Enolase 1, a Moonlighting Protein, as a Potential Target for Cancer Treatment

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Abstract

Enolase 1 (ENO1) is a moonlighting protein, function as a glycolysis enzyme, a plasminogen receptor and a DNA binding protein. ENO1 play an important role in the process of cancer development. The transcription, translation, post-translational modifying activities and the immunoregulatory role of ENO1 at the cancer development is receiving increasing attention. Some function model studies have shown that ENO1 is a potential target for cancer treatment. In this review, we provide a comprehensive overview of the characterization, function, related transduction cascades of ENO1 and its roles in the pathophysiology of cancers, which is a consequence of ENO1 signaling dysregulation. And the development of novels anticancer agents that targets ENO1 may provide a more attractive option for the treatment of cancers. The data of sarcoma and functional cancer models indicates that ENO1 may become a new potential target for anticancer therapy.

Key words: Enolase 1; Moonlighting Protein; transduction cascades; Cancer treatment

Introduction

As a moonlight protein, Enolase 1 (ENO1) performs multiple biochemical functions. ENO1 catalyzing 2-phosphoglycerate (2-PGA) to phosphoenolpyruvic acid (PEP), a vital step in the glycolysis, plays an important role in several pathophysiological processes (1). ENO1, anchored on the cell membrane, servers as a receptor for activating plasminogen to stimulates the migration and invasion ability of cells (2). Furthermore, ENO1 also displays multiple binding capacity to the DNA, mRNA (3), LncRNA and tRNA(CUULys) to regulate the gene transcription and translation in cancer cells (4,5). Since the energy of cancer cells mainly rely on the glycolysis, so that ENO1 has been developed as an important target in cancer treatment.

The expression levels of ENO1 are closely associated with several diseases, including Alzheimer’s disease (6), diabetes (7), hypoxic-ischemic encephalopathy (8), as well as some types of cancers (9–16). ENO1 has been found to be overexpressed in more than 10 types of human cancer, and is related with metabolic reprogramming of cancer cells and cancer associated transduction cascades. And, the expression of ENO1 in a variety of cancer types have been found to be dependent on the pathophysiological conditions. In the cancer metabolic reprogramming, ENO1 stimulates cancer cells to create energy largely by disintegration of glucose in a non-oxidative manner rather than typical oxidative phosphorylation (17,18). The switching of ENO1 location is related to the pathology of cancer and inflammatory. The inflammatory stimulation could induce ENO1 translocation from the cytosolic to the cell membrane (7,19).
As the important pathophysiological biomarker in the cancers, targeting ENO1 or ENO1-mediated signaling pathways has been received the most attention. Over the years, several ENO1 inhibitors have performed promising results of the pharmacological-based study and preclinical trials (20,21). As the non-enzymatic active site inhibitor of ENO1, ENOblock reflects cytotoxicity in hypoxic and normal conditions on colon cancer cells(22). In addition, a series of enzymatic active inhibitors of ENO1, such as phosphonoacetohydroxamate acid (PhAH), (1,5-dihydroxy-2-oxopyrrolidin-3-yl)phosphonic acid (SF2312), deoxy-SF2312, Methyl-SF2312, POMSF, POMHEX and its derivatives, show potential anticancer in the functional cancer models (21,23). Furthermore, overexpression of ENO1 contributes to immune response and autoimmunity in the patients (24–27). The recently studies show that ENO1 is also an effective target for anticancer immunotherapy (28). ENO1 binds with guanylate binding protein to negative regulate T cell signaling by interfering with early T Cell Receptor signaling (29). And the post-translational modifying activities is also critical to the function of ENO1 in the immunity (26,30,31). A landmark study suggests that citrullinated ENO1 peptides could be an effective vaccines for cancer immunotherapy (32). In our previous study, we found that ENO1 is a novel cancer therapeutics (33).

The structure and function of ENO1

The cytogenetic location of ENO1 is located at the locus 1p36.23. The ENO1 gene spans about 18 kb and consists of twelve exons and eleven introns. Hypoxia-response elements (HREs), the sequence NCGTG, are located in the ENO1 promoter. The expression of ENO1 is also regulated by hypoxia-inducible factor 1 (HIF-1) recognizing to HREs in the genome for responding the hypoxia stress (34). Because of ENO1 gene lacks the canonical TATA box, so that the ENO1 possessing multiple transcription starts sites (35). The ENO1 gene encodes both ENO1 and c-myc binding protein (MBP-1), and mounting shreds of evidence have indicated that both ENO1 and MBP-1 play pivotal roles in tumorigenesis. The isoform of ENO1 gene, MBP-1 expression is regulated by the protein kinase B (AKT)/ eukaryotic initiation factor 2α (eIF2α) transduction cascades to response cell stress. The study implies that alternative splicing of ENO1 mRNA could be controlled by the AKT/eIF2α axis (36). There are three flexible active site loops of ENO1 are named as L1 loops (residues 36-43), L2 loops (residues 156-162) and L3 loops (residues 262-270) in crystal structures of ENO1, respectively(37). And two magnesium ions bind to Glu292, Asp317, Asp244 and Ser39, they are the key active site residues of ENO1. In these studies, the target drug is always interacting with the activity residues of ENO1.

As moonlighting protein, ENO1 performs distinct functions and involves in multiple cellular processes in cells. As phosphopyruvate hydratase, ENO1 participates in the 9th step of glycolysis and is responsible for the catalysis of the conversion of 2-PGA to PEP, which in turn is converted by pyruvate kinase to pyruvate. The Ser41 of ENO1 show closer to the phosphate group of the 2-PGA and directly interacts with the second Mg2+ of ENO1 (38). As a plasminogen receptor, ENO1 converts plasminogen to plasmin, which involves in the metastatic spread of cancer from the primary tumor to a remote site. In the intravascular and pericellular fibrinolytic system, ENO1 activates plasminogen (PLG) on the cell surface of several cell-types, such as leukocytes and neurons (39). The Lys345, Glu211 and the metal cations of ENO1 forms an essential part of the reaction mechanism to activate the plasminogen. The Lys345 of ENO1 plays an important role in capturing the R-proton of plasminogen, and then Glu211 is protonated and hydrogen bonded to the a-hydroxyyl group of plasminogen (40). The putative plasminogen-binding motif of ENO1 (residues: 250-256) and three hydrophobic and two hydrophilic residues play a critical role in the enzyme capacity of ENO1 (1). As a DNA binding protein, the residues of 97-237 are required for binding capacity of ENO1 with c-Myc promoter to repress tumorigenic (41).

Post-translational modifications of ENO1 play an important role in the function change of ENO1

Post-translational modifications (PTMs) of ENO1 are the important mechanism of regulation of protein function and cellular transduction cascades events that orchestrate biological functions in mammalian cells. Acetylation, phosphorylation, succinylation, hydroxyisobutyrylation (Khib) and methylation of ENO1 can alter the catalytic activity, localization, protein stability and binding capacity of ENO1.
Investigation of these modification patterns in different human cancer cells have provided insights into its important role in pathophysiological processes (42). As shown in the Table 1, the 44 modified residues of ENO1 have been mention, identified and investigated in the decades.

Table 1. The post-modification residues position and the function of ENO1

| Ontology               | Residues position(s) | Reference | Function change                        |
|------------------------|----------------------|-----------|----------------------------------------|
| N-acetylserine         | 2                    | Jacome et al. (43) | unknown                               |
| N6-acetyllysine        | 5, 60, 64, 71, 89, 92, 126, 193, 199, 202, 228, 233, 256, 257, 281, 283, 285, 326, 335, 343, 406, 420 | Substrate binding capacity             |
| N6-succinyllysline     | 60, 80, 81, 89, 228,335, 420 |  |                           |
| Phosphoserine          | 27, 115, 282, 254, 263, 272, 282, 291 | Dephoure et al. (50), Zhou et al. (51), Bian et al.(52), Mayya et al. (53), Tertyt Yang et al. (54) | Cellular glycolysis                      |
| Phosphotyrosine        | 44, 287              | Rush et al. (55), Ross SH et al. (56) | Cytotoxic T cells capacity             |
| N6-malonyllysine       | 233, 420             | Peng et al. (57), Nishida et al. (58) | unknown                               |
| Initiator methionine removed | 1                    | Bienvenut et al. (59) | unknown                               |
| Argininemethylation    | 50                   | Zakrzewicz et al. (30) | Cell invasion and migration            |
| Khib                   | 58, 228, 281, 343    | Huang et al. (60), Dong et al. (48) | Cellular glycolysis                    |
| Phosphoglycerylation-lysine | 343               | Moellering et al. (61) | Cellular glycolysis                    |
| Citrullination         | 15, 253              | Katherine Cook et al. (32) | Auto-immune                           |
| Ubiquitin-like modifiers | Unknown              | Giannakopoulos et al. (46), Wong et al. (62), Peng et al. (63) | unknown                               |

The enzyme activity of ENO1 is always modulated by the various PTMs in the glycolysis. Acetylation modified of protein is one of the major PTMs (43,44). Acetylation lysine residue 335 of ENO1 by histone deacetylase 11 (HDAC11) causes loss of ENO1 activity and suppress the glycolysis in tumor cells (64). Acetylation lysine residues 257 and 283 of ENO1 could disrupt both the electrostatic binding potential and the geometry of the binding site and perturbing substrate binding capacity of ENO1 to inhibited the glycolysis in *Synechococcus elongates* (45,65). Moreover, the enzyme activity of ENO1 and others are regulated by acetylation and succinylation modified in metabolism, thus mediating the anticancer drug effect of dichloroacetate is related with the glycosis (66). The succinylation and desuccinylation of ENO1 is co-regulated by carnitine palmitoyltransferase I (CPT1A) and sirtuin 5 (SIRT5), respectively. Kiran Kurmi et al. indicate that CPT1A inhibits enzymatic activity of ENO1 by succinylation of ENO1 to promote hematopoietic cell proliferation under glutamine depletion. And the succinylation of ENO1 lysine residues 80, 81 and 335 are also the important sites for the catalytic activity of ENO1 (49,67). Khib may directly mediate environmental influences on the epigenome and biological processes. To regulate the glycolysis of colorectal carcinoma cell HCT116, the surface lysine residues of ENO1 could be Khib modified by p300 protein (60). However, the NAD-dependent protein deacetylase (CobB) regulates the catalytic activities of ENO1 by removing ENO1K343hib and ENO1K326ac simultaneously in prokaryotes. That meaning that the Khib of ENO1 play an important role of regulating of cellular glycolysis and cell growth (48). Previous studies have shown that phosphoglycerate kinase (pgK) of metabolism enzymes play an important role of regulating the glycolytic process and glucose metabolism (68–70). Enhancing pgK modification of ENO1K343 causes the decreasing of enzyme activities of ENO1 (61). The acetylation, succinylation, Khib and pgK modification play critical role for the catalysis capacity of ENO1 in the glycolysis.

As a direct substrate of the serine/threonine-protein kinase (ULK1), the phosphorylation of ENO1 protect cells from ROS-associated cell death (54). Furthermore, ENO1 phosphorylation is also regulated by JAK-independent signaling controlled PIP3 levels and AKT activity in cytotoxic T cells (56). Arginine residues 50 of ENO1 is methylated by protein arginine methyltransferase 5 (PRMT5) that plays an important role of regulating of cellular metabolism enzymes play an important role of regulating of cellular metabolism enzymes (30). The interferon (IFN)-stimulated gene product 15 modification (ISGylated) of ENO1 is found in the human lung adenocarcinoma epithelial cell line A549 and other bacterial (46,62,63), however the function of ISGylated ENO1 remain unclear. The PTMs of protein is closely related with the function and structure of protein, the details of PTMs of ENO1 might reveal some function characterization in the change of structure. As shown in the Table 1, the single residue might be different modified types. And the most modifications of ENO1 are related with the substrate binding capacity, catalyst activity, cellular glycolysis and cell invasion and migration in cancer. The enzyme activity of ENO1 is always modulated by the various PTMs in the glycolysis. Acetylation modified of protein is one of the major PTMs (43,44). Acetylation lysine residue 335 of ENO1 by histone deacetylase 11 (HDAC11) causes loss of ENO1 activity and suppress the glycolysis in tumor cells (64). Acetylation lysine residues 257 and 283 of ENO1 could disrupt both the electrostatic binding potential and the geometry of the binding site and perturbing substrate binding capacity of ENO1 to inhibited the glycolysis in *Synechococcus elongates* (45,65). Moreover, the enzyme activity of ENO1 and others are regulated by acetylation and succinylation modified in metabolism, thus mediating the anticancer drug effect of dichloroacetate is related with the glycosis (66). The succinylation and desuccinylation of ENO1 is co-regulated by carnitine palmitoyltransferase I (CPT1A) and sirtuin 5 (SIRT5), respectively. Kiran Kurmi et al. indicate that CPT1A inhibits enzymatic activity of ENO1 by succinylation of ENO1 to promote hematopoietic cell proliferation under glutamine depletion. And the succinylation of ENO1 lysine residues 80, 81 and 335 are also the important sites for the catalytic activity of ENO1 (49,67). Khib may directly mediate environmental influences on the epigenome and biological processes. To regulate the glycolysis of colorectal carcinoma cell HCT116, the surface lysine residues of ENO1 could be Khib modified by p300 protein (60). However, the NAD-dependent protein deacetylase (CobB) regulates the catalytic activities of ENO1 by removing ENO1K343hib and ENO1K326ac simultaneously in prokaryotes. That meaning that the Khib of ENO1 play an important role of regulating of cellular glycolysis and cell growth (48). Previous studies have shown that phosphoglycerate kinase (pgK) of metabolism enzymes play an important role of regulating the glycolytic process and glucose metabolism (68–70). Enhancing pgK modification of ENO1K343 causes the decreasing of enzyme activities of ENO1 (61). The acetylation, succinylation, Khib and pgK modification play critical role for the catalysis capacity of ENO1 in the glycolysis.

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**ENO1 participates in the important transduction cascades of cancer**

**Hypoxia-induced ENO1 expression**

Hypoxia microenvironment leads to the expansion of aggressive clones of heterogeneous tumor cells, and promotes a lethal phenotype in the tumor (71). ENO1 is required for maintaining the Warburg effect of cancer cells. ENO1 responds to the hypoxic microenvironments in the rapid proliferation
cells. HREs of ENO1 gene is a key transcription element in the expression of ENO1. The H3K9 acetylation at HREs of ENO1 gene are significantly increased by mucin-1 (MUC1) overexpression in a hypoxia-dependent manner, and the mRNA of ENO1 show a significant increase (72). Moreover, ENO1 also is temporally distinct transcriptional regulation by HIF1α-HIF1β binding with HREs of ENO1 to increase the expression of ENO1 (73,74). Furthermore, the expression of ENO1 and other glycolysis genes also can be regulated by sineoculis homeobox homolog 1 (SIX1), which interact with acetyltransferase 1 (HBO1) and Nuclear receptor coactivator 3 (AIB1) to affect Warburg effect in cancer (75). In addition, SIX1 glycolytic function is directly repressed by microRNA-548a-3p. The miR-548a-3p/SIX1 axis regulates aerobic glycolysis by alteration expression of glycolysis genes in cancer cells (75). Knockdown of ENO1 increases ROS as well as tricarboxylic acid (TCA) cycle activity, induces a decline in nucleotide base synthesis and promotes cellular senescence mainly through the sorbitol in cancer cell lines (76). Zhang et al. indicate that the phosphoglycerate mutase–enolase–pyruvate kinase interacts with voltage-dependent anion channel protein (VDAC1) of the outer mitochondrial membrane and the triose phosphate translocator of the chloroplast, and the interaction is considerably more efficient than the free enzymes of the extra-plastidial preparation, as well as prevents the use of substrate by competing reactions such as phosphoenolpyruvate carboxylase and amino acid biosynthesis in the plant. And lacking enolase compromised the movement of their mitochondria as well as the degree of mitochondria-chloroplast colocalization, so that the interaction of enolase with mutase/pyruvate kinase is important for promoting a highly efficient coordination of the major energy systems of the plant cell (77). However, mitochondrial ENO1 interacting with VDAC1 prevents Ca2+-induced loss of mitochondrial transmembrane potential, swelling of matrix and release of cytochrome c to regulate cardiomyocyte apoptosis, a critical regulator of the mitochondrial cell death pathway in the mammalian cells (78). The interaction of ENO1 with others glycolysis enzymes should be clarified in the mammalian, that may be a critical mechanism for the tumorigenesis.

Elevation of ENO1 could promotes glycolysis, cellular proliferation and migration via activating the MAPK/ERK pathways. Dr. Chen et al. show that cytotoxin-associated gene A (CagA) activates the Src and MAPK/ERK to induce the elevation of expression of ENO1 in H. pylori-mediated gastric cancer. U0126 (selective inhibitor of both MEK1/2) and PP1 (inhibitor of Src kinase) can attenuate the upregulation of ENO1 in the wild-type H. pylori CagA+/-infected cell (79).

**ENO1 and PI3K/AKT**

AKT is a serine/threonine kinase that is involved in mediating various biological responses, such as inhibition of apoptosis and proliferation. The phosphoinositide 3-kinase (PI3K)/AKT signaling pathway is the major pathway for driving cancer cells to favor glycolysis over mitochondrial oxidation. Sun et al. demonstrate that ENO1 is a positive regulator of the PI3K/AKT axis for promoting gastric cancer cell proliferation and metastasis (80). Otherwise, Chen et al. elucidate that ENO1/AKT/PI3K signaling axis is mediated by WW Domain Binding Protein 2 (WBP2) modulating the expression and glycolysis activity of ENO1 to regulate the proliferation and metastatic ability of glioma cells (81). Furthermore, overexpression of ENO1 significantly increase the levels of β-catenin and phosphorylated focal adhesion kinase (FAK), PI3K, and AKT. They indicate that ENO1 is an upstream signal factor modulating the FAK/PI3K/AKT axis in non-small-cell lung carcinoma (NSCLC) to promote glycolysis, proliferation, migration and invasion of cancer cells (82). ENO1 contribute to the subsequent cellular response of cancer and may be a critical effector in PI3K/AKT pathway.

**ENO1 and AMPK/mTOR**

AMP-activated protein kinase (AMPK) respond on the external stress by gauging the AMP and ATP levels of the cell. In cancer, ENO1 induces the ATP production in HCT116 cells. Furthermore, ENO1 also promotes growth, tumorigenesis, migration, invasion and metastasis of colorectal cancer cells by AMPK/mTOR pathway. Mechanically, ENO1 inhibits phosphorylation of AMPKα while increases the phosphorylation of mTOR concomitantly to promote tumorigenesis and metastasis (83). However, the overexpression of ENO1 regulate proliferation, de-differentiation, resistance to apoptosis and the hypoxia-induced metabolic shift via the AMPKα/AKT/GSK3β in human pulmonary artery smooth muscle cells (PASMC). ENO1 up-regulate phosphorylation levels of AKT (T308 and S473) and GSK3β via the major kinase AMPK (84). Phosphorylated AMPK (T172) is observed in ENO1-deleted D423 glioma cell line, which induce growth inhibition and subsequent apoptosis (23,85).

**ENO1 and Wnt/β-catenin**

Canonical Wnt signaling controls key developmental gene expression programs by increasing nuclear and cytoplasmic β-catenin. Some studies demonstrate that ENO1 functions as an oncogene in
bladder cancer to regulate cell cycle and apoptosis by regulating β-catenin. Overexpression of ENO1 can up-regulate β-catenin as well as its downstream targets cyclin D1 to promote cell growth and proliferation (82,86). In addition, ENO1, protein disulfide isomerase family A member 3 (PDIA3) and podoplanin (T1α) involved in β-catenin driven trans-differentiation of murine alveolar epithelial cells. The Wnt10a and Wnt10b represent potential Wnt ligands to active Wnt/β-catenin signaling. Activated Wnt/β-catenin signaling regulates expression of ENO1, PDIA3 and carbonyl reductase 2 (CBR2) to induce the ATII-to-ATI cell trans-differentiation (87).

ENO1 and LncRNA

The expression regulation and dysfunction of ENO1 have a closely relationship with the hypoxic metabolic environment in the cancer development. And the long non-coding RNA is also an important regulator for the expression of ENO1. Dr. Liu, et al., find that LncRNA P5848 is upregulated in the mimic environment of radiofrequency ablation. LncRNA P5848 promotes HCC development by upregulating ENO1. Comparatively, blocking of LncRNA P5848/ENO1 axis can attenuate growth, survival and invasion capacity of cancer cells (12). However, the other LncRNA-6195 reduces enzymatic activity of ENO1 to repress the growth of HCC via directly binding with ENO1 (13).

ENO binding proteins and the MBP-1 related pathway

ENO1 is able to activate the CD14-dependent TLR4 pathway via binding with TLR4 on monocytes involving in a dual mechanism firstly pro-inflammatory and secondly anti-inflammatory in rheumatoid arthritis in vitro (88). Wygrecka et al. indicate that the ENO1 act as a plasminogen receptor (PLGR), which could promote invasion of monocytes into lungs in mice and humans (19). Further, they find that translocation of ENO1 can be mediated by Ca2+ through SOC channels related proteins. Stromal interaction molecule 1 (STIM1)/calcium release-activated calcium channel protein 1 (ORAI1) axis regulate LPS-induced translocation of ENO1 to the cell membrane surface. And, ENO1 is released into the extracellular space in the form of exosomes, and exosomal ENO1 associate with the metastatic potential of cancer cells (89). ENO1 also plays a vital role in urokinase receptor (uPAR)/integrins pathways in pancreatic ductal adenocarcinoma. The knockdown of ENO1 in CFPAC-1 cells results in a markedly increasing expression of uPAR. The binding of uPAR with VN triggers alpha v/beta 3 integrin activates serine/threonine kinase 1-2 (ERK) and Rac family small GTPase to accumulate ROS produce and senescence of pancreatic ductal adenocarcinoma (PDA) cells (90).

As alternatively spliced nuclear isoform of the ENO1 gene, the expression of MBP-1 is regulated by the AKT/elF2α axis to response cell stress. The study implies that mRNA alternative splicing of ENO1 could be controlled by the AKT/elF2α axis (36). In additional, MBP-1 suppresses tumorigenesis and regulates the development and metastasis of cancer cells by regulating genes expression, including c-Myc, COX-2 and ERBB2 (91). Moreover, MBP-1 regulates prostate cancer cell growth by inducing proteasome mediated degradation of the MEK5 to inhibit the MEK5/MAPK7-mediated activation of MEF2C and NF-kb (92). However, the mechanism of alternative splicing changes in the gene of ENO1 remain unclear in the cell stress response.

Targeting ENO1 in cancer therapy

The identification of oncogenic ENO1 has triggered the development of small molecule inhibitors and others agents targeting ENO1 for cancer treatment. The group of Florian L Muller have developed a serial of ENO1 inhibitors which perform antitumor activity in vitro and in vivo.

ENOblock

ENOblock (1) (Figure 4), non-glycolytic inhibitor of ENO1, induce nuclear translocation of ENO1 and down-regulation of phosphoenolpyruvate carboxy kinase to inhibit the growth and migration of cancer cells (7,93,94). Haaglim et al. identify that 1 can alleviate the pathology of diet-induced obesity by regulating the expression of srebp-1a and srebp-1c, PCK1, TNF-α and IL-6 (95). In our pervious study, granulin A increased glucose uptake of cells to inhibit the growth of HCC (94,96).

Derivatives of phosphonoacetohydroxamate

The substrate PEP coordinates with only one Mg2+ ion of ENO1 in the active site. Florian L Muller et al. develop a serious of derivatives from natural product SF-2312 (2) (Figure 4) which mimics the substrate phosphate group of 2-PGA. And the 2 perform the inhibition of glioma cells growth (23). HEX (3) and POMHEX (4) (Figure 4) is substrate-competitive enolase inhibitor with a Ki of 63 nM for ENO2 versus 250 nM for ENO1 (20). Mechanically, 4 diffuses into cell rapidly, and is hydrolyzed by high intracellular carboxylesterase activity and is trapped in the negatively charged mono-ester form. The mono-ester form of 4 is converted to the fully active through the action of phosphodiesterases-4 to inhibit
the proliferation of cells. Moreover, the ATP of glioma cells is reduced by 4, which indicate that 4 can block glycolysis in the tumor. As inhibitor of ENO1, the derivatives of 2 preferentially inhibits glycolysis in ENO1-null or isogenic ENO1-rescued glioma cells. ENO1-intact glioma cells can recover more quickly from enolase inhibition. 4 can eradicate intracranial orthotopic ENO1-deleted GBM tumors in mouse, despite sub-optimal pharmacokinetic properties. In addition, overexpression of ENO1 have been found in many cancer types. ENO1 is more important than other isoform in the glycolysis, and should be a potential target for cancer treatment.

The ginsenoside derivatives

Huang et al. synthesizes two ginsenoside derivatives, 20(R)-Rh2E2 (5) and 20(S)-Rh2E2 (6), which are structurally modified from parental compound 20(S)-Rh2 (97,98). The ginsenoside derivatives could inhibit the growth of cancer in vitro and in vivo by energy metabolism intervention, and the tumor bearing mouse are administrated with 20(S)-Rh2E2 up to 320 mg/kg/day and survived with no observable toxicity. Mechanically, the ginsenoside derivatives could down-regulated the expression of ENO1 in the cancer cells. And the growth of glycolysis and inhibition of invasion is the result of ENO1 downregulation in the cancer. The results shown that 5 and 6 might be the metabolic inhibitors of cancer with low toxicity for normal cells.

The other compounds targeting ENO1

The library of compounds has been screening by docking, and some chemicals show better binding energies values than the substrate 2-PG in computer-aided identification of potential tool. These compounds such as ZINC1304634 (7), ZINC16124623 (8), ZINC17027262 (9), tropolone (10), pyridine (11) and hydroxyquinoline (12) (Figure 4) may be considered promising anticancer agents for further development, and
could fight the metabolism of cancer by inhibiting ENO1 (99,100). Furthermore, upregulation of ENO1 by CagA can be attenuated by U0126 (13) (selective inhibitor of both MEK1/2) and PPI1 (14) (inhibitor of Src kinase). As shown above, ENO1 act as upstream or downstream regulators in the important signalling. The therapeutic inhibitor targeting to the ENO1 might offer the promising new treatment options for cancer treatment.

[Image: Figure 3. The schematic diagram illustrated that (A) ENO1 binding with TLR4, uPAR and Plg-Rs, and (B) MBP-1 (ENO1) mediated signal transduction. (1) ENO1 is able to activate the CD14-dependent TLR4 pathway via binding with TLR4 on monocytes involving in a dual mechanism firstly pro-inflammatory and secondly anti-inflammatory. (2) As a plasminogen receptor (PLGR), ENO1 could binding with Plg-Rs to promote invasion of monocytes into lungs in mice and humans. (3) ENO1 interact with uPAR to activates serine/threonine kinase 1-2 (ERK) and Rac family small GTPase to accumulate ROS produce and senescence of pancreatic ductal adenocarcinoma cells. (4) alternative splicing of ENO1 mRNA could be controlled by the AKT/eIF2α axis. (5) MBP-1 regulates prostate cancer cell growth by inducing proteasome mediated degradation of the MEK5 to inhibit the MEK5/MAPK7-mediated activation of MEF2C and NF-κB. (6) MBP-1 suppresses tumorigenesis and regulates the development and metastasis of cancer cells by regulating genes expression, including c-Myc, COX-2 and ERBB2.]

ENO1 in the immunotherapies of cancer

ENO1 and antibody of ENO1 in the serum is associated with the clinical stage of cancer. The expression of ENO1 and antibody of ENO1 in the serum can be detected in some of oral squamous cell carcinoma patients (31,101). Meanwhile, there is a positive correlation between auto-antibody levels of ENO1 and expression of phosphorylated ENO1 or ENO1 in tumor tissue, suggesting that ENO1 might be a biomarker of cancer (16,102–105). The ENO1 are upregulation in the majority of clinically relevant cancers, and serum ENO1 auto-antibody have been reported in wide range of cancers. Zhang et al. demonstrate that serum ENO1 auto-antibody levels is associated with the clinical stage of lung cancer in the patients (104). The other study implies that serum ENO1 antibody levels are significantly higher in the lung cancer than those in the control or in the benign lung disease groups (P < 0.001). Serum ENO1 auto-antibody levels in the benign lung disease group is significantly higher than those in the control group (P < 0.05). Serum ENO1 auto-antibody levels in the patients in stages I and II were significantly higher than those in the patients with advanced stages III and IV (P < 0.01). These result show that auto-antibody level of ENO1 is associated with the clinical stage of lung cancer. But the detail mechanism is remaining unclear. The another study also identify that the decreasing of serum ENO1 antibody is a marker in late stage of NSCLC, SCLC and breast cancers (28). The levels of ENO1 auto-antibody is significantly decreased in both Stage IV of NSCLC and SCLC when compared with those of the normal individuals (P < 0.001). In Stage IV breast cancer, the data show that the patients have dramatically lower levels of the ENO1 auto-antibody (P < 0.001) than healthy individuals (n = 99) or female controls (n = 49). The study indicates that the titer change of ENO1 is a key factor associating with tumor malignancy of NSCLC, SCLC and breast cancer patients (28). The ENO1 may be a potential marker for the cancer immunotherapy.

The Sun et al. have shown the expression of ENO1 is positively associated with Ki67 expression in the tumor tissues (106). However, the expression of ENO1 is negatively correlated with p53 expression in the tumor tissues. And expression of ENO1 is significantly associated with unfavorable survival. The level of ENO1 (HR=2.469; 95% Cl: 1.348-4.522; P=0.003) is an independent prognostic for pancreatic patients by Cox multivariate regression analysis. The DNA vaccination of ENO1 could slow progression of PDA by promoting the respond of Th17 T cell, while inhibits expansion of MDSC and Treg cell in the PDA tumor-bearing mice (107). In recently, both DNA vaccination of ENO1 and the combination treatment of DNA vaccination of ENO1 and gemcitabine treatment have shown a significant reduction of PDA lesions compared with untreated in the pancreatic cancer-prone KC mice. They find that the combination of DNA vaccination of ENO1 and gemcitabine shows a significant reduction of circulating CD4 and CD8 T cells, which supports the increasing infiltration of these cells in tumor lesions in the pancreatic cancer-prone KC mice. And CT also could increase the coordinated immune response to DNA vaccination of ENO1 and the combination treatment in the KC mice with PDA (108).
In recently, the preclinical evidence demonstrates that targeting ENO1 induces plasmacytoid dendritic cells triggered T and NK cell mediated anti-multiple myeloma activity (99). Arghya et al. indicate that ENO1 inhibitor could increase plasmacytoid dendritic cells to activate multiple myeloma specific CD8+ T cells and NK cell against autologous tumor cells. As well as the combination of ENO1 inhibitor and anti-PD-L1 antibody or HDAC6 inhibitor synergistically enhances the activity of autologous multiple myeloma specific CD8+ T cells (109). The further research indicates that modification peptides deriving from ENO1 protein can stimulate the CD8+ T cells and CD4+ T cells, peripheral blood mononuclear cells (PBMCs) and tumor infiltrating lymphocytes (TILs) to release cytokine, such as like IFN-γ, TNF-α, as well as the expression of activation marker like CD107a (31). The vaccine targeting to ENO1 have shown encouraging results in the animal model (32).

Cook Katherine et al. indicate targeting post translation modified epitope of ENO1 may provide a new strategy for generation tumor specific immune responses. They show that citrullinated epitopes from ENO1 can be used in vaccines to induce potent CD4+ T cell responding to an anti-tumor effect with minimal cross reactivity to healthy tissue. Transgenic mice model demonstrate that immunization with citrullinate ENO1 peptide can simultaneously induce CD4-mediated IFN-γ responses. The citrullinated ENO1 peptides provides a significant advantage survival in the mice model. The study also implies that the CD4+ T cells indirect recognize tumor, and direct recognition of the citrullinated epitopes on MHC class II in the tumor is more potent for cancer therapy. Moreover, the studies also identify that
healthy human has a repertoire of CD4+ T cells capable of responding to the citrullinate ENO1 peptide. Citrullinated ENO1 epitope can be used to generate strong Th1 responses that can preferentially target tumor cells (32). The studies of ENO1-related immune respond in the cancer is a promised thought for cancer immune therapeutics. In the further, the ENO1 related immune in the cancer development should be further investigated to assess the immunotherapies of cancer.

Conclusion and future perspective

ENO1 is almost 47.2 kilo Dalton. With other two isozyme subunits ENO2 and ENO3, ENO1 could form homodimers heterodimers. The alternative splicing of ENO1 mRNA results in an isoform protein that could binding with the promotor of c-myc to regulating the cell proliferation. The composition of the secondary structure in ENO1 are 21 α-helices, 13 β-sheets, and 4 random coils. Large number of evidences the ENO1 plays a critical role in human cancer and is inappropriately activated in a large fraction of cancers through a variety of different mechanisms. Metabolic reprogramming of tumor associate with anabolic glucose metabolism enzyme, such as ENO1, PGD, GLU1, PKM1/2 and so on. ENO1 can dramatically disparately responds on the cell stress and induces signaling reprogramming due to the distinct change of ENO1, such as alternative splicing, translocation, PTMs of ENO1 and so on. Increased ENO1 expression is detected in major of tumor tissues, and is positive correlation with multi-drug-resistance of cancer cell and poor survive of patients. In recent years, comprehensive biochemical and functional studies have elucidated that ENO1 is a key target for therapeutic development.

Alternative splicing of ENO1 may be an exquisite sense for cancer development. In contrast, translocation of ENO1 and over-expression of ENO1 on the membrane can promote tumor development and metastasis. Inflammatory stimulation also induces translocation of ENO1 from the cytosolic to cell membrane. The switching of ENO1 location is related with pathology of cancer development and inflammatory (19). The transcription, translation, PTMs of ENO1 and even translocation alteration of ENO1 is associated with the rearrange of cellular process. However, the mechanism of how ENO1 transfer in the cell surface remains unclear, and which factor is the required for cancer development should be further authenticated. So that the future clinical trials of novel strategies targeting ENO1 should include careful pharmacodynamics assessment, through paired pretreatment and on-treatment tumor biopsies to ascertain the specific transduction

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3-yl phosphoryl)bis(oxy))bis(2,2-dime thylpropanoate); PRMT5: Protein arginine methyltransferase 5; PTMs: Post-translational modifications; SCLC: Small cell lung cancer; STIR5: Sirtuin 5; S6X1: Sineoelus homeobox homolog 1; Srebp: Sterol regulatory element-binding proteins; STIM1: Stromal Interaction Molecule 1; T1a: podoplanin; TCA: Tricarboxylic acid; TCR: T cell receptor; TILs: Tumor infiltrating lymphocytes; TLR4: Toll Like Receptor 4; TNF: Tumor Necrosis Factor; ULK1: Serine/threonine-protein kinase; uPAR: Urokinase receptor; VDAC1: Voltage-dependent anion-selective channel 1; VN: vitronectin; WBP2: WW Domain Binding Protein 2.

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Author Contributions

Material preparation and writing—original draft preparation, Gan Qiao; writing—review and editing, Xiaoliang Chen and Xiukun Lin; investigation and resources, Gan Qiao, Xiaoliang Chen, Ye Tian and Anguo Wu.; visualization, Gan Qiao and Anguo Wu; project administration, funding acquisition, conceptualization and supervision, Xiukun Lin. All authors have read and agreed to the published version of the manuscript.

Competing Interests

The authors have declared that no competing interest exists.

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