Basic Study

Combination immunotherapy with Survivin and luteinizing hormone-releasing hormone fusion protein in murine breast cancer model

Himani Garg, Rohit Singh Hada, Jagdish C Gupta, G P Talwar, Shweta Dubey

ORCID number: Himani Garg (0000-0002-4672-8289); Rohit Singh Hada (0000-0002-3236-3047); Jagdish C Gupta (0000-0002-9456-0097); G P Talwar (0000-0002-3421-1303); Shweta Dubey (0000-0002-0457-9692).

Author contributions: Gupta JC and Dubey S shared corresponding authorship; Garg H and Hada RS conducted and analysed the experiments; Gupta JC and Dubey S designed the study and wrote the manuscript; Talwar GP edited the manuscript.

Institutional animal care and use committee statement: All procedures involving animals were reviewed and approved by the Institutional Animal Care and Use Committee of the Amity Institute of Pharmacy, Amity University, Noida, Uttar Pradesh, India (IACUC protocol number: AIP/CPSC/PRO/01/2015).

Conflict-of-interest statement: To the best of our knowledge, no conflict of interest exists.

ARRIVE guidelines statement: The authors have read the ARRIVE guidelines, and the manuscript was prepared and revised according to the ARRIVE guidelines.

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Abstract

AIM To investigate the therapeutic potential of two recombinant proteins, Survivin and luteinizing hormone-releasing hormone (LHRH) fusion protein [LHRH(6leu)-LTB] for immunotherapy of breast cancer.

METHODS Murine 4T-1 breast cancer model was used to evaluate the efficacy of recombinant proteins in vivo. Twenty four Balb/c mice were divided into 4 groups of 6 mice each. Recombinant Survivin and LHRH fusion protein, alone or in combination, were administered along with immunomodulator Mycobacterium indicus pranii (MIP) in Balb/c mice. Unimmunized or control group mice were administered with phosphate buffer saline. Each group was then challenged with syngeneic 4T-1 cells to induce the growth of breast tumor. Tumor growth was monitored to evaluate the efficacy of immune-response in preventing the growth of cancer cells.

RESULTS Preventive immunization with 20 µg recombinant Survivin and MIP was effective in suppressing growth of 4T-1 mouse model of breast cancer (P = 0.04) but 50 µg dose was ineffective in suppressing tumor growth. However, combination of Survivin and LHRH fusion protein was more effective in suppressing tumor growth (P = 0.02) as well as metastasis in vivo in comparison to LHRH fusion protein as vaccine antigen alone.

CONCLUSION Recombinant Survivin and MIP suppress tumor growth significantly. Combining LHRH fusion protein with Survivin and MIP enhances tumor suppressive effects
marginally which provides evidence for recombinant Survivin and LHRH fusion protein as candidates for translating the combination cancer immunotherapy approaches.

Key words: Immunotherapy; Survivin; Luteinizing hormone-releasing hormone fusion protein; Combination immunotherapy; Breast cancer

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Core tip: Targeting Survivin for treatment of cancer is an emerging trend in cancer immunotherapy. In this study we optimized the dose of recombinant full-length Survivin for tumor growth inhibition. Since luteinizing hormone-releasing hormone (LHRH) is also known to support breast cancer in case of premenopausal women, we combined recombinant LHRH fusion protein with Survivin and *Mycobacterium indicus pranii* and obtained a positive anti-tumor response. We report our results of using recombinant Survivin protein and LHRH fusion protein as potential vaccine antigens for immunotherapy of murine model of breast cancer which was developed by injecting 4T-1 mammary cell line in syngeneic Balb/c mice. Immunized mice challenged with syngeneic 4T-1 cells exhibited suppression of tumor growth and metastasis in lungs in comparison to control mice.

INTRODUCTION

Combination methods that combine two or more therapeutic agents have become a principal treatment modality for breast cancer, especially for metastatic breast cancer[1]. The rationale behind combination approach is to enhance therapeutic response by targeting multiple pathways involved in the complex process of oncogenesis. However, none of the empirically defined combination of drugs such as taxanes/anthracyclins has been able to overcome the problems of drug resistance and metastasis in breast cancer and impart a satisfactory toxicity profile when administered to patients[2,3]. In order to overcome the drawbacks associated with prevalent drug combinations, immunotherapy is proposed as a viable approach for developing targeted and safe treatment options for cancer[4,5]. Advancements in molecular mechanisms of cancer cell survival have led to identification of new targets which can be employed for immunotherapy[6].

Cancer immunotherapy approach using immunomodulators, monoclonal antibodies or vaccines against tumor growth promoters, have shown benefit in preclinical models of many cancers and clinical trials[7-9]. However since tumor cells often develop immune evasion mechanisms[9], combination of tumor antigens may be used to counteract immune resistance by tumor cells. We hypothesize that combination immunotherapy based on potent tumor antigen and hormone, may present a viable approach to enhance the therapeutic landscape and effectively counteract the heterogeneous process of tumor development in breast cancer.

Survivin is a tumor antigen exclusively expressed on tumor cells, essential for cancer cell survival, and its overexpression is associated with aggressiveness of the disease[10,11]. It has been widely explored as a candidate antigen for cancer immunotherapy[12]. Similarly, many tumor cells from breast, prostate, ovary or endometrial origin are hormone dependent for their survival[13]. Therefore, immune system mediated neutralization of hormones is also a key therapeutic strategy for hormone dependent cancers[14,15]. Immunization against hormones such as luteinizing hormone-releasing hormone (LHRH) using LHRH based peptide vaccines have shown efficacy as vaccine antigens in preclinical models of hormone dependent cancers such as breast and prostate cancer[16].

We have previously shown that preventive immunization with a combination of recombinant Survivin protein and an immunomodulator *Mycobacterium indicus pranii*
(MIP) provides protection to Balb/c mice when challenged with syngeneic mammary tumor 4T-1 cells\textsuperscript{[16]}. This communication describes the studies undertaken to determine the optimum dose of Survivin antigen for maximum tumor suppressive effect. Also investigated is the synergy between Survivin and LHRH fusion protein as vaccine antigens for immunotherapy of murine model of breast cancer. 4T-1 murine breast cancer cells not only display Survivin as tumor antigen but also have receptors for LHRH\textsuperscript{[17,18]}. Thus active immunization against both the Survivin and LHRH, is likely to neutralize Survivin and also make LHRH unavailable for uptake by the cancer cells, hence generate enhanced protective immune response directed against the cancer cells.

\textbf{MATERIALS AND METHODS}

\textit{Cloning and expression of recombinant mouse Survivin and LHRH fusion protein}

Survivin and LHRH fusion protein, were used as vaccine antigens. These antigens were expressed as recombinant proteins using \textit{E. coli} based host-vector systems as described previously\textsuperscript{[16,19]}. Survivin and LHRH fusion protein were purified as approximately 17 kDa and 14 kDa size proteins, respectively using affinity based Ni-NTA chromatography. Purified proteins were adsorbed on alum before administration as immunogens in mice. MIP was used as immunomodulator along with the recombinantly made vaccine antigens.

\textit{Immunogenicity and efficacy of Survivin vaccine in vivo}

Eight-10 week old inbred Balb/c mice were used for conducting the immunogenicity and efficacy studies of the Survivin vaccine alone in vivo. For determining the optimal Survivin dose, study was carried out in 3 groups of mice. Mice were divided into 3 groups randomly with 6 animals in each group. Group 1 was administered phosphate buffer saline (PBS) buffer. While group 2 mice was administered alum adsorbed 20 µg Survivin vaccine (+ MIP), group 3 mice received alum adsorbed 50 µg Survivin dose along with MIP. Five \times 10^6 cells of heat-killed MIP were used in each dose wherever required. Each group of mice was given 2 boosters of its respective dose at 15 d interval after the primary immunization. Sixty days after the last immunization, mice in all the groups were challenged with 3.0 \times 10^5 4T-1 murine breast cancer cells subcutaneously. Third booster of the vaccine was administered on day 18th after the challenge, when tumors were palpable. Tumor size was measured every alternate day till the end of the study. Tumor volume was measured bi-dimensionally with digital vernier calipers and tumor volume calculated using the formula: (a \times b^2)/2, where “a” is the largest diameter and “b” is the perpendicular diameter. Tumor volumes were measured till day 35 post challenge with 4T-1 cells. Results are expressed as mean ± SD of tumor volume for each group.

\textit{Immunogenicity and efficacy of combination of Survivin and LHRH fusion protein in comparison to Survivin and LHRH fusion protein alone}

Studies were designed in 4 groups of mice each consisting of 6 of 8-10 week old inbred Balb/c mice. Recombinant antigen(s) were administered intramuscularly along with heat-killed 5.0 \times 10^6 cells of MIP. Group 1 of mice received 20 µg of Survivin protein alone, group 2 received 20 µg of LHRH(6leu)-LTB (LTB- heat-labile enterotoxin of \textit{E. coli}) protein alone and group 3 received combination of 20 µg of Survivin and 20 µg of LHRH(6leu)-LTB proteins. Each group of mice was given 2 boosters of its respective vaccine dose at 15 d interval after the primary immunization. A control group of mice was administered PBS buffer. Fifty-two days after first dose, mice were inoculated with 4T-1 breast cancer cells (3.0 \times 10^5 cells in 100 µL of PBS buffer) subcutaneously on right flank. It was recorded as day 0 of tumor challenge. Tumors were palpable in mice in 18 d after tumor cell inoculation. Third booster of vaccine antigen(s) along with immunomodulator MIP was given intramuscularly at the same day when tumors were palpable. Tumor volume was measured till the end of the study. Mice were sacrificed at the end of the study and lungs were examined for the presence of pulmonary metastases.

\textit{Estimation of Interferon-γ levels in sera}

Sera were collected from mice and assayed for Interferon gamma levels using commercially available enzyme linked immunosorbent assay (ELISA) kit (E Biosciences, United States) as per the manufacturer’s protocol. Briefly, 96-well microtitre plate was coated with 100 µL/well of IFN-γ capture antibody at a dilution of 1:1000 and incubated overnight at 4 °C. Plates were then washed thrice with PBS buffer (PBS buffer supplemented with 0.05% Tween-20) and then blocked with ELISA diluent (250 µL/well) at 37 °C for 2 h. Standard was serially diluted in ELISA diluent
to obtain concentration in the range of 15-2000 pg/mL and 100 µL of standard of each concentration was added to the corresponding wells. Serum samples were pooled for each group of mice, diluted in ELISA diluent and added to the respective wells at 100 µL/well. Plate was then incubated for 2 h at 37 °C. Biotinylated anti-IFN-γ detection antibody was added to each well (100 µL/well) at a dilution of 1:1000 followed by incubation of 100 µL/well of Avidin-HRP diluted at 1:250 for 30 min. After washing, TMB was added in each well and incubated in dark for 15 min. Reaction was stopped with 2N H$_2$SO$_4$, and absorbance was read at 450 nm. Standard curve was prepared by plotting the average absorbance of each standard on the vertical axis versus the corresponding IFN-γ standard concentration on the horizontal axis and fitted using 4 parameter logistic (4PL) regression analysis. Actual concentration of IFN-γ in mouse serum samples was calculated by extrapolating OD values using the 4PL curve.

**ELISA**

ELISA technique was used for the analysis of immunogenicity of the recombinant proteins in mice. ELISA plate (Nunc, Thermo Fisher Scientific, United States) was coated with 100 µL (5 µg/mL concentration) of recombinant Survivin or LHRH(6leu)-LTB protein, and incubated overnight at 4 °C. Plate was washed thrice with 10 mmol/L PBS, pH 7.4 supplemented with 0.05% Tween 20 followed by blocking with 2% Bovine serum albumin (BSA) in PBS buffer. Plate was washed with PBST buffer thrice. One hundred micro litre of the serum diluted at 1:5000 in PBS buffer, was added to the wells of the plate, and incubated for 1 h at 37°C. HRP-labeled goat anti-mouse antibody was added at a dilution of 1:10000 to each well and color was developed by adding 100 µL of O-phenylenediamine (OPD) solution containing Hydrogen Peroxide. The reaction was stopped by addition of 50 µL of 2N H$_2$SO$_4$. Plate was read at 492 nm in an ELISA plate reader (ELX 800MS, Biotek, United States) and results were expressed as mean ± SD of absorbance values.

**Ethics statement**

Pathogen free female Balb/c mice used in the study were procured from National Institute of Nutrition, Hyderabad. Animals were housed at animal facility of Amity University Uttar Pradesh and were given ad libitum access to water and food. All experimental procedures were in strict agreement with committee for the purpose of control and supervision of experiments on animals (CPCSEA) guidelines for the care and use of laboratory animals and approved by institutional animal ethics committee (IAEC) of Amity University Uttar Pradesh.

**Statistical analysis**

Data was analyzed using two tailed student’s t test. $P$ value < 0.05 was reported as significant. Data was analyzed using graph pad prism software.

**RESULTS**

**Optimum dose of Survivin antigen dose for the effective inhibition of tumor growth in 4T-1 model**

We reported previously that the recombinant Survivin vaccine is efficacious in preventing the growth of 4T-1 based murine model of breast cancer[16]. Increase in vaccine dose from 5 µg to 10 µg, led to better immunogenicity which resulted in significant reduction in tumor suppression[16]. Therefore, it was decided to test if it was possible to employ doses higher than previously used 10 µg dose to inhibit the tumor maximally. Thus the studies were designed to evaluate the efficacy of the vaccine at 20 µg and 50 µg doses. Pathogen-free mice of similar weight (6 mice/group) were first immunized with alum adsorbed recombinant Survivin vaccine along with immune-modulator MIP. Mice were then challenged with syngeneic 4T-1 cells and observed for the tumor growth in presence of pre-formed anti-Survivin antibodies. Three immunizations of alum adsorbed recombinant Survivin and MIP were administered at 15 d interval following which 4T-1 cancer cells were injected at day 90. A booster of the Survivin vaccine was given at day 108 when tumors were palpable. Tumor volume was measured and continued till 35 d post injection of cancer cells. Figure 1 shows the volume of the tumor developed in mice immunized with 20 µg and 50 µg of Survivin protein administered along with MIP. It is evident from the Figure that significant decrease in tumor volume was observed in mice immunized with 20 µg dose of the vaccine ($P = 0.04$) in comparison to control mice. However increase in vaccine dose to 50 µg did not lead to any significant inhibition in tumor volume thus reflecting dose-inhibition related effects. Figure 2 shows the IFN-γ levels determined in the sera collected from immunized
mice. Survivin at 20 µg dose induced higher levels of IFN-γ in comparison to 50 µg dose, which is in accordance with the observed efficacy of the anti-Survivin antibodies in preventing the growth of tumor at 20 µg dose. The mice in each group did not show any visible adverse effects of the treatment.

**Active immunization against a combination of Survivin and LHRH fusion protein**

After finding the optimal dose of Survivin vaccine, studies were advanced to investigate the additive effect of LHRH vaccine, if any, in terms preventing the 4T-1 breast cancer cells. Studies were carried out in 4 groups of pathogen free mice (6 mice/group). A group of mice was immunized by combination of alum adsorbed Survivin and LHRH(6leu)-LTB proteins along with immunomodulator MIP. Two groups of mice were injected with Survivin alone (+ MIP) and LHRH(6leu)-LTB alone (+ MIP), respectively. Mice receiving the PBS buffer formed the control group. Mice were immunized thrice at 15 d interval with combination or individual antigens following which 4T-1 tumor cells were injected at 50th day. Boosters of the vaccine antigens were administered when the tumors became palpable. The tumor size was measured post 35 d of tumor cell challenge. Though LHRH(6leu)-LTB alone prevented the growth of tumor size in comparison to control group mice but did not reach statistical significance, inhibition was more significant with Survivin alone ($P = 0.04$). Combining it with LHRH fusion protein led to marginally better tumor suppression, $P = 0.02$ (Figure 3). There were no adverse effects of the treatment observed in the mice of each group.

**Evaluation of metastasis of primary tumor to lungs**

Mice challenged with the tumor cells were followed to evaluate the efficacy of the active immunization by Survivin and LHRH fusion protein alone or in combination in terms of preventing cancer metastasis. The secondary growth of the cancer was studied by counting the number of nodules in the lungs of the mice challenged with the tumor cells. Table 1 shows the mean of number of nodules counted in various groups at the end of the study. Mice immunized with the combination of Survivin and LHRH(6leu)-LTB along with MIP did not show any visible nodules in the lungs 30 d after challenge with 4T-1 cells whereas mice immunized with either Survivin + MIP or LHRH(6leu)-LTB + MIP showed presence of tumor nodules in lungs. Thus combination of Survivin and LHRH(6leu)-LTB along with immunomodulator MIP is capable of controlling the pulmonary metastasis besides suppressing the primary tumor growth.

**Characterization of total IgG response**

Sera collected from each group of mice were checked for the total IgG titre against the respective antigen(s) pre and post challenge with tumour cells. Both Survivin and LHRH(6leu)-LTB antigens were able to induce antibody response when used to immunize the mice alone. Combined immunization with both the antigens, also led to induction of both anti-Survivin and anti-LHRH antibodies thus confirming their immunogenicity (Figure 4).

**DISCUSSION**

Tumor antigens have been proposed as important therapeutic targets for cancer immunotherapy[20]. Immunotherapy approaches incorporating Survivin as tumor antigen have shown efficacy for elimination of cancer cells. Various approaches such as peptide vaccines[21-23], multi epitope vaccines[24,25] or DNA vaccines incorporating Survivin as vaccine antigen[26,27], have shown efficacy in preclinical studies. It has also been suggested that combination of Survivin derived peptides with immunomodulators or immunomodulatory cytokines such as IFN alpha induces a significant overall survival in patients with advanced or recurrent urothelial cancer[28]. We have also previously reported that combination of Survivin along with MIP has potent dose dependent anti-tumor effects in a mouse model of breast carcinoma[16].

Our data suggests that MIP alone is not effective in preventing tumor growth (data not shown) however combining MIP with tumor antigen such as Survivin gives significant tumor suppression. MIP may work as immunomodulator and serves to enhance the immune response against Survivin antigen.

We have demonstrated in the present study that increasing the dose of recombinant Survivin to 20 µg induces most effective tumor suppression in murine 4T-1 model of breast cancer. Interestingly, further increasing the dose of antigen does not induce a tumor suppressive effect. Suppression of immune response at supraoptimal doses of antigen has also been previously reported[29-31]. We hypothesize that high dose Survivin antigen may induce a suboptimal or anergic immune response which may
not be able to inhibit the tumor growth.

Interferon-γ secreted by Th1 CD4+ T, NK cells, CD8+ T cells, APCs and B-cells is known for its anti-tumor effects\cite{32}. It has role in rejection of transplanted tumors and inhibition of formation of spontaneous tumors\cite{32,33}. Data presented in the present studies also show higher levels of IFN-γ at 20 µg dose of Survivin than the 50 µg dose which is in agreement with observed better suppression of tumor with 20 µg dose. Previous studies have confirmed the important role of humoral immune response in providing anti-tumor immunit\cite{34}. Our studies also indicate that antigen specific antibodies were induced after immunization with tumor antigens which possibly have contributed to tumor suppression. A recent study by Fenstermaker et al.\cite{36} has also demonstrated therapeutic effect of monoclonal antibody against Survivin in vivo in murine glioma model.

Many of the premenopausal breast cancers are also hormone-dependent which rationalizes the use of LHRH agonists as an effective treatment option in breast cancers\cite{37}. Various combinations of LHRH agonists such as cisplatin loaded LHRH nanoparticles which have shown good anti-tumor response in 4T1 breast tumor model by accumulating higher cisplatin in tumors\cite{18}. Another conjugate of LHRH, Pt-Mal-LHRH (activated cisplatin linked to LHRH peptide with a malonate linker) showed targeted delivery to cells expressing LHRH receptors. Treatment of 4T1 breast cancer with Pt-Mal-LHRH reduced tumor volume and metastasis to lungs\cite{38}.

We evaluated the possibility of using an immunotherapy based on a combination of Survivin and LHRH fusion protein in 4T-1 murine model of breast cancer. LHRH(6leu)-LTB, a vaccine developed for the immunotherapy of androgen dependent cancers, was used as an immunogen in combination with Survivin along with immunomodulator, MIP. The idea was to generate anti-LHRH antibodies to neutralize LHRH making it unavailable for intake by the breast cancer cells through the receptors. It was observed that efficacy of the combination was marginally better than the Survivin alone but significantly better when only LHRH(6leu)-LTB was used along with MIP. More importantly, combination was best at preventing metastasis of primary tumors to lungs.

Our results show promising outcome in a prophylactic scenario. We undertook the prophylactic approach for two reasons: (1) to investigate if the immunization induces a tumor protective response in vivo; and (2) this prophylactic approach also closely mimics the clinical situation where primary tumors have been removed or adjuvant treatment for cancer patients has been initiated. In both these settings, the patients undergo surgery. Though patients are rendered free of tumor, chances of recurrence are very high. If translation of LHRH fusion protein and Survivin based combination approach is successful, it may find applications in such clinical scenario. We have also investigated the therapeutic potential of this approach by immunizing the mice carrying palpable 4T-1 tumor, which has showed similar suppression of tumor growth (data not shown).

In conclusion, we have shown the Survivin (+ MIP) at a dose of 20 µg is most effective in preventing the growth of 4T-1 breast tumor cells in mice. Incorporation of anti-LHRH vaccine exercises a synergistic effect. A schematic diagram depicting the strategy used in the present study has been shown as Figure 5. Although our results show the protective role of combination immunotherapy with Survivin and LHRH antigens in murine tumor model, yet the study is limited by the fact the results need to be validated in other tumor models and further translational development needs to
be undertaken for appropriate human application. Though it did not lead to much benefit in preventing the growth of primary tumor, it was highly effective in blocking the pulmonary metastasis. Our data suggests that combination of Survivin and LHRH fusion protein may hold immense promise for further development of immunotherapeutic approaches in management of breast cancers. The combination enhances immune response that is involved in inhibiting tumor growth, thus it will be effective in immune competent organisms and has to be supplemented with other therapies for use in immune compromised individuals. Furthermore, investigating the molecular mechanism of action of the combination leading to tumor inhibition may also lead to development of novel targeted therapies for cancer.
Table 1  Number of nodules in the lungs of mice challenged with 4T-1 breast cancer cells post immunization with PBS buffer, Survivin, LHRH(6leu)-LTB and combination of Survivin and LHRH(6leu)-LTB

| Immunization group                  | Average No. of lung nodules |
|------------------------------------|-----------------------------|
| PBS (untreated)                     | > 20                        |
| Survivin                           | 9                           |
| LHRH(6leu)-LTB                     | 9                           |
| Survivin + LHRH(6leu)-LTB          | 0                           |

LHRH: Luteinizing hormone-releasing hormone.

Figure 3  4T-1 Breast tumor volume in mice immunized with PBS buffer, 20 µg Survivin + Mycobacterium indicus pranii, 20 µg luteinizing hormone-releasing hormone + Mycobacterium indicus pranii and combination of 20 µg Survivin and 20 µg luteinizing hormone-releasing hormone along with Mycobacterium indicus pranii. Tumor volume is measured at different time points from day 19<sup>th</sup> when tumor was palpable. LHRH: Luteinizing hormone-releasing hormone.

Figure 4  Graph representing the comparative level of total IgG antibody measured in mice immunized with Survivin (+ Mycobacterium indicus pranii), LHRH(6leu)-LTB (+ Mycobacterium indicus pranii) and combination of Survivin and LHRH(6leu)-LTB (+ Mycobacterium indicus pranii). The serum was collected pre tumor challenge (Day -7), on tumor challenge (Day 0) and at the end of the study (Day 38). The total IgG titer was determined against respective antigens in each group. Groups of mice receiving Survivin alone and combination were checked for anti-Survivin antibody titers. Similarly, anti-LHRH titres were checked in mice in groups corresponding to LHRH(6leu)-LTB alone and combination. LHRH: Luteinizing hormone-releasing hormone.
Figure 5  Schematic representation of the study of anti-tumor effect of Survivin, luteinizing hormone-releasing hormone fusion protein individually and in combination. The study was conducted on four groups of mice: Untreated or control, Survivin alone, LHRH(6leu)LTB alone and Survivin + LHRH(6leu)LTB. Individual recombinant proteins were purified and used as immunogen along with Mycobacterium indicus pranii as an immunomodulator in murine model. The mice from all the groups were further challenged with tumor cells, 4T1 and tumor development was observed in each group. LHRH fusion protein was not effective in suppressing tumor alone. However, the combination of both Survivin and LHRH(6leu)LTB inhibited tumor growth substantially followed by Survivin alone. LHRH: Luteinizing hormone-releasing hormone.

ARTICLE HIGHLIGHTS

Research background
Tumor cells often develop immune evasion mechanisms, thus combination of tumor antigens may be used to counteract immune resistance. Survivin is a tumor antigen exclusively expressed on tumor cells, essential for cancer cell survival, and its overexpression is associated with aggressiveness of the disease. Similarly, many tumor cells from breast, prostate, ovary or endometrial origin are hormone dependent for their survival. Immunization against hormones such as luteinizing hormone-releasing hormone (LHRH) using LHRH based peptide vaccines have shown efficacy as vaccine antigens in preclinical models of hormone dependent cancers such as breast and prostate cancer. This communication describes the studies undertaken to determine the optimum dose of Survivin antigen for maximum tumor suppressive effect and the synergy between Survivin and LHRH as vaccine antigens for immunotherapy of murine model of breast cancer, 4T-1.

Research motivation
We undertook the prophylactic approach for two reasons: (1) to investigate if the immunization induces a tumor protective response in vivo and (2) this prophylactic approach also closely mimics the clinical situation where primary tumors have been removed or adjuvant treatment for cancer patients has been initiated. Major issues in cancer treatment are the recurrence of tumor after surgery and chemoresistance. Our study was aimed at targeting these issues and developing a strategy, which may prevent tumor spread and recurrence and can overcome resistance. Generating immune response in the body against the tumor antigens will be a safer way of treatment. Anti-Survivin approach may help in overcoming the problem of resistance to therapy.

Research objectives
The aim of this study was to evaluate the possibility of using an immunotherapy based on a combination of Survivin and LHRH fusion protein in 4T-1 murine model of breast cancer. LHRH(6leu)-LTB was used as an immunogen in combination with Survivin along with an immunomodulator, Mycobacterium indicus pranii (MIP). It was observed that efficacy of the combination was marginally better than the Survivin alone but significantly better when only LHRH(6leu)-LTB was used along with MIP. More importantly, combination was best at preventing metastasis of primary tumors to lungs. In most cancer cases, the patients undergo surgery. Though patients are rendered free of tumor, chances of recurrence are very high. If translation of LHRH fusion protein and Survivin based combination approach is successful, it may find applications in such clinical scenario.

Research methods
Survivin and LHRH fusion protein were used as vaccine antigens. These antigens were expressed as recombinant proteins using E. coli based host-vector systems. Purified proteins were adsorbed on alum before administration as immunogens in mice. MIP was used as immunomodulator along with the recombinantly made vaccine antigens. Inbred Balb/c mice were used for conducting the immunogenicity and efficacy studies of the Survivin vaccine in vivo. Tumor volume was measured bi-dimensionally with digital vernier calipers and results are expressed as mean ± SD of tumor volume for each group. Anti-tumor efficacy of combination of Survivin and LHRH fusion protein in comparison to Survivin and LHRH fusion protein alone was also determined. Sera were collected from mice and assayed for Interferon gamma levels using ELISA kit. ELISA was also performed for the analysis of immunogenicity of the recombinant proteins in mice.

Research results
We have shown that Survivin (+ MIP) at a dose of 20 µg is most effective in preventing the growth of 4T-1 breast tumor cells in mice and incorporation of anti-LHRH vaccine exercises a synergistic effect. The study is limited by the fact that the results need to be validated in other tumor models and further translational development needs to be undertaken for appropriate human application. The combination will be effective in immune competent organisms and has to be supplemented with other therapies for use in immune compromised individuals. Furthermore, investigating the molecular mechanism of action of the combination leading to tumor inhibition may also lead to development of novel targeted therapies for cancer.

Research conclusions
Our results show the protective role of combination immunotherapy with Survivin and LHRH antigens in murine tumor model. Though it did not lead to much benefit in preventing the growth of primary tumor, it was highly effective in blocking the pulmonary metastasis. Combination of Survivin and LHRH fusion protein may hold immense promise for further development of immunotherapeutic approaches in management of breast cancers.

REFERENCES
1. Zanardi E, Bregni G, de Braud F, Di Cosimo S. Better Together: Targeted Combination Therapies in Breast Cancer. Semin Oncol 2015; 42: 887-895 [PMID: 26615133 DOI: 10.1055/s-0035-1556323]
2. Biganzoli L, Cufer T, Bruning P, Coleman R, Ducheateau L, Calvert AH, Gamucci T, Twelves C, Fargeot P, Epelbaum R. Doxorubicin and paclitaxel versus doxorubicin and cyclophosphamide as first-line chemotherapy in metastatic breast cancer: The European Organization for Research and Treatment of Cancer 10961 Multicenter Phase III Trial. J Clin Oncol 2002; 20: 3114-3121 [PMID: 12118025 DOI: 10.1200/JCO.2002.11.085]
3. Jassem J, Pietrowski T, Phuzantisa A, Jelic S, Corbunova V, Mrsc-Krmotic Z, Berzins J, Nagykalna T, Wigler N, Renard J, Doxorubicin and paclitaxel versus fluorouracil, doxorubicin, and cyclophosphamide as first-line therapy for women with metastatic breast cancer: final results of a randomized phase III multicenter trial. J Clin Oncol 2001; 19: 1707-1715 [PMID: 11251000 DOI: 10.1200/JCO.2001.19.6.1707] 
4. Morrissey KM, Yuraszeck TM, Li CC, Zhang Y, Kasichayanula S. Immunotherapy and Novel Combinations in Oncology: Current Landscape, Challenges, and Opportunities. Clin Transl Sci 2016; 9: 89-104 [PMID: 26924066 DOI: 10.1111/cts.12391]
5. Khalil DN, Smith EL, Brentjens RJ, Wolchok JD. The future of cancer treatment: immunomodulation, CARs and combination immunotherapy. Nat Rev Clin Oncol 2016; 13: 394 [PMID: 27118494 DOI: 10.1038/nrclinonc.2016.65]
6. Wang RF, Wang HY. Immune targets and neoantigens for cancer immunotherapy and precision medicine. Cell Res 2017; 27: 11-37 [PMID: 28025978 DOI: 10.1038/cr.2016.159]
7. Farkoma S, Diamandis EP, Blasutig IM. Cancer immunotherapy: the beginning of the end of cancer? BMC Med 2016; 14: 73 [PMID: 27151159 DOI: 10.1186/s12916-016-0622-5]
8. Weiner LM, Surana R, Wang S. Monoclonal antibodies: versatile platforms for cancer immunotherapy. Nat Rev Immunol 2010; 10: 317-327 [PMID: 20414205 DOI: 10.1038/nri2744]
9. Vinay DS, Ryan EP, Pawelec G, Talib WH, Stagg J, Elkord E, Lichter T, Deckert WK, Whelan RL, Kumara HMCS. Immune evasion in cancer: Mechanistic basis and therapeutic strategies. Semin Cancer Biol 2015; 35 Suppl: S185-S198 [PMID: 25818339 DOI: 10.1016/j.semcancer.2015.03.004]
10. Cheng KY, Wang ZL, Gu QY, Hao M. Survivin Overexpression Is Associated with Aggressive Clinicopathological Features in Cervical Carcinoma: A Meta-Analysis. PLoS One 2016; 11: e0155177 [PMID: 27764226 DOI: 10.1371/journal.pone.0155177]
11. Chuwa AH, Sone K, Oda K, Ikeda Y, Fukuda T, Wada-Hiraike O, Inaba K, Makii C, Takeuchi M,
Oki S. Significance of survivin as a prognostic factor and a therapeutic target in endometrial cancer. Gynecol Oncol 2016; 141: 564-569 [PMID: 2709211 DOI: 10.1016/j.ygyno.2016.04.003]

Boullisa LF, Savaila P, Bonney S, Orchard L, Wickenden H, Lee C, Smiths E, Banham AH, Mills KJ, Orchard K. Identification of survivin as a promising target for the immunotherapy of adult B-cell acute lymphoblastic leukemia. Oncotarget 2017; 9: 3853-3866 [PMID: 29423088 DOI: 10.18632/oncotarget.23380]

Capper CP, Rae JM, Auchus RJ. The Metabolism, Analysis, and Targeting of Steroid Hormones in Breast and Prostate Cancer. Horm Cancer 2016; 7: 149-164 [PMID: 26985690 DOI: 10.1017/hoc.2016.11]

Junco JA, Basalto R, Fuentes F, Bover E, Reyes O, Pimentel E, Calzada L, Castro MD, Arteaga N, López Y. Gonadotropin releasing hormone-based vaccine, an effective candidate for prostate cancer and other hormone-sensitive neoplasms. Adv Exp Med Biol 2008; 617: 381-387 [PMID: 18497385 DOI: 10.1007/978-0-387-69080-3_60]

Junco JA, Poschke P, Zuna I, Ehemann V, Fuentes F, Bover E, Pimentel E, Basalto R, Reyes O, Calzada L. Immunotherapy of prostate cancer in a murine model using a novel GnRH based vaccine candidate. Vaccine 2007; 25: 8460-8468 [PMID: 18022737 DOI: 10.1016/j.vaccine.2007.09.059]

Garg H, Gupta JC, Talwar GP, Dubey S. Immunotherapy approach with recombinant survivin adjuvanted with alum and MIP suppresses tumor growth in murine model of breast cancer. Prep Biochem Biotechnol 2018; 48: 264-269 [PMID: 29355462 DOI: 10.1080/10826069.2018.1425710]

Tahter A, Dinarvand R, Aghadi F, Khormalizadeh MR, Atyabi F. The in vivo antitumor activity of LHRRGD targeted methotrexate-human serum albumin nanoparticles in 4T1 tumor-bearing Balb/c mice. Int J Pharm 2012; 431: 183-189 [PMID: 22531853 DOI: 10.1016/j.ijpharm.2012.04.031]

Li M, Tang Z, Zhang Y, Lv S, Li Q, Chen X. Targeted delivery of cisplatin by LHRRGD-peptide conjugated dextran nanoparticles suppresses breast cancer growth and metastasis. Acta Biomater 2015; 18: 132-143 [PMID: 25735801 DOI: 10.1016/j.actbio.2015.02.022]

Gupta JC, Hada RS, Saihat P, Talwar GP. Development of a novel recombinant LHRRGD fusion protein for therapy of androgen and estrogen dependent cancers. Protein Expr Purif 2017; 134: 132-138 [PMID: 28410993 DOI: 10.1016/j.pep.2017.04.003]

Srinivasan R, Wolchok JD. Tumor antigens for cancer immunotherapy: therapeutic potential of xenogeneic DNA vaccines. J Transl Med 2004; 2: 12 [PMID: 15090064 DOI: 10.1016/S1479-5876(02)00123-1]

Ciesielksi MJ, Kozbor D, Castanaro CA, Barone TA, Fenstermaker RA. Therapeutic effect of a T helper cell supported CTL response induced by a survivin peptide vaccine against murine cerebral glioma. Cancer Immunol Immunother 2008; 57: 1827-1835 [PMID: 18436666 DOI: 10.1007/s00262-008-0510-9]

Fenstermaker RA, Ciesielksi MJ, Qiu J, Yang N, Frank CL, Lee KP, Mechtler LR, Belal A, Ahluwalia MS, Hutson AD. Clinical study of a survivin long peptide vaccine (SurVaxM) in patients with recurrent malignant glioma. Cancer Immunol Immunother 2016; 65: 1339-1352 [PMID: 27576783 DOI: 10.1007/s00262-016-1808-x]

Yang Z, Wang L, Wang H, Shang X, Niu W, Li J, Wu Y. A novel mimovirus vaccine containing survivin epitope with adjuvant IL-15 induces long-lasting cellular immunity and high antitumor efficiency. Mol Immunol 2008; 45: 1674-1681 [PMID: 18053418 DOI: 10.1016/j.molimm.2007.10.026]

Idenoue S, Hirohashi T, Torigoe T, Sato Y, Tamura Y, Harui H, Yamamoto M, Kurotaki T, Tsuruma T, Asanuma H. A potent immunogenic general cancer vaccine that targets survivin, an inhibitor of apoptosis proteins. Clin Cancer Res 2005; 11: 1474-1482 [PMID: 15746049 DOI: 10.1158/1078-0432.CCR-03-0817]

Lennerz V, Gross S, Gallerani E, Sesso C, Mach N, Boehm S, Hess D, von Boehm L, Knuth A, Ochsnebein AF. Immunologic response to the survivin-derived multi-epitope vaccine EMD640744 in patients with advanced solid tumors. Cancer Immunol Immunother 2014; 63: 381-394 [PMID: 24487961 DOI: 10.1007/s00262-013-1516-5]

Ladser A, Ljungberg K, Tufvesson H, Tazzani M, Roos AK, Quest AF, Kiessling R. Intradermal DNA electroporation induces survivin-specific CTLs, suppresses angiogenesis and confers protection against mouse melanoma. Cancer Immunol Immunother 2010; 59: 81-92 [PMID: 19526360 DOI: 10.1007/s00262-009-0725-4]

Xiang R, Mizutani N, Luo Y, Chiodoni C, Zhou H, Mizutani M, Ba Y, Becker JC, Reisfeld RA. A DNA vaccine targeting survivin combines apoptosis with suppression of angiogenesis in lung tumor eradication. Cancer Res 2005; 65: 533-561 [PMID: 15605399]

Tanaka T, Kitamura H, Inoue R, Nishida S, Takahashi-Takaya A, Kawami S, Torigoe T, Hirohashi Y, Tsukamoto T, Sato N. Potential survival benefit of anti-apoptosis protein: survivin-derived peptide vaccine with and without interferon alpha therapy for patients with advanced or recurrent uterine carcinoma—results from phase I clinical trials. Clin Deo Immunol 2013; 2013: 262967 [PMID: 24363758 DOI: 10.11155/2013/262967]

Gotsman I, Israeliti D, Alper R, Rabani E, Engelhardt D, Ilan Y. Induction of immune tolerance toward tumor-associated-antigens enables growth of human hematoma in mice. Int J Cancer 2002; 97: 52-57 [PMID: 11774243 DOI: 10.1002/ijc.1576]

Suzuki G, Kawase Y, Koyasu S, Yahara I, Kobayashi Y, Schwartz RH. Antigen-induced suppression of the proliferative response of T cell clones. J Immunol 1988; 140: 1359-1365 [PMID: 2456125]

Michalet MC, Saltel F, Flacher M, Revillard JP, Genestier L. Cathepsin-dependent apoptosis triggered by supraoptimal activation of T lymphocytes: a possible mechanism of high dose tolerance. J Immunol 2004; 172: 5405-5414 [PMID: 15100281 DOI: 10.4049/jimmunol.172.9.5405]

Ikedo H, Old LJ, Schreiber RD. The roles of IFN-gamma in protection against tumor development and cancer immunotherapy. Cytokine Growth Factor Rev 2002; 13: 95-109 [PMID: 11909366 DOI: 10.1016/S1359-6101(01)00038-7]

Blankenstein T, Qin Z. The role of IFN-gamma in tumor transplantation immunity and inhibition of chemical carcinogenesis. Curr Opin Immunol 2003; 15: 148-154 [PMID: 12633663 DOI: 10.1016/S0952-7915(03)00047-4]

Dunn GP, Old LJ, Schreiber RD. The immunobiology of cancer immunosurveillance and immunoeediting. Immunity 2004; 21: 137-148 [PMID: 15380995 DOI: 10.1016/j.immuni.2004.06.012]
35 Tsou P, Katayama H, Ostrin EJ, Hanash SM. The Emerging Role of B Cells in Tumor Immunity. Cancer Res 2016; 76: 5597-5601 [PMID: 27634765 DOI: 10.1158/0008-5472.CAN-15-3421] 
36 Fenstermaker RA, Figel SA, Qiu J, Barone TA, Dharma SS, Winograd EK, Galbo PM, Wiltsie LM, Ciesielski MJ. Survivin Monoclonal Antibodies Detect Survivin Cell Surface Expression and Inhibit Tumor Growth In Vivo. Clin Cancer Res 2018; 24: 2642-2652 [PMID: 29540489 DOI: 10.1158/1078-0432.CCR-17-2778] 
37 Goel S, Sharma R, Hamilton A, Beith J. LHRH agonists for adjuvant therapy of early breast cancer in premenopausal women. Cochrane Database Syst Rev 2009; 4: CD004562 [PMID: 19821328 DOI: 10.1002/14651858.CD004562.pub4] 
38 Calderon LE, Keeling JK, Rollins J, Black CA, Collins K, Arnold N, Vance DE, Ndinguri MW. Pt-Mal-LHRH, a Newly Synthesized Compound Attenuating Breast Cancer Tumor Growth and Metastasis by Targeting Overexpression of the LHRH Receptor. Bioconjug Chem 2017; 28: 461-470 [PMID: 27997127 DOI: 10.1021/acs.bioconjchem.6b00610]
