Effects of Exercise Combined with Finasteride on Hormone and Ovarian Function in Polycystic Ovary Syndrome Rats

Chuyan Wu,1 Feng Jiang,2 Ke Wei,3 Feng Lin,1 and Zhongli Jiang1

1Department of Rehabilitation Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China
2Neonatal Department, Obstetrics and Gynecology Hospital of Fudan University, Shanghai, China
3Medical Service Section, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China

Correspondence should be addressed to Zhongli Jiang; 32081537@qq.com

Received 7 August 2018; Revised 25 January 2019; Accepted 6 February 2019; Published 13 March 2019

Academic Editor: Vito A. Giagulli

Copyright © 2019 Chuyan Wu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Exercise can reduce androgen and insulin levels in polycystic ovary syndrome (PCOS) patients. Finasteride is also presumed to improve the developing follicle environment. Therefore, the aim of this study was to observe the effects of the combination of exercise and finasteride therapy on hormone levels and ovarian morphology in rats with polycystic ovary syndrome. Forty female rats were randomly divided into five groups (n = 8 each group): the PCOS sedentary group (P-Sed), PCOS exercise group (P-Ex), PCOS finasteride and sedentary group (P-FSed), and PCOS finasteride and exercise group (P-FEx), and healthy, age-matched rats were used as controls (CO). The results indicated that the levels of FINS in the P-FEx group were significantly lower than those in the P-Sed and P-FSed groups, while the ratio of fasting blood glucose (FBG)/fasting serum levels of insulin (FINS) in the P-FEx group was significantly higher than that in the P-Sed and P-FSed groups. Compared to the P-FEx group, serum levels of TT (total testosterone) in the P-Sed and P-FSed groups were significantly increased. The thickness of the follicular membrane and the number of atresia follicles in the P-FEx and CO groups were significantly lower than those in the P-Sed group, but there was no significant difference between the P-Ex and P-Sed groups. These results show that the combined usage of exercise and finasteride does not alter the effects of exercise on increasing insulin sensitivity and reducing androgen levels. There is also a synergistic effect of exercise and finasteride on the morphology of the ovary, including a reduced number of atresia follicles and thickness of the follicular membrane.

1. Introduction

Polycystic ovary syndrome (PCOS) is a common endocrine syndrome among women of childbearing age and seriously affects the lives of such women [1]. Hyperandrogenemia and hyperinsulinemia are two major endocrine characteristics of polycystic ovary syndrome [2]. Additionally, acne and hirsutism are very painful complaints in adolescent women with PCOS [3, 4]. Our previous studies confirmed that 5α-reductase activity was enhanced in women with PCOS. Increased 5α-reductase activity has been shown to be associated with idiopathic hirsutism, androgenic alopecia, and acne. There are many types of drugs available for the treatment of hirsutism and acne, including oral contraceptives, antiandrogens, and reductase inhibitors [5]. Reductase inhibitors are widely used in clinical applications, including the commonly used drug finasteride. Finasteride, an antiandrogen drug, is a competitive inhibitor of type 2 5α-reductase, which converts testosterone into the stronger dihydrotestosterone. Small doses of finasteride are often given intermittently to adolescent PCOS women. Studies have shown that it can effectively improve the hairy symptoms of PCOS women and has no effect on metabolic parameters [6].

Exercise is a first-line treatment recommended for polycystic ovary syndrome that has been widely used in patients of all ages [7–9]. Exercise can reduce the levels of androgen and insulin in PCOS and improve the ovarian endocrine environment, which plays an important role in promoting ovulation. Finasteride is also presumed to improve the developing follicle environment. It has been shown to improve the
hormonal environment around the follicle and block the development of atresia follicles [10].

In the treatment of PCOS in adolescent women, sometimes, it is necessary to combine finasteride with exercise therapy. Therefore, the purpose of this study was to investigate the effects of exercise combined with finasteride on hormone levels and ovarian morphology in rats with polycystic ovary syndrome.

2. Methods

The experimental protocols were approved by the Institutional Animal Care and Use Committees of Nanjing Medical University (permit number: NJMU/IA-CUC_20120210_01) and strictly followed. Forty female Wistar rats (50-70 g, 21 days old) were obtained (Beijing Vital River Laboratory Animal Technology Co., Beijing, China) and cared for according to the Guiding Principles for the Care and Use of Laboratory Animals (National Research Council of People’s Republic of China, 2010). The rats were housed individually in an animal facility under controlled conditions (12:12 h light:dark cycle, with the light period from 8:00 am to 8:00 pm) and were given access to water. The PCOS rat model was generated by injecting testosterone propionate for 28 days and feeding a high-fat diet according to our previous studies [2]. After 28 days, the PCOS rats were randomly divided into four groups (n = 8 per group): a sedentary group (P-Sed), an exercise group (P-Ex), a finasteride and exercise group (P-Fex), and a finasteride and sedentary group (P-Fsed). Additionally, healthy age-matched rats (n = 8) were used as controls (CO). Finasteride (5 mg/kg/day) was administered by gavage in the P-Fsed and P-Fex groups once daily [11]. All the rats were sacrificed at the end of exercise between 8:00 am and 11:00 am under 12-h fasting conditions with 3% pentobarbital abdominal anesthesia followed by cervical dislocation. Ovary samples and blood were collected from all the animals.

2.1. Exercise Protocol. The exercised rats were forced to swim with no load for 120 min/day, 6 days/week for two weeks in a barrel filled with water at a depth of 40-50 cm maintained at 33–35°C [2, 7]. Each rat was given enough area to swim freely. The nonexercise rats’ cages were kept near the barrels to equalize the effects of exercise noise.

2.2. Measurements of Body Weight and Body Length. The body weight and length of rats were measured weekly from 21 days of age. The body length of rats was defined as the distance from the nose to the anus. The Lee obesity index was used to reflect body fat as a parameter [LI = body weight (g) / \( \sqrt[3]{1000/\text{body length (cm)}} \)] [2, 12]. The bilateral ovaries of each rat were weighed, and the mean values were regarded as the ovary weight. The ovarian organ index is calculated as ovary weight/body weight \( \times 10^{-3} \) [2, 7].

2.3. Serum Analysis. Following intervention, all PCOS groups (n = 32) and the control group (n = 8) were fasted for 12 h after 8:00 pm. The blood was taken from the canthus vein and used for a fasting insulin determination. The blood was taken from the tip of the tail, and blood glucose was measured by fast test paper using the Surestep Plus glucose meter (Johnson, United States). After blood glucose was measured, 3% pentobarbital was injected into the abdominal cavity, and 1-5 ml of blood from the abdominal aorta was collected and used to determine hormone levels. The blood samples obtained from rats fasting for 12 h were centrifuged at 2500 g for 10 min and stored at –80°C. Fasting serum levels of insulin (FINS), total testosterone (TT), sex hormone-binding globulin (SHBG), and dihydrotestosterone (DHT) were determined using commercial ELISA kits (Jiancheng Bioengineering Institute, Nanjing, P.R. China). The free androgen index (FAI) was calculated as follows: FAI = (TT \times 100)/SHBG [13].

2.4. Ovarian Morphological Analysis. After separation of the bilateral ovaries, one side of the ovary was quickly put into liquid nitrogen and then stored at –80°C. After weighing, the other side of the ovary was fixed in neutral formalin solution for 24 h and then embedded in paraffin. One ovary of each rat was sectioned at the center along the long axis of the ovary and serially sectioned at 4 μm from the center. We chose three serial sections with the largest areas for counting the numbers of preantral, antral, and atretic follicles and corpora lutea [7, 14]. All the sections were photographed with a camera (Leica D4000, Germany). The area of the largest section (at \( \times 40 \) magnification), the number of corpora lutea (at \( \times 100 \) magnification), and the thickness of the theca cell layer and granulosa cell layer (the average from 3 visual fields of follicles at \( \times 200 \) magnification) were measured with Image-Pro Plus 6.0.

3. Statistical Analysis

SPSS statistical software (version 16.0, SPSS, Chicago, IL, USA) was used to analyze the data, and the data were described as the mean ± SD. All the data were analyzed by one-way ANOVA, and P < 0.05 was considered significantly different. When the ANOVA revealed significant differences among the three experimental groups, a post hoc analysis was performed by using Bonferroni’s method of multiple comparisons (10 repeated tests; P < 0.005 was considered significant for each test).

4. Results

4.1. The Effect of Exercise Combined with Finasteride on Body Weight, Body Length, Body Mass Index, and Ovarian Organ Index in the PCOS Rats. Body weight was significantly higher in the P-FEx than in the P-Sed group, and body length was significantly greater in the P-Ex, P-FEx, and CO groups than in the P-Sed group. However, the Lee index in the P-Ex and P-FEx groups was significantly lower than that in the P-Sed group, and it was significantly higher in the P-Sed and P-Fsed groups than in the CO group. The ovary organ indexes in the P-Sed and P-Fsed groups were both significantly lower than in the CO group (Table 1).

4.2. The Effects of Exercise Combined with Finasteride on Serum Insulin and Androgens. There were no significant differences in the levels of FBG among these groups. The levels
of FINS in the P-Sed and P-FSed groups were significantly higher than those in the other three groups, while the ratios of FBG/FINS in the P-Sed and P-FSed were significantly lower than the other three groups. In comparison with the P-Ex, P-FEx, and CO groups, the serum levels of TT in the P-Sed and P-FSed groups were significantly greater. The serum levels of SHBG in the P-Sed group were significantly lower compared with those of the other groups. The levels of FAI in the P-Sed and P-FSed groups were significantly higher than those in the P-EX and CO groups (Table 2). There was no difference in the serum DHT among these groups.

4.3. Observations of Ovary Slices under the Microscope. In the CO, antral follicles at different stages and a large number of corpora lutea were observed. The antral follicles at different stages were observed in the P-Ex and P-FEx groups, and a small amount of luteal formation was found in the P-Ex group. The P-Sed group showed typical cystic follicular structure and many preantral and atresia follicles, while the P-FSed group showed different stages of antral follicle structure and several typical cystic structures (Figure 1).

4.4. Morphological Analysis of the Ovary in Each Group. Compared with the CO and P-Ex groups, the ovarian and luteal areas in the P-Sed and P-FSed groups were significantly smaller. The luteal area of the P-FEx group was also significantly smaller compared to the P-Fsed and P-Sed groups. The thickness of the granular layer and the number of atretic follicles in the CO and P-FEx groups were significantly lower than those in the P-Sed group, and the numbers of antral follicles in the P-Sed and P-FSed groups were significantly lower than those in the CO group. The number of corpora lutea, the ratio of the corpus luteum area to ovarian area, the ratio of the number of corpora lutea to the total number of follicles, and the ratio of the corpus luteum area to ovarian area in the P-Sed and P-FSed groups were significantly lower than those in the CO and P-Ex groups, while the ratio of the number of atretic follicles to the total number of follicles was increased significantly in the P-FSed group compared to the CO, P-Ex, and P-FEx groups. The ratio of the corpus luteum area to ovarian area and the ratio of the number of atretic follicles in the P-FEx group were also significantly higher than those in the P-Sed group (Table 3).

Table 1: Comparisons of weight, length, Lee index, and ovary organ index.

|                  | CO      | P-Sed   | P-Ex    | P-FSed  | P-FEx   |
|------------------|---------|---------|---------|---------|---------|
| Body weight (g)  | 199.17  | 185.00  | 190.71  | 202.83  | 205.17  |
|                  | ± 15.4  | ± 5.83  | ± 6.73  | ± 13.36 | ± 9.04  |
| Body length (cm) | 20.88   | 19.31   | 20.47   | 20.08   | 20.87   |
|                  | ± 0.99  | ± 0.72  | ± 0.64  | ± 0.56  | ± 0.27  |
| Lee index        | 280.07  | 295.27  | 281.39  | 292.53  | 282.62  |
|                  | ± 16.14 | ± 9.30  | ± 8.33  | ± 6.62  | ± 4.23  |
| Ovary organ index | 0.35    | 0.24    | 0.30    | 0.24    | 0.30    |
|                  | ± 0.04  | ± 0.05  | ± 0.03  | ± 0.02  | ± 0.02  |

Values are the means ± SD. *Compared with the CO group, P < 0.005. †Compared with the P-Sed group, P < 0.005.

PCOS is an endocrine syndrome with high clinical heterogeneity [15–17]. This syndrome often begins during puberty [18]. Symptoms of the disease in adolescence include irregular menstruation, hirsutism, and acne [19]. These clinical manifestations greatly affect the body and mind of adolescent females, which causes a psychological burden of self-abasement and forms a vicious cycle further aggravating the disease. Both exercise and antiandrogen therapies are common methods used to treat polycystic ovary syndrome [16, 20–22]. However, it is not clear how the combination of these two factors affects the female hormone levels and the ovary. The PCOS rat model is characterized by hyperinsulinemia and hyperandrogenemia [2, 23]. The morphology of the ovaries includes the morphology of a polycystic follicle, which consists of a thin layer of follicle granules, a thickening of the follicular membrane, and increasing follicular follicles [2].

Numerous animal experiments and human studies have confirmed that exercise treatment can reduce weight [21] and improve hyperinsulinemia and high androgen levels [22] in individuals exhibiting PCOS. In addition, the ovaries can be restored to promote ovulation by exercise. Among the antiandrogen drugs, finasteride, a type 2 reductase inhibitor, has been shown by many studies to exhibit a higher than average safety level [19, 24–26]. Compared with the type 1 reductase inhibitor dutasteride, previous experiments with finasteride have proved that there is no significant difference in clinical efficacy and related parameters [27, 28]. However, recent studies have shown that taking dutasteride can increase body fat and decrease insulin sensitivity, whereas finasteride has no such effect and can improve insulin resistance [29, 30]. In the present study, the finasteride group showed no significant difference in body mass index; however, it showed an increase in body weight compared with the nonfinasteride group.

Finasteride alone cannot improve insulin resistance in PCOS rats. Fortunately, it also does not aggravate insulin resistance and it did not affect the action of exercise to improve insulin resistance in the P-FEx group. Finasteride inhibited testosterone conversion to dihydrotestosterone, resulting in an increase in total testosterone in the two finasteride groups. In addition, a large number of population experiments have confirmed that sex hormone-binding protein was closely related to impaired glucose tolerance [31, 32]. In this study, it was also found that the levels of sex hormone-binding protein in the two exercise groups were significantly higher than those in the two sedentary groups. The current study also confirmed that finasteride has no significant effect on improving insulin resistance.
Table 2: Levels of insulin and androgen in each group.

|          | CO      | P-Sed   | P-Ex    | P-FSed  | P-FEx   |
|----------|---------|---------|---------|---------|---------|
| FBG (mmol/l) | 3.62 ± 0.99 | 3.41 ± 0.48 | 3.47 ± 0.30 | 3.05 ± 0.49 | 3.33 ± 0.53 |
| FINS (μIU/ml) | 11.10 ± 0.79 | 12.76 ± 1.10<sup>a</sup> | 11.10 ± 0.87<sup>b</sup> | 12.42 ± 1.40<sup>abc</sup> | 11.04 ± 1.11<sup>bd</sup> |
| FBG/FINS   | 0.33 ± 0.09  | 0.26 ± 0.04<sup>a</sup> | 0.32 ± 0.05<sup>b</sup> | 0.24 ± 0.02<sup>abc</sup> | 0.31 ± 0.07<sup>bd</sup> |
| TT (nmol/l) | 34.26 ± 2.11 | 38.52 ± 3.64<sup>a</sup> | 34.56 ± 5.37<sup>b</sup> | 38.39 ± 1.13<sup>abc</sup> | 36.66 ± 1.42<sup>bd</sup> |
| DHT (nmol/l) | 30.18 ± 2.39 | 30.29 ± 3.01  | 31.62 ± 2.28 | 30.03 ± 4.12 | 30.25 ± 2.59 |
| SHBG (nmol/l) | 52.78 ± 2.71 | 44.76 ± 1.08<sup>a</sup> | 51.85 ± 5.22<sup>b</sup> | 48.53 ± 4.22 | 50.03 ± 2.74<sup>b</sup> |
| FAI (nmol/l) | 68.06 ± 3.81 | 85.95 ± 7.18<sup>a</sup> | 66.96 ± 10.09<sup>b</sup> | 79.26 ± 7.04<sup>abc</sup> | 73.24 ± 6.78 |

Values are the means ± SD. <sup>a</sup>Compared with the CO group, P < 0.005. <sup>b</sup>Compared with the P-Sed group, P < 0.005. <sup>c</sup>Compared with the P-Ex group, P < 0.005. <sup>d</sup>Compared with the P-FSed group, P < 0.005.

Figure 1: Ovarian morphology of rats (×100). (a) Control. Various stages of follicles are seen. (b) PCOS. Many preantral follicles (P) and atretic follicles (AF) with enlargement of stroma (arrow) and cystic follicles (C) are seen in the section. (c) PCOS plus exercise. Many antral follicles (A), several atretic follicles (AF), and several corpora lutea (CL) can be seen in the section. (d) PCOS plus finasteride. Different stages of antral follicles and several typical cystic structures are seen in the section. (e) PCOS plus exercise and finasteride. Several corpora lutea (CL) can be seen in the section.
In a population study in 2004, 31 PCOS women were given oral finasteride or the same dose of placebo. The experiment concluded that the PCOS women who took finasteride did not show a significant difference in ovarian capacity compared with the control group [33]. In our study, the PCOS rats in the P-FEx group showed a significant reduction in the area of the corpus luteum. Although the area of the ovary was decreased, there was no significant difference compared with the P-Ex group. The growth of follicles is affected by aromatase and reductase and is particularly affected by their metabolites, estradiol and dihydrotestosterone. Follicular growth is the result of interactions between endocrine, autocrine, and paracrine factors. High levels of DHT in the follicle block the cell signaling pathway. Studies have shown that androgen receptor blockers and testosterone antibodies inhibit follicular atresia [10, 34]. Our experimental results showed that the P-FEx rats compared with the P-Sed rats primarily displayed decreased atresia follicles and decreased follicle film thickness, which is consistent with the inference that finasteride can reduce follicle atresia. However, the improvement of ovarian morphology in these two aspects was not obvious in the P-Ex group. A possible mechanism is that the growth and development of follicles can be affected by competition between aromatase and 5α-reductase.

In conclusion, this study investigated the obesity PCOS rat model induced by testosterone propionate combined with a high-fat diet. Exercise plus finasteride or either intervention alone was used to observe the effects of exercise, finasteride, and their combination on serology and ovarian morphology in the PCOS rats. The results showed that the combined use of finasteride and exercise did not alter the effect of exercise on increasing insulin sensitivity and reducing androgen levels. Furthermore, there is a synergistic effect of finasteride and exercise on the morphology of the ovary as the number of atresia follicles, and the thickness of the follicular membrane can be reduced. However, due to the small number of animals tested and the difference with the model of a chronic hyperandrogenism as in PCOS women, the effect of finasteride on the PCOS patient population needs further study.

**Abbreviations**

PCOS: Polycystic ovary syndrome  
FBG: Fasting blood glucose  
FINS: Fasting serum levels of insulin  
TT: Total testosterone  
SHBG: Sex hormone-binding globulin  
DHT: Dihydrotestosterone  
FAI: Free androgen index

**Data Availability**

The data used to support the findings of this study are available from the corresponding author upon request.

**Ethical Approval**

The experimental protocols were approved by the Institutional Animal Care and Use Committees of Nanjing Medical University (permit number: NJMU/IA-CUC_20120210_01) and strictly followed.

**Disclosure**

The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Conflicts of Interest**

The authors have no relevant conflict of interest to disclose.
Authors’ Contributions

Chuyan Wu designed the study, performed the experiments, analyzed the data, and wrote the manuscript; Feng Jiang performed the experiments, analyzed the data, and critically reviewed the manuscript; Ke Wei and Feng Lin performed the experiments and critically reviewed the manuscript; Zhongli Jiang interpreted the results of the experiments and critically reviewed and edited the manuscript.

Acknowledgments

We thank Xianhui Lu and Jin Gao for their assistance with this study. This study was supported by the National Nature Science Foundation of China (81501947). The support from the Priority Academic Program Development of Jiangsu Higher Education Institutions was also appreciated.

References

[1] C. Wu, K. Wei, and Z. Jiang, “5α-reductase activity in women with polycystic ovary syndrome: a systematic review and meta-analysis,” Reproductive Biology and Endocrinology, vol. 15, no. 1, p. 21, 2017.
[2] C. Wu, F. Lin, S. Qiu, and Z. Jiang, “The characterization of obese polycystic ovary syndrome rat model suitable for exercise intervention,” PloS One, vol. 9, no. 6, article e91155, 2014.
[3] J. A. Balfour and K. McClellan, “Topical efollornithine,” American Journal of Clinical Dermatology, vol. 2, no. 3, pp. 197–201, 2001.
[4] J. C. Harper, “Hormonal therapy for acne using oral contraceptive pills,” Seminars in Cutaneous Medicine and Surgery, vol. 24, no. 2, pp. 103–106, 2005.
[5] P. Moghetti and V. Toscano, “Treatment of hirsutism and acne in hyperandrogenism,” Best Practice & Research Clinical Endocrinology & Metabolism, vol. 20, no. 2, pp. 221–234, 2006.
[6] M. V. Tartagni, H. Alrasheed, G. R. Damiani et al., “Intermittent low-dose finasteride administration is effective for treatment of hirsutism in adolescent girls: a pilot study,” Journal of Pediatric and Adolescent Gynecology, vol. 27, no. 3, pp. 161–165, 2014.
[7] S. Qiu, C. Wu, F. Lin, L. Chen, Z. Huang, and Z. Jiang, “Exercise training improved insulin sensitivity and ovarian morphology in rats with polycystic ovary syndrome,” Hormone and Metabolic Research, vol. 41, no. 12, pp. 880–885, 2009.
[8] F. Giallauria, S. Palomba, L. Maresca et al., “Exercise training improves autonomic function and inflammatory pattern in women with polycystic ovary syndrome (PCOS),” Clinical Endocrinology, vol. 69, no. 5, pp. 792–798, 2008.
[9] A. Khademi, A. Alleyassin, M. Aghalosseini, L. Tabatabaeefar, and M. Amini, “The effect of exercise in PCOS women who exercise regularly,” Asian Journal of Sports Medicine, vol. 1, no. 1, pp. 35–40, 2010.
[10] M. Tartagni, E. Cicinelli, G. De Pergola et al., “Effect of finasteride on ovulation induction in nonresponder (hyperandrogenic) polycystic ovary syndrome (PCOS) women,” Fertility and Sterility, vol. 94, no. 1, pp. 247–249, 2010.
[11] M. Kumazaki, H. Ando, K. Ushijima et al., “Influence of dosing time on the efficacy and safety of finasteride in rats,” The Journal of Pharmacology and Experimental Therapeutics, vol. 338, no. 2, pp. 718–723, 2011.
[12] M. Morifuji, C. Sanbongi, and K. Sugiyama, “Dietary soy protein intake and exercise training have an additive effect on skeletal muscle fatty acid oxidation enzyme activities and mRNA levels in rats,” British Journal of Nutrition, vol. 96, no. 3, pp. 469–475, 2006.
[13] Z. Zhou, R. Ni, Y. Hong et al., “Defining hyperandrogenaemia according to the free androgen index in Chinese women: a cross-sectional study,” Clinical Endocrinology, vol. 77, no. 3, pp. 446–452, 2012.
[14] G. Liu, F. Shi, U. Blas-Machado et al., “Ovarian effects of a high lactose diet in the female rat,” Reproduction, Nutrition, Development, vol. 45, no. 2, pp. 185–192, 2005.
[15] B. C. Fauser, B. C. Tarlatzis, R. W. Rebar et al., “Consensus on women’s health aspects of polycystic ovary syndrome (PCOS): the Amsterdam ESHRE/ASRM-Sponsored 3rd PCOS Consensus Workshop Group,” Fertility and Sterility, vol. 97, no. 1, pp. 28–38.e25, 2012.
[16] N. Naderpoor, S. Shorakae, B. de Courten, M. L. Misso, L. J. Moran, and H. J. Teede, “Metformin and lifestyle modification in polycystic ovary syndrome: systematic review and meta-analysis,” Human Reproduction Update, vol. 22, no. 3, pp. 408-409, 2016.
[17] T. Li, H. Mo, W. Chen et al., “Role of the PI3K-Akt signaling pathway in the pathogenesis of polycystic ovary syndrome,” Reproductive Sciences, vol. 24, no. 5, pp. 646–655, 2017.
[18] M. B. Eriksen, D. Glinborg, M. F. Nielsen et al., “Testosterone treatment increases androgen receptor and aromatase gene expression in myotubes from patients with PCOS and controls, but does not induce insulin resistance,” Biochemical and Biophysical Research Communications, vol. 451, no. 4, pp. 622–626, 2014.
[19] C. N. Jayasena and S. Franks, “The management of patients with polycystic ovary syndrome,” Nature Reviews Endocrinology, vol. 10, no. 10, pp. 624–636, 2014.
[20] A. Benrick, M. Maliqueo, S. Miao et al., “Resveratrol is not as effective as physical exercise for improving reproductive and metabolic functions in rats with dihydrotestosterone-induced polycystic ovary syndrome,” Evidence-Based Complementary and Alternative Medicine, vol. 2013, Article ID 964070, 13 pages, 2013.
[21] L. J. Moran, C. B. Lombard, S. Lim, M. Noakes, and H. J. Teede, “Polycystic ovary syndrome and weight management,” Women’s Health, vol. 6, no. 2, pp. 271–283, 2010.
[22] C. L. Harrison, C. B. Lombard, L. J. Moran, and H. J. Teede, “Exercise therapy in polycystic ovary syndrome: a systematic review,” Human Reproduction Update, vol. 17, no. 2, pp. 171–183, 2011.
[23] L. Manners, S. Cajander, A. Holmang et al., “A new rat model exhibiting both ovarian and metabolic characteristics of polycystic ovary syndrome,” Endocrinology, vol. 148, no. 8, pp. 3781–3791, 2007.
[24] A. Petrone, R. M. Civitillo, L. Galante et al., “Usefulness of a 12-month treatment with finasteride in idiopathic and polycystic ovary syndrome-associated hirsutism,” Clinical and Experimental Obstetrics & Gynecology, vol. 26, no. 3–4, pp. 213–216, 1999.
[25] P. Moghetti, F. Tosi, A. Tosti et al., “Comparison of spironolactone, flutamide, and finasteride efficacy in the treatment of hirsutism: a randomized, double blind, placebo-controlled trial,” The Journal of Clinical Endocrinology & Metabolism, vol. 85, no. 1, pp. 89–94, 2000.
[26] A. Tolino, A. Petrone, F. Sarnacchiaro et al., “Finasteride in the treatment of hirsutism: new therapeutic perspectives,” *Fertility and Sterility*, vol. 66, no. 1, pp. 61–65, 1996.

[27] Y. Kotelevtsev, M. C. Holmes, A. Burchell et al., “11β-hydroxysteroid dehydrogenase type 1 knockout mice show attenuated glucocorticoid-inducible responses and resist hyperglycemia on obesity or stress,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 26, pp. 14924–14929, 1997.

[28] N. M. Morton, M. C. Holmes, C. Fiévet et al., “Improved lipid and lipoprotein profile, hepatic insulin sensitivity, and glucose tolerance in 11β-hydroxysteroid dehydrogenase type 1 null mice,” *Journal of Biological Chemistry*, vol. 276, no. 44, pp. 41293–41300, 2001.

[29] J. K. Dowman, L. J. Hopkins, G. M. Reynolds et al., “Loss of 5α-reductase type 1 accelerates the development of hepatic steatosis but protects against hepatocellular carcinoma in male mice,” *Endocrinology*, vol. 154, no. 12, pp. 4536–4547, 2013.

[30] R. Upreti, K. A. Hughes, D. E. Livingstone et al., “5α-reductase type 1 modulates insulin sensitivity in men,” *The Journal of Clinical Endocrinology & Metabolism*, vol. 99, no. 8, pp. E1397–E1406, 2014.

[31] D. A. Ehrmann, R. B. Barnes, R. L. Rosenfield, M. K. Cavaghan, and J. Imperial, “Prevalence of impaired glucose tolerance and diabetes in women with polycystic ovary syndrome,” *Diabetes Care*, vol. 22, no. 1, pp. 141–146, 1999.

[32] S. H. Golden, A. S. Dobs, D. Vaidya et al., “Endogenous sex hormones and glucose tolerance status in postmenopausal women,” *The Journal of Clinical Endocrinology & Metabolism*, vol. 92, no. 4, pp. 1289–1295, 2007.

[33] W. M. Arie, V. R. Bagnoli, and A. M. da Fonseca, “Effects of finasteride on the morphology of polycystic ovaries,” *International Journal of Gynaecology & Obstetrics*, vol. 87, no. 1, pp. 52–53, 2004.

[34] A. J. Zeleznik, S. G. Hillier, and G. T. Ross, “Follicle stimulating hormone-induced follicular development: an examination of the role of androgens,” *Biology of Reproduction*, vol. 21, no. 3, pp. 673–681, 1979.