Improving our understanding of cloud chemistry depends on achieving better chemical characterization (90% of the organic carbon [OC] fraction remains uncharacterized) and, consequently, assessing the reactivity of this complex system. In this manuscript, we report for the first time the concentrations of 16 amino acids (AAs) in 25 cloud water samples. The concentrations of individual AAs ranged from a few nM up to ~2.0 μM, and the average contribution of AAs corresponded to 9.1% (4.4 to 21.6%) of the dissolved OC (DOC) concentration. Considering their occurrence and concentrations, AAs were expected to represent an important hydroxyl radical (HO•) sink in aqueous cloud samples. In this work, we estimated that approximately 17% (from 7 to 36%) of the hydroxyl radical-scavenging ability of the DOC could be attributed to the presence of AAs, whereas comparing the AAs suggested that an average of 51% (from 22 to 80%) of their reactivity with HO• could account for the presence of tryptophan. These results clearly demonstrate that the occurrence and reactivity of AAs must be considered to better estimate the chemical composition and oxidant capacity of the cloud aqueous phase.
The concentrations found here were relatively similar to those reported in rainwater by Gorzelska et al. Hypothesizing that enzymatic hydrolysis could be not-negligible. This hypothesis was corroborated by recent works revealing that enzymatic hydrolysis could be not-negligible. Milne and Zika proposed different hypotheses about aerosol samples, excluding thermal hydrolysis but suggesting that enzymatic hydrolysis could be not-negligible. This hypothesis was corroborated by recent works revealing the presence of living microorganisms and impact on the oxidant capacity and organic speciation in cloud water. The direct photolysis of peptides and photosensitized reaction pathways could be responsible for their fragmentation under actinic radiation. Thus, in environmental samples, both forms of AAs could be detected, but the concentration of DFAAs (uncombined forms) is expected to be lower because this is the preferred form for the uptake of nitrogen compounds by microorganisms. Recent works have shown that AAs dissolved in cloud water could be transformed by solar radiation and reaction with oxidative species (i.e., hydroxyl radicals), leading to the production of different functionalized and oxidized products and short-chain CAs.

In this manuscript, we report the AA concentrations in 25 cloud water samples collected at puy de Dôme Mountain (France) during 11 cloud events. Chromatographic separation coupled with chemical complexation and fluorescence detection was used to determine the concentration of DFAAs. To the best of our knowledge, this study reports, for the first time, the concentrations of free AAs in cloud water. Comparing the reactivities of the AAs with those of naturally occurring CAs and dissolved OC (DOC) suggested that the presence and reactivity of AAs should be considered in the future to better assess the effect of cloud aqueous-phase chemistry on the organic matter transformation mediated by the hydroxyl radical (HO·).

Results and Discussions
Quantification of AAs. The concentrations of 16 AAs were determined in 25 individual samples from 11 cloud events collected during two campaigns in March/April and November 2014 at the top of puy de Dôme mountain (1465 m a.s.l.) in France. The cloud droplet sampling was performed using a one-stage cloud droplet impactor, as previously described. The sampling times ranged from 120 to 180 min depending on the liquid water content, which reflects the mass of water in a cloud in a specified amount of dry air and was in the range of 0.1–0.2 g m⁻³. Single-AA concentrations ranged from 5 nM (corresponding to the analytical detection limit) to ~2.0 μM (Supplementary Table S1).

Figure 1 illustrates the concentrations of each AA in the cloud samples, showing that the average concentrations of ILE, PHE, SER and TRP exceeded 0.25μM (see Table 1 for the AA abbreviations and relative concentrations). The median values of the single-AA concentrations were lower than 0.5μM, and concentrations of ILE and TRP were the highest, with an average value of 0.3 and 0.4μM respectively. The total AA concentration, defined as the median value of the sum of the AA concentration in each sample, ranged from 1 to 4μM. The average concentrations of AA in cloud water samples were lower than those measured in fog water by Zhang et al. (2.6–99μM, mean ± 1σ = 20 ± 27μM) and in dew water samples in Germany by Scheller et al. (0.5–110μM, mean = 22.5μM). The concentrations found here were relatively similar to those reported in rainwater by Gorzelska et al. (≈ 0.4μM) and Mopper and Zika (6.5μM). Fog and dew water were expected to be more concentrated than cloud water because they are influenced relatively strongly by local sources and lead to less dilution of particle compared to the cloud droplets. The sampling procedure may also have contributed to this difference because of the potential evaporation of the liquid sample. The high TRP concentration cannot be directly correlated

![Figure 1. Distribution of each AA in the cloud samples.](image-url)
to the hydrolysis of proteinaceous matter. However, TRP was recently demonstrated to be strongly correlated with the formation of high-molecular-weight compounds with spectroscopic characteristics similar to those of HULIS, which can also be considered as a potential TRP reservoir. Muller et al. evaluated TRYptophan LIke Substances (TRYLIS) in rain using fluorescence emission. These authors found that the TRP concentration was higher in samples from marine origins, suggesting that the presence of TRYLIS promoted the development of ice-precipitation at ‘warmer’ temperatures. Nevertheless, the detection of this compound in the fluorescence matrix was not suitable for the discrimination of free and combined TRP.

The proportion of AAs to the DOC ranged from 4.4 to 21.6% of the total carbon (with an average value of 9.1%) in 25 samples, corresponding to an average estimated concentration of 211 ± 19 μg C L⁻¹ (see Table 1 for the average AA concentrations). Deguillaume et al. suggested that the characterization of the organic matter in cloud water remains incomplete because approximately 11% and 1% of the DOC is represented by short-chain CAs (formate, acetate, oxalate, malonate and succinate) and aldehydes, respectively. In Fig. 2 the average contributions of CAs, aldehydes and AAs to the DOC are presented, and the second pie plot shows the distribution of a single AA relative to the total distribution.

Table 1. Average single-AA concentration (nM) and corresponding average carbon concentration expressed in μg C L⁻¹. Values are presented with the standard deviation determined by analysing 25 samples.
Figure 3. Ratio of the scavenging rate constants of HO* (expressed in s⁻¹) with AAs and short-chain CAs (CAA/AC) without (filled circles) and with (empty circles) formate (calculated using equation 1). The error bars were determined by considering the uncertainties associated with AA quantification in each sample.

Reactivity with HO*: Competition among AAs, CAs and DOC. The reactivity of AAs with hydroxyl radicals can be calculated and compared with those of most relevant short-chain CAs (formate, acetate, oxalate, malonate and succinate) always measured in the cloud aqueous phase. This competition (CAA/AC) can be evaluated by considering the ratios between the scavenging rate constants of HO* by AA (k_HO*,AA) and CA (k_HO*,CA) using Equation (1):

\[ C_{AA/AC} = \frac{k_{HO*,AA}}{k_{HO*,CA}} \]  

(1)

where \( k_{HO*,AA} \) and \( k_{HO*,CA} \) are determined as the product of the second-order rate constant of HO* with single AAs and CAs (\( k_{II,AA} \) or \( k_{II,CA} \)) and their concentrations, respectively. Based on the average CA concentration in the aqueous cloud phase reported by Deguillaume et al. (Supplementary Table S2) and the AA concentrations determined in each sample, CAA/AC can be calculated by considering the contribution (or not) of formate to the CAs and a pH of 5.0 (see Supplementary Tables S3 and S4 for the pKa values and HO* reactivities of each species and the DOC concentrations in each cloud sample, respectively). As shown in Fig. 3, for the comparison without formate, the AA contribution was between 7 and 26 times larger than that of the CAs. Interestingly, for all samples, the scavenging abilities (corresponding to the pseudo-first order decay constant between considered species and hydroxyl radical) of the AAs were at least one order of magnitude higher than those of the short-chain CAs. In terms of the proportion of formate among the short-chain CAs, the CAA/AC value ranged from 0.12 to 0.42. These findings seem reasonable given the high reactivity of formate with HO* (\( k_{II,formate} = 3.2 \times 10^8 \text{M}^{-1}\text{s}^{-1} \)). Nevertheless, at lower pH values, the contribution of formate was expected to be relatively small; indeed, at pH 4.0, approximately 37% of formate exists as its acidic form (i.e., formic acid, pKa = 3.76), which is less reactive towards HO* (\( k_{II,formic acid} = 1.3 \times 10^8 \text{M}^{-1}\text{s}^{-1} \)).

Figure 4 shows the contributions of all AAs, TRP and DOC to the scavenging rate constant of the hydroxyl radical in terms of their average concentrations. Up to 51% of the total reactivity of HO*, which is expressed as \( k_{II,TRP} \), was attributable to TRP (\( k_{II,TRP} = 6.76 \pm 3.98 \times 10^5 \text{s}^{-1} \)) among all AAs (\( k_{II,AA} \sim 1.28 \pm 0.51 \times 10^4 \text{s}^{-1} \)). Interestingly, the concentration of TRP was expected to be lower than those of other AAs because of its high reactivity with HO* (\( k_{II,TRP} \)). The oxidation of TRP in cloud water led to the formation of formate and acetate in 20% yields, with other nitrogen derivatives also likely produced (up to 60% of the uncharacterized carbon). These findings demonstrated that TRP oxidation could be responsible for a large fraction of the HO* scavenged in natural samples.

Moreover, the relative contributions of AAs to the DOC’s reactivity with HO* can be estimated using Equation (2):

\[ k_{II,\text{DOC}} = k_{II,\text{DOC}[DOC]} \]  

(2)

where \( k_{II,\text{DOC}} \) and [DOC] are the second-order rate constant between HO* and DOC estimated by Arakaki et al. to be \( 3.8 \times 10^9 \text{L (mol C)}^{-1}\text{s}^{-1} \) and the concentration of DOC, which varied from 1.0 mg C L⁻¹ to 8.6 mg C L⁻¹, with an average of 2.91 mg C L⁻¹ (Supplementary Table S4). In our samples, \( k_{II,\text{DOC}} \) was in the range 0.32–2.72 \( \times 10^5 \text{s}^{-1} \). This value is similar to those previously determined by Arakaki and Faust, who reported a HO* scavenging rate range of 0.95–4.2 \( \times 10^5 \text{s}^{-1} \) in 7 cloud water samples collected from Whiteface Mountain. From the quantification of hydroxyl radicals reactivity toward AAs in each cloud sample (\( k_{II,AA} \)) determined considering the second order rate constant between AA and HO* expressed in L (mol C)⁻¹ s⁻¹ and AA...
concentration ([AA]) in mol C L⁻¹ (see Table S3) we can argue that 7% to 36% of the total hydroxyl radicals scavenged by DOC were attributable to the presence of AAs (see Supplementary Table S5 for the calculations performed for each cloud sample).

Finally, AAs constituted a large fraction of the OC in cloud waters and play an important role in the scavenging of generated hydroxyl radicals. The origins of these AAs remain unknown, and their concentrations could not be explained by considering only particle dissolution during cloud droplet formation. This observation suggests the presence of in-situ sources (i.e., macromolecule oxidation or microorganism metabolism) that should be investigated in the future. However, our findings implied that the oxidant capacity of this medium (mainly related to HO• reactivity) was strongly influenced by the presence of AAs, which can both act as sink but also modify the hydroxyl radical photogeneration processes via iron (Fe²⁺/Fe³⁺) complexation in cloud water. In fact, the complexation of ferrous and ferric ions with amino acids in water has been reported. Stability constants of Fe-amino acids complexes result in some case (i.e., for Glycine, Tryptophan, Valine) to be higher that the value reported for Fe-carboxylate complexes. We demonstrated that the presence of AAs in cloud waters must be accounted for to facilitate predicting the cloud water oxidizing capacity using multiphase chemistry models.

**Experimental Materials and methods.**

**AA analysis.** Cloud water samples were filtered immediately after collection using a 0.45-µm polytetrafluoroethylene (PTFE) filter to eliminate any microorganisms or particles. The solutions were stored at 255 K in the dark before analysis. The derivatization adopted was based on a previous method described by Ishida et al. and adapted to the cloud water analysis of primary AAs. Essentially, the -NH₂ groups of primary AAs react with o-phthalaldehyde (OPA) to form a fluorescent derivative in the presence of a thiol group (here, mercaptopropionic acid [MPA]), which acts as a reaction catalyst. Secondary AAs were previously reported to be unreactive with OPA. After reacting with OPA, the cloud water samples were analysed by high-performance liquid chromatography (HPLC; Shimadzu Nexera equipped with an autosampler/pretreatment unit [SEL-30AC]). For this purpose, 30 µL of MPA (7.7 mM in 0.1 M borate buffer), 15 µL of OPA (15.0 mM in 0.1 M borate buffer) and 5 µL of sample were mixed in a vial, and after 35 min, which is the length of time required for complete complexation, 20 µL of the solution was injected. Chemicals were purchased from Sigma-Aldrich and all solutions were prepared using MilliQ water (≥18.2 MΩ cm). The derivatized compounds were separated with an HPLC column (Shimadzu Shim-pack XR-ODS; 3.0 × 100 mm, Ø 2.2 µm porous particles) and eluted at a flow rate of 0.7 mL min⁻¹ using a gradient program with two eluents: eluent A (10 mM phosphate buffer, pH 6.8) and eluent B (acetonitrile, methanol and water 45:45:10 v/v/v). The gradient elution was as follows: initially, 10% of (B); a linear gradient to 75% (B) within 15 min; a faster increase of (B) to 100% in 1 min (corresponding to 16 min of elution); and constant (B) for an additional 5 min. A fluorescence detector (RF-20A XS) was used to detect AA-OPA derivatives at an excitation wavelength (λex)/emission wavelength (λem) 350/450 nm (Fig. 5) shows the HPLC chromatograms of samples 2 and 9).

The standard solution was prepared by diluting the AA-S-18 AA standard solution (Sigma Aldrich) in 0.1-M HCl (Supplementary Figure S1). Cloud water samples were analysed without further pre-treatment.

Each analysis was evaluated and monitored in terms of the reproducibility of the peak retention times and peak heights and the linearity of the calibration curve. The method’s limit of detection (LOD) was estimated to be on the order of 5 nM with the injection of 20 µL of sample. Blank experiments (MilliQ water subjected to the entire derivatization process) were performed and confirmed that the AA concentration in water was always lower than the LOD.

In this work, only the DFAA concentration was quantified. The samples were subjected to acidic hydrolysis according to the method described by Fountoulakis et al. (1.5 mL of sample combined with 6 M HCl in an ampoule and then heated at 100 °C for 60 min). However, both the combined and free AAs were hydrolysed,
Figure 5. Example HPLC chromatograms obtained from samples 2 and 9.

and the concentration was below the LOD. No significant differences in the AA concentration were found using filtered vs. unfiltered samples or before and after freezing.

Calibration curves (single amino acid area vs. injected concentration) were performed using a cloud water matrix in order to consider the interferences on the signal in amino acids quantification (Figure S2).

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Author Contributions
A.B. collected cloud water, contributed to the scientific discussion, provided the experimental work, main figures and chemical identification with technical assistance of G.V. for HPLC method development. L.D. supervised the cloud aqueous collection and participated to the discussion. G.M. and M.B. supervised the work, wrote the manuscript text and main figures. All authors reviewed the manuscript.

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