Original Article

Biochemical and histological changes in the heart of post-partum rats exposed to Natron

Yusuf Saidu\textsuperscript{a}, Maimuna Jumai Usman\textsuperscript{a, *}, Suleiman Ahmed Isa\textsuperscript{a}, Simeon Alabi Isezuo\textsuperscript{b}, Lawal Suleiman Bilbis\textsuperscript{a}, Saddiku Malam Sahabi\textsuperscript{c}, Ahmad Bello\textsuperscript{d}, Suleiman Alhaji Muhammad\textsuperscript{a}

\textsuperscript{a} Department of Biochemistry, Usman Danfodiyo University, P.M.B. 2346, Sokoto, Nigeria
\textsuperscript{b} Department of Medicine, Usman Danfodiyo University, P.M.B. 2346, Sokoto, Nigeria
\textsuperscript{c} Department of Histopathology, Usman Danfodiyo University, P.M.B. 2346, Sokoto, Nigeria
\textsuperscript{d} Department of Biochemistry and Molecular Biology, Federal University, Birnin Kebbi, Nigeria

\textbf{A R T I C L E  I N F O}

Article history:
Received 2 August 2017
Accepted 9 December 2017
Available online 11 December 2017

Keywords:
Peripartum cardiomyopathy
Natron
Myocyte hypertrophy
Postpartum
Wistar rat

\textbf{A B S T R A C T}

\textit{Background:} The customary puerperal practice of Natron consumption has been identified as one of the predisposing factors in the etiology of peripartum cardiomyopathy (PPCM). This study was designed to investigate the effect of Natron in postpartum Wistar albino rats.

\textit{Methods:} A total of 30 postpartum Wistar rats were exposed to different doses (50 mg/kg, 100 mg/kg, 200 mg/kg and 300 mg/kg) of Natron for 28 days. After the treatment, we carried out biochemical analyses and histological evaluations of kidney, liver and heart.

\textit{Results:} The study revealed that the exposure of postpartum rats to 100 mg/kg of Natron and above significantly (\(p < 0.05\)) increases the cardiac markers; myoglobin, creatine kinase-MB, troponin I and T as compared with control. The result of liver function indicated no significant difference in alanine aminotransferase, aspartate aminotransferase, gamma-glutamyltransferase, albumin and total protein of the Natron treated groups as compared with control. However, at higher doses, the levels of total protein, globulin and alkaline phosphatase activity were significantly increased in comparison to the control. There was no significant difference in the kidney function markers of the treatment groups as compared with control. Histological examinations revealed no changes in the kidney of the treated groups. Mild portal triaditis was observed in the liver of the treated rats. The heart of the rats administered \(\geq 100\) mg/kg of Natron showed myocyte hypertrophy.

\textit{Conclusion:} The study demonstrated that the administration of Natron for 28 days caused changes in the heart of postpartum rats and thus may contribute to the pathogenesis of PPCM.

© 2017 Published by Elsevier B.V. on behalf of Cardiological Society of India. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

1. Introduction

Peripartum cardiomyopathy (PPCM) is an infrequent form of dilated cardiomyopathy that causes heart failure occurring typically between the last month of pregnancy and up to five months after giving birth.\textsuperscript{1} The disease is characterized by systolic dysfunction of the heart with a left ventricular ejection fraction of less than 0.45\textsuperscript{2} which could increase the risk of atrial and ventricular arrhythmias, blockage of blood vessels by clot as well as cardiac death. As a result, the heart muscle cannot contract vehemently enough to supply adequate amounts of blood needed by the vital organs of the body.\textsuperscript{3,4} The exact incidence of PPCM is still unknown worldwide, but the accepted incidence is 1 per 3000–4000 births which are more common in West Africa\textsuperscript{5} and Haiti.\textsuperscript{6} The disease is common among the Hausa tribe in Nigeria, which accounts for about 13% of all female admissions.\textsuperscript{7} Studies have shown that the incidence of PPCM varies between 1 in 100 to 1 in 15,000 live births, and this could be ascribed to divergent definitions of PPCM and referral pattern in different nations.\textsuperscript{8,9,10} Unique customary puerperal practices such as postpartum consumption of excess Natron and daily hot water bath for 40 days\textsuperscript{11} by the Hausa and Fulani women in northern Nigeria have been hypothesized as contributing factors to the incidence of PPCM in Nigeria. Isezuo and Abubakar\textsuperscript{12} reported that these
practices are still common in Sokoto and have been suggested to cause volume overload and heart failure. These puerperal practices could cause the initial accumulation of fluid due to the increase of the peripheral resistance. As a result, the volume expansion overburdens the dilated heart leading to myocardial damage.\(^\text{12}\)

Natron (known as kanwa) is a hydrated sodium carbonate with water of crystallization, which contains traces of sodium chloride and silicon dioxide.\(^\text{13}\) Different varieties exist which differ in composition with deference to their locations.\(^\text{14}\) It is also widely used in Nigeria as a cooking ingredient in soups. Natron has been reported for the treatment of various diseases such as skin diseases, reproductive disorders, and stomach pain.\(^\text{15}\) Therefore, the study was designed to investigate the hypothesis that consumption of Natron as a puerperal practice is a predisposing factor in the pathogenesis of PPCM using Wistar albino rats.

2. Materials and methods

2.1. Chemicals and reagents

Analytical graded chemicals were used for the study. Assay kits were obtained from Agappe Diagnostics Ltd, India and WKEA Med Supplies, China.

2.2. Natron

The Natron was sourced from Sokoto Central Market, Nigeria, pulverized into a fine powder using pestle and mortar, sieved and stored in a clean airtight container until required. It was identified and authenticated at the Department of Chemistry, Usmanu Danfodiyo University, Sokoto, Nigeria. Appropriate measurement of different concentrations per kg body weight was prepared daily for the administration.

2.3. Experimental animals

This study was conducted in accordance with the standard set for the Care and Use of Laboratory Animals. The protocol was approved by the Ethics Committee on Animal Use of the Usmanu Danfodiyo University, Sokoto, Nigeria. The rats were purchased from the Nigerian Institute for Trypanosomiasis Research, Kaduna, Nigeria. The animals were kept in cages under supervision in the animal house with environment temperature and controlled light (21 ± 2°C, 12 h cycles of light/dark). They were allowed free access to feed and tap water ad libitum before and during the experimental period. The rats were grown into maturity and co-habited with male rats which were separated after the female rats got pregnant. The rats were randomly grouped after delivery.

2.4. Experimental protocol

Thirty postpartum rats (weighing between 160 and 180 g) were used for the study. The animals were randomly divided into 5 groups of 6 rats each and were fed pelleted growers' feed (Vital Feeds, Jos, Nigeria). Group I served as control and was given distilled water, group II, III, IV and V were administered 50 mg/kg, 100 mg/kg, 200 mg/kg and 300 mg/kg, respectively, of Natron solution by oral gavage using cannula for 4 weeks. At the end of the experimental period, the animals were anesthetized and blood sample was collected for biochemical assays. The rats were then euthanized; the heart, kidney and liver were collected for histological assessment. The blood samples were allowed to clot and centrifuged at 4000 rpm for 5 min and serum was then collected using a Pasteur pipette for biochemical assays.

2.5. Cardiac function biomarkers

The activities of lactate dehydrogenase (LDH), creatine kinase-MB (CK-MB), creatine kinase-NAC (CK-NAC), myoglobin, troponin T and I levels were measured using commercial kits according to the manufacturer’s instructions. CK-MB, LDH and CK-NAC kits were purchased from Agappe Diagnostics Ltd, India. Myoglobin, troponin T and I ELISA kits were products of WKEA Med Supplies, China. Microplate reader (RT6000, Germany) was used for the assay.

2.6. Liver function biomarkers

Serum bilirubin\(^\text{16}\), alanine transaminases (ALT) and aspartate transaminases (AST)\(^\text{17}\), alkaline phosphatase (ALP)\(^\text{18}\), total Protein\(^\text{19}\) and albumin\(^\text{20}\) were estimated as described previously. The method of Szasz\(^\text{21}\) was used for the estimation of gamma-glutamyltransferase (GGT). The parameters were assayed using Semi-auto Chemistry Analyser (RT9200, China).

2.7. Kidney function biomarkers

Serum Urea was estimated according to the method of Young\(^\text{22}\) while the method of Bartels and Bohmer\(^\text{23}\) was used for the estimation of serum creatinine by Semi-auto Chemistry Analyser (RT9200, China). Flame photometer (Jenway PPP7, UK) was used for the estimation of sodium and potassium.\(^\text{24}\) The method relied on the principle of Atomic Emission Spectrophotometry, which the intensity of atomization is measured with a photocell in a selected wavelength range corresponding to the given element. Chloride ion and bicarbonate were determined as described previously.\(^\text{25,26}\)

2.8. Tissue processing for histopathologic study

The organs (heart, kidney, and liver) of the rats were harvested and preserved in 10% formalin for histologic analysis. The organs were fixed in 10% buffered formalin for 72 h. The tissues were then dehydrated in alcohol of graded concentrations and embedded in paraffin. Embedded tissues were cut into sections of 5 μm thick and stained with hematoxylin and eosin for microscopic assessment. Photomicrographs of the samples were then taken.

2.9. Data analysis

The results are expressed as the mean ± standard error of the mean (SEM) with six rats per group. Data were analyzed using SPSS version 20. The differences between the five groups were compared using one way ANOVA, followed by Turkey multiple comparison tests. p < 0.05 was used as statistically significant.

3. Results

The result of the effect of Natron on cardiac function markers of postpartum rats is presented in Table 1. The result indicated that Natron significantly (p < 0.05) increase the activities of CK-MB and myoglobin, troponin I and troponin T levels at higher doses of the Natron as compared with control. There was no significant difference in the activities of CK-MB and troponin I and T levels at 50 mg/kg of Natron but the myoglobin level was significantly higher in comparison to the control. The activities of lactate dehydrogenase in the Natron exposed groups at a dose of 100 mg/kg and above were higher than the control.
Table 1
Effect of Natron on Cardiac Function Markers of Postpartum Rats.

| Biomarkers | Groups          | I Control  | II 50 mg/kg | III 100 mg/kg | IV 200 mg/kg | V 300 mg/kg |
|------------|-----------------|------------|-------------|---------------|--------------|-------------|
| LDH (U/L)  |                 | 10.88 ± 0.89a | 10.77 ± 0.90a | 11.27 ± 1.44a | 12.07 ± 0.70a | 14.27 ± 1.37a |
| Myoglobin (ng/mL) |     | 0.92 ± 0.09a | 1.32 ± 0.04a | 1.77 ± 0.08a  | 1.96 ± 0.11a  | 3.14 ± 0.13a  |
| CK-MB (ng/mL) |   | 6.60 ± 0.54a  | 8.39 ± 1.47a  | 13.61 ± 0.44b | 12.49 ± 0.69b | 11.74 ± 0.29b |
| CK-NAC (ng/mL) | | 32.41 ± 2.25a | 33.80 ± 4.91a | 45.06 ± 2.84b | 43.43 ± 3.35b | 47.55 ± 1.86b |
| TnI (ng/mL) |                   | 1.42 ± 0.03a | 1.45 ± 0.06a | 1.50 ± 0.09b  | 1.70 ± 0.07b  | 1.77 ± 0.07b  |
| TnT (µg/L) |                 | 25.75 ± 0.82a | 25.55 ± 0.90a | 28.68 ± 0.14a | 30.87 ± 0.36a | 34.99 ± 0.75a |

Values are expressed as mean ± Standard Error of Mean (n = 6). Mean value having different superscript letters in rows are significantly different (p < 0.05).

LDH = Lactate Dehydrogenase; CK-MB = Creatinine Kinase (Isoform); CK-NAC = Creatinine Kinase (N- Acetyl Cysteine); TnI = Troponin I; TnT = Troponin T.

Table 2
Effect of Natron on Kidney Function Parameters of Postpartum Rats.

| Parameters  | Groups          | I Control  | II 50 mg/kg | III 100 mg/kg | IV 200 mg/kg | V 300 mg/kg |
|-------------|-----------------|------------|-------------|---------------|--------------|-------------|
| Urea (mg/dL)|                 | 5.57 ± 0.36 | 4.75 ± 0.42 | 5.23 ± 0.49   | 4.35 ± 0.25  | 5.31 ± 0.54  |
| Creatinine (mg/dL) |   | 1.03 ± 0.43  | 0.98 ± 0.23 | 1.08 ± 0.08   | 1.09 ± 0.05  | 1.21 ± 0.04  |
| Na⁺ (mMol/L) |       | 133.50 ± 0.56 | 133.83 ± 0.94 | 132.67 ± 1.65 | 133.83 ± 1.74 | 133.73 ± 1.19 |
| K⁺ (mMol/L) |                 | 4.02 ± 0.13  | 4.12 ± 0.19  | 3.92 ± 0.09   | 4.12 ± 0.02  | 4.12 ± 0.18  |
| Cl⁻ (mMol/L) |               | 94.67 ± 1.09 | 94.17 ± 1.01 | 94.83 ± 1.19  | 96.00 ± 1.71 | 96.00 ± 0.68 |
| HCO₃⁻ (mMol/L) |     | 25.33 ± 1.33  | 23.50 ± 0.85  | 24.50 ± 1.20  | 23.17 ± 0.70 | 23.67 ± 0.76 |

Values are expressed as mean ± Standard Error of Mean (n = 6). Mean values of all the parameters are not statistically different as compared with control at p < 0.05. Na⁺ = sodium ion, K⁺ = potassium ion, Cl⁻ = chloride ion, HCO₃⁻ = bicarbonate.

The result of the effect of Natron on kidney function parameters of postpartum rats is presented in Table 2. The result showed no significant difference in urea, creatinine, and electrolytes of the exposed groups as compared with control.

The result of the effect of Natron on liver function biomarkers of postpartum rats is presented in Table 3. The result indicated no significant difference was observed in AST, ALT, Albumin, GGT and total bilirubin of treatment groups as compared with control. The treatments with ≥100 mg/kg significantly (p < 0.05) increase the activity of ALP. The levels of total protein and globulin were also increased at high doses of Natron.

The results of the histology of the heart, liver, and kidney are presented in Figs. 1–3, respectively. The heart of the rats exposed to ≥100 mg/kg body weight of Natron showed myocyte hypertrophy. There was no effect of the administered substance on the kidney of the postpartum rats. However, mild to moderate portal triaditis was observed in the liver, which was more pronounced in the group administered 200 mg/kg of Natron.

4. Discussion
Peripartum cardiomyopathy is uncommon, potentially life-threatening disease, which presents with symptoms of heart failure mainly due to left ventricular systolic dysfunction.22 This occurs in women of childbearing age with no preexisting heart disease during the last month of pregnancy or in the first five months postpartum.3 This in study, the effect of Natron at different doses was investigated in postpartum rats after 28 days treatment. Exposure of postpartum rats to the Natron increased myoglobin, troponin I and T levels and creatine kinase-MB and creatine kinase-NAC activities. These markers, especially cardiac troponins are intricate intracellular contractile-regulating proteins of the heart muscles that are important for calcium-mediated regulation of heart muscle contraction.28 Elevated levels of troponin I and T could reflect a direct effect of Natron that subsequently leads to myocyte hypertrophy observed in our study as shown by the results of heart histology. The increase activities of CK release from the myocardium may be associated with

Table 3
Effect of Natron on Liver Function Biomarkers of Postpartum Rats.

| Biomarkers | Groups          | I Control  | II 50 mg/kg | III 100 mg/kg | IV 200 mg/kg | V 300 mg/kg |
|------------|-----------------|------------|-------------|---------------|--------------|-------------|
| AST (U/L)  |                 | 191.11 ± 1.15a | 190.81 ± 0.79a | 193.03 ± 0.78a | 192.16 ± 1.00a | 193.27 ± 1.54a |
| ALT (U/L)  |                 | 170.65 ± 1.68a | 170.17 ± 1.16a | 170.19 ± 2.21a | 171.07 ± 2.41a | 169.32 ± 1.57a |
| ALP (U/L)  |                 | 535.65 ± 1.46a | 535.56 ± 3.88a | 566.79 ± 2.99a | 564.66 ± 1.74a | 569.16 ± 3.36a |
| TP (g/dL)  |                 | 6.16 ± 0.43a  | 6.36 ± 0.26a  | 7.84 ± 0.23a  | 7.05 ± 0.28a  | 7.56 ± 0.23a  |
| ALB (g/dL) |                 | 2.64 ± 0.06a  | 2.62 ± 0.07a  | 2.71 ± 0.04a  | 2.57 ± 0.06a  | 2.67 ± 0.05a  |
| GLO (g/dL) |                 | 3.53 ± 0.41a  | 3.74 ± 0.22a  | 5.13 ± 0.24a  | 4.48 ± 0.31a  | 4.90 ± 0.27a  |
| TBIL (mg/dL)|               | 0.35 ± 0.04a  | 0.32 ± 0.08a  | 0.31 ± 0.06a  | 0.23 ± 0.06a  | 0.22 ± 0.03a  |
| GGT (U/L)  |                 | 0.23 ± 0.12a  | 0.03 ± 0.01a  | 0.30 ± 0.20a  | 0.17 ± 0.11a  | 0.34 ± 0.16a  |

Values are expressed as mean ± Standard Error of Mean (n = 6). Mean Value Having Different Superscript Letters in rows are significantly different (p < 0.05).

ALT = Alanine Aminotransferase; AST = Aspartate Aminotransferase; ALP = Alkaline Phosphatase; TP = Total Protein; ALB = Albumin; GLO = Globulin; TBIL = Total Bilirubin; GGT = γ-Glutamyl Transferase.
membrane damage or left ventricular hypertrophy, which could be secondary to inflammatory responses due to Natron assault. The progressive loss of myocyte through apoptotic or necrotic cell death has been recognized as a key mechanistic pathophysiology in the progression of heart dysfunction in dilated cardiomyopathy and heart failure. The finding of this study is consistent with our earlier study of which administration of dried lake salt (kanwa) to postpartum rats caused cardiac chamber dilation, hypertrophy and focal atrophy. High circulating cardiac troponin I (cTnI) have been shown to be associated with larger left ventricular, left atrial and right ventricular diameters which correlated with higher mortality rates in idiopathic dilated cardiomyopathy, demonstrating a link between cTnI levels and cardiac remodeling. PPCM shared similar clinical characteristics as dilated cardiomyopathy, such as ventricle dilation and systolic dysfunction, but the former is thought to be dilated cardiomyopathy activated by pregnancy. During pregnancy, physiological changes due to the hemodynamic load caused by the rise in the blood volume bring about physiological hypertrophy. Consequently, the chamber dimensions would increase blood volume with a compensatory increase in wall thickness to handle the increased hemodynamic load and to ease cardiac output. These physiological changes in PPCM differ because an increase in chamber dimension does not increase the wall thickness proportionately in response to increased hemodynamic load. Therefore, it is evident from this study that Natron could cause volume overload without a compensatory increase in wall thickness and couples with oxidative stress is probably the Fig. 1. Photomicrograph of the Heart. (a) and (b) control and group administered 50 mg/kg of Natron showing normal histologic structures of heart cells and connective tissues. (c), (d) and (e) are groups administered 100, 200 and 300 mg/kg of Natron, respectively with arrows showing myocyte hypertrophy. Hematoxylin & Eosin ×20. CT- connective tissue, CN-cell nucleus.
mechanism underlying the changes observed in the Natron treated rats. Increased oxidative stress levels have been reported in both animal models and human PPCM patients.34

The measurement of activities of various enzymes in the tissues and body fluids plays a significant role in disease investigation and diagnosis.35 ALT, AST, and ALP are biomarkers used to predict hepatic injury. Although ALT is a more specific marker of liver damage. The serum levels of ALT, AST, and ALP are usually elevated in conditions associated with injuries or diseases affecting the liver, which leads to the release of these hepatocellular enzymes into the bloodstream.36 The significant increase in total protein, globulin and ALP activities due to Natron might be due to mild portal triaditis observed in the liver as revealed by the result of histology. Furthermore, increased in ALP activities could also be from the circulating placental ALP isoform or the tissue non-specific isoform. The histological analysis of the kidneys revealed no changes due to Natron treatment as glomeruli and renal tubule were normal just as the control. Treatment of postpartum rats with Natron caused significant alterations in the heart as evidenced by the results of biochemical and histological analyses.

5. Conclusion

The result of this study revealed that the exposure of postpartum rats to Natron for 28 days caused myocyte hypertrophy and changes in the heart biomarkers. It can be concluded that
consumption of Natron at a higher dose may be a predisposing factor in the pathogenesis of PPCM.

Conflict of interest

The authors have not declared any conflict of interest.

Acknowledgement

The authors thank TETFUND and Federal University, Birnin Kebbi, for providing the grant used for this work.

References

1. Sliwa K, Fett J, Elkayam U. Peripartum cardiomyopathy. Lancet. 2006;368:687–693.
2. Hibbard JU, Lindheimer M, Lang RM. A modified definition for peripartum cardiomyopathy and prognosis based on echocardiography. Obstet Gynecol. 1999;94:311–316.
3. Elkayam U, Akhter MW, Singh HS, et al. Pregnancy-associated cardiomyopathy: clinical characteristics and a comparison between early and late presentation. Circulation. 2005;111:2050–2055.
4. Murali S, Baldisseri MR. Peripartum cardiomyopathy. Crit Care Med. 2005;33:340–346.
5. Pearson GD, Veille JC, Rahimtoola S, et al. Peripartum cardiomyopathy: national heart, lung, and blood institute and office of rare diseases (National Institutes of Health) workshop recommendations and review. J Am Med Assoc. 2000;28:1183–1188.
6. Ferriere M, Saraz E, Bouhour JR, et al. Cardiomyopathy in the peripartum period: current aspects. A multicenter study. 11 cases. Arch Mal Coeur Vaiss. 1990;83:1563–1569.
7. Cenac A, Djibo A. Postpartum cardiac failure in Sudanese-Sahelian Africa: clinical prevalence in western Niger. Am J Trop Med Hyg. 1998;58:319–323.
8. Davidson NM, Parry EHO. Peripartum cardiac failure. Q J Med. 1978;47:431–461.
9. Lampert MB, Lang RM. Peripartum cardiomyopathy. Am Heart J. 1995;130:860–870.
10. Demakis JG, Rahimtoola SH, Sutton GC, et al. Natural course of peripartum cardiomyopathy. Circulation. 1971;44:1053–1061.
11. Isezuo AS, Akubakar AS. Epidemiology of peripartum cardiomyopathy. Erhn Dis. 2007;17:[68_TD$DIFF]228–233.
12. Sanderson JE, Adesanya CO, Anjorn P, Parry EHO. Post-partum cardiac failure – heart failure due to cardiac overload. Am Heart J. 1979;97:613–621.
13. Ekosse GE. X-ray diffraction study of kanwa used as active ingredient in achu soup in Cameroon. Afr J Biotechnol. 2010;9:7928–7929.
14. Ekanem EJ, Harrison GFS. Assessment of Lake salt for sourcing dietary minerals. Niger J Chem Res. 1997;2:[69_TD$DIFF]33–38.
15. Alawa CBI, Adamu AM, Ehoche OW, Lamidi OS, Oni OO. Performance of Bunaji Bulls fed maize stover supplemented with urea and local mineral lick (Kanwa). J Agric Environ. 2000;1:35–42.

16. Jendrassik I, Grof P. Simplified photometric methods for the determination of bilirubin. Biochem Zchr. 1938;297:81–89.

17. Reitman S, Frankel S. A calorimetric method for the determination of serum glutamic oxaloacetic and glutamic pyruvic transaminases. Am J Clin Pathol. 1957;28:56–63.

18. Sood R. Medical Laboratory Technology. Method and Interpretation. New Delhi: Jaypee Brothers Medical Publishers Ltd; 1999:488.

19. Lowry OH, Rosenbrough NT, Farr AL, Randall RJ. Protein measurements with the Folin phenol reagent. J Biol Chem. 1951;193:265–275.

20. Doumas BT, Watson W, Briggs HG. Albumin standards and the measurement of serum albumin with bromcresol green. Clin Chim Acta. 1971;31:87–96.

21. Szasz GUMZ. New substrates for measuring gamma-glutamyl transpeptidase activity. Z Klin Chem Biochem. 1974;12:228.

22. Young DS. Effects of drugs on clinical laboratory test. Ann Clin Biochem. 1997;34:579–581.

23. Bartels H, Bohm M. Micro determination of creatinine. Clin Chim Acta. 1971;32:81–85.

24. Willebrands AF. The determination of sodium and potassium in blood serum and urine by means of the flame photometer. Rec Trav Chim Pays Bas. 1950;69:799–821.

25. Glerach JL, Frazier RG. Spectrophotometric determination of chloride in sweat and serum with diphenylcarbazone. Anal Chem. 1958;30:1142–1146.

26. Sibnner BH, Caillouette J. Improved method for the bedside determination of bicarbonate in serum. JAMA. 1954;155:644–648.

27. Pyatt JR, Dubey G. Peripartum cardiomyopathy: current understanding, comprehensive management review and new developments. Postgrad Med J. 2011;87:34–39.

28. Li X, Luo R, Jiang R, et al. The prognostic use of serum concentrations of cardiac troponin-I, CK-MB and myoglobin in patients with idiopathic dilated cardiomyopathy. Heart Lung: J Acute Crit Care. 2014;43:219–224.

29. Logeat D, Beyne F, Cusson C, et al. Evidence of cardiac myolysis in severe nonischemic heart failure and the potential role of increased wall strain. Am Heart J. 2001;141:247–253.

30. Muhammad AS, Saidu Y, Bilbis LS, Onu A, Isezuo SA, Sahabi S. Effect of dried lake salt (Kanwa) on lipid profile and heart histology of female albino rats. NJ Basic Appl Sci. 2014;22:73–78.

31. Bolien IA, Van Deel ED, Kuster DW, Van Der Velden J. Peripartum cardiomyopathy and dilated cardiomyopathy: different at heart. Front Physiol. 2015;5:531.

32. Umar S, Nadadur R, Jorga A, Amjedi M, Matoni H, Eghbali M. Cardiac structural and hemodynamic changes associated with physiological heart hypertrophy of pregnancy are reversed postpartum. J Appl Physiol. 2012;113:1253–1259.

33. Gaasch WH, Zile MR. Left ventricular structural remodeling in health and disease: with special emphasis on volume, mass, and geometry. J Am Coll Cardiol. 2011;58:1733–1740.

34. Hilfiker-Kleiner D, Kaminski K, Podewski E, et al. A cathepsin D-cleaved 16 kDa form of prolactin mediates postpartum cardiomyopathy. Cell. 2007;128:589–600.

35. Malomo SO. Toxicological implication of ceftriaxone administration in rats. Niger J Biochem Mol Biol. 2000;15:33–38.

36. Pagana KD, Pagana TJ. Mosby’s Manual of Diagnostic and Laboratory Tests. USA: St Louis, Mosby Elsevier; 2006:40.