the last ten years, in our country, there has been an increase in the consumption of raw fish due to the introduction of new eating habits from countries outside Europe (sushi, sashimi, herring, pickled anchovies, etc.), increasing the risk of contracting parasitic diseases, in particular the anisakiasis (Audicana et al., 2002). Anisakiasis is a disease caused by nematodes of the genus Anisakis, belonging to the Anisakidae family, with Pseudoterranova, Contracaecum and Hysterobothrium genus. Human infestation occurs by ingestion of thirdstage Anisakis larvae present in raw fish and cephalopods but also undercooked, marinated, pickled, smoked or salted. These parasites are found, at the adult stage, in the abdomen of marine mammals (whales, seals, dolphins), more precisely in the stomach where they are visible to the naked eye. In fish, which are intermediate hosts, the larval form are normally found in the coelomic cavity where they can found free or encysted or adherent to the various organs and tissues (Mattucci and Nascetti, 2008). In the past you may encounter traumatic injuries, and fibrinous adhesions due to the presence of the parasite that passes from the gastrointestinal tract to the coelomic cavity to reach the flesh. In humans the anisakiasis may manifest as an acute form with a predominantly gastric localisation, characterised by nausea, vomiting and epigastric pain that occur after 4-6 h after ingestion of fish parasitised. We distinguish localised forms where you can discover granulomas in the submucosa, with the presence of eosinophils (eosinophilic granulomas) and diffuse forms characterised by an edematous infiltration that affects the entire gastric or intestinal wall called eosinophilic Phlegmon (Mattucci et al., 2011).

It is also an intestinal form that develops after about 7 days post infection with abdominal pain, nausea, vomiting, fever, diarrhea and fecal occult blood. The parasites, however, introduced into the body, induce an increase in immunoglobulins of class E (IgE) with IgE-mediated reactions such as urticaria, angioedema, anaphylaxis, asthma (Urban et al., 1992). The finding of a positive specific IgE in a certain percentage of asymptomatic subjects and in patients with urticaria, angioedema or anaphylaxis triggered by foods not parasitised, has led to speculation that there may be an initial sensitisation to Anisakis, asymptomatic, with the appearance of new symptoms after exposure to allergens (Mattucci et al., 2013; Audicana and Kennedy, 2008; Daschner et al., 2012). Because of all these implications on human health, the Italian legislation has put in place, in 1992 an initial regulatory approach of the Health Ministry Circular No. 10 of 11/03/1992 and subsequent Ordinance 12/05/1992, which recommended to the fishermen and operators involved in the handling of fresh fish to provide for a timely gutting for fish with size exceeding 18 cm belonging to the species Lepidopus caudatus, Engraulis encrasicholus, Scomber scombrus, Merluccius merluccius, Sardina pilchardus, Bramidae brama. Fish or parts thereof that clearly have parasites should not be placed on the market. More recently, the European Community, with the enactment of the so-called Hygiene Package and in particular with Regulation (UE) No. 1276/2011 defines the following: Food business operators placing on the market the following fishery products derived from fish or cephalopod molluscs: (a) fishery products intended to be consumed raw; or (b) marinaded, salted and any other treated fishery products, if the treatment is insufficient to kill the viable parasite. Must ensure that the raw material or finished product undergo a freezing treatment in order to kill viable parasites that may be a risk to the health of the consumer. For parasites other than trematodes the freezing treatment must consist of lowering the temperature in all parts of the product to at least: (a) -20°C for not less than 24 h; or (b) -35°C for not less than 15 h (European Commission, 2011).

In addition, with the spread of fashion consumption of raw fish, the Ministry of Health with prot. n. 4379 of 17/02/2011 specifies that, as foreseen in the Regulation (EC) n. 1020/2008, also in retail must be complied with the specific requirements of parasites and those relating to the visual examination for their research in accordance with the provisions in Regulation (EC) No 853/2004. According to the above note, the food business operator if it were to carry out operations of filleting and/or slicing should submit the same to visual inspection as required by Regulation (EC) No 2074/2005. If the OSA would produce raw or almost raw fishery products using chilled, he must demonstrate that the treatments used, whatever they are, ensure the killing of all parasites that may be present in the product ready for consumption. To give support to the emerging problems regarding the presence of Anisakis in fish products and to carry out the monitoring of the Italian seas has been activated the National Reference Centre for Anisakiasis (C.Re.NA) at headquarters of Institute Zooprofilactic Sperimental of Sicily (IZSSI). In the present study larval forms of
Anisakidae collected in fishing products sampled in Sicily were identified by molecular analysis.

Materials and Methods

This study was conducted on the samples received at the Center from October 2012 to February 2013. In this period were analysed 231 fish (Lepidopus caudatus, Engraulis encrasicolus, Merluccius merluccius, Sarda pilchardus, Scomber scombrus, Longing longing, Trachurus trachurus, Mullus surmuletus), sampled caught in Sicilian seas (Fishing Areas 37), parasitised by a total of 1299 larvae of the Family Anisakidae. 157 of them (67.96%) were parasitised by larvae of the Anisakidae family while 74 (32.03%) were not parasitised. The larvae, taken from viscera (n. 1296), organs and muscles (n. 3) of fish were counted with the help of the stereo microscope and identified at genus level by light microscopy. The Anisakis spp. larvae recovered and identified were then preserved in 70% ethanol until DNA extraction, the DNA and muscles (n. 3) of fish were counted with the help of the stereo microscope and identified at genus level by light microscopy. The Anisakis spp. larvae recovered and identified were then preserved in 70% ethanol until molecular analysis based on polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP). In this molecular technique, after the DNA extraction, the DNA regions of ITS1, ITS2 and 5.8 SrRNA was amplified by PCR using the primer pair NC5/NC2 (Zhu et al., 1998) and the amplified was subjected to enzymatic restriction using the enzymes HinI and Hha previously shown to be diagnostic for the species of Anisakis.

Different species of Anisakis give rise to restriction fragments that allow the species identification (Anisakis pegreffii, Anisakis physeteris, Anisakis typica, Anisakis simplex C, Anisakis ziphidarium and Anisakis simplex sensu strico) (D’Amelio et al., 2000). The DNA was also purified and subjected to sequencing to confirm the identification through homology with the sequences found in GeneBank. Several larvae were collected from fish caught in the Norwegian Sea and morphologically identified at genus level by light microscopy as Pseudoterranova.

Results and Discussion

Of the 231 fish received, 1299 larvae were collected and were identified by microscop-