Traditional Uses, Phytochemistry, Pharmacology, and Quality Control of Dendrobium officinale Kimura et. Migo

Wenhua Chen, Jiemiao Lu, Jiahao Zhang, Jianjun Wu, Lilong Yu, Luping Qin* and Bo Zhu *

School of Pharmaceutical Sciences, Zhejiang Chinese Medical University, Hangzhou, China

Dendrobium officinale, a well-known plant used as a medicinal and food homologous product, has been reported to contain various bioactive components, such as polysaccharides, bibenzyls, phenanthrenes, and flavonoids. It is also widely used as a traditional medicine to strengthen "Yin", nourish heart, tonify five viscera, remove arthralgia, relieve fatigue, thicken stomach, lighten body, and prolong life span. These traditional applications are in consistent with modern pharmacological studies, which have demonstrated that D. officinale exhibits various biological functions, such as cardioprotective, anti-tumor, gastrointestinal protective, anti-diabetes, immunomodulatory, anti-aging, and anti-osteoporosis effects. In this review, we summarize the research progress of D. officinale from November 2016 to May 2021 and aim to better understand the botany, traditional use, phytochemistry, and pharmacology of D. officinale, as well as its quality control and safety. This work presents the development status of D. officinale, analyzes gaps in the current research on D. officinale, and raises the corresponding solutions to provide references and potential directions for further studies of D. officinale.

Keywords: Dendrobium officinale, phytochemistry, pharmacology, quality control, traditional use

INTRODUCTION

The genus Dendrobium (Orchidaceae) includes more than 1,000 species (Teixeira and Ng, 2017). Among them, Dendrobium officinale Kimura et. Migo is a precious medicinal plant recorded in Chinese Pharmacopoeia, which is widely used as a traditional Chinese medicine (Editorial Board of China Pharmacopoeia Committee, 2020). Moreover, there is a synonym of "Dendrobium catenatum Lindl." in the plant list (http://ipni.org/urn:lsid:ipni.org:names:628140-1). It is well known that D. officinale is distributed in several countries around the world, such as the United States, Japan, and Australia. In particular, D. officinale exhibits a broader distribution in the different regions of China, including Anhui, Zhejiang, Hunan, Fujian, Guangxi, Sichuan, and Yunnan Provinces (Guo et al., 2020a). Due to the overexploitation and depletion of its wild plant resources, it has been considered as a secondary endangered plant in the “China Plant Red Data Book” (Fu, 1992). On the other hand, the artificial cultivation technology of D. officinale has made a significant breakthrough (Cheng Y et al., 2019). However, the existing cultivation resources of D. officinale are mixed, which results in the uneven product quality and the unsound evaluation system, greatly affecting the practical and reasonable development and utilization of D. officinale.

Pharmacological results published in the literature have revealed multiple promising bioactivities of D. officinale, including cardioprotective (Xiao et al., 2018), anti-tumor (Guo et al., 2019), gastrointestinal protective (Liu et al., 2020), anti-diabetes (Zeng et al., 2020),...
immunomodulatory (Huang et al., 2018), anti-aging (Liang et al., 2017), and anti-osteoporosis (Wang et al., 2018) effects. Besides, it is usually prescribed as one of the ingredients of herbal formula, such as “Tin maple crystal” (Tiepfengdoujing: D. officinale and American ginseng), for regulating the gastrointestinal tract in traditional and contemporary clinical practice (Qin et al., 2019). According to currently available phytochemical investigations, the bioactive chemical components of D. officinale mainly consist of polysaccharides (Zhao et al., 2019), bibenzyls (Zhao et al., 2020), phenanthrenes (Liu et al., 2019), flavonoids (Xing et al., 2018), and alkaloids (Chu et al., 2019). In contrast, according to the literature, only few of these compounds have been evaluated in bioactivity assays (Zhao et al., 2018a; Liu et al., 2018; Xing et al., 2018; Chu et al., 2019; Liu et al., 2019; Lee et al., 2020; Zhao et al., 2020). Additionally, although D. officinale is utilized as an effective matter to treat various diseases, its quality control and safety have yet to be defined.

The present review provides an up-to-date and comprehensive literature analysis of D. officinale on the basis of botany, traditional applications, phytochemistry, pharmacology, quality control, and safety, which may provide new insights into the development and applications of D. officinale as a novel therapeutic agent for the prevention and treatment of diseases.

**MATERIALS AND METHODS**

All the information available about D. officinale was obtained from scientific databases, including Web of Science, PubMed, Google Scholar, Baidu Scholar, Springer, Sci Finder, and ScienceDirect, CNKI, from November 2016 to May 2021, and classic books of Chinese herbal medicines. The keywords included Dendrobium officinale, Dendrobium catenatum, ethnopharmacology, phytochemistry, biological activity, pharmacology, clinic, traditional uses, safety, quality control, toxicology, and other related words. In this paper, D. catenatum was also written as D. officinale. However, the other species in Dendrobium genera were excluded.

**BOTANY**

According to the Chinese Pharmacopoeia (Editorial Board of China Pharmacopoeia Committee, 2020), D. officinale can be twisted into a spring or spiral shape, which can be further heated and dried to prepare Tiepfengdou. D. officinale typically has 2–6 whorls with a length of 3–12 cm and a diameter of 0.2–0.4 cm after straightening. The surface color of D. officinale is yellow-green or light golden yellow, with fine longitudinal wrinkles, obvious nodes. Moreover, sometimes residual gray-white leaf sheaths can be seen on the nodes. The short fibrous roots remaining at the stem base can be observed at one end (Flora of China Editorial Committee, 2009).

D. officinale is solid and fragile with a flat section while showing gray-white to gray-green color and slightly horny (Figure 1).

“Flora of China” records: D. officinale grows upright to a height of 9–35 cm and a thickness of 2–4 mm and is usually cylindrical. The stem is vertical without branches while having many nodes above the middle and 3–5 leaves alternate. The leaves are biseriate, papery, and oblong-lanceolate. The apex is obtuse and slightly hooked. The leaf-sheaths often exhibit purple spots. Its inflorescence racemes usually come from the upper part of old stems, with deciduous leaves and 2–3 flowers. The inflorescence axis is bent back and is 2–4 cm long. Sepals and petals show
yellow-green color and comparable sizes, which are usually end-rounded and have five veins (Flora of China Editorial Committee, 2009). The flowering period is from March to June (Zhao et al., 2018a; Liu et al., 2018).

D. officinale is well-known for its nutritional and medicinal value (Flora of China Editorial Committee, 2009), which often grows in semi-humid mountain rocks over 1,600 m and is widely distributed in Anhui, Zhejiang, Fujian, Guangxi, Sichuan, and Yunnan Provinces in China. Moreover, D. officinale displays a broad geographical distribution in other countries or regions, including India, Australia, Japan, and United States (Guo et al., 2020a).

**TRADITIONAL USES**

D. officinale is mainly used as traditional medicine and healthy food in China. D. officinale was initially recorded in "Shennong’s Herbal Classic" (《神农本草经》, Ming Han Dynasty, A.D. 25–220), which has the ability to strengthen “Yin”, tonify five viscera, nourish heart, remove arthralgia, relieve fatigue, thickening stomach, lighten the body, and prolong life span. In “Variorum of Classic of Herbology” (《本草经集注》, Liang Dynasty, A.D. 502–557), the appearance of D. officinale was mentioned to be rough, fine, and solid with a golden color. Moreover, the quality is better if the shape is like grasshopper legs. In "Records of Famous Doctors" (《名医别录》, Weijin Dynasty, A.D. 220–450), D. officinale was recorded to be non-toxic and could be widely used to nourish the essence, reinforce kidney, calm the stomach, build muscles, expel evil heat, relieve foot and knee pain, and remove convulsion. In “Theory of Property” (《药性论》, Tang Dynasty, A.D. 618–907), it was stated that D. officinale could replenish "Qi" and remove heat. Moreover, Herbal Medicines for Kaibao (《开宝本草》, Song Dynasty, A.D. 960–1,279) also had similar description regarding the medical benefits of D. officinale as described above. In addition, it was also recorded in "Amplified Herbology" (《本草衍义》, Song Dynasty, A.D. 960–1,127) that D. officinale could treat asthenia heat in the stomach. In “Compendium of Herbology” (《本草纲目》, Ming Dynasty, A.D. 1,552–1,578), D. officinale was described as a therapeutic approach for fever, spontaneous sweating, ulcer, and pus. According to the record in “Herbal classics” (《本草经疏》, Ming Dynasty, A.D. 1,552–1,578), D. officinale shows the function to remove dampness of spleen and stomach. It was reported that D. officinale could treat deficiency of spleen and stomach and relieve internal heat or fever in "Herbal Medicines for Mengquan" (《本草蒙筌》, Ming Dynasty, A.D. 1,552–1,578). In “Essential Herbs” (《本草备要》, Qing Dynasty, A.D. 1,636–1,912), it was documented that D. officinale could invigorate spleen and kidney. Similarly, in "Herbal Medicines for Qiuzhen" (《本草求真》, Qing Dynasty, A.D. 1,636–1,912), it was registered that D. officinale has the ability to enter the spleen to remove deficiency heat and kidney astringency. Additionally, it was recorded that D. officinale could mitigate stomach deficiency and heat (inflammation) and check for polydipsia in “Match Medica” (《得配本草》, Qing Dynasty, A.D. 1,636–1,912). The summary of the traditional medicinal uses is shown in Table 1.

**Table 1**: Summary of traditional medicinal uses of D. officinale.

| Traditional use | Part used | Ref. |
|----------------|-----------|-----|
| Removing arthralgia, tonifying five viscera, relieving fatigue, strengthening Yin, thickening stomach, lightening body, and prolonging life | Stem | Shennong’s Herbal Classic (《神农本草经》) (Dong Han Dynasty, A.D. 25–220) |
| Raw stone, fine and solid, color of gold, and the shape of grasshopper legs is better | Stem | Variorum of Classic of Herbology (《本草经集注》) (Liang Dynasty, A.D. 502–557) |
| Nourishing the essence, reinforcing kidney, calming stomach, building muscles, expelling evil heat, relieving foot and knee pain, and removing convulsion | Stem | Records of Famous Doctors (《名医别录》) (Weijin Dynasty, A.D. 220–450) |
| Replenishing Qi and removing heat | Stem | Theory of Property (《药性论》) (Tang Dynasty, A.D. 618–907) |
| Nourishing the essence, calming stomach, building muscles, expelling skin evil heat, relieving foot and knee pain | Stem | Herbal Medicines for Kaibao (《开宝本草》) (Song Dynasty, A.D. 960–1,279) |
| Treating asthenia heat in stomach | Stem | Amplified Herbology (《本草衍义》) (Song Dynasty, A.D. 960–1,127) |
| Treating fever, spontaneous sweating, ulcer, and pus | Stem | Compendium of Herbology (《本草纲目》) (Ming Dynasty, A.D. 1,552–1,578) |
| Removing dampness of spleen and stomach | Stem | Herbal classics (《本草经疏》) (Ming Dynasty, A.D. 1,552–1,578) |
| Treating deficiency of spleen and stomach, and relieve internal heat or fever | Stem | Herbal Medicines for Mengquan (《本草蒙筌》) (Ming Dynasty, A.D. 1,552–1,578) |
| Invigorating spleen and kidney | Stem | Essential Herbs (《本草备要》) (Qing Dynasty, A.D. 1,636–1,912) |
| Entering the spleen to remove deficiency heat and kidney astringency | Stem | Herbal Medicines for Qiuzhen (《本草求真》) (Qing Dynasty, A.D. 1,636–1,912) |
| Removing stomach deficiency heat and check polydipsia | Stem | Match Medica (《得配本草》) (Qing Dynasty, A.D. 1,636–1,912) |
TABLE 2: The main compounds isolated from *D. officinale*.

| Class            | No | Name                                           | Formula     | Plant parts       | Ref.                     |
|------------------|----|------------------------------------------------|-------------|-------------------|--------------------------|
| Bibenzyls        | 1  | 4,4'-Dihydroxy-3,5-dimethoxybibenzyl           | C₁₆H₁₈O₄   | Leaves            | Ren et al. (2020a); Ren et al. (2020b) |
|                  | 2  | 3,4'-Dihydroxy-5-methoxybibenzyl               | C₁₆H₁₈O₃   | Stems             | Liu et al. (2018)         |
|                  | 3  | 3,4-Dihydroxy-4',5-dimethoxybibenzyl           | C₁₆H₁₈O₄   | Leaves            | Ren et al. (2020a); Ren et al. (2020b) |
|                  | 4  | Gigantol                                       | C₁₈H₂₁O₄   | Stems, Leaves     | He et al. (2020); Ren et al. (2020b) |
|                  | 5  | Moscatin                                       | C₁₈H₂₁O₄   | Leaves            | Ren et al. (2020a); Ren et al. (2020b) |
|                  | 6  | Tristin                                        | C₁₆H₁₈O₄   | Stems             | He et al. (2020)          |
|                  | 7  | Eriarin                                        | C₁₆H₂₀O₄   | Stems             | Peng et al. (2020)        |
|                  | 8  | 3,4,4'-Trihydroxy-5-methoxybibenzyl            | C₁₆H₁₈O₃   | Stems             | Chu et al. (2019)         |
|                  | 9  | 3-Hydroxy-4',5-dimethoxybibenzyl               | C₁₆H₁₈O₃   | Stems             | Chu et al. (2019)         |
|                  | 10 | Gigantol                                       | C₁₆H₁₈O₄   | Stems             | He et al. (2020)          |
|                  | 11 | Moscatin                                       | C₁₈H₂₁O₄   | Stems             | He et al. (2020)          |
|                  | 12 | Dihydroresveratrol                             | C₁₄H₁₀O₃   | Stems             | Liu et al. (2018)         |
| Phenanthrenes    | 23 | Ephemeranthol A                                | C₁₆H₁₈O₄   | Stems             | Cui et al. (2019); He et al. (2020) |
|                  | 24 | Eriarchin                                      | C₁₆H₁₈O₄   | Stems             | Cui et al. (2019)         |
|                  | 25 | Orinol                                         | C₁₆H₁₈O₄   | Stems             | Cui et al. (2019)         |
|                  | 26 | 2,4,7-Trihydroxy-9,10-dihydrophenanthrene      | C₁₄H₁₀O₃   | Stems             | Cui et al. (2019)         |
|                  | 27 | Conflurarin                                     | C₁₆H₁₈O₄   | Stems             | Cui et al. (2019)         |
| Penypropanoids   | 28 | 2,7-Dihydroxy-3,4-dimethoxyphenanthrene        | C₁₆H₁₈O₄   | Stems             | Cui et al. (2019)         |
|                  | 29 | 1-O-Caffeoyl-β-D-glucoside                     | C₁₅H₁₈O₆   | Flowers           | Zhang et al. (2019)       |
|                  | 30 | 1-O-p-Coumaroyl-β-D-glucoside                  | C₁₅H₁₈O₆   | Flowers           | Zhang et al. (2019)       |
|                  | 31 | ethyl-p-Hydroxyhydrocinnamate                   | C₁₁H₁₂O₃   | Leaves            | Ren et al. (2020a)        |
|                  | 32 | ω-Hydroxypropoglaucione                         | C₁₈H₂₂O₄   | Stems             | Zhe et al. (2020)         |
|                  | 33 | trans-3,4,5-Trimethoxy-cinnamyl alcohol         | C₁₈H₂₂O₄   | Stems             | Cui et al. (2019)         |
|                  | 34 | Dihydroconiferylhydro-p-coumarate              | C₁₈H₂₂O₄   | Stems             | Ye et al. (2017)          |
|                  | 35 | 6-Hydroxy-3-oxo-a-ionol                        | C₁₈H₂₂O₃   | Stems             | Zhe et al. (2020)         |
|                  | 36 | Scopalone                                       | C₁₁H₁₀O₄   | Stems             | Peng et al. (2020)        |
|                  | 37 | Syringaresinol                                  | C₂₀H₂₂O₆   | Stems, Leaves     | Ye et al. (2017); Ren et al. (2020a) |
|                  | 38 | Magnoliner C                                   | C₂₀H₂₄O₁₄  | Stems             | Cui et al. (2019)         |
|                  | 39 | Oficinalisolate                                 | C₂₀H₂₄O₁₄  | Stems             | Cui et al. (2019)         |
|                  | 40 | Moelinoside A                                  | C₂₀H₂₄O₁₁  | Stems             | Cui et al. (2019)         |
|                  | 41 | Phoroneisol-4-O-β-D-glucopyranoside            | C₂₀H₂₄O₁₄  | Stems             | Cui et al. (2019)         |
|                  | 42 | 5,5'-Dimethoxy-lanecresinol                    | C₂₀H₂₄O₁₆  | Stems             | He et al. (2020)          |
|                  | 43 | Lyoneresinol                                   | C₂₀H₂₄O₁₆  | Stems             | He et al. (2020)          |
|                  | 44 | (+)-Syringaresinol-4'-O-β-D-glucopyranoside    | C₂₀H₂₄O₁₃  | Stems             | Chen et al. (2018)        |
| Flavonoids       | 45 | Apigenin                                       | C₁₅H₁₀O₅   | Roots, Stems, Leaves | Yu et al. (2018); Ren et al. (2020c) |
|                  | 46 | Naringenin                                     | C₁₅H₁₀O₅   | Stems             | Ye et al. (2017); Peng et al. (2020) |
|                  | 47 | Naringin                                       | C₂₀H₂₄O₁₄  | Stems             | Zhou et al. (2018)        |
|                  | 48 | 2-Hydroxyxaringerin                            | C₁₅H₁₀O₅   | Roots, Stems, Laves | Ren et al. (2020c)        |
|                  | 49 | Phorolentin                                    | C₁₅H₁₀O₅   | Roots, Stems      | Ren et al. (2020c)        |
|                  | 50 | Eldodicytol                                    | C₁₅H₁₀O₅   | Stems             | Lv et al. (2017)          |
|                  | 51 | Chrysoscelol                                   | C₁₅H₁₀O₅   | Stems             | Lv et al. (2017)          |
|                  | 52 | Quercetin                                      | C₁₅H₁₀O₅   | Stems             | Lv et al. (2017)          |
|                  | 53 | Taxelolin                                      | C₁₅H₁₀O₅   | Stems             | Lv et al. (2017)          |
|                  | 54 | Isorhamnetin                                   | C₁₅H₁₀O₅   | Stems             | Lv et al. (2017)          |
|                  | 55 | Rutin                                          | C₁₅H₁₀O₅   | Stems, Leaves, Flowers | Ye et al. (2017); Zhou et al. (2018); Ren et al. (2020a); Zhang et al. (2019) |
|                  | 56 | Cosmosin                                       | C₁₂H₁₀O₁₀  | Flowers           | Ren et al. (2020c)        |
|                  | 57 | Genin-7-0-gentlobioside                         | C₁₂H₁₀O₁₂  | Stems             | Yu et al. (2018)          |
|                  | 58 | Pelargonidin 3,5-O-diglucoside                 | C₁₂H₁₀O₁₂  | Stems             | Yu et al. (2018)          |
|                  | 59 | Pelargonidin 3-O-rutinoside                    | C₁₂H₁₀O₁₂  | Stems             | Yu et al. (2018)          |
|                  | 60 | Malvidin 3-O-glucosid                          | C₁₂H₁₀O₁₂  | Stems             | Yu et al. (2018)          |
|                  | 61 | Vicenin I                                      | C₁₂H₁₀O₁₂  | Roots, Stems, Leaves, Flowers | Ye et al. (2017); Ren et al. (2020c); Zhang et al. (2017b) |

(Continued on following page)
### TABLE 2 | The main compounds isolated from *D. officinale*.

| Class | No. | Name | Formula | Plant parts | Ref. |
|-------|-----|------|---------|-------------|------|
|      |     |      |         |             |      |
| 51   |     |      |         |             |      |
| 52   |     |      |         |             |      |
| 53   |     |      |         |             |      |
| 54   |     |      |         |             |      |
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| 97   |     |      |         |             |      |
| 98   |     |      |         |             |      |
| 99   |     |      |         |             |      |
| 100  |     |      |         |             |      |
| 101  |     |      |         |             |      |
| 102  |     |      |         |             |      |
| 103  |     |      |         |             |      |

**Alkaloids**

| 104  |     | Anosmine | C_{11}H_{17}N_{2} | Stems | Chen et al. (2018) |
| 105  |     | Benzoisothiazol | C_{5}H_{4}N_{5}S | Leaves | Ren et al. (2020a) |
| 106  |     | 3,5-Dimethoxyphenethyllamines | C_{12}H_{13}NO_{3} | Stems | Zhi et al. (2018) |
| 107  |     | N-p-coumaroyltyramine | C_{10}H_{11}NO_{4} | Stems | Chui et al. (2019) |
| 108  |     | N-trans-p-fenuryltyramine | C_{11}H_{17}NO_{4} | Stems | Chui et al. (2019) |
| 109  |     | Malic acid | C_{4}H_{6}O_{3} | Stems | Chen et al. (2018) |
| 110  |     | Fumaric acid | C_{4}H_{4}O_{4} | Stems | Ye et al. (2017) |
| 111  |     | Succinic acid | C_{4}H_{6}O_{4} | Stems | Ye et al. (2017) |
| 112  |     | Syringic acid | C_{10}H_{10}O_{3} | Leaves | Ren et al. (2020a) |
| 113  |     | Protocatechuic acid | C_{9}H_{6}O_{3} | Leaves | Ren et al. (2020a) |
| 114  |     | p-Hydroxybenzoic acid | C_{7}H_{6}O_{3} | Leaves | Ren et al. (2020a) |
| 115  |     | p-Hydroxybenzoic acid | C_{7}H_{6}O_{3} | Leaves | Ren et al. (2020a) |
| 116  |     | Palmitic acid | C_{16}H_{32}O_{2} | Leaves | Ren et al. (2020a) |

(Continued on following page)
China, 2013). In traditional Chinese therapies, *D. officinale* is often combined with other Chinese herbal medicines, including *Adenophora stricta*, *Ophiopogon japonicus*, *Paecia lactiflora*, *Astragalus membranaceus* and *Polygonatum odoratum*. In recent years, pharmacological research mainly focused on the cardioprotective (Xiao et al., 2018), anti-tumor (Guo et al., 2019), gastrointestinal protective (Liu et al., 2020), and anti-diabetes (Zeng et al., 2020) effects of *D. officinale*. Therefore, it is encouraged to combine the traditional pharmacological actions of *D. officinale* and advanced research technologies in order to explore the potential mechanisms and establish the foundation for its clinical applications in the future.

**PHYTOCHEMISTRY**

Extensive phytochemistry studies have demonstrated that *D. officinale* mainly contains polysaccharides, bibenzyls (1–22), phenanthrenes (23–28), phenylpropanoids (29–44), flavonoids (45–103), alkaloids (104–108), acids (109–116), and others (117–126). Among them, polysaccharides, bibenzyls, and flavonoids are considered the main bioactive compounds responsible for various pharmacological properties and therapeutic efficacy of *D. officinale*. All compounds identified in *D. officinale* are summarized and shown in Table 2, and the corresponding structures are shown in Figures 2–8.

**Polysaccharides**

Polysaccharides are the main medicinal components of *D. officinale*, with a wide range of medicinal properties. A large number of polysaccharides have been isolated from *D. officinale*, including DWDO1, DWDO2, DWDO3, FWDOP1, FWDOP2, FWDOP3 (Yu et al., 2018), DOP-50, DOP-60, DOP-70 (Xing et al., 2018), DO (Ma et al., 2018), DOPA-1 (Wei et al., 2018), DOP1-DES, DOP2-DES (Liang J et al., 2018), LDP1-1 (Yang et al., 2020), UDP-1, FLP-1, and FDP-1 (Liang K. L. et al., 2018), which are identified by various analytical technologies, such as high performance liquid chromatography (HPLC), high performance gel permeation chromatography (HPAEC), gas chromatography-mass spectrometry (GC-MS), and gel permeation chromatography (GPC). The molecular weights of *D. officinale* polysaccharides range from 30 to 1,415 kDa (Liang J et al., 2018; Ma et al., 2018; Wei et al., 2018; Yu et al., 2018). It has been demonstrated that *D. officinale* polysaccharides are composed of glucose, mannose, galactose, xylose, arabinose, ribose and rhamnose (Tian et al., 2019). Among them, glucomannan is considered to be the main component of *D. officinale* polysaccharides with 1,4-β-D-Manp and 1,4-β-D-GlcP, comprising acetyl groups in varying degrees and positions with or without branches (Liang et al., 2019). It should be mentioned that a lot of factors may impact the biological activity of polysaccharides, including molecular structure, main chain composition, molecular branching degree, the configuration of the main chain, and chemical modification ( Yue et al., 2017; Huang et al., 2020 ). Importantly, according to the literature, polysaccharides have exhibited various therapeutic potentials, including cardioprotective (Zhang et al., 2017a; Su et al., 2021), anti-tumor (Wei et al., 2018; Zhao et al., 2019), gastrointestinal protective (Ke et al., 2020; Zhang et al., 2020), immunomodulatory (Huang et al., 2018), anti-aging (Liang et al., 2017; Wei et al., 2017), and pulmonary protective effects (Chen et al., 2020).

**Bibenzyls**

Bibenzyl components in *D. officinale* have attracted a lot of attention due to their promising anti-tumor properties (He et al., 2020). To date, 19 bibenzyls have been isolated, which were identified predominantly from the stems and leaves of *D. officinale*. The typical parent nucleus structure of bibenzyls presented in the stems of *D. officinale* is two benzene rings, which can be replaced by different substituents. Some bibenzyls (4, 5, and 7) were found to have significant anti-tumor activity (Tian et al., 2019; Zhao et al., 2020), which attracted further research and development. The chemical structures of these bibenzyls are shown in Figure 2.

**Phenanthrenes**

The phenanthrenes are abundantly found in the stems of *D. officinale*, including ephemeralanthol A (23), erianthridin (24), orchinol (25), 2, 4, 7-trihydroxy-9, 10-dihydrophenanthrene (26), confusarin (27), and 2,7-dihydroxy-3,4-dimethoxyphenanthrene (28). Notably, orchinol (25) possesses anti-tumor activity, which can potentially be used to develop new anti-tumor drugs (Zhao et al., 2018a). The chemical structures of these phenanthrenes are shown in Figure 3.

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**Table 2** | The main compounds isolated from *D. officinale*.

| Class | No | Name | Formula | Plant parts | Ref. |
|---|---|---|---|---|---|
| Others | 117 | Filtriodioside A | C_{30}H_{29}O_{13} | Stems | Chen et al. (2018) |
| | 118 | Flickrintmoside B | C_{31}H_{30}O_{13} | Stems | Chen et al. (2018) |
| | 119 | Lololidine | C_{17}H_{18}O_{3} | Leaves | Ren et al. (2020a) |
| | 120 | 1-Glycerol linolenate | C_{18}H_{32}O_{4} | Leaves | Ren et al. (2020a) |
| | 121 | Densiflorol A | C_{18}H_{20}O_{2} | Leaves | Ren et al. (2020a) |
| | 122 | 2-Butoxyethyl linolenate | C_{19}H_{22}O_{4} | Leaves | Ren et al. (2020a) |
| | 123 | Catechol | C_{6}H_{8}O_{2} | Leaves | Ren et al. (2020a) |
| | 124 | Octadecaenoic acid-2,3-dihydroxypropyl ester | C_{28}H_{48}O_{4} | Leaves | Ren et al. (2020a) |
| | 125 | Stigmaster-5-en-3β-ol-7-one | C_{29}H_{48}O_{3} | Stems | Zhu et al. (2020) |
| | 126 | Dendrofindiaphenol B | C_{25}H_{38}O_{6} | Stems | Liu et al. (2018) |
Phenylpropanoids
Phenylpropanoid compounds refer to natural organic compounds with one or more C₆-C₃ units in the basic parent nucleus, mainly including simple phenylpropanoids, coumarins, and lignans. To date, 16 phenylpropanoids (29–44) have been identified in *D. officinale*, including simple phenylpropanoids (29–35), coumarin (36), and lignans (37–44). The chemical structures of these phenylpropanoids are presented in Figure 4.

Flavonoids
It has been reported that flavonoids belong to a large group of secondary metabolites in *D. officinale*, which possess anti-tumor activity (Xing et al., 2018). Most flavonoids isolated and identified from the roots, stems, leaves, and flowers of *D. officinale* are C-glycosides, while the rest of flavonoids are O-glycosides. It is well known that the foundational skeletons of flavonoids are apigenin, vitexin, quercetin, and kaempferol. Nowadays, mass spectrometry technologies coupled with liquid chromatography, such (HPLC-ESI-MS) (Ye et al., 2017), ultra-high-performance liquid chromatography (UPLC-ESI-MS/MS) (Zhou et al., 2018), and UPLC-quadrupole time of flight mass spectrometry (UPLC-QTOF-MS) (Yu et al., 2018), have been widely implemented for the identification and quantification of these flavonoid compounds. Interestingly, it is generally believed that glucoside derivatives (anthocyanins) delphinidin 3,5-O-diglucoside and cyanidin 3-O-glucoside are responsible for

**Figure 4** Structures of bibenzyls (1-22) isolated from *D. officinale*. 
the red color stems of D. officinale (Yu et al., 2018). In addition, the transcriptome and component metabolism analyses have become the typical approaches to investigate flavonoid biosynthesis mechanisms in D. officinale (Lei et al., 2018). The chemical structures of these flavonoids are shown in Figures 5–7.

Alkaloids
Alkaloids are a class of nitrogenous organic compounds possessing various biological activities. They are also active constituents of Dendrobium plants. It is worth noting that the regulation of genes related to alkaloid biosynthetic pathways through comparative transcriptomic analysis has gradually become a research interest (Jiao et al., 2018). The chemical structures of these alkaloids are shown in Figure 8.

Organic Acids
Organic acids are a kind of acid organic compounds containing a carboxyl group. Eight types of organic acids have been identified in D. officinale. The chemical structures of these organic acids are displayed in Figure 9.

Other Compounds
Some other types of compounds have also been isolated from the stems and leaves of D. officinale, including filifimbioside A (117), flickinflimoside B (118) (Chen et al., 2018), loliolide (119), 1-glycerol linolenate (120), densiflorol A (121), 2-butoxyethyl linolenate (122), catechol (123) (Ren et al., 2020a), octadecadienoic acid-2,3-dihydroxypropyl ester (124) stigmast-5-en-3β-ol-7-one (125) (Zhu et al., 2020), and dendrofindaphenol B (126) (Liu et al., 2018). The chemical structures of these compounds are shown in Figure 10.

PHARMACOLOGY
D. officinale has been found to possess multiple biological functions, including cardioprotective (Zhang et al., 2017a; Su et al., 2021), anti-tumor (Guo et al., 2019), gastrointestinal protective (Liu et al., 2020), anti-diabetes (Zeng et al., 2020), immunomodulatory (Huang et al., 2018), anti-aging (Liang et al., 2017), and anti-osteoporosis (Wang et al., 2018) effects. Among them, modern pharmacological studies of D. officinale majorly focus on its cardioprotective, anti-tumor, gastrointestinal protective, and hypoglycemic effects. The biological activities of D. officinale and corresponding mechanisms are shown in Figure 11.

Cardioprotective Activity
Cardiovascular diseases are the major causes of mortality globally, and the main cause of more than 10 million people death. Among cardiovascular diseases risk factors, cardiomyopathy and high blood pressure are the most common ones. Oral administration of D. officinale fine powders at the doses of 0.09, 0.18, and a very high dose of 1.1 g/kg for 30 days protected isoproterenol (ISO)-induced cardiac hypertrophy, indicated by the decreased myocardial collagen synthesis, increased myocardial fibrosis and ventricular remodeling, and significantly reduced levels of atrial natriuretic peptide (ANP), brain natriuretic peptide (BNP), and cardiac troponin I (cTn-I) in plasma related to the model group (ISO = 5 mg/kg) (Xiao et al., 2018). In addition, it was well known that diabetic cardiomyopathy was a typical cardiovascular complication mediated via hyperglycemia. One study indicated that D. officinale water-soluble extracts prevented diabetic cardiomyopathy and might be a candidate for therapeutic use (Zhang et al., 2016). D. officinale water extracts (75, 150, and 300 mg/kg) intragastrically once daily for 2 weeks could protect left anterior descending coronary artery (LAD)-induced myocardial ischemia through decreasing creatine kinase (CK)-MB, lactate dehydrogenase (LDH), malondialdehyde (MDA), and increasing superoxide dismutase (SOD) and Meis 1 levels (Dou et al., 2016). Treatment with D. officinale stems polysaccharide DOP-GY at the doses of 6.25, 12.5, and 25 μg/ml exhibited protective effects on hydrogen peroxide (H₂O₂)-induced H9c2 cardiomyocyte apoptosis via phosphatidylinositol/protein kinase B (PI3K)/Akt and mitogen-activated protein kinase (MAPK) signaling pathways, as demonstrated by the decreased levels of LDH, lipid peroxidation damage (LPD), reactive oxygen species (ROS), and pro-apoptosis protein (Zhang et al., 2017a).
Another study demonstrated that *D. officinale* polysaccharides could decrease malondialdehyde levels, increase SOD activities, and inhibit the generation of intracellular ROS in H9c2 cardiomyocytes (Zhao et al., 2017). Besides, Yue et al. (2017) obtained a novel homogeneous heteroxylan from alkali-extracted *D. officinale* crude polysaccharide (S32), which possessed significant anti-angiogenic effects (S32, 13.51 μM) on human microvascular endothelial cells (HMEC-1) by inhibiting their migration and disruption of tube formation in a dose-dependent manner, compared with a vehicle group (Yue et al., 2017). However, some of these experimental doses were too high and there was a lack of positive controls.

It has been suggested that *D. officinale* has cardioprotective activity by treating hypertension. It was reported that blood pressure was significantly reduced after treating with *D. officinale* ultrafine powder (DOFP) at very high doses of 200 and 400 mg/kg for 20 weeks exhibited anti-hypertensive activity on overeating greasy-induced metabolic hypertension in rats by inhibiting the activation of lipopolysaccharide/toll-like receptor 4 (LPS/TLR4) signal pathway, as demonstrated by the decreased levels of total cholesterol (TC), triglyceride (TG), low-density lipoprotein cholesterol (LDL-c), LPS, C-reactive protein (CRP), interleukin 6 (IL-6), TLR4, myeloid differentiation factor (MyD88), IL-1β, and tumor necrosis factor alpha (TNF-α) and the increased levels of high-density lipoprotein cholesterol (HDL-c) and nitric oxide (NO) relative to valsartan (8 mg/kg)-treated positive control (Su et al., 2021). Moreover, Yan et al. (2019a) reported that the effective component of alcohol extract of *D. officinale* in the treatment...
of metabolic hypertension was apigenin flavonoid glycosides (Yan et al., 2019a). In contrast, more investigations are still needed to reveal the underlying mechanism of the anti-hypertensive effect of apigenin flavonoid glycosides. The results showed that treatment with D. officinale flowers at a very high dose of 3.1 g/kg for 6 weeks could significantly improve vascular diastolic function by reducing systolic blood pressure and mean arterial pressure in high glucose and fat compound alcohol-induced hypertensive rats, inhibiting the thickening of thoracic aorta and the loss of endothelial cells, reducing plasma content of endothelin 1 (ET-1) and thromboxane B2 (TXB2), and increasing the content of prostacycline (PGI2) and NO, compared with model control group and valsartan (5.7 mg/kg) positive control group (Liang K. L et al., 2018). Treating with D. officinale ultrafine powder DOFP could improve the intestinal flora and increase the production, transportation, and utilization of short-chain fatty acid (SCFA), activate the intestinal-vascular axis SCFA-GPCR43/41 pathway, increase vascular endothelial function, and finally decrease the blood pressure in alcohol, and high sugar and fat diets (ACHSFD)-induced metabolic hypertension model rats (Li et al., 2021). These results suggested that D. officinale may have a potential clinical application in the treatment of hypertension. In this study, however, the optimal dose, constituents, and side effects of D. officinale are not assessed. Moreover, further detailed clinical trials should be employed to assess the value of D. officinale as a drug for the treatment of hypertension. In addition, this evidence is still tenuous; no double-blind trials involving D. officinale have been performed, and more evidence from randomized controlled trials is required to elucidate other mechanisms that may be responsible for anti-hypertension effects.

Although D. officinale possesses a potential therapeutic effect on cardiovascular diseases, especially cardiomyopathy and hypertension (Figure 12), more in-depth investigations on its
Effective monomer compounds, molecular mechanism, and clinical trials are warranted to identify effective cardioprotective agents with minimized side effects.

**Anti-Tumor Activity**

As a type of traditional medicine and ordinary food, *D. officinale* has benefits on human health supported by its effectiveness in the prevention and treatment of cancer diseases (Guo et al., 2019). Several studies have reported all crude extracts, polysaccharides, and other pure compounds isolated from *D. officinale* exhibited anti-tumor activities (Wei et al., 2018; Guo et al., 2019; Zhao et al., 2020).

Administration of *D. officinale* methanol extracts at a dose of 0.25, 0.5, and 1 mg/ml could inhibit the growth of SMMC-7721, BEL-7404 cells, and primary liver cancer cells, and promote their apoptosis via activating mitochondria apoptosis pathway and suppressing the Wnt/β-catenin pathway (Guo et al., 2019). Similarly, another study demonstrated that *D. officinale*...
polysaccharide (DOPA-1) induced HepG-2 cell apoptosis by influencing mitochondrial function, ROS production, and apoptosis-related protein expression (Wei et al., 2018). Importantly, a rat study also confirmed the anti-tumor effects of *D. officinale* in vivo. The 2-weeks administration of *D. officinale* polysaccharide DOP at very doses of 2.4, 4.8, and 9.6 g/kg suppressed 1-methyl-2-nitro-1-nitrosoguanidine-(MNNNG)-induced (150 µg/ml) precancerous lesions of gastric cancer in rats via modulating Wnt/β-catenin pathway and altering endogenous serum metabolites (Zhao et al., 2019). In addition to crude extracts and polysaccharides, it is more evident that gigantol (4), moscatilin (5), erianin (7), orchinol (25), and isoviolanthin (71) isolated from *D. officinale* are also responsible for the anticancer activity of *D. officinale* (Xing et al., 2018; Liu et al., 2019; Lee et al., 2020; Zhao et al., 2020). Gigantol (4) was found to repress invasiveness and growth of SW780, 5,637, and T24 human bladder cancer cells by inhibiting the Wnt/epithelial-mesenchymal transition (EMT) signaling (Guo et al., 2019). Likewise, moscatilin (5) was demonstrated to induce apoptosis in FaDu human head and neck squamous carcinoma cells (HNSCC) via c-Jun N-terminal kinase (JNK) signaling pathway (Lee et al., 2020). The anti-tumor function of erianin (7) was investigated by two independent studies. The results suggest that erianin (7) induced cell apoptosis through the
ERK pathway in nasopharyngeal carcinoma (NPC) (Liu et al., 2019) while suppressing the growth of bladder cancer cells EJ and T24 through JNK pathways with the IC50 values of 65.04 and 45.9 nM, respectively (Zhu et al., 2019). Moreover, it was demonstrated that orcinol (25) exhibited strong cytotoxic activity in HI-60 and THP-1 cells with the IC50 values of 11.96 and 8.92 μM, respectively (Lee et al., 2020). Additionally, isoviolanthin (71) was revealed to suppress transforming growth factor (TGF-β1)-induced EMT through the regulation of TGF-β/Smad and PI3K/Akt/mTOR signaling pathways in HepG2 and Bel-7402 hepatocellular carcinoma (HCC) cells (Xing et al., 2018). However, the dose of D. officinale polysaccharide DOP was too high in the treatment of gastric cancer, attention should be paid to the possible side-effects in clinical applications.

Overall, the anti-tumor mechanisms of D. officinale are mainly attributed to promoting tumor cell apoptosis, inhibiting tumor...
cell proliferation, repressing tumor cell migration and invasion, and improving body immunity (Figure 13). The studies described above suggest that the compounds identified in *D. officinale* demonstrate significant inhibitory effects on different types of tumor cells, such as SMMC-7721, BEL-7404, HepG-2, SW780, and HCC cell lines. Further investigations may explore the structural modification and the structure-activity relationship (SAR) studies for these bioactive compounds of *D. officinale*, thereby facilitating the further discovery and development of new anti-tumor candidates, along with the characterization of the key regulatory genes, metabolic pathways, and heterologous biosynthesis pathways of active ingredients from *D. officinale*. The currently published studies on the anti-tumor effects of the compounds from *D. officinale* are mainly focused on in vitro and in vivo experiments, while clinical studies have yet to be conducted and the exact molecular mechanisms remain elusive.

**Gastrointestinal Protective Effect**

*D. officinale* is traditionally used to nourish “Yin” and thicken stomach. *D. officinale* extracts have thus been frequently applied to treat gastrointestinal diseases as a traditional Chinese medicine. It was reported that polyphenols from fermentation liquid of *D. officinale* improved intestinal health via the regulation of intestinal microbiota and their metabolites, thereby relieving oxazolone-induced intestinal inflammation in Zebrafish (Gong et al., 2020). Moreover, *D. officinale* is often used in combination with other traditional Chinese medicines to achieve its better therapeutic effects. It was found that the mixture of *D. officinale* and American ginseng at a dose of (0.32 × the dog's weight × 6 g)/12 kg could function as a probiotic agent to enhance SCFA-producing genera and reverse gut dysbiosis (Liu et al., 2020). Likewise, the combination of *D. officinale* and other Chinese herbal medicine, such as *Acanthopanax senticosus*, *Panaxnotoginseng*, *Didymocarpus hancei* and *Valeriana officinalis* can also alleviate gastric mucosal injury (Guo, 2020b). The 14-days treatment of *D. officinale* glucomannans at 0.16 g/kg produced more SCFAs (mainly acetate and butyrate) in cecum and colon (Shi et al., 2020). It should be noted that polysaccharides LDOP-1 isolated from *D. officinale* could protect ethanol-induced gastric mucosal injury in vitro (250, 125, and 62.5 mg/ml for 2 h) and in vivo (100 and a very high dose of 400 mg/kg for 30 days) by regulating AMP-activated protein...
kinase (AMPK)/mTOR signaling pathway demonstrated by the increased levels of p-AMPK, light chain 3β (LC-3β), heme oxygenase-1 (HO-1) and Beclin-1, the decreased levels of p-mTOR and p62, and the reversed levels of caspase3, Bax, and Bcl-2 detected both in vitro and in vivo (Ke et al., 2020). Furthermore, it was suggested that D. of fici nale polysaccharide (200 mg/kg/d) exhibited protective effects against DSS-induced colitis by inhibiting pro-inflammatory cytokines TNF-α, IL-6, and IL-1β in the colonic mucosa, modulating the abundance of gut microbiota, and promoting the production and utilization of SCFAs in the colon (Zhang et al., 2020). However, positive control or dose-dependent effect analysis was not performed, which might require further validation. In addition, the structure of the correlation between polysaccharide administration and health outcomes, as well as the functional role of the polysaccharides themselves has not been examined in depth.

**Anti-Diabetes Activity**

Diabetes is a common disease with glucose metabolism disorder, which seriously affects human health around the world (Ye et al., 2017). Numerous ethnomedicinal studies supported the traditional use of *D. officinale* for clearing heat, nourishing “Yin”, benefiting the stomach, and promoting body fluid, thereby leading *D. officinale* to become an essential medicine to treat “Xiao Ke” disease (Diabetes) for thousands of years (Yan et al., 2019b).

Over the past few years, the hypoglycemic effect of *D. officinale* has become an attractive research field. One study has confirmed that the administration of water extracts from the stems of *D. officinale* at the doses of 75, 150, and a very high dose of 300 mg/kg for 12 weeks decreased serum insulin, TC, TG, LDL-c, and increase HDL-c in high-fat diet/streptozotocin (HFD/STZ) (30 mg/kg)-induced diabetic mice in a dose-dependent manner (Zeng et al., 2020). The possible mechanism of *D. officinale* water extracts can be associated with improving lipid transport and suppressing insulin resistance and fibrosis via EMT. Another study similarly found that a 4-weeks treatment with *D. officinale* water extracts at very high doses of 350 and 700 mg/kg elevated the liver glycogen synthesis, energy, and amino acid metabolism as well as taurine-mediated defense against oxidative stress in STZ-induced diabetic mice (Zheng et al., 2017). It was found that the

**FIGURE 11 |** The bioactivities of main compounds isolated from *D. officinale* and their underlying mechanisms.
The treatment of *D. officinale* stem’s water extracts at very high doses of 10 and 20 g/kg for 4 weeks ameliorated insulin resistance by decreasing toll-like receptors (TLRs) and inflammatory response in STZ-induced diabetic rats in comparison with the treatment of dimethyl biguanide (DMBG, 0.0042 g/kg) as a positive control (Zhao and Han 2018b). Moreover, it was reported that the administration of *D. officinale* at the doses of 0.2, 0.4, and 0.8 g/kg for 8 weeks protected against diabetic kidney lesions.
in STZ-induced (60 mg/kg) rats via suppressing vascular endothelial growth factor (VEGF), glucose transporter 1 (GLUT-1), and connective tissue growth factor (CTGF) relative to the irbesartan (17.5 mg/kg)-treated positive control (Chang et al., 2019). Additionally, a previous study also revealed that the hypoglycemic effect of the 3-months combined treatment of *D. officinale* (10 g/d) and metformin (0.5 g/d) was superior to the single-use (Wu et al., 2017). However, these studies have important limitations—one only or two doses of *D. officinale* are used in these studies. In addition, the evaluation of *D. officinale* doses that are much higher, and it has no translational value from a therapeutic viewpoint.

It is evident that the hypoglycemic effect of *D. officinale* is closely related to the inhibition of α-glucosidase and α-amylase (Zhu et al., 2020). Notably, 3,4′-dihydroxy-5-methoxybibenzyl (2) and dihydroresveratrol (12) isolated from *D. officinale* extracts have been reported to exhibit hypoglycemic activity in an α-glucosidase inhibitory assay with IC_{50} values of 36.05 ± 0.67 and 159.59 ± 0.86 μM, respectively, relative to the acarbose as a positive control (IC_{50} = 152.86 ± 1.43 μM) (Liu et al., 2018). Likewise, 3,4-dihydroxy-4′,5-dimethoxybibenzyl (3), 3,4′-trihydroxy-5-methoxybibenzyl (8), dendrocandin U (15), N-p coumaroyltyramine (107), and N-trans-p-feruloyltyramine (108) have been identified as α-glucosidase inhibitors with IC_{50} values of 199.5 μM, 9.46 mM, 403.4, 0.4 and 234.1 μM, respectively, compared with acarbose (IC_{50} = 763.5 μM). Furthermore, the inhibitory effect of 3,4-dihydroxy-4′,5-dimethoxybibenzyl (3) on α-amylase (an IC_{50} value of 5.99 mM) was weaker than acarbose (IC_{50} = 0.12 μM) (Chu et al., 2019). Several studies have demonstrated the definite hypoglycemic effect of *D. officinale*, while further investigations are required to identify the specific bioactive components responsible for this activity and clarify the hypoglycemic mechanism of *D. officinale*.

**Immunomodulatory Activity**

A large number of experiments have provided evidence for the immunomodulatory activity of *D. officinale*. Polysaccharide is the main active component for immunoregulation. Oral administration of 0.25% *D. officinale* polysaccharide DOW-5B (w/v) for 25 days displayed significant immunomodulatory effects via increasing the content of butyrate, immunoglobulin M (IgM), IL-10, and TNF-α in vivo (Li et al., 2020a). Another study also demonstrated that *D. officinale* polysaccharide DOP-1-1 stimulated immunity by enhancing the level of nuclear factor kappa-B (NF-κB) while inhibiting the level of IkBα through TLR4 signaling (Huang et al., 2018). Moreover, it was found that *D. officinale* polysaccharides (FDP-1) treatments, ranging from 12.5 to 200 μg/ml, exhibited immunomodulatory activity through increasing cell proliferation and NO and IL-1β production in a dose-dependent manner (Tian et al., 2019). Neither positive control nor dose-effect analysis in vivo was assessed in these studies. In addition, there is a general lack of systematic research on the relationship between immune activity and structure of *D. officinale* polysaccharides.

**Anti-Aging Activity**

The oral administration with a very high dose of 1 g/kg of *D. officinale* juice and a very high dose of 0.32 g/kg of *D. officinale* polysaccharide for 9 weeks exhibited an anti-aging effect in D-galactose-induced (0.125 g/kg) aging mice, supported by the significantly increased contents of SOD, glutathione peroxidase (GSH-Px) and total antioxidant capacity (T-AOC) in serum, as well as the enhanced SOD level in heart, liver, kidney, and cerebrum (Liang et al., 2017). Similar to this study, the treatment with *D. officinale* polysaccharide (DOP) at the doses of 50 and 100 mg/kg for 4 weeks exhibited more potent anti-fatigue activity than *Rhodiola rosea* extract as a positive control in BALB/c mice, as revealed by the increased TG (or fat) mobilization and the decreased lipid oxidation (LOD) and cell variability of T and B lymphocytes in the weight-loaded swimming test (Wei et al., 2017). However, the underlying mechanisms by which *D. officinale*’s anti-aging effects remain unclear. Additionally, the current pharmacological research lacks component analysis and clinical pharmacological experiments.

**Anti-Osteoporosis Activity**

A previous study revealed that the administration of *D. officinale* water extracts at very high doses of 150, 300, and 600 mg/kg for 13 weeks prevented ovariectomy (O VX)-induced bone loss in Wistar rats by decreasing the levels of TG, alkaline phosphatase (ALP), and bone mineral density (BMD) in comparison with Xian-Ling-Gu-Bao capsule (240 mg/kg) as a positive control (Wang et al., 2018). Meanwhile, *D. officinale* water extract treatments (10, 40, 80 μg/ml) were found to suppress receptor activator expression of the nuclear factor-kB ligand (RANKL)-induced osteoclastogenesis in RAW264.7 cells (Wang et al., 2018). In addition, *D. officinale* polysaccharide DOP treatments at very high doses of 200 or 400 μg/ml exhibited an anti-osteoporosis effect through the activation of NRF2 signaling, thereby attenuating adipogenic differentiation and promoting osteogenic differentiation in BMSCs (Peng et al., 2019). Notably, no reports about the anti-osteoporosis effect of small molecule compounds from *D. officinale* have been documented so far.

**Protective Effects on Liver and Kidney**

One study has demonstrated that the treatment with *D. officinale* ethanol extracts at the doses of 4.375 and 17.5 mg/kg for 9 weeks prevented liver and kidney damage in hyperuricemic rats by suppressing the protein levels of NF-κB and TLR4 compared with the model group (0.15% ademin, 10% yeast extract, and 89.85% standard diet) (Lou et al., 2020). Likewise, another study revealed that *D. officinale* flower water extracts (50, 100, 150, and 200 mg/kg) showed protective effects on alcohol-induced (10 ml/kg) liver injury by its anti-steatosis, anti-oxidative, and anti-inflammatory effects (Wu et al., 2020). However, additional evidence from randomized controlled trials is required to identify other regulatory mechanisms that may be responsible for the protective effects on liver and kidney. The bioactive constituents of these extracts also remain unknown.
Other Activities
In addition to the bioactivities described above, *D. officinale* was also found to have other therapeutic effects, such as neuroprotective effect, anti-photoaging effect, and pulmonary protective function. For instance, in hypoxic-ischemic brain damage (HIBD) neonatal rat model (vehicle group, normal saline = 10 ml/kg), the administration of aqueous extracts of *D. officinale* at the doses of 75, 150, and 300 mg/kg for 14 days suppressed the neuronal apoptosis by reducing cleaved caspase-3 and Bax, increasing Bcl-2, enhancing the expression of neurotrophic factors and K + –Cl–cotransporter 2 (KCC2), and decreasing the expression of hypoxia-inducible factor-1α (HIF-1α) and histone deacetylase 1 (HDAC1), leading to neuroprotective effects in neonatal rats against HIBD (Li and Hong, 2020b). *D. officinale* protocorm treatments at the doses of 10, 25, and 50 mg/ml exerted an anti-photoaging effect through decreasing erythema and protected skin from dryness by increasing CAT, SOD, and GSH-Px expression levels and decreasing thioharbitoric acid reactive substances (TBARS) and MMPs levels relative to the model group (UV irradiation) and positive control group (UV irradiation and a formulation of matrixyl) (Mai et al., 2019). In addition, *D. officinale* polysaccharides prevented lung injury by ameliorating cigarette smoke-induced mucus hypersecretion and viscosity by decreasing the expression of mucin-5AC (MUC5AC) mRNA and secretory protein in vitro and in vivo (Chen et al., 2020).

QUALITY CONTROL
The wild resources of *D. officinale* have gradually decreased, while the supply of artificially cultivated *D. officinale* has increased correspondingly. Consequently, wild resource collection has progressively become a non-mainstream, and the artificial cultivation mode occupies a dominant position in *D. officinale* industry (Ni et al., 2018). According to the description in the Editorial Board of Chinese Pharmacopoeia (2020) edition, the peak area ratio of mannose to glucose, moisture content, total ash content, ethanol extract, and polysaccharide content should reach 2.4–8.0%, less than 12.0%, more than 6.0, 6.5, and 25.0% of *D. officinale* stem, respectively (China Pharmacopoeia Committee, 2020). However, the quality of *D. officinale* may be affected by regions, tissues, harvest time, cultivation techniques, growth years, endophytes, and others which may disturb the long growth cycle (Cheng J et al., 2019). For instance, Li et al. (2017a) found that the contents of polysaccharides in *D. officinale* from Yunnan, Fujian, Jiangsu, and Zhejiang were variable (Li et al., 2017b). Among them, the highest content of polysaccharides was 54.42% in Zhejiang, followed by 43.26% in Fujian (Li et al., 2017a). Previous studies have revealed that the contents of polysaccharides in stems, leaves, and flowers of *D. officinale* are 34.61%, 23.51%, and 13.47%, respectively (Cao et al., 2018). It was also found that the polysaccharide content of *D. officinale* is gradually increased during the entire flower-opening process from buds to full bloom, in which the polysaccharide content is the highest in the full bloom stage (13.75%), followed by the micro bloom stage (11.52%). In contrast, the lowest content appears in the bud stage (9.50%) (Huang et al., 2017). In addition, *D. officinale* is usually planted on trees (fixed to the trunk with fine twine). Therefore, the polysaccharide content of *D. officinale* is significantly affected by different auxiliary tree species. Those planted on the evergreen tree *Phoebe zhennan* as the accessory hosts exhibit the highest polysaccharide content (37.8%) substantially different from those grown on the *Michelia ilsonii*, *Davidia nvolucrate* and *Taxus chinsens* var. *mairei* (Gu and Xie, 2021). Moreover, the polysaccharide content of 5-year-old *D. officinale* stems is the highest, followed by 3-year-old, 4-year-old, 2-year-old, and 1-year-old ones (Qin et al., 2017). Besides, endophytes play an important role in promoting the accumulation of polysaccharides. For example, DO14 (*Pestalotiopsis* sp.) isolated from *D. officinale* treated with 240 mg/L protein-polysaccharide fractions (PPF) can promote the accumulation of polysaccharides. This endophytic fungus could be used as biological fertilizer to improve the yield and quality of *D. officinale* (Zhu et al., 2018). In addition, exploiting suitable artificial-sheltered cultivation mode, screening the best cultivation substrate, and developing aseptic germination technology could be utilized to enhance the quality production of *D. officinale* in current agronomical practices (Cheng Y et al., 2019; Zuo et al., 2020).

It is well known that the polysaccharides’ bioactivities vary from different sources, production regions, and cultivation conditions of *D. officinale*. Moreover, it has been reported that the bioactivities of *D. officinale* polysaccharides are related to their chemical characteristics and advanced structures. However, *D. officinale* polysaccharides are macromolecular compounds with large molecular weight and complex structure, thereby generating a great challenge to implement analytical technologies (Ma et al., 2018). Therefore, it is necessary to establish a safe and effective quality assessment method for the quality control and clinical application of *D. officinale*. Herein, the qualitative and quantitative analytic methods of *D. officinale* polysaccharides are summarized and discussed.

There are several rapid and accurate methods for quantitative estimation of natural polysaccharides and their different fractions in *D. officinale* (Wu et al., 2018a). Generally, the large *D. officinale* polysaccharides are acid hydrolyzed into oligosaccharides, and then LC-MS is used to separate and characterize the products efficiently. The previous results indicate that the variations in the mass values of different peaks present structural differences of various metabolized products. Moreover, MS can be used to explore the oligosaccharide hydrolysates of *D. officinale* polysaccharides in detail from the aspects of identity, structure, and properties (Ma et al., 2018). An oligosaccharide-marker approach by labeling them with fluorescence reagent paminobenzoic acid ethyl ester (ABEE) (Te-Man-ABEE and Pen-Man-ABEE) was recently applied for quality assessment of *D. officinale* polysaccharides using UHPLC-QTOF-MS (Wong et al., 2019). The results revealed that the two oligosaccharide markers exhibited a satisfactory linearity relationship with *D. officinale* polysaccharides (R$^2$ ≥ 0.997) in the range of 0.68–16.02 μg. These markers also revealed satisfactory precision (relative standard deviation, RSD <7.0%) and recovery (91.41–118.30%) in unknown sample
determination. It is speculated that the oligosaccharide-marker method is a simple, rapid, and reliable approach for the qualitative and quantitative determination of specific polysaccharides from *D. officinale* and other herb formulas (Wong et al., 2019).

Although single-component quality assessment can be used to control the quality of natural Chinese herbal medicine (Wei et al., 2020), the chemical compositions of *D. officinale* are complex. *D. officinale* contains a lot of effective nutritional compositions, including flavonoids, crude fiber, amino acids, proteins, and fat. These active components of *D. officinale* are affected by tissues and harvest times. A previous study found that the contents of flavonoids in the stems, leaves, and flowers of *D. officinale* were 0.052%, 0.251%, and 1.835%, respectively (Li et al., 2019a). Another research reported that the content of total flavonoids in the flowers of *D. officinale* is the highest at the full flowering stage (1.66%) followed by the bract stage (1.52%) and micro flowering stage (1.41%) (Huang et al., 2017). It should be noted that the contents of five representative flavonoid glucosides from *D. officinale* in 25 batches with different sources were determined by UHPLC-ESI-MS/MS, where principal component analysis (PCA) and hierarchical cluster analysis (HCA) were applied (Ye et al., 2017). The content of crude fiber in the autumn stem is 59.7% higher than that in the spring stem, and the fiber in the autumn leaf is 122.9% higher than that in the spring leaf (Li et al., 2017a). The total protein content of *D. officinale* stems in autumn is 10.9% higher than that in spring, while that of the leaves in autumn is 9.3% higher than that in spring. The results also revealed that the total protein content in the leaves is higher than that in the stems, and the ratio of total protein in the autumn leaves is 67.03 mg/g, which is 31.4% higher than that in the autumn stems (51.02 mg/g). The content of fat in the stems and leaves of *D. officinale* ranges from 10.0 to 15.0 mg/g. The average content of fat in the leaves (13.90 mg/g) is slightly higher than that in the stems (11.50 mg/g) (Liao et al., 2018). In particular, Wu and Feng (2019) revealed that the content of polysaccharides is the highest in those samples collected from October to the following March (Wu and Feng, 2019). Yu et al. (2014) found that it is more reasonable to harvest *D. officinale* at biennials pre-bloom than at specific harvesting months according to the content of polysaccharides (Xu et al., 2021). According to the Editorial Board of Chinese Pharmacopoeia (2020) edition, however, *D. officinale* should be harvested from November to the following March (China Pharmacopoeia Committee, 2020).

Additionally, there are some noteworthy scientific gaps, which can be resolved from the following aspects. First, the genuine, defective, and counterfeit varieties of *D. officinale* are mixed together, so there is an urgent need to breed varieties with better agronomic characters, high yield, high quality, and intense stress resistance. (Guo et al., 2019). Second, it is also an effective way to ensure the quality of *D. officinale* by controlling pesticide residues and using endophytes to control diseases and pests (Yu et al., 2014; Zhou et al., 2018). Third, at present, chemical methods are primarily used to evaluate the quality of *D. officinale*, while the quality evaluation of traditional Chinese medicine is closely related to its biological activity. Therefore, the evaluation methods are suggested to be improved by combining chemical approaches and bioassays, providing a foundation for the industrial production and clinic use of *D. officinale* (Wei et al., 2020).

**SAFETY**

The stems and leaves of *D. officinale* were approved by National Health and Family Planning Commission (NHFPC) of People’s Republic (PR) of China to be utilized as a novel food material on Jan 15, 2013 and Jan 15, 2017 (National Health and Family Planning Commission of the People’s Republic of China, 2013; National Health and Family Planning Commission of the People’s Republic of China, 2017), respectively. Therefore, several safety studies of *D. officinale* have been performed. It has been reported that acute toxicity test (12.0 g/kg), genetic toxicity tests ( Ames test, micronucleus test of bone marrow, and sperm shape abnormality test in mice) (1,000, 2000, and 4,000 mg/kg), and 90-days feeding test (1.08, 1.67, and 5.00 g/kg) in rats were employed to assess the safety of the stems from *D. officinale*. These results indicated that *D. officinale* was a type of health food product without noticeable toxicity, genetic toxicity, and mutagenicity within the range of the test doses (Li Z. et al., 2019). Moreover, oral administration with *D. officinale* stems at the doses of 25, 1,250, and 2,500 mg/kg did not exhibit any apparent effect on pregnant rats or deformity effect on fetal rats (Qin et al., 2019). Likewise, oral administration with the leaves and flowers of *D. officinale* at the doses of 0, 2.0, 4.0, and 6.4 g/kg for 90 days did not exhibit apparent adverse effects on sperm quality and testicular tissue morphology in parent and offspring rats (Fu et al., 2017; Fu et al., 2020a). Besides, *D. officinale* flowers (0, 2.0, 6.4 g/kg) had no apparent adverse effects on pregnant and offspring rats before birth (Fu et al., 2020b).

*D. officinale* is considered to have edible and medicinal values. However, due to its thermal tonic property, it is prohibited for patients with wind-heat cold, dampness, and allergies, teenagers, and pregnant women. According to the Pharmacopoeia of the People’s Republic of China (China Pharmacopoeia Committee, 2020), the dosage of 6–12 g/d of *D. officinale* is appropriate, and intake of trace elements recommended by the Food and Drug Administration of the United States will not induce poisonous effects.

**CONCLUSION AND FUTURE PROSPECTS**

*D. officinale*, as a medicinal or food homologous product, plays a crucial role in healthcare. This study summarizes and updates the botany, traditional uses, bioactive components, pharmacology, quality control, and safety of *D. officinale*. Available data indicate that over 120 compounds have been isolated and identified from *D. officinale*, including polysaccharides, bifenzyls, flavonoids, alkaloids, phenanthenes, etc. *D. officinale* is associated with multiple beneficial pharmacological properties, such as cardioprotective, anti-tumor, gastrointestinal protective,
anti-diabetes, anti-aging, and anti-osteoporosis activities. Furthermore, it is evident that *D. officinale* is a non-toxic, which can be listed as a toxicologically safe functional food. However, its clinical applications has been rarely described, and critical improvements are still required for its industrial applications.

Firstly, phytochemical studies have demonstrated that *D. officinale* mainly contains polysaccharides, bibenzyls, phenanthrenes, and flavonoids while little is known about the analysis and function of organic compounds such as protein and fatty acid. Growing evidence has shown that structure-based drug design plays a vital role in developing novel drugs, and a series of strategies can be adopted to obtain effective therapeutic drugs. Further studies need to be conducted to isolate and identify more compounds from *D. officinale* with novel structures, emphasizing on bioactivity-guided, structurally modified, and chemically synthesized molecules. Besides, most of the *D. officinale* containing health products are mainly derived from its stems rich in chemical compounds, while non-medicinal parts are rarely exploited. Therefore, it may be interesting to extend the research to the non-medicinal parts of the inexpensive flowers, leaves, and roots of *D. officinale* to ensure the fully utilization of its edible and medicinal values (Zhang et al., 2017a).

Second, *D. officinale* is traditionally used to relieve fatigue, nourish “Yin”, heart, and stomach, and expel evil heat, which is closely related to its preventive effects on cardiac, gastrointestinal and diabetes diseases according to modern pharmacology. Pharmacological research of the cardioprotective, gastrointestinal tract and anti-diabetes protective effects of *D. officinale* mainly concentrates on its crude extracts and polysaccharides. However, the optimal dose, constituents, and side effects of *D. officinale* are not assessed. Besides, there is a lack of in-depth study on the mechanism of action of *D. officinale* monomeric compounds or the specific mechanism of *D. officinale* in animals is not comprehensive enough. Therefore, the high-quality and well-designed *in vivo, in vitro*, and clinical studies are encouraged to be carried out and to explore the molecular mechanisms and relationship between active chemical constituents and potential cardioprotective, gastrointestinal tract regulatory and anti-diabetes effects.

Third, the dosage of *D. officinale* used in previous studies were different (ranging from 4.375 mg/kg to 20 g/kg). It is not possible to define an exact upper cut-off dose, and the test dose needs to be pharmacologically relevant. In many cases, 100–200 mg/kg extracts for *in vivo* studies should be assumed as the upper limit for meaningful pharmacological studies. For pure compounds, a much lower dose range (e.g., 30–50 μM) should be considered for *in vitro* studies (Heinrich et al., 2020). In some cases, the pharmacological activity of *D. officinale* is present only with doses that might be too high for clinical use. According to the Chinese Pharmacopoeia, 6–12 g/day administration of *D. officinale* extract is common for human, but any clinical application at such doses should be accompanied with conservative safety insurance. Besides, the acute toxicity and sub-toxicity assessments of *D. officinale* were mostly carried out based on animal experiments. Furthermore, comprehensive placebo-controlled and double-blind clinical trials are necessary to provide sufficient evidence ensuring drug efficacy and patient safety.

Finally, *D. officinale* possesses various biological activities, which has been applied as health care medicine, health food, and health tea. In addition, with the development of analytical techniques and quality control methods, such as the improvement and update in chromatography techniques and molecular identification methods, new quality markers and quality control measures are likely to be adopted for better quality assessment of Chinese herbal medicine in the future (Leong et al., 2020).

In conclusion, *D. officinale* is one of the most popular medicinal and food homologous products in China. Modern pharmacology investigations have revealed its cardioprotective, gastrointestinal protective, anti-diabetes, and anti-aging effects, which robustly support its traditional application in nourishing “Yin”, heart, and stomach, expelling evil heat, and relieving fatigue. This paper provides a full-scale review about the progress of botany, traditional uses, phytochemistry, pharmacology, quality control, and toxicology of *D. officinale*. The information summarized in this work can provide a foundation for further applying the medicinal and edible value of *D. officinale* in the future.

**AUTHOR CONTRIBUTION**

WC collated documents and wrote the manuscript; JL and JZ helped to perform the arrangement of tables and pictures; JW and LY helped to check chemical structure formula; LQ and BZ contributed significantly to analysis and manuscript preparation. All authors have read and approved the final version of the manuscript.

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| Term | Definition |
|------|------------|
| D. officinale | *Dendrobium officinale* |
| HPLC | High performance liquid chromatography |
| HPAEC | High performance gel permeation chromatography |
| GC-MS | Gas chromatography-mass spectrometry |
| GPC | Gel permeation chromatography |
| HPLC-ESI-MS | High performance liquid chromatography electrospray ionization mass spectrometry |
| UHPLC | Ultra-high performance liquid chromatography |
| QTOF | Quadrupole time of flight mass spectrometry |
| ISO | Isoproterenol |
| ANP | Atrial natriuretic peptide |
| BNP | Brain natriuretic peptide |
| cTNI | Cardiac troponin I |
| LAD | Anterior descending coronary artery |
| CK | Creatine kinase |
| LDH | Lactate dehydrogenase |
| MDA | Malondialdehyde |
| SOD | Superoxide dismutase |
| H2O2 | Hydrogen peroxide |
| PI3K | Phosphatidylinositol 3-kinase |
| Akt | Protein kinase B |
| MAPK | Mitogen-activated protein kinase |
| LPD | Lipid peroxidation damage |
| ROS | Reactive oxygen species |
| HMEC-1 | Human microvascular endothelial cells |
| DOFP | *D. officinale* ultrafine powder |
| LPS | Lipopolysaccharide |
| TLR4 | Toll-like receptor 4 |
| TC | Total cholesterol |
| TG | Triglyceride |
| LDL-c | Low-density lipoprotein cholesterol |
| CRP | C-reactive protein |
| IL-6 | Interleukin 6 |
| MyD88 | Myeloid differentiation factor |
| TNF-α | Tumor necrosis factor alpha |
| HDL-c | High-density lipoprotein cholesterol |
| NO | Nitric oxide |
| ET-1 | Endothelin 1 |
| TXB2 | Thromboxane B2 |
| PGI2 | Prostacyclin |
| SCFA | Short-chain fatty acid |
| ACHSFD | Alcohol, and high sugar and fat diets |
| MNNG | Methyl-2-nitro-1-nitrosoguanidine |
| EMT | Epithelial-mesenchymal transition |
| HNSCC | Human head and neck squamous carcinoma cells |
| JNK | Jun N-terminal kinase |
| NPC | Nasopharyngeal carcinoma |
| TGF | Transforming growth factor |
| HCC | Hepatocellular carcinoma |
| SAR | Structure-activity relationship |
| AMPK | AMP-activated protein kinase |
| LC-3β | Light chain 3β |
| HO-1 | Heme oxygenase-1 |
| HFD/STZ | High-fat diet/streptozotocin |
| TLRs | Toll-like receptors |
| DMBG | Dimethylbiguanide |
| VEGF | Vascular endothelial growth factor |
| GLUT-1 | Glucose transporter 1 |
| CTGF | Connective tissue growth factor |
| HMEC-1 | Human microvascular endothelial cells |
| LPS | Lipopolysaccharide |
| T-AOC | Total antioxidant capacity |
| DOP | *D. officinale* polysaccharide |
| LOD | Lipid oxidation |
| OVX | Ovariectomy |
| ALP | Alkaline phosphatase |
| BGP | Bone glucose protein |
| ACP | Acid phosphatase |
| BMD | Bone mineral density |
| RANKL | Receptor activator expression of the nuclear factor-kB ligand |
| Nrf2 | Nuclear factor E2-related factor 2 |
| KCC2 | K + -Cl -cotransporter 2 |
| HIF-1α | Hypoxia-inducible factor-1α |
| HDAC1 | Histone deacetylase 1 |
| HIBD | Hypoxic-ischemic brain damage |
| TBARS | Thioarbituric acid reactive substances |
| MUC5AC | Mucin-5AC |
| ABEE | Aminobenzoic acid ethyl ester |
| RSD | Relative standard deviation |
| PCA | Principal component analysis |
| HC | Hierarchical cluster analysis |
| NHFPC | National health and family planning commission |
| PR | People’s republic |