Gene Mutation Analysis in Papillary Thyroid Carcinoma Using a Multi-Gene Panel in China

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Purpose: To detect low-frequency mutation in the 57 genes of small panels that are associated with developing thyroid cancer in papillary thyroid carcinoma (PTC) patients and provide patients with precise-targeted therapy.

Patients and Methods: This study included 144 patients diagnosed with PTC who underwent total thyroidectomy and lymph node dissection in the central area of the neck between May 2017 and October 2018. We performed ultra-deep sequencing of 57 genes from 144 patients and detected the 57 genes mutations with bioinformatics.

Results: There were 698 mutations in 45 genes from 138 PTC patients. A high frequency of mutations was detected in the RBM10 gene (44%) and TERT (43%), and some hotspot mutations, such as RBM10:p.E119D and TERT:p.P112fs, were also found.

Conclusion: Ultra-deep sequencing of small gene panels can find some low-frequency mutation genes, which can provide targeted therapy for patients.

Keywords: papillary thyroid carcinoma, thyroid cancer, BRAF, gene panel

Introduction

Thyroid tumors are the most common malignant tumors of the endocrine system, and their incidence has been increasing in the recent decades. Currently, there are some target drugs that can effectively treat PTC, and next-generation sequencing (NGS) can be used for targeted therapy. In order to make better informed treatment decisions for patients with thyroid cancer, we need to detect the gene mutations, especially those associated with potential metastasis. Additionally, it is necessary to identify the different characteristics of oncogenes and tumor suppressor genes related to thyroid tumorigenesis and potential targeting. In the past, due to cost and several other factors, whole-exome sequencing was rarely used to detect thyroid cancer-related genes. In recent years, there have been a variety of multi-gene panels for thyroid tissue examination.1 Technological advances, including NGS methods, have made it possible to use tumor DNA to screen a large number of gene loci from existing clinical samples; the NGS panels can enhance the capture of gene information, and when detecting 2 or more genes, it is more cost-effective for first-generation sequencing. High-throughput massive parallel sequencing technology and bioinformatics can be used to screen for tumor activation and suppressor gene mutations, including point mutations, indels, and copy number aberrations.2

PTC mutations include the MAPK pathway elements, such as the BRAF, HRAS, KRAS, and NRAS genes, and the most important tumor suppressor TP53 gene.3
Approximately one-third of the PTCs have the BRAF V600E mutation. In addition to BRAF mutations, PAX8/PPARG and RET/PTC gene rearrangements are also common in PTC mutations. Thus, different treatment options would be needed for various mutations in the genes associated with PTC. Simultaneous detection of known driver genes and new driver genes, combined with histological findings, will provide more accurate clinical results. In addition, this method may have potential applications in the supplementary diagnosis of suspected thyroid disease.

In this study, we aimed to screen 57 genes mutations related to with developing thyroid cancer in 144 PTC patients in the Shanxi province, China.

**Materials and Methods**

This is a prospective study. We evaluated 144 patients diagnosed with PTC who underwent total thyroidectomy and lymph node dissection in the central area of the neck between May 2017 and October 2018. A total of 144 patients with PTC were included in this study, including 59 men and 85 women within the age range of 21–56 years, with an average age of 48.5±3.5 years. We collect the patient’s age, gender, lymph node metastasis status and BRAF V600E, TERT gene mutation status and other data to analyze the correlation between gene mutation and clinicopathological characteristics. This study was conducted in accordance with the Declaration of Helsinki.

We studied the genomic DNA of surgical samples of PTC. The samples were fresh tissues obtained from the operation. All surgically removed thyroid tissue samples were subjected to detailed histological examination. All patients were diagnosed with PTC. The study was approved by the Research Ethics Committee of Shanxi Provincial People’s Hospital (NO.201992), and all patients provided written informed consent.

The freshly collected tissues were sent to Youxun for genetic testing, and the Illumina NextSeq 500 high-throughput sequencer (Beijing, China) was used for next-generation sequencing (NGS). The sequencing covered 57 genes (Table 1) closely related to thyroid cancer, including part of the exons and the relevant genes containing regions involving single base mutation, insertion or deletion of a small fragment, gene copy number variation, gene fusion, and other types of variation.

**Data Analysis**

Using the FastQC software, raw sequencing of data was done for quality control and low-quality sequences were removed to obtain high-quality sequences; the BWA software was used to align high-quality sequences to the human reference genome (hg19), and Qualimap2 was used for QC (quality control) for alignment. The Mutect2 algorithm in GATK was used to identify raw somatic mutations. To filter germine mutation, we filter out raw somatic mutation with a population frequency >0.1, the mutations in dbSNP. To filter false somatic mutations, we filter raw somatic mutation with a sequencing depth <50, and the number of mutation reads <5. The Oncotator software was used to annotate the mutations, and the mutual exclusion and co-mutation were analyzed with maftools. Search the target drug for thyroid cancer in the Civic database, and analyze the gene mutation in the Civic database.

**Results**

Analysis of the correlation between gene mutation and patients’ clinicopathology. According to the type of gene mutation, it is divided into non-BRAF V600E single-gene mutation group, BRAF V600E single-gene mutation group, non-TERT single-gene mutation group, and TERT single-gene mutation group. The results of univariate

| AKT1 | AKT2 | ALK | APC | ATM | AXIN1 | BRAF |
|------|------|-----|-----|-----|-------|------|
| CCDC6 | CDKN1B | CDKN2A | CDKN2B | CDKN2C | CHEK2 | CTNNB1 |
| EGFR | EBF1AX | EML4 | ERBB2 | EZH1 | FGF2 | GNAS |
| HRAS | IDH1 | KIT | KRAS | MEDI | MEN1 | MET |
| NCOA4 | NCIOR1 | NF1 | NF2 | NOTCH1 | NRAS | NTRK1 |
| OFD1 | NCO2 | PDGFRB | PIK3CA | PPARG | PRKAR1A | PDGFR |
| PTEN | PAX8 | RB1 | RBM10 | RET | SPOP | TFG |
| STRN | RAC1 | TERT | TG | TP53 | TSHR | TRK1 |
| ZNF148 | TERT | TF | TGF | TPI3 | ZNF148 |

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**Table 1** List of Gene in the Panel

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**Table 2** Correlation Between BRAF Mutation and Patients’ Clinicopathology

| Characteristics       | No Gene Mutation (n=42) | BRAF V600E Mutation (n=102) | \( \chi^2/F \) | \( P \) |
|-----------------------|-------------------------|-----------------------------|----------------|--------|
| Gender                |                         |                             |                |        |
| Male                  | 22                      | 37                          | 3.191          | 0.074  |
| Female                | 20                      | 65                          |                |        |
| Average age (years)   |                         |                             | 0.034          | 0.852  |
| ≤45                   | 17                      | 43                          |                |        |
| >45                   | 25                      | 59                          |                |        |
| Pathology (PTC)       |                         |                             | 2.024          | 0.154  |
| Non-PTMC              | 18                      | 57                          |                |        |
| PTMC                  | 24                      | 45                          |                |        |
| Lesion location       |                         |                             | 0.012          | 0.91   |
| Unilateral            | 33                      | 81                          |                |        |
| Bilateral             | 9                       | 21                          |                |        |
| Lymph node metastasis |                         |                             | 0.013          | 0.909  |
| Yes                   | 28                      | 69                          |                |        |
| No                    | 14                      | 33                          |                |        |

**Table 3** Correlation Between TERT Mutation and Patients’ Clinicopathology

| Characteristics       | No Gene Mutation (n=75) | TERT Mutation (n=69) | \( \chi^2/F \) | \( P \) |
|-----------------------|-------------------------|----------------------|----------------|--------|
| Gender                |                         |                      |                |        |
| Male                  | 34                      | 25                   | 1.231          | 0.2672 |
| Female                | 41                      | 44                   |                |        |
| Average age (years)   |                         |                      | 1.61           | 0.204  |
| ≤45                   | 35                      | 25                   |                |        |
| >45                   | 40                      | 44                   |                |        |
| Pathology (PTC)       |                         |                      | 24.87          | 0      |
| Non-PTMC              | 54                      | 21                   |                |        |
| PTMC                  | 21                      | 48                   |                |        |
| Lesion location       |                         |                      | 5.33           | 0.021  |
| Unilateral            | 65                      | 49                   |                |        |
| Bilateral             | 10                      | 20                   |                |        |
| Lymph node metastasis |                         |                      | 0.804          | 0.369  |
| Yes                   | 48                      | 49                   |                |        |
| No                    | 27                      | 20                   |                |        |

Sequencing Overview

The reads number of 144 samples ranged from 4,915,148 to 33,632,576. The align rate of the samples reached more than 99%. As the reads increased, the average coverage also increased (151X-3721X). The reads number and coverage relationship are shown in Figure 1. As the reads number increased, the coverage also increased significantly.

Mutation Frequency Analysis

The mutations identified by the Mutect2 software were filtered germline mutation and low-quality somatic mutation. Finally, a total of 114 exon regions of 57 genes were detected. Six hundred and eighty-nine mutations were detected at 314 position in 45 genes in 138 samples (6 samples were not detected mutation in 57 genes of this panel). The frequency of these mutations was unevenly distributed (0.11–99.84%), as shown in Figure 2. There were 372 mutations with a frequency of <1%, accounting for 53.99% of the mutations, and 439 with <5% frequency, accounting for 63.71%. Most of these mutations were low frequency. These low-frequency mutation sites are difficult to detect by whole-genome sequencing or low-depth whole-exome sequencing (<500X). Using ultra-high-depth panel sequencing can help identify these low-frequency sites. Since most mutations are of low

analysis showed that TERT mutation was related to Lesion location (p=0.021), and it was related to Pathology (p=0). See Tables 2 and 3.
frequency, a Droplet Digital PCR (ddPCR) with high sensitivity is required for verification.

From the Civic database, it was found that 14 genes of thyroid cancer had relevant drug information, as shown in Table 4. The targeted drug site BRAF.p.V600E was found in 102 patients, in which the variant allele frequency (VAF) ranged from 0.245% to 40.27%, among them there were 30 patients with a mutation frequency of <0.5%. For other targeted drug sites, we found that one patient was found to have an NRAS.p.Q61R mutation (11.81%), and another patient was found to have an HRAS.p.G12C (3.2%) mutation, which two patients can benefit from targeted drugs from the Civic database.

Overview of Gene Mutations in the PTC Patients

An overview of all mutations in 144 patients is shown in Figure 3. There were 138 samples mutated in the 57 genes of this panel. Mutations in the BRAF gene were detected in many samples. Mutations were found in up to 74% of the samples, see Figure 4, and all of these were V600E mutations. The mutation frequency of these mutation sites is generally biased. The low-frequency mutations need further verification along with the RBM10 gene mutation, which was observed as a mutation hotspot RBM10: p.E119D (41.13%) in this study, as shown in Figure 5. Finally, the TERT gene also showed mutations in 50% of the samples and was also observed as a mutation hotspot TERT: p.P112fs (37.59%). Figure 6 shows the TERT mutation distribution.

Mutually Exclusive and Co-Mutation Analysis

We observed three pairs of mutually exclusive mutations in BRAF-CDKN2C (p = 0.01273), MED12-BRAF (p = 0.0356), and BRAF-BRAF (p = 0.0372). There were many co-mutated genes, with the most significant being the STK11 and AXIN1 genes (p=6.81 e −06). See Figure 7.

Discussion

Although most PTCs can be successfully treated with radioiodide and levothyroxine suppression after complete surgical intervention, there is still a certain proportion of PTCs that are resistant to treatment and cause comorbidities and death. From an analysis at the molecular level, it has been further clarified that the pathogenesis of PTC is significantly related to thyroid invasion, advanced disease, lymph node metastasis, and tumor recurrence. A multi-gene panel can be used for gene analysis to understand the changes in tumor suppressor genes.15

Now, liquid biopsy can non-invasively reflect the status of tumors in the body, and provide a strong basis for early diagnosis of tumors, individualized treatment monitoring and prognostic judgment. According to the types of tumor-related substances, liquid biopsy covers circulating tumor cells (CTCs), circulating tumor DNA (ctDNA), extracellular vesicles (EVs) and circulating tumor RNA (ctRNA) and other detection targets.16 The detection of thyroid cancer indicators through liquid biopsy technology can effectively assess the risk of thyroid cancer in the subject, so that they can receive more timely and effective treatment, so as to reduce the risk of thyroid cancer. At the same time, after routine examination and diagnosis, the patient can be diagnosed by liquid biopsy technology, and the tumor cell growth and development status and metastasis in the body can be evaluated and diagnosed to further confirm the specific condition of the patient. At the same time, the doctor can perform the diagnosis on the patient’s body and understanding in many aspects, adjusting the diagnosis and treatment plan, and making relevant preventive diagnosis and treatment measures, so that the patient’s condition can be effectively controlled.

At present, droplet digital PCR (ddPCR) is considered to be the preferred technique for detecting rare mutations in liquid biopsy samples.17 Recent studies have shown that ddPCR can
be successfully used to monitor treatment response by quantifying BRAF and RAS mutants in cancer patient samples.

On the other hand, Co-amplification PCR (COLD-PCR) based on lower denaturation temperature is an improved PCR method that allows the preferential amplification of rare mutant alleles within the target amplicon. Recent studies have shown that the sensitivity of cold PCR to detect mutant variants is 10–100 times higher than that of standard PCR. At present, various versions of cold PCR have been developed and successfully used to detect mutant genes, including KRAS, HRAS, NRAS, EGFR, TP53 and BRAF.

| Gene | Variant | Disease | Drugs | Evidence Level |
|------|---------|---------|-------|----------------|
| RET  | M918T   | Thyroid Gland Medullary Carcinoma | Motesanib | B |
| RET  | C634W   | Thyroid Gland Medullary Carcinoma | Motesanib | D |
| RET  | M918T   | Thyroid Gland Medullary Carcinoma | JAK2 Inhibitor AZD1480 | D |
| BRAF | V600E   | Thyroid Gland Medullary Carcinoma | Motesanib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | M918T   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | AKAP9-BRAF | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| NRAS | Q61     | Thyroid Gland Follicular Carcinoma | Perifosine, Temsirolimus | D |
| PAX8 | PAX8-PPARG | Thyroid Gland Follicular Carcinoma | Perifosine, Temsirolimus | D |
| TERT | C228T   | Thyroid Gland Papillary Carcinoma | Perifosine, Temsirolimus | D |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Perifosine, Temsirolimus | D |
| TERT | Promoter Mutation | Thyroid Gland Cancer | Everolimus | C |
| TERT | Promoter Mutation | Thyroid Gland Cancer | Everolimus | C |
| TERT | C228T   | Thyroid Gland Cancer | Everolimus | C |
| RET  | M918T   | Thyroid Gland Medullary Carcinoma | Sorafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | AKAP9-BRAF | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| NRAS | Q61     | Thyroid Gland Follicular Carcinoma | Perifosine, Temsirolimus | D |
| PAX8 | PAX8-PPARG | Thyroid Gland Follicular Carcinoma | Perifosine, Temsirolimus | D |
| TERT | C228T   | Thyroid Gland Papillary Carcinoma | Perifosine, Temsirolimus | D |
| RET  | M918T   | Thyroid Gland Medullary Carcinoma | Sorafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
| BRAF | V600E   | Thyroid Gland Papillary Carcinoma | Vemurafenib | B |
The discovered mutations may have diagnostic and prognostic implications and help develop therapies for these potential molecular drivers. BRAF mutations are present in 45% to 80.8% of PTCs and are related to regional metastasis and extrathyroidal expansion. Once thyroid cancer is highly suspected or diagnosed, risk factors must be considered, such as the clinical risk factors and tumor invasion behavior, patient’s age and sex, initial size and location of the tumor, presence or absence of lymph nodes and/or distant metastasis, cytology and mutations data, and patient preferences. A positive BRAF mutation test indicates that the probability of malignancy is close to 100%. This may help guide the scope of thyroidectomy.

Co-mutation has a synergistic effect on the occurrence and development of PTC and malignant biological behaviors, which can lead to increased tumor invasiveness, and can promote tumor progression to a certain extent, leading to poor prognosis. Co-mutation can be compared with mutations data, and patient preferences. A positive BRAF mutation test indicates that the probability of malignancy is close to 100%. This may help guide the scope of thyroidectomy.
in a single gene. Make PTC more aggressive (prone to extra-thyroid invasion, lymph node metastasis and distant metastasis), worse prognosis, recurrence and death. Co-mutation and exclusion analysis in human tumors are very important for cancer biology studies and treatment design.

In short, using small panel sequencing, genes can be ultra-deeply sequenced. In this study, the mutation coverage depth was 1000X on an average, and many ultra-low-frequency mutations could be detected. Some low-frequency mutations can provide clinicians with important targeted drug medication information. The most important BRAF.p.V600E locus analyzed in this study had multiple mutations, all with very low frequency. Through sequencing of the small panel, we found that the frequency of many gene mutations is much higher than that reported in the previous studies. In previous studies of thyroid cancer, BRAF was the most significant mutation, with a mutation frequency of 56.9% in previous studies, which is higher in this study. It is possible that the use of ultra-deep sequencing may also be a characteristic of the Chinese population. In previous studies, the NRAS gene ranked second, with a mutation frequency of only 9.3%. In this study, its...
Figure 6 TERT mutation distribution.

Figure 7 Overview of mutational mutual exclusion and co-mutation. *Means p < 0.05, Means P < 0.1.
mutation frequency was only 1%. However, the frequency of the second most mutated gene, RBM10, was as high as 44%, which was reported as 0.3% in the previous studies. Moreover, some hotspot mutations, such as the RBM10: p.E119D (41.3%), were found at low frequencies, and the highest mutation frequency of this locus was only 1.721%. For mutations in this type of locus, verification needs to be done by ddPCR in a large number of samples. If this locus is a true positive site, it must be an important target for thyroid cancer, similar to the BRAF.p.V600E locus.

**Conclusion**

The diagnosis of uncertain thyroid lesions is a challenge in cytopathological practice. Some molecular markers, including the most studied BRAF and TERT promoter mutations, have been shown to have prognostic value. Panel sequencing can identify low-frequency mutations and detect additional mutations. In the era of targeted therapy, understanding the molecular characteristics of tumors is crucial for selecting the most appropriate anti-tumor drugs.

Panel sequencing can not only provide clinicians with more detailed references for targeted drug therapy but can also be very helpful for the investigation of driver genes in thyroid cancer.

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**Disclosure**

The authors declare that they have no competing interests.

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