A three-dimensional spatial mapping approach to quantify fine-scale heterogeneity among leaves within canopies

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Abstract

Plants are able to survive a variety of environmental conditions due to their ability to detect and respond to their environment. Although ultraviolet (UV) light has the potential to damage DNA and photosynthetic machinery, plants can minimize its negative effects by synthesizing UV-absorbing compounds (Li et al., 1993; Landry et al., 1995) such as flavonoids, hydroxycinnamic acids (HCAs), and mycosporine-like amino acids (Beggs and Wellmann, 1994; Cocksell and Knowland, 1999; Agati et al., 2013). These phenolic secondary metabolites can protect plants from UV damage by decreasing the transmittance of UV photons through tissue. Given that UV-absorbing compounds are energetically expensive to produce (Weing et al., 2004), most plant species plastically upregulate their production in response to UV-exposure (Lois, 1994; Dixon and Paiva, 1995). When these compounds are energetically expensive yet adaptive (i.e., when they increase fitness in the presence of UV, but decrease fitness in the absence of UV), natural selection will favor individuals that optimally allocate these compounds to cells that shield valuable tissues from UV exposure (Weing et al., 2004). For example, UV-absorbing compounds are concentrated within trichomes and/or epidermal cells on the adaxial surface of leaves, which decreases the transmittance of UV-light into the mesophyll cells beneath (Cen and Bornman, 1993; Reuber et al., 1996; Schnitzler et al., 1996; Burchard et al., 2000; Tattini et al., 2000; Bilger et al., 2001; Agati et al., 2002). The reduction in epidermal transmittance in turn reduces damage to photosynthetic machinery (Tevini et al., 1991; Kolb et al., 2001) and DNA (Mazza et al., 2000). There is also evidence that plants nonrandomly distribute UV-absorbing compounds within their reproductive organs (Day and Demchik, 1996a, 1996b) and seeds (Griffen et al., 2004).

The concentration of UV-absorbing compounds may also vary among leaves within canopies. Previous studies have subdivided leaves into broad categories (i.e., “unshaded vs. shaded” or “upper vs. lower canopy”) and found higher concentrations of UV-absorbing compounds in “unshaded” or “upper” leaves (Lovelock et al., 1992; McKee, 1995). However, using categorical variables to quantify leaf position does not sufficiently capture the differences in microenvironments experienced by leaves throughout the entire three-dimensional space of a canopy. For example, leaves on the exterior of canopies alter both the quantity and distribution of UV-absorbing compounds, thereby providing individuals that optimize allocation of these compounds to regions where they are most beneficial.

Methods

We use a three-dimensional spatial mapping approach based on spherical polar coordinates to examine the fine-scale spatial distributions of photosynthetically active radiation (PAR) and the concentration of ultraviolet (UV)-absorbing compounds ($A_{UVA}$) among leaves within the canopies of black mangroves (Avicennia germinans).

Results

Linear regressions revealed that interior leaves received less PAR and produced fewer UV-absorbing compounds than leaves on the exterior of the canopy. By allocating more UV-absorbing compounds to the leaves on the exterior of the canopy, black mangroves may be maximizing UV-protection while minimizing biosynthesis of UV-absorbing compounds.

Discussion

Three-dimensional spatial mapping provides an inexpensive and portable method to detect fine-scale differences in environmental and biological traits within canopies. We used it to understand the relationship between PAR and $A_{UVA}$ but the same approach can also be used to identify traits associated with the spatial distribution of herbivores, pollinators, and pathogens.

Key words: Avicennia germinans; spatial mapping; spherical polar coordinates; UV-absorbing compounds.
and quality of light received by leaves on the interior of canopies (Le Roux et al., 2001; Baldocchi et al., 2002; Valladares, 2003). Although multiple methods exist for quantifying leaf position within a canopy, they are often challenging to implement due to both financial and/or logistical constraints in the field. For example, digitizing methods (i.e., sound propagation: Sinoquet et al., 1991; Room et al., 1996; magnetic field current induction: Sinoquet et al., 1998; Le Roux et al., 2001; Everhart et al., 2011; and LiDAR: Greaves et al., 2015; Magney et al., 2016; Swatantran et al., 2016) are expensive, require electrical power supplies, and fail to access the inner canopy (Magney et al., 2016). Moreover, digitizing methods generate enormous data sets that often require discrete variable distinctions such as flush numbering, stem and branch classifications, or primary and lateral leaf groups to clearly observe trends within the canopy (Sinoquet et al., 1998).

We introduce a simple, inexpensive, and portable method based on a spherical polar coordinate system to quantify the exact location of leaves within the canopy of black mangrove (Avicennia germinans (L.) Stearn) trees. By quantifying leaf position with three continuous variables (radial distance, zenith angle, and azimuthal angle) and a set of mathematical combination variables (radial and vertical canopy depths), we are able to use linear regressions to determine how light quantity changes throughout the canopy and how leaf position within the canopy affects the concentration of UV-absorbing compounds. Black mangrove trees were selected for this study for several reasons. First, as inhabitants of the tropics, black mangroves receive higher levels of UV radiation than plants inhabiting temperate regions (Frederick et al., 1989; Madronich, 1993). Moreover, this species inhabits coastlines and is rarely shaded by taller trees. Consequently, black mangroves have had the opportunity to adapt to this high-UV environment by evolving strategies that minimize UV damage (Lovelock et al., 1992; Weing et al., 2004). Black mangrove trees are also an ideal species because their canopies are large enough to create a multitude of microenvironments, but small enough to sample thoroughly.

**MATERIALS AND METHODS**

**Plant system**—The four black mangrove (A. germinans) trees included in our study were of similar sizes and occurred as single isolated trees along the beach on the west side of Hummingbird Cay, a privately owned island located approximately 13 km west of Georgetown in the Great Exuma island chain of the Bahamas. Geospatial coordinates for the trees are as follows: (23°27′35.2800″N, 75°56′39.8040″W), (23°27′36.8400″N, 75°56′39.8520″W), (23°27′37.2188″N, 75°56′39.8148″W), and (23°27′43.1964″N, 75°56′39.8256″W).

**Quantifying leaf position using spherical polar coordinates**—A spherical polar coordinate system was used to quantify the spatial location of 49–50 leaves within each canopy. This coordinate system uses three continuous variables ($\rho$, $\theta$, $\phi$) to specify leaf location. Approximately 12 leaf samples were collected per quadrant of each canopy, which yielded spatial locations with many combinations of $\rho$, $\theta$, and $\phi$. $\rho$ is the radial distance or the direct distance of a point from a fixed origin, $\theta$ is the zenith angle measured from a fixed vertical direction, and $\phi$ is the azimuthal angle, measured as the angle relative to a fixed direction on a reference plane that passes through the origin and is orthogonal to the vertical direction (Fig. 1). The origin is defined as the trunk of the tree at the base of the canopy. Radial distance ($\rho$) was measured by looping a piece of twine around the trunk of the tree at the base of the canopy. A flexible tape measure was then secured to the twine with 0 cm positioned at the base of the canopy. The twine was drawn to the center of the leaf location, and the direct distance from the trunk to the leaf was measured to the nearest centimeter.

To measure the angular components ($\theta$, $\phi$) of the spatial location, two measurement protractors were prepared from printed 360° protractor images. The protractors were secured to two layers of folder stock, cut to shape, and then laminated. Materials were selected on the basis of availability, ease of construction, and field-portability. The protractor used to measure zenith angle ($\theta$) was prepared by cutting the first 360° protractor in half along the 0° line. A second cut (~55 mm) was made, originating from the flat end of the corresponding 180° protractor, along the 90° line (Appendix 1). The protractor was positioned such that the flat edge was flush with the tree trunk and 0° was oriented directly vertical. Measurements were recorded to the nearest degree (with a spacing of 1.75 mm per degree) and ranged from 0–90°. The protractor used to measure azimuthal angle ($\phi$) was prepared using the second 360° protractor. A ~100 mm cut was made along the 0° line toward the center of the protractor. Three additional cuts (~55 mm) originating from the origin of the protractor were made along the 90°, 180°, and 270° lines (Appendix 2). The single long cut allowed the protractor to be pulled around the base of the canopy, and the three shorter cuts provided space to position the base of the canopy at the origin. The protractor was then secured, leveled using a bubble level, and oriented with 0° positioned at due north using a compass. Measurements of $\phi$ were recorded to the nearest degree and ranged from 0–360°.

The 180° zenith protractor was fitted perpendicular to the azimuthal protractor using the partial cut along 90° line of the zenith protractor (Appendix 3). The angular coordinates were measured using the direct path of the twine to the leaf location relative to the two protractors. Using this method, we are able to spatially map the canopy at the resolution scale of individual leaves. Measurement errors in $\rho$, $\theta$, and $\phi$ are ±1 cm, ±1°, and ±1°, respectively, and are below the scale of individual leaves.

**Quantifying PAR and UV-absorbing compounds**—After measuring the spherical polar coordinates of each leaf, the amount of photosynthetically active radiation (PAR) it received was determined by placing the sensor of a handheld light meter (LI-250; LI-COR, Lincoln, Nebraska, USA) at the adaxial leaf surface. To minimize variation in full sun (unobstructed by leaves) PAR, we only collected data between 12:00 p.m. and 3:00 p.m. on sunny days and we sampled all 49–50 leaves within a single canopy on the same day. Full sun PAR during data collection ranged from 1566 to 1719 µmol m$^{-2}$ s$^{-1}$. Collecting data from a subset of leaves from each tree each day would have required that we spend more time sampling each day and consequently would have increased the range of full sun PAR levels during sampling.

Following the measurement of PAR, a leaf disk (31.2 mm$^{2}$) was collected from the middle of the leaf at its widest part using a cork borer. Disks were placed in individual 5-mL Eppendorf tubes containing 4 mL of a 90:1:1

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http://www.bioone.org/loi/apps/2017/5/11/1700056
Spatial distribution of PAR—The amount of PAR received by the leaves ranged from 76.5 to 1718.7 μmol m⁻² s⁻¹ and depended on the leaf position (Table 1, Fig. 3A). Overall, leaves on the exterior of the canopy received more PAR than leaves on the interior. Although PAR was negatively correlated with $R_D$ ($β = -10.6088$, $P < 0.0001$), it was not significantly correlated with the other spatial variables $θ$ and $ϕ$ or any interaction term combinations of the spatial variables (Table 1). The strong dependence of PAR on $R_D$ is consistent with an internal shading model within the canopy, where the pigments within outer leaves reduce the amount of radiation that reaches the interior leaves.

Spatial distribution of UV-absorbing compound concentrations ($A_{300}$) —The concentration of UV-absorbing compounds varied among leaves throughout the canopy (i.e., individual $A_{300}$ values ranged from 0.301 to 1.089). This variation was explained in part by the position of the leaf within the canopy (Table 1, Fig. 3B). Much like the observed spatial trends in PAR, $A_{300}$ was negatively correlated with $R_D$; leaves at the exterior of the tree canopy contained higher concentrations of UV-absorbing compounds than leaves on the interior of the canopy ($β = -0.0041$, $P < 0.0001$). Also similar to PAR, $A_{300}$ was not significantly correlated with any of the other spatial variables or the interaction terms.

Vertical canopy depth ($V_D$) —Both PAR and $A_{300}$ were negatively correlated with $V_D$ ($P < 0.0001$; Table 2). To understand this observation, we examined the independent spatial variables that can contribute to $V_D$ (i.e., $R_D$ and $θ$). Although PAR and $A_{300}$...
were both correlated with $R_D$, neither showed a significant correlation with $\theta$ or the $R_D \cdot \theta$ interaction term in the linear regression analysis, indicating that the effect of $R_D$ on PAR and $A_{300}$ was independent of $\theta$. Consequently, the observed correlation of PAR and $A_{300}$ with $V_D$ was driven by $R_D$ and was not significantly influenced by $\theta$.

**DISCUSSION**

We have developed a simple, economical, and portable method based on a spherical polar coordinate system to quantify the exact location of leaves within the canopy. By defining leaf position with three continuous variables in spherical coordinates, our three-dimensional spatial mapping technique can detect fine-scale heterogeneity in both environmental conditions (i.e., PAR) and biological traits (i.e., $A_{300}$) throughout the entire canopy of individual trees. Data collected using this method can be analyzed using multiple linear regressions to determine the degree to which each spatial variable (i.e., $R_D$, $\theta$, and $\phi$) and combination of spatial variables (i.e., their interaction terms) explain the variation observed.

In our application of this method, we found that the spatial variation in both PAR and $A_{300}$ within the canopy of black mangrove ($A.\text{germinans}$) trees was due to $R_D$. The shared $R_D$-dependence of PAR and $A_{300}$ suggests that the spatial distribution of light within canopies could determine the spatial distribution of UV-absorbing compounds. Multiple studies have observed increased production of photoprotective flavonoids and HCAs upon increased light exposure for plant species of diverse geographical origins (McKee, 1995; Schnitzler et al., 1996; Tegelberg et al., 2001; Izaguirre et al., 2007; Sullivan et al., 2007; El Morchid et al., 2014; Nascimento et al., 2015). In a manipulative growth chamber experiment, black mangrove seedlings grown in high light (339 ± 4 μmol m$^{-2}$ s$^{-1}$) had 52% more phenolics than those grown in low light (37 ± 1 μmol m$^{-2}$ s$^{-1}$) (McKee, 1995). The observed significant negative correlations of PAR and $A_{300}$ with $V_D$ in our study suggest that the light microenvironment and the biosynthesis of UV-absorbing compounds are also coupled in adult trees in the field. Moreover, these results provide a plausible framework for explaining how the canopy distributes UV-absorbing compounds. Rather than produce a uniform concentration of UV-absorbing compounds across the entirety of the canopy, black mangroves preferentially allocate more UV-absorbing compounds to the outermost leaves, which receive more incoming solar radiation.

Other methods have been used to quantify the fine-scale environmental and biological heterogeneity within canopies. For example, leaf position has been quantified using leaf area index (Hirose and Werger, 1987; Pierce and Running, 1988; Ellsworth and Reich, 1993; Bréda, 2003; Weiss et al., 2004; Li et al., 2014; Magney et al., 2016) and vertical canopy depth (Ellsworth and Reich, 1993; Sinoquet et al., 1998; Magney et al., 2016). However, leaf area index methods often require either direct harvesting or the absorbance at 300 nm ($A_{300}$) within the canopy based on the combined leaf measurements from four black mangrove ($Avicennia\text{germinans}$) trees on a unitless Cartesian scale. Orientation of the composite canopy relative to the cardinal directions is displayed along the x-y plane.

![Fig. 3. Three-dimensional spatial distribution of (A) photosynthetically active radiation (PAR) and (B) UV-absorbing compound concentration as indicated by the absorbance at 300 nm ($A_{300}$) within the canopy based on the combined leaf measurements from four black mangrove ($Avicennia\text{germinans}$) trees on a unitless Cartesian scale. Orientation of the composite canopy relative to the cardinal directions is displayed along the x-y plane.](http://www.bioone.org/loi/apps)
Overall, we have demonstrated that three-dimensional mapping can be used to probe the structure-property relationships of both environmental (i.e., PAR) and biological (i.e., $A_{300}$) measurements. The significant negative correlation of $R_D$ with both PAR and $A_{100}$ in our study provides vital clues regarding how the canopy structure efficiently balances the need for photoprotection with the corresponding energetic cost of biosynthesis. Using the spherical polar coordinate system to quantify leaf position, however, is not limited to studying the relationship between light and pigments. This simple, inexpensive, and portable method can also be used to more thoroughly understand how plants are affected by and respond to any abiotic or biotic factor, including quantifying how intraplant variation in leaf or floral traits is linked to spatial patterns of herbivory, the colonization and proliferation of microbes, and the effectiveness of pollinators.

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Appendix 1. Zenith angle ($\theta$) protractor construction template. Appropriate cuts and their respective orders are indicated by the dashed lines and numbers. Image reproduced from https://commons.wikimedia.org/wiki/File:Rapporteur.svg#filelinks (Rapporteur.svg, Autiwa) under a CC BY-SA 3.0 license (https://creativecommons.org/licenses/by-sa/3.0/legalcode). Overlaid images by J. D. Patterson.

Appendix 2. Azimuthal angle ($\phi$) protractor construction template. Appropriate cuts and their respective orders are indicated by the dashed lines and numbers. Image reproduced from https://commons.wikimedia.org/wiki/File:Rapporteur.svg#filelinks (Rapporteur.svg, Autiwa) under a CC BY-SA 3.0 license (https://creativecommons.org/licenses/by-sa/3.0/legalcode). Overlaid images by J. D. Patterson.

Appendix 3. Schematic illustrating the arrangement of the zenith ($\theta$) protractor and azimuthal ($\phi$) protractor at the base of the canopy.