An updated and illustrated dichotomous key for the Chagas disease vectors of *Triatoma brasiliensis* species complex and their epidemiologic importance

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Abstract

In the subfamily Triatominae, *Triatoma* exhibits the largest number of species, which are arranged in complexes. For the *T. brasiliensis* species complex, recent investigations based on results of geometric morphometrics combined with phylogeny have provided evidence that it should be composed of seven species: *T. brasiliensis*, *T. babiensis*, *T. juazeirensis*, *T. lenti*, *T. melanica*, *T. petrocchiae*, and *T. sherlocki*, in which *T. brasiliensis* is divided in two subspecies: *T. b. brasiliensis* and *T. b. macromelasoma*. A taxonomic key is presented to identify each taxon. Among members of this complex, *T. b. brasiliensis* is the most important in an epidemiologic context, due to its high prevalence in natural infection by *Trypanosoma*

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cruzi combined with a pronounced adaptation to domiciliary habitats. However, some members may be currently invading and colonizing homes, a process known as domiciliation. Therefore, the key presented here may be potentially useful for researchers as well as those involved in vector control measures.

**Keywords**
kissing bugs, morphological key, species group

**Introduction**

Chagas disease is an infection caused by the etiologic agent *Trypanosoma cruzi* (Chagas, 1909), a protozoan transmitted to humans and other mammals through the feces of infected hematophagous insects of subfamily Triatominae. Currently, this group consists of more than 150 species, of which more than 65 are found in Brazil (Costa and Lorenzo 2009; Galvão 2014; Oliveira and Alevi 2017; Oliveira et al. 2018). In addition to vector-borne transmission (the main route), several other modes are known, including congenital transmission, accidental intake of contaminated food (e.g., açai juice and sugarcane juice), organ transplantation, blood transfusion, breastfeeding, and laboratory accidents (WHO 2017). In the subfamily Triatominae, *Triatoma* exhibits the largest number of species, which are arranged in complexes, a kind of grouping that was initially based in varied rationalities, as ecology, geographic distribution, cytogenetics, among others (see Oliveira et al. 2017). Phylogenetic reconstruction based on multiple mitochondrial genes did not recover *Triatoma* as a monophyletic unit (Gardim et al. 2014; Justi et al. 2014). Despite this, it is a genus with several species of epidemiological importance, as *T. infestans* in several countries of South America, *T. dimidiata* in Central America and *T. brasiliensis*, *T. pseudomaculata* and *T. sordida* in Brazil (Coura 2015). Justi et al. (2014) presented a comprehensive phylogenetic study of Triatomini, proposing the species complexes should be composed by natural groups.

*Triatoma brasiliensis* species complex represents a monophyletic unit (Oliveira et al. 2017) and was first suggested as a group (Costa et al. 2013) based on data on morphology (Costa et al. 1997), biology (Costa and Marchon-Silva 1998), crossing experiments (Costa et al. 2003b), ecology (Costa et al. 1998; Almeida et al. 2009), isoenzymes (Costa et al. 1997), dispersal abilities (Almeida et al. 2012), and DNA variation analyses (Monteiro et al. 2004). Recent cytogenetic, morphological and molecular studies (Alevi et al. 2013, 2014, 2015, 2018; Mendonça et al. 2014, 2016; Oliveira et al. 2017) have shown that other species, in addition to those previously identified (Costa et al. 2013), should be included in the *T. brasiliensis* complex, which currently consists of *T. brasiiliensis* Neiva, 1911, *T. bahiensis* Sherlock & Serafim, 1967, *T. juazeirensis* Costa & Felix, 2007, *T. lenti* Sherlock & Serafim, 1967, *T. melanica* Neiva & Lent, 1941, *T. petrochiae* Pinto & Barreto,
1925, and *T. sherlocki* Papa et al., 2002, in which *T. brasiliensis* is divided in two subspecies *T. b. brasiliensis* Neiva, 1911 and *T. b. macromelasoma* Galvão, 1956.

To date, most of measures to combat the transmission of Chagas disease have been focused on vector control. Defining the taxonomic status and correctly identifying vectors of the *T. brasiliensis* complex is crucial to the success of surveillance actions, because each species exhibits its own epidemiological importance (Costa et al. 2003a, 2013). Because the *T. brasiliensis* species complex suffered rearrangements after phylogenetic and morphometric studies, an update of the dichotomous key by Costa et al. (2013) for members of the *T. brasiliensis* complex is proposed, according to the new consensus for defining this group.

**Materials and methods**

Most of insects studied here are deposited in the Entomological Collection of Oswaldo Cruz Institute (CEIOC), Oswaldo Cruz Foundation, Rio de Janeiro, Brazil. The type species were always checked if possible, as previously detailed (Costa et al. 2013). For the newly included members (*T. petrocchiae*, *T. lenti*, and *T. bahiensis*) material from the insectary of Araraquara was also used that was deposited in the Dr Jose Maria Soares Barata Triatominae Collection (CTJMSB) of the São Paulo State University Julio de Mesquita Filho (UNESP), School of Pharmaceutical Sciences (FCFAR), Araraquara, São Paulo, Brazil. Insects from this insectary were also used for taking the photographs. The terminology of Lent and Wygodzinsky (1979) is followed.

**Results**

According to Lent and Wygodzinsky (1979), the genus *Triatoma* comprises species in most cases with less than 30 mm. Other features include femora denticulate or not; ventral connexival plates distinct, although narrow in some cases. The sides of abdomen are rarely membranous, with membrane connecting dorsal and ventral connexival plates. The posterior process of pygophore is narrowly tapering apically. Members of *T. brasiliensis* species complex share a combination of characteristics: (i) they compose a natural group of (ii) inhabitants of rocky outcrops, (iii) distributed in semi-arid zones of Brazilian Northeast in the Caatinga Biome. The only exception is *T. melanica*, which can be also found in connections between Caatinga and Cerrado in the state of Minas Gerais. They are spread in many states, such as Bahia (BA), Ceará (CE), Maranhão (MA), Minas Gerais (MG), Paraíba (PB), Pernambuco (PE), Piauí (PI), and Rio Grande do Norte (RN).

A pictorial dichotomous key for the *T. brasiliensis* species complex was built up as follows:
Brachypterous specimens (short wings for both genders), hemelytra not extending beyond the posterior margin of urotergite VI; legs unusually long; overall color dark brown to black, connexiva and femora with reddish orange markings ........................................................................................................... *T. sherlocki* (BA)

Macropterous specimens, hemelytra reaching or almost reaching urotergite VII........................................................................................................... 2

Short first antennal segment, not close to reaching apex of clypeus .......... ................................................................. *T. petrocchiae* (BA, CE, PB, PE, RN)

First antennal segment reaching or almost reaching the level of apex of clypeus ............................................................................................................... 3
Updated key for *Triatoma brasiliensis* complex

3 Pronotum with 1+1 pale colored areas or stripes ........................................... 4

– Pronotum with entirely dark anterior lobe .................................................. 6

4 Pronotum with 1+1 narrow brownish-yellow stripes; membrane of hemelytra with lumen of cells partially darkened............... *T. b. macromelasoma* (PE)

– Pronotum with 1+1 broad, elongated brownish yellow areas; membrane of hemelytra with lumen of cells entirely darkened or not ....................... 5

5 Pronotum with 1+1 brownish yellow areas extending from the posterior portion of anterior lobe to posterior lobe; femora with broad brownish yellow rings; membrane of hemelytra with lumen of cells not darkened; males with fossula spongiosa on fore tibia only............... *T. b. brasiliensis* (CE, MA, PB, PI, RN)

– Pronotum with 1+1 brownish yellow areas only on posterior lobe; femora with narrow brownish yellow rings; membrane of hemelytra with lumen of
cells entirely darker; males with fossula spongiosa on fore tibiae..............
..............................................................................................................T. melanica (BA, MG)

6 Pronotum with black anterior lobe and wrinkled posterior lobe; rarely with few inconspicuous brownish yellow marks; light yellow corium with dark areas of variable extent; dark legs with light colored areas on trochanter........
..............................................................................................................T. juazeirensis (BA)

– Pronotum entirely black, non-granular, with anterolateral angles short and apically rounded; corium and clavus dark brown to black, and dark brown membrane; legs uniformly black.................................................................7

7 Scutellum with posterior portion of central depression pointed; first abdominal segment without prominences; anterior region of prothorax (near the stridulatory sulcus) with a depression in ventral view; posterior region of stridulatory sulcus with rounded and well defined edges; mesothorax smooth and rounded .................................................................T. lenti (BA)
– Scutellum with posterior portion of central depression rounded; first abdominal segment with two lateral prominences; no depression on anterior region of prothorax; mesothorax with a central longitudinal projection, rectangular in shape .................................................................T. bahiensis (BA)
Discussion

The epidemiological profile of Chagas disease has changed, both in endemic areas and in other regions due to human migration, resulting in dissemination to countries where there is no vector transmission (Coura 2013; Dias et al. 2014). In Brazil, despite the control of *T. infestans*, the main Chagas disease vector in the past (Dias et al. 2002; Moncayo and Silveira 2017) each geographic region presents distinct challenges due to different ecological profiles of the distinct vectors and also due to dramatic environmental modifications. For instance, oral transmissions have been observed in the north region of Brazil (Coura 2013). In the south of Brazil, the persistent *T. rubrovaria* requires intensive monitoring actions (Almeida et al. 2002) and in the northeast region, species of the *T. brasiliensis* complex have been showing rapid changes in their behavior and ecology due to environmental anthropization (Costa et al. 2014). Therefore, in endemic areas, monitoring the synanthropic behavior of Chagas disease vectors is a challenge (Costa 1999; Costa and Lorenzo 2009). This can be illustrated by the case of *T. sherlocki*: a species described as sylvatic was later found invading and colonizing domiciles in a quarry mining community in a remote area of Gentio do Ouro, Bahia state (Almeida et al. 2009). Therefore, a comprehensive taxonomic key is crucial to be used by researchers and by those involved in vector control (Lent and Wygodzinsky 1979).

Members of the *T. brasiliensis* complex have been found in 12 Brazilian states and show mainly allopatric and parapatric distribution patterns, (Costa et al. 2003a, 2014; Gurgel-Gonçalves et al. 2012; Mendonca et al. 2016). However, some species of this complex are known to be occasionally found in sympathy, as *T. b. brasiliensis*, *T. b. macromelasoma*, and *T. juazeirensis* (Costa et al. 2014, 2016) which are all sympatric with the newly included *T. petrocchiae* (Oliveira et al. 2017), which renders geography alone as an imperfect tool for confidently identifying species. Other species may also be found later to be sympatric with each other. For instance, a hybrid zone between *T. b. brasiliensis* and *T. juazeirensis* was found (Costa et al. 2009, 2016), highlighting the utility of this key in detecting intermediate forms between them.

Studies on members of the complex have demonstrated that *T. b. brasiliensis* is the most important species in epidemiological terms. This species exhibits high intra-domestic infestation and infection rates (Costa et al. 2003a), which led Lilioso et al. (2017) to attribute a possible role to this species in a recent Chagas disease outbreak in Rio Grande do Norte State (Vargas et al. 2018). Additionally, via molecular markers, the existence of perennial and uncontrollable foci has been demonstrated in the sylvatic areas of populations with high *T. cruzi* prevalence (Almeida et al. 2008, 2016).

We recommend disseminating a version of this document in Portuguese to those involved in vector control measures. However, despite the contribution presented here, we still face some taxonomic challenges regarding this complex. There is no available key to differentiate immature stages for all members of this complex, which may complicate the correct identification of these forms. As mentioned above (Oliveira et al. 2017), some members are sympatric (e.g., *T. petrocchiae* and *T. brasiliensis*), and if immature forms of *T. petrocchiae* are found in domiciles, it may be operationally recorded as *T. brasiliensis* during regular vector inspections, because this last species is
the most frequently found in domiciles wherever it occurs. Therefore, a comprehensive taxonomic key is a crucial tool for use by researchers and by those involved in vector control, which should include also immature forms.

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**References**

Alevi KCC, Mendonça PP, Pereira NP, Guerra AL, Facina CH, da Rosa JA, de Azeredo Oliveira MTV (2013) Distribution of constitutive heterochromatin in two species of triatomines: *Triatoma lenti* Sherlock and Serafim (1967) and *Triatoma sherlocki* Papa, Jurberg, Carcavallo, Cerqueira & Barata, (2002). Infection, Genetics and Evolution 13: 301–303. https://doi.org/10.1016/j.meegid.2012.11.011

Alevi KCC, de Oliveira J, Moreira FFF, Jurberg J, da Rosa JA, de Azeredo-Oliveira MTV (2015) Chromosomal characteristics and distribution of constitutive heterochromatin in the Matogrossensis and Rubrovaria subcomplexes. Infection, Genetics and Evolution 33: 158–162. https://doi.org/10.1016/j.meegid.2015.04.024

Alevi KCC, Pinotti H, De Araújo RF, De Azeredo Oliveira MTV, Da Rosa JA, Mendonça VJ (2018) Hybrid collapse confirms the specific status of *Triatoma bactiens* Sherlock & Serafim, 1967 (Hemiptera, Triatominae), an endemic species in Brazil. American Journal of Tropical Medicine and Hygiene 98: 475–477. https://doi.org/10.4269/ajtmh.17-0270

Alevi KCC, Rosa JA, Azeredo-Oliveira MTV (2014) Cytotaxonomy of the *Brasiliensis* subcomplex and the *Triatoma brasiliensis* complex (Hemiptera: Reduviidae: Triatominae). Zootaxa 3838: 583–589. https://doi.org/10.11646/zootaxa.3838.5.7

Almeida CE, Duarte R, Do Nascimento RG, Pacheco RS, Costa J (2002) *Triatoma rubrovaria* (Blanchard, 1843) (Hemiptera, Reduviidae, Triatominae) II: Trophic resources and ecological observations of five populations collected in the State of Rio Grande do Sul, Brazil. Memorias do Instituto Oswaldo Cruz 97: 1127–1131. https://doi.org/10.1590/S0074-02762002000800011

Almeida CE, Faucher L, Lavina M, Costa J, Harry M (2016) Molecular Individual-Based Approach on *Triatoma brasiizensis*: Inferences on Triatomine Foci, *Trypanosoma cruzi* Natural Infection Prevalence, Parasite Diversity and Feeding Sources. PLoS Neglected Tropical Diseases 10. https://doi.org/10.1371/journal.pntd.0004447

Almeida CE, Folly-Ramos E, Peterson AT, Lima-Neiva V, Gumiel M, Duarte R, Lima MM, Locks M, Beltrão M, Costa J (2009) Could the bug *Triatoma sherlocki* be vectoring Chagas disease in small mining communities in Bahia, Brazil? Medical and Veterinary Entomology 23: 410–417. https://doi.org/10.1111/j.1365-2915.2009.00822.x
Almeida CE, Oliveira HL, Correia N, Dornak LL, Gumiel M, Neiva VL, Harry M, Mendonça VJ, Costa J, Galvão C (2012) Dispersion capacity of Triatoma sherlocki, Triatoma juazeirensis and laboratory-bred hybrids. Acta Tropica 122: 71–79. https://doi.org/10.1016/j.actatropica.2011.12.001

Almeida CE, Pacheco RS, Haag K, Dupas S, Dotson EM, Costa J (2008) Inferring from the Cyt B gene the Triatoma brasiliensis Neiva, 1911 (Hemiptera: Reduviidae: Triatominae) genetic structure and domiciliary infestation in the State of Paraíba, Brazil. American Journal of Tropical Medicine and Hygiene 78: 791–802.

Chagas C (1909) Nova tripanozomiase humana: estudos sobre a morfolojia e o ciclo evolutivo do Schizotrypanum cruzi n. gen., n. sp., ajente etiologico de nova entidade morbida do homem. Memorias do Instituto Oswaldo Cruz 1: 159–218.

Costa J (1999) The synanthropic process of Chagas disease vectors in Brazil, with special attention to Triatoma brasiliensis Neiva, 1911 (Hemiptera, Reduviidae, Triatominae) population, genetical, ecological, and epidemiological aspects. Memorias do Instituto Oswaldo Cruz 94: 239–241. https://doi.org/10.1590/S0074-02761999000700038

Costa J, Almeida CE, Dotson EM, Lins A, Vinhaes M, Silveira AC, Beard CB (2003a) The Epidemiologic Importance of Triatoma brasiliensis as a Chagas Disease Vector in Brazil: A Revision of Domiciliary Captures during 1993–1999. Memorias do Instituto Oswaldo Cruz 98: 443–449. https://doi.org/10.1590/S0074-02762003000400002

Costa J, Almeida CE, Dujardin JP, Beard CB (2003b) Crossing Experiments Detect Genetic Incompatibility among Populations of Triatoma brasiliensis Neiva, 1911 (Heteroptera, Reduviidae, Triatominae). Memorias do Instituto Oswaldo Cruz 98: 637–639. https://doi.org/10.1590/S0074-02762003000500009

Costa J, Bargues MD, Neiva VL, Lawrence GG, Gumiel M, Oliveira G, Cabello P, Lima MM, Dotson E, Provance DW, Almeida CE, Mateo L, Mas-Coma S, Dujardin JP (2016) Phenotypic variability confirmed by nuclear ribosomal DNA suggests a possible natural hybrid zone of Triatoma brasiliensis species complex. Infection, Genetics and Evolution 37: 77–87. https://doi.org/10.1016/j.meegid.2015.10.025

Costa J, Barth OM, Marchon-Silva V, De Almeida CE, Freitas-Sibajev MGR, Panzera F (1997) Morphological Studies on the Triatoma brasiliensis Neiva, 1911 (Hemiptera, Reduviidae, Triatominae) Genital Structures and Eggs of Different Chromatic Forms. Memorias do Instituto Oswaldo Cruz 92: 493–498. https://doi.org/10.1590/S0074-02761997000400009

Costa J, Cordeiro Correia N, Neiva VL, Gonçalves TCM, Felix M (2013) Revalidation and redeciption of Triatoma brasiliensis macromelasoma Galvão, 1956 and an identification key for the Triatoma brasiliensis complex (Hemiptera: Reduviidae: Triatominae). Memorias do Instituto Oswaldo Cruz 108: 785–789. https://doi.org/10.1590/0074-0276108062013016

Costa J, Dornak LL, Almeida CE, Peterson AT (2014) Distributional potential of the Triatoma brasiliensis species complex at present and under scenarios of future climate conditions. Parasites and Vectors 7. https://doi.org/10.1186/1756-3305-7-238

Costa J, Freitas-Sibajev MGR, Marchon-Silva V, Pires MQ, Pacheco RS (1997) Isoenzymes Detect Variation in Populations of Triatoma brasiliensis (Hemiptera: Reduviidae: Triatominae). Memorias do Instituto Oswaldo Cruz 92: 459–464. https://doi.org/10.1590/S0074-02761997000400002
Costa J, Lorenzo M (2009) Biology, diversity and strategies for the monitoring and control of triatomines - Chagas disease vectors. Memorias do Instituto Oswaldo Cruz 104: 46–51. https://doi.org/10.1590/S0074-02762009009000008

Costa J, Marchon-Silva V (1998) Periodo de intermuda e resistência ao jejum de diferentes populações de *Triatoma brasiliensis* (Hemiptera: Reduviidae: Triatominae). Entomol Vectores 5: 23–34.

Costa J, Peterson AT, Dujardin JP (2009) Morphological evidence suggests homoploid hybridization as a possible mode of speciation in the Triatominae (Hemiptera, Heteroptera, Reduviidae). Infection, Genetics and Evolution 9: 263–270. https://doi.org/10.1016/j.meegid.2008.12.005

Costa J, Ribeiro De Almeida J, Britto C, Duarte R, Marchon-Silva V, Pacheco RDS (1998) Ecotopes, Natural Infection and Trophic Resources of *Triatoma brasiliensis* (Hemiptera, Reduviidae, Triatominae). Memorias do Instituto Oswaldo Cruz 93: 7–13. https://doi.org/10.1590/S0074-02761998000100002

Coura JR (2013) Chagas disease: Control, elimination and eradication. Is it possible? Memorias do Instituto Oswaldo Cruz 108: 962–967. https://doi.org/10.1590/0074-0276130565

Coura JR (2015) The main sceneries of Chagas disease transmission. The vectors, blood and oral transmissions - A comprehensive review. Memorias do Instituto Oswaldo Cruz 110: 277–282. https://doi.org/10.1590/0074-0276140362

Dias FBS, Quartier M, Diotaiuti L, Mejia G, Harry M, Lima ACL, Davidson R, Mertens F, Lucotte M, Romaña CA (2014) Ecology of *Rhodnius robustus* Larrousse, 1927 (Hemiptera, Reduviidae, Triatominae) in *Attalea* palm trees of the Tapajós River Region (Pará State, Brazilian Amazon). Parasites and Vectors 7. https://doi.org/10.1186/1756-3305-7-154

Dias J, Silveira A, Schofield C (2002) The Impact of Chagas Disease Control in Latin America - A Review. Mem Inst Oswaldo Cruz Rio de Janeiro 97: 603–612. https://doi.org/10.1590/S0074-027620000500002

Galvão C (2014) Vetores da doença de Chagas no Brasil. Sociedade Brasileira de Zoologia, 289 pp.

Gardim S, Almeida CE, Takiya DM, Oliveira J, Araújo RF, Cicarelli RMB, Da Rosa JA (2014) Multiple mitochondrial genes of some sylvatic Brazilian *Triatoma*: Non-monophyly of the *T. brasiliensis* subcomplex and the need for a generic revision in the Triatomini. Infection, Genetics and Evolution 23: 74–79. https://doi.org/10.1016/j.meegid.2014.01.024

Gurgel-Gonçalves R, Galvão C, Costa J, Peterson AT (2012) Geographic distribution of Chagas disease vectors in Brazil based on ecological niche modeling. Journal of Tropical Medicine. https://doi.org/10.1155/2012/705326

Justi SA, Russo CAM, Mallet JR dos S, Obara MT, Galvão C (2014) Molecular phylogeny of Triatomini (Hemiptera: Reduviidae: Triatominae). Parasites & vectors 7: 149. https://doi.org/10.1186/1756-3305-7-149

Lent H, Wygodzinsky P (1979) Revision of the Triatominae (Hemiptera, Reduviidae), and their significance as vectors of Chagas disease. Bulletin of the American museum of natural history 163: 123–520.

Lilioso M, Folly-Ramos E, Rocha FL, Rabinovich J, Capdevielle-Dulac C, Harry M, Marcet PL, Costa J, Almeida CE (2017) High *Triatoma brasiliensis* densities and *Trypanosoma cruzi* prevalence in domestic and peridomestic habitats in the State of Rio Grande do Norte,
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Brazil: The source for Chagas disease outbreaks? American Journal of Tropical Medicine and Hygiene 96: 1456–1459. https://doi.org/10.4269/ajtmh.16-0823

Mendonça VJ, Alevi KCC, Medeiros LM de O, Nascimento JD, de Azeredo-Oliveira MTV, da Rosa JA (2014) Cytogenetic and morphologic approaches of hybrids from experimental crosses between Triatoma lenti Sherlock & Serafim, 1967 and T. sherlocki Papa et al., 2002 (Hemiptera: Reduviidae). Infection, Genetics and Evolution 26: 123–131. https://doi.org/10.1016/j.meegid.2014.05.015

Mendonca VJ, Alevi KCC, Pinotti H, Gurgel-Goncalves R, Pita S, Guerra AL, Panzera F, De Araujo RF, De Azeredo-Oliveira MTV, Da Rosa JA (2016) Revalidation of Triatoma bahiensis Sherlock & Serafim, 1967 (Hemiptera: Reduviidae) and phylogeny of the T. brasiliensis species complex. Zootaxa 4107: 239–254. https://doi.org/10.11646/zootaxa.4107.2.6

Moncayo A, Silveira AC (2017) Current epidemiological trends of Chagas disease in Latin America and future challenges: Epidemiology, surveillance, and health policies. In: American Trypanosomiasis Chagas Disease: One Hundred Years of Research, 2nd ed., 59–88. https://doi.org/10.1016/B978-0-12-801029-7.00004-6

Monteiro FA, Donnelly MJ, Beard CB, Costa J (2004) Nested clade and phylogeographic analyses of the Chagas disease vector Triatoma brasiliensis in Northeast Brazil. Molecular Phylogenetics and Evolution 32: 46–56. https://doi.org/10.1016/j.ympev.2003.12.011

Oliveira J, Alevi KCC (2017) Taxonomic status of Panstrongylus herreri Wygodzinsky, 1948 and the number of Chagas disease vectors. Revista da Sociedade Brasileira de Medicina Tropical 50: 434–435. https://doi.org/10.1590/0037-8682-0125-2017

Oliveira J, Ayala JM, Justi SA, da Rosa JA, Galvão C (2018) Description of a new species of Nesotriatoma Usinger, 1944 from Cuba and revalidation of synonymy between Nesotriatoma bruneri (Usinger, 1944) and N. flavida (Neiva, 1911) (Hemiptera, Reduviidae, Triatominae). Journal of Vector Ecology 43: 148–157. https://doi.org/10.1111/jvec.12294

Oliveira J, Marcet PL, Takiya DM, Mendonça VJ, Belintani T, Bargues MD, Mateo L, Chagas V, Folly-Ramos E, Cordeiro-Estrela P, Gurgel-Gonçalves R, Costa J, da Rosa JA, Almeida CE (2017) Combined phylogenetic and morphometric information to delimit and unify the Triatoma brasiliensis species complex and the Brasiliensis subcomplex. Acta Tropica 170: 140–148. https://doi.org/10.1016/j.actatropica.2017.02.020

Vargas A, Malta JMAS, Costa VM da, Claudio LDG, Alves RV, Cordeiro G da S, Aguiar LMA, Percio J (2018) Investigação de surto de doença de Chagas aguda na região extra-amazônica, Rio Grande do Norte, Brazil, 2016. Cadernos de saúde publica 34: e00006517. https://doi.org/10.1590/0102-311X00006517

WHO (2017) Integrating neglected tropical diseases. World Health Organization: Fact Sheet March 2017.