Promising Antioxidant Activity of *Erythrina* Genus: An Alternative Treatment for Inflammatory Pain?

Tania Jiménez-Cabrera 1, Mirandeli Bautista 1,*, Claudia Velázquez-González 1, Osmar Antonio Jaramillo-Morales 2,*, José Antonio Guerrero-Solano 1,*, Thania Alejandra Urrutia-Hernández 1 and Minarda De la O-Arciniega 1,*

1 Institute of Health Sciences, Autonomous University of the State of Hidalgo, San Agustín Tlaxiaca 55679, Hidalgo, Mexico; taniaJimenez@uah. edu.mx (T.-C.); claudia@uah.edu.mx (C.-V.-C.); gsNutricional@gmail.com (J.A.G.-S.); thania.Urrutia956@uah.edu.mx (T.A.-U.-H.)
2 Nursing and Obstetrics Department, Life Sciences Division, Campus Irapuato-Salamanca, Ex Hacienda El Copal, Km. 9 Carretera Irapuato-Silao, A.P 311, Irapuato 36500, Guanajuato, Mexico; oa.jaramillo@ugto.mx

*Correspondence: mibautista@uah.edu.mx (M.B.); mina@uah.edu.mx (M.D.I.O.-A.)

Abstract: The negative impact that oxidative stress has on health is currently known. The complex mechanism of free radicals initiates a series of chain reactions that contribute to the evolution or development of different degenerative disorders. Likewise, these disorders are usually accompanied by inflammatory processes and, therefore, pain. In this sense, reactive oxygen species (ROS) have been shown to promote the nociceptive process, but effective treatment of pain and inflammation still represents a challenge. Over time, it has been learned that there is no single way to relieve pain, and as long as there are no other alternatives, the trend will continue to apply multidisciplinary management, such as promote the traditional use of the *Erythrina* genus to manage pain and inflammation. In this sense, the *Erythrina* genus produces a wide range of secondary metabolites, including flavanones, isoflavones, isoflavones, and pterocarpans; these compounds are characterized by their antioxidant activity. Phenolic compounds have demonstrated their ability to suppress pro-oxidants and inhibit inflammatory signaling pathways such as MAPK, AP1, and NFκB. Although there is preclinical evidence supporting its use, the pharmacological effect mechanisms are not entirely clear. Nowadays, there is a fast advancement in knowledge of the disciplines related to drug discovery, but most of nature’s medicinal potential has not yet been harnessed. This review analyzes the decisive role that the *Erythrina* genus could play in managing inflammatory pain mediated by its compounds and its uses as an antioxidant.

Keywords: *Erythrina*; antioxidant; inflammatory pain; prenylated flavonoids

1. Molecular Origin of Inflammatory Pain

Pain is traditionally defined as a complex sensory and emotional experience associated with actual or potential tissue damage or described in terms of such damage (International Association for the Study of Pain) [1–3]. It is a complex process that involves neuronal signaling pathways between the peripheral nervous system (PNS) and the central nervous system (CNS) [1,3]. The transduction of noxious stimuli (those that actually or potentially damage tissues) is carried out by a nociceptor, creating an electrophysiological neuronal signal encoded in the form of an action potential that is transmitted to the CNS. The acute injury is associated with a first, well-localized pain sensation transduced and transmitted by nociceptors. Although pain is one of the body’s most important adaptation and protection mechanisms, the degree of tissue damage leads to the release of inflammatory mediators that bind to its receptors, triggering an enzymatic cascade [1]. Thus, inflammatory pain is generated by an increase in sensitivity due to the cellular response associated with tissue damage, promoting the influx of activated cells such as macrophages, lymphocytes, and mast cells that release inflammatory mediators such as bradykinin, Hþ ions, ATP, purines,
prostaglandin E2, leukotrienes, cytokines, nerve growth factor (NGF), sympathetic amines, and oxidative stress products present in the membrane of nociceptors [4] (Figure 1). NGF released by activated macrophages acts directly on peptidergic C fibers that express the TrkA receptor, a key component of peripheral sensitization. Macrophages release cytokines such as interleukin-6 (IL-6), IL-1β, tumor necrosis factor α (TNFα) which in turn contribute to peripheral sensitization through increased local production of proalgesic agents such as bradykinin, prostaglandins and increased release of NGF [1]. Prostaglandins are synthesized by consecutive reactions initiated by the phospholipase A2 enzyme that causes the release of arachidonic acid (AA) from cell membranes. Cyclooxygenase-2 (COX2) metabolizes arachidonic acid to prostaglandin G2 (PGG2) and prostaglandin H2 (PGH2) which is ultimately converted to PGE2 by prostaglandin E synthase (PTGES) [5]. PGE2 acts on all four E-prostanoid (EP) receptor subtypes (EP1-4). In peripheral tissue, PGE2 modulates pain sensitivity by sensitizing primary afferents. Sensitizes ion channels involved in pain, namely transient receptor potential vanilloid 1 channel, tetrodotoxin-sensitive Na+ channels, and purinergic P2X3 channels, also enhances the release of neurotransmitters. So, it is a crucial lipid mediator of inflammatory responses that causes pain hypersensitivity [6].

The pathological effects of ROS, IL-1β, TNFα, and IL-6 are due to the activation of various pro-inflammatory signaling pathways including the three (ERK, JNK, and p38) mitogen-activated protein kinases (MAPK), NFκB, AP1, and JAK/STAT [7]. There is evidence that mitochondrial dysfunction induced by oxidative and nitrosative stress leads to peripheral and central sensitization. Mammalian nerves are especially susceptible to free radicals, both oxygen (ROS) and nitrogen (RNS), due to the high content of phospholipids and axonal mitochondria; in addition to having a weak antioxidant system and other hand, some studies on antioxidant supplementation in animal models show that hydroxyl radicals (OH), superoxide (O₂⁻), and nitric oxide (ON) may have a role in peripheral sensitization due to a deleterious effect on lipids and nucleic acids, protein carbonylation, and therefore the involvement of organelles and antioxidant enzymes [8]. Furthermore, ROS have also been shown to act as signaling molecules in a wide variety of cellular processes, including proliferation and survival (MAP kinases and PI3 kinase) and the regulation of antioxidant genes (Ref-1 and Nrf-2) [7]. PI3K catalyzes the synthesis of the second messenger phosphatidylinositol 3,4,5-triphosphate (PIP3) from phosphatidylinositol 4,5-bisphosphate (PIP2), wherein the membrane-bound PIP3 serves as a signaling molecule to recruit proteins containing the pleckstrin homology (PH) domain. These PH domain

![Diagram of the process of pain and peripheral sensitization leading to inflammatory pain.](image_url)
proteins, such as the phosphoinositide-dependent protein kinase (PDK) and protein kinase B (AKT) serine/threonine kinases are thus activated and mediate further downstream signaling events. Both PI3K and MAPK are similarly regulated by ROS at the oxidative interface, where protein phosphatases are directly oxidized by ROS, resulting in sustained activation of signaling pathways [7]. According to the International Association for the Study of Pain (IASP), although comprehensive epidemiological data are not worldwide available, almost 50% of adults suffer from more than one type of pain [9]. Pain is associated with most of the diseases, however, particularly the skin, joints, and intestines are susceptible to the development of inflammatory pain, and its prevalence is increased [10]. Therefore, the effective management of pain and inflammation represents a challenge in clinical research, as the scientific discipline of pain management is a relatively new field of research [4].

Nowadays, the general population suffers different types of collateral damage, which leads to the need to find more effective drugs, with fewer side effects and greater accessibility, to eliminate the inflammatory process and the associated pain. Furthermore, nociceptive stimuli do not always respond to common analgesics or NSAIDs, so other therapies or therapeutic options [11] as natural products, are used. They represent a desirable approach for developing new drugs, particularly useful in patients with inflammatory pain [12] the antioxidant activity of natural products assumes a decisive role in the management of inflammation and accompanying pain [13]. Antioxidants already known, such as vitamin E, resveratrol, or quercetin have shown tables analgesic and anti-inflammatory properties due to the properties that their chemical structure confers on them [14,15]. Although today there is rapid growth and advancement in knowledge of the disciplines related to drug discovery, the medicinal potential of most of nature has not yet been harnessed [12].

The species of genus *Erythrina*, have a great variety of medicinal properties. It has been widely used in folk therapies due to their curare and hypnotic functions and their associated pharmacological effects, including sedatives, hypotensives, neuromuscular blockers, and central nervous system (CNS) depressants [16,17]. Besides, they have been also used to treat microbial infections [18], inflammation [19], amenorrhea, headache, eye problems, female sterility, liver disorders, asthma, and malaria diseases [20–22].

2. Ethnomedicinal Use of the Genus *Erythrina*

This genus is a member of the legume family (*Fabaceae*), subfamily *Papilionoideae*. It comprises at least 120 species most of which are trees and some perennials with large woody roots [23]. These species are collectively called “coral trees”, alluding to the flowers, characteristics of the genus, which are commonly bright red [24]. The place of origin of the genus *Erythrina* is not exactly known, but it is suggested that it was probably in South America, since most of the supposed “primitive” groups within the genus are found there. 70 species are recognized in the Neotropics, 38 in Africa and Madagascar, and 12 in Asia and Australia [23].

Although there is local ethnobotanical data on the use of the genus *Erythrina* to relieve pain and inflammation, few preclinical studies to evaluate the effect have been published. An even smaller amount describes the biological activity and therapeutic potential of the genus for this purpose. Furthermore, the relationship between antioxidant properties and anti-inflammatory effect has not been analyzed. Therefore, this review will focus on the ethnopharmacological analysis of the genus *Erythrina* and the decisive role that it could play in the management of inflammatory pain.

There are some papers that documented the ethnomedicinal use of different species from *Erythrina* genus in the treatment of pain and/or inflammation, such as: *E. abyssinica*, *E. caffra* [25–30] and *E. arborences* [31,32] are the most used species in traditional medicine. Table 1 shows a summary of the species studied the part of the tree used, the method of preparation and their ethnomedicinal use. The bark and leaves are the common part of the plant used for medicinal purposes. The decoction is the habitual form of preparation, the liquid obtained is ingested or applied externally on the affected area.
Table 1. Species of the genus *Erythrina* used in traditional medicine for the relief of pain or inflammation.

| Species           | Part Used      | Folk Use                               | Administration                          | Reference       |
|-------------------|----------------|----------------------------------------|-----------------------------------------|-----------------|
| *E. abyssinica*   | Bark           | Inflammation, backache, pain and cramps lower belly | Decoction, external use, extract drunk and boiled in milk. | [25,27,33]      |
| *E. arborencens*  | Branch, seed and leaves | Bone fracture and back pain          | Paste/fomentation, decoction oral, juice of leaves | [31,32]         |
| *E. caffra*       | Bark, leaves and roots | Sprains, aches              | Decoction oral, eardrops and plaster    | [26,28,29]      |
| *E. caffra*       | Stem bark and leaves | Toothache and earache            | Oral infusion                           | [30]            |
| *E. edulis*       | Bark           | Headache                               | Aqueous infusion drink                   | [34]            |
| *E. humeana*      | Bark           | Sprains                                | Decoction, external use, massage with ointment | [26]            |
| *E. senegalensis* | Bark           | Inflammation and Backache            | Decoction, external use, massage with ointment | [22]            |
| *E. variegata*    | Leaves and bark | Fever, body ache, chronic bronchitis and otalgia | Decoction, oral | [35]         |

3. Preclinical Studies of Pain and Inflammation from the Genus *Erythrina*

Table 2 shows preclinical studies on the analgesic and anti-inflammatory effect of species of the genus *Erythrina* genus. *E. variegate* was the most studied species, both for its analgesic and anti-inflammatory effect [36–40], followed by *E. velutina* and *E. mulungu* [41–43]. Mostly, high polarity solvents (ethanol, water, and methanol) were used to obtain the extracts. This suggests that the biological activity demonstrated in the various studies, was largely due to polar compounds. Mostly, high polarity solvents (ethanol, water, and methanol) were used to obtain the extracts. This suggests that the biological activity demonstrated in the various studies was largely due to polar compounds. Two flavanones (Sigmoidin A and B), a prenylated flavonoid (abyssinone V-4′-methyl ether), a prenylisoflavone (warangalone), and a pterocarpane (Erycristagallin) (Figure 2), demonstrated anti-inflammatory properties at doses of 300–600 mg/kg [44–46], all compounds demonstrated marked anti-inflammatory efficacy. The anti-inflammatory suggested mechanisms include the inhibition of prostaglandins and cyclooxygenases. The most widely used reference drugs in pain models were morphine and diclofenac. While for the inflammation models it was Indomethacin, diclofenac, and dexamethasone. Although in some cases, acetylsalicylic acid and Pentazocin were used. In general, all preclinical studies agree that each species studied is shown to have an analgesic and/or anti-inflammatory effect [36,40,41,46]. Antagonistic effects with histamine and/or serotonin were also mentioned [40,46]. Although their causes are not clarified, blocking of HRs and 5-HT receptors are related [47]. Likewise, the participation of antioxidant activity in the regulation of anti-inflammatory and analgesic processes through the inhibition of nitric oxide (NO) is highlighted [45,48,49]. According to the authors, the compounds involved in these mechanisms are mainly flavonoids. However, it is also mentioned that alkaloids erysotrine, erysotrine hypophorine, reduced the number of inflammatory cells in lung tissue, mainly eosinophils and lymphocytes. Possibly due to the decrease of IL-4 and IL-5, which stimulate the maturation of eosinophils in the bone marrow and recruit these cells to the tissues. In turn, this can impact the modulation of the synthesis and release of inflammatory mediators, such as prostaglandins, nitric oxide, and cytokines such as IL-1 and TNF α [19]. Docking studies shows that phaseollin of *Erythrina variegata* has the best fitness score against the COX-1 which is 56.64 and 59.63 for COX-2 enzyme [50–57]. However, is required to delve into the possible mechanisms of action, as well as the phytoconstituents and their relationship with the biological activity [58].
Table 2. Preclinical studies on the analgesic and anti-inflammatory effect of species of the genus Erythrina.

| Species          | Extract                  | Part                | Model                                                                 | Reference |
|------------------|--------------------------|---------------------|----------------------------------------------------------------------|-----------|
| *E. addisoniae*  | EtOAc and MeOH           | Stem bark           | Inhibition of leukotriene B4 production from rat polymorphonuclear leukocytes. Cyclooxygenase-1 (COX-1) activity from human platelets. PLA2 induced paw oedema in mouse. TPA-induced mouse ear oedema. | [46]      |
| *E. lysistemon*  | Ethanol and ethyl acetate| Leaves and bark     | Cyclooxygenase-1 inhibition                                           | [48]      |
| *E. latissima*   | Ethanol and ethyl acetate| Leaves and bark     |                                                                   |           |
| *E. humeana*     | Ethanol and ethyl acetate| Leaves and bark     |                                                                   |           |
| *E. zeyheri*     | Ethanol and ethyl acetate| Leaves and bark     |                                                                   |           |
| *E. indica*      | MeOH                     | Leaves              | Carrageenan-induced hind paw edema                                   | [59]      |
| *E. droogmansina*| Ethyl acetate and MeOH   | Root bark           | Carrageenan-induced hind paw edema Ear edema induced by xylene Cotton pellet-induced granuloma | [45]      |
| *E. crist-galli* | EtOH (70%) Dichlorometane| Aerial parts        | Writhing test, Formalin test, Hot-plate                             | [60]      |
| *E. mildbraedii* | Ethyl acetate            | Root bark           | Carrageenan-induced hind paw edema PLA2 induced paw oedema in mouse | [57]      |
| *E. mulungu*     | EtOH and EtOH 30%        | Flowers and stem bark| Ovalbumin (OVA)-induced asthma in mice Dextran induced paw edema     | [19,41, 61]|
| *E. senegalensis*| Aqueous and EtOH (70%)   | Bark and roots      | Writhing test, Egg albumin induced paw edema in rats. Hot-plate      | [62,63]   |
| *E. sigmoidea*   | Chloroform               | Bark                | Inhibition of leukotriene B4 production from rat polymorphonuclear leukocytes. Cyclooxygenase-1 (COX-1) activity from human platelets. PLA2 induced paw oedema in mouse. TPA-induced mouse ear oedema. | [49]      |
| *E. variegata*   | MeOH, EtOH (95%), EtOH and Aqueous. | Leaves and bark | Writhing test, Tail-flk Carrageenan-induced hind paw edema Cotton pellet induced granuloma Hot plate HRBC membrane stabilization | [36,39, 40]|
| *E. velutina*    | EtOH (30%) and Aqueous    | Stem bark and leaves| Writhing test, Formalin test, Hot-plate Carrageenan-induced hind paw edema | [41,62]   |
Figure 2. Main chemical structures are involved in the inhibition of the inflammatory process. Sigmoidin A and B (1), Erycristagallin (2), Abyssinone V-4′-methyl ether (3), Waragalone (4), Mildbenone (5), Mildbone (6), 2″-O-galloyl orientin (7), Neobavaisoflavone (8) and Hypaphorine (9). The chemical structures were created with the ACD/ChemSketch Freeware program.

4. Radical Scavenging Activity in the Model In Vitro Systems

Various methods are used to investigate the antioxidant property of different samples. Can be classified so in vitro and in vivo antioxidant models [64]. The antioxidant activity of the extracts and/or compounds of *E. abyssinica*, *E. livingstoniana* and *E. mildbraedii* it proved in different studies [51,55,56,65,66]. In vitro radical scavenging assay (DPPH) was the most widely used in vivo test to determine the capacity of free radical scavenging (Table 3). In most studies, a similar and even higher activity was obtained than the positive controls (Trolox, BHA, ascorbic acid and quercetin) [58,65–68]. Reduction of Fe ions was also evaluated, an assay often used as an indicator of electron donation activity (FRAP). Additionally, the in vivo antioxidant activities of the enzymes SOD, CAT, and GSH were measured to evaluate the hepatoprotective potential of *Erythrina indica*, *senegalensis*, and × neillii, in rats. Where the activities of antioxidant enzymes were restored (*p* < 0.05) [69]. Inhibition of lipoxygenase and xanthine oxidase, enzymes that participate in the production of reactive oxygen species and pro-inflammatory agents were other tests used [55,56,70]. Likewise, the decrease in lipid peroxidation (TBARS) and the inhibition of NO were used to evaluate the antioxidant properties [52,53,71,72]. Among the compounds responsible
for these activities, Eryvarin H, Abyssinone V, mildbone, mildbenone, 7,3’-dihydroxy-4’-methoxy-5’-(3-methylbut-2-enyl)flavanone, erylivingstone H, 7,3’,4’-trihydroxyflavanone, trans-3,4,2’,4’-tetrahydroxychalcone, Eryvarin J and erycrisagallin [46,55,65,68,73–75]. Flexible molecular docking on heme oxygenase, an important stress protein that is involved in cellular protection, antioxidant and anti-inflammatory activities, showed with 2”-O-galloyl orientin forming four binding interactions with residues, Arg 136 (two interactions), Met34 and Gly139 [39]. On the other hand, it has been reported that the compound abyssinone V increases oxidative stress and reduces stress resistance in the Caenorhabditis elegans model [76]. However, many antioxidant compounds are also evaluated for their cytotoxic activity that promotes apoptosis favoring a pro-oxidant environment. This is highly dependent on the used concentrations of the compound. However, studies are required to help clarify this activity.

Table 3. Antioxidant activity of species from genus Erythrina.

| Species          | Part                  | Identified Extract or Compounds                                                                                   | Model          | Reference          |
|------------------|-----------------------|------------------------------------------------------------------------------------------------------------------|----------------|--------------------|
| *E. abyssinica*  | Stem bark and root    | Erycristagallin (4), 3-hydroxy-9-methoxy-10-(3,3-dimethylallyl) pterocarpene and 7,3’,4’-trihydroxy-5’-prenylflavanone (Abyssinone VII) | DPPH           | [50,51]            |
| *E. burttii*     | Root bark             | Burttinol-A and burttinol-C, and the 2-arylbenzofuran derivative burttinol-D                                     | DPPH           | [77]               |
| *E. crista-galli*| Bark                  | Alkaloids, erythraline, erythrinine and hypaphorine                                                               | Inhibitory activity on LPS-induced nitric oxide (NO)     | [72]               |
| *E. droogmansiana* | Root bark             | Genistein, 3-(3’,4’-methelenedioxyphenyl)-2,3-epoxypropanol, asperphenamate, Erydroogmansin B, vogelin C, Isolupalbigenin and erypostyrene | DPPH and FRAP  | [78,79]            |
| *E. edulis*      | Seeds                 | Protein concentrate from the seed flour                                                                          | ABTS, DPPH and ORAC | [80]               |
| *E. indica*      | Leaves and stem bark  | Methanol extract                                                                                                | DPPH) and Nitric oxide scavenging assay                  | [52,53]            |
| *E. livingstoniana* | Stem bark and twing  | 7,3’-dihydroxy-4’-methoxy-5’-(3-methylbut-2-enyl) flavanone, 7, 3’,4’,3’-trihydroxyflavanone and trans-3,4,2’,4’-tetrahydroxychalcone | DPPH           | [65,81]            |
| *E. variegata*   | Leaves and bark       | Methanolic extract and crude polysaccharides                                                                     | DPPH, FRAP and TEAC | [67,82]            |
| *E. mildbraedi*  | Roots and bark        | Flavanone (mildbone), chalcone (mildbenone) and Pterocarpene (Erycristagallin)                                | DPPH           | [55,56,83]         |
| *E. senegalensis* | Stem bark and leaves  | Hydroalcoholic extract fraction 3 (polyphenols and flavonoids) and Methanol extract                              | DPPH, ABTS and FRAP | [84,85]            |
| *E. sigmoidea*   | Stem bark             | Methanol extract; Flavanones, Sigmoidin A and Sigmoidin B                                                        | DPPH           | [49,67]            |
| *E. stricta*     | Leaves                | Hydromethanolic extract was In vitro xanthine oxidase inhibitory activity All                                   | DPPH           | [70]               |
| *E. suberosa*    | Flowers               | Methanol extract                                                                                                | DPPH and Nitric oxide scavenging assay                  | [69]               |
| *E. vogelii*     | Leaves                | Ethanol extract                                                                                                | DPPH           | [67]               |
| *E. neillii*     | Leaves                | Methanol total extract and its fractions                                                                       | ORAC           | [58]               |
As mentioned earlier an increase in free radicals exacerbates the inflammatory response. Likewise, it has been observed that supplementation with antioxidants in animal models can decrease peripheral sensitization caused by ROS. For which it is suggested that compounds with antioxidant activity of the genus *Erythrina* may play an important role in the modulation of inflammatory pain.

Although medicinal plants often have many different uses within and between cultures, much remains to be investigated about the species of this genus in terms of their potential in treating pain and inflammation.

5. Molecular Mechanisms of Anti-Inflammatory Activity of the Genus *Erythrina*

Medicinal plants and their secondary metabolites, such as polyphenols and alkaloids, have long been considered valuable sources of natural remedies for the treatment of human diseases [86]. At least 25% of modern medicines come directly or indirectly from plant origin [12,87]. However, even today with the rapid growth and advance in the knowledge of disciplines related to drug discovery, it may be that most of nature’s medicinal potential has yet to be tapped [12].

Such is the case of the genus *Erythrina*, which is made up of a great variety of species and characterized by producing a wide range of secondary metabolites. At least 370 flavonoid compounds have been isolated, including flavones, flavonols, flavanones, chalcones, isoflavans, isoflav-3-enos, neo flav-3-ene, isoflavanones, isoflavones, pterocarpans, coumestanes, arylcoumarins, coumaryl benin chromones that include flavoflavones, isoflavones, isoflavanones, pre-C-erythrine alkaloids and approximately 143 alkaloids distributed mainly in seeds, stem, bark, leaves and flowers [37,88].

Pain is always associated with the region where the inflammation is located and can become chronic if the inflammation is not relieved quickly [1]. Several mediators are involved in the inflammatory pathway, prostaglandins, leukotrienes, cytokines, platelet-activating factor, and chemokines [89]. Likewise, during the inflammatory process reactive oxygen and nitrogen species are also produced along with different proteases that can cause tissue damage, fibrosis, and cell proliferation, which can contribute to the chronicity of inflammation [90]. Behind these processes, there is a complex signaling network between the immune system and injured tissue [91]. The deepening of the knowledge of inflammatory pain will allow to optimize and accelerate the development of innovative therapeutic targets of natural origin [92].

The *Erythrina* alkaloids have been of interest for their structural characteristics and their variety of biological activities [93,94]. The dihydro-β-erythroidine alkaloid was used to characterize the nicotinic acetylcholine receptors (nAChRs) [94], a preferential antagonist of the α4β2 nicotinic receptor subunit that acts as a competitive inhibitor like erisothrin, erisopine, erisodine [59]. nAChRs are involved in several central nervous system (CNS) disease states, including depression, schizophrenia, attention deficit hyperactivity disorder, Alzheimer’s and Parkinson’s diseases, substance abuse, and pain [95,96]. On the other hand, it has been suggested that nAChRs may represent viable targets for new analgesics [96,97]. These receptors are widely distributed throughout the CNS, expressing themselves in neurons and non-neuronal cells [98]. In recent years, α7 nAChRs in macrophages has been shown to regulate inflammation, activating the “cholinergic anti-inflammatory pathway” [98–100]. There is accumulating evidence suggesting that α7 nAChR agonists and modulators are promising targets for the treatment of chronic inflammatory pain [101]. The treatment with *E. mulungu* extract significantly reduced the levels of pro-inflammatory cytokines, as well as the infiltration of inflammatory cells in lung tissue. The main compounds identified in the extract were erisothrin, erisothrin-N-oxide and hypaphorine (Figure 1). The cholinergic anti-inflammatory pathway allows the suppression of inflammation, it was characterized by its effects on the release of cytokines by macrophages. This pathway allows the suppression of inflammation by vagal efferents depending on α7 nAChRs [99]. Hypaphorine, an anti-inflammatory compound [102] has been isolated from many *Erythrina* species and other plant species [98,102–104]. A study carried out
by Aswad 2017 determined that hypaphorine is one of the molecules that is considered as a potential candidate for an anti-inflammatory drug. Hypophorine present in *Vaccaria segetalis* demonstrated downregulation of cyclooxygenase-2 (COX-2) and inducible nitric oxide synthase (iNOS). Furthermore, it delayed LPS-induced phosphorylation of ERK, and immunofluorescence staining revealed that *Vaccaria* hypophorine eliminated nuclear translocation of NFκB in LPS-treated RAW 264.7 cells [102]. During inflammation, the action of α7 nAChR is associated with the entry of calcium and the interruption of the stimulation of nuclear factor κB (NFκB) [105]. It is possible that *Eryhrina* hypaphorine regulates the inflammatory process through α7 nAChRs, activating a cholinergic anti-inflammatory pathway.

Several studies have shown that the anti-inflammatory activity of some polyphenols depends on their ability to suppress pro-inflammatory signaling pathways such as MAPK, AP1, and NFκB, and in turn, this ability is associated with the ability to restore a suitable redox environment [106,107]. Among the members of the MAPK cascades, apoptosis signal-regulated kinase 1 (ASK1) is an upstream MAPKKK that regulates the JNK and p38 MAPK pathways. ASK1 is activated under various stress conditions including oxidative stress. ASK1-deficient mouse embryonic fibroblasts were decreased JNK and p38 MAPK activation. ROS-activated ASK1 mediates p38 signaling pathway leading to nonapoptotic outcomes that probably favor the increase of pro-inflammatory cells [7]. Flavonoids are molecules of interest due to their biological effects observed in vitro. Their potential utility as antibiotic agents have been validated [18] anti-allergy [108] anti-diarrhea [109] antiulcer [110] anti-inflammatory [49,90] and analgesics [60]. At first, it was considered that the main mechanism of action of antioxidant compounds lays in their ability to scavenge radicals directly. Although the mechanisms that participate in these processes have not been studied in depth. The possible mechanisms may be related to effects on intracellular and intercellular signaling pathways, regulation of nuclear transcription factors, fat metabolism, and modulation in the synthesis of inflammatory [111]. Since the inhibition of pro-inflammatory enzymes, such as cyclooxygenase-2 (COX-2), lipooxygenase (LOX) and inducible nitric oxide synthase (iNOS) has been demonstrated. Pretreatment of primary chondrocytes and cartilage explants with Imperatorin, a plant secondary metabolite belonging to the family of furanocoumarins, suppressed the production of iNOS and NO, blocking IL-1β-induced phosphorylation of the ERK-MAPK/AP1 signaling pathway [106]. Inhibition of protein kinases such as phosphoinositol kinase, protein kinase C, phosphatidylinositol kinase has been documented, tyrosine kinase, or cyclin-dependent kinase-4. As well as the activation of phase II detoxifying pathways through the activation of factor 2 related to erythroid nuclear factor 2 (Nrf2). Additionally. Additionally, several flavonoids can decrease the expression of different pro-inflammatory cytokines/chemokines, including TNFα, IL-1β, IL-6, IL-8, and the monocyte chemoattractant protein-1 (MCP-1), in different types of cells, such as RAW macrophages, Jurkat T cells, and peripheral blood mononuclear cells [90,112]. Phenolic compounds isolated and characterized from *E. neillii* exhibited the highest antioxidant activity, principally 2”-O-galloyl orientin, followed by 2”-O-galloyl vitexin. Additionally. Additionally, flexible molecular docking on heme oxygenase (HO-1), an important stress protein that is involved in cellular protection, antioxidant and anti-inflammatory activities, justified the antioxidant activity of the isolated compounds [58].

One of the pathways implicated in the control of inflammation Nrf2 that controls the expression of antioxidant response element-regulated antioxidant and cyto-protective genes, such as NAD(P)H: quinone oxidoreductase 1 (NQO1), γ-glutamyl cysteine synthetase catalytic subunit (GCLC), and heme oxygenase (HO)-1 [113]. From what can be suggested, 2”-O-galloyl orientin, maybe a potential activator of Nrf2 and therefore play a fundamental role in the treatment of inflammatory pain.

The hydroalcoholic extracts of *E. indica* and *E. suberosa*, have shown percentages of inhibition of nitric oxide 21.5% to 89%. Given the polarity of the solvent used for the extraction, the participation of phenolic compounds in the reduction of nitric oxide was suggested [54,69]. The production of NO and prostaglandins is regulated by iNOS and
COX-2. iNOS is distinguished by generating more NO than the constituent members, it is involved in the development and maintenance of central and peripheral sensitization in inflammatory and neuropathic pain. NO can act as a neurotransmitter that affects spinal nociceptive processing in various pain models. COX-2 in macrophages increases in an oxidizing environment and, in turn, increases inflammatory responses [114]. So too, the ethanol and ethyl acetate bark extracts of *E. caffra*, *E. latissima*, and *E. lysistemon* exhibited an important cyclooxygenase inhibiting activity. *E. caffra* and *E. lysistemon* displayed inhibition of more than 90% for both the 50 and 500 mg/mL doses, this suggested the presence of potent compounds in the bark, as flavonoids. *E. caffra* is one of *Erythrina* species most frequently used by traditional healers to relieve inflammation [48]. On the other hand, the administration of ethanolic extract of leaves from *E. neillii* at 100, 250, and 500 mg/kg in the methotrexate-intoxicated rats significantly mitigated lipid peroxidation activity, with a significant decrease of malondialdehyde (MDA) in the hepatic tissue, and a significant increase of GSH and SOD activity, in a dose-dependent manner. Additionally, a significant decrease in the hepatic tissue content of TNF-α was demonstrated [71]. Likewise, the GSH, SOD, and CAT content significantly increased (*p* < 0.05 to *p* < 0.001) in the groups treated with the methanol extract of *E. indica* leaves [52]. All this compared with silymarin a potent antioxidant. It was suggested that flavones such Liquiritin, derived from plant licorice, increased SOD, CAT, GSH-PX enzymatic activity through activating the Nrf2/keap1 pathway and attenuation the ERK1/2/NF-κB pathway [115]. Although it has been documented that the pathological effects of ROS on inflammation are due to the activation of several pro-inflammatory signaling pathways such as mitogen-activated protein kinases (MAPK) [105], recently shown that some prenylated flavonoids induce the expression of HO-1 activating Nrf2 through the p38 MAPK pathway [113,116,117].

NF-κB is a transcription factor that plays an important role in the transcription of genes, which are involved in immune and inflammatory responses. It was recently observed that Toll-like receptor 7 contributes to neuropathic pain by activating NF-κB in primary sensory neurons and subsequently induced the release of inflammatory mediators in immune cells [118]. NF-κB is released from a complex with I-κB and migrates to the nucleus where it binds to the κB enhancer element and induces transcription of its target genes, such as COX-2, iNOS, TNF-α, IL-1β, and IL-6, chemokines and adhesion molecules [27,114]. Treatment with an *E. speciosa* methanol extract (ESLE) induced a significant reduction in the immunoexpression of NFκB, COX-2, iNOS, and the pro-inflammatory marker, TNF-α in doses of 50–100 mg/kg. Additionally, it increased the levels of GSH and catalase. This study concludes that ESLE exerted a gastroprotective effect through the synergistic anti-inflammatory and antioxidant activity of its various compounds, such as flavonoids (orientin, isoorientin, vitexin, isovitexin, and luteolin), alkaloids (hypaphorine), and saponins [88]. It has been seen that an oxidative environment produces a positive regulation of cytokines and macrophages, increasing the inflammatory response mediated by COX-2. The cytokines released can act directly on the primary afferent sensory neurons increasing the permeability of sodium (Na+) sensitizing the capsaicin receptors (TRPV1). These effects facilitate peripheral sensitization. Likewise, the oxidative environment can activate brain regions that transmit pain signals from other brain regions, which are responsible for nociceptive processing, such as the rostral ventrolateral medulla (RMV) [119]. The methanolic extract from the leaves of *E. senegalensis* has recently been seen, it showed a protective effect against oxidative stress in SC-1 fibroblasts and THP-1 macrophages. Observing the highest antioxidant activity against free radicals (IC50 = 44.86 µg/mL [ABTS]; 291.1 µg/mL [DPPH]) and intracellular ROS induced by 2,2′-Azobis (2-amidinopropane) dihydrochloride (AAPH) in macrophages. This effect was comparable to that of the positive control, Trolox [70]. The presence of compounds such as kaempferol, rutin, and rotenone is suggested as possible, responsible for the effect, however, confirmation of the finding is required [85]. However, neobavaisoflavone (Figure 1), an isoflavone previously isolated
from the root of *E. senegalensis* and *E. excelsa* [120] and found in *Psoralea corylifolia* L., has been shown to have significant anti-inflammatory and antioxidant activity [121,122].

The sources that precede the generation of reactive oxygen species (ROS) can be both endogenous and exogenous. B (XO) located on the outer surface of the plasma membrane and in the cytoplasm. It catalyzes the oxidation of hypoxanthine to xanthine and then to uric acid. The production of xanthine and XO is greatly increased during ischemia, accompanied by the loss of antioxidant enzymes. O$_2^-$ is an electron acceptor and XO cofactor, thus generating O$_2^-$ and H$_2$O$_2$, one of the main ROS in ischemia, causing damage to ischemic cells and different tissues [123]. Gout, a common and complex form of arthritis that can affect anyone, is characterized by sudden and intense attacks of pain, swelling, redness, and tenderness in the joints. Its treatment consists of increasing uric acid excretion or reducing uric acid production [21]. Xanthine oxidase inhibitors (XOI) are particularly useful, as they have minor side effects compared to uricosuric and anti-inflammatory agents (NSAIDs and corticosteroids). However, allopurinol, the only clinically used XOI, also causes side effects such as rash, low blood cells, and hypersensitivity syndrome. In this context, it has been shown that the chloroform fraction of *E. stricta* leaves inhibits xanthine oxidase in a concentration-dependent manner. Although in vitro inhibition was moderate compared to allopurinol, at higher doses (21.2 y 100 µg/mL) XO was significantly inhibited. It was suggested that the presence of phenolic and flavonoid content in the extract contributed to the inhibition of XO [70]. For their part, [53], concluded that the methanolic extract of the stem bark of *E. indica* has a strong activity in the inhibition of XO. Additionally, it has been shown that the methanol extract of the root has an important antioxidant activity and a strong inhibition of NO [54]. Some of the compounds isolated in this species include isoflavones such as genistein, wighteone, alpinumisoflavone, dimethylalpinumisoflavone, 8-prenilerithrinin C, and erisenegalensein E, from stem bark [55], compounds that may be responsible for the effect. Mildbone and mildbenone, flavanone and chalcone from *E. mildbraedii* exhibited significant antioxidant and moderate LOX inhibition activities [55,56] participates in the eicosanoid cascade during the inflammatory response, using arachidonic acid as a substrate, for the synthesis of leukotrienes (LT) and other oxidized lipids intermediates [124]. So, these compounds acted as an anti-inflammatory. At least three inhibitory mechanisms of LOX have been recognized, among these, some compounds act on an essential iron atom at the site, affecting its oxidation state (redox inhibitors) and binding directly to the iron atom (chelators), affecting the cycle catalytic. This non-selective antioxidant mechanism [124] may be associated with the antioxidant capacity of *E. mildbraedii* compounds and be responsible for the observed effect. The authors suggest that the high antioxidant activity of mildbone is mainly related to the second bond dissociation enthalpy (BDE) of a second hydrogen atom transfer from i-OH phenoxyl radical to the free radical [55]. The BDEs of the respective phenol bonds correlate with the antioxidant effect of these compounds. It was found that phenols with low BDE values lead to clearly higher stabilizations [125]. Mildbone with lower IC50 and BDE had greater antioxidant effects than mildbenone [55]. These values are generated by group Electrondonators (EDG). Hydroxy groups (–OH) represent EDG and therefore lead to lower BDE values. This makes it clear that high antioxidant activity is mainly related to the number and positions of OH groups located in ring B [125,126].

Prenylated flavonoids are a subclass of flavonoids, combining a flavonoid skeleton with a lipophilic prenyl side chain. They are compounds of low abundance in nature. Prenylation provides these compounds with improved biological activity. Increases the lipophilicity of flavonoids, providing a greater affinity for biological membranes and a better interaction with target proteins [127,128]. It was found that pterocarpan erycristagallin decreased the edema induced by phospholipase A2 by 51% in the first 30 min, however, the effect disappeared 60 min after application. This suggests that the effect may be an indirect inhibition of the enzyme. In rat peritoneal leukocytes, the application of erycristagallin inhibited the production of leukotriene B4 (metabolite 5-lipoxygenase). This
compound showed purifying properties, inhibiting the stable free radical DPPH by 96% at a concentration of 50 AM, while the reference drug quercetin produced inhibition of 92% at the same dose. Previous studies have suggested that different antioxidant agents and free radical scavengers can reduce 5-lipoxygenase activity through a mechanism that interferes with divalent ions involved in catabolism of arachidonic acid [129,130]. Besides, some prenylated flavonoids have been shown to have the ability to inhibit COX or LOX activity, depending on substitution patterns. Sigmoidins A and B, prenylated flavanones from *E. sigmoidea*, contain a catechol group in ring B and a 2′,5-diphenyl group (Sigmoidin A) or a 2′-prenyl group (sigmoidin B). At a concentration of 100 nm, sigmoidin A clearly inhibited leukotriene B4 production in rat polymorphonuclear leukocytes by 100%, while the same dose of sigmoidin B only reduced production by 44%. In PLA2-induced paw edema, sigmoidin B showed a clear inhibitory effect against the induction of edema (59% at 60 min) while sigmoidin A had only a mild effect at 30 min. Since sigmoidin B did not affect arachidonate metabolism, it was suggested that it affected histamine release [49]. Another prenylated flavonoid, the prenylisoflavone warangalone isolated from the bark of *E. addisoniae*, is a potent inhibitor of protein kinase A and showed marked anti-inflammatory efficacy on phospholipase A2-induced paw edema and 12-induced ear edema. O-tetradecanoylphorbol 13-acetate in mice [46]. Prenylated flavonoids have been reported to act as anti-inflammatory, through five mechanisms that include antioxidant and radical scavenging activities; regulation of the activities of cells related to inflammation; modulation of the activities of the enzymes of arachidonic acid metabolism (phospholipase A2, cyclooxygenase, lipoxygenase) and nitric oxide synthase; modulation of the production of other pro-inflammatory molecules and the modulation of the expression of pro-inflammatory genes [128,131].

At least 370 flavonoid compounds have been identified, in the genus *Erythrina*. Among them, several prenylated flavonoids [114,132–137] that due to their characteristic structures, have a better interaction with the target molecules. However, the role of these compounds in inflammatory pain has not been explored. Prenylated flavonoids have a very restricted application due to their lower abundance in the environment. However, the lack of preclinical studies on these compounds present in the species of the genus *Erythrina* indicates that most of the medicinal potential of this genus has not yet been exploited.

6. Methods
6.1. Search Strategy

An organized search for the ethnomedicinal use of the genus *Erythrina* in the treatment of pain and inflammation was conducted, as well as the preclinical studies performed.

The search was carried out systematically using MeSH (Medical Subject Headings) terms and “keywords”. First, we define the related MeSH terms: “Anti-inflammatory agents”, “analgesics/therapeutic use” “analgesics [Pharmacological action]”, “pain management”, “ethnobotany” “medical plant”, “ethnopharmacology” “antioxidant activity” and “flavonoids”, then each term was combined with *Erythrina*. Subsequently, keywords such as: “pain relief” “antinociceptive effect” “anti-inflammatory effect”, “ethnopharmacological studies” and “antioxidants” were combined with “Erythrina”.

6.2. Inclusion Criteria

All articles published from 2000 to May 2020, found in the scientific information sources ScienceDirect, Medline (Pubmed) and Springer link, were considered.

A selection of titles was made, from which the abstracts were read and those that met the necessary characteristics were retrieved. The following criteria were included for the selection of documents.

In the case of the ethnomedicinal reports, the documents that expose the use of the different parts of species of the genus *Erythrina* in the relief of pain and/or inflammation were selected.

In preclinical studies, studies that describe species of the genus *Erythrina* and models to evaluate analgesic and/or anti-inflammatory activity (in vivo and in vitro), including dose,
reference drug, possible mechanisms of action, as well as the main metabolites associated with the effect were selected.

Regarding antioxidant activity, articles were selected that mentioned the type of test used, the species studied and the type of extract and/or compounds evaluated. Items that did not meet the requirements were discarded.

7. Conclusions

Inflammatory pain, as a pathological phenomenon, has been established throughout history as a public health problem. There are several pro-inflammatory mediators involved in the complex web of the process. However, the data presented here show that the phytoconstituents of the genus Erythrina have the potential capacity to modulate different therapeutic targets and those effects could be associated with their antioxidant properties. Several compounds with antioxidant, analgesic, and anti-inflammatory effects have been identified. Among them, several prenylated flavonoids that, due to their characteristic structures, have a better interaction with the target molecules. However, the lack of preclinical studies on these compounds and the species of the genus Erythrina indicates that most of the medicinal potential of this genus has not yet been explored. Likewise, it is necessary to delve into the molecular mechanisms involved in both effects.

Author Contributions: T.J.-C. and M.D.I.O.-A., original draft preparation, drafted the figures and tables, performed bibliographic research; M.D.I.O.-A., C.V.-G., O.A.J.-M., J.A.G.-S. and T.A.U.-H. wrote the manuscript and formal analysis; M.B. and T.J.-C. conceived the topic, wrote the manuscript and retrieved the funding. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: One of the authors (T.J.-C.) is grateful for a scholarship provided by the National Council of Science and Technology of Mexico (CONACyT).

Conflicts of Interest: The authors declare that there are no conflicts of interest in the publication of this article.

Abbreviations

| Abbreviation | Description |
|--------------|-------------|
| ROS          | Reactive oxygen species |
| MAPK         | Mitogen-Activated Protein Kinase |
| API          | Activator protein 1 |
| NF-kB        | Nuclear factor kappa-light-chain-enhancer of activated B cells |
| ERK          | Extracellular signal-regulated kinases |
| JNK          | Jun N-Terminal Kinase |
| JAK          | Janus kinases |
| STAT         | Signal Transducer and Activator of Transcription |
| IASP         | International Association for the Study of pain |
| NSAIDs       | Nonsteroidal anti-inflammatory drugs |
| CNS          | Central Nervous System |
| NO           | Nitric Oxide |
| DPPH         | 2,2-diphenyl-1-picrylhydrazyl |
| BHA          | Butylated hydroxyanisole |
| NAChRs       | Nicotinic Acetylcholine Receptors |
| COX-2        | Cyclooxygenase 2 |
| iNOS         | Nitric oxide synthase, inducible |
| LOX          | Lipoxigenases |
| Nrf2         | Nuclear factor erythroid 2-related factor 2 |
| TNFα         | Tumor Necrosis Factor alpha |
| IL-1β        | Interleukin-1-beta |
| IL-6         | Interleukin-6 |
| IL-8         | Interleukin-8 |
| HO-1         | Heme Oxygenase-1 |
| GCLC         | γ-glutamyl cysteine synthetase catalytic subunit |
| NAD(P)H      | Nicotinamide adenine dinucleotide phosphate |
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