A comparative proteomics study of a synovial cell line stimulated with TNF-\(\alpha\)

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To elucidate the pathogenesis of rheumatoid arthritis (RA), we used proteomic analysis to determine the protein profile in a synovial cell line, MH7A, established from patients with RA. Proteins were extracted from MH7A cells that were or were not stimulated with tumor necrosis factor-\(\alpha\) (TNF-\(\alpha\)), and then analyzed on a liquid chromatography/mass spectrometry system equipped with a unique long monolithic silica capillary. On the basis of the results of this proteomic analysis, we identified 2650 proteins from untreated MH7A cells and 2688 proteins from MH7A cells stimulated with TNF-\(\alpha\). Next, we selected 269 differentially produced proteins that were detected only under TNF-\(\alpha\) stimulation, and classified these proteins by performing gene ontology analysis by using DAVID as a functional annotation tool. In TNF-\(\alpha\)-stimulated MH7A cells, we observed substantial production of plasminogen-activator inhibitor 2 and apoptosis-regulating proteins such as BH3-interacting domain death agonist, autophagy protein 5, apolipoprotein E, and caspase-3. These results indicate that the upregulation of plasminogen-activator inhibitor 2 and apoptosis-regulating proteins in synovial cells in response to TNF-\(\alpha\) stimulation might represent a predominant factor that contributes to the pathogenesis of RA.

Rheumatoid arthritis (RA) is characterized by a proliferation of synovial cells that leads to joint destruction [1]. The pathogenesis of RA comprises a preclinical stage involving the generation of autoantibodies, an initiation stage in which synovial inflammation emerges, and a clinical stage dominated by synovial inflammation and joint destruction [2]. In RA pathogenesis, a central role is played by cytokines derived from macrophages and fibroblast-like synoviocytes (FLS), such as tumor necrosis factor-\(\alpha\) (TNF-\(\alpha\)) and interleukin-6 (IL-6) [3]. RA FLS have been described as transformed cells and they share morphologic features with tumor cells, such as resistance to apoptosis, potentially due to somatic mutations in p53 [4], and RA FLS display and retain an invasive capacity against articular cartilage [5]. Although FLS are the major synovial producers of IL-6, a key pathogenic cytokine [6,7], macrophages are likely the major source of synovial TNF-\(\alpha\), a critical driver of the synovial inflammation that occurs in RA [4,7]. However, macrophages produce TNF-\(\alpha\) only transiently [8] and FLS are the major responders to TNF-\(\alpha\), which

Abbreviations
FLS, fibroblast-like synoviocytes; GO, gene ontology; IL-6, interleukin-6; LC, liquid chromatography; MS, mass spectrometry; PAI, plasminogen-activator inhibitor; RA, rheumatoid arthritis; TNF-\(\alpha\), tumor necrosis factor-\(\alpha\).
suggest that TNF-α-stimulated FLS are crucial for the pathogenesis of RA [9]. Therefore, we sought to identify the functional protein profile in FLS stimulated by TNF-α.

Proteome analysis is an indispensable technique for comprehensively analyzing differences in protein production. One of the critical purposes of clinical proteome analysis is the identification of biomarkers for disease diagnosis and therapeutic intervention: the biomarkers are identified by analyzing the proteome profiles of disease states and other physiologic states [10]. Recent high-throughput and sensitive mass spectroscopy (MS) based on proteome-analysis technology has provided a novel methodology for investigating the physiology and pathophysiology of numerous medicinal samples. In the case of protein separation coupled with MS analysis, two-dimensional polyacrylamide gel electrophoresis (2D-PAGE) has been widely used, but the low-throughput performance of 2D-PAGE could become a disadvantage in analyses that require the examination of several samples. The shotgun method is an alternative approach used in proteome analysis that involves liquid chromatography (LC) and tandem MS [11]. Previously, we constructed a system that exhibits ultraperformance in chromatographic separation achieved with a long monolithic silica capillary column; this system has enabled, for example, successful one-shot identification of 1658 proteins produced in Mesorhizobium loti in nodule or free-living conditions [12]. Thus, we adapted this nano LC-MS/MS proteome-analysis system for investigation into the mechanism of RA onset.

In this study, we stimulated a transformed FLS cell line, MH7A [13–16], with TNF-α and analyzed the intracellular protein profile by using an LC-MS/MS system equipped with a 500-cm-long monolithic silica capillary column. We observed substantial production of plasminogen-activator inhibitor (PAI) 2 and several apoptosis-regulating proteins in TNF-α-stimulated MH7A cells. Thus, TNF-α-stimulated upregulation of PAI-2 and apoptosis-regulating proteins in synovial cells might play a crucial role in the pathogenesis of RA.

Materials and methods

Cell line

The human MH7A synovial cell line (Riken, Saitama, Japan), which originated from intra-articular soft tissue of the knee joints of an RA patient, was established by transfecting cells with SV40 T antigen [13–16]. MH7A cells were cultured in Pure Coat 6-well plates (BD, Franklin Lakes, NJ, USA) with RPMI 1640 medium (Sigma, St. Louis, MO, USA) containing 10% heat-inactivated fetal bovine serum (Whittaker, Walkersville, MD, USA) and 100 U·mL⁻¹ penicillin and 100 μg·mL⁻¹ streptomycin (Invitrogen, Carlsbad, CA, USA). The culture plates were incubated for 24 h at 37°C in an atmosphere of 5% CO₂ in air. Subsequently, the plates were incubated for another 24 h in the presence or absence of 100 ng·mL⁻¹ TNF-α.

Protein preparation

Culture medium was aspirated from culture plates and the cells were washed twice with ice-cold PBS, after which ice-cold PBS was added to the wells and cells were removed from the plates by using a cell-scaper. These collected cells were transferred to conical tubes and total proteins were extracted using a Qproteome Mammalian Protein Prep Kit (Qiagen, Hilden, Germany).

Trypsin digestion

We mixed 1 mL of the protein supernatant with 135 μL of 200 mM triethylammonium bicarbonate (TEAB), 165 μL of distilled water, and 15 μL of 200 mM tris-(2-carboxyethyl) phosphine (Thermo Fisher Scientific, Waltham, MA, USA). The mixture was incubated at 55°C for 1 h, after which 15 μL of 375 mM iodoacetamide was added to the solution and incubated for 30 min. Next, the reactants were mixed with 3 mL of ice-cold acetone and incubated at −20°C for 2 h to precipitate proteins. Last, the precipitated proteins were resuspended in 100 μL of TEAB and mixed with 2 μL of 1 μg·μL⁻¹ sequencing-grade modified trypsin (Promega, Madison, WI, USA) at 37°C overnight.

LC-MS/MS analysis

Proteome analyses were performed using an LC-MS system (LC, UltiMate 3000 RSLCnano System, and MS, LTQ Velos Orbitrap mass spectrometer; Thermo Fisher Scientific) that was equipped with a long monolithic silica capillary column (500 cm long, 0.1 mm ID; Kyoto Monotech, Kyoto, Japan). Tryptic digests (5 μL) were injected and separated through reversed-phase chromatography at a flow rate of 500 nL·min⁻¹; the gradient was produced by changing the mixing ratio of these two eluents: A, 0.1% (v/v) formic acid, and B, 80% (v/v) acetonitrile containing 0.1% (v/v) formic acid. The gradient was started with 5% B, increased to 45% B for 600 min, further increased to 95% B to wash the column, and then returned to the initial condition and held for re-equilibration. The separated analytes were detected on a mass spectrometer with a full scan range of 350–1500 m/z. For data-dependent acquisition, the method was set to automatically analyze the top-10 most intense ions observed in the MS scan. An ESI voltage of 2.3 kV was applied directly to the LC buffer distal to the
chromatography column by using a MicroTee. The temperature of the ion transfer tube was set to 280 °C.

**Data analysis**

All samples were subjected to triplicate LC-MS/MS analysis, and the combined spectrometry data were used for protein identification. Proteins were identified by using MASCOT (Matrix Science, London, UK) against the assembly 21 protein database at SwissProt (2002–2015 UniProt Consortium, EMBL-EBI) that contains 20210 sequences, with a precursor mass tolerance of 20 ppm, fragment tolerance of 0.8 Da, and strict specificity allowing for up to one missed cleavage. For trypsin digestion, carbamidomethylation of cysteine was set as a fixed modification, and oxidation of methionine was set as a dynamic modification. The data were then filtered at a q-value ≤ 0.01 corresponding to 1% false-discovery rate on a spectral level. Triplicate analyses were performed for each sample of three biological replicates. The protein identification threshold was as follows; proteins identified using more than one unique peptide and three peptides; if not, proteins identified using a single unique peptide for each biological replicate. For gene ontology (GO) analysis, the DAVID annotation tool was used. A significant enrichment score means that the number of identified proteins belonging to their annotation term is significantly enriched comparing to the number of genes belonging to that. For comparing the protein amount of each gene, exponentially modified protein abundance index (emPAI) [17] was calculated with MACOT. An emPAI offers approximate, semiquantitative value. It is developed by Ishihama et al. [17] and emPAI values were calculated from number of identified peptides per protein and identifiable peptides per protein.

**Results**

**Screening for differentially produced proteins**

We conducted three independent experiments by using cultured MH7A cells that were untreated or stimulated with TNF-α. On the basis of the results of the proteome analysis, we identified 2650 proteins from untreated MH7A cells and 2688 proteins from TNF-α-stimulated MH7A cells, and then selected 269 differentially produced proteins that were detected under TNF-α stimulation (Fig. 1).

**GO analysis of differentially produced proteins**

The proteins that were differentially produced following stimulation with TNF-α were classified according to GO analysis performed by using DAVID as the

| Enrichment score | Category                              | Enrichment score | Category                              |
|------------------|---------------------------------------|------------------|---------------------------------------|
| 4.28             | Protein localization                   | 5.35             | RNA transport                         |
| 3.05             | Negative regulation of apoptosis      | 4.91             | Protein localization                  |
| 2.37             | Regulation of apoptosis                | 2.23             | Cellular protein catabolic process    |
| 2.18             | Protein localization in mitochondrion | 1.55             | Maintenance of protein location       |
| 1.87             | Apoptosis                              | 1.47             | Mitosis                               |
| 1.76             | Heme metabolic process                | 1.19             | Phosphate metabolic process           |
| 1.56             | T cell activation                     | 1.13             | GPI anchor metabolic process          |

Fig. 1. Summary of protein identification in MH7A cells. (A) Venn diagram of identified proteins; (B) gene ontology analysis of proteins identified with/without TNF-α stimulation.
The enrichment scores obtained for protein localization, negative regulation of apoptosis, regulation of apoptosis, protein localization in mitochondria, apoptosis, heme metabolic process, and T-cell activation were 4.28, 3.05, 2.37, 2.18, 1.87, 1.76, and 1.56, respectively (Fig. 1).

Plasminogen-activator inhibitor 2 and apoptosis-regulating proteins

Among the differentially produced proteins analyzed, we detected TNF-α signaling proteins such as nuclear factor-kappa-B (NF-κB) and AP-1 complex subunit σ1A, and synovial cell growth proteins such as transforming growth factor-β1 and IL-6 receptor-β. We also observed substantial production of PAI-2 and apoptosis-regulating proteins such as BH3-interacting domain death agonist (BID), autophagy protein 5 (ATG5), apolipoprotein E (ApoE), and caspase-3 in TNF-α-stimulated MH7A cells (Table 1).

Discussion

In TNF-α-stimulated MH7A cells, we detected NF-κB and AP-1 complex subunit σ1A. This suggested that TNF-α-induced intracellular signaling (Fig. 2) was
operating in the MH7A cells used in this experiment [18].

In MH7A cells stimulated with TNF-α, PAI-2 was also produced. Abnormalities of the synovial lining layer might represent an extremely early feature of synovitis in RA. Even in an early clinical stage, the surface of the synovial lining layer is frequently covered with fibrin deposits generated from the activation of the fibrinolytic system by synovial fluid. The synovial lining layer is completely replaced by a fibrin cap, and in highly inflamed tissues, this fibrin can extend into the sublining layers.

Proteolytic digestion of the fibrin clot is mediated by plasminogen activation. The plasminogen activation system has been widely suggested to play a crucial pathogenic role in destructive joint disease [19–21]. Busso et al. [22] reported that the concentrations of urokinase-type plasminogen activator (u-PA), PAI-1, PAI-2, and u-PA receptor were markedly higher in RA patients than in osteoarthritis patients. The u-PA, u-PA receptor, and PAI proteins produced in increased amounts were distributed mainly in the synovial lining area of proliferative and invasively growing synovial tissue in RA patients [22]. Our detection of substantial PAI-2 production in MH7A cells after TNF-α stimulation potentially supports these previous observations (Table 1).

We detected markedly increased production of apoptosis-regulating proteins such as ApoE, ATG5, and BID in TNF-α-stimulated MH7A cells (Table 1). ApoE might play a role in RA. A recent study demonstrated that ApoE-deficient mice were resistant to the development of collagen-induced arthritis [23]. Synovial fluid samples collected from the inflamed joints of RA patients contain several citrullinated proteins. Citrullinated ApoE is a newly identified antigen in RA synovial fluid, and only a limited number of the protein’s citrullinated epitopes are targeted by the immune system in RA [24]. Thus, the increased ApoE production in TNF-α-stimulated MH7A cells observed here suggests that the induction of citrullinated ApoE and autoimmune reactions could be involved in RA development (Fig. 2).

Autophagy is a highly conserved lysosome-mediated catabolic process and a homeostatic process that degrades unnecessary or dysfunctional cellular organelles and recycles nutrients. ATG5 is a critical protein required for autophagy at the stage of autophagosome-precursor synthesis. In addition to regulating autophagosome formation, ATG5 might play an important role in apoptosis [25–27]. Moreover, ATG5 regulates the production of inflammatory cytokines, the functions of antigen-presenting cells, and the clearance of apoptotic cells, which indicates that ATG5 might contribute to the pathogenesis of RA [28–31]. Our finding that ATG5 production was increased in TNF-α-stimulated MH7A cells suggests that apoptosis induction and immune-function regulation by this protein could underlie the development of RA (Fig. 2).

BID is a pro-apoptotic BH3-only Bcl-2 homolog that is involved in “extrinsic” apoptosis: BID is activated by cleavage only in response to cell-surface receptors that transmit apoptotic signals initiated by the specific ligand granzyme B produced by cytotoxic cells [32]. Peripheral blood neutrophils that were dying spontaneously as a result of apoptosis (i.e., through “intrinsic” apoptosis) showed no BID cleavage despite efficient caspase-3 activation. By contrast,
prominent BID cleavage was detected in neutrophils that were exposed to granzyme B/perforin or in neutrophils in which death receptor was activated (with anti-Fas or TNF-α/cycloheximide). In RA synovial fluid, BID cleavage was closely associated with the cleavage of caspase-3 [33]. The increased BID production in TNF-α-stimulated MH7A cells observed in this study suggests that RA development might involve BID activation-dependent induction of apoptosis (Fig. 2).

Previously, TNF-α was demonstrated to induce prolonged activation of NF-κB signaling and transcription of mRNAs encoding IL-6 and chemokines and chemokine receptors such as CXCL8/IL-8 and CCL5/RANTES [34]. A comparative proteomics study previously showed that proteins related to vasculature development were upregulated in RA FLS as compared with the corresponding levels in normal FLS [35]. However, this study has demonstrated for the first time that the plasminogen activation system is upregulated in synovial cells after TNF-α stimulation (Fig. 2).

In conclusion, we detected substantial production of PAI-2 and several apoptosis-regulating proteins in TNF-α-stimulated MH7A cells. Our results indicate that these upregulated proteins might contribute to the pathogenesis of RA.

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Author contributions

S.S., M.U., and T.I. conceived and designed the project, M.K., S.A., H.M., Y.T., S.K., and M.K. acquired the data, S.A., H.S., and W.A. analyzed and interpreted the data, S.S., M.U., and T.I. wrote the paper.

References

1 McInnes IB and Schett G (2011) The pathogenesis of rheumatoid arthritis. N Engl J Med 365, 2205–2219.
2 Firestein GS (2003) Evolving concepts of rheumatoid arthritis. Nature 423, 356–361.
3 Feldmann M (2009) Translating molecular insights in autoimmunity into effective therapy. Annu Rev Immunol 27, 1–27.
4 Bartok B and Firestein GS (2010) Fibroblast-like synoviocytes: key effector cells in rheumatoid arthritis. Immunol Rev 233, 233–255.
5 Müller-Ladner U, Kriegsmann J, Franklin BN, Matsumoto S, Geiler T, Gay RE and Gay S (1996) Synovial fibroblasts of patients with rheumatoid arthritis attach to and invade normal human cartilage when engrafted into SCID mice. Am J Pathol 149, 1607–1615.
6 Guerne PA, Zuraw BL, Vaughan JH, Carson DA and Lotz M (1989) Synovium as a source of interleukin 6 in vitro. Contribution to local and systemic manifestations of arthritis. J Clin Invest 83, 585–592.
7 Firestein GS and Zvaifler NJ (1990) How important are T cells in chronic rheumatoid synovitis? Arthritis Rheum 33, 768–773.
8 Ivashkiv LB (2011) Inflammatory signaling in macrophages: transitions from acute to tolerant and alternative activation states. Eur J Immunol 41, 2477–2481.
9 Armaka M, Apostolaki M, Jacques P, Kontoyiannis DL, Elewaut D and Kollia G (2008) Mesenchymal cell targeting by TNF as a common pathogenic principle in chronic inflammatory joint and intestinal diseases. J Exp Med 205, 331–337.
10 Anderson NG and Anderson NL (1996) Twenty years of two-dimensional electrophoresis: past, present and future. Electrophoresis 17, 443–453.
11 Aoki W, Ueda T, Tatsukami Y, Kitahara N, Morisaka H, Kuroda K and Ueda M (2013) Time-course proteomic profile of Candida albicans during adaptation to a fetal serum. Pathog Dis 67, 67–75.
12 Tatsukami Y, Nambo M, Morisaka H, Kuroda K and Ueda M (2013) Disclosure of the differences of Mesorhizobium loti under the free-living and symbiotic conditions by comparative proteome analysis without bacteroid isolation. BMC Microbiol 13, 180.
13 Miyazawa K, Mori A and Okudaira H (1998) Establishment and characterization of a novel human rheumatoid fibroblast-like synoviocyte line, MH7A, immortalized with SV40 T antigen. J Biochem 124, 1153–1162.
14 Shibasaki S, Karasaki M, Gräslund T, Nygren PÅ, Sano H and Iwasaki T (2014) Inhibitory effects of H-Ras/Raf-1–binding affibody molecules on synovial cell function. AMB Express 4, 82.
15 Shibasaki S, Tsunemi S, Kitano S, Sekiguchi M, Sano H and Iwasaki T (2014) Differential regulation of c-Met signaling pathways for synovial cell function. Springerplus 3, 554.
16 Shibasaki S, Fujita A, Usui C, Watanabe S, Kitano S, Sano H and Iwasaki T (2012) Effect of transient expression of fluorescent protein probes in synovial and myoblast cell lines. Springerplus 1, 36.
17 Ishihama Y, Oda Y, Tabata T, Sato T, Nagasu T, Rappisilber J and Mann M (2005) Exponentially modified protein abundance index (emPAI) for estimation of absolute protein amount in proteomics by
the number of sequenced peptides per protein. Mol Cell Proteomics 4, 1265–1272.
18 Dulos J, Kaptein A, Kavelaars A, Heijnen C and Boots A (2005) Tumour necrosis factor-α stimulates dehydroepiandrosterone metabolism in human fibroblast-like synoviocytes: a role for nuclear factor-κB and activator protein-1 in the regulation of expression of cytochrome p450 enzyme 7b. Arthritis Res Ther 7, R1271–R1280.
19 Hamilton JA (1982) Plasminogen activator activity of rheumatoid and nonrheumatoid synovial fibroblasts. J Rheumatol 9, 834–842.
20 Inman RD and Harpel PC (1986) Alpha 2-plasmin inhibitor-plasmin complexes in synovial fluid. J Rheumatol 13, 535–537.
21 Kikuchi H, Tanaka S and Matsuo O (1987) Plasminogen activator in synovial fluid from patients with rheumatoid arthritis. J Rheumatol 14, 439–445.
22 Busso N, Pèlat V, So A and Sappino AP (1997) Plasminogen activation in synovial tissues: differences between normal, osteoarthritis, and rheumatoid arthritis joints. Ann Rheum Dis 56, 550–557.
23 Asquith DL, Miller AM, Hueber AJ, Liew FY, Sattar N and McInnes IB (2010) Apolipoprotein E-deficient mice are resistant to the development of collagen-induced arthritis. Arthritis Rheum 62, 472–477.
24 van Beers JJ, Schwarte CM, Stammen-Vogelzangs J, Oosterink E, Božič B and Prujin GJ (2013) The rheumatoid arthritis synovial fluid citrullinome reveals novel citrullinated epitopes in apolipoprotein E, myeloid nuclear differentiation antigen, and β-actin. Arthritis Rheum 65, 69–80.
25 Kametaka S, Matsuura A, Wada Y and Ohsumi Y (1996) Structural and functional analyses of APG5, a gene involved in autophagy in yeast. Gene 178, 139–143.
26 Kuma A, Hatano M, Matsu M, Yamamoto A, Nakaya H, Yoshimori T, Ohsumi Y, Tokuhisa T and Mizushima N (2004) The role of autophagy during the early neonatal starvation period. Nature 432, 1032–1036.
27 Pyo JO, Jang MH, Kwon YK, Lee HJ, Jun JI, Woo HN, Cho DH, Choi B, Lee H, Kim JH et al. (2005) Essential roles of Atg5 and FADD in autophagic cell death: dissection of autophagic cell death into vacuole formation and cell death. J Biol Chem 280, 20722–20729.
28 Tal MC, Sasai M, Lee HK, Yordy B, Shadel GS and Iwasaki A (2009) Absence of autophagy results in reactive oxygen species-dependent amplification of RLR signaling. Proc Natl Acad Sci USA 106, 2770–2775.
29 Jouan N, Takeshita F, Kobiyama K, Sawano A, Miyawaki A, Xin KQ, Ishii KJ, Kawai T, Akira S, Suzuki K et al. (2007) The Atg5–Atg12 conjugate associates with innate antiviral immune responses. Proc Natl Acad Sci USA 104, 14050–14055.
30 Qu X, Zou Z, Sun Q, Luby-Phelps K, Cheng P, Hogan RN, Gilpin C and Levine B (2007) Autophagy gene-dependent clearance of apoptotic cells during embryonic development. Cell 128, 931–946.
31 Lee HK, Mattei LM, Steinberg BE, Alberts P, Lee YH, Chervonsky A, Mizushima N, Grinstein S and Iwasaki A (2010) In vivo requirement for Atg5 in antigen presentation by dendritic cells. Immunity 32, 227–239.
32 Li H, Zhu H, Xu CJ and Yuan J (1998) Cleavage of BID by caspase 8 mediates the mitochondrial damage in the Fas pathway of apoptosis. Cell 94, 491–501.
33 Romero V, Fert-Bober J, Nigrovic PA, Darrah E, Haque UJ, Lee DM, van Eyk J, Rosen A and Andrade F (2013) Immune-mediated pore-forming pathways induce cellular hypercitrullination and generate citrullinated autoantigens in rheumatoid arthritis. Sci Transl Med 5, 209ra150.
34 Lee A, Qiao Y, Grigoriev G, Chen J, Park-Min KH, Park SH, Ivashkiv LB and Kalliolia GD (2013) TNFα induces sustained signaling and a prolonged and unremitting inflammatory response in rheumatoid arthritis synovial fibroblasts. Arthritis Rheum 65, 928–938.
35 Wang JG, Xu WD, Zhai WT, Li Y, Hu JW, Hu B, Li M, Zhang L, Guo W, Zhang JP et al. (2012) Disorders in angiogenesis and redox pathways are main factors contributing to the progression of rheumatoid arthritis: a comparative proteomics study. Arthritis Rheum 64, 993–1004.