Mycoplasma pneumoniae (MP) is the most common cause of childhood bacterial pneumonia. Although macrolide is known to be effective as a first-line therapy, the proportion of macrolide resistance in MP pneumonia has strikingly increased during recent 2 decades in East Asia. This is challenging to physicians since they have to decide more often whether to use secondary treatment. Diagnostic methods to detect macrolide-resistance of MP are currently not available in Korean hospitals. Even in the diagnosis of MP infection, both serologic and molecular test have limitation: inability to differentiate current illness from carriage or asymptomatic infection. Combining these 2 diagnostic methods and excluding infection caused by other respiratory pathogens allow a more reliable diagnosis. This effort is even more demanding in recent years to keep children from unnecessary exposure to secondary antibiotics. Although several observational studies have reported that tetracycline and fluoroquinolone, which are considered in the treatment of refractory MP pneumonia, have efficacy of shortening the duration of fever and respiratory symptoms, those findings need to be proven by well-designed prospective studies. The use of tetracycline and fluoroquinolone in children is generally tolerable, as supported by many observational data. However, since concerns about side effects still remain, careful consideration about benefits and risks is needed to decide their use.

**Key words:** Mycoplasma pneumoniae, Child, Fluoroquinolone, Tetracycline

**Introduction**

Mycoplasma pneumoniae (MP) is the most common cause of childhood bacterial pneumonia, and is a major consideration when choosing empirical antibiotics for treatment of community acquired pneumonia in children.1) Epidemics usually occur in the endemic area every 3–7 years,2) especially common in East Asia. In Korea, the recent epidemics of MP pneumonia were in 2011 and 2015.3) Mycoplasmas lack cell walls and are intrinsically resistant to antimicrobial agents that target cell wall synthesis, such as β-lactams.4,5) Macrolide and tetracycline, which inhibit protein synthesis by acting on bacterial ribosomes, and fluoroquinolone, which inhibits bacterial DNA replication, are intrinsically active against MP. Macrolide is primarily used for the treatment of children due to the potential toxicity of tetracycline and fluoroquinolone in young age.6) MP infection is known to respond well to macrolide antibiotics. Recently, however, there has been an increased incidence of MP infections caused by macrolide-resistant bacteria, which is associated with the development of severe progressive pneumonia.

Although the use of secondary antibiotics (tetracycline and fluoroquinolone), corticosteroids, or immunoglobulin has been considered as an alternative treatment for macrolide-resistant cases,7,8) there have been some concerns about their efficacy and safety. In the current situation where there are limitations to the diagnosis of macrolide-resistant MP infections or even MP infections, there is an increasing concern about overuse and misuse as the use of these drugs increases during the epidemic.

In this review, we will discuss what to consider when diagnosing refractory MP pneumonia, and the efficacy and safety of second-line drugs for the proper use of second-line therapy.

**Macrolide-resistant MP and recent epidemiology**

MP has macrolides-binding sites to nucleotides in domains II and/or V of 23S ribosomal RNA (rRNA) in the 50S bacterial ribosomal subunit. Macrolide blocks the growth of the peptide chain at the early stages of protein synthesis and inhibits the
assembly of new large ribosomal subunits, resulting in gradual depletion of functional ribosomes in the cell. Several point mutations (positions 2063, 2064, 2067, and 2617 in the peptidyl transferase loop of 23S rRNA) and insertions or deletions in ribosomal proteins L4 and L22 are involved in macrolide resistance in MBs. The most common transition mutation is A2063G followed by A2064G. The MP strains that carry the A2063G or A2064G mutation at the domain V of 23S rRNA exhibited very high minimal inhibitory concentrations (MICs) against all macrolides.

The macrolide resistance of MP, which has emerged from the 2000s, gradually increased in frequency and reached more than 90% in 2011 depending on the region (Table 1). In Korea, it was reported that clonal expansion of P1 type 1 and sequence type (ST) 3 MP with macrolide resistance contributed to the spread. Similar patterns have been observed in neighboring countries, Japan and China, that P1 type 1 and ST3 or multiple-locus variable-number tandem repeat analysis type 4-5-7-2 has predominated in MP infection and has contributed to the spread. In the United States has been gradually rising.9,16,17) The macrolide resistance rate still remains low, but the resistance rate in Europe and America, where the incidence of MP is not high, the macrolide-resistance rate still remains low, but the resistance rate in the United States has been gradually rising.9,16,17)

Why should we distinguish between macrolide-resistant MP and refractory MP infection?

1. Difficulties in laboratory diagnosis of macrolide-resistant MP infection

The diagnosis of macrolide-resistant MP infection can be made by measuring the MIC of strains isolated by culture or by detecting the point mutation of 23S rRNA by polymerase chain reaction (PCR). The MICs of each antimicrobial can be determined by a microdilution method. The criteria for drug-resistant MP, proposed by the Japanese Society for Mycoplasmology, is MICs of ≥16 µg/mL for erythromycin, clarithromycin, and azithromycin. Among the 36 isolates of macrolide-resistant MP from Korean children, 2010–2015, the MIC50/MIC90 values for erythromycin, clarithromycin, and azithromycin were 16/128 µg/mL, 64/128 µg/mL, and 8/16 µg/mL, respectively, which were higher than those for 23 macrolide-susceptible MP isolates, with MIC50/MIC90 values of 0.001/0.002 µg/mL for erythromycin, 0.001/0.002 µg/mL for clarithromycin, and 0.001/0.001 µg/mL for azithromycin. In a recent study on MP isolates retrieved from Japanese children, 2011–2016, MIC50/MIC90 of erythromycin, clarithromycin, and azithromycin for macrolide-resistant MP was >128/128, >128/128, and 32/64 µg/mL, respectively, while those for

| Nation       | Study          | Study period | Cases studied | Macrolide resistance (%) | ST/MLVA type (%) | P1 type | Identified mutations among MRMP (%)                |
|--------------|----------------|--------------|---------------|---------------------------|------------------|---------|---------------------------------------------------|
| China        | Guo et al.62)  | 2014         | 341           | 69.2                      | ND               | Type 1, 87.7%; type 2, 12.3% | A2063G (58.4%), A2064G (7.3%) |
|              | Qu et al. 63)  | 2010-2012    | 88            | 94.3                      | M4-5-7-2 (86.4%), M3-5-6-2 (9.1%); M4-5-7-2 predominated in MRMP | Type 1, 92.0%; type 2, 8.0%; type 1 predominated in MRMP | A2063G (100%) |
|              | Xue et al.64) | 2016         | 186           | 76.3                      | M4-5-7-2 (72.1%), M3-5-6-2 (22.1%); M4-5-7-3 (4.1%); M4-5-5-2 (12.2%); M3-6-6-2 (0.6%) | Type 1, 78.6%; type 2, 21.4% | A2063G (74.1%), mixed (2.7%) |
|              | Yan et al.65) | 2014-2015    | 200           | 90.5                      | M4-5-7-2 (87.5%), M3-5-6-2 (10.0%); M4-5-6-2 (1.0%); M4-5-7-3 (1.0%); M4-5-5-2 (0.5%) | Type 1, 89.5%; type 2, 10.5% | A2063G (89%), A2064G (1%); A2063T (0.5%) |
|              | Yan et al.66) | 2013-2017    | 423           | 91.0                      | M4-5-7-2 (87.5%), M3-5-6-2 (11.3%); M4-5-6-2 (0.5%); M4-5-7-3 (0.5%); M4-5-5-2 (0.2%); M3-5-6-2 increasing across the study period; M4-5-7-2 predominated in MRMP | Type 1, 88.2%; type 2, increasing across the study period | Not specified |
|              | Yuan et al.67) | 2016         | 120           | 90                        | ND               | ND      | A2063G (90%)                                     |
|              | Zhao et al.68) | 2014-2016    | 81            | 65.4                      | M4-5-7-2 (51.9%), M3-5-6-2 (39.5%) | Type 1 associated with MR; type 2 increasing, but MRMP proportion of type 2 isolates also increasing | A2063G (98.1%), A2064G (1.9%) |
|              | Zhao et al.69) | 2015–2018    | 1,045         | 29.4                      | ND               | ND      | Not specified                                    |
| Nation           | Study                  | Study period | Cases studied | macrolide resistance (n) | Prevalence of macrolide resistance (%) | ST/MLVA type (%) | P1 type | Identified mutations among MRMP (%) |
|------------------|------------------------|--------------|---------------|--------------------------|----------------------------------------|------------------|---------|------------------------------------|
| Japan            | Akashi et al.          | 2016–2017    | 222           | 65.3                     | ND                                     | ND               | ND      | Not specified                      |
|                  | Ando et al.            | 2002-2016    | 417           | 7-23 (2002-2005)/41-57 (2006-2010)/66-73 (2011-2012)/76 (2015-2016) | Prevalence not specified MRMP: ST3 (74.6%), ST19 (11.4%) NSMP: ST3 (52.6%), ST14 (28.4%) | ND               | A2063G (88.1%), A2063T (5.4%), A2064G (5.4%), A2063C (0.6%), C2617A (0.6%) |
|                  | Katsukawa et al.       | 2011-2017    | 419           | 71-82 (2011-2014)/21-50 (2015-2017) | ND                                     | Type 1 predominated in 2011–2014, type 2 increased from 2015 | A2063G (49.9%) |
|                  | Kawasaki et al.        | 2015-2016    | 137           | 58.4                     | ND                                     | ND               | A2063G (98.7%), A2064G (1.3%) |
|                  | Kenri et al.           | 2006-2019    | 554           | 0 (2006-2007)/92.2 (2012)/2.6 (2018)/40 (2019) | Type and prevalence not defined | Type 1 (>80%) in 2011–2012, but type 2 increased in 2015-2016, dominated after 2017 | ND |
|                  | Morozumi et al.        | 2002-2019    | 105           | 11.3                     | ST3 and ST14 dominated during 2002-2016, ST7 and ST33 dominated during 2018-2019 | Type 2 (ST33, ST34, ST7) dominated during 2018-2019, type 1 (ST3, ST17, ST19) dominated during 2012-2016 | A2063G (100%, 2018-2019) |
|                  | Suzuki et al.          | 2016-2017    | 25            | 28                       | ND                                     | Type 2, 52.2% (no MRMP) | ND      | A2063G (98.3%), A2064G (1.7%) |
| Republic of Korea| Lee et al.             | 2000-2016    | 146           | 3.4 (2006-2007)/54.1 (2010-2012)/84.4 (2014-2016) | ST3 (74.7%), ST14 (15.1%) | Type 1 (70.6%), type 2 (29.4%); type 1 has decreased (75% [2006-2007] to 50% [2014-2016]) | A2063G (94.7%) |
|                  | Lee and Lee            | 2019-2020    | 148           | 61.5                     | ND                                     | ND               | A2063G (98.3%), A2064G (1.7%) |
| Taiwan           | Hung et al.            | 2017-2019    | 226           | 77                       | MSMP: ST14 (53.8%), ST17 (28.8%), ST3 (11.5%), MRMP: ST3 (50%), ST17 (39.7%), ST14 (6.9%) | Type 1 dominated in MRMP, Type 2 dominated in MSMP | A2063G (82.8%), A2063T (16.1%), A2064G (1.1%) |
|                  | Lu et al.              | 2016         | 180           | 24                       | M4-5-7-2 (52%, associated with MRMP), M4-5-7-3 (27%), M1-5-6-2 (15%) | ND               | A2063G (86%) |
|                  | Yang et al.            | 2010-2017    | 471           | 24                       | ND                                     | ND               | A2063G (91%) |
| Germany          | Dumke and Ziegler      | 2016-2018    | 166           | 3                        | ND                                     | ND               | A2064 |
| Russia           | Voronina et al.        | 2015-2018    | 117           | 7                        | M3-5-6-2 (56.8%), M4-5-7-2 (43.2%) | ND               | A2063G (7%) |
| Slovenia         | Kogoj et al.           | 2006-2015    | 872           | 0.8                      | M3-5-6-2, M3-6-6-2 predominated in 2009-2011 (77.0%), M4-5-7-2 in 2013-2015 (71.6%) | Type 2 predominated → type 1 predominated after 2013 | Not specified |
| Spain            | Rivaya et al.          | 2013-2017    | 138           | 8                        | M4-5-7-2 (50.1%), M3-5-6-2 (29.2%) | ND               | Not specified |
| Sweden           | Gullby et al.          | 1996-2017    | 578           | 0.2                      | M3-5-6-2 (42.4%), M4-5-7-2 (37.4%), M3-6-6-2 (14.9%), M4-5-7-2 predominated in 2016-2017 | Type 2 (57.8%), type 1 (42.2%); type 1 predominated in 2016-2017 | A2063G |
| Switzerland      | Wagner et al.          | 2014-2017    | 163           | 9                        | ND                                     | ND               | A2063G (60%), A2064G (33.3%) |
| USA              | Waites et al.          | 2015-2018    | 306           | 6.9                      | ND                                     | ND               | A2063G (81.5%), A2064G (14.8%) |
|                  | Xiao et al.            | 2012-2018    | 446           | 8.3                      | M3-5-6-2 (41.7%), M4-5-7-2 (35.3%), M3-6-6-2 (16.6%) | Type 2 (59.8%), MRMP proportion 12.9% in type 1 and 5.5% in type 2 | A2063G (86.5%) |
| Colombia         | Copete et al.          | 2011-2013    | 42            | 0                        | ND                                     | Type 2 (89.3-96.1%) | Not specified |
| South Africa     | Carrim et al.          | 2012-2015    | 33            | 0                        | M3-5-6-2 (44%), M3-6/6/2 (38%), M4-5-7-2 (18%) | Type 2 (61%), type 1 (29%) | Not specified |

MRMP, macrolide-resistant *Mycoplasma pneumoniae*; MSMP, macrolide-susceptible *Mycoplasma pneumoniae*; ST, sequence type; MLVA, multiple-locus variable-number tandem repeat analysis; ND, not done.

*Adults' data included.*
macrolide-sensitive MP was 0.004/0.008, 0.002/0.004, and 0.00025/0.0005, respectively. However, this test is not useful to clinical practice because it requires long time (>2 weeks) for bacterial growth.

The PCR method can be performed to identify macrolide resistance using cultured MP isolates or DNA extracted from nasopharyngeal samples. The PCR products can be sequenced to identify the transitions in domain V of the 23S rRNA gene that are associated with macrolide resistance. Multiplexed PCR test has been widely used to detect MP from respiratory samples in many hospitals in Korea, but the sequencing test to identify macrolide resistance is not available in clinical practice. In Japan, it is recommended that clinicians should assess the efficacy of macrolides and consider to collect samples available to clarify the susceptibility of the causative organism in the absence of defervescence 48–72 hours after the start of macrolide therapy. It is needed to introduce clinically available tests to detect macrolide resistance of MP.

2. Uncertain impact of macrolide resistance on clinical course

Since the first macrolide resistance of MP was isolated from clinical samples obtained from children with pneumonia in Japan in 2000, it has been suggested that macrolide resistance can be associated with severe respiratory symptoms in children. However, the clinical relevance of resistant strains is largely debated, because it is not clarified whether resistant strains are related with severe or prolonged disease. A recent study reported that some of macrolide-resistant MP pneumonia patients who received macrolide treatment showed clinical failure with progress of disease. In contrast, it has been reported that macrolide resistance alone does not affect clinical course. A multicenter study showed that the time to defervescence after initiation of macrolide therapy in MP pneumonia children was longer in the macrolide-resistant group than in the macrolide-susceptible group (5.7 days vs. 4.1 days). However, multivariate logistic regression analysis revealed that the presence of extra-pulmonary signs, homogeneous lobar consolidation, and parapneumonic effusion was responsible for prolonged fever ≥7 days following macrolide treatment regardless of macrolide resistance. In a Taiwan study, while intensive care unit (ICU) cases (n=34) were associated with more pleural effusion, longer fever duration, and longer hospital stay, compared with non-ICU cases (n=291), macrolide resistance was similar in ICU and non-ICU groups.

What to consider when diagnosing MP pneumonia

As the need for secondary drugs increases, there are concerns about their safety: not only adverse reactions from them, but also their misuse and overuse. Efforts to diagnose MP infection more carefully are necessary to avoid unnecessary exposure to antibiotics or additional therapeutic agents.

1. Serologic diagnosis

Diagnosis of MP infection in routine clinical practice has been mainly based on serology, since bacterial culture of this organism takes long time and lacks sensitivity. Several specific antibody tests can be used to measure serum antibodies to MP. The complement fixation test, which was the standard serologic method for the diagnosis of MP infection, is currently not widely used due to nonspecific reactions and lack of sensitivity. Alternative formats adapted for commercial serologic assays include indirect immunofluorescence assay, particle agglutination (PA) assay, and enzyme-linked immunoassay (EIA). The PA assay was the most widely used method in Korea until recently, because it is easy to perform and provide quantitative results with acceptable sensitivity. However, the PA assay has some disadvantages: ambiguity in the determination of agglutination, nonspecific reactions, and inability to discriminate between IgG and IgM. The EIA can detect IgG and IgM separately, which can be somewhat helpful to distinguish current from past infections. Recently, many diagnostic laboratories have replaced the PA tests with EIA for demonstration of MP antibodies. The current widely used IgM EIA test has the advantage of being able to diagnose using a single sample, but it is likely to have a false-positive result and can be detected for a long time after infection. Therefore, in Japans Guiding Principle, even if IgM is positive, it is recommended to confirm the diagnosis using IgG tests from paired samples. Testing paired samples is practically impossible and is not always necessary for routine MP pneumonia care. In the case of refractory pneumonia, testing IgG from paired samples can be used to confirm the diagnosis of MP infection since it is often more than 1–2 weeks after the onset of illness when determining whether to change the treatment.

2. Diagnosis using molecular detection

Molecular diagnostic tests using PCR are more advantageous than culture tests and serologic tests since they can detect the presence of bacterial DNA quickly and more sensitively. In addition, PCR assays can be used to diagnose MP infection earlier than serology tests, particularly in young children and immunocompromised patients who do not produce antibodies. The PCR can also play a complementary role in interpreting the results of serologic tests to exclude the past infections. Because both serology and PCR may occasionally give non-overlapping false-negative results, a combination of these 2 diagnostic methods would be helpful.

However, PCR testing using nasopharyngeal specimen or sputum may be positive also in the presence of upper respiratory carriage or asymptomatic infection. PCR cannot distinguish actual infection from carriage or asymptomatic infection. Unfortunately, there is no current diagnostic test that is able to differentiate between asymptomatic carriage and symptomatic infection.

3. Asymptomatic carriage of MP in the upper respiratory tract

Like many other respiratory pathogens, MP can be carried
asymptomatically in upper respiratory tract. Recent studies have demonstrated that asymptomatic carriage of MP is highly prevalent. In a Dutch study, MP DNA was detected in 21% of asymptomatic healthy children in 16% of children with respiratory symptoms. The MP detection rates using culture were not significantly different between asymptomatic and symptomatic children (1.0% vs. 1.6%). Among symptomatic children, the prevalence of MP by real-time PCR did not differ significantly between children with an upper respiratory infection and a lower respiratory infection (15.9% vs. 15.6%). This suggests that among pneumonia patients with positive MP PCR in nasopharyngeal specimens, some of them are caused by a pathogen other than MP and MP only plays a role as a colonizer. A longitudinal observation of asymptomatic MP carriers revealed that MP bacteria were present for up to 4 months without causing disease. This prolonged shedding of MP seems to be related with that natural immunity against MP does not persist for a long time after MP infection and frequent reinfection is possible in the same person.

4. Considering infections caused by other respiratory pathogens

When patients with pneumonia who have been empirically treated with macrolide have fever and have no clinical improvement, the possibility of infection with other pathogens should be considered as well as that of macrolide-resistant MP infection. Children with MP infection were found to simultaneously carry many kinds of pathogens in their upper respiratory tract (Table 2). These pathogens include the bacteria Staphylococcus pneumoniae, Staphylococcus aureus, Moraxella catarrhalis, and Haemophilus influenzae, and the viruses influenza A/B, human metapneumovirus, respiratory syncytial virus, parainfluenzavirus, rhinovirus, coronavirus, bocavirus, and adenovirus. Recent studies have reported coexistence of MP with other pathogens in children with MP pneumonia. Other pathogens were simultaneously found in 20.5%–56.1% of the patients with MP pneumonia. However, the impact of coinfections in MP pneumonia on disease severity is not yet determined.

Recently, multiplexed PCR tests for bacterial or viral pathogen of respiratory infections have been widely used in many hospitals, which can be helpful to rule out the possibility of coinfection. However, it can require multiple nasopharyngeal swabs that make young patients and their caregivers discomfort, and additional cost. Since virus panel testing is currently not covered by national health insurance for patients of other ages than neonates, it may not be applied to all patients requiring the tests.

### Efficacy of secondary antimicrobial treatment in refractory MP pneumonia

As macrolide-resistant MP infections have been increasing, the needs and concerns for secondary antibiotic treatment have been rising. In 2 recent randomized studies of children with macrolide-refractory MP pneumonia, the duration of fever in the group with azithromycin plus minocycline was shorter than that in the control group with azithromycin-only. In a Japanese study of patients with macrolide-resistant MP pneumonia, the 48-hour defervescence rate was 2%, 100%, and 0% in the macrolide, minocycline, and tosufloxacin groups, respectively. This was similar to the results of other observational studies that showed 48-hour defervescence rates in the minocycline group (87%–91%) and macrolide group (44%–46%). However, the 48-hour defervescence rate of the tosufloxacin group was conflicting with those of other studies (69%). In an observational study of children with MP pneumonia who had persistent fever beyond
72 hours of macrolide treatment, the secondary treatments such as doxycycline and levofloxacin did not shorten the duration of fever or hospitalization compared to prolonged macrolide treatment.44)

It still remains controversial whether delayed effective antimicrobial treatment is associated with prolonged and/or more severe disease. Some studies reported that delayed treatment was associated with prolonged fever after appropriate treatment, fulminant disease, and extrapulmonary manifestations.23,45) In contrast, other studies demonstrated that patients with macro-lide-resistant MP pneumonia responded to macrolide treatment without treatment failure.46)

### Safety issues of secondary antimicrobial treatment

Table 3 describes the dosage and approved age of antibiotics used for the treatment of MP pneumonia.

#### 1. Tetracyclines

Tetracyclines are generally safe in children although concerns about adverse reactions, such as gastrointestinal disturbances, esophagitis, photosensitivity, tooth discoloration, still remain.6,47) Most concerning side effect is incorporation into calcifying tissues at the time of their administration. The affinity for mineralizing tissue leads to incorporation into calcifying tissues, tissues at the stage of tooth mineralization, and activity of the mineralization process.52) However, previous studies showed that short cycles with limited courses of treatment (less than 6 courses, 6 days per course) caused negligible tooth discoloration in children treated in the first 5 years of life.53)

Minocycline is more likely to cause other central nervous system effects (e.g., dizziness, lack of concentration, ataxia, vertigo, and tinnitus associated with weakness, nausea, and vomiting) and pigmentation of various body sites. Although overall adverse reactions are reported more frequently for minocycline, minocycline and doxycycline are generally well tolerated in young children.38)

#### 2. Fluoroquinolones

Fluoroquinolones have not been widely accepted as safe drug for use in children because of the observation that young animals consistently develop damages on articular surfaces of cartilage after exposure to these drugs. However, fluoroquinolones are generally well-tolerated as supported by recent reports on musculoskeletal safety concerns. Retrospective and prospective assessments of children exposed to fluoroquinolones have not clearly demonstrated the development of lesions consistent with those observed in young animals.54,55) A recent comparative, unblinded, observational surveillance study has monitored the occurrence of predefined musculoskeletal disorders (arthralgia, arthritis, tendinopathy, and gait abnormality) in children participating in prospective, randomized efficacy trials. Levofloxacin was well tolerated during and for 1 month after therapy.50) Even though the incidence of musculoskeletal disorders was greater in levofloxacin group compared with comparator group at 2 months (2.1% vs. 0.9%; P=0.04) and 12 months (3.4% vs. 1.8%; P=0.03) after starting therapy,49) 85% of these disorders consisted of reports of arthralgia, without any objective signs of joint disease documented for the children by the investigator at the time of the follow-up visit examination. Moreover, the results of following-up to 5 years after treatment revealed no

### Table 3. Recommended treatments for pediatric patients of M. pneumoniae pneumonia

| Drug | Route of administration | Drug dose (mg/kg/day), divided dose/day | Maximal dose | Treatment period | Age limit |
|------|-------------------------|----------------------------------------|--------------|-----------------|-----------|
| Azithromycin | Oral or IV | Day 1, 10 mg/kg/day, #1; Day 2–5, 5 mg/kg/day, #2 | Day 1, 500 mg/day; Day 2–5, 250 mg/day | 3 Days | - |
| Clarithromycin | Oral or IV | 15 mg/kg/day, #2 | 1,000 mg/day | 10 Days | - |
| Roxithromycin | Oral | 5–8 mg/kg/day, #2 | 300 mg/day | 10 Days | - |
| Levofloxacin | Oral or IV | Oral: <5 years of age: 16–20 mg/kg/day, #2; ≥5 years of age: 8–10 mg/kg/day, #1; adolescents with mature musculoskeletal system, 500 mg/day, #1; IV: <5 years of age: 16–20 mg/kg/day, #2; ≥5 years of age: 8–10 mg/kg/day, #1 | 750 mg/day; 5 days (750 mg/day) | ≤18 Years of age | - |
| Tosufloxacin | Oral | 12 mg/kg/day, #2 | 360 mg/day | 7–14 Days | ≤18 Years of age |
| Doxycycline | Oral | 4 mg/kg/day, #2 | 200 mg/day | 10 Days | <12 Years of age |
| Minocycline | Oral or IV | 2–4 mg/kg/day, #2 | 400 mg/day | 7–14 Days | <12 Years of age |

#IV, intravenous.

44) Tosufloxacin is approved for pediatric use in Japan. 46) Doxycycline and minocycline are approved for use in children ≥8 years of age in the United States.
clinically detectable difference between levofloxacin- and comparator-treated children in musculoskeletal toxicity.\(^{57}\) It suggested that risks of cartilage injury with levofloxacin appear to be uncommon, are clinically undetectable during 5 years, or are reversible.\(^{57}\)

Other potential adverse reactions of fluoroquinolones, albeit very uncommon in children, include central nervous system adverse effects (seizures, headaches, dizziness, lightheadedness, sleep disorders, and hallucinations) and peripheral neuropathy. However, recent studies have reported that the rates of neurologic events were statistically similar between fluoroquinolone- and comparator-treated children.\(^{36,38}\)

Although there has been no compelling published evidence that supports the occurrence of sustained injury to developing bones or joints in children treated with currently available fluoroquinolones, the U.S. Food and Drug Administration (FDA) analysis of ciprofloxacin safety data suggests the possibility of increased musculoskeletal adverse events. The rate of arthropathy in the ciprofloxacin group exceeded that of the comparator group by more than 6%.\(^{59}\) In 2016, FDA restricted the use of fluoroquinolones even in adults only if no other alternative treatment exists.\(^{60}\) It is because of the potential risks of disabling side effects involving tendons, muscles, joints, nerves, and the central nervous system.

No compelling published evidence to date supports the occurrence of sustained injury to developing bones or joints in children treated with available fluoroquinolone agents; however, FDA analysis of ciprofloxacin safety data suggests the possibility of increased musculoskeletal adverse events.

Tosufloxacin is an oral fluoroquinolone developed by Toyama Chemical Co. Ltd. (Tokyo, Japan) in 1990. The granule form was released for administration to children with otitis media or pneumonia in Japan in January 2010. In Japan, the use of tosufloxacin for children is recommended only for pneumonia when a resistant bacterial infection is suspected and other antimicrobial agents are expected to be ineffective. A Japanese guiding principle has recommended that tosufloxacin may be considered for patients with MP pneumonia who do not respond to macrolide even in children.\(^{21}\) Although no adverse reaction to the musculoskeletal system has been reported in children yet, it has not been approved for use in children in other countries than Japan due to a potential risk of inducing cartilage and joint toxicity in children. Additionally, because the increased use of fluoroquinolones has a risk of emerging resistant bacteria,\(^{61}\) decisions about antibiotic use should be made carefully.

Therefore, when prescribing a fluoroquinolone to a child or adolescent, the risks and benefits of this drug should be reviewed and discussed with patients and their caregivers or legal guardians. They should be aware of side effects, including rash, diarrhea, joint or tendon pain, confusion, or numbness or tingling of extremities while taking the antibiotic.

Conclusions

It is challenging for clinicians to treat pediatric patients with refractory MP pneumonia. Because diagnostic testing for macrolide-resistant MP infection is currently not clinically available, careful diagnosis is necessary using a combination of serology and molecular detection. In addition, efforts to consider carriage, asymptomatic infection, and infection by other pathogens are necessary to avoid unnecessary exposure to drugs. While some studies have suggested a good efficacy of second-line antimicrobials, further well-designed studies are needed to prove their efficacy on refractory MP pneumonia. Although second-line antimicrobials are generally tolerable, careful decision should be given taking into account the benefits and risks of their use. If prescribed to pediatric patients, monitoring on its adverse events is necessary.

Footnotes

Conflicts of interest: No potential conflict of interest relevant to this article was reported.

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