BluePrint breast cancer molecular subtyping recognizes single and dual subtype tumors with implications for therapeutic guidance

Midas M. Kuilman1 · Architha Ellappalayam1 · Andrei Barcaru1 · Josien C. Haan1 · Rajith Bhaskaran1 · Diederik Wehkamp1 · Andrea R. Menicucci2 · William M. Audeh2 · Lorenza Mittempergher1 · Annuska M. Glas1

Received: 28 March 2022 / Accepted: 27 July 2022 / Published online: 19 August 2022
© The Author(s) 2022

Abstract
Purpose BluePrint (BP) is an 80-gene molecular subtyping test that classifies early-stage breast cancer (EBC) into Basal, Luminal, and HER2 subtypes. In most cases, breast tumors have one dominant subtype, representative of a single activated pathway. However, some tumors show a statistically equal representation of more than one subtype, referred to as dual subtype. This study aims to identify and examine dual subtype tumors by BP to understand their biology and possible implications for treatment guidance.

Methods The BP scores of over 15,000 tumor samples from EBC patients were analyzed, and the differences between the highest and the lowest scoring subtypes were calculated. Based upon the distribution of the differences between BP scores, a threshold was determined for each subtype to identify dual versus single subtypes.

Results Approximately 97% of samples had one single activated BluePrint molecular subtype, whereas ~3% of samples were classified as BP dual subtype. The most frequently occurring dual subtypes were the Luminal-Basal-type and Luminal-HER2-type. Luminal-Basal-type displays a distinct biology from the Luminal single type and Basal single type. Burstein’s classification of the single and dual Basal samples showed that the Luminal-Basal-type is mostly classified as ‘luminal androgen receptor’ and ‘mesenchymal’ subtypes, supporting molecular evidence of AR activation in the Luminal-Basal-type tumors. Tumors classified as Luminal-HER2-type resemble features of both Luminal-single-type and HER2-single-type. However, patients with dual Luminal-HER2-type have a lower pathological complete response after receiving HER2-targeted therapies in addition to chemotherapy in comparison with patients with a HER2-single-type.

Conclusion This study demonstrates that BP identifies tumors with two active functional pathways (dual subtype) with specific transcriptional characteristics and highlights the added value of distinguishing BP dual from single subtypes as evidenced by distinct treatment response rates.

Keywords Genomic testing · BluePrint · Breast cancer · Single and dual subtypes · Molecular subtypes

Abbreviations
BP BluePrint
ER Estrogen receptor

Midas M. Kuilman and Architha Ellappalayam have contributed equally as first authors.

Lorenza Mittempergher
lorenza.mittempergher@agendia.com

Annuska M. Glas
annuska.glas@agendia.com

1 Department of Research and Development, Agendia N.V, Radarweg 60, 1043 NT Amsterdam, The Netherlands
2 Department of Medical Affairs, Agendia Inc, 22 Morgan, Irvine, CA 92618, USA
**Introduction**

Breast cancer (BC) is a heterogenous disease with respect to clinical, histopathological, and molecular features. Based on clinical behavior and genomic characteristics, multiple methods have been utilized to categorize BC into distinct subgroups, be it with clinical subtyping for hormone receptor (HR) protein status or more recently with molecular subtyping based on RNA assays [1–4].

Clinical subtyping relies on well-established immuno-histochemistry (IHC) and fluorescence in situ hybridization (FISH) staining that determines estrogen receptor (ER), progesterone receptor (PR), and human epidermal growth factor receptor 2 (HER2) status [5, 6]. The BluePrint (BP) 80-gene subtyping assay was developed to bridge clinical pathology and molecular subtyping, by using IHC-based receptor status and mRNA expression, resulting in a molecular diagnostic array with predictive value [1, 7]. Each of the three subtypes determined by BP (Basal-type, Luminal-type, and HER2-type) is scored according to their respective gene signatures (consisting of 28, 58, and 4 genes, respectively) reflecting specific functional pathways, with the highest score determining the subtype [1, 7]. In most cases, the highest score is significantly higher than the score of the other two subtypes, indicating a strong dominance of a single pathway activation in the tumor (so-called single subtype). However, in rare instances, the difference between the highest score and the second-highest score is statistically indiscernible, indicating that these tumors might be characterized by multiple activated pathways (dual subtype). Having a deeper understanding of which pathways are activated may help understanding the specific biology of BP dual subtypes that distinguish them from the single subtypes.

In addition to the standard BP subtypes (Basal-type, Luminal-type, and HER2-type), other studies have identified expression-based subtypes, which include normal-like, claudin-low, triple positive, and triple-negative [3, 8–11] types. Among others, Burstein and colleagues further classified the clinical triple-negative breast cancer (TNBC) subtype into basal-like immuno-activated (BLIA), basal-like immuno-suppressed (BLIS), luminal androgen receptor (LAR), and mesenchymal-like (MES) [12]. Therefore, assessing further the differences between the BP scores may help identifying additional subtypes previously not detected by standard BP. Also, understanding the biological characteristics of BP dual subtypes may help in guiding more effective treatment plans.

**Materials and methods**

**Data**

For this study, only data and no samples were collected, and all patient data were fully anonymized according to the ‘General Data Protection Regulation’ (GDPR) and the ‘Health Insurance Portability and Accountability Act’ (HIPAA) and are in compliance with the ‘Data Protection Act.’ This study was a retrospective analysis of (internal) studies between 2015 and 2020. These studies included those previously described in Beumer et al. [13], the FLEX registry trial (NCT03053193), the Neoadjuvant Breast Registry Symphony Trial (NBRST) (NCT01479101), and the Multi-Institutional Neo-adjuvant Therapy MammaPrint Project I (MINT) trial (NCT01501487). Most samples comply with MammaPrint (MP) eligibility criteria [14, 15], stage I, II, or operable stage III breast cancer, tumor diameter ≤ 5 cm, and up to three positive lymph nodes, with any ER/PR/HER2 status. Microarray processing was performed following standard procedure at Agendia [7] (Supplementary methods). Agendia’s customized diagnostic arrays were either a targeted array or a full genome array, as previously described [7, 13, 16].

Of the 15,580 samples analyzed with BP, 7985 had full-genome expression data available of which 1978 with clinicopathological information (Table 1). All samples analyzed with the targeted array had clinicopathological information available (Table 1b).

The Neoadjuvant Breast Registry Symphony Trial (NBRST) [17–19] classified BC patients according to MP and BP and compared it with conventional IHC/FISH subtyping to predict treatment sensitivity. From the entire NBRST trial dataset (n = 1060), a subset that received HER2-targeted therapy (n = 289) was used to evaluate the association between the dual subtypes and response to HER2-targeted therapy. The NBRST trial protocol was approved by Institutional Review Boards at all participating sites (ClinicalTrials.gov NCT01479101). All patients consented to participation in the study and clinical data collection. Part of the anonymized data (BP results and IHC) used in this study was generated from early-stage BC patients collected from standard diagnostic testing and was only used to identify potential dual subtypes and not for any gene expression analysis. The data from studies can be shared by the authors upon reasonable request.

**BluePrint single and dual-subtype classification**

Standard BP scores of 15,580 samples were calculated followed by dual-subtype classification, which was based on bootstrap technique [20], and multi-modality detection. Details on the procedure can be found in the Supplementary methods and Figure S1 (Fig. S1a).

**Conventional subtype classification**

Clinicopathological information was available for 9573 of 15,580 samples, including IHC HR status for ER
and PR, Ki-67, and IHC/FISH HER2 status (Table S1). Tumors with at least 1% positivity for either ER or PR were classified HR-positive (HR+), otherwise HR-negative (HR−). Tumors with HER2 IHC 0, 1+ or 2+ (FISH non-amplified) score were considered HER2-negative (HER2−) while tumors with HER2 IHC 2+ (FISH amplified) and 3+ score were considered HER2-positive (HER2+).

**Burstein classification**

An algorithm published by Burstein et al., stratifies TNBCs into different subtypes by gene expression analyses of 80 signature genes. This algorithm was used to classify the Basal-single-type and Luminal-Basal-type samples into BLIA, BLIS, LAR, and MES [12].

**Software and statistics**

Gene expression analysis was performed on full-genome microarray data \( n = 7985 \) using limma \( \text{(v3.2)} \) [21]. Hallmark and Oncogenic gene sets from the Molecular Signatures Database \( \text{v7.2} \) were used for gene set enrichment analysis (GSEA) [22]. Genes were ranked based on the effect size ratio using the Cohen’s \( D \) effect size [23]. Differentially expressed genes (DEG) were considered significant with a \( p \) value \( \leq 0.05 \) and a \( \log_2 \) fold change \( \geq 1 \).

Computational analysis and visualization were performed using R \( \text{(v3.6.1)} \) [24]. Principal component analysis (PCA) was performed using the “prcomp” package \( \text{(v3.6.2)} \) and visualized using “ggplot” \( \text{(v3.3.2)} \) [26]. Unpaired, two-sample t-tests were used to measure if the means of ER, PR, and Ki-67 positivity were significantly different between single and dual subtypes. Chi-square test of Independence was used to test for differences of categorical variables within the Burstein classification (BLIA, BLIS, LAR, and MES) and a multivariate logistic regression analysis for response to therapy (pathological complete response, pCR) between single and dual subtypes. Molecular subtype classification algorithms were used from the “Genefu” package [27].
Results

BluePrint single and dual subtype classification

Molecular subtyping of patient tumors (n = 15,580) was performed at Agendia using the BP 80-gene assay as previously described [1, 7]. We applied the dual subtype classification method (see “Methods” for details) to assess the presence of multiple activated pathways. Most tumors were classified as single subtype (n = 15,087, 96.8%), followed by 449 (2.9%) tumors classified as dual subtype, and 44 (0.3%) tumors as triple subtype (Table 1). The most common dual subtypes in this dataset were the Luminal-Basal-type and the Luminal-HER2-type. These had sufficient numbers for downstream analyses while HER2-Basal-type and Luminal-HER2-Basal-type were not sufficient in size [28] and not further analyzed (Table 1a).

To note, only 1.9% of Luminal-type tumors were identified as dual subtype, whereas this was the case for 9.6% of the Basal-type and 23.8% of the HER2− type tumors. Since our dataset was largely HR+HER2− (Table S1), in order to estimate the dual subtype prevalence in the overall BC clinical population, we iteratively created subsets representing expected distributions of clinical subtypes (https://seer.cancer.gov/statfacts/html/breast-subtypes.html) [29] (70% HR-HER2−) and we detected 4.92% dual subtypes (95% CI 4.91–4.93) (Fig. S2).

Principal component analyses using BluePrint reveals similarities between subtypes

To understand the similarities between single and dual subtypes, we performed PCA based on the BP gene expression signatures (Fig. 1a–e). We observed a clear distinction of single subtypes shown in the first two principal components (Fig. 1a–c). Luminal-Basal-type cluster separately from both Basal-single-type (Fig. 1d) and Luminal-single-type (Fig. 1e), conversely, Luminal-HER2-type (Fig. 1e, f) are more closely related with both Luminal-single-type (Fig. 1b) and HER2-single type (Fig. 1c).

Differential gene expression analysis highlights differences between BluePrint dual and single subtypes

Differential expression analysis using full-genome data (n = 7985) was performed to evaluate global transcriptional differences between single and dual subtypes. As expected from the PCA, when compared with their corresponding single subtypes, more DEGs were found for the Luminal-Basal-type (446 DEGs) (Fig. 2a, b) than for the Luminal-HER2-type (151 DEGs) (Fig. 2e, f).

Among the up-regulated genes in Luminal-Basal-type (vs. both Basal-single-type and Luminal-single-type) were present MUCLI, a known tumor suppressor gene [30], and CLCA2, a negative regulator of cancer cell migration and invasion [31].

Among the most up-regulated genes in Luminal-HER2-type compared with Luminal-single-type tumors, we found GRB7, TCAP, and ERBB2 which belong to the HER2 amplicom and are known to be overexpressed in pathologically confirmed HER2 tumors [32]. Indeed, these genes were also up-regulated in HER2-single-type tumors (Fig. 2e). When comparing Luminal-HER2-type with HER2-single-type, ESR1 was found to be upregulated, similarly as in Luminal-single-type tumors. Additionally, Luminal-HER2-type tumors were mainly classified as either Luminal B (n = 34/99, 34%) or HER2 enriched (n = 44/99, 44%) using the intrinsic subtype classifier of the “Genefu” [27, 33]. Together, these data suggest that both ER and HER2 are activated in Luminal-HER2-type tumors.

Differences between BluePrint single and dual subtypes may impact therapy response pathways

A better understanding of the underlying biological characteristics of the dual subtypes may come from analyzing gene pathway regulation.

Comparison of Luminal-Basal-type with Basal-single-type revealed upregulation of two estrogen response (ESR) and one androgen response (AR)-related gene sets (Fig. 2c). Same ESR gene sets were downregulated in Luminal-Basal-type versus Luminal-single-type, indicating that Luminal-Basal-type has intermediate ER levels. Conversely, AR was upregulated in Luminal-Basal-type, versus both the Basal-single-type and Luminal-single-type. G2M and E2F pathways [34, 35] were either downregulated or upregulated in Luminal-Basal-type compared with Basal-single-type and Luminal-single-type, respectively, indicating that Luminal-Basal-type are less proliferative than Basal-single-type, but more proliferative than Luminal-single-type tumors. Taken together, Luminal-Basal-type tumors show a distinct biology from their single counterparts with decreased proliferation than Basal-single-type and AR activation.

Compared with single HER2-single-type tumors, a Luminal-HER2 type shows downregulation of MAPK (MEK and RAF) signaling pathways and ER activation (Fig. 2g). Clinical characteristics of the single and dual BP subtypes and their response to therapy may confirm these hypotheses and provide additional insights.
Fig. 1 Principle component analysis using the three BluePrint signature gene sets (Basal-type, \( N = 28 \), panels a and d; Luminal-type, \( N = 58 \), panels b and e; HER2-type, \( N = 4 \), panels c and f). Clustering of Basal-single-type, Luminal-single-type, and HER2-single-type samples based on BluePrint signature genes. a–c shows coloring of single subtype samples (blue, Luminal-single-type; green, HER2-single-type; red, Basal-single-type) whereas the dual subtype samples are colored grey. d–f shows this in reverse where the dual subtype samples are colored (yellow, Luminal-HER2-type; pink, Luminal-Basal-type) and the single subtypes are shown in grey. The ellipses reported in each subfigure illustrate the 90% confidence intervals of the single- and dual subtype samples.
Fig. 2  Differential gene expression analysis between BluePrint single and dual subtypes. The x-axis and y-axis report the Log2 fold change and the FDR-adjusted p-values (−Log10(FDR)), respectively. Number of tumor samples used for the analysis are shown in between brackets in titles. Significance thresholds of ≤ 0.05 FDR and a log2 fold change of ≥ 1 were used. Red and blue dots illustrate significant differentially expressed genes. The strongest differentially expressed genes are labeled (abs(logFC) ≥ 2 or −Log10 adj p-value ≥ 50). Differentially expressed genes are identified in the following comparisons: a Luminal-Basal-type versus Basal-single-type, b Luminal-Basal-type versus Luminal-single-type, c Luminal-Basal-type versus Basal-single-type, d Luminal-Basal-type versus Luminal-single-type, e Luminal-HER2-type versus HER2-single-type, and f Luminal-HER2-type versus Luminal-single-type. Similarly, differentially expressed pathways are shown between c Luminal-Basal-type versus Basal-single-type, d Luminal-Basal-type versus Luminal-single-type, g Luminal-HER2-type versus HER2-single-type, and h Luminal-HER2-type versus Luminal-single-type. FDR = false discovery rate, UP = upregulated, DN = downregulated.
BluePrint dual subtypes present clear clinicopathological differences from single subtypes

Standard BP Luminal-, HER2−, and Basal-type tumors were further stratified using the single–dual subtyping classification (Fig. 3a). Additionally, conventional clinical subtypes (based on IHC HR staining (ER, PR) and HER2 status) were further classified into BP single subtypes or dual subtypes (Fig. 3b, c).

Majority of HR-HER2− tumors were classified as Basal-single-type ($n=150/176$) (Fig. 3b), while only 26 were dual subtype of which 22 were Luminal-Basal-type.

Most of the HR+ HER2− tumors were classified as Luminal-single-type ($n=4285/4548$), but interestingly, 3% ($n=152/4548$) was classified as Basal-single-type, which corresponds to more than half of all Basal-single-types identified by BP ($n=152/265$) (Fig. 3b). Of the HR+ HER2− with a dual subtype, majority was Luminal-Basal-type ($n=57/86$) (Fig. 3c).

Most HR+ HER2+ tumors were classified as either Luminal-single-type ($n=165/272$) or as HER2-single-type ($n=61/272$) (Fig. 3b) with the most frequent dual subtype being the Luminal-HER2-type ($n=30/34$) (Fig. 3c).

Luminal-single-type tumors had the highest IHC ER expression levels with the lowest levels observed in Basal-single-type tumors (Fig. 3d). Dual subtypes showed intermediate ER expression, compared to their single counterparts (Fig. 3d). ER low positive tumors (1–10% IHC) were mostly found in the Basal-single-type ($n=62/147, 42\%$) and in the Luminal-single-type ($n=58/147, 39\%$) (Fig. 3d). However,

Fig. 3  a Sankey plot showing the further stratification of the standard BluePrint (BP) Basal, Luminal, and HER2 subtypes with full-genome microarray data available, into the BP single and dual subtypes. b Sankey plot illustrating the re-classification of clinical-based subtypes (based on hormone receptors (HR) and human epidermal growth factor receptor 2 (HER2) status) to BP-based single-type molecular subtypes (Basal-single-type, Luminal-single-type, HER2-single-type). c Further stratification of the same clinical-based subtypes as in (b) to the BP-based dual subtypes (Luminal-HER2-type, Luminal-Basal-type, HER2-Basal-type, and Luminal-HER2-Basal-type). d, e Boxplots reporting for each single and dual subtype category (x-axis), the level and spread of estrogen receptor and Ki67 positivity based on Immunohistochemistry assessment (y-axis). Significant differential positivity between ER and Ki67 was assumed at a $p$-value $<0.05$ determined with a t-test between subtype categories. To note, for 4511 of the 9573 tumor samples with clinical annotation, HR and HER2 status were not available (Table S1)
considering the differences in sample size of the subtypes, a larger fraction of Basal-single-type (24%) was found to be ER low positive, compared with other subtypes. Proliferation measured by % Ki-67 positivity was significantly higher in Luminal-Basal-type and Luminal-HER2-type compared with Luminal-single-type, but significantly lower than Basal-single-type and HER2-single-type (Fig. 3e). Indeed, there were significantly more Luminal-Basal-type (n = 34/47, 72.3%; p value < 0.001) than Basal-single-type tumors (59/173, 34.1%) with Ki67 < 30%, threshold recently proposed for the so-called TNBC low proliferation (TNLP) tumors [36] (Fig. 3e).

Burstein LAR and MES subtypes are identified using BluePrint dual subtype classification

Since Luminal-Basal-type displays different transcriptional characteristics than Luminal-single-type and Basal-single-type, we classified them using the Burstein classifier to better understand their biology. Indeed, we found a significant association between BP single/dual subtypes and the Burstein BLIA, BLIS, LAR, and MES subtypes [12] (p value < 0.001) with the Basal-single-type classified mostly as BLIA or BLIS, whereas the Luminal-Basal-type as LAR or MES (Fig. 4a), irrespective of their standard BP subtype (Fig. 4b).

When using the PAM50 [3, 33] intrinsic subtype classifier of the "Genefu" [27] package, the Luminal-Basal-type tumors were mostly classified as HER2 enriched (HER2-e) (Table S3a). When comparing "Luminal-Basal/HER2-e" against "Luminal-Basal/ non-HER2-e", common biomarkers for HER2 molecular classification were not differentially expressed (Table S3b). Indeed, ~98% of Luminal-Basal-type tumors were clinically HER2− (Fig. 3c).

BluePrint dual subtype classification of the NBRST dataset shows refined prediction to therapy

Our findings indicate that the Luminal-HER2-type shares clinical and genomic features with Luminal-single-type and HER2-single-type and previous studies suggest that HR and HER2 co-expression is associated with endocrine and HER2-targeted therapy resistance [37, 38]. Therefore, to better understand how Luminal-HER2-type relates to HER2-targeted therapy response, we analyzed the NBRST dataset (see Methods for details) [18] and selected only pathologically confirmed HER2+ tumors (n = 289) with gene expression and HER2-targeted therapy response data available [either Trastuzumab (T) only or with Pertuzumab (P)]. Patient tumors were stratified using the BluePrint dual subtype classification (Fig. 5).

BP HER2-single-type showed higher pCR rate to chemotherapy (C) + T compared to Luminal-HER2-type (61.3% vs. 23.8%, p = 0.032) (Fig. 5a). Although not significant (also due to lower numerosity of Luminal-HER2-type), this trend remained for patients that received additional P (Fig. 5b). Response rates of Luminal-HER2-type tumors was higher, but not significantly different than for Luminal-single-type. Instead, a significant higher response rate was observed for receptor (LAR), or Mesenchymal (MES).

![Fig. 4 BluePrint (BP) dual subtype classification compared with Burstein’s classification of triple-negative breast cancer tumors [12]. a The inner circle contains percentages of the BP Basal-single-type and BP Luminal-Basal-type. The outer circle illustrates the corresponding Burstein classification into Basal-like immuno-activated (BLIA), Basal-like immuno-suppressed (BLIS), Luminal androgen](image-url)
the HER2-single-type compared to the Luminal-HER2-type, after correcting for HR status, tumor stage, tumor grade, and therapy in a multivariate logistic regression analysis (p value = 0.006, Table S2).

Discussion

Molecular subtyping using the standard BP 80-gene assay enables to discern the tumor subtype by the underlying functional pathways and not merely by HR and HER2 status [1, 7]. In most cases, the assay identifies a single, dominant activated pathway distinctive of a Luminal-, Basal-, or HER2-type tumor. This information often confirms the pathologically defined subtype but in many cases further classifies tumors from their initial clinical subtype into a different molecular subtype. This phenomenon has clinical implications for the treatment of patients, perhaps most notably in the ER+/Basal and HER2+/Luminal-HER2-Basal subtypes which have been previously described [18, 19, 39, 40].

The vast majority of the breast cancer tumors analyzed in this study using the BP test show a single activated pathway (i.e., single BP subtype) (97%); however, less frequently, they exhibit multiple activated pathways (i.e., dual or triple subtype) (3%), as we showed in a preliminary analysis [41]. Notably, this dataset mostly reflects a HR + population but upon sampling the data based on observed frequencies of clinical subtypes, such a percentage raises to ~5%. Importantly, the single and dual assessment performed on the NBRST dataset and also reported for the TRAIN2 [42] and APHINITY [43] patient cohorts show a higher number of dual subtypes, ranging from 11 to 30%, indicating that the dual subtype classification might have a greater clinical impact on a HER2+ population and that the potential clinical utility should be found in specific subgroups rather than in the entire EBC population. The analysis on the NBRST dataset was performed on limited numbers of dual subtypes (n = 32); however, the size was sufficient to generate statistically powerful results.

Overall, in this manuscript, we aimed to provide a better understanding of the biological diversity of EBC and these results should be taken with caution with respect to any immediate change in clinical management.

Next, by analyzing whole-transcriptomic data, we set out to understand if and how dual subtypes were distinct from single subtypes. For the analysis, we focused on the...
Luminal-Basal-type and Luminal-HER2-type tumors as the other dual subtypes were limited in size.

Neither the Basal nor the Luminal BP template genes were able to fully capture the biology of the Luminal-Basal-type tumors. The majority of tumors expressing typical Basal gene patterns are TNBC by pathology [44], and it is known that there is a large overlap between BP Basal subtypes and TNBCs. Therefore, we applied the TNBC Burstein classifier on the Basal-single-type and Luminal-Basal-type. Basal-single-type tumors were mostly classified as BLIA and BLIS while Luminal-Basal-type tumors were more likely to be either LAR or MES. Genes described by Burstein et al. to be up-regulated in the LAR subtype, such as DHRS2, AGR2, FOXA1, AR, and MUCL1, were indeed higher expressed in Luminal-Basal-type compared with the Basal-single-type samples. Since the majority of Luminal-Basal-type tumors were classified as LAR, and according to Burstein et al., those patients derive benefit from traditional anti-estrogen or anti-androgen therapy, we could speculate that Luminal-Basal-type cancers would benefit from such treatment as well. Furthermore, ADH1B and FABP4 genes were up-regulated in Luminal-Basal-type samples compared with Basal-single-type samples. The upregulation of these genes is typical of the MES subtype, which is characterized by the dysregulation of cell cycle and DNA damage repair pathways. On the contrary, BLIS subtype-specific genes, HORMAD1, SOX10, SERPINB5, and FOXC1, were up-regulated in Basal-single-type samples compared with Luminal-Basal-type samples. Therefore, we could hypothesize that among the Basal-single-type samples, two subgroups are present which are indiscernible with the current dual subtype classification, but might have a different prognosis according to Burstein et al. and require additional analyses. Notably, majority of the Luminal-Basal-type showed a Ki67 positivity below 30% which might indicate that they share features with the TNLP tumors recently described by Bhargava and colleagues [36]. Additionally, no large agreement was found between any of the dual subtypes and the normal-like [3, 4] (Table S3) or claudin-low classifications [9, 27] (data not shown). Conversely, BluePrint Basal-, Luminal-, and HER2-single type classifications were largely concordant with the intrinsic subtypes (> 90%) (see Table S3). Interestingly, and perhaps unexpectedly, the Luminal-Basal-type tumors were mostly classified as HER2-e intrinsic subtype, possibly due to the absence of Luminal- and Basal-type biology in the BP Luminal-Basal-type.

Luminal-HER2-type samples consistently showed patterns of both ER and HER2 activation (by expression and IHC/FISH), which may suggest similarities to the clinically triple-positive tumors [10]. Expression of both ER and HER2 may lead to receptor crosstalk which has often been associated with resistance to both endocrine and HER2-targeted therapies [45]. However, down-regulation of the MAPK-related gene sets MEK and RAF may indicate no downstream activation of the HER2 pathway. Therefore, Luminal-HER2-type tumors are unlikely fueled through the HER2 pathway alone and HER2-targeted therapies might not be as effective as in the HER2-single-type tumors. This suggestion is strengthened by the observation in the NBRST data that Luminal-HER2-type tumors have a significantly lower pCR rate to neoadjuvant chemotherapy including HER2-targeted agents compared with HER2-single-type tumors (p-value < 0.032). This is supported by preliminary subanalysis of the TRAIN2 [42, 46] and APHINITY [43, 47, 48] trial datasets, suggesting that BluePrint HER2-single-type tumors derive the most benefit from HER2 dual-targeted treatment [43].

It has been suggested that clinically triple-positive tumors develop endocrine resistance as downstream-activated MAPK inhibits ER transcription and phosphorylates ER [38]; however, in this study, Luminal-HER2-type tumors may be only driven by the ER pathway, as MAPK is down-regulated compared with HER2-single-type tumors and not significantly different from that of Luminal-single-type tumors. Further analysis on Luminal-HER2-type samples treated with endocrine therapy is required to investigate and confirm this hypothesis.

Conclusion

Our study showed that by further dissecting the BP scores, it is possible to identify a small proportion of EBCs that have dual-activated BP pathways. These dual subtypes display specific transcriptional and clinicopathological features supporting the idea that they represent a different biological subgroup than their single counterparts. Most dual BP subtypes are either Luminal-Basal-type or Luminal-HER2-type.

The Luminal-Basal-type shows lower proliferation levels compared with the Basal-single-type and AR activation. Interestingly, using the Burstein classification, Luminal-Basal tumors are mostly classified as LAR and MES subtypes.

The Luminal-HER2-type resembles features of both the Luminal-single-type and HER2-single-type. However, patients with Luminal-HER2-type tumors have a lower pCR rate after receiving HER2-targeted therapies in addition to chemotherapy compared with patients with a HER2-single-type.

Taken together, BP dual classification shows potential clinical utility in helping treatment decision for a limited, but still relevant, fraction of EBC patients with dual subtypes that may benefit from additional or alternative targeted therapies. Even though molecular subtyping is not yet standardly used in routine clinical diagnostics, increasing number of evidences are emerging indicating that...
molecular subtypes should become part of breast cancer management [49]. In this light, results presented here further support the need toward such transition and implementation.

Future work will be focused on further confirming and prospectively validating the findings described here in additional independent datasets.

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1007/s10549-022-06698-x.

Acknowledgements We are indebted to Patricia Dauer and Jeff Fens-ster for their thorough review of the paper.

Author’s contributions MMK analyzed the data, interpreted and visualized the results, and wrote the manuscript. AE collected and analyzed the data, interpreted the results, and wrote the manuscript. AB analyzed the data, interpreted the results, and reviewed the manuscript. JCH interpreted the results and reviewed the manuscript. RB collected the data and reviewed the manuscript. DW designed the research questions. ARM collected the data and reviewed the manuscript. WMA interpreted the results and reviewed the manuscript. LM designed and supervised the research study, interpreted the results, and wrote the manuscript. AG designed and supervised the research study, interpreted the results, and reviewed the manuscript.

Funding This research did not receive any special grant from funding agencies in the public, commercial, or for not-for-profit sectors. NBRST trial was funded by Agendia Inc.

Declarations

Conflict of interest All authors (MMK, AE, AB, JCH, RB, DW, ACM, WMA, LM, and AMG) are non-commercial employees of Agendia, the commercial entity that markets the 80-gene signature as BluePrint. AMG is named inventor on the patent for the 80-gene signature used in this study. No writing assistance was utilized in the production of this manuscript.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

References

1. Krijgsman O, Roepman P, Zwart W et al (2012) A diagnostic gene profile for molecular subtyping of breast cancer associated with treatment response. Breast Cancer Res Treat 133:37–47. https://doi.org/10.1007/s10549-011-1683-z

2. Walldeen B, Storhoff J, Nielsen T et al (2015) Development and verification of the PAM50-based Prosigna breast cancer gene signature assay. BMC Med Genomics 8:1–14. https://doi.org/10.1186/s12920-015-0129-6

3. Perou CM, Sorlie T, Eisen MB et al (2000) Molecular portraits of human breast tumours. Nature 406:747–752. https://doi.org/10.1038/35021093

4. Sorlie T, Perou CM, Tibshirani R et al (2001) Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. Proc Natl Acad Sci USA 98:10869–10874. https://doi.org/10.1073/pnas.191367098

5. Press O, Guzman R, Cervantes M et al (2014) Characterization of HER2 status by fluorescence in situ hybridization (FISH) and immunohistochemistry (IHC). Springer, New York

6. Badve SS, Baehner FL, Gray RP et al (2008) Estrogen- and progesterone-receptor status in ECOG 2197: comparison of immunohistochemistry by local and central laboratories and quantitative reverse transcription polymerase chain reaction by central laboratory. J Clin Oncol 26:2473–2481. https://doi.org/10.1200/JCO.2007.13.6424

7. Mittempenger L, Delahaye LJ, Witteveen AT et al (2020) Performance characteristics of the BluePrint® breast cancer diagnostic test. Transl Oncol 13:100756. https://doi.org/10.1016/j.tranon.2020.100756

8. Dai X, Li T, Bai Z et al (2015) Breast cancer intrinsic subtype classification, clinical use and future trends. Am J Cancer Res 5:2929–2943

9. Prat A, Guo H, Ganesan P et al (2010) Phenotypic and molecular characterization of the claudin-low intrinsic subtype of breast cancer. Breast Cancer Res 12:68–86. https://doi.org/10.1186/bcr2635

10. Vici P, Pizzuti L, Natoli C et al (2015) Triple positive breast cancer: a distinct subtype? Cancer Treat Rev 41:69–76. https://doi.org/10.1016/j.ctrv.2014.12.005

11. Prat A, Adamo B, Cheang MCU et al (2013) Molecular characterization of basal-like and non-basal-like triple-negative breast cancer. Oncologist 18:123–133. https://doi.org/10.1634/theoncologist.2012-0397

12. Burstein MD, Tsimelzon A, Poage GM et al (2015) Comprehensive genomic analysis identifies novel subtypes and targets of triple-negative breast cancer. Clin Cancer Res 21:1688–1698. https://doi.org/10.1158/1078-0432.CCR-14-0432

13. Beumer I, Witteveen A, Delahaye L et al (2016) Equivalence of MammaPrint array types in clinical trials and diagnostics. Breast Cancer Res Treat 156:279–287. https://doi.org/10.1007/s10549-016-3764-5

14. Cardoso F, VeerBogaerts LJ et al (2016) 70-gene signature as an aid for treatment decisions in early breast cancer. N Engl J Med 375:717–729. https://doi.org/10.1056/NEJMoa1602253

15. Piccart M, van’t Veer LJ, Pouncer C et al (2021) 70-gene signature as an aid for treatment decisions in early breast cancer: updated results of the phase 3 randomised MINDACT trial with an exploratory analysis by age. Lancet Oncol 22:476–488. https://doi.org/10.1016/S1470-2045(21)00007-3

16. Glas AM, Floore A, Delahaye LJMJ et al (2006) Converting a breast cancer microarray signature into a high-throughput diagnostic test. BMC Genomics 7:1–10. https://doi.org/10.1186/1471-2164-7-278

17. Whitworth P, Beitsch P, Mislowsky A et al (2017) Chemosensitivity and endocrine sensitivity in clinical luminal breast cancer patients in the prospective neoadjuvant breast registry symphony trial (NBRST) predicted by molecular subtyping. Ann Surg Oncol 24:669–675. https://doi.org/10.1245/s10434-016-5600-x

18. Whitworth P, Stork-Sloots L, de Snoo FA et al (2014) Chemosensitivity predicted by BluePrint 80-gene functional subtype and MammaPrint in the prospective neoadjuvant breast registry
symphony trial (NBRST). Ann Surg Oncol 21:3261–3267. https://doi.org/10.1245/s10434-014-3908-y

19. Whitworth P, Pellicane J V, Baron P, et al (2021) 5-year outcomes in the NBRST trial: preoperative MammaPrint and BluePrint breast cancer subtype is associated with neoadjuvant treatment response and survival. San Antonio Breast Cancer Symposium

20. Efron BTR (1993) An introduction to the Bootstrap Boca Raton. Chapman & Hall/CRC

21. Ritchie ME, Shipman B, Wu D et al (2015) Limma powers differential expression analyses for RNA-sequencing and microarray studies. Nucleic Acids Res 43:e47. https://doi.org/10.1093/nar/gk007

22. Subramanian A, Tamayo P, Mootha VK et al (2005) Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. Proc Natl Acad Sci U S A 102:15545–15550. https://doi.org/10.1073/pnas.0506580102

23. Cohen J (1998) Statistical power analysis for the behavioral sciences, 2nd Edn. Routledge, New York

24. R Core Team (2019) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria

25. Sigg CD, Buhmann JM (2008) Expectation-maximization for sparse and non-negative PCA. In: Proceedings of 25th international conference on machine learning, pp 960–967. https://doi.org/10.1145/1390156.1390277

26. Hadley Wickham (2016) ggplot2: Elegant graphics for data analysis. https://ggplot2.tidyverse.org. Springer, New York

27. Gendoo DMA, Ratanasirigulchai N, Schröder MS et al (2016) Genefu: an R/Bioconductor package for computation of gene expression-based signatures in breast cancer. Bioinformatics 32:1097–1099. https://doi.org/10.1093/bioinformatics/btv693

28. Wei C, Li J, Bumgarner RE (2004) Sample size for detecting differentially expressed genes in microarray experiments. BMC Genomics 5:1–10. https://doi.org/10.1186/1471-2164-5-87

29. Harbeck N, Renault-Llorca F, Cortes J et al (2019) Breast cancer. Nat Rev Dis Prim. https://doi.org/10.1038/s41572-019-0111-2

30. Conley SJ, Bosco EE, Tice DA et al (2016) HER2 drives Mucin-like 1 to control proliferation in breast cancer cells. Oncogene 35:4225–4234. https://doi.org/10.1038/onc.2015.487

31. Sasaki Y, Koyama R, Maruyama R et al (2012) CLCA2, a target of the p53 family, negatively regulates cancer cell migration and invasion. Cancer Biol Ther 13:1512–1521. https://doi.org/10.1089/cbt.2012.0239

32. Nadler Y, González AM, Camp RL et al (2009) Growth factor receptor-bound protein-7 (Grb7) as a prognostic marker and therapeutic target in breast cancer. Ann Oncol 20:1577–1586. https://doi.org/10.1093/annonc/mdp346

33. Parker JS, Mullins M, Cheang MCU et al (2009) Supervised risk predictor of breast cancer based on intrinsic subtypes. J Clin Oncol. https://doi.org/10.1001/jco.2008.18.1370

34. Oshi M, Takahashi H, Tokumaru Y et al (2020) G2m cell cycle pathway score as a prognostic biomarker of metastasis in estrogen receptor (Er)-positive breast cancer. Int J Mol Sci. https://doi.org/10.3390/ijms2120282921

35. Hollern DP, Swiatnicki MR, Remhink JP et al (2019) E2F1 drives breast cancer metastasis by regulating the target gene FGFI13 and altering cell migration. Sci Rep 9:1–13. https://doi.org/10.1038/s41598-019-47218-0

36. Srivastava P, Wang T, Clark BZ, et al (2022) Clinical-pathologic characteristics and response to neoadjuvant chemotherapy in triple-negative low Ki-67 proliferation (TNLP) breast cancers. NPJ Breast Cancer. https://doi.org/10.1038/s41523-022-00415-z

37. Loubser G, Collignon J, Jerusalem G (2016) Resistance to therapy in estrogen receptor positive and human epidermal growth factor 2 positive breast cancers: progress with latest therapeutic strategies. Ther Adv Med Oncol 8(6):429–449. https://doi.org/10.1177/1758834016665077

38. Schiff R, Massarweh SA, Shou J et al (2004) Cross-talk between estrogen receptor and growth factor pathways as a molecular target for overcoming endocrine resistance. Clin Cancer Res 10:331S–336. https://doi.org/10.1158/1078-0432.ccr-031212

39. Groenendijk FH, Treece T, Yoder E, et al (2019) Estrogen receptor variants in ER-positive basal-type breast cancers responding to therapy like ER-negative breast cancers. NPJ Breast Cancer. https://doi.org/10.1038/s41523-019-0109-7

40. Beitsch P, Whitworth P, Baron P et al (2017) Pertuzumab/trastuzumab/CT versus trastuzumab/CT therapy for HER2+ breast cancer: results from the prospective neoadjuvant breast registry symphony trial (NBRST). Ann Surg Oncol 24:2539–2546. https://doi.org/10.1007/s10434-017-5863-x

41. Kuilman* M, Ellappalamay* A, Mittempergher L, et al (2020) BluePrint molecular subtyping recognizes single and dual subtype tumors with implications for therapeutic guidance. European Breast Cancer Conference

42. Liefaard MC, Voort A Van Der, Sanders J, et al (2021) Effect of pertuzumab plus neoadjuvant trastuzumab-based chemotherapy in early-stage HER2-positive breast cancer according to BluePrint molecularly defined breast cancer subtypes. San Antonio Breast Cancer Symposium

43. Krop IE, Mittempergher L, Paulson JN, et al (2020) Abstract PD3-01: BluePrint performance in predicting pertuzumab benefit in genomically HER2-positive patients: a biomarker analysis of the APHINITY trial. San Antonio Breast Cancer Symposium, San Antonio

44. Badve S, Dabbs DJ, Schnitt SJ et al (2011) Basal-like and triple-negative breast cancers: a critical review with an emphasis on the implications for pathologists and oncologists. Mod Pathol 24:157–167. https://doi.org/10.1038/modpathol.2010.200

45. Giuliano M, Trivedi MV, Schiff R (2013) Bidirectional crosstalk between the estrogen receptor and human epidermal growth factor receptor 2 signaling pathways in breast cancer: molecular basis and clinical implications. Breast Care 8:256–262. https://doi.org/10.1159/000354253

46. van Ramshorst MS, van der Voort A, van Werkhoven ED et al (2021) Effect of dual HER2 blockade for HER2-positive breast cancer (TRAIN-2): a multicentre, open-label, randomised, phase 3 trial. Lancet Oncol 19:1630–1640. https://doi.org/10.1016/S1470-2045(18)30570-9

47. von Minckwitz G, Procter M, de Azambuja E et al (2021) Neoadjuvant chemotherapy with or without anthracyclines in the presence of dual HER2 blockade for HER2-positive breast cancer (TRAIN-2): a multicentre, open-label, randomised, phase 3 trial. Lancet Oncol 19:1630–1640. https://doi.org/10.1016/S1470-2045(18)30570-9

48. van Ramshorst MS, van der Voort A, van Werkhoven ED et al (2021) Effect of dual HER2 blockade for HER2-positive breast cancer (TRAIN-2): a multicentre, open-label, randomised, phase 3 trial. Lancet Oncol 19:1630–1640. https://doi.org/10.1016/S1470-2045(18)30570-9

49. Prat A, Chaudhury A, Solovieff N et al (2021) Correlative biomarker analysis of the APHINITY trial. San Antonio Breast Cancer Symposium, San Antonio

50. Badve S, Dabbs DJ, Schnitt SJ et al (2011) Basal-like and triple-negative breast cancers: a critical review with an emphasis on the implications for pathologists and oncologists. Mod Pathol 24:157–167. https://doi.org/10.1038/modpathol.2010.200

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.