Analysis of serum circulating MicroRNAs level in Malaysian patients with gestational diabetes mellitus

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Gestational diabetes mellitus (GDM) is a severe global issue that requires immediate attention. MicroRNA expression abnormalities are possibly disease-specific and may contribute to GDM pathological processes. To date, there is limited data on miRNA profiling in GDM, especially that involves a longitudinal study. Here, we performed miRNA expression profiling in the entire duration of pregnancy (during pregnancy until parturition and postpartum) using a miRNA-polymerase chain reaction array (miRNA-PCRArray) and in-silico analysis to identify unique miRNAs expression and their anticipated target genes in Malay maternal serum. MiRNA expression levels and their unique potential as biomarkers were explored in this work. In GDM patients, the expression levels of hsa-miR-193a, hsa-miR-21, hsa-miR-23a, and hsa-miR-361 were significantly increased, but miR-130a was significantly downregulated. The area under the curve (AUC) and receiver operating characteristic (ROC) curve study demonstrated that hsa-miR-193a (AUC = 0.89060 ± 0.04347, P = 0.0001), hsa-miR-21 (AUC = 0.89500 ± 0.04411, P = 0.0001), and miR-130a (AUC = 0.6939 ± 0.05845, P = 0.0025) had potential biomarker features in GDM. In-silico analysis also revealed that KLF (Kruppel-Like family of transcription factor), ZNF25 (Zinc finger protein 25), AFF4 (ALF transcription elongation factor 4), C1orf143 (long intergenic non-protein coding RNA 2869), SRSF2 (serine and arginine rich splicing factor 2), and ZNF655 (Zinc finger protein 655) were prominent genes targeted by the common nodes of miR23a, miR130, miR193a, miR21, and miR361. Our findings suggest that circulating microRNAs in the first trimester has the potential for GDM screening in the Malay population.

Gestational diabetes mellitus (GDM) is defined as any degree of glucose intolerance discovered during pregnancy1. Furthermore, GDM is a risk factor for maternal and fetal morbidity during pregnancy1. Babies born to women with GDM are more likely to have an excessive birth weight, known as macrosomia (weighing more than 4 kg), which can lead to juvenile obesity, type 2 diabetes, and/or cardiovascular disease later in life2. Globally, 21.3 million pregnancies are associated with hyperglycemia, with 18.4 million pregnancies related to GDM3. It has been shown that GDM is a complex disease with multiple etiologies4. Studies have shown that the development of GDM may be the result of a combined genetic and environmental effect, but the exact cause is still unknown5. GDM is diagnosed at the end of the second or early third trimester, depending on the physiological findings, and the generally accepted time for screening is the end of the second trimester, that is, between 24 and 28 weeks of pregnancy6. Over the past few decades, several studies have shown that aberrant expression of miRNAs is associated with pregnancy complications and the progression of GDM7, but longitudinal data on miRNA profiling in GDM is scarce. MiRNAs also function in blood glucose homeostasis and insulin production and secretion8. Furthermore, miRNAs have been reported to be present in biological fluids such as plasma/serum of diabetic patients and are highly stable there which can be easily detected and measured9. MicroRNA are short, single stranded RNA that post-translationally regulate gene expression10. miRNAs produced in the

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miR-193a, hsa-miR-21, hsa-miR-23a, hsa-miR-130a were significantly upregulated and hsa-miR-361, hsa-miR-126, hsa-miR-129 were significantly downregulated in GDM patients. In the second trimester, hsa-miR-let7-i, hsa-miR-126, hsa-miR-129, were significantly upregulated and hsa-miR-125, hsa-miR-129–2, hsa-miR-130a, hsa-miR-34 and hsa-miR-375 were significantly downregulated. In the third trimester, upregulation was observed with hsa-miR-let7e, hsa-miR-107, hsa-miR-361 and hsa-miR-370 while downregulated miRNAs were hsa-miR-125, hsa-miR-129 and hsa-miR-130a. In the postpartum period, data analysis revealed that two microRNAs hsa-miR-194 and hsa-miR-24 are significantly upregulated and three miRNAs hsa-miR-125, hsa-miR-129, and hsa-miR-375 are significantly downregulated. A scatter plot and expression diagram of relative fold change showed the dysregulation patterns of miRNAs in a separate phase of GDM and control groups (Figs. 1 and 2). Implementation of screening tests such as miRNAs evaluation during early pregnancy, offers an opportunity to identify women at risk of GDM and to evaluate intervention strategies on pregnancy outcome and the long-term health of both mother and baby. Therefore, we focused more on the results from the first trimester. Dysregulated microRNAs in the first trimester are presented in Table 3.

ROC curve analysis. The diagnostic evaluation of ROC curve based on distribution of ΔCt of dysregulated microRNAs in the 1st trimester in GDM patients revealed that the area under the curve (AUC) for hsa-miR-193a, hsa-miR-21, hsa-miR-23a, hsa-miR-361 and hsa-miR-130a were 0.8906 ± 0.04470 (P < 0.0001, 95% CI = 0.8–0.9), 0.8950 ± 0.04411 (P < 0.0001, 95% CI = 0.8–0.9), 0.6337 ± 0.08262 (P = 0.1124, 95% CI = 0.4–0.7), 0.6510 ± 0.08722 (P = 0.07, 95% CI = 0.4–0.8) and 0.7222 ± 0.07450 (P = 0.0083, 95% CI = 0.5–0.8) respectively. The discriminatory capability (P < 0.05) of miR-193a, miR-21 and miR-130a had the highest area under the curve (AUC) (Fig. 3A–E). The data indicated that three microRNAs namely, hsa-miR-193a (Sensitivity of 91% and Specificity of 70%, P = 0.0001), hsa-miR-21 (Sensitivity of 83% and Specificity of 79%, P = 0.0001) and hsa-miR-130a (sensitivity of 79% and Specificity 54%, P = 0.0025) fit the biomarker role and have a potential diagnosis value for GDM detection in the first trimester of pregnancy. Further analysis revealed a decreasing AUC to 0.84 ± 0.021 when were incorporated into a 3-miRNA model (hsa-miR-193a, hsa-miR-21, and hsa-miR-130a significance (Fig. 3F). Furthermore, based on regression analysis, a significant correlation had been revealed between miR-21 and miR-130a (r = 0.3921, P = 0.05) whereas here was not observed significant association between miR-21 and miR-13a (r = 0.3146, P = 0.1279) (Fig. 3G).

## Table 1. Demographics and clinical data of the study population. Data presented as mean ± SD and differences in demographical data were evaluated using ANOVA and T-test. P<0.05 = significant.

| Characteristics | GDM (24) | Control (24) | P-Value |
|-----------------|----------|--------------|---------|
| Maternal age/Years | 31.42 ± 4.836 | 30.58 ± 3.844 | 0.4971 |
| Weight | 62.92 ± 14 | 49.36 ± 8.047 | 0.0005 |
| BMI (kg/m2) | 27.63 ± 4.388 | 21.11 ± 3.086 | 0.0001 |
| FPG | 5.375 ± 2.144 | 4.208 ± 0.2701 | 0.011 |
| 2-h glucose mmol/l | 7.5 (6–8.5) | 6.0 (5.1–6.4) | <0.0001 |
| Family history (diabetes) | 20 (maternal) | 3 | 0.0001 |
| Birth weight/kg | 3.270 (3100–3.5) | 3.370 (3.150–3.700) | 0.167 |
In-silico target analysis. In silico analysis revealed the interaction between miRNAs and their target genes involved in the first trimester of GDM. In this analysis, KLF, ZNF25, AFF4, SRSF2, C1orf143 and ZNF655 were prominent genes targeted by the common nodes miR23, miR130a, miR193, miR21, and miR361 (Fig. 4).

Discussion
GDM is an increasingly common condition and can cause serious and lifelong harmful complications for both mother and child. There is great interest in the potential role of miRNAs as regulators of biological processes, mediators of tissue crosstalk, and biomarkers of GDM12. MiRNAs participate in various mechanisms associated with pregnancy and GDM. In addition, in maternal blood, circulating miRNAs are persistent and detectable. As a result, they are potential biomarker candidates for non-invasive pregnancy problems diagnostic tests. Studies have shown that dysregulated miRNAs are associated with pregnancy complications, suggesting the potential use as prognostic markers for GDM disease13,14. Furthermore, previous studies have reported that serum or plasma miRNAs are differentially expressed between GDM patients and controls.

In this study, we attempted to identify the differentially expressed serum-derived miRNAs and the molecular interactions of the in-silico miR target genes that may be involved in GDM. Furthermore, the potential characteristics of dysregulated miRNAs were investigated. GDM is diagnosed at the end of the second or beginning of the third trimester3, leaving no time to prevent pathological changes that could occur during the first and second trimester. Thus, miRNAs detection as early as in the first trimester is crucial and was our main study objective. Our results showed that miR130a is significantly downregulated in the GDM sample, while miR193a, miR21, miR23a and miR361 were significantly upregulated in the first trimester.

One of the prominent and thought-provoking results of the current research is the biological behaviour of miR-130a, which has decreased expression in the first, second, and third trimesters of pregnancy while there were no changes in expression level in the postpartum period (Fig. 2). MiR130a affects a variety of cellular processes, from inhibition of glucose uptake, mitochondrial function, oxidative stress to fetal development15,16. In general, this microRNA has a dual role in biological processes. Due to its cellular functions, miR-130a expression is dysregulated in a variety of pathologies, including oral squamous cell carcinoma17, ovarian epithelial cell carcinoma, thyroid eye disease18, cervical cancer19, and acute myeloid leukemia20. It was also shown that upregulated miR-130a reduces intracellular ATP levels in the pancreatic beta cell21. Furthermore, previous studies found upregulation of circulating miR-130a has a crucial role in diabetes-related complications22. In a study by Meng et al., decreased miR-130a in endothelial progenitor cells from diabetes mellitus was reported to contribute to impaired EPC (Endothelial progenitor cell dysfunction) function via its target RunX323.

MiR23a identified from this study, has been proposed as a potential biomarker for early diagnosis of prediabetes and type 2 diabetes and is particularly useful in distinguishing between undiagnosed and prediabetes24. In a research study that conducted by Liron Yoffe et al25, miR-23a was in upregulated pattern that the ROC value (AUC = 0.89 and accuracy = 0.90) revealed its biomarker characteristics in Spain and Italy countries patients with GDM (Liron Yoffe et al. 2019). Furthermore, our finding was in agreement with Yang et al. in which we found that miR-23a could function as a potential biomarker in the first trimester of GDM. On the other hand, upregulated miR193a has been shown to play a vital role in the pathogenesis of placenta accreta spectrum development mediated by the target in EFNB2 gene via the EMT signaling pathway26. In addition to that, elevated miR193a was reported as a potentially new biomarker for the diagnosis of diabetic nephropathy27. Although miR193a was

| Dysregulated miRNAs | Trimesters | 1st Fold change | 2nd Fold change | 3rd Fold change | Post-partum Fold change |
|---------------------|-----------|----------------|----------------|----------------|------------------------|
| miR-130a            | −0.32     | 0.006          | −0.43          | 0.004          | −0.34                  | 0.007                  |
| miR-193a            | 3.1       | 0.03           |                |                |                        |                       |
| miR-21              | 7.4       | 0.007          |                |                |                        |                       |
| miR-23a             | 2.14      | 0.01           |                |                |                        |                       |
| miR-361             | 2.9       | 0.008          |                |                | 1.77                   | 0.001                  |
| miR-let7i           | 0.71      | 0.008          |                |                |                        |                       |
| miR-125             | −0.047    | 0.008          | −0.42          | 0.008          | −0.21                  | 0.01                   |
| miR-126             | 0.55      | 0.04           |                |                |                        |                       |
| miR-129-2           | −0.42     | 0.021          |                |                |                        |                       |
| miR-129             | 0.62      | 0.01           | −0.29          | 0.002          |                        |                       |
| miR-34              | −0.22     | 0.003          |                |                | −0.31                  | 0.021                  |
| miR-375             | −0.48     | 0.009          |                |                | −0.31                  | 0.021                  |
| miR-let7e           | 0.59      | 0.004          |                |                |                        |                       |
| miR-107             | 0.57      | 0.002          |                |                |                        |                       |
| miR-370             | 0.61      | 0.004          | −0.33          | 0.043          |                        |                       |
| miR-194             | 13.27     | 0.02           |                |                |                        |                       |
| miR-24              | 4.85      | 0.04           |                |                |                        |                       |

Table 2. Dysregulated microRNAs in each trimester and post-partum period of GDM.
Literatures proven that miR21 has a significant biological route in a variety of diseases, although, as a molecular diagnostic marker can therapeutically regulate type 2 diabetes and pancreatic cancer. However, it is still unclear how miR21 is linked to GDM. One plausible explanation could be that miR21 promotes glucose uptake through induction of the PPARα gene in GDM patients and inhibits cell proliferation and infiltration. Gestational-adjusted expression of miR-21 has been reported to be positively associated with GDM. Sexually dimorphic miRNA expression during pregnancy in the human placenta was reported by Amy E. Flowers et al. They found that miR361 was differentially expressed and upregulated in women of the 1st and 3rd trimesters. One of the significant points in this research is the results of the binary logistic regression and correlation analysis. BLR shows that the accumulation of the effect of three microRNAs causes a significant decrease in their biomarker power, which can be due to the difference in the expression value of microRNA genes in our study population. In addition, the results of the correlation analysis among three microRNAs signature showed a significant relationship between the expression of miR-21 and miR-130a only, so that the increase of miR-21 is associated with the decrease of miR-130a in GDM patients in the first trimesters of pregnancy, and based on our knowledge, these findings are for the first report in the Malay population.

Overall, our study data investigated differentially expressed microRNAs in GDM patients and we showed the potential role of hsa-miR-193a, hsa-miR-21 and hsa-miR-130a as biomarker but the role of these dysregulated microRNAs in the pathogenesis of GDM is still not clearly understood.

**Figure 1.** Scatter plot analysis of the relative expression of thirty-seven miRNAs compared between control and GDM patients. (A) = first trimester, (B) = second trimester (C) = third trimester (D) = post-partum. Color scheme: red color indicates upregulated miRNAs, while blue indicates downregulated miRNAs. The miRNAs that are significantly altered between the two groups (control and GDM) are located above. The diagonal black line corresponds to \( P < 0.05 \). Analysis was performed using online statistical analysis by Qiagen.
This research has several limitations. Since miRNA profiling experiments are relatively expensive, they are frequently conducted on a small number of research participants. Furthermore, there are very few miRNAs profiling studies in GDM, especially for longitudinal research. Only four published reports on miRNA profiling in GDM were discovered during our initial search. However, three were only done during the first trimester, while a fourth was done throughout both the first and second trimesters. Data on miRNA expression in different trimesters are therefore hard to come by. This work is limited by the absence of follow-up on the functional aspects of the miRNA findings. In addition, we did not compare the circulating miRNAs with the placental miRNAs. This is due to the religious belief that disapproved tissue sampling. Future research must investigate the functional role of these miRNAs and their correlation with GDM parameters. In addition, it is essential to investigate the placenta miRNAs for comparative study.

The routine use of miRNAs as screening tools has enormous promise for assisting in the earlier detection and treatment of GDM by dietary changes or pharmacological intervention. Their clinical usefulness in different diseases has been shown in an increasing number of research. MiRNA profiling during GDM is currently inconclusive, partly because of the scarcity of data and the lack of consistency among research findings. Before circulating miRNAs are used in therapeutic settings, there are numerous analytical and pre-analytical hurdles that must be overcome. Pre- and post-analytical procedures should be standardized to improve reproducibility between experiments. Large prospective cohort studies should also be conducted to examine the factors influencing miRNA expression and determine whether they may be candidates for diagnostic or prognostic information.

Conclusions
In summary, this study identified three miRNAs that are differently expressed in GDM women and potentially serve as biomarkers. It is hypothesized that these microRNAs regulate genes including ZNF25, KLF, and SRSF2. Our findings emphasize the significance of these microRNAs, in particular their role in the first trimester of GDM.
| miR-130a | SRSF2, CLIP1, GJA1, CPEB1, SLAIN1, SKD1A1, ESR1, ACVR1, TSC1, RPN6K5A, MDM4, KLF, IGF1, RAP2C, ACSL4, PIK3CB, ZBTB30, MYB1, DDX6, FAM155A, KRTBD8, ZBTB18, ZFYVE9, TBL1XR1, LGALS3, ATG16L1, R060, CHST1, MEC2P, FMR1, CNOT6, SOS2, DYNC1L2, SEC5BP2, LAMPA, Runx3 | Inhibition of glucose uptake Impaired mitochondrial function and oxidative stress which affects fetal development | Decreased/−3.12 | 0.006548 |
| miR-193a | SRSF2, MAPPK10, PIGA, DCAF7, RAPGEF6, KLF, AFF4, EFRN2 | Promote trophoblast migration and invasion | Increase/3.1 | 0.032241 |
| miR-21 | C1orf143, ZNF25, YYD1, PRDM11, FASLG, ZNF367, VCL, SKF2, TGFBLP2P2 | Down regulated of miR-21 Inhibit Cell Proliferation and Infiltration, increased miR-21 is associated with pregnancy complications | Increase/7.4 | 0.007121 |
| miR-23a | ZNF25, ZNF99, SEMA6D, FAM234B, TRIL, INTU, ZNF138, AUN, TAB3, SEC23PI, ZBTB43, PDE7A, PPARCGLA, PEDRA, TOPI, KLF, TED51, FUT9, SESN3, DNAJC6, PPM1K, VGCL3, SFT2D1, ZNF716, C2orf69, ATP11C, NEK6, LPR, ARHGAP20, MRC1, ETNK1, WBP2, FAM126B, TMPO, PPRPAR, HEXIM1, CNT3, PRK4, PTEN, TNC2, SLCA44, MUC3, RAB8B, CAC1, PPE, ZNF117, RPS2, CNSK1G3, ZFH4X, SLCA1, MAP4K4, ROBO2, PRTG, NFIP1, RPR22, CTSC, SCG5, SEC23A, CDC40, BORA, CNOT6L, NCOA2, NUP506, NACC2, ZNF665 | Potential biomarkers for GDM in the first trimester | Increase/2.14 | 0.018739 |
| miR-361 | PIK3CG, TEAP2B, ZMAT3, RANBP17, AFF4, ZNF655, C1orf143 | Ultimately is a sex specific early markers of extremely low gestational age | Increase/2.9 | 0.000063 |

Table 3. MiRNAs role and their putative and predictive target genes in the first trimester of GDM.

Methods

Participant characteristics. A case–control study was conducted on a total of 1122 women (267 GDMs and 855 controls) who had spontaneous deliveries at the University of Malaya Medical Center (UMMC) from April 2014 to June 2016. All participants were characterized by pregnancy, were nonsmokers, and did not abuse alcohol. The selection criteria for the study were the age of the mother between the ages of 18 and 45 and the diagnosis of gestational diabetes by a trained doctor. The age of mothers between the ages of 18 and 45 with normal pregnancies served as a control. Abnormal fetal, still giving birth, sickle cell anemia, thalassemia or other hemoglobinosis, and other pregnancies such as lupus, hypertension, thyroid disease, cardiovascular disease, transplantation, kidney disease, asthma or other serious disease Women diagnosed with previous conditions, drug abuse, a history of smoking and depression, and carriers of blood-borne infections were excluded. Screening was performed between the 24th and 28th weeks of gestation using the Modified Oral Glucose Tolerance Test (mOGTT). Universal screening includes pregnant women with a BMI greater than 27 kg/m², previous giants weighing 4 kg or more, previous GDM, one-time relatives with diabetes, a history of unexpected prenatatal fetal death, and birth defects. It was performed on pregnant women with a medical history Positive. GDM is defined as fasting plasma glucose (FPG) (>5.1) and 75 g mOGTT plasma glucose (≥7.8). Substantial risk women with normal initial screening results were subjected to a repeat mOGTT at 4–6 weeks later. The GDM screening followed the National Obstetric Registry (NOR) guideline. The control consisted of uncomplicated pregnant mothers who gave birth to a baby between 38 and 41 weeks. Trained personnel assessed the physical health of the mother and gestational age was calculated from the first day of the last menstrual cycle or from the patient’s early ultrasound scan results. All study protocols followed the medical ethics from the Universiti Malaya Medical Centre (Ethics Committee/IRB Reference Number: 982.3), and informed consent was obtained from all participants.

Serum RNAs extraction and cDNA synthesis. Twenty-four women with GDM were included from the pool randomly. Maternal samples were collected within first, second, third trimesters and 2–6 months postpartum. Twenty-four healthy pregnant women served as controls, taken randomly from the pool. The sample size was estimated based on publication by Kok et al. Total RNAs were extracted from serum samples using the miRNeasy serum/plasma kit and synthesized into cDNA using the miScript II Rt kit according to the manufacturer’s protocol (Qiagen, Mississauga, Ontario, Canada).

MiRNAs expression profiling. A pathway-focused miScript miRNA PCR Array Human Diabetes (Qiagen, Mississauga, Ontario, Canada) was used in combination with miScript SYBR Green PCR kit (Qiagen) to profile miRNA expression in a 96-well plate using a Step One Plus™ real time PCR detection system (Applied Biosystem, California, USA) following the cycling conditions recommended by the manufacturer’s instructions. Amplification conditions were 15 min at 95 °C, followed by 40 cycles of 15 s at 94 °C, 30 s at 55 °C, and 30 s at
Figure 3. ROC curve analysis. The figure shows the comparison of five dysregulated microRNAs and their specific AUC based on their ΔCt distribution in the first trimester of pregnancy (A–E). Cumulative Graf of ROC analysis of three combined microRNAs and Binary logistic regression (BLR) analysis (E and F). The ROC curve is derived from Graf Pad Prism version 8. Normality distribution of samples was performed using Shapiro–Wilk test. P-value calculated using non-parametric analytical test.
70 °C. A total of 37 miRNAs involved in GDM phenotype were selected for the PCR array. Six snoRNA/snRNA, including SNORD61, SNORD68, SNORD72, SNORD95, and SNORD96A were selected as housekeeping genes. Furthermore, miRTC and PPC genes served as reverse transcription control and primer assay positive control, respectively. PCR array reactions were conducted in triplicate repeats. The data were analyzed using online Gene Globe data analysis software version 2.1.0 (https://geneglobe.qiagen.com/). The validity and accuracy of array PCR were confirmed using reverse transcription-qPCR for five randomly selected miRNAs in triplicate.

**ROC curve analysis.** A receiver operating characteristic (ROC) which is a plot of the true positive rate (Sensitivity) in function of the false positive rate (100-Specificity) for different cut-off points of a parameter was performed to evaluate the potential microRNAs biomarker characteristics.

**Insilico target genes Identification and bioinformatic analysis.** Target Scan Human Release 8.0 and miRDB were used to obtain predicted or previously experimentally validated miRNA target genes. Interaction network analysis was performed using the Cytoscape 3.7.1 tool to verify all potential interactions between the identified target gene and potential functional mediators.

**Data availability**
The datasets generated during and/or analyzed are available from the corresponding author on reasonable request.

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**Figure 4.** *In-silico* analysis of dysregulated microRNAs in the first trimester. Green and purple colors indicate dysregulated microRNAs and their common target genes, respectively. ZNF25, RANBP17, CLORF43, ZNF655, AFF4, KLF and SRSF2 were targeted in a common node of microRNAs. Interaction pathway provided by Cytoscape tool version 3.7.1.
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Author contributions
S.J. reviewed the results and drafted the manuscript. S.M.Z., Z.M. designed the study plan. S.Z.O. recruited the patients and provided constructive protocol review. S.J., H.K., Y.F.P. participated in the total RNA isolation, miRNA profiling and qRT-PCR analyses. S.J., R.V. performed the data analyses and prepared the figures. S.M.Z., S.Z.O. reviewed and conducted critical revision of the manuscript for important intellectual content. All authors read and approved the final manuscript.

Competing interests
The authors declare no competing interests.

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