Effect of memantine on calcium signaling in hippocampal neurons cultured with β-amyloid

V.M. Shkryl, V.V. Ganzha, E.A. Lukyanetz

O.O. Bogomoletz Institute of Physiology, National Academy of Sciences of Ukraine, Kyiv; e-mail: elena@biph.kiev.ua

Alzheimer’s disease (AD) is the most common type of dementia and is characterized by accumulating amyloid (Aβ) plaques and neurofibrillary tangles in the brain. Excessive stimulation of glutamate receptors, mainly NMDA-type, causes intense entry of calcium ions into cells and is a key early step in glutamate-induced excitotoxicity, resulting in many neurological diseases, including AD. Memantine, an NMDA receptor antagonist, blocks NMDA receptors and reduce the influx of calcium ions into neuron. In our experiments, we have modeled AD on cultured rat hippocampal neurons to test the effects of memantine on calcium signaling in neurons. Our results show that the neuroprotective effect of memantine could be provided not only through the inhibition of NMDA receptor current but also through the suppression of voltage-dependent Ca\(^{2+}\) channels, most likely L-type. This study suggests that NMDA receptor antagonist memantine can protect hippocampal neurons from calcium overloading induced by Aβ1–42 amyloid exposure via blocking Ca\(^{2+}\) channels.

Keywords hippocampal neuron culture; Aβ-amyloid; NMDA; memantine; calcium; calcium homeostasis; Alzheimer’s disease.

INTRODUCTION

Alzheimer’s disease (AD) is clinically characterized by progressing cognitive impairment, memory loss, and behavioral changes. Currently, more than 26.6 million people worldwide suffer from this disease, and this number is growing sharply every year. By 2050, the number of patients is expected to quadruple to more than 106 million globally, and an estimated 1 from 85 people will be subject to the disease [1]. For this reason, both scientific and clinical studies covering all aspects of AD have become extremely relevant and have expanded greatly in recent decades. So far, the etiology of AD remains unknown. However, it was shown many factors had been involved in AD, and several hypotheses were proposed to explain the onset and progression of the neurodegenerative process observed in this disorder. One such hypothesis is “the amyloid hypothesis”, which supports the idea that accumulation of neurotoxic forms of beta-amyloid peptide (Aβ) leads to amyloid plaques, which play a special role in the pathogenesis of AD [2, 3]. Finally, Aβ plaques cause neuronal loss. Recent evidences suggest that soluble Aβ oligomers, rather than plaques, are a major cause of synaptic dysfunction leading to neurodegeneration. The soluble Aβ oligomer has been shown can interact with glutamatergic N-methyl-D-aspartate (NMDA) receptors and other proteins involved in maintaining glutamate homeostasis as uptake and release.

Calcium plays a particularly important role in neuronal cells, where it mediates numerous vital physiological processes and plays a central role in controlling synaptic plasticity [4]. For normal functioning, neurons require extremely precise control of Ca\(^{2+}\) concentration in certain compartments. One of the common factors underlying AD pathogenesis is a violation of the regulation of Ca\(^{2+}\) in neurons. Constant disturbances in Ca\(^{2+}\) signaling have significant consequences for neurons’ health and functionality throughout the organism’s life.
and form the basis of “the calcium hypothesis” in AD [5]. At rest, cytosolic Ca\(^{2+}\) is maintained at a low nanomolar concentration by pumps, buffers, and transport mechanisms. Ca\(^{2+}\) entry from the extracellular environment is carried out by activation of voltage-gated Ca\(^{2+}\) channels (VGCC), ionotropic glutamate channels (NMDA), and acetylcholine receptors [6]. Besides, intracellular depots – mitochondria and endoplasmic reticulum (ER) – regulate calcium in the cell [7-9].

It is known that memantine, a selective antagonist of NMDA-receptors, is used in medical practice for treatment of different types of dementia, including AD [10, 11]. We previously tested meantime action on memory tests in rats [12-15]. These experiments have shown that memantine markedly increases the fulfillment of a conditioned reflex in old and young rats, which indicated its effect on memory processes. Therefore, this study aimed to test memantine’s effects on calcium signaling in cultured hippocampal neurons treated with Aβ1-42 (AD modeling).

**METHODS**

All experimental procedures were performed following the European Commission Directive (86/609/EEC) and were approved by the local Animal Ethics Committee of the Bogomoletz Institute of Physiology (Kyiv, Ukraine). All efforts were made to minimize the number and suffering of animals used. Studies were performed on rat hippocampal culture neurons using a common technique described earlier [16, 17]. In our studies, we used a control set of cells, which were incubated in standard conditions – nutrient medium, which included 90% of the Minimum Essential Media (MEM, Sigma-Aldrich, USA), 2.2 g/l NaHCO\(_3\), 10% horse serum (Gibco, USA), 10 μg/ml insulin and antibiotics: 50 IU/ml benzylpenicillin sodium and 50 μg/ml streptomycin sulfate. Prolonged culturing of CNS cells in vitro is widely used for AD modeling [18, 19]. In our study, the model was obtained by 24-hour incubation of hippocampal culture neurons with Aβ1-42 (Sigma-Aldrich, USA) at a final concentration of 2 μM.

We used the electric field stimulation (EFS) of neurons to depolarize cell membrane and fill endoplasmic reticulum with Ca\(^{2+}\). Field stimulated Ca\(^{2+}\) transients were induced with a standard protocol by lowering two parallel platinum electrodes (20÷25 mm apart) into the chamber in near contact with the surface and then passing 15 current pulses at 15 Hz of 1 msec between the electrodes, yielding fields of \(\sim 12\div 15\ \text{V/cm}\) across the surface of the coverslip [20]. The resistance between medium and cells is negligible as electrodes are not in direct contact with cells, and EFS is delivered to cells through the medium.

To measure intracellular calcium concentration, we used fluorescent microscopy and calcium-sensitive dye Fura-2 AM as was described previously [21, 22]. Before the experiment, neurons were stained in a solution of Fura-2 AM dye (5 μM) for 30 min. Coverslips with neurons were then washed and placed in the chamber with the experimental solution included in mM: NaCl – 140.0; KCl – 2.0; CaCl\(_2\) – 2.0; MgCl\(_2\) – 2.0; HEPES – 10.0; pH = 7.4. To induce Ca\(^{2+}\) transient we used depolarization solution, its composition was equivalent to bath solution described above except that 50 mM of NaCl was replaced by KCl. Using a digital video camera, we recorded changes in the level of fluorescence intensity of Fura-2 dye in the soma of neurons at 340 and 380 nm excitation. Cell M software (Olympus, Japan) was used for data collection, and data analysis was performed using the IDL programming environment (ITT Visual Information Solutions). We performed further data analysis, and the ratio of 340 to 380 nm fluorescence intensity (ratio; F340/F380) was calculated. We estimated the data according to the protocol described in [23] to calculate F340/F380 by subtracting the resulting background level (calculated outside the cells). Dynamic changes in this indicator assessed changes in the level of free calcium in neurons’ cytosol.
The obtained results were processed using variation statistics using Origin software (Microcall Inc., USA). Numerical data are given as mean ± mean error. The data were tested to be normally distributed. Intergroup comparison was performed using analysis of variance (ANOVA). If intergroup differences were found, the Tukey test was used. The results were considered statistically significant at $P < 0.05$.

RESULTS

In the first series of experiments, we used hippocampal culture neurons in control conditions. We used the protocol of experiments to apply 3 EFS stimuli (1 s duration of each) with 3 s intervals between them, Fig. 1A. These were necessary to fill intracellular depots. After that, we applied caffeine (10 mM) for 5 s to induce $Ca^{2+}$ release from the ER stores by ryanodine receptors (RyRs) and estimate their contribution to the $Ca^{2+}$ signaling of these cells. Accordingly, after the next three-time EFS, we applied a high KCl solution (50 mM) to depolarize the membrane and induce $Ca^{2+}$ transients evoked by the activity of voltage-operated $Ca^{2+}$ channels, Fig. 1A.

Further, we conducted the same set of the protocol but in the presence of 20 μM memantine to test its effect on the mentioned participants of the $Ca^{2+}$ signaling in the same tested neurons. As it can be seen, the application of memantine in extracellular solution led to a decrease in calcium loading in endoplasmic depot and decreased the amplitude of calcium transient during depolarization of the membrane by KCl but slightly influenced the basal free calcium.

In the next series of experiments, we used the same protocol but tested neurons, which were incubated 24 h before the experiments with...
5 μM Aβ1-42. An example of such registrations is presented in Fig. 1B. We first measured calcium signal changes before memantine was added to the external solution and then measured from the same cell in the presence of 20 μM of this reagent. Such a form of experiment reduced the differences between the registrations’ conditions and allowed a convincing study of memantine’s effect. As can be seen from the presented registrations of calcium signal changes, 20 μM memantine induced a significant decrease in values of Ca²⁺ transients evoked by 5 s depolarization (50 mM KCl) and 5 s 10 mM caffeine application compared to the control.

Statistics data of observed effects are presented in Fig. 2. Thus, the ratio value of basal level of free calcium under control conditions was 1.05 ± 0.07 (n = 15) and 0.95 ± 0.02 (n = 14) in the presence of 20 μM memantine. These values in the neurons which were previously incubated for 24 h with 2 μM Aβ1-42 in culturing medium, were 0.97 ± 0.04 (n = 20) and 1.01 ± 0.03 (n = 16) in the presence of 20 μM memantine.

The level of free calcium (in ratio value) before the second Ca²⁺ transient caused by EFS under control conditions was 1.07 ± 0.04 (n = 15) and 1.02 ± 0.02 (n = 14) in the presence of 20 μM memantine. These values in the cells, which were previously incubated for 24 h in culture medium with 2 μM Aβ1-42 (Aβ), were 1.17 ± 0.07 (n = 20) and 1.07 ± 0.04 (n = 16) in

---

*Fig. 2. Diagrams represent statistical data of memantine’s effect on the changes in the level of intracellular calcium concentration in value of ratio. Part A is mean values of the basal Ca²⁺ level before induction of the second Ca²⁺ transient caused by EFS; part B is the mean values of the peak of the second Ca²⁺ transient caused by EFS; part C is the mean values of the peak of signal evoked by 5 s application of 10 mM caffeine; part D is the mean values of the peak of the Ca²⁺ transient evoked by the application of the depolarizing solution (50 mM KCl; 5 s). For all cases, data represent values in control conditions (control), during application of 20 μM memantine (mem) and for case 24 h preincubation of neurons with 2 μM Aβ1-42 (Aβ). *P < 0.05
the presence of 20 μM memantine, as shown in the diagram 2A.

Memantine almost unchanged the peak value of Ca\(^{2+}\) transients caused by EFS. Thus, the mean peak value of Ca\(^{2+}\) transients was reduced from 1.69 ± 0.04 (n = 15) in control to 1.64 ± 0.08 (n = 14) after memantine application. Under β-amyloid treatment, this parameter was significantly reduced from 1.56 ± 0.05 (n = 19) in control conditions to 1.39 ± 0.02 in the presence of memantine (n = 16, P < 0.05). The data are presented in Fig. 2B.

Statistical data concerning RyRs endoplasmic depo involvement in Ca\(^{2+}\) responses are presented in Fig. 2C. We used caffeine, an agonist of RyRs, to induce Ca\(^{2+}\) release from the endoplasmic reticulum, which can appear as Ca\(^{2+}\) transient during registrations. As shown in Fig. 2C, the peak values of calcium signals induced by 10 mM caffeine application were significantly reduced in the presence of 20 μM memantine in comparison with control values. They decreased from 1.56 ± 0.24 (n = 14) in control to 1.24 ± 0.12 (n = 12, P < 0.05) after memantine action. In the neurons, which were previously treated with β-amyloid, this value was reduced from 2.16 ± 0.36 (n = 17) in control to 1.57 ± 0.25 (n = 16, P < 0.01) after memantine application.

We also calculated statistical data concerning the peaks of Ca\(^{2+}\) transients induced by 5 s depolarizing solution (50 mM KCl) applications. They also were significantly reduced from 3.71 ± 0.34 (n = 15) in control to 2.95 ± 0.16 (n = 14, P < 0.05) after 20 μM memantine applications. In the neurons previously incubated with Aβ1-42, this parameter was also noticeably decreased by memantine, Fig. 2D. These values were reduced from 5.21 ± 0.35 (n = 19) in control to 4.02 ± 0.36 (n = 16, P < 0.05) after memantine application.

DISCUSSION

Glutamate is the major fast excitatory neurotransmitter and is involved in almost all CNS functions, especially in the cortex and hippocampus: 70% of all excitatory synapses in the CNS use glutamate as a neurotransmitter. Ionotropic glutamate receptors, responsible for rapid neural communication at excitatory synapses, contain three subfamilies: α-amino 3-hydroxy 5-methyl 4-isoxazole-propionic acid (AMPA), kainate receptors, and NMDA receptors. Among them, NMDA receptor ion channels are the most permeable to Ca\(^{2+}\), which can, in turn, function as a second messenger in different signaling pathways [24].

The hippocampus has a high density of glutamate NMDA receptors and is extremely important for learning and memory. Normally, they are activated only in certain physiological processes, such as the induction of synaptic plasticity, and are capable of long-term potentiation of synaptic transmission, which is considered one of the key mechanisms for providing higher functions: learning, memory, behavioral and other reactions.

NMDA receptor has been implicated as a mediator of neuronal damage associated with many neurological disorders, including ischemia, epilepsy, brain injury, dementia, and neurodegenerative disorders such as AD. Pathological elevations in glutamate levels and other disturbances that alter the resting membrane potential (e.g., metabolic disturbances) may cause over-stimulation of NMDA receptors, which induces excitotoxicity and promotes cell death. This process underlies the potential mechanism of neurodegeneration that occurs in AD.

Under normal synaptic transmission, NMDAR channels are blocked by Mg\(^{2+}\) inside the channel and are activated only for a short period. However, under pathological conditions, the normal Mg\(^{2+}\) block of the ion channels is removed and abnormally enhances the activity of NMDA receptors. Excessive receptor activation leads to an excessive influx of Ca\(^{2+}\) into the neuron, which then triggers various processes that can lead to necrosis or apoptosis [25]. Various pathologies, such as Aβ oligomer action,
oxidative stress, mitochondrial dysfunction, elevated glutamate concentrations, and neuronal inflammation, have been associated with hypersensitivity and activity glutamergic system, leading to neuronal dysfunction and cell death in AD [26].

Memantine, a non-competitive NMDA receptor antagonist, blocks the NMDA ion channels with fast kinetics. Complete depolarization of the membrane leads to memantine removing from the channel, which provides normal synaptic transmission. It is suggested that these properties underlie memantine’s ability to ensure the receptor’s normal physiological functioning while disrupting receptor pathological activation. It has also been shown that NMDA receptor blockade alleviates Aβ-induced degeneration in rat hippocampal neurons [24]. Preclinical data suggest that NMDA receptor-mediated excitotoxicity may be associated with the effects of abnormal Aβ deposition during AD.

In our experiments, we tested the effects of memantine on the components of calcium intracellular signaling, such as basal Ca$^{2+}$ level in the cytoplasm, changes in Ca$^{2+}$ transients evoked by membrane depolarization with EFS, activation of voltage-depended Ca$^{2+}$ channels by high KCl solution and Ca$^{2+}$ release from ryanodine receptors of the endoplasmic reticulum. As well, we estimated these parameters in neurons, which were treated with Aβ1-42 (cell model of AD). It is proved that Aβ1-42 increased all tested parameters in our experiments except for EFS’ amplitude, which had a tendency to be decreased, Fig. 2. Thus, the ratio ($F_{340}/F_{380}$) of basal level of Ca$^{2+}$ increased by 8.7%, the peak of KCl-transient – 40.32%, caffeine peak – 40.2%, whereas the EFS peak was slightly decreased by 7.7%. These data show that Aβ1-42 significantly affects membrane voltage-dependent channels, increasing the flow of calcium into the cell and, consequently, more filling of the ER-depot and the release of calcium from it into the cytoplasm during RyRs agonist action.

Our experiments have shown that memantine decreased all tested indicators in control conditions and neurons treated with the Aβ1-42 (AD model) but with different degrees. It only tended to decrease the basal level, and this value was in the range not exceeding 8%. Also, the effect was weakly expressed in the case of EFS, it did not exceed 3% in control and was more noticeable in cells treated with Aβ1-42 (~11%). The most pronounced effect of memantine was found in the case of caffeine and KCl applications. It was 20.5% in the case of ER-depots and 20.7% in the case of Ca$^{2+}$-channels. In the cells treated with Aβ1-42, these values were some more – 28.2 and 22.8% correspondingly, Fig. 2C, D.

Interestingly, the effects of Aβ1-42 and memantine on calcium peaks at EFS were significantly less pronounced than those at the depolarization of membrane by KCl. It was previously shown that EFS could affect calcium channels, Ca$^{2+}$-stores, stretch channels, Na$^{+}$ channels [27]. It was also known that EFS is better to use mainly for modulation of electrosecretory coupling at the nerve endings where it can be better investigated by evoking synaptic release with electrical pulses. We suppose that EFS evoked electrical pulses are very short in duration to produce remarkable membrane depolarization. Our experiments show that EFS is not well suited for studying the chemical compound effects, but it can be successfully used mainly to fill ER depots with Ca$^{2+}$.

Although it is known that memantine is an antagonist of NMDA receptors, our research has shown that it can partially block L-type calcium channels of hippocampal neurons in the concentration of 20 μM. This observation is in line with data obtained on a single retinal ganglion cell of adult frogs Rana temporaria [28]. In these experiments, the authors have shown that memantine (30±45 μM) largely inhibited the L-type Ca$^{2+}$ channels. Their results suggested that memantine’s neuroprotective effect could be provided not only through the inhibition of NMDA receptor currents but also through suppressing L-type channels. Our results obtained on rat hippocampal neurons completely support
The above-mentioned conclusion that memantine also suppresses voltage-dependent Ca\(^{2+}\)-channels, which are most likely of L-type.

Acknowledgments. This work was supported from funds of National Academy of Sciences of Ukraine to support the development of priority areas of research SRN (state registration number) 0118U007344, 0120U001281 and 0116U004470.

The authors of this study confirm that the research and publication of the results were not associated with any conflicts regarding commercial or financial relations, relations with organizations and/or individuals who may have been related to the study, and interrelations of co-authors of the article.

REFERENCES

1. Prasansuklab A, Tencomnao T. Amyloidosis in Alzheimer’s disease: The toxicity of amyloid beta (Aβ), mechanisms of its accumulation and implications of medicinal plants for therapy. Evid Based Complement Alternat Med. 2013;2013:413808.

2. Tyshchenko YN, Lukyanetz EA. The role of beta-amyloid in norm and at Alzheimer’s disease. Fiziol Zh. 2020;66(6):88-96.

3. Kravenska Y, Nieznanska H, Nieznanski K, Lukyanetz E, Szewczyk A, Koprowski P. The monomers, oligomers, and fibrils of amyloid-β inhibit the activity of mitoBKCa channels by a membrane-mediated mechanism. Biochimica et Biophysica Acta (BBA) - Biomembranes. 2020;1862(9):183337.

4. Pchitskaya E, Popugaeva E, Bezprozvanny I. Calcium signaling and molecular mechanisms underlying neurodegenerative diseases. J Biol Chem. 2018;70:87-94.

5. Khachaturian ZS. Hypothesis on the regulation of cytosol calcium concentration and the aging brain. Neurobiol Aging. 1987;8(4):345-6.

6. Demuro A, Parker I, Stutzmann GE. Calcium signaling and amyloid toxicity in Alzheimer disease. J Biol Chem.
Effect of memantin on calcium signaling in hippocampal neurons cultured with β-amyloid