H19 gene polymorphisms and Wilms tumor risk in Chinese children: a four-center case-control study

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Wenya Li
Zhengzhou University First Affiliated Hospital

ORCiD: https://orcid.org/0000-0001-6638-8675

Rui-Xi Hua
Sun Yat-sen University First Affiliated Hospital

Mi Wang
Guangzhou Women and Children's Medical Center

Da Zhang
Zhengzhou University First Affiliated Hospital

Jinhong Zhu
Guangzhou Women and Children's Medical Center

Songyang Zhang
Zhengzhou University First Affiliated Hospital

Yang Yang
Zhengzhou University First Affiliated Hospital

Jiwen Cheng
Xi'an Jiaotong University Second Affiliated Hospital

Haixia Zhou
Wenzhou Medical University Second Affiliated Hospital and Yuying Children's hospital

Jiao Zhang

fcczhangj7@zzu.edu.cn Corresponding Author
ORCiD: https://orcid.org/0000-0002-4977-263X

Jing He
Guangzhou Women and Children's Medical Center
Abstract

Background: Wilms tumor is the most common pediatric renal cancer. However, genetic bases behind Wilms tumor remain largely unknown. H19 is a critical maternally imprinted gene. Previous studies indicated that Single nucleotide polymorphisms (SNPs) in the H19 can modify the risk of several human malignancies. Epigenetic errors at the H19 locus lead to biallele silencing in Wilms tumors. Genetic variations in the H19 may be related to Wilms tumor susceptibility.

Methods: We conducted a four-center study to investigate whether H19 SNPs was a predisposing factor to Wilms tumor. Three polymorphisms in the H19 (rs2839698 G>A, rs3024270 C>G, rs217727 G>A) were genotyped in 355 cases and 1070 cancer-free controls, using Taqman method. Odds ratios (ORs) and 95% confidence intervals (CIs) were calculated to evaluate the strength of the associations.

Results: We found that all of these three polymorphisms were significantly associated with Wilms tumor risk alterations. Carriers of 1, 2 and 1-2 risk genotypes were inclined to develop Wilms tumor compared with those without risk genotype [adjusted odds ratio (OR)=1.36, 95% confidence interval (CI)=1.02-1.80, P =0.037; adjusted OR=1.84, 95% CI=1.27-2.67, P =0.001; adjusted OR=1.50, 95% CI=1.17-1.92, P =0.002, respectively]. The stratified analysis further revealed that rs2839698 AA, rs217727 AA, and 1-2 risk genotypes could strongly increase Wilms tumor risk among children above 18 months of age, males, and with clinical stage I+II disease.

Conclusion: Our findings indicate that genetic variations in the H19 may confer Wilms tumor risk.

Background

Wilms tumor, also known as nephroblastoma, is derived from the pluripotent embryonic kidney precursor. Wilms tumor is the most common renal malignancy in children, accounting for 85% of pediatric renal tumors [1-3]. It is characterized by early diagnosis and male predominance worldwide, with incidence varying by race [4]. The prevalence of Wilms tumor is similar in black and white children [5], but is around half in East Asian children, about three per million [4]. In China, the frequency of Wilms tumor is around 3.3 per million, ranking the fifth in the incidence of malignant tumors in children aged 0 to 4 years [6]. Besides, about 1-3% of Wilms tumor have a family history, probably due to rare germline mutations and incomplete expressiveness [7]. Environmental factors
and immigration factors seem not to play a prominent role in etiology [1, 4, 8]. The survival rate of Wilms tumor is more than 90% after excluding some high-risk cases with anaplastic histology, bilateral lesions and recurrent diseases [9]. However, up to 25% of survivors reported severe chronic health problems [10]. Moreover, late diagnosis and high recurrence rates in patients are reported in underdeveloped regions [11], based on the difficulty of stratification of increasingly refined tumor subtypes and the high cost of chemoradiotherapy for high-risk tumors [9]. Therefore, to improve the outcomes, it is of great significance to enhance prevention and early diagnosis by developing accurate biomarkers to identify high-risk individuals.

As a critical maternally imprinted gene, the H19 was discovered successively in different laboratories in the 1980s. This gene located on chromosome 11p15.5 in humans is composed of five exons and four introns [12]. The expression of H19 is highly increased in many embryos and decreased after birth [13]. H19 gene encodes a long non-coding RNA, which may have tumor-inhibiting functions [14]. More and more evidence indicates that the H19 gene is essential for human tumor growth from different biological processes [15]. Studies have shown that the H19 gene was upregulated in lung cancer, gastric cancer, colon cancer, retinoblastoma, thyroid cancer and breast cancer [15–20]. However, the up-regulated expression of the H19 gene can inhibit pituitary tumor cell proliferation in vitro and in vivo [21]. Consistently, H19 gene expression decreased in most hepatoblastomas [22]. Studies have shown that epigenetic errors at the H19 gene site in early embryonic development may result in the silencing of the double-alleles in Wilms tumor, thereby affecting the imprinting of parental alleles [23]. Matthew K Iyer et al. found many IncRNAs overlapping disease-associated SNPs [24]. Previous genomics studies have demonstrated that SNPs in several genes are associated with the risk of Wilms tumor [25–27]. It has been reported that H19 rs2839698 G > A, rs3024270 C > G or rs217727 G > A polymorphism is not associated with neuroblastoma susceptibility in the whole study population, while in stratified analysis, girls with rs3024270 GG genotype had an increased risk of neuroblastoma [28]. To date, no publication has been reported on the association between H19 gene polymorphisms and Wilms tumor susceptibility. In this study, we scrutinized the association of several H19 gene SNPs (rs2839698, rs3024270, and rs217727) and Wilms tumor risks based on a four-center study of
Chinese children.

Materials And Methods

Study subjects

The cases were enrolled in this project according to previously reported criteria [29-31]. In brief, 355 Wilms tumor cases and 1,070 healthy controls were included in this study (Supplemental Table 1). The 355 cases were from four medical centers (Guangzhou Women and Children’s Medical Center, The First Affiliated Hospital of Zhengzhou University, The Second Affiliated Hospital, and Yuying Children’s Hospital of Wenzhou Medical University, and Second Affiliated Hospital of Xi’an Jiao Tong University). All the control groups were selected from the same region as cases during the same period. Patients’ age, sex, and clinical stages were collected by trained medical staff. We conducted this study following the approval of the Institutional Review Board of the participating hospitals. All the participants’ parents provided signed informed consent before the examination.

Polymorphism analysis

Each subject donated about 2 mL of peripheral blood for DNA extraction using a TIANamp Blood DNA Kit (TianGen Biotech Co. Ltd., Beijing, China). Three H19 gene polymorphisms (rs2839698 G>A, rs3024270 C>G, rs217727 G>A) were chosen for genotyping [28]. We genotyped the gene polymorphisms using Taqman real-time PCR [32, 33]. The randomized and blinded process method was adopted while genotyping all samples. Approximately 10% random selection samples were re-genotyped, and the genotype concordance rate was 100%.

Statistical analysis

Departures from Hardy-Weinberg equilibrium (HWE) for the selected SNPs in controls were evaluated using a goodness-of-fit $\chi^2$ test. Allele frequencies and demographic variables between the two groups were assessed by the $\chi^2$ test. Risk associations between genotypes and Wilms tumor were determined from a logistic regression analysis. The ORs, 95% CIs, and the corresponding $P$ value for each SNP were calculated with adjustment for age and gender. All statistical calculations were implemented with the utilization of SAS software version 9.4 (SAS Institute, Cary, NC). Two-sided statistical tests were employed in this study. The significance threshold was defined as $P<0.05$. 
Results

**Associations between H19 gene polymorphisms and Wilms tumor susceptibility**

The detailed characteristics of all the subjects were shown in Supplementary Table 1. A total of 355 patients and 1068 healthy controls were successfully genotyped. The genotype frequencies of the three selected H19 gene polymorphisms and their associations with Wilms tumor susceptibility were presented in Table 1. We observed the genotype frequency distributions of the selected H19 gene polymorphisms were no significant deviation with the Hardy-Weinberg equilibrium (\(P=0.245\) for rs2839698 G>A, \(P=0.138\) for rs3024270 C>G, \(P=0.992\) for rs217727 G>A polymorphism) in controls. In single-locus analysis, we observed that all three polymorphisms were significantly associated with Wilms tumor risk individually. Specifically, the risk estimates for the these SNPs were as follows: the rs2839698 G>A polymorphism (AG vs. GG: adjusted OR=0.74, 95% CI=0.57-0.96, \(P=0.024\); AA vs. GG: adjusted OR=1.52, 95% CI=1.05-2.22, \(P=0.027\); AA vs. GG/AG: adjusted OR=1.75, 95% CI=1.23-2.50, \(P=0.002\)), the rs3024270 C>G polymorphism (CG vs. CC: adjusted OR=0.61, 95% CI=0.46-0.81, \(P=0.0007\); CG/GG vs. CC: adjusted OR=0.73, 95% CI=0.57-0.95, \(P=0.018\); GG vs. CC/CG: adjusted OR=1.38, 95% CI=1.05-1.82, \(P=0.023\)), and the rs217727 polymorphism (AG vs. GG: adjusted OR=0.76, 95% CI=0.58-0.99, \(P=0.035\)).

While analyzing the combined effect of risk genotypes, we found that subjects carrying 1 or 2 risk genotypes had a significantly increased Wilms tumor risk when compared with those without risk genotypes (adjusted OR=1.36, 95% CI=1.02-1.80, \(P=0.041\); and adjusted OR=1.84, 95% CI=1.27-2.67, \(P=0.001\)). Moreover, we found that subjects with 1-2 risk genotypes were significantly more likely to develop Wilms tumor than subjects carrying no risk genotypes (adjusted OR=1.50, 95% CI=1.17-1.92, \(P=0.002\)).

**Stratification analysis**

We then performed a stratified analysis to explore how age, gender, and clinical stage influence the association between selected polymorphisms and Wilms tumor susceptibility (Table 2). Compared to the rs2839698 GG/AG genotype, the risk effects of AA genotypes were more predominant in children above 18 months of age (adjusted OR=1.73; 95% CI=1.09-2.74, \(P=0.020\)), female (adjusted OR=1.94,
95% CI=1.11-3.39, \( P=0.021 \), male (adjusted OR=1.63, 95% CI=1.02-2.58, \( P=0.040 \)), and those with clinical stage I+II disease (adjusted OR=1.83, 95% CI=1.20-2.79, \( P=0.005 \)). Consistently, with the rs217727 GG/AG genotype as references, AA genotype carriers was associated with an increased risk of Wilms tumor for children above 18 months of age (adjusted OR=1.65; 95% CI=1.06-2.58, \( P=0.027 \)), male (adjusted OR=1.60, 95% CI=1.01-2.54, \( P=0.047 \)), clinical stage I+II cases (adjusted OR=1.60, 95% CI=1.05-2.44, \( P=0.029 \)). However, no association was observed between rs3024270 and Wilms tumor susceptibility in subgroups defined by age, sex, and clinical stages.

We also interrogated the cumulative effects of these SNPs on Wilms tumor risk in the stratified analysis. We found that the presence of 1-2 risk genotypes was significantly associated with the risk of Wilms tumor in children above 18 months of age (adjusted OR=1.66; 95% CI=1.21-2.27, \( P=0.002 \)), male (adjusted OR=1.59, 95% CI=1.14-2.21, \( P=0.006 \)), and clinical stage I+II patients (adjusted OR=1.64, 95% CI=1.21-2.22, \( P=0.002 \)) when compared with those of 0 risk genotype.

**Discussion**

In the current hospital-based case-control study, we demonstrated the association of three H19 gene polymorphisms with Wilms tumor susceptibility. This article was the first report indicating that H19 SNPs were related to Wilms tumor risk.

Non-coding RNAs are known to play central roles in the dynamic control of transcriptional and gene expression [34]. LncRNAs contribute to the pathogenesis of various cancers by participating in the control of cell cycle, proliferation, differentiation, and apoptosis [35, 36]. H19 gene is the only imprinted gene that can encode lncRNA and play a role in the mRNA level [37]. In this study, the three SNPs we screened were all located in the H19 gene [38]. So far, 10 polymorphisms in H19 have been identified as predisposing factors to various cancer types, among which the rs217727 has been most frequently studied, followed by rs2839698 [39]. H19 plays an essential role in the up-regulation of the expression of breast cancer, bladder cancer, gastric cancer, and other tumors; other than that, mutations in the H19 gene coding sequence are also closely related to tumors, despite unknown regulatory mechanisms [12, 40]. The following evidence suggests that SNPs may affect the expression and function of the H19 gene. The rs2839698 polymorphism may influence the folding
structures of lncRNA H19 and change the target microRNAs of lncRNA H19, thereby increasing the risk of colorectal cancer [41]. Verhaegh et al. found that the folding structure of rs217727 and rs2839698 of lncRNA H19 was different under TT and CC genotypes, and both the T and C genotype of them had a significantly decreased risk of bladder cancer [42]. What’s more, the rs217727 CT + TT genotype was associated with a lower risk of breast cancer in women who were pregnant more than twice [43]. The above results indicated that the H19 gene encoding the SNP variation might lead to changes in the secondary structure of lncRNA H19, subsequently altering the biological characteristics of lncRNA H19 and the occurrence and development of tumors. The lncRNA H19 could be a potential diagnostic and prognostic marker in the development of tumors [44], and the different genotypes of SNPs might facilitate an individualized diagnosis of cancer.

There are other explanations of the relationship between lncRNA H19 and tumors. The miR675 signal axis plays a vital role in tumorigenesis, which is a microRNA, embedded in the H19 gene’s first exon [45]. Li et al. first demonstrated that miR675 promoted liver carcinogenesis through the cascade of miR675-HP1α-EGR1-H19-PKM2 signaling and clearly demonstrated that miR675 overexpression stimulated liver cancer cell growth, vice versa [46]. Wu et al. revealed that lncRNA H19 promoted laryngeal squamous cell carcinoma (LSCC) progression via miR-148a-3p and DNMT1, indicating that H19 plays the role of microRNA sponge in promoting tumor development [47]. Another study suggested that the effect of H19 in GC is mediated by the direct upregulation of ISM1 and the indirect suppression of CALN1 expression via miR-675 [48]. Additionally, functional SNP rs217727 in H19 is highly likely to be involved in breast cancer development in hormone-signaling pathways [49].

In stratified analysis, Wilms tumor risk of rs2839698 variant GG/AG genotypes was more evident in subgroups of age above 18 months, female, male, and clinical stage I + II cases. The same genotype is also associated with an increased risk of gastrointestinal cancer [39]. Similar results were obtained in rs217727 GG/AG except in gender considerations, only males. In addition, previous data showed that stratified analysis of rs217727 C > T showed both dominant and recessive effects associated with increased risk of oral squamous cell carcinoma (OSCC) and lung cancer [39]. Our results further revealed the critical influence of G and A genotype in H19 rs217727. These facts may partially explain
the apparent imbalance of the analyzed SNPs. We did not find any associations between the rs3024270 genotype and Wilms tumor in stratified analysis.

There are potential limitations of the current study: 1) the relatively small sample size and lacking participants from different ethnic groups, 2) the consideration of only three polymorphisms without potential function, and 3) unknown living environmental factors on.

To conclusion, we verified that the rs2839698 G > A, rs3024270 C > G, rs217727 G > A polymorphisms were significantly associated with the risk of Wilms tumor. Further stratified data showed that older children, early clinical stage and gender were risk factors. Thus, the results of our study should be verified in studies with larger samples from different ethnicities.

Abbreviations
SNP: Single nucleotide polymorphism; OR: Odds ratio; CI: confidence interval;
HWE: Hardy-Weinberg equilibrium; PCR: Polymerase chain reaction;
IncRNA: long non-coding RNA

Declarations

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Authors’ contributions

All authors contributed to data analysis, drafting or revising the article, gave final approval of the version to be published, and agree to be accountable for all aspects of the work.

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Availability of data and materials
The datasets during and/or analyzed during the current study available from the corresponding author on reasonable request.

Ethics approval and consent to participate
The study was approved by the Institutional Review Board of the participating hospitals. All the participants’ parents provided signed informed consent before the examination.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

ORCID
Jing He, https://orcid.org/0000-0002-1954-2892
Wenya Li, https://orcid.org/0000-0001-6638-8675

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Tables
### Table 1. Associations between H19 polymorphisms and Wilms tumor risk

| Genotype            | Cases (N=355) | Controls (N=1068) | \( p^a \) | Crude OR (95% CI) | \( P \) |
|---------------------|---------------|-------------------|---------|-------------------|-----|
| rs2839698 (HWE=0.245) |               |                   |         |                   |     |
| GG                  | 174 (49.01)   | 488 (45.69)       | 1.00    |                   |     |
| AG                  | 127 (35.77)   | 480 (44.94)       | 0.74 (0.57-0.96) | 0.025 |
| AA                  | 54 (15.21)    | 100 (9.36)        | 1.52 (1.04-2.20) | 0.029 |
| Additive            |               |                   | 0.0008  |                   |     |
| Dominant            | 181 (50.99)   | 580 (54.31)       | 1.06 (0.89-1.27) | 0.537 |
| Recessive           | 301 (84.79)   | 968 (90.64)       | 0.88 (0.69-1.11) | 0.277 |
| rs3024270 (HWE=0.138) |               |                   |         |                   |     |
| GG                  | 120 (33.80)   | 290 (27.15)       | 0.61 (0.46-0.81) | 0.0007 |
| AG                  | 141 (39.72)   | 556 (52.06)       | 1.02 (0.74-1.41) | 0.888 |
| AA                  | 94 (26.48)    | 222 (20.79)       | 0.98 (0.83-1.16) | 0.826 |
| Additive            |               |                   | 0.0003  |                   |     |
| Dominant            | 235 (66.20)   | 778 (72.85)       | 0.73 (0.56-0.95) | 0.017 |
| Recessive           | 261 (73.52)   | 846 (79.21)       | 1.37 (1.04-1.81) | 0.026 |
| rs2177277 (HWE=0.992) |               |                   |         |                   |     |
| GG                  | 177 (49.86)   | 486 (45.51)       | 0.76 (0.59-0.99) | 0.039 |
| AG                  | 130 (36.62)   | 469 (43.91)       | 1.17 (0.80-1.70) | 0.426 |
| AA                  | 48 (13.52)    | 113 (10.58)       | 0.97 (0.81-1.16) | 0.733 |
| Additive            |               |                   | 0.039   |                   |     |
| Dominant            | 178 (50.14)   | 582 (54.49)       | 0.84 (0.66-1.07) | 0.155 |
| Recessive           | 307 (86.48)   | 955 (89.42)       | 1.32 (0.92-1.90) | 0.131 |
| Combined effect of risk genotypes \(^c\) |               |                   |         |                   |     |
| 0                   | 211 (59.44)   | 732 (68.54)       | 1.00    |                   |     |
| 1                   | 92 (25.92)    | 237 (22.19)       | 1.35 (1.01-1.79) | 0.041 |
| 2                   | 52 (14.65)    | 99 (9.27)         | 1.82 (1.26-2.64) | 0.001 |
| Trend               |               |                   | 1.35 (1.14-1.60) | 0.0005 |
| 0                   | 211 (59.44)   | 732 (68.54)       | 1.00    |                   |     |
| 1-2                 | 144 (40.56)   | 336 (31.46)       | 1.49 (1.16-1.91) | 0.002 |

\( ^a \) \( \chi^2 \) test for genotype distributions between Wilms' tumor patients and controls.

\( ^b \) Adjusted for age and gender.

\( ^c \) Risk genotypes were carriers with rs2839698 AA, rs3024270 GG and rs217727 AA genotypes.

### Table 2. Stratification analysis for association between H19 genotypes and Wilms' tumor susceptibility.

| Variables          | rs2839698 (case/control) | Adjusted OR\(^a\) (95% CI) | \( p^a \) | rs3024270 (case/control) | Adjusted OR\(^a\) (95% CI) | \( p^a \) |
|--------------------|---------------------------|-----------------------------|---------|---------------------------|-----------------------------|-----|
| Age, month         |                           |                             |         |                           |                             |     |
| ≤18                | 104/382                   | 21/43                       | 1.76 (0.99-3.10) | 92/335                   | 33/90                       | 1.32 (0.84-2.10) |
| >18                | 197/586                   | 33/57                       | **1.73 (1.09-2.74)** | 169/511                   | 61/132                      | **1.41 (0.99-2.01)** |
| Gender             |                           |                             |         |                           |                             |     |
| Female             | 140/412                   | 23/35                       | **1.94 (1.11-3.39)** | 121/361                   | 42/86                       | **1.46 (0.96-2.23)** |
| Male               | 161/556                   | 31/65                       | **1.63 (1.02-2.58)** | 140/485                   | 52/136                      | **1.32 (0.91-1.91)** |
| Clinical stage     |                           |                             |         |                           |                             |     |
| I+II               | 117/968                   | 34/100                      | **1.83 (1.20-2.79)** | 156/846                   | 55/222                      | **1.35 (0.95-1.89)** |
| III+IV             | 108/968                   | 18/100                      | 1.66 (0.96-2.85) | 93/846                    | 33/222                      | 1.36 (0.89-2.07) |

\( ^a \) Adjusted for age and gender, omitting the corresponding stratify factor.
Supplementary Files

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Supplemental Table 1-Feb4.doc