The arrival of commercial bioprinters – Towards 3D bioprinting revolution!

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Abstract: The dawn of commercial bioprinting is rapidly advancing the tissue engineering field. In the past few years, new bioprinting approaches as well as novel bioinks formulations have emerged, enabling biological research groups to demonstrate the use of such technology to fabricate functional and relevant tissue models. In recent years, several companies have launched bioprinters pushing for early adoption and democratisation of bioprinting. This article reviews the progress in commercial bioprinting since the inception, with a particular focus on the comparison of different available printing technologies and important features of the individual technologies as well as various existing applications. Various challenges and potential design considerations for next generations of bioprinters are also discussed.

Keywords: Bioprinter; bioprinting; bioinks; 3D printing; rapid prototyping; biofabrication; tissue engineering

1. Introduction to 3D Bioprinting

1.1 Definition

The advent of 3D bioprinting has opened exciting new possibilities for tissue engineering, reconstruction and in vitro drug testing studies[1,2]. This additive manufacturing-based (AM) biotechnology utilises bioinks[3] along with living cells (cell aggregates or tissue spheroids), to spatially construct 3D functional structures without pre-fabricated scaffolds. Before the more generic term of bioprinting, organ printing was first coined in 1999 and was defined as computer-aided, jet-based 3D tissue-engineering of living human organs[4]. Since then, a host of other printing technologies have been developed as well as new bioink formulations[5,6]. Lately, bioprinting is defined as “computer-aided transfer processes for patterning and assembly of living and non-living materials with a prescribed 2D or 3D organization to produce bio-engineered structures serving in regenerative medicine, pharmacokinetics, and basic cell biology studies[7]. The sole purpose of bioprinting is to engineer a fully integrated and functionally restored biological environment[8] which could be achieved either through scaffold printing and subsequent cell seeding or direct cell printing. The scaffold-based fabrication approach involves mimicking of the extracellular matrix (ECM) by printing a temporary and biodegradable supporting structure[9]. Incorporation of live cells into the bioink increases/magnifies the complexity involved in printing but promises a more homogeneous distribution of cells as compared to dropwise seeding over a 3D printed scaffold such that the resulting three-dimensional cellular constructs better mimic complex biological functionalities found in native tissues and organs.
1.2 Market, Research and Patent Landscape

Bioprinting is a revolutionary tissue engineering (TE) strategy that holds immense potential as a manufacturing platform for fabrication of in vitro tissue. A market research report by BCC Research pegged the global bioprinting at US$ 263.8 million in 2015. It forecasted the market to reach US$ 295 million in 2016 and shoot to $1.8 billion by 2021. This growth is calculated at a compound annual growth rate (CAGR) of 43.9% from 2016 to 2021. Another report valued the global 3D bioprinting market at US$ 682 million in 2016 with significant future increases. It forecasted the market to reach US$ 2.6 billion by 2024. The market growth is expected to be driven by new printing technologies as well as the expansion of new applications beyond the medical field. However, medical applications (including toxicity screening, organ transplants) are expected to retain the largest growth at 30%.

Bioprinting as a field has been very technology intensive. John Hornick and Kai Rajan evaluated the 3D bioprinting patent landscape and found, as of June 2016, 950 patents and pending applications filed by more than a hundred companies. This diverse list included both small enterprises and MNCs based in various geographical locations, suggesting a strong global interest and drive. In terms of the patent portfolio, the leaderboard was occupied by Organovo followed by Koninklijke Philips and Wake Forest University respectively. A very detailed scientometric and patentometric analyses of the field of 3D bioprinting was carried out by Rodriguez-Salvador et al. using competitive technology intelligence methodology. The countries currently leading the patent race are China and USA while Organovo and Tsinghua University came out as the leading institutions (Figure 1). The patent analysis from the year Jan 2000 – July 2016 revealed at least 345 patent families (PFs). The overall proportion of patent applications–patents granted–inactive patents stood at 70%–17%–13%. Biomaterials and Biofabrication are the most popular scientific journals for publishing. The four major knowledge clusters identified are: tissue engineering (163 PFs), tissue or organ (67), polylactic acid (46) and 3D printer (45). Eight key drivers identified by the experts in the field are: (a) tissue engineering, (b) 3D bioprinting system, (c) bioinks, (d) fibres and scaffolds, (e) human body models, (f) regenerative medicine, (g) pharmaceutical research and (h) vascularization. The linkage between those knowledge clusters and the key drivers have been demonstrated. As of now, companies within the bioprinting industry can be broadly categorized into three categories—companies selling only commercial systems, companies providing the bioprinted tissue and companies providing bioprinting system as a service.

1.3 Common 3D Bioprinting Technologies

Almost all the current bioprinters make use of the traditional material deposition techniques (extrusion or ink-jet) or modern optics-based/light-based (laser-assisted or stereolithography) for printing (Table 1).

1.3.1 Extrusion Bioprinting

The extrusion-based approach is the most common technology implemented by the majority of commercial 3D bioprinters primarily due to cheaper assembly and operational costs. The technique facilitates extrusion of cylindrical filaments of bioink, employing either a pneumatic (air pressure), mechanical (piston) or solenoid (electrical pulses) control. Extrusion-based bioprinters enjoy clear advantages such as greater deposition and printing speed, but their higher throughput comes at the expense of lower resolution as compared to the other technologies.

1.3.2 Inkjet Bioprinting

Inkjet bioprinting is a concept directly borrowed from the conventional paper printing. The printing setup includes a reservoir of ink, where an acoustic wave is produced to dispense a polymeric solution through the nozzle. This acoustic wave can either be

Table 1. Comparison of bioprinting approaches

| Parameters           | Microextrusion | Inkjet | Laser-assisted | Stereolithography |
|----------------------|----------------|--------|----------------|-------------------|
| Material viscosity   | 30 to > 6 × 10⁷ mPa/s | 3.5 – 12 mPa/s | 1 – 300 mPa/s | No limitation     |
| Crosslinking strategy| Photocuring, thermal, chemical | Photocuring, chemical | Photocuring, chemical | Photocuring, chemical also |
| Cell viability       | 40% – 80% | > 85% | > 95% | > 85% |
| Cell density         | High | Low | Medium | Medium |
| Printing speed       | Slow | Fast | Moderate | Fast |
| Printing resolution  | Medium | High | High | High |
| Cost of printer      | Medium | Low | High | Low |
generated through air bubbles or piezoelectric actuator\cite{16}. Since the technique requires a low-viscous bioink for successful deposition, an additional crosslinking step is accompanied in most of the cases\cite{15}.

1.3.3 Laser-assisted Bioprinting

Laser-assisted bioprinting is an emerging technology that allows bioprinting of cells and hydrogels with a cell-level resolution (Figure 2C). The technique has shown great potential towards the realisation of scaffold-free 3D cell systems\cite{17}. It could be classified into two broad categories:

- Laser-guided direct cell printing: Manipulating the difference in refractive index of the cells and suspending solution, the laser traps and guides these cells in the desired direction\cite{18}.

![Figure 1](image)

**Figure 1.** Distribution of 3D bioprinting patents (Adapted from\cite{19}): (A) The top 10 assignee institutions as per the total number of patent families from 1st Jan 2000 till 1st July 2016. Three institutions are tied for the tenth position; (B) The top 10 most countries in which priority filing was done for the first patent of a patent family. Four countries are tied for the tenth position.
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Figure 2. Schematics of various types of bioprinting technologies: (A) Extrusion bioprinting; (B) Inkjet bioprinting; (C) Laser-assisted bioprinting and; (D) SLA bioprinting

- Laser-induced direct cell printing: The laser beam is focussed on the aqueous bioink via a laser-energy absorbing membrane. This phenomenon generates a vapour bubble, resulting in a mechanical pressure enough to extrude the bioink on the substrate [19].

1.3.4 Stereolithography Bioprinting

The basic principle of Stereolithography is selective crosslinking of a photosensitive biomaterial using a light source, which propagates into the printing solution, thereby activating the photoinitiator and crosslinking the polymeric chains together (Figure 2D) [20]. Several motors are coupled together to facilitate the movement of this light source along X-Y plane, whereas that of the printing support in the Z axis. This arrangement ensures a conventional layer-by-layer fabrication of the desired three-dimensional structure [21].

1.4 Applications of 3D Bioprinting

Bioprinting enables a wide variety of applications, encompassing many industry sectors.

1.4.1 Regenerative Medicine (Engineered organs for transplantation)

The holy grail of tissue engineering is to achieve a fully functional organ which can be transplanted into a patient. However, there are many complex challenges which are hindering fabrication of functional organs: (a) vascularization to the extent of capillaries (b) integrating all the various cell types to achieve complex organotypic biology (c) a stable structural and mechanical integrity (d) innervation (e) maturation of printed construct inside a bioreactor and long-term stable functions.

The bioprinted skin models mostly involved two cell types, fibroblasts and keratinocytes and did not take into consideration melanocytes, the pigment-producing cells until Ng et al. [26] demonstrated 3D in vitro pigmented human skin constructs by incorporating melanocytes via a two-step drop-on-demand bioprinting strategy. Solid organs are very challenging to print because they require innervation, vascularization, and have dense biomass. However, many attempts have been made for bioprinting solid tissues such as bone, cartilage, tendon, lung, liver, cardiac and neural etc. Kang et al. [36] using their specially customised integrated tissue-organ printer (ITOP) demonstrated that it is possible to print centimetre scale tissue/organ constructs of the mandible and calvarial bone, cartilage and skeletal muscle. Detailed list of bioprinted tissues and organs have been covered elsewhere [37].
1.4.2 Drug Discovery and Drug Development

Bioprinting can advance pharmaceutics under four different broad categories\textsuperscript{[38]}: (a) Drug delivery (b) Drug screening (c) Microarrays and high-throughput screenings and (d) Absorption, distribution, metabolism and excretion (ADME) assays. Compared to other in vitro models, 3D printed human tissues have better spatial control of cells, in vivo-like tissue microarchitecture, scalability, easier handling, co-culture capability, cell-cell and cell-matrix interactions and low risk of cross-contaminations\textsuperscript{[2]}. Bioprinting potentially can also ensure controlled delivery of growth factors and possibly genes, an important consideration for longer tissue cultivation\textsuperscript{[39]}. Additionally, these printed constructs should be open to complicated phenotypic assays used inside a person including biochemistry, histology and various ‘omics’ methods\textsuperscript{[40]}. The co-printing technology of multiple materials facilitates even closer replication of cellular microenvironment, opening novel approaches for drug screening. To date, simplified in vitro models of liver\textsuperscript{[1,32,41,42]} and kidney\textsuperscript{[43]} have been successfully fabricated using bioprinting.

1.4.3 Disease Modelling

The premise of modelling disease in vitro is to get the diseased model as similar to that in the actual human body and bioprinting can essentially advance this research. One example is the cancer metastasis process; the mechanical properties (such as stiffness) and composition of a tumour microenvironment often undergo quick changes\textsuperscript{[44,45]}, making it challenging to diagnose and treat. Bioprinting can enable the fabrication of various structural aspects of a tumour thereby creating a realistic tumour microenvironment with heterogeneous cells and orders of complexity complete with vasculature\textsuperscript{[40,45]}. To date, bioprinting has been employed to create blood vessels of different diameters to study cancer cell migration\textsuperscript{[46]} as well as uniform tumour spheroids with hollow necrotic cores\textsuperscript{[47]}. The cardiovascular disorder is another prevalent modern life-style dependent disease which can be modelled using bioprinting\textsuperscript{[48]}

1.4.4 Bio-hybrid Robotics

In recent times, there has been a gradual shift towards soft robotics from the conventional one\textsuperscript{[49]}. Researchers have been cautiously inching towards secundum naturam (according to nature) for a better understanding of the intricate organization in living beings, and their subsequent biomimicry\textsuperscript{[4]}. This interest germinates from the desire to tackle more complex, unpredictable environments, which demands higher order of mechanical intelligence. Hybrid bio-bots are defined as an intelligent assembly of highly deformable materials that can be activated by an external stimulus to generate desired motions\textsuperscript{[50,51]}. Bashir et al.\textsuperscript{[52]} first conceptualised the notion of 3D printed “bio-bots” by introducing a class of miniaturized walking biological machines, powered by beating cardiac cells. Through precise patterning of cell-laden hydrogels, it is possible to “build” robots that are powered by cells.

1.4.5 Other Emerging Applications

Another upcoming application for bioprinting lies in the consumer product domain. AM has recently emerged as a promising strategy for designing food materials with complex geometry, detailed patterning, and customized nutritional value\textsuperscript{[53]}. One such revolutionary approach has been the production of tissue engineered meat products through drop-on-demand deposition. Researchers at the University of Missouri–Columbia have manipulated AM-based tissue engineering for the development of comestible food products as an excellent source of protein\textsuperscript{[54]}. Their patents have primarily focused on initiating an alternate technology to meet the growing demand for consumable meat, which so far has been exclusively catered by the overburdened livestock industry\textsuperscript{[54,55]}

In an interesting study, Schroeder et al.\textsuperscript{[56]} made a softer artificial electric organ (power source) by getting inspired from an electric eel. The construct was made up of gradients of ions sandwiched between tiny polyacrylamide hydrogel compartments which were bounded by a repeating sequence of cation- and anion-selective hydrogel membranes. This electric organ is soft, transparent and flexible in comparison to traditional batteries. In future, such artificial electric organs could potentially power advanced implant materials such as pacemakers and implantable sensors etc.

Green bioprinting\textsuperscript{[57]} is defined as an AM approach which involves processing of cells from the plant kingdom for studying secondary metabolites production and monitoring methods. Plant bioprinting can also potentially revolutionize many applications like, production of plant-based materials from printed tissues, chimeric grafting, in situ/in vivo bioprinting for repairing tissues and printing designer plant-based food\textsuperscript{[56]}

2. Commercialization of 3D Bioprinting Technology

The expanded applications of bioprinting have driven the development of bioprinting processes as well as platforms over the last few years (Figure 3). Many new companies have emerged to exploit this upcoming industry, and there are three key areas/business models here: (a) directly selling the bioprinters, (b) providing contractual bioprinting services and (c) directly entering
with a partnership with a client who has a tissue/organ model of interest. All these companies are based on their unique value proposition of providing bioprinting services or partnerships for manufacturing of functional tissues. In the following section 2.2, we have highlighted all the bioprinting companies (in alphabetical order) which are currently in operation to our knowledge. The detailed specifications of printer dimensions, nozzle diameter, build volume, resolution of printing, materials that can be printed, additional interesting features have been duly summarized in a table (Table 2). High-resolution images of these printers have been included in Figures 4–6 (courtesy of the companies). We have also included a chronological order of important events/technological advancements in the field of bioprinting (Figure 7) which shows the tremendous progress made just under two decades. Lastly, we have also summarized the current challenges and future innovations which could be integrated with a bioprinter. We hope this review will benefit uninitiated researchers who want to venture into the exciting and revolutionary world of bioprinting as well as those experienced researchers who may not be aware of the sheer number of the companies in bioprinting.

### 2.1 The Case Study of the Industry Pioneer-Organovo

In 2003, Boland et al. patented the first ink-jet technology for printing viable cells. The team subsequently secured a US$ 5 million National Science Foundation (NSF) grant which led to their first major patent on Novo Gen MMX Bioprinter™ (Figure 5B) platform soon after in 2005. The first 3D bioprinting company, Organovo came into existence in 2007. Organovo partnered with Invetech (Australia) to launch their bioprinting system-Novogen MMX Bioprinter™ in 2009.

Organovo had their first breakthrough in 2010 when working with a grant from National Institutes of Health (NIH); the company successfully bioprinted fully cellular blood vessels using only primary human cells. Organovo’s ExVive™ 3D Bioprinted Human Liver Tissue Models debuted in 2014. The tissues demonstrate histological and functional similarity to the native liver with consistent expression of albumin, ATP and CYP3A4 activity up to 28 days. The tissue model has been used to demonstrate the clinical effect of drugs such as Valproic acid and Monocrotaline. In 2016, Organovo launched ExVive™ Human Kidney Tissue, a fully 3D bioprinted human tissue consisting of an apical layer of polarized primary renal proximal tubule epithelial cells (RPTECs), supported by a collagen IV-rich tubulointerstitial interface of primary renal fibroblasts and endothelial cells. ExVive™ Human Kidney Tissue exhibits in vivo like barrier function, expression of renal transporters and secretion of important enzyme gamma-glutamyltransferase (GGT). This bioprinted kidney tissue when exposed to the drug Cisplatin releases kidney injury biomarkers as well as depicts transporter-dependent (OCT2) drug uptake.

Apart from commercial tissues, Organovo also offers two kinds of partnerships: custom tissue partnerships (co-develop in vitro tissues for drug screening, disease modelling and therapy) as well as technology access partnerships (access to their Novogen Bioprinter™ to experts on a specific tissue/disease model). To date, Organovo has inked significant multi-year collaborations with Merck (drug screening) and with world’s largest...
cosmetics company, L’Oréal (screening of cosmetics in printed dermal tissue).  

2.2 Commercial 3D Bioprinting Companies

Since the launch of Organovo, many other bioprinting companies have entered the market. Many of them offer bioprinting systems for research use with several them also selling proprietary bioinks while others such as Tevido have focussed on recreating human tissue. In this section, we highlight the technology and differentiating factor of each of these companies.

3D Bioprinting Solutions (3dbio) successfully 3D printed world’s first animal thyroid gland in March 2015 and transplanted into a living mouse. With plans to fabricate artificial thyroid and kidney, 3dbio has also been collaborating with Russia’s national space agency, United Rocket and Space Corporation (URSC) to create a magnetic 3D bioprinter that can fabricate artificial tissues in the International Space Station. 3dbio is a subsidiary of Vivax Bio, a global biotech company. Vivax Bio offers FABION (Figure 5C) bioprinter which 5 nozzles, 3 of which are specially designed to dispense bioink and spheroids of any type or size as well as cell suspensions or other biomaterials.

3Dynamic Systems (3DS), a UK-based 3D Bioprinting company offers two 3D bioprinters called 3Dynamic Alpha (Figure 4F) and Omega, with the focus of constructing 3D transplantable bone and complex tissue constructs for injured patients. 3DS have partnered with Bioink Solutions to offer a new gelatin-based bioink (Gel4Cell®).

Aether introduced its first bioprinter Aether 1 (Figure 4K) in March 2016. The system boasts a multitude of printing capabilities where 10+ different materials, including viscous pastes, gels, ceramics, filament and oils can be brought in use simultaneously. The other optional features include high-resolution motors, a laser system for ultra-high-precision cutting and engraving, CNC milling machine, photocrosslinking UV LED, microscope, 3D electronics printer, and universal modular fabrication device.

Advanced Solutions Life Sciences (ASLS) offers two...
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Figure 5. Medium-sized Bioprinters: (A) 3DDiscovery™ (courtesy of RegenHU, Fribourg, Switzerland); (B) NovoGen MMX Bioprinter™ (courtesy of Organovo, San Diego, United States); (C) FABION (courtesy of 3D Bioprinting Solutions, Russia); (D) 3D-Bioplotter® (courtesy of Envision TEC, Gladbeck, Germany); (E) BioScaffolder (courtesy of GeSim, Radeberg, Germany); (F) RX1™ BIOPRINTER (courtesy of Aspect Biosystems, Vancouver, Canada); (G) BIO X (courtesy of CELLINK, Sweden); (H) INVIVO (courtesy of Rokit, Seoul, Korea); (I) Allevi 6 (courtesy of Allevi, Philadelphia, US).

systems: BioAssemblyBot (3D bioprinter) (Figure 6A) and BioBot™ Basic (Figure 4J). The six-axis robotic arm of BioAssemblyBot is well complemented by the option to load up to ten independent delivery systems during a single print run, thus facilitating the fabrication of more versatile biological 3D scaffolds. The key highlight lies in the form of its software interface: Tissue Structure Information Modeling (TSIM). Both systems also offer features such as rotational stage movement and automated material change.

**Allevi** (previously called BioBots) has a range of printers (Figure 4A and Figure 5I) featuring pneumatically-controlled dual-extrusion print heads and visible light technology[71]. Allevi’s systems also employ an open concept for easy switching between bioinks and a temperature-controlled extruder head (25°C to 120°C). Allevi 6 is WiFi enabled and includes a sophisticated array of sensors and computer vision which automatically tunes the printing parameters for materials[72]. Allevi also offers a wide-range of commercial bioinks[73].

**Aspect Biosystems** have been using their proprietary Lab-on-a-Printer™ microfluidic technology in the latest RX1™ bioprinting platform (Figure 5F)[74]. Equipped with a coaxial flow-focusing technology, which ensures direct extrusion of biological fibres with varied diameters. The system was used to demonstrate the fabrication of an artificial airway, termed 3DBioRingTM. The airway consists of contractile smooth muscle tissue comprising of primary human airway smooth muscle cells. The airway tissue has shown appropriate and reproducible contractions to physiological stimuli (histamine) and dilations in response to pharmacological stimuli (B2-agonist)[75].
Bio3D Technologies launched its first modular bioprinter in August 2014. The key highlights of their technology include an anti-vibration levitating platform, multiple interchangeable printing heads, nozzle-to-platform auto-alignment and remote viewing and control. One of the models, Bio3D Explorer, stands out as being the first foldable bioprinter designed for easy transportation.[76]

BIOLIFE4D is an upcoming biotech firm founded in 2015, with headquarters at Illinois (USA). The company aspires to 3D bioprint a fully functional, patient-specific heart for safe and affordable organ transplantation.[77] They are a strong team of business leaders and biomedical researchers, currently financed through equity crowdfunding in their scientific endeavour.[78] The BIOLIFE4D technology intends to use adult induced pluripotent stem cells (iPSCs) for 3D bioprinting a human heart, after conducting a thorough MRI scan to obtain the specific dimensions for its construction.

CELLINK markets both bioprinters and a wide-range of bioinks.[79,80] In fact, CELLINK was the first to commercialize bioinks for bioprinting applications. INKREDIBLE (Figure 4D) is a pneumatic-based extrusion bioprinter with dual print heads and a UV LED system. In 2017, CELLINK launched BIO X (Figure 5G). Some of the key highlights of BIO X include exchangeable print heads, UV curing tool head, cool pneumatic head, HD camera tool head and printbed temperature control (4–60 °C). Bio X is a fully standalone product which offers two different ways of ensuring clean printing chambers: (a) The HEPA H14 filter positive air system which retains germs and other dust particles and (b) UVC germicidal lamps to sterilize the printing chamber.

Cyfuse Biomedical K. K developed their bioprinter Regenova® (Figure 6B) in partnership with Kanazawa-based Japanese medical-device manufacturer, Shibuya Kogyo Co. Ltd. In 2016, the company also collaborated with San Diego-based Cell Applications Inc. allowing them to create scaffold-free, 3D-engineered tissues using the novel Kenzan bioprinting method.[81] Regenova applies a novel robotic approach that enables the fabrication of three-dimensional cellular structures by placing cellular spheroids in a temporary array of fine needles; a methodology has been termed as “Kenzan”. The company in collaboration with Cell Applications...
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EnvisionTEC 3D-Bioplotters (Figure 5D) can be used in sterile biosafety cabinets and feature automated nozzle cleaning process, external temperature sensor ports and layer by layer photographic log[82].

GeSiM is a spin-off from a publicly funded German research laboratory, Helmholtz-Zentrum Dresden-Rossendorf (HZDR). BioScaffolder 3.1 (Figure 5E) features a piezoelectric micro-pipetting system, on the fourth z-axis allowing non-contact dispensing of small drops in the pico- and nanoliter scale[83]. Other add-on alternatives on BioScaffolder include melt electrospinning and a camera to measure strut dimensions.

nScrypt has a patented Micro Dispense Direct Write technology (MDDW)[84]. nScrypt highlights 3Dn 300 TE Series (Figure 6D) uses a positive pressure pump, SmartPump™ to achieve resolutions as small as 15 microns. Along with nTtip™ comes in a wide range of standard sizes from 12.5 microns (inner diameter) 125 microns, the company has demonstrated the possibility to print living cells with near 100% cell viability[85].

Ourobotics’ 8 materials with options to incorporate additional features such as UV, laser and drill for construction of more intricate and challenging architectures[86].

Pensées designed their Vitarix™ Bioprinter (Figure 4G) to work on a patented articulation-based approach instead of Cartesian coordinates[87]. Pensées has also been developing alginate-based bioinks.

Poietis utilises a technology portfolio from INSERM and University of Bordeaux. The company specializes in 4D laser-assisted bioprinting technology and has collaborations with L’Oréal for developing bioprinting hair follicle[88] and BASF for bioprinted skin models[89]. Equipped with an eight-axis motion, their NGB 17.03 (Figure 6C) bioprinting platform is capable of 3D printing with a resolution of a single cell[90]. In early 2018, Poietis has commercially launched the first bioprinted human full skin model Poieskin® fabricated using their NGB bioprinter[91].

REGEMAT 3D successfully bioprint osteochondral tissue using their REGEMAT 3D V1 (Figure 4E) bioprinting system which incorporates three printing technologies: Individually Pore Filling (IPF), Injection Filling (IF) and Fused Deposition Modelling (FDM)[92]. IPF enhances the viability and survival of the cells when working with high temperature thermoplastics whereas IF fills the different printed volumes when working with small injury sites. It offers users to fully configure their bioprinters.

regenHU offers two different models of commercial bioprinters: BioFactory™ (Figure 6F) and 3DDiscovery (Figure 5A)[93]. Key highlights of the systems include a high degree of customizability and a dispensing protocol.
resolution of nanolitre. Both printers can also be further customized to incorporate modified laser or photo-crosslinking devices, multiple dispensing units and a wide variety of software suites. RegenHU has also launched biomaterials for 3D tissue printing: BioInk®, OsteoInk® and Stark®.[96] Regenovo Biotechnology Co., Ltd. markets Bio-Architect® (Figure 6E).[97] The ability to print in a sterilized environment at temperatures ranging from -5 °C to 260 °C is undoubtedly a key highlight of the Regenovo 3D bioprinter.

Revotek’s T-Series™ 3D Bio Printers come with proprietary injection nozzles and specialized, modular Rollovesselar™ platforms which print scaffold-free 3D bio-vascular structures using Biosynsphere™ ink, a proprietary formulation.[98] The Biosynsphere™ ink itself is created using another encapsulation machine made by Revotek. The system comes complete with in-house software, RevoCloud™ which can convert 2D images such as CT scans and MRIs into 3D representations. Working with researchers from West China Hospital at Sichuan University, Revotek has been successful in embedding 3D printed blood vessels into simian test subjects.[50] A 2-centimetre segment of the abdominal artery was replaced with a 3D printed blood vessel in 30 rhesus monkey, where the stem cell bioink was prepared from the autologous adipose mesenchymal stem cells (ADSCs) of the monkeys.[99] The printer currently employs a print head with two nozzles and boasts of printing ten-centimetre blood vessels in just two minutes.[99]

ROKIT received funding from Korean government[100] and partnered with top Korean Universities to develop their bioprinter ROKIT INVIVO (Figure 5H). Major highlights of the system include the interactive user interface (through Android OS system and WiFi connectivity) along with the customised nozzle setup where the user can select from- extrusion, mechanical dispensing and hot melt pneumatic dispensing. INVIVO can print a wide array of biomaterials ranging from PLGA, PCL, PLLA to collagen or gelatin-based hydrogels with cell mixtures.

Seraph Robotics was born out of Cornell University. Scientist™ 3D printer (Figure 4B), the latest model of Seraph Robotics, is based on Fab@Home Model 3 research platform which allows detailed customisation[101]. The printer also comes with UV LED accessory at various wavelengths (365 or 385 nm) for cross-linking. Scientist™ 3D printer is programmed with XDFL instead of the usual G-Code.

SE3D is funded by National Science Foundation (US). The company has been collaborating with high schools to offer hands-on 3D printing curriculum while providing bioprinting tools and training to research labs working in tissue engineering[102]. The key highlight of SE3D is its r3bEL 3D bioprinter (Figure 4H). The printer is based on open-source programs and includes a swappable multi-tool printhead, comprising of a hydrogel extruder (20–100°C) and a fused filament extruder (up to 260°C). SE3D also supplies a wide range of ready-to-use BioKits, with biomaterials and reagents required for initiating educational bioprinting in classrooms.[103]

SunP Biotech International offers several categories of bioprinters (ALPHA-BP11, tower-style ALPHA-CPT1 (Figure 6G), desktop ALPHA-CPD1 and mini ALPHA-CPM1).[104] They have developed a unique low-temp printing technology in ALPHA-BP11 which integrates lyophilisation in 3D printed microfilaments. This results in controlled macro pores (a few hundred microns) for tissue scaffolds and micro pores (10–50 microns) in the printed microfilament. Apart from regular printable scaffold materials, the company also provides personalized bioinks for various in vitro models such as tumours, iPSCs and hepatocytes[105].

Te Vido biodevices utilise patented technology from Clemson University.[106] TeVido’s first product on offer is a bioprinted nipple-areola graft for breast reconstruction. The clinical trials are predicted in 2–3 years’ time.[107] TeVido’s second offering is targeted for Vitiligo patients where they want to print skin tissues which would potentially reduce the contrast in colours.[108] Te Vido biodevices currently does not offer any commercial bioprinters in the market.

### 2.3 Analysis of Commercial Bioprinting Companies

Bioprinting as a technology has a global presence, and it is clearly demonstrated by the locations of those companies (Figure 8). The majority (almost 40%) of the bioprinting companies are based in North America, and rest are split in between Europe and Asia. Almost three-quarters of all the bioprinting companies have been established within the last 6 years. In fact, 20% of all the companies have been in operation for less than 3 years, and this shows the dynamism of this sector. Organovo, the first bioprinting-only company started operating only in the last 10 years, while much older companies like EnvisionTEC, GeSiM and nScrypt (who have been in business for more than 15 years) had their core applications focus on non-bioprinting related technologies. Extrusion-based printing mode is undoubtedly the most popular mode (Figure 9) in the current bioprinters owing to their ease of operation and lower set-up and maintenance costs. The most common form a biomaterial gets printed is in the hydrogel form, and microextrusion is ideally suited for this. More than half of the printing heads in the commercially available
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Table 2. Summary table of commercially available bioprinters
(* IPF: Individually Pore Filling; IF: Injection Filling; FDM: Fused Deposition Modelling; NA: Not available; #: Information unavailable, Unless additional materials are highlighted, all extrusion-based bioprinters are at least capable of printing hydrogels)

| Company                   | Model          | Technology and Materials | Nozzle Diameter (μm) | Build Volume (mm)  | Resolution (μm) | Number of printing heads | Temperature Control |
|---------------------------|----------------|--------------------------|----------------------|--------------------|----------------|--------------------------|---------------------|
|                           |                |                          |                      |                    |                |                          |                     |
|                           |                |                          |                      |                    |                |                          |                     |
| 3Dynamic Systems          | Alpha          | Extrusion                | *                    | 150 × 150 × 60     | ±75 (Accuracy) | 1                        | *                   |
|                           | Omega          |                          |                      | 210 × 140 × 60     | ±50 (Accuracy)  |                          |                     |
| Advanced Life Sciences    | BioAssemblyBot | Extrusion                | *                    | 250 × 300 × 250    | *               | 8 Interchangeable tools   | Tool: 5–110 °C      |
|                           | Biobot         |                          |                      |                    |                |                          | Bed: 5–110 °C       |
|                           |                |                          |                      |                    |                |                          |                     |
| Aether                    | Aether1        | Extrusion, Droplet jetting, FDM | 50                   | 315 × 229 × 132    | 1.055 0.00043  | 10+ Pneumatic: 8 FDM: 2   | Anodized aluminum heated syringe mount and stage |
| Allevi                    | Allevi1        | Extrusion (hydrogel and thermopolymer) | 150                   | 90 × 130 × 60      | 10 10         | 1                        | Nozzle: 4–200 °C Bed: NA |
|                           | Allevi2        | Extrusion                | 150                   | 90 × 90 × 90       | 5.5 5         | 2                        | Nozzle: RT–160 °C Bed: NA |
|                           | Allevi6        | Extrusion Thermoplastics, hydrogels | *                    | *                  | 1 1           | 6                        | Nozzle: 4–200 °C Bed: RT–120 °C |
| Bio3D Technologies        | Explorer       | Extrusion                | *                    | *                  | 5 5           | Up to 4                   | *                   |
| CELLINK                   | INKREDIBLE+    | Extrusion                | 100                   | 130 × 80 × 100     | 10 10         | 2                        | Nozzle: Heating Bed: NA |
| BIO X                     | Extrusion, Inkjet, FDM | *                        | 130 × 90 × 70      | 1 1               | 3 Heated pneumatic: 2 Piston-driven: 1 | Nozzle: 4–130 °C Bed: 4–60 °C |
| Cyfuse biomedical         | Regenova       | Kenzan method            | 170 (Needle)         | 10 × 20 × 18 (9 × 9) 20 × 30 × 18 (26 × 26) | –             | Up to 2 types of NA spheroids |                     |
| Envision TEC              | Bioplotter Starter | Extrusion, Hydrogel, thermoplastic, ceramic, metal paste | 100                | 150 × 150 × 140    | 1 1           | 2 Only high temperature | Nozzle: 30–250 °C Bed: NA |
|                           | Bioplotter Developer |                         | 100             | 150 × 150 × 140    | 1 1           | 3 High and low | Nozzle: 0–250 °C Bed: Available |
|                           | Bioplotter Manufacturer |                     | 100                      | 150 × 150 × 140    | 1 1           | 5 High and low | Nozzle: 0–250 °C Bed: -10 – 80°C |
| GeSiM                     | BioScaffolder 3.1 | Extrusion, Inkjet, Melt electrospinning | *                    | 310 × 200 (Area)  | 2 10          | 3                        | Nozzle: * – 120°C Bed: NA |
| nScrypt                   | 3Dn 300 TE series | Extrusion                | *                    | 300 × 300 × 150    | 0.01 0.5      | 4                        | *                   |
|                           |                |                          |                      |                    | ±5 (Accuracy)  |                          |                     |
| Pensees                   | VitarixTM      | Extrusion                | 200                   | 100 × 60 (Area)    | ±10 (Accuracy) | 2                        | NA                  |
| REGEMAT 3D V1             | IPF, IF, FDM * |                          | 100                   | 150 × 150 × 110    | 150 0.4       | FDM: 2 IPF/IF: up to 8 | *                   |
### Table 2. Continued.

| Company          | Model               | Technology and Materials | Nozzle Diameter (μm) | Build Volume (mm) | Resolution (μm) | Number of printing heads available | Temperature Control |
|------------------|---------------------|--------------------------|----------------------|-------------------|-----------------|-------------------------------------|---------------------|
| regenHU          | 3DDiscovery™        | Extruion, Inkjet Hydrogel, thermopolymer | *                    | *                 | 130 × 90 × 60   | 5                    | 5                   | Up to 9 technologies   | Nozzle: 0–80 °C  
|                  | Bench-Top           |                          |                      |                   |                 |          | Bed: 0–80 °C               |                     |
| Biofactory       | 1st                 | Extruion, Inkjet Hydrogel, thermopolymer | *                    | *                 | 60 × 60 × 60     | 5                    | 5                   | Up to 8               | Nozzle: 5–80 °C  
|                  |                     |                          |                      |                   |                 |          | Bed: 5–80 °C               |                     |
| Regenova         | Bio-Architect Lite  | Extruion                | *                    | 160 × 160 × 150   | *               | 10                   | 10                  | NA                  | Nozzle: RT–300 °C  
|                  |                     |                          |                      |                   |                 |          | Bed: NA                    |                     |
|                  | Bio-Architect Pro   | Extruion                | *                    | 160 × 160 × 150   | 1                | 1                    | 1                   | NA                  | Nozzle: -5–260 °C  
|                  |                     |                          |                      |                   |                 |          | Bed: -5 °C–RT              |                     |
|                  | Bio-Architect WS    | Extruion                | *                    | 170 × 170 × 150   | 1                | 1                    | 1                   | NA                  | Nozzle: -10 – 260 °C |
|                  |                     |                          |                      |                   |                 |          | Bed: -5 °C–RT              |                     |
| ROKIT            | INVIVO              | Bio-dispenser, FDM      | Hydrogel: 50 FDM: 200 | *                 | *               | 100 × 100 × 90   | *                   | 10                  | Nozzle: -10 – 80 °C  
|                  |                     |                          |                      |                   |                 |          | Bed: -4 – 80 °C            |                     |
| Sc3D             | r3EL MINI           | Extruion                | 100                  | 130 × 120 (Area)  | *               | 1                    | 1                   | NA                  | Only for FDM 25 – 260 °C |
|                  | r3EL X              | (Hydrogel and thermopolymer) | 100                | 200 × 200 (Area)  | 2               | 100                  | 2                   | Interchangeable tools |                     |
| Seraph Robotics  | Scientist™ Piston-driven or pneumatic extrusion, Traditional FDM | *                    | *                 | *                 | *               | *                    | *                   | Up to 4              | Nozzle: -3.6 – 80 °C  
|                  |                     |                          |                      |                   |                 |          | Bed: -3.6 – 150 °C         |                     |
| SunP Biotech     | ALPHA-BP11          | Extruion                | 100                  | *                 | *               | *                    | *                   | 2 – 4               | Nozzle: RT – 80 °C  
|                  |                     |                          |                      |                   |                 |          | Bed: -30 °C – RT           |                     |
|                  | ALPHA-CPT1          | Electro-mechanical       | 100                  | *                 | *               | *                    | *                   | 2 – 4               | Nozzle: RT – 37 °C  
|                  |                     |                          |                      |                   |                 |          | Bed: NA                    |                     |
|                  | ALPHA-CPD1          | Extruion                | 50                   | 170 × 285 × 70    | 5                | 5                    | 5                   | 1 – 3               | Nozzle: RT – 37 °C  
|                  |                     |                          |                      |                   |                 |          | Bed: NA                    |                     |
|                  | ALPHA-CPM1          | Extruion                | 100                  | 120 × 160 × 50    | 5                | 5                    | 5                   | 1 – 2               | NA                  |                     |

Bioprinters are microextrusion-based, and the majority are pneumatically controlled. The number of extrusion-based print heads could vary from 1 to about 10 across various bioprinters.

The second most popular print head in bioprinters is FDM-based where they can extrude biocompatible thermoplastics mainly for fabricating scaffolds of good mechanical strength. Only a handful of companies offer ink-jet printing (Figure 9) of cells owing to the technical challenge of getting uniform droplets as well as the practical challenge of getting higher cell densities with them. Although laser-assisted printing ensures the best printing resolution as well as the viability of printed cells, it is the least common print head in the market owing to the cost associated with the laser itself with only one company focussing on laser-assisted bioprinters. Apart from the regular print heads, there has been the development of special print heads/printing technologies in bioprinting. RX1™ Bioprinter (Aspect Biosystems) offers microfluidic print head which ensures higher resolution as well as flexible co-axial flow focusing technology. Regenova Bioprinter (Cyfuse Biomedical) uses the proprietary Kenzan method to generate cellular spheroids. T-Series 3D Bio Printers (Revotek) prints scaffold-free vascular constructs.

A majority of the bioprinters follow the usual 3D printers cartesian-based coordinate system which helps the system to decide where and how to move. They usually have a square print bed which runs along the Y-axis, while the X-axis carries the print head. The Z-axis is for upward or downward motion of the print bed. Ideally if each of the axes is controlled independently then the assembly is very intuitive, easy to operate and would ensure reliable operation. However
many of the bioprinters operate on a Cartesian system which utilizes sliding rails; this in itself is prone to contamination from particles caused by friction.

BioBot™ Basic (ASLS) is a polar coordinate based bioprinting system which offers a rotational stage movement apart from the usual 3-axis motion. Vitarix™ Bioprinter (Pensées) offers a third kind of system called as delta coordinate system with a circular print bed. The extruders are suspended in air above the print bed by three arms in a triangular configuration (thus the name “delta”). This suspension arguably eliminates any contamination from particles caused by friction during its motion.

The nozzle diameters across the extrusion print-heads vary from 50–200 microns. The printing resolution of the bioprinters primarily depends on the printing mode as well the reliability of the associated motor system, but ideally it can go to as low as 1 um across all the 3 axes. The regular extrusion-based print heads have also been made possible to operate at lower temperatures (sub-zero, e.g., 4 deg C) and are being sold as cooled pneumatic heads. Many of the bioprinters are now offering a high-definition (HD) camera tool head to document the work, as well to keep track of the printing process. Many of the companies also sell bioinks which could be natural-based, synthetic-based, sacrificial, biocompatible thermoplastics, or tissue/in vitro model specific formulations.

2.4 Potential Improvements to the Current Bioprinters—Future Design Considerations

Although there has been many new bioprinting technologies and innovations in the field, there remain key challenges which need to be overcome in order to realize the tissue manufacturing process. Ozbulat et al. have articulated many of such limitations with different bioprinting technologies. This section discusses additional features which may be incorporated into a bioprinter to make the printing process faster, flexible, efficient and more predictable.

- Flexibility in the direction of printing: Vertical printing is not always desirable. For example, complex neural tissues demand more skewed architecture. The proposed solution would be engineering bioprinters with multiple robotic arms. In this regard, it is worthwhile to note that BioAssemblyBot (ASLS), which is a 6-axis robot arm that can carry out not only additive printing but also contour printing. However, significant progress is still needed in terms of the ability to directly fabricate using biologically relevant hydrogels, with tighter control of the microstructure and anisotropy in 3D. A
newly lab-developed 3D bioprinting technique called freeform reversible embedding of suspended hydrogels (FRESH) can ensure printing of hydrogels in complex 3D structures.

- Incorporation of co-axial extrusion system: Co-axial extrusion system will facilitate simultaneous chemical crosslinking while 3D printing.
- Scaffold-free tissues: In recent years, the notion of scaffold-free tissue engineering has gained traction, utilizing the self-organization and self-assembly based approaches. Spheroid printing technologies (for example, Kenzan) are expected to be the future of tissue engineering. Kenzan method does not use any gel (ECM) however in case of some tissues, external biochemical and mechanical cues may be crucial for their development and maturation.
- Tissue maturation: Cell-material constructs need optimal culturing conditions in terms of the physio-chemical environment in order to become functional tissue. Currently, none of the bioprinters has any bioreactors associated with them which can support the printed tissue construct for further maturation. For successful tissue maturation and large-scale implantations, this needs to be given serious thought.
- Integration of inline sensors/modules: Sensors could be integrated with the bioprinters for enhanced data gathering and analysis: During the actual printing process, it would be of real value to gauge real-time measurements of parameters such as temperature of printing chamber, humidity in printing chamber, pressure in extruder, speed and fidelity of the printing process etc. Since printing process usually takes hours, dynamic readouts using sensors can ensure remote observation and control too.
- In situ/in vivo bioprinting modules: An in vitro bioprinting approach may pose many inherent limitations regarding its clinical applicability in some cases like (a) ethical issues (b) fixation of fragile living constructs inside body (c) mismatch between the shape of the prefabricated construct and the actual defect and (d) maturation of the construct in a bioreactor before implantation. A possible solution is in vivo bioprinting (used interchangeably with in situ bioprinting), which will involve the de novo tissues/organs to be directly printed and positioned at the wound/defect site. This method would be highly effective in the treatment of tissues/organs that can be easily arrested and immobilized during bioprinting, e.g., the musculoskeletal system, craniofacial skeletal defects etc. For this to come true, portable bioprinters need to be designed and tested. There have already been some efforts in this direction, including the development of a hand-held bioprinting device called BioPen and the concept of Robotic arm based printer.

3. Conclusion

The advent of commercial bioprinting has revolutionized the field of tissue engineering. Within a short span we have at least 26 companies who are working in bioprinting business and out of which at least 23 are selling their bioprinters in the market. The industry now has a worldwide reach with companies based in almost all the major markets. Some of the companies also offer their bioprinters as services and others offer avenues for collaborations/partnerships. Hence a researcher has a multitude of choices, either to purchase bioprinting system (based on his/her applications requirements and budgetary constraints) or to enter into collaborations with the company of choice or to contract out the printing process. The fact that a simple and reasonably reliable extrusion-based bioprinting system is available for under five thousand US dollars makes it an attractive option for most of the small academic labs or individual researchers.

While this is a young industry with most ground-breaking developments happening only in the last 10 years, there has been tremendous growth. Extrusion bioprinting has emerged as the most popular choice owing to its simplicity and relatively lesser costs as compared to other modes. There is much potential in SLA bioprinting for manufacturing of tissues, but no commercial bioprinter can do it yet; although many of the printers do have UV light-source for curing of hydrogels. Currently, many labs around the world are trying to develop non-toxic cocktails of photoinitiators and novel biomaterials/composites which could be cured around visible light wavelengths for SLA bioprinting. The cost barrier for laser-assisted bioprinting is very high, and hence so far only one company is offering it. The development of first 6-axis robotic bioprinter, as well as the desktop polar coordinate-based system, has only demonstrated how much flexibility and...
sophistication are still possible. The incorporation of IOT elements and possibly AI (artificial intelligence) in future could only make the systems smarter. Integration of a bioreactor with a bioprinter would be key for natural maturation of printed tissue constructs.

The most important component in the bioprinting apart from the bioprinter is the bioink itself which supports the cell growth, differentiation and proliferation leading to maturation of the printed tissue. Several bioprinter companies have expanded their portfolio by selling bioinks; some of these inks are well-known, well-researched biomaterials while others are proprietary materials/formulations. This has enabled the companies to offer a complete solution to potential clients. Another crucial element for the optimum function of bioprinting hardware is its harmony with the respective controlling software. So far some of the low-cost bioprinters have been using open source software (e.g. Repetier-Host), and this could have reliability issues. Going forward, the printer having the most dedicated hardware-software integration would have the edge over the rest. With so many companies already jostling for space in this bioprinting business, coming decade is bound to see more intense competitions with the launch of upgraded and smarter bioprinters as well as the development of more biomimetic bioinks. Whether these commercial bioprinters get upgraded into high-throughput tissue manufacturing systems, only time will tell. Whether there will be a clear winner who is going to take the lion share of the bioprinting device market is to be seen. The regulatory aspect also needs to keep up with the pace of the device and material development in this field.

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DC and MWN conceptualized the manuscript and DC, MWN and SA wrote the manuscript. DC and SA made the figures and tables. MWN edited the manuscript.

Conflict of Interest
All authors reported no conflict of interest.

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