Enteric fever in two vaccinated travellers to Latin America

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A previously healthy 24-year-old woman (patient A) and her partner, a previously healthy 25-year-old man (patient B), presented to our clinic with fever after travelling in Latin America. Patient A reported an 11-day history of daily fever to 40°C, chills, diffuse muscle aches and joint pains, bilateral throbbing headache, insomnia and watery, nonbloody diarrhea up to 12 times daily. Patient B presented with a 12-day history of daily fevers to 41°C, chills, headache and extreme fatigue.

Three days before their presentation, they had returned from a six-month trek through rural and urban areas of Peru, Ecuador, Chile, Argentina and Bolivia. They had sought travel advice before their trip and had received vaccinations for typhoid fever (injectable Vi polysaccharide vaccine), yellow fever and hepatitis A. They had received atovaquone/proguanil for malaria prophylaxis, to which they had been nonadherent.

The last week of their trip had been spent in Lima, Peru, where they had both begun to feel ill. Six days before the onset of illness, they had spent several days in a remote village in the jungle near Rurrenabaque, Bolivia, living among local residents, eating local foods and drinking local water. They had also been bitten extensively by mosquitoes and possibly other arthropods. On day four of patient A’s illness (day five of patient B’s illness), they sought care at a local hospital in Lima. Thick and thin blood films were used to screen for malaria, and serum agglutination antibody tests were used to screen for typhoid and paratyphoid fevers and brucellosis. The results of the patients’ blood tests were normal, with the exception of mild thrombocytopenia seen in the sample from patient A. They were discharged from the hospital with a probable diagnosis of dengue fever and were given no specific treatment. Both patients continued to feel ill, with spiking temperatures, headache and fatigue. Patient A also continued to have myalgia and arthralgia, though her diarrhea began to improve. By the time patient A arrived in Canada (eight days after the start of her illness), she was having five diarrheal stools per day. Both patients had loss of appetite but were drinking large amounts of water.

On examination, the patients appeared ill, although both were afebrile, alert and oriented. Patient A’s blood pressure was 100/50 mm Hg; patient B’s blood pressure was 90/60 mm Hg. Physical examination showed that patient A had mild tenderness to palpation of the left upper quadrant of the abdomen without clinical evidence of splenomegaly, as well as a diffuse, blotchy, erythematous, macular rash across her upper and lower extremities, chest and abdomen. A physical examination of patient B was unremarkable.

Laboratory investigations from their initial visit to our clinic (day 1) are summarized in Table 1. Notably, the results from thick and thin blood smears and dipstick assays screening for malaria were negative. The patients returned to our clinic immediately after providing samples of their blood. Because the results of the tests for malaria were negative, each patient was given a 10-day course of ciprofloxacin for empiric management of presumed enteric fever. They were asked to return to the clinic the next day (follow-up 1, day 2) for another malaria screening and reassessment. The results of laboratory investigations done during each of their three follow-up visits are summarized in Table 1.

By the third day of the course of ciprofloxacin, both patients began to feel better, reporting peak temperatures of 38–38.5°C in the previous
### Table 1: Results of laboratory investigations for a previously healthy 24-year-old woman (patient A) and 25-year-old man (patient B) who presented with fever after travelling in South America

| Investigation               | Normal values | Patient | Initial visit on day 1 | Follow-up on day 2 | Follow-up on day 7 | Follow-up on day 14 |
|-----------------------------|---------------|---------|------------------------|--------------------|--------------------|---------------------|
| Hemoglobin, g/L             | 120–140       | A       | 136                    | 123                | 123                | 121                 |
|                             | 140–160       | B       | 145                    | 136                | —                  | 138                 |
| WBC, 10⁹/L                  | 3.5–11.0      | A       | 3.0                    | 4.2                | 3.9                | 4.4                 |
|                             |               | B       | 8.2                    | 7.7                | —                  | 4.4                 |
| Platelets, 10⁹/L            | 150–400       | A       | 135                    | 143                | 309                | 388                 |
|                             |               | B       | 279                    | 270                | —                  | 473                 |
| AST, U/L                    | < 37          | A       | 380                    | 301                | 54                 | 23                  |
|                             |               | B       | 82                     | 75                 | 45                 | 24                  |
| ALT, U/L                    | < 37          | A       | 228                    | 217                | 92                 | 26                  |
|                             |               | B       | 99                     | 112                | 101                | 52                  |
| ALP, U/L                    | 20–140        | A       | 162                    | 128                | 86                 | 69                  |
|                             |               | B       | 59                     | 52                 | 60                 | 65                  |
| Bilirubin, µmol/L           | < 20          | A       | 8                      | 7                  | 5                  | 6                   |
|                             |               | B       | 10                     | 11                 | 11                 | 11                  |
| Creatinine, µmol/L          | 45–100        | A       | 62                     | 58                 | 54                 | —                   |
|                             |               | B       | 76                     | 98                 | 66                 | —                   |
| Sodium, µmol/L              | 135–145       | A       | 126                    | 129                | 137                | —                   |
|                             |               | B       | 129                    | 131                | 138                | —                   |
| Potassium, µmol/L           | 3.5–4.5       | A       | 3.4                    | 4.1                | 4.2                | —                   |
|                             |               | B       | 5                      | 4                  | 4.8                | —                   |
| Blood culture               | None          | A       | Pending                | GNB                | Salmonella enterica* | No further speciation |
|                             |               | B       | Pending                | GNB                | Salmonella species* | S. enterica ser. Paratyphi* |
| Malaria screen              | Negative      | A       | Negative               | Negative           | —                  | —                   |
|                             |               | B       | Negative               | Negative           | —                  | —                   |
| Dengue serology             | Nonreactive   | A       | Pending                | Pending            | Pending            | IgM nonreactive, IgG reactive |
|                             |               | B       | Pending                | Pending            | Pending            | IgM nonreactive, IgG nonreactive |
| Rickettsial serology        | Negative      | A       | Pending                | Pending            | Pending            | Negative |
|                             |               | B       | Pending                | Pending            | Pending            | Negative |
| Urinalysis                  | Normal        | A       | Normal                 | —                  | —                  | —                   |
|                             |               | B       | Normal                 | —                  | —                  | —                   |
| Stool culture†              | Negative      | A       | Pending                | Negative           | —                  | S. enterica ser. Paratyphi B (repeat stool culture) |
|                             |               | B       | Pending                | Salmonella species | S. enterica Paratyphi | S. enterica ser. Paratyphi B |
| Chest radiography           | Normal        | A       | Normal                 | —                  | —                  | —                   |
|                             |               | B       | Normal                 | —                  | —                  | —                   |
| Abdominal ultrasound        | Normal        | A       | —                      | —                  | Normal             | —                   |
|                             |               | B       | —                      | —                  | —                  | —                   |

Note: ALP = alkaline phosphatase, ALT = alanine transaminase, AST = aspartate transaminase, GNB = gram-negative bacilli, HB = hemoglobin; IgG = immunoglobulin G; IgM = immunoglobulin M; WBC = white blood cell.

*Susceptible to amoxicillin, ciprofloxacin and trimethoprim/sulfamethoxazole.
†The results of follow-up stool cultures obtained three months post-treatment were negative for both patients.
48 hours. Their chills had stopped and their
headaches and myalgia had resolved, as had
patient A’s diarrhea. Both patients still had loss of
appetite and felt fatigued, and patient A continued
to have mild tenderness on the left side of her
abdomen. By the fourth day of treatment, the
patients were no longer febrile. By the seventh
day of treatment, both patients had regained their
appetites and energy levels. Fourteen days after
starting treatment, the patients felt normal. The
final diagnosis was enteric fever due to Salmonella
enterica serotype Paratyphi B.

**Discussion**

Enteric fever due to either Salmonella enterica
serotype Typhi (“typhoid fever”) or Salmonella
enterica serotypes Paratyphi A, B or C (“para-
typhoid fever”), is one of the more common
causes of fever in the returned traveller. In single-
centre and multicentre observational studies of
illness in returned travellers, enteric fever has
been shown to account for 2%–7% of such ill-
nesses, and it is generally among the top five
specific etiologic diagnoses (Box 1).\(^1\)–\(^6\) This
foodborne and waterborne illness has highest
relative risks among people travelling to visit
friends and relatives on the Indian subconti-
nent;\(^1\)\(^\_\)\(^6\)\(^\_\) however, travellers to all developing
countries, regardless of purpose, are at theoretical
risk (Figure 1).\(^9\)

A history of travel to the Indian subcontinent
in a febrile returned traveller should raise suspi-
cion of enteric fever. Of 416 cases of imported
typhoid in the United Kingdom over a three-year
period, 70% were from India and Pakistan.\(^11\) In
the observational analysis by Freedman and col-
leagues, typhoid fever was a major contributor to
systemic febrile illness without an identifiable
organ focus among people who had returned
from southcentral Asia.\(^7\)

Of 149 patients with documented S. enterica
ser. Paratyphi A in the United States in 2005–
2006 for whom epidemiologic information was
known, 90% had recently travelled to south
Asia.\(^11\) In a number of Asian countries, S. enter-
ica ser. Paratyphi A is becoming increasingly
responsible for enteric fever — in some regions,
it accounts for up to 50% of instances of the dis-
ease.\(^12\) However, S. enterica ser. Paratyphi B was
reported in five American travellers, four of whom
had recently visited Latin America.\(^11\)

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**Box 1: Conditions that commonly present
as fever in a returned traveller**\(^1\)–\(^6\)

- Malaria (20%–30%)
- Acute traveller’s diarrhea or gastroenteritis
  (10%–20%)
- Respiratory tract infection (10%–15%)
- Dengue fever (5%)
- Enteric fever due to Salmonella enterica
  serotypes Typhi or Paratyphi (2%–7%)
- Infections of the skin and soft tissue (2%–11%)
- Rickettsioses (3%)
- Acute infection of the urinary tract or sexually
  transmitted infection (2%–3%)
- Viral hepatitis (3%)
- Mononucleosis- or viral-like syndrome
  (4%–25%)

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**Figure 1:** Geographic distribution of typhoid fever. Reproduced with permission from Crump et al.\(^9\)
Are typhoid and paratyphoid clinically distinguishable?

Enteric fever caused by *S. enterica* ser. Paratyphi was formerly believed to cause a milder clinical picture than typhoid fever, although this is now known to be incorrect.\textsuperscript{13,14} Recent reports have shown that paratyphoid fever is clinically indistinguishable from typhoid fever,\textsuperscript{13,14} and that almost two-thirds of patients with enteric fever caused by *S. enterica* ser. Paratyphi A required admission to hospital.\textsuperscript{11}

After ingesting an infectious dose of *S. enterica* ser. Typhi (> 1000 organisms) or *S. enterica* ser. Paratyphi in water or food, an asymptomatic incubation period of 7–14 days typically ensues.\textsuperscript{15} Fever heralds the onset of bacteremia, which is often accompanied by frontal headache, myalgia, anorexia, nausea, vomiting, abdominal pain and constipation or diarrhea, as seen in our patients. Fever becomes high and is sustained over time. Enteric fever may be complicated by gastrointestinal bleeding in 10% of patients who have been unwell for more than two weeks before the start of treatment (Box 2).\textsuperscript{15} Gastrointestinal perforation and encephalopathy are more severe sequelae of untreated typhoid fever; fortunately, these outcomes occur in less than 3% of patients.\textsuperscript{15} Relapse of illness two to three weeks after fever has abated occurs in up to 10% of patients.\textsuperscript{19}

Chronic fecal carriage, defined as excretion of *S. enterica* in the stool for at least one year, occurs in 1%–4% of patients, and is more common among patients with gallstones,\textsuperscript{15} though the risk is lowest after treatment with fluoroquinolones. The eradication of these organisms is important from a public health perspective, but also because chronic carriage of *S. enterica* ser. Typhi (and Paratyphi A) has a small associated risk of biliary carcinoma.\textsuperscript{16}

What tests are used to confirm enteric fever?

Enteric fever should be excluded, along with competing diagnoses (Box 1),\textsuperscript{1,6} in returned travellers with history of fever, regardless of their immunization status. A culture of the bone marrow is the gold-standard diagnostic test for enteric fever;\textsuperscript{17} however, owing to its practical limitations and invasiveness, it is rarely performed. Rather, cultures of the blood and stool, though less sensitive than cultures of the bone marrow, are usually used for diagnostic and epidemiologic purposes. Blood cultures yield positive results from 60%–80% of patients with acute enteric fever, and results of stool cultures are positive 30% of the time.\textsuperscript{15} Serology plays no role in the diagnosis of enteric fever in resource-limited settings. Results of laboratory investigations that are compatible with a diagnosis of enteric fever include leukopenia, thrombocytopenia, and mild-to-moderate elevation of hepatic transaminase levels.\textsuperscript{15} However, the results of investigations, including the complete blood count, are often normal, even for a patient who looks ill.

Cultures of the blood and stool take time, and their results may be falsely negative. When the index of suspicion is high and a diagnosis of malaria has been ruled out, empiric administration of antibiotics (Table 2), the choice of which is based on epidemiology, is a reasonable approach.

What are the best options for management?

Enteric fever is usually treated with either oral or parenteral antibiotics (Table 2). Severe enteric fever characterized by gastrointestinal bleeding or perforation, neuropsychiatric complications (as outlined in Box 2) or cardiovascular complications such as shock, myocarditis or endocarditis, necessitates inpatient management of care and treatment with parenteral antibiotics. In addition, patients with severe enteric fever complicated by delirium, stupor, coma or shock have been shown to benefit from dexamethasone in randomized controlled trials.\textsuperscript{15,18}

### Box 2: Complications of enteric fever\textsuperscript{15}

**Gastroenterologic**
- Gastrointestinal bleeding and hemorrhage
- Gastrointestinal perforation
- Hepatitis
- Cholecystitis
- Relapse of illness
- Chronic carriage of *Salmonella enterica*

**Neuropsychiatric**
- Encephalopathy
- Delirium
- Meningitis

**Respiratory**
- Pneumonia

**Cardiovascular**
- Shock
- Myocarditis
- Electrophysiologic abnormalities
- Endocarditis (rarely, such as in patients with underlying rheumatic or congenital heart disease)

**Hematologic**
- Anemia
- Thrombocytopenia
- Disseminated intravascular coagulation
Antimicrobial resistance is an increasing concern in the management of enteric fever. Up to 87% of isolates of S. enterica ser. Paratyphi from south Asia that were tested at the Centers for Disease Control and Prevention in 2005–2006 showed resistance to nalidixic acid, which confers reduced clinical response to fluoroquinolones. Multidrug resistance (i.e., resistance to ampicillin, chloramphenicol and trimethoprim/sulfamethoxazole) and resistance to nalidixic acid is particularly common among strains of S. enterica ser. Typhi from south and southeast Asia. In general, multidrug resistance among isolates of S. enterica ser. Paratyphi is rare.\(^1^9\)

Fluoroquinolones have emerged as the favoured antibiotic in susceptible cases of enteric fever because they reduce the length of time the fever lasts, they have high clinical cure rates, and they show a reduced risk of long-term fecal carriage of the Salmonella organism.\(^1^5\) Systematic reviews of randomized controlled trials have shown higher rates of clinical failure among patients given treatment with ceftriaxone versus fluoroquinolones, which is the basis for using parenteral fluoroquinolone as the first line of treatment for complicated cases.\(^2^0\) In instances of fluoroquinolone resistance, systematic reviews of randomized controlled trials support the use of azithromycin orally as a practical first-line option for uncomplicated cases.\(^2^1\) Randomized controlled trials of treatment for typhoid fever have shown that fevers typically abate four to seven days after appropriate treatment has started; failure of a fever to diminish quickly is not an indication to change antibiotics.

### Table 2: Options for the antibiotic treatment of enteric fever caused by Salmonella enterica serotypes Typhi or Paratyphi\(^1^3,1^7\)

| Susceptibility profile of organism | Drug                          | Duration of treatment, d |
|-----------------------------------|-------------------------------|--------------------------|
| Uncomplicated enteric fever       |                               |                          |
| Fully susceptible                 | Ciprofloxacin                 | 5–10                     |
|                                   | Amoxicillin                   | 14                       |
|                                   | Trimethoprim/sulfamethoxazole | 14                       |
| Multidrug resistant*              | Ciprofloxacin                 | 5–10                     |
|                                   | Cefixime                      | 7–14                     |
| Quinolone resistant               | Azithromycin                  | 5–7                      |
|                                   | Cefixime                      | 7–14                     |
| Severe or complicated enteric fever (parenteral antibiotics)† |                               |                          |
| Fully susceptible                 | Ciprofloxacin                 | 10–14                    |
|                                   | Amoxicillin                   | 14                       |
|                                   | Trimethoprim/sulfamethoxazole | 14                       |
| Multidrug resistant*              | Ciprofloxacin                 | 10–14                    |
|                                   | Ceftriaxone                   | 10–14                    |
| Quinolone resistant               | Ceftriaxone                   | 10–14                    |
|                                   | Cefotaxime                    | 10–14                    |
|                                   | Ciprofloxacin†                | 14                       |

*Resistant to ampicillin, chloramphenicol and trimethoprim/sulfamethoxazole; more common in isolates from south and southeast Asia.
†Once the patient is able to tolerate oral medications, step down to a corresponding oral agent, such as ciprofloxacin, cefixime, trimethoprim/sulfamethoxazole or azithromycin, depending on the susceptibility profile of the organism, is appropriate.
‡High doses of fluoroquinolones for a longer period (e.g. 750 mg of ciprofloxacin twice daily for 14 d) may be used to treat enteric fever caused by a quinolone-resistant isolate with the caveat that the fever may not subside as quickly. Quinolone resistance is more common in isolates from south and southeast Asia.

### Table 3: Vaccines for typhoid that have been licensed for use in Canada\(^2^2\)

| Formulation                                      | Route of administration and schedule | Recommended minimum age of patient | Contraindications                                                                 | Booster schedule |
|--------------------------------------------------|--------------------------------------|------------------------------------|-----------------------------------------------------------------------------------|------------------|
| Live attenuated Ty21a                             |                                       |                                    |                                                                                   |                  |
| Vivotif (Berna Biotech)                          | 1 capsule, orally, every 2 d × 4 doses | ≥ 5 yr                             | Immune suppression, concurrent use of antibiotics, inflammatory bowel disease     | Every 5 yr       |
| Vivotil (Berna Biotech)                          | 1 sachet, orally, every 2 d × 3 doses  | ≥ 3 yr                             | Immune suppression, concurrent use of antibiotics, inflammatory bowel disease     | Every 5 yr       |
| Vi capsular polysaccharide (Typhim Vi and ViVaxim, Sanofi Pasteur; Typherix, GlaxoSmithKline) | 1 injection, intramuscularly         | ≥ 2 yr; ≥ 16 yr for ViVaxim        | Hypersensitivity to components of the vaccine                                    | Every 2–3 yr     |
How efficacious are currently available vaccines?

Two vaccine formulations, one oral (Ty21a) and one injectable (Vi polysaccharide), are licensed for prevention of enteric fever and confer partial protection against \textit{S. enterica} ser. Typhi (Table 3). A systematic review and meta-analysis showed that the cumulative three-year efficacy of Ty21a is 51% (95% confidence interval [CI] 36%–62%) under circumstances of ongoing exposure; the corresponding efficacy for the Vi polysaccharide vaccine is 55% (95% CI 30%–70%). The Ty21a live oral vaccine has also been shown to confer some protection against \textit{S. enterica} ser. Paratyphi. They had eaten and drunk local foods and water under the expectation of protection against enteric fever. They were therefore surprised in their scope of efficacy compared with the injectable Vi formulations.

Strict adherence to food and water hygiene may prevent transmission of enteric fever to travellers; however, such vigilance is difficult for long-term travellers to rural areas of the developing world. Thus, vaccination should be recommended for most travellers to tropical and subtropical destinations.

Given that the efficacy of vaccination is limited, a history of immunization against typhoid should not preclude exclusion of enteric fever as a diagnostic possibility in returned travellers with fever, and the limitations of these vaccines should be emphasized in the pretravel setting. Although the two travellers described in this report had received the injectable Vi polysaccharide vaccine, which lacks efficacy against \textit{S. enterica} ser. Paratyphi, they had eaten and drunk local foods and water under the expectation of protection against enteric fever. They were therefore surprised by the diagnosis.

The experience of these two patients underscores the importance of taking a full travel and epidemiologic history when returned travellers present with fever. Enteric fever remains a common cause of fever in returned travellers, even for those who report having been immunized.

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