Differential Influence of Age on the Relationship between Genetic Mismatch and A(H1N1)pdm09 Vaccine Effectiveness

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Abstract: Assessment of influenza vaccine effectiveness (VE) and identification of relevant influencing factors are the current priorities for optimizing vaccines to reduce the impacts of influenza. To date, how the difference between epidemic strains and vaccine strains at genetic scale affects age-specific vaccine performance remains ambiguous. This study investigated the association between genetic mismatch on hemagglutinin and neuraminidase genes and A(H1N1)pdm09 VE in different age groups with a novel computational approach. We found significant linear relationships between VE and genetic mismatch in children, young adults, and middle-aged adults. In the children’s group, each 3-key amino acid mutation was associated with an average of 10% decrease in vaccine effectiveness in a given epidemic season, and genetic mismatch exerted no influence on VE for the elderly group. We demonstrated that present vaccines were most effective for children, while protection for the elderly was reduced and indifferent to vaccine component updates. Modeling such relationships is practical to inform timely evaluation of VE in different groups of populations during mass vaccination and may inform age-specific vaccination regimens.

Keywords: A(H1N1)pdm09; influenza vaccine effectiveness; age-related effects; genetic mismatch

1. Introduction

Since the emergence of the pandemic H1N1 influenza in early 2009, the swine-origin A(H1N1)pdm09 virus has replaced the seasonal H1N1 subtype and become one of the four major circulating viruses underlying seasonal influenza epidemics [1]. The new H1N1 pandemic virus is antigenically distinct from the former seasonal H1N1 virus [2]. In response, the new subtype is introduced as one of the components in the trivalent or quadrivalent seasonal influenza vaccines [3]. As influenza viruses are evolving rapidly, the World Health Organization (WHO) annually reviews and recommends influenza vaccine compositions for vaccine updates.

Typically, vaccine effectiveness (VE) estimation is conducted by retrospective investigation in the middle and late stage during epidemics or at the end of the flu season [4,5]. The current VE exhibits high season-to-season variations, mostly due to the genetic mismatch
of the circulating strains and the vaccine virus [6]. Apart from survey-based VE studies, the hemagglutination inhibition (HI) test is a standard method to evaluate the antigenic distance [7]. The HI test detects the prevention of binding of the epitopes in the hemagglutinin protein to the sialic acid receptors on red blood cells [8]. Studies based on antigenic distance showed that past vaccinations might have a negative or positive interference on current vaccine efficacy depending on the antigenic relatedness among the previous vaccine virus, current vaccine virus and the circulating strains [9–11]. Sequence-based methods were also proposed to estimate the VE using various genetic mismatch summary statistics [12,13]. These sequence-based studies showed a clear trend that a weaker VE is associated with a larger genetic mismatch of the circulating strains and vaccine viruses. However, the existing genetic-VE relationship is developed for the all-age group. Although it is clear that the seniority is associated with a reduced vaccine-induced immune response [14–17], it is unclear whether genetic mismatch will generate a similar VE response in age generations. Therefore, in this study, based on the previous statistical framework [13], we quantified the effect of genetic mismatch on observed VE in different age groups.

2. Materials and Methods

2.1. Genetic Data

Protein sequences of human influenza A(H1N1)pdm09 virus were retrieved from the global initiative on sharing all influenza data (GISAID) [18], with sampling dates ranging from 1 January 2009 to 31 December 2019. The sequences with age information of the host were collected. We stratified the samples into five groups according to the commonly adopted scheme in VE studies, namely: (1) children: age ≤ 8 years; (2) adolescents: 8 years < age ≤ 17 years; (3) young adults: 17 years < age ≤ 49 years; (4) middle-aged adults: 49 years < age ≤ 64 years; and (5) the elderly: age > 64 years [5]. Duplicated strains were removed. For the model building samples, in total 5232 hemagglutinin (HA) and neuraminidase (NA) strains were retained with sample origins of the United States and Canada. The independent validation set included a total number of 3470 strains from the United Kingdom, Germany, Spain, Italy, France, Greece, Sweden, Mexico, Japan, and Hong Kong. Multiple sequence alignment was performed with Clustal X [19]. The detailed sample size of genetic data is shown in Supplementary Materials Table S1.1. All sequence data used in the analysis are acknowledged in the Supplementary Acknowledgment Table.

2.2. Vaccine Effectiveness Data

The VE data were collected from published studies in the target countries. The inclusion criteria of the studies were: (1) VE evaluation (or corresponding odds ratio) was reported for A(H1N1)pdm09; (2) original research of VE was conducted with a test-negative design by observational studies; (3) RT-PCR method was used to confirm the positive status of infection; (4) patients were recruited by predefined illness criteria; and (5) age-stratified VE outcomes were reported. The exclusion criteria included: (1) the study was restricted to a population with special conditions such as asthma patients or pregnant women; (2) interim reports and, if relevant, final reports were available. The eligible studies between 2009 and 2020 are listed in Supplementary Materials Table S1.2.

2.3. Statistical Methods

In previous research, we proposed the effective mutation distance (EMD) as a summary statistic to evaluate genetic mismatch between vaccine strains and circulating strains [13]. The EMD was defined by the Hamming distance on the effective mutations (EMs) underlying seasonal influenza epidemics [20–22], which could be interpreted as the amino acid substitutions that contributed to the viral escape from herd immunity. Thus, the EMs served as a rational basis to evaluate the genetic distance associated with vaccine response. The list of EM sites for the A(H1N1)pdm09 can be found in Supplementary Materials Table S1.3. We calculated EMDs for HA and NA genes separately in each epidemic season [13]. The generalized linear regression model was fitted to evaluate the relationship between the
EMD and the VEs for each age group. Repeated-measures analysis of variance (ANOVA) and the Friedman test were respectively used to assess the difference in observed VE and genetic measure across age groups. Paired t-test and Wilcoxon signed-rank test were used to perform pairwise comparisons. Statistical significance was declared if p-value < 0.05. All analyses were conducted using R statistical software (version 4.0.3) [23].

3. Results

**VE, EMD and Genetic Effect Size across Age Groups**

First, we compared the observed VE and genetic mismatch of the isolated viruses to the vaccine strain in all age groups. As expected, the observed VEs of the young adults, middle-aged and elderly groups were significantly lower compared with the children and adolescent groups (p-value = 0.01 for two-sided paired t-test, see Figure 1a). Secondly, the genetic measure in terms of the EMD of the HA gene showed no statistically significant difference between the adults and the younger age groups (p-value = 0.26 for Wilcoxon test, Figure 1b). Next, we conducted linear regression analysis within each age stratum, and the effect size of the HA-EMD inferred per key substitution’s associated reduction in VE. Figure 2 plots the age-stratified linear relationship between the EMD and VE. In the children, young adults and middle-aged groups, the estimated per substitution effect on the reduction of VE was 3.44%, 3.57%, and 3.95% in absolute terms, respectively (p-values < 0.01, Figure 2a,c,d). That is to say, every three key amino acid mutations will induce a decrease in vaccine effectiveness by an average of ~10% in a given epidemic season. For adolescents, HA mismatch was weakly associated with VE, and per mismatch yielded an average of a 2.25% decrease in VE (p-value = 0.10, Figure 2b). We found that genetic mismatch exerted no influence on VE for the elderly group (p-value = 0.83, Figure 2e).

![Figure 1](image-url). Comparison of H1N1 vaccine effectiveness (VE) and effective mutation distance (EMD) among age groups. Panel (a): distribution of the observed VE in different age groups. VE in the younger groups was higher than adults (paired t-test p-value = 0.01). Panel (b): EMD distribution in different generations. No statistical difference in EMD between the younger groups and the adult groups is present (Wilcoxon test p-value = 0.26). In panels (a) and (b), dashed lines represent the mean observed VE and the median EMD of the children’s group.

In each age group, independent validation data points were overlaid on the estimated relationship between HA-EMD and VE (Figure 2). In the children, young adults and middle-aged groups in which the genetic effect was significant, validation data showed close clustering around the predicted line, with an average mean absolute error (MAE) of 13.4%. The intercept of the fitted regression implied the highest VE achievable for existing influenza H1N1 vaccines when there was no genetic mismatch on key loci (EMD = 0), the
range of which was covered by the observed data. Figure 2 showed that the children’s group would result in a maximum VE of 80.5% [95% CI: 67.9–93.1%] if a perfect match of vaccine virus is present, while this optimal value was slightly lower, 76.6% [95% CI: 66.7–86.5%], for the young adults and 74.0% [95% CI: 56.4–91.6%] for the middle-aged adult group, respectively. The highest VE was 71.0% [95% CI: 53.1–88.9%] for the adolescent group. In the elderly population, no linear relationship was observed between genetic mismatch and VE, and the grand mean of VE was 44.4% [95% CI: 19.8–69.0%], regardless of the matching level between the vaccines and circulating strains.

Figure 2. The relationship between vaccine effectiveness (VE) of A(H1N1)pdm09 and effective mutation distance (EMD) on hemagglutinin (HA) gene, in different age groups: (a) children; (b) adolescents; (c) young adults; (d) middle-aged adults; (e) the elderly. Green dots: model building samples from the United States and Canada; pink cross: independent validation samples from the United Kingdom, Germany, Spain, Italy, France, Greece, Sweden, Mexico, Japan, and Hong Kong. In the first four age groups, clear linear relationships were observed between VE and the EMD; the validation samples largely followed the relationship estimated by the North American samples (a–d). In the elderly population, no statistically significant association is present between VE and the EMD.

We also conducted analysis using the EMD of the NA gene and VE (Supplementary Materials Figure S2.1). NA-EMD showed a generally consistent pattern of effect size distribution, in which the children, young adults, and the middle-aged groups had the largest genetic effect size on VE, while the VE of the elderly group was not related to vaccine matching levels. In general, the magnitudes of NA genetic effect were lower compared with HA-EMD.

4. Discussion

This study explored the role of genetic mismatch on vaccine performance in different age groups for the influenza A(H1N1)pdm09 virus. We identified a significant negative association between the genetic mismatch measure and observed VE among the children, young adults, and middle-aged groups. We reported a weak association in the adolescent group, and the relationship does not appear statistically evident in the elderly group. For an ideal genetically matched vaccine, the 0–8 years old group would exhibit the highest vaccine protection effect, while the age > 64 years group would not show differential response to the alterations of vaccine components.

Our findings characterized impaired immunity in the elderly population by the relationship between genetic mismatch and vaccine response. Previous studies reported relatively lower vaccine performance in the senior group [15,16]. One possible explana-
The main limitation of this study is that only genetic factors were considered in the analysis, and in-host immunological measurements were not included as a constraint by limited resources.

5. Conclusions

We demonstrated that influenza vaccine effectiveness was evidently enhanced in non-elderly groups when genetic matching was improved, while vaccine protection for the elderly was generally lower and indifferent to vaccine component updates. Through evaluating the genetics and vaccine response relationship, this study indicates the importance of optimizing vaccine components to match circulating viruses, and a special vaccination scheme is needed to increase the protection of the elderly population.

Supplementary Materials: The following are available online at https://www.mdpi.com/article/10.3390/v13040619/s1. Table S1.1: Genetic data sample size-number of strains. Table S1.2: Influenza A(H1N1)pdm09 vaccine effectiveness in Northern Hemisphere, 2009-2019. Table S1.3: List of Effective Mutation (EM) sites on HA and NA genes of A(H1N1)pdm09. Figure S2.1: The relationship between H1N1 VE and EMD on neuraminidase (NA), in different age groups.

Author Contributions: M.H.W. conceived the study, L.C., M.H.W. and S.Z. wrote the manuscript. L.C carried out the analysis. J.L., H.Z., R.W.Y.C., M.K.C.C., Z.C. and P.K.S.C. revised the manuscript. B.C.Y.Z. approved the paper. All authors have read and agreed to the published version of the manuscript.

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**Conflicts of Interest:** M.H.W. and B.C.Y.Z. are shareholders of Beth Bioinformatics Co., Ltd. B.C.Y.Z. is a shareholder of Health View Bioanalytics Ltd. All other authors declare no competing interests.

**References**

1. Kramer, F.; Smith, G.J.D.; Fouchier, R.A.M.; Peiris, M.; Kedzierska, K.; Doherty, P.C.; Palese, P.; Shaw, M.L.; Treanor, J.; Webster, R.G.; et al. Influenza. Nat. Rev. Dis. Prim. 2018, 4, 3. [CrossRef]

2. Garten, R.J.; Davis, C.T.; Russell, C.A.; Shu, B.; Lindstrom, S.; Balish, A.; Sessions, W.M.; Xu, X.; Skepner, E.; Deyde, V.; et al. Antigenic and Genetic Characteristics of Swine-Origin 2009 A(H1N1) Influenza Viruses Circulating in Humans. Science 2009, 325, 197–201. [CrossRef]

3. Yamayoshi, S.; Kawaoka, Y. Current and future influenza vaccines. Nat. Med. 2019, 25, 212–220. [CrossRef]

4. Skowronski, D.M.; Zou, M.; Sabaiduc, S.; Murti, M.; Olsha, R.; Dickinson, J.A.; Gubbay, J.B.; Croxen, M.A.; Charest, H.; Jassem, A.; et al. Interim estimates of 2019/20 vaccine effectiveness during early-season co-circulation of Influenza A and B viruses, Canada, February 2020. Eurosurveillance 2020, 25. [CrossRef]

5. Flannery, B.; Kondor, R.J.G.; Chung, J.R.; Gaglani, M.; Reis, M.; Zimmerman, R.K.; Nowalk, M.P.; Jackson, M.L.; Jackson, L.A.; Monto, A.S.; et al. Spread of Antigenically Drifted Influenza A(H3N2) Viruses and Vaccine Effectiveness in the United States During the 2018–2019 Season. J. Infect. Dis. 2020, 221, 8–15. [CrossRef]

6. Paules, C.I.; Sullivan, S.G.; Subbarao, K.; Fauci, A.S. Chasing Seasonal Influenza—The Need for a Universal Influenza Vaccine. N. Engl. J. Med. 2018, 378, 7–9. [CrossRef]

7. Smith, D.J.; Lapedes, A.S.; De Jong, J.C.; Bestebroer, T.M.; Rimmelzwaan, G.F.; Osterhaus, A.D.M.E.; Fouchier, R.A.M. Mapping the Antigenic and Genetic Evolution of Influenza Virus. Science 2004, 305, 371–376. [CrossRef]

8. Smith, D.J.; Forrest, S.; Ackley, D.H.; Perelson, A.S. Variable efficacy of repeated annual influenza vaccination. Proc. Natl. Acad. Sci. USA 1999, 96, 14001–14006. [CrossRef]

9. Skowronski, D.M.; Chambers, C.; De Serres, G.; Sabaiduc, S.; Winter, A.-L.; Dickinson, J.A.; Gubbay, J.B.; Fonseca, K.; Drews, S.J.; Charest, H.; et al. Serial Vaccination and the Antigenic Distance Hypothesis: Effects on Influenza Vaccine Effectiveness During A(H3N2) Epidemics in Canada, 2010–2011 to 2014–2015. J. Infect. Dis. 2017, 215, 1059–1099. [CrossRef] [PubMed]

10. Linderman, S.L.; Chambers, B.S.; Zost, S.J.; Parkhouse, K.; Li, Y.; Herrmann, C.; Ellebedy, A.H.; Carter, D.M.; Andrews, S.F.; Zheng, N.-Y.; et al. Potential antigenic explanation for atypical H1N1 infections among middle-aged adults during the 2013–2014 influenza season. Proc. Natl. Acad. Sci. USA 2014, 111, 15798–15803. [CrossRef] [PubMed]

11. Pan, K.; Subieta, K.C.; Deem, M.W. A novel sequence-based antigenic distance measure for H1N1, with application to vaccine effectiveness and the selection of vaccine strains. Protein Eng. Des. Sel. 2010, 24, 291–299. [CrossRef]

12. Cao, L.; Lou, J.; Zhao, S.; Chan, R.W.; Chan, M.C.; Chong, M.K.K.; Wu, W.K.K.; Chan, P.K.; Wei, Y.; et al. Characterization of the evolutionary dynamics of influenza A H3N2 hemagglutinin. BioRxiv 2020. [CrossRef]

13. Shu, Y.; McCauley, J. GISAID: Global initiative on sharing all influenza data—From vision to reality. Eurosurveillance 2017, 22, 13. [CrossRef]

14. Larkin, M.A.; Blackshields, G.; Brown, N.P.; Chenna, R.; Mcgettigan, P.A.; McWilliam, H.; Valentin, F.; Wallace, I.M.; Wilm, A.; Lopez, R.; et al. Clustal W and Clustal X version 2.0. Bioinformatics 2007, 23, 2947–2948. [CrossRef]

15. Wang, M.H.; Lou, J.; Cao, L.; Zhao, S.; Chan, P.K.; Chan, M.C.; Chong, M.K.; Wu, W.K.K.; Chan, R.W.; Wei, Y.; et al. Influenza Virus Vaccination Elicits Poorly Adapted B Cell Responses in Elderly Individuals. Cell Host Microbe 2019, 25, 357–366.e6. [CrossRef] [PubMed]

16. Sasaki, S.; Sullivan, M.; Narvaez, C.F.; Holmes, T.H.; Furman, D.; Zheng, N.-Y.; Nishtala, M.; Wrammert, J.; Smith, K.; James, J.A.; et al. Limited efficacy of inactivated influenza vaccine in elderly individuals is associated with decreased production of vaccine-specific antibodies. J. Clin. Investig. 2011, 121, 3109–3119. [CrossRef] [PubMed]

17. Henry, C.; Zheng, N.-Y.; Huang, M.; Cabanov, A.; Rojas, K.T.; Kaur, K.; Andrews, S.F.; Palm, A.-K.E.; Chen, Y.-Q.; Li, Y.; et al. Influenza Virus Vaccination Elicits Poorly Adapted B Cell Responses in Elderly Individuals. Cell Host Microbe 2019, 25, 357–366.e6. [CrossRef] [PubMed]

18. Rolfs, M.A.; Flannery, B.; Chung, J.R.; O’Halloran, A.; Garg, S.; Belongia, E.A.; Gaglani, M.; Zimmerman, R.K.; Jackson, M.L.; Monto, A.S.; et al. Efforts of Influenza Vaccination in the United States During the 2017–2018 Influenza Season. Clin. Infect. Dis. 2019, 69, 1845–1853. [CrossRef] [PubMed]

19. Shu, Y.; McCauley, J. GISAID: Global initiative on sharing all influenza data—From vision to reality. Eurosurveillance 2017, 22, 13. [CrossRef]

20. Wang, M.H.; Lou, J.; Cao, L.; Zhao, S.; Chan, P.K.; Chan, M.C.; Chong, M.K.; Wu, W.K.K.; Chan, R.W.; Wei, Y.; et al. Characterization of the evolutionary dynamics of influenza A H3N2 hemagglutinin. BioRxiv 2020. [CrossRef]
22. Lou, J.; Zhao, S.; Cao, L.; Chong, M.K.; Chan, R.W.; Chan, P.K.; Zee, B.C.; Yeoh, E.-K.; Wang, M.H. Predicting the dominant influenza A serotype by quantifying mutation activities. *Int. J. Infect. Dis.* 2020, 100, 255–257. [CrossRef]

23. Team, R.C. *R: A Language and Environment for Statistical Computing*; GBIF: Copenhagen, Denmark, 2013.

24. McElhaney, J.E. Influenza vaccine responses in older adults. *Ageing Res. Rev.* 2011, 10, 379–388. [CrossRef]

25. Gostic, K.M.; Bridge, R.; Brady, S.; Viboud, C.; Worobey, M.; Lloyd-Smith, J.O. Childhood immune imprinting to influenza A shapes birth year-specific risk during seasonal H1N1 and H3N2 epidemics. *PLoS Pathog.* 2019, 15, e1008109. [CrossRef] [PubMed]

26. Sanyal, M.; Holmes, T.H.; Mæcker, H.T.; Albrecht, R.A.; Dekker, C.L.; He, X.-S.; Greenberg, H.B. Diminished B-Cell Response After Repeat Influenza Vaccination. *J. Infect. Dis.* 2019, 219, 1586–1595. [CrossRef] [PubMed]

27. Vatti, A.; Monsalve, D.M.; Pacheco, Y.; Chang, C.; Anaya, J.-M.; Gershwin, M.E. Original antigenic sin: A comprehensive review. *J. Autoimmun.* 2017, 83, 12–21. [CrossRef] [PubMed]

28. Ciabattini, A.; Nardini, C.; Santoro, F.; Garagnani, P.; Franceschi, C.; Medaglini, D. Vaccination in the elderly: The challenge of immune changes with aging. *Semin. Immunol.* 2018, 40, 83–94. [CrossRef]

29. Wilkinson, K.; Wei, Y.; Szwajcer, A.; Rabbani, R.; Zarychanski, R.; Abou-Setta, A.M.; Mahmud, S.M. Efficacy and safety of high-dose influenza vaccine in elderly adults: A systematic review and meta-analysis. *Vaccine* 2017, 35, 2775–2780. [CrossRef]

30. Ranjeva, S.; Subramanian, R.; Fang, V.J.; Leung, G.M.; Ip, D.K.M.; Perera, R.A.P.M.; Peiris, J.S.M.; Cowling, B.J.; Cobey, S. Age-specific differences in the dynamics of protective immunity to influenza. *Nat. Commun.* 2019, 10, 1–11. [CrossRef]

31. Meade, P.; Kuan, G.; Strohmeier, S.; Maier, H.E.; Amanat, F.; Balmaseda, A.; Ito, K.; Kirkpatrick, E.; Javier, A.; Gresh, L.; et al. Influenza Virus Infection Induces a Narrow Antibody Response in Children but a Broad Recall Response in Adults. *mBio* 2020, 11. [CrossRef]

32. Ando, S. Estimation of the Effectiveness of Quadrivalent Influenza Vaccines by Distinguishing Between Influenza A (H1N1)pdm09 and Influenza A (H3N2) Using Rapid Influenza Diagnostic Tests During the 2018-2019 Season. *Intern. Med.* 2020, 59, 933–940. [CrossRef] [PubMed]

33. Dormitzer, P.R.; Galili, G.; Castellino, F.; Golding, H.; Khurana, S.; Del Giudice, G.; Rappuoli, R. Influenza vaccine immunology. *Immunol. Rev.* 2010, 239, 167–177. [CrossRef] [PubMed]