SN-DONOR TRIAZINE-3(2H)-THIONE CHELATING LIGAND AND TRANSITION METAL COMPLEXES: SYNTHESES, CHARACTERIZATION, In Vitro ANTIBACTERIAL EVALUATION AND ELECTROCHEMICAL BEHAVIOUR

Manoj Kumar1, *, Tanuja Kumari2, Jyoti Joshi2, Sunil Chhimpa2, P.J. John2, Raj Rajeshwar Malinda3 and Bidya S. Joshi1, *  

1, *Department of Chemistry, University of Rajasthan, Jaipur-302004, Rajasthan India.  
2Department of Chemistry, Malaviya National Institute of Technology, Jaipur-302017, Rajasthan, India.  
2Department of Zoology, Centre for Advanced Studies, University of Rajasthan, Jaipur-302004, Rajasthan, India.  
3NUS Lifesciences Private Limited, Ghaziabad, Uttar Pradesh 201102, India.  
*E-mail: kmanoj.ru@gmail.com

ABSTRACT

Triazine-based heterocycles constitute a class of pertinent heterocyclic compounds that are found ubiquitously in numerous pharmaceuticals, functional materials, and chiral catalysts or ligands. The synthesis and further physicochemical assessment of metal complexes viz. [Cu(ptt)2]2, [Ni(ptt)2]2H2O, [Zn(ptt)2]2H2O, [Cd(ptt)2]2H2O (PTTH: 5-Phenyl-1,2,4-triazine-3(2H)-thione) are delineated in this account. The prepared complexes were identified quantitatively and qualitatively by a variety of physicochemical techniques, namely elemental analysis, FT-IR, LC-MS and 1HNMR spectroscopy. Spectral data show that Cu and Ni complexes comprise a binuclear framework, in which the organic molecule simulates as a mono-functional bidentate SN- donor and with two of the four moieties of the molecule performing a bridge to coordinates the central metal ions with the sulphur atom while a mononuclear structure is obtained in the ZnII and CdII complexes. Subsequently, FT-IR and LC-MS spectra also confirmed the coordinated aqua molecules in the NiII, ZnII and CdII complexes. The ligand and its complexes exhibited considerable antimicrobial activities against the Gram-positive bacteria (Bacillus subtilis: MTCC-121) as well as Gram-negative bacteria (Escherichia coli: MTCC-40). In the present study, NiII complex showed the highest antibacterial activity with 33 mm diameter of zone of inhibition. Electrochemical reduction behavior of ligand (PTTH), CuII and NiII complexes was explored at glassy carbon electrode using cyclic voltammetry.

Keywords: Triazine, Transition Metal Complex, Spectra, Antimicrobial Activity, Inhibition Zone, Cyclic Voltammetry.
thione form in the solid and the thiol form in some solutions. In their anionic mode, both the thionate sulfur and thioamide nitrogen atom are functional to coordinate with metal centers in several manners, according to the “Pearson’s principle”, as a N,S-bidentate chelating agent or as an individual basis deprotonated bidentate anions or as a bridging agent between two metal centers. Therefore, there is indeed a tunable capability for adopting various co-ordinations mode with the panoptic range of metal ions, to give stable, intensely colored and biologically active metal derivatives. Furthermore, due to the electronic delocalization, which is strengthened on deprotonation, these ligands are immensely versatile. This fact, together with the presence of different kinds of donor atoms like N, or S, several coordination modes are possible which subjected to the metal coordination preferences; therefore, a specific ligand can demonstrate different coordination behaviour\textsuperscript{18-21}. Moreover, triazine derivatives have traditionally found utile applications in electrochemical science as multi-step oxidation-reduction systems during exploration from cyclic voltammetry\textsuperscript{16, 22-25}.

We thus, set out to characterize the chelation properties and propensity of 5-Phenyl-1,2,4-triazine-3(2H)-thione (PTTH) toward the formation of a variety of transition metal complexes over the insertion of metal ion (M\textsuperscript{II}=Ni, Cu, Zn, Cd) in a more systematic fashion. These compounds were carried out to evaluate the antibacterial activities against Gram (+) and Gram (-) Human bacteria. All newly synthesized compounds exhibited a high degree of inhibition zone towards both tested Human pathogenic microorganisms. Besides, the electrochemical properties of ligand and metal complexes have been examined in DMF at a glassy carbon working electrode employing an electrochemical analyser to monitor the structural changes along with electron transfer.

**EXPERIMENTAL**

**Materials and Methods**

Thiosemicarbazide (98\%) was purchased commercially source from Sigma- Aldrich. The starting material phenylglyoxal hydrate was prepared as standard method\textsuperscript{26}. Solvents were dried before using according to standard procedure\textsuperscript{27}. The transition metal salts used were in their hydrated form as received and double distilled water was used throughout the study. The melting point of the synthesized compounds was determined by the ‘melting point apparatus’ using the air atmosphere. FT-Infrared Spectra of ligand and its transition metal complexes were taken in dry KBr pellets using in the range 400–4000 cm\textsuperscript{−1} with a SHIMADZU model 8400 FT-IR spectrophotometer.\textsuperscript{1}H (300.1 MHz) Nuclear Magnetic Resonance spectra were collected on a Bruker 300 NMR spectrometer in DMSO-d\textsubscript{6} solution using TMS as an internal reference. LC-MS spectra of compounds were recorded with a Waters Q-TOF micro mass. Carbon, hydrogen, nitrogen and sulphur were analyzed with an elemental analyser. Standard Gravimetric methods were employed to find out the metal contents\textsuperscript{27}. All the compounds (1-5) have been screened against pathogenic bacterial strains of *Bacillus subtilis* (MTCC-121) and *Escherichia coli* (MTCC-40) by disc diffusion assay method\textsuperscript{30-31}. Electrochemical assessments were performed with model 1230 A [SR 400] (CHI instrument, Bee Cave, TX, USA) electrochemical analyzer using glassy carbon as the working electrode, a platinum wire as the auxiliary electrode (counter electrode), and Ag/AgCl as the reference electrode. Cyclic voltammetry studies of neat ligand, Ni\textsuperscript{II} and Cu\textsuperscript{II} complexes were carried out on 0.001 M solutions in dimethylformamide containing 0.1 M KCl as the supporting electrolyte; double distilled water, obtained from laboratory distillation assembly was used to prepare Britton-Robinson (BR) buffer solution of different pH (2 to 10) used throughout the voltammetric study.

**Pre-treatment of the Glassy Carbon Electrode**

The working electrode was polished before each experiment with Alumina slurry in water to furnish the reproducible electrode surface and to ameliorate sensitivity and resolution of voltammetric elevations. The procedure was performed at room temperature and a nitrogen atmosphere was maintained over the solution during the measurements.

**Synthesis of Ligand (PTTH)**

5-Phenyl-1,2,4-triazine-3(2H)-thione was synthesized by reported procedure\textsuperscript{28-29} with some modifications. A mixture of phenylglyoxal hydrate (1.52g; .01M), thiosemicarbazide (1.00 g; .011M) and 10% aqueous solution of potassium carbonate was refluxed at 30–35°C temperature for $\frac{1}{2}$ hr. After cooling the solution acidified with hydrochloric acid, a red precipitate was obtained which crystallized from ethanol and water.
The band corresponding to the C=S moiety has moved to lower frequencies (by 15–30 cm⁻¹) authenticated by the appearance of new ν(M-N, M-S) bands at 505–550 and 400–455 cm⁻¹ of fresh M─N and M-S coordination bond take place during the complexation. This is further suggested by the absence of ν(S–H) vibrations, tell us that mercapto triazine exhibits thiol-thione tautomerism. The absence of these bands (ν(NH, N=N, S-H)) was also corroborated by the downfield shift in the spectra of the complexes corresponding to ν(N-H, S-H) band. Further, in the spectra of all of the complexes, these bands (N-H, S-H) were disappeared.

**RESULTS AND DISCUSSION**

**In vitro Antimicrobial Assay**

To screen the biological activities of the ligand (PTTH) and their transition metal complexes, an in-vitro antibacterial assay was employed against the Gram-positive *Bacillus subtilis* (MTCC-121) and Gram-negative *Escherichia coli* (MTCC-40) as the method previously described. The concentrations used for the test were 1mg/ml and 2mg/ml of the compounds.

**FT-Infra Red Spectroscopy**

The significant FT-IR spectral data of the organic molecule and its metal complexes are disclosed in Table 1 showed some characteristic stretching bands at: 3163, 1540, 1595, cm⁻¹ assigned to N-H, N=N and C≡N of triazine ring of free ligand (PTTH), respectively. Additionally, a weak broadband appeared just about 2610 cm⁻¹ due to ν(S–H) vibrations, tell us that mercapto triazine exhibits thiol-thione tautomerism. Further, in the spectra of all of the complexes, these bands (N-H, S-H) were disappeared. The absence of these bands suggesting the deprotonation of thio triazine molecule and further constitution of fresh M─N and M-S coordination bond take place during the complexation. This is further authenticated by the appearance of new ν(M-N, M-S) bands at 505-550 and 400–455 cm⁻¹ respectively. The band corresponding to the C=S moiety has moved to lower frequencies (by 15-30 cm⁻¹) due to
decreasing of the bond order (and increasing of bond length) of carbon–sulphur bond result from coordination of the S to central metal ions. Further, the bands of C=N, and N=N of triazine moiety are moved to the lower frequency because of complexation in between, however, the other bands such as C=C, C-H of phenyllic ring did not express any quite shifting because they are not participating in the complexation to the metal ion. Thus, It deduced that the organic molecule chelated through amine N and thione S-atoms to the metal ion leading to the constitution of four-member chelate ring, and the triazine rings are virtually planar. The appearance of coordinated aqua molecules in the NiII, ZnII and CdII complexes are further expressed by a broad through band in the region 3250–3370 cm\(^{-1}\) is assigned.

### 1\(^{1}\)H-NMR Spectroscopy

The NMR spectra of the thio 1,2,4-triazine nucleus were reported for \(^1\)H. The parent compound (PTTH) showed signals at 9.7, 11.9, and 7.4-8.2 ppm (in DMSO-d\(_6\)) which were attributed to N=CH, NH, and phenyllic protons (5H). The \(^1\)HNMR of all the prepared compounds exhibit a multiplet only in the aromatic zone corresponding to the phenyl ring of the particular compound. \(^1\)H NMR spectra of metal complexes indicate the absence of the NH signal which was recorded at 11.9 ppm in the free ligand, supporting the coordination between the metal ion and Nitrogen atom leading to the formation of new M–N bond. The fresh M-N coordination in the complexes is too supported by an upfield shift in the position of the proton signal of N=CH, of the cyclic triazine moiety, compared to the neat ligand.

| Compound | \(v(N-H)\) | \(v(C=O)\) | \(v(C-S)\) | \(v(N=N)\) | \(v(H_2O/OH^-)\) | \(v(M-N)\) | \(v(M-S)\) |
|----------|-------------|-------------|-------------|-------------|-----------------|-------------|-------------|
| PTTH     | 3163        | 1595        | 840         | 1540        | --              | --          | --          |
| (Cu(ptt)\(_2\))\(_2\) \(2H_2O\) | --          | 1580        | 820         | 1535        | --              | 502         | 414         |
| (Ni(ptt)\(_2\))\(_2\) \(2H_2O\) | --          | 1575        | 825         | 1525        | 3270            | 510         | 405         |
| Zn(ptt)\(_2\))\(_2\) \(2H_2O\) | --          | 1580        | 815         | 1535        | 3285            | 522         | 408         |
| Cd(ptt)\(_2\))\(_2\) \(2H_2O\) | --          | 1575        | 820         | 1510        | 3365            | 500         | 412         |

### LC-MS Spectroscopy

The prominent molecular ion peaks in the mass spectra of compounds 1 to 5 established the molecular formula and supported the proposed structures (Fig.-2) of the complexes as obtained. The mass spectrum of neat ligand displays the parental ion peak \([M]^+\) at \(m/z = 189\) (78%) and a weak peak at \(m/z = 190\) due to \(^{13}\)C and/or \(^{15}\)N isotopes. CuII complex exhibits a molecular ion peak at \(m/z = 878.50\) (calc. M. Wt. = 877.97) with an intensity of 9% which is equivalent to its molecular weight (Fig.-3). In the mass spectrum of a molecule, peaks are attributed to fragmentation obtained from the rapture of various bonds in a particular molecule. Also, in the mass spectrum of other complexes molecular ion peak observed at \(m/z = 905, 477\) and 525 are attributed to \([Ni(ptt)\(_2\))\(_2\)\(2H_2O, Zn(ptt)\(_2\)\(2H_2O\) and Cd(ptt)\(_2\)\(2H_2O\) respectively. Cu and Ni complexes display a peak corresponding to the fragment \([M_2L_3]^+\) suggesting a binuclear species. Analytical data further indicating the non-appearance of chloride groups in the compounds. If the complexes bear two metal ions they should contain four molecules of ligand to acquire neutrality. The process to get a binuclear species as it may be because of the formation of the sulfur bridges, take place in many cyclic thiosemicarbazone complexes\(^{23-24}\). The mass spectrum of CuII complex expresses a peak regarding to a fragment \([Cu(ptt)\(_3\)]^+\) at 628.10(30%) indicating the existence of a molecule comprising one copper and three deprotonated molecules of the ligand. The base peak at \(m/z = 439.04\) (100%) is due to \(C_3H_7CuN_3S_2\). The other positive ions give the peaks at, 471.02, 252, 145, and 103.97 mass numbers. The intensities of these peaks suggest an idea regarding the stabilities of the fragments. Mass spectra of Zn and Cd complexes show a peak corresponding to the fragment \([Zn(ptt)\(_3\)]^+\) and \([Cd(ptt)\(_3\)]^+\) respectively, which indicates the existence a mononuclear comprising one metal ion and two deprotonated molecules of the ligand.

### Biological Evaluation

The ligand (PTTH) and the complexes (2-5) exhibited variable antibacterial activities against both Bacillus subtilis and Escherichia coli, are summarised in Table-2. As expected, Negative control, and
DMSO, did not exhibit any antibacterial properties. Maximum inhibition activity, thus showed the most effective against E. coli that compounds 1-5 has Zone of Inhibition (ZoI) 33.0, 27.0, 24.0, 22.0 and 21.0 mm, and further B. subtilis has ZoI 13.0, 12.0, 11.0 and 10.0 mm, respectively.

Thus, we observed that, compounds 1-5, with both the stock concentration, were active against both pathogenic bacterial strains. The nickel complex has demonstrated the highest zone of inhibition against E. coli at 33 mm. The disc diffusion assay showed that, compounds 1-5 have antibacterial properties at various concentrations. Comparatively Ni\(^{II}\), Cu\(^{II}\), Zn\(^{II}\) and Cd\(^{II}\) complexes of thio(1,2,4-)triazine have shown wider ZoI than free ligand, therefore, it does indicate the high spectrum of antibacterial activity against pathogenic strains such as E. coli, of these metal complexes.

### Table-2: Inhibition Zone (mm) of Ligand and their Metal Complexes against Pathogenic Bacteria

| S. No. | Compound   | Concentration of compound (mg/ml) | Diameter growth of inhibition zone (mm) | Antibacterial activity DMSO (Control) |
|--------|------------|----------------------------------|----------------------------------------|--------------------------------------|
| 1      | PTTH       | 1                                | 14 21                                  | ++伙伴关系                           |
| 2      | (Ni(ptt)\(_2\))\(_2\).2H\(_2\)O | 1 2                             | 30 33 09 12                           | +++伙伴关系                           |
| 3      | (Cu(ptt)\(_2\)) | 1 2                            | 15 22 08 10                           | +++伙伴关系                           |
| 4      | Zn(ptt)\(_2\).2H\(_2\)O | 1 2                            | 18 27 11 12                           | +++伙伴关系                           |
| 5      | Cd(ptt)\(_2\).2H\(_2\)O | 1 2                            | 22 24 08 11                           | +++伙伴关系                           |

(-)=no inhibition; (+)=zone size 6-8mm; (++)=zone size 9-14mm; (+++)=zone size 15-20mm; (++++)=zone size >20 mm
These results indicate the antibacterial activity of the ligands more prominent and significant while coordinating with the transition metal ions. We therefore speculate, this increased antimicrobial activity thus could be considered because of the efficient diffusion of the metal complexes in a bacterial cell or further could be interacting with them as consistent with previous studies.\textsuperscript{30-31} In metal complexes,
Overton’s concept and Tweedy’s chelation theory can further explain the enhanced biological activities as described previously\(^3^2\). As consistent, our results show the dose-dependent patterns as the growth is pace down with lower concentration, while the more inhibitory effect with the higher concentration in both bacterial strains. Furthermore, ZoI of Ni complex thus suggests the higher antibacterial activity as compared to the other synthesized compounds and therefore it could potentially be used for new therapeutic targets.

**Fig.-6:** Comparison of diameters of zone of inhibition for synthesized compounds (1-5) against Gram (-) bacterium (*E. coli*); and Gram (+) bacterium (*B. subtilis*) at two different concentrations.

**Electrochemical Assessments**

Electrochemical reduction properties of ligand, Cu\(^{II}\) and Ni\(^{II}\) complexes were studied at glassy carbon electrode using cyclic voltammetry. Cyclic voltammograms depicted unambiguous irreversible cathodic peaks at -0.560, -0.521 and -0.503 V for ligand, Ni\(^{II}\) and Cu\(^{II}\) complex respectively which can be attributed to the reduction of the groups present in the compound (vs. Ag/AgCl reference electrode in BR buffer). No peaks were observed on the reverse scan, indicating the irreversibility of electrode processes of said compounds. The design of cyclic voltammograms evinced the irreversible nature of reduction of ligand, Ni\(^{II}\) and Cu\(^{II}\) complexes as portrayed in Fig.-7. The Cyclic voltammogram of ligand (PTTH) between 0.0 and -1.0 V in DMF depicted an unambiguous irreversible cathodic peak at -0.560 V (\(E_{1/2} = -0.450\) V), which can be attributed to the reduction of the group present in the compound (vs. Ag/AgCl reference electrode in BR buffer) (Fig.-7). The Cyclic voltammograms of Cu\(^{II}\) (between 0.0 and -0.7 V) and Ni\(^{II}\) (between 0.0 and -0.8 V; Fig.-7) in DMF also exhibited an unambiguous irreversible cathodic peak at -0.503 (\(E_{1/2} = -0.402\) V) and -0.521 (\(E_{1/2} = -0.417\) V) respectively, corresponds to the Cu(II)/Cu(I) and Ni(II)/Ni(I).

**Fig.-7:** Cyclic voltammogram of the ligand and Ni\(^{II}\) complexes.
CONCLUSION

SN-donor 1,2,4-triazine ligand (PTTH) and its Ni$^{II}$, Cu$^{II}$, Zn$^{II}$, and Cd$^{II}$ complexes were successfully prepared and further investigated through elemental analysis, $^1$H-NMR, FT-IR and LC-MS spectral studies. The results indicate that, ligand acts as a mono-functional bidentate and it coordinates via deprotonated nitrogen and sulphur donor to the metal ion to afford the corresponding complexes 2 to 5 giving a four-member stable chelate ring. Antibacterial activities towards two bacterial strains, namely Bacillus subtilis (MTCC-121) and Escherichia coli (MTCC-40) were screened, and the results revealed significantly more activity for these newly synthesized derivatives as compared to free 1,2,4-triazine (PTTH) ligand. Therefore, it is suggested that the coordination of said ligand (PTTH) with different metal ions makes them stronger bacteriostatic agents, thus inhibiting the growth of bacteria more than the parent ligand. Furthermore, preliminary electrochemical results confer good prospects to elucidate the possible cathodic process for ligand, Ni$^{II}$ and Cu$^{II}$ complexes.

ACKNOWLEDGEMENT

The authors are thankful to the Head, Department of Chemistry, for providing the necessary research facilities. This study received financial assistance from the University Grant Commission, New Delhi through JRF during the research work progressed. We are also grateful to USIC (University Science Instrumentation Centre), MNIT Jaipur and Therachem lab Jaipur for providing analytical and spectral data.

REFERENCES

1. A. S. Abd-El-All, A. A. Labib, H. A. Mousa, F. A. Bassyouni, K. H. Hegab, M. A. El-Hashash, S. R. Atta-Allah, W. H. AbdEl-Hady and S. A. M. Osman, Journal of Applied Sciences Research, 9(1), 469 (2013).
2. G. A. Elmegeed, A. R. Baiuomy, M. M. Abdelhalim and H. Y. Hana, Arch Pharm Chemistry in Life Sciences, 343, 261(2010), DOI:10.1002/ardp.200900244
3. A. G. Banerjee, N. Das, S. A. Shengule, R. S. Srivastava and S. K. Shrivastava, European Journal of Medicinal Chemistry, 101, 81(2015), DOI:10.1016/j.ejmech.2015.06.020
4. D. R. Kerzarea and P. B. Khedekar, Journal of Pharmaceutical Science and Bioscientific Research, 6(1), 144 (2016).
5. M. A. El-Hashash, S. A. Rizk and A. A. El-Sayed, Journal of Advances in Chemistry, 13(12), 6130 (2017), DOI:10.24297/jac.v13i12.6188
6. T. E. Olalekan, A. S. Ogunlaja and G. M. Watkins, Heteroatom Chemistry, 1(2019), DOI:10.1155/2019/9203435
7. M. M. Mashaly, H. F. El-Shafiyy, S. B. El-Maraghy and H. A. Habib, Spectrochimica Acta Part A, 61, 1853 (2005), DOI:10.1016/j.saa.2004.06.056
8. R. M. Zaki, A. M. K. El-Dean, S. M. Radwan and A. F. Saber, Journal of the Brazilian Chemical Society, 29(12), 2482 (2018), DOI:10.21577/0103-5053.20180127
9. S. Dadashpour, T. T. Kucukkilinc, O. U. Tan, K. Ozadali, H. Irannejad and S. Emami, Arch Pharm Chemistry in Life Sciences, 348, 179(2015), DOI:10.1002/ardp.201400400
10. I. Karpenko, S. Deev, O. Kiselev, V. Charushin, V. Rusinov, E. Ulomsky, E. Deeva, D. Yanvarev, A. Ivanov, O. Smirnova, S. Kochetkov, O. Chupakhin, M. Kukhanova. Antimicrobial Agents and Chemotherapy, 54(5), 2017 (2010), DOI:10.1128/AAC.01186-09
11. M. Mojzych, Z. Bernat, Z. Karczmarzyk, J. Matysiak and A. Fruziński, Molecules, 25, 1(2020), DOI:10.3390/molecules25010221
12. M. S. T. Makki, R. M. A. Rahman and O. A. A. Ali, International Journal of Organic Chemistry, 5(3), 153 (2015), DOI:10.4236/ijoc.2015.53017
13. K. Ban, S. Duffy, Y. Khakham, V. M. Avery, A. Hughes, O. Montagnat, K. Katneni, E. Ryan and J. B. Baell, Bioorganic & Medicinal Chemistry Letters, 20, 6024(2010), DOI:10.1016/j.bmcl.2010.08.065
14. D. Branowska, J. Ławecka, M. Sobiczewski, Z. Karczmarzyk, W. Wysocki, E. Wolińska, E. Olender, B. Mirowska, A. Perzyńska, A. Bielsawska and K. Bielsawska, Monatsh Chem, 149, 1409(2018), DOI:10.1007/s00706-018-2206-y
15. N. P. Prajapati and H. D Patel, *Synthetic Communications*, 49, 2767(2019), DOI: 10.1080/00397911.2019.1649432
16. S. Chandra, Sangeetika and S. Thakur, *Transition Metal Chemistry*, 29, 925(2004), DOI: 10.1080/00397911.2019.1649432
17. U. M. Aswar, P. P. Kalshetti, S. M. Shelke, S. H. Bhosale and S. L. Bodhankar, *Asian Pacific Journal of Tropical Biomedicine*, 2(12), 992 (2012), DOI:10.1006/jpbb.2001.1512-1543
18. I. F. Nassar, *Journal of Heterocyclic Chemistry*, 50(1), 129 (2013), DOI:10.1002/jhet.1022
19. G. G. Berest, O. Y. Voskoboynik, S. I. Kovalenko, O. M. Antypenko, I. S. Nosulenko, A. M. Katsev and O. S. Shandrovska, *European Journal of Medicinal Chemistry*, 46(12), 6066(2011), DOI:10.1016/j.ejmech.2011.10.022
20. H. A. Saad, M. M. Youssef and M. A. Mosselhi, *Molecules*, 16(6), 4937(2011), DOI: 10.3390/molecules16064937
21. A. Gouranourimi, M. Ghasemzadeh, S. Bahemmat, B. Neumuller and R. Tonner, *Monatsh Chem.*, 146, 57 (2015), DOI: 10.1007/s00706-014-1280-z
22. M. A. Blanco, E. Lopez-Torres, M. A. Mendiola, E. Brunet and M. T. Sevilla, *Tetrahedron*, 58, 1525 (2002), DOI:10.1016/S0040-4020(02)00016-9
23. E. Lopez-Torres and M. A. Mendiola, *Polyhedron*, 24, 1435(2005), DOI:10.1016/j.poly.2005.03.093
24. E. Lopez-Torres, M. A. Mendiola and C. J. Pastor, *Inorganic Chemistry*, 45(7), 3103(2006), DOI:10.1021/ic052009d
25. E. Lopez-Torres, M. A. Mendiola and C. J. Pastor, *Polyhedron*, 25, 1464(2006), DOI:10.1016/j.poly.2005.10.013
26. H. A. Riley and A. R. Gray, Organic Syntheses, 15, 67 (1935), DOI:10.15227/orgsyn.015.0067
27. A. I. Vogel, Text Book of Quantitative Chemical Analysis, London:Longmans, Addison Wesley, (1999).
28. K. C. Joshi, K. Dubey and A. Dandia *Heterocycles*, 16 (9), 1545(1981).
29. M. Tisler, *Croatica Chemica Acta*, 32, 123(1960).
30. S. Beniwal, S. Chhipa, D. Gaur, P. J. John, Y. Singh and J. Sharma, *Applied Organometallic Chemistry*, 31(10), (2016), DOI:10.1002/aoc.3725
31. A. W. Bauer, W. M. M. Kirby, J. C. Sherris and M. Turck, *The American Journal of Clinical Pathology*, 45, 493(1966).
32. B. G. Tweedy, *Phytopatology*, 55, 910(1964).