Coexisting opportunities and challenges: In which scenarios can minimal/measurable residual disease play a role in advanced non-small cell lung cancer?

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Abstract

Curative therapy was not previously available for patients with advanced non-small cell lung cancer (NSCLC); thus, the concept of minimal/measurable (or molecular) residual disease (MRD) was not applicable to these patients. However, advances in targeted and immunotherapy have revolutionized the treatment landscape for patients with advanced NSCLC, with emerging evidence of long-term survival and even the hope of complete remission (CR) by imaging examination. The latest research shows that patients with oligometastatic lung cancer can benefit from local treatment. After removing the lesions, the choice of follow-up therapy and monitoring of the lesions could remain uncertain. MRD plays a role in identifying early-stage NSCLC patients with high risks of recurrence and determining adjuvant therapy after radical treatment. In recent years, evidence has been accumulating regarding the use of circulating cell-free tumor DNA (ctDNA) to assess MRD in solid tumors. This study discussed the possible applications of ctDNA-based MRD monitoring in advanced NSCLC and described the current challenges and unresolved problems in the application of MRD in advanced NSCLC.

Keywords: Minimal residual disease; non-small cell lung cancer; circulating tumor DNA

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Introduction

Lung cancer is a leading cause of cancer-related deaths worldwide (1). Approximately 75% of patients with non-small cell lung cancer (NSCLC) are diagnosed after the disease has reached an advanced and unresectable stage (2), at which point surgical resection is not a feasible option and the prognosis is very poor (3). Thus, there is an unmet medical need to develop strategies to improve the cure rate of patients with advanced lung cancer.

Minimal/measurable residual disease (MRD, also known as molecular residual disease), refers to the small number of cancer cells remaining in the body after cancer treatment that do not respond or are resistant to treatment, which cannot be detected by traditional imaging [including positron emission tomography/computed tomography (PET/CT)] or laboratory methods [microscopic observation of cells and/or tracking of abnormal serum protein markers (tumor biomarker) in the blood]. MRD in solid tumors is now referred to as measurable residual disease in the National Comprehensive Cancer Network (NCCN) guidelines (4). While these residual cancer cells may be small in number and do not cause any signs or symptoms at that time, they represent the continued existence of cancer and the possibility of clinical progression (5). In addition to describing the clinical significance of the ability to evaluate tumors, MRD can also describe the sensitivity of detection techniques and the understanding of tumors at the molecular level. Therefore, the blood-based assessment of MRD is desirable. Recent advances in fluid biopsies have allowed the ultrasensitive
detection of circulating tumor cells (CTCs) and circulating cell-free tumor DNA (ctDNA) derived from residual or occult micrometastatic lesions in patients with solid tumors. Data from several clinical studies have reported that CTCs and ctDNA following curative treatment strongly predict recurrence in multiple tumor types (6-8). In patients with distant metastasis, high levels of CTCs and ctDNA are also associated with a poor prognosis (9).

The shift from monitoring dominant metastatic lesions to monitoring MRD may change the way we manage patients with solid tumors. For patients with early-stage lung cancer, MRD has been widely used to evaluate the risk of recurrence and inform the choice of systemic adjuvant therapy after radical local treatment (RLT) (8,10). MRD is mainly used in advanced lung cancer to assess disease-free status (no evidence of disease, NED), meaning that no sign of residual tumor is found by using the existing examination methods after treatment, indicating that tumors that can be found at this stage have been “completely removed” from the patient’s body. For example, 1) advanced lung cancer patients with an effect evaluation of complete remission (CR) after chemotherapy; 2) oligometastatic disease (OMD) after surgery; and 3) no evidence of active disease based on existing imaging techniques.

**Advanced lung cancer patients with CR by imaging examination**

At present, the detection of disease recurrence mainly depends on traditional imaging examinations, including CT and PET. Although technological advances have greatly improved imaging performance in recent decades, there remain several limitations (11). The most crucial disadvantage of these imaging techniques is that they detect only space-occupying lesions but not MRDs. In recent years, the development of tyrosine kinase inhibitors (TKIs) for patients harboring certain molecular aberrations and immunotherapy, such as immune checkpoint inhibitors (ICIs), have revolutionized the treatment of advanced NSCLC. Patients with advanced NSCLC have achieved CR rates of 1%–7% after targeted therapy or immunotherapy (12-32) ([Table 1](#table1)). Notably, the 5-year follow-up data showed CR rates of patients who completed long-term immunotherapy (35 cycles/2 years) as high as 15% and 10% in the KEYNOTE-010 and KEYNOTE-024 studies, respectively (23,29). However, among patients who have achieved CR, monitoring for disease recurrence is a major clinical problem when lesions on imaging modalities have completely resolved. Clinical studies have indicated that treatment with programmed death-ligand 1 (PD-L1) blockade can produce durable responses in patients with NSCLC; however, the optimal treatment duration and the availability and duration of “drug holidays” remain unknown. Drug resistance is a major challenge for patients treated with targeted therapies. However, the monitoring of drug resistance when there are no visible lesions in imaging remains a challenge. Therefore, we urgently need a less invasive but more sensitive tool to accurately detect MRD to improve patient survival rates and quality of life.

Although research is lacking on MRD in patients with advanced lung cancer who have achieved CR, ctDNA-based MRD detection following curative-intent treatment has been widely proven to be a predictor of progression-free survival (PFS) in patients with advanced lung cancer. A prospective study showed that ctDNA analysis of long-term responders (PFS≥12 months) to PD-L1 blockade may differentiate those who will achieve ongoing benefit from those at risk of eventual progression. The best overall responses (BORs) according to the Response Evaluation Criteria in Solid Tumors (RECIST) criteria were CR (11 patients), partial response (PR, 15 patients), and stable disease (SD, 5 patients). At the surveillance time point, 27 of 31 patients had undetectable ctDNA, 25 (93%) of whom remained progression-free in the subsequent follow-up (range: 4.76–24.21 months). In contrast, all four patients with detectable ctDNA eventually progressed [Fisher P=0.0001; positive predictive value =1; 95% confidence interval (95% CI): 0.51–1; negative predictive value =0.93; 95% CI: 0.80–0.99] (33). Another prospective study showed that, among 65 patients with locally advanced NSCLC, freedom from progression 24 months after chemoradiation therapy (CRT) was much higher in the ctDNA MRD-negative group than that in the ctDNA MRD-positive group in both the consolidation ICI (87.5% vs. 0%, P<0.0001) and no consolidation ICI (100% vs. 0, P=0.0006) cohorts. Although the difference was not significant, these results suggested that patients with positive MRD may benefit from consolidation ICI (P=0.04), while patients with negative ctDNA after CRT may not (P=0.23). ctDNA MRD monitoring following curative-intent treatment strongly predicted recurrence, with a mean lead time of 4.1 months between ctDNA detection and radiographic progression (6). These results
suggested that ctDNA analysis may be helpful for personalized treatment and recurrence risk monitoring in patients who have achieved a radiographic CR after initial treatment.

**Advanced lung cancer patients with OMD**

OMD refers to a state with a limited number of metastatic sites, a concept first proposed by Hellman and Weichselbaum in 1995. This state is considered an intermediate status between locally advanced and widely metastatic phases (34). The definition of OMD remains controversial; most retrospective studies defined OMD as a synchronous metastasis based on M1b staging of the eighth edition of The International Association for the Study of Lung Cancer (IASLC) Classification (within 6 months of diagnosis) or up to three metastatic cerebral metastases (35); however, some are defined as no more than five metastases (36). Stage M1b is defined as the presence of one metastasis in a single organ; the median overall survival (mOS) of patients with stage M1b disease (11.4 months) is similar to that of patients with one metastasis to the contralateral lung (stage M1a) (11.8 months), whereas mOS of patients with multiple lesions in one other organ in addition to the primary site (7.0 months) is similar to that for plurimetastatic patients (6.2 months) (37). The European Organization of Research and Treatment of Cancer (EORTC) consensus defines OMD as up to five metastases and three organs (38). In contrast, the Canadian consensus defines OMD as three or fewer to five or fewer metastases and up to six extracranial lesions (39). Analysis of patients with metastatic NSCLC at all sites showed that approximately 30%-50% of the advanced lung cancers were oligometastatic at the initial diagnosis (36,40,41).

Growing evidence has been recognized in the European Society for Medical Oncology (ESMO) and NCCN guidelines, which recommend consideration of RLT to all visible disease sites as an option for selected patients with OMD (42-44).

A phase II prospective trial in which 49 patients with stage IV NSCLC were randomized to LCT arm (local consolidation therapy with radiotherapy/surgery) or MT/O arm (maintenance therapy or observation) after first-line systemic therapy. The result showed that the LCT arm prolonged the median PFS (14.2 months vs. 4.4 months, P=0.022) and OS (41.2 months vs. 17.0 months, P=0.017) (35).

In general, adjuvant treatment can be used for MRD or micrometastatic lesions that may not be detected by traditional imaging methods; thus, the combination of RLT (surgery and radiotherapy) and adjuvant treatment can effectively reduce local recurrence and improve survival rates. A single-arm study of locally ablative therapy followed by sequential pembrolizumab adjuvant therapy in patients with oligometastatic lung cancer showed that ICI adjuvant therapy significantly improved the median PFS (19.1 months) compared to the historical median PFS (6.6 months) (P=0.005) (45). As the existing routine evaluation methods for solid tumors do not consider the evaluation of MRD, adjuvant therapy may be used “blindly”, and its success or failure is retrospectively evaluated after years of follow-up. Improved methods to detect MRD and/or micrometastatic disease will help to identify patients who might benefit from adjuvant therapies and those who may not (8). The research on this topic has mainly focused on early-stage NSCLC. Studies have shown that detection of ctDNA MRD after radical treatment can reliably identify patients with final recurrence, with a sensitivity of 93% and a specificity of 96%, which is superior to those for standard imaging monitoring (10).

The concept of OMD points to a special patient population in whom RLT may provide a better prognosis than that for patients with widely metastatic NSCLC. However, unlike patients with early-stage lung cancer who undergo RLT, patients with OMD may have a greater tendency for subsequent metastasis. This holds new promise for the treatment of patients with oligometastatic lung cancer and is also accompanied by significant challenges. For example, the sequence of systemic therapy vs. local therapy remains controversial (46,47). There are no studies on predictive markers to help determine patients with OMD who may benefit from postoperative adjuvant therapy. However, existing studies have provided evidence that the combination of RLT and systemic therapy is safe in patients with OMD (40,48) and can prolong survival (41). MRD monitoring can provide information as part of a more individualized clinical decision-making process and can help to improve patient quality of life (49). For patients with MRD, systemic treatment may be considered following RLT. For patients without detected MRD, a “drug holiday” may be possible, with continued monitoring of MRD (Figure 1).

**Other lesions that cannot be evaluated by imaging**

Imaging examinations are routinely used for clinical cancer
staging and treatment response tracking. However, since gross disease is difficult to distinguish from inflammation or fibrosis changes induced by therapy (especially radiotherapy and immunotherapy) (50), it can be challenging to monitor the disease in following clinical scenarios such as cavities, fibrosis, and scars. Second, pseudopropgression may occur after immunotherapy (51). Third, while lesions remain stable, PET-CT may show no metabolic activity after treatment. Although the target lesions are still visible on imaging, the local response is considered complete in these cases, as these lesions may not be active (52).

The incidence of pseudopropgression in NSCLC is about 3%−5% (53,54). In these cases, the amount of ctDNA would decrease or remain stable, whereas the levels would increase in true progression, with a sensitivity of 90%–100% (55). ctDNA MRD can be used as a supplementary method in cases in which it is difficult to distinguish inflammatory changes or fibrosis from true progression using imaging methods.

**Challenges and future prospects**

Although MRD can be used to evaluate lesions beyond current imaging methods, which may change the treatment patterns of advanced lung cancer, some unresolved problems remain.

First, there is a lack of forward-looking experiments. While data published to date indicate that ctDNA MRD is a reliable prognostic biomarker, there is a lack of data supporting ctDNA as a predictive biomarker in advanced NSCLC. Some prospective cohort studies showed that ctDNA analysis of patients with NSCLC who achieved CR, PR, or SD ≥1 year from PD-L1 blockade may differentiate those who will achieve ongoing benefit from those at risk of eventual progression (33) and that consolidation ICI therapy improves outcomes for NSCLC patients with MRD detected after CRT (6), which provides preliminary evidence to support the predictive power of ctDNA testing. There is also a lack of studies on MRD in patients with advanced NSCLC. However, several clinical trials are currently underway to verify the role of ctDNA-guided MRD assessment in patients with early-stage NSCLC (NCT04585477, NCT04642469, NCT04367311, NCT03774758, and NCT04585490).

Second, there remain limitations in detection technology, including that for MRD, the application of next-generation sequencing (NGS), changes in ctDNA level, lack of a standardized platform, and so on. MRD detection and monitoring based on bone marrow samples is a well-established procedure for the management of hematological malignancies (56). However, it is challenging in patients with solid tumors, as the sampling of tissue that might contain disseminated tumor cells (DTCs) (usually not located in the bone marrow fluid), such as post-treatment lung lobar specimens, is too invasive. Recent advances in detecting MRD with CTCs and ctDNA have identified alternative strategies to identify patients at high risk of relapse who instead require adjuvant therapy (8,10,57). ctDNA is more commonly used in clinical research to monitor the MRD of solid tumors, as it is easier to separate, more stable, and higher in proportion in the bloodstream, with higher sensitivity than that for CTCs (6,58). The current laboratory techniques and ctDNA
Table 1 CR rates of patients with advanced NSCLC in major clinical trials

| Variables | Clinical trial | Phase | Treatment | Patients | CR rate of target lesions [% (n/N)] | ORR [% (n/N)] | Reference |
|-----------|----------------|-------|-----------|----------|------------------------------------|---------------|-----------|
| **Target therapy** | | | | | | | |
| **First-line** | | | | | | | |
| | FLAURA (NCT02296125) | III | Osimertinib | EGFR mutation | 3 (7/279) | 80 (223/279) | (12) |
| | NEJ009 (UMIN000006340) | III | Gefitinib vs. Gefitinib + Carboplatin | EGFR mutation | 3 (5/173) vs. 4 (7/172) | 67 (116/173) vs. 84 (144/172) | (13) |
| | NEJ026 (UMIN000017069) | III | Erlotinib + Bevacizumab vs. Erlotinib | EGFR mutation | 7 (8/112) vs. 4 (4/112) | 72 (81/112) vs. 66 (74/112) | (14) |
| | ARCHER 1050 (NCT01774721) | III | Dacomitinib vs. Geftinib | EGFR mutation | 5 (12/227) vs. 2 (4/225) | 75 (170/227) vs. 72 (161/225) | (15) |
| **Second-line and above** | | | | | | | |
| | AURA3 (NCT02151981) | III | Osimertinib | EGFR mutation | 1 (4/279) | 71 (198/279) | (16) |
| | ARROW (Chinese subgroup) (NCT03037385) | I/II | Pralsetinib (BLU-667) | RET mutation | 3 (1/32) | 56 (18/32) | (18) |
| | LIBRETTO-001 (NCT03157128) | I/II | Selpercatinib | RET mutation | 2 (2/105) | 64 (67/105) | (19) |
| | CodeBreak 100 (NCT03600883) | II | Sotorasib | KRAS G12C mutation NSCLC | 2 (3/124) | 37.1 (46/124) | (20) |
| | NCT02122913 and NAVIGATE (NCT02576431) | I | Larotrectinib | NTRK mutation | 15 (2/13) | 77 (10/13) | (21) |
| | CRYSTALS (NCT02609776) | I | Amivantamab (JNJ-6372) | EGFR-Exon20ins mutation | 3 (4/81) | 40 (32/81) | (22) |
| **Immunotherapy** | | | | | | | |
| **First-line** | | | | | | | |
| | KEYNOTE-024 (NCT02122913) | III | Pembrolizumab vs. Chemotherapy | PD-L1 + (TPS≥ 50%) advanced NSCLC | – | 44.8 (69/154) vs. 27.8 (42/151) | (23) |
| | KEYNOTE-024 (5-year follow-up data) (NCT02122913) | III | Pembrolizumab | Up to 35 cycles/2 years of pembrolizumab | 10 (4/39) | 82 (32/39) | (24) |
| | KEYNOTE-042 (NCT02220894) | III | Pembrolizumab | PD-L1 + (TPS ≥ 1%) NSCLC | – | 39 (118/299) | (25) |
| | KEYNOTE-799 (NCT03631784) | II | Pembrolizumab + Concurrent chemoradiotherapy | IIIA-C NSCLC | 4 (7/13) | 70 (121/173) | (26) |
| | CheckMate-227 (NCT02477826) | III | Nivolumab + Ipilimumab | PD-L1 + (TPS≥ 1%) | 5.8 (23/396) | 36 (143/396) | (27) |
| | IMpower130 (NCT02367781) | III | Atezolizumab + Chemotherapy | IV NSCLC | 5 (22/453) | 46 (207/453) | (28) |
| | IMpower133 | I/III | Atezolizumab + EP | ES-SCLC | 2.5 (5/201) | 60.2 (121/201) | (29) |
| **Second-line and above** | | | | | | | |
| | KEYNOTE-010 (5-year follow-up data) (NCT02576431) | III | Pembrolizumab | Up to 35 cycles/2 years of pembrolizumab | 15 (12/79) | 94.9 (75/79) | (30) |
| | CheckMate-017 (NCT01642004) | III | Nivolumab | Metastatic squamous cell lung cancer | 0.7 (1/135) | 20 (27/135) | (31) |
| | CheckMate-057 (NCT01673867) | III | Nivolumab | Metastatic non-squamous NSCLC | 1.4 (4/292) | 19 (56/292) | (32) |
| | CheckMate-032 (NCT01928394) | I/II | Nivolumab | Recurrent SCLC | 0.9 (1/109) | 12 (13/109) | (33) |

CR, complete remission; NSCLC, non-small cell lung cancer; EGFR, epidermal growth factor receptor; RET, ret proto-oncogene; KRAS, Kirsten rat sarcoma viral oncogene homolog; PD-L1, programmed death-ligand 1; TPS, tumor proportion score; ES, extensive stage; ORR, overall response rate.
analysis include sequencing, polymerase chain reaction (PCR), and NGS (59). Increasing observable events (variables) (60) and combined methylation analysis methods can improve the sensitivity of MRD detection (61).

In summary, although the definition of MRD for solid cancer is imperfect, the ctDNA detection criteria are uncertain, and there remain many problems to be solved in MRD itself, it is a feasible scientific hypothesis to guide the systemic treatment of locally advanced or advanced lung cancer, as shown in Table 1. For patients who cannot undergo surgery but achieve radiographic CR after treatment (chemoradiation, target treatment, and/or immune treatment), MRD surveillance can help to determine the need for maintenance therapy and select the population that will benefit from consolidation therapy with ICI. The use of adjuvant therapy guided by MRD monitoring after local therapy for OMD can reduce the treatment burden for patients without detectable MRD and allow patients to enjoy “drug holidays”. Regarding the challenges caused by cavity, fibrosis, scar formation, and false progression caused by emerging treatment forms such as immunotherapy to traditional imaging evaluation, the detection of ctDNA MRD can also be used to help determine the prognosis and formulate further treatment strategies. Monitoring ctDNA MRD mutations is currently used to detect treatment response, emergence of resistance, and metastatic progression (62). Therefore, research is needed on MRD-based treatment strategies in patients with locally advanced/advanced lung cancer to provide accurate consolidation treatment plans and extend the duration of CR.

The emergence of new therapies provides realistic hope for the effective treatment, or even cure, of patients with advanced lung cancer. Whether disease assessment through MRD can change our understanding of advanced lung cancer and if it is possible to determine the prognosis and precise intervention for advanced lung cancer based on MRD detection to change the treatment strategy for advanced lung cancer remain to be determined. We look forward to the design of relevant clinical studies to answer these questions.

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Footnote

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References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. CA Cancer J Clin 2020;70:7-30.
2. Moeller B, Balagamwala EH, Chen A, et al. Palliative thoracic radiation therapy for non-small cell lung cancer: 2018 Update of an American Society for Radiation Oncology (ASTRO) Evidence-Based Guideline. Pract Radiat Oncol 2018;8:245-50.
3. Qin K, Hou H, Liang Y, et al. Prognostic value of TP53 concurrent mutations for EGFR-TKIs and ALK-TKIs based targeted therapy in advanced non-small cell lung cancer: a meta-analysis. BMC Cancer 2020;20:328.
4. Benson AB, Venook AP, Al-Hawary MM, et al. Colon Cancer, Version 2. 2021, NCCN Clinical Practice Guidelines in Oncology. J Natl Compr Canc Netw 2021;19:329-59.
5. Pantel K, Alix-Panabières C. Tumour microenvironment: informing on minimal residual disease in solid tumours. Nat Rev Clin Oncol 2017;14:325-6.
6. Moding EJ, Liu Y, Nabet BY, et al. Circulating tumor DNA dynamics predict benefit from consolidation immunotherapy in locally advanced non-small-cell lung cancer. Nat Cancer 2020;1:176-83.
7. Reinert T, Scholer LV, Thomsen R, et al. Analysis of circulating tumour DNA to monitor disease burden following colorectal cancer surgery. Gut 2016;65:625-34.
8. Wu C, Lee C, Wu C, et al. Circulating tumor cells as a tool of minimal residual disease can predict lung cancer recurrence: A longitudinal, prospective trial. Diagnostics (Basel) 2020;10:144.
9. Nieva J, Wendel M, Luttgen M, et al. High-definition imaging of circulating tumor cells and associated cellular events in non-small cell lung cancer patients: a longitudinal analysis. Phys Biol 2012;9:016004.
10. Chaudhuri AA, Chabon JJ, Lovejoy AF, et al. Early detection of molecular residual disease in localized lung cancer by circulating tumor DNA profiling. Cancer Discov 2017;7:1394-403.
11. Pieterman RM, van Putten JW, Meuzelaar JJ, et al. Preoperative staging of non-small-cell lung cancer...
with positron-emission tomography. N Engl J Med 2000;343:254-61.
12. Soria JC, Ohe Y, Vansteenkiste J, et al. Osimertinib in untreated EGFR-mutated advanced non-small-cell lung cancer. N Engl J Med 2018;378:113-25.
13. Hosomi Y, Morita S, Sugawara S, et al. Gefitinib alone versus gefitinib plus chemotherapy for non-small-cell lung cancer with mutated epidermal growth factor receptor: NEJ009 study. J Clin Oncol 2020; 38:115-23.
14. Saito H, Fukuhara T, Furuya N, et al. Erlotinib plus bevacizumab versus erlotinib alone in patients with EGFR-positive advanced non-squamous non-small-cell lung cancer (NEJ026): interim analysis of an open-label, randomised, multicentre, phase 3 trial. Lancet Oncol 2019;20:625-35.
15. Wu YL, Cheng Y, Zhou X, et al. Dacomitinib versus gefitinib as first-line treatment for patients with EGFR-mutation-positive non-small-cell lung cancer (ARCHER 1050): a randomised, open-label, phase 3 trial. Lancet Oncol 2017;18:1454-66.
16. Papadimitrakopoulou V, Wu Y, Ahn MJ, et al. Randomized phase III study of osimertinib vs platinum-pemetrexed for EGFR T790M-positive advanced NSCLC (AURA3). J Thorac Oncol 2017;12:S5-S6.
17. Gainor JF, Curigliano G, Kim DW, et al. Registrational dataset from the phase I/II ARROW trial of pralsetinib (BLU-667) in patients (pts) with advanced RET fusion+ non-small-cell lung cancer (NSCLC). J Clin Oncol 2020;38(15 suppl):9515.
18. Drilon A, Oxnard GR, Tan DSW, et al. Efficacy of selpercatinib in RET fusion-positive non-small-cell lung cancer. N Engl J Med 2020;383:813-24.
19. Hong DS, Kuo J, Sacher AG, et al. CodeBreak 100: Phase I study of AMG 510, a novel KRAS G12C inhibitor, in patients (pts) with advanced solid tumors other than non-small cell lung cancer (NSCLC) and colorectal cancer (CRC). J Clin Oncol 2020;38 (15 suppl):3511.
20. Roth JA, Carlson JJ, Xia F, et al. The potential long-term comparative effectiveness of larotrectinib and entrectinib for second-line treatment of TRK fusion-positive metastatic lung cancer. J Manag Care Spec Pharm 2020;26:981-6.
21. Hong DS, Fakih MG, Strickler JH, et al. KRAS G12C inhibition with sotorasib in advanced solid tumors. N Engl J Med 2020;383:1207-17.
22. Reck M, Rodriguez-Abreu D, Robinson A, et al. Updated analysis of KEYNOTE-024: Pembrolizumab versus platinum-based chemotherapy for advanced non-small-cell lung cancer with PD-L1 tumor proportion score of 50% or greater. J Clin Oncol 2019;37:537-46.
23. Brahmer J, Rodriguez-Abreu D, Robinson AG, et al. LBA51 KEYNOTE-024 5-year OS update: First-line (1L) pembrolizumab (pembro) vs platinum-based chemotherapy (chemo) in patients (pts) with metastatic NSCLC and PD-L1 tumour proportion score (TPS) ≥50%. Ann Oncol 2020;31:S1181-2.
24. Wu YL, Zhang L, Fan Y, et al. Randomized clinical trial of pembrolizumab vs chemotherapy for previously untreated Chinese patients with PD-L1-positive locally advanced or metastatic non-small-cell lung cancer: KEYNOTE-042 China Study. Int J Cancer 2021;148:2313-20.
25. Jabbour SK, Park K, Cohn D, et al. Phase 2 trial of first-line pembrolizumab with platinum doublet chemotherapy and radiotherapy in patients (pts) with unresectable, locally advanced stage III non-small-cell lung cancer (NSCLC): KEYNOTE-799. J Clin Oncol 2019;37(15 suppl):TPS8575.
26. Hellmann MD, Ciuleanu TE, Pluzanski A, et al. Nivolumab plus ipilimumab in lung cancer with a high tumor mutational burden. N Engl J Med 2018;378:2093-104.
27. West H, McCleod M, Hussein M, et al. Atezolizumab in combination with carboplatin plus nab-paclitaxel chemotherapy compared with chemotherapy alone as first-line treatment for metastatic non-squamous non-small-cell lung cancer (IMpower130): a multicentre, randomised, open-label, phase 3 trial. Lancet Oncol 2019;20:924-37.
28. Horn L, Mansfield AS, Szczesna A, et al. First-line atezolizumab plus chemotherapy in extensive-stage small-cell lung cancer. N Engl J Med 2018;379:2220-9.
29. Herbst RS, Garon EB, Kim DW, et al. Long-term outcomes and retreatment among patients with previously treated, programmed death-Ligand 1-positive, advanced non-small-cell lung cancer in the KEYNOTE-010 study. J Clin Oncol 2020;38:1580-90.
30. Brahmer J, Reckamp KL, Baas P, et al. Nivolumab
versus docetaxel in advanced squamous-cell non-small-cell lung cancer. N Engl J Med 2015;373:123-35.

31. Borghaei H, Brahmer J, Horn L, et al. Nivolumab vs docetaxel in advanced NSCLC: CheckMate 017/057 2-Y update and exploratory cytokine profile analysis: Track: Immunotherapy. J Thorac Oncol 2016;11 (10 suppl):S237-8.

32. Ready N, Farago AF, de Braud F, et al. Third-line nivolumab monotherapy in recurrent SCLC: CheckMate 032. J Thorac Oncol 2019;14:237-44.

33. Hellmann MD, Nabet BY, Rizvi H, et al. Circulating tumor DNA analysis to assess risk of progression after long-term response to PD-(L)1 blockade in NSCLC. Clin Cancer Res 2020;26:2849-58.

34. Hellman S, Weichselbaum RR. Oligometastases. J Clin Oncol 1995;13:8-10.

35. Gomez DR, Tang C, Zhang J, et al. Local consolidative therapy vs. maintenance therapy or observation for patients with oligometastatic non-small-cell lung cancer: Long-term results of a multi-institutional, phase II, randomized study. J Clin Oncol 2019;37:1558-65.

36. Miyawaki T, Wakuda K, Kenmotsu H, et al. Proposing synchronous oligometastatic non-small-cell lung cancer based on progression after first-line systemic therapy. Cancer Sci 2021;112:359-68.

37. Eberhardt WE, Mitchell A, Crowley J, et al. The IASLC lung cancer staging project: Proposals for the revision of the M descriptors in the forthcoming eighth edition of the TNM classification of lung cancer. J Thorac Oncol 2015;10:1515-22.

38. Dingemans AC, Hendriks LEL, Berghmans T, et al. Definition of synchronous oligometastatic non-small cell lung cancer — A consensus report. J Thorac Oncol 2019;14:2109-19.

39. Laurie SA, Banerji S, Blais N, et al. Canadian consensus: oligoprogressive, pseudoprogressive, and oligometastatic non-small-cell lung cancer. Curr Oncol 2019;26:e81-e93.

40. Spaggiari L, Bertolaccini L, Facciolo F, et al. A risk stratification scheme for synchronous oligometastatic non-small cell lung cancer developed by a multicentre analysis. Lung Cancer 2021;154:29-35.

41. Gauvin C, Krishnan V, Kaci I, et al. Survival impact of aggressive treatment and PD-L1 expression in oligometastatic NSCLC. Curr Oncol 2021;28:593-605.

42. Planchard D, Popat S, Kerr K, et al. Metastatic non-small cell lung cancer: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. Ann Oncol 2018;29(suppl 4):iv192-iv237.

43. Ettinger DS, Wood DE, Aisner DL, et al. Non-Small Cell Lung Cancer, Version 5. 2017, NCCN Clinical Practice Guidelines in Oncology. J Natl Compr Canc Netw 2017;15:504-35.

44. Kozower BD, Larner JM, Detterbeck FC, et al. Special treatment issues in non-small cell lung cancer: Diagnosis and management of lung cancer, 3rd ed: American College of Chest Physicians evidence-based clinical practice guidelines. Chest 2013;143(5 suppl): e369S-e99S.

45. Bauml JM, Mick R, Ciunci C, et al. Pembrolizumab after completion of locally ablative therapy for oligometastatic non-small cell lung cancer: A phase 2 trial. JAMA Oncol 2019;5:1283-90.

46. Couñago F, Luna J, Guerrero LL, et al. Management of oligometastatic non-small cell lung cancer patients: Current controversies and future directions. World J Clin Oncol 2019;10:318-39.

47. Magnuson WJ, Lester-Coll NH, Wu AJ, et al. Management of brain metastases in tyrosine kinase inhibitor — naïve epidermal growth factor receptor — mutant non-small-cell lung cancer: A retrospective multi-institutional analysis. J Clin Oncol 2017;35:1070-7.

48. Blake-Cerda M, Lozano-Ruíz F, Maldonado-Magos F, et al. Consolidative stereotactic ablative radiotherapy (SABR) to intrapulmonary lesions is associated with prolonged progression-free survival and overall survival in oligometastatic NSCLC patients: A prospective phase 2 study. Lung Cancer 2021;152:119-26.

49. Villaruz LC, Kubicek GJ, Socinski MA. Management of non-small cell lung cancer with oligometastasis. Curr Oncol Rep 2012;14:333-41.

50. Zaheer SN, Whitley JM, Thomas PA, et al. Would you bet on PET? Evaluation of the significance of positive PET scan results post-microwave ablation for non-small cell lung cancer. J Med Imaging Radiat Oncol 2015;59:702-12.

51. Beer L, Hochmair M, Prosch H. Pitfalls in the radiological response assessment of immunotherapy. Memo 2018;11:138-43.
52. Dewas S, Bibault JE, Mirabel X, et al. Prognostic factors affecting local control of hepatic tumors treated by Stereotactic Body Radiation Therapy. Radiat Oncol 2012;7:166.

53. Fujimoto D, Yoshioka H, Kataoka Y, et al. Pseudoprogression in previously treated patients with non-small cell lung cancer who received nivolumab monotherapy. J Thorac Oncol 2019;14:468-74.

54. Ferrara R, Mezquita L, Texier M, et al. Hyper-progressive disease in patients with advanced non-small cell lung cancer treated with PD-1/PD-L1 inhibitors or with single-agent chemotherapy. JAMA Oncol 2018;4:1543-52.

55. Lee JH, Long GV, Menzies AM, et al. Association Between circulating tumor DNA and pseudoprogression in patients with metastatic melanoma treated with anti-programmed cell death 1 antibodies. JAMA Oncol 2018;4:717-21.

56. Radich J, Yeung C, Wu D. New approaches to molecular monitoring in CML (and other diseases). Blood 2019;134:1578-84.

57. Coakley M, Garcia-Murillas I, Turner NC. Molecular residual disease and adjuvant trial design in solid tumors. Clin Cancer Res 2019;25:6026-34.

58. Ng C, Di Costanzo G, Tosti N, et al. Genetic profiling using plasma-derived cell-free DNA in therapy-naïve hepatocellular carcinoma patients: a pilot study. Ann Oncol 2018;29:1286-91.

59. Gauri S, Ahmad M. ctDNA detection in microfluidic platform: A promising biomarker for personalized cancer chemotherapy. J Sensors 2020;2020:1-10.

60. Abbosh C, Birkbak NJ, Wilson GA, et al. Phylogenetic ctDNA analysis depicts early-stage lung cancer evolution. Nature 2017;545:446-51.

61. Liu MC, Oxnard GR, Klein EA, et al. Sensitive and specific multi-cancer detection and localization using methylation signatures in cell-free DNA. Ann Oncol 2020;31:745-59.

62. Kilgour E, Rothwell D, Brady G, et al. Liquid biopsy-based biomarkers of treatment response and resistance. Cancer Cell 2020;37:485-95.