ArrayExpress update—trends in database growth and links to data analysis tools

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ABSTRACT

The ArrayExpress Archive of Functional Genomics Data (http://www.ebi.ac.uk/arrayexpress) is one of three international functional genomics public data repositories, alongside the Gene Expression Omnibus at NCBI and the DDBJ Omics Archive, supporting peer-reviewed publications. It accepts data generated by sequencing or array-based technologies and currently contains data from almost a million assays, from over 30 000 experiments. The proportion of sequencing-based submissions has grown significantly over the last 2 years and has reached, in 2012, 15% of all new data. All data are available from ArrayExpress in MAGE-TAB format, which allows robust linking to data analysis and visualization tools, including Bioconductor and GenomeSpace. Additionally, R objects, for microarray data, and binary alignment format files, for sequencing data, have been generated for a significant proportion of ArrayExpress data.

INTRODUCTION

The ArrayExpress Archive of Functional Genomics Data (1) is one of the major international repositories for functional genomics high throughput data, supporting publications as well as various data generating consortia. It stores functional genomics data derived from high throughput sequencing (HTS) and microarray-based experiments. Users come to ArrayExpress to (i) find functional genomics experiments that might be relevant to their research; (ii) retrieve information describing these experiments and the data associated with them; (iii) retrieve data for including in their own local data warehouses or added value databases; and (iv) submit their own data supporting a peer-reviewed publication.

Once submitted, data may be kept in ArrayExpress as private for a limited period of time, typically during the peer-review process of the related publication. Upon submission, an accession number is assigned to it and access to the data is restricted to providers/reviewers via a login system. The submitter specifies the release date and the data becomes public either when the accession number associated with the data is cited in a publication or at the set release date, whichever comes first.

All submissions are automatically checked for compliance to the Minimum Information About a Microarray Experiments (MIAME) (2) or Minimum Information about Sequencing Experiments (MINSEQE – http://www.fged.org/projects/minseqe/) guidelines, for microarray and sequencing-based experiments, respectively. The MIAME/MINSEQE scores associated with an experiment are displayed in the ArrayExpress interface and provided to submitters.

In addition to the data submitted directly to ArrayExpress, data from the Gene Expression Omnibus (GEO) (3) are imported to provide users with a single access to most of the functional genomics data available in the public domain. All data are organized, and available for download, in a structured and standardized format, MAGE-TAB (4), which also facilitates linking to open source analysis environments such as Bioconductor (5) and GenomeSpace (http://www.genomespace.org). A format conversion tool, from GEO SOFT to MAGE-TAB (6), is run on all GEO HTS and microarray data. The conversion is successful in 83% of cases; there are various reasons why this conversion may fail, including
failure to parse SOFT files correctly or failure to retrieve the associated data files and we are constantly working with GEO to increase the success rate. All HTS data are exchanged with GEO and a data sharing agreement with the DDBJ Omics Archive is also in place.

For all experiments, the column labels describing the sample (e.g. disease) and its characteristics (e.g. type II diabetes) are mapped to the EBI’s Experimental Factor Ontology (EFO) (8) and the data loaded into ArrayExpress. This allows consistent query results to be returned from direct submissions as well as imported data. As data are curated for Gene Expression Atlas use (9), they are reloaded into ArrayExpress with enriched annotation.

The ArrayExpress user interface allows users to search for experiments of interest by keywords and ontology terms, which enable semantically driven searches of the experimental metadata: for instance searching with the EFO term ‘cancer’ will also find experiments investigating ‘leukemia’ even if ‘cancer’ is not mentioned explicitly. Both US and UK spelling is supported.

DATA GROWTH TO A MILLION ASSAYS

Over the last 2 years, the database content has grown from 13 000 experiments and 370 000 assays, to over 30 000 experiments and almost a million assays. Approximately 20% of the data were submitted directly to ArrayExpress; the rest are imported from GEO weekly.

Although HTS-based experiments account for only 6% of the entire database content, the proportion of new HTS submissions has been growing exponentially over the last few years, from 2% in 2009 to 6% in 2010, 7% in 2011 and 15% in 2012. Nevertheless, the total number of assays associated with HTS-based experiments is still only 3%, reflecting the fact that HTS experiments are typically smaller than microarray-based experiments. If we look at a breakdown of the HTS data by application, 50% of the experiments used RNA-seq only, 32% ChIP-seq only and the remaining experiments either utilized more than one application or used DNA-seq for genotyping, copy number variation detection or methylation profiling.

For HTS data, ArrayExpress stores processed data and metadata describing the sample properties and the experimental design, including experimental variables and protocols, whereas raw sequence data are stored in the European nucleotide archive (ENA) (10) and linked from ArrayExpress. For datasets that require controlled access, the raw sequence data are stored in, and should be submitted directly to, the European Genome-phenome Archive (EGA – www.ebi.ac.uk/ega).

LINKS TO DATA ANALYSIS TOOLS

Approximately 50 GB of data are downloaded every day from ArrayExpress, by an average of 1000 different users. To simplify the interface between ArrayExpress and analytical platforms, we are now providing links to popular analytical tools such as Bioconductor and GenePattern (11), as well as developing robust internal pipelines for HTS data processing.

To facilitate loading microarray data from ArrayExpress into Bioconductor, we have pre-generated R objects for 16 250 out of 25 000 gene expression microarray experiments with raw data files available. A revised version of the Bioconductor package ArrayExpress (12) is used with default parameters. The package has been updated to support popular data formats including Affymetrix and Agilent. More than 85% of Affymetrix data in the repository have downloadable R objects. Older submissions, other technologies and experiments with only processed data available can still be loaded in R, but require user-specified settings for the package to recognize the data format, so loading must be supervised by a user. All pregenerated R objects are now available through the ArrayExpress interface and can be easily loaded into Bioconductor for downstream analysis. More R objects will be created for experiments in ArrayExpress as more data arrive, and the R package will be maintained and extended for this purpose.

Direct links are now provided to GenomeSpace (http://www.genomespace.org), a data analysis environment that makes it possible for users to move data smoothly between popular bioinformatics tools. From ArrayExpress, the user can, with a single click, load a dataset into GenomeSpace, provided that he/she has a registered account with GenomeSpace. Once logged in, the user will be able to utilize the data analysis tools available through GenomeSpace, including GenePattern, Galaxy (13) and Cytoscape (14), to perform data analysis.

For HTS data, the Bioconductor package ArrayExpressHTS (15) and the R-workbench (http://www.ebi.ac.uk/Tools/rcloud/) are used to generate binary alignment (BAM) format files (16). BAM files contain sequence alignment data and can be displayed using the Ensembl genome browser (17), through a direct link from ArrayExpress. So far approximately 1200 BAM files are available for 125 RNA-seq experiments, for 14 different species, with over half of these data studying human and a quarter mouse. The BAM file generation has been done for experiments for which: (i) the sample–data relationship information is available and contains details such as the library strategy and the experiment type (i.e. RNA-seq); (ii) the raw sequence reads (FASTQ files) are deposited in ENA and a valid link to the ENA entry is present; and (iii) the annotation for the reference genome is available in Ensembl.

In addition, 3000 datasets from ArrayExpress have been analysed and the results of this analysis are presented through the Gene Expression Atlas (9), a separate EBI database, which helps users to (i) find out whether the expression of a gene (or a group of genes with a common gene attribute, e.g. GO term) change(s) across all the experiments or (ii) discover which genes are differentially expressed in a particular biological condition of interest.

CONTINUOUS USER INTERFACE IMPROVEMENTS

The ArrayExpress user interface has been continuously improved since the repository was established in
Recent additions include the sample–data relationship viewer (Figure 1), which provides an overview of all samples used in an experiment and their characteristics, the experimental variables (factors) investigated and the data files associated with each sample.

Other improvements include (i) improved array designs browsing and querying for; (ii) specific features for HTS data display; (iii) better organization of the species drop-down filter, and (iv) improved performance for retrieving and visualizing large experiments.

The ArrayExpress user documentation has recently been updated and several online courses, covering how to search, interpret and submit data to ArrayExpress, can be found on the EBI e-Learning portal, Train online (http://www.ebi.ac.uk/training/online/).

**FUTURE DEVELOPMENTS**

We are currently developing a new submission tool, optimized for supporting HTS data submissions; this new tool is based on the community developed annotation tool Annotare (19) and will be released in 2013.

Like all other major EBI data resources, ArrayExpress is working toward deeper integration in the overall EBI infrastructure, in particular with the BioSample Database (20), the Gene Expression Atlas and the sequence databases ENA, EGA and Ensembl. We will continue this integration effort to ensure that our users can obtain a systems level view of the data stored at EBI by easily navigating through our resources.

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