Alzheimer’s disease: a mathematical model for onset and progression

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In this article we propose a mathematical model for the onset and progression of Alzheimer’s disease based on transport and diffusion equations. We regard brain neurons as a continuous medium and structure them by their degree of malfunctioning. Two different mechanisms are assumed to be relevant for the temporal evolution of the disease: i) diffusion and agglomeration of soluble polymers of amyloid, produced by damaged neurons and ii) neuron-to-neuron prion-like transmission. We model these two processes by a system of Smoluchowski equations for the amyloid concentration, coupled to a kinetic-type transport equation for the distribution function of the degree of malfunctioning of neurons. The second equation contains an integral term describing the random onset of the disease as a jump process localized in particularly sensitive areas of the brain. Our numerical simulations are in good qualitative agreement with clinical images of the disease distribution in the brain which vary from early to advanced stages.

Keywords: Alzheimer’s disease; transport and diffusion equations; Smoluchowski equations; numerical simulations.

1. Introduction

Alzheimer’s disease (AD) is one of the most common late life dementia, with huge social and economic impact (Mattson, 2004; Blennow et al., 2006; Hurd et al., 2013). Its global prevalence, about 24 millions in 2011, is expected to double in 20 years Reitz et al. (2011). In silico research, based on mathematical modelling and computer simulations (Good & Murphy, 1996; Cruz et al., 1997; Urbanc et al., 1999; Murphy & Pallitto, 2000; Edelstein-Keshet & Spiross, 2002; Raj et al., 2012; Achdou et al., 2013;
Helal et al., 2013) effectively supplements in vivo and in vitro research. We present a multiscale model for the onset and evolution of AD which accounts for the diffusion and agglomeration of amyloid-β (Aβ) peptide (amyloid cascade hypothesis; (Haass & Selkoe, 2007; Karran et al., 2011)), and the spreading of the disease through neuron-to-neuron transmission (prionoid hypothesis, Braak & Del Tredici, 2011).

Indeed, to cover such diverse facets of AD in a single model, different spatial and temporal scales must be taken into account: microscopic spatial scales to describe the role of the neurons, macroscopic spatial and short temporal (minutes, hours) scales for the description of relevant diffusion processes in the brain and large temporal scales (years, decades) for the description of the global development of AD. The way in which we combine distinct scales in a single model forms the core and major novelty of the article.

Following closely the biomedical literature on AD, we briefly describe the processes which we shall include in our model. In the neurons and their interconnections, several microscopic phenomena take place. It is largely accepted that beta amyloid (Aβ), especially its highly toxic oligomeric isoforms Aβ40 and Aβ42, play an important role in the process of the cerebral damage (the so-called amyloid cascade hypothesis; Karran et al., 2011). In this note we, focus on the role of Aβ42 in its soluble form, which recently has been suggested to be the principal cause of neuronal death and eventually dementia (Walsh & Selkoe, 2007). At the level of the neuronal membrane, monomeric Aβ peptides originate from the proteolytic cleavage of a transmembrane glycoprotein, the amyloid precursor protein (APP). By unknown and partially genetic reasons, some neurons present an unbalance between produced and cleared Aβ (we refer to such neurons as damaged neurons). In addition to this, it has been proposed that neuronal damage spreads in the neuronal net through a neuron-to-neuron prion-like propagation mechanism (Braak & Del Tredici, 2011; Raj et al., 2012).

On the other hand, macroscopic phenomena take place at the level of the cerebral tissue. The monomeric Aβ produced by damaged neurons diffuses through the microscopic tortuosity of the brain tissue and undergoes a process of agglomeration, leading eventually to the formation of long, insoluble amyloid fibrils, which accumulate in spherical deposits known as senile plaques. Moreover, soluble Aβ shows a multiple neurotoxic effect: it induces a general inflammation that activates the microglia (the resident immune cells in the central nervous system) which in turn secretes proinflammatory innate cytokines (Griffin et al., 1998) and, at the same time, increases intracellular calcium levels (Good & Murphy, 1996) yielding ultimately apoptosis and neuronal death.

The model we present is a conceptual interdisciplinary construction based on clinical and experimental evidence, yielding in particular numerical simulations and related graphs, that can be compared with time-dependent trajectories of AD biomarkers (see e.g. Jack et al., 2010, 2013). In particular, Fig. 2 fits the core of the model proposed in Jack et al. (2010) for the temporal progression of the abnormalities in AD biomarkers, which identifies two subsequent periods:

- a first period of β-amyloidosis characterized prevalently by reductions in cerebrospinal fluid (CSF) Aβ42 and increased amyloid plaque formation (biomarkers of this first period in our model correspond to CSF-Aβ42 and PIB-PET, Pittsburgh compound B - Positron Emission Tomography);
- a second one characterized by neuronal dysfunction and neurodegeneration (for this period, we only take into account the structural magnetic resonance imaging (MRI)).

Of particular medical interest is the initial stage of the second period, which is commonly referred to as mild cognitive impairment (MCI): see, e.g. Petersen et al. (2009).
2. Mathematical model

Highly toxic oligomeric isoforms of beta amyloid, $\text{A}^\beta_{40}$ and $\text{A}^\beta_{42}$, cause cerebral damage. Here we restrict our attention to $\text{A}^\beta_{42}$ (shortly $\text{A}^\beta_{42}$ in the sequel) in soluble form, generally considered the principal cause of neuronal death and dementia (Walsh & Selkoe, 2007). Monomeric $\text{A}^\beta$ peptides originate from proteolytic cleavage of a transmembrane glycoprotein, the APP. In AD, neurons progressively present an unbalance between produced and cleared $\text{A}^\beta$, but the underlying mechanism is still largely unknown. On the other hand, it was proposed that neuronal damage spreads in the neural pathway through a neuron-to-neuron prion-like propagation mechanism (Braak & Del Tredici, 2011; Raj et al., 2012).

Soluble $\text{A}^\beta$ diffuses through the microscopic tortuosity of the brain tissue and undergoes an aggregation process. Eventually, this leads to the formation of long, insoluble fibrils, accumulating in spherical deposits known as senile plaques. Soluble $\text{A}^\beta$ has a multiple neurotoxic effect (Good & Murphy, 1996; Griffin et al., 1998). In our model, we do not enter the details of the brain tissue, we neglect the action of the $\tau$-protein, we simplify the role of microglia and neglect its multifaceted mechanism (see, e.g. Edelstein-Keshet & Spiross, 2002; Quinlan & Straughan, 2005). We simply assume that high levels of soluble amyloid are toxic for neurons.

We identify a portion of the cerebral tissue with a three-dimensional region $\Omega$ and $x \in \Omega$ indicates a generic point. Two temporal scales are needed to simulate the longitudinal evolution of the disease over a period of years: a short (i.e. rapid) $s$-scale (unit time coincides with hours) for the diffusion and aggregation of $\text{A}^\beta$ Meyer-Luehmann et al. (2008) and a long (i.e. slow) $t$-scale (unit time coincides with several months) for the progression of AD, so $\Delta t = \varepsilon \Delta s$ for a small constant $\varepsilon \ll 1$.

We denote the molar concentration of soluble $\text{A}^\beta$ polymers of length $m$ at point $x$ and time $s$ by $u_m(x, s)$, with $1 < m < N$. That of clusters of oligomers of length $\geq N$ (fibrils) is denoted by $u_N(x, s)$ and may be thought as a medical parameter (the plaques), clinically observable through PIB-PET (Nordberg, 2008).

To model the aggregation of $\text{A}^\beta$ $m$-polymers ($1 < m < N$) we follow Achdou et al. (2013),

\[\text{[variation in (short) time]} = \text{[diffusion]} + \text{[agglomeration]},\]

which, in mathematical terms, leads to the Smoluchowski equation with diffusion:

\[
\partial_s u_m = d_m \nabla^2 u_m + \left[ \frac{1}{2} \sum_{j=1}^{m-1} a_{j,m-j} u_j u_{m-j} - u_m \sum_{j=1}^{N} a_{m,j} u_j \right]. \tag{2.1}
\]

where $d_m > 0$, $m = 1, \ldots, N$, and $a_{i,j} = a_{j,i} > 0$, $i, j = 1, \ldots, N$ (giving the factor $\frac{1}{2}$ in (2.1)).

We refer to Franchi & Tesi (2012) and Achdou et al. (2013) for an extensive discussion of (2.1). For reasons related to the model, we can assume that the diffusion coefficients $d_m$ are small when $m$ is large, since big assemblies do not move. In fact, the diffusion coefficient of a soluble peptide scales approximately as a reciprocal of the cube root of its molecular weight (see Goodhill, 1997 and also Nicholson & Syková, 1998).

Applications of the Smoluchowski equation to the description of the agglomeration of $\text{A}^\beta$ amyloid appear in Murphy & Pallitto (2000). In this article, the authors compare experimental data, obtained in vitro, with numerical simulations based on the Smoluchowski equation (without diffusion) in order to describe the process leading to insoluble fibril aggregates from soluble amyloid. The form of the coefficients $a_{i,j}$ (the coagulation rates) we use has been considered in vitro by Murphy & Pallitto (see
According to formula (13) in Murphy & Pallitto (2000), the coagulation rates \emph{in silico} in our equations take the form

\begin{equation}
 a_{i,j} = \text{const.} \frac{1}{i+j} \left( \frac{\ln(i/d) + v_i}{i} + \frac{\ln(j/d) + v_j}{j} \right),
\end{equation}

where \( i, j \) are the lengths of the fibrils, \( d \) is their diameter, and \( v_i = 0.312 + 0.565(i/d)^{-1} - 0.1(i/d)^{-2} \).

The physical arguments leading to formula (2.2) rely on sophisticated statistical mechanics considerations (see also Tomsky & Murphy, 1992).

Since \( d \) can be assumed very small, without loss of generality we can assume \( v_i = v \) for \( i = 1, \ldots, N-1 \). Thus we can replace (2.2) by

\begin{equation}
 a_{i,j} = \text{const.} \frac{1}{i+j} \left( \frac{\ln(i/d) + v}{i} + \frac{\ln(j/d) + v}{j} \right) = \frac{1}{ij} \left( v + |\ln d| + O(\ln N) \right).
\end{equation}

Since \( N \) is finite, in our numerical simulations we use a slightly approximate form of these coefficients, taking

\begin{equation}
 a_{i,j} = \alpha \frac{1}{ij}, \quad \text{where} \; \alpha > 0.
\end{equation}

Smoluchowski equations with diffusion have already been considered in the literature (without reference to \( A\beta \) amyloid and AD) with diverse boundary conditions: see, for instance, Drake (1972) for a general introduction, and Wrzosek (1997); Amann (2000); Amann & Weber (2001); Laurençot & Mischler (2002); Amann & Walker (2005).

Neurons produce \( A\beta \) monomers, whence the equation for \( u_1 \) contains a source term \( \mathcal{F} \):

\begin{equation}
 \partial_t u_1 = d_1 \nabla^2 u_1 - u_1 \sum_{j=1}^{N} a_{1,j} u_j + \mathcal{F}.
\end{equation}

Since fibrils are assumed not to move, the equation for \( u_N \) has no diffusion term and takes the form (see (4) in Achdou \textit{et al.}, 2013):

\begin{equation}
 \partial_t u_N = \sum_{j+k \geq N, k < N} a_{j,k} u_j u_k.
\end{equation}

It is coherent with experimental data to assume \( a_{N,N} = 0 \) for large \( N \). This is equivalent to saying that large oligomers do not aggregate with each other.

The justification of the condition \( j, k < N \) in (2.6) requires a few more words. In fact, we must remember that the meaning of \( u_N \) differs from that of \( u_m, m < N \), as well as the identity

\begin{equation}
 \frac{1}{2} \sum_{j+k \geq N, k < N} a_{j,k} u_j u_k = \frac{1}{2} \sum_{j+k \geq N} a_{j,k} u_j u_k - u_N \sum_{j=1}^{N} a_{N,j} u_j.
\end{equation}
The idea is that \( u_N \) should describe the sum of the densities of all the ‘large’ assemblies. We assume that large assemblies exhibit all the same coagulation properties and do not coagulate with each other. Let us briefly show how (2.6) is obtained: we start by writing the exact Smoluchowski equation for all \( m \geq 1 \) using \( \tilde{u}_m \) instead of \( u_m \) in order to avoid confusion, i.e. nothing but the PDE in (2.1) with \( m \) ranging from 2 to \( \infty \). We have

\[
\frac{\partial}{\partial t} \tilde{u}_m = d_m \nabla^2 \tilde{u}_m - \tilde{u}_m \sum_{j=1}^{N} a_{m,j} \tilde{u}_j + \frac{1}{2} \sum_{j=1}^{m-1} a_{j,m-j} \tilde{u}_j \tilde{u}_{m-j},
\]

(2.8)

where, coherently with our assumptions, we assume

i) \( d_m = d_N \) for \( m \geq N \);

ii) \( a_{m,j} = a_{N,j} \) for \( m \geq N \). In particular, if \( m, j \geq N \), \( a_{N,j} = a_{N,N} = 0 \).

Therefore, if \( m \geq N \), (2.8) becomes

\[
\frac{\partial}{\partial t} \tilde{u}_m = d_N \nabla^2 \tilde{u}_m - \tilde{u}_m \sum_{j=1}^{N-1} a_{N,j} \tilde{u}_j + \frac{1}{2} \sum_{j=1}^{m-1} a_{j,m-j} \tilde{u}_j \tilde{u}_{m-j},
\]

(2.9)

Now we sum up (2.9) for \( m \geq N \), and we set for a while \( v := \sum_{m \geq N} \tilde{u}_m \). We want to show precisely that \( v \) satisfies (2.6) (satisfied by \( u_N \)). By i), we have

\[
\frac{\partial v}{\partial t} = d_N \nabla^2 v - \sum_{m \geq N} \tilde{u}_m \sum_{j=1}^{N-1} a_{N,j} \tilde{u}_j + \frac{1}{2} \sum_{m \geq N} \sum_{i=1}^{m-1} a_{i,m-i} \tilde{u}_i \tilde{u}_{m-i}
\]

:= \( d_N \nabla^2 v - I_1 + \frac{1}{2} I_2 \)

It is clear that

\[
I_1 = \sum_{m \geq N} \tilde{u}_m \sum_{j=1}^{N-1} a_{N,j} \tilde{u}_j = v \sum_{j=1}^{N-1} a_{N,j} \tilde{u}_j,
\]

which is precisely the second term in (2.7), since \( a_{N,N} = 0 \). As for \( I_2 \), if we set \( j := i \) and \( k := m - i \), we obtain the first term in (2.7). Finally, if set \( u_m = \tilde{u}_m \) for \( m < N \) and \( u_N = v \) we recover the PDE in (2.6), as desired.

We model the degree of malfunctioning of a neuron with a parameter \( a \) ranging from 0 to 1: \( a \) close to 0 stands for ‘the neuron is healthy’, whereas \( a \) close to 1 for ‘the neuron is dead’. This parameter, although introduced for the sake of mathematical modelling (see also Raj et al., 2012), can be compared with medical images from Fluorodeoxyglucose PET (FDG-PET Mosconi et al., 2010).

Given \( x \in \Omega, t \geq 0, \) and \( a \in [0, 1], \)

\[
f(x, a, t) \, da
\]
indicates the fraction of neurons close to \( x \) with degree of malfunctioning at time \( t \) between \( a \) and \( a + da \).

The progression of AD occurs at the long time scale \( t \), over decades, and is determined by the deterioration rate, \( v = v(x, a, t) \), of the health state of the neurons:

\[
\partial_t f + \partial_a (fv[f]) = 0. \tag{2.10}
\]

Here \( v[f] \) indicates that the deterioration rate depends on \( f \) itself. The onset of AD will be included in a subsequent step.

We assume that

\[
v[f] = \int_{\Omega \times [0, 1]} K(x, a, y, b)f(y, b, t) \, dy \, db + S(x, a, u_1(x, s), \ldots, u_{N-1}(x, s)). \tag{2.11}
\]

The integral term describes the possible prion-like propagation of AD through the neural pathway. Malfunctioning neighbours are harmful for a neuron’s health state, while healthy ones are not:

\[
\begin{align*}
K(x, a, y, b) &\geq 0 \quad \forall \, x, y \in \Omega, \, a, b \in [0, 1], \\
K(x, a, y, b) &= 0 \quad \text{if } a > b.
\end{align*}
\]

Typically

\[
K(x, a, y, b) = G(x, a, b)H(x, y)
\]

with, e.g.

\[
G(x, a, b) = C_G(b - a)^+, \quad H(x, y) = h(|x - y|),
\]

where \((\cdot)^+\) denotes the positive part \((x^+ := \max\{0, x\})\), while \(h(r)\) is a nonnegative and decreasing function, which vanishes at some \( r = r_0 \) and satisfies \( \int_{|y|<r_0} h(|y|) \, dy = 1 \). In the limit \( r_0 \to 0 \), (2.11) reduces to

\[
v[f] = \int_0^1 G(x, a, b)f(x, b, t) \, db + S(x, a, u_1(x, s), \ldots, u_{N-1}(x, s)). \tag{2.12}
\]

Since we aim at a minimal effective model, we avoid precise assumptions on the underlying biological processes expressed by \( K \).

The term \( S \geq 0 \) in (2.11) and (2.12) models the action of toxic A\(\beta\) oligomers, ultimately leading to apoptosis. For example

\[
S = C_S(1 - a)\left(\sum_{m=1}^{N-1} m u_m(x, s) - \overline{U}\right)^+
\]

The threshold \( \overline{U} > 0 \) indicates the minimal amount of toxic A\(\beta\) needed to damage neurons, assuming that the toxicity of soluble A\(\beta\)-polymers does not depend on \( m \). In reality length dependence has been
observed Ono et al. (2009), but, to our best knowledge, quantitative data are only available for very short molecules (see Ono et al., 2009, Table 2). For long molecules, any analytic expression would be arbitrary.

Since Aβ monomers are produced by neurons and the production increases if neurons are damaged, we choose in (2.5)

$$\mathcal{F} = \mathcal{F}(f) = C\int_0^1 (\mu_0 + a)(1 - a)f(x, a, t)\, da.$$  \hspace{1cm} (2.14)

The small constant $\mu_0 > 0$ accounts for Aβ production by healthy neurons (dead neurons do not produce amyloid).

To describe the onset of AD we assume that in small, randomly chosen parts of the cerebral tissue, concentrated for instance in the hippocampus, the degree of malfunctioning of neurons randomly jumps to higher values due to external agents or genetic factors. This leads to an additional term in the equation for $f$,

$$\partial_t f + \partial_a (fv[f]) = J[f],$$

where

$$J[f] = \eta \left( \int_0^1 P(t, a \rightarrow a)f(x, a, t)\, da - f(x, a, t) \right) \chi(x, t).$$ \hspace{1cm} (2.15)

$P(t, a \rightarrow a)$ is the probability to jump from state $a$ to state $a \in [0, 1]$ (obviously, $P(t, a \rightarrow a) = 0$ if $a < a$), $\chi(x, t)$ describes the random jump distribution, and $\eta$ is the jump frequency. In most of our numerical tests we choose

$$P(t, a \rightarrow a) \equiv P(a \rightarrow a) = \begin{cases} \frac{2}{1 - a} & \text{if } a \leq a \leq \frac{1 + a}{2} \\ 0 & \text{otherwise,} \end{cases}$$

that is we neglect randomness and we set $\chi(x, t) \equiv \chi(x)$ concentrated in the hippocampus. For a simulation with a random jump distribution, see Fig. 10.

To model the phagocytic activity of the microglia as well as other bulk clearance processes Iliff et al. (2012), we add to (2.1) and (2.5) a term $-\sigma_m u_m$, where $\sigma_m > 0$. This leads to the system

\[
\begin{cases}
\partial_t f + \partial_a (fv[f]) = J[f], \\
\varepsilon \partial_t u_1 = d_1 \nabla^2 u_1 - u_1 \sum_{j=1}^N a_{1,j} u_j + \mathcal{F}(f) - \sigma_1 u_1, \\
\varepsilon \partial_t u_m = d_m \nabla^2 u_m + \frac{1}{2} \sum_{j=1}^{m-1} a_{m-j} u_{m-j} u_m - u_m \sum_{j=1}^N a_{m,j} u_j - \sigma_m u_m \quad (2 \leq m < N), \\
\varepsilon \partial_t u_N = \frac{1}{2} \sum_{k,j=N}^{j+k=N} a_{j,k} u_j u_k, \quad (2 \leq m < N)
\end{cases}
\]

\hspace{1cm} (2.16)
where \( v[f] \) is given by (2.11) or (2.12) (with \( s \) replaced by \( \varepsilon^{-1} t \)), \( \mathcal{F}[f] \) by (2.14), and \( J[f] \) by (2.15). Since we are interested in longitudinal modelling, we assume that initially, at \( t = 0 \), the brain is healthy, with a small uniform distribution of soluble amyloid.

### 3. Problem setting and discretization of the equations

In this section, we detail the structure of the domain and the boundary conditions which we will use, in Section 4, to produce numerical simulations. We also discuss the discretization of the equations (2.16).

#### 3.1 Physical domain and boundary conditions

We consider the two-dimensional transverse section of the brain illustrated in Fig. 1. Since approximating a real brain section is a quite complicated issue, for the sake of simplicity we schematize it as a box \( \Omega \subset \mathbb{R}^2 \), \( \Omega = [0, L_x] \times [0, L_y] \), with two inner rectangular holes representing the sections of the cerebral ventricles. We also identify, close to the front part of the ventricles, the two sections of the hippocampus, which we represent as two small circles. Unlike the cerebral ventricles, the sections of the hippocampus are meant as actual portions of the domain \( \Omega \), not as holes.

On the outer boundary of \( \Omega \), say \( \partial \Omega_{\text{out}} \), we assume vanishing normal polymer flow. Therefore, we impose a homogeneous Neumann condition for the diffusing amyloid polymers:

\[
-d_m \varepsilon \nabla u_m \cdot \mathbf{n} = 0 \quad \text{on} \quad \partial \Omega_{\text{out}}, \quad m = 1, \ldots, N - 1,
\]  

(3.1)

![Fig. 1](image-url)  

**Fig. 1.** Left: A real transverse section of the brain (reproduced from Miller, 2006 with kind permission of the publisher). Right: Two-dimensional schematization for numerical purposes. Black dots are the internal nodes of the numerical grid, where discretized equations are solved, while white dots are boundary nodes, where boundary conditions are imposed.
\( n \) being the outward normal unit vector to \( \partial \Omega_{\text{out}} \). Notice that no boundary condition is required for the concentration \( u_N \) of the fibrillar amyloid, since its equation does not feature space dynamics (cf. the last equation in (2.16)).

On the inner boundary of \( \Omega \), say \( \partial \Omega_{\text{in}} \), that is the boundaries of the cerebral ventricles, we model the removal of \( \text{A}\beta \) from cerebrospinal fluid (CSF) through the choroid plexus (cf. Iliff et al., 2012; Serot et al., 2012) by an outward polymer flow proportional to the concentration of the amyloid. For this, we impose a Robin boundary condition of the form:

\[
- \frac{d_m}{\varepsilon} \nabla u_m \cdot n = \beta u_m \quad \text{on} \quad \partial \Omega_{\text{in}}, \quad m = 1, \ldots, N - 1,
\]

with \( \beta > 0 \) a constant.

We discretize \( \Omega \) by means of a two-dimensional structured orthogonal grid, whose points have coordinates \( x_{ij} = (x_i, y_j) = (i \Delta x, j \Delta y) \) with \( \Delta x = L_x/N_x, \Delta y = L_y/N_y \) being the numbers of discretization points in the \( x \) and \( y \)-direction, respectively, and \( i = 0, \ldots, N_x, j = 0, \ldots, N_y \). See Fig. 1.

We also introduce a time lattice \( t_n = n \Delta t, n = 0, 1, 2, \ldots \).

Letting \( (u_m)_{ij}^n \approx u_m(x_{ij}, t_n) \) denote an approximation of the concentration of the \( m \)-polymers of amyloid in the point \( x_{ij} \in \Omega \) at time \( t_n \), on the numerical grid the Neumann boundary condition (3.1) becomes simply:

\[
\begin{align*}
(u_m)_{0,j}^n &= (u_m)_{1,j}^n, \\
(u_m)_{N_x,j}^n &= (u_m)_{N_x-1,j}^n, \\
(u_m)_{i,0}^n &= (u_m)_{i,1}^n, \\
(u_m)_{i,N_y}^n &= (u_m)_{i,N_y-1}^n
\end{align*}
\]

Concerning the Robin boundary condition (3.2), we discretize the components of the gradient via the forward Euler formula, then we take into account the orientation of the vector \( n \) as indicated in Fig. 1 to find:

- along the left boundary of each cerebral ventricle

\[
(u_m)_{ib,j}^n = \frac{(u_m)_{ib-1,j}^n}{1 + \varepsilon \beta \Delta x / d_m},
\]

where \( ib \in \{0, \ldots, N_x\} \) denotes the grid index in the \( x \)-direction such that \( x_{ib} = i_b \Delta x \) is the abscissa of the boundary;

- along the right boundary of each cerebral ventricle

\[
(u_m)_{ib,j}^n = \frac{(u_m)_{ib+1,j}^n}{1 + \varepsilon \beta \Delta x / d_m};
\]

- along the lower boundary of each cerebral ventricle

\[
(u_m)_{ijb}^n = \frac{(u_m)_{ijb-1}^n}{1 + \varepsilon \beta \Delta y / d_m},
\]
where \( j_y \in \{0, \ldots, N_y\} \) denotes the grid index in the y-direction such that \( y_{j_y} = j_y \Delta y \) is the ordinate of the boundary;

- along the upper boundary of each cerebral ventricle

\[
(u_m)^n_{i,j_b} = \frac{(u_m)^n_{i,j_b+1}}{1 + \varepsilon \beta \Delta y / d_m}.
\]

### 3.2 Discretization of the Smoluchowski equations

In order to approximate the equations for the \( u_m \)'s, \( m = 1, \ldots, N - 1 \), we use a fractional step procedure in time: first we solve the diffusion and reaction parts, then we add the coagulation and possibly the source (for \( u_1 \) parts).

Adopting a Finite Difference discretization of the Laplace operator \( \nabla^2 \) we obtain the scheme:

\[
\begin{cases}
(u_m)^*_{i,j} = (u_m)^n_{i,j} + \frac{\Delta t}{\varepsilon} \left( d_m \frac{(u_m)^n_{i,j-1} - 2(u_m)^n_{i,j} + (u_m)^n_{i,j+1}}{\Delta x^2} + d_m \frac{(u_m)^n_{i,j-1} - 2(u_m)^n_{i,j} + (u_m)^n_{i,j+1}}{\Delta y^2} \right) - \sigma_m (u_m)^n_{i,j} \\
(u_m)^{n+1}_{i,j} = (u_m)^*_{i,j} + \frac{\Delta t}{\varepsilon} \left( \frac{1}{2} \sum_{h=1}^{m-1} a_{h,m-h}(u_h)^{n}_{i,j} (u_{m-h})^{n}_{i,j} - (u_m)^n_{i,j} \sum_{h=1}^{N} a_{m,h}(u_h)^{n}_{i,j} \right),
\end{cases}
\]

where \((u_m)^*_{i,j}\) denotes the temporary solution computed after the first fractional time step. For an alternative Finite Element discretization of Smoluchowski equations see e.g., Achdou et al. (2013).

The scheme above applies to all inner nodes \( x_{i,j} \) of the numerical grid (that means \( 1 \leq i \leq N_x - 1, 1 \leq j \leq N_y - 1 \) excluding furthermore the indexes \( i_b, j_b \) identifying the inner boundary \( \partial \Omega_m \) and to the A\(\beta \) \( m \)-polymers with \( m = 2, \ldots, N - 1 \). Because of the adopted approximation of the diffusion part, the following constraint on the time and space steps has to be enforced for the stability of the numerical scheme:

\[
\Delta t \leq \frac{\varepsilon}{4} \cdot \frac{\min\{\Delta x^2, \Delta y^2\}}{\max_{1 \leq m \leq N-1} d_m}.
\]

For \( m = 1 \) the equation is discretized in a similar way but for the addition of the source term \( F \). We refer the reader to the next subsection for discretization methods of the integral contained in it.

Finally, for \( m = N \) the equation is actually an ODE, which we approximate by the explicit Euler formula:

\[
(u_N)^{n+1}_{i,j} = (u_N)^n_{i,j} + \frac{\Delta t}{2\varepsilon} \sum_{h+k \geq N} a_{h,k}(u_h)^{n}_{i,j} (u_k)^{n}_{i,j}.
\]

### 3.3 Discretization of the equation for \( f \)

In the interval \([0, 1]\), which constitutes the domain of the variable \( a \), we introduce a Finite Volume partition made of \( N_a \) cells of the form \([a_{i-1/2}, a_{i+1/2}]\) with central point \( a_i = (k - \frac{1}{2}) \Delta a \), where \( \Delta a = \frac{1}{N_a} \). The cell index \( k \) runs from 1 to \( N_a \). Then we discretize the first equation in (2.16) using again a fractional step procedure in time.
First, we solve the homogeneous transport part by means of the push-forward scheme introduced by e.g. Piccoli & Tosin (2011); Tosin & Frasca (2011), which is particularly suited to deal with non-local fluxes. Denoting \( f_{i,j,k}^n \approx f(x_{ij}, a_k, t_n) \), we have:

\[
f_{i,j,k}^n = f_{i,j,k}^n - \frac{\Delta t}{\Delta a} \left( f_{i,j,k}^n |v_{i,j,k}^n| - f_{i,j,k-1}^n (v_{i,j,k-1}^n)^+ - f_{i,j,k+1}^n (v_{i,j,k+1}^n)^- \right)
\]

(3.4)

where \( v_{i,j,k}^n \approx v(x_{ij}, a_k, t_n) \) indicates an approximation of the deterioration rate of the neurons and \((\cdot)^-\) is the negative part \((x^- := \max\{0, -x\})\). Here, we compute \( v_{i,j,k}^n \) by approximating the integral contained in the expression (2.12) via a zeroth order Euler formula and then adding the expression (2.13):

\[
v_{i,j,k}^n = \sum_{h=1}^{N_a} G(x_{ij}, a_k, a_h) f_{i,j,h}^n \Delta a + C_S (1 - a_k) \left( \sum_{m=1}^{N-1} m(u_m)^n_{ij} - \mathcal{U} \right)^+.
\]

If the form \( G(x, a, b) = C_G (b - a)^+ \) is used then in the formula above, we simply have \( G(x_{ij}, a_k, a_h) = C_G (a_h - a_k)^+ \).

The stability of the scheme (3.4) requires that the grid steps \( \Delta a, \Delta t \) be linked by the following CFL condition:

\[
\Delta t \leq \frac{\Delta a}{\max |v_{i,j,k}^n|}.
\]

(3.5)

Second, we update the values \( f_{i,j,k}^n \) by including the jump process:

\[
f_{i,j,k}^{n+1} = f_{i,j,k}^n + \eta \Delta t \left( \sum_{h=1}^{N_a} P_k f_{i,j,h}^n \Delta a - f_{i,j,k}^n \right) \chi_{i,j}^n,
\]

where we have denoted \( P_k := P(a_h \to a_k) \) and \( \chi_{i,j}^n := \chi(x_{ij}, t_n) \).

### 3.4 Final choice of the time step

On the whole, the time step of the complete numerical scheme has to comply with both the parabolic and the hyperbolic constraints (3.3), (3.5), respectively. Therefore, it is ultimately chosen as:

\[
\Delta t \leq \min \left\{ \frac{\varepsilon}{4} \cdot \min \{\Delta x^2, \Delta y^2\} \cdot \frac{\Delta a}{\max_{i,j,k} |v_{i,j,k}^n|}, \frac{\Delta a}{\max_{i,j,k} d_m^{1,N-1}} \right\}
\]

at each time iteration of the numerical scheme.

### 3.5 Computing physiological indicators

Several macroscopic (aggregate) quantities can be computed out of the results of model (2.16). In Section 4, the outputs of the simulations will be discussed in terms of a few of such quantities, which can be directly compared with real clinical images and known qualitative time evolution of AD.
The macroscopic distribution of neuron malfunctioning \( A = A(x, t) \) is computed over the cerebral domain \( \Omega \) as the local average of the degree of malfunctioning \( a \):

\[
A(x, t) := \int_0^1 a f(x, a, t) \, da,
\]

which is numerically approximated as

\[
A(x_{ij}, t_n) \approx A_{ij}^n = \sum_{k=1}^{N_a} a_k f_{ij,k}^n.
\]

Following Jack et al. (2013), we relate then the ‘local brain atrophy’ \( \phi(x, t) \) to the average neuron malfunctioning \( A \) as:

\[
\phi(x, t) := \max \left\{ 0, \frac{A(x, t) - A_0}{1 - A_0} \right\},
\]

\( A_0 \in (0, 1) \) being a threshold of malfunctioning over which the brain is considered locally atrophic. The corresponding numerical approximation is

\[
\phi_{ij}^n = \max \left\{ 0, \frac{A_{ij}^n - A_0}{1 - A_0} \right\}.
\]

Next we define the ‘global brain atrophy’ in time \( \Phi = \Phi(t) \) as the average of \( \phi \) over the whole domain \( \Omega \), i.e.

\[
\Phi(t) := \frac{1}{|\Omega|} \int_{\Omega} \phi(x, t) \, dx,
\]

\(|\Omega|\) denoting the area of \( \Omega \), which is numerically approximated as

\[
\Phi^n := \frac{1}{|\Omega|} \sum_{i=0}^{N_x-1} \sum_{j=0}^{N_y-1} \phi_{ij}^n \Delta x \Delta y,
\]

In this formula, we conventionally consider \( \phi_{ij}^n = 0 \) if the grid point \( x_{ij} \) does not belong to the domain \( \Omega \), i.e. if it is a point inside the cerebral ventricles.

The ‘total concentration of soluble amyloid’ \( U_S = U_S(t) \) in the brain occipital region, to be related to the \( A\beta \) concentration found in the CSF by clinical exams (\( A\beta \)), is given by:

\[
U_S(t) := \frac{1}{|\Omega|} \int_{\tilde{\Omega}} \sum_{m=1}^{N-1} m \mu_m(x, t) \, dx,
\]

where \( \tilde{\Omega} \subset \Omega \) is a subdomain located in the bottom part of \( \Omega \), entirely contained in the region below the cerebral ventricles. Assuming for simplicity that it is a rectangle as well, whose grid coordinates are
comprised between two indexes $0 < \hat{i}_1 < \hat{i}_2 < N_x$ in the x-direction and between $j = 0$ and $j = \hat{j} > 0$ in the y-direction, we obtain the numerical values of $U_S$ as:

$$U^n_S := \frac{1}{|\Omega|} \sum_{i=\hat{i}_1}^{\hat{i}_2-1} \sum_{j=0}^{\hat{j}-1} \sum_{m=1}^{N_y-1} m(u_m)^n_{ij} \Delta x \Delta y = \frac{1}{(\hat{i}_2 - \hat{i}_1)\hat{j}} \sum_{i=\hat{i}_1}^{\hat{i}_2-1} \sum_{j=0}^{\hat{j}-1} \sum_{m=1}^{N_y-1} m(u_m)^n_{ij},$$

where we have used that $|\Omega| = (\hat{i}_2 - \hat{i}_1)\hat{j} \Delta x \Delta y$.

Finally, the ‘average quantity of brain Aβ deposits’ in time is:

$$U_N(t) := \frac{1}{|\Omega|} \int_{\Omega} N u_N(x, t) \, dx,$$

which is naturally discretized as

$$U_N(t_n) \approx U^n_N = \frac{1}{|\Omega|} \sum_{i=0}^{N_x-1} \sum_{j=0}^{N_y-1} N(u_N)^n_{ij} \Delta x \Delta y$$

by letting conventionally $(u_N)^n_{ij} = 0$ if $x_{ij} \notin \Omega$ (inside the cerebral ventricles).

4. Numerical results

To begin with, we provide a typical output of the numerical simulations. In Fig. 2, we plot the evolution of three crucial biomarkers of AD (as a function of the computational time):

- the CSF Aβ42 (purple dashed curve);
- the average quantity of brain Aβ42 deposits (red solid curve);
- the global brain atrophy (blue dash-dot curve).

All curves are normalized to their maxima. The values of the constants used in the simulation are specified in the figure caption.

The level of Aβ42-deposition (red solid curve) grows rapidly, reaches its maximum and then stabilizes. The purple dashed curve, corresponding to CSF-Aβ42, decreases after having reached a peak. The blue dash-dot curve corresponds to the brain atrophy and increases in time as expected. The graphs in Fig. 2 can be well illustrated by the following quote from Jack et al. (2010):

The initiating event in AD is related to abnormal processing of β-amyloid peptide, ultimately leading to formation of Aβ plaques in the brain. This process occurs while individuals are still cognitively normal. Biomarkers of brain β-amyloidosis are reductions in CSF Aβ42 and increased amyloid PET tracer retention. After a lag period, which varies from patient to patient, neuronal dysfunction and neurodegeneration become the dominant pathological processes. Biomarkers of neuronal injury and neurodegeneration are increased CSF tau and structural MRI measures of cerebral atrophy. Neurodegeneration is accompanied by synaptic dysfunction, which is indicated by decreased fluorodeoxyglucose uptake on PET. We propose a model that relates disease stage to AD biomarkers in which Aβ biomarkers become abnormal first before neurodegenerative biomarkers and cognitive
symptoms, and neurodegenerative biomarkers become abnormal later and correlate with clinical symptom severity.

The plots we obtain should be compared with the clinical graphs in Bateman et al. (2012), Jack et al. (2013), Yau et al. (2015), and with the data of Fleisher et al. (2012) and Reiman et al. (2012). For the reader’s convenience, we reproduce here a picture from Jack et al. (2013), see Fig. 3, and a picture from Yau et al. (2015), see Fig. 4.

There is a satisfactory agreement between the plots of the qualitative temporal behaviour of the biomarkers and those obtained from clinical data. Observe that not only the shapes of the curves are comparable (CSF Aβ corresponds to CSF Aβ_{42}, brain Aβ deposits correspond to Amyloid PET and global brain atrophy corresponds to MRI + FDG PET) but also the temporal order of the events is in good agreement with clinical data.

Obviously the details of the numerical output depend on the choice of the constants used in the mathematical model. Performing a considerable amount of numerical runs with different values of the constants in the model, we have reached the conclusion that, at least qualitatively, the behaviour of the solutions does not depend on the precise choice of those constants, as long as their variation is restricted to reasonable ranges. In other words, the values of the constants taken in Fig. 2 can be considered as an indication for the order of magnitude. For example, the longitudinal graphs of the biomarkers CFS-Aβ, brain Aβ deposits and brain atrophy are — in this sense — qualitatively stable under variations of C_S, C_F, C_G and α.

It is particularly instructive to consider the constants \( \bar{U} \) in (2.13) and \( \beta \) in (3.2). We recall that \( \bar{U} \) is a threshold value for the minimal amount of toxic Aβ necessary to damage neurons (see (2.13)). In
Fig. 2 we have used the value $\overline{U} = 0.1$, but if we make it considerably larger, e.g. $\overline{U} = 1$ (the remaining constants are unchanged), then the threshold becomes so high that the illness does not develop at all.

The constant $\beta$ enters the model through condition (3.2) at the boundary of the cerebral ventricles. Smaller values of $\beta$ mean that less $A\beta$ is removed from the CSF through the choroid plexus. Fig. 5 shows what happens if we change it into $\beta = 0.01$: the three curves are moved to the left and become steeper: the illness starts earlier and develops faster. Recalling that in Figs. 2 and 5 we have plotted values which are normalized with respect to their maximal values, one could wonder how the latter ones depend on $\beta$. It turns out that the maximal values of CSF $A\beta$ and the brain atrophy are essentially independent of $\beta$. The $A\beta$ deposits (the plaques), however, increase by a factor 6 if $\beta$ is changed from 1 to 0.01. This result
is compatible with our modelling Ansatz (in accordance with the medical literature) that plaques are not toxic (even healthy brains may contain plaques).

The comparison of the cases $\beta = 1$ and $\beta = 0.01$ becomes even clearer when we create spatial plots of $f$ and of the distribution and density of the cerebral plaques at fixed computational times $t = T$. The plots of $f$ at different times are meant to be compared with FDG-PET images (see e.g. Fleisher et al., 2012). More precisely, we take a schematic image of a transverse section of the brain and attribute different colours to those parts of the brain where probabilistically the level of malfunctioning lies in different ranges. As in the FDG-PET, the red corresponds to a healthy tissue. Here, AD originates only from the hippocampus and propagates, at the beginning, along privileged directions (such as those corresponding to denser neural bundles) mimicked by two triangles.

In Figs. 6 and 7 we compare plots of $f$ at, respectively, times $T = 34$ and $T = 52$ for the two different values of $\beta = 0.01$ and $\beta = 1$. Figures 6 and 7 do not only confirm the temporal acceleration of the development of the illness for smaller values of $\beta$ but also show that the spatial pattern and heterogeneity become less evident as $\beta$ becomes smaller. Since experimental data suggest a strong spatial heterogeneity of the illness, this could indicate the potential importance of the removal of $A\beta$ through the choroid plexus to slow down the temporal development of AD.

In Fig. 8, we plot the plaques’ distribution for the two different values of $\beta = 0.01$ and $\beta = 1$ and at $T = 52$. This figure confirms the strong increase of the plaques when $\beta$ becomes smaller.

We stress that, though our images represent ‘a mean value’ of brain activity instead of a single patient’s brain activity, still they show a good agreement with clinical neuroimaging: compare Figs. 6 and 7 with Fig. 9 below.
Looking for more realistic images, we have to take into account randomness of the spatial distribution of the sources of the disease. For example, we have performed some runs where the AD does not only originate from the hippocampus but also from several sources of Aβ randomly distributed in the occipital part of the brain. We report the outputs of such runs in Fig. 10. The random distributed sources appear as the small blue spots.
Fig. 8. Density of plaques for $\beta = 0.01$ (left), $\beta = 1$ (right), $T = 52$.

Fig. 9. FDG-PET images showing patterns of metabolic activity: an elderly individual with no dementia (left) and with AD (right). Reproduced from Miller (2006) with permission.

5. Discussion and future research directions

We have presented a new mathematical model for the onset and evolution of AD. The model is characterized by a high level of flexibility, which potentially allows one to simulate different modelling hypotheses and compare them with clinical data. In fact, the model provides a flexible tool to test in the future alternative hypotheses on the evolution of the disease. In this article, we have chosen some specific aspects of the illness, such as the aggregation, diffusion and removal of $A\beta$, the possible spread of neuronal damage in the neural pathway, and, to describe the onset of AD, a random neural deterioration mechanism.
Numerical simulations are compared with clinical data and, although oversimplified and restricted to a two-dimensional rectangular section of the brain, they are in good qualitative agreement with the spread of the illness in the brain at various stages of its evolution. In particular, our model captures the cerebral damage in the early stage of MCI.

There are multiple future research developments in quite different directions, each of which requires substantial research efforts. We mention some of them.

Further development of the model is needed and should be carefully guided by clinical data. The constants appearing in the equations should be well calibrated to optimize quantitative agreement with clinical data. Simulations should become more realistic, in a three-dimensional domain which matches the geometric characteristics of the brain.

The true challenge in AD research is a breakthrough which allows one to develop effective therapies to stop or slow down the evolution of AD, possibly in an early stage of the illness. Also effective mathematical models can give a contribution in this direction. For example, a certain sensibility of the numerical output to the value of the constant $\beta$ in (3.2), which models the removal of $A\beta$ through the choroid plexus, spontaneously leads to the question whether dialysis mechanisms can be introduced to enhance $A\beta$-removal artificially. Most probably, a serious answer to this question requires, in addition to a detailed comparison with clinical data, a more refined modelling of the removal which takes into account the transport of soluble $A\beta$ by the cerebral fluid.

Finally, some mathematical effort is necessary to check the mathematical correctness (well posedness) of the model.

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