A Mechanistic Overview on Impact of Dietary Fibres on Gut Microbiota and Its Association with Colon Cancer

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Abstract: Colorectal cancer (CRC) is an abnormal growth that occurs in the rectum or rectal portion. In 2020, an anticipated 104,610 new cases of colon illness and 43,340 new cases of rectal abnormal growth were expected in the United States. Red and processed meat, body bloatedness, belly fatness, and binge drinking expands the occurrence of colorectal disease. Dietary fibres contribute to faecal bulking, but they are break down by gut bacteria and produce metabolites such as short-chain fatty acids (SCFAs). SCFAs are chemical compounds that are mostly made up of acetate, propionate, and butyrate. Acetate and butyrate help to control mucus production and discharge, and thus, protect the gut mucosa. Reduced mucus secretion/increased bacterial catabolism, and fermentation of amino acids resulted in an increase of potentially detrimental metabolites such as branched-chain fatty acids, ammonia, amines, and N-nitroso complex components. Vital roles of fibres include reduction in the time that carcinogens encounter the intestinal lumen and promotion of healthy gut microbiota as well as modification of the host metabolism. The present review focuses on a brief introduction to various dietary fibres and specialised metabolites that can possess beneficial effect on CRC, as well as presenting our current, detailed understanding of various dietary fibres along with their potential effects on gut microbiota and its association with the colon cancer. A comprehensive discussion is also included, advocating the dietary fibre-enriched diet.

Keywords: carbohydrates; colorectal cancer; diet; flavonoids; probiotics; SCFA

1. Introduction

Cancer is a serious public health issue that affects people globally. It is a disease in which abnormal cells expand uncontrollably, and is among the leading cause of death, with an estimated 10 million deaths projected by 2020 [1]. In 2022, the United States is predicted to have 1,918,030 new cancer cases and 609,360 cancer deaths, with around 350 deaths daily due to lung cancer [2]. The coronavirus disease 2019 (COVID-19) pandemic had a negative impact on cancer detection and therapy in 2020 [2,3].

Colorectal cancer (CRC) is the occurrence of tumour development in the colon or rectum. The colon, rectum (coloanum), and anus are the final sections of the gastrointestinal (GI) tract [1]. In the United States, an estimated 104,610 new cases of colon cancer and 43,340 new cases of rectal cancer were diagnosed in 2020. In the same year an estimated 53,200 people would die from CRC, with 3640 men and women below 50 years [4]. There is lack of concrete statistics on mortalities from colon and rectal cancers separately, due to the fact that around 40% of rectal cancer deaths are falsely claimed as colon cancer instead of rectal cancer on death certificates [4]. According to reports, due to ageing, human advancement, and population growth, the global number of new CRC cases is expected to reach 3.2 million in 2040 globally [5]. The increased prevalence of CRC is mostly caused by excessive exposure to environmental and genetic factors such as familial adenomatous polyposis, Lynch syndrome, MUTYH-associated polyposis, etc. [6]. This can also be associated with lifestyle changes [5].
Various Risk Factors of Colon Cancer

According to a recent report by the World Cancer Research Fund/American Institute for Cancer Research expert panel, eating a lot of red and processed meat, body fatness, belly fatness, and binge drinking raises the risk of CRC [7]. CRC may be prevented by drinking milk and eating full grains [7]. Intake of vegetables and possibly fruits have an indirect relationship with the risk of colon cancer. Male sex, age, race/ethnicity, body mass index (BMI), height, diabetes, dietary antioxidants, folate, calcium, postmenopausal hormone use, and smoking are all widely listed risk factors for CRC [8].

2. Dietary Fibres

Dietary fibres are defined as eatable carbohydrate polymers with three or more monosaccharide units that are resistant to action of endogenous digestive enzymes, and thus, are neither metabolised nor absorbed in the small intestine [9]. These are edible carbohydrate polymers present in foods such as fruits, vegetables, legumes, and cereals; (ii) edible raw food materials through physical, enzymatic, or chemical degradation and synthetic carbohydrate polymers with a proven biological benefit [9]. The World Health Organization (WHO) recommends consuming at least 25 g of fibre each day [10]. Various food and health groups recommend diets high in vegetables, fruit, and whole-grain cereals to satisfy the guidelines. For adults, most countries recommend 25–35 g of dietary fibre per day, with recommendations ranging from 18–38 g per day [9]. Although most national authorities use this definition, there are some differences among these definitions. These differences mainly concern (1) whether non-carbohydrates such as cellulose and other substances found in cell walls linked to polysaccharides are considered dietary fibres and (2) the required minimum quantity of carbohydrate monomers [9]. The present review provides a detailed understanding of various dietary fibres along with their potential effects on gut microbiota and their association with colon cancer.

2.1. Types and Characteristics of Dietary Fibres

Dietary fibres are classified as polysaccharides (non-starch polysaccharides (NSPs), resistant starch (RS), and resistant oligosaccharides (Ros)), soluble as well as insoluble in nature [9]. Fibres of an insoluble nature of the carbohydrate moiety, such as cellulose and hemicellulose, enter the colonic region and are gradually or not digested by gut bacteria and produce faecal bulking. Unlike ROs, most soluble NSPs, particularly polymers with a high molecular weight, such as guar gum, certain pectins, β-glucans, and psyllium, are viscous, and can form a gel structure in the intestinal tract. This delays glucose and lipid absorption and influences post-prandial metabolism [11].

Most soluble fibres do not facilitate to faecal bulking, but they are digested by gut bacteria and produce metabolites such as short-chain fatty acids (SCFAs) [11]. SCFAs are chemical compounds that are mostly made up of acetate, propionate, and butyrate. SCFAs are important regulators of the host’s metabolism, immunological system, and cell growth. SCFAs are abundant in the cecum and proximal colon, where they are utilised as an energy source by colonocytes (particularly butyrate), but they can also be transferred to the interstitial blood circulation via the portal vein and act on the hepatocytes and peripheral tissues [12]. Even though SCFA levels in the peripheral circulation are modest, it has become well known that they function as chemical messengers and regulate a variety of physiological functions in the host [12]. Soluble fibres increase food transit time in the intestine and are easily fermented into gases in the colon. Insoluble fibres can either be metabolically inert, providing bulk to intestinal contents, or they can be fermented in the large intestine [13]. Insoluble fibres have the effect of speeding up the passage of food through the intestine [13]. Soluble fibres bond to bile acids in the small intestine, preventing them from being absorbed. As a result, cholesterol levels of blood are reduced [14]. Soluble fibres also inhibit sugar absorption, normalise blood cholesterol levels, and create SCFA in the colon as by-products of fermentation, which have a wide range of physiological
functions. Although insoluble fibres are linked to a lower incidence of diabetes, the exact mechanism remains uncertain [15].

2.2. Sources of Fibres

A promise that a food is high in fibre, or any claim likely to have the similar meaning for the consumer, may only be made if the product has at least 30 g of fibre per 500 g or at least 3 g of fibre per 100 kcal (418 kJ) [9]. In the human small intestine, dietary fibres are the indigestible matter of plant food. Poly/oligosaccharide cellulose and other plant-derived compounds make up dietary fibres [16]. Raw chickpea seeds have a total dietary fibre content (DFC) of 18–22 g per 100 g [17]. Soluble and insoluble DFC is around 4–8 and 10–18 g per 100 g of raw chickpea seed, respectively [16]. A study on total dietary fibre of various gram and peas seeds showed that soaking can significantly enhance total dietary fibre and soluble dietary fibres [13]. A summary on dietary fibre content and their energy in some widely used food is presented in Table 1 [18].

Table 1. Dietary fibre and energy content of some selected fibre-rich foods.

| Plant Food                | Dietary Fibre (g/100 g) | Energy (KJ/100 g) |
|---------------------------|-------------------------|-------------------|
| Almond                    | 13.06 ± 0.31            | 2549 ± 4          |
| Avocado                   | 6.69                    | 604               |
| Barley                    | 15.64 ± 0.64            | 1321 ± 19         |
| Black gram (Whole)        | 20.41 ± 0.06            | 1219 ± 5          |
| Broad beans               | 8.63 ± 0.15             | 123 ± 4           |
| Carrot (Red)              | 4.49 ± 0.19             | 160 ± 19          |
| Chickpea                  | 25.22 ± 0.39            | 1201 ± 9          |
| Drumstick leaves          | 8.21 ± 0.19             | 282 ± 27          |
| Field bean (Black)        | 23.40                   | 1155              |
| Finger millet (Ragi)      | 11.18 ± 1.14            | 1342 ± 10         |
| Fresh peas                | 6.32 ± 0.26             | 340 ± 19          |
| Gingelly seeds (Black)    | 17.16 ± 0.19            | 2124 ± 8          |
| Gooseberry                | 7.75 ± 0.64             | 99 ± 19           |
| Guava (White Flesh)       | 8.59 ± 0.05             | 135 ± 5           |
| Ladyfinger                | 4.08 ± 0.20             | 115 ± 5           |
| Pear                      | 4.48 ± 0.08             | 157 ± 3           |
| Pearl millet (Bajra)      | 11.49 ± 0.62            | 1456 ± 18         |
| Pistachio nuts            | 10.64 ± 0.16            | 2257 ± 10         |
| Quinoa                    | 14.66                   | 1374              |
| Red gram (whole)          | 22.84 ± 0.43            | 1146 ± 10         |
| Sapota                    | 9.60 ± 0.57             | 307 ± 18          |
| Sesbania grandiflora      | 8.60                    | 295               |
| Soya bean (Brown)         | 21.55 ± 0.66            | 1596 ± 11         |
| Wheat flour               | 11.36 ± 0.29            | 1340 ± 7          |

3. Imbalance of Gut Microbiota and Health Disorders

Firmicutes, Bacteroidetes, Actinobacteria, and Proteobacteria are the major groups in the human intestinal microbiome. Gut microbiosa digest indigestible food, produce nutrients such as vitamins, detoxify metabolites, influence the immune response, provide signals for epithelial cell renewal and mucosal integrity maintenance, and exude antimicro-
bial compounds [19]. The microbiome (which includes bacteria, viruses, fungus, and other microorganisms) governs health, and microbiota changes can lead to disease. Alteration in the intestinal microbiome allow environmental risk factors to develop and perpetuate CRC [20,21]. This could be because microbiome influence metabolism and immune function. Evidence suggests that the alterations in the gut microbiota occur early in CRC and can be used to identify those who are at risk for colorectal adenoma, a precursor lesion to CRC [22]. Thus, microbiota alteration can be utilised as biomarker for the early diagnosis of CRC [22]. There is a definite link between the GI microbiome and the progression of chronic non-communicable diseases, which shows the importance for development of strategies to enhance human health by targeting the GI microbiota.

4. Symbiotic Relationship between Dietary Fibres and Microbiota

4.1. Human Gut Microbiota

The human gut is home to trillions of bacteria from all kingdoms of life, all of which are necessary for the growth and physiology of the host. This “gut microbiota” is a complicated ecosystem to communicate with one another and with the human to influence vital biological function, such as regulating juvenile growth [23,23], maturation of the immune system, and protection against some infectious agents [24]. In addition, it helps in carbohydrate and lipid metabolism that regulate and maintains the host’s energy storage [25,26].

4.2. Understanding of the Host-Microbiota System

The microbiome helps to maintain homeostasis in all our body’s tissues. Symbiotic relationships exist between vertebrates and the large and complex microbial populations that populate their gastrointestinal tracts. The general interaction between people and their microbiota may be described as mutualistic [27], and the balance of gut microbial ecosystems (eubiosis) is a fundamental concept. Disturbance of gut microbiota results in the onset of a variety of chronic diseases [28]. Pathogenic changes in the microbiome’s profile and functions are referred to as dysbiosis. Changes in the number of good intestinal bacteria can result in long-term inflammation and the formation of carcinogenic chemicals, which can lead to neoplasia [19].

4.3. Bacterial Signaling for Virulence

The gut bacteria convert dietary and endogenous substances into metabolites that allow communication with host’s peripheral organs and tissues. If any changes in gut bacteria composition have been connected to specific illnesses, altering gut bacteria composition by dietary changes may be a potential therapy option [29]. Dietary fibre confers resistance against the colonisation of the gut with multidrug-resistant bacteria and increases the number of bacteria that convert fibre into SCFA [30]. The intestinal microbiota and the gut mucosal immune system have a homeostatic relationship, and disturbance of this interaction can lead to illnesses [31]. The Western lifestyle comprises a diet low in microbiota-accessible carbohydrates (MACs), which leads to a lower participation and functionality of microbioata as compared with the non-Westernised population, which includes a high fibre in their diet. Immunological dysregulation caused by interactions between resident microorganisms and the host could explain a number of disorders involving inflammation as a common cause [32].

5. Impact of Dietary Fibres on Gut Bacteria

Diet has a big influence on the gut microbiota’s composition, variety, and richness. Different aspects of a diet have a time-dependent effect on gut bacterial ecosystems [33]. Diet is the simplest therapeutic intervention, since it is the easiest to change [34]. Bacteroides, Prevotella, and Ruminococcus were found to be the most prevalent varieties, or “enterotypes”, in a study of gut microbial populations [34]. Variation in the ratio of Prevotella/Bacteroides was also seen across industrialised and unindustrialised human populations, implying that these bacterial communities are influenced by long-term dietary differences, for instance,
mutton (which drives *Bacteroides* in Westernised populations) and dietary fibre (which drives *Prevotella* in non-Westernised populations) [35].

*Prevotella* plays a crucial role in nurturing healthy gut microbiota. The impact of nutrition on the microbiome was revealed by comparing the gut bacteria of children from rural (Burkina Faso (BF)) and urban (Italy) communities [35]. In both groups, the gut microbiota composition changed significantly after breastfeeding. In BF children, there was a considerable rise in the population of microbiota from the genera *Prevotella* and *Xylanibacter*, which was associated with greater amounts of faecal SCFAs, indicating their ability to break down complex carbohydrates [35]. These variations are irrespective of race, as the gut microbial populations of BF children residing in urban, rather than rural, parts of BF resembled those of Italian children. The modification in the gut microbiota population is linked with the fact that when people relocate to cities, they are introduced to a Westernised way of lifestyle, which includes availability to high-saturated fat and high-sugar foods. As a result, children in urbanised areas of BF have bacteria that are better suited to metabolise meat protein, fat, and sugar-rich foods, whereas children in rural areas have bacteria that are better suited to digest fibre and carbohydrate fermentation from vegetables (enrichment in *Prevotella*, *Treponema*, and *Succinivibrio*). Surprisingly, the microbiomes of BF children living in cities were equivalent to those of Italian children, indicating that nutrition had a significant impact independent of host genetics [35].

Hadza is a modern human community who reside in an important geographic region for human evolution research and hunt for resources [36]. It was found that Hadza hunter-gatherer people have a greater level of microbial abundance and biodiversity than Italian city dwellers. These people’s gut microbiota exhibited higher *Bacteroidetes* and lower *Firmicutes* species richness, and they almost completely lacked the *Actinobacteria* phylum, with minor amounts of *Bifidobacterium*. Interestingly, Hadza SCFA production was linked with higher propionate levels, whereas the Italian cohort had higher butyrate levels [36]. Environmental lifestyle changes, rather than host genetics, affected gut microbiota diversity, and industrialisation results in a considerable loss of species, according to these findings [37].

Antibiotic use, therapeutic practices (e.g., caesarean sections), and sanitation are the underlying causes for a decreasing variety of microbiota [32]. For instance, the gut microbiota was reported to be altered after caesarean section [38]. The fact that babies delivered naturally had greater gut bacterial counts at one month of age than babies delivered by caesarean section shows that natural birth promotes and starts the colonisation of the gut by microorganisms [39]. Poor personal hygiene and unsanitary living circumstances in poorer nations can contribute to the spread of infectious diseases. Circadian disorder caused by travel, shift work, or other factors affects gut health and changes the bacteria communities in the gut [39,40]. The long-term effects of various antibiotics include reduced microbial diversity, altered *Bacteroidetes/firmicutes* ratio, overgrowth of *Clostridium difficile*, and increased expansion of opportunistic pathogens, such as *Salmonella typhimurium*, *Escherichia*, and *Klebsiella* species [41].

5.1. Probiotic-Prebiotic Relationship of Dietary Fibres

Probiotics, prebiotics, and synbiotics are all based on the same concept, i.e., to develop meals that increase “good” bacteria in the intestinal lumen after consumption [42]. This can be accomplished by adding either health-promoting “probiotic” bacteria or fermentable “prebiotic” carbohydrates that are indigestible but fermentable [42]. The functions of the probiotic bacteria added to food include the reduction of potentially pathogenic bacteria and/or toxic metabolites in the intestine, restoration of GI motility, and modulation of the immune response. Conversely, so-called prebiotic food components should support the growth of beneficial bacteria of the indigenous intestinal microbiota of humans, and also improve survival of probiotic bacteria which have been consumed at the same moment [43]. Prebiotic polysaccharides are dietary fibres that are fermented by the large intestinal microbiota rather than processed by human enzymes. As a result, they improve the
health of the intestinal lumen mucosa by increasing biomass, faeces weight, and regulate frequency of defecation, reducing constipation and enhancing the health of the intestinal lumen mucosa [44,45].

5.2. Microbial Metabolism of Dietary Fibres and Functional Implication

Dietary fibres are a good source of energy for the bacteria that live in the cecum and colon. Under certain intestinal conditions, anaerobic bacteria activate their machinery, which includes important enzymes and metabolic pathways that may digest complex carbohydrates and produce metabolites such as SCFAs. SCFAs are chemical compounds that are mostly made up of acetate, propionate, and butyrate, which are necessary for the health of the intestinal mucosa and are otherwise unavailable in the diet. The colonic microbiota’s ability to ferment fibre for their own metabolic needs, leading in the generation of luminal SCFAs [12,46,47]. A study by Roediger [48] demonstrated that colonocytes choose butyrate over glucose as a source of energy, laying the groundwork for the now well-known symbiotic relationship between colonic microorganisms and colonic mucosal health.

5.3. Effect of Dietary Fibres and SCFA on Host

Mucus generation and secretion are stimulated by dietary fibres and SCFAs. Both acetate and butyrate help to control mucus production and discharge. Acetate- and propionate-producing bacteria, Bacteroides thetaiotaomicron, enhance goblet cell development and mucin-related genes expression. On the other hand, Faecalibacterium prausnitzii, an acetate consumer and butyrate generator, reduces the influence of acetate on mucus and inhibits mucus overproduction, allowing the gut epithelium to maintain its correct shape and composition [49]. Moreover, dietary fibres can mechanically increase mucus secretion by the intestinal epithelium [50].

Long-term dietary fibre deficiency has been linked to an increase in mucin-degrading bacteria such as Akkermansia muciniphila, which affects the mucus barrier [51]. When the diet lacks dietary fibres, some gut bacteria adapt their metabolism to utilise mucin glycans by activating the gene expression of mucin-degrading enzyme [52].

A Western diet (low fibre content) given to mice increased the permeability of the inner mucosal layer and decreased the development rate, making the mucus penetrable and perhaps increasing infection susceptibility [29]. A study conducted on obese mice showed that a small inulin dose (1%) or Bifidobacterium longum (a prebiotic with bifidogenic properties) was beneficial in mucus problems. The permeability of the inner mucosal layer was restored by the supplementation of insulin, whereas the mucus growth rate defect was restored by B. longum supplementation, indicating that the two criteria are independent and may be governed by distinct mechanisms. Obese animals did not benefit from the administration of either 1% inulin or B. longum [53]. High inulin intake (20%), on the other hand, prevented microbial invasion, enhanced intestinal health, and contributed to the relief of low-grade inflammation in obese mice [53]. A previous study revealed that colonocytes caused beta-oxidation of butyrate aerobically, resulting in an anaerobic environment in the gut [54]. Because butyrate-producing bacteria are particularly sensitive to oxygen, their numbers are reduced even further, resulting in a reduction in butyrate production. This feedforward loop results in enhanced luminal oxygen levels, allowing proteobacteria such as Escherichia coli and Salmonella enterica serovar Typhimurium to flourish. This unique approach not only explains most of the diseases linked with a low-fibre diet, but also provides a molecular explanation for why lower microbiota richness is reported both in humans and mice having low-fibre diet.

6. Diet Manipulation, Gut Microbiota, and Colon Cancer

Humans and their gut microbiota coexist as mutualists, but this connection can become pathological in some cases, such as obesity, diabetes, atherosclerosis, inflammatory bowel diseases (IBD), and cancer [33]. CRC is thought to be linked to localised inflammation. Though inflammation cannot initiate oncogenesis on its own, it is thought to be a crucial...
contributor. Previous research has revealed that gut inflammation and CRC oncogenesis can be modulated by food, gut microbiota, and gut environment, and these variables may be adjustable variables in modifying CRC outcomes [55]. SCFA is a useful metabolite that is produced under the action of local biota that ferments fibre and other indigestible starches as they move from the small intestine into the large colon. These SCFAs, in turn, have antineoplastic, anti-inflammatory, and physiological properties.

### 6.1. Causal Relationship between Microbiota, Dietary Fibre, and Colon Cancer

Low dietary fibre consumption reduces microbial diversity, SCFA generation, and shifting gut microbial metabolism to less favourable substrates, such as dietary and endogenously provided proteins [56,57]. This might promote mucus-degrading bacteria, which might harm the host [58]. The generation of total SCFAs and butyrate was dramatically reduced when human volunteers were fed a high-protein, low-carbohydrate diet [56,59]. However, the fermentation of amino acids resulted in an increase of potentially harmful metabolites such as branched-chain fatty acids, ammonia, N-nitroso compounds, p-cresol, sulphides, indoles compounds, and hydrogen sulphide [60]. These metabolites’ cytotoxic as well as pro-inflammatory properties contribute to the development of chronic illnesses, especially CRC. Butyrate-producing bacteria are lower in CRC patients than in healthy people, suggesting that a significant structural imbalance of the gut bacteria is produced by a decrease in butyrate bio-makers and an increase in pathogenic organisms [61]. Reduced dietary fibre intake is linked to a higher risk of CRC [62]. As a result, it is not surprising that low-fat diets high in fibre-rich grain products, vegetables, and fruits have health claims allowed by the FDA for a possible reduction in the risk of developing some types of cancer. According to the findings, increasing dietary fibre in African Americans modified the microbiota and enhanced butyrogenesis, resulting in lower cancer risk indicators is depicted in Figure 1 [63].

![Figure 1](image)

**Figure 1.** The key role of high and low dietary fibres on gut microbiota, diversity, and function on host health: action of dietary fibres on the progression of colon cancer takes place, high intake of dietary fibres will change into SCFAs, such as propionate, butyrate, and acetate, under the action of healthy gut microbiota as well as inhibit TNF, NB-kF, and early colonic lesion and regulate microbiota health. Moreover, due to intake of low dietary fibres, the gut microbiota is altered, resulting in colonic lesion due to a disturbance in mucus-degrading enzymes and bacteria [64]. ↓: decreasing, ↑: increasing.
6.2. Effect of Dietary Fibre on Colon Cancer

CRC and ulcerative colitis are two common bowel illnesses that affect the descending distal colon, which is the principal location for fermentation of protein into their amino acids [60]. IBD can also lead to CRC [65], which is the third most common cancer [66]. CRC is linked to food habits, smoking, physical activity, and hereditary and environmental variables [66]. Americans consumed two to three times more animal protein and fat, while Africans consumed more carbohydrate and fibre, primarily in the form of resistant starch. African Americans had more polyps and greater rates of mucosal proliferation on colonoscopy when evaluated using Ki67 epithelial cell staining, showing its potential application as a cancer risk biomarker [63,63]. A plant-based cuisine includes vegetables, fruits, cereals, nuts, and legumes, most of which are made up of considerable amounts of olive oil, with moderate use of fish, seafood, or dairy, and limited intake of meat and alcohol (mostly red wine), which is the nutritional dimension of the Mediterranean lifestyle [67]. Indian meals are also beneficial to one’s health, because they have been linked to a lower prevalence of numerous ailments. Increased consumption of high-fibre meals or fibre supplements decreases blood pressure, improves blood glucose, assists weight loss, and reduces the risk of CRC [68].

These differences in the dietary pattern were associated to major changes in microbiota (the genus Bacteroides dominated in Americans, and the genus Prevotella in Africans) and metabolic phenotype derivatives [69]. Africans had larger quantities of starch break-down, carbohydrate fermenters, and butyrate makers, as well as their metabolites derivatives [69]. In Americans, there were more potentially pathogenic proteobacteria (Escherichia and Acinetobacter) and bile acid deconjugators and their derivatives [69]. The products of fibre fermentation, particularly butyrate, have been shown to be anti-inflammatory and antineoplastic in numerous studies [70–72] and secondary bile acids (BAs), the by-products of bacterial bile acid conjugation, were reported as carcinogenic [73].

Dietary fibres bind conjugated primary BAs may act as a medium for gut bacteria that produce non-conjugated BAs via the bile salt hydrolase (Bsh). These can attach to dietary fibres and then be processed by bacteria that have the activity of 7-alpha dehydroxylation, resulting in secondary BAs. Dietary fibre’s ability to bind secondary BAs indicates that they may be involved in controlling BA levels in the gut. This structural connection may influence host physiology by either avoiding the build-up of toxic BAs that can lead to polyps and CRC or by boosting the destruction of BAs that can trigger G protein-coupled receptor 5 (TGR5) and promote Glucagon-like peptide-1 (GLP-1) production [56]. GLP-1 has separate effects on cell growth and survival, and chronic activation of GLP-1 receptor (GLP-1R) signalling causes colon cancer in mice, which can be prevented by using a GLP-1R antagonist [74].

High-fibre food products, nuts, avocados, and eggs, which are high in monosaccharides, peptides, and amino acids, as well as monounsaturated and polyunsaturated fatty acids and SCFAs nutrients, appear to influence GLP-1 secretion, This might enhance associated beneficial outcomes in healthy individuals [75]. Furthermore, bacterial digestion of dietary fibres releases minerals and phenolic compounds that can be absorbed by the distal intestine [56]. This highlighted two potential mechanisms for diet-related cancer risk: dietary fibre’s preventive action in increasing butyrogenesis and dietary fat’s promotional effect in promoting bile acid production by the liver. According to the findings of two previously published human investigations, dietary changes in fibre and fat content have a significant influence on the gut microbes within 2–3 days [33,33,76]. Shifting African Americans to a high-fibre, low-fat meal for two weeks resulted in an extreme reduction in colonic mucosal inflammation and suppression of secondary bile acid synthesis. The association and role of bile acid, protein, and high fat in cancer progression is depicted in Figure 2.
This is not just because probiotics in food or therapeutically used microbes are usually proinflammatory and proneoplastic properties of protein fermentation and bile acid deconjugated and bacteriocines), agglutination of infective bacteria, and strengthening of the intestinal decreased intestinal pH, generation of bactericidal chemicals (e.g., organic acids, H₂ in infection of other organs [85]. The mechanisms for these effects are unknown, including decreased intestinal permeability, and bacterial translocation, or by providing bioactive or regulatory metabolites [77–79]. Probiotics are “live microorganisms that, when administered by using a GLP-1R antagonist [74].

Figure 2. The association and role of bile acid, proteins, and high fat in cancer progression. Food acts differently under the influence of various microbiota as good and bad bacteria, and their effect on the progression of colorectal cancer. With an imbalanced high-fat, high-meat, low-fibre diet, the proinflammatory and proneoplastic properties of protein fermentation and bile acid deconjugated residues predominate, leading to increased colon cancer risk. H2S: Hydrogen sulphide, AA: Amino acid, NH2: Amines, BA: Bile acid. ↓: decreasing, ↑: increasing.

6.3. Probiotic

Probiotic bacteria not only just influence the intestinal microbiota in the large intestine, but also have an impact on other organs, either by altering immunological parameters, intestinal permeability, and bacterial translocation, or by providing bioactive or regulatory metabolites [77–79]. Probiotics are “live microorganisms that, when administered in adequate amounts, confer a health benefit on the host”, according to the FAO/WHO 2001 [80,81]. Most of the health benefits associated to probiotic microorganisms are linked to the GI tract, either directly or indirectly (i.e., mediated by the immune system) [82,83]. This is not just because probiotics in food or therapeutically used microbes are usually taken by mouth [82,83] however, interactions with the host’s unique microbiota or immunocompetent cells of the intestinal lumen typically determine the mechanisms and potency of a probiotic effect. The gut (or the gut-associated lymphatic system (GALT)) is the largest immunologically competent organ, and the growth and composition of the indigenous microbiota determines the immune system’s maturation and optimal development from birth [31,84].

6.3.1. Probiotics and Their Role on Gut Microbiota

Probiotics may improve or prevent gut inflammation and other intestinal or systemic disease phenotypes by restoring the composition of the gut microbiome and introducing beneficial functionalities to gut microbial communities [79]. Many probiotic bacteria strains have been found to alter the microbiota of small intestine and/or limit pathogen colonisation of the gut; in addition, pathogenic bacteria translocation through intestinal lining and infection of other organs [85]. The mechanisms for these effects are unknown, including decreased intestinal pH, generation of bactericidal chemicals (e.g., organic acids, H₂O₂, and bacteriocines), agglutination of infective bacteria, and strengthening of the intestinal
mucosa’s barrier function, which are some of the unproven mechanisms underlying these effects [86]. Competition for microbial fermentation substrates or receptors on the mucosal cellular surface, release of gut-protective compounds such as arginine, glutamine, SCFA, CLA, SCFA, and CLA [46,87–90]. In addition to their immunomodulatory features, probiotics’ potential utility in the prevention or treatment of diarrhoea or IBD has been investigated [91].

Antibiotic-associated diarrhoea traveller’s diarrhoea and diarrhoea caused by other causes are substantially reduced when probiotics are used [92] The most well-documented probiotic benefit is protection from immunostimulatory bacteria yeasts, or symptoms relief and shortening of acute infections [79]. A reduced frequency of diarrhoea, GI, and respiratory tract infections was reported after two months of daily administration of L. casei-bearing fermented milk and milk constitute a probiotic Lactococcus lactis (Lactobacillus rhamnosus) [93]. The consequence of probiotic bacteria on traveller’s diarrhoea has yielded mixed results, possibly due to differences in probiotic strains, travelled countries, local microbiota, traveller’s specified (eating) habits, or the method of probiotic administration (before or during travel, as a capsule or a fermented dairy product). In some research, participants who consumed the probiotic had fewer or shorter episodes of diarrhoea [94].

In many studies, taking probiotic strains before and during antibiotic treatment reduced the duration and/or frequency of antibiotic-associated diarrhoea episodes, as well as the severity of symptoms [92,95]. Chemo- and radiotherapy frequently cause significant immune system and intestinal microbiota disruptions, as well as diarrhoea and/or increased fungus (Candida albicans) cell counts in the GI and other organs. Probiotic microorganisms were given before and during chemo side effects in rodents [96]. Although it has not been proven whether frequent consumption of probiotics has useful or helpful effects on HIV patients, it has been established that probiotic product are well digested by these individuals [97]. Similarly, in lactose malabsorbers, fermented milk products improve lactose digestion and prevent intolerance symptoms [98]. This action is mostly due to the presence of microbial galactosidase in fermented milk products containing live bacteria, which survives passage via or by way of the stomach before being freed in the small intestine to facilitate lactose hydrolysis [98].

6.3.2. Synbiotic Food

Synbiotics are probiotic and prebiotic blends that benefit the host by enhancing the survival and implant of live microbial dietary additives into the host’s GI tract. However, this phrase should be used only when there is actual “synergistic” mutual reinforcement. This condition is not met by most foods that include both probiotic microorganisms and prebiotic carbohydrates [99].

7. Cancer Prevention and Dietary Fibres

CRC is thought to be linked to localised inflammation. Previous research has revealed that gut inflammation and CRC oncogenesis can be modulated by food, gut microbiota, and gut environment, and they might serve as adjustable variables in modifying CRC outcomes [55]. Colonocytes, unlike the rest of our body’s cells, obtain their energy from butyrate and other SCFAs instead of glucose [100]. Chundakkattumalayil et al., in their study on male Balb/c mice, showed a decreased expression of beta-glucuronidase with increase butyrate production and improved colon motility and TNF- levels. The levels were found to be lower after using synbiotics and prebiotics at a dose of 1 gm/100 mL [101]. Another study on colon cancer through the Cantharellus cibarius crude polysaccharides showed COX-1 and COX-2 inhibition in the colon cancer cells proliferation at a dose of 100 µg/mL [102]. A resistant starch diet enhanced choline acetyltransferase expression along with butyrate synthesis and alleviate colon motility in rats emphasised the importance of SCFAs such as butyrate in colon health [103]. Furthermore, butyrate enemas reduced the levels of pro-inflammatory cytokines IL-1, IL-6, and TNF in peripheral blood,
which was found to be interfered by butyrate-induced NF-kB transcription activity attenuation [104]. Hence, butyrate seems to be able to regulate both fundamental colon function and alter locoregional inflammation. In addition, the link between gut bacteria and butyrate levels, as well as their relationship to CRC, is gaining attention. Additional research has indicated that gut microbiota can help CRC cell models and patients have better outcomes. *Lactobacillus casei* and *Lactobacillus rhamnosus* supernatants have been found to suppress the aggression and progression of CRC in in vitro studies [105]. Though the exact mechanism is unknown, it is known that *L. casei* and *L. rhamnosus* supernatants reduce the activity of matrix metalloproteinase-9 (MMP-9) while increasing the tight junction protein zona occludens-1 levels (ZO-1). Both proteins govern colonocytes‘ extracellular matrix (ECM), and MMP-9 is thought to be used by malignant colonocytes to breakdown the ECM to spread to nearby healthy group of cells. MMP-9 is, of course, associated with nodal metastases in patients with CRC [105].

In adenomatous polyposis coli (APC) gene mutant in vivo mouse model studies, *Lactiplantibacillus plantarum* has also been shown to reduce the progression of CRC. Over 80% of CRC cases contain a mutation in the APC gene. APC-mutated mice administered oral enemas of *L. plantarum* had a slower progression of CRC. The levels of IL-6, IL-17, and TNF in the blood were reduced after treatment with these enemas. Enema therapy also boosted gut microbial diversity and the abundance of Firmicutes and Actinobacteria, mimicking the gut ecology of wild type mice. Microbial variety was found to be less in the non-treated APC mouse model, with greater levels of Proteobacteria and Bacteroides [106]. Earlier, it was discovered that *L. plantarum* enema reduced inflammatory cytokines and beta-catenin protein production in colonocyte nuclei. The Wnt signalling pathway, which is a critical regulator in oncogenesis in APC-mutated CRC, is modulated by beta-catenin levels [3,100]. It has been established that *Limosilactobacillus reuteri* inhibits nuclear factor-kappaB (NF-B)-dependent gene products that control cell survival and proliferation. It slowed down the proliferation of cancer cells by inhibiting NF-kB activation caused by tumour necrosis factors (TNF), including NF-kB-dependent reporter gene expression in a dose- and time-dependent manner [107,108].

**In Vitro, In Vivo, and Clinical Studies of Dietary Fibres on Colon Cancer**

In a previous study the count of total aberrant crypt foci, sialomucin-producing ACF (SIM-ACF), and mucin-depleted foci (MDF) in the distal colon were considerably reduced by 10% djulis (*Chenopodium formosanum*) plus $5 \times 10^6$ cfu L. of acidophilus/g (DLA) and 10% djulis plus $5 \times 10^7$ cfu L. of acidophilus/g (DHA). DLA and DHA also regulated apoptosis-related proteins and suppressed the expression of proliferating cell nuclear antigen (PCNA) and COX-2. The synbiotic as adjuvant of djulis with *Lactobacillus crispatus* (L. acidophilus) inhibited colon carcinogenesis [109]. After treating ApcMin/+ mice with Gynostemma pentaphyllum saponin (GpS), the polyps in ApcMin/+ mice were effectively reduced. In *Bifidobacterium animalis* culture, GpS increased the expression of a wide variety of genes encoding biogenesis and metabolic processes. Furthermore, colonisation with *B. animalis* significantly lowers the polyp load in ApcMin/+ mice [110].

In another study, *Myrciaria jaboticaba* seed extract inhibited amylase, glucosidase, and ACE-I activities and was more harmful to cancer cells than normal cells in a mouse model. It was mixed into yoghurts at various concentrations, and total phenolic content, AA (amino acid), and EI increased in a dose-dependent manner and influenced the gut bacterial microbiota, acting as a prebiotic [111]. It was discovered that polysaccharides extracted from *N. commune* (NVPS) prevent mice from colitis-associated colon carcinogenesis. It significantly reduced the number and size of tumours and the incidence of intestinal tumours, by dramatically increasing the number of SCFA-yielding genera, such as butyric acid-producing genera (*Butyricicoccus*, *Butyrivibrio*, and *Butyricimonas*), and acetic acid-yielding genera (*Butyricimonas* (*Lachnospiraceae UCG 001, Lachnospiraceae UCG 006, and Blautia*) [112].
On the basis of clinical data, twenty patients were given 20 g/day of *Plantago ovata* seeds orally for three months, which eradicated colorectal cancer and significantly increased butyrate (and acetate) production, as well as increased faecal butyrate concentrations by 42% in patients diminished for colonic cancer [112]. Another study showed that Yacón (*Smallanthus sonchifolius*) flour as a source of fructooligosaccharides lowered intraluminal pH, lactulose/mannitol ratio, (TNF-α)/interleukin (IL)-10 ratio, and secretory immunoglobulin A levels in rats with CRC. It could promote and protect gut health. A study conducted by Nowacka-Jechalke et al. on colon cancer through the *Cantharellus cibarius* crude polysaccharides showed inhibition of both COX-1 and COX-2 activity at 100 µg/mL dose [102].

**Jujube polysaccharides** were found to restore the gut microbiota composition, making them interesting prebiotic options for the prevention and treatment of colorectal cancer. It displayed prebiotic-like properties by positively modifying gut bacteria and influencing KEGG (Kyoto encyclopedia of genes and genomes) metabolic pathways and ATP-binding cassette (ABC) transporters that contribute to host health. It may help to prevent colon cancer by reversing the gut dysbiosis caused by colitis. There was also a considerable reduction in Firmicutes/Bacteroidetes [113]. The beneficial effect of soluble dietary fibre from *Musa paradisiaca* inflorescence (PIF)-fermented adjuvant with *Lactobacillus casei* (Lactobacillus casei) and *Bifidobacterium bifidum* on HT29 colon cancer cells was also demonstrated [114].

The details of various in vitro and in vivo studies on colon cancer are presented in Table 2.

| Scientific Name and Family | Dose | Phytoconstituents | In-Vivo/In-Vitro | Effect/MoA | Reference |
|----------------------------|------|-------------------|-----------------|------------|-----------|
| *Acacia gum* (Fabaceae)    | 1 gm/100 mL Distilled water | Agarose, 7,3′,4′-Trihydroxysolavone, noscapine, tetrahydro-papaveroline, costunolide, hesperidin | BALB/C mice | ↑ butyrate ↓ TNF-α ↓ β-glucuronidase, ↓ NF-kB. | [101,115] |
| *Cantharellus cibarius* (Cantharellaceae) | (10, 25, 50 and 100 µg/mL) Polysaccharides (1,4-linked-β-D-glucose) | Colon cancer cell | Inhibit the proliferation of colon cancer cells | [102] |
| *Chenopodium formosanum* (Amaranthaceae) | 44 g dry leaves for a 60 kg per day per person Phenolics, flavonoids | Carcinogen induced rat model | ↓ Aberrant Crypt Foci (ACF) cells ↓ Mucin-depleted Foci (MDF) cells, inhibit cell proliferation, ↓ mucins secretion | [109] |
| *Gynostemma pentaphyllum* (Thunb.) | 0.01 to 0.3 mg/mL | Gypenosides or gynosaponins | In ApMin /+ mice | ↑ SCF, ↓ Polyps, ↓ Sulphur | [110,116] |
| *Lactulose* | 2 gm galacto-oligosaccharides | AOM/DSS induced CRC | ↑ SCF ↓ Firmicutes ↑ Bacteroidetes ↑ Phascolarctobacterium genus | [117] |
| *Myrciaria jaboticaba* (Myrtaceae) | 50 and 100 mg/L Castalagin, vescalagin, procyanidin, anthocyanins and ellagic acid | Rat with induced carcinogenesis | ↓ Firmicutes ↓ Bacteroidetes ↓ β-glucosidase, β-galactosidase, mucinase, and nitroreductase inhibit proliferation of cancer cells | [111] |
| *Nostoc commune* (Nostocaceae) | 100, 200 mg kg⁻¹ Polysaccharides (glucose, arabinose, xylose, mannose, and galactose) | AOM/DSS-induced CRC C57BL/6 J mice | ↑ SCF ↓ Firmicutes ↓ Bacteroidetes | [112,118] |
Table 2. Cont.

| Scientific Name and Family | Dose                  | Phytoconstituents                                                                                                                                                                                                 | In-Vivo/In-Vitro                                                                 | Effect/MoA                                                                 | Reference |
|----------------------------|-----------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------|---------------------------------------------------------------------------|----------|
| *Smallanthus sonchifolius* (Asteraceae) | 10 g/kg Yacon         | Flour-fructooligosaccharidesgallic acid, beta-sitosterol, behenic acid, kaempferol, quercetin, vanillic acid, hexadecanoic acid                                                                                   | Wistar rats                                                                    | ↑SCF TNF-α/IL-10 ratio ↓ PH ↑ sIgA, anti-inflammatory, cytokines, mainly IL-10. | [119–121]|
| *Ziziphus jujuba* Mill. (Rhamnaceae) | 1000 mg kg⁻¹ JP       | Jujube polysaccharides (Glucose)                                                                                                                                                                                   | AOM/DSS 1-induced CRC C57BL/6 mice                                            | ↓Firmicutes ↑Bacteroidetes ↑SCF Regulate dysbiosis                          | [113,122]|
| *Musa paradisiaca* (Musaceae) | 905.75–1510.88 µL     | Dietary fibre and polyphenols                                                                                                                                                                                    | HT29 cell line                                                                | ↑ROS apoptosis HT29 Cell                                                  |          |
| Inulin-rich grains         | 15.7% in chorizo and 10% in baked ham |                                                                                                                                                                                                              | AOM/DSS 1-induced CRC                                                         | ↑Firmicutes ↑Bacteroidetes                                                  | [123]    |

8. Mechanistic Attributes of Dietary Fibres on Cancer

Increased carbohydrate intake promotes bacterial cell growth, resulting in laxative effects and faster colon transit times. Protein degradation and the accumulation of potentially pathogenic chemicals such as ammonia, phenols, amines, and hydrogen sulphide in the colon are decreased as transit periods are shortened. Dietary fibre has several health benefits, including laxation, mineral absorption, anticancer, lipid metabolism, anti-inflammatory, and anticancer effects [124]. The chemical degradation of dietary fibre into SCFAs inside the colon is responsible for many of these health advantages. The colonic bacteria produce these SCFAs, and an equation describing total carbohydrate fermentation inside the colon was established [125,126]:

$$59 \text{C}_6\text{H}_{12}\text{O}_6 + 38 \text{H}_2\text{O} \rightarrow 60 \text{acetate} + 22 \text{propionates} + 18 \text{butyrate} + 96 \text{CO}_2 + 256 \text{H}^+$$

Dietary fibre improves viscosity, faecal bulking reduces proteolytic fermentation time and the amount of interaction between potential carcinogens and mucosal cells [127,128]. Furthermore, dietary fibre could bind/excrete possible luminal carcinogens (e.g., secondary bile acids) as well as lower faecal pH in the colon [124,129]. In addition, dietary fibre is a source of vitamins, minerals, and carbohydrates, and it produce digested energy [130]. Phytochemicals such as phenolics, carotenoids, lignans, beta-glucan, and inulin are linked to dietary fibre [130,131]. It has the potential to protect the GI tract from oxidative stress [132]. Bioactive compounds originating from dietary fibre sources can be found in considerably higher concentrations in the intestinal lumen than in plasma or other tissues, suggesting that these phytochemicals may help to prevent colon cancer.

SCFAs are ligands that bind to specific G-protein coupled receptors (GPCRs) on colonic and immune cells, in addition to providing energy [133]. As a result, they can serve as signalling molecules in the large intestine, reducing the production of proinflammatory cytokines and increasing the total number of regulatory T (Treg) cells [134]. The primary GPCRs that bind particularly to SCFAs are GPRC43 (FFAR2), GPRC41 (FFAR3), and GPRC109A. The GPRC109A appears to be more selective to butyrate than GPRC41 and GPRC43, which can bind to butyrate, propionate, and acetate [133]. These receptors are responsible for the majority of anticarcinogenic changes in the gut microbiome [135–138]. SCFAs, for instance, activate GPRC43 on (Treg) cells, causing them to expand and inhibit procarcinogenic inflammation [139]. A study conducted on mice treated with 50% chitin-glucan and 50% raw potato starch as a prebiotic in colon-specific polyposis revealed that the GPRC109A expression was enhanced, and the tumour numbers were reduced when butyrate levels increased [135].
The acetate, propionate, and butyrate are the three primaries colonic SCFAs, with total SCFA concentrations in colonic content exceeding 100 mmol/L [47]. The molar proportion of SCFA components is determined by the content of the diet intake and the gut microbiota. Acetate accounts for 60% to 75% of total SCFA and is produced by a variety of bacteria in the colon, with reductive acetogenesis accounting for around one-third of the total [140]. Propionate and butyrate are formed by specific bacterial groups that are of great interest due to their health benefits. The most important propionate-yielding microbiota in the human colon are still being discovered, and various metabolic routes for propionate production have been identified [141,142].

Cluster IV bacteria associated to *Faecalibacterium prausnitzii* and cluster XIVa bacteria related to *Eubacterium rectal* and *Roseburia spp.* account for 7% to 24% of total gut microbiota in healthy persons [143]. In rat colonic smooth muscle, acetate increases normal crypt cell proliferation but decreases the frequency of spontaneous longitudinal muscular contractions [144]. Through interactions with the GPCR43 and GPCR41 in adipose tissue and immune cells, acetate improves ileal motility, enhanced colonic blood flow, and plays a function in adipogenesis and the host immune system [145,146]. Acetate lowers lipopolysaccharide-stimulated (TNF), (IL)-6, and nuclear factor (NF)-kβ levels while increasing peripheral blood antibody production in several organs [147]. Propionate decreases human colon cancer cell proliferation and differentiation by hyperacetylating histone proteins and stimulating apoptosis, which may protect against carcinogenesis [148,149]. Propionate also inhibits the formation of pro-inflammatory cytokines (TNF-α and NF-kβ) in a variety of organs [150,151]. Butyrate has potent anti-inflammatory activities, which are likely due to reduction of TNF-α production, NF-kβ activation, and IL-8, -10, and -12 expression in immunological and colonic epithelial cells [152,153].

Adenomatous polyposis coli (APC) gene interaction with beta-catenin was discovered, and loss of APC function led to excessive T-cell factor (TCF4)/beta-catenin signalling [154]. These studies proved that Wnt signalling and human CRC are directly related. In colorectal cancer, loss of APC is the primary regulator of Wnt signalling [155]. Along with APC, mutations in the R-spondin/Lgr5/RNF43 module have also been linked to the promotion of Wnt-dependent tumour growth. Only 19% of CRC cases have been reported to have harmful RNF43 mutations, which are exclusive of APC alterations [156]. Because RNF43 mutant CRC are highly dependent on Wnt secretion, they can be treated with drugs that specifically target Wnt secretion [3,155].

### Effect of SCFAS on Cell Cycle and Colorectal Cancer

Butyrate restrict HDACs, allowing for histone hyperacetylation, which result into the transcription of a number of genes, including p21/Cip1 and cyclin D3 [157]. Butyrate inhibits migration and invasion of cancer cells by raising antimetastasis genes expression and reducing the activation of pro-metastatic genes at 0.5 or higher mmol/L concentrations [158,159]. Dietary fibre helps to prevent colon cancer in its early stages. Carbohydrates may prevent colonocytes against genotoxicity caused by high-protein, high-fat Western diets. As a result, resistant starch reduces DNA damage in colonocytes expressed by single strand breaks by 70% [160]. If this DNA damage is not repaired, colonic carcinogenesis can occur, and resistant starch protects against it [161]. The increased generation of SCFAs, as well as lower phenol and ammonia levels, may explain the preventive role of resistant starch against such DNA changes [160]. Butyrate, one of the SCFAs, has been shown to have a biological effect on neoplastic colonic cells [162]. Dietary sodium gluconate raises the level of butyrate and lowers the number of colon cancers [163]. Oral administration of bacteria that produce butyrate *Butyrivibrio fibrisolvens* increased the level of butyrate in the colon and rectum and decreased the production of aberrant crypt foci, an early colonic lesion [164].
9. Dietary Modification

Dietary modification can result in significant differences in the risks and occurrences of a variety of malignancies. Natural dietary compounds obtained from fruits, vegetables, and herbs have recently gained a lot of attention as chemo preventive and chemotherapeutic medicines all around the world [165]. Many researchers have endorsed the strategy of cancer prevention, employing non-toxic, new plant-derived medicines. Most plants contain flavonoids, which have a wide range of physiological actions, including antioxidant, anticarcinogenic, antibacterial, immune modulating, anti-inflammatory, and antiviral properties [165–168].

Fisetin (3,3′,4′,7-tetrahydroxyflavone) is a phytoconstituent which is a flavanol derivatives found in strawberries, apples, persimmons, grapes, onions, and cucumbers in concentrations ranging from 3 to 165 µg/g [166]. Antioxidants derived from food such as fisetin are being studied for their health-promoting properties, including their potential significance in cancer chemoprevention [165]. Camptothecins and their analogue are a group of terpene alkaloids derived from the Chinese plant Camptotheca acuminata (Nyssaceae), which have shown to be effective against colorectal cancer (CRC). CRC is a serious health problem that affects people all around the world. Despite major advances in treatment, this condition continues to cause severe morbidity and mortality [5]. Therefore, a diet rich in dietary fibres may possess a highly beneficial role in cancer prevention in general and CRC in particular.

10. Conclusions

Much literature suggests that dietary fibre consumption plays a crucial impact in general metabolic health, via key pathways such as T reg. cells, Wnt signalling pathway, and GPCRs. There are clear links between dietary fibre consumption and a variety of diseases, including cardiovascular disease, colon health, gut motility, and CRC risk. The gut microbiota plays a key role in the positive benefits of dietary fibre, including appetite management, metabolic activities, and chronic inflammatory pathways. The deficiency of dietary fibre in the typical Western diet is caused by several factors. A large population have been adapted to our modern surroundings, lifestyles, diets, eating habit, and consumption of ultra-processed meals. Our gut microbiota, similar to our bodies, did not evolve to embrace this nutritional maladaptive behaviour. Our diet serves as a predictor of general health and wellbeing, including various advantages mediated by our gut bacteria. Many features of our food are troubling (such as excessive consumption of carbohydrates and fat), and the evident lack of dietary fibre in our modern diet is particularly concerning. As food consumers, preference for high-fibre foods over fibre-poor ultra-processed foods is likely to have a significant positive impact on our future health and wellbeing, and will eventually influence food companies’ strategic commercial plans, with likely improvements in the fibre content of processed foods.

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