Demonstration That CobG, the Monooxygenase Associated with the Ring Contraction Process of the Aerobic Cobalamin (Vitamin B$_{12}$) Biosynthetic Pathway, Contains an Fe-S Center and a Mononuclear Non-heme Iron Center*

Susanne Schroeder$^{1,1}$, Andrew D. Lawrence$^{1,1}$, Rebekka Biedendieck$^{1}$, Ruth-Sarah Rose$^{1}$, Evelyne Deery$^{1}$, Ross M. Graham$^{2}$, Kirsty J. McLean$^{2}$, Andrew W. Munro$^{4}$, Stephen E. J. Rigby$^{1}$, and Martin J. Warren$^{1}$

From the $^1$Protein Science Group, Department of Biosciences, University of Kent, Canterbury, Kent CT27NJ, United Kingdom, the $^2$School of Medicine and Pharmacology (M704), University of Western Australia, Fremantle Hospital, Fremantle, 6160 Western Australia, Australia, and the $^4$Faculty of Life Sciences, Manchester Interdisciplinary Biocentre, University of Manchester, Manchester M1 7DN, United Kingdom

The ring contraction process that occurs during cobalamin (vitamin B$_{12}$) biosynthesis is mediated via the action of two enzymes, CobG and CobJ. The first of these generates a tertiary alcohol at the C-20 position of precorrin-3A by functioning as a monooxygenase, a reaction that also forms a gamma lactone with the acetate acid side chain on ring A. The product, precorrin-3B, is then acted upon by CobJ, which methylates at the C-17 position and promotes ring contraction of the macrocycle by catalyzing a masked pinacol rearrangement. Here, we report the characterization of CobG enzymes from Pseudomonas denitrificans and Brucella melitensis. We show that both contain a [4Fe-4S] center as well as a mononuclear non-heme iron. Although both enzymes are active in vivo, the P. denitrificans enzyme was found to be inactive in vitro. Further analysis of this enzyme revealed that the mononuclear non-heme iron was not reducible, and it was concluded that it is rapidly inactivated once it is released from the bacterial cell. In contrast, the B. melitensis enzyme was found to be fully active in vitro and the mononuclear non-heme iron was reducible by dithionite. The reduced mononuclear non-heme was able to react with the oxygen analogue NO, but only in the presence of the substrate precorrin-3A. The cysteine residues responsible for binding the Fe-S center were identified by site-directed mutagenesis. A mechanism for CobG is presented.

Vitamin B$_{12}$ is the antipernicious anemia factor (1), whose isolation was first described over 60 years ago (2, 3), and whose subsequent structure determination by x-ray crystallography revealed that it is a cobalt-containing modified tetrapyrrole (4). As such it belongs to the same family of compounds as heme, chlorophyll, siroheme, and coenzyme F$_{430}$ (5). The major structural differences between vitamin B$_{12}$ and these other family members include the shrunken macrocyclic ring that represents the corrin ring component of the vitamin and the presence of upper and lower axial ligands for the centrally chelated metal ion. There has been some considerable interest in the biosynthesis of vitamin B$_{12}$ as there are no chemical or biological parallels for the removal and extrusion of the integral meso carbon atom that is required for the contraction of the macrocycle (6).

The coenzyme form of vitamin B$_{12}$ is adenosylcobalamin (Fig. 1), which is synthesized by one of the most elaborate and complex biosynthetic pathways known in biological systems requiring somewhere around thirty enzymatic steps for its complete de novo synthesis (5). Cobalamin biosynthesis is further complicated by the presence of two similar though biochemically distinct pathways that are referred to as the aerobic and anaerobic routes (7–10). These pathways differ in their requirement for molecular oxygen and the timing of cobalt insertion. The aerobic pathway requires molecular oxygen to facilitate the ring contraction process prior to cobalt insertion, whereas on the anaerobic route there is no requirement for oxygen but early cobalt insertion is an essential prerequisite for ring contraction.

Along the aerobic pathway for cobalamin biosynthesis, the ring contraction process is mediated by two enzymes called CobG and CobJ (11, 12) (Fig. 1). Initially, CobG hydroxylates the C-20 position of precorrin-3A and generates a gamma lactone structure with the acetate side chain of ring A to give precorrin-3B (13). CobG utilizes molecular oxygen, which was first shown when it was demonstrated that the enzyme was inactive under anaerobic conditions (12). It was only when the enzyme was incubated with precorrin-3A in the presence of air that any activity was observed. Subsequently, elegant labeling experiments with $^{18}$O$_{2}$ revealed that one atom of oxygen was incorporated into the C-20 position (14, 15). Moreover, the absolute stereochemistry of precorrin-3B was determined, revealing that the C-20 hydroxyl is cis to the oxygen terminus of the gamma-lactone at C-1 (16). This reaction prepares the macrocycle for contraction by CobJ via a pinacol-type rearrangement. The extruded carbon frag-
ment is eventually lost as acetic acid. These steps are outlined in Fig. 1.

Despite the characterization of the reaction catalyzed by CobG, comparatively little is known about the enzyme itself. CobG is a 459-amino acid protein, with a molecular mass of 46 kDa, and displays similarity to nitrite and sulfite reductases (11). Sulfite reductase, which catalyzes the six-electron reduction of sulfite to sulfide, requires both a [4Fe-4S] cluster and siroheme for activity. In this unusual enzyme, the two cofactors are covalently linked through a shared cysteine residue (17–19).

On the basis that purified CobG was found to have a slight coloration and that atomic absorption analysis of the purified protein indicated that it contained 4.6 mol of Fe and 4 mol of S per mol of protein, it was suggested that the protein is likely to contain both a [4Fe-4S] cluster and a mononuclear non-heme iron as well as a [Fe-S] cluster (9). In this report, we provide the first biophysical evidence that CobG does indeed contain both a mononuclear non-heme iron as well as a [Fe-S] cluster. We also show that the substrate must bind in close proximity to these metalloprosthetic groups. Moreover, the cysteine residues required for Fe-S formation are confirmed and the presence of the Fe-S center is shown to be essential for catalytic activity of the enzyme.

**EXPERIMENTAL PROCEDURES**

**Chemicals and Reagents**—Most chemicals were purchased from Sigma. Other materials were provided by the following suppliers: restriction enzymes, modification enzymes, and the vector pGEM-T Easy were from Promega and New England Biolabs UK; chelating-Sepharose fast flow resin was from Amersham Biosciences, Little Chalfont, Bucks, UK; pET3a, pET14b, pLysS, and pETcoco2 were from Novagen, Madison, WI; tryptone and yeast extract were from Oxoid, Basingstoke, UK; sodium dithionite was from Roche, Poole, UK; and primers were from Invitrogen, Paisley, UK.

**Recombinant DNA Techniques**—All strains and plasmids used in this study are listed in Table 1. DNA manipulations were performed in *Escherichia coli* JM109 using established laboratory procedures. Plasmid DNA was isolated using the QIAprep Miniprep kit (Qiagen). DNA fragments were isolated from 0.7% (w/v) or 1.0% (w/v) agarose gels and purified by using the QIAquick Gel Extraction kit (Qiagen). PCR products were purified by using the QIAquick PCR purification kit (Qiagen). DNA sequencing was performed at GATC Biotech, Germany.

**Cloning of cobG from Brucella melitensis and Pseudomonas denitrificans**—*B. melitensis cobG* (*Bmei0715*) was amplified by PCR from genomic DNA from *B. melitensis* strain 16 M. Primers contained the restriction sites AseI and SpeI (underlined) and the sequences (5'-3') were actattaatatgcgcaacgctccttttccc and actactagttattgatgtcgggcgttcag. The resulting PCR product was ligated into the pGEM-T Easy vector. The cobG insert was excised using the AseI and SpeI sites and ligated into the NdeI and SpeI sites of the vectors pET-14b and pET-3a. The *P. denitrificans cobG* was amplified by PCR using the plasmid pCR427 as a template and was directly cloned into the NdeI and SpeI sites of pET14b. The sequences of the cobG inserts were confirmed by sequencing.

**Site-directed Mutagenesis**—Site-directed mutagenesis was performed using the QuikChange II XL site-directed mutagenesis kit (Stratagene) to generate point mutations in the plasmid pRS003. The sequences of the resulting plasmids were confirmed by DNA sequencing. The mutagenic primers used are listed in Table 2.

**Recombinant Protein Overproduction and Purification**—For protein overproduction, the *E. coli* strain BL21(DE3) pLysS was transformed with the appropriate plasmid. The resulting strain was grown at 37 °C in LB supplemented with ampicillin (100 mg/liter) and chloramphenicol (34 mg/liter). Once the cell density reached $A_{600} \approx 0.6$, protein overproduction was induced.
with the addition of 0.4 mM isopropyl-1-thio-β-D-galactopyranoside, and the cells were left to grow overnight at 16 °C before harvesting by centrifugation at 3,500 × g for 15 min at 4 °C (Beckman Coulter, JI30). The pellet was gently resuspended in binding buffer (20 mM Tris-HCl, pH 8.0, containing 0.5 mM NaCl, 5 mM imidazole), and the cells were broken either by one passage through a cell disrupter (Stansted Fluid) operating at 12,000 psi or by sonication (Sonics Vibracell Ultrasonic Processor). Cell debris was removed by centrifugation at 35,000 × g for 20 min at 4 °C. The resulting supernatant was transferred to an anaerobic glove box (Belle Technology), maintained at least 2 ppm oxygen, and applied to a metal affinity chromatography column charged with Ni²⁺. The column was washed with buffer containing 20 mM imidazole before elution with 400 mM imidazole.

In Vivo Assays—The in vivo activity of CobG was determined through complementation using a strain containing all the necessary genes for the production of cobyclic acid in E. coli with the exception of cobG. The wild type and mutant cobG genes were excised from the vector pET14b using the XbaI and HindIII restriction sites and ligated into the SpeI and HindIII sites of pLysS. An E. coli strain containing the plasmid pET3a-cobG was transformed with the resulting plasmids and restoration of cobyric acid production in E. coli except for cobG was successful. The activity of CobG was determined in vivo for 20 min at 4 °C. The resulting supernatant was for 15 min at 4 °C. The resulting supernatant was

**In Vitro Assays**—CobG activity was measured using a coupled multi-enzyme system where the production of hydrogenobyrinic acid (HBA)³ could be easily monitored. To achieve this, a plasmid (pSS200) was constructed that harbored all the genes for cobyric acid production in E. coli with the exception of cobG. pET3a vector carrying all genes required for hydrogenobyrinic acid production in E. coli [AI(DG)FMKLEHNSTRWQ] was transformed with pETcoco2[AI(DG)FMKLEHNSTRWQ] and pET3a-cobG. The resulting plasmids and restoration of cobyric acid production in E. coli were confirmed. The wild type and mutant cobG genes were excised from the vector pET14b using the XbaI and HindIII restriction sites and ligated into the SpeI and HindIII sites of pLysS. An E. coli strain containing the plasmid pET3a-cobG was transformed with the resulting plasmids and restoration of cobyric acid production in E. coli was confirmed.

**Table 2**

Primer sites are underlined and the nucleotides exchanged are shown in capital letters.

| Primer name | Sequence forward primer | Sequence reverse primer |
|-------------|-------------------------|-------------------------|
| BmelcobG C42A Nael | ggcgccaggtccGGGcAaggcttc | gagagccgttccGCGcactgtgcc |
| BmelcobG C358A Nael | ctggtgccccggGCGtcagttc | cgcttgtccacgttcGCGcactgttg |
| BmelcobG C364A Nael | ggtggcaggtcGCGcactgttc | cgagagccgttccGCGcactgttg |
| BmelcobG C394A Nael | gggcgcaggtcGCGcactgttc | cgagagccgttccGCGcactgttg |
| BmelcobG C398A Putl | gggcgcaggtcGCGcactgttc | cgagagccgttccGCGcactgttg |

The abbreviations used are: HBA, hydrogenobyrinic acid; ALA, 5-aminolevulinic acid; NO, nitric oxide; SAM, S-adenosyl-L-methionine; EPR, electron paramagnetic resonance.
genes necessary for the production of HBA except cobG. This plasmid, pSS200, was derived from pED420 and pRS005 by cloning a MfeI/SpeI fragment of pRS005 into pED420, which had been cut with the same enzymes. Cleared lysates of an E. coli strain harboring the plasmid pSS200 were incubated with crude extract of strains overproducing CobG in 20 mM Tris-HCl buffer, pH 8.0, containing 100 mM NaCl, 720 μg/ml S-adenosyl-L-methionine (SAM), 120 μg/ml 5-aminolevulinic acid (ALA), and 100 μg/ml NADPH for 18–20 h under aerobic conditions in the dark at 4 °C.

Preparation of Precorrin-3A—Precorrin-3A was synthesized in a multi-enzyme reaction containing HemB, HemC, HemD, CobA, and CobI. The proteins were purified from two E. coli strains harboring the plasmids pRS004 and pED408 by metal affinity chromatography under both aerobic and anaerobic conditions. The purified proteins migrated as a single band with a molecular mass of around 50 kDa when analyzed by SDS-PAGE (Fig. 2b). Purified fractions containing either CobG protein were dark green-brown in color and exhibited a broad absorbance around 415 nm (Fig. 2a).

Isolation of HBA—HBA was purified in its free acid form by initially heating either incubations or extracts containing the compound to 80 °C for 15 min. The extracts were then applied to a small DEAE-Sephadex column and the HBA eluted in 1 M NaCl. The HBA was quantified by high performance liquid chromatography on a BDS Hypersil C-18 column (4.6 × 250 mm; Thermo Electron Corporation) run on a Agilent 1100 series LC/MSD Trap equipped with a diode array detector and eluted at a flow rate of 1 ml min⁻¹ with a gradient of acetonitrile in 1 M ammonium acetate.

Electron Paramagnetic Resonance (EPR) Spectroscopy—EPR spectra were obtained using a Bruker ELEXSYS E500 spectrometer operating at X-band. Temperature control was maintained using an Oxford Instruments ESR900 cryostat linked to an ITC503 controller. Experimental conditions were as given in the figure captions. Nitric oxide gas was formed by the reaction of sodium nitrite with FeSO₄ (1.2 mM) in H₂SO₄ (1.8 M) under anaerobic conditions and collected in a syringe. Nitric oxide gas was injected into a sealed vial (2-ml HPLC vial) containing 300 μl of sample. Samples were incubated for 5 min before being transferred into EPR tubes and rapidly frozen in liquid nitrogen.

Redox Potentiometry—Redox titrations were performed in a Belle Technology glove box under a nitrogen atmosphere as described previously (21–23).

RESULTS

For the purpose of this research the CobG proteins from B. melitensis and P. denitrificans were studied. Details of the cloning of the respective cobG genes into suitable expression vectors are given in the experimental section. The cloning allowed the two proteins to be produced recombinantly in E. coli as His-tagged fusions.

Purification of Recombinant CobG—Purifications of the P. denitrificans and B. melitensis CobG proteins were achieved by metal affinity chromatography under both aerobic and anaerobic conditions. The purified proteins migrated as a single band with a molecular mass of around 50 kDa when analyzed by SDS-PAGE (Fig. 2b). Purified fractions containing either CobG protein were dark green-brown in color and exhibited a UV-visible spectrum indicative of an iron-sulfur (Fe-S) protein with a broad absorbance around 415 nm (Fig. 2a). Preparations typically exhibited an A₄00/A₃20 ratio between 0.28 and 0.36. When the CobG proteins were purified aerobically, the intensity of the absorbance around 415 nm decreased with time, over a period of a few hours, suggesting that the iron-sulfur center was sensitive to oxygen.

His-tagged CobG Is Active in E. coli—There are no cobG mutants available that can be used in complementation studies. We therefore made a recombinant E. coli strain with all the genetic material to allow the biosynthesis of HBA except for cobG. HBA is the first stable ring-contracted molecule that is easily isolated from the aerobic pathway. This strain accumulates precorrin-3A as it does not have the capacity to make the C-20 hydroxylated and gamma lactone-containing intermediate, precorrin-3B. However, transformation of this E. coli strain with a plasmid containing either the B. melitensis or P. denitrificans cobG resulted in it being able to make HBA (Fig. 2c). This demonstrates that both enzymes are active as recombinant His-tagged proteins within E. coli.

The activity of the CobG proteins was also investigated in vitro. Here, cell lysates enriched with the enzymes required for uroporphyrinogen III synthesis (HemB, -C, and -D) and all the enzymes for HBA synthesis (CobA, -I, -J, -T, -M, -K, -L, -E, -H, and the B. melitensis CobG. In the presence of the B. melitensis CobG, the assay yielded HBA, which was identified by HPLC and mass spectrometry. In contrast, however, the P. denitrificans CobG was found to be inactive. Thus, it would appear that P. denitrificans CobG is very unstable once it is released from the intact bacterial cells.

FIGURE 2. Characterization of the CobG proteins used in this study. a, UV-visible spectrum of B. melitensis CobG, where an absorption of around 445 nm is indicative of a [4Fe-4S] center. b, inset, SDS gel showing the purified P. denitrificans and B. melitensis CobG proteins. Both proteins migrate with a molecular mass of around 50 kDa. c, spectrum of HBA synthesized in an in vitro assay that contained CobA, -I, -J, -T, -M, -K, -L, -E, -H, and the B. melitensis CobG.
Characterization of P. denitrificans CobG—The UV-visible absorption spectrum of the purified P. denitrificans CobG is characteristic of a Fe-S protein, and the broad absorption in the region of 415 nm is indicative of a [4Fe-4S] center (24, 25). However, to confirm the presence of a putative Fe-S center and to determine the type of center present ([2Fe-2S], [3Fe-4S], or [4Fe-4S]) an electron paramagnetic resonance (EPR) study was conducted. EPR samples were prepared from CobG that had been purified anaerobically in a glove box and to a concentration of >250 μM. The EPR spectrum of the dithionite reduced CobG, recorded at 15 K, shows features at g = 2.048 and g = 1.927 (Fig. 3a). This EPR signal was not observed at 70 K or in oxidized samples, which precludes it being a [2Fe-2S] or [3Fe-4S] center because of the differences in the properties of the delocalized electrons in the two types of cluster. A [3Fe-4S] center is easily discounted as such centers contain an even number of electrons in the reduced state and thus do not give rise to an EPR signal when reduced. Furthermore their EPR spectra are very weakly axial and appear as an almost isotropic line at g ≈ 2.02 in the oxidized state [3Fe-4S] (26). It is more difficult to distinguish signals arising from [2Fe-2S] and [4Fe-4S]1+, the paramagnetic reduced states of the [2Fe-2S] and [4Fe-4S] clusters. Both give rise to axial, pseudo-axial or rhombic spectra with at least one g value below 2.0 due to g-anisotropy (27). However, due to the nature of the antiferromagnetic coupling (exchange and double exchange coupling) there are several low lying energy levels available in the [4Fe-4S]1+ system that do not exist in the [2Fe-2S]1+ system, see (27) for further discussion of energy levels in Fe-S systems. The availability of these energy levels promotes electronic relaxation, leading to very short electron spin relaxation times (28). These relaxation times are so short as to effectively preclude the observation of the [4Fe-4S]1+ signal at temperatures above 40 K. Collectively, the measured g values and the observed temperature dependence of the signal allow the spectrum to be safely assigned to a [4Fe-4S]1+ cluster (24, 29).

CobG Contains a Mononuclear Non-heme Iron Center—Because CobG does not contain a bound heme or flavin cofactor it has been speculated that the activation of molecular oxygen by CobG may be achieved through a mononuclear non-heme iron center (9). Using EPR spectroscopy it has been possible to confirm that CobG does indeed contain a bound mononuclear non-heme iron center. The EPR spectrum, which was observed in both the oxidized and dithionite-reduced samples, with g = 4.08 (Fig. 3b) is assigned to a high spin (total spin S = 5/2) monomeric non-heme Fe3+ ion in a rhombic ligand field. The EPR line shapes of high spin mononuclear Fe3+ centers are dominated by zero field splitting (30). These rise from the effects of the ligand arrangement around the ion on the energies of the five electronic orbitals populated by the unpaired electrons. Different ligand arrangements lift the degeneracy in the energies of these orbitals in different ways, thus zero field splitting is indicative of the ligand geometry at the ion and is very sensitive to changes in this geometry. The zero field splitting is described by two parameters, the axial, D, and rhombic, E, zero field splitting parameters. When E/D ≈ 1/3, indicating a rhombic ligand field, the EPR spectrum consists of a single line at g ≈ 4.3. The line width of the EPR spectrum we observed is somewhat larger than might be expected for a typical mononuclear Fe3+ ion center. This may arise from E and D values that give E/D of slightly less than 1/3, deviation from ideal rhombic ligand field, or from heterogeneity in the sample leading to an overlap of many different spectra with slightly different E/D. To be catalytically active the iron center would need to be reduced to Fe2+ prior to the binding and activation of dioxygen. However, reduction of the mononuclear non-heme iron, which would be observable as a loss of the EPR signal since Fe2+ (S = 2) is EPR silent, could not be achieved using either sodium dithionite, sodium borohydride, or ascorbate as the reducing agent. Furthermore, the addition of precorrin-3A or either α-ketoglutarate or pterin, which are known to be cofactors for mononuclear non-heme iron proteins (31), had no effect on the EPR spectra and/or the reduction of the iron center by dithionite. Conversely, by this method it was not possible to determine if any Fe2+ was already present in the sample. The lack of reducibility of this iron species may explain why the P. denitrificans enzyme was found to be inactive when it was assayed in vitro.

Midpoint Redox Potential for the P. denitrificans CobG—The midpoint redox potential for the P. denitrificans CobG was determined by redox potentiometry using anaerobic spectroelectrochemical methods, as described in previous studies (21–23). As we have observed recently in studies of the 4Fe-4S center in the PduT protein from Citrobacter freundii (32) and also in our work on the Bacillus megaterium CbiX protein (33), there are only small changes in the optical spectrum of PduT as the iron-sulfur cluster is reduced from the [4Fe-4S]2+ to the [4Fe-4S]1+ form. Nonetheless, CobG was readily and reversibly titrated between these forms, enabling an accurate determination of the midpoint potential of the cluster as −212 mV (Fig. 4).

Characterization of B. melitensis CobG—X-band EPR spectra of purified CobG protein showed no signals under anaerobic conditions. However, taking the sample out of the glove box and exposing it to air for 10 min prior to its EPR analysis gave rise to a signal at g = 4.31 (Fig. 5b). This signal is narrow and well defined unlike signals that are normally attributed to
adventitious or free iron. It arises from a mononuclear high spin ferric iron center with a rhombic geometry (30), as described above for the enzyme from P. denitrificans. This EPR spectrum is typical of high spin mononuclear non-heme Fe3+/H11001 centers with E/D1/3. Despite the presence of oxygen, there is no evidence for the presence of a low spin ferric peroxide species with g values of 2.19, 2.12, and 1.95 (34), or a rhombic O2-coordinated species with g values of 5.9 and 7.5 (35). Such species have been observed previously in studies of bleomycin and chemical models of mononuclear non-heme iron centers, though they are rarely observed in proteins (36, 37). The addition of dithionite to the anaerobic protein gave rise to the spectrum of Fig. 5a. This axial spectrum with g values 2.04 and 1.93 was not be observed at 70 K, indicating that it derives from a [4Fe-4S]1+ cluster (27). Thus the enzyme isolated anaerobically possesses a ferrous mononuclear non-heme iron center and a [4Fe-4S]2 cluster. Although the dithionite-reduced protein contains a [4Fe-4S]1+ cluster and an S = 2 high spin ferrous iron, there is no evidence for a strong magnetic interaction between these two centers as the form and behavior of the [4Fe-4S]1+ signal is as expected for an isolated 4Fe-4S center.

The addition of 1–2 equivalents of precorrin-3A to the anaerobically purified protein had no effect on the EPR spectrum. The presence of precorrin-3A did, however, affect both the EPR signals previously observed in its absence and described above (Fig. 6, a and b). These small but reproducible
Characterization of CobG

![Image](60x575 to 288x734)

**FIGURE 7.** X-band EPR spectra of *B. melitensis* CobG treated with NO. **a,** in the absence and **b,** in the presence of precorrin-3A. Experimental conditions: microwave power, 0.5 milliwatt; modulation amplitude, 10G; temperature, 15K; each spectrum is the average of six scans.

**TABLE 3**

| Mutant of CobG | Activity | Iron sulfur center present |
|----------------|----------|---------------------------|
| C42A           |          |                           |
| C358A          | -        |                           |
| C364A          | -        |                           |
| C394A          | -        |                           |
| C398A          | -        |                           |
| F390A          | +        |                           |
|                |          |                           |

Effects suggest that precorrin-3A does not bind directly to the mononuclear non-heme iron or Fe-S centers. Note that the small but reproducible effect on the mononuclear non-heme iron signal further supports our assertion that this signal arises from a protein-bound mononuclear non-heme iron site and not from adventitiously bound or free iron.

Nitric oxide is an oxygen analogue that forms a paramagnetic iron-nitrosyl species upon binding to a reduced mononuclear non-heme iron (38). Binding of NO to the mononuclear non-heme iron in CobG would provide evidence that this cofactor is indeed the site of oxygen activation. The addition of 5 equivalents of NO to the anaerobic enzyme does not give rise to signals in the g = 4 region (Fig. 7a). However, in the presence of precorrin-3A the same concentration of NO does produce a weak signal at g = 4.28, Fig. 7b, which is typical of ferro-NO complexes. Such complexes have been observed in protocatechuate 4,5-dioxygenase, lipoxygenase, isopenicillin N-synthase, and other iron-sulfur center-containing proteins, and no EPR spectra could be detected consistent with the absence of a Fe-S center (data not shown). In contrast, the iron-sulfur center remained bound in both the C42A and H390A mutants, as deduced by UV/VIS spectroscopy and EPR, and both these mutant CobG enzymes retained activity in the *in vitro* assay system. These findings suggest that the cysteine residues in the C terminus of the protein coordinate the [4Fe-4S] center and that the iron-sulfur center plays an important role in the activity of *B. melitensis* CobG.

**DISCUSSION**

CobG has been successfully overproduced as an N-terminal His-tag fusion protein in *E. coli* and purified by metal affinity chromatography using a column loaded with Ni^{2+}. The UV-visible spectrum of the purified material was indicative of the presence of an iron-sulfur center, which was later confirmed by EPR spectroscopy. The EPR spectrum of the iron-sulfur center was only obtainable after reduction of the protein with dithionite. Significantly, when the temperature was raised from 15 to 70 K the signal disappeared. This marked temperature dependence allowed the signal to be assigned to a [4Fe-4S] center (27).

Previously, the presence of this redox center had been implied but never shown experimentally. Site directed mutagenesis suggests that the Fe-S center is formed between four conserved cysteine residues at the C-terminal region of the protein, cysteines 358, 364, 394, and 398. The presence of the Fe-S center appears to be essential for the activity of the enzyme, as demonstrated by our activity complementation and optical studies on the relevant cysteine replacement mutant proteins.

EPR spectroscopy was also used to identify a bound Fe^{3+} mononuclear non-heme iron center in *P. denitrificans* CobG that is a possible site for oxygen activation. To bind and activate dioxygen this site would need to be reduced to Fe^{2+}, which is...
then EPR silent. However, this mononuclear non-heme iron could not be reduced by dithionite or borohydride even in the presence of a wide range of known cofactors for mononuclear non-heme oxygenases or substrate (precorrin-3A). Thus, the center in *P. denitrificans* CobG may be either kinetically/structurally blocked to reduction by these agents, or else have a quite negative reduction potential. In *B. melitensis* CobG the mononuclear non-heme iron site is isolated in the Fe$^{2+}$/H$_{11001}$ oxidation state and the addition of oxygen is required to generate the Fe$^{3+}$/H$_{11001}$ state with a *g* = 4.31 EPR signal. This signal is very similar to that observed in ferric lipoxygenase and protocatechuate 4,5-dioxygenase (39, 40). The mononuclear non-heme Fe$^{3+}$/H$_{11001}$ signal is affected by the addition of precorrin-3A, although the effect is slight, suggesting that it does not arise from direct coordination of the iron by the tetrapyrrrole, but rather that precorrin-3A binds close to the mononuclear non-heme iron site as in lipoxygenases (44). Direct binding of substrates or inhibitors to the mononuclear non-heme Fe$^{3+}$ center, as is observed in protocatechuate 4,5-dioxygenase, affects the zero field splitting parameters E and D as a new ligand arrangement around the Fe$^{3+}$ ion is created. In protocatechuate 4,5-dioxygenase such binding gives rise to a rhombic spectrum with *g* = 6.4, 5.5, and 2.0 (40). This is obviously a much larger effect than is observed here. Nitric oxide, which is an oxygen analogue, will form a paramagnetic iron-nitrosyl species upon binding to a Fe$^{2+}$/H$_{11001}$ mononuclear non-heme iron. The EPR spectrum of *B. melitensis* CobG in the presence of low concentrations of NO, however, failed to show an iron-nitrosyl signal. Significantly, though, the addition of precorrin-3A to a sample containing the same concentration of NO did exhibit such an iron-nitrosyl signal at *g* = 4.28. This suggests that precorrin-3A gates access of NO, and hence also oxygen, to the mononuclear non-heme iron site. Analysis of the peptide sequence of CobG does not reveal any of the typical motifs involving two histidine side chains that

---

**FIGURE 8. Model and mechanism of CobG.** Diagram illustrating the coordination and topology of the bound cofactors within sulfite reductase (a) and the putative model of CobG (b). The mechanism is outlined in c.
have been identified in other iron-containing monoxygenases (45).

The activity of CobG could be detected in vivo using a series of E. coli strains containing incomplete B12 biosynthetic pathways. All attempts to detect enzymatic activity in cell free systems using the P. denitrificans CobG, using either cell lysates or the purified enzyme, failed. In contrast, the CobG from B. melitensis was active both in vivo and in vitro. The likely explanation for the difference in activity between the two CobG enzymes is that the P. denitrificans CobG contains a sensitive mononuclear non-heme iron that appears to become colloidal once the enzyme is released from the internal environment of the cells. In contrast, the B. melitensis CobG has a mononuclear non-heme iron that behaves as expected for a cofactor that binds molecular oxygen.

Sequence analysis indicates that CobG has similarity with assimilatory sulfite and nitrite reductases. Sulfite and nitrite reductase both contain [4Fe-4S] clusters that are linked to the iron at the center of the siroheme cofactor through a shared cysteine bridge (Fig. 8a). By comparison with CobG, it can be envisaged that precorrin-3A may bind in a similar manner to siroheme, in close proximity to the Fe-S cluster. As indicated by the EPR, the mononuclear non-heme iron is also likely to be located nearby to the bound substrate and the Fe-S center in a position such to promote the formation of precorrin-3B following the activation of oxygen (Fig. 8b). A model for the proposed mechanism is illustrated in Fig. 8c. Mechanistically, the mononuclear non-heme iron is likely to bind molecular oxygen, which via the addition of an electron from the [4Fe-4S] center will generate a peroxy and then a Fe(III)-O2− which via the addition of an electron from the [4Fe-4S] center nuclear non-heme iron is likely to bind molecular oxygen, a mechanism is illustrated in Fig. 8c.

From an evolutionary perspective, it is probable that sulfite reductases and CobG have arisen through the process of patchwork evolution, where both have retained a shared protein framework, yet have specialized to catalyze different processes (47). An alternative enzyme system to CobG is found in some of the Rhodobacteraceae, where a flavoprotein (CobZ) acts as the monooxygenase (48).

In summary, this research has shown that CobG contains both a [4Fe-4S] cluster and a bound mononuclear non-heme iron. Much work is still required to confirm the role of the redox cofactors within the mechanism of this intriguing enzyme.

Acknowledgments—The B. melitensis genomic DNA was a generous gift from Jean-Jacques Letesson, Facultes Universitaires Notre-Dame de la Paix, Belgium. The plasmid pCR427 was a generous gift from Dr. Charles Rossener (Texas A & M University).

REFERENCES
1. Minot, G. R., and Murphy, W. P. (1983) J. Am. Med. Assoc. 250, 3328–3335
2. Rickes, E. L., Brink, N. G., Koniusz, F. R., Wood, T. R., and Folkers, K. (1948) Science 107, 396–397
3. Smith, E. L. (1948) Nature 161, 638–639
4. Hodgkin, D. C., Pickworth, J., Robertson, I. H., Trueblood, K. N., Prosen, R. I., and White, I. (1955) Nature 176, 325–328
5. Warren, M. I., Raux, E., Schubert, H. L., and Escalante-Semerena, J. C. (2002) Nat. Prod. Rep. 19, 390–412
6. Battersby, A. R. (1994) Science 264, 1551–1557
7. Blanche, F., Thibaut, D., Debussche, L., Hertle, R., Zipfel, F., and Muller, G. (1993) Angew. Chem. Int. Ed. Engl. 32, 1651–1653
8. Raux, E., Schubert, H. L., Roper, J. M., Wilson, S. W., and Warren, M. J. (1999) Bioorg. Chem. 27, 100–118
9. Roessner, C. A., Santander, P. J., and Scott, A. I. (2001) Vitam. Horm. 61, 267–297
10. Scott, A. I. (1994) Heterocycles 39, 471–476
11. Debussche, L., Thibaut, D., Cameron, B., Crouzet, J., and Blanche, F. (1993) J. Bacteriol. 175, 7430–7440
12. Scott, A. I., Roessner, C. A., Stolowich, N. J., Spencer, J. B., Min, C., and Ozaki, S. I. (1993) FEBS Lett. 331, 105–108
13. Debussche, L., Thibaut, D., Danzer, M., Debu, F., Frechet, D., Herman, F., Blanche, F., and Vuillehmoge, M. (1993) J. Chem. Soc. Chem. Commun. 1100–1103
14. Spencer, J. B., Stolowich, N. J., Roessner, C. A., Min, C. H., and Scott, A. I. (1993) J. Am. Chem. Soc. 115, 11610–11611
15. Spencer, J. B., Stolowich, N. I., Santander, P. J., Pichon, C., Kajiwara, M., Tokiwa, S., Takatori, K., and Scott, A. I. (1994) J. Am. Chem. Soc. 116, 4991–4992
16. Stolowich, N. J., Wang, J. J., Spencer, J. B., Santander, P. J., Roessner, C. A., and Scott, A. I. (1996) J. Am. Chem. Soc. 118, 1657–1662
17. Christner, J. A., Munck, E., Janick, P. A., and Siegel, L. M. (1981) J. Biol. Chem. 256, 2098–2101
18. Crane, B. R., Siegel, L. M., and Getzoff, E. D. (1995) Science 270, 59–67
19. Janick, P. A., and Siegel, L. M. (1982) Biochemistry 21, 5397–5398
20. Raux, E., Lanois, A., Levillayer, F., Warren, M. J., Brody, E., Rambach, A., and Thermes, C. (1996) J. Biol. Chem. 271, 753–767
21. Dutton, P. L. (1978) Methods Enzymol. 54, 411–435
22. Munro, A. W., Noble, M. A., Robledo, L., Daff, S. N., and Chapman, S. K. (2001) Biochemistry 40, 1956–1963
23. Ost, T. W., Miles, C. S., Munro, A. W., Murdoch, J., Reid, G. A., and Chapman, S. K. (2001) Biochemistry 40, 13421–13429
24. Lovenberg, W. (1973) Iron-Sulphur Proteins, Vol II, pp. 195–237, Academic Press, New York
25. Bertini, I., Ciurli, S., and Luchinat, C. (1995) Structure and Bonding, pp. 1–53, Springer, Berlin/Heidelberg
26. Johnson, M. K., Duderstadt, R. E., and Duin, E. C. (1999) Adv. Inorg. Chem. 47, 1–82
27. Mousseau, J. M., and Lamotte, B. (1998) Coord. Chem. Rev. 180, 1537–1614
28. Bertrand, P., Gayda, J.-P., and Rao, K. K. (1982) J. Chem. Phys. 76, 4715–4719
29. Sweeney, W. V., and Rabinowitz, J. C. (1980) Annu. Rev. Biochem. 49, 139–161
30. Solomon, E. I., Brunold, T. C., Davis, M. I., Kemsley, J. N., Lee, S. K., Lehnert, N., Neese, F., Skulan, A. I., Yang, Y. S., and Zhou, J. (2000) Chem. Rev. 100, 235–350
31. Neidig, M. L., and Solomon, E. I. (2005) Chem. Comm. 5843–5863
32. Parsons, J. B., Dinesh, S. D., Devey, E., Leech, H. K., Brindley, A. A., Heldt, D., Frank, S., Smales, C. M., Lunsford, H., Rambach, A., Gass, M. H., Bleloch, A., McClean, K. J., Munro, A. W., Rigby, S. E., Warren, M. J., and Prentice, M. B. (2008) J. Biol. Chem. 283, 14366–14375
33. Leech, H. K., Raux, E., McClean, K. J., Munro, A. W., Robinson, N. I., Borrelly, G. P., Malten, M., Jahn, D., Rigby, S. E., Heathcote, P., and Warren, M. J. (2003) J. Biol. Chem. 278, 41900–41907
34. Girerd, J. J., Banse, F., and Sima, A. J. (2000) Metal-Oxo and Metal-Peroxo Species in Catalytic Oxidations 97, 145–177
35. Sima, A. J., Banse, F., Mialane, P., Boussac, A., Un, S., Kargar-Grisel, T., Bouchoux, G., and Girerd, J. J. (1999) Eur. J. Inorg. Chem. 993–996
36. Burger, R. M., Peisach, J., and Horowitz, S. B. (1981) J. Biol. Chem. 256, 11636–11644
37. Sima, A. J., Banse, F., Girerd, J. J., Wieghardt, K., and Bill, E. (2001) Inorg. Chem. 40, 6538–6540
38. Ryle, M. J., and Hausinger, R. P. (2002) Curr. Opin. Chem. Biol. 6, 193–201
39. Nelson, M. J. (1987) J. Biol. Chem. 262, 12137–12142
40. Arciero, D. M., Lipscomb, J. D., Huynh, B. H., Kent, T. A., and Münck, E. (1983) J. Biol. Chem. 258, 14981–14991
41. Chen, V. J., Orville, A. M., Harpe, M. R., Frolik, C. A., Surerus, K. K., Münck, E., and Lipscomb, J. D. (1989) J. Biol. Chem. 264, 21677–21681
42. Jackson, T. A., Yikilmaz, E., Miller, A.-F., and Brunold, T. C. (2003) J. Am. Chem. Soc. 125, 8348–8363
43. Bian, S. M., and Cowan, J. A. (1999) Coord. Chem. Rev. 192, 1049–1066
44. Gaffney, B. J. (1996) Annu. Rev. Biophys. Biomol. Struct. 25, 431–459
45. Costas, M., Mehn, M. P., Jensen, M. P., and Que, L., Jnr. (2004) Chem. Rev. 104, 939–986
46. Eschenmoser, A. (1988) Angew. Chem. 27, 5–39
47. Bartlett, G. J., Borkakoti, N., and Thornton, J. M. (2003) J. Mol. Biol. 331, 829–860
48. McGoldrick, H. M., Roessner, C. A., Raux, E., Lawrence, A. D., McLean, K. J., Munro, A. W., Santabarbara, S., Rigby, S. E., Heathcote, P., Scott, A. I., and Warren, M. J. (2005) J. Biol. Chem. 280, 1086–1094