Excellent histological results in terms of articular cartilage regeneration after spheroid-based autologous chondrocyte implantation (ACI)

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Abstract
Purpose Traumatic lesions of articular cartilage represent a crucial risk factor for osteoarthritis. Even if several strategies exist to treat such damages, the optimal solution has not yet been found. A new strategy represents the scaffold-free spheroid-based autologous chondrocyte transplantation. In this method, spheroids of chondrocytes are synthesized after chondrocyte isolation and expansion, followed by the implantation in a second intervention.

Methods Fine Jamshidi-needle biopsies from five patients (one from each patient, Ø 2 mm) treated with a spheroid-based autologous chondrocyte implantation (ACI) after traumatic lesions of the articular cartilage of the knee were analysed histologically and immunohistologically for collagen II, collagen X and aggrecan expression. The indication for a second look arthroscopy was given by arthrofibrosis or meniscus-lesions, respectively. The time between ACI and second-look arthroscopy ranged between 6 and 16 months.

Results In all patients, the histological examinations revealed an avascular cartilage tissue with a homogenic extracellular matrix. The subchondral bone neither showed bleeding, necrosis nor hypertrophy. A homogenous alcian blue staining indicated high amounts of mucopolysaccharides and glycosaminoglycans. Collagen II staining was highly positive, whereas collagen X staining was negative in every patient, ruling out hypertrophic chondrocyte differentiation. In addition, intense aggrecan staining indicated a strong expression of this extracellular matrix component.

Conclusion The present case series represents the first histological and immunohistological analyses of spheroid-based ACI in humans. Spheroid-based ACI revealed excellent histological results regarding the regeneration of hyaline articular cartilage. These results indicate that spheroid based ACI is a promising strategy for treating traumatic lesions of the articular cartilage of the knee.

Keywords Cartilage · Autologous chondrocyte transplantation · Chondrocytes · Spheroids · MACI

Introduction

Hyaline cartilage has a unique capacity to answer pressure transformations and is thus critical for the proper function of the musculoskeletal system [11]. However, it has limited regeneration capacity after trauma, leading to degenerative changes of the traumatized cartilage. These changes represent a risk factor for early development of osteoarthritis, resulting in the complete destruction of the joint.

Different strategies have been developed to treat cartilage lesions, such as mosaic arthroplasty, microfracture, autologous matrix-induced chondrogenesis (AMIC) and autologous chondrocyte implantation (ACI) [10, 12]. These strategies were shown to significantly reduce relevant clinical
burdens of traumatic cartilage lesions, such as pain and immobility. Moreover, the necessity of total joint replacement could be postponed.

Currently, several Matrix-based ACI (MACI) are in clinical use [1, 9, 18, 19]. However, until today it is not yet clear, which is the optimal one, especially with respect to hyaline cartilage regeneration [4]. Bentley et al. compared ACI with mosaic arthroplasty and revealed excellent clinical, radiological, and histological results [6]. Zeifang et al. compared ACI with MACI and reported no significant differences regarding the IKDC-score, Tegner-activity and the SF-36, but a significantly better result in the Lysholm-score for the ACI-P [20]. A trial comparing periosteal flap covered (ACI-P) and collagen membrane-covered ACI (ACI-C) could not detect histological differences in Safranin-O staining between ACI-P and ACI-C [13, 15]. However, a recent trial reports a high number of failures in osteoarthritic knees and postulate to better profile patients before undergoing the MACI [3].

The spheroid-based ACI represents a relatively new technique. It requires two surgical interventions, one for harvesting chondrocytes and one for the implantation of the generated chondrocyte spheroids [2, 14]. The spheroids are implanted into the defect after debridement and adhere within 20 min at the implantation site. The safety and efficacy of this technique could be demonstrated by clinical studies [5, 17]. However, until today there are no publications regarding the histological outcome after this treatment. In the present case study, we show for the first time histological and immunohistological results of regenerated hyaline cartilage-like tissue after spheroid-based ACT. The present results are of clinical importance as the regenerated cartilage tissue clearly resembles normal hyaline cartilage tissue, which gives relevant implications for its functional properties.

Materials and methods

Ethical approval for the re-evaluation was given by the Institutional Review Board of the University Regensburg (ID-number: 19-1558-104).

Samples were collected from five patients (one biopsy from each patient) treated with spheroid-based ACI (Codon, Teltow, Germany) due to traumatic lesions followed by re-arthroscopy. All defects were localized on the medial femur condyle. Implantation was performed via mini-arthrotomy (Fig. 1). Re-arthroscopy was necessary on the basis of clinical indications. In three patients a lesion of the medial meniscus, in one patient arthrofibrosis and in one patient persisting pain of unknown origin. Therefore, time-points of re-arthroscopy were not homogenous in the present report. Every patient agreed with a transplant-biopsy in written form to ensure histological analyses of a potential cartilage hypertrophy versus hypertrophy of the subchondral bone. Biopsies were taken from the central part of the implant using a 2 mm Jamshidi-needle. Specimens were immediately fixed in formalin. The patient characteristics are summarized in Table 1.

Histological and immunohistological staining

Standardized and automated methods were used in all procedures. The specimens were decalcified, dehydrated and embedded in paraffin (Shandon Pathcentre, Thermo Electron Corporation, USA). Sections of 5 µm thickness were deparaffinized and histological staining was performed with haematoxylin/eosin and alcian blue. Immunohistological detection was performed for collagen II, collagen X and aggrecan (Table 2) in a fully automated system (Histostainer Plus, Dako, Hamburg, Germany).

Histological evaluation

Specimens were histologically analysed. The existence of hyaline cartilage-like tissue, fibrous tissue and potential vascularization were examined. Furthermore, the potential hypertrophy of the subchondral bone was described.

Immunohistological evaluation

The evaluation was performed by two independent double-blinded investigators (D.G. and C.B.) using a visual-based scheme in which the stained area and the intensity of the staining were scored semi-quantitatively using a wide-spread
Knee Surgery, Sports Traumatology, Arthroscopy

The staining intensity was evaluated as zero (negative/no staining), + (slight positive), + + (moderate) and + + + (strongly positive). A positive control (+ + + /strongly positive) and a negative control (0/negative) were used as references. No test of significance was possible due to the very low number of patients. Therefore, the focus was placed on the exact description of the tissue. The findings were documented by whole slide imaging using a Digital Microscope and Scanner Device (M8, PreciPoint, Freising, Germany).

Results

Histological and histochemical results

Histologically, an avascular cartilage tissue with an intact cartilage-bone border interface was observed in the regenerated tissue in all patients. Cells within the regenerated tissue showed a round and chondrocytic phenotype with cluster-formation. In three patients the typical layer-based architecture with flattened chondrocytes in the apical layer was detectable. The alcian blue staining revealed intensive and homogeneous staining of the extracellular matrix in all patients, which represents an indicator for the presence of glycosaminoglycans and mucopolysaccharides. In three of five patients, the staining was intensive in the basal half of the biopsy and decreased in the apical direction, corresponding to the three patients with the typical layer-based structure. The other two patients showed a homogeneous alcian blue staining. There were no signs for degenerative changes, such as demarked collagen fibres in the tissue. Hypertrophy of the subchondral bone was not detected in any of the five analysed patients.

Immunohistological results

Immunohistologically, one biopsy showed an intensive and homogeneous expression of collagen II within the extracellular matrix. Two biopsies showed an intensive expression of collagen II in the basal and middle zone of the biopsy with a slight decrease of intensity in the apical half of the specimen. In one patient, higher expression in the apical zone could be detected than in the middle and basal zone. One patient was not evaluable due to loss of the cartilage from the biopsy after performing the histological staining. The immunohistological staining for aggrecan revealed a homogenous strong expression in all patients. No spatial differences of the aggrecan positivity could be detected in the basal, middle or apical zone of the biopsies. The immunohistological staining for collagen X antigen was negative in four patients. In one patient no more cartilage tissue was seen in the biopsy. The results of the immunohistological evaluation are summarized in Table 1. The histological and immunohistological findings are documented in Fig. 2.

Discussion

The present study demonstrates for the first time that the regenerated cartilage tissue after spheroid-based ACI is histologically, histochemically and immunohistochemically very similar to hyaline articular cartilage. The typical layer-based architecture with a deep, middle and apical zone was identified in three patients. In addition, these three patients demonstrated a typical apical zone with two to three layers of flattened chondrocytes and a decrease of alcian blue-positive glycosaminoglycans, which is well

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Table 1 Results of the histological and immunohistological stainings

| Sex F/M | Age [Y] | Time of biopsy [mos] | Defect localization | HE (N=5) | COL II (N=4) | COL X (N=4) | ACAN (N=5) |
|---------|--------|---------------------|--------------------|---------|-------------|-------------|-----------|
| Range   | 1/4    | 37–47               | 5½–16              | MFC II (3x) | + +         | + + (Basal) | Negative  |
|         |        |                     |                    | MFC r (2x)  | + + +       | + (Apikal)  | + + +     |
| Ø [± SD]| 43 [± 4]| 11 [± 7]            |                    |          | + +         | +           | Negative  |

Biopsies from five patients were collected at different time points after ACI. Staining intensity of haematoxylin (HE), Collagen II (COL II), Collagen X (COL X) and Aggrecan (ACAN) stained paraffin sections was assessed (negative/+ +/+ + + /strongly positive)

Table 2 Antibodies used for histological and immunohistological stainings

| Antigen | Supplier information | Dilution | Blocking buffer/antibody diluent |
|---------|----------------------|----------|---------------------------------|
| Collagen II | Nr. II-Il6B3, DSHB, Iowa, USA | 1:40 | TNB buffer |
| Collagen X | Dako EnVision Flex, HRP, Dab, K8010, Dako, Glostrup, Denmark | 1:50 | Citrate buffer |
| Aggrecan | MS MAB Anti-Hu-Aggrecan, Invitrogen, Carlsbad/CA, USA | 1:1000 | Citrate buffer |

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The biopsy and decreased in the apical direction, corresponding to the three patients with the typical layer-based structure. The other two patients showed a homogeneous alcian blue staining. There were no signs for degenerative changes, such as demarked collagen fibres in the tissue. Hypertrophy of the subchondral bone was not detected in any of the five analysed patients.
known from initial articular cartilage. Specimens with high amounts of glycosaminoglycans, high expression of collagen type II and cells with chondroid phenotype, but without the typical layer-based architecture was defined as “very similar to hyaline articular cartilage”, which was present already 5 ½ months after ACI and almost of similar structure in the specimen which was taken 16 months after ACI. These findings implicate a good regenerative potential of the spheroids, making them an attractive alternative to other methods where changes in the regenerative tissue and long-term failures have been described [3, 6]. Long-term follow-up studies should clarify if spheroid-based ACI also reveals a good long-term performance with respect to proper cartilage regeneration and good long-term preservation of the regenerated tissue. For that purpose, histological analyses are the method of choice, since clinical or radiological examinations were not able to verify the quality of the regenerated tissue [7, 16, 20]. However, further long-term follow-up trials are mandatory for a better understanding of the cellular mechanisms involved in cartilage regeneration and preservation, as well as to give a proper base for the comparison with single-stage techniques as e.g. microfracture, which may be easier to perform [18]. From the clinical point of view, a recent study demonstrated a substantial improvement of various clinical outcomes parameters after 36 months after matrix-associated, spheroid-based ACI compared to microfracturing [14]. Nevertheless, the ACI should also prove, that the regenerated tissue is indeed a noticeable improvement in tissue quality which is mandatory for long-term function.

In this study biopsies taken between 5 ½ and 16 months after ACI clearly demonstrated the regeneration of predominantly hyaline cartilage-tissue. Nevertheless, the limited number of specimens without a homogenous study cohort is a major limitation of the present case-study. Further clinical trials with a larger number of patients have to be performed to consolidate these results and to generate statistically significant data [3]. The indication for the biopsy in this report was given by a potential hypertrophy of the subchondral bone, which had to be excluded by histomorphological analyses.

**Conclusion**

The present study represents the first histological and immunohistological analysis of spheroid-based ACI. It shows that spheroid-based ACI yields excellent histological results in terms of regeneration with predominantly hyaline articular cartilage.
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Compliance with ethical standards

Conflict of interest Authors declare that they have no conflict of interest.

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Ethical approval This study is approved by the Institutional Review Board of the University Regensburg (ID-number: 19-1558-104).

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References

1. Abelow SP, Guillen P, Ramos T (2006) Arthroscopic technique for matrix-induced autologous chondrocyte implantation for the treatment of large chondral defects in the knee and ankle. Oper Tech Orthop 16:257–261
2. Anderer U, Libera J (2002) In vitro engineering of human autogenous cartilage. J Bone Miner Res 17:1420–1429
3. Andriolo L, Reale D, Di Martino A, Zaffagnini S, Vannini F, Ferruzzi A, Filardo G (2019) High rate of failure after matrix-assisted autologous chondrocyte transplantation in osteoarthritic knees at 15 years of follow-up. Am J Sports Med 47(9):2116–2122
4. Bartlett W, Skinner JA, Gooding CR et al (2005) Autologous chondrocyte implantation versus matrix-induced autologous chondrocyte implantation for osteochondral defects of the knee: a prospective, randomised study. J Bone Joint Surg Br 87:640–645
5. Becher C, Laute V, Fickert S et al (2017) Safety of three different product doses in autologous chondrocyte implantation: results of a prospective, randomised, controlled trial. J Orthop Surg Res 12:71
6. Bentley G, Biant LC, Carrington RW et al (2003) A prospective, randomised comparison of autologous chondrocyte implantation versus mosaicplasty for osteoarticular defects in the knee. J Bone Joint Surg Br 85:223–230
7. Bhosale AM, Kuiper JH, Johnson WE (2009) Midterm to long-term longitudinal outcome of autologous chondrocyte implantation in the knee joint. Am J Sports Med 37:131–138
8. Biant LC, Bentley G, Vijayan S et al (2014) Long-term results of autologous chondrocyte implantation in the knee for chronic chondral and osteochondral defects. Am J Sports Med 42:2178–2783
9. Britberg M (2010) Cell carriers as the next generation of cell therapy for cartilage repair: a review of the matrix-induced autologous chondrocyte implantation procedure. Am J Sports Med 38:1259–1271
10. Chahla J, Stone J, Mandelbaum BR (2019) How to manage cartilage injuries? Arthroscopy 35(10):2771–2773. https://doi.org/10.1016/j.arthro.2019.08.021
11. Eckstein F, Hudelmaier M, Putz R (2006) The effects of exercise in human articular cartilage. Anat 208:491–512
12. Everhart JS, Jiang EX, Poland SG, Du A, Flanagan DC (2019) Failures, reoperations, and improvement in knee symptoms following matrix-assisted autologous chondrocyte transplantation: a meta-analysis of prospective comparative trials. Cartilage. https://doi.org/10.1177/1947603519870861
13. Gooding CR, Bartlett W, Bentley G et al (2006) A prospective, randomised study comparing two techniques of autologous chondrocyte implantation for osteochondral defects in the knee: periosseum covered versus type I/III collagen covered. Knee 13:203–210
14. Hoburg A, Niemeyer P, Laute V, Zinser W, Becher C, Kolombe T, Fay J, Pietsch S, Kuzma T, Widuchowski W, Fickert S (2020) Matrix-associated autologous chondrocyte implantation with spheroid technology is superior to arthroscopic microfracture at 36 months regarding activities of daily living and sporting activities after treatment. Cartilage 1:1947603519897290. https://doi.org/10.1177/1947603519897290
15. McCarthy HS, Roberts S (2013) A histological comparison of the repair tissue formed when using either Chondrogide® or periosteum during autologous chondrocyte implantation. Osteoarth Cartilage 21:2048–2057
16. Nawaz SZ, Bentley G, Briggs TW, Carrington RW et al (2014) Autologous chondrocyte implantation in the knee: mid-term to long-term results. J Bone Joint Surg Am 96:824–830
17. Niemeyer P, Laute V, John T et al (2016) The effect of cell dose on the early magnetic resonance morphological outcomes of autologous cell implantation for articular cartilage defects in the knee. A randomized clinical trial. Am J Sports Med 44:2005–2014
18. Russilies M, Behrens P, Wünsch L et al (2002) A cell-seeded bio-composite for cartilage repair. Ann Anat 184:317–323
19. Schuette HB, Kraeutler MJ, McCarty EC (2017) Matrix-assisted autologous chondrocyte transplantation in the knee: a systematic review of mid- to long-term clinical outcomes. Orthop J Sports Med. https://doi.org/10.1177/2325967117T0290
20. Zeifang F, Oberle D, Nierhoff C et al (2009) Autologous chondrocyte implantation using the original periosteum-cover technique versus matrix-associated autologous chondrocyte implantation: a randomized clinical trial. Am J Sports Med. 38:924–933

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