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Conjugative Plasmids Disseminating CTX-M-15 among Human, Animals and the Environment in Mwanza Tanzania: A Need to Intensify One Health Approach

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Abstract: Background: Globally, bla_{CTX-M-15} beta-lactamases are the most popular extended spectrum beta-lactamase alleles that are widely distributed due to its mobilisation by mobile genetic elements in several compartments. We aimed to determine the conjugation frequencies and replicon types associated with plasmids carrying bla_{CTX-M-15} gene from Extended Spectrum Beta-lactamase producing isolates in order to understand the dissemination of resistance genes in different compartments.

Material and methods: A total of 51 archived isolates carrying bla_{CTX-M-15} beta-lactamases were used as donors in this study. Antibiotic susceptibility tests were performed as previously described for both donors and transconjugants. Conjugation experiment was performed by a modified protocol of the plate mating experiment, and plasmid replicon types were screened among donor and transconjugant isolates by multiplex Polymerase Chain Reaction in a set of three primer panels.

Results: The conjugation efficiency of plasmids carrying bla_{CTX-M-15} was 88.2% (45/51) with conjugation frequencies in the order of 10^{-1} to 10^{-9} and a 100% transfer efficiency observed among E. coli of animal origin. Majority of donors (n = 21) and transconjugants (n = 14) plasmids were typed as either Inc FIA or Inc FIB. Resistance to non-beta-lactam antibiotics was transferrable in 34/45 (75.6%) of events. Ciprofloxacin, tetracycline and sulphamethoxazole-trimethoprim resistance was co-transferred in 29/34 (85.3%) such events. Gentamicin resistance was transferred in 17/34 (50%) of events. Conclusions: Majority of plasmids carrying bla_{CTX-M-15} were conjugatively transferred by IncF plasmids along with non-beta lactam resistance. There is a need for more research on plasmids to understand how plasmids especially multi replicon plasmids interact and the effect of such interaction on conjugation. One Health approach is to be intensified to address antimicrobial resistance which is a public health threat.

Keywords: conjugation; CTX-M-15; replicon; plasmid; non-beta lactam antibiotics; One Health

1. Introduction

The increasing trend of antimicrobial resistance is intensified by mobile genetic elements that harbour resistance genes [1]. The effect of these elements is extensively reported among bacteria of the Enterobacteriaceae family where multi drug resistance (MDR) is high. The CTX-M extended-spectrum beta-lactamases (ESBL) are the most successful MDR determinants [2], with over 100 alleles in five distinct phylogenetic groups [3]. The ecological success of CTX-M-ESBL attributes to the enzymes’ spread both clonally and horizontally [3–5] in multiple hosts that include Acinetobacter spp., Enterobacter spp., E. coli, P. aeruginosa, K. pneumoniae and P. mirabilis.

In natural environments, ESBL enzymes are chromosomally mediated by the selection pressure induced by beta-lactamase-producing soil organisms [6,7] or the irrational use
of third-generation cephalosporins [6,8], however as previously reviewed [9], plasmid-mediated ESBL resulted from transposon-mediated insertion of different \( \text{bla}_{\text{CTX-M}} \) genes in bacteria chromosome. Specifically, the precursors of plasmid-mediated \( \text{bla}_{\text{CTX-M-15}} \) are environmental \( \text{Kluyvera} \) spp. whose chromosomal CTX-M clusters are incorporated into the chromosome of host bacteria by mobilising elements such as \( \text{ISEcp1} \) or \( \text{ISCR1} \). Moreover, the location of \( \text{ISEcp1} \) upstream \( \text{bla}_{\text{CTX-M}} \) genes together with multiple inverted repeats downstream the gene facilitates the expression and ongoing transposition of \( \text{bla}_{\text{KLU}} \) genes that result to various CTX-M enzymes, including plasmid-mediated \( \text{bla}_{\text{CTX-M-15}} \) [9,10]. The mobilisation potential of \( \text{ISEcp1} \) for chromosome-linked multi-resistant determinants in other members of \( \text{ enterobacteriaceae} \) increases with the additional possession of \( \text{ISCR1} \), another mobile genetic element (MGE) embedded in a Class 1 integron that mobilises unrelated CTX-M groups in similar or different species.

As vectors and carriers of AMR genes, plasmids are responsible for the intracellular accumulation and intercellular transfer of these genes by the process of conjugation [11]. In such cases, high conjugation rates ensure the stable long-term persistence of plasmids and associated AMR genes in minimal fitness costs even in the absence of selection pressure [12–14]. A multidrug resistance phenomenon is observed when these plasmids are associated with other MGE possessing different resistance determinants and code for adaptive traits such as virulence or metal resistance genes among host bacteria strains [15,16].

The globally disseminated O25: H4-ST131 \( \text{E. coli} \) clone producing CTX-M-15 is by conjugative \( \text{IncF} \) plasmids that are frequently recovered from hospital and community settings [17]. In Tanzania, the prevalence of bacteria producing ESBL ranges between 25 and 50 percent [18], with \( \text{bla}_{\text{CTX-M-15}} \) as the predominant allele in both community [19] and hospital settings [20]. The gene is also observed among companion and domestic animals and the environment combined with quinolone and aminoglycoside resistance genes [21,22]. Therefore, as demonstrated by its discovery in a novel \( \text{Enterobacter} \) spp. [23] and location in multiple plasmid types such as \( \text{IncF} \), \( \text{IncY} \) and \( \text{IncHI1} \), there is a possibility of an extensive variation in the epidemiology of \( \text{bla}_{\text{CTX-M-15}} \) carrying plasmids in Tanzania.

The presence of \( \text{bla}_{\text{CTX-M-15}} \) gene in multiple \( \text{E.coli} \) clones of human, animal and the environment of Tanzania [24], and limited information on the persistence of the gene’s alleles in any compartments can lead to the acquisition, transmission and evolution of new resistant strains even among non-conjugative bacteria. Since plasmids facilitate the spread of AMR genes in different compartments efforts to understand their spread and establishment in these settings is unquestionable. This study has improved our understanding of the importance of \( \text{IncF} \) plasmids in disseminating multidrug-resistant determinants among human, animal and environmental settings. It has further highlighted the importance of collaborative One Health based efforts that focus on animal and human health as critical when addressing the global threat.

2. Results
2.1. Isolates Characteristics

\( \text{Escherichia coli} \) was the only species isolated in both human and animals. The environment included isolates from soil and fresh water fish and comprised of \( \text{E. coli} \), \( \text{K. pneumoniae} \), \( \text{C. braakii} \) and \( \text{E. cloacae} \) species (Table 1).

2.2. Conjugation Efficiency of \( \text{bla}_{\text{CTX-M-15}} \) Gene among Isolates of Human, Animals and the Environment

Among 51 \( \text{bla}_{\text{CTX-M-15}} \) positive donor isolates, 45 (88.2%) transferred plasmids by conjugation with a transfer rate (transconjugants per donor cells) ranging from 4.8 \( \times \) 10\(^{-1}\) to 1.5 \( \times \) 10\(^{-9}\) as observed from a human and environment isolate, respectively (Table 2).
### Table 1. Bacteria species distributed among donor isolates of human, animal and environment.

| Sample Origin | Sample Type | Frequency n% | Species | Species n (%) | Total n (%) |
|---------------|-------------|--------------|---------|---------------|-------------|
| Human         | Human       | 22 (43.14)   | *E. coli* | 22 (43.1)     | 22 (43.14)  |
| Goat          |             | 1 (1.96)     | *E. coli* | 1 (1.96)      |             |
| Animal        | Pig         | 3 (5.88)     | *E. coli* | 3 (5.88)      |             |
| Dog           |             | 6 (11.76)    | *E. coli* | 6 (11.76)     | 12 (23.52)  |
| Chicken       |             | 2 (3.92)     | *E. coli* | 2 (3.92)      |             |
| Environment   | Soil        | 6 (11.76)    | *E. coli* | 6 (11.76)     |             |
| Fish          |             | 11 (21.57)   | *K. pneumonia* | 3 (5.88) | 17 (33.32) |
|               |             |              | *C. braakii* | 2 (3.92) |             |
|               |             |              | *E. cloacae* | 4 (7.84) |             |
|               |             |              |          |               |             |
| Total (n)     |             | 51 (100)     |          |               | 51 (100)    |

### Table 2. Conjugation efficiency of human, animal and environment donor isolates.

| Sample ID | Source        | Species | Conjugation Frequency |
|-----------|---------------|---------|-----------------------|
| CN1       | Fish          | *E. cloacae* | $8.2 \times 10^{-5}$ |
| CN2       | Fish          | *E. cloacae* | $2.3 \times 10^{-4}$ |
| CN3       | Fish          | *E. cloacae* | $5.2 \times 10^{-5}$ |
| CN4       | Fish          | NIL      |                       |
| CN5       | Fish          | *C. braakii* | $7.5 \times 10^{-6}$ |
| CN6       | Fish          | *E. coli* | $7.6 \times 10^{-3}$ |
| CN7       | Fish          | *E. coli* | NIL                   |
| CN8       | Fish          | *K. pneumonia* | $2.0 \times 10^{-5}$ |
| CN9       | Fish          | *K. pneumonia* | $4.2 \times 10^{-4}$ |
| CN10      | Fish          | *K. pneumonia* | $3.3 \times 10^{-5}$ |
| CN11      | Fish          | *C. braakii* | $9.4 \times 10^{-4}$ |
| CN12      | Pig           | *E. coli* | $4.7 \times 10^{-5}$ |
| CN13      | Pig           | *E. coli* | $2.6 \times 10^{-6}$ |
| CN14      | Pig           | *E. coli* | $9.8 \times 10^{-5}$ |
| CN15      | Local chicken | *E. coli* | $4.7 \times 10^{-5}$ |
| CN16      | Local chicken | *E. coli* | $8.4 \times 10^{-7}$ |
| CN17      | Goat          | *E. coli* | $4.1 \times 10^{-6}$ |
| CN18      | Dog           | *E. coli* | $2.1 \times 10^{-5}$ |
| CN19      | Dog           | *E. coli* | $1.2 \times 10^{-7}$ |
| CN20      | Dog           | *E. coli* | $5.0 \times 10^{-5}$ |
| CN21      | Dog           | *E. coli* | $1.1 \times 10^{-6}$ |
| CN22      | Dog           | *E. coli* | $6.0 \times 10^{-4}$ |
| CN23      | Dog           | *E. coli* | $9.6 \times 10^{-6}$ |
| CN24      | Environment   | *E. coli* | $1.5 \times 10^{-9}$ |
| CN25      | Environment   | *E. coli* | $2.6 \times 10^{-7}$ |
| CN26      | Environment   | *E. coli* | $3.5 \times 10^{-6}$ |
| CN27      | Environment   | *E. coli* | $2.9 \times 10^{-7}$ |
| CN28      | Environment   | *E. coli* | $6.1 \times 10^{-6}$ |
| CN29      | Environment   | *E. coli* | $7.2 \times 10^{-3}$ |
| CN30      | Human         | *E. coli* | $1.0 \times 10^{-3}$ |
| CN31      | Human         | *E. coli* | $4.7 \times 10^{-4}$ |
| CN32      | Human         | *E. coli* | $2.1 \times 10^{-4}$ |
| CN33      | Human         | *E. coli* | $4.0 \times 10^{-5}$ |
| CN34      | Human         | *E. coli* | $5.4 \times 10^{-5}$ |
| CN35      | Human         | *E. coli* | $4.8 \times 10^{-1}$ |
| CN36      | Human         | *E. coli* | $1.7 \times 10^{-4}$ |
| CN37      | Human         | *E. coli* | $3.5 \times 10^{-7}$ |
| CN38      | Human         | *E. coli* | $8.1 \times 10^{-5}$ |
| CN39      | Human         | *E. coli* | $1.2 \times 10^{-5}$ |
Table 2. Cont.

| Sample ID | Source | Species | Conjugation Frequency |
|-----------|--------|---------|-----------------------|
| CN40      | Human  | E. coli | 2.7 × 10^{-5}         |
| CN41      | Human  | E. coli | 2.4 × 10^{-7}         |
| CN42      | Human  | E. coli | NIL                   |
| CN43      | Human  | E. coli | 5.5 × 10^{-6}         |
| CN44      | Human  | E. coli | 4.4 × 10^{-6}         |
| CN45      | Human  | E. coli | 2.9 × 10^{-6}         |
| CN46      | Human  | E. coli | NIL                   |
| CN47      | Human  | E. coli | 2.1 × 10^{-5}         |
| CN48      | Human  | E. coli | 1.2 × 10^{-4}         |
| CN49      | Human  | E. coli | 1.1 × 10^{-7}         |
| CN50      | Human  | E. coli | NIL                   |
| CN51      | Human  | E. coli | NIL                   |

NIL: no conjugation.

2.3. Transferrable Resistance of Non-Beta-Lactam Phenotype among Isolates of Human, Animal and the Environment

Table 3 and Figure 1 presents a summary of non-beta lactam resistant phenotypes transferred along the \(\text{bla}_{\text{CTX-M-15}}\) gene. A total of 45 plasmids successful transferred the gene to transconjugants. Non-beta-lactam resistance phenotypes were observed in 34/45 (75.6%) transconjugants. Donor resistance to ciprofloxacin (CIP), tetracycline (TE) and trimethoprim-sulphamethoxazole (SXT) was observed in 46/51 (90.2%), 47/51 (92.2%) and 48/51 (94.1%) of events, respectively, and was co-transferred in 29/34 (85.3%) of such events. Gentamicin was the least transferred with a frequency of 17/34 (50.0%).

Table 3. Antibiotic resistance phenotypes of donors and transconjugants of human animals and the environment.

| Sample No. | Source | Species          | Donor’s Non-B-lactam Resistance Phenotype |
|------------|--------|------------------|------------------------------------------|
| CN1        | Fish   | E. cloacae       | SXT *, CIP *, CN *, TE *                 |
| CN2        | Fish   | E. cloacae       | CIP, SXT, CN, TE                         |
| CN3        | Fish   | E. cloacae       | CIP *, SXT *, TE *, CN *                 |
| CN4        | Fish   | E. cloacae       | CIP, CN, TE, SXT                         |
| CN5        | Fish   | C. braakii       | CIP *, SXT *, CN *, TE *                 |
| CN6        | Fish   | E. coli          | CIP, SXT, CN, TE                         |
| CN7        | Fish   | E. coli          | CIP, TE                                  |
| CN8        | Fish   | K. pneumoniae    | CIP *, SXT *, CN *, TE *                 |
| CN9        | Fish   | K. pneumoniae    | CIP *, SXT *, CN *, TE *                 |
| CN10       | Fish   | K. pneumoniae    | CIP, SXT, CN, TE                         |
| CN11       | Fish   | C. braakii       | CIP, SXT, CN, TE                         |
| CN12       | Pig    | E. coli          | CIP *, SXT *, TE *                       |
| CN13       | Pig    | E. coli          | TE, CIP, CN                              |
| CN14       | Pig    | E. coli          | CIP *, SXT *, TE *, CN *                 |
| CN15       | Local chicken | E. coli   | CIP, SXT, CN, TE                       |
| CN16       | Local chicken | E. coli   | CIP, SXT, CN, TE                       |
| CN17       | Goat   | E. coli          | SXT, TE *, CN, CIP                       |
| CN18       | Dog    | E. coli          | SXT                                       |
| CN19       | Dog    | E. coli          | SXT *, CIP *, TE, CN                     |
| CN20       | Dog    | E. coli          | CIP *, SXT *, TE *                       |
| CN21       | Dog    | E. coli          | CIP *, SXT *, TE *, CN *                 |
| CN22       | Dog    | E. coli          | CIP *, CN *, TE *, SXT *                 |
| CN23       | Dog    | E. coli          | SXT, TE, CN, CIP                         |
| CN24       | Environment | E. coli   | SXT *, CIP *, TE *                      |
| CN25       | Environment | E. coli   | SXT, TE, CIP                           |
| CN26       | Environment | E. coli   | CIP *, SXT *, TE *                      |
Table 3. Cont.

| Sample No. | Source     | Species | Donor's Non-B-lactam Resistance Phenotype |
|------------|------------|---------|----------------------------------------|
| CN27       | Environment| E. coli | CIP *                                  |
| CN28       | Environment| E. coli | CIP *, SXT *, CN *, TE *               |
| CN29       | Environment| E. coli | CN, CIP *, SXT *, TE *                 |
| CN30       | Human      | E. coli | TE *, CIP *, CN, SXT *                |
| CN31       | Human      | E. coli | CIP *, SXT *                          |
| CN32       | Human      | E. coli | SXT *, CIP *                          |
| CN33       | Human      | E. coli | TE *, CN *, CIP *, SXT *              |
| CN34       | Human      | E. coli | SXT *, TE *, CN *, CIP                |
| CN35       | Human      | E. coli | CIP *, CN *, SXT *, TE *              |
| CN36       | Human      | E. coli | CIP *, CN *, SXT *, TE *              |
| CN37       | Human      | E. coli | CIP *, CN *, SXT *, TE *              |
| CN38       | Human      | E. coli | SXT *, TE *, CIP *, CN *              |
| CN39       | Human      | E. coli | SXT, TE, CIP *, CN *                 |
| CN40       | Human      | E. coli | SXT *, TE *                          |
| CN41       | Human      | E. coli | SXT, TE *, CIP *, CN                 |
| CN42       | Human      | E. coli | SXT, CIP, CN, TE                     |
| CN43       | Human      | E. coli | CN *, CIP *, SXT *, TE *             |
| CN44       | Human      | E. coli | SXT, TE, CIP                         |
| CN45       | Human      | E. coli | SXT, TE, CIP, CN                     |
| CN46       | Human      | E. coli | TE, SXT                               |
| CN47       | Human      | E. coli | SXT *, TE *, CIP, CN                 |
| CN48       | Human      | E. coli | SXT *, TE *, CIP, CN                 |
| CN49       | Human      | E. coli | CIP *, CN *, SXT *, TE *             |
| CN50       | Human      | E. coli | SXT, TE                              |
| CN51       | Human      | E. coli | CN, CIP, SXT, TE                     |

* Transferable resistance; SXT: Trimethoprim-sulphamethoxazole, CIP: Ciprofloxacin, TE: tetracycline, CN: Gentamicin.

Figure 1. Transfer success of \( \text{bla}_{\text{CTX-M-15}} \) among \( E. \text{coli} \) isolates of human, animals and the environment.
2.4. Replicon Types of Plasmids Carrying bla<sub>CTX-M-15</sub>

Common replicon types were FIA (n = 11) and FIB (n = 27) that occurred as single transferrable replicons in 14 events. Inc A/C and Y replicons were minor, and each was typed once. Of all the 14 typed transconjugant plasmids, 7 replicons were observed in both donors and transconjugants, while 20/27 donor replicons were not observed in respective transconjugants (Table 4), (Figure 2).

Table 4. Replicon types of plasmids carrying bla<sub>CTX-M-15</sub> among donors and transconjugants.

| Sample Source | Conjugation Efficiency | Conjugation Range | Donor’s Plasmid Replicon | Transconjugant Replicon Type |
|---------------|------------------------|-------------------|--------------------------|----------------------------|
| Human         | 1.2 x 10^-4            | FIB               | FIA                      | Nil                        |
| Human         | 8.1 x 10^-5            | FIA, FIB          | FIB                      | Nil                        |
| Dog           | 5.0 x 10^-5            | FIB               | FIB                      | Nil                        |
| Human         | 5.4 x 10^-5            | 10^-6-10^-3       | FIB                      | FIB                        |
| Human         | 2.1 x 10^-4            | FIB               | FIB                      | Nil                        |
| Environment   | 7.2 x 10^-3            | FIB               | FIB                      | Nil                        |
| Dog           | 1.1 x 10^-6            | FIB               | FIB                      | Nil                        |
| Human         | 1.7 x 10^-4            | FIB               | FIB                      | Nil                        |
| Dog           | 9.6 x 10^-6            | no rep            | FIB                      | Nil                        |
| Human         | 1.2 x 10^-4            | 10^-7-10^-4       | no rep                   | Nil                        |
| Human         | 4.7 x 10^-4            | no rep            | FIB                      | Nil                        |
| Environment   | 2.9 x 10^-7            | no rep            | FIB                      | Nil                        |
| Fish          | 2.3 x 10^-4            | no rep            | FIB                      | NA                         |
| Human         | NIL                    | no rep            | NA                       | NA                         |
| Human         | NIL                    | 0                 | no rep                   | NA                         |
| Human         | NIL                    | no rep            | no rep                   | NA                         |
| Human         | NIL                    | no rep            | no rep                   | NA                         |
| Fish          | NIL                    | no rep            | NA                       | NA                         |
| Fish          | 4.2 x 10^-4            | A/C, FIA          | no rep                   | NA                         |
| Pig           | 2.6 x 10^-6            | FIA               | no rep                   | NA                         |
| Human         | 5.5 x 10^-6            | FIA               | no rep                   | NA                         |
| Pig           | 6.0 x 10^-4            | FIA               | no rep                   | NA                         |
| Human         | 9.8 x 10^-5            | FIA               | no rep                   | NA                         |
| Human         | 2.9 x 10^-6            | FIA               | no rep                   | NA                         |
| Human         | 4.0 x 10^-5            | FIA               | no rep                   | NA                         |
| Human         | 4.8 x 10^-1            | 10^-9-10^-1       | FIA                      | no rep                     |
| Dog           | 1.2 x 10^-7            | FIB               | no rep                   | NA                         |
| Human         | 3.5 x 10^-7            | FIB               | no rep                   | NA                         |
| Environment   | 1.5 x 10^-9            | FIB               | no rep                   | NA                         |
| Environment   | 2.6 x 10^-7            | FIB               | no rep                   | NA                         |
| Human         | 4.4 x 10^-6            | FIB               | no rep                   | NA                         |
| Environment   | 3.5 x 10^-6            | FIB               | no rep                   | NA                         |
| Human         | 2.1 x 10^-5            | FIB               | no rep                   | NA                         |
| Fish          | 7.5 x 10^-6            | no rep            | no rep                   | NA                         |
| Fish          | 9.4 x 10^-4            | no rep            | no rep                   | NA                         |
| Human         | 2.7 x 10^-5            | no rep            | no rep                   | NA                         |
| Local chicken | 4.7 x 10^-3            | no rep            | no rep                   | NA                         |
| Pig           | 4.7 x 10^-5            | no rep            | no rep                   | NA                         |
| Human         | 2.4 x 10^-7            | no rep            | no rep                   | NA                         |
| Fish          | 3.3 x 10^-5            | no rep            | no rep                   | NA                         |
| Fish          | 2.0 x 10^-5            | 10^-7-10^-3       | no rep                   | no rep                     |
| Fish          | 7.6 x 10^-3            | no rep            | no rep                   | no rep                     |
| Human         | 1.1 x 10^-7            | no rep            | no rep                   | no rep                     |
| Fish          | 5.2 x 10^-5            | no rep            | no rep                   | no rep                     |
| Goat          | 4.1 x 10^-6            | no rep            | no rep                   | no rep                     |
| Environment   | 6.1 x 10^-6            | no rep            | no rep                   | no rep                     |
| Local chicken | 8.4 x 10^-7            | no rep            | no rep                   | no rep                     |
| Human         | 1.0 x 10^-3            | no rep            | no rep                   | no rep                     |
| Fish          | 8.2 x 10^-5            | no rep            | no rep                   | no rep                     |

NIL: no conjugation, NA: no transconjugants, no rep: no typable replicon.
2.5. Transfer Success of \( \text{bla}_{\text{CTX-M-15}} \) among \( \text{E. coli} \) Isolates

Table 5 shows the percentage transfer of \( \text{bla}_{\text{CTX-M-15}} \) among \( \text{E. coli} \) donor isolates. Out of 42 \( \text{E. coli} \) donors, 37 (88.1%) successfully transferred the gene, accounting for an 82.1% of all transconjugants. All \( \text{E. coli} \) originating from animals transferred the gene successfully.

| Source      | \( \text{E. coli} \) Donors n | \( \text{E. coli} \) Transconjugants n (%) |
|-------------|-------------------------------|------------------------------------------|
| Human       | 22                            | 18 (81.8)                                |
| Animal      | 12                            | 12 (100.0)                               |
| Environment | 8                             | 7 (87.5)                                 |
| Total       | 42                            | 37 (88.1)                                |

3. Discussion

In this study, we aimed to determine conjugation frequencies and type plasmids carrying the \( \text{bla}_{\text{CTX-M-15}} \) gene from human, animal and environment ESBL producing isolates. The study is epidemiologically important in understanding the pattern and possibly predict the flow of AMR from different sources. As presented in (Table 1), \textit{Escherichia coli} was the dominant bacteria species from all sourced samples. The successful colonisation of \( \text{E. coli} \) in human and animal gastrointestinal tract (GIT) have been reported previously [25], the GIT can serve as exchange hotspots and reservoirs of antimicrobial resistance genes. Likewise, \textit{Escherichia coli} and \textit{Klebsiella pneumoniae} are frequently isolated in infections associated...
with CTX-M-15 in hospitals [26] and the community, including households [27], aquatic environment [28] and the soil [29].

The transfer efficiency of \( \text{bla}_{\text{CTX-M-15}} \) among isolates in this study was higher (88%) [Table 2], than that reported by Zurfluh and colleagues [30], where a 38.3% efficiency was observed, however, despite the varying frequencies of transfer, the reported efficiency is slightly lower than that previously reported (100%) for randomly selected hospital originating isolates [20]. It is reported that high conjugation rates above thresholds compensate fitness costs and establish a long-term persistence of plasmids in multiple hosts [13,31,32] through maintaining successive generations of bacteria with adaptive traits. Therefore, the high transfer efficiency is a fitness advantage that improves the persistence and dissemination potential of \( \text{bla}_{\text{CTX-M-15}} \) ESBL in human, animal and the environment interface. Moreover, transfer failure for some isolates—CN4, CN7, CN42, CN46, CN50 and CN51, could possibly be due to the gene’s integration in the chromosome [33] or transposition events that prevent plasmid mobility.

As summarised in Table 3 and Figure 1, transferable multidrug resistance phenotypes were also observed. The conjugative spread of \( \text{bla}_{\text{CTX-M-15}} \) gene by IncF plasmids along with tetracycline, aminoglycoside and quinolones have been reported [34]. These plasmids harbor several combinations of resistance determinants and transfer them to human, animals and environment isolates through the ecological interaction of bacteria in these settings. Moreover, the genetic environment of \( \text{bla}_{\text{CTX-M-15}} \) is dominated by multiple antibiotic resistance genes such as \( \text{aac}(6\prime)-\text{Ib-cr}, \text{tet}(A, B), \text{qnrS}, \text{qnr} \) and \( \text{sul} \) genes [35–37], whose phenotypic expression denotes the existing selection pressure for these antibiotics. Such selection can increase their transfer rate and possibly account for the high co-transfer of non-beta lactam antibiotics observed in this study.

We also observed single replicon IncF plasmids as common vectors of \( \text{bla}_{\text{CTXM-15}} \), (Table 4), (Figure 2). Replicon typing of plasmids carrying antimicrobial resistance genes is important for detecting, tracing or monitoring the spread of antimicrobial resistance. These observations are in line with findings in the same setting [20] and elsewhere [30,38] where multireplicon FIA and FIB plasmids were reported to carry multiple resistance genes including \( \text{bla}_{\text{CTX-M-15}} \). As in previous studies, IncY plasmids and Inc A/C carrying \( \text{bla}_{\text{CTX-M-15}} \) in association with quinolone and aminoglycoside genes were also reported [22,39,40]. Together these findings support the diverse nature of plasmids adapted to spread and maintain \( \text{bla}_{\text{CTX-M-15}} \) gene.

The absence of donor replicons in respective transconjugants might have resulted from conjugation failure, multi-replicons (undetected by the method used) that destabilise and prevent the transfer of some resident replicons [41], and prior plasmid dependent mutations (which do not occur in transconjugants) that may have altered backbone genes of donor plasmids and obscure the detection of existing replicons [32,39,41]. In addition, and as a shortcoming, the PBRT technique used in detecting plasmid replicons can give false-negative results when replicon sequences go undetected by the primer sets used, target replicon sequences undergo mutation through transpositional alterations and the unknown existence of new replicons in such plasmid [42].

Lastly, we observed that all animal originating \( \text{E. coli} \) isolates transferred \( \text{bla}_{\text{CTX-M-15}} \) to respective transconjugants (Table 5), (Figure 3). These findings were also supported by a recent review [34], that human and animal originating \( \text{E.coli} \) are adapted to disseminate ESBL genes by IncF plasmids. The colonisation and infection of animals by \( \text{E. coli} \) maximises microbial interactions between non-pathogenic and pathogenic commensal \( \text{E.coli} \) in either companion or food-producing animals and facilitate the exchange of materials between them through conjugation. In addition, the increasing use of antibiotics in animals could select and transfer resistant pathogenic bacteria from animals to human and the environment with huge cost implications. Since AMR is a public health threat, the highest transfer rate observed in animal originating \( \text{E. coli} \) calls for integrated efforts to address AMR with experts from veterinary, human and ecological fields. It further implies that
animals may serve as dual targets for studies focusing on the horizontal transfer and evolution of antimicrobial resistance.

Figure 3. Transfer success of bla\textsubscript{CTX-M-15} among E. coli isolates by source.

4. Materials and Methods

4.1. Study Isolates

All isolates used as donors in this study were obtained from the Catholic University of Health and Allied Sciences (CUHAS) in Mwanza Tanzania. A total of 51 bla\textsubscript{CTX-M-15} positive isolates were purposively selected and activated overnight in Luria Bertani (LB) broth at 37 °C ready for use in conjugation and PBRT techniques.

Among the 51 isolates, twenty-two bla\textsubscript{CTX-M-15} positive isolates were obtained from a study that reported the magnitude of fecal carriage and diversity of ESBL genotypes among human residing in rural communities of Mwanza Tanzania [19], 12 other bla\textsubscript{CTX-M-15} positive isolates were from a study that reported the fecal carriage of ESBL among companion and domestic farm animals that included pigs, chicken, dogs and goats [21]. The remaining 17 environmental isolates were obtained from a study that investigated the presence of bla\textsubscript{CTX-M-15} from muddy soils and gut contents of freshwater fish from Lake Victoria in Mwanza Tanzania [22].

4.2. Antibiotic Susceptibility Testing

Susceptibility testing of all donor isolates and the resulting transconjugants was performed by the disk diffusion method on Mueller Hinton agar as recommended by the Clinical and Laboratory Standard Institute [43]. Antibiotics tested were tetracycline (30 µg), gentamicin (30 µg), ciprofloxacin (5 µg) and trimethoprim/sulphamethoxazole (1.25/23.75 µg) (Hi-media, India).
4.3. Conjugation Experiment

A total of 51 known bla\textsubscript{CTX-M-15} positive isolates and \textit{Escherichia coli} J53 ((F\textsuperscript{−}, met, pro, Az\textsuperscript{−})-a mutant strain of \textit{E. coli} [44] obtained from the Institute of Medical Microbiology, Giessen, Germany, were used as donors and recipient strain, respectively. As previously described [20], we performed conjugation experiments with some modifications. Shortly, the recipient strain was prepared by streaking \textit{Escherichia coli} J53 in Luria Bertani (LB) plates supplemented with 100 \(\mu\)g/mL NaN\textsubscript{3} (LB++), while donor strains were selected in LB plates supplemented with 2 \(\mu\)g/mL cefotaxime only (LB+). From these, fresh overnight donor and recipient strains were prepared by picking single colonies emulsified in 10 mL LB broth and incubated overnight at 37 \(^\circ\)C in a 150 rpm shaking incubator. After exactly 12 h, equal volumes (500 \(\mu\)L) of donor and recipient strains were immediately mixed in 1.5 mL eppendorf tubes previously labeled transconjugant (Tc) while 1000 \(\mu\)L of donor strain were added in fresh tubes of similar volume-to be separately selected on LB+ and LB++ plates as respective controls. All tubes were incubated at 37 \(^\circ\)C for 15 min, vortexed briefly, centrifuged at 12,000 \(g\) for 2 min and the pellet re-suspended in fresh 1000 \(\mu\)L LB broth. Finally, 0.1 mL of \(10^{-1}\) to \(10^{-4}\) transconjugant cultures were double selected on LB plates supplemented with 100 \(\mu\)g/mL NaN\textsubscript{3} and 2 \(\mu\)g/mL cefotaxime. Conjugation efficiency was reported as transconjugants per donor cells, with the denominator obtained from an initial volume of 100 \(\mu\)L.

4.4. Genomic Extraction of Donor and Transconjugants DNA

Donor and transconjugant genomic DNA was extracted using a previously described chelex protocol with slight modifications [45]. First, 5 \(\mu\)L of proteinase K (10 mg/mL) were added into tubes containing 100 \(\mu\)L fresh LB emulsified colonies. In the same tubes, 300 \(\mu\)L of chelex buffer (Qiagen GmbH, Hilden, Germany) was added consecutively. The mixture was incubated for 3 hr at 55 \(^\circ\)C before adding 85 \(\mu\)L of 5 M NaCl and vortexed for 15 s to precipitate proteins. The supernatant was centrifuged at 13,000 \(\times\) g for 10 min followed by the addition of 300 \(\mu\)L of 100% cold ethanol and a 5 min centrifugation at 13,000 \(\times\) g that precipitated and pelleted the DNA. Lastly, the pellet was rinsed by pouring off the remaining fluid, adding 500 \(\mu\)L of 70% ethanol, centrifuging at 13,000 \(\times\) g for 5 min and leaving the pellet to air dry at 55 \(^\circ\)C for 10 min. The DNA was then re-suspended in 50 \(\mu\)L nuclease-free water. Nanodrop (Thermo Scientific, Wilmington, DE) was used to check the quantity of the DNA, while the quality was confirmed by electrophoresis in 1.5% (w/v) agarose gel using TAE buffer. The obtained DNA samples were used in typing plasmid replicons or stored at \(-20\) \(^\circ\)C.

4.5. PCR Based Replicon Typing

Targeted genes were amplified by a simplified version of the previously described PBRT technique [42]. Shortly, the eight Polymerase Chain Reaction (PCR) panels illustrated by Caratoli and colleagues [46], were reduced to three [42], (Table 6). PCR was performed using a readily reconstituted master mix according to manufacturer’s instructions (New England BioLabs, Inc. Beverly, MA) under the following conditions; 5 min at 94 \(^\circ\)C; 30 cycles of 30 s at 94 \(^\circ\)C, 30 s at 60 \(^\circ\)C and 90 s at 72 \(^\circ\)C; then a final extension of 5 min at 72 \(^\circ\)C. Amplicons were visualised on 1.5% tris-acetate EDTA agarose gels alongside a 100 bp DNA ladder (New England BioLabs, Inc. Beverly, MA). The sample was considered positive for replicon gene (s) if an amplicon of the expected band size was observed.
Table 6. Primers used in PCR based replicon typing of donor and transconjugant plasmids.

| Primer Panel/Target | Direction | Primer Sequence | Annealing Temp (°C) | Amplicon Size (bp) |
|---------------------|-----------|-----------------|---------------------|-------------------|
| **Panel 1**         |           |                 |                     |                   |
| B/O                 | F         | 5′-gcgggtccggaagccagaaac-3′ | 60                   | 159               |
|                     | R         | 5′-tgctggcttccggaagcttcga-3′ | 60                   | 262               |
| FIC                 | F         | 5′-gtgaacctggccagtgaggaaggg-3′ | 60                   | 534               |
|                     | R         | 5′-tctgcgttccgcagaaactag-3′ | 60                   |                   |
| A/C                 | F         | 5′-gacaacaaacacaaagaccttgga-3′ | 60                   | 465               |
|                     | R         | 5′-tacaagcagcgagcagcagc-3′ | 60                   |                   |
| P                   | F         | 5′-ctagggcctggacacgag-3′ | 60                   |                   |
|                     | R         | 5′-tgctgtttgtgcttcataa-3′ | 60                   |                   |
| T                   | F         | 5′-aatcgaacacagcgagcttg-3′ | 60                   | 750               |
|                     | R         | 5′-cgcctagtccaggttggac-3′ | 60                   |                   |
| **Panel 2**         |           |                 |                     |                   |
| K/B                 | F         | 5′-gcgggtccggaagccagaaac-3′ | 60                   | 160               |
|                     | R         | 5′-tcttcacagccgcagcagaa-3′ | 60                   |                   |
| W                   | F         | 5′-ctaaggaacacaaacacccgcag-3′ | 60                   | 242               |
|                     | R         | 5′-gagttcgccagcagaaactag-3′ | 60                   |                   |
| FIIr                | F         | 5′-ctgctgaactcagctgcgag-3′ | 60                   | 270               |
|                     | R         | 5′-tacaagcagcgagcagcagc-3′ | 60                   |                   |
| FIA                 | F         | 5′-ctagggcctggacacgagcag-3′ | 60                   | 462               |
|                     | R         | 5′-atcaacctccatcctcgccagag-3′ | 60                   | 702               |
| FIB                 | F         | 5′-gcgggtccggaagccagaaac-3′ | 60                   |                   |
|                     | R         | 5′-tctgcgttccgcagaaactag-3′ | 60                   |                   |
| Y                   | F         | 5′-aatcgaacacagcgagcttg-3′ | 60                   | 765               |
|                     | R         | 5′-gcgggtccggaagccagaaac-3′ | 60                   |                   |
| **Panel 3**         |           |                 |                     |                   |
| I1                  | F         | 5′-gcgggtccggaagccagaaac-3′ | 60                   | 139               |
|                     | R         | 5′-tgctggcttccggaagcttcga-3′ | 60                   | 270               |
| FrepB               | F         | 5′-tgctggcttccggaagcttcga-3′ | 60                   | 376               |
|                     | R         | 5′-gcgggtccggaagccagaaac-3′ | 60                   | 559               |
| X                   | F         | 5′-gcgggtccggaagccagaaac-3′ | 60                   | 644               |
|                     | R         | 5′-tgctggcttccggaagcttcga-3′ | 60                   | 785               |

5. Conclusions

Majority of plasmids carrying blaCTX-M-15 were conjugatively transferred by IncF plasmids along with non-beta lactam resistance. The heterogeneous nature of these plasmids continuously maintains and reserves the blaCTX-M-15 gene in these settings. The 100% transfer efficiency among E. coli of animal origin is of concern since the networked interaction of animals with human and their environment continuously exchange and reserve resistance determinants in this interface. Therefore, there is a need for more research to understand the interaction and spread of mobile elements circulating in animals, One Health approach is to be intensified to further address AMR as a public health threat.

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**Abbreviations**

| Abbreviation | Definition |
|---------------|------------|
| AMR           | Antimicrobial resistance |
| CREATESS      | Centre for Research, Agricultural advancement, Teaching Excellence and Sustainability in Food and Nutritional Security |
| CUHAS         | Catholic University of Health and Allied Sciences |
| DNA           | Deoxyribonucleic acid |
| EDTA          | Ethylene Diaminetetracetic acid |
| ESBL          | Extended spectrum beta-lactamase |
| GIT           | Gastrointestinal tract |
| LB            | Luria Bertani |
| MGE           | Mobile genetic element |
| NaCl          | Sodium Chloride |
| NaN3          | Sodium azide |
| PCR           | Polymerase chain reaction |
| TAE           | Tris acetate EDTA |

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