DOPAMINE RECEPTOR IN ANTERIOR BYSSUS RETRACTOR MUSCLE OF MYTILUS EDULIS

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Abstract—Effects of dopamine, N-methyl-, ethyl- and propyl-derivatives of dopamine, and alpha- and beta-adrenoceptor stimulants on catch contraction of anterior byssus retractor muscle of Mytilus edulis were tested. The test drugs except the beta-adrenoceptor stimulants relaxed catch contraction. Dopamine was most active and substitution of amino group in dopamine with ethyl and propyl decreased activity considerably. The concentration-action curves of dopamine, its derivatives and norepinephrine shifted in parallel with application of haloperidol but were not influenced by the alpha- and beta-adrenoceptor antagonists. These results suggest that relaxation of catch contraction by catecholamines is mediated through a dopamine receptor. This muscle is considered to be suitable for a study of the dopamine receptor.

Dopamine relaxes catch contraction in the anterior byssus retractor muscle of Mytilus edulis and is present in the ganglia of Mytilus (1, 2). However, little is known of dopamine receptors in this muscle. We investigated the effects of dopamine and its derivative on catch contraction of this muscle and compared the findings with the effects of alpha- and beta-adrenoceptor stimulants.

MATERIALS AND METHODS

Sea mussels, Mytilus edulis L., collected from the east side of Tokyo bay were used. Mytilus edulis were stored in aerated artificial sea water (NaCl 456 mM, KCl 11 mM, CaCl₂2H₂O 11 mM, MgCl₂6H₂O 48 mM and Tris-HCl 25 mM; pH 7.8–8.0) at a temperature of about 10°C and used within 7 days after collection. Muscle bundles (about 1 mm in diameter) dissected from the anterior byssus retractor muscle under a pair of forceps were suspended in 10 ml organ bath filled with artificial sea water bubbled with air and kept at 23 to 25°C. Responses to drugs were recorded isotonically under a tension of 0.2 g. After a 1 hr immersion, the muscle was exposed to acetylcholine (10⁻⁴ M) for 2 min and washed with artificial sea water. We used the muscle in which the relaxation at 5 min after washout of acetylcholine was less than 25% of the maximum relaxation which was obtained by application of serotonin (10⁻⁶ M). The test drugs were applied at 5 min after washout of acetylcholine and relaxations following a 10 min exposure to the test drugs were estimated.
muscle had been exposed to the test drug for 10 min. These relaxations were expressed as a percent of the maximum relaxation by serotonin (10^{-6} M) (3). After a 2 min exposure to acetylcholine (10^{-4} M) and washing with artificial sea water, one of the antagonists was applied for 5 min and the test drug was given for 10 min in the presence of the antagonist in order to assess the antagonism between the test drugs and antagonists. The pA2-values of the antagonist were calculated from parallel shifts of the concentration-action curves of the test drugs (4). Drugs used: Dopamine hydrochloride, N-methyl(epinine), N-ethyl- and N-propyl-dopamine hydrochlorides, which were synthesized by Takagi et al. (5), apomorphine hydrochloride (Sankyo), norepinephrine bitartrate (Wako), isoprenaline hydrochloride (Sigma), trimetoquinol hydrochloride (Tanabe), prazosin hydrochloride (Pfizer), propranolol hydrochloride (Sumitomo-Kagaku) and serotonin creatinine sulfate (Wako), all in powder form. All concentrations used were presented as salt forms.

RESULTS

Preliminary tests in this study suggested the possibility that the variance between the responses to the test drugs in muscle bundles obtained from different groups of Mytilus edulis was larger than that within responses of muscle bundles from the same group of sea mussels. Therefore, the results in our series of experiments represent data obtained from mussels at the same time of year and from one group of sea mussels.

After exposure to acetylcholine and washing with artificial sea water, the muscle spontaneously relaxed to 23.0±2.5% (mean ±S.E.) (Figs. 1–3). The maximum responses to all the test drugs were significantly smaller than responses to serotonin. At present we had no explanation for these phenomena (Figs. 1 and 2). Dopamine was the most potent relaxant while isoprenaline (up to 3×10^{-4} M) and trimetoquinol (up to 3×10^{-4} M), beta-adrenoceptor stimulants were inactive. Apomorphine, a dopamine receptor stimulant was a much less active relaxant than dopamine (Fig. 2). The responses to norepinephrine and dopamine...
were not influenced by an alpha-adrenoceptor antagonist, prazosin (10⁻⁷ M) and a beta-adrenoceptor antagonist, propranolol (10⁻⁶ M) in sufficient concentrations to block the responses of the mammalian smooth muscles through the corresponding receptors (Fig. 3). These results suggest that the alpha-and beta-adrenoceptors do not play an important role in relaxing processes of this muscle induced by the test drugs. The concentration-action curves of dopamine, N-ethyl- and N-propyl-dopamines and norepinephrine were shifted in parallel by a specific dopamine antagonist, haloperidol (3×10⁻⁵ M), thereby suggesting a competitive antagonism (Figs. 1 and 2). The pA₂-values of haloperidol were 6.1±0.21 against dopamine, 6.0±0.23 against N-ethyl dopamine, 6.2±0.24 against norepinephrine. The values are presented as means±S.E. of 6 experiments. Substitution of the amino group in dopamine with ethyl and propyl resulted in a considerable reduction in the activity, while N-methyl-dopamine (epinine) was as active as dopamine (Fig. 1).

**DISCUSSION**

Dopamine is reportedly released from inhibitory nerves and the catch contraction changes to a response of relaxation (1, 2, 6). However, little is known of dopamine receptors in the anterior byssus retractor muscle of Mytilus edulis. The beta-adrenoceptor stimulants, isoprenaline and trimetoquinol did not relax the catch contraction and relaxation by dopamine and norepinephrine was inhibited by the dopamine antagonist, haloperidol but not by the alpha-adrenoceptor antagonist, prazosin and the beta-adrenoceptor antagonist, propranolol. The present results coincide with the findings of others (7), as related to isolated arteries, including renal, mesenteric and femoral, and suggest that relaxation of this muscle by catecholamines is mainly mediated through the dopamine receptors.

The pA₂-values of haloperidol to dopamine coincided with a negative logarithm of Kᵢ-values for haloperidol against the specific binding of ³H-dopamine to human caudate, calf caudate and rat striatum (8). Being active in this muscle, apomorphine was much weaker. This result is in agreement with findings on the dopamine receptor in vascular smooth muscle (9). Substitution of the amino group with hydrocarbons larger than methyl yields compounds which have a lesser effect on the dopamine receptors in this muscle. Thus, amine or methylamine would probably be suitable for the cationic head as this head is probably important for the dopamine-receptor interaction (5). Trimetoquinol did not stimulate the dopamine receptors. On the basis of the above results alone, it would appear that the conformation of dopamine required to interact with its receptors is unrelated to the interrelationship between catecholamine and nitrogen in
trimetoquinol.

Since responses via the alpha- and beta-adrenoceptors were not apparent, this muscle is probably suitable for studies on the dopamine receptors.

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