Acute and Repeated 28-Day Oral Dose Toxicity Studies of *Thymus vulgaris* L. Essential Oil in Rats

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Abstract

*Thymus vulgaris* L. (thyme), a member of the *Lamiaceae* family, is a grassy perennial plant that grows in many parts of the world. It is aromatic and rich in essential oil. Native to southern Europe, it is widely used in traditional medicine across the globe. In Peru, an infusion of this plant is used to improve cerebral and circulatory functions and to ward against intestinal parasites, whereas the essential oil is used as an antiseptic (1).

There are several studies that have reported the pharmacological activity of the essential oil of *Thymus vulgaris*, such as: antibacterial activity against *Staphylococcus aureus*, *Escherichia coli*, *Bacillus cereus*, *Proteus vulgaris*, *Listeria monocytogenes*, *Propionibacterium acnes*, *Clostridium perfringens*, *Enterococcus faecalis*, *Salmonella typhimurium*, and *Yersinia enterolitica* (2-7); antifungal activity against...
Candida albicans, Trichophyton mentagrophytes, Fusarium sp., Aspergillus flavus, and Aspergillus niger (8-11); antiparasitic effects against protoscoleces and cysts of Echinococcus granulosus (12) and Trypanosoma cruzi (13); cytotoxic activity against cells of human prostate carcinoma, human lung carcinoma, and human breast cancer (4); hepatoprotective effect against hepatic damage induced by acetaminophen in mice (14).

Recently, a study evaluated the potential of thyme essential oil and its individual components to reduce cholinergic deficits related to neurodegenerative and psychiatric disorders. The study revealed that thyme oil increases neurotransmission by modulating acetylcholine (Ach) synaptic levels and activity of the nicotinic acetylcholine receptor, orchestrated by the positive regulation of the cho-1, unc-17, and unc-50 genes. Likewise, the active components para-cymene (1-methyl-4-propan-2-ylbenzene) and the combination of thymol and gamma-terpinene exhibited similar effects, with possible improvement in cholinergic dysfunction (15).

For several medicinal plants, information regarding their toxicity is limited (16,17). Despite the fact that important pharmacological activities of thyme essential oil have been reported, no oral toxicity study of thyme essential oil has been published. However, oral toxicity studies of thymol, the main constituent of the essential oil of Thymus vulgaris, have already been reported in another study (18). Therefore we set out to evaluate the acute and repeated oral toxicity of Thymus vulgaris essential oil for 28 days in a murine model.

MATERIALS AND METHODS

Plant material. Thymus vulgaris L. was collected in the city of Lima, Peru. A sample was taken to the Natural History Museum of the National University of San Marcos for taxonomic identification (No. 184-USM-2016).

The essential oil was obtained from fresh leaves using steam distillation in a Clevenger-type apparatus (19). The oil was separated and dehydrated with anhydrous Na$_2$SO$_4$, filtered, and stored in an amber glass bottle under refrigeration at a temperature of 4°C until used.

Experimental animals and housing. Albino Holtzman rats were obtained from the National Health Institute, Peru. They were kept in cages for one week prior to commencing the study to allow for acclimatization to laboratory conditions. The animals’ housing was maintained under controlled environmental conditions (12-hr light/dark cycle) and temperature (22 ± 3°C). They were fed ad libitum with commercial rat feed and drinking water.

All the animal experiments were performed in accordance with institutional protocols and the guideline for care and use of laboratory animals (20). All research protocols were submitted to, and approved by, the School of Medicine Research Ethics Committee, National University of San Marcos, Peru (Act No. 158).

Acute oral toxicity studies. Acute toxicity at a single dose was evaluated according to the Organization for Economic Co-operation and Development (OECD) guideline, method 423. This method is a step-by-step procedure that begins with the maximum dose of 2,000 mg/kg bw and, then depending on the mortality and/or morbidity of the animals, is lowered to 300, 50, or 5 mg/kg bw doses to allow a judgment of the test substance’s acute toxicity (21). All the animals were fasted overnight before commencing the experiment. The experiment was conducted on six female rats (160 ± 10 g bw), randomly assigned to two groups (n = 3), which each received a single dose of essential oil (300 or 2,000 mg/kg bw). The animals were observed individually during the first 30 min, with special attention during the first four hours, then daily throughout the 14 days of the experiment. Signs and symptoms of toxicity were recorded. Observations focused on the determination of death and time of occurrence, signs and symptoms of toxicity from the beginning and through the duration of the experiment, including changes in skin, fur, mucous membranes and eyes, respiratory and circulatory systems, central nervous and autonomic systems, somatomotor activity and behavior. Special attention was paid to the potential occurrence of tremors, seizures, salivation, diarrhea, lethargy, drowsiness, and coma. To conclude the experiment, the animals were sacrificed by inhalation of ethyl ether. This was followed by necropsy and macroscopic pathological study of the stomach, liver, spleen, lungs, kidneys, esophagus, brain, and small intestine. Finally, the organs were studied by microscopic examination.

Repeated dose 28-day oral toxicity study. The subchronic oral toxicity study was performed in accordance with the instructions by OECD test guideline-407 (22) with slight modifications, using 20 female rats (160 ± 10 g bw) and 20 male rats (170 ± 10 g bw). The animals were randomly assigned to four groups (n = 10: five female and five male). Each rat in Group I (control group) received only the vehicle. Groups II, III, and IV received the essential oil of Thymus vulgaris in repeated oral doses of 100, 250, and 500 mg/kg bw, respectively, for 28 days. The animals were dosed at approximately the same time each day. The animals were monitored for signs of toxicity and mortality twice a day (a.m. and p.m.) throughout the experimental period of 28 days. The terminal weight of each animal was recorded weekly throughout the course of the experiment. On day 29, blood samples were collected from the rats via intracardiac puncture, under anesthesia with ethyl ether, and assessed for hematological and biochemical parameters.
The animals were sacrificed by decapitation under anesthesia with intraperitoneal sodium pentobarbital (40 mg/kg). Organs were fixed in 10% formalin for histopathological examination.

**Body and organ weight measurement.** After treatments, the body weight of each rat in the experiment was measured and recorded. The heart, lungs, liver, spleen, stomach, kidney, and testes or uterus were excised immediately after sacrifice, trimmed of fat and connective tissue, blotted with filter paper and weighed. The relative organ weights [ratio of organ weight and the animal’s body weight (at the end of experiment) × 100] were calculated.

**Biochemical parameters.** Biochemical parameters were evaluated using a Semi-Automatic Biochemical Analyzer, model EMP-168 (Ivdiaignostik, Emperor Medical, Shenzhen, China), according to the manufacturer’s specifications. The levels of aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase, total protein, bilirubin, cholesterol, triglycerides, high-density lipoprotein (HDL), low-density lipoprotein (LDL), total albumin, glucose, urea, and creatinine were determined.

**Hematological assay.** Hematological assays were performed using an Automatic Hematology Analyzer KT-6400 (Genius, Med Equipment, Guangzhou, China). At the end of the experiment hematocrit, hemoglobin concentration, erythrocyte count, total and differential leukocyte counts, and platelet count were evaluated.

**Histopathological analysis.** All the animals were subjected to necropsy at the end of the oral toxicity study (or earlier in the case of death). The organs were preserved in 10% formalin and fixed for three days, dehydrated, embedded in paraffin, sectioned at 5 μm, and stained with hematoxylin/eosin. Slides of organs taken from all animals were examined microscopically and photographed with a light microscope (BX53, Olympus, Tokyo, Japan) at 100× and 400× magnification.

**Statistical analyses.** The data were expressed as mean ± SD of five animals in each group and were analyzed using one-way analysis of variance (ANOVA), followed by the Tukey’s test. The results were considered significant when \( p < 0.05 \). IBM’s SPSS version 19 was used for all statistical analyses.

**RESULTS**

**Acute oral toxicity.** With the single dose of *Thymus vulgaris* essential oil (2,000 mg/kg bw), the three rats in the group showed immediate signs of toxicity, including hyperactivity and burning nose. After 10 min, ataxia, hypersalivation, and respiratory distress were observed. After four hours, one rat had recovered, while the other two rats worsened and went into lethargy with hypothermia, and finally coma. Death occurred within 24 hr. After these results were obtained, a single oral dose of 300 mg/kg bw was administered to another group of rats. The three animals in the group all showed slight manifestations that completely disappeared within 3 hr.

Histopathological examination of organs showed alterations in the lungs, where polymorph nuclear infiltrates, hemosiderin macrophages, and interstitial space thickening were observed. In the kidneys, a polymorph nuclear infiltrate was observed in the pyelocaliceal area, and the liver showed a slight fatty infiltration. In the stomach, spleen, heart, and brain no alterations were observed.

**Repeated dose 28-day oral toxicity studies.** All rats survived the 28-day treatment period and showed no apparent signs of toxicity.

**Effect of essential oil on body and organ weight of rats.** There were no significant changes in body weight in female rats treated with the essential oil of *Thymus vulgaris* when compared to the control group. However, in...
the male rats, a significant change occurred in the fourth week of the 500 mg/kg bw dose group, with body weight decreasing from 275.00 ± 11.36 g (control group) to 242.67 ± 11.93 g ($p < 0.05$) (Fig. 1). The relative weight of the heart, lungs, liver, spleen, stomach, kidney, and testes of male rats, or uterus of female rats treated in the 28-day period did not show any significant changes when compared with the control group (Table 1).

### Effect of essential oil on biochemical parameters.

The biochemical parameters of both female and male rats showed no significant changes after the 28-day toxicity test when compared to the control group (Table 2).

### Effect of essential oil on hematological parameters.

The results of the hematological parameters of control rats and those treated daily for 28 days with the essential oil of *Thymus vulgaris* are shown in Table 3. These results show that there were no significant changes to any of the parameters in the treated groups when compared with the control group.

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**Table 1.** Effect of *Thymus vulgaris* essential oil on the relative weight of the organs in rats treated for 28 days.

| Organ     | Dose group (mg/kg/day) |
|-----------|------------------------|
|           | Control | 100 | 250 | 500 |
| Males     |         |     |     |     |
| Heart     | 0.35 ± 0.03 | 0.33 ± 0.07 | 0.35 ± 0.11 | 0.36 ± 0.03 |
| Lungs     | 0.72 ± 0.09 | 0.73 ± 0.14 | 0.81 ± 0.10 | 0.80 ± 0.06 |
| Liver     | 3.13 ± 0.16 | 3.14 ± 0.60 | 2.97 ± 0.40 | 2.95 ± 0.18 |
| Spleen    | 0.35 ± 0.04 | 0.34 ± 0.17 | 0.31 ± 0.05 | 0.35 ± 0.02 |
| Stomach   | 0.54 ± 0.08 | 0.55 ± 0.05 | 0.55 ± 0.21 | 0.57 ± 0.11 |
| Kidney    | 0.40 ± 0.05 | 0.43 ± 0.08 | 0.42 ± 0.07 | 0.44 ± 0.07 |
| Testis    | 1.02 ± 0.09 | 1.07 ± 0.14 | 1.02 ± 0.11 | 1.07 ± 0.19 |
| Females   |         |     |     |     |
| Heart     | 0.34 ± 0.14 | 0.35 ± 0.09 | 0.33 ± 0.07 | 0.35 ± 0.11 |
| Lungs     | 0.91 ± 0.13 | 0.90 ± 0.22 | 0.92 ± 0.23 | 0.89 ± 0.26 |
| Liver     | 2.93 ± 0.24 | 3.17 ± 0.57 | 3.28 ± 0.20 | 3.05 ± 0.22 |
| Spleen    | 0.41 ± 0.11 | 0.42 ± 0.27 | 0.39 ± 0.03 | 0.44 ± 0.05 |
| Stomach   | 0.49 ± 0.14 | 0.49 ± 0.14 | 0.53 ± 0.12 | 0.54 ± 0.26 |
| Kidney    | 0.35 ± 0.08 | 0.38 ± 0.17 | 0.40 ± 0.21 | 0.41 ± 0.19 |
| Uterus    | 0.49 ± 0.04 | 0.51 ± 0.12 | 0.48 ± 0.07 | 0.49 ± 0.18 |

Values are expressed as Mean ± SD. No significant difference was observed when compared with controls.

**Table 2.** Biochemical parameters of rats after treatment with repeated oral doses of *Thymus vulgaris* essential oil for 28 days.

| Parameter         | Dose group (mg/kg/day) |
|-------------------|------------------------|
|                   | Control | 100 | 250 | 500 |
| Males             |         |     |     |     |
| AST (IU/L)        | 145.00 ± 6.55 | 155.67 ± 7.02 | 159.67 ± 4.51 | 162.00 ± 8.89 |
| ALT (IU/L)        | 60.33 ± 12.86 | 73.00 ± 7.94 | 74.33 ± 9.50 | 75.67 ± 10.02 |
| Alkaline phosphatase (IU/L) | 173.67 ± 6.03 | 190.33 ± 8.74 | 193.67 ± 12.50 | 194.67 ± 9.87 |
| Total bilirubin (mg/dL) | 0.61 ± 0.12 | 0.65 ± 0.25 | 0.68 ± 0.08 | 0.67 ± 0.07 |
| Total protein (g/dL) | 7.17 ± 0.50 | 7.23 ± 0.60 | 7.37 ± 0.57 | 7.47 ± 0.32 |
| Albumin (g/dL)    | 3.63 ± 0.35 | 3.89 ± 0.09 | 3.97 ± 0.21 | 4.03 ± 0.15 |
| Cholesterol (mg/dL) | 68.40 ± 5.05 | 62.43 ± 8.16 | 64.00 ± 7.21 | 62.13 ± 10.43 |
| Triglycerides (mg/dL) | 95.27 ± 11.07 | 84.03 ± 6.99 | 81.47 ± 6.63 | 80.53 ± 2.84 |
| HDL (mg/dL)       | 35.10 ± 6.71 | 35.80 ± 6.01 | 35.07 ± 4.65 | 35.10 ± 3.15 |
| LDL (mg/dL)       | 14.25 ± 2.84 | 9.82 ± 2.02 | 12.64 ± 2.19 | 11.92 ± 2.77 |
| Glucose (mg/dL)   | 118.10 ± 7.41 | 118.67 ± 12.58 | 119.00 ± 6.56 | 123.33 ± 7.23 |
| Urea (mg/dL)      | 35.50 ± 2.29 | 34.83 ± 3.19 | 37.23 ± 3.95 | 38.17 ± 2.25 |
| Creatinine (mg/dL) | 0.75 ± 0.18 | 0.73 ± 0.06 | 0.84 ± 0.07 | 0.83 ± 0.05 |
| Females           |         |     |     |     |
| AST (IU/L)        | 130.33 ± 5.51 | 131.00 ± 6.56 | 134.33 ± 6.03 | 136.67 ± 7.09 |
| ALT (IU/L)        | 58.67 ± 9.71 | 60.33 ± 8.39 | 66.00 ± 12.17 | 67.00 ± 8.19 |
| Alkaline phosphatase (IU/L) | 174.00 ± 9.64 | 194.67 ± 9.87 | 180.33 ± 12.50 | 192.33 ± 6.03 |
| Total bilirubin (mg/dL) | 0.62 ± 0.14 | 0.58 ± 0.13 | 0.63 ± 0.08 | 0.64 ± 0.09 |
| Total protein (g/dL) | 7.23 ± 0.67 | 7.43 ± 0.15 | 7.47 ± 0.21 | 7.50 ± 0.89 |
| Albumin (g/dL)    | 3.80 ± 0.20 | 3.95 ± 0.15 | 4.07 ± 0.21 | 4.17 ± 0.22 |
| Cholesterol (mg/dL) | 72.23 ± 10.28 | 72.50 ± 6.73 | 69.33 ± 6.66 | 70.10 ± 5.46 |
| Triglycerides (mg/dL) | 89.07 ± 6.21 | 82.52 ± 9.98 | 79.17 ± 8.89 | 77.33 ± 7.02 |
| HDL (mg/dL)       | 36.03 ± 4.65 | 36.47 ± 4.22 | 34.47 ± 4.08 | 35.83 ± 1.89 |
| LDL (mg/dL)       | 18.39 ± 5.24 | 19.53 ± 3.14 | 19.03 ± 3.95 | 18.80 ± 4.11 |
| Glucose (mg/dL)   | 120.07 ± 6.26 | 117.67 ± 8.02 | 122.47 ± 6.43 | 127.07 ± 13.11 |
| Urea (mg/dL)      | 37.93 ± 1.69 | 34.93 ± 4.78 | 33.40 ± 3.22 | 32.63 ± 4.22 |
| Creatinine (mg/dL) | 0.78 ± 0.06 | 0.77 ± 0.06 | 0.81 ± 0.07 | 0.74 ± 0.05 |

Values are expressed as Mean ± SD. No significant differences were observed when compared with controls.

AST, aspartate aminotransferase; ALT, alanine aminotransferase; ALP, alkaline phosphatase; HDL, high-density lipoprotein; LDL, low-density lipoprotein.
Table 3. Hematological parameters of rats after treatment with repeated oral doses of *Thymus vulgaris* essential oil for 28 days

| Parameter                  | Control | 100       | 250       | 500       |
|----------------------------|---------|-----------|-----------|-----------|
| RBC (× 10^6/µL)            | 7.25 ± 0.12 | 7.13 ± 0.28 | 7.21 ± 0.23 | 7.50 ± 0.29 |
| WBC (× 10^3/µL)            | 8.30 ± 0.95 | 8.20 ± 0.60 | 8.70 ± 0.95 | 8.60 ± 1.85 |
| Hemoglobin (g/dL)          | 14.87 ± 0.80 | 14.80 ± 0.95 | 15.17 ± 0.65 | 14.60 ± 0.87 |
| Hematocrit (%)             | 46.73 ± 2.05 | 46.43 ± 2.77 | 47.07 ± 2.53 | 46.33 ± 4.51 |
| Neutrophils (%)            | 18.00 ± 2.00 | 19.33 ± 3.51 | 23.67 ± 4.51 | 21.67 ± 3.06 |
| Eosinophils (%)            | 2.67 ± 0.58 | 3.00 ± 1.73 | 2.33 ± 1.53 | 3.00 ± 1.00 |
| Basophils (%)              | 0.67 ± 0.58 | 1.00 ± 1.00 | 1.00 ± 1.00 | 0.67 ± 0.58 |
| Monocytes (%)              | 3.33 ± 0.58 | 3.00 ± 1.00 | 2.67 ± 0.58 | 3.33 ± 1.15 |
| Lymphocytes (%)            | 75.33 ± 2.52 | 73.67 ± 4.51 | 70.33 ± 5.03 | 71.33 ± 4.04 |
| Platelets (× 10^3/µL)      | 700.67 ± 32.08 | 638.00 ± 72.38 | 674.00 ± 42.58 | 681.67 ± 25.66 |

Values are expressed as Mean ± SD in each sex group. Differences between the treated group and the control group were not significant.

RBC, Red Blood Cell; WBC, White Blood Cell.

Fig. 2. Photomicrographs of the sections from liver, lung, stomach, and esophagus of rats treated with vehicle (control) and *Thymus vulgaris* essential oil over 28 days. H-E staining, 40×.
Histological findings. Microscopic examination of the organs from rats sacrificed after the 28-day treatment period showed no alteration in the kidney and spleen. However, a mild inflammatory infiltrate was observed in the liver in 70% of the cases for all three dose levels; 100, 250, and 500 mg/kg bw. Lungs of rats given doses of 100 and 250 mg/kg bw of essential oil showed a moderate inflammatory infiltrate in 65% of cases; whereas with a dose of 500 mg/kg, the inflammatory infiltrate was severe, and in 45% of the cases hemorrhagic foci were also observed. The stomach showed a mild acute inflammatory infiltrate in one case (10%) at a dose of 250 mg/kg bw, and in two cases (20%) at 500 mg/kg bw (Fig. 2). In the esophagus, only one case (10%) of moderate subepithelial inflammatory infiltrate was observed at a dose of 500 mg/kg bw (Fig. 2).

DISCUSSION

The 2,000 mg/kg bw single oral dose caused the death of two rats (2/3) within 24 hr, preceded by the rapid onset of toxicity manifestations in the respiratory, central nervous, and autonomic systems. The intense burning of the nose observed in the animals could be due to the fact that the respiratory tract is innervated by primary sensory afferent nerves, which are activated by mechanical and chemical stimuli. The transient receptor potential cation channel, subfamily A, member 1 (TRPA1) channels are nonselective cation channels that are activated by a range of natural substances (23). When an odorous substance is sufficiently pungent, the trigeminal nerve, or vagal afferent C fibers, in the nose, mouth, and eyes are activated. This sensory system, called the common chemical sense (CCS), may have evolved as a warning system for potentially hazardous chemicals. It evokes sensations, such as irritation, tickling, burning, warming, cooling, and stinging in the nasal and oral cavities, and in the cornea, via pain receptors and the trigeminal nerve (24).

Histopathological alterations in the lungs of dead rats in the acute study are related to interstitial pneumonitis of chemical origin (consistent with capillaritis). This is probably due to the direct effect of the essential oil on the lung tissue, as it is a volatile compound. Essential oil is primarily eliminated via the airways, meaning that inflammatory infiltrates in the lung tissue are also related to TRPA1 activation, which is highly expressed in the sensory nerve cells that project vagal C fibers into the airways and lungs, where they play a role in the inflammatory response (23). In the 28-day toxicity study, concentration-dependent inflammatory infiltrates were also observed, being severe in the group with the highest dose (500 mg/kg bw), where some cases with hemorrhagic foci were also observed (Fig. 3).

In a previous study we found that the main constituent of Thymus vulgaris essential oil is thymol, which represents 46.47% of the total constituents. Other constituents are γ-terpinene (20.27%), p-cymene (15.80%), α-terpinene (2.84%), β-myrcene (1.91%); and in small amounts, α-thujene, α-pinene, camphene, α-phellandrene, d-limonene, eucalyptol, caryophyllene, germacrene D, γ-cadinene, and γ-murolene (13).

It has been reported that thymol induces TRPA1 activation (25), so it is likely that the effects produced in the rats’ respiratory system was, in part, due to the presence of this compound. It has also been reported that thymol presynaptically enhances spontaneous excitatory transmission (increases the release of L-glutamate) in lamina II (gelatinous substance) neurons by activating TRPA1 channels (26). This would explain, in part, the initial hyperactivity shown by the rats during the acute study. Likewise, it has been shown that both the essential oil of Thymus vulgaris and thymol inhibit the activity of acetyl cholinesterase (27), which may be related to the hyper-salivation that was observed after the administration of the 2,000 mg/kg bw dose.

The manifestation of symptoms before death suggest that the toxic effect occurs mainly at the central nervous system (CNS) level, which leads to lethargy, coma, and death. The CNS is particularly vulnerable to toxic substances because it has very limited capacity to regenerate, and if the nerves are damaged they cannot be recovered. Behavioral changes may include deficits in cognitive function, effects on mood or sleep (CNS), and muscle conditions, such as weakness, numbness, and alterations in motor coordination (CNS and SNP) (28). Some of these manifestations were observed in this research after the administration of 2,000 mg/kg bw of essential oil. Since there were no alterations in the brain tissue of rats that received the lower dose (300 mg/kg bw) of essential oil of Thymus vulgaris, it is possible that the neurotoxicity is related to an alteration of the neurotransmitters. Both CNS stimulation and CNS depression are manifestations of functional neurotoxicity, as they can alter the normal state of cognition, alertness, and coordination. The common effects of CNS depression include reduced respiratory and heart rate, blood pressure, temperature, and alertness, as well as partial loss of motor coordination. An overdose of depressant drugs can lead to loss of consciousness, coma, and death. Typical mechanisms of CNS depression include facilitation of GABA_A and/or opioid activity, and inhibition of adrenergic activity and/or acetylcholine (28). With regard to this, it has been shown that thymol potentiates human GABA_A receptors (29).

The histopathological alterations found in the kidneys during the acute oral toxicity study may be due to spontaneous lesions—the most common and important in rodents being chronic nephropathy. This lesion starts between three and six months of age and manifests through basophilic renal
tubules, widening of the glomeruli membranes, interstitial fibrosis, cellular infiltrates foci, and glomerulosclerosis (30). In the 28-day toxicity study, histological changes were not observed, and serum urea and creatinine levels were also unaltered (Table 1).

The hepatic histopathological study, performed after the acute toxicity test, showed no alterations; whereas in the study with repeated doses over 28 days, only mild inflammatory infiltration was observed (Fig. 2). This is related to the results of the biochemical parameters, in which AST, ALT, alkaline phosphatase, total bilirubin, total protein, and albumin were within normal ranges (Table 2). However, the mild inflammatory infiltrate observed in the stomach (Fig. 2), and the moderate sub-epithelial inflammatory infiltrate in the esophagus (Fig. 2), in a minimum number of cases, could be related to the oro gastric tube that was used for the administration of the Thymus vulgaris essential oil.

Body weight is an important indicator of toxicity. However, in the 28-day period of treatment with repeated doses of Thymus vulgaris, body weight was unaltered in female rats. Whereas in males there was a significant decrease in the last week of treatment, but only in the 500 mg/kg bw dose group (Fig. 1). Likewise, the relative weight of the organs was unaltered (Table 1). Therefore, the effect of treatment on weight is not relevant. On the other hand, the hematological parameters were not altered (Table 3), and the other biochemical parameters, such as total cholesterol, triglycerides, HDL, LDL, and glucose, remained unchanged (Table 2).

In other in vitro toxicity studies evaluating cell damage at the DNA level produced by thymol, the results were diverse. One study showed that this compound is not genotoxic in mammalian cells (31), but another study reported that thymol induced structural chromosomal aberrations and changes in frequency of micronuclei in human peripheral lymphocytes (32). Moreover, genotoxicity was dependent on the thymol concentration, so that in low concentrations did not cause DNA damage in human lymphocytes, but in higher concentrations produced a significant increase in damage (33). A similar result was observed in Chinese hamster lung fibroblast V79 cells, indicating a lack of thymol clastogenic activity in biologically relevant concentrations (34). In addition, thymol did not show any mutagenic activity in Salmonella with the Ames test, nor did it exhibit a genotoxic effect in the Caco-2 cell line of human colon carcinoma (35). The essential oil of Thymus vulgaris did not induce significant DNA damage in vitro in the human embryonic lung cells HEL 12469 (7).

There are no previous reports of acute oral toxicity from repeated doses of Thymus vulgaris essential oil, and an average oral rat LD₅₀ = 1,220 mg/kg bw dose was reported only for thymol (18). According to the results obtained from the oral acute toxicity test, and in accordance with the Globally Harmonized Classification System (GHS), established by the OECD (21), the essential oil of Thymus vulgaris falls into category 4 (> 300-2,000 mg/kg bw), with an LD₅₀ cut-off = 1,000 mg/kg bw. This result, according to Tisserand and Boyd (18), is classified as moderately toxic.

Our studies reveal that the essential oil of Thymus vulgaris has moderate oral toxicity according to the results of the acute test. Whereas the results of the 28-day oral toxicity test suggest that the no-observed-adverse effect level (NOAEL) is greater than 250 mg/kg/day. These results could be of interest in promoting the research of other in vitro and in vivo toxicity tests to establish the safety of thyme oil. It is advisable not to use high doses to avoid possible harmful effects to health.

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CONFLICT OF INTEREST

The authors declare that there are no conflicts of interest.

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