Abstract. The aim of the present study was to investigate the effects of propranolol and isoproterenol on the growth curve of infantile hemangioma endothelial cells (IHECs) in vitro and determine the functions of the β-adrenergic receptor in the pathogenesis of infantile hemangioma. IHECs were divided into three groups: The control group, the propranolol group (PG) and the isoproterenol group (IG). The PG and IG were administered with high, medium and low concentrations of the corresponding drugs. The cell growth in each group was determined using the MTT assay. A high propranolol concentration resulted in the inhibition of cell growth. By comparison, isoproterenol promoted cell growth. Within a specific time-frame (72-96 h), high drug concentrations (20 µg/ml) elicited strong effects on the cells. At certain concentrations, propranolol inhibited cell growth once the proliferation stage of IHECs had been affected for a specific length of time, whereas isoproterenol yielded opposite results. The β-adrenergic receptor elicits an important effect in the pathogenesis of infantile hemangioma.

Introduction

At present, the nomenclature of skin vascular tissue tumors remains inconsistent, particularly in vascular tissue tumors. Different classifications have also been used unsatisfactorily. In 1996, the International Society of Vascular Anomalies divided the vascular birthmarks into two categories: Vascular tumors and vascular malformations; infantile hemangioma, for example, is classified as a vascular tumor (1,2). Infantile hemangioma is one of the most common benign tumors in infants and young children; these tumors are superficially located in the head and neck area (3). A few cases have also been observed in the mucosa, muscle, bone tissue and internal organs, which are affected in terms of appearance and function. In extreme cases, these superficial tumors threaten the life of the patient. Different theories, including the heredity and gene mutation theory, the placental chorionic cell ectopia theory, the endothelial progenitor cell or stem cell theory and the angiogenesis imbalance theory (4-14), have been proposed regarding the pathogenesis of hemangioma; however, no decisive conclusion has been obtained to date. Propranolol is a nonselective β-adrenergic receptor agonist. In 2008, a study proposed that propranolol elicits significant adverse effects on hemangioma (15); thus, propranolol has been increasingly used as the first-line treatment against hemangioma. Therefore, the functions of the β-adrenergic receptor in the development and progression of vascular tumors have attracted increased attention. Isoproterenol is a β-adrenergic receptor antagonist. A previous study demonstrated that isoproterenol promoted angiogenesis in vitro, indicating that the β-adrenergic receptor is important in the occurrence and development of vascular tumors (16).

In the present study, specific concentrations of propranolol and isoproterenol were used to compare the effects on infantile hemangioma endothelial cells (IHECs) in an in vitro cultivation environment of IHECs. This experiment was conducted to further investigate the functions of the β-adrenergic receptor in the development and progression of vascular tumors.

Subjects and methods

Subjects. A hemangioma specimen was resected clinically from a proliferating hemangioma on the forehead of a nine-month-old female patient. This study was conducted in accordance with the Declaration of Helsinki and with approval from the Ethics Committee of The First Affiliated Hospital of Xinjiang Medical University (Urumqi, China). Written informed consent was obtained from the guardian of the infant.

Primary cultivation of IHECs. The resected hemangioma specimen was immediately placed and stored in RPMI-1640 serum-free medium and promptly transferred to a laboratory laminar flow cabinet. Following the removal of the
supernatant, the specimen was placed in a sterile Petri dish and rinsed with double-antibody phosphate-buffered saline (PBS; HyClone Laboratories, Inc., South Logan, UT, USA) twice. The dish was replaced and washed three times with PBS. Specimen trimming was performed in the Petri dish to remove the epidermis and the connective tissues, prior to the specimen being cut into 1x1 mm sections and washed twice with PBS. A 0.25% trypsin (HyClone Laboratories, Inc.) solution was used to digest the cells for 3-4 h at 37°C with constant agitation. The RPMI-1640 medium, containing 20% fetal bovine serum (Gibco®-BRL, Grand Island, NY, USA), was then added to terminate digestion. The mixture was filtered and centrifuged (179 x g for 5 min) and the supernatant was subsequently discarded. Another portion of the RPMI-1640 medium was used to resuspend the precipitate. The supernatant was centrifuged (179 x g, 5 min) and discarded. Endothelial cell growth medium 2 (EGM-2), containing vascular endothelial growth factor (VEGF), hydrocortisone, ascorbic acid, human alkaline fibroblast growth factor B, human insulin-like growth factor and epidermal growth factor (Lonza Ltd., Basel, Switzerland), was added to the mixture, which was then transferred into a 25-cm² flask and placed in an incubator with CO₂ at 37°C. After 24 h, the cells adhered to the flask walls. The medium was changed at intervals of 2-3 days. Generation passage was performed when the cells covered 70-80% of the flask bottom. All of the procedures were completed under sterile conditions in the laminar flow cabinet.

Identification of IHECs. The primary cells were obtained and digested with 0.25% trypsin solution. The digested cells were then centrifuged (179 x g, 5 min) and EGM-2 was used to resuspend the precipitate. The resulting suspension was transferred to the slide of a 10-cm dish and developed at 37°C for 37°C with constant agitation. The RPMI-1640 medium, containing 20% fetal bovine serum (Gibco®-BRL, Grand Island, NY, USA), was then added to terminate digestion. The mixture was filtered and centrifuged (179 x g for 5 min) and the supernatant was subsequently discarded. Another portion of the RPMI-1640 medium was used to resuspend the precipitate. The supernatant was centrifuged (179 x g, 5 min) and discarded. Endothelial cell growth medium 2 (EGM-2), containing vascular endothelial growth factor (VEGF), hydrocortisone, ascorbic acid, human alkaline fibroblast growth factor B, human insulin-like growth factor and epidermal growth factor (Lonza Ltd., Basel, Switzerland), was added to the mixture, which was then transferred into a 25-cm² flask and placed in an incubator with CO₂ at 37°C. After 24 h, the cells adhered to the flask walls. The medium was changed at intervals of 2-3 days. Generation passage was performed when the cells covered 70-80% of the flask bottom. All of the procedures were completed under sterile conditions in the laminar flow cabinet.

In the blank control group, PBS was used as the secondary antibody. In the blank control group, PBS was used as the primary antibody.

Determination of cell growth curves using the MTT assay. The second-generation cells were spread evenly on a 96-well plate at a concentration 2×10⁴ cells per well. EGM-2 (~200 µl) was added and the cells were incubated for 24 h. The medium was subsequently changed and eight cells from each row of wells were observed daily. Approximately 20 µl MTT solution (5 mg/ml; Sigma, St. Louis, MO, USA) was added daily into each well and cultivation was continued for 4 h, prior to the supernatant being discarded. Approximately 150 µl dimethyl sulfoxide (DMSO; Sigma) was then added, and the system was agitated for 10 min. A microplate reader (Thermo Fisher Scientific, Inc., Rockford, IL, USA) was used to determine the absorbance of each well at a wavelength of 490 nm for 10 consecutive days. In the cell growth curve, time was set as the abscissa and the average absorbance was set as the ordinate.

Effect of propranolol on cell growth curves. Propranolol (Tianjin Lisheng Pharmaceutical Co., Ltd., Tianjin, China) was dissolved in DMSO and then diluted with EGM-2 to prepare working solutions at three concentrations: 10, 15 and 20 µg/ml. The final DMSO concentration was 0.16%. Following the preparation, the solutions were filtered with a 0.22-µm filter and sub-packaged.

The second-generation cells were obtained for digestion, centrifugation and counting. These cells were evenly spread on a 96-well plate at a cell concentration of 2×10⁴ cells per well; 200 µl EGM-2 was added and the cells were incubated for 24 h. Following adhesion of the cells to the walls, the medium was replaced with the 10, 15 and 20 µg/ml working solutions, respectively. Blank EGM-2 (without propranolol) and 0.16% DMSO-containing EGM-2 were used as the controls, with each treatment replicated four times. The cells were cultured in a CO₂ incubator, and the absorbance was determined using the MTT assay at 24, 48, 72 and 96 h; simultaneously, the morphological changes were observed and recorded. The non-parametric test of the multiple-sample-related measurements was performed to compare the difference in absorbance between each concentration and the control group. The average absorbance curve of the cells at each concentration was plotted, with time as the abscissa and average absorbance as the ordinate.

Effects of isoproterenol on cell growth curves. The effects of isoproterenol were observed following the procedures used to determine the effects of propranolol; however, isoproterenol was used instead of propranolol.

Statistical analysis. The non-parametric test of the multiple-sample-related measurements used in the data analysis was performed using SPSS 17.0 (SPSS, Inc., Chicago, IL, USA).

Results

Cultivation of IHECs. After the 24-h primary cultivation of IHECs, the cells gradually adhered to the walls, appearing...
round to polygonal in shape. Between days 2 and 3, the cell number gradually increased, and between days 4 and 5 the cells, half of which appeared polygonal, gradually fused. Between days 8 and 9, the cells gradually became fusiform and funicular, and blood vessel-like structures were visible in certain regions. Generation passage was performed when the cells occupied 80% of the bottom of the container (Fig. 1).

**Identification of IHECs.** Upon conducting immunohistochemistry, it was observed that the cells were polygonal and spindle-shaped with brownish-yellow stained cytoplasm; the nucleus remained unstained, indicating a positive result. These results were consistent with those in previous studies (17,18), in which vwf was present in the cytoplasm; therefore, it was confirmed that these cells were IHECs (Fig. 2A). A similar staining method was used to determine the presence of VEGFR-2 in the cytoplasm, yielding the same positive results as the previous experiment and confirming that the cells were IHECs (Fig. 2B).

**Determination of the growth curve of the IHECs.** The MTT assay was used to determine the IHEC growth curves. The results indicated a slow increase in the average absorbance of the cells on days 1 and 2. This parameter rapidly increased between days 3 and 5 but slightly decreased at day 6. On days 7 and 8, absorbance increased, prior to decreasing and then reached a plateau. Absorbance decreased gradually between days 9 and 10 (Fig. 3). This result was consistent with those from other studies (17,18).

**Effect of propranolol on IHECs.** In the *in vitro* culture environment, the absorbance of IHECs was measured using the MTT assay following cultivation in 10, 15 and 20 µg/ml propanol working solution, respectively, for 24, 48, 72 and 96 h. SPSS 17.0 (SPSS, Inc.) software was used for the statistical analysis of the absorbance at each concentration. The results showed no significant difference in the average absorbance of the IHECs among the blank, the DMSO and the three propranolol concentration groups in the 24- to 48-h time-frame (P>0.05). However, a difference in absorbance was observed in the 72- to 96-h time-frame. The inter-group comparison revealed that while no significant differences were observed among the control, the DMSO and the 10 and 15 µg/ml propranolol groups, a significant difference was identified between the 20 µg/ml group and the blank group (P<0.05). A change was also observed in the cell morphology: The IHECs in each well initially proliferated and then adhered to the walls 24 h before the medium was replaced (Fig. 4A). Comparing this result with the conditions in the well after 96 h and the replaced medium, it was found that the morphological characteristics of IHECs had changed (Fig. 4B). In particular, the IHECs appeared round or almost round, the cellular space increased and the cell number was low. The stacked line chart of IHEC absorbance under different concentrations of propranolol is shown in Fig. 4C.

**Effect of isoproterenol on IHECs.** In the *in vitro* culture environment, the average absorbance of the IHECs was
measured using the MTT assay following cultivation in 5, 10 and 20 µg/ml isoproterenol working fluid, respectively, for 24, 48, 72 and 96 h. Isoproterenol-free medium was used as the blank control group. No significant differences were found in the average absorbance of the IHECs among the groups in the 24-to 72-h time-frame (P>0.05); however, an absorbance difference was observed at 96 h among the blank group and the isoproterenol-containing groups. The inter-group comparison revealed that a high isoproterenol concentration corresponded to a high absorbance (P<0.05). The stacked line chart of IHEC absorbance under different concentrations of isoproterenol is shown in Fig. 5.

Discussion

Vascular tumors, including infantile hemangioma, delayed pyogenic granuloma and other rare vascular tumors occurring during infancy and early childhood, are characterized by a proliferation of endothelial cells, while vascular malformations are abnormalities of vascular morphogenesis. Vascular tumors can readily occur in the neonatal period; however, unlike infantile hemangioma, vascular malformations do not grow rapidly nor regress spontaneously in the first year following birth (19).

Infantile hemangioma can be divided into the proliferation period (three to 12 months old), the regression period (one to three years old) and the regression completion period (three to seven years old) (20). Although infantile hemangioma in certain patients undergoes self-regression, hemangioma in the proliferation period reproduces rapidly. This condition can affect the function of important surface organs and internal vital organs and can potentially be life-threatening (19).

The present study targeted infantile hemangioma in the proliferation period (nine months old), and a surgically resected specimen was obtained for the primary cultivation of the IHECs in EGM-2. A previous study on the pathogenesis of hemangioma indicated that growth factors, including VEGFs, fibroblast growth factors and human insulin-like growth factors, have an important function in the formation of vascular tumors (21). The medium used in this study contained various growth factors and selectively promoted the growth of IHECs. In the primary cultivation of IHECs at the proliferation period, normal medium is not able to satisfy the requirements of cell growth, and the medium containing specific cytokines is more suitable. The growth curve of the IHECs shows that the cells grew slowly between days 1 and 2, and that growth increased rapidly between days 3 and 5, decreased slightly at day 6 and increased again between days 7 and 8. A plateau was ultimately reached and growth decreased between days 9 and 10.

Since the discovery of propranolol for the treatment of hemangioma, propranolol has been widely used on affected children and achieved favorable results (22,23). Although the therapeutic mechanism remains unclear, the involvement of the β-adrenergic receptor has been considered. In the present study, two drugs, mutually antagonistic towards each other, were used to investigate the functions of the β-adrenergic receptor in the development of vascular tumors.

Following the intervention with different propranolol-concentration mediums, the results revealed that the low-concentration groups (10 and 15 µg/ml) had no significant adverse effects on the IHECs. In the high-concentration group (20 µg/ml), absorbance decreased between the 72- and 96-h time-points compared with the other groups, indicating that cell growth was inhibited.
observations also showed that cell growth was significantly limited following propranolol action. The growth curves showed that the average absorbance in the high-concentration group was decreased compared with that in the blank control group, indicating that cell growth was suppressed by propranolol. The degree of suppression corresponded to the drug concentration.

Isoproterenol is a non-selective β-adrenergic receptor agonist that exhibits a mutually antagonistic effect with propranolol. The present study identified that, following culture for 24-72 h with a certain concentration of isoproterenol-containing medium, negligible effects were observed in the IHECs of each group. In addition, no significant differences were observed in the absorbance among the groups. After 96 h, a high concentration of isoproterenol corresponded to a high absorbance in each group, indicating that isoproterenol elicited a stimulating effect on the IHECs, with an activity degree that was proportional to the concentration.

The present study revealed that propranolol elicits a positive therapeutic effect on infantile hemangioma; however, the mechanism remains unclear. Considering different theories, including the cytokine, signal transduction and receptor theories, the mechanism involved in such effects has yet to be determined. This study used two mutually antagonistic drugs to act on the β-adrenergic receptor. The results showed the contrasting effects of these drugs on IHECs. The results further indicated that the β-adrenergic receptor plays a role in infantile hemangiomas and that the therapeutic effect of propranolol was largely induced via the β-adrenergic receptor. However, further investigation is required to determine the exact mechanism.

References

1. Enjolras O and Mulliken JB: Vascular tumors and vascular malformation (new issues). Adv Dermatol 13: 375-423, 1997.
2. Mulliken J and Enjolras O: Congenital hemangiomas and infantile hemangioma: missing links. J Am Acad Dermatol 50: 875-882, 2004.
3. Cahill AM and Nijs EL: Pediatric vascular malformations: pathophysiology, diagnosis, and the role of interventional radiology. Cardiovasc Intervent Radiol 34: 691-704, 2011.
4. Ittineang T, Tan ST, Guthrie S, Tan CE, Mchtyre BC, Brasch HD and Day DJ: A placental chorionic villous mesenchymal core cellular origin for infantile haemangiomat. J Clin Pathol 64: 870-874, 2011.
5. Régnier S, Dupin N, Le Danff C, Wassef M, Enjolras O and Araarini S: Endothelial cells in infantile haemangiomas originate from the child and not from the mother (a fluorescence in situ hybridization-based study). Br J Dermatol 157: 158-160, 2007.
6. Zhang GY, Yi CG, Li X, Liang ZQ, Wang RX, Liu DE, Zhang LM, Meng CY and Guo SZ: Proliferation hemangiomas formation through dual mechanism of vascular endothelial growth factor mediated endothelial progenitor cells proliferation and mobilization through matrix metalloproteases 9. Med Hypotheses 70: 815-818, 2008.
7. Takahashi K, Mulliken JB, Kozakewich HP, Rogers RA, Folkman J and Ezekowitz RA: Cellular markers that distinguish the phases of hemangioma during infancy and childhood. J Clin Invest 93: 2357-2364, 1994.
8. Peng Q, Matsuda T and Hirst SJ: Signaling pathways regulating interleukin-13-stimulated chemokine release from airway smooth muscle. Am J Respir Crit Care Med 169: 596-603, 2004.
9. Queto T, Vasconcelos ZF, Luz RA et al: G-CSF suppresses allergic pulmonary inflammation, downmodulating cytokine, chemokine and eosinophil production. Life Sci 88: 830-838, 2011.
10. Gordillo GM, Onat D, Stockinger M, Roy S, Atalay M, Beck FM and Sen CK: A key angiogenic role of monocyte chemoattractant protein-1 in hemangioendothelioma proliferation. Am J Physiol Cell Physiol 287: C866-C873, 2004.
11. Eum SY, Maghni K, Hamid Q, Eidelberg DH, Campbell H, Isogai S and Martin JG: Inhibition of allergic airways inflammation and airway hyperresponsiveness in mice by dexamethasone: role of eosinphils, IL-5, eotaxin, and IL-13. J Allergy Clin Immunol 111: 1049-1060, 2003.
12. North PE, Waner M, Mizeracki A and Mihm MC Jr: GLUT-1: a newly discovered immunohistochemical marker for juvenile hemangiomas. Hem Pathol 31: 99-200, 2000.
13. Bauland CG, Smit JM, Bartelink LR, Zondervan HA and Spauw PH: Hemangioma in the newborn: increased incidence after chorionic villus sampling. Prenat Diagn 30: 913-917, 2010.
14. Boye E, Yu Y, Paranya G, Mulliken JB, Olsen BR and Bischoff J: Clonality and altered behavior of endothelial cells from hemangiomas. J Clin Invest 107: 745-752, 2001.
15. Léauté-Labrèze C, Dumas de la Roque E, Hubiche T, Boralevi F, Thambo JB and Tateb A: Propranolol for severe hemangiomas of infancy, N Engl J Med 358: 2649-2651, 2008.
16. Ji Y, Chen S, Li K, Xiao X, Zheng S and Xu T: The role of β-adrenergic receptor signaling in the proliferation of hemangioma-derived endothelial cells. Cell Div 8: 1, 2013.
17. Ma G, Lin XX, Jin YB, et al: Isolation, culture and identification of endothelial cells in infantile hemangioma. Zhonghua Zheng Zhi Ke Za Zhi 24: 144-147, 2008 (In Chinese).
18. Ma G, Lin XX, Jin YB, et al: Culture and cryopreservation of endothelial cells in vitro in infantile hemangioma. Zhonghua Zheng Xing Wai Ke Za Zhi 24: 197-200, 2008 (In Chinese).
19. Bologna JL, Jorizzo JL and Rapini RP: Dermatology, 2nd Edition. Saunders (W.B) Co. Ltd, Philadelphia, PA, USA, pp1905-1937, 2012.
20. Léauté-Labrèze C, Prey S and Ezzedine K: Infantile haemangioma: part I. Pathophysiology, epidemiology, clinical features, life cycle and associated structural abnormalities. J Eur Acad Dermatol Venereol 25: 1245-1253, 2011.
21. Takahashi K, Mulliken JB, Kozakewich HP, Rogers RA, Folkman J and Ezekowitz RA: Cellular markers that distinguish the phases of hemangioma during infancy and childhood. J Clin Invest 93: 2357-2364, 1994.
22. Sans V, de la Roque ED, Berge J, Grenier N, et al: Propranolol for severe infantile hemangiomas: follow-up report. Pediatrics 124: e423-e431, 2009.
23. Sommers Smith SK and Smith DM: Beta blockade induces apoptosis in cultured capillary endothelial cells. In Vitro Cell Dev Biol Anim 38: 298-304, 2002.