AMPLIFICATION OF CYCLINE D1, C-MYC AND EGFR ONCOGENES IN TUMOUR SAMPLES OF BREAST CANCER PATIENTS

Nasta Tanić1, Vedrana Milinković2, Tatjana Dramičanin1, Milica Nedeljković2, Tijana Stanković2, Zorka Milovanović4, Snežana Sušnjar5, Verica Milošević6, Branka Šošić-Jurjević6, Radan Đodović7, Nikola Tanić2

1Department of Radiobiology and Molecular Genetics, Institute of Nuclear Sciences »Vinča«, University of Belgrade, Belgrade, Serbia
2Department of Neurobiology, Institute for Biological Research »Siniša Stanković«, University of Belgrade, Belgrade, Serbia
3Department of Experimental Oncology, Institute for Oncology and Radiology of Serbia, Belgrade, Serbia
4Pathology Department, Institute for Oncology and Radiology of Serbia, Belgrade, Serbia
5Department of Medical Oncology, Institute for Oncology and Radiology of Serbia, Belgrade, Serbia
6Department of Cytology, Institute for Biological Research »Siniša Stanković«, University of Belgrade, Belgrade, Serbia
7Department of Surgical Oncology, Institute for Oncology and Radiology of Serbia, Belgrade, Serbia

Summary

Background: Breast cancer is the most common form of cancer in women. It arises from multiple genetic changes in oncogenes and tumor suppressor genes. Among so far studied oncogenes relatively few, including epidermal growth factor receptor (EGFR), cyclinD1 (CCND1) and c-myc, have been found to play an important role in progression of this type of human malignancy. The aim of this study was to examine the prognostic potential of CCND1, c-myc and EGFR amplification and their possible cooperation in breast carcinogenesis.

Methods: Copy number analyses of CCND1 and c-myc genes were done by TaqMan based quantitative real time PCR. Amplification status of EGFR was determined by differential PCR.

Address for correspondence:
Nikola Tanić
University of Belgrade
Institute for Biological Research
Department of Neurobiology
Bulevar Despota Stefana 142, 11060 Belgrade, Serbia
tel.: +381 11 2078 410
Fax: +381 11 2761 433
e-mail: nikolata@ibiss.bg.ac.rs; nikolata@sbb.rs

List of Abbreviations: EGFR, epidermal growth factor receptor; HER, human epidermal growth factor receptor; QUART, quadrantectomy; SMIR, subcutaneous mastectomy; PCR, polymerase chain reaction; HT, hormone therapy; RT, radiotherapy; CHT, chemotherapy.
Results: Amplification of CCND1, c-myc and EGFR oncogene has been found in 20.4%, 26.5% and 26.5% of breast cancer cases, respectively. Analysis showed that amplification of CCND1 oncogene was significantly associated with the stage II of disease while amplification of EGFR gene was significantly associated with overexpression of HER-2/neu. Tumour stage and expression of HER-2/neu appeared to be significant predictors of patient’s outcome. Stage I patients lived significantly longer then stage III patients (p=0.04) while patients with HER-2/neu overexpression had worse prognoses and lived significantly shorter (p=0.001). Finally, survival of patients who underwent hormone therapy only was significantly longer (p=0.001) then survival of the rest of patients.

Conclusions: Amplification of CCND1 or EGFR oncogene is associated with the progression of breast cancer and bad prognosis. No co-ordination in amplification of CCND1, c-myc and EGFR oncogenes were established in this cohort of breast cancer patients.

Keywords: breast cancer; oncogenes; cycline D1, c-myc, EGFR

Introduction

Breast cancer is the most common form of cancer in women. It comprises 22% of all cancers (1) and is second only to lung cancer as a cause of cancer related death in women (2). It is a heterogeneous disease arising from multiple genetic changes in oncogenes and tumour suppressor genes with pivotal roles in the control of cell proliferation, differentiation and death. Alterations of these genes lead to clonal expansion with subsequent acquisition of invasive and metastatic phenotypes.

Numerous oncogenes have been characterized in human cancers, but relatively few have been found to play an important role in promotion and progression of breast cancer. Among them are epidermal growth factor receptor (EGFR), cyclin D1 and c-myc.

EGFR (also known as HER1) is a member of the human epidermal growth factor receptor (HER) family of transmembrane receptor tyrosine kinases that is linked to growth control, cell adhesion, mobility and apoptosis (3). Its role in breast tumors is complicated by the fact that its function may vary according to important clinical features like estrogen receptor (ER) and HER2 status (4). Namely, high expression of EGFR has been reported to be associated with low expression of ER (5).

Cyclin D1 is the product of the CCND1 gene and plays the central role in the regulation of progression from the G1 to the S phase of the cell cycle through the formation of active enzyme complexes with cyclin-dependent kinases Cdk4 and Cdk6 (6). Consequently, deregulation of cyclin D1 gene expression or function contributes to loss of normal cell cycle control during carcinogenesis. Strong evidence implicates cyclin D1 amplification and overexpression as a driving force in human breast cancer (7).

c-myc protein is a transcription factor which participates in most aspects of cellular function, including replication, growth, metabolism, differentiation, and apoptosis (8). Most, if not all, types of human malignancy have been reported to have amplification and/or overexpression of c-myc oncogene.

Thus, amplification and overexpression of these oncogenes and oncogene products are the major mechanisms through which these genes participate in carcinogenesis. A drawback of many studies of oncogenes in human breast cancer is that usually only one oncogene was evaluated. Based on a series of unselected cases, in the present study we aimed to examine the possible prognostic potential of the amplification of CCND1, c-myc and EGFR oncogenes. Moreover, we aimed to determine whether these oncogenes cooperate in breast carcinogenesis. Furthermore we studied whether adjuvant therapies such as chemotherapy and endocrine treatment or no treatment at all had any impact on survival among oncogene amplified breast cancer patients.

Material and Methods

Patients

This prospective study comprised of 49 primary breast cancer tissue samples. 21 patients underwent modified radical mastectomy, 19 underwent quadrantectomy (QUART) and 9 underwent subcutaneous mastectomy (SMIR) at the Institute of Oncology and Radiology of Serbia. All relevant clinical parameters (age, tumor size, lymphonodal status, disease free
survival, overall survival) were retrieved from patients’ medical records.

Collected tumor specimens and corresponding normal tissue were formalin-fixed, paraffin-embedded and hematoxylin-eosin (HE) stained. Histological type and grade of each carcinoma sample were determined after hematoxylin-eosin staining. The carcinomas were graded (I – III) according to Scarff-Bloom-Richardson scoring system (9).

For each obtained tumor sample written consent and approval were acquired according to the ethical standards laid down in the 1964 Declaration of Helsinki, the International Ethical Guidelines for Biomedical Research Involving Human Subjects (CIOMS), Geneva 1993, and the Guidelines for Good Clinical Practice CPMP/ICH/135/95, September 1997.

Immunohistochemistry

Labelled streptavidin-biotin-LSAB+ method together with immunoperoxidase was used according to the recommended procedure for commercial primary monoclonal mouse antibody: Anti-Human ERO clone (1:50; Clone 1D5; Dako) and Anti-Human PR clone (1:50; Clone PgR 636; Dako), as well as for polyclonal rabbit antibody Anti-Human c-erbB2/HER2 Oncorprote (1:300; Dako) with Dako LSAB+TM+/ HRP kit (K0679). Slices were contrasted with Mayer hematoxylin.

The evaluation of steroid receptors (ER, PR) was based on the scoring system which included percentage of stained malignant nuclei (0-5) and their intensity of staining (0–3); positive (high expression) cases were with score ≥4 while negative (low expression) cases were with score <4 (10). HER2 status was determined using DAKO scoring system and HER2 positive status was defined if IHC score was 2+/3+ (11).

Copy number analysis by quantitative real time PCR

Genomic DNA was extracted from 49 fresh frozen tumor and corresponding normal tissue samples according to the standard phenol/chloroform extraction procedure described by Sambrook and colleagues (12). The quality of the extracted DNA was verified by agarose gel electrophoresis and the concentrations (12). The quality of the extracted DNA was verified by agarose gel electrophoresis and the concentrations (12). The quality of the extracted DNA was verified by agarose gel electrophoresis and the concentrations (12). The quality of the extracted DNA was verified by agarose gel electrophoresis and the concentrations (12). The quality of the extracted DNA was verified by agarose gel electrophoresis and the concentrations (12).

Copy number analyses of CCND1 and c-myc genes were done by quantitative real time PCR using TaqMan based assays. Assays included forward and reverse primers for c-myc and CCND1 oncogenes as well as highly specific 6-Fam-TAMRA labeled probes for them. Primers and probe for CCND1 gene were as follows: F 5'-GGACGACGAGAAGGTGATACAG-3'; R 5'-CACAGTCAGCCAGGTTTAA-3'; Probe 6-FAM-5'-.CAGCCCTTGTGTTACGGCCTTTTGTAG-3'-TAMRA. For the analysis of c-myc gene, the following primers and probe were used: F 5'-GGACGACGAGAAGGTGATACAG-3'; R 5'-CCAGGCTTCTTGCAGAGAAGCAGTTT-3'; TaqMan Probe 6-FAM-5'-.AGAAGC CGTCACCATAAGCTTGG-3'-TAMRA.

RNase-P was used as the internal control, reference gene (accession # 4316831, Applied Biosystems).

Each sample was prepared in duplicate, in total reaction volume of 20 µL, with primers /probe ratio 3:1 (0.1 µmol/L probe : 0.3 µmol/L primers), 1x TaqMan Master Mix and 150ng of tested DNA. Each reaction contained normal DNA controls. Control samples were used as calibrators. PCR reactions were carried out in the ABI Prism 7500 Sequence Detection System at 50 °C for 2 minutes, 95 °C for 10 minutes, followed by 40 cycles at 95 °C for 15 seconds, and 60 °C for 1 minute. The experimental threshold was calculated based on the mean baseline fluorescence signal from cycles 3 to 15 plus 10 standard deviations. A mean value of each Ct duplicate was used for further calculation. Each run included a no-template control, as well. The obtained results were analyzed by RQ Study Add ON software for 7500 v 1.3 SDS instrument with a confidence level of 95% (p<0.05).

Differential PCR

Amplification status of EGFR oncogene was determined by differential PCR (D-PCR) that engaged two pairs of primers, one for the target gene (EGFR) and the other for the reference gene (β-actin). The primer sequences were as follows: EGFR_F 5'-AGCAGTCCTTCTCTTTCCAGAGTATT-3' and EGFR_R 5'-AACCCTTCACTGACGAAA-3' for EGFR, and ACTB_F 5'-CCAGTCTCTTCCCTTGGAGGGCT-3' and ACTB_R 5'-CCAGTCTCTTCCCTTGGAGGGCT-3' for the ACTB. D-PCR was performed in the total reaction volume of 25 µL with 150 ng of DNA, 1× PCR buffer (50 mMol/L KCl, 10 mMol/L Tris–HCl, pH 8.3, 0.01% gelatin), 1.5 mMol/L MgCl, 0.2 mMol/L each dNTP, 1 µmol/L each of four primers, 1U TaqPolymerase. Thermal cycling included thirty repeats of denaturation at 95 °C/1 minute, annealing at 58 °C/1 minute before and final extension (72 °C/10 minutes) after the repeating temperature steps. Generated PCR products were applied to 9% polyacrylamide gel for electrophoresis, stained with silver-nitrate, photographed and analyzed by ImageQuant 5.2 by comparing the median pixel intensity in the same lane (sample). When median pixel intensity of EGFR band was equal or higher then 25% of median pixel intensity of ACTB, it was interpreted as gene amplification.

Unauthenticated Download Date | 3/8/18 2:36 PM
Statistical analysis

Significant differences between the data sets were determined by STATISTICA 6.0 software (StatSoft, Inc., Tulsa, USA). The correlations between clinicopathological parameters and amplification of c-myc, CCND1 and EGFR genes were evaluated using Fisher exact test. Survival analyses were performed using Kaplan & Meier product-limit method. The log rank test was used to assess the significance of the difference between pairs of survival probabilities. Overall survival was calculated from the day after surgery to the last follow-up examination or death of the patient. Statistical differences were considered significant when p was < 0.05 (*).

Results

Patient cohort and treatment

We examined breast cancer specimens from 38 postmenopausal and 11 premenopausal women for the amplification status of c-myc, CCND1 and EGFR oncogenes. In total of 49 patients, 27 patients had breast carcinomas with histology of invasive ductal carcinoma, while 22 were invasive lobular carcinomas. Patients’ characteristics are summarized in Table I. Among clinical and histopathological characteristics, stage and HER-2/neu expression were significant predictors of patient’s outcome (Figures 1A and 1B). Namely, stage I patients lived significantly longer then stage III patients (p=0.04) while patients with HER-2/neu overexpression had worse prognoses and lived significantly shorter (p=0.001). Most of the samples were steroid receptor (ER and/or PR) positive (96%) but, nevertheless, patients were on different regimens of treatment: 5 of them were on hormone therapy (HT), 8 on combined hormone and radiotherapy (HT+RT), 4 on combined hormone and chemotherapy (HT+CHT), 17 on combined hormone, chemotherapy and radiotherapy (HT+CHT+RT) and 15 on other therapeutic protocols (CHT only, RT only, combined CHT and RT). Kaplan-Meier survival curves were generated to evaluate the effects of these treatment regimens on survival. The survival of patients who underwent hormone therapy only was significantly longer (p=0.001) then survival of the rest of patients (Figure 1C).

Amplification of c-myc, CCND1 and EGFR oncogenes

We determined amplification status of CCND1 and c-myc oncogenes by Quantitative Real Time PCR. Our results revealed that 10 out of 49 samples (20.4%) possessed three to 14-fold amplification of CCND1 oncogene. c-myc gene was amplified three to 10-folds in 13/49 breast cancer samples (26.5%). Further analysis by Fisher exact test showed that amplification of CCND1 oncogene was significantly associated with the stage II of breast cancer patients (Table II). On the contrary, amplification of c-myc gene did not show correlation with tumour stage (Table II) or any other clinical or pathohistological characteristics of patients (data not shown) including breast cancer subtype, ER or HER2/neu status.

Amplification of EGFR oncogene was assessed by differential PCR and we found that it was amplified in 13 patients (26.5%). Interestingly, amplification of EGFR gene was significantly associated with overexpression of HER-2/neu (Table II). In other words, EGFR expressing tumours were more likely to overexpress HER-2/neu. Association with any other clinical or pathohistological characteristic was not obtained.

In order to reveal possible association among three studied oncogenes, we further analyzed whether there were co-alterations between any of them. Our results showed that there was no co-operation among CCND1, c-myc and EGFR gene alterations. Namely, CCND1 and c-myc were co-amplified in four samples (8%), as were CCND1 and EGFR while c-myc and EGFR were co-altered in only two samples (4%). All three of them were amplified in just one breast cancer sample.

Further, we analyzed possible association between each oncogene alteration and survival and did not find anything of significance (Figure 2). Finally, neither of them had any significant influence on response to any applied therapeutic protocol.

Table I Patients’ characteristics.

| Age in years (mean) | 32 – 82 (61) |
| Follow-up in months (mean) | 36 – 110 (31) |
| Estrogen receptor status | | |
| positive | 47 |
| negative | 2 |
| HER-2/neu status | | |
| positive | 19 |
| negative | 30 |
| Stage* | | |
| I | 12 |
| II | 20 |
| III | 12 |
| Grade | | |
| g 1 | 0 |
| g 2 | 44 |
| g 3 | 5 |
| Lymph node metastasis | | |
| Yes | 27 |
| No | 22 |
| TOTAL | 49 |

* – unavailable data for 5 patients
Figure 1 Kaplan–Meier survival curves. (A) Impact of tumor stage on patients’ survival. Patients with stage I tumors lived significantly longer compared to stage III (p=0.04); (B) Impact of HER2/neu on patients’ survival. Patients with positive status of HER2/neu receptor lived significantly shorter (p=0.001); (C) Impact of therapy on patients’ survival. Patients receiving hormone therapy lived significantly longer compared to all other groups (p=0.001).

Figure 2 Impact of oncogenes amplification on patients’ survival. (A) Patients without amplification of CCND1 had tendency for better survival; (B) Amplification of c-myc seems to have no impact on patients’ survival (C) Patients without EGFR amplification had tendency for better survival.
Table II Association between amplified oncogenes, stage and HER2/neu status.

| Parameter                        | Total aNP | CCND1 | c-myc | EGFR |
|----------------------------------|-----------|-------|-------|------|
|                                  | NP | % | p | NP | % | p | NP | % | p |
| Total                            | 49 | 10 | 20.4 | 13 | 26.5 | | 13 | 26.5 | |
| Stage b                          |   |    |      |    |       | |
| I                                | 12 | 0  | 0    | 4  | 33.3 | p1 | 0.04c | 2  | 16.7 | p1 | 0.16 |
| II                               | 20 | 6  | 30.0 | 6  | 30.0 | p2 | 0.55 | 8  | 40.0 | p2 | 0.32 |
| III                              | 12 | 3  | 25.0 | 3  | 25.0 | p3 | 0.50 | 3  | 25.0 | p3 | 0.50 |
| HER2/neu status                  |    |    |      |    |       | |
| positive                         | 19 | 6  | 31.6 | 4  | 21.1 | p3 | 0.11 | 8  | 42.1 |  |
| negative                         | 30 | 4  | 15.4 | 9  | 30.0 |  |

a Number of patients; b unavailable data for 5 patients c Bold indicates statistically significant values; p1 – statistical significance between stages I and II; p2 – statistical significance between stages II and III; p3 – statistical significance between stages I and III

Discussion

Gene amplification is an important mechanism of oncogene activation and is crucial for the development and progression of cancer. In the present study we used quantitative real time PCR and differential PCR to study gene copy number alterations of CCND1, c-myc and EGFR oncogenes in a cohort of 49 primary breast cancer patients.

CCND1 oncogene was amplified in 20.4% of analyzed breast cancer samples which is consistent with previously reported frequencies of 13%–20%. Quantification results, which revealed that amplification of CCND1 was three to 14-fold, are consistent with the same reports (7, 13). Interestingly, amplification of CCND1 gene was significantly associated with the progression of breast cancer, both of ductal and lobular subtypes. Specifically, it was associated with the stage II of the disease. It is important to emphasize that cyclin D1 is always strongly overexpressed when amplified (14–16). Contrary to our findings, immunohistochemical studies of preneoplastic lesions demonstrated that overexpression of cyclin D1 had been already apparent in hyperplasia and increased with increasing malignancy (17). On the other hand, in situ hybridisation studies suggest that cyclin D1 overexpression occurs at the transition from in situ to invasive carcinoma (18). Our results imply that amplification of CCND1 oncogene could be the marker of the progression to stage II of the disease. In addition, studies on primary breast cancers indicate that overexpression of cyclin D1 was confined to specific phenotypes, implying different roles in different subtypes of the disease. Lobular carcinoma appears to universally overexpress cyclin D1 (19), while overexpression in ductal carcinoma is confined almost exclusively to estrogen receptor positive cases (20). We have not observed any difference in CCND1 amplification between lobular and ductal breast carcinomas but we are limited in discussing this issue since 96% of our samples are ER positive.

The need for cooperativity with other oncogenic hits is entirely expected for an authentic human oncogene (7), which was the reason why we analyzed the amplification of c-myc and EGFR in the same cohort of samples. c-myc gene was amplified in 26.5% of analyzed samples, slightly above reported frequencies (21, 22), but did not show any co-ordination with CCND1 amplification. Barnes and Gillette (23) have also shown that CCND1 gene is amplified in cases were c-myc is not. Although this reciprocal amplification seems to be consistent with the observation that c-myc represses the transcription of cyclin D1 (24), direct evidence is still lacking for a reciprocal relationship between the expression of c-myc and that of cyclin D1.

The role that epidermal growth factor receptor plays in breast cancer has been a subject of intensive study and controversy. Some retrospective immunohistochemical studies have indicated that EGFR overexpression in primary tumors is an indicator of poor prognosis (25), whereas other similar studies have failed to establish such a link (26). Collectively, these studies suggest that EGFR is expressed in 18–35% of breast cancers (27). Our study revealed that EGFR oncogene was amplified in 26.5% of examined samples. No co-alteration with c-myc and CCND1 was observed. However, amplification of EGFR gene showed significant association with the expression of HER-2/neu. The largest and the most comprehensive study analyzing EGFR expression in breast cancer patients showed that EGFR expression was positively correlated with HER-2/neu overexpression, which has been associated with bad prognosis (28). Our results confirm this finding since Kaplan-Meier survival analysis showed that patients with HER-2/neu overexpression had worse prognoses and lived significantly shorter.
Finally, the most promising therapy in this patient’s cohort was endocrine (hormone) therapy. We looked for an oncogene signature in the background and found that samples of patients who underwent endocrine therapy did not have amplifications of CCND1 and EGFR oncogenes. This was not a surprise since the overexpressions of both, CCND1 and EGFR, have been associated with the resistance to hormone therapy and chemotherapy in a number of studies (29, 30). However, the therapy groups analyzed in the present study are small, and therefore we recommend careful interpretation of the findings.

In conclusion, amplification of CCND1 or EGFR oncogenes is associated with the progression of breast cancer and bad prognosis. No co-ordination in amplification of CCND1, c-myc and EGFR oncogenes was established in this cohort of breast cancer patients.

Acknowledgments. This work has been funded by the Ministry of Education, Science and Technological Development, Republic of Serbia, grant # III41031 and grant # ON173049.

Conflict of interest statement

The authors stated that there are no conflicts of interest regarding the publication of this article.

References

1. Parkin DM. International variation. Oncogene 2004; 23: 6529–40.
2. Stewart SL, King JB, Thompson TD, Friedman C, Wingo PA. Cancer mortality surveillance – United States. 1990–2000, MMWR, Surveill Sum 2004; 53: 1–108.
3. Yarden Y, Sliwkowski MX. Untangling the ErbB signalling network. Nat Rev Mol Cell Biol 2001; 2: 127–37.
4. Hoadley KA, Weigman VJ, Fan C, Sawyer LR, He X, Troester MA, et al. EGFR associated expression profiles vary with breast tumor subtype. BMC Genomics 2007; 8: 258.
5. Witton CJ, Reeves JR, Going JJ, Cooke TG, Bartlett JM. Expression of the HER1-4 family of receptor tyrosine kinases in breast cancer. J Pathol 2003; 200: 290–7.
6. Sherr CJ. Cancer cell cycles. Science 1996; 274: 1672–7.
7. Andrew Arnold, Papanikolaou A. Cyclin D1 in Breast Cancer Pathogenesis. J Clin Oncol 2005; 23: 4215–24.
8. Liao DJ, Dickson RB. c-Myc in breast cancer. Endocrine-Related Cancer 2000; 7: 143–164.
9. Bloom HJ, Richardson WW. Histological grading and prognosis in breast cancer. A study of 1409 cases of the HER1-4 family of receptor tyrosine kinases in breast cancer. J Pathol 2003; 200: 290–7.
10. Leake R, Barnes D, Pinder S, Ellis I, Anderson L, Anderson T, et al. Immunohistochemical detection of steroid receptors in breast cancer: a working protocol. J Clin Pathol 2000; 53: 634–5.
11. HercepTestTM, For determination of HER2 protein overexpression. Catalog Products and Services, DAKO 2007; 86–7.
12. Barnes DM, Gillett CE. Cyclin D1 in breast cancer. Breast Cancer Research and Treatment 1998; 52: 1–15.
13. Jansen-Durr P, Meichle A, Steiner P, Pagano M, Finke K, Botz J, et al. Differential modulation of cyclin gene expression by MYC. Proc Natl Acad Sci USA 1995; 90: 3685–9.
24. Newby JC, A’Hern RP, Leek RD, Smith IE, Harris AL, Dowsett M. Immunohistochemical assay for epidermal growth factor receptor on paraffin-embedded sections: validation against ligand binding assay and clinical relevance in breast cancer. Br J Cancer 1995; 71: 1237–42.

25. Tsutsui S, Ohno S, Murakami S, Hachitanda Y, Oda S. Prognostic value of epidermal growth factor receptor (EGFR) and its relationship to the estrogen receptor status in 1029 patients with breast cancer. Breast Cancer Res Treat 2002; 71: 67–75.

26. Pawlowski V, Revillion F, Hebbar M, Hornez L, Peyrat JP. Prognostic value of the type I growth factor receptors in a large series of human primary breast cancers quantified with a real-time reverse transcription-polymerase chain reaction assay. Clin Cancer Res 2000; 6: 4217–25.

27. Rimawi MF, Shetty PB, Weiss HL, Schiff R, Osborne CK, Chamness GC, et al. Epidermal Growth Factor Receptor Expression in Breast Cancer Association with biologic phenotype and clinical outcomes. Cancer 2010; 116: 1234–42.

28. Lundgren K, Brown M, Pineda S, Cuzick J, Salter J, Zabaglo L, et al. Effects of cyclin D1 gene amplification and protein expression on time to recurrence in postmenopausal breast cancer patients treated with anastrozole or tamoxifen: a TransATAC study. Breast Cancer Research 2012; 14: R57

29. Giltnane JM, Ryden L, Cregger M, Bendahl PO, Jirstrom K, Rimm L. Quantitative measurement of epidermal growth factor receptor is a negative predictive factor for tamoxifen response in hormone receptor positive premenopausal breast cancer. J Clin Oncol 2007; 25: 3007–14.

Received: August 15, 2013

Accepted: September 10, 2013