Study of bicyclomycin biosynthesis in *Streptomyces cinnamoneus* by genetic and biochemical approaches

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The 2,5-Diketopiperazines (DKPs) constitute a large family of natural products with important biological activities. Bicyclomycin is a clinically-relevant DKP antibiotic that is the first and only member in a class known to target the bacterial transcription termination factor Rho. It derives from cyclo-(l-isoleucyl-l-leucyl) and has an unusual and highly oxidized bicyclic structure that is formed by an ether bridge between the hydroxylated terminal carbon atom of the isoleucine lateral chain and the alpha carbon of the leucine in the diketopiperazine ring. Here, we paired in vivo and in vitro studies to complete the characterization of the bicyclomycin biosynthetic gene cluster. The construction of in-frame deletion mutants in the biosynthetic gene cluster allowed for the accumulation and identification of biosynthetic intermediates. The identity of the intermediates, which were reproduced in vitro using purified enzymes, allowed us to characterize the pathway and corroborate previous reports. Finally, we show that the putative antibiotic transporter was dispensable for the producing strain.

2,5-Diketopiperazines (DKPs) are a class of molecules characterized by the presence of the piperazine-2,5-dione ring obtained by the condensation of two alpha amino acids. Specialised metabolites with a DKP scaffold are produced by a wide range of microorganisms and present various interesting biological properties, including antibacterial, antifungal, antiviral or antitumoral activity. Among the DKPs, bicyclomycin (1), also known as aizumycin or bicozamycin (Fig. 1a), is a highly oxidized, clinically-relevant molecule with a unique mechanism of action. The ability of bicyclomycin to disrupt the activity of the bacterial termination factor Rho has been used to treat enteropathogenic *Escherichia coli* that causes diarrhoea in humans and livestock animals. Moreover, the use of bicyclomycin in combination with bacteriostatic concentrations of protein synthesis inhibitors dramatically enhances its bactericidal properties and potential clinical utility.

Bicyclomycin was first discovered as a natural product of *Streptomyces cinnamoneus* (= *Streptomyces sapporoensis*). *Streptomyces aizunensis* and *Streptomyces griseoflavus* have since been reported as producers. In bacteria, the synthesis of the DKP core can be catalysed either by nonribosomal peptide synthetases (NRPSs) or by cyclodipeptide synthases (CDPSs). At the outset of our investigation into the biosynthesis of bicyclomycin only the first and last intermediate were proposed: cyclo-(l-isoleucyl-l-leucyl) (cIL) (2) and dihydrobicyclomycin (3), respectively. During the final steps of our work, which focused on *S. cinnamoneus*, several studies on bicyclomycin biosynthesis that used in vitro approaches or heterologous expression were published, identifying biosynthetic genes and proposing a biosynthetic pathway for bicyclomycin in both Actinobacteria and Proteobacteria.

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Our work provides in vivo data that corroborate the results obtained in vitro on the biosynthetic pathway of bicyclomycin, establish the stereochemistry of several biosynthetic intermediates and demonstrate the dispensable nature of the transporter associated with the biosynthetic gene cluster in S. cinnamoneus.

**Results**

**Functional analysis of the bicyclomycin biosynthetic gene cluster in S. cinnamoneus.** Having generated and analysed a draft genome sequence of the bicyclomycin producer S. cinnamoneus, we had identified as candidates to direct bicyclomycin biosynthesis the same genes as those identified by other groups studying this topic. These genes constitute a cluster composed of a CDPS gene, five genes encoding 2-oxoglutarate/iron-dependent dioxygenases (2OG/Fe dioxygenases), one gene encoding a cytochrome P450 monoxygenase, and one gene encoding a transporter belonging to the Major Facilitator Superfamily (MFS). The cluster is flanked by genes with divergent functions that are not involved in bicyclomycin production, including genes with predicted roles in chitin degradation or the biosynthesis of an unrelated polyketide.

To provide unambiguous evidence for the involvement of the biosynthetic gene cluster (BGC) and decipher the role of individual genes in bicyclomycin biosynthesis, we attempted recombinant gene expression by introducing the complete gene cluster into Streptomyces coelicolor (strain M1154) or Streptomyces lividans (strain TK24). For these experiments, a plasmid encoding the complete bcm was generated in E. coli (Fig. 1b). The introduction of pJWe14 into M1154 or TK24 did not result in the heterologous production of bicyclomycin, even though for each strain, nine different culture media were tested (data not shown).

We therefore decided to investigate the role of the bcm cluster directly in S. cinnamoneus. Accordingly, we tested ten culture media for bicyclomycin (1) production by S. cinnamoneus and observed that only growth in MP5 resulted in reliable amounts detected in culture supernatants by HPLC/ELSD (Fig. 2b); the chemical standard (Santa Cruz Biotechnology, Ref. sc-391755) was used for comparison (Fig. 2a). In these culture conditions, bicyclomycin production began after two days and the amount increased over an eleven-day period (Supplementary Fig. S1). Deletion of the whole bcm cluster from S. cinnamoneus (Δbcm::aphII mutant, Supplementary methods) abolished bicyclomycin production (Fig. 2c). Furthermore, bicyclomycin production could be restored in the mutant S. cinnamoneus strain using the pJWe14 vector which carries the entire bcm cluster in its native configuration (Figs. 2d,e).

To study the involvement of each gene in bicyclomycin biosynthesis, we undertook their in-frame deletion. As chromosomal gene deletion is difficult and time-consuming in S. cinnamoneus, the pJWe14 vector appeared an excellent tool to study the bcm cluster. By PCR-targeting followed by excision of the resistance cassette used for gene replacement, we constructed in E. coli a set of pJWe14 derivatives, each of them with an in-frame deletion of one of the bcm biosynthetic gene, from bcmA to bcmG. To avoid as much as possible effect on downstream gene expression and to preserve translation signals and possible promoters, the first 30 nucleotides and the last 90 nucleotides of each gene were conserved in the in-frame deletion mutants. These derivatives were then introduced into the S. cinnamoneus Δbcm::aphII mutant. The culture supernatants of each Δbcm::aphII mutant harbouring modified bcm BGCs were analysed by HPLC/ELSD. Deletion of each of the six predicted biosynthetic genes bcmB-G resulted in the abolishment of bicyclomycin production. Furthermore, we constructed a vector expressing the bcmA gene alone under the control of its native promoter and showed that it rescued cIL (2) production. LC/MS analysis demonstrated that the accumulated metabolite has a retention time, m/z value and fragmentation pattern similar to an authentic standard of 2 (Supplementary Fig. S2 and Supplementary Table S1). No other cycloleupeptide was detected in this experiment. Taken together, these data suggest that each of the seven genes from bcmA to bcmG is required for bicyclomycin biosynthesis. A representative chromatogram from this experiment is shown in Fig. 2f, chromatograms corresponding to all deletion mutants are presented in Supplementary Fig. S3.

**In vivo characterization of the bicyclomycin biosynthetic pathway.** HPLC chromatograms corresponding to the culture supernatants of S. cinnamoneus Δbcm::aphII harbouring an ectopic copy of the bcm BGC intact or with in-frame deletion of each of the genes encoding tailoring enzymes (bcmB, C, D, E, F, or G) were analysed. They showed the appearance of additional peaks in the case of chromatograms corresponding to

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**Figure 1.** Bicyclomycin and its biosynthetic gene cluster. (a) Structure of the bicyclomycin molecule; (b) schematic representation of the bicyclomycin biosynthetic gene cluster and its flanking regions. The predicted functions of the proteins encoded by the cluster are: BcmH: MFS transporter; BcmA: cycloleupeptide synthase; BcmB, C, E, F, G: 2OG/Fe dioxygenase; BcmD: cytochrome P450 monoxygenase. The blue line indicates the extent of the insert cloned in pJWe14. The structure of the bicyclomycin molecule was drawn using ChemDraw version 18.0.0.231.
the different mutants (e.g. Fig. 2f and Supplementary Fig. S3). LC/MS/MS analyses of the same samples allowed the detection of several products whose fragmentation patterns indicate that they are related to the bicyclomycin scaffold (Supplementary Data Sheet 1). These products could be intermediates in bicyclomycin biosynthesis, as previously detected by Bradley et al. for intermediates 2 and 3 or shunt products resulting from the promiscuous activity of some tailoring enzymes on substrate analogues that accumulate in the absence of proper metabolic flux. Some of these products were also detected in trace amounts by LC/MS analysis in the culture supernatants of S. cinnamoneus Δbcm::aphII harbouring an intact copy of the bcm BGC. We considered that only these products were intermediates in bicyclomycin biosynthesis (Table 1).

We detected 2 in the supernatants of all our constructions in varying amounts, ranging from traces to quantifiable amounts in terms of UV214 peak area (Supplementary Data sheet 1). Intermediate 2 specifically accumulates...
in the bcmE-deleted mutant supernatant, suggesting the involvement of BcmE in the first modification of 2. We also detected and identified intermediate 3 thanks to its molecular weight and fragmentation pattern, consistent with previous reports (Supplementary Table S1, Supplementary Data Sheet 1). As 3 is observed specifically in the supernatant of the bcmF-deleted mutant, we could assign BcmF the role of dehydrogenation of the isoleucyl side chain of 3 to produce bicyclomycin (1). Comparison of the compounds detected in the different strain supernatants enabled us to identify two compounds of molecular weight 288 (products 4 and c2, Supplementary Data Sheet 1), likely to be the penultimate precursor to bicyclomycin. They are both present in trace amounts in the presence of the whole cluster and in the bcmF-deleted strain, and in quantifiable amounts in terms of UV \_\alpha peak area in the bcmD-deleted strain. Similarly, product 5 of molecular weight 274, is the only product found in both the whole-cluster-harbouring strain and the bcmF-, bcmD- and bcmB-deleted strains. It specifically accumulates in the bcmB-deleted strain, suggesting that BcmB catalyses the reaction from 5 to 4 (or to c2, see the next section). A unique compound, product 6, whose molecular weight of 258 is compatible with the role of precursor to 5, is found in both the whole-cluster-harbouring strain and the bcmG-deleted strain. Product c4 (molecular weight of 256), which also accumulates in the bcmG-deleted strain was not considered as an intermediate to bicyclomycin, as it contains a further dehydrogenation not compatible with 5 as the next intermediate. The last intermediate would be product 7 (molecular weight of 242), which accumulates in the bcmC-deleted strain and is also present in varying amounts in the whole-cluster-harbouring strain and the bcmG-, bcmD- and bcmB-deleted strains.

All together, our in vivo results allowed to propose the sequence in which each Bcm enzyme is involved in the pathway of biosynthesis, consistent with the generation of a highly oxidized cyclic dipetide. This sequence is in agreement with recently reported in vitro studies. The candidate intermediates are listed in Table 1 by ascending molecular weight values derived from LC/MS experiments (Supplementary Table S1, Supplementary Data Sheet 1).

The fragmentation pattern of MS2, MS3 and MS4 data (Supplementary Data Sheet 1) also allowed making hypotheses on the reactions catalysed by Bcm enzymes, as explained in Supplementary information §1.3. However, ambiguities remained for some of these reactions (Supplementary Fig. S4). These hypotheses are compatible with data reported previously.

**In vitro characterization of the cIL tailoring pathway and stereochemical analysis of some intermediates.** In order to correlate the biosynthetic intermediates identified from in vivo analyses to the compounds previously identified in in vitro studies, we characterized the activity of the tailoring enzymes BcmB, BcmC, BcmD, BcmE, BcmF and BcmG in vitro. An N-terminal histidine-tagged recombinant form of each protein was produced in E. coli and purified using nickel affinity chromatography. Single or multiple purified enzymes were reacted with cIL (2) or purified intermediates. The reaction mixtures and purified products were analysed by HPLC and ESI/MS, respectively (Supplementary Figs. S5–S8). The fragmentation patterns of the products obtained in vitro were compared to those of the products detected in vivo with mutant strains to check their concordance (Supplementary Table S1). The final assignments of product structures were determined by \textsuperscript{1}H, \textsuperscript{13}C NMR spectroscopy (Supplementary Information and Supplementary Figs. S9–S20). In addition to planar chemical structure elucidation, NMR analysis yielded information on the stereochemistry of compounds (Fig. 3). The observation of NOEs between H\_\textalpha protons in DKP ring indicated a syn orientation of amino acid side chains for products 2, 5, 6 and 7. The determination of the configuration of new stereogenic centres in products 5 and 4 also relied on a NOE- and J-based NMR strategy. The stereochemical analysis was based on the Karplus-type dihedral angle dependence of homonuclear vicinal coupling constants (\textsuperscript{1}J\textsubscript{HH}) and heteronuclear coupling constants (\textsuperscript{13}J\textsubscript{CH}). In addition, two-bond carbon-proton coupling constants (\textsuperscript{3}J\textsubscript{CH}) provided useful conformational and configurational information in oxygen-substituted two-carbon fragments. The results are summarized in Table 2 and detailed below. The identified compounds with their stereochemistry are positioned in the bicyclomycin biosynthetic pathway (Fig. 3).

**The in vitro product of BcmE catalysis is 7.** We began our in vitro studies by focusing on the transformation of synthetic cIL by the predicted 2OG/Fe dioxygenase in the presence of iron (II) and 2-oxoglutarate. BcmE was the only 2OG/Fe dioxygenase that efficiently transformed cIL (2) into a novel product. MS/MS fragmentation pattern of the product of the BcmE-catalysed transformation of 2 did correspond to 7 (Supplementary Table S1), which accumulated in supernatants of the \textDelta bcmC strain. Finally, NMR analyses confirmed that BcmE catalyses the hydroxylation of cIL at the Ile \textalpha C\textbeta to form 7. Neither BcmF nor BcmB were able to transform cIL (data not
Table 2. Results of in vitro characterization of tailoring enzymes encoded by the bcm cluster. “U” means unidentified products that did not correspond to any product observed in culture supernatants of the S. cinnamoneus strains (Supplementary data sheet 1).

| Substrate | Enzyme | Observed products | Relevant Supplementary information |
|-----------|--------|-------------------|-----------------------------------|
| cIL (2)   | BcmE   | 7                 | Supplementary Fig. S5A for the reaction of BcmE on cIL (2). | BcmG Supplementary Fig. S5C for the reaction of BcmG on cIL (2). |
| cIL (2)   | BcmC   | U                 | Supplementary Fig. S5A for the reaction of BcmC on cIL (2). |
| cIL (2)   | BcmG   | U                 | Supplementary Fig. S5C for the reaction of BcmG on cIL (2). |
| cIL (2)   | BcmE + BcmC | 7, 6            | Supplementary Fig. S5A for the reaction of BcmE + BcmC on cIL (2). |
| cIL (2)   | BcmE + BcmG | 7, U            | Supplementary Fig. S5A for the reaction of BcmE + BcmG on cIL (2). |
| cIL (2)   | BcmE + BcmC + BcmG | 7, 6, 5, U | Supplementary Fig. S5A for the reaction of BcmE + BcmC + BcmG on cIL (2). |
| cIL (2)   | BcmE + BcmC + BcmG + BcmB | 7, 6, 5, 4, c2, U | Supplementary Fig. S5A for the reaction of BcmE + BcmC + BcmG + BcmB on cIL (2). |
| 4         | BcmD   | 3                 | Supplementary Fig. S7 for the reaction of BcmD on cIL (2). |
| 3         | BcmF   | bicyclomycin (1)  | Supplementary Fig. S8 for the reaction of BcmF on cIL (2). |

6 is the next intermediate, synthesized by BcmC. We next analysed BcmE and BcmC or BcmG reaction mixtures that contained cIL as a substrate. A new product was observed in significant quantities when BcmE and BcmC were combined and reacted with cIL (2). The MS/MS characteristics of this product are identical to 6, previously observed in vivo when the ΔbcmG strain was grown in MP5. NMR analyses were used to assign 6 to the hydroxylated product of 7 at the Cγ position of Leu side chain (Fig. 3), in agreement with previous studies12. No compound corresponding to c4 could be detected in in vitro experiments, suggesting that this compound which accumulates in large quantities in the ΔbcmG strain is out of the biosynthesis pathway. When BcmE was combined with BcmG, 7 was slowly converted into a new product; however, mass analysis of this product was not correlated to any metabolite observed in supernatants of bcm mutant strains.

5 is the fourth biosynthetic intermediate. As BcmG was the next enzyme in the pathway based on our in vivo analyses and published in vitro experiments11,12, we incubated cIL with BcmE, BcmC and BcmG, and observed that the disappearance of 6 was concomitant with the appearance of a novel product, that matched product 5 accumulating in the bcmB-deleted strain. The chemical structure and stereochemistry of 5 were confirmed by NMR analysis (Supplementary table S6). For 5, the measurement of J_{H_6-H_5}, J_{H_6-C_5}, and J_{H_6-C_3} provided unambiguous stereospecific assignment of H3 methylenic protons and indicated the predominance of a major Cα–C3 rotamer of the leucyl side chain, with a χ1 angle of −60°. The analysis of 2J_{CH} coupling constants involving H3 protons and Cγ, Cβ, Cb1 and Cb2 carbons, together with NOEs is consistent with a R configuration of Cγ atom and a favoured rotamer around Cβ–Cγ bond adopting a χ2 angle (Cα–Cβ–Cγ–O atoms) of +60°.

Incubation of cIL with BcmE, BcmC, BcmG and BcmB results in the bicyclic metabolite 4. Finally, the four-enzyme mixture of BcmE, BcmC, BcmG and BcmB was able to convert 2 to 4. The metabolite matched the one which accumulated in the ΔbcmD strain, and NMR analysis of the enzyme-generated product was used to assign the structure of 4 (Supplementary table S7). For 4, the analysis of NOEs in combination with J_{H_6-H_5} and J_{H_6-C_3} heteronuclear coupling constants of H3 proton with all carbon atoms of leucyl side chain are in agreement with 5 configurations of both Cα and Cβ atoms and the presence of major rotamers around Cα–C3 and Cβ–Cγ bonds. Product 2 was also detected in in vitro experiments, in much smaller quantities than 4. It could not be isolated in quantities compatible with NMR analyses.

4 is efficiently transformed in dihydrobicyclomycin (3) by BcmD. We then incubated 4 with the cytochrome P450 BcmD in the presence of NADPH and the spinach electron transfer chain system (incubation time 24 h). BcmD catalysed the transformation of 4 to a new product with m/z value and fragmentation pattern consistent with 3 (dihydrobicyclomycin). This result confirms that 4 is a true precursor of dihydrobicyclomycin, c2 being a by-product of the biosynthesis pathway.

BcmF generates a vinylidene to complete the biosynthesis. Finally, when 3 was incubated with BcmF in the presence of iron (II) and 2-oxoglutarate, the molecule was transformed into a new product with m/z value and fragmentation pattern consistent with bicyclomycin (1), in agreement with the vinylidene formation.

In vivo detection of a shunt pathway in the bcmE deletion mutant. We observed in vivo that the strain devoid of BcmE accumulates, in addition to cIL (2), some products in large quantities including c26, c27, c28, c29, c31 and c32 (Supplementary Data sheet 1 and Supplementary Table S2). Products c26, c27, c29, c31...
Δbcm::aphII (in red), S. cinnamoneus Δbcm::aphII + pMC16 (clone 1 in blue, clone 2 in green), S. cinnamoneus Δbcm::aphII + pJWe21 (in pink). The bicyclomycin (1) peak is indicated by an arrow. The fact that this peak corresponds to bicyclomycin was confirmed by LC/MS/MS analysis.

and c32 have the same mass as compounds identified in vitro in the BcmC shunt pathway by Meng et al.12 (compounds 10, 11, 12, 14a and 14b according to their numbering). Moreover, their relative retention times in our chromatographic conditions are the same as those observed by Meng et al. and their fragmentation patterns are compatible with the structure proposed by Meng et al.12 Another product, c28, might be a desaturation intermediate between the products 11 and 12 characterized by Meng et al.12. These products are not detected in the other mutant strains in which BcmE is present. This indicates that, in vivo, in the absence of BcmE, BcmC or BcmG could act on cIL (2) leading to the shunt pathway observed in vitro by Meng et al.12.

BcmH is dispensable for bicyclomycin export in S. cinnamoneus. BcmH is predicted to be a MFS transporter, and when expressed in E. coli, BcmH confers resistance to bicyclomycin. To test the role of BcmH in bicyclomycin export in the producing organism, the bcmH gene was inactivated on the pJWe14 plasmid, yielding pMC16. As a precautionary measure against the loss of an indispensable self-resistance gene, another plasmid, pJWe21, carrying the bcmH gene under the control of its native promoter was also constructed. We then attempted the construction of two strains derived from the Δbcm mutant: one containing only pMC16, the other containing pJWe21 and pMC16. As these two strains were obtained, we then compared bicyclomycin production by both strains. Similar amounts of bicyclomycin were found in the culture supernatants of both strains (Fig. 4). We also examined by HPLC cell-free extracts of both strains and no bicyclomycin was detected. These results indicate that BcmH is non-essential for the survival of the strain in conditions of bicyclomycin production and is also dispensable for export, which is novel in the light of previous publications11.

Finally, we examined the transcription of the bcm genes by RT-PCR analysis, using RNA extracted after seven days of cultivation, a time at which bicyclomycin is produced by S. cinnamoneus. The results (Supplementary Fig. S21) show that while the bcm biosynthetic genes (bcmA-G) are transcribed, bcmH is not. These gene expression data are in agreement with the nonessential phenotype observed for the bcmH-deleted strain during bicyclomycin production.

Discussion
Our study combined genetic manipulations of the BGC for bicyclomycin that is encoded by S. cinnamoneus with in vitro enzyme characterization to complement and confirm in vivo the pathway proposed by Meng and coworkers12.

By creating and characterizing in frame gene deletions for each of the biosynthetic genes, we were able to correlate the metabolic intermediates that accumulated in vivo to the products of in vitro reactions. Our work is unique in that we deleted the full gene cluster, and replaced it with native and modified ectopic copies of it, generating robust in vivo data for each step in bicyclomycin biosynthesis. Moreover, we established the stereochromy of some of the intermediates (4 and 5). This pathway is the most complex CDPS-dependent DKP biosynthetic pathway characterized to date with six tailoring enzymes involved in the conversion of the cyclodipeptide cIL (2).

Concerning the reactions catalysed by the tailoring enzymes, hydroxylations (as the one introduced by BcmC, D, E and G) are one of the canonical reactions in the repertoire of both O2/O Fe dioxygenases and cytochrome P450 monoxygenases; desaturations (like the one catalysed by BcmF) are also well documented20–22. The reaction catalysed by BcmB results in hydroxylation and ether bridge formation, and is original but not unheard of. Meng et al. proposed that it processed through at least two BcmB-catalyzed reactions, a desaturation and an epoxidation, followed by the nucleophilic attack of the CO carbon atom involved in the epoxide functional group by the oxygen atom of the hydroxylated Ile C8.

Meng et al. reported that in vitro BcmC was more active on cIL (2) than BcmE, this reaction being the first of a shunt pathway involving also BcmG and BcmB. We did not observe this activity of BcmC on 2 in our in vitro results, as we have seen almost no substrate depletion over one hour incubation of 2 with BcmC (Supplementary Fig. S5A) and complete substrate transformation with BcmE (Supplementary Fig. S5B). The difference could be
explained by different in vitro reaction conditions, as the buffer type, substrate and enzyme concentrations were substantially different. In particular, the substrate to enzyme ratio was 10 times lower in the conditions used by Meng et al. 12. We observed in vivo in the bcmE-deleted mutant a shunt pathway similar to the one observed in vitro by Meng et al. 12. This confirms that BcmC, BcmG and BcmB are also promiscuous in vivo. Paterson et al. 13 also observed that overincubation of 6 with BcmG leads to the supplementary oxidation reactions performed by this enzyme that are not normally observed in vivo. We made similar observation in vitro but did not characterize the compound obtained in these conditions. Other compounds related to bicyclomycin such as c2 or c4, not structurally characterized in this study, also accumulate in large quantities in some deletion mutant strains. Taken together, these observations made in vitro or in vivo indicate that some of the Bcm enzymes are promiscuous. This opens the possibility to generate novel bicyclomycin derivatives in vivo as well as in vitro by combinatorial biosynthetic approaches.

Concerning the regulation of bicyclomycin production, no putative regulatory gene is located in the vicinity of the BGC. However, bicyclomycin production is regulated, as indicated by its detection in only one out of ten growth media tested. Moreover, heterologous expression of the pathway with the native promoters (encoded by pJWe14) was not observed in S. coelicolor or S. lividans, consistent with previous attempts 12. Only when we introduced pJWe14 into a mutant strain of S. cinnamoneus lacking the cluster, was bicyclomycin production observed. In contrast to attempts to express the gene cluster using the native nucleotide sequence, the addition of the strong constitutive promoter ermEp upstream of bcmA did afford bicyclomycin production in Streptomyces 31. All these observations suggest that a regulatory system, present in S. cinnamoneus and missing in the strains tested for heterologous expression, is required for the expression of the bcm biosynthetic genes from their native promoter(s).

BGCs homologous to the bcm cluster are present in Actinobacteria or Proteobacteria 11,13 and Supplementary Fig. S22. One of these homologous BGC from P. aeruginosa SCV026 has been shown to direct the biosynthesis of bicyclomycin. There are no putative regulators near the bcm cluster of S. cinnamoneus and no regulatory processes are known for this pathway or bicyclomycin production. More generally, a regulatory gene for CDPS-encoding BGCs has only been identified for pulcherriminic acid. This regulator is a member of the MarR family and is encoded by a gene that is adjacent to the biosynthetic genes in Bacillus subtilis while its homologue in Bacillus licheniformis is not encoded in the vicinity of the gene cluster 23–25.

The initial assignment of bcmH as a resistance gene is derived from overexpression experiments performed in E. coli 41. However, our results indicate that BcmH, a member of the Major Facilitator Superfamily, is not required for bicyclomycin export. Specifically, we showed that bcmH is dispensable in a strain producing bicyclomycin, and that it is not always transcribed at the same time as the rest of the bcm genes. Indeed, heterologous expression of the bcm cluster from S. cinnamoneus lacking bcmH in S. coelicolor was recently achieved. 12. An orthologue of bcmH is not universally conserved within or near BGCs predicted for bicyclomycin in other Actinobacteria. For instance, no bcmH orthologue is present near the orthologues of bcmA-bcmG in Streptomyces ossamyceticus 46 and Supplementary Fig. S22. Other examples of transporter-encoding gene associated with specialized metabolite biosynthetic genes, but dispensable for export have already been described. For instance, in Streptomyces venezuelae, the jadL gene associated with the jadomycin BGC is dispensable for efficient jadomycin export 47. In Streptomyces spp., a number of bcmH homologues exist, including some whose products resemble the MFS exporter isolated from a spontaneous E. coli mutant with resistance to bicyclomycin 47. For example, the MFS encoded by the S. cinnamoneus BLA24_RS22250, which is not associated with the bcm BGC, shares 33% amino acid sequence identity with the bicyclomycin resistance protein recovered from E. coli. In contrast, BcmH and the E. coli transporter share only 20% identity. Therefore, the export of bicyclomycin in the producing organism could be due to redundant export mechanisms, one of which might involve BcmH.

It should be noted that bicyclomycin is not active against Streptomyces and most other actinobacteria 28,29 (with the exception of Micrococcus luteus 48) even though the ATPase activity of Rho is sensitive in vitro as demonstrated in the case of S. lividans 31. Rho from S. cinnamoneus shares 78% protein sequence identity to that of S. lividans and all the positions predicted to be involved in the interaction with bicyclomycin 12 are conserved between both proteins (Supplementary Fig. S23). In addition to this genus-level similarity, the Streptomyces proteins are identical to E. coli wild-type Rho at 18 of the 24 positions whose amino acid substitution confers resistance to bicyclomycin 13. The remaining six variable positions include relatively conservative amino acid substitutions. Moreover at these six positions, the residues are identical in S. lividans and S. cinnamoneus. Based on the sequence similarities, we assume that the Rho from S. cinnamoneus is also sensitive to bicyclomycin. The fact that bicyclomycin production is not detrimental to the Streptomyces producing strain can be explained by an efficient export of the antibiotic but also by the fact that Rho might be non-essential in Streptomyces, as for instance in B. subtilis 49.

Additional studies are required to understand the regulation of bicyclomycin biosynthesis, export, and resistance in the producing organisms. The bicyclomycin pathway is the most complex CDPS-dependent DKP biosynthetic pathway characterized so far, with six tailoring enzymes. The characterization of this pathway and the further study of its enzyme promiscuity may provide insight for the biosynthesis of bicyclomycin analogues and also for the use of these enzymes for the modification of other cyclodeptides in combinatorial biosynthesis.

Material and Methods

Bacterial strains, culture conditions and strain manipulation. Bacterial strains are listed in Supplementary Table S3. Escherichia coli strains were manipulated using standard laboratory procedures 85,86. When needed, the following compounds were added to E. coli cultures at the following concentrations: ampicillin, 100 mg/l; apramycin, 50 (liquid media) or 75 (solid media) mg/l; chloramphenicol, 30 mg/l; kanamycin, 25 mg/l; hygromycin, 50 (liquid media) or 100 (solid media) mg/l; spectinomycin, 100 mg/l; isopropyl β-D-1-thiogalactopyranoside (IPTG), 1 mM. S. cinnamoneus was manipulated using standard laboratory procedures 97. Spores were prepared on Gruau medium (wheat groats, 30 g/l; glucose, 5 g/l; yeast extract, 5 g/l; pH adjusted to 7.2

Material and Methods

Bacterial strains, culture conditions and strain manipulation. Bacterial strains are listed in Supplementary Table S3. Escherichia coli strains were manipulated using standard laboratory procedures 85,86. When needed, the following compounds were added to E. coli cultures at the following concentrations: ampicillin, 100 mg/l; apramycin, 50 (liquid media) or 75 (solid media) mg/l; chloramphenicol, 30 mg/l; kanamycin, 25 mg/l; hygromycin, 50 (liquid media) or 100 (solid media) mg/l; spectinomycin, 100 mg/l; isopropyl β-D-1-thiogalactopyranoside (IPTG), 1 mM. S. cinnamoneus was manipulated using standard laboratory procedures 97. Spores were prepared on Gruau medium (wheat groats, 30 g/l; glucose, 5 g/l; yeast extract, 5 g/l; pH adjusted to 7.2
with NaOH; agar, 20 g/l) (Mellouli, personal communication). Conjugations between E. coli ET12567pUZ8002 and S. cinnamoneus were carried out by the modified Kieser et al. protocol17, using MS medium complemented with 10 mM MgCl₂ and a 3 ml antibiotic overlay of SNA medium containing fosfomycin instead of nalidixic acid. Exconjugants were patched on MS containing the selective antibiotic(s), fosfomycin and ampicillin. Antibiotic concentrations used for Streptomyces were as follow: ampicillin, 30 mg/l; apramycin, 30 mg/l; fosfomycin, 50 mg/l; kanamycin, 25 mg/l; hygromycin, 50 mg/l.

Plasmids and DNA manipulations. All plasmids used in this study are listed in Supplementary Table S4. PCR reactions for cloning applications were carried out using Phusion (Thermo Fisher Scientific) or Q5 (New England Biolabs) high fidelity polymerases according to manufacturers’ protocol for GC-rich DNA, reaction mixtures being supplemented with 1 M of betaine. PCR reactions for the control of constructions and during RT-PCR were carried out using the DreamTaq DNA Polymerase (Thermo Fisher Scientific) supplemented with 1x Q Solution (Qiagen). All primers are listed in Supplementary Table S5. Primers were provided either by Integrated DNA Technologies or Eurofins. PCR products were purified using the NucleoSpin Gel and PCR Clean-up kit (Macherey-Nagel). Plasmid isolation from E. coli was performed using the NucleoSpin Plasmid kit (Macherey-Nagel). Restriction enzymes and DNA-modifying enzymes were supplied by Thermo Fisher Scientific. Vector DNA was systematically dephosphorylated before ligation. Digestion products were isolated on agarose gels and purified with the NucleoSpin Gel and PCR Clean-up kit (Macherey-Nagel). Ligation reactions were performed using T4 DNA Ligase (either Thermo Fisher Scientific or Promega). Chemocompetent E. coli DH5α cells were used for transformation by ligation products or plasmids unless stated otherwise. Streptomyces genomic DNA was extracted as previously described18, with the exception of DNA used for genome sequencing. DNA isolation for the sequencing of the S. cinnamoneus ATCC 21532 genome was carried out as previously described19. The sequence obtained for the bcm gene cluster and its flanking regions were identical to those already deposited in DDBJ/ENA/GenBank11,13.

Bioinformatic analyses. In silico analyses of sequences and primer design were made using the Clone Manager 9 Professional Edition Software (Scientific & Educational Software). Other analyses were performed using BLAST, run locally, or on the NCBI web server and AntiSMASH 239. Multiple sequence alignments were done with Multalin40.

Deletion of the whole bcm cluster. The entire bicyclomycin biosynthetic gene cluster was replaced in a double homologous recombination event with a kanamycin resistance cassette. Details of the procedure are given in Supplementary Information.

Plasmid construction. Briefly, for the in vivo characterization of the bicyclomycin BGC, the bcmA gene with its native promoter and the whole bicyclomycin BGC were cloned into a pRT801 derivative, yielding respectively pjWe20 and pjWe14 vectors. The plasmid pjWe14 was used to generate derivatives with individual in-frame deletions for each of the bcm genes, yielding plasmids pMC2, pMC4, pMC6, pMC8, pMC12, pMC14 and pMC16 where the bcmB, bcmC, bcmD, bcmE, bcmF, bcmG, bcmA or bcmH gene were deleted, respectively. For the bcmH mutant complementation, the bcmH gene was cloned with its native promoter into pOSV806 vector yielding pjWe21. The plasmids obtained were transferred to appropriate S. cinnamoneus strains by conjugation.

For the in vitro characterization of decoration enzymes, genes bcmB-G were individually cloned into the pET15b E. coli expression vector. The recombinant proteins had an N-terminal 6xHis tag.

Details of cloning procedure are provided in Supplementary Information. Constructions were verified by sequencing (Genewiz).

Chemical synthesis of cIL. Briefly, 1 g (5.5 mmoles) of H-L-Ile-OMe.HCl (Novabiochem) was dissolved in 15 ml of dimethylformamide at room temperature and 935 µl (5.5 mmoles) of disopropylamine were added. After freezing at 4°C, 1.37 g (5.5 mmole) of N-Boc-L-Leu-OH were added, followed by 1.13 g (5.5 mmole) of dicyclohexylcarbodiimide. After incubation for 1 h at 4°C, the reaction mixture was incubated overnight at room temperature, and the rest of the process was performed according to 41. 528 mg of final product (2.32 mmoles, yield 42%) was obtained, dissolved in DMSO and shown to be homogeneous by HPLC analysis using a dC18 column (Atlantis, dC18, 150 x 4.6 mm, 3 µm). The chemical structure of cIL was validated by NMR (Supplementary Information and Supplementary Figs. S9–S10).

In vitro characterization of tailoring enzymes. For the in vitro characterization of tailoring enzymes, His-tagged proteins BcmB to G were produced in the E. coli BL21-AI strain and purified by nickel affinity chromatography. Enzymatic assays were performed at 30°C in 1 ml reaction volume in the presence of 1 µM purified protein, 200 µM of diketopiperazine substrate and the necessary cofactors and co-substrates. The reaction mixtures were analysed by HPLC and ESI-MS. Reaction procedures were optimized and scaled up, and major products were purified and analysed by NMR. Details of experimental procedures are given in Supplementary Information.

Production assays in S. cinnamoneus strains. All production experiments were carried out as follows. A 10 ml pre-culture in MP5 medium20 was launched in 50 ml Falcon tube on an orbital shaker (180 rpm) at 28°C. The pre-culture was inoculated with either spores or mycelium fragments. After 48 h, 1 ml of pre-culture was used to inoculate 25 ml of MP5 medium in a 500 ml baffled Erlenmeyer flask, and the culture was incubated 7 days on
an orbital shaker (180 rpm) at 28 °C. Supernatants were subsequently analysed by HPLC or LC-MS (for details, see Supplementary Information).

**HPLC and LC-MS analysis of the S. cinnamoneus culture supernatants.** Culture supernatants obtained after centrifugation of the culture were filtered on 0.2 μm MininPrep filter (Whatman). When following “method A”, the filtered supernatants were analyzed on an Atlantis T3 column (250 mm x 4.6 mm, 5 μm, 30 °C) using an Agilent 1200 HPLC instrument equipped with a quaternary pump. Samples were eluted with iso-ocratic 0.1% HCOOH in H2O (solvent A) / 0.1% HCOOH in CH3CN (solvent B) (95:5) at 1 ml/min for 5 min followed by a gradient to 100% solvent B over 45 min. Molecules that were present in the supernatant were detected by an Alltech 3300 evaporative light scattering detector (ELSD) (Grace).

When necessary, an analysis of the cell content was made. The mycelium pellet obtained after centrifugation of 2 ml of culture, was washed twice with 2 ml of water, suspended in 400 μl of water and broken down with glass beads using the FastPrep-24 apparatus (MP Biomedicals). A cell-free extract was obtained after centrifugation (13,000 g, 4 °C, 30 min) to remove cell debris.

LC-MS analyses were performed using an Agilent 1100 HPLC coupled via split system to an Esquire HCT ion trap mass spectrometer (Bruker) set in positive and negative modes. Multiple analyses were made. Some of them were made using “method A”; some other using the conditions described by15 for CDPS 1–47 and called “method B”.

**Data availability**
All data generated and analysed during this study are included in this published article and its supplementary information files.

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Author contributions

J.W., P.B., M.G., J-L.P. and E.D. designed the research strategy. P.B.; O.L., M.G. and J-L.P. obtained the funding. J.W., M.C., I.C., P.B., A.R., C.S., L.C., E.F. and S.L. performed the experiments. J.W., I.C., A.R., O.L., J-L.P. and E.D. analysed the data. J.W., I.C., A.R., O.L., J-L.P. and E.D. wrote the initial draft. All the authors contributed to and approved the final manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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