Natural Organic Matter Removal from Raw Surface Water: Benchmarking Performance of Chemical Coagulants through Excitation-Emission Fluorescence Matrix Spectroscopy Analysis

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Abstract: Chemical disinfection of surface waters has been proven effective in minimizing the risk of contamination by water-borne pathogens. However, surface waters contain natural organic matter (NOM) which, upon chemical disinfection, is readily converted into hazardous disinfection-by-products. Hence, NOM removal from these waters is critical. Chemical coagulation is a readily implementable technology to minimize these undesired side-effects by NOM removal. Herein, capabilities of ferric chloride (FeCl₃) and polyaluminum chloride (PACl) as pre-treatment for NOM abatement from natural raw surface water have been benchmarked. Excitation-emission fluorescence matrix (EEM) spectroscopy characterization of NOM fractions demonstrated high removal efficiency. A two-level full factorial design was employed to analyze the effects of coagulant dosage and initial pH on the removal of turbidity, humic acid-like substances and fulvic acid-like substances from the raw water. Higher removal of ~77% NOM was attained with PACl than with FeCl₃ (~72%). Optimization through response surface methodology showed that the initial pH—coagulant dosage interaction was significant in removing NOM and turbidity for both PACl and FeCl₃. These results identify the opportunity for coagulation technologies to prevent and minimize disinfection-by-products formation through NOM removal.

Keywords: coagulation; excitation-emission fluorescence matrix; full factorial design; natural organic matter; physical water treatment; raw surface water

1. Introduction

Natural organic matter (NOM) is a complex mixture of organic compounds consisting of aromatic, aliphatic, phenolic, and quinolic functional groups [1]. In surface water, NOM accounts for 50–80% of humic substances [2]. Disinfection by-products (DBPs) can be formed when NOM reacts with active chlorine species during the drinking water treatment process [3–5]. Trihalomethanes and haloacetic acids are two popular groups of halogenated DBPs currently regulated by various drinking water standards [6]. Thus, minimizing NOM content prior to disinfection treatment can be identified as an urgent need to avoid DBP yield [7]. Different technologies such as photocatalysis [8], electrochemical treatments [9], the Fenton process [10], adsorption [11], ionic exchange [12] or coagulation [13] have been
explored to remove NOM from drinking water sources prior to disinfection. However, coagulation treatment presents the most techno-economically viable option, and it is therefore the most widely used in drinking water treatment plants worldwide.

Coagulation–flocculation is a process in drinking water treatment which aims to remove turbidity, color, and pathogens from raw water \[14,15\]. Coagulation is a physical-chemical process that involves the destabilization of repulsive forces between negatively charged organic matter by positively charged metals (aluminum or iron), resulting in the subsequent agglomeration and settling of organic pollutants \[16\]. Iron and aluminum salts in monomeric or polymeric forms are the typical coagulants used in large-scale drinking water treatment \[17,18\]. Iron-based coagulants, typically ferric chloride (FeCl\(_3\)), have been found to reduce dissolved organic carbon by about 29–70\% \[19\]. Aluminum-based coagulants, on the other hand, are the most common coagulants in the drinking water industry \[20\]. Polyaluminum chloride (PACl) can address the drawbacks of the traditional alum, (Al\(_2\)(SO\(_4\))\(_3\)), since it is less sensitive to pH and temperature variation \[21\]. Moreover, PACl has high amounts of positively charged polycations which are very effective in neutralizing the negatively charged colloidal particles, thus increasing colloidal destabilization \[22,23\].

Previous research articles demonstrated excellent performance on NOM abatement by coagulation treatment. Aluminum salts have been the most employed coagulant source \[19,24,25\]. Recent reports suggest that residual aluminum (III) concentrations may be associated with neurodegenerative diseases such as Alzheimer’s \[26\]. In this work, we benchmark the performance of iron (III) and aluminum (III) regarding their capabilities to remove NOM due to the relevance of transitioning to the use of alternative coagulants non-containing aluminum. Furthermore, the interaction between different variables is not discussed in the literature that evaluates the effect of discrete aspects one at a time. In the present study, a two-level full factorial design was utilized to benchmark the removal of NOM fractions by chemical coagulation with FeCl\(_3\) and PACl coagulants. This study enables understanding of the principles of different variables’ interaction and their synergies on the enhancement of NOM abatement by coagulation. An excitation-emission fluorescence matrix (EEM) was used to characterize the NOM fractions before and after coagulant application. The contributions of initial pH and coagulant dosage in NOM removal were evaluated. The effects of each factor, as well as the interaction effects, the adequacy of the model to describe the experimental data, and the optimum values of the parameters were also examined.

2. Materials and Methods

2.1. Chemicals and Raw Surface Water

Ferric chloride (FeCl\(_3\), 99.99\%), which is a yellowish solid with a hexagonal crystalline structure that is used as iron (III) coagulant source, was provided by Sigma-Aldrich. Polyaluminum chloride (PACl, 30\%), an inorganic polymer difficult to structurally characterize with a suggested Keggin ion structure and is used as an aluminum (III) coagulant source, was supplied by Nanning Chemical Engineering Co. Ltd., Guangxi, China). Both salts were dissolved in water, yielding iron hydroxides and aluminum hydroxides commonly used in coagulation treatments for drinking water. Hydrochloric acid (HCl, 37\%, ENSURE) and sodium hydroxide (NaOH, 98\%, Shimakyu’s Pure Chemicals) were used as received. Stock solutions were prepared by dissolving known amounts of reagents in deionized water (18.2 M\(\Omega\) cm, Millipore). The raw surface water used in all experiments was obtained from a water treatment plant in Taiwan which had an initial water quality summarized in Table 1.

2.2. Coagulation Experiments

Coagulation tests were conducted in a jar test apparatus (PB-700, Phipps & Bird). The initial pH of the water sample was adjusted according to the experimental design using 0.50 M NaOH and 0.50 M HCl. The water sample was subjected to 1 min rapid mixing at 100 rpm to ensure homogeneous distribution of the NOM particles in solution. Coagulant
was added to the water sample and the solution was subjected to rapid mixing at 100 rpm for 1 min, followed by a flocculation process through slow mixing at 30 rpm for 15 min, and sedimentation for 30 min. Aliquots of 10 mL samples were collected during treatment for further analysis and characterization.

Table 1. Characteristics of raw surface water.

| Parameter                        | Value          |
|----------------------------------|----------------|
| pH                               | 4.8 ± 0.1      |
| Turbidity (NTU)                  | 9.5 ± 0.2      |
| Zeta potential (mV)              | −17.6 ± 0.6    |
| Fulvic acid-like substances: Ex/Em = 200 − 250/380 − 550 (au) | 2830 ± 60      |
| Humic acid-like substances: Ex/Em = 250 − 400/380 − 550 (au) | 2260 ± 50      |

A full factorial design was used to understand the influence of operational variables and the interaction between factors during the coagulation process. The response surfaces generated were employed to identify optimum operational parameters [27]. The effect of each factor on the response was determined by the design along with the interaction, which is the effect of a factor that varies with the change in the level of other factors [28]. The two-level full factorial experimental design considered two levels per factor coded as ‘high’ and a ‘low’ level of ‘+1’ and ‘−1’, respectively. The experimental matrix consisted of 2 k runs where k is the number of factors/variables evaluated. However, a potential concern in the use of two-level full factorial design (FFD) is the assumption of linearity in the effect of the factors: (A) initial pH and (B) coagulant dosage. The variables and levels of the experimental design are presented in Table 2, where A and B are the equivalent values in coded forms. Protection against curvature is done by adding center points, whereby the design also obtains an independent estimate of error [29]. In addition, the method can easily be upgraded to response surface designs for optimization of process parameters [30]. The FFD was carried out using Design-Expert 6.0.8.

Table 2. Experimental ranges and level of the factors in the factorial design.

| Variables                        | Coded Symbols | Coded Levels |
|----------------------------------|---------------|--------------|
| Initial pH                       | A             | −1 1         |
| Coagulant dose (mg L⁻¹ Al or Fe) | B             | 6 8          |

2.3. Analytical Methods

Initial pH was determined using a pH meter (PC-310, Suntex, Hong Kong, China). Turbidity was measured by a turbidity meter (2100 Q, Hach, Loveland, CO, USA). A zeta potential analyzer (ZetaPlus, Brookhaven, NY, USA) was utilized to determine the zeta potential of the water samples. An excitation emission fluorescence matrix (EEM) was employed to evaluate the NOM fractions (F-4500FL spectrophotometer, Hitachi, Tokyo, Japan). EEM spectroscopy is a sensitive and selective technique that can rapidly process data on NOM fractions without sample preparation [31]. EEM can analyze the structure and functional groups in a molecule, and determine the difference between sources of chromophoric dissolved organic material that has similar wavelengths [32]. Excitation emission matrix peaks are divided into five regions—aromatic proteins such as tyrosine and tryptophan are located in region I and region II, fulvic acid-like substances (FLS) can be found in region III, microbial by-product-like materials in region IV, and humic acid-like substances (HLS) in region V [33]. Fluorescence intensity decreases with increasing molecular size. The presence of fluorescence peaks is associated with linearly condensed aromatic rings and other unsaturated bond systems and has a high degree of conjugation with macromolecules. The difference between the initial and final EEM peak intensities can be linked to NOM removal efficiency of emerging water treatment solutions [34]. Al-
though pH adjustment may alter the fluorescence intensities, it does not affect the removal efficiency of the coagulation/flocculation process, as reported by earlier studies [35]. EEM plots were created by scanning excitation wavelength from 200 to 400 nm and emitting fluorescence from 280 to 550 nm with 10 nm steps. The steps for excitation and emission were set at 10 nm and the scan speed at 500 nm s\(^{-1}\). A fluorometer’s response to a blank solution was subtracted from the fluorescence spectra recorded for each sample to account for the Raleigh scattering.

The decrease in the fluorescence intensity peaks in the excitation-emission matrix was used as an indicator of NOM removal efficiency in the experiment. In previous studies, peak intensities were also used as an indication of NOM fraction removal [32,34]. Equation (1) was utilized to determine the NOM fraction removal (%):

\[
\text{NOM fraction removal} (\%) = \left(\frac{F_{0\text{max}} - F_{S\text{max}}}{F_{0\text{max}}}\right) \times 100
\]

where \(F_{0\text{max}}\) is the maximum intensity of each fluorescence component of raw water NOM, and \(F_{S\text{max}}\) is the maximum intensity of each fluorescence component of water NOM of the sample after sedimentation [35].

3. Results and Discussion

3.1. Natural Organic Matter Fractions Removal by Coagulation Treatment

Figure 1 shows the EEM contour plots before and after treatment using FeCl\(_3\) and PACl. The red-colored regions in sections III and V correspond to fulvic and humic acid-like species, respectively [33]. Transition to lighter colors (i.e., yellow and green) is associated with a notorious decrease in the concentration of these species. The high intensity in region III prior to treatment denotes a large presence of fulvic-like species in raw water samples (cf. Figure 1a,c). Likewise, in region V, the red spot located at the lower left side near region III denotes significant amounts of humic-like species. Intensity reduction of EEM signals was evident after the coagulation process where the peaks identified in the regions III and V which correspond to fulvic and humic acid-like disappeared after coagulation treatment independently of the coagulant species employed. The significant reduction in intensity after treatment, depicted by the EEM contours after coagulation treatment, allows us to infer effective NOM abatement with a better performance associated to iron (III) salts [18,35].

3.2. Model Fitting and Statistical Analyses

Table 3 summarizes the experimental results of the 2 k full factorial design obtained for the removal of turbidity, humic, and fulvic acid-like species at the different coded levels. Similar results were observed for the duplicate of the central point at pH 7 and coagulant dose of 25 mg L\(^{-1}\) with less than 2% error (runs 3 and 6), which corroborates that the pure error of coagulation experiments is negligible. The data for the experimental matrix were then modeled and statistically validated.

| Run | pH | Coagulant Dose (mg L\(^{-1}\)) | HLS | FLS | Turbidity |
|-----|----|-------------------------------|-----|-----|-----------|
|     | 1  | 6                             | 10  | FeCl\(_3\) | PACl | FeCl\(_3\) | PACl | FeCl\(_3\) | PACl |
| 2   | 6  | 40                            | 64.90 | 77.02 | 53.51 | 75.27 | 63.45 | 69.23 |
| 3   | 7  | 25                            | 62.70 | 48.97 | 72.00 | 49.97 | 76.87 | 64.52 |
| 4   | 8  | 10                            | 69.55 | 78.46 | 73.87 | 73.94 | 72.12 |
| 5   | 8  | 40                            | 51.74 | 67.91 | 64.34 | 66.56 | 67.94 | 71.96 |
| 6   | 7  | 25                            | 50.49 | 69.37 | 63.08 | 60.14 | 74.13 | 71.98 |
Figure 1. Excitation emission fluorescence matrix (EEM) contour plots of raw water before and after coagulation treatment: (a) natural raw water prior FeCl$_3$ treatment, (b) after FeCl$_3$ treatment, (c) natural raw water prior PACl treatment, (d) after PACl treatment.

Analysis of variance (ANOVA) was utilized to determine the statistical significance of the factors and their corresponding goodness of fit. Table 4 presents the values for the regression coefficients (RC), sum of squares (SS), standard error (SE), F-value (F), and p-value ($p$). The p-value is the probability value that determines the significance of the effect of each factor in the model [36]. Fisher’s test was used to determine the significance of the variables where the degree of significance was ranked based on the value of the F-ratio—the larger the value of $F$, the smaller the value of “Prob > F”. This translates to the greater significance of the corresponding model and the individual coefficient [36]. The confidence level used to determine the statistical significance of the factors is 95%, which means that the p-value should be less than or equal to 0.05 for the effect to be statistically significant [37]. Upon elimination of the insignificant terms, the final empirical models based on statistical analyses were defined.
Table 4. Statistical analysis of the results of the factorial experimentation.

| Response          | Factor | RC | SS  | SE  | F-Value | p-Value |
|-------------------|--------|----|-----|-----|---------|---------|
|                   |        | FeCl₃| PACl| FeCl₃| PACl    | FeCl₃   | PACl    | FeCl₃| PACl    |
| HLS Removal       | A      | -4.38| 3.19| 76.65| 40.70   | 0.44    | 0.52    | 98.11| 38.19   | 0.064 | 0.102  |
|                   | B      | 6.70 | -2.56| 179.68| 26.21   | 0.44    | 0.52    | 230.01| 24.69   | 0.042 | 0.127  |
|                   | AB     | 7.80 | 11.56| 243.52| 534.07  | 0.44    | 0.52    | 311.70| 504.10  | 0.036 | 0.028  |
|                   | Curvature | 92.02| 7.46| 0.77 | 0.89 | 117.78 | 7.00 | 0.059 | 0.230  |
| FLS Removal       | A      | -1.41| 4.58| 7.95 | 84.00   | 0.45    | 0.53    | 10.02| 74.66   | 0.195 | 0.070  |
|                   | B      | 11.59| -4.40| 537.31| 77.53   | 0.45    | 0.53    | 676.89| 68.91   | 0.025 | 0.070  |
|                   | AB     | 2.34 | 8.25| 22.00| 272.09  | 0.45    | 0.53    | 27.71| 241.85  | 0.120 | 0.041  |
|                   | Curvature | 7.46 | 61.38| 0.77 | 0.92 | 9.39   | 72.34  | 0.201 | 0.075  |
| Turbidity Removal | A      | 2.57 | 4.79| 26.47| 91.45   | 0.067   | 0.057   | 1466.54| 7183.51 | 0.017 | 0.008  |
|                   | B      | 7.73 | 1.08| 238.36| 4.65   | 0.067   | 0.057   | 13233.08| 363.49 | 0.005 | 0.033  |
|                   | AB     | 1.02 | 3.43| 4.14 | 47.10   | 0.067   | 0.057   | 229.43| 3679.75 | 0.042 | 0.011  |
|                   | Curvature | 2.26 | 0.20| 0.12 | 0.098  | 125.32  | 15.72   | 0.057 | 0.157  |

The response surface equation defined for the use of FeCl₃ as coagulant for each response is defined by Equations (2)–(4):

Humic acids removal (%) = 9.42 + 6.70B + 7.80AB  
Fulvic acids removal (%) = 61.35 + 2.34AB  
Turbidity removal (%) = 72.73 + 7.73B  

Meanwhile, identical modelling approaches were employed to define the model equation for the coagulation treatment using PACl. Equations (5)–(7) describe the response surfaces for the removal of humic-like species, fulvic-like species, and turbidity.

Humic acids removal (%) = 66.27 + 8.25AB  
Fulvic acids removal (%) = 67.20 + 4.58A + 4.40B + 8.25AB  
Turbidity removal (%) = 71.67 + 4.79A + 1.08B + 3.43AB  

It can be seen in Table 5 that the values of adj- \( R^2 \) are greater than 0.98, which indicates that the variability of new data is expected to be about 98%. At 95% confidence level, the coefficients of determination (\( R^2 \)) for all responses obtained from coagulants FeCl₃ and PACl were more than 0.99. The value of \( R^2 \) is a measure of the proportion of total variability by the model, where values close to 1 and at least 0.80 mean that the model is a good fit [36]. This implies that the generated model was sufficient in closely estimating the experimental HLS, FLS, and Turbidity removal efficiency [39]. Previous studies suggested the use of adj- \( R^2 \), a statistic that is adjusted for the size of the model, i.e., the number of factors, to evaluate the adequacy of the model in order to prevent the potential problem wherein the value of \( R^2 \) tends to increase as factors are added to the model [36]. Adequate precision, included in Table 5, pertains to the signal to noise ratio and typically has a value greater than 4, which implies that the signal is desirable. On the other hand, the coefficient of variation (CV) is the standard deviation calculated as a percentage of the mean, with values no greater than 10% [36]. It was observed that all statistical results presented in Table 5 demonstrate the good adequacy of the estimated model to fit all the responses. This was also further revealed by the predicted–observed plots in Figure 2, which enables inferring the fair agreement of predicted responses to the experimental data for both coagulants, FeCl₃ and PACl.
Table 5. Statistical validation from analysis of variance for the surface response models.

| Statistics | HLS Removal | FLS Removal | Turbidity Removal |
|------------|-------------|-------------|-------------------|
|            | FeCl₃ | PACl | FeCl₃ | PACl | FeCl₃ | PACl |
| $R^2$      | 0.9988 | 0.9987 | 0.9912 | 0.9995 | 0.9985 |
| $R^2_{adj}$| 0.9953 | 0.9948 | 0.9647 | 0.9978 | 0.9940 |
| Adeq. Precision | 59.06 | 42.30 | 29.14 | 15.36 | 119.09 |
| CV (%)     | 1.15 | 1.23 | 1.42 | 1.64 | 0.27 | 0.69 |

Figure 2. EEM predicted vs. actual plots of the experiment with coagulant (a) FeCl₃ and (b) PACl for each response: (◇) humic acid-like species, (⋄) fulvic acid-like species, and (△) turbidity.

3.3. Effect of Initial pH, Coagulant Dosage, and Their Interaction

Figure 3 illustrates the percentage of contribution of each term defined by Equations (2)–(7) on the NOM removal performance using different coagulants (i.e., FeCl₃ or PACl). Note that the results refer to the effects of initial water pH (A) and coagulant dosage (B) on the coagulation process.

Figure 3. Percent contribution of each equation model term on the overall NOM removal by coagulation using (a) FeCl₃ or (b) PACl. Factors: A refers to initial pH, B corresponds to coagulant dose, AB is the term that represents the synergistic interaction between both variables, and curvature of the model.

Figures 4 and 5 show the contour plots of the main factors, initial water pH and coagulant dosage, and interaction effects on the NOM removal percentage after using FeCl₃ and PACl coagulants, respectively. As shown in Figure 3a, high percentage contributions of coagulant dosage were obtained for FLS and turbidity removals (93.36% and 87.90%, respectively.)
respectively) when FeCl₃ was used as the coagulant. This trend is attributed to the initial pH being set at near neutral levels, where charge neutralization occurs. This implies that, for all pH values considered, all runs were expected to yield high removal efficiencies. It was observed in Figure 4 that the removal efficiency improved as the coagulant dosage was increased. This is attributed to the increase in the amount of FeCl₃ that hydrolyzed to positive ferric species and subsequently interacted with the negatively charged NOM fractions to form larger complexes [2]. NOM fraction removal efficiency was reported to be constant at a coagulant dosage greater than 40 mg L⁻¹ [25]. At a high coagulant dosage, FeCl₃ could not effectively remove NOM because only a portion of Fe could interact with NOM to form Fe–NOM complexes. Meanwhile, the excess of iron dosed will form negatively charged Fe(OH)₃ flocs and Fe(OH)₄⁻ instead of desired Fe-NOM complex [40].

![Figure 4. Contour plots showing the percent removal attained during coagulation treatment in function of FeCl₃ coagulant dose and pH.](image1)

![Figure 5. Contour plots showing the percent removal attained during coagulation treatment in function of PACl coagulant dose and pH.](image2)

On the other hand, when using PACl, the NOM removal was mainly due to the interaction of initial pH and coagulant dosage (AB) as deduced from Figure 2b. Note that Figure 5 also shows high interaction effects on the parameters considered when PACl coagulant was used. It is evident in the contour plots that pH affected the removal efficiencies. The highest removal percentages were recorded at 40 mg L⁻¹ coagulant dosage and an initial pH of 8, with values of 76% and 81% for fulvic acids and turbidity, respectively. It can be noted that the removal of fulvic at pH 6 and 8 are almost the same, which may be due to the solubility of fulvic acids in both alkaline and acidic regions, while humic acids are only soluble in the alkali region [41]. High PACl dosage is not effective in removing turbidity and NOM fractions because only a portion of Al reacts with NOM fraction to form an Al–NOM fraction complex. This is due to the formation of polymer bridges between particles that caused the destabilization of Al-NOM complex, resulting in the repulsion between particles at excessive dosage [42].

Analysis of turbidity evolution shows interesting trends in the function of coagulant species employed and operational conditions of coagulation treatment, as can be seen...
in Figure 6. The turbidity spiked up upon addition of FeCl₃ coagulant and 1 min rapid mixing, gradually reduced during the flocculation process of 30 min slow mixing, and dropped to near zero at the end of the 30 min sedimentation. Increase in turbidity can be explained by the formation of iron hydroxide flocs after coagulant addition, which removes suspended solids during its settlement [43,44]. Analysis of zeta potential changes during treatment shows a gradual change from initial negative values of $-17.09 \pm 1.03$ to zero. These results that show a gradual increase in zeta potential towards zero can be attributed to charge neutralization.

Interestingly, a spike in zeta-potential at high doses of FeCl₃ of 40 mg L⁻¹ can be observed, which can be explained by the positive zeta potential of iron hydroxide flocs [40]. The iron coagulation process is controlled by charge neutralization mechanism, which is said to occur at pH 6 and >7 where NOM is most negative [28]. This trend demonstrates the dual role of pH not only on the coagula formation but also on the natural speciation of NOM in function of the pH. NOM is composed by a complex mixture of fulvic and humic acids of different molecular weights and different functional groups that are susceptible to be deprotonated (i.e., carboxylic groups) [19,45,46]. The ratio of the different charged and non-charged species is determined by the respective pKₐ value of each organic acid. A higher density of negatively charged species will require higher doses to induce removal mechanisms ruled by charge destabilization and adsorption/complexation. The addition of positive coagulant disrupts the negatively charged NOM fractions and produces coagulant-NOM flocs [19,46], which also assists in the removal of solids from the suspension. The removal is therefore dependent not only on the formation of metal hydroxides as coagulants but also on the charge distribution of organic species in as a function of pH. Identical mechanisms are associated for PACl coagulant agent [47,48]. Generally, charge neutralization occurs at around neutral pH when aluminum salts are used. At neutral pH, the cationic hydrolysis products are only a small portion of the total soluble Al, while aluminate ions are the dominant form. Colloidal hydroxide particles are suggested to be effective charge-neutralizing species and may be positively charged up to pH 8, which explains the high removal up to pH 8 [29]. At neutral pH, coagulants are said to be prone to further hydrolysis and polymerization into medium polymer species. A quick comparison between turbidity abatement can be conducted between the results described in Figure 6a,b. Note that independent of experimental conditions, FeCl₃ outperforms PACl in turbidity reduction. However, PACl presents better performance on the abatement of NOM (i.e., humic and fulvic acid-like species), which highlights the use of PACl as an efficient approach to minimize the risk of disinfection by-products formation through NOM oxidation during disinfection stages. Therefore, these results encourage the use of PACl to remove NOM from raw waters.

![Figure 6](image-url)
Figure 6. Turbidity vs. time during coagulation by (a) FeCl₃ and (b) PACl, and zeta potential vs. time during coagulation by (c) FeCl₃ and (d) PACl.

3.4. Optimization of Process Parameters

In order to improve the coagulation process, optimization of parameters was defined from the contour surfaces. The desirability function method was used to determine the most desirable condition in the responses [28]. This method can combine multiple responses to generate a response called desirability function. Desirability function ranges from 0 to 1, with the desired value closest to 1 [36].

The initial pH and coagulant dosage were set within a range, whereas the removal responses were set to maximum levels. Figure 7 shows the desirability plot for all the responses, with an overall desirability of 1 at an initial pH of 8 and a coagulant dosage of 40 mg L⁻¹. Optimized experiments allowed for attaining a maximum removal of 69.6% of...
humic acids, 73.9% of fulvic acids, and 84.0% of turbidity when using FeCl$_3$ as the coagulant. On the other hand, for PACI slightly higher removals of NOM, with percentage removals of 78.5% for humic acids, and 75.6% for fulvic acids, and lower turbidity reduction of 80.9% were attained. These results suggest superior performance of PACI as the coagulant to trap and precipitate NOM during coagulation treatment.

![Figure 7. Contour plot for desirability in coagulation using coagulant (a) FeCl$_3$ and (b) PACI.](image)

4. Conclusions

Factorial design was used to evaluate FeCl$_3$ and PACI coagulants in removing NOM fractions from raw surface water by chemical coagulation. Humic and fulvic acid-like species were successfully characterized by an excitation-emission fluorescence matrix (EEM), both qualitatively and quantitatively. The effects of coagulant dosage and initial pH were also determined. The FeCl$_3$ coagulant dosage had the highest contribution to NOM and turbidity removal. This was explained by the hydrolyzation of FeCl$_3$ to positive ferric ion and the interaction with the negatively charged NOM fractions at a pH range between 6 and 8. For PACI, initial pH had the highest contribution on the removal of turbidity, while coagulant dosage and initial pH had the highest contribution for HLS and FLS treatment. This trend correlation was attributed to the solubility of humic-acid like species in alkaline media and the solubility of fulvic acid-like species at acidic and alkali conditions. The optimum operational condition was determined to be at an initial pH of 8.0 and coagulant dosage of 40 mg L$^{-1}$. The optimum removals when FeCl$_3$ was used were 69.55, 73.87, and 84.05% for humic acids, fulvic acids, and turbidity, respectively. On the other hand, optimum removals of 78.46, 75.63, and 80.98% for HLS, FLS, and turbidity, respectively, were obtained for PACI. These results identify PACI as the desired coagulant species to minimize NOM content in raw water prior to disinfection treatments.

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