Research Article

TTC7B Is a Novel Prognostic-Related Biomarker in Glioma Correlating with Immune Infiltrates and Response to Oxidative Stress by Temozolomide

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Background. Gliomas are one of the most prevalent malignant brain tumors. Hence, identifying biological markers for glioma is imperative. TTC7B (Tetratricopeptide Repeat Domain 7B) is a gene whose role in cancer is currently identified. To this end, we examined the TTC7B expression as well as its prognostic significance, biological roles, and immune system impacts in patients with glioma.

Methods. We evaluated the function of TTC7B in GBM and LGG through the published CGGA (Chinese Glioma Genome Atlas) and TCGA (The Cancer Genome Atlas) databases. CIBERSORT and TIMER were used to analyze the link between TTC7B and immune cells, while R was used for statistical analysis. In addition, Transwell analysis, including migration and invasion assays, was performed to identify the relationship between TTC7B and temozolomide.

Results. Low expression of TTC7B was observed in GBM and LGG. 1p/19q codeletion, IDH mutation, chemotherapy, and grade were found to have a significant correlation with TTC7B. Besides, low TTC7B expression was linked with low overall survival (OS) in both GBM and LGG. In the Cox analysis, TTC7B was found to independently function as a risk element for OS of patients with glioma. Furthermore, CIBERSORT analysis demonstrated a positive link between TTC7B and multiple immune cells, especially activated NK cells. Transwell analysis, including migration and invasion assays, revealed that temozolomide reduced the migration and invasion capacity of glioma cells and increased the expression of TTC7B.

Conclusion. In all, TTC7B could serve as a promising prognostic indicator of LGG and GBM, and is closely associated with immune infiltration and response to oxidative stress by temozolomide.

1. Introduction

Gliomas are among the most prevalent primary brain tumors in adults, accounting for over 70% of malignant brain tumors [1]. They have been categorized into three types: astrocytomas, oligodendrogliomas, and ependymomas on the basis of their histological characteristics and specific. World Health Organization (WHO) grades I-IV, which reflect the degree of malignancy [2–4]. The study of molecular mechanisms has led to a deeper understanding of gliomas. The codeletions of the chromosome arms 19q and1p, along with the molecular characterization of the primary brain tumors like IDH were included in gliomas as per the 2016 report of the WHO [3].

Although the current standard interventions, such as surgery, chemotherapy, and radiation, have improved the prognosis of patients with glioma [5, 6], it is still dismal. The local recurrence of tumor is closely associated with tumor heterogeneity, and the immune microenvironment of malignant tumor is a major reason for the failure of the treatment of malignant glioma [7]. Hence, new treatments for glioma are critical [8]. The molecular processes controlling the metabolism of glioma are rapidly developing, and are evidenced by a series of recent technological
developments [9]. It is important to urgently elucidate these molecular mechanisms to develop new therapies to avail diagnosis and treatment of glioma.

In cellular signaling, metabolism, and epigenetics, reactive oxygen species (ROS) are essential regulators. The altered metabolism of cancer cells is generally characterized by increased glycolysis and ROS levels [10]. Growth of tumors and inflammation can further increase ROS, shifting the redox balance towards oxidation. When ROS levels are low to moderate, they may act as signaling molecules, induce DNA mutations, and inactivate tumor suppressor genes. When ROS are present at high levels, they cause cellular damage and death, a principle that has been exploited in cancer treatments involving ionizing radiation (IR) and chemotherapy [11].

TTC7B (Tetratricopeptide Repeat Domain 7B) is a protein-coding gene, which is linked to several diseases, including hypomyelinating leukoencephalopathy and trichohepatoenteric syndrome 1 [12]. Nonetheless, TTC7B has not been reported in patients with glioma to date. Moreover, the relation of TTC7B with the immune cell infiltration status in LGG and GBM is unrecognized. Temozolomide is an essential medication for glioma [13]. Hence, for the first time, in the current study we analyzed the association of TTC7B with glioma.

2. Materials and Methods

2.1. Downloading and Preprocessing of Glioma Datasets. Clinical and transcriptome data of patients with glioma were

| Table 1: Baseline of CGGA patients information. |
|-----------------------------------------------|
| Total | Low-expression | High-expression | $\chi^2$ | p   |
|-------|----------------|-----------------|---------|-----|
| PRS_type |                |                 |         |     |
| Primary | 502            | 204             | 298     | 55.4292 | 0.0000 |
| Recurrent | 222            | 151             | 71      |       |
| Secondary | 25             | 20              | 5       |       |
| Grade |                |                 |         |     |
| WHO II | 218            | 45              | 173     | 186.0704 | 0.0000 |
| WHO III | 240            | 98              | 142     |       |
| WHO IV | 291            | 232             | 59      |       |
| Gender |                |                 |         |     |
| Male | 442            | 146             | 161     | 11.8564 | 0.0006 |
| Female | 307            | 229             | 213     |       |
| Age |                |                 |         |     |
| <=41 | 342            | 139             | 203     | 2.4806 | 0.1553 |
| >41 | 407            | 236             | 171     |       |
| Radio_status |        |                 |         |     |
| No | 124            | 69              | 55      | 174.5180 | <0.001 |
| Yes | 625            | 306             | 319     |       |
| Chemo_status |       |                 |         |     |
| No | 229            | 94              | 135     | 51.2404 | <0.001 |
| Yes | 520            | 281             | 239     |       |
| IDH_mutation_status |       |                 |         |     |
| Wildtype | 366            | 274             | 92      | 0.4197 | 0.5171 |
| Mutant | 383            | 101             | 282     |       |
| 1p19q_codeletion_status | |                 |         |     |
| Noncodel | 594            | 349             | 245     | 176.9779 | <0.001 |
| Codel | 155            | 26              | 129     |       |

| Table 2: The sequences of primer pairs for the target genes. |
|-------------------------------------------------------------|
| Gene | Forward primer sequence (5′-3′) | Reverse primer sequence (5′-3′) |
|------|---------------------------------|-------------------------------|
| TTC7B | CCGTCACCCACAGATCACC | CATGGACGGAGGCTCTCTG |
| RNF112 | CTTTCTGGGAGAAGGCA | CCAGGTGGCACAACATCTCC |
| NME5 | TGGAGATATCAATGCCCTCACCT | CCAATACGTAGCCAAAAATCACAGG |
| GAPDH | AATGGGCAGCCTTGGAAA | GCCCAATACGACAAATCACAGAG |

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| GAPDH | AATGGGCAGCCTTGGAAA | GCCCAATACGACAAATCACAGAG |
Overall survival

Logrank $p = 6.6e^{-08}$
$HR$ (high) = 0.5
$p$ (HR) = $1.2e^{-07}$

$n$ (high) = 338
$n$ (low) = 338

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**Figure 1**: Continued.
downloaded from the TCGA (http://cancergenome.nih.gov/) and CGGA (http://www.cgga.org.cn/) database. From the CGGA database, the WHO grade, radiotherapy and chemotherapy status, 1p/19q codeletion, and IDH mutation status of 2,000 glioma samples were obtained [14]. In all, 325 specimens (dataset ID: mRNAseq_325) and 693 specimens (dataset ID: mRNAseq_693) of RNA-seq data comprised the 1,018 samples. The clinicopathological information and informed consent were obtained for all the samples. The work was authorized by the Institutional Review Board of Tiantan Hospital. From the TCGA database, 703 samples were downloaded, including 698 tumor samples and 5 paracarcinoma samples [15, 16]. After implementing data preprocessing in different datasets, a correlation analysis was conducted between the clinical variables and TTC7B expression. Table 1 shows the comprehensive clinical data and the

| PRS_type  | pvalue | Hazard ratio          |
|-----------|--------|-----------------------|
|           | <0.001 | 1.961 (1.671 – 2.302) |
| Histology | 0.077  | 0.671 (0.431 – 1.044) |
| Grade     | <0.001 | 2.693 (1.970 – 3.683) |
| Gender    | 0.592  | 1.054 (0.870 – 1.275) |
| Age       | 0.054  | 1.215 (0.996 – 1.482) |
| Radio     | 0.319  | 0.872 (0.666 – 1.142) |
| Chemo     | 0.005  | 0.709 (0.557 – 0.902) |
| IDH_mutation | <0.001 | 0.594 (0.472 – 0.747) |
| 1p19q_codeletion | <0.001 | 0.378 (0.271 – 0.527) |
| TTC7B     | 0.005  | 0.847 (0.754 – 0.952) |

**Figure 1:** (a) Differential expression of TTC7B in GBM and LGG. (b) Survival curve analyzed by GEPIA in different TTC7B expression levels. (c, d) Multivariate and univariate Cox analysis of clinical-pathological factors and TTC7B expression. (e) The time-dependent receiver operating characteristic (ROC) curves of survival at 1-, 3-, and 5-years.
clinicopathological features of patients in the CGGA database. The R software (version 4.0.2) was utilized to carry out the gene expression and survival analyses. The Strawbery Perl software and the R software (version 4.0.2) enabled all the preprocessing procedures.

2.2. GEPIA Investigation of Survival and Expression. The Gene Expression Profiling Interactive Analysis (GEPIA) (http://gepia.cancer-pku.cn/index.html), an online repository, was employed to assess the link between TTC7B expression and survival of patients with glioma. The GEPIA 'survival' modules were used to evaluate the link between TTC7B expression and prognosis of patients with glioma. Furthermore, boxplots were used to depict the difference in TTC7B expression between the normal and tumor samples, with the disease conditions (normal or tumor) serving as variables.

2.3. Multivariate and Univariate Cox Model. Multivariate and univariate Cox analyses were conducted to evaluate

Figure 2: (a) Differences in TTC7B expression distribution in various WHO grades (dataset ID: mRNAseq_325). (b) Differences in TTC7B expression distribution in distinct WHO grades (dataset ID: mRNAseq_693).
Figure 3: Continued.
the correlation of 1p/19q codeletion, treatment response, and overall survival (OS) with TTC7B expression. The survival program in R (version 4.0.2) was utilized to acquire the CGGA statistical analysis.

2.4. CIBERSORT Analysis. CIBERSORT is an extensively used algorithm for evaluating the cellular composition of intricate tissues according to their gene expression patterns because it produces findings that are consistent and predictable in majority of cancer cases. The LM22 signature-based algorithm was adopted upon data entry of the gene expression profiles exhibiting standard annotations to the CIBERSORT website application (http://cibersort.stanford.edu/). A further step was downloading the LM22, which is an annotated gene profile matrix representing the 22 distinct kinds of immune cells. This was accomplished via the use of the CIBERSORT online resource. CIBERSORT was employed to detect distinct kinds of immune cells, such as T cells, B cells, macrophages, natural killer cells, myeloid subsets, and dendritic cells, accurately and sensitively [17–20]. Data were classified based on the median TTC7B expression levels into high and low TTC7B expression groups to assess the differences in the proportion of immune cells between these groups.

**Figure 3:** The expression of TTC7B in CGGA dataset. (a) IDH status-stratified distribution (dataset ID: mRNAseq_325). (b) IDH status-stratified distribution (dataset ID: mRNAseq_693). (c) The 1p/19q-codeletion status distribution (dataset ID: mRNAseq_325). (d) The 1p/19q-codeletion status distribution (dataset ID: mRNAseq_693).
Figure 4: Continued.
Figure 4: Continued.
2.5. TIMER Database Analysis. For the inclusive analysis of the TIICs, such as B cells, neutrophils, dendritic cells, CD4⁺ T cells, CD8⁺ T cells, and macrophages, the TIMER database (https://cistrome.shinyapps.io/timer/) was utilized by using the RNA-seq expression profile data [21, 22]. We used the “Gene” module plots to evaluate the link of TTC7B expression and immune infiltrate level with tumor purity.

2.6. Single-Cell Analysis. The Tabula Muris tool (https://tabula-muris.ds.czbiohub.org/) contains over 100,000 single-cell transcriptomes of 20 different tissues and organs. Through this database, we examined the associations between TTC7B expression levels and various types of cells and tissues, including endothelial cells and T lymphocytes. We also used fluorescence-activated cell sorting (FACS) to analyze the links between TTC7B expression and distinct types of cells.

2.7. Gene Set Enrichment Analysis. The Gene Set Enrichment Analysis (GSEA) (https://www.gsea-msigdb.org/) includes GO and KEGG pathway analysis, and was undertaken to examine the signaling pathways associated with TTC7B [23]. To evaluate the biological coherence and connections between each anticipated module, which was formed by correlating differently expressed mRNAs with distinct GO subsets, we performed GO analysis. An investigation of significant pathways linked to TTC7B expression was carried out using the KEGG analysis.

2.8. Quantitative RT-PCR. Total RNA was extracted from cell lines using the TRIzol reagent (Sigma-Aldrich, St. Louis, MO, USA). Then, 2 μg RNA from each sample was subjected to quantitative reverse transcription-polymerase chain reaction using FastStart Universal SYBR® Green Master (Roche, USA) on a Roche LightCycler 480 PCR System (Roche, USA). The cDNA was used as a template in a 20 μl reaction volume (10 μl of PCR mixture, 0.5 μl of forward and reverse primers, 2 μl of cDNA template, and an appropriate volume of water). PCR reactions were performed as follows: Cycling conditions started with an initial DNA denaturation step at 95°C for 30 s, followed by 45 cycles at 94°C for 15 s, at 56°C for 30 seconds, and at 72°C for 20 seconds. Each sample was examined in triplicate. Threshold cycle (CT) readings were collected and normalized to glyceraldehyde-3-phosphate dehydrogenase (GAPDH) levels in all samples using the 2−ΔΔCT method. The mRNA expression levels of tumor tissues were compared with those of normal tissue controls. The sequences of primer pairs for the target genes are shown in Table 2.

2.9. Cell Culture and Drugs. Human glioma cell lines U-87 and U-251 were obtained from ATCC (Beijing Beina Chuanqian Biotechnology Institute), and cultured in F12 and DMEM medium containing 10% fetal bovine serum (Gibco, Carlsbad, CA, USA), respectively. Both cell lines were stored in a humidified incubator at 37°C with 5% CO₂. Temozolomide was procured from MCE (USA, HY-17364), and dissolved in dimethyl sulfoxide (DMSO, Beyotime). Finally, it was cocultured with cells at a concentration of 20 μM/ml.
2.10. Transwell Assay. Transwell assay was performed to assess the migration and invasion of glioma cells (U-87, U-251). Briefly, $5 \times 10^4$ cells were inoculated into chambers coated (for invasion) or uncoated (for migration) with Matrigel (BD Biosciences, San Jose, CA). Serum-free medium was added to the upper layer and a complete DMEM medium was added to the lower layer. After 24 hours of incubation, migrating or invading cells were fixed.
with 4% paraformaldehyde, stained with 0.1% crystal violet, and counted under a light microscope.

3. Results

3.1. Relationship between TTC7B Expression and Glioma Survival Rates. TTC7B was found to be expressed at low levels in both, GBM [num (T) = 163 and num (N) = 207] and LGG [num (T) = 518 and num (N) = 207] (Figure 1(a)). Furthermore, low TTC7B expression was linked to an unfavorable OS [num (high) = 338 and num (low) = 338, P < 0.001; Figure 1(b)]. A bipartite technique was utilized to classify the TTC7B expression levels in tumor and adjoining normal specimens into two groups, namely, high- and low-expression groups.

3.2. TTC7B Expression as an Independent Predictive Indicator for Individuals with Glioma. In univariate analysis, variables such as TTC7B expression (P < 0.001), PRS_type (P < 0.001), histology (P < 0.001), grade (P < 0.001), 1p19q_codeletion (P < 0.001), and IDHmutation (P < 0.001) provide confirmation that TTC7B independently functions as a biological marker for patients with glioma (Figure 1(c)). Similarly, in the multivariate model, factors linked to TTC7B expression (P = 0.005), notably, PRS_type (P < 0.001), IDH_codeletion (P < 0.001), 1p19q_codeletion (P < 0.001), and grade (P < 0.001), demonstrated that TTC7B independently served as a prognostic marker (Figure 1(d)). These findings demonstrate that TTC7B independently functions as a predictive marker for glioma and is highly correlated with several other variables associated with glioma. Moreover, the AUC of TTC7B expression was 0.57, 0.60, and 0.62 for 1-, 3-, and 5-year survival, respectively (Figure 1(e)). This illustrates that TTC7B has a satisfactory prognostic performance in anticipating the survival of patients with glioma.

3.3. The Association of TTC7B Expression with 1p19q Codeletion and IDH1 Phenotype Status in CGGA. The connection between TTC7B expression level and survival was evaluated in two separate datasets depending on the WHO grade and IDH1 phenotype. The relationship between TTC7B expression pattern and WHO grade was examined and compared in two separate datasets (IDs: mRNAseq 693 and mRNAseq 325). Both datasets demonstrated a significant relationship between TTC7B expression pattern and WHO grade in gliomas (Figures 2(a) and 2(b)). These
Figure 7: Continued.
findings suggest that increased malignancy of glioblastoma was linked to attenuated TTC7B expression. Moreover, TTC7B expression was considerably downregulated in the IDH wild type group compared with the IDH mutant group, predicated on the two datasets characterized by IDH mutation status (Figures 3(a) and 3(b)) and computed using the ANOVA algorithm. TTC7B expression was remarkably reduced in the 1p/19q noncodeletion (noncodel) group (Figures 3(c) and 3(d)) when compared with the 1p/19q codeletion (T-test) group. These findings illustrate that TTC7B expression level was reduced in the IDH mutant and 1p19q codeletion groups.

3.4. TTC7B Overexpression Was Associated with a Favorable Chance of Survival in Primary Glioma. To examine the link between TTC7B expression and survival status (prognosis) in patients with WHO-graded glioma, a thorough survival analysis was conducted using the two CGGA datasets. TTC7B overexpression in dataset 1 (ID: mRNAseq_325) predicted favorable outcomes in primary glioma ($P < 0.001$; Figure 4(e)). Thus, we infer unequivocally that TTC7B overexpression level was correlated with better survival prognosis of patients with primary glioma.

3.5. Analysis of Multivariate Integrated Survival Data from the CGGA. To evaluate the therapeutic significance of TTC7B, factors such as chemotherapy (Figure 4(a)), radiation (Figure 4(b)), IDH1 phenotypes (Figure 4(c)), and 1p/19q codeletion status (Figure 4(d)) were included in the multivariate analysis. In the 1p/19q-codeletion status group, lower TTC7B expression levels with 1p/19q codeletion predicted favorable survival outcomes compared with high TTC7B expression in patients without 1p/19q codeletion (Figure 4(d)). Additionally, TTC7B expression and chemotherapy were used to investigate the links between the survival probabilities. Elevated expression of TTC7B in the absence of chemotherapy was linked to favorable survival outcomes, whereas decreased expression of TTC7B in the
Figure 8: Continued.
Figure 8: Continued.
The presence of chemotherapy was associated with unfavorable survival results (Figure 4(a)). TTC7B seems to be substantially linked to all factors studied so far ($P < 0.0001$).

### 3.6. Association between TIICs and TTC7B Expression

We evaluated the correlation between TIICs of glioma and TTC7B expression level. Based on the CGGA specimens, the infiltration degree of immune cells, such as monocytes, activated mast cells, and T follicular helper cells, was considerably elevated in the high-expression group compared with the low-expression group. Further, based on the TCGA database, the infiltration degree of immune cells, such as plasma cells, T follicular helper cells, naïve B cells, and eosinophils, was considerably elevated in the high-expression group compared with the low-expression group. In addition, the CGGA (Figure 5(a)) and TCGA database (Figure 5(b)) revealed that NK cell activation ($P < 0.0001$) was significantly reduced in the TTC7B high expression group.

### 3.7. TTC7B Expression Is Linked to the Degree of Immune Infiltration and OS in GBM and LGG Derived from TIMER

TIMER database was used to investigate whether the immune invasion level of glioma is related to TTC7B expression. In addition, we selected the TTC7B expression negatively linked to LGG purity. We identified a positive link between CD8$^+$ T cell infiltration and TTC7B expression ($r = 0.099$, $p = 3.03e-02$) (Figure 6(a)). In addition, the expression of TTC7B was intimately linked to the immune-infiltrating microenvironment of macrophages, CD4$^+$ T cells, and B cells in LGG. Furthermore, accumulation rates of GBM and LGG were found to be associated with DCs, macrophages, T cells neutrophils, and B cells (Figure 6(b)).

### 3.8. Investigation of TTC7B Expression and Cells from Various Organs by Single-Cell Analysis

We used the Tabula Muris database to study the relationship of TTC7B expression with cells. As demonstrated in Figure 7(a), brain tumors were linked to endothelial cells, basal cells, luminal epithelial cells of the mammary gland, stromal cells, and t-SNE of FACS cells. TTC7B was primarily linked to endothelial cells, basal cells, luminal epithelial cells of the mammary gland, and stromal cells, as illustrated in Figure 7(b).

### 3.9. Analysis of TTC7B-Related Pathways Using GSEA

We conducted GO and KEGG pathway analyses to probe into the probable bioactivities of TTC7B. We selected three
pathways that were strongly linked with \textit{TTC7B} expression and discovered that \textit{TTC7B} is tightly linked to cellular metabolism and pathways. The KEGG pathway analysis illustrated that the MAPK, GnRH, and inositol phosphate metabolism signaling pathways exhibited a positive link to the elevated expression of \textit{TTC7B}. In contrast, the three inversely linked categories were systemic lupus erythematosus, ECM receptor interaction, and cytokine-cytokine receptor interaction (Figure 7(c)). GO analysis revealed that \textit{TTC7B} regulates response to oxidative stress (Figure 7(d)).

3.10. \textit{TTC7B} Inhibits the Migration and Invasion of Glioma Cells. To assess the role of \textit{TTC7B} in glioma progression, we examined \textit{TTC7B} mRNA expression using qRT-PCR. PCR results indicated a decreased \textit{TTC7B} expression in glioma patients. Transwell analysis (in vitro), including migration and invasion assays, revealed that migration and invasion capacity of glioma cells were reduced and \textit{TTC7B} expression was increased with the use of temozolomide (Figure 8).

4. Discussion

Previous investigations have never identified the involvement of \textit{TTC7B} gene in cancer. Hence, in the current study, we aimed to evaluate the role of \textit{TTC7B} gene as prognostic biomarker of gliomas. We demonstrated that differences in the degree of \textit{TTC7B} expression are linked to prognosis of patients with glioma. Additionally, \textit{TTC7B} expression was found to independently serve as a prognostic marker for a positive prognosis. Moreover, the expression patterns of \textit{TTC7B} were shown to be substantially correlated with a variety of clinical parameters, particularly pathological stage and tumor status. Furthermore, we found that \textit{TTC7B} expression in glioma is linked to the expression of a variety of immune biomarkers and the degree of immune infiltration. Hence, the findings of the current investigation showed that \textit{TTC7B} could have possible effects on tumor immunotherapy and might function as a promising cancer-related biological marker.

Using GEPIA, an online database, we discovered a link between \textit{TTC7B} expression and prognosis of patients with glioma. The upregulated expression of \textit{TTC7B} was shown to be linked to a positive prognosis. We obtained information from the TCGA database to better investigate the underlying processes and roles of \textit{TTC7B} expression in cancers. \textit{TTC7B} expression was shown to be associated with several clinical parameters, including pathological stage and tumor status, according to a statistical analysis conducted utilizing R-4.0.2. The results of multivariate analysis illustrated that \textit{TTC7B} expression independently serves as a predictive factor for prognosis of patients with glioma. This research also compared the similarities between \textit{IDH1} and \textit{TTC7B} expressions. As indicated by WHO, \textit{IDH1} phenotypes serve as a unique diagnostic technique for clinical usage and categorization of diffuse gliomas among adults is mostly determined by \textit{IDH1} mutation status [3, 24, 25]. In this study, we investigated the differences between \textit{IDH1} wild type and \textit{IDH1-R132} mutant groups. The elevated expression of \textit{TTC7B} inhibits the progression of glioma to a malignant state, as evidenced by the reduced survival of the \textit{IDH1-R132} mutant group. Additionally, by comparing radiotherapy with chemotherapy, we were able to demonstrate the potential applications of \textit{TTC7B}. Hence, the potential of \textit{TTC7B} as a molecular predictor of prognosis of glioma was investigated in this research.

CIBERSORT analysis revealed a significant explicit link between \textit{TTC7B} expression and NK cell infiltration levels in glioma. A similar pattern was seen in connections between gene biomarkers of various immune cells and \textit{TTC7B} expression. This suggests that \textit{TTC7B} has an important role in modulating the tumor immune microenvironment of glioma. Using the CIBERSORT algorithm we discovered that the proportion of NK cells was elevated in the high-expression group compared with the low-expression group. NK cells are viable immune effectors [26]. According to previous studies, NK cells may generate cytokines, including IFN-\textgamma and TNF-\textalpha, which can suppress the progression, proliferation, and invasiveness of gliomas [27–32]. As a result, the favorable effects of \textit{TTC7B} on glioblastoma are consistent with the role of high levels of NK cells, suggesting that \textit{TTC7B} may have an impact on the OS of patients with glioma. Nevertheless, controlled experiments and multicenter clinical trials are required in the future to get a more precise understanding of the interaction between \textit{TTC7B} and NK cells in vivo.

From GSEA analyses, the high-\textit{TTC7B} expression group exhibited substantial enrichment of oxidative stress-related gene sets. In the previous research, in response to ionising radiation (IR) and chemotherapy, reactive oxygen species (ROS) are produced and are responsible for the mutagenic and cytotoxic effects of these agents. Increasing ROS levels cause DNA damage, lipid oxidation, and protein oxidation, ultimately leading to tumour cell death. Prior research identified putative pathways that might explain the association of \textit{TTC7B} expression with better prognosis. Meanwhile, from qRT-PCR and Transwell analysis, \textit{TTC7B} inhibits glioma cell migration and invasion, and temozolomide treatment increases \textit{TTC7B} expression.

5. Conclusion

Overall, \textit{TTC7B} serves as a predictive biological marker with prospective applications, and is associated with the immune infiltration and oxidative stress of gliomas. It might also function as a novel target for the regulation of immunosuppression. With the absence of a biological validation being performed, the study has several limitations. To elucidate its role in glioma, further clinical and experimental studies are required. However, it is expected that large sample sizes from CGGA and TCGA will help the subsequent investigation of gliomas.

Data Availability

The data used in this article is downloaded from the public databases TCGA (http://cancergenome.nih.gov/) and CGGA (http://www.cgga.org.cn/). Readers can download and use it for free.
Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

Zhenhua Chen, Shasha Cui, and Yong Dai contributed equally to this work.

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