Distribution of testate amoebae in bryophyte communities in São Miguel Island (Azores Archipelago)

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Abstract

Background

Testate amoebae are a polyphyletic group of protists living preferentially in soils, freshwaters and wetlands. These Protozoa have a worldwide distribution, but their presence and diversity in the Azores (a remote oceanic archipelago) is poorly known, with only twelve taxa recorded so far. The published information reflects occasional collections from sporadic field visits from naturalists to São Miguel Island, mainly in the nineteenth century. To overcome this limitation, a standardised survey was carried out on the Island, sampling different types of habitats from several localities to provide the distribution and information on species ecology of testate amoebae.
New information

In this study, 43 species of testate amoebae were recorded (within a total of 499 occurrences), belonging to two orders of Protista (26 Arcellinida and 17 Euglyphida). The most frequently occurring testate amoebae were *Euglypha strigosa*, *Trinema lineare*, *Euglypha rotunda*, *Assulina muscorum* and *Cyclopyxis eurystoma*. The most diverse genus was *Euglypha* (six species). A total of 38 species are new records for the Azores Archipelago. These data help to improve knowledge of the geographical distribution of testate amoebae in the northern hemisphere and their diversity in the Azores Archipelago.

Keywords

biodiversity, community ecology, island, moss, Protozoa, substratum specificity

Introduction

Testate amoebae are a polyphyletic group of small size [ranging from 7 to 500 µm long (Clarke 2003)] protists, enclosed within a xenosomal or idiosomic shell or test, made from proteinaceous, calcareous and/or siliceous material (Mitchell et al. 2008), with one or two oral apertures. They have a worldwide distribution, occurring in aquatic and terrestrial systems (Smith et al. 2007). In aquatic environments, they play an important role, especially in material cycling and energy flow (Glime 2017), while in terrestrial habitats, they play a crucial role in carbon and nitrogen cycling (Puppe et al. 2015). Due to their importance and sensitivity to environmental changes in both systems, they have been frequently used as bioindicators of environmental quality or stress or ecosystem resilience (Mitchell et al. 2003, Nguyen-Viet et al. 2007, Zapata et al. 2008). In addition, their shells are usually well preserved in sedimentary records and remain nearly unchanged over time. As the species composition of these protists depends on environmental conditions, they are frequently used to reconstruct the past climate and environment (Ellison 1995, Mitchell et al. 2001, Mitchell et al. 2008). The increasing use of testate amoebae in palaeoecological studies in the last decades demands the knowledge of modern assemblages for comparative analysis (Mitchell et al. 2001, Charman 2001, Booth 2002, Swindles et al. 2015) and for the establishment of their functional traits (Mitchell et al. 2014, Amesbury et al. 2016, Marcisz et al. 2020).

Despite their great importance, current knowledge of testate amoebae in the Azores Archipelago is limited when compared to other groups (e.g. Borges et al. 2010, Raposeiro et al. 2012, Pereira et al. 2014) and previous studies are fragmented and unsystematic. Interest in Azorean testate amoebae started almost two centuries ago with the work of Ehrenberg (1854), which reported three protist species *Difflugia azorica*, *Difflugia oligodon* and *Trinema enchelys*, found in soil collected on São Miguel Island. *Difflugia azorica* was described by Ehrenberg (1871) as an endemic species, although the diagnosis can be applied to many species of the genus and may correspond to a variety of *Difflugia pyriformis*. Leidy (1879), in his work “Fresh-water rhizopods of North America”, quotes the
species again, without adding any comment and *Difflugia oligodon* was described by Ehrenberg (1844) on the basis of samples from Kurdistan and a 10-word diagnosis.

Later, with the Challenger expedition that took place from 1872 to 1876 and which had a brief passage to São Miguel Island, the Irish naturalist Archer (Archer 1874) studied samples from Lake Furnas and reported six new taxa. By the end of the nineteenth century, Gerne (1888) and Barrois (1896) published several papers on freshwater biota of the Azores including some additions to testate amoebae fauna (Table 1).

| Taxa (Current name)       | Ehrenberg 1854 | Archer 1874 | Gerne 1888 | Barrois 1896 | Gadea 1979 |
|---------------------------|----------------|-------------|------------|--------------|------------|
| *Arcella vulgaris* Ehrenberg, 1830 |               |             |            |               | X          |
| *Centropyxis aculeata* (Ehrenberg, 1830) Stein, 1857 as *Echinopyxis aculeata* |               |             | X           | X            |            |
| *Centropyxis constricta* (Ehrenberg, 1841) Penard, 1890 |               |             |            | X            | X          |
| *Centropyxis sp.* |               |             |            | X            |            |
| *Difflugia acuminata* Ehrenberg, 1838 |               | X           |            | X            |            |
| *Difflugia globulosa* Dujardin, 1837 as *D. globularis* |               |             |            |              |            |
| *Difflugia mitriformis* Wallich, 1864 |               |             |            | X            |            |
| *Difflugia oligodon* Ehrenberg, 1844 |               |             | X           |              |            |
| *Difflugia pyriformis* Perty, 1849 as *D. azorica* |               |             | X           | X            |            |
| *Euglypha sp.* |               |             |            |              | X          |
| *Euglypha acanthophora* (Ehrenberg, 1841) Perty, 1849 as *E. alveolata* |               |             | as *E. alveolata* |            |            |
| *Nebela collaris* (Ehrenberg, 1848) Leidy, 1879 s.l. |               |             |            | X           | X          |
| *Quadrulella symmetrica* (Wallich, 1863) Kosakyan et al., 2016 as *Difflugia symmetrica* |               |             |              | X            |            |
| *Trinema enchelys* (Ehrenberg, 1838) Leidy, 1879 | X           |             |            |              | as *T. acinus* | X | X |

Almost 100 years later, Gadea (1979), in his work on nematodes living in mosses, recorded three genera (*Centropyxis*, *Euglypha* and *Plagiostoma*). Of the three genera reported, *Plagiostoma* seems to be a misprint and refers to *Centropyxis plagiostoma* or genus *Plagiopyxis*. Since that time, no studies have been carried out on testate amoebae in the Archipelago.

The main objective of this data paper is to provide a record of the diversity and detailed distribution of testate amoebae in São Miguel Island (Azores Archipelago, Portugal).
Additional information on species ecology is also discussed. Our purpose is to release this valuable dataset since no similar datasets have been previously published for Azores and it constitutes a relevant tool for comparison for ecologists studying, for example, biogeographic patterns or climate change and as modern analogues for environmental reconstructions on oceanic islands in paleoecological studies.

Project description

Title: Records of testate amoebae in São Miguel Island (Azores Archipelago)

Personnel: Collections were undertaken and occurrence data recorded during 2020 in São Miguel Island. The collectors were Martin Souto, Vitor Gonçalves and Pedro Miguel Raposeiro. Identification was done by Martin Souto and Xabier Pontevedra-Pombal. Production and analysis of scanning electron microscopy images was done by Xabier Pontevedra-Pombal.

Study area description: The Azores is an oceanic archipelago located in the middle of the North Atlantic, about 1500 and 2100 km off the coast of Portugal (Europe) and North America, respectively (Fig. 1).

Native forests cover less than 10% of the total area, mostly at elevations > 800 m a.s.l. (Borges et al. 2010, DDRF 2014), being a priority habitat in the Natura 2000 network (Guimarães and Olmeda 2008). Dominant tree species of this endemic forest are
Juniperus brevifolia (Seub.) Antoine, Laurus azorica (Seub.) Franco and Ilex azorica Loes. with a close canopy in which a great diversity of ferns and mosses is found (Elias et al. 2016). Cryptomeria japonica (Thunb. ex L.f.) D. Don forest occupies about 22% of the land area in the Azores (representing 60% of forest plantation area (Cruz et al. 2007), located especially at elevations > 400 m a.s.l. (DDRF 2014). These Japanese cedar forests are very dense, limiting the development of ferns and some mosses. The bryophyte communities present under the canopy of this forest is dominated by Leucobryum juniperoides (Brid.) Müll. Hal., Marchantia paleacea Bertol., Trichocolea tomentella (Ehrh.) Dumort, Thuidium tamariscinum (Hedw.) Schimp. and Hypnum cupressiforme Hedw. Peatlands, mainly located in depressions in high elevation areas and cover an area of 3000 ha (Mendes and Dias 2017), are characterised by the strong development of different species of Sphagnum and other bryophytes (Dias and Mendes 2007). Apart from their ecological importance, peatlands are, together with lakes (e.g. Hernandez et al. 2017, Raposeiro et al. 2017, Vázquez-Loureiro et al. 2019), the best paleoecological archives available in the Azores. Due to the existence of active volcanoes, São Miguel Island is particularly rich in hydrothermal vent fields (Gaspar et al. 2015). Biological communities of wetlands located close to these hydrothermal sites are influenced by higher temperatures and CO₂-rich mineral waters. In this specific habitat, plant communities consist mainly of vascular plants, such as Juncus effusus L. and Equisetum telmateia Ehrh. Hydrothermal carbonisation of different wetland biomass wastes allows for the development of a rich community of bryophytes characterised by species that tolerate extreme conditions (Elmarsdóttir et al. 2015), such as lawn communities of Sphagnum spp, Calliergon sp. and Polytrichum sp. According to (Porteiro 2000), São Miguel Island has 33 lakes, located at a range of between 260 m (Azul and Verde) and 830 m in altitude (Éguas Norte). In general, the dominant vegetation of lake shores are J. effusus, Osmunda regalis L. Agrostis sp., surrounded by hygrophyte shrub communities of Calluna vulgaris. The most abundant bryophytes are Rhytiadiadelphus squarrosus (Hedw.) Warnst, H. cupressiforme and several Sphagnum species. The presence of dense carpets of Fissidens sp. or Campylopus sp. is more common in open areas.

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Sampling methods

Study extent: This study covers 16 sampling locations on São Miguel Island (Fig. 1, Table 2), encompassing several habitat types (native forest, Cryptomeria forest, lake shore, peatland and hydrothermal vents). Within these habitats, different species of bryophytes were subsampled in triplicate, from a total of 46 moss subsampling locations, comprising a total of 138 samples.
Table 2.
Habitat characteristics and location of the sixteen studied localities in São Miguel

| Cod. | Locality                     | Latitude (Nº) | Longitude (Wº) | Alt. (m) | Date    | Habitat                                                                 |
|------|------------------------------|---------------|----------------|----------|---------|-------------------------------------------------------------------------|
| Loc. 1 | Lagoa Azul                  | 37.97833      | -25.84638      | 260      | 04- Feb. | Lake shore with degraded aquatic vegetation                              |
| Loc. 2 | Lagoa Verde                 | 37.89056      | -25.80194      | 260      | 04- Feb. | Strongly altered and degraded aquatic vegetation                         |
| Loc. 3 | Lagoa do Canário            | 37.83540      | -25.75898      | 755      | 04- Feb. | Aquatic vegetation with *Sphagnum* and *Thuidium* communities            |
| Loc. 4 | Lagoa do Caldeirão Norte    | 37.82340      | -25.75049      | 775      | 04- Feb. | Lake shore and shrub communities rich in *Sphagnum*                     |
| Loc. 5 | Lagoa do Carvão             | 37.82360      | -25.74241      | 700      | 15- May. | Lake shore and shrub communities                                          |
| Loc. 6 | Lagoa da Prata              | 37.80652      | -25.73641      | 520      | 15- May. | Peatlands with *Sphagnum* and *Cryptomeria* forest                       |
| Loc. 7 | Alto da Barrosa             | 37.76333      | -25.50056      | 800      | 27- May. | Shrub communities of *Calluna* with epiphytic bryophytes                |
| Loc. 8 | Lagoa do Fogo               | 37.76669      | -25.48024      | 580      | 22-Jun.  | Lake shore and *Cryptomeria* forest                                      |
| Loc. 9 | Ribeira da Praia            | 37.72583      | -25.46822      | 180      | 05- May. | *Fissidens* communities                                                  |
| Loc. 10 | Lombadas                   | 37.77745      | -25.46167      | 600      | 11-May.  | Hydrothermal, native and *Cryptomeria* forest                            |
| Loc. 11 | Lagoa de São Brás          | 37.79306      | -25.41278      | 600      | 27-Feb.  | Aquatic vegetation surrounded by *Cryptomeria* forest                   |
| Loc. 12 | Lagoa do Areeiro           | 37.76324      | -25.42743      | 580      | 13-Feb.  | Aquatic vegetation surrounded by *Cryptomeria* forest                   |
| Loc. 13 | Lagoa do Congro             | 37.75522      | -25.40737      | 700      | 15-May.  | Lake shore and *Fissidens* communities                                  |
| Loc. 14 | Ribeira do Caldeirão       | 37.81769      | -25.26294      | 600      | 27-May.  | Native and *Cryptomeria* forest                                         |
| Loc. 15 | Planalto dos Graminhais     | 37.80056      | -25.23944      | 900      | 10-Jun.  | Native forest and peatlands                                             |
| Loc. 16 | Ribeira do Guilherme        | 37.79972      | -25.20194      | 580      | 09- March.| Native forest                                                            |

**Sampling description:** Testate amoebae collections were taken at the beginning of the growing season, between spring (February - March) and summer 2020 (May - June). In each location, several types of vegetation were chosen for their homogeneity and
abundance (e.g. forest, lowland shrub, shrubland, wetland, peatland, riparian communities (Fig. 2)) and within these communities, the most abundant bryophytes were sampled. In each subsampling site, three homogeneous subplots (10 × 10 cm) were chosen, defining a total 138 sampling points (Table 2).

**Step description:** Testate amoebae were sorted by fragmenting, washing and stirring of 10 × 10 cm of a wet mass of moss material into 1 litre of distilled water and then sieved through a 300 μm mesh size to remove large moss particles. The samples were concentrated by sedimentation and stored in vials with 50% alcohol at 4°C. A small aliquot of each sample was stored as a reference collection.

A drop of each sample (three subplots) was mounted on a semi-permanent slide and all testate amoebae were identified at 200x and 400x magnification using a compound microscope Leica DM2500. All measurements were made on photomicrographs (Leica DFC495 camera) of at least than 10 specimens, using image analysis software (Leica Application Suite version 3.8.0).

In order to obtain Scanning Electronic Microscopy (SEM) images, an aliquot was dried on aluminium supports with a carbon film. They were metallised with Iridium (40 nm) in a BioRad Microscience ion plating system and examined in a Zeiss Ultra Plus field emission microscope, at 5 kV high electric voltage.

The identification of testate amoebae was based on Ogden and Hedley 1980, Todorov and Bankov 2019. The classification at higher ranks follows Adl et al. (2019). Identification of the bryophyte species follow Smith and Smith (2004). Accepted names and authorities for vascular plants and bryophytes were checked in [http://www.theplantlist.org](http://www.theplantlist.org) (June 2020).
Comparison of species richness (S) amongst different habitats was tested using one-way analysis of variance (ANOVA). Tukey’s honest significant difference (HSD) test was used as the multiple comparison post-hoc test when significant differences were identified in the ANOVA.

**Geographic coverage**

**Description:** The Azores is an oceanic archipelago located in the middle of the North Atlantic, about 1500 and 2100 km off the coast of Portugal (Europe) and North America, respectively (Fig. 1). The Archipelago is comprised of nine volcanic islands that are divided into three groups: the western group (Corvo and Flores Islands), the central group (Faial, Pico, Graciosa, São Jorge and Terceira Islands) and the eastern group (São Miguel and Santa Maria Islands). São Miguel is the largest Island, with an area of 746 km² and approximately 45% of the Island is between 300-800 m a.s.l., with a maximum elevation of 1103 m. The climate in the Azores is temperate oceanic, with regular and abundant rainfall, with high levels of relative humidity (up to 95% in high elevation native forests), ensuring moderate thermal variations throughout the year (Brito de Azevedo et al. 1999). Mean annual temperatures range between 14 and 18°C, while the mean annual precipitation is between 740 and 2400 mm (Marques et al. 2008, Hernández et al. 2016).

**Coordinates:** 37.704 and 37.917 Latitude; -25.857 and -25.125 Longitude.

**Taxonomic coverage**

**Description:** Testate amoebae found on São Miguel Island

**Taxa included:**

| Rank          | Scientific Name                  |
|---------------|----------------------------------|
| order         | Arcellinida                       |
| family        | Arcellidae                        |
| species       | *Arcella arenaria* Greeff, 1866   |
| species       | *Arcella catinus* Penard, 1890    |
| family        | Netzeliidae                       |
| species       | *Cyclopyxis eurystoma* Deflandre, 1929 |
| species       | *Cyclopyxis kahl* Deflandre, 1929 |
| infraorder    | Incertae sedis Sphaerothericina   |
| species       | *Trigonopyxis arcula* (Leidy, 1879) Penard, 1912 |
| family        | Diffugiidae                       |
| species       | *Diffugia bacillifera* Penard, 1890 |
| species                        | family                      | order                        |
|-------------------------------|-----------------------------|------------------------------|
| *Difflugia glans* Penard, 1902 | Centropyxidae               | Incertae sedis Arcellinida   |
| *Difflugia elegans* Penard, 1890 |                            |                              |
| *Centropyxis aerophila* Deflandre, 1929 |                  |                              |
| *Centropyxis constricta* (Ehrenberg, 1841) Penard, 1890 |                  |                              |
| *Centropyxis discoides* (Penard, 1890) Deflandre, 1929 |                  |                              |
| *Centropyxis elongata* (Penard, 1890) Tomas, 1959 |                  |                              |
| *Alabasta militaris* (Penard, 1890) Duckert, Blandenier, Kosakyan & Singer, 2018 | Hyalospheniidae |                            |
| *Nebela collaris* (Ehrenberg, 1848) Leidy, 1879 |                  |                              |
| *Padaungiella lageniformis* (Penard, 1890) Lara & Todorov, 2012 |                  |                              |
| *Padaungiella tubulata* (Brown, 1911) Lara & Todorov, 2012 |                  |                              |
| *Planocarina carinata* (Archer, 1867) Kosakyan et al., 2016 |                  |                              |
| *Quadruplella symmetrica* (Wallich, 1863) Kosakyan et al., 2016 |                  |                              |
| *Heleopera rosea* Penard, 1890 | Heleoperidae                |                              |
| *Heleopera sphagni* Leidy, 1874 | Microchlamyidae            |                              |
| *Pyxidicula cymbalum* Penard, 1902 |                  |                              |
| *Phryganeella acropodia* (Hertwig & Lesser, 1874) Hopkinson, 1909 | Cryptodiffugiiidae |                              |
| *Cryptodiffugia oviformis* Penard, 1890 |                  |                              |
| *Argynnia caudata* (Leidy, 1879) |                  |                              |
| *Argynnia dentistoma* (Penard, 1890) |                  |                              |
| *Physochila griseola* Penard, 1911 |                  |                              |
| *Euglypha acanthophora* (Ehrenberg, 1841) Perty, 1849 | Euglyphidae               |                              |
| *Euglypha cristata* Leidy, 1874 |                  |                              |
| species | *Euglypha filifera* Penard, 1890 |
|---------|----------------------------------|
| species | *Euglypha laevis* (Ehrenberg, 1845) Perty, 1849 |
| species | *Euglypha rotunda* Wailes, 1911 |
| species | *Euglypha strigosa* (Ehrenberg, 1871) Leidy, 1879 |
| species | *Tracheleuglypha dentata* (Vejdovsky, 1882) Deflandre, 1928 |
| family | Assulinidae |
| species | *Assulina muscorum* Greeff, 1888 |
| family | Cyphoderiidae |
| species | *Cyphoderia ampulla* (Ehrenberg, 1840) Leidy, 1879 |
| family | Sphenodermiidae |
| species | *Sphenoderia fissirostris* Penard, 1890 |
| species | *Trachelocorythion pulchellum* (Penard, 1890) Bonnet, 1979 |
| family | Trinematidae |
| species | *Corythion constricta* (Certes, 1889) Jung, 1942 |
| species | *Corythion dubium* Taranek, 1881 |
| species | *Playfairina valkanovi* Golemansky, 1966 |
| species | *Trinema complanatum* Penard, 1890 |
| species | *Trinema enchelys* (Ehrenberg, 1838) Leidy, 1879 |
| species | *Trinema lineare* Penard, 1890 |

**Temporal coverage**

**Notes:** 04-02-2020 through to 22-06-2020

**Usage licence**

**Usage licence:** Open Data Commons Attribution License

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**Data resources**

**Data package title:** Checklist of testate amoebae in São Miguel Island (Azores Archipelago)
Resource link:  http://ipt.gbif.pt/ipt/resource?r=tecamebas

Alternative identifiers:  https://www.gbif.org/dataset/13c79ceb-0ceb-424f-b000-7991d7c49834

Number of data sets:  1

**Data set name:** Checklist of testate amoebae in São Miguel Island (Azores Archipelago)

**Data format:** Darwin Core Archive

**Description:** This paper presents data distribution of testate amoebae in São Miguel Island (Azores Archipelago) collected during 2020. The dataset has been published as a Darwin Core Archive (DwC-A), which is a standardised format for sharing biodiversity data as a set of one or more data tables (Souto et al. 2021). The core data table contains 16 events (eventID), 499 occurrences (occurrenceID) with 43 taxa (taxonID). The number of records in the data table is illustrated in the IPT link. This IPT archives the data and thus serves as the data repository. The data and resource metadata are available for downloading in the downloads section.

| Column label       | Column description                                                                 |
|--------------------|------------------------------------------------------------------------------------|
| id                 | Identifier of the record, coded as a global unique identifier                      |
| locality           | Name of the locality where the event occurred                                      |
| continent          | Continent of the sampling site                                                    |
| country            | Country of the sampling site                                                       |
| island             | Island from the Island Group of the sampling site                                  |
| islandGroup        | Island group of the sampling site                                                  |
| eventID            | Identifier of the event, unique for the dataset                                    |
| occurrenceID       | Identifier of the occurrence, coded as a global unique identifier                 |
| type               | The nature of the resource                                                        |
| Habitat            | Habitat sampled                                                                    |
| basisOfRecord      | The specific nature of the data record                                             |
| samplingProtocol   | Sampling protocol                                                                  |
| recordedBy         | Person who collected the specimens                                                 |
| identifiedBy       | Person who identified the specimens                                                |
| eventDate          | Time interval when the event occurred                                              |
| taxonID            | The identifier for the set of taxon information (data associated with the Taxon class). Specific identifier to the dataset |
Additional information

Analysis

This study presents 499 testate amoebae (Protista) occurrences in 46 sampled sites (16 localities) in São Miguel Island, belonging to 43 species from 25 genera, 14 families and two orders. The order Euplyphida, represented by five families, accounted for 58.5% of the total occurrences and the order Arcellinida 41.5% of the total occurrences.

The families with the highest number of occurrences were Euglyphidae (121), Hyalospheniidae (54), Heleoperidae (38), Centropyxidae (35) and Assulinidae (32). Additionally, the families with the highest number of taxa were Euglyphidae and Trinematidae (7), followed by Hyalospheniidae (6) and Centropyxidae (5). The families with lower occurrences (< 5) were Cyphoderiidae (4) and Microchlamyidae (1). The genera with the highest number of occurrences were *Euglypha* (106), *Trinema* (82), *Heleopera* (38) and *Centropyxis* (35). The other 21 genera had less than 35 occurrences. The genera with the highest number of taxa were *Euglypha* (6) and *Centropyxis* (4). *Euglypha strigosa*, *Trinema lineare* and *Euglypha rotunda* were the most frequent species occurring in 39, 37 and 34 sites, respectively. *Assulina muscorum* (32 sites), *Cyclopyxis eurystoma* (27 sites),
Corythion dubium (26 sites), Trinema complanatum (25 sites) and Euglypha laevis (24 sites) were amongst the most ubiquitous testate amoebae (Figs 3, 4).

A total of six taxa occurring at only one sampling site were considered rare (Fig. 5). These included Cryptodifflugia oviformis; Euglypha filifera; Physochila griseola; Planocarina carinata; Pyxidicula cymbalum and Trachelocorythion pulchellum. Another 14 taxa were
considered occasional, occurring in two to five sampling sites. These included species, such as *Alabasta militaris*, *Argynnina dentistoma*, *Cyclopyxis kahli*, *Argynnina caudata*, *Corynthion constricta*, *Difflugia bacillifera*, *Difflugia elegans*, *Euglypha acanthophora*, *Arcella catinus*, *Centropyxis constricta*, *Cyphoderia ampulla*, *Euglypha cristata*, *Padaungiella tubulata* and *Trigonopyxis arcula*. Species richness significantly differed amongst habitats (one-way ANOVA, P < 0.05). Overall species richness was in the order Natural forest > Peatland > Cryptomeria forest > Lake > Hydrothermal, but significant differences were found only between Native forest and Lake habitats (Fig. 6).

Figure 5. doi
Percentage of occurrence of each species for the 46 samples and their distribution amongst habitats (proportion of occurrences is represented in colour bars)

Figure 6. doi
Boxplot of species richness in each sampled habitat. Different letters indicate significant differences (Tukey Test, p < 0.05).
Discussion

There are very few inventories of testate amoebae in the Azores, therefore the overall species richness of testate amoebae is unknown. Here, we present the first systematic study that explored the distribution of testate amoebae in bryophyte communities mainly in forest habitats. Forty-three species are recorded for São Miguel Island, 38 of these being new records for the Azores Archipelago. However, we must take into account the numerous cryptic taxa that testate amoebae present and some taxonomic uncertainty (Roland et al. 2017). For example, amongst Arcellinid, the *Nebela tincta–bohemica–collaris* species complex is a problematic group having very similar species (Heal 1964, Gilbert et al. 2003, Kosakyan et al. 2013). It is possible that what we identify here as *Nebela collaris* s.l. may include several taxa and, for this reason, it appears as the most abundant species. Another genus with similar difficulties is *Quadrulella* (Kosakyan et al. 2016). Considering the complexity of these groups, more detailed taxonomic work and more morphometric studies, combined with genetic approaches, such as the molecular barcoding effort, are needed to characterise this species complex.

The three most representative families in terms of species richness, Centropyxidae, Euglyphidae and Trinematidae, are, in general, the most commonly registered in other oceanic archipelagos (Smith 1986, Smith 1992, Beyens et al. 1990, Mieczan and Adamczuk 2015, Golemansky 2016) and in other parts of the world (Beyens et al. 1986, Bobrov et al. 1999, Mazei and Belyakova 2011, Acosta-Mercado et al. 2012, Šatkauskienė 2014). Considering the species richness for different testate amoebae genera in the distinct habitats studied, *Euglypha* and *Trinema* were the most frequent, followed by *Corythion* and *Centropyxis* (Fig. 5).

The 43 taxa recorded to the Azores is higher than what was reported to other oceanic archipelagos, such as the Canary Islands (10 species), Balearic Islands (15) and Island of Annobón (30 species). Testate amoebae assessment in the Balearic and Canary Islands was focused on mosses under *Pinus* forests of drier characteristics (Gracia 1965a, Gracia 1965b), while on the Island of Annobón (Equatorial Guinea), the work was performed on forest epiphyte mosses (Gracia 1963). However, these numbers cannot be used to draw conclusions about testate amoebae species richness in each archipelago since sampling efforts, habitats and approaches were different. In this context, it is essential to increase the sampling effort on other archipelagos, as well as to survey multiple habitats in order to find a greater diversity of testate amoebae.

The testate amoebae assemblages in São Miguel Island were composed mainly by genus with a cosmopolitan distribution which are also known from other oceanic islands. For example, in the comparable Annobón tropical rainforest, situated much further south (Gracia 1963), the diversity of testate amoebae is very similar (share 20 species). The cosmopolitan character of testate amoebae assemblages is also found in the mainland counterparts: the tropical mountain rainforest in Ecuador presented taxa geographically widespread with only nine species (6.7%) being considered tropical (Krashevska 2009). However, these biogeographic conclusions may be biased, on the one hand because of the number of cryptic species that exist and, on the other hand, because of the lack of
habitat diversity surveyed. The information regarding protist communities came from \textit{Sphagnum} moss on peatlands (Glime 2017, Lamentowicz and Mitchell 2005). Only a few studies were made in other types of bryophytes assemblages, mainly in northern areas including Devon Island (Beyens et al. 1990), Greenland (Beyens et al. 1986), Russia (Mazei and Belyakova 2011) or in sub-Antarctic areas (Smith 1992), such as Adelaide Island (Smith 1986), South Shetland Islands (Golemansky 2016), King George Island (Mieczan and Adamczuk 2015). The biogeographical situation of the Azores Archipelago between the Nearctic and the Palearctic offers unique possibilities to study the distribution of these organisms and their ability to colonise islands. This is the case of \textit{Argynnia caudata} present in the Azores and with a tropical/subtropical distribution.

Moss biotopes are very abundant in the Azores, where extant bryoflora comprises about 430 species of mosses and hepatics (Sjögren 2003). The subtropical forest from the Azores is more or less constantly humid and warm and supports a very rich assemblage of moss species, including a higher proportion of endemic species (Sjögren 2001). The highest species richness of testate amoebae (n = 17, 40 species) occurred in native forest habitats and corresponds to epiphytic bryophytes that grow abundantly on the bark of living trees/shrubs. This alliance Echinodion prolixi Sjn. 93 is established in part of the native forest-phytocenoses at high altitudes (600 m), dominated in the tree-layer of \textit{Laurus}, \textit{Erica}, \textit{Juniperus} and \textit{Ilex} (Sjögren 2003). In these epiphytic bryophytes, belonging to genera \textit{Frullania} or \textit{Scapania}, the most frequent species associated with these mosses were \textit{Corythion dubium}, \textit{Centropyxis aerophila} and \textit{Euglypha rotunda}. Although, native forest communities are highly degraded in São Miguel, where the best-preserved area corresponds to the high eastern part of the Island (Fig. 1, Loc.: 14, 15 and 16) and in many places have been replaced by lowland shrub and peatlands (Fig. 2, habitats 1 and 5). In fact, peatland habitats can be considered in the Azores as an extension of these wet mountain forests, where the more abundant mosses are \textit{Polytrichum juniperinum} Hedw. and \textit{Sphagnum} spp. Despite that, they only share 42.5% of testate amoebae species, especially from genus \textit{Euglypha} and a lower species diversity was observed (n = 2, 17 species). However, these results must be regarded with caution, because of the low number of replicates collected and analysed from peatlands.

Most of the natural vegetation of the Island has been replaced by pastures and \textit{Cryptomeria} forests (Fig. 2, habitat 2). These \textit{Cryptomeria} forests are the third most sampled habitat (n = 12, 31 species). Despite being conifer monocultures, the ecotone areas maintain a similar diversity of testate amoebae, when compared to native forest and it is easy to find native bryophyte communities, such as the case of \textit{Breutelia azorica} (Mitt.) Cardot. The most common species of bryophyte \textit{Leucobryum juniperoides} grows in very dense, glaucous green, swollen cushions or hummocks. Some hummocks in woodland can be massive and colonised by other bryophytes and vascular plants. This eosinophilic moss which grows mainly at the base of \textit{Cryptomeria} trunks, due to its dense growth structure, constitutes a favourable habitat for a high diversity of testate amoebae. In fact, this moss \textit{Breutelia azorica}, presents the most diverse assemblages of testate amoebae within the \textit{Cryptomeria} forest. Species from \textit{Nebela collaris} complex (60%), \textit{Euglypha strigosa}, \textit{Assulina muscorum} and \textit{Trinema lineare} dominated in cushion moss \textit{Leucobryum}. The
testate amoebae communities that are richest in species are those that develop on pleurocarpic mosses and Jungermannialian hepatics in forest habitats (Figs 5, 6).

The most frequent genera shared on these terrestrial and semi-terrestrial habitats are *Nebela* and *Euglypha*, which are less represented in the aquatic systems. According to Lansac-Tôha et al. (2007), these genera possess fragile shells, which limit their occurrence in more dynamic environments, especially lakes and hydrothermal vents. It is possible that these are eurytopic species or that they are more abundant because they have greater access to their food resources in these terrestrial habitats.

Bryophytes assemblages on lake shores presented a high diversity of testate amoebae (n = 13, 33 species). Acarcarpic mosses, like *Fissidens* or *Campylopus* in more open areas near lakes, maintain less developed testate amoebae communities (Fig. 6). However, the genera *Sphagnum* and *Rhytidiadelphus squarrosus*, especially in more waterlogged areas, maintain rich testate communities (n = 13, 33 species). Several works consider aquatic environments and sediments to be the preferred habitat for these organisms, yet they end up functioning as a data collector for the surrounding ecosystems and many species are actually terrestrial (Glime 2017). In order to better understand this issue, it is important to further study species’ autoecology and habitat preferences.

On hydrothermal vents, lower diversity of testate amoebae was observed (n =3, 14 species). These vents develop an abundant bryophyte extension (Fig. 2), characterised by species that tolerate extreme conditions, such as *Sphagnum* spp, *Calliergon* sp. and *Polytrichum juniperinum*. The most frequent species are *Quadrulella symmetrica* and *Trinema lineare*. One possible explanation to this fact could be explained by their shell shape and composition, which are stiffer and more resistant, allowing their presence and permanence in this extreme habitat.

**Final remarks**

Here, we presented the first study that explored the distribution of testate amoebae of different habitats from the Azores Archipelago, mainly in São Miguel Island. Moreover, this work indicates that there are typical species on the different sampled habitats. This a matter of concern on islands, where large areas of native forest have been replaced by exotic forest and changes in land uses, driven by human activities, will affect population dynamics. In order to better understand the complexity of these habitats, population dynamics and species specificity need to be carried out. Larger datasets located in different islands and habitats are required to better understand how these communities respond to environment changes. Additionally, molecular barcoding is a useful tool, not only for species identification, but also for studying evolutionary and ecological processes. The results of this study provide indications that testate amoebae assemblages are habitat specific and therefore constitute a promising group for paleoenvironmental reconstruction of Azorean ecosystems. Future studies in drier ecosystems, coastal areas and hydrothermal zones may reveal and offer us a greater diversity of these organisms.
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Author contributions

Martin Souto, Vitor Gonçalves and Pedro Miguel Raposeiro conceived the study and carried out the sampling campaign. Identification was done by Martin Souto and Xabier Pontevedra-Pombal. Production and analysis of scanning electron microscopy images was done by Xabier Pontevedra-Pombal. Martin Souto and Pedro Miguel Raposeiro wrote the paper with inputs from all authors. All authors agree with the final version of the paper.

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