Enhanced Biogas Production of Cassava Wastewater Using Zeolite and Biochar Additives and Manure Co-Digestion

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Abstract: Currently, there are challenges with proper disposal of cassava processing wastewater, and a need for sustainable energy in the cassava industry. This study investigated the impact of co-digestion of cassava wastewater (CW) with livestock manure (poultry litter (PL) and dairy manure (DM)), and porous adsorbents (biochar (B-Char) and zeolite (ZEO)) on energy production and treatment efficiency. Batch anaerobic digestion experiments were conducted, with 16 treatments of CW combined with manure and/or porous adsorbents using triplicate reactors for 48 days. The results showed that CW combined with ZEO (3 g/g total solids (TS)) produced the highest cumulative CH4 (653 mL CH4/g VS), while CW:PL (1:1) produced the most CH4 on a mass basis (17.9 mL CH4/g substrate). The largest reduction in lag phase was observed in the mixture containing CW (1:1), PL (1:1), and B-Char (3 g/g TS), yielding 400 mL CH4/g volatile solids (VS) after 15 days of digestion, which was 84.8% of the total cumulative CH4 from the 48-day trial. Co-digesting CW with ZEO, B-Char, or PL provided the necessary buffer needed for digestion of CW, which improved the process stability and resulted in a significant reduction in chemical oxygen demand (COD). Co-digestion could provide a sustainable strategy for treating and valorizing CW. Scale-up calculations showed that a CW input of 1000–2000 L/d co-digested with PL (1:1) could produce 9403 m3 CH4/yr using a 50 m3 digester, equivalent to 373,327 MJ/yr or 24.9 tons of firewood/year. This system would have a profit of $5642/yr and a $47,805 net present value.

Keywords: methane; fermentation; dairy; poultry; absorbent

1. Introduction

Eutrophication and organic pollution resulting from poor management of wastes from food processing industries, such as cassava processing industries, is a major problem in many developing countries [1,2]. Cassava (Manihot esculenta) is a starch-containing root crop of global importance that can be processed into food, feed, and other non-food products [3]. The cassava processing industry is a key industry in many developing countries, especially in Africa, but also in parts of Latin America and Asia. In 2017, the African region contributed 55% of the global production of cassava, equivalent to 121 million tons, with 25–37% of the crop discarded as waste in the form of peels and pulp [4,5], and approximately 60,000 L of effluent generated from each ton of cassava tubers processed [3,6]. Meanwhile, more than 70% of cassava production in sub-tropical and tropical regions of the world is conducted by small and medium-scale farmers [7,8]. These small and medium-scale cassava industries lack the capacity to treat the large waste streams resulting from daily cassava roots processing, which can lead to environmental degradation and pollution of nearby water bodies [9]. Additionally, farmers
often depend solely on firewood as a source of energy to process cassava, which has associated negative environmental impacts.

Bioenergy from organic wastes materials through anaerobic digestion (AD) can be used to produce renewable energy from this organic-rich wastewater, while reducing the concentrations of organic pollutants [10]. Cassava wastewater (CW) has a high organic loading, with high concentrations of chemical oxygen demand (COD), biochemical oxygen demand (BOD), and total solids (TS), as well as a low pH [3,11]. In parts of Thailand, Brazil, Vietnam, and India, CW has been managed using stabilization ponds, aerobic systems, and AD [12]. Reported concerns associated with digestion of CW are the low nitrogen concentration and rapid acidification (low pH) of CW [13,14]. Co-digestion with a nitrogen-rich substrate, such as manure, could decrease the carbon-to-nitrogen (C:N) ratio and provide buffering capacity for stabilizing the pH in order to increase methane (CH\textsubscript{4}) production. Previous studies have investigated cassava peels and pulps co-digested with livestock wastes [2,15], digestion of cassava starch effluent with separation of the acidogenic and methanogenic phases [16], re-circulation of methanogenic sludge [17], dolomitic limestone addition to increase alkalinity [13], and use of up flow anaerobic sludge blanket (UASB) digestion processing [18,19].

Porous adsorbents, such as biochar, zeolite, and activated carbon, have been used to enhance CH\textsubscript{4} production and general AD processes [20]. Biochar is a carbonaceous material obtained from agricultural biomass through pyrolysis and gasification. Mumme et al. reported that biochar is relatively cheaper to manufacture than other adsorbents, which has increased interest in land application of biochar, and more recently, inclusion in AD processing [21]. The use of biochar as an additive in AD has not been fully investigated, and no work has been done with biochar and AD of CW. There is potential for biochar to enhance the operational stability of the AD process and increase the quality of the digestate produced.

Zeolite has also been reported to possess favorable characteristics for microorganism adhesion [22], with the capacity to induce ion exchange during AD due to the presence of Na\textsuperscript{+}, Ca\textsuperscript{2+}, and Mg\textsuperscript{2+} cations in its crystalline structure. These properties could be useful for improving AD of wastewaters with high concentrations of nitrogen, such as poultry manure, as it prevents process inhibition. Application of natural zeolites as support media in digesters treating wastewaters has been reported to increase microbial population density and provide greater opportunity for microbial growth and attachment, cross feeding, co-metabolism, and interspecies hydrogen and proton transfer [22].

A prior study by Montalvo et al. reported that the addition of natural zeolite at doses between 2 and 4 g/L increased CH\textsubscript{4} production, with increasing inhibition at doses >6 g/L [23]. The use of porous materials, such as natural zeolites, to create surface area for microbial communities and increase retention of high biomass concentrations in the digestion of wastewater has been documented [22]. Zeolite was employed to enhance energy recovery, in the form of hydrogen (H\textsubscript{2}), from cassava-ethanol wastewater during the dark fermentation process [24]. To our knowledge, zeolite additions have not been used during the digestion of CW to enhance CH\textsubscript{4} production.

General implementation and adoption of large-scale biogas technology in most African countries have been limited due to the high costs associated with investments and operations of AD systems [25], especially when there is an additional cost of transporting wastes to offsite AD reactors. The availability of agricultural biomass, which is abundant in the rural cassava industry, along with the large volume of CW generated daily in this industry, would provide larger quantities of organic material to be treated onsite using AD. This study focused on investigating the impact of the co-digestion of CW with selected livestock manures and porous adsorbents on biogas production for potential implementation and adoption in cassava industries. The objectives of the study were to: (1) characterize cassava wastewater as a substrates for AD and identify appropriate substrates for co-digestion, (2) investigate the CH\textsubscript{4} potential of cassava digestion, with and without co-digestion with manure, biochar, and zeolite, in terms of cumulative production and retention time, (3) characterize the wastewater transformations during digestion in terms of organic and nutrient transformations, and (4) analyze the economic viability and environmental impact of employing digestion for the rural cassava industry.
2. Materials and Methods

2.1. Substrate and Inoculum Collection and Preparation

2.1.1. Cassava Wastewater Substrate

The cassava tubers were obtained from a farmer’s market in Adelphi, Maryland, USA. The cassava tubers were manually peeled and soaked for 5 days in the laboratory using 1 L/kg of deionized water to replicate the rural cassava processing steps for ‘fufu’ production, a popular African dish derived from fermented cassava paste. Fermented tubers were manually squeezed, and the cassava wastewater (CW) was collected and used for the experiments. The substrate characteristics and the experiment design are given in Tables 1 and 2. The CW had a COD range of 29.8–33.4 g/L, volatile solids (VS) of 17.3 g/kg, total solids (TS) of 17.8 g/kg, and a pH of 5.5.

| Table 1. | Experimental design showing grams (g) of substrate addition into each triplicate 250 mL reactor. |

| Treatment | Cassava Wastewater (CW) (g) | Zeolite (ZEO) (g) | Biochar (B-Char) (g) | Poultry Litter (PL) (g) | Dairy Manure (DM) (g) | Inoculum (g) | Water (g) |
|----------|-----------------------------|------------------|---------------------|------------------------|---------------------|--------------|-----------|
| CW + PL + ZEO (HC) | 28.9 | 3.4 | 0.0 | 0.8 | 0.0 | 92.1 | 28.2 |
| CW + PL + B-Char (HC) | 28.9 | 0.0 | 3.4 | 0.8 | 0.0 | 92.1 | 28.2 |
| CW:PL (1:1) | 28.9 | 0.0 | 0.0 | 0.8 | 0.0 | 92.1 | 28.2 |
| CW:PL (2:1) | 38.6 | 0.0 | 0.0 | 0.5 | 0.0 | 92.1 | 18.9 |
| CW + DM + ZEO (HC) | 28.9 | 3.4 | 0.0 | 0.0 | 4.4 | 92.1 | 24.7 |
| CW:DM (1:1) | 28.9 | 0.0 | 0.0 | 0.0 | 4.4 | 92.1 | 24.7 |
| CW:DM (2:1) | 38.6 | 0.0 | 0.0 | 0.0 | 2.9 | 92.1 | 16.5 |
| CW + ZEO (HC) | 57.8 | 0.3 | 0.0 | 0.0 | 0.0 | 92.1 | 0.1 |
| CW + ZEO (HC) | 57.8 | 1.5 | 0.0 | 0.0 | 0.0 | 92.1 | 0.1 |
| CW + B-Char (LC) | 57.8 | 0.0 | 0.3 | 0.0 | 0.0 | 92.1 | 0.1 |
| CW + B-Char (HC) | 57.8 | 0.0 | 1.5 | 0.0 | 0.0 | 92.1 | 0.1 |
| CW-only | 57.8 | 0.0 | 0.0 | 0.0 | 0.0 | 92.1 | 0.0 |
| Inoculum-only | 0.0 | 0.0 | 0.0 | 0.0 | 0.0 | 91.9 | 0.0 |

| Table 2. | Characterization of the substrates, cassava wastewater (CW), poultry litter (PL), and dairy manure (DM) and the inoculum source, including total solids (TS), volatile solids (VS), chemical oxygen demand (COD), pH, total Kjeldahl nitrogen (TKN), total phosphorus (TP), and the carbon to nitrogen ratio (C:N). |

| TS (g/kg) | VS (% TS) | COD (g/L) | pH | TKN (mg N/L) | TP (mg P/L) | C:N Ratio |
|----------|-----------|-----------|-----|--------------|-------------|-----------|
| CW | 17.8 ± 0.7 | 97.2 ± 0.7 | 33.7 ± 0.8 | 5.53 | 375 | 222 | 27.8 |
| PL | 776 ± 1 | 80.0 ± 0.2 | NA | 8.25 | 3675 | 1245 | 13.0 |
| DM | 131 ± 2 | 87.5 ± 0.6 | NA | 7.33 | 3450 | 603 | 15.2 |
| Inoculum | 29.5 ± 0.1 | 73.6 ± 7.0 | 25.1 ± 0.3 | 7.55 | 3050 | 1225 | 3.91 |

2.1.2. Dairy and Poultry Manure Substrates and Inoculum Source

The dairy manure (DM) used as a co-substrate was obtained from the 100-cow dairy at the US Department of Agriculture (USDA) Beltsville Agricultural Research Service (ARS) in Beltsville, MD. Poultry litter (PL) was obtained from a poultry (broiler) farm at the University of Maryland Extension—Talbot county, Easton Maryland. The poultry litter consisted of poultry droppings and beddings from wood shavings. Both manure substrates were collected onsite and stored at 4 °C before use. The inoculum used for the experiment was digestate of a complete mixed wastewater sludge digester (Alexandria, VA, USA) and was stored at 4 °C prior to use.

2.1.3. Biochar and Zeolite Additives

Two porous materials, biochar (B-Char) and clinoptilolite zeolite (ZEO), were added to the CW as co-treatments. The biochar (B-Char) substrate was derived from corn stover prepared through pyrolysis under an O₂-free atmosphere at 500 °C, with a holding time of 10 min (ArtiCHAR, Prairie
City, Iowa, USA). The biochar particle size varied from 841 mm to <74 mm, with a VS and TS of 690 and 980 g/kg, respectively. The zeolite was a high purity 97% clinoptilolite zeolite produced at Amargosa Valley (Nye county, NV, USA). The zeolite used was in the form of granules with an angular shape and gray color. The pore diameter was between 4.0–7.0 angstroms.

2.2. Experimental Design

A batch digestion experiment was conducted based on the biochemical methane potential (BMP) test following methods by Moody et al. [26]. The digestion tests were conducted at the University of Maryland’s Department of Environmental Science and Technology (ENST) Water Quality Laboratory (College Park, MD USA). Prior to starting BMP tests, the TS and VS for CW, PL, DM, and inoculum were determined and used to combine the co-substrate ratios based on VS. The experiment was designed for 16 treatments, with three replicates for each treatment (48 total digestion reactors). Each digestion reactor consisted of a 250 mL serum bottle, with the substrates and inoculum loaded at a 2:1 inoculum to substrate ratio (ISR) based on VS and operated in mesophilic conditions (35 °C). For all treatments, an equal volume of inoculum (92.1 g) was added to each triplicate reactor.

The biochar (B-Char) and zeolite (ZEO) treatments were prepared using a low concentration (LC) (0.5 g adsorbent/g TS of substrate) and a high concentration (HC) (3 g adsorbent/g TS of substrate) added to 57.82 g CW. For the manure co-digestion treatments, 3.42 g of PL and 3.26 g of DM were digested alone and co-digested with 28.91 g CW with and without the HC of ZEO. Additionally, PL was co-digested with 28.91 g CW and the HC of B-Char. Inoculum-only reactors were also incubated, and the CH₄ production from the inoculum was subtracted from each treatment to account for residual CH₄ production from organics in the inoculum.

Prior to incubation, the headspace in each vessel was purged with N₂ for three minutes to ensure anaerobic conditions and immediately capped with a rubber septum, and the bottles were placed on a shaker (120 rpm) in a controlled environmental chamber at 35 °C for 48 days. The daily biogas volume was measured by volumetric displacement using a graduated, gas-tight, wet-tipped 50 mL glass syringe inserted through the top of the rubber septum. Biogas production was quantified volumetrically at normal temperature and pressure conditions using a glass gas-tight syringe, equilibrated to atmospheric pressure [26]. All CH₄ production values are reported in normal temperature and pressure conditions (1 atm and 20 °C).

2.3. Analytical Methods

The pH of substrates and inoculum were determined with an Accumet AB 15 pH meter (Fisher Scientific, Hampton, NH). For all samples, TS (Method 2540B) and VS (Method 2540E) concentrations were determined using standard methods for the examination of water and wastewater [27]. Total Kjeldahl nitrogen (TKN) and total phosphorus (TP) samples were analyzed on a Lachat autoanalyzer (Quikchem 8500, Hach Company, Loveland, CO, USA) using QuikChem methods 13-107-06-2-D for TKN and 13-115-01-1-B for TP. The COD concentration was measured using a Hach DR 5000 spectrophotometer (Hach Company, Loveland, CO, USA).

The carbon content of the CW, PL, DM, and inoculum were calculated using the equation from Adams et al. [28], where % Carbon = % VS/1.8.

Biogas was analyzed for CH₄ and CO₂ content by injecting 0.10 mL of gas sample using a luer-lock, gas-tight syringe into an Agilent HP 7890 A gas chromatograph (Agilent Technologies, Santa Clara, CA, USA) equipped with a thermal conductivity detector (TCD) and single HP porous layer open tubular (PLOT) Q column with an injection temperature of 250 °C, a detector temperature of 250 °C, an oven temperature of 60 °C, and conveyed using He gas at a flow rate of 8.6 mL He/min [10].

2.4. Statistical Analysis

Cumulative CH₄ production was analyzed using analysis of variance (ANOVA) to determine significantly differences, with p-values < 0.05 considered significant. Tukey’s honestly significant
difference (HSD) post-hoc tests were performed for multiple comparisons between variables based on different digestion periods during the 48 days of the experiment. All results presented in the tables and charts are average values with standard error (SE).

3. Results and Discussion

3.1. Characterization of Substrate and Inoculum

The pH of the CW substrate was between 5.5 and 6.5 (Table 2). Some studies have reported lower pH values for CW, ranging from 3.9–4.5 [29]. The pH of mixed substrates before and after AD was within the ideal pH range (6.5–8) for CH₄ production [30]. The TS and VS of the cassava wastewater (17.8 and 17.3 g/L, respectively) was 75.8% and 60.4% lower than PL respectively, and 11.3 and 9.7% lower than DM, respectively. As the CW was a liquid wastewater, it was a more dilute waste stream than the manure substrates and had comparatively less TS and VS.

The TKN and TP of the CW were 375 and 222 mg/L respectively (Table 2), whereas, the PL had higher TKN and TP values (3675 and 1245 mg/L, respectively), which were similar to DM (3450 and 603 mg/L, respectively). The pH of the CW substrate was between 5.5 and 6.5 (Table 2). Some studies have reported lower pH values for CW observed in this study was consistent with findings from others CW studies [1,2,8]. The C:N ratio of the CW substrate was 27.8, which was lower than DM, respectively. As the CW was a liquid wastewater, it was a more dilute waste stream than the manure substrates and had comparatively less TS and VS.

The low nitrogen content of CW observed in this study was consistent with the value 29.1 reported by Lin et al. [31]. The carbon to nitrogen ratio is a key factor affecting anaerobic digestion [32], with C:N ratios between 25 and 30 reported as the most suitable for CH₄ production [2,31]. The C:N ratio for PL and DM in this experiment was 13.0 and 15.2 respectively, which were lower than the optimal conditions, while the CW was higher. When the substrates were combined, the C:N ratio of the mixtures were 20.4 and 21.1, for PL and DM, respectively.

3.2. Effect of Livestock Manure Co-Digestion with Cassava Wastewater on Biogas Production

3.2.1. Cumulative CH₄ Production Based on VS Addition into the Digestion Reactor

After 48 days of digestion, the cumulative CH₄ production (on a per g VS-basis) from CW-only (620 mL CH₄/g VS) was 15.8% higher than co-digestion of CW:DM at a 2:1 ratio (522 mL CH₄/g VS; p-value < 0.001; Figure 1; Table 3). The CW-only digestion had 5.8% higher CH₄ production than CW:PL at a 2:1 ratio (590 mL CH₄/g VS), but this difference was not statistically significant (p-value = 0.864). Similarly, CW-only was 14.4% and 25.6% higher respectively, than CW co-digested at a 1:1 ratio with DM and PL (461 and 531 mL CH₄/g VS; p-values < 0.001 and 0.001, respectively).

Table 3. Reductions in volatile solids (VS) and chemical oxygen (COD) during digestion for the substrate and inoculum in each reactor. The cumulative methane (CH₄) production is given using two normalizations: per g VS added and per g of total substrate added. Superscript letters (a through g) indicate significant differences within each column at p-value < 0.05.

| Substrate | VS Reduction (%) | Influent COD (g/L) | COD Reduction (%) | Cumulative CH₄ (ml CH₄/g VS) | Cumulative CH₄ (ml CH₄/g Substrate) |
|-----------|------------------|--------------------|-------------------|-------------------------------|-----------------------------------|
| CW-only   | 65.5 ± 0.1 a     | 29.6 ± 0.4 ab      | 40.6 ± 2.9 a      | 620 ± 6.0 abc                 | 107 ± 0.1 a                       |
| CW + B-Char (HC) | 37.5 ± 0.1 b     | 41.9 ± 0.7 c      | 23.6 ± 7.5 b      | 611 ± 27 bc                   | 10.6 ± 0.5 a                      |
| CW + B-Char (LC)  | 62.7 ± 0.3 a     | 34.9 ± 2.4 d      | 48.8 ± 5.3 a      | 611 ± 16 a                    | 10.6 ± 0.3 a                      |
| CW + ZEO (HC)    | 66.0 ± 1.9 a     | 33.7 ± 0.8 de     | 49.2 ± 1.6 a      | 653 ± 4 a                     | 11.3 ± 0.1 ab                     |
| CW + ZEO (LC)    | 66.2 ± 2.6 a     | 32.8 ± 1.0 de     | 46.2 ± 1.0 a      | 634 ± 6 b                     | 11.0 ± 0.1 ab                     |
| CW + PL + B-Char (HC) | 6.88 ± 1.4 e   | 41.6 ± 0.6 f      | 31.1 ± 1.2 g      | 471 ± 16 d                    | 15.9 ± 0.5 fce                    |
| CW + PL + ZEO (HC) | 61.4 ± 0.6 a     | 31.6 ± 2.1 a      | 44.0 ± 3.4 a      | 518 ± 8 e                     | 17.4 ± 0.3 fce                    |
| CW + DM + ZEO (HC) | 57.3 ± 0.2 a     | 25.8 ± 0.3 f      | 21.9 ± 6.1 b      | 473 ± 5 d                     | 14.2 ± 0.2 fbe                    |
| CW:PL (1:1)      | 63.5 ± 0.6 b     | 28.8 ± 2.0 b      | 42.7 ± 3.2 a      | 531 ± 10 e                    | 17.9 ± 0.3 c                      |
| CW:PL (2:1)      | 64.8 ± 0.5 a     | 28.9 ± 0.3 b      | 39.6 ± 4.9 a      | 590 ± 6 c                     | 15.1 ± 0.2 fce                    |
| CW:DM (1:1)      | 59.9 ± 1.0 a     | 29.5 ± 0.9 ab     | 37.4 ± 0.5 a      | 461 ± 17 d                    | 13.8 ± 0.5 bce                   |
| CW:DM (2:1)      | 63.4 ± 0.3 a     | 32.5 ± 1.9 a      | 47.8 ± 2.7 a      | 522 ± 14 e                    | 12.6 ± 0.3 bce                   |
| DM-only          | 48.4 ± 5.3 d     | 28.0 ± 0.7 b      | 20.9 ± 2.9 b      | 100 ± 5 f                     | 22.9 ± 1.1 i                      |
| PL-only          | 63.0 ± 8.4 a     | 28.2 ± 1.3 b      | 20.7 ± 0.1 b      | 156 ± 3 g                     | 193 ± 4 h                        |

a negative value indicates a percent increase due to addition of biochar not included in pre-COD.
When CH₄ production is normalized on a VS basis, the efficiency of the organic material to CH₄ conversion process is shown. These results show that CW can be co-digested or digested alone, and co-digestion of CW with manure resulted in similar or slightly lower CH₄ production efficiency values (5.8%–25.6% decrease with co-digestion). It should be noted that the inoculum included in the digestion reactor helped to lower the C:N ratio from 27.8 in the CW substrate to 21.7 in the digestion reactor with the inoculum and CW mixture, with an increase in the pH value from 5.53 to 7.75 due to inoculum inclusion. The significance of pH as a key determining factor for AD process, especially in full-scale continuous reactors were highlighted in Calabrò et al. [33,34]. In field conditions, it would be important to have a viable inoculum source for initiating digestion and to consider a co-digestion material that can help to neutralize the low pH and is high in nitrogen to ensure that the microbes are not nitrogen-limited.

3.2.2. Cumulative CH₄ Production Based on the Mass of Substrate Added to the Digestion Reactor

Due to the high VS concentration of the manure substrates, the PL-only reactors had an order of magnitude higher CH₄ production on a mass basis (193 mL CH₄/g substrate) than DM-only (22.9 mL CH₄/g substrate) and all CW reactors (Table 3). The DM and PL manure substrates had 84.8% to 97.2% higher VS concentrations respectively, than the CW substrate (Table 2). Digestion of CW yielded 83.9% and 74.8% more cumulative CH₄ (on a VS-basis) than DM-only and PL-only digestion (100 and 156 mL CH₄/g VS, respectively; Table 2), as the organic matter in the CW substrate was converted into CH₄ more efficiently than the manure substrates, likely due to the more recalcitrant nature of the VS in the complex manure substrates compared to the cassava wastewater. The CW:PL (1:1) had the highest CH₄ production (17.9 mL CH₄/g substrate) of the CW co-digestion treatments (on a mass basis). The PL co-substrate had higher CH₄ production efficiencies than CW co-digested with DM at both the 1:1 and 2:1 ratios (p-values = 0.017 and 0.025, respectively), indicating that a continuously-fed CW digestion system would benefit from co-digesting with PL due to the high organic loading of the PL substrate.
3.2.3. Cumulative CH₄ Production Based on Digestion Period

While the overall CH₄ production efficiency from the CW-only was higher than the manure substrates, the CW-only treatment produced 118 mL CH₄/g VS (19% of the cumulative CH₄) in the first 15 days of the 48-day digestion period (Table 4; Figure 2). During this first third of the digestion period (Days 1–15), the CH₄ production from the CW-only treatment was significantly lower (38.6%) than CW:DM at 1:1 (265 mL CH₄/g VS) and 32.6% lower CW:PL at 1:1 (274 mL CH₄/g VS; p-values < 0.001). When the ratio of CW to manure was doubled (2:1), the CH₄ production from Days 1–15 in the CW:PL (2:1) and CW:DM (2:1) reactors was 263 and 211 mL CH₄/g VS respectively, which was 25.7 and 21.5% higher than CW-only (p-value < 0.001 and 0.002, respectively).

Table 4. Cumulative methane (CH₄) during the 48-day digestion period for designated time periods, with the percent of the total cumulative CH₄ production in parenthesis. Superscript letters (a through g) significant differences within each column at p-value < 0.05.

| Treatment | 9 Days | 15 Days | 20 Days | 37 Days | 48 Days |
|-----------|--------|---------|---------|---------|---------|
| CW-only   | 26.8 (4.3%) abc | 118 (19%) a | 364 (58.7%) ab | 598 (96.4%) abc | 620 (100%) abc |
| CW + B-Char (HC) | 39.9 (6.5%) def | 265 (43.4%) b | 403 (66%) bc | 589 (96.4%) bc | 611 (100%) bc |
| CW + B-Char (LC) | 37.8 (6.2%) def | 314 (48.1%) c | 425 (65.2%) c | 634 (97.1%) c | 653 (100%) c |
| CW + ZEO (HC) | 39.3 (6%) def | 189 (29.8%) d | 370 (58.4%) d | 614 (96.8%) d | 634 (100%) d |
| CW + ZEO (LC) | 31.5 (5%) de | 400 (64.8%) e | 419 (68.9%) e | 459 (77.4%) e | 471 (100%) e |
| CW + PL + B-Char (HC) | 273 (57.2%) g | 255 (49.2%) b | 370 (71.4%) b | 497 (95.9%) f | 518 (100%) e |
| CW + PL + B-Char (LC) | 373 (71.5%) e | 197 (41.7%) d | 323 (68.5%) d | 447 (94.7%) d | 473 (100%) d |
| CW + PL + ZEO (HC) | 43.2 (8.3%) d | 255 (49.2%) b | 370 (71.4%) b | 497 (95.9%) f | 518 (100%) e |
| CW + PL + ZEO (LC) | 47.3 (9.4%) e | 197 (41.7%) d | 323 (68.5%) d | 447 (94.7%) d | 473 (100%) d |
| CW + DM + ZEO (HC) | 36.2 (5.5%) e | 263 (44.7%) b | 366 (62%) b | 571 (96.8%) c | 590 (100%) c |
| CW + DM + ZEO (LC) | 46.5 (10.1%) d | 263 (57.6%) b | 353 (66.6%) b | 439 (95.3%) b | 461 (100%) d |
| CW:DM (1:1) | 25.4 (4.9%) bc | 211 (40.5%) d | 336 (64.3%) d | 484 (92.8%) ef | 522 (100%) e |
| CW:PL (2:1) | 34.8 (34.6%) e | 48.4 (48.2%) f | 62.6 (62.3%) e | 89.1 (88.8%) g | 100 (100%) f |
| PL-only | 92.3 (59%) h | 114 (72.9%) * | 128 (82%) f | 148 (95%) b | 156 (100%) g |

Figure 2. Cumulative methane (CH₄) production based on volatile solids (VS) added to each reactor for cassava wastewater (CW) digested alone and co-digested with poultry litter (PL), dairy manure (DM), zeolite (ZEO), and biochar (B-Char) at low and high concentrations (LC and HC) at five time points (Days 9, 15, 20, 37, and 48) in the 48-day digestion period.
There was no significant difference in cumulative CH$_4$ production between CW:PL at the 1:1 and 2:1 ratios (p-value = 0.087). The CW:PL (2:1) had 4.1% less CH$_4$ in the first 15 days of digestion, showing a slight decrease in lag phase during digestion without a significant effect on the overall CH$_4$ production potential (Figure 2). A similar trend was observed when comparing the CW:DM at 1:1 and 2:1 ratios, with 265 and 211 mL CH$_4$/g VS respectively, in the first 15 days of digestion, which were not significantly different (p-value of 0.061).

Generally, co-digestion of CW with PL or DM reduced the lag phase for CH$_4$ production. The highest rate of CH$_4$ production occurred within the first two weeks, which is consistent with the results from Witarsa and Lansing [30], where a large percentage of CH$_4$ production from digestion of separated and unseparated dairy manure (DM) occurred in the first 16 days of a 216 day digestion period (40% and 36%, respectively). Cassava wastewater (CW-only) generated 19% of the total cumulative CH$_4$ in the first 15 days of our 48-day digestion period, while DM-only generated 48.2% of the total cumulative CH$_4$ and CW:DM (1:1) generated 57.6% of the cumulative CH$_4$ production in the first 15 days.

The observed increase in CH$_4$ production with co-digestion of CW within the first two weeks was consistent with other findings, which emphasized the advantages of co-digestion over single digestion of substrates [2,32,35–37]. Panichnumsin et al. [2] examined the potential of co-digestion of cassava pulp and swine manure in a semi-continually fed stirred tank reactor in mesophilic conditions (37 °C) at a constant organic loading rate of 3.5 kg VS/m$^3$d for 15 days and reported a 41% increase in CH$_4$ yield compared with digestion of swine manure alone. Similar to our study, a batch experiment conducted by Riano et al. [37] at 35 °C for 55 days reported that co-digestion of winery wastewater (10–40%) and swine manure increased CH$_4$ production by 45–75% and improved digestion stability compared to digestion of swine manure alone.

Abouelenien et al. [35] co-digested, poultry manure (PM) with mixed agricultural wastes comprised of coconut wastes, cassava wastes, and coffee grounds. The cassava waste used in their study was root residue and wet cake from cassava, while our study utilized cassava wastewater. Similar to our study, co-digestion was conducted under mesophilic conditions (35 °C) and saw an increase in CH$_4$ yield of up to 50% (506 mL CH$_4$/g VS) compared to PM-only after 40 days of digestion. Their results were comparable to our study, with cumulative CH$_4$ production of 531 mL CH$_4$/g VS for CW:PL (1:1) after 48 days, which was significantly (p-value = 0.001) higher than PL-only, which yielded only 156 mL CH$_4$/g VS. Contrary to our findings, Abouelenien et al. [37] reported an elongation of the lag phase due to co-digestion, which was attributed to the complex organic matter in the mixed agricultural wastes compared to the PM substrate. Whereas in our study, a reduction in lag phase was recorded due to the liquid state of the CW substrate, which was more readily accessible for the rate-limiting hydrolysis phase of digestion.

### 3.3. Impact of Porous Adsorbent on AD of Cassava Wastewater

#### 3.3.1. Zeolite Addition with Cassava Wastewater Digestion

Digesting CW and a high concentration (HC) of zeolite (CW + ZEO-HC) produced the highest cumulative CH$_4$ (653 mL CH$_4$/g VS) for all treatments after 48 days, followed by the treatment with a lower concentration (LC) of zeolite (CW + ZEO-LC), which produced 634 mL CH$_4$/g VS, with no significant different between the two zeolite concentrations (p-value = 1.00; Figure 1; Table 2). The two porous adsorbents used in this study at the HC were also not significantly different (p-value = 0.50), with the cumulative CH$_4$ produced from CW + ZEO-HC (653 mL CH$_4$/g VS) only slightly higher than CW + B-Char-HC (611 mL CH$_4$/g VS).

After 9 days of digestion, a significantly higher percentage of the total CH$_4$ production (37.8%) was observed in CW + ZEO-HC compared to CW-only (Table 3; Figure 2). This observation is consistent with Milan et al., where doses of zeolite between 2 and 4 g/L increased CH$_4$ production of swine manure and of zeolite, while doses above 6 g/L inhibited the process [38]. In our study, the effect of
3.3.2. Biochar Addition with Cassava Wastewater Digestion

Similarly, an increase in CH\textsubscript{4} production with a shortened lag phase was observed due to biochar addition (Figure 2). The effect of biochar addition in reducing the lag phase in AD has been previously reported [39,40]. Jang et al. showed a 24.9% increase in cumulative CH\textsubscript{4} (467 mL CH\textsubscript{4}/g VS) with 10 g/L of biochar compared to 1 g/L of biochar (395 mL CH\textsubscript{4}/g VS) with mesophilic conditions and 40 days of digestion [40]. Our findings showed that on Day 15, CW + B-Char-HC had 76.8% more CH\textsubscript{4} production than the CW-only treatment, which Jang et al. suggested was due to the high alkalinity of biochar enhancing CH\textsubscript{4} production and shortening the lag phase [40].

Comparing the concentrations of porous adsorbent added, there was a significant difference in cumulative CH\textsubscript{4} production between the low and high concentrations of biochar (p-value < 0.001) on Day 15 of digestion (Figure 2). In the first two weeks of digestion, CW + B-Char-HC yielded 265 mL CH\textsubscript{4}/g VS, while CW + B-Char-LC yielded 133 mL CH\textsubscript{4}/g VS, illustrating the decrease in lag phase with an increase in the quantity of biochar added.

Comparing the LC and HC of zeolite showed no significant difference (p-value = 1.000) after 48 days of digestion. The ZEO-LC and ZEO-HC at Day 15 produced 8.1% and 4.7% more CH\textsubscript{4} than B-Char-LC and HC, respectively. Yet, by 48 days, there were no significant differences between LC and HC of B-Char and ZEO (p-values = 0.974 and 1.000, respectively). The observed lag in digestion in the mixtures containing CW alone or low concentrations of zeolite or biochar could be as a result of the rapid acidification of CW and inadequate buffer to provide the necessary buffer for microbial community and methanogens, and thus, a longer lag phase for microbial recovery.

The combined effects of manure and biochar showed that CW + PL + B-Char produced significantly more CH\textsubscript{4} at Days 15 and 20 (273 and 400 mL CH\textsubscript{4}/g VS, respectively; p-values < 0.001) than CW + B-Char (39.9 and 265 mL CH\textsubscript{4}/g VS, respectively). After Day 20, the daily CH\textsubscript{4} production of CW + PL + B-Char decreased, while CW + B-Char increased and resulted in higher cumulative CH\textsubscript{4} over 44 days. While the addition of PL or DM to CW increased CH\textsubscript{4} production in the first 15 days of digestion, the CW + PL + B-Char-HC treatment yielded 400 mL CH\textsubscript{4}/g VS (84.8% of total cumulative CH\textsubscript{4}) in the first 15 days, with this reduction is lag phase likely attributed to the combined presence of biochar and poultry litter.

The ability of biochar to catalyze digestion by providing surface area for the colonization of the microbial cell was previously reported in a review by Mumme et al. [21]. The CW substrate used in our experiment contained a low pH and when co-digested with biochar and manure showed an improved AD process due to the buffer provided by manure and biochar [20], as observed in the first two weeks of our experiment.

3.4. Volatile Solids and COD Reduction during Digestion

Chemical oxygen demand (COD) and VS reduction is associated with CH\textsubscript{4} production. The substrate mixture containing CW + ZEO-HC showed the highest VS and COD reductions during digestion (66% and 49%), which corresponded with the highest cumulative CH\textsubscript{4} production (Table 2). Similar trends were reported in previous work [18,29]. Jiraprasertwong et al. used cassava wastewater in a three-stage up flow anaerobic sludge blanket (UASB) reactor and showed a steady reduction in COD removal with increasing COD loading and an increasing biogas production up to 15 kg COD/m\textsuperscript{3}d (one reactor) and 10 kg/m\textsuperscript{3}d (two reactors) [18]. For comparison, our batch study had a one-time COD loading for each substrate tested that ranged from 25 to 43 kg COD/m\textsuperscript{3}, respectively.
3.5. Scale-Up Model

A medium size cassava factory in Nigeria processes 3000–6000 kg of cassava tubers per day, yielding 1000–2000 L CW/d. The size of the digester needed to co-digest CW with PL was calculated to be 50 m³ (40 m³ liquid and 10 m³ biogas headspace), as shown in Figure 3. The quantity of PL added to digester would vary from 56 kg/d during high cassava production (March to October) to 28 kg/d during low cassava production (November to February) to maintain a 1:1 ratio (by VS). The hydraulic retention time (HRT) would vary from 20 days during high cassava production to 40 days during low cassava production, which should result in 78.8% to 96.5% of the cumulative CH₄ production is more pronounced below the mesophilic range (<25 °C) [30]. The digester design also includes a greenhouse covering the digester, which our previous research has shown can significantly increase (6.8–24.5 °C) the digester temperature and help maintain a more consistent digestion temperature throughout the day [44]. Assuming a 10-year lifetime and 8% discount rate, the discounted capital investment would be $8617. Based on the expected annual CH₄ production, the system would have a yearly profit of $5642/yr. The net present value (NPV) is calculated as $47,805, which does not take into consideration the value of the digestate, which can provide valuable nutrients to produce cassava and/or other crops. The price of adding zeolite or biochar was not included, since these additives

**Figure 3.** Scale-up model for a cassava processing factory, with two 25 m³ digesters plumbed in series to treat cassava wastewater and poultry manure with the utilization of the digestate for fertilizer.
may not be available and did not increase the overall biogas production, only decreased the lag phase associated with biogas production. If a higher throughput is desired or if the pH is not stabilized, the addition zeolite or biochar could be added to the full-scale system, if available.

4. Conclusions

Anaerobic digestion of cassava wastewater was shown to be viable, with CH$_4$ production enhanced by additions of zeolite and biochar. Co-digestion with dairy and poultry manure decreased the lag phase of digestion but did not increase overall CH$_4$ production, likely due to the more recalcitrant materials within the manure feedstock compared to CW. While CH$_4$ production was more efficient with CW-only (higher CH$_4$/g VS included), more gross energy production (CH$_4$/g substrate added) occurred with the manure substrates due to the higher VS content of these substrates compared to the relatively dilute CW. Poultry litter generally contributed to higher CH$_4$ production when digested with CW compared to the use of DM, likely due to the higher N content of the PL raising the low C:N value of CW. All combinations of DM and PL showed that adding CW increased their overall CH$_4$ potential compared to the mono-digestion of PL or DM-only.

Co-digesting CW with PL with or without biochar in a typical rural cassava processing industry can significantly enhance the valorization of CW by yielding more CH$_4$ in less time and an estimated profit of >$5000/yr, with the creation of valuable fertilizer. The reduction in COD achieved through the digestion of CW could contribute significantly to reducing pollution of surface water due to indiscriminate disposal of untreated CW, as currently practiced in many rural settings. The potential application of digestate for land treatment should be further explored as a means of adding value to the overall cassava processing and value chain.

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