Comparison of Different Bleaching Protocols Efficacy on Artificially Tea-Stained Teeth up to 24 Months Post-Treatment

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Abstract

Objective: In vitro investigation of four bleaching protocols efficiency, on artificially tea-stained human teeth, up to 24-month post-treatment.

Methods: 100 intact, extracted, human incisors were randomly divided (n=4x25/group) and underwent black tea staining. The bleaching protocols applied per group were: 1. (BH, at-home) 10%CaP for 5 days (120min/day), repeated twice, 2. (BO1, in-office) 40%HP for 3 successive applications (15 min/application), repeated twice, 3. (BO1, in-office) 40%HP for 3 successive applications (15 min/application), repeated 3 times and 4. (BH+BO1, at-home/in-office) starting with BH, interval of 2 days, followed by BO1. The color was measured before bleaching(t0), right after staining(t1), after each bleaching procedure(t2), 3(t3), 6(t4) and 24 months(t5) after treatment, with a colorimeter (Braive Instruments) in CIEL*a*b*system. ΔΕ* values were calculated relative to t0. Statistical analysis was performed by Mann-Whitney U and Friedman and Wilcoxon Signed Ranks tests (α=0.01).

Results: All bleaching protocols, at t2 induced increase in L*, return of a* to before-staining values, shifting of b* to blue (BO1, BO2) and improvement of tooth color (ΔΕ*) with no statistically significant difference among protocols. All color coordinates presented gradual rebound at t3. Regarding ΔΕ*, tooth color reached after-staining values for BH, BO1, BO2 up to 24 months, while bleaching effect was partially preserved for BH+BO1 group.

Conclusion: At-home and in-office bleaching protocols provided similar whitening effect on tea-stained teeth, immediate after application. However, rebound effect started at 3 and progressed up to 24 months post-treatment. The combination of at-home and in-office treatment showed relative better potential in bleaching color preservation.

Keywords: Tooth bleaching agents; Tooth discolorations; Carbamide peroxide; Hydrogen peroxide

Introduction

The percentage of patients who seek improvement of their smile's appearance by tooth color change increases exponentially. As a result, a wide range of bleaching procedures has been recommended for tooth whitening [1]. Numerous randomized clinical trials evaluate the bleaching efficacy on teeth which fall into the natural tooth color range. However, due to the high number of variables contributed to the bleaching techniques (e.g. chemical agents and concentrations, different application periods, number of sessions, time intervals between bleaching sessions, at-home or in-office procedure) a clear conclusion cannot be drawn in terms of which protocol induces the most reliable and long-lasting color change [2-5]. Some studies report that at-home bleaching provides better and more stable whitening effect than in-office protocols, others support the opposite, while similar results for both techniques have been also published [3,6,7]. Even a systematic review could not answer the relative questions that exist [8].

The aforementioned data are pertain to the bleaching of teeth with natural colors. Teeth with discoloration due to wine, coffee, tea, cola consumption or smoke, present an additional challenge. It should be highlighted that any bleaching impact seen in the natural tooth color may not be obtained for discolored teeth, because the latter one's present different reactions to the bleaching protocols [9]. Clinical trials with respect to color change
in discolored teeth after bleaching are rare and these do not lead to certain conclusions [10,11]. The majority of in vitro studies, which investigate the relation between extrinsic discoloration and bleaching, are focused on immediate and not on long-term effects of various bleaching treatments, using bovine and not human teeth [12-17]. However, in general, the color rebound through the post-bleaching period is considered as one of the most serious problems [18-20]. Therefore, the long-lasting color change provided by a bleaching protocol offers a considerable advantage. In addition, although bovine teeth or their enamel slabs are often utilized in experimental studies and show similarities with the human teeth, it is under consideration if these are reliable substitutes of the human teeth [21]. Furthermore, to the best of our knowledge, the in vitro studies conducted for stained teeth compare different single in-office or at-home bleaching protocols or compare the first versus the second technique [22,23]. Thus, examination of a combination of an in-office and at-home protocol for bleaching treatment emerges.

Thus, the objective of this study was to in vitro compare the efficiency of in-office, at-home and in-office followed by at-home bleaching protocols on artificially tea-stained human teeth for a period up to 24-month post-treatment. The null hypothesis was that no differences exist among the protocols in terms of color changes induced immediate after bleaching and that there is no statistically significant color reverse of stained bleached teeth through time regardless of the bleaching protocols. Lightness that gradually blends with the surrounding tooth color. The color improvement seemed to be more noticeable in orange/brown discolorations compared to the dark brown/black ones. The different response of stains to bleaching agents, might be explained by the different stain history of each tooth (type, depth); therefore, the bleaching agent may not be as efficacious in different stained lesions.

Materials and Methods

The ethics committee of Athens dental school approved this in vitro trial.

The bleaching products used in this study are shown in Table 1.

| Materials | Serial Number | Composition |
|-----------|---------------|-------------|
| Opalescence PF (Ultradent, 505 West Ultradent Drive South Jordan, USA) (at-home agent) | (474338,2014-09) (B7L44,2014-10) | 10% carbamide peroxide, potassium nitrate and fluoride |
| Opalescence Boost (Ultradent, 505 West Ultradent Drive South Jordan, USA) (in-office agent) | (B7L1X,2014-03) (B7QNX,2014-05) | 40% hydrogen peroxide, potassium nitrate and fluoride |

Sample preparation

One hundred intact human central and lateral incisors, extracted for periodontal reasons, were selected. After extraction, each tooth was cleaned with a polishing brush (Kerr Dental) and paste (Clean Polish, Kerr), which was then thoroughly removed with air-water spray and kept in a thymol solution (0.02% thymol in saline) at room temperature (22°C). Then, the teeth were numbered at the root surface with a permanent black marker. The numbers were then covered with a clear nail polish. For staining, all teeth were immersed in a black tea solution (M&S Fair Trade Earl Grey), at room temperature (22±2°C) for 7 days, which was renewed daily. The solution was prepared by boiling 2gr black tea in 100ml of distilled water, for 5 minutes. After staining, teeth were randomly allocated into four groups (n=25). Four different bleaching protocols per group were applied, [BH: bleaching at-home, B01: bleaching in-office simulating 2 visits, B02 bleaching in-office simulating 3 visits, BH+B01: combination of at-home and in-office bleaching] (Table 2).

| Group | Bleaching Material & (Group) | Protocol |
|-------|-----------------------------|----------|
| (BH)  | Opalescence PF (BH)         | Application of the agent for 5 days continuously (120min/day) with trays followed by two days without application. This sequence was repeated two (2) times more (at-home procedure). |
| (B01) | Opalescence Boost (B01)     | Three (3) successive applications of the agent, directly on the teeth, for 15 minutes per session/week. This sequence was repeated for two (2) consecutive weeks (in-office procedure). |
| (B02) | Opalescence Boost (B02)     | Three (3) successive applications of the agent, directly on the teeth, for 15 minutes per session/week. This sequence was repeated for three (3) weeks, with an interval of one week without application between the sessions/weeks (in-office procedure). |
| (BH+B01) | Opalescence PF + Opalescence Boost (BH+B01) | First, the protocol of group BH is followed and then after an interval of two (2) days, the protocol of group B01 is applied (at-home + in-office procedures). |
In groups 2 and 3 the bleaching agent was directly applied on tooth surfaces while in groups 1 and 4 through custom made trays. For tray preparation, the teeth were placed in jaw models (Frasaco GmbH, Tettnang, Germany) using a VPS impression material (Variotime, Heraeus/Kulzer). Following, custom made trays with reservoirs were manufactured out of these jaw models, over plaster casts, after taking alginate impressions (Cavex). The reservoirs were made with a light-cured resin (LC Block-Out Resin, Ultradent) formed in a layer of 1 mm over the labial surface. A groove along the tooth neck (to prevent leakage of material during the splint usage) was also formed, with a fine round bur (005 SKU: CB1 of Drendel & Zweiling). The trays were formed with a thermoplastic material (Soft-Tray Sheets 0.9mm, 5”x5”, Ultradent). The limits of the trays were placed 1 mm below the anatomical neck of the teeth. At the intervals, during and after the completion of bleaching cycles, the teeth of all groups were kept in artificial saliva solution (Saliva Orthana, Luboran -AS Pharma, 24 (t5) months after treatment. Before each color measuring, the teeth were rinsed with spray-water for 5 seconds and dried with air for 5 seconds. Each recording was taken from the middle third of the labial surface of the teeth. A customized silicone matrix was used to achieve measurements from the same tooth position while this was placed in the colorimeter. Color changes (ΔΕ*), were calculated at each time recording relative to the measurement before bleaching (t0), based on the equation

\[ \Delta E* = \sqrt{(\Delta L*)^2 + (\Delta a*)^2 + (\Delta b*)^2} \]

**Statistical analysis**

After applying normality tests, dependent variables (L*, a*, b* and ΔΕ*) were not normally distributed, thus non-parametric tests (Mann-Whitney U and Friedman test) were used to investigate possible associations between the independent (groups and time) and the outcome variables. Also, a post-hoc analysis of the results using Wilcoxon Signed Ranks test was performed. Statistical analysis was performed using the program SPSS, IBM v21 and significance level was set at 1% (p-value= 0.01) to reduce Type I (by chance) error as much as possible.

**Results**

The statistical power of the sample was very satisfactory (> 0.95), with a strong impact effect (effect size=0.87). The values of color parameters L*, a* and b* for the four groups, at all-time measurements are presented in Tables 3 & 4, accordingly and for ΔΕ* in Table 5. Tea staining caused statistically significant decrease in L*, increase in a* and no effect in b* coordinate. Statistically higher L* parameters were obtained right after bleaching (t2) relative to the values recorded after staining (t1), in all groups. Concerning L* at t3 and t4 assessments, statistically decrease was observed relative to the values obtained immediately after bleaching application (t2). At 24 months post-bleaching period (t5), statistically significant reduction in L* compared to the immediate after bleaching values (t2) was observed for BH and BH+BO1 protocols, whereas BO1 and BO2 subjected no statistical reduction. In respect of a*, the four bleaching protocols provided lower values relative to the staining analogues. At three (t3) and six (t4) months recordings, a gradual increase in a* values were detected. At the final measurement (t5), a* values in all groups were statistically higher than the after bleaching (t2) and the staining (t1) correspondence ones.

**Table 3:** Mean values (first line) and standard deviations (second line) of the L*, a* parameters for the bleaching protocols tested at all time measurements*.

| Group  | t0       | t1       | t2       | t3       | t4       | t5       | t0       | t1       | t2       | t3       | t4       | t5       |
|--------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|----------|
| (BH)   | 61.67 ± 3.14 | 59.29 ± 3.52 | 49.89 ± 3.04 | 47.26 ± 3.95 | 54.52 ± 3.85 | 5.40 ± 2.29 | 7.40 ± 2.45 | 4.62 ± 2.06 | 7.56 ± 2.73 | 8.08 ± 2.38 | 11.9 ± 2.14 |
| (BO1)  | 59.0 ± 3.07   | 49.38 ± 3.86 | 50.24 ± 4.58 | 49.12 ± 4.40 | 56.67 ± 4.76 | 5.024 ± 2.47 | 8.05 ± 2.18 | 6.13 ± 2.61 | 7.26 ± 2.72 | 7.56 ± 2.46 | 12.77 ± 2.46 |
| (BO2)  | 63.33 ± 3.65   | 51.76 ± 3.54 | 50.78 ± 3.94 | 57.19 ± 4.36 | 4.036 ± 2.61 | 8.20 ± 2.14 | 6.66 ± 2.45 | 7.95 ± 2.71 | 8.90 ± 2.84 | 11.66 ± 2.14 |
| (BH+BO1)| 63.30 ± 3.49 | 50.16 ± 3.54 | 55.74 ± 3.26 | 6.48 ± 2.58 | 12.10 ± 4.57 | 6.03 ± 2.32 | 8.91 ± 2.63 | 7.95 ± 2.72 | 12.98 ± 2.67 |

*Same numbers within the rows for each parameter (L*, a*) indicate statistically significant differences between each group/row (Friedman test and post-hoc Wilcoxon Signed Ranks test, p<0.01).

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Table 4: Mean values (first line) and ± standard deviations (second line) of the b* parameter for bleaching protocols tested at all time measurements*.

| Group     | t₂   | t₃   | t₄   | t₅   |
|-----------|------|------|------|------|
| (BH)      | 7.35±1.82 | 7.94±1.73 | 8.08±1.75 | 6.62±1.45 | 5.63±1.61 | 8.36±1.47 |
| (B01)     | 5.56±1.98 | 6.10±1.48 | 4.53±1.52 | 5.47±1.67 | 4.81±1.87 | 7.06±2.07 |
| (B02)     | 7.02±2.13 | 6.85±2.20 | 5.61±2.23 | 5.53±2.10 | 5.78±1.94 | 8.38±4.11 |
| (BH+B01)  | 6.69±2.18 | 7.92±2.14 | 5.52±2.30 | 5.77±2.15 | 5.38±2.48 | 7.66±2.11 |

*Same numbers within the rows indicate statistically significant differences between time points for each group/row (Friedman test and post-hoc Wilcoxon Signed Ranks test, p<0.01). Same letters within the columns indicate statistically significant differences among the Groups (1, 2, 3, 4) for each time point/column (to, t₁, t₂, t₃, t₄) (Mann-Whitney U test, p<0.01). No letters are noted because no differences were revealed.

Table 5: Mean values and ± standard deviations of the ΔE* parameter for the bleaching protocols tested at all time measurements*.

| Group     | t₁   | t₂   | t₃   | t₄   | t₅   |
|-----------|------|------|------|------|------|
| (BH)      | 9.53±5.61 | 5.20±4.05 | 12.59±6.43 | 15.22±3.37 | 11.11±6.37 |
| (B01)     | 10.36±4.27 | 5.87±5.04 | 9.48±3.17 | 10.77±3.37 | 9.17±4.27 |
| (B02)     | 11.67±5.93 | 7.31±4.66 | 11.19±6.33 | 12.43±5.94 | 10.53±4.75 |
| (BH+B01)  | 12.41±5.31 | 4.88±2.89 | 9.84±4.55 | 11.71±4.61 | 9.61±3.61 |

* Same numbers within the rows indicate statistically significant differences between time points for each group/row (Friedman test and post-hoc Wilcoxon Signed Ranks test, p<0.01). Same numbers within the rows indicate statistically significant differences between time points for each group/row (Friedman test and post-hoc Wilcoxon Signed Ranks test, p<0.01). No letters are noted because no differences were revealed.

Bleaching procedures statistically decreased b* parameters compared to the staining ones, except in BH group. No statistically significant changes were noticed in all groups through up to six months post-treatment (t3 and t4) relative to their bleaching values recorded (t2). After storage for 24 months, b* presented higher values (t5 vs. t2) in B01, B02 and BH+B01 protocols whereas no difference observed for BH. After bleaching (t2), the four protocols presented statistically significantly lower ΔE* values compared to the after staining (t1) corresponding data. Through post-bleaching time (t3, t4, t5), ΔE* values were increased relative to the immediate after bleaching recordings (t2), reaching statistically significantly higher values for all protocols. Furthermore, the values had no statistically significant difference relative to the staining ones with the exception of BH+B01 treatment which was kept lower. No significant differences concerning ΔE* were noticed among the bleaching treatments applied at all time intervals.

Discussion

In general, intrinsic and extrinsic dyes influence the natural color of teeth. Regarding the extrinsic dyes, tobacco, dietary intakes, mouth rinses, metal salts, medication function as colorings [24,25]. Tea is one of the most popular drinks and at the same time, it is considered as a possible extrinsic staining of the dentition, mostly because of the tannin which is a polyphenol macromolecule [26-28]. It is thought that the staining caused by tea is external and not internal because long length organic chains can hardly penetrate through enamel to reach the dentin [29]. Deposition of tannins on the outer tooth surface, which may bind to the enamel hydroxyapatite is believed to further promote tooth discoloration [30]. Macromolecules and dyes absorbed onto tooth surface, decrease the light reflectance and as a result, the teeth turn darker [31]. Bleaching technique, apart from natural tooth color on the oxidative reaction induced by OH and OOH radicals - that are released from hydrogen or carbamide peroxide - and the tooth chromophore molecules. The latter ones are broken to smaller molecules, resulting in changing of the tooth optical properties [32]. This reaction is complex, so it has not clearly documented, up to now, how this whitening process functions [33]. In addition, the bleaching response seems to be differently influenced by each particular colorant, because their composition is not similar [34,35]. Therefore, the response of each stain to the various bleaching protocols is not predictable.
Experimental models, which simulate clinical conditions, have been designed through the years to study the efficiency of whitening methods on stained teeth. The most commonly used stains are coffee, red wine, tea, blood and chlorhexidine [16,35-38].

The experimental tea-staining procedure followed in the current investigation is an accelerated approach, which has been applied by Kielbassa et al. [39]. In this study and concerning the teeth used, those of human origin were preferred than bovine ones. Although Attia et al. [15] reported that bovine and human dental substrates behave similarly during the bleaching process, the staining of human teeth and their bleaching implementation provide a better depiction of clinically relevant conditions. Furthermore, only incisor teeth were used for morphological similarity reasons among the samples and for convenience in color measurements by spectrometer due to their large extended flat buccal surface. Under the conditions of the current study, the storage media of the specimens played a significant role because the samples should be kept up to 2 years for the sequence of color recordings. Therefore, artificial saliva was employed instead of any other agent. It is established that mineral salts lost during bleaching can be replaced by the natural saliva [40,41]. The immediate after bleaching outcome is influenced by dehydration, therefore, one-week post-recording is commonly conducted to assign the validity of each bleaching protocol [18,42]. This specific period of measurement was skipped in the current study because the long-term color stability and not the immediate bleaching effect was the main objective.

In laboratory and clinical bleaching studies, commercially available shade color guides (e.g. Vita Classical, Vita 3D Master) are commonly utilized for color recording. As far as the present study is concerned, a colorimeter was chosen because shade guides are associated with serious drawbacks [43,44]. On the contrary, colorimeters present low risk of error and high rates of accuracy and repeatability between measurements [23,45,46]. Generally, for tooth color shade, the variation of L* is more significant, followed by that of b* and finally that of a* [47,48]. Moreover, L* values, which depict the lightness/darkness appears to be the most relevant parameter to make comparisons under experimental conditions [45]. Lower L* values, obtained in the current study after tea staining, indicate tooth darkening. The a* value changes were in the direction of redness (higher values) and not greens, results which are in agreement with Lee et al. [49]. The polyphenols contained in black tea are the a-flavins, which have red color and are mostly related with the staining capacity of tea [50]. The tea-flavins are another kind of polyphenol compound found in black tea, and although these present a yellow color, significant shifting to this direction (b*) was not detected in the current study. On the contrary, change of b* values to yellow axis was observed in Sulieman et al. study [16]. Differences in experimental conditions between the two investigations probably cause the controversial results. In Sulieman et al. [16] study, tea staining was performed through cut dentin and not only through enamel. Moreover, the different concentrations of the a-flavins in several tea brands used and the different immersed time chosen could be some additional contributor factors.

According to the findings of this study, the null hypothesis was confirmed concerning the absence of difference in whitening effect among the four bleaching protocols after the end of the procedure and it was rejected regarding the color preservation through post-treatment time. Bleaching, regardless of the methods used in the current study, considerably increased lightness (L*) immediately after the application period and L* was at first reduced after tea staining of the teeth. Changes in L* values, individually, cannot determine the protocol with the best whitening performance because each group presented different initial (t0) and after staining (t1) L* values. However, the percentage difference per group (t1-t2/t1) can function as an indicator. The relative values were as follows: BH (11.7%), BO1 (10.5%), BO2 (14%) and BH+BO1 (25.8%) L* increase. It is obvious that the combined at-home/in-office technique is appeared as the most intense bleaching procedure. However, for all groups, L* started to drop down already after 3 months post-bleaching, with a different rate for each protocol. Specifically, although the two in-office techniques presented L* rebound, these were proved more efficient to preserve the whitening effect than the at-home and the at-home/in-office techniques.

Regarding a*, after bleaching, the values reversed to the initial ones (before staining) per group. After bleaching, through time, gradual rebound was observed relative to the after control, regardless the group. At 24-month assessment, higher values were recorded than those of control for each stained group. It is rather difficult to explain the phenomenon. The aging of the end products of the oxidative reaction as well as, the remaining tea chromophore molecules may provide shifting to the green direction. Although tea-staining had no influence on all b* coordinates, bleaching decreased the correspondence b* values per group except of BH, leading to blueness. Through time, b* presented graduate increase up to values higher than the after bleaching and higher than those of the natural teeth (initial values) except of BH. Although, generally, the higher the ΔE* difference, the greater the color change, in clinical point of view, the calculation of ΔE* by itself has no practical significance. It should be combined with the clinical perception threshold (PT: perceptibility threshold), which stands for the color difference that is clinically distinct from/to the 50% of the observers [51]. In this study, the recommended value of 2.7 for the PT was used for comparison [52,53]. According to our results, ΔE* values obtained either through the tea-staining process or bleaching were higher than the limit of 2.7 value. Therefore, in any combination between time intervals, and for all groups the color difference recorded was clinically distinct from/to the 50% of the observers.

ΔE* shows the magnitude of the change and not the direction of the change in the three coordinates of the CIELab system.
Bleaching, upon ending of the procedure, improved color (ΔΕ) relative to stained analogues, with no difference among the protocols. Opalescence PF (BH group) based on 10% carbamide peroxide is considered as milder bleaching agent than Opalescence Boost, which contains 40% hydrogen peroxide (BO1 and BO2 groups). Therefore, it should expect that Opalescence Boost could obtain stronger immediate color change [54,55]. On the other hand, generally in bleaching protocols, it is not clear if higher concentrations provide better whitening results or a combination of application time and agent influences the bleaching effect [56-58]. The significantly longer application time of Opalescence PF (1.800 min) compared to Opalescence Boost (90 for BO1 and 135 min for BO2) may be associated with the absence of difference concerning the whitening effect of the protocols. Therefore, time may be the dominant variable for color change for the tea-stained teeth and not the concentration of the bleaching agent.

The additional application of a strong agent after at-home procedure (BH+BO1) did not show the potential for further whitening outcome. As far as, the majority of chromophore molecules have been broken by oxygen within the first phase of the protocol (BH), the influence of the second phase (BO1) may be negligible. However, it is interesting that this protocol achieved more successful long-lasting effect than all the others applied. Indeed, the bleaching effect was short-lasting for all the protocols. The teeth started to recover their color three months after the end of the treatment. After twenty-four months, their color reached that received after staining, except of those treated by at-home/ in-office procedure, which preserved percentage of the bleaching effect. Although the bleaching effect on stained teeth has been investigated in some studies, serious differences are identified in comparison with the experimental conditions followed by this study [12-17,39]. Type and preparation of samples exposed dental tissue, staining type and procedure, color recording method, bleaching protocols, follow-up period etc. are some of the differences that can be mentioned. The wide diversity of the factors involved causes great difficulties for comparison of our findings with those of previous studies.

**Conclusion**

Within the limitations of this study the following conclusions can be derived:

a) Artificially tea-staining led to decrease in L* values of teeth, shifting of a* to redness and no changes in b* values.

b) All bleaching protocols, immediate after completion of the procedure, induced: a) improvement in lightness (L*) ranged from 11,7% for at-home protocol to 25,8% for the combined at-home/in-office one, b) reverse of a* values to before staining status, c) shifting of b* values to blue direction for in-office and at-home methods and, d) improvement of tooth color (ΔΕ) with no difference among the protocols.

c) Through post-treatment period, L*, a* and b* coordinates presented gradual rebound starting after three (3) up to twenty-four (24) months.

d) Within 24-month post-treatment, the tooth color (ΔΕ*) reached the after-staining values for in-office and at-home protocols. Bleaching effect was partially preserved only for at-home/in-office procedure.

e) All ΔΕ values obtained either through the tea-staining or the bleaching process were clinically distinct from the 50% of the observers (>2.7).

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