Selective Vulnerability of Nerve Demyelination in Chronic Acquired Multifocal Polyneuropathy

Xiajun Zhou¹, Ying Zhu², Desheng Zhu¹, Lu Han¹, Qianyun Liu¹, Yong Hao¹, Zezhi Li¹ and Yangtai Guan¹*

¹Department of Neurology, China
²Shanghai International Medical Center, Department of Neurology, China

*Corresponding author: Yangtai Guan, Department of Neurology, Shanghai, China

ARTICLE INFO

Introduction: Multifocal motor neuropathy (MMN), Lewis-Sumner syndrome (LSS) and many of chronic inflammatory demyelinating polyradiculoneuropathy (CIDP) are representatives of acquired multifocal polynyepathy and characterized by conduction block (CB).

Objective: This retrospective study aimed to investigate the selective vulnerability and the demyelinating pattern of MMN, LSS and CIDP with CB (CIDP-CB) nerves.

Methods: Fifteen LSS subjects (107 nerves), 24 MMN subjects (176 nerves) and 17 CIDP-CB subjects (110 nerves) were included. Their clinical information, blood and cerebrospinal fluid tests were collected, and nerve conduction were performed on median, ulnar, radial, peroneal and tibial nerves. CB, temporal dispersion (TD), distal motor latency (DML), F-wave latency were recorded and nerve conduction velocity (NCV), terminal latency index (TLI) and modified F-wave ratio (MFR) were calculated.

Results: CB was more likely to occur around elbow (p<0.01) while less likely to be found between wrist and elbow in CIDP-CB (p<0.05). Tibial nerve CB was also frequently observed in MMN (p<0.05). CIDP-CB had a prolonged DML in all nerves and its slow motor nerve velocity (MCV) of upper limb was remarkable when CB nerves were excluded.

Conclusion: We reported the difference of selective segmental vulnerability of ulnar and tibial nerves in LSS, MMN and CIDP-CB. The different CB distribution could help in differentiation.

Abbreviations: ADQ: Abductor Digiti Quinti; AH: Abductor Hallucis; ANOVA: One-way Analysis of Variance APB: Abductor Pollicis Brevis; CB: Conduction Block; CIDP: Chronic Inflammatory Demyelinating Polyradiculoneuropathy; CMAP: Compound Muscle Action Potential; CSF: Cerebrospinal Fluid; DML: Distal Motor Latency; EDB: Extensor Digitorum Brevis; EFNS: European Federation of Neurological Societies; EIP: Extensor Indicis Proprius; LSS: Lewis-Sumner Syndrome; MCV: Motor Conduction Velocity; MFR: Modified F-wave Ratio; MMN: Multifocal Motor Neuropathy; NCV: Nerve Conduction Velocity; PML: Proximal Motor Latency; PNS: Peripheral Nerve Society; SD: Standard Deviation; TD: Temporal Dispersion; TLI: Terminal Latency Index

Introduction

The pathological features of peripheral nerve myelin lesion are known as diffuse and focal demyelination. As a hallmark of peripheral nerve focal demyelination, conduction block (CB) has been found in a series of both acquired and hereditary demyelinating neuropathies [1-4]. Multifocal motor neuropathy (MMN) and Lewis-Sumner syndrome (LSS) are the most representative chronic acquired focal neuropathies, while many of the classic chronic inflammatory demyelinating polyradiculoneuropathy

| Received:  June 06, 2019 | Published: June 13, 2019 |

Citation: Xiajun Zhou, Ying Zhu, Desheng Zhu, Lu Han, Qianyun Liu, Yong Hao, Zezhi Li, Yangtai Guan. Selective Vulnerability of Nerve Demyelination in Chronic Acquired Multifocal Polyneuropathy. Biomed J Sci & Tech Res 18(5)-2019. BJSTR. MS.ID.003213. Keywords: Multifocal Motor Neuropathy; Lewis-Sumner Syndrome; Chronic Inflammatory Demyelinating Neuropathy; Conduction Block; Demyelination
(CIDP) also presents with CB [5]. Distinct from common peripheral neuropathies, the motor impairments in these focal neuropathies are clinically uneven, i.e., non-length dependent and asymmetric [6,7]. Their limb onset selectivity is well-known [8], however, not efficient in reciprocal differentiation. Different nerve involvement among demyelinating polyneuropathies was proposed by the morphological study, which suggested the selective vulnerability of nerves [9]. The selectivity was observed in other neuropathies [10,11], but hasn’t been fully studied in focal demyelination by electrophysiology methods. Thus, in this study, by using of a bunch of well-established electrophysiological indicators, e.g., distal motor latency (DML), terminal latency index (TLI), F-wave latency and nerve conduction velocity (NCV) as well as CB, we aimed to investigate the selective vulnerability and the demyelinating pattern of nerves and segments in patients with MMN, LSS and CIDP with CB (CIDP-CB).

**Materials and Methods**

**Subjects**

This was a retrospective cross-section study performed under the principles of the Declaration of Helsinki and was approved by the Institutional Review Board of Renji Hospital Shanghai Jiaotong University School of Medicine. Written informed consent was obtained from the subjects for publication of this paper. Fifteen LSS subjects, 24 MMN subjects and 17 CIDP-CB subjects admitted at Renji Hospital Shanghai Jiaotong University School of Medicine from 2013 to 2017 were included in the study. A total of 107 nerves of LSS subjects, 176 nerves of MMN subjects, and 110 nerves of CIDP-CB subjects were analyzed. The LSS and CIDP diagnoses were based on the 2010 European Federation of Neurological Societies (EFNS)/Peripheral Nerve Society (PNS) guideline on CIDP [12], and the MMN diagnose was based on the 2010 EFNS/PNS guideline on MMN [13]. The exclusion criteria were peripheral neuropathy family history; alcohol abuse; toxic and neurotoxic exposure; tumor; metabolic disorders including pathoglobulinaemia (diabetic mellitus and impaired glucose tolerance); paraproteinemia (monoclonal gamma-globin related polyneuropathy).

**Clinical Profiles and Examinations**

General information of gender, age and height were recorded. Detailed clinical history was investigated to collect disease duration and onset and determine neurological manifestations of weakness, numbness and atrophy. Meticulous physical examinations were performed to determine tendon reflex as well as other positive neurological signs. Blood routine test, liver and kidney function, blood electrolytes, thyroid function, vitamin B₁₂, folic acid and blood glucose were checked to meet the inclusion and exclusion criteria. Gangliosidosisis antibodies examination were semi-quantitative by blot and was fulfilled by KingMed Diagnostics (Guangdong, China). A panel of various kinds of anti-Gangliosidosis (GM1, GM2, GM3, GD1a, GD1b, GQ1b, GD1b) was used and positive result was determined by visible band in GM1 area. Blood immunofixation electrophoresis was tested for subjects to meet the exclusion terms. Lumbar puncture was performed under the informed consent of subjects and cerebrospinal fluid (CSF) was quantitatively analyzed to determine albumin protein concentration.

**Nerve Conduction**

All subjects received nerve conduction test by Keypoint. net (Natus, CA, USA) in a quiet room with temperature >20°C. Surface electrodes were used to record wave form on fully relaxed muscle after stimulation. Median, ulnar, radial, peroneal, peroneal superficial, tibial and sural nerve were selected for evaluation (sensory nerve data were notshown). Multiple sites were stimulated for every nerve to obtain sufficient data and calculate velocities. For median nerve, recording was placed on abductor pollicis brevis (APB), and stimulation was conducted on wrist and elbow. For ulnar nerve recording was placed on abductor digiti quinti (ADQ), and stimulation was conducted on wrist, below elbow, above elbow and Erb’s point. For radial nerve recording was placed on extensor indicis proprius (EIP), and stimulation was conducted on forearm, below elbow and upper arm. For peroneal nerve, recording was placed on extensor digitorum brevis (EDB), and stimulation was conducted on ankle, below fibular head and above fibular head. For tibial nerve, recording was placed on abductor hallucis (AH), and stimulation was conducted on ankle and popliteal fossa. Normal values and ranges of our lab were adopted. Negative peak amplitude, curve area and duration were recorded to determine CB and temporal dispersion (TD). A definite CB was defined as area reduction on proximal vs. distal stimulation of at least 50%, distal compound muscle action potential (CMAP) must be >20% of the lower limit of normal and >1 mV, and increase of proximal to distal CMAP duration must be ≤30%.13 Nerve variant especially Martin-Gruber anastomosis was carefully excluded by additional stimulation. Temporal dispersion was defined as >30% duration increase between the proximal and distal CMAP.

Take-off latency and distance were recorded and NCV was thus calculated. Distal distance (dD), DML, and NCV was used to calculate TLI by the following formula: TLI=(dD/DML)/NCV. F-wave was performed during a bout of stimulation recorded on distal muscle and the first F-wave latency (F) was recorded. DML, proximal motor latency (PML) which is the latency recorded from the second stimulation site from distal, and F were used to calculate modified F-wave ratio (MFR) by the formula:

\[
MFR = \left( \frac{F + DML - 2 \times PML - 1}{2 \times (PML - DML)} \right)
\]

**Statistical Analysis**

Median, ulnar, radial, peroneal and tibial nerves were selected for studying CB and TD distributions. Median, ulnar, peroneal and tibial nerves were selected for comparing other conduction parameters. Normal distribution data were described as mean (x) ± standard deviation (SD), and skewed distribution data were described as median (25% percentile – 75% percentile).
Particularly, data of the same type were kept the same description in the tables for comparability. Pearson and Fisher chi-square tests were used to analyze the proportion and composition of different groups. Intragroup composition was analyzed by binomial exact test. One-way analysis of variance (ANOVA) and group t-test were used for data with normal distribution and standard homogeneity of variance, otherwise Kruskal Wallis test and Wilcoxon Ranksum test were chosen instead. All statistics were performed by Stata 15.1 (StataCorp, TX, USA) and p<0.05 indicated significant differences.

**Results**

**General Information of Subjects**

Gender, age and height were commensurate among different groups (p>0.05). Intragroup gender composition was significant in LSS group (p=0.035) and in MMN group (p=0.007), but not significant in CIDP-CB group (p=0.332). MMN group showed a longer disease duration compared to other groups (p<0.05). Pure upper limb onset and sensory involvement were less probable in CIDP-CB group compared to MMN group (p<0.05). There was no significant difference found in other clinical symptoms or signs among groups (p>0.05). CSF albumin protein showed a gradient elevation from MMN to LSS to CIDP-CB (p<0.001). Anti-GM1 was most likely to be detected in MMN group (p<0.001). General and clinical comparisons are shown in Table 1.

**CB And TD Distribution of Nerves**

CB was more likely to occur around elbow (p<0.01) while less likely to be found between wrist and elbow in CIDP-CB (p<0.05). CB was found in more than half of tibial nerve examined in MMN and it was significant (p<0.01). No significant difference of CB occurrence was found in other sites. TD was found in 7 of 107 nerves in LSS, 12 of 176 nerves in MMN and 8 of 110 nerves in CIDP-CB. The comparison was statistically insignificant (p=0.977). CB distribution is shown in Table 2.

**Table 1:** General and clinical profiles of LSS, MMN and CIDP-CB subjects.

| Characteristic                      | LSS (n=15) | MMN (n=24) | CIDP-CB (n=17) | p Value |
|-------------------------------------|------------|------------|----------------|---------|
| Gender                              |            |            |                | 0.501   |
| Female; s (%)                       | 3 (20.0)   | 5 (20.8)   | 6 (35.3)       |         |
| Male; n (%)                         | 12 (80.0)  | 19 (79.2)  | 11 (64.7)      |         |
| Age (years); ±SD                    | 44.2±15.1  | 42.7±12.5  | 51.6±16.4      | 0.142   |
| Height (cm); ±SD                    | 169.0±7.3  | 168.0±5.7  | 164.6±8.7      | 0.183   |
| Duration (months)*,**; median (P25-P75) | 10 (3-12)  | 24 (12-39) | 6 (5-12)       | 0.033   |
| Onset                               |            |            |                |         |
| Upper Limbs***; n (%)               | 10 (66.7)  | 16 (66.7)  | 5 (29.4)       | 0.036   |
| Lower Limbs; n (%)                  | 2 (13.3)   | 2 (8.3)    | 3 (17.6)       | 0.693   |
| Upper and Lower Limbs; n (%)        | 3 (20.0)   | 6 (25.0)   | 9 (52.9)       | 0.084   |
| Numbness***; n (%)                  | 8 (53.3)   | 6 (25.0)   | 12 (70.6)      | 0.013   |
| Weakness; n (%)                     | 12 (80.0)  | 20 (83.3)  | 14 (82.3)      | 0.174   |
| Atrophy; n (%)                      | 5 (33.3)   | 8 (33.3)   | 1 (58.5)       | 0.093   |
| Hyopreflexia; n (%)                 | 15 (100)   | 24 (100)   | 17 (100)       | NA      |
| CSF Albumin Protein (mg/dl)*,**; **; ±SD | 629.9±184.0 | 398.5±171.6 | 995.2±416.9    | <0.001  |
| Positive anti-GM1***; n (%)         | 2 (13.3)   | 18 (75)    | 2 (11.8)       | <0.001  |

Note:
CIDP-CB, Chronic inflammatory demyelinating polyneuropathy with conduction block; CSF, Cerebrospinal Fluid; GM1, Monosialotetrahexosyl ganglioside; LSS, Lewis-Sumner syndrome; MMN, Multifocal motor neuropathy; NA, Not applicable.

*p<0.05 between LSS subjects and MMN subjects; ** p<0.05 between LSS subjects and CIDP-CB subjects; *** p<0.05 between MMN subjects and CIDP-CB subjects.
**Table 2:** Nerve distribution of CB.

| Nerve and Position          | LSS     | MMN     | CIDP-CB    | p Value |
|-----------------------------|---------|---------|------------|---------|
|                             | No. of Nerves with CB / n (%) | No. of Nerves with CB / n (%) | No. of Nerves with CB / n (%) |         |
| Median                      | 15 / 29 (51.7) | 27 / 44 (61.4) | 12 / 30 (40.0) | 0.195   |
| Wrist - Elbow               | 11 / 28 (39.3) | 10 / 44 (22.7) | 3 / 28 (10.7)  | 0.042   |
| Ulnar                       | 6 / 28 (21.4)  | 3 / 44 (6.8)   | 22 / 28 (78.6) | 0.003   |
| Wrist - below Elbow**       | 9 / 10 (90.0)  | 20 / 33 (60.6) | 10 / 14 (71.4) | 0.677   |
| below Elbow - above Elbow***| 6 / 28 (21.4)  | 3 / 44 (6.8)   | 22 / 28 (78.6) | 0.003   |
| above Elbow - Erb’s point   | 6 / 28 (21.4)  | 3 / 44 (6.8)   | 22 / 28 (78.6) | 0.003   |
| Radial                      | 2 / 7 (28.6)   | 2 / 12 (16.7)  | 2 / 2 (100.0)  | 0.081   |
| Forearm - above Elbow       | 2 / 5 (40.0)   | 3 / 11 (27.3)  | 0 / 0 (0.0)    | 1.000   |
| above Elbow - upper Arm     | 5 / 23 (21.7)  | 9 / 38 (23.7)  | 6 / 25 (24.0)  | 0.980   |
| Peroneal                    | 3 / 28 (21.4)  | 3 / 44 (6.8)   | 22 / 28 (78.6) | 0.003   |
| Ankle - below Fibular head  | 4 / 20 (20.0)  | 20 / 38 (52.6) | 4 / 25 (16.0)  | 0.005   |

Note:

CB, Conduction Block; CIDP-CB, Chronic inflammatory demyelinating polyneuropathy with conduction block; LSS, Lewis-Sumner syndrome; MMN, Multifocal motor neuropathy.

* p<0.05 between LSS nerves and MMN nerves; ** p<0.05 between LSS nerves and CIDP-CB nerves; *** p<0.05 between MMN nerves and CIDP-CB nerves.

**Conduction Indicators of Nerves**

DML was significantly prolonged in CIDP-CB in all nerves (p<0.05), while only ulnar nerve TLI prolonging was observed in LSS (p<0.05). No F-wave latency or MFR difference was found in any nerve. Ulnar nerve NCV differences were found among groups (p<0.05). When nerves with CB were excluded, decrease of upper limb nerve NCV was statically significant (p<0.05). Lower limb CMAP was lower in CIDP-CB (p=0.01). Electrophysiological findings of the indicators are shown in Table 3.

**Table 3:** Electrophysiological findings of peripheral nerve demyelination among LSS, MMN and CIDP-CB nerves.

| Indicator and Nerve | LSS     | MMN     | CIDP-CB   | p Value |
|---------------------|---------|---------|-----------|---------|
| DML (ms)            |         |         |           |         |
| Median*****          | 3.4 (3.15-3.8) | 3.67 (3.2-4.24) | 4.75 (3.5-5.4) | 0.006   |
| Ulnar*****           | 2.71 (2.5-3.07) | 2.75 (2.35-3.11) | 3.87 (2.62-4.38) | 0.004   |
| Peroneal***          | 4.3 (3.7-5.17) | 4.75 (4.3-5.5) | 5.9 (4.6-7.1) | 0.004   |
| Tibial***            | 4.31 (3.86-5.19) | 4.4 (3.65-5.3) | 5.65 (4.51-6.70) | 0.047   |
| TLI                  | 0.33 (0.3-0.4) | 0.34 (0.26-0.39) | 0.29 (0.25-0.39) | 0.334   |
| Ulnar***             | 0.40 (0.36-0.51) | 0.35 (0.30-0.41) | 0.36 (0.29-0.42) | 0.038   |
| Peroneal             | 0.41 (0.33-0.46) | 0.44 (0.32-0.52) | 0.36 (0.28-0.47) | 0.267   |
| Tibial               | 0.42 (0.36-0.50) | 0.43 (0.37-0.51) | 0.35 (0.29-0.48) | 0.172   |
| F-wave latency (ms)  |         |         |           |         |
| Median               | 29.95 (28.4-NP) | 48.7 (29.15-NP) | 39.6 (30.3-57.3) | 0.595   |
| Ulnar                | 32.35 (27.9-NP) | 33.7 (27.1-NP) | 45.25 (33.1-NP) | 0.243   |
| Peroneal             | 54.9 (48.3-NP) | 66.75 (51.5-NP) | 69.8 (58.6-NP) | 0.141   |
| Tibial               | 52.8 (49.45-67) | 51.5 (70.3-NP) | 55.15 (51.75-70.15) | 0.236   |
| MFR                  | 1.81 (1.65-NP) | 2.81 (1.87-NP) | 1.84 (1.51-2.63) | 0.050   |
| Ulnar                | 2.16 (1.76-NP) | 3.06 (2.36-NP) | 3.03 (2.32-NP) | 0.147   |
|                  | Peroneal | Tibial | MCV (m/s) | Median | Ulnar*** | Peroneal | Tibial | MCV (nerves without CB) (m/s) | Median*** | Ulnar**** | Peroneal | Tibial | Distal CMAP (mV) | Median | Ulnar | Peroneal***** | Tibial***** |
|------------------|----------|--------|-----------|---------|-----------|----------|--------|-------------------------------|-----------|-----------|----------|--------|----------------|--------|-------|---------------|------------|
| Peroneal         | 3.19 (2.21-NP) | 3.69 (2.42-NP) | 4.53 (2.88-NP) | 0.519   | 50.95 (41.4-53.8) | 42.3 (39.2-45.7) | 43.4 (39.8-45.6) | 52.4 (51.3-55.4) | 52.2 (49.1-55.1) | 42.8 (40.2-45.7) | 43.7 (42.05-45.6) | 6.3 (5.1-8.2) | 7 (4.9-7.65) | 2.8 (1.3-3.9) | 7.85 (4.9-10.1) |
| Tibial           | 1.92 (1.49-2.29) | 2.25 (1.63-NP) | 2.09 (1.63-2.82) | 0.479   | 49.65 (36.75-55.8) | 40.4 (36.5-44.15) | 43.3 (39.1-46.3) | 54.8 (53-59) | 57.25 (51.7-60.5) | 41.3 (37.4-44.5) | 46.1 (39.1-48.3) | 6.55 (3.4-9.4) | 7.65 (4.35-8.95) | 3.3 (1-4.9) | 7.3 (3.7-10.5) |

Note:
CB, Conduction block; CMAP, Compound muscle action potential; CIDP-CB, Chronic inflammatory demyelinating polyneuropathy with conduction block; DML, Distal motor latency; LSS, Lewis-Sumner syndrome; MCV, Motor conduction velocity; MFR, Modified F-wave ratio; MMN, Multifocal motor neuropathy; NP, No potential elicited; TLI, Terminal latency index. Results are show as median (P25-P75). No. of nerves: LSS, Median=29, Ulnar=28, Peroneal=23, Tibial=20; MMN, Median=44, Ulnar=44, Peroneal=38, Tibial=38; LSS, Median=30, Ulnar=28, Peroneal=25, Tibial=25.

* p<0.05 between LSS nerves and MMN nerves; ** p<0.05 between LSS nerves and CIDP-CB nerves; *** p<0.05 between MMN nerves and CIDP-CB nerves.

Discussion

MMN and LSS were characterized by male predominant, upper limb onset and different sensory involvement [14-17]. In this study we also found a male predominant composition in the MMN group and LSS group and it was different in the CIDP-CB group. High upper limb onset proportion and rare sensory involvement in MMN were also proved and could help in differentiating CIDP-CB. In addition, we found MMN presented slowly progressive long duration, low CSF protein and frequent positive anti-GM1, while LSS and CIDP presented progressive duration, moderate-high CSF protein and rare positive anti-GM1. Clinical findings were confirmatory and consistent with previous study [18,19]. Using electrophysiological indicators, we presented a horizontal comparison of LSS, MMN and CIDP-CB nerves. Selective vulnerability of nerve is particularly interesting and controversial in focal neuropathy. Ulnar nerve has been suggested useful in diagnosing acute demyelinating neuropathy [20], and tibial nerve was hypothesized to be especially involved in MMN [21], while other study found no difference in CB distribution [22]. In this study, noting that wrist to elbow segment of ulnar nerve conduction was least likely to be blocked in CIDP-CB compared to LSS, while the CB likelihood of segment around ulnar was vice versa in CIDP-CB compared to MMN. Another CB distribution characteristic we found was the frequent tibial nerve CB in MMN. Although upper limb is more frequently affected in MMN, superimposing of tibial nerve focal demyelination implies inclination of MMN diagnose. These findings suggest that ulnar and tibial CB may be critical in differentiation, and to our knowledge hasn’t been reported in chronic focal demyelinating diseases.

The mechanism of nerve selectivity of chronic focal demyelinating diseases hasn’t been fully investigated. Recently, a common etiology discovered in demyelinating diseases with CB was the detection of autoantibodies, e.g., anti-neurofascin 140/186, anti-neurofascin 155 [23,24]. As a target, the neurofascin expression in node and paranode sites determines the impairment pattern of conduction failure and is associated with CB [25]. A reasonably hypothesis is that the uneven distribution of nerve expression of neurofascin may contribute to the nerve selectivity.

Distal vulnerability was suggested in typical CIDP previously [26,27], while, CIDP-CB, as a subtype, has rarely been analyzed and compared solely. Although prolonged DML in all CIDP-CB nerves was in accordance with the distal predominant pattern, the similar values of TLI, a comparison of distal and middle segments [28], of most nerves among groups implied that the distal impairments might commensurate with middle segment when the existence of CB was taken into consideration. The CB also masked the different diffuse demyelinating of median nerve as only ulnar nerve MCV was lower in CIDP-CB. When the nerves with CB were excluded, upper...
limb MCV decrease in CIDP-CB emerged. In the conduction study, we proved that CIDP-CB showed many common features with typical CIDP comparing to MMN and LSS and this highlighted its diffused demyelinating feature in median and ulnar nerves.

**Conclusion**

We reported the difference of selective segmental vulnerability of ulnar and tibial nerves in LSS, MMN and CIDP-CB, and proved more diffuse demyelinating features in CIDP-CB which distinguished it from LSS and MMN. The different nerve demyelination distribution and pattern could help in differentiation. This study was limited by insufficient proximal and distal conduction data and small sample. A prospective cohort and immunological mechanism study are anticipated.

**References**

1. Chad DA, Hamner K, Sargent J (1986) Slow resolution of multifocal weakness and fasciculation: a reversible motor neuron syndrome. Neurology 36(9): 1260-1263.

2. Chen SD, Li ZX, Guan YT, Zhou XJ, Jiang JM, et al. (2011) A novel mutation of gap junction protein β1 gene in X-linked Charcot-Marie-Tooth disease. Muscle Nerve 43(6): 887-892.

3. Créange A, Shahrizaila N, Salhi H, Lefaucheur JP, Yuki N (2014) Association of antibodies to ganglioside complexes and conduction blocks in axonal Guillain-Barré syndrome presenting as acute motor conduction block neuropathy. J Peripher Nerv Syst 19(2): 115-120.

4. Lewis RA, Sumner AJ, Brown MJ, Ashby AK (1982) Multifocal demyelinating neuropathy with persistent conduction block. Neurology 32(9): 958-964.

5. Dimachkie MM, Barohn RJ, Katz J (2013) Multifocal motor neuropathy, multifocal acquired demyelinating sensory and motor neuropathy, and other chronic acquired demyelinating polyneuropathy variants. Neurol Clin 31(2): 533-555.

6. Whitesell J (2010) Inflammatory neuropathies. Semin Neurol 30(4): 356-364.

7. Lewis RA (2007) Neuropathies associated with conduction block. Curr Opin Neurol 20(5): 525-530.

8. Rajabally YA, Chavada G (2009) Lewis-sumner syndrome of pure upper-limb onset: diagnostic, prognostic, and therapeutic features. Muscle Nerve 39(2): 206-220.

9. Kemsnowd A, Pitarkoikil K, Gold R, Yoon MS (2015) Bochum ultrasound score allows distinction of chronic inflammatory from multifocal acquired demyelinating polyneuropathies. J Neurol Sci 348(1-2): 211-215.

10. Gwathmey KG, Burns TM, Collins MP, Dyck PJ (2014) Vasculitic neuropathies. Lancet Neurol 13(1): 67-82.

11. Takahashi S, Chum M, Kimpinski K (2017) Electrodiagnostic Characterization of Hereditary Neuropathy with Liability to Pressure Palsies. J Clin Neurosci 28(3): 119-124.

12. (2010) Joint Task Force of the EPNS and the PNS. European Federation of Neurological Societies/Peripheral Nerve Society Guideline on management of chronic inflammatory demyelinating polyradiculoneuropathy: report of a joint task force of the European Federation of Neurological Societies and the Peripheral Nerve Society—First Revision. J Peripher Nerv Syst 15(1): 1-9.

13. (2010) Joint Task Force of the EPNS and the PNS. European Federation of Neurological Societies/Peripheral Nerve Society guideline on management of multifocal motor neuropathy. Report of a joint task force of the European Federation of Neurological Societies and the Peripheral Nerve Society—first revision. Journal of the peripheral nervous system. J Peripher Nerv Syst 15(4): 295-301.

14. Rajabally YA, Chavada G (2009) Lewis-sumner syndrome of pure upper-limb onset: diagnostic, prognostic, and therapeutic features. Muscle Nerve 39(2): 206-220.

15. Katz JS, Saperstein DS (2001) Asymmetric Acquired Demyelinating Polyneuropathies: MMN and MADSM. Curr Treat Options Neurol 3(2): 119-125.

16. Saperstein DS, Amato AA, Wolfe GI, Katz JS, Nations SP, et al. (1999) Multifocal acquired demyelinating sensory and motor neuropathy: the Lewis-Summer syndrome. Muscle Nerve 22(5): 560-566.

17. Lawson VH, Arnold WD (2014) Multifocal motor neuropathy: a review of pathogenesis, diagnosis, and treatment. Neuropsychiatr Dis Treat 10: 567-576.

18. Bles Z, Blaabjerg M (2017) Cerebrospinal fluid findings in Guillain-Barré syndrome and chronic inflammatory demyelinating polyneuropathies. Handb Clin Neurol 146: 125-138.

19. Nowack DG, Teener JW (2012) Multifocal motor neuropathy. Semin Neurol 32(5): 500-505.

20. Ahdab R, Noureldine MHA, Mohammed K, Nader M, Zouari HG, et al. (2018) The ulnar ratio as a sensitive and specific marker of acute inflammatory demyelinating polyneuropathy. Clin Neurophysiol 129(8): 1699-1703.

21. Bentes C, de Carvalho M, Evangelista T, Sales-Luís ML (1999) Multifocal motor neuropathy mimicking motor neuron disease: nine cases. J Neurol Sci 169(1-2): 75-79.

22. Verschueren A, Azulay JP, Attarian S, Bocquart J, Pelli-Sliskier JF, et al. (2005) Lewis-Summer syndrome and multifocal motor neuropathy. Muscle Nerve 31(1): 86-94.

23. Delmont E, Manso C, Querol L, Cortese A, Berardinielli A, et al. (2017) Autoantibodies to nodal isoforms of neurofascin in chronic inflammatory demyelinating polyneuropathy. Brain 140(7): 1851-1858.

24. Koike H, Nishi R, Ikeu S, Kawagashira Y, Iijima M, et al. (2018) Restoration of a Conduction Block after the Long-term Treatment of CIDP with Anti-neurofascin 155 Antibodies: Follow-up of a Case over 23 Years. Intern Med 57(14): 2061-2066.

25. Rinaldi S, Bennett DLH (2014) Pathogenic mechanisms in inflammatory and paraproteinemic peripheral neuropathies. Curr Opin Neuro 27(5): 541-551.

26. Kuwabara S, Iose S, Mori M, Mitsuwa S, Sawai S, et al. (2015) Different electrophysiological profiles and treatment response in "typical" and "atypical" chronic inflammatory demyelinating polyneuropathy. J Neurol Neurosurg Psychiatry 86(10): 1054-1059.

27. Rajabally YA, Narasimhan M (2011) Distribution, clinical correlates and significance of axonal loss and demyelination in chronic inflammatory demyelinating polyneuropathy. Eur J Neurol 18(2): 293-299.

28. Vahdatpour B, Khosrawi S, Chatraei M (2016) The role of median nerve terminal latency index in the diagnosis of carpal tunnel syndrome in comparison with other electrophysiological parameters. Adv Biomed Res 5: 110.
