Wolfram Syndrome: A Case Report and Review of Clinical Manifestations, Genetics Pathophysiology, and Potential Therapies

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1. Background

Wolfram syndrome (WS) is a form of monogenic diabetes that typically presents with diabetes mellitus in childhood and optic atrophy by the age of 16. The prevalence of WS has been estimated between 1 in 770,000 in the United Kingdom [1] and 1 in 100,000 in North America [2]. This entity was first described in 1938 by Wolfram and Wagener [3]. Many individuals with WS eventually develop diabetes insipidus and deafness, hence the acronym, DIDMOAD (Diabetes Insipidus, Diabetes Mellitus, Optic Atrophy, and Deafness). Additional morbidities include hypogonadism, infertility, hypopituitarism [4], cerebellar ataxia, peripheral neuropathy, dementia, psychiatric illness, and urinary tract problems [5–7].

Previously hypothesized to be a mitochondrial disorder, it is now known that classical WS is the result of autosomal recessive mutations affecting the WFS1 gene, which is implicated in endoplasmic reticulum (ER) function. Autosomal dominant mutations in WFS1 have been reported to cause WS-like diseases characterized by diabetes, low frequency sensorineural hearing loss, psychiatric illness, variable optic atrophy, and WFS1-related low frequency sensorineural hearing loss [8–10].

This paper reviews recent advances in the understanding of the genetics and pathophysiology of WS and summarizes
promising therapeutic management options directed at preserving \( \beta \)-cell function and slowing progressive decline in neurologic function.

2. Case Presentation

A 39-year-old woman of mixed Northern European descent was diagnosed with gestational diabetes (GDM) at sixteen weeks of gestation during her first and only pregnancy. Age was her only risk factor for GDM. She was treated with insulin, up to a maximum of 25 units per day, to maintain euglycemia during pregnancy. Her pre-insulin-treatment A1c was 6.1\% (43 mmol/mol) (upper limit normal range A1c < 6.1\%, 43 mmol/mol). A healthy female neonate weighing 3374 g was delivered at 39 weeks of gestation by cesarean section, due to transverse lie. Three months postpartum, the patient met criteria for diabetes mellitus based on a 75 g oral glucose tolerance test; her 2-hour glucose value was 13 mmol/L (234 mg/dl) and fasting glucose 6.0 mmol/L (108 mg/dl). Following pregnancy, her daily insulin requirements dropped, to 6–12 units per day (0.12 to 0.24 units/kg/day).

Currently, 11 years after the birth of her daughter, her quarterly A1c values remain less than 7.0\% (53 mmol/mol). Her 2-hour postprandial glucose values usually remain less than 6.5 mmol/L (117 mg/dl) despite only administering pre-meal insulin when her meals contain more than 60 g of carbohydrates. She consistently administers intermediate insulin at bedtime. She has never experienced diabetic ketoacidosis or severe hypoglycaemia requiring the assistance of another person. In the last year, she has begun to experience the need to drink water when swallowing food.

Her past medical history was significant for optic atrophy, diagnosed at age 10 which progressed to severe visual impairment over the following 8 years. From the age of 25 years, the patient complained of dizziness, unsteady gait with minor falls, and high-frequency sensorineural hearing loss.

Family history revealed one healthy sister, a brother with trisomy 21 who died at the age of 2 years from a congenital heart defect, and late onset deafness in her mother and maternal grandmother (in their seventies). Her father died of prostate cancer. There was no family history of diabetes mellitus or optic atrophy and no consanguinity.

On examination, her BMI was 17.8 kg/m\(^2\). She reported a low BMI throughout her life. She had bilateral optic atrophy with pale discs and impaired colour vision. Visual acuity was 20/400 in both eyes, with bilateral central scotomas. She had mild dysdiadochokinesia without dysmetria. She was unable to perform tandem gait for more than a few steps. She notably lacked typical findings associated with diabetes mellitus type 2, such as obesity, hypertension, and acanthosis nigricans.

Brain MRI revealed marked cerebellar vermis and hemispheric atrophy, as well as brainstem atrophy, particularly involving the pons. There was minimal change on MRI of her brain over a 10-year interval.

Antiglutamic acid decarboxylase and anti-islet cell antibodies were negative. Nutritional and vitamin deficiencies were ruled out with biochemical testing.

Genetic testing for autosomal dominant optic atrophy (OPA1) and spinocerebellar ataxia 1–8 and 17 were negative. Clinical sequencing of the WFS1 gene was performed through the Casey Eye Institute. Two mutations were identified. The first variant was c.2590G>T (p.E864*) , a truncating mutation previously reported in patients affected with WS. The second variant was c.977C>T (p.A326V) and has been previously described in psychiatric patients [10] but has not yet been described as a known causative mutation in WS. This variant was predicted as likely damaging by some in silico prediction models (Polyphen 2) but also predicted as tolerated by other models (SIFT, PROVEAN). Targeted sequencing was performed for these two variants on a first-degree relative, and this family member was found to have only one of the two mutations. This segregation analysis therefore confirmed that the mutations were in trans in the patient and therefore affecting both maternally and paternally inherited alleles. Mitochondrial DNA sequencing had been considered to assess for maternally inherited diabetes and deafness (MIDD) but was not pursued once testing for WFS1 returned positive. A comparison between WS and MIDD is provided in Table 1.

An experimental therapeutic trial of a dipeptidyl peptidase-4 inhibitor (DPP-4) was offered in the hope of extending neurologic and \( \beta \)-cell function; however the patient declined this.

3. Discussion

This patient has typical features of classical WS, including optic atrophy, diabetes mellitus, hearing loss, and cerebellar ataxia. However, she is remarkable for her late age of onset of diabetes and slow progression of pancreatic \( \beta \)-cell loss. She has survived for more than 12 years past the average life-expectancy, as the typical median age of death for WS is 39 years (range 25–49 years) [1]. One of her genetic variants c.2590G>T (p.E864*) has been reported previously in patients affected with WS and is a truncating loss of function mutation. Her other variant c.977C>T (p.A326V), resulting in a missense mutation, was described in psychiatric patients, but not as a causative mutation in WS [20]. However, this variant affects a highly conserved residue and is rare, with an allele frequency of less than 2.47 × 10\(^{-5}\); we feel this provides support for pathogenicity [21] and may be the cause of the mild phenotype. Thus, the presence of biallelic variants in WFS1 involving a known pathogenic truncating mutation and a rare missense variant, particularly in the context of her clinical findings, support the diagnosis of WS in our patient. It is possible, but less likely, that the true second pathogenic variant in WFS1 was missed with current sequencing technology.

3.1. Clinical Manifestations of WS. Diabetes mellitus is not the presenting clinical feature of WS in greater than 20\% of patients. In nearly 15\% of patients, the combination of both diabetes mellitus and optic atrophy is not yet present by 18 years of age [11]. In fact, less than one-third of patients meet the full clinical syndrome of Diabetes Insipidus, Diabetes Mellitus, Optic Atrophy, and Deafness, indicating that current clinical ascertainment criteria (early onset diabetes
Table 1: Comparison of our patient to WS and MIDD.

| Case patient | Wolfram syndrome | Maternally inherited diabetes mellitus and deafness |
|--------------|------------------|-----------------------------------------------------|
| Onset of diabetes mellitus | Diagnosed at the age of 39 years with gestational diabetes mellitus | Present in 98% [11]. Average age of diagnosis 6 years [1, 5] | Average age of diagnosis 37 years [12] |
| Anti-GAD and anti-islet cell antibodies | Absent | Absent | Absent |
| Diabetic ketoacidosis at presentation | Absent | 3% [2] | No data reported |
| Retinal disease | Absent | 35% diabetic retinopathy after 15 years [13] | Macular pattern dystrophy [14] |
| Renal disorders | Absent | 8% [13] | Focal segmental glomerulosclerosis with hyalinised glomeruli, myocyte necrosis in afferent arteries, and small arteries [15] |
| Optic atrophy | Diagnosed at the age of 10 years | Present in 82% [11]. Average age of diagnosis 10-11 years [5, 11] | No macular retinal dystrophy more common [14] |
| Sensorineural hearing loss | Diagnosed at the age of 25 years | Present in 48% [11]. Average age of diagnosis 16 years [1, 5] | Diagnosed between 2 and 61 years, frequently precedes diagnosis of DM, mean age of onset of hearing loss 33.2 years [15] |
| Diabetes insipidus | Absent | Present in classical WS 38% [11]. Average age of diagnosis 14-15 years [1, 5]. Absent in WS (type 2) [16] | Not routinely screened for |
| Neurological manifestations | Present, symptomatic by the age of 25 years | Present in 53% [5]. Average age of diagnosis 15 years [5] cebellar ataxia, peripheral neuropathy, and dementia | Mitochondrial encephalomyopathy, lactic acidosis and stroke-like episodes (MELAS) [17] |
| Urological manifestations | Absent | Present in 19% [11]. Average age of diagnosis 12–20 years [1, 5] | Not routinely screened for |
| Gastrointestinal manifestations | Absent | Gastrointestinal dysmotility in 24% [1]. Severe gastrointestinal ulcer and bleeding in WS (type 2) [18] | Gastrointestinal dysmotility |
| Psychiatric manifestations | Absent | 39% [5] | Depression, dementia, and psychosis [19] |
| Median age of death | n/a | 39 years [1] | No reported data |
| Cause of death | n/a | Neurological complications [1, 2] | Lactic acidosis, renal failure [19] |

Glycaemic control tends to be better in those with WS than individuals with type 1 diabetes. In an age and diabetes duration-matched comparison between WS and type 1 diabetes mellitus patients, A1c was lower for WS, 7.72 ± 0.21% versus 8.99 ± 0.25%, respectively, p = 0.002 (60.9 mmol/mol versus 74.8 mmol/mol), as we were total daily insulin requirements (0.71 ± 0.07 versus 0.88 ± 0.04 International Unit/kg/day, p = 0.0325) [13]. The lower daily insulin requirements per kg of body weight in WS patients suggest either greater pancreatic β-cell reserve or better insulin sensitivity when compared to matched individuals with type 1 diabetes. Our patient has very low insulin requirement suggesting good insulin sensitivity and considerable residual β-cell reserve.

Between II and 29 percent of WS patients encounter metabolic complications, such as severe hypoglycaemia causing coma or seizure. Severe hypoglycaemia was more
commonly reported in WS patients with coexistent neurological symptoms (9 of 31) compared to WS patients without coexistent neurological symptoms (3 of 28) [5]. Thus, coexistent neurological dysfunction in patients with WS appears to predispose to the high rate of severe hypoglycaemia observed.

In addition to diabetes, other endocrinological abnormalities such as diabetes insipidus are present in approximately 38% [11]. Other abnormalities include primary gonadal atrophy in males and menstrual irregularities and delayed menarche in females [1]. Short stature and growth hormone deficiency have been reported, as has hypopituitarism, which are believed to be due to hypothalamic dysfunction [4].

In the largest cohort series to date, neurological symptoms were present in 53% of patients by an average age of 15 years. The majority of symptoms were related to the brainstem and cerebellum, specifically, cerebellar ataxia (45%), peripheral neuropathy (39%), cognitive impairment (32%), epilepsy (26%), and lastly dysarthria, dysphagia, and nystagmus in 10% [5]. Brain MRI was abnormal in 54%, including atrophy of the cerebrum, cerebellum, and brainstem [5] as was observed in the woman described here.

3.2. Genotype Phenotype Correlations in WS. Genotype phenotype correlations in WS are unclear, with many reported potentially pathogenic variants [22] and considerable phenotypic variability in patients. Therefore, predicting and prognosticating the natural history of disease for an individual patient is not recommended. However, trends are emerging. In 2013 De Heredia et al. systematically reviewed 412 published WS cases in the literature, of which 337 had confirmed WFS1 mutations [11]. Using a genotype classification scheme based on the predicted amount of residual expression of defective WFS1 protein, they suggested that patients with genotypes that are likely to lead to absent protein production were more likely to have earlier onset diabetes, and possibly earlier onset optic atrophy, than patients with residual protein expression [11]. Genotype phenotype correlations are less evident for other features. Our patient had later onset of diabetes, which may in part be due to the presence of a milder missense mutation.

3.3. Pathophysiology of WS. WFS1 maps to chromosome 4p16.1 [22, 23]. The gene product (a transmembrane glycoprotein localized primarily to the ER [24]) functions to maintain homeostasis in the ER, the cellular organelle responsible for the folding of secretory proteins, such as insulin. Endocrine cells are particularly vulnerable to ER stress due to their rapid changes in secretory protein expression levels [25]. When ER homeostasis is disrupted, misfolded and unfolded proteins accumulate, leading to a state of ER stress [26]. The unfolded protein response is a response to ER stress in which cellular apoptosis may be triggered if the stress cannot be relieved [26] (Figure 1). ER stress is believed to play a role in β-cell dysfunction and apoptosis in type 1 and 2 diabetes mellitus and in other monogenetic forms of diabetes [27]. In WS, functional WFS1 protein deficiency alters IP3R-mediated ER calcium release, disrupting cytoplasmic calcium homeostasis [28] (Figure 1). Additionally, calpain-2, a calcium-dependent proapoptotic cellular protease, may play a role in ER stress-induced apoptosis through increased cytoplasmic calcium levels [29–31] (Figure 1). Calcium-calpain-2 pathway overactivation is thought to contribute to pancreatic β-cell dysfunction and apoptosis in diseases such as type 2 diabetes mellitus and WS [30, 32]. Cagaline et al. 2016 demonstrated that disruption in cytoplasmic calcium homeostasis in neurons also dysregulates mitochondrial dynamics which results in lower ATP levels. This is thought to hinder neuronal development and survival [28]. This likely explains the mitochondrial phenotype associated with WS. Interestingly, elevated free fatty acid levels, which are often present in type 2 diabetes mellitus and the metabolic syndrome, have been shown to promote activation of the calcium-calpain 2 pathway which promotes cellular apoptosis [33].

A second (ever rarer) type of WS (WS type 2) has a similar phenotype to classical WS. The causative gene for WS (type 2), CISD2, encodes an ER small protein, implicated in structural integrity and functional cross-talk between the ER and mitochondria [38]. Manifestations of WS (type 2) include diabetes mellitus, peptic ulcers, prolonged bleeding time, and neurodegenerative features [38–41]. The phenotypic overlap with classical WS is likely due to the overlapping function of the CISD2 and WFS1 gene products [7].

3.4. Promising Therapeutic Considerations for Those with WS. WS mutations lead to increased ER stress, altered cytoplasmic calcium, and dysregulation of mitochondria, which inhibits cellular growth and survival. Treatments which attempt to reduce ER stress may improve cell survival, notably, neural and pancreatic β-cell survival. In 2014, Lu et al. demonstrated that dantrolene could prevent apoptosis of WS patients’ neural progenitor cells [34]. Dantrolene inhibits ryanodine receptors in the ER and functions to suppress efflux of calcium from the ER to the cytosol (Figure 1) [42]. Hepatotoxicity, even with sporadic short-term use, is a known side effect of this drug that may range from asymptomatic transaminase elevations to fulminant hepatic failure [16]. Previous reports of dantrolene-related fatal hepatotoxicity have been associated with daily doses greater than 300 mg/day [42, 43]. More recently it has been suggested that lower daily doses (i.e., <200 mg/day) may be safely used in patients without coexisting liver dysfunction or coingestion of hepatotoxic medications [16]. It is important to systematically evaluate dantrolene safety prior to it becoming usual care for WS because reduced cytosolic calcium in the setting of neuronal stimulation in WS in vitro neuronal models has been linked with poor mitochondrial function [28]. A phase I clinical trial is currently investigating safety of long term use (A Clinical Trial of Dantrolene Sodium in Pediatric and Adult Patients with Wolfram Syndrome, ClinicalTrials.gov, NCT02829268).

In 2006, Yusta et al. showed that a glucagon-like peptide-1 receptor (GLP-1R) agonist interfered with the ER unfolded protein response, resulting in decreased apoptotic signalling and increased cell survival (Figure 1) [44, 45]. In a mouse model of classical WS, the GLP-1R agonist exenatide effectively treated hyperglycaemia [35]. Treatment of a patient with WS (type 2) with exenatide was associated with a 70% reduction in daily insulin dose, improved glycaemic control, and a 7-fold increase in maximal insulin secretion.
Endoplasmic reticulum

Unfolded proteins

ER stress

PKA/cAMP activation

ATF-6 activation

GLP-1R activation

Figure 1: Hypothesized molecular pathophysiology of WS. Under situations of stress, such as hyperglycaemia and elevated free fatty acids levels, unfolded and misfolded proteins accumulate [31]. Endoplasmic reticulum (ER) transmembrane proteins sense the stress and activate the unfolded protein response (UPR) [31]. The UPR may culminate in either an adaptive response which decreases the workload on the ER or a maladaptive response (as occurs in chronic hyperglycaemia or WS) which culminates in cellular apoptosis [31]. In healthy cells, the transmembrane protein (WFS1) complexes with activating transcription factor-6 (ATF-6) and directs ATF-6 to ubiquitin-mediated proteasome degradation [31]. This serves to negatively regulate the UPR [31]. In WFS1 deficient cells ATF-6 is no longer under negative inhibition and is permitted to constitutively activate genes that promote cellular apoptosis and decrease insulin gene expression [31]. ER calcium channels, such as the ryanodine receptor (RyR), but most importantly the inositol triphosphate receptor (IP3R), permit efflux of calcium from the ER to the cytosol [16, 28]. It is believed that increased cytoplasmic calcium levels activate the calcium-dependent protease, calpain-2, which promotes cellular apoptosis [16, 34]. Potential therapeutic targets include molecules which inhibit calcium efflux from the ER, such as dantrolene, via inhibition of the RyR [35], and rapamycin and pioglitazone, via inhibition of IP3R [34]. In neuronal WS models, cytosolic calcium appears to be increased under resting conditions and reduced under stimulated conditions [28]. This disruption in cytoplasmic calcium homeostasis also dysregulates mitochondrial dynamics which leads to lower ATP levels [28]. This is thought to hinder neuronal development and survival [28]. Under periods of ER stress, pancreatic ER kinase (PERK), a transmembrane ER protein, becomes activated and through the action of protein kinase A (PKA) and cyclic-AMP (cAMP) results in the phosphorylation of translation initiation factor 2α (eIF2α) (not shown) [36]. This in turn results in increased production of activating transcription factor 4 (ATF4) which increases the expression of genes for ER stress recovery [31, 36, 37]. Furthermore, phosphorylated eIF2α leads to decreased overall protein synthesis and therefore to reduction in the ER protein load (not shown) [36]. Glucagon-like peptide-1 receptor (GLP-1R) activation, acting downstream of PERK, decreases the phosphorylation of eIF2α via the PKA/cAMP pathway, in order to ameliorate the decrease in protein synthesis that would otherwise occur (not shown) [36, 37]. This mechanism of action of GLP-1R activity facilitates a faster resumption of protein synthesis following ER stress (not shown) [36, 37].

Therefore GLP-1R agonists have potential as therapeutic agents in patients with WS because of their role in decreasing ER stress mediated pancreatic β-cell apoptosis. Another strategy would be to use a DPP-4 inhibitor of the enzyme that deactivates glucagon-like peptide-1 (GLP-1), thus increasing GLP-1 levels.

In 2009, the role of pioglitazone, a thiazolidinedione, was studied in WS knockout mice that ordinarily develop insulin-dependent diabetes by an average age of 8 weeks [47]. These mice exposed to pioglitazone were found to be protected from pancreatic β-cell apoptosis and to be almost completely protected from the development of diabetes mellitus [47]. It has been hypothesized that pioglitazone inhibits inositol triphosphate receptor (IP3R) release of calcium from the endoplasmic reticulum (Figure 1) [32]. Pioglitazone and other thiazolidinediones may have a therapeutic role in the management of patients with WS. However, recent concerns linking thiazolidinediones use with increased risk of heart failure [48], osteoporosis [36, 37], and bladder cancer [49] make them a less attractive option than GLP-1R agonists or DPP-4 inhibitors. Furthermore, Cagaliniec et al. suggest that reducing cytosolic calcium has negative effects on mitochondrial dynamics [28].

In vitro models of WS (type 2) have shown the potential of iron chelation therapy to preserve neuronal cell and β-cell function [46]. Rapamycin is thought to reduce cytoplasmic calcium by a mechanism similar to pioglitazone (Figure 1) [32] but side effects and expense of rapamycin make it a less promising therapeutic option to investigate. Valproate has been shown to reduce ER stress-induced apoptosis in a model of diabetic nephropathy [50]. The molecular mechanisms for its diverse epigenetic effects on certain ER stress-related diseases have yet to be elucidated [51]. Valproate should be avoided in reproductive age women because of its
known teratogenicity. We are unaware of any clinical trials of valproate in WS but it has been designated an orphan drug for the treatment of WS [52].

In addition to the targeted drug approaches described above, gene based therapies which include adeno-associated virus [53] and Clustered Regularly Interspaced Short Palindromic Repeats (CRISPR) technology [7] are also being pursued. Mesencephalic astrocyte-derived neurotrophic factor (MANF) is also being tested as a method of preserving and proliferating existing β-cells and neurons [7].

4. Conclusions

The woman described in this report has biallelic mutations in WFS1, supporting a diagnosis of classical WS. She developed diabetes at a much older age than is typical of classical WS, had a successful pregnancy, continues to have significant residual β-cell function, and has outlived the median age of death for WS by over 12 years. Her more favourable clinical course may in part be due to the presence of a milder missense mutation. Recent advances in the pathophysiology of WS have informed potential targeted therapeutics aimed at reducing associated morbidities. Treatments which attempt to reduce ER stress or improve mitochondrial function may improve neurologic and β-cell survival. As a greater understanding of this rare monogenic disorder is gained, valuable insight may be gained into other ER stress and mitochondrial disorders and β-cell preservation for individuals with type 1 and 2 diabetes and various neurodegenerative diseases.

Conflicts of Interest

Dr. Suchowersky has research grants from Abbvie, Teva, Biotie, and Apopharma. She receives honoraria from Upto-Date and Springer. The other authors have no conflicts of interest to declare.

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