Dear Editor,

Red blood cell (RBC) genotyping is recommended to limit alloimmunization in patients with sickle cell disease or thalassemia [1]. RBC genotyping can resolve the serologic weak D phenotype and inconclusive RhD typing in obstetrics and identify the RBC phenotype in patients with autoantibodies or a positive direct antiglobulin test (DAT) [2]. DNA-based RBC typing has better accuracy and provides more information on RBC antigens than typical phenotyping [1].

We report a challenging case of a pregnant patient in whom a lack of compatible RBCs, based on RBC genotyping, suggested the presence of anti-Dib and anti-E specificities. To our knowledge, extended RBC genotyping was used to obtain compatible RBC units for the first time in Korea.

A 36-year-old Korean woman (gravida 2, para 0) came to Pusan National University Hospital for prenatal care. She had a history of blood transfusion at the age of 12 years. Her blood type was B and RhD positive, and she tested positive for irregular antibodies at 30 weeks of gestation. However, we could not determine the types of the unexpected antibodies; moreover, detection of compatible RBCs was difficult due to the presence of unexpected antibodies. We obtained one unit of compatible packed RBCs (matched for Rh, Kell, Duffy, and Kidd) for practicable 350-unit cross matching and one unit of whole blood from autologous blood donation. The study protocol was approved by the institutional review board of Pusan National University Hospital, and written informed consent was obtained from the patient. Extended genotyping of 37 RBC antigens with the ID CORE™ kit (Progenika Biopharma-Grifols, Bizkaia, Spain) that is based on Luminex xMAP technology predicted the rare blood type Di(a+b−) (Table 1). We hypothesized that the lack of a compatible blood product was due to the presence of anti-Dib antibodies. The patient delivered a female infant by elective cesarean section at 36 weeks of gestation. She received an intraoperative transfusion of autologous whole blood (one unit) and compatible packed RBCs (one unit) one day post-surgery. The Central Laboratory of the Swiss Red Cross in Bern, Switzerland, confirmed the presence of anti-Dib and anti-E antibodies in the patient’s serum after delivery. The patient and her infant were discharged from the hospital on day 5 after the cesarean section.

The infant had a birth weight of 2,590 g. Her day 14 laboratory findings were as follows: hemoglobin, 9.74 mmol/L; hematocrit, 0.46 fraction; reticulocyte count, 0.01 fraction; total bilirubin, 107.24 µmoL/L; direct bilirubin, 18.81 µmoL/L; blood group O, RhD positive. DAT was positive (1+) for polyspecific anti-hu-
man IgG. Although the neonate’s serum tested negative for unexpected antibodies, the eluate prepared from her RBCs tested positive in the unexpected antibody screening. We suspected the presence of anti-Dib antibodies because all panels, except the auto-control, were reactive in antibody screening. Blood group typing of the infant was not performed as the sample volume was too low.

Dib is a high-frequency antigen (HFA) in most populations. Anti-Dib can cause hemolytic transfusion reactions and serious hemolytic disease in fetuses and newborns [3]. In a report from Korea, the prevalence of the predictive phenotype Di(a+b−) was 0.7% (3/419) in healthy Korean donors [4]. Anti-Dib was observed in four cases of hemolytic disease in newborns [5-8] and two adult cases in which no matched blood products could be found for orthopedic surgery [9]. In another report, Anti-Dib were detected through a reaction with blood cells for the identification of an unexpected antibody with the Di(a+b−) phenotype; the genotype of the Diego blood type could not be identified [7]. In another case, Dib was detected through genotyping of the Diego blood type by direct sequencing of SLC4A1, which encodes the erythroid band 3 protein anion exchanger 1 (AE1) glycoprotein. A mutation resulting in a single amino acid change in this protein resulted in the production of the Diego antigen [3, 9].

Hospitals typically perform time-consuming serologic tests on site to locate antigen-negative RBC units [10]. Antibodies against HFAs may be difficult to identify due to a lack of negative panel cells; thus, identifying compatible antigen-negative blood can be challenging. In this case, reactions with all panel cells were positive (except for the auto-control). As it is difficult to identify antibodies in the hospital, samples are typically sent to a reference blood bank or laboratory for antibody identification through additional tests; however, this process is time-consuming. In this case, we could predict anti-Dib antibodies through extended RBC genotyping because we suspected antibodies against HFAs. RBC genotyping can be performed immediately at the hospital. In conclusion, in cases in which antibody identification and detection of compatible RBCs are difficult, antibodies against HFAs can be suspected. Extended RBC genotyping is a useful tool to identify specific anti-HFA antibodies and identify compatible RBCs for patients with these antibodies in transfusion medicine laboratories at hospitals.

Authors’ Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

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