Research Article

Synthesis and In Vitro Inhibition Effect of New Pyrido[2,3-d]pyrimidine Derivatives on Erythrocyte Carbonic Anhydrase I and II

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In vitro inhibition effects of indolylchalcones and new pyrido[2,3-d]pyrimidine derivatives on purified human carbonic anhydrase I and II (hCA I and II) were investigated by using CO₂ as a substrate. The results showed that all compounds inhibited the hCA I and hCA II enzyme activities. Among all the synthesized compounds, 7e (IC₅₀ = 6.79 μM) was found to be the most active compound for hCA I inhibitory activity and 5g (IC₅₀ = 7.22 μM) showed the highest hCA II inhibitory activity. Structure-activity relationships study showed that indolylchalcone derivatives have higher inhibitory activities than pyrido[2,3-d]pyrimidine derivatives on hCA I and hCA II. Additionally, methyl group bonded to uracil ring increases inhibitory activities on both hCA I and hCA II.

1. Introduction

Carbonic anhydrase (CA, EC 4.2.1.1) is a ubiquitous zinc enzyme. Basically, there are several mammalian cytosolic forms (CA-I, CA-II, CA-III, CA-VII, and CA-XIII), four membrane-bound forms (CA-IV, CA-IX, CA-XII, and CA-XIV), one mitochondrial form (CA-V), and a secreted CA form (CA-VI) [1, 2]. They all catalyze a very simple physiological reaction, the interconversion between carbon dioxide and the bicarbonate ion, and are thus involved in crucial physiological processes connected with respiration and transport of CO₂/bicarbonate between metabolizing tissues and the lungs, pH and CO₂ homeostasis, electrolyte secretion in a variety of tissues/organs, biosynthetic reactions (such as the gluconeogenesis, lipogenesis, and ureagenesis), bone resorption, calcification, tumorigenicity, and many other physiologic or pathologic processes [1–3]. CA inhibitors have now been a mainstay of human clinical intervention for several decades, with at least 25 clinically used drugs that are CA inhibitors [4]. Although there are many studies on this enzyme, the CA enzyme family continues to capture the attention of drug discovery scientists and clinicians as the knowledge regarding the therapeutic implications associated with this enzyme class continues to grow [4, 5].

Indoles are one of the most important nitrogen containing heterocyclic molecules, found extensively in biological system which play vital role in biochemical processes. Indole ring constitutes an important template for drug design such as the classical nonsteroidal anti-inflammatory drugs (NSAIDs) indomethacin and indoxole [6]. Further indole derivatives have been reported to possess promising biological activities including analgesic [7], antipyretic [8], antifungal [9], anti-inflammatory [10,11], antitumor [12], anticonvulsant [13], and selective COX-2 inhibitory activities [14]. Thus the efficient synthesis of novel substituted indole derivative compounds still represents highly pursued target.

Pyrido[2,3-d]pyrimidines have received considerable attention over the past years because of their wide range of biological activities, which include antitumor [15], antibacterial [16], anti-inflammatory [17], and antifungal activities...
[18], and also act as cyclin-dependent kinase 4 inhibitors [19]. Also these compounds are considered to be important for synthetic drugs (e.g., barbituric acid derivatives), chemotherapeutic agents (e.g., sulfadiazine), and agricultural chemicals [20].

In this study, a series of 7 indolylchalcone and 11 new pyrido[2,3-d]pyrimidine derivatives containing indole ring were synthesized and their effects on human carbonic anhydrase (hCA) I and II were evaluated. Additionally, structure-activity relationship was examined.

2. Materials and Methods

2.1. General Chemistry. Melting points were taken on a Barnstead Electrothermal 9200. IR spectra were measured on a Shimadzu Prestige-21 (200 VCE) spectrometer. 

2.2. Synthetic Procedures and Spectral Data

**IH-Indole-3-carbaldehyde (2).** Phosphorous oxychloride (1 mL) was added dropwise to cold anhydrous DMF (3 mL) and the mixture was stirred at 0°C for 1 h. The indole (1.17 g), dissolved in anhydrous DMF, was added dropwise to above formylation complex solution at below 10°C. The mixture was warmed to 35–40°C and stirred for 1 hour. Then NaOH (0.5 g NaOH, 14.6 mL water) was added. The mixture was warmed to 100°C and stirred for 1 h; then it was cooled, filtered, and dried in vacuum for overnight.

Pink powder was obtained in 94% yield. 

**N-Methyl Indole-3-carboxaldehyde (3).** K₂CO₃ (0.95 g) and CH₃OH (2 mL) were added to solution of indole-3-carboxaldehyde (1 g) in 10 mL DMF. The mixture was stirred at 100°C for 4 hours and then cooled and poured onto ice-water. The precipitate was filtered and dried in vacuum oven.

White solid was obtained in 95% yield. 

**Synthesis of Indolylchalcone Derivatives (5a–g).** A solution of NaOH (40%, 5 mL) was added to mixture of 1-methyl-1H-indole-3-carbaldehyde (1 mmol) 3 and aceto phenone derivatives (1 mmol) 4a–g in absolute ethanol. The mixture was stirred at room temperature for 2 hours. Then it was poured into ice-cold water, neutralized with acid, filtered, and washed with water. The filtrate was dried in vacuum oven.

(E)-1-(3,4-Dimethoxyphenyl)-3-(1-methyl-1H-indol-3-yl) prop-2-en-1-one (5a). Yield: 75%, yellow powder, mp: 271°C, IR (KBr): 3089.9, 3008.9, 2910.5, 2839.2, 1645.2, 1597.0, 1556.5, 1373.3, 1255.6, 1166.9, 1022.7, 804.3 µ (cm⁻¹); 

1H NMR (CDCl₃, 300 MHz) δ: 8.08 (IH, d, J = 15.5 Hz), 8.01–8.03 (IH, dd, J₁ = 2.0 Hz, J₂ = 6.1 Hz), 7.71–7.74 (IH, dd, J₁ = 1.8 Hz, J₂ = 8.5 Hz), 7.66 (IH, d, J = 1.8 Hz), 7.37 (IH, d, J = 15.5 Hz), 7.47 (IH, s), 7.30–7.39 (3H, m), 6.95 (IH, d, J = 8.5 Hz), 3.99 (3H, s), 3.97 (3H, s), 3.84 (3H, s) ppm; 

13C NMR (CDCl₃, 75 MHz) δ: 189.1, 152.8, 149.2, 138.4, 138.1, 134.7, 132.3, 126.3, 123.3, 122.7, 121.7, 121.0, 116.6, 113.1, 110.8, 110.4, 110.1, 56.3, 56.2, 33.5 ppm; LC-MS (m/z): 322.57 [MH⁺]

(E)-1-(4-Methoxyphenyl)-3-(1-methyl-1H-indol-3-yl) prop-2-en-1-one (5b). Yield: 65%, yellow powder, mp: 254°C, IR (KBr): 3128.5, 3054.6, 2935.6, 2841.1, 1649.1, 1598.9, 1373.3, 1253.7, 1166.9, 1026.1 µ (cm⁻¹); 

1H NMR (CDCl₃, 300 MHz) δ: 8.00–8.10 (4H, m), 7.57 (IH, d, J = 15.5 Hz), 7.46 (IH, s), 7.30–7.37 (3H, m), 7.00 (2H, d, J = 8.8 Hz), 3.89 (3H, s), 3.83 (3H, s) ppm; 

13C NMR (CDCl₃, 75 MHz) δ: 187.2, 161.2, 136.4, 136.1, 132.7, 130.1, 128.7, 124.3, 121.5, 119.7, 119.0, 114.9, 111.9, 111.2, 108.4, 53.7, 31.5 ppm; LC-MS (m/z): 293.00 [MH⁺]

(E)-3-(1-Methyl-1H-indol-3-yl)-1-p-tolylprop-2-en-1-one (5c). Yield: 85%, dark yellow powder, mp: 240°C, IR (KBr): 3105.5, 3022.7, 2914.4, 1647.2, 1579.7, 1556.5, 1371.3, 1280.7, 1174.6, 1029.9, 804.3 µ (cm⁻¹); 

1H NMR (CDCl₃, 300 MHz) δ: 8.08 (IH, d, J = 15.5 Hz), 8.00–8.03 (1H, dd, J₁ = 2.0 Hz, J₂ = 6.5 Hz), 7.97 (2H, d, J = 8.0 Hz), 7.55 (1H, d, J = 15.5 Hz), 7.46 (IH, s), 7.31–7.39 (3H, m), 7.29 (2H, d, J = 8.0 Hz), 3.84 (3H, s), 2.44 (3H, s) ppm; 

13C NMR (CDCl₃, 75 MHz) δ: 190.4, 143.1, 138.5, 138.4, 136.7, 134.7, 129.4, 128.6, 126.3, 123.5, 121.7, 121.0, 117.1, 113.2, 110.4, 33.5, 21.9 ppm; LC-MS (m/z): 276.25 [MH⁺]

(E)-1-(4-Chlorophenyl)-3-(1-methyl-1H-indol-3-yl)prop-2-en-1-one (5d). Yield: 96%, yellow powder, mp: 248°C, IR (KBr): 3103.4, 3085.2, 2908.8, 2807.7, 1645.2, 1580.7, 1371.9, 1282.6, 1029.9, 1008.7, µ (cm⁻¹); 

1H NMR (CDCl₃, 300 MHz) δ: 8.09 (IH, d, J = 15.5 Hz), 7.98–8.02 (3H, m), 7.46–7.52 (4H, m), 7.32–7.40 (3H, m), 3.85 (3H, s) ppm; 

13C NMR (CDCl₃, 75 MHz) δ: 189.6, 139.5, 138.7, 138.5, 137.6, 135.3, 129.9, 129.0, 126.3, 123.5, 121.9, 121.1, 116.4, 113.1, 110.5, 33.6 ppm; LC-MS (m/z): 296.61 [MH⁺]

(E)-3-(1-Methyl-1H-indol-3-yl)-1-phenylprop-2-en-1-one (5e). Yield: 86%, light yellow powder, mp: 227°C, IR (KBr): 3095.7, 3055.2, 2933.7, 1643.3, 1581.6, 1554.6, 1462.0, 1371.3, 1278.8, 1213.2, 1076.2 µ (cm⁻¹); 

1H NMR (CDCl₃, 300 MHz) δ: 8.09 (IH, d, J = 15.2 Hz), 8.00–8.06 (3H, m), 7.47–7.57 (5H, m), 7.29–7.39 (3H, m), 3.84 (3H, s) ppm; 

13C NMR (CDCl₃, 75 MHz) δ: 190.9, 139.3, 138.9, 138.4, 135.0, 132.4, 128.7, 128.5, 126.3, 123.4, 121.8, 121.0, 117.0, 113.1, 110.4, 33.5 ppm; LC-MS (m/z): 262.26 [MH⁺]

(E)-3-(1-Methyl-1H-indol-3-yl)-1-(4-nitrophenyl)prop-2-en-1-one (5f). Yield: 88%, orange powder, mp: 278°C, IR (KBr):
or (E)-3-(1-Methyl-1H-indol-3-yl)-1-(3-nitrophenyl)prop-2-en-1-one (5g). Yield: 87%, light yellow powder, mp: 385–389 °C, 1H NMR (CDCl$_3$, 300 MHz) δ: 8.32–8.39 (3H, m), 8.18 (1H, s), 8.13–8.16 (1H, m), 8.07 (1H, d, J = 15.5 Hz), 7.63 (1H, d, J = 15.5 Hz), 7.58 (1H, s), 7.29–7.35 (3H, m), 3.88 (3H, s) ppm; 13C NMR (CDCl$_3$, 75 MHz) δ: 183.4, 140.0, 136.0, 134.4, 131.0, 125.1, 121.5, 120.6, 119.5, 118.5, 117.3, 116.2, 110.6, 108.3, 105.8, 29.0 ppm; LC-MS (m/z): 307.29 [MH$^+$].

Synthesis of Pyrido[2,3-d]pyrimidines Derivatives (7a–k). A mixture of 6-aminouracil derivatives (1 mmol) 6a or 6b, chalcone derivatives 5a–g (1 mmol), and NaOH (1 mmol) in 30 mL absolute ethanol was refluxed for 18 hours. The mixture was cooled and poured into ice-cold water. The precipitate was filtered, washed with water, and dried in vacuum oven for overnight. The crude products were recrystallized from ethanol.

7-(3,4-Dimethoxyphenyl)-1,3-dimethyl-5-(1-methyl-1H-indol-3-yl)pyrido[2,3-d]pyrimidin-2,4(1H,3H)-dione (7f). Yield: 20%, yellow powder, mp: 335–336 °C, 1H NMR (CDCl$_3$, 300 MHz) δ: 7.70–7.75 (3H, m), 7.54 (1H, d, J = 8.2 Hz), 7.51 (1H, s, NH), 8.98 (1H, t, J = 1.7 Hz), 8.66 (1H, d, J = 8.2 Hz), 8.32–8.35 (1H, dd, J = 1.5 Hz, J = 8.2 Hz), 7.82 (1H, d, J = 8.2 Hz), 7.79 (2H, s), 7.50 (1H, d, J = 8.2 Hz), 7.42 (1H, J = 7.6 Hz), 7.20 (1H, t, J = 7.0 Hz), 7.15 (2H, m), 3.88 (3H, s), 3.96 (3H, s), 3.98 (3H, s), 3.90 (3H, s), 3.42 (3H, s) ppm; 13C NMR (CDCl$_3$, 75 MHz) δ: 161.2, 157.3, 152.8, 152.6, 150.0, 149.8, 138.8, 138.4, 137.5, 134.0, 131.9, 130.9, 127.2, 125.5, 120.8, 120.7, 119.6, 118.3, 113.3, 110.9, 108.9, 33.5, 28.9 ppm; LC-MS (m/z): 458.18 [MH$^+$].

7-(3,4-Dimethoxyphenyl)-1,3-dimethyl-5-(1-methyl-1H-indol-3-yl)pyrido[2,3-d]pyrimidin-2,4(1H,3H)-dione (7d). Yield: 20%, yellow powder, mp: 340–341 °C, IR (KBr): 3078.3, 2937.9, 2839.2, 1692.3, 1647.1, 1419.6, 1329.4, 1220.9, 1134.1, 1022.2 ν (cm$^{-1}$); 1H NMR (CDCl$_3$, 300 MHz) δ: 7.77–7.83 (3H, m), 7.54 (1H, d, J = 8.2 Hz), 7.51 (1H, s, NH), 7.41 (1H, d, J = 8.2 Hz), 7.31 (1H, d, J = 7.0 Hz), 7.18 (1H, t, J = 7.0 Hz), 6.98 (1H, d, J = 8.2 Hz), 3.99 (3H, s), 3.96 (3H, s), 3.94 (3H, s), 3.90 (3H, s), 3.42 (3H, s) ppm; 13C NMR (CDCl$_3$, 75 MHz) δ: 161.0, 158.3, 152.5, 151.8, 151.4, 149.4, 148.3, 136.9, 130.8, 130.5, 127.4, 122.3, 120.8, 120.7, 119.8, 118.4, 112.8, 111.2, 110.3, 110.1, 106.1, 56.2, 56.1, 33.4, 30.3, 28.7 ppm; LC-MS (m/z): 482.66 [MH$^+$].
118.5, 112.6, 109.9, 106.2, 33.2, 30.2, 28.4, 21.4 ppm; LC-MS (m/z): 411.33 [MH⁺].

7-(4-Chlorophenyl)-1,3-dimethyl-5-(1-methyl-1H-indol-3-yl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (7h). Yield: 76%, yellow powder, mp: 358-359°C, IR (KBr): 3064, 3045, 2943, 1701, 1654 ν (cm⁻¹); ¹H NMR (CDCl₃, 300 MHz) δ: 8.07 (2H, d, J = 8.5 Hz), 7.73 (1H, s), 7.52–7.55 (2H, m), 7.47 (2H, d, J = 8.5 Hz), 7.41 (1H, d, J = 8.2 Hz), 7.29 (1H, t, J = 7.5 Hz), 7.18 (1H, t, J = 7.0 Hz), 3.90 (3H, s), 3.88 (3H, s), 3.42 (3H, s) ppm; ¹³C NMR (CDCl₃, 75 MHz) δ: 161.0, 157.5, 152.7, 151.8, 148.8, 136.9, 136.8, 136.2, 131.0, 129.3, 128.9, 127.3, 122.4, 120.8, 119.8, 118.8, 112.6, 110.2, 106.8, 33.5, 30.5, 28.7 ppm; LC-MS (m/z): 432.37 [MH⁺].

1.3-Dimethyl-5-(1-methyl-1H-indol-3-yl)-7-phenylpyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (7i). Yield: 32%, yellow powder, mp: 320°C, IR (KBr): 3074, 3170.9, 1703.1, 1656.8, 1519.9, 3157.4, 3049.4, 2943.5, 1701, 1654, 1585.4, 1519.9, 1434.2, 852.5 ν (cm⁻¹); ¹H NMR (CDCl₃, 300 MHz) δ: 8.13–8.15 (2H, dd, J = 2.0 Hz, J₂ = 7.5 Hz), 7.77 (1H, s), 7.49–7.56 (5H, m), 7.41 (1H, d, J = 8.2 Hz), 7.29 (1H, t, J = 7.0 Hz), 7.18 (1H, t, J = 7.0 Hz), 3.91 (3H, s), 3.90 (3H, s), 3.42 (3H, s) ppm; ¹³C NMR (CDCl₃, 75 MHz) δ: 161.1, 158.7, 152.6, 151.8, 148.8, 137.8, 136.9, 131.0, 130.6, 129.1, 127.6, 127.4, 122.4, 120.7, 119.9, 119.1, 112.7, 110.1, 106.6, 33.5, 30.5, 28.7 ppm; LC-MS (m/z): 397.51 [MH⁺].

1.3-Dimethyl-5-(1-methyl-1H-indol-3-yl)-7-(4-nitrophenyl)pyrido[2,3-d]pyrimidine-2,4(1H,3H)-dione (7j). Yield: 60%, light orange powder, mp: 347–349°C, IR (KBr): 3157.4, 3049.4, 2943.5, 1701, 1658.7, 1585.4, 1519.9, 1434.2, 852.5 ν (cm⁻¹); ¹H NMR (CDCl₃, 300 MHz) δ: 8.52 (2H, d, J = 79 Hz), 8.37 (2H, d, J = 79 Hz), 7.92 (1H, s), 7.82 (1H, s), 7.53 (1H, d, J = 8.2 Hz), 7.42 (1H, d, J = 79 Hz), 7.23 (1H, t, J = 71 Hz), 7.08 (1H, t, J = 70 Hz), 3.90 (3H, s), 3.75 (3H, s), 3.24 (3H, s) ppm; ¹³C NMR (CDCl₃, 75 MHz) δ: 160.8, 155.3, 152.8, 151.6, 148.9, 148.7, 137.2, 134.4, 132.0, 129.2, 127.2, 124.6, 121.7, 120.7, 119.8, 118.6, 113.3, 110.9, 108.1, 33.4, 30.5, 28.8 ppm; LC-MS (m/z): 442.19 [MH⁺].

2.3. Preparation and Purification of Hemolysate from Blood Red Cells. Blood samples (25 mL) were taken from healthy human volunteers. They were anticoagulated with acid-citrate-dextrose and centrifuged at 2000 g for 20 min at 4°C, and the supernatant was removed. The packed erythrocytes were washed three times with 0.9% NaCl and then haemolysed in cold water. The ghosts and any intact cells were removed by centrifugation at 2000 g for 25 min at 4°C, and the pH of the haemolysate was adjusted to pH 8.5 with solid Tris-base. The 25 mL haemolysate was applied to an affinity column containing L-tyrosine-sulfonamide-sepharose-4B [21] equilibrated with 25 mM Tris-HCl/0.1M Na₂SO₄ (pH 8.5). The affinity gel was washed with 50 mL of 25 mM Tris-HCl/22 mM Na₂SO₄ (pH 8.5). The hCA isozymes were then eluted with 0.1 M NaCl/25 mM Na₂HPO₄ (pH 6.3) and 0.1 M CH₃COONa/0.5 M NaClO₄ (pH 5.6), which recovered hCA I and hCA II, respectively. Fractions of 3 mL were collected and their absorbance was measured at 280 nm.

2.4. CA Enzyme Assay. CA activity was measured by the Maren method which is based on determination of the time required for the pH to decrease from 10.0 to 7.4 due to CO₂ hydration [22]. The assay solution was 0.5 M Na₂CO₃/0.1 M NaHCO₃ (pH 10.0) and Phenol Red was added as the pH indicator. CO₂-hydration activity (enzyme units (EU)) was calculated by using the equation t₀ – tₑ/tₑ, where t₀ and tₑ are the times for pH change of the nonenzymatic and the enzymatic reactions, respectively.

2.5. In Vitro Inhibition Studies. For the inhibition studies of indolylchalcone and pyrido[2,3-d]pyrimidine derivatives, different concentrations of these compounds were added to the assay with CO₂. Activity percentage values of CA for different concentrations of each pyrimidine derivatives were determined by regression analysis using Microsoft Office 2000 Excel. CA enzyme activity without these compounds was accepted as 100% activity.

3. Results and Discussion

3.1. Chemistry. The indolylchalcone derivatives 5a–g, prepared by the condensing various acetophenones and indolylaldehyde 3 with NaOH as a base, were reacted with 3-methyl-6-aminouracil 6a and 6-aminouracil 6b to get pyrido[2,3-d]pyrimidine derivatives (7a–k) at high yields. The large J value (15.5 Hz) clearly reveals the E-geometry for the chalcones.

3.2. Biological Evaluation of Indolylchalcone and Pyrido[2,3-d]pyrimidine Derivatives for hCA I and hCA II Inhibitory Activities. For evaluating the hCA I and II inhibitory effect, all compounds were subjected to hCA I and II inhibition assay with CO₂ as a substrate. The result showed that all synthesized compounds (5a–g and 7a–k) inhibited the hCA I and hCA II enzyme activities.

The IC₅₀ values and inhibition constants of 5a–g and 7a–k analogues against hCA I and hCA II were summarized in Table 1 and the IC₅₀ graphs were given in Figure 1.

We have determined the IC₅₀ values of 6.79–26.21 μM for the inhibition of hCA I and 7.22–31.10 μM for the inhibition of hCA II. Among all compounds, 7f (IC₅₀ = 6.79 μM) was found to be the most active one for hCA I inhibitory
activity and 5g (IC$_{50}$ = 7.22 μM) showed the highest hCA II inhibitory activity. 5b (IC$_{50}$ = 7.42 μM) was found to be the most active one for hCA I inhibitory activity and 5g (IC$_{50}$ = 7.22 μM) showed the highest hCA II inhibitory activity for the indolylchalcone derivatives. Among the pyrido[2,3-d]pyrimidine derivatives, 7e (IC$_{50}$ = 6.79 μM) showed the highest hCA I inhibitory activity and 7g (IC$_{50}$ = 7.57 μM) showed the highest hCA II inhibitory activity.

It was reported that 1,4-dihydropyrimidinone substituted diarylurea compounds were synthesized and their effects on the hCA I and II enzyme activities were examined. Their minimum concentrations to achieve 50% inhibition were between 66.23 and 197.70 μM for hCA I, 63.09 and 169.71 μM for hCA II [23]. It is evident that the indolylchalcone and pyrido[2,3-d]pyrimidine derivatives, synthesized in this work, showed better hCA I and II inhibitory activities than 1,4-dihydropyrimidinone substituted diarylurea compounds.

### 3.3. Structure-Activity Relationships (SAR)

Generally, we have seen that indolylchalcone derivatives have higher inhibitory activities than pyrido[2,3-d]pyrimidine derivatives on hCA I and hCA II. The following structure-activity relationship (SAR) observations can be drawn from the data.

(a) For the indolylchalcone derivatives, the presence of one electron-donating group (methoxy) bonded to paraposition of phenyl ring (5b) increased inhibitory activity on hCA I. Electron-withdrawing group (nitro) bonded to metaposition of phenyl ring (5g) has the highest hCA II inhibitory activity (IC$_{50}$ = 7.22 μM).

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**Table 1:** Inhibitory effect of indolylchalcone (5a–g) and pyrido[2,3-d]pyrimidine derivatives (7a–k) on hCA I and hCA II.

| Compound | hCA I IC$_{50}$ (μM) | hCA II IC$_{50}$ (μM) |
|----------|-----------------|-----------------|
| 5a       | 8.34            | 8.88            |
| 5b       | 7.42            | 10.35           |
| 5c       | 13.07           | 12.28           |
| 5d       | 8.20            | 8.26            |
| 5e       | 12.84           | 9.15            |
| 5f       | 10.87           | 9.31            |
| 5g       | 8.38            | 7.22            |
| 7a       | 16.29           | 19.42           |
| 7b       | 11.56           | 12.06           |
| 7c       | 21.09           | 31.10           |
| 7d       | 12.14           | 13.66           |
| 7e       | 6.79            | 8.06            |
| 7f       | 26.23           | 25.40           |
| 7g       | 7.61            | 7.57            |
| 7h       | 12.36           | 24.67           |
| 7i       | 8.72            | 8.14            |
| 7j       | 10.19           | 9.56            |
| 7k       | 22.30           | 26.68           |
Figure 1: IC_{50} graphics of indolylchalcone (5a–g) and pyrido[2,3-d]pyrimidine (7a–k) derivatives on hCA I and hCA II.
Sulfonamides are coordinated to the zinc (II) ion within the hCA active site, whereas their organic scaffold fills the entire active site cavity, making an extensive series of van der Waals and polar interactions with amino acid residues delimiting this cavity [24,25]. As the synthesized compounds are very bulky and do not contain a classical zinc-binding group [4], it can be hypothesized that they are not able to bind near the zinc ion showing a different mechanism of action. Structural studies of the complexes that these compounds form with the human isof orm II could clarify this important issue.

4. Conclusions

In conclusion, series of 7 indolylchalcone and II new pyrido[2,3-d]pyrimidine derivatives containing indole ring were synthesized. Their activities as hCA I and hCA II inhibitors and structure-activity relationships were examined. All compounds inhibited both hCA I and hCA II enzyme activities. Most of compounds containing electron-donating groups at phenyl ring were gener ally stronger inhibitors of hCA I and hCA II. Additionally, methyl group bonded to 3-position of uracil ring generally increased inhibitory activities on both hCA I and hCA II. Thus, the present study revealed that the type and position of substituent of the phenyl and uracil rings could be exploited to modulate the CA inhibitors efficacy.

In summary, enzyme inhibition is an important issue for drug design [26–28]. Our results showed that new pyrido[2,3-d]pyrimidine derivatives inhibited the hCA I and II enzyme activity. Therefore, the compounds here investigated are likely to be adopted as good candidates as drugs and may be taken for further evaluation in in vivo studies.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

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