Perilesional Inflammation in Neurocysticercosis - Relationship Between Contrast-Enhanced Magnetic Resonance Imaging, Evans Blue Staining and Histopathology in the Pig Model

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Abstract

Background

Disease manifestations in neurocysticercosis (NCC) are frequently due to inflammation of degenerating Taenia solium brain cysts. Exacerbated inflammation post anthelmintic treatment is associated with leakage of the blood brain barrier (BBB) using Evans blue (EB) staining. How well EB extravasation into the brain correlates with magnetic resonance imaging (MRI) using gadolinium (Gd) enhancement as a contrast agent and pericystic inflammation was analyzed in pigs harboring brain cysts of Taenia solium.

Methodology/Principal Findings

Three groups of 4 naturally infected pigs were assessed. The first and second groups were treated with both praziquantel plus albendazole and sacrificed two and five days post treatment, respectively. A third untreated group remained untreated. Pigs were injected with EB two hours prior to evaluation by Gd-enhanced T1-MRI, and euthanized. The EB staining for each cyst capsule was scored (EB grades were 0: 0%; 1: up to 50%; 2: over 50% but less than 100%; 3: 100%). Similarly, the Gd enhancement around each cyst was qualitatively and quantitatively scored from the MRI. The extent of pericystic inflammation on histology...
was scored in increasing severity as IS1, IS2, IS3 and IS4. Grade 3 EB staining and enhancement was only seen in treated capsules. Also, treated groups had higher Gd intensity than the untreated group. Grades of enhancement correlated significantly with Gd enhancement intensity. EB staining was correlated with Gd enhancement intensity and with IS4 in the treated groups. These correlations were stronger in internally located cysts compared to superficial cysts in treated groups.

**Significance**

EB staining and Gd enhancement strongly correlate. The intensity of enhancement determined by MRI is a good indication of the degree of inflammation. Similarly, EB staining highly correlates with the degree of inflammation and may be applied to study inflammation in the pig model of NCC.

**Author Summary**

Neurocysticercosis (NCC) is a frequent parasitic infection of the human brain in developing countries. The symptomatology of human NCC after antiparasitic treatment is generally related to inflammation. The presence and degree of enhancement after intravascular injection of the contrast agent gadolinium in magnetic resonance imaging (MRI) is commonly considered an evidence of blood brain barrier (BBB) leakage. Experimentally, the presence and degree of extravasation of Evans blue (EB) after intravascular injection into the tissues of the brain is a direct measure of blood brain barrier leakage. The BBB leakage of gadolinium in neurocysticercosis is commonly used as an indirect measure of inflammation but has never been experimentally proven. Here we evaluated the relationship between contrast T1-MRI, EB staining and histology findings in naturally infected pigs. There was a strong correlation between EB staining, contrast MRI and histopathology findings after antiparasitic treatment. This correlation was stronger when cysts were internally located in the brain than in superficial cysts partly located in the subarachnoid space (meninges). Contrast-enhanced MRI is a non invasive tool used in diagnosis and follow up of NCC patients. This study shows that the use of EB staining allows for the same conclusions as when using MRI post-treatment, and that both techniques correlate with histopathology findings. These results support the use of EB staining to study NCC using the porcine model as well as validate MRI enhancement to assess brain inflammation in patients.

**Introduction**

Neurocysticercosis (NCC), infection of the brain by the larval stage of the parasite *Taenia solium*, is a common cause of epilepsy in endemic countries [1]. Imaging studies are essential to diagnose NCC. Magnetic resonance imaging (MRI) is more sensitive than computed axial tomography (CT), providing better anatomical differentiation, superior visualization of small lesions, edema, vascular enhancement and tissue changes; CT scan is more sensitive for the detection of calcified lesions [2, 3].

Cysts that have formed in the brain parenchyma can remain quiescent for a period of months to years. At some point, either as a result of the natural course of disease or because of
cysticidal treatment, the host mounts a focal inflammatory immune response to the cyst resulting in parasite degeneration [1]. A contrast-enhanced MRI at this time shows a hyperdense ring in the adjacent capsule surrounding the cysticercus, reflecting blood brain barrier (BBB) disruption [4]. Previous experiments by our group using a T. solium naturally-infected pig model [5, 6] measured the degree of capsular inflammation and the extravasation of Evans blue dye (EB) into the inflamed capsules of degenerating cysts. The presence of EB capsular staining, a measure of BBB disruption, correlates with the degree of pericystic inflammation [6].

Among imaging techniques, fluid-attenuated inversion recovery (FLAIR) and contrast-enhanced T1-MRI using gadolinium as contrast agent are the most useful for diagnostics and follow up, as they provide more detail about the stage of the inflammatory response and the evolution of the damage suffered by the parasite [3]. In the evaluation of human NCC, the degree of enhancement in the contrast T1-MRI protocol is commonly assumed to be a measure of the amount of inflammation present in specific lesions [4]. Since both contrast enhancement and EB staining reflect BBB dysfunction, we assessed whether these markers correlate between themselves and to pericystic brain inflammation as determined on histology. A significant correlation would support the use of enhancement on MRI as a measure of inflammation in swine and by analogy to human NCC.

**Materials and Methods**

**Study design and animals**

We compared pericystic EB staining, gadolinium enhancement on MRI, and histological findings following cysticidal treatment of pigs naturally infected with T. solium brain cysts. An individual cyst was considered the unit of analysis for this study. Twelve pigs naturally infected with T. solium cysts from endemic Peruvian highland villages, confirmed by positive tongue examination [7], were imaged by MRI to confirm brain infection and then randomly divided in three groups of four pigs each. One group remained untreated as a untreated, and the other two groups were treated with albendazole (Zentel, GlaxoSmithKline, Peru) at 15 mg/kg daily until sacrifice plus a single day treatment of praziquantel (Helmiben, Farmindustria, Peru) at 75 mg/kg divided into three doses of 25 mg/kg every two hours on the first day [7]. Pigs in the treatment groups were sacrificed two (n = 4) and five (n = 4) days after treatment.

**Post-treatment procedures**

Immediately before sacrifice, all pigs were anesthetized with an intramuscular injection of a mixture of ketamine (Ket-A-100 50 mg/kg, Agrovet Market SA, Peru) and xylazine (Dormi-Xyl 2mg/kg, Agrovet Market SA, Peru) through a catheter inserted into the marginal ear artery and kept latent by very slow normal saline drip. Then they were infused through the ear catheter with EB dye first [5], and after 45 minutes with Gadolinium diethylene triaminopentaacetic acid (Gd-DTPA) at 0.1 mmol/kg for contrast-enhanced brain MRI. Shortly after the MRI, the pigs were perfused intraaortally for 15 minutes with normal saline solution using a peristaltic pump and euthanized with a lethal IM dose of 120 mg/kg of pentobarbital. The pigs’ brains were retrieved at necropsy and examined macroscopically and by histology.

**Evans Blue staining**

A 2% EB (Sigma–Aldrich, St. Louis, MO) solution in normal saline was administered by the EB catheter as reported [5, 6] and allowed to circulate for 2 hours under additional sodium pentobarbital sedation (Halatal, Agrovet Market SA Peru, at 25 mg/kg every 45 min).
Grading of EB staining. Increase of BBB permeability was assessed by the degree of extravasation of EB into the host tissue around the cysticercus (capsule). Cyst capsules were scored qualitatively according to distribution of the color per cyst. EB staining grades were 0: 0%; 1: up to 50% of the capsule was blue; 2: over 50% of the capsule was blue; 3: 100% of the capsule was blue (Fig 1).

MRI examinations
Sedated pigs were placed on a surgical table in a side-lying position. Pre and post-contrast MRI were performed on a 3-Tesla scanner (Philips Achieva, Best, The Netherlands) including axial, coronal, and sagittal TFE (Turbo field echo). T1-weighted gradient-echo images were taken with 1–4 mm section thickness, 7 milliseconds (ms) of repetition time, 4 ms echo time, 8° flip angle, 270kHz pixel bandwidth and matrix = 256–480 pixels.

MRI imaging interpretation and analysis. Post-contrast MR images were used to examine and classify cyst lesions using qualitative and quantitative measures. Positive enhancement is defined as a post-contrast hyperintense signal along the edge of a cyst. Qualitative evaluation was based upon the percent of cyst involved in the signal; all consecutive images (1–10) that belonged to each cyst were individually graded in a way similar as used for EB staining: 0 (no enhancement); 1 (up to 50% of the capsule enhanced); 2 (over 50% of the capsule enhanced), or 3 (100% of the capsule enhanced) (Fig 2). A global, single grade was assigned to each cyst by the following criteria: grades 0 and 3 required all of its images to be grade 0 or 3; grade 1 was given when at least one image was grade 1 but none was grade 2 or 3; cysts with images in any other combination were grade 2.

A quantitative measurement of Gd enhancement intensity was also built using the open access image-processing program FIJI (FIJI/ImageJ, NIH, Bethesda, Maryland, USA) to convert digital imaging and communication in medicine (DICOM) scan images to tagged image file format (TIFF). Brightness and contrast in the TIFF scan images were first adjusted to define and delineate pericystic lesion areas, and then the gray values in each pixel within these selections were normalized using the percentage of a continuous scale from 0 (black) to 256 (brightest). Then, the average gray value from all pixels within the selection of each image was

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Finally, we calculated the average of intensity of Gd enhancement in all pericystic section images in order to obtain a single value for each cyst using R program v3.2.2 [8].

**Brain examinations**

Immediately upon extraction, brains were placed on dry ice, which helped with the slicing of the samples into 1-cm coronal sections. The anterior and posterior surfaces of each section were photographed.

**Cyst location.** Cysticerci were characterized as superficial or deep cysts. Superficial cysts are those in a cortical location with a region exposed to meninges and another embedded in parenchyma, mainly in gray matter [9]. Deep cysts were those totally embedded into the brain parenchyma (white and gray matter), with close proximity to the brain center (coordinates x, y, z = 0, 0, 0).

**Histology.** All capsules were excised from the brains. And each biopsy from the right brain hemispheres was separately fixed in 10% neutral buffered formalin (3.7% formaldehyde in PBS, pH 7.2). Fixed biopsies were embedded in paraffin and sliced in coronal 4 μm-thick sections. Conventional hematoxylin-eosin stains were performed on every slide and two sections of the same cyst were examined with conventional light microscopy. Microphotographs taken with 15X magnification were taken with a Carl Zeiss stereoscope and assembled using AxioVision software (Rel. 4.6, Zeiss, Oberkochen, Germany) to obtain a single large image ("cyst map"). Biopsies from the left hemispheres were saved for parallel studies.

**Inflammatory response.** As in previous studies, the areas of inflammatory response around the cyst were categorized in inflammatory stages (IS) 1 to 4, where IS1 was a thin layer of collagen with scarce or null immune cells, IS2 was a thicker layer of collagen with more non organized immune cells, IS3 was a typical granulomatous reaction with abundant immune cells distributed in layers (an epithelioid cells or eosinophil-rich layer near to the cyst wall with few multinucleated giant cells) and IS4 was similar to IS3 but with higher numbers of eosinophils (identified by their red staining granules) and with an increase of multinucleated giant cells distributed in a layer close to the cyst wall with the parasite structure clearly damaged (Fig 3). Since the IS around a cyst was usually not homogeneous, the proportions (percents) of the perimeter presenting each inflammatory stage were recorded using the cyst map [6].

**Statistical analysis**

Spearman’s rank correlation was used to evaluate the correlation between the qualitative and quantitative measure of Gd enhancement and the relationship between the grades of EB staining (ordinal variable) with quantitative measure of Gd enhancement in each treatment group, and the correlation between each inflammatory stage (IS1 to IS4 extension expressed in

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**Fig 3. Inflammatory stages.** cw = Cyst wall, c = collagen, i = immune cells infiltrate, ci = collagen plus immune cells infiltrate, bp = brain parenchyma. A shows IS1 with few immune cells into the collagen layer. B shows IS2 where the immune cells are more abundant. C shows IS3 where the number of immune cells were increased and ordered in layers. D shows IS4 where multinucleated giant cells are increased, the cyst wall seen damaged and a near layer of eosinophils.

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percentage as a four continuous variables) with the grades of EB staining and quantitative enhancement measure. The non-parametric Wilcoxon-Mann Whitney test was used to compare Gd enhancement intensity and EB grades between the different locations of the cyst in each individual treatment group. All the analyses were performed using the R v3.2.2 and graphs were created using ggplot package [8]. Differences were considered significant at p < 0.05.

Ethical statement

The study was conducted in accordance with the National Institutes of Health/AALC guidelines, and was reviewed and approved by the Institutional Animal Care and Use Committee Animal Ethics of the Universidad Peruana Cayetano Heredia (assurance Number: A5146-0).

Results

Cyst distribution per animal and per brain hemisphere

A total of 328 brain cysts from 12 naturally infected pigs were assessed in this study. The parasitic load per pig brain was widely distributed, ranging from 1 to 45 cysts in the untreated group, from 10 to 29 cysts in PZQ+ABZ 2d and from 4 to 152 cysts in the group allocated to PZQ+ABZ 5d. Pigs in the PZQ+ABZ 5d group had more cysts (n = 192) than did pigs in the untreated group (n = 73) or in the PZQ+ABZ 2d group (n = 63). The distribution of cysts in both hemispheres was similar in each pig, a total of 165 cysts were found in the right hemispheres of study pigs and 163 cysts were found in the left hemispheres. All brain cysts (n = 328) were used for macroscopic assessment of Evans blue uptake and also for Gd enhancement on MRI (Table 1).

Evans Blue staining

Cysts with clear capsules (grade 0) were seen only in untreated animals; whereas EB grade 3 capsules (completely blue) were seen only in animals treated with antiparasitic drugs (Table 2). Capsules from treated groups had significantly more EB staining than untreated capsules (PZQ+ABZ 2d versus untreated group, p = 0.02 and PZQ+ABZ 5d versus untreated group, p < 0.001 and p < 0.001 for comparisons of EB staining between: PZQ+ABZ 2d versus untreated group, PZQ+ABZ 5d versus untreated group, and PZQ+ABZ 5d vs PZQ+ABZ 2d, respectively.

Table 1. Cyst numbers and distribution in study pigs.

| Characteristics                          | Untreated (n = 4) | PZQ+ABZ 2d (n = 4) | PZQ+ABZ 5d (n = 4) | Total (n = 12) |
|-------------------------------------------|------------------|--------------------|-------------------|---------------|
| Brain cyst load (per pig)                 | 73 (1, 11, 16, 45) | 63 (10, 11, 13, 29) | 192 (4,13, 23, 152) | 328           |
| Total cyst distribution (right / left hemisphere distribution per pig) | 30/43 (0/1, 6/5, 5/11, 19/26) | 34/29 (7/3, 5/6, 7/6,15/14) | 101/91 (3/1, 9/4, 13/10, 76/76) | 165/163       |

Table 2. Grades of Evans blue staining in brain cysts by treatment group. Values represent numbers of brain cyst capsules in each stratum.

| Evans blue staining (Cyst capsules, %) | Untreated | PZQ+ABZ 2d | PZQ+ABZ 5d |
|---------------------------------------|-----------|------------|------------|
| Grade 0                               | 6 (8%)    | 0 (0%)     | 0 (0%)     |
| Grade 1                               | 34 (47%)  | 5 (8%)     | 10 (5%)    |
| Grade 2                               | 33 (45%)  | 19 (30%)   | 144 (75%)  |
| Grade 3                               | 0 (0%)    | 39 (62%)   | 38 (20%)   |

p = 0.02, p< 0.001 and p< 0.001 for comparisons of EB staining between: PZQ+ABZ 2d versus untreated group, PZQ+ABZ 5d versus untreated group, and PZQ+ABZ 5d vs PZQ+ABZ 2d, respectively.
p < 0.001). However the capsules from PZQ+ABZ 2d had more EB staining than those in the PZQ+ABZ 5d group (p < 0.001).

The apparent decrease in EB staining from day 2 to day 5 (proportional increase in EB grade 2 capsules with fewer grade 3 capsules) may be due to one pig contributing 128 out of the 144 grade 2 capsules in day 5 (S1 Table).

Gadolinium enhancement in MRI

Similarly to the EB staining findings, cyst capsules without Gd enhancement (grade 0 of enhancement) were seen only in the untreated group, whereas completely enhanced capsules (grade 3) were seen only in treated animals. Capsules in treated groups had more Gd enhancement than capsules from the untreated group (p < 0.001), however there were marginal differences between both treated groups (p = 0.06) (Table 3). Again, although the data suggest a maximum effect on day 2, this may have been caused by many grade 2 cysts in the PZQ+ABZ 5d group that came from the same pig (S2 Table). Quantitative measurements of Gd enhancement by cyst were highly correlated to the above-described qualitative assessment (Table 4). From here on, all analyses shown refer to the quantitative measurement of enhancement, which is less subjective.

Correlation between EB staining and Gd enhancement in MRI

We analyzed the relationship between the different grades of EB staining and Gd enhancement intensity at the level of individual cysts per treatment group. Gd enhancement intensity had a significant tendency to increase with EB grades in all groups (Table 5, Fig 4).

Table 3. Qualitative grade assessment of Gd enhancement in MRI of brain cysts by treatment group. Values represent numbers of brain cyst capsules in each stratum.

| Gadolinium enhancement (Cyst capsules, %) | Untreated | PZQ+ABZ 2d | PZQ+ABZ 5d |
|-----------------------------------------|-----------|------------|------------|
| Grade 0                                 | 4 (6%)    | 0 (0%)     | 0 (0%)     |
| Grade 1                                 | 58 (79%)  | 10 (16%)   | 22 (11%)   |
| Grade 2                                 | 11 (15%)  | 39 (62%)   | 161 (84%)  |
| Grade 3                                 | 0 (0%)    | 14 (22%)   | 9 (5%)     |

p < 0.001, p < 0.001 and p = 0.06 for comparisons of Gd enhancement between PZQ+ABZ 2d versus untreated group, PZQ+ABZ 5d versus untreated group, and PZQ+ABZ 5d vs PZQ+ABZ 2d, respectively.

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Table 4. Correlation between qualitative and quantitative Gd enhancement measures in brain cysts by treatment group. Values represent medians and ranges of quantitative Gd enhancement intensity measure of brain cysts in each stratum.

| Gadolinium enhancement (qualitative) | Grade 0 | Grade 1 | Grade 2 | Grade 3 |
|--------------------------------------|---------|---------|---------|---------|
| Untreated *                          | 18.30 (16.63–24.11) | 34.43 (11.23–40.15) | 35.31 (22.05–42.0) | 0 (0–0) |
| PZQ+ABZ 2d **                        | 0 (0–0)  | 30.96 (25.86–43.73) | 34.51 (28.86–42.29) | 41.37 (36.01–45.81) |
| PZQ+ABZ 5d ***                       | 0 (0–0)  | 27.01 (17.95–41.85) | 36.09 (16.49–48.49) | 41.01 (24.95–51.54) |

* r = 0.223, p = 0.0582  
** r = 0.6098, p < 0.001  
*** r = 0.3898, p < 0.001

Spearman’s correlation between quantitative and qualitative Gd enhancement.

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Table 5. Correlation between EB staining and Gd enhancement intensity (quantitative) in MRI (n = 328 cysts). Data in each cell represent median and range for cysts in the stratum.

| Gadolinium enhancement intensity | Evans blue staining | Grade 0 | Grade 1 | Grade 2 | Grade 3 |
|---------------------------------|--------------------|--------|--------|--------|--------|
| Untreated *                     |                    | 18.93  (16.63–24.56) | 31.77  (11.23–39.51) | 36.73  (22.44–42.0) | 0 (0–0) |
| PZQ+ABZ 2d **                   |                    | 0 (0–0) | 28.86  (25.86–32.84) | 31.44  (29.31–35.75) | 38.87  (30.75–45.81) |
| PZQ+ABZ 5d ***                  |                    | 0 (0–0) | 20.88  (14.99–23.90) | 35.07  (21.23–45.60) | 41.25  (29.12–51.54) |

* $r = 0.5960$, $p < 0.001$

** $r = 0.7860$, $p < 0.001$

*** $r = 0.5415$, $p < 0.001$

Spearman's correlation between quantitative Gd enhancement and EB staining.

Fig 4. Quantitative assessment of perycistic Gd enhancement intensity on MRI by grades of EB staining and treatment group.

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EB staining and Gd enhancement by cyst location

Since the inflammatory response can vary according to cyst location, we performed a stratified analysis between deep and superficial cysts (S3 Table). Considering the location of all cysts (328), there were 214 superficial and 114 deep cysts. Superficial cysts were more frequent in the PZQ+ABZ 5d group, (153 of 192), while deep and superficial cysts had similar frequency in the untreated (30 of 73) and the PZQ+ABZ 2d groups (31 of 63).

Within strata of treatment group, EB staining grades were significantly higher in deep cysts than in superficial cysts. These differences were statistically significant in the untreated group (p<0.001), where the majority of deep cysts were grade 2 (26/43, 60%) whereas superficial cysts were grade 1 (19/30, 64%). The same significant difference was observed in PZQ+ABZ 5d group (p<0.001), where the majority of deep cysts were grade 3 (30/39, 77%), whereas the superficial cysts were grade 2 (135/153, 88%). In PZQ+ABZ 2d group we found a marginal significance (p = 0.052) between deep (25/32, 78% were grade 3) and superficial cysts (17/31, 55% were grade 2). Gd enhancement intensity (quantitative measure) was significantly higher in deep cysts than in superficial cysts in the PZQ+ABZ 2d group (median 40.01 versus 33.02, p<0.001) and in the PZQ+ABZ 5d group (40.7 versus 35.07, p<0.001), but not in the untreated group (34.38 versus 33.57, p = 0.213).

Histopathological assessment

Since the left brain hemispheres were reserved for molecular biology studies, only those cysts located in right brain hemispheres (n = 165) were available for microscopic assessment. Right hemisphere cysts were similar to those in the left hemisphere in terms of grade of EB staining or intensity of Gd enhancement (S4 Table). From these, 113 cysts (69%) from 11 pigs had available slides showing complete parasite structures (cyst wall and scolex with adjacent brain with immune response) and were thus used for histopathological studies in order to correlate inflammatory findings with the corresponding EB staining and MRI findings.

The distribution of inflammatory stages by cyst and treatment group was consistent with EB staining and Gd enhancement in the entire cyst population, with no extent of IS4 in untreated cysts and few extents of IS1 and IS2 in cysts from treated pigs (Table 6).

When assessed within this population of 113 cysts, we first correlated each inflammatory stage with EB staining and Gd enhancement intensity in each treatment group. A positive and significant correlation was found only between intensity enhancement and IS4 (Spearman rank correlation; r = 0.363, p = 0.041) in the PZQ+ABZ 2d group. There were significant and positive correlations between IS3 and Gd enhancement intensity in the PZQ+ABZ 5d group (Spearman rank correlation; r = 0.358, p = 0.006) and between IS4 and EB staining (Spearman rank correlation; r = 0.578 p<0.001) and intensity enhancement (Spearman rank correlation; 0.50, p<0.001) in the PZQ+ABZ 5d. Also, IS4 was significantly higher in deep cysts than in superficial cysts in the PZQ+ABZ 5d group (median 10 versus 0, p<0.001) (Fig 5).

Table 6. Grading of inflammatory stage (histopathological assessment) by treatment group. Data in each cell represent median and range for cysts in the stratum.

| Inflammatory stages (n = 113) | Untreated Median of IS (range) | PZQ+ABZ 2d Median of IS (range) | PZQ+ABZ 5d Median of IS (range) |
|------------------------------|-------------------------------|---------------------------------|---------------------------------|
| IS 1                         | 0 (0–28)                      | 0 (0–0)                         | 0 (0–21)                        |
| IS 2                         | 60.5 (0–100)                  | 0 (0–40)                        | 0 (0–100)                       |
| IS 3                         | 37.5 (0–100)                  | 90 (0–100)                      | 78 (0–100)                      |
| IS 4                         | 0 (0–0)                       | 10 (0–100)                      | 5 (0–100)                       |

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Discussion

The study of inflammation due to degeneration of *T. solium* cysts (natural or induced by anti-parasitic treatment) in NCC is limited by the lack of suitable animal models. Brain cysts can be found in the natural intermediate host, the pig, and studied by brain imaging (particularly MRI), or histopathology (macroscopically by assessing EB leakage, or by standard histological or immunohistochemistry techniques). MRI has been used in pigs naturally infected with NCC, providing important information on lesion characteristics (localization, parasite load, localization and stage of lesions, inflammation and perilesional edema, etc.) [10–14], suggesting its usefulness to evaluate treatment schemes in the pig model. Disruption of BBB is associated with high levels of pro-inflammatory markers and inflammatory cell recruitment into pericystic tissue [6].

The injection of gadolinium as an MRI contrast agent reveals alterations in the integrity of the BBB in various neurological diseases (multiple sclerosis, strokes, acute ischemic brain injury, brain tumors, encephalomyelitis, etc.) [15–17]. The same mechanism is observed when the EB stain is used to evaluate alterations in BBB permeability. Our group has applied EB staining to mark BBB disruption in porcine NCC, demonstrating that treatment with praziquantel induces inflammation, cyst damage and BBB leakage at 2 and 5 days (48 and 120 h); we also have demonstrated that the BBB leakage is accompanied with expression of pro-inflammatory and immunoregulatory cytokines [5, 6]. In this study we evaluated the correlation between EB staining and contrast enhancement on MRI in regards to the inflammation and BBB leakage in the pig model of NCC. Our results show that EB extravasation around cysts as a marker of BBB disruption is equivalent to gadolinium enhancement on contrast T1 MRI, reflecting the location and intensity of pericystic inflammation areas. There was a strong correlation between both techniques and the histopathological findings; the location and intensity of gadolinium enhancement, as well as EB staining, reflected the degree and areas of histologically assessed inflammation. Using grades of EB staining and Gd enhancement also allowed for the understanding of the evolution of BBB disruption in the pericystic tissue as the immune response and local inflammation increased in response to anthelmintic treatment. Gd enhancement and EB staining increased markedly at 2 and 5 days post-treatment, which

Fig 5. Correlation between EB staining, MRI findings and histology in deep and superficial cysts from PZQ+ABZ 5d group. A and F shows EB staining grade 3 cysts (black arrows) in macroscopic photographs. B and G show enhanced cysts (white arrows) in coronal T1-MRI. C and H show hematoxylin & eosin images (25X magnification; bar: 500 μm) where “e” indicates the scolex. C shows a deep cyst with 100% of IS4 extension (red line) whereas H shows a superficial cyst with a 10% of IS4 extension (red line). D and I are the 400X magnification (bar: 50 μm) of the circle (host-parasite interface) in C and H, respectively, and show IS4 (cw = cyst wall with damage, bp = brain parenchyma, i = infiltrate, mainly with eosinophils and lymphocytes). E and J show 1000x magnification (bar: 10 μm) of D and I (m = multinucleated giant cell, eo = eosinophils cells).

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strongly correlated with greater perilesional inflammation. These results are consistent with other studies that evaluated the disruption of the BBB in brain tumors [18] and may explain the increase of symptoms in patients after the onset of antiparasitic treatment [19].

Chronic inflammation by *Taenia solium* cysts may cause increased angiogenesis in the granuloma around the cysts [20]. These neo-vessels may become more susceptible to vascular leakage after anthelminthic treatment due to the stimulus by pro-inflammatory molecules, thus the correlation of EB staining and Gd enhancement with the severe inflammatory response in NCC. In brain tumors there is a strong association between contrast enhancement and tumor neovascularization [21–23]. The association between brain inflammation and BBB disruption was also described in neurodegenerative diseases as Alzheimer's disease and multiple sclerosis. In these diseases, the disruption of BBB integrity is associated with extravasation of proinflammatory molecules into the brain that cause damage in the nervous cells [24–25].

The inflammatory response varies according to the location of the parasite in the brain in relation to the parenchyma or the meninges; it is more evident in cysts surrounded by parenchyma [9]. In this study, deep (parenchymal) cysticerci showed stronger contrast enhancement in relation to areas where the inflammatory response was severe (IS3 or IS4), whereas superficial cysts frequently had a thin, weak enhancement signal and moderate or less severe inflammation in histological examinations (IS2 with some areas of IS3). This suggests enhancement is a marker of more intense and destructive inflammation as seen in other studies [26]. These findings are also consistent with our previous histological findings in the same model [9].

Our study had some limitations. First, the parasite load by pig was very variable, leading us to adjust analyses to account for this variability in the multivariate models. Second, the correlation with histopathology studies only used cysts located in the right hemisphere, although we found no reason to suspect any systematic differences between cysts in the left and right brain hemispheres [27], as seen in previous studies. Third, our analyses rely in two histological sections of cysts to determine the immune response; the two sections may not be representative of the inflammation around the whole cyst. Despite these limitations, the strong correlation between Gd enhancement on MRI and EB staining suggests that both methods are reliable in the evaluation of perilesional inflammation and BBB disruption in the porcine model of NCC and can be used alternatively or in combination.

Supporting Information

**S1 Table. Grades of Evans blue staining in brain cysts per pigs by treatment group.** Values represent numbers of brain cyst capsules in each pig. (DOCX)

**S2 Table. Qualitative grade assessment of gadolinium enhancement in MRI of brain cysts per pigs by treatment group.** Values represent numbers of brain cyst capsules in each pig. (DOCX)

**S3 Table. Comparisons of Evans Blue staining and gadolinium enhancement intensity between cyst locations.** Median values and ranges are shown for Gd enhancement intensity. EB staining grades are shown as number of cyst capsules and percentages. n = Number of cyst capsules. p* = Wilcoxon-Mann Whitney test used to compare EB staining grades and enhancement intensity between superficial and deep cysts in each treatment group. (DOCX)

**S4 Table. Distribution of cyst characteristics between both hemispheres.** Median values and ranges are shown for Gd enhancement intensity. EB staining grades are shown as
number of cyst capsules and percentages. n = Number of cyst capsules. hem = hemisphere. 
p = Wilcoxon-Mann Whitney test was used to compare EB staining grades and Gd enhancement intensity between cysts located in right and in left hemispheres in each treatment group.

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Conceived and designed the experiments: JAB HHG TEN CC. Performed the experiments: JAB CC JCa AVC DS JCh. Analyzed the data: CC HHG TEN JAB. Contributed reagents/materials/analysis tools: HHG JAB CGG TEN AEG SM. Wrote the paper: CC HHG TEN SM CGG JAB.

**References**

1. Garcia HH, Rodriguez S, Friedland JS; The Cysticercosis Working Group in Peru. Immunology of Taenia solium taeniasis and human cysticercosis. Parasite Immunol. 2014; 36(8): 388–96. doi:10.1111/pim.12126 PMID: 24962350
2. Garcia HH, Del Brutto OH. Imaging findings in neurocysticercosis. Acta Trop. 2003 Jun; 87(1):71–8. PMID: 12781380
3. Ng SH, Tan TY, Fock KM. The value of MRI in the diagnosis and management of neurocysticercosis. Singapore Med J. 2000 Mar; 41(3):132–4. PMID: 11063199
4. Chang KH, Lee JH, Han MH, Han MC. The role of contrast-enhanced MR imaging in the diagnosis of neurocysticercosis. AJNR Am J Neuroradiol. 1991 May-Jun; 12(3):509–12. PMID: 2058504
5. Guerra-Giraldez C, Marzal M, Cangalaya C, Balboa D, Orrego MÁ, Paredes A, Gonzales-Gustavson E, Arroyo G, García HH, González AE, Mahanty S, Nash TE; cysticercosis working group in Peru. Disruption of the blood-brain barrier in pigs naturally infected with Taenia solium, untreated and after anthelmintic treatment. Exp Parasitol. 2013; 134(4):443–6. doi: 10.1016/j.exppara.2013.05.005 PMID: 23684909
6. Mahanty S, Orrego MA, Mayta H, Marzal M, Cangalaya C, Paredes A, Gonzales E, Arroyo G, Gonzalez AE, Guerra-Giraldez C, Garcia HH, Nash TE. Post-treatment vascular leakage and inflammatory responses around brain cysts in porcine neurocysticercosis. PLoS Negl Trop Dis. 2015; 9(3): e0003577. doi: 10.1371/journal.pntd.0003577 PMID: 25774662
7. Gonzalez AE, Bustos JA, Jimenez JA, Rodriguez ML, Ramirez MG, Gilman RH, Garcia HH; Cysticercosis Working Group in Peru. Efficacy of diverse anthelmintic treatments for cysticercosis in the pig model. Am J Trop Med Hyg. 2012 Aug; 87(2):292–6. PMID: 22855760
8. R Core Team (2015) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. http://www.R-project.org/.
9. Cangalaya C, Zimic M, Marzal M, González AE, Guerra-Giraldez C, Mahanty S, Nash TE, Garcia HH; Cysticercosis Working Group in Peru. Inflammation Caused by Praziquantel Treatment Depends on the Location of the Taenia solium Cysticercus in Porcine Neurocysticercosis. PLoS Negl Trop Dis. 2015 Dec 11; 9(12):e0004207. doi: 10.1371/journal.pntd.0004207 PMID: 26658257
10. Gupta RK, Awasthi R, Garg RK, Kumar N, Gupta PK, Singh AK, Sahoo P, Paliwal VK, Prasad KN, Pandey CM, Rathore RK. T1-weighted dynamic contrast-enhanced MR evaluation of different stages of neurocysticercosis and its relationship with serum MMP-9 expression. AJNR Am J Neuroradiol. 2013 May; 34(5):997–1003. doi:10.3174/ajnr.A3346 PMID: 23179648

11. Chawla S, Husain N, Kumar S, Pal L, Tripathi M, Gupta RK. Correlative MR imaging and histopathology in porcine neurocysticercosis. J Magn Reson Imaging. 2004 Aug; 20(2):15. PMID:2598111

12. Flisser A, Gonzalez D, Shkurovich M, Madrazo I, Correa D, Rodriguez-Carbayo J, Cohen S, Rodriguez-del-Rosal E, Collado M, Fernandez B, et al. Praziquantel treatment of porcine brain and muscle Taenia solium cysticercosis. 1. Radiological, physiological and histopathological studies. Parasitol Res. 1990; 76(3):263–9. PMID:349005

13. de Aluja AS, Gonzalez D, Rodriguez Carbajal J, Flisser A. Histological description of tomographic images of Taenia solium cysticerci in pig brains. Clin Imaging. 1989; 13(4):292–8. PMID:2598111

14. Gonzalez D, Rodriguez-Carbayo J, Aluja A, Flisser A. Cerebral cisticercosis in pigs studied by computed tomography and necropsy. Vet Parasitol. 1987; 26(1–2):55–69. PMID:340005

15. Bronen RA, Sze G. Magnetic resonance imaging contrast agents: theory and application to the central nervous system. J Neurosurg. 1990 Dec; 73(6):820–39. Review. PMID:2230966

16. Kobayashi Shigeru, Meira Adam, Uchidab Kenzo and Hayakawac Katsuhiko. American Journal of Neuroradiology. Imaging of Intraneural Edema by Using Gadolinium-Enhanced MR Imaging: Experimental Compression Injury.

17. Broom KA, Anthony DC, Blamire AM, Waters S, Styles P, Perry VH, Sibson NR. MRI reveals that early changes in cerebral blood volume precede blood-brain barrier breakdown and overt pathology in MS-like lesions in rat brain. J Cereb Blood Flow Metab. 2005 Feb; 25(2):204–16. PMID:15678123

18. Garcia HH, Gonzalez I, Lescano AG, Bustos JA, Zimic M, Escalante D, Saavedra H, Gavidia M, Rodriguez L, Najar E, Umeres H, Pretell EJ; Cysticercosis Working Group in Peru. Efficacy of combined anti-parasitic therapy with praziquantel and albendazole for neurocysticercosis: a double-blind, randomised controlled trial. Lancet Infect Dis. 2014 Aug; 14(8):687–95. doi:10.1016/S1473-3099(14)70779-0 PMID: 24999157

19. Sikasunge CS, Johansen MV, Phiri IK, Willingham AL 3rd, Leifsson PS. The immune response in Taenia solium neurocysticercosis in pigs is associated with astroglial, axonal degeneration and altered blood-brain barrier permeability. Vet Parasitol. 2009 Mar 23; 160(3–4):242–50. doi:10.1016/j.vetpar.2008.11.015 PMID: 19117683

20. Long OM. Capillary ultrastructure and the blood brain barrier in human malignant brain tumors. J Neurosurg 1970; 32:127–144 PMID:5411991

21. Nishio S, Ohta M, Abe M, et al. Microvascular abnormalities in ethylnitrosourea (ENU)-induced rat brain tumors: structural basis for altered blood brain barrier function. Acta Neuropathol (Berl) 1983; 59: 1–10 37.

22. Nystrom S. Pathological changes in blood vessels of human glioblastoma multiforme. Comparative studies using plastic casting, angiography, light microscopy and electron microscopy, and with reference to some other brain tumors. Acta Pathol Microbial Scand [Suppl] 1960; 49(137): 1–83.

23. Sun D, Newman TA, Perry VH, Weller RO. Cytokine-induced enhancement of autoimmune inflammation in the brain and spinal cord: implications for multiple sclerosis. Neuropathol Appl Neurobiol. 2004 Aug; 30(4):374–84. PMID: 15305983

24. Disruption in the Blood-Brain Barrier: The Missing Link between Brain and Body Inflammation in Bipolar Disorder? Patel Jay P., Frey Benicio N. Neural Plast. 2015; 2015: 708306. Published online 2015 May 13. doi: 10.1155/2015/708306 PMID: 26075104

25. Smirmiotopoulos JG, Murphy FM, Rushing EJ, Rees JH, Schroeder JW. Patterns of contrast enhancement in the brain and meninges. Radiographics. 2007 Mar-Apr; 27(2):525–51. Review. PMID: 17374867

26. Mkupasi EM, Ngowi HA, Sikasunge CS, Leifsson PS, Johansen MV. Distribution and histopathological changes induced by cysts of Taenia solium in the brain of pigs from Tanzania. J Helminthol. 2014; 6:1–6.