Research paper

Morphological and genomic evidence for a new species of Corallorhiza (Orchidaceae: Epidendroideae) from SW China

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A R T I C L E  I N F O
Article history:
Received 11 November 2020
Received in revised form 2 January 2021
Accepted 7 January 2021
Available online 23 January 2021

Keywords:
Corallorhiza sinensis
Oreorchis
Sichuan province
Plastome
Phylogeny

A B S T R A C T
Corallorhiza sinensis, a new species of mycoheterotrophic orchid from western Sichuan, China, is described and illustrated based on molecular and morphological evidence. It is morphologically similar to Corallorhiza trifida, but can be distinguished by bigger flowers, both sepals and petals with 3 veins, and longer lateral lobes of lip. To distinguish the new Corallorhiza species and explore its phylogenetic position within subtribe Calypsoinae, this study employed sequences of the nuclear ribosomal DNA (nrDNA) and whole plastome assembled from the genome skimming approach. The plastome is 148,124 bp in length, including a pair of inverted repeats (IRs) of 26,165 bp, a large single-copy region (LSC) of 82,207 bp, and a small single-copy region (SSC) of 13,587 bp. Further, phylogenetic analyses were performed using nrDNA sequence and 79 coding sequences (CDSs) from 26 complete plastomes of subtribe Calypsoinae. The phylogenetic tree based on nrDNA sequence suggested that Corallorhiza is a monophyletic group, and strongly support C. sinensis as sister to the rest species of Corallorhiza. The plastid tree showed that 10 Corallorhiza species grouped into two clades and C. sinensis is most closely related to the North American C. striata and C. bentfeyi instead of Oreorchis foliosa and O. angustata in the same clade. The plastid tree and nrDNA tree indicate Oreorchis is a paraphyletic. Although the topological conflicts are displayed between plastome and nrDNA phylogenies of C. sinensis, it is still the most closely related to Corallorhiza. Comparative analysis showed that C. sinensis populations are characteristic of the intermediate morphological traits between Corallorhiza and Oreorchis. The finding of this new species from China shed new light on the phylogeny of Oreorchis and Corallorhiza.

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1. Introduction

Orchidaceae is one of the largest families of flowering plants, comprising about 736 known genera and 28,000 species worldwide (Chase et al., 2015; Christenhusz and Byng, 2016). During an investigation on wild orchid resources in western Sichuan, China in 2020, a distinctive orchid species was discovered in the Siguniang Mountains National Nature Reserve under shady Populus forests. Some morphological characters such as lack of leaf and coralloid rhizome immediately caught our attention, and it is morphologically similar to the species in the genus Corallorhiza. Corallorhiza Gagnebin is the largest temperate genus of leafless putative heterotrophs in Orchidaceae, comprising 11 species that are restricted to the northern hemisphere of the New World, except for a single circumboreal species, C. trifida Chatelain (Chase et al.,...
According to the *Flora of China*, only *C. trifida* has been recorded in China, distributed in Gansu, Guizhou, Hebei, Jilin, Nei Mongol, Qinghai, Sichuan, Xinjiang (Chen and Stephan, 2009). We compared the new collection with *C. trifida*, and discovered that it can be distinguished from the latter by bigger flowers, both sepals and petals with 3 veins, and longer lip lateral lobes. We further compared it with other congeneric species. It is also similar to *S. striata* Lindley and *C. bentleyei* Freudenstein in sharing a distinctive morphological character: lack a mentum (i.e. a vestigial nectar spur at the base of the perianth), but can be easily distinguished from these two species by its lip broad oblong and obviously 3-lobed the toward base. After carefully comparing the plant with all previously known congeneric taxa, it could not be matched with any known species. Hence, our collection might be a new species of *Corallorhiza*.

Chase et al. (2015) presented a revised classification of Orchidaceae and divided the family into five subfamilies, namely Apostasioideae, Vanillioideae, Cyropipedioideae, Orchidoideae, and Epidendroideae. The genus *Corallorhiza* was placed in the subtribe Calypsoideae (Epipendroideae, Epidendroideae) which contains 13 genera, namely *Corallorhiza*, *Aplectrum*, *Calypso* Salisb., *Chargniensis* S.S. Chien, *Kitigorchis* Lindl., *Lindleya* Pohl, *Rchb. f.*, *D. danxiaoricris* J.W. Zhai, F.W. Xing & Z.J. Liu, *Epiphippanthus* Rchb. f., *Govaenia* Lindl., *Oreorchis* Lindl., *Tippulatoria* Nutt., and *Yoenia* Maxim. Freudenstein et al. (2017) reconstructed phylogenetic relationships among subtribe Calypsoideae. Based on matK marker, results showed *Corallorhiza* was paraphyletic, but the support for the node was relatively weak. In the results of ITS sequence and combined analyses (matK + ITS sequences), *Corallorhiza* was shown to be monophyletic with strong support. An earlier study based upon combined ITS, plastid datasets, and morphological character matrix strongly support *Oreorchis and Corallorhiza* are both monophyletic groups, but the *Corallorhiza* is far from *Oreorchis* (Zhao et al., 2013). In recent molecular phylogenetic studies based on the plastomes, however, *Corallorhiza* is still non-monophylectic, and the *Oreorchis* and *Corallorhiza* were nested together and formed a single clade (Kim et al., 2020; Li et al., 2020). In the taxonomic revision of *Oreorchis* (Pearce and Cribb, 1997), except for *Kitigorchis itoana*, the remaining species of *Kitigorchis* were transferred into *Oreorchis*. Moreover, Pearce and Cribb (1997) remarked that *Kitigorchis* be a monotypic genus intermediate between the *Oreorchis* and *Corallorhiza*, and suggested that *K. itoana* has coral-like rhizomes quite different from the subterranean organs of *Oreorchis*. Interestingly, the final study revealed that *K. itoana* and *O. indica* were conspecific (Yukawa et al., 2003). In fact, *O. indica* often possesses coralloid rhizomes (Suetsumu et al., 2020). An earlier study has shown that coralloid rhizomes occur in the *Aplectrum hyemale* (Muhl. ex Willd.) Nutt. (MacDougal and Dufrenoy, 1944). Thus the habit has happened many times in Calypsoideae. Similarly, in the Calypsoideae, a previous study also demonstrated that leafless habit has happened many times (Freudenstein et al., 2017). Suetsumu et al. (2020) suggested that *O. indica* is a partially mycoheterotrophic orchid, and the full mycoheterotrophy evolved after the establishment of partial mycoheterotrophy, rather than through direct shifts from autotrophy in the *Oreorchis-Corallorhiza* clade. In the genus *Corallorhiza*, it is well known that *C. trifida* is partial mycoheterotrophy (Zimmer et al., 2008). Due to the phylogenetic relationship of *Oreorchis* and *Corallorhiza* remaining uncertain, we hypothesize that our collection might be a new species of *Corallorhiza*. Here we describe and illustrate the new species *Corallorhiza sinensis*. To distinguish the new *Corallorhiza* species, we sequenced its complete plastome. Its plastome was analyzed in comparison with other members of subtribe Calypsoideae. Finally, we reconstructed its phylogenetic relationship within subtribe Calypsoideae based upon the nrDNA sequence and 79 coding sequences (CDSs).

2. Materials and methods

2.1. Morphological analyses

Morphological characters of the new species were based on the study of living plants and herbarium specimens. Measurements of flowers and rhizomes were taken from living plants. Photographs of *Corallorhiza sinensis* and *C. trifida* were taken in the field and are included in Figs. 1 and 2, respectively. A total of 11 diagnostic characteristics of these two species are compared in Table 1. Measurements of *C. trifida* were based on the combination of *Flora of China* (Chen and Stephan, 2009) and observation on living plants. Given that the phylogenetic relationship of *Oreorchis* and *Corallorhiza* remains uncertain, we compared the new collection with the members of *Oreorchis* as well and found the new species resembles *O. patens* and *O. foliosa*, var. *indica*. To better distinguish the new species, we also presented its supplementary photograph (Fig. S1). Furthermore, the photographs of *O. patens* and *O. foliosa*, var. *indica* were combined in the Fig. S2, which will provide important morphological evidence for this study. The voucher specimens of the new species were deposited in the Herbarium of Wuhan Botanical Garden, Chinese Academy of Sciences (HIB).

2.2. Specimens examined of *Oreorchis*

*Oreorchis patens* Lindl., China: Hunan, Sangzhi county, July-4-1998, *Beijingdul* 2362 (PE, O1340734).

*Oreorchis foliosa*, var. *indica* (Lindl.) N. Pearce & P.J. Cribb, China: Xizang, June-2-1972, *Xizang Herbal Medicine Investigation Team* 89 (PE, 00850746).

2.3. DNA extraction and sequencing

Purified total DNA of *Corallorhiza sinensis* was fragmented to construct short-insert libraries for sequencing (Illumina HiSeq, 2000 platform from the Biomarker Technologies, Inc.), and 6 GB of reads was obtained.

2.4. Plastome and nrDNA assembly, annotation

The complete plastome of *Corallorhiza sinensis* was assembled using GetOrganelle v.1.7.1 with appropriate parameters: K-mer is “21,45,65,85,105”, the Word size is 0.6 (Jin et al., 2020). Five key steps were included in the workflow: 1. Mapping reads to seed and assembling seed-mapped reads for parameter estimation; 2. Recruiting more target-associated reads through extending iterations; 3. Conducting de novo assembly; 4. Roughly filtering for target-like contigs; 5. Identifying target contigs and exporting all configurations (Bankevich et al., 2012; Camacho et al., 2009; Jin et al., 2020; Langmead and Saltberg, 2012). Final assembly graphs were visualized by Bandage to assess the completeness of the final graph (Wick et al., 2015). The nrDNA sequence (18S–ITS1–5.8S–ITS2–26S) of *C. sinensis* was assembled using GetOrganelle v.1.7.1 with default parameters.

Initially, PCA (Qu et al., 2019) was utilized to annotate the complete plastome of *Corallorhiza sinensis*, with the *C. trifida* (GenBank: MN998435) selected as the reference, and then manually adjusted protein-coding genes and exons boundaries using software Geneious 11.0.4 (Kease et al., 2012). The final plastome and nrDNA sequence of *C. sinensis* have been submitted to GenBank under accession numbers MW191791 and MW413395, respectively.
Gene map of the plastome of *C. sinensis* was drawn online using OGDRAW (Lohse et al., 2007).

### 2.5. Genome features and comparative plastid genomic analysis

Length of the whole plastome of *Corallorhiza sinensis*, numbers of genes, and categories of Genes were analyzed in Geneious 11.0.4 (Kearse et al., 2012). The software MEGA (Kumar et al., 2016) was used for calculating the guanine-cytosine (GC) content. In order to further identify potential variations between *C. sinensis* and other *Corallorhiza* species, the genome rearrangement analyses of ten *Corallorhiza* sequences were performed using Mauve (Darling et al., 2004) plugin in Geneious 11.0.4 (Kearse et al., 2012). Geneious 11.0.4 (Kearse et al., 2012) was used to detect the contraction/

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**Fig. 1.** Photographs of *Corallorhiza sinensis*. (A) habit; (B) individual; (C) root; (D) front view of flowers; (E) lateral view of flowers; (F) anatomy of flowers (a – dorsal sepal, b – petal, c – lateral sepal, d – lip); (G) lateral and front view of lips.
expansion of the inverted repeat regions (IRs), and the graph of expansions/contractions was visualized using Adobe Illustrator.

2.6. Phylogenetic analysis

To estimate the evolutionary history of *Corallorhiza trifida* within Calypsoinae, phylogenies were reconstructed by Maximum likelihood (ML) and Bayesian Inference (BI) analyses using the nrDNA sequence and coding sequences (CDSs), the total of 79 CDSs from 27 plastomes were used for constructing the phylogenetic relationship of the members within Calypsoinae (Table S1). Lost genes and pseudogenes were treated as missing data. All nrDNA sequences and plastomes were downloaded from the NCBI database except *C. sinensis*. In the plastid tree and nrDNA tree, *Cattleya crispata* and *Chysis bractescens* are selected as outgroup, respectively. We extracted 79 CDSs using Geneious 11.0.4 (Kearse et al., 2012). Under the codon model, each single CDS matrix was aligned and edited by MAFFT (Katoh and Standley, 2013) and Gblocks (Talavera and Castresana, 2007) plugin in PhyloSuite (Zhang et al., 2020), respectively, and each matrix is eventually concatenated into one supermatrix by PhyloSuite (Zhang et al., 2020). ModelFinder was utilized to select the best-fit model for

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**Fig. 2.** Photographs of *Corallorhiza trifida*. (A) individual; (B) habit; (C) root; (D) front view of flowers; (E) lateral view of flowers; (F) anatomy of flowers (a – dorsal sepal, b – petal, c – lateral sepal, d – lip); (G) lateral and front view of lips.
accurate phylogenetic estimates (Kalyaanamoorthy et al., 2017). ML analysis was performed using IQ-TREE with 1000 bootstrap replicates (Nguyen et al., 2015). BI analysis was performed in MrBayes 3.2.2 (Ronquist et al., 2012) using the Markov chain Monte Carlo (MCMC) method, with two independent runs for 10 million generations (Number of Chains is four). Phylogenetic trees were sampled every one thousand generations, the first 25% of trees generated were discarded as burn-in and the remaining trees were used to construct majority-rule consensus tree (Ronquist et al., 2012). Finally, the phylogenetic trees were edited using FigTree 1.4 (https://github.com/rambaut/figtree).

3. Results

3.1. Taxonomic treatment

3.1.1. Corallorhiza sinensis C.W. Hu & Q.F. Wang (Figs. 1 and 3)

Type: China. Sichuan: Xiaojin County, Siguniang Mountains National Nature Reserve, 102°44′31.5″E, 30°58′26.5″N, under Populus forests ca. 2984 m in elevation, 9 Jun 2020, S. Peng, J. X. Yang et J. J. Wang YJX-0060 (holotype: HIB, isotype: HIB).

Diagnosis: *Corallorhiza sinensis* resembles *C. trifida*, but prominently differs from the latter in having bigger flowers, both petals and sepals with 3 veins, and lateral lobes of lip. It is also similar to *C. striata* and *C. bentleyi* by sharing a distinctive morphological character: mentum absent, but can be easily distinguished by its lip broadly dilated toward apex, ca. 7 × 5 mm.

3.1.2. Description

Perennial mycoheterotrophic herbs, 10–46 cm tall. Rhizome fleshy, extensively branching, coralloid, nodiferous, internodes 2–4 mm long, often brown at the base of branch and beige at the apex of branch, diameter ca. 5 mm. Stems erect, cylindrical. Inflorescence light reddish brown, glabrous, leafless, covered with 3 or 4 membranous tubular sheaths from base to middle; sheaths amplexicaul, reddish brown, 2–12 cm long; rachis 5–18 cm, laxly 5–13-flowered; floral bracts small, lanceolate, apex acuminate, 1–3.5 mm long. Flowers light reddish brown, pedicel and ovary 9–12 mm long, torsion at base. Dorsal sepal arching forward, narrowly oblong-lanceolate, ca. 12 × 2 mm, 3-veined, apex acuminate; lateral sepals similar to dorsal sepal, slightly oblique, 3-veined, ca. 10 × 2 mm, connate at base, apex obtuse or acute. Petals long ovate-lanceolate, ca. 11 × 4 mm, 3-veined, apex obtuse or acute; lip broadly oblong, pure white with a yellowish claw, ca. 8 × 6 mm, obviously 3-lobed toward base; lateral lobes 2, erect, lanceolate but slightly curved, ca. 3 × 1 mm; middle lobe oblong to broadly dilated toward apex, ca. 7 × 5 mm, margin entire and slightly wavy at apex, with 2 distinct basal lamellae. Column yellowish, ca. 8 mm long, arcuate, basally with ridges or poorly developed auricles; ovary ca. 8.5 mm; mentum absent. Fruit not seen.

3.1.3. Etymology

The specific epithet “sinensis” refers to China, because the new species is endemic to China.

3.1.4. Distribution, habitat and conservation status

Currently, *Corallorhiza sinensis* is known only from Siguniang Mountains National Nature Reserve in Xiaojin County, Sichuan, China. The plants grow in shady *Populus* forests, flowering from May to June. Only two small populations of the new species were discovered, each population comprises approximately 12 mature individuals. Based on a previous study, *Corallorhiza* cannot be cultivated reliably because of the require of fungal associations for growth (Freudenstein, 1997). A preliminary assessment of the risk of extinction was conducted according to the International Union for Conservation of Nature (IUCN) Red List Categories and Criteria (https://www.iucn.org/), we thereby consider that *C. sinensis* should be classified as critically endangered (CR).

3.1.5. Additional specimens examined (paratypes)

China. Sichuan: Xiaojin County, Siguniang Mountains National Nature Reserve, grows Populus forests, elev. 2984 m, 102°48′21″E, 30°59′15.6″N, 8 Jun 2020, S. Peng, J. X. Yang et J. J. Wang P5-00166 (HIB).

3.2. Characteristics of the *Corallorhiza sinensis* plastome

The complete plastome of *Corallorhiza sinensis* is 148,124 bp in length, and exhibits a typical quadripartite structure, consisting of a large single copy (LSC) region of 82,207 bp and a small single copy (SSC) region of 82,207 bp, which were separated by a pair of 26,165 bp inverted repeat regions (IRs). The gene map of *C. sinensis* plastome is presented in Fig. 4. The gene composition in plastome of *C. sinensis* could be divided into four categories: genes related to photosynthesis, genes related to self-replication, protein-coding genes with unknown functions, and other genes. A total of 109
Fig. 3. Illustrations of Corallorhiza sinensis. (A) individual; (B) front view of flowers; (C) lateral view of flowers; (D) column and ovary; (E) anatomy of flowers; (F) anther cap; (G) pollinia. Drawn by Jing Tian.
unique genes were identified in the plastome, it contains 68 protein-coding genes, 30 tRNAs, and four rRNAs (Table 2). Among these genes, 15 genes are annotated as pseudogenes (ccsA, ndhA, ndhB, ndhC, ndhJ, ndhK, petA, petB, psaB, psaC, rbcL, rpoB, ycf1, rpl22). A total of 21 genes were duplicated in the IR regions, including eight tRNA genes (trnA-UGC, trnH-GUG, trnI-CAU, trnI-GAU, trnL-CAA, trnN-GUU, trnR-ACG, trnV-GAC), four rRNA genes (rrn4.5, rrn5, rrn16, rrn23), and nine PCGs (ndhB, ndhC, rpl2, rpl22, rpl23, rps7, rps19, rps12, ycf1). A total of seven protein-coding genes and six tRNA genes contain a single intron, only clpP and ycf3 contain two introns.

3.3. Comparative analysis of the plastomes

A comparison of the Corallorhiza and Oreorchis plastomes was conducted in the Table 3. The full-length plastome of the 10 Corallorhiza species ranged from 124,482 bp (C. bentleyi) to 151,031 bp (C. macrantha), and the C. bentleyi has the shortest plastome. The range of length variation among members of the Corallorhiza for the LSC and SSC region is obvious. Further, we also compared them with publicated Oreorchis plastomes (Table 3). The length of the plastome ranges from 158,256 bp in O. patens to 158,654 bp in O. angustata, obviously larger than Corallorhiza. In addition, the results indicated that Oreorchis plastome have longer LSC region than Corallorhiza. Comparative analysis of gene content showed that the number of PCGs in Oreorchis is very similar to most angiosperms, but it is very low in some Corallorhiza members. As our results displayed, GC content and the number of tRNA and rRNA genes are highly conserved in all these plastomes.

Expansion and contraction of the IRs were compared among Calypsoinae (Fig. S3). The LSC/IRb boundary information of Corallorhiza sinensis is similar to that of ten plastomes, including Changnienia amoena, Calypso bulbosa, Cremastra appendiculata, two Corallorhiza species (C. bentleyi and C. striata var. striata), three Oreorchis species (O. angustata, O. foliosa, and O. foliosa var. indica), and two Tipularia species (T. josephi and T. szchuanica). The gene rpl22 spanned the IRb/LSC region and the length of

Fig. 4. Circular gene map of the plastid genome of Corallorhiza sinensis. Genes drawn within the circle are transcribed clockwise, while those drawn outside are transcribed counterclockwise. Genes are color-coded according to their functional groups.
Table 2
Gene contents in the plastid genome of Corallorhiza sinensis.

| Category, Group of Genes | Gene Names |
|--------------------------|------------|
| Photosynthesis:          |            |
| Subunits of ATP synthase | atpB, atpA, atpE, atpF, atpA*, atpB*, atpF, atpB, atpH, atpA |
| Subunits of NADH dehydrogenase | ndhA, ndhB(x2), ndhC, ndhD(x2), ndhE, ndhF, ndhH |
| Cytochrome b/f complex    | petA, petB, petD, petC, petE, petF, petN |
| Subunits of photosystem I | psaA, psaB, psaC, psaA, psaB, psaF, psaA, psaB, psaC, psaF |
| Subunits of photosystem II | psbA, psbB, psbC, psbD, psbE, psbF, psbH, psbI, psbK, psbL, psbM, psbN, psbT, psbZ |
| Large subunit of ribulose | rbcL* |
| Other genes              |            |
| Subunit of Acetyl-CoA-carboxylase | accD |
| c-type cytochrome synthesis gene | ccaT* |
| Envelope membrane protein | cemA |
| Protease                 | cdpP** |
| Translational initiation | infA |
| Matrase                  | matK |
| Self-replication         |            |
| Large subunit of ribosome | rpl14, rpl16*, rpl22(x2), rpl20, rpl22b(x2), rpl32, rpl33, rpl36 |
| DNA dependent RNA polymerase | rpoA, rpoB, rpoC1*, rpoC2 |
| Small subunit of ribosome | rps2, rps3, rps4, rps5(x2), rps8, rps11, rps12*(x2), rps14, rps15, rps16*, rps18, rps19(x2) |
| tRNA Genes               | trnA-UGC, trnH-GUG(x2), trnK-UUU, trnL-UGU, trnL-CAA(x2), trnL-UGZ(x2), trnT-UGU, trnT-ACC(x2), trnU-UCU, trnV-GCU, trnZ-GAG, trnZ-UGA, trnZ-UGC |
| rRNA Genes               | trnL-UAG, trnM-CGU, trnM-UGU, trnM-CAL, trnM-UGA, trnR-ACG(x2), trnR-UCU, trnS-ACG, trnS-UGA, trnS-GCA, trnS-UCC, trnS-UGA, trnS-UCG, trnS-UGC, trnT-UCU, trnT-UCU, trnV-GAC(x2), trnV-UCG, trnW-CCA, trnY-GUA |

| Unknown function         |            |
| Conserved open reading frames | ycf1(x2), ycf2, ycf3**, ycf4 |

Note: * indicates the pseudogene; ** indicates trans-spliced gene; # indicates one of them is a pseudogene; * genes containing a single intron; ** genes containing two introns; (x2) genes present as two copies in the IR regions.

Table 3
Statistics on the basic features of the plastid genomes of species from Corallorhiza and Oreorchis.

| Species                          | Accession no. | Length (bp) | LSC (bp) | SSC (bp) | IR (bp) | GC content (%) | No. of PCGs | No. of tRNA | No. of rRNA |
|----------------------------------|---------------|-------------|----------|----------|---------|----------------|------------|-------------|-------------|
| Corallorhiza sinensis            |               |             |          |          |         |                |            |             |             |
| C. bulbosa                       | KM390013      | 148,643     | 82,851   | 12,368   | 27,712  | 37.1           | 74         | 38          | 8           |
| C. mertensiana                   | KM390018      | 147,941     | 81,109   | 13,774   | 26,529  | 36.8           | 60         | 38          | 8           |
| C. wisteriana                    | KM390020      | 146,437     | 82,590   | 11,743   | 26,172  | 37.1           | 73         | 38          | 8           |
| C. odontorhiza                   | KM390021      | 147,317     | 82,259   | 13,508   | 25,775  | 37.0           | 73         | 38          | 8           |
| C. trifida                       | MN990434      | 149,408     | 83,171   | 14,397   | 25,920  | 37.2           | 74         | 38          | 8           |
| C. macrantha                     | NC_025660     | 151,031     | 84,263   | 12,544   | 27,112  | 37.2           | 72         | 38          | 8           |
| C. striata var. striata          | NC_040978     | 141,202     | 75,881   | 12,999   | 26,161  | 36.3           | 50         | 37          | 8           |
| C. bentleyi                      | NC_040979     | 124,482     | 63,581   | 7,781    | 26,560  | 36.6           | 45         | 39          | 8           |
| C. maculata                      | NC_046814     | 146,198     | 79,612   | 13,018   | 26,784  | 36.8           | 55         | 38          | 8           |
| Oreorchis angustata              | MN990443      | 158,654     | 86,294   | 18,312   | 27,024  | 37.0           | 86         | 38          | 8           |
| O. patens                        | MN990436      | 158,256     | 86,253   | 18,341   | 26,831  | 37.0           | 86         | 38          | 8           |
| O. foliosa var. indica           | MN990440      | 158,599     | 86,061   | 18,616   | 27,437  | 36.9           | 82         | 38          | 8           |
| O. foliosae                      | MN990441      | 158,496     | 86,061   | 18,361   | 27,037  | 36.9           | 82         | 38          | 8           |

The result of the plastome structure comparison showed that there is one small inversion in Corallorhiza sinensis but none in other members of Corallorhiza and Oreorchis (Fig. S4). The inversion occurred in the LSC region between trnE-UCG and psbD genes, and was approximately 700 bp in length. Gene trnI-GGU was contained in the inversion region. We also detected an obvious inversion in plastome of C. maculata var. maculata and C. maculata. Furthermore, two inversions were detected in D. singhiana.

3.4 Phylogenetic analysis

Phylogenetic result of CDSs dataset showed Corallorhiza sinensis is sister to the clade comprised of C. striata var. vreelandii + C. striata var. striata + C. cf. striata + C. bentleyi + C. striata var. involuta with strong support in both Maximum likelihood and Bayesian Inference (ML: BS = 100; BI: PP = 1.00) (Fig. 5). In the plastid tree, Corallorhiza did not form
a monophyletic clade within Calypsoinae. Both ML and BI phylogenetic trees strongly indicated that the Oreorchis is nested within Corallorhiza (ML: BS = 100; BI: PP = 1.00). However, phylogenetic analysis based on nrDNA sequence suggest that Corallorhiza is a monophyletic group, and strongly support C. sinensis as sister to the rest species of Corallorhiza (ML: BS = 98; BI: PP = 1.00) (Fig. 6).

4. Discussion

The study is the first time to report new species of mycoheterotrophic orchid from western Sichuan: Corallorhiza sinensis. Previous studies indicated the members of Oreorchis possess coralloid rhizomes (Pearce and Cribb, 1997; Suetsugu et al., 2020), while the new species exhibits more abundant coralloid rhizomes than Oreorchis. C. sinensis present many synapomorphies that involved in Corallorhiza, such as leafless, no visible green tissue in all individuals. Despite the populations are characteristic of the intermediate morphological traits between Corallorhiza and Oreorchis, it most closely resembles the Corallorhiza. The complete plastome of C. sinensis was sequenced, assembled, and analyzed in comparison with these members of subtribe Calypsoinae. The plastome of C. sinensis shows a typical quadripartite structure, as reported for most angiosperms (Khayi et al., 2020; Wu et al., 2020; Zhu et al., 2020). A total of 109 unique genes were identified in the plastome, which contains 68 protein-coding genes, 30 tRNAs, and four rRNAs. C. sinensis further shows evidence of some photosynthesis-related genes either lost or pseudogenized, which is consistent with some previous studies of Corallorhiza (Barrett and Davis, 2012; Kim et al., 2020; Li et al., 2020). In the current study, a common feature of the plastome of Corallorhiza and Oreorchis species was that IR regions were more conserved than the LSC and SSC regions. C. sinensis is closely related to Corallorhiza in terms of the length of whole plastome. Comparative analysis of plastomes showed that C. sinensis has one small inversion occurred in the LSC region. However, the larger inversions appear in Danxiaorchis singchiana (Lee et al., 2020), C. maculata var. maculata (Barrett et al., 2014) and C. maculata (Kim et al., 2020), which had been reported. IR boundaries of C. sinensis present obvious expansion in these members of Calypsoinae. Phylogenetic analysis based on coding sequences (CDS) from 26 Calypsoinae members strongly supports Corallorhiza sinensis to be sister to the clade consisting of C. striata var. vreelandii + C. striata var. striata + C. cf. striata + C. bentleyi + C. striata var. involuta. All members of the sister clade of C. sinensis are distributed in North America (Freudenstein, 1997, 1999). C. sinensis is extremely similar to these members in sepals and petals morphology, but can be easily distinguished by its lip broad oblong and obviously 3-lobed toward base. In the clade consisting of C. sinensis, C. striata complex, and C. bentleyi, their mentum is absent. According to a monograph of Corallorhiza (Freudenstein, 1997), Corallorhiza was divided into two clades by the mentum absent or present. In current phylogenetic trees, red and green clade indicates mentum absent and present, respectively. The monograph indicated that the mentum is one of the characters to distinguish C. striata and other congeneric species (Freudenstein, 1997), thus the character presents a possible synapomorphy for the clade. Our phylogenetic
result is consistent with the previous studies (Kim et al., 2020; Li et al., 2020), that Corallorhiza is not a monophyletic group, because the genus Oreorchis is nested within the clade of Corallorhiza with strong bootstrap support (BS = 100) and posterior probability (PP = 1.00). In the Calypsoinae, however, phylogenetic analysis based on the nrDNA sequence suggests that Corallorhiza is a monophyletic group and strongly supports C. sinensis as sister to Corallorhiza (ML: BS = 98; BI: PP = 1.00). Although the topological conflicts are presented between plastome and nrDNA phylogenies of C. sinensis, it is still the most closely related to Corallorhiza. Both the plastid tree and nrDNA tree indicate Oreorchis is paraphyletic. The finding of this new species from China shed new light on the phylogenetic relationship of Oreorchis and Corallorhiza. Finally, we treat it here as a new species of Corallorhiza based upon morphological and molecular evidence.

Author Contributions

JXY, SP, JJW and HY collected these materials; GWH and QFW identified species; SX D, YW, JT, and HY designed and performed the experiments; JXY, SP, and JJW performed the analyses; JT drew the illustrations of the species; JY and SP wrote the manuscript; GWH and QFW revised the paper. All authors read and approved the final manuscript.

Declaration of competing interest

The author declares no conflict of interest.

Acknowledgements

The authors are grateful to the Project of Orchid Biodiversity Survey of China from National Forestry and Grassland Administration. We also thank professor Xiao-Hua Jin for confirming the identification and encouraging us to publish this new species, and staffs from Siguniangshan National Nature Reserves for their help during the field work. We acknowledge the National Wild Plant Germplasm Resource Center for all kinds of support. This work was supported by the grants from the Second Tibetan Plateau Scientific Expedition and Research (STEP) program (2019QZKK0502), the National Natural Science Foundation of China (31970211).

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.pld.2021.01.002.

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