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**Introduction**

Schizophrenia and bipolar disorder are characterized by psychotic symptoms, mood dysregulation and cognitive dysfunction, and are regarded as complex genetic disorders with mainly unknown underlying pathophysiology. Despite the high heritability, the genetic susceptibility factors are only partly understood. Large genome-wide association studies (GWAS) have provided the opportunity to identify new genes and genetic variation in new pathways.\(^1\) Using appropriate sample sizes, GWAS have identified several disease-associated gene variants,\(^2\)\(^-\)\(^4\) and in schizophrenia, these studies indicate the presence of neurodevelopmental and immune-related mechanisms.\(^3\)\(^,\)\(^5\)

One of the findings in schizophrenia is association to the gene transcription factor 4 (**TCF4**), located on chromosome 18q21, coding for a basic helix–loop–helix transcription factor. First, Stefansson et al.\(^6\)\(^,\)\(^7\) combined data from several GWAS and reported seven single nucleotide polymorphisms (SNPs) associated with schizophrenia at a genome-wide level, including rs9960767 located in an intron of **TCF4** ($P = 4.1 \times 10^{-8}$). A study of Han Chinese patients replicated this association through another SNP (rs2958182, in high linkage disequilibrium (LD) with rs9960767).\(^8\) Rs4309482, situated intergenically downstream of **TCF4** and upstream of **CCDC68**, was reported in another large GWAS.\(^9\) Finally, in recent mega-analyses, two new **TCF4** SNPs (rs17512836 (intron 3 of **TCF4**) and rs24966547 (in high LD with rs4309482)) were identified and further supported **TCF4** as a disease gene for schizophrenia.\(^5\) Interestingly, **TCF4** has also been associated with bipolar disorder,\(^8\) which is in line with the growing evidence supporting overlapping genetic factors in schizophrenia and bipolar disorders\(^9\),\(^10\) and corresponding to the overlapping clinical and neurocognitive features.\(^11\) It is possible that **TCF4** risk loci confer risk for psychosis-related phenotypes across diagnostic boundaries. **TCF4** variations have been shown to be responsible for Pitt–Hopkins syndrome characterized by severe mental retardation, developmental delay, microcephaly, hyper-ventilation episodes and characteristic dysmorphisms.
The participants are part of a larger Norwegian study sample (Thematically Organized Psychosis (TOP) Study), which is a collaborative study involving the University of Oslo and all the Hospitals in the Oslo region, funded by the University, Regional Health Authorities and the Research Council of Norway. A total of 596 patients with psychotic disorders or affective disorders according to DSM-IV and 385 healthy controls were included. The patients were divided into three groups: (1) schizophrenia spectrum disorders, referred to as schizophrenia in the following: encompassing schizophrenia (n = 225), schizoaffective disorder (n = 23) and schizophreniform disorder (n = 52), (2) bipolar spectrum disorder, referred to as bipolar disorder in the following: encompassing bipolar disorder type 1 (n = 131), bipolar disorder type 2 (n = 60), bipolar disorder not otherwise specified (n = 13) and major depressive disorder with psychosis (n = 19) and (3) psychosis classified as ‘other psychosis’, that is, non-schizophrenia or bipolar disorder (n = 73). Main inclusion criteria in addition to the mentioned DSM-IV disorders were age between 18 and 65 years, absence of mental retardation and no history of major brain trauma.

The group of healthy controls (n = 385) was from the same catchment area as the patient group and randomly selected from national statistical records (www.ssb.no). The healthy control sample was evaluated with a clinical interview about severe mental disorder symptoms and the Primary Care Evaluation of Mental Disorders (PRIME-MD).20 The subjects were excluded if they or any of their close relatives had a lifetime history of a severe psychiatric disorder (schizophrenia, bipolar disorder or major depression), if they had an unstable medical condition known to interfere with brain function, or substance abuse or dependency in the past 3 months. A detailed description of the sample has been reported elsewhere.21 Demographic and clinical data are presented in Table 1.

In the described sample, a total of 895 participants had both neurocognitive and genotype data, 542 had mRNA data and genotype data, 480 had both brain imaging and genotype data and a total of 717 had both mRNA and clinical data. Furthermore, a total of 300 patients with schizophrenia had clinical data and genotype data. For details, see Supplementary Tables S1 and S2 online.

The study was approved by The Norwegian Scientific-Ethical Committees and the Norwegian Data Protection Agency, and the biobank was approved by the Health Authorities. Written informed consent was given from all participants before inclusion.

Clinical characterization. Clinical assessment was carried out by trained physicians and clinical psychologists. A standardized protocol of operational instruments was used.22 Diagnosis was based on the Structured Clinical Interview for DSM-IV Axis I disorders (SCID-I).23 Diagnostic reliability was found satisfactory, with overall agreement for DSM-IV diagnostic categories of 82% with κ = 0.77 (95% confidence interval: 0.60–0.94).22 In addition, information from follow-up visits was used to secure correct diagnoses. In order to specifically address core psychotic features, the following assessments were used: Current positive and negative psychiatric symptoms were assessed using the Positive and Negative Syndrome Scale (PANSS).24 Psychosocial functioning in patients was assessed with the Global Assessment of Functioning scale, split version.25 Age at onset was obtained from interview and/or hospital records. For the current analysis, phenotypes related to neurodevelopment (age at onset and positive and negative symptoms (PANSS positive and negative sum-score)) were selected.17 All subjects reported their alcohol and drug intake in the past 2 weeks before the interview.

**Methods and materials**

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### Neuropsychological testing.
All participants were tested with a clinically relevant neuropsychological battery. For the current analysis, cognitive tests that have revealed cognitive dysfunction in schizophrenia and bipolar disorder were selected.\(^1\)\(^1\) The test battery included the following tests: general cognitive function measured using the Wechsler Abbreviated Scale of Intelligence (WASI).\(^2\)\(^6\) Mental speed was evaluated by the Digit Symbol Coding Test (WAIS-III).\(^2\)\(^7\) Executive function was evaluated by the Inhibition and Inhibition/Switching on Color-Word Interference Test, and by Verbal Fluency Letter, Verbal Fluency Category and Category Switching from the Verbal Fluency Test (Delis Kaplan Executive Function System (D-KEFS)).\(^2\)\(^8\) Verbal memory was measured by the total learning score (trials A1-5) from the California Verbal Learning Test- Second Edition (CVLT-II).\(^2\)\(^9\) Low scores equal poorer performance for all tests apart from the Word-Interference Color Test.

### MR image acquisition and processing.
A subsample of 480 participants underwent MRI scanning on the same 1.5T scanner (Siemens Magnetom Sonata scanner, Siemens Medical Solutions, Erlangen, Germany) equipped with a standard head coil at the Oslo University Hospital, Oslo, Norway between 2003 and 2009.\(^3\)\(^0\) After a conventional 3-plane localizer, two sagittal T1-weighted magnetization-prepared rapid gradient echo volumes were acquired with the Siemens tl3d1_ns pulse sequence (TE 3.93 ms, TR 2730 ms, TI = 1000 ms, flip angle = 7°; FOV = 24 cm, voxel size = 1.33 × 0.94 × 1 mm\(^3\), number of partitions = 160). Acquisition parameters were optimized for increased gray/white matter image contrast. All brain scans were read by a specialist in neuroradiology and were found to be free from organic brain pathology (for example, tumors, bleeding, infarcts and obstructive hydrocephalus).

The image files in DICOM format were transferred to a Linux workstation for morphometric analysis. Images were corrected for non-linear warping caused by gradient coil non-linearities, using tools developed through the Morphometry Biomedical Informatics Research Network,\(^3\)\(^1\) and the two T1-weighted images were rigid body registered to each other (motion corrected). The FreeSurfer 3.0.2 software package ([http://surfer.nmr.mgh.harvard.edu](http://surfer.nmr.mgh.harvard.edu)) was used to obtain measures of subcortical volumes\(^3\)\(^2\) and cortical area and thickness.\(^3\)\(^3\)\(^3\)\(^4\) Summary measures were generated and used as structural phenotypes.\(^3\)\(^5\) For more details, see Supplementary Information online.

We included seven morphometric summation measures\(^3\)\(^5\) shown to be abnormal in schizophrenia and bipolar disorders:\(^3\)\(^0\) frontal cortical area and thickness, temporal cortical area and thickness, hippocampal volume, cerebellar volume and total ventricular volume (comprising the lateral and inferior lateral, third and fourth ventricle). Based on the finding of microcephaly in patients with Pitt–Hopkins syndrome, we also investigated total brain volume.

### Genotyping and SNP selection.
The sample was genotyped at Expression Analysis (Durham, NC, USA) using the Affymetrix Genome-Wide Human SNP Array 6.0. (Affymetrix, Santa Clara, CA, USA). Quality control was performed using PLINK.\(^3\)\(^6\) Details about genotyping and quality control procedures are provided in supplementary information. This resulted in a sample of 1377 individuals with suitable genotype data available; of the 909,622 markers on the array, 525,117 autosomal markers remained. The total genotyping rate in remaining individuals was 0.993. We extracted SNPs from the TCF4 gene region based on UCSC coordinates ± 20 kb kb (chr 18, 52869562-53323185 bp (hg19)) leaving a total of 59 SNPs (see Supplementary Table S3 online). Five additional SNPs in or near the TCF4 gene were selected because of association with schizophrenia (rs12966547, rs2958182, rs9960767, rs4309482 and rs17512836).\(^4\)\(^–\)\(^7\) They were imputed by IMPUTE2\(^3\)\(^7\) using 1000 genomes CEU sample as reference. LD blocks are defined by confidence bounds on the normalized measure of allelic association (D')\(^3\)\(^8\) (see Supplementary Figure S1 online).
RNA measures. Total RNA was isolated from blood using the Tempus 12-Port RNA Isolation kit (Applied Biosystems; Ambion, Austin, TX, USA) and quantified using the ND-1000 spectrophotometer (NanoDrop Technologies, Wilmington, DE, USA).

Quantitative real-time RT-PCR was performed by using the ABI Prism 7900 HT sequence detection system (Applied Biosystems, Foster City, CA, USA). RNA (1 μg) was reverse-transcribed using a high-capacity cDNA RT kit (Applied Biosystems), and quantitative RT-PCR was performed using Taqman Gene Expression Master Mix (Applied Biosystems). The Assays-on-Demand products purchased from Applied Biosystems contained Taqman minor groove binder probes (6-FAM dye-labeled) combined with the primers for the genes of interest: TCF4 (Hs00162613_m1), An Assay-on-Demand product for eukaryotic ACTB (ACTB: Hs99999903_m1) was used as endogenous control. Details about RNA quantification procedure are provided in supplementary information online.

Statistical analysis. Potential differences in TCF4 mRNA level between the diagnostic groups and control group were analyzed using an analysis of covariance in the PASW software package for Windows, version 18.0. (PASW, Chicago, IL). TCF4 mRNA level was entered as the dependent variable, diagnostic group as the independent variable. Age was significantly correlated with TCF4 mRNA level (Pearson \( r = 0.14, P < 0.005 \)), and was entered as a covariate in the analysis. Independent sample \( t \)-test showed no statistical effect of gender on TCF4 mRNA level (\( t(714) = 0.99, P > 0.05 \)). As psychotropic medication may affect the TCF4 mRNA level, a separate one-way analysis of variance was conducted to explore the effect of medication. The patient sample was divided into six non-overlapping groups based on their prescription: (1) Antipsychotics; 2: Lithium; 3: Anticonvulsiva; 4: Antidepressants; 5: Combinational treatment of more than one type of psychotropic; 6: Unmedicated; see Supplementary Table S4 online). Partial correlation (\( r \)) controlling for age was used to investigate how positive and negative symptoms related to TCF4 mRNA as a previously study had found a correlation between delusions and TCF4 mRNA.30

Associations between TCF4 SNPs and core psychotic phenotypes were tested through the additive model of linear regression in PLINK.36 The whole sample was investigated for association to cognitive measures, MRI and mRNA. Additionally, associations with psychosis symptoms and age at disease onset were investigated in the whole subset of patients. Each analysis controlled for gender, age at sampling and diagnostic group (‘schizophrenia’, ‘bipolar’, ‘other psychosis’ or control). The effect in the schizophrenia group only, controlling for age and gender, was also analyzed.

Our primary focus was the five SNPs (rs12966547, rs9960767, rs2958182, rs4309482 and rs17512836) implicated in schizophrenia. Firstly, we analyzed these SNPs with general cognitive function, mental speed, executive function and verbal memory, then PANSS and age at onset. Using the same linear model, we investigated association to the eight morphometric summation measures: frontal cortical area and thickness, temporal cortical area and thickness, hippocampal volume, cerebellar volume, total ventricular volume and total brain volume.

Correction for multiple testing. A Bonferroni correction method was used to correct for multiple testing across independent variables after taking the correlation between genotypes and phenotypes into account. LD blocks determined by confidence bounds on the normalized measure of allelic association (D’) define number of independent SNPs.38 Several of the 19 phenotypes in the current study are highly correlated (see correlation matrix, Supplementary Table S5 online) and subdivision into phenotype families is one method to account for this.40 The phenotype families in the current study are: (1) TCF4 mRNA, (2) positive and negative symptoms, (3) cognitive tests performance, and (4) brain morphological summary measures. Owing to evidence that rs12966547, rs9960767, rs2958182, rs4309482 and rs17512836 are associated with schizophrenia and our a priori hypothesis for an association between the risk SNPs and key phenotypes in psychosis, Bonferroni correction for four independent SNPs (see Supplementary Figure S1A online) and four independent phenotype families were performed and significance level was set to \( P < 0.0051 \) in these analyses. The 59 TCF4 SNPs were tested for the same core psychosis phenotypes mentioned above. The \( P \)-values for the SNP–phenotype analyses were corrected for multiple testing with Bonferroni correction for 14 independent SNPs (see Supplementary Figure S1B online) and four independent phenotype families and the significance level was set to \( P < 0.00089 \).

Results

TCF4 mRNA expression. We measured TCF4 mRNA expression in the blood of patients with schizophrenia, bipolar disorder, other psychoses and controls. There was a significant difference in TCF4 mRNA expression level between the groups after correcting for age (\( F(3,711) = 8.23, P = 2.2 \times 10^{-5} \)), with a higher relative expression in patient groups compared with controls (Figure 1). Post hoc Tukey tests revealed significantly higher relative mRNA level in all individual patient groups in comparison to controls: schizophrenia (2.22%, \( P = 9.0 \times 10^{-5} \)), bipolar disorder (1.78%, \( P = 0.005 \)) and other psychoses (1.87%, \( P = 0.049 \)) (see Supplementary Table S6 online). No significant difference between the individual patient groups was observed (see Supplementary Table S6 online). Medication status (see Supplementary Table S4 online) was significantly associated with TCF4 mRNA level (\( F(5,499) = 2.7, P = 0.020 \)). Post hoc Tukey tests did not show any significant mRNA differences between the medication groups. Comparison of mRNA level in all unmedicated patients (\( n = 74 \)) and controls (\( n = 212 \)) revealed a trend of higher mRNA level in patients (\( t(284) = 1.37, P = 0.17 \)). Next, positive symptoms (\( r = 0.10, P = 0.027 \)) and negative symptoms (\( r = 0.13, P = 0.004 \)) were positively correlated with TCF4 mRNA level in total sample and negative symptoms (\( r = 0.16, P = 0.016 \)) in schizophrenia sample (see Supplementary Table S7 online). To investigate if the mRNA differences could be associated with TCF4 gene variants, we tested SNPs previously associated with schizophrenia: rs12966547, rs9960767, rs2958182, rs4309482 and rs17512836 (the ‘schizophrenia
SNP analysis). We also tested a set of 59 additional SNPs in the TCF4 gene, which we called the ‘exploratory analysis’. Each analysis was performed in the entire sample as well as in schizophrenia patients separately. Nominal significant associations between TCF4 SNP variants and TCF4 mRNA expression levels were found (see Supplementary Table S7 online) but did not remain significant after correcting for multiple testing. No significant eQTLs for TCF4 mRNA were found in our data (data not shown).

**TCF4 SNPs and clinical characteristics.** Nominal significant results from the ‘schizophrenia SNP analysis’ are presented in Table 2 and Figure 2. To investigate whether the TCF4 SNPs were associated with cognitive dysfunction, another sub-phenotype of psychosis characteristic of abnormal neurodevelopment. In the ‘schizophrenia SNP analysis’, risk alleles of the tightly linked rs12966547 and rs4309482 were associated with poorer performance on Verbal Fluency in total sample after correction for multiple testing (Table 2). In addition, risk alleles of rs12966547 and rs4309482 were associated with poorer verbal fluency in patients with schizophrenia and risk allele of rs2958182 was associated with better performance on Color-Word Interference test (inhibition) in patients with schizophrenia, but these findings did not survive correction for multiple testing.

In the ‘exploratory analysis’, the minor alleles of rs1377243 and rs2958188 and major allele of rs1942265, rs7241077, rs1893431, rs4800988, rs1261134, rs1788027, rs1660242 and rs1788014) from sequential LD blocks were associated with less negative symptoms in patients with schizophrenia (Table 3). There was no significant association with positive symptoms.

**TCF4 SNPs and structural brain MRI measures.** We tested if TCF4 SNPs were associated with MRI morphometric summary measures in brain regions found to be abnormal in psychosis. In the ‘schizophrenia SNP analysis’, the risk alleles of rs12966547 and rs4309482 were associated with larger ventricular volumes and the risk allele of rs9960767 was associated with a larger hippocampus and smaller

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**Figure 1** Relative TCF4 mRNA expression between the diagnostic groups. The figure shows significantly lower TCF4 mRNA expression in healthy controls (HC, n = 212) compared with schizophrenia (ScZ, n = 232, \( P = 8.6 \times 10^{-8} \)). **Bipolar disorder (BD, n = 200, \( P = 0.005 \)) and *other psychoses (OP, n = 73, \( P = 0.049 \)). Mean values with ± one standard error are presented.

**Table 2** Results from the ‘schizophrenia SNPs’ and phenotypes analyses

| SNP       | \( R^2 \) | Localisation | Minor allele | MAF       | Risk allele | Phenotype     | Sample | N    | Beta    | SE    | P      | Corr   |
|-----------|----------|--------------|--------------|-----------|-------------|---------------|--------|------|---------|-------|--------|--------|
| rs12966547 | 0.998    | 52752017     | A            | 0.436     | G\(^2\)      | Verbal fluency\(^a\) | All    | 890  | -1.75   | 0.55  | 0.0014 | 0.022  |
| rs4309482  | 0.981    | 52750469     | G            | 0.444     | A\(^1\)      | Verbal fluency\(^a\) | All    | 480  | -2.56   | 1.18  | 0.031  | 0.50   |
| rs2958182  | 0.938    | 53149021     | A            | 0.373     | T\(^9\)      | Verbal fluency\(^a\) | All    | 890  | -1.81   | 0.55  | 0.0010 | 0.016  |
| rs9960767  | 0.949    | 53155002     | C            | 0.056     | C\(^1\)      | Verbal fluency\(^a\) | All    | 480  | -2.73   | 1.19  | 0.023  | 0.37   |

**Abbreviations:** MAF, minor allele frequency; N, number; SE, standard error; SNP, single nucleotide polymorphism.  
\(^a\)Inhibition score. \(^b\)Beta is estimated by increasing number of risk alleles. \(^p\)-Values in bold remained significant after controlling for multiple testing across independent genotypes and phenotype families. \(^\delta\)Ripke et al. \(^\alpha\)Verbal Fluency Letter from the Verbal Fluency Test (Delis Kaplan Executive Function System (D-KEFS)). \(^\beta\)Steinberg et al. \(^\gamma\)Li et al. \(^\alpha\)Inhibition from the Color-Word Interference Test (Delis Kaplan Executive Function System (D-KEFS)). \(^\phi\)Stefansson et al. Nominal significant results (\( P < 0.05 \)) from the linear regression analysis of schizophrenia SNPs, correcting for diagnosis and gender.
Table 3 Results of the ‘Exploratory Analysis’ of TCF4 SNPs and phenotypes

| LD block | SNP | Minor allele | Phenotype | Sample | N  | Mean (s.d.) | Mean (s.d.) | Mean (s.d.) | Effect size | P  | Corr* |
|----------|-----|--------------|-----------|--------|----|-------------|-------------|-------------|-------------|----|-------|
| 1        | rs1261086 | C | Negative symptoms | Scz  | 282 | 13.51 (6.10) | 15.51 (6.67) | 16.94 (6.67) | -0.21 | 0.00038 | 0.021 |
| 2        | rs1261076 | C | Negative symptoms | Scz  | 282 | 13.69 (6.03) | 15.22 (6.20) | 17.91 (7.36) | -0.24 | 0.00039 | 0.0022 |
| 3        | rs1942265 | G | Negative symptoms | Scz  | 282 | 13.23 (5.94) | 14.86 (6.02) | 16.85 (7.15) | -0.20 | 0.00059 | 0.033 |
| 4        | rs7224107 | A | Negative symptoms | Scz  | 282 | 13.23 (5.94) | 14.86 (6.02) | 16.85 (7.15) | -0.20 | 0.00059 | 0.033 |
| 5        | rs1895431 | A | Negative symptoms | Scz  | 282 | 13.47 (6.31) | 15.02 (5.09) | 17.26 (7.41) | -0.22 | 0.00017 | 0.0096 |
| 6        | rs4800988 | A | Negative symptoms | Scz  | 282 | 13.23 (5.94) | 14.86 (6.02) | 16.85 (7.15) | -0.20 | 0.00059 | 0.033 |
| 7        | rs1261134 | T | Negative symptoms | Scz  | 282 | 13.72 (6.30) | 14.81 (5.85) | 17.33 (7.37) | -0.22 | 0.00011 | 0.0062 |
| 8        | rs1788027 | T | Negative symptoms | Scz  | 275 | 13.89 (6.14) | 14.87 (6.14) | 16.05 (7.22) | -0.23 | 0.00077 | 0.0043 |
| 9        | rs1760242 | T | Negative symptoms | Scz  | 281 | 14.05 (5.97) | 14.85 (6.39) | 17.17 (7.03) | -0.21 | 0.00045 | 0.025 |
| 10       | rs1788014 | G | Negative symptoms | Scz  | 282 | 13.51 (6.10) | 15.51 (6.67) | 16.94 (6.67) | -0.21 | 0.00038 | 0.021 |
| 11       | rs1377243 | T | California verbal learning | Scz  | 227 | 11.15 (2.84) | 11.81 (2.58) | 12.83 (2.59) | -0.23 | 0.00044 | 0.025 |

| —        | rs9636107 | G | Age at onset | Patients | 594 | 25.29 (8.49) | 27.38 (9.74) | 29.19 (10.47) | 0.14 | 0.00049 | 0.027 |
| —        | rs1396445 | A | Temporal cortical area | All | 480 | 11.00 (2.73) | 11.94 (2.56) | 12.82 (2.59) | -0.25 | 0.00014 | 0.0078 |
| 6        | rs2958188 | T | Age at onset | Patients | 227 | 11.00 (2.73) | 11.94 (2.56) | 12.82 (2.59) | -0.25 | 0.00014 | 0.0078 |
| 9        | rs1261086 | C | Negative symptoms | Scz  | 282 | 13.51 (6.10) | 15.51 (6.67) | 16.94 (6.67) | -0.21 | 0.00038 | 0.021 |
| 9        | rs682245  | T | Cerebellar volume | All  | 106 | 146700 (4050) | 42480 (4231) | 43380 (4511) | -0.12 | 0.00083 | 0.046 |
| 9        | rs1751422 | G | Temporal cortical area | All | 480 | 110000 (127100) | 120000 (134500) | 120000 (134500) | -0.12 | 0.00067 | 0.038 |
| 9        | rs12458118| A | Cerebellar volume | Scz | 106 | 146700 (13250) | 135600 (13020) | 132500 (16150) | 0.28 | 0.00074 | 0.041 |
| 9        | rs9951280 | G | Cerebellar volume | Scz | 106 | 146700 (13250) | 135600 (13020) | 132500 (16150) | 0.28 | 0.00074 | 0.041 |
| 9        | rs10871582 | A | Temporal cortical area | All | 480 | 41520 (3982) | 42450 (4279) | 43370 (4515) | -0.13 | 0.00056 | 0.031 |

Abbreviations: LD, linkage disequilibrium; N, number; P, uncorrected P-value; s.d., standard deviation; SNP, single nucleotide polymorphism.

*Mean and standard deviation in homoygotes for minor allele; *Heterozygotes. **Homoygotes for major allele. ***Standardized regression coefficient. **Significant P-values after controlling for multiple testing across independent genotypes and phenotype families. Of note, six remained significant after controlling for all the 19 phenotypes. **California Verbal Learning test- Second edition (CVLT-II), total learning score (trial A1-5). Scz: Schizophrenia group. Patients: all patients. All: total sample with patients and controls. Results surviving correction for multiple testing in the linear regression analysis of TCF4 SNPs and phenotypes, correcting for diagnosis and gender.

ventricles, but these findings did not remain significant after correction for multiple testing. The ‘exploratory analysis’ showed that the minor allele of three SNPs (rs41396445, rs17514242 and rs10871582) in the same LD block were associated with reduced temporal cortical area in total sample. Out of these, minor alleles of two SNPs (rs17514242 and rs10871582) were associated with reduced total brain volume in total sample. The major alleles of three SNPs (rs682245, rs12458118 and rs9951280) in the same LD block were associated with decreased cerebellar volume in patients with schizophrenia (Table 3).

Discussion

The main findings of the present study were upregulated TCF4 expression level in all psychotic diagnostic groups and that TCF4 risk variants associate with poorer verbal fluency in total sample. The exploratory analysis revealed that TCF4 variants associated with negative symptoms, cognitive dysfunction and cerebellar volume in the schizophrenia sample, age at onset in total patient sample and variants correlating with temporal cortical area and brain volume in total sample. This further implicates TCF4 in psychosis pathophysiology, and indicates a specific role in the development of clinical phenotypes related to aberrant neurodevelopment. To our knowledge, this is the first study on known TCF4 risk variants of schizophrenia and hitherto uncharacterized variants targeting psychosis phenotypes at different levels, including the peripheral TCF4 mRNA.

The analysis of known schizophrenia TCF4 risk variants revealed that the two linked rs12966547 and rs4309482 are associated with poorer executive function in the form of verbal fluency. Interestingly, reduced cognitive functioning has been associated with enlarged ventricular volume, corroborating our finding that the same risk variants, rs12966547 and rs4309482, were associated with larger ventricular volume even though this did not survive correction for multiple testing. Enlarged ventricular volume is one of the most consistent findings in schizophrenia, also found in bipolar disorder and Pitt–Hopkins syndrome. This indicates that these TCF4 risk SNPs might, at least partially, cause this common subphenotype in schizophrenia.

The pattern of associations in the non-schizophrenia risk variants Target SNP analysis corroborates a relationship between TCF4 and changes in brain morphology. Several SNPs in one haploblock were associated with cerebellar volume, temporal cortical area or brain volume. The Allan Brain Atlas shows TCF4 expression in several parts of the brain, most pronounced in hippocampus and cerebellum (http://www.brain-map.org). This is consistent with a role of TCF4 in early neurodevelopment and embryogenesis previously shown in animal models as well as concordant with microcephaly as a core characteristic of Pitt–Hopkins syndrome.

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mutations (exons 9–19) are identified. Furthermore, we region of the gene in which Pitt–Hopkins syndrome–causing affect negative symptoms are situated in the exon-dense negative symptoms in schizophrenia sample; TCA, temporal cortical area in total volume in total sample; CV, cerebellar volume in schizophrenia sample; CVLT, California Verbal Learning test in schizophrenia sample; Mbp, mega base pair; NS, negative symptoms in schizophrenia sample; TCA, temporal cortical area in total sample.

Figure 2 A TCF4 linkage disequilibrium map and a Manhattan plot with negative log $P$ values (uncorrected) from the exploratory single nucleotide polymorphism–phenotype analysis. Only phenotypes remaining significant after multiple testing is illustrated. AO, age at onset in total patient sample; BV, brain volume in total sample; CV, cerebellar volume in schizophrenia sample; CVLT, California Verbal Learning test in schizophrenia sample; Mbp, mega base pair; NS, negative symptoms in schizophrenia sample; TCA, temporal cortical area in total sample.

as common variants in CDK5RAP2 and MCPH1 are associated with reduced brain volume and cortical area, while mutations cause primary microcephaly.

Psychotic disorders are a heterogenous group of diseases and division into subgroups based on symptomatology might be desirable. The consistent effect of the TCF4 SNPs of several sequential haploblocks on negative symptoms in schizophrenia patients might be a path to define a subgroup of patients with more pronounced negative symptoms as previously suggested. Interestingly, the variants that affect negative symptoms are situated in the exon-dense region of the gene in which Pitt–Hopkins syndrome–causing mutations (exons 9–19) are identified. Furthermore, we found that one SNP was associated with both an earlier age at onset and worse verbal learning. Cognitive decline and early age at onset are predictors of poor prognosis in schizophrenia. The pattern of clinical symptoms and cognitive dysfunction resembles what has been regarded as neurodevelopmental characteristics, and is consistent with an effect of TCF4 on abnormal neurodevelopment. The consistent pattern of effect across different SNPs on independent clinical, cognitive and brain imaging phenotypes suggests that the current findings are not due to chance.

TCF4 mRNA levels were significantly higher in the patient groups compared with controls. Measurements in the blood allow for larger sample sizes and better standardization of technical procedures, but it might not reflect mRNA levels in the brain. It has previously been reported that the availability of an accessible tissue, such as blood, with similar gene expression to a more inaccessible tissue, such as brain, may advance research in neuropsychiatric disorders. Sullivan et al. reported that, on a transcriptome level, whole blood shares significant gene expression similarities with brain tissue. Thus, even though it was not perfectly correlated, gene expression in blood might be a useful surrogate marker for gene expression in the central nervous system. Our observation of increased level of TCF4 mRNA in patients with schizophrenia is consistent with increased cerebellar expression of TCF4 in schizophrenia patients shown in a genomic convergence analysis on post-mortem cerebellar cortices, indicating that in the case of TCF4, blood and brain expression might correlate. Microarrays of gene expression in blood is a practical method to use in psychotic patients and the technique has become increasingly accepted for studying biomarkers for neuropsychiatric diseases. For example, gene expression in peripheral blood mononuclear cells in patients with multiple sclerosis was correlated to disease activity. This is in line with our finding of a correlation between mRNA level and severity of positive and negative symptoms in patients with schizophrenia.

The result of increased TCF4 mRNA in psychotic patients should be interpreted with caution as an effect on mRNA levels of prescribed psychotropic medications cannot be excluded. We did find an overall effect of medication on mRNA level in patients, with the highest level in patients on antipsychotics or treated with several types of psychotropics, even though it did not survive correction for multiple testing. Thus, the observed effect might be a mirror of medication or reflect a more severe form of psychosis in patients on regular medical treatment. The trend of higher TCF4 mRNA level in unmedicated patients compared with controls does corroborate an effect regardless of medication, though. Another small-size (41 patients) study observed less TCF4 expression in blood from patients with higher rates of delusions. TCF4 mRNA level was positively correlated with positive and negative symptoms in our sample. This is in line with our finding of several SNPs associated with negative symptoms, which might reflect a difference in TCF4 expression in patients with high level of positive and negative symptoms and suggests that TCF4 variants may contribute to more pronounced symptoms. The reason for the apparent discrepancy between Kurian et al. and our findings might reflect different assumptions. While they compared TCF4 mRNA level in patients with schizophrenia with high level of delusions and patients with no delusions, our sample is generally in a more chronic phase of disease. Further, our sample is much larger and more heterogeneous and includes bipolar disorder and other psychoses, while they include different ethnicities. Therefore, before TCF4 mRNA can be used as a biomarker for psychosis, any putative effect of the level of positive and
negative symptoms and medication on TCF4 mRNA level needs further investigation.

Two recent studies have found allele-specific expression of TCF4 in post-mortem human brains.\textsuperscript{65,66} No significant association between mRNA level and the common TCF4 polymorphisms was found in our data after correction for multiple testing, but there were nominal associations. The lack of significant support for cis-regulatory effect could be due to Type II error, as there are many factors that affect expression levels and may mask the effect. It may also be due to differences in eQTLs in blood and brain tissues. Given the observed small differences in transcript levels and the deleteriousness of hemizygosity for TCF4 observed in Pitt-Hopkins syndrome, it seems reasonable that evolutionary constraint on variants that alter expression levels might exist. Another option is that the locus only exerts its regulatory effect in a developmental context, or is organ specific. The gene expression in utero and postnatal life has recently been examined, and the brain TCF4 mRNA level is highest in the early brain development in utero, decreases in childhood and stabilizes in early adulthood.\textsuperscript{57} Thus, epigenetic factors from early life may have a continued effect on expression\textsuperscript{69} and together with a decrease in variation explain the lack of association between GWAS SNPs and TCF4 mRNA level in adults.\textsuperscript{67} It is also possible that none of the current TCF4 SNPs tagged the eQTLs.

Taken together, these results support earlier findings of TCF4 as an important gene in psychosis etiology, and suggest that TCF4 variants may lead to a subgroup of psychosis with clinical, cognitive and brain structural abnormalities related to neurodevelopmental mechanisms.

Conflict of interest
The authors declare no conflict of interest.

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Supplementary Information accompanies the paper on the Translational Psychiatry website (http://www.nature.com/tp)