Rising trends towards the development of oral herbal male contraceptive: an insight review

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Abstract

Background: Acknowledging population control to be an essential step for global health promotes wide research study in the area of male contraception. Although there are a great number of synthetic contraceptives available in the market, they have plenty of adverse effects. Different potential strategies for male contraception were investigated over a long period time consisting of hormonal, chemical, and immunological interventions, although these methods showed good antifertility results with low failure rates relative to condoms.

Main text: This review is based upon the concept of herbal contraceptives which are an effective method for controlling the fertility of animals and humans. This review has highlighted herbal medicinal plants and plant extracts which have been reported to possess significant antifertility action in males. The review considers those plants which are used traditionally for their spermicidal and antispermatogetic activities and imbalance essential hormones for fertility purposes and plants with reported animal studies as well as some with human studies for antifertility effect along with their doses, chemical constituents, and mechanism of action of the antifertility effect of the plants. This review also explains the phases of sperm formation, hormone production, and the mechanism of male contraceptives.

Conclusion: As far as the relevance of the current review is discussed, it might be quite useful in generating monographs on plants and recommendations on their use. A lot of the plant species listed here might appear promising as effective alternative oral fertility-regulating agents in males. Therefore, significant research into the chemical and biological properties of such less-explored plants is still needed to determine their contraceptive efficacy and also to possibly define their toxic effects so that these ingredients can be utilized with confidence to regulate male fertility. The new inventions in this field are necessary to concentrate on modern, more potent drugs with less harmful content and that are self-administrable, less costly, and entirely reversible.

Keywords: Antifertility, Family planning, Herbal contraceptives, Herbs, Mechanisms, Oral male contraceptives, Overpopulation

Background

Today, overpopulation is a matter of extreme concern for developed nations along with developing countries [1, 2]. In the year 2011, the world population was estimated at 6,928,198, 253 and increasing rapidly at a rate of 83 million citizens per year [3]. Among developing countries, India is densely populated and it is estimated that it will reach about 9.2 billion by the year 2050 [1, 2]. The year 2012 witnessed population figures reaching 1,210,193,422 [1] with an increment of 18 million to the total population every year [2]. According to India’s population in 2019, figures are 1,372,717,495 [4]. Increasing population leads to an increase in the demand for resources like water and food, starvation, malnutrition, and consumption of natural resources. Since natural resources are limited, control of the increasing population is a mandatory step [5]. Family planning is an easy and important tool for controlling population burden [6]. On other hand, in
the USA, the unwanted pregnancy rate is approximately 45% with the help of the various contraceptive options provided to women [7]. Although female contraceptives are much effective in preventing unplanned pregnancy, giving high yielding results, it cannot be used by a greater proportion of sex due to their profuse adverse effects [8].

Talking about extreme measures, the World health Organization has initiated a population control program that includes trials linked to traditional medical activities [2]. Many methods for induction of infertility are implemented over a long period that involves biochemical, biological, and immunological pathways [9] with the least impact but minimal inadequacy [5]. During ancient times, the human reproduction system was not fully established. Hence, the progress in research could not be put into practice due to unclear mechanisms related to human hormones [10].

**Main text**

**Methods and materials**

The present study has been geared up with wide-ranging facts of curative plants inhabiting all over the world concerning their accepted tradition by countless ethnic groups for fertility regulation in males. To date, no examination has analyzed the dose, constituents of elements, and mechanism of action of the antifertility effect of plants.

So, information concerning this article has been systematically gathered from the sources of scientific literature, including PubMed, Google Scholar, Science Direct, and Scopus. Simply applicable studies available in the English language were considered. The botanical and English names are considered after validation from available text and database. The criteria followed for the choice of information in this evaluation deliberate folk plant:

1. Found in Ancient Indian medicines
2. With recorded animal studies for effects on infertility as well as those with human studies of antifertility effect

Plants, their parts, or their extracts traditionally used for spermicidal and antispermatogenic activities and those that imbalance essential hormones for fertility purposes have been considered as antifertility agents. Furthermore, compounds isolated from plants with attributed potential for fertility regulation are also classified into 6 categories:

1. Phytoconstituents with spermicidal activity
2. Phytoconstituents with antispermatogenic activity
3. Phytoconstituents acts through Sertoli cells
4. Phytoconstituents acts through Leydig cells
5. Phytoconstituents with antimotility activity
6. Phytoconstituents acts by unbalancing hormones

The following keywords were used to search the literature in the data sources: oral male contraceptives, herbal contraceptives, antifertility, and male contraception.

**The need of male contraceptives-a boon for society!!!**

Acknowledging population control to be an essential step for global health promotes an opportunity for a large-scale research study in the field of male contraceptives [11]. Male contraceptives originated with the use of a condom in ancient times in Imperial Rome. Researches on male contraception initiated with the sole objective of taking easy targets to the male reproductive system by stopping either the sperm or the testis to function [10]. Although there are a great number of synthetic contraceptives available in the market, they have plenty of adverse effects [5].

Moreover, there is a rise in the need for male contraceptives to avoid unwanted pregnancies because not many men wish to take responsibility for family planning [8]. Also due to the adverse effects of synthetic male contraceptives, the quest for a modern, more effective, more reliable, and less expansive approach is the priority as well as objectives for the pharmaceutical and medical sciences not to forget an unusual self-administration and long-lasting effect of male contraceptives [9]. The new inventions in this field often concentrate on modern, more potent drugs with less harmful content and that are self-administrable, less costly, and entirely reversible [12].

**Why not herbals???

Since ancient times, plants are always regarded as a potent source of nourishment that we require for staying healthy along with their valuable components commonly used for food and nutrition, beverages, cosmetics, dyes, medicines, etc. Herbs are excellent examples of being one of the richest sources of nutrients that aim at protecting and restoring a healthy life [13]. As specified by the World Health Organization (WHO) statistics, almost 65–80% of the world’s population relies on plant species and their health care products due to the lack of modern facilities and poor conditions. There is a total of 422,000 plant species that have been recorded all over the world, out of which 20,000 species are acclaimed as wild edible species and less than 20,000 of the same community is consumed as a food supplement for 90% of people around the world, contributing to almost 25% of drug formulations from plants or their extracts [14]. The herbal preparations have been used as an oral tradition. It is becoming more popular and useful in modern times as demand for natural remedies/medicines is increasing.
every day because of the belief of people that they do not have any adverse effects, a boon in disguise [15]. Continuing the traditional system of medicines, more than 35,000 plant species are being used worldwide for medicinal purposes. Following which, more than 80% of the world population is turning to herbal preparations that contain plant extracts for primary health care [2].

Herbal contraceptives are plant-based contraceptives which are effective methods for controlling the fertility of animals and humans [16]. The chemical constituents of plants such as flavonoids, terpenes, tannins, quinines, diterpenoids, and lactones are apprehended to possess antifertility action through a different mechanism [17]. Different potential mechanisms for male contraception have been studied over a long period of time consisting of hormonal, chemical, and immunological strategies [2, 18] though these methods have shown better results of antifertility effects with minimum failure rates than condoms.

In recent years, plants have been reported to be used in the regulation of male fertility because of the better compatibility with the human body, better cultural acceptability, and lesser adverse effects giving it an upper hand [19]. As a result, herbal products attract scientists as a primary source of naturally occurring antifertility agents due to little or negligible adverse signs [17]. For instance, in India, several herbal plants have been reported to have antifertility effects that act through the mechanism by suppression of spermatogenesis or by prevention of implantation [20].

Phases for sperm formation

Testicular carries out two primary goals:

1. Testosterone production
2. Spermatogenesis (origin of haploid germ cells) [8] (Fig. 1)

Flow sheet for production of hormones [21] (Fig. 2)

Pituitary gonadotrophins regulate the functions of the testicles, with luteinizing hormones (LH) acting on the testosterone and producing interstitial cells and the follicle-stimulating hormone (FSH) affecting the cells of the seminiferous tubules. Including the seminiferous epithelium’s structural elements, the movement of nutritional growth factor to the haploid germ cells with a near junction in the epithelium for consecutive cells is known as the “blood-testis barrier.” Well-performing Sertoli cells supply sufficient mitogens, distinct factors, and energy sources to the growing germ cell, as well as shielding them from the host’s own immune system from harmful agents. The number and function of the present Sertoli cells define the spermatogenesis rate and quality [22].

Male contraceptives might work as follows:

1. Suppress sperm production by antispermagenic
2. Prevention of maturation of sperm
3. Prevention of the flow of sperm through vas deferens
4. Prevention of deposition of the sperm [1]
Phytoconstituents with spermicidal agents

Ideal spermicidal characteristics include the following: would rapidly and irreversibly achieve sperm immobilization, are not harmful to the vaginal and penile mucosa, do not have any side effects on the growing fetus that are free from long-term topical and systemic toxicity, and should not be ingested systemically [23]. A typical lipid bilayer consisting of outer, intra, and transmembrane proteins is a key component of the plasma membrane of sperms. This lipid bilayer associate with saponin molecules, impacting cell membrane glycoproteins and altering ionic movement across the membrane, resulting in many plants (Phytolacca dodecadra, Calendula officinalis, and Acacia caesia) differing [24].

Sperm immobilization can be caused by the acid pH of lemon juice through the denaturation of ATPase dyein [23].

Phytoconstituents with antispermatogenic activity

The spermatogenesis process involves a complex process:

1. Spermatocytogenesis
2. Spermatidogenesis
3. Spermiogenesis

A diploid spermatogonium undergoes mitotic division in the process of spermatocytogenesis and develops two diploids known as primary spermatocytes. Every primary spermatocyte divides into two haploid secondary spermatocytes by meiosis. Spermiogenesis is the process of spermatid differentiation into mature sperm. It indicates interference in the steroidogenesis when the cholesterol level rises and sudanophilic lipid accumulates [25].

Phytoconstituents acts through Sertoli cells

Sertoli cells are columnar with oval or pear nuclei and thin mitochondria; at the base of their cytoplasm, they have lipofuscin and lipid droplets. The main feature of Sertoli cell structural support for germ cell development is the blood test barrier, which is situated between neighboring Sertoli cells in close junctions. Sertoli cells play an important part in the process of spermatogenesis and adult life as a whole. The plant extracts kill the viability and work of Sertoli cells and have various effects 

Fig. 2 Release and production of different hormones from glands
on spermatogenesis, such as reducing the nuclear and cytoplasmic volume and vacuolizing Sertoli cells [26].

**Phytoconstituents acts through Leydig cells**

Leydig cells are polyhedral with a large prominent nucleus, an eosinophilic cytoplasm, and various vesicles packed with lipids. The hormone-releasing gonadotropin, secreted and synthesized by the hypothalamus, produces and releases LH and FSH from the pituitary gland. LH induces the production of testosterone in the testis Leydig cells [27].

**Phytoconstituents with antimotility activity**

The sperm passes through three sections of the caput, corpus, and cauda epididymis that are important for sperm maturation [28]. Therefore, the production and secretion of proteins through the epididymis and the completion of various morphological, biochemical, and motile properties during the transformation from epididymis are important for the spermatozoa’s full capacity to fertilize [29].

**Phytoconstituents acts by unbalancing hormone**

Hypothalamus, pituitary gland, and testis secrete the various hormones which regulate spermatogenesis [30]. The Leydig cells synthesize and secrete the major male sex hormone testosterone under the influence of pituitary gonadotropin luteinizing hormone (LH). Several plant products are considered to contain enzymes that are used in androgen synthesis [31]. Quassia Amara’s blunt methanol extract shows lower levels of testosterone, LH, and hormone-stimulating follicles (Fig. 3) (Table 1)

Epidemiological studies have shown that the use of combination oral contraceptives increases the possibility of brain thrombosis; enhances the serum triglyceride, high-density lipoprotein (HDL), and cholesterol levels; and also increases family mortality related to cardiovascular diseases as well as malignant tumors in any organ, low resistance to glucose or insulin, diarrhea, abdominal pain, fatigue, hypertension, and menstrual shifts [2].

Patents available are shown Table 2.
| Sr. no. | Botanical name | Common name | Family | Part used | Subject | Dose | Active constituents | Duration of administration (days) | Mechanism of action | Antifertility activity | References |
|--------|----------------|-------------|--------|-----------|---------|------|---------------------|----------------------------------|----------------------|----------------------|-------------|
| 1.     | Acacia auriculiformis Benth. | Ear leaf | Fabaceae | Seeds | Human sperm | 0.35 mg/ml | Acacic acid lactone | – | Disintegrate sperm plasma membrane and immobilize sperm | – | [32] |
| 2.     | Aegle marmelos (L.) Corrêa Bael | Leaves | Rutaceae | Rat | | 200 and 300 mg/kg B.W. /day | Coumarins, tannins, phenols, rutin | 60 | Suppress gonadotropic hormone | Reversible | [33] |
| 3.     | Albizia lebbeck (L.) Benth | Pods | Fabaceae | Rat | | 100 mg/kg B.W. | Labbekanin-E, saponins | 60 | Reduce sperm density and sperm motility and decrease testes and prostate size | – | [34] |
| 4.     | Allium sativum L. | Bulb | Lillicae | Human sperm | | 0.25 and 0.5 g/ml | Allitridum | – | Disrupt membrane architecture | Irreversible | [20] |
| 5.     | Cananga odorata (Lam.) Hook. f. & Thomson | Root bark | Annonaceae | Rat | | 1 g/kg B.W./day | 52-kd protein | 60 | Decrease androgen production, increase 3-hydroxy-3-methyl-glutaryl-CoA (HMG CoA) reductase activity, and decrease 3β-hydroxy steroid dehydrogenase enzyme activity | Reversible | [35] |
| 6.     | Cestrum parqui (Lam.) L'Hér. | Leaves | Solanaceae | Human semen | | 40, 62.5, 100, 150, and 250 μg/ml | Saponin | – | Disrupt sperm plasma membrane sterol | – | [36] |
| 7.     | Chenopodium album L. | Lamb's quarters | Chenopodiaceae | Fruits | Rat/rabbit | 2 mg/ml | Oleanolic acid, glucuronic acid | – | Disintegrate sperm plasma membrane and cause the dissolution of acrosomal cap causing sperm death | – | [37] |
| 8.     | Chromolaena odoratum (L.) Tivra | Leaves | Compositae | Rat | | 250 and 500 mg/kg B.W. | – | 14 | Decrease biomolecule concentration and disrupt seminiferous tubules | – | [38] |
| 9.     | Colebrookia oppositifolia | Leaves | Lamaceae | Rat | | 100 and 200 mg/kg | 5,6,7,4′-tetramethoxy flavones, 5,6,7-trimethoxy flavones, 5,7, 4′-trihydroxy flavones 3-o-glucoside | 56–70 | | – | [39] |
| 10.    | Juniperus phoenica (L.) | Ripe red cones | Cupressaceae | Rat | | 400 and 800 mg/kg | α-Pinene, δ-3-carone, β-phellandrene | 21 | Inhibit LH and gonadotropin-liberating hormone | – | [40] |
| 11.    | Mollugo pentaphylla L. | Aerial part | Molluginaceae | Human sperm | | 10, 30, 100, and | Mollugogenol-A (saponin) | – | Plasma membrane-losing osmoregulatory properties and | – | [41] |
| Sr. no. | Botanical name | Common name | Family      | Part used | Subject | Dose       | Active constituents | Duration of administration (days) | Mechanism of action | Antifertility activity | References |
|---------|----------------|-------------|-------------|-----------|---------|------------|--------------------|----------------------------------|---------------------|----------------------|------------|
| 12.     | Quassia amara  | Bitterwood  | Simaroubaceae| Stem wood | Rat     | 0.1, 1.0, | Quassin, 2-methoxycanthin-6-one | 56                 | Decrease serum level testosterone, LH, and FSH | Reversible | [42]       |
|         | L.             |             |             |           |         | and 2 mg/kg B.W. |                     |                                  |                     |                      |            |
| 13.     | Sapindus        | Reetha      | Sapindaceae | –         | Human   | 0.05%, 0.1%, | Saponins, digitonin | –                               | Disruption and erosion of membrane | –         | [32]       |
|         | mukorossi       |             |             |           | Semen   | 1.25%, and 5%|                     |                                  |                     |                      |            |
| Gaertn. |                |             |             |           |         |             |                     |                                  |                     |                      |            |
| 14.     | Terminalia      | Chebulic     | Combretaceae| Dry fruits| Rat     | 50 and 100 mg/ | Anthraquinones, ellagittamic acid, 42,4- | 60     | Inhibit acrosomal enzyme and sperm hyaluronidase enzyme | Reversible | [43]       |
|         | chebula         | myrobala    |             |           |         | kg/day     | 4,2,4-chebulyl-β-D-glucopyranose, ellagic acid, gallic acid |                     |                                  |                     |            |
| Retz.   |                |             |             |           |         |             |                     |                                  |                     |                      |            |
| 15.     | Tinospora       | Guduchi     | Menispermaceae| Stem   | Rat     | 100 mg/ | –                        | 60     | Reduce plasma level of testosterone and inhibit glycolysis in spermatozoa | –         | [44]       |
|         | cordifolia (Wild)|            |             |           |         | rat/day   |                     |                                  |                     |                      |            |
|         | (Willd)         |             |             |           |         |             |                     |                                  |                     |                      |            |
| 16.     | Ziziphus        | Ber         | Rhamnaceae  | Barks    | Human semen | 0.1 and 0.5 mg/ml | Saponin | 20 s–20 min | Disrupt lipid within sperm membrane | –         | [45]       |
|         | mauritiana      |             |             |           |         |            |                     |                                  |                     |                      |            |
| Lam.    |                |             |             |           |         |            |                     |                                  |                     |                      |            |

**Phytoconstituents with antispermatogenic activity**

1. *Bacopa monnieri* (L.) Wettst.  
   *Brahmi*  
   Scrophulariaceae  
   All part  
   Mice  
   250 mg/kg B.W. /day  
   28 and 56  
   Decrease fructose level and inhibit spermatogenesis  
   Reversible  
   [46]

2. *Barleria prionitis* L.  
   *Vajra-danti*  
   Acanthaceae  
   Root  
   Rat  
   100 mg/kg  
   Barlerin, acetyl barlerin, apigenin-7-o-glucoside  
   60  
   Reduce glycogen, protein, and sialic acid content and deplete germinal and Leydig cell constituents  
   –  
   [47]

3. *Cannabis sativa* L.  
   *Ganja*  
   Cannabinaceae  
   Seeds  
   Rat  
   20 mg/day  
   Cannabinoids  
   20  
   Act on cannabinoid receptors  
   –  
   [48]

4. *Chrysophyllum albidum* G.Don  
   *White star apple, vdara*  
   Compodeoidea  
   Root bark  
   Rat  
   100 and 200 mg/kg  
   Alkaloids, tannis, saponin, phenol, flavonoids  
   147  
   Reduce gonadotropins level (FSH and LH) and inhibit spermatogenesis  
   –  
   [49]

5. *Citrullus colocynthis* (L.) Schrad.  
   *Tumba*  
   Cucurbitaceae  
   Root  
   Rat  
   50, 100, and 200 mg/kg B.W./day  
   Hentriacontane, n-octacosanol, 1,2,6-hexa-cosanediol  
   60  
   Inhibit pituitary gonadotropin secretion and reduce sialic acid and protein  
   Reversible  
   [50]

6. *Crotalaria juncea* L.  
   *Sunn hemp*  
   Papilionaceae  
   Seed  
   Mice  
   25 mg/100 g/day  
   –  
   30  
   Reduce seminiferous tubular fluid and decrease protein content, FSH, and LH  
   –  
   [51]

7. *Cuminum Jeera*  
   *Apiceae*  
   Seed  
   Rat  
   100 mg/  
   –  
   60  
   Sloughing or death of epithelial  
   –  
   [52]
| Sr. no. | Botanical name | Common name | Family | Part used | Subject | Dose | Active constituents | Duration of administration (days) | Mechanism of action | Antifertility activity | References |
|---------|----------------|-------------|--------|-----------|---------|------|---------------------|---------------------------------|---------------------|---------------------|------------|
| 8.      | Curcuma longa L. | Haldi       | Zingiberaceae | Rhizomes | Mice    | 600 mg/kg B.W./day | –                  | 56 and 84 | Inhibit gonadotropin secretion and decrease serum level | Reversible [53] |
| 9.      | Fadogia agrestis Schweinf. ex Hiern | Nagbitenga | Rubiaceae | Stem | Rat | 18, 50, and 100 mg/kg B.W. | Alkaloids, anthraquinones, flavonoids, saponin | 28 | Increase cholesterol level and reduce glycogen content | Reversible [54] |
| 10.     | Hibiscus rosa-sinensis L. | Gudhal | Malvaceae | Flower | Mice | 150 and 300 mg/kg | –                  | 20 | Decrease androgen synthesis and reduce spermatogenic element | – [55] |
| 11.     | Lepidium meyenii Walp. | Maca | Brassicaceae | Root | Rat | 66.7 mg/ml | –                  | 14 | Enhance epididymal weight and reduce stages I-VI of seminiferous epithelium | – [56] |
| 12.     | Leptadenia hastata | Cheila | Asclepiadaceae | Leaves and stem | Rat | 100, 200, 400, and 800 mg/kg | –                  | 60 | Reduce Leydig cell and imbalance LH, prolactin, and testosterone serum level hormones | – [57] |
| 13.     | Momordica charantia L. | Karela | Cucurbitaceae | Seed | Rat | 25 mg/100 g B.W. | –                  | 35 | Inhibit gonadotrophins (FSH) and enhance cholesterol level and sudanophilic lipids | – [58] |
| 14.     | Mondia whitei (Hook.f) Skeels | La racine | Periplocaeae | Root | Rat | 500 and 1000 mg/kg B.W. | Steroids, triterpenes | 30 | Reduce intratesticular concentration of cholesterol | Reversible [59] |
| 15.     | Morinda lucida Benth. | Brimstone tree | Rubiaceae | Leaves | Rat | 400 mg/kg/day | Anthraquinones, anthraquinols | 28 and 91 | Reduce serum testosterone level and inhibit acetylcholinesterase | Reversible [60] |
| 16.     | Mucuna urens L. | Ibaba | Cannabaceae | Seed | Rat | 70, 140, and 210 mg/kg B.W. | Flavonoids, anthranoid, anthraquinones, polyphenols | 14 | Inhibit endogenous gonadotrophic activity | – [61] |
| 17.     | Ocimum gratissimum L. | African basil, ram tulsi | Lamiaceae | Leave | Mice | 11–88 mg/kg | Eugenol, citral, linalol, charvicol, thymol, geraniol | 7, 14 and 28 | Deplete Leydig and Sertoli cells and destroy cell membrane | – [62] |
| 18.     | Parkinsonia aculeate L. | Viliyat babul | Caesalpinaceae | Stem bark | Rat | 50, 100, and 200 mg/rat/day | α-Amyrin acetate, β-amyron acetate, 6-hydroxypentacosylpentanoate ethynoma decanoate, 6-hydroxytritriacont-3-one | 60 | Reduce testosterone level and Leydig cell diameter and seminiferous tubular diameter | – [63] |
| 19.     | Piper nigrum L. | Long pepper | Piperaceae | Fruits | Mice | 25 and 100 mg/ | Piperine | 20 and 90 | Reduce sialic acid level and decrease fructose concentration | Reversible [64] |
| Sr. no. | Botanical name | Common name | Family | Part used | Subject | Dose | Active constituents | Duration of administration (days) | Mechanism of action | Antifertility activity | References |
|--------|----------------|-------------|--------|-----------|---------|------|---------------------|----------------------------------|---------------------|-------------------|------------|
| 20.    | Ruta graveolens L. | Rue | Rutaceae | Leave | Rat | 500 mg/kg B.W./day | – | Reduce serum androgen level and degenerate Leydig cells | – | – | [65] |
| 21.    | Semecarpus anacardium | Bilawa | Anacardiaceae | Fruit | Rat | 100, 200, and 300 mg/kg/day | – | Decrease sialic acid content and androgen production (LH) | – | – | [66] |
| 22.    | Terminalia bellirica (Gaertn.) Roxb. | Baheda | Combretaceae | Fruit | Rat | 10 and 25 mg/100 g B.W. | Triphala | Reduce androgen level and increase cholesterol level | – | – | [67] |
| 23.    | Thevetia peruviana | Lucky nut, Mexican oleander | Apocynaceae | Stem bark | Rat | 100 mg/rat/day | α-Amyrin acetate, lupeol acetate, α-amyrin, β-amyrin, lupeol, thevetigenin | 60 | Deform and impair Leydig cell and reduce androgen concentration | – | – | [68] |

**Phytoconstituents acts through Sertoli cells**

1. *Azadirachta indica* A.Juss. | Neem | Meliaceae | Leaves | Rat | 100 mg/rat/day | – | Degenerate germ cells | Reversible | [69] |
2. *Dendrophthoe falcate* (L.f.) Ettingsh. | Banda | Loranthaceae | Stem | Rat | 100 mg/kg B.W./day | Quercitrin (quercetin-3-o-rhamnoside), kaempferol, rutin | 60 | Decrease seminiferous tubular fluid and reduce androgen synthesis and sialic acid | – | – | [70] |
3. *Thespesia populnea* (L.) Sol. ex Corrêa | Tulip tree | Malvaceae | Leaves | Mice | 400 mg/kg B.W. | – | Elongate seminiferous tubules | – | – | [71] |
4. *Tripterygium wilfordi* Hook. f. | Yellow vine root | Celastraceae | Root | Rat | 100 mg/kg/day | – | Degenerative changes of seminiferous epithelium and reduce reproductive cells in testes | Irreversible | – | [72] |

**Phytoconstituents acts through Leydig cells**

1. *Berberis chitria* Buch.-Ham. ex Lindl. | Daruhaldi | Berberidaceae | Root | Dog | 30 mg/kg/day | Palmitine hydroxide | 60 | Decrease postmeiotic germ cells and decrease androgen binding protein of Sertoli cells via FSH | – | – | [73] |
2. *Calotropis procera* | Camelweed | Asclepiadaceae | Roots | Gerbil/ rabbit | 25 mg/kg B.W. | Calotropin | 30 | Suppress testicular function by decreasing androgenic parameter | – | – | [74] |
3. *Garcinia cambogia* | Malabar tamarind | Cluciaceae | Seed | Rat | 100 and 200 mg/kg B.W. | Biflavonoid, xanthone | 42 | Enhance interstitial spaces and reduce Leydig cells in interstitial space and seminiferous tubules contraction | – | – | [75] |
4. *Malvaviscus conzattii* Greenm. | Turk's cap mallow | Malvaceae | Flower | Rat | 800 mg/kg B.W./day | – | Reduce germ cells and impairs function of epididymides | – | – | [76] |
| Sr. no. | Botanical name | Common name | Family | Part used | Subject | Dose | Active constituents | Duration of administration (days) | Mechanism of action | Antifertility activity | References |
|--------|----------------|-------------|--------|-----------|---------|------|--------------------|-------------------------------|-------------------|---------------------|------------|
| 5.     | Martynia annua L. | Scorpion Martyniaceae | Root | Rat | 50, 100, and 200 mg/kg B.W. | – | Reduce serum concentration of LH and testosterone | 30 | Reversible | [77] |
| 6.     | Ocimum sanctum L. | Tulsi Lamiaceae | Fresh leaves | Rabbit | 2 g/day | – | Reduce pH, mucoprotein, and alkaline phosphatase and make non-viable spermatozoa | 30 | Reversible | [21] |
| Phytoconstituents with antimotility activity | | | | | | | | | |
| 1.     | Carica papaya L. | Papita Caricaceae | Seeds | Monkey | 50 mg/kg B.W./day | – | Hasten sperm transport leading to ejaculation and affect composition of epididymal fluid and their enzymes on spermatozoa | 360 | Reversible | [78] |
| 2.     | Echinops echinatus Roxb. | Utakatira, oonkateli | Roots | Rat | 50, 100, and 200 mg/kg B.W./day | Echinopsine, echinopsidine, echinozolinone | 60 | Reduce concentration of protein in the cauda epididymis and testicular glycogen level and reduce ascorbic acid content of the adrenal gland | – | [79] |
| 3.     | Gossypum herbaceum | Cotton Malvaceae | Fruit | Rabbit | 20 mg/day | Gossypol acetic acid | 84 | – | [80] |
| 4.     | Lagenaria breviflora (Benth.) Roberty | Molina Cucurbitaceae | Whole fruit | Rat | 1000, 2000, 4000, and 8000 mg/kg B.W. | – | Degenerate seminiferous tubules | 14 | – | [81] |
| Phytoconstituents acts by unbalancing hormones | | | | | | | | | |
| 1.     | Abelmoschus esculentus (L.) Moench | Okra Malvaceae | Fruit | Rat | 70 mg/kg B.W./day | Flavonoids, saponins | 28 | Reduce serum testosterone level and spermatogenesis | Reversible | [82] |
| 2.     | Abrus precatorius L. | Coral bead vine, rosary pea, ratti Leguminosae | Seed | Rat | 250 mg/kg | Abridine | 30 and 60 | Impair function of sperm plasma membrane, suppress oxidative/energy metabolism, and reduce sperm motility | Reversible | [83] |
| 3.     | Bulbine natalensis Baker | Bulbine Asphodelaceae | Stem | Rat | 25, 50, and 100 mg/kg B.W. | Alkaloids, tannins, anthraquinones | 7 | Reduce serum testosterone and progesterone levels | – | [84] |
| 4.     | Curcuma longa L. | Haldi Zingiberaceae | Rhizomes | Rat | 500 mg/kg/day | – | Decrease androgen synthesis and Leydig cell nuclei diameter and inhibit Leydig cell function | 60 | – | [85] |
| Sr. no. | Botanical name | Common name | Family | Part used | Subject | Dose | Active constituents | Duration of administration (days) | Mechanism of action | Antifertility activity | References |
|---------|----------------|-------------|--------|-----------|---------|------|---------------------|----------------------------------|----------------------|----------------------|------------|
| 5.      | Psoralea corylifolia L. | Babchi | Leguminosae | Seeds | Rat | 10 g/kg B.W. | Corylin, bavachin, psoralen, isopsoralen, psoralidin | 84 | Decrease serum testosterone and FSH levels and suppress pituitary-testicular axis | – | [86] |
| 6.      | Stevia rebaudiana | Sugar leaf | Asteraceae | Leave | Rat | 2 ml/rat | Stevioside | 60 | Decrease androgen level | Irreversible | [87] |
| 7.      | Syzygium aromaticum (L.) Merr. & L.M. Perry | Lavang | Myrtaceae | Flower buds | Mice | 15, 30, and 60 mg/kg B.W. | Eugenol, β-caryophyllene | 35 | Destroy germ cells and inhibit spermatogonia | – | [88] |
Conclusion
Different potential strategies for male contraception were investigated over a long period time consisting of hormonal, chemical, and immunological interventions, although these methods showed good antifertility results with low failure rates relative to condoms. The present study showed the literature data that there is majority of plants, which are traditionally used as antifertility agents and their effects have not been thoroughly studied on animals. The majority of tests have been performed on conscious animals and relatively few tests have had their efficacy confirmed in humans. Herbal contraceptives are safer and cheaper methods for population overcoming. To summarize, a lot of the plant species listed here might appear promising as effective alternative oral fertility-regulating agents in males. Among plant parts, leaves have been maximally utilized for controlling fertility. As far as the relevance of the current review is discussed, it might be quite useful in generating monographs on plants and recommendations on their

Table 2 List of patents on a male contraceptive

| Summary of invention                                                                 | Patent number           | Inventor/assignee                                      |
|-------------------------------------------------------------------------------------|-------------------------|--------------------------------------------------------|
| Substituted acylanilides and methods of use thereof                                  | AU2015264895B2          | Dalton, James, Miller, Duane D.                        |
| Styrene maleic anhydride based formulation for male contraception and prostate cancer | EP 2 268 290 B1          | Guha, Sujoy Kumar                                      |
| Lonidamine analogs and their use in male contraception and cancer treatment         | EP 2 502 624 A1         | Chakrassali, Georg, Jakkaraj, Tash                    |
| Orally active 7-alpha-alkyl androgens                                              | EP1212345B1             | Louw Van Der, Leysen, Buma Bursi                      |
| Methods of making the 4-n-butyloclohexanoic and the undecanoic acid esters of (7 alpha, 11 beta)-dimethyl-17 beta-hydroxy-4-estren-3-one and their medical use | EP1272196B1             | Blye, Kim                                              |
| Oral pharmaceutical composition comprising 15-hydroxytestosterone and its analogues  | EP1551415B1             | Bunschoten, Coelingh Bennink, Van Der Linden          |
| Male contraceptive formulation comprising norethisterone                           | US20020103176A1         | Eberhard Nieschlag, Axel Kamischke, Michael Oettel, Alexander Ruebig, Ekkerhard Schillinger, Habenicht Ursula-Friederike |
| Male contraceptive method and composition                                          | US20020164368A1         | Ronald Zimmerman                                      |
| Androgen as a male contraceptive and non-contraceptive androgen replacement         | US20020193359A1         | Alfred J. Moo-Young                                   |
| Reversible infertility in male mice following oral administration of alkylated imino sugars: a non-hormonal approach to male contraception | US20040019082A1         | Aamoud C. Van Der Spoel, Mylvaganam Jeyakumar, Terry D. Butters, Raymond A. Dewk, Frances M. Platt |
| Non-hormonal compositions and methods for male contraception                         | US20190290615A1         | Guillaume Ei Glau, Mehdi Ei Glau, Philippe Perrin, Stéphane Droupy, Véronique Agathon--Meriau |
| Reversible male contraception                                                       | US4252798               | Donald J. Tindall                                     |
| Male contraceptive steroids and methods of use                                      | US4297350               | John C. Babcock; J. Allan Campbell, Thomas J. Lobl,  |
| Oral male contraceptive                                                             | US4381298               | Patricia B. Coulson, Sheffield Dr.                    |
| Male contraceptive implant                                                          | US5733565               | Alfred J. Moo-Young, Saleh I. Saleh                   |
| Male contraceptives                                                                  | US5854254               | Susan H. Benoff                                       |
| Buccal drug delivery system for use in male contraception                           | US6180682               | Virgil A. Place                                       |
| High-strength testosterone undecanoate compositions                                 | US9480690               | Chandrashekar Gilyar, Basavaraj Chickmath, Nachiannan Chidambaram, Srinivasan Venkateshwaran |
| Male contraceptive comprising a prolactin inhibitor and a sex steroid               | WO1999066953A1          | Lincoln, Kirkton Cottages, WU                         |
| A pulmonary drug delivery composition containing a progestogen and androgen for use in a contraceptive method in males | WO2003068315A1          | Coelingh Bennink, Van Der Linden                      |
| Novel spermicidal and anti-infective contraceptive device                            | WO2007074478A1          | Jain Rajesh, Jindal Kour Chand                         |
| Substituted (5,6)-dihydronaphraalenyl compounds as reversible male contraceptives  | WO2008137081A1          | Wolgemuth Debra J., Reczek Peter R.                   |
| Inhibitors of eppin/semenogelin binding as male contraceptives                      | WO2009042565A2          | O’rand Michael G., Widgren Esther Elaine, Richardson Richard, Temple Brenda |
| Non-hormonal male contraceptive agents and methods using same                       | WO2016205339A1          | Yan Wei                                                |
| Piperidine-dione derivatives for use as contraceptives                               | WO2018211276A1          | Sieng Bora, Lundvall, Steffi, Claudia Alejandra       |
use. Therefore, this review has highlighted the significant antifertility activity of herbal medicinal plants and their extracts. Moreover, this review can concentrate the interest of researchers on toxicity studies of phytoconstituents and their clinical trials, which may serve as an alternate potential antifertility agents with milder or fewer side effects and can be developed into suitable contraceptive formulations. Therefore, significant research into the chemical and biological properties of such less-explored plants is still needed to determine their contraceptive efficacy and also will possibly define their toxic effects so that these ingredients can be utilized with confidence to regulate male fertility.

Abbreviations
LH: Luteinizing hormones FSH: Follicle-stimulating hormone
hDL: High-density lipoprotein B/W: Body weight

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Authors’ contributions
We declare that this work was done by the authors named in this article: SY conceived and designed the study. AY carried out the literature collection of the data and writing of the manuscript. AY and SY assisted in the data analysis and corrected the manuscript. The authors read and approved the final manuscript.

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