Rheumatoid arthritis and osteogenesis imperfecta: is there a genetic causal association?

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Dear Editor,

We read with great interest the article by Damian et al. [1], in which the authors presented for the first time the coexistence of osteogenesis imperfecta (OI) and rheumatoid arthritis (RA) [1]. More than 90% of all OI cases are caused by mutations in the alpha-1 type I collagen (COL1A1) and collagen type I alpha 2 (COL1A2) genes, leading to quantitative and/or qualitative defects in type I collagen [2, 3]. Despite joint pain, stiffness, and instability related to osteoarthritis have been reported in OI patients, especially affecting weight-bearing joints of the lower extremities [4], the concomitance of OI and symptomatic inflammatory joint disease is very uncommon. In the survey performed by McKiernan et al. [4], nearly half of 111 patients with type 1 OI reported a diagnosis of “non-inflammatory arthritis” (usually osteoarthritis). In contrast, only 4% of patients were affected by inflammatory arthritis (psoriatic and rheumatoid). Only three cases of RA associated with OI have been described so far.

A 43-year-old female was referred to our Clinic with atraumatic joint pain, stiffness, and thirteen fragility fractures occurring from infancy to the date. She was treated with alendronate, zoledronic acid (ZA), and oral cholecalciferol due to a type I OI and adalimumab monotherapy for seropositive RA. Genetic testing to confirm OI was carried out by a t-NGS analysis of a multigene panel composed of > 5000 genes associated with Mendelian diseases. The molecular analysis identified a pathogenic heterozygous missense variant in COL1A1 gene, NM_000088:c.769 G>A, p.(Gly257Arg) (rs72645321; HGMD ID: CM960320). Of note, this variant was predicted to induce a large splicing change [5] by activating a cryptic acceptor splice site, as predicted by the human splicing finder at Genomnis webtool (https://hsf.genomnis.com/home). The impaired splicing was predicted to produce an aberrant mRNA (NM_000088.3:c.751_770del) with a subsequent frameshift at protein level, p.(Gly251Asnfs*29). Of note, COL1A1 mutations that result in premature termination codons not only may reduce type I collagen synthesis, but also may impact chain structure and processing, posttranslational modifications, and extracellular matrix interactions (Fig. 1) [1].

In Damian’s case, the mutation of COL1A1 gene, c.3399del, p.(Ala1134Profs*105), occurred in the major ligand-binding region (MLBR) 3; in our case, the variant c.769 G>A, p.(Gly257Arg) is located in a protein region between the MLBR 1 and 2, within the putative binding site of interleukin-2 and α2β1-integrin [6]. MLBR regions are crucial for collagen self-assembly, and they are important

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for interactions of collagen monomers with integrins, matrix metallocproteinases, fibronectin, and cartilage oligomeric matrix protein [7], leading to receptor-mediated signaling impairment. The bond between collagens and integrins plays a key role in the pathogenesis of RA. In addition, endoplasmic reticulum stress has been described in both RA and OI, probably caused by proteins misfolding or excess that can push cells to apoptosis or autoantigen generation, consequently activating inflammation [8]. These findings can make us speculate that systemic inflammation plays a key role in the development of both OI and RA and can modify the clinical presentations of these conditions.

Declarations

Ethics approval All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Consent to participate Informed consent was obtained from the patient included in the study for the publication of any potentially identifiable images or data included in this article.

Conflicts of interest None.

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References

1. Damian LO, Zmarandache CD, Vele P, Albu A, Belizna C, Craciun A (2020) Osteogenesis imperfecta and rheumatoid arthritis: is there a link? Arch Osteoporos 5(1):40. https://doi.org/10.1007/s11657-020-0681-3
2. Warman ML, Cormier-Daire V, Hall C et al (2011) Nosology and classification of genetic skeletal disorders: 2010 revision. Am J Med Genet A 155A:943–968. https://doi.org/10.1002/ajmg.a.37365
3. Tournis S, Dede AD (2018) Osteogenesis imperfecta - a clinical update. Metabolism 80:27–37. https://doi.org/10.1016/j.metabol.2017.06.001
4. McKiernan FE (2005) Musculoskeletal manifestations of mild osteogenesis imperfecta in the adult. Osteoporos Int 16:1698–1702. https://doi.org/10.1007/s00198-005-1905-5

5. Xiong HY, Alipanahi B, Lee LJ et al (2015) RNA splicing The human splicing code reveals new insights into the genetic determinants of disease. Science 347:1254806

6. Di Lullo GA, Sweeney SM, Korkko J, Ala-Kokko L, San Antonio JD (2002) Mapping the ligand-binding sites and disease-associated mutations on the most abundant protein in the human, type I collagen. J Biol Chem 277:4223–4231. https://doi.org/10.1074/jbc.M110709200

7. Maasalu K, Nikopensius T, Koks S et al (2015) Whole-exome sequencing identifies de novo mutation in the COL1A1 gene to underlie the severe osteogenesis imperfecta. Hum Genomics 9(1):6. https://doi.org/10.1186/s40246-015-0028-0

8. Kabala PA, Angiolilli C, Yeremenko N et al (2017) Endoplasmic reticulum stress cooperates with Toll-like receptor ligation in driving activation of rheumatoid arthritis fibroblast-like synoviocytes. Arthritis Res Ther 19(1):207. https://doi.org/10.1186/s13075-017-1386-x

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