Splenic switch-off as a predictor for coronary adenosine response: validation against 13N-ammonia during co-injection myocardial perfusion imaging on a hybrid PET/CMR scanner

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Abstract: BACKGROUND Inadequate coronary adenosine response is a potential cause for false negative ischemia testing. Recently, the splenic switch-off (SSO) sign has been identified as a promising tool to ascertain the efficacy of adenosine during vasodilator stress cardiovascular magnetic resonance imaging (CMR). We assessed the value of SSO to predict adenosine response, defined as an increase in myocardial blood flow (MBF) during quantitative stress myocardial perfusion 13 N-ammonia positron emission tomography (PET). METHODS We prospectively enrolled 64 patients who underwent simultaneous CMR and PET myocardial perfusion imaging on a hybrid PET/CMR scanner with co-injection of gadolinium based contrast agent (GBCA) and 13N-ammonia during rest and adenosine-induced stress. A myocardial flow reserve (MFR) of > 1.5 or ischemia as assessed by PET were defined as markers for adequate coronary adenosine response. The presence or absence of SSO was visually assessed. The stress-to-rest intensity ratio (SIR) was calculated as the ratio of stress over rest peak signal intensity for splenic tissue. Additionally, the spleen-to-myocardium ratio, defined as the relative change of spleen to myocardial signal, was calculated for stress (SMRstress) and rest. RESULTS Sixty-one (95%) patients were coronary adenosine responders, but SSO was absent in 18 (28%) patients. SIR and SMRstress were significantly lower in patients with SSO (SIR: 0.56 ± 0.13 vs. 0.93 ± 0.23; p < 0.001 and SMRstress: 1.09 ± 0.47 vs. 1.68 ± 0.62; p < 0.001). Mean hyperemic and rest MBF were 2.12 ± 0.68 ml/min/g and 0.78 ± 0.26 ml/min/g, respectively. MFR was significantly higher in patients with vs. patients without presence of SSO (3.07 ± 1.03 vs. 2.48 ± 0.96; p = 0.038), but there was only a weak inverse correlation between SMRstress and MFR (R = -0.378; p = 0.02) as well as between SIR and MFR (R = -0.356; p = 0.004). CONCLUSIONS The presence of SSO implies adequate coronary adenosine-induced MBF response. Its absence, however, is not a reliable indicator for failed adenosine-induced coronary vasodilatation.

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Abstract

Background: Inadequate coronary adenosine response is a potential cause for false negative ischemia testing. Recently, the splenic switch-off (SSO) sign has been identified as a promising tool to ascertain the efficacy of adenosine during vasodilator stress cardiovascular magnetic resonance imaging (CMR). We assessed the value of SSO to predict adenosine response, defined as an increase in myocardial blood flow (MBF) during quantitative stress myocardial perfusion 13 N-ammonia positron emission tomography (PET).

Methods: We prospectively enrolled 64 patients who underwent simultaneous CMR and PET myocardial perfusion imaging on a hybrid PET/CMR scanner with co-injection of gadolinium based contrast agent (GBCA) and 13N-ammonia during rest and adenosine-induced stress. A myocardial flow reserve (MFR) of > 1.5 or ischemia as assessed by PET were defined as markers for adequate coronary adenosine response. The presence or absence of SSO was visually assessed. The stress-to-rest intensity ratio (SIR) was calculated as the ratio of stress over rest peak signal intensity for splenic tissue. Additionally, the spleen-to-myocardium ratio, defined as the relative change of spleen to myocardial signal, was calculated for stress (SMRstress) and rest.

Results: Sixty-one (95%) patients were coronary adenosine responders, but SSO was absent in 18 (28%) patients. SIR and SMRstress were significantly lower in patients with SSO (SIR: 0.56 ± 0.13 vs. 0.93 ± 0.23; p < 0.001 and SMRstress: 1.09 ± 0.47 vs. 1.68 ± 0.62; p < 0.001). Mean hyperemic and rest MBF were 2.12 ± 0.68 ml/min/g and 0.78 ± 0.26 ml/min/g, respectively. MFR was significantly higher in patients with vs. patients without presence of SSO (3.07 ± 1.03 vs. 2.48 ± 0.96; p = 0.038), but there was only a weak inverse correlation between SMRstress and MFR (R = -0.378; p = 0.02) as well as between SIR and MFR (R = -0.356; p = 0.004).

Conclusions: The presence of SSO implies adequate coronary adenosine-induced MBF response. Its absence, however, is not a reliable indicator for failed adenosine-induced coronary vasodilatation.

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Background
Due to its rapid onset of action, its short half-life and the good tolerability and safety profile, adenosine is a widely used vasodilator for ischemia testing in myocardial perfusion imaging (MPI) using single photon emission computed tomography (SPECT), positron emission tomography (PET), and cardiovascular magnetic resonance imaging (CMR). However, induction of maximal coronary vasodilation is a crucial prerequisite for obtaining high diagnostic accuracy concerning the detection of obstructive coronary artery disease (CAD) [1–4]. Whether this condition is achieved remains difficult to assess in clinical routine because commonly used markers such as the hemodynamic response to adenosine (i.e., a decrease in systolic blood pressure and/or increase in heart rate) are unreliable as they are prone to procedure-related confounders such as anxiety or adenosine-induced side effects [5, 6]. Recently, the splenic switch-off (SSO) sign has been proposed as a more objective and direct marker of adequate adenosine response during stress CMR [7]. SSO has been defined as a visible decrease in splenic signal intensity during adenosine stress as compared to rest, presumably due to reduced splenic blood flow which may be mediated through reactive sympathetic vasoconstriction after adenosine-induced hypotension [8].

Contrary to modalities relying on the detection of relative regional differences in myocardial perfusion, such as CMR and SPECT, PET MPI allows for absolute quantification of myocardial blood flow (MBF) and calculation of myocardial flow reserve (MFR) [9]. Adenosine-induced coronary vasodilation has been shown to increase MBF during PET MPI up to fourfold over resting baseline [10]. Hence, PET-derived MBF quantification is a reliable standard of truth for assessing adequate coronary adenosine response. Furthermore, the introduction of novel PET/CMR devices, incorporating both modalities in a single hybrid scanner, offers the unique possibility of simultaneous assessment and cross-validation of both modalities during co-injection of gadolinium-based contrast agents (GBCA) and 13N-ammonia [11]. Most importantly, the simultaneity of PET and CMR data acquisition during the same adenosine stimulus ensures an identical physiological state of the coronary and extra-cardiac vasculature. We sought to test the hypothesis that SSO constitutes a reliable predictor of adequate coronary adenosine response. MFR served as the standard of truth and was assessed on a PET/CMR device with co-injection of 13N-ammonia and GBCA during adenosine stress.

Methods
Study design and population
Data of this prospective single-center study were derived from ongoing PET/CMR projects. We assessed patients who underwent cardiac PET/CMR for evaluation of CAD or cardiomyopathy. Written informed consent was obtained from all participants, and the study protocol was approved by the local ethics committee (KEK-ZH-Nr. 2014-0187 and BASEC-Nr. 2018-00170). Patients aged ≥18 years without any contraindications against CMR (e.g., implanted cardiac devices, claustrophobia, known GBCA allergy, severe renal impairment), adenosine (e.g., asthma, atrioventricular block), or PET (e.g., pregnancy or breastfeeding) were included. This study was partially funded by the Swiss National Science Foundation (SNSF Project Nr. 175640).

Hybrid PET/CMR perfusion imaging
CMR and PET datasets were acquired using a hybrid PET/CMR device incorporating a 3 T CMR and a latest-generation PET scanner with time-of-flight (TOF) (Signa PET/MR, GE Healthcare, Waukesha, Wisconsin, USA). All patients were asked to refrain from caffeine intake for at least 12 h before the examination. The stress protocol consisted of 6 min of adenosine infusion with a weight-adapted dose of adenosine (140 μg/kg/min). A body mass index adapted dose of 13N-ammonia (i.e., 200–600 megabecquerels (MBq) and a weight-adapted dose of GBCA (Gadovist, Bayer AG, Berlin, Germany) (0.1 mmol/kg) was simultaneously injected 3 min into adenosine stress. Dynamic PET data acquisition consisted of 21 frames (i.e., 9 × 10-s, 6 × 15-s, 3 × 20-s, 2 × 30-s, and 1 × 120-s), followed by an electrocardiogram (ECG)-gated static acquisition over 10 min. Resting perfusion imaging was performed using an identical acquisition protocol after a minimum of 15 min following the stress acquisition according to Society for Cardiovascular Magnetic Resonance (SCMR) guidelines [12]. All PET data were acquired in 3D mode and reconstructed using TOF reconstruction with VUE Point FX (2 iterations and 16 subsets) and 5-mm Hanning filter. Standard Dixon-based maps were used for attenuation correction [13]. Summed semi-quantitative myocardial PET tracer uptake and quantitative MBF was obtained from stress and rest scans and analyzed using QPET (Version 2015, Cedars-Sinai Medical Center, Los Angeles, California, USA). Datasets were examined in consensus by two experienced readers regarding the presence of ischemia and/or scar.

Keywords: Splenic switch-off, Cardiovascular magnetic resonance imaging, Myocardial perfusion imaging
CMR stress and rest perfusion scans were acquired using three left ventricular short-axis slices per cardiac cycle (basal, mid, and apical levels) each of 10 mm slice thickness and ECG-gated breath-hold protocol. A T1 weighted fast gradient echo sequence with short TR and TE (TR 3.3 ms TE 1.2 ms, flip angle 20°) was used for stress and rest perfusion with a typical acquired voxel size of 2.9 × 2.9 mm² and typical matrix size of 128 × 128 (frequency × phase). A 90-degree non-selective saturation preparation pulse was applied prior to every acquired slice with a saturation delay time of 100 ms. A parallel imaging acceleration factor of 2 was used to reduce slice acquisition time. Heart rate (HR) and blood pressure were monitored during the stress and rest scans. A HR increase ≥ 10 beats per minute (bpm) was regarded as a positive HR response to adenosine [14]. However, as HR response has been shown to be influenced by several parameters like age and sex [15], we used MFR as assessed by PET to define true coronary adenosine response. Taking into account a reported day-to-day intra-subject variability of MFR of up to 20% [16], we defined an MFR of > 1.5 or the presence of ischemia on PET as markers for adequate coronary adenosine response.

Visual and quantitative assessment of SSO
Presence of SSO was defined as a visually perceivable lower splenic enhancement on stress than on rest first-pass CMR images, as previously reported [7]. Visual analysis was performed independently by two readers in a blinded fashion. In cases of disagreement between the readers, the decision was made by consensus. Inter-rater variability was calculated using Cohen’s Kappa (k). Grey-scale values were normalized, and identical thresholds were used for stress and rest images for better comparability using a commercially available software package (Circle 42, Version 5.6.4, Circle Cardiovascular Imaging, Calgary, Canada).

Myocardial signal intensity curves during first-pass perfusion were derived from CMR. In addition, regions of interest were drawn in the spleen. Corrected peak signal intensity of the spleen was calculated for stress and rest by subtracting baseline intensity (pre-contrast) from peak intensity after first-pass perfusion. Stress-to-rest intensity ratio (SIR) was calculated as the ratio between corrected peak signal intensity of stress and rest images for splenic tissue as previously reported (Fig. 1) [14]. Finally, spleen-to-myocardium intensity ratios for stress (SMR stress) were calculated, defined as the ratio of increase of spleen activity to myocardial intensity during stress and rest, respectively, to ensure standardized evaluation (Fig. 1).

Statistical analysis
Descriptive statistics were used to examine clinical data. Values are presented as averages ± standard deviations or as total numbers and as percentages, where applicable. Patients were stratified into groups according to the presence or absence of SSO and according to a positive or negative HR response (i.e., ≥ or < 10 bpm increase upon adenosine stress). Comparison between the groups was performed using a Chi2-test in nominal or students’ t-test and Mann–Whitney-test in normally distributed and non-parametric variables, respectively. Correlation analysis between SIR and MFR, SMR and MFR, as well as between the HR increase from rest to stress and MFR was performed using Pearson analysis. A cut-off value for SIR and SMR for predicting the presence of SSO was calculated using receiver operating characteristic (ROC) analysis. Statistical analysis was performed using SPSS (version 22, Statistical Package for the Social Sciences, International Business Machines, Inc., Armonk, New York, USA).

Results
Baseline characteristics
A total of 67 patients were included in our study. The spleen was visible within the CMR field-of-view in 64 (96%) patients. The remaining 3 (4%) patients were excluded from further investigations. Baseline characteristics and PET findings are given in Table 1.

13N-ammonia PET myocardial perfusion imaging
The mean injected activity of 13N-ammonia during PET was 297 ± 98 MBq and 463 ± 103 MBq for stress and rest imaging, respectively. Mean hyperemic and rest MBF were 2.1 ± 0.7 ml/min/g and 0.8 ± 0.3 ml/min/g, respectively. Mean MFR was 2.9 ± 1.0. Three (5%) patients with an MFR ≤ 1.5 and without any signs of ischemia neither in PET nor CMR were classified as adenosine non-responders. Three (5%) additional patients with severe coronary artery disease had an MFR ≤ 1.5 but presented with clear signs of ischemia in 13 N-ammonia PET and CMR and were therefore classified as adenosine responders.

SSO and quantitative assessment of splenic perfusion
SSO was present in 46 (72%) patients. Inter-rater reliability was excellent (κ = 0.906; p < 0.001). Among the three adenosine non-responders, two patients did not present with SSO, but one patient showed SSO. MFR differed significantly in patients with vs. patients without presence of SSO (3.1 ± 1.0 vs. 2.5 ± 1.0; p = 0.038). Splenic stress-to-rest SIR was significantly lower in
patients with vs. patients without presence of SSO (SIR: 0.56 ± 0.13 vs. 0.93 ± 0.23; p < 0.001).

ROC analysis yielded an optimal cut-off value of SIR for predicting presence of SSO of 0.71 (sensitivity: 94%; specificity: 94%; AUC = 0.947). SMR stress was significantly lower in patients with vs. patients without presence of SSO (1.1 ± 0.5 vs. 1.7 ± 0.6; p < 0.001). ROC analysis yielded an optimal cut-off value of SMR stress for predicting presence of SSO of 1.53 (sensitivity: 61%; specificity: 85%; AUC = 0.76).

A weak correlation was found between MFR and SIR (R = -0.356; p = 0.004) as well as between MFR and SMR stress (R = -0.378; p = 0.02). By contrast, there was no correlation between SMR stress or SIR and the total increase of HR during adenosine (SMR stress: R = -0.115; p = 0.37, SIR: R = -0.143; p = 0.26).

**HR response**

HR increased significantly from rest to stress (62 ± 10 vs. 74 ± 19 bpm, p < 0.001), and 29 (45%) patients had a positive HR increase of ≥ 10 bpm. There was a trend towards higher MFR in patients with vs. patients without a positive HR response (3.2 vs 2.7 ml/min/g; p = 0.053) but there was no difference in the proportion of patients with and without a positive HR response among patients with or without SSO (p = 0.23).

**Discussion**

To the best of our knowledge, this is the first study investigating the performance of SSO to predict coronary adenosine response on a hybrid PET/CMR device with co-injection of GBCA and 13N-ammonia using PET-derived MFR as the standard of truth. While we found that 95% of the patients were coronary adenosine
responders, SSO was absent in 28% of the patients. Furthermore, there was only a weak correlation between SMR and MFR as well as between SIR and MFR. Hence, from our results, we conclude that the absence of SSO should not necessarily lead to the conclusion that a patient failed to respond to adenosine. By contrast, however, the presence of SSO in case of normal stress perfusion CMR is a strong indicator for a true negative finding. Our findings are in line with the results of Kuijpers et al. who demonstrated that SSO failed to predict adenosine non-responders after caffeine intake [17]. By contrast, Manisty et al. assessed a sub-population of the CE-MARC study, comparing 35 false negative CMR scans (defined as normal CMR but significant coronary lesion in quantitative coronary angiography) with 65 true negative CMR scans in a blinded fashion [7, 18]. They found that the absence of SSO was almost four times more likely in patients with a false negative CMR than in patients with a true negative finding. Hence, the authors concluded that absence of SSO may be a predictor of inadequate coronary response to adenosine infusion [7]. However, one limitation of this study was the highly ambiguous standard of truth consisting solely of the hemodynamic response. By contrast, by performing co-injection on a PET/MR device, the current study uses a solid standard of reference in the form of PET-derived MFR.

In our study, SSO was present in 72% of the patients, a proportion which is slightly lower than previously described by Manisty et al. (90%) [7] and Hosking et al. (89%) [14]. However, our calculated thresholds of 0.71 for SIR for predicting visually perceivable SSO was higher than in the study by Hosking et al., which reported a threshold of 0.4 [14]. This more sensitive approach should have led to an even higher presence of SSO. Of note, however, both Manisty et al. [7] and Hosking et al.

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### Table 1 Baseline characteristics and comparison of PET results of patients with positive vs negative splenic switch off

|                        | All patients | SSO present n = 46 (72%) | SSO absent n = 18 (28%) | p-Value |
|------------------------|--------------|--------------------------|-------------------------|---------|
| **Baseline characteristics** |              |                          |                         |         |
| Mean age (years)       | 51 ± 20      | 51.4 ± 18                | 49.2 ± 24               | 0.69    |
| Male gender            | 52 (81)      | 37 (80)                  | 15 (83)                 | 0.30    |
| Weight (kg)            | 80 ± 11      | 81 ± 10                  | 75 ± 12                 | 0.047   |
| Height (cm)            | 175 ± 8      | 175 ± 8                  | 176 ± 8                 | 0.65    |
| Body mass index (kg/m2)| 26 ± 3       | 27 ± 3                   | 24 ± 3                  | 0.006   |
| **Risk factors**       |              |                          |                         |         |
| Diabetes mellitus      | 5 (7.8)      | 5 (11)                   | 0 (0)                   | 0.145   |
| Dyslipidaemia          | 27 (42)      | 21 (46)                  | 6 (34)                  | 0.37    |
| Hypertension           | 25 (39)      | 20 (43)                  | 5 (28)                  | 0.247   |
| Family history         | 16 (25)      | 12 (26)                  | 4 (23)                  | 0.748   |
| Smoking                | 21 (33)      | 17 (37)                  | 4 (23)                  | 0.259   |
| **Medication**         |              |                          |                         |         |
| Aspirin                | 24 (38)      | 17 (37)                  | 7 (39)                  | 0.886   |
| Betablocker            | 18 (28)      | 15 (33)                  | 3 (17)                  | 0.202   |
| ACE inhibitor          | 24 (38)      | 18 (39)                  | 6 (34)                  | 0.667   |
| Statin                 | 27 (42)      | 19 (41)                  | 8 (45)                  | 0.819   |
| **Medical history**    |              |                          |                         |         |
| Myocardial infarction  | 10 (16)      | 7 (15)                   | 3 (17)                  | 0.886   |
| Stenting               | 18 (28)      | 13 (28)                  | 5 (28)                  | 0.969   |
| CABG                   | 7 (11)       | 4 (9)                    | 3 (170)                 | 0.358   |
| **13N-ammonia PET**    |              |                          |                         |         |
| Ischemia               | 15 (23)      | 9 (20)                   | 6 (34)                  | 0.242   |
| Scar                   | 33 (52)      | 22 (48)                  | 11 (61)                 | 0.1     |
| Stress MBF (ml/min/g)  | 2.12 ± 0.68  | 2.2 ± 0.66               | 1.93 ± 0.72             | 0.23    |
| Rest MBF (ml/min/g)    | 0.78 ± 0.26  | 0.77 ± 0.27              | 0.81 ± 0.24             | 0.65    |
| MFR                    | 2.91 (± 1)   | 3.07 ± 1.03              | 2.48 ± 0.96             | 0.038   |

Values given are average ± standard deviations or absolute numbers and percentages (in brackets).

ACE angiotensin converting enzyme, CABG coronary artery bypass graft, CAD coronary artery disease, MBF myocardial blood flow, MFR myocardial flow reserve.

SSO splenic switch-off
increased the injected dose of adenosine in patients who failed to show hemodynamic changes during stress. By contrast, in the present study, we infused a fixed dose of 140 ug/kg/min. The literature on the adequate dose of adenosine reveals conflicting results with some studies raising the question whether adenosine infusion at a standard rate of 140 ul/kg/min induces maximal coronary vasodilation in all patients [19, 20]. Other studies have demonstrated that increasing the dose of adenosine beyond 140 ug/kg/min does not result in further coronary vasodilation [21, 22], but only induces a more aggravated peripheral hemodynamic response [20], potentially yielding a higher rate of SSO but without a positive effect on the coronary arteries. Hemodynamic response, as well as SSO are peripheral side effects induced by A2B receptors due to aortic hypotension with consequent reflex tachycardia and vasoconstriction of vascular beds in order to maintain mean arterial pressure. By contrast, coronary vasodilatation is induced through stimulation of A2A receptors [2]. In view of the different receptors, however, it may seem inadequate to draw conclusions as to the extent of coronary vasodilation through extrapolation of any observed peripheral effects. Moreover, accurate assessment of the splenic perfusion itself is difficult and may be hampered due to the spleen’s physiologically inhomogeneous perfusion of the red and white pulp. Hence, areas of lower and higher perfusion are simultaneously assessed by CT, ultrasound, and CMR during the arterial phase of perfusion [23, 24]. Finally, spleen visualization and analysis during myocardial perfusion imaging are currently not standardized. In the present study, one patient without SSO was classified as an adenosine responder as per the standard of reference. Of note, however, the SIR in this patient was 0.68 which lies slightly below the calculated SIR-threshold (i.e., 0.71) necessary to allow for visual recognition of SSO. This finding underlines the potential shortcomings and inherent uncertainties of a visual assessment of SSO. However, the future clinical value of SSO should be put into perspective as the advent of vasodilators such as regadenoson, which are more specific to A2A receptors and may confer a better safety profile than adenosine, may gradually reduce the clinical value of the latter in the future.

The present study extends our knowledge and understanding of the SSO sign. While our results question its value as a reliable marker of inadequate adenosine response, future prospective studies—ideally placebo- or caffeine-controlled—are also needed. It is noteworthy and important, however, that our results on the other hand also carry the reassuring message that the presence of SSO is a strong marker for adequate coronary adenosine response, therefore increasing a readers’ confidence in case of a normal CMR myocardial perfusion scan.

Limitations
In the present study, we used a binary system to classify patients into responders vs. non-responders. This may be seen as an over-simplification in light of some studies suggesting a more graduated adenosine response, depending on receptor density and responsiveness of adenosine receptors [25].

Conclusion
The presence of SSO implies adequate coronary adenosine-induced MBF response. Its absence, however, is not a reliable indicator for failed adenosine-induced coronary vasodilatation.

Abbreviations
CAD: Coronary artery disease; ECG: Electrocardiogram; HR: Heart rate; GBCA: Gadolinium based contrast agent; MFB: Myocardial blood flow; MBq: Megabecquerels; MFR: Myocardial flow reserve; MPI: Myocardial perfusion imaging; PET: Positron emission tomography; ROC: Receiver operating characteristic; SIR: Stress-to-rest intensity ratio; SMR: Spleen-to-myocardium ratio; SSO: Splenic switch off; TOF: Time-of-flight.

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Authors’ contributions
DP analyzed and interpreted the data, was involved in patient recruitment and imaging, and was involved in manuscript drafting. EvF and AB were involved in data analysis and interpretation as well as with conceptualization, data management. Additionally, EvF was involved in patients recruitment and imaging. AG, CK, MS, MM, DB, CG, and AP were involved in study design and manuscript reviewing. PK supported the study in terms of conceptualization and academic advice and by providing the human resources. TF and RB performed overall supervision and were involved in all aspects of the study. All authors read and approved the final manuscript.

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Availability of data and materials
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate
This study was approved by the ethics committee of Zurich (KEK-ZH-Nr. 2014-0187 and BASEC-Nr. 2018-00170). All patients provided written informed consent.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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References

1. Dilisizian V, Bacharach SL, Beanlands RS, et al. ASCN imaging guidelines/ SNMMI procedure standard for positron emission tomography (PET) nuclear cardiology procedures. J Nucl Cardiol. 2016;23:1187–226.

2. Hentzova M, Duval W, Einstein AJ, Travin MI, Verberne HJ. ASCN imaging guidelines for SPECT nuclear cardiology procedures: Stress, protocols, and tracers. J Nucl Cardiol. 2016;23:606–39.

3. Nguyen K-L, Bandettini WP, Shanbhag S, Leung SW, Wilson JR, Arai AE. Safety and tolerability of regadenoson CMR. Eur Heart J Cardiovasc Imaging. 2014;15:753–60.

4. Writing Committee M, Acrf Task Force M, ACcf/ACR/AHA/NASCI/SCMR. Expert consensus document on cardiovascular magnetic resonance a report of the American College of Cardiology Foundation Task Force on Expert Consensus Documents. Circulation. 2010;2010(12):2462–508.

5. Mishra RK, Dorbala S, Logsetty G, et al. Quantitative relation between hemodynamic changes during intravenous adenosine infusion and the magnitude of coronary hyperemia: implications for myocardial perfusion imaging. J Am Coll Cardiol. 2005;45:533–8.

6. Amanullah AM, Berman DS, Kiat H, Friedman JD. Usefulness of hemodynamic changes during adenosine infusion in predicting the diagnostic accuracy of adenosine technetium-99m sestamibi single-photon emission computed tomography (SPECT). Am J Cardiol. 1997;79:1319–22.

7. Manisty C, Ripley DP, Herrey AS, et al. Splenic switch-off: a tool to assess stress adequacy in adenosine perfusion cardiac MR imaging. Radiology. 2015;276:732–40.

8. Rooney MW, Crystal GJ, Salem MR, Paulissian R. Influence of nifedipine on systemic and regional hemodynamics during adenosine-induced hypotension in dogs. Anesth Analg. 1989;68:261–9.

9. Hutchins GD, Schweiger M, Rosenspire KC, Knviokapich J, Schelbert H, Kuhl DE. Noninvasive quantification of regional blood flow in the human heart using N-13 ammonia and dynamic positron emission tomographic imaging. J Am Coll Cardiol. 1990;15:1032–42.

10. Chan SY, Brunkcn RC, Czernin J, et al. Comparison of maximal myocardial blood flow during adenosine infusion with that of intravenous dipyridamole in normal men. J Am Coll Cardiol. 1992;20:979–85.

11. Nazir MS, Ismail TF, Reyes E, Chiribiri A, Kaufmann PA, Plein S. Hybrid positron emission tomography–magnetic resonance of the heart: current state of the art and future applications. Eur Heart J Cardiovasc Imaging. 2018;19:962–74.

12. Fratz S, Chung T, Greil GF, et al. Guidelines and protocols for cardiovascular magnetic resonance in children and adults with congenital heart disease: SCMR expert consensus group on congenital heart disease. J Cardiovasc Magn Reson. 2013;15:51.

13. Lassen ML, Rasul S, Beitzke D, et al. Assessment of attenuation correction for myocardial PET imaging using combined PET/MRI. J Nucl Cardiol. 2017. https://doi.org/10.1007/s12350-017-1118-2.

14. Hosking A, Koulouroudias M, Zemrak F, et al. Evaluation of splenic switch off in a tertiary imaging centre: validation and assessment of utility. Eur Heart J Cardiovasc Imaging. 2017;18:1216–21.

15. Gebhard C, Messerli M, Lohmann C, et al. Sex and age differences in the association of heart rate responses to adenosine and myocardial ischemia in patients undergoing myocardial perfusion imaging. J Nucl Cardiol. 2018. https://doi.org/10.1007/s12350-018-1276-x.

16. Kikutungvan D, Johnson NP, Roby AE, Patel MB, Kirkeeide R, Gould KL. Routine clinical quantitative rest stress myocardial perfusion for managing coronary artery disease: clinical relevance of test-retest variability. JACC Cardiovasc Imaging. 2017;10:565–77.

17. Kuipers D, van Dijk R, van Assen M, et al. Disagreement between splenic switch-off and myocardial T1-mapping after caffeine intake. Int J Cardiovasc Imaging. 2018;34:625–32.

18. Greenwood JP, Maredia N, Younger JF, et al. Cardiovascular magnetic resonance and single-photon emission computed tomography for diagnosis of coronary heart disease (CE-MARC): a prospective trial. Lancet. 2012;379:453–60.

19. Wilson RF, Wyche K, Christensen BV, Zimmer S, Laxson DD. Effects of adenosine on human coronary arterial circulation. Circulation. 1990;82:1595–606.

20. Karamitsos TD, Tsusi NA, Francis JM, Holloway CJ, Myerson SG, Neubauer S. Feasibility and safety of high-dose adenosine perfusion cardiovascular magnetic resonance. J Cardiovasc Magn Reson. 2010;12:66.

21. De Bruyne B, Pijls Nico HJ, Barbato E, et al. Intracoronary and intravenous adenosine 5′-triphosphosphate, adenosine, papaverine, and contrast medium to assess fractional flow reserve in humans. Circulation. 2003;107:1877–83.

22. Kaufmann PA, Gnecki-Ruscone T, di Terlizzi M, Schäfers KP, Lüscher TF, Camici PG. Coronary Heart Disease in Smokers. Circulation. 2010;82:1595–606.

23. Palas J, Matos AP, Ramalho M. The spleen revisited: an overview on systemic and regional hypoperfusion in dogs. Anesth Analg. 1989;68:261–9.

24. Rabushka LS, Kawashima A, Fishman EK. Imaging of the spleen: CT with supplemental MR examination. Radiographics. 1994;14:307–32.

25. Martin EA, Nicholson WT, Eisenach JH, Charkoudian N, Joyner MJ. Influence of adenosine receptor antagonism on vasodilator responses to adenosine and exercise in adenosine responders and nonresponders. J Appl Physiol. 2006;101:1678–84.

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