Multifaceted Roles of Ferroptosis in Lung Diseases

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Ferroptosis is a distinct type of programmed cell death (PCD) that depends on iron and is characterized by the accumulation of intracellular iron, exhaustion of glutathione, deactivation of glutathione peroxidase, and promotion of lipid peroxidation. Recently, accumulated investigations have demonstrated that ferroptosis is strongly correlated with the initiation and development of many lung diseases. In this review, we summarized the contribution of ferroptosis to the pathologic process of lung diseases, namely, obstructive lung diseases (chronic obstructive pulmonary disease, asthma, and cystic fibrosis), interstitial lung diseases (pulmonary fibrosis of different causes), pulmonary diseases of vascular origin (ischemia-reperfusion injury and pulmonary hypertension), pulmonary infections (bacteria, viruses, and fungi), acute lung injury, acute respiratory distress syndrome, obstructive sleep apnea, pulmonary alveolar proteinosis, and lung cancer. We also discussed the therapeutic potential of targeting ferroptosis for these lung diseases.

Keywords: ferroptosis, lung disease, infection, injury, cancer

INTRODUCTION

The term “ferroptosis” was first coined by Dixon et al. (2012) to define a previously unknown form of programmed cell death (PCD) elicited by erastin in Ras-mutant cells in 2012. It is dependent on iron and is characterized by the accumulation of intracellular iron, exhaustion of glutathione, deactivation of glutathione peroxidase, and promotion of lipid peroxidation (Figure 1). Iron can catalyze the formation of free radicals from reactive oxygen species (ROS) via the Fenton reaction, which is the reduction of H2O2 by a single electron to produce a hydroxyl radical. Ferroptosis is distinct from other types of cell death, such as apoptosis, autophagy, pyroptosis, or necrosis, in morphology, genetics, metabolism, and molecular biology. The specific morphology of ferroptosis includes intact cytomembranes, cellular shrinkage, enhanced mitochondrial membrane density, reduced or absent mitochondrial cristae, crumpling of the mitochondrial membrane, rupture of the outer membrane, and regular nucleus size without concentrated chromatin (Dixon et al., 2012; Yang and Stockwell, 2016). However, apoptosis is characterized by typical apoptotic cellular bodies without rupture of cell membranes, while necrosis is characterized by the occurrence of cell swelling, nucleus concentration, fragmentation and dissolution, chromatin staining and flocculence, and organelle enlargement or fragmentation. In terms of pathological processes, cell apoptosis increases ROS, Ca2+, and pH levels, which activates cysteiny1 aspartate-specific proteinase and releases cytochrome c to promote caspase-3/7. During cell necrosis, the inflammatory response is induced by the activation of several signaling pathways, namely, Toll-like receptor 4/Myeloid...
differentiation primary response gene 88 (TLR4/Myd88)-dependent tumor necrosis factor (TNF) and receptor-interacting protein kinases (RIPK). Ferroptosis is regulated by glutathione peroxidase 4 (GPX4), which directly converts lipid hydroperoxides (L-OOH) to nontoxic lipid alcohols (L-OH) (Yang et al., 2014; Forcina and Dixon, 2019; and Stockwell et al., 2020). Later, a shred of evidence showed that p53 influences ferroptosis, it was reported and demonstrated that p53 is a key regulator of both classical ferroptosis pathways and noncanonical ferroptosis pathways, such as arachidonate 12-lipoxygenase (ALOX12) and iPLA2β. (Jiang et al., 2015a; Liu and Gu, 2021; Liu and Gu, 2022). Besides, ferroptosis is manipulated by several signaling pathways, such as p62-kelch-like ECH-associated protein (Keap1)-nuclear factor erythroid 2-related factor 2 (Nrf2)/heme oxygenase-1 (HO-1) (Sun et al., 2016; Dodson et al., 2019) and acyl-CoA synthetase long-chain family member 4 (ACSL4) (Doll et al., 2017). In addition, ferroptosis can be prevented by iron chelation or by the use of a lipophilic antioxidant but not by inhibitors of other forms of cell death. Aberrant regulation of ferroptosis has been implicated in disease pathogenesis in the heart, kidney, brain, liver, and lung (Zheng and Conrad, 2020). The contributions of ferroptosis to the pathologic process of multiple lung diseases have been recognized. In addition, many interventions, including ferroptosis inducers and inhibitors, iron chelators, lipid peroxidation inhibitors, and antioxidants, have been introduced into ferroptosis-related lung diseases (Table 1). In this review, we will summarize the recent understanding of ferroptosis in lung diseases and provide a new angle for future research on the pathologic process and clinical treatment of lung diseases (Figure 2).

**Chronic Obstructive Pulmonary Disease**
At present, chronic obstructive pulmonary disease (COPD) is the fourth leading cause of morbidity and mortality worldwide and is still increasing (Barnes et al., 2015; Barnes et al., 2019). It is characterized by chronic airway inflammation, lung destruction, and remodeling, resulting in irreversible airflow obstruction. Cigarette smoke (CS) exposure is the main risk factor for COPD due to its high concentration of ROS. The consequent cellular oxidative stress provokes inflammation, cell senescence, and death. Early studies have demonstrated that accumulated iron and ferritin and increased serum ferritin and nonheme iron were observed in lung epithelial cells and alveolar macrophages during exposure to CS (Wesselius et al., 1994; Ghio et al., 2008a).
TABLE 1 | Reagents and mechanisms of ferroptosis in lung diseases.

| Disease                  | Reagent     | Proposed mechanism                                                                 | References                                                                 |
|--------------------------|-------------|------------------------------------------------------------------------------------|---------------------------------------------------------------------------|
| COPD                     | Deferoxamine| Chelate iron and block iron-dependent lipid peroxidation                           | Ghio et al. (2008a); Yoshida et al. (2019); Tang et al. (2021a)            |
|                          | Ferrostatin-1| Inhibit lipid peroxidation                                                          | Yoshida et al. (2019); Tang et al. (2021a); Lian et al. (2021)            |
|                          | NAC         | Recover intracellular cysteine                                                     | Tang et al. (2021a)                                                       |
|                          | Curcumin    | Activate GPX4; Upregulate SLC7A11                                                  | Liu et al. (2022a)                                                       |
|                          | Dihydroquercetin| Regulate NRF2                                                                        |                                                                           |
| Asthma                   | Deferoxamine| Chelate iron and block iron-dependent lipid peroxidation                           | Wenzel et al. (2017)                                                     |
|                          | Ferrostatin-1| Inhibit lipid peroxidation                                                          | Wenzel et al. (2017); Yang and Shang, (2022)                              |
|                          | 3-methyladenine| Upregulate system Xc-; Activate GPX4                                               | Yang and Shang, (2022)                                                   |
|                          | Acupuncture | Downregulate SLC3A2 and ATP1A3                                                      | Tang et al. (2021c)                                                      |
|                          | Erastin     | Block cysteine import, inducing GSH depletion and GPX4 inactivation                | Wu et al. (2020)                                                        |
|                          | RSL3        | Covalently inhibit GPX4, causing accumulation of lipid peroxidation                | Wu et al. (2020)                                                        |
|                          | Artesunate  | Activate ferritinophages to increases iron abundance                                | Wu et al. (2020)                                                        |
| Cystic fibrosis          | Deferoxamine| Chelate iron and block iron-dependent lipid peroxidation                           | Maniam et al. (2021)                                                    |
|                          | Ferrostatin-1| Inhibit lipid peroxidation                                                          | Maniam et al. (2021)                                                    |
| Pulmonary fibrosis       | Deferoxamine| Chelate iron and block iron-dependent lipid peroxidation                           | (Cheng et al., 2021; Takahashi et al., 2021)                              |
|                          | Liproxstatin-1| Inhibit lipid peroxidation                                                          | He et al. (2021)                                                        |
| Ischemia–Reperfusion Injury | Liproxstatin-1| Inhibit lipid peroxidation                                                         | Xu et al. (2020)                                                        |
|                          | Rosiglitazone| Inhibition of ACSL4                                                                 | Xu et al. (2020)                                                        |
|                          | Irisin      | Regulate Nrf2/HO-1 axis; activates GPX4; inhibition of ACSL4                        | Wang et al. (2022a)                                                     |
|                          | Lidoceaine  | Regulate the p38/MAPK; increase FTH1 and GPX4; decrease Tf                         | Ma et al. (2022)                                                        |
|                          | Deferoxamine| Chelate iron and block iron-dependent lipid peroxidation                           | Lui et al. (2013)                                                       |
|                          | Pirfenidone | Chelate iron and block iron-dependent lipid peroxidation                           | Liu et al. (2005)                                                       |
| Pulmonary hypertension   | Ferrostatin-1| Inhibit lipid peroxidation                                                          | Xie et al. (2022)                                                       |
|                          | Deferoxamine| Chelate iron and block iron-dependent lipid peroxidation                           | Jaxenosky et al. (2015)                                                 |
| Infection: P. aeruginosa | Ferrostatin-1| Inhibit lipid peroxidation                                                          | Ousingsawat et al. (2021)                                               |
|                          | Bacilaein   | Suppress peroxidation of polyunsaturated fatty acids                                | Dar et al. (2022)                                                       |
|                          | CoQ10       | Inhibit lipid peroxidation                                                          | Ousingsawat et al. (2021)                                               |
|                          | Idebenone   | Inhibit lipid peroxidation                                                          | Ousingsawat et al. (2021)                                               |
| Pulmonary tuberculosis   | Ferrostatin-1| Inhibit lipid peroxidation                                                          | Amaral et al. (2019)                                                    |
|                          | Glutathione | Recover intracellular cysteine                                                      | Pan et al. (2020)                                                       |
|                          | Vitamin E   | Inhibit lipid peroxidation                                                          | Seyedrezazzadeh et al. (2008)                                           |
| Infection: Gram-negative bacteria | Liproxstatin-1| Inhibit lipid peroxidation                                                          | Klobucar et al. (2021)                                                 |
| COVID-19                 | Vitamin C   | Inhibit lipid peroxidation                                                          | Chavarría et al. (2021)                                                 |
|                          | Vitamin E   | Inhibit lipid peroxidation                                                          | Chavarría et al. (2021)                                                 |
|                          | NAC         | Recover intracellular cysteine                                                      | Chavarría et al. (2021); Jaiswal et al. (2020); Chavarría et al. (2021) |
|                          | Melatonin   | Chelate iron and block iron-dependent lipid peroxidation                           | Chavarría et al. (2021)                                                 |
|                          | Deferoxamine| Chelate iron; inhibit viral replication; immunomodulation; downregulate hepcidin I| Dalamaga et al. (2020)                                                  |
|                          | Deferriprone| Chelate iron; inhibit viral replication; immunomodulation                           | Dalamaga et al. (2020)                                                  |
|                          | Deferasirox | Chelate iron; inhibit viral replication; immunomodulation                           | Dalamaga et al. (2020)                                                  |
|                          | Lactoferrin | Bind iron and inhibit viral replication                                              | Carota et al. (2021)                                                    |
|                          | Methemoglobin reductase| Glutathione| Recover intracellular cysteine                                                      | Muhoberac, (2020)                                                     |
|                          | Glutathione | Recover intracellular cysteine                                                      | Silvagno et al. (2020)                                                  |
| Infection: Zygomycetes   | Melatonin   | Chelate iron; inhibit microbial activity                                            | Sen, (2021)                                                             |
|                          | Deferoxamine| Chelate iron and block iron-dependent lipid peroxidation                           | Kontogiorgi et al. (2010); Álvarez et al. (2013)                         |

(Continued on following page)
Mitochondrial dysfunction and endoplasmic reticulum stress are usually observed in the cytoplasm, and ferroptosis occurs in bronchial epithelial cells (Park et al., 2019). The increased ferritinophagy mediated by nuclear receptor coactivator 4 (NCOA4) and reduction of glutathione peroxidase 4 (GPX4) led to the accumulation of free iron and lipid peroxidation during CS exposure. Moreover, GPX4−/− mice showed significantly higher degrees of lipid peroxidation and an enhanced COPD phenotype than wild-type mice, whereas these phenotypes could be attenuated in GPX-transgenic mice (Dowdle et al., 2014; Yoshida et al., 2019). PM2.5 is another risk factor for COPD. Increased cellular iron content and ROS production in human endothelial cells were observed after inhaling PM2.5 particles, while the levels of glutathione (GSH) and nicotinamide adenine dinucleotide phosphate (NADPH) decreased. Iron overload and redox imbalance caused by TFRC and ferritin dysregulation are the major inducers of ferroptosis (Wang and Tang, 2019). The abovementioned investigations indicated that ferroptosis is involved and plays a crucial damaging role in COPD (Mizumura and Gon, 2021), and searching for an accurate inhibitor of ferroptosis to delay the progression and prevent the occurrence of COPD is pivotal to the forthcoming research. Experimental interventions, such as the iron chelator deferoxamine, the ferroptosis inhibitor ferrostatin-1, and suppression of lipid peroxidation by GPX4, could effectively reduce lipid peroxidation, upregulate GSH and NADPH levels, and inhibit ferroptosis (Ghio et al., 2008a; Yoshida et al., 2019; Tang et al., 2021a; and Lian et al., 2021). Moreover, recent reports revealed that antioxidants, such as N-acetyl-L-cysteine (NAC) and curcumin, could improve the reduction of GSH and reduce lipid peroxidation (Tang et al., 2021a), while dihydroquercetin could inhibit ferroptosis in lung epithelial cells by activating the Nrf2-mediated pathway (Liu et al., 2022a).

### Table 1 (Continued) Reagents and mechanisms of ferroptosis in lung diseases.

| Disease | Reagent | Proposed mechanism | References |
|---------|---------|--------------------|------------|
| Deferiprone | Chelate iron; inhibit microbial activity with better tissue penetration capacity | Kontogiorgioghes et al. (2010); Álvarez et al. (2013) |
| Deferoxin | Chelate iron; inhibit microbial activity | Kontogiorgioghes et al. (2010); Álvarez et al. (2013) |
| Infection: A. Fumigatus | Deferoxamine | Chelate iron and block iron-dependent lipid peroxidation | Hsu et al. (2018) |
| | Deferiprone | Chelate iron and block iron-dependent lipid peroxidation | Hsu et al. (2018) |
| | Deferoxin | Chelate iron and block iron-dependent lipid peroxidation | Hsu et al. (2018) |
| Acute lung injury/acute respiratory distress syndrome | iASPP | Regulate p53 and NRF2 | Li et al. (2020a) |
| | Ferrostatin-1 | Inhibit lipid peroxidation | Liu et al. (2020) |
| | Liproxstatin-1 | Inhibit lipid peroxidation | Li et al. (2020a) |
| | Panaxadil | Regulate KEAP1/NRF2/HO-1 axis; activates GSH and GPX4 | Li et al. (2021b) |
| | 4-octyl itaconate | Regulate NRF2/HO-1 axis; activates GSH and GPX4 | He et al. (2022b) |
| | Obacunone | Regulate NRF2/HO-1 axis; activates GSH and GPX4 | Li et al. (2022) |
| | Puerarin | Activate SLC7A11, GPX4, and FTH1; inhibit NOX1 to reduce lipid ROS generation | Xu et al. (2021a) |
| | Acupuncture | Activate SLC7A11, GPX4, and FTH1 | Zhang et al. (2022) |
| Lung cancer/metastasis | Erastin | Block cysteine import, inducing GSH depletion and GPX4 inactivation | Huang et al. (2018); Pan et al. (2019) |
| | RSL3 | Covalently inhibit GPX4, causing accumulation of lipid peroxidation | Zhang et al. (2020b); Li et al. (2021b) |
| | Cisplatin | Block cysteine import, inducing GSH depletion and GPX4 inactivation | Guo et al. (2018) |
| | PRL93938 | Regulate NRF2/HO-1 axis; GSH depletion and GPX4 inactivation | Li et al. (2021) |
| | Imidazole ketone erastin | Block cysteine import, inducing GSH depletion and GPX4 inactivation | Ye et al. (2020) |
| | Sorafenib | Block cysteine import, inducing GSH depletion and GPX4 inactivation | Li et al. (2020b) |
| | Levobupivacaine | Increases iron abundance and mediate iron-dependent lipid peroxidation | Meng et al. (2021) |
| | BEBT-908 | Regulate p53 and STAT1 | Fan et al. (2021) |
| | iFSP1 | Inhibit FSP1−CoQ10−NAD(P)H system | Doli et al. (2019) |
| | Erlarin | Induce GSH depletion and mediate iron-dependent lipid peroxidation | Chen et al. (2020b) |
| | Sanguinarine | Inhibit GSH and GPX4 | Xu et al. (2022) |
| | Dihydroisotanshinone I | Inhibit GSH and GPX4 | Wu et al. (2021) |

**Asthma**

Asthma is a chronic inflammatory respiratory disorder that results in intermittent episodes of wheezing, breathlessness, chest stuffiness, and cough. The distinguishing features of the
disease are sporadic and reversible airway obstruction, chronic bronchial inflammation, bronchial smooth muscle cell hypertrophy and hyperreactivity, and increased mucus secretion (Papi et al., 2018). The function of type-2 helper T (Th2) cells is critical to the pathologic process of asthma. Upon activation, Th2 cells produce cytokines and active mediators, such as interleukin-4 (IL-4), which stimulate IgE secretion by B cells, IL-5 triggers eosinophils, and IL-13 arouses mucus and IgE production. Meanwhile, mast cells, macrophages, neutrophils, and T cells are activated and gather in the airways and secrete inflammatory mediators, resulting in chronic airway inflammation (Hammad and Lambrecht, 2021). Continuous studies have demonstrated that airway epithelial cells from asthmatic patients actively produced 15-lipoxygenase-1 (15-LOX1) and 15-hydroxyeicosatetraenoic acid (15-HETE). The reports demonstrated that 15LOX1-phosphatidylethanolamine-binding protein 1 (PEBP1) complex activation consistently stimulates IL-13/IL-4-mediated Th2 inflammation, including mucin 5AC (MUC5AC), periostin (POSTN), and chemokine (C-C motif) ligand 26 (CCL26). 15-LOX1-PEBP1 can activate autophagy and ferroptosis. Besides, there was a positive correlation between the colocalization of 15LOX1/PEBP1 puncta in freshly brushed airway epithelial cells and the fraction of exhaled nitric oxide (FeNO) in asthmatic patients. Ferroptosis inducing agents, namely, erastin, RSL3, and artesunate (ART),

![FIGURE 2 | Ferroptosis in lung diseases. Ferroptosis contributes greatly to the pathogenesis of lung diseases, including chronic obstructive pulmonary disease, asthma, cystic fibrosis, pulmonary fibrosis, ischemia-reperfusion injury, pulmonary infections, acute lung injury and acute respiratory distress syndrome, and lung cancer and metastasis.](image-url)

Eosinophils are vital in allergic disorders, such as asthma, and prompting eosinophil death, which effectively weakens inflammation (Hammad and Lambrecht, 2021). The increased number and prolonged survival time of eosinophils in asthma suggest the selected targeting of ferroptosis as a promising therapeutic strategy for airway inflammation. Ferroptosis-inducing agents, namely, erastin, RSL3, and artesunate (ART),
were proven to induce eosinophil ferroptosis and collaborate with dexamethasone to promote eosinophil death. Targeted administration of eosinophil ferroptosis appeared to relieve allergic airway inflammation and might cut the dosage and reduce the side effects of glucocorticoids in an OVA-induced asthma model (Wu et al., 2020). The concept of inducing ferroptosis of specific immune cells to diminish airway inflammation and alleviate mediator production is promising but needs to be investigated for its clinical potential in future studies of asthma.

**Cystic Fibrosis**

Cystic fibrosis (CF) is an autosomal recessive genetic disease that causes dysfunction of ion transport in the exocrine glands (Ratjen and Döring, 2003). It is the most common fatal genetic disorder among white populations. The disease is due to defects in the cystic fibrosis transmembrane conductance regulator (CFTR) gene, resulting in abnormalities in chloride transport in epithelial transport affecting fluid secretion in exocrine glands and respiratory, gastrointestinal, and reproductive tracts. Recurrent respiratory infection and pancreatic dysfunction are the two most important clinical symptoms (Prentice et al., 2021). In patients with CF, the excretion of chloride ions in the airway is reduced and the resorption of sodium and water is increased, which results in dehydration of mucous layer coating epithelial cells, loss of mucociliary function, mucus blockage, and mucous plugging. A recent report demonstrated that CF airway epithelial cells exposed to ferric ammonium citrate (FAC) and the ferroptosis inducer erastin are susceptible to ferroptosis compared to isogenic CFTR-corrected airway epithelial cells in vitro (Maniam et al., 2021). Ferrostatin-1 and deferoxamine are able to inhibit ferroptosis by alleviating iron accumulation and lipid peroxidation and thus, may be potential therapeutic interventions for CF. On the other hand, patients with CF commonly develop respiratory infections caused by *Pseudomonas aeruginosa*, *Staphylococcus aureus*, *Hemophilus influenzae*, and *Burkholderia cepacia*. Among these, *P. aeruginosa* is an important opportunistic pathogen responsible for the ferroptotic cell death in airways, as discussed later in the section on pulmonary infections.

**Obstructive Sleep Apnea**

Obstructive sleep apnea (OSA) is a respiratory disease characterized by an intermittent nocturnal decrease in oxygenated hemoglobin and sleep interruption (Patel et al., 2019). The main feature of OSA is chronic intermittent hypoxia (CIH) from the view of pathophysiology. Cardiovascular diseases, liver diseases, and metabolic diseases are the major health risks to OSA patients. Although there is no report of ferroptosis caused by hypoxia-reperfusion in lung diseases, oxidative stress, and lipid peroxidation have been reported to induce liver injury in OSA (Chen et al., 2020a; Chen et al., 2021a). Both reports revealed that ferroptosis mediated liver injury induced by CIH in rodent models, as evidenced by the increase in lipid peroxidation, the decrease in GPX4 expression, and the increase in ACSL4 expression.

**Pulmonary Fibrosis**

Idiopathic pulmonary fibrosis (IPF) refers to a refractory and irreversible progressive pulmonary fibrotic disorder without clear etiology. The characteristic feature of IPF is patchy but progressive bilateral interstitial fibrosis, leading to severe hypoxemia and cyanosis in advanced cases (Lederer et al., 2018). Recurrent epithelial activation and injury is the leading cause of IPF occurrence. Epithelial repair deficiency and inflammation at damage sites give rise to the vigorous proliferation of fibroblasts and myofibroblasts, resulting in typical fibroblastic foci. Ferroptosis-related genes, which includes neuroblastoma ras viral (v-ras) oncogene homolog (NRAS) and MUC1, but not limited to them, were elevated in bronchoalveolar lavage fluid and cells in IPF patients, which suggests the participation of ferroptosis in the disease and may be used for prognostic prediction of IPF (Li et al., 2021a; He et al., 2021; He et al., 2022a).

Pulmonary fibrosis (PF) is an interstitial lung disease that derives from long-term inhalation of CS and dust, such as asbestos, silica, and coal, the use of drugs, such as bleomycin and amiodarone, accidental exposure to paraquat, or lung injury are caused by radiation therapy. A previous study showed that erasin promoted transforming growth factor β1 (TGF-β1)-induced fibroblast-to-myofibroblast differentiation by promoting ROS and lipid peroxidation and hindering GPX4 expression, resulting in collagen accumulation and destruction of the alveolar structure, while ferrostatin-1 may inhibit this process (Gong et al., 2019). A recent report demonstrated that the upregulated long noncoding RNA (lncRNA) zinc finger antisense 1 (ZFAS1) is positively correlated with SLC38A1 expression in bleomycin-induced PF rat lung tissue and in TGF-β1-induced human fetal lung fibroblast cells. Experiments have shown that inhibition by knockdown or silencing of the lncRNA ZFAS1 can significantly attenuate lipid peroxidation and inflammation, thus, inhibiting ferroptosis and progression of PF induced by bleomycin (Yang et al., 2020). Another study demonstrated that SET domain bifurcated 1 (SETDB1) and H3K9me3 expression was downregulated in a bleomycin-induced PF rat model, leading to the induction of epithelial-mesenchymal transition and increased lipid ROS, ferrous ions, and ferroptosis (Liu et al., 2022b). On the other hand, inducing ferroptosis by erasin could elicit iron accumulation, ROS production, epithelial-mesenchymal transition, and autophagy in lung epithelial cell lines, as evidenced by upregulated microtubule-associated protein 1A/1B-light chain 3 (MAP1LC3) and Beclin 1 (BECN1) (Han et al., 2021; Sun et al., 2021). Links between pulmonary fibrosis and ferroptosis are complicated and still under investigation. Experimental interventions, such as deferoxamine, effectively reduced iron accumulation and lipid peroxidation, thus, inhibiting ferroptosis in a bleomycin-induced PF cell and mouse model (Cheng et al., 2021; Takahashi et al., 2021).

Asbestos is a silicate mineral containing iron, magnesium, and calcium with a core of SiO₂. It has been established that asbestos fibers tend to accumulate near the mesothelial cell layer, where they produce ROS, leading to DNA damage and potential carcinogenic mutations. Workers exposed to asbestos can...
develop lung cancer and malignant mesothelioma. Iron homeostasis is changed among patients with asbestososis, as evidenced by the accumulation of iron, ferritin, divalent metal transporter 1 (DMT1), and ferroportin 1 (FPN1) in the lung autopsies. Iron chelators, including defereroxamine and phytic acid, were utilized in asbestos- and silica-induced PF animal models, which shows that procollagen and inflammation could partly reversed, whereas glutathione had no effect.

Paraquat (PQ) is an economic and effective herbicide that could cause multiple organ acute injury, lung fibrosis, multiple organ failure, and death due to unintentional or intentional inhalation, ingestion, and percutaneous absorption. Evidence of the connection between PQ poisoning and ferroptosis is emerging, confirmed by the formation of high-energy oxygen free radicals and lipid peroxidation (Rashidipour et al., 2020). The hypothesis that ferroptosis inhibitors might be introduced in PQ-induced PF was raised and needs to be confirmed.

Radiation-induced lung fibrosis (RILF) is a life-threatening complication after radiotherapy of chest tumors. Collagen deposition, decreased GPX4 expression, and increased ROS were observed in the mouse model, indicating that ferroptosis was involved when exposed to radiation. The ferroptosis inhibitor liproxstatin-1 alleviated RILF by activating the Nrf2 pathway in this experimental study (Li et al., 2019). Therefore, strategies for regulating iron metabolism and controlling ferroptosis can be exploited to delay the progression of PF in future experimental and clinical practice.

**Lung Ischemia-Reperfusion Injury**

An ischemia injury arises at the initial stage of vascular compromise, however, airway epithelial cells are relatively resistant to transient hypoxia. Reperfusion injury is caused by recovery of blood flow and is related to overwhelming damage. Multiple organ failure might arise in serious cases. Although the underlying mechanisms of reperfusion injury are not fully understood, they involve the production of free radicals, neutrophil infiltration, and the secretion of inflammatory mediators, namely, cytokines, chemokines, and complements. Clinical disorders associated with the development of ischemia-reperfusion include pulmonary thromboembolism and thrombosis, lung resection and trauma, lung transplantation, and other operations. Increased iron content and lipid peroxidation accumulation, together with decreased GPX4 and elevated ACSL4 expression, were detected in ischemia-reperfusion lungs. Treatment with liproxstatin-1 to inhibit ferroptosis, administration of the ACSL4 inhibitor rosiglitazone before ischemia, and ACSL4 knockdown ameliorated lung ischemia-reperfusion injury by protecting against ferroptotic damage in animal and cell models (Xu et al., 2020). Irisin, a novel muscle-derived myokine, was reported to suppress ferroptosis in lung ischemia-reperfusion damage in vitro and vivo, as confirmed by lower ROS, malondialdehyde (MDA), and iron accumulation, along with alterations in GPX4 and ACSL4 (Wang et al., 2022a). Lidocaine attenuated inflammation, apoptosis, and ferroptosis in a lung epithelial cell line by controlling the p38/mitogen-activated protein kinase (MAPK) pathway (Ma et al., 2022).

Furthermore, there were elevated iron concentrations in alveolar fluid and tissue in human lung allografts (Baz et al., 1997), which could increase the risk of lung allografts to iron radicals, ROS, fibrosis, and chronic rejection. An early study demonstrated that HO-1 expression was elevated in human lung allografts with acute cellular rejection and obliterator bronchiolitis (Lu et al., 2002). Recipient iron overload and hyperferritiniemia were also associated with poor prognosis after lung transplantation (Pugh et al., 2005). Administrations for optimizing iron homeostasis before and after lung transplantation were considered. Different iron chelators, namely, defereroxamine, were used in lung preservation and alleviated oxidative stress after transplantation (Lui et al., 2013). Pirfenidone significantly reduced the deposition of iron, increased the expression of HO-1, and alleviated fibrosis and collagen deposits (Liu et al., 2005), demonstrating anti-ferroptosis, anti-fibrotic and antioxidative properties in preventing chronic airway rejection in a rat model.

Targeted inhibition of ferroptosis may be a potential way to protect against ischemia-reperfusion injury. By reducing the oxidative stress response, the cell damage caused by ferroptosis can be avoided to reduce the occurrence of complications.

**Pulmonary Hypertension**

Pulmonary hypertension (PH) is usually secondary to a decrease in vessel diameter or an increase in blood flow in the pulmonary vascular bed (Vonk Noordegraaf et al., 2016). Less commonly, PH caused by unknown causes is called primary or idiopathic PH. At present, the dysfunction of pulmonary endothelial and/or vascular smooth muscle cells is considered to be the potential basis for most forms of PH. The proliferation of endothelial and smooth muscle cells leads to thickening of the intima and media with narrowing of the lumina in the entire pulmonary arterial tree. Dysregulated iron homeostasis in the pathological mechanism of PH is not clear and fully recognized. Investigations have established that pulmonary vascular function could be affected by intracellular iron deficiency in pulmonary artery smooth muscle cells (Rhodes et al., 2011), and rats fed on an iron-deficient diet showed substantial pulmonary vascular remodeling and muscularization, medial hypertrophy, perivascular inflammatory cell infiltration strongly associated with rising pulmonary artery pressure (PAH), and right ventricular hypertrophy (Cotroneo et al., 2015), which suggests that iron metabolism participates in the homeostasis of the pulmonary vasculature and that abnormal iron metabolism takes part in the occurrence and development of PH. In contrast, iron deposition was noted in lung tissue sections and was highly correlated with advanced PH in IPF patients (Kim et al., 2010). Moreover, the administration of the iron chelator defereroxamine prevented pulmonary vascular remodeling in chronic hypoxia-induced PH rats (Wong et al., 2012). Whether iron deficiency or overload leads to PH or is merely a consequence is still under debate. Recently, bioinformatic analyses of unregulated ferroptosis-associated genes were performed, and the targets and potential drugs were predicted. However, there were contradictions between the dysregulation and function of some key genes, such as SLC7A11, in these...
studies (Zhang et al., 2021; Zou et al., 2021). In another recent study, ferroptosis of pulmonary artery endothelial cells (PAECs) was reported to play a critical role in the progression of PH in a monocrotaline-induced model, proven by increased lipid peroxidation, cellular iron concentrations, mitochondrial damage, abnormal expression of GXP4, ferritin heavy chain 1 (FTH1), and NADPH oxidase 4 (NOX4), followed by activation of inflammatory factors and pulmonary artery remodeling. Ferroptosis suppression by ferrostatin-1 postponed lung vascular remodeling and protected right ventricle function in PH (Xie et al., 2022). Based on these findings, a thorough understanding of ferroptosis in PH is needed, and treating PH with medicines based on ferroptosis regulation might be promising in the future.

**Pulmonary Infections**

Deaths from pulmonary infections in the form of pneumonia are not rare worldwide. The epithelial surfaces of the lung are consistently exposed to open air containing microbial contaminants, and other common lung diseases and unhealthy lifestyles, such as smoke and alcohol use, render the lung parenchyma vulnerable to virulent organisms.

Although previously discussed, *P. aeruginosa* is associated with infections in CF patients, and it is most commonly seen in the hospital environment. Based on reports, *P. aeruginosa* can express lipoxygenase (pLoxA) after infection, oxidize host arachidonoyl-phosphatidylethanolamine (ETE-PE) to pro-ferroptotic 15-hydroperoxy-arachidonyl-PE (15-HpETE-PE), and cause ferroptosis of human bronchial epithelial cells (Dar et al., 2018). In addition, *P. aeruginosa* can degrade the host GPX4 defense by stimulating lysosomal chaperone-mediated autophagy (CMA) (Dar et al., 2021). Baicalein inhibits LOX-mediated ferroptotic pathways, which may be a possible target for the treatment of respiratory infections (Dar et al., 2022). In another study, knockdown of anoctamin 1 (ANO1 or TMEM116A), antioxidants, such as coenzyme Q10 (CoQ10) and idebenone, and ferrostatin-1 attenuated *Pseudomonas* aeruginosa-induced cell death in CF bronchial epithelial cells (Ousingsawat et al., 2021).

Pulmonary *tuberculosis* (TB) is the major infectious disease within the spectrum of chronic pneumonia that seriously endangers human health. The World Health Organization (WHO) estimates that TB causes 6% of the world’s deaths and is now deteriorating the condition more worldwide. Immunity to *tuberculosis* infection is mainly driven by Th1 cells, which trigger macrophages to kill bacteria (Jasenosky et al., 2015). Re-exposure to *Mycobacterium tuberculosis* (Mtbb) or reactivation of the infection in a previously sensitized host stimulates a swift definitive reaction, but hypersensitivity also increases tissue necrosis and tissue destruction. Mtbb-induced macrophage death was related to a decrease in GSH and GPX4 levels and an increase in free iron, mitochondrial superoxide, and lipid peroxidation, which indicates that ferroptosis was involved in tissue necrosis in Mtbb infection (Amaral et al., 2019). Mtbb also increased the expression of HO-1, which may in turn facilitate Mtbb survival and growth as a consequence of increased iron availability (Yang et al., 2022). The destruction of macrophages by Mtbb and bacterial load were reduced after ferrostatin-1 treatment (Amaral et al., 2019). The primary anti-tuberculosis drugs isoniazid (INH) and rifampicin (LFP) usually exhaust GSH and cause lipid peroxidation and ferroptosis of hepatocytes during liver metabolism. GSH replenishment prevented injury, while iron supplementation augmented the ferroptosis process (Pan et al., 2020). Vitamin E (vit E) was used to intervene in *tuberculosis* patients in clinical trials. The results showed that MDA levels were reduced, and the plasma total antioxidants were improved in *tuberculosis* patients after 2 months of administration (Seyedrezazadeh et al., 2008). This can be explained by the fact that vit E can reduce nonheme iron from ferric iron to ferrous iron to inhibit 15-LOX1 (Hinman et al., 2018), therefore, preventing the ferroptosis pathway regulated by 15-LOXI (Kagan et al., 2017).

Ferroptosis inhibitors were also investigated for the treatment of resistant pathogens, such as liproxstatin-1, which was proven to interact with lipopolysaccharide in the outer leafllet to disrupt the integrity of the outer membrane of Gram-negative bacteria and potentially serve as an outer membrane permeabilizing compound (Klobucar et al., 2021).

The coronavirus disease 2019 (COVID-19) outbreak caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is an ongoing global health emergency, and the pathologic process of the virus has not been completely elucidated. The clinical symptoms are mostly cough, fever, chest distress, muscle aches, fatigue, dyspnea, and headache (Huang et al., 2020a; Wang et al., 2020). Conditions including acute respiratory distress syndrome (ARDS), septic shock, severe metabolic acidosis, and a hypercoagulable state are life-threatening in severe cases with COVID-19, and respiratory failure is the most common cause of death. The pathological lesions include hemoglobinopathy, hypoxia, and hyperferritinemia (Zhao et al., 2020b; Cavezzi et al., 2020; and Colafrancesco et al., 2020). Cellular iron overload is believed to play a pivotal role in COVID-19 infection. Elevated serum ferritin can further aggravate systemic inflammation, which is closely related to poor prognosis (Huang et al., 2020b; Edeas et al., 2020; and Zhou et al., 2020). Considering that the expression of transferrin and the risk of severe cases in male patients were higher than those in female patients and increased with age, this relationship could explain the higher infection rate and mortality in elderly male COVID-19 patients. Transferrin could deliver iron to cells by binding the transferrin receptor and mediating endocytosis. The higher levels of the soluble transferrin receptor in COVID-19 patients confirmed this possibility (Duca et al., 2021). It remains unclear whether hyperferritinemia is a systemic marker or a key modulator in the pathogenesis of COVID-19. Iron is engaged in several biological processes involving DNA, RNA, and ATP synthesis. There is evidence that the replication of SARS-CoV-2 depends on iron-containing enzymes. Regulating host iron metabolism can play an antiviral role and prolong cell survival. Iron is also responsible for SARS-CoV-2 replication, which include key steps, such as ATP hydrolysis (Jia et al., 2019). Moreover, SOD and lipid peroxidation levels were significantly elevated in COVID-19 patients, while GSH and GPX4 levels were decreased (Silvagno et al., 2020). Changes in iron metabolism,
GSH depletion, GPX4 inactivation, and upregulation of lipid peroxidation may lead to the hypothesis that ferroptosis may be triggered after SARS-CoV-2 infection, which results in damage to multiple organs. Early treatments with the antioxidants vitamin C (Vit C), vitamin E (Vit E), N-acetylcysteine (NAC) and melatonin (MT) with pentoxifylline were reported to scavenge ROS, supply GSH, and delay the aggression and death of COVID-19 (Jaiswal et al., 2020; Chavarria et al., 2021). Iron chelators, such as deferiprone, deferasirox, deferoxamine exhibited iron chelating, antiviral, and immunomodulatory effects in vitro and in vivo. Critically ill patients can benefit from orally given deferasirox or intravenously given deferoxamine. Several mechanisms are involved in this process: a. inhibiting viral replication; b. reducing iron availability; c. upregulating B cells; d. increasing neutralizing antibody titer; e. inhibiting endothelial cell inflammation; and f. preventing lung fibrosis and pulmonary recession by reducing pulmonary iron accumulation (Cavezzi et al., 2020; Dalamaga et al., 2020). However, there have been debates on the use of deferoxamine or hепcidin antagonists, whereas it was declared that deferoxamine could also downregulate the expression of hepcidin I (Abobaker, 2021; Garrick and Ghio, 2021). Lactoferrin is a naturally occurring, nontoxic glycoprotein that contains more pertinence to binding iron than transferrin and has demonstrated antiviral efficacy against different kinds of viruses in vitro, including SARS-CoV, and might potentially serve as a preventative and adjunct treatment for COVID-19 (Carota et al., 2021). Theoretically, drugs, for instance, methemoglobin reductase, ascorbic acid (Vit C), and GSH were inferred to reduce ferric iron to ferrous iron in hemoglobin to restore its ability to combine with oxygen to alleviate the symptoms of hypoxia in COVID-19 patients (Muhoberac, 2020). In conclusion, reducing cellular iron and replenishing the level of reductants are the most basic treatment methods to lessen tissue injury in COVID-19 patients. These drugs also ought to be investigated in future clinical studies to confirm their safety and effectiveness.

Mucormycosis is caused by a fungus called Zygomycetes. It is a life-threatening opportunistic mycosis that is generally confined to immunocompromised patients, especially those with hematolymphoid malignancies or severe neutropenia, those receiving corticosteroids, or other immunosuppressive drugs, allogeneic stem cell transplant recipients, poorly controlled diabetes mellitus (DM), and those with an iron overload state. Evidence suggests that iron metabolism and fungal endothelial cell interactions play an important role in the pathogenesis of mucormycosis (Hamilos et al., 2011). In a recent report, both infected and recovered COVID-19 patients were promptly infected with mucormycetes (Pasrija and Naime, 2022). The reason might be that COVID-19 patients with elevated serum levels of available iron were susceptible to mucormycosis, and these infections are highly angioinvasive, since the pathogens could acquire iron from the host and interact with endothelial cells lining blood vessels (Pasrija and Naime, 2022). In addition, SARS-CoV-2 increased the susceptibility of patients to mucormycosis by augmenting the virulence factors of the Mucor species. Iron chelator therapy may be advantageous to treat the infection by correcting and inappropriately supplying the fungus with iron. However, there was evidence that the new oral iron chelators deferiprone and deferasirox, better than deferoxamine, could deteriorate the growth of fungi both in vitro and in animal models. The iron liberated from deferoxamine was likely transported into the fungus by the high-affinity iron permease, thus, promoting infections. Deferiprone showed the highest antimicrobial activity and tissue penetration capacity, particularly access to the brain (Kontoghiorghes et al., 2010; Álvarez et al., 2013). Melatonin (MT) is an iron chelator, calmodulin blocker, and inhibitor of myeloperoxidase along with an inhibitor of ferroptosis and pyroptosis. By correcting MT deficiency, the enhancement of fungal virulence in COVID-19 patients was alleviated because MT could hinder the iron acquisition of Mucor species and prevent their morphological transformation from yeast to the virulent hyphal form (Sen, 2021).

Aspergillus fumigatus is the most common airborne fungal pathogen and is accountable for invasive aspergillosis in immunocompromised hosts. The acquisition of iron is important for the growth of Aspergillus fumigatus. This fungus synthesizes and secretes triacetylfusarinine C to capture iron and accumulates ferricrocin and hydroxyferricrocin to store iron for hyphae and conidia. Meanwhile, it decreased the expression of the iron importer DMT1 and the transferrin receptor and iron exporter FPN1 (Seifert et al., 2008). In lung transplant recipients, Aspergillus fumigatus infection can be life-threatening. The microhemorrhage-related iron content in the graft might be the main determining factor of invasion and virulence of infection, and progressive graft rejection was related to the increase in ferric iron concentration. Iron chelation, including deferiprone, deferasirox, and deferoxamine, maybe a potential therapy for Aspergillus fumigatus (Hsu et al., 2018), but the effects can be paradoxical, thus, chelators must be chosen carefully.

**Acute Lung Injury/Acute Respiratory Distress Syndrome**

The pulmonary infiltrates in ALI are caused by damage to the alveolar-capillary membrane, consisting of the microvascular endothelium and the alveolar epithelium. The acute results of damage include increased vascular permeability and alveolar flooding, impaired diffusion capacity, and extensive surfactant abnormalities. ALI can progress to more severe diffuse alveolar damage and is known as ARDS in the setting of sepsis, severe trauma, or a diffuse lung infection. The clinical features are the emergence of life-threatening respiratory dysfunction, cyanosis, and hypoxemia that is refractory to oxygen therapy and rapidly progresses to multisystem organ failure. Neutrophils and their products, e.g., oxidants, proteases, platelet-activating factor, and leukotrienes, play a crucial role in the pathologic process of ARDS by causing damage to the alveolar-capillary membrane. On the other hand, the endogenous antiproteases, antioxidants, and anti-inflammatory cytokines that counteract the destruction and balance determine the degree of tissue injury and severity of clinical symptoms in ARDS. The characteristic finding in ARDS is the presence of hyaline membranes...
consisting of fibrin-rich edema fluid admixed with remnants of necrotic epithelial cells, particularly lining the distended alveolar ducts.

In ALI mouse models induced by intestinal ischemia-reperfusion and oleic acid, mitochondrial shrinkage, and mitochondrial membrane rupture were noted in type II alveolar epithelial cells (AEC2). The characteristic indicators of ferroptosis, namely, iron overload, GSH depletion and MDA accumulation, and downregulated GPX4 and ferritin in lung tissue were also detected (Zhou et al., 2019; Dong et al., 2020). In a mouse model of ALI-induced by intestinal ischemia-reperfusion, Nrf2 expression can inhibit ferroptosis via modulation of telomerase reverse transcriptase (TERT), HO-1, and SLC7A11 levels (Dong et al., 2020; Dong et al., 2021). In another study, the overexpression of inhibitor of apoptosis stimulating protein of p53 (iASPP) and Nrf2 exhibited therapeutic effects. iASPP inhibited ferroptosis and alleviated tissue injury, depending on Nrf2/hypoxia-inducible factor 1 (HIF1) signal transduction (Li et al., 2020a), Nrf2 upregulation also alleviated GPX4 decreases and attenuated signal transducer and activator of transcription-3 (STAT3) phosphorylation (pSTAT3). STAT3 enhanced the antioxidant capacity through SLC7A11 activation, thereby attenuating the development of ferroptosis during the disease (Qiang et al., 2020). In lipopolysaccharide (LPS)-induced ALI animal models and bronchial epithelial cell lines, the levels of MDA, 4-hydroxynonenal (4-HNE), and total iron were dramatically decreased, indicating that ferroptosis was involved in LPS-induced ALI (Liu et al., 2020). Ferroptosis inhibitors, including ferrostatin-1 and liproxstatin-1, showed a protective effect in these ALI/ARDS models. In addition, a System Xc- (xCT) derived from Panax ginseng root was utilized in LPS-induced ALI/ARDS, and the results showed that PX lessened the pathological lesions in mouse lungs, inhibiting ferroptosis by upregulating the Kelch-like ECH-associated protein 1 (KEAP1)/Nrf2/HO-1 pathway (Li et al., 2021b). Similarly, 4-ocetyl itaconate (4-OI) and obacunone (OB) were reported to significantly alleviate lung injury, increase GSH and GPX4, and reduce malonaldehyde and lipid peroxidation by activating Nrf2 and HO-1 in vivo (He et al., 2022b; Li et al., 2022). Puerarin (PUE) (Xu et al., 2021a), silencing or knockdown of mixed lineage kinase 3 (MLK3) (Chen et al., 2022), lipocalin-2 (Wang et al., 2022b) and Jumonji domain-containing 3 (JMJD3) (Peng et al., 2021), and electroacupuncture (EA) (Zhang et al., 2022) presented novel targets for the treatment of LPS-induced ALI/ARDS. These studies preliminarily confirmed that ferroptosis is intricately connected to ALI/ARDS and that it can be a novel therapeutic target.

**Pulmonary Alveolar Proteinosis**

Pulmonary alveolar proteinosis (PAP) is an infrequent disease characterized by the aggregation of surfactant and phospholipids in the distal airways and alveoli. The causative factors in primary PAP are still unknown, but it is postulated to be an autoimmune disorder since the emergence of antibodies to granulocyte-macrophage colony-stimulating factor (GM-CSF). Disturbance of iron homeostasis in epithelial cells and macrophages in the lung has been reported in idiopathic PAP patients. Several early reports demonstrated that the contents of iron, transferrin, transferrin receptor, lactoferrin, and ferritin were remarkably elevated in lavage from PAP patients in comparison with healthy controls, while the concentrations of ascorbate, glutathione, and urate were remarkably lower. The cells of PAP patients accumulated significant iron and ferritin. Immunohistochemistry for the lung tissue revealed the accumulation of ferritin in the lower respiratory tract of PAP patients (Ghio et al., 2008b; Shimizu et al., 2011). This led to the hypothesis that ferroptosis might participate in the development of PAPs; however, there is neither direct evidence to prove the correlation between ferroptosis and PAPs nor intervention experiments to evaluate the effect of medicines based on ferroptosis regulation on PAPs.

**Lung Cancer**

Lung cancer has the highest incidence rate and mortality rate in the world. Accumulated studies have found that ferroptosis has a close relationship with lung cancer and tumor metastasis, and lung cancer cells are in a state of ferroptosis inhibition.

a System Xc- (xCT) is a cystine/glutamate antiporter consisting of SLC3A2 and SLC7A11, which is accountable for exporting glutamate and importing cysteine. By upregulating SLC3A2 and SLC7A11, lung cancer cells could enhance their antioxidant effect, inhibit the occurrence of ferroptosis, and increase drug resistance to inducers of ferroptosis, such as imidazole ketone estrogen (IKE) and cisplatin (DDP) (Huang et al., 2005; Ma et al., 2021; Tabnak et al., 2021; and Wang et al., 2021).

b The antioxidant enzymes GPXs, GPX8, and GPX4 in particular, have been identified to play an extensive and broad role in the pathological process of cancers (Zhang et al., 2020a). Overexpression of GPX4 promoted the proliferative capacity of lung cancer cells and inhibited ferroptosis, whereas RSL3 hindered GPX4 activity and limited the proliferation, migration, invasion, and angiogenesis of lung cancer cells (Zhang et al., 2020a; Tabnak et al., 2021). Besides, these antioxidants can protect metastasizing cancer cells in both circulations and the metastatic niche to resist ferroptosis (Liu et al., 2021a). Targeting the GPX4 pathway may provide a new strategy for treating lung cancer growth and metastasis.

c The tumor-suppressive activity of p53 has been proposed and proven after decades of intensive study. Mutation of p53 cannot repress SLC7A11 and promote ferroptosis (Jiang et al., 2015b). In addition, the p53 P47S polymorphism, commonly found in people of African descent, is also defective in promoting ferroptosis and repressing tumor development (Leu et al., 2019). Targeting p53 can potentially improve the efficiency of lung cancer treatment by mediating ferroptotic responses.

d Ferroptosis suppressor protein 1 (FSP1) is a ferroptosis inhibitor that is independent of the GPX4 pathway and could suppress ferroptosis via CoQ10 (Doll et al., 2019). FSP1 is decorated with cardamom acylation by regulating NADPH to reduce CoQ10, producing lipophilic free radicals to capture free antioxidants to prevent lipid
peroxidation to suppress ferroptosis (Doll et al., 2019). High expression of FSP1 could lead to increased resistance in lung cancer cells (Doll et al., 2019).

e MicroRNAs (miRNAs) are a class of noncoding RNAs (ncRNAs) with a length of 18–25 nucleotides that could impede protein translation by regulating their target mRNAs. Tumor suppressor miRNAs are negatively regulated in cancers and usually target oncopgenic proteins. Downregulated tumor suppressor miRNAs that induce ferroptosis, such as miR-302a-3p and miR-324-3p, can promote survival and proliferation, and their overexpression sensitizes resistant cells to ferroptosis and increases the sensitivity of chemotherapeutic drugs (Deng et al., 2021; Wei et al., 2021).

f LncRNAs engage in the occurrence and development of non-small-cell lung cancers (NSCLCs) by mediating ferroptosis. LIN00472 (P53RRA) activates the p53 pathway by interacting with Ras GTPase-activating protein-binding protein 1 (G3BP1) and induces ferroptosis (Mao et al., 2018). P53RRA promotes erastin-induced growth inhibition and increases the cellular iron and lipid ROS concentrations in NSCLC cells (Mao et al., 2018). Low expression of P53RRA removed p53 and weakened ferroptosis in NSCLC (Mao et al., 2018). In addition, overexpressed LIN00336 acted as a crucial ferroptosis inhibitor in lung cancer by lowering cellular iron, lipid peroxidation, and mitochondrial superoxide through ELAV-like RNA-binding protein 1 (ELAVL1) interactivity, a novel regulator of ferroptosis (Wang et al., 2019). LIN00336 also served as an endogenous sponge of MIR6852 as a circulating extracellular DNA (ceRNA) to increase cystathionine-β synthase (CBS) expression and inhibit ferroptosis in lung cancer (Wang et al., 2019). Overexpression of LIN00336 limited RSL3-induced ferroptosis in lung adenocarcinoma cells.

These studies established that resistance to ferroptosis was enhanced from multiple aspects in lung cancer and metastasis. The first thing that comes to mind is that the promising ferroptosis inducers erastin and RSL3 can inhibit the biological process of System Xc- and GPXs (Huang et al., 2018; Pan et al., 2019; Zhang et al., 2020b; Liu et al., 2021b). Other signaling pathways can be involved in these ferroptosis inducers. For example, erastin can activate the p53 signaling pathway, subsequently inhibiting the expression of SCL7A11 posttranscriptionally and subsequently inducing ferroptosis (Huang et al., 2018). DDP, a classical chemotherapeutic drug, was recently proven to trigger ferroptosis in NSCLC by inhibiting the activity of the GSH-GPX system and promoting the therapeutic effect together with erastin (Guo et al., 2018). Similarly, an analog of erastin, PRLX93936, can induce ferroptosis via GPX4 inhibition when combined with DDP (Li et al., 2021). Novel ferroptosis inhibitors, such as IKE and sorafenib, can inhibit the cystine/glutamate transporter, exhaust GSH, and increase sensitivity to radiotherapy (Li et al., 2020b; Ye et al., 2020). The local anesthetic levobupivacaine could increase p53 expression, enhance ferroptosis, and inhibit tumor growth in NSCLC (Meng et al., 2021). The dual PI3K/HDAC inhibitor BEBT-908 could activate immunogenic ferroptosis by hyperacetylating and activating p53 in lung cancer cells (Fan et al., 2021). An FSP1 inhibitor (iFSP1) could reverse FSP1-mediated drug resistance by increasing cellular sensitivity to ferroptosis and thus, promoting PCD in lung cancer cells (Doll et al., 2019). The application of erianin (Chen et al., 2020b), sanguinarine (SAG) (Xu et al., 2022), and dihydroisotanshinone I (DT) (Wu et al., 2021) also repressed tumor growth and prevented metastasis in vivo and in vitro. In addition, targeted delivery of ncRNAs is also considered a promising anticancer strategy (Tabnak et al., 2021), and the application of delicate nanotechnology has recently attracted extensive attention due to its specific physicochemical properties (Chen et al., 2021b). Follow-up research can carry out more effective and precise interventions in the above and other regulatory pathways, regulating cellular sensitivity to ferroptosis, inhibiting the growth and metastasis of lung cancer cells, and prolonging the survival of patients.

CONCLUSION AND PERSPECTIVES

As a unique form of PCD, ferroptosis has received growing attention and interest since its first report in 2012 (Dixon et al., 2012). With the progress of ferroptosis-related research, it has been revealed that ferroptosis plays a crucial role in many pathological changes in lung diseases. Among these findings, the dysregulation of ferroptosis in lung cancer has been more widely and deeply explored (Xu et al., 2021b; Yu et al., 2021). In this review, we provide a current understanding and views of ferroptosis in lung diseases, especially beyond lung cancer, from the aspects of the molecular basis and the corresponding therapeutic significance. Impressive efforts have been made to reveal the potential pathological mechanisms of ferroptosis in diseases, such as COPD, asthma, and pulmonary fibrosis, which provide fresh thinking for the treatment of these diseases in a large number of suffering patients. However, the following questions remain to be addressed for further clinical development of ferroptosis-targeted therapies. First, are there other potential ferroptosis regulators and mechanisms? Current thinking suggests that p53 and GPX4 are two main ferroptosis-regulatory mechanisms but are not mutually exclusive. Second, what is the optimal timing, dose, and route of administration for treatment targeting ferroptosis in specific pathological lung conditions and disease stages? Persistent usage, overdose, and systemic administration of drugs that inactivate ferroptosis might theoretically increase cancer occurrence. Third, what physiological role does ferroptosis play in lung disease? Ferroptosis is involved in the occurrence of asthma and the development of inflammation, thus, some studies have shown that inhibition of ferroptosis can be developed as a therapeutic target, while ferroptosis of eosinophils could result in sensitivity to dexamethasone and relieve the symptoms of asthmatic patients. Hence, an appropriate treatment for different types of lung disease must be carefully chosen. Fourth, in treating lung cancer and metastasis, can we activate ferroptosis specifically in cancer cells without affecting healthy cells? Inhibitions of specific ferroptotic targets are better choices than targeting p53 or GPX4.
to preserve their comprehensive influence on tumor suppression and antioxidation in normal cells. It is of great significance and value to further study the pathogenesis of ferroptosis in lung diseases because of insufficient understanding, to determine sensitive biological indicators, and reliable therapeutic targets. More efficient and specific regulation of cellular ferroptosis is pivotal to further investigations.

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YL collected and analyzed the data, conceived the study, and wrote the manuscript. YY collected and analyzed the data. YY collected and analyzed the data and designed the study. All authors commented on previous versions of the manuscript, read and approved the final manuscript.

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