**Introduction**

Management of some advanced and metastatic cancers is a challenging area in modern oncology because of poor prognosis, drug resistance development, and few available therapeutic options beyond first line. These obstacles have prompted clinical investigation for novel anticancer compounds, such as those from natural sources, targeting the proposed model of cancer hallmarks. Tumor angiogenesis is still the major targeted and most important cancer hallmark allowing for tumor cell invasion [1–3]. It has recently gained more interest in clinical development of anticancer drugs with more than 5000 clinical trials recorded on the U.S. National Institutes of Health database (www.clinicaltrials.gov) as well as more than 94,000 articles according to the PubMed/Medline database (Fig. 1). In addition, tumor angiogenesis and lymphangiogenesis have become an important research area over the last decades for their crucial role as an essential component for the growth and spread of cancer cells [3, 4]. Angiogenesis-mediated metastatic spread through the bloodstream worsens the prognosis of most aggressive solid tumors [5–8]. A large amount of literature has shown the link between tumor angiogenesis, metastasis, and overall survival (OS) in patients with advanced cancer [9, 10]. Of note, blood vessels of tumors differ from normal resting blood vessels; this difference makes tumor blood vessels a reliable target for tumor-directed treatment. Classically, vascular-targeted therapeutic agents may act by neutralizing the angiogenic proteins, inhibiting their synthesis by cancer cells, directly inducing endothelial cell (EC) apoptosis, inhibiting endothelial receptors of angiogenic proteins [11, 12], or by disrupting tubulin network of the established tumor vasculature.
and therefore tumor blood flow interruption [13–17]. The nutritional supply of tumor tissues is under the regulation of various cytokines that (1) increases the permeability and the widening of tumor blood vessels, (2) stimulates proteolytic enzymes and cellular infiltration, (3) rebuilds blood vessels, and (4) protects ECs from apoptosis. Hypoxia increases the production of proangiogenic factors by tumor cells, macrophages, and other immune cells from the tumor microenvironment [18–20]. Targeting tumor vessels with natural compounds such as trabectedin, plitidepsin, and combretastatins, which induce tumor starvation, has gained space and interest in cancer drug development for advanced cancers [21–26]. These compounds showed significant improvement in clinical outcomes and might become an alternative therapeutic for these challenging cancers according to recent human clinical trials. This critical review analyzes the clinical relevance of using natural compounds targeting tumor vessels alone or in combination with other anticancer drugs with a focus on recent clinical trials as well as the importance of predictive biomarkers for their optimal use in the management of advanced and metastatic cancers.

Targeting the Tumor Vasculature by Natural Compounds: Perspectives from Clinical Trials

Natural compounds are a major source of small molecular weight angiogenesis inhibitors [23, 27]. Considering these perspectives, there have been extensive studies on natural compounds that showed potent and promising antiangiogenic activity as well as destabilizing microtubule dynamics (an important player in tumor vasculature) [28–30]. These compounds act through multiple cell-signaling pathways and decrease the development of resistance by cancer cells [31–34]. A number of compounds derived from natural sources were screened in order to study their potential as antiangiogenesis inhibitors (Fig. 2). Published clinical trials and real-world studies showed promising results in the advanced setting using natural compounds as a single agent or in combination with other anticancer drugs.

Trabectedin (ET-743)

Trabectedin (Yondelis®, PharmaMar®) is an alkaloid originally derived from Ecteinascidia turbinata (a Caribbean tunicate) and one of the most successful marketed natural drugs for relapsed ovarian cancer (OC) and soft-tissue sarcoma (STS) [22, 38–40]. Trabectedin has been shown to have antiangiogenic properties in several preclinical studies by activation of TIMP1 (tissue inhibitor of metalloproteinase 1) and TIMP2 (tissue inhibitor of metalloproteinase 2) as well as exerting multimodal targeting of other cancer signaling pathways [41–43]. Recently, in a neoadjuvant phase II trial for myxoid liposarcoma, trabectedin exhibited a decrease in the arborizing tumor vascularity [44]. Furthermore, trabectedin was found to generate DNA damage, strong inhibition of transcription, replication, and late apoptosis. These observations are consistent with the use of trabectedin alone or in combination with other anticancer drugs. This review discusses the mechanisms of action of trabectedin and provides insights into the perspectives of using trabectedin in advanced cancers.

Figure 1. Evolution of published PubMed-indexed articles from 2000 to 2017 and clinical trials related to angiogenesis research. Data for this figure were extracted from PubMed/Medline and ClinicalTrials.gov [accessed 12 September 2017 and analyzed by Excel (Microsoft Office 2007™)].
and cytokine production related to tumor-associated macrophages (TAMs) which are involved in cell survival, migration, and metastasis [22, 45–47]. Trabectedin has also been shown to alter the DNA repair genes including BRCA1 (breast cancer type 1 susceptibility gene) and BRCA2 (breast cancer type 2 susceptibility gene), which launches new perspectives as a chemotherapy option for patients with mutated ovarian and breast cancer (BC) [48]. More than 70 clinical trials registered in the US National Institutes of Health database are currently ongoing evaluating trabectedin in phase I/III studies especially for STS and OC (Fig. S1).

Phase I trials

As a single agent, trabectedin has been used in various schedules to determine the safety, tolerability, maximum tolerated dose, and recommended doses for phase II studies. In this direction, sequential cohorts of patients with advanced solid tumors were enrolled and treated with doses ranging from 0.46 to 0.80 mg/m² (1-h schedule) and 0.30 to 0.65 mg/m² (3-h schedule) [49]. The most observed side events in these two schedules included neutropenia, fatigue, and transaminase elevation, which translates into an acceptable safety profile. Furthermore, maximum tolerated dose for trabectedin given weekly was 0.58 mg/m² as a 3-h infusion and 0.61 mg/m² as a 1-h infusion [49]. This safety profile was confirmed in another phase I study enrolling pediatric patients with refractory and relapsed solid malignancies [50]. In this study, recommended dose of trabectedin in children with advanced solid tumors was 1.5 mg/m² given intravenously over 24 h every 3 weeks (q3 wk). Trabectedin was combined with dexamethasone and granulocyte colony-stimulating factor (G-CSF) support to prevent hepatotoxicity and neutropenia [50]. In Japanese patients with STS, trabectedin was given at a starting dose of 0.9 mg/m² as a 24-h infusion q3 wk. This dose was escalated to 1.2 mg/m² and then to 1.5 mg/m² using a “3 + 3” design [51]. Frequent grade 3 or 4 adverse events including elevation of aminotransferase and neutropenia were observed. In this cohort, the recommended dose for evaluating trabectedin in phase II studies for tissue sarcoma was 1.2 mg/m² [51].

Combination of trabectedin with other anticancer drugs, namely gemcitabine, capecitabine, carboplatin, paclitaxel, and doxorubicin, showed feasible administration, manageable toxicities (including neutropenia, thrombocytopenia, and hepatotoxicity as frequent adverse events), and preliminary antitumor activity in a wide spectrum of advanced solid tumors, which was previously reported in single-arm trials [52–57]. Recently, Sessa et al. [58] evaluated cisplatin at a fixed dose of 75 mg/m² 1-h infusion followed by a dose-escalation schedule using trabectedin 3-h infusion both administered on day 1 q3 wk in patients with advanced cancers. This combination was well tolerated with reversible side events and concordant with known toxicities of trabectedin. Based on this, trabectedin at 0.60 mg/m² was declared the recommended dose for further phase II investigations [58]. Furthermore, data from an open-label phase I trial investigating docetaxel (60 or 75 mg/m²; 1-h intravenous infusion) given q3 wk followed by escalating doses of trabectedin (0.4–1.3 mg/m² by 3-h intravenous infusion) showed a favorable toxicity profile and no evidence of drug–drug interactions [59]. In conclusion, trabectedin as single agent or in combination is well tolerated, which merits further evaluation in randomized phase II–III trials. Serious adverse events such as neutropenia and transaminase elevation can be prevented by concomitant use of prophylactic G-CSF and corticosteroids.

Phase II trials

Moving from these encouraging evidences from phase I trials, trabectedin was used in numerous phase II trials to assess its preliminary efficacy in STS, translocation-related sarcoma, OC, BC, and other advanced solid tumors.

Soft-tissue sarcoma

In STS, which represents a very heterogeneous cancer, initial nonrandomized evaluation of trabectedin demonstrated durable objective responses and relatively high survival rate in previously treated patients with recurrent or metastatic STS with disease progression despite prior conventional therapy (doxorubicin and ifosfamide) [60–62]. After these favorable data, Demetri et al. [63] randomized 270 patients with unresectable or metastatic liposarcoma or leiomyosarcoma after failure of anthracycline-based chemotherapy. This study assigned patients to one of two trabectedin regimens including 1.5 mg/m² 24-h intravenous infusion once q3 wk versus 0.58 mg/m² 3-h intravenous infusion every week for 3 weeks [63]. A documented superiority in disease control was seen with the q3 wk 24-h schedule compared to 3 h every week for 3 weeks including improved median progression-free survival (PFS) (3.3 months vs. 2.3 months; HR: 0.755; 95% CI: 0.574–0.992; P = 0.0418) and median OS (13.9 months vs. 11.8 months; HR: 0.843; 95% CI: 0.653–1.090; P = 0.1920), respectively [63]. Based on these results, trabectedin was then approved in Europe by the European Medicines Agency (EMA) and in other parts of the world and recently in the United States for treating patients with advanced sarcoma after failure of standard therapy based on ifosfamide and doxorubicin.
Combination therapy with dexamethasone was found to ameliorate the safety of trabectedin by reducing the myelosuppression and hepatotoxicity in patients with recurrent advanced STS, and it is now required in all trabectedin clinical trials [66]. In an open-label randomized and controlled phase II study of the Spanish Group for Research on Sarcoma (GEIS), Martin-BROTO et al. [67] compared trabectedin 1.1 mg/m² plus doxorubicin 60 mg/m² with doxorubicin alone at a dose of 75 mg/m². PFS was the primary endpoint; results did not show superiority of the combination. However, this exploratory translational study found that FAS (fae cell surface death receptor, also known as CD95) and p53 (phosphoprotein p53) are potent prognostic biomarkers for trabectedin evaluation which warrant more examination in the future clinical trials [67]. Similarly, according to the TRUSTS trial, first-line trabectedin did not show superiority compared to doxorubicin which is still the standard treatment of metastatic STS [68]. In addition to these safety data, these phase II trials in STS have also provided important perspectives on predictive biomarkers. Italiano et al. [69] analyzed tumor tissue makers including ERCC1 (excision repair cross-complementation group 1), ERCC5 (excision repair cross-complementation group 5; also known as XPG), and BRCA1 SNPs and their transcripts in 113 patients with advanced STS as well as their association with 6-month PFS and OS based on log-rank tests (Fig. S4). High expression of these three biomarkers was significantly associated with improved PFS and OS, which can be a potential signature for trabectedin response [69]. For the moment, trabectedin is a promising approach in the second-line setting of metastatic STS. Of note, trabectedin combined with doxorubicin showed also potent antitumor activity in advanced uterine or soft-tissue leiomyosarcoma [70] and significantly reduced the risk of death and disease progression in advanced translocation-related sarcoma [71].

Ovarian cancer

The SENDO trial inaugurated the use of trabectedin in women with OC that relapsed after platinum–taxane first-line therapy [72]. Trabectedin was administered at a dose of 1.3 mg/m² as a 3-h intravenous infusion q3 wk to 51 patients either sensitive (n = 29) or resistant (n = 30). Objective response rate was 43% and 7% in platinum-sensitive patients and platinum-resistant patients, respectively [72]. Following these preliminary results, another phase II study was conducted to evaluate the activity of single-agent trabectedin at 0.58 mg/m² as a 3-h infusion weekly every 4 weeks (q4 wk) in previously treated OC patients with platinum-based chemotherapy (66 platinum-sensitive and 81 platinum-resistant) [73]. In the sensitive cohort, in the 62 evaluable patients, the overall response rate (ORR) was 29% and the median PFS was 5.1 months. In the resistant/refractory cohort, 79 patients were evaluable and the ORR was 6.3% with a median PFS of 2.0 months [73]. In summary, this trial found trabectedin effective in the platinum-sensitive cohort of patients with OC. Later, to confirm these results in this platinum-sensitive population, Del Campo et al. [74] randomized 108 patients to receive two dosing schedules of trabectedin at a dose of 1.5 mg/m² over 24 h (n = 54; arm 1) or 1.3 mg/m² over 3 h (n = 53; arm 2). In platinum-sensitive and advanced OC, trabectedin was well tolerated and effective in both schedules—ORR was 38.9% in arm 1 and 35.8% in arm 2, and median time to progression (TTP) was 6.2 and 6.8 months for arm 1 and arm 2, respectively. In addition, most responses were seen in patients with serous histology (62.5%) and with large tumor size (<5 cm) (77.5%) [74]. Del Campo et al. attempted to compare clinical activity in a pooled analysis enrolling these three phase II trials. The investigators demonstrated that the q3 wk regimen over 3 h is superior to the weekly schedule in terms of efficacy (median PFS: 5.6 vs. 2.8 months) [75]. In this regard, a large randomized phase III trial showed interesting results for trabectedin as second-line treatment for OC (see further). Considering the amount of evidence suggesting high response rate with trabectedin in OC patients with BRCA1 phenotype (defined as the group of patients with documented repeated previous responses to platinum-based therapy), a phase II study (MITO 15 trial) was conducted in this population [76]. One hundred patients with recurrent BRCA-mutated OC were enrolled in the trial and treated with 1.3 mg/m² q3 wk over 3 h with trabectedin as single agent. As expected, platinum-sensitive patients (n = 46) had better response compared to platinum-resistant population (n = 48; ORR: 47.8% vs. 31.2%). A key problem in this study arose after finding that there was no statistically significant difference between patients considering the BRCA gene mutational status. The authors hypothesized that such unexpected results may be due to the small number of recruited patients as well as the possible presence of other genes related to the DNA repair system [76] (reviewed by Monk et al. [48]). A definitive answer to this question may be provided by the recently initiated phase III trial that will include patients with recurrent OC and BRCA-mutated and/or BRCA1 phenotype (NCT02891004, MITO-23).

Breast cancer

The first study investigating the activity of single-agent trabectedin as an alternative therapy in advanced BC included patients who have previously received palliative
therapy with taxanes or anthracyclines [77]. Trabectedin was given to 27 patients at a dose of 1.5 mg/m² over 24 h q3 wk. The ORR was 14%, three confirmed partial responses and one unconfirmed partial response were observed, and median TTP was 2.14 months [77]. In similar conditions, Goldstein et al. [78] compared trabectedin 3-h infusion at 1.3 mg/m² q3 wk or 0.58 mg/m² weekly for 3 of 4 weeks in a randomized multicenter phase II trial. Trabectedin modestly prolonged PFS and median TTP in the q3 wk arm compared with the weekly arm [78].

Data obtained from in vitro studies supported the use of trabectedin in BRCA-mutated BC which is more sensitive to DNA-damaging therapies [48, 79]. Based on these pharmacogenomic findings, 38 patients with confirmed germline BRCA-mutated metastatic BC were treated with trabectedin 1.3 mg/m² over 3 h q3 wk [80]. Median PFS was 3.9 months, and eight enrolled patients (21%) showed modifications in tumor volume. Unfortunately, the trial was closed due to low recruitment rate, and these results are immature [80]. Remarkably, a further very recent subset analysis of a phase II trial found that trabectedin had a high antitumor activity in advanced BC patients with BRCA2 mutations [81]. In addition to BRCA genes, another phase II trial analyzed Xeroderma pigmentosum G gene (XPG), which is involved in the DNA repair, to predict the efficacy of trabectedin in advanced BC [82]. In this study, 44 patients with hormone receptor-positive, HER2-negative, advanced BC were treated with trabectedin at a dose of 1.3 mg/m² over 3-h infusion q3 wk and stratified according to the XPG status (high XPG (>3) or low XPG (≤3)) [82]. This schedule was found to have limited efficacy in this population (only one complete response in low-XPG cohort), and the predictive value of the XPG expression was not noted. Therefore, the reason for the negative results when using gene signatures as prognostic and predictive tools in trabectedin efficacy is currently not completely understood, and large phase III trials are awaited. Promising data from these trials provided optimistic prospects in advanced BC with the currently available armamentarium salvage agents for this indication. Other phase II trials for other advanced tumors are listed in Table 1.

**Phase III trials**

**Ovarian cancer**

Based on several experimental studies showing synergistic effects between doxorubicin and trabectedin and early favorable safety–efficacy studies [22, 52, 88, 89], a large randomized, controlled phase III trial (OVA-301) was conducted to assess the efficacy of combining liposomal doxorubicin (PLD) given at 30 mg/m² with trabectedin infused intravenously at a dose of 1.1 mg/m² q3 wk (arm 1) versus PLD alone at 50 mg/m² q4 wk (arm 2) [90]. A total of 672 women were randomly allocated to the two arms: 329 received PLD alone and 334 received the combination. Patients were stratified according to platinum resistance, and those that discontinued treatment were excluded from the analysis. Importantly, in the arm containing trabectedin the ORR (35.3% vs. 22.6%) and median PFS (7.3 months vs. 5.8 months) were significantly improved. Furthermore, platinum-sensitive patients had better PFS with the combination arm compared to PLD alone (9.2 months vs. 7.5 months). In contrast, this doublet did not show any benefit in the platinum-resistant cohort, which is still a poor prognostic subgroup of patients with OC (see Fig. S5 for survival curves) [90]. Additionally, an exploratory analysis was performed to evaluate whether 13 selected predictive biomarkers influenced clinical benefit [91]. High expression of nibrin, which is involved in DNA double-strand break repair, was found to be significantly associated with worse survival of patients with serous OC treated with trabectedin + PLD [91]. At this time, trabectedin is not approved in the United States for treating OC in second line. Additional clinical trials including patients with platinum-sensitive advanced or relapsed OC are undergoing, evaluating the combination of trabectedin with PLD versus carboplatin/PLD (phase III INOVATYON trial, NCT01379989) and combination of trabectedin with PLD versus PLD alone (phase III ORCHYD trial, NCT01846611).

**Soft-tissue sarcoma**

In metastatic translocation-related sarcomas, Blay et al. [92] conducted a multicenter, randomized phase III trial to evaluate first-line trabectedin (arm 1; 1.5 mg/m² 24 h q3 wk) versus doxorubicin/ifosfamide (arm 2; doxorubicin 75 mg/m² q3 wk, or doxorubicin 60 mg/m² plus ifosfamide (range: 6–9 g/m³) q3 wk). Sixty-one patients were randomly assigned to arm 1 and 60 to arm 2, but only 88 were treated after pathology review confirmation. The primary endpoint was PFS. No statistically significant difference was observed in the two arms in terms of PFS and OS in this setting [92]. In advanced liposarcoma or leiomyosarcoma, Demetri et al. [93] randomized 518 previously pretreated patients to receive trabectedin at a starting dose of 1.5 mg/m² over 24 h (n = 345) versus dacarbazine at a starting dose of 1 g/m² as a 20- to 120-min infusion (n = 173). Interestingly, trabectedin reduced the death and risk of progression by 45% compared with dacarbazine (median PFS: 4.2 vs. 1.5 months). In addition, this trial demonstrated that trabectedin is superior in terms of disease control in comparison with standard dacarbazine in patients with advanced liposarcoma and leiomyosarcoma.
after failure of first-line chemotherapy, despite nonstatistically significant OS benefit (13% reduction in risk of death; median OS: 12.4 vs. 12.9 months) [93]. It should be noted that the positive results from this pivotal trial were the main reason for trabectedin approval in the United States by the US Food and Drug Administration (FDA) and in other parts of the world. Additionally, considering retrospective studies published until this time, trabectedin showed interesting efficacy in routine clinical practice [94–98]. Moreover, from a medico-economic point of view, trabectedin was found to be cost-effective as an orphan drug for patients with short life expectancy [99]. Overall, these data from trabectedin clinical trials (Table 2 and Table S1) are of potential clinical relevance as salvage therapy and promising drug in STS and OC.

### Combretastatin

Combretastatin (CBT) is an antiangiogenic agent derived from *Combretum caffrum*, a South African bush willow [101]. It belongs to the family of stilbenes, which possess antiangiogenic properties that cause a vascular shutdown in cancer tissues, leading to tumor necrosis by altering the cytoskeleton of ECs and, therefore, their proliferation and migration (Fig. S3) [35, 102]. In this context, CBT is currently undergoing several clinical trials by Mateon Therapeutics®, Inc., NCI (National Cancer Institute), and the Leukemia and Lymphoma Society for leukemia, thyroid cancer, OC, gastrointestinal cancers, and other solid tumors (Table S2). CBT showed excellent antiangiogenic activity in preclinical studies by interfering with microtubule dynamics [103–105].

Several clinical trials have shown CBT clinical safety in different tumors, both alone and in combination with chemotherapy or targeted therapies. Previously, to study safety and tolerability of CBT, Rustin et al. [106] conducted a phase Ib trial that enrolled and treated 46 advanced cancer patients with CBT (10-min infusion, at doses ranging from 36 to 60 mg/m²), 20 h before paclitaxel, carboplatin, or paclitaxel, followed by carboplatin. Grade 3 or 4 neutropenia, thrombocytopenia, grade 1–3

### Table 1. Published phase II trials of trabectedin in other advanced cancers.

| Author (year) | Regimen and enrollment | Indication | Findings |
|---------------|------------------------|------------|----------|
| Belli et al. (2016) [83] NCT01339754 | Trabectedin 1.3 mg/m² as a 3-h continuous intravenous infusion every 3 weeks (q3 wk) \((n = 25)\) | Metastatic pancreatic cancer | Median PFS and OS were 1.9 months and 5.2 months, respectively. Grade >2 neutropenia was seen in 44% of patients. Tissue analysis of 17 patients enrolled in this study identified 30 predictive genes associated with better prognosis. Single-agent trabectedin had negligible antitumor activity as salvage therapy for advanced pancreatic adenocarcinoma |
| Michaelson et al. (2012) [84] NCT00147212 | Cohort A: weekly 3-h infusion at 0.58 mg/m² for 3 of 4 weeks \((n = 33)\). Cohort B1: 24-h infusion at 1.5 mg/m² q3 wk \((n = 5)\). Cohort B2: 24-h infusion at 1.2 mg/m² q3 wk \((n = 20)\). | Metastatic castration-resistant prostate cancer | Median TTP was 1.5 months in cohort A and 1.9 months in cohort B2. At 1.2 mg/m² q3 wk and 0.58 mg/m² weekly, nausea, fatigue, neutropenia, and transaminase elevation were the most observed side events. This two schedules showed modest activity in metastatic castration-resistant prostate cancer |
| Blay et al. (2004) [85] NCT00003939 | Trabectedin was given at 1.5 mg/m² per cycle as a 24-h infusion q3 wk \((n = 28)\) | Advanced gastrointestinal stromal tumors (GIST) | Trabectedin at 1.5 mg/m² is not effective in advanced GIST. Grades 3–4 neutropenia, thrombocytopenia and transaminase elevation were the most observed side events at this dose |
| McMeekin et al. (2009) [86] NCT00050440 | Trabectedin as a 3-h infusion q3 wk at a starting dose of 1.3 mg/m² with dexamethasone pretreatment \((n = 50)\) | Persistent or recurrent endometrial cancer (PREC) | Median TTP and PFS were both 1.8 months, and median OS was 6.7 months. Nausea, asthenia, vomiting, and transaminase elevation were the most frequent side events. Single-agent trabectedin displayed minimal antitumor activity in this pretreated patients with PREC |
| Paz-Ares et al. (2007) [87] | Trabectedin over a 3-h intravenous infusion q3 wk from 1650 μg/m² to 1300 μg/m² \((n = 21)\) | Advanced colorectal cancer | This schedule is well tolerated in pretreated advanced colorectal cancer at 1.3 mg/m². Trabectedin as single agent is not effective in metastatic colorectal cancer |

PFS, progression-free survival; OS, overall survival; TTP, time to progression.
hypertension, and grade 1–3 pain were the typically observed toxicities. Response rates were seen in 22% of patients with OC, esophageal cancer, small-cell lung cancer, and melanoma. This dose-escalation trial found this regimen containing CBT well tolerated with adequate premedication [106]. Combined with radiation, CBT was found to be well tolerated in another phase Ib trial enrolling 39 patients with non-small-cell lung cancer (NSCLC), prostate cancer, and squamous cell carcinoma of the head and neck [107]. Later, to assess the safety and pharmacokinetics of intravenous CBT as a single dose, He et al. conducted an open-label, nonrandomized, and dose-escalation (5, 10, 20, 33, 50, 65, and 85 mg/m² over 30 min) phase I trial enrolling 25 patients with refractory solid tumors. Doses of ≤ 65 mg/m² given as 30-min infusions were found to be the maximum tolerated dose in this cohort of East Asian patients [108]. Moreover, adding CBT to carboplatin and paclitaxel (63 mg/m² minimum 18 h before carboplatin–paclitaxel repeated q3 wk) was found to be well tolerated in another phase Ib trial enrolling 39 patients with non-small-cell lung cancer (NSCLC), prostate cancer, and squamous cell carcinoma of the head and neck [107]. Later, to assess the safety and pharmacokinetics of intravenous CBT as a single dose, He et al. conducted an open-label, nonrandomized, and dose-escalation (5, 10, 20, 33, 50, 65, and 85 mg/m² over 30 min) phase I trial enrolling 25 patients with refractory solid tumors. Doses of ≤ 65 mg/m² given as 30-min infusions were found to be the maximum tolerated dose in this cohort of East Asian patients [108]. Moreover, adding CBT to carboplatin and paclitaxel (63 mg/m² minimum 18 h before carboplatin–paclitaxel repeated q3 wk) was found to be well tolerated in another phase Ib trial enrolling 39 patients with non-small-cell lung cancer (NSCLC), prostate cancer, and squamous cell carcinoma of the head and neck [107]. Later, to assess the safety and pharmacokinetics of intravenous CBT as a single dose, He et al. conducted an open-label, nonrandomized, and dose-escalation (5, 10, 20, 33, 50, 65, and 85 mg/m² over 30 min) phase I trial enrolling 25 patients with refractory solid tumors. Doses of ≤ 65 mg/m² given as 30-min infusions were found to be the maximum tolerated dose in this cohort of East Asian patients [108]. Moreover, adding CBT to carboplatin and paclitaxel (63 mg/m² minimum 18 h before carboplatin–paclitaxel repeated q3 wk) was found to be well tolerated in another phase Ib trial enrolling 39 patients with non-small-cell lung cancer (NSCLC), prostate cancer, and squamous cell carcinoma of the head and neck [107]. Later, to assess the safety and pharmacokinetics of intravenous CBT as a single dose, He et al. conducted an open-label, nonrandomized, and dose-escalation (5, 10, 20, 33, 50, 65, and 85 mg/m² over 30 min) phase I trial enrolling 25 patients with refractory solid tumors. Doses of ≤ 65 mg/m² given as 30-min infusions were found to be the maximum tolerated dose in this cohort of East Asian patients [108]. Moreover, adding CBT to carboplatin and paclitaxel (63 mg/m² minimum 18 h before carboplatin–paclitaxel repeated q3 wk) was found to be well tolerated in another phase Ib trial enrolling 39 patients with non-small-cell lung cancer (NSCLC), prostate cancer, and squamous cell carcinoma of the head and neck [107]. Later, to assess the safety and pharmacokinetics of intravenous CBT as a single dose, He et al. conducted an open-label, nonrandomized, and dose-escalation (5, 10, 20, 33, 50, 65, and 85 mg/m² over 30 min) phase I trial enrolling 25 patients with refractory solid tumors. Doses of ≤ 65 mg/m² given as 30-min infusions were found to be the maximum tolerated dose in this cohort of East Asian patients [108]. Moreover, adding CBT to carboplatin and paclitaxel (63 mg/m² minimum 18 h before carboplatin–paclitaxel repeated q3 wk) was found to be well tolerated in another phase Ib trial enrolling 39 patients with non-small-cell lung cancer (NSCLC), prostate cancer, and squamous cell carcinoma of the head and neck [107]. Later, to assess the safety and pharmacokinetics of intravenous CBT as a single dose, He et al. conducted an open-label, nonrandomized, and dose-escalation (5, 10, 20, 33, 50, 65, and 85 mg/m² over 30 min) phase I trial enrolling 25 patients with refractory solid tumors. Doses of ≤ 65 mg/m² given as 30-min infusions were found to be the maximum tolerated dose in this cohort of East Asian patients [108]. Moreover, adding CBT to carboplatin and paclitaxel (63 mg/m² minimum 18 h before carboplatin–paclitaxel repeated q3 wk) was found to be well tolerated in another phase Ib trial enrolling 39 patients with non-small-cell lung cancer (NSCLC), prostate cancer, and squamous cell carcinoma of the head and neck [107]. Later, to assess the safety and pharmacokinetics of intravenous CBT as a single dose, He et al. conducted an open-label, nonrandomized, and dose-escalation (5, 10, 20, 33, 50, 65, and 85 mg/m² over 30 min) phase I trial enrolling 25 patients with refractory solid tumors. Doses of ≤ 65 mg/m² given as 30-min infusions were found to be the maximum tolerated dose in this cohort of East Asian patients [108]. Moreover, adding CBT to carboplatin and paclitaxel (63 mg/m² minimum 18 h before carboplatin–paclitaxel repeated q3 wk) was found to be well tolerated in another phase Ib trial enrolling 39 patients with non-small-cell lung cancer (NSCLC), prostate cancer, and squamous cell carcinoma of the head and neck [107]. Later, to assess the safety and pharmacokinetics of intravenous CBT as a single dose, He et al. conducted an open-label, nonrandomized, and dose-escalation (5, 10, 20, 33, 50, 65, and 85 mg/m² over 30 min) phase I trial enrolling 25 patients with refractory solid tumors. Doses of ≤ 65 mg/m² given as 30-min infusions were found to be the maximum tolerated dose in this cohort of East Asian patients [108]. Moreover, adding CBT to carboplatin and paclitaxel (63 mg/m² minimum 18 h before carboplatin–paclitaxel repeated q3 wk) was found to be well tolerated in another phase Ib trial enrolling 39 patients with non-small-cell lung cancer (NSCLC), prostate cancer, and squamous cell carcinoma of the head and neck [107]. Later, to assess the safety and pharmacokinetics of intravenous CBT as a single dose, He et al. conducted an open-label, nonrandomized, and dose-escalation (5, 10, 20, 33, 50, 65, and 85 mg/m² over 30 min) phase I trial enrolling 25 patients with refractory solid tumors. Doses of ≤ 65 mg/m² given as 30-min infusions were found to be the maximum tolerated dose in this cohort of East Asian patients [108]. Moreover, adding CBT to carboplatin and paclitaxel (63 mg/m² minimum 18 h before carboplatin–paclitaxel repeated q3 wk) was found to be well tolerated in another phase Ib trial enrolling 39 patients with non-small-cell lung cancer (NSCLC), prostate cancer, and squamous cell carcinoma of the head and neck [107]. Later, to assess the safety and pharmacokinetics of intravenous CBT as a single dose, He et al. conducted an open-label, nonrandomized, and dose-escalation (5, 10, 20, 33, 50, 65, and 85 mg/m² over 30 min) phase I trial enrolling 25 patients with refractory solid tumors. Doses of ≤ 65 mg/m² given as 30-min infusions were found to be the maximum tolerated dose in this cohort of East Asian patients [108].
a therapeutic alternative. However, these encouraging results on the safety profile of CBT have pushed further investigations in phase II/III trials. Until now, there are three phase III trials conducted by Mateon Therapeutics®, Inc. [FACT, FACT2, and Focus trials—NCT00507429, NCT01701349, and NCT02646139, respectively] and still recruiting or withdrawn prior to enrollment.

**Ombrabulin (AVE8062)**

Ombrabulin is a soluble analogue of CBT and acts with a similar mechanism of action [36, 114, 115]. In preclinical studies, ombrabulin strongly inhibited tumor growth and reduced tumor blood flow by destabilizing tumor vasculature, and potentiated synergistically the action of chemotherapy drugs and radiation [116–119]. Ombrabulin was evaluated in four phase I trials and two randomized phase II/III trials by Sanofi® [NCT01263886 (DISRUPT) and NCT00699517, respectively]. The first one evaluated ombrabulin in combination with paclitaxel–cisplatin in the treatment of advanced STS, and the second evaluated ombrabulin in combination with cisplatin in patients with metastatic NSCLC. Additional trials for other advanced cancers are still ongoing (see Table S3). Ombrabulin (30-min infusion) was evaluated in a phase I dose-escalation study (from 6 to 60 mg/m²) which enrolled 105 patients with advanced solid tumors [120]. At 60 mg/m², the most common adverse events were considered manageable and included grade 3 abdominal pain (at 50 mg/m²), grade 3 tumor pain, and grade 3 hypertension. The recommended dose for phase II studies was 50 mg/m². Importantly, the levels of circulating ECs, VEGF (vascular endothelial growth factor), and MMP-9 (matrix metalloproteinase 9) were found to be significantly increased 6–10 h after intravenous infusion, making them potential biomarkers for investigating ombrabulin efficacy [120]. Similarly, a Japanese phase I trial enrolled 15 patients with advanced solid tumors and used q3 wk 30-min infusion of single-agent ombrabulin up to 50 mg/m² (15.5, 25, 35, or 50 mg/m²) [121]. In this trial, ombrabulin was well tolerated with limited cardiovascular side effects. Furthermore, in three additional phase I trials, ombrabulin has recently shown manageable and comparable overlapping toxicities to those observed in previous safety/pharmacokinetic studies [122–124]. Von Pawel et al. [125] randomized 176 patients with metastatic NSCLC in a phase II trial (DISRUPT) to receive ombrabulin (n = 88, 35 mg/m²) or placebo (n = 88) followed by taxane–platinum doublet q3 wk as first-line therapy. Adding ombrabulin to standard treatment did not show improved PFS when compared to placebo (5.65 vs 5.45 months). Moreover, the two groups had similar OS—11.0 months [125]. In a multinational, double-blind, placebo-controlled phase III trial, Blay et al. randomized 355 patients with metastatic STS previously treated with anthracycline and ifosfamide. One hundred and seventy-six patients were randomized to ombrabulin plus cisplatin and 179 patients to placebo plus cisplatin [126]. Combining ombrabulin with cisplatin significantly improved PFS (median 1.54 months vs. 1.41 months) but did not demonstrate a sufficient clinical benefit, and no improvement in OS was seen (see Fig. 3 for survival curves) [126]. Unfortunately, these disappointing results have forced the discontinuation of developing ombrabulin and its future remains uncertain. Of note, this pivotal study was the only phase III trial conducted on ombrabulin.

**Bryostatin-1**

Bryostatin-1 is a highly oxygenated macrocyclic lactone isolated from the marine invertebrate *Bugula neritina* [127]. Bryostatin-1 is a potent inhibitor of mammalian target of rapamycin (mTOR) and competitor of protein kinase C (PKC) regulatory domain, a downstream effector of mTOR that plays a key role in cancer cell metabolism, autophagy, cell cycle progression, and angiogenesis as well [128–130]. Importantly, bryostatin-1 exhibited interesting in vitro and in vivo anticancer activities alone or combined with other antiproliferative agents [127, 131]. Currently, there are 41 safety/dose-finding studies conducted by the National Cancer Institute and registered in the U.S. National Institutes of Health database for HIV, Alzheimer, and various cancers including melanoma, pancreatic cancer, kidney cancer, lymphoma, and ovarian and other gynecological cancers (Table S4). Previous phase I/II trials tested bryostatin-1 as monotherapy or associated with cytotoxic drugs on a large spectrum of both solid cancers and hematologic malignancies. Results showed that bryostatin-1 was associated with serious side events including hyponatremia, thrombocytopenia, myalgias, arthralgia, local phlebitis, and fatigue. Also, these trials failed to show clinical benefit with bryostatin-1 therapy which precludes further phase III trials [132–140]. Recently, combination of weekly bryostatin-1 (25 μg/m²) and paclitaxel (90 mg/m²) has shown consistent results with other previous studies [141]. In this phase II trial, there were no confirmed responses among 19 patients with locally advanced pancreatic cancer and the response rate did not exceed 30% [141]. In another recent phase II trial, combination of bryostatin-1 (45 μg/m², 72-h continuous infusion) and cisplatin (50 mg/m²) q3 wk was found moderately active (median PFS and OS were 3 and 8.2 months, respectively) in patients with recurrent or persistent OC [142]. Fortunately, a recent phase I study of bryostatin-1 (20 μg/m² q4 wk) and temsirolimus (an approved inhibitor mTOR for the treatment of metastatic renal cell carcinoma) showed...
optimistic results of PKC inhibition as well as mTOR signaling blockade by acting on both p70-S6 serine/threonine kinase (mTOR complex 1) and mTOR complex 2 in patients with advanced STS and metastatic renal cell carcinoma [143]. It seems that developing selective inhibitors of PKC by modifying the existing bryostatin-1 using structure–activity relationship studies may potentially enhance the biological activity of temsirolimus (for details, see reviews by Mochly-Rosen et al. [144], Martin-Liberal et al. [145], and Garg et al. [146]).

Most of these phase I-II trials were canceled because of serious toxicities (especially myalgias) and because development of bryostatin-1 is no longer available for clinical investigation. However, bryostatin-1 appears to have a promising future in HIV/AIDS antiviral “shock and kill” therapy [147].
Plinabulin (NPI-2358)

Plinabulin is an antimicrotubule agent and belongs to the halimide class, which is derived from a marine fungus (*Aspergillus ustus*) [148]. Mechanistically, it induces loss of cytoskeletal dynamics, structure, and cell–cell cohesion in immature ECs and causes a selective disruption of tumor vascular with a similar mechanism of action to that of CBT [149]. Preclinical investigations have shown that NPI-2358 causes direct cytotoxicity to rapidly proliferating ECs and/or cancer cells by inducing apoptosis through cleavage of poly(ADP-ribose) polymerase (PARP), activation of caspase-3, caspase-8, and caspase-9, and triggering phosphorylation of the stress response protein JNK (c-Jun N-terminal kinase) [150]. Only two phase I trials have published data for using plinabulin in advanced NSCLC and lymphoma [151, 152]. Mita et al. initiated the first phase I clinical trial (Nereus Pharmaceuticals®, Inc., NCT00322608) to assess the safety/pharmacokinetics and to determine the recommended dose of plinabulin for phase II trial in patients with advanced cancers. A favorable safety profile including nausea, vomiting, fever, fatigue, pain, and elevation of transient blood pressure was seen in the majority of the 38 enrolled patients. Interestingly, using DCE-MRI, this team showed a significant decrease in tumor blood flow (K Trans) at 30 mg/m^2 [151]. More details about K Trans and other biomarkers for investigating therapy response to antiangiogenic drugs can be found in two recent reviews by El Bairi et al. [153] and O’Connor et al. [154]. The 30 mg/m^2 dose was selected for further phase II trial in combination with standard chemotherapy. Recently, these data were confirmed by another phase I/II trial (Nereus Pharmaceuticals, Inc., NCT00630110) using a “3 + 3” dose-escalation design [152]. Thirteen patients with advanced NSCLC that had progressed after first-line chemotherapy were enrolled. The pharmacokinetic evaluation did not show any drug–drug interactions, and 30 mg/m^2 of plinabulin with 75 mg/m^2 of docetaxel was recommended for future studies [152]. Other phases II/III clinical studies have recently started and are in the process of recruiting patients for lymphoma and lung cancer (Table S5).

Plitidepsin (Aplidin®)

Plitidepsin is a marine compound also known as dehydrodidembin B. It is a hydrophobic cyclic depsipeptide (solubilized using Cremophor®) isolated from the Mediterranean tunicate *Aplidium albicans* [155–157]. Plitidepsin showed potent pleiotropic actions against a broad spectrum of cancer cell lines and xenograft animal models [157]. Effects of plitidepsin encompass angiogenesis, cell cycle arrest, apoptosis by caspase-3, p38/MAPK, impairment of the eukaryotic elongation factor 1A2 (eEF1A2), subsequent PARP fragmentation, tumor stroma regulation, and sustained JNK activation as well [155, 158, 159]. Importantly, JNK activation was demonstrated to be a useful biomarker for evaluating plitidepsin clinical response in vivo [160]. Preclinical studies have shown that plitidepsin inhibits angiogenesis by blocking the secretion of VEGF, MMP-2, MMP-9, and FGF-2 [161–163]. In addition, combination of plitidepsin with radiation therapy exhibited a potential bystander effect in in vitro studies [164]. Plitidepsin was introduced into clinical investigation since 1995 for patients with advanced malignancies [155] and is currently undergoing eight clinical trials (Table S6).

Plitidepsin was evaluated in eight phase I studies (four for plitidepsin alone and four in combination with other chemotherapeutics and targeted agents) for the treatment of several advanced solid tumors and hematologic malignancies. A dose-escalation Fibonacci scheme was used to determine the recommended dose for phase II trials and safety profile (Table 3). Overall, the safety profile of plitidepsin was considered manageable and included reversible myalgia with oral carnitine treatment, transaminase elevation, fatigue, vomiting, and nausea. The recommended
Table 3. Published primary endpoints for phase I trials conducted on plitidepsin.

| Author/Year | Cancer type                  | Regimen and administration                                      | Side Events                                                                 | Recommended dose for phase II studies |
|-------------|------------------------------|-----------------------------------------------------------------|----------------------------------------------------------------------------|---------------------------------------|
| Aspeslagh et al. (2017) [165] | Refractory solid tumors or lymphomas (n = 44) | Plitidepsin + sorafenib (or gemcitabine) | Arm 1: sorafenib/plitidepsin Intravenous plitidepsin at 1.8 mg/m², day 1, day 8, day 15, and, and every 4 weeks (q4 wk) (+ oral administration of sorafenib at two dose levels: day 1: 200 mg twice daily and day 2: 400 mg twice daily) Arm 2: gemcitabine/plitidepsin Intravenous infusion of plitidepsin (day 1–day 2: 1.8 mg/m²; day 3: 2.4 mg/m²; day 4: 3 mg/m²) (+ gemcitabine: day 1: 750 mg/m², day 2–day 4: 1000 mg/m²). Both agents were administered on day 1, day 8, day 15, and q4 wk | Grade 4 thrombocytopenia Palmar–plantar erythrodysesthesia and erythema Nausea, vomiting, and fatigue Grade 2 transaminase increase | 2.4 mg/m² (for combination with gemcitabine) Plitidepsin dose for combination with sorafenib was not defined because of a sponsor decision |
| Aspeslagh et al. (2016) [166] | Refractory solid tumors (n = 13) | Plitidepsin + bevacizumab | Intravenous administration of plitidepsin at three dose levels (2.8 mg/m², n = 3; 3.8 mg/m², n = 4; and 4.8 mg/m², n = 6) with bevacizumab (10 mg/kg) at days 1 and 15 of a 28-day cycle | Grade 3 fatigue | 3.8 mg/m² |
| Salazar et al. (2011) [167] | Lymphomas and advanced solid tumors (n = 20) | Plitidepsin + carboplatin | Weekly plitidepsin (1-h intravenous infusion, days 1, 8, and 15; dose escalation starting from 1.8 mg/m² + carboplatin (1-h intravenous infusion, day 1, after plitidepsin) | Grade 3 transaminase increase | 2.4 mg/m² |
| Geoerger et al. (2012) [168] | Children with refractory or relapsed solid tumors (n = 41) | Plitidepsin alone | 3-h intravenous infusion of plitidepsin every 2 weeks (one cycle), standard “3 + 3” design with dose escalation starting from 4, 5 and 6 mg/m² | Myalgia, nausea, and vomiting Elevated creatine phosphokinase and transaminase | 5 mg/m² |
| Plummer et al. (2013) [169] | Advanced melanoma (n = 28) | Plitidepsin + dacarbazine | 1-h intravenous infusion of plitidepsin on days 1, 8, and 15, followed by dacarbazine as a 1-h intravenous infusion on day 1, in cycles of 4 weeks | Grade 4 neutropenic sepsis Grade 3 (fatigue, vomiting, diarrhea, hypersensitivity, respiratory tract infection, weakness, transaminase increase) | 2.4 mg/m² |
| Faivre et al. (2005) [170] | Advanced malignancies (n = 67) | Plitidepsin alone | 24-h intravenous 0.2 mg/m² of plitidepsin every 2 weeks was selected as the starting dose; a modified Fibonacci scheme was used for dose escalation | Grade 4 panmyelopenia Grade 2–3 creatine phosphokinase elevation Grade 1–2 myalgia and muscle weakness (at 6 mg/m²) Grade 3 myotoxicity (at 5 mg/m², reversible with oral carnitine) Nausea, vomiting, diarrhea, asthenia, and transaminase increase | 5 and 7 mg/m² (+carnitine) |
| Izquierdo et al. (2008) [171] | Metastatic solid tumors or non-Hodgkin’s lymphomas (n = 48) | Plitidepsin alone | 1-h intravenous infusion of plitidepsin (starting from 0.4 mg/m²) given weekly for 3 consecutive weeks during 4-week treatment cycle | Mild-moderate myalgia, increased creatine phosphokinase levels Grade 3/4 increases in hepatic enzyme levels Fatigue, vomiting, nausea, anorexia, injection site reaction, and pain | 3.2 mg/m² |
| Maroun et al. (2006) [172] | Refractory solid tumors (n = 37) | Plitidepsin alone | 1-h intravenous infusion of plitidepsin (starting dose of 0.08 mg/m²) given daily for 5 days q3 wk | Nausea, vomiting, and diarrhea Myalgia, fatigue, and skin rash | 1.2 mg/m² |
dose ranged from 1.2 to 5 mg/m² (7 mg/m² + carnitine) [165–172]. In phase II trials (summarized in Table 4), plitidepsin showed disappointing activities on several advanced malignancies including liposarcoma, melanoma, thyroid carcinoma, lung cancer, renal cell carcinoma, and other refractory hematologic cancers. Earlier in 2008, Peschel et al. [181] conducted a multicenter phase II single-arm study that evaluated plitidepsin efficacy using 3-h continuous infusion of 5 mg/m² biweekly in 21 patients with pretreated locally advanced or metastatic NSCLC. The lack of significant antitumor activity of this single agent in NSCLC (median PFS and OS were 1.2 months and 4.3 months, respectively) was the main cause to close this trial [181]. Later, similarly, weekly single-agent plitidepsin at a dose of 3.2 mg/m² as second-lined therapy showed no antitumor activity in treating patients with relapsed small-cell lung cancer (PFS: 1.3 months; median OS: 4.8 months) [177]. The same activity was seen in

Table 4. Published phase II trials of plitidepsin.

| Author/year | Regimen and enrollment | Indication | Response rate (RR) | Progression-free survival (PFS) | Overall survival (OS) |
|-------------|------------------------|------------|-------------------|-------------------------------|---------------------|
| Toulmonde et al. (2015) [173] | Plitidepsin alone (5 mg/m² on days 1-15 and day 28) (n = 24) | Advanced dedifferentiated liposarcoma | – | 1.6 months (median) | 9.2 months (median) |
| Plummer et al. (2013) [169] | Plitidepsin (2.4 mg/m² on days 1, 8, and 15 every 4 weeks (q4 wk) + dacarbazine (800 mg/m² q4 wk) (n = 38) | Advanced melanoma | 21.4% | 3.3 months (median) | – |
| Ribrag et al. (2013) [174] | Plitidepsin (1-h intravenous infusion of 3.2 mg/m² administered weekly on days 1, 8, and 15 q4 wk (n = 67) | Relapsed/refractory non-Hodgkin’s lymphoma | 20.7% | 1.6 months (median) | 10.2 months (median) |
| Baudin et al. (2010) [175] | Plitidepsin alone (5 mg/m² as a 3-h intravenous infusion every 2 weeks (q2 wk) (n = 16) | Unresectable advanced medullary thyroid carcinoma | – | 5.3 months (median) | Not reached |
| Mateos et al. (2010) [176] | Plitidepsin (5 mg/m² as a 3-h intravenous infusion q2 wk, and 19 of them added dexamethasone 20 mg/day on days 1–4) (n = 51) | Relapsed and refractory multiple myeloma | 13% (plitidepsin) and 22% (plitidepsin + dexamethasone) | 2.3 months (plitidepsin) and 3.8 months (plitidepsin + dexamethasone) | 16.7 months (median, for plitidepsin) and not reached for the second arm |
| Eisen et al. (2009a) [177] | Plitidepsin (1-h weekly intravenous infusion of 3.2 mg/m²) (n = 20) | Relapsed small-cell lung cancer | – | 1.3 months | 4.8 months (median) |
| Eisen et al. (2009b) [178] | Plitidepsin (3 h of continuous intravenous infusion of 5 mg/m² q2 wk (n = 37) | Advanced and relapsed or progressed malignant melanoma | – | 1.3 months (median) | 3.5 months (median) |
| Dumez et al. (2009) [179] | Plitidepsin (3-h continuous intravenous infusion of 5 mg/m² q2 wk) (n = 21) | Locally advanced or metastatic transitional cell carcinoma of the urothelium | – | 1.4 months (median) | 2.3 months (median) |
| Schöffski et al. (2009) [180] | 24-h intravenous infusion q2 wk. Arm A (5 mg/m², without L-carnitine) Arm B (7 mg/m² with L-carnitine) (n = 38) | Unresectable advanced renal cell carcinoma | – | 2.1 months (both arms) | 7.0 months (arm A) and 7.6 months (arm B) |
| Peschel et al. (2008) [181] | 3-h continuous intravenous infusion of plitidepsin at a dose of 5 mg/m², q2 wk (n = 21) | Locally advanced or metastatic non-small-cell lung cancer | – | 1.2 months (median) | 4.3 months (median) |
metastatic transitional cell carcinoma of the bladder with 1.4 months and 2.3 months in terms of PFS and OS, respectively [179], in unresectable advanced medullary thyroid carcinoma [175], and recently in liposarcoma (median PFS: 1.6 months; OS: 9.2 months) [173]. As yet, a minor clinical response was seen with biweekly plitidepsin at a dose of 5 mg/m² in advanced melanoma (median PFS: 1.3 months; median OS: 3.5 months) [178]. In combination with dacarbazine (800 mg/m²) in first-line therapy for advanced melanoma, plitidepsin at 2.4 mg/m² showed some promising results (response rate: 21.4%; PFS: 3.3 months) and probably deserves more investigation in patients with BRAF-wild-type melanoma as demonstrated by another recent phase II trial [169]. When considering hematologic malignancies, plitidepsin 5 mg/m² (in a 3-h infusion biweekly) given in combination with oral dexamethasone showed enhanced and reproducible activity in 47 patients with relapsed and refractory multiple myeloma [176]. Moreover, in a multicenter phase II trial, plitidepsin showed encouraging results in noncutaneous peripheral T-cell lymphoma (an aggressive non-Hodgkin’s lymphoma) [174]. This single-arm trial used plitidepsin at a dose of 3.2 mg/m² as a 1-h infusion weekly on days 1, 8, and 15 q4 wk. The ORR reached 20.7% including four partial responses and two complete responses [174]. Because of these findings in hematologic cancers, plitidepsin (Aplidin®) has been granted an orphan drug designation by the Swiss Agency for Therapeutic Products, the EMA, and the FDA for the treatment of patients with multiple myeloma [182]. In a recent randomized phase III trial (ADMYRE, NCT01102426) that enrolled 255 heavily pretreated patients with relapsed/refractory and refractory multiple myeloma, plitidepsin (intravenous 5 mg/m²) in combination with dexamethasone (40 mg per os) was compared to dexamethasone alone (40 mg per os). Interestingly, adding plitidepsin was found to significantly reduce the risk of progression or death by 35% [183]. Additional randomized studies combining plitidepsin with other anticancer drugs as well as data on predictive biomarkers are awaited.

**Mechanisms of Drug Resistance**

Despite promising clinical benefits of tumor vessel targeting by these bioactive compounds, drug resistance is inevitable and more invasive tumor regrowth was seen in patients treated with these agents [184, 185]. This problem needs to be resolved by deep understanding of various mechanisms of resistance which are currently considered as potential targets for combinatorial approaches (Fig. 5) [184,

---

**Figure 5.** Mechanisms of drug resistance to antivascular-targeted therapy. For details, see text. Ang-2, angiopoietin-2; FGF-2, fibroblast growth factor 2; G-CSF, granulocyte colony-stimulating factor; HIF-1, hypoxia-inducible factor 1; MMPs, matrix metalloproteinases; SDF-1, stromal cell-derived factor-1; VEGF, vascular endothelial growth factor.
In addition to the already-known drug efflux mechanism and somatic mutations in drug targets, it was noted from previous clinical trials, as well as in vivo studies, that cancer cells survive in the periphery of tumors after a single dose of disrupting agent’s therapy (the viable rim concept), ensuring the regrowth of the tumor after treatment [184, 188]. This type of resistance was explained by the fact that these cells are able to receive oxygen and nutrients to supply their metabolic needs from the surrounding normal tissues. Vascular disrupting agents such as those mentioned earlier lead to the vascular shutdown in the center of the tumors. However, the peripheral vessels remain intact because of the differences observed in their architecture in these two regions. Therefore, combining vascular disrupting agents with other antiangiogenics such as monoclonal antibodies which act on the peripheral blood vessels (bevacizumab, for example) might be promising [184]. In this perspective, two clinical trials [111, 189] used fosbretabulin with bevacizumab and found deep vascular damages maintained by this combined regimen. However, these encouraging results were limited by the increased risk of developing hypertension and more associations to be further investigated. In addition, acute mobilization of bone marrow-derived progenitors of endothelial cells after treatment by vascular-targeted agents mediated by chemotactic proteins such as VEGF, G-CSF, and SDF-1 (stromal cell-derived factor-1, also known as CXCL-12) was also noted as a driving mechanism of resistance [184, 190–192]. After differentiation, these endothelial cells ensure the restoration of tumor revascularization which explains a rapid regrowth and metastases after treatments. Importantly, pharmacologic targeting of these previous chemokines enhanced the antivascular efficacy and reduced the tumor blood flow and the viable rim size [191]. Cancer cell survival and resistance are also dependent on tumor hypoxia caused by vascular shutdown [184, 193]. Consequently, tumor cells including the viable rim reprogram their metabolism into high glucose uptake and

**Box 1. Recommended articles and books of particular interest**

| Review articles and perspectives | DOI                                                                 |
|----------------------------------|----------------------------------------------------------------------|
| Hollebecque A, Massard C, Soria JC. Vascular disrupting agents: a delicate balance between efficacy and side effects. *Curr Opin Oncol*. 2012;24(3):305–315. | https://doi.org/10.1097/CCO.0b013e32835249de |
| Ribatti D. Novel angiogenesis inhibitors: addressing the issue of redundancy in the angiogenic signaling pathway. *Cancer Treat Rev*. 2011;37:344–352. | https://doi.org/10.1016/j.ctrv.2011.02.002 |
| Hoff PM, Machado KK. Role of angiogenesis in the pathogenesis of cancer. *Cancer Treat Rev*. 2012;38(7):825–833. | https://doi.org/10.1016/j.ctrv.2012.04.006 |
| Rodríguez-Antona C, Taron M. Pharmacogenomic biomarkers for personalized cancer treatment. *J Intern Med*. 2015;277(2):201–217. | https://doi.org/10.1111/joim.12321 |
| Albini A, Tosetti F, Li VW, et al. Cancer prevention by targeting angiogenesis. *Nat Rev Clin Oncol*. 2012;9(9):498–509. | https://doi.org/10.1038/nrclinonc.2012.120 |
| Atanassov AG, Waltenberger B, Pferschy-Wenzig EM, et al. Discovery and resupply of pharmacologically active plant-derived natural products: A review. *Biotechnol Adv*. 2015;33(8):1582–1614. | https://doi.org/10.1016/j.biotechadv.2015.08.001 |
| Nalejska E, Mączyńska E, Lewandowska MA. Prognostic and Predictive Biomarkers: Tools in Personalized Oncology. *Mol Diagn Ther*. 2014;18(3):273–284. | https://doi.org/10.1007/s40291-013-0077-9 |
| Pritzker KP. Predictive and prognostic cancer biomarkers revisited. *Expert Rev Mol Diagn*. 2015;15(8):971–974. | https://doi.org/10.1586/14737159.2015.1063421 |
| Mita MM, Sargsyan L, Mita AC, et al. Vascular-disrupting agents in oncology. *Expert Opin Investig Drugs*. 2013;22:317–328. | https://doi.org/10.1517/13543784.2013.759557 |
| Renfró LA, An MW, Mandrekar SJ. Precision oncology: A new era of cancer clinical trials. *Cancer Lett*. 2017;387:121–126. | https://doi.org/10.1016/j.canlet.2016.03.015 |
| Wu G, Wilson G, George J, et al. Overcoming treatment resistance in cancer: current understanding and tactics. *Cancer Lett*. 2017;28:387:69–76. | https://doi.org/10.1016/j.canlet.2016.04.018 |
| Siemann DW. The Unique Characteristics of Tumor Vasculature and Preclinical Evidence for its Selective Disruption by Tumor-Vascular Disrupting Agents. *Cancer Treat Rev*. 2011;37(1):63–74. | https://doi.org/10.1016/j.ctrv.2010.05.001 |

**Books**

| Author | Title | Publisher |
|--------|-------|-----------|
| Kim SK | Handbook of Anticancer Drugs from Marine Origin | Springer International Publishing; 2015 |
| Saeidinia S | New approaches to natural anticancer drugs | Springer International Publishing; 2015 |
| Mehta JL, Mathur P, Dhalla NS (Eds) | Biochemical Basis and Therapeutic Implications of Angiogenesis | Springer International Publishing, New York; 2017 |
Box 2. Useful links and databases

Dietmar W. Siemann (PhD) Laboratory (a pioneer in the field of antivascular disrupting therapies) http://radonc.med.ufl.edu/researchlabs/cancer-biology-laboratories/dietmar-siemann/
The Angiogenesis Foundation https://angio.org/
The Metastasis Research Society http://www.metastasis-research.org/
The French Angiogenesis Society http://angio.genes.fr/index_en.html
ANGIOGENES: A knowledge database for angiogenesis* http://angiogenes.uni-frankfurt.de/
PubAngioGen database† http://www.megabionet.org/aspd/
Centre for Microvascular Research (William Harvey Research Institute (WHRI)) http://www.whri.qmul.ac.uk/research/inflammation224-microvascular-research
Tumor angiogenesis and vascular biology laboratory at Mayo Clinic http://www.mayo.edu/research/labs/tumor-angiogenesis-vascular-biology
Laboratory of Angiogenesis and Vascular Metabolism (VIB Center for Cancer Biology (CCB)) http://www.vib.be/en/research/scientists/Pages/Peter-Carmeliet-Lab.aspx
Angiogenesis and Natural Products Laboratory at the University of Cambridge http://www.giga.ul.ac.be/cms/c_22070/en/molecular-angiogenesis-laboratory-home
Ph.D. positions in angiogenesis research https://www.findaphd.com/search/phd.aspx?keywords=angiogenesis
The International Natural Product Sciences Taskforce http://inpst.net
Useful link for Postdoc positions
Useful link for research grants and funding http://www.postgraduatefunding.com/
Open access articles from PubMed Central® related to angiogenesis methods https://www.ncbi.nlm.nih.gov/pmc/?term=Angiogenesis+methods
Jobs and Ph.D. positions related to angiogenesis research from ResearchGate® website https://www.researchgate.net/search.Search.html?type=job&query=Angiogenesis
Clinical trials on angiogenesis https://clinicaltrials.gov/ct2/results?cond=angiogenesis&term=&cntry1=&state1=&recrs

* Details can be found here: https://www.nature.com/articles/srep32475.
† Details can be found here: https://academic.oup.com/nar/article-lookup/doi/10.1093/nar/gku1139.

Box 3. Helpful websites

National Comprehensive Cancer Network® https://www.nccn.org/
U.S. National Institutes of Health clinical trials database https://clinicaltrials.gov/
European Society for Medical Oncology (ESMO) http://esmo.org/
ESMO Clinical Practice Guidelines http://www.esmo.org/Guidelines
ESMO OncologyPRO* http://oncologypro.esmo.org/
Targeted oncology® http://www.targetedonc.com/
OnClive® http://www.onclive.com/
American Society of Clinical Oncology (ASCO) https://www.asco.org/
ASCO Clinical Practice Guidelines https://www.asco.org/practice-guidelines/quality-guidelines/guidelines
Professional networking site of the ASCO http://connection.asco.org/
The Conquer Cancer Foundation® https://www.conquer.org/
American Cancer Society (ACS) https://www.cancer.org/
American Association for Cancer Research (AACR) http://www.aacr.org/Pages/Home.aspx
eCanCer† http://ecancer.org/
European CanCer Organisation (ECCO) http://www.ecco-org.eu/
European Association for Cancer Research (EACR) https://www.eacr.org/

* A scientific and educational tool offering the latest information in new cancer treatments, prevention strategies and research.
† A vast online free knowledge bank to the whole oncology community (available in both English and Spanish).
glycolysis to generate their rapid energetic needs for proliferation and invasion by acquisition of epithelial-to-mesenchymal transition phenotype, a concept known as “Warburg effect” (for reviews, see Liberti et al. [194], Muz et al. [195], and Chen et al. [196]). Under hypoxic conditions, tumors cells activate HIF-1 (hypoxia-inducible factor 1), which in turn acts on many angiogenic pathways including VEGF and other redundant pathways (predominantly FGF-2 and angiopoietin-2 (Ang-2)) [184]. Currently, various clinical trials investigating potent HIF-1 inhibitors are ongoing [197]. Recently, TAMs were also suggested as drivers of resistance to vascular targeting as well as poor prognosis [198–200]. Tumor-infiltrating macrophages are highly proangiogenic and were found to be associated with the recovery of tumor vessels after therapeutic vascular shutdown [201, 202]. Recruitment of TAMs promotes fast repair of vascular damages and necrosis via the secretion of VEGF, MMPs, and other cytokines [184]. They also increase the mobility of cancer cells and their metastatic ability which explain the observed rapid occurrence of metastases after antivascular therapy and therefore a possible target for drug discovery [184, 203]. More recently, another new mechanism was discussed as a potential targetable driver of resistance based on the pericyte coverage of tumor vessels [204, 205]. In this perspective, Chen et al. [204] used a prodrug approach to selectively target the tumor pericytes of the peripheral blood vessels via abrogation of their cytoskeleton and observed complete regression of tumors as a surrogate therapy to overcome resistance to these agents. However, along these lines and in light of this growing body of evidence, it seems that cancer cells adapt to any developed therapy in “real-time” conditions which needs more smart combined therapies.

**Conclusion and Perspectives**

Antivascular-targeted therapeutic modalities using biologically active natural compounds are currently under investigation in various randomized clinical trials. Results from these studies might expand the use of these agents, particularly in combination with other potent anticancer compounds. Because of serious toxicities and lack of efficacy, clinical investigation of bryostatin-1 and ombrabulin is discontinued. Notably, among dozens of antivascular natural compounds registered in the NIH database, marine-derived trabectedin (Yondelis®) successfully arrived into the market for some advanced sarcomas and platinum-sensitive recurrent OC. Results from clinical trials investigating plitidepsin, plinabulin, and combretastatin derivatives are still ongoing. Promisingly, new naturally developed antivascular molecules and their derivatives, especially tubulin-binding agents, have provided potential perspectives in in vitro, in vivo, and early-phase clinical studies (summarized in Table S7) and may be used in the future in developing novel anticancer drugs. In order to better understand the different effects of these compounds and the physiological angiogenesis effects on tumor development, a crucial in-depth study is needed. Much of the investigative work has so far been done in cancer chemoprevention, but there is still much more to be done in the use of new approaches, namely the use of natural compounds as starvation tactics against advanced cancers (additional information and useful data can be found in Boxes 1–3 and Boxes S1 and S2). The following points may guide the future development of these promising molecules: (1) Vascular disruption by natural compounds is an attractive strategy for cancer drug discovery; (2) in the era of evidence-based medicine, randomized and controlled trials must be the design of all future clinical trials using this strategy to improve their quality; (3) clearly, it is necessary to reduce toxicities associated with these agents as well as to identify predictive biomarkers that will allow investigators to select patients who are most likely to benefit from this treatment; and finally, (4) deep understanding of molecular mechanisms of vascular-targeted resistance and characterization of the organotypic vasculature of each cancer at the molecular and cellular levels is necessary.

**Acknowledgments**

The authors wish to thank: Prof. Heshu Rahman and Dr. Hind Kadiri (PhD student, Cifre UTT – SILSEF, Laboratoire de Nanotechnologie et d’Instrumentation Optique, Institut Charles Delaunay - UMR CNRS 6279, Université de Technologie de Troyes, France) for providing plagiarism reports before the submission. We are grateful to Teresa Amaral (MD, PhD), medical oncologist at the Interdisciplinary Skin Cancer Center (Dept. of Dermatology, University Medical Center Tuebingen, Tuebingen, Germany) and to Prof. Amar Sellam for the revision of the English of our manuscript. We are also much obliged to Dr. Ikram Dib (PhD), for the preparation of the chemical structures of the natural compounds discussed in this review. In addition, we are grateful to Servier® laboratories for providing their useful open source service (Servier Medical Art).

**Conflict of Interest**

None declared.

**References**

1. Strilic, B., and S. Offermanns. 2017. Intravascular survival and extravasation of tumor cells. Cancer Cell 32:282–293.
2. Carmeliet, P., and R. K. Jain. 2011. Molecular mechanisms and clinical applications of angiogenesis. Nature 473:298–307.

3. Hanahan, D., and R. A. Weinberg. 2011. Hallmarks of cancer: the next generation. Cell 144:646–674.

4. Ribatti, D. 2017. Epithelial-mesenchymal transition in morphogenesis, cancer progression and angiogenesis. Exp. Cell Res. 353:1–5.

5. Bielenberg, D. R., and B. R. Zetter. 2015. The unique characteristics of angiogenesis and angiogenic growth factors in NSCLC. Cancer 117:3889–3899.

6. van der Zee, J. A., C. H. van Eijck, W. C. Hop, H. van den Bergh, B. M. Dicheva, A. L. Seynhaeve, et al. 2011. Angiogenesis: a prognostic determinant in pancreatic cancer? Eur. J. Cancer 47:2576–2584.

7. Saharinen, P., L. Eklund, K. Pulkki, P. Bono, and K. Alitalo. 2011. VEGF and angiopoietin signaling in tumor angiogenesis and metastasis. Trends Mol. Med. 17:347–362.

8. Cao, Z., B. Shang, G. Zhang, L. Miele, F. H. Sarkar, Z. Wang, et al. 2013. Tumor cell-mediated neovascularization and lymphangiogenesis contrive tumor progression and cancer metastasis. Biochim. Biophys. Acta 1836:273–286.

9. Wan, L., K. Pantel, and Y. Kang. 2013. Tumor metastasis: moving new biological insights into the clinic. Nat. Med. 19:1450–1464.

10. Dudley, A. C. 2012. Tumor endothelial cells. Cold Spring Harb. Perspect Med. 2a006536.

11. Weis, S. M., and D. A. Cheresh. 2011. Tumor angiogenesis: molecular pathways and therapeutic targets. Nat. Med. 17:1359–1370.

12. Bayless, K. J., and G. A. Johnson. 2011. Role of the cytoskeleton in formation and maintenance of angiogenic sprouts. J. Vasc. Res. 48:369–385.

13. Pérez-Pérez, M. J., E. M. Priego, O. Bueno, M. S. Martins, M. D. Canela, and S. Lickens. 2016. Blocking blood flow to solid tumors by destabilizing tubulin: an approach to targeting tumor growth. J. Med. Chem. 59:8685–8711.

14. Mita, M. M., L. Sargsyan, A. C. Mita, and M. Spear. 2013. Vascular-disrupting agents in oncology. Expert Opin. Investig. Drugs 22:317–328.

15. Siemann, D. W. 2011. The unique characteristics of tumor vasculature and preclinical evidence for its selective disruption by tumor-vascular disrupting agents. Cancer Treat. Rev. 37:63–74.

16. Porcù, E., R. Bortolozzi, G. Basso, and G. Viola. 2014. Recent advances in vascular disrupting agents in cancer therapy. Future Med. Chem. 6:1485–1498.

17. De Palma, M., D. Biziato, and T. V. Petrova. 2017. Microenvironmental regulation of tumour angiogenesis. Nat. Rev. Cancer 17:457–474. https://doi.org/10.1038/nrc.2017.51

18. López-Soto, A., S. Gonzalez, M. J. Smyth, and L. Galluzzi. 2017. Control of metastasis by NK cells. Cancer Cell 32:135–154.

19. Silva, E. A., and D. J. Mooney. 2010. Effects of VEGF temporal and spatial presentation on angiogenesis. Biomaterials 31:1235–1241.

20. Mousa, S. A., T. Sudha, and P. J. Davis. 2017. Anti-angiogenesis therapy and its combination with chemotherapy: impact on primary tumor and its metastasis. Shaker A. Mousa and Paul J. Davis. Anti-Angiogenesis Strategies in Cancer Therapies. New York, United States. Elsevier. 147–163.

21. Teplinsky, E., and T. J. Herzog. 2017. The efficacy of trabectedin in treating ovarian cancer. Expert Opin. Pharmacother. 18:313–323.

22. El Bairi, K., E. L. EL-Meghawry, H. Rahman, G. Abdelkarim, and A. Najda. 2016. Natural products against cancer angiogenesis. Tumour Biol. 37:14513–14536.

23. Lin, Z., Q. Zhang, and W. Luo. 2016. Angiogenesis inhibitors as therapeutic agents in cancer: challenges and future directions. Eur. J. Pharmacol. 793:76–81.

24. Harvey, A. L., R. Edrada-Ebel, and R. J. Quinn. 2015. The re-emergence of natural products for drug discovery in the genomics era. Nat. Rev. Drug Discov. 14:111–129.

25. Rodrigues, T., D. Reker, P. Schneider, and G. Schneider. 2016. Counting on natural products for drug design. Nat. Chem. 8:531–541.

26. Diniz, C., I. Suliburska, and I. M. Ferreira. 2017. New insights into the antiangiogenic and proangiogenic properties of dietary polyphenols. Mol. Nutr. Food Res. 61:1600912. https://doi.org/10.1002/mnfr.201600912

27. Wang, Z., C. Dabrosin, X. Yin, M. M. Fuster, A. Arreola, W. K. Rathmell, et al. 2015. Broad targeting of angiogenesis for cancer prevention and therapy. Semin. Cancer Biol. 35(Suppl):S224–S243.

28. Bates, D., and A. Eastman. 2016. Microtubule destabilising agents: far more than just antimitotic anticancer drugs. Br. J. Clin. Pharmacol. 83:255–268.

29. Guan, Y. Y., X. Luan, Q. Lu, Y. R. Liu, P. Sun, M. Zhao, et al. 2016. Natural products with antiangiogenic and antivasculogenic mimicry activity. Mini Rev. Med. Chem. 16:1290–1302.

30. Hussain, S. A., A. A. Sulaiman, C. Balch, H. Chauhan, Q. M. Alhadidi, and A. K. Tiwari. 2016. Natural polyphenols in cancer chemoresistance. Nutr. Cancer 68:879–891.
42. Brodowicz, T. 2014. Trabectedin in soft tissue angiogenesis. Angiogenesis 19:287–295.

43. Jaroch, K., M. Karolak, P. Górski, A. Jaroch, A. Krajewski, A. Ilnicka, et al. 2015. Combretastatins: in vitro structure-activity relationship, mode of action and current clinical status. Pharmacol. Rep. 68:1266–1275.

44. Dossi, R., R. Frapolli, S. Di Giandomenico, L. Paracchini, F. Bozzi, S. Brich, et al. 2015. Antiangiogenic activity of trabectedin in myxoid liposarcoma: involvement of host TIMP-1 and TIMP-2 and tumor thrombospondin-1. Int. J. Cancer 136:721–729.

45. Gronchi, A., B. N. Bui, S. Bonvalot, S. Pilotti, S. Ferrari, P. Hohenberger, et al. 2012. Phase II clinical trial of neoadjuvant trabectedin in patients with advanced localized myxoid liposarcoma. Ann. Oncol. 23:771–776.

46. Germano, G., R. Frapolli, C. Belgiovine, A. Anselmo, S. Pesce, M. Liguori, et al. 2013. Role of macrophage targeting in the antitumor activity of trabectedin. Cancer Cell 23:249–262.

47. Galmarini, C. M., M. D’Incalci, and P. Allavena. 2014. Trabectedin and plitidepsin: drugs from the sea that strike the tumor microenvironment. Mar. Drugs 12:719–733.

48. Monk, B. J., D. Lorusso, A. Italiano, S. B. Kaye, M. Aracil, A. Tanović, et al. 2016. Trabectedin as a chemotherapy option for patients with BRCA deficiency. Cancer Treat. Rev. 50:175–182.

49. Forouzesh, B., M. Hidalgo, Q. Chu, A. Mita, M. Mita, G. Schwartz, et al. 2009. Phase I and pharmacokinetic study of trabectedin as a 1-or 3-hour infusion weekly in patients with advanced solid malignancies. Clin. Cancer Res. 15:3591–3599.

50. Chuk, M. K., A. Aikin, T. Whitcomb, B. C. Widemann, P. Zannikos, E. Bayever, et al. 2012. A phase I trial and pharmacokinetic study of a 24-hour infusion of trabectedin (Yondelis®, ET-743) in children and adolescents with relapsed or refractory solid tumors. Pediatr. Blood Cancer 59:865–869.

51. Ueda, T., S. Kakunaga, M. Ando, K. Yonemori, H. Sugiuira, K. Yamada, et al. 2014. Phase I and pharmacokinetic study of trabectedin, a DNA minor groove binder, administered as a 24-h continuous infusion in Japanese patients with soft tissue sarcoma. Invest. New Drugs 32:691–699.

52. von Mehren, M., R. J. Schilder, J. D. Cheng, E. Temmer, T. M. Cardoso, F. G. Renshaw, et al. 2008. A phase I study of the safety and pharmacokinetics of trabectedin in combination with pegylated liposomal doxorubicin in patients with advanced malignancies. Ann. Oncol. 19:1802–1809.

53. Messersmith, W. A., A. Ijimino, D. Ettinger, D. Laheru, J. Brahmer, D. Lansey, et al. 2008. Phase I trial of weekly trabectedin (ET-743) and gemcitabine in patients with advanced solid tumors. Cancer Chemother. Pharmacol. 63:181–188.

54. Sessa, C., A. Perotti, C. Noberasco, F. De Baud, E. Gallerani, S. Cresta, et al. 2009. Phase I clinical and pharmacokinetic study of trabectedin and doxorubicin in advanced soft tissue sarcoma and breast cancer. Eur. J. Cancer 45:1153–1161.

55. Chu, Q., A. Mita, B. Forouzesh, A. W. Tolcher, G. Schwartz, A. Nieto, et al. 2010. Phase I and pharmacokinetic study of sequential paclitaxel and trabectedin every 2 weeks in patients with advanced solid tumors. Clin. Cancer Res. 16:2656–2665.

56. Gore, L., E. Rivera, M. Basche, S. L. Moulder-Davis, G. Schwartz, A. Tanovic, et al. 2016. Phase I combination study of trabectedin and capcetabine in patients with advanced malignancies. Invest. New Drugs 30:1942–1949.
57. Vidal, L., M. Magem, C. Barlow, B. Pardo, A. Florez, A. Montes, et al. 2012. Phase I clinical and pharmacokinetic study of trabectedin and carboplatin in patients with advanced solid tumors. Invest. New Drugs 30:616–628.

58. Sessa, C., G. Del Conte, A. Christinat, S. Cresta, A. Perotti, E. Gallerani, et al. 2013. Phase I clinical and pharmacokinetic study of trabectedin and cisplatin given every three weeks in patients with advanced solid tumors. Invest. New Drugs 31:1236–1243.

59. von Mehren, M., M. Bookman, N. J. Meropol, L. M. Weiner, E. Sherman, J. Li, et al. 2015. Phase I study of the safety and pharmacokinetics of trabectedin with doxetaxel in patients with advanced malignancies. Cancer Chemoth. Pharmacol. 75:1047–1055.

60. Garcia-Carbonero, R., J. G. Supko, J. Manola, M. V. Seiden, D. Harmon, D. P. Ryan, et al. 2004. Phase II and pharmacokinetic study of ecteinascidin 743 in patients with progressive sarcomas of soft tissues refractory to chemotherapy. J. Clin. Oncol. 22:1480–1490.

61. Yovine, A., M. Riofrio, J. Y. Blay, E. Brain, J. Alexandre, C. Kahatt, et al. 2004. Phase II study of ecteinascidin-743 in advanced pretreated soft tissue sarcoma patients. J. Clin. Oncol. 22:890–899.

62. Le Cesne, A., J. Y. Blay, I. Judson, A. Van Oosterom, J. Verweij, J. Radford, et al. 2005. Phase ii study of et-743 in organisation for the research and treatment of cancer (EORTC) soft tissue and bone sarcoma group trial. J. Clin. Oncol. 23:576–584.

63. Demetri, G. D., S. P. Chawla, M. Von Mehren, P. Ritch, L. H. Baker, J. Y. Blay, et al. 2009. Efficacy and safety of trabectedin in patients with advanced or metastatic liposarcoma or leiomyosarcoma after failure of prior anthracyclines and ifosfamide: results of a randomized phase II study of two different schedules. J. Clin. Oncol. 27:4188–4196.

64. Carter, N. J., and S. J. Keam. 2010. Trabectedin: a review of its use in soft tissue sarcoma and ovarian cancer. Drugs 70:355–376.

65. Administration USFaD, FDA news release: FDA approves new therapy for certain types of advanced soft tissue sarcoma. Available at http://www.fda.gov/NewsEvents/PressAnnouncements/ucm468832.htm; (accessed 15 July 2017).

66. Paz-Ares, L., A. López-Pousa, A. Poveda, C. Balañá, E. Ciruelos, J. Bellmunt, et al. 2012. Trabectedin in pre-treated patients with advanced or metastatic soft tissue sarcoma: a phase II study evaluating co-treatment with dexamethasone. Invest. New Drugs 30:729–740.

67. Martin-Broto, J., A. L. Pousa, R. de las Peñas, X. García del Muro, A. Gutierrez, J. Martinez-Trufero, et al. 2016. Randomized phase II study of trabectedin and doxorubicin compared with doxorubicin alone as first-line treatment in patients with advanced soft tissue sarcomas: a Spanish Group for Research on Sarcoma study. J. Clin. Oncol. 34:2294–2302.

68. Bui-Nguyen, B., J. E. Butrynski, N. Penel, J. Y. Blay, N. Isambert, M. Milhem, et al. 2015. A phase Ib multicentre study comparing the efficacy of trabectedin to doxorubicin in patients with advanced or metastatic untreated soft tissue sarcoma: the TRUSTS trial. Eur. J. Cancer 51:1312–1320.

69. Italiano, A., A. Laurand, A. Laroche, P. Casali, R. Sanfilippo, A. Le Cesne, et al. 2011. ERCC5/XPG, ERCC1, and BRCA1 gene status and clinical benefit of trabectedin in patients with soft tissue sarcoma. Cancer 117:3445–3456.

70. Pautier, P., A. Floquet, C. Chevreau, N. Penel, C. Guillemet, C. Delcambre, et al. 2015. Trabectedin in combination with doxorubicin for first-line treatment of advanced uterine or soft-tissue leiomyosarcoma (LMS-02): a non-randomised, multicentre, phase 2 trial. Lancet Oncol. 16:457–464.

71. Kawai, A., N. Araki, H. Sugiu, R. T. Ueda, T. Yonemoto, M. Takahashi, et al. 2015. Trabectedin monotherapy after standard chemotherapy versus best supportive care in patients with advanced, translocation-related sarcoma: a randomised, open-label, phase 2 study. Lancet Oncol. 16:406–416.

72. Sessa, C., F. De Braud, A. Perotti, J. Bauer, G. Curiglino, C. Nofrasci, et al. 2005. Trabectedin for women with ovarian carcinoma after treatment with platinum and taxanes fails. J. Clin. Oncol. 23:1867–1874.

73. Krasner, C. N., D. S. McMeekin, S. Chan, P. S. Braly, F. G. Renshaw, S. Kaye, et al. 2007. A Phase II study of trabectedin single agent in patients with recurrent ovarian cancer previously treated with platinum-based regimens. Br. J. Cancer 97:1618–1624.

74. Del Campo, J. M., A. Roszak, M. Bidzinski, T. E. Ciuleanu, T. Hogberg, M. Z. Wojtukiewicz, et al. 2009. Phase II randomized study of trabectedin given as two different every 3 weeks dose schedules (1.5 mg/m2 24 h or 1.3 mg/m2 3 h) to patients with relapsed, platinum-sensitive, advanced ovarian cancer. Ann. Oncol. 20:1794–1802.

75. Del Campo, J. M., C. Sessa, C. N. Krasner, J. B. Vermorken, N. Colombo, S. Kaye, et al. 2013. Trabectedin as single agent in relapsed advanced ovarian cancer: results from a retrospective pooled analysis of three phase II trials. Med. Oncol. 30:435.

76. Lorusso, D., G. Scambia, S. Pignata, R. Sorio, G. Amadio, S. Lepori, et al. 2016. Prospective phase II trial of trabectedin in BRCA1-mutated and/or BRCA1ness phenotype recurrent ovarian cancer patients: the MITO 15 trial. Ann. Oncol. 27:487–493.
77. Zelek, L., A. Yovine, E. Brain, F. Turpin, A. Taamma, M. Riforio, et al. 2006. A phase II study of Yondelis (trabectedin, ET-743) as a 24-h continuous intravenous infusion in pretreated advanced breast cancer. Br. J. Cancer 94:1610–1614.

78. Goldstein, L. J., J. Gurtler, S. A. Del Prete, S. Tjulandin, V. F. Semiglazov, E. Bayever, et al. 2014. Trabectedin as a single-agent treatment of advanced breast cancer after anthracycline and taxane treatment: a multicenter, randomized, phase II study comparing 2 administration regimens. Clin. Breast Cancer 14:396–404.

79. Soares, D. G., A. E. Escargueil, V. Poindessous, A. Sarasin, A. de Gramont, D. Bonatto, et al. 2007. Replication and homologous recombination repair regulate DNA double-strand break formation by the antitumor alkylator eteincasidc 743. Proc. Natl. Acad. Sci. USA 104:13062–13067.

80. Delalogue, S., R. Wolp-Diniz, T. Byrski, J. L. Blum, A. Goncalves, M. Campone, et al. 2014. Activity of trabectedin in germline BRCA1/2-mutated metastatic breast cancer: results of an international first-in-class phase II study. Ann. Oncol. 25:1152e8.

81. Ghoutani, A., S. Delaloge, P. Lardelli, C. Kahatt, T. Byrski, J. L. Blum, et al. 2017. Higher antitumor activity of trabectedin in germline BRCA2 carriers with advanced breast cancer as compared to BRCA1 carriers: a subset analysis of a dedicated phase II trial. Breast 34:18–23.

82. Awada, A., J. Cortés, M. Martín, P. Aftimos, M. Oliveira, S. López-Tarruella, et al. 2016. Phase 2 study of trabectedin in patients with hormone receptor–positive, HER-2–negative, advanced breast carcinoma according to expression of xeroderma pigmentosum G gene. Clin. Breast Cancer 16:364–371.

83. Belli, C., L. Piemonti, M. D’Incalci, M. Zucchetti, L. Porcu, S. Cappio, et al. 2016. Phase II trial of salvage therapy with trabectedin in metastatic pancreatic adenocarcinoma. Cancer Chemoer. Pharmacol. 77:477–484.

84. Michaeelson, M. D., J. Bellmunt, G. R. Hudes, S. Goel, R. J. Lee, P. W. Kantoff, et al. 2012. Multicenter phase II study of trabectedin in patients with metastatic castration-resistant prostate cancer. Ann. Oncol. 23:1234–1240.

85. Blay, J. Y., A. Le Cesne, J. Verweij, M. Scru, C. Szyneae, S. Bonvalot, et al. 2004. A phase II study of ET-743/trabectedin (Yondelis®) for patients with advanced gastrointestinal stromal tumours. Eur. J. Cancer 40:1327–1331.

86. McMeekin, D. S., A. Lisynskaya, M. Crispens, A. M. Oza, P. Braly, D. Doering, et al. 2009. Single-agent trabectedin as second-line therapy of persistent or recurrent endometrial cancer: results of a multicenter phase II study. Gynecol. Oncol. 114:288–292.

87. Paz-Ares, L., F. Rivera-Herreros, E. Díaz-Rubio, M. García, E. Casado, R. Cubedo, et al. 2007. Phase II study of trabectedin in pretreated patients with advanced colorectal cancer. Clin. Colorectal Cancer 6:522–528.

88. Takahashi, N., W. W. Li, D. Banerjee, K. W. Scotto, and J. R. Bertino. 2001. Sequence-dependent enhancement of cytotoxicity produced by Ecteinascidin 743 (ET-743) with doxorubicin or paclitaxel in soft tissue sarcoma cells. Clin. Cancer Res. 7:3251–3257.

89. Meco, D., T. Colombo, P. Ubezio, M. Zucchetti, M. Zaffaroni, A. Riccardi, et al. 2003. Effective combination of ET-743 and doxorubicin in sarcoma: preclinical studies. Cancer Chemother. Pharmacol. 52:131–138.

90. Monk, B. J., T. J. Herzog, S. B. Kaye, C. N. Krasner, J. B. Vermorken, F. M. Muggia, et al. 2010. Trabectedin plus pegylated liposomal Doxorubicin in recurrent ovarian cancer. J. Clin. Oncol. 28:3107–3114.

91. Monk, B. J., S. B. Kaye, A. Poveda, T. J. Herzog, M. Aracil, A. Nieto, et al. 2014. Nibrin is a marker of clinical outcome in patients with advanced serous ovarian cancer treated in the phase III OVA-301 trial. Gynecol. Oncol. 132:176–180.

92. Blay, J. Y., M. G. Leahy, B. B. Nguyen, S. R. Patel, P. Hohenberger, A. Santoro, et al. 2014. Randomised phase III trial of trabectedin versus doxorubicin-based chemotherapy as first-line therapy in translocation-related sarcomas. Eur. J. Cancer 50:1137–1147.

93. Demetri, G. D., M. von Mehren, R. L. Jones, M. L. Hensley, S. M. Schuetze, A. Staddon, et al. 2016. Efficacy and safety of trabectedin or dacarbazine for metastatic liposarcoma or leiomyosarcoma after failure of conventional chemotherapy: results of a phase III randomized multicenter clinical trial. J. Clin. Oncol. 34:786–793.

94. Fayette, J., H. Boyle, S. Chabaud, B. Favier, C. Engel, P. Cassier, et al. 2010. Efficacy of trabectedin for advanced sarcomas in clinical trials versus compassionate use programs: analysis of 92 patients treated in a single institution. Anticancer Drugs 21:113–119.

95. Gounaris, I., H. M. Hatcher, D. Davidson, K. Sherbourne, S. Alam, K. A. Zaki, et al. 2014. Trabectedin for advanced soft tissue sarcomas: a single institution experience. Future Oncol. 10:1843–1851.

96. Le Cesne, A., I. Ray-Coquard, F. Duffaud, C. Chevreau, N. Penel, B. Bui Nguyen, et al. 2015. Trabectedin in patients with advanced soft tissue sarcoma: a retrospective national analysis of the French Sarcoma Group. Eur. J. Cancer 51:742–750.

97. Daupin, J., P. Paubel, J. Fillon, I. Debrix, D. G. Soares, and J. P. Lotz. 2016. Trabectedin for sarcomas...
in daily clinical practice: analysis of 45 patients treated in a French institution. J. Chemother. 28:494–499.

98. Adam, J. P., F. Boumedién, N. Letarte, and D. Provencher. 2017. Single agent trabectedin in heavily pretreated patients with recurrent ovarian cancer. Gynecol. Oncol. 147:47–53. https://doi.org/10.1016/j.gyono.2017.07.123

99. Daupin, J., P. Paubel, D. Grazziotin-Soares, and J. P. Lotz. 2017. A medico-economic study of trabectedin compared with end-stage treatment in soft tissue sarcomas. J. Cancer Policy 12:55–60.

100. Monk, B. J., J. A. Blessing, D. G. Street, C. Y. Muller, J. J. Burke, and M. L. Hensley. 2012. A pharmacokinetic and safety study of single dose intravenous combretastatin A4 phosphate in patients with refractory solid tumours. Br. J. Cancer 106:231–237.

101. Ng, Q. S., H. Mandeville, V. Goh, R. Alonzi, J. Milner, D. Carnell, et al. 2012. Phase Ib trial of radiotherapy in combination with combretastatin-A4-phosphate in patients with non-small-cell lung cancer, prostate adenocarcinoma, and squamous cell carcinoma of the head and neck. Ann. Oncol. 23:231–237.

102. He, X., S. Li, H. Huang, Z. Li, L. Chen, S. Ye, et al. 2011. A pharmacokinetic and safety study of single dose intravenous combretastatin A4 phosphate in Chinese patients with refractory solid tumours. Br. J. Clin. Pharmacol. 71:860–870.

103. Hori, K. 2012. Starvation tactics for solid tumors: tumor blood flow interruption via a combretastatin derivative (Cderiv), and its microcirculation mechanism. Cancer Metastasis Rev. 31:109–122.

104. Patil, P. O., A. G. Patil, R. A. Rane, P. C. Patil, P. K. Deshmukh, S. B. Bari, et al. 2015. Recent advancement in discovery and development of natural product combretastatin-inspired anticancer agents. Anticancer Agents Med. Chem. 15:955–969.

105. Greene, L. M., M. J. Meegan, and D. M. Zisterer. 2015. Combretastatins: more than just vascular targeting agents? J. Pharmacol. Exp. Ther. 355:212–227.

106. Rustin, G. J., G. Shreeves, P. D. Nathan, A. Gaya, T. S. Ganesan, D. Wang, et al. 2010. A Phase Ib trial of CA4P (combretastatin A-4 phosphate), carboplatin, and paclitaxel in patients with advanced cancer. Br. J. Cancer 102:1355–1360.

107. Zweifel, M., G. C. Jayson, N. S. Reed, R. Osborne, B. Hassan, J. Ledermann, et al. 2011. Phase II trial of combretastatin A4 phosphate, carboplatin, and paclitaxel in patients with platinum resistant ovarian cancer. Ann. Oncol. 22:2036–2041.

108. Patterson, D. M., M. Zweifel, M. R. Middleton, P. M. Price, L. K. Folkes, M. R. Stratford, et al. 2012. Phase I clinical and pharmacokinetic evaluation of the vascular-disrupting agent OXi4503 in patients with advanced solid tumors. Clin. Cancer Res. 18:1415–1425.

109. Nathan, P., M. Zweifel, A. R. Padhani, D. M. Koh, M. Ng, D. J. Collins, et al. 2012. Phase I trial of combretastatin A4 phosphate (CA4P) in combination with bevacizumab in patients with advanced cancer. Clin. Cancer Res. 18:3428–3439.

110. Liu, P., Y. Qin, L. Wu, S. Yang, N. Li, H. Wang, et al. 2014. A phase I clinical trial assessing the safety and tolerability of combretastatin A4 phosphate injections. Anticancer Drugs 25:462–471.

111. Garon, E. B., J. D. Neidhart, N. Y. Gabrail, M. R. de Oliveira, J. Balkisson, and F. Kabbninar. 2016. A randomized Phase II trial of the tumor vascular disrupting agent CA4P (fosbretabulin tromethamine) with carboplatin, paclitaxel, and bevacizumab in advanced nonsquamous non-small-cell lung cancer. Onco. Targets Ther. 9:7275–7283.

112. Liu, P., Y. Qin, L. Wu, S. Yang, N. Li, H. Wang, et al. 2014. A phase I clinical trial assessing the safety and tolerability of combretastatin A4 phosphate injections. Anticancer Drugs 25:462–471.

113. Demers, B., P. Vrignaud, and M. Bissery. 2006. In vivo synergy combining oxaliplatin with AVE8062, a vascular disrupting agent. J. Clin. Oncol. 24(Suppl. 18):13074.

114. Dark, G. G., S. A. Hill, V. E. Prise, et al. 1997. Combratstatin A-4, an agent that displays potent and selective toxicity toward tumor vasculature. Cancer Res. 57:1829–1834.

115. Quatrale, A. E., L. Porcelli, A. Gnoni, G. Numico, A. Paradiso, and A. Azzariti. 2014. New vascular disrupting agents in upper gastrointestinal malignancies. Curr. Med. Chem. 21:1039–1049.

116. Price, L. K. Folkes, M. R. Stratford, et al. 2012. Phase I clinical and pharmacokinetic evaluation of the vascular-disrupting agent OXi4503 in patients with advanced solid tumors. Clin. Cancer Res. 18:3428–3439.

117. Delmonte, A., and C. Sessa. 2013. Phase I safety,
pharmacokinetic and pharmacodynamic evaluation of the vascular disrupting agent ombrabulin (AVE8062) in patients with advanced solid tumors. Clin. Cancer Res. 19:4832–4842.

121. Murakami, H., T. Kurata, Y. Onozawa, J. Watanabe, A. Ono, T. Takahashi, et al. 2014. An open-label, dose-escalation, safety, and pharmacokinetics phase I study of ombrabulin, a vascular disrupting agent, administered as a 30-min intravenous infusion every 3 weeks in Japanese patients with advanced solid tumors. Cancer Chemother. Pharmacol. 73:623–630.

122. Takahashi, S., K. Nakano, T. Yokota, K. Shitara, K. Muro, Y. Sunaga, et al. 2016. Phase 1 study of ombrabulin in combination with cisplatin (CDDP) in Japanese patients with advanced solid tumors. Jpn. J. Clin. Oncol. 46:1000–1007.

123. Bahleda, R., C. Sessa, G. Del Conte, et al. 2014. Phase I clinical and pharmacokinetic study of ombrabulin (AVE8062) combined with cisplatin/docetaxel or carboplatin/paclitaxel in patients with advanced solid tumors. Invest. New Drugs 32:1188–1196.

124. Eskens, F. A., P. Tresca, D. Tosi, L. Van Doorn, H. Fontaine, A. Van der Gaast, et al. 2014. A phase I pharmacokinetic study of the vascular disrupting agent ombrabulin (AVE8062) and docetaxel in advanced solid tumours. Br. J. Cancer 110:2170–2177.

125. von Pawel, J., V. Gorbounova, M. Reck, D. M. Kowalski, A. Allard, M. Chadjaa, et al. 2014. DISRUPT: a randomised phase 2 trial of ombrabulin (AVE8062) plus a taxane–platinum regimen as first-line therapy for metastatic non-small cell lung cancer. Lung Cancer 85:224–229.

126. Blay, J. Y., Z. Pápai, A. W. Tolcher, A. Italiano, D. Cupissol, A. López-Pousa, et al. 2015. Ombrabulin plus cisplatin versus placebo plus cisplatin in patients with advanced soft-tissue sarcomas after failure of anthracycline and ifosfamide chemotherapy: a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet Oncol. 2015:531–540.

127. Kollár, P., J. Rajchard, Z. Balounová, and J. Pazourek. 2014. Marine natural products: Bryostatins in preclinical and clinical studies. Pharm. Biol. 52:237–242.

128. Pópulo, H., J. M. Lopes, and P. Soares. 2012. The mTOR signalling pathway in human cancer. Int. J. Mol. Sci. 13:1866–1918.

129. Huang, K., and D. C. Fingar. 2014. Growing knowledge of the mTOR signaling network. Semin. Cell Dev. Biol. 36:79–90.

130. Kim, Y. C., and K. L. Guan. 2015. mTOR: a pharmacologic target for autophagy regulation. J. Clin. Invest. 125:25–32.

131. Wang, Y., J. Zhang, Q. Wang, T. Zhang, Y. Yang, Y. Yi, et al. 2013. Bryostatin 5 induces apoptosis in acute monocytic leukemia cells by activating PUMA and caspases. Eur. J. Pharmacol. 718:340–349.

132. Brockstein, B., B. Samuels, R. Humeric-khouse, et al. 2001. Phase II studies of bryostatin-1 in patients with advanced sarcoma and advanced head and neck cancer. Invest. New Drugs 19:249–254.

133. Pfister, D. G., J. McCaffrey, A. J. Zahalasky, G. K. Schwartz, E. Lis, W. Gerald, et al. 2002. A phase II trial of bryostatin-1 in patients with metastatic or recurrent squamous cell carcinoma of the head and neck. Invest. New Drugs 20:123–127.

134. Ku, G. Y., D. H. Ilson, L. H. Schwartz, M. Capanu, E. O’Reilly, M. A. Shah, et al. 2008. Phase II trial of sequential paclitaxel and 1 h infusion of bryostatin-1 in patients with advanced esophageal cancer. Cancer Chemother. Pharmacol. 62:875–880.

135. Ajani, J. A., Y. Jiang, J. Faust, B. B. Chang, L. Ho, J. C. Yao, et al. 2006. A multi-center phase II study of sequential paclitaxel and bryostatin-1 (NSC 339553) in patients with untreated, advanced gastric or gastroesophageal junction adenocarcinoma. Invest. New Drugs 24:353–357.

136. Roberts, J. D., M. R. Smith, E. J. Feldman, L. Cragg, M. M. Millenson, G. J. Roboz, et al. 2006. Phase I study of bryostatin 1 and fludarabine in patients with chronic lymphocytic leukemia and indolent (non-Hodgkin’s) lymphoma. Clin. Cancer Res. 12:5809–5816.

137. Winegarden, J. D., A. M. Mauer, T. F. Gajewski, P. C. Hoffman, S. Krauss, C. M. Rudin, et al. 2003. A phase II study of bryostatin-1 and paclitaxel in patients with advanced non-small cell lung cancer. Lung Cancer 39:191–196.

138. Blackhall, F. H., M. Ranson, J. A. Radford, et al. 2001. A phase II trial of bryostatin 1 in patients with non-Hodgkin’s lymphoma. Br. J. Cancer 84:465–469.

139. Madhusudan, S., A. Protheroe, D. Propper, C. Han, P. Corrie, H. Earl, et al. 2003. A multicentre phase II trial of bryostatin-1 in patients with advanced renal cancer. Br. J. Cancer 89:1418–1422.

140. Smith, B. D., R. J. Jones, E. Cho, J. Kowalski, J. E. Hoffman, S. Krauss, C. M. Rudin, et al. 2003. A phase II study of bryostatin-1 and paclitaxel in patients with advanced non-small cell lung cancer. Lung Cancer 39:191–196.

141. Lam, A. P., J. A. Sparano, V. Vinciguerra, A. J. Ocean, P. Christos, et al. 2010. Phase II study of paclitaxel plus the protein kinase C inhibitor bryostatin-1 in advanced pancreatic carcinoma. Am. J. Clin. Oncol. 33:121–124.

142. Morgan, R. J. Jr, L. Leong, W. Chow, D. Gandara, P. Frankel, A. Garcia, et al. 2012. Phase II trial of bryostatin-1 in combination with cisplatin in patients with recurrent or persistent epithelial ovarian cancer: a
California cancer consortium study. Invest. New Drugs 30:723–728.

143. Plimack, E. R., T. Tan, Y. N. Wong, M. M. von Mehren, L. Malizzia, et al. 2014. A phase I study of temsirolimus and bryoastatin-1 in patients with metastatic renal cell carcinoma and soft tissue sarcoma. Oncologist 19:354–355.

144. Mochly-Rosen, D., K. Das, and K. V. Grimes. 2012. Protein kinase C, an elusive therapeutic target? Nat. Rev. Drug Discov. 11:937–957.

145. Martin-Liberal, J., A. J. Cameron, J. Claus, I. R. Judson, P. J. Parker, and M. Linch. 2014. Targeting protein kinase C in sarcoma. Biochim. Biophys. Acta 1846:547–559.

146. Garg, R., L. G. Benedetti, M. B. Ahera, H. Wang, M. Abba, and M. G. Kazaniezt. 2014. Protein kinase C and cancer: what we know and what we do not. Oncogene 33:5225–5237.

147. Schwartz, C., S. Bouchat, C. Marban, V. Gautier, C. Van Lint, O. Rohr, et al. 2017. On the way to find a cure: purging latent HIV-1 reservoirs. Biochem. Pharmacol. 146:10–22. https://doi.org/10.1016/j.bcp.2017.07.001

148. Spear, M. A., P. LoRusso, A. Mita, and M. Mita. 2011. Vascular disrupting agents (VDA) in oncology: advancing towards new therapeutic paradigms in the clinic. Curr. Drug Targets 12:2009–2015.

149. Ferrer, E., J. Bolos, and R. Castaner. 2010. Plinabulin: tubulin polymerization inhibitor vascular-disrupting agent oncolytic. Drugs Future 35:11–15.

150. Singh, A. V., M. Bandi, N. Raje, P. Richardson, M. A. Palladino, D. Chauhan, et al. 2011. A novel vascular disrupting agent plinabulin triggers INK-mediated apoptosis and inhibits angiogenesis in multiple myeloma cells. Blood 117:5692–5700.

151. Mita, M. M., M. A. Spear, L. K. Yee, A. C. Mita, E. I. Heath, K. P. Papadopoulos, et al. 2010. Phase 1 first-in-human trial of the vascular disrupting agent plinabulin (NPI-2358) in patients with solid tumors or lymphomas. Clin. Cancer Res. 16:5892–5899.

152. Millward, M., P. Mainwaring, A. Mita, K. Federico, G. K. Lloyd, N. Reddinger, et al. 2012. Phase 1 study of the novel vascular disrupting agent plinabulin (NPI-2358) and docetaxel. Invest. New Drugs 30:1065–1073.

153. El Bairi, K., M. Amrani, A. H. Kandhro, and S. Afqir. 2017. Prediction of therapy response in ovarian cancer: where are we now? Crit. Rev. Clin. Lab. Sci. 54:233–266.

154. O’Connor, J. P., A. Jackson, G. J. Parker, C. Roberts, and G. C. Jayson. 2012. Dynamic contrast-enhanced MRI in clinical trials of antivascular therapies. Nat. Rev. Clin. Oncol. 9:167–177.

155. Alonso-Álvarez, S., E. Pardal, D. Sánchez-Nieto, M. Navarro, M. D. Caballero, M. V. Mateos, et al. 2017. Plitidepsin: design, development, and potential place in therapy. Drug Des. Devel. Ther. 11:253–264.

156. Xing, H., M. Tong, N. Jiang, X. Zhang, H. Hu, H. Pan, et al. 2017. Antitumor bioactive peptides isolated from marine organisms. Clin. Exp. Pharmacol. Physiol. 44:1077–1082. https://doi.org/10.1111/1440-1681.12808

157. Danu, A., C. Willekens, and V. Ribrag. 2013. Plitidepsin: an orphan drug. Expert Opin. Orphan Drugs 1:569–580.

158. Morande, P. E., S. R. Zanetti, M. Borge, P. Nannini, C. Jancic, R. F. Bezares, et al. 2012. The cytotoxic activity of Aplidin in chronic lymphocytic leukemia (CLL) is mediated by a direct effect on leukemic cells and an indirect effect on monocyte-derived cells. Invest. New Drugs 30:1830–1840.

159. Losada, A., M. J. Muñoz-Alonso, C. García, P. A. Sánchez-Murcia, J. F. Martínez-Leal, J. M. Domínguez, et al. 2016. Translation elongation factor eEF1A2 is a novel anticancer target for the marine natural product plitidepsin. Sci. Rep. 6:35100.

160. Muñoz-Alonso, M. J., E. Álvarez, M. J. Guillén-Navarro, M. Pollán, P. Avilés, C. M. Galmarini, et al. 2013. c-Jun N-terminal kinase phosphorylation is a biomarker of plitidepsin activity. Mar. Drugs 11:1677–1692.

161. Caers, J., E. Menu, H. De Raeve, D. Lepage, E. Van Valkenborgh, B. Van Camp, et al. 2008. Antitumour and antiangiogenic effects of Aplidin® in the 5TMM syngeneic models of multiple myeloma. Br. J. Cancer 98:1966–1974.

162. Taraboletti, G., M. Poli, R. Dossi, L. Manenti, P. Borsotti, G. T. Faircloth, et al. 2004. Antiangiogenic activity of aplidine, a new agent of marine origin. Br. J. Cancer 90:2418–2424.

163. Brogioni, M., S. V. Marchini, E. Galliera, P. Borsotti, G. Taraboletti, E. Erba, et al. 2003. Aplidin®, a new anticancer agent of marine origin, inhibits vascular endothelial growth factor (VEGF) secretion and blocks VEGF-VEGFR-1 (flt-1) autocrine loop in human leukemia cells MOLT-4. Leukemia 17:52–59.

164. Rockwell, S., and Y. Liu. 2010. Aplidin as a potential adjunct to radiation therapy: in vitro studies. Int. J. Radiat. Biol. 86:63–70.

165. Aspelasgh, S., M. Stein, R. Bahleda, A. Hollebeque, G. Salles, E. Gyan, et al. 2017. Phase I dose-escalation study of plitidepsin in combination with sorafenib or gemcitabine in patients with refractory solid tumors or lymphomas. Anticancer Drugs 28:341–349.

166. Aspelasgh, S., A. Awada, P. Affimos, R. Bahleda, A. Varga, and J. C. Soria. 2016. Phase I dose-escalation
study of plitidepsin in combination with bevacizumab in patients with refractory solid tumors. Anticancer Drugs 27:1021–1027.

167. Salazar, R., R. Plummer, A. Oaknin, A. Robinson, B. Pardo, A. Soto-Matos, et al. 2011. Phase I study of weekly plitidepsin as 1-hour infusion combined with carboplatin in patients with advanced solid tumors or lymphomas. Invest. New Drugs 29:1406–1413.

168. Geoerger, B., E. J. Estlin, I. Aerts, P. Kearns, B. Gibson, N. Corradi, et al. 2012. A phase I and pharmacokinetic study of plitidepsin in children with advanced solid tumours: an Innovative Therapies for Children with Cancer (ITCC) study. Eur. J. Cancer 48:289–296.

169. Plummer, R., P. Lorigan, E. Brown, R. Zaucha, V. Moiseyenko, L. Demidov, et al. 2013. Phase I-II study of plitidepsin and dacarbazine as first-line therapy for advanced melanoma. Br. J. Cancer 109:1451–1459.

170. Faivre, S., S. Chièze, C. Delbaldo, N. Ady-Vago, C. Guzman, L. Lopez-Lazaro, et al. 2005. Phase I and pharmacokinetic study of aplidine, a new marine cyclodepsipeptide in patients with advanced malignancies. J. Clin. Oncol. 23:7871–7880.

171. Izquierdo, M. A., A. Bowman, M. García, D. Jodrell, M. Martínez, B. Pardo, et al. 2008. Phase I clinical and pharmacokinetic study of plitidepsin as a 1-hour weekly intravenous infusion in patients with advanced solid tumors. Clin. Cancer Res. 14:3105–3112.

172. Maroun, J. A., K. Belanger, L. Seymour, S. Matthews, J. Roach, J. Dionne, et al. 2006. Phase I study of aplidine in a daily 5 one-hour infusion every 3 weeks in patients with solid tumors refractory to standard therapy. A National Cancer Institute of Canada Clinical Trials Group study: NCIC CTG IND 115. Ann. Oncol. 17:1371–1378.

173. Toulmonde, M., A. Le Cesne, S. Piperno-Neumann, N. Penel, C. Chevreau, and F. Duffaud. 2015. Aplidin in patients with advanced dedifferentiated liposarcomas: a French Sarcoma Group Single-Arm Phase II study. Ann. Oncol. 26:1465–1470.

174. Ribrag, V., D. Caballero, C. Fermé, E. Zucca, R. Arranz, J. Briones, et al. 2013. Multicenter Phase II study of plitidepsin in patients with relapsed/refractory non-Hodgkin's lymphoma. Haematologica 98:357–363.

175. Baudin, E., J. P. Droz, L. Paz-Ares, A. T. van Oosterom, M. Cullell-Young, and M. Schlumberger. 2010. Phase II study of plitidepsin 3-hour infusion every 2 weeks in patients with unresectable advanced medullary thyroid carcinoma. Am. J. Clin. Oncol. 33:83–88.

176. Mateos, M. V., M. T. Cibeira, P. G. Richardson, F. Prosper, A. Oriol, I. de la Rubia, et al. 2010. Phase II clinical and pharmacokinetic study of plitidepsin 3-hour infusion every two weeks alone or with dexamethasone in relapsed and refractory multiple myeloma. Clin. Cancer Res. 16:3260–3269.

177. Eisen, T., N. Thatcher, S. Leyvraz, W. H. Miller, F. Couture, P. Lorigan, et al. 2009. Phase II study of weekly plitidepsin as second-line therapy for small cell lung cancer. Lung Cancer 64:60–65.

178. Eisen, T., I. Thomas, W. H. Jr Miller, M. Gore, P. Wolter, P. Kavan, et al. 2009. Phase II study of biweekly plitidepsin as second-line therapy in patients with advanced malignant melanoma. Melanoma Res. 19:185–192.

179. Dumez, H., E. Gallardo, M. Garcia, F. Rivera, J. Tabernero, M. Cullell, et al. 2009. Phase II randomized study of Plitidepsin (Aplidin), alone or in association with L-carnitine, in patients with unresectable advanced renal cell carcinoma. Mar. Drugs 7:451–463.

180. Schöffski, P., V. Guillem, M. Garcia, F. Rivera, J. Tabernero, M. Cullell, et al. 2009. Phase II randomized study of Plitidepsin (Aplidin), alone or in association with L-carnitine, in patients with unresectable advanced renal cell carcinoma. Mar. Drugs 7:57–70.

181. Peschel, C., J. T. Hartmann, A. Schmittel, C. Bokemeyer, F. Schneller, U. Keilholz, et al. 2008. Phase II study of plitidepsin in pretreated patients with locally advanced or metastatic transitional cell carcinoma of the urothelium. Mar. Drugs 6:357–369.

182. PharmaMar®, Press release. 2017. Available at https://www.pharmamar.com/newsroom/press-releases/ (accessed 10 August 2017).

183. PharmaMar®, Press release. 2016. Available at https://www.pharmamar.com/newsroom/press-releases/ (accessed 10 August 2017).

184. Wu, X. Y., W. Ma, K. Gurung, and C. H. Guo. 2013. Mechanisms of tumor resistance to small-molecule vascular disrupting agents: treatment and rationale of combination therapy. J. Formos. Med. Assoc. 112:115–124.

185. Giuliano, S., and G. Pagès. 2013. Mechanisms of resistance to anti-angiogenesis therapies. Biochimie 95:1110–1119.

186. Close, A. 2016. Antiangiogenesis and vascular disrupting agents in cancer: circumventing resistance and augmenting their therapeutic utility. Future Med. Chem. 8:443–462.

187. Clémenson, C., C. Chargari, and E. Deutsch. 2013. Combination of vascular disrupting agents and ionizing radiation. Crit. Rev. Oncol. Hematol. 86:143–160.

188. Nguyen, L., T. Fifis, C. Malcontenti-Wilson, L. S. Chan, P. N. L. Costa, M. Nikfarjam, et al. 2012. Spatial morphological and molecular differences within...
solid tumors may contribute to the failure of vascular disruptive agent treatments. BMC Cancer 12:522.

189. Monk, B. J., M. W. Sill, J. L. Walker, C. J. Darus, G. Sutton, K. S. Tewari, et al. 2016. Randomized phase II evaluation of bevacizumab versus bevacizumab plus fosfomycin in recurrent ovarian, tubal, or peritoneal carcinoma: an NRG Oncology/Gynecologic Oncology Group Study. J. Clin. Oncol. 34:2279–2286.

190. Taylor, M., F. Billiot, V. Marty, V. Rouffièac, P. Cohen, E. Tournay, et al. 2012. Reversing resistance to vascular-disrupting agents by blocking late mobilization of circulating endothelial progenitor cells. Cancer Discov. 2:434–449.

191. Shaked, Y., A. Ciarrrocchi, M. Franco, C. R. Lee, S. Man, A. M. Cheung, et al. 2006. Therapy-induced acute recruitment of circulating endothelial progenitor cells to tumors. Science 313:1785–1786.

192. Moschetta, M., Y. Mishima, I. Sahin, S. Manier, S. Glavey, A. Vacca, et al. 2014. Role of endothelial progenitor cells in cancer progression. Biochim. Biophys. Acta 1846:26–39.

193. Das, V., J. Steřánková, M. Hajdúch, and J. H. Miller. 2015. Role of tumor hypoxia in acquiring of resistance to microtubule-stabilizing drugs. Biochim. Biophys. Acta 1855:172–182.

194. Liberti, M. V., and J. W. Locasale. 2016. The warburg effect: how does it benefit cancer cells? Trends Biochem. Sci. 41:211–218.

195. Muz, B., P. de la Puente, F. Azab, and A. K. Azab. 2015. The role of hypoxia in cancer progression, angiogenesis, metastasis, and resistance to therapy. Hypoxia (Auckl) 11:83–92.

196. Chen, X., Y. Qian, and S. Wu. 2015. The warburg effect: evolving interpretations of an established concept. Free Radic. Biol. Med. 253–263.

197. https://www.clinicaltrials.gov/ct2/results?term=HIF&cond=Cancer&recrs=b&recrs=a&recrs=f&recrs=d&recrs=g&recrs=e&age_v=&gndr=&type=Intr&rslt=&Search=Apply (accessed 18 February 2018).

198. Guo, C., A. Buranych, D. Sarkar, P. B. Fisher, and X.-Y. Wang. 2013. The role of tumor-associated macrophages in tumor vascularization. Vasc. Cell 5:20.

199. De Palma, M., and C. E. Lewis. 2013. Macrophage regulation of tumor responses to anticancer therapies. Cancer Cell 23:277–286.

200. Ruffell, B., and L. M. Coussens. 2015. Macrophages and therapeutic resistance in cancer. Cancer Cell 27:462–472.

201. Welford, A. F., D. Biziato, S. B. Coffelt, S. Nucera, M. Fisher, F. Pucci, et al. 2011. TIE2-expressing macrophages limit the therapeutic efficacy of the vascular-disrupting agent combretastatin A4 phosphate in mice. J. Clin. Invest. 121:1969–1973.

202. Henze, A.-T., and M. Mazzone. 2016. The impact of hypoxia on tumor-associated macrophages. J. Clin. Invest. 126:3672–3679.

203. Noy, R., and J. W. Pollard. 2014. Tumor-associated macrophages: from mechanisms to therapy. Immunity 41:414–4161.

204. Chen, M., X. Lei, C. Shi, M. Huang, X. Li, B. Wu, et al. 2017. Pericyte-targeting prodrug overcomes tumor resistance to vascular disrupting agents. J. Clin. Invest. 127:3689–3701.

205. Shenoy, A. K., Y. Jin, H. Luo, M. Tang, C. Pampol, R. Shao, et al. 2016. Epithelial-to-mesenchymal transition confers pericyte properties on cancer cells. J. Clin. Invest. 126:4174–4186.

Supporting Information

Additional supporting information may be found in the online version of this article:
Table S1. Trabectedin phase III trials.
Table S2. Combretastatin clinical trials.
Table S3. Ombrabulin clinical trials.
Table S4. Completed clinical trials of bryostatin-1.
Table S5. Plinabulin clinical trials.
Table S6. Plitidepsin clinical trials.
Table S7. Promising anti-vascular natural compounds and their derivatives tested in (pre)clinical studies for cancer drug discovery.

Figure S1. Disposition of trabectedin clinical trials by phase number.
Figure S2. Disposition of OVA-301 phase III study.
Figure S3. Mechanism of action of combretastatin.
Figure S4. Survival of patients with soft tissue sarcoma according to tumor tissue markers.
Figure S5. Survival of ovarian cancer patients treated with trabectedin.
Box S1. Useful list of Medline-indexed and highly accessed journals studying angiogenesis and related oncology clinical trials.
Box S2. Additional useful reviews of particular interest.