Accuracy of circulating microRNAs in diagnosis of sepsis: a systematic review and meta-analysis

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Abstract

Objectives: The aim of this study was to systematically assess the accuracy of circulating microRNAs (miRNAs) as a promising biomarker for sepsis via a meta-analysis.

Methods: PubMed, Cochrane Library, Embase, Web of Science, Scopus, and Ovid databases were searched up to April 3, 2020. The Quality in Prognostic Studies (QUADAS-2) tool was used to assess methodological quality. The pooled sensitivity (Sen), specificity (Spe), positive or negative likelihood ratios (PLR or NLR), diagnostic odds ratio (DOR), curve, and area under the curve (AUC) were calculated with 95% confidence interval (95% CI). The overall accuracy (OA) of miRNAs, procalcitonin (PCT), and C-reactive protein (CRP) was analyzed by the chi-square test.

Results: A total of 22 records were eligible for systematic review, including 2210 sepsis, 426 systemic inflammatory response syndrome (SIRS), and 1076 healthy controls (HC). The pooled Sen, Spe, and DOR of miRNAs were 0.80 (95% CI 0.75–0.83), 0.85 (95% CI 0.80–0.89), and 22 (15–32), respectively. The DOR of PCT and CRP were 17 (95% CI 4–68) and 7 (95% CI 1–48), respectively. The OA value of miRNAs (79.02%) and PCT (76.95%) were higher than CRP (61.22%) (P < 0.000). The subgroup analysis indicated that miRNAs in adults, serum type, downregulation of miRNA expression, criteria of Sepsis-3, internal reference of non-U6, and dysregulation expression of miR-223 had superior diagnostic accuracy. In addition, there was no significant publication bias among the included studies. Fagan’s nomogram showed valuable clinical utility.

Conclusions: Our meta-analysis indicated that the level of circulating miRNAs, particularly the miR-223, could be used as an indicator for sepsis.

Keywords: Sepsis, SIRS, miRNAs, Diagnosis, Biomarkers

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**Introduction**

Sepsis is defined as a life-threatening condition of organ dysfunction resulted from a dysregulated host response to infection [1]. Recent epidemiologic data have shown an increasing incidence of sepsis and septic shock with high mortality [2]. Although recent data suggested a declining trend in mortality, longer-term morbidity and decreased health-related quality of life remain a serious problem [3]. Additionally, the clinical syndrome of sepsis is difficult to diagnose. A previous study reported 50% of patients with sepsis were not correctly classified in the USA [4]. Early diagnosis of sepsis is crucial for improving the survival rate; however, traditional screening tools and biomarkers lack specificity [5].

MicroRNAs (miRNAs) are small non-coding RNAs, associated with the presence and severity of sepsis. Dysregulation of several miRNAs, such as miR-146a, miR-223, miR-15a, miR-16, and miR-150, was found in the peripheral blood of sepsis patients [6-9]. Previous studies have demonstrated the expression of miRNAs in sepsis had specific compartment characteristics, and serum-derived exosome miRNAs were considered to be a significant diagnosis marker and survival prediction factor [10]. In addition, miRNAs had shown to be involved in the regulation of the exacerbated inflammation, endothelial dysfunction, and coagulation cascade in sepsis [11], miR-15a, miR-125b, and miR-146a have been shown to prevent NF-κB activation in sepsis by repressing TRAF6 and IRAK expression [10, 11]. However, the expression level of miRNAs showed differences in sepsis due to different population characteristics and normalization methodologies among different studies [11]. Therefore, a comprehensive and systematic meta-analysis of currently available data will facilitate the understanding of the diagnostic accuracy of these miRNAs in sepsis.

**Methods**

**Search strategy and selection criteria**

This meta-analysis was presented according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses statement (PRISMA) [12]. Six databases including PubMed, Cochrane Library, Embase, Web of Science, Scopus, and Ovid were searched by the end of April 3, 2020. The retrieval terms included “sepsis” or “Severe Sepsis” or “Pyemia” or “Septicemia” and “micro RNA” and “microRNAs” or “miRNA.”

Literatures were considered eligible for inclusion using the following criteria: (1) the target population consisted of one or more circulating miRNAs for sepsis; (2) sufficient data to generate true positive (Tp), true negative (Tn), false positive (Fp), and false negative (Fn) directly or indirectly; and (3) either retrospective or prospective design studies. Studies that are excluded in our meta-analysis met the following criteria: (1) reviews, letters, correspondence, expert opinions, and editorial; (2) animal or in vitro studies; (3) duplicated information; and (4) insufficient information regarding the diagnosis value.

**Data collection**

Two investigators (XMS and WY) independently screened the records retrieved from the search after deduplication by title and abstract. Full-text studies that satisfied all the inclusion criteria were further screened for eligibility by the same investigators. Any discrepancies were resolved by a third reviewer (JJZ). Data extraction was performed by two review authors (JJZ and YCH) independently, including author and year of publication, sample characteristics, diagnostic criteria, illness severity (APACHE II and SOFA score), and numbers of Tp, Tn, Fp, and Fn observations.

**Assessment of methodologic quality**

Quality Diagnostic Accuracy Studies-2 (QUADAS-2) was used to assess the methodological qualities of the included studies by independent authors (XMS and WY) [13]. Disagreements were resolved by a third reviewer (JPT).

**Data synthesis and statistical analysis**

The threshold effect was evaluated before data synthesis. The diagnostic index tests, including sensitivity (Sen), specificity (Spe), positive or negative likelihood ratios (PLR or NLR), diagnostic odds ratio (DOR), summary receiver operating characteristic (SROC), curve, and area under the curve (AUC), were quantified with 95% confidence interval (95% CI). Meta-analysis was performed using a bivariate random-effects model to estimate the summary diagnostic indexes. Heterogeneity was explored using meta-regression models, and subgroup analyses were further analyzed according to varied factors. Publication bias was evaluated by Deeks’ funnel plot, and clinical utility was evaluated by Fagan’s nomogram. STATA version 14 (STATA Corp, College Station, TX, USA) was used for all statistical analyses. The differences between the overall accuracy (OA), that is the proportion of Tp and Tn in all evaluated cases, of miRNAs, procalcitonin (PCT), and C-reactive protein (CRP) were analyzed by the chi-square test using SPSS Statistics 22 (IBM, China).

**Results**

**Search results and methodological qualities of included studies**

Our search yielded 2061 references through 6 electronic databases. After removing 1978 duplicates, animal or in vitro studies, irrelevant articles, and excluded article
format, a total of 83 full-text references were screened for eligibility. Finally, 22 records were included in our systematic review [6–9, 14–31]. The detailed flow diagram of the study selection process is shown in Fig. 1.

The methodological qualities of the included articles were evaluated according to the QUADAS-2 criteria (Supplementary Figure 1). Risk of bias is mainly derived from the index test. The conduct or interpretation of the index test introduced high risk in 6 records [7, 8, 17, 19, 23, 28] and unclear risk in 13 articles [6, 14–16, 18, 20, 21, 25–27, 29–31].

Characteristics of included studies
A total of 2337 patients, including 14 studies of systemic inflammatory response syndrome (SIRS), 2 studies of local infections, and 14 studies of healthy controls (HC), provided data regarding the diagnostic accuracy of miRNAs. The characteristics of the included studies and patients are presented in Table 1 and Supplemental Table 1. Single miRNAs were reported in all studies. Among the 29 studies, 8 studies were about the expression of miRNAs in children, and 21 studies were about adults. In addition, there were 15 studies of serum specimens and 13 studies of plasma specimens.

Threshold effect
The Spearman correlation coefficient for miRNAs was 0.127 (P = 0.503) using a rank correlation test. The shape of the ROC plot was not arm and shoulder shape (Supplementary Figure 2a). These results indicated the heterogeneity among the included studies was not caused by the threshold effect [32].

Diagnostic accuracy of miRNAs, procalcitonin, and C-reactive protein for sepsis
The random-effects model was used to synthesize the data due to significant heterogeneity among the 30 studies (I² = 86.49% for Sen, I² = 78.03% for Spe, I² = 100% for DOR). Outliers, falling outside 95% CI, could be the main source of heterogeneity [Supplementary Figure 2b].

The pooled Sen and Spe were 0.80 (95% CI 0.75–0.83) and 0.85 (95% CI 0.80–0.89), respectively (Fig. 2). The PLR and NLR were 5.3 (95% CI 4.0–6.9) and 0.24 (95% CI 0.20–0.29), respectively. The DOR of pooled studies was 22 (15–32), and the AUC for SROC was 0.89 (95% CI 0.86–0.92), indicating a high overall accuracy of circulating miRNAs for sepsis (Table 2 and Fig. 3a, b).

In all of these studies, 4 studies analyzed the diagnostic accuracy of PCT for sepsis; the pooled Sen and Spe were 0.73 (95% CI 0.66–0.79) and 0.86 (95% CI 0.61–0.96), respectively. The PLR and NLR were 5.2 (95% CI 1.6–16.8) and 0.31 (95% CI 0.23–0.43), respectively. The DOR of pooled studies was 17 (95% CI 4–68), and the AUC for SROC was 0.74 (95% CI 0.70–0.77) (Table 2).

Furthermore, 4 studies analyzed the diagnostic accuracy of CRP for sepsis; the pooled Sen and Spe were 0.77 (95% CI 0.73–0.81) and 0.71 (95% CI 0.29–0.94), respectively. The DOR of pooled studies was 7 (95% CI 1–48), and the AUC for SROC was 0.77 (95% CI 0.73–0.81) (Table 2).
| First author | Year | Population | Region | miRNAs | Regulation mode | Specimen | DC | Internal reference types in qRT-PCR | Sample size | Diagnostic power | Ref. |
|--------------|------|------------|--------|--------|-----------------|----------|----|------------------------------------|-------------|------------------|------|
| Wang JF      | 2010 | Adults     | China  | miR-146a | Downregulated   | Serum    | 1  | mmu-miR-295                      | Sepsis 50    | SIRS 30          | 63.3 | 100  | 0.804 | 10 |
| Wang JF      | 2010 | Adults     | China  | miR-223 | Downregulated   | Serum    | 1  | mmu-miR-295                      | Sepsis 50    | SIRS 30          | 80.0 | 100  | 0.858 | 10 |
| Wang H       | 2012 | Adults     | China  | miR-15a  | Downregulated   | Serum    | 1  | U6                                 | Sepsis 166   | SIRS 32          | 68.3 | 94.4 | 0.858 | 10 |
| MA Y         | 2013 | Adults     | UK     | miR-150  | Upregulated     | Whole blood | 2  | None                              | Sepsis 23    | SIRS 22          | 72.7 | 85.7 | 0.830 | 10 |
| MA Y         | 2013 | Adults     | UK     | miR-4772-5p | Downregulated  | Whole blood | 2  | None                              | Sepsis 23    | SIRS 22          | 68.2 | 71.4 | 0.760 | 10 |
| Wang L       | 2013 | NR         | China  | miR-146a | Downregulated   | Plasma   | NR | U6                                 | Sepsis 14    | SIRS 14          | 60.0 | 87.5 | 0.813 | 10 |
| Wu YH        | 2013 | Children   | China  | miR-223 | Upregulated     | Plasma   | 2  | cel-miR-39                        | Sepsis 35    | SIRS 20          | 82.9 | 85.0 | 0.919 | 14 |
| Wu YH        | 2013 | Children   | China  | miR-146a | Upregulated     | Plasma   | 2  | cel-miR-39                        | Sepsis 40    | SIRS 20          | 77.5 | 77.0 | 0.815 | 15 |
| Wu YH        | 2013 | Children   | China  | miR-223 | Upregulated     | Plasma   | 2  | cel-miR-39                        | Sepsis 40    | SIRS 20          | 77.5 | 55.0 | 0.678 | 15 |
| Liu CL       | 2015 | Children   | China  | miR-223 | Upregulated     | Plasma   | 2  | None                              | Sepsis 49    | HC 50            | 83.5 | 86.0 | 0.904 | 16 |
| Yang YL      | 2015 | Adults     | China  | miR-499 | Upregulated     | Plasma   | 2  | None                              | Sepsis 112   | HC 20            | 86.7 | 90.8 | 0.838 | 17 |
| Han Y        | 2016 | Adults     | China  | miR-143 | Upregulated     | Serum    | 1  | U6                                 | Sepsis 103   | SIRS 95          | 78.6 | 91.6 | 0.910 | 18 |
| Lau C        | 2016 | Adults     | China  | miR-155-5p | Upregulated  | Serum    | 3  | None                              | Sepsis 105   | HC 35            | 85.3 | 80.6 | 0.855 | 19 |
| Lan C        | 2016 | Adults     | China  | miR-133a-3p | Upregulated | Serum   | 3  | None                              | Sepsis 105   | HC 35            | 97.9 | 54.8 | 0.769 | 19 |
| Xu J         | 2017 | Adults     | China  | miR-195 | Upregulated     | Serum    | 1  | U6                                 | Sepsis 54    | SIRS 41          | 75.6 | 70.4 | 0.776 | 20 |
| Rahmel T     | 2018 | Adults     | Germany | miR-122 | Upregulated     | Serum    | 3  | cel-miR-54                        | Sepsis 108   | SIRS 20          | 58.3 | 95.0 | 0.760 | 21 |
| Wu XL        | 2018 | Adults     | China  | miR-223 | Upregulated     | Plasma   | 1  | U6                                 | Sepsis 187   | HC 186           | 56.6 | 86.6 | 0.754 | 22 |
| Abou El-Khier| 2018 | Adults     | Egypt  | miR-122 | Upregulated     | Serum    | NR | cel-miR-39                        | Sepsis 25    | LI 25            | 100  | 100  | 1.000 | 23 |
| Guo HL       | 2019 | Adults     | China  | miR-495 | Downregulated   | Serum    | 2  | U6                                 | Sepsis 105   | HC 100           | 89.5 | 83.0 | 0.915 | 24 |
| Karam RA     | 2019 | Children   | Egypt  | miR-146a | Downregulated   | Serum    | 2  | U6                                 | Sepsis 55    | HC 60            | 86.6 | 56.6 | 0.803 | 25 |
| Zhang WP     | 2019 | Adults     | China  | miR-7110-5p | Upregulated | Plasma   | 3  | miR-16                            | Sepsis 44    | HC 21            | 84.1 | 90.5 | 0.813 | 26 |
| Zhang WP     | 2019 | Adults     | China  | miR-223-3p | Upregulated  | Plasma   | 3  | miR-16                            | Sepsis 44    | HC 21            | 82.9 | 100  | 0.964 | 26 |
| Lin YJ       | 2019 | Adults     | China  | miR-210 | Upregulated     | Plasma   | 1  | None                              | Sepsis 125   | HC 110           | 81.0 | 80.9 | 0.852 | 27 |
| Lin YJ       | 2019 | Adults     | China  | miR-494 | Downregulated   | Plasma   | 1  | None                              | Sepsis 125   | HC 110           | 80.9 | 72.1 | 0.847 | 27 |
| Lin YJ       | 2019 | Adults     | China  | miR-205 | Downregulated   | Plasma   | 1  | None                              | Sepsis 125   | HC 110           | 78.6 | 90.5 | 0.86  | 27 |
| Zhu XP       | 2019 | Adults     | China  | miR-125b | Upregulated     | Plasma   | 2  | U6                                 | Sepsis 120   | HC 120           | 40.2 | 80.0 | 0.658 | 28 |
| Liu GZ       | 2020 | Children   | China  | miR-181a | Downregulated   | Serum    | 2  | U6                                 | Sepsis 102   | LI 50            | 83.3 | 84.0 | 0.893 | 29 |
| Rabab F      | 2020 | Children   | Egypt  | miR-101 | Upregulated     | Serum    | 2  | U6                                 | Sepsis 50    | SIRS 30          | 84.0 | 84.0 | 0.908 | 30 |
| Rabab F      | 2020 | Children   | Egypt  | miR-187 | Upregulated     | Serum    | 2  | U6                                 | Sepsis 50    | SIRS 30          | 72.0 | 76.0 | 0.789 | 30 |
| Xu HM        | 2020 | Adults     | China  | miR-19b-3p | Downregulated | Serum    | 2  | U6                                 | Sepsis 103   | HC 98            | 85.4 | 85.7 | 0.921 | 31 |

qRT-PCR Quantitative RT-PCR, DC Diagnostic criteria, SIRS Systemic inflammatory response syndrome, HC Healthy controls, LI Local infections, Sen Sensitivity, Spe Specificity, AUC Area under the curve, Ref. References, NR No report
The OA value of miRNAs (79.02%) and PCT (76.95%) were higher than CRP (61.22%) \( (P < 0.000) \). The OA value confirmed no significant difference in the diagnostic accuracy between miRNAs and PCT \( (P = 0.373) \) (Fig. 3c).

**Diagnostic accuracy of miR-223**

miR-223 was reported in 6 studies of collected researches. The pooled Sen, Spe, AUC, and DOR were 0.77 (95% CI 0.67–0.84), 0.91 (95% CI 0.73–0.97), 0.87 (95% CI 0.84–0.90), and 33 (95% CI 8–142), respectively (Table 2).

**Meta-regression and subgroup analysis**

The heterogeneity was explored using multiple univariable bivariate meta-regression models. The effect of each covariate on Sen was estimated separately from that on Spe (Fig. 4). The diagnostic criteria did not have an effect on miRNA Sen and Spe \( (P > 0.05) \), and the covariate of the population did not have an effect on Spe.

![Fig. 2 Forest plots of pooled sensitivity (Sen) and specificity (Spe) of circulating miRNAs for the diagnosis of sepsis. a Sen, b Spe](image)

**Table 2** Diagnostic accuracy of miRNAs, PCT, and CRP for sepsis

| Index   | Number of studies | Number of patients | Sen (95% CI) | Spe (95% CI) | AUC (95% CI) | OA (%) |
|---------|-------------------|--------------------|--------------|--------------|--------------|--------|
| miRNAs | 30                | 3914               | 0.80 (0.75–0.83) | 0.85 (0.80–0.89) | 0.89 (0.80–0.89) | 79.02  |
| PCT     | 4                 | 347                | 0.73 (0.66–0.79) | 0.86 (0.61–0.96) | 0.86 (0.40–0.69) | 76.95  |
| CRP     | 4                 | 245                | 0.77 (0.73–0.81) | 0.91 (0.29–0.94) | 0.83 (0.73–0.87) | 61.22  |
| miR-223 | 6                 | 732                | 0.77 (0.67–0.84) | 0.91 (0.29–0.94) | 0.83 (2.52–27.9) | 79.25  |

miRNAs: microRNAs, PCT: Procalcitonin, CRP: C-reactive protein, 95% CI: 95% confidence intervals, Sen: Sensitivity, Spe: Specificity, AUC: Area under the curve, OA: Overall accuracy.

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addition, the rest of the covariates were statistically significant for Sen and Spe.

Subgroup analyses of miRNAs were further performed due to obvious heterogeneity. As shown in Table 3, among studies examining only adult patients, the Sen and Spe of circulating miRNAs were 0.80 (0.73–0.85) and 0.88 (0.82–0.92), respectively. The DOR of studies using serum samples was higher than that of studies using plasma samples (32 versus 17). The Sen, Spe, and DOR of miRNAs in patients with sepsis reference to Sepsis-3 were higher than those in patients reference to Sepsis-1 or Sepsis-2. Non-U6, as an internal control to normalize the expression levels of miRNAs, showed superiority compared with U6. The pooled Sen, Spe, PLR, NLR, DOR, and AUC in the non-U6 group were 0.79 (0.70–0.86), 0.96 (0.82–0.99), 19.5 (3.9–79.8), 0.22 (0.15–0.32), 89 (15–518), and 0.90 (0.87–0.92), respectively. In the subgroup of control groups, the pooled Sen and Spe of miRNAs for sepsis were 0.74 (0.69–0.78) and 0.76 (0.76–0.88) versus SIRS and 0.83 (0.76–0.88) and 0.82 (0.76–0.87) versus HC, respectively.

Publication bias
A funnel graph was performed to assess the likelihood of publication bias. Deeks’ test indicated no potential publication bias in all the variables analyzed in this study (Supplementary Figure 2c).

Clinical utility of index test
Fagan’s nomogram was used to assess the post-test probabilities. When the pre-test probability was set at 20%, the post-test probability arrived at 57% with PLR of 5 and 6% with NLR of 0.24 (Supplementary Figure 2d).

Discussion
Sepsis is a significant public health problem, with high mortality and long-term morbidity [3]. Good evidence of a mortality benefit in the early diagnosis and treatment of sepsis and septic shock [5]. However, it is not always easy to distinguish sepsis and SIRS in the early stage and is even sometimes impossible. A number of studies have investigated miRNAs as efficient biomarkers in various
phases of sepsis [11]. Therefore, the present study comprehensively evaluated the diagnostic accuracy of circulating miRNAs in sepsis patients via a systematic review and meta-analysis.

In this study, we found the Sen, Spe, and DOR of circulating miRNAs for diagnosis of sepsis were higher than PCT and CRP. The OA value of miRNAs and PCT were significantly higher than CRP, while no significant difference was found between miRNAs and CRP. The results of the subgroup analysis showed adults and patients diagnosed by reference to Sepsis-3 had a higher value of DOR. In addition, improvement of miRNA accuracy for the detection of sepsis was observed in the subgroup of serum type, internal reference of non-U6, and downregulation of miRNA expression.

Although different screening tools and biomarkers, such as white cell count, neutrophil count, interleukin 6 (IL-6), CRP, and PCT, have been used for sepsis diagnosis, none of them is proven to be specific [5]. Previous studies demonstrated CRP lack specificity for sepsis due to an increase in non-infectious inflammatory conditions [33]. The levels of PCT could help to guide the antibiotic therapy in patients with sepsis, while the heterogeneous data of PCT failed to provide guidance for the benefits of sepsis patients [34, 35]. Current literature suggested miRNAs played a vital role in the pathophysiology of sepsis and could distinguish the various phases of sepsis [6–9, 11, 14–31]. Several miRNAs, such as miR-146-a, miR-125b, and miR-223, were positively or negatively associated with sepsis severity [22, 25, 28]. In this present study, the AUC of miRNAs was 0.89 (0.86–0.92), which is higher than that of PCT [0.74 (0.70–0.77)] and CRP [0.77 (0.73–0.81)]. However, the OA value showed no significant difference between miRNA and PCT. Notably, a number of miRNAs have proved to be not only as diagnostic markers (miR-15a, miR-16, miR-223, miR-499-5p) but also as prognostic markers (miR-193b, miR-483-5p, and miR-574-5p) for sepsis [6–9, 36]. These results indicated circulating miRNAs could have the ability to monitor the progress of sepsis.

**Fig. 4 Meta-regression and subgroup analysis**

| Unvariable Meta-regression & Subgroup Analyses |
|-----------------------------------------------|
| Sen  | Spe  | DOR |
| Adult Yes | No | No | No | No | No |
| China Yes | No | No | No | No | No |
| Upregulated Yes | No | No | No | No | No |
| Serum Yes | No | No | No | No | No |
| Sepsis3 Yes | No | No | No | No | No |
| U6 Yes | No | No | No | No | No |
| SIRS Yes | No | No | No | No | No |
| Size100 Yes | No | No | No | No | No |

| Sensitivity (95% CI) |
|----------------------|
| 0.67 (0.59–0.75) |
| 0.93 (0.87–0.98) |

| Specificity (95% CI) |
|----------------------|
| 0.69 (0.60–0.78) |
| 0.97 (0.93–1.00) |

*p<0.05, **p<0.01, ***p<0.001
Despite the efficacy of miRNAs, there was obvious heterogeneity in this meta-analysis. Patients diagnosed by reference to Sepsis-3 had higher Sen, Spe, and DOR of miRNAs than that of patients reference to Sepsis-1 or Sepsis-2. As a deeper understanding of the biology of sepsis, the criteria of Sepsis-3 introduced a framework based on susceptibility, pathogen, dysregulated host response, and organ dysfunction [35]. The different specimen types could have impacted the conflicting results. Our results found the accuracy of miRNAs for sepsis diagnosis in the serum samples was higher than that in the plasma. Contrastingly, other disease studies reported the expression of miRNAs in the plasma was higher than the serum due to more proteins in the plasma [37]. The same may be true of internal reference types. However, there has been no unified standard for

| Subgroups                  | Number of studies | Cases number of sepsis | Cases number of controls | Sen (95% CI) | Spe (95% CI) | PLR (95% CI) | NLR (95% CI) | DOR (95% CI) | AUC (95% CI) |
|----------------------------|-------------------|------------------------|--------------------------|--------------|--------------|--------------|--------------|--------------|--------------|
| Population                 |                   |                        |                          |              |              |              |              |              |              |
| Adults                     | 21                | 1902                   | 1283                     | 0.80 (0.73–0.85) | 0.88 (0.82–0.92) | 6.4 (4.4–9.4) | 6.4 (4.4–9.4) | 28 (17–46)  | 0.91 (0.88–0.93) |
| Children                   | 8                 | 421                    | 280                      | 0.81 (0.77–0.85) | 0.77 (0.67–0.84) | 3.47 (2.4–5.0) | 0.24 (0.19–0.30) | 14 (8–24)   | 0.82 (0.79–0.86) |
| Region                     |                   |                        |                          |              |              |              |              |              |              |
| China                      | 23                | 2003                   | 1368                     | 0.80 (0.75–0.84) | 0.85 (0.80–0.89) | 5.4 (4.0–7.1) | 0.24 (0.19–0.30) | 23 (15–33)  | 0.89 (0.86–0.92) |
| Non-China                  | 7                 | 334                    | 209                      | 0.80 (0.67–0.89) | 0.84 (0.69–0.93) | 5.1 (2.4–11.0) | 0.24 (0.13–0.43) | 21 (7–71)   | 0.89 (0.86–0.91) |
| Regulation mode            |                   |                        |                          |              |              |              |              |              |              |
| Upregulated                | 20                | 1544                   | 1031                     | 0.80 (0.74–0.85) | 0.83 (0.78–0.87) | 4.8 (3.5–6.4) | 0.24 (0.18–0.32) | 20 (13–32)  | 0.89 (0.86–0.91) |
| Downregulated              | 10                | 793                    | 546                      | 0.80 (0.73–0.85) | 0.89 (0.80–0.94) | 7.3 (4.1–13.1) | 0.23 (0.18–0.29) | 32 (18–56)  | 0.89 (0.86–0.91) |
| Specimen                   |                   |                        |                          |              |              |              |              |              |              |
| Serum                      | 15                | 1231                   | 711                      | 0.82 (0.76–0.88) | 0.87 (0.79–0.93) | 6.4 (3.9–10.5) | 0.20 (0.15–0.27) | 32 (19–54)  | 0.91 (0.88–0.93) |
| Plasm                      | 13                | 1060                   | 822                      | 0.77 (0.70–0.82) | 0.84 (0.78–0.88) | 4.7 (3.4–6.6) | 0.28 (0.21–0.37) | 17 (10–29)  | 0.88 (0.84–0.90) |
| Diagnostic criteria        |                   |                        |                          |              |              |              |              |              |              |
| Sepsis-3                   | 5                 | 406                    | 132                      | 0.85 (0.69–0.94) | 0.87 (0.69–0.95) | 6.8 (2.8–16.1) | 0.17 (0.08–0.35) | 40 (20–81)  | 0.93 (0.90–0.95) |
| Sepsis-1/2                 | 23                | 1892                   | 1406                     | 0.78 (0.73–0.82) | 0.84 (0.79–0.88) | 4.8 (3.7–6.2) | 0.26 (0.22–0.32) | 18 (13–26)  | 0.87 (0.84–0.90) |
| Internal reference types   |                   |                        |                          |              |              |              |              |              |              |
| U6                         | 12                | 1109                   | 856                      | 0.76 (0.68–0.83) | 0.83 (0.77–0.87) | 4.4 (3.3–5.8) | 0.29 (0.21–0.39) | 15 (9–24)   | 0.87 (0.84–0.89) |
| Non-U6                     | 9                 | 436                    | 207                      | 0.79 (0.70–0.86) | 0.96 (0.82–0.99) | 19.5 (3.9–97.8) | 0.22 (0.15–0.32) | 89 (15–518) | 0.90 (0.87–0.92) |
| Control groups             |                   |                        |                          |              |              |              |              |              |              |
| SIRS                       | 14                | 806                    | 426                      | 0.74 (0.69–0.78) | 0.87 (0.79–0.92) | 5.7 (3.5–9.4) | 0.30 (0.26–0.36) | 19 (11–33)  | 0.82 (0.78–0.85) |
| HC                         | 14                | 1404                   | 1076                     | 0.83 (0.76–0.88) | 0.82 (0.76–0.87) | 4.6 (3.4–6.1) | 0.21 (0.15–0.30) | 22 (14–35)  | 0.89 (0.86–0.91) |
| Sample size                |                   |                        |                          |              |              |              |              |              |              |
| > 100                      | 15                | 1746                   | 1181                     | 0.81 (0.73–0.87) | 0.83 (0.77–0.87) | 4.7 (3.6–6.2) | 0.23 (0.17–0.33) | 20 (13–30)  | 0.89 (0.86–0.91) |
| < 100                      | 15                | 591                    | 396                      | 0.78 (0.74–0.82) | 0.88 (0.79–0.94) | 6.7 (3.7–12.2) | 0.24 (0.20–0.30) | 27 (13–58)  | 0.85 (0.82–0.88) |

miRNAs MicroRNAs, 95% CI 95% confidence intervals, Sen Sensitivity, Spe Specificity, PLR Positive likelihood ratios, NLR Negative likelihood ratios, DOR Diagnostic odds ratio, AUC Area under the curve, SIRS Systemic inflammatory response syndrome, HC Healthy controls
the normalization of miRNA. Thus, multicenter standardization researches need to be explored for better elucidation of miRNAs as a promising biomarker in sepsis.

Our results showed dysregulation expression of miR-223 showed a superior diagnostic achievement. miR-223, as a pro-inflammatory factor, is involved in several signaling pathways that control inflammatory responses and infection reaction activation, such as negative regulation of STAT3 and IL-6 expression during sepsis [38–40]. Wu et al. [22] showed miRNA-223 expression was positively correlated with APACHE II score as well. Of note, Wang et al. [6] suggested that miR-223 in serum was downregulated among sepsis patients, while other five studies demonstrated miR-223 in the plasm was upregulated [14–16, 22, 26]. In fact, the conflicting results of miR-223 were also reported in hepatitis and hepatocellular carcinoma [39]. The possible explanations were that the expression of miR-223 was different among different sepsis stages and different experimental methods. However, the sample sizes for the evaluation of miR-233 were not large enough. Therefore, more additional researches are needed to assess the benefit of miR-233 during the course of sepsis.

Despite miRNAs as a promising diagnostic biomarker for sepsis, there are still several potential limitations. First, heterogeneity was observed in our meta-analysis. However, there was no significant threshold effect. Further subgroup analyses improved the heterogeneity. Second, miRNAs in the included patients were confirmed after sepsis diagnosis by the clinic criteria, which may add population selection bias. In addition, the included studies did not evaluate the correlation between miRNA expression and illness severity in detail. Furthermore, we could not control the statistical methods among the included studies, which may influence the results of the meta-analysis.

Conclusions

In conclusion, circulating miRNAs, especially for miR-223, are potential markers for distinguishing sepsis from SIRS and HC. Better results could be obtained for adults and patients diagnosed by reference to Sepsis-3. Further large and well-designed studies should be explored to identify the role of promising miRNAs in sepsis.

Supplementary Information

Supplementary information accompanies this paper at https://doi.org/10.1186/s40560-020-00497-6.

Additional file 1: Supplementary Figure 1. A summary of methodological qualities included articles using the QUADAS-2 criteria.

Additional file 2: Supplementary Figure 2. The ROC plot, Deeks’ funnel plot, and Fagan’s Nomogram of the diagnostic meta-analysis. (a) ROC plot; (b) Galbraith; (c) Deeks’ funnel plot; (d) Fagan’s Nomogram.

Additional file 3: Supplemental Table 1. The research type and severity characteristics of 22 included studies

Abbreviations

miRNAs: MicroRNAs; QUADAS-2: Quality in Prognostic Studies; Sen: Sensitivity; Spe: Specificity; PLR: Positive likelihood ratios; NLR: Negative likelihood ratios; DOR: Diagnostic odds ratio; AUC: Area under the curve; OA: Overall accuracy; SROC: Summary receiver operating characteristic; 95% CI: 95% confidence intervals; PCT: Procalcitonin; CRP: C-reactive protein; SIRS: Systemic inflammatory response syndrome; HC: Healthy controls; IL-6: Interleukin 6

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Authors’ contributions

The work presented here was carried out in collaboration between all authors. WY and YQQ developed the concept and designed the study. XMS and WY carried out the literature research and study selection. JJZ and YCH co-worked on the associated data collection. The qualities of the included studies were carried out by XMS and WY. Disagreements were resolved by JPT and LZ. Data synthesis and analysis were carried out by WY, JPT, and ZZ. The manuscript was written by WY and corrected by XMS and JJZ. All authors discussed the results and implications and commented on the manuscript at all stages. The authors read and approved the final manuscript.

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Availability of data and materials

All data generated or analyzed during this study are included in this published article and its supplementary information files.

Ethics approval and consent to participate

Not applicable.

Consent for publication

All authors have seen and approved the content and fulfill the journal’s requirements for authorship.

Competing interests

The authors declare that they have no competing interests.

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