Pancreatic ductal adenocarcinoma (PDAC) is a genomically diverse, prevalent, and almost invariably fatal malignancy. Although conventional genetically engineered mouse models of human PDAC have been instrumental in understanding pancreatic cancer development, these models are much too labor-intensive, expensive, and slow to perform the extensive molecular analyses needed to adequately understand this disease. Here we demonstrate that retrograde pancreatic ductal injection of either adenoviral-Cre or lentiviral-Cre vectors allows titratable initiation of pancreatic neoplasias that progress into invasive and metastatic PDAC. To enable in vivo CRISPR/Cas9-mediated gene inactivation in the pancreas, we generated a Cre-regulated Cas9 allele and lentiviral vectors that express Cre and a single-guide RNA. CRISPR-mediated targeting of Lkb1 in combination with oncogenic Kras expression led to selection for inactivating genomic alterations, absence of Lkb1 protein, and rapid tumor growth that phenocopied Cre-mediated genetic deletion of Lkb1. This method will transform our ability to rapidly interrogate gene function during the development of this recalcitrant cancer.

[Keywords: CRISPR; genome editing; mouse model; pancreatic cancer]

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Pancreatic ductal adenocarcinoma (PDAC) is an almost uniformly lethal tumor type that is projected to become the second leading cancer killer in the United States by 2030 [Rahib et al. 2014]. PDAC patients have a 5-year survival rate of ~5%, underscoring the need for novel approaches to accelerate the molecular characterization of this disease. Although high-prevalence mutations have been identified in pancreatic cancer, these tumors also
incure low-frequency mutations and genomic alterations, interact with their extensive and complex stromal environment, and undergo poorly characterized changes in their expression programs (Biankin et al. 2012; Waddell et al. 2015). Despite the potential importance of these molecular and cellular changes, very little is known about how these alterations contribute to the development of metastatic and therapy-refractory PDAC. Given the inability to test gene function in human cancers in vivo, genetically engineered mouse models represent tractable and biologically relevant systems with which to interrogate the molecular determinants of each stage of pancreatic cancer development.

Identification of the mutations that drive the development of human pancreatic cancer combined with the ability to alter gene function in mice has enabled the development of genetically engineered murine PDAC models. Transgenic expression of Cre-recombinase in pancreatic cells ofloxP-Stop-loxP [LSL]KrasG12D knock-in mice [KrasLSL-G12D/+] results in deletion of the transcriptional/translational Stop element, expression of oncogenic KrasG12D, and development of lesions that closely resemble early stage human pancreatic intraepithelial neoplasms (PanINs) (Hingorani et al. 2003). Concomitant expression of a point mutant p53 allele, deletion of p53, deletion of Cdkn2a, and/or deletion of Smad4 allow[s] for the development of invasive and metastatic PDAC (Aguirre et al. 2003; Hingorani et al. 2005; Bardeesy et al. 2006a,b; Gidekel Friedlander et al. 2009; Whittle et al. 2015). These in vivo models have been instrumental in our understanding of the genetic determinants of cancer progression as well as the functional interactions of neoplastic cells with the immune system and stromal environment. However, using conventional genetically engineered autotransplanted mouse models of PDAC to interrogate gene function is complicated by inherent practical and biological limitations of these systems. Existing mouse models typically fail to model the adult onset of pancreatic cancer and induce genomic alteration in nearly every cell in the pancreas. In the decade since the first genetically engineered PDAC models were developed, few technical advances have been made, and generating the mice required to investigate a gene of interest in the established PDAC models remains a time-consuming and costly endeavor (Aguirre et al. 2003; Hingorani et al. 2003, 2005; Saborowski et al. 2014).

Systems that enable in vivo functional interrogation of genes in pancreatic cancer without the financial and temporal cost of generating new mouse alleles and incorporating them into increasingly complex mouse models could have an extremely broad impact on pancreatic cancer research. To functionally investigate the molecular changes that drive each step of pancreatic cancer development, it would be desirable to have a system in which the timing of tumor initiation and the number of lesions that form in the adult pancreas can be controlled, the number of germline-encoded alleles is minimized, and genes of interest can be eliminated without having to generate a conditional allele and breed it into a complex genetically engineered mouse model.

Here we describe methods for the direct delivery of viral vectors to the pancreas and transgenic mouse lines to allow CRISPR/Cas9-mediated genomic alterations in pancreatic cells in vivo. These systems allow titratable initiation of pancreatic tumors in adult mice and functional interrogation of candidate genes in pancreatic cancer in vivo.

Results

Retrograde pancreatic ductal injection of adenoviral-Cre (Adeno-Cre) leads to infection of pancreatic cells

Current genetically engineered mouse models of human PDAC are limited by their complete reliance on germline conditional and transgenic Cre alleles that lead to widespread recombination in the majority of the epithelial cells, most often within the embryonic pancreas. Since viral-Cre infection of the lung, prostate, bladder, and muscle can initiate cancer in adult tissues (Jackson et al. 2001; Kirsch et al. 2007; Puzio-Kuter et al. 2009, Cho et al. 2014), we considered how best to deliver viral-Cre to the pancreas without infecting other cells within the peritoneal cavity. To specifically initiate recombination in cells within the adult pancreas while keeping the virus contained within the target organ, we used retrograde pancreatic ductal injection. In this procedure, the virus is directly injected into the pancreas through the common bile duct that drains the biliary and pancreatic fluid into the duodenum (Fig. 1A,B; Supplemental Movie 1; Taniguchi et al. 2003; Wang et al. 2006). Injection of adenoviral-CMV-Cre [Ad-Cre] into Cre reporter mice (R26LSL-Tom+) [Madisen et al. 2010] led to sporadic labeling of mostly acinar cells within the pancreas, with no detectable infection of cells within the liver, intestine, or any other organs outside the pancreas (Fig. 1C; Supplemental Fig. 1A–C; data not shown).

Retrograde ductal injection of Adeno-Cre and lentiviral-Cre (Lenti-Cre) initiates pancreatic tumors

Retrograde ductal injection of KrasLSL-G12D+/+,R26LSL-Tom+ [K7] mice with Ad-Cre led to acinar-to-ductal metaplasia [ADM] and the formation of hundreds of Tomatopositive PanIN lesions, which expressed cytokeratin 19 [CK19] (Fig. 1D,E). These PanIN lesions had abundant apical mucin with small oval nuclei, and none of the lesions developed beyond PanIN1 over a 6-mo period (Fig. 1D–F; Supplemental Table 1; Supplemental Fig. 1E,F).

To determine whether retrograde ductal injection could initiate cancer when oncogenic KrasG12D expression was combined with loss of the p53 tumor suppressor, we injected Ad-Cre into the pancreata of KrasLSL-G12D+/−,p53lox/fox;R26LSL-Tom+ [KPT] mice (Fig. 1G–J). At early time points, KPT mice infected with Ad-Cre already harbored numerous ADMs and PanIN lesions of both early and advanced grades (Fig. 1G,H; Supplemental Fig. 2A–E). At later time points, these mice became
cachectic and jaundiced and displayed poor grooming due to the development of multifocal pancreatic tumors (Fig. 1I,J; Supplemental Fig. 2F,G; Supplemental Table 1).

KPT mice infected with Ad-Cre (1 × 10⁸ to 5 × 10⁶ IU) lived between 2 and 6 mo depending on the viral titer and developed invasive ductal adenocarcinomas similar to those observed in the conventional transgenic-Cre-based models (Fig. 1I,J; Supplemental Fig. 2F,H; Bardeesy et al. 2006a). Tomatopositive, CK19positive ductal adenocarcinomas were characterized by the formation of glandular structures. These tumors often had extensive, collagen-rich extracellular matrix deposition and abundant, Tomato-negative desmoplastic stromal responses (Fig. 1J; Supplemental Fig. 2F).

Ad-Cre infection of mice with a conditional point mutant p53 allele (Kras<sup>LSL-G12D</sup>/p53<sup>LSL-R172H/fox</sup>) also initiated PDAC with overall similar histological features [Supplemental Fig. 2I]. Finally, neither Ad-Cre infection of p53<sup>flox/flox</sup>; R26<sup>LSL-Tom</sup> mice, which lack the oncogenic Kras allele, nor control adenovirus (adenoviral-CMV-FLPo [Ad-FLPo]) infection of KPT mice resulted in any pancreatic lesions (Supplemental Table 1; Supplemental Fig. 2G; data not shown).

To specifically target acinar cells in the pancreas, we generated an adenoviral vector that contains a 2.3-kb acinar cell-specific Ptf1a enhancer and Ela1 minimal promoter driving Cre [Ad-Ptf1a-Cre]. Retrograde ductal injection of R26<sup>LSL-Tom</sup> mice with Ad-Ptf1a-Cre demonstrated the specificity of this promoter for acinar cells [Supplemental Fig. 3A–D]. KPT mice infected with Ad-Ptf1a-Cre developed PanINs and invasive metastatic PDAC, consistent with our results using Ad-CMV-Cre and consistent with acinar cells within the adult pancreas being...
The use of lentiviral vectors to initiate pancreatic cancer would offer a modular system to rapidly alter genes in developing tumors due to their stable genomic integration. Retrograde ductal injection of Lenti-Cre into \textit{R26LSL-Tom}^+ mice led to labeling of mostly acinar cells (Supplemental Fig. 4A–C). KT mice infected with Lenti-Cre developed exclusively low-grade PanINs (Supplemental Fig. 4D). KPT mice infected with Lenti-Cre developed PanINs and PDAC 2–6 mo after tumor initiation (Fig. 1K–N; Supplemental Fig. 2J,K). Viral-Cre-initiated pancreatic tumors ranged from well differentiated to undifferentiated, and the spectrum of differentiation within primary tumors and metastases was similar for Adeno-Cre and Lenti-Cre (Fig. 1C,D,F,J; Supplemental Table 1; data not shown). In many cases, Ad-Cre-induced and Lenti-Cre-induced tumors developed in the head of the pancreas and were accompanied by evidence of gallbladder obstruction and jaundice, a frequent end-stage complication of pancreatic cancer in patients (Supplemental Table 1; Supplemental Fig. 2G; Kruse 2010; Boulay and Parepally 2014).

Chronic pancreatitis is associated with an increased incidence of PDAC in humans, and experimental evidence from mouse models suggests that adult cells may be refractory to transformation in the absence of pancreatic inflammation [Guerra et al. 2007; Raimondi et al. 2010]. Therefore, we assessed whether retrograde ductal viral infection induces pancreatitis. Both Ad-Cre and Lenti-Cre infection induced acute pancreatitis with morphological evidence of ADM and focal replacement of acinar cells by infiltrating mononuclear cells [Supplemental Fig. 3E; Supplemental Fig. 4D]. The induction of pancreatitis is consistent with the efficient tumor initiation observed in adult animals following retrograde ductal injection of viral-Cre.

\textbf{Use of virally induced PDAC to investigate the kinetics and genetics of metastatic progression}

A key attribute of human pancreatic cancer is its invasive growth and propensity for widespread metastasis. We used the bright Tomato reporter to mark neoplastic cells in virally induced tumors, thereby allowing for quantification of disseminated tumor cells (DTCs) in the peritoneal cavity and micrometastases and macrometastases in KT and KPT mice (Fig. 2A–G). Tomato\textsuperscript{positive} DTCs could also be easily identified in the peritoneal cavity of some late time point KPT mice (Fig. 2A,D,G). KPT mice developed widespread metastases in the liver, diaphragm, lung, and mesenteric lymph nodes, all of which are prevalent sites for human PDAC metastasis [Fig. 2B–G, Supplemental Fig. 5; Supplemental Table 1; Yachida and Iacobuzio-Donahue 2009]. Despite the high total tumor burden in most Ad-Cre-infected and Lenti-Cre-infected KPT mice, the number of DTCs in the peritoneal cavity was quite variable, most consistent with the notion that the primary tumors must progress to a cellular state capable of dissemination (Fig. 2D,G). The absence of DTCs and metastases in KT mice was consistent with the presence of only homogeneously low-grade PanIN lesions and suggests that other engineered tumor suppressor alterations or spontaneous cell state alterations are a prerequisite for neoplastic cell dissemination.

\textbf{Figure 2.} Diverse expansion potential and metastatic ability of viral-Cre-initiated PDAC. \textit{[A]} DTCs can be detected in the peritoneal cavity of Ad-Cre-infected KPT mice. Viable [DAPI\textsuperscript{negative}, lineage-negative [CD31, CD45, Ter119, F4/80]negative cells are shown. \textit{[B,C]} Retrograde ductal Ad-Cre tumor initiation in \textit{KPT} mice leads to widespread metastasis. A dotted outline of liver (Liv) lobes is shown in \textit{B}. (C) Representative histology of liver metastasis. Bars: B, 3 mm; C, 100 mm. \textit{[D]} Quantification of DTCs and metastases in Ad-Cre-infected \textit{KPT} mice. Data represent the number of mice with the indicated lesion/total number of mice of that genotype. For DTCs, the range of the total DTC number is shown in parentheses. \textit{PerC} indicates metastases in the peritoneal cavity. \textit{[E,F]} KPT mice with Lenti-Cre-induced PDAC can develop multifocal, multigorgan metastatic disease. (Liv) Liver. Bars: E, 2 mm; F, 100 mm. \textit{[G]} Quantification of DTCs and metastases in Lenti-Cre-infected \textit{KPT} and \textit{KPT} mice. Data are represented as in \textit{D}. \textit{[H–K]} Injection of Ad-Cre (\textit{HJ}) or Lenti-Cre (\textit{KL}) into \textit{KrasLSL-G12D}\textsuperscript{+/−}\textit{p53flox/flox}\textit{R26Mot/Mot} [\textit{KP, R26Mot/Mot}]; \textit{[KP, R26Mot/Mot]} mice initiate diverse color labeling of early lesions (\textit{HJ}), few of which progress into tumors (\textit{J}). \textit{[K]} A single, clonally derived, late time point primary tumor (\textit{L}) seeded multiple liver metastases (\textit{K}). (Duodenum; Liv liver. Bars, 3 mm.)
Identifying the clonal origin of PDACs and determining tumor–metastasis relationships

Conventional transgenic Cre-driven and Cre[ER]-driven PDAC models lack the ability to distinguish between a tumor mass derived from a single transformed cell and a mass of collision tumors. To assess the clonality of viral Cre-initiated tumors, we recently generated a multicolor Cre reporter in which recombination leads to the expression of RFP, CFP, or YFP (Caswell et al. 2014). Retrograde ductal Ad-Cre infection of R26<sup>Moter/Moter</sup> homozygous mice generated cells with each of the six color combinations (Supplemental Fig. 6A). Infection of Kras<sup>LSL-G12D</sup>/p53<sup>Box/Box</sup>R26<sup>Moter/Moter</sup> (KP,R26<sup>Moter/Moter</sup>) mice generated large numbers of ADMs and PanINs, a relatively small number of which progressed into individual tumors within the pancreatic cancer mass (Fig. 2H,I; Supplemental Fig. 6B–E). Both Ad-Cre-infected and Lenti-Cre-infected KP,R26<sup>Moter/Moter</sup> mice developed clonally derived monochromatic tumors (Fig. 2J; Supplemental Fig. 6F,G). Some viral-Cre-infected KP,R26<sup>Moter/Moter</sup> mice progressed to have one dominant tumor, and, whenever metastases were present, they were the same color as the largest primary PDAC (Fig. 2J,K, Supplemental Fig. 6E–G).

Generation and characterization of Cre-regulated Cas9 mice and lentiviral vectors for single-guide RNA (sgRNA) and Cre expression

The ability to induce targeted genomic deletions in pancreatic cancer in vivo would facilitate the investigation of all aspects of pancreatic tumorigenesis. To enable in vivo somatic genomic editing in mouse models of human cancer, we generated single insertion transgenic mice through site-directed integration of a LSL-Cas9 cassette into the H11 locus (H11<sup>LSL-Cas9</sup>) (Fig. 3A,B; Supplenentary Fig. 7A; Tasic et al. 2011). Expression of Cre in fibroblasts from H11<sup>LSL-Cas9</sup> mice induced recombination of the Stop cassette and led to expression of Cas9 RNA and protein (Fig. 3C,D; data not shown). To characterize the potential for Cas9 expression from the H11<sup>LSL-Cas9</sup> allele in vivo, we intercrossed H11<sup>LSL-Cas9</sup> and CMV-Cre deleter mice to generate an H11<sup>LSL-Cas9</sup> allele. H11<sup>LSL-Cas9</sup> mice had widespread expression of Cas9 across all organs tested (Supplemental Fig. 8).

To deliver a synthetic sgRNA and Cre into pancreatic cells, we generated dual promoter lentiviral vectors that contain a U6-driven sgRNA and also express Cre (Supplemental Fig. 7B). Given the frequent down-regulation and genomic loss of Lkb1/STK11 in human PDAC (Morton et al. 2010; Waddell et al. 2015), the predisposition of Peutz-Jeghers syndrome patients to develop pancreatic tumors (Su et al. 1999), and the impact of Lkb1 inactivation in other pancreatic tumor models (Hezel et al. 2008; Morton et al. 2010; Lo et al. 2012), we chose Lkb1 as an initial target for CRISPR/Cas9-mediated somatic alteration in the adult pancreas. To assess the impact of Lkb1 loss on pancreatic cancer initiation and growth, we generated and screened Lenti-sgRNA/Cre vectors targeting Lkb1 [Supplemental Fig. 7B–D]. Infection of H11<sup>LSL-Cas9</sup>/m<sup>Cre</sup> fibroblasts with Lenti-sgLkb1/Cre led to the formation of insertions and deletions (indels) in Lkb1 at the targeted location, thus validating both our lentiviral sgRNA vector and the H11<sup>LSL-Cas9</sup> transgenic allele (Fig. 3E; Supplemental Fig. 7E).

Somatic genome engineering enables rapid generation of genetically defined pancreatic cancer mouse models

To assess the impact of Lkb1 deletion on pancreatic cancer, we performed retrograde ductal injections of KT;H11<sup>LSL-Cas9</sup>/, control KT, and KT;Lkb1<sup>flex/flox</sup> mice with Lenti-sgLkb1/Cre as well as KT;H11<sup>LSL-Cas9</sup>/mice with lentiviral vectors containing negative control sgRNAs. Control KT mice infected with Lenti-sgLkb1/Cre as well as KT;H11<sup>LSL-Cas9</sup>/mice infected with a Lenti-sgRNA/Cre vector containing either a nontargeting sgRNA (sgNT) or an sgRNA targeting an inert region of the genome [sgNeo] formed only rare Tomato<sup>positive</sup> lesions (Fig. 3F–H). Both Lenti-sgLkb1/Care-infected KT;H11<sup>LSL-Cas9</sup>/ and KT;Lkb1<sup>flex/flox</sup> mice had extensive tumor growth as early as 2 mo after tumor initiation (Fig. 3I,J, respectively).

All three groups of negative control mice developed only rare PanIN lesions within almost completely normal pancreata (Fig. 4A–C; Supplemental Fig. 9A; data not shown). The tumors that formed in Lenti-sgLkb1/Cre-infected KT;H11<sup>LSL-Cas9</sup>/ mice were cystic lesions comprised of Tomato<sup>positive</sup>, CK19<sup>positive</sup> tall cuboidal to columnar epithelial cells with otherwise bland cytological features (Fig. 4D; Supplemental Fig. 9B). Some smaller lesions also contained high levels of mucin (Supplemental Fig. 9B). Importantly, these features were histologically indistinguishable from those found in KT;Lkb1<sup>flex/flox</sup> mice infected with Lenti-sgLkb1/Cre (Fig. 4E; Supplemental Fig. 9C). Lenti-sgLkb1/Cre-infected KT;H11<sup>LSL-Cas9</sup>/ mice also had a substantially higher tumor burden when compared with all three groups of negative control mice (Fig. 4F).

FACS-sorted Tomato<sup>positive</sup> cells from Lenti-sgLkb1/Cre-infected KT;H11<sup>LSL-Cas9</sup>/mice had recombinated the H11<sup>LSL-Cas9</sup> allele and expressed Cas9 RNA and protein (Supplemental Fig. 9D,E; data not shown). Indels at the targeted Lkb1 locus were specifically detected in the neoplastic cells from these tumors (Fig. 5A,B, Supplemental Fig. 9F). The indels were all frameshift mutations or large exon deletions that included the splice acceptor or donor regions of Lkb1 exon 6, further supporting the strong selective advantage of Lkb1 inactivation (Fig. 5B, C). Consistent with the presence of CRISPR/Cas9-induced frameshift mutations and large deletions, Lkb1 protein was absent from most neoplastic cells in the Lenti-sgLkb1/Cre-induced tumors in KT;H11<sup>LSL-Cas9</sup>/ mice (Fig. 5D,E; Supplemental Fig. 9G).

Discussion

The development of systems that accelerate our ability to investigate pancreatic carcinogenesis at the molecular
level will be a critical step toward overcoming the dismal rate of successful treatment and low survival rate of patients with this recalcitrant cancer. The ability to induce pancreatic cancer using viral vectors will be instrumental in understanding the mechanisms that sustain tumor growth, lead to metastatic spread, and drive drug resistance. Our CRISPR/Cas9-based model should allow any gene of interest to be inactivated in pancreatic cancer in vivo without the need to generate any new mouse alleles. The method developed in this study could have a profound impact on both basic and translational pancreatic cancer research. Collectively, these methods will enable a more rapid and complete understanding of the molecular regulators of all aspects of pancreatic tumorigenesis and complement the strengths of existing genetically engineered models, human patient-derived xenograft models, and studies on human and murine cell lines.

Retrograde ductal injection of Adeno-Cre and Lenti-Cre vectors allows titratable pancreatic tumor initiation in the adult pancreas. This removes the requirement for the transgenic Cre[ER] alleles used in conventional genetically engineered pancreatic cancer models and enables sparse rather than widespread expression of oncogenic Kras and deletion of tumor suppressor genes, more closely recapitulating the initiating events in human PDAC. The inclusion of a fluorescent Cre reporter in transgenic Cre[ER]-induced PDAC models leads to the fluorescent labeling of not only the neoplastic cells but also most of the nontransformed pancreatic epithelial cells, making unequivocal distinction of neoplastic from normal cells difficult.

The ability to induce PDAC in Cre-lox models without having to include a transgenic Cre line will make the molecular investigation of pancreatic cancer more rapid and less expensive (Supplemental Table 2). Additionally, with the ease of generating pancreatic tumors now approaching that of lung tumors (via intranasal or intratracheal injection), comparing the impact of the same genetic alterations on each cancer type should become easier.
standard practice and will uncover the commonalities and differences between these lethal adenocarcinomas.

PDAC models often generate tumor masses of unknown clonal origin. By incorporating a multicolor fluorescent reporter, we were able to mark individual clonal lesions and identify the relationship between primary tumors and metastases. Unexpectedly, some late time point mice had only one or two large cancers, underscoring the dramatic heterogeneity in expansion potential of pancreatic lesions initiated with identical engineered genetic events. Clonal lineage tracing using diverse fluorescent labeling or lentivirally mediated nucleotide barcoding (Caswell et al. 2014) may facilitate experiments aimed at understanding early steps in the metastatic process and enable the identification of primary tumors that have and have not gained the ability to seed metastases.

The identity of the cell types capable of giving rise to pancreatic cancer is an area of active investigation and considerable debate (Guerra et al. 2007; Habbe et al. 2008; Gidekel Friedlander et al. 2009; Kopp et al. 2012; Puri et al. 2015). The Adeno-Cre and Lenti-Cre that we used in this study mostly had ubiquitous promoters (CMV and PGK, respectively), which led to recombination in acinar, ductal, and islet cells in the pancreas, although Lenti-
Cre had a very strong preference for acinar cells [Supplemental Fig. 4B]. While a vast majority of pancreatic cancer studies have used non-cell type-specific promoters to drive Cre, models using transgenic and knock-in Cre(ER) alleles to enable cell type-specific expression of oncogenic Kras and tumor suppressor loss have been used to identify cells that possess the ability to give rise to PDAC [Guerra et al. 2007; Habbe et al. 2008; Gidekel Friedlander et al. 2009; Kopp et al. 2012; Puri et al. 2015]. However, comprehensive studies that target diverse genomic alterations to a greater array of defined cell types are simply not feasible using current transgene-based systems. The use of adenoviral vectors with cell type-specific promoters has greatly accelerated the investigation of the cell of origin of lung adenocarcinoma and small cell lung cancer [Sutherland et al. 2011, 2014], suggesting that similar approaches could rapidly advance our understanding of the precise cell type–genotype combinations that can give rise to PDAC.

Soft tissue sarcomas can be generated in KrasLSL-G12D; p53flox mice by viral-Cre infection of cells within the muscle [Kirsch et al. 2007], therefore, we initially suspected that this would be a major impediment for generating viral-induced PDAC models. In fact, although retrograde ductal injection of Ad-Cre did induce extrapancreatic sarcomas in over half of the KPT mice, these were rarely problematic. PDAC initiation in KPT mice with an adenoviral vector containing the pancreatic-specific Ptf1a enhancer driving Cre expression solved this issue. We also found that many fewer Lenti-Cre-injected mice developed sarcomas, consistent with the limited ability of lentiviral vectors to induce sarcomas [Supplemental Table 1; DuPage et al. 2012]. We envision that retrograde pancreatic ductal injection could become a ubiquitous technique for the generation of PDAC in adult mice. Importantly, the post-doctoral fellow who performed these surgeries (S.-H. Chiou) had no prior surgical experience and quickly mastered this technique. We experienced ~10% mortality within the first week after surgery (19 out of 173 mice) and achieved nearly 100% successful injection as determined by the presence of numerous Tomato-positive cells in the pancreata of infected mice.

Integrating the CRISPR/Cas9 system into the pancreatic cancer biologists’ toolkit via our viral and transgenic approaches should make loss-of-function experiments in pancreatic cancer in vivo no more difficult than altering those genes in vitro. This system should enable the rapid functional investigation of any gene of interest and allow genetic screens in pancreatic cancer in vivo. Investigating panels of putative tumor suppressor genes, factors that influence malignant progression, genes involved in cancer cell–stromal cell interactions, and genes that may modulate therapy responses could all be facilitated by this system. Conceptually, sgRNA-directed Cas9 cutting should also enable the rapid generation of structural genomic alterations observed in PDAC patients, and the ability to encode multiple sgRNAs on viral vectors will allow multiplexed gene inactivation in pancreatic cancer models [Maddalo et al. 2014; Platt et al. 2014].

Given the relative simplicity of generating pancreatic neoplasms and pancreatic cancer with these systems as well as the rapid and simple nature with which loss-of-function models can be generated using in vivo CRISPR/Cas9 genome editing in the pancreas, we believe that this virally based PDAC model will provide new avenues to further understand the mechanistic basis of many aspects of pancreatic carcinogenesis.

Materials and methods

Mice

KrasLSL-G12D, p53flox p53LSL-R172H, Lkb1flox, Rosa26Molety, CMVCre, and Rosa26LSL-Tomato (ai9), mice have been described (Schwenk et al. 1995; Jackson et al. 2001; Jonkers et al. 2001; Olive et al. 2004; Madisen et al. 2010; Nakada et al. 2010; Caswell et al. 2014). H11LSL-Cas9 site-specific knock-in mice [The Jackson Laboratory, stock no. 026816] were generated by microneural injection of a targeting plasmid and capped 4C31 mRNA into C57BL/6 H11attP3a/P3xzygotes [Tasic et al. 2011] followed by implantation into pseudopregnant CD1 recipient mice. H11Cas9 mice with constitutive Cas9 expression [The Jackson Laboratory, stock no. 027650] were generated by crossing H11LSL-Cas9 mice to CMV-Cre mice [The Jackson Laboratory, stock no. 006054]. The Stanford Institutional Animal Care and Use Committees approved all animal studies and procedures.

Tumor initiation and quantification

Tumors were initiated by retrograde pancreatic ductal injection of mice with viral vectors as previously described [Supplemental Movie 1; Taniguchi et al. 2003]. Briefly, laparotomy was performed on anesthetized mice, and half of the total bowel mass was gently pulled out of the abdomen, exposing the duodenum. A microclip was placed on the cystic duct close to the gallbladder, and a 30-gauge needle was inserted through the sphincter of Oddi into the common bile duct. One-hundred-fifty microliters of viral vector was injected over the course of 1–2 min. Subsequently, the microclip was removed, the duodenum and bowels were placed back into the abdomen, the muscle layer was sewn shut, and the skin was stapled with sterile suture clips. Once proficient, the entire procedure takes <20 min per mouse.

Generation of lentiviral vectors

Ad-Cre and Ad-FLPo were purchased from the University of Iowa Gene Transfer Core. Lenti-PGK-Cre and Lenti-U6-sgRNA/PGK-Cre vectors were generated using standard methods and are available from Addgene [see the Supplemental Material for Addgene IDs]. To assess the efficiency of genome editing with LentisgRNA/Cre vectors, we generated LSL-YFP cells that stably express Cas9 by infecting LSL-YFP cells with an MSCV-Cas9/Puro vector followed by puromycin selection.

Histologic preparation, immunohistochemistry, and tumor analysis

Samples were fixed in 4% formalin and paraffin-embedded. Immunohistochemistry was performed on 4-μm sections with the ABC Vectastain kits [Vector Laboratories] with antibodies to Tomato [Rockland Immunochemicals, 600-401-379], Muc5a [mouse monoclonal [45M1], Abcam], CK19 [TROMA-III, Developmental Studies Hybridoma Bank], SMA [mouse monoclonal [1A4], Sigma-Aldrich], and Lkb1 [Cell Signaling, 13031P]. Sections were developed with DAB and counterstained with
hematoxylin. Hematoxylin and eosin, Masson’s trichrome, Alcian blue, and periodic acid-Schiff were performed using standard methods. All histological assessments were conducted by a board-certified pathologist (G.E. Kim).

**Western blotting**

Tail tip fibroblasts infected with viral-Cre and ex vivo tissue samples were homogenized and lysed with RIPA buffer. FACS-isolated cells were lysed with RIPA buffer. Denatured samples were run on a 4%–12% Bis-Tris gel (NuPage) and transferred onto PVDF membrane. Membranes were immunoblotted using primary antibodies against Hsp90 (BD Transduction Laboratories, 610419), Lkb1 [Cell Signaling, 13031P], actin [clone AC-15, Sigma-Aldrich, A1978], Cas9 [Novus Biologicals, NB2-36440], Flag [Sigma-Aldrich, F1804], and secondary HRP-conjugated anti-mouse (Santa Cruz Biotechnology, sc-2005S) and anti-rabbit (Santa Cruz Biotechnology, sc-2004R) antibodies.

**Assessing CRISPR/Cas9-induced indels**

The GeneArt genomic cleavage detection (GCD) assay (Life Technologies) was used to analyze CRISPR/Cas9-mediated cleavage of the Lkb1 locus following the manufacturer’s instructions. To further characterize the spectrum and frequency of Lkb1 mutations resulting from CRISPR/Cas9 targeting both in vitro and in vivo, Lkb1 amplicons were TOPO TA-cloned [Invitrogen], and ∼35 clones from each sample were Sanger-sequenced.

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