Baculovirus vector-mediated transfer of NIS gene into colon tumor cells for radionuclide therapy

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Abstract

AIM: To investigate the feasibility of radionuclide therapy of colon tumor cells by baculovirus vector-mediated transfer of the sodium/iodide symporter (NIS) gene.

METHODS: A recombinant baculovirus plasmid carrying the NIS gene was constructed, and the viruses (Bac-NIS) were prepared using the Bac-to-Bac system. The infection efficiency in the colon cancer cell line SW1116 of a green fluorescent protein (GFP) expressing baculovirus (Bac-GFP) at different multiplicities of infection (MOI) with various concentrations of sodium butyrate was determined by flow cytometry. An in vitro cytotoxicity assay was also conducted after infection of SW1116 cells with Bac-NIS. Iodine uptake of Bac-NIS infected SW1116 cells and inhibition of this uptake by sodium perchlorate was examined, and the effect of Bac-NIS-mediated $^{131}$I in killing tumor cells was evaluated by cell colony formation tests.

RESULTS: Infection and transgene expression in SW1116 with Bac-GFP were significantly enhanced by sodium butyrate, as up to 72% of SW1116 cells were infected with the virus at MOI of 400 and sodium butyrate at 0.5 mmol/L. No obvious cytotoxicity was observed under these conditions. Infection of SW1116 with Bac-NIS allowed uptake of $^{131}$I in these tumor cells, which could be inhibited by sodium perchlorate. The viability of SW1116 cells infected with Bac-NIS was significantly lower than with Bac-GFP, suggesting that NIS gene-mediated $^{131}$I uptake could specifically kill tumor cells.

CONCLUSION: Baculovirus vector-mediated NIS gene therapy is a potential approach for treatment of colon cancer.

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Key words: Colon cancer; Baculovirus; Sodium iodide symporter; Radionuclide therapy; Iodine radioisotopes

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meanwhile, chemotherapy shows a relatively high incidence of severe side effects. Therefore, it is important to find alternative ways to improve the prognosis of patients with colon cancer.

The cloning of the sodium/iodide symporter (NIS) gene and subsequent studies of its properties have resulted in a new approach to targeted radiotherapy of malignant cancers. NIS is a transmembrane glycoprotein located in the basal membrane of thyroid follicular cells. It can effectively co-transport two Na+ and one I- into cells[3], and this transportation of iodine is an active process against the chemical gradient. The uptake of radioactive 131I can be achieved by the expression of the NIS protein in tumor cells via vector-mediated gene transfer, and the radiotherapeutic destruction of the tumor occurs by emission of β rays from 131I.

Recently, several researchers have treated tumors by delivering the NIS gene via vectors based on retroviruses[4], adenoviruses[5] and lentiviruses[6], and achieved a certain level of therapeutic effect. For the purpose of gene transfer, an ideal vector would have high transduction efficiency and robust protein expression so that the uptake of radioactive 131I in tumor cells will reach an effective radiotherapeutic dose. The adenovirus vector system does in fact have high transduction efficiency, but its high immunogenicity and cytotoxicity prevent its further application in vivo. The major concern regarding the retroviral or lentiviral system is that the transgenes are easily integrated into the genomic DNA of host cells, which may cause changes in the target genome expression and result in oncogenicity in vivo. Moreover, long-term gene expression is not necessary in therapeutic gene-mediated radiotherapy.

We previously performed a pilot study using a baculovirus vector to deliver the NIS gene as a reporter[7]. Our results showed that the large NIS gene could be inserted into the recombinant baculovirus. A follicular thyroid cancer cell line, FTC-133, infected with recombinant green fluorescent protein (GFP) baculovirus previously showed a high expression of GFP, which lasted approximately seven days[8]. Considering the above characteristics, we reasoned that a recombinant NIS gene baculovirus may be beneficial for mediating 131I uptake and the killing of tumor cells. Furthermore, since baculoviruses can only infect arthropods, this vector is highly safe for mammals. In fact, recombinant baculoviruses have been widely used in mammalian cells as a gene transfer vector because of these advantages[9]. However, no reports are available on baculovirus vector-mediated radiotherapy for cancer.

In this study, the recombinant sodium/iodide symporter gene baculovirus (Bac-NIS) was generated and used to infect colon cancer cells to investigate the expression and function of NIS in these cells and to provide an experimental basis for the NIS gene-mediated targeted radiotherapy of colorectal cancer.

MATERIALS AND METHODS

Construction of baculovirus vectors
The baculovirus vector pFBGFPR (kindly provided by the Molecular Institute of the University of Hong Kong) was digested with restriction endonucleases Acc I and Age I. The digested ends of the vector were filled in with Klenow fragment, yielding a 6 kb PFB fragment which was then purified. The target gene was prepared by digesting the recombinant plasmid pCDNA-NIS (kindly provided by Professor Sissy Jhiang of Ohio University, USA) with restriction endonucleases BamH I and Xho I and filled in with Klenow kinase to obtain the 2.3 kb NIS gene fragment. The purified NIS target gene fragment was ligated to the phosphatase-treated PFB vector using T4 bacteriophage DNA ligase. The resulting plasmid was transformed into DH15a competent bacteria (Tiangen Biotech Co., Ltd., Shanghai) and screened on ampicillin Luria Bertani (LB) culture plates. White colonies were selected and screened for the correct orientation of the inserted target gene fragment by sequencing. The confirmed recombinant baculovirus pFBNIS plasmid DNA was amplified and purified.

Preparation of recombinant NIS and GFP baculoviruses
The baculovirus vector plasmid pFBGFPR and the constructed recombinant plasmid pFBNIS were transformed into DH10 competent bacteria (Invitrogen). The successfully transpositioned white colonies were selected and amplified. Bacmid plasmids expressing the NIS gene were extracted and transfected into Spodoptera frugiperda (sf9) cells using Lipofectamine 2000 (Invitrogen). The supernatant of the transfected cells containing the virus was collected 5-7 d later. SF9 cells were infected at a multiplicity of infection (MOI) of 0.1 and harvested after 5-7 d in culture by centrifuging at 10000 × g for 15 min. The resulting supernatant was collected, centrifuged again at a high speed of 10000 × g at 4°C for 5 min, sterilized with a 0.45 μm filter and preserved at 4°C in the dark. Plaque assays were used to determine the titer of baculovirus stocks.

SW1116 cell culture
SW1116 (human Caucasian colon adenocarcinoma) was cultured in DMEM (Gibco-BRL) supplemented with 10% fetal bovine serum (FBS; Gibco-BRL) and 1% penicillin/streptomycin (Gibco-BRL) in a humidified environment with 5% CO2 at 37°C.

Baculovirus infection of SW1116 cells
SW1116 cells were seeded in 6-well or 24-well plates with DMEM with 10% FBS at least 24 h before infection. The medium was changed to fresh serum-free DMEM just before infection. After infection with the baculovirus vector at indicated MOIs, the cells were incubated with serum-free DMEM for 4 h, followed by addition of FBS to the final concentration of 10%. All infections were performed in triplicate.

Effects of sodium butyrate on the transduction efficiency and fluorescence intensity of SW1116 cells
SW1116 cells were seeded in 6-well plates at a density of 1 × 105 cells/well and cultured with DMEM without serum for 24 h. Recombinant green fluorescent protein
gene baculovirus (Bac-GFP) was then added at an MOI of 400, and sodium butyrate was added at different concentrations (0, 0.5, 1 and 2 mmol/L) simultaneously. All experiments were performed in triplicate. Cells were incubated at 37℃ for 4 h after treatment, and FBS was added to the final concentration of 10% and incubated for another 24 h. The expression of GFP and the mean fluorescence intensity were determined by flow cytometry. The excitation wavelength of blue laser was 488 nm, and the detection wavelength was 520 nm.

**Infection efficiency of Bac-GFP in SW1116 cells**
SW1116 cells were seeded in 6-well plates at a density of 1 × 10^4 cells/well, and cultured with serum-free DMEM for 24 h. Bac-GFP was added at the MOI of 0, 100, 200, or 400. The final concentration of sodium butyrate was 0.5 mmol/L, and cells without sodium butyrate were used as negative controls at various MOI of Bac-GFP. All experiments were performed in triplicate. After treatment, the cells were incubated at 37℃ for 4 h, and FBS was added to reach the final concentration of 10% and incubated at 37℃ for another 24 h. The expression of GFP was observed under an inverted phase-contrast fluorescence microscope. The expression of GFP and the mean fluorescence intensity were detected by flow cytometry.

**Effects of Bac-GFP infection and sodium butyrate on the cytotoxicity of SW1116 cells**
SW1116 cells were seeded in 6-well plates at a density of 1 × 10^4 cells/well and cultured with serum-free DMEM for 24 h. Bac-GFP was added at the MOI of 0, 50, 100, 200 or 400. For testing the effect of sodium butyrate, the final concentration was 0, 0.5, 1, and 2 mmol/L with Bac-GFP at an MOI of 400, and wells without sodium butyrate were used as the control well at an MOI of 400. After treatment, the cells were incubated at 37℃ for 4 h, and FBS was added to reach the final concentration of 10% and incubated at 37℃ for another 24 h. Twenty microliters of 5 g/L MTT (Beyotime Co., Ltd., Shanghai) were added to each well and incubated for another 4 h. Subsequently, 100 μL DMSO was added, and the plate was gently shaken to completely dissolve the blue-violet precipitate. The absorption at 490 nm (A_{490}) was determined using a microplate reader. Cell viability = test well A_{490}/control well A_{490} × 100%.

**Determination of iodine uptake of SW1116 cells infected with Bac-NIS**
SW1116 cells were seeded in 6-well plates at a density of 5 × 10^4 cells/well and cultured with serum-free DMEM for 24 h. Bac-NIS was added at the MOI of 400. The final concentration of sodium butyrate was 0.5 mmol/L. All experiments were performed in triplicate. After treatment, cells were incubated at 37℃ for 4 h, and FBS was added to reach the final concentration of 10% and incubated at 37℃ for another 24 h. Twenty microliters of 5 g/L MTT (Beyotime Co., Ltd., Shanghai) were added to each well and incubated for another 4 h. Subsequently, 100 μL DMSO was added, and the plate was gently shaken to completely dissolve the blue-violet precipitate. The absorption at 490 nm (A_{490}) was determined using a microplate reader. Cell viability = test well A_{490}/control well A_{490} × 100%.

**Inhibition of iodine uptake by NaClO**
SW1116 cells were seeded in 6-well plates at a density of 5 × 10^4 cells/well and cultured with serum-free DMEM for 24 h. Bac-NIS and Bac-GFP were added at the MOI of 400 respectively. The final concentration of sodium butyrate was 0.5 mmol/L, and Bac-GFP was used as negative controls. All experiments were performed in triplicate. After treatment, cells were incubated at 37℃ for 4 h, FBS was then added to reach the final concentration of 10% and incubated at 37℃ for another 24 h. Cells were washed twice with bHBSS solution, and 0.5 mL bHBSS solution containing 0.1 μCi Na^{125}I with 1 μmol/L NaI were added into each well with cold bHBSS solution three times, and incubated for another 20 min with 1 mL 100% cold ethanol. The samples were then collected to determine the radioactive cpm by a well γ-counter.

**125I-mediated killing of tumor cells in cell colony formation test**
SW1116 cells were inoculated in 6-well plate at a density of 5 × 10^4 cells/well and cultured with serum-free DMEM for 24 h. Bac-NIS and Bac-GFP were added at an MOI of 400 with sodium butyrate at 0.5 mmol/L respectively. Bac-GFP was used as the negative control. The same volume of bHBSS solution was added into a blank well as a control. All experiments were performed in triplicate. After treatment, cells were incubated at 37℃ for 4 h, and FBS was added to reach the final concentration of 10% and incubated at 37℃ for another 24 h. Cells were washed twice with bHBSS solution. After 0.5 mL bHBSS solution containing 0.1 μCi Na^{125}I was added into each well with cold bHBSS solution three times and incubated for another 20 min with 1 mL 100% cold ethanol. The samples were then collected to determine the radioactive cpm by a well γ-counter.
twice with bHBSS solution, digested, counted, seeded in a 6-well plate (1000 cells/well) and incubated at 37 ℃ for additional 7 d. Cells were then washed three times with bHBSS solution and stained with crystal violet solution (Sigma). Cell viability was determined in the infected cells by counting colonies visible to the naked eyes and calculated as a percentage of the colonies that were treated with only bHBSS [8].

Statistical analysis
All data are presented as mean ± SD. Differences were considered significant at P < 0.05. Statistical analysis was performed using SPSS 11.0 software.

RESULTS
Sodium butyrate enhances infection efficiency of Bac-GFP and expression of the reporter protein
The transduction efficiencies of baculovirus in SW1116 cells were 39%, 71%, 80% and 83% at an MOI of 400 with concentrations of sodium butyrate of 0, 0.5, 1 or 2 mmol/L, respectively. This result indicated that sodium butyrate could significantly improve the transduction efficiency of baculovirus in SW1116 cells, achieving high levels of transduction at concentrations above 0.5 mmol/L. The fluorescence intensity of the infected SW1116 cells increased along with sodium butyrate in a dose-dependent manner, suggesting that the expression of the reporter protein was enhanced by the sodium butyrate (Figure 1).

Transduction efficiency of Bac-GFP at various MOI in SW1116 cells
The transduction efficiency (Figure 2A) of baculovirus (Bac-GFP) in the SW1116 tumor cells gradually increased, and the fluorescence intensity (Figure 2B) was also enhanced with increasing MOI. Additionally, sodium butyrate could significantly improve the transduction efficiency and fluorescence intensity of tumor cells by Bac-GFP at different MOI (Figure 2). The sodium butyrate-mediated transduction efficiency of baculovirus inoculated at the MOI of 400 in SW1116 cells was 72%.

Cytotoxicity of SW1116 cells infected with baculovirus at different MOIs with various concentrations of sodium butyrate
There was no obvious change in cell viability with the dose escalation of recombinant baculovirus (Figure 3A). There was no significant cell death observed at the highest MOI of 400. These results suggested that the baculovirus itself had no obvious cytotoxic effects on the tumor cells.
On the other hand, the tumor cell viability decreased with increasing concentrations of sodium butyrate with virus at the MOI of 400 (Figure 3B). However, the cell viability did not change significantly when the concentration of sodium butyrate was lower than 0.5 mmol/L.

**Determination of iodine uptake in infected SW1116 cells**

The radiiodine uptake of tumor cells increased with increasing doses of Bac-NIS, and this uptake was significantly enhanced by sodium butyrate (Figure 4).

**Dynamic determination of $^{125}$I uptake of SW1116 cells**

The radiiodine uptake of infected tumor cells increased over the incubation time with radiiodine which reached the peak at 30 min with Bac-NIS at the MOI of 400 and sodium butyrate at 0.5 mmol/L (Figure 5). This result suggested that the NIS protein expressed in transfected tumor cells could perform its normal function in iodine uptake.

**Inhibition of iodine uptake by NaClO$_4$**

NIS-mediated iodine uptake in thyroid tissues can be specifically inhibited by ClO$_4^-$. In the present study, we tested the ability of ClO$_4^-$ at various concentrations to inhibit the iodine uptake by tumor cells expressing the NIS protein. Our results showed that the radiiodine uptake of SW1116 cells was significantly reduced with escalating doses of ClO$_4^-$. (Figure 6), indicating that the function of the expressed NIS protein in the infected tumor cells could also be inhibited by the presence of NaClO$_4$.

**$^{131}$I-mediated killing of tumor cells**

In order to evaluate the killing effect of $^{131}$I in infected tumor cells, a cell colony formation test was performed with cells infected by Bac-GFP as the control. The results showed that the cell viability of SW1116 cells infected with Bac-NIS was reduced to about 20%, and that of SW1116 cells infected with Bac-GFP (70%) was similar to that of the non-infected group (76%) (Figure 7). These observations suggested that NIS-mediated tumor cells could be effectively and specifically killed by $^{131}$I.

**DISCUSSION**

Since 1996 when Dai et al$^{[9]}$ and Smanik et al$^{[10]}$ cloned the
Meanwhile, the November 14, 2010

rus expressing the NIS gene due to the rapid clearance of iodine in infected rat prostate carcinoma cells by a retrovirus-infected hepatoma cell line (MH3924A) and screened stable cell lines expressing the NIS gene from cervix cancer, breast cancer and prostate carcinoma by adenovirus-mediated NIS gene transfer or viral vectors in recent years. Boland experiments provided solid evidence for the feasibility of radionuclide therapy for colon cancer.

Presently, viral vectors most commonly used for gene transfer are based on adenoviruses, retroviruses and lentiviruses. Adenoviruses can induce strong host immune responses, and antibodies against adenovirus has been found in 97% of the population. These pre-existing antibodies against the viral vector can inhibit the efficiency of gene transfer. Furthermore, transgenes carried by lentiviruses and retroviruses can randomly insert into the genomic DNA of the host cells and potentially activate oncogenes in normal tissues.

Therefore, it is critical to choose a proper vector system to deliver the gene of interest in gene therapy. The baculovirus vector system has a high transduction efficiency and protein expression in mammalian cells. It is a replication competent vector that displays less cytotoxicity to mammalian cells when compared with other viral vectors. Because the baculovirus DNA does not integrate into the host genome, baculovirus vectors have an extremely high biological safety profile. Since baculoviruses also have very species-specific tropisms among the invertebrates, there would be virtually no pre-existing neutralizing antibody and specific T cells in mammals to prevent the application of these vectors in vivo. Meanwhile, the packaging capacity of baculovirus is about 38 kb, allowing insertion and expression of large genes, and high titer virus stocks can easily be produced. All these advantages show that the baculovirus is a promising vector for use in gene transfer applications.

In this study, we successfully cloned and purified baculovirus with the NIS gene driven by the cytomegalovirus immediate-early gene promoter. Some studies have shown that the transduction efficiency of recombinant baculovirus can be increased by adding sodium butyrate in SW1116 cells. Infection of Bac-NIS in SW1116 cells showed that it can uptake iodine, and that this process can specifically inhibit the host sodium perchlorate. This suggested that the NIS protein expressed by baculovirus functioned similarly to the natural NIS protein. The proliferation of SW1116 cell was significantly lower in cells infected with Bac-NIS than those infected with Bac-GFP, indicating that tumor cells infected with Bac-NIS could specifically uptake \(^{131}\text{I}\), and as a result the emitted \(\beta\)-rays could kill the target cells effectively. These experiments provided solid evidence for the feasibility of radionuclide therapy for colon cancer.

The expression level of an exogenous gene positively correlates with the amount of stable transgenes in host cells, and the regulation of the transgene in mammalian cells is the key to its expression. Modifications of the transgenes such as acetylation, methylation and/or the chromatin compactness may play important roles in the expression of the transgenes carried by the baculovirus. Sodium butyrate is a four-carbon short-chain fatty acid derived from dietary fiber in the intestinal tract via bacte-
rial fermentation. The concentration of sodium butyrate is very high in the intestinal tract of healthy individuals especially those with a high-fiber diet. Sodium butyrate is a histone deacetylase inhibitor that has been shown to relax chromatin by inducing the acetylation of chromosomes which can in turn facilitate gene expression. Furthermore, it can also significantly improve the transduction efficiency of baculovirus and the expression level of exogenous genes[9]. In this study, we found that sodium butyrate could significantly improve the transduction efficiency of baculovirus and the expression of the NIS transgene in SW1116 cells which led to a significant killing effect (viability = 20%) of SW1116 tumor cells. Therefore, baculovirus-mediated gene transfer of the NIS gene allows targeted radiotherapy for intestinal tumors.

The therapeutic effect of radiiodine therapy depends on the absorbance of target tissues which are affected by the effective half-life of $^{131}$I in 1 tumor. However, iodine uptake in SW1116 cells mediated by NIS gene transfer is rapidly cleared from the body. This is different from the observation in thyroid cells due to the absence of thyroid peroxidase (TPO) in SW1116 cells. Therefore, the organification of inorganic iodine taken up by SW1116 cells to produce thyroid hormone for storage cannot take place, ultimately affecting the therapeutic effect for tumors that can be achieved with $^{131}$I. Thus, an important issue in NIS gene-mediated tumor therapy is the improvement of the retention of $^{131}$I in tumor cells. Huang et al.\[6] transfected NIS and the TPO gene together into non-small cell lung cancer cell lines and found that the rate of clearance of iodine was reduced. However, it is still controversial whether or not the organification of iodine mediated by TPO would increase the retention of $^{131}$I.\[21]

Various approaches to improve radiotherapy have been reported. It has been shown that NIS-mediated Re (Rhenium) radiating $\beta$-rays\[22] and At (Astatine) radiating $\alpha$-rays\[23,24] in various tissues resulted in better radiotherapeutic effects than with $^{131}$I due to higher radiation energy deposition. Additionally, as the serum complement is the primary inhibitory factor of baculovirus-mediated gene transfer, some researchers have created certain modifications in baculovirus to prevent inactivation by complement in serum. For example, human decay-accelerating factor expressed on the envelope of the virion could render recombinant baculovirus resistant to the complement system, and the expression of transgene could be detected when the modified baculovirus was injected into murine hepatic parenchyma.\[24] The inhibition of serum inactivation can also be achieved by injecting complement inhibitors such as cobra-venom factor prior to baculovirus vector delivery.\[25] Thus, baculovirus vectors will provide a new approach for the gene therapy of malignant tumors especially when the related techniques are improved.

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