Mutational status of main driver genes influences the prognosis of stage I–III lung adenocarcinoma patients underwent radical surgery

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Background: The therapeutic strategies and prognosis of local advanced and metastatic lung cancer have been extensively investigated. However, the prognosis of early-stage lung cancer patients undergoing radical surgery has not been fully studied due to the difficulties in follow-up and assessment.

Methods: We recruited 447 stage I–III lung adenocarcinoma (LUAD) patients who underwent radical surgery and investigated the influence of main driver gene mutations and clinicopathological factors on patient overall survival (OS). Cancer tissue samples were collected retrospectively and mutational status and tumor mutational burden (TMB) were determined by whole-exome sequencing (WES).

Results: Distinct stage-dependent mutational frequency was revealed in main driver genes including EGFR, TP53, KRAS, STK11, ATM and NF1. Patients with TP53 mutations exhibited a trend of better survival than those with wild type TP53 (P=0.066), and STK11 mutations exhibited worse survival in stage III patients (P=0.031). EGFR mutations eliminated the across-stage difference in survival, which was still present in other wild type and mutant driver genes. Furthermore, patients with wild type TP53 appeared to have significantly worse survival than patients with other wild type driver genes in stage I (P<0.001). TMB cannot stratify the survival of LUAD patients in stage I–III. Age, gender, smoking status, smoking years, prior cancer history and cancer location had no stratification effect on patient survival, while T grading (P<0.001) and N grading (P<0.001) had significant stratification on survival.

Conclusions: TP53, EGFR and STK11 mutational status influenced the prognosis of stage I–III LUAD. T and N grading also stratified the patient survival. T grading was an independent risk factor.

Keywords: Lung cancer; non-small cell lung cancer (NSCLC); adenocarcinoma; tumor mutational burden (TMB); EGFR

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Introduction

Lung cancer is the most common types of cancer with highest morbidity and mortality in the world (1). Great progress has been made in the development of therapeutic strategies for local advanced or metastatic lung cancer, including targeted therapy, immunotherapy and combined therapy (2). Late-stage lung cancer has attached much attention and clinical research resource due to the therapeutic difficulties and its high mortality and complications. In contrast, early stage lung cancer, especially for those who have opportunity for radical surgery, was not the focus of systematic therapy, although post-surgical adjuvant therapy is needed for these patients. Recent advances in adjuvant therapy by tyrosine kinase inhibitors (TKIs) for surgically resected lung cancer rendered new therapeutic strategies for early-stage lung cancer patients and enhanced patients recurrent free survival (RFS) and potentially overall survival (OS) (3-5).

The correlation between mutational landscape and patient prognosis in late-stage lung cancer patients has been extensive investigated in various scenarios including targeted therapy, immunotherapy and combined therapy (6,7). Main driver gene mutations and a series of markers, including EGFR, KRAS, TP53 and TMB have been suggested as stratification markers for therapeutic response or prognosis in systematic therapy (8,9). Since radical surgery is the main therapeutic strategy for early-stage lung cancer, the correlation between these markers and the response or prognosis of early-stage patients has not been thoroughly investigated.

In order to clarify the relationship between main driver gene mutational status and the prognosis of early-stage lung cancer patients, we designed a study and recruited 447 stage I–III Chinese lung adenocarcinoma (LUAD) patients who underwent radical surgery, and examined the mutational landscape of these patients by whole-exome sequencing (WES). We investigated the stratification of prognosis by main driver gene mutations and discovered interesting correlation between mutational status and prognosis. The clinicopathological factors were also examined with the prognosis of these patients.

We present the following article in accordance with the REMARK reporting checklist (available at https://dx.doi.org/10.21037/tcr-21-240).

Methods

Ethical statement

The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the ethics committee of Yuebei People's Hospital and the First Affiliated Hospital of Jinan University (NO. 2019YBPH0518) and individual consent for this retrospective analysis was waived.

Study design, patients and samples

The study was designed and implemented in Yuebei People’s Hospital and the First Affiliated Hospital of Jinan University. Cancer tissue and blood samples were collected retrospectively. The study was designed to include stage IA-IIIB (T1-4, N0-2, M0) LUAD patients who received radical surgery as the first-line therapy (Table 1). The staging of lung cancer patients was based on the Eighth Edition of the TNM Classification of Lung Cancer. The primary outcomes included the difference in OS among different groups of patients based on mutational status, stage, tumor mutational burden (TMB) and clinicopathological factors. The secondary outcome involved stage-dependent mutational frequency. Patients were followed up to 7,248 days (median at 658 days, ranging from 4 to 7,248 days). Samples from 447 qualified lung adenocarcinoma patients were obtained and the demographic and clinicopathological information are shown in Table 1. Samples consisted of frozen tissue from surgery or needle biopsy and FFPE samples. Samples with cancer cell content more than 20% were sent for WES test. All technicians were blinded to subject clinical information. Confirmation of pathological types was based on diagnosis from imaging examinations and subsequent pathological examinations. None of the subjects received chemotherapy, radiotherapy, targeted therapy, or immunotherapy before tissue samples were collected. The somatic sequencing data presented in this study were from lung tumor tissue DNA, with germline sequencing data from the corresponding peripheral blood lymphocytes (PBLs) as the control.

Sample preparation, targeted NGS and data processing for WES

DNA was extracted from the fresh tissue, FFPE tissue or needle biopsy samples using the QIAamp DNA Tissue Kit (QIAGEN, Valencia, CA, USA) following the manufacturer’s instructions. 2 ml of peripheral blood was collected and DNA was extracted from PBLs using the RelaxGene Blood DNA system (Tiangen Biotech Co., Ltd., Beijing, China). Both cancer tissue and white blood
Table 1 Demographic and clinicopathological information for all patients in this study

| Items                      | Number of patients |
|----------------------------|--------------------|
| Sex                        |                    |
| Male                       | 207                |
| Female                     | 240                |
| Age                        |                    |
| 30–39                      | 3                  |
| 40–49                      | 35                 |
| 50–59                      | 117                |
| 60–69                      | 149                |
| 70–79                      | 122                |
| ≥80                        | 21                 |
| T grading                  |                    |
| T1                         | 152                |
| T2                         | 235                |
| T3                         | 43                 |
| T4                         | 17                 |
| N grading                  |                    |
| N0                         | 304                |
| N1                         | 84                 |
| N2                         | 59                 |
| Clinical staging           |                    |
| I                          | 279                |
| II                         | 124                |
| III                        | 44                 |
| Location                   |                    |
| Upper lung                 | 266                |
| Middle lung                | 26                 |
| Lower lung                 | 155                |
| Cancer history             |                    |
| Prior malignancy           | 80                 |
| No prior malignancy        | 367                |
| Smoking status             |                    |
| Non-smoker                 | 142                |
| Smoker <40 packs/year      | 151                |
| Smoker ≥40 packs/year      | 154                |

Table 1 (continued)

| Items                      | Number of patients |
|----------------------------|--------------------|
| Smoking years              |                    |
| 0 years (non-smoker)       | 142                |
| <33 years                  | 149                |
| ≥33 years                  | 156                |
| Mutational status          |                    |
| EGFR                       |                    |
| mut                        | 176                |
| WT                         | 271                |
| TP53                       |                    |
| mut                        | 191                |
| WT                         | 256                |
| KRAS                       |                    |
| mut                        | 72                 |
| WT                         | 375                |
| STK11                      |                    |
| mut                        | 27                 |
| WT                         | 420                |
| ATM                        |                    |
| mut                        | 29                 |
| WT                         | 418                |
| NF1                        |                    |
| mut                        | 34                 |
| WT                         | 413                |
| Adjuvant therapy           |                    |
| Chemotherapy               | 53                 |
| Chemo/radiotherapy         | 22                 |
| TKI therapy                | 48                 |
| Unknown                    | 324                |

Cell genomic DNA was quantified with the Qubit 2.0 Fluorometer and the Qubit dsDNA HS assay kit (Thermo Fisher Scientific, Inc., Waltham, MA, USA) according to manufacturer’s instructions. In brief, fragmented genomic DNA underwent end-repairing, A-tailing and ligation with
indexed adapters sequentially, followed by size selection using Agencourt AMPure XP beads (Beckman Coulter Inc., Brea, CA, USA), and DNA fragments were used for library construction with the KAPA Library Preparation kit (Kapa Biosystems, Inc., Wilmington, MA, USA) according to the manufacturer’s protocol. Hybridization-based target enrichment was conducted with standard WES panel (HaploX Biotechnology, Shenzhen, China) for cancer tissue sequencing. Seven to eight PCR cycles, depending on the amount of DNA input, were performed on Pre-LM-PCR Oligos (Kapa Biosystems, Inc.) in 50 μL reactions. The DNA sequencing was then performed on the Illumina Novaseq 6000 system according to the manufacturer’s instructions at an average depth of 300x.

Sequencing data were de-multiplexed and aligned to the hg38 genome (GRCh38) using Burrows-Wheeler Aligner (http://bio-bwa.sourceforge.net/) version 0.7.15-r1140 with default settings. Pileup files for properly paired reads with mapping quality ≥60 were generated using Samtools (http://www.htslib.org/). Somatic variants were determined using VarScan2 (http://varscan.sourceforge.net/). Allele frequencies were calculated for all Q30 bases. Using a custom Python script, previously identified tumor DNA mutations were intersected with a Samtools mpileup file generated for each sample, and the number and frequency were then calculated for each mutation. A mutation was declared if ≥5 mutant reads were identified, and ≥1 mutant read was found on each strand. Matched genomic DNA from white blood cells was used as the control.

Statistical analysis

Statistical analysis was performed, and figures were plotted with GraphPad PRISM version 5.0 software (GraphPad Software, Inc, La Jolla, CA 92037, USA). Student t-test or non-parametric test was performed when two groups were compared, and ANOVA and post hoc tests were performed when three or more groups were compared. Chi-square test and Fisher test were performed when rate or percentage was compared for significance. Kaplan-Meier analysis was performed and the survival curves were compared using the Log-rank test. ‘*’ represents P<0.05 (significant), ‘**’ represents P<0.01 (highly significant), and ‘***’ represents P<0.001 (very highly significant). Tissue TMB was calculated by dividing the total number of tissue non-synonymous SNV and INDEL variations (allele frequency > 2%) by the full length of the WES panel.

Results

Mutational status of main driver genes influenced the prognosis of stage I–III LUAD patients

The mutational status of stage I–III LUAD patients was examined first. Distinct stage-dependent mutational frequency was revealed for main driver genes. Figure 1A shows the distinct mutational frequency (size of dots) and variant allele frequency (color of dots) across many genes. The highly mutated driver genes include TP53, KRAS, EGFR, etc. We then examined the mutational frequency of representative driver genes for each stage. It can be seen from Figure 1B that the mutational frequency of EGFR in stage I was significantly higher than that of the stage II and III. In contrast, the TP53 mutational frequency increased with the elevation of stage, while KRAS exhibited higher frequency in stage II than stage I and III. Other driver genes, such as STK11, ATM and NF1, also exhibited characteristic mutational frequency across stages. These observations suggest that the main driver genes were differentially mutated across stages of LUAD in Chinese population.

We further investigated whether the distinct stage-dependent mutational frequency was correlated with patient long-term survival. Figure 2 shows the stratification of patient survival by mutational status (wild type or mutant) across each stage for the main driver genes. When all stage I–III patients were involved (Figure 2A), no significant stratification by mutational status was observed for main driver genes, although a trend of better survival for wild type EGFR (P=0.117) and mutant TP53 (P=0.066) can be found. When stage I, II or III was studied individually (Figure S1), significant stratification was only observed in STK11 in stage III (P<0.031), although a trend of stratification was also observed in other genes and other stages, as indicated by the P values in individual panels. Cancer stage is a widely accepted stratification factor for survival, and higher stage generally leads to worse survival. We therefore compared the across-stage survival in wild type and mutant driver genes (Figure 2B,C). It is interesting to find that the stratification by stage vanished in EGFR mutant (Figure 2B, first row), while still existed in other genes, regardless of mutational status. These observations suggested that TP53, STK11 and EGFR mutations may affect the long-term survival of LUAD patients.

The stratification by different driver genes was also investigated. Figure 3 shows the comparison of survival
across wild type (upper row) or mutant (lower row) driver genes. It was interesting to find that stage I–III patients with wild type \textit{TP53} exhibited significantly worse survival (P<0.001) than patients with other wild type genes. Further analysis showed that this difference in survival originated from the worse survival in stage I patients with wild type \textit{TP53}, not the stage II and stage III patients. In contrast, no significant difference in survival was observed across mutant driver genes in stage I–III patients (Figure 3 lower row).

However, a trend of worse survival was observed in stage I with \textit{EGFR} or \textit{TP53} mutations than those with \textit{KRAS} or \textit{ATM} mutations (P=0.05).

Tumor mutational burden (TMB) is a widely accepted marker in first and multiple lines of immunotherapy for local advanced or metastatic lung cancer. Here we examined the correlation between TMB and survival in stage I–III LUAD patients. No significant difference in TMB was observed across stage I, II and III (Figure 4A). When TMB

![Figure 1](https://example.com/figure1.png)  
**Figure 1** The mutational frequency of main driver gene mutations in lung adenocarcinoma. (A) demonstration of the mutational frequency and variant allele frequency (VAF) of mutations in adenocarcinoma. Size of dots indicates mutations frequency and color indicates the VAF. (B) the mutational frequency of \textit{EGFR}, \textit{TP53}, \textit{KRAS}, \textit{STK11}, \textit{ATM} and \textit{NF1} and a comparison of frequency among stage I–III cancer. *, P<0.05; **, P<0.01; ***, P<0.001.
Figure 2 Stratification of prognosis based on Kaplan-Meier analysis of main driver gene mutations. Stratification of survival in stage I–III adenocarcinoma patients based on EGFR, TP53, KARS, STK11, ATM and NF1 mutational status (A). Comparison of stage-related survival according to the mutational status (B,C). P values are labeled for those with significant or close-to-significant differences.
=10 was defined as the threshold for high and low TMB, no statistically significant stratification by TMB was observed in stage I–III patients or each individual stage (Figure 4B), although a trend of better survival with high TMB may exist in stage III (P=0.165).

Clinicopathological factors influenced the prognosis of stage I–III patients

The influence of patient demographic factors, lifestyle factors, cancer history and cancer pathological factors on
survival was also investigated. It can be seen from Figure 5 that age and gender had no stratification effect on survival, although a trend of younger patients with better survival may exist (P=0.179). Although smoking status is a definite risk factor for lung cancer incidence, it appeared to have no influence on patient survival. The duration of smoking may have an influence (P=0.111), but this may be related to patient age. Furthermore, prior cancer history and tumor location did not affect the patient survival. However, patients with ≤5 cm tumor (T1 and T2) had better survival than those with >5 cm tumor (T3 and T4) (P<0.001), and patients with no lymph node metastasis (N0) had better survival than those with lymph node metastasis (N1 and N2) (P<0.001). These observations suggest that T and N grading predicted the survival of stage I–III LUAD patients.

Both univariate and multivariate regression analyses were performed with clinicopathological and mutational factors, including clinical stage, pathological T grading, pathological N grading, EGFR mutations, TP53 mutations and STK11 mutations. It can be clearly seen from Table 2 that clinical stage and pathological T grading were risk factors in univariate analysis, while pathological T grading was the only independent risk factor for stage I–III LUAD patients in multivariate analysis, with a hazard ratio (HR) of 1.675 (95% CI: 1.148–2.442).

**Discussion**

**Influence of main driver gene mutational status on patients therapeutic response and prognosis**

The influence of driver gene mutations in local advanced or metastatic lung cancer has been intensively investigated in targeted therapy and immunotherapy. EGFR mutations have been correlated with better survival as TKIs can be used for patients with the mutations (10). KRAS mutations are also targets for TKIs in both colorectal cancer and lung cancer (11,12). Since systematic therapy was not conventionally used for patients with early stage lung cancer undergoing radical surgery, the roles of these driver gene mutations in therapy and survival of these patients have not been investigated in detail. In this study, we investigated the potential roles of main driver gene mutations in OS stratification in stage I–III LUAD patients.

We identified distinct stage-dependent mutational
frequency for main driver genes in this study in Chinese population, suggesting that the frequency of mutations in each stage was not consistent. EGFR appeared to have higher mutational frequency in stage I than stage II and III, while the mutational frequency of TP53 and ATM increased with stage, and stage II had the highest mutational frequency in KRAS, STK11 and NF1. The reason is not clear for the stage-dependent difference and across-gene differences in mutational frequency, but it may be related to distinct clonal expansion during carcinogenesis, reflecting the Darwin evolution and subsequent parallel evolution in tumor development (13,14). Some clones containing EGFR may expand very early in carcinogenesis, and therefore exhibited a high mutational ratio in early stage cancers, but its ratio decreased when other clones start to expand at later cancer stages, such as those with TP53 and ATM. Clones with KRAS, STK11 and NF1 may preferentially expand at stage II, and parallel evolved at later stages.

The influence of the distinct stage-relevant mutational frequency was investigated in our study. It was interesting to find that EGFR mutations eliminated the difference in survival among stage I–III patients, which appeared to be a protective effect for patients with EGFR mutations at higher stages, while this effect was not seen in other wild type and mutant genes. Patients involved in this study had conventional post-surgical adjuvant therapy but not TKI-based therapy (3-5), therefore, it can be suggested this effect was not due to post-surgical treatment but EGFR itself. However, recent strong evidence suggested disease-free survival was significantly longer among patients who received Osimertinib as post-surgical adjuvant therapy than among those who received placebo for stage IB to IIIA EGFR mutation-positive non-small cell lung cancer (NSCLC) (3-5). This opens a new era for adjuvant TKI therapy in early stage lung cancer patients with radical surgery, which may significantly enhance PFS and OS for these patients.

It was also interesting to find that wild type TP53 exhibited worse survival than their counterparts of other genes in stage I alone, while this was not that obvious in mutant TP53. TP53 mutations also exhibited a trend of better survival than its wild type. These observations suggested that wild type TP53 was an indicator for worse survival in stage I LUAD. Indeed, the amount of TP53 mutations was found to be positively correlated with TMB, which has been suggested as a predicting indicator.
for survival of early-stage lung cancer patients (15,16). Therefore, less TP53 mutations may indicate poorer survival in this scenario. The correlation among main driver gene mutations and the roles of their co-mutations in survival prediction are worth more investigation in early stage patients.

**Influence of TMB on patient prognosis**

TMB has been extensively reported as one effective marker for stratifying the therapeutic response in first-line or multiple lines of immunotherapy in local advanced and metastatic cancer patients including lung malignancies (15,17). This is mainly due to the fact that mutational burden is in positive correlation with the amount of cancer neoantigens, generating tumor immunogenicity and conditioning the response to ICIs (6). Patients with higher TMB were found to benefit more from immunotherapy than those with lower TMB (18). More recently, it was reported that TMB subtyping can predict the prognosis of first-line chemotherapy in local advanced and metastatic NSCLC patients, in which higher TMB correlated with worse PFS and OS in chemotherapy (19). In contrast to the extensive investigation of TMB in systematic therapy, its roles in early-stage lung cancer with radical surgery has not been fully defined. Although we did not find significant difference in survival between high and low TMB patients in our study, a trend of higher TMB with better survival can be observed, which is consistent with a recent study with large TCGA cohort (20). This is interesting in early-stage lung cancer patients with radical surgery, since our observation and recent evidence suggest that TMB itself is capable of predicting the OS of these patients, regardless of the strategy of post-surgical adjuvant therapy. Similar correlation can also be observed in stage II colon cancer patients with surgical resection, in which mismatch repair deficiency and high TMB have been utilized as good prognostic biomarkers (21). The reason for this correlation has not been clearly defined, but it was suggested that high neoantigen burden and intense TIL infiltration may be associated with favorable survival outcomes in early-stage lung cancer (22,23). Indeed, high TMB may reflect the individual immunogenicity and mediate the profile of tumor-host immune interactions.

**Influence of clinicopathological factors on patient prognosis**

Smoking is a widely accepted risk factor for lung cancer and increases lung cancer incidence. However, whether smoking affects the patient’s long-term survival has not been fully investigated. We found no difference in OS between non-smoker and smoker in stage I–III LUAD patients, but this does not mean that smoking has no influence on survival of lung cancer patients, as evidence suggest that ever-smokers exhibited significantly lower RFS rate than never smokers (24). We also found a trend of better OS in patients with less smoking years. Since young smokers tend to have fewer smoking years, it is not known whether the trend was caused by age effect.

It appeared that previous cancer history did not affect the survival of stage I–III LUAD patients. The possibility of second primary lung cancer should always be considered

| Variables          | Category       | Univariate HR | 95% CI       | P value | Multivariate HR | 95% CI       | P value |
|--------------------|----------------|---------------|--------------|---------|----------------|--------------|---------|
| Clinical stage     | I, II, II      | 1.170         | 0.951–1.438  | 0.020   | 1.182          | 0.852–1.634  | 0.310   |
| Pathological T     | T1, T2, T3, T4 | 1.379         | 1.086–1.751  | 0.000   | 1.675          | 1.148–2.442  | 0.000   |
| Pathological N     | N0, N1, N2     | 0.902         | 0.576–1.413  | 0.161   | 0.879          | 0.539–1.433  | 0.194   |
| EGFR mutation      | Mutant/WT      | 0.932         | 0.716–1.190  | 0.538   | 0.964          | 0.738–1.259  | 0.788   |
| TP53 mutation      | Mutant/WT      | 0.892         | 0.767–1.039  | 0.142   | 0.874          | 0.731–1.016  | 0.127   |
| KRAS mutation      | Mutant/WT      | 1.032         | 0.721–1.477  | 0.864   | 1.056          | 0.743–1.481  | 0.795   |
| STK11 mutation     | Mutant/WT      | 1.034         | 0.725–1.238  | 0.596   | 1.046          | 0.695–1.256  | 0.629   |
| ATM mutation       | Mutant/WT      | 0.971         | 0.536–1.762  | 0.924   | 0.934          | 0.506–1.729  | 0.882   |
| NF1 mutation       | Mutant/WT      | 1.035         | 0.654–1.638  | 0.884   | 1.013          | 0.628–1.617  | 0.835   |
when new lesions are found in patients with previous cancer history. It has been reported that patients with secondary lung cancer associated other primary malignancies have non-inferior survival than those with single lung cancer (25). However, metachronous primary cancers may represent higher susceptibility to cancer for individuals, and more intensive screening for other cancers should be considered. If cancers are found at early stages and appropriate therapeutic strategies are implemented, the survival may not be different to those patients with only one primary cancer. Moreover, for those with family history of cancers, screening for germline mutations should be considered, as it has been shown that pathogenic or likely pathogenic germline mutations increased the risk of lung cancer (26).

Location appeared to have no influence on patient survival in our study, while it was reported that a small proportion of lung cancer patients have synchronous multiple primary lung cancer (SMPLC) or intrapulmonary metastasis, representing two distinct conditions of multiple lung lesions (27-29). One recent meta-analysis found that the pooled 5-year OS was 45% for SMPLC patients and 62% for stage I lung cancer, which was different from the 5-year OS of 93% of patients with multifocal ground-glass/lepidic (GG/L) lung cancers. The poor prognostic factors for SMPLC were lymph node metastasis and pneumonectomy, whereas histology, laterality, sublobar resection and adjuvant therapy were not found to influence the outcome (27). Therefore, it seems that the long-term prognosis of SMPLC patients after surgery is dependent on staging and metastasis, and is acceptable in patients with early-stage disease. SMPLC should be distinguished from multifocal GG/L lung cancer to perform accurate surgical evaluation.

T and N grading have long been known to be factors affecting the survival of lung cancer patients (30). We found in this study that patients with less than 5 cm lesions and with no lymph node metastasis had significantly better survival, which confirmed that early stage cancer with no lymph node metastasis have better survival than late stage cancer with lymph node metastasis. Furthermore, T grading was the only independent risk factor for stage I–III LUAD patients, although TP53, EGFR and STK11 mutational status, T and N grading all appeared to affect the prognosis. This observation suggested that N grading may be correlated with T grading to influence prognosis.

This study had some limitations. The follow-up information was not complete in this study. This was because a large majority of patients left our hospital after surgery and went back to local hospitals for subsequent systematic therapy, including chemotherapy, chemoradiotherapy and TKI therapy (Table 1). This made it difficult for us to collect information on therapeutic response assessment, progression free survival and cancer recurrence. Therefore, it was not possible to ensure enough statistical confidence for stratification of survival by postsurgical therapy or progression free survival due to the incomplete information.

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**Footnote**

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**Ethical Statement:** The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the ethics committee of Yuebei People’s Hospital and the First Affiliated Hospital of Jinan University
(NO. 2019YBPH0518) and individual consent for this retrospective analysis was waived.

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