Pre-S Deletion and Complex Mutations of Hepatitis B Virus Related to Young Age Hepatocellular Carcinoma in Qidong, China

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Abstract

Background/Aim: To investigate the roles of biomedical factors, hepatitis B virus (HBV) DNA levels, genotypes, and specific viral mutation patterns on the progression of hepatocellular carcinoma (HCC) patients below 40 years of age in Qidong, China.

Methods: We conducted a case-control study within a cohort of 2387 male HBV carriers who were recruited from August, 1996. The HBV DNA sequence was determined in 49 HCC and 90 chronic hepatitis (CH) patients below 40 years of age. Mutation exchanges during follow-up in 32 cases were compared with 65 controls with paired serum samples. In addition, a consecutive series of samples from 14 HCC cases were employed to compare the sequences before and after the occurrence of HCC.

Results: After adjustment for age, history of cigarette smoking and alcohol consumption, HBeAg positive, HBV DNA levels ≥4.00 log10 copies/mL, pre-S deletion, T1762/A1764 double mutations, and T1766 and/or A1768 mutations were associated with risk of young age HCC. Moreover, the presence of an increasing number of HBV-related mutations (pre-S deletion, T1762/A1764, and T1766 and/or A1768 mutations) was associated with an increased risk of young age HCC. Paired samples analysis indicated that the increased HCC risk for at-risk sequence mutations were attributable to the persistence of these mutations, but not a single time point mutation. The longitudinal observation demonstrated a gradual combination of pre-S deletion, T1762/A1764 double mutations, and T1766 and/or A1768 mutations during the development of HCC.

Conclusion: High HBV DNA levels and pre-S deletion were independent risk factors of young age HCC. Combination of pre-S deletion and core promoter mutations increased the risk and persistence of at-risk sequence mutations is critical for HCC development.

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Introduction

Hepatocellular carcinoma (HCC) is the fifth most common cancer and the third most common cause of cancer-related death in the world [1]. Etiologically, majority of HCC develops in chronic hepatitis B virus (HBV) carriers, especially in East Asia and sub-Saharan Africa, where HBV is endemic. Previous studies have shown that chronic HBV infection was associated with the development of HCC in 60% of patients [2]. It is generally accepted that HBV played a major causative role in the development of HCC in humans [3]. Identification of risk factors for HCC and stratification of patient risk are very important to guide future surveillance strategy. The current recommendations most frequently applied for screening patients with HCC are published by the American Association for the Study of Liver Diseases (AASLD) [4]. The recommendations advise HCC screening Asian male HBV patients elder than 40 years and Asian female HBV patients elder than 50 years. Based on this guideline, young patients (under the age of 40 years) could be excluded from cancer screening programs. However, recent studies have reported a significant prevalence and worse prognosis in young HCC patients [5–7]. The cost-effectiveness for screening all the HBV carriers below 40 years of age need to be proved. The alternative strategy was to screen the high-risk subjects in this particular age group of HBV carriers.

The pathogenesis of HCC in HBV infection has been extensively investigated, and various viral risk factors have been identified. Recently, high viral load, pre-S deletion, T1653 mutation in enhancer II (EnhII), V1753 mutation, and T1762/ A1764 double mutations in basal core promoter (BCP) have been found to be associated with the development of HCC in several reports [8–14]. However, to the best of our knowledge, there was no study primarily focused on HBV mutations in young HCC patients. It has been postulated that there may be different
mechanisms of hepato-carcinogenesis according to the age distribution of patients [15]. The data are largely lacking in this group of patients.

The township of Qidong is one of the highest endemic regions for chronic HBV infection and HCC in China. This case-control study was conducted within a large cohort of male HBV carriers in Qidong. The goal of the present study was to assess the risk of specific complex mutation patterns with other viral factors in the development of young HCC (under the age of 40 years).

Methods

Study Population

The analysis used data and stored samples from a prospective cohort in Qidong, Jiangsu Province, China [16]. From August 1 to September 30, 1996, a total of 18,000 males aged 20 to 65 years, who were living in 17 towns of Qidong, were invited to participate in this HCC screening study. The health examination at study entry included abdominal ultrasonography (US) and serological tests for hepatitis B surface antigen (HBsAg), hepatitis B e antigen (HBeAg), anti-hepatitis C virus (HCV), serum levels of alanine aminotransferase (ALT), and serum alpha-fetoprotein (AFP). A total of 2387 males who were seropositive for HBsAg and free of HCC at recruitment were followed up until October 2006. US, conventional liver function, and AFP levels were tested every 6–12 months. Patients underwent intensive surveillance with computed tomography (CT), magnetic resonance imaging (MRI), and/or hepatic angiography if there was any suspicious abnormality on US or the AFP level was greater than 20 ng/mL. At recruitment, each study participant provided informed consent and a structured questionnaire on sociodemographic characteristics, habits of alcohol and tobacco consumption. Serum samples collected at interview were stored at −70°C before analysis. This study was approved by the research ethics committee at Zhongshan Hospital, Fudan University, Shanghai, China.

Cases and Controls

The data of HCC were obtained from medical records and searches of computer files of death certification and cancer registry systems. To ensure complete ascertainment, we also contacted relatives to identify cases. As a prospective study, in order to eliminate the possible influence of undiagnosed HCC at recruitment, we excluded the HCC cases that were diagnosed within the first two years of follow-up. Our analysis was restricted to HCC diagnosed from October 1, 1998, to September 30, 2006. During this follow-up period, we confirmed 199 incident HCC patients. A total of 73 patients diagnosed with HCC within the first two years of follow-up were excluded from the analysis, 19 were younger than 40 years. The diagnosis of HCC was based on the following criteria: a histopathological examination; 1 imaging technique and a serum AFP level ≥400 ng/mL; or a positive lesion detected by at least 2 different imaging techniques (US, CT, MRI, and hepatic angiography). Several cases qualified based on more than 1 criterion. Of the 199 patients with HCC, 57 were younger than 40 years (young HCC). For each case, we randomly selected two chronic hepatitis (CH) patients as controls from the cohort of HBsAg carriers who were alive and had not been diagnosed with HCC throughout the follow-up period. The controls were individually matched to the cases by age (within 2 years). A total of 114 controls were recruited. Subjects were excluded if they had poor sequence data (3 cases and 5 controls) or polymerase chain reaction (PCR) failure resulted from low quantity of DNA (below about 500 copies/ml; the detection limit of our nested PCR assay) (4 cases and 14 controls) or a history of antiviral therapy (1 cases and 5 controls). Consequently, a total of 49 cases and 90 controls were included in the analysis.

Serology

Serum HBsAg, HBeAg and anti-HCV antibody were tested by commercially available enzyme immunoassay kits (Shanghai Kehua Bio-engineering Co., Ltd., China). Serum ALT level was determined by ultraviolet-lactate dehydrogenase (UV-LDH) method (Shanghai Kehua Bio-engineering Co. Ltd.). The serum HBV DNA levels were determined using the Fluorescein quantitative polymerase chain reaction (FQ-PCR) detection system (Taqmen; Roche US), according to the manufacturer’s instructions. The lower limit of detection was 500 copies/mL.

Nested Polymerase Chain Reaction and Direct Sequencing of the EnvII/BCP/PC and pre-S Regions

HBV DNA was extracted from 200 μL serum samples using the commercial Kit (Shanghai Shenyou Biotech Company, China). HBV genes of the EnhII/BCP/PC regions were amplified by nested PCR. First-round PCR primers were 5’-CAGCTTGTTTTGTCGGCAGC-3’ (nt 1286-1305) and 5’-GAGTACTC-GACAGAAGCTCC-3’ (nt 2083-2063). PCR reaction was carried out in 50 μL containing 5 μL 10 × buffer, 4 μL 2.5 mmol/L deoxynucleoside triphosphates (dNTP), 2 μL 10 μmol/L sense and antisense primers, 1.5 U PlatinumTaq DNA polymerase (Invitrogen, shanghai, China). First-round PCR was performed as follows: 95°C for 2 min; 95°C for 30 sec, 56°C for 30 sec, and 68°C for 3 min for 35 cycles; and finally, 68°C for 10 min. 2 μL of the first-round PCR product was reamplified by the same PCR condition as the first-round reaction. Second-round PCR primers were 5’-GAGAAGCTCCAACGAGTCC-3’ (nt 1941-1922) and 5’-CCCAAGAAGCTCGGAATTC-3’ (nt 2747-2766). For pre-S region sequence analysis, pre-S genes were amplified under the same PCR condition described above except the primers were used. First-round PCR primers were 5’-AAAATTAATTATGTCTGCTAGG-3’ (nt 2627-2647) and 5’-GAGAAGCTCCAACGAGTCC-3’ (nt 269-251). Second-round PCR primers were 5’-TTTACACACGAGTCC-3’ (nt 2174-2116) and 5’-GAGTACTAGACTCTGTGGTATTG-3’ (nt 253-232). All necessary precautions to prevent cross-contamination were taken, and negative controls were included in each assay. Amplified products were directly sequenced in both the forward and reverse directions using an ABI 3700 sequencer and commercial kit (Applied Biosystems, Foster City, CA).

HBV Genotyping

HBV genotypes were determined by comparing the sequence of EnhII/BCP/PC and pre-S regions with a set of standard sequences obtained from GenBank. Phylogenetic tree was constructed by software MEGA version 3.1.

Statistical Analysis

Data are presented as means ± SD, proportions, or median (range). To compare the values between the two groups, χ² or Fisher exact tests were performed for categorical variables and Mann-Whitney U tests were used for continuous variables with skewed distributions, respectively. Binary unconditional logistic regression models were used to estimate the odds ratios (ORs) of HCC associated with HBV-related factors and corresponding 95% confidence intervals (CIs). Potential confounders including age, history of cigarette smoking and alcohol consumption were adjusted. Multivariate analyses with stepwise logistic regression
were used to determine the independent factors associated with HCC. All statistical tests were two sided. \( P < 0.05 \) was considered statistically significant. All statistical analyses were performed using SPSS 11.5 for Windows (SPSS Inc., Chicago, IL).

**Results**

Clinical Features and Virologic Characteristics of Young HCC Patients and Controls

The clinical features and virologic characteristics for patients with and patients without HCC were presented in Table 1. There were no statistically significant differences in age, the histories of cigarette smoking and alcohol consumption between HCC patients and controls.

After adjustment for age, history of cigarette smoking and alcohol consumption, the OR for ALT elevation (>45 U/L) was 1.567 (95% CI, 0.769–3.194); seropositivity for HBeAg, 2.326 (95% CI, 1.092–4.955). Compared with participants having serum HBV DNA levels of less than 4.00 log\(_{10}\) copies/mL, the adjusted OR was 3.107 (95% CI, 1.035–9.328) for participants with serum HBV DNA levels of 4.00–5.99 log\(_{10}\) copies/mL; 6.040 (95% CI, 1.759–20.743), 6.00 log\(_{10}\) copies/mL or greater. A significant biological gradient of HCC risk by HBV DNA level from less than 4.00 log\(_{10}\) copies/ml to 6.00 log\(_{10}\) copies/mL or greater was observed (Table 1).

Genotype C dominated the HBV types in Qidong. HCC patients and control subjects showed the similar distribution pattern for genotype. When we examined HBV DNA sequences in the pre-S and EnhII/BCP/PC regions, pre-S deletion, T1762/A1764, and T1766 and/or A1768 mutations were significantly associated with HCC, showing adjusted ORs from 2.295 to 3.167 (Table 1). The most frequently occurring mutation was T1762/A1764 double mutations. T1653, V1753, A1896, and A1899 mutations were not associated with a higher risk for developing young HCC. Among 139 patients, 25 were infected with pre-S deletion mutation. Compared to control patients, patients with HCC had significant higher frequencies of pre-S deletions. Among the 14 pre-S deletion mutations in the HCC group, three occurred in pre-S1, nine in the 5' half of the pre-S2 region and four cases had mutations that removed the pre-S2 initiation codon and adjacent sequences. Two of 14 had two deletions (one in pre-S1 and another in the 5’ half of the pre-S2 region). In contrast, of the HBV deletion mutations in the control groups, three occurred in the pre-S1 region, five in the pre-S2 region and three had mutations that removed the pre-S2 initiation codon and adjacent sequences. Compared to patients without pre-S deletion, patients with pre-S deletion had higher proportions of T1766 and/or A1768 mutations \([11/25 (44.0\%) \text{ vs. } 9/114 (7.9\%)\), \(P<0.001\].

Multivariate Analysis on the Risk Factors for Young HCC

Unconditional logistic regression analyses showed that HBeAg positive, high viral load (≥4 log\(_{10}\) copies/mL), and three sequence mutations (listed in Table 1) were significantly associated with the subsequent risk of young HCC. On further calculation using stepwise logistic regression analysis, the followings were found to be independent risk factors of young HCC: high HBV DNA levels and presence of pre-S deletion (Table 2).

Association between Young HCC Risk and the Presence of Specific Mutation Patterns

A statistical analysis of the 3 at-risk mutation combinations (pre-S deletion, T1762/A1764 mutations, and T1766 and/or A1768 mutations) was performed in the analysis of the combined risk for HCC. Our data showed that any 2 or 3 mutation combinations...
Discussion

Chronic HBV infection is the main cause of HCC, especially in the Chinese population. The average age at onset of HBV associated HCC is 50 years of age, approximately 10 years earlier than that of onset of HCV-associated HCC [17,18]. The target population for HCC screening is usually limited to the elderly and the younger age group is therefore neglected in the screening program. However, the incidence of HCC in patients younger than 40 years, especially in high risk populations, is relatively high [19,20]. Only a small number of articles have so far been written about the epidemiologic, clinical, and histopathologic features of HCC in patients younger than aged 40 years. The relationship between HBV mutation and the development of HCC in this group was rarely demonstrated. The current study may help to clarify the HCC risk factors in young age HBV carriers.

In this case control study from a cohort of HBV carriers, a significantly positive association between high viral load, HBeAg positive, specific sequence mutation and HCC was observed in young males after adjusting for age, history of cigarette smoking, and history of alcohol consumption. Multivariate analysis demonstrated that high HBV DNA levels and presence of pre-S deletion were independent factors associated with the development of HCC in young age HBV carriers. Recently, the role of HBV DNA levels in predicting the progression to HCC has been reported in many studies from mainland of China, Hong Kong, Taiwan, and Japan [8,9,21]. Similar to the REVEAL study [8], we found that HBV DNA levels $\geq 4.00 \log_{10}$ copies/mL started to have increased risk of HCC and HBV DNA levels $\geq 6.00 \log_{10}$ copies/mL had a further incremental HCC risk. Consistent with previous studies [13,22], our study has shown the relationship between the presence of pre-S deletion and risk of HCC. The mutations in the BCP region have been widely studied. The relationship between T1762/A1764 double mutations and risk of HCC has been demonstrated in two large cohort studies [10,23]. Recently, T1766/A1768 mutations have been reported to be associated with the development of HCC [16,24]. The magnitude of the ORs of HCC associated with the presence of the BCP mutations is generally 2- to 3-fold after adjusting for age, history of cigarette smoking and alcohol consumption. Most previous studies were significantly associated with the development of HCC. Compared to patients with wild-type HBV, patients with a single at-risk mutation (OR 2.834; 95% CI, 1.160–6.920), 2 mutation combinations (OR, 4.024; 95% CI, 1.187–13.646), 3 mutation combinations (OR, 8.938; 95% CI, 1.762–45.338) and had a higher risk of young HCC. Table 3 demonstrated a significant biological gradient of HCC risk by number of at-risk mutations.

Persistence of Sequence Mutation and Young HCC Risk

To analyze association between persistence of a HBV mutation and HCC risk, serum samples at recruitment were also retrieved for sequence analysis. Of all included 139 cases, 32 HCC patients and 65 controls had adequate serum samples collected at recruitment for sequence analysis. The median time interval between the dates of the recruitment samples and the dates of the follow-up samples was 5.7 years (range, 2 to 7.8 years) for cases and 5.2 years (range, 2.5 to 7.5 years) for controls. For at-risk sequence mutations identified to be associated with HCC, detection of a high-risk mutation at both time points was significantly associated with an increased risk of HCC after adjusting for age, history of cigarette smoking, and history of alcohol consumption, while no association with HCC was observed at paper that paper was due to the negative PCR product. There

Table 2. Multivariate analysis of risk factors for the development of HCC.

| Factors                  | Odds ratio (95% CI) | P-value |
|--------------------------|---------------------|---------|
| Serum HBV DNA levels     | 1.925 (1.322–2.803) | 0.001   |
| Presence of pre-S deletion | 3.979 (1.459–10.850) | 0.007  |

Table 3. Association between HCC and the presence of specific mutation patterns of pre-S deletion, T1762/A1764 double mutations, and T1766 and/or A1768 mutations.

| Number of mutation | HCC patients | Controls | Adjusted odds ratio* (95% CI) | P-value |
|--------------------|--------------|----------|-------------------------------|---------|
| No mutation        | 9 (18.4)     | 39 (43.3) | 0.297 (0.128–0.691)           | 0.005   |
| 1                  | 25 (51.0)    | 39 (43.3) | 2.834 (1.160–6.920)           | 0.022   |
| 2                  | 9 (18.4)     | 9 (10.0)  | 4.024 (1.187–13.646)          | 0.025   |
| 3                  | 6 (12.3)     | 3 (3.3)   | 8.938 (1.762–45.338)          | 0.008   |

*Adjusted for age, history of cigarette smoking, history of alcohol consumption.

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Table 4. Adjusted ORs for HCC associated with persistence of sequence mutations of HBV.

| Variable                  | HCC patients | Controls | Adjusted odds ratio* | P-value |
|---------------------------|--------------|----------|----------------------|---------|
|                           | at baseline  | at follow-up | n = 32 (%) | n = 65 (%) | (95% CI) |         |
| Pre-S deletion            | -- / --      | 21 (65.6) | 57 (87.7)           | 1.00 (reference) |         |
|                           | -- / -- or t / -- | 3 (9.4) | 3 (4.6)           | 2.673 (0.477–14.983) | 0.254 |
|                           | + / +        | 8 (25.0) | 5 (7.7)           | 4.392 (1.277–15.106) | 0.019 |
| T1653                     | -- / --      | 20 (62.5) | 53 (81.5)           | 1.00 (reference) |         |
|                           | -- / -- or t / -- | 3 (9.4) | 5 (7.7)           | 1.472 (0.309–7.003) | 0.627 |
|                           | + / +        | 9 (28.1) | 7 (10.8)           | 3.356 (1.089–10334) | 0.035 |
| V1753                     | -- / --      | 21 (65.6) | 46 (70.8)           | 1.00 (reference) |         |
|                           | -- / -- or t / -- | 4 (12.5) | 7 (10.8)           | 1.198 (0.293–4.900) | 0.802 |
|                           | + / +        | 7 (21.9) | 12 (18.5)           | 1.294 (0.435–3.853) | 0.643 |
| T1762/A1764               | -- / --      | 7 (21.9) | 29 (44.6)           | 1.00 (reference) |         |
|                           | -- / -- or t / -- | 6 (18.8) | 10 (15.4)           | 2.415 (0.636–90168) | 0.195 |
|                           | + / +        | 19 (59.4) | 26 (40.0)           | 3.203 (1.129–9.088) | 0.029 |
| T1766 and/or A1768        | -- / --      | 23 (71.9) | 59 (90.8)           | 1.00 (reference) |         |
|                           | -- / -- or t / -- | 3 (9.4) | 3 (4.6)           | 2.648 (0.469–14.954) | 0.270 |
|                           | + / +        | 6 (18.8) | 3 (4.6)           | 5.297 (2.110–23.188) | 0.027 |
| A1896                     | -- / --      | 11 (34.4) | 27 (41.5)           | 1.00 (reference) |         |
|                           | -- / -- or t / -- | 8 (25.0) | 11 (16.9)           | 1.506 (0.468–4.850) | 0.492 |
|                           | + / +        | 13 (40.6) | 25 (38.5)           | 1.378 (0.501–3.788) | 0.535 |
| A1899                     | -- / --      | 28 (87.5) | 60 (92.3)           | 1.00 (reference) |         |
|                           | -- / -- or t / -- | 2 (6.2) | 2 (3.1)           | 2.097 (0.272–16.149) | 0.477 |
|                           | + / +        | 2 (6.2) | 3 (4.6)           | 1.517 (0.225–10.211) | 0.668 |

A total of 32 cases and 65 controls who had a baseline blood sample and a blood sample collected at follow-up were included in analysis.

* = absence; + = presence.

Adjusted for age at recruitment, history of cigarette smoking, history of alcohol consumption.

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Table 5. Longitudinal observation of specific mutation patterns of pre-S deletion, T1762/A1764, and T1766 and/or A1768 mutations in 14 HCC patients.

| Sample | At baseline | 2–4 years before HCC | HCC |
|--------|-------------|-----------------------|-----|
| #1     | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #2     | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #3     | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #4     | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #5     | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #6     | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #7     | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #8     | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #9     | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #10    | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #11    | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #12    | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #13    | ○ . . . . . | ○ . . . . . | ○ . . . . . |
| #14    | ○ . . . . . | ○ . . . . . | ○ . . . . . |

○ = wild-type; < = pre-S deletion, T1762/A1764 double mutations, or T1766 and/or A1768 mutations.

Most previous studies on the relationship of HBV mutation and the risk of HCC were conducted with the use of samples taken after the diagnosis of cancer. In this investigation, we also examined the HBV mutations in serum samples at recruitment and after HCC diagnosis. The association between the presence of the HBV mutations and HCC risk could be evaluated from samples collected at recruitment and the diagnosis of HCC in combination. Base exchanges in nucleotide of HBV during follow-up appeared in about 6 to 25% of the subjects. We also found that detection of HBV mutations in both the baseline and subsequent
samples at diagnosis of HCC was associated with substantially higher risk than detection of this mutation at a single time point. Thus, the increased HCC risk for carriers of HBV strains harboring the mutations is most likely a result of persistence of such mutations. It is biologically reasonable that pre-S deletion and BCP mutations could contribute to the process of HCC. The HBV envelope is composed of 3 forms of HBV surface antigen: large (coded for by the pre-S1/pre-S2/S gene), middle (the pre-S2/S gene), and small (the S gene) protein. The pre-S regions play an essential role in the interaction with the immune responses because they contain several epitopes for T or B cells [25,26]. In persistent HBV infection, immune epitope deletion mutants occur, escape the host immune surveillance, and lose important functional sites. The deletion over the pre-S gene may affect the expression of middle and small surface proteins, resulting in intracellular accumulation of large surface protein and viral particles, formation of ground glass hepatocytes. These deletion mutations accumulate in the endoplasmic reticulum and cause endoplasmic reticulum stress signals. Through endoplasmic reticulum stress signaling pathways, the pre-S mutant large HBV surface antigens can induce oxidative stress and lead to oxidative DNA damage of HBV infected hepatocytes. Presence of the oxidative DNA lesions stimulates DNA repair activity; the induced mutagenesis occurs in the genome [27,28]. It has been proposed that BCP mutations may diminish the production of HBeAg and increase viral replication, which theoretically results in increased host immune responses against the virus, therefore increasing hepatocyte apoptosis and degeneration, which leads to liver injury [29,30]. In addition, this mutation in BCP may alter the binding ability of trans-regulating nuclear factors (such as CCAAT/enhancer-binding protein-α, the ubiquitous transcription factor Sp1, and hepatocyte nuclear factor 4) and may also lead to amino acid alterations of X protein, affect the function of the X protein, interfere with cell growth control and DNA repair and may contribute to the process of multiple steps in hepatocarcinogenesis [31,32].

The strengths of this study include the sequence analysis of a series of serum samples from a community-based cohort study, repeated sequence analysis provides data on the long-term stability of viral sequence and helps clarify the temporal relationship between a sequence mutation and the occurrence of HCC. Furthermore, this association between viral factors and HCC is unlikely to be biased by the effect of antiviral therapy because the proportion of participants in this cohort who received such therapy was very low (<1%) and no participants with a history of such therapy were included in the analysis. There are also several limitations in this study. First, although CH patients in the control group were age matched with those in the HCC group, the possibility of developing malignancy in the future cannot be denied. Second, the generalizability of the results is limited because all the study subjects were males, a larger cohort and a longer follow-up time are needed for a similar study in females.

In conclusion, this current study further supports the view that Chronic HBV carriers below 40 years of age should not be neglected and should be included surveillance programs for HCC, especially those high risk subjects infected with HBV of complex sequence mutations. These HBV mutations may serve as useful biomarkers for predicting the clinical outcomes of young patients with chronic hepatitis B. Modifications of regular HCC screening guidelines could be expected to result in earlier disease detection and improved prognosis in young patients.

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Author Contributions

Conceived and designed the experiments: XS LQ. Performed the experiments: LQ TL. Analyzed the data: XK. Contributed reagents/materials/analysis tools: TC ZN. Wrote the paper: LQ.

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