Evaluation of Skeletal Muscle Dysfunction Associated With Acute Inflammation by Electrical Impedance Myography: A Case Report on Skeletal Muscle Dysfunction After Cardiac Surgery and Literature Review

Hiroki Sato 1, 2, 3, Takao Nakamura 4

1. Department of Radiological Technology, Graduate School of Health Sciences, Okayama University, Okayama, JPN 2. Department of Rehabilitation Medicine, Kawasaki Medical School, Kurashiki, JPN 3. Department of Rehabilitation Center, Kawasaki Medical School Hospital, Kurashiki, JPN 4. Graduate School of Health Sciences, Okayama University, Okayama, JPN

Corresponding author: Hiroki Sato, h0306.pt@gmail.com

Abstract

Electrical impedance myography (EIM) is an evaluation technique for skeletal muscles that uses electrical impedance technology. Recent reviews have shown that EIM is useful as a method to assess changes in skeletal muscle quality and quantity with aging. These may be utilized for functional changes in inflammatory skeletal muscles, such as disease and operation. In this report, the impedance parameters using EIM present perioperative skeletal muscle changes in patients after cardiac surgery. In addition, we will describe the efficacy of EIM in skeletal muscle dysfunction due to inflammation or disease. This study aimed to elucidate the efficacy of EIM in acute inflammation-associated skeletal muscle dysfunction.

Introduction

Bioelectrical impedance technology has been used in the medical field to assess body composition, such as body water, skeletal muscle, and fat content [1]. Recently, a diagnostic technique called electrical impedance myography (EIM) has been proposed, which measures the intrinsic impedance of skeletal muscle using these techniques [2]. In EIM, a variety of numerical values are calculated, including the phase angle (PA) and the ratio of intracellular to extracellular fluid resistance (Ri/Rc), the beta parameter (β) that represents tissue uniformity, and the center frequency (fC). These parameters have been shown to be related to muscle fiber mass and fascial function in skeletal muscles [2]. We investigated the effectiveness of skeletal muscle quality and quantitative assessment of the impedance parameters of the upper and lower limb measured by direct segmental multifrequency bioelectrical impedance analysis and found that Ri/Rc correlates with muscle mass and beta parameters with muscle mass indices and that each has an independent effect on muscle strength [3]. Thus, PA, which reflects the state of cells, may be applied to evaluate quality and quantity by separating them into their constituent components.

Skeletal muscle function is altered not only by aging but also by diseases and systemic inflammation [4]. Skeletal muscle dysfunction associated with systemic inflammation, especially after cardiac surgery, has been shown to cause a severe functional decline in a short period of time [5]. However, there is no standardized evaluation method immediately after surgery because general conditions and pain make conventional muscle strength evaluation difficult. Although EIM is likely to be effective for such changes in skeletal muscle function in the acute phase, no studies have investigated changes in skeletal muscle function in the acute phase using impedance parameters by EIM.

This case report presents perioperative changes in skeletal muscle in patients after cardiac surgery using impedance parameters by EIM. In addition, EIM insights into acute inflammation and disease-induced skeletal muscle dysfunction are discussed.

Case Presentation

A 56-year-old Japanese male with severe aortic regurgitation (maximum velocity of blood flow at the aortic valve opening, 2.3 m/s; aortic valve area, 1.54 cm²; ejection fraction, 36%) and dilatation of the ascending aorta (5.1 cm) on echocardiography was indicated for surgery. Concomitant diseases included hypertension, dyslipidemia, and diabetes mellitus (type II). He worked at his office and was independent in his daily life. Rehabilitation was initiated before surgery. Before surgery, physical function tests by the physiotherapist...
revealed a body weight of 72.0 kg, grip strength of 38.0 kg, leg extension strength of 138.0 Nm, skeletal muscle index (SMI) of 8.1 kg/m², short physical performance battery (SPPB) of 12 points, and no abnormalities in physical function. Skeletal muscle function was evaluated using the rectus femoris muscle. Muscle thickness and intensity were measured using ultrasonography (SonoSite M-turbo, Fujifilm, Japan) [6]. EIM was measured using a bioelectrical impedance spectroscopy unit (SFB7, Impedimed, Australia) to measure PA and β (details of measurement and analysis methods are given in the Appendix).

Approximately one week of preoperative rehabilitation was followed by Bentall surgery (extracorporeal circulation time, 212 min; anesthesia time, 497 min; operation time, 408 min). Rehabilitation (e.g., early mobilization and respiratory rehabilitation) was started the day after surgery. On a postoperative day 1 (POD 1), the patient was weaned from a ventilator, and on POD 3, the patient was able to walk 100 m. There were no complications, the patient became independent in daily life on POD 7, and the patient was discharged from the hospital on POD 18. Physical function was evaluated in POD 14, and skeletal muscle function was evaluated preoperatively and in PODs 1, 3, 5, 7, and 14.

The physical function evaluation of POD 14 showed a body weight of 66.7 kg, grip strength of 36.5 kg, leg extensor strength of 126.0 Nm, SMI of 7.8 kg/m², and SPPB of 12 points. Table 1 shows the evaluation of perioperative skeletal muscle function. In the evaluation of skeletal muscle function, ultrasonography showed a decrease in the thickness of the rectus femoris muscle from 1.81 cm preoperatively to 1.64 cm on POD 14. The increase in muscle intensity peaked on POD 5 and gradually improved after POD 7. In the EIM, PA decreased from 14.6° before surgery to 11.1° on POD 14. The β peaked at POD 3, and the POD 14 level improved to the same level as the preoperative level. Increased from baseline (preoperatively) to 0.32 and 0.36 on POD 5 and POD 14, respectively.

| US  | Preoperative | POD 1 | POD 3 | POD 5 | POD 7 | POD 14 |
|-----|--------------|-------|-------|-------|-------|--------|
| MT (cm) | 1.81 | 1.60 | 1.71 | 1.65 | 1.65 | 1.64 |
| MI | 36.1 | 53.3 | 56.2 | 52.6 | 48.6 | 40.5 |
| EIM | | | | | | |
| PA (°) | 14.6 | 14.0 | 13.5 | 12.7 | 13.0 | 11.1 |
| β | 0.793 | 0.748 | 0.722 | 0.761 | 0.773 | 0.797 |
| Ri /Re | 0.32 | 0.31 | 0.31 | 0.34 | 0.34 | 0.36 |
| fc (kHz) | 24.2 | 28.3 | 28.3 | 38.7 | 36.6 | 35.5 |

**TABLE 1: Ultrasonography and EIM of the rectus femoris during perioperative**

US: ultrasonography, MT: muscle thickness, MI: muscle intensity, EIM: electrical impedance myography, PA: phase angle, β: beta parameter, Ri /Re: ratio of intracellular fluid resistance to extracellular fluid resistance, fc: central relaxation frequency, POD: postoperative day

**Discussion**

To the best of our knowledge, this is the first report to evaluate EIM for inflammatory skeletal muscle dysfunction after surgery. Previous studies have shown that the impedance parameters measured by EIM are associated with age-related changes in skeletal muscle and neuromuscular diseases [2]. In this case, acute changes in skeletal muscle function caused by inflammatory reactions after thoracotomy were serially evaluated using ultrasonography and EIM to determine the efficacy of EIM. In addition to the changes in skeletal muscle mass and quality measured using ultrasonography, PA decreased markedly. A noteworthy finding is that the changes in PA-constituted β occurred independently and were more sensitive than the quantity and quality indices of skeletal muscle assessed with ultrasonography.

Skeletal muscle damage associated with acute inflammation is caused by infiltration of inflammatory cytokines, leading to a decrease in fascial function and an increase in non-contractible tissue within the muscle [4]. Changes in skeletal muscle quality due to these disorders cause marked muscle weakness in a short period of time. After cardiac surgery, muscle weakness of >10% during POD 7 is associated with inflammatory muscle protein catabolism [5]. As in the previous study, muscle weakness and a decrease in muscle mass of approximately 10% were observed in this case after the operation. Muscle intensity increased more at POD 1 and gradually improved after peaking at POD 5. Meanwhile, the EIM results showed that PA gradually decreased from 14.6° (preoperatively) to 13.0° (a decrease of 11.2%) and 11.1° (a decrease of...
The equipment used was a Strength Ergo 5 (BK-ERG-051, Mitsubishi Electric, Japan). The measurement was performed in isokinetic mode at a rotation speed of 50 rpm and five consecutive drives, and then the peak
torque (Nm) was calculated.

**Electrical impedance myography**

A Bioelectrical Impedance Spectroscopy Unit (SFB7, Impedimed, Australia) was used. The electrodes used for measurement were plastic sprint materials (Easy Foam, Sakai Medical, Japan), and handheld electrodes with an arc-shaped electrode mounting surface were used. The electrode mounting surface was 80 mm (R 66 mm arc) × 150 mm wide, and the tape electrodes (Ag-AgCl, 10 mm × 80 mm) were mounted in a four-electrode arrangement. The distance between the potential electrodes was 100 mm, and the distance between the potential electrodes was 20 mm. A hard-type jelly (Conductor Transmission Gel, Chattanooga) was used during the measurement to avoid skin pressure and resistance. The rectus femoris muscle was measured from the anterior superior iliac spine to the midpoint of the upper patellar margin, Unified. For the analysis of impedance, parameters were calculated by arc optimization using the Cole-Cole model using 172 points of resistance and reactance with frequencies in the range of 10-300 kHz, and PA were calculated from the numerical values in the same manner as in a previous study [3]. The EIM was measured twice, and the average value was calculated.

**Additional Information**

**Disclosures**

**Human subjects:** Consent was obtained or waived by all participants in this study. **Conflicts of interest:** In compliance with the ICMJE uniform disclosure form, all authors declare the following: **Payment/services info:** All authors have declared that no financial support was received from any organization for the submitted work. **Financial relationships:** All authors have declared that they have no financial relationships at present or within the previous three years with any organizations that might have an interest in the submitted work. **Other relationships:** All authors have declared that there are no other relationships or activities that could appear to have influenced the submitted work.

**References**

1. Ling CH, de Craen AJ, Slagboom PE, Gunn DA, Stokkel MP, Westendorp RG, Maier AB: Accuracy of direct segmental multi-frequency bioimpedance analysis in the assessment of total body and segmental body composition in middle-aged adult population. Clin Nutr. 2011, 30:610-5. 10.1016/j.clnu.2011.04.001

2. Clark BC, Rutkove S, Lupton EC, Padilla CJ, Arnold WD: Potential utility of electrical impedance myography in evaluating age-related skeletal muscle function deficits. Front Physiol. 2021, 12:669664. 10.3389/fphys.2021.669664

3. Sato H, Nakamura T, Kusuhara T, Kenichi K, Kuniyusako K, Kawashima T, Hanayama K: Effectiveness of impedance parameters for muscle quality evaluation in healthy men. J Physiol Sci. 2020, 70:53. 10.1186/s12576-020-00780-z

4. Lomhle P, Guttridge DC: Inflammation induced loss of skeletal muscle. Bone. 2015, 80:131-42. 10.1016/j.bone.2015.05.015

5. Iida Y, Yamazaki T, Arima H, Kowate T, Yamada S: Predictors of surgery-induced muscle proteolysis in patients undergoing cardiac surgery. J Cardiol. 2016, 68:536-41. 10.1016/j.jjcc.2015.11.011

6. Sato H, Kuniyusako K, Kobara K, et al.: Verification of the accuracy of measuring the muscle cross-sectional area and muscle intensity of the rectus femoris using ultrasonography. Japanese J Compr Rehabil Sci. 2018, 9:66-72. 10.11356/ijcrs.9.66

7. Kapur K, Nagy JA, Taylor RS, Sanchez B, Rutkove SB: Estimating myofiber size with electrical impedance myography: a study in amyotrophic lateral sclerosis mice. Muscle Nerve. 2018, 58:713-7. 10.1002/mus.26187

8. Shefror JM, Rutkove SB, Caress JB, et al.: Reducing sample size requirements for future ALS clinical trials with a dedicated electrical impedance myography system. Amyotroph Lateral Scler Frontotemporal Degener. 2018, 19:555-61. 10.1080/21678421.2018.1510008

9. Rutkove SB, Kapur K, Zaidman CM, et al.: Electrical impedance myography for assessment of Duchenne muscular dystrophy. Ann Neurol. 2017, 81:622-32. 10.1002/ana.24874

10. Jia Li, Sanchez B, Rutkove SB: The effect of profound dehydration on electrical impedance of mouseskeletal muscle. Annu Int Conf IEEE Eng Med Biol Soc. 2014, 2014:514-7. 10.1109/EMBC.2014.6945641

11. McLester CN, Dewitt AD, Rooks R, McLester JR: An investigation of the accuracy and reliability of body composition assessed with a handheld electrical impedance myography device. Eur J Sport Sci. 2018, 18:765-71. 10.1080/17461391.2018.1448458

12. Li Z, Chen L, Zhu Y, Wei Q, Liu W, Tian D, Yu Y: Handheld electrical impedance myography probe for assessing Carpal Tunnel syndrome. Ann Biomed Eng. 2017, 45:1572-80. 10.1007/s10439-017-1819-3

13. Boujemaa H, Verbouven K, Hendriks M, et al.: Muscle wasting after coronary artery bypass graft surgery: impact on post-operative clinical status and effect of exercise-based rehabilitation. Acta Cardiol. 2020, 75:406-10. 10.1007/s10554-019-008055