Identification of novel mutations in SARS-COV-2 isolates from Turkey

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Abstract

SARS-CoV-2 was first detected in the city of Wuhan, Hubei Province, China. In this study, we identified 11 unique mutations in viral SARS-COV-2 isolates from Turkey. Nine of them cause structural alterations in the S protein, nsp2, nsp3, nsp4 and nsp12 regions. The mutations identified here might have significant functional implications that need to be addressed in future studies in the context of vaccine engineering and therapeutic interventions. Moreover, transmission and phylogenetic analysis revealed multiple independent sources of introductions of SARS-CoV-2 into Turkey and a close relationship to the isolates from Saudi Arabia.

The first case of the novel coronavirus outbreak in humans was reported in Wuhan, China. The disease was named COVID-19 by WHO, and the virus was named SARS-CoV-2 (severe acute respiratory syndrome coronavirus 2) by the International Committee on Taxonomy of Viruses [1]. Since the emergence of COVID-19, based on the WHO report of July 8, 2020, more than 11,591,595 confirmed cases were reported in 147 countries, with 537,859 deaths, due to rapidly spreading SARS-CoV-2 [2]. Genetically different coronaviruses are spread in birds, humans and other mammals and can lead to severe diseases of the intestine, liver, nervous system, and respiratory system. Turkey straddles eastern Europe and western Asia and is a major travel hub. According to the World Health Organization’s report dated April 24, 2020, Turkey ranks sixth in the European region in terms of COVID-19 disease, after Spain, Italy, Germany, the United Kingdom, and France [3]. The first case of COVID-19 in Turkey was reported on March 11, 2020, and two months after the first case, on May 11, 2020, the Turkish Ministry of Health declared that the number of COVID-19 cases had reached 139,771, with 3841 deaths [4]. In the light of the coronavirus outbreak, the present study was designed with the aim to characterize notable genetic features of SARS-CoV-2 from Turkey and to identify some novel mutations in the spike protein (S), nucleocapsid protein (N), and non-structural proteins (nsp2, nsp3, nsp4, nsp6, nsp12/RdRP). Furthermore, transmission and phylogenetic analysis were also conducted to provide significant insight into the spread of the virus within Turkey.

For analysis, a total of 80 genome sequences of virulent strains from Turkey that had been uploaded to the NCBI (https://www.ncbi.nlm.nih.gov/genbank) and GISAID (https://www.gisaid.org/) databases as of May 4, 2020, were retrieved and compared to genome sequences from Saudi Arabia, Iran, America, China, Pakistan, Denmark, Spain, and Italy (Table S1). These sequences were first aligned using the Clustal W program, and a phylogenetic tree was constructed based on this alignment using the maximum-likelihood method in MEGA X software with 1000 replicates [5]. For identification of mutations, sequences of SARS-CoV-2 isolates were compared with a reference sequence (MN908947.1) from Wuhan. The secondary structure of the S protein and nonstructural proteins of SARS-COV-2 was predicted using CFSSP (Chou and Fasman secondary structure prediction), an online server [6].
Based on mutation analysis, 59 out of 80 isolates from Turkey contained a signature 23,403A > G (D614G) mutation in the spike glycoprotein (S), which is clearly indicative of a very frequent mutation (73%). Most samples with the D614G mutation were strongly associated with two other mutations (3037 C > T and 14,408C > T) in ORF1ab region (Table 1). These co-occurring mutations have been described recently as a characteristic of one of the major SARS-CoV-2 variants occurring in Europe. Regarding the ORF1ab region, we also identified previously reported single-nucleotide polymorphisms (SNPs) at positions 14,408 (C > T), 3037 (C > T), 11,083 (G > T), 1397 (G > A), 18,877 (C > T), 1059 (T > A) and 8782 (C > T). The 14,408C > T (P4715L) and 3037 C > T (F106F) variants in ORF1ab were found to occur at high frequency and are presumed to be linked, causing mutations in the RNA-dependent RNA polymerase (RdRP/nsp12) and nsp3 gene, respectively. RdRP/nsp12 is a key component of the replication/transcription machinery, and therefore, the leucine mutation at position 4715 of RdRP/nsp12 may potentially affect its function, possibly increasing the viral mutation rate. Moreover, the proline-to-leucine mutation was consistently observed in previous reports as a frequent mutation in Europe (51.6%) and North America (58.1%) [7, 8]. The variation C3037T has been reported to cause a synonymous mutation in the region encoding nsp3 and was seen in 57 isolates (71%) from Turkey. Consistent with other studies, C3037T, C14408T and A23403G were the most common mutations (73%) that were found together in isolates from Turkey.

Other key variations observed in the present report includes 25,563G > T (Q57H) in ORF3a, along with a consecutive series of three variations at positions 28,881 (G > A), 28,882 (G > A) and 28,883 (G > C) in the N protein. The triple mutation 28,881–28,883, which results in a change of two amino acids, 203–204: RG > KR, is known to play a critical role in virion assembly and structure and had been found frequently in US strains [9]. In present study, this tri-nucleotide mutation in the N protein was observed in eight samples from Turkey, including one from a patient (EPI_ISL_429870) with a history of travel to Saudi Arabia. This mutation was accompanied by the mutations 241:C > T, 3037:C > T and 14,408:C > T. The missense mutation G11083T, conferring an amino acid change from leucine (L) to phenylalanine (F) in non-structural protein 6 (nsp6) at position 3606, was present in 25 samples. It has been observed in Spain, Italy, and Iran, as well in this study (Table S2). Previously, it was reported as an infrequent mutation from Japan, the Netherlands, and Australia [10, 11]. Another substitution (G > A) was found within the region of ORF1a encoding nsp2 at position 1397 and was seen in 26% (21/80) of the isolates, resulting in an amino acid change from valine to isoleucine (V378I) that would not affect the isoelectric point. In a study by Pachetti et al. [8], the V378I substitution was mainly observed in isolates from Oceania and less frequently in Asia and North America.

Our secondary structure prediction analysis also highlighted 11 unique mutations in viral SARS-COV-2 isolates from Turkey in the spike (S) protein and non-structural proteins (nsp2, nsp3, nsp4, and nsp12/RdRP) (Table 1). Of these 11 mutations, nine of them have been found to be involved in structural alterations at different sites. Three of them (A771V, T1238I and G1251V) cause alterations in the structure of the S protein, while the rest of them induce structural changes in nsp2 (A206T, R207C, T265I), nsp3 (A1824V), nsp4 (M2796I) and nsp12 (A4489V).

Among novel mutants in the S protein, a substitution mutation (I468V) at position 468 was found in four Turkish isolates (EPI_ISL_437316, EPI_ISL_437322, EPI_ISL_437323 and EPI_ISL_437324) in the receptor-binding domain (RBD) of the spike protein that was not reported previously in strains from any other country. At this site, isoleucine (I) was replaced by valine (V) at position 468. Since both amino acids are hydrophobic in nature, having C beta branched residues, this mutation is unlikely to cause a functional change in the protein, as predicted by our secondary structure analysis (Fig. 1a). Nevertheless, if this site is prone to mutate further, it is possible that such mutations could affect the binding of the S protein to ACE2 receptor. Similarly, two strains (EPI_ISL_437314, EPI_ISL_437316) contained a missense mutation at position 771 exchanging alanine (A) for valine (V) in the S protein. Secondary structure predictions demonstrated that this could disrupt the helix structure, favoring a β-sheet at positions 770 and 771 due to addition of (Fig. 1b). The alanine-to-valine substitution as residue 771 was previously seen in one Belgian strain [12].

Additionally, a threonine (polar amino acid)-to-isoleucine (non-polar amino acid) substitution (T1238I) and a change of glycine to a bulkier valine at position 1251 (G1251V) were specifically found in strains EPI_ISL_429870 and EPI_ISL_437317, respectively. A secondary structure prediction revealed that a change of threonine to isoleucine makes this position hydrophobic and favors the formation of an additional helix at position 1238 (Fig. 1c), while the G1251V mutation favors the formation of six additional sheets from position 1448 to 1253 and a helix at site 1254 (Fig. 1d). Earlier reports have suggested that substitution mutations of threonine to isoleucine and alanine to valine in the Ebola virus (EBOV) glycoprotein (GP) increase the infectivity in humans [13, 14]. Therefore, it is possible that the mutant residues at positions 771, 1238 and 1254 in the spike protein of EBOV may lead to an alteration in the way the spike interacts with the receptor, changing the infectivity of the virus, as these mutations lie in the S2 subunit of the S protein.
Table 1 Mutations identified in 80 SARS-COV-2 genome sequences from Turkey. Unique mutations in the S protein, nsp2, nsp3, nsp4, nsp12 are indicated by *

| Mutations | Amino acid change | Gene | Type of mutation | Number of isolates |
|-----------|-------------------|------|------------------|--------------------|
| 23,403 A > G | D614G | S | Missense | 59 |
| 14,408 C > T | P4715L | ORF1ab | Missense | 58 |
| 3037C > T | ORF1ab | Synonymous | 57 |
| 241 C > T | 5'UTR | Non coding | 42 |
| 25563G > T | Q57H | ORF3a | Missense | 31 |
| 18877C > T | L6205L/L280L | ORF1ab | Synonymous | 29 |
| 11083G > T | L3606F | ORF1ab | Missense | 25 |
| 28688 T > C | N | Synonymous | 23 |
| 28,881 G > A | R203K | N | Missense | 23 |
| 28882G > A | N | Synonymous | 23 |
| 28,883 G > C | G204R | N | Missense | 23 |
| 29742G > T | 3'UTR | Non coding | 22 |
| 1397G > A | V378I | Orf1ab | Missense | 21 |
| 26735C > T | M | Synonymous | 13 |
| 884C > A | R207S | Orf1ab/nsp2 | Missense | 11* |
| 8653G > T | M2796I | Orf1ab/nsp4 | Missense | 11* |
| 19839 T > C | Orf1ab | Synonymous | 11 |
| 7765C > A | Orf1ab/nsp3 | Synonymous | 09* |
| 17690C > T | S5809L | Orf1ab | Missense | 08 |
| 228C > T | 5'UTR | Missense | 08 |
| 28854C > T | S194L | N | Missense | 08* |
| 22444C > T | D294D | S | Synonymous | 08* |
| 2113C > T | Orf1ab/nsp2 | Synonymous | 07* |
| 12809C > T | L4182F | ORF1ab | Missense | 04 |
| 10702C > T | Orf1ab | Synonymous | 04 |
| 26551 T > C | V10A | M | Missense | 04 |
| 22964A > T | I468V | S | Missense | 04* |
| 5736 C > T | A1824V | Orf1ab/nsp3 | Missense | 03* |
| 13476C > T | ORF1ab | Synonymous | 03* |
| 3903C > T | P1213L | ORF1ab/nsp3 | Missense | 03* |
| 881G > A | A206T | ORF1ab/nsp2 | Missense | 03* |
| 25611C > A | ORF3a | Synonymous | 03 |
| 8782 C > T | Orf1ab | Synonymous | 02* |
| 1059C > T | T265I | ORF1ab | Missense | 02* |
| 23874C > T | A771V | S | Missense | 02* |
| 25275C > T | T1238I | S | Missense | 01* |
| 23929C > T | Y789Y | S | Synonymous | 01* |
| 22468G > A | T302T | S | Synonymous | 01* |
| 25314G > T | G1251V | S | Missense | 01* |
| 2416C > T | Y717Y | ORF1ab/nsp2 | Synonymous | 01* |
| 13730C > T | A4489V | ORF1ab | Missense | 01 |
| 28144 T > C | ORF1ab | Missense | 01 |
| 2997C > T | ORF1ab | Missense | 01 |
| 25775C > T | A927V | M | Missense | 01 |
| 5477 C > T | H1738Y | ORF1ab | Missense | 01 |
| 6402C > T | P2046L | ORF1ab | Missense | 01 |
| 12741C > A | T4159V | ORF1ab/nsp8 | Missense | 01 |
| 2997C > T | S911F | ORF1ab | Missense | 01 |
Among the synonymous SNPs in the S protein, the C23929T (Y789Y) mutation was observed in only one Turkish sequence (EPI_ISL_455719) in the S gene. It was reported previously with high prevalence in Indian strains (39.13%) and in one US strain (EPI_ISL_436,898) [7, 12], but not in any other European strains so far. C22444T (S protein) and C28854T substitution (N protein) were observed to be uncommon co-occurring mutations in viral isolates from Turkey (8 samples) and Saudi Arabia (9 samples) (Table S2), which may provide evidence for a travel-associated origin of these mutations.

The ns2 protein is postulated to play an important role in the host cell survival pathway via its interaction with prohibitin (PHB) and prohibitin 2 (PHB2) [15]. The A206T, R207C and T265I mutations were predicted to cause structural alterations within the ns2 domain (Fig. 1e, f and g). A change of a nonpolar amino acid (alanine) to a polar amino acid (threonine) at position 206 (G881A mutation) in the ns2 protein resulted in the predicted loss of an α-helix at positions 203, 204 and 206 with the addition of a β-sheet at positions 203 and 204 (Fig. 1e), and this mutation appeared only in three sequences (EPI_ISL_480239, EPI_ISL_428723, EPI_ISL_429863) from Turkey. Similarly, an 884C > T mutation, which resulted in the change of arginine (R) to cysteine (C) at position 207 was present in 11 Turkish isolates and one Pakistani isolate (MT240479.1) (Table S2). The R207C mutation resulted in the replacement of an α-helix with a sheet structure at position 204 and an additional turn at position 210 (Fig. 1f). Furthermore, two Turkish samples (EPI_ISL_437309; EPI_ISL_437315), and one strain each from Spain (EPI_ISL_428688) and Denmark (EPI_ISL_437668) had a T265I mutation, which resulted in the addition of sheet structure at position 266 when threonine is substituted by isoleucine at position 265 (Fig. 1g).

Since all of these mutations were identified in the ns2 domain, possibly causing structural alterations, it is essential to consider the mutational spectrum when designing new antiviral therapeutics targeting the viral ORF1ab. In addition, a synonymous mutation (2416C > T) within ns2 was found in two infected individuals (EPI_ISL_428712; EPI_ISL_437332) in Turkey with a history of travel from Iran, and in three samples (EPI_ISL_468162; EPI_ISL_468161; EPI_ISL_468163; EPI_ISL_468164) from Turkey with a history of travel from Iran.

Table 1 (continued)

| Mutations | Amino acid change | Gene | Type of mutation | Number of isolates |
|-----------|------------------|------|-----------------|-------------------|
| 13376A>G  | T4371A           | ORF1ab | Missense        | 01                |
| 19484C>T  | A6407V           | ORF1ab | Missense        | 01                |
| 1437C>T   | S391F            | ORF1ab | Missense        | 01                |
| C7834T    |                  | ORF1ab | Synonymous      | 01                |
| 944G>A    | G227S            | ORF1ab | Missense        | 01                |
| 26735C>T  | M                | Synonymous |        | 01                |
| 14178C>T  |                  | ORF1ab | Synonymous      | 01                |
| 25549C>T  | L53F             | ORF3a  | Missense        | 01                |
| 16247C>T  | A5328V           | ORF1ab | Missense        | 01                |
| 6078C>T   | A1938V           | ORF1ab | Missense        | 01                |
| 16616C>T  | T5451I           | ORF1ab | Missense        | 01                |
| 27354A>G  |                  | ORF6   | Synonymous      | 01                |
| 20464G>A  | D6734N           | ORF1a  | Missense        | 01                |
| 9667T>C   |                  | ORF1ab | Synonymous      | 01                |
| 29197C>T  |                  | N      | Synonymous      | 01                |
| 7303C>T   |                  | ORF1b  | Synonymous      | 01                |
| 3464C>T   | H1067Y           | ORF1b  | Missense        | 01                |
| 5192C>T   |                  | ORF1b  | Synonymous      | 01                |
| 20629C>T  | H6789Y           | ORF1b  | Missense        | 01                |
| 20799T>C  |                  | ORF1b  | Synonymous      | 01                |
| 28391C>T  | R40C             | N      | Missense        | 01                |
| 25611C>A  |                  | ORF3a  | Synonymous      | 03                |
| 20668G>A  | A6802T           | ORF1b  | Missense        | 01                |
| 27476C>T  | T28I             | ORF7a  | Missense        | 01                |

Fig. 1 Prediction of secondary structure in the S protein, ns2, ns3, ns4, ns12 regions. a–d Mutations in the S protein. e–g Mutations in ns2; h–i Mutations in ns3; j Mutations in ns4; k Mutations in ns12. Small rectangular boxes indicate mutated residues. Differences in secondary structure between Wuhan and Turkish isolates are indicated by black boxes.
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A. Mutation in RBD of S protein
   - Wuhan-Hu-1
   - EPI_ISL_437324
   - 1409V

B. Mutation in S Protein
   - Wuhan-Hu-1
   - EPI_ISL_437314
   - A771V

C. Mutation in S protein
   - Wuhan-Hu-1
   - EPI_ISL_429870
   - T1238I

D. Mutation in S protein
   - Wuhan-Hu-1
   - EPI_ISL_437317
   - G1251V

E. Mutation in Nsp2 region
   - Wuhan-Hu-1
   - EPI_ISL_428723
   - A206T

F. Mutation in Nsp2 region
   - Wuhan-Hu-1
   - EPI_ISL_437324
   - R207C

G. Mutation in Nsp3 region
   - Wuhan-Hu-1
   - EPI_ISL_437309
   - T265I

H. Mutation in Nsp3 region
   - Wuhan-Hu-1
   - EPI_ISL_437329
   - A3182V

I. Mutation in Nsp4 region
   - Wuhan-Hu-1
   - EPI_ISL_437334
   - P1213L

J. Mutation in Nsp12 region
   - Wuhan-Hu-1
   - EPI_ISL_455719
   - A4489V
EPI_ISL_468160) from Pakistan. Previously, the T265I mutation was detected exclusively in the American population at a frequency of 43% making it a signature SNP for the USA, whereas it was found at very low frequency in Asia (4.8%) [15]. Likewise, a nucleotide change (C > T) was observed at position 2113 in seven Turkish samples and two isolates from Saudi Arabia, but not in any other sequence in this report.

In SARS-CoV, nsp3 has been proposed to work with nsp4 and nsp6 to induce double-membrane vesicles (DMVs), which serve as an important component of the replication/transcription complex [16]. In this report, some novel variants in the nsp3 region were also detected at positions 3903 (C > T), 5736 (C > T), and 7765 (C > A). Two of these were missense mutations (C3903T and C5736T) resulting in changes from proline to leucine (P1213L) and alanine to valine (A1824V), respectively, and each was exclusively seen in isolates from Turkey, while C7765A was a silent mutation and was observed in 11% (9/80) of Turkish samples and only one viral isolate from Saudi Arabia (EPI_ISL_437463) (Table S2). In mutant A1824V, there is a loss of a turn and the addition of a sheet structure at position 1825 (Fig. 1h) that might have significant functional implications, whereas no substantial change in secondary structure was observed for mutant P1213L (Fig. 1i).

Apart from frequent mutations, two mutations in the nsp4 gene (C8782T and G8653T) and one in the nsp12/RdRP gene (13,730C > T) were found to be completely unique to Turkey. A silent mutation at position 8782 (C > T) within nsp4 was found in two infected individuals (EPI_ISL_428718 and EPI_ISL_437317) from Turkey and in seven samples from Spain, where it was an infrequent mutation; however, it was present at high frequency in isolates from Oceania and North America in previous reports [8]. The G8653T mutation in the nsp4 gene, resulting in a change of methionine (M) to isoleucine (I) at position 2796 was found in 11 Turkish strains but not observed in any other viral isolates from Europe so far. However, this mutation was detected by Joshi and Paul [18] in nine Indian samples and in two isolates from Kuwait. In the nsp12/RdRP region, an alanine/valine substitution was observed at site 4489 in a single sequence (EPI_ISL_455719, Turkey/Mardin) on April 9, 2020, that was not found in any other sequence in this report. In a previous report by Maitra et al. [19], this unique mutation was also found in two infected individuals in India.

Further, our secondary structure prediction analysis showed that the M2796I mutation causes changes in the secondary structure of nsp4 in which the α-helix is replaced with a β-sheet structure at position 2795 and a turn at position 2798 (Fig. 1j), which might affect the interaction between nsp3 and nsp4 and thus the replication of the virus.
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The side chain of valine is larger than that of alanine, and substitution of valine in nsp12 resulted in the potential loss of an α-helix at positions 4486, 4487, and 4488 with the addition of a β-sheet from positions 4486 to 4490 (Fig. 1k). Therefore, this substitution might have functional consequences that can potentially affect the replication and mutagenic capability of SARS-CoV-2.

Importantly, it would be interesting to investigate the effects of these substitutions in the non-structural proteins, as one of a previous study has suggested that an alanine-to-valine substitution in the non-structural protein NS2A of Zika virus impairs viral RNA synthesis and results in attenuation of the virus [20]. Similarly, a valine substitution in the RdRP protein in Indian SARS-CoV-2 isolates has been shown to cause a structural alteration that impairs packing of the protein [21]. Thus, functional characterization of the mutations investigated in our study needs to be carried out to understand their role and to develop strategies for vaccine design.

We found that 13 viral isolates from Turkey harboring 23,403 A > G, 3037 C > T, and 14,408 C > T mutations were from individuals who had a history of travel from Saudi Arabia (Table 2). Of these, six cases were reported from the city of Ankara while the remaining ones were from Aksaray, Sakarya, Tekirdag, Kocaeli, Kastamonu, Konya and Afyon. Similarly, six samples were from individuals who had a history of travel from Iran and most of these cases (5/6) were from Istanbul and were listed in GISAID. Among Iran-travel-linked isolates, four samples (EPI_ISL_437319, EPI_ISL_437324, EPI_ISL_437325, and EPI_ISL_437327) contain the mutations (G1397A, T28688C and G29742T, a Europe-based introduction), while the remaining two samples (EPI_ISL_437326 and EPI_ISL_437332) lacked these mutations (Table 2). However, the G1397A and T28688C substitutions were also observed in one patient who had travelled to Taiwan. This suggests that COVID-19 infection spread to Turkey from multiple countries, especially from Saudi Arabia and Iran. In a similar analysis, Eden et al. [22] provided evidence of SARS-CoV-2 being exported to Australia from Iran. Our findings may also contribute to a better understanding of the diversity of circulating SARS-COV-2 strains and origin of imported cases in Turkey.

Phylogenetic analysis of SARS-COV-2 genome sequences suggested that the Turkish strain is closely related to isolates from Saudi Arabia, suggesting a common origin (Fig. 2). However, the sequences from Turkey were dispersed throughout the phylogenetic tree, indicating multiple independent introductions into the country. In the phylogram, two distinct clades were categorized as cluster 1
and cluster 2. Cluster 1 contained the mutations 1397G > A, 11,083G > T, 28,688 T > C, and 29,742G > T, while the second cluster had 23,403 A > G, 3037 C > T, and 14,408 C > T as the dominant mutations. Moreover, the majority of viral isolates in Turkey showed L type characteristics and formed a monophyletic clade, while the S type was present in limited numbers. An SNP with “T” at position 28,144 encoding leucine was classified as L type, while “C” at this position encoding serine is referred as S type. It has been suggested that the L type is more aggressive and contagious than the S type [12]. Thus, it appears that the L type is predominant in the Turkish population. In addition, samples having travel connection to Saudi Arabia and Iran showed a monophyletic origin within their respective clusters in the phylogenetic tree.

In conclusion, functional characterization of novel mutations investigated in our study needs to be carried out to understand the exact role of these variations. Furthermore, awareness of the above-mentioned mutations might be useful for the identification of less-virulent strains and the development of vaccines against a large repertoire of strains. Phylogenetic and transmission analysis revealed that the spread of SARS-COV-2 to Turkey was due to multiple independent introductions and that viral isolates from Saudi Arabia and Turkey are closely related to one another.

Compliance with ethical standards

Competing interests The authors declare that they have no competing interests.

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