Review

c-Cbl: An Important Regulator and a Target in Angiogenesis and Tumorigenesis

Chimera L. Lyle 1, Mostafa Belghasem 2 and Vipul C. Chitalia 1,3,*

1 Department of Medicine, Boston University Medical Center, Boston, MA 02118, USA; clyle@bu.edu
2 Department of Pathology and Laboratory Medicine, Boston University Medical Center, Boston, MA 02118, USA; mbelgha@bu.edu
3 Boston Veterans Affairs Healthcare System, Boston, MA 02118, USA
* Correspondence: vichital@bu.edu; Tel.: +617-638-7330

Received: 7 April 2019; Accepted: 10 May 2019; Published: 23 May 2019

Abstract: Casitas B lineage lymphoma (c-Cbl) is a multifunctional protein with a ubiquitin E3 ligase activity capable of degrading diverse sets of proteins. Although previous work had focused mainly on c-Cbl mutations in humans with hematological malignancies, recent emerging evidence suggests a critical role of c-Cbl in angiogenesis and human solid organ tumors. The combination of its unique structure, modular function, and ability to channelize cues from a rich network of signaling cascades, empowers c-Cbl to assume a central role in these disease models. This review consolidates the structural and functional insights based on recent studies that highlight c-Cbl as a target with tantalizing therapeutic potential in various models of angiogenesis and tumorigenesis.

Keywords: angiogenesis; tumors; ubiquitination; proteasomal degradation; c-Cbl; RTK; non-RTK; VEGFR; FGFR; PDGFR; EGFR; c-Met; Wnt signaling; β-catenin; PLCγ1

1. Introduction

The formation of new blood vessels is fundamental to health and disease[1–5]. This formation can occur by two processes: vasculogenesis or angiogenesis. In vasculogenesis, new vessels are created from angioblasts or other precursor cell, which then differentiate into endothelial cells forming lumens and primordial blood vessels. In contrast, in angiogenesis preexisting vasculature is the source of new capillary formation. Angiogenesis is a dynamic process orchestrated by endothelial cells (ECs), their associated perivascular supporting cells (vascular smooth muscle cells and pericytes), and immune cells. Dysregulation of angiogenesis has been recognized as a commonality in a myriad of pathological disorders including cancer [6], peripheral artery disease (PAD) [7], diabetic retinopathy [8], rheumatoid arthritis [9], and inflammatory bowel disease (IBD), etc. [10].

Angiogenesis is regulated by a delicate and dynamic balance of growth factors, intracellular signaling pathways, and extracellular components. Several mediators are involved in this process, including basic fibroblast growth factor (BFGF) [11,12], transforming growth factor (TGF)-α [13], TGF-β [14,15], tumor necrosis factor (TNF)-α [16,17], vascular endothelial growth factor VEGF [2,18–20], β-catenin [21–23], and Phospholipase C gamma (PLCγ1) [24–27]. The growth factors secreted from a medley of different cell types including endothelial cells, smooth muscle cells, fibroblasts, platelets, and immune cells coordinate this process. Physiologic and pathologic stimuli such as chronic inflammation, injury, hypoxia, and cancer further modulate specific growth factors within the extracellular milieu to influence the angiogenesis. This review specifically describes the role of Casitas B lineage lymphoma (c-Cbl) in different models of angiogenesis.
2. c-Cbl Family of Ubiquitin E3 Ligases

Casitas B-lineage lymphoma (c-Cbl) is a member of the CBL family of proteins and is comprised of three homologues [28,29]. c-Cbl is an ubiquitously expressed mammalian gene that plays a vital role in fundamental cellular functions including cell survival, migration and proliferation. v-Cbl (viral casita B-lineage lymphoma), the first member of the Cbl family, was discovered in 1989 [30]. It is a 357 amino acid protein from the Cas NS-1 murine retrovirus, which was found to induce pre-B cell lymphomas and myelogenous leukemia in mice obtained from the Lake Casitas, California [30]. Further studies showed that CAS NS-1 originated from the ecotropic (a virus that can only infect mouse or rat cells) CAS-Br-M virus resulting from continuous recombination with a cellular oncogene and endogenous retroviral sequences [31]. The truncated virally encoded protein mutant was named v-Cbl to distinguish it from normal mouse c-Cbl (906–973 amino acid), which failed to induce the formation of tumors [28,30–32]. Another homologue of c-Cbl is Cbl-b, which is expressed in normal and malignant mammary epithelial cells, hematopoietic cell lines/tissues, as well as various normal tissue lines [33–36].

Structurally, c-Cbl has several regions that encode functionally distinct protein domains (Figure 1). An N-terminal tyrosine kinase binding (TKB) domain consisting of four helix bundles, an EF hand and a SH2 domain. It binds specifically to the receptor and non-receptor tyrosine kinases. The RING finger domain is the E3 ubiquitin ligase and mediates ubiquitination of its targets. The C terminus has a proline rich domain that represents the protein-protein interaction domain and mediates the interaction of c-Cbl with various targets. Three exposed tyrosines in this region Tyr700, Tyr731 and Tyr744, modulate the interaction of c-Cbl with downstream signaling molecules such as Akt and other proteins. The C-terminal ubiquitin associated domain (UBA) functions as the site of ubiquitin binding and also facilitates the dimerization of c-Cbl [22]. v-Cbl retains the TKB domain, but lacks the RING finger and the C-terminal domains. Studies have shown that a deletion mutation in a region close to the RING finger results in c-Cbl becoming oncogenic [37]. Cbl-b is a larger protein than c-Cbl and contains an additional 69 amino acids at the C-terminus. 70Z-c-Cbl is a naturally occurring mutation lacking a 17-amino acids region within the linker domain upstream of the RING finger. It lacks the E3 ligase activity and serves as a dominant negative for several of c-Cbl’s targets [21,38].

![Figure 1. The primary structure and domain organization of human Casitas b-lineage lymphoma family proteins. c-Cbl, Cbl-b and Cbl-c proteins contain highly conserved tyrosine kinase-binding (TKB), linker (L), RING finger (RF) and proline-rich (PR) domains. 4H: Four-helix bundle; EF: EF hand, SH2: Src-homology 2; UBA: Ubiquitin-associated domain.](image)

In addition to the adaptor function of c-Cbl, activity of c-Cbl in angiogenesis is in large part driven by its E3 ubiquitin ligase function [39,40]. Ubiquitination is a critical post translational protein modification (PTM) [41]. It regulates fundamental cellular processes and is characterized by the covalent binding of the C-terminal glycine located on the ubiquitin molecule to a lysine residue on a substrate protein. Polyubiquitinated proteins are targeted for lysosomal or proteasomal degradation [42–45].
This specificity to the ubiquitination process is imparted by E3 ubiquitin ligase that recognize the substrate and also interacts and transfers E2 ubiquitin to the substrate.

The bulk of c-Cbl’s cellular effects are linked to its RING finger and E3 ubiquitin function. Studies have shown that the ubiquitin activity of c-Cbl is regulated by Tyr371, a key tyrosine residue. Recent X-ray crystal structural and point mutagenesis studies have provided interesting insight into c-Cbl structure and functional analysis [28,46]. Dou et al solved the crystal structures of human c-CBL; a c-CBL–substrate peptide complex and a phosphorylated-Tyr371-CBL–E2–substrate peptide complex and compared them with the known structure of a c-CBL–E2–substrate peptide complex [29]. Structural and biochemical analyses showed that c-CBL adopted an autoinhibited RING conformation such that the binding surface of the RING finger exhibited reduced affinity for E2. c-Cbl ubiquitination activity was induced by phosphorylation of a linker helix region (LHR) Tyr371. Tyr371 phosphorylation induced a conformational change in the LHR domain to eliminate autoinhibition and flipped the RING domain to convert the RING domain into an enhanced E2-binding module. These results demonstrated the importance of Tyr371’s phosphorylation in regulating c-Cbl’s activity.

c-Cbl is a RING finger E3 ubiquitin ligase and targets several pro-angiogenic mediator proteins for degradation including receptor and non-receptor tyrosine kinases (c-Src, c-MET, EGFR, PDGFR, etc), mediators of Wnt signaling (β-catenin) [21,22,47,48], and VEGF signaling (PLCγ1) [25,38]. Thus, c-Cbl is able to both modulate and regulate angiogenesis. The significance of c-Cbl is underscored by the fact that endothelial cells from c-Cbl knockout mice exhibited increased cell proliferation and tube formation following stimulation with VEGF. Mice with a c-Cbl knockout showed increased evidence of physiologic angiogenesis in several models such as chorioalloantoic membrane angiogenesis [25,38], zebrafish model [22], pathologic angiogenesis in tumor-induced angiogenesis [25], and retinal neovascularization [24].

3. c-Cbl as a Major Regulator of Angiogenesis

3.1. c-Cbl and β-Catenin

Wingless/Integrated (Wnt) signaling is a critical mediator of angiogenesis [49–51]. The canonical as well as the noncanonical Wnt signaling pathways are activated by engagement of Wnt ligands to their cogent receptors SFRP and Frizzled. The central mediator of canonical Wnt signaling is β-catenin, which translocates into the nucleus of endothelial cells and induces transcription of several pro-angiogenic mediators such as VEGF-A and IL-8 [50]. In contrast, the noncanonical Wnt signaling is β-catenin independent and is critical for both the transcriptional and non-transcriptional cellular responses [52]. For example, the planar cell polarity (PCP) pathway, a non-canonical Wnt pathway, regulates cell polarity by controlling gene expression and rearrangements in the cytoskeleton via the effects of c-Jun N-terminal kinase (JNK) and Ras homologue gene family, member A [53]. The Wnt-Ca^{2+} pathway is mediated through the heterotrimeric G proteins [54] These proteins activate phospholipase C (PLC) resulting in intracellular storage Ca^{2+} release and transcription of genes responsible for cell migration and cell fate [54].

The canonical Wnt pathway is inhibited by noncanonical Wnt signaling [55,56]. β-catenin–TCF dependent transcription has been shown to be inhibited through TCD phosphorylation via CaMKII-TAK1-NLK pathway activation. Similarly, NFAT has been shown to suppress β-catenin transcription [55,57]. While several angiogenic disorders are linked to mutations in the canonical Wnt signaling pathway [49,51,58–61], disorders involving cellular polarity and motility result from disruptions in the noncanonical Wnt signaling pathway.

Under normal circumstances, canonical Wnt signaling is constitutively inhibited by the degradation of β–catenin (Wnt-off phase) [58]. Binding of the Wnt ligand to its cogent receptor stabilizes β–catenin, which undergoes nuclear translocation (Wnt-on phase). The pro-angiogenic function of Wnt signaling is largely driven by nuclear β–catenin in Wnt-on phase. Work of Chitalia et al demonstrated that c-Cbl
directly interacted and suppressed β–catenin in both Wnt-off and Wnt-on phase in the endothelial cells in different subcellular compartments [21] (Figure 2).

**Figure 2.** c-Cbl regulates angiogenesis through Wnt signaling. Constitutively Wnt signaling is suppressed in endothelial cells. In absence of Wnt ligand, β-Catenin is downregulated by in a destruction complex in the cytosol consisting of Adenomatous Polyposis Coli (APC), where β-Catenin gets phosphorylated by GSK3β. This results in β-catennin ubiquitination and proteasomal degradation by E3 ligases such as c-Cbl and β-TrCP, etc. Wnt ligand activates Frz receptors, which dissociates the destruction complex, β-catenin undergoes nuclear translocation and activates the Wnt target genes such as IL-8 and VEGF-A to induce angiogenesis. Wnt activation results in phosphorylation of c-Cbl, its nuclear translocation, wherein it ubiquitinates nuclear β-catenin to switch-off the Wnt signaling and angiogenesis.

The site of interaction of c-Cbl and β–catenin imparted this functional modularity to c-Cbl. The C-terminus of c-Cbl interacts with the central region of β–catenin containing Armadillo repeat region and targets the cytosolic β –catenin to proteasomal degradation in Wnt-off phase [21]. In Wnt-on phase, c-Cbl undergoes nuclear translocation in a manner similar to that of β–catenin, degrades nuclear β–catenin, and suppresses transcription induction of pro-angiogenic VEGF-A and IL-8 cytokines. Shivanna et al. further showed that activation of the Wnt signaling pathway promoted the phosphorylation of c-Cbl at tyrosine 731(Tyr-731) [22]. Phosphorylation of Tyr731 increased its dimerization through its UBA domain as well as the nuclear translocation of c-Cbl and also increased its interaction with the nuclear β–catenin. This event augmented the degradation of nuclear active β–catenin in the Wnt-on phase and suppressed Wnt signaling and angiogenesis. c-Cbl silenced cells exhibited augmented Wnt activity in endothelial cells and increased nuclear β–catenin, increased tube formation in an in vitro angiogenesis assay [21].

In line with the crystal structure of c-Cbl, its regulation of nuclear β–catenin requires phosphorylation of Tyr371 [48]. Inactivating mutations of Tyr371 compromised the ability of c-Cbl to target nuclear β–catenin. The c-Cbl tyrosine 371 (Y371H) mutant interacts with but fails to ubiquitinate nuclear β–catenin. The nuclear localization of the c-Cbl TyrY371H mutant contributes to its dominant
negative effect on nuclear β–catenin. A transgenic Wnt-8 zebrafish model showed c-Cbl Y371H mutant augmented Wnt/β–catenin signaling, increased Wnt target genes and angiogenesis [48]. Taken together, this body of work demonstrated that c-Cbl regulated angiogenesis through Wnt signaling.

3.2. c-Cbl and VEGF Signaling

VEGF signaling is critical for angiogenesis as well as vasculogenesis. VEGF-A stimulates VEGF receptors 1 and 2 (VEGFR-1 and VEGFR-2) on endothelial cells to initiate a diverseness of biological responses that serve to increase endothelial cell survival, migration, and differentiation [18–20,62,63]. The binding of VEGF to its receptor VEGFR-2 activates another pivotal mediator of angiogenesis: PLCγ1. PLCγ1 plays an important role in mediating cellular signaling, cell migration, and actin cytoskeleton remodeling. The importance of PLCγ1 in angiogenesis was proven by the targeted deletion of PLCγ1, which resulted in early embryonic lethality between E (embryonic day) 9.5 and E10.5 due to significantly impaired vasculogenesis and erythrogenesis. Inactivation of PLCγ1 in zebrafish is also shown to be required for the function of VEGF and arterial development [22,64,65]. Pharmacological manipulation of PLCγ1 by compound U73122 inhibited endothelial cell tube formation in vitro [25,66] and angiogenesis in vivo in CAM assay [24]. In fact, activating mutations of PLCγ1 are detected in angiosarcoma that may regulate the angiogenic phenotype of the tumors [26,37,67,68].

Biochemical and functional studies have uncovered the effects of c-Cbl and PLCγ1 on VEGF signaling [24,25,38,66,69,70]. c-Cbl interacts with PLCγ1 via its proline rich region in an non-inducible manner. However, c-Cbl interacts with VEGFR-2 via its N-terminal TKB domain with the activation of VEGF receptor signaling [38]. Specifically, the TKB domain of c-Cbl interacts with the phosphorylated VEGFR-2 at Tyr1052 and Tyr1057 [25,38]. The co-recruitment of c-Cbl and PLCγ1 to VEGFR-2 results in the formation of a complex that suppresses the tyrosine phosphorylation of PLCγ1. Although PLCγ1 undergoes ubiquitination, intriguingly it escapes proteolysis.

Since c-Cbl suppressed VEGFR-2 signaling, loss of c-Cbl activity should augment angiogenesis. Indeed, the loss of c-Cbl was found to robustly activate PLCγ1, while increasing the release of intracellular calcium [25]. Taken together, these results demonstrated c-Cbl is a suppressor protein of angiogenesis via modulation of PLCγ1 activation and VEGF signaling.

3.3. c-Cbl Regulation of VEGFR-2 through Epsin

The epsins are a family of highly conserved endocytic adaptor proteins that function in the assembly of clathrin-coated vesicles and thus play important physiological roles in cellular function, such as recognizing or binding to ubiquitinated cargo and functioning as coordinators of endocytosis by interacting with the elements of the endocytic machinery (AP2 and clathrin) [71]. The epsin protein consists of an epsin NH(2)-terminal homology (ENTH) domain (responsible for promoting phospholipid interaction), two clathrin binding sequences, multiple AP2 binding sites, and multiple EPS15 homology (EH) domain binding motifs. Epsin also contains multiple ubiquitin interacting motifs (UIMs) that bind ubiquitin moiety [72].

Epsins regulate VEGFR-2 via their ubiquitin interacting motif (UIM) and augment its degradation thereby attenuating VEGFR-2 signaling [73,74]. This degradation is accomplished through c-Cbl. Song et al. demonstrated that the binding of Epsin via its UIM to the ubiquitin moieties on activated VEGFR-2 elicited the ubiquitination of epsin by c-Cbl. This process resulted in polyubiquitination and proteasomal degradation of activated VEGFR-2 by c-Cbl [73].

3.4. c-Cbl and Other Receptor Tyrosine Kinases

c-Cbl regulates several tyrosine kinases including Tie-2, Eph and c-Src. The following section covers those tyrosine kinase receptors that play an important role in angiogenesis and are regulated by c-Cbl.
3.4.1. c-Cbl and Angiopoietin(1/2)-Tie2 Axis

The Tie2 receptor is a transmembrane tyrosine kinase highly expressed on the surface of endothelium. The ligands of Tie2 are Angiopoietin 1 and 2 and exhibit opposite functions to each other, such that their relative expression in the local milieu determine the activity of Angiopoietin(1/2)-Tie2 axis [75]. This signaling axis is considered to be an important regulator of endothelial permeability and angiogenesis [5,75–77]. The current model suggests that in the quiescent vasculature, Tie2 is phosphorylated at tyrosine residues in its intracellular domain and remains constitutively active. Activated Tie2 promotes the barrier function and anti-inflammation. In inflammation, Angiopoietin 2 is upregulated at tyrosine residues in its intracellular domain and remains constitutively active. Activated Tie2 promotes the barrier function and anti-inflammation. In inflammation, Angiopoietin 2 is upregulated in endothelial cells and released from platelets to antagonize Angiopoietin-1. This event results in the inhibition of Tie2.

The role of both Tie2 and Angiopoietin-1 in developmental angiogenesis became evident when mice knockouts for the genes died in utero from defects in cardiac development and blood vessel formation [75,78,79]. Tie2 as well as Angiopoietin-1 are expressed in normal human arteries and veins [80]. Further studies demonstrated that Angiopoietin-1 but not Angiopoietin-2, was chemotactic for endothelial cells and augmented endothelial cell migration for angiogenesis. On the other hand, Angiopoietin-2 inhibited migration of endothelial cells toward angiopoietin-1.

c-Cbl downregulates Angiopoietin(1/2)-Tie2 axis and interacts with Tie-2 in a manner dependent on Angiopoietin-1 stimulation [81]. This interaction results in the internalization of Angiopoietin and polyubiquitination of Tie-2 [81]. While inhibition of Angiopoietin (1/2)-Tie2 axis by c-Cbl is likely to suppress angiogenesis, further work is needed to elucidate the importance of Angiopoietin(1/2)-Tie2 axis in pathologic angiogenesis models.

3.4.2. c-Cbl and Eph Receptors

Along with ephrin ligands, Eph receptors play a key role in a variety of processes during embryonic development. Mice lacking Ephrin-B2 and double mutants deficient in EphB2 and EphB3 receptor signaling die in utero before embryonic day 11.5 (E11.5) due to defects in the remodeling of the embryonic vascular system. These mice show defects in the boundaries between arterial and venous domains [82]. These findings were consistently observed in other models, which showed that Ephrin-B2 regulated angiogenesis and lymphangiogenesis through VEGFR-3 signaling [83].

On the other hand, EphA2 regulation of angiogenesis is dependent on VEGFR-2 signaling [84]. In a retinal neovascularization model, EphrinA1 stimulated retinal microvascular endothelial cell proliferation, migration, tube formation and Ca2+ flux, and angiogenesis in rat aortic rings even in the milieu of VEGFR-2 inhibition. EphA2 is a target of c-Cbl [83,85]. The negative regulation of EphA2 mediated by c-Cbl was dependent on the activity of EphA2, as c-Cbl failed to downregulate the kinase inactive mutant of EphA2. The dominant negative mutant 70Z-Cbl also failed to suppress EphA2. Taken together, these data indicate the role of c-Cbl in Eph-mediated angiogenesis.

3.4.3. c-Cbl and c-Src

c-Src is a member of the Src family of non-receptor tyrosine kinases. A large body of literature indicates their integral role in tumorigenesis and several of them also enhance angiogenesis [42,86,87]. For example, Donnini et al. observed that a pyrazolo-pyrimidine-derived c-Src inhibitor suppressed vascular endothelial growth factor production and signaling, thereby reducing angiogenesis as well as the survival of squamous carcinoma cells [88]. c-Cbl polyubiquitinates c-Src [89,90].

4. c-Cbl in Tumor-Mediated Angiogenesis

Tumors manipulate host blood flow to ensure their growth and further exploit the blood vessels for metastasis. Thus, angiogenesis is an integral part of tumor growth [1,5,6,10,13]. Specific models of tumor angiogenesis have demonstrated the role of c-Cbl in this process. Typically, tumor cells are implanted in mice as a xenograft (on the back of mice) or an orthografts (at the site of its natural growth.
For example, colon cancer cells injected in mice and the vasculature within the tumor mass serves as a metrics for tumor angiogenesis. Meyer et al subcutaneously injected syngeneic B16F mouse melanoma cells into c-CBL knockout mice. They hypothesized that compared to wild-type c-Cbl mice, the tumors of c-CBL knockout mice would exhibit higher angiogenesis [25]. In fact, results showed significantly increased tumor growth in c-Cbl knockout mice when compared to the wild type mice; concurrent with increased tumor angiogenesis. This model specifically demonstrates the role of c-Cbl in tumor angiogenesis.

5. c-Cbl in Myocardial Ischemia

Critical coronary artery obstruction due to atherothrombotic coronary artery disease results in myocardial ischemia or angina and remains the leading cause of death in the world [91]. Myocardial ischemia is followed by reperfusion and remodeling of the cardiac myocytes. Given the role of c-Cbl in angiogenesis, Rafiq et al investigated c-Cbl in recovery from myocardial ischemia and demonstrated that mice lacking c-Cbl showed an increased functional recovery following myocardial ischemia [92]. A significant reduction in myocardial cell apoptosis was noted in these mice which augmented cardiac function post injury [92]. Loss of c-Cbl was correlated with an enhanced expression of angiogenic factors such as VEGF and its receptor VEGFR-2, as well as increased expression of epidermal growth factor receptor (EGFR) and focal adhesion kinase (FAK) [92]. Taken together, the loss of c-Cbl was cardio protective and elicited this effect by preventing or reducing the ubiquitination and degradation of various critical angiogenic factors.

6. c-Cbl in Retinal Neovascularization

Choroidal neovascularization retinal angiogenesis is considered the pathognomonic feature of age-related macular degeneration [24,93–95]. VEGF signaling has been identified as an important contributor to this disease [95–97]. c-Cbl knockout mice were subjected to laser-induced choroidal neovascularization (CNV) formation using the laser photocoagulation method [24,25]. The mice were followed by fundus photography and angiography to document the size and leakage from the CNV in these lesions using fluorescein angiography. c-Cbl knockout (KO) status was associated with a two-fold greater number of leaking CNV lesions compared with the wild-type mice. The CNV lesions were more aggressive and a higher number of lesions displayed confluence in c-Cbl KO mice. This data strongly suggested that c-Cbl is a negative regulator of CNV. While both of the above models suggest the role of c-Cbl in a model of pathological angiogenesis, its role in other models such as hind limb ischemia remains to be investigated.

7. Therapeutic Potential of c-Cbl

The quintessential role of the ubiquitination-proteasomal system in fundamental cell processes has attracted intense attention of the pharmaceutical industry. In fact, this system is already a validated therapeutic target for several diseases including cancer and is explored in other diseases such as heart conditions, etc. [98,99].

Currently, the inhibitors of ubiquitin-proteasome are the first line of therapy for paraproteinemias such as multiple myeloma [100]. Several preclinical studies and phase I clinical trials are underway for a range of compounds for variety of indications including liquid and solid cancers [98]. The inhibition of the ubiquitination-proteasomal system exerts a wide variety of effects such as cell death from the accumulation of ubiquitinated immunoglobin protein, induction of senescence, and anti-proliferative and immunomodulatory effects. While these global inhibitors of the ubiquitination-proteasomal system have paved the way for developing viable drug targets, targeting of E3 ubiquitin ligase remains an attractive proposal to drive specificity while minimizing the off-target effects.

c-Cbl represents an attractive target from several perspectives. For example, in the area of cancer, c-Cbl suppresses both tumorigenesis and angiogenesis (Figure 3). In colorectal cancer, c-Cbl suppresses the growth of colon cancer cells by downregulating Wnt signaling (a critical driver of colorectal cancer)
and suppressing nuclear β–catenin (a hallmark of Wnt activation) [47]. In fact, the expression of c-Cbl in colorectal tumors inversely correlated with the overall survival of patients [48]. c-Cbl also suppresses other mitogenic proteins such as c-MET, PDGFR and FGF that are upregulated in colorectal and other cancers [10,42,101]. At the same time, c-Cbl inhibits tumor-induced angiogenesis by targeting the Wnt/β–catenin pathway, VEGFR-2 signaling, Angiopoietin(1/2)-Tie2 axis, and other tyrosine kinases [25]. c-Cbl is highly expressed in immune cells such as T-lymphocytes and myeloid cells. In fact, a global c-Cbl knockout was characterized by immune cell proliferation and splenomegaly [102,103]. Recent explosion in the field of immune oncology has underscored the profound influence these cells and their receptors have on tumor growth [104,105]. Alteration of the tumor milieu by modulation of c-Cbl can exert an additional benefit, this area remains to be explored.

Recent explosion in the field of immune oncology has underscored the profound influence these cells and their receptors have on tumor growth [104,105]. Alteration of the tumor milieu by modulation of c-Cbl can exert an additional benefit, this area remains to be explored.

Figure 3. c-Cbl is an attractive therapeutic target for angiogenesis and tumorigenesis. In the endothelial cells of vessels supplying the tumor, Wnt activation results in secretion of pro-angiogenic cytokines IL-8 and VEGF-A that augment tumor-induced angiogenesis. In colon cancer epithelial cells, inactivating mutations in APC tumor suppressor stabilizes β-Catenin and allows its nuclear translocation. This event results in the transcription of pro-proliferative and pro-oncogenic genes such as Cyclin D1 and c-Myc to augment oncogenic activity. c-Cbl, by ubiquitinating nuclear β-Catenin in endothelial cells and in colon cancer cells, suppresses the growth of blood vessels and tumors. In addition, c-Cbl suppresses these processes by downregulating other pathways such VEGF signaling (PLCγ1) and mitogenic signaling (RTK and non-RTKs).

There are potential solutions regarding the concerns around targeting c-Cbl. Diseases such as cancer will warrant activation of c-Cbl for its therapeutic value. In general, activation of a protein is considered challenging compared to inhibiting a protein. However, c-Cbl’s crystal structure provides an intriguing possibility of addressing this limitation [29]. Constitutively c-Cbl remains in an autoinhibited conformation. The activation of E3 ubiquitin ligase is impacted by phosphorylation of Tyr371. The phosphorylation status of Tyr371 is important for its ability to suppress Wnt signaling.
in cells and animals [48]. It is conceivable that targeting the kinases or phosphatases modulating the phosphorylation of this particular tyrosine residue can be exploited to drive therapeutic benefit. c-Cbl is ubiquitously expressed protein and its manipulation can have off-target effects. This limitation can be circumvented by technologies that precisely target cell-types or organs. Biomarkers accompanying a therapeutic strategy, helps to narrow the patient population to maximize the benefit and offset the off-target effects. In this case, targeting of c-Cbl can be linked to its activity in the organ of interest and the antibody-based detection of phosphorylation of Tyr371 can potentially serve as a marker of c-Cbl activity. Development of such resources will enrich the understanding of the pattern of activity of c-Cbl in relationship to development of a disease model. Eventually, such know-how will drive the therapeutic potential of c-Cbl.

8. Conclusions
c-Cbl as a deterrent of angiogenesis is both interesting and highly compelling. Regulation of key angiogenic factors by c-Cbl underscores its importance in this fundamental biological process. Independent of its suppressive effect on angiogenesis, c-Cbl’s ability to regulate tumor growth and potentially modulate immune cells to orchestrate potent growth inhibitory effects on tumors, makes it an appealing target for cancer therapeutics. Further work in some of the underexplored aspects of c-Cbl biology such as biomarkers of its activity and it’s role in other models of angiogenesis (such as hind limb ischemia and a therapeutic strategy to specifically target it), will allow realization of its true potential as a critical regulating node in the fabric of several disease processes.

Author Contributions: C.L.L. and V.C.C. drafted the entire manuscript, M.B. designed the figures and edited the manuscript.

Funding: This work was funded by R01CA175382, R01 HL132325 and Evans Faculty Merit award (VCC); R01 HL132325 (MB) and T32 Research Training T32 DK007053-44 (CL)

Acknowledgments: Authors thank Elysia Heilig for proof reading the manuscript.

Conflicts of Interest: Authors report no conflict of interest.

References
1. Simons, M.; Ware, J.A. Therapeutic angiogenesis in cardiovascular disease. Nat. Rev. Drug Discov. 2003, 2, 863. [CrossRef] [PubMed]
2. Abhinand, C.S.; Raju, R.; Soumya, S.J.; Arya, P.S.; Sudhakaran, P.R. VEGF-A/VEGFR2 signaling network in endothelial cells relevant to angiogenesis. J. Cell Commun. Signal. 2016, 10, 347–354. [CrossRef]
3. Cantatore, F.P.; Maruotti, N.; Corrado, A.; Ribatti, D. Angiogenesis Dysregulation in the Pathogenesis of Systemic Sclerosis. Biomed. Res. Int. 2017, 2017, 5345673. [CrossRef]
4. Devi, L.; Pothana, L.; Goel, S. Dysregulation of angiogenesis-specific signalling in adult testis results in xenograft degeneration. Sci. Rep. 2017, 7, 2605. [CrossRef] [PubMed]
5. Patan, S. Vasculogenesis andangiogenesis. Cancer Treat. Res. 2004, 117, 3–32.
6. Carmelletti, P.; Jain, R.K. Angiogenesis in cancer and other diseases. Nature 2000, 407, 249–257. [CrossRef]
7. Kim, H.-J.; Jang, S.Y.; Park, J.-I.; Byun, J.; Kim, D.-I.; Do, Y.-S.; Kim, J.-M.; Kim, S.; Kim, B.-M.; Kim, W.-B.; et al. Vascular endothelial growth factor-induced angiogenic gene therapy in patients with peripheral artery disease. Exp. Amp; Mol. Med. 2004, 36, 336. [CrossRef]
8. Talia, N.C.; Alfaro, V.D., III; John, B.K.; Eric, P.J. Diabetic Retinopathy and Angiogenesis. Curr. Diabetes Rev. 2009, 13, 8–13. [CrossRef]
9. Koch, A.E. Angiogenesis as a target in rheumatoid arthritis. Ann. Rheum. Dis. 2003, 62, ii60. [CrossRef]
10. Danese, S.; Sans, M.; de la Motte, C.; Graziani, C.; West, G.; Phillips, M.H.; Pola, R.; Rutella, S.; Willis, J.; Gasbarrini, A.; et al. Angiogenesis as a Novel Component of Inflammatory Bowel Disease Pathogenesis. Gastroenterology 2006, 130, 2060–2073. [CrossRef] [PubMed]
11. Haugsten, E.M.; Wiedlocha, A.; Olsnes, S.; Wescbe, J. Roles of Fibroblast Growth Factor Receptors in Carcinogenesis. Mol. Cancer Res. 2010, 8, 1439. [CrossRef]
12. Tiong, K.H.; Mah, L.Y.; Leong, C.-O. Functional roles of fibroblast growth factor receptors (FGFRs) signaling in human cancers. *Cell Death 2013*, 18, 1447–1468. [CrossRef]

13. Schreiber, A.B.; Winkler, M.E.; Derynck, R. Transforming growth factor-alpha: A more potent angiogenic mediator than epidermal growth factor. *Science 1986*, 232, 1250. [CrossRef]

14. Akhurst, R.J.; Derynck, R. TGF-β signaling in cancer—a double-edged sword. *Trends Cell Biol. 2001*, 11, S44–S51. [CrossRef] [PubMed]

15. Derynck, R.; Akhurst, R.J.; Balmain, A. TGF-β signaling in tumor suppression and cancer progression. *Nat. Genet. 2001*, 29, 117. [CrossRef]

16. Fajardo, L.F.; Kwan, H.H.; Kowalski, J.; Prionas, S.D.; Allison, A.C. Dual role of tumor necrosis factor-alpha in angiogenesis. *Am. J. Pathol. 1992*, 140, 539–544. [PubMed]

17. Leibovich, S.J.; Polverini, P.J.; Shepard, H.M.; Wiseman, D.M.; Shively, V.; Nuseir, N. Macrophage-induced angiogenesis is mediated by tumour necrosis factor-α. *Nature 1987*, 329, 630–632. [CrossRef]

18. Claesson-Welsh, L.; Welsh, M. VEGFA and tumour angiogenesis. *J. Intern. Med. 2013*, 273, 114–127. [CrossRef]

19. Matsumoto, K.; Ema, M. Roles of VEGF-A signalling in angiogenesis, regeneration, and tumours. *J. Biochem. 2014*, 156, 1–10. [CrossRef] [PubMed]

20. Melincovici, C.S.; Bosca, A.B.; Susman, S.; Marginean, M.; Mihu, C.; Istrate, M.; Moldovan, I.M.; Roman, A.L.; Mihu, C.M. Vascular endothelial growth factor (VEGF)—key factor in normal and pathological angiogenesis. *Rom. J. Morphol. Embryol. Rev. Roum. De Morphol. Et Embryol. 2018*, 59, 455–467.

21. Chialita, V.; Shivanna, S.; Martorell, J.; Meyer, R.; Edelman, E.; Rahimi, N. c-Cbl, a ubiquitin E3 ligase that targets active β-catenin: A novel layer of Wnt signaling regulation. *J. Biol. Chem. 2015*, 288, 23505–23517. [CrossRef] [PubMed]

22. Shivanna, S.; Harrold, I.; Shashar, M.; Meyer, R.; Kiang, C.; Francis, J.; Zhao, Q.; Feng, H.; Edelman, E.R.; Rahimi, N.; et al. The c-Cbl ubiquitin ligase regulates nuclear β-catenin and angiogenesis by its tyrosine phosphorylation mediated through the Wnt signaling pathway. *J. Biol. Chem. 2015*, 290, 12537–12546. [CrossRef] [PubMed]

23. Zhang, Y.; Xia, M.; Jin, K.; Wang, S.; Wei, H.; Fan, C.; Wu, Y.; Li, X.; Li, X.; Li, G.; et al. Function of the c-Met receptor tyrosine kinase in carcinogenesis and associated therapeutic opportunities. *Mol. Cancer 2018*, 17, 45. [CrossRef]

24. Husain, D.; Meyer, R.D.; Mehta, M.; Pfeifer, W.M.; Chou, E.; Navruzbekov, G.; Ahmed, E.; Rahimi, N. Role of c-Cbl-dependent regulation of phospholipase Cgamma1 activation in experimental choroidal neovascularization. *Invest. Ophthalmol. Vis. Sci. 2010*, 51, 6803–6809. [CrossRef]

25. Meyer, R.D.; Husain, D.; Rahimi, N. c-Cbl inhibits angiogenesis and tumor growth by suppressing activation of PLCgamma1. *Oncogene 2011*, 30, 2198–2206. [CrossRef]

26. Prenen, H.; Smeets, D.; Mazzone, M.; Lambrechts, D.; Sagaert, X.; Sciot, R.; Debiec-Rychter, M. Phospholipase C gamma 1 (PLCG1) R707Q mutation is counterselected under targeted therapy in a patient with hepatic angiosarcoma. *Oncotarget 2015*, 6, 36418–36425. [CrossRef] [PubMed]

27. Wang, Y.; Wu, J.; Wang, Z. Akt binds to and phosphorylates phospholipase C-gamma1 in response to epidermal growth factor. *Mol. Biol. Cell 2006*, 17, 2267–2277. [CrossRef]

28. Swaminathan, G.; Tsygankov, A.Y. The Cbl family proteins: Ring leaders in regulation of cell signaling. *J. Cell Physiol. 2006*, 209, 21–43. [CrossRef]

29. Dou, H.; Buetow, L.; Hock, A.; Sibbet, G.J.; Vosden, K.H.; Huang, D.T. Structural basis for autoinhibition and phosphorylation-dependent activation of c-Cbl. *Nat. Struct. Mol. Biol. 2012*, 19, 184–192. [CrossRef]

30. Langdon, W.; Hartley, J.; Klinken, S.; K Ruscetti, S.; Morse, H. v-cbl, an Oncogene from a Dual-Recombinant Murine Retrovirus that Induces Early B-Lineage Lymphomas. *Proc. Natl. Acad. Sci. USA 1989*, 86, 1168–1172. [CrossRef]

31. J Blake, T.; G Heath, K.; Langdon, W. The truncation that generated the v-cbl oncogene reveals an ability for nuclear transport, DNA binding and acute transformation. *EMBO J. 1993*, 12, 2017–2026. [CrossRef]

32. Blake, T.J.; Shapiro, M.; Morse, H.C., 3rd; Langdon, W.Y. The sequences of the human and mouse c-cbl proto-oncogenes show v-cbl was generated by a large truncation encompassing a proline-rich domain and a leucine zipper-like motif. *Oncogene 1991*, 6, 653–657.

33. Chiang, Y.J.; Kole, H.K.; Brown, K.; Naramura, M.; Fukuhara, S.; Hu, R.J.; Jang, I.K.; Gutkind, J.S.; Shevach, E.; Gu, H. Cbl-b regulates the CD28 dependence of T-cell activation. *Nature 2000*, 403, 216–220. [CrossRef]
34. Lutz-Nicoladoni, C.; Wolf, D.; Sopper, S. Modulation of Immune Cell Functions by the E3 Ligase Cbl-b. *Front. Oncol.* 2015, 5, 58. [CrossRef]

35. Mukhopadhyay, C.; Triplett, A.; Bargar, T.; Heckman, C.; Wagner, K.-U.; Naramura, M. Casitas B-cell lymphoma (Cbl) proteins protect mammary epithelial cells from proteotoxicity of active c-Src accumulation. *Proc. Natl. Acad. Sci. USA* 2016, 113, E8228–E8237. [CrossRef]

36. Mohapatra, B.; Zutshi, N.; An, W.; Goetz, B.; Arya, P.; Bielecki, T.A.; Mushtaq, I.; Storck, M.D.; Meza, J.L.; Band, V. An essential role of CBL and CBL-B ubiquitin ligases in mammary stem cell maintenance. *Development* 2017, 144, 1072–1086. [CrossRef]

37. Smit, L.; Borst, J. The Cbl family of signal transduction molecules. *Crit. Rev. Oncog.* 1997, 8, 359–379.

38. Singh, A.J.; Meyer, R.D.; Navruzbekov, G.; Shelke, R.; Duan, L.; Band, H.; Leeman, S.E.; Rahimi, N. A critical role for the E3-ligase activity of c-Cbl in VEGFR-2-mediated PLCgamma1 activation and angiogenesis. *Proc. Natl. Acad. Sci. USA* 2007, 114, E8228–E8237. [CrossRef]

39. Rahimi, N. The ubiquitin-proteasome system meets angiogenesis. *Mol. Cancer* 2012, 11, 538–548. [CrossRef] [PubMed]

40. Rahimi, N.; Costello, C.E. Emerging roles of post-translational modifications in signal transduction and angiogenesis. *Proteomics* 2015, 15, 300–309. [CrossRef] [PubMed]

41. Rape, M. Ubiquitylation at the crossroads of development and disease. *Nat. Rev. Mol. Cell Biol.* 2017, 19, 59. [CrossRef] [PubMed]

42. Taher, T.E.; Tjin, E.P.; Beuling, E.A.; Borst, J.; Spaargaren, M.; Pals, S.T. c-Cbl is involved in Met signaling in B cells and mediates hepatocyte growth factor-induced receptor ubiquitination. *J. Immunol.* 2002, 169, 3793–3800. [CrossRef] [PubMed]

43. Leestemaker, Y.; Ovaa, H. Tools to investigate the ubiquitin proteasome system. *Drug Discov. Today Technol.* 2017, 26, 25–31. [CrossRef] [PubMed]

44. El Hokayem, J.; Amadei, C.; Obeid, J.P.; Nawaz, Z. Ubiquitination of nuclear receptors. *Clin. Sci.* 2017, 131, 917–934. [CrossRef] [PubMed]

45. Gallo, L.H.; Ko, J.; Donoghue, D.J. The importance of regulatory ubiquitination in cancer and metastasis. *Cell Cycle* 2017, 16, 634–648. [CrossRef]

46. Buettow, L.; Tria, G.; Ahmed, S.F.; Hock, A.; Dou, H.; Sibbet, G.J.; Svergun, D.I.; Huang, D.T. Casitas B-lineage lymphoma linker helix mutations found in myeloproliferative neoplasms affect conformation. *Bmc Biol.* 2016, 14, 76. [CrossRef]

47. Shashar, M.; Siwak, J.; Tapan, U.; Lee, S.Y.; Meyer, R.D.; Parrack, P.; Tan, J.; Khatri, M.; Francis, J.; Zhao, Q.; et al. c-Cbl mediates the degradation of tumorigenic nuclear beta-catenin contributing to the heterogeneity in Wnt activity in colorectal tumors. *Oncotarget* 2016, 7, 71136–71150. [CrossRef]

48. Kumaradevan, S.; Lee, S.Y.; Richards, S.; Lyle, C.; Zhao, Q.; Tapan, U.; Jiangliu, Y.; Ghumman, S.; Walker, J.; Belghasem, M.; et al. c-Cbl Expression Correlates with Human Colorectal Cancer Survival and Its Wnt/beta-Catenin Suppressor Function Is Regulated by Tyr371 Phosphorylation. *Am. J. Pathol.* 2018, 188, 1921–1933. [CrossRef]

49. Zhan, T.; Rindtorff, N.; Boutros, M. Wnt signaling in cancer. *Oncogene* 2017, 36, 1461–1473. [CrossRef]

50. Reis, M.; Liebner, S. Wnt signaling in the vasculature. *Exp. Cell Res.* 2013, 319, 1317–1323. [CrossRef]

51. Nusse, R. Wnt signaling in disease and in development. *Cell Res.* 2005, 15, 28–32. [CrossRef]

52. Gomez-Orte, E.; Saenz-Narciso, B.; Moreno, S.; Cabello, J. Multiple functions of the noncanonical Wnt pathway. *Trends Genet.* 2013, 29, 545–553. [CrossRef]

53. Wang, Y. Wnt/Planar cell polarity signaling: A new paradigm for cancer therapy. *Mol. Cancer* 2009, 8, 2103. [CrossRef]

54. Kohn, A.D.; Moon, R.T. Wnt and calcium signaling: beta-catenin-independent pathways. *Cell Calcium* 2005, 38, 439–446. [CrossRef]

55. Sugimura, R.; Li, L. Noncanonical Wnt signaling in vertebrate development, stem cells, and diseases. *Birth Defects Res. Part Cembryo Today Rev.* 2010, 90, 243–256. [CrossRef]

56. Van Amerongen, R. Alternative Wnt pathways and receptors. *Cold Spring Harbor Persp. Biol.* 2012, a007914. [CrossRef]

57. Ishitani, T.; Kishida, S.; Hyodo-Miura, J.; Ueno, N.; Yasuda, J.; Waterman, M.; Shibuya, H.; Moon, R.T.; Ninomiya-Tsuji, J.; Matsumoto, K. The TAK1-NLK mitogen-activated protein kinase cascade functions in...
the Wnt-5a/Ca(2+) pathway to antagonize Wnt/beta-catenin signaling. *Mol. Cell. Biol.* 2003, 23, 131–139. [CrossRef]

58. Logan, C.Y.; Nusse, R. The Wnt signaling pathway in development and disease. *Annu. Rev. Cell Dev. Biol.* 2004, 20, 781–810. [CrossRef]

59. Ducharte, Y.; Kim, Y.M.; Kahn, M. The Wnt signaling pathway in cancer. *Crit. Rev. Oncol. Hematol.* 2016, 99, 141–149. [CrossRef]

60. Karimiania, A.; Majidinia, M.; Bannazadeh Baghi, H.; Yousefi, B. The crosstalk between Wnt/beta-catenin signaling pathway with DNA damage response and oxidative stress: Implications in cancer therapy. *Dna Repair* 2017, 51, 14–19. [CrossRef]

61. Foulquier, S.; Daskalopoulos, E.P.; Lluri, G.; Hermans, K.C.M.; Deb, A.; Blanksteijn, W.M. WNT Signaling in Cardiac and Vascular Disease. *Pharm. Rev.* 2018, 70, 68–141. [CrossRef]

62. Liao, H.J.; Kume, T.; McKay, C.; Xu, M.J.; Ihle, J.N.; Carpenter, G. Absence of erythrogenesis and vasculogenesis in Plcg1-deficient mice. *J. Biol. Chem.* 2002, 277, 9335–9341. [CrossRef]

63. Rahimi, N. A role for protein ubiquitination in VEGFR-2 signalling and angiogenesis. *Curr. Vasc. Pharm.* 2017, 19, 557–569. [CrossRef] [PubMed]

64. Siveen, K.S.; Prabhu, K.; Krishnankutty, R.; Kuttkrishnan, S.; Tsakou, M.; Alali, F.Q.; Dermime, S.; Mohammad, R.M.; Uddin, S. Vascular Endothelial Growth Factor (VEGF) Signaling in Tumour Vascularization: Potential and Challenges. *Curr. Protein Pept. Sci.* 2018, 19, 589–597. [CrossRef] [PubMed]

65. Lawson, N.D.; Mugford, J.W.; Diamond, B.A.; Weinstein, B.M. Phospholipase C gamma-1 is required for normal postnatal neovascularization. *Circ. Res.* 2018, 122, 947–958. [CrossRef] [PubMed]

66. Meyer, R.D.; Latz, C.; Rahimi, N. Comparative structure-function analysis of VEGFR-1 and VEGFR-2: What have we learned from chimeric systems? *Ann. NY Acad. Sci.* 2003, 995, 200–207. [CrossRef]

67. Sen, A.; Madhivanan, K.; Mukherjee, D.; Aguilar, R.C. The epsin protein family: coordinators of endocytosis and signaling. *Biomol. Concepts* 2012, 3, 117–126. [CrossRef] [PubMed]

68. Oldham, C.E.; Mohney, R.P.; Miller, S.L.; Hanes, R.N.; O’Bryan, J.P. The ubiquitin-interacting motifs target the endocytic adaptor protein epsin for ubiquitination. *Curr. Biol.* 2002, 12, 1112–1116. [CrossRef]

69. Song, H.; Pasula, S.; Brophy, M.; Tessneer, K.L.; Wu, H.; Dong, Y.; Chen, H. Abstract 15705: 1346–1351. [CrossRef] [PubMed]

70. Rahimi, N. Recruitment and activation of phospholipase Cgamma1 involved in brain disorders. *Prog. Neurobiol.* 2013, 93, 155–179. [CrossRef] [PubMed]

71. Sen, A.; Madhivanan, K.; Mukherjee, D.; Aguilar, R.C. The epsin protein family: coordinators of endocytosis and signaling. *Biomol. Concepts* 2012, 3, 117–126. [CrossRef] [PubMed]

72. Oldham, C.E.; Mohney, R.P.; Miller, S.L.; Hanes, R.N.; O’Bryan, J.P. The ubiquitin-interacting motifs target the endocytic adaptor protein epsin for ubiquitination. *Curr. Biol.* 2002, 12, 1112–1116. [CrossRef]

73. Song, H.; Pasula, S.; Brophy, M.; Tessneer, K.L.; Wu, H.; Dong, Y.; Chen, H. Abstract 15705: 1346–1351. [CrossRef] [PubMed]

74. Asahara, T.; Chen, D.; Takahashi, T.; Fujikawa, K.; Kearney, M.; Magner, M.; Yancopoulos George, D.; Isner Jeffrey, M. Tie2 Receptor Ligands, Angiopoietin-1 and Angiopoietin-2, Modulate VEGF-Induced Postnatal Neovascularization. *Curr. Res.* 1998, 83, 233–240. [CrossRef] [PubMed]
78. Chatterjee, S.; Naik, U.P. Pericyte-endothelial cell interaction. Cell Adhes. Migr. 2012, 6, 157–159. [CrossRef] [PubMed]
79. Milam, K.E.; Parikh, S.M. The angiopoietin-Tie2 signaling axis in the vascular leakage of systemic inflammation. Tissue Barriers 2015, 3, e957508. [CrossRef]
80. Witzenbichler, B.; Maisonpierre, P.C.; Jones, P.; Yancopoulos, G.D.; Isner, J.M. Chemotactic properties of angiopoietin-1 and -2, ligands for the endothelial-specific receptor tyrosine kinase Tie2. J. Biol. Chem. 1998, 273, 18514–18521. [CrossRef]
81. Wehrle, C.; Van Slyke, P.; Dumont, D.J. Angiopoietin-1-induced ubiquitylation of Tie2 by c-Cbl is required for internalization and degradation. Biochem. J. 2009, 423, 375. [CrossRef]
82. Adams, R.H.; Wilkinson, G.A.; Weiss, C.; Diella, F.; Gale, N.W.; Deutsch, U.; Risau, W.; Klein, R. Roles of ephrin ligands and EphB receptors in cardiovascular development: demarcation of arterial/venous domains, vascular morphogenesis, and sprouting angiogenesis. Genes Dev. 1999, 13, 295–306. [CrossRef]
83. Wang, Y.; Nakayama, M.; Pitulescu, M.E.; Schmidt, T.S.; Bochenek, M.L.; Sakakibara, A.; Adams, S.; Davy, A.; Deutsch, U.; Lüthi, U.; et al. Ephrin-B2 controls VEGF-induced angiogenesis and lymphangiogenesis. Nature 2010, 465, 483. [CrossRef]
84. Ferrara, L.A.D.E.; Fassbender, E.; Shen, S.; Woolfenden, A.; Qu, Y.; Poor, S.; Anderson, K.; Jaffee, B. EphA2 Stimulation of Angiogenesis is Dependent on VEGFR2. Investig. Ophthalmol. Vis. Sci. 2013, 54, 5613.
85. Wang, Yj.; Ota, S.; Katoaka, H.; Kanamori, M.; Li, Z.y.; Band, H.; Tanaka, M.; Sugimura, H. Negative regulation of EphA2 receptor by Cbl. Biochem. Biophys. Res. Commun. 2002, 296, 214–220. [CrossRef]
86. Irby, R.B.; Yeatman, T.J. Role of Src expression and activation in human cancer. Oncogene 2000, 19, 5636. [CrossRef]
87. Wheeler, D.L.; Iida, M.; Dunn, E.F. The role of Src in solid tumors. Oncologist 2009, 14, 667–678. [CrossRef]
88. Donnini, S.; Monti, M.; Castagnini, C.; Solito, R.; Botta, M.; Schenone, S.; Giachetti, A.; Ziche, M. Pyrazolo-pyrimidine-derived c-Src inhibitor reduces angiogenesis and survival of squamous carcinoma cells by suppressing vascular endothelial growth factor production and signaling. Int. J. Cancer 2007, 120, 995–1004. [CrossRef]
89. Yokouchi, M.; Kondo, T.; Sanjay, A.; Houghton, A.; Yoshimura, A.; Komiya, S.; Zhang, H.; Baron, R. Src-catalyzed Phosphorylation of c-Cbl Leads to the Interdependent Ubiquitination of Both Proteins. J. Biol. Chem. 2001, 276, 35185–35193. [CrossRef]
90. Lee, G.W.; Park, J.B.; Park, S.Y.; Seo, J.; Shin, S.H.; Park, J.W.; Kim, S.J.; Watanabe, M.; Chun, Y.S. The E3 ligase C-CBL inhibits cancer cell migration by neddylyating the proto-oncogene c-Src. Oncogene 2018, 37, 5552–5568. [CrossRef]
91. Marzillii, M.; Merz, C.N.B.; Boden, W.E.; Bonow, R.O.; Capozza, P.G.; Chilian, W.M.; DeMaria, A.N.; Guarini, G.; Huqi, A.; Morrone, D.; et al. Obstructive Coronary Atherosclerosis and Ischemic Heart Disease: An Elusive Link! J. Am. Coll. Cardiol. 2012, 60, 951. [CrossRef]
92. Rafiq, K.; Kolpakov Mikhail, A.; Seqqat, R.; Guo, J.; Guo, X.; Qi, Z.; Yu, D.; Mohapatra, B.; Zutshi, N.; An, W.; et al. c-Cbl Inhibition Improves Cardiac Function and Survival in Response to Myocardial Ischemia. Circulation 2014, 129, 2031–2043. [CrossRef]
93. Ambati, J.; Fowler, B.J. Mechanisms of age-related macular degeneration. Neuron 2012, 75, 26–39. [CrossRef]
94. Yoshida, T. Molecular mechanism of choroidal neovascularization in age-related macular degeneration. Nippon Ganka Gakkai Zasshi 2007, 111, 881–891.
95. Yang, S.; Zhao, J.; Sun, X. Resistance to anti-VEGF therapy in neovascular age-related macular degeneration: A comprehensive review. Drug Des. Dev. 2016, 10, 1857–1867. [CrossRef]
96. Cabral, T.; Mello, L.G.M.; Lima, L.H.; Polido, J.; Regatieri, C.V.; Belfort, R.; Mahajan, V.B. Retinal and choroidal angiogenesis: a review of new targets. Int. J. Retin. Vitr. 2017, 3, 31. [CrossRef]
97. Kwak, N.; Okamoto, N.; Wood, J.M.; Campochiaro, P.A. VEGF Is Major Stimulator in Model of Choroidal Neovascularization. Investig. Ophthalmol. Vis. Sci. 2000, 41, 3158–3164.
98. Weathington, N.M.; Mallampalli, R.K. Emerging therapies targeting the ubiquitin proteasome system in cancer. J. Clin. Investig. 2014, 124, 6–12. [CrossRef]
99. Drews, O.; Taegtmeyer, H. Targeting the ubiquitin-proteasome system in heart disease: the basis for new therapeutic strategies. Antioxid. Redox Signal. 2014, 21, 2322–2343. [CrossRef]
100. Lub, S.; Maes, K.; Menu, E.; De Bruyne, E.; Vanderkerken, K.; Van Valckenborgh, E. Novel strategies to target the ubiquitin proteasome system in multiple myeloma. Oncotarget 2015, 7, 6521–6537. [CrossRef]
101. Kaabeche, K.; Lemonnier, J.; Le Mee, S.; Caverzasio, J.; Marie, P.J. Cbl-mediated degradation of Lyn and Fyn induced by constitutive fibroblast growth factor receptor-2 activation supports osteoblast differentiation. *J. Biol. Chem.* 2004, 279, 36259–36267. [CrossRef]

102. Naramura, M.; Kole, H.K.; Hu, R.J.; Gu, H. Altered thymic positive selection and intracellular signals in Cbl-deficient mice. *Proc. Natl. Acad. Sci. USA* 1998, 95, 15547–15552. [CrossRef]

103. Murphy, M.A.; Schnall, R.G.; Venter, D.J.; Barnett, L.; Bertoncello, I.; Thien, C.B.; Langdon, W.Y.; Bowtell, D.D. Tissue hyperplasia and enhanced T-cell signalling via ZAP-70 in c-Cbl-deficient mice. *Mol. Cell. Biol.* 1998, 18, 4872–4882. [CrossRef]

104. Sharpe, A.H.; Pauken, K.E. The diverse functions of the PD1 inhibitory pathway. *Nat. Rev. Immunol.* 2018, 18, 153–167. [CrossRef]

105. Pardoll, D.M. The blockade of immune checkpoints in cancer immunotherapy. *Nat. Rev. Cancer* 2012, 12, 252–264. [CrossRef]

© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).