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Estimating uncertainty in density surface models

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ABSTRACT

Providing uncertainty estimates for predictions derived from species distribution models is essential for management but there is little guidance on potential sources of uncertainty in predictions and how best to combine these. Here we show where uncertainty can arise in density surface models (a multi-stage spatial modelling approach for distance sampling data), focussing on cetacean density modelling. We propose an extensible, modular, hybrid analytical-simulation approach to encapsulate these sources. We provide example analyses of fin whales Balaenoptera physalus in the California Current Ecosystem.

Subjects Conservation Biology, Marine Biology, Statistics, Natural Resource Management, Spatial and Geographic Information Science

Keywords Density surface models, Distance sampling, Uncertainty quantification, Spatial modelling, Species distribution modelling, Model uncertainty, Environmental uncertainty

INTRODUCTION

Reliable estimates of uncertainty in abundance are essential for management and conservation of biological populations. One of the most popular methods of estimating abundance is distance sampling (Buckland et al., 2001), which uses data collected on the distances between sampler and observation to estimate the detection probability. This detection probability can be used in design-based estimates (via a Horvitz-Thompson-like estimator) or in model-based estimates to obtain abundance and density estimates. Here we focus on density surface models (DSMs; Hedley & Buckland, 2004; Miller et al., 2013), a model-based approach to obtain spatially-explicit abundance and density estimates. Spatially-explicit models allow managers and scientists to ask more fine-grained questions of their data. For example, marine species applications include spatial abundance estimation (Becker et al., 2016; Becker et al., 2017; Forney et al., 2015), habitat preference (Cañadas & Hammond, 2008; Torres, Read & Halpin, 2008; Hazen et al., 2017), spatial prioritization (Winiarski et al., 2014), risk assessment (Gilles et al., 2016; Redfern et
and as a tool to assess potential impacts on cetaceans as required by government regulations (Roberts et al., 2016; Mannocci et al., 2017b). Ensuring that the major sources of uncertainty are included in final estimates allows managers and policy makers to make the best possible decisions by allowing them to evaluate the degree to which they can believe their results or the impact of proposed management actions. Failing to correctly account for uncertainty may lead to management strategies that have dire consequences, especially for endangered species.

Our interest in uncertainty quantification comes from the legal requirements surrounding the US Marine Mammal Protection Act of 1972 (MMPA), which bans the intentional “take” (disturbance or harm) of marine mammals but permits activities that may incidentally take them, provided that the number of takes is estimated with suitable methods and found to be sufficiently small. Uncertainty is built-in to the calculation of this limit (potential biological removal, PBR; e.g., Taylor et al., 2000) as it uses the 20% quantile of the distribution of abundance to give a minimum abundance estimate.

Every seven years the US Navy must apply for a “Letter of Authorization” to conduct peacetime testing and training activities that may take cetaceans, e.g., during the use of tactical SONAR. The US Navy uses simulations to assess the impact of sound from SONAR and other sources on cetaceans, and quantifies the take of each affected stock (US Department of the Navy, 2017). A primary input to these simulations are spatially explicit models of cetacean density (Roberts et al., 2016; Becker et al., 2016). In this and similar processes, it is critical that the major sources of uncertainty are clearly accounted for and propagated through the different phases of impact modelling.

In this article we propose an approach to uncertainty characterization and estimation that will help practitioners in two ways: (i) we give a checklist of the major sources of uncertainty, so possible pitfalls can be considered before a survey is conducted and appropriate field methods can be adapted in advance; (ii) we provide a framework to estimate the combined uncertainty from various sources, once data are collected.

In this article we adopt a modular, simulation-based approach to uncertainty estimation for DSMs. Rather than deriving a single, complex analytical expression, we use posterior simulation (sometimes referred to as “parametric bootstrapping”) where possible. Our approach uses the (posterior) distribution of the model parameters and samples from that distribution, from which we can obtain corresponding predictions. The variation in these predictions represents the uncertainty in our model. Simulation-based approaches can be easier to understand, less technically and notationally demanding, and more easily parallelizable. Our approach includes uncertainty from each model component while estimating covariance between components where possible. It is conceptually and computationally tractable, while avoiding potential pitfalls that occur with other approaches like the non-parametric bootstrap (see ‘Discussion’).

The article is structured as follows: ‘Density Surface Models’ gives a brief overview of the density surface modelling framework. ‘Characterising Components of Uncertainty’ then lists the various sources of uncertainty that may be present in these models and gives a summary of current methods to estimate uncertainty for each. ‘Combining Uncertainty from Multiple Model Components’ presents our framework for integrating uncertainty
and we apply this to line transect survey data of fin whales (*Balaenoptera physalus*) in the California Current Ecosystem in ‘Example: Fin Whales in the California Current’. Extensions and future work are outlined in ‘Discussion’.

## DENSITY SURFACE MODELS

DSMs are multi-stage species distribution models (SDMs), where biases in the observation process(es) are first addressed, and then a spatial model is fitted to the resulting corrected data. An overview of DSM methodology is given in *Miller et al. (2013)* (see also *Hedley & Buckland, 2004*). A typical DSM might take into account changes in detectability resulting from the distance between sampler and observation using distance sampling methods (*Buckland et al., 2001*). A detection function is fitted to the distances between the sampler and collected observations, and from that detection function a probability of detection (unconditional on distance), \( \hat{p} \), is estimated. This probability is then used as an offset (along with the effort expended) in a generalized additive model (*Wood, 2017*), which might have the following mathematical form:

\[
E(n_j) = A_j \hat{p}_j \exp\left[ \beta_0 + f_{xy}(x_j, y_j) + f_{SST}(SST_j) \right],
\]

where \( j \) indexes sample units. In the case of point transects, \( j \) indexes the points; for line transects, the longer transects are cut into segments and then indexed by \( j \) (without loss of generality we refer to sample units as segments henceforth). \( A_j \) is the area of segment \( j \), \( \hat{p}_j \) is the estimated probability of detection in that segment. \( n_j \) is the number of observations (individuals or groups) in segment \( j \). We assume that \( n_j \) is distributed according to some count distribution (e.g., Tweedie or negative binomial) and \( \beta_0 \) is the intercept. In this example \( f_{xy} \) is a smooth function of location and \( f_{SST} \) is a smooth of sea surface temperature; more generally, any number of smooth terms can be added (denoted \( f \) with subscript for the covariate).

We can adapt (Eq. (1)) to include information about other observation processes. These could include additional data collected either during the survey or during some other period. For example, we may want to include whether an animal is available to be detected (a problem for animals which dive under the water such as seabirds, cetaceans or pinnipeds). Availability can be addressed as another offset multiplier in Eq. (1), \( \hat{u}_j \). We may also want to relax the distance sampling assumption that animals on the trackline are detected with certainty, e.g., by applying mark-recapture distance sampling (*Burt et al., 2014*), in which we use data from multiple observers to estimate \( g(0) \), the probability of detecting an animal on the trackline in segment \( j \). Again this quantity can be included in the offset of Eq. (1). We can extend Eq. (1) to include these additional estimates as follows:

\[
E(n_j) = A_j \hat{p}_j \hat{g}(0) \hat{u}_j \exp\left[ \beta_0 + f_{xy}(x_j, y_j) + f_{SST}(SST_j) \right],
\]

DSMs are commonly developed using multiple years of survey data, both to increase sample size and capture a wider range of environmental conditions, and then model predictions are made on finer temporal scales to evaluate seasonal and inter-annual
differences in abundance and distribution (Becker et al., 2012; Forney et al., 2012; Redfern et al., 2019). For this reason, accounting for these biases is important, especially in the case where we are potentially combining data from multiple sources (e.g., Miller et al., 2021), as we need to ensure that any spatial heterogeneity that the GAM models is due to spatial effects, not down to unaddressed biases in the data. Model predictions can be made on days, weeks, or months, depending on the scale of the ecological question and the variability of the study ecosystem (Mannocci et al., 2017a).

CHARACTERISING COMPONENTS OF UNCERTAINTY

Our aim is to think about uncertainty estimation in a modular way. We first give a taxonomy and our general strategies for including uncertainty from model components into our estimates, before moving on to review available methods for uncertainty estimation for the components we have seen so far.

We are generally concerned with two types of components: ones which have covariance with the spatial model and those which do not; we refer to these as coincident and non-coincident, respectively. Non-coincident components can use the delta method (Seber, 1987) and add their squared coefficient of variation to that of the model. This is easily justified if the estimate is from a different place and time (e.g., an estimate of availability from the literature for this species). The non-coincident case trivially includes the case where there are no covariates in that model component. If estimates are coincident in space and time, our general strategy here is to absorb this into the spatial model if possible, using the method of Bravington, Miller & Hedley (2021). A typical example is that detection functions will usually be coincident as they are estimated from data collected during the survey, in that case it is likely that weather conditions that affect detectability (e.g., sea state) vary in space, so there will be covariance between the detectability and spatial model that must be accounted for.

Methods for uncertainty estimation are more developed for some model components than others. We begin by looking at those which are fairly mature in some depth before moving onto areas that are still in development or have not been applied in the DSM context yet.

GAM (smooths, smoothing parameters, model structure)

Smooths in the spatial model (such as \( f_{xy} \) and \( f_{SST} \)) are estimated and so have associated model-based uncertainties from standard GAM theory, based on their basis function coefficients (\( \hat{\beta} \)). We can also incorporate uncertainty in the smoothing parameter(s), \( \lambda \), which dictate how wiggly the smooths should be.

GAMs fitted using the popular R package mgcv using restricted maximum likelihood (REML) are empirical Bayes estimates (Wood, 2017, Section 6.2.6), so we have an approximate posterior distribution \( \beta | \lambda \sim N(\hat{\beta}, V_{\hat{\beta}}) \) (where \( \hat{\beta} \) are the estimated GAM coefficients and \( V_{\hat{\beta}} \) is their corresponding covariance matrix). Taking samples from this distribution we can make predictions of abundance, then make appropriate summaries over these predictions to obtain estimates of the variance, intervals, or other desired uncertainty statistics for the spatiotemporal extents that are useful for species management.
In order to move between the statements about parameters to statements about predictions (and then total abundance estimates) we form a matrix, \( \tilde{X} \), that maps the model parameters to the linear predictor, \( \eta \). We can then multiply \( \tilde{X} \) by a sample from \( N(\hat{\beta}, V_{\hat{\beta}}) \) to obtain predictions on the linear predictor scale. Applying the link function gives us predictions on the response scale (\( y = g^{-1}(\eta) = g^{-1}(\tilde{X}\hat{\beta}) \)). A simulation-based approach also takes into account the situation where our summary is a non-linear function of the linear predictor (e.g., the log link function) \( (Wood, 2017, \text{Section 7.2.6}) \).

For example, to depict uncertainty geographically, we can perform a number of simulations (\( B \)) for the area of interest and summarize predictions on a per-grid-cell basis using the following algorithm:

1. For \( b = 1, \ldots, B \):
   (a) Simulate from \( N(\hat{\beta}, V_{\hat{\beta}}) \), to obtain \( \hat{\beta}_b \).
   (b) Calculate predicted abundance for each prediction grid cell for this \( \hat{\beta}_b \), \( \hat{N}^*_b = g^{-1}(\tilde{X}\hat{\beta}_b) \).
   (c) Store \( \hat{N}^*_b \), a vector of abundances, with one element per cell.
2. For each grid cell, calculate the empirical variance, percentiles, or statistic of interest for the \( \hat{N}^*_b \)s.

In practice \( B \) does not have to be particularly large. \( Marra, Miller \& Zanin (2012) \) achieved reasonable results with \( B = 100 \), though this is dependent on the summaries required. We note that here we talk about simply sampling from the multivariate normal distribution but in practice the approximation may not hold. In this case either importance sampling or a random-walk Metropolis–Hastings sampler can be used (the latter is implemented as \texttt{gam.mh} in the \texttt{mgcv} package).

Two further sources of model uncertainty from the GAM can be included in \( V_{\hat{\beta}} \) during model fitting and leave the above procedure unchanged. (i) Smoothing parameter uncertainty: estimated uncertainty in how wiggly terms should be, using the approach outlined in \( Wood, Pya \& Säfken (2016) \), we then denote the covariance matrix as \( V_{\hat{\beta}_{\hat{\lambda}}} \). (ii) Model structural uncertainty: using shrinkage smoothers (either thin plate regression splines via the \texttt{ts} basis or cubic regression splines via the \texttt{cs} basis, in \texttt{mgcv}) that shrink model terms to zero (or near zero) effect size if necessary \( (Marra \& Wood, 2011) \) as opposed to term selection via \( p \)-values or AIC. As terms are retained (but their effect sizes are shrunk), resulting uncertainty includes uncertainty about the model structure (conditional on the terms that were included).

**Variability in environmental covariates**

In dynamic environments that exhibit high environmental variability, a major contributor to uncertainty in estimates of abundance is the associated variability in population density due to movement of animals within, into, or out of the study area (e.g., \( Forney, 2000; Becker et al., 2014; Becker et al., 2019 \)). To account for the potentially large changes that can occur within short periods, model predictions need to be made over temporally-relevant time periods, which may change depending on ecosystem dynamics \( (Mannocci et al., 2017a) \). For example, in a highly dynamic ecosystem such as the California Current \( (Bograd et al., 2009) \) environmental covariates (and hence model predictions) vary dramatically over time.
scales as short as several days during coastal upwelling events. Making predictions over multiple time periods allows the model to include different oceanic states and account for the variability in environmental covariates. One can think of this as being analogous to the procedure above but generating predictions from the realized (observed) distribution of possible environmental covariates, rather than the model parameters.

In ecosystems with limited seasonal or inter-annual variability or for models over larger areas and longer time periods, this environmental variability may be small, as illustrated for the Central North Pacific (Forney et al., 2015). Thus, both ecosystem factors and modelling scales will play a role in determining the relative importance of variability in environmental covariates.

**Detectability**

If we assume that detectability only varies at the segment level (e.g., with covariates representing weather conditions, e.g., Beaufort wind force scale), and not at the level of an individual observation, then we can use the variance propagation method of Bravington, Miller & Hedley (2021) to include the uncertainty about the detection function in $\hat{V}_β$ (the GAM covariance matrix). Briefly, the method consists of refitting the GAM including a random effect that has the covariance from the detection function model ($\hat{V}_θ$). We then obtain a modified covariance matrix, incorporating uncertainty in both detection function and GAM ($\hat{V}_{θθ}$ or $\hat{V}_{θλ}$ if smoothing parameter uncertainty is also included). If detectability varies at the level of the observations (e.g., with group size or behavioural state), we may be able to apply the factor-smooth model of Bravington, Miller & Hedley (2021), categorising the detection covariate and then applying the variance propagation procedure. Finally, if detectability is constant (i.e., $p_j = \hat{p}$), we can simply apply the delta method. We do not recommend using the delta method in any other case, as then covariance between detectability and spatial model are ignored.

**Trackline detectability**

Relaxing the assumption that all objects on the trackline are observed (“$g(0) = 1$”) can be handled in a number of ways (Hiby, 1999; Barlow, 1995; Burt et al., 2014; Barlow, 2015). Most rely on having an additional observer or observers who are independent (or conditionally independent) of the main observer team. The proportion of observations missed by the main team can then be used to correct $g(0)$. For the mark-recapture distance sampling method where $g(0)$ is another component of the detection function model, the variance propagation approach can be used to include this uncertainty. Uncertainty around independent estimates of $g(0)$ can be included via the delta method. Other methods for trackline detectability would need to be adapted appropriately.

**Availability**

If estimates of availability are constant in space/time (either calculated from observational studies or tags) then corresponding uncertainty can be included via the delta method (Ver Hoef et al., 2014). More complex models such as Borchers et al. (2013) and Borchers & Langrock (2015), which account for $g(0)$ and availability simultaneously using auxiliary
data from tags or a modified survey protocol, could be included in the GAM with some modification of the procedures in Bravington, Miller & Hedley (2021).

**Other sources of uncertainty**

Some sources of uncertainty do not have methods that can currently be applied in a spatial context, or have not yet been applied in the DSM literature. We discuss these briefly for completeness but do not address them fully here.

To account for measurement error in the environmental covariates, the procedure proposed by Stoklosa et al. (2015) to deal with both classical and Berkson-type measurement errors could be used. These require adaptation in how the spatial model (GAM in our case) is fitted.

When using a DSM, we have substituted the usual group size and encounter rate components that occur in distance sampling for the spatial model and as such these are handled as part of the GAM above. Group size uncertainty that arises from measurement error in observers’ counts needs to be addressed at the spatial model stage as it is likely that corrections may vary according to group size and sighting conditions, which vary spatially. Such errors will also likely effect estimates of response distribution hyper-parameters (such as overdispersion). Constant correction factors for group size uncertainty could be estimated depending on species and environment (Hodgson, Peel & Kelly, 2017), but this is currently an active area of research.

There are several methods to investigate species identification uncertainty. Methods that pro-rate species identity from additional data (e.g., Johnston et al., 2015) may not always be an option (though if they are, uncertainty can be included). Joint modelling or approaches where true species identity is a latent variable (Conn et al., 2013) might be one way to allow for this in the spatial model.

**COMBINING UNCERTAINTY FROM MULTIPLE MODEL COMPONENTS**

We now focus on combining the above methods to create a single procedure for uncertainty estimation from complex DSMs (e.g., Eq. (2)). This modular workflow allows us to exclude any of the steps in the case where the model does not contain that uncertainty component. Our procedure is as follows (shown diagrammatically in Fig. 1):

1. Fit the DSM, with detectability, trackline detectability and/or availability included in the offset.
2. If any offset corrections are coincident, incorporate their uncertainty via the variance propagation method of Bravington, Miller & Hedley (2021).
3. Extract posterior estimates of the GAM parameters ($\hat{\beta}$) and related covariance matrix ($V_{\hat{\beta}}$, $V_{\hat{\beta},\hat{\lambda}}$, $V_{\hat{\beta}\hat{\theta}}$ or $V_{\hat{\beta}\hat{\theta}\hat{\lambda}}$; see ‘GAM (Smooths, Smoothing Parameters, Model Structure)’ and ‘Detectability’ for definitions).
4. Simulate $B$ samples from $\beta_b \sim N(\hat{\beta}, V^*)$ where $V^*$ is one of $V_{\hat{\beta}}$, $V_{\hat{\beta},\hat{\lambda}}$, $V_{\hat{\beta}\hat{\theta}}$ or $V_{\hat{\beta}\hat{\theta}\hat{\lambda}}$.
5. For each time period that needs to be predicted ($t = 1, \ldots, T$):
   (a) Form the prediction matrix $\tilde{X}_t$ for this time period’s prediction grid.
   (b) For $b = 1, \ldots, B$ posterior samples generated above:
Include as many sources of uncertainty as possible in GAM covariance matrix.

Generate $n_b$ from $N(\hat{\beta}, \hat{V}_{\beta\lambda\theta})$

Multiple rasters of environmental covariates

Day n
Day 2
Day 1
…

Summaries

Figure 1  Flow diagram showing our process for capturing uncertainty from multiple model components. In this case we have a GAM model of spatial counts, a detection function and environmental variability to include in the model. Once detection function uncertainty is included in the GAM, some number ($n_b$) posterior samples of model parameters ($\hat{\beta}_b$) can be generated using the GAM-plus-other-components mean ($\hat{\beta}$) and covariance matrix ($\hat{V}_{\beta\lambda\theta}$ here). These samples are used to generate potential density surfaces exploring both model parameter and environmental space. Finally, summaries can be calculated from these predictions.

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1. Calculate predicted abundance for each prediction grid cell: $\hat{N}_{b,t}^* = g^{-1}(\tilde{X}_t \hat{\beta}_b)$.
2. Store $\hat{N}_{b,t}^*$ for this iteration–time-period combination.
3. Summarize the per iteration–time-period predictions ($\hat{N}_{b,t}^*$) by computing the appropriate summary statistic (typically mean or median) and the empirical standard error of the estimates.
4. Include uncertainty from non-coincident estimates \textit{via} the delta method.

At point 5 above, we may wish to make spatial summaries: per-time-period abundances at each prediction location or average abundance across all time periods for each prediction location. We could also compute a time series of abundance. We show examples of these in the next section. We also note that the exact form of the variance estimators depends on the summary taken. Appendix S1 gives details of some common situations and their estimators.
The above algorithm can be implemented in any statistical programming language, and we include example code for the analysis in the next section with this article R. For step one we rely on functions in the R packages `distance` and `mrds` for detection function fitting and `dsm` for fitting the DSM (examples of how to use these packages for model fitting are provided at http://examples.distancesampling.org). Step two can be accomplished using the function `dsm_varprop` in `dsm`. Step three relies on functions from the `mgcv` package, `coef` and `vcov` can be used to extract the parameter estimates and covariance matrix, respectively. To sample from the posterior distribution of the parameters we can use the `rmvn` function from `mgcv`. The remaining steps can be coded in base R and we recommend interested readers consult the code provided for examples.

**EXAMPLE: FIN WHALES IN THE CALIFORNIA CURRENT**

To illustrate our approach, we use the example of fin whales in the California Current Ecosystem (CCE). Data were collected from line transect surveys conducted by NOAA’s Southwest Fisheries Science Center (SWFSC), July through early December of 1996, 2001, 2005, 2008, and 2014. Each of these surveys covered a broad area off the entire United States West Coast and, when combined, provided dense coverage of waters from the coast to approximately 556 km offshore (Fig. 2, left panel). Standardized line transect protocols were followed during all years using a team of three experienced observers stationed on the flying bridge of the ship (Barlow & Forney, 2007; Kinzey, Olson & Gerrodette, 2000). Our aims for this analysis are to estimate uncertainty around: (i) estimates of monthly abundance, (ii) a density map of fin whales in the study area, averaged over the whole time period, (iii) density maps of fin whales in the study area averaged within each year.

The analysis presented here is designed to be as close to SWFSC’s modelling process as possible, to give an idea of how our framework can be adapted in practical situations. We refer readers to the technical memos and papers below for detailed information on this process. Our model consisted of the following components.

1. Detection function from Barlow, Ballance & Forney (2011). A half-normal detection function was fitted using the R package `distance` (Miller et al., 2019) to all observations of large whales (Bryde’s, sei, blue, fin, humpback, and unidentified large whales), with covariates for ship (6 level factor), species (10 level factor), visibility, log average total school size for fin whales over all sightings and Beaufort (all continuous). Distances were truncated at 5.5 km. Fig. S1 shows the detection function.

2. Fixed estimate of \( g(0) \) from (Barlow & Forney, 2007, Table 3): \( \hat{g}(0) = 0.921 \), \( CV = 0.023 \). This estimated was based on the method developed by Barlow (1995) using conditionally independent observer data.

3. Dynamic environmental covariates taken from a 10 km resolution data-assimilative implementation of the Regional Ocean Modeling System (ROMS) in the CCE, which was produced by the University of California Santa Cruz Ocean Modeling and Data Assimilation group (Moore et al., 2011). The covariates sea surface temperature, sea surface height, mixed layer depth, and the standard deviation of each covariate within a \( 3 \times 3 \) cell box surrounding each point were extracted at three-day intervals for the 5
Figure 2  **Raw data and results from applying our procedure to the fin whale data.** Top row, left: locations of segment centroids (black dots) for the fin whale survey with observation locations (orange dots) and underlying bathymetry. Top row, middle: predicted density for fin whales in the California Current Ecosystem. Top row, right: estimated standard error for the predictions, using the procedure we outline. Black dots give locations of observations of fin whales in the centre and right plots. Bottom row: confidence surfaces derived from a log-normal approximation; left is lower 2.5%, right is upper 97.5%.

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years of sampled data. Bathymetric data (depth and standard deviation of depth) were
derived from the ETOPO1 1-arc-min global relief model (Amante & Eakins, 2009).

4. A generalized additive model to describe the spatio-temporal variation in fin whale
density as a function of a subset of the above environmental covariates. We used the
same model structure and covariate selection procedure as Becker et al. (2016), yielding
the final model:

\[ E(n_j) = A_j \tilde{g}(0) \exp\{\beta_0 + f_{sy}(x_j, y_j) + f_{SST}(SST_j) + f_{SSTSD}(SSTSD_j) + f_{SSH}(SSH_j)
+ f_{MLD}(MLD_j) + f_{year}(year_j)\}, \] (3)

where SSTSD is the standard deviation of sea surface temperature, SSH is sea surface
height, MLD is mixed layer depth and year is the year in which segment was surveyed.
All smooths used a thin plate spline basis with shrinkage (Marra & Wood, 2011). \( f_{xy} \)
was constructed using a tensor product of smooths of longitude and latitude. Model
term plots are shown in Fig. S3.

Predictions were made over grids of 11,860 cells covering the CCE, with a grid size of
0.09 degrees (≈ 10 km square). There were 821 grids giving daily values of covariates, 26
June through 6 December, for each of the 5 years, although occasionally the daily grids
had missing grid cells because the covariates were not available from the ROMS model
outputs. Based on the algorithm given in the previous section, we took the following steps
to estimate uncertainty for Eq. (3).

1. Propagate detectability uncertainty into Eq. (3) via Bravington, Miller & Hedley (2021).
2. Extract \( \hat{\beta} \) and \( V_{\hat{\beta}} \).
3. Simulate 1000 samples from \( N(\hat{\beta}, V_{\hat{\beta}}) \), to obtain \( \beta_b \) for \( b = 1, \ldots, 1,000 \).
4. For each time period that needs to be predicted (\( t = 1, \ldots, 821 \)):
   a. Form the prediction matrix \( \tilde{X}_t \) for this time period, \( t \).
   b. For \( b = 1, \ldots, 1,000 \) posterior samples:
      i. Predict abundance for each prediction grid cell: \( \tilde{N}_{b, t} = g^{-1}(\tilde{X}_t \beta_b) \).
      ii. Store \( \hat{\tilde{N}}_{b, t} \).
5. Calculate per-prediction-cell means and variances over \( \hat{\tilde{N}}_{b, t} \) for all years and for each
   month.
6. Calculate monthly means and for \( \hat{\tilde{N}}_{b, t} \) to obtain abundance time series.
7. Include \( g(0) \) uncertainty via the delta method.

This procedure encapsulates spatial model uncertainty, detection function uncertainty,
trackline detection uncertainty and spatial covariate variability.

Resulting maps of overall predictions and corresponding standard errors are shown in
Fig. 2. Yearly maps (Fig. S2) and monthly time series plots of abundance (Fig. 3)
showed increasing abundance between years (in agreement with e.g., Moore & Barlow,
2011). Uncertainty appeared to be highest in 2008 and on June 26 in particular; further
investigation showed that this was due to large mixed layer depth values (Fig. S4). On June
26, 2008 the mixed layer depth reached 137 m, whereas in the data used to build the model
the maximum was about 100 m, at which point the smooth showed increasing behaviour
(Fig. S3, bottom row left panel). There are a number of options for modellers when this
kind of extrapolation occurs, including excluding the cell from predictions, winsorizing or leaving as-is; Bouchet et al. (2020) provide an overview and practical tools for DSMs.

The data used to fit models here had some overlap with those used in Becker et al. (2016), but for a more direct comparison Figs. S5 and S6 show plots comparable to Fig. 2 and Fig. S2 but where only environmental uncertainty was included in the variance calculation procedure (as was the case in Becker et al., 2016). We note that when only environmental variability is accounted for, plots seem to show much less detail in spatial uncertainty patterns.

Similar analyses of these data were provided by Moore & Barlow (2011) and Nadeem et al. (2016). Both provided a state-space model for abundance and the latter used a spatial (only over four large areas) Ricker model to obtain abundance estimates. Neither provided spatially-explicit estimates; only stratified abundance was calculated. Detection probability and trackline detection (using estimates from Barlow & Forney, 2007), along with their associated uncertainties were included in estimates of abundance. For comparison (Fig. 4), we calculated summary abundance estimates at the yearly level, along with 95% intervals (based on a log-normal approximation).

**DISCUSSION**

Maps of point estimates (e.g., mean density over a time period) are often given prominence in articles and reports but without some measure of uncertainty they are not useful for conservation (Jansen et al., 2022). In many cases, only uncertainty from the spatial model
is reported in studies, perhaps because it could be obtained as a convenient output directly from that modelling stage, and modellers lacked ready methods for considering other sources of uncertainty. The framework we provide here allows modellers to estimate and evaluate these sources of uncertainty in their DSMs. These approaches can be adapted to other spatial modelling methodologies.

We see the delta method as the last resort when we are not able to include that uncertainty via other methods. We do not however want to discourage people from using this method if that is all that is available to them. A significant advantage of incorporating covariances via Bravington, Miller & Hedley (2021) where possible over using the delta method is that uncertainty estimates can go down as well as up (e.g., when there is negative covariance between the detection function and spatial model).

A practical benefit to using the simulation-based approach we outline here is the computation is easily parallelizable; speeding it up is simply a matter of splitting the simulation runs across a larger number of cores. Storage and retrieval of interim results may prove to be more of an issue; in our example we switched to using serialized objects (via R’s `saverDS` functionality) over text-based (comma separated value) files to improve the speed of final calculations and reduce disk space, though storing in this format still used about 68GB of disk space. More efficient binary storage options could be investigated for larger prediction grids. Iteration over time period is conceptual and was done for a practical computing consideration; we could have created a prediction matrix of all time periods at once (X rather than X_t), but this may not fit in memory so we formed one matrix for each time period. We applied the online variance calculation method of Welford.
This approach could be included in the algorithms above, avoiding storage of \( \hat{N}_b \). Appendix S1 also provides equivalent analytic expressions for the variances in some useful situations. We find the simulation framework easier to understand and reason about, so that is the main focus here.

A popular reply to the question of how to calculate uncertainty for complex models is “do a bootstrap”. What is usually meant by this is that one should resample the data with replacement, refitting models each time and calculating some summary statistic (a non-parametric bootstrap). This seems appealing but it has been shown that the use of so-called “naïve” bootstraps leads to underestimation of uncertainty. The issue stems from the fact that GAM terms are structured random effects, which have priors on them. When data are resampled, the structure (prior) is ignored so uncertainty is collapsed leaving only the sampling variation in the bootstrap resamples. Bravington, Miller & Hedley (2021) show a simple simulated example of this happening. A secondary issue is that of coming up with a resampling scheme for spatial uneven data. A sufficiently complicated bootstrap might be constructed to achieve our aims here, but we have instead focussed on an easily-implementable, modular approach.

We have proposed a framework for uncertainty estimation to allow for the inclusion of relevant sources of uncertainty in final estimates. We do not believe that all of these sources should be included in all models, at least in part because the corresponding mean model components may not be necessary (e.g., availability may not be an issue for large terrestrial mammals). There are situations in which we think the environmental variability may not be necessary (models with non-dynamic covariates) or may be less of an issue (less dynamic system), so this component can be easily omitted (or included and tested). We also note that there are multiple approaches possible for each source of uncertainty and for many there is no best practice at the moment, so our intention is not to be prescriptive. Our hope is that this framework will prompt more discussion of the issue of how to estimate these quantities, since we can now include their uncertainty final estimates.

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**ADDITIONAL INFORMATION AND DECLARATIONS**

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**Competing Interests**
Elizabeth A. Becker is employed by Ocean Associates Inc.

**Author Contributions**
- David L. Miller conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the article, and approved the final draft.
- Elizabeth A. Becker conceived and designed the experiments, performed the experiments, analyzed the data, authored or reviewed drafts of the article, and approved the final draft.
- Karin A. Forney conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Jason J. Roberts conceived and designed the experiments, performed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Ana Cañadas conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.
- Robert S. Schick conceived and designed the experiments, authored or reviewed drafts of the article, and approved the final draft.

**Data Availability**
The following information was supplied regarding data availability:

The code and small data files are available at Github: https://github.com/densitymodelling/lemur_fin_analysis/.

The processed ROMS grids are available at figshare: Miller, David; Becker, Elizabeth (2022): ROMS grids for predictions. figshare. Dataset. https://doi.org/10.6084/m9.figshare.20132291.v1.

**Supplemental Information**
Supplemental information for this article can be found online at http://dx.doi.org/10.7717/peerj.13950#supplemental-information.

**REFERENCES**
- Amante C, Eakins BE. 2009. ETOPO1 1 arc-minute global relief model: procedures, data sources and analysis. Technical Report NESDIS NGDC-24. National Geophysical Data Center, Boulder, CO.
- Barlow J. 1995. The abundance of cetaceans in California waters. Part I: ship surveys in summer and fall of 1991. *Fisheries Bulletin* 93:1–14.
- Barlow J. 2015. Inferring trackline detection probabilities, g(0), for cetaceans from apparent densities in different survey conditions. *Marine Mammal Science* 31(3):923–943 DOI 10.1111/mms.12205.
- Barlow J, Ballance LT, Forney K. 2011. Effective strip widths for ship-based line-transect surveys of cetaceans. Technical Report NOAA-TM-NMFS-SWFSC-484.
Barlow J, Forney KA. 2007. Abundance and population density of cetaceans in the California Current ecosystem. Fishery Bulletin 105(4):509–526.

Becker EA, Forney KA, Ferguson MC, Barlow J, Redfern JV. 2012. Predictive modeling of cetacean densities in the california current ecosystem based on summer/fall ship surveys in 1991–2008. Technical Report NMFS-SWFSC-499. La Jolla, CA.

Becker E, Forney K, Fiedler P, Barlow J, Chivers S, Edwards C, Moore A, Redfern J. 2016. Moving towards dynamic ocean management: how well do modeled ocean products predict species distributions? Remote Sensing 8(2):149 DOI 10.3390/rs8020149.

Becker E, Forney K, Foley D, Smith R, Moore T, Barlow J. 2014. Predicting seasonal density patterns of California cetaceans based on habitat models. Endangered Species Research 23(1):1–22 DOI 10.3354/esr00548.

Becker E, Forney K, Redfern JV, Barlow J, Jacox MG, Roberts JJ, Palacios DM. 2019. Predicting cetacean abundance and distribution in a changing climate. Diversity and Distributions 25(4):626–643 DOI 10.1111/ddi.12867.

Becker E, Forney K, Thayre BJ, Debich AJ, Campbell GS, Whitaker K, Douglas AB, Gilles A, Hoopes R, Hildebrand JA. 2017. Habitat-based density models for three cetacean species off southern California illustrate pronounced seasonal differences. Frontiers in Marine Science 4:121 DOI 10.3389/fmars.2017.00121.

Bograd SJ, Schroeder I, Sarkar N, Qiu X, Sydeman WJ, Schwing FB. 2009. Phenology of coastal upwelling in the California Current. Geophysical Research Letters 36(1):101602.

Borchers DL, Langrock R. 2015. Double-observer line transect surveys with Markov-modulated Poisson process models for animal availability: availability modeling on double-observer surveys. Biometrics 71(4):1060–1069 DOI 10.1111/biom.12341.

Borchers DL, Zucchini W, Heide-Jørgensen MP, Cañadas A, Langrock R. 2013. Using hidden markov models to deal with availability bias on line transect surveys: line transect availability bias. Biometrics 69(3):703–713 DOI 10.1111/biom.12049.

Bouchet PJ, Miller DL, Roberts JJ, Mannocci L, Harris CM, Thomas L. 2020. dsmextra: extrapolation assessment tools for density surface models. Methods in Ecology and Evolution 11(11):1464–1469 DOI 10.1111/2041-210X.13469.

Bravington MV, Miller DL, Hedley SL. 2021. Variance propagation for density surface models. Journal of Agricultural, Biological and Environmental Statistics 26:306–323 DOI 10.1007/s13253-021-00438-2.

Buckland S, Anderson D, Burnham K, Laake J, Borchers D, Thomas L. 2001. Introduction to distance sampling: estimating abundance of biological populations. Oxford, UK: Oxford University Press.

Burt ML, Borchers DL, Jenkins KJ, Marques TA. 2014. Using mark-recapture distance sampling methods on line transect surveys. Methods in Ecology and Evolution 5(11):1180–1191 DOI 10.1111/2041-210X.12294.
Cañadas A, Hammond P. 2008. Abundance and habitat preferences of the short-beaked common dolphin Delphinus delphis in the southwestern Mediterranean: implications for conservation. *Endangered Species Research* 4:309–331 DOI 10.3354/esr00073.

Conn PB, McClintock BT, Cameron MF, Johnson DS, Moreland EE, Boveng PL. 2013. Accommodating species identification errors in transect surveys. *Ecology* 94(11):2607–2618 DOI 10.1890/12-2124.1.

Forney KA. 2000. Environmental models of cetacean abundance: reducing uncertainty in population trends. *Conservation Biology* 14(5):1271–1286 DOI 10.1046/j.1523-1739.2000.99412.x.

Forney K, Becker E, Foley D, Barlow J, Oleson E. 2015. Habitat-based models of cetacean density and distribution in the central North Pacific. *Endangered Species Research* 27(1):1–20 DOI 10.3354/esr00632.

Forney K, Ferguson M, Becker E, Fiedler P, Redfern J, Barlow J, Vilchis I, Ballance L. 2012. Habitat-based spatial models of cetacean density in the eastern Pacific Ocean. *Endangered Species Research* 16(2):113–133 DOI 10.3354/esr00393.

Gilles A, Viquerat S, Becker EA, Forney KA, Geelhoed SCV, Haelters J, Nabe-Nielsen J, Scheidat M, Siebert U, Sveegaard S, Beest FM, Bemmelen R, Aarts G. 2016. Seasonal habitat-based density models for a marine top predator, the harbor porpoise, in a dynamic environment. *Ecosphere* 7(6):e01367.

Hazen EL, Palacios DM, Forney KA, Howell EA, Becker E, Hoover AL, Irvine L, DeAngelis M, Bograd SJ, Mate BR, Bailey H. 2017. WhaleWatch: a dynamic management tool for predicting blue whale density in the California Current. *Journal of Applied Ecology* 54(5):1415–1428 DOI 10.1111/1365-2664.12820.

Hedley SL, Buckland ST. 2004. Spatial models for line transect sampling. *Journal of Agricultural, Biological, and Environmental Statistics* 9(2):181–199 DOI 10.1198/1085711043578.

Hiby L. 1999. The objective identification of duplicate sightings in aerial survey for porpoise. In: Garner GW, Amstrup SC, Laake JL, Manly BF, McDonald LL, Robertson DG, eds. *Marine mammal survey and assessment methods*. Rotterdam: Balkema, 179–189.

Hodgson A, Peel D, Kelly N. 2017. Unmanned aerial vehicles for surveying marine fauna: assessing detection probability. *Ecological Applications* 27(4):1253–1267 DOI 10.1002/eap.1519.

Jansen J, Woolley SNC, Dunstan PK, Foster SD, Hill NA, Haward M, Johnson CR. 2022. Stop ignoring map uncertainty in biodiversity science and conservation policy. *Nature Ecology & Evolution* 6:828–829 DOI 10.1038/s41559-022-01778-z.

Johnston A, Thaxter CB, Austin GE, Cook AS, Humphreys EM, Still DA, Mackay A, Irvine R, Webb A, Burton NH. 2015. Modelling the abundance and distribution of marine birds accounting for uncertain species identification. *Journal of Applied Ecology* 52(1):150–160 DOI 10.1111/1365-2664.12364.
Kinze D, Olson P, Gerrodette T. 2000. Marine mammal data collection procedures on research ship line-transect surveys by the Southwest fisheries science center. Technical Report LJ-00-08. National Marine Fisheries Service, La Jolla, CA.

Mannocci L, Boustany AM, Roberts JJ, Palacios DM, Dunn DC, Halpin PN, Viehman S, Moxley J, Cleary J, Bailey H, Bograd SJ, Becker EA, Gardner B, Hartog JR, Hazen EL, Ferguson MC, Forney KA, Kinlan BP, Oliver MJ, Perretti CT, Ridoux V, Teo SLH, Winship AJ. 2017a. Temporal resolutions in species distribution models of highly mobile marine animals: recommendations for ecologists and managers. Diversity and Distributions 23(10):1098–1109 DOI 10.1111/ddi.12609.

Mannocci L, Roberts JJ, Miller DL, Halpin PN. 2017b. Extrapolating cetacean densities to quantitatively assess human impacts on populations in the high seas: cetacean densities in the high seas. Conservation Biology 31(3):601–614 DOI 10.1111/cobi.12856.

Marra G, Miller DL, Zanin L. 2012. Modelling the spatiotemporal distribution of the incidence of resident foreign population: spatiotemporal smoothing of resident Foreign population. Statistica Neerlandica 66(2):133–160 DOI 10.1111/j.1467-9574.2011.00500.x.

Marra G, Wood SN. 2011. Practical variable selection for generalized additive models. Computational Statistics & Data Analysis 55(7):2372–2387 DOI 10.1016/j.csda.2011.02.004.

Miller DL, Burt ML, Rexstad EA, Thomas L. 2013. Spatial models for distance sampling data: recent developments and future directions. Methods in Ecology and Evolution 4(11):1001–1010 DOI 10.1111/2041-210X.12105.

Miller DL, Fifield D, Wakefield E, Sigourney DB. 2021. Extending density surface models to include multiple and double-observer survey data. PeerJ 9:e12113 DOI 10.7717/peerj.12113.

Miller DL, Rexstad E, Thomas L, Marshall L, Laake JL. 2019. Distance sampling in R. Journal of Statistical Software 89(1):1–28.

Moore AM, Arango HG, Broquet G, Edwards C, Veneziani M, Powell B, Foley D, Doyle JD, Costa D, Robinson P. 2011. The Regional Ocean Modeling System (ROMS) 4-dimensional variational data assimilation systems. Progress in Oceanography 91(1):50–73 DOI 10.1016/j.pocean.2011.05.003.

Moore JE, Barlow J. 2011. Bayesian state-space model of fin whale abundance trends from a 1991–2008 time series of line-transect surveys in the California current: Bayesian trend analysis from line-transect data. Journal of Applied Ecology 48(5):1195–1205 DOI 10.1111/j.1365-2664.2011.02018.x.

Nadeem K, Moore JE, Zhang Y, Chipman H. 2016. Integrating population dynamics models and distance sampling data: a spatial hierarchical state-space approach. Ecology 97(7):1735–1745 DOI 10.1890/15-1406.1.

Redfern JV, Mckenna MF, Moore TJ, Calambokidis J, Deangelis ML, Becker EA, Barlow J, Forney KA, Fiedler PC, Chivers SJ. 2013. Assessing the risk of ships striking large whales in marine spatial planning: assessing ship-strike risk. Conservation Biology 27(2):292–302 DOI 10.1111/cobi.12029.
Redfern JV, Moore TJ, Becker EA, Calambokidis J, Hastings SP, Irvine LM, Mate BR, Palacios DM. 2019. Evaluating stakeholder-derived strategies to reduce the risk of ships striking whales. *Diversity and Distributions* 25(10):1575–1585 DOI 10.1111/ddi.12958.

Roberts JJ, Best BD, Mannucci L, Fujioka E, Halpin PN, Palka DL, Garrison LP, Mullin KD, Cole TVN, Khan CB, McLellan WA, Pabst DA, Lockhart GG. 2016. Habitat-based cetacean density models for the US Atlantic and Gulf of Mexico. *Scientific Reports* 6(1):22615 DOI 10.1038/srep22615.

Seber G. 1987. *Estimation of animal abundance*. Charles Griffin Book. Oxford, UK: Oxford University Press, Incorporated.

Stoklosa J, Daly C, Foster SD, Ashcroft MB, Warton DI. 2015. A climate of uncertainty: accounting for error in climate variables for species distribution models. *Methods in Ecology and Evolution* 6(4):412–423 DOI 10.1111/2041-210X.12217.

Taylor BL, Wade PR, De Master DP, Barlow J. 2000. Incorporating uncertainty into management models for marine mammals. *Conservation Biology* 14(5):1243–1252 DOI 10.1046/j.1523-1739.2000.99409.x.

Torres LG, Read AJ, Halpin P. 2008. Fine-scale habitat modeling of a top marine predator: do prey data improve predictive capacity. *Ecological Applications* 18(7):1702–1717 DOI 10.1890/07-1455.1.

US Department of the Navy. 2017. Quantifying acoustic impacts on marine mammals and sea turtles: methods and analytical approach for phase III training and testing. Technical report prepared by Space and Naval Warfare Systems Center Pacific, San Diego and Naval Undersea Warfare Center, Newport. Technical report. US Department of the Navy, Washington, D. C., USA.

Ver Hoef JM, Cameron MF, Boveng PL, London JM, Moreland EE. 2014. A spatial hierarchical model for abundance of three ice-associated seal species in the eastern Bering Sea. *Statistical Methodology* 17:46–66 DOI 10.1016/j.stamet.2013.03.001.

Welford BP. 1962. Note on a method for calculating corrected sums of squares and products. *Technometrics* 4(3):419–420 DOI 10.1080/00401706.1962.10490022.

Winiarski KJ, Miller DL, Paton PW, McWilliams SR. 2014. A spatial conservation prioritization approach for protecting marine birds given proposed offshore wind energy development. *Biological Conservation* 169:79–88 DOI 10.1016/j.biocon.2013.11.004.

Wood SN. 2017. Generalized additive models. An Introduction with R. In: *Texts in statistical science*. Second Edition. Boca Raton, FL, USA: CRC Press.

Wood SN, Pya N, Säfken B. 2016. Smoothing parameter and model selection for general smooth models. *Journal of the American Statistical Association* 111(516):1548–1563 DOI 10.1080/01621459.2016.1180986.