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Next-Generation Sequencing of an 88-Year-Old Specimen of the Poorly Known Species *Liagora japonica* (Nemaliales, Rhodophyta) Supports the Recognition of *Otohimella* gen. nov.

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**Abstract**

*Liagora japonica* is a red algal species distributed in temperate regions of Japan. This species has not been collected from its type locality on the Pacific coast of Japan since 1927 and seems to have become extinct in this area. For molecular characterization of *L. japonica*, we extracted DNA from the topotype material of *L. japonica* collected in 1927, analyzed seven genes using Illumina next-generation sequencing, and compared these data with sequences from modern samples of similar red algae collected from the Japan Sea coast of Japan. Both morphological and molecular data from modern samples and historical specimens (including the lectotype and topotype) suggest that the specimens from the Pacific and Japan Sea coasts of Japan should be treated as a single species, and that *L. japonica* is phylogenetically separated from the genus *Liagora*. Based on the phylogenetic results and examination of reproductive structures, we propose *Otohimella japonica* gen. et comb. nov., characterized morphologically by diffuse carposporophytes, undivided carposporangia, and involucral filaments initiated only from the cortical cell on the supporting cell.
Introduction

Taxonomic studies are based on type specimens, which are permanently attached to taxonomic names. The type specimens of red algal seaweeds are basically specimens housed in herbaria. To determine the correct names of modern collections, type specimens have been used for comparisons, including habit and anatomy. In many cases, morphological information from type specimens is not sufficient for taxonomic studies, because many type materials are fragmented and lack reproductive structures required for comparison with modern collections. Instead of type materials, detailed morphological observations and DNA sequencing from topotypes newly collected from their type locality have been used for taxonomic studies [1, 2]. However, in some cases the coastal environment of type localities has changed and the species are missing. The most reliable method to determine the correct name of modern collections involves sequencing of type specimens and comparing them to related sequences from field-collected material [3, 4]. However, DNA fragments of less than 300 base pairs can be obtained typically [3, 4], and thus complete gene sequences required for phylogenetic analyses are not available. Many type specimens were established more than 50 years ago, and the DNA has become highly fragmented due to deterioration over time [5]; therefore, Sanger sequencing is not reliable with old DNA samples. Recently, next-generation sequencing (NGS) has been used to examine DNA sequences from old samples [6]. For red algal seaweeds, Hughey et al. [7] performed NGS to determine the complete plastid and mitochondrial genomes from 140-year-old type specimens of the bangiophycean species of *Pyropia* using the published complete organelle genome data from *Pyropia* spp. for reference mapping and sequence assembly. However, in the case of Florideophyceae, the largest group of red algae, limited genomic data are available as references in NGS, compared with bangiophycean red algae, and thus old DNA sequences have not been determined previously by NGS for molecular phylogenetic analyses.

The red algal genus *Liagora* is the largest genus of Liagoraceae and is widely distributed in warm temperate to tropical regions of the Atlantic, Indian, and Pacific Oceans [8]. Recent molecular analyses suggest that *Liagora* is polyphyletic, and the generic concept of *Liagora* sensu stricto has been revised substantially in recent years [9–11]. Although three new genera, *Macrocarpus*, *Neoizziella*, and *Titanophycus*, have since been separated from *Liagora* sensu lato [10, 12], the generic positions of some species of *Liagora* remain unresolved.

*Liagora japonica* Yamada was originally described based on plants collected from Misaki, Kanagawa Prefecture, on the Pacific side of Japan in 1903 [13]. According to our investigations of specimens housed in the Herbarium of the Graduate School of Science, Hokkaido University (SAP) and the Department of Botany, National Museum of Nature and Science, Japan (TNS), *L. japonica* has not been collected from its type locality since 1927, and the species has not been collected from the Pacific coast of Japan since 1960 (Figure A in S1 File and Table A in S2 File). The coast of the Miura Peninsula (which includes the type locality of *L. japonica*) has experienced environmental disturbances several times over the past 100 years, including vertical displacement during the 1923 Kanto Earthquake [14], landfill in Tokyo Bay in the 1950s [15], and construction of marinas for the 1964 Summer Olympics in Tokyo [16]. Several seaweeds have disappeared from the peninsula and its vicinity [15, 16], and it seems that *L. japonica* has become extinct at its type locality and vicinity.

Recently, we collected samples similar to *L. japonica* from Sado Island and Oki Island in the Japan Sea (Figure A in S1 File Tables A and B in S2 File). The collection sites are located on the side opposite the Pacific coast of Honshu, Japan, and the reproductive structures of modern samples do not correspond completely to those originally described by Yamada [13]. Therefore, it remains unclear whether modern samples collected from the Japan Sea side of Japan belong to *L. japonica*. To determine the identity of *L. japonica*, we extracted DNA from
topotype material of *L. japonica* collected in 1927 and sequenced nuclear-encoded 28S ribosomal RNA (28S rRNA), plastid-encoded photosystem I P700 chlorophyll a apoprotein A1 (psaA), plastid-encoded ribulose-1,5-bisphosphate carboxylase/oxygenase (rbcL) genes, and the universal mitochondrial barcode marker cytochrome oxidase 1 (COI) using Illumina NGS and determined the phylogenetic position of the type material of *L. japonica*. We also examined herbarium specimens of lectotype and topotype as well as formalin-preserved samples used by Chihara and Yoshizaki [17] for anatomical observations. Both morphological characteristics and molecular analyses indicated that the specimens from the Pacific and Japan Sea coasts are the same species and can therefore be referred to as *L. japonica*. However, our molecular analyses indicated that *L. japonica* is incorrectly placed in *Liagora* and requires assignment to a new genus. We address the generic placement of *L. japonica* and propose a new genus, *Otohimella* gen. nov.

**Materials and Methods**

**Ethics statement**

We collected *Liagora japonica* from Sado Island and Oki Island in Japan. Collection locations and details are shown in Table B in S2 File. The collection site in Sado Island is not a protected area, while the coast of Oki Island lies within the ordinary zone of Daisen-Oki National Park. According to the National Park Act in Japan, there are no restrictions collecting marine algae from the ordinary zones of national parks. In addition, no specific permission was required for the locations on Sado Island and Oki Island.

The specimens of *L. japonica* housed in herbaria were subjected to morphological and molecular investigations. Official permission for this study was obtained from the Herbarium of the Graduate School of Science, Hokkaido University (SAP), and the Department of Botany, National Museum of Nature and Science, Japan (TNS).

**Morphological observations**

Specimens were preserved in Silica gel for DNA extraction, or 10% formalin/seawater for anatomical observations. Voucher herbarium specimens were deposited at the Department of Botany, National Museum of Nature and Science, Japan (TNS). Lectotype and topotype specimen of *L. japonica*, and formalin preserved sample collected from Tsushima used for Chihara and Yoshizaki [17] were also studied. Specimens on herbarium sheets, or preserved in formalin seawater were examined under an Olympus BX50 microscope (Olympus, Japan) after the material had been rehydrated and decalcified (in some cases stained with 1% aniline blue, acidified with 1% HCl and mounted in 50% aqueous Karo syrup (Englewood Cliffs, NJ, USA) with 3% formaldehyde to prevent microbial growth. Drawings were made with the aid of a camera lucida.

**DNA extraction and sequencing procedures for modern specimens**

For phylogenetic analyses, partial 28S rRNA, rbcL genes and COI were sequenced with Sanger sequencing from two modern samples (Table B in S2 File). DNA extractions and sequencing procedures were performed in S1 Text.

**DNA extraction from the historical herbarium specimens**

DNA was extracted from each of the herbarium specimens, ranging in age from 88 years (collected in 1927) to 57 years (collected in 1958), using a class 100 NK System Clean Bench VSF-1600RA (NK Systems, Tokyo, Japan) to prevent microbial and human contamination.
Approximately a 1 × 5 mm² section of material was ruptured manually using a Handy Pestle® (Toyobo, Osaka, Japan) in liquid nitrogen, and DNA was subsequently extracted using a QIA-GEN® Genomic-tip 20/G according to the manufacturer’s protocol (Qiagen, Valencia, CA, USA). The DNA concentration was determined using a Quant-iT dsDNA HS assay kit with a Qubit fluorometer (Life Technologies, Carlsbad, CA, USA).

Aliquot of DNA (40–80 ng) from herbarium specimens were sheared to a target peak size of 500 bp using the Covaris S220 Focused-Ultrasonicator system (Covaris, Woburn, MA, USA) according to the manufacturer’s recommendations. To generate DNA sequencing libraries for high-throughput DNA sequencing, the NEBNext Ultra DNA Library Prep kit for Illumina (New England Biolabs) was used, according to the manufacturer’s instructions with the exception of performing the 10–12 cycles of PCR. The amplified library products (size range, 250–600 bp) were isolated on agarose gels and purified using the NucleoSpin Gel and PCR Clean-up kit (TaKaRa, Kyoto, Japan).

The paired-end reads were generated on the Illumina MiSeq platform using the MiSeq Reagent Kit version 2 (Illumina, San Diego, CA, USA). FASTQ files were generated using the MiSeq Reporter software version 2.3.32 (Illumina). Raw sequence reads, 10,452,985 base pairs (bp) for the 57-year-old Wakayama prefecture specimen and 9,335,303 bp for the 88-year-old Kanagawa prefecture specimen, were generated.

Quality filtering of Illumina sequence data

We discarded the Illumina MiSeq reads that contained ambiguous nucleotides or were mapped to PhiX genomic sequences using Bowtie 2 version 2.2.3 with the default parameters [18]. Subsequently, we removed the adapter sequences from the reads, using Cutadapt version 1.2.135, and low-quality regions with a Phred-like quality score <17 within the 3’ end of the reads. In addition, we discarded reads that were < 20 bp in length or were associated with an average Phred-like quality score < 25. The quality filtering yielded high-quality reads: 10,057,049 bp for the 57-years-old specimen and 8,913,992 bp for the 88-year-old specimen.

Identification of Liagora japonica sequences

The sequences derived from the L. japonica genomes were identified based on two different methods to avoid sequencing error. The first was a read-based method and the other a scaffold-based method.

Read-based method. The high-quality MiSeq reads derived from L. japonica genomes were identified as follows. (1) An in-house nucleotide sequence database consisting of the phylogenetic marker gene sequences of Nemaliales including Liagoraceae and its relatives (designated as in-house Nemaliales database) was constructed by combining previously reported sequence data (Table C in S2 File). (2) All of the high-quality reads were subjected to BLASTN 2.2.27 searches [19] against the in-house Nemaliales database with an E-value < 0.01. (3) The reads that matched the sequences in the in-house Nemaliales database were subjected to BLASTN 2.2.27 searches against the GenBank nucleotide database (September 2014) with an E-value < 0.01. (4) The reads that matched the sequences from Rhodophyta in the GenBank nucleotide database were regarded as genome fragments of L. japonica. The target read coverages of L. japonica are shown in Tables D and E in S2 File.

MiSeq reads identified as sequences belonging to nuclear genes or organelle genes of L. japonica were used for reconstruction of each gene sequence as follows. (1) In each sample, the identified reads were assembled independently for each gene using CAP3 [20]. (2) CLUSTALW was used with the default parameters to generate multiple alignments of the assembled contigs and singletons with reference gene sequences in the in-house Nemaliales database [21].
(3) Partially matched false-positive reads were removed manually by checking the multiple alignment. (4) In each sample, each gene sequence of *L. japonica* was reconstructed manually from the multiple alignment.

**Scaffold-based method.** The high-quality MiSeq reads of each sample were assembled using IDBA-UD version 1.1.0 with the following parameters:—mink 20—maxk 120—step 5 (Table F in S2 File) [22]. Assembled scaffolds were subjected to BLASTN 2.2.27 searches against the in-house Nemaliales database with an *E*-value < 0.0001. The scaffolds that matched the sequences in the in-house Nemaliales database were subjected to BLASTN 2.2.27 searches against the GenBank nucleotide database with an *E*-value < 0.001. The scaffolds that matched the sequences from Rhodophyta in the GenBank nucleotide database were regarded as genome fragments of *Liagora japonica*. The sequences similar to the reference nuclear gene sequences or putative organelle gene sequences were obtained from the scaffolds by (1) identifying a similar region within the reference gene sequence from the BLASTN results and (2) extracting a ±500 bp region of the BLASTN-aligned region. CLUSTAL W was used with default parameters to generate multiple alignments of the extracted sequences with reference gene sequences from the in-house Nemaliales database. Partially matched false-positive sequences were removed manually by checking the multiple alignment. In each sample, each gene sequence of *L. japonica* was manually reconstructed from the multiple alignment. The results of CLUSTALW and manual refinement of phylogenetic marker genes of two samples were checked, and the read coverage was calculated by Bowtie2 (version 2.2.6) mapping of the corresponding sample reads. The read alignments (BAM files) are available from the web server (http://liagora.paleogenome.jp).

**Phylogenetic analysis**

We sequenced two *psaA*, four *rbcL*, four 28S rRNA, and four COI gene sequences from modern samples and historical herbarium specimens of *L. japonica* (Table B in S2 File). *PsaA, rbcL,* and 28S rRNA genes were selected to infer the phylogeny of Liagoraceae, and the COI gene was selected to assess the effectiveness of DNA barcoding. The sequence data of Liagoraceae, available from GenBank, were compiled. *PsaA* sequences for 55 taxa, *rbcL* sequences for 99 taxa, and COI sequences for 22 taxa were aligned using CLUSTAL W [21]. The 28S rRNA gene sequences for 51 taxa of Nemaliales were aligned using CLUSTAL W [21] and were refined based on published secondary structures of the 28S rRNA gene of *Palmaria palmata* (Linnaeus) Kuntze [23] using SeaView 4.1 [24]. The ambiguous regions of the alignments were removed. Samples with identical nucleotide sequences were treated as a single operational taxonomic unit (OTU). As the Liagoraceae has been resolved previously as a monophyletic group [25], four to five other families belonging to the Nemaliales were designated as outgroups for *psaA, rbcL,* combined *psaA* and *rbcL,* and 28S rRNA gene analyses. Based on the results of *psaA, rbcL,* combined *psaA* and *rbcL,* and 28S rRNA gene analyses, the genera *Cumagloia, Hommersandidiophycus,* and *Nemalion* were designated as outgroups for COI analysis. For confirmation of the identification of 18S rRNA, *psaB,* and *psbA* genes derived from the historical specimens of *L. japonica,* phylogenetic analyses based on those genes were performed. The alignments used for the present phylogenetic analyses are available from TreeBASE at https://treebase.org/treebase-web/search/study/summary.html?id=18829 (matrix accession number S18829).

Phylogenetic analyses of the aligned sequences from each dataset were subjected to Bayesian inference (BI) and maximum likelihood (ML) analysis. The substitution models applied to BI analyses are shown in Table G in S2 File. BI analysis was performed using MrBayes 3.2.1 [26], as described previously [27]. Four chains of Markov chain Monte Carlo (MCMC)
iterations were carried out for 2,000,000 or 3,000,000 generations, keeping one tree every 500 generations. Convergence of log-likelihood and parameter values was assessed in Tracer version 1.4. [28]. A burn-in sample of 5,000–7,500 trees was removed before constructing the majority rule consensus tree, and the remaining trees were used to calculate a 50% majority-rule tree and to determine the posterior probabilities (PP) of the individual branches. The Bayesian analyses are summarized in Table G in S2 File. ML analysis was performed using RAxML version 7.0.4 software [29]. The GTR+I+Γ model was applied to each dataset in the analysis. Bootstrap values (BP) for ML analysis were calculated based on 1000 pseudoreplicates. The p distances and K2P genetic distances for each pair of liagoracean species were calculated using PAUP 4.0b10 [30].

### Nucleotide sequence accession numbers

The Sanger sequence datasets have been submitted to DDBJ under accession numbers LC066217 to LC066223, and LC066521 to LC066532 and LC093491 to LC093498, and the Illumina sequence datasets have been submitted to the DDBJ Short Read Archive under accession number DRA003813. IDBA-UD assemblies of two samples can be accessed under BCQK01000001-BCQK01275014 (suzuki-1) and BCQL01000001-BCQL01381344 (suzuki-2), respectively.

### Nomenclature acts

The electronic version of this article in Portable Document Format (PDF) in a work with an ISSN or ISBN will represent a published work according to the International Code of Nomenclature for algae, fungi, and plants, and hence the new names contained in the electronic publication of a PLOS ONE article are effectively published under that Code from the electronic edition alone, so there is no longer any need to provide printed copies.

In addition, new names contained in this work have been submitted to IPNI, from where they will be made available to the Global Names Index. The IPNI LSIDs can be resolved and the associated information viewed through any standard web browser by appending the LSID contained in this publication to the prefix http://ipni.org/. The online version of this work is archived and available from the following digital repositories: PubMed Central, LOCKSS.

### Results

#### Sequences determined from historical specimens

Based on limited publicly available sequence data from the Nemaliales, including Liagoraceae (Table C in S2 File), reliable sequences from *Liagora japonica* were determined for four genes: 28S rRNA, *psaA*, *rbcL*, and COI (Fig 1, Figures B-E in S1 File). Another three genes, nuclear-encoded 18S rRNA, putative plastid-encoded photosystem I P700 chlorophyll a apoprotein A2 (*psaB*), and putative plastid-encoded photosystem II core 32 kDa protein (*psbA*) genes of potentially nemalialean affinity, were obtained from the historical specimens (Figure F in S1 File, Table B in S2 File). Both the read-based and scaffold based methods, the determined sequences of the seven genes were the same. The *psaA*, *psaB*, *psbA*, *rbcL*, and COI gene sequences were complete, while only partial sequences of the 28S rRNA and 18S rRNA genes were available. The length of the 28S and 18S rRNA gene sequences were 82.8% and 91.5%, respectively, of the published complete 28S and 18S rRNA gene sequence of *P. palmata* [23].

#### Phylogenetic markers

Analysis of the *rbcL* and COI sequences identified few polymorphisms (Figures B and E in S1 File) among populations of *L. japonica* from Misaki, type locality, Wakayama, Oki Island, and
Fig 1. Bayesian tree based on combined *psaA* and *rbcL* gene sequences. Numbers on the branches indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP). Note that the trifurcation (asterisk) represents lack of bifurcation with 0.50 or more PP values in BI.

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Sado Island. The rbcL gene of *L. japonica* showed 0–4 bp variation. The p distance among the species of the genus *Liagora* sensu stricto used for rbcL was 5.5–10.9%, whereas that among populations of *L. japonica* was < 0.3%. The COI gene for *L. japonica* showed 0–9 bp variation. The p distance among the species of the genus *Liagora* sensu stricto used for COI was 7.8–13.5%, whereas that among populations of *L. japonica* was < 1.5%. The p distances and K2P genetic distances among the liagoracean species used in COI analysis are shown in Tables H and I in S2 File. Both the p distances and K2P genetic distances generated similar results.

**Phylogenetic analyses**

As psaA, rbcL, and 28S rRNA genes have been used for phylogenetic analyses of Liagoraceae [9–11, 25], we performed, psaA, rbcL, combined psaA and rbcL, and 28S rRNA gene analyses. To determine whether the third nucleotides of the codons of the psaA and rbcL genes exhibit saturation of substitutions, p distances among liagoracean species based on the third nucleotide of codons were compared with those of the first and second nucleotides of the same codons (Figure G in S1 File). The data indicated that these third nucleotide positions have been saturated with substitutions in liagoracean species, but not in *L. japonica* or four related genera: Akalaphycus, Macrocarpus, Neoizziella, and Titanophycus.

The topologies of individual psaA, rbcL, and combined psaA and rbcL trees, except the position of *L. japonica*, were basically similar to previous phylogenetic analyses for Liagoraceae [9–11, 25, 31, 32]. The phylogenetic trees generated by BI and ML showed the same topologies; therefore, we present only the BI tree topology (Fig 1, Figures B and C in S1 File). BI topologies of the individual psaA and rbcL trees (Figures B and C in S1 File) were similar to the combined psaA and rbcL data (Fig 1), except for the position of Neoizziella, but with weaker statistical support. In the individual rbcL tree, *L. japonica* and *Macrocarpus* were sisters to each other, with low support (< 0.95 PP and 59% BP), while in the individual psaA and combined psaA and rbcL trees, *L. japonica* and Neoizziella were sisters to each other, with moderate to high support (0.98–0.99 PP and 79–81% BP). In the combined psaA and rbcL tree, the monophyly of each genus belonging to Liagoraceae was highly supported (1.00 PP and > 95% BP), excluding Iziella. *Liagora japonica*, *Macrocarpus*, and Neoizziella formed a monophyletic clade with full support (1.00 PP and 100% BP), whereas *L. japonica* was clearly separated from *Liagora* sensu stricto.

The topology of the 28S rRNA gene tree, except the position of *L. japonica* was similar to previous phylogenetic analyses for Liagoraceae [10, 12, 33]. In the tree based on the 28S rRNA gene, *Liagora japonica* was separated from *Liagora* sensu stricto; however, the boundaries of the genera belonging to Liagoraceae were not resolved (Figure D in S1 File).

**Morphological observations of modern samples**

Thalli were found on rocks at a depth of approximately 1–3 m (Fig 2A). The thalli were erect, 4–7 cm in height, and composed of 2–3 terete axes branched subdichotomously to 5–7 orders, arising from a small discoid holdfast, loosely calcified, pinkish or reddish brown in color (Fig 2A and 2B).

The thalli were multiaxial and composed of assimilatory filaments and medullary filaments (Fig 2C). The assimilatory filaments were subdichotomously branched 3–4 times. The upper parts of the assimilatory filaments were composed of ellipsoidal cells, which were 10–18 μm long and 5–8 μm wide. The lower parts were composed of elongated cells, which were 30–60 μm long and 4–6 μm wide.
Fig 2. Morphological examination of modern samples of *Liagora japonica* Yamada. A: Habit (TNS-AL 195934). Scale bar = 1.0 cm. B: Herbarium specimen (TNS-AL 185628). Scale bar = 2.0 cm. C: Assimilatory filaments (TNS-AL 182118). Scale bar = 50 μm. D: Spermatangia (s) cut off from spermatangial parental cell (spc) (TNS-AL 190026). Scale bar = 10 μm. E: 4-celled carpogonial branch (TNS-AL 190026). cp = carpogonium. sc = supporting cell. Scale bar = 10 μm. F: 5-celled carpogonial branch (TNS-AL 182118). Scale bar = 30 μm. G: An early post-fertilization stage showing gonimoblast initial (gi), gonimoblast cells (arrowhead), and involucral filaments (arrows) (TNS-AL 182118). Scale bar = 20 μm. H: A later post-fertilization stage showing developing gonimoblast cells (arrowheads) and involucral filament (arrows) (TNS-AL 182118). Scale bar = 30 μm. I: Young carposporophyte showing the growth of the gonimoblast cells (arrowheads) and involucral filaments (arrows) (TNS-AL 182118). Note that the cells of carpogonial branch (cb) are not fused. Scale bar = 50 μm. J: Mature carposporophyte showing diffuse carposporophyte (TNS-AL 182118). Note that the cells of carpogonial branch remain distinct. Scale bar = 50 μm. K: Carposporangia (TNS-AL 190026). Note that carposporangia are not divided. Scale bar = 10 μm.

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Gametophytes were monoecious. Spermatangial parent cells derived from apical cells and spermatangia were cut off terminally (Figs 2D and 3A). One or, rarely, two spermatangia were cut off from each spermatangial parental cell.

Carpogonial branches were curved slightly, 4- or 5-celled, and borne on the middle part of assimilatory filaments (Figs 2E and 2F, 3B and 3C). After presumed fertilization, the carpogonium divided transversely to produce a gonimoblast initial, after which the gonimoblast initial cut off primary gonimoblast cells obliquely (Fig 2G). Meanwhile, the involucral filaments were initiated mostly from cortical cells on the supporting cell (Figs 2G, 3D and 3E). Gonimoblast initials cut off second and third gonimoblast cells, and the gonimoblast developed radially (Figs 2H, 3D and 3E). At an early stage of gonimoblast development, growth of the involucral filaments and gonimoblast cells became dominant (Fig 2I). At maturity, the gonimoblast cells were embedded in and intermingled with the involucral filaments, and carposporophytes were diffuse (Figs 2J and 3F). The cells of the carpogonial branch were not fused through carposporophyte development (Figs 2G–2J and 3D–3F). Carposporangia were elliptical to oblong, 10–15 μm long and 5–6 μm wide (Fig 2K).

Morphological observations of historical materials
Based on habit and vegetative anatomies, the lectotype and topotype specimens examined corresponded to the descriptions of Yamada [13], while samples collected from Tsushima fit those of Chihara and Yoshizaki [17]. Thalli were erect and bushy, consisting of 3–5 main axes, subdi- chotomously branched, with 3–9 orders of branching, 7–18 cm in height, arising from a discoid holdfast with moderately calcified branches (Fig 4A and 4B). After fertilization, cells of the carpogonial branch were not fused. At an early stage of gonimoblast development, gonimoblast cells were loosely elongated, and involucral filaments intermingled with gonimoblast cells (Fig 4C). Mature carposporophytes were not observed in the lectotype or topotype, while those of Tsushima were diffuse (Fig 4D).

Discussion
The rbcL and COI have been used for species delineation of Nemaliales, including Liagoraceae [9, 11, 34]. Analyses of rbcL and COI sequences suggest that modern and historical specimens collected from four localities of both the Japan Sea and Pacific sides of Japan, including topotypes, are very closely related. The typical intraspecific divergence of rbcL of the Liagoraceae based on p distances is < 0.4% [31], while that of COI of the Nemaliales is < 1.0% [34]. Liagora japonica showed < 0.3% intraspecific divergence of rbcL, which was within the range of that of Liagoraceae [31], while the intraspecific divergence of COI was < 1.5%, which was higher than typical intraspecific divergence of the Nemaliales [34]. The typical minimum interspecific divergence of COI of the Liagoraceae is 5.6% [32], and thus the intraspecific divergence of L. japonica is much lower than the interspecific divergence. Thus, all specimens should be identified as a single species: L. japonica.

However, the structures of carposporophytes from modern samples differed from those originally described by Yamada [13]. Yamada [13] presented a drawing of compact carposporophytes, whereas carposporophytes of the modern samples were diffuse. Furthermore, Chihara and Yoshizaki [17] observed L. japonica collected from Tsushima, Japan, and also presented a drawing of compact carposporophytes, similar to Yamada [13]. However, the present examination of lectotype and topotype specimens, as well as specimens used by Chihara and Yoshizaki [17], showed elongated gonimoblast cells and involucral filaments intermingled with gonimoblast cells, similar to the carposporophyte development of diffuse carposporophytes (Table 1). The habit and vegetative structures of the modern samples collected from the Japan Sea.
Fig 3. Reproductive structures of modern samples of *Liagora japonica* Yamada. A: Spermatangia (s) cut off from spermatangial parental cell (spc) (TNS-AL 190026). Scale bar = 10 μm. B: 4-celled carpogonial branch (TNS-AL 190026). cp = carpogonium. sc = supporting cell. Scale bar = 10 μm. C: 5-celled carpogonial branch (TNS-AL 190026). Scale bar = 10 μm. D: An early post-fertilization stage showing gonimoblast cells (arrowhead) and involucral filaments (arrows) (TNS-AL 190026). Scale bar = 10 μm. E: A later post-fertilization stage showing developing gonimoblast cells (arrowheads) and involucral filaments (arrows) (TNS-AL 182118). Scale bar = 30 μm. F: Mature carposporophyte showing diffuse carposporophyte (TNS-AL 182118). Scale bar = 50 μm.

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correspond to the original description of Yamada [13]. Thus, Yamada [13] and Chihara and Yoshizaki [17] did not recognize diffuse carposporophytes, because those of the genus *Liagora* sensu lato were not recognized until the 1990s [35, 36].

Liagoracean genera have been recognized based on the characteristics of the structures of carposporophyte development including (1) carpogonial branches and whether they form a fusion cell; (2) the origin of the involucral filaments, a specially produced sterile cell; (3) association of the gonimoblast with paraphyses, a specially produced assimilatory filament around the carposporophyte; (4) compact or diffuse carposporophytes; and (5) division of carposporangia [9–12, 34]. Lin et al. [9–11] suggested that the genus *Liagora* is polyphyletic and should be separated into several genera based on the characteristics of carposporophyte development. Recent taxonomic studies of *Liagora* sensu stricto, including the type species *L. viscosa* (Forsskål) C. Agardh, suggested that *Liagora* sensu stricto is characterized by fused carpogonial branching and diffuse carposporophyte [9, 37]. Three new genera, *Macrocarpus*, *Neoizziella*, and *Yoshizakia* including species with unfused carpogonial branches and diffuse carposporophytes, were separated from...
Liagora sensu stricto [9, 31]. The other liagoracean genera, Akalaphycus, Patenocarpus, and Stenopeltis, also produce diffuse carposporophytes; however, they produce paraphyses [33, 38]. Liagora japonica produces unfused carpogonial branches and diffuse carposporophytes without paraphyses, which are similar to those of Macrocarpus, Neoizziella, and Yoshizakia. Macrocarpus is characterized by divided carposporangia, while Neoizziella is characterized by the position of involucral filaments, which are produced from cortical cells in the vicinity of the supporting cell [9]. Liagora japonica differs from Macrocarpus and Neoizziella by undivided carposporangia.

Table 1. Morphological comparison among the description of Yamada [13] and Chihara and Yoshizaki [17], type specimens, and related samples with Liagora japonica.

| Specimen | Yamada [13] | Chihara and Yoshizaki [17] | OJ5 | OJ1 | OJ3 | OJ4 | OJ10 |
|----------|-------------|-----------------------------|-----|-----|-----|-----|------|
| Locality | Misaki      | Tsushima                    | Misaki | Misaki | Sado Island | Oki Island | Tsushima |
| Accession No. of rbcL gene sequence | - | - | LC066217 | LC066219 | LC066220 | - |
| Fusion of the cells of carpogonial branch | Not observed | Absent | Absent | Absent | Absent | Absent | Absent |
| Development of gonimoblast cells | Not observed | Not observed | Elongated | Elongated | Elongated | Elongated | Elongated |
| Development of involucral filaments | Intermingled with gonimoblast cells | Intermingled with gonimoblast cells | Intermingled with gonimoblast cells | Intermingled with gonimoblast cells | Intermingled with gonimoblast cells | Intermingled with gonimoblast cells |
| Shape of carposporophyte | Rather loose | Globular aggregation | Not observed | Not observed | Diffuse | Diffuse | Diffuse |

aVoucher specimen of Chihara & Yoshizaki [17].
bYamada’s [13] Fig.10D shows compact carposporophyte.

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Table 2. Comparisons of features distinguishing among Liagora japonica, Liagora sensu stricto, and the genera with diffuse carposporophyte belonging to Liagoraceae.

| Liagora japonica | Liagora sensu stricto | Akalaphycus | Macrocarpus | Neoizziella | Patenocarpus | Stenopeltis | Yoshizakia |
|------------------|-----------------------|-------------|-------------|-------------|--------------|-------------|------------|
| Type species     | -                     | L. viscida  | A. setchelliae | M. perennis | N. asiatica  | P. paraphysiferus | S. gracilis | Y. indopaciica |
| Association of gonimoblast with paraphyses | Absent | Absent | Present | Absent | Absent | Present | Present | Absent |
| Fusion of cells of carpogonial branch after fertilisation | Absent | Present | Absent | Absent | Absent | Absent | Absent | Absent |
| Involucral filament | Present | Present | Absent | Present | Present | Absent | Present | Present |
| Origin of involucral filaments | Cortical cell on the supporting cell | Cortical cells in the vicinity of the supporting cell or cortical cell on the supporting cell | N.A. | Cortical cell on the supporting cell | Cortical cells in the vicinity of the supporting cell | Cortical cells in the vicinity of the supporting cell | N.A. | Cortical cell on the supporting cell |
| The gonimoblast intermingled with the involucral filaments | Present | Present | N.A. | Present | Present | Present | Present | N.A. |
| Division of carposporangia | Absent | Absent or present | Absent | Present | Absent | Absent | Absent | Absent |

References This study [10, 31, 37] [34] [9] [9] [39] [33] [31]

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and involucral filaments produced from the cortical cell on the supporting cell (Table 2). The critical features of carposporophyte development of *L. japonica* are most similar to those of *Yoshizakia*. *Yoshizakia* is characterized by involucral filaments, which are not intermingled with gonimoblast filaments. *Liagora japonica*, however, has involucral filaments that intermingle with gonimoblast filaments. Individual *psaA* and *rbcl*, and combined *psaA* and *rbcl* analyses suggested that *L. japonica*, *Macrocarpus*, and *Neoizziella* form a monophyletic clade separated from *Liagora sensu stricto*. However, *L. japonica* was not included in the clade of *Macrocarpus* or *Neoizziella*. Unfortunately, *psaA* sequence data for the species of *Yoshizakia* are not yet available, but individual *rbcl* analyses suggest that *L. japonica* is clearly separated from *Yoshizakia*. Both morphological and molecular results suggest that *L. japonica* is a distinct genus in the Lia-goraceae, and we propose a new genus, *Otohimella* Mas. Suzuki, to accommodate this species.

**Taxonomic Treatment**

*Otohimella* Mas. Suzuki gen. nov.

Description: Thalli are moderately calcified, and 5 – 18 cm in height, arising from a discoid holdfast with a short stipe. Thalli are subdichotomously branched to 5 or 6 orders. The cells of assimilatory filaments are ovoid to ellipsoidal and borne on colorless medullary filaments. Gametophytes are monoeocious. Spermatangia are produced terminally on spermatangial parent cells. Carpogonial branches are slightly curved, and 4- or 5-celled. After fertilization, cells of the carpogonial branch are not fused through carposporophyte development. The involucral filaments are initiated mostly from the cortical cell on the supporting cell and eventually intermingled with the gonimoblast filaments. Mature carposporophytes are diffuse. Carposporangia are formed at the distal ends of gonimoblast filaments and are not divided.

Generitype: *Otohimella japonica* (Yamada) Mas. Suzuki, T. Segawa, Hi. Mori et Nozaki comb. nov.

Basionym of *O. japonica*: *Liagora japonica* Yamada. Sci. Pap. Inst. Algol. Res., Fac. Sci., Hokkaido Imp. Univ. 2: 16–17 Fig 9 and 10, Pl. IV (1938).

Etymology: Named for Otohime, a sea goddess in Japanese mythology.

**Conclusion**

NGS can be used to determine DNA sequences from historical herbarium specimens containing highly fragmented DNA molecules. The liagoracean species are non-model organisms; thus, there are no useful complete genome sequences in the GenBank database for reference mapping and sequence assembly of NGS. However, we were able to determine seven gene sequences from historical herbarium specimens of *L. japonica* using NGS with limited reference sequence data. Four of the seven genes can be used for phylogenetic analyses and species identification. This study showed that sequencing of historical specimens using NGS is a powerful tool for systematics and identification of not only model, but also non-model, organisms.

We addressed the identity of *L. japonica* based on both morphological and molecular data, including those of the lectotype and topotype specimens. The species had been considered extinct on the Pacific Ocean side of Japan. However, we showed that *L. japonica* survives on the Japan Sea side of Japan. Further, we propose *Otohimella japonica* gen. et comb. nov. based on this species.

**Supporting Information**

S1 File. Figure A. Geographical distribution of *Liagora japonica* based on the herbarium specimen deposited in SAP and TNS. Detail of collection data is shown in Tables A and B in S2 File. Figure B. Bayesian tree based on *psaA* gene sequences. Numbers on the branches
indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP).

**Figure C. Bayesian tree based on rbcL gene sequences.** Numbers on the branches indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP). Note that the trifurcation (asterisk) represents lack of bifurcation with 0.50 or more PP values in BI.

**Figure D. Bayesian tree based on 28S rRNA gene sequences.** Numbers on the branches indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP). Note that the trifurcation (asterisk) represents lack of bifurcation with 0.50 or more PP values in BI.

**Figure E. Bayesian tree based on COI gene sequences.** Numbers on the branches indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP). Note that the trifurcation (asterisk) represents lack of bifurcation with 0.50 or more PP values in BI.

**Figure F. Bayesian tree based on 18S rRNA (A), psaB (B), and psbA (C) gene sequences.** Numbers on the branches indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP). Note that the trifurcation (asterisk) represents lack of bifurcation with 0.50 or more PP values in BI.

**Figure G. Comparison of p distances among the liagoracean species based on the first and second nucleotides of codons and based on the third nucleotide of codons in the combined psaA and rbcL dataset used for the present phylogenetic analyses (Fig 1).** Red diamonds indicate p distances among "Liagora" japonica, Akalaphycus, Macrocarpus, Neoizziella, and Titanophyca.

(PDF)

**S2 File.** Table A. Collection and herbarium information for specimens of Liagora japonica used in the morphological analyses that have no molecular data. Table B. Collection locations and details, and GenBank accession numbers of samples used in the psaA, psaB, psbA, rbcL, COI, 18S rRNA, and 28S rRNA genes analyses. Table C. GenBank accession numbers of species used in the identification of Liagora japonica sequences from old herbarium specimens. Table D. Target read coverage of Liagora japonica (sample ID: suzuki-2; sample No.: OJ1). Table E. Target read coverage of Liagora japonica (sample ID: suzuki-1; sample No.: OJ2). Table F. De novo assembly statistics. Table G. Summary for the Bayesian analyses on the basis of psaA, psaB, psbA, rbcL, 18S rRNA, and 28S rRNA datasets. Table H. Matrix of p distances of the liagoracean species used in the COI analysis. Table I. Matrix of Kimura 2-parameter (K2P) genetic distances of the liagoracean species used in the COI analysis.

(DOC)

**S1 Text.** The DNA extraction and sequencing procedures for modern specimens.

(DOCX)

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**Author Contributions**
Conceived and designed the experiments: MS TS HN. Performed the experiments: MS TS AA. Analyzed the data: HM TS RO MS. Contributed reagents/materials/analysis tools: MS TS HM AA AK HS TK TA KK HK HN. Wrote the paper: MS TS HM HN.

**References**

1. Boo GH, Park JK, Boo SM. *Gelidiophycus* (Rhodophyta: Gelidiales): A new genus of marine algae from East Asia. Taxon 2013; 62: 1105–1116.

2. Lin S-M, Yang W-C, Huisman J, De Clerck O, Lee WJ. Molecular phylogeny of the widespread *Martensia fragilis* complex (Delesseriaceae, Rhodophyta) from the Indo-Pacific region reveals three new species of *Martensia* from Taiwan. Euro J Phycol 2013; 48: 173–187.

3. Carlile AL, Cho TO, Waaland JR. The conspecificity of *Ceramium pacificum* and *Ceramium washingtoniense* (Ceramiaceae, Rhodophyta). Phycologia 2010; 49: 336–344.

4. Hind KR, Gabrielson PW, Lindstrom SC, Martone PT. Misleading morphologies and the importance of sequencing type specimens for resolving coralline taxonomy (Corallinales, Rhodophyta): *Pachyarthron cretaceum* is *Corallina officinalis*. J Phycol 2014; 50: 760–764. doi:10.1111/jpy.12205 PMID: 26988460

5. Dadney J, Meyer M, Pääbo S. Ancient DNA damage. Cold Spring Harb Perspect Biol 2013; 5: a012567. doi:10.1101/cshperspect.a012567 PMID: 23729639

6. Thalmann O, Shapiro B, Cui P, Schuenemann VJ, Sawyer SK, Greenfield DL, et al. Complete Mitochondrial Genomes of Ancient Canids Suggest a European Origin of Domestic Dogs. Science 2013; 342: 871–874. doi:10.1126/science.1243650 PMID: 24233726

7. Hughey JR, Gabrielson PW, Rohmer L, Tortolani J, Silva M, Miller KA, et al. Minimally destructive sampling of type specimens of *Pyropia* (Bangiales, Rhodophyta) recovers complete plastid and mitochondrial genomes. 2014; Sci Rep 4: 5113. doi:10.1038/srep05113 PMID: 24894641

8. Guiry MD, Guiry GM. AlgaeBase. World-wide electronic publication, National University of Ireland, Galway. 2015; 10: 28. Available: http://www.algaebase.org.

9. Lin S-M, Yang S-Y, Huisman JM. Systematics of *Liagora* with diffuse gonimoblasts based on *rbcL* sequences and carposporophyte development, including the description of the new genera *Neoizziella* and *Macrocarpus* (Liagoraceae, Rhodophyta). Eur J Phycol 2011; 46: 249–262.

10. Lin S-M, Yang S-Y, Huisman JM. Systematic revision of the genus *Liagora* and *Izziella* (Liagoraceae, Rhodophyta) from Taiwan based on molecular analyses and carposporophyte development, with the description of two new species. J Phycol 2011; 47: 352–365. doi:10.1111/j.1529-8817.2011.00965.x PMID: 27021867

11. Lin S-M, Huisman JM, Ballantine DL. Revisiting the systematics of *Ganonema* (Liagoraceae, Rhodophyta) with emphasis on species from the northwest Pacific Ocean. Phycologia 2014; 53: 37–51.

12. Huisman JM, Saunders GM, Sherwood AR. Recognition of *Titanophycus*, a new genus based on *Liagora valida* Harv. (Liagoraceae, Nemaliales). In: Huisman JM, editors. Algae of Australia—Nemaliales: ABRS; 2006. pp. 116–119.

13. Yamada Y. The species of *Liagora* from Japan. Sci Pap Inst Algol Res, Fac Sci, Hokkaido Imp Univ 1938; 2: 1–34.

14. Yoshida A, Harada M, Odawara K. Vertical displacement of the seabed of Sagami Bay at the 1923 Kanto earthquake. Bull Hot Springs Res Inst Kanagawa Pref 2012; 44: 17–28.

15. Tanaka J, Otori Y. Marine algae collected from Honmoku, Yokohama in Tokyo Bay in 1945–1954. Nat Hist Rep Kanagawa 1988; 19: 105–109.

16. Yokosawa T. Sourui-Saisyuti-Ananai Enoshima (Fujisawa, Kanagawa). Jpn J Phycol 2008; 56: 213–216.

17. Chihara M, Yoshizaki M. Reproductive system of *Liagora japonica* (Nemaliales, Rhodophyta). Bull Natn Sci Mus Tokyo 1972; 15: 395–401.

18. Langmead B, Salzberg SL. Fast gapped-read alignment with Bowtie 2. Nat Methods 2012; 9: 357–359. doi:10.1038/nmeth.1923 PMID: 22388286
19. Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, Miller W, et al. Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. Nucleic Acids Res 1997; 25: 3389–3402. PMID: 9254694

20. Huang X, Madan A. CAP3: A DNA sequence assembly program. Genome Res 1999; 9: 868–877. PMID: 10508846

21. Larkin MA, Blackshields G, Brown NP, Chenna R, McGettigan PA, McWilliam H, et al. Clustal W and Clustal X version 2.0. Bioinformatics 2007; 23: 2947–2948. PMID: 17846036

22. Peng Y, Leung HC, Yiu SM, Chin FY. IDBA-UD: a de novo assembler for single-cell and metagenomic sequencing data with highly uneven depth. Bioinformatics 2012; 28: 1420–1428. doi: 10.1093/bioinformatics/bts174 PMID: 22495754

23. Wuyts J, van den Peer Y, Winkelmans T, De Wachter R. The European database on small subunit ribosomal RNA. Nucleic Acids Res 2002; 30: 183–185. PMID: 11752288

24. Galtier N, Gouy M, Gautier C. 1996. SEAVIEW and PHYLO_WIN: Two graphic tools for sequence alignment and molecular phylogeny. Comput Appl Biosci 1996; 12: 543–548.

25. Lin S-M, Rodríguez-Prieto C, Huisman JM, Guiry MD, Payri C, Nelson WA, et al. A phylogenetic re-appraisal of the family Liagoraceae sensu lato (Nemaliales, Rhodophyta) based on sequence analyses of two plastid genes and postfertilization development. 2015; J Phycol 51: 546–559. doi: 10.1111/jpy.12301 PMID: 26986669

26. Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, et al. MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. Syst Biol 2012; 61: 539–542. doi: 10.1093/sysbio/sys029 PMID: 22357727

27. Suzuki M, Nozaki H, Terada R, Kitayama T, Hashimoto T, Yoshizaki M. Morphology and molecular relationships of Liagora leptophylla comb. nov. (Rhodymeniales, Rhodophyta) from Japan. Phycologia 2012; 51: 479–488.

28. Rambaut A, Drummond AJ. Tracer v1.4. 2007; Available: http://beast.bio.ed.ac.uk/Tracer.

29. Stamatakis A, Hoover P, Rougemont J. A rapid bootstrap algorithm for the RAxML web-servers. Syst Biol 2008; 75: 758–771.

30. Swoford DL. Paup*. Phylogenetic Analysis Using Parsimony (* and Other Methods). Version 4. Sinauer Associates, Sunderland, MA; 2002.

31. Lin S-M, Huisman JM, Payri C. Characterization of Liagora ceranoides (Liagoraceae, Rhodophyta) on the basis of rbcL sequence analyses and carposporophyte development, including Yoshizakia indopacifica gen. et sp. nov. from the Indo-Pacific region. Phycologia 2013; 52: 161–170.

32. Popolizia TR, Schneider CW, Lane CE. A molecular evaluation of the Liagoraceae sensu lato (Nemaliales, Rhodophyta) in Bermuda including Liagora nesophila sp. nov. and Yamadaelia grassyi sp. nov. J Phycol 2015; 51: 637–658. doi: 10.1111/jpy.12306 PMID: 26986788

33. Huisman JM, Abbott IA, Sherwood AR. Large subunit rDNA gene sequences and reproductive morphology reveal Stenopeltis to be a member of the Liagoraceae (Nemaliales, Rhodophyta), with a description of Akalaphycus gen. nov. Eur J Phycol 2004; 39: 257–272.

34. Le Gall L, Saunders GW. Establishment of a DNA-barcode library for the Nemaliales (Rhodophyta) from Canada and France uncovers overlooked diversity in the species Nemalion helminthoides (Velley) Batters. 2010; Cryptogamie Algol 31: 403–421.

35. Abbott IA. A new "tetrasporangial" species of Liagora (Rhodophyta, Nemaliales) from Hawaii. Chinese J Limnol Oceanogr 1995; 13: 343–347

36. Huisman JM, Wynne MJ. Liagora tsengii sp. nov. (Liagoraceae, Nemaliales) from the Lesser Antilles, West Indies. Bot Mar 1999; 42: 219–225.

37. Huisman JM. The type and Australian species of the red algal genera Liagora and Ganonema (Liagoraceae, Nemaliales). Austral Syst Bot 2002; 15: 773–838.

38. Yoshizaki M. The structure and reproduction of Patenocarpus paraphysiferus gen. et sp. nov. (Dermo-nemataceae, Nemaliales, Rhodophyta). Phycologia 1987; 26: 47–52.
Figure A. Geographical distribution of *Liagora japonica* based on the herbarium specimen housed in SAP and TNS. Detail of collection data is shown in S1 and S2 Tables.
Figure B. Bayesian tree based on psaA gene sequences. Numbers on the branches indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP).
Figure C. Bayesian tree based on *rbcl* gene sequences. Numbers on the branches indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP). Note that the trifurcation (asterisk) represents lack of bifurcation with 0.50 or more PP values in BI.
Figure D. Bayesian tree based on 28S rRNA gene sequences. Numbers on the branches indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP). Note that the trifurcation (asterisk) represents lack of bifurcation with 0.50 or more PP values in BI.
Figure E. Bayesian tree based on COI gene sequences. Numbers on the branches indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP). Note that the trifurcation (asterisk) represents lack of bifurcation with 0.50 or more PP values in BI.
Figure F. Bayesian tree based on 18S rRNA (A), psaB (B), and psbA (C) gene sequences. Numbers on the branches indicate the corresponding posterior probabilities (PP, left) from Bayesian analysis and bootstrap values (BP, right) from maximum likelihood analysis. Only the PP (≥ 0.95) and BP (≥ 50%) are shown. The thick branches represent highest statistic supports (1.00 PP and 100% BP). Note that the trifurcation (asterisk) represents lack of bifurcation with 0.50 or more PP values in BI.
Figure G. Comparison of $p$ distances among the liagoracean species based on the first and second nucleotides of codons and based on the third nucleotide of codons, in the combined *psaA* and *rbcL* dataset used for the present phylogenetic analyses (Fig. 1). Red diamonds indicate $p$ distances among “Liagora” *japonica* and four related genera: *Akalaphycus*, *Macrocarpus*, *Neoizziella*, and *Titanophycus*. 
Table A. Collection and herbarium information for specimens of *Liagora japonica* used in the morphological analyses that have no molecular data.

| Sample No. | Collection data |
|------------|-----------------|
| OJ5        | Misaki (35°15' N, 139°36' E), Miura, Kanagawa Prefecture, Japan; May 1903; TI; lectotype |
| OJ6        | Oura (34°40' N, 138°56' E), Shimoda, Shizuoka Prefecture, Japan; 15 May 1952; coll. Mitsuo Chihara; TNS-AL 047542 |
| OJ7        | Kisami (34°39' N, 138°55' E), Shimoda, Shizuoka Prefecture, Japan; 26 Mar. 1959; TNS-AL 029956 |
| OJ8        | Hamashima (34°17' N, 136°45' E), Mie Prefecture, Japan; 1 Apr. 1938; coll. Michitaro Higashi; TNS-AL 158161 |
| OJ9        | Kushimoto (33°26' N, 135°45' E), Higashimuro County, Wakayama Prefecture, Japan; 29 Apr. 1960; coll. Torao Yamamoto; TNS-AL 150505 |
| OJ10       | Tsutsu (34°06' N, 129°10' E), Tsushima, Nagasaki Prefecture, Japan; 17 Mar. 1969; coll. Mitsuo Chihara & Makoto Yoshizaki; TNS-AL 029955\(^a\) |
| OJ11       | Nishiura (34°06' N, 129°10' E), Tsutsu, Tsushima, Nagasaki Prefecture, Japan; 15 Mar. 1969; coll. Mitsuo Chihara & Makoto Yoshizaki; TNS-AL 029954 |
| OJ12       | Uragawa (38°12' N, 138°29' E), Sado, Niigata Prefecture, Japan; 21 Feb. 2003; coll. Masahiro Suzuki; TNS-AL 185627 |
| OJ13       | Uragawa (38°12' N, 138°29' E), Sado, Niigata Prefecture, Japan; 7 Nov. 2003; coll. Masahiro Suzuki; TNS-AL 185628 |
| OJ14       | Tamasaki (38°09' N, 138°27' E), Sado, Niigata Prefecture, Japan; 26 Jul. 1998; coll. Makoto Yoshizaki; TNS-AL 190026 |
| OJ15       | Sai (41°27' N, 140°52' E), Shimokita County, Aomori Prefecture, Japan; 25 Oct. 1987; coll. Taiju Kitayama; SAP 53016\(^b\) |

\(^a\)Voucher specimen of Chihara and Yoshizaki (1972).

\(^b\)165 bp of *rbcL* sequence is identical to LC066217.
Table B. Collection locations and details, and GenBank accession numbers of samples used in the plastid-encoded \textit{psaA}, \textit{psaB}, \textit{psbA}, \textit{rbcL}, the universal mitochondrial barcode marker COI, and nuclear-encoded 18S \textit{rRNA} and 28S \textit{rRNA} gene analyses.

| Sample No. | Collection data | \textit{psaA} | \textit{psaB} | \textit{psbA} | \textit{rbcL} | COI   | 18S \textit{rRNA} | 28S \textit{rRNA} |
|------------|-----------------|------------|------------|----------|----------|------|----------------|----------------|
| OJ1        | Misaki (35°15' N, 139°36' E), Miura, Kanagawa | LC066222 | LC093495 | LC093497 | LC066217 | LC066521 | LC066529 | LC066525 |
|            | Prefecture, Japan; May 1927; coll. Yukio Yamada; SAP88755; topotype | | | | | | | |
| OJ2        | Nada (33°49' N, 135°10' E), Gobou, Wakayama | LC066223 | LC093496 | LC093498 | LC066218 | LC066522 | LC066530 | LC066526 |
|            | Prefecture, Japan; 20 Apr. 1958; coll. Torao Yamamoto; SAP 28242 | | | | | | | |
| OJ3        | Uragawa (38°12' N, 138°29' E), Sado, Niigata | LC066219 | LC066523 | LC066531 | LC066527 | | | |
|            | Prefecture, Japan; 6 Aug. 2011; coll. Masahiro Suzuki; TNS-AL 182118 | | | | | | | |
| OJ4        | Jodogaura (36°18' N, 133°20' E), Okinoshimacho, Oki County, Shimane Prefecture, Japan; 26 Jul. 2014; coll. Akira Kurihara; TNS-AL 195934 | LC066220 | LC066524 | LC066532 | LC066528 | | | |
Table C. GenBank accession numbers of species used in the identification of *Liagora japonica* sequences from historical herbarium specimens.

| Species                        | Reference                        | psaA  | psaB  | psbA  | rbcL | COI   | 28S rRNA | 18S rRNA |
|--------------------------------|----------------------------------|-------|-------|-------|------|-------|----------|----------|
| **Nemaliales**                 |                                  |       |       |       |      |       |          |          |
| *Akalaphycus liagoroides* (Yamada) Huisman, I.A. Abbott & A.R. Sherwood | [1]                              |       |       |       |      | KC134343 |          | KC157584 |
| *Akalaphycus setchelliae* (Yamada) Huisman, I.A. Abbott & A.R. Sherwood | [2]                              |       |       |       |      | GU357697 |          |          |
| **Ganonema farinosum** (J.V. Lamouroux) K.C. Fan & Yung C. Wang | [3]                              |       |       |       |      |       |          |          |
| *Gloiocallis dendroidea* (P. Crouan & H. Crouan) Showe M. Lin, Huisman & D.L. Ballantine (as *Ganonema dendroides*) | [3]                              |       |       |       |      |       |          |          |
| *Hommersandiophycus borowitzkæ* (Huisman) | [3]                              |       |       |       |      |       |          |          |
| *Hommersandiophycus clavatus* (Yamada) | [3]                              |       |       |       |      |       |          |          |
Table C. Continued.

| Species                                      | Reference | psaA | psaB | psbA | rbcL | COI   | 28S rRNA | 18S rRNA |
|----------------------------------------------|-----------|------|------|------|------|-------|----------|----------|
| *Hommersandiophycus pectinatus* (Collins &   |           |      |      |      |      | HQ603226 |          |          |
| Hervey) Popolizio, C.W. Schneider & C.E. Lane                                         |
| *Hommersandiophycus samaensis* (C.K. Tseng) Showe M. Lin & Huisman                  | [3]       |      |      |      |      | KF667102 |          |          |
| *Izziella formosana* (Yamada) Showe M. Lin, S.-Y. Yang & Huisman                    | [2]       |      |      |      |      | GU357688 |          |          |
| *Izziella hommersandii* Showe M. Lin, S.-Y. Yang & Huisman                           | [2]       |      |      |      |      | GU357690 |          |          |
| *Izziella kuroshioensis* Showe M. Lin, S.-Y. Yang & Huisman                          | [2]       |      |      |      |      | GU357684 |          |          |
| *Izziella orientalis* (J. Agardh) Huisman & Schils                                 | [4]       |      |      |      |      | HQ422594 |          |          |
| *Liagora albicans* J.V. Lamouroux          | [5]       |      |      |      |      | HQ901786 |          |          |
### Table C. Continued.

| Species | Reference | *psaA* | *psaB* | *psbA* | *rbcL* | COI | 28S rRNA | 18S rRNA |
|---------|-----------|--------|--------|--------|--------|-----|----------|----------|
| *Liagora albicans* J.V. Lamouroux | [4] | | | | | HQ422860 | | |
| | [4] | | | | | HQ422866 | | |
| | [4] | | | | | HQ422978 | | |
| *Liagora boergesenii* Yamada | [2] | | | | | GU357679\(^a\) | | |
| | [4] | | | | | HQ422649 | | |
| *Liagora ceranoides* J.V. Lamouroux | [2] | | | | | GU357681 | GU357669 | |
| *Liagora distenta* (Mertens ex Roth) J.V. Lamouroux | [6] | | | | | HQ603225 | | |
| *Liagora donaldiana* I.A. Abbott & Huisman | [4] | | | | | HQ423078 | | |
| *Liagora harveyana* Zeh | [2] | | | | | HM572263 | | |
| | [7] | | | | | DQ873275 | | |
| *Liagora japonica* Yamada (OJ3) | This study | | | | | LC066219 | LC066523 | LC066527 | LC066531 |
| *Liagora japonica* Yamada (OJ4) | This study | | | | | LC066220 | LC066524 | LC066528 | LC066532 |
| *Liagora julieae* I.A. Abbott & Huisman | [4] | | | | | HQ422852 | | |
### Table C. Continued.

| Species                          | Reference | psaA   | psaB   | psbA   | rbcL   | COI     | 28S rRNA | 18S rRNA |
|----------------------------------|-----------|--------|--------|--------|--------|---------|----------|----------|
| *Liagora mannarensis* V. Krishnamurthy & Sundararajan | [8]       |        |        |        |        |         |          |          |
| *Liagora viscida* (Forsskål) C. Agardh | [2]       |        |        |        |        | GU357678 | GU357670 |          |
|                                  | [6]       |        |        |        |        |         |          |          |
| *Liagora sp.*                    | [4]       |        |        |        |        | HQ422634 |          |          |
|                                  | [4]       |        |        |        |        | HQ422780 |          |          |
|                                  | [4]       |        |        |        |        | HQ422954 |          |          |
|                                  | [4]       |        |        |        |        | HQ422956 |          |          |
| *Macrocarpus perennis* (I.A. Abbott) Showe M. Lin, S.-Y. Yang & Huisman (as *L. perennis*) | [5]       |        |        |        |        | HQ901783 |          |          |
| *Neoizziella asiatica* Showe M. Lin, S.-Y. Yang & Huisman | [5]       |        |        |        |        | HQ901777 |          |          |
| *Neoizziella divaricata* (C.K. Tseng) Showe M. Lin, S.-Y. Yang & Huisman | [5]       |        |        |        |        | HQ901781 |          |          |
| Species                                           | Reference | psaA     | psaB     | psbA     | rbcL   | COI     | 28S rRNA | 18S rRNA |
|--------------------------------------------------|-----------|----------|----------|----------|--------|---------|----------|----------|
| Neoizziella divaricata (C.K. Tseng) Showe M.     | [4]       |          |          |          |        | HQ423117|          |          |
| Lin, S.-Y. Yang & Huisman                       |           |          |          |          |        |         |          |          |
| Nemalion multifidum (Lyngbye) Chauvin (as Nemalion sp.) | Unpublish | DQ787598|          |          |        |         |          |          |
| Nemalion multifidum (Lyngbye) Chauvin            | [1]       |          |          |          |        |         |          | KC157579 |
| P.G. Parkinson                                   |           |          |          |          |        |         |          | KC157581 |
| Nothogenia fastigiata (Bory) Ino & Tak. Tanaka   | [2]       |          |          |          |        | GU357695|          |          |
| Stenopeltis gracilis (Yamada & Tak. Tanaka)      |           |          |          |          |        |         |          |          |
| Itono & Yoshizaki                                 |           |          |          |          |        |         |          |          |
| Titanophycus setchellii (Yamada) Showe M.        | [2]       |          |          |          |        | GU357694|          | GU357674 |
| Lin, S.-Y. Yang & Huisman (as L. setchellii)     |           |          |          |          |        |         |          |          |
| Titanophycus validus (Harvey) Huisman, G.W.      | [2]       |          |          |          |        | GU357692|          | GU357672 |
| Saunders & A.R. Sherwood                         |           |          |          |          |        |         |          |          |
| Trichogloeopsis mucosissima (Yamada) I.A.        | [3]       |          |          |          |        |         |          | KF667107 |
| Abbott & Doty                                    |           |          |          |          |        |         |          |          |
| Species                                      | Reference | psaA | psaB | psbA | rbcL | COI       | 28S rRNA | 18S rRNA |
|----------------------------------------------|-----------|------|------|------|------|-----------|----------|----------|
| Trichogloeopsis pedicellata (M. Howe) I.A.   | [3]       |      |      |      |      | KF667108  |          |          |
| Abbott & Doty                                |           |      |      |      |      |           |          |          |
| Yoshizakia indopacifica Showe M. Lin,       | [9]       |      |      |      |      | JX878374  |          |          |
| Huisman & C. Payri                           |           |      |      |      |      |           |          |          |
| Nemaliophycidae                              |           |      |      |      |      |           |          |          |
| Acrochaetium savianum (Meneghini) Nägeli    | Unpublish | DQ787597 |     |      |      |           |          |          |
| ed                                           |           |      |      |      |      |           |          |          |
| Ballia callitriche (C. Agardh) Kützing       | Unpublish | DQ787595 |     |      |      |           |          |          |
| ed                                           |           |      |      |      |      |           |          |          |
| Batrachospermum gelatinosum (Linnaeus) De    | Unpublish | DQ787596 |     |      |      |           |          |          |
| Candolle                                     | ed        |      |      |      |      |           |          |          |
| Palmaria palmata (Linnaeus) F. Weber & D.   | Unpublish | DQ787599 |     |      |      |           |          |          |
| Mohr                                         | ed        |      |      |      |      |           |          |          |
| Thorea violacea Bory                         | [10]      |      |      |      |      | AY119712  |          |          |
| Species                        | Reference | *psaA*   | *psaB*   | *psbA*   | *rbcL*   | COI     | 28S rRNA | 18S rRNA |
|-------------------------------|-----------|----------|----------|----------|----------|---------|----------|----------|
| **Corallinophycidae**         |           |          |          |          |          |         |          |          |
| *Calliarthron tuberculatum* (Postels & Ruprecht) E.Y. Dawson | [11]      | KC153978^b | KC153978^b |          |          |         |          |          |
| **Rhodymeniophycidae**        |           |          |          |          |          |         |          |          |
| *Chondrus crispus* Stackhouse | [11]      | HF562234^b | HF562234^b |          |          |         |          |          |
| *Gracilaria Salicornia* (C. Agardh) E.Y. Dawson | [12]      | KF861575^b | KF861575^b |          |          |         |          |          |
| *Grateloupia taiwanensis* Showe M. Lin & H.Y. Liang | [13]      | KC894740^b | KC894740^b |          |          |         |          |          |

^aThe sequence is identical with *Dotyophycus yamadae* (Ohmi & Itono) I.A. Abbott & Yoshizaki (JX878366).

^bComplete genome of plastid.
Table D. Target read coverage of *Liagora japonica* (sample ID: suzuki-2; sample No.: OJ1).

|                        | psaA  | psaB  | psbA  | rbcL  | COI     | 18S rRNA | 28S rRNA |
|------------------------|-------|-------|-------|-------|---------|----------|----------|
| Reconstructed gene sequence length (bp) | 2259  | 2205  | 1083  | 1467  | 1599    | 2675     | 1619     |
| Average read coverage per base            | 538.8 | 438.1 | 577.3 | 673.9 | 321.3   | 198      | 217.5    |
| (Total length of mapped reads)/2 (bp)     | 1217746 | 966283 | 625515.5 | 988827 | 514100  | 529928   | 352419.5 |
| Average mapped read length (bp)           | 180.5 | 181.8 | 180.4 | 183.7 | 184.2   | 165.9    | 170.5    |
| Standard deviation of mapped read length (bp) | 69    | 67.8  | 68.8  | 69.6  | 68.9    | 68.9     | 67.7     |
| Total number of mapped pairs to the reference gene | 6744  | 5314  | 3466  | 5382  | 2790    | 3194     | 2066     |
| Number of mapped identical sequence pairs between F and R read | 4382  | 3379  | 2252  | 3569  | 1852    | 1806     | 1493     |
| Number of mapped different sequence pairs between F and R read | 2362  | 1935  | 1214  | 1813  | 938     | 1388     | 573      |
| Number of mapped different sequence pairs with only F or R read was mapped to the reference gene | 422   | 397   | 168   | 148   | 102     | 136      | 35       |
| Percent of identical sequence pairs in total mapped pairs to the reference gene | 64.9  | 63.5  | 64.9  | 66.3  | 66.3    | 56.5     | 72.2     |
Table E. Target read coverage of *Liagora japonica* (sample ID: suzuki-1; sample No.: OJ2).

|                        | psaA | psaB | psbA | rbcL | COI  | 18S rRNA | 28S rRNA |
|------------------------|------|------|------|------|------|----------|----------|
| Reconstructed gene sequence length (bp) | 2259 | 2205 | 1083 | 1467 | 1599 | 2675     | 1619     |
| Average read coverage per base            | 76.9 | 60.5 | 86   | 101.1| 128.4| 27.5     | 25.2     |
| (Total length of mapped reads)/2 (bp)     | 173875 | 133476 | 93274.5 | 148404 | 205554 | 73802.5  | 40874.5  |
| Average mapped read length (bp)           | 146.7| 149.3| 141.9| 147  | 153.1| 137.9    | 138      |
| Standard deviation of mapped read length (bp) | 57.5 | 57.9 | 57   | 56.2 | 62.3 | 54.4     | 48.6     |
| Total number of mapped pairs to the reference gene | 1185 | 894  | 657  | 1009 | 1342 | 535      | 296      |
| Number of mapped identical sequence pairs between F and R read | 975  | 735  | 533  | 841  | 1101 | 390      | 259      |
| Number of mapped different sequence pairs between F and R read | 210  | 159  | 124  | 168  | 241  | 145      | 37       |
| Number of mapped different sequence pairs with only F or R read was mapped to the reference gene | 25   | 19   | 20   | 7    | 25   | 19       | 2        |
| Percent of identical sequence pairs in total mapped pairs to the reference gene | 82.2 | 82.2 | 81.1 | 83.3 | 82   | 72.8     | 87.5     |
Table F. De novo assembly statistics.

| Sample ID (Sample No.) | Number of high quality reads (pairs) | Number of >500 bp scaffolds | Largest scaffolds (bp) | N50 length (bp) |
|------------------------|-------------------------------------|----------------------------|------------------------|-----------------|
| suzuki-1 (OJ2)         | 10,057,049                          | 101,376                    | 45,375                 | 619             |
| suzuki-2 (OJ1)         | 8,913,992                           | 158,568                    | 32,912                 | 670             |
Table G. Summary for the Bayesian analyses on the basis of *psaA*, *psaB*, *psbA*, *rbcL*, COI, 18S rRNA, and 28S rRNA datasets.

|                  | *psaA + rbcL* | *psaA* | *rbcL* | COI     | 28S rRNA | 18S rRNA | *psaB* | *psbA* |
|------------------|---------------|--------|--------|---------|----------|----------|--------|--------|
| Number of taxa   | 45            | 49 (55)*1 | 72 (99)*1 | 22 (21)*1 | 49 (51)*1 | 19 (21)*1 | 8 (9)*1 | 16     |
| Number of nucleotides (bp) | *psaA*: 1407, 1392 | 1317 | 579 | 2082*2 | 1453*2 | 2205 | 834 |
|                  | *rbcL*: 1374  |        |        |         |          |          |        |        |
| Included in analysis |               |        |        |         |          |          |        |        |
| Substitution model | *psaA*: 1st codons | 1st codons | 1st codons | 1st codons | GTR+I+G | GTR+I+G | 1st codons | 1st codons |
| selected*3       | (GTR+1+G), 2nd | (GTR+1+G), 2nd | (GTR+1+G), 2nd | (GTR+G), 2nd | (GTR+G), 2nd | (GTR+G), 2nd | (SYM+G), 2nd |        |
|                  | (GTR+1+G), 3rd | +I+G, 3rd codons | 3rd codons | codons (GTR+G) | (GTR+1+G), 3rd | codons (GTR+G) |        |        |
|                  | (GTR+1+G)     | (GTR+1+G) | (GTR+1+G) |          |          |          |        |        |
|                  | *rbcL*: 1st codons |               |        |         |          |          |        |        |
|                  | (GTR+1+G), 2nd |               |        |         |          |          |        |        |
|                  | (GTR+1+G), 3rd |               |        |         |          |          |        |        |

*1 indicates values are corrected by 10% and 5%; *2 indicates values are corrected by 15%; *3 indicates selected substitution models.
|                  | psaA + rbcL | psaA | rbcL | COI    | 28S rRNA | 18S rRNA | psaB | psbA |
|------------------|-------------|------|------|--------|----------|----------|------|------|
| MCMC generations | 2,000,000   | 2,000,000 | 3,000,000 | 2,000,000 | 2,000,000 | 2,000,000 | 2,000,000 | 2,000,000 |
| Average standard deviation of split | 0.003956 | 0.006071 | 0.009248 | 0.002545 | 0.003587 | 0.008603 | 0.001131 | 0.001975 |

*1 The numbers within parentheses indicate original number of taxa including the samples with identical nucleotide sequences.

*2 Numbers of aligned sites.

*3 Each substitution model was selected by hierarchical likelihood ratio test using MrModeltest 2.3 [14].
Table H. Matrix of $p$ distances among the liagoracean species used in the COI analysis.

|    | 1     | 2     | 3     | 4     | 5     | 6     | 7     | 8     | 9     |
|----|-------|-------|-------|-------|-------|-------|-------|-------|-------|
| 1  | Liagora japonica OJ1 | -     |       |       |       |       |       |       |       |
| 2  | Liagora japonica OJ2 | 0.00518135 | -     |       |       |       |       |       |       |
| 3  | Liagora japonica OJ3, OJ4 | 0.01554404 | 0.01381693 | -     |       |       |       |       |       |
| 4  | Neoizziella divaricata HQ423117 | 0.13644214 | 0.13816926 | 0.13816926 | -     |       |       |       |       |
| 5  | Liagora donaldiana HQ423078 | 0.12953368 | 0.13298792 | 0.13126080 | 0.12953368 | -     |       |       |       |
| 6  | Liagora boergesenii HQ422649 | 0.15889464 | 0.15716752 | 0.15716752 | 0.14335060 | 0.13471502 | -     |       |       |
| 7  | Liagora sp. HQ422634 | 0.15544042 | 0.15371330 | 0.15371330 | 0.14853196 | 0.13298792 | 0.01381693 | -     |       |
| 8  | Izzia_orientalis HQ422594 | 0.16234888 | 0.16407600 | 0.16062176 | 0.17616580 | 0.14335060 | 0.17962003 | 0.17098446 | -     |
| 9  | Liagora albicans HQ422978 | 0.15544042 | 0.15025906 | 0.14853196 | 0.15889464 | 0.14680484 | 0.18307427 | 0.17271157 | 0.12435233 |
| 10 | Liagora albicans HQ422866 | 0.14853196 | 0.14335060 | 0.14162348 | 0.16062176 | 0.14680484 | 0.18652850 | 0.17616580 | 0.12607944 | 0.01899827 |
| 11 | Liagora albicans HQ422860 | 0.16234888 | 0.16062176 | 0.16407600 | 0.16062176 | 0.14680484 | 0.18652850 | 0.17962003 | 0.12435233 | 0.11917099 |
| 12 | Liagora distenta HQ6033225 | 0.15198618 | 0.15198618 | 0.15371330 | 0.16925734 | 0.16580310 | 0.18652850 | 0.17962003 | 0.12435233 | 0.11917099 |
| 13 | Liagora julieae HQ422852 | 0.15556994 | 0.15556994 | 0.15037246 | 0.15385078 | 0.15396003 | 0.18666779 | 0.17975710 | 0.12443700 | 0.11410315 |
| 14 | Liagora viscida HQ6033227 | 0.16407600 | 0.16925734 | 0.16234888 | 0.16580310 | 0.16753022 | 0.18307427 | 0.17616580 | 0.12089810 | 0.11226252 |
| 15 | Liagora sp. HQ422956 | 0.16925734 | 0.16925734 | 0.17271157 | 0.15716752 | 0.16753022 | 0.18998273 | 0.18652850 | 0.13989638 | 0.12435233 |
| 16 | Liagora sp. HQ422954 | 0.15544042 | 0.15544042 | 0.15716752 | 0.16234888 | 0.15198618 | 0.16234888 | 0.15544042 | 0.13816926 | 0.12607944 |
| 17 | Liagora sp. HQ422780 | 0.15371330 | 0.15371330 | 0.15025906 | 0.16925734 | 0.16062176 | 0.16580310 | 0.16234888 | 0.14680484 | 0.12953368 |
| 18 | Hommersandiophycus pectinatus HQ603226 | 0.16753022 | 0.16234888 | 0.16580310 | 0.17271157 | 0.16234888 | 0.17616580 | 0.17443869 | 0.18825561 | 0.16062176 |
Table H. Continued.

|   | 10  | 11  | 12  | 13  | 14  | 15  | 16  | 17  | 18  |
|---|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| 1. | *Liagora japonica* OJ1 |   | | | | | | | |
| 2. | *Liagora japonica* OJ2 |   | | | | | | | |
| 3. | *Liagora japonica* OJ3, OJ4 |   | | | | | | | |
| 4. | *Neoizziella divaricata* HQ423117 |   | | | | | | | |
| 5. | *Liagora donaldiana* HQ423078 |   | | | | | | | |
| 6. | *Liagora boergesenii* HQ422649 |   | | | | | | | |
| 7. | *Liagora* sp. HQ422634 |   | | | | | | | |
| 8. | *Izziella_orientalis* HQ422594 |   | | | | | | | |
| 9. | *Liagora albicans* HQ422978 |   | | | | | | | |
| 10. | *Liagora albicans* HQ422866 |   | | | | | | | |
| 11. | *Liagora albicans* HQ422860 | 0.07944732 | 0.18247199 | | | | | | |
| 12. | *Liagora distenta* HQ603225 | 0.11917099 | 0.13298792 | 0.19327102 | | | | | |
| 13. | *Liagora julieae* HQ422852 | 0.11409407 | 0.11581545 | 0.13137032 | 0.18247199 | | | | |
| 14. | *Liagora viscida* HQ603227 | 0.11053541 | 0.13126080 | 0.11571676 | 0.12101270 | 0.18247199 | | | |
| 15. | *Liagora* sp. HQ422956 | 0.12262522 | 0.11398964 | 0.13298792 | 0.13138475 | 0.12435233 | 0.18247199 | | |
| 16. | *Liagora* sp. HQ422954 | 0.12607944 | 0.12780656 | 0.11226252 | 0.12790917 | 0.1398638 | 0.11917099 | 0.18247199 | |
| 17. | *Liagora* sp. HQ422780 | 0.12780656 | 0.13471502 | 0.13298792 | 0.12274799 | 0.11917099 | 0.13471502 | 0.12953368 | 0.18247199 |
| 18. | *Hommersandiophycus pectinatus* HQ603226 | 0.16234888 | 0.16753022 | 0.17789292 | 0.19011766 | 0.19516407 | 0.17443869 | 0.16753022 | 0.18134715 |
Table I. Matrix of Kimura 2-parameter (K2P) genetic distances among the liagoracean species used in the COI analysis.

|                | 1       | 2       | 3       | 4       | 5       | 6       | 7       | 8       | 9       |
|----------------|---------|---------|---------|---------|---------|---------|---------|---------|---------|
| 1. Liagora japonica OJ1 |  | -       |         |         |         |         |         |         |         |
| 2. Liagora japonica OJ2 | 0.00520838 | -       |         |         |         |         |         |         |         |
| 3. Liagora japonica OJ3, OJ4 | 0.01576537 | 0.01398917 | -       |         |         |         |         |         |         |
| 4. Neoizziella divaricata HQ423117 | 0.15241556 | 0.15464273 | 0.15448944 | -       |         |         |         |         |         |
| 5. Liagora donaldiana HQ423078 | 0.14292258 | 0.14725161 | 0.14498559 | 0.14283040 | -       |         |         |         |         |
| 6. Liagora boergesenii HQ422649 | 0.18217263 | 0.17981967 | 0.17960888 | 0.16009565 | 0.15005483 | -       |         |         |         |
| 7. Liagora sp. HQ422634 | 0.17747775 | 0.17514674 | 0.17494684 | 0.16679217 | 0.14785218 | 0.01401142 | -       |         |         |
| 8. Izzie1a orientalis HQ422594 | 0.18406512 | 0.18636397 | 0.18189923 | 0.20175615 | 0.15938573 | 0.20703724 | 0.19524424 | -       |         |
| 9. Liagora albicans HQ422978 | 0.17557563 | 0.16878819 | 0.16643274 | 0.17974012 | 0.16420557 | 0.21166740 | 0.19744845 | 0.13709708 | -       |
| 10. Liagora albicans HQ422866 | 0.16654603 | 0.15987927 | 0.15758295 | 0.18202844 | 0.16420557 | 0.21649836 | 0.20214331 | 0.13925691 | 0.01936862 |
| 11. Liagora albicans HQ422860 | 0.18446855 | 0.18216440 | 0.18663678 | 0.18202844 | 0.18611885 | 0.20897038 | 0.19953614 | 0.15721501 | 0.08336194 |
| 12. Liagora distenta HQ603225 | 0.17104045 | 0.17104045 | 0.17343797 | 0.19362989 | 0.1818993 | 0.20688269 | 0.13709708 | 0.13126765 |         |
| 13. Liagora julieae HQ422852 | 0.17561246 | 0.17561246 | 0.16893576 | 0.17323226 | 0.17277952 | 0.21591265 | 0.20640452 | 0.13668761 | 0.12446852 |
| 14. Liagora viscida HQ603227 | 0.18600610 | 0.19291864 | 0.18383019 | 0.18818893 | 0.19072685 | 0.21107565 | 0.20164141 | 0.13303448 | 0.12273866 |
| 15. Liagora sp. HQ422956 | 0.19318236 | 0.19318236 | 0.19801950 | 0.17746221 | 0.19072685 | 0.21967866 | 0.21493928 | 0.15687987 | 0.13776125 |
| 16. Liagora sp. HQ422954 | 0.17557563 | 0.17557563 | 0.17771989 | 0.1843273 | 0.17031585 | 0.18461565 | 0.17544200 | 0.15464273 | 0.13913736 |
| 17. Liagora sp. HQ422780 | 0.17343797 | 0.17343797 | 0.16878819 | 0.19347382 | 0.18189923 | 0.18854196 | 0.18394423 | 0.16455002 | 0.14418115 |
| 18. Hommersandiophycus pectinatus HQ603226 | 0.18993880 | 0.18316668 | 0.18760966 | 0.19657850 | 0.18311524 | 0.20259441 | 0.20022474 | 0.21760963 | 0.18104354 |
|   | Liagora japonica OJ1                      | Liagora japonica OJ2                     | Liagora japonica OJ3, OJ4                          | Neoizziella divaricata HQ423117 | Liagora donaldiana HQ423078 | Liagora boergesenii HQ422649 | Liagora sp. HQ422634                  | Iziella_orientalis HQ422594          | Liagora albicans HQ422978            | Liagora albicans HQ422866               | Liagora albicans HQ422860 | Liagora distenta HQ603225            | Liagora julieae HQ422852               | Liagora viscida HQ603227             | Liagora sp. HQ422956                    | Liagora sp. HQ422954                    | Liagora sp. HQ422780                    | Hommersandiophycus pectinatus HQ603226 |
|---|------------------------------------------|----------------------------------------|--------------------------------------------------|---------------------------------|-------------------------------|-------------------------------|-----------------------------------|--------------------------------------|----------------------------------|----------------------------------------|-----------------------------------|------------------------------------------|----------------------------------------|----------------------------------------|----------------------------------------|----------------------------------------|----------------------------------------|
|10 |                                          |                                        |                                                  |                                 |                               |                               |                                   |                                      |                                  |                                        | 0.08537807                          | 0.13126765                          | 0.12445746                            | 0.12062895                            | 0.13558733                            | 0.13913736                            | 0.14198335                            | 0.18329073                            |                                        |
|11 |                                          |                                        |                                                  |                                 |                               |                               |                                   |                                      |                                  |                                        | -                                 | -                                        |                                        |                                        |                                        |                                        |                                        |                                        |
|12 |                                          |                                        |                                                  |                                 |                               |                               |                                   |                                      |                                  |                                        | 0.12637727                          | 0.14655268                          | 0.14652897                            | 0.14654747                            | 0.14877491                            | 0.14130187                            | 0.15050350                            | 0.20341456                            |                                        |
|13 |                                          |                                        |                                                  |                                 |                               |                               |                                   |                                      |                                  |                                        | 0.14652897                          | -                                 | -                                        |                                        |                                        |                                        |                                        |                                        |                                        |
|14 |                                          |                                        |                                                  |                                 |                               |                               |                                   |                                      |                                  |                                        | -                                 |                                        |                                        |                                        |                                        |                                        |                                        |                                        |
|15 |                                          |                                        |                                                  |                                 |                               |                               |                                   |                                      |                                  |                                        | -                                 |                                        |                                        |                                        |                                        |                                        |                                        |                                        |
|16 |                                          |                                        |                                                  |                                 |                               |                               |                                   |                                      |                                  |                                        | -                                 |                                        |                                        |                                        |                                        |                                        |                                        |                                        |
|17 |                                          |                                        |                                                  |                                 |                               |                               |                                   |                                      |                                  |                                        | -                                 |                                        |                                        |                                        |                                        |                                        |                                        |                                        |
|18 |                                          |                                        |                                                  |                                 |                               |                               |                                   |                                      |                                  |                                        | -                                 |                                        |                                        |                                        |                                        |                                        |                                        |                                        |
Additional References

1. Scott FJ, Saunders GW, Kraft GT. *Entwisleia bella*, gen. et sp. nov., a novel marine ‘batrachospermaceseous’ red alga from southeastern Tasmania representing a new family and order in the Nemaliophycidae. Euro. J. Phycol. 2013; 48: 398–410.

2. Lin S-M, Yang S-Y, Huisman JM. Systematic revision of the genus *Liagora* and *Izziella* (Liagoraceae, Rhodophyta) from Taiwan based on molecular analyses and carposporophyte development, with the description of two new species. J. Phycol. 2011; 47: 352–365.

3. Lin S-M, Huisman JM, Ballantine DL. Revisiting the systematics of *Ganonema* (Liagoraceae, Rhodophyta) with emphasis on species from the northwest Pacific Ocean. Phycologia 2014; 53: 37–51.

4. Sherwood AR, Kurihara A, Conklin KY, Sauvage T, Presting GG. The Hawaiian Rhodophyta Biodiversity Survey (2006-2010): a summary of principal findings. BMC Plant Biol. 2010: 10: 258.

5. Lin S-M, Yang S-Y, Huisman JM. Systematics of *Liagora* with diffuse gonimoblasts based on *rbcL* sequences and carposporophyte development, including the description of the new genera *Neoizziella* and *Macrocarpus* (Liagoraceae, Rhodophyta). Euro. J. Phycol. 2011; 46: 249–262.

6. Le Gall L, Saunders GW. Establishment of a DNA-barcode library for the Nemaliales (Rhodophyta) from Canada and France uncovers overlooked diversity in the species *Nemalion helminthoides* (Velley) Batters. Cryptogamie Algol. 2010; 31: 403–421.

7. Huisman JM, Saunders GM, Sherwood AR. Recognition of *Titanophycus*, a new genus based on *Liagora valida* Harv. (Liagoraceae, Nemaliales). In: Huisman JM, editors. Algae of Australia – Nemaliales: ABRS; 2006. pp. 116-119.

8. Huisman JM, Harper JT, Saunders GW. Phylogenetic study of the Nemaliales (Rhodophyta) based on large-subunit ribosomal DNA sequences supports
segregation of the Scinaiaceae fam. nov. and resurrection of *Dichotomaria* Lamarck. Phycol Res 2004; 52: 224–234.

9. Lin S-M, Huisman JM, Payli C. Characterization of *Liagora ceranoides* (Liagoraceae, Rhodophyta) on the basis of *rbcL* sequence analyses and carposporophyte development, including *Yoshizakia indopacifica* gen. et sp. nov. from the Indo-Pacific region. Phycologia 2013; 52: 161–170.

10. Yoon HS, Hackett JD, Bhattacharya D. A single origin of peridinin- and fucoxanthin-containing plastids in dinoflagellates through tertiary endosymbiosis. PNAS 2002; 99: 11724-11729.

11. Janouskovec J, Liu SL, Martone PT, Carre W, Leblanc, C, et al. Evolution of red algal plastid genomes: ancient architectures, introns, horizontal gene transfer, and taxonomic utility of plastid markers. PLOS ONE 2013; 8: e59001.

12. Campbell MA, Presting GP, Bennett MS, SherwoodAR. Highly conserved organellar genomes in the Gracilariales as inferred using new data from the Hawaiian invasive alga *Gracilaria salicornia*. Phycologia 2014; 53: 109-116.

13. Depriest MS, Bhattacharya D, Lopez-Bautista JM. The Plastid Genome of the Red Macroalga *Grateloupia taiwanensis* (Halymeniaceae). PLOS ONE 2013; 8: e68246.

14. Nylander JAA. MrModeltest 2.1. Program distributed by the author. Evolutionary Biology Centre, Uppsala University, Uppsala; 2004.
S1 Text. The DNA extraction and sequencing procedures for modern specimens.

The modern specimens used in molecular analyses are listed in S1 Table. Total DNA was extracted from field-collected specimens dried by silica gel using the DNeasy Plant Mini Kit (QIAGEN, Tokyo, Japan) following the instructions of the manufacturer. The total DNA was used as a template for polymerase chain reaction (PCR) amplification using a TOYOBO KOD FX Neo (TOYOBO CO. LTD., Oosaka, Japan). Primers used for PCR amplification were: 28S rRNA gene: T01

\(5'-\text{TAAAGCATATCAGTAAGCGGAG-3'}\) – V

\(5'-\text{CGTATCGGCCAGTCTGCTTACC-3'}\), F449

\(5'-\text{CCCGAAGATGGGAACACTATG-3'}\) – G \(5'-\text{CACCACTCCTACTC-3'}\), T04 \(5'-\text{GCAGGACCGTGCCCATGGAAGT-3'}\) - 28F

\(5'-\text{CAGAGCACTGGCCAGAAAATC-3'}\), and T05

\(5'-\text{GCAACGGKCAAAGGGAATCC-3'}\) - T15

\(5'-\text{TGATAGGAAGAGCCCGACATCA-3'}\) [1, 2]; 18S rRNA gene: SR1

\(5'-\text{CCTGGTTGATCTCCGCA-3'}\) - SR9 \(5'-\text{AACTAAGACGCCATGCAC-3'}\),

and SR4 \(5'-\text{AGCCCGCAGTTCCAGCT-3'}\) - SR12

\(5'-\text{CCTTCYGCAGGTCCTACCT-3'}\) [3]; \textit{rbcL}: F8

\(5'-\text{GGYGTAATCCATGCGWAAATG-3'}\) - R1150
(5’-GCATTTGWCCACARTGAATACC-3’) and F645
(5’-ATGMGHTGGAAAGAAAGATT-3’) - R1381 (5’-
ATCTTTCCATAAATCTARAGC-3’) [4]; COI: GazF1
(5’-TCAACAAATCATAAAGATATTGG-3’) - GazR1
(5’-ACTTCTGGATGTCCAAAAAYCA-3’) [5]. The temperature-cycling protocol was: 28S rRNA gene: 2 min at 94°C for an initial denaturation step, followed by 35 cycles of 15 sec denaturation at 94°C, 30 sec primer annealing at 55°C, and 1 min extension at 68°C, with a final 7 min extension at 72°C, and then a hold at 4°C; rbcL and COI: 2 min at 94°C for an initial denaturation step, followed by 35 cycles of 15 sec denaturation at 94°C, 30 sec primer annealing at 46°C and 1 min extension at 68°C, with a final 7 min extension at 72°C, and then a hold at 4°C. The amplified DNA fragments were purified using QIAquick PCR Purification Kit (QIAGEN, Tokyo, Japan). Cycle-sequencing with BigDye Terminator v3.1 Cycle Sequencing Kit (Applied Biosystems, Tokyo, Japan) was carried out in a 7.5 μl volume of reaction: 2 μl of BigDye Terminator v3.1 Reaction Mix, 10-30 ng/ml of PCR product, 10 pmol of primer, and dH2O to 7.5 μl. The cycle-sequencing program consisted of an initial step at 97°C for 2 min, 25 sequencing cycles (97°C for 10 s, 50°C for 25 s, 60°C for 2 min). The BigDye-labeled PCR products were ethanol-precipitated following the manufacturer's
protocol and completely sequenced using ABI PRISM 310 Genetic Analyzer (Applied Biosystems, Tokyo, Japan). Reverse and direct chromatograms were assembled using the program GeneStudio™ Professional Ver. 2.2. (GeneStudio, Inc.).

**Additional References**

1. Freshwater DW, Fredericq S, Bailey JC. Characteristics and utility of nuclear-encoded large-subunit ribosomal gene sequences in phylogenetic studies of red algae. Phycol Res 1999; 47: 33–38.

2. Harper JT, Saunders GW. The application of sequences of the ribosomal cistron to the systematics and classification of the florideophyte red algae (Florideophyceae, Rhodophyta). Cah Biol Mar 2001; 42: 25–38.

3. Nakayama T, Watanabe S, Mitsui K, Uchida H, Inouye I. The phylogenetic relationship between the Chlamydomonadales and Chlorococcales inferred from 18SrDNA sequence data. Phycol Res 1996; 44: 47-55.

4. Wang HW, Kawaguchi S, Horiguchi T, Masuda M. Reinstatement of Grateloupia catenata (Rhodophyta, Halymeniaceae) on the basis of morphology and rbcL sequences. Phycologia 2000; 39: 228–237.

5. Saunders GW. Applying DNA barcoding to red macroalgae: a preliminary appraisal
holds promise for future applications. Phil Trans R Soc B 2005; 360: 1879-1888.