Granulocyte Colony-Stimulating Factor Combined with Methylprednisolone Improves Functional Outcomes in Rats with Experimental Acute Spinal Cord Injury

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OBJECTIVES: To evaluate the effects of combined treatment with granulocyte colony-stimulating factor (G-CSF) and methylprednisolone in rats subjected to experimental spinal cord injury.

METHODS: Forty Wistar rats received a moderate spinal cord injury and were divided into four groups: control (no treatment); G-CSF (G-CSF at the time of injury and daily over the next five days); methylprednisolone (methylprednisolone for 24 h); and G-CSF/Methylprednisolone (methylprednisolone for 24 h and G-CSF at the time of injury and daily over the next five days). Functional evaluation was performed using the Basso, Beattie and Bresnahan score on days 2, 7, 14, 21, 28, 35 and 42 following injury. Motor-evoked potentials were evaluated. Histological examination of the spinal cord lesion was performed immediately after euthanasia on day 42.

RESULTS: Eight animals were excluded (2 from each group) due to infection, a normal Basso, Beattie and Bresnahan score at their first evaluation, or autophagy, and 32 were evaluated. The combination of methylprednisolone and G-CSF promoted greater functional improvement than methylprednisolone or G-CSF alone (p < 0.001). This combination also exhibited a synergistic effect, with improvements in hyperemia and cellular infiltration at the injury site (p < 0.001). The groups displayed no neurophysiological differences (latency p=0.85; amplitude p=0.75).

CONCLUSION: Methylprednisolone plus G-CSF promotes functional and histological improvements superior to those achieved by either of these drugs alone when treating spinal cord contusion injuries in rats. Combining the two drugs did have a synergistic effect.

KEYWORDS: Spinal Cord Injuries; Wistar Rats; Granulocyte Colony-Stimulating Factor; Methylprednisolone.

INTRODUCTION

Researchers worldwide struggle to find evidence of substances or procedures that can reduce the devastating consequences of spinal cord injuries, which affect up to 80 per million people worldwide, with 500,000 new cases per year (1). Most research is focused on surgical treatment, rehabilitation or physical training and particularly on pharmacological therapies aimed at restoring neuronal plasticity and function by reducing the effects of secondary lesions and stimulating tissue regeneration (2-5). Estrogen has been found to reduce pro-inflammatory activity and protect neurons (6,7). The antioxidant effect of melatonin has also been extensively investigated, but only in low-quality experimental studies (8). Methylprednisolone has proven clinically effective (9,10); however, its use as a first-line treatment has recently been questioned due to the high incidence of complications and adverse events, especially when higher doses are used (11). The use of methylprednisolone has thus been reevaluated (12) but is still widely accepted (13). Stem-cell transplantation (14,15) has also been investigated.

Granulocyte colony-stimulating factor (G-CSF) is a glycoprotein that acts as a growth-stimulating factor for hematopoietic progenitor cells (16,17). G-CSF is used to treat neutropenia and to mobilize hematopoietic stem cells in cases of bone marrow transplantation (16). Studies show that G-CSF also has non-hematopoietic functions and that it can increase tissue regeneration in organs such as the brain and heart (17,18).

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G-CSF has also been shown to produce neurological improvements in experimental models of spinal cord injury (17,19-21). In the acute phase of injury, G-CSF mobilized bone marrow cells to the injured spinal cord, where it suppressed neuronal and oligodendrocyte apoptosis, the expression of inflammatory cytokines such as tumor necrosis factor-alpha (TNF-α) and interleukin-1 beta (IL-1 beta) (22), and lipid peroxidation (23), in addition to stimulating the production of neurotrophic factors (21). In the subacute phase of traumatic spinal cord injury, G-CSF stimulated angiogenesis (24) and suppressed glial scar formation (25). In two phase I/IIa studies, some neurological recovery was observed in most patients with spinal cord injuries (26,27).

It is also possible that some drugs may act synergistically to alleviate spinal cord injury (28), and synergy between pharmacological and physical factors may improve trauma recovery (2,29). Indeed, the best strategy in situations with a complex pathophysiology, as in the case of spinal cord injury, likely involves treatments that employ different pathogenic mechanisms (29). However, no study has yet investigated the effects of combining methylprednisolone and G-CSF, which can be investigated using an experimental model of spinal cord injury in rats.

Therefore, our objective was to evaluate the effects of G-CSF associated with methylprednisolone on functional, neurophysiological and histological outcomes in a standardized experimental rat model of spinal cord injury.

### MATERIALS AND METHODS

#### Ethics

This experimental study was carried out in strict accordance with all international guidelines on handling and controlling pain or suffering in the care and use of animals in research. According to our laboratory protocols, all animals are handled and stimulated to move prior to the experiments so that they can become accustomed to the researchers and to the motor function evaluation procedures. *Ad libitum* feeding and hydration are maintained during studies carried out in the lab, and appropriate cages (40 x 60 cm) and housing are provided, with a maximum of five animals per cage in a space with controlled temperature and humidity. The animals were submitted to surgical and experimental procedures and were euthanized under general anesthesia.

The study protocol was evaluated and approved by the Research Ethics Committee of the institution (Comissão Científica do Instituto de Ortopedia e Traumatologia e pela Comissão de Ética para Análise de Projetos de Pesquisa do Hospital das Clinicas da Faculdade de Medicina da Universidade de São Paulo (CAPPesq – HC-FMUSP; permit number: IOT no. 965).

#### Study design, sample size and experimental animals

This is a controlled study with Wistar rats, which were divided into four groups. Sample size was based on previously published studies using approximately 10 animals per group (9,29). In total, 40 male Wistar rats, aged 20 to 21 weeks old and initially weighing 300 to 340 g, were used. All animals came from the same university vivarium and were weighed at the beginning and at the end of the study. All rats were clinically evaluated at baseline to ensure that they had normal motricity and good health. Animals with macroscopic evidence of abnormalities in the spinal cord or functional problems were excluded. Animals that died shortly after the experimental spinal cord lesion, or animals with autophagy or mutilation behaviors, were also excluded from the study. Normal motricity (21 points on the Basso, Beattie and Bresnahan (BBB) scale), even after the experimental spinal cord lesion, was also an exclusion criterion, since normal functioning indicated that the procedure failed.

The 40 animals were randomly allocated (using sequence generator software and a ratio of 1:1:1:1) into five groups, with 10 animals per group. The Control Group (CG) received no treatment at all after the experimental spinal cord injury. The Methylprednisolone Group (MG) was treated with 30 mg/kg of intravenous methylprednisolone at 10 min, 6 h and 24 h after the spinal cord lesion, according to the protocol described by Park et al. (30). The G-CSF group received treatment with 15 mcg/kg of subcutaneous G-CSF after the lesion was induced and then daily for 5 days, according to the method described by Kawabe et al. (24). Finally, the G-CSF-M group received G-CSF at the moment the experimental lesion was induced, followed by daily treatment with G-CSF for 5 days, as well as methylprednisolone 24 h after the lesion was induced. G-CSF and methylprednisolone were provided at 10 min, 6 h and 24 h after the spinal cord lesions were given to this group, at the same doses used for the other two groups.

#### Procedures

All animals were operated on by a single surgeon. The animals were anesthetized using 50 mg/kg of pentobarbital administered intraperitoneally. The anesthetic effect was expected to begin within five minutes and to last approximately two hours, which was long enough for the spinal cord injury procedure to be carried out. The level of anesthesia was determined by evaluating the withdrawal reflex to tail compression. After spinal cord injury, the animals were treated with oral tramadol (15 mg/kg) dissolved in water (31).

All animals underwent a laminectomy under anesthesia. A moderate contusion spinal cord lesion was produced at T10, as described previously (29), using an NYU-Impactor device with a 10-g impact rod from a 12.5 mm-height to compress the spinal cord for 15 s. This procedure produced a spinal cord lesion that created loss of locomotor function.

After surgery and spinal cord lesion, the animals were transferred to chambers with controlled temperature (25°C to 28°C), where they remained for 30 min. The bladders were emptied by external compression. Antibiotic prophylaxis was performed with cephalothin administered subcutaneously (25 mg/kg) immediately after the lesion and once a day for seven subsequent days. All rats were kept in the same vivarium and under the same controlled environmental conditions for 42 days before being euthanized by a lethal dose of pentobarbital (140 mg/kg).

The spinal cord was removed for necropsy. A segment from T8 to T12 (approximately 2.5 cm-long) was fixed and prepared for histological analysis as described previously (29). Axial slices were cut 2 mm apart to span the injured area and were extended 1 cm proximally and caudally from the center of the lesion. After fixation and production of paraffin blocks, 5-μm-thick sections were produced and stained with hematoxylin-eosin for histological evaluation.

#### Outcomes evaluation

The pathologist responsible for the histological analysis (not one of the authors), who was blinded to the experimental
groups, scored the sections for the presence of necrosis, hemorrhage, hyperemia, neural tissue degeneration, and infiltrate as follows: 0 (absent), 1 (mild), 2 (moderate) and 3 (extensive).

Function was evaluated using the BBB rating scale (29,32,33). This 21-point scale ranges from complete paraplegia (score 0) to normal neurological function (score 21) and was applied for 4 to 5 min on days 2, 7, 14, 21, 28, 35, and 42 after spinal cord lesioning. The BBB score on the 42nd day after injury was assessed simultaneously by two trained observers who were blinded to experimental conditions and to each other’s evaluations. In cases of disagreement, the lower score was recorded for analysis.

On the 42nd day after lesioning, anesthetized rats (pentobarbital, 50 mg/kg) underwent a motor evoked potential (MEP) exam according to the method of Letaif et al. (6), which was performed by a researcher who was blinded to the experimental conditions, with electrodes positioned in the semitendinosus and the biceps muscles of the thigh. Amplitude and latency data were recorded.

Statistical analysis

Data are presented as means and standard deviations in the descriptive analysis. The primary outcome of this study was the BBB score on the 42nd day after injury. The histological variables and MEP results were secondary outcomes.

For inferential statistics, the data behaviors were observed for possible outliers. Once identified, outliers were submitted to the Dixon Q test for possible rejection. Subsequently, the data were subjected to the Kolmogorov-Smirnov normality test. If the data showed a normal distribution, parametric tests, e.g., Student’s t-test and analysis of variance (ANOVA), were performed. The number of factors was adjusted for each situation. Parametric analysis was also performed when the distribution was asymmetric. When the asymmetry was to the right, the data were transformed by logarithm to the base of 10, and when it was to the left, the data were raised to the square.

The BBB scores were compared using a mixed effects model with two factors, four groups and six weeks of evaluation, while considering repeated measures over time. A possible effect of interaction between these factors was also evaluated. The mixed effects model was adjusted by considering matrices of equal covariance in the different groups, as well as in unstructured form.

For the MEP results, the distribution of amplitude and latency data was initially compared between the four groups using the Kruskal-Wallis test. Next, ANOVA was used to compare the amplitude and latency means in different groups. The histological variables were also compared between groups using the Kruskal-Wallis test.

We started with a null hypothesis considering a 5% probability of type I error. Statistical analyses were performed using the Statistical Package for Social Sciences (SPSS), version 19.0zx.

RESULTS

A total of 40 animals were operated on, but two rats had to be excluded from each group. In the CG, one rat was excluded due to failure of the experimental lesion (i.e., the rat showed normal function after the procedure), and the second was excluded for autophagy on the 22nd postoperative day.

### Table 1 - Basso, Beattie, and Bresnahan (BBB) rating scores for hindlimbs, according to the evaluation times after spinal cord injury: mean, minimum (min) and maximum (max) values, standard deviations (SD), degrees of freedom (DF) and results of the chi-squared and Kruskal-Wallis tests.

| Day | Group | N | Mean | SD | Min | Max | $\chi^2$ | DF | p |
|-----|-------|---|------|----|-----|-----|--------|-----|----|
| 2   | CG    | 16 | 0.31 | 0.60 | 0.0  | 2.0  | 0.905  | 3   | 0.824 |
|     | MG    | 16 | 0.31 | 0.60 | 0.0  | 2.0  |        |     |    |
|     | G-CSF | 16 | 0.50 | 0.73 | 0.0  | 2.0  |        |     |    |
|     | G-CSF-M | 16 | 0.44 | 0.73 | 0.0  | 2.0  |        |     |    |
| 7   | CG    | 16 | 1.13 | 0.62 | 0.0  | 3.0  | 4.938  | 3   | 0.176 |
|     | G-CSF | 16 | 0.63 | 0.89 | 0.0  | 2.0  |        |     |    |
|     | G-CSF-M | 16 | 1.00 | 1.71 | 0.0  | 5.0  |        |     |    |
| 14  | CG    | 16 | 4.94 | 2.11 | 1.0  | 8.0  | 16.634 | 3   | 0.001 |
|     | MG    | 16 | 4.38 | 2.33 | 1.0  | 9.0  |        |     |    |
|     | G-CSF | 16 | 3.13 | 1.59 | 1.0  | 5.0  |        |     |    |
|     | G-CSF-M | 16 | 6.38 | 1.78 | 3.0  | 8.0  |        |     |    |
| 21  | CG    | 16 | 6.06 | 2.69 | 2.0  | 11.0 | 16.126 | 3   | 0.001 |
|     | MG    | 16 | 7.63 | 3.32 | 1.0  | 14.0 |        |     |    |
|     | G-CSF | 16 | 5.44 | 1.71 | 2.0  | 8.0  |        |     |    |
|     | G-CSF-M | 16 | 8.81 | 2.14 | 5.0  | 12.0 |        |     |    |
| 28  | CG    | 16 | 9.56 | 1.82 | 7.0  | 12.0 | 21.408 | 3   | <0.001 |
|     | MG    | 16 | 7.88 | 1.36 | 5.0  | 10.0 |        |     |    |
|     | G-CSF | 16 | 11.38 | 3.46 | 5.0  | 15.0 |        |     |    |
|     | G-CSF-M | 16 | 7.56 | 2.78 | 5.0  | 12.0 | 21.204 | 3   | <0.001 |
| 35  | CG    | 16 | 10.13 | 1.63 | 8.0  | 12.0 |        |     |    |
|     | MG    | 16 | 9.31 | 1.62 | 8.0  | 12.0 |        |     |    |
|     | G-CSF | 16 | 13.19 | 3.37 | 8.0  | 18.0 |        |     |    |
|     | G-CSF-M | 16 | 10.38 | 1.59 | 8.0  | 12.0 |        |     |    |
| 42  | CG    | 16 | 8.69 | 2.09 | 6.0  | 12.0 | 21.644 | 3   | <0.001 |
|     | MG    | 16 | 11.00 | 1.83 | 8.0  | 14.0 |        |     |    |
|     | G-CSF | 16 | 14.13 | 3.59 | 8.0  | 18.0 |        |     |    |

CG = control group; MG = methylprednisolone group; G-CSF = granulocyte colony-stimulating factor group and G-CSF-M = group receiving G-CSF and methylprednisolone.
In the MG group, animals were excluded due to infection on the 8th and 10th postoperative days. In the G-CSF group, one animal was excluded due to lesion failure (i.e., normal BBB) and the other due to infection on the 5th postoperative day. Finally, in the G-CSF-M group, animals were excluded due to lesion failure and autophagy (on the 10th postoperative day).

The final sample consisted of eight animals in each group and therefore 16 limbs. Because the BBB scores in both limbs of the same animal were considered equivalent, Table 1 shows the data on the BBB evaluation for 16 cases per group at seven evaluation times. From the 14th day on, the BBB scores differed significantly among the groups. The highest score was obtained in the G-CSF-M group, with a difference of 1.44 points on the 14th day and 5.44 on the 42nd day from the score in the CG (untreated). Multiple (post-hoc) comparisons are shown in Table 2 and confirm the superior recovery of the G-CSF-M group.

Because the MEP results were also equivalent in the forelimbs and hindlimbs, this evaluation was performed for 16 individuals per group. The MEP results in the forelimbs did not differ significantly, either for latency ($p=0.09$) or amplitude values ($p=0.20$), and the hindlimbs also showed no significant differences ($p=0.85$ and 0.08 respectively; Table 3).

Table 4 shows the results of the histological analysis. No significant difference was observed for necrosis, but the other histological variables were significantly different among the groups. The multiple comparison test (Mann-Whitney) showed significant differences for hyperemia, hemorrhage, degeneration of neural tissue and infiltration, as shown in Table 5. Figure 1 shows examples of histological evaluations and an example of a normal spinal cord.

**DISCUSSION**

One of the reasons why traumatic spinal cord injury is such a devastating condition is that it leads to chronic impairment and disability: it impairs the function of young accident victims in the productive phase of their lives, and with the aging of the population, it is also increasingly affecting the elderly (1,34). Despite numerous advances in diagnostic technology, rehabilitation and surgical decompression therapies,
which have reduced morbidity and mortality, improvements in the therapeutic approach to secondary lesions are still mostly experimental. No treatment has yet to provide a cure or to facilitate complete functional recovery for patients with spinal cord injury. In the present study, we explored whether a combination of substances will have a better effect on functional and histological recovery than one isolated drug, possibly due to synergistic effects or to the fact that different substances may act in different pathways as part of a complex physiopathological process. This hypothesis proved to be accurate: the combination of G-CSF and methylprednisolone proved to be better than the control treatment and either treatment alone, according to the functional evaluations from the second week of treatment in rats. The functional results were also better than those observed in other studies using G-CSF alone (20,31).

Primary spinal cord injury develops into a secondary injury via several biological processes, resulting in the impairment of viable tissues and leading to necrosis and neuronal apoptosis (16). The goal of pharmacological therapy is to reduce or avoid secondary injury by inhibiting inflammation, lipid peroxidation and excitotoxicity (35). Several drugs have been studied in experimental and clinical trials (2,9). Many drugs showed promising results in experimental animal studies, but few have proven effective in humans. Of these, methylprednisolone, tirilazad mesylate, naloxone, and GM-1 ganglioside were evaluated in human clinical trials in patients with spinal cord injury (36). Methylprednisolone was the most extensively studied drug (10,36). Although its mechanism of action is not fully known, methylprednisolone has been shown to stabilize membranes and preserve the hematospinal cord barrier, potentially reducing vasogenic edema. It also increases spinal cord blood flow, changes the electrolyte concentrations at the injury site, inhibits endorphin release, reduces free radical availability, and reduces the inflammatory response (11,36). Despite its worldwide acceptance in the treatment of spinal cord injury, methodological problems in clinical trials and new evidence from clinical studies and literature reviews have raised concerns about the small effect sizes of the clinical benefits, as well as the risks of complications associated with the use of methylprednisolone (36). Therefore, new studies should be conducted with promising drugs, including but not limited to methylprednisolone, tirilazad mesylate, naloxone and

**Table 3** - Latency and amplitude values in the motor-evoked potential exam (means, standard deviations, minimum and maximum values) of hindlimbs of rats subjected to experimental spinal cord lesion and comparisons between groups using the chi-squared and Kruskal-Wallis tests.

| Hindlimbs | Groups | N (limbs) | Mean (ms) | SD | Min | Max | χ² | DF | p |
|-----------|--------|-----------|-----------|----|-----|-----|-----|-----|---|
| **Latency** | | | | | | | | | |
| CG | 16 | 27.83 | 8.77 | 15.80 | 40.30 | 6.626 | 3 | 0.85 |
| MG | 16 | 23.71 | 3.74 | 16.30 | 29.10 |
| G-CSF | 16 | 22.36 | 7.30 | 12.50 | 34.20 |
| G-CSF-M | 16 | 21.30 | 8.20 | 12.40 | 40.30 |
| **Amplitude** | | | | | | | | | |
| CG | 16 | 26.86 | 14.78 | 6.00 | 50.10 | 6.898 | 3 | 0.075 |
| MG | 16 | 43.98 | 65.22 | 4.70 | 276.80 |
| G-CSF | 16 | 64.71 | 92.85 | 10.90 | 312.10 |
| G-CSF-M | 16 | 97.43 | 115.58 | 4.70 | 483.00 |

CG = control group; MG = methylprednisolone group; G-CSF = granulocyte colony-stimulating factor group and G-CSF-M = group receiving G-CSF and methylprednisolone.

**Table 4** - Histological analysis scores according to the variables necrosis, hemorrhage, hyperemia, degeneration, and cellular infiltrate: mean, minimum (min) and maximum (max) values, standard deviations (SD), degrees of freedom (DF) and chi-squared and Mann-Whitney test results.

| Variable | Group | N | Mean score (0-3) | SD | Min | Max | χ² | DF | p |
|----------|-------|---|-----------------|----|-----|-----|-----|-----|---|
| Necrosis | CG | 8 | 1.33 | 0.87 | 0 | 3 | 6.858 | 3 | 0.077 |
| MG | 8 | 0.92 | 0.5 | 0 | 2 |
| G-CSF | 8 | 1.04 | 0.46 | 0 | 2 |
| G-CSF-M | 8 | 0.83 | 0.48 | 0 | 2 |
| Hemorrhage | CG | 8 | 1.54 | 0.72 | 1 | 3 | 14.23 | 3 | 0.003 |
| MG | 8 | 1.08 | 0.65 | 0 | 2 |
| G-CSF | 8 | 0.95 | 0.55 | 0 | 2 |
| G-CSF-M | 8 | 0.83 | 0.48 | 0 | 2 |
| Hyperemia | CG | 8 | 1.83 | 0.64 | 1 | 3 | 22.51 | 3 | <0.001 |
| MG | 8 | 1.38 | 0.65 | 0 | 2 |
| G-CSF | 8 | 1.25 | 0.61 | 1 | 3 |
| G-CSF-M | 8 | 0.96 | 0.46 | 0 | 2 |
| Degeneration | CG | 8 | 1.92 | 0.65 | 1 | 3 | 16.9 | 3 | 0.001 |
| MG | 8 | 1.42 | 0.65 | 0 | 2 |
| G-CSF | 8 | 1.29 | 0.62 | 0 | 2 |
| G-CSF-M | 8 | 1.08 | 0.65 | 0 | 2 |
| Cellular infiltrate | CG | 8 | 1.92 | 0.78 | 1 | 3 | 19.01 | 3 | <0.001 |
| MG | 8 | 1.29 | 0.81 | 0 | 3 |
| G-CSF | 8 | 1.54 | 0.59 | 1 | 3 |
| G-CSF-M | 8 | 0.92 | 0.65 | 0 | 2 |

CG = control group; MG = methylprednisolone group; G-CSF = granulocyte colony-stimulating factor group and G-CSF-M = group receiving G-CSF and methylprednisolone.
Table 5 - Multiple comparisons between groups in relation to histological evaluation (Mann-Whitney test).

| Groups     | Comparison | Group | p     |
|------------|-----------|-------|-------|
| Hyperemia  | CG        | CG    | 1.00  |
|            | MG        | 0.608 |
|            | G-CSF     | 0.002 |
|            | G-CSF-M   | <0.001|
|            | MG        | 1.000 |
|            | G-CSF     | 0.311 |
|            | G-CSF-M   | 0.011 |
|            | G-CSF     | 0.002 |
|            | MG        | 0.311 |
|            | G-CSF     | 1.00  |
|            | G-CSF-M   | 0.077 |
|            | G-CSF     | <0.001|
|            | MG        | 0.011 |
|            | G-CSF     | 0.077 |
|            | G-CSF-M   | 1.000 |
| Hemorrhage | CG        | CG    | 1.00  |
|            | MG        | 0.047 |
|            | G-CSF     | 0.005 |
|            | G-CSF-M   | <0.001|
|            | MG        | 0.047 |
|            | G-CSF     | 1.000 |
|            | G-CSF-M   | 0.466 |
|            | G-CSF     | 0.144 |
|            | G-CSF-M   | 0.419 |
|            | G-CSF     | <0.001|
|            | MG        | 0.144 |
|            | G-CSF     | 0.419 |
|            | G-CSF-M   | 1.000 |
|            | G-CSF     | 0.003 |
|            | G-CSF-M   | <0.001|
|            | MG        | 0.019 |
|            | G-CSF     | 0.448 |
|            | G-CSF-M   | 0.076 |
| Degeneration| CG       | CG    | 1.00  |
|            | MG        | 1.000 |
|            | G-CSF     | 0.448 |
|            | G-CSF-M   | 0.076 |
|            | G-CSF     | 0.003 |
|            | MG        | 1.000 |
|            | G-CSF     | 0.003 |
|            | G-CSF-M   | 1.000 |
|            | G-CSF     | 0.016 |
|            | MG        | 0.016 |
|            | G-CSF     | 0.088 |
|            | G-CSF-M   | <0.001|
|            | MG        | 1.000 |
|            | G-CSF     | 0.288 |
|            | G-CSF-M   | 0.089 |
|            | G-CSF     | 0.088 |
|            | MG        | 0.288 |
|            | G-CSF     | 1.00  |
|            | G-CSF-M   | 0.002 |
|            | G-CSF     | <0.001|
|            | MG        | 0.002 |
|            | G-CSF     | 0.002 |
|            | G-CSF-M   | 1.000 |

CG = control group; MG = methylprednisolone group; G-CSF = granulocyte colony-stimulating factor group and G-CSF-M = group receiving G-CSF and methylprednisolone.
scale, that was superior to the improvement obtained with either methylprednisolone or G-CSF alone. The association of methylprednisolone and G-CSF also had a synergistic effect that resulted in an improvement in hyperemia and cellular infiltration at the lesion site.

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AUTHOR CONTRIBUTIONS

Teixeira WG conceived and designed the experiments and methodology, performed the experiments, with data curation, formal analysis and investigations, analyzed the data, contributed to the writing and editing of the manuscript, contributed to project and resources/funding administration. Cristante AF conceived and designed the experiments and methodology, analyzed the data, contributed to the writing and editing of the manuscript, contributed to project and resources/funding administration. Marcon RM conceived and designed the experiments and methodology, performed the experiments, with data curation, formal analysis and investigations, analyzed the data. Bispo G and Ferreira R performed the experiments, with data curation, formal analysis and investigations. Barros-Filho TE conceived and designed the experiments and methodology, analyzed the data, contributed to the writing and editing of the manuscript, contributed to project and resources/funding administration.

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Figure 1 - Examples of hematoxylin-eosin-stained histological slides showing the area of the spinal cord lesion. A - intense degeneration of neural tissue (200x magnification); B - intense hyperemia (400x magnification); C - moderate degree of infiltration and cystic degeneration (100x magnification) and D - an area close to the spinal cord lesion with no alterations (100x magnification).
Teixeira WG et al.

G-CSF and methylprednisolone in SCI

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