Original Article

Apical Extrusion of Irrigants in Immature Permanent Teeth by Using EndoVac and Needle Irrigation: An In Vitro Study

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Abstract

Objective: Immature teeth have a large apical opening and thin divergent or parallel dentinal walls; hence, with conventional needle irrigation there is a very high possibility of extrusion. This study was done to compare the apical extrusion of NaOCl in an immature root delivered using EndoVac and needle irrigation.

Materials and Methods: Eighty freshly extracted maxillary central incisors were decoronated followed by access cavity preparation. Modified organotypic protocol was performed to create an open apex; then, the samples were divided into four groups (n=20): EndoVac Microcannula (group I), EndoVac Macro cannula (group II), NaviTip irrigation needle (group III) and Max-i-Probe Irrigating needle (group IV); 9.0 ml of 3% sodium hypochlorite was delivered slowly over a period of 60 seconds. Extruded irrigants were collected in a vial and analysed statistically.

Results: Group I, group III and group IV showed 100% extrusion (20/20) but group II showed only 40% extrusion (8/20). The difference in this respect between group II and other groups was statistically significant (P<0.001). With regards to the volume of extrusion, group II had only 0.23 ml of extruded irrigant. Group I extruded 7.53 ml of the irrigant. Group III and group IV extruded the entire volume of irrigant delivered.

Conclusion: EndoVac Macro cannula resulted in the least extrusion of irrigant in immature teeth when compared to EndoVac Microcannula and conventional needle irrigation.

Key Words: Open apex; Extrusion; Irrigation

INTRODUCTION

Endodontic management of necrotic immature permanent teeth is often difficult, as such teeth lack an apical stop and are also weak and fragile [1,2]. Traditionally, the treatment of choice in these teeth has been apexification using calcium hydroxide or mineral trioxide aggregate (MTA) [3]. Apexification procedures do not result in continued root development.
A barrier is induced or created in the apical region against which obturation is performed; but these roots still remain short and are susceptible to fracture [4]. Hence, currently there is a shift in focus towards a more biological treatment called revascularization that allows for continued root development. Numerous case reports and case series on successful revascularization of necrotic immature teeth have been recently reported [5-8].

Three critical components identified to contribute to the successful outcome of this procedure are the presence of stem cells, signalling molecules (growth factors) and a 3-dimensional physical scaffold [9]. Revascularization procedures involve disinfecting the root canal by using sodium hypochlorite (NaOCl), and placement of an intracanal medicament. After 2-3 weeks, an intentional over instrumentation is done to induce bleeding into the root canal. Once a clot is formed inside the root canal, a cervical barrier with mineral trioxide aggregate is placed [4]. Root canal irrigants that are used for disinfection in immature teeth should not cause any damage to the stem cells. Trevino et al. [10] stated that NaOCl used for disinfecting the immature root canals should be used very carefully, as any extrusion of this irrigant can result in damage to these stem cells. Immature teeth have a large apical opening and thin divergent or parallel dentinal walls and hence with conventional needle irrigation there is a very high possibility of extrusion.

The EndoVac system (Discus Dental, Culver City, CA, USA) was introduced in 2007 and was designed to safely deliver irrigants to the apical terminus of the root canals. The device consisted of a delivery/evacuation tip that is attached to a syringe containing the irrigant and high speed suction of the dental chair. Using a combination of macro or micro cannulae attached to the suction device, the irrigant is introduced into the pulp chamber and is then pulled by negative pressure down the canal into the tip of the cannula and then removed through a coronally positioned suction hose [11, 12].

Apical negative pressure system has proven to be the safest when compared to other irrigation systems, in terms of extrusion in teeth with mature apices [11]. Fukumoto et al. evaluated the intracanal aspiration technique for irrigation and found it to be superior for smear layer removal, when compared with conventional irrigation. They also reported that this intracanal aspiration technique resulted in limited extrusion of irrigants beyond the apical foramen in canine teeth where the apex of the root was resected [13]. To date, there are no studies done on extrusion of irrigants in immature teeth using EndoVac. Hence, the aim of this study was to compare the apical extrusion of NaOCl in immature roots delivered using EndoVac and needle irrigation. In addition, this study quantified the volume of irrigant extruded in immature permanent teeth.

**MATERIALS AND METHODS**

Eighty freshly extracted adult permanent maxillary central incisors were used. The study design was analyzed and approved by the Institutional Review Board of Meenakshi Animal Dental College, Meenakshi University, Tamil Nadu, India. Collection, storage, sterilization, and handling of extracted teeth followed Occupational Safety and Health Administration guidelines and regulations. Each tooth was radiographed to confirm the presence of a single canal and all the teeth were decoronated and the root length was standardized to 17 mm. Endodontic access was achieved and the canals were negotiated with an ISO size #15 stainless steel K file (MANI Inc, Toshigi-Ken, Japan) until the tip just appeared through the apical foramen.

**Preparation of Modified Root Canal Organotypic Model**

The teeth were then instrumented under constant sterile saline irrigation using peeso reamer (MANI Inc, Toshigi-Ken, Japan) from
size 1-4 to create an apical opening of 1.3 mm in diameter and simulate an open/immature apex. Trevino et al. [10] suggested the organotypical model for creating immature apex wherein LSX instrument to the size of 130 was used. Since a peeso reamer size 4 corresponds to size 130, it was used to create an immature apex. The samples were then divided randomly into four groups of 20 teeth each based on the irrigant delivery system.

The recommended protocol for using apical negative pressure irrigation system includes 2 main phases namely, macroirrigation and microirrigation [12]. Since this study aimed to evaluate the apical extrusion of irrigants in open apices, the manufacturer’s recommended protocol was modified. The protocol suggested by Nester Cohenca et al. was followed in this study [14].

**Group I: (EndoVac Microcannula)**
Irrigation with EndoVac (Discus Dental, Culver City, CA, USA) system was started by placing the microcannula at 16 mm. Then, 9.0 ml of 3% sodium hypochlorite was delivered slowly over a period of 60 seconds through the master delivery/evacuation tip that was placed above the access opening.

**Group II: (EndoVac Macrocannula)**
Irrigation with EndoVac (Discus Dental, Culver City, CA, USA) system was started by placing macrocannula at 16 mm. Then, 9.0 ml of 3% sodium hypochlorite was delivered slowly over a period of 60 seconds through the master delivery/evacuation tip that was placed above the access opening.

**Group III: (Conventional NaviTip irrigating needle)**
Irrigation with NaviTip (Ultradent products Inc. India) 29 gauge needle was done by placing the needle 2 mm short of the working length.
Then, 9.0 ml of 3% sodium hypochlorite was delivered slowly over a period of 60 seconds.

**Group IV: (Conventional Max-i-Probe Irrigating needle)**
Irrigation with Max-i-Probe (Dentsply India Pvt. Ltd. New Delhi, India) was done by placing the needle 2 mm short of the working length. Then, 9.0 ml of 3% sodium hypochlorite was delivered slowly over a period of 60 seconds.

**Experimental model to collect the extruded irrigants**
Experimental model used in this study was based on the Myers and Montegomery model [15]. A vial with a rubber stopper was taken and a hole was created with a heated instrument in the center of the rubber stopper. The tooth was then inserted under pressure into the rubber stopper which was fixed to the cementoenamel junction using cyanoacrylate glue. Another glass vial (smaller in size than the previous vial) that fits tightly into the suspended apical portion of the root was used as the collecting container.

The whole assembly of rubber stopper with the collecting container was inserted into the main vial. To equalize the pressure, a 23-gauge needle was inserted into the rubber stopper (Figure 1). The irrigation protocol was performed in each group and the irrigant extruded peripherically was collected. The volume of extruded irrigant was measured by using a calibrated collection vial.

The root canal diameter was very large when compared to the needle diameter (this may be the reason why the entire irrigant fluid escaped without wetting the canal).

**Statistical analysis**
Presence or absence of extrusion of irrigant in each group was determined. In addition to this, the volume of irrigant extruded in each group was calculated. For the extrusion of irrigants, Fisher’s exact test was used to compare the proportions. The mean values in groups I and II were statistically analyzed using independent sample t-test.
In the present study, the level of significance was set at 0.05. The statistical analysis was conducted using SPSS 17 software (SPSS Inc, Chicago, Ill., USA).

RESULTS
All the samples in group I, group III and group IV showed extrusion (100%) while only 8 samples in group II showed extrusion (40%). There was a statistically significant difference (P< 0.001) between group II and other groups (Table 1).

[Table 1. The intergroup comparison for the presence or absence of extrusion of irrigants]

| Extrusion | Group I | Group II | Group III | Group IV | Total | χ²-Value | P-Value |
|-----------|---------|----------|-----------|----------|-------|----------|---------|
| Yes       | 20 (100.0) | 8 (40.0) | 20 (100.0) | 20 (100.0) | 68 (85.0) | 42.35 | <0.001 |
| No        | 0 (0.0) | 12 (60.0) | 0 (0.0) | 0 (0.0) | 12 (15.0) | | |
| Total     | 20 (100.0) | 20 (100.0) | 20 (100.0) | 20 (100.0) | 80 (100.0) | | |

DISCUSSION
Various stem cells that may play a role in revascularization are dental pulp stem cells (DSPSCs), stem cells from PDL or bone and stem cells from the apical papilla (SCAP) [16, 17]. Of all these stem cells, SCAP are thought to play a major role as they are in close proximity to the apical root canal [18]. It is speculated that during over instrumentation these SCAP get seeded into the root canal. These SCAP allow revascularization to occur in the presence of a blood clot which acts as a scaffold and also provides growth factors [19, 20]. Disinfection of the root canal in immature teeth during the revascularization procedure is mainly carried out by the use of irrigants and by placement of intracanal medicaments. NaOCl and triple antibiotic paste are the most commonly used irrigants and medicaments by most authors [21].

Mechanical instrumentation is usually avoided in these teeth, as it might further weaken the thin dentinal walls and also result in the formation of smear layer [4].
The SCAP are in close proximity to the root tip [18]. Any extrusion of NaOCl during irrigation is likely to damage these cells; which are vital for revascularization [10].

Nestar Cohenca et al. reported that the EndoVac macrocannula was efficient in disinfecting immature teeth in dogs. They further stated that it may be safe to use EndoVac in immature teeth to prevent extrusion [14]. To date, there are no studies done on extrusion of irrigants in immature teeth using EndoVac. Hence, this study was done to compare the apical extrusion of NaOCl while using Endo-Vac and needle irrigation systems. In addition, this study quantified the volume of irrigant extruded in immature permanent teeth.

The model used in this study was based on the Myers and Montegomery model used for testing extrusion in case of mature roots [15]. Trevino et al. suggested the organotypic model for creating an immature apex wherein LSX instrument to the size of 130 was used [10]. In this study, we followed a modified organotypic model protocol with peeso reamers (size 1–4), which was used to prepare a parallel-walled canal space with constant 1.3 mm diameter without destroying the structural integrity of the root. The results of the present study revealed that all four groups showed extrusion of irrigants. Extrusion was seen in all samples (100%) in groups I, III and IV; whereas, in group II (EndoVac macrocannula) extrusion was seen in only 8 out of 20 samples (40%). Group II had significantly less extrusion when compared to the other groups. Our results were similar to those of an earlier study by Ross Paton et al, who reported less extrusion of irrigants with EndoVac in teeth with mature apices, where extrusion was seen in 2 out of 24 samples (8.33%) [22].

In terms of volume of extrusion, group II (EndoVac macrocannula) extruded significantly lower volume of irrigant when compared to the other groups. Group I extruded less volume of irrigants when compared to groups III and IV.

In groups III (closed ended) and IV (open ended), samples leaked the entire 9 ml of irrigants used, in spite of the needle being placed 2 mm short of the working length. Recently, Zoi Psimma et al. used point conductivity probe method to assess irrigant extrusion. They reported that the open ended needle caused more extrusion than closed ended needle and the extrusion of irrigants decreased as the needles were moved away from the working length [23]. In our study both open ended and closed ended needles caused extrusion of the entire volume of irrigants used. The mean volume of extrusion was 7.53 and 0.23 ml for microcannula and macrocannula groups, respectively. EndoVac macrocannula extruded only 0.23 of 9 ml delivered but it is still not clear whether this small amount will cause any significant damage to the stem cells and needs further investigation.

Both EndoVac microcannula and macrocannula are connected to the chair-side suction apparatus. Less extruded volume in macrocannula may be due to the large diameter (0.55 mm) of the opening of the macrocannula and the presence of this opening at the tip of the cannula in contrast to the microcannula which is of smaller diameter (0.10 mm) and has only side vents.

We used an open ended model to test the extrusion as the worst case scenario. However, in clinical situations some resistance to extrusion will be provided by the periapical tissue and bone.

Table 2. Using independent sample t-test to compare the mean volume (ml) of irrigants extruded between groups

| Groups     | N  | Mean | Std. Deviation | P-Value |
|------------|----|------|----------------|---------|
| Group I    | 20 | 7.53 | 0.472          | <0.001  |
| Group II   | 20 | 0.23 | 0.302          |         |
Recently, it has been established that even 1 microgram of triple antibiotic paste can be toxic to stem cells; hence, to avoid the extrusion of sodium hypochlorite it is better to use the EndoVac macrocannula for irrigation in an immature apex [24].

CONCLUSION

Within the limitations of this study, EndoVac macrocannula (apical negative pressure technique) resulted in the least extrusion of irrigant beyond the apex in immature teeth when compared to EndoVac microcannula and conventional needle irrigation.

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