Effect of Hormonization Treatment on Yield Quantity and Quality, Contents of Biologically Active Compounds, and Antioxidant Activity in ‘Einset Seedless’ Grapevine Fruits and Raisins

Magdalena Kaplan ¹, Kamila Klimek ²,*, Ewa Jabłońska-Ryś ³,*, Aneta Slawińska ³ and Anna Stój ⁴

1 Institute of Horticulture Production, University of Life Science, 28 Głęboka Street, 20-612 Lublin, Poland; magdalena.kaplan@up.lublin.pl
2 Department of Applied Mathematics and Informatics, University of Life Science, 28 Głęboka Street, 20-612 Lublin, Poland
3 Department of Plant Food Technology and Gastronomy, University of Life Science, 8 Skromna Street, 20-704 Lublin, Poland; ewa.jablonska-rys@up.lublin.pl (E.J.-R.); aneta.slawinska@up.lublin.pl (A.S.)
4 Department of Biotechnology, Microbiology and Human Nutrition, Faculty of Food Science and Biotechnology, University of Life Sciences in Lublin, 8 Skromna Street, 20-704 Lublin, Poland; anna.stoj@up.lublin.pl
* Correspondence: kamila.klimek@up.lublin.pl; Tel.: +48-734-858-363

Abstract: In this study, we determined the effect of hormonization treatment on yield quantity and quality, content of biologically active compounds, and antioxidant activity in fruits and raisins of ‘Einset Seedless’ grapevine. Field studies were conducted in 2017 at Nobilis Vineyard (50°39’ N; 21°34’ E) in the Sandomierz Upland. Analytical studies were carried out in the Laboratory of the University of Life Sciences in Lublin. Hormonized fruits and raisins, which were dried at 40 °C in a food dryer for 7 days, were the experimental material. It was shown that the application of the hormonization treatment had a significant effect on yield size and quality. The hormonization treatment and the form of plant material analyzed had a significant effect on the content of biologically active compounds and the antioxidant activity in ‘Einset Seedless’ grapevine fruits and raisins. The concentration of applied gibberellic acid had a significant effect on the levels of acidity, content of anthocyanins, and antioxidant activity determined with the FRAP and DPPH methods. The application of the multivariate analysis technique showed that, in the fresh fruits and raisins, the level of biologically active compounds and antioxidant activity in the case of the 200 mg GA₃ L⁻¹ concentration and in the control combination was similar but differed significantly in the case of the 300 mg GA₃ L⁻¹ application.

Keywords: seedless grape; gibberellic acid (GA₃); yield; FRAP; DPPH; anthocyanins; correlation

1. Introduction

Of all the grapes produced in the world, 80% of the harvest is used to make wine, 13% are dessert grapes, and the rest are used to make raisins [1]. A prerequisite for the profitability of dessert grape production is to adapt the size of its fruits to the high demands of the market, i.e., to obtain the best quality. An important attribute of dessert grapes is also the absence of seeds [2].

Of all the seedless cultivars known in Poland and recommended for outdoor cultivation, ‘Einset Seedless’ is the best, mainly due to its prolificacy, frost resistance, and berry flavor. The fruits of this variety can be used for raisin production and fresh consumption as dessert grapes. However, the natural berry weight of ‘Einset Seedless’ (±2 ÷ 3 g) is not sufficient for commercial use as table grapes [3].

In recent years, new agrotechnical possibilities have emerged to produce very good-quality fruits of these cultivars through exogenous application of gibberellic acid, which
stimulates cell division, accelerates flowering, and increases fruit size [4–6]. Many studies have shown that the use of gibberellins in parthenocarpic fruit production is very effective [7–12]. This is very important, because customers expect grapes that are uniform and reproducible, have equal size and shape and uniform coloration of all berries, and have increased resistance during transport. In addition to improving the yield size and quality of parthenocarpic grapevine varieties, gibberellic acid applications significantly influence berry hardness and elasticity [13]. GA$_3$-treated grapes are more resistant to cracking caused by rain, especially at harvest time. Conscious application of gibberellic acid can optimize the yield of seedless grape cultivars by increasing fruit size and improving grape structure. The effectiveness of the treatment itself largely depends on the timing of the treatment, taking into account the developmental stage of the flowers or buds, the concentration of gibberellic acid applied, and the conditions after application [14].

To increase berry and cluster size and compactness, gibberellic acid has been used for many years in countries with long cultivation traditions: in India [15], USA [16,17], Thailand [18], Brazil [19,20], Greece [21], Chile [22], Spain [23], Poland [6,11,12,24], and Jordan [25]. Despite many experiments and studies, there are no clear guidelines on the dose and number of applications of this compound.

Due to its taste and dietetic properties, grapevine fruit is a valuable source of biologically active substances, i.e., vitamins (A, B1, B2, C, PP), minerals (potassium, phosphorus, calcium, iron, boron, magnesium), pectins, dyes, tannins, oils, easily assimilable carbohydrates, amino acids, fruit acids, and fiber. The most important group of health-promoting compounds present in grapes comprises polyphenols: flavonoids, phenolic acids, flavones, flavonols, flavanones, catechins, and anthocyanin pigments [26–29].

Grape consumption significantly reduces the risk of cardiovascular disease and cancer [30–32]. This is due, among other things, to the presence of secondary metabolites such as polyphenols, which have antioxidant, anti-inflammatory, anticancer, antiplatelet, neuroprotective, vasodilator, and immune system-enhancing effects. This is related to the ability of polyphenols to modulate and induce signaling pathways [27,33–38]. Furthermore, polyphenols inactivate free radicals and chelate divalent metal ions, thereby reducing their oxidative potential [39].

The qualitative and quantitative composition and distribution as well as the antioxidant activity of polyphenols in grapes are highly variable and depend on the species, cultivar, location in the berries (skin, pulp, seeds, juice), climate and soil conditions in which the bush grows (light exposure, temperature, soil type), agronomic treatments (irrigation, fertilization, use of growth regulators), harvest date, berry maturity, yield and berry size, postharvest conditions, and storage and processing techniques [37,40–44]. Polyphenols are responsible for the main sensory characteristics of plant-derived products and beverages, determining their appearance, especially color, and flavor properties, such as bitterness, astringency, and aroma [45,46].

The aim of this study was to determine the effect of hormonization on yield quantity and quality, the content of biologically active compounds, and antioxidant activity in ‘Einset Seedless’ grapevine fruits and raisins.

2. Materials and Methods

Field studies were conducted in 2017 at Nobilis Vineyard (50°39’ N; 21°34’ E) in the Sandomierz Upland. The field experiment was set up in a randomized block design, including 3 combinations with five replications, comprising plots of five grapevines each. Parthenocarpic vines of the ‘Einset Seedless’ cultivar (‘Fredonia’ × ‘Canner’ [47]) were planted in the spring of 2003 at a spacing of 2.0 × 1.0 m (5000 units·ha$^{-1}$) on a loess-derived podzolic soil containing 2.1% of organic matter. The shrubs were managed with a scaffold consisting of poles and four metal wires stretched at heights of 70, 110, 150, and 190 cm. The plants were managed as a single Guyot twine with a 40 cm high trunk, a single approximately 0.9 m long bed, and a single two-eyed pivot. During the experiment, regular protection against diseases, pests, and weeds was carried out according to the current
The vines were not irrigated, and the soil pH was 6.2. At the budburst stage, Hydrocomplex fertilizer (12N-11P-18K) was applied by sprinkling at a dose of 300 kg·ha$^{-1}$, and the other macro- and microelements were supplemented by foliar application as needed. In the field experiment, the effect of gibberellic acid concentration (GA$_3$) on the quantity and quality of ‘Einset Seedless’ grapevine fruit yield was evaluated. The solution was prepared using 99% gibberellic acid (Acros OrganicsTM, Thermo Fisher Scientific, Geel, Belgium) and the SILWET GOLD (Chembura Europe Limited, Warsaw, Poland) wetting agent and adhesion promoter at a concentration of 0.015%. The control consisted of plants whose clusters were not treated with gibberellic acid. The solution was prepared just before the treatment. Inflorescences were sprayed with a hand sprayer in the morning, thoroughly covering the stems and flowers. On average, 50 mL of the prepared solution was sufficient to cover all the clusters on one bush very thoroughly. The solutions were applied at concentrations of 200 or 300 mg·L$^{-1}$. The treatment was performed once during full flowering (70–80% of developed flowers). The level of gibberellic acid concentrations was selected based on the results of our previous research.

In the experiment, the quality of the crop was assessed by analyzing the following parameters: number and weight of bunches, total yield per 1 ha, weight of berries, number of berries per cluster, length of clusters, and length and width of berries. The mean weight and length of clusters were determined by weighing and measuring 15 typical clusters, with 5 clusters randomly taken from each plant. The average weight, number, length, and width of the berries were determined by weighing, counting, and then measuring the berries from five average-sized clusters from each replicate.

The study was conducted in 2017 in the Laboratory of the Department of Plant Food Technology and Gastronomy, Faculty of Food Science and Biotechnology, University of Life Sciences in Lublin. Grapes of the ‘Einset Seedless’ cultivar (‘Fredonia’ × ‘Canner’ [47], subjected to hormonal treatment with gibberellic acid GA$_3$ were the experimental material.

2.1. Drying

The samples of the grapes were dried at 40 °C in a food dehydrator (Ezidri Ultra FD 1000, Liberec, Czech Republic) for 7 days.

2.2. Total Acidity, Extraction, and Dry Weight Content

Acidity was measured by titration with 0.1 N NaOH according to PN-EN 12147:2000 [48]. The measurements were taken with a class A burette. The level of total titratable acidity was expressed as tartaric acid. Soluble solids were measured using a DR201-95 refractometer (Kruss, Germany). The results were reported as °Bx at 20 °C (PN-A-75101-02:1990/Az1:2002 [49]). The dry weight was determined by drying at 70 °C until constant weight was achieved according to PN-A-75101-03:1990 [50].

2.3. Extraction Procedure

The extraction procedure was based on the method described by Kaczmarska et al. [51], with some modifications. One gram of homogenized plant materials (grapes or a 1:1 w/w mixture of dried raisins and water) was mixed with 5 mL of 80% (v/v) ethanol. The slurry was shaken for 30 min at room temperature at a speed of 200 rpm. The obtained extracts were centrifuged (MPW 350-R) at 3755×g for 15 min. Supernatants were used for the determination of antioxidant activity and total phenolic content.

The extracts for anthocyanin determination were prepared as follows. Mixed grapes or raisins (2 g) were extracted with 10 mL of 80% ethanol acidified with HCl (pH 2) and sonicated for 20 min (Grant, Royston, UK). After centrifugation (4200×g for 20 min) (MPW-350R, MPW, Warszawa, Poland), supernatants were concentrated by a rotary evaporator (Laborota 4000 Efficient; Heidolph, Schwabach, Germany) and dissolved in 10 mL of acidified ethanol.
2.4. Determination of Total Phenolic Content

The measurement of total phenolics was carried out according to the methodology developed by Singleton and Rossi [52]. The extracts (0.2 mL) were mixed with tenfold diluted Folin and Ciocalteu reagent (0.8 mL). The mixtures were incubated for 3 min at room temperature. Then, 1.25 mL of 7% Na₂CO₃ was added, and the mixtures were left to stand for 30 min in the dark at room temperature. The absorbance was read at 725 nm (Helios Gamma; Thermo Fisher Scientific, Waltham, MA, USA) against a blank sample. The phenolic content was expressed as gallic acid equivalent (GAE) per 100 g of fruit fresh mass (FW).

2.5. Determination of Ferric Reducing Antioxidant Power (FRAP)

The ferric reducing antioxidant power of the fruit extracts was determined using a FRAP assay following the method proposed by Benzie and Strain [53]. The FRAP reagent was prepared by mixing 300 mM acetate buffer at pH 3.6 with 2,4,6-tri(2-pyridyl)-1,3,5-triazine (TPTZ) (Sigma-Aldrich, St. Louis, MO, USA) solution (10 mM TPTZ in 40 mM HCl) and a 20 mM FeCl₃·6H₂O solution at a 10:1:1 ratio. Assay solutions were prepared by mixing 1.9 mL of the FRAP reagent with 0.1 mL of the fruit extract. After 15 min of incubation (water bath W610; LABOPLAY, Bytom, Poland) at 37°C in the dark, the absorbance was measured at 593 nm using a Helios Gamma apparatus (Thermo Fisher Scientific, Waltham, MA, USA). The results were reported as µmol of Trolox equivalents (TE) per 1 g of fruit fresh mass (FW).

2.6. Determination of DPPH Radical-Scavenging Activity

The DPPH (2,2-diphenyl-1-picrylhydrazyl) radical-scavenging activity was assayed following the method developed by Choi et al. [54]. Each 0.2 mL of the ethanolic extract was mixed with 0.8 mL of a 0.2 mM DPPH ethanolic solution. The mixture was shaken (TK3S; Kartell, Noviglio, Italy) for 15 s and left to stand for 15 min in the dark. The absorbance was measured at 520 nm using a Helios Gamma apparatus (Thermo Fisher Scientific, Waltham, MA, USA). The antioxidant activity was expressed as µmol of Trolox equivalents (TE) per 1 g of fruit fresh mass (FW).

2.7. Determination of Total Anthocyanin Content

Total anthocyanin content (TA) was determined using the pH differential method proposed by Wrolstad [55] with some modifications. Then, 250 µL of the extract was diluted with 1 mL of two different buffers: 0.025 M potassium chloride pH 1.0 and 0.4 M sodium acetate pH 4.5. After 15 min of incubation at room temperature, the absorbance (A) was measured at 526 and 700 nm (Helios Gamma; Thermo Fisher Scientific, Waltham, MA, USA). The results were calculated as follows:

\[ A = \left( A_{526} - A_{700} \right) \text{pH 1.0} - \left( A_{526} - A_{700} \right) \text{pH 4.5} \]

The content of total anthocyanins (TA) was calculated as follows:

\[ TA = \left( A \times MW \times DF \times 1000 \right) / (\varepsilon \times P), \]

where A is absorbance, DF is the dilution factor, ε is the molar absorptivity coefficient (20,200 cm⁻¹ mg⁻¹ for malvidin 3-glucoside–M3G) [56], MW is the molecular weight (493.2 for M3G), and P is the cuvette path length (1 cm). The total anthocyanin content was expressed as mg M3G 100 g⁻¹ of the fresh weight of grapes or raisins.

All the measurements were taken in triplicate.

The results obtained in this test were statistically analyzed using two-factor analysis of variance and Tukey confidence intervals. Inference was based on a significance level of \( p < 0.05 \). Correlations between qualitative grape parameters were estimated by counting Pearson correlation coefficients. Multivariate analysis techniques were applied to show similarities within groups such that homogeneous objects were in grouped one cluster. The
results of cluster analysis were summarized using a dendrogram. All statistical analyses were performed using SAS Enterprise Guide 5.1 software.

3. Results and Discussion

On average, the number of clusters per bush, irrespective of the combination used, was over 19 and did not differ significantly between each other (Table 1).

Table 1. Effect of gibberellic acid on the size yield of ‘Einset Seedless’ grapevine.

| Combination | Cluster Vine⁻¹ | Yield kg Vine⁻¹ | Cluster | Berries Cluster⁻¹ | Berry Weight, g |
|-------------|-----------------|-----------------|---------|-------------------|----------------|
| Control     | 19.30 ± 0.25 A⁺ | 2.34 ± 0.22 C   | 121.2 ± 12.24 C | 65.2 ± 6.04 B   | 1.86 ± 0.11 B |
| 200 mg GA₃ L⁻¹ | 19.28 ± 0.16 A  | 2.58 ± 0.13 B   | 133.6 ± 7.58 B  | 67.5 ± 2.51 A⁺  | 1.98 ± 0.08 A⁺ |
| 300 mg GA₃ L⁻¹ | 19.26 ± 0.15 A  | 2.76 ± 0.16 A   | 143 ± 5.73 A⁺   | 69.4 ± 2.79 A   | 2.06 ± 0.08 A⁺ |
| p-value     | 0.5203          | <0.0001         | <0.0001         | 0.0048           | <0.0001         |

* Mean values marked with the same letters do not differ significantly at p > 0.05; ns—not significant. A B C is an indication and the difference between averages.

The gibberellic acid applications had a significant effect on the grapevine yield. The GA₃ concentration significantly modified the yield, which increased as the GA₃ concentration increased. Similar relationships were shown by Dimovska et al. [2], who evaluated ‘Flame Seedless’ (Vitis vinifera L.) after application of 5, 10, and 20 mg·GA₃·L⁻¹.

The hormonal treatment had a significant effect on the grape weight, as the gibberellic acid-treated grapes were significantly heavier than the control. A significant effect of the gibberellic acid concentration on the assessed quality parameter was found, as the clusters treated with 200 mg·GA₃·L⁻¹ were significantly heavier than those treated with 300 mg·GA₃·L⁻¹. A beneficial effect of gibberellic acid on cluster weight was shown by Habrooks et al., Lu, Casanova et al. and Abu-Zahra [16,17,23,25].

The exogenous applications of gibberellic acid had a beneficial effect on the number of berries per cluster; this effect in the case of application of 300 mg·GA₃·L⁻¹ was significant. There was no significant effect of the applied concentration on the trait studied.

The weight of one berry ranged from 1.86 to 2.06 g and differed significantly among the studied combinations (Table 1). The statistical analysis showed a significant effect of the hormonization treatment on the studied trait, while there was no significant effect of the concentration of applied gibberellic acid on the weight of one berry. This is consistent with the results reported by other researchers, e.g., Habrooks and Mortensen [16], Zabadal and Dittmer [57], and Casanova et al. [23], who showed that gibberellic acid application exerted a significant effect on berry weight. A negative effect of GA₃ applied at doses of 50 and 300 mg·GA₃·L⁻¹ was shown by Lu [17].

The hormonization treatment significantly influenced the size of grapes, which was reflected in the length and width of the cluster (Table 2). The concentration of gibberellic acid had a significant effect on the grape length, whereas no such relationship was shown in the case of the grape width.

Table 2. Effect of gibberellic acid on the quality of yield of ‘Einset Seedless’ grapevine cultivar.

| Combination     | Cluster | Berry |
|-----------------|---------|-------|
|                 | Length, cm | Width, cm | Length, mm | Width, mm |
| Control         | 15.32 ± 0.08 A⁺ | 9.10 ± 0.29 B  | 17.88 ± 0.22 C | 15.28 ± 0.43 C |
| 200 mg GA₃ L⁻¹ | 15.92 ± 0.78 B  | 9.76 ± 1.15 A  | 18.74 ± 0.18 B  | 15.6 ± 0.46 B  |
| 300 mg GA₃ L⁻¹ | 16.68 ± 1.01 A   | 10.58 ± 1.62 A⁺ | 19.58 ± 0.82 A⁺ | 15.98 ± 0.37 A⁺ |
| p-value         | <0.0001       | <0.0001       | <0.0001       | <0.0001       |

* Mean values marked with the same letters do not differ significantly at p > 0.05; ns—not significant. A B C is an indication and the difference between averages.
The obtained results indicate that the hormonization treatment had a significant effect on the length and width of the 'Einset Seedless' grape berries, as the gibberellic acid-treated clusters formed significantly longer and wider berries than the control (Table 2). The concentration of gibberellic acid had a significant effect on the studied quality parameters.

Dimovska et al. [2] evaluating the above parameters obtained different results at most of the GA3 concentrations applied.

The drying efficiency of the 'Einset Seedless' fruits differed significantly between the evaluated combinations (Figure 1). It was shown that the evaluated parameter in the control grapes (24.48%) and in grapes treated with gibberellic acid at the dose of 200 mg GA3 L\(^{-1}\) (24.53%) did not differ significantly between each other and was significantly higher than in fruits treated with 300 GA3 L\(^{-1}\) (23.72).

![Figure 1. Drying efficiency of 'Einset Seedless' fruit.](image)

Table 3 presents a two-way analysis of variance. The first example (A) concerns the effect of the GA3 concentration level, on the assessed parameters, regardless of the type of raw material, and in case (B), the impact of the type of plant material was considered at the level of the parameters assessed, regardless of the combination used. This combination was considered the mean of the controls, 200 mg GA3 L\(^{-1}\) and 300 mg GA3 L\(^{-1}\). The hormonization treatment significantly reduced the dry matter of the 'Einset Seedless' grapevine fruits. There was no significant effect of the concentration of gibberellic acid on the studied trait. The fresh fruits had significantly lower dry weight than after drying (Table 3, Figure 2).

### Table 3. Effect of the hormone treatment on the content of biologically active compounds and antioxidant activity in 'Einset Seedless' fruits and raisins.

| Concentration (A) | Dry Mass * | Total Acidity | Extract | Total Phenolic Content | FRAP | DPPH | Anthocyanin |
|-------------------|------------|---------------|---------|------------------------|------|------|------------|
| Control           | 53.32 ± 31.93 \(^{A}^{\text{**}}\) | 1.30 ± 0.66 \(^{B}\) | 49.19 ± 30.16 \(^{A}\) | 61.79 ± 38.71 \(^{A}\) | 2.95 ± 1.49 \(^{A}\) | 1.35 ± 0.61 \(^{A}^{B}\) | 18.69 ± 14.91 \(^{A}\) |
| 200 mg GA3 L\(^{-1}\) | 52.16 ± 31.71 \(^{B}\) | 1.34 ± 0.66 \(^{B}\) | 46.22 ± 27.97 \(^{B}\) | 62.67 ± 42.16 \(^{A}\) | 2.89 ± 1.53 \(^{A}\) | 1.38 ± 0.68 \(^{A}\) | 25.69 ± 21.86 \(^{A}\) |
| 300 mg GA3 L\(^{-1}\) | 52.17 ± 31.16 \(^{B}\) | 1.45 ± 0.75 \(^{A}\) | 47.25 ± 28.69 \(^{B}\) | 55.83 ± 40.06 \(^{A}\) | 2.38 ± 1.31 \(^{B}\) | 1.17 ± 0.63 \(^{B}\) | 16.09 ± 13.83 \(^{B}\) |

| Type of plant material | p-value | | | | | | |
|------------------------|---------|------------|---------|------------------------|------|------|------------|
| Grapevine fruits       | <0.0001 | 0.0048     | 0.0006  | <0.0001               | 0.0008 | 0.0328 | <0.0001   |
| Raisins                |         |            |         |                        |       |      |            |

* Dry mass %, Total acidity (g tartaric acid 100 g\(^{-1}\) FW), Extract (°Bx), Total phenolic content (mg GAE 100 g\(^{-1}\) FW), FRAP (μmol TE g\(^{-1}\) FW), DPPH (μmol TE g\(^{-1}\) FW), Anthocyanin (mg MSG 100 g\(^{-1}\) FW), Mean values marked with the same letters do not differ significantly at p > 0.05, ns—not significant. A B is an indication and the difference between averages.
Grape acidity is correlated with flavor due to the presence of tartaric and malic acids, which account for up to 90% of the acids contained in these fruits [58,59]. The total acidity of the hormone-treated fruits differed significantly. It was shown that the control fruits and fruits treated with 200 mg GA3·L−1 had significantly higher total acidity than the fruits sprayed with gibberellic acid at a concentration of 300 mg GA3·L−1. It was found that the fresh fruits had significantly lower total acidity than after drying (Table 3, Figure 2). In the available literature, the information on the effect of hormonization treatment on the total acidity level is quite inconsistent. A study conducted by Koka [60] showed no significant effect of GA3 application on the acidity of ‘Cardinal’ grapes. An adverse effect of hormonization treatment on the acidity level was shown by Al-Atrushy [61] and Rachna and Singh [62].
Sugars are one of the most important components that determine the quality of fruit, responsible for its sweet taste. The ratio of sugars to organic acids in fruit determines the final taste of grapes [59]. The statistical analysis showed that, regardless of the GA$_3$ concentration, the extract level in the hormonized fruits was significantly lower than in the control (Table 3, Figure 2), which confirms previous studies [24]. There was no significant effect of the concentration of gibberellic acid on the studied trait, and similar relationships were shown during observations in 2010–2012 [12]. Dimovska et al. [2] applied GA$_3$ at concentrations of 5, 10, and 20 mg·GA$_3$·L$^{-1}$, and did not show its significant effect on extract content in ‘Flame Seedless’ fruits. In the study conducted by Al-Atrusha [61], the total sugar level increased significantly with an increasing GA$_3$ concentration. The present study showed that the level of extract in the fresh fruits was 21.14%, which was significantly lower than that in the raisins (73.96%).

The antioxidant activity of grapevine raw materials is related to the presence of secondary metabolites, such as phenolic acids, anthocyanins, flavonoids, and tannins. The phenolic content in the ‘Einset Seedless’ grapevine fruits was not significantly affected by the hormonization treatment and the concentration of GA$_3$ applied. It was shown that the raisins obtained from the fruits of the studied grapevine cultivar were characterized by more than four times significantly higher level of phenolic compounds than the fresh fruits (Table 3, Figure 2).

The antioxidant activity of the fruit extracts determined with the FRAP and DPPH methods significantly depended on the concentration of GA$_3$. Fruits treated with 200 mg·GA$_3$·L$^{-1}$ were characterized by significantly higher antioxidant activity than those treated with 300 mg·GA$_3$·L$^{-1}$. The level of antioxidant activity expressed by the DPPH parameter in the control combination did not differ significantly from the others, while similar relationships were shown between the control and application of 200 mg·GA$_3$·L$^{-1}$ in the case of the FRAP parameter. The fresh fruits were characterized by significantly lower antioxidant activity than the raisins (Table 3, Figure 3). Gougoulias and Masheva [63] showed a positive effect of the hormonization treatment on the increase (from 16 to 42%) in the level of antioxidant activity of seedless grape varieties.

The hormone treatment significantly increased the anthocyanin content only when 200 mg·GA$_3$·L$^{-1}$ was applied (Table 3, Figure 2). The study conducted by Dimovska et al. [2] showed no significant effect of the hormonization treatment on the level of these compounds in fruits of the ‘Flame Seedless’ cultivar. Fresh fruits had more than seven times significantly higher levels of anthocyanins than raisins.

The article presents a Pearson’s correlation analysis in order to determine the influence or lack thereof between the parameters if the chemical composition of biologically active compounds of ‘Einset Seedless’ grape. Pearson’s coefficient for biologically active compounds in the ‘Einset Seedless’ grapevine fruits indicates a strong significant correlation between the DPPH parameter and FRAP, and the total phenolic content and the DPPH and FRAP parameters. A strong significant negative correlation was observed between dry weight and anthocyanin content, total acidity and phenolic content, and DPPH and FRAP parameters (Table 4).

The analysis of Pearson’s coefficient for parameters determining biologically active compounds in the ‘Einset Seedless’ grapevine raisin showed a strong significant correlation between DPPH and FRAP, the total phenolic level and the DPPH and FRAP parameters, and the anthocyanin content and FRAP. A strong negative correlation was observed between total acidity and FRAP. A strong correlation relationship between the considered features within the clusters was expected. Higher extract content in fruit and products made from them is closely related to a higher dry matter value. In turn, the antioxidant properties, determined by FRAP and DPPH methods, largely depend on the overall content of phenolic compounds, including anthocyanins. Such dependencies have been demonstrated in many research studies, including by Kaczmarska et al. [50] (Table 5).
Table 4. Correlation coefficient for biologically active compounds in the ‘Einset Seedless’ fruits and their significance.

|                     | Dry Mass | Total Acidity | Extract | Total Phenolic Content | FRAP   | DPPH   | Anthocyanin |
|---------------------|----------|---------------|---------|------------------------|--------|--------|------------|
| Dry mass            | 1        | −0.4317       | 0.4952  | 0.2689                 | 0.1743 | 0.1580 | −0.7140    |
|                     |          | 0.2460        | 0.1753  | 0.4496                 | 0.6537 | 0.6847 | 0.0307     |
| Total acidity       | −0.4317  | 0.2460        | −0.3123 | −0.7513                | −0.6011 | −0.5993 | −0.0511    |
|                     |          |               | 0.4132  | 0.0196                 | 0.0869 | 0.0881 | 0.8962     |
| Extract             | 0.4952   | −0.3123       | 0.0453  | 0.9078                 | 0.0360 | 0.1147 | −0.5596    |
|                     | 0.1753   | 0.4132        | 1       | 0.9267                 | 0.7690 | 0.1172 |            |
| Total phenolic content | 0.2897 | −0.7513       | 0.0453  | 1                      | 0.8967 | 0.9054 | 0.4008     |
|                     | 0.4496   | 0.0196        | 0.9078  | 1                      | 0.0011 | 0.0008 | 0.2851     |
| FRAP                | 0.4952   | −0.7513       | 0.0453  | 0.9078                 | 0.8967 | 0.9054 | 0.4008     |
|                     | 0.1753   | 0.0196        | 0.9078  | 1                      | 0.0011 | 0.0008 | 0.2851     |
| DPPH                | 0.4952   | −0.7513       | 0.0453  | 0.9078                 | 0.8967 | 0.9054 | 0.4008     |
|                     | 0.1753   | 0.0196        | 0.9078  | 1                      | 0.0011 | 0.0008 | 0.2851     |
| Anthocyanin         | −0.7140  | −0.0511       | −0.5596 | 0.4008                 | 0.4244 | 0.4053 | 0.2792     |
|                     | 0.0307   | 0.8962        | 0.1172  | 0.2851                 | 0.2549 | 0.2792 |            |

The sum of PC (PC1 and PC2) of the total trait variable for the fresh ‘Einset Seedless’ fruits was 100% (for PC1, 62.99%, and for PC2, 37.01%, respectively) (Figure 4). Considering the analysis of biologically active compounds and antioxidant activity of the fresh fruits, a similarity was observed between certain parameters, which formed four groups. The first group is the relationship between dry weight and extract, while the second group is the relationship between the total phenolic content and the DPPH and FRAP parameters. Groups three and four are anthocyanins and total acidity.

The sum of PC of the total variable for the analyzed grape raisins ‘Einset Seedless’ was 100% (for PC1 73.76% and for PC2 26.24%, respectively) (Figure 5). The analysis of the biologically active compounds and antioxidant activity of raisins revealed a similarity between certain parameters, which formed three groups. The first one is the relationship between total phenolic content, anthocyanins, and the DPPH and FRAP parameters. The second relationship was shown between dry weight and extract. The third group was the total acidity.

Table 5. Correlation coefficient for biologically active compounds in the ‘Einset Seedless’ raisins and their significance.

|                     | Dry Mass | Total Acidity | Extract | Total Phenolic Content | FRAP   | DPPH   | Anthocyanin |
|---------------------|----------|---------------|---------|------------------------|--------|--------|------------|
| Dry mass            | 1        | −0.4883       | 0.7712  | 0.0200                 | 0.4528 | 0.1225 | 0.4119     |
|                     |          | 0.1823        | 0.0150  | 0.9953                 | 0.2210 | 0.7535 | 0.2707     |
| Total acidity       | −0.4883  | 0.1823        | 0.0355  | −0.2716                | −0.6458 | −0.3074 | −0.7914    |
|                     |          |               | 0.9928  | 0.4795                 | 0.0063 | 0.4211 | 0.0111     |
| Extract             | 0.7712   | 0.0035        | 1       | −0.0130                | 0.1596 | −0.0429 | −0.1010    |
|                     | 0.0150   | 0.9928        | 1       | 0.9236                 | 0.6818 | 0.9127 | 0.7960     |
| Total phenolic content | 0.0200 | −0.2716       | −0.0130 | 1                      | 0.7952 | 0.9286 | 0.2885     |
|                     | 0.9593   | 0.4795        | 0.9736  | 1                      | 0.0104 | 0.0003 | 0.4515     |
| FRAP                | 0.4528   | −0.6458       | 0.1596  | 0.7952                 | 0.8669 | 0.6691 | 0.0487     |
|                     | 0.2210   | 0.0063        | 0.6818  | 0.0104                 | 0.0025 | 0.0005 | 0.0487     |
| DPPH                | 0.1225   | −0.3074       | −0.0429 | 0.8669                 | 1      | 0.4209 | 0.2593     |
|                     | 0.7535   | 0.4211        | 0.9127  | 0.0003                 | 0.0025 | 0.0005 | 0.2593     |
| Anthocyanin         | 0.4119   | −0.7914       | −0.0011 | 0.2885                 | 0.6691 | 0.4209 | 1          |
|                     | 0.2707   | 0.0111        | 0.7960  | 0.4515                 | 0.0487 | 0.2593 | 1          |
As can be seen from the presented dendrogram (Figure 6) for biologically active compounds and antioxidant activity of the fruits of the studied grapevine cultivar, there are three clusters and an object determining the level of total acidity, which is not included in the dendrogram. A high similarity is shown by group 1—dry matter, total phenolic level, and extract; group 2 are anthocyanins; and group 3 are FRAP and DPPH parameters.

The presented dendrogram (Figure 7) allowed us to determine the similarity of the level of biologically active compounds and antioxidant activity of the ‘Einset Seedless’ raisins. Based on the results, two pairs of clusters showing clear similarities were identified. The analysis revealed that anthocyanins together with the FRAP parameter showed a clear similarity, as did the acidity level and the DPPH parameter. These compounds formed a common object, indicating a high similarity. Similar relationships were shown for the extract and dry matter contents, which formed an object combining with the total phenolic content. The formed objects merge into one at the distant level.
The second relationship was shown between dry weight and extract. The third group was the total acidity.

**Figure 5.** PCA analysis of 'Einset Seedless' raisins against biologically active compounds.

As can be seen from the presented dendrogram (Figure 6) for biologically active compounds and antioxidant activity of the fruits of the studied grapevine cultivar, there are three clusters and an object determining the level of total acidity, which is not included in the dendrogram. A high similarity is shown by group 1—dry matter, total phenolic level, and extract; group 2 are anthocyanins; and group 3 are FRAP and DPPH parameters.

**Figure 6.** Branching-tree diagram for biologically active compounds and antioxidant activity of 'Einset Seedless' fruits irrespective of the concentration of gibberellic acid.

The presented dendrograms (Figures 8 and 9) allowed us to determine the similarity of the effect of the concentrations of gibberellic acid on the content of biologically active compounds and antioxidant activity of the 'Einset Seedless' fruits and raisins. Irrespective of the type of the examined raw material, it was shown that the level of biologically active compounds and antioxidant activity in the case of the concentration of 200 mg·L\(^{-1}\) GA\(_3\) and the control combination was similar but differed substantially in the case of the application of 300 mg·GA\(_3\)·L\(^{-1}\). The above-mentioned relationships occurred in the case of both fresh fruits and raisins.
The hormonization treatment and the form of the plant material analyzed had a significant effect on the content of biologically active compounds and antioxidant activity. Grapes treated with gibberellic acid were characterized by a significantly higher yield, as well as weight, length, and width of grapes, than that of the control. There was a significant effect of the concentration of gibberellic acid applied on the yield, weight, and length of grapes as well as the length and width of berries. Grapes treated with 300 mg·GA$_3$·L$^{-1}$ were characterized by significantly better parameters of the yield size and quality than those exposed to 200 mg·GA$_3$·L$^{-1}$.

The hormonization treatment and the form of the plant material analyzed had a significant effect on the content of biologically active compounds and antioxidant activity of the ‘Einset Seedless’ fruits and raisins. The hormonization treatment significantly reduced the level of dry matter and extract clearly no significant differences were found between the ‘Einset Seedless’ fruits and raisins. The hormonization treatment significantly reduced the level of dry matter and extract clearly no significant differences were found between the ‘Einset Seedless’ fruits and raisins. The concentration of gibberellic acid had a significant effect on the levels of acidity, anthocyanins, and antioxidant activity determined with the FRAP and DPPH methods. The increase in GA$_3$ concentration from 200 mg · L$^{-1}$ to 300 mg · L$^{-1}$ had a negative impact on the size of the parameters tested but it was significantly different in the case of the 300 mg · L$^{-1}$ GA$_3$. The above-mentioned relationships occurred in the case of both fresh fruits and raisins.

### Figure 8
Branching-tree diagram for the hormonal treatment and the concentration of gibberellic acid applied in ‘Einset Seedless’ fruits.

### Figure 9
Branching-tree diagram for the hormonal treatment and the concentration of gibberellic acid applied in ‘Einset Seedless’ raisins.

### 4. Conclusions
The application of the hormonization treatment in the ‘Einset Seedless’ grapevine cultivar had a significant effect on yield quantity and quality. Grapes treated with gibberellic acid were characterized by a significantly higher yield, as well as weight, length, and width of grapes, than that of the control. There was a significant effect of the concentration of gibberellic acid applied on the yield, weight, and length of grapes as well as the length and width of berries. Grapes treated with 300 mg·GA$_3$·L$^{-1}$ were characterized by significantly better parameters of the yield size and quality than those exposed to 200 mg·GA$_3$·L$^{-1}$.
between 200 mg GA$_3$·L$^{-1}$ and 300 mg GA$_3$·L$^{-1}$. The concentration of gibberellic acid had a significant effect on the levels of acidity, anthocyanins, and antioxidant activity determined with the FRAP and DPPH methods. The increase in GA$_3$ concentration from 200 mg·L$^{-1}$ to 300 mg·L$^{-1}$ had a negative impact on the size of the parameters tested.

The fresh fruits were characterized by significantly lower levels of dry matter, acidity, extract, phenols, and antioxidant activity determined with the FRAP and DPPH methods than the raisins. A reverse relationship was shown in the case of the anthocyanin levels.

The application of the multivariate analysis technique showed that, in both the fresh fruits and the raisins, the level of biologically active compounds and antioxidant activity in the case of the 200 mg GA$_3$·L$^{-1}$ concentration and the control combination was similar, but it was significantly different in the case of the 300 mg GA$_3$·L$^{-1}$ application.

Author Contributions: Conceptualization: M.K., K.K., E.J.-R., A.S. (Aneta Ślawińska) and A.S. (Anna Stój); data curation: M.K., K.K., E.J.-R., A.S. (Aneta Ślawińska) and A.S. (Anna Stój); formal analysis: M.K. and K.K.; funding acquisition: M.K. and K.K.; investigation: M.K. and K.K.; methodology: M.K., K.K. and E.J.-R.; project administration: M.K. and K.K.; resources: E.J.-R., A.S. (Aneta Ślawińska) and A.S. (Anna Stój); software: K.K.; supervision: M.K., K.K. and E.J.-R.; validation: E.J.-R., A.S. (Aneta Ślawińska) and A.S. (Anna Stój); visualization: M.K. and K.K.; writing—original draft preparation: M.K., K.K., E.J.-R., A.S. (Aneta Ślawińska) and A.S. (Anna Stój); writing—review and editing: M.K. and K.K. All authors have read and agreed to the published version of the manuscript.

Funding: Statutory activity University of Life Sciences in Lublin.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study, in the collection, analyses, or interpretation of data, in the writing of the manuscript, or in the decision to publish the results.

References
1. Yang, J.; Martinson, T.E.; Liu, R.H. Phytochemical profiles and antioxidant activities of wine grapes. Food Chem. 2009, 116, 332–339. [CrossRef]
2. Dimovska, V.; Petropulos, V.I.; Salamovska, A.; Ilieva, F. Flame Seedless grape variety (Vitis vinifera L.) and different concentrations of gibberellic acid (GA$_3$). Bulg. J. Agric. Sci. 2014, 20, 137–142.
3. Myśliwiec, R. Uprawa Winorośli; Plantpress: Kraków, Poland, 2009; p. 161.
4. Khan, M.; Hafeez-ur-Rahman, A.; Ahmed, M.; Abbas, G.; Ahmed, N. Effect of gibberellic acid on growth and fruit yield of grape cultivar ‘flame seedless’. Int. J. Biol. Biotech. 2009, 6, 265–268.
5. Nampil, R.; Chen, B.-S.; Chen, C.C.; Yang, Y.S. Effect of GA$_3$ and CPPU on berry size of seedless grapes. Hort. NCHU 2010, 35, 53–64.
6. Kaplan, M.; Najda, A.; Baryła, P.; Klimek, K. Effect of gibberellic acid dose and number of treatments on yield components of ‘Einset Seedless’ grapevine cultivar. Hort. Sci. 2017, 44, 195–200.
7. Sečer, M. Natural growth regulator’s physiological effects and researches about this area. Derim 1989, 6, 109–124.
8. Bora, R.K.; Sarma, C.M. Effect of gibberellic acid and Cycocel on growth, yield and protein content of pea. Asian J. Plant Sci. 2006, 5, 324–330.
9. Korkutal, I.; Bahar, E.; Gökhan, O. The characteristics of substances regulating growth and development of plants and the utilization of gibberellic acid (GA$_3$) in viticulture. World J. Agric. Sci. 2008, 4, 321–325.
10. Kaplan, M. Wpływ kwasu giberelinowego i kwasu 2-naftoksyoctowego na wielkość i jakość plonu winorośli odmiany ‘Einset Seedless’. Zesz. Probl. Postęp. Nauk Rol. 2009, 539, 299–305.
11. Kaplan, M. The effect of the method of application of growth regulators on fruit quality of ‘Einset Seedless’ Grape (Vitis sp. L.). Acta Agrobot. 2011, 64, 189–196. [CrossRef]
12. Kaplan, M. Wpływ stężenia kwasu giberelinowego oraz terminu zabiegu na wielkość i jakość plonu winorośli (Vitis sp. L.) odmiany ‘Einset Seedless’. Episteme 2013, 20, 327–338.
13. Yamada, M.; Yamane, H.; Kurihara, A.; Nagata, K.; Yoshinaga, K.; Hikarawa, N.; Sato, A.; Iwanawi, H.; Ozawa, T.; Sumi, T.; et al. New grape variety Sunny Rouge. Bull. Natl. Inst. Fruit Tree Sci. Jpn. 2003, 2, 33–42.
14. Dokouzlan, N. Influence of Gibberellic Acid Berry Sizing Sprays on Crimson Seedless Table Grape; University of California Cooperative Extension, Tulare County, Agriculture and Natural Resources: Oakland, CA, USA, 1999; Available online: http://cetulare.ucdavis.edu/pubgrape/tb897.htm (accessed on 11 October 2021).
15. Dass, H.C.; Randhawa, G.S. Response of certain seeded Vitis vinifera varieties to gibberellin application at post bloom stage. Am. J. Enol. Vitic. 1968, 19, 56–62.

16. Halbrook, M.C.; Mortensen, J.A. Influence of gibberellic acid and various management practices on berry; seed and cluster development in ‘Orland Seedless’ grape. Proc. Fla. State Hort. Soc. 1987, 100, 312–315.

17. Lu, J. Application of gibberellic acid on grape cultivar ‘Orlando Seedless’. Proc. Fla. State Hort. Soc. 1996, 109, 246–247.

18. Surasak, N.; Choopong, S. The Improvement of Grape Quality and Production: Fruiting Responses of Some Grape Varieties to Gibberellic Acid. Kasetsart J. Nat. Sci. 1988, 22, 229–237.

19. Pommer, C.V. Characteristic of seedless grape cv. Maria as affected by girdling and gibberellic acid. Bragantia 1995, 54, 151–159. [CrossRef]

20. Formolo, R.; Rufato, L.; Kretzschmar, A.A.; Schlemer, C.; Mendes, M.; Marcon Filho, J.L.; Lima, A.P. Gibberellic acid and cluster thinning on seedless grape ‘BRS Clara’ in Caxias do Sul; Rio Grande do Sul State; Brazil. Acta Hort. 2010, 884, 467–471. [CrossRef]

21. Korkas, E.; Nerantzis, E.; Kourtidou-Tymba, P.; Banilas, G. The effect of gibberellic acid application at different phenological growth stages on yield and quality parameters of ‘Sultanina’ table grapes (Vitis vinifera L.) in Greece. Part I. At development of flower cluster and at fruit set bloomtime. Vitic. Enol. Sci. 1999, 54, 44–53.

22. Pérez, F.J.; Gómez, M. Possible role of soluble inverte in the gibberellic acid berry-sizing effect in Sultana grape. Plant Growth Regul. 2000, 30, 111–116. [CrossRef]

23. Casanova, L.; Casanova, R.; Moret, A.; Agusti, M. The application of gibberellic acid increases berry size of ‘Emperatiz’ seedless grape. Span. J. Agric. Res. 2007, 9, 919–927. [CrossRef]

24. Kaplan, M.; Najda, A.; Klimek, K.; Borowy, A. Effect of Gibberellic Acid (GA3) Inflorescence Application on Content of Bioactive Compounds and Antioxidant Potential of Grape (Vitis, L.) ‘Einset Seedless’ Berries. S. Afr. J. Enol. Vitic. 2019, 40, 1–10. [CrossRef]

25. Abu-Zahra, T.R. Berry size of ‘Thompson Seedless’ as influenced by the application of gibberellic acid and cane girdling. Pak. J. Bot. 2010, 42, 1755–1760.

26. Mazza, G. Anthocyanins in grapes and grape products. Crit. Rev. Food Sci. Nutr. 1995, 35, 341–371. [CrossRef]

27. Pezzuto, J.M. Grapes and human health: A perspective. J. Agric. Food Chem. 2000, 58, 6777–6784. [CrossRef]

28. Krońskiak, M.; Gąsłoł, M.; Banach, P.; Pytel, A. Wybrane parametry jakościowe winogron uprawianych w Polsce południowej. Żywność. Nauka Technol. Jakość 2009, 4, 116–121.

29. Katalinić, V.; Mozina, S.S.; Skroza, D.; Generalić, I.; Abramović, H.; Miloš, M.; Ljubenkov, I.; Piskernik, S.; Pezo, I.; Terpnic, P.; et al. Polyphenolic profile; antioxidant properties and antimicrobial activity of grape skin extracts of 14 Vitis vinifera varieties grown in Dalmatia (Croatia). Food Chem. 2010, 119, 715–723. [CrossRef]

30. Arts, I.; Hollman, P. Polyphenols and disease risk in epidemiologic studies. Am. J. Clin. Nutr. 2005, 81, 317–325. [CrossRef]

31. Erdman, J.; Balentine, D.; Arab, L.; Beecher, G.; Dwyer, J.T.; Folts, J.; Hamly, J.; Hollman, P.; Keen, C.L.; Mazza, G.; et al. Flavonoids and heart health. Proceedings of the ILSI North America Flavonoids Workshop. J. Nutr. 2007, 137, 718–737. [CrossRef]

32. Leifert, W.R.; Abeywardena, M.Y. Cardio protective actions of grape polyphenols. Nutr. Res. 2008, 28, 729–737. [CrossRef]

33. Frankel, E.N. Natural phenolic antioxidants and their impact on health. In Antioxidant Food Supplements in Human Health; Packer, L., Ed.; Academic Press: London, UK, 1999; pp. 385–392.

34. Stevenson, D.E.; Hurst, R.D. Polyphenyly phytocemicals: Just antioxidants or much more? Cell. Mol. Life Sci. 2007, 64, 2900–2916. [CrossRef] [PubMed]

35. Dohadwala, M.; Vita, J.A. Grapes and cardiovascular disease. J. Nutr. 2009, 139, 1788–1793. [CrossRef] [PubMed]

36. Crozier, A.; Del Rio, D.; Clifford, M.N. Bioavailability of dietary flavonoids and phenolic compounds. Mol. Asp. Med. 2010, 31, 446–467. [CrossRef]

37. Xia, E.; Deng, G.F.; Guo, Y.J.; Li, H.B. Biological activities of polyphenols from grapes. Int. J. Mol. Sci. 2010, 11, 622–646. [CrossRef]

38. Vislocky, L.M.; Fernandez, M.L. Biomedical effects of grape products. Nutr. Rev. 2010, 68, 656–670. [CrossRef]

39. Scalbert, A.; Manach, C.; Morand, C.; Remesy, C.; Jimenez, L. Dietary polyphenols and the prevention of diseases. Crit. Rev. Food Sci. Nutr. 2005, 45, 287–306. [CrossRef]

40. Kim, D.O.; Jeong, S.W.; Lee, C.Y. Antioxidant capacity of phenolic phytochemicals from various cultivars of plums. Food Chem. 2003, 81, 321–326. [CrossRef]

41. Peña-Neira, A.; Dueñas, M.; Duarte, A.; Hernández, T.; Estrella, I.; Loyola, E. Effects of ripening stages and of plant vegetative rigor on the phenolic composition of grapes (Vitis vinifera L.) cv. Cabernet Sauvignon in the Maipo Valley (Chile). Vitis 2004, 43, 51–57.

42. Jiang, H.; Ji, B.P.; Liang, J.F.; Zhou, F.; Yang, Z.W.; Zhang, G.Z. Changes of contents and antioxidant activities of polyphenols during fruit development of four apple cultivars. Eur. Food Res. Technol. 2006, 223, 743–748. [CrossRef]

43. Montalegre, R.R.; Peces, R.; Vozmediano, J.L.; Gascuena, J.M.; Romero, G.E. Phenolic compounds in skins and seeds of ten grape Vitis vinifera varieties grown in a warm climate. J. Food Compos. Anal. 2006, 19, 687–693. [CrossRef]

44. Orak, H.H. Total antioxidant activities; phenolics; anthocyanins; polyphenoloxidase activities of selected red grape varieties and their correlation. Sci. Hort. 2007, 111, 235–241. [CrossRef]

45. Tomás-Barberán, F.A.; Espin, J. Phenolic compounds and related enzymes as determinants of quality in fruits and vegetables. J. Sci. Food Agric. 2001, 81, 853–867. [CrossRef]

46. Es-Safi, N.; Ghidouche, S.; Ducrot, P.H. Flavonoids: Hemisynthesis; reactivity; characterization and free radical scavenging activity. Molecules 2007, 12, 2228–2258. [CrossRef] [PubMed]
47. Reisch, B.I.; Remaly, G.W.; Pool, R.M.; Watson, J.P. ‘Einset Seedless’ grape. *HortScience* 1986, 21, 155–156.
48. The Polish Committee for Standardization (PKN). *Fruit and Vegetable Products. Preparation of Samples and Testing Methods. Determination of Total Extract; PN-A-75101-02:1990/Az1:2002*; Polish Committee for Standardization: Warszawa, Poland, 2002.
49. The Polish Committee for Standardization (PKN). *Fruit and Vegetable Products. Preparation of Samples and Testing Methods. Determination of Dry Matter Content by Gravimetric Method; PN-A-75101-03:1990*; Polish Committee for Standardization: Warszawa, Poland, 1990.
50. Kaczmarska, E.; Gawroński, J.; Jabłońska-Ryś, E.; Zalewska-Korona, M.; Radzki, W.; Sławinska, A. General combining ability and heterosis regarding the phytochemical properties in strawberry (*Fragaria × ananassa*) hybrids. *Plant Breed.* 2017, 136, 111–118. [CrossRef]
51. The Polish Committee for Standardization (PKN). *Fruit and Vegetable Juices. Determination of Titratable Acidity; PN-EN 12147:2000*; Polish Committee for Standardization: Warszawa, Poland, 2000.
52. Singleton, V.L.; Rossi, J.A. Colorimetry of total phenolics with phosphomolybdic acid–phosphotungstic acid reagents. *Am. J. Enol. Vitic.* 1965, 16, 144–158.
53. Benzie, I.F.F.; Strain, J.J. The ferric reducing ability of plasma (FRAP) as a measure of ‘antioxidant power’: The FRAP assay. *Anal. Biochem.* 1996, 239, 70–76. [CrossRef]
54. Choi, Y.; Lee, S.M.; Chun, J.; Lee, H.B.; Lee, J. Influence of heat treatment on the antioxidant activities and polyphenolic compounds of shiitake (*lentinus edodes*) mushroom. *Food Chem.* 2006, 99, 381–387. [CrossRef]
55. Wrolstade, R.E. *Color and Pigment Analyses in Fruits Products; Oregon State University Agriculutural Experiment Station Bulletin:* Corvallis, OR, USA, 1976; p. 624.
56. Gordillo, B.; Sigurdson, G.T.; Lao, F.; González-Miret, M.L.; Heredia, F.J.; Giusti, M.M. Assessment of the color modulation and stability of naturally copigmented anthocyanin-grape colorants with different levels of purification. *Food Res. Int.* 2018, 106, 791–799. [CrossRef]
57. Zabadal, T.J.; Dittmer, T.W. Gibberellic acid sprays increase berry size and reduce shot berry of ‘Vanessa’ grapevine. *J. Am. Pomol. Soc.* 2000, 54, 130–133.
58. Laszlo, J.C.; Saayman, D. Optimum harvesting stage for Sultanina as table grape. *Decid. Fruit Grow.* 1990, 40, 101–105.
59. Topalovic, A.; Mikulic-Petkovsek, M. Changes in sugars; organic acids and phenolics of grape berries of cultivar Cardinal during ripening. *J. Food Agric. Environ.* 2010, 8, 223–227.
60. Kok, D. Grape growth; anthocyanin and phenolic compounds content of early ripening Cv. Cardinal table grape (*V. vinifera* L.) as affected by various doses of foliar biostimulant applications with gibberellic acid. *Erwerbs-Obstbau* 2017, 58, 1–7. [CrossRef]
61. Al-Atrashy, S.M.M. Effect of GA3 dose and frequency on yield and quality of ‘Zark’ grape. *Jordan J. Agric. Sci.* 2016, 12, 1183–1191.
62. Arora, R.; Singh, S. Effect of gibberellic acid on periodical changes in bio-chemical composition of ber cv. *Umran. HortFlora Res. Spectr.* 2013, 2, 25–29.
63. Gougoulias, N.; Masheva, L. Effect of gibberellic acid (GA3) on polyphenols content and antioxidative activity of some table grape varieties of *Vitis vinifera* L. *Oxid. Commun.* 2010, 33, 652–660.