Monocyclic Phenolic Acids; Hydroxy- and Polyhydroxybenzoic Acids: Occurrence and Recent Bioactivity Studies

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Abstract: Among the wide diversity of naturally occurring phenolic acids, at least 30 hydroxy- and polyhydroxybenzoic acids have been reported in the last 10 years to have biological activities. The chemical structures, natural occurrence throughout the plant, algal, bacterial, fungal and animal kingdoms, and recently described bioactivities of these phenolic and polyphenolic acids are reviewed to illustrate their wide distribution, biological and ecological importance, and potential as new leads for the development of pharmaceutical and agricultural products to improve human health and nutrition.

Keywords: polyphenols; phenolic acids; hydroxybenzoic acids; natural occurrence; bioactivities

1. Introduction

Phenolic compounds exist in most plant tissues as secondary metabolites, i.e. they are not essential for growth, development or reproduction but may play roles as antioxidants and in interactions between the plant and its biological environment. Phenolics are also important components of the human diet due to their potential antioxidant activity [1], their capacity to diminish oxidative stress-induced tissue damage resulted from chronic diseases [2], and their potentially important properties such as anticancer activities [3-5].
The structure of phenolics consists of an aromatic ring carrying one (phenol) or more hydroxyl (polyphenol) moieties. Several classes can be distinguished according to the number of phenol rings and to the structural elements that join these rings [6]. Two main groups of polyphenols, termed flavonoids and non-flavonoid polyphenols, have been adopted in the literature [7]. The flavonoid group, including flavanones, flavones, dihydroflavonols, flavonols, flavan-3-ols, isoflavones, anthocyanidins, proanthocyanidins and chalcones, comprises those compounds with a C6-C3-C6 structure (Figure 1).

Figure 1. Flavonoids and phenolic acids.

The non-flavonoid polyphenols can be classified based on their carbon skeleton into the following subgroups: simple phenols, benzoic acids, hydrolysable tannins, acetophenones, phenylacetic acids, cinnamic acids, lignans, coumarins, benzophenones, xanthones, stilbenes, and secoiridoids.

Phenolic acids have a carboxyl group attached or linked to benzene ring [8]. Two classes of phenolic acids can be distinguished depending on their structure: benzoic acid derivatives (i.e. hydroxybenzoic acids, C6-C1) and cinnamic acid derivatives (i.e. hydroxycinnamic acids, C6-C3) [9] (Figure 1).

This review will cover the natural occurrence and recently described biological activities of monocyclic hydroxy- and polyhydroxybenzoic acids. Research published prior to the last ten years will not be included as considerable efforts have been made already to cover those findings [e.g., 10-12]. Many hydroxybenzoic acids have not been discussed here due to their lack of known bioactivities.

2. Results and Discussion

3-Hydroxybenzoic acid (1, Figure 2) is found in common plants such as grapefruit (Citrus paradisi), olive oil (Olea europaea) [13], and medlar fruit (Mespilus germanica) [14]. It has glucosylating activity [15]. p-Hydroxybenzoic acid (4-hydroxybenzoic acid, 2, Figure 2) has been isolated from many sources including carrots (Daucus carota) [16], oil palm (Elaeis guineensis) [17], grapes (Vitis vinifera), and numerous other species including east African satinwood (Fagara macrophylla), yellow-leaf tree (Xanthophyllum rubescens), peroba (Paratecoma peroba), taheebo (Tabebuia impetiginosa), red sandalwood (Pterocarpus santalinus), southern catalpa (Catalpa bignonioides), Chinese chastetree (Vitex negundo) [18], betel palm (Areca catechu), Cuban royal palm (Roystonea regia) [19], and medlar (Mespilus germanica) [14]. It shows antifungal, antimutagenic,
antisickling, estrogenic [20], and antimicrobial [17] activities. $p$-Hydroxybenzoic acid has a growth stimulation effect on the freshwater green alga *Pseudokirchneriella subcapitata* [21].

**Figure 2.** 3-Hydroxybenzoic acid (1) and 4-Hydroxybenzoic acid (2).

Pyrocatechuic acid (2,3-dihydroxybenzoic acid, 3, Figure 3) occurs in rhododendrons (*Rhododendron* spp.) and other members of the heather family such as winter heath (*Erica carnea*) and teaberry (*Gaultheria procumbens*), yellow gentian (*Gentiana lutea*) and the related European centaury (*Catharanthus roseus*) [22]. It is also produced by algal, bacterial and fungal microorganisms such as marine-derived actinomycetes [23], the green alga *Spongiochloris spongiosa*, the cyanobacterium *Anabaena doliolum* [24], and the other bacteria *Streptomyces* sp., *Acinetobacter calcoaceticus*, *Brucella abortus*, *Aspergillus sojae*, and *Bacillus* sp., and the fungi *Rhizobium* sp. and *Penicillium roquefortii*. Pyrocatechuic acid is an antioxidant [25], a radical scavenger [23], and a siderophore [26]. It has some natural derivatives such as the 3-$O$-$\beta$-$D$-glucopyranoside isolated from totally unrelated plants such as the gentian relative *Geniostoma antherotrichum*, the common periwinkle (*Vinca minor*), and the mustard relative *Boreava orientalis*; and 2-hydroxy-3-methoxybenzoic acid from a crocus (*Colchicum decaisnei*) and a birch (*Betula pendula*).

Gentisic acid (2,5-dihydroxybenzoic acid, 4, Figure 3) also has a widespread occurrence, being found in citrus fruits (*Citrus* spp.), grapes (*Vitis vinifera*), Jerusalem artichoke (*Helianthus tuberosus*), sesame (*Sesamum indicum*), gentians (*Gentiana* spp.), red sandalwood (*Pterocarpus santalinus*), rose gum (*Eucalyptus grandis*), saxifrage (*Saxifraga* spp.), and olive (*Olea europaea*) [13]. In addition to being an analgesic, anti-inflammatory, anti-rheumatic, antiarthritic, and cytostatic agent, gentisic acid inhibits low-density lipoprotein oxidation in human plasma [27]. It is believed that gentisic acid has an effective role in the anticarcinogenetic activity of China-rose hibiscus (*Hibiscus rosa-sinensis*) extract [28]. A recent study has shown that gentisic acid is a Fibroblast Growth Factor (FGF) inhibitor [29].

Many derivatives of gentisic acid are found naturally, such as 5-$O$-(1-carboxyethyl) in aster (*Aster indicus*), 5-methylether in cowslip (*Primula veris*), 2-$O$-[\(\beta\)-D-glucopyranosyl-(1→3)-3-hydroxybenzoyl] in marsh felwort (*Lomatogonium rotatum*), 5-$O$-[4-hydroxy-3,5-dimethoxybenzoyl-(→5)-\(\beta\)-D-apiofuranosyl-(1→2)-\(\beta\)-D-glucopyranoside] (albizinin) in Indian albizia (*Albizia lebbek*), 5-$O$-[\(\beta\)-D-apiofuranosyl-(1→2)-\(\beta\)-D-glucopyranoside] in sensitive-plant (*Mimosa pudica*), 5-$O$-[\(\beta\)-D-apiofuranosyl-(1→2)-\(\beta\)-D-xylopyranoside] in the legume *Spatholobus suberectus*, 5-(6-galloylglucoside) in sawtooth oak (*Quercus acutissima*), 5-$O$-[4-hydroxy-3-methoxy-benzoyl-(→6)-\(\beta\)-D-glucopyranoside] in squirrel’s-foot fern (*Davallia mariesii*), 5-$O$-[2,3-dihydroxy-5-\(\beta\)-D-glucopyranoside] in cassia (*Cassia absus*), Chinese goldthread (*Coptis chinensis*), and sensitive-plant (*Mimosa pudica*), 5-
xyloside in Indian coral-tree (Erythrina indica), 2-O-β-D-glucopyranoside (orbicularin) in cotoneaster (Cotoneaster orbicularis), 5-O-β-xylopyranosyl, 5-O-[(5"-O-E-(4"-O-threo-guaiacylglycerol)-feruloyl]-β-apiofuranosyl-(1→2)-β-xylopyranosyl] and 1-O-[E-(4"-O-threo-guaiacylglycerol)-feruloyl]-3-O-β-galacturonopyranosyl glycerol in barrel medic (Medicago truncatula) [30].

α-Resorcylic acid (3,5-dihydroxybenzoic acid, 5, Figure 3) is a constituent of peanuts (Arachis hypogaea), chickpeas (Cicer arietinum), red sandalwood (Pterocarpus santalinus), and hill raspberry (Rubus niveus). It has nematocidal activity [31].

**Figure 3.** Pyrocatechuic acid (3), Gentisic acid (4), and α-Resorcylic acid (5).

Salicylic acid (2-hydroxybenzoic acid, 6, Figure 4) occurs in such diverse plants as willow bark (Salix spp.), poplar (Populus pseudo-simonii), Voodoo lily (Sauromatum guttatum), gumweed (Grindelia spp.), and medlar (Mespilus germanica) [14]. It is also produced by the bacterium Pseudomonas cepacia. Salicylic acid has keratolytic, anti-inflammatory, antipyretic, analgesic, antiseptic, and antifungal properties for several skin conditions such as dandruff and seborrhoeic dermatitis, ichthyosis, psoriasis, acne, etc. [32]. It functions as a hormonal mediator of plant resistance responses to environmental stress and pathogen attacks [33,34].

6-Methylsalicylic acid (2-hydroxy-6-methylbenzoic acid, 7, Figure 4) is a polyketide derivative occurring in narrow-leaf yerba-santa (Eriodictyon angustifolium). It is also produced as a mold metabolite by Phyllosticta and Penicillium spp. [35]. 6-Methylsalicylic acid is a phytotoxin. It has antibacterial and antifeeding [36] activities.

β-Resorcylic acid (2,4-dihydroxybenzoic acid, 8, Figure 4) is found in red sandalwood (Pterocarpus santalinus) and the related coralwood (Adenanthera pavonina). β-Resorcylic acid has thyroid peroxidase inhibitory effect [37]. Its methyl ether derivatives are also found naturally. For example, 2-methyl ether (pluchoic acid) is a constituent of the fleabane Pluchea lanceolata and 4-methyl ether is found in leaves and stems of the unrelated legume, Anthyllis sericea.

Orsellinic acid (2,4-dihydroxy-6-methylbenzoic acid, 9, Figure 4) presents in some lichens such as Roccella, Lecanora, and Lobaria yunnanensis. It has also been isolated from cultures of the fungi Penicillium spp., Hypoxylon spp., and Chaetomium cochlodes. Orsellinic acid has antimicrobial activity. Some derivatives of orsellinic acid are found naturally, for example, the 2-O-β-D-glucopyranoside in cloves (Syzygium aromaticum), the 2-methyl ether (isoeverninic acid) in the lichen Lecanora gangaleoides, and the 4-methyl ether (everninic acid) in the honey mushroom Armillaria mellea.
Figure 4. Salicylic acid (6), 6-Methylsalicylic acid (7), \(\beta\)-Resorcylic acid (8), and Orsellinic acid (9).

| Number | Compound Description | Structural Formula |
|--------|----------------------|--------------------|
| 6      | Salicylic acid (R1=R2=H) | ![Salicylic acid](COOH) OH R1 R2 |
| 7      | 6-Methylsalicylic acid (R1=CH3, R2=H) | 6-Methylsalicylic acid (R1=CH3, R2=H) |
| 8      | \(\beta\)-Resorcylic acid (R1=H, R2=OH) | \(\beta\)-Resorcylic acid (R1=H, R2=OH) |
| 9      | Orsellinic acid (R1=CH3, R2=OH) | Orsellinic acid (R1=CH3, R2=OH) |

Protocatechuic acid (3,4-dihydroxybenzoic acid, 10, Figure 5) found in Spanish heath (Erica australis), dog rose (Rosa canina), Korean spruce (Picea koraiensis), gum-tree (Eucalyptus grandis), the Traditional Chinese Medicine (TCM) herb shensi (Picrorhiza kurrooa), ferns, buckwheat (Fagopyrum spp.), alder (Alnus spp.), onion and garlic and relatives (Allium spp.), Japanese pepper (Zanthoxylum piperitum) [38], another TCM herb danshen (Salvia miltiorrhiza) [39], sharp-leaf galangal (Alpinia oxyphylla) [40], sea buckthorn (Hippophae rhamnoides) [41], Japanese honeysuckle (Lonicera japonica) [42], mulberry (Morus alba) [43], and medlar (Mespilus germanica) [14]. It has been found to have several bioactivities such as antifungal, antihepatotoxic, anti-inflammatory, antioxidant [25,44], free radical scavenger, cytotoxic [42], chemopreventive, apoptotic [45-47], platelet aggregation inhibitor, neuroprotective [40], and LDL oxidation inhibitor [38]. Protocatechuic acid is the major metabolite of anthocyanins [48,49]. Many protocatechuic acid glucosides are also found naturally. For example the 3-O-\(\beta\)-glucopyranoside is reported in lobelia (Lobelia sessilifolia), the 4-O-\(\beta\)-glucopyranoside in turnip fern (Angiopteris lygodiifolia) and in the oriental and American cockroaches (Blatta orientalis and Periplaneta americana) perhaps coming from their diet rather than endogenously produced, dracunculifoside B in the groundsel relative Baccharis dracunculifolia, and the 4-O-(4-O-methyl-\(\beta\)-D-glucopyranoside) in Japanese climbing fern (Lygodium japonicum).

Vanillic acid (4-hydroxy-3-methoxybenzoic acid, 11, Figure 5) occurs in many plants such as prickly ash (Fagara spp.), Japanese alder (Alnus japonica), spiny oleaster (Elaeagnus pungens), Spanish heath (Erica australis), upland cotton (Gossypium mexicanum), Chinaberry (Melia azedarach), oriental ginseng (Panax ginseng), Korean peroba (Paratecoma koraiensis), red sandalwood (Pterocarpus santalinus), dog rose (Rosa canina), shensi (Picrorhiza kurrooa), luo shi (Trachelospermum Asiaticum), shishi (Amburana cearensis), and Shiitake mushroom (Lentinula edodes). Besides antisickling and anthelmintic activities, vanillic acid could suppress hepatic fibrosis in chronic liver injury [50,51]. It is also found to be an inhibitor of snake venom 5'-nucleotidase [52].

Figure 5. Protocatechuic acid (10), Vanillic acid (11), and Isovanillic acid (12).

| Number | Compound Description | Structural Formula |
|--------|----------------------|--------------------|
| 10     | Protocatechuic acid (R1=R2=H) | ![Protocatechuic acid](COOH) OR1 OR2 |
| 11     | Vanillic acid (R1=CH3, R2=H) | Vanillic acid (R1=CH3, R2=H) |
| 12     | Isovanillic acid (R1=H, R2=CH3) | Isovanillic acid (R1=H, R2=CH3) |

Isovanillic acid (3-hydroxy-4-methoxybenzoic acid, 12, Figure 5) is a methyl ether derivative of protocatechuic acid. It is found in hortensia (Hydrangea macrophylla), Chinese endospermum tree (Endospermum chinense) [53], the orange relative Citrus changshan-huyou [54], Chinese banyan...
(Ficus microcarpa) [55], the chamomile relative Anthemis melaleuca [56], poonspar (Calophyllum polyanthum) [57], sanchi ginseng (Panax notoginseng) [58], Formosa koa (Acacia confusa) [59,60], the breadfruit relative Treculia obovoidea [61], and saffron (Crocus sativus) [62]. Isovanillic acid has antibacterial [56,61] and antioxidant [59,60] activities.

Gallic acid (3,4,5-trihydroxybenzoic acid, 13, Figure 6) is a widespread phytochemical that occurs in tallow-tree (Allanblackia floribunda), the mangosteen relative Garcinia densivenia, bridelia (Bridelia micrantha), sappanwood (Caesalpinia sappan), elephant-apple (Dillenia indica), cinnabar ebony (Diospyros cinnabarina), peroba (Paratecom peroba), guava (Psidium guajava), water-berry (Syzygium cordatum), staghorn sumac (Rhus typhina), tamarisk (Tamarix nilotica), grape (Vitis vinifera), witch-hazel (Hamamelis virginiana) [63], and red toon (Toona sinensis) [64]. It has uses as astringent and styptic. Besides having antineoplastic and bacteriostatic activities, gallic acid possesses antimelanogenic and antioxidant properties [65]. A phenolic fraction from evening primrose (Oenothera biennis) containing gallic acid showed anti-tumour activity [66]. Gallic acid has been proposed to be a candidate for treatment of brain tumours as it suppresses cell viability, proliferation, invasion, and angiogenesis in human glioma cells [72], although the cytotoxic effects of tannins are generally not specific to tumour cells. Gallic acid induced HeLa cervical cancer cells death [73]. Many gallic acid derivatives (as phenolic acids) are naturally occurring. This includes 3-O-β-D-glucopyranoside (3-glucogallic acid) from rhubarb (Rheum spp.), 3-O-(6-galloylglucoside) from rhubarb and great burnet (Sanguisorba officinalis), 3-O-[β-D-apiofuranosyl-(1→6)-β-D-glucopyranoside] (or mudanoside B) from tree peony (Paeonia suffruticosa), 4-O-(6-galloylglucoside) from rhubarb, 3-O-dodecanoyl (3-lauroylgallic acid) with antioxidant and antimicrobial activities from the palm tree Satakentia liukuensis, 3-methyl ether from the geranium Geranium collinum and the knotweed relative mu liao (Atraphaxis frutescens), 3-methyl-5-O-sulfate (as salts) from sea-heath (Frankenia laevis) and tamarisk (Tamarix amplexicaulis), 3-methyl-4-O-[3,4-dihydroxy-5-methoxybenzoyl-(→6)-β-D-glucopyranoside] (or bistortaside A) from bistort (Polygonum bistorta), 3-methyl-5-O-β-D-glucopyranoside from the dogbane relative Tabernaemontana cymosa, 3-methyl ether from the cashew relative Poupartia axillaris and the related smooth sumac (Rhus glabra), 3-ethyl ether from emblic (Phyllanthus emblica), and 4-ethyl ether from mimosas (Mimosa hamata, Mimosa rubicula), logwood (Haematoxyllum campechianum), strawberry-tree (Arbutus unedo), cider tree (Eucalyptus gunnii), black myrobalan (Terminalia chebula) and the toxic legume Elephantorrhiza elephantina.

Syringic acid (4-hydroxy-3,5-dimethoxybenzoic acid, 14, Figure 6) occurs in many natural sources including Chinese catalpa (Catalpa ovata), garden balsam (Impatiens balsamina), New Jersey tea (Ceanothus americanus), Citrus spp., soybean (Glycine max), saxifrages (Saxifragaceae), thyme (Thymus vulgaris), summer savory (Satureja hortensis), hyssop (Hyssopus officinalis), rosemary (Rosmarinus officinalis) [74], pot marigold (Calendula officinalis) [75], tinder fungus (Phellinus igniarius) [76], golden eye grass (Curculigo orchioides) [77], date (Phoenix dactylifera) [78], sea hibiscus (Hibiscus tiliaceus) [79], Natal mahogany (Trichilia emetica) [80], birch conk (Inonotus
obliquus) [81], chickory (Cichorium intybus) [82], finger millet (Eleusine coracana) [83], woad (Isatis tinctoria) [84], clove (Syzygium aromaticum) [85], shiitake (Lentinula edodes) [50], the African medicinal shrub Anisophyllea dichostyla [86], French tamarisk (Tamarix gallica) [87], the Brazilian medicinal tree Caraipa densifolia [88], propolis (resinous materials gathered by bees from tree buds, sap flows and various other botanical sources, obtained in this case from Turkey) [89], rhododendrons (Rhododendron spp.) [90], medlar (Mespilus germanica) [14], and several other cereal grains such as barley, maize, millet, oat, rice, rye, sorghum, and wheat [91]. Besides being an antioxidant, syringic acid has antibacterial [84] and hepatoprotective [50,51] activities.

Digallic acid (\([3,4\text{-dihydroxy-5-[(3,4,5-trihydroxybenzoyl)oxy]benzoic acid}\), 15, Figure 6) is isolated from sweet acacia (Acacia farnesiana), gum arabic (Acacia arabica), dawn redwood (Metasequoia glyptostroboides), chinkapin (Castanopsis spp.), oriental white oak (Quercus aliena) [92], mango (Mangifera indica) [93], Chinese sumac (Rhus chinensis) [94], wild granadilla (Adenia cissampeloides) [95], black myrobalan (Terminalia chebula) [96], and mastic (Pistacia lentiscus) [97]. It is an HIV reverse transcriptase inhibitor. Digallic acid has cytotoxic/antiapoptotic activity [3]. It also shows antigenotoxic and antioxidant activities [97].

**Figure 6.** Gallic acid (13), Syringic acid (14), and Digallic acid (15).

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\begin{align*}
13. \text{ Gallic acid (R}_1\text{=R}_2\text{=H)} \\
14. \text{ Syringic acid (R}_1\text{=R}_2\text{=CH}_3) \\
15. \text{ Digallic acid (R}_1\text{=H, R}_2\text{=gallate)}
\end{align*}
\]

Lunularic acid (\([2\text{-hydroxy-6-[(2-(4-hydroxyphenyl)ethyl)benzoic acid}\), 16, Figure 7) has been isolated from hortensia (Hydrangea macrophylla), the liverworts Lunularia cruciata [98], Riella spp., Marchantia polymorpha, Blasia pusilla, and Riccia spp. [99], and celery (Apium graveolens) [100]. It has growth inhibitory and dormancy-inducing effects for lower plants [101]. It has also shown fungicidal, algicidal and antihyaluronidase activities [102].

Hydrangeic acid ([2-hydroxy-6-[2-(4-hydroxyphenyl)ethenyl]benzoic acid], 17, Figure 7) is a stilbenecarboxylic acid constituent of hortensia (Hydrangea macrophylla) [98]. Hydrangeic acid possesses anti-diabetic activity and lowers blood glucose, triglyceride and free fatty acid levels [103].

**Figure 7.** Lunularic acid (16) and Hydrangeic acid (17).

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\begin{align*}
16. \text{ Lunularic acid} \\
17. \text{ Hydrangeic acid (1',2'-E-didehydro)}
\end{align*}
\]

Pinosylvic acid ([2,4-dihydroxy-6-styrylbenzoic acid], 18, Figure 8) is another stilbenecarboxylic acid found in climbing skullcap (Scutellaria scandens). The leaves of this plant are traditionally used to treat wounds and swelling by insects [104].
4-O-Methylpinosylvic acid (2-hydroxy-4-methoxy-6-styrylbenzoic acid, 19, Figure 8) is the methyl ether derivative of pinosylvic acid found in leaves of pigeon pea (Cajanus cajan) [105]. The 4-O-β-D-glucopyranoside derivative of pinosylvic acid, called gaylussacin (20, Figure 8), is found in black huckleberry (Gaylussacia baccata), dangleberry (Gaylussacia frondosa) and climbing skullcap (Scutellaria scandens) [106].

**Figure 8.** Pinosylvic acid (18), 4-O-Methylpinosylvic acid (19), and Gaylussacin (20).

Anacardic acid (6-pentadecyl-2-hydroxybenzoic acid, 21, Figure 9) is a phenolic lipid; one of the 6-alkylated-2-hydroxybenzoic acids. The name “anacardic acid” is also used for a mixture of different 6-alkylated-2-hydroxybenzoic acids in which the alkyl chain is either saturated or unsaturated. Anacardic acid is found in cashew (Anacardium occidentale) [107], ginkgo (Ginkgo biloba), sumac (Rhus javanica), zonal geranium (Ozoroa mucronata), pistachio (Pistacia vera), the Thai medicinal tree Knema elegans, heart-leaf philodendron (Philodendron scandens), California figwort (Scrophularia californica), and cuachalalate (Amphipterygium adstringens). A mixture of anacardic acids showed antibacterial action against methicillin-resistant *Staphylococcus aureus* (MRSA) [108]. Some anacardic acids have also been found to be lipoxygenase inhibitors [109]. Anacardic acids prevent generation of superoxide radicals by inhibiting xanthine oxidase [110]. Anacardic acid has bioactivity against Colorado potato beetle (*Leptinotarsa decemlineata*) larvae [111]. An anacardic acid mixture has shown anti-*Helicobacter pylori* activity [112].

Ginkgolic or ginkgoic acid ([2-hydroxy-6-(8-pentadecenyl)benzoic acid], 22, Figure 9) is a derivative of anacardic acid isolated from ginkgo [113,114] and cashew [115]. Besides antitumor and antitubercular activities, ginkgolic acid inhibits protein SUMOylation. Small ubiquitin-related modifier proteins (SUMO) control several cellular functions, which can be related to cancer and neurodegenerative diseases [116].

**Figure 9.** Anacardic acid (21) and Ginkgolic acid (22).

Turgorins such as turgorin A (23, Figure 10) are Periodic Leaf Movement Factor (PLMF) substances isolated from honey locust (*Gleditsia triacanthos*) (PLMF1, PLMF3-6), karoo-thorn (*Acacia karroo*) (PLMF1-2), sensitive-plant (*Mimosa pudica*) (PLMF1), yellow wood-sorrel (*Oxalis*
stricta) (PLMF3), silk tree (*Albizia julibrissin*) (K-PLMF1), black locust (*Robinia pseudoacacia*), and hairy Indian mallow (*Abutilon grandifolium*). They are believed to be substances that control thigmonastic (touch-sensitive) and nyctinastic (diurnal light and temperature-sensitive) leaf movements [117]. Recent studies have shown that nyctinastic leaf movement is not regulated by plant hormones but rather by chemicals that differ depending on the plant species [118,119]. For example, the potassium salt of PLMF1 is the leaf-closing substance for *Mimosa pudica* [120].

**Figure 10.** Turgorin A (23).

![Turgorin A](image)

Merulinic acid A (24, Figure 11) is a phenolic lipid isolated from basidiomycetes such as *Hapalopilus mutans* [121], *Phlebia radiata*, and *Merulius tremellosus*. It has antibacterial activity, for example against *Arthrobacter citreus*, *Bacillus subtilis*, *Corynebacterium insidiosum*, *Micrococcus roseus*, and *Sarcina lutea* [122]. Merulinic acid A has pronounced promotory and/or inhibitory activities on biological membranes as an amphiphilic molecule [123].

**Figure 11.** Merulinic acid A (24).

![Merulinic acid A](image)

Platencin (25, Figure 12) and its analogs (platencin A1-A4) were isolated from the bacterium *Streptomyces platensis* [124-126]. They have been found to be dual FabF and FabH inhibitors of bacterial fatty acid biosynthesis enzymes, dubbed ‘Superbug challengers’ [127]. Superbugs are bacteria resistant to almost all antibiotics. Platencin shows broad-spectrum antibacterial activity against gram-positive pathogens such as *S. aureus*, MRSA, macrolide- and Linezolid-resistant *S. aureus*, Vancomycin intermediate *S. aureus*, Vancomycin-resistant *enterococci* and *Streptococcus pneumonia* [128].

Platensimycin (26, Figure 12) is another superbug challenger produced by *Streptomyces platensis* isolated from soil [129,130]. Platensimycin is an inhibitor of cellular lipid biosynthesis and active against gram-positive bacteria including MRSA [131,132].
Lasalocid (Lasalocid A, 27, Figure 13) is an ionophorous (transport-inducing) [133] antibiotic produced by *Streptomyces lasaliensis*. Its sodium salt is used as an antiprotozoal in veterinary practice for the prevention of coccidiosis [134].

Cannabidiolic acid (28, Figure 14) is a cannabinoid from marijuana (*Cannabis sativa*) [135-136]. It is a selective cyclooxygenase-2 inhibitor [137], TRPA1 (a member of the transient receptor potential channel family) and TRPV1 (a member of the transient receptor potential vanilloid family) agonist and TRPM8 (a member of the transient receptor potential cation channel family) antagonist [138]. Cannabidiolic acid exerts anti-proliferative actions [139].

Cajaninstilbene acid (3-hydroxy-4-prenyl-5-methoxystilbene-2-carboxylic acid, 29, Figure 15) is a stilbenecarboxylic acid found in pigeon pea (*Cajanus cajan*) [140]. It has hypotriglyceridic and hypoglycaemic activities [141,142]. Besides being a good antioxidant [143,144], cajaninstilbene acid has potential use in the treatment of postmenopausal osteoporosis [145]. It also showed anti-inflammatory, impermeability (not permitting fluids to pass through) and analgesic effects [146].
Isocajaninstilbene acid (6-hydroxy-4-methoxy-3-prenyl-2-styrylbenzoic acid, 30, Figure 15) is an isoprenylated stilbene-2-carboxylic acid also found in the leaves of pigeon pea [105,147].

**Figure 15. Cajaninstilbene acid (29) and Isocajaninstilbene acid (30).**

![Chemical structures of Cajaninstilbene acid and Isocajaninstilbene acid](image)

**29. Cajaninstilbene acid (R₁=H, R₂=prenyl)**

**30. Isocajaninstilbene acid (R₁=prenyl, R₂=H)**

3. Conclusions

The structural features common to the 30 compounds described in this review are the presence of benzoic and phenolic functional groups on a core monocyclic carbon skeleton. This does not imply a common biosynthetic origin. Many of these compounds arise from the shikimic acid pathway that starts with the coupling of phosphoenolpyruvate and d-erythrose-4-phosphate to give the core 6-membered ring with one carboxyl and three hydroxyl substituents. However, other molecules with similar functionality, such as the orsellinic acids, cannabidiolic acid and 6-methylsalicylic acid, are biosynthesized through the acetate pathway via polyketide intermediates. This indicates that the source organisms have a variety of routes by which these monocyclic phenolic acids can be synthesized.

By providing detailed descriptions of the source organisms for these monocyclic phenolic acids, we have endeavored to demonstrate that unlike many secondary metabolites which have a very restricted distribution in the bacterial, algal, fungal, and plant (and to a much lesser and generally secondary extent, animal) kingdoms, many of the compounds discussed here are found in a wide diversity of unrelated plant, algal, fungal, and bacterial species. Since, as secondary metabolites, their biosynthesis arises from mutations in the genes coding for enzymes involved in the biosynthesis of primary metabolites, a wide distribution in distantly related or unrelated species suggests that the mutations occurred early in phylogeny and are highly conserved and/or they occurred more recently and frequently across the taxa, and have been conserved. In either case, their frequent occurrence suggests that many of these phenolic acids confer advantages to the survival of the source organisms.

Despite their various biosynthetic origins, many of these molecules have been shown in experimental studies to have similar biological functions. For example, they have antioxidant, antimutagenic and even leaf movement regulating agents that protect the organism that produces them from the oxidative stress created by metabolism and their physical environment. They also have antiviral, antibacterial (bactericidal, bacteriostatic), algicidal, plant growth regulating, phytotoxic, antifungal, antiprotozoal, nematicidal, insecticidal, antifeedant, and mammalian estrogenic, keratolytic, platelet aggregation inhibiting, hypoglycemic, cytotoxic, and neurotoxic activities that may serve to protect the organism that biosynthesizes them from competing, pathogenic, and herbivorous organisms in their biological environment.
The diverse biological functions of these monocyclic phenolic acids suggest potential pharmacological activities. Thus, this review of the structures, occurrence and activities of phenolic acids can provide not only ecological insights but leads for the development of natural and derivative pharmaceutical and agricultural chemicals with implications for significant benefits to human health and nutrition.

The focus of this review on the last 10 years of peer-reviewed publications has shown that the study of the chemistry, occurrence, biological and pharmacological functions of the monocyclic phenolic acids continues to be a very active and dynamic field of investigation. From this it is reasonable to predict that many novel compounds and applications remain to be discovered.

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References

1. Martin, K.R.; Appel, C.L. Polyphenols as dietary supplements: A double-edged sword. *Nutr. Dietary Suppl.* 2010, 2, 1-12.
2. Bravo, L. Polyphenols: Chemistry, dietary sources, metabolism, and nutritional significance. *Nutr. Rev.* 1998, 56, 317-333.
3. Harris, C.S.; Mo, F.; Migahed, L.; Chepelev, L.; Haddad, P.S.; Wright, J.S.; Willmore, W.G.; Arnason, J.T.; Bennett, S.A.L. Plant phenolics regulate neoplastic cell growth and survival: a quantitative structure-activity and biochemical analysis. *Can. J. Physiol. Pharmacol.* 2007, 85, 1124-1138.
4. Huang, W.Y.; Cai, Y.Z.; Zhang, Y.B. Natural Phenolic Compounds From Medicinal Herbs and Dietary Plants: Potential Use for Cancer Prevention. *Nutr. Cancer* 2010, 62, 1-20.
5. Liu, R.H. Potential synergy of phytochemicals in cancer prevention: Mechanism of action. *J. Nutr.* 2004, 134, 3479S-3485S.
6. Stalikas, C.D. Extraction, separation, and detection methods for phenolic acids and flavonoids. *J. Sep. Sci.* 2007, 30, 3268-3295.
7. de la Rosa, L.A.; Alvarez-Parrilla, E.; Gonzalez-Aguilar, G.A. *Fruit and Vegetable Phytochemicals—Chemistry, Nutritional Value, and Stability*, 1st ed.; Wiley-Blackwell: Ames, IA, USA, 2010.
8. Lafay, S.; Gil-Izquierdo, A. Bioavailability of Phenolic acids. *Phytochem. Rev.* 2008, 7, 301-311.
9. Robbins, R.J. Phenolic acids in foods: An overview of analytical methodology. *J. Agr. Food Chem.* 2003, 51, 2866-2887.
10. Harborne, J.B.; Baxter, H.; Moss, G.P. *Phytochemical Dictionary—A Handbook of Bioactive Compounds from Plants*, 2nd ed.; Taylor & Francis: London, UK, 1999; p. 976.
11. Harborne, J.B., Ed. *Plant Phenolics*. Academic Press: San Diego, CA, USA, 1989.
12. CRC-DNP. *The Dictionary of Natural Products*. Chapman & Hall: London, UK, 2010.
13. Bendini, A.; Cerretani, L.; Carrasco-Pancorbo, A.; Gomez-Caravaca, A.M.; Segura-Carretero, A.; Fernandez-Gutierrez, A.; Lercker, G. Phenolic molecules in virgin olive oils: a survey of their
sensory properties, health effects, antioxidant activity and analytical methods. An overview of the last decade. Molecules 2007, 12, 1679-1719.

14. Gruz, J.; Ayaz, F.A.; Torun, H.; Strand, M. Phenolic acid content and radical scavenging activity of extracts from medlar (Mespilus germanica L.) fruit at different stages of ripening. Food Chem. 2011, 124, 271-277.

15. Ford, C.M.; Hoj, P.B. Multiple glucosyltransferase activities in the grapevine Vitis vinifera L. Aust. J. Grape and Wine Res. 1998, 4, 48-58.

16. Sirca, D.; Mitra, A. Accumulation of p-hydroxybenzoic acid in hairy roots of Daucus carota 2: Confirming biosynthetic steps through feeding of inhibitors and precursors. J. Plant Physiol. 2009, 166, 1370-1380.

17. Chong, K.P.; Rossall, S.; Atong, M. In Vitro Antimicrobial Activity and Fungitoxicity of Syringic Acid, Caffeic Acid and 4-hydroxybenzoic Acid against Ganoderma Boninense. J. Agr. Sci. 2009, 1, 15-20.

18. Ling, W.C.; Ahmad, F.; Mat Ali, R. Luteolin and 4-Hydroxybenzoic Acid from the Leaves Vitex negundo L. Malaysian J. Sci. 2005, 24, 133-135.

19. Chakraborty, M.; Das, K.; Dey, G.; Mitra, A. Unusually high quantity of 4-hydroxybenzoic acid accumulation in cell wall of palm mesocarps. Biochem. Syst. Ecol. 2006, 34, 509-513.

20. Pugazhendhi, D.; Pope, G.S.; Darbre, P.D. Oestrogenic activity of p-hydroxybenzoic acid (common metabolite of paraben esters) and methylparaben in human breast cancer cell lines. J. Appl. Toxicol. 2005, 25, 301-309.

21. Kamaya, Y.; Tsuboi, S.; Takada, T.; Suzuki, K. Growth stimulation and inhibition effects of 4-hydroxybenzoic acid and some related compounds on the freshwater green alga Pseudokirchneriella subcapitata. Arch. Environ. Contam. Toxicol. 2006, 51, 537-541.

22. Muljono, R.A.B.; Darsono, F.L.; Scheffer, J.J.C.; Verpoorte, R. Assay of 2,3-dihydroxybenzoic acid and related compounds in plant materials by high-performance liquid chromatography. J. Chromatogr. A 2001, 927, 39-45.

23. Sugiyama, Y.; Hirota, A. New Potent DPPH Radical Scavengers from a Marine-Derived Actinomycete Strain USF-TC31. Biosci. Biotechnol. Biochem. 2009, 73, 2731-2734.

24. Onofrejova, L.; Vasickova, J.; Klejdus, B.; Stratil, P.; Misurcova, L.; Kraacmar, S.; Kopecky, J.; Vacek, J. Bioactive phenols in algae: The application of pressurized-liquid and solid-phase extraction techniques. J. Pharm. Biomed. Anal. 2010, 51, 464-470.

25. Sroka, Z.; Cisowski, W. Hydrogen peroxide scavenging, antioxidant and anti-radical activity of some phenolic acids. Food Chem. Toxicol. 2003, 41, 753-758.

26. Parent, M.A.; Bellaire, B.H.; Murphy, E.A.; Roop, R.M.; Elzer, P.H.; Baldwin, C.L. Brucella abortus siderophore 2,3-dihydroxybenzoic acid (DHBA) facilitates intracellular survival of the bacteria. Microb. Pathog. 2002, 32, 239-248.

27. Ashidate, K.; Kawamura, M.; Mimura, D.; Tohda, H.; Miyazaki, S.; Teramoto, T.; Yamamoto, Y.; Hirata, Y. Gentisic acid, an aspirin metabolite, inhibits oxidation of low-density lipoprotein and the formation of cholesterol ester hydroperoxides in human plasma. Eur. J. Pharmacol. 2005, 513, 173-179.

28. Sharma, S.; Khan, N.; Sultana, S. Study on prevention of two-stage skin carcinogenesis by Hibiscus rosa sinensis extract and the role of its chemical constituent, gentisic acid, in the
29. Fernandez, I.S.; Cuevas, P.; Angulo, J.; Lopez-Navajas, P.; Canales-Mayordomo, A.; Gonzalez-Corrochano, R.; Lozano, R.M.; Valverde, S.; Jimenez-Barbero, J.; Romero, A.; Gimenez-Gallego, G. Gentisic Acid, a Compound Associated with Plant Defense and a Metabolite of Aspirin, Heads a New Class of in vivo Fibroblast Growth Factor Inhibitors. J. Biol. Chem. 2010, 285, 11714-11729.

30. Stochmal, A.; Kowalska, I.; Janda, B.; Perrone, A.; Piacente, S.; Oleszek, W. Gentisic acid conjugates of Medicago truncatula roots. Phytochemistry 2009, 70, 1272-1276.

31. Sultana, N.; Akhter, M.; Khatoon, Z. Nematicidal natural products from the aerial parts of Rubus niveus. Nat. Prod. Res. 2010, 24, 407-415.

32. Lin, A.N.; Nakatsui, T. Salicylic acid revisited. Int. J. Dermatol. 1998, 37, 335-342.

33. Shah, J. The salicylic acid loop in plant defense. Curr. Opin. Plant Biol. 2003, 6, 365-371.

34. Loake, G.; Grant, M. Salicylic acid in plant defence-the players and protagonists. Curr. Opin. Plant Biol. 2007, 10, 466-472.

35. Wattallachaisaereekul, S.; Lantz, A.E.; Nielsen, M.L.; Andresson, O.S.; Nielsen, J. Optimization of heterologous production of the polyketide 6-MSA in Saccharomyces cerevisiae. Biotechnol. Bioeng. 2007, 97, 893-900.

36. Stuart, A.E.; Brooks, C.J.W.; Prescott, R.J.; Blackwell, A. Repellent and antifeedant activity of salicylic acid and related compounds against the biting midge, Culicoides impunctatus (Diptera: Ceratopogonidae). J. Med. Entomol. 2000, 37, 222-227.

37. Haschek, W.M.; Wallig, M.A.; Rousseaux, C. Fundamentals of Toxicologic Pathology. 2nd ed.; Academic Press: San Diego, CA, USA, 2010.

38. Hur, J.M.; Park, J.G.; Yang, K.H.; Park, J.C.; Park, J.R.; Chun, S.S.; Choi, J.S.; Choi, J.W. Effect of methanol extract of Zanthoxylum piperitum leaves and of its compound, protocatechuic acid, on hepatic drug metabolizing enzymes and lipid peroxidation in rats. Biosci. Biotechnol. Biochem. 2003, 67, 945-950.

39. Pan, Y.L.; Zhang, L.; Chen, G.N. Separation and determination of protocatechuic aldehyde and protocatechuic acid in Salvia miltiorrhiza by capillary electrophoresis with amperometric detection. Analyst 2001, 126, 1519-1523.

40. An, L.J.; Guan, S.; Shi, G.F.; Bao, Y.M.; Duan, Y.L.; Jiang, B. Protocatechuic acid from Alpinia oxyphylla against MPP+-induced neurotoxicity in PC12 cells. Food Chem. Toxicol. 2006, 44, 436-443.

41. Gutzeit, D.; Wray, V.; Winterhalter, P.; Jerz, G. Preparative isolation and purification of flavonoids and protocatechuic acid from sea buckthorn juice concentrate (Hippophae rhamnoides L. ssp rhamnoides) by high-speed counter-current chromatography. Chromatographia 2007, 65, 1-7.

42. Yip, E.C.H.; Chan, A.S.L.; Pang, H.; Tam, Y.K.; Wong, Y.H. Protocatechuic acid induces cell death in HepG2 hepatocellular carcinoma cells through a c-Jun N-terminal kinase-dependent mechanism. Cell Biol. Toxicol. 2006, 22, 293-302.
43. Fukuji, T.S.; Tonin, F.G.; Tavares, M.F.M. Optimization of a method for determination of phenolic acids in exotic fruits by capillary electrophoresis. *J. Pharm. Biomed. Anal.* 2010, 51, 430-438.

44. Vari, R.; D'Archivio, M.; Filesi, C.; Carotenuto, S.; Scazzocchio, B.; Santangelo, C.; Giovannini, C.; Masella, R. Protocatechuic acid induces antioxidant/detoxifying enzyme expression through JNK-mediated Nrf2 activation in murine macrophages. *J. Nutr. Biochem.* 2010, in press.

45. Guan, S.; Jiang, B.; Bao, Y.M.; An, L.J. Protocatechuic acid suppresses MPP+-induced mitochondrial dysfunction and apoptotic cell death in PC12 cells. *Food Chem. Toxicol.* 2006, 44, 1659-1666.

46. Liu, Y.M.; Jiang, B.; Bao, Y.M.; An, L.J. Protocatechuic acid inhibits apoptosis by mitochondrial dysfunction in rotenone-induced PC12 cells. *Toxicol. In Vitro* 2008, 22, 430-437.

47. Yin, M.C.; Lin, C.C.; Wu, H.C.; Tsao, S.M.; Hsu, C.K. Apoptotic Effects of Protocatechuic Acid in Human Breast, Lung, Liver, Cervix, and Prostate Cancer Cells: Potential Mechanisms of Action. *J. Agr. Food Chem.* 2009, 57, 6468-6473.

48. Galvano, F.; Vitaglione, P.; Volti, G.L.; Di Giacomo, C.; Gazzo, D.; Vanella, L.; La Faucci, L.; Fogliano, V. Protocatechuic acid: The missing human cyanidins' metabolite. *Mol. Nutr. Food Res.* 2008, 52, 386-387.

49. Vitaglione, P.; Donnarumma, G.; Napolitano, A.; Galvano, F.; Gallo, A.; Scalfi, L.; Fogliano, V. Protocatechuic acid is the major human metabolite of cyanidin-glucosides. *J. Nutr.* 2007, 137, 2043-2048.

50. Itoh, A.; Isoda, K.; Kondoh, M.; Kawase, M.; Kobayashi, M.; Tamesada, M.; Yagi, K. Hepatoprotective Effect of Syringic Acid and Vanillic Acid on Concanavalin A-Induced Liver Injury. *Biol. Pharm. Bull.* 2009, 32, 1215-1219.

51. Itoh, A.; Isoda, K.; Kondoh, M.; Kawase, M.; Watari, A.; Kobayashi, M.; Tamesada, M.; Yagi, K. Hepatoprotective Effect of Syringic Acid and Vanillic Acid on CCl4-Induced Liver Injury. *Biol. Pharm. Bull.* 2010, 33, 983-987.

52. Dhananjaya, B.L.; Nataraju, A.; Gowda, C.D.R.; Sharath, B.K.; D'Souza, C.J.M. Vanillic acid as a novel specific inhibitor of snake venom 5'-nucleotidase: A pharmacological tool in evaluating the role of the enzyme in snake envenomation. *Biochem.-Moscow* 2009, 74, 1315-1319.

53. Li, X.; Lin, L.; Wu, P.; Liu, M.; Wei, X. Chemical Constituents from Barks of *Endospermum chinense* Benth. *J. Trop. Subtrop. Bot.* 2007, 1, Q946.

54. Zhao, X.; Ye, X.; Zhu, D. Isolation and identification of chemical constituents from peels of *Citrus changshan-huyou* Y.B.Chang. *J. Peking Univ.* 2009, 5, 575-577.

55. Lai, Y.; Zhou, H. Isolation and Identification of the Chemical Constituents of *Folium Fici Microcarpa*. *Food Drug* 2008, 3, 11-13.

56. Saroglou, V.; Karioti, A.; Rancic, A.; Dimas, K.; Koukoulitsa, C.; Zervou, M.; Skaltsa, H. Sesquiterpene Lactones from *Anthemis melanolepis* and Their Antibacterial and Cytotoxic Activities. Prediction of Their Pharmacokinetic Profile. *J. Nat. Prod.* 2010, 73, 242-246.

57. Zhong, H.; Ruan, J.L.; Yao, Q.Q. Two new 4-arylcoumarins from the seeds of *Calophyllum polyanthum*. *J. Asian Nat. Prod. Res.* 2010, 12, 562-568.

58. Komakine, N.; Okasaka, M.; Takashi, Y.; Kawazoe, K.; Murakami, K.; Yamada, Y. New dammarane-type saponin from roots of *Panax notoginseng*. *J. Nat. Med.* 2006, 60, 135-137.
59. Wu, J.H.; Tung, Y.T.; Wang, S.Y.; Shyur, L.F.; Kuo, Y.H.; Chang, S.T. Phenolic antioxidants from the heartwood of Acacia confusa. J. Agr. Food Chem. 2005, 53, 5917-5921.
60. Tung, Y.T.; Wu, J.H.; Kuo, Y.H.; Chang, S.T. Antioxidant activities of natural phenolic compounds from Acacia confusa bark. Bioresour. Technol. 2007, 98, 1120-1123.
61. Kuete, V.; Metuno, R.; Ngameni, B.; Tsafack, A.M.; Ngandeu, F.; Fotso, G.W.; Bezabih, M.; Etoa, F.X.; Ngadjui, B.T.; Abegaz, B.M.; Beng, V.P. Antimicrobial activity of the methanolic extracts and compounds from Treculia obovoidea (Moraceae). J. Ethnopharmacol. 2007, 112, 531-536.
62. Li, C.Y.; Lee, E.J.; Wu, T.S. Antityrosinase principles and constituents of the petals of Crocus sativus. J. Nat. Prod. 2004, 67, 437-440.
63. Wang, H.F.; Provan, G.J.; Helliwell, K. Determination of hamamelitannin, catechins and gallic acid in witch hazel bark, twig and leaf by HPLC. J. Pharm. Biomed. Anal. 2003, 33, 539-544.
64. Chen, H.M.; Wu, Y.C.; Chia, Y.C.; Chang, F.R.; Hsu, H.K.; Hsieh, Y.C.; Chen, C.C.; Yuan, S.S. Gallic acid, a major component of Toona sinensis leaf extracts, contains a ROS-mediated anti-cancer activity in human prostate cancer cells. Cancer Lett. 2009, 286, 161-171.
65. Kim, Y.J. Antimelanogenic and antioxidant properties of gallic acid. Biol. Pharm. Bull. 2007, 30, 1052-1055.
66. Dalla Pellegrina, C.; Padovani, G.; Mainente, F.; Zoccatelli, G.; Bissoli, G.; Mosconi, S.; Veneri, G.; Peruffo, A.; Andrighetto, G.; Rizzi, C.; Chignola, R. Anti-tumour potential of a gallic acid-containing phenolic fraction from Oenothera biennis. Cancer Lett. 2005, 226, 17-25.
67. Agarwal, C.; Tyagi, A.; Agarwal, R. Gallic acid causes inactivating phosphorylation of cdc25A/cdc25C-cdc2 via ATM-Chk2 activation, leading to cell cycle arrest, and induces apoptosis in human prostate carcinoma DU145 cells. Mol. Cancer Ther. 2006, 5, 3294-3302.
68. Kaur, M.; Velmurugan, B.; Rajamanickam, S.; Agarwal, R.; Agarwal, C. Gallic Acid, an Active Constituent of Grape Seed Extract, Exhibits Anti-proliferative, Pro-apoptotic and Anti-tumorigenic Effects Against Prostate Carcinoma Xenograft Growth in Nude Mice. Pharm. Res. 2009, 26, 2133-2140.
69. Liu, Z.J.; Schwimer, J.; Liu, D.; Lewis, J.; Greenway, F.L.; York, D.A.; Woltering, E.A. Gallic acid is partially responsible for the antiangiogenic activities of Rubus leaf extract. Phytother. Res. 2006, 20, 806-813.
70. Gupta, N.; Gupta, S.; Mahmood, A. Gallic acid inhibits brush border disaccharidases in mammalian intestine. Nutr. Res. 2007, 27, 230-235.
71. Kratz, J.M.; Andrighetti-Frohner, C.R.; Leal, P.C.; Nunes, R.J.; Yunes, R.A.; Trybala, E.; Bergstrom, T.; Barardi, C.R.M.; Simoes, C.M.O. Evaluation of anti-HSV-2 activity of gallic acid and pentyl gallate. Biol. Pharm. Bull. 2008, 31, 903-907.
72. Lu, Y.; Jiang, F.; Jiang, H.; Wu, K.; Zheng, X.; Cai, Y.; Katakowski, M.; Chopp, M.; To, S.S.T. Gallic acid suppresses cell viability, proliferation, invasion and angiogenesis in human glioma cells. Eur. J. Pharmacol. 2010, 641, 102-107.
73. You, B.R.; Moon, H.J.; Han, Y.H.; Park, W.H. Gallic acid inhibits the growth of HeLa cervical cancer cells via apoptosis and/or necrosis. Food Chem. Toxicol. 2010, 48, 1334-1340.
74. Zgorka, G.; Glowniak, K. Variation of free phenolic acids in medicinal plants belonging to the Lamiaceae family. J. Pharm. Biomed. Anal. 2001, 26, 79-87.
75. Matysik, G.; Wojciak-Kosior, M.; Paduch, R. The influence of Calendulae officinalis flos extracts on cell cultures, and the chromatographic analysis of extracts. *J. Pharm. Biomed. Anal.* 2005, 38, 285-292.

76. Mo, S.Y.; Wang, S.J.; Zhou, G.X.; Yang, Y.C.; Li, Y.; Chen, X.G.; Shi, J.G. Phelligidins C-F: Cytotoxic pyrano[4,3-c][2]benzopyran-1,6-dione and furo[3,2-c]pyran-4-one derivatives from the fungus Phellinus igniarius. *J. Nat. Prod.* 2004, 67, 823-828.

77. Wu, Q.; Fu, D.X.; Hou, A.J.; Lei, G.Q.; Liu, Z.J.; Chen, J.K.; Zhou, T.S. Antioxidative phenols and phenolic glycosides from Curculigo orchioides. *Chem. Pharm. Bull.* 2005, 53, 1065-1067.

78. Al-Farsi, M.; Alasalvar, C.; Morris, A.; Baron, M.; Shahidi, F. Comparison of antioxidant activity, anthocyanins, carotenoids, and phenolics of three native fresh and sun-dried date (Phoenix dactylifera L.) varieties grown in Oman. *J. Agr. Food Chem.* 2005, 53, 7592-7599.

79. Chen, J.J.; Huang, S.Y.; Duh, C.Y.; Chen, I.S.; Wang, T.C.; Fang, H.Y. A new cytotoxic amide from the stem wood of Hibiscus tiliaceus. *Planta Med.* 2006, 72, 935-938.

80. Germano, M.P.; D'Angelo, V.; Biasini, T.; Sanogo, R.; De Pasquale, R.; Catania, S. Evaluation of the antioxidant properties and bioavailability of free and bound phenolic acids from Trichilia emetica Vahl. *J. Ethnopharmacol.* 2006, 105, 368-373.

81. Nakajima, Y.; Sato, Y.; Konishi, T. Antioxidant small phenolic ingredients in Inonotus obliquus (persoon) Pilat (Chaga). *Chem. Pharm. Bull.* 2007, 55, 1222-1226.

82. Atta Ur, R.; Zareen, S.; Choudhary, M.I.; Akhtar, M.N.; Khan, S.N. alpha-glucosidase inhibitory activity of triterpenoids from Cichorium intybus. *J. Nat. Prod.* 2008, 71, 910-913.

83. Chethan, S.; Dharmesh, S.M.; Malleshi, N.G. Inhibition of aldose reductase from cataracted eye lenses by finger millet (Eleusine coracana) polyphenols. *Bioorg. Med. Chem.* 2008, 16, 10085-10090.

84. Kong, W.K.; Zhao, Y.L.; Shan, L.M.; Xiao, X.H.; Guo, W.Y. Thermochemical studies on the quantity - Antibacterial effect relationship of four organic acids from Radix Isatidis on Escherichia coli growth. *Biol. Pharm. Bull.* 2008, 31, 1301-1305.

85. Rastogi, S.; Pandey, M.M.; Rawat, A.K.S. High-Performance Thin-Layer Chromatography Densitometric Method for the Simultaneous Determination of Three Phenolic Acids in Syzygium aromaticum (L.) Merr. & Perry. *J. AOAC Int.* 2008, 91, 1169-1173.

86. Khallouki, F.; Hull, W.E.; Owen, R.W. Characterization of a rare triterpenoid and minor phenolic compounds in the root bark of Anisophyillea dichystyla R. Br. *Food Chem. Toxicol.* 2009, 47, 2007-2012.

87. Gulcin, I.; Bursal, E.; Sehitoglu, M.H.; Bilsel, M.; Goren, A.C. Polyphenol contents and antioxidant activity of lyophilized aqueous extract of propolis from Erzurum, Turkey. *Food Chem. Toxicol.* 2010, 48, 2227-2238.
90. Sharma, N.; Sharma, U.K.; Gupta, A.P.; Sinha, A.K. Simultaneous determination of epicatechin, syringic acid, quercetin-3-O-galactoside and quercitrin in the leaves of *Rhododendron* species by using a validated HPTLC method. *J. Food Compos. Anal.* 2010, 23, 214-219.

91. Dykes, L.; Rooney, L.W. Phenolic compounds in cereal grains and their health benefits. *Cereal Foods World* 2007, 52, 105-111.

92. Jin, Y.S.; Heo, S.I.; Lee, M.J.; Rhee, H.I.; Wang, M.H. Free radical scavenging and hepatoprotective actions of Quercus aliena acorn extract against CCl4-induced liver. *Free Radic. Res.* 2005, 39, 1351-1358.

93. Masibo, M.; He, Q. Major mango polyphenols and their potential significance to human health. *Compr. Rev. Food. Sci. Food Saf.* 2008, 7, 309-319.

94. Abbasi, A.M.; Khan, M.A.; Ahmad, M.; Zafar, M.; Khan, H.; Muhammad, N.; Sultana, S. Medicinal plants used for the treatment of jaundice and hepatitis based on socio-economic documentation. *Afr. J. Biotechnol.* 2009, 8, 1643-1650.

95. Njoku, O.V.; Obi, C. Phytochemical constituents of some selected medicinal plants. *African J. Pure Appl. Chem.* 2009, 3, 228-233.

96. Ma, H.B.; Diao, Y.P.; Zhao, D.Y.; Li, K.; Kang, T.G. A new alternative to treat swine influenza A virus infection: extracts from *Terminalia chebula* Retz. *Afr. J. Microbiol. Res.* 2010, 4, 497-499.

97. Bhouri, W.; Derbel, S.; Skandrani, I.; Boubaker, J.; Bouhlel, I.; Sghaier, M.B.; Kilani, S.; Mariotte, A.M.; Dijoux-Franca, M.G.; Ghedira, K.; Chekir-Ghedira, L. Study of genotoxic, antigenotoxic and antioxidant activities of the digallic acid isolated from *Pistacia lentiscus* fruits. *Toxicol. Vitro* 2010, 24, 509-515.

98. Eckermann, C.; Schroder, G.; Eckermann, S.; Strack, D.; Schmidt, E.; Schneider, B.; Schroder, J. Stilbenecarboxylate biosynthesis: a new function in the family of chalcone synthase-related proteins. *Phytochemistry* 2003, 62, 271-286.

99. Asakawa, Y. Chemosystematics of the *Hepaticae. Phytochemistry* 2004, 65, 623-669.

100. Zhou, K.; Wu, B.; Zhuang, Y.; Ding, L.; Liu, Z.; Qiu, F. Chemical constituents of fresh celery. *Zhongguo Zhong Yao Za Zhi* 2009, 34, 1512-1515.

101. Yoshikawa, H.; Ichiki, Y.; Sakakibara, K.; Tamura, H.; Suiko, M., The biological and structural similarity between lunularic acid and abscisic acid. *Biosci. Biotechnol. Biochem.* 2002, 66, 840-846.

102. Asakawa, Y. Biologically active compounds from bryophytes. *Pure Appl. Chem.* 2007, 79, 557-580.

103. Zhang, H.L.; Matsuda, H.; Yamashita, C.; Nakamura, S.; Yoshikawa, M. Hydrangeic acid from the processed leaves of *Hydrangea macrophylla* var. *thunbergii* as a new type of anti-diabetic compound. *Eur. J. Pharmacol.* 2009, 606, 255-261.

104. Shang, X.F.; He, X.R.; He, X.Y.; Li, M.X.; Zhang, R.X.; Fan, P.C.; Zhang, Q.L.; Jia, Z.P. The genus *Scutellaria* an ethnopharmacological and phytochemical review. *J. Ethnopharmacol.* 2010, 128, 279-313.

105. Yannai, S. *Dictionary of Food Compounds- Additives, Flavors, and Ingredients*, 1st ed.; Chapman & Hall/CRC: Boca Raton, FL, USA, 2004; p. 1784.

106. Malikov, V.M.; Yuldashev, M.P. Phenolic compounds of plants of the *Scutellaria* genus. Distribution, structure, and properties. *Chem. Nat. Compd.* 2002, 38, 473-519.
107. Philip, J.Y.N.; Francisco, J.D.; Dey, E.S.; Buchweishaija, J.; Mkayula, L.L.; Ye, L. Isolation of Anacardic Acid from Natural Cashew Nut Shell Liquid (CNSL) Using Supercritical Carbon Dioxide. *J. Agr. Food Chem.* **2008**, *56*, 9350-9354.

108. Kubo, I.; Nihei, K.I.; Tsujimoto, K. Antibacterial action of anacardic acids against methicillin resistant *Staphylococcus aureus* (MRSA). *J. Agr. Food Chem.* **2003**, *51*, 7624-7628.

109. Ha, T.J.; Kubo, I. Lipoxygenase inhibitory activity of anacardic acids. *J. Agr. Food Chem.* **2005**, *53*, 4350-4354.

110. Kubo, I.; Masuoka, N.; Ha, T.J.; Tsujimoto, K. Antioxidant activity of anacardic acids. *Food Chem.* **2006**, *99*, 555-562.

111. Schultz, D.J.; Olsen, C.; Cobbs, G.A.; Stolowich, N.J.; Parrott, M.M. Bioactivity of anacardic acid against Colorado potato beetle (*Leptinotarsa decemlineata*) larvae. *J. Agr. Food Chem.* **2006**, *54*, 7522-7529.

112. Castillo-Juarez, I.; Rivero-Cruz, F.; Celis, H.; Romero, I. Anti-Helicobacter pylori activity of anacardic acids from *Amphipterygium adstringens*. *J. Ethnopharmacol.* **2007**, *114*, 72-77.

113. van Beek, T.A.; Montoro, P. Chemical analysis and quality control of *Ginkgo biloba* leaves, extracts, and phytopharmaceuticals. *J. Chromatogr. A* **2009**, *1216*, 2002-2032.

114. van Beek, T.A. Chemical analysis of *Ginkgo biloba* leaves and extracts. *J. Chromatogr. A* **2002**, *967*, 21-55.

115. Trevisan, M.T.S.; Pfundstein, B.; Haubner, R.; Wurtele, G.; Spiegelhalder, B.; Bartsch, H.; Owen, R.W. Characterization of alkyl phenols in cashew (*Anacardium occidentale*) products and assay of their antioxidant capacity. *Food Chem. Toxicol.* **2006**, *44*, 188-197.

116. Fukuda, I.; Ito, A.; Hirai, G.; Nishimura, S.; Kawasaki, H.; Saitoh, H.; Kimura, K.; Sodeoka, M.; Yoshida, M. Ginkgolic Acid Inhibits Protein SUMOylation by Blocking Formation of the E1-SUMO Intermediate. *Chem. Biol.* **2009**, *16*, 133-140.

117. Ueda, M.; Nakamura, S. Chemistry and biology of plant leaf movements. *Angew. Chem.-Int. Edit.* **2000**, *39*, 1400-1414.

118. Ueda, M.; Nakamura, Y. Chemical basis of plant leaf movement. *Plant Cell Physiol.* **2007**, *48*, 900-907.

119. Ueda, M.; Sugimoto, T.; Sawai, Y.; Ohnuki, T.; Yamamura, S. Chemical studies on plant leaf movement controlled by a biological clock. *Pure Appl. Chem.* **2003**, *75*, 353-358.

120. Ueda, M.; Shigemori, H.; Sata, N.; Yamamura, S. The diversity of chemical substances controlling the nyctinastic leaf-movement in plants. *Phytochemistry* **2000**, *53*, 39-44.

121. Sontag, B.; Dasenbrock, J.; Arnold, N.; Steglich, W. Metabolites from the wood-rotting basidiomycete *Hapalopilus mutans* (*Aphyllophorales*). *Eur. J. Org. Chem.* **1999**, *5*, 1051-1055.

122. Zjawiony, J.K. Biologically active compounds from *aphyllophorales* (polypore) fungi. *J. Nat. Prod.* **2004**, *67*, 300-310.

123. Stasiuk, M.; Jaromin, A.; Kozubek, A. The effect of merulinic acid on biomembranes. *Biochim. Biophys. Acta-Biomembr.* **2004**, *1667*, 215-221.

124. Wang, J.; Kodali, S.; Lee, S.H.; Galgoci, A.; Painter, R.; Dorso, K.; Racine, F.; Motyl, M.; Hernandez, L.; Tinney, E.; Colletti, S.L.; Herath, K.; Cummings, R.; Salazar, O.; Gonzalez, I.; Basilio, A.; Vicente, F.; Genilloud, O.; Pelaez, F.; Jayasuriya, H.; Young, K.; Cully, D.F.; Singh,
S.B. Discovery of platencin, a dual FabF and FabH inhibitor with in vivo antibiotic properties. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 7612-7616.

125. Zhang, C.W.; Ondeyka, J.; Dietrich, L.; Gailliot, F.P.; Hesse, M.; Lester, M.; Dorso, K.; Motyl, M.; Ha, S.N.; Wang, J.; Singh, S.B. Isolation, structure and biological activities of platencin A(2)-A(4) from *Streptomyces platensis*. *Bioorg. Med. Chem.** 2010*, *18*, 2602-2610.

126. Singh, S.B.; Ondeyka, J.G.; Herath, K.B.; Zhang, C.W.; Jayasuriya, H.; Zink, D.L.; Parthasarathy, G.; Becker, J.W.; Wang, J.; Soisson, S.M. Isolation, enzyme-bound structure and antibacterial activity of platencin A(1) from *Streptomyces platensis*. *Bioorg. Med. Chem. Lett.** 2009*, *19*, 4756-4759.

127. Jayasuriya, H.; Herath, K.B.; Zhang, C.; Zink, D.L.; Basilio, A.; Genilloud, O.; Diez, M.T.; Vicente, F.; Gonzalez, I.; Salazar, O.; Pelaez, F.; Cummings, R.; Ha, S.; Wang, J.; Singh, S.B. Isolation and structure of platencin: A FabH and FabF dual inhibitor with potent broad-spectrum antibiotic activity. *Angew. Chem.-Int. Edit.** 2007*, *46*, 4684-4688.

128. Palanichamy, K.; Kaliappan, K.P. Discovery and Syntheses of "Superbug Challengers"-Platensimycin and Platencin. *Chem.-Asian J.** 2010*, *5*, 668-703.

129. Wang, J.; Soisson, S.M.; Young, K.; Shoop, W.; Kodali, S.; Galgoci, A.; Painter, R.; Parthasarathy, G.; Tang, Y.S.; Cummings, R.; Ha, S.; Dorso, K.; Motyl, M.; Jayasuriya, H.; Ondeyka, J.; Herath, K.; Zhang, C.W.; Hernandez, L.; Allocco, J.; Basilio, A.; Tormo, J.R.; Genilloud, O.; Vicente, F.; Pelaez, F.; Colwell, L.; Lee, S. H.; Michael, B.; Felcetto, T.; Gill, C.; Silver, L.L.; Hermes, J.D.; Bartizal, K.; Barrett, J.; Schmatz, D.; Becker, J.W.; Cully, D.; Singh, S.B. Platensimycin is a selective FabF inhibitor with potent antibiotic properties. *Nature** 2006*, *441*, 358-361.

130. Singh, S.B.; Jayasuriya, H.; Ondeyka, J.G.; Herath, K.B.; Zhang, C.W.; Zink, D.L.; Tsou, N.N.; Ball, R.G.; Basilio, A.; Genilloud, O.; Diez, M.T.; Vicente, F.; Pelaez, F.; Young, K.; Wang, J. Isolation, structure, and absolute stereochemistry of platensimycin, a broad spectrum antibiotic discovered using an antisense differential sensitivity strategy. *J. Am. Chem. Soc.** 2006*, *128*, 11916-11920.

131. Habich, D.; von Nussbaum, F. Platensimycin, a new antibiotic and "superbug challenger" from nature. *ChemMedChem** 2006*, *1*, 951-954.

132. Manallack, D.T.; Crosby, I.T.; Khakham, Y.; Capuano, B. Platensimycin: A promising antimicrobial targeting fatty acid synthesis. *Curr. Med. Chem.** 2008*, *15*, 705-710.

133. Edrington, T.S.; Callaway, T.R.; Varey, P.D.; Jung, Y.S.; Bischoff, K.M.; Elder, R.O.; Anderson, R.C.; Kutter, E.; Brabban, A.D.; Nisbet, D.J. Effects of the antibiotic ionophores monensin, lasalocid, laildlomycin propionate and bambermycin on *Salmonella* and *E-coli* O157: H7 in vitro. *J. Appl. Microbiol.** 2003*, *94*, 207-213.

134. Armson, A.; Thompson, R.C.A.; Reynoldso, J.A. A review of chemotherapeutic approaches to the treatment of cryptosporidiosis. *Expert Rev. Anti-infective Ther.** 2003*, *1*, 297-305.

135. Izzo, A.A.; Borrelli, F.; Capasso, R.; Di Marzo, V.; Mechoulam, R. Non-psychotropic plant cannabinoids: new therapeutic opportunities from an ancient herb. *Trends Pharmacol. Sci.** 2009*, *30*, 515-527.
136. Sirikantaramas, S.; Taura, F.; Morimoto, S.; Shoyama, Y. Recent advances in *Cannabis sativa* research: Biosynthetic studies and its potential in biotechnology. *Curr. Pharm. Biotechnol.* **2007**, *8*, 237-243.

137. Takeda, S.; Misawa, K.; Yamamoto, I.; Watanabe, K. Cannabidiolic acid as a selective cyclooxygenase-2 inhibitory component in cannabis. *Drug Metab. Dispos.* **2008**, *36*, 1917-1921.

138. De Petrocellis, L.; Vellani, V.; Schiano-Moriello, A.; Marini, P.; Magherini, P.C.; Orlando, P.; Di Marzo, V. Plant-derived cannabinoids modulate the activity of transient receptor potential channels of ankyrin type-1 and melastatin type-8. *J. Pharmacol. Exp. Ther.* **2008**, *325*, 1007-1015.

139. Ligresti, A.; Moriello, A.S.; Starowicz, K.; Matias, I.; Pisanti, S.; De Petrocellis, L.; Laezza, C.; Portella, G.; Bifulco, M.; Di Marzo, V. Antitumor activity of plant cannabinoids with emphasis on the effect of cannabidiol on human breast carcinoma. *J. Pharmacol. Exp. Ther.* **2006**, *318*, 1375-1387.

140. Kong, Y.; Zu, Y.G.; Fu, Y.J.; Liu, W.; Chang, F.R.; Li, J.; Chen, Y.H.; Zhang, S.; Gu, C.B. Optimization of microwave-assisted extraction of cajanustilbene acid and pinostrobin from pigeonpea leaves followed by RP-HPLC-DAD determination. *J. Food Compos. Anal.* **2010**, *23*, 382-388.

141. Inman, W.D.; Hoppe, D.C. Compositions containing hypotriglycerideremically active stilbenoids. W. I. P. O. I. Bureau: Geneva, Switzerland, 2002.

142. Inman, W.D.; Hoppe, D.C. Compositions containing hypoglucomically active stilbenoids. W. I. P. O. I. Bureau: Geneva, Switzerland, 2003.

143. Kong, Y.; Fu, Y.J.; Zu, Y.G.; Liu, W.; Wang, W.; Hua, X.; Yang, M. Ethanol modified supercritical fluid extraction and antioxidant activity of cajanustilbene acid and pinostrobin from pigeonpea [*Cajanus cajan* (L.) Millsp.] leaves. *Food Chem.* **2009**, *117*, 152-159.

144. Wu, N.; Fu, K.; Fu, Y.J.; Zu, Y.G.; Chang, F.R.; Chen, Y.H.; Liu, X.L.; Kong, Y.; Liu, W.; Gu, C.B., Antioxidant Activities of Extracts and Main Components of Pigeonpea [*Cajanus cajan* (L.) Millsp.] Leaves. *Molecules* **2009**, *14*, 1032-1043.

145. Sun, S.M.; Song, Y.M.; Liu, J. Studies on the pharmacology of Cajanin preparation. *Chin. Trad. Herbal Drug*. **1995**, *26*, 147-148.

146. Zheng, Y.Y.; Yang, J.; Chen, D.H.; Sun, L. Effects of stilbene extracts from *Cajanus cajan* L. on ovariectomy-induced bone loss in rats. *Acta Pharmacol. Sin.* **2007**, *42*, 562-567.

147. Cooksey, C.J.; Dahiya, J.S.; Garratt, P.J.; Strange, R.N. 2 Novel Stilbene-2-carboxylic acid Phytoalexins from *Cajanus-Cajan*. *Phytochemistry* **1982**, *21*, 2935-2938.

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