Case report

A del(X)(p11) carrying SRY sequences in an infant with ambiguous genitalia

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Abstract

Background: SRY (sex-determining region, Y) is the gene responsible of gonadal differentiation in the male and it is essential for the regular development of male genitalia. Translocations involving the human sex chromosomes are rarely reported, however here we are reporting a very rare translocation of SRY gene to the q-arm of a deleted X chromosome. This finding was confirmed by cytogenetic, fluorescent in situ hybridization (FISH) and polymerase chain reaction (PCR).

Case presentation: A 7-month infant was clinically diagnosed as an intersex case, with a phallus, labia majora and minora, a blind vagina and a male urethra. Neither uterus nor testes was detected by Ultrasonography. G-banding of his chromosomes showed 46.X,del(X)(p11) and fluorescent in situ hybridization (FISH) analysis showed a very small piece from the Y chromosome translocated to the q-arm of the del(X). Polymerase chain reaction (PCR) analysis revealed the presence of material from the sex-determining region Y (SRY) gene.

Conclusion: It is suggested that the phenotype of the patient was caused by activation of the deleted X chromosome with SRY translocation, which is responsible for gonadal differentiation.

Background

The SRY (sex-determining region Y), which is normally located in the distal part of the short arm of the Y chromosome is a genetic ‘master switch’ of gonadal differentiation [1], the product of which is present in the male genital ridge before testis formation and is required for the regular development of male genitalia [2]. SRY encodes a transcription factor that is a member of the high mobility group (HMG)-box family of DNA binding proteins and in mammals triggers the development of undifferentiated gonads towards a testicular phenotype [1,3]. In humans, zygotes bearing mutations in SRY develop into XY females [4,5], while XX individuals with the presence of SRY typically show a normal male phenotype [6], but may occasionally show ambiguous genitalia [7]. One example is testicular regression syndrome (XY gonadal regression syndrome), in which there are no gonads, and variable development of Mullerian and Wolfian ducts depending...
on the stage of fetal development at which the embryonic testis involutes; in most cases this happens before Mullerian tissues have regressed and before testosterone synthesis has started, the etiology is unknown but affected siblings have been reported [8].

Case presentation

Case history
A 7-month old infant, raised as a male, was clinically diagnosed as an intersex case. On examination the patient showed a big phallus, labia majora and minora, a blind vagina and a male urethra (Fig. 1). Ultrasonographic investigations did not detect the presence of uterus or testes. The patient was referred for chromosomal analysis.

Cytogenetic analysis
Patient peripheral blood was subjected to short-term culturing in RPMI 1640 medium for 72 hours. After metaphase arrest through exposure to Colcemide, cells were harvested, treated with hypotonic solution, and then fixed with methanol and acetic acid according to standard procedures. The harvested cells were dropped on clean slides and stained with Wright's stain, for chromosome banding [9]. The clonality criteria and the karyotypic descriptions were according to the ISCN recommendations [10].

Fluorescent in situ hybridization (FISH)
FISH with whole chromosome painting, and X centromeric probe, and the SRY gene specific probe (Vysis, Naperville, IL) was applied to fixed metaphases cells according to standard procedures [11].

Analysis of 11 metaphase cells showed 46,X,del(X)(p11) in all cells, which is a deletion within the breakpoint in band 11 in the p arm of one of the X chromosomes (Fig 2).

PCR analysis corroborated the presence of SRY material in the peripheral blood of the patient (Fig 4).

Conclusion
Translocations involving the human sex chromosomes are rarely reported. One of the best-known consequences of such exchanges is sex reversal in 46,XX males and some 46,XY females, due to exchange in the paternal germ line of terminal portions of Xp and Yp, including the SRY gene [13]. The presence of the SRY gene in the normal male zygote leads to the development of male genital organs and the absence of this gene leads to the development of the female genitalia. Patients who carry a structural abnormality of the X chromosome and ambiguous genitalia have provided opportunities to elucidate the genotype/phenotype correlation in relation to the X and Y chromosome content and X chromosome inactivation. At least

Figure 1
The external genitalia of the 7-month Sudanese infant who was clinically diagnosed as intersex.
one previous intersex case with an Xp deletion has been reported [14]. However, SRY sequences could not be detected in that patient and the authors suggested that the phenotype resulted from unmasking of recessive mutations in Xp by activation of the abnormal X chromosome in the majority of cells. In the present case, G-banding showed one ostensibly normal X chromosome and one X chromosome with deletion of most of the short arm. However, FISH and PCR analyses showed that there was also a translocation of SRY gene material to the long arm of abnormal X, which could potentially lead to development of normal male genitalia. In the present case, development of normal male genitalia presumes a skewed inactivation, allowing expression of the SRY in the gonadal cell population [15].

We hypothesize that the incomplete masculinisation of our patient is due to activation of the abnormal X to which SRY sequences had been translocated in a subpopulation of gonadal cells. This is the first case of this rare intersex condition reported from the Sudan.

**Competing interests**
The author(s) declare that they have no competing interests.
Figure 4
The PCR analysis showing the presence of SRY material in patient blood. Lane 1 patient sample, lane 2 positive control, lane 3 negative control, lane 6 123 DNA markers.

Authors’ contributions
ME has collected the sample, cultured it, and harvested metaphase chromosomes; ME has also participated in the design and coordination of the study and drafted the manuscript. DG and TN performed the cytogenetic analysis; DG has performed FISH. DG and TN as also participated in the design of the study and helped to draft the manuscript. SA and ME has performed the PCR analysis, and helped to draft the manuscript. TA has performed the ultrasoundography and also helped to draft the manuscript. AE, and IF participated in the design of the study, and helped to draft the manuscript. All authors have given final approval of the final version of this paper.

Note
Table 1: Primer, DNA sequence, and temperature used in DNA analysis

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