FOXM1 predicts disease progression in non-muscle invasive bladder cancer

Sebastien Rinaldetti1,3 · Ralph Wirtz2 · Thomas Stefan Worst3 · Arndt Hartmann4 · Johannes Breyer5 · Lars Dyrskjot6,7 · Philipp Erben3

Received: 31 May 2018 / Accepted: 19 June 2018 / Published online: 29 June 2018
© The Author(s) 2018

Abstract

Purpose The proto-oncogene forkhead box M1 (FOXM1) is associated with poor survival in many cancers. The impact of FOXM1 expression on progression-free survival (PFS) of non-muscle invasive bladder cancer (NMIBC) has not yet been investigated. The differential expression of FOXM1 between the different molecular NMIBC subtypes has further been assessed.

Methods Transcript levels of FOXM1 and MKI67 were determined in 460 NMIBC patients (UROMOL cohort) by RNA-Seq and validated in silico by the Chungbuk and Lund cohort (n = 277). FOXM1 and MKI67 cutoffs were identified by the minimal \( p \) value method. Variables were evaluated by multivariable Cox regression analyses in order to identify independent predictors.

Results FOXM1 is an independent predictor for PFS superior to current histological, clinical and molecular staging methods. Patients with high FOXM1 expression have a 6- to 8-fold higher risk of progression in multivariable analysis \( (p < 0.03) \). Highest transcript levels were found in the Class 2 and genomically unstable molecular NMIBC subtype \( (p < 0.03) \). The proto-oncogene further positively correlated with tumor grade and stage. NMIBCs with high FOXM1 expression showed a PFS advantage when treated with intravesical BCG instillation.

Conclusion FOXM1 is a highly prognostic marker for disease progression of NMIBC superior to current histological, clinical and molecular staging methods and MKI67. It is mainly expressed in the Class 2 and genomically unstable molecular bladder cancer subtypes. Its role in drug resistance development makes FOXM1 valuable biomarker for NMIBC risk stratification.

Keywords Molecular subtypes · FOXM1 · Progression-free survival · Biomarker · Bladder cancer · MKI67

Background

Non-muscle invasive tumors represent 75% of patients diagnosed with transitional cell carcinoma. Transurethral resection of bladder tumors (TURB), intravesical mitomycin C and bacillus Calmette–Guerin (BCG) instillation represent...
the current standard treatments for non-muscle invasive bladder cancer (NMIBC) (Babjuk et al. 2017). As 50–80% of pTa NMIBC have cancer recurrence and 10–30% of pT1 and CIS patients show disease progression, clinicopathologic parameters are insufficient for disease prediction (Prout et al. 1992; Ark et al. 2014). Given the biologic heterogeneity of bladder cancer, survival and progression varies even within the same stage. Thus molecular biomarkers are needed to improve prediction of treatment response or even for clinical decision-making in the sense of preemptive biomarkers (Youssef and Lotan 2011; Kluth et al. 2015). Based on recent data, the most promising results were provided by gene expression signatures of TURB samples and by liquid biopsies from urine or blood (Contreras-Sanz et al. 2017; Robertson et al. 2017). The elucidation of valuable invasive or non-invasive biomarkers or drug targets is still at its beginning.

The role of the proto-oncogene forkhead-box M1 (FOXM1) in carcinogenesis and drug resistance development is already well established and has been validated in many cancer types (Dai et al. 2015). FOXM1 originates from the forkhead gene family, was first identified in Drosophila and is characterized by a conserved 100-amino acid DNA-binding motif. It is involved as regulator in embryogenesis and numerous developmental processes (Ye et al. 1997). In adult organs, FOXM1 is mainly expressed in proliferating cells or induced by growth factor release. In this context, it is not surprising that FOXM1 serves as proto-oncogene in most cancers and aberrant expression or mutations constitute the origin of many treatment resistance mechanisms (Kwok et al. 2010; Kalin et al. 2011; Saba et al. 2016). Although FOXM1 is exclusively expressed in dividing cells, its targeting may result in many side effects given its involvement in angiogenesis, cell migration and epithelial–mesenchymal transition (Halasi and Gartel 2013). We have recently shown that FOXM1 is a predictor for overall and disease-specific survival in muscle invasive bladder cancer (MIBC) superior to the TNM staging system and MKI67 (Rinaldetti et al. 2017). A recent TCGA study further underlined the role of FOXM1 as regulator in MIBC (Robertson et al. 2017). As MKI67 is considered as the gold standard biomarker for proliferation and prognosis, the impact of FOXM1 needs yet again to be compared with the later (Rodríguez-Alonso et al. 2002).

Recent findings showed that bladder cancer can be subclassified in molecular subtypes with some similarities to breast cancer subtypes (Choi et al. 2014; Hedegaard et al. 2016; Robertson et al. 2017). These findings open the doors for personalized treatment concepts similar to those in breast cancer. That is why the subtype-specific expression of FOXM1 will be analyzed in this study. As the FOXM1 signaling network represents a valuable and promising target for further cancer treatment personalization, we here investigate its clinical impact in three cohorts with a total of 737 NMIBC patients.

**Methods**

### Clinicopathologic characteristics

Patients and clinicopathologic data of 460 NMIBC patients from a European multicenter prospective study (UROMOL cohort) were investigated (Hedegaard et al. 2016). All samples have a carcinoma cell percentage > 50. Clinicopathologic characteristics are summarized in Tables 1 and 2. Sample collection procedures were published before (Hedegaard et al. 2016). All patients gave informed consent and the study was approved in all countries by institutional review boards or ethical committees. Expression data and clinicopathologic information from the Chungbuk (n = 104, GSE13507) and Lund (n = 173, GSE32894) cohort were used for validation (Kim et al. 2010; Sjödahl et al. 2012). All MIBC (T2–T4) patients were excluded.

### Gene expression analyses

Gene expression analyses of FOXM1 and MKI67 are based on paired-end RNA-Seq (101 + 7 + 101 bp) analysis on an Illumina HiSeq 2000. Data were normalized as described before and log2 transformed. Cluster assignments were performed by ConsensusClusterPlus using the programming

| Table 1 Clinicopathologic characteristics of the UROMOL cohort | Median value (range) or absolute value (%) |
|---------------------------------------------------------------|-------------------------------------------|
| Age                                                          | 69 (23–96)                                 |
| Male                                                         | 357 (78%)                                  |
| Grade                                                        |                                           |
| High grade                                                   | 176 (38%)                                  |
| Low grade                                                    | 277 (60%)                                  |
| PUNLMP                                                       | 7 (2%)                                     |
| Stage                                                        |                                           |
| CIS                                                          | 3 (1%)                                     |
| Ta                                                           | 345 (75%)                                  |
| T1                                                           | 112 (24%)                                  |
| Growth pattern                                               |                                           |
| Papillary                                                    | 414 (97%)                                  |
| Other (mixed, solid, unknown)                                | 11 (3%)                                    |
| BCG treatment                                                | 88 (20%)                                   |
| FOXM1 expression                                             | 1.67 (0–5.7)                               |
| MKI67 expression                                             | 2.74 (0.26–6.24)                           |
| Follow-up duration (months)                                  | 33 (0–75)                                  |
software R (version 3.2.2, R Foundation for Statistical Computing, Vienna, Austria) (Hedegaard et al. 2016).

Expression data of the Chungbuk and Lund cohort were based on Illumina human-6 v2.0 expression beadchip and Affymetrix Human Gene 1.0 ST Array analyses, respectively. Processed gene expression data, as used in the respective studies, were downloaded from the Gene Expression Omnibus database (Kim et al. 2010; Sjödahl et al. 2012).

Statistical analyses

Cutoffs for FOXM1 and MKI67 were identified by the minimum p value method in order to stratify patients in a high- and low-risk group (Budczies et al. 2012). The hazard ratio of the cutoffs was estimated by multivariable Cox regression analysis. The Wald forward algorithm was used for testing significance. The stepwise entry criterion for covariates was $p < 0.05$ and the removal criterion consisted in $p > 0.10$. Association between variables was analyzed using Kruskal–Wallis test, Mann–Whitney $U$ test or Spearman’s rank correlation. Kaplan–Meier estimates together with the log-rank test were used for survival analysis. In case the median survival rates could not be determined, they were specified as mean survival rates. The unadjusted significance level of 0.05 was considered for all statistical tests. Statistical analyses were performed using SPSS (version 20, IBM, Armonk, NY, USA).

Results

High correlation of FOXM1 with stage, grade and MKI67 expression

In both the UROMOL and Lund cohort FOXM1 was significantly higher in T1 vs Ta NMIBCs (Figs. 1a, 3, $p < 0.001$). Further, the expression correlated positively with tumor grade in the UROMOL, Chungbuk and Lund cohort (Figs. 1a, 2a, $p < 0.001$). The same tendency was observed for MKI67 (data not shown). This is in accordance with the high spearman correlation between FOXM1 and MKI67 of the UROMOL, Chungbuk and Lund cohorts (Spearman coefficient: 0.88 / 0.86 / 0.77, respectively, $p < 0.001$). The 12-gene risk score developed for predicting progression (Dyrskjøt et al. 2017) positively correlated with the FOXM1 expression ($p < 0.001$, Fig. 1a).

FOXM1 is superior in predicting PFS than MKI67 and TNM

The cutoff levels for FOXM1 and MKI67 have been optimized for progression-free survival (PFS). For the Lund cohort no cutoff could be determined as duration data for PFS were not available. Poor PFS correlated with high FOXM1 or MKI67 expression. Cutoff values for risk stratification were calculated by the minimal $p$ value method. A FOXM1 cutoff of 2.41 (median 1.66, 95% CI 1.72–1.91, Fig. 1c) and an MKI67 cutoff of 5.1 (median 1.67, 95% CI 1.72–1.91, figure not shown) have been determined for the UROMOL cohort. A FOXM1 cutoff of 8.27 (median 8.07, 95% CI 8.20–8.47) and a MKI67 cutoff of 8.04 (median 7.6, 95% CI 7.68–7.92, figure not shown) were chosen for the Chungbuk cohort (Fig. 2b, Kaplan–Meier plot for MKI67 not shown). In both the UROMOL and Chungbuk cohort, these FOXM1 and MKI67 cutoffs allowed a distinct risk stratification.

The molecular and clinicopathologic parameters of the UROMOL cohort, tested by univariable Cox regression analysis, are summarized in Table 3. The single variables retained by the multivariable Cox regression model as independent PFS predictors were age, stage and FOXM1 with a hazard ratio (HR) of 5.7 (95% CI 2.45–12.31, $p < 0.001$, Table 3). Indeed, the FOXM1 high-risk group showed a mean PFS of 57 months ($n = 114$, 95% CI 51.78–61.90) in contrast to the prolonged mean PFS of 73 months from the low-risk group ($n = 346$, 95% CI 72.08–74.35, $p < 0.001$, Fig. 1c).

The impact of FOXM1 has further been validated in silico by the independent Chungbuk cohort. The covariates age, grade, BCG instillation therapy, FOXM1 and

| Table 2 | Clinicopathologic characteristics of the Chungbuk cohort |
|---------|----------------------------------------------------------|
| Chungbuk cohort: clinicopathologic characteristics ($n = 104$) | Median value (range) or absolute value (%) |
| Age | 67 (24–88) |
| Male | 87 (84%) |
| Grade | |
| High grade | 18 (17%) |
| Low grade | 86 (83%) |
| Stage | |
| Ta | 24 (23%) |
| T1 | 80 (77%) |
| Intravesical therapy | 56 (54%) |
| FOXM1 expression | 8.08 (7.25–10.34) |
| MKI67 expression | 7.62 (6.92–9.66) |
| Follow-up duration (months) | 55 (2–137) |
MKI67 cutoff were included in the multivariable analysis (Table 4). However, the only covariate retained by the Cox regression model was the FOXM1 cutoff with an HR of 8.53 (95% CI 1.79–40.72, \( p = 0.007 \)). The FOXM1 high-risk (\( n = 44 \)) versus low-risk group (\( n = 60 \)) showed a mean PFS of 98 months (95% CI 80.78–116.12) versus 132 months (95% CI 125.68–138, Fig. 2b). The same cutoff was tested for overall survival. The FOXM1 high-risk vs low-risk group (\( n = 60 \)) showed a median OS of 70 months (95% CI 51.82–89.64) versus 135 months (95% CI 72.16–197.78, Fig. 2b).

**Aberrant expression of FOXM1 within molecular subtypes**

The NMIBC subtypes developed by the UROMOL cohort were described recently by Hedegaard et al. They subclassified the NMIBC into three molecular subtypes: Class 1 (luminal), Class 2 (luminal CIS-like) and Class 3 (basal-like). FOXM1 was exclusively overexpressed in the Class 2 subtype (Fig. 1b, \( p < 0.001 \)). The classification into luminal and basal based on the BASE47 signature from Damrauer et al. (2014) showed an overexpression of FOXM1.
Fig. 2 Chungbuk cohort: a differential expression of FOXM1 based on tumor stage and grade. b Kaplan–Meier plots of the Chungbuk cohort for progression-free survival and overall survival associated with the FOXM1 risk stratification

Table 3 Uni- and multivariable analyses of the UROMOL cohort with progression-free survival as endpoint

| Variables                                      | Univariable Cox regression analyses | Multivariable Cox regression analyses |
|------------------------------------------------|------------------------------------|--------------------------------------|
|                                                 | HR  95% CI  p value                | HR  95% CI  p value                  |
| UROMOL cohort                                   |                                    |                                      |
| FOXM1 high vs low risk                          | 9.8  4.40–22.03 < 0.001            | 5.71  2.45–12.31 < 0.001             |
| MKI67 high vs low risk                          | 10.6  4.07–27.72 < 0.001           |                                      |
| Subtype (luminal cis-like vs rest)              | 5.5  1.21–8.30  0.019              |                                      |
| BASE47                                         | 0.88  0.43–1.78  0.717              |                                      |
| CIS signature                                   | 4.50  1.73–11.73  0.002             |                                      |
| Gene 12 risk signature                          | 2.85  1.31–6.18  0.008              |                                      |
| Lund subtype (infiltrated vs rest)             | 0.14  0.04–0.46  0.001              |                                      |
| Stage (Ta, CIS vs T1)                          | 9.22  4.24–20.05 < 0.001           | 5.28  2.36–11.82 < 0.001             |
| Grade (low grade vs rest)                      | 0.23  0.11–0.50 < 0.001            |                                      |
| Age                                            | 1.06  1.02–1.10  0.005              | 1.04  1.00–1.08  0.04                |
| Sex                                            | 0.82  0.37–1.83  0.630              |                                      |
| BCG treatment                                   | 0.57  0.20–1.64  0.300              |                                      |

Bold values indicate statistical significance (p < 0.05)

HR hazard ratio, CI confidence interval
in the luminal subtype (Fig. 1b, \( p = 0.005 \)). The molecular subtypes previously defined by Sjödahl et al. on the Lund cohort, included the subtypes described in Fig. 3. FOXM1 showed a distinct overexpression in the genomically unstable and SCC-like molecular subtype \(( p < 0.001)\). The NMIBC samples from the UROMOL cohort had additionally been clustered according to the main Lund subtypes: urobasal, infiltrated and genomically unstable. Also in the UROMOL cohort, the genomically unstable subtype showed highest FOXM1 transcript levels (Fig. 1b, \( p < 0.001)\).

**Table 4** Uni- and multivariable analyses of the Chungbuk cohort with progression-free survival as endpoint

| Variables                        | Univariable Cox regression analyses | Multivariable Cox regression analyses |
|----------------------------------|-------------------------------------|--------------------------------------|
|                                  | HR 95% CI  \( p \) value            | HR 95% CI  \( p \) value            |
| Chungbuk cohort                  |                                      |                                      |
| FOXM1 high vs low risk           | 8.53 1.79–40.72  \( \text{0.007} \) | 8.53 1.79–40.72  \( \text{0.007} \) |
| MKI67 high vs low risk           | 6.07 1.69–21.80  \( \text{0.006} \) |                                      |
| Intravesical therapy             | 3.61 0.94–13.83  0.061               |                                      |
| Stage (Ta vs T1)                 | 1.41 0.35–5.64  0.625               |                                      |
| Grade (low vs high grade)        | 3.53 0.88–14.17  0.075               |                                      |
| Age                              | 1.06 0.10–1.12  0.060               |                                      |
| Sex                              | 27.78 0.02–3.24 \times 10^{4}  0.356 |                                      |

Bold values indicate statistical significance \(( p < 0.05)\)

\( HR \) hazard ratio, \( CI \) confidence interval

**Fig. 3** Lund cohort: correlation of FOXM1 with clinical and molecular characteristics of the Lund cohort
FOXM1 serves as prognostic marker for BCG instillation therapy

Following the FOXM1 risk stratification, NMIBCs were further divided in patients with and without intravesical BCG instillation treatment \((p<0.001, \text{Fig. 4})\). Patients with poorest PFS received no instillation treatment and had high FOXM1 expression. However, patients with FOXM1 overexpression benefited most from BCG instillation treatment and constitute an intermediate risk group. However, the BCG therapy had no impact on NMIBC patients with low FOXM1 transcript levels. The Kaplan–Meier estimates showed no differences in PFS in FOXM1 low-risk patients with and without BCG treatment (Fig. 4).

Discussion

In this study, the prognostic and clinical impact of FOXM1 has been investigated retrospectively in a multicenter study and has been validated in published datasets. In order to evaluate its translational benefit, FOXM1 has been compared with MKI67 and relevant clinicopathologic parameters in multivariable analyses. Validation was performed by in silico data based on the Lund and Chungbuk cohort. FOXM1 showed a high correlation with MKI67, underlining its known role in cell proliferation and migration (Hamurcu et al. 2016). High FOXM1 transcript levels correlated with high tumor stage and grade. However, we managed to demonstrate that FOXM1 is an independent predictor for PFS superior to the TNM staging system and MKI67. Indeed, patients of the high-risk group characterized with high FOXM1 expression showed a 6- to 8-fold higher risk of progression \((p<0.001)\).

The UROMOL low-risk group comprised 75% of the patients, whereas in the Chungbuk cohort 58% of patients were included. This can be explained by the significantly higher portion of T1 NMIBCs of the Chungbuk cohort.

FOXM1 showed a distinct subtype-specific overexpression in the Class 2, genomically unstable and SCC-like subtypes \((p<0.001, \text{Figs. 1b, 3})\). As published previously, the Class 2 subtype from Hedegaard at al. overlapped with the Lund subtypes ‘genomically unstable’ and ‘SCC-like’ (Hedegaard et al. 2016). FOXM1 is suspected to play a phenotype-determining role in the development of the molecular bladder cancer subtypes and promote its aggressiveness (Eriksson et al. 2015). The Class 2 subtype, characterized by an APOBEC-related mutational signature and the upregulation of the ERBB gene family, is known to have poorest PFS rate in NMIBCs (Hedegaard et al. 2016). Thus the overexpression of FOXM1 in this specific subtype seems plausible. FOXM1 is well known for its resistance development against many chemotherapies especially cisplatin. Thus, patients with progression of Class 2 tumors may not benefit from platinum-based chemotherapies. Patients with high FOXM1 expression seemed to profit most from BCG instillation therapy. However, only 8 patients of 346 patients from the low-risk FOXM1 group experienced progression, limiting the statistical power for the low-risk group. Thus, further studies for investigating the impact of BCG instillation therapy on patients with low FOXM1 expression are required.

In recent data, FOXM1 has also been shown to be an independent predictor for OS and disease-specific survival in muscle-invasive bladder cancer, with a subtype-specific expression in the luminal subtype (Rinaldetti et al. 2017). Thus FOXM1 has global impact on bladder cancer given its role in both muscle invasive and non-muscle invasive tumors. Up- and downstream FOXM1 regulators (e.g., FOXO3, PI3k, AKT) may be valuable drug targets and should be further explored also in bladder tumors (Yao et al. 2017).

This study has an exploratory character. FOXM1 needs to be further validated in prospective clinical studies in order to evaluate its impact in MIBC resistance development against platinum-based chemotherapies and to validate its prognostic role in NMIBC BCG instillation treatment. In this study two different quantification platforms (RNA-Seq vs microarray) allowed a distinct FOXM1 risk stratification. In order to translate these findings into clinics, a standardizable FOXM1 qPCR screening is needed for future studies (Rinaldetti et al. 2017).

The impact on MIBC and NMIBC as prognostic biomarker superior to clinicopathologic parameters and MKI67, raises FOXM1 to a crucial biomarker for...
molecular grading and to valuable drug target in bladder cancer (Radhakrishnan and Gartel 2008).

Conclusions

FOXM1 is a highly prognostic marker for bladder cancer disease progression. It is mainly expressed in Class 2 and genomically unstable molecular bladder cancer subtype. As FOXM1 is a druggable proto-oncogene, the elucidation of its impact on bladder cancer survival may contribute to a further personalization of future NMIBC or MIBC therapy.

Funding No funding.

Compliance with ethical standards

Conflict of interest All authors declare to have no COIs.

Ethical approval All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Informed consent Informed consent was obtained from all individual participants included in the study.

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

Ark JT, Keegan KA, Barocas DA et al (2014) Incidence and predictors of understaging in patients with clinical T1 urothelial carcinoma undergoing radical cystectomy. BJU Int 113:894–899. https://doi.org/10.1111/bju.12245

Babjuk M, Böhle A, Burger M et al (2017) EAU guidelines on non-muscle-invasive urothelial carcinoma of the bladder: update 2016. Eur Urol 71:447–461. https://doi.org/10.1016/j.eururo.2016.05.041

Budczies J, Klauschen F, Sinn BV et al (2012) Cutoff Finder: a comprehensive and straightforward Web application enabling rapid biomarker cutoff optimization. PLoS One 7:e51862. https://doi.org/10.1371/journal.pone.0051862

Choi W, Porten S, Kim S et al (2014) Identification of distinct basal and luminal subtypes of muscle-invasive bladder cancer with different sensitivities to frontline chemotherapy. Cancer Cell 25:152–165. https://doi.org/10.1016/j.ccr.2014.01.009

Contreras-Sanz A, Roberts ME, Seiler R, Black PC (2017) Recent progress with next-generation biomarkers in muscle-invasive bladder cancer. Int J Urol Off J Jpn Urol Assoc 24:7–15. https://doi.org/10.1111/iju.13193

Dai J, Yang L, Wang J et al (2015) Prognostic value of FOXM1 in patients with malignant solid tumor: a meta-analysis and system review. Dis Markers 2015:352478. https://doi.org/10.1155/2015/352478

Damrauer JS, Hoadley KA, Chism DD et al (2014) Intrinsic subtypes of high-grade bladder cancer reflect the hallmarks of breast cancer biology. Proc Natl Acad Sci USA 111:3110–3115. https://doi.org/10.1073/pnas.1318376111

Dyrskjøt L, Reintier T, Algaia F et al (2017) Prognostic impact of a 12-gene progression score in non-muscle-invasive bladder cancer: a prospective multicentre validation study. Eur Urol 72:461–469. https://doi.org/10.1016/j.eururo.2017.03.040

Eriksson P, Aine M, Veerla S et al (2015) Molecular subtypes of urothelial carcinoma are defined by specific gene regulatory systems. BMC Med Genomics 8:25. https://doi.org/10.1186/s12920-015-0101-5

Halasi M, Gartel AL (2013) Targeting FOXM1 in cancer. Biochem Pharmacol 85:644–652. https://doi.org/10.1016/j.bcp.2012.10.013

Hamurcu Z, Ashour A, Kahraman N, Ozpolat B (2016) FOXM1 regulates expression of eukaryotic elongation factor 2 kinase and promotes proliferation, invasion and tumorigenesis of human triple negative breast cancer cells. Oncotarget 7:16619–16635. https://doi.org/10.18632/oncotarget.7672

Hedegaard J, Lamy P, Nordentoft I et al (2016) Comprehensive transcriptional analysis of early-stage urothelial carcinoma. Cancer Cell 30:27–42. https://doi.org/10.1016/j.cell.2016.05.004

Kalin TV, Ustiyan V, Kalinichenko VV (2011) Multiple faces of FoxM1 transcription factor: lessons from transgenic mouse models. Cell Cycle Georget Tex 10:396–405. https://doi.org/10.4161/cc.10.3.14709

Kim W-J, Kim E-J, Kim S-K et al (2010) Predictive value of progression-related gene classifier in primary non-muscle invasive bladder cancer. Mol Cancer 9:3. https://doi.org/10.1186/1476-4598-9-3

Kluth LA, Black PC, Bochner BH et al (2015) Prognostic and prediction tools in bladder cancer: a comprehensive review of the literature. Eur Urol 68:238–253. https://doi.org/10.1016/j.eururo.2015.01.032

Kwok JM-M, Peck B, Monteiro LJ et al (2010) FOXM1 confers acquired cisplatin resistance in breast cancer cells. Mol Cancer Res MCR 8:24–34. https://doi.org/10.1158/1541-7786.MCR-09-0432

Prout GR, Barton BA, Griffin PP, Friedell GH (1992) Treated history of noninvasive grade 1 transitional cell carcinoma. The National Bladder Cancer Group. J Urol 148:1413–1419

Radhakrishnan SK, Gartel AL (2008) FOXM1: the Achilles’ heel of cancer? Nat Rev Cancer 8:e1; author reply e2. https://doi.org/10.1038/nrc2233-c1

Rinaldetti S, Wirz RM, Worst TS et al (2017) FOXM1 predicts overall and disease specific survival in muscle-invasive urothelial carcinoma and presents a differential expression between bladder cancer subtypes. Oncotarget 8:47595–47606. https://doi.org/10.18632/oncotarget.17394

Robertson AG, Kim J, Al-Ahmadie H et al (2017) Comprehensive molecular characterization of muscle-invasive bladder cancer. Cell 171:540–556.e25. https://doi.org/10.1016/j.cell.2017.09.007

Rodríguez-Alonso A, Pita-Fernández S, González-Carreró J, Nogueira-March JL (2002) p53 and ki67 expression as prognostic factors for cancer-related survival in stage T1 transitional cell bladder carcinoma. Eur Urol 41:182–188 (discussion 188–189)

Saba R, Alsayed A, Zacny JP, Dudek AZ (2016) The role of Forkhead Box Protein M1 in breast cancer progression and resistance to therapy. Int J Breast Cancer 2016:9768183. https://doi.org/10.1155/2016/9768183

Sjödahl G, Lauss M, Lövgren K et al (2012) A molecular taxonomy and system review. Dis Markers 2015:352478. https://doi.org/10.1155/2015/352478
Yao S, Fan LY-N, Lam EW-F (2017) The FOXO3-FOXM1 axis: A key cancer drug target and a modulator of cancer drug resistance. Semin Cancer Biol. https://doi.org/10.1016/j.semcancer.2017.11.018

Ye H, Kelly TF, Samadani U et al (1997) Hepatocyte nuclear factor 3/fork head homolog 11 is expressed in proliferating epithelial and mesenchymal cells of embryonic and adult tissues. Mol Cell Biol 17:1626–1641

Youssef RF, Lotan Y (2011) Predictors of outcome of non-muscle-invasive and muscle-invasive bladder cancer. Sci World J 11:369–381. https://doi.org/10.1100/tsw.2011.28