Straightforward synthesis of bis-tetraazacycloalkanes: towards new potential CXCR4 antagonists?†

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We report herein an efficient and general method for the synthesis of new bismacroyclic compounds, structural analogues of biscyclam AMD3100, in which the two macrocycles are linked together through carbon atoms of the cycles. Several representatives of this new class of biscyclic derivatives were prepared by reacting C-aminomethyl-13aneN4 with aromatic dialdehydes. Preliminary in vitro studies were performed to evaluate the affinity of these compounds towards the co-receptor CXCR4.

1 Introduction

The CXCR4 chemokine receptor is a seven transmembrane helix protein, member of the G-protein-coupled receptor (GPCR) superfamily, whose own natural ligand is SDF-1α or CXCL12.1–4 The axis CXCR4/CXCL12 is implicated in the regulation of numerous biological processes in the human body that result in a variety of normal physiological responses, such as chemotaxis, cell survival and proliferation, intracellular calcium flux, and gene transcription. CXCR4 is also involved in many downstream pathways, such as inflammation,5 mobilisation of stem cells,6 and development of cancer cells metastasis.7–8 Furthermore, CXCR4 is a co-receptor for T-cell tropic strains of human immunodeficiency virus-1 (HIV-1) and allows fusion and entry of the virus into human white blood cells. Targeting specifically these receptors represents therefore a huge potential for many applications, such as characterization of a pathology, drug development, staging and restaging, following a therapy response, delineation and quantification of tumors in oncology. Nowadays, only one bis-cyclam derivative, AMD3100,9,10 is clinically used in oncology, in combination with G-CSF (granulocyte colony stimulating factor) to mobilize haematopoietic stem cells and allow harvesting.11,12 Several AMD like compounds, differing by the size of the macrocycle and the nature of the linker, have been synthesized and studied,11,12–15 highlighting the tremendous importance of these parameters on the affinity of the bismacroyclic compound for its target. Thus, it is of major interest to devise new synthetic routes for the construction of bismacroyclic scaffolds analog to AMD3100, which could present better therapeutic properties.16,17

In this study we report the synthesis of a series of new bis-tetraazacycloalkanes, in which the two macrocycles (13aneN4) are linked together through carbon atoms of the cycles. To this end, we decided to use C-aminomethyl macrocycles, already described by our group.16–18 Beside the fact that these precursors may lead to a new family of AMD analogues with enhanced properties, this approach is synthetically appealing since it represents a straightforward route towards bismacroyclic compounds. Indeed, it is a very attractive alternative to the synthesis of AMD like derivatives in which the bridge between the two macrocycles is linked to nitrogen atoms of the cycles, which usually involves tedious purification steps and/or protection/deprotection sequences in order to differentiate one out of the four nitrogen atoms. We also report inhibition studies of anti-CXCR4 mAb binding and of SDF-1α chemotaxis activity by AMD3100 and its analogs.

2 Results and discussion

2.1 Synthesis

In previous work we have demonstrated that the primary amine function of C-aminomethyl-13aneN4 (1) readily reacts with one equivalent of aromatic aldehydes to form cyclic aminal derivatives involving an adjacent nitrogen atom of the cycle. Reduction of these aminal compounds with NaBH₄ gave access to the...
corresponding macrocycles selectively alkylated on the pendant amino methyl group. It might be expected that such peculiar reactivity can be exploited to synthesize bismacrocycles by reacting the C-aminomethyl macrocycle (1) with dialdehydes. Indeed, by using 0.5 equivalent of various dialdehydes (isophthalaldehyde, terephthalaldehyde, pyridine-2,6-dicarbaldehyde, 2,2'-oxydibenzaldehyde), the corresponding bismacro cyclic bisaminal derivatives were obtained in good yields (up to 82%) (Scheme 1).

The reduction of these compounds with NaBH₄ followed by acidic treatment gave the hydrochloride salts of the aimed bismacrocyclic bisaminal derivatives 5–8 as white powders.

The NMR spectra of the isolated bismacro cyclic bisaminal derivatives 2–4 are very complicated due to the formation of a mixture of different isomers. Monocrystals of pure 2a were prepared after slow crystallization of 2 in diethylether (Fig. 1, left). The structure of compound 4a was also determined by X-ray diffraction (Fig. 1, right). An exhaustive description of each compound is given in the ESI†.

Both compounds crystallize identically, they present the same symmetry (monoclinic) and the same conformation with a water molecule in the asymmetric unit. A RMSD_max of 0.4 Å was found by superimposing both structures. Cremer–Pople parameters, which describe the conformation of a puckered ring in a quantitative mathematically well-defined manner, indicate that the five membered aminal rings have a slightly distorted chair geometry [compound 2a: N1–C9–C10–N5–C11, Q = 0.360(3) Å, ϕ = 312.4(4)°, N6–C18–N10–C19–C20, Q = 0.345(2) Å, ϕ = 226.6(4)°; compound 4a: N4–C7–C10–N5–C11, Q = 0.348(4) Å, ϕ = 314.1(7)°, N7, N7–C17–N8–C19–C18, Q = 0.363(4) Å, ϕ = 24.5(6)]°]. These five membered rings impose the half conformation of a part of the polyazacycloalkane. The second shape of the molecule is governed by intra- and intermolecular hydrogen bonds (see ESI†).

The characterization of the three different isomers was not further investigated since they all give the same compound after opening the aminal bridges by NaBH₄. The new bismacro cyclic 5–8 were fully characterized using mass spectrometry, ¹H and ¹³C NMR and elemental analysis. It is noteworthy that the number of signals in the ¹³C NMR spectra of these compounds is low. This feature can of course be explained by an expected symmetry of the molecule. However, the presence of two identical chiral carbon atoms in these bismacro cyclics should give raise to the formation of three stereoisomers, i.e. a meso compound and a couple of enantiomers since the starting C-aminomethyl macrocycle 1 was used as a racemic mixture. The simplicity of the NMR spectra seems to prove that the reaction is stereospecific, leading to either the meso compound or a racemic mixture of the two enantiomers.

![Scheme 1 Synthesis of bistetraazacycloalkanes.](image-url)

**Scheme 1** Synthesis of bistetraazacycloalkanes.

**Fig. 1** ORTEP views of compound 2a (left) and compound 4a (right). Thermal ellipsoids are drawn at 50% probability level. Water molecule is omitted for clarity.
If the method described herein appears as a powerful route for the preparation of bis-13aneN4 derivatives, it must be noted that some unexpected results have been obtained with two other diahdehydes. When using ortho-phthalaldehyde, the formation of the bismacrocycle was not observed. NMR and mass spectrometry data suggest the formation of a [1 + 1] aminal adduct 9 containing a lactam moiety (Scheme 2).

The structure of compound 9 was confirmed by X-ray diffraction of the monocrystal grown from a hexane/dichloromethane (99 : 1) mixture (Fig. 2). A 6-membered ring (N1–C9–C8–C10–N5–C11) with a slightly distorted chair conformation is formed [Cremer–Pople parameters: Q = 0.5717(19) Å, φ = 10.9(2)°, ϕ = 32.5(11)°]. An intramolecular hydrogen bond imposes the conformation of the nitrogen atoms N2 and N3.

This reactivity can be explained by the close proximity of the two aldehyde functions in the rigid ortho-phthalaldehyde. Indeed, after the formation of the cyclic aminal resulting from the reaction of the primary amine of the starting macrocycle with one aldehyde, the second carbonyl group is attacked preferentially by the secondary nitrogen atom of the aminal cycle rather than by the primary amine of a neighbor macrocycle. This intramolecular reaction is thermodynamically favored by the formation of a five membered hemi-acetal ring which can not undergo a dehydration step but is oxidized into a lactam function, although no oxidizing reagent was used during the reaction, apart from the oxygen from air.

The expected bisaminal bismacrocycle was also not obtained when reacting C-aminomethyl-13aneN4 (1) with 2,9-dicarboxaldehyde-1,10-phenanthroline (2,9-DCP). Again, this could be attributed to intra- and/or intermolecular reaction of the aldehyde with different amine functions, giving side products. These results prompted us to devise another route for the preparation of the desired bismacrocycles. In order to avoid the intramolecular reaction of the second aldehyde with secondary amine functions, we decided to use as starting material the bisaminal protected macrocycle 10 which is actually the precursor of C-aminomethyl macrocycle 10 (Scheme 3).

In this case the cyclic aminal cannot be formed and the reaction with 2,9-dicarboxaldehyde-1,10-phenanthroline followed by reduction with NaBH₄ gives the protected bismacrocycle 11. This compound was finally treated by HCl 37% in ethanol to yield the aimed bis-macrocycle 12.

### 2.2 Biological studies

Cellular binding assays of new AMD3100 analogues to the CXCR4 receptor were investigated in a competition assay with an anti-CXCR4 monoclonal antibody (mAb) (12G5 conjugated to phycoerythrin). Jurkat cells, a T-lymphocyte cell line which expresses high level of CXCR-4 expression, were used and binding assays of new compounds 2(a–c), 3(a–c), 4(a–c), 5, 6, 7, and AMD3100 (as reference) to the cells were analysed by flow cytometry. On the whole, synthesized compounds were less efficient than AMD3100, which maximum inhibition concentration was 1 μg mL⁻¹. Compounds 2(a–c), 3(a–c), 4(a–c) and 7, didn’t show any inhibition potency. However, at 5 μg mL⁻¹, the analogs 5 and 6 could inhibit the binding of the mAb (41.16% ± 8.1 and 54.83% ± 2.5, respectively) on Jurkat T cells (Fig. 3). This result was confirmed using DAUDI cells (data not shown). These preliminary results confirm the importance of both the nature of the macrocycle and of the bridge between the two primary amine functions of the starting macrocycle.
limit the formation of undesired compounds. A has been used instead of C-aminomethyl-13aneN4 in order to prepare the aimed bismacrocycles, the protected macrocycle really in hydrogen bonds with protonated cyclam of AMD3100. The carboxylate groups of the receptor residues form three (one strong, one intermediate and one weak) hydrogen bonds with protonated cyclam of AMD3100.

Changing the geometry of the macrocycle and the spacer can really influence the binding of the compounds, yielding in a decrease of the resulting interactions for the receptor. Therefore, not surprisingly, 5 and 6, which are structurally more resembling to AMD3100, present the highest affinity for the CXCR4 receptor.

Since the interaction between SDF-1 and CXCR4 has been shown to direct tumor cells to organ sites with high levels of SDF-1 expression, the effect of AMD3100 and its analogs on inhibiting CXCL12-induced migration of Jurkat T cells was estimated by a classical chemotaxis assay. As expected, Jurkat T cells migrated in response to CXCL12, even if only 20% migrated because of a low % of Jurkat T cells bound the chemokine. After AMD3100 treatment (10 µg mL⁻¹), chemotactic activity of Jurkat cells was totally inhibited. However, this property of AMD3100 to inhibit CXCL12-induced chemotaxis through CXCR4 was compromised by structural modifications as none of the analogs could strongly SDF-1-induced migration of Jurkat T cells (Fig. 4).

### 3 Conclusion

The work described herein reports the synthesis of a new family of bis-tetraazacycloalkanes formed by two same 13aneN4 units connected by an aromatic spacer. The reaction of two equivalents of C-aminomethyl-13aneN4 with commercial dialdehydes in ethanol followed by reduction with NaBH₄ resulted in the formation of bismacroyclic compounds, structural analogs of AMD3100. In some cases, with ortho-phthalaldehyde or 2,9 DCP, unexpected lactam or side products were obtained. To prepare the aimed bismacrocycles, the protected macrocycle has been used instead of C-aminomethyl-13aneN4 in order to limit the formation of undesired compounds. Affinity assays with the CXCR4 receptor have been performed and showed an effective binding to the receptor for compounds 5 and 6. Further work is ongoing for the preparation of metal complexes of these bismacrocycles. Indeed, previous studies have shown that metal ions incorporation could increase the affinity of bismacrocycles such as AMD3100 or analogs for its receptor.²³⁻²⁵

### 4 Experimental section

#### 4.1 Instruments

All analyses were performed at the “Plateforme d’Analyses Chimiques et de Synthèse Moléculaire de l’Université de Bourgogne”. High resolution and accurate mass measurements were carried out using a Bruker MALDI-TOF, microTOF-Q™ ESI-TOF (Electro Spray Ionization-Time of Flight) and a Thermo Scientific™ LTQ Orbitrap mass spectrometer. ²H and ¹³C NMR spectra were recorded on a Bruker 300 and 500 NMR spectrometer. Chemical shifts were reported in parts per million using tetramethylsilane (TMS) as the internal standard. The following abbreviations are used; s: singlet, d: doublet, t: triplet, q: quartet, m: multiplet, b: broad.

X-ray equipment and refinement are detailed in ESL.† Data CCDC 1476681, 1476682 and 1476683 contain the supplementary crystallographic data for compound 2a, 4a and 9 respectively.

#### 4.2 Biologicals

**Chemokine and antibodies.** Recombinant human SDF-1 (CXCL12) was purchased from PeproTech (Neuilly-Sur-Seine, France), Fluorokine® biotinylated human SDF-1, monoclonal anti-human CXCR4-phycocerythrin (PE) antibody (clone 12G5) and mouse IgG2A isotype control-PE were purchased from R&D Systems.

**Cell lines.** Daudi cell line was provided by OncoDesign and Jurkat cell line (clone E6-1) was obtained from the American Type Culture Collection (ATCC) (Molsheim, France). Cells were maintained in RPMI 1640 supplemented with 10% FBS, 2 mM L-glutamine, 100 µg mL⁻¹ penicillin and 100 µg mL⁻¹ streptomycin.

**Flow cytometry.** Daudi or Jurkat cells (105 cells in 25 µL) were incubated with or without AMD3100 or its analog molecules at different concentrations for 15 min on ice. 12G5 mAb (5 µg mL⁻¹) was then added for 30 min on ice. Cells were washed twice with buffer and acquired on a flow cytometer (LSR II, Becton Dickinson, San Jose, CA, USA). To determine the presence of SDF-1 on Jurkat cells, Fluorokine cells, Fluorokine® kit was used. Briefly, 10 µL of biotinylated human SDF-1 was added to 25 µL of cells (4 × 10⁵ per mL) for 30 min on ice. 10 µL of avidin-FITC was then added and cells were further incubated for 30 min on ice. Cells were washed twice with Cell Wash Buffer and re-suspended for flow cytometry acquisition.

**Chemotaxis.** Jurkat cells were serum-starved in migration medium (RPMI 1640 containing 1% BSA, 10 mM HEPES buffer, pH 6.9) for 4 h. Migration assays were performed in duplicate in transwell plates of 6.5 mm diameter, with 5.0 µm pore polycarbonate membrane (VWR international, France). A volume of 0.6 mL migration medium, with or without 100 ng mL⁻¹ of recombinant human SDF-1, was added to the lower chamber of
Reagents, solvents and chemicals were purchased from Sigma-Aldrich and were used as supplied without further purification. Compounds 1 and 10 were purchased from Chematech SA.

1,3-Bis[tetradecahydro-1H-imidazo[1,5-d][1,4,7,10]tetraazacyclododecane-3-yl]benzene (2a–c). 0.31 g of isophthalaldehyde (2.3 mmol) was added to a solution of 1 (1.0 g, 4.65 mmol) in ethanol (20 mL) at room temperature. The mixture was stirred at reflux during 2 hours. A mixture of compounds 2a, 2b, 2c was obtained as a colorless oil (m = 0.85 g, 1.60 mmol, yield = 70%). Crystals of 2a could be obtained by recrystallization in diethyl ether. 1H NMR of mixture of 2a-c (300 MHz, CDCl3, 300 K) δ (ppm): 1.42–3.31 (m, 46H), 4.29 (bs, 1H), 7.21–8.21 (m, 4H). 13C{1H} NMR (75 MHz, CDCl3, 300 K) δ (ppm): 46.6, 46.8, 46.9, 47.3, 47.5, 47.8, 48.0, 48.1, 49.2, 49.3, 49.4, 49.8, 50.0, 50.1, 50.3, 50.5, 50.6, 50.8, 51.0, 51.1, 51.2, 51.3, 51.5, 51.6, 51.7, 51.8, 51.9, 52.0, 52.4, 52.6, 52.7, 52.8, 52.9, 53.0, 53.5, 53.6, 53.8, 54.0. HRMS-MALDI-TOF (m/z): calculated: 529.152 [M + H]+. Elemental analysis: C28H56N10 clastoid (100 mL). Compound 6 was obtained as a white powder (m = 0.80 g, 0.88 mmol, yield = 55%). 1H NMR (300 MHz, CDCl3, 300 K) δ (ppm): 2.16 (bs, 5H), 2.84–3.73 (m, 37H), 4.34 (m, 4H), 7.59 (bs, 4H). 13C{1H} NMR (75 MHz, CDCl3, 300 K) δ (ppm): 24.7, 43.8, 45.4, 46.5, 46.6, 48.4, 49.7, 50.8, 54.2, 54.3, 133.4 (*2), 134.2. ESI-TOF (m/z): calculated: 571.432, found: 571.456 [M + K]+. Elemental analysis: C28H35N10, 10HCl, 4H2O, calculated: C (34.69%), H (7.69%), N (14.45%), found: C (34.74%), H (8.09%), N (14.18%).

1,1′-(1,3-Phenylenylene)bis(N-(1,4,7,10-tetraazacyclododecane-5-yl)-methyl)ethanamine, hydrochloride salt (7). This compound was synthesized according to the procedure used for the synthesis of 5 starting from compound 3a-c (0.85 g, 1.62 mmol) and NaBH4 (0.61 g, 16.10 mmol). Compound 7 was obtained as a white powder after protonation by HCl 37% (m = 0.5 g, 0.56 mmol, yield = 80%). 1H NMR (600 MHz, CDCl3, 300 K) δ (ppm): 1.05 (bs, 1H), 2.08 (m, 5H), 2.70–4.01 (m, 41H), 4.43 (bs, 5H), 7.40 (m, 2H), 7.83 (m, 1H). 13C{1H} NMR (150 MHz, CDCl3, 300 K) δ (ppm): 22.3, 41.2, 43.1, 44.2, 44.3, 46.0, 47.6, 48.4, 51.6, 51.7, 123.4, 139.5, 150.4. ESI-TOF (m/z): calculated: 266.732, found: 267.738 [M + 2H]+.

2,2′-(Oxybis(2,1-phenylene))bis(N-(1,4,7,10-tetraazacyclododecane-5-yl)methyl)ethan-1-amine) (8). 0.2 g of 2,2′-oxydibenzaldehyde (0.90 mmol) was added to a solution of compound 1 (0.38 g, 1.76 mmol) in ethanol (20 mL). The mixture was stirred at room temperature during 2 hours. 0.34 g of NaBH4 (9.00 mmol) was added to the mixture, which was stirred at room temperature during 12 hours. Then the solvent was removed and the residual oil was taken up in dichloromethane (50 mL). The insoluble salts were removed by filtration on CLARCEL. After evaporation of the solvent, the colorless oil was taken in ethanol and protonated by HCl 37%. The precipitate was filtered and washed with diethyl ether (2 × 20 mL). Compound 5 was obtained as a white powder (m = 0.85 g, 0.90 mmol, yield = 58%). 1H NMR (300 MHz, CDCl3, 300 K) δ (ppm): 2.09 (bs, 4H), 2.79–3.69 (m, 39H), 4.29 (m, 4H), 7.54 (m, 4H). 13C{1H} NMR (75 MHz, CDCl3, 300 K) δ (ppm): 16.8, 22.2, 41.2, 45.9, 47.2, 48.3, 51.7, 51.8, 130.2, 131.1, 131.4, 131.8. ESI-TOF (m/z): calculated: 532.469, found: 533.479 [M + H]+. Elemental analysis: C28H45N10, 10HCl, 4H2O, calculated: C (36.74%), H (7.49%), N (15.30%), found: C (36.92%), H (7.71%), N (14.90%).
solid (calculated: 734.522, found: 733.246 [M]+.

136.6 (yield = 4H), 8.16 (m, 2H). 13C{1H} NMR (75 MHz, CDCl3, 300 K) δ (ppm): 28.0, 45.7, 47.7, 48.2, 48.9, 50.9, 51.1, 51.7, 51.9, 53.3, 77.2, 124.2, 124.9, 129.3, 130.9, 133.5, 141.3, 166.7. HRMS-MALDI-TOF (m/z): calculated = 329.221, found: 329.701 [M]+. Elemental analysis: C18H27N5O.

Calculated: C (65.62%), H (8.26%), N (21.26%); found: C 51.1, 52.3, 56.4, 57.1, 122.2, 125.8, 127.7, 136.5, 145.2, 160.8. HRMS-MALDI-TOF (m/z): calculated = 634.490, found: 635.503 [M + H]+.

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