Exploring the Impact of Climatic Variables on Arecanut Fruit Rot Epidemic by Understanding the Disease Dynamics in Relation to Space and Time

Balanagouda Patil 1,2,*  Vinayaka Hegde 2  Shankarappa Sridhara 3  Hanumappa Narayanaswamy 1  Manjunatha K. Naik 1, Kiran Kumar R. Patil 4, Hosahatti Rajashekara 5 and Ajay Kumar Mishra 6,*

1 Department of Plant Pathology, University of Agricultural and Horticultural Sciences, Shivamogga 577255, Karnataka, India; swamyareca@gmail.com (H.N.); manjunaik2000@yahoo.co.in (M.K.N.)
2 Division of Crop Protection, ICAR-Central Plantation Crops Research Institute, Kasaragod 671124, Kerala, India; vinayaka.hegde@icar.gov.in
3 Center for Climate Resilient Agriculture, University of Agricultural and Horticultural Sciences, Shivamogga 577255, Karnataka, India; sridharas1968@gmail.com
4 Department of Agricultural Economics, University of Agricultural and Horticultural Sciences, Shivamogga 577255, Karnataka, India; kiranecon@gmail.com
5 Division of Crop Protection, ICAR-Directorate of Cashew Research, Puttur 574202, Karnataka, India; rajashekara.h@icar.gov.in
6 Khalifa Centre for Genetic Engineering and Biotechnology, United Arab Emirates University, Al Ain P.O. Box 15551, United Arab Emirates

* Correspondence: balupat007@gmail.com (B.P.); ajaymishra24@uaeu.ac.ae (A.K.M.)

Abstract: To understand the spatio-temporal dynamics and the effect of climate on fruit rot occurrence in arecanut plantations, we evaluated the intensity of fruit rot in three major growing regions of Karnataka, India for two consecutive years (2018 and 2019). A total of 27 sampling sites from the selected regions were monitored and the percentage disease intensity (PDI) was assessed on 50 randomly selected palms. Spatial interpolation technique, ordinary kriging (OK) was employed to predict the disease occurrence at unsampled locations. OK resulted in aggregated spatial maps, where the disease intensity was substantial (40.25–72.45%) at sampling sites of the Malnad and coastal regions. Further, Moran’s I spatial autocorrelation test confirmed the presence of significant spatial clusters (p ≤ 0.01) across the regions studied. Temporal analysis indicated the initiation of disease on different weeks dependent on the sampling sites and evaluated years with significant variation in PDI, which ranged from 9.25% to 72.45%. The occurrence of disease over time revealed that the epidemic was initiated early in the season (July) at the Malnad and coastal regions in contrary to the Maidan region where the occurrence was delayed up to the end of the season (September). Correlations between environmental variables and PDI revealed that, the estimated temperature (T), relative humidity (RH) and total rainfall (TRF) significantly positively associated (p = 0.01) with disease occurrence. Regression model analysis revealed that the association between Tmax, RH1 and TRF with PDI statistically significant and the coefficients for the predictors Tmax, RH1 and TRF are 1.731, 1.330 and 0.541, respectively. The information generated in the present study will provide a scientific decision support system, to generate forecasting models and a better surveillance system to develop adequate strategies to curtail the fruit rot of arecanut.

Keywords: arecanut; fruit rot disease; spatio-temporal pattern; ordinary kriging; climatic indicators; epidemiology; spatial dependency; GLM

1. Introduction

Arecanut (Areca catechu L.) is a tropical industrial tree species that plays a crucial role in providing livelihood and economic security to millions of people in India. It is one of...
the most predominant industrial crops in Southern and South-East countries, including India, China, Indonesia and Malaysia [1]. Areca nut cultivation generates direct and indirect job opportunities, creates a diverse array of materials to several small- and medium-scale industries and plays a critical role in conservation of the soil and ecosystem [2]. Even though the areca nut cultivation is location-specific, its consumption is widespread throughout the country. The areca nut-based industries primarily act as a major sole source of income to six million areca nut farmers and stakeholders [3]. Evidence has been suggested that areca nut has different pharmacological benefits, such as anti-parasitic, bacterial, fungal, inflammatory and analgesic effects [4].

Areca nut is majorly grown in the southern and northeastern parts of the country, which includes Karnataka, Kerala, Tamil Nadu, West Bengal and Assam, which shares 90% of the total area and production [5]. Although the areca nut production is restricted to a few states, the major industrial products are globally shared, and about 700 million consume areca nuts world-wide [6,7]. India contributes about 57% of the global areca nut production and the remaining 43% was shared among China, Bangladesh, Myanmar and other countries. Irrespective of the areca nut growing regions, nuts were generally harvested upon maturity in the December–January months. Despite areca nut having industrial importance, the maximum potential is highly prone to attack by a wide range of biotic and abiotic stresses at all the growth stages. Areca nut production is majorly impaired by various fungal diseases. Among them, fruit rot stands at the topmost position, which results in the complete death of the palms either individually or for the whole plantation, which accounted for about 10–90% of the potential production [8]. Losses were over 75% or complete damage of the palm in Karnataka during 1910 [9] and further resulted in a significant economic loss of 15–20% in 1918 at Kerala [10–12], 50–90% in 1956 at parts of Malabar region [13] and 72–350 kg nuts ha$^{-1}$ in 1985 in North Canara [14] was estimated.

The occurrence of fruit rot disease (FRD) is primarily confined to traditional and regularly growing tracts belongs to Karnataka, Kerala, Tamil Nadu, West Bengal, Assam, parts of Andaman Islands and Meghalaya states. Recently, areca nut cultivation is continuously increasing due to its higher industrial and economic benefits in non-traditional growing tracts (Maidan regions), resulting in the spread and survival of the pathogen in these areas. For the first time, fruit rot was reported from the erstwhile Mysore province [15] and later from the endemic areas of Dakshina Kannada (South Canara) and Uttara Kannada (North Canara) in Karnataka state and disease-prone coastal tracts (Malabar and Cochin) of Kerala [9]. Recently, the disease has been increasing over a period of time due to the buildup of inoculum load within the gardens and micro-climatic conditions favoring disease spread and development [9,16].

The occurrence of a pathogen, Phytophthora meadii, which McRae (1918) associated with disease, was documented from areca nut-growing areas of Kerala and Karnataka. $P. meadii$ is an oomycetous hemi-biotrophic fungus that has harmful effects on areca nut by producing various symptoms at different stages of the palms [17–19]. The knowledge about the spatio-temporal patterns and the spread of the disease in disease-prone regions will provide valuable information to design suitable approaches for effective management of the disease [20]. Studies conducted to understand the spatio-temporal patterns of areca nut FRD and factors responsible for its spread are lacking. Information on the epidemiology and impact of weather parameters on FRD in these regions play a critical role in managing this disease due to the development of new virulent pathotypes of hemi-biotrophic oomycete fungus $P. meadii$ in the recent past [21,22].

Several factors might have contributed to the increased buildup of the disease, such as establishing new areca nut gardens in new regions, cultivating susceptible varieties across these areas and the unexpected change in climate and the frequent devastating cyclones [23]. Apart from these, the infection ability of $P. meadii$ on areca nut strongly depends on the virulence of pathogen that prevailed across the regions of areca nut cultivation.
Earlier studies on epidemiology and the survival ability of the pathogen revealed that *P. meadii* causing fruit rot perpetuates as a dormant mycelium either in the treetop on infected and dried bunches or rotted buds present on affected palms or in the top soil in the form of oospores [9,11,14,19,24]. However, precise information on the epidemiological cycle, development of epidemics and their mode of spread needs to be assessed through appropriate experimentation. The initial inoculum and intra-plot microclimate play a pivotal role in the development of epidemics.

Hence, there is a need to understand the level of the primary inoculum and its distribution between different survival sites. The description of the fruit rot epidemics regarding space and time scales in arecanut-growing areas may provide valuable information on the pathogen dissemination and epidemiological consequences. These kinds of studies are lacking at a regional scale in major arecanut-growing areas. Similar studies have been conducted in other plantation crops to describe the spatio-temporal patterns of black pod rot in cocoa at the regional scale in Cameroon. Recently, the spatial and temporal patterns of black pod rot disease were investigated in cocoa farms where the pathogen was already well established [25,26]. Further, experiments have mainly concentrated on point of infection on the pods by the pathogen in Nigeria [27], Brazil [28], the Solomon Islands [29] and the Ivory Coast [30].

The present study aimed to generate information on the spatio-temporal dynamics of FRD over different arecanut growing regions of Karnataka, India and to identify the environmental factors favoring the disease initiation and development. This study also aimed at assessing the possible impacts of climatic indicators on the regional occurrence of fruit rot epidemics as an aid for the adoption of efficient management tactics.

2. Materials and Methods

2.1. Ethical Declaration

No specific permission was required for the proposed nature of work. At every monitored sampling site, the arecanut growers permitted us to collect FRD-infected samples and to perform the sampling process. The investigation conducted during two consecutive years (2018 and 2019) did not involve any endangered or protected species of plants or animals. The fixed-plot surveys were conducted across the varied arecanut growing regions of Karnataka, and subsequent investigations were conducted at the Indian Council of Agricultural Research-Central Plantation Crops Research Institute, Kasaragod, Kerala, India.

2.2. Description of the Study Area

The field sampling and disease scoring was performed in three varied arecanut growing regions of Karnataka, India viz., coastal, Malnad and Maidan regions, respectively (Figure 1). The patterns of FRD regarding space and time scales were examined through continuous monitoring for disease (fixed-plot) in two successive seasons (Kharif 2018 and 2019). The description and particulars of the experimental area is provided in Table 1. The sampling points were selected randomly by considering uniformity in the PDI over a large area, and their distribution was chosen based on disease prevalence.

The agro-climatic regions were chosen based on their variations in climate and topographic profiles. Nine sampling sites were selected from each region to assess the disease intensity. The arecanut palms were 20–45 years old and were planted at a spacing of 2.7 × 2.7 m (1371 palms hectare⁻¹) in all the plantations considered for this study. The gardens were managed using no-tillage practices, and weed control was achieved with herbicidal treatments or manually.
The disease intensity of FRD in each garden was assessed weekly by visual observation of typical symptoms on palms along the rows throughout the entire garden. The locations of each diseased palm were mapped on the average monthly determined observations. Estimation of disease was done on randomly chosen and tagged fallen nuts per palm (n = 50) across the 18 weeks and then converted into the PDI. The PDI of fruit rot of arecanut was determined by the standard Formula (1) and disease rating scale (1–6) developed by Sastry and Hegde [31] as shown in Table 2.

\[
\text{Percent Disease Intensity (PDI)} = \frac{\text{Sum of numerical ratings}}{\text{Total number of palms observed} \times \text{Maximum rating}} \times 100
\]  

(1)

2.3. Estimation of Disease Variables

The climatic diversity and ecological profile of the three regions of Karnataka considered during this study.

Table 1. The climatic diversity and ecological profile of the three regions of Karnataka considered during this study.

| Characteristics | Malnad | Coastal | Maidan |
|-----------------|--------|---------|--------|
| Agro-ecological zones | The Western Ghats and Hot-Humid region (19.2) | Coastal Plain, Hot Humid Region (19.2) | Sub-Humid plain region (19.1) |
| Agro-climatic Region | Ghat region | West Coast Plains | Southern plateau |
| Agro-climatic Zone | Hilly (Malnad) zone | Coastal Zone | Northern transition zone |
| Altitude (m) | 1119 m | 69 m | 631 m |
| Latitude (Boundary) | 13°55′20.10″ N | 12°50′04.02″ N | 13°18′44.72″ N |
| Longitude (Boundary) | 75°34′14.73″ E | 75°14′54.92″ E | 75°46′13.10″ E |
| Annual rainfall (mm) * | 1812.5 | 2559.4 | 906.3 |
| Average temperature (°C) * | 25.6 °C | 27.6 °C | 29.8 °C |
| Soil type | Laterite clayey soils | Red Laterite soils | Red clay soils |

* Average values of two consecutive years 2018 and 2019.
Table 2. Description of the disease rating scale used to assess the PDI of FRD in arecanut [31].

| Scale | Description                                      |
|-------|--------------------------------------------------|
| 1     | 1–10% nut fall from bunches                      |
| 2     | 11–25% nut drop                                  |
| 3     | 26–50% nut drop                                  |
| 4     | 51–75% nut drop + spread of the disease to bunch stalk |
| 5     | 76–100% nut drop + spread of the disease to the main stalk of the bunch |
| 6     | Crown death                                      |

The PDI observations were collected weekly from the June to September months of monsoon season, totaling 18 weeks of observations per site.

2.4. Conceptual Framework of the Study

A conceptual framework regarding the procedures used to describe the spatio-temporal analyses and processes utilized during the study is presented in Figure 2. It provides a succinct summary of the methodology applied in this investigation and consists of data retrieval, analysis and processing using GIST tools.

2.5. Temporal Progression and Statistical Analysis of FRD at the Regional Scale

The PDI data was analyzed using the Kruskal–Wallis test in ‘R statistical software (version R-3.6.1)’ to determine the statistical difference observed among the sampling sites \((n = 9)\) in each region. Significant variation in PDI at sampling sites was assessed...
Temporal analysis was performed to understand the progression of the disease over sampling points among each region based on observations of disease intensity at weekly intervals during two consecutive years. Column graphs were plotted by using PDI against standard weeks at sampling sites in each region then dataset was compared among the years of experimentation and plots were prepared using ‘Graph-pad Prism (Version 9.0.1, GraphPad Software Inc. San Diego, CA, USA)’.

Generalized Linear Model (GLM)

The dependent variable PDI is count data that failed to follow a normal distribution. The influence of predictors, such as arecanut growing regions (Maidan, Malnad and coastal), 18 weeks starting from the June month to the end of September month and time element (years) on PDI cannot be estimated through multiple linear regression. Hence, a generalized linear model (GLM) with a Poisson or negative binomial regression with log link was tried.

Areanut growing regions having three categories are captured through two dummies, i.e., $D_1$ and $D_2$. $D_1$ takes the value ‘1’ and $D_2$ ‘0’ for Malnad region, $D_1 = 0$ and $D_2 = 1$ for the coastal region, and the control group was represented by $D_1 = 0$ and $D_2 = 0$. Though sampling weeks considered was from the first week of June to the last week of September, the disease occurrence was apparent only from the sixth week. Hence, the 6th to the 18th weeks were considered for model analysis and captured through dummies.

There were 13 categories; hence, 12 dummies were used, i.e., $D_1 = 1$, $D_2 = 0$, $D_3 = 0$, $D_6 = 0$, $D_7 = 0$, $D_8 = 0$, $D_9 = 0$, $D_{10} = 0$, $D_{11} = 0$, $D_{12} = 0$, $D_{13} = 0$ and $D_{14} = 0$ to represent the sixth sample week. The same analogy was followed to create the dummy matrix for the sample weeks. The influence of the time element, i.e., 2018 and 2019, was captured through dummy variable $D_{15}$ assuming a ‘0’ value for 2018 and ‘1’ for 2019. SAS software version 9.4M7 was used for the estimation of Poisson regression and negative binomial regression.

The deviance expected to be 1 is called ‘equi-dispersion’; if it is more than one, called ‘over-dispersion and less than one, called ‘under-dispersion. Generalized Linear Model (GLM) negative binomial regression with log link is given below.

$$\log(\text{PDI}) = \beta_1 + \beta_2 D_1 + \beta_3 D_2 + \beta_4 D_3 + \beta_5 D_4 + \beta_6 D_5 + \beta_7 D_6 + \beta_8 D_7 + \beta_9 D_8 + \beta_{10} D_9 + \beta_{11} D_{10} + \beta_{12} D_{11} + \beta_{13} D_{12} + \beta_{14} D_{13} + \beta_{15} D_{14} + \beta_{16} D_{15}$$

PDI = Percentage Disease Intensity; D1 and D2 represent dummy variables for the Malnad and coastal regions, respectively; whereas, D3–D14 indicted the dummy variables from 6th to 18th weeks, and D15 described the dummy variable for year.

2.6. Spatial Dynamics of FRD at Regional Scale

The spatial patterns of FRD distribution in varied regions of Karnataka were characterized by employing geostatistical tools viz., ordinary kriging (OK) approach and spatial autocorrelation. First, the OK tool was used to create spatial maps to predict FRD patterns across the regions that helps to identify significant clusters of disease in arecanut-growing areas. Secondly, the identified disease-prone areas were optimized through autocorrelation analysis and these analyses were performed using ArcGIS 10.3 version software. The region-wise sample data collected were tabulated indicating the latitude, longitude and PDI. Since PDI data was log $(x + 1)$ transformed to understand the distribution of data set.

The OK approach was employed to estimate the PDI values of the spatial phenomenon at non-sampled locations, such as the disease intensity of FRD at locations $(X_1, X_2, \ldots, X_n)$ are $(Z_1, Z_2, \ldots, Z_n)$. The objective of spatial data view is to predict the $Z$ value at unsampled points of $X$. Theoretically, kriging approach was used to optimize the surface area to predict random parameter $Z$ at different unsampled locations [32,33]. A surface map of disease
intensity of fruit rot was generated with the help of robust OK technique and can be mathematically expressed as following Equation (2):

\[ Z(x_0) = \sum_{i=1}^{n} \lambda_i z(x_i) \]  

(2)

where \( z \) is the PDI at locations \( x_i \) and \( x_0 \); \( n \) is the number of neighbors taken into consideration; \( x_i \) and \( x_0 \) are the sites observations recorded; \( \lambda_i \) are the weights; and \( z(x_i) \) is the value of \( z \) at \( x_i \) \[34–37\].

2.7. Spatial Autocorrelation

To assess the degree of spatial autocorrelation among the neighboring plots having infected palms, Moran’s \( I \) statistic was utilized to compute the spatial dependence between experimental sites in each region \[38–40\]. The neighborhood category was taken into consideration as that of the nearest neighbors among different locations \[41–43\]. The interpretation of the spatial autocorrelation output was performed through Moran’s \( I \) test and inferred using a \( p \)-value, which estimated the existence of significant spatial clusters. The following Equation (3) was used to determine the spatial dependency is as mentioned below:

\[ I = \frac{Z}{Z} \sum_{i=1}^{n} W - Z \]  

(3)

where \( I \) is the statistic for site \( i \), \( Z \) indicates the difference in PDI values at \( i \), and \( n \) is the average PDI for three regions calculated separately. \( W \) is the weight matrix that, in this case, only considered neighbors that share a common border or vertex.

2.8. Monitoring of Climatic Variables

An attempt was made to investigate the effect of environmental variables on FRD epidemics on arecanut plantations. The PDI was recorded from a farmer’s garden and research stations in varied regions of Karnataka. Environmental variables, such as the minimum and maximum temperature (\( T_{\text{max}} \); \( T_{\text{min}} \), °C), first half (morning) and second half (evening) Relative Humidity (RHI; RHII, %), Total Rainfall (TRF, mm), Number of rainy days (RD) and Wind Speed (WS, km/h), of the sampling points were collected from the nearest meteorological observatory maintained by the Agricultural and Horticultural Research Station (AHRS) belonging to the University of Agricultural and Horticultural Sciences, Shivamogga, Karnataka, India as well as from the Karnataka State Natural Disaster Monitoring Centre (KSNDMCC), Bengaluru, Karnataka, India for two consecutive years (2018 and 2019).

The daily data were converted into weekly averages for \( T_{\text{max}}, T_{\text{min}}, \text{RHI}, \text{RHII} \) and \( \text{WS} \) whereas weekly totals were considered for TRF and RD. The combined data from the sampling sites in each region during 2018 and 2019 were gathered to analyze the correlation between disease intensity and climatic factors using ‘R statistical software (version R-3.6.1)’. A correlation was performed to investigate whether the micro-climate conditions were similar within or between gardens.

Further, we conducted multiple linear regression model to estimate the relationship between FRD (dependent variable, PDI) and seven independent variables (\( T_{\text{max}}, T_{\text{min}}, \text{RHI}, \text{RHII}, \text{WS}, \text{TRF} \) and \( \text{RD} \)), which often called as predictors or explanatory variables. The dataset combined from 2018 and 2019 were considered to estimate the predictable impact of explanatory parameters on response variable and also used to infer causal relationship between both the variables. The regression analysis was conducted in ‘R statistical software (version R-3.6.1)’. Confidence and prediction intervals were displayed at 95%, which gives the range of likely values from the mean response and single observation.

3. Results

3.1. Disease Intensity of Fruit Rot on Areca nut in Karnataka

The pathogen infection was found throughout the sampling points (except few points in Maidan areas) over the regions with varying disease intensity (PDI), which significantly
differed across the studied regions of Karnataka. A significant difference in PDI was observed among sampling points in the Malnad region, where S2 exhibited a higher disease proportion than other points. In contrast, sampling sites in the coastal region did not show considerable differences in disease intensity that indicated almost similar trend was observed across the region. Disparately, no disease was observed in few sampling plots of the Maidan region (S1, S7, S8 and S9) even though the remaining plots exhibited slight variation with minimum disease proportion (Figure 3).

Figure 3. Box and whisker plot representing intensity (PDI) of fruit rot across different regions of Karnataka. The middle bar = median, box = inter-quartile range (25th–75th percentile) and whiskers (error bars) above and below the box represents the 90th and 10th percentiles.

3.2. Temporal Analysis of FRD

FRD progression over time in the Malnad region during both years substantially differed and was not consistent among the sampling plots for disease intensity (Figure 4). In all the plots, the disease intensity began during the sixth week of observation (second fortnight of July) in both seasons. Then, the disease progression linearly increased over
weeks of observations and from the initiation of disease development. Further, it progressed consistently with the varied proportion of disease until the end of the season.

Figure 4. The temporal progress of arecanut fruit rot PDI during two consecutive years at sampling sites in each region. Each point was the mean of 18 weeks of PDI data at different sampling sites \((n = 9)\) collected during 2018 and 2019. Graphs represented the mean, range and standard deviation of PDI, and the vertical bar represents the standard error of means.

Whereas, at the coastal region, the disease intensity was seen early (first week of July) compared to other regions irrespective of sampling sites where arecanut palms showed initiation of disease. Further, it was distributed virtually throughout the plots and neighboring places with different disease proportions (Figure 4). From the fifth week onwards, the disease was initiated tremendously up to the end of August. However, decreased disease intensity was noticed during the 15th to 18th week of disease monitoring.

Contrastingly, at the five experimental sites of the Maidan region, disease intensity progression seemed to exhibit a gradient, which was less evident at other regions (Figure 4). During both seasons, the disease to be initiated from the 12th week of observation and disease occurrence gradually increased over time with slight variation in disease proportion. In some of the Maidan region sampling sites, the disease was not noticed during both seasons after 18 weeks of observation.

3.3. Generalized Linear Model: Negative Binomial Regression with Log Link

We constructed a GLM model, which explained the FRD intensity through the proposed explanatory variables and then applied Akaike Information Criteria (AIC) to confirm the better fit of the GLM model: negative binomial regression with log link.
The estimates of the GLM negative regression model with log link and their standard errors are presented in Table 3 and Supplementary Figure S1. The AIC was lower in negative binomial regression (5216) compared to Poisson regression (infinite). The other criteria that reiterated negative binomial regression for the PDIS are deviance factors. The residual deviance, similar to the residual sum of squares in usual multiple regression, is expected to be low/minimum. The residual deviance in the case of negative binomial regression was 1.4, which was substantially lower than Poisson regression with the deviance of 5.6. The results outlined below are pertinent to negative binomial regression.

Table 3. Summary and estimates of GLM negative binomial regression model with log link.

| Covariates | Estimates | Standard Error | Rate Ratio | z Value | Pr (>|z|) | Significance |
|------------|-----------|----------------|------------|---------|----------|--------------|
| Intercept  | 1.971     | 0.136          | 7.179      | 14.533  | 2 × 10^{-16} | ***          |
| D1         | 2.384     | 0.088          | 10.843     | 26.951  | 2 × 10^{-16} | ***          |
| D2         | 2.189     | 0.089          | 8.924      | 24.701  | 2 × 10^{-16} | ***          |
| D3         | -2.465    | 0.186          | 0.085      | -13.229 | 2 × 10^{-16} | ***          |
| D4         | -1.838    | 0.179          | 0.159      | -10.291 | 2 × 10^{-16} | ***          |
| D5         | -1.592    | 0.176          | 0.204      | -9.026  | 2 × 10^{-16} | ***          |
| D6         | -1.407    | 0.175          | 0.245      | -8.046  | 8.58 × 10^{-16} | ***          |
| D7         | -1.276    | 0.174          | 0.279      | -7.34   | 2.14 × 10^{-16} | ***          |
| D8         | -0.986    | 0.172          | 0.373      | -5.73   | 1.01 × 10^{-13} | ***          |
| D9         | -0.729    | 0.171          | 0.482      | -4.269  | 1.97 × 10^{-5} | ***          |
| D10        | -0.398    | 0.169          | 0.672      | -2.35   | 0.0188   | *            |
| D11        | -0.293    | 0.169          | 0.746      | -1.733  | 0.0831   |              |
| D12        | -0.206    | 0.169          | 0.813      | -1.223  | 0.2212   |              |
| D13        | -0.131    | 0.169          | 0.878      | -0.775  | 0.4385   |              |
| D14        | -0.006    | 0.168          | 0.994      | -0.036  | 0.9716   |              |
| D15        | 0.050     | 0.069          | 1.052      | 0.728   | 0.4668   |              |

Significance codes: ***—0.001, *—0.05, '.'—0.1 and no marking—1. (Dispersion parameter for Negative Binomial (1.4024) family taken to be 1). Null deviance: 1876.83 on 701 degrees of freedom. Residual deviance: 933.88 on 686 degrees of freedom. AIC: 5216, Number of Fisher Scoring iterations: 1.

All the regression coefficients excepting dummy variables representing 14th, 15th, 16th and 17th sample weeks and time elements were statistically significant at less than the level of significance (1%). The obtained estimate of disease intensity for the Malnad region irrespective of sample weeks and sample years was 2.38. The rate ratio for the same was estimated by taking its antilog as 10.80, indicating that disease intensity in the Malnad region is 10.80-times more than the control group, i.e., the Maidan region. Similarly, the rate ratio for the coastal region was estimated at 8.94, reflecting that the disease intensity in the coastal region was 8.94-times more than the control group (Maidan region).

Concerning sample weeks, irrespective of regions and years, the PDI indicated by the rate ratio for the sixth week reflected that disease intensity was lower by 91% compared to the 18th week (control week). With the progression in the sample weeks, the likelihood of disease intensity increased. For instance, the disease intensity in the 10th week was 72% lower than the 18th week, while it was 1% lower in the 17th week compared to the 18th week (control). Year element has a less significant influence on the response variable with a rate ratio of 1.05, indicating that PDI was 5% higher in 2019 than in 2018.

The negative binomial regression model is additive in the log scale, and the additive model fits reasonably well for the data considered in the study. The predicted values of PDI across sample weeks for all the three regions and differences between Malnad and Maidan; coastal and Maidan are presented in Table 4. The regional differential/gradient of tenfold translates the PDI to 6.62 in the sixth week and reaches 77.84 in the 18th week. Thus, it could be inferred that the absolute effect of regions measured in the original scale increases with sampling weeks. The same analogy holds for the coastal region wherein the regional differential/gradient of eightfold translates PDI to 5.45 in the sixth week and reaches 64.06 in the 18th week.
Table 4. Predicted values of PDI for arecanut growing regions across the sampled weeks.

| Regions/Weeks    | 6th  | 7th  | 8th  | 9th  | 10th | 11th | 12th | 13th | 14th | 15th | 16th | 17th | 18th |
|------------------|------|------|------|------|------|------|------|------|------|------|------|------|------|
| Malnad region    | 6.62 | 12.39| 15.85| 19.07| 21.72| 29.04| 37.55| 52.28| 58.08| 63.32| 68.31| 77.38| 77.84|
| Coastal region   | 5.45 | 10.19| 13.04| 15.69| 17.88| 23.90| 30.91| 43.03| 47.80| 52.12| 56.22| 63.68| 64.06|
| Maidan region    | 0.61 | 1.14 | 1.46 | 1.76 | 2.00 | 2.68 | 3.46 | 4.82 | 5.36 | 5.84 | 6.30 | 7.14 | 7.18 |

3.4. Dynamics of FRD over Space

Spatial interpolated maps generated using ordinary kriging (OK) showed the spatial aggregation pattern of disease intensity over weeks (n = 18) of observation in the varied regions (n = 3). Among the studied areas, FRD was widely distributed and spatially aggregated in the Malnad and coastal regions compared to the Maidan region, which had random occurrence of FRD.

Spatial Interpolation through the OK Approach

The results of OK are depicted through color-coded maps where the darker color (red) represents a higher disease intensity of FRD and lighter color (green) indicates a lower disease variable. Of the total interpolated surface at the Malnad region during 2018 and 2019, 2.5% and 6% showed higher disease intensity ranged from 61–70%, respectively.

The disease intensity up to 51–60%, 41–50% and 31–40% was observed in 22.3%, 25% and 7.5% of the total surface, respectively, indicating spatial aggregation of disease during 2018. Subsequently, around 60%, 12% and 3% were observed in 2019 (Figure 5A). This pattern of spatial clusters evidenced that the disease intensity was varied among sampling sites of the region with varying disease proportions and identified risk areas highly vulnerable to the disease spread and occurrence.

Figure 5. Cont.
Interpolated maps represent disease intensity of FRD using ordinary kriging approach at Malnad (A), coastal (B) and Maidan (C) regions of Karnataka during 2018 and 2019.

From the appearance of disease until the end of the period during both seasons, the disease intensity displayed an aggregated pattern of disease occurrence in the coastal region.
The maximum surface area during 2018 (about 90%) exhibited disease intensity between 41% and 50%, and the remaining (10%) areas witnessed disease proportion between 51% and 60% representing potential risk areas (Figure 5B). Throughout the season of 2019, 50% of the interpolated surface area depicted the disease intensity of >51% and 41%, respectively.

When looking at the distribution of disease intensity overall plots, the Maidan region indicated an unequal or scattered pattern. The disease intensity ranged from 21 to 30% in 2018 than 2019, which was relatively lesser than other studied regions. Surprisingly, around four sampling sites in the region comprised no disease intensity (40% interpolated areas), which is non-obvious compared to other regions depicting less disease-prone or risk areas.

In 2018, 11–20% of disease intensity was observed in 55% of the total interpolated surface, and the remaining 5% of areas consisted of a disease proportion of about 21–30% (Figure 5C). The disease intensity during 2019 varied between 11% and 20%, which shared at 60% of the total surface, and only 1% had disease intensity ranged from 21–30%. It seems that general trend was maintained across the regions in both years.

3.5. Spatial Autocorrelation

The Moran’s $I$ statistical test on the PDI for 18 standard weeks during 2018 and 2019 at the observational sampling points in combined regional data were depicted in Table 5. Moran’s $I$ spatial autocorrelation confirmed the presence of a significant ($p < 0.05$) spatial clustering pattern among PDI of FRD from the sixth week onwards. At all sampling sites ($n = 27$), up to the fifth standard week, the Moran’s $I$ statistic value was zero, indicating no distinct clusters observed across the studied areas (Table 5), which is also reflected in interpolation results. After the sixth week onwards, a strong spatial dependency among 27 sampling sites was witnessed and significantly exhibited distinct spatial clusters ($p < 0.05$). Furthermore, the spatial distribution pattern depicted the similar clusters indicating that the same cluster category was bounded on another cluster category with Moran’s $I$ statistic close to 1.

| Standard Week | 2018 | 2019 |
|---------------|------|------|
|               | Moran’s $I$ Statistic | $p$-Value (0.05) | Moran’s $I$ Statistic | $p$-Value (0.05) |
| W1            | 0.00 | 0.00 | 0.00 | 0.00 |
| W2            | 0.00 | 0.00 | 0.00 | 0.00 |
| W3            | 0.00 | 0.00 | 0.00 | 0.00 |
| W4            | 0.00 | 0.00 | 0.00 | 0.00 |
| W5            | 0.00 | 0.00 | 0.00 | 0.00 |
| W6            | 0.151 | 0.160 | $-0.005$ | 0.820 |
| W7            | 0.591 | 0.06 | 0.460 | 0.022 |
| W8            | 0.650 | 0.01 | 0.562 | 0.014 |
| W9            | 0.656 | 0.001 | 0.559 | 0.001 |
| W10           | 0.639 | 0.02 | 0.550 | 0.0027 |
| W11           | 0.682 | 0.001 | 0.516 | 0.0074 |
| W12           | 0.657 | 0.005 | 0.498 | 0.001 |
| W13           | 0.620 | 0.0001 | 0.522 | 0.0057 |
| W14           | 0.600 | 0.006 | 0.524 | 0.005 |
| W15           | 0.630 | 0.008 | 0.552 | 0.002 |
| W16           | 0.660 | 0.0041 | 0.584 | 0.008 |
| W17           | 0.700 | 0.0036 | 0.576 | 0.009 |
| W18           | 0.715 | 0.0075 | 0.623 | 0.002 |
3.6. Influence of Climatic Variables on Fruit Rot Epidemics

3.6.1. Correlations between Fruit Rot Intensity and Climatic Determinants

For all the environment variables (n = 7), a strong relationship between points were observed in all the plots. Bivariate correlation analysis revealed a significant (p > 0.01) correlation between environmental variables and FRD intensity across the sampling sites of Karnataka.

The results from the combined dataset, the correlation between T\textsubscript{max} and T\textsubscript{min} (r = 0.80, p = 0.01), RH (r = 0.88, p = 0.01), WS (r = 0.79, p = 0.01), TRF (r = 0.88, p = 0.01) and FRD occurrence was strong positive and significant (p > 0.01) but correlation of RD with disease intensity was slightly positive (p > 0.01) (Figure 6). The correlation results output clearly indicated that FRD has strong relation with temperature, relative humidity and rainfall across the sampling sites. This variation in prevailed environmental situations and microclimate within or between gardens suggest significance of the weather parameters in FRD epidemics at various sampling sites in Karnataka.

Furthermore, the plot indicating the maximal PDI of each of the 27 sites against integrated climatic variables over the 18 weeks period inferred in Figure 7. This clearly deciphered and supported the results of correlation analysis as well as allowed us to understand the impact of climatic indicators against PDI across the sampling sites.

Figure 6. Correlation analysis between FRD intensity and climatic conditions across the 27 sampling sites of Karnataka. T\textsubscript{max} (Maximum Temperature), T\textsubscript{min} (Minimum Temperature), RH1 (Morning Relative Humidity), RH2 (Evening Relative Humidity), WS (Wind Speed), TRF (Total Rainfall), RD (Number of Rainy Days) and PDI (Percentage Disease Intensity). Significance codes: *** —0.001, ** —0.01, * —0.05.

Furthermore, the plot indicating the maximal PDI of each of the 27 sites against integrated climatic variables over the 18 weeks period inferred in Figure 7. This clearly deciphered and supported the results of correlation analysis as well as allowed us to understand the impact of climatic indicators against PDI across the sampling sites.
Figure 6. Correlation analysis between FRD intensity and climatic conditions across the 27 sampling sites of Karnataka. T max (Maximum Temperature), T min (Minimum Temperature), RH1 (Morning Relative Humidity), RH2 (Evening Relative Humidity), WS (Wind Speed), TRF (Total Rainfall), RD (Number of Rainy Days) and PDI (Percentage Disease Intensity). Significance codes: ***—0.001, **—0.01, *—0.05.

Furthermore, the plot indicating the maximal PDI of each of the 27 sites against integrated climatic variables over the 18 weeks period inferred in Figure 7. This clearly deciphered and supported the results of correlation analysis as well as allowed us to understand the impact of climatic indicators against PDI across the sampling sites.

Figure 7. Graph representing the maximum PDI and integrated climatic variables across the 27 sampling sites and deciphered the clustering of level of PDI with climatic indicators.

3.6.2. Multiple Linear Regression Model

Table 6 represents the parameter estimates to understand the possible associations between occurrences of FRD and climatic covariates along with corresponding confidence interval (CI). Based on combined dataset (2018 and 2019), there was considerable association was noticed between climatic variables and PDI across the studied areas. The significant association between T max, RH1 and TRF with response variable (PDI) was determined. In these results p-value (significance) for T max, RH1 and TRF is \( \leq 0.001 \), which is less than significance level of 0.05.

Table 6. The estimated effects of climatic indicators on FRD analyzed through multiple linear regression model.

| Covariates | \( \beta \) | \( t \)-Value | Sig. | 95% Confidence Interval (CI) |
|-----------|----------|-------------|------|-----------------------------|
| (Constant) | 168.51   | 3.885       | 0.001 *** | 72.241 210.316 |
| T max     | 1.731    | 4.514       | 0.001 *** | 0.795 6.235 |
| T min     | 1.006    | 8.213       | 0.030 **  | −6.473 20.852 |
| RH1       | 1.330    | 4.418       | 0.001 *  | −18.150 24.21 |
| RH2       | 1.064    | 1.791       | 0.013 **  | −3.381 1.856 |
| WS        | 0.452    | 0.435       | 0.055 *   | −1.005 4.526 |
| TRF       | 0.541    | −0.673      | 0.001 **  | −2.583 1.627 |
| RD        | −0.604   | −2.182      | 0.117 *   | −2.851 0.901 |
| Region    | 2.065    | 1.935       | 0.001 *** | 0.126 3.172 |

\( T_{\text{max}} \) (Maximum Temperature), \( T_{\text{min}} \) (Minimum Temperature), RH1 (Morning Relative Humidity), RH2 (Evening Relative Humidity), WS (Wind Speed), TRF (Total Rainfall), RD (Number of Rainy Days). Sig. indicates the level of significance at (\( p = 0.05 \)); and \( \beta \) represents the estimated coefficients of variables. Significance codes: ***—0.001, **—0.01, *—0.05.

This indicated that the association between T max, RH1 and TRF with PDI statistically significant. The coefficients for the predictors T max, RH1 and TRF are 1.731, 1.330 and 0.541, respectively. This indicated that the FRD intensity increases by 1.731, 1.330 and 0.541 for every 1 unit increase in T max, RH1 and TRF. The sign of the coefficient is positive, which
revealed that as $T_{\text{max}}$, RH1 and TRF increased, FRD intensity also increased (Table 6 and Supplementary Figure S2). The remaining predictors, such as $T_{\text{min}}$, RH2, WS and RD, were found slightly associated with response variable as level of significance is more than 0.05. This represented that the occurrence of FRD is independent on $T_{\text{min}}$, RH2, WS and RD factors.

The overall results from regression model analysis demonstrated that, maximum temperature, relative humidity and rainfall pattern were found to be key determining factors in prevalence and spread of the FRD in arecanut plantations over studied sites of Karnataka, India.

4. Discussion
4.1. Spatial Pattern Analysis of FRD

The studied regions were varied by climatic profiles, altitudinal characteristics and initial inoculum of the pathogen, which decides the distribution of fruit rot epidemic across the regions. The disease pattern displayed a gradient across the regions as confirmed by ordinary kriging (OK) and spatial dependency, which represented the nature of disease distribution at sampling sites throughout investigated areas.

The spatial clustering was varied throughout the regions as evidenced by spatial autocorrelation analysis, which had significant spatial clusters between weeks at all the sampling sites ($n = 27$). The spatial analysis and dependency exhibited an aggregated pattern of infections by the pathogen at Malnad and coastal regions; however, Maidan regions had observed random patterns. The findings of our study are corroborated with similar results on the spatio-temporal patterns of bud rot on coconut, which observed in multiple sites and spread across the studied area with regular and contagious or cluster-distribution patterns [44].

Ordinary kriging (OK) generated identical FRD prediction surfaces at Malnad and coastal regions but Maidan region had lesser predicted surface area and disease demonstrated relatively higher spatial correlation indicating spatial clusters. Due to presence of spatial dependency, the kriging approach was considered the most suitable surface interpolation technique than IDW due to distance-based outcomes; hence, OK generated maps were recommended in this study [45–47].

The relative geographical distribution of FRD showed the potential risk areas in the studied regions of Karnataka. The surface interpolated maps demonstrated the higher potential risk areas mainly concentrated in high altitude areas on Western-Ghats (Malnad tracts), Plateau and hilly areas and Seashore (coastal belt). By considering the results from OK demonstrated that, the topographical effects highly influenced disease distribution. A decade ago, disease was increasing to non-traditional plain areas (Maidan region) as revealed in OK surface maps, including arecanut plantations established along the riverside (Figure 8).

The variations in the aggregation of spatial effects between sampling sites at Malnad and coastal regions might be credited to the difference in the age of the palms; hence, gardens at Malnad and coastal regions were established before long back (>25 years older) than other Maidan regions (recently established) would be highly vulnerable to attack by the pathogen ($P. \text{meadii}$). Over the years, the initial disease intensity that existed might have disseminated throughout the arecanut plantations; however, the random pattern of disease distribution was witnessed at Maidan region.

This idea is further supported by data, which showed that in the plantations >20 years old and well established, 16 samples (88%) out of 18 tested positive for $P. \text{megakarya}$ [48]. The intra-plot environment, especially the presence of other plantation crops, such as cocoa, black pepper and banana, as potential intercrops between the rows of arecanut at the Malnad and coastal region influenced the inoculum buildup and spread within the garden due to increased relative humidity.
results on the spatio-temporal patterns of bud rot on coconut, which observed in multiple sites and spread across the studied area with regular and contagious or cluster-distribution patterns [44].

Ordinary kriging (OK) generated identical FRD prediction surfaces at Malnad and coastal regions but Maidan region had lesser predicted surface area and disease demonstrated relatively higher spatial correlation indicating spatial clusters. Due to presence of spatial dependency, the kriging approach was considered the most suitable surface interpolation technique than IDW due to distance-based outcomes; hence, OK generated maps were recommended in this study [45–47].

The relative geographical distribution of FRD showed the potential risk areas in the studied regions of Karnataka. The surface interpolated maps demonstrated the higher potential risk areas mainly concentrated in high altitude areas on Western-Ghats (Malnad tracts), Plateau and hilly areas and Seashore (coastal belt). By considering the results from OK demonstrated that, the topographical effects highly influenced disease distribution. A decade ago, disease was increasing to non-traditional plain areas (Maidan region) as revealed in OK surface maps, including arecanut plantations established along the riverside (Figure 8).

Deviations in disease patterns over space were attributed to rainfall patterns, congenial environmental situations, development of microclimate within gardens, dispersal of the inoculum within palms, movement the infectious spores between-palms, fields and neighboring gardens; management practices. It was evident that, the disease was highly correlated with temperature, relative humidity and rainfall pattern at Malnad and coastal regions. It was hypothesized that the presence of initial inoculum is the key factor for the spatio-temporal development of FRD at the plantation level [26]. The temperature has significantly influenced the occurrence of \( P. \) megakarya on cocoa plantations at a lower temperature between 23 and 26 °C [49,50], and the survival ability of \( P. \) megakarya showed that the soil is not only medium also the existence of intra-tree dispersal might be credited [51].

When compared to between and within the years, the location of infection clusters was consistent in coastal and Malnad regions. In contrast, it was not consistent in the Maidan region. When looking at the data for the Maidan region, there did not have particular area that was consistently infected (random spatial effect was witnessed). This was reflected in the microclimate data, which did not show a plot having optimal microclimatic conditions favoring pathogen development.

In Malnad and coastal regions, microclimate was most conducive for the disease development in the plot where infections were consistently noticed. These areas receive more shade in the morning than other areas, which could induce dew on the surface of arecanut bunches for a prolonged period, facilitating the germination of spores of pathogen. These results were consistent with the reports demonstrated the persistent dew on the pod surface for up to 3 h after sunrise is ideal for black pod rot disease development [52]. Primary inoculum initiates disease and influences the development of epidemic thought-out the year was demonstrated on cocoa plantations [53].
4.2. Temporal Dynamics of FRD

It was expected that, over time, an increase in infections to larger areas with disease intensity was observed at Malnad and coastal regions. However, this was not the case in the Maidan region, where the disease was observed on a limited scale. Over time, disease progression across regions revealed that secondary infections are most often derived from primary ones than other infections. However, most infected palms generated secondary or primary inoculum, contributing to them being infected within the same year. The lack of increase of the disease over time is thus because secondary inoculum mainly causes self-infection but fails to disseminate between palms in the garden. This supports the hypothesis of Ten Hoopen et al., 2009 [26], demonstrating that the primary inoculum is the prime factor that favors the development of an epidemic over time at the plantation level.

Although variability in the distribution of rainfall and quantity could explain differences in disease intensity between years, it was remained relatively low in plots ($n = 9$) of the Maidan region. Two potential explanations could be attributed, weekly phytosanitation and fungicide treatments, to consider when interpreting the results. In Malnad and coastal regions, fungicidal applications were taken on a preventive basis. In both cases, the subsequent year showed a decrease in disease intensity; however, this decrease was quite substantial in the case of the Maidan region because of late occurrence of rainfall. However, rainfall is the prime explanatory variable for temporal dynamics of disease and is mainly linked to the number of infected palms within a garden [25,49,54].

As discussed above, hilly, Western-Ghats, Seashore areas had considerable variation in altitude, which characterized a microclimate with lower temperature, continuous erratic rainfall and higher humidity favors FRD development and spread. In arecanut-growing areas, these kinds of terrains are aggregated spatially, which resulted spatial patterns in disease intensity. The aggregations of FRD at a regional scale across districts and taluks were unequal due to the nature of disease prevalence. This was partially confirmed by the observation of no strong spatial autocorrelation at the Maidan region. The lack of spatial dependence at Maidan region and the strong spatial correlation was at Malnad and coastal over weeks across regions suggested that the presence of initial inoculum and microclimate prevailed in gardens (which could be from infected bunches, treetop and various pathogen structures surviving in the soil) might have played a significant role in the fruit rot epidemics on arecanut in major producing areas of Karnataka.

4.3. Implications in FRD Management

The pattern of spatio-temporal distribution of fruit rot on arecanut has very significant implications. Due to aggregated patterns of fruit rot at a regional scale, the disease monitoring attention should be focused on altitude and weather conditions. The distribution of disease and relatively stable high probable risk areas over the years would help us to design suitable disease management approaches.

High-risk areas are usually Western-Ghats (Malnad), hilly and mountainous areas (coastal), often located at the junction of several districts or states. Therefore, we should promote a professional pest control system for large-scale disease prevention. Most importantly the farmers and extension agents could use the spatial distribution maps generated in our study in devising precautionary measures and formulating management strataums to regulate the further dissemination of disease to neighboring districts or states. The highly vulnerable areas to disease were identified in our study, indicating that farmers in those areas should resort management approaches to curtail the disease effectively.

5. Conclusions

We demonstrated the spatial and temporal dynamics of FRD at various sampling sites across different regions (Malnad, coastal and Maidan) of Karnataka. The present study constitutes the first documentation of spatial and temporal dynamics by employing geostatistical and GLM analysis to generate gradient spatio-temporal maps across the regions.
The spatial pattern and autocorrelation on disease intensity at twenty-seven sampling plots across regions displayed random and aggregated effects over the evaluated seasons.

Further, temporal dynamics revealed that the early initiation of the epidemic was witnessed at the Malnad and coastal regions in contrary to the Maidan tracts where the occurrence was delayed up to the end of the season. The regional occurrence of FRD was greatly influenced by the average temperature (25–30 °C), relative humidity (>90%) and erratic rainfall (>1000 mm) as evident from the correlation and regression analysis. Furthermore, the results indicated that arecanut plantations with the presence of initial inoculum exhibited the maximum disease proportion over space and time as revealed in the temporal analysis. However, more attention should be focused on the reduction of initial inoculum in the gardens. The actual surface interpolated maps generated in this study would help to develop suitable management approaches, which rely on the reduction of further spread of FRD to neighboring areas. The epidemiological information on FRD in this investigation will be useful in developing forecast models as well as a decision support system as an aid for the design and implementation of control strategies for the disease.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/jof8070745/s1, Figure S1. Summary and estimates of the generalized linear model (GLM) with negative binomial regression model with log link, including different model parameters (count, ARC and linear (LN)). Figure S2. Regression analysis on the regional occurrence of FRD pitting climatic predictors in Malnad (A), coastal (B) and Maidan (C) regions of Karnataka.

Author Contributions: Conceptualization, methodology, formal analysis, writing—original draft preparation, writing—review and editing, B.P. and S.S.; supervision, project administration, investigation, V.H., M.K.N., H.N. and A.K.M.; formal analysis, K.K.R.P.; and writing—review and editing, H.R. and A.K.M. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: The data presented in this study are available on request from the corresponding author.

Acknowledgments: This is the collaborative doctoral research work of first author and we greatly acknowledge University of Agricultural and Horticultural Sciences, Shivamogga; ICAR-Central Plantation Crops Research Institute, Kerala, India and Biology Centre of the CAS, Institute of Plant Molecular Biology (IPMB) for providing necessary technical, financial and other guidance. We would like to thank the Director of Khalifa Center for Genetic Engineering and Biotechnology (KCGEB) for publication support.

Conflicts of Interest: The authors declare no conflict of interest.

References
1. Amudhan, M.S.; Begum, V.H.; Hebbbar, K.B. A review on phytochemical and pharmacological potential of Areca catechu L. seed. Int. J. Pharm. Sci. Res. 2012, 3, 4151–4157. [CrossRef]
2. Chowdappa, P.; Sharma, P.; Anandaraj, M.; Khetarpal, R.K. Diseases of Plantation Crops; Indian Phytopathological Society: New Delhi, India, 2014.
3. Rajagopal, V.; Balasimha, D. Areca nut; Central Plantation Crops Research Institute: Kasaragod, India, 2004; pp. 1–7.
4. Gilani, A.H.; Ghayur, M.N.; Saify, Z.S.; Ahmed, S.P.; Choudhary, M.I.; Khalid, A. Presence of cholinomimetic and acetylcholinesterase inhibitory constituents in betel nut. Life Sci. 2004, 75, 2377–2389. [CrossRef] [PubMed]
5. Kulkarni, A.R.; Mulani, R.M. Indigenous palms of India. Curr. Sci. 2004, 86, 1598–1603.
6. Guo, J.; Xie, H.; Mao, S.; Liang, M.; Wu, H. Efficacy of hyaluronidase combined with corticosteroids in treatment of oral submucous fibrosis: A meta-analysis of randomized controlled clinical trials. J. Oral Pathol. Med. 2020, 49, 311–319. [CrossRef] [PubMed]
7. Peng, W.; Liu, Y.; Wu, N.; Sun, T.; Yan, X.; Xiang, G.Y.; Wu, W. Areca catechu L. (Arecales): A review of its traditional uses, botany, phytochemistry, pharmacology and toxicology. J. Ethnopharmacol. 2015, 164, 340–356. [CrossRef]
8. Saraswathy, N. Diseases and disorders. In Areca nut; Rajagopal, V., Balasimha, D., Eds.; Central Plantation Crops Research Institute: Kasaragod, India, 2004; pp. 134–169.

9. Coleman, L.C. Diseases of the areca palm. In Koleroga; Mycological Series, Bulletin No. 2; Department of Agriculture: Mysore, Karnataka State, India, 1910; p. 92.

10. Coleman, L.C.; Rao, M.K.V. The Cultivation of Areca Palm in Mysore; Department of Agriculture: Bangalore, India, 1918; p. 32.

11. Kamath, M.N. Introductory Plant Pathology; Prakash Publishing House: Pune, India, 1956; p. 200.

12. Nambiar, K.K. Areca Nut Cultivation in India; Indian Council of Agricultural Research: New Delhi, India, 1956; p. 32.

13. Koti Reddy, M.; Anandaraj, M. Koleroga of arecanut. In The Proceedings of the Workshop on Phytophthora Diseases of Tropical Cultivated Plants, Kasaragod, India, 19–23 September 1980; Nambiar, K.K.N., Ed.; Central Plantation Crops Research Institute: Kasaragod, India, 1980; pp. 71–79.

14. Sastry, M.N.L.; Hedge, R.K. Taxonomic identity of arecanut Phytophthora isolates from the gardens of Sirsi, Uttara Kannada. In Areca nut Research and Development; Shama Bhat, K., Radhakrishnan Nair, C.P., Eds.; Central Plantation Crops Research Institute: Kasaragod, India, 1985; pp. 92–94.

15. Butler, E.J. Some diseases of palm. Agric. J. Indian 1906, 1, 299–310.

16. Balanagouda, P.; Sridhara, S.; Shil, S.; Vinayaka, H.; Naik, M.K.; Narayanaswamy, H.; Siva, K.B. Assessment of the spatial distribution and risk associated with fruit rot disease in Areca catechu L. J. Fungi 2021, 7, 797. [CrossRef]

17. Dutta, P.K.; Hegde, R.K. Studies on two Phytophthora diseases (Koleroga of arecanut and black pepper wilt) in Shimoga district, Karnataka State. Plant Pathol. 1987, 5, 12–29.

18. Santhakumari, P.; Hegde, R.K. Studies on Phytophthora diseases of plantation crops. Plant Pathol. 1987, 5, 1–2.

19. Saraswathy, N. Studies on Phytophthora spp. on Arecanut and Arecanut Based Cropping Systems. Ph.D. Thesis, Mangalore University, Karnataka, India, 1994.

20. Gilligan, C.A. A test for randomness by soilborne pathogens. Phytopathology 1985, 73, 300–303. [CrossRef]

21. Chowdappa, P. Phytophthora Causing Black Pod Disease of Cocoa (Theobroma cacao L.) in South India. Ph.D. Thesis, Mangalore University, Mangalagangothri, Mangalore, India, 1995.

22. Prathibha, V.H.; Hegde, V.; Sharadraj, K.M.; Suresh, K.R. Evaluation of fungicides and biocontrol agents against Phytophthora meadii infecting arecanut. Bioscan 2016, 11, 1547–1550.

23. Jose, C.T.; Balasimha, D.; Kannan, C. Yield loss due to fruit rot (Mahali) disease of arecanut in Karnataka. Indian J. Areca nut Spices Med. Plants 2008, 10, 45–51.

24. Uppal, V.N.; Desai, M.K. Koleroga disease of arecanut. Curr. Sci. 1939, 8, 122–124.

25. Ndoumbé-Nkeng, M.; Elombagn, M.I.B.; Bidzanga, N.L.; Sache, I.; Cilas, C. Spatio temporal dynamics on a plot scale of cacao black pod rot caused by Phytophthora megakarya in Cameroon. Eur. J. Plant Pathol. 2017, 147, 579–590. [CrossRef]

26. Ten Hoopen, G.M.; Sounigo, O.; Babin, R.; Yédé; Dikwe, G.; Cilas, C. Spatial and temporal analysis of a Phytophthora megakarya epidemic in a plantation in the Centre of Cameroon. In Proceedings of the 16th International Cacao Research Conference, Bali, Indonesia, 16 November 2009.

27. Opeke, L.K.; Gorentz, A.M. Phytophthora pod rot: Symptoms and environmental cause. In Phytophthora Disease of Cocoa; Gregory, P.H., Ed.; Longman: London, UK, 1998; pp. 117–125.

28. Medeiros, A.G. Sporulation of Phytophthora meadii (Butl.) in Relation to Epidemiology and Control of Cocoa Black Pod Disease. Ph.D. Thesis, University of California, Riverside, CA, USA, 1976.

29. Jackson, G.V.; Newhook, F.J. Sources of Phytophthora palmivora inoculum in Salomon Islands cocoa plantations. Trans. Brit. Mycol. Soc. 1978, 71, 239–249. [CrossRef]

30. Babacauh, K.D. Facteurs déterminant the localisation des lésions de pourriture brune des cabosses du cacaoyer (Theobroma cacao L.). I. Age de la cabosse et localisation of the lesion. Café Cacao Thé 1983, 27, 195–208.

31. Sastry, M.N.L.; Hedge, R.K. Phytophthora associated with arecanut (Areca catechu Linn.) in Uttara Kannada, Karnataka. Curr. Sci. 1987, 56, 367–368.

32. Madden, L.V.; Hughes, G.; van den Bosch, F. The Study of Plant Diseases Epidemics; The American Phytopathological Society: St. Paul, MN, USA, 2007.

33. Santra, P.; Chopra, U.; Chakraborty, D. Spatial variability of soil properties and its application in predicting surface map of hydraulic parameters in an agricultural farm. Curr. Sci. 2008, 95, 937–945.

34. Armstrong, M.; Boufassa, A. Comparing the robustness of ordinary kriging and lognormal kriging: Outlier resistance. Math. Geol. 1988, 20, 447–457. [CrossRef]

35. Stein, M.L. Interpolation of Spatial Data: Some Theory for Kriging; Springer Science & Business Media: New York, NY, USA, 2012.

36. Rhodes, H.R. Multivariate and Geostatistical Analyses of Phase-I Eastern Lake Survey Data. Ph.D. Thesis, University of Arizona, Tucson, AZ, USA, 1993.

37. Tomczak, M. Spatial interpolation and its uncertainty using automated anisotropic inverse distance weighting (IDW)—Cross-validation and jackknife approach. J. Geogr. Inf. Deci. Anal. 1998, 2, 18–30.

38. Bivan, R.S.; Pedesma, E.J.; Gomez-Rubio, V. Applied Spatial Data Analysis with R; Springer: Berlin/Heidelberg, Germany, 2008; p. 373.

39. Cliff, A.D.; Ord, J.K. Spatial Processes: Models and Applications; Pion: London, UK, 1981; p. 266.

40. Ristaino, J.B.; Gumpertz, M.L. New frontiers in the study of dispersal and spatial analysis of epidemics caused by species in the genus Phytophthora. Annu. Rev. Phytopathol. 2000, 38, 541–576. [CrossRef] [PubMed]
41. Reynolds, K.M.; Madden, L.V. Analysis of epidemics using spatio-temporal autocorrelation. *Phytopathology* **1988**, *78*, 240–246. [CrossRef]

42. Rémond, F.; Cilas, C.; Vega-Rosales, M.I.; Gonzalez, M.O. Méthodologie’ méchantillonnage pour estimer les attaques des baies du caféier par les scolytes (HypothenemushampeiFerr.). *Café Cacao Thé* **1993**, *37*, 35–52.

43. Johnston, K.; Ver Hoeff, J.M.; Krivoruchko, K.; Lucas, N. *Using ArcGIS Statistical Analyst*; Environmental System Research Institute: New York, NY, USA, 2001.

44. Juan-Bachiller, N.S. Distribution and progression of *Phytophthora* bud rot disease of coconut in selected areas in the Philippines. In *Diversity and Management of Phytophthora in Southeast Asia*; BPA Print Group Pty Ltd.: Melbourne, Australia, 2004.

45. Yao, X.; Fu, B.; Lü, Y.; Sun, F.; Wang, S.; Liu, M. Comparison of four spatial interpolation methods for estimating soil moisture in a complex terrain catchment. *PLoS ONE* **2013**, *8*, 546–560. [CrossRef]

46. Gong, G.; Mattevada, S.; O’bryant, S.E. Comparison of the accuracy of kriging and IDW interpolations in estimating groundwater arsenic concentrations in Texas. *Environ. Res.* **2014**, *130*, 59–69. [CrossRef]

47. Shahbeik, S.; Afzal, P.; Moarefvand, P.; Qumar, M. Comparison between ordinary kriging (OK) and inverse distance weighted (IDW) based on estimation error. *Arab. J. Geo.* **2014**, *7*, 3693–3704. [CrossRef]

48. Ndoungué-Nkeng, M.; Petchayo, S.; Techou, Z.; Nana, W.G.; Nembot, C.; Fontem, D.; Ten Hoopen, G.M. The impact of soil treatments on black pod rot (caused by *Phytophthora megakarya* of cacao in Cameroon. *Bio. Cont.* **2018**, *123*, 9–17. [CrossRef]

49. Ndoumbé-Nkeng, M.; Efombagn, M.I.B.; Nyassi, S.; Nyemb, E.; Sache, I.; Cilas, C. Relationships between cocoa *Phytophthora* pod rot disease and climatic variables in Cameroon. *Can. J. Plant Pathol.* **2009**, *31*, 309–320. [CrossRef]

50. Puig, A.S.; Ali, S.S.; Strem, M.;icher, R.; Gutierrez, O.A.; Bailey, B.A. The differential influence of temperature on *Phytophthora megakarya* and *Phytophthora palmivora* pod lesion expansion, mycelia growth, gene expression, and metabolite profiles. *Physiol. Mol. Plant Pathol.* **2018**, *102*, 95–112. [CrossRef]

51. Ali, S.S.; Amoako-Atta, I.; Bailey, R.A.; Strem, M.D.; Schmidt, M.; Akrofi, A.Y.; Surujdeo-Maharaj, S.; Kolawole, O.O.; Begoude, D.B.A.; Ten Hoopen, G.M.; et al. PCR-based identification of cacao black pod causal agents and identification factors possibly contributing to *Phytophthora megakarya* field dominance in West Africa. *Plant Pathol.* **2016**, *65*, 1095–1108. [CrossRef]

52. Monteith, L.J.; Butler, D.R. Dew and Thermal lag: A model for cocoa pods. *Quart. J. Met. Soc.* **1979**, *105*, 207–215. [CrossRef]

53. Nembot, C.; TakamSoh, P.; Ten Hoopen, G.M.; Dumont, Y. Modeling the temporal evolution of cocoa black pod rot disease caused by *Phytophthora megakarya*. *Math. Meth. Appl. Sci.* **2018**, *41*, 8816–8843. [CrossRef]

54. Deberdt, P.; Mtegue, C.V.; Tondje, P.R.; Bon, M.C.; Ducamp, M.; Hurard, C.; Begoude, B.A.D.; Ndoumbe-Nkeng, M.; Hebbar, P.K.; Cilas, C. Impact of environmental factors, chemical longicide and biological control on cacao pod production dynamics and black pod disease (*Phytophthora megakarya*) in Cameroun. *Biol. Cont.* **2008**, *44*, 149–159. [CrossRef]