Basic Study

Effect of prolonged omeprazole administration on segmental intestinal Mg\(^{2+}\) absorption in male Sprague-Dawley rats

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Abstract

BACKGROUND

The exact mechanism of proton pump inhibitors (PPIs)-induced hypomagnesemia (PPIH) is largely unknown. Previous studies proposed that PPIH is a consequence of intestinal Mg\(^{2+}\) malabsorption. However, the mechanism of PPIs-suppressed intestinal Mg\(^{2+}\) absorption is under debate.

AIM

To investigate the effect of 12-wk and 24-wk omeprazole injection on the total, transcellular, and paracellular Mg\(^{2+}\) absorption in the duodenum, jejunum, ileum, and colon of male Sprague-Dawley rats.

METHODS

The rats received 20 mg/kg.d subcutaneous omeprazole injection for 12 or 24 wk. Plasma and urinary Mg\(^{2+}\), Ca\(^{2+}\), and PO\(_4^{3-}\) levels were measured. The plasma concentrations of 1α,25-dihydroxyvitamin D\(_3\) (1α,25(OH)\(_2\)D\(_3\)), parathyroid hormone (PTH), fibroblast growth factor 23 (FGF-23), epidural growth factor (EGF), and insulin were also observed. The duodenum, jejunum, ileum, and colon of each rat were mounted onto individual modified Using chamber setups to study the rates of total, transcellular, and paracellular Mg\(^{2+}\) absorption simultaneously. The expression of transient receptor potential melastatin 6 (TRPM6) and cyclin M4 (CNNM4) in the entire intestinal tract was also measured.

RESULTS

Single-dose omeprazole injection significantly increased the intraluminal pH of the stomach, duodenum, and jejunum. Omeprazole injection for 12 and 24 wk induced hypomagnesemia with reduced urinary Mg\(^{2+}\) excretion. The plasma Ca\(^{2+}\)
was normal but the urinary Ca\(^{2+}\) excretion was reduced in rats with PPIH. The plasma and urinary PO\(_{4}\)\(^{3-}\) levels increased in PPIH rats. The levels of 1α,25(OH)\(_{2}\)D\(_{3}\), and FGF-23 increased, whereas that of plasma EGF decreased in the omeprazole-treated rats. The rates of the total, transcellular, and paracellular Mg\(^{2+}\) absorption was significantly lower in the duodenum, jejunum, ileum, and colon of the rats with PPIH than in those of the control rats. The percent suppression of Mg\(^{2+}\) absorption in the duodenum, jejunum, ileum, and colon of the rats with PPIH compared with the control rats was 81.86%, 70.59%, 69.45%, and 39.25%, respectively. Compared with the control rats, the rats with PPIH had significantly higher TRPM6 and CNNM4 expression levels throughout the intestinal tract.

**CONCLUSION**

Intestinal Mg\(^{2+}\) malabsorption was observed throughout the intestinal tract of rats with PPIH. PPIs mainly suppressed small intestinal Mg\(^{2+}\) absorption. Omeprazole exerted no effect on the intraluminal acidic pH in the colon. Thus, the lowest percent suppression of total Mg\(^{2+}\) absorption was found in the colon. The expression levels of TRPM6 and CNNM4 increased, indicating the presence of a compensatory response to Mg\(^{2+}\) malabsorption in rats with PPIH. Therefore, the small intestine is an appropriate segment that should be modulated to counteract PPIH.

**Key words:** Adverse effect; Colon; Mg\(^{2+}\) absorption; Proton pump inhibitors-induced hypomagnesemia; Small intestine

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**INTRODUCTION**

Mg\(^{2+}\) is an essential ion that mediates several physiological functions in the brain, lung, heart and vessel, pancreas and liver, muscle, and bone\(^{1,2}\). Mg\(^{2+}\) disturbance has been implicated in the pathophysiological mechanisms of several diseases in those organs; therefore, Mg\(^{2+}\) supplement is a potential therapeutic regime in many of these diseases\(^{3,4}\). Over 99% of total body Mg\(^{2+}\) is stored in the bone, muscle, and soft tissues\(^{5,6}\). The remaining 1% is found in the plasma, which is tightly regulated by intestinal absorption and renal excretion\(^{7,8}\). Intestinal epithelium absorbs Mg\(^{2+}\) through paracellular passive and transcellular active mechanisms\(^{9}\). Transcellular Mg\(^{2+}\) absorption requires the activities of luminal transient receptor potential melastatin 6 (TRPM6) and basolateral cyclin M4 (CNNM4), whereas paracellular absorption is regulated by tight junction-associated claudin proteins. However, the study of transepithelial Mg\(^{2+}\) transport is limited by the poor understanding on the mechanism of intestinal Mg\(^{2+}\) absorption\(^{9}\). Previous studies proposed that paracellular and transcellular Mg\(^{2+}\) absorption exclusively proceeds in the small and large intestines, respectively\(^{10,11}\). In addition, the expression of TRPM6 mRNA is...
barely detected in mouse duodenum\textsuperscript{[4]}. Conversely, another report demonstrated paracellular and transcellular Mg\textsuperscript{2+} absorption in the duodenum\textsuperscript{[4]}. Immunofluorescence imaging clearly showed the expression and localization of TRPM6 protein in the brush-border villi of the duodenum\textsuperscript{[4]}. CNNM4 protein was also identified in small intestinal epithelium\textsuperscript{[4]}. Therefore, the simultaneous study on the transcellular and paracellular Mg\textsuperscript{2+} absorption in the duodenum, jejunum, ileum, and colon will provide data on the mechanism of intestinal Mg\textsuperscript{2+} absorption.

Proton pump inhibitor (PPI)-induced hypomagnesemia (PPIH) has been reported since 2006\textsuperscript{[5-9]}. Approximately 61\% and 29\% of patients with PPIH have PPIs prescription for at least 5 and 10 years, respectively\textsuperscript{[10]}. However, PPIH was also diagnosed in patients who used PPIs for 2 months\textsuperscript{[11]}. Clinical observations suggested that PPIH is a consequence of intestinal Mg\textsuperscript{2+} malabsorption\textsuperscript{[12,13]}, the mechanism of which is currently under debate. Previous studies proposed that PPIs suppress colonic Mg\textsuperscript{2+} absorption in normal and PPIH mice\textsuperscript{[13]}. However, the effect of PPIs administration on colonic Mg\textsuperscript{2+} absorption had not been investigated\textsuperscript{[14,15]}. Moreover, the stimulation of colonic Mg\textsuperscript{2+} absorption by using inulin fibers could not normalize plasma Mg\textsuperscript{2+} level in those PPIH mice\textsuperscript{[11]}. A recent study has reported that omeprazole injection for 24 wk induces systemic Mg\textsuperscript{2+} deficiency and hypomagnesemia in male Sprague-Dawley rats\textsuperscript{[16]}. The rate of transcellular and paracellular duodenal Mg\textsuperscript{2+} absorption is suppressed in rats with PPIH\textsuperscript{[13]}. However, the effect of prolonged PPIs administration on jejunal, ileal, and colonic Mg\textsuperscript{2+} absorption in a PPIH model remains unknown.

The present study aimed to observe the paracellular and transcellular Mg\textsuperscript{2+} transport across the duodenum, jejunum, ileum, and colon in control and prolonged omeprazole-treated male Sprague-Dawley rats. The expression of TRPM6 and CNNM4 proteins in each intestinal segment was also studied. Plasma and urinary Mg\textsuperscript{2+}, Ca\textsuperscript{2+}, and PO\textsubscript{4}\textsuperscript{3−} levels were measured. The plasma concentrations of hormones that modulate Mg\textsuperscript{2+} homeostasis, such as 1α,25-dihydroxyvitamin D\textsubscript{3} (1α,25(OH)\textsubscript{2}D\textsubscript{3}), parathyroid hormone (PTH), fibroblast growth factor 23 (FGF-23), epidermal growth factor (EGF), and insulin\textsuperscript{[3,18-21]}, were also determined. Given that 16.7 rat days are equivalent to 1 human year\textsuperscript{[22]}, the animals were treated with 20 mg/kg omeprazole daily for 12 and 24 wk which is comparable to 5 and 10 human years, respectively.

**MATERIALS AND METHODS**

**Animals**

This study was performed in strict compliance with the Animal for Scientific Purposes Act of Thailand and in accordance with Ethical Principles and Guidelines for the Use of Animals for Scientific Purposes, National Research Council of Thailand. All experimental procedures were approved by the Ethics Committee on Animal Experiment of Burapha University, Thailand. Male Sprague-Dawley rats (9 weeks old) were randomly divided into three experimental groups: control, 12-wk omeprazole, and 24-wk omeprazole treatments. The rats were acclimatized for 7 days and fed with standard pellet chow and reverse osmosis water given ad libitum. The health, body weight, and food intake were monitored and recorded daily.

The effect of single-dose subcutaneous omeprazole injection (20 mg/kg; Ocid\textsuperscript{®} IV; Zydus Cadila, India) on intraluminal gastrointestinal pH was observed. The pellet chow was removed 4 h before and then retrieved 30 min after sham or omeprazole injection. At 2 and 24 h after administration, the stomach, duodenum, jejunum, ileum, cecum, and proximal colon were removed under thiopental anesthesia (70 mg/kg; Anesthal, Jagsonpal Pharmaceuticals Ltd., India). The intraluminal pH levels of the stomach, duodenum, jejunum, ileum, cecum, and proximal colon were measured by diagnostic test strips (MCOLORFAST\textsuperscript{TM} pH-Indicator Strips, Merck-Millipore, German).

To study the effect of prolonging omeprazole injection, control and 24-wk omeprazole-treated rats received subcutaneous sham or omeprazole (20 mg/kg) injection daily for 24 wk. The rats in the 12-wk-omeprazole-treated group received sham injection daily for 12 wk followed by subcutaneous omeprazole injection for 12 wk. At 24 h prior to the experimental endpoint, the rats were housed in a metabolic cage to collect food and water intake. Urinary and fecal output was also collected. The rats were anesthetized, blood was collected from the left ventricle, and the rats were subsequently sacrificed. The duodenum, jejunum, ileum, and colon were rapidly collected. The plasma concentrations of 1α,25(OH)\textsubscript{2}D\textsubscript{3}, PTH, FGF-23, EGF, and insulin were determined by Labhouse Chonburi Co. Ltd., Thailand. Plasma and urine Mg\textsuperscript{2+}, Ca\textsuperscript{2+}, and PO\textsubscript{4}\textsuperscript{3−} concentrations were also measured.
Magnesium flux measurement

The total, paracellular, and transcellular Mg\(^{2+}\) flux was studied in accordance with the method described by Thongon et al.\(^{[5]}\). In brief, the duodenum, jejunum, ileum, and colon from individual rats were dissected into two pieces, which then were rapidly mounted onto individually modified Ussing chamber setups. Intestinal samples were bathed and equilibrated for 10 min with physiological bathing solution containing (in mmol/L) 118 NaCl, 4.7 KCl, 1.25 CaCl\(_2\), 23 NaHCO\(_3\), 12 D-glucose, 2.5 L-glutamine, and 2 D-mannitol (osmolality of 290-295 mmol/kg H\(_2\)O and pH of 7.4). The solution was maintained at 37 °C and continuously gassed with 5% CO\(_2\) in 95% O\(_2\). The first piece of each intestinal segment was subjected to study the rate of total Mg\(^{2+}\) transport. The apical solution of each Ussing chamber setup was substituted with a Mg\(^{2+}\)-containing bathing solution supplemented with (in mmol/L) 40 MgCl\(_2\), 2.5 CaCl\(_2\), 4.5 KCl, 12 D-glucose, 2.5 L-glutamine, 115 mannitol, and 10 HEPES (pH 7.4). Meanwhile, the basolateral solution was substituted with a Mg\(^{2+}\)-free bathing solution containing (in mmol/L) 1.25 CaCl\(_2\), 4.5 KCl, 12 D-glucose, 2.5 L-glutamine, 250 D-mannitol, and 10 HEPES pH 7.4. To study the rate of paracellular Mg\(^{2+}\) transport, another piece of each intestinal segment was incubated with Mg\(^{2+}\) channel blocker Co(III)hexaaamine (1 mmol/L; Sigma, St. Louis, MO, United States)\(^{[23]}\) to inhibit transcellular Mg\(^{2+}\) flux. The Mg\(^{2+}\)-containing and Mg\(^{2+}\)-free bathing solutions were also used to study paracellular Mg\(^{2+}\) transport. At 30, 60, and 120 min after solution replacements, a 100 µL solution was collected from the basolateral and apical sides. The Mg\(^{2+}\) concentration and Mg\(^{2+}\) flux rates were determined as previously described by Thongon and Krishnamra\(^{[23]}\). The rate of transcellular Mg\(^{2+}\) transport was calculated by subtracting the rate of total Mg\(^{2+}\) transport with the rate of paracellular Mg\(^{2+}\) transport from the same intestinal segment of the individual rats.

Western blot analysis

Western blot analysis was performed as previously described\(^{[5]}\). Epithelial cells of the duodenum, jejunum, ileum, and colon were collected and lysed in Pierce\(^{®}\) Ripa Buffer (Thermo Fisher Scientific Inc.) with 10% v/v protease inhibitor cocktail (Sigma). After being sonicated, centrifuged, and heated, 50 µg samples were loaded and separated on SDS-PAGE gel and then transferred to a polyvinylidene difluoride membrane. The membrane was blocked and probed with 1:1000 primary antibodies (Santa Cruz Biotechnology) raised against CNNM4 and TRPM6. The membranes were also reprobed with 1:5000 anti-β-actin monoclonal antibodies (Santa Cruz Biotechnology). The membranes were then washed with Tris-buffered saline (TBS) containing 0.1% Tween-20 (Thermo Fisher Scientific Inc.) with 10% v/v protease inhibitor cocktail (Sigma). After rinsing, the membranes were probed with 1:10000 HRP-conjugated secondary antibodies (Santa Cruz Biotechnology), visualized by Thermo Scientific SuperSignal\(^{®}\) West Pico Substrate (Thermo Fisher Scientific Inc.), and captured on CL-XPosure Film (Thermo Fisher Scientific Inc.). Densitometric analysis was performed using ImageJ for Mac Os X.

Statistical analysis

Results were expressed as mean ± SE. Two sets of data were compared using the unpaired Student’s t-test. One-way ANOVA with Dunnett’s post-test was used for comparison of multiple sets of data. All data were analyzed by GraphPad Prism for Mac Os (GraphPad Software Inc., San Diego, CA, United States).

RESULTS

Single-dose omeprazole injection increased gastrointestinal pH

Our previous study showed that 20 mg/kg oral gavage and subcutaneous omeprazole administration markedly suppress gastric acid secretion\(^{[9]}\). In the present study, the rats were subjected to prolonged omeprazole administration (24 wk). Therefore, subcutaneous administration that often causes minimal pain or discomfort was chosen. The injection site was changed to avoid the repeated damage or irritation of the rat tissue.

These experiments were performed to observe the effect of single-dose 20 mg/kg subcutaneous omeprazole injection on gastric acid secretion and gastrointestinal pH. Intraluminal pH of the stomach, duodenum, jejunum, ileum, cecum, and proximal colon were measured at 2 and 24 h after injection. In the vehicle-treated control group, the intraluminal pH levels of the stomach, duodenum, jejunum, ileum, cecum, and colon were 1.83 ± 0.31, 5.83 ± 0.30, 6.33 ± 0.21, 7.67 ± 0.42, 6.50 ± 0.22, and 6.67 ± 0.31, respectively. Omeprazole significantly increased gastric, duodenal, and jejunal pH at 24 h after injection (Figure 1). However, omeprazole did not affect the intraluminal acidic environment of the cecum and colon. These results indicated that 20 mg/kg omeprazole injection daily effectively suppressed the gastric acid secretion and...
increased the intraluminal pH of the stomach, duodenum, and jejunum through 24 wk of treatment.

**Metabolic characteristic of omeprazole-treated rats**

As demonstrated in Figure 2A, all rats showed equal growth after the 24 wk of the experiment (Figure 2A). Food intake (Figure 2B) and fecal excretion (Figure 2D) of all experimental groups were equal. Water intake (Figure 2C) and urine excretion (Figure 2D) significantly increased in the omeprazole-treated groups.

**Omeprazole-induced hypomagnesemia**

The 12- and 24-wk-omeprazole-treated rats had significantly reduced plasma (Figure 3A) and urinary Mg\(^{2+}\) concentration (Figure 3D). The plasma concentrations of the 12- and 24-omeprazole-treated groups were 1.41 ± 0.08 mg/dL and 1.37 ± 0.14 mg/dL respectively, which were lower than the reference range of plasma Mg\(^{2+}\) concentration (1.7-2.4 mg/dL). Therefore, omeprazole induced hypomagnesemia in our rat model. In addition, the urinary Mg\(^{2+}\) concentrations of the 12- and 24-omeprazole-treated groups were 1.26 ± 0.72 mg/dL and 1.48 ± 0.52 mg/dL, respectively, which were also lower than the normal reference of 1.7-3.0 mg/dL. While the plasma Ca\(^{2+}\) concentration (Figure 3B) did not change, the urinary Ca\(^{2+}\) concentrations of the 12- (1.63 ± 0.28 mg/dL) and 24-wk-omeprazole-treated (1.42 ± 0.23 mg/dL) groups were significantly lower than those of the control group (4.06 ± 0.87 mg/dL) (Figure 3E). The plasma (Figure 3C) and urinary phosphate concentrations (Figure 3F) of the 12- and 24-omeprazole-treated groups significantly increased in comparison to its corresponding control group.

**Hormonal change in PPIH rats**

In consideration that 1α,25(OH)\(_2\)D\(_3\), PTH, FGF-23, EGF, and insulin modulate Mg\(^{2+}\) homeostasis\(^{[3,18-21]}\), their plasma concentrations in the rats with PPIH were determined. The plasma 1α,25(OH)\(_2\)D\(_3\), PTH, FGF-23 (Figure 4A) and FGF-23 (Figure 4C) concentrations of the 24-wk-omeprazole-treated rats significantly increased compared with those of the control group. The plasma PTH (Figure 4B) and insulin (Figure 4E) of all experimental groups showed no difference. The 12- and 24-wk-omeprazole-treated groups significantly increased compared with those of the control group. The plasma 1α,25(OH)\(_2\)D\(_3\), PTH, FGF-23, EGF, and insulin modulate Mg\(^{2+}\) homeostasis\(^{[3,18-21]}\).

**Segmental intestinal Mg\(^{2+}\) transport in PPIH rats**

Previous research proposed that paracellular passive and transcellular active Mg\(^{2+}\) transport exclusively occur in the small and large intestines, respectively. In the present study, the total, paracellular, and transcellular Mg\(^{2+}\) transport rates of the vehicle-treated control group were 27.68 ± 1.36 nmol/h/cm\(^2\), 23.04 ± 1.19 nmol/h/cm\(^2\), and 4.65 ± 0.59 nmol/h/cm\(^2\), respectively, in the duodenum (Figure 5A); 31.00 ± 1.19 nmol/h/cm\(^2\), 23.73 ± 1.22 nmol/h/cm\(^2\), and 7.27 ± 0.81 nmol/h/cm\(^2\), respectively, in the jejunum (Figure 5B); 30.77 ± 0.94 nmol/h/cm\(^2\), 22.23 ± 0.88 nmol/h/cm\(^2\), and 8.53 ± 0.58 nmol/h/cm\(^2\), respectively, in the ileum (Figure 5C); and 19.77 ± 0.99 nmol/h/cm\(^2\), 9.83 ± 0.51 nmol/h/cm\(^2\), and 9.93 ± 0.52 nmol/h/cm\(^2\), respectively, in the colon (Figure 5D). In the 12-wk-omeprazole-treated rats, the total, paracellular, and transcellular Mg\(^{2+}\) transport rates were 8.55 ± 1.27 nmol/h/cm\(^2\), 5.78 ± 1.03 nmol/h/cm\(^2\), and 2.77 ± 0.53 nmol/h/cm\(^2\), respectively, in the duodenum (Figure 5A); 12.36 ± 0.79 nmol/h/cm\(^2\), 8.47 ± 0.57 nmol/h/cm\(^2\), and 3.88 ± 0.42 nmol/h/cm\(^2\), respectively, in the jejunum (Figure 5B); 12.39 ± 0.76 nmol/h/cm\(^2\), 9.01 ± 0.45 nmol/h/cm\(^2\), and 3.39 ± 0.44 nmol/h/cm\(^2\), respectively, in the ileum (Figure 5C); and 15.41 ± 0.90 nmol/h/cm\(^2\), 7.39 ± 0.33 nmol/h/cm\(^2\), and 2.65 ± 0.36 nmol/h/cm\(^2\), respectively, in the colon (Figure 5D). In the 24-wk-omeprazole-treated rats, the total, paracellular, and transcellular Mg\(^{2+}\) transport rates were 5.02 ± 0.51 nmol/h/cm\(^2\), 3.59 ± 0.59 nmol/h/cm\(^2\), and 1.43 ± 0.29 nmol/h/cm\(^2\), respectively, in the duodenum (Figure 5A); 9.13 ± 0.75 nmol/h/cm\(^2\), 6.48 ± 0.45 nmol/h/cm\(^2\), and 2.65 ± 0.36 nmol/h/cm\(^2\), respectively, in the jejunum (Figure 5B); 9.40 ± 0.40 nmol/h/cm\(^2\), 7.43 ± 0.21 nmol/h/cm\(^2\), and 1.97 ± 0.34 nmol/h/cm\(^2\), respectively, in the ileum (Figure 5C); and 12.01 ± 0.56 nmol/h/cm\(^2\), 6.29 ± 0.36 nmol/h/cm\(^2\), and 5.71 ± 0.21 nmol/h/cm\(^2\), respectively, in the colon (Figure 5D). In the small intestinal segment, Mg\(^{2+}\) was absorbed mainly through the paracellular route (Figure 5A-C and E). By contrast, the large intestine absorbed Mg\(^{2+}\) through the paracellular and transcellular routes in a comparable amount (Figure 5D). The rate of transcellular Mg\(^{2+}\) transport was the highest in the colon and the lowest in the duodenum (Figure 5F). In the 12- and 24-wk-omeprazole-treated rats, the total, paracellular, and transcellular Mg\(^{2+}\) transport rates were significantly lower in the duodenum, jejunum, ileum, and colon than in those of the corresponding vehicle-treated control rats (Figure 5A-D). These results suggest that prolonged omeprazole injection suppresses Mg\(^{2+}\) absorption throughout the intestinal tract.
**Figure 1** Effect of subcutaneous omeprazole injection on rat gastrointestinal pH. Intraluminal pH was measured by using test strips after 2 or 24 h after omeprazole administration. *a* $P < 0.05$, *b* $P < 0.01$, *c* $P < 0.001$, vs the control group ($n = 6$).

**Segmental intestinal TRPM6 and CNNM4 expression in PPIH rats**

Regarding our recent results on Mg$^{2+}$ transport throughout the intestinal tract, these series of experiment aimed to study the expression of TRPM6 and CNNM4 in the duodenum, jejunum, ileum, and colon. As demonstrated in Figure 6, TRPM6 protein was detected in the duodenum, jejunum, ileum, and colon of all experimental groups. The level of TRPM6 expression was the lowest in the duodenum and the highest in the colon of the control (Figure 6A), 12-wk- (Figure 6B), and 24-wk-omeprazole (Figure 6C)-treated rats. The 12- and 24-wk-omeprazole-treated groups had significantly higher TRPM6 expression in the duodenum, jejunum, ileum, and colon than the vehicle-treated control group (Figure 7). Similar to TRPM6, CNNM4 protein was detected throughout the intestinal tract (Figure 8). The expression of CNNM4 protein significantly increased in the duodenum, jejunum, ileum, and colon of the 12- and 24-wk-omeprazole-treated groups compared with those of the control group.

**DISCUSSION**

The present study showed paracellular and transcellular Mg$^{2+}$ transport in the duodenum, jejunum, ileum, and colon. The rate of total Mg$^{2+}$ transport was shown (in order of highest to lowest) in the jejunum, ileum, duodenum, and colon. Small intestinal epithelium absorbed Mg$^{2+}$ mainly through the paracellular route. A comparable rate of paracellular and transcellular Mg$^{2+}$ transport was shown in the colon.

The mechanism by which prolonged PPIs administration induce hypomagnesemia is currently unclear. In the present study, 12- and 24-wk omeprazole injection induced systemic Mg$^{2+}$ depletion and hypomagnesemia. Similar to previous reports in PPIH patients[8,9,12-16], urinalysis of a recent PPIH rat model revealed reduced urinary Mg$^{2+}$ excretion (less than 8.5 mg/dL), excluding urinary Mg$^{2+}$ loss. The depletion of stored Mg$^{2+}$ was also demonstrated in patients[8] and rats[9] with PPIH. Our results clearly showed that transcellular and paracellular Mg$^{2+}$ transport mechanisms were markedly suppressed in the entire intestinal tract of rats with PPIH. Regarding the rate of total Mg$^{2+}$ absorption, the length of intestinal segment, and diameter (Table 1)[25], the small intestine was the major intestinal segment for Mg$^{2+}$ absorption. The percent suppression of total Mg$^{2+}$ absorption in the duodenum, jejunum, ileum, and colon of the rats with PPIH rats was 81.86%, 70.59%, 69.45%, and 39.25% (Table 1), respectively. The percent suppression can be calculated using the following formula: $100 - [(\text{total Mg}^{2+} \text{ absorption of 24-wk-omeprazole group} \times 100) / \text{total Mg}^{2+} \text{absorption of the control group}]$. Therefore, the small intestine is the major affected organs for the adverse effect of prolonged PPI administration. The stimulation of small intestinal Mg$^{2+}$ absorption in PPIH is probably normalizing plasma Mg$^{2+}$ level and requires further study. The present study demonstrated a higher expression level of TRPM6 protein in the colon compared with the small intestinal segment. The up-regulation of TRPM6 and CNNM4 expression in the rats with PPIH was also higher in the colon. The rate of transcellular Mg$^{2+}$ absorption was the highest in the colon of the control and PPIH rats. However, the lowest percent suppression of total Mg$^{2+}$ absorption was found in the colon of the rats with PPIH because single-dose
omeprazole injection had no effect on intraluminal acidic environment in the colon. Our results could explain why the stimulation of colonic Mg$^{2+}$ absorption could not normalize plasma Mg$^{2+}$ level in the previous PPIH mouse model[13].

The overexpression of TRPM6 and CNNM4 proteins in the entire intestinal tract suggested the compensatory response in rats with PPIH. However, the rate of transcellular Mg$^{2+}$ absorption was significantly suppressed in the duodenum, jejunum, ileum, and colon of the rats with PPIH. Hess et al[26], reported two common single-nucleotide polymorphisms (SNPs) in the TRPM6 gene that increase the risk for PPIH. These SNPs may explain why the overexpression of TRPM6 and CNNM4 could not increase transcellular Mg$^{2+}$ absorption.

PPIH rats revealed normal plasma but reduced urinary Ca$^{2+}$ concentration. Increment of plasma 1α,25(OH)$_2$D$_3$ in the rats with PPIH should stimulate intestinal Ca$^{2+}$ absorption, renal tubular Ca$^{2+}$ reabsorption, and bone resorption to regulate plasma Ca$^{2+}$ concentration. Bone resorption not only increases plasma Ca$^{2+}$ but also PO$_4^{3-}$ levels, which trigger FGF-23 release. Plasma 1α,25(OH)$_2$D$_3$ also stimulates FGF-23 release. FGF-23 further suppresses renal tubular PO$_4^{3-}$ reabsorption, which increases urinary PO$_4^{3-}$ excretion[27].

Plasma 1α,25(OH)$_2$D$_3$, FGF-23, and EGF levels are altered in rats with PPIH. However, the hormonal regulation of plasma Mg$^{2+}$ level is largely unknown. In addition, the data from the study of hormonal control of intestinal Mg$^{2+}$ absorption are often confusing and conflicting. Previous research proposed that 1α,25(OH)$_2$D$_3$ stimulates intestinal Mg$^{2+}$ uptake[28]. In addition, 1α,25(OH)$_2$D$_3$ treatment for 7 d exerts no effect on intestinal Mg$^{2+}$ absorption in male C57BL/6 mice[11]. 1α,25(OH)$_2$D$_3$ increases plasma and urinary Mg$^{2+}$ levels[11] possibly through increasing bone resorption. In consideration that 1α,25(OH)$_2$D$_3$ increases urinary Mg$^{2+}$ excretion[11], renal Mg$^{2+}$ wasting is probably involved in the development of hypomagnesemia in prolonged PPI administration[11]. Hypomagnesemia increases plasma FGF-23 level[29].
Figure 3 Effect of omeprazole on plasma and urinary Mg\(^{2+}\), Ca\(^{2+}\), and PO\(_{4}^{3-}\) levels. A: Plasma Mg\(^{2+}\); B: Plasma Ca\(^{2+}\); C: Plasma PO\(_{4}^{3-}\); D: Urinary Mg\(^{2+}\); E: Urinary Ca\(^{2+}\); F: Urinary PO\(_{4}^{3-}\) levels of control, 12 wk-omeprazole-treated, and 24 wk-omeprazole-treated groups. \(^aP<0.05\), \(^bP<0.01\), vs the control group (n = 6).

Khuituan et al.\(^{[29]}\), demonstrated that FGF-23 markedly suppresses intestinal Ca\(^{2+}\) absorption. Magnesiotropic hormone EGF stimulates renal Mg\(^{2+}\) reabsorption to increase plasma Mg\(^{2+}\) level\(^{[20]}\). However, the direct effect of 1\(\alpha\),25(OH)\(_{2}\)D\(_3\), FGF-23, and EGF on segmental intestinal Mg\(^{2+}\) absorption requires further study.

In conclusion, our recent study confirmed the adverse effect of prolonged PPI injection on plasma Mg\(^{2+}\) levels. In specific, PPIs can inhibit intestinal Mg\(^{2+}\) absorption. A higher level of suppression was shown in the small intestine than in the other organs. Therefore, the stimulation of small intestinal Mg\(^{2+}\) probably normalizes plasma Mg\(^{2+}\) in PPIH.
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### Table 1 Rate of total Mg\textsuperscript{2+} absorption, segmental length, and diameter of the duodenum, jejunum, ileum, and colon of the rats

| Segment      | Control | 24-wk omeprazole | Level of suppression | Length (mm)\textsuperscript{[29]} | Diameter (mm)\textsuperscript{[29]} |
|--------------|---------|------------------|----------------------|-----------------------------------|-----------------------------------|
| Duodenum     | 27.68 ± 1.36 | 5.02 ± 0.51 | 81.86% | 95-100 | 2.5-3 |
| Jejunum      | 31.01 ± 1.19 | 9.12 ± 0.75 | 70.59% | 900-1350 | 4.5 |
| Ileum        | 30.77 ± 0.94 | 9.40 ± 0.40 | 69.45% | 25-35 | 3.5 |
| Colon        | 19.76 ± 0.98 | 12.01 ± 0.56 | 39.25% | 90-110 | 3.10 |

Level of suppression was a percentage decrement of total Mg\textsuperscript{2+} absorption in 24-wk omeprazole-treated rats compared with the corresponding control rats.

Figure 4 Effect of omeprazole on plasma 1α,25-dihydroxyvitamin D\textsubscript{3}, parathyroid hormone, fibroblast growth factor 23, epidermal growth factor, and insulin concentrations. A: Plasma 1α,25-dihydroxyvitamin D\textsubscript{3}; B: Plasma parathyroid hormone; C: Plasma fibroblast growth factor 23; D: Plasma epidermal growth factor; E: Plasma insulin of control, 12 wk-omeprazole-treated, and 24 wk-omeprazole-treated groups. \textsuperscript{a}P < 0.05, \textsuperscript{b}P < 0.01, \textsuperscript{c}P < 0.001, vs the control group (n = 6). 1α,25(OH)\textsubscript{2}D\textsubscript{3}: 1α,25-dihydroxyvitamin D\textsubscript{3}; PTH: Parathyroid hormone; FGF-23: Fibroblast growth factor 23; EGF: Epidermal growth factor.
Figure 5  Effect of omeprazole on segmental intestinal Mg\textsuperscript{2+} absorption. A-D: Rate of total, paracellular, and transcellular Mg\textsuperscript{2+} transport of control, 12 wk-omeprazole-treated, and 24 wk-omeprazole-treated groups (A: Duodenum; B: Jejunum; C: Ileum; D: Colon); E: the rate paracellular; F: transcellular Mg\textsuperscript{2+} transport of control, 12 wk-omeprazole-treated, and 24 wk-omeprazole-treated groups. \textsuperscript{a}P < 0.05, \textsuperscript{b}P < 0.01, \textsuperscript{c}P < 0.001, vs the corresponding control group (n = 6). Para: Paracellular; Trans: transcellular.
Figure 6  Transient receptor potential melastatin 6 expression in entire intestinal tract. A: Control; B: 12 wk-omeprazole-treated; C: 24 wk-omeprazole-treated groups. The quantitative immunobloting analysis and representative densitometric analysis of transient receptor potential melastatin 6 expression in duodenum, jejunum, ileum, and colon. *P < 0.05, **P < 0.01, ***P < 0.001, vs the corresponding duodenal segment (n = 6). TRPM6: Transient receptor potential melastatin 6.

Figure 7  The effect of omeprazole on transient receptor potential melastatin 6 expression in entire intestinal tract. A: Quantitative immunobloting analysis of transient receptor potential melastatin 6 expression in duodenum, jejunum, ileum, and colon. B: Duodenum; C: Jejunum; D: Ileum; E: Colon. Representative densitometric analysis of transient receptor potential melastatin 6 expression in duodenum, jejunum, ileum, and colon of control, 12 wk-omeprazole-treated, and 24 wk-omeprazole-treated groups. *P < 0.05, **P < 0.01, ***P < 0.001, vs its corresponding vehicle-treated group (n = 5). TRPM6: Transient receptor potential melastatin 6.
Figure 8 The effect of omeprazole on cyclin M4 expression in entire intestinal tract. A: Quantitative immunobloting analysis of cyclin M4 expression in duodenum, jejunum, ileum, and colon; B: Duodenum; C: Jejunum; D: Ileum; E: Colon. Representative densitometric analysis of cyclin M4 expression in duodenum, jejunum, ileum, and colon of control, 12 wk-omeprazole-treated, and 24 wk-omeprazole-treated groups. *P < 0.05, †P < 0.01, ‡P < 0.001, vs its corresponding vehicle-treated group (n = 5). CNNM4: Cyclin M4.

ARTICLE HIGHLIGHTS

Research background
Dietary intake is the sole source of Mg²⁺ in humans, and intestinal absorption plays a vital role in the regulation of normal Mg²⁺ balance. Previous case reports suggested that intestinal Mg²⁺ malabsorption is a major pathophysiological mechanism in proton pump inhibitor (PPI)-induced hypomagnesemia (PPIH).

Research motivation
The exact mechanism of PPI-inhibited intestinal Mg²⁺ absorption is still controversial. In addition, a simultaneous study on transcellular and paracellular Mg²⁺ absorption in the duodenum, jejunum, ileum, and colon of normal and PPIH had not been performed.

Research objectives
The present study aimed to observe the rate of paracellular and transcellular Mg²⁺ transport across the duodenum, jejunum, ileum, and colon in control and prolonged omeprazole-treated male Sprague-Dawley rats. Magnesiotropic hormones and proteins were measured.

Research methods
The rats received subcutaneous omeprazole injection for 12 or 24 wk. The duodenum, jejunum, ileum, and colon of each rat were mounted onto individual modified Ussing chamber setups to study the rates of total, transcellular, and paracellular Mg²⁺ absorption simultaneously. Magnesiotropic hormones and proteins were observed.
Research results
Hypomagnesemia and hypomagnesuria were demonstrated in the PPIs-treated rats. Plasma 1α,25-dihydroxyvitamin D₃ and fibroblast growth factor 23 increased, whereas plasma epidermal growth factor level decreased in the omeprazole-treated rats. We clearly showed paracellular and transcellular Mg²⁺ absorption in the duodenum, jejunum, ileum, and colon of the control rats. Prolonged PPI treatment significantly inhibited transcellular and paracellular Mg²⁺ absorption in the duodenum, jejunum, ileum, and colon. High transient receptor potential melastatin 6 and cyclin M4 expression in the entire intestinal tract of the PPI-treated rats were demonstrated.

Research conclusions
Prolonged PPI administration markedly inhibits Mg²⁺ absorption throughout the entire length of intestinal tract and lead to systemic Mg²⁺ deficiency.

Research perspectives
PPIs mainly suppressed Mg²⁺ absorption in the small intestine. The stimulation of small PPIs mainly suppressed Mg²⁺ absorption in the small intestine. The stimulation of small

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