Genetic alterations in anaplastic thyroid carcinoma and targeted therapies (Review)

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Abstract. Thyroid cancer is the most common type of endocrine malignancy, and its incidence is increasing. Anaplastic thyroid cancer (ATC), referring to undifferentiated subtypes, is considered to be aggressive and associated with poor prognosis. Conventional therapies, including surgery, chemotherapy and radioiodine therapy, have been used for ATC, but these do not provide any significant reduction of the overall mortality rate. The tumorigenesis, development, dedifferentiation and metastasis of ATC are closely associated with the activation of various tyrosine cascades and inactivation of tumor suppressor genes, including B-Raf proto-oncogene, serine/threonine kinase V600E, phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit α, tumor protein 53 mutations and telomerase reverse transcriptase mutation. These pathways exert their functions individually or through a complex network. Identification of these mutations may provide a deeper understanding of ATC. A variety of tyrosine kinase inhibitors have been successfully employed for controlling ATC growth in vitro and in xenografts. Certain novel compounds are still in clinical trials. Multi-kinase inhibitors provide a novel approach with great potential. This systematic review determined the prevalence of the major genetic alterations and their inhibitors in ATC.

Contents

1. Introduction
2. PI3K/Akt/mTOR pathway in ATC
3. PI3K inhibitors
4. mTOR inhibitors
5. Dual PI3K/mTOR inhibitors
6. AKT inhibitors
7. TP53 in ATC
8. Introduction of wild-type p53
9. ‘Correction’ of mutant p53 protein
10. hTERT and BRAFV600E in advanced thyroid cancer
11. TERT inhibitors and BRAF inhibitors
12. Other agents and approaches
13. Conclusion

1. Introduction

Thyroid cancer is the most prevalent type of endocrine malignancy and accounts for 1% of cancer cases worldwide. Based on the degree of differentiation, thyroid cancers are categorized as differentiated thyroid cancer (DTC, including papillary thyroid carcinoma (PTC) and follicular thyroid carcinoma), as well as poorly-differentiated thyroid carcinoma (PDTC) and anaplastic thyroid cancer (ATC) (1). Most patients with DTC maybe effectively cured with standard primary treatments and have an excellent prognosis (2). Although ATC only accounts for 2% of thyroid cancer cases, it is responsible for more than half of all thyroid cancer mortalities due to its aggressive behavior and resistance to conventional therapies (3,4).

Recent molecular pathological studies have indicated that activation of various tyrosine cascades and inactivation of the tumor protein (TP) 53 tumor suppressor gene may induce the progression and dedifferentiation of ATC (5). According to a study on 516 patients from The Cancer Genome Atlas (TCGA) and Memorial Sloan Kettering Cancer Center (MSKCC) database, poorly differentiated thyroid carcinoma (including ATC) had more mutations in TP53, telomerase reverse transcriptase (TERT) and PI3K than PTC. The signaling pathways may exert their functions individually or through synergy with
other pathways (6,7). Besides conventional chemotherapy, multi-kinase-targeted inhibitors are emerging as novel therapeutic strategies (8). The aim of this review was to determine the prevalence of the major genetic alterations and report on the emerging kinase-targeted therapies in ATC.

2. PI3K/Akt/mTOR pathway in ATC

The PI3K pathway has a key role in regulating cell growth, proliferation and survival. This pathway is upregulated in two ways, firstly by the binding of the p85 subunit of PI3K to the subunits of activated tyrosine residues present on an activated growth factor receptor and secondly via direct recognition and combination of RAS and P110 (9). mTOR is a regulatory protein of the PI3K/Akt/mTOR pathway and its activation results in the phosphorylation of 4E-binding protein (4EBP1) and ribosomal protein S6 (S6k1) both of which regulate the transcription and translation of critical growth genes (10). Willems et al (11) reported that the activation of mTOR was higher in lymph node metastases than in primary thyroid cancers. Phosphorylated (p)-Akt then activates a variety of downstream factors, including glycolgen synthase kinase, Bad, the forkhead box family of transcription factors, p27 and Mdm2. The downstream targets of p-Akt have been demonstrated to increase cell proliferation, motility, protein synthesis and gluconeogenesis, as well as to inhibit apoptosis (12,13). p-Akt also affects the nuclear proteins including transcription factors and nuclear receptors, to form a unique signaling network (14).

The MSKCC and TCGA contains data for 516 thyroid cancer patients (84 PDTCs, 33 ATC cases and 399 PTC cases) and indicated that mutations of PI3K/Akt/mTOR (including mutations of phosphatidylinositol-4,5-bisphosphate 3-kinase catalytic subunit α (PIK3CA) and phosphatase and tens in homolog (PTEN), and of PIK3CA, PIK3CG,PIK3C3, PIK3R1, PIK3R2, AKT3, TSC subunit 1 (TSC1), TSC2 and mTOR) were more frequent in PDTCs and ATCs than in PTCs (Fig. 1A). Analysis of the association between PIK3CA and overall survival among the 117 PDTC and ATC cases indicated that patients with mutations of PIK3CA had an obviously shorter survival time (10.3 vs. 116.69 months; Fig. 1B and C). Kunstman et al (15) and Landa et al (16) performed next-generation sequencing of thyroid cancer patients and the results revealed that mutations of PI3K/Akt/mTOR were more frequent in ATCs than in other types of thyroid cancer.

All of the above suggests that the PI3K/mTOR pathway may be closely linked to thyroid cancer. Current research focuses on multiple inhibitors that interfere with different nodes of the PI3K pathway for use in combination with traditional therapy in clinical trials (Fig. 2).

3. PI3K inhibitors

NVP-BKM120 (Buparlisib), an inhibitor of pan-class I PI3K, has been proven to have a role in tumor suppression in a variety of cell lines and xenograft models, with or without PI3K alterations (17). NVP-BKM120 has a bioavailability of >90% and a half-life of 40 h according to a study on breast cancer (18). It has been proven that NVP-BKM120 suppresses downstream factors of PI3K, including downregulation of p-Akt and p-S6R and cancer stem markers. A previous study by our group suggested that synergistic action of BKM120 and Prima-I

4. mTOR inhibitors

RAD001 (everolimus), a rapamycin analog, is an allostERIC mTORC1 inhibitor. RAD001 has a therapeutic effect in tumors harboring alterations in the mTOR pathway (23). RAD001 has been tested in a phase II clinical trial (24) including 28 patients with progressive metastatic or locally advanced radioactive refractory DTC and 7 patients with ATC. The results indicated that 17 patients (65%) achieved stable disease (SD) as the best response, with 15 (58%) exhibiting SD lasting for >24 weeks. Furthermore, treatment with RAD001 increased progression-free survival in patients with metastatic cancer. As only 7 patients with ATC were included, it was not possible to draw any definite conclusions. The results of that study on patients with ATC were disappointing, with none of the patients benefiting from treatment. Certain pre-clinical trials evaluating the anti-tumor activity of everolimus, alone or in combination, are ongoing, suggesting that mTOR inhibitors including everolimus may be a promising treatment option for thyroid cancer (25).

5. Dual PI3K/mTOR inhibitors

BEZ235, a dual PI3K/mTOR inhibitor, reduces PI3K, mTORC1 and mTORC2 kinase activity, and consistently inactivates downstream signaling factors via competitive binding to the ATP binding domain of these enzymes (26,27). For dual PI3K/mTOR inhibition, BEZ235 has demonstrated a better effect in terms of therapeutic resistance and synergistic effects. BEZ235 has been effectively evaluated in vitro on several thyroid cancer cell lines derived from major pathological types, with ATC exhibiting the greatest sensitivity. BEZ235 generally induces cell cycle arrest at the G0/G1 phase, and also causes apoptosis in the most sensitive thyroid cell lines. The experiments also established that daily treatment with BEZ235 at a dose of 50 mg/kg significantly reduced the growth of xenograft tumors composed of 8505C ATC cells, without any toxicity observed (27). In addition, BEZ235 was able to increase the expression of Na⁺/I⁻ symporter (NIS) and other thyroid-specific genes, leading to an increase in or the restoration of the sensitivity to radioactive iodine (RAI) (28).

6. AKT inhibitors

Akt is a central and essential point in the PI3K pathway, and is therefore a promising target for anti-cancer therapies. Akt
inhibitors maybe grouped into various classes, including ATP-competitive inhibitors (GSK690693 and Afuresertib), allosteric inhibitors (MK-2206 and PH-316) and irreversible inhibitors (LL-AF101) (29). MK-2206 is a selective inhibitor of all Akt isoforms, decreasing p-Akt Thr308 and p-Akt Ser473 levels as well as downstream phosphorylation. MK-2206 was demonstrated exert dose- and time-dependent effects on different thyroid cancer cells lines. It potently inhibited the proliferation of all thyroid cancer cells in a low-micromolar range (IC50 mostly below or around 0.5 µM). When used synergistically with temsirolimus, MK2206 was able to completely overcome the side effect of mTOR inhibition of Akt (30) therefore, MK-2206 is a good candidate for further investigation as a treatment for ATC.

7. TP53 in ATC

TP53 lies at the center of a large network of apoptosis-, invasion- and stem cell-associated genes. The p53 protein is composed of a core DNA-binding domain (DBD) and regulatory domains, serving as a major barrier against tumorigenesis (31). Of note, numerous in vitro and xenograft models...
have confirmed that p53 mutation not only abolishes the tumor suppressive function, but also often acquires new tumorigenic driver activities. This negative role was termed gain-of-function (GOF). Inactivation of p53 has been considered a hallmark of advanced thyroid tumors (32). TP53 mutations have been identified in 28.2% of PTCs and in 1% of ATCs and PDTCs. Among the 33 ATC and 84 PDTC patients from MSKCC dataset, the mutation group had a median survival time of only 10 months (124.08 in the wild-type TP53 group; Fig. 3A and B). Mutant p53 enhances signaling through receptors such as transforming growth factor β receptor, epidermal growth factor receptor and MET. Unlike wild-type p53, mutant p53 protein has been verified to escape proteasome-dependent degradation, leading to its hyper-stabilization in tumors (33). The degradation resistance of mutant p53 presents a fundamental problem for therapeutic intervention in tumors with mutant p53.

Restoration of p53 function is essential for promoting the sensitivity to chemotherapeutic drugs or radiation therapy. This hypothesis is supported by a study in which redifferentiation and restoration of cellular responses to physiological stimuli were achieved after re-expression of wild-type p53 in ATC (34). Table I summarizes the small molecules that restore the function of the tumor suppressor gene p53.

8. Introduction of wild-type p53

Restoration of wild-type p53 function may be achieved by introduction of an intact complementary DNA copy of the p53 gene using a suitable viral vector, in most cases an adenoviral vector [recombinant adenovirus-p53 (Adp53)]. ONYX-015, the most prominent and clinically evaluated replication-competent Adp53 vector, has been proven to have a tumor-specific effect, and proliferates effectively in p53-mutant, but not in p53 wild-type cells (35). Furthermore, suppression of p21 via p21-targeting micro-ribonucleic acids may effectively induce Adp53-mediated apoptosis and autophagy in human cancer cells (36). Subsequent studies demonstrated that a higher clinical efficacy was achieved when ONYX-015 was combined with conventional chemotherapeutic drugs or radiation. Numerous clinical trials have been performed in different types of advanced cancers patients (37,38). However, numerous patients still respond poorly (39). Gendicine (rAd-p53), a first-generation gene therapeutic, has been licensed for clinical use for head and neck malignancies in China. In randomized trials on nasopharyngeal and pancreatic carcinoma, better control was observed when Gendicine (weekly intra-tumoral injections) was combined with radiation or chemoradiation therapy (40). However, to date, these vectors have not been widely used in patients worldwide. The precise mechanism of action and clinical anti-tumor effect of these vectors requires further exploration.

9. Enhancement of the functionality of endogenous wild-type p53

As mentioned above, the p53 pathway is most likely also disrupted in a large fraction of wild-type p53-carrying tumors. Mdm2, a critical negative regulator acting via ubiquitin-mediated p53 degradation, is frequently overexpressed in wild-type TP53-carrying tumors, leading to resistance to p53 gene therapy for cancer (41). Furthermore, mutant p53 is overexpressed in numerous tumors due to a lack of sufficient amounts of Mdm2 to trigger p53 degradation. For these reasons, Mdm2 has been an important therapeutic target. Different strategies for targeting Mdm2 and/or inhibition of p53-Mdm2 binding
have been designed, including Nutlins, reactivation of p53 and induction of tumor cell apoptosis (RITA) and inhibitor of Hdm2 HLI98ubiquitin ligase (HLI 373), which have the ability to increase p53 levels and transcriptional activity (42). Nutlin-3a stabilizes and activates p53 by blocking p53 binding to Mdm2, leading to the expression of downstream genes of p53, including p21, BAX and p53 upregulated modulator of apoptosis (43). RITA binds to the N-terminus of p53 and causes a configurable change, resulting in accumulation of p53 and upregulation of its target genes. RITA induces apoptosis in wild-type p53-harboring cancer cells, but has little side effects on normal cells (44). The availability of these therapies raises hope for the treatment of wild-type TP53-carrying tumors, with fewer side effects than traditional chemotherapeutic drugs.

10. ‘Correction’ of mutant p53 protein

As a result of the unfolding of the DBD, mutant p53 loses its role as a tumor suppressor gene and promotes the development of tumor progression. Therefore, pharmacological compounds to reactivate mutant p53 through changing it to the wild-type conformation and activating its transcription have been developed. These small-molecular drugs, including CP-31398, WR1065, PRIMA-1, PRIMA-1MET (APR-246), Ellipticine and MIRA-1, have demonstrated positive effects in cancer, including induction of massive apoptosis, inhibition of invasion and tumor stem cell suppression (45). PRIMA-1MET is able to not only restore the wild-type conformation of mutant p53, but also that of mutant N-terminal transactivation domain of TAp63γ (TAp63γ γ) and N-terminal transactivation domain of TAp73β (TAp73β) in tumor cells (46). Messina et al (47) successfully validated that PRIMA-1MET prevented the GOF effect of mutant p53 and increased the expression of thyroid-specific differentiation markers, including Tg and NIS, in thyroid cancer cells. PRIMA-1MET is less effective in wild-type or null p53 thyroid cell lines (BC-PAP and Hth-74 cell viability was significantly reduced at 1 µM; by contrast, Prima-1 at up to 20 µM had no effect on TPC-1 and SW-1736 cells). The use of PRIMA-1MET in combination with irradiation and novel targeted tyrosine kinase inhibitors is a novel research hotspot.

11. Human TERT and B-Raf proto-oncogene, serine/threonine kinase (BRAF)\textsuperscript{V600E} in advanced thyroid cancer

Telomerase has a key role in cellular immortality and tumorigenesis. Its catalytic subunit is TERT. Accumulating evidence indicates that TERT promoter mutations are associated with aggressive, metastatic and thyroid stem cell phenotypes (48,49). In the dataset of TGCA database, ~40% of PDTCs and 73% of ATCs harbored TERT promoter mutations as compared with 9% of PTCs. In addition, TERT promoter mutations were rarely detected in normal parenchyma or in benign lesions (50,51). BRAF, a member of the RAF family, is a serine-threonine kinase. The BRAF\textsuperscript{V600E} mutation, a crucial stimulator of the mitogen-activated protein kinase pathway, is associated with the radioiodine resistance due to block of NIS (Na\textsuperscript{+}/I\textsuperscript{-}) symporter expression. BRAF\textsuperscript{V600E} is more frequently encountered in the dedifferentiated subtype (52). TERT promoter mutation is closely combined with BRAF\textsuperscript{V600E} and RAS mutations. Studies have indicated that the coexistence of somatic mutations, BRAF\textsuperscript{V600E} and TERT C228T, is strongly associated with aggressive phenotypes, poor prognosis and recurrence of ATC (53). Analysis of the clinical data of the 84 PDTC and 33 ATC cases from MSKCC revealed that co-existence of BRAF and TERT mutations has a synergistic effect on aggressiveness.
in thyroid carcinoma development. Tumors with both mutations are associated with a high risk of recurrence and shorter overall survival (Fig. 4A and B). Charles et al (54) reported that in the mouse model, PIK3CA was unable to drive thyroid tumorigenesis independently; however, in mice with both PIK3CA and BRAF^{V600E} mutations, ATC was observed. Of note, in ATC cases featuring BRAF^{V600E} in combination with PIK3CA or TP53 mutations, few, if any, mutations in other known ATC-associated genes were observed. The mechanism resulting in progression from DTC to ATC has been a subject of frequent study with accumulation of mutations in recognized malignancy-associated genes identified. Therefore, combination of multiple targeted drugs may achieve an effective breakthrough in the treatment of ATC.

### 12. TERT inhibitors and BRAF inhibitors

Vemurafenib (PLX4032) and Dabrafenib (GSK2118436), highly selective for BRAF^{V600E}-mutant cells, have been approved by the US Food and Drug Administration for melanoma therapy (55). The first report of using vemurafenib to treat metastatic papillary thyroid carcinoma was in 2013. Vemurafenib appears to have a promising clinical efficacy in patients with metastatic PTC (56). In a phase II clinical trial, 51 patients (patients with RAI-refractory PTC) were enrolled. Treatment with BRAF inhibitors for redifferentiation and RAI reuptake in BRAF^{V600E}-mutant thyroid cancer was also assessed. The clinical results suggested that the effect was better at first use (57). Marten and Gudena (58) reported on an ATC patient

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**Table I. Compounds that induce reactivation of mutant p53.**

| Type of drug                            | Drug                  | Mechanism                                                                 |
|----------------------------------------|-----------------------|---------------------------------------------------------------------------|
| Adenovirus gene therapy                | Advexin               | Exogenous import and increase of wide-type p53 expression                 |
|                                        | ONYX-015              | Exogenous import and increase of wide-type p53 expression                 |
|                                        | CP-31398              | Stabilize the DNA-binding core domain induce conformational change       |
| Compounds that induce reactivation of | PRIMA-1               | Bind to thiol groups in the core domain and restore wide-type conformation|
| mutant p53                             | PRIMA-1^{Met}         | Conformational change                                                     |
|                                        | RITA                  | Restore p53 transcriptional activity                                       |
| Compounds that deplete mutant p53     | 17-AAG                | Hsp90 inhibitors, increase the mutant p53 degradation                     |
|                                        | Geldanamycin          | Disrupt the HDAC6/Hsp90/mutant p53 complex                                |
|                                        | LBH589                |                                                                           |
|                                        | SAHA                  |                                                                           |

Hsp, heat shock protein; HDAC, histone deacetylase; RITA/NSC 652287; 17-AAG/Tanespimycin; LBH589/Panobinostat/NVP-LBH589; SAHA/Vorinostat, suberanilo hydroxamic acid.

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**Figure 4.** TERT and BRAF mutations in thyroid cancers. (A) Kaplan-Meier survival in PDTCs and ATCs with log-rank P-values (P=0.001). (B) Overall median survival analysis of 117 PDTC and ATC cases. PDTC, poorly differentiated thyroid tumor; ATC, anaplastic thyroid cancer; Cum, cumulative; BRAF, B-Raf proto-oncogene, serine/threonine kinase; TERT, telomerase reverse transcriptase; mut, mutation.
with an early clinical response, demonstrating a decrease in the subcutaneous metastasis, but follow-up computed tomography at 2 months after initiation of vemurafenib revealed rapid progression of the disease with metastases to the central nervous system and esophagus and progression of pulmonary metastasis. Other BRAF inhibitors, including PLX-4720 and MLN2480, are also being tested in the clinic, but have not been evaluated in thyroid cancer. Clinical data indicate that these inhibitors significantly improve response rates and overall survival in patients with BRAF\textsuperscript{V600E}-mutant metastatic melanoma (59,60). Maggisano et al (61) proved that inhibition of TERT expression by small interfering RNA significantly depressed the proliferation and invasion of ATC cells. BIBR1532, a selective telomerase inhibitor, decreases native and recombinant human telomerase activity, leading to senescence of human cancer cells. A study by Bu et al (62) reported that BIBR1532 effectively decreased the invasion, migration and angiogenesis of PTC cells \textit{in vivo} and \textit{in vitro}. However, at present, studies assessing the efficacy of BIBR1532 in the treatment of ATC are rare.

13. Other agents and approaches

NF-\kappa B allows thyroid cancer cells to acquire invasive properties and undergo metastasis by upregulating the expression of matrix metalloproteinases and urokinase-type plasminogen activator (63). Triptolide has been used in preclinical studies on ATC, and has demonstrated an inhibitory effect on angiogenesis and invasion (64). More clinical research is required to evaluate the synergistic effect with other chemotherapeutic agents for thyroid cancer. The Wnt-\beta-catenin signaling pathway is associated with cell adhesion and differentiation (65). Accumulation of \beta-catenin may induce nuclear transport and combine with certain transcription factors, which may activate the transcription of downstream factors, including c-myc and cyclin D1. The Wnt-\beta-catenin pathway may also regulate the level of cyclin D1, which is associated with lymph node metastases in thyroid cancer patients. Dickkopf-1, a selective inhibitor of the Wnt-\beta-catenin pathway, effectively inhibits the proliferation and migration of several thyroid cancer cell lines by regulating Wnt-\beta-catenin and E-cadherin expression (66). Certain genes regulate each other and form networks to produce a biological effect. Further signaling pathways are also associated with the development and progression of ATC. Sorafenib is a multikinase inhibitor that targets vascular endothelial growth factor receptor (VEGFR), platelet-derived growth factor receptor (PDGFR) and RAF. In the phase II trial of sorafenib, 10 patients with ATC were enrolled and no objective responses were observed (67). Another two trials assessing sorafenib in thyroid carcinomas included six ATC patients and similarly no responses were achieved (68,69). These results call into question the benefit of this agent in ATC patients. Another study involved 15 ATC patients treated with pazopanib (a multikinase inhibitor targeted on VEGFR, PDGFR and c-kit) in a single-arm, phase II study, with no responses obtained according to the Response Evaluation Criteria in Solid Tumors (70). Lenvatinib, a newer, small-molecule VEGFR inhibitor, was tested in differentiated thyroid cancer, medullary thyroid carcinoma and ATC in a phase II study. Only 11 patients with ATC were recruited; however, the results were encouraging, with three patients exhibiting a partial response according to the Response Evaluation Criteria in Solid Tumors, seven with SD and one patient exhibiting progression of disease (71). Lenvatinib has been approved in the USA for the treatment of DTC, but it is approved for all subtypes of thyroid cancer in Japan (67).

Genomic and epigenetic alterations are now being exploited as molecular targets in thyroid cancer treatment. The abnormalities of the histones in post-translational modification have been demonstrated in thyroid cancer development, and they are regarded as promising molecular targets for the patients that are resistant to conventional therapies. Compounds/drugs that reverse the effects of histone modifications by modulating the pattern of histone acetylation/methylation have been tested in preclinical models of thyroid cancer to identify their effects on tumor cell proliferation and/or invasiveness, and their ability to re-differentiate tumor cells and restore their ability to accumulate radioactive iodine (72,73).

14. Conclusion

Precision medical therapy is an exciting technique for individualized tumor treatment based on the patient's features and tumor characteristics. ATC is one of the most aggressive human cancer types and is associated with a low survival rate. Dysfunction of signaling pathways may increase the proliferation, dedifferentiation and metastasis of thyroid cancer. Inhibition of protein tyrosine kinases has been indicated to hold great promise. These drugs may not only provide a better understanding of tumor biological characteristics, but also more optimistic outcomes in the future. Combination of tyrosine kinases inhibitors with traditional radiotherapy and chemotherapy may represent an improved treatment strategy.

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Availability of data and materials

The datasets used and/or analyzed during the present study are available from the corresponding author on reasonable request.

Authors' contributions

LZ and ZL contributed to the design and conception of the study, and revised it carefully for important intellectual content. YZ was responsible for acquiring the data by screening the papers identified on Pubmed and TCGA. RW revised the study critically for important intellectual content. KZ and ZL were involved in drafting the study. YZ analyzed and interpreted the data. All authors read and approved the final manuscript.
Ethics approval and consent to participate
Not applicable.

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Competing interests
The authors declare that they have no competing interests.

References
1. O'Neill JP and Shaha AR: Anaplastic thyroid cancer. Oral Oncol 49: 702-706, 2013.
2. Jin S, Borkhau O, Bao W and Yang YT: Signaling pathways in thyroid cancer and their therapeutic implications. J Clin Med 8: 286-306, 2019.
3. Hsu KT, Yu XM, Audhwa YA, Jaume JG, Lloyd RV, Miyamoto S, Prolla TA and Chen H: Novel approaches in anaplastic thyroid cancer therapy. Oncologist 19: 1148-1155, 2014.
4. Keutgen XM, Sadowski SM and Kebebew E: Management of anaplastic thyroid cancer. Gland Surg 4: 44-51, 2015.
5. Saito Y, Mafani AW, Burman KD and Prabhakar BS: Genetic aberrations and alterations in signaling cascades implicated in the pathogenesis of anaplastic thyroid cancer. Biochim Biophys Acta Rev Cancer, 2018 (Epub ahead of print).
6. Guerra A, Di Crescenzo V, Garzi A, Cinelli M, Carlonmagno C, Tonacchera M, Zeppe P and Vitale M: Genetic mutations in the treatment of anaplastic thyroid cancer: A systematic review. BMC Surg 13 (Suppl 2): S44, 2013.
7. Xu B and Ghoseein R: Genomic landscape of poorly differentiated and anaplastic thyroid carcinoma. Endocr Pathol 27: 209-212, 2016.
8. Perri F, Pezzullo L, Chiofalo MG, Lastoria S, Di Gennaro F, Scarpati GD and Capanogo F: Targeted therapy: A new hope for thyroid carcinomas. Crit Rev Oncol Hematol 94: 55-63, 2015.
9. Bartholomeusz C and Gonzalez-Angulo AM: Targeting the PI3K/Akt/mTOR axis in cancer therapy. Expert Opin Ther Targets 16: 121-130, 2012.
10. Saji M and Ringel MD: The PI3K-Akt-mTOR pathway in initiation and progression of thyroid tumors. Mol Cell Endocrinol 321: 20-28, 2010.
11. Willems L, Tamburini J, Chapuis N, Lacombe C, Mayeux P and Willems L: PI3K and mTOR signaling pathways in cancer: New data on targeted therapies. Curr Opin Oncol 14: 129-138, 2002.
12. Cao F, Zhang C, Han W, Gao XJ, Liu SY, Ma C and Zou XP: Clinical evaluation of epithelial-mesenchymal transition and cancer stem cell phenotypes. J Clin Endocrinol Metab 97: 2956-2965, 2012.
13. Liu R, Liu D, Trink E, Bojdani E, Ning G and Xing M: The specific-activity inhibitor MK2206 selectively inhibits thyroid cancer cells harboring mutations that can activate the PI3K/Akt pathway. J Clin Endocrinol Metab 96: E577-E855, 2011.
14. Pflaum J, Smitha TD, Smit JW, Georgescu CE and Netae-Maier RT: PI3K/Akt/mTOR: A promising therapeutic target for non-medullary thyroid carcinoma. Cancer Treat Rev 41: 707-713, 2015.
15. Nitulescu GM, Margina D, Juzenas P, Peng Q, Oluaro OT, Saloustros E, Fenga C, Spanidios DA, Libra M and Tsatsakis AM: Akt inhibitors in cancer treatment: The long journey from drug discovery to clinical use (Review). Int J Oncol 40: 869-885, 2016.
16. Liu R, Liu D, Trink E, Bojdani E, Ning G and Xing M: The specific-activity inhibitor MK2206 selectively inhibits thyroid cancer cells harboring mutations that can activate the PI3K/Akt pathway. J Clin Endocrinol Metab 96: E577-E855, 2011.
17. Pflaum J, Smitha TD, Smit JW, Georgescu CE and Netae-Maier RT: PI3K/Akt/mTOR: A promising therapeutic target for non-medullary thyroid carcinoma. Cancer Treat Rev 41: 707-713, 2015.
18. Nitulescu GM, Margina D, Juzenas P, Peng Q, Oluaro OT, Saloustros E, Fenga C, Spanidios DA, Libra M and Tsatsakis AM: Akt inhibitors in cancer treatment: The long journey from drug discovery to clinical use (Review). Int J Oncol 40: 869-885, 2016.
19. Liu R, Liu D, Trink E, Bojdani E, Ning G and Xing M: The specific-activity inhibitor MK2206 selectively inhibits thyroid cancer cells harboring mutations that can activate the PI3K/Akt pathway. J Clin Endocrinol Metab 96: E577-E855, 2011.
20. Pflaum J, Smitha TD, Smit JW, Georgescu CE and Netae-Maier RT: PI3K/Akt/mTOR: A promising therapeutic target for non-medullary thyroid carcinoma. Cancer Treat Rev 41: 707-713, 2015.
21. Nitulescu GM, Margina D, Juzenas P, Peng Q, Oluaro OT, Saloustros E, Fenga C, Spanidios DA, Libra M and Tsatsakis AM: Akt inhibitors in cancer treatment: The long journey from drug discovery to clinical use (Review). Int J Oncol 40: 869-885, 2016.
22. Liu R, Liu D, Trink E, Bojdani E, Ning G and Xing M: The specific-activity inhibitor MK2206 selectively inhibits thyroid cancer cells harboring mutations that can activate the PI3K/Akt pathway. J Clin Endocrinol Metab 96: E577-E855, 2011.
23. Pflaum J, Smitha TD, Smit JW, Georgescu CE and Netae-Maier RT: PI3K/Akt/mTOR: A promising therapeutic target for non-medullary thyroid carcinoma. Cancer Treat Rev 41: 707-713, 2015.
24. Nitulescu GM, Margina D, Juzenas P, Peng Q, Oluaro OT, Saloustros E, Fenga C, Spanidios DA, Libra M and Tsatsakis AM: Akt inhibitors in cancer treatment: The long journey from drug discovery to clinical use (Review). Int J Oncol 40: 869-885, 2016.
41. Wade M, Li YC and Wahl GM: MDM2, MDMX and p53 in oncogenesis and cancer therapy, Nat Rev Cancer 13: 83-96, 2013.
42. Oren M, Tal P and Rotter V: Targeting mutant p53 for cancer therapy. Aging (Albany NY) 8: 1159-1160, 2016.
43. Selivanova G: Wild type p53 reactivation: From lab bench to clinic. FEBS Lett 588: 2628-2638, 2014.
44. Urso L, Calabrese F, Favaretto A, Conte P and Passello G: Critical review about MDM2 in cancer: Possible role in malignant mesothelioma and implications for treatment. Crit Rev Oncol Hematol 97: 220-230, 2016.
45. Puca R, Nardinocchi L, Porru M, Simon AJ, Rechavi G, Leonetti C, Givol D and D’Orazio G: Restoring p53 active conformation by zinc increases the response of mutant p53 tumor cells to anticancer drugs. Cell Cycle 10: 1679-1689, 2011.
46. Bykov VJ and Wiman KG: Mutant p53 reactivation by small molecules makes its way to the clinic. FEBS Lett 588: 2622-2627, 2014.
47. Messina RL, Sanfilippo M, Vella V, Pandini G, Vigneri P, Nicolosi ML, Giani F, Vigneri R and Frasca F: Reactivation of p53 mutants by prima-1 [corrected] in thyroid cancer cells. Int J Cancer 130: 2259-2270, 2012.
48. Liu X, Bishop J, Shan Y, Pai S, Liu D, Murugan AK, Sun H, El-Naggar AK and Xing M: Highly prevalent TERT promoter mutations in aggressive thyroid cancers. Endocr-Relat Cancer 20: 603-610, 2013.
49. Melo M, da Rocha AG, Vinagre J, Batista R, Peixoto J, Tavares C, Celestino R, Almeida A, Salgado C, Eloy C, et al: TERT promoter mutations are a major indicator of poor outcome in differentiated thyroid carcinomas. J Clin Endocrinol Metab 99: E754-E765, 2014.
50. Liu R and Xing M: TERT promoter mutations in thyroid cancer. Endocr-Relat Cancer 23: R143-R155, 2016.
51. Jin A, Xu J and Wang Y: The role of TERT promoter mutations in postoperative and preoperative diagnosis and prognosis in thyroid cancer. Medicine (Baltimore) 97: e11548, 2018.
52. Dong H, Shen WZ, Yan YJ, Yi JL and Zhang L: Effects of BRAF(V600E) mutation on Na(+)/I(-) symporter expression in papillary thyroid carcinoma. J Huazhong Univ Sci Technolog. Med Sci 36: 77-81, 2016.
53. Shi X, Liu R, Qu S, Zhu G, Bishop J, Liu X, Sun H, Shan Z, Wang E, Luo Y, et al: Association of TERT promoter mutation 1,295,228 C>T with BRAF V600E mutation, older patient age, and distant metastasis in anaplastic thyroid cancer. J Clin Endocrinol Metab 100: E632-E637, 2015.
54. Charles RP, Silva J, Iezza G, Phillips WA and McMahon M: Activating BRAF and PIK3CA mutations cooperate to promote anaplastic thyroid carcinogenesis. Mol Cancer Res 12: 979-986, 2014.
55. Zhang W: BRAF inhibitors: The current and the future. Curr Opin Pharmacol 23: 68-73, 2015.
56. Kim KB, Cabanillas ME, Lazor AJ, Williams MD, Sanders DL, Ilagan JL, Nolop K, Lee RJ and Sherman SI: Clinical responses to vemurafenib in patients with metastatic papillary thyroid cancer harboring BRAF(V600E) mutation. Thyroid 23: 1277-1283, 2013.
57. Brose MS, Cabanillas ME, Cohen EE, Wirth LJ, Richel T, Yue H, Sherman SI and Sherman EF: Vemurafenib in patients with BRAF(V600E)-positive metastatic or unresectable papillary thyroid cancer refractory to radioactive iodine: A non-randomised, multicentre, open-label, phase 2 trial. Lancet Oncol 17: 1272-1282, 2016.
58. Marten KA and Gudena VK: Use of vemurafenib in anaplastic thyroid carcinoma: A case report. Cancer Biol Ther 16: 1430-1433, 2015.
59. Cabanillas ME, Patel A, Danysh BP, Dadu R, Kopez S and Falchook G: BRAF inhibitors: Experience in thyroid cancer and general review of toxicity. Horm Cancer 6: 21-36, 2015.
60. Lim AM, Taylor GR, Fellowes A, Cameron L, Lee B, Hicks RJ, McArthur GA, Angel C, Solomon B and Rischin D: BRAF Inhibition in BRAFV600E-Positive Anaplastic Thyroid Carcinoma. J Nal Compr Canc Netw 14: 249-254, 2016.
61. Maggisi V, Celano M, Lombardo GE, Lepore SM, Sponzillo M, Rosignolo F, Verrienti A, Baldan F, Puxeddu E, Durante C, et al: Silencing of hTERT blocks growth and migration of anaplastic thyroid cancer cells. Mol Cell Endocrinol 448: 34-40, 2017.
62. Bu R, Siraj AK, Divya SP, Kong Y, Parvathareddy SK, Al-Rasheed M, Al-Obaiasi KAS, Victoria IG, Al-Sobhi SS, Al-Dawish M, et al: Telomerase reverse transcriptase mutations are independent predictor of disease-free survival in middle eastern papillary thyroid cancer. Int J Cancer 142: 2028-2039, 2018.
63. Namba H, Saenko V and Yamashita S: Nuclear factor-kB in thyroid carcinogenesis and progression: A novel therapeutic target for advanced thyroid cancer. Arq Bras Endocrinol Metabol 51: 843-851, 2007.
64. Zhu W, He S, Li Y, Qiu P, Shu M, Ou Y, Zhou Y, Leng T, Xie J, Zheng X, et al: Anti-angiogenic activity of triptolide in anaplastic thyroid carcinoma is mediated by targeting vascular endothelial and tumor cells. Vascul Pharmacol 52: 46-54, 2010.
65. Sastre-Pemon A and Santisteban P: Wnt-independent role of β-catenin in thyroid cell proliferation and differentiation. Mol Endocrinol 28: 681-695, 2014.
66. Wang L, Shao YY and Ballock RT: Thyroid hormone interacts with the Wnt/beta-catenin signaling pathway in the terminal differentiation of growth plate chondrocytes. J Bone Miner Res 22: 1988-1995, 2007.
67. Ito Y, Onoda N, Ito KI, Sugitani I, Takahashi S, Yamaguchi I, Kabu K and Tsukada K: Sorafenib in Japanese patients with locally advanced or metastatic medullary thyroid carcinoma and anaplastic thyroid carcinoma. Thyroid 27: 1142-1148, 2017.
68. Gupta-Abraham V, Troxel AB, Nellore A, Puttaswamy K, Redlinger M, Ransone K, Mandel SJ, Flaherty KT, Loevner LA, D’Owyer PJ and Brose MS: Phase II trial of sorafenib in advanced thyroid cancer. J Clin Oncol 26: 4714-4719, 2008.
69. Kloos RT, Ringel MD, Knopp MV, Hall NC, King M, Stevens R, Liang J, Wakely PE Jr, Vasko VV, Sajj M, et al: Phase II trial of sorafenib in metastatic thyroid cancer. J Clin Oncol 27: 1675-1684, 2009.
70. Bible KC, Suman VJ, Menefee ME, Smallridge RC, Molina JR, Maples WJ, Karlin NJ, Traynor AM, Kumar P, Goh BC, et al: A multiinstitutional phase 2 trial of pazopanib monotherapy in advanced anaplastic thyroid cancer. J Clin Endocrinol Metab 97: 3179-3184, 2012.
71. Takahashi S, Kiyota N, Yamazaki T, Chayahara N, Nakano K, Inagaki L, Toda K, Enokida T, Minami H, Imamura Y, et al: A Phase II study of the safety and efficacy of lenvatinib in patients with advanced thyroid cancer. Future Oncol 15: 717-726, 2019.
72. Sasanakietkul T, Murtha TD, Javid M, Korah R and Carling T: Epigenetic modifications in poorly differentiated and anaplastic thyroid cancer. Mol Cell Endocrinol 469: 23-37, 2018.
73. Celano M, Mio C, Sponzillo M, Verrienti A, Bulotta S, Durante C, Damante G and Russo D: Targeting post-translational histone modifications for the treatment of non-medullary thyroid cancer. Mol Cell Endocrinol 469: 38-47, 2018.