First Case Report of Possible Sarcocystis truncata-induced Food Poisoning in Venison

Takayuki Ota¹, Yoshio Nakano¹, Tsuyoshi Mizuno¹, Ayako Shiozaki¹, Yoshikazu Hori¹, Kazuki Yamanishi¹, Kana Hayakawa¹, Takahiro Hayakawa¹, Tokuzo Fujimoto¹, Chiaki Nakamoto², Kei Maejima³, Yasuhiko Wada³, Fumio Terasoma⁴ and Takahiro Ohnishi⁵

Abstract:
Along with the increase in consumption of raw animal meat, the prevalence of food poisoning is increasing. A 67-year-old Japanese man had eaten raw venison 4 hours prior to the beginning of vomiting. Many white cysts were discovered in the venison, with numerous bradyzoites being detected after the cysts were punctured. The presence of the Sarcocystis spp. 18S rRNA gene was detected by polymerase chain reaction, and Sarcocystis truncata was isolated from the venison. Sarcocystis truncata has not previously been identified in sika deer (Cervus nippon) in Japan. This is the first report of possible Sarcocystis truncata-induced food poisoning following consumption of venison.

Key words: Sarcocystis truncata, food poisoning, venison

(Intern Med 58: 2727-2730, 2019) (DOI: 10.2169/internalmedicine.2817-19)

Introduction

The habit of consuming raw animal meat is common in some countries. As this practice increases, the prevalence of food poisoning or paragonimiasis is also increasing. With recent increases in the population of sika deer (Cervus nippon) (1) and the increasing popularity of wild game cuisine, the incidence of food poisoning or paragonimiasis following ingestion of raw venison has been increasing in Japan. A variety of zoonotic pathogens may be found in wild game meat, including venison (2). Sarcocystis is one such pathogen, living as a parasite in the muscles and digestive tracts of domestic animals (3).

Aoki et al. isolated S. sybellensis and S. wapiti from venison that had caused food poisoning and revealed by immunohistochemical staining that both had a protein that reacted with an antibody against a 15-kDa protein from S. fayeri (4). It is widely known that Sarcocystis spp. are a causative pathogen of food poisoning from horse meat. However, Sarcocystis infection of venison has rarely been reported.

This is the first report describing a case of food poisoning possibly caused by S. truncata in venison.

Case Report

A 67-year-old Japanese man, suffering from abdominal pain, vomiting, watery diarrhea, and fever, visited a hospital by ambulance. He had experienced no episodes of cough, sputum, or trauma in the immediate past, and his symptoms had occurred suddenly an hour before he presented to the hospital. The patient had no history of operation. Despite suffering from hepatitis C virus infection, he had never been hospitalized for hepatic diseases. He lived in a rural area surrounded by forests and had a habit of consuming 700 mL of beer a day.

The patient had eaten raw venison 4 hours before the start of vomiting. The deer had been hunted 8 hours prior to consumption and was stored at room temperature. Upon admission, the patient was febrile with a temperature of 37.3°C, blood pressure of 108/83 mmHg, and a pulse rate of 70/min. Vomiting and watery diarrhea continued in the emergency department.

¹Department of Internal Medicine, Kinan Hospital, Japan, ²Nursing, Kinan Hospital, Japan, ³Tanabe Public Health Center, Japan, ⁴Wakayama Prefectural Research Center of Environment and Public Health, Japan and ⁵National Institute of Health Science, Japan

Received: February 12, 2019; Accepted: March 12, 2019; Advance Publication by J-STAGE: May 22, 2019
Correspondence to Dr. Takayuki Ota, FZG05232@nifty.ne.jp
The results of a physical examination were normal, except for a remarkable increase in intestinal peristaltic sounds. The laboratory examination results are shown in Table. The serum levels of creatinine, urea acid, and total protein were found to be elevated, whereas the serum sodium level was decreased.

The stool culture was negative, with no enteric pathogens, ova, or parasites detected. *Shigella* and other enteritis pathogens were also absent. The differential diagnosis of *Salmonella*, *Escherichia coli*, hepatitis E virus, *Shigella*, *Enterobacteriaceae*, *Leptospira*, *Listeria*, and *Campylobacter* was considered. However, with the exception of *Escherichia coli*, infection symptoms from these pathogens would be expected to present 24 hours after infection. Besides this patient, two other people who had eaten the same venison displayed identical symptoms, and food poisoning was diagnosed in both cases. Considering the possibility of *Sarcocystis* infection, we asked the public health center to examine the venison for pathogens.

An examination by stereoscopic microscopy revealed the presence of white cysts in the venison muscle tissue (Fig. 1A). The cysts were 1 to 2 mm long and approximately 0.1 mm wide (Fig. 1B and C). Many bradyzoites were detected after puncturing the cysts (Fig. 2). The presence of the *Sarcocystis* spp. 18S rRNA gene was detected by polymerase chain reaction, and *Sarcocystis truncata* was isolated from the venison. We extracted proteins from the cysts isolated from the venison and determined their amino acid sequences, identifying a toxic 15-kDa protein (Fig. 3). No cysts were detected in the venison liver tissue.

No antibiotics were administered. Sufficient infusion was performed. On day 3, the patient improved clinically and became afebrile. He was therefore discharged.

### Discussion

*Sarcocystis* is a zoonotic pathogen that lives as a parasite in the muscles and digestive tracts of domestic animals (3). Carnivorous animals act as final hosts for these parasites and excrete *Sarcocystis* sporocysts in their feces. In herbivorous

---

**Table. Laboratory Data on Admission.**

| Hematological analysis | Biochemistry       |
|------------------------|--------------------|
| WBC 8.700 μL           | AST 54 U/L         |
| Neutro 78.4 %          | ALT 31 U/L         |
| Lym 17.2 %             | γ-GTP 41 IU/L      |
| Eosino 0.3 %           | TP 8.2 g/dL        |
| Mono 3.9 %             | Alb 4.4 g/dL       |
| Baso 0.2 %             | LDH 276 IU/L       |
| RBC 465 ×10^6/μL       | ALP 305 IU/L       |
| Hb 14.0 g/dL           | CK 161 IU/L        |
| Ht 42.3 %              | CHE 185 IU/L       |
| PLT 18.3 ×10^6/μL      | TB 0.6 mg/dL       |
|                        | CRP 0.05 mg/dL     |
| Immunology             | PG 114 mg/dL       |
| HBs-Antigen negative   | Cr 1.23 mg/dL      |
| HBV-Antibody positive  | BUN 19.7 mg/dL     |
| HCV-Antibody positive  | UA 10.7 mg/dL      |
|                        | Na 142 mEq/L       |
|                        | K 3.3 mEq/L        |
|                        | Cl 104 mEq/L       |

Neutro: neutrocyte, Lym: lymphocyte, Eosino: eosinocyte, Mono: monocyte, RBC: red blood cell, Ht: hematocrit, PLT: platelet, HB: hepatitis B virus, HCV: hepatitis C virus, AST: aspartate transaminase, ALT: alanine transaminase, γ-GTP: γ-glutamyl transpeptidase, TP: total protein, Alb: albumin, LDH: lactate dehydrogenase, ALP: alkaline phosphatase, CK: creatine kinase, CHE: cholinesterase, TB: total bilirubin, CRP: C-reactive protein, PG: plasma glucose, Cr: creatinine, BUN: blood urea nitrogen, UA: uric acid, Na: Natrium, K: Kalium, Cl: chlorine

---

**Figure 1.** *Sarcocystis truncata* in venison muscle tissue (A), cysts of *Sarcocystis truncata* isolated from the venison causing food poisoning (B, C).
Figure 2. Bradyzoites of Sarcocystis truncata isolated from the venison causing food poisoning.

animals, which are its intermediate hosts, Sarcocystis species are present within cysts in the musculature, containing proliferating bradyzoites.

Many species of Sarcocystis have already been identified in other animals, including S. cruzi, S. hirsuta, and S. bovihominis in cattle; S. miescheriana, S. porcifelis, and S. suihominis in pigs; and S. tenella and S. mihoensis in sheep (3, 5-10). Among these, S. bovihominis and S. suihominis are known to be transmitted to humans, which are their final host (11), and cause a type of intestinal sarcocystosis after infection (12).

Infection with Sarcocystis in humans can result in a type of intestinal sarcocystosis (12). After eating raw beef containing the S. hominis cysts, patients in Spain experienced nausea, vomiting, abdominal discomfort, abdominal pain, and frequent diarrhea. Sporocysts of S. hominis were detected in a specimen of the patients’ diarrheal feces, confirming the diagnosis (13).

In the present case, the patient developed symptoms within 4 hours of ingestion, with recovery after 24 hours. Besides this patient, two people who had eaten the same venison showed similar symptoms. Given the duration, rapidity of onset, and clinical symptoms, we suspected food poisoning. However, multiple stool cultures on enriched and selective media gave negative results.

Recently, S. fayeri in horses (13) and Sarcocystis spp. in deer (4) have been reported as new food poisoning agents that induce gastrointestinal symptoms, such as nausea, vomiting, and diarrhea, within 24 hours of ingestion, similar to this case. We therefore suspected food poisoning to have been caused by Sarcocystis spp. Using polymerase chain reaction, the presence of the Sarcocystis spp. 18S rRNA gene was detected. Furthermore, Sarcocystis truncata was isolated from the venison.

To date, S. sybillensis, S. wapiti, and S. hofmanni have been identified in Japanese sika deer (C. nippon) (4, 14). S. truncata has been identified in white-tailed sea eagle (15) and red deer (C. elaphus) in Norway (16) but never in sika deer (C. nippon) in Japan. In 2018, S. truncata was identified in sika deer farmed in Lithuania (17). It is therefore surprising that S. truncata would be found in these animals in Japan.

Since 1969, the toxicity of Sarcocystis has been studied experimentally. Rabbits injected with Sarcocystis cysts developed rapid respiration, depression, and paralysis and eventually died (18). The protein extracts of S. gigantea are toxic to rats and mice (19, 20). A toxic protein fraction has also been isolated from S. cruzi cysts in beef (21). This protein had a molecular mass of approximately 15-kDa; however, precise studies on this toxic protein have not been performed.

A related 15-kDa protein produced by Sarcocystis spp. present in horsemeat evoked enteropathogenic changes in an animal model (22). Kamata et al. found that, following intravenous administration, a similar protein from S. fayeri induced diarrhea and lethal toxicity in rabbits and that it also exhibited enterotoxicity in an ileal loop test (13). This toxin showed homology with the actin-depolymerizing factor from Toxoplasma gondii and Eimeria tenella. Irikura et al. showed that a recombinant version of the same 15-kDa protein induced enterotoxicity in a rabbit ileal loop test (23). These findings showed that the 15-kDa protein from S. fayeri originating from horse meat was a diarrheal toxin functioning as a food poisoning agent in humans. We suspected that one or more toxins originating from S. truncata are able to induce pathological changes similar to those caused by S. cruzi and confirmed this by isolating a toxic 15-kDa protein from a strain isolated from the patient in the present case. It is reasonable to suspect that this protein was the cause of the gastrointestinal symptoms in this patient.

The cysts of Sarcocystis species lose viability following rapid freezing at -20°C, heating at 70°C for 1 minute, and in 2.0% salt for 1 day (14). Therefore, adequate freezing and heating are needed for the cooking and storage of venison in order to reduce the risk of food poisoning. The 15-kDa protein might be a useful indicator for determining the possibility of food poisoning induced by Sarcocystis spp. Further studies will be necessary to reveal whether or not the 15-kDa protein is responsible for the enterotoxicity induced by Sarcocystis detected in venison.

Some people prefer to serve their meat rare or raw, allowing consumers to inadvertently ingest live Sarcocystis cysts. Sarcocystis cysts in meat are therefore a risk factor for public health. To prevent food poisoning caused by venison, it is essential to understand how to inactivate the responsible

Figure 3. Immunoblotting pattern of the 15-kDa protein in Sarcocystis spp. M: Molecular weight marker, 1: control (uninfected horse meat), 2: horse meat infected with Sarcocystis, 3: venison infected with Sarcocystis truncata (our case).
toxin during storage. In *S. fayeri*, this can be accomplished by freezing (22). In the present case, the cause of food poisoning was failure to treat the venison at the proper temperature.

This is the first report to describe a case of food poisoning presumably caused by *S. truncata* from venison. While examining patients suffering from venison-related food poisoning, the possibility of infection by *Sarcocystis* spp. should be kept in mind.

The authors state that they have no Conflict of Interest (COI).

Acknowledgement
The authors would like to acknowledge the significant contribution of Akiko Yamazaki (Cooperative Department of Veterinary Medicine, Faculty of Agriculture, Iwate University, Japan) in providing the 15-kDa protein antibody.

References
1. Ministry of the Environment. [The population estimation of nihon deer and wild boar by statistical method] [Internet]. [cited 2018 Feb 22]. Available from: http://www.env.go.jp/press/files/jp/26914.pdf
2. Ruiz-Fons F. A review of the current status of relevant zoonotic pathogens in wild swine (*Sus scrofa*) populations: changes modulating the risk of transmission to humans. Transbound Emerg Dis 64: 68-88, 2017.
3. Markus MB. *Sarcocystis* and sarcocystosis in domestic animals and man. Adv Vet Sci Comp Med 22: 159-193, 1978.
4. Aoki K, Ishikawa K, Hayashi K, et al. An outbreak of suspected food poisoning related to deer meat containing *Sarcocystis* cysts. Jpn J Food Microbiol 30: 28-32, 2013 (in Japanese, Abstract in English).
5. Ashford RW. The fox, *Vulpes vulpes*, as a final host for *Sarcocystis* of sheep. Ann Trop Med Parasitol 71: 29-34, 1977.
6. Dubey JP, Streitel RH, Stromberg PC, Toussaint MJ. *Sarcocystis fayeri* sp. n. from the horse. J Parasitol 63: 443-447, 1977.
7. Fayer R, Esposito DH, Dubey JP. Human infections with *Sarcocystis* species. Clin Microbiol Rev 28: 295-311, 2015.
8. Saito M, Shibata Y, Kobayashi T, Kobayashi M, Mubo K, Itagaki H. Ultrastructure of the cyst wall of *Sarcocystis* species with canine final host in Japan. J Vet Med Sci 58: 861-867, 1996.
9. Saito M, Shibata Y, Kubo M, Itagaki H. *Sarcocystis mihoensis* sp. n. from sheep in Japan. J Vet Med Sci 59: 103-106, 1997.
10. Saito M, Shibata Y, Ohno A, Kubo M, Shimura K, Itagaki H. *Sarcocystis suihominis* detected for the first time from pigs in Japan.
11. Guo M, Dubey JP, Hill D, et al. Prevalence and risk factors for *Toxoplasma gondii* infection in meat animals and meat products destined for human consumption. J Food Prot 78: 457-476, 2015.
12. Fayer R. *Sarcocystis* spp. in human infections. Clin Microbiol Rev 17: 894-902, 2004.
13. Kamata Y, Saito M, Irikura D, et al. A toxin isolated from *Sarcocystis fayeri* in raw horsemeat may be responsible for food poisoning. J Food Prot 77: 814-819, 2014.
14. Honda M, Sawaya M, Taira K, et al. Effects of temperature, pH and curing on the viability of *Sarcocystis*, a Japanese sika deer (*Cervus nippon centralis*) parasite, and the inactivation of their diarrheal toxin. J Vet Med Sci 80: 1337-1344, 2018.
15. Gjerde B, Vikrøren T, Hammes IS. Molecular identification of *Sarcocystis halleri* n. sp., *Sarcocystis lari* and *Sarcocystis truncata* n. sp. based on mitochondrial cox1 sequences. Parasitology 141: 441-452, 2014.
16. Rudaitytė-Lukošienė E, Prakash P, Butkauskas D, Kukšienė L, Veptaitė-Monstavičė I, Servienė E. Morphological and molecular identification of *Sarcocystis* spp. from the sika deer (*Cervus nippon*), including two new species *Sarcocystis frondea* and *Sarcocystis nipponi*. Parasitol Res 117: 1305-1315, 2018.
17. Mandour AM. Studies on the toxicity of *Sarcocystis*. J Med Microbiol 2: 361-363, 1969.
18. Al-Hyali NS, Aljawady MA, Mohammad-Fakhri MA. The influence of some physio-chemical properties of sarcotoxin in rats. J Anim Vet Adv 9: 302-305, 2010.
19. Al-Hyali NS, Khalil LY, Aljawady MA. Sarcotoxin effect on leukocytic finding and phagocytic activity in mice. J Anim Vet Adv 8: 2395-2398, 2009.
20. Saito M, Taguchi K, Shibata Y, Kobayashi T, Shimura K, Itagaki H. Toxicity and properties of the extract from *Sarcocystis cruzi* cysts. J Vet Med Sci 57: 1049-1051, 1995.
21. Harada S, Furukawa M, Tokuoka E, et al. Control of toxicity of *Sarcocystis fayeri* in horsemeat by freezing treatment and prevention of food poisoning. Shokuhin Eiseigaku Zasshi (J Food Hyg Soc Japan) 54: 198-203, 2013 (in Japanese, Abstract in English).
22. Irikura D, Saito M, Sugita-Konishi Y, et al. Characterization of *Sarcocystis fayeri*’s actin-depolymerizing factor as a toxin that causes diarrhea. Genes Cells 22: 825-835, 2017.

The Internal Medicine is an Open Access journal distributed under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License. To view the details of this license, please visit (https://creativecommons.org/licenses/by-nc-nd/4.0/).