Review

Essentials Oils from Brazilian *Eugenia* and *Syzygium* Species and Their Biological Activities

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Abstract: The *Eugenia* and *Syzygium* genera include approximately 1000 and 1800 species, respectively, and both belong to the Myrtaceae. Their species present economic and medicinal importance and pharmacological properties. Due to their chemical diversity and biological activity, we are reporting the essential oils of 48 species of these two genera, which grow in South America and found mainly in Brazil. Chemically, a total of 127 oil samples have been described and displayed a higher intraspecific and interspecific diversity for both *Eugenia* spp. and *Syzygium* spp., according to the site of collection or seasonality. The main volatile compounds were sesquiterpene hydrocarbons and oxygenated sesquiterpenes, mainly with caryophyllane and germacrane skeletons and monoterpenes of mostly the pinane type. The oils presented many biological activities, especially antimicrobial (antifungal and antibacterial), anticholinesterase, anticancer (breast, gastric, melanoma, prostate), antiprotozoal (*Leishmania* spp.), antioxidant, acaricidal, antinociceptive and anti-inflammatory. These studies can contribute to the rational and economic exploration of *Eugenia* and *Syzygium* species once they have been identified as potent natural and alternative sources to the production of new herbal medicines.

Keywords: *Eugenia* spp; *Syzygium* spp; Myrtaceae; essential oil variability; mono- and sesquiterpenes; biological properties

1. Introduction

From the latter 19th century through the second half of the 20th century, *Syzygium* species were included in the *Eugenia* genus. However, anatomical and morphological analyses provided evidence that these genera were not closely linked, and a distinction between individuals belonging to these two genera has been attributed [1]. *Eugenia* and *Syzygium* genera belong to the Myrtaceae family, included in the Myrtales order, Rosidae clade, and Malvidae subclade [2]. In recent studies, these genera have been classified into subgenera; *Eugenia* includes *Eugenia*, *Hexachlamys*, and *Pseudeugenia* subgenera [3], while the *Syzygium* genus encompasses *Syzygium*, *Acmena*, *Sequestratum*, *Perikion*, *Anetholea*, and *Wesa* subgenera [4].
Eugenia L. has approximately 1000 species, occurring in Central and South America, and few in the African continent [5]. They can be shrubs or small trees, twigs glabrous or pubescent when young. Leaves are opposite, petiolate; lamina gland-dotted, intramarginal vein visible. Inflorescence is axillary; solitary flowers, fascicled or rarely in triads; pubescent peduncles; pubescent bracts, persistent. Flowers are 4-merous, buds usually turbinate; rounded sepals, sparsely pubescent; petals ± orbicular, gland-dotted, and ciliate margins—stamens in multiple whorls on a broad staminal disc; ovary 2-locular, ovules several to many, radiating from a centrally-located axile placenta; style about as long as the stamens or slightly longer; stigma not dilated. Fruits are succulent berries; sepals persistent; seed 1–2, cotyledons of uniform texture, partly fused [6].

Eugenia species have great ecological relevance because their fruits are sources of food for birds, mammals, and reptiles [7]. Moreover, this genus stands out due to the commercial exploitation of edible fruits, wood, essential oils, and plants for ornamental purposes, and its pharmacological potential [8,9]. For example, infusions and teas of leaves, fruits, and trunk bark of Eugenia brasiliensis are used in Amazon folk medicine to treat stomach diseases, as an antirheumatic, anti-inflammatory, antidiarrheal, and diuretic. In contrast, decoctions of twigs of Eugenia supraaxillaris (syn. Eugenia axillaris) are used as antirheumatic and bathing by women after childbirth [10].

Syzygium Gaertn. has 1800 species [11], mainly found in the southern and Southeast Asia, southern China, Australia, Malaysia and New Caledonia, and some in East Africa, Madagascar, Mascarenes islands, southwest Pacific islands, Taiwan, and southern Japan [12]. There are only four species occurring in South America: Syzygium aqueum (syn. Eugenia aqua, Jambosa aquea), S. cumini (syn. E. jambolana, Syzygium jambolanum), S. jambos (Eugenia jambos, E. monantha, Syzygium merrillii, S. monanthum), and S. malaccense [13].

Although Syzygium species are not native to the Americas, their species are well spread throughout South America in afforestation and decoration. Syzygium cumini and S. malaccense are two of the most widespread and popularly known exotic species throughout the Brazilian territory and commonly found in indigenous villages in the sub-spontaneous state amid natural vegetation [14].

Syzygium species can be trees, with thick, granular bark; twigs usually glabrous. Leaves are opposite, entire, penninerved, usually gland-dotted; lateral nerves united, forming a clear or faint intramarginal vein. Flowers are bisexual, in terminal or axillary corymbose cymes or panicles; calyx tube hemispherical, globose, or turbinate, tube produced above the summit of the ovary, lobes four or five, ovate to suborbicular, imbricate; petals four or five, orbicular, pellucid-glandular; stamens numerous, filaments inflexed in bud; staminal disc broad or absent; anthers globose; ovary inferior, two-celled; ovules few to several in each cell; style 1, subulate, stigma simple. The fruit is a berry, one-celled; seeds few [15].

Syzygium is one of the most common tree genera in the forest ecosystem, presenting nectariferous flowers, often in mass and typically fleshy fruits; it is used as food by birds, insects, and small and large mammals [1,12]. Its species present economic and medicinal importance [16], and pharmacological proprieties being a potential source for pharmachemistry studies [17,18]. Traditional communities use the infusions and decoction leaves of Syzygium cumini and S. aqueum to treat diabetes, and stomach pains and dysentery, respectively [19,20].

Chemometrics analysis, such as Principal cluster analysis (PCA) and Hierarchical cluster analysis (HCA), is the most used method to group essential oils from many samples. PCA is often the first step in multivariate data analysis. The score plot is a graphical representation that provides information about relationships between samples and chemical compounds. In turn, HCA is a significant pattern recognition technique used as a preliminary evaluation of a given data set. In HCA, samples are grouped based on information that describes their relationship by similarities, such as distance and correlation. HCA is plotted in branched structures with a defined hierarchy (called dendrogram), which permits a qualitative visualization (in two-dimensional space) of grouping among samples [21].
Due to the chemical and biological importance of *Eugenia* and *Syzygium* species, in this review, we report 127 chemical compositions of 48 species of these two genera growing in South America (Figure 1) and their biological activities.

![Geographical distribution in South America of Eugenia and Syzygium specimens based on their essential oil studies.](image)

**Figure 1.** Geographical distribution in South America of *Eugenia* and *Syzygium* specimens based on their essential oil studies. This map was built by the authors using the information of the plant occurrence, available in each bibliographic reference. Abbreviation list: BR: Brazil, VE: Venezuela, CO: Colombia.

2. Bibliographic Search Criteria

The bibliographic research was performed using the databases Google Scholar, Pubmed, Science Direct, Medline, and Scopus. The keywords applied were “*Eugenia*”, “*Syzygium*” and “essential oils”; and “volatile compounds”, and “essential oils”. Some unusual or incorrect botanical names were updated based on The Plant List” (http://www.theplantlist.org/).

3. Volatile Profiles

*Eugenia* essential oils are characterized by intra- and interspecific chemical diversity, with the predominance of cyclic sesquiterpenes, follow by monoterpenes, and without phenylpropanoids as the main compound (See Appendices A and B) [22]. However, dimethylxanthoxylin (73.2 and 83.0%), an acetophenone derivative, was identified by GC–MS and NMR techniques as the major constituent in EOs of two specimens of *E. protenta* McVaugh collected in Santarém Novo, western Pará state located in the Brazilian Amazon [23].

Some essential oils of *Eugenia* species have shown great interspecific chemical variation. For example, *Eugenia protenta* EOs from Pará state (Brazil) occurring at three chemical profiles rich in the cyclic sesquiterpenes with eudesmane, elemene, and germacrane skeletons as selin-11-en-4α-ol.
was rich in viridiflorol (17.7%), followed by \( \beta \) (29.9%) [41]. Additionally, the oils of (19.3%) [39]. In addition, (9.6-28.2%) [33]; and rich in a mixture of oxygenated sesquiterpenes as spathulenol (12.7%), \( \beta \) (15.8–34.5%) and (27.3%) as main constituents [25].

The minority compounds were monoterpenes with pinane (\( \alpha \)) displayed the sesquiterpene (\( \alpha \)) presented caryophyllene oxide (22.2%) and (\( \beta \))-cubebene (10.9%). At the same time, it leaves oil composition at the vegetative (22.3%) and flowering (18.9%) stages. Additionally, the flowers’ oil was characterized by the predominance of sesquiterpene hydrocarbons (33.2%), such as (E)-caryophyllene (12.8%), \( \alpha \)-cadinol (10.1%), and (\( \tau \))-murolol (8.9%) and (\( \tau \))-murolol (8.5%) [34].

Essential oils of (Mart.) DC. leaves from central and southern regions of Brazil displayed the sesquiterpene (E)-caryophyllene as the main compound (14.8% and 24.0%, respectively). The minority compounds were monoterpenes with pinane (\( \beta \)-pinene and \( \alpha \)-pinene), menthane (\( \alpha \)-terpineol and limonene), caryophyllane (\( \alpha \)-humulene and caryophyllene oxide), and cadinane skeletons (\( \delta \)-cadinene, \( \alpha \)-copaene), displaying variability in their amounts [35,36].

The oils of (L.) DC. EOs from Brazil were classified in four chemical profiles as follows: caryophyllane profile, with a significant content of the sesquiterpenes (E)-caryophyllene (9.8–16.8%) and caryophyllene oxide (20.5–28.6%); cadinane profile, characterized by the presence of \( \alpha \)-cadinol (14.7%); aromadendrane profile, with the predominance of the sesquiterpenes globulol (9.8%), germacrene B (7.9%) [24]; and pinane profile with \( \beta \)-pinene (27.8%) and \( \alpha \)-pinene (27.3%) as main constituents [25].

Oils of (Cambess. (syn. ) present two chemical profiles, three specimens collected in Rio de Janeiro and São Paulo states (Brazilian southeast) were shown to be rich in \( \alpha \)-pinene (15.8–34.5%) and \( \beta \)-pinene (11.0–34.1%) [26–28]. The specimen from Santa Catarina (Brazil southern) was rich in viridiflorol (17.7%), followed by \( \beta \)- (13.2%) and \( \alpha \)-pinene (11.2%) [29]. ( ) oils from Santa Catarina state (Brazil southern) showed high amounts of cyclic sesquiterpene hydrocarbons with germacrene/caryophyllane/cadinane skeleton such as bicyclogermacrene (7.2–14.3%), germacrene D (1.6–8.6%), (E)-caryophyllene (6.4–8.0%), and \( \delta \)-cadinene (4.9–7.2%) [30,31].

(Eugenia brasiliensis) Lam. leaf oils collected in the south and southeast Brazil displayed the occurrence of three chemical profiles. Its oils can be rich in bicyclic monoterpenes as \( \beta \)-pinene (10.4%) and \( \alpha \)-pinene (10.3%) [32]; or \( \alpha \)-pinene (18.8–35.5%), \( \beta \)-pinene (11.0–14.4%), followed by 1,8-cineol (9.6–28.2%) [33]; and rich in a mixture of oxygenated sesquiterpenes as spathulenol (12.7%), \( \tau \)-cadinol (8.7%) and viridiflorol (7.1%) [29]. In addition, (E. brasiliensis) fruits showed different volatile profiles according to their genotypic variety. The oils extracted from the yellow variety showed as the main compound \( \alpha \)-pinene (15.4%), myrcene (10.7%), and \( \alpha \)-terpineol (10.2%), while oils from purple variety presented caryophyllene oxide (22.2%) and \( \alpha \)-cadinol (10.4%) [33].

A stems oil of (O.Berg from Paraná state, Brazil, was rich in oxygenated sesquiterpenes (83.4%), characterized by caryophyllene oxide (17.2%), globulol (16.5%), \( \tau \)-murolol (16.8%), \( \alpha \)-cadinol (12.1%) and 1-epi-cubenol (10.9%). At the same time, it leaves oil composition at the vegetative, and flowering stages had only small quantitative differences, with globulol as the main compound at the vegetative (22.3%) and flowering (18.9%) stages. Additionally, the flowers’ oil was characterized by the predominance of sesquiterpene hydrocarbons (33.2%), such as (E)-caryophyllene (12.8%), \( \alpha \)-cadinol (10.1%), \( \alpha \)-pinene (8.9%) and \( \tau \)-murolol (8.5%) [34].

The moisture of plant material can influence in the EOs yield and composition. Leaves of (O.Berg dried by three different processes (in natura, at room temperature, and drying by forced air circulation) showed qualitative and quantitative variations in their oil constituents. The main compounds were \( \alpha \)-copaene (1.7–10.6%), \( \beta \)-bisabolene (1.9–17.4%), (\( \alpha \))-bergamotene (0.0–10.1%), spathulenol (7.2–10.9%) and germacrene D (2.4–13.3%). Besides, there was a clear difference between the oil compositions from its leaves and flowers, which this last was rich in (\( \alpha \))-bergamotene (29.9%) [41].
The EOs of leaves of *E. langsdorffii* O.Berg presented *epi-*longipinanol (13.6%), γ-eudesmol (12.3%), limonene (11.8%), and 10-*epi-γ*-eudesmol (10.6%) as the major constituents, while their fruits showed 10-*epi-γ*-eudesmol (35.7%), 1,10-di-*epi*-cubebol (15.6%) [42].

The leaf oil of *E. patrisii* Vahl was characterized by presence of cyclic sesquiterpenes with cadinane and caryophyllane skeletons including *trans*-cadin-1,4-diene (16.5%), *trans*-muurola-3,5-diene (13.3%), (*E*)-caryophyllene (11.1%) and α-cubebene (9.8%) [25]. However, its aerial parts (leaves and fine stems) had a mixture of acyclic and cyclic oxygenated sesquiterpenes as (*E*,*E*)-farnesol (34.5%), (*E*,*Z*)-farnesol (23.2%), caryophylla-4(12)-8(13)-dien-5β-ol (15.6%) as major constituents [37].

Different chemical oil profiles of *E. punicifolia* (Kunth) DC. were found. Specimens collected in Brazilian northeast showed linalool (44.0–61.2%), (*E*)-caryophyllene (16.2–22.7%), and α-terpineol (6.7–8.8%) as the main compounds [43]. Specimens sampled in Brazilian Amazon presented the sesquiterpene hydrocarbons (*E*)-caryophyllene (9.9%), bicylogermacrene (8.7%), germacrene D (5.4%) and the monoterpenes (*E*)-β-ocimene (5.5%) as their primary constituents [25]. Additionally, in another plant sample from Brazilian southeast predominated sesquiterpenes with cadinane/germacrane skeleton as α-cadinol (10.6%), 10-*epi-γ*-eudesmol (10.2%), paradisiol (9.0%), and 7-*epi*-α-selinene (6.8%) [28].

Essential oil from leaves of *E. pyriformis* Cambess. collected in southern Brazil showed two chemical profiles: The first one (1) showing sesquiterpenes with cadinane/germacrane-type skeletons as α-cadinol (14.0%), δ-cadinene (12.4%), τ-cadinol (11.9%), and bicyclogermacrene (10.2%) [30]; and the second (2) presenting the monoterpenes β-pinene (0.2–25.7%), limonene (0.2–22.0%), and 1,8-cineole (0.6–14.7%), with pinane and menthane-type skeletons, and the sesquiterpene caryophyllene oxide (3.9–21.3%) with a caryophyllane-type skeleton [44]. Additionally, flowers EO was characterized by caryophyllane/cadinane sesquiterpene-types, as (*E*)-caryophyllene (22.8%), germacrene D (15.3%), bicyclogermacrene (8.4%), and α-cadinol (7.2%) [44]. Additionally, fruits oil of *E. pyriformis* showed caryophyllene oxide (16.2%), limonene (12.4%), α-terpineol (5.4%), α-cadinol (5.4%) as the main compounds [44].

Leaf oils of *E. sulcata* Spring ex Mart. from Brazil may be characterized by two different chemical profiles: the first one (1) presented (*E*)-caryophyllene (15.0–24.6%), α-pinene (17.2–34.2%), and β-pinene (1.7–10.9%) [32,45,46], and the second (2) showed 1,8-cineole (19.0%), α-pinene (16.9%), and β-pinene (14.5%) [28]. Another sample of fine stems oil was rich in (*E*)-caryophyllene (18.8%), spathulenol (8.8%), and *trans*-calamene (7.0%) with minor amounts of α- and β-pinene [46].

There is a significant chemical variability on the essential oils of *Eugenia uniflora* L., the most studied *Eugenia* species [22], with the occurrence of about nine chemotypes, rich in cyclic oxygenated sesquiterpenes, with a germacranype skeleton, as the curzerene (thermally produced in situ during the oil processing) [47–50], atractylone [51,52], selina-1,3,7(11)-trien-8-one and selina-1,3,7(11)-trien-8-one epoxide [48,49,53,54], germacrene [55,56], and in association with other cyclic sesquiterpene hydrocarbons [48,49,51,56–58].

Although various *Eugenia* species and specimens have been widely studied concerning their leaf essential oil compositions, about 26 species were sampled only once to furnish their oil compositions. These plant samples exist mainly in Brazil, and the species that included monoterpene hydrocarbons preferably were *E. dimorpha* O.Berg (α-pinene, 22.4%) [59], *E. multicostata* D.Legrand (α-pinene, 16.1% and s pathulenol 10.7%) [32], *E. speciosa* Cambess. (α-pinene, 47.3% and limonene, 23.0%) [32], and *E. uruguayensis* Cambess. (α-pinene, 23.5% and β-pinene, 11.8%) [27]; and that included sesquiterpene hydrocarbons were *E. acutata* Miq. (*E*)-caryophyllene, 26.93%) [60], *E. bacopari* D.Legrand (δ-cadinene, 15.8% and aromadendrene, 12.2%) [59], *E. burkartiana* (D.Legrand) D.Legrand (bicyclogermacrene, 14.2%, germacrene D, 8.8% and (*E*)-caryophyllene, 7.8%) [59], *E. florida* DC. (*E*-caryophyllene, 14.5% and β-elemene, 1.8%) [61], *E. hiemalis* Cambess. (bicyclogermacrene, 37.7%) [62], *E. pitya* (O.Berg) Nied. (germacrene D, 29.3% and bicyclogermacrene, 22.4%) [32], *E. platysema* O.Berg (β-selinene, 17.9%, *allo*-aromadendrene, 12.6% and 7-*epi*-α-selinene, 10.4%) [27], *E. rambod* D.Legrand (β-elemene, 10.6%, bicyclogermacrene, 9.7% and (*E*)-caryophyllene, 6.2%) [27], *E. repanda* O.Berg ((*E*)-caryophyllene, 16.3%, α-humulene, 10.2% and bicyclogermacrene, 9.4%) [61], *E. stictopetala* Biomolecules 2020, 10, 1155 5 of 36
Moreover, eight species were found to have oxygenated sesquiterpenes as the major constituents: *E. candollea* DC. (1-epi-cubenol, 77.59%) [60], *E. egensis* DC. (5-hydroxy-trans-calamenene, 35.8%) [37], *E. expansa* Spring ex Mart. (spathulenol, 15.4% and *E. polystachya* Mart. ex DC. (spathulenol, 16.02% and *E. xiriricana* Mattos (spathulenol, 15.4% and *β-pinene, 14.1%) [32]. In addition, three species presented other constituents such as esters, carboxylic acid, and oxygenated monoterpenes as main compounds. They were *E. catharinaensis* D. Legrand (ethyl palmitate, 10.5%; *trans*-α-bergamotene, 6.5%; and *β-pinene, 5.9%) [59], *E. stigmata* DC. (physeteric acid, 90.5%) [62], and *E. triquetra* O. Berg (from Venezuela) (linool, 17.5%, limonene, 16.9%, *α-pinene, 11.6%, and *β-pinene, 8.7%) [65].

Chemical profiles of oils from *Syzygium* species growing in South America are poorly described in the literature. We found seven *Syzygium cuminii* accessions (specimens) growing in Brazil. It is known that the collection site and habitat affect the chemical composition of essential oils. The oils of *S. cumini* (L.) Skeels (syn. *Eugenia cuminii* and *E. jambolana*) collected in Brazil (Ceará, Maranhão, and Rio de Janeiro states) showed *α-pinene* (22.2–48.0%) as the main constituent [66–69], while another oil of *S. cumini* collected in Minas Gerais state, Brazil, showed *α-humulene* (25.2%) and (E)-caryophyllene (16.0%) as the predominant compounds [70].

### 4. Seasonal Variation of Oil Composition

Some studies have shown variation in the chemical composition and yield of EOs of *Eugenia* and *Syzygium* affected by seasonality. Samples of fresh leaves of *E. astringens* (syn. *Eugenia rotundifolia*), with Brazil’s occurrence, were collected every three months. The oil was mainly composed of cyclic monoterpane and sesquiterpene hydrocarbons, as *α-pinene* (19.7–34.4%), *β-pinene* (20.6–34.1%), and (E)-caryophyllene (3.6–11.7%). The rain precipitation data displayed a significant positive and negative correlation between the (E)-caryophyllene and *α-pinene* content, respectively [26].

Two oil profiles of *E. brasiliensis* were monitored according to their seasonal and circadian variations. A specimen collected in Santa Catarina state, Brazil, showed *α-pinene* (1.77–15.94%), *β-pinene* (2.98–11.24%), spathulenol (8.10–18.17%), 1-epi-cubenol (4.83–7.46%), and *β-cadinol* (10.38–15.30%) as main volatile compounds. Spathulenol was the most abundant in the spring (16.02%) and summer (18.17%), *β-cadinol* in the autumn (12.83%), and *α-pinene* (15.94%) in the winter [71]. The oil of the specimen collected in São Paulo state, Brazil, presented spathulenol (7.0–18.0%), *trans*-α-bergamotene (2.6–19.0%), *α-thujene* (4.0–11.5%), and *β-selinene* (2.3–8.5%) as primary constituents, with a significant content variation on the seasonality study. The monoterpenes amount was higher in July, September, and November, mainly for *α-thujene* (10–11%) and *β-pinene* (6–8%), which were detected in low proportion in the other collection months. On the other hand, the number of oxygenated sesquiterpenes increased from January (45.0%) to May (56.2%) [72].

The oil composition of leaves of *E. pyrifera* was monthly monitored and showed as main compounds the monoterpens *β-pinene* (0.0–25.7%), limonene (0.2–22.0%), and 1,8-cineole (0.6–14.7%), followed by the sesquiterpene, caryophyllene oxide (3.9–21.3%). During the annual study, the monoterpens were predominant in nine months, excepting April, October, and December, where the sesquiterpens showed concentrations higher than 70%. The monoterpene hydrocarbon *β-pinene* was the principal constituent in January (25.0%), presented meaningful content in other months, but it was absent in October [44].

Two different leaf oil profiles of *E. uniflora* were monitored during the year seasonal changes. The oil with profiles rich in curzerene (34.0–53.1%) [73], and selin-1,3,7(11)-trien-8-one (43.0%) plus selin-1,3,7(11)-trien-8-one epoxide (20.0–29.0%) [54] have a clear seasonal influence, however these profiles did not interconvert themselves. The curzerene content showed no significant difference
between the dry (42.7%) and rainy (40.8%) seasons, presenting a high level of similarity by HCA analysis [73]. On the other hand, the selin-1,3,7(11)-trien-8-one plus selin-1,3,7(11)-trien-8-one epoxide profile showed a separation into two different clusters for the oils: Cluster I included samples collected in the dry season, and it was characterized by the highest percentage of spathulenol (10.0%) and caryophyllene oxide (4.0%). Cluster II grouped samples collected in the rainy season showed selina-1,3,7(11)-trien-8-one epoxide (29%) as the principal constituent. Additionally, another two subclusters separated the rainy sampling months. The first one characterized by a high percentage of selina-1,3,7(11)-trien-8-one (46.4%), and the second that was rich in selina-1,3,7(11)-trien-8-one epoxide (37.0%) [54].

The leaf oils of Syzygium jambos (L.) Alston, collected in different year seasons, showed an environmental influence due to the foliar nutrients (N, Mn, Co, Fe, S, and Mg), and soil nutrients (Na, Al, S and H+ and Al). The main compounds were (E)-caryophyllene (0–10.86%), α-humulene (0.0–7.1%), α-zingiberene (0.97–17.73%), caryophyllenyl alcohol (1.48–17.14%), caryolan-8-ol (0.0–10.75%), caryophyllene oxide (0.0–5.05%), thujopsan-2-α-ol (1.01–12.19%), and heneicosane (1.73–18.0%) [74].

5. Differences in Oil Composition and Extractions Methods

Differences in EO’s yields and chemical composition may be associated with the extraction technique employed and its conditions. Some studies compared the extraction methods and reported their influence in yields and chemical compositions of oils [74]. The leaf oil of E. involucrata collected in Rio de Janeiro (Brazil) was rich in β-elemene (41.8%), bicyclogermacrene (28.4%) and (E)-caryophyllene (6.7%) when extracted by CO2 supercritical fluid. However, the oil showed quantitative variations of β-elemene (42.4%), bicyclogermacrene (23.0%), and (E)-caryophyllene (13.4%) when extracted by hydrodistillation [37].

6. Biological Activities

6.1. Antibacterial and Antifungal Activity

In the last years, antimicrobial resistance to antibiotics has been increasing due to the adaptive evolution of bacteria and fungi. For this reason, research focused on the potential of new antimicrobial agents based on natural products has been explored, particularly the essential oils [75].

Several published works have reported the potential of Eugenia and Syzygium EOs against Gram-positive and Gram-negative bacteria. Among the Gram-positive strains, Staphylococcus aureus, S. mitis, S. sanguinis, S. epidermis, S. saprophyticus, Listeria monocytogenes, Streptococcus equi, S. mutans, S. sobrinus, Bacillus cereus, B. subtilis, Enterococcus faecalis, Mycobacterium bovis, Kocuria rhizophila were evaluated. Concerning the Gram-negative strains, Prevotella nigrecens, Porphyromonas gingivalis, Escherichia coli, Pseudomonas aeruginosa, Enterobacter aerogenes, Salmonella spp., S. enteritidis, S. typhimurium, and Neisseria gonorrhoeae were included.

Essential oils from Eugenia astringens, E. beaurepaireana, E. brasiliensis, E. chlorophylla, E. stipitata, E. uniflora, and Syzygium cumini were tested against Staphylococcus aureus strains and showed minimal inhibitory concentrations (MICs) ranged from 119.2 to 56,000 µg/mL [29,34,57,68,76,77].

Eugenia uniflora oils showed low activity when tested against Staphylococcus epidermis (MIC 7500 µg/mL) and S. equi (MIC 7500 µg/mL) [51]. The oils of E. involucrata displayed activity against Streptococcus mitis (MIC 400 µg/mL), S. sanguinis (MIC 400 µg/mL), Prevotella nigrecens (MIC 100 µg/mL), and Porphyromonas gingivalis (MIC 100 µg/mL) [39]. The oils of E. astringens (MIC 477 µg/mL), E. beaurepaireana (MIC 556.6 µg/mL), E. brasiliensis (MIC 624.9–1000 µg/mL) and E. stipitata (inhibition halo 11 mm, tetracycline standard 7 mm) presented antibacterial activity against Escherichia coli [29,68,71,77]. The oil of E. dysenterica showed activity against Cryptococcus neoformans var. neoformans (MIC 250 µg/mL) and C. neoformans var. gattii (MIC 250 µg/mL) [35]. Oils of E. astringens (MIC 470 µg/mL), E. beaurepaireana (MIC 270 µg/mL), E. brasiliensis (MIC 500–1000 µg/mL), E. stipitata
Additionally, oils of *Eugenia uniflora* presented a moderate activity against *Listeria monocytogenes* (MIC 1040 µg/mL) [57], differentially of *E. stipitata* oil (inhibition halo 12 mm, tetracycline standard that showed no inhibition) [77]. Furthermore, the oils of *Eugenia uniflora* showed activity against *Streptococcus mutans* (MIC 50 µg/mL), *S. sobrinus* (MIC 50 µg/mL), and *Kocuria rhizophila* (MIC 500 µg/mL) [34]. However, it was not active against *Enterobacter aerogenes* (MIC 3.125%) and *Salmonella typhimurium* (MIC 3.125%) [78]. With respect to antifungal activity, the oils of *E. uniflora* displayed fungicidal activity against *Paracoccidioides brasiliensis* with MICs between 62.5 and 500 µg/mL [49]; and many *Candida* species, as *Candida dubliniensis* (MIC 230 µg/mL), *C. tropicalis* (MIC 900 µg/mL), *C. albicans* (MIC 1800 µg/mL), *C. glabrata* (MIC 930 µg/mL), *C. parapsilosis* (MIC 3750 µg/mL), *C. grubii* serotype A (MIC 450 µg/mL), *C. gattii* serotype C (MIC 1800 µg/mL), *C. gattii* serotype B (MIC 220 µg/mL), *C. neoformans* serotype D (MIC 110 µg/mL), *C. lipolytica* (MIC 93.7 µg/mL), and *C. guilliermondii* (MIC 109.4 µg/mL) [51,57]. Additionally, the oils of *E. chlorophylla* showed activity against *Candida albicans* (MIC 500 µg/mL) and *C. tropicalis* (MIC 500 µg/mL) [34].

6.2. Acetylcholinesterase Inhibition

Acetylcholinesterase (AChE) acts in the final step of the nervous impulse transmission by hydrolysis of acetylcholine (ACh), a neurotransmitter. AChE inhibitors reduce the ACh level in the brain and increase its concentration in the synapses [79]. In this sense, these inhibitors represent major compounds approved for clinical use in the symptomatic management of Alzheimer’s disease and other neurodegenerative disorders [80]. In recent years, natural compounds from essential oils have shown a high anticholinesterase potential [79].

There are few studies on *Eugenia* and *Syzygium* oils with this focus in South America. The essential oil of *Eugenia sucula*, collected in Rio de Janeiro, Brazil, rich in (E)-caryophyllene (24.6%), showed antiacetylcholinesterase activity (IC$_{50}$ 4.66 µg/mL) by the Elman colorimetric method when compared to physostigmine (IC$_{50}$ 0.59 µg/mL), the positive control [46]. Moreover, the oil of *E. verticillata* (syn. *E. riedeliana*) from Sào Paulo (Brazil), rich in valerianol (28.1%), showed an IC$_{50}$ of 67.3 µg/mL [64]. On the other hand, the oil of *E. brasiliensis* sampled in Santa Catarina, Brazil, with a seasonal variation of α-pinene (1.77–15.94%), β-pinene (2.98–11.24%), spathulenol (8.10–18.17%), 1-epi-cubenol (4.83–7.46%) and τ-cadinol (10.38–15.30%), displayed low antiacetylcholinesterase activity (IC$_{50}$ >1000 µg/mL), when compared to the control galanthamine (IC$_{50}$ 6.93 µg/mL) [71].

6.3. Cytotoxic Activity

The anticancer potential of essential oils has been widely studied, aiming to mitigate the resistance development to multiple drugs and side effects associated with antitumor drugs currently used [81]. Essential oils of *E. egensis*, *E. flavescens*, *E. patrisii*, *E. polystachya* and *E. uniflora* showed cytotoxic activity against human cancer cell lines. The main human carcinoma cell lines evaluated were colorectal (HCT-116), gastric (AGP-01), and melanoma (SKMEL-19) [37,48].

The essential oil of *E. uniflora*, sampled in Brazil, where the selin-1,3,7(11)-trien-8-one, selin-1,3,7(11)-trien-8-one epoxide, and curzerene were the primary constituents, showed high cytotoxic activity against the colorectal (HCT-116), gastric (AGP-01) and melanoma (SKMEL-19) strains, with IC$_{50}$ values between 8.73 to 16.26 µg/mL [48]. However, another *E. uniflora* oil chemotypes, where predominated a mixture of caryophyllene oxide, selin-1,3,7(11)-trien-8-one, selin-1,3,7(11)-trien-8-one epoxide, germacrene B, curzerene and (E)-caryophyllene, did not show cytotoxic activity against these cells [48]. Furthermore, oils of *E. flavescens*, *E. patrisii*, and *E. polystachya*, rich in γ-bisabolene, (2E,6E)-farnesol and germacrene D, displayed activity against colorectal carcinoma (HCT-116) with IC$_{50}$ values of 13.9, 16.4 and 10.3 µg/mL, respectively. On the other hand, the oil of *E. egensis*, rich in 5-hydroxy-cis-calemene (35.8%) did not show cytotoxic activity against this cell line (IC$_{50}$ > 25 µg/mL) [37].
6.4. Antiprotozoal Activity

The protozoal or parasitic diseases induce significant morbidity and mortality, being endemic to developing countries. Thus, affordable drugs have serious side effects, high cost, and low effectiveness. Moreover, there is an increased need to expand the investigations for new drug development [82]. In this way, essential oils could be promising sources of antiprotozoal agents, opening perspectives to discover more effective drugs of vegetal origin, in the treatment of diseases caused by protozoa [83].

The essential oils of *E. uniflora* and *S. cumini* showed anti-Leishmanial activity. *Eugenia uniflora* oil from Brazil, rich in curzerene (47.3%), displayed significant anti-Leishmanial activity against promastigote (IC50, 3.04 μg/mL) and amastigote (IC50 1.92 μg/mL) forms of *Leishmania amazonensis*. The oil was 20 times more toxic to amastigotes than to healthy macrophages. Although the anti-Leishmanial activity was not mediated by nitric oxide production, the authors have suggested that macrophage activation may be involved in the anti-Leishmanial activity of *Eugenia uniflora* essential oil, as evidenced by increases in both the phagocytic capacity and lysosomal activity [47]. Moreover, the essential oil of *Syzygium cumini* collected in Maranhão state (Brazil), rich in a-pinene (31.85%), showed anti-Leishmanial activity against promastigote form of *L. amazonensis* (IC50 60 μg/mL) [67].

6.5. Antioxidant Activity

The antioxidants usually act as free radical scavengers, preventing oxidative stress, decreasing the possibility of chronic and degenerative diseases, controlling the autoxidation, and interrupting the propagation of free radicals, or by inhibiting the formation of free radicals via different mechanisms. Some aromatic plants are sources of natural antioxidants [83–86]. The antioxidant activity of *Eugenia* essential oils has been evaluated by radical scavenging DPPH, ABTS, and TLC methods, lipid peroxidation by β-carotene/linoleic acid, and the iron ion reduction power.

The oil of *E. involucrata*, obtained by supercritical fluid extraction (SFE), showed inhibition of 93.6% to the β-carotene oxidation, while its oil extracted by hydrodistillation was shown to be inactive, with only 6.4% of inhibition. The main compounds were β-elemene (41.8–42.4%), bicyclogermacrene (23.0–28.4%), and (E)-caryophyllene (6.7–13.4%), in both oil samples. The higher activity of the sample obtained by the extraction using SFE can be attributed to the nonvolatile compounds present in the extract [38,87]. Additionally, its fruit oil was rich in (E)-caryophyllene (10.1%), spathulenol (7.8%), and β-bisabolene (7.2%), displaying a significant antioxidant activity by TLC-DPPH method [40].

The oils of *E. uniflora* collected in the Brazilian Amazon with different chemical profiles showed more antioxidant activity in the DPPH method than using the β-carotene/linoleic acid system [48]. Two different samples with higher content of caryophyllene oxide, selin-1,3,7(11)-tri-en-8-one, selin-1,3,7(11)-tri-en-8-one epoxide, and curzerene (50.6%) presented an inhibition of DPPH radical of 45.1% (228.3 μg/mL) and 42.8% (217.0 mg/mL), respectively [47]. The leaf oils rich in curzerene (36.2–53.1%) exhibited an antioxidant capacity varying from moderate (42.6%) to high (64.2%), according to seasonality and corresponding to 186.9 mg/mL and 400.3 mg/mL, respectively [76]. The leaf oils rich in germacrene B (18.4%), curzerene (13.4%) and (E)-caryophyllene (9.1%) exhibited an inhibition of 40.6% (205.6 mg/mL), and the oil samples rich in selin-1,3,7(11)-tri-en-8-one (32.6–43.1%) and selin-1,3,7(11)-tri-en-8-one epoxide (21.7–30.4%), displayed a DPPH radical scavenging of 30.3–35.3%, corresponding to 153.5–178.8 mg/mL [48]. Another specimen of *E. uniflora*, collected in South Brazil, with an oil rich in germacrene B (21.2%) and selin-1,3,7(11)-tri-en-8-one epoxide (19.3%), displayed low activity in the DPPH method (IC50 833 μg/mL) and high activity to reduce ABTS radical (IC50 8.1 μg/mL). The results have suggested that its antioxidant activity mechanism is based on electron transfer [57]. Additionally, the EO from fruits, composed of hexadecanoic acid (11.7%), (E)-β-ocimene (7.4%), and α-selinene (7.2%) showed a high antioxidant potential by TLC-DPPH method [40].

The essential oil of *E. egesis*, rich in 5-hydroxy-cis-calamenene (35.8%), displayed a strong DPPH radical scavenging (79.6%, 216.5 mg/mL, 177.6 mg, BHA, mL), about five times less than the standards Trolox and BHA [37]. On the other hand, the *E. flavescens* oil, rich in (E)-γ-bisabolene (35.0%) and β-bisabolene (34.7%), showed a moderate DPPH radical scavenging (45.1%, 122.6 mg/mL,
100.6 mgBHAЕ/mL). Additionally, the E. patrisii oil, rich in (2E,6Z)-farnesol (34.5%) and (2E,6E)-farnesol (23.2%), presented a moderate inhibition of 40.9% (111.2 mgTE/mL, 91.3 mgBHAЕ/mL) [37]. The oil of E. brasiliensis displayed a low activity against DPPH (IC_{50} > 500 μg/mL), β-carotene/linoleic acid (1.67–14.05%), and iron reducing power (60.37–94.32 mg.AA/g) methods [71]. Similarly, the oil of E. polystachya showed a low activity (11.5%) against the DPPH method [37].

**6.6. Other Biological Activities**

The oils of E. langsdorff from Brazil showed promise as a natural acaricide, with high fumigation potential observed for the leaf oil rich in epi-longipinanol (13.6%), γ-eudesmol (12.3%), limonene (11.8%), and 10-epi-γ-eudesmol (10.6%), and the fruit oil, where predominated 10-epi-γ-eudesmol (35.7%) and 1,10-di-epi-cubenol (15.6%). The LC_{50} (median lethal concentration) values determined for the leaf and fruit oils were 1.7 μL/L and 3.06 μL/L in air. However, in the experiment based on the residual contact, the effect of the leaf oil (LC_{50} 21.90 μL/L) was less toxic than the fruit oil (LC_{50} 12.25 μL/L) [42].

The oils of E. stictopetala (LC_{50} 230 μg/mL) [63] and E. triquetra (64.9 μg/mL) were active as larvicidal against Aedes aegypti larvae [65]. The pure essential oil of E. sulcata applied as a topical treatment on the arthropods Oncopeltus fasciatus and Dysdercus peruvianus, caused a mortality rate of 96.7% and 80.0%, respectively [45].

The essential oils of E. uniflora presented antinociceptive activity (ED_{50} 218.6 mg/kg), and hypothermic [52] and hepatoprotective effects induced by acetaminophen [88] in vivo assays with mice. Moreover, the Syzygium cumini oil from Rio de Janeiro state, Brazil, rich in α-pinene (22.2%), (Z)-β-ocimene (10.2%), (E)-caryophyllene (9.45%) showed anti-inflammatory activity by lipopolysaccharide-induced pleurisy model, with eosinophils inhibition of 67% [66]. Additionally, S. cumini oil from São Luís state, Brazil, rich in α-pinene (31.85%), (Z)-β-ocimene (28.98%), and (E)-β-ocimene (11.71%), presented molluscicide potential against Biomphalaria glabrata (LC_{50} 90 μg/mL) [67]. The distribution of reports on the biological activity of Eugenia and Syzygium species with occurrence in South America are represented in Figure 2.

![Figure 2. Distribution of reports on biological activity of Eugenia and Syzygium species.](image-url)
7. Conclusions

The Eugenia and Syzygium species from South America have high chemical variability and several biological activities in their essential oils. Based on the present study, these variations can be attributed to different geographic occurrences or seasonal changes. Despite the medicinal use of Eugenia species in traditional therapy, few reports have been made about the biological activity of its essential oils (less than 5% of species in Brazil). On the other hand, the composition of oils from a large number of species (127 samples) is already known, representing 12.2% of the total species distributed in Brazil. The variations in the chemical profile of these species indicated the importance of optimizing the protocols for collecting, processing, and extracting plant material. The standardization of these essential oils can contribute to the economic and commercial exploitation of bioactive products from aromatic plants.

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Abbreviations

| Abbreviation | Definition |
|--------------|------------|
| ABTS        | 2,2′-azino-bis (3-ethylbenzothiazoline-6-sulfonic acid) |
| ACh         | Acetylcholine |
| AChE        | Acetylcholine esterase |
| BHA         | Butylated hydroxyanisole |
| BR          | Brazil |
| CO          | Colombia |
| DPPH        | 2,2-Diphenyl-1-picrylhydrazyl (radical) |
| EDS         | Median Effective dose |
| EO          | Essential oils |
| GC–MS       | Gas chromatography–mass spectrometry |
| HCA         | Hierarchical Cluster Analysis |
| HD          | Hydrodistillation |
| IC50        | Median inhibitory concentration |
| LC50        | Median lethal concentration |
| mg.AA/g     | Milligram of ascorbic acid per gram |
| mg.BHAE/mL  | Milligram BHA equivalent per gram |
| mgTE/g      | Milligram trolox equivalent per gram |
| MMIC        | Minimum inhibitory concentration |
| MMC         | Minimum microbicidal concentration |
| NI          | Unidentified compound |
| NMR         | Nuclear magnetic resonance |
| PCA         | Principal component analysis |
| SD          | Steam distillation |
| SFE         | Supercritical fluid extraction |
| SPME        | Solid Phase Micro Extraction |
| spp.        | Species (plural) |
| TLC         | Thin-layer chromatography |
| VE          | Venezuela |
Appendix A

Table A1. Species of *Eugenia* and *Syzygium* in South America: essential oil composition and biological activity.

| Species         | Occurrence                          | Essential Oil | Primary Components (>5%)                                                                 | Essential Oil Bioactivity                                                                 | Ref. |
|-----------------|-------------------------------------|----------------|------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------|------|
| *E. acutata*    | Valinhos, São Paulo, Brazil         | Leaf (SD)      | *E*-caryophyllene (26.93%), δ-cadinene (6.01%), germacrene D (5.29%)                      | —                                                                                       | [60] |
| *E. arenosa*    | Manuel Viana, Rio Grande do Sul, Brazil | Leaf (HD)      | farnesyl acetate (70.4%), aromadendrene (11.7%), globulol (7.1%)                         | —                                                                                       | [32] |
| *E. astringens* | Macaé, Rio de Janeiro, Brazil       | Leaf (HD)      | α-pinene (15.8%), 1-epi-cubenol (11.9%), α-eudesmol (10.6%), selin-11-en-4x-ol (9.4%)   | —                                                                                       | [28] |
| *E. astringens* | Rio de Janeiro, Rio de Janeiro, Brazil | Leaf (HD)      | α-pinene (19.7-34.4%), β-pinene (20.6-34.1%), *E*-caryophyllene (3.6-11.7%)               | —                                                                                       | [26] |
| *E. astringens* | Juréia, São Paulo, Brazil           | Leaf (HD)      | α-pinene (24.7%), *E*-caryophyllene (14.6%), β-pinene (11.0%), myrcene (7.7%)             | —                                                                                       | [27] |
| *E. astringens* | Santo Amaro da Imperatriz, Santa Catarina, Brazil | Leaf (HD) | viridiflorol (17.7%), δ-pinene (13.2%), α-pinene (11.2%), aromadendrene (6.9%) | Antibacterial, broth dilution assay (*Staphylococcus aureus*, MIC 119.2 µg/mL; *Pseudomonas aeruginosa*, MIC 447.0 µg/mL; *Escherichia coli*, MIC 477.0 µg/mL) | [29] |
| *E. bacopari*   | Dom Pedro de Alcântara, Rio Grande do Sul, Brazil | Leaf (HD) | δ-cadinene (15.8%), aromadendrene (12.2%), viridiflorene (7.9%), globulol (6.8%)         | —                                                                                       | [59] |
| *E. beaurepaireana* | Morro dos Sul, Rio Grande do Sul, Brazil | Leaf (HD) | bicyclogermacrene (14.3%), germacrene D (8.6%), δ-cadinene (7.2%), *E*-caryophyllene (6.4%) | —                                                                                       | [30] |
| *E. beaurepaireana* | Santo Amaro da Imperatriz, Santa Catarina, Brazil | Leaf (HD) | *E*-caryophyllene (8.0%), bicyclogermacrene (7.2%), valencene (5.5%), δ-cadinene (4.9%), spathulenol (4.9%), viridiflorol (4.9%) | Antibacterial, broth dilution assay (*Staphylococcus aureus*, 1110 µg/mL; *Pseudomonas aeruginosa*, MIC 278.3 µg/mL; *Escherichia coli*, MIC 556.6 µg/mL) | [29] |
| *E. biflora*    | Belém, Pará, Brazil                  | Leaf (HD)      | caryophyllene oxide (28.6%), *E*-caryophyllene (16.8%), τ-murrolol (8.4%), 1-epi-cubenol (6.4%), α-cadinol (6.2%) | —                                                                                       | [24] |
Table A1. Cont.

| Species       | Occurrence           | Essential Oil | Primary Components (>5%)                                                                 | Essential Oil Bioactivity                  | Ref. |
|---------------|----------------------|---------------|------------------------------------------------------------------------------------------|--------------------------------------------|------|
| *E. biflora*  | Belém, Pará, Brazil  | Leaf (HD)     | caryophyllene oxide (20.5%), E-caryophyllene (11.4%), *epi*-α-murrolol (7.6%), *1-epi*-cubenol (6.6%), α-cadinol (6.3%) | —                                          | [24] |
| *E. biflora*  | Belém, Pará, Brazil  | Leaf (HD)     | E-caryophyllene (9.8%), globulol (9.8%), germacrene B (7.9%), α-cadinol (7.5%)             | —                                          | [24] |
| *E. biflora*  | Belém, Pará, Brazil  | Leaf (HD)     | α-cadinol (14.7%), τ-murrolol (8.3%), caryophyllene oxide (7.5%), copaborneol (5.6%)        | —                                          | [24] |
| *E. biflora*  | Maracanã, Pará, Brazil | Leaf (HD)    | E-caryophyllene (15.4%), germacrene D (6.6%), α-copaene (6.0%), bicyclogermacrene (5.8%)   | —                                          | [25] |
| *E. biflora*  | Maracanã, Pará, Brazil | Leaf (HD)    | β-pinene (27.8%), α-pinene (27.3%), limonene (6.7%)                                      | —                                          | [25] |
| *E. brasiliensis* | Blumenau, Santa Catarina, Brazil | Leaf (HD) | β-pinene (10.4%), α-pinene (10.3%), spathulenol (7.7%), τ-cadinol (7.1%)                   | —                                          | [32] |
| *E. brasiliensis* | Florianópolis, Santa Catarina, Brazil | Leaf (HD) | α-pinene (1.77-15.94%), β-pinene (2.98-11.24%), spathulenol (8.10-18.17%), *1-epi*-cubenol (4.83-7.46%), τ-cadinol (10.38-15.30%) | Antibacterial, broth dilution assay (Staphylococcus saprophyticus, MIC 500-1000 µg/mL; Escherichia coli, MIC 1000 µg/mL; Pseudomonas aeruginosa, MIC 500-1000 µg/mL) Antioxidant (DPPH IC_{50} > 500 µg/mL; β-carotene/linoleic acid, 1.67-14.05%; iron reducing power, 60.37-94.32 mg AA/g) Enzyme inhibitory (acetylcholinesterase, IC_{50} >1000 µg/mL) | [71] |
| *E. brasiliensis* | Moji-Guaçu, São Paulo, Brazil | Leaf Purple Variety (HD) | α-pinene (18.8%), β-pinene (11.0%), 1,8-cineol (9.6%), limonene (8.6%), τ-cadinol (6.8%) | —                                          | [33] |
| *E. brasiliensis* | Moji-Guaçu, São Paulo, Brazil | Leaf Yellow Variety (HD) | α-pinene (33.5%), 1,8-cineol (28.2%), β-pinene (14.4%), myrcene (5.5%) | —                                          | [33] |
Table A1. Cont.

| Species       | Occurrence                             | Essential Oil       | Primary Components (>5%)                        | Essential Oil Bioactivity                                                                 | Ref.  |
|---------------|----------------------------------------|---------------------|-------------------------------------------------|------------------------------------------------------------------------------------------|-------|
| *E. brasiliensis* | Moji-Guaçu, São Paulo, Brazil          | Fruit Yellow Variety (HD) | α-pinene (15.4%), myrcene (10.7%), α-terpineol (10.2%), β-pinene (9.3%) | —                                                                                       | [33]  |
| *E. brasiliensis* | Santo Amaro da Imperatriz, Santa Catarina, Brazil | Leaf (HD)           | spathulenol (12.7%), τ-cadinol (8.7%), viridiflorol (7.1%), 1-c-p-cubenol (6.3%), α-cadinol (6.6%) | Antibacterial, broth dilution assay (*Staphylococcus aureus*, MIC 156.2 µg/mL; *Pseudomonas aeruginosa*, MIC 624.9 µg/mL; *Escherichia coli*, MIC 624.9 µg/mL) | [29]  |
| *E. brasiliensis* | São Paulo, São Paulo, Brazil            | Leaf (HD) b          | spathulenol (7.0-18.0%), trans-α-bergamotene (2.6-19.0%), α-thujene (4.0-11.5%), β-selinene (2.3-8.5%) | —                                                                                       | [72]  |
| *E. brasiliensis* | Moji-Guaçu, São Paulo, Brazil           | Fruit Purple Variety (HD) | caryophyllene oxide (22.2%), α-cadinol (10.4%), τ-cadinol (9.9%), β-bisabolene (9.6%) | —                                                                                       | [33]  |
| *E. burkartiana* | Parque do Turvo, Rio Grande do Sul, Brazil | Leaf (HD)           | bicyclogermacrene (14.2%), germacrene D (8.8%), E-caryophyllene (7.8%), 3E-hexenol (6.7%) | —                                                                                       | [59]  |
| *E. candelleana* | Rio de Janeiro, Rio de Janeiro, Brazil  | Leaf (SD)            | 1-c-p-cubenol (77.59%), δ-elemene (13.87%), muurola-4,10(14)-dien-1β-ol (8.68%), globulol (5.52%), α-cadinol (5.26%) | —                                                                                       | [60]  |
| *E. catharinensis* | Blumenau, Santa Catarina, Brazil        | Leaf (HD)            | ethyl palmitate (10.5%), trans-α-bergamotene (6.5%), β-selinene (5.9%) | —                                                                                       | [59]  |
| *E. chlorophylla* | Curitiba, Paraná, Brazil                | Fine Stem (HD)       | caryophyllene oxide (17.2%), τ-muurolol (16.8%), globulol (16.5%), α-cadinol (12.1%), 1-c-p-cubenol (10.9%) | Antimicrobial, agar diffusion assay and broth dilution assay (*Kocuria rhizophila*, MIC 500 µg/mL; *Staphylococcus aureus*, MIC 500 µg/mL; *Streptococcus mutans*, MIC 50.0 µg/mL; *Streptococcus sobrinus*, MIC 50.0 µg/mL) | [34]  |
| Species                  | Occurrence                        | Essential Oil       | Primary Components (>5%)                                                                 | Essential Oil Bioactivity                                                                 | Ref. |
|--------------------------|-----------------------------------|---------------------|------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------|------|
| *E. chlorophylla*        | Curitiba, Paraná, Brazil          | Leaf (HD)           | globulol (22.5%), 1-10-di-epi-cubenol (9.8%), τ-muurolol (8.1%), caryophyllene oxide (6.4%) | Antimicrobial, agar diffusion assay and broth dilution assay                               | [34] |
|                          |                                   |                     |                                                                                          | (Kocuria rhizophila, MIC 500, 50.0 µg/mL; Staphylococcus aureus, MIC 500, 50.0 µg/mL;       |      |
|                          |                                   |                     |                                                                                          | Streptococcus mutans, MIC 50.0 µg/mL;                                                    |      |
|                          |                                   |                     |                                                                                          | Streptococcus sobrinus, MIC 50.0 µg/mL;                                                   |      |
|                          |                                   |                     |                                                                                          | *Candida albicans*, MIC 50.0 µg/mL;                                                      |      |
|                          |                                   |                     |                                                                                          | *Candida tropicalis*, MIC 50.0 µg/mL                                                     |      |
| *E. chlorophylla*        | Curitiba, Paraná, Brazil          | Leaf at flowering   | globulol (18.9%), α-cadinol (9.4%), 1-epi-cubenol (8.1%), E-caryophyllene (8.1%),         | Antimicrobial, agar diffusion assay and broth dilution assay                               | [34] |
|                          |                                   | stage (HD)          | caryophyllene oxide (5.8%)                                                                | (Kocuria rhizophila, MIC 500 µg/mL; Staphylococcus aureus, MIC 500 µg/mL; Streptococcus |      |
|                          |                                   |                     |                                                                                          | mutans, MIC 50.0 µg/mL;                                                                  |      |
|                          |                                   |                     |                                                                                          | Streptococcus sobrinus, MIC 50.0 µg/mL;                                                   |      |
|                          |                                   |                     |                                                                                          | *Candida albicans*, MIC 50.0 µg/mL                                                       |      |
|                          |                                   |                     |                                                                                          | *Candida tropicalis*, 50.0 µg/mL                                                         |      |
| *E. chlorophylla*        | Curitiba, Paraná, Brazil          | Flower (HD)         | E-caryophyllene (12.8%), α-cadinol (10.1%), caryophyllene oxide (8.9%),                    | Antimicrobial, agar diffusion assay and broth dilution assay                               | [34] |
|                          |                                   |                     | τ-muurol (8.5%)                                                                           | (Kocuria rhizophila, MIC 500 µg/mL; Staphylococcus aureus, MIC 500 µg/mL; Streptococcus |      |
|                          |                                   |                     |                                                                                          | mutans, MIC 50.0 µg/mL;                                                                  |      |
|                          |                                   |                     |                                                                                          | Streptococcus sobrinus, MIC 50.0 µg/mL;                                                   |      |
|                          |                                   |                     |                                                                                          | *Candida albicans*, MIC 50.0 µg/mL                                                       |      |
|                          |                                   |                     |                                                                                          | *Candida tropicalis*, 50.0 µg/mL                                                         |      |
| *E. copacabanensis*      | Rio de Janeiro, Rio de Janeiro,    | Leaf (SD)           | 1,10-di-epi-cubenol (14.24%), caryophyllene oxide (6.75%), epi-α-cadinol (5.03%),         | —                                                                                        | [60] |
|                          | Brazil                             |                     | γ-cadinene (4.17%)                                                                         |                                                                                          |      |
| *E. copacabanensis*      | Rio de Janeiro, Rio de Janeiro,    | Leaf (HD)           | β-pinene (30.4%), α-pinene (20.2%), E-caryophyllene (10.3%)                                | —                                                                                        | [89] |
|                          | Brazil                             |                     |                                                                                          |                                                                                          |      |
| *E. dimorpha*            | Morro Santana, Rio Grande do Sul,  | Leaf (HD)           | α-pinene (22.4%), α-humulene (12.9%), 1,8-cineole (9.9%)                                   | —                                                                                        | [59] |
|                          | Brazil                             |                     |                                                                                          |                                                                                          |      |
| *E. dysenterica*         | Catalão, Goiás, Brazil             | Leaf (HD)           | E-caryophyllene (14.8%), α-humulene (11.0%), α-terpineol (6.1%), limonene (5.5%),         | Antibacterial, broth dilution assay                                                      | [35] |
|                          | Brazil                             |                     | δ-cadinene (5.8%), caryophyllene oxide (5.4%)                                             | (Cryptococcus neoformans var neoformans, MIC 250 µg/mL; Cryptococcus neoformans var gattii, |      |
|                          |                                   |                     |                                                                                          | MIC 250 µg/mL                                                                            |      |
| *E. dysenterica*         | Senador Canedo Santa Catarina,     | Leaf (HD)           | E-caryophyllene (18.0%), β-pinene (9.3%), α-pinene (9.0%), limonene (7.8%), Z-β-ocimene (5.9%) | —                                                                                        | [36] |
Table A1. Cont.

| Species          | Occurrence                  | Essential Oil | Primary Components (>5%)                                                                 | Essential Oil Bioactivity                                                                 | Ref. |
|------------------|-----------------------------|---------------|------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------|------|
| E. dysenterica   | Campo Alegre de Goiás, Goiás, Brazil | Leaf (HD)     | E-caryophyllene (24.0%), δ-cadinene (13.0%), α-copaene (9.6%), α-pinene (6.4%), caryophyllene oxide (4.5%) | —                                                                                         | [36] |
| E. egensis       | Marabá, Pará, Brazil       | Aerial parts (HD) | 5-hydroxy-Z-calamenene (35.8%), E-caryophyllene (8.9%), trans-muurola-3,5-diene (5.9%), trans-calamenene (6.1%), trans-cadin-1,4-diene (6.3%), ledol (5.0%) | Antioxidant, DPPH assay (79.6%, 216.5 mgTE/mL, 177.6 mgBHAE/mL) Cytotoxic (HCT-116 human colorectal carcinoma, IC₅₀ > 25 µg/mL; MRC5 human fibroblast IC₅₀ > 25 µg/mL; AGP-01 human gastric adenocarcinoma, IC₅₀ > 25 µg/mL; SKMEL-19 human melanoma, IC₅₀ > 25 µg/mL) | [37] |
| E. expansa       | Peruibe, São Paulo, Brazil | Leaf (HD)     | spathulenol (12.1%), E-caryophyllene (9.2%), caryophyllene oxide (8.7%), α-copaene (5.0%) | —                                                                                         | [32] |
| E. flavescens    | Maracanã, Pará, Brazil     | Leaf (HD)     | germacrene D (14.5%), bicyclogermacrene (11.7%), δ-cadinene (5.7%)                      | —                                                                                         | [25] |
| E. flavescens    | Parauapebas, Pará, Brazil  | Aerial parts (HD) | E-γ-bisabolene (35.0%), β-bisabolene (34.7%), E-iso-γ-bisabolene (5.1%)                  | Antioxidant, DPPH assay (45.1%, 122.6 mgTE/mL, 100.6 mgBHAE/mL) Cytotoxic (HCT-116 human colorectal carcinoma, IC₅₀ 13.99 µg/mL; MRC5 human fibroblast IC₅₀ 14.0 µg/mL; AGP-01 human gastric adenocarcinoma, IC₅₀ > 25 µg/mL; SKMEL-19 human melanoma, IC₅₀ > 25 µg/mL) | [37] |
| E. florida       | Derrubadas, Rio Grande do Sul, Brazil | Leaf (HD)     | E-caryophyllene (14.5%), β-elemene (11.8%), α-copaene (7.9%), α-cadinol (5.8%)             | —                                                                                         | [61] |
| E. gracillima    | Porto Rico, Paraná, Brazil  | Leaf (HD)     | globulol (8.7%), viridiflorene (6.9%), epi-globulol (6.8%), spathulenol (5.9%)            | —                                                                                         | [61] |
| E. hiemalis      | Morro Santana, Rio Grande do Sul, Brazil | Leaf (HD)     | bicyclogermacrene (37.7%), E-caryophyllene (7.4%), germacrene D (7.0%), globulol (5.8%) | —                                                                                         | [62] |
| Species          | Occurrence            | Essential Oil | Primary Components (>5%)                                                                                                                                                                                                 | Essential Oil Bioactivity                                                                                      | Ref.  |
|------------------|-----------------------|---------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------|-------|
| *E. involucrata* | Urupema, Santa Catarina, Brazil | Leaf (SFE)    | β-elemene (41.8%), bicyclogermacrene (28.4%), E-caryophyllene (6.7%), germacrene D (7.4%)                                                                                                                                   | Antioxidant (β-carotene/linoleic acid assay, at 1 mg/mL, 93.8%)                                                | [38]  |
| *E. involucrata* | Urupema, Santa Catarina, Brazil | Leaf (HD)     | β-elemene (42.4%), bicyclogermacrene (23.0%), E-caryophyllene (13.4%)                                                                                                                                                | Antioxidant (β-carotene/linoleic acid assay, at 1 mg/mL, 6.41%)                                               | [38]  |
| *E. involucrata* | Uberlândia, Minas Gerais, Brazil | Leaf (HD)     | spathulenol (21.4%), bicyclogermacrene (19.3%), E-caryophyllene (8.6%), caryophyllene oxide (7.7%)                                                                                                                  | Antibacterial, broth dilution assay (*Streptococcus mitis*, MIC 400 µg/mL; *S. sanguinis*, MIC 400 µg/mL; *Prevotella nigrescens*, MIC 100 µg/mL; *Porphyromonas gingivalis*, MIC 100 µg/mL) | [39]  |
| *E. involucrata* | Pelotas, Rio Grande do Sul, Brazil, Fruit (HD) | Fruit (HD) | E-caryophyllene (10.1%), spathulenol (7.8%), β-bisabolene (7.2%), γ-eudesmol (6.0%)                                                                                                                                    | Antioxidant, DPPH assay (TLC method)                                                                           | [40]  |
| *E. joenssonii*  | Morro Santana, Rio Grande do Sul, Brazil | Leaf (HD)     | 5-epi-paradisiol (8.4%), δ-selinene (7.9%), β-selinene (7.2%), neointermedeol (6.3%), α-cadinol (5.3%)                                                                                                                 | —                                                                                                               | [59]  |
| *E. klotzschiana* | Portelândia, Goiás, Brazil | Leaf (HD)     | α-copaene (10.6%), α-humulene (5.5%), spathulenol (8.7%), caryophyllene oxide (7.4%), τ-muurolol (5.4%), α-cadinol (6.2%)                                                                                             | —                                                                                                               | [41]  |
| *E. klotzschiana* | Portelândia, Goiás, Brazil | Leaf (HD)     | β-bisabolene (14.0%), spathulenol (10.9%), caryophyllene oxide (6.3%), bicyclogermacrene (5.4%)                                                                                                                          | —                                                                                                               | [41]  |
| *E. klotzschiana* | Portelândia, Goiás, Brazil | Leaf (HD)     | β-bisabolene (17.4%), germacrene D (13.3%), α-humulene (10.2%), α-trans-bergamotene (10.1%), spathulenol (7.2%)                                                                                                       | —                                                                                                               | [41]  |
| *E. klotzschiana* | Portelândia, Goiás, Brazil | Flower (HD)   | α-trans-bergamotene (29.9%), germacrene D (12.1%), β-bisabolene (10.2%)                                                                                                                                             | —                                                                                                               | [41]  |
| Species       | Occurrence                          | Essential Oil | Primary Components (>5%)                                                                                      | Essential Oil Bioactivity                                                                 | Ref. |
|---------------|-------------------------------------|---------------|---------------------------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------|------|
| *E. langsdorfi* | Brasilia, Distrito Federal, Brazil  | Leaf (HD)     | \( \text{epi-}\text{longipinanol (13.6%)}, \text{γ-eudesmol (12.3%)}, \text{limonene (11.8%)}, \text{10-epi-γ-eudesmol (10.6%)}, \text{maaliol (6.2%)} \) | Acaricidal (Tetranychus urticae, Fumigation LC\(_{50}\) 1.79 µL/L; residual contact LC\(_{50}\) 21.90 µL/L) | [42] |
| *E. langsdorfi* | Brasilia, Distrito Federal, Brazil  | Fruit (HD)    | \( \text{10-epi-γ-eudesmol (35.7%)}, \text{1,10-di-epi-cubenol (15.6%)}, \text{caryophyllene oxide (7.5%)}, \text{epi-}\text{longipinanol (7.3%)}, \text{isolongifolan-7-α-ol (7.1%)} \) | Acaricidal (Tetranychus urticae, Fumigation LC\(_{50}\) 3.06 µL/L; residual contact LC\(_{50}\) 12.25 µL/L) | [42] |
| *E. multicostata* | Dom Pedro de Alcântara, Rio Grande do Sul, Brazil | Leaf (HD) | \( \text{α-pinene (16.1%)}, \text{spathulenol (10.7%)}, \text{epi-globuol (7.8%)}, \text{β-pinene (7.3%)} \) | —                                                                                         | [32] |
| *E. patrisii* | Maracanã, Pará, Brazil               | Leaf (HD)     | \( \text{trans-cadin-1,4-diene (16.5%)}, \text{trans-muurola-3,5-diene (13.3%)}, \text{E-caryophyllene (11.1%)}, \text{α-cubebeene (9.8%)} \) | —                                                                                         | [25] |
| *E. patrisii* | São Geraldo do Araguaia, Pará, Brazil | Aerial parts (HD) | \( \text{2E,6E-Farnesol (34.5%)}, \text{2E,6Z-Farnesol (23.2%)}, \text{caryophylla-4(12)-8(13)-dien-5-β-ol (15.6%)} \) | Antioxidant, DPPH assay (40.9%, 111.2 mgTE/mL, 91.3 mgBHAEL/mL) Cytotoxic (HCT-116 human colorectal carcinoma, IC\(_{50}\) 16.4 µg/mL; MRC5 human Fibroblast IC\(_{50}\) 18.1 µg/mL; AGP-01 human gastric adenocarcinoma, IC\(_{50}\) > 25 µg/mL; SKMEL-19 human melanoma, IC\(_{50}\) > 25 µg/mL) | [37] |
| *E. pitanga*  | Manuel Viana, Rio Grande do Sul, Brazil | Leaf (HD)     | \( \text{germacrene D (29.3%)}, \text{bicyclogermacone (22.4%)}, \text{E-β-ocimene (10.5%)}, \text{E-caryophyllene (5.1%)} \) | —                                                                                         | [32] |
| *E. platyna*  | Blumenau, Santa Catarina, Brazil     | Leaf (HD)     | \( \text{β-selinene (17.9%)}, \text{allo-aromadendrene (12.6%)}, \text{7-epi-α-selinene (10.4%)} \) | —                                                                                         | [27] |
| Species     | Occurrence                  | Essential Oil | Primary Components (>5%)                                                                 | Essential Oil Bioactivity                                                                 | Ref.  |
|-------------|-----------------------------|---------------|------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------|-------|
| E. pluriflora | Aparados da Serra, Rio Grande do Sul, Brazil | Leaf (HD)     | $E$-nerolidol (24.6%), $\alpha$-pinene (24.0%), 1,8-cineole (12.7%), $\alpha$-terpineol (5.8%) | —                                                                                         | [27]  |
| E. polystachya | Parauapebas, Pará, Brazil   | Aerial parts (HD) | germacrene D (18.4%), ishwarane (15.7%), 7-epi-$\alpha$-selinene (7.5%), bicyclogermacrene (5.1%), $\alpha$-ylangene (5.0%) | Antioxidant, DPPH assay (11.5%) Cytotoxic (HCT-116 human colorectal carcinoma, $IC_{50}$ 10.3 µg/mL; MRC5 human fibroblast $IC_{50}$ > 25 µg/mL; AGP-01 human gastric adenocarcinoma, $IC_{50}$ > 25 µg/mL; SKMEL-19 human melanoma, $IC_{50}$ > 25 µg/mL) | [37]  |
| E. protenta  | Maracanã, Pará, Brazil      | Leaf (HD)     | germacrene D (15.1%), $\beta$-elemene (12.8%), $\delta$-cadinene (9.3%), $E$-caryophyllene (8.3%), bicyclogermacrene (5.9%), $\alpha$-cadinol (5.5%) | —                                                                                         | [25]  |
| E. protenta  | Santarém Novo, Pará, Brazil | Leaf (HD)     | dimethylxanthoxylin (73.2%), limonene (5.9%)                                              | —                                                                                         | [23]  |
| E. protenta  | Santarém Novo, Pará, Brazil | Leaf (HD)     | dimethylxanthoxylin (83.0%)                                                              | —                                                                                         | [23]  |
| E. protenta  | Santarém Novo, Pará, Brazil | Leaf (HD)     | selin-11-en-4$\alpha$-ol (18.3%), $\beta$-elemene (16.9%), $\beta$-selinene (5.6%), $\delta$-cadinene (5.5%), $\gamma$-selinene (5.1%) | —                                                                                         | [23]  |
| E. protenta  | Magalhães Barata, Pará, Brazil | Leaf (HD)     | selin-11-en-4$\alpha$-ol (14.4%), $\beta$-elemene (12.3%), $\alpha$-selinene (6.5%), cubenol (5.7%) | —                                                                                         | [23]  |
| E. protenta  | Maracanã, Pará, Brazil      | Leaf (HD)     | $\beta$-elemene (9.2%), germacrene D (7.8%), $\alpha$-cadinol (8.1%)                     | —                                                                                         | [23]  |
| E. protenta  | Maracanã, Pará, Brazil      | Leaf (HD)     | germacrene D (15.1%), bicyclogermacrene (11.8%), $\delta$-cadinene (5.8%), $\alpha$-copaene (5.1%), $E$-caryophyllene (5.1%) | —                                                                                         | [23]  |
| Species      | Occurrence                        | Essential Oil  | Primary Components (>5%)                                                                 | Essential Oil Bioactivity | Ref. |
|--------------|-----------------------------------|----------------|-----------------------------------------------------------------------------------------|----------------------------|------|
| *E. protenta* | Maracanã, Pará, Brazil            | Leaf (HD)      | δ-cadinene (15.4%), germacrene D (11.5%), δ-elemene (8.5%), E-caryophyllene (7.9%), α-copaene (6.2%) | —                          | [23] |
| *E. protenta* | Algodoal island, Maracanã, Pará, Brazil | Leaf (HD)      | germacrene D (15.6%), δ-cadinene (8.5%), α-cadinol (6.9%), bicyclogermacrene (6.8%), τ-muurolol (5.9%), E-caryophyllene (5.7%) | —                          | [23] |
| *E. punicifolia* | Serra Negra, Pernambuco, Brazil | Leaf (HD)      | linalool (44.0%), E-caryophyllene (22.7%), α-terpineol (8.8%)                          | —                          | [43] |
| *E. punicifolia* | Brejo da Madre de Deus, Pernambuco, Brazil | Leaf (HD)      | linalool (61.2%), E-caryophyllene (16.2%), α-terpineol (6.7%)                          | —                          | [43] |
| *E. punicifolia* | Maracanã, Pará, Brazil            | Leaf (HD)      | E-caryophyllene (9.9%), bicyclogermacrene (8.7%), E-β-ocimene (5.5%), germacrene D (5.4%) | —                          | [25] |
| *E. punicifolia* | Macaé, Rio de Janeiro, Brazil     | Leaf (HD)      | α-cadinol (10.6%), 10-epi-γ-eudesmol (10.2%), paradisiol (9.9%), 7-epi-α-selinene (6.8%) | —                          | [28] |
| *E. pyriformis* | Porto Alegre, Rio Grande do Sul, Brazil | Leaf (HD)      | α-cadinol (14.0%), δ-cadinene (12.4%), τ-cadinol (11.9%), bicyclogermacrene (10.2%)   | —                          | [30] |
| *E. pyriformis* | Curitiba, Paraná, Brazil          | Leaf (HD) b    | β-pinene (0.0-25.7%), limonene (0.2-22.0%), caryophyllene oxide (3.9-21.3%), 1,8-cineole (0.6-14.7%) | —                          | [44] |
| *E. pyriformis* | Curitiba, Paraná, Brazil          | Flower (HD)    | E-caryophyllene (22.8%), germacrene D (15.3%), bicyclogermacrene (8.4%), α-cadinol (7.2%), spathulenol (6.8%) | —                          | [44] |
| *E. pyriformis* | Curitiba, Paraná, Brazil          | Fruits (HD)    | caryophyllene oxide (16.2%), limonene (12.4%), α-terpineol (5.4%), α-cadinol (5.4%)   | —                          | [44] |
Seasonal Variation of Oil Composition

During the annual study, the oxygenated sesquiterpenes increased from January (45.0%) to May (56.2%) [72]. On the other hand, the number of monoterpenes was higher in July, September, and November, mainly for specimens collected in São Paulo state, Brazil, presented spathulenol (7.0–18.0%), Epi-β-pinene (16.0%) as the predominant compounds [70].

It was the principal constituent in January (25.0%), presented meaningful content in other months, but it was absent in October [44]. A specimen collected in Santa Catarina state, Brazil, showed variations. A specimen collected in São Paulo state, Brazil, presented spathulenol (7.0–18.0%), Epi-β-pinene (16.0%) as the predominant compounds [70].

Chemical profiles of oils from Syzygium cumini (L.) Skeels (syn. Eugenia cumini), a species native to Brazil’s occurrence, were collected every three months. The oil was mainly composed of cyclic sesquiterpenes, with a significant content variation on the seasonality study. The monoterpenes amount was higher in the collections made in Minas Gerais state, Brazil, whereas the sesquiterpene content was higher in the collections made in São Paulo state, Brazil. The oxygenated sesquiterpenes presented a significant increase from January (45.0%) to May (56.2%) [72].

The oil composition of leaves of Eugenia spp. was monitored according to their seasonal and circadian variations. A specimen collected in Santa Catarina state, Brazil, showed variations. A specimen collected in São Paulo state, Brazil, presented spathulenol (7.0–18.0%), Epi-β-pinene (16.0%) as the predominant compounds [70].

Some studies have shown variation in the chemical composition and yield of EOs of Eugenia species. The oil composition of leaves of Eugenia spp. was monitored according to their seasonal and circadian variations. A specimen collected in Santa Catarina state, Brazil, showed variations. A specimen collected in São Paulo state, Brazil, presented spathulenol (7.0–18.0%), Epi-β-pinene (16.0%) as the predominant compounds [70].

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Table A1. Cont.

| Species | Occurrence | Essential Oil | Primary Components (>5%) | Essential Oil Bioactivity | Ref. |
|---------|------------|---------------|---------------------------|--------------------------|------|
| E. ramboi | Torres, Rio Gande do Sul, Brazil | Leaf (HD) | β-elemonol (10.6%), bicyclogermacrene (9.7%), E-caryophyllene (6.2%) | — | [27] |
| E. repanda | Porto Rico, Paraná, Brazil | Leaf (HD) | E-caryophyllene (16.3%), α-humulene (10.2%), bicyclogermacrene (9.4%), germacrene D (7.5%), β-elemonol (7.3%) | — | [61] |
| E. selloi | Rio de Janeiro, Rio de Janeiro, Brazil | Leaf (HD) | bicyclogermacrene (15.2-24.3%), germacrene D (5.7-18.7%), E-caryophyllene (6.7-12.5%), viridiflorol (3.5-9.1%) | — | [26] |
| E. speciosa | Itapuã, Rio Grande do Sul, Brazil | Leaf (HD) | α-pinene (47.3%), limonene (23.0%), γ-terpinene (2.5%) | — | [32] |
| E. stictopetala | São Luís, Maranhão, Brazil | Leaf (HD) | γ-elemene (17.48%), E-caryophyllene (16.46%), bicyclogermacrene (8.11%), β-pinene (7.08%), germacrene D (5.64%) | Larvicidal (Aedes aegypti, LC50 230 µg/mL) | [63] |
| E. stigmatosa | Peruibe, São Paulo, Brazil | Leaf (HD) | physeteric acid (90.5%) | — | [62] |
| E. stipitata | São Miguel island, Açores, Portugal | Leaf (HD) | E-caryophyllene (22.7%), caryophyllene oxide (15.4%), α-pinene (14.1%), 4,8-α-epoxy-caryophyllene (10.9%) | Antibacterial, agar diffusion assay (Staphylococcus aureus, 14 mm; Pseudomonas aeruginosa, 11 mm; Listeria monocytogenes, 12 mm) | [77] |
| E. stipitata | Recife, Pernambuco, Brazil | Fruit (SPE) | germacrene D (38.3%), β-pinene (15.2%), α-pinene (10.4%), NI (12.6%) | — | [90] |
| E. stipitata | Florencia-Caquetá, Colombia | Fruit (SD) | limonene (81.0%), myrcene (5.5%) | — | [91] |
| E. sulcata | Restinga de Jurubatiba, Rio de Janeiro, Brazil | Leaf (HD) | E-caryophyllene (24.6%), α-pinene (17.2%), β-pinene (10.9%), 1,8-cineole (5.6%), α-humulene (5.1%) | Enzyme inhibitory (acetylcholinesterase, IC50 4.66 µg/mL); Insecticidal (Dysdercus peruvianus and Oncopeltus fasciatus) | [45, 46] |
| Species          | Occurrence                  | Essential Oil | Primary Components (>5%)                                                                 | Essential Oil Bioactivity                    | Ref. |
|------------------|-----------------------------|---------------|------------------------------------------------------------------------------------------|---------------------------------------------|------|
| *E. sulcata*     | Jureia, São Paulo, Brazil   | Leaf (HD)     | α-pinene (34.2%), *E*-caryophyllene (15.0%), globulol (11.8%), spathulenol (7.0%)          | —                                          | [32] |
| *E. sulcata*     | Restinga de Jurubatiba, Rio de Janeiro, Brazil | Fine stem (HD) | *E*-caryophyllene (18.8%), spathulenol (8.8%), calamenene (7.0%), γ-cadinene (5.8%), α-humulene (5.6%), caryophyllene oxide (5.6%) | —                                          | [46] |
| *E. sulcata*     | Macaé, Rio de Janeiro, Brazil | Leaf (HD)     | 1,8-cineole (19.0%), α-pinene (16.9%), β-pinene (14.5%), caryophyllene oxide (9.8%)       | —                                          | [28] |
| *E. triquetra*   | Pueblo Hondo, Táchira, Venezuela | Leaf (HD)     | linalool (17.5%), limonene (16.9%), α-pinene (11.6%), β-pinene (8.7%)                      | Larvicidal (*Aedes aegypti*, LC$_{50}$ 64.8 ppm) | [65] |
| *E. uruguayensis* | Torres, Rio Grande do Sul, Brazil | Leaf (HD)     | α-pinene (23.5%), β-pinene (11.8%), *E*-caryophyllene (9.5%), caryophyllene oxide (6.4%)   | —                                          | [27] |
| *E. uniflora*    | Pelotas, Rio Grande do Sul, Brazil | Leaf (HD)     | germacrene B (21.2%), selin-1,3,7-(11)-triene-8-one epoxide (19.3%), *E*-caryophyllene (12.6%), germacrene A (11.6%), germacrene D (11.4%), selin-1,3,7-(11)-triene-8-one (9.7%) | Antimicrobial, agar diffusion assay and broth dilution assay (*Staphylococcus aureus*, MIC 800 µg/mL; *Listeria monocytogenes*, MIC 1040 µg/mL; *Candida lipolytica*, MIC 93.7 µg/mL; *C. guilliermondii*, MIC 109.4 µg/mL). Antioxidant (DPPH radical scavenging assay, IC$_{50}$ 833 µg/mL; ABTS IC$_{50}$ 8.1 µg/mL). Hepatoprotective effect in rats | [57], [58] |
| *E. uniflora*    | São Lourenço do Sul, Rio Grande do Sul, Brazil | Leaf (HD)     | NI (38.28%), *E*-caryophyllene (8.18%), spathulenol (7.71%), 9,10-dehydro-iso-longifolene (6.21%), viridiflorol (5.78%), *allo*-aromadendrene (5.66%), dihydro-*cis*-α-copaene-8-ol (5.24%), δ-cadinene (5.19%), τ-cadinol (5.08%) | Antibacterial, broth dilution assay (*Enterobacter aerogenes* MIC 3.125%; *Salmonella typhimurium* (MIC 3.125%)) | [78] |
| Species | Occurrence                      | Essential Oil | Primary Components (>5%) | Essential Oil Bioactivity                                                                 | Ref. |
|---------|--------------------------------|---------------|---------------------------|------------------------------------------------------------------------------------------|------|
| **E. uniflora** | São Luis, Maranhão, Brazil | Leaf (HD)     | curzerene (47.3%), γ-elemene (14.2%), trans-β-elemone (10.4%), β-elemene (5.5%) | Antileishmanial (*L. amazonensis* promastigotes, IC₅₀ 3.04 µg/mL; *L. amazonensis* amastigotas, IC₅₀ 1.92 µg/mL) | [47] |
|         | Osasco, São Paulo, Brazil | Leaf (SD)     | atractilone (26.78%), curzerene (17.96%), germacrene B (9.31%) | Antimicrobial, agar diffusion assay and broth dilution assay (*Streptococcus equi*, MIC 7500 µg/mL; *Staphylococcus epidermis*, MIC 7500 µg/mL; *Candida dubliniensis*, MIC 230 µg/mL; *C. tropicalis*, MIC 900 µg/mL; *C. albicans*, MIC 1800 µg/mL; *C. glabrata*, MIC 930 µg/mL; *C. parapsilosis*, MIC 3750 µg/mL; *C. grubii* serotype A, MIC 450 µg/mL; *C. gattii* serotype C, MIC 1800 µg/mL; *C. gattii* serotype B, MIC 220 µg/mL; *C. neoformans* serotype D, MIC 110 µg/mL; *Saccharomyces cerevisiae*, MIC 220 µg/mL) | [51] |
| **E. uniflora** | Águas de Santa Bárbara, São Paulo, Brazil | Leaf (SD)     | selina-1,3,7-(11)-trien-8-one (34.0%), selina-1,3,7-(11)-trien-8-one epoxide (17.0%), germacrene B (10.5%), curzerene (5.5%), E-caryophyllene (5.0%) | — | --- |
| **E. uniflora** | Botucatu, São Paulo, Brazil | Leaf (HD)     | selina-1,3,7-(11)-trien-8-one (30.1%), selina-1,3,7-(11)-trien-8-one epoxide (21.89%), E-caryophyllene (6.1%) | Antibacterial, broth dilution assay (*Staphylococcus aureus*, MIC 55,600.0 µg/mL; *Staphylococcus aureus* – meticillin resistant, MIC 56,000.0 µg/mL; *Staphylococcus aureus* – meticillin sensitive, MIC 50,800.0 µg/mL; *Salmonella spp.*, MIC 92,400.0 µg/mL; *Salmonella enteritidis*, MIC 92,400.0 µg/mL; *Salmonella typhimurium*, MIC 92,400.0 µg/mL; *Escherichia coli*, MIC 84,300.0 µg/mL; *Pseudomonas aeruginosa*, MIC 92,400.0 µg/mL) | [76] |
| **E. uniflora** | Seropédica, Rio de Janeiro, Brazil | Young leaf (HD) | germacrone (37.86%), curzerene (16.6%), germacrene B (13.59%), E-caryophyllene (6.02%) | — | [92] |
| **E. uniflora** | Seropédica, Rio de Janeiro, Brazil | Old leaf (HD)  | curzerene (22.37%), furanodiene (18.99%), germacrene B (14.39%), E-caryophyllene (9.35%) | — | [92] |
Table A1. Cont.

| Species        | Occurrence          | Essential Oil        | Primary Components (>5%)                                                                 | Essential Oil Bioactivity                                                                 | Ref. |
|----------------|---------------------|----------------------|-----------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------|------|
| *E. uniflora*  | Anápolis, Goiás, Brazil | Aerial parts (HD) b   | selina-1,3,7-(11)-tri-en-8-one (43.0%), selina-1,3,7-(11)-tri-en-8-one epoxide (20.0-29.0%), spathulenol (7.5-10.0%) | — | [54] |
| *E. uniflora*  | Belém, Pará, Brazil | Aerial parts (HD)     | germacrene (32.8%), curzerene (30.0%), germacrene B (15.6%) | Antioxidant, DPPH radical scavenging assay (30.3%, 153.5 mg.ET/mL); Antioxidant, β-carotene/linoleic acid assay at 1 mg/mL (10.8%) | [55] |
| *E. uniflora*  | Santarém, Pará, Brazil | Leaf (HD)             | selina-1,3,7(11)-tri-en-8-one (32.6%), selina-1,3,7(11)-tri-en-8-one epoxide (30.4%), E-caryophyllene (5.4%), germacrene B (5.0%) | Antioxidant, DPPH radical scavenging assay (35.3%, 178.8 mg.ET/mL); Antioxidant, β-carotene/linoleic acid assay at 1 mg/mL (23.0%); Cytotoxic (HCT-116 human colon carcinoma, IC₅₀ > 25 µg/mL; MRC5 human fibroblast IC₅₀ > 25 µg/mL; AGP-01 human gastric adenocarcinoma, IC₅₀ > 25 µg/mL; SKMEL-19 human melanoma, IC₅₀ > 25 µg/mL) | [48] |
| *E. uniflora*  | Belém, Pará, Brazil | Leaf (HD)             | selina-1,3,7(11)-tri-en-8-one (43.1%), selina-1,3,7(11)-tri-en-8-one epoxide (21.7%), germacrene B (5.9%) | Antioxidant, DPPH assay (45.1%, 228.3 mg.ET/mL); Antioxidant, β-carotene/linoleic acid assay at 1 mg/mL (16.8%); Cytotoxic (HCT-116 human colon carcinoma, IC₅₀ > 25 µg/mL; MRC5 human fibroblast IC₅₀ > 25 µg/mL; AGP-01 human gastric adenocarcinoma, IC₅₀ > 25 µg/mL; SKMEL-19 human melanoma, IC₅₀ > 25 µg/mL) | [48] |
| *E. uniflora*  | Bahia, Brazil       | Leaf (SPME)           | germacrene B (9.5%), γ-elemene (8.5%), β-elemene (7.4%), E-caryophyllene (7.1%), germacrene D (5.5%), γ-muurolene (5.3%), germacrene (5.1%) | — | [58] |
| Species       | Occurrence               | Essential Oil  | Primary Components (>5%)                                                                 | Essential Oil Bioactivity                                                                                   | Ref. |
|--------------|--------------------------|----------------|------------------------------------------------------------------------------------------|-----------------------------------------------------------------------------------------------------------|------|
| **E. uniflora** | Belém, Pará, Brazil      | Leaf (HD)      | curzerene (50.6%), germacrene B (5.3%)                                                  | Antioxidant, DPPH assay (42.8%, 217.0 mg.ET/mL); Antioxidant, β-carotene/linoleic acid assay at 1mg/mL (20.1%) | [48] |
|              |                          |                | curzerene (34.4-53.1%), germacrone (0.2–10.5%), globulol (1.5–7.4%), germacrene B (0.1–7.5%), spathulenol (0.5–7.0%), viridiflorol (0.8–6.2%), β-eleme (1.8–5.8%) | Antioxidant, DPPH assay (42.6-64.2%, 186.9-400.3 mg.ET/g)                                                | [73] |
|              | Belém, Pará, Brazil      | Leaf (HD)      | germacrone B (18.4%), curzerene (13.4%), E-caryophyllene (9.1%), γ-eleme (7.8%), β-eleme (7.4%) | Antioxidant, DPPH assay (40.6%, 205.6 mg.ET/mL); Antioxidant β-carotene/linoleic acid (26.3%); Cytotoxic (HCT-116 human colon carcinoma, IC50 9.28 µg/mL; MRC5 human fibroblast IC50 14.95 µg/mL; AGP-01 human gastric adenocarcinoma, IC50 8.73 µg/mL; SKMEL-19 human melanoma, IC50 15.42 µg/mL) | [48] |
|              | Bahia, Brazil            | Leaf (SPME)    | germacrone B (9.0%), γ-eleme (8.1%), γ-muurolone (7.7%), germacrene D (6.1%), β-eleme (5.0%) | —                                                                                                           | [58] |
|              | Bahia, Brazil            | Leaf (SPME)    | germacrone B (9.6%), β-eleme (9.3%), γ-eleme (8.0%), germacrene D (6.5%), E-caryophyllene (5.2%), γ-muurolone (5.0%) | —                                                                                                           | [58] |
|              | Rio de Janeiro, Brazil   | Leaf (HD)      | atractilone, 3-furanoeudesmenes                                                          | Antinociceptive (ED50 218.6 mg/kg) and hypothermic in mouse model                                           | [52] |
|              | Rio de Janeiro, Brazil   | Leaf (HD)      | curzerene (50.2%), β-eleme (5.9%)                                                        | —                                                                                                           | [50] |
|              | Goiás, Brazil            | Leaf (HD)      | germacrone B (21.6%), curzerene (20.5%), germacrone (17.3%), E-caryophyllene (8.7%)    | Antifungal, broth dilution assay (Paracoccidioides brasiliensis, MIC 500 µg/mL; MFC 500 µg/mL)               | [49] |
| Species        | Occurrence                  | Essential Oil | Primary Components (>5%)                                                                 | Essential Oil Bioactivity                                                                 | Ref.          |
|----------------|-----------------------------|---------------|------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------|--------------|
| *E. uniflora*  | Goiás, Brazil               | Leaf (HD)     | curzerene (42.6%), germacrone (13.5%), germacrone D (8.8%), germacrene A (7.4%), E-caryophyllene (7.0%) | Antifungal, broth dilution assay (*Paracoccidioides brasiliensis*, MIC 62.5 µg/mL; MFC 125 µg/mL) | [49]         |
| *E. uniflora*  | Goiás, Brazil               | Leaf (HD)     | selin-1,3,7-(11)-trien-8-one (48.2%), selin-1,3,7-(11)-trien-8-one epoxide (19.3%), atractilone (11.7%), germacrene B (5.9%) | Antifungal, broth dilution assay (*Paracoccidioides brasiliensis*, MIC 250 µg/mL; MFC 250 µg/mL) | [49]         |
| *E. uniflora*  | Pelotas, Rio Grande do Sul, Brazil | Fruits (HD) | hexadecanoic acid (11.7%), E-β-ocimene (7.4%), α-selinene (7.2%), germacrene B (7.2%), bicyclogermacrene (5.2%) | Antioxidant, DPPH assay (TLC method)                                                      | [40]         |
| *E. verticillata* | Caraguatatuba, São Paulo, Brazil | Leaf (HD)     | valerianol (28.1%), 10-epi-γ-eudesmol (12.6%), E-caryophyllene (10.9%), α-selinene (6.1%), δ-cadinene (5.7%) | Enzyme inhibitory (acetylcholinesterase, IC₅₀ 67.3 µg/mL)                               | [64]         |
| *E. xiriricana* | Peruibe, São Paulo, Brazil  | Leaf (HD)     | spathulenol (15.4%), β-pinene (14.1%), globulol (8.6%), α-pinene (7.8%)                    | —                                                                                         | [32]         |
| *S. cumini*    | Rio de Janeiro, Rio de Janeiro, Brazil | Leaf (HD)     | α-pinene (22.2%), Z-β-ocimene (10.2%), E-caryophyllene (9.45%), limonene (7.3%), α-terpineol (7.00%), E-β-ocimene (5.88%), α-humulene (5.5%) | Anti-inflammatory, lipopolysaccharide-induced pleurisy model, an eosinophil was inhibited in 67% | [66]         |
| *S. cumini*    | São Luís, Maranhão, Brazil  | Leaf (HD)     | α-pinene (31.85%), Z-β-ocimene (28.98%), E-β-ocimene (11.71%), β-pinene (5.57%), E-caryophyllene (5.02%) | Antileishmanial (*Leishmania amazonensis* promastigotes, IC₅₀ 60 µg/mL); Molluscicide (*Biomphalaria glabrata*, LC₅₀ 90 µg/mL) | [67]         |
| *S. cumini*    | Crato, Ceará, Brazil        | Leaf (HD)     | α-pinene (48.1%), E-nerolidol (8.7%), nerol (7.1%), nonanol (6.8%) a                       | Antibacterial, broth dilution assay (*Staphylococcus aureus*, MIC 128 µg/mL; *Escherichia coli* MIC > 1024 µg/mL) | [68]         |
| *S. cumini*    | Crato, Ceará, Brazil        | Leaf (HD)     | α-pinene (30.0%), E-β-ocimene (26.8%), β-ocimene (11.1%), β-pinene (8.3%), limonene (6.5%), β-Fenchol (7.3%) a | —                                                                                         | [69]         |
| Species   | Occurrence                        | Essential Oil | Primary Components (>5%)                                                                 | Essential Oil Bioactivity                              | Ref. |
|-----------|-----------------------------------|---------------|------------------------------------------------------------------------------------------|--------------------------------------------------------|------|
| *S. cumini* | Juiz de Fora, Minas Gerais, Brazil | Leaf (HD)     | α-humulene (25.24%), *E*-caryophyllene (16.00%), α-terpineol (9.08%), globulol (5.23%) | Anti-inflammatory; Antibacterial (*Mycobacterium bovis*) | [70] |
| *S. cumini* | Porto Rico, Paraná, Brazil        | Seed (HD) b   | *E*-caryophyllene (6.4-42.5%), α-humulene (6.6-22.2%), caryophyllene oxide (5.3-37.3%), humulene epoxide II (2.1-17.1%) | —                                                       | [93] |
| *S. jambos* | Rio Verde, Goiás, Brazil          | Leaf (HD) b   | *E*-caryophyllene (0–10.86%), α-humulene (0.0-7.07%), α-zingiberene (0.97–17.73%), hydroxytoluenebuthyled (3.44–32.82%), caryophyllenyl alcohol (1.48–17.14%), caryolan-8-ol (0–10.75%), caryophyllene oxide (0–5.05%), tujopsan-2-α-ol (1.01–12.19%), heneicosane (1.73–18.0%) a | —                                                       | [74] |

* = The analysis is doubtful, based on known retention indices (RI), b = seasonal study, SD = steam distillation, HD = hydrodistillation, SPME = solid phase microextraction, NI = unidentified compound, IC50 = median inhibitory concentration, MIC = minimum inhibitory concentration, MMC = minimum microbicide concentration, LC50 = median lethal concentration.
Appendix B

Figure A1. Major monoterpene constituents found in Eugenia and Syzygium essentials oils.

Figure A2. Major sesquiterpene hydrocarbons found in Eugenia and Syzygium essentials oils.

Figure A2. Cont.
Figure A2. Major sesquiterpene hydrocarbons found in Eugenia and Syzygium essentials oils.

Figure A3. Cont.
Figure A3. Major oxygenated sesquiterpenes found in Eugenia and Syzygium essentials oils.
Figure A3. Major oxygenated sesquiterpenes found in Eugenia and Syzygium essentials oils.

Figure A4. Other major constituents found in Eugenia and Syzygium essentials oils.
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