Pallister-Killian syndrome: clinical, cytogenetic and molecular findings in 15 cases

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Abstract

Background: Pallister Killian syndrome (PKS, OMIM 601803) is a rare genetic disorder with a distinct phenotype caused by tissue-limited mosaicism tetrasomy of the short arm of chromosome 12, which usually cytogenetically presents as an extra isochromosome 12p. Wide phenotypic variability in PKS has been reported, ranging from pre-to perinatal death due to multiple congenital anomalies, especially diaphragmatic hernia, and classic phenotypes including seizures, severe developmental delay, macrosomia at birth, deafness, and distinct dysmorphic features, such as coarse face, temporal alopecia, a small nose with anteverted nostrils, long philtrum, and hypo-/hyper-pigmented streaks on the skin.

Results: Karyotypes obtained from cultured peripheral lymphocytes of 13 cases, who were diagnosed as PKS, were normal, while karyotypes obtained from cultured skin samples and buccal mucosa revealed the supernumerary mosaic i(12p). Mosaic karyotype was found in both fibroblast and buccal mucosa in 14 of 15 patients in our series, whereas in one stillbirth, following the clinical diagnosis of PKS, skin and buccal smear samples were taken, and all karyotypes from cultured fibroblasts revealed a supernumerary i(12p), while I-FISH study showed 60% mosaicism in mucosal cells.

Conclusions: We here share the clinical, cytogenetic and molecular cytogenetic findings of 15 cases with PKS phenotype and the parental origin of seven i(12p) identified by molecular analyses. To our knowledge, this is the largest series of PKS patients with parental origin study from a single center. We believe that our study makes a significant contribution to the literature because we specifically found no differences in the phenotypes of cases with either a maternal or paternal origin of the extra element and differential imprinting appeared not to be a factor.

Keywords: OMIM 601803, Pallister-Killian syndrome, Somatic mosaicism, Mosaic tetrasomy 12p, Isochromosome 12p, Parental origin, Small supernumerary marker chromosome

Background

Pallister-Killian syndrome (PKS) was first described an adult phenotype by Pallister in 1977 and independently a childhood phenotype by Killian and Teschler-Nicola in 1981 [1, 2]. In both instances, cytogenetic examinations of blood cells did not reveal any abnormality. Later observations by Schinzel in 1991 confirmed a distinct phenotype. In 1987, PKS was shown to be caused by somatic mosaicism of an extra chromosome, which was not primarily not detected since mosaicism was limited to fibroblasts and not present in lymphocytes [3, 4]. Using banded karyotypes, and after several mis-interpretations, the extra element was identified as an isochromosome 12p.

The clinical phenotype of PKS is quite homogeneous for the dysmorphic pattern, but highly variable for developmental delay, congenital malformations, and survival. There is a wide range of congenital malformations that occur rarely including anal atresia/stenosis and atlanto-occipital fusion. Areas of hyper- or hypo-pigmentation are often found and reflect the mosaic pattern of the skin, with different proportions of hyperdiploid cells [5].

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The developmental delay in patients with PKS is quite variable, and although most reported patients with PKS have severe to profound mental retardation, milder phenotypes have also been reported [6–13]. A large number of patients with i(12p) were reported in the literature, and also their clinical and cytogenetic information routinely posted in a database (http://ssmc-tl.com/ssMC.html http://ssmc-tl.com/chromosome-12.html http://ssmc-tl.com/chromosome-12.html#iso) [14]. Here, we report the cytogenetic, and molecular cytogenetic findings in 15 cases with PKS phenotype and parental origin analysis results of 7 cases with i(12p) chromosome. To our knowledge, this series is the first series from Turkey and the largest series of PKS patients with parental origin study from a single center.

Methods
All cases were evaluated at Medical Genetics Department outpatients’ clinics, Istanbul Medical Faculty, Istanbul University. Cytogenetic and molecular cytogenetic analyses were performed in the laboratory of the department. Before the implementation of the FISH technique, in presence of the clinical features of PKS, cytogenetic studies were performed from cultured fibroblasts from skin biopsies in parallel to the peripheral lymphocyte cultures. Following the introduction of the interphase FISH (I-FISH) study using chromosome 12 centromeric probe (D12Z3, Aquarius®, Cytocell Cambridge, UK), I-FISH test on mucosal cells from buccal smear were applied in parallel to the lymphocyte cultures (Fig. 1).

De novo supernumerary i(12p) was further confirmed by FISH using chromosome 12 specific wcp, centromeric and subtelomeric probes.

To determine the parental origin and formation mechanism, 7 cases with i(12p) were investigated with gel-electrophoresis technique using 15 microsatellite markers on 12p (D12S352, D12S99, D12S77, D12S269, D12S320, D12S364, D12S1303, D12S373, D12S301, D12S1669, D12S1650, D12S799, D12S823, D12S1053, D12S345,) and 2 markers on 12q (D12S72, D12S43).

Results
Clinical findings
Fifteen cases were cytogenetically evaluated due to their phenotypic features (Fig. 2).

The mean age at the time of diagnosis was about 2.5 years of age (ranging between one day-7 years 10/12). Karyotypes of fibroblast cultures in 15 cases revealed the supernumerary mosaic i(12p), while cultured peripheral blood lymphocytes of those cases were normal. The 12 p tetrasomic metaphases were observed 10% to 100% of cells analyzed in fibroblasts. Although the level of +i(12p) in cultured fibroblasts was low (11%), I-FISH indicated higher level of i(12p) (40%) in the mucosal cells in one case. In one stillborn, following the clinical diagnosis of PKS, skin and buccal smear samples were taken, and a supernumerary i(12p) was present in all metaphases of cultured fibroblasts, while I-FISH study showed mosaicism of 60% in mucosal cells (Table 1).

The clinical findings of the cases with tetrasomy 12p are summarized in Table 2. Almost all cases with tetrasomy 12p had coarse facial features, bitemporal alopecia, anteverted nostrils and everted lower lip, hypertelorism, epicanthus, long philtrum and hypo/hyperpigmented skin changes. Intellectual disability was observed in all patients over 8 months of age except one case with near normal intelligence at adult age (case 7), and three newborns. Parental origin was determined by STR analysis as maternal in 6 cases (85.7%), paternal in one case (14.22%).

Discussion
Classical PKS is a rare condition caused by mosaic tetrasomy 12p presenting with an additional isochromosome of 12p, which is usually not present in blood lymphocyte cultures, but can be shown in skin
fibroblasts [3, 4] and other tissues such as buccal smears, chorionic villi and amniocytes. The clinical phenotype is characterized by a pattern of dysmorphic features, hypo- or hyperpigmentation areas of the skin, progressive coarsening of the face with advancing age and moderate to severe-profound intellectual disability accompanied with different types of seizures. Congenital malformations have been found in a minority of live births.

The dysmorphic pattern is age-dependent and includes brachycephaly, temporal balding during the first years of life, a short nose with flat bridge and anteverted nostrils giving the false impression of hypertelorism, microstomia, progressive macroglossia, prominent everted lower lip, small mandible and short neck. Supernumerary nipples are frequent as are hypoplastic male external genitalia. Hands and fingers, feet and toes are proportionately small. Pigmentation anomalies of the skin are due to mosaicism with different expression of pigmentation genes with predominance of one or the other cell line. This is well known for mosaicism in general [14–23].

Although there are limited number of reports on the mosaicism of the isochromosome in peripheral blood cells in PKS patients by I-FISH, microarray studies, and karyotype analysis [24–27]. Studies have shown that phytohaemagglutinin used in lymphocyte cultures promotes the growth of the normal cells, which leads to under-representation or disappearing of the abnormal cells [28–31]. Furthermore tissues with a shorter turnover period seem to lose the cells with additional chromosomes, either due to longer duration of cell division, the instability of (submicroscopically) dicentric chromosomes or the abnormal cell line going through apoptosis or necrosis at a higher rate than the normal cells [32].

Fig. 2 Facial appearance of the patients. Alopecia/Sparse temporal hair, hypo- or hyper pigmented areas and other stigmata such as epicanthic folds and slightly low-set ears, prominent forehead, hypertelorism, up-sloping palpebral fissures, anteverted nostrils, long philtrum, everted lower lip, short neck and short hands and toes
At present I-FISH using chromosome 12 centromeric probes on buccal mucosa cells is a very useful, non-invasive, reliable, rapid and cost-effective test for the definite diagnosis [33, 34]. It is suggested that all live born tetrasomy 12 cases are mosaic, even it could not be shown in one tissue. Therefore, it is helpful to investigate different tissues by different techniques to enlighten the mosaicism like in our case 11 (Table 2). However, in all similar cases from the literature only one or at most two different tissues had been analysed and thus mosaicism in non-examined tissues or even in a second skin biopsy from another area of the body was not excluded. In a few cases with clinical diagnosis of PKS normal karyotype both in blood and fibroblast cultures derived from one or two skin biopsies mosaicism could not be excluded.

**Early cytogenetic misinterpretations**

Prior to and during the early banding era of cytogenetics, several patients were published as tetrasomy 21 (an aberration which seems incompatible with life) [35–39] and the extra chromosome was also classified as chromosome 12 with deletion of about ¾ of the long arm [40].

**Mechanism(s) of formation of the additional i(12p)**

Molecular marker analysis favored the following steps: Initial maternal meiosis II nondisjunction (evidenced by centromere-near homozygosity of the maternal markers) leading to trisomy 12. Trisomy 12 zygotes will spontaneously abort; the few exceptions are those conceptuses in which a secondary structural aberration, isochromosome formation, results in loss of the acentric long arms and subsequent formation of an +i(12p) [41–45]. Ravel et al. published a mosaic case with trisomy 12 and +i(12p) diagnosed prenatally and proposed a formation mechanism for +i12p [46]. Pauli et al. discussed the mechanisms of formation based on the data of their monozygotic twins and suggested that prezygotic or postzygotic error (both before the twinning event) in the inner cell mass may have happened. [47].

The increased mean maternal age at conception suggests this mechanism [43]. In a few cases paternal origin of the i(12p) could be demonstrated [25, 42, 48], obviously caused by a different cascade of events. To the best of our knowledge, there seems to be no difference in phenotype between the cases of maternal versus paternal origin of the extra element and thus no effect of differential imprinting [6, 10].

**Karyotype-phenotype correlation**

It has been suggested that no correlation could be established between the proportion of tetrasomic cells and the severity of clinical presentation [5]. Our observation supports this suggestion, because of the technical limitations, the severity of the phenotype was independent from the degree of mosaicism in karyotyping. However, fibroblast cultures were analyzed from only one biopsy in all cases. As mentioned above, the incidence of hyperdiploid cells may grossly vary among

Table 1 Karyotypes and FISH findings in different tissues

| Case no | Karyotypes on cultured lymphocytes | Karyotype on fibroblast | % | I-FISH results on buccal swap sample | % | Parental origin |
|---------|----------------------------------|------------------------|---|-------------------------------------|---|----------------|
| 1       | 46,XY                            | mos 46,XY/47,XY,+i(12)(p10)[33/17] | 34 | –                                  |   | P              |
| 2       | 46,XX                            | mos 46,XX/47,XX,+i(12)(p10)[18/2] | 10 | –                                  |   | M              |
| 3       | 46,XX                            | mos 46,XX/47,XX,+i(12)(p10)[100/11] | 9.9 | .ish (D12Z2x2/D12Z2x3)[50/13] | 20.6 | M              |
| 4       | 46,XX                            | mos 46,XX/47,XX,+i(12)(p10)[28/12] | 30 | –                                  |   | M              |
| 5       | 46,XY                            | mos 46,XY/47,XY,+i(12)(p10)[65/35] | 35 | –                                  |   | M              |
| 6       | 46,XX                            | mos 46,XX/47,XX,+i(12)(p10)[24/9] | 27.3 | –                                  |   | M              |
| 7       | 46,XX                            | mos 46,XX/47,XX,+i(12)(p10)[15/5] | 25 | –                                  |   | M              |
| 8       | 46,XX                            | mos 46,XX/47,XX,+i(12)(p10)[31/4] | 11.4 | .ish (D12Z2x2/D12Z2x3)[64/41] | 39 | M              |
| 9       | 46,XY                            | mos 46,XY/47,XY,+i(12)(p10)[21/9] | 30 | .ish (D12Z2x2/D12Z2x3)[5/13] | 72 | P              |
| 10      | 46,XY                            | mos 46,XY/47,XY,+i(12)(p10)[20/20] | 50 | .ish (D12Z2x2/D12Z2x3)[25/30] | 54.5 | M              |
| 11      | –                                | 47,XY,+i(12)(p10) | 100 | .ish (D12Z2x2/D12Z2x3)[40/60] | 60 | M              |
| 12      | 46,XY                            | mos 46,XY/47,XY,+i(12)(p10)[25/5] | 16.6 | .ish (D12Z2x2/D12Z2x3)[7/25] | 78.1 | M              |
| 13      | 47,XY,+mar.ish(14/22)(q10;q10)mat | 48,XY,+mar.ish(14/22)(q10;q10)mat | 100 | –                                  |   | M              |
| 14      | –                                | mos 46,XY/47,XY,+i(12)(p10)[45/40] | 47 | .ish (D12Z2x2/D12Z2x3)[71/68] (F) | 49 | F              |
| 15      | 46,XY                            | –                                | mos.ish (D12Z2x2/D12Z2x3)[180/20] | 10 | B              | 51.4 | M              |

F Fibroblast, BS Buccal swap, M Maternal, P Paternal
| Case No | 1  | 2  | 3  | 4  | 5  | 6  | 7  | 8  | 9  | 10 | 11 | 12 | 13 | 14 | 15 |
|---------|----|----|----|----|----|----|----|----|----|----|----|----|----|----|----|
| Sex     | m  | f  | f  | f  | f  | f  | f  | f  | f  | m  | m  | m  | m  | m  | m  |
| Age at diagnosis (days/months (m)/year) | 7 | 10 | 6 | 12 | 10 | 2 | 10 | 2 | 11 | 8 | 1 | 10 | 22 | 11 | 1 |
| Gestation age at birth | ‘term’ | 38 | 37 | 40 | 39 | ‘term’ | ‘term’ | 36 | 33 | 41 | 37 | ‘term’ | 40 | 32 | 38 |
| Maternal/paternal age at conception | 18/25 | 32/33 | 22/22 | 23/28 | 38/38 | 19/25 | 35/2 | 31/3 | 21/23 | 33/31 | 28/29 | 39/33 | 32/32 | 33/41 |
| Weight/length at birth | 2100/7 | 3500/47 | 3500/50 | 2600/45 | 3450/54 | 4250/? | 3250/? | 2250/45 | 2500/44 | 4750/? | 2950/49 | 3800/ | 3200/53 | 2800/? | 3830/53 |
| Intellectual disability | + | + | + | + | + | + | + | + | - (near normal) | + | ? | + | ? | + | + |
| Seizures | – | – | – | – | – | – | – | – | – | – | – | – | – | – | – |
| Hypotonia | + | + | + | + | + | + | + | + | – | – | – | – | + | + | + |
| Prominent forehead | + | + | – | + | + | – | + | + | – | – | – | + | + | + | + |
| Bitemporal alopecia/sparse temporal hair | + | + | + | + | + | + | + | + | – | – | – | + | + | + | + |
| Coarse facial features | + | + | + | + | + | + | + | + | + | + | + | + | + | + | + |
| Flat occiput | + | – | – | – | – | – | – | – | – | – | – | – | – | – | – |
| Hypertelorism | + | + | – | + | + | – | + | + | – | – | – | + | + | + | + |
| Epicanthus | – | – | – | – | – | – | – | – | – | – | – | + | + | + | + |
| Dysmorphic ears | + | + | – | – | – | + | + | + | – | + | + | + | + | + | + |
| Short nose | – | – | – | + | – | + | – | + | – | – | – | – | + | + | + |
| Anteverted nostrils | + | – | + | + | + | + | + | + | + | + | + | + | + | + | + |
| Long philtrum | + | – | – | + | + | + | + | + | + | + | + | + | + | + | + |
| Macroglossia | – | – | – | – | – | – | – | – | – | – | – | – | – | – | – |
| Micrognathia | – | – | – | – | – | + | – | – | – | – | – | + | + | + | + |
| Everted lower lip | + | + | – | + | + | + | + | + | + | + | + | + | + | + | + |
| Short neck | – | + | + | + | + | + | + | + | – | – | – | + | + | + | + |
| Accessory nipples | – | – | – | + | – | – | – | – | – | – | – | – | – | – | – |
| Hypo/hyperpigmented areas of skin | +/+ | +/+ | +/+ | +/+ | +/+ | +/+ | +/+ | –/+ | –/+ | –/+ | –/+ | +/+ | +/+ | +/+ | –/+ |
| CHD | – | – | – | PDA, PFO, VSD | – | – | – | – | PDA | – | PFO, PDA | – | AS | – |

*Abbr: CHD congenital heart defect, PDA patent ductus arteriosus, PFO patent foramen ovale, VSD ventricular septal defect, ASD atrial septal defect*
Conclusion
Phenotypic and cytogenetic variability of PKS may be challenging for the differential diagnosis. Karyotyping on lymphocytes usually do not reveal the supernumerary i(12p). Therefore, we suggest that in cases with suspicion of clinical diagnosis of PKS, when pigmentation anomalies of the skin and temporal balding is noted as accompanying features, I-FISH on buccal mucosa cells and karyotyping from peripheral lymphocytes should be performed as a first tier test.

Abbreviations
CNS: Central Nervous System; FISH: fluorescence in situ hybridization; I-FISH: Interphase FISH; PKS: Pallister-Killian syndrome; STR: Short Tandem Repeats

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Availability of data and materials
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors’ contributions
BK, HK, AG, ZOU, GT, UA, and SB wrote the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate
These work were a part of routine clinical genetic evaluation. Written informed consent was obtained from the parents for all genetic tests.

Consent for publication
The parents of patients gave written informed consent allowing the publication of this manuscript and any accompanying images. A copy of the written consent is available and can be sent if desired.

Competing interests
The authors declare that they have no competing interests.

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