Cerebrospinal fluid–suppressed T2-weighted MR imaging at 7 T for human brain

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Purpose: T2-weighted lesional imaging is most commonly performed using inversion recovery turbo spin echoes. At 7 T, however, this acquisition is limited for specific absorption rate and resolution. This work describes and implements a method to generate CSF-suppressed T2-weighted imaging.

Methods: The strategy uses a driven equilibrium spin-echo preparation within an inversion recovery with multiple 3D gradient-echo imaging blocks. Images are combined using the self-normalization approach, which achieves CSF suppression through optimized timing of individual blocks and minimizes sources of variation due to coil receptivity, T2*, and proton density. Simulations of the magnetization-prepared fluid-attenuated inversion recovery gradient-echo (MPFLAGRE) method over T1 and T2 relaxation values are performed, and in vivo demonstrations using an 8 × 2 transceiver array in healthy controls are shown.

Results: The specific absorption rate of the calculated MPFLAGRE sequence is 11.1 ± 0.5 W (n = 5 volunteers), which is 74 ± 2% of the US Food and Drug Administration guidelines. This method acquires both contrasts for CSF suppression with detection of long T2 components and T2-weighted imaging in a single acquisition. In healthy controls, the former contrast generates increased signal in the cortical rim and ependyma. A comparison is shown with a conventional 3D SPACE fluid-attenuated inversion recovery gradient-echo acquisition, and sensitivity to pathology is demonstrated in an epilepsy patient.

Conclusion: As applied with the 8 × 2 transceiver, the MPFLAGRE sequence generates both whole-brain contrast suitable for lesional and T2-weighted imaging at 7 T in fewer than 10 minutes within the US Food and Drug Administration’s specific absorption rate guidelines.

KEYWORDS
7T, FLAIR, MP2RAGE, SAR, T2 weighted

1 | INTRODUCTION

The increase in SNR at 7 T relative to 3 T has been used by numerous groups for higher-accuracy physiological and metabolic MRI. However, although high-resolution T1-weighted structural imaging has been applied quickly, T2-weighted and fluid attenuated inversion recovery (FLAIR) imaging at 7 T has been less implemented—most likely
because of the increased power deposition and poor transmission homogeneity due to the need for multiple spin echoes. Parallel transmit arrays have mitigated inhomogeneity problems; for example, RF shimming and a dual-row transmitter array can achieve 15% SD $B_1^+ \text{ over the human brain}$ (Supporting Information Figures S1 and S2). However, given the squared dependence of voltage with frequency at 7 T, turbo spin-echo acquisitions or the use of adiabatic pulses still result in high specific absorption rate (SAR), such that $T_2^*$-weighted imaging covering the entire brain in acceptable scan times remains challenging. The variable flip angle turbo spin-echo sequence provides a reasonable solution to control power deposition; however, it is sensitive to $B_1^+$ inhomogeneity, with several groups proposing design of parallel transmit pulses as a function of k-space position to generate more homogeneous excitation and inversion profiles. With this approach for turbo spin echo, however, achieving consistent contrast and motion insensitivity can be problematic by virtue of the dependence on accurate $B_1^+$ maps and applied $B_1$ variation between k-points. Alternatively, to altogether avoid multiple refocusing pulses, a driven equilibrium spin-echo strategy to longitudinally encode $T_2$ contrast (i.e., $T_2^*$ preparation) has been useful. To reduce the sensitivity of the acquisition to $B_1^+$ inhomogeneity at 7 T, Dyvorne and Balchandani implemented the $T_2$ preparation with an adiabatic spin-echo module in a multislab approach to generate excellent $T_2$-weighted whole-brain coverage at 0.8-mm$^3$ isotropic resolution (approximate 5.5-minute acquisition). The FLAIR imaging at 7 T, however, remains challenging.

To suppress the CSF signal, the now commonly used calculated MP2RAGE images from Marques provides an insightful $T_1$-based approach. This self-correcting normalization combines 2 images acquired at different delays after an inversion to give a calculated signal within the range of $[-0.5, 0.5]$. With a reconstruction in the complex domain (Eq. 1), the calculated signal retains sensitivity to the inversion recovery, while eliminating $B_0$-based $T_2^*$ phase effects and correcting for common factors of $M_0$ and $B_1^+$. Inspection of this reconstruction shows that the largest signal (+0.5) is seen when the intensities between the 2 images are equal, and the smallest signal (−0.5) when the intensities are equal and inverted in sign. For CSF, the calculated signal returns in the negative range (−0.5 to 0), whereas the white matter (WM) and gray matter (GM) return in the positive range (0 to +0.5). In the MP2RAGE acquisition, the sign and timings of the individual imaging readouts are optimized to generate a suppressed CSF intensity of −0.5 and high contrast between WM and GM. The equation of self-normalization reconstruction is

$$R = \text{real} \left( \frac{S_1^* S_2}{|S_1|^2 + |S_2|^2} \right)$$

where $S_1$ and $S_2$ are the GRE signal at inversion recovery delays T11 and T12, respectively; and the “$*$” operator is a complex conjugator.

As discussed by O’Brien, the MP2RAGE reconstruction provides a much flatter $T_1$-weighted image (than the non-reference-corrected MPRAGE), which enables high-consistency gray-white-CSF segmentation. More importantly, however, it is recognized that the self-correcting strategy can be applied to other acquisitions that acquire multiple readout blocks with a common preparation sequence.

In this report we describe the incorporation of a longitudinal $T_2$ preparation module with a multiblock inversion recovery 3D acquisition to achieve a CSF-suppressed $T_2^*$-weighted image for 7T use in the detection of brain pathology. Our preliminary data demonstrate that the proposed sequence MPFLAGRE (magnetization-prepared fluid attenuated gradient echo) achieves this goal. The sequence uses an inversion recovery with a $T_2$ spin-echo preparation and multiple short gradient-echo imaging blocks, with the self-correcting normalization generating the calculated MPFLAGRE image. Cerebrospinal fluid suppression is achieved through appropriate timing of TI for $T_1$ weighting. In this report, we show Bloch simulations to examine the sequence’s dependence on $T_1$ and $T_2$ and demonstrate its performance with a transceiver array at 7 T. Because of the longitudinal $T_2$ preparation that is performed with conventional adiabatic refocusing pulses, this sequence efficiently generates whole-brain $T_1$ and $T_2^*$-weighted coverage in single sequence that is well within SAR guidelines.

2.1 Two-block sequence (MPFLAGRE-2)

To generate $T_2$ FLAIR contrast, our method introduces $T_2$ weighting into the $T_1$-weighted MP2RAGE sequence using a longitudinal $T_2$ encoding module performed after the initial inversion (Figure 1). The $T_2$ weighting is performed with a nonselective spin-echo module (90°-180°-180°-90° of duration TE) and is followed by multiple short 3D gradient-echo (readout) blocks. The MPFLAGRE-2 uses 2 blocks, with the first block performed immediately after the spin echo, block $S_1$. Another signal block, $S_2$, is performed at another timepoint in the $T_1$ recovery, which with increasing delay from the $S_1$ block, reflects primarily $T_1$ weighting. Equations (2) and (3) give the expressions for the signal for the 2-block sequence:

$$S_1 \propto M_0 e^{-i\pi/|T_2|} B_1^+ sin\alpha_1$$

$$S_2 \propto \left[ \frac{e^{-i\pi/2M_0}}{\mu M_0} \sin M_0 + \left( 1 - e^{-i\pi/2M_0} \right) \right] A_{\text{center}} R_{\text{center}}$$

$$S = \text{real} \left( \frac{S_1^* S_2}{|S_1|^2 + |S_2|^2} \right)$$
S_x \propto M_0 e^{-\pi t_1/T_2} B_1 \sin \alpha_z \left\{ \left[1 - e^{-\pi t_2/T_2} + e^{-\pi t_2/T_2} R_1^{\text{Nesp}} \right] e^{-\pi t_1/T_1} + e^{-\pi t_1/T_1} A_1^{\text{Nesp}} \left[1 - e^{-\pi t_1/T_1} - \frac{M_0}{M_x} e^{-\pi t_1/T_1} \right] \right\}

\text{where } R_1^{\text{Nesp}} = \left(1 - e^{-\pi \frac{t_1}{T_1}}\right) \frac{1 - (\cos a_{\text{esp}}) e^{-\pi t_1/T_1}}{1 - \cos (a_{\text{esp}}) e^{-\pi t_1/T_1}},

A_1^{\text{Nesp}} = \left(1 - e^{-\pi \frac{t_1}{T_1}}\right) \left(1 - \cos a_{\text{esp}} e^{-\pi t_1/T_1}\right)^{N_{\text{esp}}},

\frac{R_1}{R_{0.5}} = \text{real} \left( \frac{-S_1 S_2}{|S_1|^2 + |S_2|^2} \right). \tag{4}

The normalization from Eq. (4) for the 2 signals gives a high 0.5 value when the 2 input intensities, \(S_1\) and \(S_2\), are equal and of opposite sign, and a lesser value when the intensities are unequal. Thus, CSF suppression can be achieved through appropriate timing for the \(S_2\) block to give a CSF signal intensity that is different from the \(S_1\) block. Figure 2 shows Bloch simulations of the 2-block sequence performed over the \(T_1, T_2\) parameter space. For comparison, simulation of the MP2RAGE is shown in Figure 2A using timings of \(t_1/t_2/\text{TR} = 0.9/2.6/5\) seconds and tip angles of \(5^\circ\) and \(9^\circ\). As expected from the MP2RAGE, the base images \((S_1, S_2)\) and the calculated intensity \(R_{1/2}\) (from Eq. 1) is primarily dependent on the \(T_1\) with minimal \(T_2\) dependence (a small dependence results from the finite duration of the adiabatic inversion pulse).

In comparison, Figure 2B shows simulations of the MPFLAGRE-2 sequence, including the \(T_2\) preparation module applied early in the \(T_1\) recovery and using a parameter set of \(t_1/t_2/\text{TR}/\TE = 0.1/1.6/5/0.085\) seconds and tip angles of \(5^\circ\) and \(9^\circ\). The calculated image (Eq. 4) shows that the CSF signal \((T_1, T_2)\) taken at \([4.3, 0.9]\) seconds is substantially lower than normal \(R_{1/2} = 0.18\), which is substantially lower than normal \(R_{1/2}^{\text{GM}}\) and \(R_{1/2}^{\text{WM}}\). The \(T_1, T_2\) values for tissue components used in these simulations are \(\text{CSF} = 4.3\) seconds, \(0.9\) seconds; \(\text{GM} = 2.0\) seconds, \(60\) ms; \(\text{WM} = 1.2\) seconds, \(60\) ms. It should be stated that the strategy for MPFLAGRE can also be applied with the spin-echo application applied late (rather than early) in the \(T_1\) recovery. Although there are differences between the early or late \(T_2\) preparation (Supporting Information Figure S3), the strategy remains similar (i.e., to optimize the timings of the \(T_2\) and \(T_1\) weighted blocks used with...
the self-normalization to achieve CSF suppression due to $T_1$ differences while maintaining $T_2$ sensitivity. This report focuses on the $T_2$ weighting applied early in the $T_1$ recovery.
2.2 | Three-block and 4-block sequences (MPFLAGRE-3 and MPFLAGRE-4)

The use of multiple readout blocks of acquisition in combination with both $T_1$ and $T_2$ preparation in the MPFLAGRE sequence provides additional flexibility. Two aspects of the multiple-block acquisition are considered. First, although the MPFLAGRE-2 sequence is able to suppress CSF, Figure 2B shows that there is relatively limited dynamic range for the calculated $R_{1/2}$ image. To increase this sensitivity with $T_2$, we recognize that the $T_2$ preparation induces a change in the longitudinal magnetization and thus the rate of $T_1$ recovery. Given the typically slow $T_1$ recovery, the $T_2$-dependent effects on amplitude and $T_1$ recovery rate thus transiently persist after the spin echo. To maximize the effect of the spin echo, we can then acquire an additional signal block (Figures 1 and 3A) and sum the 2 succeeding image blocks, $S_1$ and $S_2$. Analytically, this can be expressed as a sum of Eqs. (2) and (3), with simplifications including the small values for $t_{i2}$, $t_{i1}$, identical tip angles $\alpha$ for $S_1$ and $S_2$ (see Eqs. (2) and (3) for other terms, omitting the $B_1^*$, tip angles, density, and $T_2^*$ factors), and extracting the TE dependent terms:

$$S_1 + S_2 \propto -e^{-te/T_2} e^{M_{ss}/M_0} e^{-t_{i1}/T_1 A} N/2 (1 + e^{-t_{i2}/T_1 A}) + f (M_{ss}, A^N, R^N).$$

With this sign inversion to maintain the desired tissue sensitivity, a third (delayed) block $S_3$ is then used as the reference image for the self-correcting normalization according to Eq. (6):

**FIGURE 3** Bloch simulations of a 4-block acquisition. A, Time course of $I_z$ amplitudes (5 tissue components: blue, CSF; black, GM; red, WM; GM and WM with long $T_2$ values for pathology). The $[T_1, T_2]$ values for CSF are assigned at 4.3 seconds, 0.9 seconds; GM at 2.0 seconds, 60 ms; and WM at 1.2 seconds, 60 ms. B, Signal block 1 intensity is strongly $T_2$ weighted, shown over the $T_1, T_2$ parameter space. C, Signal block 2 intensity. D, Summed signal blocks 1+2 show the combined $T_1$ and $T_2$ sensitivity. E,F, Signal blocks 3 and 4 are strongly influenced by $T_1$. All signal intensities are plotted on a scale of $[-0.1, 0.1]$
As seen from Eq. (5) and simulated in Figures 3B-D, the summed block $S_{1+2}$ increases the weighting of the $-e^{-ti_2/T_1}A_N$ term from 1 to $1 + e^{-ti_2/T_1}A_N$. Consistent with this, simulation of the summed block strategy shows that the increase in dynamic range for the $R_{(1+2)/3}$ image arises from a drop in the calculated signal for normal tissue, whereas the pathologic long $T_2$ values “saturate” at 0.5 (Figure 4). Thus, the persistence of the spin-echo effect is a function of $T_1$ (WM returns more rapidly than GM) (i.e., $R_{(1+2)/3}$ depends on the $T_1$, as $R^{GM,normal}_{(1+2)/3}$ is larger [brighter] than $R^{WM,normal}_{(1+2)/3}$). With the adjacent block sum of $S_1$ and $S_2$, given the same numerical $T_2$ range is present between normal and pathology for WM and GM (approximately 60 ms to 160 ms), the absolute increase in calculated intensity between normal to pathologic $T_2$ values is similar between WM and GM: $R^{GM,normal}_{(1+2)/3} - R^{GM,pathol}_{(1+2)/3}$ and $R^{WM,normal}_{(1+2)/3} - R^{WM,pathol}_{(1+2)/3}$.

The second aspect of multiple-block acquisitions arises from the timing and use of the delayed reference image (e.g., through the addition of a fourth imaging block $S_4$) (Figure 4). Combining the $S_1$ and $S_4$ images using Eq. (4) generates a calculated $R_{1/4}$ that, by virtue of their common $T_1$ factors, largely eliminates $T_1$ dependence to generate a solely $T_2$-weighted image (which can be estimated from the simple 2-block analysis in Eqs. and under the conditions of long $ti_2$ and short $ti_1$). For completeness, we also show the calculated $R_{(1+2)/4}$ similar to $R_{(1+2)/3}$. This shows an enhanced dynamic range over $T_2$ but with mild $T_1$ sensitivity. Figure 4 shows the resulting simulation, including the calculated $R_{1/3}$ and $R_{(1+2)/3}$ (CSF-suppressed, $T_2$-weighted) and $R_{1/4}$ and $R_{(1+2)/4}$ ($T_2$-weighted) images.

\[
R_{(1+2)/3} = \text{real}\left(\frac{-(S_1 + S_2)^*S_3}{|S_1 + S_2|^2 + |S_3|^2}\right).
\]
2.3 | $B_1^+$ dependence

Although the self-correcting normalization eliminates $B_1^-$ variation, variation in $B_1^+$ is not wholly corrected. Figure 5 considers the $B_1^+$ dependence of the sequence when multiple acquisition blocks are used to calculate the signal. The transceiver array exhibits 15% SD over the brain when combined with RF shimming (Supporting Information). Simulation results from a MPFLAGRE-4 sequence are shown over the range of $T_2$ values, using $B_1^+$ values at ±15% and ±30% of the optimum $B_1^+$ value used (750 Hz). The CSF shows the greatest effect with $B_1^+$ variation, although the $R_{CSF}$ intensity remains at or below the normal-tissue WM intensity $R_{WM}$. Normal GM and WM show increased $R_{GM}$ and $R_{WM}$, increasing by less than or equal to 25%. However, it should be noted that in the range of normal $T_2$ values for GM and WM (40 ms-70 ms), the calculated intensity is steeply rising with $T_2$, and that within a 15% erroneous $B_1^+$, a less than 10-ms $T_2$ increase will give the same $R_{GM}$ intensity (i.e., the same signal intensity is equivalent to less than a 10-ms $T_2$ rise).

2.4 | Pathologic $T_2$ values

As the goal of the FLAIR sequence is to detect tissues with long (and likely pathologic) $T_2$ values, we need to have the values of pathologic relaxation at 7 T to properly optimize the parameters of the MPFLAGRE sequence. Although such values have not been widely reported, a 3T estimate is available, which found a less than 20% increase in hippocampal $T_2$ in epilepsy versus control. We estimate that to leave adequate “head room” for pathology, the calculated image values for normal GM and WM tissue need to be less than 0.30, with CSF intensities at or below the normal tissue values; these values were achieved in these optimizations.

3 | METHODS

We used a Siemens whole-body 7T Magnetom 8-channel multiple transmit system with body gradient coil and an 8 x 2 transceiver array for all acquisitions. All studies were approved by the institutional review board. The transceiver array was driven in coil pairs using 8 one-to-two splitters such that coils at equivalent azimuthal positions from the 2 rows are driven by the same RF transmit channel, with independent reception from all 16 channels. A fixed phase shift between the 2 rows is used to ensure constructive addition of the RF across rows and to maximize spatial coverage. $B_1$ shimming was performed in all subjects, requiring about 3.5 minutes (including 2.5 minutes of $B_1$ mapping acquisitions).
In control subjects, a mean $B_1^+$ of 17.6 uT with a SD of 10.4 ± 1.8% over the brain using a maximum voltage for every RF channel of less than 170 V (n = 8 subjects); Supporting Information Figures S1 and S2). As a result, conventional pulses are used, including hyperbolic secant pulses with $\mu = 10$ and 4 time constants. With a single $T_2$ preparation in the MPFLAGRE-4 acquisition and $2\pi$ refocusing pulses, the acquisition has a global SAR of 74 ± 2% of the US Food and Drug Administration’s guidelines of 3.2 W/kg (using 5 kg for adult head mass), as determined from the Siemens calibrated directional coupler measurements. A total of 5 control subjects were studied (Table 1), with mean height and weight of 62.1 ± 11 kg and 168.1 ± 8.1 cm (3 males, 2 females). A GRAPPA factor of 3 was used, achieving a $0.7 \times 0.7 \times 1.2$ mm (nominal volume $0.6 \text{ mm}^3$) resolution, with an acquisition time of 9.5 minutes. The phase encoding for the imaging blocks were either linear or center out, with the latter used to maintain maximal $T_2$ weighting at the center of $k$-space. All timings are thus reported as the duration between the center of the inversion recovery to the $k$-space center acquisition.

4 | RESULTS

4.1 | MPFLAGRE-2

Figure 6 shows the data acquired from a healthy control using the MPFLAGRE-2 sequence. For comparison, Figure 6A,B shows the data without and with the $T_2$ preparation, respectively. The timings in Figure 6A are set to create MP2RAGE-like contrast ($t_1/t_2/TR = 0.9/2.67/5$ seconds, tip angle = 5° and 9°), and as expected, the acquired and calculated images are all strongly $T_1$ weighted. Figure 6B,C shows the MPFLAGRE-2 data, acquired with $t_1/t_2/TR/TE = 0.14/1.65/5/0.100$ seconds using pulse angles of 5° and 9°. Figure 6C shows the base images with $T_2$ contrast dominating the $S_1$ image. In $S_1$, the WM-GM contrast is small; this is due to the dynamic range of the signal intensity governed by the high proton density CSF signal and the small WM-GM variation in $T_2$ at 7 T. Consistent with simulation, there is little $T_2$ contrast in the CSF-suppressed $R_{1/2}$ image (Figure 6B).

4.2 | MPFLAGRE-4

As the MPFLAGRE-3 acquisition is very similar to that of MPFLAGRE-4, Figure 7 shows the data from the MPFLAGRE-4 sequence. Shown are both the CSF-suppressed (Figure 7A) and $T_2$-weighted (Figure 7B) calculated images, over a range of [−0.5, 0.5]. The $T_2$-weighted images $R_{1/4}$ and $R_{1+(2)/4}$ again show the limited dynamic range of $T_2$ variation at 7 T between WM and GM. The difference between the calculated $R_{1/3}$ and $R_{1+(2)/3}$ images (showing the increase in dynamic range in total signal) is consistent with the simulation, and is a result of decreased calculated signal in normal WM and GM in $R_{1+(2)/3}$. With this effect being different between WM and GM, the $R_{1+(2)/3}$ images show slightly more GM-WM contrast in comparison with $R_{1/3}$. Although this initial report is not intended to evaluate pathology, Figure 8 shows the performance of the MPFLAGRE-4 (same acquisition parameters as in control from Figure 7) in 2 patients (epilepsy, left neocortical temporal lobe, anaplastic astrocytoma). In comparison to a clinical 3T FLAIR (Figure 8C), the epilepsy patient shows bright signal in the lateral cortex and hippocampus consistent with clinical data. The tumor patient shows that the bright MPFLAGRE signal in the temporal lobe extends into the posterior thalamus. The 3T studies were performed on a GE Signal Discovery MR750, with an inversion recovery for the epilepsy $T_1$ image; $T_1$ optimized fast spin echo for the tumor $T_1$ image; and both FLAIRs acquired with 2D fast spin echoes ($\text{TR/TE} = 11.6$ seconds/2.5 seconds/154 ms, resolution = $0.43 \times 0.43 \times 3.3 \text{ mm}^3$ [epilepsy] and $\text{TR} = 8.7$ seconds/2.2 seconds/150 ms, resolution = $0.63 \times 0.63 \times 5 \text{ mm}^3$ [tumor]).

4.3 | Comparison with 3D SPACE and contrast-to-noise ratio

For whole-brain coverage, the MPFLAGRE sequence is compared with the 3D variable flip angle SPACE acquisition. As reported by Visser, the SPACE acquisition benefits substantially from 2D acceleration ($2.5 \times 2.5$) but still requires a long TR. Our implementation used a GRAPPA factor of 4; however, a TR of 10 seconds was needed to reduce SAR to within the US Food and Drug Administration.

| TABLE 1 | Signal-to-noise ratio, contrast-to-noise ratio, and SAR for the MPFLAGRE sequence |
|---------|-------------------------------------------------------------|
|         | SNR            | CNR            | SAR (FDA max. 15 W) |
|         | GM      | WM    | CSF  | GM-CSF | WM-CSF |          |
| 3D SPACE | 8.6    | 12.0  | 5.6  | 0.20   | 4.21   | 11.6 W (77%) |
| 3D MPFLAGRE-4 $R_{1/3}$ | 13.7  | 30.4  | 7.0  | 6.17   | 8.00   | 10.7 W (70%)   |
| 3D MPFLAGRE-4 $R_{1/3}$, N=5 | 17.0 ± 2.3 | 35.2 ± 6.1 | 5.4 ± 18 | 6.63 ± 1.0 | 6.94 ± 1.0 | 11.1 ± 0.5 W (74%) |

Abbreviations: CNR, contrast-to-noise ratio; FDA, US Food and Drug Administration.
Administration’s guidelines, resulting in an acquisition of 19.7 minutes with TR/TI/TE of 10.1 seconds/2.35 seconds/200 ms. Figure 9 shows the mildly brighter GM, characteristic of the residual $T_1$ weighting. However, this contrast is $B_1^+$-sensitive, and the consistency of the image intensity is relatively poor, reflecting the variation in $B_1^+$ (and $B_1^+$). Matched in resolution ($0.7 \times 0.7 \times 1.2 \text{ mm}^3$) with the MPFLAGRE, Table 1 provides the contrast-to-noise ratio (CNR) and SNR between the SPACE and $R_{1/3}$ images for a single volunteer (single session). For region of
interest measurements, contiguous $4 \times 4 \times 1$ pixel blocks were taken from the centrum semi-ovale, thalamus, and posterior ventricle for WM, GM, and CSF. The CNR and SNR were calculated in 5 healthy volunteers (Table 1) using the following equation:

$$
\text{CNR} = \frac{(R_a - R_b)}{\sqrt{\left(\sigma_a^2 + \sigma_b^2\right) / 2}}, \quad \text{SNR} = \frac{R_a}{\sigma_a}.
$$

**FIGURE 8** Data from an epilepsy (A) and brain tumor patient (B). For both, the coregistered 7T MP2RAGE, MPFLAGRE-4 ($S_{(1+2)/3}$), 3T clinical $T_1$, and FLAIR images are shown. For the epilepsy patient, increased signal intensity is identified in the lateral left temporal lobe (thick arrows, axial and coronal) and is consistent with clinical data. Note from the coronal image that there is also increased signal in the left hippocampus (thin arrow, coronal), which is characteristic of the local network involvement in temporal lobe epilepsy. The 3T FLAIR from this patient, acquired within 3 months of the 7T images, was interpreted as negative. For the tumor patient, all 7T and 3T images were acquired within 1 week. The arrows identify the increased MPFLAGRE signal in the left temporal lobe extending into the posterior thalamus.

5 | DISCUSSION

5.1 | $T_1$ and $T_2$ dependence of the MPFLAGRE signal

As shown in simulation and implementation, the MPFLAGRE sequence is able to generate $T_2$-weighted images with controlled $T_1$ effects. The data show that the $T_2$ preparation is effective and the self-correcting normalization with multiple
acquisition blocks enables modulation of the $T_1$ relaxation effects. The $T_1$ and $T_2$ behavior is better understood by recognizing the balance between the $T_2$-weighted $S_1$ and reference (non-$T_2$-weighted) images used in the normalization. In the case in which the common $T_1$ relaxation factor contributes to both $S_1$ and the reference image (e.g., $S_4$ in the MPFLAGRE-4), the normalization results in negligible $T_1$ dependence regardless of the $T_2$ value (Figure 4D).

However, if the reference image $S_4$ is acquired with some residual $T_2$ (in addition to $T_1$) weighting, the normalization cannot cancel the $T_1$ weighting over all $T_2$ values. Tissues with very long $T_2$ will be transparent to the spin-echo weighting, and generate a $T_1$-weighted image. Tissues with extremely short $T_2$ will approach 0.0 for all $T_1$ values. Signal from tissues with intermediate relaxation values will vary depending on the specific acquisition parameters and the relaxation values, and thus is a target for optimization. As discussed, suppressing CSF is achieved by optimizing the timings of the 2 blocks (either $S_1$ and $S_2$ in MPFLAGRE-2 or $S_1$ and $S_3$ in MPFLAGRE-3 and MPFLAGRE-4) to acquire substantially dissimilar signal intensities, ideally generating a signal in the 0 to -0.5 range. With lesional imaging, the goal is to generate significant calculated $R_{T2w,Ref}$ difference between normal and longer (pathologic) $T_2$ values. Applying the $T_2$ weighting before the $T_1$ recovery null, we recognize that pathologic $T_2$ values will result in “less $T_1$ later shift” than tissues with normal $T_2$ values. Thus, to generate an increase in calculated signal from tissue with pathology, the reference image needs to be acquired after the null, sign inverted and larger in absolute amplitude compared with normal $T_2$ values (i.e., as exemplified with the MPFLAGRE-2 timing).

The limited dynamic range of the 2-block acquisition arises from the amplitudes of signal in the $T_2$-weighted and reference images. With the MPFLAGRE-3 and MPFLAGRE-4, we expand this range and take advantage of the temporal persistence of the $T_2$-dependent $T_1$ later shift effect through the summation of the 2 adjacent blocks. This later shift effect is larger for shorter $T_2$ components (i.e., normal $T_2$) rather than pathologic $T_2$ values (Figure 3). Thus, the sign-sensitive summation of 2 adjacent blocks after the spin echo will result in an apparent decrease in amplitude for normal $T_2$ components and give decreased $R_{(1+2)/3}$, whereas the pathologic long $T_2$ components effectively saturate with $R_{(1+2)/3}$ at 0.5.

With the shorter $T_1$ of normal WM, the $R_{WM}^{normal}$ falls more than $R_{GM}^{normal}$. However, because of the similar $T_2$ values of WM and GM at 7 T, the absolute increases in $R_{GM}^{normal} - R_{GM}^{pathol}$ and $R_{WM,normal}^{normal} - R_{WM,normal}^{pathol}$ are similar for both tissues; this effect is also seen with the simpler MPFLAGRE-2 sequence.
5.2 Specific absorption rate and CNR of the MPFLAGRE sequence

From a SAR perspective, the use of a single nonselective adiabatic spin echo allows the MPFLAGRE sequence to function with high efficiency. With the MPFLAGRE-4 sequence, over n = 5 adult control volunteers, the 6-minute average SAR was maintained at 74% ± 2% of the US Food and Drug Administration’s guidelines. The CNR between WM-GM is approximately 1.0 (data not shown), which is consistent with the small differences in their T2 at 7 T. For GM-CSF the CNR is 6.63 ± 1.0, and for WM-CSF the CNR is 6.94 ± 1.0, which again is consistent with the similar intensities between WM and GM. In spite of the combination of multiple images used in this approach, which may otherwise be expected to increase variability, the coefficient of variance(s) in the MPFLAGRE data are comparatively low at less than 15%, reflecting their nonrandom source of variability. As a result, the SNR and CNR are higher with the MPFLAGRE sequence in comparison with the variable flip angle acquisition.

Inspection of Figures 5 and 6 show that there is enhanced signal intensity at the cortical rim and ependyma. This is consistent with the report of von Veluw,23 who concluded that increased T2 contributes to their enhanced FLAIR signal intensity. Although quantitative T2 data do not exist on these tissue components, a long T2 for the cortical rim is not surprising, given that it contains the comparatively poorly vascularized molecular layer I of the neocortex. Similarly, the high signal intensity from the ependyma is likely related to its function as the neuroepithelial layer around the ventricle that contributes to the production and regulation of CSF.

5.3 Caveats with the MPFLAGRE sequence

The primary challenge with this sequence is its B1+ sensitivity, as shown in Figure 9. A 15% increase or decrease in B1+ results in increased intensity R1/3 by less than 20% for WM and less than 28% for GM, although this effect decreases at longer T2. However, this effect arises from the steep rise of R over the normal range of T2 values. From a viewpoint of a given R1/3 intensity, a 15% B1+ variation results in a small decline in apparent T2 by about 10 ms for GM (about 5 ms for white matter). With 30% B1+ inhomogeneity, these effects increase substantially, with the T2 sensitivity curves broadly shifting to lower T2 values (i.e., there can be erroneously bright signal [> 0.30] for relatively normal T2 values). In practice, our experience with the transceiver with the adiabatic refocusing pulses has not been shown to be a major problem for this over the entire head. Nonetheless, this sensitivity can be improved with incorporation of an adiabatic excitation spin-echo module, such as that used by Dyvorne and Balchandani,16 although at added SAR cost.

With the CSF suppression generated from the normalization strategy performed between the multiple acquisition blocks, acquisition of differing resolution images but at the same delay times can change the resulting contrast. However, for the given parameters there is excellent consistency between different subjects, as indicated in Table 1 and the patient data. Nonetheless, this effect can be minimized with use of additional acceleration methods in 2 or 3 dimensions. In addition, magnetization-transfer effects, which are not modeled here, can differentially affect the signal between the Si,j blocks and thus contribute to the contrast.24 Notably, with magnetization-transfer effects typically decreasing the signal intensity in high-density (normal) tissue, this would result in an enhancement of the apparent MPFLAGRE sensitivity to pathology (the pathologic longer T2 would exhibit a lesser magnetization-transfer effect). Finally, the need for relatively long TR still limits the entire acquisition at approximately 10 minutes. However, with generation of both the T2 and CSF-suppressed T2-weighted images in a single matched image, the MPFLAGRE remains an efficient acquisition.

6 CONCLUSIONS

In summary, we have developed and demonstrated the MPFLAGRE sequence as a flexible acquisition that builds on the self-correcting normalization strategy17 to generate T2 and CSF-suppressed T2 weighted contrast at 7 T. This is performed by incorporating a longitudinal spin-echo weighting within the context of an inversion recovery. With appropriate timing and combination of multiple imaging blocks during the inversion recovery, it is possible to control the T1 dependence and specifically suppress CSF. Not surprisingly, the T2-weighted image, even when corrected for M0, B1+ and T2*, displays relatively little contrast between WM and GM, reflecting the small to minimal difference in T2 at 7 T. As applied with a transceiver array, the sequence functions well within SAR guidelines. Overall, as the CSF-suppressed T2-weighted contrast is designed for sensitivity to pathology, additional work with patients will be necessary to further optimize the parameters at 7 T.

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SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section at the end of the article.

FIGURE S1 Two configurations are shown for driving the 8 × 2 transceiver from 8 independent RF channels. In configuration A, a single RF channel drives 2 adjacent coils within a row. In configuration B, a single RF channel drives 2 longitudinally adjacent coils.

FIGURE S2 Experimental data showing performance of the 2 configurations. For each configuration, the MP2RAGE and B1 maps are shown. In configuration B, 2 RF distributions can be defined to excite the “homogeneous” volume and a “ring” volume.

FIGURE S3 Bloch simulations of a late T2 preparation 2-block acquisition, S1 and S2, A. The time course of Iz amplitudes is shown (5 tissue components: T1, T2 values for CSF are assigned at 4.3 seconds, 60 ms; red, WM with inversion; pathologic T2). Note that the time axis is not linear, with each unit representing a simulation step. B. The surface plots of the signal amplitudes are shown over the [T1, T2] parameter space for S1, S2, and the calculated signal R1/2 (using Eq. 1). The calculated R1/2 shows the CSF suppression and signal enhancement over the pathologic range of T2. The black dots on the surface plots indicate the 5 tissue components: T1, T2 values for CSF are assigned at 4.3 seconds, 0.9 seconds; GM at 2.0 seconds, 60 ms; and WM at 1.2 seconds, 60 ms.

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