Effect of high temperature on *Wolbachia* density and impact on cytoplasmic incompatibility in confused flour beetle, *Tribolium confusum* (Coleoptera: Tenebrionidae)

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Abstract

**Objectives:** Environmental constraints, especially temperature, have been identified as a key in understanding host-symbiont relationships, as they can directly impact the fitness of the symbiont population and the host development. Here we investigated the effect of temperature during the host development on the density of intracellular bacteria of the *Wolbachia*, wTcon strain within the confused flour beetle, *Tribolium confusum*. The wTcon can induce a complete cytoplasmic incompatibility (CI) in *T. confusum* beetles; therefore, we observed the effect of heat stress on the symbiont-mediated CI.

**Results:** The density of CI inducing *Wolbachia* in the *Tribolium confusum* is temperature-specific. Our observation of the beetles reared in five different temperatures (30–34 °C) measured the highest *Wolbachia* density at 30–31 °C and lowest at 34 °C within a single insect generation. In this species, changes in the density of *Wolbachia* related to higher temperature did not influence CI. However, the fertility of beetles reared in higher temperatures showed a substantial decrease in the number of laid and hatched eggs. Thus, we can confirm the effect of high temperature on lowering the wTcon density and no impact on induction of cytoplasmic incompatibility (CI) in *T. confusum* beetles.

**Keywords:** *Wolbachia* density, Fertility, Cytoplasmic incompatibility, Heat stress

Introduction

Environmental factors are of primary importance in the development and survival of the host-endosymbiont relationship. In particular, the temperature directly impacts the ecological and evolutionary dynamics of populations and the individual's infection development and pathogen virulence [1, 2]. The influence of temperature in various symbiotic model systems may help predict the evolution of local adaptations to regulate infection density [3]. However, the temperature can have a specific effect on both the symbiont and the host separately; the influence on the symbiotic interaction relies on the type of symbiosis between host and symbiont [1, 2]. For example, short exposure to high temperatures in pea aphids eliminates all or most of their bacterial symbiont, *Buchnera aphidiciola*. This resulted in drastically lower fecundity and reduced thermal resistance of hosts due to a deficiency in the production of essential amino acids derived from the obligatory symbiont [4].

*Wolbachia*, arguably the most common animal endosymbiont in nature, is a maternally inherited intracellular bacteria belonging to Alphaproteobacteria, present in arthropods and nematodes [5–7]. *Wolbachia* lineages are classified into 17 supergroups (A–H) based on their
divergence in molecular phylogenetic analyses, which differ in their host range and type of symbiosis, spanning from mutualistic to parasitic [8, 9]. The Wolbachia-host symbiosis can affect the host fitness, especially by manipulating reproduction, e.g., due to feminization, parthenogenesis, male-killing, or cytoplasmic incompatibility (CI) to eventually increase their spread in the host population [6, 10]. The effect of temperature on Wolbachia has always been of considerable interest, as it may influence endosymbiotic density and completeness of CI [11], yet this effect may vary among endosymbiotic strains and hosts. Previous studies showed that extremely high and low temperatures could be lethal for the symbiont [12, 13]. In Drosophila bifasciata, lower Wolbachia density has been recorded at elevated temperature (26 °C) [14]; however, D. simulans males favor low temperature (19 °C) in terms of the infection density, especially during larval development [15]. Interestingly, Wolbachia is even able to manipulate the temperature preference of its hosts [16].

Here we explore the effects of high temperature by comparing Wolbachia density in naturally infected (MN61) and uninfected (HP70) stocks of confused flour beetle, Tribolium confusum, from Kansas, USA. Both T. confusum stocks may differ in their genetic background, and as such, this could influence the fecundity of crosses between them. Previous studies demonstrated the presence of complete CI and reproductive isolation between the beetle populations [17]. One follow-up study reported the probability of the existence of two different Wolbachia strains [5]; nevertheless, Fialho and Stevens [18] showed that out of eight different stocks of T. confusum, all were infected with a single and common CI inducing strain [18–21]. Here, we investigated the effect of high temperature on the density of Wolbachia infection in T. confusum adults. High temperature has a significant impact on Wolbachia density, which might influence the host reproduction, and as such, we investigated (i) how symbiont density is affected by heat, (ii) the impact of heat on CI, and (iii) if high temperature influences sex ratio of the host regarding Wolbachia infection.

Main text
Methods
Insect biology and rearing
In this study, two stocks of Tribolium confusum (Coleoptera: Tenebrionidae) beetles were compared, being either infected (MN61) or uninfected (HP70) with Wolbachia. The beetle’s stock was established from adults and transported from the Stored Product Insect and Engineering Research center of USDA in Kansas, USA. They were stored in container boxes with a feed medium containing a small proportion of brewer yeast (5%) in type 405 wheat flour and maintained at 30 °C and 65±5%RH under a 16:8 (D: L) cycle. Later, beetles were sexed at the pupal stage based on their urogomphi morphology [22].

DNA extraction and detection of Wolbachia
Single adults were removed from their stock containers for DNA extraction with Roboklon tissue and bacterial DNA kit (Roboklon GmbH, Berlin, Germany). Polymerase chain reaction (PCR) was carried out using primer pairs wspF (5′-GCAGCATATACGCAATCTTCA) and wspR (5′-GCATCATCCTTAGCCCGCCTTAT) [20]. PCR was performed in thermocyclers in a total volume of 25 µl (12.5 µl DreamTaq PCR master mix, 0.5 µl for each forward and reverse primer, and 10.5 µl distilled water). The PCR thermal profile used was—one cycle (the 30 s 94 °C) followed by 35 cycles (15 s 94 °C, 30 s 53 °C) and one cycle (30 s 72 °C). Gel electrophoresis demonstrated DNA bands in 1% agarose gel stained in GelRed [23].

Effect of heat on Wolbachia density
Six young infected adults (3–5 days old—3 ♀ and 3 ♂), which were kept at the rearing temperatures of 30–34 °C, were tested to measure the relative Wolbachia density for two consecutive generations by carrying out real-time PCR on Rotor-Gene Q (Qiagen, Hilden, Germany). Each sample was pipetted two times into a 72-well plate and run with two sets of primers and two technical replicates for each sample. As for the first set, a specific pair for Tribolium confusum is Tco261F23 (CAGGATGAA CTGTTTACC) and Tco474R25 (GTAGGTCGATATTA ATTACTG), along with a specific TaqMan probe (FAM-ATCATCTAATATCGCTACGGAGG-TAMRA) to identify T. confusum were used. PCR amplification in a final reaction volume of 20 µl contained 10 µl Premix ExTaq (Probe qPCR, 2×) (ThermoFisher Scientific, MA, USA), 0.4 µl specific forward primer, 0.4 µl specific reverse primer, 0.8 µl TaqMan probe, 7.4 µl ddH2O, and 1 µl template DNA. The PCR cycler conditions were an initial denaturation at 95 °C for the 30 s, followed by 35 cycles of 95 °C for 5 s, 60 °C for 34 s, and a final extension at 72 °C for 10 min [24]. Other sets of primers were designed to detect Wolbachia wsp gene as wspF (5′-GCA GCATATATCAGCAATCTTCA) and wspR (5′-GCATCATCCTTAGCCCGCCTTAT) as well as specific designed TaqMan probe (5′-FAM-TGGTAGCTATGATC TCTGCCGAGAAGAAGG-TAMRA). Real-time quantitative PCR reactions with a total volume of 20 µl contained 10 µl Premix ExTaq (Probe qPCR, 2×) (ThermoFisher Scientific, MA, USA), 0.2 µl specific forward primer, 0.2 µl specific reverse primer, 0.4 µl TaqMan probe, 8.4 µl ddH2O, and 2 µl template DNA. The temperature regime was as follows: 30 s at 95 °C for initial denaturation, then 40 cycles of 95 °C for 5 s, 60 °C for 34 s, and with a final
extension at 72 °C for 10 min [21]. PCR was carried out to attain the crossing point (Cp) values for these markers of each beetle. Differences between the crossing point (ΔCp) of the Wolbachia and Tribolium confusum primers were calculated and then transformed by 2^ΔCp to reach the relative estimates of Wolbachia density [25].

**Test for cytoplasmic incompatibility (CI)**
Beetles reared at 30–34 °C were used to determine the effect of Wolbachia density on CI expression in incompatible crosses under heat stress. Crosses were performed in four combinations, using 3–5 days old males and virgin females (w⁺ × w⁺, w⁺ × w⁻, w⁻ × w⁺, and w⁻ × w⁻) with three cross-replicates per combination. After 3 days, the number of eggs for each cross and assigned temperature were calculated for 30 days. Subsequently, eggs were placed in separate vials containing flour medium and were checked for hatchability. The presence of CI and the level of incompatibility were estimated from this data using temperature as the only independent variable.

**Test for reproduction and sex ratio**
The results from the last experiment, and the numbers of emerging male and female adults (in the pupal stage), were recorded for each vial and temperature every day for 30 days. This data allowed us to estimate the reproduction and survival rate plus the sex ratio (% females) of beetles in five different temperatures.

**Statistical analysis**
For statistical analysis, one-way ANOVA and Tukey post-hoc tests were conducted in JMP v16.2.0 (SAS Institute Inc., Cary, NC, USA) to assess the effect of temperature on the density of endosymbiont bacteria, Wolbachia (Additional file 1), and also its impact on the fertility of T. confusum females. As for the sex ratio, the number of eggs for a pair of beetles was assessed for each temperature (mean ± SD) by the Tukey HSD test, p < 0.05 (Additional file 2).

**Results and discussion**

**Effect of heat on Wolbachia density**
Females and males of T. confusum were reared in five different temperatures from 30 °C as a favorable developmental temperature for the host to 34 °C, the highest temporal degree that the host survived. The relative density of Wolbachia in individual beetles varies with temperature. In two consecutive generations, the density of Wolbachia was not significantly different (F1: F₁,29 = 5.61, p-value = 0.002, F2: F₉,29 = 2.30, p-value = 0.057), however there was an obvious reduction of density for the beetles reared at 34 °C, in comparison with those reared at 30 °C (F1: p-value = 0.0352, F2: p-value = 0.001) and 31 °C (F1: p-value = 0.001) (Fig. 1).

Lu et al. [26] reported Wolbachia density differences between sexes, and we thus analyzed the results of the two sexes separately. However, our results showed no significant difference in the density of males regardless of the temperature that they reared at for two consecutive generations (F1: p-value = 0.46, F2: p-value = 0.45).

**Impact of heat on cytoplasmic incompatibility**
Fecundity can be influenced by the temperature, so we first tested the effect of higher temperature on the fecundity of the virgin females and males in four different crosses respectively (w⁺ × w⁺, w⁺ × w⁻, w⁻ × w⁺, and w⁻ × w⁻). After counting the number of laid eggs for each cross in five different temperatures, no
noticeable differences in the number of laid eggs in all four crosses were found (F1: $F_{4,59} = 8.78$, p-value < 0.001, F2: $F_{4,59} = 5.795$, p-value = 0.0006), except for an obvious decrease of egg production for the beetles reared at 34 °C, in comparison with those reared at 30 °C (F2: p-value = 0.001), 31 °C (F1: p-value = 0.026, F2: p-value = 0.001) and 33 °C (F1: p-value = 0.001) (Fig. 2a, b). However, temperature had no significant impact on the number of laid eggs in crosses of the beetles which were reared at 30–33 °C (F1: $F_{3,15} = 2.256$, p-value < 0.05, F2: $F_{3,15} = 3.027$, p-value < 0.05). In addition, the number of produced eggs for four different crosses regardless of what temperature they reared at, were not significant in both generations (F1: $F_{3,19} = 0.53$, p-value < 0.05, F2: $F_{3,19} = 0.874$, p-value < 0.05).

Afterward, we tested the effect of temperature on the completeness of CI. As for the number of the hatched eggs, the same results applied to the number of hatched eggs by a drastic reduction when reared at the highest survival temperature (34 °C) in all crosses (F1: $F_{4,44} = 9.88$, p-value < 0.0001, F2: $F_{4,44} = 16.56$, p-value < 0.0001). Although a study reported a reduced number of progenies and hatched eggs from the crosses between uninfected males and females in comparison with the other two crosses [21], our results cannot validate this finding (F1: $F_{2,44} = 0.201$, p-value = 0.818, F2: $F_{2,44} = 1.02$, p-value = 0.377). As for the expression of CI under higher temperatures, we found that a complete CI occurred even in higher temperatures (33–34 °C), and no hatched egg could be detected from incompatible crosses of infected males, and uninfected females reared in different temperatures; thus, heat has no significant effect on CI. To some degree, the number of hatched eggs in crosses of both infected males and females ($w^+ \times w^+$) is higher than the hatched eggs for infected females and uninfected males ($w^+ \times w^-$), which may result from the loss of the ability of infected females to restore compatibility with infected males (Fig. 2c, d).

![Box plots showing the effect of high temperature on the number of laid eggs for F1 (a) and F2 (b) in four cross combinations and hatch proportion in three cross combinations for F1 (c) and F2 (d) of Tribolium confusum beetles](image-url)
Additionally, heat stress appears to have no significant effect on the sex ratio, which for both sets of beetles reared under optimal (30 °C) and stress temperatures (34 °C) varied between 0.5 and 0.6, confirming the suggestion that $w^T$ has no significant role in altering the sex ratio [20, 21].

**Conclusion**

Based on our results, higher temperature alters the density of $w^T$ in the *Tribolium confusum*. We showed that among the five different temperatures beetles were reared at, the highest *Wolbachia* density was reported at 30–31 °C, which is also the most favorable temperature range for the host development, while at 34 °C, the density of $w^T$ decreased to a great extent. Furthermore, the fertility of adult females was strongly reduced at 34 °C, with a drastic reduction in the number of laid and hatched eggs. However, based on our study (but with a rather low number of replicates), we found that CI was intact even in the mating crosses between the adult beetles, reared at the highest temperature. Therefore, this might interpret as the low-density requirement of $w^T con$ in the case of inducing CI in *T. confusum*, although this demands further experiments.

**Limitations**

Changes in environmental conditions can affect the infection dynamics and the interaction of *Wolbachia* with their host and, as a result, the ability of the host to adapt in a wild population. Although our findings suggest a spatial and/or seasonal difference in *Wolbachia* densities based on our experiments, field data with wild populations would be essential to understand the effects in a natural setting. Due to time and space constraints, we limited our replicates, which should be resolved in further experiments.

**Abbreviations**

- PCR: Polymerase chain reaction; qPCR: Quantitative polymerase chain reaction; CI: Cytoplasmic incompatibility.

**Supplementary Information**

The online version contains supplementary material available at https://doi.org/10.1186/s13104-022-06123-y.

**Additional file 1.** Relative density of $w^T$ for male and females of *T. confusum*, reared at different temperature based on Cq value for two consecutive generations as (a) F1 and (b) F2.

**Additional file 2.** Results of crosses between *Wolbachia*-infected and uninfected *Tribolium confusum*, rearing continuous at 30 °C (a), 31 °C (b), 32 °C (c), 33 °C (d), 34 °C (e). Statistical analysis by one-way ANOVA and Tukey/Kramer test (P = 0.05) (mean ± standard error).

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**Author contributions**

YG carried out study design, the experiment, molecular laboratory work, and data analyses. CB carried out the study design. YG drafted the manuscript. Both authors read and approved the final manuscript.

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**Availability of data and materials**

All data generated or analyzed during this study are included in this published article and its Additional files.

**Declarations**

**Ethics approval and consent to participate**

Not applicable.

**Consent for publication**

Not applicable.

**Competing interests**

The authors declare that they have no competing interests.

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