Differential Interstrain Susceptibility to Vertebrobasilar Dolichoectasia in a Mouse Model

Y.-Q. Zhu, H. Xing, D. Dai, D.F. Kallmes, and R. Kadirvel

ABSTRACT

BACKGROUND AND PURPOSE: Vertebrobasilar dolichoectasia is characterized by arterial elongation, dilation, and tortuosity and leads to high risks of ischemic stroke. Our aim was to investigate the differential susceptibility to elastase-induced vertebrobasilar dolichoectasia induction in 2 different mouse strains.

MATERIALS AND METHODS: Elastase (25 mU) was injected into the cisterna magna in C57BL/6J (n = 36) and 129/SvEv (SV129) (n = 36) mice. Control animals were injected with heat-inactivated elastase (n = 12 for each strain). At 3, 7, 14, and 28 days after elastase injection, MICROFIL polymer perfusion was performed. The arterial tortuosity index and the percentage increase in diameter were calculated for the basilar artery. Arterial samples were processed for conventional histologic examination, immunostaining, and matrix metalloproteinase expression. A ≧50% increase in diameter and a tortuosity index of ≧10 for the basilar artery were used to indicate success in achieving vertebrobasilar dolichoectasia.

RESULTS: Successful vertebrobasilar dolichoectasia induction was noted in 67% (18 of 27) of the C57BL/6J strain versus 0% (0 of 19) of the SV129 strain (P < .001). Vertebrobasilar dolichoectasia was not observed in sham-operated controls. Both the tortuosity index and diameter increase for the basilar artery were greater in the C57BL/6J strain compared with the SV129 strain [56.3% ± 16.4% versus 21.1% ± 21.6% for diameter, P < .001; 17.4 ± 7.6 versus 10.4 ± 3.8 for tortuosity index, P < .001]. Expression of pro-matrix metalloproteinase-2 and pro- and active matrix metalloproteinase-9 was increased in elastase-injected C57BL/6J animals compared with elastase-injected SV129 animals (P = .029, 0.029, and 0.029, respectively). Inflammation scores were significantly higher in C57BL/6J animals versus SV129 animals (P < .001). C57BL/6J subjects demonstrated arterial wall dilation and elongation characterized by internal elastic lamina disruption, muscular layer discontinuity, inflammatory cell infiltration, and high matrix metalloproteinase expression in the media.

CONCLUSIONS: C57BL/6J mice demonstrated greater susceptibility to vertebrobasilar dolichoectasia induction than SV129 mice.

ABBREVIATIONS: BA = basilar artery; MMP = matrix metalloproteinase; TI = tortuosity index; VBD = vertebrobasilar dolichoectasia
VBD induced via microsurgical injection of porcine elastase into the cisterna magna to induce morphologic changes similar to human VBD. The aims of the current study were to elucidate the morphologic and molecular differences in VBD between C57BL/6J and 129/SvEv (SV129) mouse strains.

**MATERIALS AND METHODS**

**Elastase Injection Procedure**

C57BL/6J \( (n = 48) \) and SV129 \( (n = 48) \) female mice \( (6–8 \) weeks of age \) (Charles River Laboratories, Wilmington, Massachusetts) were used in the current study and were divided into a test group \( (n = 36 \) for each strain) and a control group \( (n = 12 \) for each strain). The use of animals and procedures was reviewed and approved by the Institutional Animal Care and Use Committee in Mayo Clinic.

The mouse model of VBD was induced as previously described.\(^6\) Using a 10-μL micro NanoFil syringe (World Precision Instruments, Sarasota, Florida) with a 36-ga beveled needle, we injected 25 mU of porcine elastase (Worthington Biochemical, Lakewood, New Jersey) in 2.5 μL of phosphate-buffered saline (10 μL/μL) into the cisterna magna under a dissection microscope. Control mice were injected with inactivated elastase.

**MICROFIL Perfusion and Tissue Harvest**

Animals in the test group of each strain were randomly assigned to 4 subgroups \( (n = 8 \) at each time point) and were sacrificed at 3, 7, 14, and 28 days after elastase injection. Control mice \( (n = 8 \) for each strain) were sacrificed at 28 days. The MICROFIL (Flow-Tech, Cockeysville, Maryland) perfusion was performed at follow-up as described previously.\(^6\) After MICROFIL perfusion, the whole brain along with the cerebral vascular trees was harvested and fixed in 10% buffered formalin at room temperature at least 24 hours before imaging. Animals designated for zymography analysis were perfused with saline; the basilar artery was harvested at 14 days and snap frozen in liquid nitrogen \( (n = 8 \) for each strain; \( n = 4 \) for each control and test group).

**Morphometric Analysis**

Under the dissecting microscope, visual inspection of the arteries at the skull base was performed to determine the presence of tortuosity and enlargement compared with the control samples. The vascular trees were then photographed by using the MicroPublisher 5.0 RTV camera (QImaging; http://www.qimaging.com/) attached to the dissection microscope; Q-Capture Pro 7 software (QImaging) with inner calibration was used to capture the images. Morphometric analysis was performed for the BA. We selected the area of maximal dilation for diameter measurements (exterior side to exterior side) from the MICROFIL-perfused gross images by using Image-Pro Plus software (Media Cybernetics, Bethesda, Maryland). The percentage increase in arterial diameter \( (\text{individual measured diameter} \div \text{average diameter of control samples}) \div \text{average diameter of control samples} \) was determined. The previously validated tortuosity index \( (\text{TI})^7 \) was applied to calculate the TI for the current study. TI was defined as the following: \( [(\text{actual length of the vessel} \div \text{straight-line length of same vessel} – 1)] \times 100 \). Successful VBD induction was defined as a TI \( \geq 10 \) and a \( \geq 50\% \) increase in BA diameter compared with control samples.

**Histologic Examination**

Harvested samples were fixed in 10% neutral-buffered formalin for \( >48 \) hours, dehydrated by using a graded ethanol series \( (70\%–100\%) \), and embedded in paraffin. Transverse sections containing the BA were taken at \( 4-\mu \text{m} \) thickness and stained with hematoxylin-eosin and elastic fiber stains (Verhoeff-van-Gieson) to assess internal elastic lamina disruption in the media. Serial cross-sections of the BA were used to evaluate inflammation reaction on the basis of inflammation scores.\(^7\) The score was defined as follows: \( 0 = \) no inflammatory cell infiltration; \( 1 = \) minimal or mild: scant, scattered inflammatory cell infiltration; \( 2 = \) patchy but localized or limited inflammatory cells; \( 3 = \) marked, attenuated, diffuse inflammatory cell infiltration. Samples were also immunostained by using the rabbit polyclonal smooth-muscle α-actin antibody \( (1:200; \text{Abcam, Cambridge, Massachusetts}) \), rabbit polyclonal CD45 antibody (leukocyte common antigen; \( 1:200 \); Abcam), mouse monoclonal antimacrophage antibody (clone MAC387; \( 1:200 \); Thermo Fisher Scientific, Waltham, Massachusetts), rabbit polyclonal matrix metalloproteinase (MMP)-9 antibody \( (1:100; \text{Abcam}) \), and rabbit monoclonal MMP-12 antibody (clone EP1261Y; \( 1:200 \); Abcam) by using the immunofluorescent technique. Double immunofluorescent staining was performed by using anti-CD45 (leukocytes) and antimacrophages to assess whether macrophages were included in leukocytes. Negative controls were performed by omitting primary antibodies. Statistical analysis of the histopathologic images was performed with ImagePro Plus software in at least 5 randomly selected high-power \( (\times 400) \) tubulointerstitial fields from each section. Two pathologists assessed every pathologic index in each section and reached agreement by consensus in a blinded manner.

**Gelatin Zymography for MMPs**

Soluble proteins were extracted from the BA of elastase-injected \( (n = 4 \) for each strain) and heat-inactivated elastase-injected \( (n = 4 \) for each strain) groups at 14-day follow-up. Protein samples \( (20 \mu \text{g per lane}) \) were separated by \( 10\% \) zymogram gel (Bio-Rad Laboratories, Hercules, California). Gels were washed with renaturation buffer (Bio-Rad) for 1 hour and then incubated for 48 hours at \( 37^\circ \text{C} \) in development buffer (Bio-Rad). The gels were stained with 0.5% Coomassie blue R-250 (Thermo Fisher Scientific). The gelatinolytic bands, representing the activities of MMPs, were scanned and analyzed densitometrically by using ImageJ software (National Institutes of Health, Bethesda, Maryland).

**Data Analysis and Statistical Methods**

Animals that had neurologic complications or did not survive for specified follow-up time points were excluded from the analysis, as were the MICROFIL-perfused samples that did not show proper casting as indicated by air bubbles or partial filling in the cast.

Effect of follow-up time and strain on percentage BA increase and TI were assessed with robust analysis of variance by using the Huber M-estimator. Variables were then dichotomized into successful or unsuccessful injections for percentage BA increase and TI.
Table 1: Success of VBD formation in C57BL/6J and SV129 mouse strains*

| Follow-Up Time Point (Days) | C57BL/6J | SV129 |
|-----------------------------|----------|-------|
| Total No. | No. Successful | Success Rate (95% CI) | Total No. | No. Successful | Success Rate (95% CI) | P Value |
| 3 | 7 | 4 | 0.57 (0.18–0.90) | 5 | 0 | 0.00 (0.00–0.52) | | |
| 7 | 8 | 6 | 0.75 (0.35–0.97) | 3 | 0 | 0.00 (0.00–0.71) | | |
| 14 | 7 | 4 | 0.57 (0.18–0.90) | 6 | 0 | 0.00 (0.00–0.46) | | |
| 28 | 5 | 4 | 0.80 (0.28–1.0) | 5 | 0 | 0.00 (0.00–0.52) | | |
| Total No. | 27 | 18 | 0.67 (0.46–0.84) | 19 | 0 | 0.00 (0.00–0.18) | <.001 |

*Successful VBD was defined as the percentage increase in basilar artery diameter of ≥50% and a tortuosity index of ≥10. The number and proportion of mice showing successful VBD for each strain and time point are displayed. Ninety-five percent Clopper-Pearson CIs are displayed for each estimate. Proportion of successes was compared between strains using the Fisher exact test.
in the elastase-injected group as that in the C57BL/6J at all follow-up time points. Acute and/or chronic inflammatory cell infiltration was found surrounding and/or within the vessel walls in samples taken at days 3, 7, and 14, but not at day 28 or in control samples. We did not observe positive staining for MMP-9, MMP-12, CD45, or MAC387 at any time point.

Inflammation scores in C57BL/6J mice were 2.2 ± 0.84, 1.67 ± 0.58, 3.0 ± 0.0, and 1.5 ± 0.58 at 3, 7, 14, and 28 days, respectively; in SV129 mice, they were 1.0 ± 0.82, 0.5 ± 1.0, 1 ± 1.41, and 0.0 ± 0.0 at 3, 7, 14, and 28 days, respectively. Inflammation scores were significantly different between strains at all time points (P < .001). Within-strain comparisons showed a difference between the dates of sacrifice for the C57BL/6J strain, but not for the SV129 strain (P = .03 and P = .38, respectively).

**Gelatin Zymography**

Gelatin zymography analysis showed that the activities of pro- and active MMP-2 and MMP-9 were dramatically increased in the VBD samples in C57BL/6J mice compared with sham-operated controls and elastase-injected SV129 mice at 14 days after elastase injection. The activities of pro-MMP-2, active MMP-2, pro-MMP-9, and active MMP-9 were higher in elastase-injected C57BL/6J groups compared with elastase-injected SV129 mice and sham-operated C57BL/6J and SV129 groups (Fig 5). Samples from both test and control SV129 mice did not show the activities of either MMP. Wilcoxon rank sum test results were significant for pro-MMP-2 and pro- and active MMP-9 (P = .03, 0.03, and 0.03, respectively), but not active MMP-2 (P = .07).
after elastase infusion is associated, at least in part, with genetically determined inherited traits. These current findings are important not only in identifying key elements relevant to the development of VBD but also in providing a potential pathway for future discovery of genetic determinants of VBD susceptibility, possibly through linkage analysis of different strains.

The relationship among inflammation, MMPs, and aneurysms has been widely reported in aneurysm models and clinical cases.9-14 The disruption of the internal elastic lamina by elastase injection could be the initial factor for induction of VBD. Subsequent infiltration of macrophages and inflammatory cells in the medial layer with secretion of both MMP-9 (collagenase) and MMP-12 (macrophage elastase) followed by degradation of extracellular matrix and elastic lamellae could be leading to the progression of VBD in the C57BL/6J strain. The low level of inflammatory cells and undetectable activities of MMPs in SV129 mice could be associated with less BA dilation and elongation compared with the C57BL/6J strain.

A prospective clinical trial focused on MMP activities, and polymorphism in intracranial arterial dolichoectasia revealed that plasma levels of MMP-2 and MMP-9 were not associated with dolichoectasia.15 Furthermore, dolichoectasia was strongly connected to lower levels of MMP-3 and 5A/6A polymorphism of the promoter region of MMP-3. This study was in contradiction to our findings; however, we did not measure MMP levels in plasma. It is possible that specific genetic factors may lead to VBD.

Similar to our own findings, Fujii et al16 revealed that a wide divergence in susceptibility to aneurysmal dilation occurred between C57BL/6J and SV129 mice in an elastase-induced abdominal aortic aneurysm (AAA) model. In particular, C57BL/6J mice were termed “AAA susceptible,” and SV129 mice, “AAA resistant.” Following elastase perfusion, the SV129 mice had an extent of dilation of 100.0%, whereas the C57BL/6J mice demonstrated an extent of dilation of 156%. The outcross strain between 129/SvEv and C57BL/6J (ie, B6xSvEv F1 heterozygotes) showed an overall extent of dilation of 156%, in between the 2 parent strains. These observations suggest that the susceptibility to form abdominal aortic aneurysms following elastase perfusion is associated, at least in part, with genetically determined inherited traits. It is expected that future investigations based on this information will help define the inherited genetic elements that might influence aneurysmal dilation. Meanwhile, other studies showed differences between these mouse strains in susceptibility to ischemic injury and insulin resistance.16,17

Our study has several limitations. In the previous study, the success of achieving the intracranial dolichoectasia was defined as the
The arterial dilation and tortuosity index were calculated by using 2D images; however, we believed that 2D images could well reflect its 3D tortuosity because we had found that the vertebrobasilar artery mainly goes on the pons and its tortuosity occurred parallel to the surface of pons and in 1 plane. In addition, the pressure perfusion was not performed for MICROFILL polymer injections; instead, manual perfusion was performed. However, extreme care was taken to avoid the variations in the perfusion fixation among animals. In this study, we used a referred dosage for the C57BL/6J strain, which might not be the optimal dosage for the 129/SvEv strain and could be a partial reason for differences. We performed neither bone marrow transplant experiments nor quantitative loci trait analysis to characterize VBD susceptibility between strains. We did not use any inflammation or MMP inhibitors or gene knockout animals to identify the roles of inflammation and MMPs in the pathogenesis of VBD. We did not observe any MMP expression in the SV129 strain, which could be related to the lower quantity (20 μg) of total protein used for zymography experiments. A robust longitudinal study is critical in elucidating the mechanisms of VBD pathobiology.

CONCLUSIONS
C57BL/6J mice demonstrated greater susceptibility to VBD induction than SV129 mice.

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