Association of Serum T cell Immunoglobulin Domain and Mucin-3 and Interleukin-17 with Systemic Lupus Erythematosus

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Background:  Previous studies have shown that T cell immunoglobulin domain and mucin-3 (Tim-3) and interleukin-17 (IL-17) are implicated in the development of several autoimmune diseases. However, it is unclear whether these proteins contribute to the pathogenesis of systemic lupus erythematosus (SLE). The purpose of this study was to evaluate SLE patient serum Tim-3 and IL-17 levels, and to assess correlations between these proteins and major clinical parameters of SLE.

Material/Methods:  Overall, 55 SLE patients and 55 healthy controls were recruited in a case-control study. Serum Tim-3 and IL-17 levels were quantified using an enzyme-linked immunosorbent assay (ELISA) kit.

Results:  Serum Tim-3 and IL-17 levels in SLE patients were significantly elevated relative to healthy controls (all \( P<0.05 \)). Serum Tim-3 levels were significantly lower in SLE patients with nephritis than in those SLE without nephritis (\( P<0.05 \)), while no statistically significant correlation between serum IL-17 and nephritis was detected (\( P>0.05 \)). Serum Tim-3 with IL-17 levels were positively correlated in SLE patients (\( r=0.817, P<0.01 \)); however, no statistically significant correlation was found between serum IL-17 and systemic lupus erythematosus disease activity index (SLEDAI) scores in those with SLE (all \( P>0.05 \)). In addition, serum Tim-3 was associated with central lesions in SLE patients, while there were no significant correlations between serum Tim-3 or IL-17 levels and other SLE clinical parameters.

Conclusions:  Increased serum Tim-3 and IL-17 levels and their clinical associations in SLE patients suggest their possible role in this disease.

MeSH Keywords:  Immunoglobulin Allotypes • Interleukin-17 • Lupus Vasculitis, Central Nervous System

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Background

Systemic lupus erythematosus (SLE) is a typical autoimmune disease, characterized by a large number of autoantibodies and the deposition of immune complexes, with diverse clinical manifestations causing persistent illness [1]. SLE prevalence is estimated to be up to 241/100 000 people, mainly affecting women of childbearing age [2]. Regulatory imbalances in helper T cells and their cytokines play important roles in the pathogenesis of SLE [3].

Interleukin-17A (IL-17), a major effect factor of Th17 cells, is a precursor to proinflammatory cytokines identified in recent years that are secreted by CD4+ T cells. Elevated IL-17 levels are associated with a higher experimental autoimmune encephalomyelitis (EAE) risk [10], increased multiple sclerosis (MS) severity [11], serious rheumatoid arthritis (RA) inflammation [12], and higher SLE disease activity [13], while decreased IL-17 levels can lead to lupus or RA-like symptoms [14]. However, the reliability of these findings remains controversial [15]. Therefore, the exact role of IL-17 in SLE warrants additional investigation.

Herein, to further examine the roles of Tim-3 and IL-17 in the pathogenesis of SLE, we evaluated serum Tim-3 and IL-17 levels in patients with SLE. Moreover, we analyzed their correlations with major clinical parameters.

Material and Methods

Subjects

A total of 110 participants, including 55 SLE patients and 55 healthy controls, were recruited from the Affiliated Yijishan Hospital of Wannan Medical College. SLE was defined based on the classification criteria revised by the American College of Rheumatology (ACR) in 1997 [16]. Disease activity was assessed based on the Systemic Lupus Erythematosus Disease Activity Index 2000 (SLEDAI-2K) [17]. Patients in the more active SLE group were those with SLEDAI-2K score >4, while those with less active SLE were those with a SLEDAI-2K score ≤4 [18,19]. Nephritis diagnosis was made according to the ACR criteria, which were as follows: 1) persistent proteinuria ≥0.5 g/day 2) the presence of active cellular casts, or 3) biopsy evidence of lupus nephritis. Exclusion criteria for all patients were as follows: a) patients with other connective tissue diseases, b) patients with malignancies, c) patients with other autoimmune diseases, and d) patients with viral infectious diseases. Patients were also excluded if they did not meet the inclusion criteria. Patients were randomly selected from among eligible cases in the ward of the hospital's Rheumatology Department based on patient bed numbers and a random number table. Sample sizes were calculated based on the previous study of Montaigne et al. [20]. Our hypothesis was that there would be a 50% (SD 51%) relative difference in serum Tim-3 levels between SLE patients and controls. We therefore recruited 55 patients and 55 controls to achieve a power of 80% for a significance level of 5% with a 2-tailed test in this study. The healthy control group was age- and sex-matched with the SLE group. Disease duration was calculated since the time of diagnosis. General demographic information and clinical parameters were obtained through epidemiological interviews and hospital records.

This study was approved by the Ethics Committee of Wannan Medical College, and informed consent was obtained from all participants.

Serum isolation and enzyme-linked immunosorbent assay (ELISA)

Serum was separated from 5 ml of whole blood collected from all participants, and was stored at −80°C. Serum levels of Tim-3 and IL-17 were assayed via quantitative sandwich ELISAs, following the manufacturer’s instructions for the Tim-3 and IL-17 kits (R&D Systems, Inc.). All ELISA results are expressed as cytokine concentrations (pg/ml). Serum from SLE patients and controls were analyzed together in the same laboratory, and the laboratory personnel were blind to patient disease status.

Statistical analysis

Numerical data conforming to normal distributions are presented as mean± standard deviation (SD), while those not normally distributed are presented as medians (interquartile range, IQR). Mann-Whitney rank sum tests or t tests were performed to estimate differences between groups for continuous variables.
### Table 1. The general characteristics of study subjects.

|                       | SLE group | Control group |
|-----------------------|-----------|---------------|
| **Number**            | 55        | 55            |
| **Age (years)**       | 37.7±13.6 | 37.4±12.2     |
| **Female, n (%)**     | 52 (94.55)| 52 (94.55)    |
| **Duration (years)**  | 4.42 (0.07, 9.00) | NA          |
| **SLEDAI**            | 16.00 (10.00, 20.00) | NA          |
| **Lupus Nephritis, n (%)** | 30 (54.55) | NA          |
| **Arthritis, n (%)**  | 26 (47.27) | NA          |
| **Rash, n (%)**       | 30 (54.55) | NA          |
| **Alopecia, n (%)**   | 24 (43.64) | NA          |
| **Central lesions, n (%)** | 12 (21.82) | NA          |
| **Visually impairment, n (%)** | 5 (9.10) | NA          |
| **Oral ulcers, n (%)**| 11 (20.00) | NA          |
| **Fever, n (%)**      | 37 (67.27) | NA          |
| **Headache, n (%)**   | 6 (10.91)  | NA          |
| **Thrombocytopenia, n (%)** | 12 (21.82) | NA          |
| **Leukopenia, n (%)** | 8 (14.55)  | NA          |
| **C3**                | 0.65 (0.47, 0.84) | NA          |
| **C4**                | 0.10 (0.04, 0.21) | NA          |
| **ESR**               | 41.00 (22.00, 68.50) | NA          |
| **CRP**               | 5.86 (1.98, 17.22) | NA          |
| **IgA**               | 2.39 (1.63, 3.27) | NA          |
| **IgG**               | 12.77 (8.86, 20.91) | NA          |
| **IgM**               | 1.01 (0.70, 1.38) | NA          |
| **Corticosteroids≤30mg/day, n (%)** | 28 (50.91) | NA          |
| **Corticosteroids>30mg/day, n (%)** | 18 (32.73) | NA          |
| **Anti-dsDNA, n (%)** | 35 (63.64) | NA          |
| **Anti-Sm, n (%)**    | 24 (43.64)  | NA          |
| **Anti-SSA, n (%)**   | 39 (70.91)  | NA          |
| **Anti-SSB, n (%)**   | 9 (16.36)   | NA          |
| **Anti-RNP, n (%)**   | 24 (43.64)  | NA          |
| **Anti-Ribosomal P, n (%)** | 18 (32.73) | NA          |
| **Antimalarial, n (%)** | 27 (49.09) | NA          |
| **Naproxine, n (%)**  | 49 (89.09)  | NA          |
| **Methotrexate**      | 8 (14.55)   | NA          |
| **Cyclophosphamide**  | 9 (16.36)   | NA          |

SLE – systemic lupus erythematosus; SLEDAI-2K – systemic lupus erythematosus disease activity index 2000; NA – not applicable. Numerical data conforming to the normal distribution are presented as means± standard deviation (SD); otherwise data are presented as medians (interquartile range, IQR).
Differences in categorical variables among groups were assessed using the chi-squared test or Fisher’s exact test. Correlation analyses were performed using Spearman’s rank correlation coefficient. Statistical analyses were conducted using SPSS software version 18.0 (SPSS, Inc, Chicago, IL). A 2-tailed \( P < 0.05 \) was viewed as significantly different.

**Results**

### Study subject demographics

The general characteristics of study subjects are presented in Table 1. The age (\( P=0.903 \)) and sex (\( P=1.000 \)) between the SLE group and the control group were not statistically different. The average disease duration and SLEDAI-2K scores for the SLE group were 4.42 years and 16.00 years, respectively. Participants with lupus nephritis accounted for 54.55% of SLE patients.

| Group                        | Number | Tim-3 (pg/ml) | IL-17 (pg/ml) |
|------------------------------|--------|---------------|---------------|
| SLE without nephritis        | 25     | 292.54 (277.85–370.70) | 4.89 (4.18–5.43) |
| SLE with nephritis           | 30     | 236.65 (206.57–288.02)* | 4.37 (3.65–5.23) |
| Less active SLE              | 6      | 198.40 (169.11–264.00) | 3.70 (3.02–4.80) |
| More active SLE              | 49     | 270.62 (222.25–332.35) | 4.71 (3.93–5.35) |

*\( P < 0.05 \) vs. SLE without nephritis.

**Table 2.** Comparison of Tim-3 and IL-17 levels between different SLE patient subgroups.

**Figure 1.** Comparison of serum Tim-3 and IL-17 levels between SLE patients and healthy controls. (A) The serum level of Tim-3 in the SLE and healthy control groups, and (B) the serum level of IL-17 in the SLE and healthy control groups. The serum concentrations (median) of Tim-3 and IL-17 in the SLE group were significantly higher than in the healthy control group (both \( P<0.05 \)).

**Figure 2.** Correlation of serum Tim-3 level with IL-17 level in SLE patients. The individual dots represent paired values of serum Tim-3 and IL-17 levels in SLE patients. The line shown is a linear univariate correlation. A significant positive correlation was observed between the serum levels of Tim-3 and IL-17 (\( r_s=0.817, P<0.001 \)).
Comparison of serum Tim-3 and IL-17 levels between SLE patients and healthy controls, and within SLE patient subgroups

The average levels of serum Tim-3 for SLE patients and controls were 252.71 (214.61, 332.33) and 229.42 (159.40, 297.03) pg/ml, respectively. The average levels of serum IL-17 for these 2 groups were 4.67 (3.74, 5.33) and 4.13 (3.00, 4.91) pg/ml, respectively. The levels of Tim-3 ($P=0.029$) and IL-17 ($P=0.022$) in SLE patients were significantly elevated relative to healthy controls (Figure 1).

The serum level of Tim-3 was significantly lower in SLE patients with nephritis as compared to SLE patients without nephritis ($P=0.020$). However, there was no significant difference in IL-17 levels between SLE patients with or without nephritis ($P=0.101$). Moreover, no significant differences in Tim-3 and IL-17 levels were identified between patients with less active SLE as compared to those with more active SLE ($P=0.344$ and $P=0.671$, respectively) (Table 2).

Correlation of serum Tim-3 level with IL-17 level in SLE patients

We analyzed the correlation between serum Tim-3 levels and IL-17 levels in SLE patients and found that these 2 factors were significantly positively correlated ($r_s=0.817$, $P<0.01$) (Figure 2), whereas they were not correlated in controls ($r_s=0.006$, $P=0.963$).

Correlations of serum Tim-3 and IL-17 levels with major clinical parameters in SLE patients

Correlations between serum Tim-3 and IL-17 levels and major clinical parameters in SLE patients were analyzed, revealing that the Tim-3 level in SLE patients with central lesions differed significantly from those in SLE patients without central lesions ($P=0.020$), but no significant differences in serum Tim-3 or IL-17 levels between other clinical parameters were observed (all $P>0.05$) (Table 3, Figures 3, 4).

Discussion

Monney et al. showed that Tim-3 acts as an immunosuppressive factor in Th1 cells [21]. IL-17 has been reported to be involved in the development of inflammation and plays an important role in autoimmune diseases [22,23]. In the present study, we identified increased serum Tim-3 and IL-17 levels in SLE patients. We further found that serum Tim-3 levels were significantly lower in SLE patients with nephritis than in those without nephritis. Moreover, a positive correlation between serum Tim-3 and IL-17 levels was observed in SLE patients. Serum Tim-3 levels in SLE patients with central lesions were significantly different from those in SLE patients without central lesions, but no significant differences in serum Tim-3 and IL-17 levels were identified for other clinical parameters in this case-control study.

Dysfunctional regulation of Th1 and Th2 cells and an imbalance in the Th1/Th2 cell ratio may be a primary mechanism of SLE development or progression [24], and SLE may also be mediated by differentiated Th1 and Th17 cells expressing Tim-3 [3]. Studies have found that Tim-3 expression in peripheral blood mononuclear cells (PBMCs) and CD3+ CD4+/CD3+ CD4– T cells in SLE patients was significantly higher relative to controls [8,25], similar to our results. This may be due to the upregulation of Tim-3 induced by the immune environment [26]. Other research

Table 3. Correlations of serum Tim-3 and IL-17 levels with quantitative clinical parameters in SLE patients.

| Parameters     | Tim-3 |          | Tim-3 |          |
|---------------|-------|----------|-------|----------|
|               | $r_s$ | $P$      | $r_s$ | $P$      |
| C3            | 0.237 | 0.081    | 0.147 | 0.284    |
| C4            | −0.027| 0.849    | −0.092| 0.510    |
| ESR           | 0.105 | 0.453    | 0.177 | 0.204    |
| CRP           | 0.129 | 0.359    | 0.139 | 0.321    |
| SLEDAI-2K     | 0.028 | 0.840    | −0.005| 0.973    |
| Disease duration | −0.213| 0.119    | −0.044| 0.751    |
| IgA           | 0.262 | 0.103    | 0.276 | 0.085    |
| IgG           | 0.211 | 0.191    | 0.118 | 0.468    |
| IgM           | 0.047 | 0.773    | −0.007| 0.964    |

SLE = systemic lupus erythematosus; Tim-3 = T cell immunoglobulin domain and mucin-3; IL-17 = interleukin-17; C3 = Complement 3; C4 = Complement 4; ESR = erythrocyte sedimentation rate; CRP = C-reactive protein; SLEDAI-2K = systemic lupus erythematosus disease activity index 2000; IgA = immunoglobulin A; IgG = immunoglobulin G; IgM = immunoglobulin M.
Comparison of serum Tim-3 levels with different categorical clinical parameters in SLE patients. SLE – systemic lupus erythematosus; Tim-3 – T cell immunoglobulin domain and mucin-3; IL-17 – interleukin-17; dsDNA – double stranded DNA; Sm – Smith; SSA – Sjögren’s syndrome-related antigen A; SSB – Sjögren’s syndrome-related antigen B; RNP – Ribonucleoprotein. Significant differences in serum Tim-3 levels were observed between SLE patients with and without central lesions ($P=0.020$). No significant differences in other clinical parameters were observed (all $P>0.05$).
Figure 4. Comparison of serum IL-17 levels with different categorical clinical parameters in SLE patients. SLE – systemic lupus erythematosus; Tim-3 – T cell immunoglobulin domain and mucin-3; IL-17 – interleukin-17; dsDNA – double stranded DNA; Sm – Smith; SSA – Sjögren’s syndrome-related antigen A; SSB – Sjögren’s syndrome-related antigen B; RNP – Ribonucleoprotein. No significant differences in serum IL-17 level were observed between patients with different clinical parameters (all $P>0.05$).
groups have found that Tim-3 mRNA levels in PBMCs from SLE patients were similar to those in healthy controls, which may be a consequence of the different levels of Tim-3 expression in different PBMC subtypes, including activated Th17 cells, macrophages/monocytes, dendritic cells, and natural killer cells [27]. Furthermore, SLE patient renal pathological grades were positively correlated with Tim-3 expression upon renal biopsy, further providing a pathological basis for the involvement of Tim-3 in the pathogenesis of SLE [28]. Both SLEDAI score and serum levels of complement proteins and C-reactive protein (CRP) can reflect SLE disease activity, and Tim-3 expression has been associated with SLEDAI score and serum levels of C3, C4m, and CRP, suggesting that Tim-3 may reflect SLE disease activity [8,29]. We have also found that central lesions are associated with Tim-3 levels, which may be related to the upregulation of Tim-3 on CD11b+ monocytes and intrinsic microglia infiltrating the central nervous system [30]. Moreover, serum Tim-3 levels were significantly lower in SLE patients with nephritis than in those without nephritis in the present study. This may be because the reduction in soluble Tim-3 is a compensatory protective response in the body to prevent excessive kidney damage. The high levels of soluble Tim-3 in the context of chronic inflammation represent a persistent response in the body due to a persistent failure to return to the non-inflamed steady state. In MRL/lpr lupus mice, a Tim-3 ligand (Galectin-9) has been linked to the severity of various rheumatic symptoms such as nephritis and arthritis due to Galectin-9-induced programmed cell death of Th1 and Th17 effector cells [31,32]. The 1516G>C single-nucleotide polymorphism (SNP) in the Tim-3 promoter region has also been associated with SLE susceptibility [33]. These studies demonstrate that Tim-3 is closely associated with the pathological mechanisms underlying SLE, acting by negatively regulating the Th1/Th17 immune response [34].

There is accumulating evidence that IL-17 is involved in autoimmune diseases such as experimental autoimmune meningoencephalomyelitis (EAE) [35], rheumatoid arthritis (RA) [12,36], autoimmune hepatitis [37], and autoimmune enteropathy [38]. Abdel Galil et al. have identified a positive correlation between IL-17 and 24-h proteinuria and high anti-ds-DNA titers in SLE patients [39]. We observed no significant association between IL-17 and auto-antibody levels or other major clinical manifestations, which may be a consequence of the small sample size and differences in efficacy of immunosuppressive drugs in the present study. Toll-like receptor 2 (TLR2) is an innate immune receptor that recognizes bacterial lipoproteins/lipopeptides, and increased expression of TLR2 promotes IL-17 expression via histone modifications, which result in IL-17 being implicated in early-onset SLE and pediatric SLE [13,40–42]. IL-17 is a major effector factor produced by Th17 cells, and in the present study we observed a significant positive correlation between Tim-3 and IL-17 levels in patient serum, which is consistent with previous work showing that Tim-3 is expressed in differentiated Th17 cells [43]. Tim-3 and IL-17 respond similarly to corticosteroids [44]. Therefore, soluble Tim-3 may be a surrogate marker for IL-17 in SLE patients. Th17 cells are proinflammatory effector CD4+ T cells, and the expression of Tim-3 on such cells has been confirmed [45]. Application of Galectin-9 can reduce levels of stimulated Th17 cells and Tim-3+ CD4+ T cells, and can inhibit IL-17 production, thereby reducing bacterial clearance [46]. Administration of antibodies against Tim-3 can significantly exacerbate the clinical and pathologic severity of EAE and can increase the number and activation level of macrophages [21,47]. The Tim-3/Galectin-9 pathway has been well established, and such signaling can lead to the apoptosis of effector CD4+ and CD8+ T cells [48]. This suggests that the Tim-3/Galectin-9 pathway is an immunosuppressive pathway in response to Th1 and Th17-mediated inflammation. In summary, CD4+ T cells differentiate into Th17 cells when activated in SLE patients, and IL-17 expression is consequently increased. Simultaneously, the expression of Tim-3 is increased and plays an immunosuppressive role via Tim-3/Galectin-9 signaling. These changes mediate the chronic inflammatory response observed in many patients, and targeted therapy influencing the Tim-3/Galectin-9 pathway is thus attractive for use in SLE patients.

The present study has several limitations. First, several SLE patients had established disease rather than being new-onset cases, and these results may therefore be affected by corticosteroids, immunosuppressive therapy, and other confounding factors. Second, the lack of a significant correlation between serum Tim-3/IL-17 levels and SLE disease activity index could be due in part to the relatively small sample size and limited statistical power of this study. Finally, distorted results in epidemiological association studies can result from potential biases in case-control studies. Consequently, further studies are needed to clarify the exact roles of Tim-3 and IL-17 in the pathogenesis of SLE.

Conclusions

Increased serum Tim-3 and IL-17 levels in SLE patients and their correlations with disease parameters suggest that they have key roles in modulating the course of this disease.

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Conflict of interest

None.
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