Danggui Buxue Tang Inhibits 2,4-Dinitrochlorobenzene: Induced Atopic Dermatitis in Mice

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Danggui Buxue Tang (DBT) is a herbal decoction that has been used in Chinese medicine to enhance qi and blood circulation. Previously, we found that DBT can suppress allergy-related asthma in mice, leading us to hypothesize that DBT might ameliorate allergy disease. In this study, we evaluated whether DBT can attenuate atopic dermatitis (AD) symptoms and have an anti-inflammatory effect on AD-like mice. The dorsal skin of female mice was shaved and sensitized cutaneously (skin smear) with 1-chloro-2,4-dinitrobenzene. Mice were then given various doses of DBT from days 14 to 29 cutaneously. DBT treatment suppressed ear swelling and skin inflammation and decreased mast cell and eosinophil infiltration into skin and ear tissue. DBT also inhibited levels of IgE and Th2-associated cytokine levels in serum. These results demonstrate that cutaneous administration of DBT reduced the development of AD-like skin lesions in mice.

1. Introduction

Atopic dermatitis (AD) is a chronic inflammatory allergic and relapsing skin disease, and its morbidity has been increasing gradually in developing and developed countries [1]. Approximately 50% of patients experience onset before the age of 5 years [2]. AD attacks are characterized by redness and itching in the responsive skin, and excessive scratching can cause skin cracking and fluid leakage [3]. In patients with chronic AD, the skin will gradually thicken and become rough, affecting its appearance. These pathological characteristics can interfere with mood and lifestyle [4].

Treatment of AD consists mainly of steroid cream applied to the skin [5]. However, steroids are immunosuppressants and do not reduce Th2 cell function, although they do suppress immunity and increase the risk of bacterial infection [6]. Furthermore, long-term topically applied steroids also lead to thinning of the skin and to cracking and bleeding [7].

Previous studies found that activation of T cells, especially Th2 cells, can lead to induction of the allergic response [8]. Th2 cells secrete IL-4 to activate B cells for IgE production, thus inducing activation of mast cells and causing the allergic reaction [9]. In addition, Th2 cells secrete IL-5 to induce eosinophil differentiation and infiltration into the allergic
Evidence-Based Complementary and Alternative Medicine

2. Evidence-Based Complementary and Alternative Medicine

2.1. Animals. All animal experimental protocols were approved by the Animal Care Committee of Chang Gung University of Science and Technology and Chang Gung University (IACUC approval number: 2012-001). Eight-week-old female BALB/c mice were purchased from the National Laboratory Animal Center (Taiwan) and housed at a consistent temperature (23 ± 2°C) in the air-controlled conventional animal room at the Animal Center of Chang Gung University.

2.2. Preparation of DBT. Danggui Buxue Tang (DBT), which contains Astragalus membranaceus (AM) and Angelica sinensis (AS) (AM:AS = 5:1), was prepared as previously described [11]. In brief, AS and AM were planted and collected from Gansu and Shanxi province, respectively, China. The roots of 100 g AS and AM were soaked in 1000 mL water and boiled for 60 minutes, respectively. Then, extracts were centrifuged, and the supernatants were lyophilized. AS and AM were mixed with excipients. The powder contained 1.1 g/g AS or 1.4 g/g AM, respectively.

2.3. Sensitization, Challenge, and Drug Treatment. The dorsal skin of the mice was shaved and sensitized by application to the skin of 0.5% 1-chloro-2,4-dinitrobenzene (DNCB, Sigma-Aldrich, St. Louis, MO, USA) as previously described [17]. Briefly, BALB/c mice were shaved of dorsal hair and 200 μL 0.5% DNCB in acetone/olive oil (3:1) was applied to the shaved area on experimental days 1–3 (Figure 1). On days 14–29, mice were challenged with 100 μL 1% DNCB and on the dorsal skin on days 14, 17, 20, 23, 26, and 29.

Mice were randomly divided into four groups (n = 8 per group): normal control mice (N group) were sensitized and challenged with normal saline; sensitized control mice (S group) were sensitized and challenged with DNCB; and experimental mice were sensitized and challenged with DNCB and topical skin treatment with 3 g/kg or 10 g/kg DBT (groups designated as HD3 and HD10, resp.). All drug treatments were applied to the backs of the mice from days 14 to 29 as described in Figure 1.

2.4. Measurement of Ear Thickness. On day 30, ear thickness was measured using a dial gauge (Mitutoyo, Tokyo, Japan) as previously described [17].

2.5. Histopathological Studies. The ears and dorsal skin were fixed in 10% formalin. The tissues were cut into 6 μm thick sections and stained with hematoxylin and eosin (H&E) and toluidine blue as previously described [17]. The sections were observed using a light microscope at 100–200x magnification, and the mast cells were measured under 10–15 high-power fields (HPFs).
2.6. Serum Collection and Splenocyte Cultures. Blood was collected and centrifuged to harvest serum. Serum and spleen cells were isolated as previously described [11], and cells (5 × 10⁶ cells/mL) were then stimulated with 2 µg/mL concanavalin A (Con A) for 2 days in RPMI 1640 medium. The supernatants were collected and cytokine levels measured by ELISA.

2.7. Measurement of Cytokine and Antibody Levels. Cytokines and antibodies were measured as previously described [18]. Supernatants of spleen cells were measured for the specific cytokines, including interferon (IFN-γ), IL-4, IL-5, and tumor necrosis factor (TNF-α) (R&D Systems, Minneapolis, MN, USA), as were levels of serum antibodies IgG1, IgG2a, and IgE (BD Biosciences, San Diego, CA, USA) by ELISA kits.

2.8. Statistical Analysis. All statistical significance was determined by one-way ANOVA, followed by Dunnett’s post hoc test. Results are presented as mean ± SEM (standard error of the mean), and P values were considered statistically significant at <0.05.

3. Results

3.1. DBT Inhibits Ear Swelling and Ameliorates AD Symptoms in AD-Like Mice. DNCB-sensitized mice clearly manifested AD-like syndromes in the skin and ear, including edema, erythema, scarring, and excoriation compared with normal mice (Figure 2(a)). Because DNCB could induce ear swelling in this experimental model, we measured ear thickness on day 30 (Figure 2(b)). Different topical doses of DBT to DNCB-sensitized mice significantly inhibited ear swelling compared with DNCB-sensitized mice (HD3: 0.49 ± 0.09 mm, P < 0.05; HD10: 0.37 ± 0.08 mm, P < 0.01, vs. the S group: 0.63 ± 0.05 mm).

3.2. DBT Decreases Eosinophil and Mast Cell Infiltration in AD-Like Mice. To investigate the effect of DBT in AD-like skin lesions, ear and skin sections were stained with H&E to examine epidermal thickness and eosinophil infiltration (Figures 3 and 4). In brief, DNCB-sensitized mice exhibited thickening of the epidermal layer and more eosinophil infiltration compared with normal mice. Topical administration of DBT significantly decreased the epidermal layer and eosinophil infiltration compared with DNCB-sensitized mice. In addition, topical administration of DBT also suppressed mast cell infiltration in the ears and skin compared with DNCB-sensitized mice (ear: HD3: 22.5 ± 6.8, P = 0.14; HD10: 18.5 ± 2.4, P < 0.01, vs. the S group: 36.0 ± 4.5, Figure 5; and skin: HD3: 33.6 ± 4.1, P < 0.01; HD10: 22.8 ± 5.6, P < 0.01, vs. the S group: 61.4 ± 5.7, Figure 6).

3.3. DBT Affects Serum Antibodies and Cytokines. Next, we examined antibody and cytokine levels to evaluate whether DBT could modulate the allergic or inflammatory response in serum (Figure 7). Topical administration of DBT significantly decreased IL-4 and TNF-α levels compared with DNCB-sensitized mice (IL-4, HD3: 25.4 ± 2.4 pg/mL, P = 0.23; HD10: 18.3 ± 2.1 pg/mL, P < 0.01, vs. S: 29.8 ± 2.9 pg/mL, resp.; and TNF-α, HD3: 17.3 ± 2.1 pg/mL, P = 0.18; HD10: 13.2 ± 1.5 pg/mL, P < 0.05, vs. S: 19.5 ± 2.1 pg/mL, resp.). In addition, DBT also significantly suppressed IgE and IgG1 levels but did not modulate the IgG2a level.

3.4. DBT Suppresses Th2-Associated Cytokines in Spleen Cell Cultures. Splenocytes were isolated and stimulated with Con A for 2 days. Topical administration of 10 g/kg DBT decreased the levels of IL-4 and IL-5 and increased IFN-γ production compared with DNCB-sensitized mice (Figure 8).

4. Discussion

AD is a chronic inflammatory skin disease, and skin lesions are characterized by erythema, eczema, and itching [4]. The repeated allergic, inflammatory skin response and the remodeling of the skin surface can lead to hardening and thickening of the skin [19]. In addition, intense pruritus can interfere with work and sleep, and roughening, edema, and hemorrhage of skin also affect its appearance [20].

Topical steroids are used to treat AD clinically, and antihistamine and antibiotic ointments often also are used to...
Figure 3: The effects of DBT on eosinophil infiltration in the ears. Ear sections were stained with H&E and analyzed for eosinophil infiltration in normal control mice (a); sensitized control mice (b); 3 g/kg DBT (HD3, c) or 10 g/kg DBT (HD10, d) (100x magnification); $n = 8$ mice per group for all groups.

Figure 4: The effects of DBT on eosinophil infiltration in skin. Skin sections were stained with H&E and analyzed for eosinophil infiltration in normal control mice (a); sensitized control mice (b); 3 g/kg DBT (HD3, c) or 10 g/kg DBT (HD10, d) (100x magnification, and amplified graph is 200x); $n = 8$ mice per group for all groups. Black arrows indicate eosinophils.
Figure 5: The effects of DBT on mast cell infiltration in ear tissue. Ear sections were stained with toluidine blue to analyze mast cell infiltration in normal control mice (a); sensitized control mice (b); 3 g/kg DBT (HD3, c) or 10 g/kg DBT (HD10, d) (100x magnification); n = 8 mice per group for all groups. Arrows indicate mast cells. The mast cells were measured under 10–15 high-power fields (HPFs) (e). Data are presented as means ± SEM. * P < 0.05, ** P < 0.01 versus sensitized control mice.

improve the chronic itching and edema and prevent bacterial infection [21]. However, the long-term use of steroids may involve side effects and cause skin thinning and brittleness or even skin cracking [22]. Previous studies found that Staphylococcus aureus has been isolated in the inflamed skin of AD patients. S. aureus can release a superantigen that induces more T cell activity and aggravates skin inflammation and allergy symptoms [23]. Hence, AD patients need to use topical antibiotics to inhibit bacterial colonization and decrease skin inflammation. Recent studies have found that S. aureus has tolerance to steroids and antibiotics [5]. Therefore, in recent years, many researchers have sought other therapies to address AD.

Traditional Chinese medicine has used acupuncture and Chinese herbal therapies to treat AD for thousands of years [24–26]. Chinese herbal medicine has a basis in accumulated clinical experience and has resulted in several herbal formulas to treat AD, including Xiao Feng San, Xiaofeng Daochi Tang, Longdan Xiegan Tang, and Yangxue Dingfeng Tang [5, 8, 27]. Previous study has found that oral administration of Xiao Feng San significantly suppresses desquamation and lichenification of the skin [28]. Another study has shown that a formula containing Flos Lonicerae (Jin Yin Hua), Herba Menthae (Bo He), Cortex Moutan (Dan Pi), Rhizoma Atractylodis (Cang Zhu), and Cortex Phellodendri (Huang Bai) and used in clinical trials in children with AD can
improve quality of life and reduce topical steroid use in moderate to severe AD patients [27]. In addition, another herb has been reported that could improve symptoms in AD-like mice [29–31].

Previous studies have reported that DBT can reduce bleomycin-induced pulmonary fibrosis in rats by boosting matrix metalloproteinases, MMP-1, MMP-9, and TIMP-1 expression [14, 23]. DBT treatment in ovariectomized rats modulates levels of follicle-stimulating hormone and luteinizing hormone in serum, and it may improve estrogen levels in women after menopause [32]. Previously, we found that DBT could improve asthma symptoms in asthmatic mice, including attenuating eosinophil infiltration and airway hyperresponsiveness and decreasing Th2 cytokine production in bronchoalveolar lavage fluid [11]. In this study, we confirmed that DBT could reduce ear swelling in AD-like mice and improve skin redness and scaling and bleeding symptoms. The herbal formula also reduced eosinophil and mast cell infiltration in the skin and ear, suppressed IgE levels in serum, and improved Th2 cytokine in serum and spleen cells.

Previous findings have suggested that Th2 cells modulate IgE-dependent mast cell activation and degranulation in allergic AD [9, 33]. More evidence has shown that Th2 cells secreting excessive IL-4 will exacerbate the symptoms of AD [8]. IL-4 predominately induces B cell activation and releases

Figure 6: The effects of DBT on mast cell infiltration in skin tissue. Skin sections were stained with toluidine blue to analyze mast cell infiltration in normal control mice (a); sensitized control mice (b); 3 g/kg DBT (HD3, c) or 10 g/kg DBT (HD10, d) (100x magnification, and amplified graph is 200x); n = 8 mice per group for all groups. Arrows indicate mast cells. The mast cells were measured under 10–15 high-power fields (HPFs) (e). Data are presented as means ± SEM. *P < 0.05, **P < 0.01 versus sensitized control mice.
Figure 7: Effect of DBT on levels of serum IL-4 (a), TNF-α (b), serum antibodies IgE (c), IgG1 (d), and IgG2a (e). Serum was centrifuged and collected; then the cytokine and antibody levels were evaluated by ELISA. Data are presented as means ± SEM. *P < 0.05, **P < 0.01 versus sensitized control mice.
**Figure 8:** Effect of DBT on cytokine levels of spleen cells. The cells were stimulated with 2 μg/mL concanavalin A (Con A) for 2 days. The supernatants were collected and assayed the levels of IFN-γ (a), IL-4 (b), and IL-5 (c). Data are presented as means ± SEM. *P < 0.05, **P < 0.01 versus sensitized control mice.

In atopic dermatitis, Th2 cells were leading to excessive IL-5 productions [8]. IL-5 stimulates bone marrow cells to produce more eosinophils, and the surfaces of differentiated eosinophils express CCR3, which binds to chemokines CCL11 and CCL24 to induce eosinophil migration and infiltration into the skin [34]. These activated eosinophils can release more eosinophil peroxidases and leukotrienes to cause skin inflammation. DBT did not promote or decrease CCL11 in serum (data not shown). Thus, it was an important finding that DBT attenuates IL-5 production in reducing eosinophil activation and migration into the skin in a mouse model of AD.

Proinflammatory cytokines also affect swelling and inflammation in AD-like skin [8]. We sought to extract mouse skin proteins and detect IL-6 and TNF-α by ELISA. However, those proinflammatory cytokines were not detected in our system (data not shown). Hence, we assayed serum and found that DBT could decrease TNF-α production to
ameliorate the inflammatory response. In this experiment, we also found that levels of IFN-γ also increased in spleen cell culture and that TNF-α and IFN-γ were mainly secreted by Th1 cells. Previous findings have suggested that atopic dermatitis is mainly Th1/Th2 imbalance [8, 10]. Dendritic cells of skin captured allergy to induce the overexpression of Th2 cells to secrete more the levels of IL-4 and IL-5. Those Th2 cytokines induce the infiltration of eosinophils and mast cells to release inflammatory mediators [10, 22]. Therefore, DBT inhibited the activity of Th1 cells, and Th1 cells also had a moderating effect in the AD-like mice. DBT was formed by Angelica sinensis and Astragalus membranaceus. Recent studies have found that topical application of Astragalus membranaceus also improve the symptom of AD in DNCB-induced mice [35]. We would investigate which compound of DBT is main effect to improve the symptom of AD in the future.

5. Conclusion

In conclusion, DBT reduces inflammatory symptoms in AD-like mice by suppressing the production of Th2-associated and proinflammatory cytokines to suppress eosinophil and mast cell infiltration.

Conflict of Interests

The authors declare that there are no competing interests.

Authors’ Contribution

Li-Wen Fang and Chao-Chun Cheng contributed equally to this paper.

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References

[1] T. Bieber, M. Cork, and S. Reitamo, “Atopic dermatitis: a candidate for disease-modifying strategy,” Allergy, vol. 67, no. 8, pp. 969–975, 2012.
[2] M. S. de Bruin Weller, A. C. Knulst, Y. Meijer, C. A. F. M. Bruinzeel-Koomen, and S. G. M. Pasmans, “Evaluation of the child with atopic dermatitis,” Clinical and Experimental Allergy, vol. 42, no. 3, pp. 352–362, 2012.
[3] Y. Tokura, “Extrinsic and intrinsic types of atopic dermatitis,” Journal of Dermatological Science, vol. 58, no. 1, pp. 1–7, 2010.
[4] R. Berke, A. Singh, and M. Guralnick, “Atopic dermatitis: an overview,” American Family Physician, vol. 86, no. 1, pp. 35–42, 2012.
[5] M. Boguniewicz and D. Y. M. Leung, “Atopic dermatitis: a disease of altered skin barrier and immune dysregulation,” Immunological Reviews, vol. 242, no. 1, pp. 233–246, 2011.
[6] S. Guglielmetti, J. K. Dart, and V. Calder, “Atopic keratoconjunctivitis and atopic dermatitis,” Current Opinion in Allergy and Clinical Immunology, vol. 10, no. 5, pp. 478–485, 2010.
[7] A. U. Tan and M. E. González, “Management of severe atopic dermatitis in children,” Journal of Drugs in Dermatology, vol. 11, no. 10, pp. 1158–1165, 2012.
[8] M. J. Turner, J. B. Travers, and M. H. Kaplan, “T helper cell subsets in the development of atopic dermatitis,” Journal of Drugs in Dermatology, vol. 11, no. 10, pp. 1174–1178, 2012.
[9] E. B. Brandt and U. Sivaprasad, “Th2 cytokines and atopic dermatitis,” Journal of Clinical & Cellular Immunology, vol. 2, no. 3, article 110, 2011.
[10] K. Eyriech and N. Novak, “Immunology of atopic eczema: overcoming the Th1/Th2 paradigm,” Allergy, vol. 68, no. 8, pp. 974–982, 2013.
[11] C.-C. Lin, C.-J. Liu, C.-Y. Chiang, W.-Y. Huang, and W.-C. Huang, “Danggui Buxue Tang attenuates eosinophil infiltration and airway hyperresponsiveness in asthmatic mice,” Annals of Allergy, Asthma & Immunology, vol. 107, no. 6, pp. 501–509, 2011.
[12] W. L. Zhang, K. Y. Z. Zheng, K. Y. Zhu et al., “Chemical and biological assessment of Angelica roots from different cultivated regions in a Chinese herbal decoction Danggu Buxue Tang,” Evidence-Based Complementary and Alternative Medicine, vol. 2013, Article ID 483286, 10 pages, 2013.
[13] J. Gao, Y. Huang, P. Li et al., “Antifibrosis effects of total glucosides of Danggui-Buxue-Tang in a rat model of bleomycin-induced pulmonary fibrosis,” Journal of Ethnopharmacology, vol. 136, no. 1, pp. 21–26, 2011.
[14] J. Lv, Z. Zhao, Y. Chen et al., “The Chinese herbal decoction dangui buxue tang inhibits angiogenesis in a rat model of liver fibrosis,” Evidence-Based Complementary and Alternative Medicine, vol. 2012, Article ID 284963, 11 pages, 2012.
[15] Y. Chen, Q. Chen, J. Lu, F. H. Li, Y. Y. Tao, and C. H. Liu, “Effects of Danggui Buxue decoction on lipid peroxidation and MMP-2/9 activities of fibrotic liver in rats,” Chinese Journal of Integrative Medicine, vol. 15, no. 6, pp. 435–441, 2009.
[16] C. C. Wang, K. F. Cheng, W. M. Lo et al., “A randomized, double-blind, multiple-dose escalation study of a Chinese herbal medicine preparation (Dang Gui Buxue Tang) for moderate to severe menopausal symptoms and quality of life in post-menopausal women,” Menopause, vol. 20, no. 2, pp. 223–231, 2013.
[17] C.-C. Chan, C.-J. Liou, P.-Y. Xu et al., “Effect of dehydroepiandrosterone on atopic dermatitis-like skin lesions induced by 1-chloro-2,4-dinitrobenzene in mouse,” Journal of Dermatological Science, vol. 72, no. 2, pp. 149–157, 2013.
[18] W.-C. Huang, M.-L. Kuo, M.-L Li, R.-C. Yang, C.-J. Liou, and J.-J. Shen, “Gynostemma pentaphyllum decreases allergic reactions in a murine asthmatic model,” The American Journal of Chinese Medicine, vol. 36, no. 3, pp. 579–592, 2008.
[19] K. B. Yarbrough, K. J. Neuhaus, and E. L. Simpson, “The effects of treatment on itch in atopic dermatitis,” Dermatologic Therapy, vol. 26, no. 2, pp. 110–119, 2013.
[20] A. Torrelo, J. Ortiz, A. Alomar, S. Ros, M. Prieto, and J. Cuervo, “Atopic dermatitis: impact on quality of life and patients’ attitudes toward its management,” European Journal of Dermatology, vol. 22, no. 1, pp. 97–105, 2012.
[21] M. S. de Bruin Weller, H. Rockmann, A. C. Knulst, and C. A. F. M. Bruinzeel-Koomen, “Evaluation of the adult patient with atopic dermatitis,” Clinical and Experimental Allergy, vol. 43, no. 3, pp. 279–291, 2013.
[22] E. L. Simpson, “Atopic dermatitis: a review of topical treatment options,” Current Medical Research and Opinion, vol. 26, no. 3, pp. 633–640, 2010.
[23] M. Niebuhr, H. Scharonow, M. Gathmann, D. Mamerow, and T. Werfel, “Staphylococcal exotoxins are strong inducers of IL-22: a potential role in atopic dermatitis,” The Journal of Allergy and Clinical Immunology, vol. 126, no. 6, pp. 1176.e1–1183.e1, 2010.
[24] H. Y. Tan, A. L. Zhang, D. Chen, C. C. Xue, and G. B. Lenon, “Chinese herbal medicine for atopic dermatitis: a systematic review,” Journal of the American Academy of Dermatology, vol. 69, no. 2, pp. 295–304, 2013.
[25] H.-H. Kim, J.-S. Yoo, T.-Y. Shin, and S.-H. Kim, “Aqueous extract of Mosla chinensis inhibits mast cell-mediated allergic inflammation,” The American Journal of Chinese Medicine, vol. 40, no. 6, pp. 1257–1270, 2012.
[26] K.-L. E. Hon, T.-F. Leung, Y. Wong et al., “A pentaherbs capsule as a treatment option for atopic dermatitis in children: an open-labeled case series,” The American Journal of Chinese Medicine, vol. 32, no. 6, pp. 941–950, 2004.
[27] K. L. Hon, B. C. Chan, and P. C. Leung, “Chinese herbal medicine research in eczema treatment,” Chinese Medicine, vol. 6, article 17, 2011.
[28] H.-M. Cheng, L.-C. Chiang, Y.-M. Jan, G.-W. Chen, and T.-C. Li, “The efficacy and safety of a Chinese herbal product (Xiao-Feng-San) for the treatment of refractory atopic dermatitis: a randomized, double-blind, placebo-controlled trial,” International Archives of Allergy and Immunology, vol. 155, no. 2, pp. 141–148, 2011.
[29] E. Cho and S. H. Cho, “Effects of Korean red ginseng extract on the prevention of atopic dermatitis and its mechanism on early lesions in a murine model,” Journal of Ethnopharmacology, vol. 145, no. 1, pp. 294–302, 2013.
[30] J. S. Hwang, J.-E. Kim, H.-T. Kim, Y.-B. Yu, and S.-H. Im, “Topical application of Taglisodog-eum inhibits the development of experimental atopic dermatitis,” Journal of Ethnopharmacology, vol. 145, no. 2, pp. 536–546, 2013.
[31] D.-Y. Kim, J.-A. Jung, T.-H. Kim, S.-W. Seo, S.-K. Jung, and C.-S. Park, “Oral administration of Uncariae rhynchophylla inhibits the development of DNFB-induced atopic dermatitis-like skin lesions via IFN-gamma down-regulation in NC/Nga mice,” Journal of Ethnopharmacology, vol. 122, no. 3, pp. 567–572, 2009.
[32] Q.-F. Xie, J.-H. Xie, T. T. X. Dong et al., “Effect of a derived herbal recipe from an ancient Chinese formula, Danggui Buxue Tang, on ovariectomized rats,” Journal of Ethnopharmacology, vol. 144, no. 3, pp. 567–575, 2012.
[33] A. J. Frew, “Allergen immunotherapy,” The Journal of Allergy and Clinical Immunology, vol. 125, supplement 2, pp. S306–S313, 2010.
[34] M. E. Rothenberg and S. P. Hogan, “The eosinophil,” Annual Review of Immunology, vol. 24, pp. 147–174, 2006.
[35] J. H. Kim, M. H. Kim, G. Yang, Y. Huh, S.-H. Kim, and W. M. Yang, “Effects of topical application of Astragalus membranaceus on allergic dermatitis,” Immunopharmacology and Immunotoxicology, vol. 35, no. 1, pp. 151–156, 2013.