Solid-State Conformational Flexibility at Work: Energetic Landscape of a Single Crystal-to-Single Crystal Transformation in a Cyclic Hexapeptoid

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ABSTRACT: We describe the energetic landscape beyond the solid-state dynamic behavior of a cyclic hexapeptoid decorated with four propargyl and two methoxyethyl side chains, namely, cyclo-(Nme-Npa₂)₂, Nme = N-(methoxyethyl)glycine, Npa = N-(propargyl)glycine. By increasing the temperature above 40 °C, the acetonitrile solvate form 1A starts to release acetonitrile molecules and undergoes a reversible single crystal-to-single crystal transformation into crystal form 1B with a remarkable conformational change in the macrocycle: two propargyl side chains move by 113° to form an unprecedented “CH-π zipper”. Then, upon acetonitrile adsorption, the “CH-π zipper” opens and the crystal form 1B transforms back to 1A. By conformational energy and lattice energy calculations, we demonstrate that the dramatic side-chain movement is a peculiar feature of the solid-state assembly and is determined by a backbone conformational change that leads to stabilizing CH···OC backbone-to-backbone interactions tightening the framework upon acetonitrile release. Weak interactions as CH···OC and CH-π bonds with the guest molecules are able to reverse the transformation, providing the energy contribution to unzip the framework. We believe that the underlined mechanism could be used as a model system to understand how external stimuli (as temperature, humidity, or volatile compounds) could determine conformational changes in the solid state.

INTRODUCTION

The dynamic behavior of the biomolecules enables efficient guest recognition and specific substrate conversion in biological processes. The aim of inexhaustible research activity in the field of molecular nanotechnology is the design and synthesis of artificial systems able to combine the recognition abilities of proteins with thermochemical stability.1−3 As peptidomimetic compounds, N-substituted polyglycines (or peptoids, see Figure 1)4 feature useful biological activities and interesting chemical properties both in solution and in the solid state.5 They may represent new motifs on which to base artificial ionophoric antibiotics.6 The biological assays indicated in some cases antifungal activity and no toxicity toward red blood cells.7 Their ion transport abilities in artificial liposomes have often been related to promising cytotoxic activity on human cancer cell lines.8,9 They were used as scaffolds for hybrid glycopeptoid systems, with outstanding multivalent effects in α-mannosidase inhibition.10 They were also tested as phase-transfer catalysts with performances comparable to crown ethers.11 Recently, the discovery of conformational isomerism in cyclic peptoids suggests the application of peptoids in asymmetric synthesis and chiral recognition.12,13 Different from with peptides, in peptoids, the side chains are attached to the nitrogen atoms of the oligoamide backbone (Figure 1).14,15 Thus, CH₂-OC hydrogen bonds take the place of ordinary NH···OC bonds in peptides.16,17

Recently, our group evidenced how environmental changes (temperature, humidity, gas pressure, etc.) may trigger the dynamic behavior of cyclic peptoids in the solid state.18 We established the solvatomorphic behavior of compound 1, a cyclic hexapeptoid decorated with four propargyl and two methoxyethyl side chains (Figure 2), which led to the discovery of two

Figure 1. Peptide vs peptoid.

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Here, we report the results of conformational and lattice energy studies of the acetonitrile solvate and desolvated crystal forms 1A and 1B, with the aim to investigate the energetic landscape beyond the peculiar dynamic behavior of 1.

Hirshfeld surfaces and energy framework analysis allowed us to analyze and visualize the contributions of intermolecular interactions, as weak CH···OC hydrogen bonds and CH-π interactions, toward the crystal packing in 1A and 1B.

Periodic quantum chemical calculations, based on HF-3c Hamiltonian, offered the possibility to further probe the structure–energy relationship in the reversible SCSC transformation from 1A to 1B.

### MATERIALS AND METHODS

For all calculations, the lengths of X–H bonds were normalized using the standard X–H distances (X = C, O, N). Thus, the reported X–H distances and X···H contacts refer to the recalculated structures and are not equal to those calculated from the original cif files.

#### Gas-Phase Energy Optimization.
Gas-phase energy optimization for the cyclopeptoid molecule in 1A and 1B were performed with Gaussian09,[22] using a polarized valence triple-zeta basis set (cc-pVTZ) and B97-D3 method, a Grimme’s modified density functional,[23] which includes the D3 empirical dispersion correction.[24]

Least-squares overlay of pairs of structures was performed using Mercury, which also provides root-mean-square deviation (RMSD) values.[25]

#### Hirshfeld Surface and Energy Framework Analysis.
Hirshfeld surface analysis[26–29] was performed with CrystalExplorer 17.5.[30] Intermolecular interactions were calculated with the B3LYP level of theory using the 6-31G(d,p) basis set for all atoms and include electrostatic, polarization, dispersion, and exchange-repulsion terms, as reported by Turner et al.[28]

To provide a graphical representation of the intermolecular interaction energies, CrystalExplorer was used to represent the energy frameworks.[32]

#### Lattice Energy Calculations.
Lattice energy calculations were performed using the CLP-Pixel package with a Pixel condensation level of 4.[33–35] The total lattice energy is partitioned into its Coulombic, polarization, dispersion, and repulsion contributions. In CLP-Pixel, the Coulombic terms are handled by Coulomb’s law and calculated by a numerical integral equivalent to the standard analytical form; the polarization terms are calculated in the linear dipole approximation, with the incoming electric field acting on local polarizationities and generating a dipole with its associated dipole separation energy; dispersion terms are simulated in London’s inverse sixth-power approximation, involving ionization potentials and polarizabilities; repulsion is presented as a modulated function of wave function overlap.

#### Periodic HF-3c Calculations.
The energy values of 1A, 1A*, and 1B were calculated using the developer version of the software Crystal14.[36] All of the calculations were performed using the low-cost empirical corrected HF-3c method[37] in the HF-3c-(0.27 s 8) scaled version, refined by some of us to compute fast and accurate molecular crystal properties.[37] Using this methodology, we run the Hartree–Fock calculations with the minimal quality basis set called MINIX[38] and three semiempirical corrections to the HF energy: (i) the inclusion of long-range London dispersion interactions, (ii) the basis set superposition error (BSSE), and (iii) short-range basis set incompleteness (SRB).

Full optimization of the crystal structures, including the unit cell parameters, was performed at Γ point using the analytical gradient method by upgrading the Hessian with the Broyden–Fletcher–Goldfarb–Shanno (BFGS) algorithm.[41–43]Crystal14 default tolerances were applied: 10−7 Ha for the energy convergence during the geometry optimization; 10−6 Ha as truncation criteria for bioeletronic integrals of the Coulomb series and 10−14 Ha as that of the exchange series. Choosing to run the calculations using just 1 k point (shrinking factor 1) was justified after comparing calculations with 10 and 30 k
RESULTS AND DISCUSSION

Molecular Structures: Comparison between 1A and 1B. Both X-ray molecular structures of 1 in crystal forms 1A and 1B show a crystallographic inversion center and exhibit a cctcct (c = cis; t = trans) sequence of distorted amide bonds (Table S1 in the Supporting Information).

In crystal form 1A, two methoxethyl and two propargyl side chains point vertically up and down with respect to the macrocycle plane, while the two remaining propargyl side chains extend horizontally in the equatorial plane. In crystal form 1B, there are two crystallographically independent cyclopeptoid molecules, named type I and type II molecules (in blue and green, respectively, in Figure 3). Type I molecules (blue) show only slight differences with respect to form 1A. Type II molecules (green) feature vertical methoxethyl side chains, while all propargyl side chains extend horizontally in the equatorial plane.

Thus, cis propargyl \( \chi_2 \) torsion angle changes from 118.1(2) to 129.4(2)° during the transformation from crystal form 1A to 1B and vice versa (Table S1 and Figure S1 in the Supporting Information).

Besides the remarkable change of orientation of two propargyl side chains, also a backbone conformational change occurs. Type II molecules (green in Figure 3) have a more extended rectangular shape with respect to type I molecules (blue in Figure 3), as is evident considering the distance between and nitrogen atoms at the corners: N1···N2 distance is 6.119(2) Å for type I molecules and increases to 6.960(2) Å for type II molecules.

Indeed, the backbone conformation in type II molecules adapts to the ideal type I \( \beta \)-turn structure of polypeptides.

We would like to point out that the shape difference between type I and II molecules may be related to the presence of shorter CO···CO distances in type I molecules: C6···O1 is 2.965(2) Å in type I molecules and 3.068(2) Å in type II molecules, C11···O1 is 3.169(2) Å in type I molecules and 3.615(2) Å in type II molecules, and C11···O2 is 3.260 Å in type I molecules and 3.480 Å in type II molecules (Figure 4 and Table S2 in the Supporting Information).

The optimized gas-phase molecular structure of type II molecules in form 1B have equivalent energy and differ from the X-ray molecular structures by RMSDs of 0.4507 and 0.4271 Å, respectively (Figure S2a,b in the Supporting Information).

The optimized gas-phase molecular structures of 1A and type I molecules in form 1B have equivalent energy and differ from the X-ray molecular structures by RMSDs of 0.4507 and 0.4271 Å, respectively (Figure S2a,b in the Supporting Information). The orientation of the moving cis propargyl side chains is completely different, being vertical and not anymore horizontal with respect to the macrocycle plane (Figure S2c,d in the Supporting Information). This suggests that the side-chain movement is a peculiar feature of the solid-state assembly and is determined by the possibility for the propargyl groups to form a mutual CH···π zipper in the solid state.

X-ray Crystal Structures, Hirshfeld Surface Analysis, and Lattice Energy Calculations. Hirshfeld surface analysis and energy calculations allowed us to detect quantitatively the main structural features of the crystal packing in crystal forms 1A and 1B. In Tables 1 and 2, the intermolecular distances (Å), angles (°), and interaction energies (kJ/mol) in the host framework of crystal forms 1A and 1B, respectively, as calculated using PIXEL and CrystalExplorer, are reported. The Hirshfeld surface analysis highlights the short contacts between the cyclic peptoid molecules: red zones correspond to the shortest intermolecular distances. As for proteins, molecular interactions in peptoid crystals may be classified as backbone-to-backbone interactions, backbone-to-side chain interactions, and side-chain-to-side chain interactions.
vertical cis propargyl side chain and the carbonyl oxygen atom of the peptoid backbone (C10−H10···O1 2.08 Å; O1···H10−C10 161.6°). In this way, the cyclopeptoid molecules stack on top of each other to form columns that develop along the shortest c-axis through vertical propargyl side chains, which act as pillars.

Moreover, cyclic peptoids interact along the a-axis through backbone-to-side chain CH···OC interactions between the carbonyl oxygen atoms and the methylene hydrogen atom of the trans methoxyethyl side chains (C13−H13B···O3 2.27 Å; O3···H13B−C13 141.2°; Motif 2; Figure 6 and Table 1).

Further, in the last motif (Motif 3; Figure 6 and Table 1), the macrocycles interact in the bc-plane by means of backbone-to-side chain interactions and side chain-to-side chain CH···OC interactions.
Table 1. List of Intermolecular Distances (Å), Angles (°), and Interaction Energies (kJ/mol) in Crystal Form 1A

| motif       | D–H···A | H–A (Å) | D–H–A (deg) | symm. op. | centroid distance (Å) | $E_{\text{Coord}}$ | $E_{\text{Pol}}$ | $E_{\text{Dip}}$ | $E_{\text{Rep}}$ | $E_{\text{Total}}$ |
|-------------|---------|---------|-------------|-----------|-----------------------|---------------------|----------------|----------------|----------------|-----------------|
| 1           | C10–H10–O1 | 2.08    | 161.6       | $x, y, -1+z$ | 8.500               | -78.0               | -30.5          | -96.5          | 111.6          | -93.6           |
| C14–H14B–C10| 2.71    | 120.2   | $x, y, 1+z$  |           |                      | -70.3               | -17.3          | -98.2          | 116.6          | -100.5          |
| 2           | C13–H13B–O3 | 2.27    | 141.2       | $-1+x, y, z$ | 9.773               | -30.4               | -15.5          | -35.9          | 40.8           | -41.0           |
|             |          |         |             |           |                      | -27.6               | -13.9          | -39.1          | 39.9           | -49.0           |
| 3           | C8–H8A–O2 | 2.33    | 138.4       | $1-x, -1/2+y, 1/2-z$ | 11.309             | -23.1               | -9.1          | -35.4          | 32.7           | -35.0           |
| C5–H5–O4    | 2.38    | 129.7   | $1-x, 1/2+y, 1/2-z$ |          |                      | -21.4               | -5.5          | -39.4          | 34.1           | -40.0           |

Table 2. List of Intermolecular Distances (Å), Angles (°), and Interaction Energies (kJ/mol) in Crystal Form 1B

| motif       | D–H···A | H–A (Å) | D–H–A (deg) | symm. op. | centroid distance (Å) | $E_{\text{Coord}}$ | $E_{\text{Pol}}$ | $E_{\text{Dip}}$ | $E_{\text{Rep}}$ | $E_{\text{Total}}$ |
|-------------|---------|---------|-------------|-----------|-----------------------|---------------------|----------------|----------------|----------------|-----------------|
| 1–I         | C10B–H10B–O1B | 2.03    | 167.5       | $x, y, -1+z$ | 8.472               | -78.9               | -31.6          | -94.7          | 112.1          | -93.1           |
|             |         |         |             |           |                      | -71.4               | -18.6          | -97.6          | 116.7          | -102.1          |
| 2–I         | C8B–H8B–O2B | 2.24    | 138.5       | $-x, 1/2+y, 1/2-z$ | 11.015             | -20.9               | -8.5          | -35.7          | 33.9           | -31.2           |
| C5B–H5B–O4B | 2.56    | 112.9   | $-x, -1/2+y, 1/2-z$ |           |                      | -19.1               | -5.4          | -40.6          | 34.8           | -38.0           |
| 3–II        | C7A–H7B–O2A | 2.13    | 169.9       | $1-x, -1/2+y, 1/2-z$ | 11.015             | -32.9               | -15.4          | -53.7          | 49.8           | -52.2           |
|             |         |         |             |           |                      | -30.1               | -10.5          | -59.1          | 51.1           | -59.6           |
| 4–II        | C10A–H10A–O4A | 2.24    | 126.7       | $x, y, -1+z$  | 8.472               | -31.7               | -9.8          | -47.9          | 38.8           | -50.7           |
| C8A–H8B–C9A | 2.71, 2.78| 163.9, 141.1| $x, y, 1+z$  |           |                      | -28.6               | -5.8          | -51.3          | 36.9           | -56.5           |
| 5–II        | C2A–H2B–O3B | 2.16    | 151.1       | $1-x, y, z$   | 8.944               | -56.9               | -21.2          | -61.1          | 69.2           | -70.0           |
| C2B–H2D–O3A | 2.23    | 154.6   | $1+x, y, z$  |           |                      | -54.3               | -15.3          | -66.1          | 72.9           | -81.2           |
| C13A–H13B–O3B | 2.50    | 137.1   | $-1+x, y, z$ |           |                      |                    |                |                |                |                |
| C13B–H13D–O3A | 2.54    | 135.8   |             |           |                      |                    |                |                |                |                |
| C13B–H13D–C14 | 2.72    | 144.5   |             |           |                      |                    |                |                |                |                |
| C8A–H8A–C5B | 2.73    | 130.8   |             |           |                      |                    |                |                |                |                |

Energy values calculated by PIXEL are displayed in plain text and those calculated by CrystalExplorer are in bold face. Interactions between host framework and acetonitrile guest molecules are indicated with an “s”.

Intermolecular interactions (C8–H8A···O2 2.33 Å; O2···H8A–C8 138.4° and C5–H5···O4 2.38 Å; O4···H5–C5 129.7°). Regarding the acetonitrile guest molecules, they occupy the voids between the macrocycle columns, forming channels parallel to the c-axis. Acetonitrile molecules are weakly bound to the host framework (Motif 4s and Motif 5s; Figure 6 and Table 1): the main interactions involve the acetonitrile methyl hydrogen atoms and backbone oxygen atoms (C17–H17C–O3 2.36 Å; O3–H17C–C17 141.8°; C17–H17B–O2 2.60 Å; O3–H17C–C17 150.2°) and the acetonitrile nitrogen atom and the methylene hydrogen atom of the trans methoxyethyl side chain (C13–H13A···N4 2.64 Å; C13–H13A···N4 124.3°). Moreover, acetonitrile molecules are mutually linked by means of the CH···π interactions (C16–H17A 2.72 Å; N4···H17A 2.75 Å; C16–H17A–C17 150.5°).

Intermolecular Interactions in Crystal Form 1B. In crystal form 1B, there are two crystallographically independent molecules (type I and type II molecules) in the asymmetric unit and therefore two Hirshfeld surfaces are considered. Intermolecular interactions may be divided into type I–type I, type II–type II, and type I–type II interactions.

The solid-state assembly of type I molecules is very similar to that observed in crystal form 1A: the Hirshfeld surfaces show the same red zones, corresponding to strong columnar interactions via CO–···HC hydrogen bonds. In Motif 1–I (Figure 7 and Table 2), the main interactions are represented by backbone-to-side chains CO–···HC interactions, involving the backbone carbonyl atom and the vertical propargyl side chains (C10B–H10B···O1B 2.03 Å; C10B–H10B···O1B 167.5°).

Motif 2–I is analogous to Motif 3 in crystal form 1A and characterized by the arrangement of the cyclic peptides in the b-plane through CH···OC interactions between the vertical propargyl side chains and the carbonyl oxygen atoms (C8B···H8D···O2B 2.24 Å; O2B···H8D–C8B 138.5°).

Notably, the columnar arrangement of type II molecules (Motif 4–II; Table 2 and Figure 7) is not the main interaction for type II molecules but is almost energetically equivalent to the Motif 3–II (Table 2 and Figure 7). This corresponds to the
layered arrangement of peptoid type II molecules (in the \(bc\)-plane) through backbone-to-backbone CH···OC interactions and side-chain CH–π interactions (Motif 3 II in Figure 7). In detail, backbone-to-backbone CH···OC interactions (C7A–H7B···O2A 2.13 Å; O2A···H7B–C7A 169.9°) along the short side of the macrocycle were recently identified as a recurring assembly mode for cyclic hexapeptoids. Moreover, this assembly mode allows the formation of a CH–π zipper (C8A–H8B···C9A 2.71 Å; C8A···H8B–C10A 2.78 Å; C8A–H8B···C9A 163.9°; C8A–H8B···C10A 141.1°) by mutual CH–π interactions between the horizontal propargyl side chains of adjacent columns. These are exactly the propargyl side chains that rotate by 113° during the SCSC transformation!

Regarding the columnar arrangement of type II molecules (Motif 4 II–II; Table 2 and Figure 7), we would like to point out that different from with type I molecules and crystal form 1A, the vertical methoxyethyl side chains work as pillars instead of the propargyl side chains, providing CH···OC interaction between methyl hydrogen atoms and the backbone oxygen atoms (C15A–H15A···O1A 2.47 Å; O1A···H15A–C15A 171.4°).

The interaction between type I and type II molecules in crystal form 1B (Motif 5 I–II; Table 2 and Figure 7) occurs along the \(a\)-axis, thus replacing Motif 2 in crystal form 1A. Side-by-side backbone interactions form a ribbon of alternating type I and type II macrocycles along the \(a\)-axis: type I backbone methylene hydrogen atoms interact with type II carbonyl oxygen atoms (C2A–H2B···O3B 2.16 Å; O3B···H2B–C2A 151.1°) and vice versa (C2B–H2D···O3A 2.23 Å; O3A···H2D–C2B 154.6°). Other backbone-to-side chain interactions and side chain-to-side chain interactions (see Motif 5 I–II in Table 2) help to stabilize the assembly.

Energy Framework Analysis. Energy framework analysis helped greatly to visualize the interaction motifs shown in Figures 6 and 7 and listed in Tables 1 and 2, respectively. In Figures 8–10, the energies between molecular pairs are represented as cylinders joining the centers of mass of the molecules, with the cylinder radius proportional to the magnitude of the interaction energy.

Inspecting Figures 8a and 9a, it is evident that in crystal form 1A the columnar arrangement of cyclic peptoids along the shortest axis (Motif 1) is the dominant motif compared to Motifs 2 and 3.

In crystal form 1B, type I molecules feature the same columnar arrangement of form 1A (see Figure 8b,a, and correspondingly Motif 1 I–I in 1B and Motif 1 in 1A in Tables 2 and 1, respectively), while type II molecules are characterized by a less efficient columnar arrangement (Motif 4 II–II; Table 2), represented by a smaller cylinder radius in Figure 8c.

Thus, the columnar arrangement is the second most favored assembly for type II molecules, the first being the layered arrangement of type II molecules in the \(bc\)-plane (Motif 3 II–II;
Figure 8c and Table 2), characterized by the formation of the CH-π zipper. By inspecting Figure 9, it is evident that interactions between cyclic peptoid molecules along the a-axis in 1A (Motif 2; Table
1) are substituted by stronger interactions between type I and type II molecules (Motif S I–II; Table 2).

Energy framework analysis helped to rationalize the changes in the unit cell volumes and lattice constants due to the SCSC transformation from 1A to 1B.

Upon acetonitrile release, the crystal shrinks and the unit cell volume reduces by 11.6%, the a- and b-axis decrease, respectively, by 8.5 and 3.1%, while the c-axis remains unchanged ($\Delta c = -0.33\%$). The evident anisotropic decrease may be explained on the basis of the crystal packing and energy framework analysis. During the transformation from 1A to 1B:

- The columnar arrangement of the macrocycles along the c-axis is preserved both in 1A and 1B.
- Half molecules of 1A adopt an extended conformation with all propargyl side chains in the macrocycle plane (type II molecules), in particular, the vertical propargyl chains in 1A rotate by 113° and become horizontal to form a CH-$\pi$ zipper in the bc-plane.

Figure 9. Energy frameworks for the host framework in 1A and 1B as viewed along the c-axis. Cylinder radius is set to 80; interaction energies with magnitudes smaller than $-10$ kJ mol$^{-1}$ were omitted.

Figure 10. Energy frameworks as viewed along the c-axis for the host framework in 1A, 1B, and virtually empty form 1A* (which does not correspond to a real intermediate). Cylinder radius is set to 80; interaction energies with magnitudes smaller than $-10$ kJ mol$^{-1}$ have been omitted.
The conformational change from type I to type II makes possible the linkage of type I and type II molecules along the \( a \)-axis through backbone-to-backbone CH···OC interactions (Motif 5 I–II; Table 3 and Figure 7), which tighten the framework along the \( a \)-axis. Indeed, the crystal packing is more efficient in 1B than in 1A, as in the former case, the packing coefficient is 0.75 and in the latter is 0.71 (Table 3).

**Periodic HF-3c Calculations.** To investigate the SCSC transformation, a set of HF-3c-(0.27 \( s_g \)) periodic calculations \(^{38} \) was carried out using a development version of the software Crystal14. \(^{36,37} \) Full optimization of the studied crystal forms was performed at \( \Gamma \) point.

The results of optimization were evaluated by overlapping the optimized structures with the structures obtained via SCXRD (Figure S3 in the Supporting Information) and comparing the unit cell values (Table S3 in the Supporting Information).

The direct comparison between the energies of 1A and 1B crystal structures is hampered by the different chemical contents of the two crystal structures. Indeed, to evaluate the energy change from 1A to 1B (eq 1), we need to take into account the change in the unit cell content, as the energy values are given in \( \text{kJ/mol} \). In form 1A, there are two cyclopeptoid molecules and four acetonitrile molecules per unit cell; in form 1B, there are four cyclopeptoid molecules per unit cell (two independent molecules in the asymmetric unit). Thus, it is necessary to halve the energy value corresponding to form 1B and consider the energy of acetonitrile molecules in the gas phase.

The transformation from 1A to 1B is an endothermic process, as reported in eq 1

\[
\Delta E = \left( \frac{E_{1B}}{2} + 4E_{\text{CH-CN}} \right) - E_{1A} = 193.9 \text{ kJ/mol}
\]

To evaluate separately the energetic contribution of the conformational change vs the removal of acetonitrile molecules, we devised a “virtual” two-stage process:

- at first, the acetonitrile molecules leave and create a “virtually” empty crystal form 1A*, which does not correspond to a real intermediate. The event is a nonfavored process, eq 2

\[
\Delta E_1 = \left[ (4E_{\text{CH-CN}}) + E_{1A^*} \right] - E_{1A} = 218.8 \text{ kJ/mol}
\]

- then, half macrocycles change conformation to give crystal form 1B, eq 3 and Figures 3 and 10

\[
\Delta E_2 = \left( \frac{E_{1B}}{2} - E_{1A^*} \right) = -24.9 \text{ kJ/mol}
\]

Thus, a small thermal input may easily overcome the energy barrier between the two crystal forms and induce the SCSC transformation; on the other hand, the presence of acetonitrile molecules may reverse the transformation mechanism.

Upon adsorption, acetonitrile molecules bind to peptoid recognition sites, as the methylene hydrogen atoms of the methoxymethyl side chains (Motif 4-s; Table 2) and carbonyl atoms O3 and O2 (Motif 5-s; Table 2), which in 1B are involved in type II–type II interactions (Motif 3 II–II; Table 3) and type I–type II interactions (Motif 5 I–II; Table 3). This triggers the cooperative movement that enables the conformational change of type II molecules into type I molecules and therefore the transformation to crystal form 1A. Acetonitrile binding energies (which sum to \(-38 \text{ kJ/mol}\)) are enough to overcome the unfavorable energy barrier from 1B to 1A* (+24 kJ/mol).

**CONCLUSIONS**

The reversible SCSC transformation of form 1A to 1B is supported by the conformational change of the cyclic peptoid molecules. Both backbone conformational change and propargyl side-chain rotation by 113° make possible the formation of stabilizing (although weak) interactions as CH···OC and CH···π bonds.

Finally, the ability of flexible organic materials to adapt to external stimuli represents an opportunity for their possible future use as biomimetic sensors and/or actuators for biomedical applications.

**ASSOCIATED CONTENT**

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.cgd.0c01244.

- Structural and computational details (PDF)
- Motion pictures (MP4)

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(49) Crystal data for 1A and 1B: 1A monoclinic, P2₁/c, a = 17.887(3) Å, b = 20.335(3) Å, c = 8.4716(12) Å, β = 93.160(2)°, V = 3076.7(8) Å³, Z = 4, Dx = 1.318 g cm⁻³, 1B monoclinic, P2₁/c, a = 17.887(3) Å, b = 20.335(3) Å, c = 8.4716(12) Å, β = 93.160(2)°, V = 3076.7(8) Å³, Z = 4, Dx = 1.318 g cm⁻³. To properly compare the lattice constants and the unit cell volume in 1A and 1B, the crystallographic a axis and the unit cell volume in form 1A must be doubled. Thus: Δa% = 100 (a_{1B} - 2·a_{1A})/2·a_{1A} = −8.5%, Δb% = −3.1%, Δc% = −0.3%, Δβ% = 2.4%, ΔV% = −11.6%.