Collective Classification of Textual Documents by Guided Self-Organization in T-Cell Cross-regulation Dynamics

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Abstract We present and study an agent-based model of T-Cell cross-regulation in the adaptive immune system, which we apply to binary classification. Our method expands an existing analytical model of T-cell cross-regulation [28] that was used to study the self-organizing dynamics of a single population of T-Cells in interaction with an idealized antigen presenting cell capable of presenting a single antigen. With agent-based modeling we are able to study the self-organizing dynamics of multiple populations of distinct T-cells which interact via antigen presenting cells that present hundreds of distinct antigens. Moreover, we show that such self-organizing dynamics can be guided to produce an effective binary classification of antigens, which is competitive with existing machine learning methods when applied to biomedical text classification.

More specifically, here we test our model on a dataset of publicly available full-text biomedical articles provided by the BioCreative challenge [34]. We study the robustness of our model’s parameter configurations, and show that it leads to encouraging results comparable to state-of-the-art classifiers. Our results help us understand both T-cell cross-regulation as a general principle of guided self-organization, as well as its applicability to document classification. Therefore, we show that our bio-inspired algorithm is a promising novel method for biomedical article classification and for binary document classification in general.

Keywords Artificial Immune Systems · Biomedical Document Classification · Data Mining · Machine Learning · Bio-inspired Computing · Complex Adaptive systems · Guided Self-Organization

1 Background

At least since the beginning of systematic genomic studies, there has been a tremendous growth of scientific publications in the life sciences [20]. Pubmed (http://pubmed.gov) now contains a growing collection of more than 19 million biomedical articles. Manually classifying these articles as relevant or irrelevant to a given topic of interest is very time consuming and inefficient for curation of new published articles [22]. Literature (or text) mining offers solutions for automatic biomedical document classification and information extraction from huge collections of text, as well as the linking of numerous biomedical databases and knowledge resources [22][11]. Because it is very important to validate and assess the quality of proposed solutions, various community-wide competitions and challenges have been organized so that automatic systems can be evaluated against human annotated data sets (e.g. TREC Genomics [12]). One such effort is the BioCreative challenge, which aims to assess biomedical literature mining in real-world scenarios [16][27][34]. Machine learning has offered a plethora of solutions to this problem [22][25], however, even the most sophisticated of solutions often overfit to the training data and do not perform as well on real-world scenarios such as that provided by BioCreative [33][37]. One of the challenges of biomedical article classification in real-world scenarios is the presence of highly unbalanced classes; typically, there are many more irrelevant than relevant documents, without prior knowledge of class proportions. This was the case of the article classification data set in the Biocreative BC2.5 challenge [34]. While participating teams (including our own team [37]) did not enter bio-inspired solutions, the unbalanced nature of classes and the presence of conceptual drift, which we showed to occur between training to test data sets [33][37], may be a good scenario to test classifiers inspired by the vertebrate immune system—which must operate un-

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under class-imbalance with permanent drift in the populations of pathogens encountered. Therefore, here we explore the feasibility of using T-Cell cross-regulation dynamics to classify biomedical articles using the real-world scenario provided by the Biocreative 2.5 data set.

The immune system (IS) is a complex biological system made of millions of cells all interacting to distinguish between self and nonself substances, to ultimately attack the latter \[^1\]. In analogy, relevant biomedical articles for a given concept need to be distinguished from irrelevant ones. To perform such a topical classification, we can use the occurrence and co-occurrence of thousands of words in a document. In this sense, words can be seen as interacting in a text in such a way as to allow us to distinguish between relevant and irrelevant documents—in analogy with the interactions among T-cells and antigens that lead to self/nonself discrimination in the immune system, as we describe below.

Our approach is based on the idea that the immune system is a distributed collection of molecular constituents with no central controller \[^{6}\]. Therefore, immune classification needs to result from a collective classification process, defined as the ability of decentralizedized systems of many components to classify situations that require global information or coordinated action \[^{21}\]. Nature is full of examples of collective classification: the dynamics of stomata cells on leaf surfaces are known to be statistically indistinguishable from the dynamics of automata that are capable of performing nontrivial classification \[^{13}\], biochemical intracellular signal transduction networks are capable of emergent classification \[^{31}\], quorum sensing in bacteria \[^{23}\] and social insects \[^{17}\], etc. We can also study collective classification in general models of complex systems such as Cellular Automata, namely by identifying regular patterns in the dynamics that store, transmit and process information \[^{4,15,18}\]. Here, instead of looking at general models of complex systems, we focus on a specific immunological model of T-Cell cross-regulation dynamics \[^{28}\]. We are interested in exploring the collective dynamics of this model to: (1) build a novel bio-inspired machine learning solution for document classification, and (2) understand how well collections of T-Cells engaged in cross-regulation perform as a classifier. The first goal entails a bio-inspired approach to computational intelligence, and the second a computational biology experiment, but both are based on artificial life principles.

It should be noticed that recent work in artificial immune systems (AIS) \[^{20}\] has lead to a few immune-inspired solutions to document classification in general \[^{7}\], however, none to our knowledge has been applied to biomedical arti-

\[^{1}\] We use the terminology of self/nonself discrimination, though perhaps a more accurate description is classification of harmless vs. harmful substances; harmless can also include antigens from bacteria that are necessary for vertebrate bodies, and harmful can also include body’s own tumor cells.
analytical model of T-cell cross-regulation. In section 4, we present our agent-based model of T-Cell cross-regulation for binary classification, here applied to document classification. In section 5, we describe the biomedical data provided by Biocreative and the feature selection method. In section 6, we study the robustness of our model on various parameter ranges and experimental setups. Finally, in section 7, we compare our model with state-of-art classifiers.

2 The Immune System as Inspiration

The vertebrate adaptive immune system (IS) is a complex network of cells that distinguishes between self and non-self substances or antigens—usually fragments of proteins that can be recognized by the immune system. When non-self antigens are discovered, an immune response to eliminate them is set in motion. Recognizing self antigens, which obviously should not lead to an (auto)immune response to eliminate them, is resolved by negative selection of T-cells which takes place in the thymus, and removes T-Cells that strongly bind to self antigens—after positive selection of T-Cells that are capable of binding with the major histocompatibility complex (MHC). It is in the thymus that T-cells develop and mature; only T-cells that have failed to bind to self antigens are released (as mature naive T-cells), while the rest of the T-cells is culled. Mature T-cells are allowed out of the thymus to detect nonself antigens. They do this by binding to antigen presenting cells (typically B-cells, macrophages and dendritic cells) that collect and present antigens via MHC after breaking them by lysosome. The specific T-cells that are able to bind to the presented antigens then stimulate B-cells that start a cascade of events leading to antibody production and the destruction of the pathogens or tumors linked to the antigens. However, it is possible that T-cells and B-cells, which are also trained in the thymus and bone marrow, mature before being exposed to all self antigens. Even more problematic is the somatic hypermutation that ensues in lymph nodes after the activation of B-cells through a process known as “clonal selection”. At this stage, it is possible to generate many mutated B-Cell clones that could bind to self antigens. Either situation can cause autoimmunity by generating T-cells capable of attacking self antigens. One way to deal with this problem is by a process called costimulation which involves the co-verification of self antigens by both T-cells and B-cells before the antigen is identified as associated with a nonself pathogen to be attacked. To further insure that the T-cells do not attack self, another type of T-cells known as regulatory T-cells, are formed in the thymus where they mature to avoid recognizing self antigens. These regulatory T-cells have the responsi-

2 A good, though already a bit dated, overview of the vertebrate immune system for the artificial life community is Hofmeyer’s.

Fig. 1 CRM interactions that define the dynamics of APC and E and R T-cells. The model assumes that APC can only form conjugates with a maximum of two T-cells. Adapted from [28].

3 The Cross-Regulation Model

The T-cell Cross-Regulation Model (CRM) [28] is a dynamical system that aims to distinguish between self and nonself protein fragments (antigens) using only four possible interaction rules amongst three cell-types: Effector T-cells (E), Regulatory T-cells (R) and Antigen Presenting Cells (APC). As their name suggests, APC present antigens for the other two cell-types, E and R, to recognize and bind to them. Effector T-cells (E) proliferate upon binding to APC, unless adjacent to regulatory T-cells (R), which regulate E by in-
hibiting their proliferation. For simplicity, proliferation of cells is limited to duplication in quantity in contrast to having a proliferation rate. T-cells that do not bind to APC die off with a certain death rate. The dynamics of the CRM depend on four interaction rules defined by the following reactions (illustrated in Fig. 1):

\[
E \xrightarrow{d_E} \{ \} \quad \text{and} \quad R \xrightarrow{d_R} \{ \}
\]

(1)

\[ A + R \rightarrow A + R \]

(2)

\[ A + E \rightarrow A + 2E \]

(3)

\[ A + E + R \rightarrow A + E + 2R \]

(4)

Reaction (1) defines \( E \) and \( R \) apoptosis with the corresponding death rates \( d_E \) and \( d_R \). The last three proliferation reactions define the maintenance of \( R \) (2), the duplication of \( E \) (3), and the maintenance of \( E \) and duplication of \( R \) (4).

Carneiro et al. [28] developed the analytical CRM to study the dynamics of a single population of T-cells (with effector and regulatory elements) that interacts with APC that present a single antigen. In [29,30], we extended the original CRM model to be able to deal with multiple populations of antigens and T-Cells using agent-based modeling. More recently, Sepulveda [35, pp 111-113] extended the original CRM model to be able to deal with multiple populations of antigens \( d \) and T-Cells using agent-based modeling. In [29,30], we extended the original CRM model to be able to deal with multiple populations of antigens and T-Cells using agent-based modeling. More recently, Sepulveda [35, pp 111-113] extended the original CRM model to be able to deal with multiple populations of antigens and T-Cells using agent-based modeling.

In biology, antigen recognition is a more complex process than mere polypeptide sequence matching, but for simplicity we limit our feature recognition to string matching. APC are organized as a list of pairs of “slots” of (textual) features, where T-cells, specific for those features, can bind. We use this antigen/feature presentation scheme of pairs of “slots” to simplify our algorithm. In future work we will study alternative feature presentation scenarios. An APC is modeled as a list of “slots” of pairs of features \( A_d \), where \( s_d = (f,g) \), \( f,g \in A_d \), and \( n_s = \frac{n_A}{2} \), \( f \) and \( g \) are sampled (without repetition) from \( A_d \) and randomly distributed exactly \( n_A \) times over the list of slots that makes up the APC.

In the natural immune system, millions of novel T-cells are randomly generated in the thymus every day to attempt to predict future antigens. In our algorithm, in contrast, we generate T-cells only for features (words) occurring in the relevant document corpus. This is reasonable because the space of meaningful words in a language is largely fixed and much smaller than the space of possible polypeptide epitopes in biology. More specifically, a document \( d \) contains a set of features \( F_d \); an artificial APC \( A_d \) that represents \( d \), presents a subset of antigens/features \( A_d \subseteq F_d \) to artificial \( E \) and \( R \) T-cells. \( E_f \) and \( R_f \) bind to a specific feature \( F \) on any APC that presents it; if \( f \in A_d \), then any available \( E_f \) or \( R_f \) in the cellular dynamics may bind stochastically to \( A_d \) as illustrated in figure 2.

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In summary, each T-cell population is specific to and can bind to only one feature presented by any APC. Implementing the algorithm as an Agent-based model (ABM) allows us to deal with the recognition and co-recognition (co-occurrence in the same document) of many features simultaneously, rather than a single one as the original CRM does.

The ABCRM uses incremental learning to first train on \( N \) labeled documents (relevant and irrelevant), which are ordered sequentially (typically by time signature) and then test on \( M \) unlabeled documents that follow in time order. Fig. 4 illustrates this stream of labeled documents (blue for relevant and red for irrelevant) followed by unlabeled grayed documents. The sequence in which documents are received affects the artificial cellular dynamics, as incoming APC
Fig. 2 To illustrate the difference between the CRM and the ABCRM, the top part of the figure represents a single APC of the CRM which can bind to a maximum of two T-Cells. The lower part represents the APC for a document \( d \) in the ABCRM, which contains many pairs of antigen/feature “slots” where pairs of T-cells can bind. In this example, the first pair of slots of the APC \( A_d \) presents the features \( f_i \) and \( f_j \); a regulatory T-cell \( R_i \) and an effector T-cell \( E_j \) bind to these slots, which will therefore interact according to reaction (4)—\( R_i \) inhibits \( E_j \) and in turn proliferates by doubling. The next pair of slots leads to the interaction of regulatory T-cells \( R_i, R_k \) that duplicate via reaction (2)...

and T-cells face a T-cell dynamics that depends on the specific documents previously encountered. Therefore, we use publication-time as the default ordering for incoming documents, and study if there is an advantage to preserving the original temporal sequence of articles (see section 6.3).

Carneiro et al. [28] show that both \( E \) and \( R \) T-cells coexist in healthy individuals assuming enough APC exist. \( R \) T-cells require adequate amounts of \( E \) T-cells to proliferate, but not too many that can out-compete \( R \) for the specific features presented by APC. “Healthy” T-cell dynamics is identified by observing the co-existence of both \( E \) and \( R \) T-cells with \( R \geq E \). “Unhealthy” T-cell dynamics is identified by observing \( E \gg R \), and should result when encountering many irrelevant features in a document—in analogy with encountering many nonself antigens.

In other words, features associated with relevant documents should have more \( R \) T-cell representatives than \( E \) ones in the artificial cellular dynamics. In contrast, features associated with irrelevant documents should have many more \( E \) than \( R \) T-cells. Therefore, when a document \( d \) contains features \( F_d \) that bind mostly to \( E \) rather than \( R \) cells, we can classify it as irrelevant—and relevant in the opposite situation (see Fig. 3).

The ABCRM is controlled by 6 parameters:

- \( E_0 \) is the initial number of Effector T-cells generated for all new features
- \( R_0^+ \) is the initial number of Regulatory T-cells generated for all new features in irrelevant and unlabeled (test) documents
- \( R_0^- \) is the initial number of Regulatory T-cells generated for all new features in relevant documents
- \( d_E \) is the death rate for Effector T-cells that do not bind to APC
- \( d_R \) is the death rate for Regulatory T-cells that do not bind to APC
- \( n_A \) is the number of total slots in which each feature \( f \) is presented on APC

When (textual) features are encountered for the first time, a fixed initial number of \( E_0 \) effector T-Cells and \( R_0 \) regulatory T-Cells is generated for every new feature \( f \). These initial values of T-cells vary for relevant and irrelevant documents in training and in test stages. More Regulatory (\( R_0^+ \)) than Effector T-cells are generated for features that occur for the first time in documents that are labeled relevant in the training stage (\( R_0^+ > E_0 \)), while fewer Regulatory (\( R_0^- \)) than Effector T-cells are generated in the case of irrelevant documents (\( R_0^- < E_0 \) see Fig. 4). Features appearing in unlabeled documents for the first time during the test stage are treated as features from irrelevant documents, assuming that new features are irrelevant (nonself) until neutralized by the collective dynamics given their co-occurrence with relevant ones.

Naturally, relevant features will occur in irrelevant documents and vice versa. However, the assumption is that relevant features tend to co-occur more frequently with other relevant features in relevant documents and similarly for irrelevant features. Therefore, the proliferation dynamics defined by the 4 reactions and guided by co-binding to APC slots is expected to correct the erroneous initial bias as we will show in section 6.4. But this self-correction has not been proven in our previous works [38,36], and it is one of the issues we test in the present work.
Fig. 4 A stream of ordered labeled documents (blue for relevant and red for irrelevant) followed by ordered unlabeled grayed documents is introduced. Each document $d$ is represented by a polyspecific APC $A_d$ that arbitrarily presents the antigens/features $f$ of $d$. APC are then dropped in the pool of T-Cell populations representing previously encountered features/antigens, which follow the cellular interaction dynamics defined by the four interaction rules (see eq (2-4)). Finally, document $d$ is classified as relevant if the majority of its features $f$ have more $R_f$ than $E_f$, and irrelevant otherwise.

Finally, to classify a document $d$, we observe the cellular interaction dynamics that results after its respective APC $A_d$ is left to interact with the various T-Cell populations. More specifically, each document is classified based on the $E$-to-$R$ ratios of all its features $f \in A_d$; this process is illustrated in Fig. 3. A detailed pseudocode of the algorithm follows:

**ABCRM Algorithm:**

**Input:** Stream of labeled and unlabeled documents  
**Output:** Labels for unlabeled documents  

**foreach document $d$ do**

Generate a list of pair slots $A_d$ presenting each $f \in A_d$ at $n_A$ randomly distributed slots. Let $C$ contain $E_f$ and $R_f$ T-cells for all features $f$ in the cellular dynamics.

**foreach $f \in A_d$ representing document $d$ do**

if $E_f \notin C$ and $R_f \notin C$ then

$E_f = E_0$ (i.e. generate $E_0$ Effector T-cells for $f$)

if $d$ is labeled relevant then

$R_f = R_0^+$ (i.e. generate $R_0^+$ Regulatory T-cells for $f$)

else

$R_f = R_0^-$ (i.e. generate $R_0^-$ Regulatory T-cells for $f$)

end

end

Update $C$ with $E_f$ and $R_f$ Let all $E_f$, $R_f$ bind specifically to matching $f$ on $A_d$.

**end**

**foreach pair of adjacent $(f, g)$ on $A_d$ do**

Apply the following interaction rules and update total number of $E$, $R$ T-cells:

$(R_f, R_g) \rightarrow R_f + R_g$

$(E_f, E_g) \rightarrow 2E_f + 2E_g$

$(E_f, R_g) \rightarrow E_f + 2R_g$

end

**foreach $R_f, E_f \in C$ that do not bind to $A_d$ do**

Cull $E_f$ and $R_f$ according to death rates $d_E$ and $d_R$

end

if $d$ is unlabeled then

Let $R(d) = \sum_{f \in A_d} \left( \frac{R_f}{\sqrt{R_f^2 + E_f^2}} \right)$ and

$E(d) = \sum_{f \in A_d} \left( \frac{E_f}{\sqrt{R_f^2 + E_f^2}} \right)$

if $R(d) \geq E(d)$ then

Classify $d$ as relevant

else

Classify $d$ as irrelevant.

end

end
5 Data and Feature Selection

The BioCreative (BC) challenge aims to assess the quality of biomedical literature mining algorithms such as article classifiers. The article classification task of BioCreative 2.5 \([34]\) was based on a training data set \((T)\) comprised of 61 full-text articles relevant \((P_T)\) to the topic of protein-protein interaction (PPI) and 558 irrelevant ones \((N_T)\). The realistic imbalance between the relevant and irrelevant instances is very challenging for common machine learning techniques, since there are few instances of the topical category of interest to generalize from. Because we cannot predict how imbalanced the validation set will be, we first search for optimal AICRM parameters on a smaller sample of the training set that is balanced in the numbers of relevant and irrelevant documents. The optimal parameters are not only useful for fine-tuning our algorithm for the best classification performance but also for studying the robustness and behavior of T-cell dynamics under several experimental setups as we will show in section 6. For this purpose, we chose the first 60 relevant and sampled 60 irrelevant articles that were published around the same date (uniform distribution between Jan and Dec 2008), and we called this subset the optimization dataset. In the test stage we use the unbalanced validation set containing 63 \(P_T\) (black) and 532 \(N_T\) (black) documents. Notice that the validation data was provided to the participants in the classification task of BioCreative 2.5 unlabeled, therefore participants had no prior knowledge of class proportions.

We pre-processed all articles by filtering out common words and porter stemming \([2]\) the remaining words which are all the potential features. We then ranked words/features \(f\) extracted from training articles \((T)\) according to two scores: the first one is the average TF.IDF \([25]\), and the second one is the separation score \(S(f) = |p_P(f) - p_N(f)|\) where \(p_P\) and \(p_N\) is the probability of a feature occurring in a relevant (irrelevant) document of the training set \(T\) \([33,37]\). Moreover, a fixed number of features renders the algorithm more scalable for larger data sets with many more features, unlike the one used for this experiment. For example, features such as “interact”, “lysat” and “transfect” were ranked above others for their high ranks according to both scores as shown in figure 6. See \([37]\) for more details about the feature extraction procedure.

6 Parameter Search and Robustness

We performed an exhaustive parameter search by training the AICRM on 60 balanced full-text articles (30 \(P_T\) and 30 \(N_T\) from BC2.5 training) and testing it on the remaining 60

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5 The list of common (stop) words includes 33 of the most common English words from which we manually excluded the word “with”, as we know it to be of importance to PPI.
balanced ones (also 30 \( P_F \) and 30 \( N_F \) from BC2.5 Training) as illustrated in figure [31]. Each run corresponds to a unique configuration of the 6 parameters of the ABCRM. The explored parameter ranges are listed in table 1 and they result in a total of 192500 unique parameter configurations for each experiment. Finally, the parameter configurations were sorted with respect to the resulting F-score measure of performance [8], which is a good measure between precision and recall when applied to balanced data [19].

We compiled the performance of the ABCRM on the entire parameter search space for four distinct experiments: (1) the effect of cell death, (2) using both training sets in contrast to using only the positive set, (3) the importance of the sequential order of articles, and (4) the automatic correction of the initial bias.

In all four experiments, we choose the 50 configurations with highest F-score measure to study the ABCRM performance, because we are interested in identifying the experimental setups that lead to higher robustness to parameter changes. We compare experimental outcomes with the paired student t-test; the null hypothesis is that the two samples are drawn from the same distribution. A p-value < 0.01 rejects the null hypothesis, establishing a statistical distinction between the data drawn from two experimental setups—in our case, the data from each experiment are the top 50 F-score values obtained. The first two experiments were initially tested [36] to choose the best experimental set up and compare it with two additional experiments [38] that are discussed in this paper.

### 6.1 Cell Death

The first experiment aims to study the effect of cell death on immune memory and classification performance. In this experiment we compare the top 50 parameter configurations according to F-score obtained using cell death (exp 1.1) to those with no cell death (exp 1.2)—while training on both self and nonself documents. We observe a notable difference in classification performance that we validate statistically (according to the criteria above) to show that using cell death improves the performance (see Fig. 8)—regardless of whether the algorithm is trained on just relevant or on both relevant and irrelevant documents (see below). Therefore we conclude that cell death, which helps in the forgetting of useless features and focuses on more recent and frequent ones, improves classification performance, which suggests that it is important for immune memory in the T-Cell cross-regulation model.

### 6.2 Training on Self and Nonself

The second experiment is conducted to show if we can rely solely on the positive set for classification, or if the performance can be improved by training on both positive and negative sets. We compare the top 50 parameter configurations according to F-score obtained using training on positive only or PU learning (experiments 2.1 and 2.2), to the previous experiments (1.1 and 1.2). This way we compare training on positive documents only, with and without cell death. The results show that using both training sets always (significantly) improves the robustness of classification performance (see Fig. 8). Although the top performance obtained for 1.1 (training on both classes with cell death) and 2.1 (training on positive documents with cell death) is equivalent with F-Score=0.85 (see table 2), the robustness as measured by the performance of the top 50 parameter sets is significantly lower for experiment 2.1 (see figure 8).

### 6.3 Sequence Order

The third experiment aims to establish how much the sequence order of processing documents impacts performance. In particular, we test if preserving the original temporal order of biomedical documents results in better performance, as this would indicate that the ABCRM can use its sequence-dependent dynamics to track the natural concept or topical drift and thus improve classification. Therefore, we compared the performance of the ABCRM when tested on a self...

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### Table 1 Parameter ranges used for optimizing the ABCRM

| Parameter | Range | Step |
|-----------|-------|------|
| \( E_0 \) | [1.7] | 1 |
| \( R_0 \) | [3,12] | 1 |
| \( R_1 \) | [3,12] | 1 |
| \( d_E \) | [0.0,0.4] | 0.1 |
| \( d_R \) | [0.0,0.4] | 0.1 |
| \( n_A \) | [2,22] | 2 |

### Table 2 Performance and parameters of top classifiers in experiment 1 regarding cell death and experiment 2 regarding training data.

| Exp. | F-Score | \( E_0 \) | \( R_0 \) | \( R_1 \) | \( d_E \) | \( d_R \) | \( n_A \) |
|------|---------|-------|-------|-------|------|------|-------|
| 1.1  | 0.85    | 2     | 11    | 10    | 0.3  | 0.2  | 18    |
| 1.2  | 0.83    | 1     | 12    | 5     | 0.0  | 0.0  | 18    |
| 2.1  | 0.85    | 1     | 17    | 8     | 0.1  | 0.0  | 8     |
| 2.2  | 0.75    | 2     | 12    | 6     | 0.0  | 0.0  | 18    |

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8 Notice that this parameter search on the provided labeled training data uses only the information available to the teams participating in Biocreative 2.5 challenge, and none of the test data whose labels were revealed post-challenge.

9 F-score = \( \frac{2 \times \text{Precision} \times \text{Recall}}{\text{Precision} + \text{Recall}} \), where Precision = \( \frac{TP}{TP + FP} \) and Recall = \( \frac{TP}{TP + FN} \). True Positives (TP) and False Positives (FP) are the classifier’s correct and incorrect predictions for relevant documents, while True Negatives (TN) and False Negatives (FN) are the correct and incorrect predictions for irrelevant documents.
The first two experiments result in four experimental setups:
1.1) training on both sets with cell death (red),
2.1) PU learning with cell death (green),
1.2) training on both sets with no cell death (blue),
and 2.2) PU learning with no cell death (yellow) are clearly distinguishable for the top 50 configurations of each experiment on the plot on the left. On the right, the horizontal lines represent the mean, the boxes represent 95% CI, and the whiskers represent standard deviation of F-scores from the top 50 parameter configurations.

Table 3 Performance and parameters of top classifiers in experiments 1.1=3.1=4.1, 3.2 and 4.2.

| Exp. | F-Score | $E_0$ | $R_k^0$ | $R_k^1$ | $d_n$ | $d_n$ | $n_x$ |
|------|---------|-------|---------|---------|-------|-------|-------|
| 1.1  | 0.85    | 2     | 1      | 10      | 0.3   | 0.2   | 18    |
| 3.2  | 0.85    | 2     | 7      | 6       | 0.0   | 0.0   | 20    |
| 4.2  | 0.86    | 3     | 8      | 7       | 0.2   | 0.1   | 14    |

In the case of shuffled sets, we produced 8 runs with distinct random document orderings; in those cases, performance is represented by central tendency.

The results of this experiment are summarized in figure 8. The second two experiments result in 5 experimental outcomes. To the left we show the top 50 parameter configurations ranked in terms of F-score for experimental setups 1.1=3.1=4.1 (red circles), 3.2 (blue pluses), 3.3 (blue crosses), 3.4 (blue diamonds), and 4.2 (green triangles). To the right we show the mean (line), 95% CI (boxes), and standard deviation (whiskers) of F-scores for the top 50 parameter configurations.

The performance of the top 50 classifiers for experimental setups 3.2, 3.3, and 3.4 is statistically indistinguishable from each other, but is significantly lower than the performance of the top 50 classifiers for experimental setup 1.1. This means that there is indeed a conceptual drift in the Biocreative 2.5 article data stream, and the ABCRM can track it better (and in a more robust manner) when publication date is used as the sequence for processing articles than when the temporal order of articles is shuffled. This also suggests that the process of T-Cell cross-regulation in the IS, as modeled here, can track changing nonself pathogens.

It should be noted that in this experiment, the partitioning of training and test data was done according to the timestamp of documents. Therefore, the documents in the test set were published after all documents in the training set. Therefore, even in the shuffled training and test sets (experimental setup 3.3), there is some preservation of temporal order. In future work we will explore experimental setups where the training and test sets are drawn from the same time-stamp distribution to better understand the effects of concept drift and how well our model can track it.

### 6.4 Initial Bias

In the fourth experiment we test the effect of the initial biases introduced when features are first encountered. The initial biases of regulatory T-cells injected in the dynamics for a new feature $f$, depend on whether the first document $d$ where the feature is encountered is labeled irrele-
vant/unknown ($R_0^e$) or relevant ($R_0^{re}$). Since features will occur in both relevant and irrelevant articles, this initial bias for a feature could be detrimental, as a feature most associated with one class could be first encountered on a document of the opposite class. Therefore, it is important to test if the dynamics of the four reactions and APC feature co-presentation that define the ABCRM can self-correct such erroneous biases. To perform this test, we altered the ABCRM algorithm such that T-cells are incremented appropriately every time a feature occurs in a document, and not just the first time the feature occurs (as the canonical algorithm does). Specifically, every time a feature $f$ occurs in a document $d$, we increment $E_f = E_f + E_0$ and $R_f = R_f + R_0^r$ if $d$ is labeled relevant and $R_f = R_f + R_0^r$ if $d$ is labeled irrelevant or unlabeled. We label this experimental set up 4.2, which was conducted with cell death and training on both positive and negative documents.

The results of this experiment are also summarized in figure 9. The performance of top classifiers obtained for experimental setups 4.1 (same as 1.1 and 3.1 that are trained on both training sets using cell death) and 4.2 (incremental experimental setup) is shown in table 3. While the best overall classifier is obtained with experimental setup 4.2, the performance of both setups is statistically indistinguishable. Indeed, using the paired student t-test as described above, we cannot reject the null hypothesis claiming that both distributions of F-scores were drawn from a similar distribution. Therefore, we conclude that this modification does not improve the performance of the ABCRM on the Biocreative data set, thus showing that the initial bias can be corrected by the ABCRM collective dynamics and does not require incrementing T-cells for all new features. Because features most associated with a given class tend to co-occur in text with other features most associated with the same class, they will also tend to be co-presented in APC and thus the relevant T-cells will proliferate with similar rates. Therefore, the dynamics of the ABCRM can self-correct initial erroneous biases from the natural textual co-occurrence of features. This shows that T-Cell cross-regulation as modeled here can self-correct initial antigen misclassification by the IS, assuming that antigens from one class (self/nonself) tend to co-occur with antigens from the same class.

7 Validation and Conclusions

To test the ABCRM on the full, unbalanced test set of the Biocreative challenge (see figure 5), thus establishing its merit as a bio-inspired biomedical literature mining classifier, we adopted the best parameter configuration from the canonical ABCRM (experimental setup 1.1=3.1=4.1, see table 3) obtained from the parameter search described above. We compared the ABCRM classifier with the multinomial Naive Bayes (NB) with boolean attributes, one of the top Naive Bayes implementations for spam detection [24], and the publicly available SVM$^{light}$ implementation of SVM applied to normalized feature counts [9]. The SVM$^{light}$ was used with its default parameter settings [9]. All classifiers were tested on the same features obtained from the same data.

Since the F-score and Accuracy are not very reliable for evaluating unbalanced classification [19], we also use the Area Under the interpolated precision and recall Curve (AUC) and Matthew’s Correlation Coefficient (MCC). The results are listed in table 4, which also includes the central tendency of the results of all systems submitted by all Biocreative 2.5 participating teams [34,37]. It should be noted that the ABCRM, NB, and SVM classifiers we tested here, used only single-word features because we wish to establish the feasibility of the method. In contrast, most classifiers submitted to the Biocreative 2.5 challenge (including another method from our group which was one of the top-performing classifiers [37]) used more sophisticated features such as bigrams and problem-specific entities. Therefore, it is not surprising that these methods as tested here performed under the mean of the challenge. Our goal was to establish the ABCRM as a new bio-inspired text classifier to be further improved in the future with more sophisticated features. When we compare its performance to NB and SVM on the exact same single-word features, the results are encouraging. Indeed, based on the given measures, while the SVM out-performed the ABCRM, the latter out-performed NB. Therefore, the dynamics of T-Cell cross-regulation lead to a competitive collective classification of biomedical articles, which we intend to develop further.

In future work we will pursue additional experiments to study concept drift, namely by investigating the ability to simultaneously train and classify documents. Given the sequence-dependent dynamics entailed by our model, there is no reason to present all test data to the cellular interaction dynamics, only after processing all training data. The model affords various possible schedules of document processing that mix training and test data which could lead to better performance. Indeed, the immune system is constantly exposed

|               | ABCRM | NB   | SVM  | Mean | StDev | Med. |
|---------------|-------|------|------|------|-------|------|
| **Precision** | 0.22  | 0.14 | 0.24 | 0.38 | 0.68  |      |
| **Recall**    | 0.65  | 0.71 | 0.94 |      |       |      |
| **F-score**   | 0.33  | 0.24 | 0.36 | 0.39 | 0.14  | 0.38 |
| **Accuracy**  | 0.71  | 0.52 | 0.74 | 0.67 | 0.30  | 0.84 |
| **AUC**       | 0.34  | 0.19 | 0.46 | 0.43 | 0.17  | 0.44 |
| **MCC**       | 0.24  | 0.13 | 0.31 | 0.31 | 0.19  | 0.33 |

Table 4: F-Score, Accuracy, AUC and MCC performance of various classifiers when training on the balanced training set of articles and testing on the full unbalanced Biocreative 2.5 test set. Also shown is the central tendency and variation of all systems submitted to Biocreative 2.5.
to self antigens (training data), and even pathogens that may be stored in long lived plasma cells and memory B-cells.

In conclusion, we observed that our method uses cell death to enhance immune memory and forget older features while focusing on more recent and frequent ones. We proved that our algorithm is capable of classification when trained on relevant features only, however the performance can be improved when trained on both classes. We also observed that algorithm adapts to the initial bias of T-cell populations generated for new features, and it performs best when tested on a sequence of articles ordered by publication date—showing that it can track concept drift in the biomedical literature.

These properties of our model also show that T-Cell cross regulation is capable of efficient collective classification of nonself antigens and suggest that T-Cell cross-regulation can naturally respond to drift in the pathogen population. Therefore T-Cell cross-regulation defined by the 4 reaction rules and co-presentation of features in APC can be seen as an effective general principle of collective classification available to populations of cells. Clearly, there is still much to do to improve the model. For biomedical literature mining applications, we need to test it with more sophisticated features (as top classifiers in the field do). For our goal of understanding T-Cell cross-regulation in the IS, we need to understand better how memory is sustained in the collective cellular dynamics; for instance, how to sustain regulatory T-Cells, which keep memory of self, in the dynamics even in the presence of very unbalanced scenarios where there are many more self or nonself instances.

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