Core–Shell Crystals of Porous Organic Cages

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Abstract: The first examples of core–shell porous molecular crystals are described. The physical properties of the core–shell crystals, such as surface hydrophobicity, CO$_2$/CH$_4$ selectivity, are controlled by the chemical composition of the shell. This shows that porous core–shell molecular crystals can exhibit synergistic properties that out-perform materials built from the individual, constituent molecules.

The preparation of new functional porous materials is an important goal in materials chemistry, with potential applications in gas storage, molecular separations, catalysis, and sensing.[1] Established classes of porous materials include extended networks and frameworks such as zeolites,[2] metal–organic frameworks (MOFs),[3] covalent organic frameworks (COFs),[4] and crosslinked polymers.[5] More recently, porous molecular solids have emerged as a new materials platform.[6] There has been much effort to increase the structural complexity of porous solids to create materials with differentiated or multiple functions, such as mixed-component MOFs[7] and epitaxial MOF thin films.[8] Another approach is to create core–shell porous materials that can integrate multiple functionalities into the core and shell layers.[9]

Core–shell MOF structures can be formed via strong coordination bonds where the outer shell layer is grown epitaxially on the surface of an inner MOF core.[10] This way, the overall material properties can be enhanced by combining different functionalities in the core and shell layers.[11] For example, the integration of a shell crystal with selective gas sorption with a core crystal with high pore volume makes it possible to combine gas selectivity with high gas storage capacity.[12] Also, core–shell nanostructures with an inner core nanoparticle encapsulated by a porous shell have been widely used for heterogeneous catalysis, where the shell material can ensure the accessibility of reactant molecules to the active metal and also improve the selectivity and stability of the catalyst.[13] However, it remains challenging to incorporate functionality in three-dimensional (3D) core–shell porous structures in a modular way, ideally via a simple solution process. The fabrication of core–shell porous solids with a defect-free, crack-free shell layer is also still a challenge.

We have developed a series of porous organic cages (POCs) with properties such as shape-specific molecular sieving,[14] underpinned by computational design methods such crystal structure prediction.[15] A distinguishing feature of POCs is that they can be dissolved in common solvents. This enables a range of processing options that are not available to porous extended networks. For example, cage nanoparticles can be prepared by mixing cage molecules of opposite chirality in solution.[16] Mix-and-match assembly strategies can also be used to make binary and ternary cocrystals.[17]

Here, we develop a simple and efficient method to assemble core–shell POC nanostructures in a modular manner. The synthesis involves the sequential addition of solutions of the $R$ and $S$ cage enantiomers that exploits chiral recognition. This solution-based mixing process yields core–shell cocrystals with exceptional control over particle size and morphology, also allowing control over surface hydrophobicity. Moreover, CO$_2$/CH$_4$ selectivity can be tuned by varying the gas selectivity of the defect-free particle shell. To our knowledge, this is the first example of porous molecular core–shell materials.

The POC molecules were synthesized via [4+6] cyclodimerization reactions.[18] CC3-R (Figure 1a, left) was synthesized from 1, 3, 5-triformylbenzene (TFB) and (1R, 2R)-1,2-dianimocyclohexane ($R$, R-CHDA).[19] In the CC3a crystal form CC3-R packs in a window-to-window fashion to create 3D diamondoid pores connected through the internal cage voids (Brunauer–Emmett–Teller surface area, $SA_{BET}$: 409 m$^2$ g$^{-1}$; Figure 1c, left).[18] The opposite $CC3$ cage enantiomer can be formed using (1S, 2S)-1,2-dianimocyclohexane ($S$, S-CHDA).

An analogous [4+6] cage molecule, CC15-R, can be synthesized from 1,3,5-triacyetylbenzene (TAB) and $R$, $R$-CHDA. CC15-R has 12 methyl groups positioned in the windows of the cage (Figure 1a, middle).[19] By itself, CC15-R does not show the preferred window-to-window packing that is observed for CC3-R because of steric interactions between these methyl groups. However, a quasiaromatic cocrystal of CC3-S and CC15-R does pack in a window-to-window fashion (Figure 1c, middle), as rationalized previously by crystal
Because the methyl groups in TR solutions, taking advantage of the lower solubility product in each case, age particles CC3 and CC3 solutions to see CC19 = (Figure 1c right). 2 and a CC3 (left), acocrystal of CC19 (right). b) Scheme showing the window-to-window packing of homochiral cages (CC3-R and CC19-R) and a quasi-racemic cocrystal of CC3-R and CC15-S. c) Connolly surface area generated using a N probe radius of 1.82 Å to show 3D diamondoid interconnected pore structures for CC3-R (left), a cocrystal of CC3-R and CC15-S (middle), and CC19-R (right).

Figure 1. a) Organic cage molecules; CC3 (left), CC15 (middle) and CC19 (right). b) Scheme showing the window-to-window packing of homochiral cages (CC3-R and CC19-R) and a quasi-racemic cocrystal of CC3-R and CC15-S. c) Connolly surface area generated using a N probe radius of 1.82 Å to show 3D diamondoid interconnected pore structures for CC3-R (left), a cocrystal of CC3-R and CC15-S (middle), and CC19-R (right).

structure prediction. Because the methyl groups in CC15 partially block the cage windows, the (CC3-S, CC15-R) cocrystal becomes selectively porous to H2 but not N2 at 77 K, 1 bar. Another cage molecule with an analogous tetrahedral architecture, first reported by Petryk et al. can be prepared by 2-hydroxy-1, 3, 5-benzenetricarbaldehyde with R, R-CHDA. We will refer to this covalent cage here as CC19 (Figure 1a, right). The disordered hydroxyl groups occupy the four cage windows. CC19-R crystallizes to form a window-to-window packing with 3D diamondoid pores, isostructural with CC3o (Figure 1c right). CC19-R shows permanent porosity to a range of gases and exhibits a type I N2 sorption isotherm with a SA BET of 514 m2 g−1 (Figure S2 in the Supporting Information).

Three different heterochiral cage compositions were used in this study: racemic CC3-R, racemic CC19-R, and quasi-racemic CC3-R, CC15-S. In each case, cage particles were fabricated by simple mixing of the corresponding R and S solutions, taking advantage of the lower solubility product of the racemic or quasi-racemiac materials. All heterochiral cage particles were crystalline and each had the same basic packing mode, as demonstrated by powder X-ray diffraction (PXRD) (Figure S3,4). The similar lattice parameters for the three different compositions suggested the potential for epitaxial growth to create core–shell structures. All cage particles showed uniform, octahedral crystal morphologies (e.g., Figure 2b). The particle size could be controlled systematically in the range 250 nm to 2 μm by varying the mixing temperature (Figure S5). To probe the potential for core–shell structure generation, we first investigated the sequential addition of CC3-R and CC3-S solutions to see whether this would make larger particles by seeded, epitaxial growth, or whether new particles would be nucleated. The particle sizes measured by dynamic light scattering (DLS) and by scanning electron microscopy (SEM) for each addition confirmed that progressively larger particles were formed (Figure S6, Table S1), suggesting epitaxial growth and the possibility of core–shell structure generation by sequential addition of solutions of distinct cages.

Next, we prepared core–shell structures using CC3 and CC19 cage molecules. The schematic structure is shown in Figure 2a; the core molecules are colored purple. Two core–shell crystal systems were prepared: CC3-Rcore/CC19-Rshell and its inverse structure, CC19-Rcore/CC3-Rshell, both using the sequential addition method described above using DCM solutions at 30 °C. The average DLS particle diameters for the core–shell cocrystals, CC3-Rcore/CC19-Rshell and CC19-Rcore/CC3-Rshell, were 744 nm and 721 nm, respectively, as compared to 212 nm and 474 nm for the CC3-R and CC19-R core seed particles (Figure S7, Table S2). This would suggest a CC19-R shell thickness of 266 nm in CC3-Rcore/ CC19-Rshell and a CC3-R shell thickness of 124 nm in CC19-Rcore/CC3-Rshell. The particle size was further verified by SEM, as shown in Figure S8. There was a good agreement between the DLS and SEM measurements. Larger crystals were required to confirm the core–shell morphology by microscopy. We therefore mixed the solutions in CHCl3 at a higher temperature (50 °C), whereupon the average particle size of the core–shell crystals was increased to 3–4 μm, as shown in Figure 2b–d: CC3-R (~2 μm) and CC19-R (1–2 μm) prepared under the same conditions (Figure S9,10). A terraced surface structure was observed by SEM (Figure 2b, Figure S11) indicating the epitaxial growth of the shell. The core–shell samples showed uniform octahedral shape morphologies without any apparent particle aggregation during the formation of the shell.

Since no contrast could be seen between the chemically-similar core and shell by TEM (Figure 2c), the morphologies
of the $\text{CC3-R}_\text{core}/\text{CC19-R}_\text{shell}$ and $\text{CC19-R}_\text{core}/\text{CC3-R}_\text{shell}$ cocrystals were explored by confocal laser scanning microscopy (CLSM). This was possible because $\text{CC19-R}_3$, unlike $\text{CC3-R}_3$, is strongly fluorescent. To visualize the layered core–shell structure, we used $\approx 5$ micrometer-sized core–shell cocrystals prepared in CHCl$_3$ at 60°C. The horizontally sliced confocal image of $\text{CC3-R}_\text{core}/\text{CC19-R}_\text{shell}$ revealed a non-fluorescent inner core ($\text{CC3-R}_3$) encapsulated by a fluorescent outer shell layer ($\text{CC19-R}_3$), as shown in Figure 3c and the corresponding 3D structural model (Movie S1). By contrast, the $\text{CC19-R}_\text{core}/\text{CC3-R}_\text{shell}$ crystals comprise a non-fluorescent $\text{CC3-R}_3$ shell encapsulating a fluorescent core ($\text{CC19-R}_3$) (Figure 3d). The intensity profiles are presented in Figure 3e,f, which correspond to the core–shell crystals shown in the horizontally sliced images (Figure 3c,d). The distance across the crystal is approximately 6 µm for $\text{CC3-R}_\text{core}/\text{CC19-R}_\text{shell}$, and this representative crystal has a non-fluorescent core of approximately 3 µm in diameter and a 1.5-µm-thick shell, as estimated from the fluorescence intensity profiles. The diameter of the $\text{CC19-R}_\text{core}/\text{CC3-R}_\text{shell}$ crystal was 4 µm with a 3 µm fluorescent core and a 500 nm non-fluorescent shell. Z-stack of CLSM images of $\text{CC3-R}_\text{core}/\text{CC19-R}_\text{shell}$ and $\text{CC19-R}_\text{core}/\text{CC3-R}_\text{shell}$ are shown in Figure S12,13. A 3D structural model for $\text{CC3-R}_\text{core}/\text{CC19-R}_\text{shell}$ was constructed based on the z-stack of CLSM analysis (SI, Movie S2).

The structural relationship between the core seed crystals, separate crystals of the shell components, and the core–shell cocrystals was further explored by synchrotron X-ray diffraction. Both $\text{CC3-R}_3$ and $\text{CC19-R}_3$ crystallized in the cubic space group F43m with unit cell parameters of $a = 24.7069$ (1) Å for $\text{CC3-R}_3$ and $a = 24.6914$ (3) Å for $\text{CC19-R}_3$. Lattice parameter matching is important in allowing the growth of the core–shell morphology. The PXRD patterns for $\text{CC3-R}_3$, $\text{CC19-R}_3$, and $\text{CC3-R}_\text{core}/\text{CC19-R}_\text{shell}$ (Figure S14) indicate that the core–shell particles retain a similar crystal packing: the core–shell cage crystals also crystallize with cubic symmetry and window-to-window packing motifs, analogous to $\text{CC3-R}_3$ and $\text{CC19-R}_3$, with a small expansion in the unit cell parameters compared to the individual racemic crystals (Table S3).

$\text{CC3-R}_\text{core}/\text{CC19-R}_\text{shell}$ demonstrates a significantly higher oxygen content as measured by X-ray photoelectron spectroscopy (XPS) due to an outer layer containing hydroxyl groups (oxygen elements), while $\text{CC19-R}_\text{core}/\text{CC3-R}_\text{shell}$ does not (Table S4). Also, the solution UV absorption spectrum for $\text{CC19-R}_3$ shows absorption peaks at 300 and 375 nm. By contrast, a $\text{CC3-R}_3$ solution exhibits no UV adsorption in this region. The absorption peaks for the core–shell, $\text{CC3-R}_{\text{core}}/\text{CC19-R}_{\text{shell}}$, as measured by dispersing the cage particles in the hexane suspension, showed a slight blue shift relative to the $\text{CC19-R}_3$ solution spectrum, while a red shift was observed for the $\text{CC19-R}_{\text{core}}/\text{CC3-R}_{\text{shell}}$ material (Figure S16). The intensity of the fluorescence excitation/emission spectra for $\text{CC19-R}_{\text{core}}/\text{CC3-R}_{\text{shell}}$ was significantly decreased as compared to $\text{CC19-R}_3$, in keeping with a fluorescent core of $\text{CC19-R}_3$ that is encapsulated by a non-fluorescent $\text{CC3-R}_3$ layer (Figure S17).

This synthetic method can also be applied to other cage molecules: for example, a core–shell crystal with racemic $\text{CC3}$ as the core and quasi-racemic $\text{CC3-RC15}$–$\text{S}$ as the shell was also prepared. The $\text{CC3-R}_3$ core crystals had an average particle size of 1–2 µm, as measured by SEM. Subsequent addition of solutions of $\text{CC3-R}_3$ and $\text{CC15}$–$\text{S}$ formed a shell, creating a $\text{CC3-R}_{\text{core}}/\text{CC15}$–$\text{CC3-R}_{\text{shell}}$ cocrystal with an average diameter of 3 µm (Figure S18, S19).

Core–shell structures can be exploited to control particle surface properties, which are important in applications such as gas storage and separation. Contact angles with water for cage crystals (1–3 µm diameter) gradually increased from $55.68 \pm 2.5^\circ$ ($\text{CC19-R}_3$) to $78.71 \pm 0.80^\circ$ ($\text{CC3-R}_3$) to $83.06 \pm 3.04^\circ$ ($\text{CC3-RC15-S}$) as the constituent cage materials become more hydrophobic (Figure S20). $\text{CC3-R}_{\text{core}}/\text{CC19-R}_{\text{shell}}$ shows a contact angle of $59.71 \pm 6.5^\circ$: that is, very close to the pure, relatively hydrophobic $\text{CC19RC15}$ material (Figure 4a), showing that the shell dominates the surface properties. Likewise, the inverse $\text{CC3-R}_{\text{core}}/\text{CC19-R}_{\text{shell}}$ cocrystal showed a contact angle of $79.01 \pm 3.1^\circ$, close to pure $\text{CC3-R}_3$. The contact angle of $\text{CC3-R}_{\text{core}}/\text{CC15}$–$\text{CC3-R}_{\text{shell}}$ is $83.40 \pm 0.87^\circ$: this material is slightly more hydrophobic due to the methyl groups in $\text{CC15}$.

Gas sorption analysis was carried out for both $\text{CC19-R}_{\text{core}}/\text{CC3-R}_{\text{shell}}$ and $\text{CC3-R}_{\text{core}}/\text{CC19-R}_{\text{shell}}$ core–shell materials. $\text{N}_2$ sorption measurements at 77 K showed very similar Type I isotherms for both core–shell cage cocrystals.
In conclusion, we have successfully prepared core–shell cage crystals. The surface chemistry is controlled by the functionality in the shell layer, thus allowing control over surface hydrophobicity. Hence, CC3, which was shown previously to have multiple practical applications,[21] can be rendered either more hydrophobic or more hydrophilic, depending on the choice of shell. A CC3-RScore/CC19-Rsshell material was shown to have a synergistic combination of CO2 sorption capacity and CO2/CH4 selectivity that surpassed either of individual constituent cages. This approach has the potential to open up new applications for porous organic cages. To give one example, CC3 crystals have been incorporated into polymers of intrinsic microporosity to form organic mixed matrix membranes (MMMs) for molecular sieving.[22] In MMMs, a good interaction between the polymer and filler components is essential, and this core–shell approach offers a new strategy for optimizing the polymer-cage particle interface. It is also possible that cage shells could be deposited from solution onto porous crystals of other materials such as MOFs, COFs and zeolites, providing that conditions can be identified to promote epitaxial growth.

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Conflict of interest

The authors declare no conflict of interest.

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[1] S. Kitagawa, R. Kitaura, S-i. Noro, Angew. Chem. Int. Ed. 2004, 43, 2334–2375; Angew. Chem. 2004, 116, 2388–2430.
[2] P. A. Wright, G. M. Pearce, Zeotites and Catalysts, Wiley-VCH, Weinheim, 2010, chap. 7, p. 171.
[3] G. Férey, Chem. Soc. Rev. 2008, 37, 191–214.
[4] C. S. Diercks, O. M. Yaghi, Science 2017, 355, 923–931.
[5] A. Thomas, Angew. Chem. Int. Ed. 2010, 49, 8328–8344; Angew. Chem. 2010, 122, 8506–8523.
[6] a) G. Zhang, M. Mastalerz, Chem. Soc. Rev. 2014, 43, 1934–1947; b) T. Hasell, A. I. Cooper, Nat. Rev. Mater. 2016, 1, 16053.
[7] a) S. M. Cohen, Chem. Rev. 2012, 112, 970–1000; b) H. Deng, C. J. Doonan, H. Furukawa, R. B. Ferreira, J. Towne, C. B. Knobler, B. Wang, O. M. Yaghi, Science 2010, 327, 846–850.
[8] P. Falkaro, K. Okada, T. Harra, K. Ikigaki, Y. Tokudome, A. W. Thornton, A. J. Hill, T. Williams, C. Doonan, M. Takahashi, Nat. Mater. 2016, 16, 342–348.
[9] a) M. L. Foo, R. Matsuda, S. Kitagawa, Chem. Mater. 2014, 26, 310–322; b) C. V. McGuire, R. S. Forgan, Chem. Commun. 2015, 51, 5199–5217; c) Y. Peng, M. Zhao, B. Chen, Z. Zhang, Y. Huang, F. Dai, Z. Lai, X. Cui, C. Tan, H. Zhang, Adv. Mater. 2017, 1705454; d) Y. Bouizi, I. Diaz, L. Rouleau, V. P. Valtchev, Adv. Funct. Mater. 2005, 15, 1955–1960.
[10] a) S. Furukawa, K. Hirai, K. Nakagawa, Y. Takashima, R. Matsuda, T. Tsuruoka, M. Kondo, R. Haruki, D. Tanaka, H. Sakamoto, S. Shimomura, O. Sakata, S. Kitagawa, Angew. Chem. Int. Ed. 2009, 48, 1766–1770; Angew. Chem. 2009, 121, 1798–1802; b) Y. Yoo, H.-K. Jeong, Cryst. Growth Des. 2010, 10, 1283–1288.
[11] a) K. Hirai, S. Furukawa, M. Kondo, H. Uehara, O. Sakata, S. Kitagawa, Angew. Chem. Int. Ed. 2011, 50, 8057–8061; Angew. Chem. 2011, 123, 8207–8211; b) K. Hirai, S. Furukawa, M. Kondo, M. Meilikhov, Y. Sakata, O. Sakata, S. Kitagawa, Chem. Commun. 2012, 48, 6472–6474.
[12] T. Li, J. E. Sullivan, N. L. Rosi, J. Am. Chem. Soc. 2013, 135, 9984–9987.
[13] a) S. Schmitt, M. Silvestre, M. Tsotsalas, A.-L. Winkler, A. Shahnas, S. Grosjean, F. Laye, H. Gliemann, J. Lahann, S. Bräse, M. Franzreb, C. Woll, ACS Nano 2015, 9, 4219–4226; b) J.-K. Sun, W.-W. Zhan, T. Akita, Q. Xu, J. Am. Chem. Soc. 2015, 137, 7063–7066; c) L. Qiu, R. McCaffrey, Y. Jin, Y. Gong, Y. Hu, H. Sun, W. Park, W. Zhang, Chem. Sci. 2018, 9, 676–680.
[14] T. Mitra, K. E. Jelfs, M. Schmidtmann, A. Ahmed, S. Y. Chong, D. J. Adams, A. I. Cooper, Nat. Chem. 2013, 5, 276–281.
[15] a) J. T. A. Jones, T. Hasell, X. Wu, J. Baesa, K. E. Jelfs, M. Schmidtmann, S. Y. Chong, D. J. Adams, A. Trewin, F. Schifftman, F. Cora, B. Slater, A. Steiner, G. M. Day, A. I. Cooper, Nature 2011, 474, 367–371; b) A. G. Slater, P. S. Reiss, A. Pulido, M. A. Little, D. L. Holden, L. Chen, S. Y. Chong, B. M. Alston, R. Clowes, M. Haranczyk, M. E. Briggs, T. Hasell, G. M. Day, A. I. Cooper, ACS Cent. Sci. 2017, 3, 734–742.
[16] T. Hasell, S. Y. Chong, K. E. Jelfs, D. J. Adams, A. I. Cooper, J. Am. Chem. Soc. 2012, 134, 588–598.
[17] T. Hasell, S. Y. Chong, M. Schmidtmann, D. J. Adams, A. I. Cooper, Angew. Chem. Int. Ed. 2012, 51, 7154–7157; Angew. Chem. 2012, 124, 7266–7269.
[18] T. Tozawa, J. T. A. Jones, S. I. Swamy, S. Jiang, D. J. Adams, S. Shakespeare, R. Clowes, D. Bradshaw, T. Hasell, S. Y. Chong, C. Tang, S. Thompson, J. Parker, A. Trewin, J. Baesa, A. M. Sowin, A. Steiner, A. I. Cooper, Nat. Mater. 2009, 8, 973–978.
[19] M. Petryk, J. Szmykowiak, B. Gierczuk, G. Spolnik, L. Popenda, A. Janiak, M. Kwit, Org. Biomol. Chem. 2016, 14, 7495–7499.
[20] J. Chun, S. Kang, N. Park, E. J. Park, X. Jin, K.-D. Kim, H. O. Seo, S. M. Lee, H. J. Kim, W. H. Kwon, Y.-K. Park, J. M. Kim, Y. D. Kim, S. U. Son, J. Am. Chem. Soc. 2014, 136, 6786–6789.
[21] A. Kewley, A. Stephenson, L. Chen, M. E. Briggs, T. Hasell, A. I. Cooper, Chem. Mater. 2015, 27, 3207–3210.
[22] A. F. Bushell, P. M. Budd, M. P. Attfield, J. T. A. Jones, T. Hasell, A. I. Cooper, P. Bernardo, F. Bazzarelli, G. Clarizia, J. C. Jansen, Angew. Chem. Int. Ed. 2013, 52, 1253–1256; Angew. Chem. 2013, 125, 1291–1294.