Copy Number Variants Associated with 14 Cases of Self-Injurious Behavior

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Abstract

Copy number variants (CNVs) were detected and analyzed in 14 probands with autism and intellectual disability with self-injurious behavior (SIB) resulting in tissue damage. For each proband we obtained a clinical history and detailed behavioral descriptions. Genetic anomalies were observed in all probands, and likely clinical significance could be established in four cases. This included two cases having novel, de novo copy number variants and two cases having variants likely to have functional significance. These cases included segmental trisomy 14, segmental monosomy 21, and variants predicted to disrupt the function of ZEB2 (encoding a transcription factor) and HTR2C (encoding a serotonin receptor). Our results identify variants in regions previously implicated in intellectual disability and suggest candidate genes that could contribute to the etiology of SIB.

Introduction

Self-injurious behavior (SIB) is a serious behavioral disorder exhibited by many individuals with autism spectrum disorder (ASD) and other forms of developmental disability. It is characterized by self-inflicted physical injury to one’s own body [1], with common topographies of SIB including head banging on hard surfaces, head and body hitting, self-biting, eye poking, self-scratching, and hair pulling. The prevalence of SIB is estimated at 5 to 17% of persons with intellectual disability [2], and approximately 50% of individuals displaying SIB are also diagnosed with ASD [3]. Left untreated, SIB usually persists and evolves to include further topographies of injury and often has significant adverse effects on child and family quality of life.
SIB likely has a multifactorial etiology, with some portion of SIB cases arising from an interaction between genetic and environmental factors during development. Prior research has established that single gene mutations cause syndromes with increased risk for SIB as part of the phenotype, including Lesch-Nyhan syndrome [4], Rett syndrome [5], Fragile X syndrome [6], and the Cornelia de Lange syndrome [7]. In some cases copy number variants (CNVs) affecting these single genes also have causal roles [6]. We hypothesized that structural variants such as chromosomal deletions and duplications may also underlie some nonsyndromic cases of SIB in individuals with ASD.

In this study we recruited participants (probands and family) from the Neurobehavioral Unit (NBU) at the Kennedy Krieger Institute, an inpatient unit for children with disabilities and severe behavior disorders. We applied high-density single nucleotide polymorphism (SNP) microarray analysis to DNA derived from these patients. The present study was restricted to children with ASD together with SIB. Based on previous findings of genetic factors associated with both ASD and SIB we hypothesized that this population represents an enrichment of single nucleotide variants (SNVs), deletions, and duplications contributing to the SIB phenotypes. Given the heterogeneity of SIB phenotypes, we expected that the variants we identified could each account for only one specific case of SIB. Due to the expected limitations in associating recurrent variants with phenotypes, we applied widely adopted criteria to aid in identifying potentially causal alterations [8].

**Materials and Methods**

All studies were performed with written informed consent and approval of the Institutional Review Board at Johns Hopkins University. Probands were recruited from the NBU at the Kennedy Krieger Institute.

**Inclusion Criteria**

Probands with an ASD and at least one topography of SIB were recruited for enrollment. Autism diagnosis was confirmed by a child psychiatrist upon admission to the NBU. Probands had at least one biological parent available for additional genetic analyses.

**Vineland Adaptive Behavior Scales**

Measures of adaptive behavior were obtained from parents for 11 of the 14 probands using the Vineland Adaptive Behavior Scales (Vineland)[9]. While other instruments generally relevant to autism are available, such as the revised Autism Diagnostic Interview (ADI-R) or Autism Diagnostic Observation Schedule (ADOS), those measures are not routinely used for the population with severe behavioral disturbances. The first or second edition of the Vineland [9] was administered depending on the time of admission to the inpatient hospital. Both editions of the Vineland measure adaptive behavior across three domains: Communication (i.e., expressive, receptive, and written language skills), Daily Living Skills (i.e., personal, domestic, and community-based skills), and Socialization (i.e., interpersonal relationships, play and leisure, and coping skills). The second edition also includes a Motor Skills domain (i.e., fine and gross motor skills). Raw scores from each domain are converted into standard scores (M = 100, SD = 15) and an adaptive behavior composite score (M = 100, SD = 15). In addition, the standard scores within each domain and the adaptive behavior composite are categorized as one of five levels of adaptive functioning: low, moderately low, adequate, moderately high, and high.
Functional behavioral assessment

Functional analyses [10] of SIB were conducted for all participants to identify the variables that evoke and maintain SIB. The functional analysis is part of the standard of care for all patients admitted to the NBU; and for assessment of SIB, in general. During the standard functional analysis, between four and six conditions (i.e., alone, attention, demand, tangible with toys, and/or tangible with food, and toy play), each typically lasting 10 minutes, are presented in a multi-element (or alternating treatments) design. Each test condition is designed to simulate an environment the individual is likely to encounter, with variables hypothesized to give rise to and maintain problem behavior manipulated systematically. Responding in those conditions is compared to a control condition simulating an enriched environment where attention and toys are freely available. Behavioral data were graphically depicted and interpreted using criteria such as those developed by Hagopian et al. that consider magnitude of effect, stability, and trend [11].

Sample collection and DNA extraction

We obtained blood samples from each proband, and from each parent or sibling we obtained a blood or saliva sample. We isolated DNA from each participant using a PureGene kit (QIA-GEN, Valencia, CA). Lymphoblast cell lines were established from some blood samples for the purpose of performing fluorescence in-situ hybridization.

SNP arrays

DNA samples were assayed using the Affymetrix GenomeWide 6.0 platform at the Johns Hopkins Genomic Analysis and Sequencing Core (Baltimore, MD). Genotyping was performed using the birdseed-v2 algorithm [12]. Data have been deposited in the Database of Genotypes and Phenotypes (dbGaP) at the National Institutes of Health (http://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000337.v1.p1).

CNV Analysis

Copy number segmentation was performed using PennCNV [13]. Only samples with a logR ratio standard deviation of less than 0.4 were analyzed. In families where trios (proband and both biological parents) were available, the PennCNV joint trio Hidden Markov Model (HMM) was used to incorporate copy number from the entire trio. Using an R script, copy number variants were annotated by intersection with Database of Genomic Variants [14], NCBI RefSeq genes [15], and The Centre for Applied Genomics Autism Chromosome Rearrangement Database (TCAG-ACRD)[16]. We defined a CNV as a hemizygous or homozygous deletion, amplification of three or more copies, and greater than 20 kilobases.

Criteria for inclusion of candidate loci

Based on guidelines developed by the American College of Medical Genetics (ACMG) [8], uniform criteria to define CNVs of potential significance were applied. That group provided evidence-based analysis showing that for individuals with unexplained ID, ASD, or multiple congenital abnormalities (MCAs), microarrays offer greater diagnostic yield (15–20%) for genetic testing than karyotyping. Our seven primary criteria indicating that a CNV is possibly pathogenic are that: (1) a CNV is altered relative to unaffected parents; (2) a CNV is similar to one in an affected relative; (3) the CNV is not completely contained within a region of genomic imbalance as defined by a CNV database of healthy individuals, employing a similarly high-resolution technology; (4) a CNV overlaps a genomic imbalance in a database of probands with
ID/DD, ASD, or MCAs; (5) a CNV overlaps a locus for a known genomic imbalance syndrome; (6) a CNV contains morbid Online Inheritance in Man (OMIM) genes; or (7) a CNV is gene rich. The International Standards for Cytogenomic Arrays (ISCA) criteria were also consistent with those of the ACMG [17]. Both the ISCA and ACMG sets of guidelines represent strategies that help interpret whether variants are pathogenic, benign, or of unknown significance. As many of these sources of evidence as possible were applied to each interpretation in this study. For example, for cases in which only one parent was available it was not always possible to confirm whether particular CNVs were inherited or de novo.

**FISH**

Fluorescence in-situ hybridization (FISH) was performed using fosmids obtained from the BACPAC Resource Center (Children's Hospital Oakland Research Institute). These were G248P8731F5 (chr2:145,173,324–145,216,003 spanning 42,680 bp in GRCh37) and G248P85468F6 (chr2:145,516,578–145,557,736 spanning 41,159 bp in GRCh37). Fosmids were nick translation labeled with spectrum green or orange dUTP (Abbott Molecular Diagnostics) and hybridized to lymphoblastoid cell lines. Local copy number was determined by counting fluorescence signals from metaphase and interphase cells.

**Database analyses**

We accessed the DECIPHER database (v5.1, GRCh37 accessed on March 8, 2012). The DECIPHER project includes information on patients with chromosomal microdeletions and microduplications [18]. We obtained a comprehensive list of cases and chromosomal loci involving those areas relevant to the current analysis. Of these, entries were excluded having a classification of “Familial inherited from normal parent” or “Unknown—parents not analyzed/uninformative.”

**Results**

Demographic and clinical summaries of the 14 studied probands are given in Table 1. The ages of probands, of whom 12 (~86%) were male, ranged from 6 to 17 years, with an average age of 12. For the sample of children included in the current study for whom the Vineland was conducted, scores in the Communication, Daily Living Skills, and Socialization domains all fell within the low range of adaptive functioning. Similarly, all children scored in the low range of adaptive functioning for the Adaptive Behavior Composite. For the Motor Skills domain (assessed with Probands 7, 8, 10, 11, 12, 13, and 14), 57.1% of the sample fell within the low range, 28.6% fell within the moderately low range, and 14.3% fell within the adequate range. Table 2 depicts the standard scores across domains.

Following interviews with caregivers and direct observation of the targeted problem behaviors, behavioral staff members established distinct operational definitions for each topography for each patient. All participants were reported to engage in at least two topographies of SIB (mean = 3.9, range = 2–7 topographies). Similar to the results obtained by Kahng et al. [19], the most commonly observed topographies of SIB included hitting one’s head with an open or closed fist or object, biting oneself (often targeting the hands, wrists, or arms), and banging one’s head against surfaces. Head-hitting, self-biting, and head-banging were reported to occur in 86%, 71%, and 64% of all 14 cases, respectively (S1 Table). We identified the targeted body regions when participants engaged in SIB, other problem behavior, and the functional analysis outcomes for all topographies of SIB (S2 Table).

We determined the percentage of functional analysis outcomes for each reported topography of SIB (n = 55) (S2 Table). It should be noted that some topographies may have been
counted in more than one functional analysis outcome category if the topography was observed to have multiple functions (e.g., if head-banging were observed to be maintained by access to attention and to preferred toys, it would be counted in the Attention, Tangible–Toy, and Multi-
ply Maintained categories). Functional analysis results for SIB are summarized in Table 3.

Genotyping was completed for a total of 40 individuals, including: (1) three father, mother, proband, and sibling quartets (Probands 1, 4, 9), (2) five father, mother, and proband trios (Probands 3, 7, 10, 12, 14), (3) one father, proband, and sibling trio (proband 6), and (4) five parent/child pairs (Probands 2, 5, 8, 11, 13). To confirm that the pedigree structures were correct as obtained from family interviews, and to identify large-scale chromosomal changes, we

Table 1. Demographic and clinical summary of probands.

| Proband | Age | Sex | Psychiatric and Medical Diagnoses |
|---------|-----|-----|----------------------------------|
| 1       | 13  | F   | Autism spectrum disorder (ASD), disruptive behavior disorder not otherwise specified (DBD NOS), stereotypic movement disorder with self-injurious behavior (SMD with SIB), moderate intellectual disability (ID), seizure disorder. |
| 2       | 16  | M   | ASD, severe ID, DBD, SMD with SIB, and a history of Stevens Johnson Syndrome |
| 3       | 12  | M   | ASD, severe ID, SMD with SIB, unspecified disturbance of conduct, and impulsive control disorder NOS |
| 4       | 9   | M   | ASD, Severe ID, agenesis of the corpus callosum, cerebral palsy, and thrombocytopenia |
| 5       | 10  | M   | ASD and moderate ID |
| 6       | 17  | M   | ASD, SMD with SIB, unspecified disturbance of conduct, and severe ID |
| 7       | 13  | M   | ASD, SMD with SIB, DBD NOS, obsessive compulsive disorder, bipolar affective disorder, and unspecified level of ID |
| 8       | 14  | M   | ASD, SMD with SIB, DBD NOS, and history of multiple small bowel ulcers of unclear etiology |
| 9       | 12  | M   | ASD, SMD with SIB, and unspecified disturbance of conduct |
| 10      | 13  | F   | ASD, SMD with SIB, DBD NOS, Mood Disorder NOS, and profound ID |
| 11      | 6   | M   | ASD, DBD NOS, SMD with SIB and moderate ID |
| 12      | 9   | M   | ASD, DBD, SMD with SIB, mood disorder, and impulse control disorder |
| 13      | 17  | M   | ASD, DBD (NOS), severe ID, SMD with SIB, and mood disorder (NOS) |
| 14      | 11  | M   | ASD, obsessive-compulsive disorder, moderate ID, and unspecified mood disorders |

Table 2. Standard scores from Vineland and Vineland-II for each domain and the adaptive behavior composite.

| Proband | Communication | Daily Living Skills | Socialization | Motor Skills | Adaptive Behavior Composite |
|---------|--------------|---------------------|---------------|-------------|------------------------------|
| 1       | <20          | <20                 | <20           | N/A         | <20                          |
| 4       | 54           | 48                  | 53            | N/A         | 52                           |
| 6       | <20          | <20                 | <20           | N/A         | <20                          |
| 7       | 45           | 50                  | 45            | 94          | 46                           |
| 8       | 48           | 36                  | 46            | 59          | 38                           |
| 9       | 57           | 57                  | 50            | N/A         | 55                           |
| 10      | 34           | 35                  | 38            | 56          | 33                           |
| 11      | 42           | 40                  | 53            | 59          | 46                           |
| 12      | 38           | 40                  | 48            | 30          | 39                           |
| 13      | 33           | 43                  | 48            | 78          | 40                           |
| 14      | 53           | 59                  | 40            | 81          | 51                           |

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analyzed the genotype data using identity-by-state (IBS) and identity-by-descent (IBD) methods. This allowed us to estimate Cotterman coefficients of relatedness of all individuals in the study. We analyzed all pairwise relationships between the 40 individuals in this study (n = 780 pairwise comparisons) using kcoeff software to analyze autosomal SNP data [20] (data not shown). This confirmed the expected number of parent-child relationships (n = 29), full siblings (n = 4), and unrelated individuals and allowed us to confirm that all 14 pedigrees were correct as annotated. We noted a slight decrease in IBD1 (with a corresponding increase in IBD0) between proband 4 and mother, consistent with a chromosomal deletion (see below).

Table 3. Participant SIB topographies and functional analysis outcomes.

| Proband | SIB topography                                      | Functional analysis outcome                                      |
|---------|-----------------------------------------------------|------------------------------------------------------------------|
| Proband 1 | Head-hitting, self-biting, body-hitting, chin-banging | Multiply maintained: Escape from demands and access to tangible stimuli |
| Proband 2 | Head-hitting, self-biting, head-banging, body-hitting, self-pinning, self-scratching, hand/wrist-banging | Multiply maintained: Access to edible stimuli and access to attention |
| Proband 3 | Head-hitting, self-biting, hand/wrist-banging       | Automatic reinforcement                                          |
|         | Head-banging                                        | Access to edible stimuli                                         |
|         | Head-banging, self-biting, head-banging             | Automatic reinforcement                                          |
|         | Head-banging on body (including foot, knee, leg)    | Low to zero rates observed in FA, function not identified         |
| Proband 5 | Head-hitting                                        | Multiply maintained                                              |
|         | Self-biting                                         | Low to zero rates observed in FA, function not identified         |
| Proband 6 | Head-banging                                        | Automatic reinforcement                                          |
|         | Head-hitting, self-biting                           | Escape from demands                                              |
| Proband 7 | Head-hitting, self-biting, body-hitting, self-scratching | Low to zero rates observed in FA, function not identified        |
| Proband 8 | Self-biting, head-banging                          | Multiply maintained: Access to tangible stimuli and automatic reinforcement |
|         | Head-biting, self-scratching, self-pinning          | Automatic reinforcement                                          |
| Proband 9 | Head-hitting, body-hitting, hair-pulling, chin-banging/pressing | Multiply maintained: Access to attention, escape from demands, automatic reinforcement |
|         | Head-banging                                        | Low to zero rates observed in FA, function not identified         |
| Proband 10 | Head-hitting, head-banging, body-hitting, self-pinning, eye-SIB | Automatic reinforcement |
| Proband 11 | Head-hitting, head-banging                          | Low to zero rates observed in FA, function not identified         |
| Proband 12 | Self-biting, hair-pulling                           | Low to zero rates observed in FA, possible access to attention    |
| Proband 13 | Head-hitting, body-hitting, skin-picking, self-scratching, body-slamming | Low to zero rates observed in FA, function not identified        |
| Proband 14 | Self-biting, body-hitting                           | Access to attention                                               |
|         | Head-banging                                        | Automatic reinforcement                                          |

Abbreviation: FA, functional analysis.

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For four of the 14 cases (Probands 1–4) we identified a CNV likely to have a causal role. We describe these in the following paragraphs. For Probands 5–14 we identified CNVs that are less likely to have causal roles because they are: (a) inherited from a normal parent, (b) located in genomic regions lacking disease-associated genes, or (c) at locations lacking any genes. In other cases only one parent was available for this study, making conclusive interpretation of genetic findings difficult and these cases were classified as negative to be consistent with conservative criteria for identifying likely causal alterations. However, the possibility of some of these alterations being causal cannot be ruled out definitively, and the true percentage of positive cases could be higher. For Probands 5–14 we describe clinical and behavioral phenotypes as well as CNVs in S1 File.

**Proband 1.** The proband was a female reported to have global developmental delay at 6 months of age with failure to meet milestones and spastic quadraparesis and was subsequently diagnosed with autism and developmental delay. Proband 1’s SIB included head-hitting, self-biting, body-hitting, and chin-banging. Results of her functional analysis suggested that her SIB was multiply maintained by escape from demands and access to preferred toys.

CNV analysis indicated two contiguous copy number variations. The first was a de novo amplification spanning 491 kb on chromosome 2q22.3, immediately followed by a heterozygous deletion region (spanning 141 kb) (Fig 1A). These copy number changes were observed with a log R ratio plot (Fig 1B). We confirmed both the amplification and deletion by FISH. Amplification (3 signals) was observed in 64% of the interphase cells counted for the proband vs. 16% observed in the control (examples shown in Fig 1C and 1E). For the deletion we counted 10 metaphase cells, each of which had only one signal in the proband (example shown in Fig 1D) and two signals were observed in the control in 5 metaphase cells (example in Fig 1F). Affymetrix analysis software determined the amplified region was from two to three copies for 127 kb immediately followed by amplification to four copies for the remaining 364 kb (see Fig 1A). However the log R ratio plot (Fig 1B) and the FISH probe in the putative region of four copies both indicate that the amplification produced a total of three copies.

The amplification region overlapped with the ZEB2 gene encoding zinc finger E-box binding homeobox 2, a DNA-binding transcriptional repressor. Loss-of-function mutations in ZEB2 cause Mowat-Wilson syndrome, an autosomal dominant disorder characterized by intellectual disability, delayed motor development, and epilepsy. However, the proband’s dysmorphism was distinct from the characteristic features of Mowat-Wilson syndrome. Only deletions, inversions, and frameshift mutations in ZEB2 have been reported in individuals with Mowat-Wilson syndrome. To this date, no amplifications of ZEB2 have been reported in the literature.

**Proband 2.** The proband was a male diagnosed with ASD, severe intellectual disability, disruptive behavior disorder, and stereotypical movement disorder with self-injurious behavior. Proband 2 engaged in head-hitting, self-biting, head-banging, body-hitting, self-pinchng, self-scratching, and hand/wrist banging. Results of his functional analysis suggested that his SIB was multiply maintained by access to preferred foods and access to attention. He had a history of Stevens-Johnson syndrome (OMIM #608579; susceptibility to severe cutaneous adverse reaction). A karyotype had been performed in 1994 at age 2, reported as 46,XY,-7,+der(7)t(7;14)(p22;q13)psu dic(14)(q13). This indicated a 7;14 translocation with trisomy for proximal 14q and possible monosomy for 7p.

Copy number segmentation, performed on the proband and his mother, indicated a large gain on 14q11.2-q13.1 from 19.4 to 34.0 Mb, spanning 14.6 Mb (Fig 2). This region included 256 RefSeq genes. Approximately 40 cases of mosaic trisomy 14 have been reported [21,22], but not the segmental trisomy observed in this proband. There were no corresponding duplications in the DECIPHER database. Trisomy 14 mosaicism is characterized by growth and
Fig 1. Evidence of a tandem amplification and heterozygous deletion at the ZEB2 locus in proband 1.

(A) Genomic landscape (from the UCSC Genome Browser, GRCh36/hg18, a ~2 Mb region spanning chr2:144,000,482-145,945,938 corresponding to all SNPs on the Affymetrix array from 144–146 Mb). (B) LogR ratio plot of amplification/deletion region for proband 1 and control (from the GRCh36/hg18, chr2:144,000,482-145,945,938). Vertical bars indicate the beginning and end of the amplification and deletion regions. Red line indicates a moving average. (C) Representative example of interphase FISH with labeled genetic variation in the ZEB2 locus, Proband 1. (D) Hemizygous deletion region, Proband 1. (E) Euploid ZEB2 locus, control. (F) Euploid deletion region, control.
psychomotor retardation, dysmorphic craniofacial features (e.g. broad nose, dysplastic ears, micrognathia), congenital heart anomalies, and genitourinary abnormalities [23]. Mental development is normal or near normal (e.g. [24]), and individuals with mosaic trisomy 14 have not been reported to display SIB-related behavior.

Other large CNVs included a 1.8 Mb amplification on chromosome 15q11.1-q11.2, a region associated with intellectual disability and developmental delay [25]; and a 131 kb hemizygous deletion on 22q11.21. This region spanned the DiGeorge syndrome critical region gene 5 and 6 (DGCR5 and DGCR6) as well as the proline dehydrogenase 1 (PRODH) gene implicated in schizophrenia and hyperprolinemia type 1.

Stevens-Johnson syndrome has been associated with HLA-class I alleles on chromosome 6p22.1 and 6p21.33. We observed no CNVs at those loci, nor did we detect any copy number change consistent with chromosome 7 segmental monosomy.

**Proband 3.** The proband was a male exhibiting SIB resistant to medication treatment since age 2. Proband 3 engaged in head-hitting, self-biting, head-banging, and hand/wrist banging. Results of his functional analysis suggested that his head-hitting, self-biting, and hand/wrist banging were maintained by automatic reinforcement. Head-banging was found to be maintained by access to preferred foods. He demonstrated hundreds of self-injurious and aggressive behaviors hourly. Diagnoses included ASD, severe intellectual disability, stereotypic movement disorder with self-injury, unspecified disturbance of conduct, and impulse control disorder not otherwise specified. Over the course of six years he was placed on 28 medication trials but no medication afforded any significant sustained reduction in self-injury or aggression. Based on careful consideration of this history and restricted treatment options, the decision was made to fosmid G248P8731F5 using lymphoblast cell lines derived from proband 1 and (E) an unaffected control. Note the presence of three copies in the patient (panel C, white arrow indicating a duplication homolog). (D) Representative example of metaphase FISH using a probe from the deletion region (fosmid G248P85468F6). (F) Metaphase FISH in a euploid control reveals the expected two chromosomal copies of this fosmid probe.

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Fig 2. Region of segmental trisomy on chromosome 14 in proband 2. (A) The region spans 14.5 Mb (GRCh37/hg19 chr14:19,487,381–33,947,341). (B) The amplification region includes 404 RefSeq genes. Additional tracks show DECIPHER regions of chromosomal imbalances (deletions in red, duplications in blue) and OMIM disease genes.

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A. Ideogram of chromosome 14

B. Amplification region

RefSeq genes

OMIM genes

DECIPHER regions
consider electroconvulsive shock therapy. This was employed successfully, significantly reducing episodes of SIB with minimal side effects and contributed to healing and salvation of vision [26].

We detected a de novo 176 kb hemizygous deletion on Xq23 upstream of the HTR2C gene encoding 5-hydroxytryptamine (serotonin) receptor 2C (Fig 3). Given the role of HTR2C in modulating synaptic transmission, this represents a candidate gene for the proband’s disease phenotype. The deletion region is flanked on the 5’ side by a gap in the GRCh38 assembly, spanning 70 kb. The deletion region resides in a potential regulatory region, extending 66 kb upstream of the 5’ end of the HTR2C gene. Within the deletion region, a CpG island corresponds to a DNase I hotspot and to a position strongly likely to have regulatory function based on data from the ENCODE project [27] (Fig 3).

**Proband 4.** The proband was an 8 1/2 year old male with severe intellectual disability, ASD, agenesis of the corpus callosum, cerebral palsy, and thrombocytopenia. Proband 4’s SIB included head-hitting, self-biting, head-banging (against hard surfaces), and head-banging against the foot, knee, or leg or other body parts. Head-hitting, self-biting, and head-banging against hard surfaces were observed to be maintained by automatic reinforcement.
Chromosomal analysis revealed 21q segmental monosomy spanning 6.5 Mb (Fig 4F). Proband 4 and his mother had a Cotterman’s coefficient k1 value of 0.9950, while all other parent/child relationships had values of 1.0 ± 0.0002. This is consistent with the slight loss of IBD1 sharing that occurs when an individual has a heterozygous chromosomal deletion that introduces a region of homozygosity.

The deletion region included 131 RefSeq genes, including seven genes annotated as disease-causing in OMIM: (1) SOD1 encoding superoxide dismutase-1, (2) MRAP encoding melanocortin 2 receptor accessory protein, (3) IFNAR2 encoding Interferon alpha, beta, and omega, receptor 2, (4) CRFB4 encoding cytokine receptor, family II, member 4, (5) IFNGR2 encoding interferon gamma receptor-2, (6) KCNE1 encoding a potassium voltage-gated channel, and (7) RUNX1 encoding runt-related transcription factor 1.

Lyle and colleagues described 13 cases of partial monosomy 21, noting that most deletions occurred in proximal or distal 21q but not in a central region from physical position ~30–36 Mb (Fig 4B) [28]. They proposed that the phenotypic consequence of deletions in this central region would be too severe to be compatible with survival. We and others described additional 21q segmental monosomy cases that partially overlap this central region; additional examples have been reported in the DECIPHER database (Fig 4C-4E). The deletion region of proband 4 is notable because it is the only one to span this central region (Fig 4F). Furthermore, the

![Image](https://example.com/image.png)

**Fig 4.** Heterozygous deletion of 6.5 Mb spanning a region of chromosome 21q in proband 4. This region is associated with agenesis of the corpus callosum. (A) We define three regions across the q arm of chromosome 21. (B) Extent of deletions reported by Lyle et al. [28]. (C) Three deletions reported by Lindstrand et al. [38]. (D) We previously reported chromosome 21 partial monosomy in three patients and additional cases from Coriell Repository [39]. (E) DECIPHER database cases spanning the chromosome. (F) Proband 4 (this study) has a hemizygous deletion that is notable for spanning the entirety of region 2, possibly accounting for the severe phenotype. (G) Chromosome 21 ideogram. This figure was adapted from [39] incorporating data on proband 4.

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deletion region contains a locus implicated in agenesis of the corpus callosum (OMIM 217990), potentially accounting for that condition in this individual.

Discussion

The present study characterized the clinical and behavioral phenotypes of 14 children with autism and extreme behavioral problems including self-injury. A candidate genetic abnormality likely to be causal was identified in four of these cases, representing a prevalence of 29%. Relatively large (>100 kb) CNVs were present in all of the other 10 cases (S1 File), but their clinical significance was less clear. Diagnostic yield using genomic microarrays is typically 10–15% for patients from cohorts having intellectual disability and/or autism. However, the diagnostic yield for individuals displaying severe behavioral problems in addition to autism and intellectual disability has not been assessed. In the present study we studied trios as well as parent/child duos, including such pedigrees because often informed consent cannot be obtained from both parents of the families we enrolled. For five probands who were not part of trios we could not confirm CNVs were de novo.

Criteria for defining candidate chromosomal CNVs as potentially causal were based on conservative ISCA guidelines [8]. While these are candidates, the present findings are not able to indicate definitively that any of these variants is causal. For Probands 2 and 4, we observed large de novo CNVs (a duplication of 14 Mb and a hemizygous deletion of 6.5 Mb, respectively). Large CNVs tend to be clinically significant, yet may be benign [17]. For Proband 4, the likelihood of the CNV having a causal (clinically significant) role is greatly increased for two reasons. First, while segmental monosomy of chromosome 21 has been observed in dozens of cases, none spans this particular band (Fig 4F). Indeed, Lyle and colleagues suggested that deletions in this particular locus most likely preclude survival, with prevalence within the population approaching zero [28]. Second, the patient had agenesis of the corpus callosum, a phenotype that has been mapped to 21q22 (OMIM %217990)[29].

To gather evidence for a causal role, approaches that are likely to be useful include: (1) identifying additional patients having a similar clinical phenotype and an overlapping CNV, (2) characterizing the functional consequences of the variants in conserved syntenic regions in animal models, and (3) characterizing the biochemical and/or cellular consequences of the variant in vitro.

In this study we focused on CNVs larger than 20 kb. In addition to the 14 Mb and 6.5 Mb CNVs of Probands 2 and 4, we describe large CNVs of unknown significance including homozygous deletions (e.g. 128 kb in Proband 5), hemizygous deletions (e.g. 526 kb in Proband 12, 453 kb in Proband 11), and amplifications to copy number 4 (e.g. 177 kb in Proband 7) or copy number 3 (e.g. 1.54 Mb in Proband 13, 323 kb in Proband 8). Empirical results from studies of apparently normal individuals indicate that 1% to 2% of all CNVs are larger than 1 Mb (see [8]). In the Database of Genomic Variants (a catalog of human genome structural variation), 0.4% of CNVs are greater than 1 Mb [30]. Thus it is possible for such large CNVs to occur in the context of an apparently normal phenotype. On the other hand, large CNVs occurring in affected individuals are commonly assumed to have causal roles, particularly when they are recurrent.

The relevance of CNVs to SIB, autism, intellectual disability, and other clinical phenotypes

The occurrence of autism together with SIB was an inclusion criterion for this study. Autism was diagnosed by clinical experts. Notably, it can be impractical to administer standard diagnostic tests for autism spectrum disorders such as the ADI-R or ADOS when patients have other impairments as severe as those of the current sample. For the CNVs described as potentially
pathogenic in the present study, the relevance to SIB, autism spectrum disorders, and ID considered individually cannot be determined at this point. In the future, it may be possible to identify individuals having various subsets of these traits and then to perform genotyping to identify CNVs. Recurrent CNVs may be associated with particular clinical phenotypes. Smith-Magenis syndrome, caused by interstitial deletions on chromosome 17p11.2, provides an example of a syndrome involving self-injurious behavior, intellectual disability, and other features. Potocki-Lupski syndrome involves duplication of the same region on chromosome 17 and has overlapping clinical features. Genotype and phenotype may be correlated: Prader-Willi syndrome (OMIM #176270) is a condition associated with both a well-characterized chromosomal change (deletion of paternal copies the imprinted SNRPN gene and other genes in the 15q11-q13 chromosomal region) and a particular topography of severe self-injurious behavior (skin-picking). There is no reported heritability for SIB, although the genetics of SIB are poorly studied.

Future directions

Whole exome or whole genome sequencing in this patient population is likely to increase the yield of variants contributing to the phenotypes. Exome sequencing studies suggest a 10% contribution from de novo single nucleotide variants [31–35] and an additional 5% contribution to ASD risk from homozygous or compound heterozygous loss-of-function variants or hemizygous X-chromosome knockouts in males [36]. Gilissen et al. [37] described a cohort of 50 patients with severe intellectual disability and their parents. These patients had not received a molecular diagnosis after both SNP arrays and exome sequencing. Additional CNVs as well as loss-of-function mutations were identified by whole genome sequencing of trios, producing a diagnostic yield of 42% (and an estimated 62% cumulative diagnostic yield including microarrays and sequencing). Thus it is likely that exome or genome sequencing will greatly increase the diagnostic yield of the cohort we are studying.

The approach we have described in this study involves detailed behavioral phenotyping of patients along with initial efforts to define the genotype. In the future, adopting this approach with a larger sample will enable us to assess the prevalence of genetic anomalies likely to be causal in this population. Defining genotypes associated with these severe phenotypes may lead to improved strategies for treatment depending on the nature of the molecular defects.

Supporting Information

S1 File. Summary of case reports and chromosomal findings based on SNP analyses for probands 5–14.

S1 Table. Description of SIB topographies and percentage of participants who engaged in them.

S2 Table. Percentage of probands who targeted specific body locations with SIB, percentage of probands who exhibited other problem behavior, and percentage of SIB topographies with a given functional analysis.

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Author Contributions
Conceived and designed the experiments: WS LH JP. Performed the experiments: LF AJ AD MAF JP. Analyzed the data: MDS LF JSL MAF WS LH JP. Contributed reagents/materials/analysis tools: AJ AD. Wrote the paper: MDS LF JSL MAF WS LH JP.

References
1. Tate BG, Baroff GS (1966) Aversive control of self-injurious behavior in a psychotic boy. Behav Res Ther 4: 281–287. PMID: 5978683
2. Cooper SA, Smiley E, Allan LM, Jackson A, Finlayson J, Mantry D, et al. (2009) Adults with intellectual disabilities: prevalence, incidence and remission of self-injurious behaviour, and related factors. J Intell Disabil Res 53: 200–216. doi: 10.1111/j.1365-2788.2008.01060.x PMID: 18444987
3. McClintock K, Hall S, Oliver C (2003) Risk markers associated with challenging behaviours in people with intellectual disabilities: a meta-analytic study. J Intell Disabil Res 47: 405–416. PMID: 12919191
4. Hall S, Oliver C, Murphy G (2001) Self-injurious behaviour in young children with Lesch-Nyhan syndrome. Dev Med Child Neurol 43: 745–749. PMID: 11730148
5. Amir RE, Van den Veyver IB, Wan M, Tran CQ, Francke U, Zoghbi HY (1999) Rett syndrome is caused by mutations in X-linked MECP2, encoding methyl-CpG-binding protein 2. Nat Genet 23: 185–188. PMID: 10508514
6. Symons FJ, Clark RD, Hatton DD, Skinner M, Bailey DB Jr. (2003) Self-injurious behavior in young boys with fragile X syndrome. Am J Med Genet A 118A: 115–121. PMID: 12655491
7. Krantz ID, McCallum J, DeScipio C, Kaur M, Gillis LA, Yaeger D, et al. (2004) Cornelia de Lange syndrome is caused by mutations in NIPBL, the human homolog of Drosophila melanogaster Nipped-B. Nat Genet 36: 631. PMID: 15146186
8. Miller DT, Adam MP, Aradhya S, Biesecker LG, Brothman AR, Carter NP, et al. (2010) Consensus statement: chromosomal microarray is a first-tier clinical diagnostic test for individuals with developmental disabilities or congenital anomalies. Am J Hum Genet 86: 749–784. doi: 10.1016/j.ajhg.2010.04.006 PMID: 20466091
9. Sparrow SS, Balla DA, Cicchetti DV (1984) Vineland Adaptive Behavior Scales. Circle Pines, MN: American Guidance Service.
10. Iwata BA, Dorsey MF, Stilfer KJ, Bauman KE, Richman GS (1994) Toward a functional analysis of self-injury. Journal of Applied Behavior Analysis 27: 197–209. PMID: 8063622
11. Hagopian LP, Fisher WW, Thompson RH, Owen-DeSchryver J, Iwata BA, Wacker DP (1997) Toward the development of structured criteria for interpretation of functional analysis data. J Appl Behav Anal 30: 313–325; quiz 326. PMID: 9210309
12. Zilberman D, Chen H, Neveu E, Dryden P, Toth L, Stutzman-Darling C, et al. (2015) Comprehensive analysis of functional diversity in the TEomes. Mol Syst Biol 11: 814. doi: 10.15252/msb.201560693
13. Wang K, Li M, Hadley D, Liu R, Gissendanner J, Grant SF, et al. (2007) PennCNV: an integrated hidden Markov model designed for high-resolution copy number variation detection in whole-genome SNP genotyping data. Genome Res 17: 1665–1674. PMID: 17921354
14. MacDonald JR, Ziman R, Yuen RK, Feuk L, Scherer SW (2014) The Database of Genomic Variants: a curated collection of structural variation in the human genome. Nucleic Acids Res 42: D986–992. doi: 10.1093/nar/gkt959 PMID: 24174537
15. Brown GR, Hem V, Katz KS, Ovetsky M, Wallin C, Ermolaeva O, et al. (2015) Gene: a gene-centered information resource at NCBI. Nucleic Acids Res 43: D36–42. doi: 10.1093/nar/gku1055 PMID: 25355515
16. Xu L, Maresh GA, Giardina J, Pincus SH (2004) Comparison of different microarray data analysis programs and description of a database for microarray data management. DNA Cell Biol 23: 643–651. PMID: 15585122

17. Kearney HM, Thorland EC, Brown KK, Quintero-Rivera F, South ST, Working Group of the American College of Medical Genetics Laboratory Quality Assurance C (2011) American College of Medical Genetics standards and guidelines for interpretation and reporting of postnatal constitutional copy number variants. Genet Med 13: 680–685. doi: 10.1097/GIM.0b013e3182217a3a PMID: 21681106

18. Swaminathan GJ, Bragin E, Chatzimichalai EA, Corpas M, Bevan AP, Wright CF, et al. (2012) DECIPHER: web-based, community resource for clinical interpretation of rare variants in developmental disorders. Hum Mol Genet 21: R37–44. PMID: 22962312

19. Kahng S, Iwata BA, Lewin AB (2002) Behavioral treatment of self-injury, 1964 to 2000. Am J Ment Retard 107: 212–221. PMID: 11966334

20. Stevens EL, Heckenberg G, Roberson ED, Baugher JD, Downey TJ, Pevsner J (2011) Inference of Relationships in Population Data Using Identity-by-Descent and Identity-by-State. Plos Genetics 7: e1002287. doi:10.1371/journal.pgen.1002287 PMID: 21966277

21. Salas-Labadia C, Lieberman E, Cruz-Alcivar R, Navarrete-Meneses P, Gomez S, Cantu-Reyna C, et al. (2014) Partial and complete trisomy 14 mosaicism: clinical follow-up, cytogenetic and molecular analysis. Mol Cytogenet 7: 65. doi: 10.1186/s13039-014-0065-8 PMID: 25276227

22. Eventov-Friedman S, Frumkin A, Bar-Oz B, Raas-Rothschild A (2015) Mosaic Trisomy 14 in a Newborn with Multiple Malformations: When Chromosomal Microarray is a Clue to Diagnosis. Isr Med Assoc J 17: 459–460. PMID:26357728

23. von Sneidern E, Lacassie Y (2008) Is trisomy 14 mosaic a clinically recognizable syndrome?—case report and review. Am J Med Genet A 146A: 1609–1613. doi: 10.1002/ajmg.a.32334 PMID: 18449929

24. Fagerberg CR, Eriksen FB, Thormann J, Ostergaard JR (2012) Trisomy 14 mosaicism: clinical and cytogenetic findings in an adult. Clin Dysmorphol 21: 45–47. doi:10.1097/MCD.0b013e32834a0436 PMID: 21857505

25. Kwasnicka-Crawford DA, Roberts W, Scherer SW (2007) Characterization of an autism-associated segmental maternal heterodisomy of the chromosome 15q11-13 region. J Autism Dev Disord 37: 694–702. PMID: 17006779

26. Wachtel LE, Reti IM, Ying H (2014) Stability of intraocular pressure after retinal reattachment surgery during electroconvulsive therapy for intractable self-injury in a 12-year-old autistic boy. J ECT 30: 73–76. doi: 10.1097/YCT.0b013e31829b2d61 PMID: 23812023

27. Consortium EP (2012) An integrated encyclopedia of DNA elements in the human genome. Nature 489: 57–74. doi:10.1038/nature11247 PMID: 22955616

28. Lyle R, Bena F, Gagos S, Gehrig C, Lopez G, Schinzel A, et al. (2009) Genotype-phenotype correlations in Down syndrome identified by array CGH in 30 cases of partial trisomy and partial monosomy chromosome 21. Eur J Hum Genet 17: 454–466. doi:10.1038/ejhg.2008.214 PMID: 19002211

29. O’Driscoll MC, Black GC, Clayton-Smith J, Sherr EH, Dobyns WB (2010) Identification of genomic loci contributing to agenesis of the corpus callosum. Am J Med Genet A 152A: 2145–2159. doi:10.1002/ajmg.a.33558 PMID: 20683985

30. (2015) Database of Genomic Variants. Available:http://dgv.tcag.ca/dgv/app/home.

31. Iossifov I, Ronemus M, Levy D, Wang Z, Hakker I, Rosenbaum J, et al. (2012) De novo gene disruptions in children on the autistic spectrum. Neuron 74: 285–299. doi:10.1016/j.neuron.2012.04.009 PMID: 22542183

32. Sanders SJ, Murtha MT, Gupta AR, Murdoch JD, Raubeson MJ, Willsey AJ, et al. (2012) De novo mutations revealed by whole-exome sequencing are strongly associated with autism. Nature 485: 237–241. doi:10.1038/nature10945 PMID: 22495306

33. De Ruinis S, He X, Goldberg AP, Poultney CS, Samocha K, Cicek AE, et al. (2014) Synaptic, transcriptional and chromatin genes disrupted in autism. Nature 515: 209–215. doi: 10.1038/nature13772 PMID: 25363760

34. Lim ET, Raychaudhuri S, Sanders SJ, Stevens C, Sabo A, MacArthur DG, et al. (2013) Rare complete knockouts in humans: population distribution and significant role in autism spectrum disorders. Neuron 77: 235–242. doi: 10.1016/j.neuron.2012.12.029 PMID: 23352160
37. Gilissen C, Hehir-Kwa JY, Thung DT, van de Vorst M, van Bon BW, Willemsen MH, et al. (2014) Genome sequencing identifies major causes of severe intellectual disability. Nature 511: 344–347. doi: 10.1038/nature13394 PMID: 24896178

38. Lindstrand A, Malmgren H, Sahlen S, Schoumans J, Nordgren A, Ergander U, et al. (2010) Detailed molecular and clinical characterization of three patients with 21q deletions. Clin Genet 77: 145–154. doi: 10.1111/j.1399-0004.2009.01289.x PMID: 19863549

39. Roberson ED, Wohler ES, Hoover-Fong JE, Lisi E, Stevens EL, Thomas GH, et al. (2010) Genomic analysis of partial 21q monosomies with variable phenotypes. Eur J Hum Genet.