EVALUATION OF ANTI-GLYCATION EFFECT AND SAFETY OF SERUM ANTI-AGING FORMULATION CONTAINING GOLD NANOPARTICLES (AUNP) USING SIDAGURI EXTRACT (SIDA RHOMBIFOLIA)

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INTRODUCTION
Aging is defined as accumulative damage that could lead to the cell, tissue, and organ disruption which happened progressively. This aging occurs in all organs, including the skin. One of the causes of skin aging is the presence of advance glycation end products (AGEs). Glycation reaction is a non-enzymatic reaction between reducing sugar and free amino acid in the protein resulting in AGEs production. The reduced free radical production could inhibited AGEs in the initial phase by reducing free radical production to reduce the formation of carbonyl or dicarbonyl groups, blocking in the propagation phase by blocking the dicarboxyl or carboxyl groups of sugar, and detoxification of reactive dicarboxyl metabolite [1].

AuNP is one of the anti-glycation agents that inhibit AGEs production by competitively binding reducing sugar in binding free amino groups of lysine and arginine that are important for glycation. By this description, AuNP could be categorized as an anti-glycation agent that inhibits in the initial phase [2]. The ability of AuNP had been proven scientifically. The gold nanoparticle has an anti-glycation activity on collagen as much as 56.3±4.2% compared to Aminoguanidine 64.0±5.7% [2]. AuNP synthesized using Sidaguri extract, resulting in 83.372% inhibition activity compared to aminoguanidine that has 79.793% inhibition activity [3]. AuNP is considered to be an anti-aging agent on cosmetics due to its anti-glycation activity.

AuNP could be synthesized using plant extract (green synthesis method). The advantages of green synthesis are relatively safe for the environment, reproducible, and cost-effective, and more stable product material [4]. Green synthesis has been performed using plant extract, vitamins, biodegradable polymers, microorganisms [4]. In extract plants, the ability to synthesize nanoparticle is due to their ability to release metal ions. Their reductive capacity relied on the hyper accumulating and reductive capacity. Flavonoid could undergo tautomeric transformation to the keto-form; this leads to releasing the reactive hydrogen atom that important to reduce metal ions in gold nanoparticle from Au3+ to Au [5]. Sidaguri is one of the wild plants found massively in a tropical environment, including Indonesia. Sidaguri has a reduction capability due to its antioxidant activity [6]. The availability and the content lead to the usage of Sidaguri extract to synthesize gold nanoparticles.

In this study, gold nanoparticle was synthetized using Sidaguri extracts and formulated into the serum, subjected to an in vitro anti-glycation test and irritancy test. An anti-glycation test was performed to optimize the formulation dose by observing the ability of serum formulation to inhibit AGEs formed during the test. Meanwhile, the irritancy test was performed using the HET-CAM test to classify further the material based on its irritancy characteristics.

MATERIALS AND METHODS
Material and chemicals
Material used during the study are Gold nanoparticle synthesized using Sidaguri extract, Xanthan Gum (Dessens Biochemical (Ordos) Ltd., China), Phenoxyethanol, Glycerin (Palm-Oleo Sdn, Malaysia), Bovine Serum Albumin (BSA) (Sigma Aldrich, Inggris), Fructose (Merck, Germany), Sodium Chloride 0.9% (B. Braun, Malaysia), Sodium Azide (Merck, Germany), Aminoguanidine HCl (Shandong Zhi Shang Chemical, China), Phosphate Buffer Saline (Merck, Germany), Bovine Serum Albumin (BSA) (Sigma Aldrich, Inggris), Phenoxyethanol, Glycerin (Palm-Oleo Sdn, Malaysia), White Leghorn Egg (Balitnak, Indonesia), Sodium Chloride 0.9% (R. Braun, Malaysia), Sodium hydroxide, distilled water and double distilled water.

The tools used are analytical weigh, Ika Eurostar High Speed Mixer (IMMlab, Francais), pHmeter (Eutetech Instrument, Singapore), Microplate reader GloMax (Promega Corporation, Madison, WI, USA), Micropippette (Eppendorf), 96 well plate (Costar), Viscometer with spindle L3 and L4 (Goel Parmer, USA), hot plate and glassware.

Serum anti-glycation formulation
AuNP was synthesized using Sidaguri extract; AuNP was prepared by reacting HAuCl 4 and Sidaguri extract for 150 min, then Sodium citrate for another minute. Then, the final product of AuNP was formulated into serum-containing xanthan gum as a thickening agent, phenoxyethanol as a preservative, and glycerine as a humectant.
Irritancy test using HET-CAM method

Fsb: The fluorescence intensity of the sample test’s blank solution
Fc: The fluorescence intensity of the negative control solution
Fs: The fluorescence intensity of the test solution
Fcb: The fluorescence intensity of the negative control’s blank solution

Leghorn species. Eggs used must be fresh (stored no longer than seven days) with a weight range of 45–70 grams. The eggs should be washed thoroughly, then incubated until day eight after laid at temperature 37.8±0.3 °C. The egg s were placed with CAM membrane in the top position, and the eggs must be rotated 180° every day. On day 8, the eggs were checked using candling light to separate the fertile and sterile egg. On day 9, the fertile eggs were tested with the sample test.

After optimizing the serum bases, two concentrations of AuNP were added to the optimized serum bases as listed in Table 2. The two formulas have tested the activity against AGEs formation using anti-glycation test.

Anti-glycation test

The dosage of AuNP used in serum was optimized using an anti-glycation test. This study used a modifications of methods by Spinola and Sutriyo [3, 7]. The sample used in this test is Aminoguanidine HCl, serum formulation containing 10% AuNP (F2.1) and serum containing 20% AuNP (F2.2).

There are two solutions prepared, a test and a blank solution. The differences between these two solutions are the blank solution did not contain fructose 0.5 M. The test solution was prepared by mixing 50 µl Bovine Serum Albumin 10 mg/ml (BSA), 50 µl fructose 0.5 M, 80 µl phosphate buffer saline (PBS) pH 7.4 containing 0.2% Natrium Azide (NaN₃) and 20 µl of a sample in the microplate. A blank solution was prepared by mixing 50 µl Bovine Serum Albumin 10 mg/ml (BSA), 80 µl phosphate buffer saline (PBS) pH 7.4 containing 0.2% Natrium Azide (NaN₃), and 20 µl of a sample in the microplate.

Then, the microplate was incubated at 37 °C for seven days. The intensity of each mixture was measured using a microplate reader by measuring the intensity of the fluorescent at 415 -445 nm emission and 365 nm excitation. The percentage of inhibition was calculated using equation 1.

\[
\%\text{inhibition} = \left(1 - \frac{F_c - F_b}{F_s - F_b}\right) \times 100\% \quad \text{equation 1}
\]

Fs: The fluorescence intensity of the sample test’s blank solution
Fcb: The fluorescence intensity of the blank solution
Fcb: The fluorescence intensity of the test solution

RESULTS

Serum formulation

The obtained serum base had a clear gel-like cosmetic form and was found to be homogenous on all formula. The differences in xanthan gum influence pH and obtained viscosity. Viscosity obtained for formula F3 was measured with different spindle due to the stark differences in viscosity with F1 and F2 that has less viscous. F3 was measured with spindle L3; meanwhile F1 and F2 were measured with spindle L4. All serum formulation evaluation result could be seen in Table 3. Based on the appearances and consistency, the considered formula was formula F1 and F3.

Irritation test using HET-CAM method

HET-CAM method performed using fertile chicken eggs from White Leghorn species. Eggs used must be fresh (stored no longer than seven days) with a weight range of 45–70 grams. The eggs should be washed thoroughly, then incubated until day eight after laid at temperature 37.8±0.3 °C. The eggs were placed with CAM membrane in the top position, and the eggs must be rotated 180 ° every day. On day 8, the eggs were checked using candling light to separate the fertile and sterile egg. On day 9, the fertile eggs were tested with the sample test.

The eggs were divided into five groups of treatment. Group 1 was given NaCl 0.9% (negative control), group 2 was given with NaOH 0.1 N (positive control), group 3 was given serum base, group 4 was given serum containing 10% AuNP, and group 5 was given serum containing 20% AuNP.

As much as 0.3 ml sample test was applied to CAM membrane. After sample test application, the time of hemorrhage, lysis, and coagulation occurred will be noted [8, 9]. Irritation index (RI) will be calculated using equation 2. Then, the mean of RI will be used to determine the irritancy level of testing material. Irritation classification could be seen in Table 2.

\[
RI = \frac{301 - \text{sec H}}{300} + 301 - \text{sec L} \quad \text{equation 2}
\]

Sec H: time when hemorrhage happened (in seconds)
Sec L: time when lysis happened (in seconds)
Sec C: time when coagulation happened (in seconds)

Note: Data given in mean±SD, n = 3

Table 1: Formula of anti-aging serum base

| Material             | Formula (%w/w) | F1   | F2   | F3   |
|----------------------|----------------|------|------|------|
| Xanthan Gum          | 0.50           | 0.75 | 1.00 |      |
| Glycerin             | 3.00           | 3.00 | 3.00 |      |
| Phenoxyethanol       | 0.80           | 0.80 | 0.80 |      |
| Distilled water      | Up to 100      | Up to 100 | Up to 100 |      |

Table 2: Formula of serum anti-aging containing AuNP

| Material          | Formula (%w/w) | F2.1 | F2.2 |
|-------------------|----------------|------|------|
| AuNP              | 10             | 20   |      |
| Serum base        | Up to 100      | Up to 100 |       |

Table 3: Material irritation classification

| Irritation index (RI value) | Material classification |
|-----------------------------|-------------------------|
| 0-0.9                       | Non-irritant             |
| 1-4.9                       | Weak to little irritant  |
| 5-8.9                       | Medium irritant         |
| 9-21                        | Strong to severe irritant|

Table 4: Physical evaluation of serum base

| Formula | Homogeneity | pH | Viscosity (at 30 rpm) |
|---------|-------------|----|----------------------|
| F1      | Homogenous  | 6.94 | 2019.53 cps (L3)     |
| F2      | Homogenous  | 6.71 | 3014.33 cps (L3)     |
| F3      | Homogenous  | 6.50 | 5508.77 cps (L4)     |

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Anti-glycation test

Glycation inhibition activity for formula 10\% (F2.1) are 68.20\pm6.86\% meanwhile formula 20\% (F2.2) resulting 74.83\pm19.91\% inhibition with the positive control are 39.09\pm11.67\%, raw data could be seen at table 5.

| Sample                      | Fluorescence intensity |
|-----------------------------|------------------------|
|                            | I          | II         | III        |
| Negative control (test)     | 662        | 662.3      | 540.7      |
| Negative control (blank)    | 200.4      | 249        | 230.8      |
| Positive control (test)     | 324.9      | 349.6      | 363.8      |
| Positive control (blank)    | 99.2       | 116.3      | 120.1      |
| Formula 2.1 (test)          | 244.9      | 241.8      | 236.1      |
| Formula 2.1 (blank)         | 134.6      | 103.8      | NA         |
| Formula 2.2 (test)          | 693.4      | 842.9      | 762.8      |
| Formula 2.2 (blank)         | 696.5      | 612.5      | 699.8      |

Based on the statistical calculation, both formulas resulting in a significantly better inhibition activity (p<0.05) than the positive control. However, the comparison between formula 10\% (F2.1) and formula 20\% (F2.2) was not significantly different. Based on the comparison, optimized dosage used for serum formulation was 10\% (F2.1).

Irritation test using HET-CAM test

| Sample                      | Before NaCl 0.9\% application | After NaCl 0.9\% application |
|-----------------------------|-------------------------------|-----------------------------|
| A1                          |                               |                             |
| B1                          |                               |                             |
| A2                          |                               |                             |
| B2                          |                               |                             |

Note: Data given as shown on the microplate reader

Table 5: Fluorescence intensity of anti-glycation activity test

Note: Data shown as mean\pmSD, n=81 (positive control, formula 2.2); n=54 (formula 2.1)
There was no hemorrhage, lysis, and coagulation formed on the egg’s CAM in groups 1, 3, 4, and 5. Hemorrhage, lysis, and coagulation were found in group 2, positive control. Hemorrhage formed averagely the first 12s, followed by lysis of blood vessel around 80–120s and coagulation formed was varied on 200–300s (fig 2. B2). All results of eye irritant potency could be seen in table 6 and fig. 2.

Table 6: Result of irritant test

| Test solution                       | Irritation score (Mean) | Irritant classification |
|-------------------------------------|-------------------------|------------------------|
| NaOH 0.9% (negative control)        | 0.0±0.00                | Non-irritant           |
| NaOH 0.1 N (positive control)       | 12.40±1.28              | Strong to severe irritant |
| Serum base                          | 0.0±0.00                | Non-irritant           |
| Serum containing 10% AuNP           | 0.0±0.00                | Non-irritant           |
| Serum containing 20% AuNP           | 0.0±0.00                | Non-irritant           |

Note: data given in mean±SD, n=4

DISCUSSION

AuNP was formulized into serum due to its high concentration than other cosmetics forms [11, 12]. The advantages of serum are easy and comfortable to use because of the minimum oil phase [13]. In this study, xanthan gum was used as a thickening agent due to its pseudo-plasticity characteristics so that the cosmetics would become smooth and soft [14]. Xanthan is non-toxic, not irritating when applied to the skin, and stable over extensive range of pH values [15]. A low concentration of xanthan gum in an aqueous solution is highly viscous and stable in changes of heat, pH, and resistance to enzymatic degradation due to its structural rigidity [14].

F1 and F2 have almost the same appearances, however, F1 was less viscous. Therefore, we considered using the F2 due to the API (AuNP) liquid form and anticipating the final product not too watery, and the F2 was selected.

AGEs are glycation reaction product derived from a non-enzymatic reaction between sugar and free amino acid in the protein. Based on AGEs’ chemical characteristics, AGEs could be classified into non-cross-linking and cross-linking. The cross-linking AGEs could be classified into fluorescent and non-fluorescent ones [16]. The inhibition mechanism of AGE synthesis could only delay or reduce AGE formation [1]. Aminoguanidine is one of the anti-glycation agents that usually used as a positive control during the anti-glycation inhibition activity test. Aminoguanidine limited AGEs’ formation by trapping products from early glycation, such as intermediate carbonyl compound [17].
Bovine serum albumin (BSA)-fructose test was one method used for detecting AGEs formed by measuring fluorescence intensity at emission and excitation wavelengths of 415-445 and 365 nm [18]. Then as mentioned before, bovine serum albumin will be acting as a model protein and fructose as the glycation agent. The condition chosen for the incubation process was similar to the normal human body (37 °C, pH 7.4). To create pH 7.4 condition, PBS was added into the mixture. NaCl was added to the PBS solution to prevent yeast from growing during the incubation process. During incubation, the plate shall be covered with aluminum foil to prevent the mixture from dried. Incubation for seven days of the mixture was expected to produce fluorescent AGEs detected using a microplate reader [19]. Theoretically, the sample solution will result in higher intensity than the blank sample due to there is little to no AGEs formation reaction in the blank. In pre-studies, gold nanoparticle synthesized with Sidaguri extract had been proven AuNP has higher inhibition activity, 83.87±4.39% than Aminoguanidine, 79.79±5.20%, and there is a significant difference in their inhibition activity [3].

HET-CAM test is used to classify material that is expected to be non-irritant or mildly irritant. HET-CAM evaluates the conjunctiva part of the eye [9, 20]. In this research, serum AuNP was predicted to be non-irritant material; therefore usage of HET-CAM for eye irritation test was performed. HET-CAM test principle observes and measures hemorrhage, lysis, and coagulation effects on the CAM when sample test applied, the test used was using RTM (Reaction Time Method) [9]. Tests were performed on day-9 when nerve tissue and pain perception have not yet been formed [20].

Based on IC50AM guidelines, the HET-CAM test is acceptable if the R.I value of 0.9% NaCl as a negative control was 0 and the R.I value of 0.1 N NaOH as a positive control was ranged between 10 and 19. In this experiment, all this requirement was met. RI value of 0.9% NaCl was 0 and RI value of 0.1 N NaOH was 12.408. Thus, both serum base and serum-containing AuNP could be presumed non-irritant to the eye with an RI value 0.0.

CONCLUSION

Serum anti-aging containing AuNP synthesized using Sidaguri extract has high anti-glycation activity compared to the positive control with dose optimized at 10%. Then, serum-containing AuNP synthesized with Sidaguri extract is a little to no irritant material. Considering the in vitro activity and safety, AuNP synthesized from Sidaguri extract could be considered to use as anti-aging cosmetics. However, the anti-aging activity and eye irritation characteristic of AuNP needs further study.

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AUTHORS CONTRIBUTIONS

All authors have contributed equally.

CONFLICT OF INTEREST

Declared none

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