Abstract: Molecular hydrogen (H₂) is now considered part of the suite of small molecules that can control cellular activity. As such, H₂ has been suggested to be used in the therapy of diseases in humans and in plant science to enhance the growth and productivity of plants. Treatments of plants may involve the creation of hydrogen-rich water (HRW), which can then be applied to the foliage or roots systems of the plants. However, the molecular action of H₂ remains elusive. It has been suggested that the presence of H₂ may act as an antioxidant or on the antioxidant capacity of cells, perhaps through the scavenging of hydroxyl radicals. H₂ may act through influencing heme oxygenase activity or through the interaction with reactive nitrogen species. However, controversy exists around all the mechanisms suggested. Here, the downstream mechanisms in which H₂ may be involved are critically reviewed, with a particular emphasis on the H₂ mitigation of stress responses. Hopefully, this review will provide insight that may inform future research in this area.

Keywords: antioxidants; heme oxygenase; hydrogen gas; hydrogenase; hydroxyl radicals; molecular hydrogen; nitric oxide; reactive oxygen species

1. Introduction

Molecular hydrogen (H₂) is now recognized to have biochemical effects in both animals [1,2] and plants [3,4]. Although it is a relatively inert gas, H₂ appears to have profound effects on cell activity, which can be harnessed to help plant growth, survival, and productivity [5–8].

Plants, particularly as they are sessile, have to endure and survive a wide range of stress challenges, both biotic and abiotic. These stresses include attack by pathogens [9] and insects [10], as well as heavy metals [11], extreme temperature [12], salt [13], and ultraviolet B light [14]. It has become apparent over many years of study that there are common molecular responses to such stresses, and these mechanisms often involve reactive oxygen species (ROS) [15] and reactive nitrogen species (RNS) [16]. These compounds include ROS such as superoxide anions (O₂⁻) and hydrogen peroxide (H₂O₂), the latter of which is a major focus of ROS signalling [17]. Importantly, ROS also include the hydroxyl radical (·OH). The most prominent RNS is nitric oxide (NO), which is known to be involved in plant cell signalling processes [18]. However, other RNS include peroxynitrite and nitrosoglutathione, both of which can act as signalling molecules [19,20]. It is also apparent that crosstalk occurs between ROS and RNS [21] as well as with other reactive signalling molecules such as hydrogen sulphide (H₂S) [22,23].

H₂ fits into this suite of reactive signalling molecules and was shown to increase the fitness of plants [24]. Suitable examples of recent papers on H₂ effects on plants include mitigation of salinity effects in barley [25] and Arabidopsis [26], and increased tolerance to cadmium in alfalfa [27]. However, exactly how H₂ interacts and has an effect is unclear. The metabolism of H₂ in plants is not a novel idea [28] and some plants are known to be significant generators of H₂, such as Chlamydomonas [29,30], whilst higher plants have been shown to produce H₂ too. Plant H₂ generation has been known for a long time [28,31], with more recent examples being reported using rice seedlings [32] and
tomato plants [33]. The role of hydrogenase enzymes and the generation of H$_2$ by plants was recently reviewed [7].

Molecular hydrogen, being a gas, is hard to use either in laboratory or environmental settings. It is extremely flammable [34], relatively insoluble [35,36], and will readily move to the gas phase. Despite this, treatment with H$_2$ is often facilitated by the production of hydrogen-rich water (HRW), which can then be applied to the soil or directly onto the foliage. If using hydroponics, the HRW can be added directly to the feed solution. Several examples of the use of HRW are included throughout this review (for example, [5,8,37]). The use of HRW is effective and easy and is commonly used to treat plants, but treatment with H$_2$ gas can also have cellular effects and is often used in animal studies, for example, with mice [38]. H$_2$ gas has been used to alter plant growth by the gaseous treatment of the soil [39]. The treatment of biological materials with H$_2$ was further discussed in previous papers [7,40].

Here, we provide a critical look at the correlation between the effect of H$_2$ and the possible modes of action, with stress responses in plants being a focus. Issues that are addressed here include both the direct and indirect actions of H$_2$ and what biological compounds H$_2$ interacts within a cell, leading to the observed responses. Once this is established, a clearer view of downstream signal transduction initiated by H$_2$ can be gained. It is hoped that this review will inform future research in this area of plant science.

2. Downstream Effects

For any molecule to be used in cell signalling, it needs to be perceived by cells and to initiate a response. For many molecules, this involves a receptor protein, which may be on the cell surface [41] or in an intracellular compartment, such as the cytoplasm [42] or nucleus [43]. Some signalling molecules are perceived by proteins not classed as receptors, such as the effect of NO on soluble guanylyl cyclase (sGC). Here, NO reacts with the iron in the heme group of the enzyme, thereby activating it [44], although the involvement of such mechanisms has been questioned in plants [45]. Alternatively, the reactive nature of ROS and RNS allows them to oxidize [46] and nitrosate [47] thiol groups on proteins, propagating the signalling needed. It is hard to envisage how H$_2$, being so small and relatively inert, can be perceived by cells. Some of the mechanisms reported and mooted are discussed below.

2.1. Effects on Reactive Oxygen Species and Antioxidant Capacity

Stress responses in plants often involve ROS metabolism. There is often an increase in ROS accumulation, which, in some cases, can initiate programmed cell death (PCD) in plants [48]. ROS accumulate in the presence of heavy metals [49], such as cadmium [50], mercury, and copper [51]. ROS also accumulate in the presence of salt, extreme temperature, and pathogens [52]. Increases in the intracellular ROS under such stress conditions are often accompanied by an increase in antioxidant levels in cells, for example, in the presence of salt [53], heavy metals [54], and extreme temperature [55]. Therefore, the modulation of ROS metabolism is crucial for stress responses: increases in ROS lead to changes in cellular function, whilst antioxidants modulate and dampen that response.

H$_2$ has been shown to be able to help plant cells mitigate stress challenge. H$_2$ can help reduce salt stress [56,57], and reduce stress due to aluminium [58,59], cadmium [60], and mercury [61]. H$_2$ also can help mitigate against drought stress [62,63] and paraquat induced oxidative stress [64].

Xie et al. [57] suggested that H$_2$ modulates plant cells’ antioxidant capacity through acting through zinc-finger transcription factor ZAT10/12. This would dampen the ROS accumulation and associated lipid peroxidation. They also suggested that H$_2$ would act on the antiporters and proton pumps responsible for exclusion of Na$^+$, particularly the protein salt overly sensitive1 (SOS1). Finally, it was suggested that both SOS1 and cytosolic ascorbate peroxidase1 (cAPX1) are molecular targets of H$_2$-mediated signalling. Additionally, Xu et al. [59] also suggested that H$_2$ may alter gene expression. In a study of
aluminium stress, they found that H$_2$ altered the ratio of gibberellin acid (GA) and abscisic acid (ABA), with the expression of genes for GA biosynthesis (GA20ox1 and GA20ox2) and for ABA breakdown (ABA8ox1 and ABA8ox2) being induced by H$_2$. H$_2$ also altered miRNA expression with downstream effects that increased superoxide dismutase (SOD) expression, increasing antioxidant levels in the cells. However, even though these findings all support the notion that H$_2$ is protecting the cells, no direct interaction with H$_2$ has been established.

As can be seen from the discussion above, both stress responses and the effects of H$_2$ can be linked to ROS metabolism and antioxidant levels in cells. Therefore, it is particularly pertinent that H$_2$ has been posited to be an antioxidant [65]. Although this study discusses the effects in H$_2$ in a clinical setting, the redox chemistry would be the same in plants cells. In an animal setting, a study showed that H$_2$ is an antioxidant against the hydroxyl radical (·OH) but has no effects against other ROS [66]. This is most significant, as it is usually hydrogen peroxide (H$_2$O$_2$) that is deemed to be the primary inter- and intracellular signal [17,67]. Of importance, the specificity of H$_2$ to scavenge ·OH has been disputed, as an in vitro study showed that H$_2$ can scavenge H$_2$O$_2$. However, H$_2$ could not scavenge superoxide anions [57]. In an experiment looking at the radiolysis of water, a negligible effect on the formation or consumption of H$_2$O$_2$ was seen when molecular hydrogen was added [68].

If, as suggested [66], the effects of H$_2$ are mediated partly by ·OH scavenging, a series of questions could be asked: How influential are the levels of hydroxyl radicals in cells, and could H$_2$ be acting through their modulation? Would this account for the effects seen?

Hydroxyl radicals are known to have effects in plant cells. Richards et al. [69] described the hydroxyl radical as being a “potent regulator in plant cell biology”. They discussed the role of this molecule in numerous physiological mechanisms in plants, including germination, control of stomatal apertures, reproduction, and adaptation to stress challenge. ·OH has also been shown to be important for ion currents in roots [70,71]. In animal cells, ·OH was shown to be upstream of mitogen-activated protein kinases (MAPKs) and transcription factors (ERK2 and NF-κB) [72], and analogous mechanisms could exist in plants. Therefore, evidence exists of ·OH acting in a positive cell signalling role, which could potentially be the target of H$_2$.

In biological systems, ROS are often the product of the sequential reduction of molecular oxygen, resulting ultimately in the 4-electron reduction to water (Equation (1)).

\[
\begin{align*}
O_2 & \rightarrow O_2^- \rightarrow H_2O_2 \rightarrow 2(\cdot OH) \rightarrow 2H_2O
\end{align*}
\]

(1)

The superoxide anion (O$_2$·−) can be produced enzymatically, for example from the action of NADPH oxidases [73]. H$_2$O$_2$ can be produced by the subsequent dismutation of O$_2$·− by the enzyme family of superoxide dismutases (SOD) [74]. ·OH can be then be subsequently produced, especially in the presence of metal ions [75,76]. This generation can be either from the Fenton reaction from H$_2$O$_2$ (Equation (2)):

\[
H_2O_2 + Fe^{2+} \rightarrow \cdot OH + HO^- + Fe^{3+}
\]

(2)

Or in the presence of transition metals through the Haber–Weiss reaction, using superoxide anions and H$_2$O$_2$ (Equation (3)):

\[
H_2O_2 + O_2^- \rightarrow \cdot OH + OH^- + O_2
\]

(3)

If the production of ROS is initiated, for example, during a stress response as discussed above, the generation of ·OH is likely to proceed. Hydroxyl radicals can be detected in plant cells [77,78], and have been found to have multiple effects.

The application of H$_2$ has mitigating influences during stress, and therefore if the effects of H$_2$ are mediated by the removal of ·OH, then it might be expected that ·OH radicals would need to be produced during these stress responses, assuming H$_2$ is working in these cases as a ·OH scavenger. It is in fact the case that ·OH can be found in these
circumstances. For example, hydroxyl radicals increase during metal ion challenge [79], a cellular challenge in which H₂ has been shown to have a beneficial effect [58–61]. In a similar manner, -OH is produced during paraquat treatment of plants [80], another situation mitigated by H₂ [64]. During chilling stress and drought stress, increases in free iron and H₂O₂ have been recorded, and this implicates hydroxyl radical generation in downstream cellular responses [81]. Once again, H₂ has beneficial effects under drought conditions [62,63], as well as chilling stress [82]. -OH and H₂ also have similar actions in heat stress [83,84]. Therefore, it can be seen that there are many stress conditions which elicit accumulation of -OH and are also relieved by the presence of H₂, suggesting that the -OH scavenging activity of H₂ is potentially responsible for the changes in cellular activity seen. This of course does not consider any spatial-temporal differences in -OH accumulation during different stresses, or plant species variations, but the correlation of -OH action and H₂ effects may be pointing to a possible mechanism.

Certainly, to support the notion that -OH removal by H₂ could be biologically significant, a look at other -OH scavengers may be useful. Such scavenging has been suggested to be useful for animal health [85], whilst in plants, mannitol has been suggested to be protective through this mechanism [81]. Sugars such as sucralose has been studied for its -OH scavenging effects in Arabidopsis [86], whilst β-carboline alkaloids [87] and more novel compounds have been used in animal systems [88]. Such studies show that there is merit in modulating -OH in cells, and therefore support the notion that such action by H₂ may be significant.

On the other hand, and importantly, it has been suggested that the reaction of H₂ with -OH is too slow to be of physiological relevance [89], although the authors were discussing clinical settings. In this paper the rate constant for the reaction of H₂ with -OH producing H₂O and H⁺ is only 4.2 × 10⁷ M⁻¹ s⁻¹ (from [90,91]). The rate constant for other radical reactions was quoted as 10⁹ M⁻¹ s⁻¹. It was suggested [89] that the -OH would react with other biomolecules before reacting with the H₂, rendering the presence of H₂ as being irrelevant. Others have doubted whether H₂ has its effects through scavenging -OH, although this is from a human health perspective [92]. Assuming this is correct, the correlation of -OH production and H₂ effects during stress responses would also be irrelevant, begging the question, if -OH scavenging is not the mechanism, what is?

It is possible that H₂ has indirect effects on antioxidant levels. There are several reports of antioxidant levels in plant cells altering on H₂ treatment. For example, this was reported in a study using black barley (Hordeum distichum L.) [93]. Antioxidant enzymes such as catalase and SOD were increased in maize [94] with similar effects in Chinese cabbage [95]. HRW was also found to maintain the intracellular redox status of plant cells through alterations the levels of reduced and oxidized glutathione (GSH and GSSG) [60]. However, the direct targets of H₂ have not been identified in such studies. Therefore, it may be that H₂ is having effects on the cells’ antioxidant capacity, which can be measured, but it may not be a direct effect on the ROS themselves.

2.2. Impact on Reactive Nitrogen Species Metabolism

RNS, such as the nitric oxide radical (NO), have been known to have important effects in plant cells for over forty years [96], although there is still some controversy of their endogenous production and action [45]. NO, like ROS are well known to be involved in plant stress responses [97], many of which are ameliorated by H₂ treatment, as discussed above. Therefore, the relationship between H₂ presence and altered RNS metabolism is worth exploring.

H₂ has been shown to have effects in nitrogen fixation [98], although this is only one facet of this complex process. Nitrogen fixation relies on many factors including nutrient availability, the soil-plant interactions, and community facilitation as exemplified by the work carried out with the alpine shrub Salix herbacea [99–101]. H₂ has also been shown to alter NO synthesis during auxin-mediated root growth [33]. Li et al. [102] reported that NO was involved in H₂-induced root growth, whilst Zhu et al. [103] also link H₂ and NO,
reporting that $\text{H}_2$ promoted NO accumulation through increases in the activities of possible synthesizing enzymes: NO synthase-like enzymes and nitrate reductase. Additionally, HRW increased NO accumulation in a study on stomatal closure [104]. On the other hand, HRW decreased NO accumulation in alfalfa [59].

It is likely that during a stress response NO and ROS are produced temporally and spatially together, and they can interact to produce downstream products. Superoxide anions and NO together can lead to the generation of the $\cdot \text{OH}$ radical [105], and as discussed above this have been mooted as a potential mechanism of $\text{H}_2$ action. However, superoxide anions and NO can react to produce peroxynitrite ($\text{ONOO}^-\cdot$) [105], which can act as a signalling molecule in its own right [106,107], possibility through alterations of amino acids [108], with tyrosine nitration being a major covalent change seen [106] which could have important downstream effects [109].

It has been reported that $\text{H}_2$ reacts with ONOO$^-$, but not NO [66,110]. Therefore, it would be unlikely that $\text{H}_2$ has direct effects in the NO signalling, per se. However, it was reported that $\text{H}_2$ reacts with peroxynitrite, which would potentially alter NO-induced signalling pathways. Despite several papers discussing the scavenging of ONOO$^-$ by $\text{H}_2$ [58,60], it has been completely ruled out by others [89]. In this paper, as well as saying that the $\cdot \text{OH}$ reaction is too slow, they report that $\text{H}_2$: (1) does not alter the rate of conversion of ONOOH to $\text{NO}_3^-$ and $\text{H}^+$; (2) does not alter the rates of ONOO$^-$-mediated tyrosine nitration; (3) does not alter the oxidative stress responses mediated by either ONOO$^-$ or $\cdot \text{OH}$. Therefore, even if effects on NO metabolism are seen, such as alterations in activities of synthesising enzymes, there appears to be no direct scavenging of RNS, or $\cdot \text{OH}$, by $\text{H}_2$ which could account for the observed cellular effects.

2.3. Stress, Heme Oxygenase and $\text{H}_2$

An enzyme mechanism that has been found to be important for $\text{H}_2$ effects in cells involves the heme oxygenase enzyme (HO-1). For example, this was shown to be involved in root development in cucumber on treatment with HRW [37]. Hydrogen-mediated tolerance to paraquat was also shown to involve heme oxygenase [64]. Similar data can be found in studies of animal systems, for example, in mice [111].

HO-1 has been shown to be involved in a range of abiotic stress responses in plants, including salt, heavy metals, UV light, and drought. Responses to stresses such as drought are complex, involving the result of many genes being expressed and the effects of gene polymorphisms, as seen with *Phaseolus vulgaris* L. [112–115], with wild types showing tolerance differences [116,117]. Resistance and tolerance to extreme temperatures are also important and involve complicated cellular responses [118–121]. Such responses are often associated with the accumulation of cellular ROS and RNS [120]. The catalytic action of HO-1 is the breakdown of heme. This is an oxygen-dependent reaction that uses NADPH as a cofactor and generates biliverdin, carbon monoxide (CO), and iron [121,122]. Interestingly, CO has been shown to be involved in signalling events in cells, and could mediate downstream effects of $\text{H}_2$, whilst iron facilitates $\cdot \text{OH}$ production, as discussed above.

However, no direct interaction between $\text{H}_2$ and HO-1 seems to have been reported. Further, no reaction has been reported between $\text{H}_2$ and CO in biological systems. Therefore, the connection between $\text{H}_2$ treatment and alterations of HO-1 activity needs to be a focus for future research.

2.4. Paramagnetic Properties and Possible Cellular Effects

The above discussion throws doubt onto many biochemical and reactive aspects of $\text{H}_2$ effects in cells. However, the physical properties of $\text{H}_2$ may also be important. Hydrogen can exist with two nuclear spin states (ortho- and parahydrogen) [123,124]. It is the interconversion between these states that may be relevant here [125]. One of the interactions discussed was with NO, which could potentially alter NO signalling. There is also the possibility of interactions with transition metals [126]. This could have a potentially significant effect on cell signalling pathways, as many enzymes involved in signal transduction have
metal prosthetic groups, including guanylyl cyclase (at least in animals), SOD, and many respiratory and photosynthetic components. Many of the aforementioned enzymes may be involved in ROS and RNS metabolism, which are important in plant responses to many stresses, with such conditions being mitigated by H$_2$, as discussed above. It is conceivable that H$_2$ may interact with the heme during the catalytic cycle of HO-1, accounting for the effects mediated by this enzyme.

This physical aspect of H$_2$ action was mooted previously [127], although experimental evidence is lacking and future research may prove this avenue wrong. However, the idea of quantum biology is not confined to H$_2$ effects, and the topic was recently reviewed [128]. It was suggested that biological processes may occur due to quantum mechanical effects. A more recent review on this topic was also published [129].

3. Discussion

H$_2$ is known to be involved in the control of cellular functions in plant cells. For example, it was reported to be involved in both phytohormone signalling and stress responses [32]. On a pragmatic note, treatment with H$_2$ in the form of HRW was suggested to be useful for delaying postharvest spoilage of fruit [5]. Therefore, it is known, like animal cells [1,130], that H$_2$ has effects, and such actions may be harnessed for future manipulation of plant growth and crop enhancement [131].

Several mechanisms of H$_2$ action have been suggested, as summarized in Figure 1.

![Possible mechanism of action of H$_2$ in cells](image)

Figure 1. Possible mechanism of action of H$_2$ in cells. The likelihood of there being effects on particular molecules is indicated (red arrows and text).

One of the significant actions of H$_2$ in biological systems was suggested to be its ·OH scavenging activity [66], as reported in animal systems [132]. A range of studies have shown that ·OH increases in cells under stressful conditions [79–81], whilst H$_2$ has been shown to have effects on such stress responses [58–61]. It may be argued that removal of ·OH by H$_2$, if it is involved in important ·OH signalling pathways, should be detrimental to cell function, although many studies have looked at scavenging ·OH as a beneficial approach to cell and organism health, both in plants and animals [81,85–87]. Hydroxyl radicals are extremely reactive, and react with kinetics that are diffusion-limited, with rate constants for a range of biomolecules being determined, including ATP and ADP [133]. ·OH radicals are known to react with proteins [134], which can lead to amino acid oxidation, crosslinking, and degradation of the polypeptide [135]. Lipids [136], carbohydrates [137],...
and DNA [138] are also -OH targets. Therefore, the scavenging activity of H\textsubscript{2} may prevent the harmful effects of -OH, which may account for some of the observed effects. However, the biggest issue is the rate constant of the reaction between H\textsubscript{2} and -OH, which is deemed to be too slow for physiological relevance [89], suggesting that the other biomolecules may react first anyway, and therefore H\textsubscript{2} would not influence the levels of oxidative stress. The same authors also ruled out reactions with peroxynitrite, as discussed above. Therefore, with H\textsubscript{2} not able to scavenge other ROS [66] and the effects of H\textsubscript{2} on both -OH and ONOO\textsuperscript{-} being ruled out [89], it appears that the scavenging role of H\textsubscript{2} may have limited effects in cells, at best.

Heme oxygenase is one enzyme that has been reported as mediating H\textsubscript{2} effects [37,64]. Although being reported in several studies, as discussed above, there is little evidence of a direct interaction which could account for the data seen. However, not all the data are negative and seemingly point to dead ends. It was reported that H\textsubscript{2} scavenged H\textsubscript{2}O\textsubscript{2} [57], which, if confirmed and can be shown to have effects in vivo, would be very significant, as H\textsubscript{2}O\textsubscript{2} is one of the major ROS signalling molecules [17,67]. However, in radiolysis experiments with H\textsubscript{2}O\textsubscript{2}, the addition of H\textsubscript{2} only had a negligible effect [68], suggesting that more research in this area would be beneficial. Another positive effect that is worth exploring is the interaction of H\textsubscript{2} with metals. It was suggested that the beneficial effects of H\textsubscript{2} may be mediated by the reduction of Fe(III), oxidized as a result of oxidative stress. However, neither iron-sulphur clusters nor heme groups were reduced by the presence of H\textsubscript{2} [89]. Even so, the effect of H\textsubscript{2} on Fe(III) is an enticing suggestion, as transition metals are widely used in biological systems, making this another area that merits further investigation.

Finally, the paramagnetic properties of hydrogen may be relevant to its biological action, as previously mooted [127]. This may include interactions with NO or transition metals, but experimental data would be needed to support this notion. There are other papers with H\textsubscript{2} in catalysis, but it is difficult to determine their relevance to biochemical reactions, as they are often conducted under non-physiological conditions, such as high pressure [139].

In conclusion, although the involvement of molecular hydrogen in plant function has been known for a long time [28], there is still considerable uncertainty surrounding the exact actions of H\textsubscript{2} in cells. Its role as a direct antioxidant is doubted, although many cellular effects have been observed, including alterations in antioxidants, changes in enzyme activity, and modulation in gene expression. What is clear is that H\textsubscript{2} may be useful for the mitigation of plant stress, so it has been proposed to have an exciting future [4,131].

Author Contributions: J.T.H. wrote the draft of the manuscript and G.R. contributed to and edited the final version. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Data Availability Statement: There were no primary data generated in this study.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Ge, L.; Yang, M.; Yang, N.-N.; Yin, X.-X.; Song, W.-G. Molecular hydrogen: A preventive and therapeutic medical gas for various diseases. Oncotarget 2017, 8, 102653–102673. [CrossRef]
2. Huang, L. Molecular hydrogen: A therapeutic antioxidant and beyond. Med. Gas Res. 2016, 6, 219–222. [CrossRef]
3. Li, C.; Gong, T.; Bian, B.; Liao, W. Roles of hydrogen gas in plants: A review. Funct. Plant Biol. 2018, 45, 783–792. [CrossRef]
4. Zeng, J.; Ye, Z.; Sun, X. Progress in the study of biological effects of hydrogen on higher plants and its promising application in agriculture. Med. Gas Res. 2014, 4, 1–4. [CrossRef]
5. Hu, H.; Li, P.; Wang, Y.; Gu, R. Hydrogen-rich water delays postharvest ripening and senescence of kiwifruit. Food Chem. 2014, 156, 100–109. [CrossRef]
6. Wilson, H.R.; Veal, D.; Whitman, M.; Hancock, J.T. Hydrogen gas and its role in cell signalling. CAB Rev. 2017, 12, 1–3. [CrossRef]
7. Russell, G.; Zulfiqar, F.; Hancock, J.T. Hydrogenases and the Role of Molecular Hydrogen in Plants. Plants 2020, 9, 1136. [CrossRef] [PubMed]
8. Wu, Q.; Su, N.; Huang, X.; Ling, X.; Yu, M.; Cui, J.; Shabala, S. Hydrogen-rich water promotes elongation of hypocotyls and roots in plants through mediating the level of endogenous gibberellin and auxin. *Funct. Plant Biol.* **2020**, *47*, 771. [CrossRef] [PubMed]

9. Dodds, P.N.; Rathjen, J.P. Plant immunity: Towards an integrated view of plant–pathogen interactions. *Nat. Rev. Genet.* **2010**, *11*, 539–548. [CrossRef]

10. Cory, J.S.; Myers, J.H. Adaptation in an insect host-plant pathogen interaction. *Ecol. Lett.* **2004**, *7*, 632–639. [CrossRef]

11. Keyster, M.; Niekirk, L.-A.; Basson, G.; Carelse, M.; Bakare, O.; Ludidi, N.; Klein, A.; Mekuto, L.; Golub, A. Decoding Heavy Metal Stress Signalling in Plants: Towards Improved Food Security and Safety. *Plants* **2020**, *9*, 1781. [CrossRef]

12. Ding, Y.; Shi, Y.; Yang, S. Molecular Regulation of Plant Responses to Environmental Temperatures. *Mol. Plant* **2020**, *13*, 544–564. [CrossRef] [PubMed]

13. Yang, Y.; Guo, Y. Unraveling salt stress signaling in plants. *J. Integr. Plant Biol.* **2018**, *60*, 796–804. [CrossRef]

14. Vanhaelwyn, L.; Prinsen, E.; Van Der Straeten, D.; Vandebussche, F. Hormone-controlled UV-B responses in plants. *J. Exp. Bot.* **2016**, *67*, 4469–4482. [CrossRef] [PubMed]

15. Choudhury, F.; Rivero, R.M.; Blumwald, E.; Mittler, R. Reactive oxygen species, abiotic stress and stress combination. *Plant J.* **2016**, *90*, 856–867. [CrossRef]

16. Sharma, A.; Soares, C.; Sousa, B.; Martins, M.; Kumar, V.; Shahzad, B.; Sidhu, G.P.; Bali, A.S.; Asgher, M.; Bhardwaj, R.; et al. Nitric oxide-mediated regulation of oxidative stress in plants under metal stress: A review on molecular and biochemical aspects. *Physiol. Plant.* **2019**, *168*, 318–344. [CrossRef] [PubMed]

17. Černý, M.; Habánová, H.; Berka, M.; Luklová, M.; Brzobohatý, B. Hydrogen Peroxide: Its Role in Plant Biology and Crosstalk with Signalling Networks. *Int. J. Mol. Sci.* **2018**, *19*, 2812. [CrossRef]

18. Umbrein, S.; Lubeja, J.; Cui, B.; Pan, Q.; Jiang, J.; Loake, G.J. Specificity in nitric oxide signalling. *J. Exp. Bot.* **2018**, *69*, 3439–3448. [CrossRef]

19. Specckmann, B.; Steinbrenner, H.; Grune, T.; Klotz, L.-O. Peroxynitrite: From interception to signaling. *Arch. Biochem. Biophys.* **2016**, *595*, 153–160. [CrossRef]

20. Ventimiglia, L.; Mutus, B. The Physiological Implications of S-Nitrosoglutathione Reductase (GSNOR) Activity Mediating NO Signalling in Plant Root Structures. *Antioxidants* **2020**, *9*, 1206. [CrossRef]

21. Lindermayr, C. Crosstalk between reactive oxygen species and nitric oxide in plants: Key role of S-nitrosoglutathione reductase. *Free. Radic. Biol. Med.* **2018**, *122*, 110–115. [CrossRef]

22. Hancock, J.; Whiteman, M. Hydrogen sulfide and cell signaling: Team player or referee? *Plant Physiol. Biochem.* **2018**, *122*, 4469–4482. [PubMed]

23. Shivaraj, S.M.; Vats, S.; Bhat, J.A.; Dhakte, P.; Goyal, V.; Kumawat, S.; Singh, A.; Prasad, M.; Sonah, H.; et al. Hydrogen and hydrogen sulfide crosstalk during heavy metal stress in plants. *Physiol. Plant.* **2019**, *168*, 435–455. [CrossRef] [PubMed]

24. Liu, F.; Jiang, W.; Han, W.; Li, J.; Liu, Y. Effects of Hydrogen-Rich Water on Fitness Parameters of Rice Plants. *Agron. J.* **2017**, *109*, 2033–2039. [CrossRef]

25. Wu, Q.; Su, N.; Huang, L.; Yu, M.; Shabala, S. Understanding the mechanistic basis of ameliorating effects of hydrogen rich water on salinity tolerance in barley. *Environ. Exp. Bot.* **2020**, *177*, 104136. [CrossRef]

26. Su, J.; Yang, X.; Shao, Y.; Chen, Z.; Shen, W. Molecular hydrogen–induced salinity tolerance requires melatonin signalling in Arabidopsis thaliana. *Plant Cell Environ.* **2021**, *44*, 476–490. [CrossRef]

27. Cui, W.; Yao, P.; Pan, J.; Dai, C.; Cao, H.; Chen, Z.; Zhang, S.; Xu, S.; Shen, W. Transcriptome analysis reveals insight into molecular hydrogen-induced cadmium tolerance in alfalfa: The prominent role of sulfur and (homo)glutathione metabolism. *BMC Plant Biol.* **2020**, *20*, 19–58. [CrossRef] [PubMed]

28. Renwick, G.M.; Giunarro, C.; Siegel, S.M. Hydrogen Metabolism in Higher Plants. *Plant Physiol.* **1964**, *39*, 303–306. [CrossRef]

29. Vargas, S.R.; Dos Santos, P.V.; Giralidi, L.A.; Zaiat, M.; Calijuri, M.D.C. Anaerobic phototrophic processes of hydrogen production by different strains of microalgae *Chlamydomonas* sp. *FEMS Microbiol. Lett.* **2018**, *365*, e00703. [CrossRef]

30. Hemschemeier, A.; Fouchard, S.; Cournac, L.; Peltier, G.; Happe, T. Hydrogen production by *Chlamydomonas reinhardtii*: An elaborate interplay of electron sources and sinks. *Planta* **2007**, *227*, 397–407. [CrossRef]

31. Sanadze, G.A. Absorption of molecular hydrogen by green leaves in light. *Fiziol. Rast.* **1961**, *8*, 555–559. [PubMed]

32. Zeng, J.; Zhang, M.; Sun, X. Molecular Hydrogen Is Involved in Phytohormone Signaling and Stress Responses in Plants. *PLoS ONE* **2013**, *8*, e71038. [CrossRef]

33. Cao, Z.; Duan, X.; Yao, P.; Cui, W.; Cheng, D.; Zhang, J.; Jin, Q.; Chen, J.; Dai, T.; Shen, W. Hydrogen Gas Is Involved in Auxin-Induced Lateral Root Formation by Modulating Nitric Oxide Synthesis. *Int. J. Mol. Sci.* **2017**, *18*, 1084. [CrossRef]

34. Jin, T.; Liu, Y.; Wei, J.; Wu, M.; Lei, G.; Chen, H.; Lan, Y. Modeling and analysis of the flammable vapor cloud formed by liquid hydrogen spills. *Int. J. Hydrogen Energy* **2017**, *42*, 26762–26770. [CrossRef]

35. Molecular Hydrogen Institute. Concentration and Solubility of H2. Available online: http://www.molecularhydrogeninstitute.com/concentration-and-solubility-of-h2 (accessed on 13 January 2021).

36. Wilhelm, E.; Battino, R.; Wilcock, R.J. Low-pressure solubility of gases in liquid water. *Chem. Rev.* **1977**, *77*, 219–262. [CrossRef]

37. Lin, Y.; Zhang, W.; Qi, F.; Cui, W.; Xie, Y.; Shen, W. Hydrogen-rich water regulates cucumber adventitious root development in a heme oxygenase-1/carbon monoxide-dependent manner. *J. Plant Physiol.* **2014**, *171*, 1–8. [CrossRef]

38. Fang, S.; Li, X.; Wei, X.; Zhang, Y.; Ma, Z.; Wei, Y.; Wang, W. Beneficial effects of hydrogen gas inhalation on a murine model of allergic rhinitis. *Exp. Ther. Med.* **2018**, *16*, 5178–5184. [CrossRef] [PubMed]
39. Dong, Z.; Wu, L.; Kettlewell, B.; Caldwell, C.D.; Layzell, D.B. Hydrogen fertilization of soils—is this a benefit of legumes in rotation? *Plant Cell Environ.* 2003, 26, 1875–1879. [CrossRef]

40. Hancock, J.T. Methods for the addition of redox compounds. In *Redox-Mediated Signal Transduction*; Hancock, J.T., Conway, M.E., Eds.; Humana: New York, NY, USA, 2019; pp. 13–25. [CrossRef]

41. Deller, M.C. Cell surface receptors. *Curr. Opin. Struct. Biol.* 2000, 10, 213–219. [CrossRef]

42. Gasc, J.M.; Baulieu, E.E. Steroid hormone receptors: Intracellular distribution. *Biol. Cell 1986*, 56, 1–6. [CrossRef]

43. Mazaírā, G.L.; Zgajnar, N.R.; Lotufo, C.M.; Daneri-Becerra, C.; Sivils, J.C.; Soto, O.B.; Cox, M.B.; Galigniana, M.D. Nuclear Receptors: A Historical Perspective. *Adv. Struct. Saf. Stud.* 2019, 1966, 1–5. [CrossRef]

44. Sandner, P.; Zimmer, D.P.; Milne, G.T.; Hobbs, A.; Stach, J.-P. Soluble Guanylate Cyclase Stimulators and Activators. In *Handbook of Experimental Pharmacology*; Springer: Berlin/Heidelberg, Germany, 2018; pp. 1–39. [CrossRef]

45. Astier, J.; Mounier, A.; Santolini, J.; Jeandroz, S.; Wendehenne, D. The evolution of nitric oxide signalling diverges between animal and green lineages. *J. Exp. Bot.* 2019, 70, 4355–4364. [CrossRef] [PubMed]

46. Smirnoff, N.; Arnaud D. Hydrogen peroxide metabolism and functions in plants. *New Phytol.* 2018, 221, 1197–1214. [CrossRef]

47. Feng, J.; Chen, L.; Zuo, J. Protein S -Nitrosylation in plants: Current progresses and challenges. *J. Integr. Plant Biol.* 2019, 61, 1206–1223. [CrossRef]

48. Petrov, V.; Hille, J.; Mueller-Roeber, B.; Gechev, T.S. ROS-mediated abiotic stress-induced programmed cell death in plants. *Front. Plant Sci.* 2015, 6, 69. [CrossRef] [PubMed]

49. Fryzova, R.; Pohanka, M.; Martinkova, P.; Cihlarova, H.; Hladky, J.; Kynicky, J. Oxidative Stress and Heavy Metals in Plants. *Rev. Environ. Contam. Toxicol.* 2015, 245, 129–156. [CrossRef]

50. Anjum, S.A.; Tanveer, M.; Hussain, S.; Bao, M.; Wang, L.; Khan, I.; Ullah, E.; Tung, S.A.; Samad, R.A.; Shahzad, B. Cadmium toxicity in Maize (*Zea mays L.*) consequences on antioxidative systems, reactive oxygen species and cadmium accumulation. *Environ. Sci. Pollut. Res.* 2015, 22, 17022–17030. [CrossRef]

51. Görski-Czekaj, M.; Borucki, W. A correlative study of hydrogen peroxide accumulation after mercury or copper treatment observed in root nodules of *Medicago truncatula* under light, confocal and electron microscopy. *Microcn* 2013, 52–54, 23–32. [CrossRef]

52. Camejo, D.; Guzmán-Cedeño, Á.; Moreno, A. Reactive oxygen species, essential molecules, during plant–pathogen interactions. *Plant Physiol. Biochem.* 2016, 103, 10–23. [CrossRef]

53. Santos, A.D.A.; Da Silveira, J.A.G.; Bonifacio, A.; Rodrigues, A.C.; Figueiredo, M.D.V.B. Antioxidant response of cowpea co-inoculated with plant growth-promoting bacteria under salt stress. *Braz. J. Microbiol.* 2018, 49, 513–521. [CrossRef] [PubMed]

54. AbdelGawad, H.; Zinta, G.; Hamed, B.A.; Selim, S.; Beemster, G.; Hozzein, W.N.; Wadaan, M.A.; Asard, H.; Abuelsoud, W.; Badredlin, A.H. Maize roots and shoots show distinct profiles of oxidative stress and antioxidant defense under heavy metal toxicity. *Environ. Pollut.* 2020, 258, 113705. [CrossRef] [PubMed]

55. Airaki, M.; Leterrier, M.; Mateos, R.M.; Valderrama, R.; Chaki, M.; Barroso, J.B.; Del Rio, L.A.; Palma, J.M.; Corpas, F.J. Metabolism of reactive oxygen species and reactive nitrogen species in pepper (*Capsicum annuum L.*) plants under low temperature stress. *Plant Cell Environ.* 2012, 35, 281–295. [CrossRef]

56. Xu, S.; Zhu, S.; Jiang, Y.; Wang, N.; Wang, R.; Shen, W.; Yang, J. Hydrogen-rich water alleviates salt stress in rice during seed germination. *Plant Soil* 2013, 370, 47–57. [CrossRef]

57. Xie, Y.; Mao, Y.; Lai, D.; Zhang, W.; Shen, W. H₂ enhances Arabidopsis salt tolerance by manipulating ZAT10/12-mediated antioxidant defence and controlling sodium exclusion. *PloS ONE* 2012, 7, e9880. [CrossRef] [PubMed]

58. Chen, M.; Cui, W.; Zhu, K.; Xie, Y.; Zhang, C.; Shen, W. Hydrogen-rich water alleviates aluminium-induced inhibition of root elongation in alfalfa via decreasing nitric oxide production. *J. Hazard. Mater.* 2014, 267, 40–47. [CrossRef]

59. Xu, D.; Cao, H.; Fang, W.; Pan, J.; Chen, J.; Zhang, J.; Shen, W. Linking hydrogen-enhanced rice aluminium tolerance with the reestablishment of GA/ABA balance and miRNA-modulated gene expression: A case study on germination. *Ecotoxicol. Environ. Saf.* 2017, 145, 303–312. [CrossRef]

60. Cui, W.; Gao, C.; Fang, P.; Lin, G.; Shen, W. Alleviation of cadmium toxicity in Medicago sativa by hydrogen-rich water. *J. Hazard. Mater.* 2013, 260, 715–724. [CrossRef]

61. Cui, W.; Fang, P.; Zhu, K.; Mao, Y.; Gao, C.; Xie, Y.; Wang, J.; Shen, W. Hydrogen-rich water confers plant tolerance to mercury toxicity in alfalfa seedlings. *Ecotoxicol. Environ. Saf.* 2014, 105, 103–111. [CrossRef]

62. Chen, Y.; Wang, M.; Hu, L.; Liao, W.; Dawuda, M.M.; Li, C. Carbon monoxide is involved in hydrogen gas-induced adventitious root development in cucumber under simulated drought stress. *Front. Plant Sci.* 2017, 8, 128. [CrossRef]

63. Jin, Q.; Zhu, K.; Cui, W.; Li, L.; Shen, W. Hydrogen-Modulated Stomatal Sensitivity to Abscisic Acid and Drought Tolerance Via the Regulation of Apoplastic pH in Medicago sativa. *J. Plant Growth Regul.* 2016, 35, 565–573. [CrossRef]

64. Jin, Q.; Zhu, K.; Cui, W.; Xie, Y.; Han, B.; Shen, W. Hydrogen gas acts as a novel bioactive molecule in enhancing plant tolerance to paraquat-induced oxidative stress via the modulation of heme oxygenase-1 signalling system. *Plant Cell Environ.* 2012, 36, 956–969. [CrossRef]

65. Hong, Y.; Chen, S.; Zhang, J.-M. Hydrogen as a Selective Antioxidant: A Review of Clinical and Experimental Studies. *J. Int. Med. Res.* 2010, 38, 1893–1903. [CrossRef] [PubMed]
66. Ohsawa, I.; Ishikawa, M.; Takahashi, K.; Watanabe, M.; Nishimaki, K.; Yamagata, K.; Katsura, K.-I.; Katayama, Y.; Asoh, S.; Ohta, S. Hydrogen acts as a therapeutic antioxidant by selectively reducing cytotoxic oxygen radicals. *Nat. Med.* 2007, 13, 688–694. [CrossRef] [PubMed]

67. Veal, E.A.; Day, A.M.; Morgan, B.A. Hydrogen Peroxide Sensing and Signaling. *Mol. Cell* 2007, 26, 1–14. [CrossRef]

68. Pastina, B.; LaVerne, J.A. Effect of Molecular Hydrogen on Hydrogen Peroxide in Water Radiolysis. *J. Phys. Chem. A* 2001, 105, 9316–9322. [CrossRef]

69. Richards, S.L.; Wilkins, K.A.; Swarbreck, S.M.; Anderson, A.A.; Habib, N.; Smith, A.G.; McAnish, M.; Davies, J.M. The hydroxyl radical in plants: From seed to seed. *J. Exp. Bot.* 2015, 66, 37–46. [CrossRef] [PubMed]

70. Pottosin, I.; Zepeda-Jazo, I.; Bose, J.; Shabala, S. An Anion Conductance, the Essential Component of the Hydroxyl-Radical-Induced Ion Current in Plant Roots. *Int. J. Mol. Sci.* 2018, 19, 897. [CrossRef]

71. Demidchik, V.; Cuiu, T.A.; Svistunenko, D.; Smith, S.J.; Miller, A.J.; Shabala, S.; Sokolik, A.; Yurin, V. Arabidopsis root K+ efflux conductance activated by hydroxyl radicals: Single-channel properties, genetic basis and involvement in stress-induced cell death. *J. Cell Sci.* 2010, 123, 1468–1479. [CrossRef]

72. Lu, W.J.; Lin, K.H.; Hsu, M.J.; Chou, D.S.; Hsiao, G.; Sheu, J.R. Suppression of NF-κB signaling by andrographolide with a novel mechanism in human platelets: Regulatory roles of the p38 MAPK-hydroxyl radical-ERK2 cascade. *Biochem. Pharmacol.* 2012, 84, 914–924. [CrossRef]

73. Qu, Y.; Yan, M.; Zhang, Q. Functional regulation of plant NADPH oxidase and its role in signaling. *Plant Signal. Behav.* 2017, 12, e1356970. [CrossRef]

74. Gill, S.S.; Anjum, N.A.; Gill, R.; Yadav, S.; Hasanuzzaman, M.; Fujita, M.; Mishra, P.; Sabat, S.C.; Tuteja, N. Superoxide dismutase—Mentor of abiotic stress tolerance in crop plants. *Environ. Sci. Pollut. Res.* 2015, 22, 10375–10394. [CrossRef]

75. Fong, K.-L.; McCoy, P.B.; Poyer, J.; Misra, H.P.; Keele, B.B. Evidence for superoxide-dependent reduction of Fe3+ and its role in enzyme-generated hydroxyl radical formation. *Chem. Interact.* 1976, 15, 77–89. [CrossRef]

76. Halliwell, B. Superoxide-dependent formation of hydroxyl radicals in the presence of iron chelates: Is it a mechanism for hydroxyl radical production in biochemical systems? *FEBS Lett.* 1978, 92, 321–326. [CrossRef]

77. Kumar, A.; Prasad, A.; Sediárová, M.; Pospíšil, P. Data on detection of singlet oxygen, hydroxyl radical and organic radical in Arabidopsis thaliana. *Data Brief* 2018, 21, 2246–2252. [CrossRef] [PubMed]

78. Chen, W.; Ding, S.; Wu, J.; Shi, G.; Zhu, A. In situ detection of hydroxyl radicals in mitochondrial oxidative stress with a nanopipette electrode. *Chem. Commun.* 2020, 56, 13225–13228. [CrossRef] [PubMed]

79. Cuypers, A.; Hendrix, S.; Dos Reis, R.A.; De Smet, S.; Deckers, J.; Gielen, H.; Jozefczak, M.; Loix, C.; Vercampt, H.; Vangronsveld, J.; et al. Hydrogen Peroxide, Signaling in Disguise during Metal Phytotoxicity. *Front. Plant Sci.* 2016, 7, 470. [CrossRef]

80. Babbs, C.F.; Pham, J.A.; Coolbaugh, R.C. Lethal Hydroxyl Radical Production in Paraquat-Treated Plants. *Plant Physiol.* 1989, 90, 1267–1270. [CrossRef]

81. Shen, B.; Jensen, R.G.; Bohnert, H.J. Mannitol Protects against Oxidation by Hydroxyl Radicals. *Plant Physiol.* 1997, 115, 527–532. [CrossRef] [PubMed]

82. Liu, F.J.; Cai, B.B.; Sun, S.N.; Bi, H.G.; Ai, X.Z. Effect of hydrogen-rich water soaked cucumber seeds on cold tolerance and its physiological mechanism in cucumber seedlings. *Sci. Agric. Sin.* 2017, 50, 881–889.

83. Yadav, D.K.; Pospíšil, P. Role of chloride ion in hydroxyl radical production in photosystem II under heat stress: Electron paramagnetic resonance spin-trapping study. *J. Bioenerg. Biomembr.* 2012, 44, 365–372. [CrossRef]

84. Chen, Q.; Zhao, X.; Lei, D.; Hu, S.; Shen, Z.; Shen, W.; Xu, X. Hydrogen-rich water pretreatment alters photosynthetic gas exchange, chlorophyll fluorescence, and antioxidant activities in heat-stressed cucumber leaves. *Plant Growth Regul.* 2017, 83, 69–82. [CrossRef]

85. Lipinski, B. Hydroxyl Radical and Its Scavengers in Health and Disease. *Oxidative Med. Cell. Longev.* 2011, 2011, 1–9. [CrossRef] [PubMed]

86. Matros, A.; Peshev, D.; Peukert, M.; Mock, H.-P.; Ende, W.V.D. Sugars as hydroxyl radical scavengers: Proof-of-concept by studying the fate of sucrose in Arabidopsis. *Plant J.* 2015, 82, 822–839. [CrossRef] [PubMed]

87. Herraz, T.; Galisteo, J. Hydroxyl radical reactions and the radical scavenging activity of β-carboline alkaloids. *Food Chem.* 2015, 172, 640–649. [CrossRef] [PubMed]

88. Sakai, T.; Imai, J.; Ito, T.; Takagaki, H.; Ui, M.; Hatta, S. The novel antioxidant TA293 reveals the role of cytoplasmic hydroxyl radicals in oxidative stress-induced senescence and inflammation. *Biochim. Biophys. Res. Commun.* 2017, 482, 1183–1189. [CrossRef]

89. Penders, J.; Kissner, R.; Koppenol, W.H. ONOOH does not react with H2: Potential beneficial effects of H2 as an antioxidant by selective reaction with hydroxyl radicals and peroxynitrite. *Free Radic. Biol. Med.* 2014, 75, 191–194. [CrossRef]

90. Buxton, G.V.; Greenstock, C.L.; Helman, P.; Ross, A.B. Critical-review of rate constants for reactions of hydrated electrons, hydrogen-atoms and hydroxyl radicals (OH/O−) in aqueous-solution. *J. Phys. Chem. Ref. Data* 1988, 17, 513–886. [CrossRef]

91. Matheson, M.S.; Rabani, J. Pulse radiolysis of aqueous hydrogen solutions. I. Rate constants for reaction of eq− with itself and other transients. II. The interconvertibility of eq− and H+. *J. Phys. Chem.* 1965, 69, 1324–1335. [CrossRef]

92. Wood, K.C.; Gladwin, M.T. The hydrogen highway to reperfusion therapy. *Nat. Med.* 2007, 13, 673–674. [CrossRef] [PubMed]

93. Guan, Q.; Ding, X.-W.; Jiang, R.; Ouyang, P.-L.; Gui, J.; Feng, L.; Yang, L.; Song, L.-H. Effects of hydrogen-rich water on the nutrient composition and antioxidative characteristics of sprouted black barley. *Food Chem.* 2019, 299, 125095. [CrossRef] [PubMed]
1. Li, C.; Huang, D.; Wang, C.; Wang, N.; Yao, Y.; Li, W.; Liao, W. NO is involved in H2-induced adventitious rooting in cucumber

2. He, H.; He, L. Heme oxygenase 1 and abiotic stresses in plants.

3. Schubert, K.R.; Evans, H.J. Hydrogen evolution: A major factor affecting the efficiency of nitrogen fixation in nodulated symbionts.

4. Wegiel, B.; Nemeth, Z.; Correa-Costa, M.; Bulmer, A.C.; Otterbein, L.E. Heme Oxygenase-1: A Metabolic Nike.

5. Wilks, A. Heme Oxygenase: Evolution, Structure, and Mechanism. Antioxid. Redox Signal.

6. Wheeler, J.A.; Cortés, A.J.; Sedlacek, J.; Cortés, A.J.; Rixen, C. Small-scale drivers: The importance of nutrient availability and snowmelt timing on performance of the alpine shrub Salix herbacea. Oecologia 2015, 180, 1015–1024.

7. Hogg, N.; Darley-Usmar, V.M.; Wilson, M.T.; Moncada, S. Production of hydroxyl radicals from the simultaneous generation of superoxide and nitric oxide. Biochem. J. 1992, 281, 419–424.

8. Vandelle, E.; Delledonne, M. Peroxynitrite formation and function in plants. Plant Sci. 2011, 181, 534–539.

9. Staszek, P.; Gniazdowska, A. Peroxynitrite induced signaling pathways in plant response to non-proteinogenic amino acids. Planta 2020, 252, 1–11.

10. Kolbert, Z.; Feigl, G.; Bordé, A.; Molnár, Á.; Erdei, L. Protein tyrosine nitration in plants: Present knowledge, computational prediction and future perspectives. Plant Physiol. Biochem. 2017, 113, 56–63.

11. Alvarez, B.; Radi, R. Peroxynitrite reactivity with amino acids and proteins. Amino Acids 2003, 25, 295–311.

12. Hanaoka, T.; Kamimura, N.; Yokota, T.; Takai, S.; Ohta, S. Molecular hydrogen protects chondrocytes from oxidative stress and indirectly alters gene expressions through reducing peroxynitrite derived from nitric oxide. Med. Gas Res. 2011, 1, 18.

13. Shen, N.Y.; Bi, J.B.; Zhang, J.Y.; Zhang, S.M.; Gu, J.X.; Qu, K.; Liu, C. Hydrogen-rich water protects against inflammatory bowel disease in mice by inhibiting endoplasmic reticulum stress and promoting heme oxygenase-1 expression. World J. Gastroenterol. 2017, 23, 1375.

14. Cortés, A.J.; Chavarro, C.M.; Madriñán, S.; This, D.; Blair, M.W. Molecular ecology and selection in the drought-related Asr gene polymorphisms in wild and cultivated common bean (Phaseolus vulgaris L.). BMC Genet. 2012, 13, 8.

15. Cortés, A.J.; This, D.; Chavarro, C.; Madriñán, S.; Blair, M.W. Nucleotide diversity patterns at the drought-related DREB2 encoding genes in wild and cultivated common bean (Phaseolus vulgaris L.). Theor. Appl. Genet. 2012, 125, 1069–1085.

16. Blair, M.W.; Cortés, A.J.; This, D. Identification of an ERECTA gene and its drought adaptation associations with wild and cultivated common bean. Plant Sci. 2016, 242, 250–259.

17. Cortés, A.J.; Monserrate, F.A.; Ramírez-Villegas, J.; Madriñán, S.; Blair, M.W. Drought Tolerance in Wild Plant Populations: The Case of Common Beans (Phaseolus vulgaris L.). PLoS ONE 2013, 8, e62898. [CrossRef]

18. Cortés, A.J.; Blair, M.W. Genotyping by Sequencing and Genome–Environment Associations in Wild Common Bean Predict Widespread Divergent Adaptation to Drought. Front. Plant Sci. 2018, 9, 128. [CrossRef]

19. Wheeler, J.A.; Cortés, A.J.; Sedlacek, J.; Karrenberg, S.; Van Kleunen, M.; Wipf, S.; Hoch, G.; Bossdorf, O.; Rixen, C. The snow and the willows: Earlier spring snowmelt reduces performance in the low-growing alpine shrub Salix herbacea. J. Ecol. 2016, 104, 1041–1050. [CrossRef]

20. López-Hernández, F.; Cortés, A.J. Last-Generation Genome–Environment Associations Reveal the Genetic Basis of Heat Tolerance in Common Bean (Phaseolus vulgaris L.). Front. Genet. 2019, 10, 954. [CrossRef]

21. Wheelier, J.A.; Hoch, G.; Cortés, A.J.; Sedlacek, J.; Wipf, S.; Rixen, C. Increased spring freezing vulnerability for alpine shrubs under early snowmelt. Oecologia 2014, 175, 219–229. [CrossRef]

22. He, H.; He, L. Heme oxygenase 1 and abiotic stresses in plants. Acta Physiol. Plant. 2013, 36, 581–588. [CrossRef]

23. Wilks, A. Heme Oxygenase: Evolution, Structure, and Mechanism. Antioxid. Redox Signal. 2002, 4, 603–614. [CrossRef]

24. Wegiel, B.; Nemeth, Z.; Correa-Costa, M.; Bulmer, A.C.; Otterbein, L.E. Heme Oxygenase-1: A Metabolic Nike. Antioxid. Redox Signal. 2014, 20, 1709–1722. [CrossRef]

25. Rychlewski, J. Magnetic effects for the hydrogen molecule in excited states: $b^3\Sigma^+$ of H$_2$. Mol. Phys. 1986, 59, 327–336. [CrossRef]
124. Rychlewski, J. Magnetic effects for the hydrogen molecule in excited states: B1 Σu+ of H2. *Phys. Rev. A Gen. Phys.* **1985**, *31*, 2091–2095. [CrossRef]

125. Steiner, U.E.; Ulrich, T. Magnetic field effects in chemical kinetics and related phenomena. *Chem. Rev.* **1989**, *89*, 51–147. [CrossRef]

126. Buntkowsky, G.; Walaszek, B.; Adamczyk, A.; Xu, Y.; Limbach, H.-H.; Chaudret, B. Mechanism of nuclear spin initiated para-H2 to ortho-H2 conversion. *Phys. Chem. Chem. Phys.* **2006**, *8*, 1929–1935. [CrossRef] [PubMed]

127. Hancock, J.T.; Hancock, T.H. Hydrogen gas, ROS metabolism, and cell signaling: Are hydrogen spin states important? *React. Oxyg. Species* **2018**, *6*, 389–395. [CrossRef]

128. Marais, A.; Adams, B.; Ringsmuth, A.K.; Ferretti, M.; Gruber, J.M.; Schuld, M.; Smith, S.L.; Sinayskiy, I.; Krüger, T.P.; et al. The future of quantum biology. *J. R. Soc. Interface* **2018**, *15*, 20180640. [CrossRef]

129. Kim, Y.; Bertagna, F.; D'Souza, E.M.; Heyes, D.J.; Johannisen, L.O.; Nery, E.T.; Pantelias, A.; Jimenez, A.S.-P.; Slocombe, L.; Spencer, M.G.; et al. Quantum Biology: An Update and Perspective. *Quantum Rep.* **2021**, *3*, 80–126. [CrossRef]

130. Huang, C.-S.; Kawamura, T.; Toyoda, Y.; Nakao, A. Recent advances in hydrogen research as a therapeutic medical gas. *Free. Radic. Res.* **2010**, *44*, 971–982. [CrossRef]

131. Wang, Y.-Q.; Liu, Y.-H.; Wang, S.; Du, H.-M.; Shen, W.-B. Hydrogen agronomy: Research progress and prospects. *J. Zhejiang Univ. Sci. B* **2020**, *21*, 841–855. [CrossRef]

132. Chuai, Y.; Gao, F.; Li, B.; Zhao, L.; Qian, L.; Cao, F.; Wang, L.; Sun, X.; Cui, J.; Cai, J. Hydrogen-rich saline attenuates radiation-induced male germ cell loss in mice through reducing hydroxyl radicals. *Biochem. J.* **2012**, *442*, 49–56. [CrossRef]

133. Halliwell, B.; Gutteridge, J.M. The deoxyribose method: A simple “test-tube” assay for determination of rate constants for reactions of hydroxyl radicals. *Anal. Biochem.* **1987**, *165*, 215–219. [CrossRef]

134. Xu, G.; Chance, M.R. Hydroxyl Radical-Mediated Modification of Proteins as Probes for Structural Proteomics. *Chem. Rev.* **2007**, *107*, 3514–3543. [CrossRef]

135. El-Bahr, S.M. Biochemistry of Free Radicals and Oxidative Stress. *Sci. Int.* **2013**, *1*, 111–117. [CrossRef]

136. Tejero, I.; González-Lafont, À.; Lluch, J.M.; Eriksson, L.A. Theoretical Modeling of Hydroxyl-Radical-Induced Lipid Peroxidation Reactions. *J. Phys. Chem. B* **2007**, *111*, 5684–5693. [CrossRef] [PubMed]

137. Gilbert, B.C.; King, D.M.; Thomas, C. The oxidation of some polysaccharides by the hydroxyl radical: An e.s.r. investigation. *Carbohydr. Res.* **1984**, *125*, 217–235. [CrossRef]

138. Dizdaroglu, M.; Jaruga, P. Mechanisms of free radical-induced damage to DNA. *Free Radic. Res.* **2012**, *46*, 382–419. [CrossRef] [PubMed]

139. Gansäuer, A.; Otte, M.; Piester, F.; Fan, C.-A. Sustainable radical reduction through catalyzed hydrogen atom transfer reactions (CHAT-reactions). *Tetrahedron* **2009**, *65*, 4984–4991. [CrossRef]