Review

Study on Supergenus *Rubus* L.: Edible, Medicinal, and Phylogenetic Characterization

Qinglin Meng 1, Hakim Manghwar 1 and Weiming Hu 1,*

Lushan Botanical Garden, Chinese Academy of Sciences, Jiujiang 332900, China; mengqinglin510@163.com (Q.M.); hakim@lsbg.cn (H.M.)
* Correspondence: huwm@lsbg.cn

Abstract: *Rubus* L. is one of the most diverse genera belonging to Rosaceae; it consists of more than 700 species with a worldwide distribution. It thus provides an ideal natural “supergenus” for studying the importance of its edible, medicinal, and phylogenetic characteristics for application in our daily lives and fundamental scientific studies. The *Rubus* genus includes many economically important species, such as blackberry (*R. fruticosus* L.), red raspberry (*R. idaeus* L.), black raspberry (*R. occidentalis* L.), and raspberry (*R. chingii* Hu), which are widely utilized in the fresh fruit market and the medicinal industry. Although *Rubus* species have existed in human civilization for hundreds of years, their utilization as fruit and in medicine is still largely inadequate, and many questions on their complex phylogenetic relationships need to be answered. In this review, we briefly summarize the history and progress of studies on *Rubus*, including its domestication as a source of fresh fruit, its medicinal uses in pharmacology, and its systematic position in the phylogenetic tree. Recent available evidence indicates that (1) thousands of *Rubus* cultivars were bred via time- and labor-consuming methods from only a few wild species, and new breeding strategies and germplasms were thus limited; (2) many kinds of species in *Rubus* have been used as medicinal herbs, though only a few species (*R. idaeus* L., *R. chingii* Hu, and *R. occidentalis* L.) have been well studied; (3) the phylogeny of *Rubus* is very complex, with the main reason for this possibly being the existence of multiple reproductive strategies (apomixis, hybridization, and polyploidization). Our review addresses the utilization of *Rubus*, summarizing major relevant achievements and proposing core prospects for future application, and thus could serve as a useful roadmap for future elite cultivar breeding and scientific studies.

Keywords: *Rubus* L.; raspberry fruit; medicinal components; phylogeny; omics

1. Introduction

*Rubus* L. is one of the most diverse and largest genera in the Rosaceae family. The genus consists of more than 700 shrubby or herbaceous species mainly distributed throughout the temperate zone of the northern hemisphere, with a few having expanded to the tropics and the southern hemisphere [1–6]. Species of the *Rubus* genus worldwide are classified into 12 subgenera [1–3]. However, Lu et al. [6] reclassified them into 8 subgenera, whereby only habitats in China were considered. There are two hypothetical centers of origin for *Rubus*: one is North America [7,8] and the other is southwestern China [9–11]. In addition, the pleasant flavor of the fresh *Rubus* fruit, its medicinal functions due to the high genetic diversity and complex phylogeny rendering it suitable for scientific studies, make *Rubus* an important and ideal genus for breeders as well as scientists [8,12,13] (Figure 1). Furthermore, the rich secondary metabolites and the bark of *Rubus* are also important raw materials for cosmetics and fiber [14].
Rubus bears aggregate drupetum fruits that have economically important edible and medicinal characteristics [12,13]. They have a pleasant flavor and have been dubbed “superfoods” due to their very high levels of secondary metabolites, such as hydrolyzable tannins, anthocyanins, polyphenols, flavanols, organic acids, and many other organic compounds [12,15–20]. In an early investigation, Moyer et al. [15] extracted multiple anthocyanins and phenols from the ripe fruits of Rubus. Based on genomic resequencing, quadrupole time-of-flight liquid chromatography, and mass spectroscopy, 29 hydrolyzable tannins and their candidate chromosomal regions were identified by Wang et al. [20]. Because of these diverse secondary metabolites, the superfood Rubus fruits can provide anti-oxidants as well as anti-cancer, anti-microbial, and anti-complement activities, in addition to having other benefits for humans [21–25].

Since the publication of Darwin’s *On the Origin of Species*, understanding the genetic basis of adaptation for the arisal of new species has been a central topic in evolutionary biology [26–29]. In the species-rich genus of Rubus, the diversity of reproductive strategy, such as through the process of hybridization, polyploidization, and apomixis, which enhances the adaptation capacity of Rubus [8,30,31]. Additionally, the various reproductive strategies also bring a huge challenge regarding the taxonomy of the Rubus genus in terms of morphological and molecular systematics [8,9,31–33]. However, in order to make better use of the wild germplasm in Rubus, it is necessary to develop a better knowledge of the clear affinity of its phylogeny. Therefore, species of the Rubus genus provide a natural experimental system for studying the fundamental mechanisms of adaptation via diverse reproductive strategies and reticulate evolutionary phylogeny.

**Figure 1.** Different aspects of Rubus species utilization. The attributes of Rubus are primarily utilized for applications involving fruit (fresh fruit, jam, and juice), medicinal compounds (fruit, leaf, and stem), and scientific studies (adaptation, reproduction, polyploidy, and evolution). In addition, among the wild Rubus species, some can also be used to produce cosmetics or fiber products. This figure was created using BioRender software.
Over the past hundred years since Focke [1] published *Species Ruborum*, our understanding of *Rubus* L. has improved, including regarding its edibility [4,12,30] and in the medicinal [4,13,15,18,20,34] and phylogenetic fields [7,8,31,35,36] (Figure 1). However, most previous studies and reviews have devoted their attention to metabolic compounds of pharmacologic interest in only a very few specific species [13,19]. This review summarizes the general outcomes for fresh fruit breeding, medicinal components, and studies into phylogenetic relationships of the *Rubus* genus; based on the latest advances in the fields of omics (genomics, transcriptomics, proteomics, and metabolomics), CRISPR/Cas, and other genome editing technologies, experimental efficiency has improved remarkably. Finally, we point out the important value of *Rubus* in fruit germplasms, medicinal research, and understanding complex phylogenetic relationships resulting from diverse adaptative reproduction strategies. In short, *Rubus* L., in the Rosaceae family, is an ideal natural “super-genus” for breeders, pharmacologists, and evolutionary biologists. Our review addresses major questions regarding how to better exploit the wild germplasm in *Rubus* species and can thus serve as a useful roadmap for future breeding and fundamental scientific studies.

2. Studies of Edible *Rubus* Species

*Rubus* L. belongs to the Rosaceae family, from which many palatable fruit cultivars have been bred, such as the woodland strawberry (*Fragaria vesca* L. var. *americana* Porter), and the domesticated apple (*Malus × domestica* Borkh.), pear (*Pyrus bretschneideri* Rehd.), and peach (*Prunus persica* (L.) Batsch) [1,9,37]. *Rubus* species are popular for their pleasant fresh fruits, including the blackberry (*R. fruticosus* L.), red raspberry (*R. idaeus* L.), and black raspberry (*R. occidentalis* L.). *Rubus* fruits are aggregate drupetum fruits with varied colors, such as red, yellow, purple, and black [12]. *Rubus* fruits have been called “superfoods” because of the very high levels of beneficial secondary metabolites that they contain, including, e.g., anthocyanins, phenolic acids, flavonoids, tannins, and other essential compounds [19,24,38–40] (Figure 2). The chemical composition of *Rubus* is influenced not only by environmental factors but also internal genetic differences. For example, studies of blackberries found that a temperate climate and higher cumulative rainfall increased the production of phenolic compounds [41]. The concentration of chemical components in *Rubus* is also related to the storage conditions, growing season or location, and maturity [42–45]. The genotype differences between species and cultivars in *Rubus* is the major internal factor that influences the divergence of chemical compositions; for instance, Skrovankova et al. [46] found that different cultivars show significant variations in the production of secondary metabolites, even when they were grown under the same environmental conditions. Furthermore, a series of studies on different *Rubus* genotypes, cultivars, and species produced results consistent with those of Skrovankova [41,44,47–49].

Species of the *Rubus* genus have been cultivated and appeared in gardens for more than 15 centuries in Europe, for example in Turkey and Rome [18,38]. To date, thousands of cultivars have been bred, and these can mainly be divided into two types: the primocane-fruiting (also called annual-fruiting) type, including Heritage, Amity, Autumn Bliss, Autumn Britten, Dinkum, and Polana cultivars; and the florican-fruiting (also called biennial-fruiting) type, including Claudia, Emily, Esta, Lauren, and Qualicum cultivars [8,12,38]. Among these, Logan, Boysen, and Marion are three elite cultivars. The major areas for growing these cultivars are Russia (125,000 t), North America (59,123 t), and Europe (43,000 t) [8,38]. Spurred by the human pursuit of fruit quality and increasing consumption, the fruit production of *Rubus* cultivars has rapidly expanded for the production of fresh fruit for use in jams and fruit juice [12,38,50,51] (Figure 1).
In the early stages of Rubus domestication, breeders commonly selected superior individuals from wild habitats [4,5]. During the nineteenth century, approximately 30 breeding projects were conducted in North America and Europe. The elite cultivars of Preussen, Cuthbert, and Newburgh were bred by crossing between different subspecies of red raspberries [30]. However, the process of domestication has vastly reduced the morphological and genetic diversities of crops [52–55]. Current cultivars are bred from crossing or the improvement of only a few wild species, i.e., red raspberry (R. idaeus L.), black raspberry (R. occidentalis L.), and blackberry (R. fruticosus L.). In order to meet the demands of visual appeal, higher yield, greater quality, excellent health benefits, and diverse adaptation for breeders, more wild germplasms of Rubus resources and advanced biotechnology should be utilized for Rubus breeding. Based on simple sequence repeat (SSR) and amplified fragment length polymorphism (AFLP) markers, the first linkage map for Rubus was constructed based on the crossing of two red raspberry cultivars, Glen Moy and Latham, from Europe and North America, respectively [56]. Bushakra et al. [57] constructed a genetic linkage map using 1218 markers by the crossing of S1 (R. occidentalis L.) and Latham (R. idaeus L.) and compared it with genomes of other genera in the rosa family, such as Fragaria L., Malus Mill., and Prunus L. That study reported a high consistency of collinearity of genomes between different genera of Rosaceae, and hundreds of new polymorphic genetic markers were found for future quantitative trait loci mapping studies. In recent years, different high-resolution markers and advanced sequencing methods have been applied for trait mapping or new wild germplasm identification in Rubus [58–62].

3. Medicinal Studies of Rubus

Rubus L. is one of the most species-rich genera in the Rosaceae family, but only a few species have been used as medicinal herbs [13,18,63]. According to the records of the ancient pharmacopoeias in Europe and China, Rubus species have been used as medicinal herbs for several centuries. The stems and leaves of blackberry (R. fruticosus L.) were soaked with white wine for use as an astringent poultice for wound healing and for difficulties during childbirth, as suggested by Hippocrates [18]. The dried unripe fruits of “Fu-Fen-Zi” (R. chingii Hu) were used to improve and enhance liver and kidney health [20,64]. More recently, as shown in Figure 2, many kinds of secondary metabolites...
with remarkable beneficial effects on humans have been extracted. Table 1 exhibits detailed information on the species, concentration, and part from which they were extracted. For instance, anthocyanins with an anti-oxidant function are extracted from the fruits of the blackberry [15,19]; flavonoids with anti-oxidant, anti-cancer, and anti-inflammatory effects are extracted from the fruits of the blackberry or Fu-Pen-Zi [16,65–67]. Other organic compounds have been identified, mainly in blackberry or Fu-Pen-Zi, such as hydrolyzable tannin, glycoprotein, organic acid, and phenolic compounds [19,21,61,68,69]. In addition, reports have indicated many other kinds of beneficial effect of these secondary metabolites on humans, for example, improving mitosis and eyesight, treating or preventing cancer, back pain, and frequent urination [23,25,70,71]. To date, most of the secondary metabolites mentioned above are considered safe according to data from limited studies [13,72–76]. For example, the extracted components from *R. niveus* Thunb. showed no statistically significant toxicity for mice [72]. Based on a cytotoxic experiment on Caco-2 cells, Ke et al. [77] found that metabolites extracted from the fruit of *R. chingii* Hu were safe and had a favourable effect on anti-cancer cells. Overall, the limited available data on the toxicity and allergenicity of *Rubus* species indicate they are safe for humans. More in-depth investigations regarding medicinal applications in pharmacology are needed.

In the genomic age, genomic sequencing of *Rubus* is already lagging behind compared to other major crops and fruits, such as rice, maize, cotton, apple, and pear [53,78–85]. However, benefiting from the quick development of the cost-effective next-generation sequencing (NGS) [86,87] and transcriptome (RNA-seq) technology [88,89], nuclear or plastid genomes have been sequenced for some important medicinal species in the *Rubus* genus [20,90–95]. Utilizing the RNA-seq data of the red raspberry (*R. idaeus* L.) fruit, Hyun et al. [90] determined the regulated candidate genes for biosynthesis of γ-aminobutyric acid and anthocyanins, which have anti-oxidant activity. Based on unripe Fu-Pen-Zi (*R. chingii* Hu) fruits, the chromosome-scale reference and genomic regions related to the biosynthetic pathway for hydrolyzable tannin (HT) have been reported [20]. Therefore, using the results of these studies, breeders could modify candidate genes or genomic regions of *Rubus* cultivars to improve the content of targeted secondary metabolites (HT, anthocyanins, etc.) using site-directed genome editing technologies such as CRISPR/Cas. More recently, in the field of crop breeding, CRISPR/Cas genome-editing technology has been used with encouraging results [96]. However, use of this speedy, proven, and precise genome editing technology has not reported in programs for breeding *Rubus*. To date, various studies taking advantage of transcriptomic analysis at different developmental stages (green; green and yellow; yellow, orange, and red) of Fu-Pen-Zi fruits have revealed that flavonoids and anthocyanins are synthesized at an early stage and their levels then decrease during subsequent development [61,97,98]. These studies also indicated that anthocyanins might not be responsible for the reddish color of ripe fruits. Thus, better characterization will require extraction of the pharmacological metabolites at the early stage of *Rubus* fruit development. Meanwhile, several studies on plastid genomes have focused on the pharmacological components of *Rubus*, i.e., *R. eucalyptus* Focke [93], *R. rufus* Focke [99], *R. longisepalus* Nakai and *R. hirsutus* Thunb. [100,101], and *R. phoenicolasius* Maxim. [94]. However, the detailed genetic basis and biosynthetic pathways of the pharmacological metabolites in *Rubus* species are still largely unclear, and further future investment and research on different aspects are needed [102–104].
Table 1. Major secondary metabolites of Rubus L.

| Secondary Metabolite | Species | Concentration * (mg/100 g) | Part          | References |
|----------------------|---------|----------------------------|---------------|------------|
| Anthocyanin          | R. chingii | 2.1~326                    | Leaf          | [20,39,40,105,106] |
|                      | R. fruticosus |                     | Fruit         |            |
|                      | R. ideaus         |                               |               |            |
|                      | R. hirsutus        |                               |               |            |
| Flavonoid            | R. chingii | 2.8~6                      | Leaf          | [20,34,40,107] |
|                      | R. occidentalis |                             | Fruit         |            |
| Phenolic compounds   | R. chingii | 13.7~1541                  | Stem          | [19,20,39,40,48,105,106,108] |
|                      | R. setchuenensis  |                              | Leaf          |            |
|                      |                     |                              | Fruit         |            |
| Organic acids        | R. chingii | 0.2~52.9                   | Stem          | [20,109]   |
|                      | R. coreanus |                               | Leaf          |            |
| Glycoprotein         | R. chingii | 14.6~81.4                  | Fruit         | [63]       |

* The concentration data were collected from multiple studies in which the extraction method, part, and species varied, and thus the data presented in the table show minimum and maximum values.

4. Phylogenetic Studies of Rubus

The Rubus genus is one of the most successful models of an adaptive and evolutionary group, with distribution worldwide except for Antarctica [1,9]. As shown in Table 2, Rubus species were classified by Focke into 12 or by Lu into 8 subgenera according to worldwide distribution or distribution in China, respectively. The classification and phylogenetic construction of Rubus is a challenging task due to phenomena such as hybridization, apomixis, polyploidization, and introgression, which happen frequently in this genus. Ploidy levels among different subgenera and species are highly differentiated [7,32,33,35,58] (Table 2). More importantly, the diverse reproductive strategies may have conferred to Rubus species the ability to occupy various habitats worldwide, and thus demonstrate reticulate evolutionary phylogeny. According to previous phylogenetic analysis based on the ndhF gene, Howarth et al. [110] suggested that the Hawaiian Islands species (R. hawaiensis A. Gray and R. macraei A. Gray) originated from different ancestors, in contradiction with the morphological results. The reticulate evolution of Rubus has been indicated in recent studies. Wang et al. [31] used multiple chloroplast and nuclear genes to investigate the phylogenetic relationships of 142 Rubus taxa, which indicated reticulate evolutionary events between different subgenera and species. A study based on approximately 1000 target genes constructed the phylogenetic tree for 87 wild Rubus taxa and three cultivars, concluding that hybridization and incomplete lineage sorting (ILS) were responsible for the low resolution and topological conflicts between different subgenera, which were not caused by insufficient molecular signals [8]. Furthermore, it has been suggested that North America might be the primary center of origin of Rubus, which then expanded into Asia and Europe and finally dispersed to Oceania via birds [8].
Table 2. List of Rubus L. subgenera.

| Subgenus         | Code | Species in Subgenus | Ploidy Level (x = 7) | References          |
|------------------|------|---------------------|----------------------|---------------------|
| Anoplobatus      | An   | 9                   | 2x                   | [8,30,111,112]      |
| Chamaebatus      | Cb   | 6 (5)               | 2x, 6x               | [8,30,31]           |
| Chamaemorus      | Cm   | 1 (1)               | 6x, 8x               | [8,30]              |
| Comaropsis       | Co   | 2                   | 4x                   | [8,30]              |
| Cylactis         | Cy   | 18 (8)              | 2x–4x                | [8,30,31]           |
| Dalibarda        | Da   | 5                   | 2x                   | [8,30]              |
| Dalibardastrum   | Da   | 15 (10)             | 4x, 6x               | [8,30,31,111–113]   |
| Idaeobatus       | Id   | 125 (83)            | 2x, 3x, 4x, 13x, 18x | [8,20,30,31,111–113]|
| Lampobatus       | Lm   | 10 (1)              | 4x                   | [8,30]              |
| Malachobatus     | Ma   | 104 (85)            | 4x, 6x, 8x, 14x      | [30,31,111–113]     |
| Orobatas         | Or   | 16                  | 6x                   | [8,30]              |
| Rubus            | Ru   | 444 (1)             | 2x–12x               | [8,30,111,112]      |

The number within brackets is the corresponding number of Rubus species in China.

Additionally, a series of studies on the phylogeny of Rubus detected conflicts in the phylogenetic affinities between plastid genes and nuclear genes in most cases [36,114,115]. The molecular and morphological toptaxies also appear inconsistent, which may result from the multiple reproductive strategies [36,111,114–117]. In general, the difficulties of morphological or molecular taxonomy in subgenera and between species are not caused by lack of characteristics or signals; the real reason may be the diverse reproductive patterns that have made this group an ideal genus for investigation of the genetic basis of different reproductive and adaptive patterns.

5. Concluding Remarks and Future Perspectives

The Rubus genus consists of more than 700 species, but only a few of them, such as blackberry (R. fruticosus L.), red raspberry (R. ideaus L.), and black raspberry (R. occidentalis L.), have been domesticated or crossed by breeders to generate elite cultivars with excellent characteristics of strong adaptability, good storage properties, and pest or disease resistance. Unfortunately, there have been relatively few molecular breeding studies on Rubus and fewer genomic resources exist compared to other types of crops and fruits. Rubus breeders can reference those studies in order to improve the breeding methods for elite cultivars of different crops and fruits.

Furthermore, the situation for medicinal cultivars of Rubus is worse, and most of the investments in pharmacology have been concerned with R. ideaus L., R. fruticosus L., and R. chingii Hu, rarely involving residual species such as R. eucalyptus Focke, R. occidentalis L., and R. phoenicolasius Maxim., which have also been used as medicinal ingredients for hundreds of years. Notably, there remains a large deficiency in the study of the basic mechanisms and genetics of the active ingredients in medicinal Rubus species. Fortunately, advances in NGS and RNA-seq technologies offer an opportunity for researchers to spend less money and labor investigating the above-mentioned problems and to quickly identify and choose high-quality germplasms.

Finally, reconstructing the phylogenetic relationships for Rubus is a task made challenging by hybridization, polyploidization, apomixis, and introgression. However, researchers can also combine consideration of morphological characteristics with omics technologies (i.e., genomics, transcriptomics, proteomics, and metabolomics) to decipher the phylogenetic and evolutionary puzzles of Rubus. Consumers in the present era are increasingly demanding tastier and healthier fresh fruits. The wild species and elite cultivars of Rubus provide ideal candidates to address this demand due to their pleasant flavor and high concentrations of secondary metabolites. However, only a few wild species have been domesticated and are used in our daily food markets and medical treatment. Therefore, in order to better exploit the abundance of Rubus wild germplasms, information on their phylogenetic relationships and genetic diversity should be clarified. This study reviewed
three major topics (edible, medicinal, and phylogenetic properties), but many challenges still exist in the utilization and research of Rubus. Working as a team and applying the latest omics strategies may open the door for developing a series of satisfactory elite germplasms for fruit and medicine, and reveal the central evolutionary phenomenon resulting in reticulate evolution.

**Author Contributions:** Conceptualization, W.H. and Q.M.; writing—original draft preparation, Q.M.; preparation of diagram and editing, Q.M. and H.M.; writing—review and editing, Q.M. and W.H.; supervision, W.H. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the Lushan Botanical Garden, Chinese Academy of Sciences, grant number 2021ZWZX11, and the National Natural Science Foundation of China, grant number 32160999.

**Institutional Review Board Statement:** Not applicable.

**Data Availability Statement:** Data sharing not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest.

**References**

1. Focke, W.O. *Species Ruborum Monographiae Generis Rubi Prodromus*; Bibliotheca Botanica; E. Schweizerbart: Stuttgart, Germany, 1910.
2. Focke, W.O. *Species Ruborum Monographiae Generis Rubi Prodromus*; Bibliotheca Botanica; E. Schweizerbart: Stuttgart, Germany, 1911.
3. Focke, W.O. *Species Ruborum Monographiae Generis Rubi Prodromus*; Bibliotheca Botanica; E. Schweizerbart: Stuttgart, Germany, 1914.
4. Jennings, D. *Raspberries and Blackberries: Their Breeding, Disease and Growth*; Academic Press: New York, NY, USA, 1988.
5. Janick, J.; Moore, J.N. *Fruit Breeding, Vine and Small Fruits*; Wiley: New York, NY, USA, 1996; Volume II, pp. 109–190.
6. Lu, L.T.; Gu, C.Z.; Li, C.L.; Alexander, C.; Bartholomew, B.; Brach, A.R.; Boufford, D.E.; Ikeda, H.; Ohba, H.; Robertson, S.A.; et al. *Rubus Linnaeus. Flora China* 2003, 9, 195–285.
7. Alice, L.A.; Campbell, C.S. Phylogeny of Rubus (rosaceae) based on nuclear ribosomal DNA internal transcribed spacer region sequences. *Ann. J. Bot.* 1999, 86, 81–97. [CrossRef] [PubMed]
8. Carter, K.A.; Liston, A.; Bassil, N.V.; Alice, L.A.; Bushakra, J.M.; Sutherland, B.L.; Mockler, T.C.; Bryant, D.W.; Hummer, K.E. Target Capture Sequencing Unravels Rubus Evolution. *Front. Plant Sci.* 2019, 10, 1615. [CrossRef] [PubMed]
9. Gu, Y.; Sun, Z.J.; Cai, J.H.; Huang, Y.S.; He, S.A. Introduction and utilization of small fruits in China, with special reference to Rubus species. *Acta Hortic.* 1989, 262, 47–56. [CrossRef] [PubMed]
10. Kalkman, C. The phylogeny of the Rosaceae. *Bot. J. Linn. Soc.* 1988, 98, 37–59. [CrossRef]
11. Guo, Q.L.; Gao, J.Y.; Yang, J.S. Analysis of Bioactive Triterpenes from Rubus chingii Hu by Cyclodextrin-Modified Capillary Electrophoresis. *Chromatographia* 2005, 62, 145–150. [CrossRef]
12. Zhu, Y.A.; Li, C.; Wang, S.; Lei, L.; Xiao, J. The complete chloroplast genome of Rubus setchuenensis, an edible and medicinal dual-purpose wild plant. *Mitochondrial DNA Part B* 2022, 7, 228–230. [CrossRef]
13. Moyer, R.A.; Hummer, K.E.; Finn, C.E.; Frei, B.; Wrolstad, R.E. Anthocyanins, phenolics, and antioxidant capacity in diverse small fruits: Vaccinium, rubus, and ribes. *J. Agric. Food Chem.* 2002, 50, 519–525. [CrossRef]
14. Guo, Q.L.; Gao, J.Y.; Yang, J.S. Analysis of Bioactive Triterpenes from Rubus chingii Hu by Cyclodextrin-Modified Capillary Electrophoresis. *Chromatographia* 2005, 62, 145–150. [CrossRef]
15. Guo, Q.L.; Yang, J.S.; Liu, J.X. Studies on Chemical Constituents in Fruits of Rubus chingii. *Chin. Pharm. J.* 2007, 42, 1141–1143.
16. Hummer, K.E.; Janick, J. Rubus Iconography: Antiquity to the Renaissance. *Acta Hortic.* 2007, 759, 89–106. [CrossRef]
17. Kaume, L.; Howard, L.R.; Devareddy, L. The blackberry fruit: A review on its composition and chemistry, metabolism and bioavailability, and health benefits. *J. Agric. Food Chem.* 2012, 60, 5716–5727. [CrossRef] [PubMed]
18. Wang, L.; Lei, T.; Han, G.; Yue, J.; Zhang, X.; Yang, Q.; Ruan, H.; Gu, C.; Zhang, Q.; Qian, T.; et al. The chromosome-scale reference genome of Rubus chingii Hu provides insight into the biosynthetic pathway of hydrolyzable tannins. *Plant J.* 2021, 107, 1466–1477. [CrossRef]
19. Sun, N.; Wang, Y.; Liu, Y.; Guo, M.-L.; Yin, J. A new ent-labdane diterpene saponin from the fruits of Rubus chingii. *Chem. Nat. Compd.* 2013, 49, 49–53. [CrossRef]
22. Zhang, T.-T.; Yang, L.; Jiang, J.-G. Bioactive comparison of main components from unripe fruits of Rubus chingii Hu and identification of the effective component. Food Funct. 2015, 6, 2205–2214. [CrossRef]
23. Zhong, B.; Guo, Q.; Zhou, G.; Fu, H.; Wan, K. Three new labdane-type diterpene glycosides from fruits of Rubus chingii and their cytotoxic activities against five tumor cell lines. Fitoterapia 2015, 102, 23–26. [CrossRef]
24. Li, K.; Xia, X.X.; Ding, Y.H.; Zhou, W.Y.; Xie, B.S.; Cai, J.B.; Xie, Y.H.; Huang, L.P. Effect of Different Extract Parts From Rubi Fructus on Improving Memory Disorder in Mice. Chin. J. Exp. Tradit. Med. Formulae 2016, 22, 142–147. [CrossRef]
25. Li, K.Y.; Zeng, M.L.; Li, Q.L.; Zhou, B.H. Identification of polyphenolic composition in the fruits of Rubus chingii Hu and its antioxidant and antiproliferative activity on human bladder cancer T24 cells. J. Food Meas. Charact. 2018, 13, 51–60. [CrossRef]
26. Rieseborg, L.H.; Willis, J.H. Plant speciation. Science 2007, 317, 910–914. [CrossRef] [PubMed]
27. Jones, F.C.; Grabherr, M.G.; Chan, Y.F.; Russell, P.; Mauceli, E.; Johnson, J.; Swofford, R.; Pirun, M.; Zody, M.C.; White, S.; et al. The genomic basis of adaptive evolution in threespine sticklebacks. Nature 2012, 484, 55–61. [CrossRef] [PubMed]
28. Theis, A.; Ronco, F.; Indermaur, A.; Saizburger, W.; Egger, B. Adaptive divergence between lake and stream populations of an East African cichlid fish. Mol. Ecol. 2014, 23, 5304–5322. [CrossRef] [PubMed]
29. Ferris, K.G.; Barnett, L.L.; Blackman, B.K.; Willis, J.H. The genetic architecture of local adaptation and reproductive isolation in sympatry within the Mimulus guttatus species complex. Mol. Ecol. 2017, 26, 208–224. [CrossRef]
30. Thompson, M.M. Survey of Chromosome Numbers in Rubus (Rosaceae: Rosoideae). Ann. Mo. Bot. Gard. 1997, 84, 128–164. [CrossRef]
31. Wang, Y.; Chen, Q.; Chen, T.; Tang, H.; Liu, L.; Wang, X. Phylogenetic Insights into Chinese Rubus (Rosaceae) from Multiple Chloroplast and Nuclear DNAs. Front. Plant Sci. 2016, 7, 968. [CrossRef]
32. Tutin, T.G.; Heywood, V.H.; Burges, N.A.; Moore, D.M.; Valentine, D.H.; Walters, S.M.; Webb, D.A. (Eds.) Flora Europaea; Cambridge University Press: Cambridge, UK, 1980; Volume 5, 452p.
33. Thompson, M.M.; Zhao, C.M. Chromosome numbers of Rubus Species in southwest China. Acta Horticult. 1993, 352, 493–502. [CrossRef]
34. Zhang, T.-T.; Wang, M.; Yang, L.; Jiang, J.-G.; Zhao, J.-W.; Zhu, W. Flavonoid glycosides from Rubus chingii Hu fruits display anti-inflammatory activity through suppressing MAPKs activation in macrophages. J. Funct. Foods 2015, 18, 235–243. [CrossRef]
35. Alice, L.A.; Eriksson, T.; Erikson, B.; Campbell, C.S. Hybridization and Gene Flow Between Distantly Related Species of Rubus (Rosaceae): Evidence from Nuclear Ribosomal DNA Internal Transcribed Spacer Region Sequences. Syst. Bot. 2001, 26, 769–778. [CrossRef]
36. Sochor, M.; Vašut, R.J.; Sharbel, T.F.; Trévniček, B. How just a few makes a lot: Speciation via reticulation and apomixis on example of European brambles (Rubus subgen. Rubus, Rosaceae). Mol. Phylogen. Evol. 2015, 89, 13–27. [CrossRef]
37. Xiang, Y.; Huang, C.-H.; Hu, Y.; Wen, J.; Li, S.; Yi, T.; Chen, H.; Xiang, J.; Ma, H. Evolution of Rosaceae Fruit Types Based on Nuclear Phylogeny in the Context of Geological Times and Genome Duplication. Mol. Biol. Evol. 2017, 34, 262–281. [CrossRef] [PubMed]
38. Kim, M.J.; Sutton, K.K.; Harris, G.K. Raspberries and Related Fruits. In Encyclopedia of Food and Health; Caballero, B., Finglas, P.M., Toldrá, F., Eds.; Academic Press: Oxford, UK, 2016; pp. 586–591.
39. Štivičar, T.; Wrobel, R.E.; Finn, C.E.; Pereira, C.B. Influence of cultivar, maturity, and sampling on blackberry (Rubus, L. Hybrids) anthocyanins, polyphenolics, and antioxidant properties. J. Agric. Food Chem. 2004, 52, 8021–8030. [CrossRef] [PubMed]
40. Nile, S.H.; Park, S.W. Edible berries: Bioactive components and their effect on human health. Nutrition 2014, 30, 134–144. [CrossRef] [PubMed]
41. Croge, C.P.; Cuquiel, F.L.; Pintor, P.; Biasi, L.A.; Bona, C. Antioxidant Capacity and Polyphenolic Compounds of Blackberries Produced in Different Climates. HortScience A Publ. Am. Soc. Hortic. Sci. 2019, 54, 2209–2213. [CrossRef]
42. Ancos, B.d.; González, E.M.; Cano, M.P. Ellagic acid, vitamin C, and total phenolic contents and radical scavenging capacity affected by freezing and frozen storage in raspberry fruit. J. Agric. Food Chem. 2000, 48, 4565–4570. [CrossRef]
43. Haffner, K.; Rosenfeld, H.J.; Skrede, G.; Wang, L. Quality of red raspberry Rubus idaeus L. cultivars after storage in controlled and normal atmospheres. Postharvest Biol. Technol. 2002, 24, 279–289. [CrossRef]
44. Mazur, S.P.; Nes, A.; Wold, A.B.; Remberg, S.F.; Aaby, K. Quality and chemical composition of ten red raspberry (Rubus idaeus L.) genotypes during three harvest seasons. Food Chem. 2014, 160, 233–240. [CrossRef]
45. Wang, S.Y.; Chen, C.-T.; Wang, C.Y. The influence of light and maturity on fruit quality and flavonoid content of red raspberries. Food Chem. 2009, 112, 676–684. [CrossRef]
46. Skrovankova, S.; Sumczynski, D.; Mieck, J.; Jurikova, T.; Sochor, J. Bioactive Compounds and Antioxidant Activity in Different Types of Berries. Int. J. Mol. Sci. 2015, 16, 24673–24706. [CrossRef]
47. Gülçin, I.; Topal, F.; Çakmakçı, R.; Bilsel, M.; Gören, A.C.; Erdogan, U. Pomological Features, Nutritional Quality, Polyphenol Content Analysis, and Antioxidant Properties of Domesticated and 3 Wild Ecotype Forms of Raspberries (Rubus idaeus L.). J. Food Sci. 2011, 76, C585–C593. [CrossRef]
48. Bobinaitė, R.; Viškelis, P.; Venskutonis, P.R. Variation of total phenolics, anthocyanins, ellagic acid and radical scavenging capacity in various raspberry (Rubus spp.) cultivars. Food Chem. 2012, 132, 1495–1501. [CrossRef] [PubMed]
49. Verma, R.; Gangrade, T.; Punasiya, R.; Ghulaxe, C. Rubus fruticosus (blackberry) use as an herbal medicine. Pharmacogn. Rev. 2014, 8, 101–104. [CrossRef] [PubMed]
50. Strik, B.; Clark, J.; Finn, C.; Banados, M. Worldwide Blackberry Production. HortTechnology 2007, 17, 205–213. [CrossRef]
51. Hassani, S.; Shariatpanahi, M.; Tavakoli, F.; Nili-Ahmadabadi, A.; Abdollahi, M. The changes of bioactive ingredients and antioxidant properties in various berries during jam processing International Journal of Biosciences | IJB. Int. J. Biosci. IJB 2015, 6, 172–179. [CrossRef]

52. Tanksley, S.D.; McCouch, S.R. Seed banks and molecular maps: Unlocking genetic potential from the wild. Science 1997, 277, 1060–1066. [CrossRef]

53. Clark, R.M.; Linton, E.; Messing, J.; Doebley, J.F. Pattern of diversity in the genomic region near the maize domestication gene tb1. Proc. Natl. Acad. Sci. USA 2004, 101, 700–707. [CrossRef]

54. Flint-Garcia, S.A. Genetics and consequences of crop domestication. J. Agric. Food Chem. 2013, 61, 8267–8276. [CrossRef]

55. Li, J.; Yuan, D.; Wang, F.; Wang, Q.; Sun, M.; Liu, Z.; Li, H.; Xu, Z.; Ma, Y.; Zhang, B.; et al. Cotton pan-genome retrieves the lost sequences and genes during domestication and selection. Genome Biol. 2021, 22, 119. [CrossRef]

56. Graham, J.; Smith, K.; MacKenzie, K.; Jorgenson, L.; Hackett, C.; Powell, W. The construction of a genetic linkage map of red raspberry (Rubus idaeus subsp. idaeus) based on AFLPs, genomic-SSR and EST-SSR markers. Theor. Appl. Genet. Theor. Angew. Genet. 2004, 109, 740–749. [CrossRef]

57. Bushakra, J.M.; Stephens, M.J.; Atmadjaja, A.N.; Lewers, K.S.; Symonds, V.V.; Udall, J.A.; Chagné, D.; Buck, E.J.; Gardiner, S.E. Construction of black (Rubus occidentalis) and red (R. idaeus) raspberry linkage maps and their comparison to the genomes of strawberry, apple, and peach. Theor. Appl. Genet. Theor. Angew. Genet. 2012, 125, 311–327. [CrossRef]

58. Randell, R.; Howarth, D.; Morden, C. Genetic analysis of natural hybrids between endemic and alien Rubus (Rosaceae) species in Hawai‘i. Conserv. Genet. 2004, 5, 217–230. [CrossRef]

59. Castillo, N.; Reed, B.M.; Graham, J.; Fernández-Fernández, F.; Bassil, N. Microsatellite Markers for Raspberry and Blackberry. J. Am. Soc. Hortic. Sci. 2010, 135, 271–279. [CrossRef]

60. Bushakra, J.M.; Lewers, K.S.; Staton, M.E.; Zhebentyayeva, T.; Sasaki, C.A. Developing expressed sequence tag libraries and the discovery of simple sequence repeat markers for two species of raspberry (Rubus L.). BMC Plant Biol. 2015, 15, 258. [CrossRef]

[PubMed]

61. Chen, Z.; Jiang, J.; Shu, L.; Li, X.; Huang, J.; Qian, B.; Wang, X.; Li, X.; Chen, J.; Xu, H. Combined transcriptomic and metabolic analyses reveal potential mechanism for fruit development and quality control of Chinese raspberry (Rubus chinii Hu). Plant Cell Rep. 2021, 40, 1923–1946. [CrossRef]

62. Zhou, Z.M.; Yan, D.M.; Wang, Y.K.; Zhang, T.; Xiao, X.R.; Dai, M.Y.; Zhang, S.W.; Liu, H.N.; Li, F. Discovery of quality markers in Rubus chinii Hu using UPLC-ESI-QTOF-MS. J. Pharm. Biomed. Anal. 2021, 203, 114200. [CrossRef] [PubMed]

63. Zeng, H.J.; Liu, Z.; Wang, Y.P.; Yang, D.; Yang, R.; Qu, L.B. Studies on the anti-aging activity of a glycoprotein isolated from Rubus chingii Hu: An overview of botany, traditional uses, phytochemistry, and pharmacology. Chin. J. Nat. Med. 2020, 18, 401–416. [CrossRef]

64. Xie, Y.H.; Lian, B.; Gong, J.H.; Tu, L.D.; Zhang, Y.T.; Huang, L.P. Preparation of Magnetic Chitosan Hyamine Microspheres and Separation of Phenolic Acids from Rubus Chinii Hu. Adv. Mater. Res. 2013, 634–638, 1347–1351. [CrossRef]

65. Cai, Y.Q.; Hu, J.H.; Qin, J.; Sun, T.; Li, X.L. Rhododendron Molle (Ericaceae): Phytochemistry, pharmacology, and toxicology. Chin. J. Nat. Med. 2018, 16, 0401–0410. [CrossRef]

66. Li, X.; Sun, J.; Chen, Z.; Jiang, J.; Jackson, A. Characterization of carotenoids and phenolics during fruit ripening of Chinese raspberry (Rubus chinii Hu). RSC Adv. 2021, 11, 10804–10813. [CrossRef] [PubMed]

67. Ding, H.Y. Extracts and constituents of Rubus chinii with 1,1-diphenyl-2-picrylhydrazyl (DPPH) free radical scavenging activity. Int. J. Mol. Sci. 2011, 12, 3941–3949. [CrossRef] [PubMed]

68. Yao, X.; Zhu, W.R.; Huang, H.L.; Zeng, Y.R.; Yu, W.W. Effective medicinal ingredients and screening of excellent germplasm in Rubus chinii. China J. Chin. Med. Mater. 2016, 46, 575–581. [CrossRef]

69. Madrigal-Gamboa, V.; Jiménez-Arias, J.; Hidalgo, O.; Quesada, S.; Pérez, A.M.; Azofeifa, G. Membrane processing effect of blackberry (Rubus adriaticus) on cytotoxic and pro-apoptotic activities against cancer cell lines. J. Food Process. Preserv. 2021, 45, e15575. [CrossRef]

70. Xu, W.; Zhao, M.; Fu, X.; Hou, J.; Wang, Y.; Shi, F.; Hu, S. Molecular mechanisms underlying macrophage immunomodulatory activity of Rubus chinii Hu polysaccharides. Int. J. Biol. Macromol. 2021, 185, 907–916. [CrossRef]

71. Tolentino, F.; Araújo, P.A.D.; Marques, E.D.S.; Petreanu, M.; Andrade, S.F.D.; Perazzo, F.F.; Rosa, P.C.P.; Maistro, E.L. In vivo evaluation of the genetic toxicity of Rubus niveus Thunb. (Rosaceae) extract and initial screening of its potential chemoprevention against doxorubicin-induced DNA damage. J. Ethnopharmacol. 2015, 164, 89–95. [CrossRef] [PubMed]

72. Kanegusuku, M.; Benassi, J.C.; Pedrosa, R.C.; Yunes, R.A.; Filho, V.C.; Maia, A.A.; de Souza, M.M.; Delle Monache, F.; Niero, R. Cytotoxic, hypoglycemic activity and phytochemical analysis of Rubus imperialis (Rosaceae). Z. Naturforsch. C J. Biosci. 2002, 57, 272–276. [CrossRef] [PubMed]

73. Om, A.-S.; Song, Y.-N.; Noh, G.; Kim, H.; Choe, J. Nutrition Composition and Single, 14-Day and 13-Week Repeated Oral Dose Toxicity Studies of the Leaves of Rubus coreanus Miquel. Molecules 2016, 21, 65. [CrossRef]

74. AlQahtani, F.S.; AlShebly, M.M.; Govindarajan, M.; Senthilmurugan, S.; Vijayan, P.; Benelli, G. Green and facile biosynthesis of silver nanocomposites using the aqueous extract of Rubus ellipticus leaves: Toxicity and oviposition deterrent activity against Zika virus, malaria and filariasis mosquito vectors. J. Asia-Pacific Entomol. 2017, 20, 157–164. [CrossRef]
Plants 2022, 11, 1211

76. Ali, N.; Shoaib, M.; Shah, S.W.A.; Shah, I.; Shuaib, M. Pharmacological profile of the aerial parts of Rubus ulmifolius Schott. BMC Complement. Altern. Med. 2017, 17, 59. [CrossRef]

77. Ke, H.; Bao, T.; Chen, W. New function of polysaccharide from Rubus chinensis Hu: Protective effect against ethyl carbamate induced cytotoxicity. J. Sci. Food Agric. 2021, 101, 3156–3164. [CrossRef]

78. Kawahara, Y.; de la Bastide, M.; Hamilton, J.P.; Kanamori, H.; McCombie, W.R.; Ouyang, S.; Schwartz, D.C.; Tanaka, T.; Wu, J.; Zhou, S.; et al. Improvement of the Orzya sativa Nipponbare reference genome using next generation sequence and optical map data. Rice 2013, 6, 4. [CrossRef] [PubMed]

79. Jiao, Y.; Peluso, P.; Shi, J.; Liang, T.; Stitzer, M.C.; Wang, B.; Campbell, M.S.; Stein, J.C.; Wei, X.; Chin, C.-S.; et al. Improved maize reference genome with single-molecule technologies. Nature 2017, 546, 524–527. [CrossRef] [PubMed]

80. Wang, M.; Tu, L.; Yuan, D.; Zhu, D.; Shen, C.; Li, J.; Liu, F.; Pei, L.; Wang, P.; Zhao, G.; et al. Reference genome sequences of two cultivated allotetraploid cottons, Gossypium hirsutum and Gossypium barbadense. Nat. Genet. 2019, 51, 224–229. [CrossRef] [PubMed]

81. Velasco, R.; Zharkikh, A.; Affourtit, J.; Dhingra, A.; Cestaro, A.; Kalyanaraman, A.; Fontana, P.; Bhatnagar, S.K.; Troggio, M.; Pruss, D.; et al. The genome of the domesticated apple (Malus × domestica Borkh.). Nat. Genet. 2010, 42, 833–839. [CrossRef] [PubMed]

82. Wu, J.; Wang, Z.; Shi, Z.; Zhang, S.; Ming, R.; Zhu, S.; Khan, M.A.; Tao, S.; Korban, S.S.; Wang, H.; et al. The genome of the pear (Pyrus bretschneideri Rehd.). Genome Res. 2013, 23, 396–408. [CrossRef] [PubMed]

83. Benz, B.F. Archaeological evidence of teosinte domestication from Guíll Naquitz, Oaxaca. Proc. Natl. Acad. Sci. USA 2001, 98, 2104–2106. [CrossRef]

84. Huang, X.; Yang, S.; Gong, J.; Feng, Q.; Zhan, Q.; Zhao, Y.; Li, W.; Cheng, B.; Xia, J.; et al. Genomic architecture of heterosis for yield traits in rice. Nature 2016, 537, 629–633. [CrossRef]

85. Chen, P.; Li, Z.; Zhang, D.; Shen, W.; Xie, Y.; Zhang, J.; Jiang, L.; Li, X.; Shen, X.; Geng, D.; et al. Insights into the effect of human civilization on Malus evolution and domestication. Plant Biotechnol. J. 2021, 19, 2206–2220. [CrossRef]

86. Varshney, R.K.; Nayak, S.N.; May, G.D.; Jackson, S.A. Next-generation sequencing technologies and their implications for crop genetics and breeding. Trends Biotechnol. 2009, 27, 522–530. [CrossRef]

87. Ray, S.; Satya, P. Next generation sequencing technologies for next generation plant breeding. Front. Plant Sci. 2014, 5, 367. [CrossRef]

88. Ramirez-Gonzalez, R.H.; Segovia, V.; Bird, N.; Fenwick, P.; Holdgate, S.; Berry, S.; Jack, P.; Caccamo, M.; Uaay, C. RNA-Seq bulked segregant analysis enables the identification of high-resolution genetic markers for breeding in hexaploid wheat. Plant Biotechnol. J. 2015, 13, 613–624. [CrossRef]

89. Julio, E.; Malpica, A.; Cotucheau, J.; Bachet, S.; Volpatti, R.; Decorps, C.; Dorlhac de Borne, F. RNA-Seq analysis of Orobanche resistance in Nicotiana tabacum: Development of molecular markers for breeding recessive tolerance from ‘Wika’ tobacco variety. Euphytica 2019, 216, 6. [CrossRef]

90. Hyun, T.K.; Lee, S.; Kumar, D.; Rim, Y.; Kumar, R.; Lee, S.Y.; Lee, C.H.; Kim, J.Y. RNA-seq analysis of Rubus idaeus cv. Nova: Transcriptome sequencing and de novo assembly for subsequent functional genomics approaches. Plant Cell Rep. 2014, 33, 1617–1628. [CrossRef]

91. VanBuren, R.; Bryant, D.; Bushakra, J.M.; Vining, K.J.; Edger, P.P.; Rowley, E.R.; Priest, H.D.; Michael, T.P.; Lyons, E.; Filichkin, S.A.; et al. The genome of black raspberry (Rubus occidentalis). Plant J. 2016, 87, 535–547. [CrossRef] [PubMed]

92. VanBuren, R.; Wai, C.M.; Colle, M.; Wang, J.; Sullivan, S.; Bushakra, J.M.; Liaeho, I.; Vining, K.J.; Dossett, M.; Finn, C.E.; et al. A near complete, chromosome-scale assembly of the black raspberry (Rubus occidentalis) genome. GigaScience 2018, 7, giz094. [CrossRef] [PubMed]

93. Zhu, Q.; Tian, Y.; Liang, W. The complete chloroplast genome sequence of Rubus eucalyptus (Rosaceae). Mitochondrial DNA Part B Resour. 2019, 4, 2976–2977. [CrossRef]

94. Zhang, G.; Liu, Y.; Hai, P. The complete chloroplast genome of Tibetan medicinal plant Rubus phoenicolasius Maxim. Mitochondrial DNA Part B Resour. 2021, 6, 886–887. [CrossRef]

95. Davik, J.; Reen, D.; Lyse, E.; Buti, M.; Rossman, S.; Alsheikh, M.; Aiden, E.L.; Dudchenko, O.; Sargent, D.J. A chromosome-level genome sequence assembly of the red raspberry (Rubus idaeus L.). PLoS ONE 2022, 17, e0265096. [CrossRef]

96. Zhu, J.K. The Future of Gene-Edited Crops in China. Natl. Sci. Rev. 2022, nwa063. [CrossRef]

97. Li, X.; Jiang, J.; Chen, Z.; Jackson, A. Transcriptomic, Proteomic and Metabolomic Analysis of Flavonoid Biosynthesis During Fruit Maturation in Rubus chinii Hu. Front. Plant Sci. 2021, 12, 706667. [CrossRef]

98. Li, X.; Sun, J.; Chen, Z.; Jiang, J.; Jackson, A. Metabolite profile and genes/proteins expression in β-citraturin biosynthesis during fruit ripening in Chinese raspberry (Rubus chinii Hu). Plant Physiol. Biochem. 2021, 163, 76–86. [CrossRef] [PubMed]

99. Huang, W.; Qiao, F.; Guo, W.; Wu, W. Characterization of the complete chloroplast genome sequence of Rubus rufus Focke (Rosaceae). Mitochondrial DNA Part B Resour. 2021, 6, 3093–3094. [CrossRef] [PubMed]

100. Park, Y.S.; Park, J.Y.; Kang, J.H.; Lee, W.H.; Yang, T.J. Diversity and authentication of Rubus accessions revealed by complete plastid genome and rDNA sequences. Mitochondrial DNA Part B Resour. 2021, 6, 1454–1459. [CrossRef] [PubMed]

101. Wang, Q.; Huang, Z.; Gao, C.; Ge, Y.; Cheng, R. The complete chloroplast genome sequence of Rubus hirsutus Thunb. and a comparative analysis within Rubus species. Genetica 2021, 149, 299–311. [CrossRef]
102. Su, X.H.; Duan, R.; Sun, Y.Y.; Wen, J.F.; Kang, D.G.; Lee, H.S.; Cho, K.W.; Jin, S.N. Cardiovascular effects of ethanol extract of Rubus chingii Hu (Rosaceae) in rats: An in vivo and in vitro approach. *J. Physiol. Pharmacol. Off. J. Pol. Physiol. Soc.* 2014, 65, 417–424.

103. He, Y.; Jin, S.; Ma, Z.; Zhao, J.; Yang, Q.; Zhang, Q.; Zhao, Y.; Yao, B. The antioxidant compounds isolated from the fruits of Chinese wild raspberry Rubus Chingii Hu. *Nat. Prod. Res.* 2018, 34, 872–875. [CrossRef]

104. Chen, Y.; Chen, Z.; Guo, Q.; Gao, X.; Ma, Q.; Xue, Z.; Ferri, N.; Zhang, M.; Chen, H. Identification of Ellagitannins in the Unripe Fruit of Rubus Chingii Hu and Evaluation of its Potential Antidiabetic Activity. *J. Agric. Food Chem.* 2019, 67, 7025–7039. [CrossRef]

105. Wang, S.Y.; Lin, H.S. Antioxidant activity in fruits and leaves of blackberry, raspberry, and strawberry varies with cultivar and developmental stage. *J. Agric. Food Chem.* 2000, 48, 140–146. [CrossRef]

106. Fu, Y.; Zhou, X.; Chen, S.; Sun, Y.; Shen, Y.; Ye, X. Chemical composition and antioxidant activity of Chinese wild raspberry (Rubus hirsutus Thunb.). *LWT Food Sci. Technol.* 2015, 60, 1262–1268. [CrossRef]

107. Han, N.; Gu, Y.; Ye, C.; Cao, Y.; Liu, Z.; Yin, J. Antithrombotic activity of fractions and components obtained from raspberry leaves (Rubus chingii). *Food Chem.* 2012, 132, 181–185. [CrossRef]

108. Sellappan, S.; Akob, C.C.; Krewer, G. Phenolic compounds and antioxidant capacity of Georgia-grown blueberries and blackberries. *J. Agric. Food Chem.* 2002, 50, 2432–2438. [CrossRef] [PubMed]

109. Lee, M.J.; Nam, J.H.; Jeong, J.H.; Rho, I.R. Effect of plant part, extraction method, and harvest time over antioxidant yield of rubus coreanus. *Pharmacogn. Mag.* 2020, 16, 455–461.

110. Howarth, D.G.; Gardner, D.E.; Morden, C.W. Phylogeny of Rubus subgenus Idaeobatus (Rosaceae) and its implications toward colonization of the Hawaiian Islands. *Syst. Bot.* 1997, 22, 433–441. [CrossRef]

111. Morden, C.W.; Gardner, D.E.; Weniger, D.A. Phylogeny and biogeography of pacific Rubus subgenus Idaeobatus (Rosaceae) species: Investigating the origin of the endemic Hawaiian raspberry *R. macraei*. *Pac. Sci.* 2003, 57, 181–197. [CrossRef]

112. Wang, Y.; Chen, Q.; Chen, T.; Zhang, J.; He, W.; Liu, L.; Luo, Y.; Sun, B.; Zhang, Y.; Tang, H.-r.; et al. Allopolyploid origin in Rubus (Rosaceae) inferred from nuclear granule-bound starch synthase I (GBSSI) sequences. *BMC Plant Biol.* 2019, 19, 303. [CrossRef] [PubMed]

113. Wang, Y.; Chen, Q.; Tang, H.; Wang, X. Phylogeny of Chinese Rubus (Rosaceae) based on nuclear ribosomal internal transcribed spacer (ITS) sequences. *Acta Hortic.* 2018, 1208, 13–24. [CrossRef]

114. Alice, L.A.; Dodson, T.M.; Sutherland, B.L. Diversity and relationships of Bhutanese Rubus (Rosaceae). *Acta Hortic.* 2008, 777, 63–70. [CrossRef]

115. Yang, J.Y.; Pak, J.-H. Phylogeny of korean rubus (rosaceae) based on its (nrDNA) and trnL/F intergenic region (cpdna). *J. Plant Biol.* 2006, 49, 44–54. [CrossRef]

116. Zhang, L.; Wang, Y.; Chen, Q.; Luo, Y.; Zhang, Y.; Tang, H.R.; Wang, X.R. Phylogenetic Utility of Chinese Rubus (Rosaceae) Based on ndhF Sequence. *Acta Hortic. Sin.* 2015, 42, 19–30. [CrossRef]

117. Imanishi, H.; Nakahara, K.; Tsuyuzaki, H. Genetic relationships among native and introduced Rubus species in Japan based on rbcL sequence. *Acta Hortic.* 2008, 769, 195–199. [CrossRef]