Fabrication of a triptolide-loaded and poly-γ-glutamic acid-based amphiphilic nanoparticle for the treatment of rheumatoid arthritis

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Abstract: Triptolide (TP) exhibits immunosuppressive, cartilage-protective and anti-inflammatory effects in rheumatoid arthritis. However, the toxicity of TP limits its widespread use. To decrease the toxic effects, we developed a novel nano-drug carrier system containing TP using poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride (PAT). PAT had an average diameter of 79±18 nm, a narrow polydispersity index (0.18), a strong zeta potential (−32 mV) and a high drug encapsulation efficiency (EEγ=48.6%) and loading capacity (EEγ=19.2%), and exhibited controlled release (t½=29 h). The MTT assay and flow cytometry results indicated that PAT could decrease toxicity and apoptosis induced by free TP on RAW264.7 cells. PAT decreased lipopolysaccharides/interferon-γ-induced cytokines expression of macrophage (P<0.05). In vivo, PAT accumulated at inflammatory joints, improved the survival rate and had fewer side effects on tumor necrosis factor α transgenic mice, compared to TP. The blood biochemical indexes revealed that PAT did not cause much damage to the kidney (urea nitrogen and creatinine) and liver (alanine aminotransferase and aspartate aminotransferase). In addition, PAT reduced inflammatory synovial tissue area (P<0.05), cartilage loss (P<0.05), tartrate-resistant acid phosphatase-positive osteoclast area (P<0.05) and bone erosion (P<0.05) in both knee and ankle joints, and showed similar beneficial effect as free TP. In summary, our newly formed nanoparticle, PAT, can reduce the toxicity and guarantee the efficacy of TP, which represents an effective drug candidate for RA with low adverse side effect.

Keywords: triptolide, rheumatoid arthritis, γ-PGA, tumor necrosis factor α transgenic mice, drug carrier system.

Introduction

Rheumatoid arthritis (RA) is an autoimmune disease that causes chronic inflammatory polyarthritis, which leads to pain and swelling, erosions of the cartilage and bone and even joint deformity. RA decreases motor function, leads to disability and diminishes quality of life.¹,² Currently, nonbiologic disease-modifying antirheumatic drugs (DMARDs) are the first-line treatment for RA. The patients will receive biologic DMARDs, such as tumor necrosis factor (TNF) inhibitors, if they did not respond to the nonbiologic DMARDs.³-⁹ Current treatments can improve symptoms and slow the progress of the disease, but only partial RA patients respond to DMARDs, and some biologics are contraindicated for others.¹⁰ Thus, there is an obvious need to develop novel effective drugs for RA due to the insufficient response, adverse effects and high cost of the present medicine treatments.¹¹

Tripterygium wilfordii Hook F. (TWHF) is clinically used as a potential source of new drugs for RA. Several clinical trials proved that the extracts of TWHF achieve better effectiveness than DMARDs monotherapy.¹²,¹³ Triptolide (TP) is an active
compound of TWHF, which exhibits immunosuppressive, cartilage-protective and anti-inflammatory effects on both humans and animals with RA. Unfortunately, the use of TP increases the risk for the occurrence of severe side effects, including nephrotoxicity, hepatic injury and bone marrow dysfunction. Development of self-assembled polymeric materials is a promising way to carry out drug delivery to achieve better therapeutic effect and minimize the side effects. These polymeric materials are prepared from a range of inorganic and organic compounds including polymers, lipids, dendrimers, carbon nanotubes and quantum dots. Among the polymers, poly-γ-glutamic acid (PGA) is stable with highly hydrophilic monoglutamate chain and can be synthesized cheaply by fermentation. Another advantage of this material is that it is nontoxic and fully biodegradable, and has been used clinically. PGA has been used as a carrier of various drugs and biomolecules, such as proteins, vaccine and genes, to treat liver tumor, glioma, bladder and lung cancer cells. However, whether PGA could be used as a carrier to treat RA was not known.

In this study, we investigated the use of PGA to wrap TP for RA to reduce the toxicity and guarantee the curative effect of TP. Previously, we used a PGA and L-phenylalanine ethylester complex to wrap TP, and found that the nanoparticle loaded with TP decreased TP toxicity in wild-type (WT) mice. In the current study, we conjugated PGA with di-tert-butyl L-aspartate hydrochloride (Asp), a hydrophobic structure that can allow the hydrophilic PGA-wrapped hydrophobic TP, to develop a new material, poly-γ-glutamic acid-grafted Asp (PA), which was utilized as a drug carrier to form a novel TP-loaded poly-γ-glutamic acid nanoparticle (PAT).

Materials and methods

Drugs and reagents

TP was purchased from Yuanye Biotech Company (Shanghai, People’s Republic of China). γ-PGA (molecular weight 300 kD) was purchased from Haining Zijingang Biotechnology Co. Ltd. Asp, penicillin–streptomycin sulfate, dimethyl sulfoxide (DMSO) and indocyanine green (ICG) were purchased from Sigma-Aldrich Co. (St Louis, MO, USA). 1-Ethyl-3-(3-dimethylaminopropyl) carbodiimide hydrochloride (EDCI) was purchased from Shanghai Medpep Company (Shanghai, People’s Republic of China). MTT and trypsin were from AMRESCO (Solon, OH, USA). 4’,6-Diamidino-2-phenylindole was purchased from Roche (Basel, Switzerland), DMEM, a cell culture medium, and fetal calf serum (FBS) were purchased from Hyclone (Logan, UT, USA). Apoptosis detection kit Annexin V-fluorescein isothiocyanate (FITC) and propidium iodide (PI) were purchased from BD (Franklin Lakes, NJ, USA).

Cells and animals

RAW264.7 cells were obtained from the cell bank of the Shanghai Institute of Biochemistry and Cell Research and were cultured in DMEM with 10% FBS and 1% penicillin–streptomycin solution at 37°C with 5% CO₂. Tumor necrosis factor α transgenic (TNFα-Tg) mice in C57BL/6 background were generated by Dr George Kollias, which were bred as heterozygotes, and the WT littermates were used as control. The TNFα-Tg mice displayed mild bone erosion and joint inflammation from 2 to 2.5 months of age. Mice (2 months old) were reproduced in the East China Normal University animal center and were housed in specific pathogen-free room. All the mice studies were performed as per the Guiding Principles for the Care and Use of Laboratory Animals according to the Regulations of the People’s Republic of China for Administration of Laboratory Animals. This study was approved by the East China Normal University Ethics Committee on Animal Resources.

Preparation of PAT

The amphiphilic copolymers consisted of hydrophilic γ-PGA backbone and hydrophobic side chain Asp. The copolymer was synthesized by coupling Asp with γ-PGA. Briefly, Asp (10 mg) was conjugated to γ-PGA (38.4 mg) through amidation reaction under the catalyst EDCI (26 mg) for 24 h to form PA at room temperature. The unconjugated Asp and catalyst EDCI were removed by ether extraction. PAT complex was obtained by adding 5 mg TP to 20 mg PA in DMSO solution, and then the solution was dropped into a baker with ultrapure water under magnetic stirring at a speed of 120 rpm. After centrifugation (12,000 rpm) for 10 min, the free TP was removed, and dry PAT nanoparticle was obtained by vacuum freeze-drying equipment (Yibokang, Beijing, People’s Republic of China). Biopolymer dispersion samples (4 mg/mL) were stored at 4°C in ultrapure water.

Characterization of PAT

The chemical structure of PAT was identified by ¹H nuclear magnetic resonance spectroscopy (NMR) spectroscopy (Bruker Avance 500 Spectrometer; Billerica, MA, USA). Dynamic light scattering (DLS; Malvern Zetasizer Nano-ZS; Malvern, UK) was used to detect the mean particle size and zeta potential of PAT at 25°C in triplicates. The particle size distribution was detected by the mean diameter and
polydispersity index (PDI). Transmission electron microscopy (TEM; JEM-2100; JEOL) was performed to study the morphological appearance of PAT.

**Drug encapsulation efficiency**

Agilent 1100 high-performance liquid chromatography (HPLC) system with a reverse-phase LiChrospher ODS C18 column was used to detect the TP encapsulation ratio. PAT (5 mg) was added in 1 mL DMSO to release TP under an oscillator for 5 min, and the solution was removed by freeze dryer (Yibokang), and then 2 mL methanol was added. The mobile phase was a mixture of acetonitrile-water (30/70, v/v), and the current speed was controlled at 1.0 mL/min. The column temperature was controlled at 30°C, and the detection wavelength was maintained at 204 nm by a variable wavelength detector. Drug encapsulation efficiency (EE) was determined using the following equation: 

\[
EE = \frac{M_1}{M_2}
\]

where \( M_1 \) represents the mass of TP in the PAT and \( M_2 \) represents the mass of TP used in the equation. The drug loading capacity (EE) was determined using the following equation: 

\[
EE = \frac{W_1}{W_2}
\]

where \( W_1 \) represents the weight of TP in the PAT and \( W_2 \) represents the total weight of PAT.

**In vivo TP release profiles**

The dialysis diffusion technique was used to study the TP release profiles. PAT (10 mg) was deposited in 2 mL phosphate buffer saline (PBS) and suspended in a dialysis bag (cut-off, 10 kD; Spectrum Laboratories Inc, Rancho Dominguez, CA, USA). Then, the dialysis bag was immersed into 50 mL PBS or saline with continuous magnetic shaking at a speed of 60 rpm at 37°C. At predetermined time point, 0.5 mL external dialysis solution was extracted for HPLC analysis, and same volume of PBS or saline was added simultaneously. A profile was plotted to show the accumulative release of TP.

**In vitro cytotoxicity**

RAW264.7 cells were grown in 96-well plates (4×10³/well). TP or PAT was mixed in culture medium at the equivalent TP concentrations of 6.25, 12.5, 25, 50, 100, and 200 nM/L and cultured for another 24, 48 and 72 h. MTT was used to quantify the cell survival rate. Briefly, each well was added with 10 μL MTT (5 mg/mL) solution and incubated for 2 h. DMSO (200 μL) was added into each well after the removal of supernatant. To evaluate the cell viability, a microplate reader (Multiskan GO; Thermo Fisher Scientific, Waltham, MA, USA) was used to detect the optical density values of each well under the wavelength of 570 nm.

**Flow cytometric analysis**

RAW264.7 cells (5×10⁵) were seeded in a 6 cm dish and treated with 100 nM TP or PAT (containing 100 nM TP) for 24 h. Then, double-staining using Annexin V-FITC/PI kit (BD) was performed to quantify apoptosis rate. Briefly, after incubation, cells were centrifuged (1,000 rpm) for 5 min and washed with PBS 3 times, and then incubated in 500 μL flow cytometric buffer added with 5 μL of PI and 5 μL of Annexin V-FITC for 30 min at 4°C. The apoptosis rate of the different groups was detected by BD FACSCalibur and analyzed with BD CellQuest Pro software.

**Real-time polymerase chain reaction (PCR)**

The expression of *TNFα, IL-1β, IL-6, iNOS* and *β-actin* (the primer sequences are listed in Table 1) genes in RAW246.7 cells was detected by quantitative real-time PCR (Q-PCR). Total RNA was isolated with the TRIzol reagent (Thermo Fisher Scientific) according to the product instruction. A NanoDrop instrument (Thermo Fisher Scientific) was used to quantify mRNA concentration from the cells. Total RNA (1 μg) from each sample was reverse transcribed into cDNA using the SYBR Green

| Genes | Sequences of primers | GenBank accession number | Product size (bp) |
|-------|-----------------|-----------------|-----------------|
| *IL-1β* | F: 5′ TGCCACCCCTTCTGACAGTAGT 3′ | NM_008361.4 | 138 |
| R: 5′ TGATTGCTGCTGGGAGATT 3′ |  |
| *iNOS* | F: 5′ GAGACAATGACTGGTGT 3′ | NM_00131922.1 | 178 |
| R: 5′ CGATGGGAGACAAAGGCG 3′ |  |
| *TNFα* | F: 5′ GATCGGGTCCTCAAGGGGATG 3′ | NM_001287601.1 | 92 |
| R: 5′ CCATTGTGTTTTGTGAGTG 3′ |  |
| *IL-6* | F: 5′ CCTCAATTTCACATGCTTCC 3′ | NM_031168.2 | 141 |
| R: 5′ CGGACTAGTTGTTGCGAGTA 3′ |  |
| *β-actin* | F: 5′ TGAGCTGCGTTTTTACACCTT 3′ | NM_007393.5 | 198 |
| R: 5′ GCCCTACCGGTTCCAGGTTT 3′ |  |

**Abbreviations:** PCR, polymerase chain reaction; F, forward; R, reverse.

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**Table 1 Sequences of primers used in real-time PCR**
I Kit (Takara, Kusatsu, Japan) according to the manual. The cDNA templates were diluted 5 times, and Q-PCR was conducted with SYBR Premix Ex Taq II (Takara) according to the instruction in a Bio-Rad CFX 96 (Hercules, CA, USA) real-time PCR system. Each experiment was performed in triplicates. The samples were heated at 94°C for 5 min, followed by 40 PCR cycles (94°C for 15 s; 56°C for 15 s; 72°C for 30 s). The gene of β-actin was used as internal control, and fold changes were counted with the 2^−ΔΔCt method.

**In vivo toxicity**

In vivo toxicity of PAT was studied in WT and TNFα-Tg mice. Two-month-old TNFα-Tg mice were divided into 4 groups (n=10/group), and treated intravenously in tail with free TP (0.15 mg/kg), PAT (0.15 mg/kg TP), PA (100 μL), or PBS (100 μL), every 3 days for 30 days. WT mice (n=10) were treated intravenously in tail with 100 μL PBS. A continuous observation for 30 days was conducted to assess the survival rate after the first injection. As an index of systemic toxicity, body weight was measured every 5 days. On the last day, mice were sacrificed, and serum samples were collected. Aspartate aminotransferase (AST), creatinine (CRE) and blood urea nitrogen (BUN) were detected using an auto-analyzer (BM704; Hitachi) to evaluate hepatic function (ALT and AST) and renal function (CRE and BUN).

**In vivo imaging of nanoparticles**

ICG (1 mg) was incubated with PA (20 mg) or with 5 mg TP in DMSO solution at room temperature for 30 min. Then, the mix was dropped into ultrapure water under magnetic stirring at a speed of 120 rpm. The ICG-modified nanoparticles (PA-ICG and PAT-ICG) were harvested after centrifugation at 5,000 rpm for 10 min at 4°C, followed by lyophilization. One hundred microliters of PA-ICG (10 mg/mL) and 100 μL of PAT-ICG (10 mg/mL) in PBS (pH 7.4) were injected into mice by intravenous tail injection. The body images (from 0.5 to 80 h) were acquired after injection using the IVIS Spectrum in vivo imaging system (Caliper, Hopkinton, MA, USA).

**Statistical analysis**

GraphPad Prism 5 software was used for statistical analyses. One-way analysis of variance test followed by a Bonferroni-corrected posttest was used for comparisons. P<0.05 was considered statistically significant.

**Results**

**Characterization of PAT**

The 5 characteristic peaks of PA in 1H NMR spectrum are shown in Figure 1: 1 was the symbolic peak of aspartic acid (d 1.44 ppm), and the remaining 4 were symbolic peaks of PGA (d 1.88, d 2.01, d 2.33 and d 4.27 ppm). The Asp loading rate for PGA could be calculated by comparing the intensities of the peaks with d 1.44 and d 4.27 ppm. A 25% Asp loading for PA (0.82 Asp per repeating units) (Figure 1A) was identified from the 1H NMR results.

The mean particle dimension of PAT was 79±18 nm with a narrow size distribution (PDI =0.18; Figure 1B) as detected...
by DLS. Nanoparticles were round in shape with a diameter of 89±23 nm (Figure 1C) as detected by TEM. The particle size was in the range suitable for enhanced permeability and retention (EPR) effect in inflammatory tissue.

Stronger zeta potentials will make the particles repel each other which can create a more stable dispersion system. A Stronger zeta potentials will make the particles repel each other which can create a more stable dispersion system.

In aqueous phase, the zeta potential border of stable particles is commonly considered as above +30 or below −30 mV. The zeta potential of the PAT developed in our study was −32 mV, which indicated that PAT was stable in aqueous phase due to strong mutual electrostatic repulsion.

In addition, the TP encapsulation rate EE$_1$ and loading capacity EE$_2$ were determined by HPLC to be 48.6% and 19.2%, respectively. The high EE$_1$ and EE$_2$ of PAT could
avoid the poor aqueous solubility and allow rapid parenteral infusion without the risk of drug precipitation.44

In vitro release profile
The slow-release formulation can hold the effective medicine concentration in the blood for a long time, which can reduce the usage frequency and the drug dose, and increase the patient compliance. In our research, the median maximum release time \( t_{1/2} \) of free TP was 1.5 h, but for PAT the \( t_{1/2} \) was about 29 h. There was 79% TP released, while only 36% TP was released by PAT at 6 h. At 120 h, 89% TP was released from PAT (Figure 1D). Compared with free TP, TP in PAT was released much more slowly. The release profile results indicated that the in vitro release of TP was burst-like, while TP release from PAT was sustainable.

PAT decreased TP-induced cytotoxic effect in the early stages and lipopolysaccharides/interferon \( \gamma \) (LPS/IFN\( \gamma \))-induced cytokines expression of macrophage
To determine the cytotoxic effect of PAT in vitro, we applied MTT assay to detect the cell survival rate and flow cytometry method to detect cell apoptosis rate of macrophage (RAW264.7). MTT assay demonstrated that TP significantly decreased the cell survival rate of macrophage in a dose-dependent manner at different time points. Compared with TP-treated macrophage, PAT, containing same TP concentration, remarkably maintained the cell viability of macrophage from 24 to 48 h. The half-inhibition concentration of PAT was much higher than that of free TP at 24 (236±39.32 vs 47±12.34 nM) and 48 h (123±11.34 vs 21±4.68 nM), which could be accounted for by the slow release of TP from PAT. But with the treatment duration increase to 72 h, PAT caused similar decrease of the cell viability of macrophage as TP (Figure 2A).

We treated macrophage with PBS, PA, TP and PAT for 24 h and found no obvious apoptosis in the PBS- or PAT-treated group. The apoptosis rate in free TP-treated group was 68.9%, while the apoptosis rate in PAT-treated group was 7.2% (Figure 2B). In addition, in PAT-treated group, there were 88% of live cells, which was higher than free TP-treated group (72%; Figure 2B). These results indicated that PAT treatment markedly improved the cell survival rate compared to free TP treatment. The result of Q-PCR indicated both TP and PAT dramatically decreased the mRNA level of TNF\( \alpha \) \( (P<0.05) \), interleukin-1\( \beta \) \( (P<0.05) \), interleukin-6 \( (P<0.05) \) and inducible nitric oxide synthase \( (P<0.05) \) induced by LPS/IFN\( \gamma \) in RAW264.7 cells (Figure 2C).

PAT targeted inflammatory joints of TNF\( \alpha \)-Tg mice
Within the near-infrared range, ICG could penetrate deep tissue and could be used for fluorescence visualization.45–47 A mixture of amphiphilic ICG48,49 and PA can self-assemble in water. To detect the retention duration of PAT in vivo and to determine the frequency of drug administration, we conjugated PA or PAT with ICG by self-assembling. PA-ICG and PAT-ICG were injected into the tail vein of 2-month-old TNF\( \alpha \)-Tg mice and WT littermates. The 2-month-old TNF\( \alpha \)-Tg mice injected with PBS and free TP were utilized as negative controls, because no bioluminescence could be detected. Bioluminescent images were taken intermittently at 0.5, 1, 4, 24, 48 and 80 h. The ICG signal spread into the internal organs immediately after injection at tail vein of both WT and TNF\( \alpha \)-Tg mice, accumulated at the knee joints of TNF\( \alpha \)-Tg mice at 24 h, reduced but remained positive at the knee joints of TNF\( \alpha \)-Tg mice at 48 h and was totally cleared in both WT and TNF\( \alpha \)-Tg mice at 80 h. In WT mice, PA-ICG did not assemble at knee joints at any time point (Figure 3A and B). These results suggested that 1) the retention duration of PAT and PA in WT and TNF\( \alpha \)-Tg mice was less than 80 h, and the frequency of drug administration should be once every 3 days, and 2) EPR effect made the nanoparticles, PA and PAT, accumulate in the inflammatory joints of TNF\( \alpha \)-Tg mice.

PAT decreased in vivo toxicity
To determine whether PAT can reduce the toxicity of TP on TNF\( \alpha \)-Tg mice, we applied PA, TP and PAT in 2-month-old TNF\( \alpha \)-Tg mice once in 3 days for 30 days, and found that the survival rate was 100% in mice treated with PBS, PA and PAT, whereas the survival rate was 70% in mice treated with 0.15 mg/kg TP (Figure 4A). Besides, 0.15 mg/kg TP brought about a 20% body weight loss in TNF\( \alpha \)-Tg mice; however, the PAT, containing same concentration of TP, caused no obvious body weight changes (Figure 4B). Only TP treatment group showed kidney (BUN and CRE) and liver (ALT and AST) damage, and no significant differences between PAT and PBS groups were detected (Table 2). These results revealed that PAT exhibited lower toxicity.

Furthermore, the HE staining was performed to detect the side effects of PAT on the liver, kidney and spleen in TNF\( \alpha \)-Tg mice, and found that the liver cells were intact in structure, and nucleus was arranged orderly without any degeneration or necrosis in WT and PA-treated TNF\( \alpha \)-Tg mice. In free TP-treated group, the livers were mottled, fragile and hyperemic, and the nuclei became ruptured and pyknotic.
Treatment of RA by TP-loaded and PGA-based nanoparticles

Figure 2 PAT reduces the cytotoxicity of TP on RAW264.7 cells. (A) Growth curves of RAW264.7 cells were determined by MTT assay after incubation with TP (200, 100, 50, 25, 12.5 or 6.25 nM) or PAT (equal to TP concentration) for 24, 48 and 72 h. The values are the mean ± SD of 8 wells. (B) The apoptosis was assessed by FACS. Annexin V+/PI− cells represent early apoptotic cells, while Annexin V+/PI+ cells represent late apoptotic cells. The percentage of both early and late apoptotic cells is shown in the lower right corner. (C) The mRNA expression of TNFα, IL-1β, IL-6 and iNOS was analyzed by real-time PCR. The values are the mean ± SD of 3 wells. *P<0.05, compared with TP; **P<0.05, compared with PBS treated groups.

Abbreviations: TP, triptolide; PAT, TP-loaded poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride; PBS, phosphate buffer saline; PA, poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride; RTTC, fluorescein isothiocyanate; TNFα, tumor necrosis factor α; IL-1β, interleukin 1 β; IL-6, interleukin 6; iNOS, inducible nitric oxide synthase; LPS/IFNγ, lipopolysaccharides/interferon γ; FACS, fluorescence-activated cell sorting; PI, propidium iodide; PCR, polymerase chain reaction.
However, the livers from PAT group exhibited normal structure, including clear cellular structures and orderly arranged cells. Similarly, free TP severely damaged the kidney, resulting in swelling, hyperemia, scattering and necrosis of renal glomerular cells and blurry borders of renal tubular epithelial cells and stenosis or atresia in lumens, while in PBS-, PA- and PAT-treated TNFα-Tg mice, the structures like renal glomerulus, interstitium and tubule were normal. Besides, in free TP-treated group, the spleen red pulp increased, whereas PAT treatment protected the spleen from the damaging effects of TP (Figure 4C). These results indicated that PAT protected the mice from the toxicity of TP.

PAT presented beneficial effect on both ankle and knee joints of TNFα-Tg mice

To determine if PAT had similar beneficial effect as TP on inflammatory arthritis, we injected PA, TP and PAT into tail veins of 2-month-old TNFα-Tg mice, because at this age, their ankle joints developed tenosynovitis. After 1-month treatment, we performed micro-CT scanning, and ABOG and TRAP staining on both ankle and knee joints of TNFα-Tg mice to investigate synovial inflammation, and bone and cartilage erosion at joints, which are the major characteristics of inflammatory arthritis.

Figure 3 PA and PAT target the inflammatory joints of TNFα-Tg mice. (A) Bioluminescent images of PA-ICG-treated WT mice, and PA-ICG- or PAT-ICG-treated TNFα-Tg mice were taken intermittently at 0.5, 1, 4, 24, 48 and 80 h after injection. (B) Fluorescence intensity of the knee joint at 24 h post-administration. The values are the mean ± SD of 6 legs from 3 mice. *P<0.05, compared with the WT + PA group.

Abbreviations: WT, wild type; PA, poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride; TNFα-Tg, tumor necrosis factor α transgenic; PAT, TP-loaded poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride; ICG, indocyanine green.
Figure 4: PAT protects mice from the toxicity of TP. (A) The survival rate of different groups show PAT increased the survival rate of TNFα-Tg mice, compared with TP treatment. (B) The body weight of different groups at the last day of treatment shows PAT significantly prevented the body weight loss of TNFα-Tg mice, compared with TP treatment. Values are the mean ± SD of 7–10 mice per group. (C) HE staining sections (magnification ×200) of the liver, spleen and kidney show PAT protected those organs from injury caused by TP. The yellow arrow indicates the hepatic nucleus, and the blue arrow indicates the renal glomerulus. *P<0.05, vs TP group.

Abbreviations: WT, wild type; TNFα-Tg, tumor necrosis factor α transgenic; PBS, phosphate buffer saline; PA, poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride; TP, triptolide; PAT, TP-loaded poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride; HE, hematoxylin–eosin.

Table 2: Biochemical parameters of blood serum

|                | ALT (mmol/L) | AST (mmol/L) | CRE (mmol/L) | BUN (mmol/L) |
|----------------|--------------|--------------|--------------|--------------|
| WT             | 83.77±7.73   | 33.07±15.12  | 178.22±14.15 | 7.18±0.42    |
| TNFα-Tg + PBS  | 69.91±20.97  | 31.64±12.52  | 165.83±23.76 | 7.54±0.72    |
| TNFα-Tg + PA   | 66.54±10.08  | 37.72±20.11  | 179.57±19.44 | 7.39±1.06    |
| TNFα-Tg + TP   | 170.87±15.32 | 160.17±14.68 | 343.70±49.90 | 13.32±2.17   |
| TNFα-Tg + PAT  | 81.58±14.12  | 38.58±18.38  | 175.36±35.76 | 7.84±1.30    |

Notes: *P<0.05, vs TNFα-Tg + PBS. *P<0.05, vs WT.

Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransferase; CRE, creatinine; BUN, blood urea nitrogen; WT, wild type; TNFα-Tg, tumor necrosis factor α transgenic; PBS, phosphate buffer saline; PA, poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride; TP, triptolide; PAT, TP-loaded poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride.
Micro-CT results indicated that mean values of bone volume of astragalus and patella significantly decreased in PBS- and PA-treated groups compared to WT mice (Figure 5A). In contrast, astragalus and patella bone volumes were significantly higher ($P < 0.05$) in TP- and PAT-treated mice than in PBS-treated TNFα-Tg mice. However, the difference between TP- and PAT-treated groups was not significant (Figure 5B). These results suggested that PAT had similar beneficial effect as TP on inflammatory arthritis.

Histological and histomorphometric analysis indicated that PA- and PBS-treated TNFα-Tg mice had conspicuous synovial inflammation, bone and cartilage destruction (Figure 6A and B) and numerous TRAP+ osteoclasts at ankle and knee joints (Figure 7A and B). All these symptoms were significantly less obvious in ankles and knee joints of PAT- and TP-injected mice.

**Discussion**

A variety of new strategies have been developed for the treatment of RA if the RA patients do not respond to the medications. Biologic agents and small-molecule drugs have been developed. Small potent molecules such as sulfasalazine, leflunomide, methotrexate, hydroxychloroquine, cyclosporine and azathioprine are widely used in the clinic but can cause unpleasant side effects. Some biologic agents including rituximab, abatacept, etanercept, infliximab, certolizumab, golimumab, adalimumab and tocilizumab are contraindicated in some patients and estimated to be cost-effective. Nano-drug is a good option to achieve cure and reduce toxicity and is a complementary therapeutic agent for RA.

In the current study, we firstly developed a novel nanodrug carrier system containing TP using γ-PGA-grafted L-Asp-OtBu, which can reduce the toxicity and guarantee the curative effect of TP for RA. Previously, PGA was conjugated with L-phenylalanine,53 leucine, tryptophan54 or carboxymethyl chitosan55 to form nanoparticles. The use of aspartic acid conjugated with PGA to form amphiphilic nanoparticles has not been reported. PA as a drug carrier was fabricated using mono-amino acid chain which helped to achieve a solid structure and reduced the leakage of TP. The TEM images indicated that the particle size of our newly formed PAT was in the range suitable for EPR effect.
in inflammatory tissue, and our experiments with PAT-ICG suggested that PAT was efficacious in targeting inflammatory joints. The zeta potential of PAT demonstrated that our nanoparticle was stable in the aqueous phase. The TP encapsulation ratio of PAT reached 48.6%, which exhibited the high encapsulation efficiency of PAT.

As a small molecule, TP could rapidly perform its therapeutic action, but it is hard to maintain its concentration in blood due to glomerular filtration and various forms of protein hydrolysis. TP release from PAT was in a sustainable manner. MTT studies suggested that PAT reduced the cytotoxicity of TP in the early stages and maintained its efficacy for a long time, which confirmed the slow-release effect of PAT. A slow-release drug formation could maintain a good drug concentration in blood and prolong the drug’s efficacy. These results suggest that PAT is stable, exhibits

Figure 6 Both TP and PAT reduce ankle and knee joint tissue inflammatory damage in TNFα-Tg mice. (A) Representative ABOG-stained sections (magnification ×40) show decreased joint tissue damage, including decreased synovial inflammation and cartilage erosion in TP- and PAT-treated mouse. (B) Quantitation of inflammatory synovial tissue area percentage of ankle and knee joints, and cartilage area percentage of ankle and knee joints. Values are the mean ± SD of 7–10 legs per group. *P<0.05, compared with PBS treated group.

Abbreviations: WT, wild type; TNFα-Tg, tumor necrosis factor α transgenic; PBS, phosphate buffer saline; PA, poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride; TP, triptolide; PAT, TP-loaded poly-γ-glutamic acid-grafted di-tert-butyl L-aspartate hydrochloride; ABOG, Alcian blue/orange G.
slow release and is suitable for EPR effect and targets inflammatory joints. Two anti-TNF monoclonal antibodies, etanercept and infliximab, have been approved by the US Food and Drug Administration and the European Medicines Evaluation Agency for RA.\(^{57}\) It was reported the TNFα-Tg mice developed an erosive arthritis with bone and cartilage destruction, and in the treatment of inflammatory bone loss disease, the administration of monoclonal antibodies against human TNF-α significantly decreased synovitis (~5-fold; \(P<0.05\) vs placebo) and potentially suppressed the pathological development.\(^{58,59}\) In the present study, PAT decreased LPS/IFN\(\gamma\)-induced cytokines expression of macrophage and showed beneficial effect on TNFα-Tg mice, as PAT reduced inflammatory area, bone erosion, cartilage loss and TRAP\(^+\) osteoclast area in knee and ankle joints. The curative effect of TP was not bothered by the carrier system, \(\gamma\)-PGA-grafted L-Asp. These results suggested that our PAT exhibited anti-inflammatory effect, similar to anti-TNF therapy.

Despite the promising bioactivities, the toxicity of TP limits its widespread clinical usage. In the current study, PAT decreased the death rate and toxicity at the liver and spleen induced by TP in mice, which suggests that PAT is safe for in vivo usage.

The emulsification of TP in PAT resulted in favorable characteristics such as high encapsulation efficiency, low energy consumption and mild preparation conditions, which are promising for more hydrophobic substances. Therefore, PAT represents a very effective drug candidate for RA with low adverse side effect.

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Disclosure
The authors report no conflicts of interest in this work.

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