Synthesis and in vitro antiproliferative activity of novel (4-chloro- and 4-acyloxy-2-butynyl)thioquinolines

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Abstract The series of new acetylenic thioquinolines containing propargyl, 4-chloro-2-butynyl, and 4-acyloxy-2-butynyl groups have been prepared and tested for antiproliferative activity in vitro against human [SW707 (colorectal adenocarcinoma), CCRF/CEM (leukemia), T47D (breast cancer)] and murine [P388 (leukemia), B16 (melanoma)] cancer lines. Most of the obtained compounds exhibited antiproliferative activity, especially compounds 8, 12, and 21 showed the ID₅₀ values ranging from 0.4 to 3.8 µg/ml comparable to that of cisplatin used as reference compounds.

Keywords Acetylenic thioquinolines · (4-Chloro-2-butynyl)thioquinolines · (4-Acylloxy-2-butynyl)thioquinolines · Antiproliferative activity

Introduction

The carbon–carbon triple bond is one of the most important functional groups in organic chemistry and pharmacology. The structure activity relationship studies suggest that introduction of alkyne motif may significantly modify the chemical, physical, and biological properties of acetylenic compounds (Ben-Zvi and Danon, 1994). Among a large group of synthetic and natural acetylenic compounds the quinolines possessing an alkyne moiety is of particular interest as many of them display important activities, namely antimicrobiological, anticancer, antiprotozoal, and antiretroviral (Fuita et al., 1998; Fakhfakh et al., 2003; Abele et al., 2002). Several acetylenic 2,5-disubstituted decahydroquinoline alkaloids of wide spectra biological activities have been isolated from the skins of frogs, toads and related amphibians (Spande et al., 1999; Michael, 2000). It should be noted that less information is available concerning the synthesis and biological evaluation of alkynylthioquinolines (Abele et al., 2002; Makisumi and Murabayashi, 1969; Boryczka, 1999). It is noteworthy that no data about the synthesis and cytotoxic activity of quinolines containing a selenocyclenyl substituent are available. The chemical and physical properties of selenium are very similar to those of sulfur but the biochemistry differs in at least two respects that distinguish them in biological systems (Aboul-Faddl, 2005). First, in biological systems selenium compounds are metabolized to more reduced states, whereas sulfur compounds are metabolized to more oxidized states; second, selenols are more acidic than thiols, and they are readily oxidized. In general, organoselenium compounds are more reactive than their sulfur analogs due to weaker C–Se bond than the C–S bond. These properties can be involved in higher activity of the Se compounds against cancer cells than S derivatives (Aboul-Faddl, 2005).

The synthetic methods for preparation of acetylenic compounds are of interest especially with regard to the synthesis of enediyne antitumor antibiotics or similar molecules (Nicolau and Dai, 1991; Grissom et al., 1996; Joshi et al., 2007; Kumar et al., 2001). Several cyclic and acyclic models have recently been developed, some of
them including pyridine and quinoline units (Rawat et al., 2001; Knoll et al., 1988; Bhattacharyya et al., 2006).

We have reported a simple and efficient method for the synthesis of 3,4-disubstituted thioquinolines, which possess one or two the same or different O, S, Se acetylenic groups. The new acetylenic thioquinolines exhibited antiproliferative activity in vitro against a broad panel of human and murine cancer cell lines comparable to cisplatin (Boryczka et al., 2002a, 2002b; Mół et al., 2006, 2008). The structure–activity relationships study show a significant correlation between the antiproliferative activity and the electronic properties expressed as $^{13}$C NMR chemical shift, lipophilicity, and molecular electrostatic potential (Boryczka et al., 2002b, 2003; Bajda et al., 2007; Boryczka et al., 2010).

It is well known that several acetylenic compounds possessing 2-butynyl motif exhibit specific pharmacological activities, although the exact role of the 2-butynyl motif in the activity of these derivatives is not fully understood (Ben-Zvi and Danon, 1994).

As an extension of our work on the development of anticancer drugs, we synthesized derivatives 6–12 and 16–25 with the aim to obtain more information about the influence of 4-chloro-2-butynyl and 4-acyloxy-2-butynyl groups on antiproliferative activity in this class of compounds.

Results and discussion

Chemistry

The synthesis of acetylenic thioquinolines 7–12 (Scheme 2) was accomplished starting with 4-chloro-3-methylthioquinoline 3 or 4-chloro-3-propargyl-thioquinoline 4 or 4-chloro-3-(4-hydroxy-2-butynylthio)quinoline 5.

Compounds 3–5 were prepared according to our previously reported methods (Boryczka et al., 2002b; Mół et al., 2008; Maśniakiewicz and Boryczka, 1993). 4-Chloroquinoline 6 was synthesized as shown in Scheme 1. The starting 1 was prepared according to our published procedure (Maśniakiewicz and Boryczka, 1993). Treatment of 1 with sodium methoxide in DMSO at 25°C gave sodium 4-chloro-3-quinolinethiolate 1-A and 4-methoxy-3-methylthioquinoline 2, which was removed by extraction. Sodium salt 1-A after S alkylation using 1-bromo-4-chloro-2-butynye gave 6 in 65% yield.

Compounds 3–5 were converted into 7–12 in 43–86% yields by nucleophilic displacement of chlorine atom by thiourea or selenourea in ethanol, hydrolysis of uronium salt 3-A and subsequent S or Se alkylation of sodium salt 3-B with 1-bromo-4-chloro-2-butynye (Scheme 2).

In order to determine whether a aclyoxy substituent at C-4 of 2-butynyl group has any significant influence on the antiproliferative activity, new compounds bearing 4-acyloxy-2-butynyl groups were prepared.

The synthesis of acetylenic thioquinolines 16–25 (Scheme 3) was accomplished starting with 4-chloro-3-(4-hydroxy-2-butynylthio)quinoline 5 or 4-(4-hydroxy-2-butynylthio)-3-propargylthioquinoline 13 or 4-(4-hydroxy-2-butynylseleno)-3-methylthioquinoline 14 or 4-(4-hydroxy-2-butynylthio)-3-methylthioquinoline 15 which were prepared according to our previously reported methods (Mół et al., 2008).

The compounds 5 and 13–15 were converted into esters 16–25 with 42–91% yields by reactions with acylating agents such as: o-phthalic anhydride, cinnamoyl chloride, and benzoyl chloride or ethyl chloroformate in dry benzene in the presence of pyridine.

The crude products were isolated from aqueous sodium hydroxide by filtration or extraction and separated by column chromatography.

Antiproliferative activity

The seventeen compounds were tested in SRB or MTT (in the case of leukemia cells) assay for their antiproliferative activity in vitro against three human cancer cell lines: SW707 (colorectal adenocarcinoma), CCRF/CEM (leukemia), T47D (breast cancer) and two murine cancer cell lines: P388 (leukemia), B16 (melanoma). The results of cytotoxic activity in vitro were expressed as an ID$_{50}$ ($\mu$g/ml), i.e., the concentration of compound, which inhibits the proliferation of 50% of tumor cells as compared to the control untreated cells. Cisplatin was applied as a referential cytotoxic agent (positive test control). A value of less than 4 $\mu$g/ml was considered as an antiproliferative activity criterion for synthetic compounds. The results of the cytotoxicity studies are summarized in Table 1, previously reported data for compounds 4-chloro-3-(4-hydroxy-2-butynylthio)-quinoline 5, 4-(4-hydroxy-2-butynylseleno)-3-methylthioquinoline 14 and 4-(4-hydroxy-2-butynylthio)-3-methylthioquinoline 15 were included for comparison (Mół et al., 2008).

In general all the compounds 6–12 containing 4-chloro-2-butynyl substituent exhibited a potent antiproliferative activity against human and murine cancer lines applied. 4-Chloro-3-(4-chloro-2-butynylthio)quinoline 6 exhibited high activity against SW707, CCRF/CEM, T47D, B16 and moderate activity against P388. As reported previously 4-chloro-3-(4-hydroxy-2-butynylthio)quinoline 5 possessed lower cytotoxic activity than 6 except activity against the cells of P388 leukemia (Mół et al., 2008).

In the series of derivatives 7–12, the replacement of methyl group by propargyl or 4-hydroxy-2-butynyl, compounds 9, 10 and 11, 12, respectively, resulted in decrease of activity. Among compounds 7–12, the selenium derivatives were more active than sulfur analogs and the
selenium compound 8 showed the most potent activity with the ID_{50} values in the range 0.4–3.5 μg/ml against all cancer lines applied.

Another noteworthy feature of the obtained compounds results was the observation that leukemia (CCRF/CEM and P388) and breast cancer (T47D) cells appear to be more sensitive to the cytotoxic effects of the compounds 7–12 than two other cancer cell lines applied with ID_{50} value of less than 4 μg/ml, which is considered as an antiproliferative activity criterion. It is important to note that the compounds 7–12 exhibited higher cytotoxic activity against breast cancer (T47D) cells than cisplatin.

The replacement of hydroxy group in 5 by hydrophthaloyloxy or cinnamoyloxy groups, compounds 16 and 17, resulted in decrease of activity.

The substitution of hydroxy group in 4-(4-hydroxy-2-butylnylthio)-3-methylthioquinoline 14 by hydrophthaloyloxy, benzoyloxy, and cinnamoyloxy, compounds 19, 21, and 24, respectively, resulted in decrease of activity except activity against the cells of B16 melanoma. A structure–activity relationships observed in compounds 19, 21, and 24 indicated that the rank order of cytotoxic activity, against all cancer lines applied, according to the nature of the acyloxy substituent is as follows: benzoyloxy > hydrophthaloyloxy > cinnamoyloxy.

The replacement of hydroxy group in 4-(4-hydroxy-2-butylnylthio)-3-methylthioquinoline 15 by hydrophthaloyloxy, cinnamoyloxy, benzoyloxy, or ethoxycarbonyloxy, compounds 18, 20, 22, and 23, also resulted in the decrease of activity. Comparing of compounds 18, 20, 22, and 23 indicated that the cytotoxic activity against SW707, CCRF/CEM, T47D, and P388 were in the order ethoxycarbonyloxy > hydrophthaloyloxy > cinnamoyloxy, benzoyloxy.

Whereas the activity of these compounds against B16 was as follows: ethoxycarbonyloxy > cinnamoyloxy > benzoyloxy > hydrophthaloyloxy. It is interesting to note that the acyloxy compounds 16–25, prepared in this study, exhibited the most potent cytotoxicity against cancer cell B16 melanoma. These results may suggest that 4-acyloxy-2-butylnyl function is important for anti-melanoma activity.
Another noteworthy feature of the obtained results was the observation that acyloxy compounds 19, 21, and 24 exhibited the most potent cytotoxicity with ID50 values of 3.1 μg/ml against B16 cancer cell line, among all the compounds (5–25) prepared in this study. The replacement of methyl group by propargyl, compounds 23 and 25, respectively, resulted in decrease of activity.

Conclusions

Novel acetylenic thioquinolines 6–12 and 16–25, possessing in positions 3 and 4, one or two, propargyl, 4-chloro-2-butynyl, or 4-acyloxy-2-butynyl groups were synthesized in good yields using 4-chloro-quinoline derivatives 3–5 and 4-hydroxy-2-butynyl derivatives 13–15 as starting material. The obtained compounds were evaluated for antiproliferative activity in vitro against three human cancer cell lines: SW707 (colorectal cancer), CCRF/CEM (leukemia), T47D (breast cancer) and two murine cancer cell lines: P388 (leukemia), B16 (melanoma). All the tested compounds showed varied activity against different cancer cell lines. As a result of the SAR, it was revealed that the nature of the acetylenic substituent at the C-3 and C-4 positions and character of the heteroatoms (Se and S) at C-4 critically influence the anticancer activity in vitro of the studied compounds. Among the prepared compounds, 8, 12, and 21 were found to be the most active, with ID50 values ranging from 0.4 to 3.8 μg/ml comparable to that of referential anticancer drug, cisplatin. It is of interest to note that the 4-acyloxy-2-butynyl function is important for anti-melanoma activity. The obtained compounds seem to be good candidate for further anticancer activity studies in vitro using a broad panel of human and murine cell lines with the aim to select compounds for studies in vivo.

Experimental

General techniques

Melting points were determined in open capillary tubes on a Boetius melting point apparatus and are uncorrected. 1H NMR (300 MHz) spectra were recorded on a Bruker MSL 300 spectrometer in CDCl3 solvents with tetramethylsilane as internal standard; chemical shifts are reported in ppm (δ) and J values in Hz. Multiplicity is designated as singlet (s), doublet (d), triplet (t), quartet (q), and multiplet (m). Mass spectra were recorded under +CI conditions on Finnigan MAT 95 using isobutane as a reagent and temperature of ion source of 200°C. Elemental C, H, and N analyses were obtained on a Carlo Erba Model 1108 analyzer. TLC was
Table 1: Structures of acetylenic thioquinolines 5–12, 14–25 and their antiproliferative activity in vitro and referential cisplatin against the cells of human and murine cancer cell lines

| Compound | Antiproliferative activity ID$_{50}$ [µg/ml] |
|----------|--------------------------------------------|
|          | Human | Murine |
|          | SW707 | CCRF/CEM | T47D | P388 | B16 |
| 5*       | ![Structure 5](image) | 6.2±1.1 | 4.1±0.3 | - | 2.4±0.0 | Neg |
| 6        | ![Structure 6](image) | 3.3±0.3 | 3.7±1.5 | 3.2±2.0 | 13.4±12.2 | 4.0±0.5 |
| R R' X   |       |       |       |       |       |
| 7        | CH$_3$ | CH$_2$C≡CCH$_2$Cl | S | 3.7±0.3 | 2.5±0.4 | 3.9±0.6 | 2.7±0.8 | 16.9±16.4 |
| 8        | CH$_3$ | CH$_2$C≡CCH$_2$Cl | Se | 3.2±0.2 | 1.7±0.8 | 0.7±0.4 | 0.4±0.3 | 3.5±0.1 |
| 9        | CH$_2$C≡CH | CH$_2$C≡CCH$_2$Cl | S | 12.9±8.2 | 2.9±0.3 | 5.8±1.3 | 3.0±1.1 | 52.8±3.8 |
| 10       | CH$_2$C≡CH | CH$_2$C≡CCH$_2$Cl | Se | 3.2±0.2 | 1.8±1.5 | 0.4±0.1 | 0.8±0.7 | 7.1±3.1 |
| 11       | CH$_2$C≡CCH$_2$OH | CH$_2$C≡CCH$_2$Cl | S | 19.8±15.1 | 3.3±1.1 | 5.3±0.7 | 5.1±3.1 | 43.4±12.0 |
| 12       | CH$_2$C≡CCH$_2$OH | CH$_2$C≡CCH$_2$Cl | Se | 3.3±0.7 | 2.6±2.2 | 1.7±1.4 | 2.0±1.7 | 3.8±0.2 |
| 14*      | CH$_3$ | CH$_2$C≡CCH$_2$OH | S | 3.4±0.1 | 0.5±0.3 | - | 2.6±1.1 | 4.2±0.6 |
| 15*      | CH$_3$ | CH$_2$C≡CCH$_2$OH | Se | 3.4±0.1 | 0.5±0.3 | - | 2.6±1.1 | 4.2±0.6 |
| R R' X   |       |       |       |       |       |
| 16       | COC$_6$H$_4$COOH | Neg | Neg | Neg | Neg | 53.8±21.6 |
| 17       | COCH═CH$_2$C$_6$H$_5$ | 48.8±4.6 | 15.4±9.9 | 22.8±22.3 | 32.8±23.4 | 11.4±9.7 |
| R R' X   |       |       |       |       |       |
| 18       | CH$_3$ | COC$_6$H$_4$COOH | S | 28.1±14.1 | 14.5±2.3 | 36.1±6.9 | 35.0±2.0 | 18.1±10.4 |
| 19       | CH$_3$ | COC$_6$H$_4$COOH | Se | 3.7±0.5 | 4.0±0.7 | 3.2±1.4 | 19.1±11.9 | 3.1±0.9 |
| 20       | CH$_3$ | COC$_6$H$_5$ | S | Neg | 22.2±9.7 | Neg | Neg | 17.8±5.4 |
| 21       | CH$_3$ | COC$_6$H$_5$ | Se | 3.6±0.1 | 1.9±1.4 | 2.2±1.1 | 3.5±0.2 | 2.1±2.1 |
| 22       | CH$_3$ | COC$_6$H$_5$ | S | 33.1±2.6 | 13.2±10.3 | 34.2±6.4 | 34.7±2.2 | 5.5±3.8 |
| 23       | CH$_3$ | COC$_6$H$_5$ | S | 66.6±2.7 | 18.1±9.1 | 71.9±16.7 | Neg | 9.4±8.2 |
| 24       | CH$_3$ | COC$_6$H$_5$ | Se | 4.7±0.5 | 17.8±17.4 | 4.3±0.9 | 43.9±18.9 | 3.1±1.3 |
| 25       | CH$_2$C≡CH | COC$_6$H$_5$ | S | Neg | 50.9±35.1 | Neg | Neg | 14.7±16.5 |
| Cisplatin | Neg | Neg | Neg | Neg | 6.0±0.3 | 1.5±0.2 | 6.7±1.3 | 1.7±0.2 | 4.7±0.2 |

Neg Negative in the concentration used; * See ref. Mól et al., 2008
performed on silica gel 60 254F plates (Merck) using a mixture of chloroform and ethanol (15:1, v/v) as an eluent. UV light and iodine accomplished visualization. Column chromatography was performed on silica gel 60, £63 µm (Merck) using a mixture of chloroform and ethanol (30:1, v/v) as an eluent. Solvents were dried and purified according to literature procedures.

Chemistry

The starting compounds: 4-chloro-3'-methylthio-3,4'-di-quinolinyl sulfide 1 (Maślankiewicz and Boryczka, 1993), 4-chloro-3-(methylthio)quinoline 3 (Maślankiewicz and Boryczka, 1993), 4-chloro-3-propargylthioquinoline 4 (Möl et al., 2006), 4-chloro-3-(4-hydroxy-2-butylnthio)quinoline 5 (Möl et al., 2008), 1-bromo-4-chloro-2-butyne (Bailey and Fujiwara, 1955) were obtained according to methods described previously.

Synthesis of 4-chloro-3-(4-chloro-2-butylnthio) quinoline 6

A mixture of 4-chloro-3'-methylthio-3,4'-di-quinolinyl sulfide 1 (0.74 g, 2 mmol) and sodium methoxide (0.32 g, 6 mmol) in 8 ml DMSO was stirred at room temperature for 15 min. The resultant solid was filtered off, washed with water, dried with anhydrous magnesium sulfate, and evaporated to give crude products 7–12 which were separated by column chromatography using chloroform/ethanol (30:1) to give pure products 7–12.

4-(4-Chloro-2-butylnthio)-3-methylthioquinoline (7) Yield 86%. Mp: 69–70°C. 1H NMR (CDCl3, 300 MHz) δ: 2.68 (s, 3H, S-CH3), 3.73 (t, J = 2.1 Hz, 2H, CH2), 3.86 (t, J = 2.1 Hz, 2H, CH2), 7.62–7.72 (m, 2H, H-6 and H-7), 8.11–8.59 (m, 2H, H-5 and H-8), 8.79 (s, 1H, H-2). CI MS m/z (rel. intensity) 294 (M + H+, 100), 282 (20), 232 (15). Anal. Calc. for C14H12ClNS: C 60.46, H 3.81, N 4.41. Found: C 60.67, H 3.32, N 3.84.

4-(4-Chloro-2-butylnseleno)-3-methylthioquinoline (8) Yield 56%. Mp: 67–68°C. 1H NMR (CDCl3, 300 MHz) δ: 2.67 (s, 3H, S-CH3), 3.62 (t, J = 2.4 Hz, 2H, CH2), 3.88 (t, J = 2.4 Hz, 2H, CH2), 7.61–7.70 (m, 2H, H-6 and H-7), 8.14–8.52 (m, 2H, H-5 and H-8), 8.76 (s, 1H, H-2). CI MS m/z (rel. intensity) 342 (M + H+, 100), 306 (35). Anal. Calc. for C14H12ClNSSe: C 49.35, H 3.55, N 4.11. Found: C 49.47, H 3.38, N 4.20.

4-(4-Chloro-2-butylnseleno)-3-(propargylthio)quinoline (9) Yield 86%. Mp: 92–93°C. 1H NMR (CDCl3, 300 MHz) δ: 2.28 (t, J = 2.7 Hz, 1H, CH), 3.74 (t, J = 2.4 Hz, 2H, CH2), 3.84 (d, J = 2.7 Hz, 2H, CH2S), 3.88 (t, J = 2.4 Hz, 2H, CH2), 7.65–7.72 (m, 2H, H-5 and H-8), 9.01 (s, 1H, H-2). CI MS m/z (rel. intensity) 318 (M + H+, 100), 282 (20), 232 (15). Anal. Calc. for C16H12ClNSSe: C 60.46, H 3.81, N 4.41. Found: C 60.67, H 3.90, N 4.30.

4-(4-Chloro-2-butylnseleno)-3-(propargylthio)quinoline (10) Yield 77%. Mp: 92–93°C. 1H NMR (CDCl3, 300 MHz) δ: 2.28 (t, J = 2.7 Hz, 1H, CH), 3.63 (t, J = 2.4 Hz, 2H, CH2), 3.82 (d, J = 2.7 Hz, 2H, CH2S), 3.89 (t, J = 2.4 Hz, 2H, CH2), 7.66–7.72 (m, 2H, H-5 and H-7), 8.07–8.53 (m, 2H, H-5 and H-8), 8.99 (s, 1H, H-2). CI MS m/z (rel. intensity) 366 (M + H+, 100), 326 (20). Anal. Calc. for C16H12ClNSSe: C 52.69, H 3.32, N 3.84. Found: C 52.77, H 3.40, N 3.68.

4-(4-Chloro-2-butylnthio)-3-(4-hydroxy-2-butylnthio) quinoline (11) Yield 58%. Mp: 103–104°C. 1H NMR (CDCl3, 300 MHz) δ: 3.75 (t, J = 2.1 Hz, 2H, CH2), 3.87–3.89 (m, 4H, 2 × CH2), 4.24 (t, J = 2.1 Hz, 2H, CH2), 7.66–7.74 (m, 2H, H-6 and H-7), 8.10–8.58 (m, 2H, H-5 and H-8), 9.02 (s, 1H, H-2). CI MS m/z (rel. intensity)
4-(4-Chloro-2-butylnylseleno)-3-(4-hydroxy-2-butynylthio)quinoline (12) Yield 43%. Mp: 99–100°C. 1H NMR (CDCl3, 300 MHz) δ: 3.64 (t, J = 2.4 Hz, 2H, CH2), 3.86–3.89 (m, 4H, 2× CH2), 4.24 (t, J = 2.4 Hz, 2H, CH2), 7.63–7.72 (m, 2H, H-5 and H-7), 8.06–8.49 (m, 2H, H-5 and H-8), 8.97 (s, 1H, H-2). CI MS m/z (rel. intensity) 396 (M + H+, 44), 310 (90), 292 (100). Anal. Calc. for C17H14ClNOS2: C 58.70, H 4.06, N 4.03. Found: C 58.62, H 4.15, N 3.86.

4-(4-Hydrophthaloyloxy-2-butylnylseleno)-3-methylthioquinoline (19) Yield 52%. Mp: 126–127°C. 1H NMR (CDCl3, 300 MHz) δ: 2.67 (s, 3H, SCH3), 3.51 (t, J = 2.4 Hz, 2H, CH2), 4.68 (t, J = 2.4 Hz, 2H, CH2), 7.52–7.89 (m, 6H, C6H5 and H-6 and H-7), 8.09–8.40 (m, 2H, H-5 and H-8), 8.78 (s, 1H, H-2). CI MS m/z (rel. intensity) 472 (M + H+, 5), 324 (100). Anal. Calc. for C22H17NO3SSe: C 56.17, H 3.64, N 2.98. Found: C 56.29, H 3.75, N 3.12.

4-(4-Benzoyloxy-2-butylnylthio)-3-methylthioquinoline (20) Yield 90%. Mp: 88–89°C. 1H NMR (CDCl3, 300 MHz) δ: 2.65 (s, 3H, SCH3), 3.74 (t, J = 2.1 Hz, 2H, CH2), 4.68 (t, J = 2.1 Hz, 2H, CH2), 7.42–7.61 (m, 7H, C6H5 and H-6 and H-7), 8.15–8.59 (m, 2H, H-5 and H-8), 8.78 (s, 1H, H-2). CI MS m/z (rel. intensity) 380 (M + H+, 100). Anal. Calc. for C23H19NO2SSe: C 66.47, H 4.52, N 3.69. Found: C 66.34, H 4.48, N 3.78.

General procedure for the synthesis of acetylenic thioquinolines 16–25

A mixture of 4-chloro-3-(4-hydroxy-2-butylnylthio)quinoline 5 (0.53 g, 2 mmol) or 4-(4-hydroxy-2-butylnylthio)-3-propargylthioquinoline 13 or (0.60 g, 2 mmol) or 4-(4-hydroxy-2-butylnylseleno)-3-methylthioquinoline 14 (0.64 g, 2 mmol) or 4-(4-hydroxy-2-butylnylthio)-3-methylthioquinoline 15 (0.55 g, 2 mmol) and pyridine (0.17 g, 2.1 mmol) and (2.1 mmol) o-phthalic anhydride or cinnamoyl chloride or benzoil chloride or ethyl chloroformate in dry benzene (8 ml) was stirred at 70°C for about 1 h (monitored by TLC until complete consumption of starting materials) and then concentrated in vacuo. The residue was separated by column chromatography using chloroform/ethanol (30:1) to give pure products 16–25.

4-Chloro-3-(4-hydrophthaloyloxy-2-butylnylthio)quinoline (16) Yield 64%. Mp: 97–98°C. 1H NMR (CDCl3, 300 MHz) δ: 3.66 (t, J = 2.1 Hz, 2H, CH2), 4.81 (t, J = 2.1 Hz, 2H, CH2), 7.31–7.64 (m, 6H, C6H5 and H-6 and H-7), 7.72–8.10 (m, 2H, H-5 and H-8), 8.28 (s, 1H, H-2). CI MS m/z (rel. intensity) 412 (M + H+, 10), 246 (100). Anal. Calc. for C21H14ClNO2S: C 61.24, H 3.43, N 3.40. Found: C 61.42, H 3.50, N 3.31.

4-Chloro-3-(4-cinnamoyloxy-2-butylnylthio)quinoline (17) Yield 60%. Mp: 123–124°C. 1H NMR (CDCl3, 300 MHz) δ: 3.84 (t, J = 2.1 Hz, 2H, CH2), 3.74 (t, J = 2.1 Hz, 2H, CH2), 6.37 (d, J = 15.9 Hz, 1H, CH), 7.39–7.73 (m, 8H, CH and C6H5 and H-6 and H-7), 8.07–8.23 (m, 2H, H-5 and H-8), 9.00 (s, 1H, H-2). CI MS m/z (rel. intensity) 394 (M + H+, 100). Anal. Calc. for C22H16ClNO2S: C 67.09, H 4.09, N 3.56. Found: C 67.25, H 3.91, N 3.62.

4-(4-Hydrophthaloyloxy-2-butylnylthio)-3-methylthioquinoline (18) Yield 50%. Mp: 96–97°C. 1H NMR (CDCl3, 300 MHz) δ: 2.64 (s, 3H, SCH3), 3.61 (t, J = 2.1 Hz, 2H, CH2), 4.63 (t, J = 2.1 Hz, 2H, CH2), 7.26–7.93 (m, 6H, C6H4 and H-6 and H-7), 8.01–8.48 (m, 2H, H-5 and H-8), 8.85 (s, 1H, H-2). CI MS m/z (rel. intensity) 424 (M + H+, 10), 276 (100). Anal. Calc. for C23H17NO2S2: C 62.39, H 4.05, N 3.31. Found: C 62.55, H 4.10, N 3.22.

4-(4-Benzoyloxy-2-butylnylthio)-3-methylthioquinoline (21) Yield 54%. Mp: 92–93°C. 1H NMR (CDCl3, 300 MHz) δ: 2.64 (s, 3H, SCH3), 3.63 (t, J = 2.4 Hz, 2H, CH2), 4.69 (t, J = 2.4 Hz, 2H, CH2), 7.42–7.99 (m, 7H, C6H5 and H-6 and H-7), 8.05–8.54 (m, 2H, H-5 and H-8), 8.75 (s, 1H, H-2). CI MS m/z (rel. intensity) 428 (M + H+, 100). Anal. Calc. for C23H18NO4SSe: C 57.87, H 4.93, N 4.03. Found: C 58.98, H 4.85, N 4.19.

4-(4-Ethoxycarbonyloxy-2-butylnylthio)-3-methylthioquinoline (22) Yield 93%. Mp: 48–49°C. 1H NMR (CDCl3, 300 MHz) δ: 1.30 (t, J = 7.2 Hz, 3H, CH3), 2.68 (s, 3H, SCH3), 3.70 (t, J = 2.4 Hz, 2H, CH2), 4.18 (q, J = 7.2 Hz, 2H, OCH2), 4.85 (t, J = 2.4 Hz, 2H, CH2), 7.61–7.73 (m, 2H, H-6 and H-7), 8.05–8.59 (m, 2H, H-5 and H-8), 8.79 (s, 1H, H-2). CI MS m/z (rel. intensity) 348 (M + H+, 100). Anal. Calc. for C24H18NO4S2: C 58.77, H 4.93, N 4.03. Found: C 58.98, H 4.85, N 4.19.

4-(4-Cinnamoyloxy-2-butylnylthio)-3-methylthioquinoline (23) Yield 91%. Mp: 82–83°C. 1H NMR (CDCl3, 300 MHz) δ: 2.68 (s, 3H, SCH3), 3.73 (t, J = 2.1 Hz, 2H, CH2), 4.57 (t, J = 2.1 Hz, 2H, CH2), 6.36 (d, J = 16.2 Hz, 1H, CH), 7.39–7.68 (m, 8H, CH and C6H5 and H-6 and H-7), 8.04–8.59 (m, 2H, H-5 and H-8), 8.80 (s, 1H, H-2). CI MS m/z (rel. intensity) 406 (M + H+, 100). Anal. Calc. for C23H19NO3SSe: C 68.12, H 4.72, N 3.45. Found: C 68.32, H 4.56, N 3.48.
4-(4-Cinnamoyloxy-2-butynylseleno)-3-methylthioquinoline (24) Yield 42%. Mp: 98–99°C. 1H NMR (CDCl₃, 300 MHz) δ: 2.67 (s, 3H, SCH₃), 3.63 (t, J = 2.1 Hz, 2H, CH₂), 4.58 (t, J = 2.1 Hz, 2H, CH₂), 6.37 (d, J = 15.9 Hz, 1H, CH), 7.39–7.69 (m, 8H, CH and C₆H₅ and H-6 and H-7), 8.02–8.53 (m, 2H, H-5 i H-8), 8.77 (s, 1H, H-2). CI MS m/z (rel. intensity) 453 (M + H⁺, 90), 256 (100). Anal. C. 61.06, H 4.23, N 3.10. Found: C 60.81, H 4.12, N 3.18.

4-(4-Cinnamoyloxy-2-butynylthio)-3-(propargylthio)quinoline (25) Yield 80%. Mp: 102–103°C. 1H NMR (CDCl₃, 300 MHz) δ: 2.27 (t, J = 2.7 Hz, 1H, CH), 3.75 (t, J = 2.4 Hz, 2H, CH₂), 3.84 (d, J = 2.7 Hz, 2H, SCH₂), 4.58 (t, J = 2.4 Hz, 2H, CH₂), 6.36 (d, J = 15.9 Hz, 1H, CH), 7.39–7.69 (m, 8H, CH and C₆H₅ and H-6 and H-7), 8.07–8.60 (m, 2H, H-5 and H-8), 9.01 (s, 1H, H-2). CI MS m/z (rel. intensity) 430 (M + H⁺, 20), 232 (100). Anal. C. 69.90, H 4.46, N 3.26. Found: C 70.12, H 4.52, N 3.38.

Antiproliferative assay in vitro

Cells

The following established in vitro cancer cell lines were applied: SW707 (human colorectal adenocarcinoma), CCRF/CEM (human leukemia), T47D (human breast cancer), P388 (mouse leukemia), and B16 (mouse melanoma). All lines were obtained from the American Type Culture Collection (Rockville, Maryland, USA) and maintained at the Cell Culture Collection of the Institute of Immunology and Experimental Therapy, Wroclaw, Poland.

Twenty-four hours before addition of the tested agents, the cells were plated in 96-well plate (Sarstedt, USA) at a density of 10⁴ cells per well in 100 l of culture medium. The cells were cultured in the opti-MEM medium supplemented with 2 mM glutamine (Gibco, Warsaw, Poland), streptomycin (50 μg/ml), penicillin (50 U/ml) (both antibiotics from Polfa, Tarchomin, Poland), and 5% fetal calf serum (Gibco, Grand Island, USA). The cell cultures were maintained at 37°C in humid atmosphere saturated with 5% CO₂.

SRB assay

The details of this technique were described by Skehan et al. (1990). The cytotoxicity assay was performed after 72-h exposure of the cultured cells to varying concentrations (from 0.1 to 100 μg/ml) of the tested agents. The compounds were dissolved in 10% DMSO to concentration of 1 mg/ml, and subsequently diluted in culture medium to reach the required concentrations. DMSO, which was used as a solvent did not exert any inhibitory effect on cell proliferation. The cells attached to the plastic were fixed by gently layering cold 50% TCA (trichloroacetic acid, Aldrich-Chemie, Germany) on the top of the culture medium in each well. The plates were incubated at 4°C for 1 h and then washed five times with tap water. The background optical density was measured in the wells filled with culture medium, without the cells. The cellular material fixed with TCA was stained with 0.4% sulforhodamine B (SRB, Sigma, Germany) dissolved in 1% acetic acid (POCh, Gliwice, Poland) for 30 min. Unbound dye was removed by rinsing (4×) with 1% acetic acid. The protein-bound dye was extracted with 10 mM unbuffered tris base (POCh, Gliwice, Poland) for determination of optical density (at 540 nm) in a computer-interfaced, 96-well microtiter plate reader Multiskan RC photometer (Labsystems, Helsinki, Finland). Each compound in given concentration was tested in triplicates in each experiment, which was repeated 3–5 times.

MTT assay

This technique was applied for the cytotoxicity screening against mouse leukemia cells growing in suspension culture. An assay was performed after 72-h exposure to varying concentrations (from 0.1 to 100 μg/ml) of the tested agents. The compounds were dissolved in 10% DMSO to concentration of 1 mg/ml, and subsequently diluted in culture medium to reach the required concentrations. DMSO, which was used as a solvent did not exert any inhibitory effect on cell proliferation. For the last 3–4 h of incubation 20 μl of MTT solution were added to each well (MTT: 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide; stock solution: 5 mg/ml). The mitochondria of viable cells reduce the pale yellow MTT to a navy blue formazan: the more viable cells are present in the background optical density was measured in the wells filled with culture medium, without the cells. The cellular material fixed with TCA was stained with 0.4% sulforhodamine B (SRB, Sigma, Germany) dissolved in 1% acetic acid (POCh, Gliwice, Poland) for 30 min. Unbound dye was removed by rinsing (4×) with 1% acetic acid. The protein-bound dye was extracted with 10 mM unbuffered tris base (POCh, Gliwice, Poland) for determination of optical density (at 540 nm) in a computer-interfaced, 96-well microtiter plate reader Multiskan RC photometer (Labsystems, Helsinki, Finland). Each compound in given concentration was tested in triplicates in each experiment, which was repeated 3–5 times.

The results of cytotoxic activity in vitro were expressed as an ID₅₀—the dose of compound (in μg/ml) that inhibits proliferation rate of the tumor cells by 50% as compared to the control untreated cells.
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