Immune checkpoint inhibitors (ICIs), despite their ability to potentiate antitumor T-cell responses, may cause various immune-related adverse events. Most cases of thrombocytopenia induced by ICIs have revealed a pathophysiologic mechanism of immune thrombocytopenia with increased platelet destruction and preserved megakaryocytes. Acquired amegakaryocytic thrombocytopenic purpura (AATP) is an unusual disorder characterized by thrombocytopenia with markedly diminished bone marrow megakaryocytes in the presence of otherwise normal hematopoiesis. AATP caused by ICIs has not been reported on. Herein, we present the case of a 79-year-old man diagnosed with squamous cell carcinoma of the lung who developed AATP after two courses of durvalumab, a drug targeting programmed death-ligand 1. Two weeks after the second cycle, his platelet count decreased to 2.1 × 10^4/μL. After the patient underwent platelet transfusion, his platelet count increased to 8.1 × 10^4/μL the next day but subsequently decreased repeatedly even after the ICI was discontinued. Six weeks after the second cycle, he developed interstitial pneumonia and was administered prednisolone (50 mg/day). However, thrombocytopenia did not improve. Bone marrow biopsy showed scarce megakaryocytes (< 1 megakaryocyte/10 high-power fields) with preservation of myeloid and erythroid series. Myelodysplasia, myelofibrosis, or metastatic lesions were not observed. Cytogenetic analysis showed a normal male karyotype of 46XY. Hence, the patient received eltrombopag, a thrombopoietin receptor agonist, and his platelet count subsequently improved. After recovery, bone marrow aspiration revealed a normal number of megakaryocytes. AATP is rarely the type of thrombocytopenia induced by ICIs and may be successfully treated with thrombopoietin receptor agonists.

Keywords: acquired amegakaryocytic thrombocytopenia, immune-related thrombocytopenia, immune checkpoint inhibitor, durvalumab, eltrombopag

INTRODUCTION

Immune checkpoint inhibitors (ICIs), despite their ability to potentiate antitumor T-cell responses, may cause various immune-related adverse events (irAEs). Hematologic irAEs (hem-irAEs) occur less frequently at a rate of 0.5% for grade ≥ 2 events (graded based on the National Cancer Institute Common Terminology Criteria for Adverse Events, version 4.03). Among the hem-irAEs, immune-related thrombocytopenia is the most common type, along with autoimmune hemolytic anemia and neutropenia, during programmed death-1 /programmed death-ligand 1 (PD-L1) treatment. Most cases of thrombocytopenia induced by ICIs have revealed an immune mechanism with increased platelet destruction and preserved megakaryocytes.

Acquired amegakaryocytic thrombocytopenic purpura (AATP) is a rare hematological disorder manifesting as thrombocytopenia with a remarkable reduction of megakaryocytes in the bone marrow and preserved myeloid and erythroid series. AATP caused by ICIs has not yet been reported on. Herein, we report the case of a patient with squamous cell carcinoma of the lung who developed AATP after two cycles of durvalumab (an ICI targeting PD-L1) and was successfully treated with eltrombopag.

CASE REPORT

A 78-year-old man with stage IV squamous cell carcinoma of the lung was treated with four cycles of nab-paclitaxel and carboplatin in April 2018. However, in March...
2019, tumor enlargement was observed in this patient. Hence, four cycles of nab-paclitaxel and carboplatin were reinitiated, and he also received radiation therapy with 60 Gy/30 fr since July 2019. He was subsequently treated with durvalumab in September 2019 (Day 0). His platelet count was $14.7 \times 10^4/\mu$L, white cell count $4,890/\mu$L with 74.5% neutrophils, and hemoglobin level 9.9 g/dL immediately before the start of durvalumab treatment. Two weeks after the first cycle, his platelet count was $10.9 \times 10^4/\mu$L, and he received a second cycle (Day 14). Two weeks after the second cycle of durvalumab, the platelet count decreased to $2.1 \times 10^4/\mu$L (Day 28). The patient’s white blood cell count was $6,960/\mu$L with 0.5% myelocytes, 68.5% neutrophils, 6.0% lymphocytes, 8.0% monocytes, 15.0% eosinophils, 1.5% basophils, and 0.5% atypical lymphocytes. The hemoglobin level was 10.8 g/dL, and blood film microscopy was negative for schistocytes. The coagulation test results were normal. He had no history of hematologic disease. His regular medication consisted only of silodosin for prostatic hypertrophy, and other than anti-PD-L1 treatment, no other medication was recently prescribed. The third cycle of durvalumab was discontinued. The patient underwent platelet transfusion successfully, considering that his platelet count increased to $8.1 \times 10^4/\mu$L the next day. Despite discontinuing ICI, platelet counts decreased after repeated platelet transfusion (Figure 1).

Six weeks after the second cycle of durvalumab (Day 56), he developed interstitial pneumonia, which was considered to be drug-induced, that is, immune-related interstitial pneumonia. He was admitted to our hospital. His platelet count was $0.7 \times 10^4/\mu$L, white cell count $10,850/\mu$L with 81.0% neutrophils, and hemoglobin level 9.6 g/dL. Treatment with prednisolone (50 mg/day) and antibiotics was initiated. Subsequently, although his dyspnea subsided, thrombocytopenia persisted. The coagulation test results were normal. Antibodies against *Helicobacter pylori* were not detected. The platelet-associated immunoglobulin G level was 243.5 ng/10^7 (normal range < 30.2 ng/10^7). The bone marrow smear test showed moderately hypocellular marrow (nucleated cell count [NCC], $12.6 \times 10^4/\mu$L [normal range, 10–25 × 10^4/\mu$L]). However, megakaryocytes were not observed on the glass slide (megakaryocyte levels < 3.0/µL [normal range, 10–49/µL]), despite the preservation of myeloid, erythroid, and lymphoid series. Myelodysplasia, myelofibrosis, or metastatic lesions were not observed (Figure 2, A). In the bone marrow core biopsy result, moderate hypocellularity of 20%–30% was observed, with significantly rare megakaryocytes (i.e., <1 megakaryocyte/10 high-power fields) (Figure 2, B–C). The thrombocytopenia was considered to be due to AATP caused by ICI. A remarkable decrease in his hemoglobin level was not noticed. Therefore, we considered AATP rather than aplastic anemia. Cytogenetic analysis of the bone marrow showed a normal male karyotype of 46XY. Prednisolone for 13 days failed to improve thrombocytopenia. We initiated eltrombopag treatment on Day 69. In response, the platelet count subsequently improved. Platelet transfusion was no longer required after the last transfusion on Day 89. Ertrombopag was tapered off on Day 155. After recovery (on Day 166), a repeat bone marrow examination revealed a normal number of megakaryocytes (NCC, $16.0 \times 10^4/\mu$L; megakaryocyte level, 28.0/µL) (Figure 2, D–F). The platelet count was in the range of 22.3–48.0 × 10^4/µL until the patient’s death in July 2020 due to exacerbation of his squamous cell carcinoma of the lung.

**DISCUSSION**

ICI treatment can lead to irAEs that involve multiple organs. In some cases, irAEs are recognized as being caused by an auto-inflammatory response driven by systemic activation of innate immunity. In other cases, they are more likely to be autoimmune in nature, with the presence of autoantibodies,
and yet in other cases, antigen-specific memory T-cell responses indicative of adaptive immunity have been documented.³

To date, most cases of thrombocytopenia induced by ICIs have been shown to be of immune origin with increased platelet destruction and preserved megakaryocytes. These patients’ thrombocytopenia was refractory to platelet transfusions and improved with corticosteroid administration. Of the 15 immune-related thrombocytopenic patients, the megakaryocyte levels were categorized as follows: 6, “elevated”; 1, “normal”; 1, “maintained”; and 7, “present” (Table 1). The platelet counts of the eight patients with information regarding platelet transfusion did not increase (Table 1). AATP caused by ICIs has

Fig. 2. Bone marrow examination at disease onset (A-C) and after recovery (D-F): (A) May-Giemsa-stained bone marrow smear showing the absence of megakaryocytes and the maintenance of myeloid, erythroid, and lymphoid series without dysplasia or leukemic cells. (B) Hematoxylin- and eosin-stained sections of bone marrow biopsy showing the absence of megakaryocytes without myelofibrosis. (C) Immunohistochemical staining for cluster of differentiation (CD) 61, a platelet glycoprotein IIa, which is expressed on megakaryocytes and platelets, showing the absence of megakaryocytes. (D) May-Giemsa-stained bone marrow smear showing megakaryocytes. (E) Hematoxylin- and eosin-stained sections of the bone marrow clot showing megakaryocytes. (F) Immunohistochemical staining for CD61 showing megakaryocytes and a myriad of platelets.
central immune cytopenia. Therefore, the patient was started on eltrombopag, which was effective for aplastic anemia derived from immune thrombocytopenic purpura (AATP). Thrombopoietin receptor agonists, such as romiplostim or intravenous immunoglobulin, are effective for AATP. Several patients have reported successful treatment with eltrombopag, although some of them were unresponsive to glucocorticoids, rituximab, or intravenous immunoglobulin. AATP is associated with central immune cytopenia, causing megakaryocyte depletion. Thrombopoietin receptor agonists, such as eltrombopag, are effective for aplastic anemia derived from central immune cytopenia. Therefore, the patient was administered eltrombopag. The patient’s outcome suggested that AATP induced by ICIs may be successfully treated with eltrombopag. However, it is possible that our patient improved spontaneously with the discontinuation of ICIs, considering that the half-life of durvalumab is approximately 18 days.

### CONFLICTS OF INTEREST

The authors declare that they have no conflicts of interest.

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**Table 1. Characteristics of immune-related thrombocytopenia induced by immune checkpoint inhibitors**

| Author *et al.* (year) | Age/sex | Agents/cycle | Platelet count $\times 10^9$/µL | IPF | Megakaryocyte | Platelet transfusion | Treatment |
|------------------------|---------|--------------|-------------------------------|-----|--------------|---------------------|-----------|
| Ahmad *et al.* (2011)  | 57/M    | Ipili/2      | 0.4                           | Not reported | Elevated | No effect | S○ ● ● |
| Kopecký *et al.* (2015)| 54/M    | Ipili/1      | 0.3                           | Not reported | &         | No effect | S○ ● ● |
| Bagley *et al.* (2016) | 34/M    | Nivo/8       | 3.3                           | Not reported | Not examined | Not used | Romiplostim○ |
| Kanameishi *et al.* (2016) | 77/F    | Nivo/2       | 0.2                           | Not reported | #         | Not examined | No effect | S+I+TRA○ |
| Le Roy *et al.* (2016), Burel *et al.* (2017) | 34/M | Nivo/6       | 0.5                           | Not reported | Elevated | No effect | S+I ○ |
| Karakas *et al.* (2017) | 78/M    | Nivo/2,Pem/2 | 1.4                           | Not reported | Not examined | Not used | S○ ● ○ |
| Pföhler *et al.* (2017) | 73/M    | Nivo/2,Pem/2 | 1.4                           | Not reported | Not examined | Not used | S○ ● ● |
| Shiu *et al.* (2017)    | 47/F    | Nivo/Ipili/  | 0.5>                          | Not reported | Not examined | Not examined | No effect | S+I+Romiplostim○ |
|                       | 45/F    | Nivo/Ipili/  | 0.8                           | Not reported | Not examined | Not examined | S+I ○ |
| Burel *et al.* (2017)  | 73/M    | PD1+Ipili/CTLA-4/ | 2.0                         | Not reported | Elevated | Not reported | Not reported |
| Jotatsu *et al.* (2018) | 62/M    | Nivo/2       | 0.16                          | IPF 9.3%       | Elevated | Not used | S○ ● |
| Hasegawa *et al.* (2019) | 82/F    | Nivo/2       | 0.2                           | Not reported | Not examined | Not examined | S+I+TRA○ |
| Mori *et al.* (2019)   | 77/M    | Nivo/1       | 0.2                           | Not reported | Maintained | No effect | S○ ● |
| Delaney *et al.* (2019) | 78/M    | Nivo/2       | 0.5                           | Not reported | Not described | No effect | S○ ○ |
| Mouri *et al.* (2020)  | 66/M    | Pem/1        | 0.3                           | Not reported | Not described | No effect | S○ ● |

$\#$ Bone marrow aspirate analysis in the seven patients with available information about platelet transfusion showed the presence of megakaryocytes.

S, steroids; I, intravenous immunoglobulin; R, rituximab; TRA, thrombopoietin receptor agonist; IPF, immature platelet fraction; Ipili, ipilimumab; Nivo, nivolumab; Pem, pembrolizumab

○, effective; ●, ineffective
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