Revision of the southern Andean genus Sadocus Sørensen, 1886 (Opiliones, Gonyleptidae, Pachyliniae)

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Abstract
Species of the genus Sadocus Sørensen, 1886 are conspicuous gonyleptids that occur in Chile and Argentina. Here, the genus is revised for the first time and the cladistic analysis based on morphological characters does not corroborate its monophyly unless a phylogenetically unrelated species is excluded (explained further on). A new classification is proposed for the seven species left in the genus and considered valid, of the 13 nominal species previously recognized. Two out of the seven valid species are considered as species inquirendae: Sadocus allermayeri (Mello-Leitão, 1945) [= Carapungue allermayeri Mello-Leitão, 1945] and Sadocus nigronotatus (Mello-Leitão, 1943) [= Carapungue nigronotatum Mello-Leitão, 1943]. The following synonymies are proposed: Sadocus bicornis (Gervais, 1849) [original combination = Gonyleptes bicornis Gervais, 1849] is a junior synonym of Sadocus asperatus (Gervais, 1847) [= Gonyleptes asperatus Gervais, 1847]; Sadocus conspicillatus Roewer, 1913, Sadocus exceptionalis (Mello-Leitão, 1946) [= Araucanoleptes exceptionalis Mello-Leitão, 1946] and Sadocus guttatus Sørensen, 1902 are junior synonyms of the valid name Sadocus polyacanthus (Gervais, 1847) [= Gonyleptes polyacanthus Gervais, 1847]; and Sadocus calcar (Roewer, 1913) [= Lycomedes calcar Roewer, 1913] is a junior synonym of the valid name Gonyleptes horridus Kirby, 1819. Sadocus brasilienis Soares & Soares, 1949 is not congeneric with Argentinean/Chilean species of the genus according to the cladistic analysis and is here synonymized with Discocyrtus catharinensis (Mello-Leitão, 1923 [= Sadocus catharinensis Mello-Leitão, 1923]).

Keywords
Argentina, Chile, harvestmen
Introduction

Harvestman systematics has advanced greatly in the last few decades, especially in the Neotropical region, with many supraspecific groups being recently revised, such as for example Stygnidae Simon, 1879 (Pinto-da-Rocha 1997), Sodreaninae Soares & Soares, 1985 (Pinto-da-Rocha and Bragagnolo 2010), Goniosomatinae Mello-Leitão, 1935 (DaSilva and Gnaspinini 2010), Hernandariinae Sørensen, 1884 (DaSilva and Pinto-da-Rocha 2010), among others. Gonyleptidae Sundevall, 1833, the largest Neotropical family in number of species, includes two taxonomically challenging and species-rich subfamilies pending revision: Gonyleptinae Sundevall, 1833 and Pachylinae Sørensen, 1884. The lack of revisions is possibly due to the considerable number of species and their great morphological variation.

Pachylinae is the most species-rich subfamily of Gonyleptidae, and is currently considered polyphyletic (Pinto-da-Rocha 2002; Pinto-da-Rocha et al. 2014; Benavides et al. 2021). A phylogenetic analysis, based on molecular data (Pinto-da-Rocha et al. 2014), recovered a clade including *Pachylus* Koch (1839), the type genus of the subfamily. This clade was named Pachylinae sensu stricto and includes mainly Chilean species. This result was the first step towards the dismemberment of this large subfamily into smaller monophyletic units.

The sister group of Pachylinae sensu stricto is a clade that includes the genus *Sadocus* Sørensen, 1886, composed of rather large-sized (5.5–13.8 mm of dorsal scutum length) and colorful harvestmen. Although conspicuous and relatively common in Chilean preserved areas, it was never revised in more than 130 years of existence. Historically (see below), the genus has been subjected to many taxonomic acts, resulting in confusing species identities. One has to use poor (for modern standards), hundred-year-old descriptions to identify a given species. In addition, similar species are difficult to distinguish, raising doubts about their identities. Therefore, the revision of *Sadocus* focuses on determining the identity of the included species, which in turn will allow further understanding of their relationships, and more precise inferences of their distribution and diversity (Acosta 2002). The goals of this article are also to test the monophyly of the genus and propose a classification based on cladistic analysis.

Historical aspects of *Sadocus* Sørensen, 1886

The history of *Sadocus* Sørensen, 1886 can be quite confusing because many of its species were described before the proposition of the genus. Therefore, this historical section mentions many species in different genera and subfamilies that were later transferred to *Sadocus* (Kury 2003; Kury et al. 2020a, b), as explained further on in this article.

Guérin-Méneville (1844) described the eldest species related to *Sadocus*, *Gonyleptes planiceps* in Cuvier’s ‘Iconographie du Règne Animal’. However, the publication of this issue was delayed, and Gervais’ (1842) “redescription” was actually published first. Gervais (1842) did credit the authorship of *G. planiceps* to Guérin-Méneville, and he
redescribed that species in 1844. Gervais also described the next five species relevant to *Sadocus*: *Gonyleptus asperatus*, *G. polyacanthoides*, and *G. polyacanthus* in 1847, and *G. bicornis* and *G. subsimilis* in 1849. Butler (1873) mistakenly proposed *G. subsimilis* as a senior synonym of *G. polyacanthoides*. A year later, Butler (1874) described *Gonyleptes funestus*. In 1884, Simon transferred *G. planiceps* to *Pachylus*.

In 1886, Sørensen proposed the monotypic genus *Sadocus*, to include the type and new species *S. vitellinosulcatus*. In 1899, Loman described *Discocyrtus calcitrosus* and *Gonyleptes platei*, which are relevant to *Sadocus*, and transferred *G. funestus* to *Discocyrtus Holmberg, 1878*.

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In 1902, Sørensen: (i) synonymized *S. vitellinosulcatus* and *G. platei* Loman, 1899 with *G. polyacanthus*; (ii) created the new genus *Lycomedes* (without indication of a type species), to which he transferred *G. asperatus*, *G. bicornis*, *D. calcitrosus*, *D. funestus* and *Pachylus planiceps*; and (iii) described *Sadocus guttatus*. On that paper, Sørensen placed these species in Gonyleptidae, but without assigning them to any subfamily. Therefore, at the beginning of the 20th century, named species relevant to *Sadocus* were placed in both *Sadocus*, comprising two species (*S. polyacanthus* and *S. guttatus*) and *Lycomedes*, with five species (*L. asperatus*, *L. bicornis*, *L. calcitrosus*, *L. funestus* and *L. planiceps*).

Roewer (1913) designated *L. asperatus* as the type species of *Lycomedes*, and placed it in the Pachylinae. In that same work, he: (i) proposed the synonymy of *G. subsimilis* and *L. calcitrosus* with *L. asperatus*; (ii) described *Lycomedes calcar*; (iii) placed *Sadocus* in the Gonyleptinae and described *S. dilatatus* and *S. conspicillatus*. In his large work of 1923, Roewer proposed *Lyomedicus* as a replacement name for *Lycomedes*, which was preoccupied, and described as new the genus *Eubalta*.

Mello-Leitão (1937) described the genus *Carampangue* for his new species *C. ingens*, placing it in the Pachylinae, and in 1943, he described *C. nigronotatum*. In that same year, Roewer (1943) described *Lyomedicus brasiliensis* for the new species *J. vastus* and placed it in the Pachylinae.

In 1945, Mello-Leitão described *Carampangue allermayeri*, and in the next year (1946), he described *Araucanoleptes* for the new species *A. exceptionalis*, and placed it in the Gonyleptinae. A few years later, Mello-Leitão (1949) synonymized *J. vastus* with the older *Carampangue ingens*. In that same year, Soares and Soares (1949) described *Lyomedicus brasiliensis*, and later, H. Soares (1968) transferred *Sadocus dilatatus* to *Lyomedicus*. During the next 30 years, only few catalogues (Cekalovic 1968, 1976, 1985) mentioned the species related to *Sadocus*. 154 years after its first description, Acosta (1996) studied the collection of type material of Pachylinae described by Roewer and found differences between Guérin-Méneville’s (1844) description of *L. planiceps* and the redescription by Roewer (1913).

At the beginning of the 21st century (prior to this study), species relevant to *Sadocus* were placed in the following genera and subfamilies: the monotypic *Araucanoleptes* (Gonyleptinae); *Carampangue* (Pachylinae), with three species (*C. allermayeri*, *C. ingens* and *C. nigronotatum*); *Lyomedicus* (Pachylinae), with seven species (*L. asperatus*, *L. bicornis*, *L. brasiliensis*, *L. calcar*, *L. dilatatus*, *L. funestus* and *L. planiceps*); and *Sadocus* (Gonyleptinae), with three species (*S. conspicillatus*, *S. guttatus*, and *S. polyacanthus*).
Kury (2003), in his complete catalogue of New World Laniatores, proposed the synonymy of *Lycomedicus*, *Carampangue*, and *Araucanoleptes* with *Sadocus*. Hence, *Sadocus* comprised 14 species (actually, there are entries for 15 species, but that of *S. subsimilis* is clearly a mistake, which should be listed as a junior synonym under *S. asperatus*). Finally, Pessoa-Silva et al. (2020), transferred *S. planiceps* to *Eubalta*. *Sadocus* hitherto was composed of 13 species (Kury et al. 2020b). In the present publication, we accept only seven species of *Sadocus* as valid.

**Materials and methods**

Material examined belongs to the following institutions (curators in parentheses) listed below:

- **AMNH** American Museum of Natural History, New York, USA (L. Prendini);
- **NHM** The Natural History Museum, London, England (J. Beccaloni);
- **CAS** California Academy of Sciences (Entomology), San Francisco, California, USA (L. Esposito);
- **MCZ** Museum of Comparative Zoology, Cambridge, Massachusetts, USA (G. Giribet);
- **MNHN** Muséum National d’Histoire Naturelle, Paris, France (M. Judson);
- **MNRJ** Museu Nacional, Universidade Federal do Rio de Janeiro, Rio de Janeiro, Brazil (A.B. Kury);
- **MZSP** Museu de Zoologia, Universidade de São Paulo, São Paulo, Brazil (R. Pin-to-da-Rocha). CGPC = Carlos Nicolau Gofferjé Private Collection was transferred to MZSP;
- **SMF** Senckenberg Research Institute and Museum, Frankfurt am Main, Germany (P. Jäger);
- **UFMG** Universidade Federal de Minas Gerais, Belo Horizonte, Brazil (A.J. Santos);
- **URMU** Museo Nacional de Historia Natural de Montevideo, Montevideo, Uruguay (M. Simó);
- **ZMB** Museum für Naturkunde Leibniz-Institut für Evolutions- und Biodiversitätsforschung, Berlin, Germany (J. Dunlop);
- **ZMUC** Zoologisk Museum Universität København, Copenhagen, Denmark (N. Scharff).

The following abbreviations are used throughout the text, including synonymic listings:

- **cat** catalogue;
- **cit** citation;
- **coll** collected;
- **desc** description;
- **eco** ecology;
- **rdesc** redescription;
- **syst** systematic discussion.
In the examined material:

- **fe** female;
- **ma** male;
- **juv** juvenile;
- **MS A–E** penis ventral plate pairs of macrosetae A–E.

[**X(y)**] where X is the character number and y, the character state.

The topological nomenclature follows Acosta et al. (2007), nomenclature of integumentary ornamentation of dorsal scutum and legs, dorsal scutum outline and ventral plate penial macrosetae follows DaSilva and Gnaspini (2010), Kury and Medrano (2016) and Kury and Villarreal (2015), respectively. Nomenclature of ovipositor morphology generally follows Townsend et al. (2015). We adopted the orientation of the captured images to reference the ovipositor lobes, because we had no topological reference after detaching it. It is unlike the penis, which has a sclerotized ventral feature, thus being easily referenced topologically. In *Sadocus*, we realized that leg IV is twisted retro-laterad from the trochanter (gradually untwisting along the femur), rendering the otherwise prolateral structures as dorsal (Fig. 3A, E). To standardize the topological nomenclature, we opted to consider (and call) those as prolateral, despite being functionally dorsal (in situ). We illustrated the external morphology using a stereomicroscope with a camera lucida and the material immersed in 70% ethanol. We prepared male and female genitalia according to Pinto-da-Rocha (1997) to take pictures using a scanning electron microscope (SEM) or to illustrate using a compound microscope with a camera lucida. The generic characteristics are not repeated in the specific (re)descriptions. Only characters differing from those of the males are listed in the female (re)descriptions. The variation on the number of tubercles on the dorsal scutum and other parts of the body or legs were included in the intraspecific variation. The color descriptions are based on specimens preserved in 70% ethanol and living photograph examples presented in the section “variation in males (or females)” under each species. Many species of *Sadocus* present a white patch on the body, commonly known as a dry-mark (Kury in DaSilva and Gnaspini 2009). It is an external serose layer of the cuticle that often forms white patches/shapes. Distribution maps for *Sadocus* species were prepared using QGIS 3.10 (QGIS.org 2019). The identification key is only for males. Synonymic listings follow Kury’s catalogue (2003), to which we add the category of its content between parentheses (see abbreviations section above). All measurements are in millimeters. We followed the view of Kury et al. (2020a) regarding the use of the correct inflection of specific epithets that are adjectives throughout the article to avoid inviting further confusion for the reader. Therefore, despite Kury himself (2003) proposing the combination *Sadocus funestis* (Butler, 1874), we use *Sadocus funestus* (Butler, 1874) as Kury et al. (2020b) in all sections (except for synonymic listing), including the historical aspects of *Sadocus*.

We chose the outgroups based on available hypotheses including Pachylinae, such as Pinto-da-Rocha et al. (2014), Hara et al. (2012) and Hara (2016). We rooted the tree in Stygnidae (*Stygnus polyacanthus* (Mello-Leitão, 1923)) based on Kury (1994). We added other taxa to account for the morphological diversity of Pachylinae sensu stricto and its sister group.
List of outgroups analyzed, with respective vouchers

Acanthopachylus aculeatus (Kirby, 1819) (Gonyleptidae: Pachylinae) (MZSP 76419)
Acanthoprocta conica Maury, 1991 (Gonyleptidae: Pachylinae) (AMNH)
Goniosoma varium Perty, 1833 (Gonyleptidae: Goniosomatinae) (MZSP 76421)
Gonyleptes horridus Kirby, 1819 (Gonyleptidae: Gonyleptinae) (MZSP 59820)
Metagyndes pulchella (Loman, 1899) (Gonyleptidae: Pachylinae) (AMNH)
Nanophareus polyhastatus Hara, 2016 (Gonyleptidae: Pachylinae) (AMNH)
Neogonylyptes docilis (Butler, 1876) (Gonyleptidae: Pachylinae) (AMNH)
Neogonylyptes karschii (Sørensen, 1902) (Gonyleptidae: Pachylinae) (AMNH)
Pachylus chilensis (Gray, 1833) (Gonyleptidae: Pachylinae) (AMNH)
Pachyloides thorellii Holmberg, 1878 (Gonyleptidae: Pachylinae) (MZSP 59880)
Roeweria bittencourti Mello-Leitão, 1923 (Gonyleptidae: Roeweriinae) (MZSP 76420)
Stygnus polyacanthus (Mello-Leitão, 1923) (Stygnidae: Stygninae) (MZSP 59951)

We include records of distribution in maps only for vials with males. We used LibreOffice Calc to edit the character matrix and TNT 1.0 (Goloboff, Farris and Nixon 2008) to perform an implicit enumeration search under parsimony using equal weights. No character was ordered. We calculated Absolute and Relative Bremer support (Bremer 1994) to evaluate the support of clades using the Bremer Support Script for TNT 1.0 written by Pablo Goloboff (available at http://tnt.insectmuseum.org/index.php/Scripts/bremer). We used Winclada 1.00.08 (Nixon 1999) to edit the tree under ACCTRAN optimization and the notation of taxon+ proposed by Amorim (1982).

Results and discussion

Cladistic analysis

To test the monophyly of Sadocus, we used a matrix of morphological characters composed of 18 taxa (13 outgroups and five ingroups) and 64 characters (Table 1). The 64 characters are distributed as follows: 18 from dorsal scutum, four from free tergites, one from the chelicera, 23 from male legs, 17 from male genitalia, and one from coloration. We only included the valid Sadocus species with known males, as most of the characters are based on armature of male leg IV and penis. The cladistic analysis resulted in a single most parsimonious tree (182 steps, L = 182; C.I. = 45; R.I. = 53, Fig. 1).

According to the retrieved tree, Sadocus is not monophyletic, as it excludes Sadocus brasiliensis (Soares & Soares, 1949). Acosta (2020) first mentioned that S. brasiliensis may not belong to this genus based on the overall distribution of the other Sadocus species. That suspicion is corroborated here, and we propose its synonymy with the Brazilian Discocyrtus catharinensis (Mello-Leitão, 1923) (see taxonomic notes in this species’ entry). In turn, the close relationship of D. catharinensis with Roeweria bittencourt
**Table 1.** List of character and character states used in the cladistic analysis. All characters of legs and genitalia refer to male.

| Character | State |
|-----------|-------|
| 1 (DS) Ocularium (Hara 2016): | 0. Divided, each eye placed onto different elevations; 1. Single. |
| 2 (DS) Ocularium, unpaired armature: | 0. Absent; 1. Present. |
| 3 (DS) Ocularium, paired armature: | 0. Absent; 1. Present. |
| 4 (DS) Anterior margin of carapace, frontal hump | 0. Inconspicuous; i.e., straight from ocularium to anterior margin of DS in lateral view; 1. Conspicuous; i.e., clear elevation from ocularium to anterior margin of DS in lateral view |
| 5 (DS) Mesotergum: placement of the maximum width | 0. Maximum width in the middle of the mesotergum; 1. Maximum width placed posteriorly to the middle of mesotergum |
| 6 (DS) Dorsal scutum length and width ratio | 0. Wider than long; 1. Longer than wide |
| 7 (DS) Area I, state of fusion | 0. Divided in right and left halves by a longitudinal groove between scutal areas I – II (even though the groove of area II slightly invades area I) 1. Divided in right and left halves by invasion of scutal area II into middle of scutal area I. |
| 8 (DS) Scutal area I, paramedian armature (Hara 2016) | 0. Absent or with similar sized granules; 1. With a pair of tubercles |
| 9 (DS) Scutal area II, paramedian paired armature (Hara 2016) | 0. Absent or with similar sized granules; 1. With a pair of tubercles |
| 10 (DS) Scutal area III, paramedian paired armature (Hara 2016) | 0. Absent or with similar sized granules; 1. With a paramedian pair of tubercles |
| 11 (DS) Scutal area IV, presence | 0. Absent; 1. Present. |
| 12 (DS) Scutal area IV, degree of division | 0. Incompletely divided; i.e., interrupted scutal groove IV; 1. Completely divided |
| 13 (DS) Scutal area IV, paramedian paired armature | 0. Absent; 1. Present. |
| 14 (DS) Lateral margin, type of integumentary ornamentation | 0. Covered with granules; 1. With tubercles, sometimes clustered |
| 15 (DS) Lateral margin, type of armature | 0. Large tubercles or apophyses; 1. similar sized tubercles |
| 16 (DS) Posterior margin of the DS, paramedian armature | 0. Absent; 1. Present. |
| 17 (DS) Posterior margin of the DS, central unpaired armature | 0. Absent; 1. Present. |
| 18 (DS) Free tergites I, paramedian armature | 0. Absent; 1. Present. |
| 19 (DS) Free tergites II, paramedian armature | 0. Absent; 1. Present. |
| 20 (DS) Free tergites II, unpaired armature | 0. Absent; 1. Present. |
| 21 (DS) Free tergites III, paramedian paired armature | 0. Absent; 1. Present. |
| 22 (Chelicerae) Chelicerae, sexual dimorphism (#30, Hara 2016) | 0. Isomorphic in both sexes 1. Large in male |
| 23 (Pedipalp) Tibia, type of retro-lateral apical seta | 0. Single; 1. Bifh |
| 24 (Leg) Coxa IV, branch of the prodorsal apophysis | 0. Single; 1. Bifh |
| 25 (Leg) Coxa IV, insertion of the prodorsal apophysis in relation to the DS main axis (Hara et al. 2012) | 0. Almost transversal; i.e., almost 90 degrees in relation to DS main axis; 1. Oblique; i.e., more than 120 degrees in relation to DS main axis; 2. Parallel to femur IV |
| 26 (Leg) Trochanter IV, retro-apical apophysis: | 0. Absent; 1. Present. |
| 27 (Leg) Leg IV, torsion that begins at the trochanter and ends at the patella | 0. Untwisted; 1. Strongly twisted; i.e., prolateral features becoming dorsal in situ and gradually untwisting towards patella; 2. Strongly twisted from coxa IV towards the patella (Fig. 3A, E). |
| 28 (Leg) Trochanter IV, Prolateral basal apophysis (# 30 Hara & Pinto–da–Rocha 2010) | 0. Absent; 1. Present. |
| 29 (Leg) Trochanter IV, dorso-median subapical apophysis | 0. Absent; 1. Present. |
| 30 (Leg) Trochanter IV, prodorsal apical apophysis | 0. Absent; 1. Present. |
| Character | State |
|-----------|-------|
| 32 (Leg) Trochanter IV, type of prodorsal apical apophysis | 0. As a wart; 1. As a hook-like pointed apophysis of large base, smoothly becoming pointed apically; 2. As a moderate size blunt cone; 3. As a finger shaped, robust apophysis, basal half of ca. uniform diameter |
| 33 (Leg) Trochanter IV, retro-dorsal apical apophysis (# 29 Hara & Pinto–da–Rocha 2010) | 0. Absent; 1. Present. |
| 34 (Leg) Trochanter IV, retro-apical armature (Hara & PdR 2010) | 0. Absent; 1. Present. |
| 35 (Leg) Trochanter IV, type of retro-apical armature | 0. Pointed tubercle; 1. Moderate apophysis (ca. a quarter of podomere width); 2. Huge apophysis (as long as podomere width). |
| 36 (Leg) Trochanter IV, proapical apophysis | 0. Absent; 1. Present. |
| 37 (Leg) Trochanter IV, length-width ratio (modified from Hara 2016) | 0. As long as wide; 1. Twice longer than wide; 2. Wider than long. |
| 38 (Leg) Femur IV, curvature in dorsal view | 0. Straight; 1. Sinuous |
| 39 (Leg) Femur IV, size of granules on retro-lateral row | 0. Similar sized granules; 1. Tubercles, twice the size of granules. |
| 40 (Leg) Femur IV, spiniform apophyses on basal half of the retro-lateral row of granules | 0. Absent; 1. Present. |
| 41 (Leg) Femur IV, the pattern of apophyses distribution at the ⅔ basal region of the retro-lateral row | 0. Just an apophysis in the basal ⅓; 1. Growing from the median region to the basal region; 2. Very high apophyses alternating with low apophyses; 3. Apophyses distributed in the median region; 4. A basal apophysis and one or more in the distal ⅔; 5. An average apophysis; 6. Decreasing from the median region to the basal region. |
| 42 (Leg) Patella IV, ventral row of granules | 0. Similar sized granules; 1. Granules becoming tubercles or spines. |
| 43 (Leg) Tibia IV, venter-basal long spine | 0. Absent; 1. Present. |
| 44 (Leg) Tibia IV, size of granules on retro-ventral row | 0. Tubercles of similar sizes; 1. Granules increasing in size apically, becoming tubercles. |
| 45 (Leg) Tibia IV, proventral row of tubercles size | 0. Tubercles of similar sizes; 1. Larger tubercles, which grows in size apically. |
| 46 (Leg) Tibia IV, size of granules on ventral row | 0. Similar sized granules; 1. Increasing in size apically. |
| 47 (Penis) Ventral plate, shape of the distal margin | 0. Straight; 1. Slightly concave; 2. Very concave, forming a “U”. |
| 48 (Penis) Ventral plate, basal lobes | 0. Inconspicuous; 1. Conspicuous. |
| 49 (Penis) Ventral plate, plate format | 0. Rectangular; 1. Hexagonal. |
| 50 (Penis) Ventral plate, number of MS C | 0. Three pairs; 1. Four pairs or more. |
| 51 (Penis) Glans, dorsal prominences in the distal region of the sac | 0. Absent; 1. Present. |
| 52 (Penis) Glans, sac texture | 0. Smooth and turgid; 1. Wrinkled. |
| 53 (Penis) Glans, latero-apical region | 0. Without projections; 1. With projections covering part or all of the pedestal in lateral view. |
| 54 (Penis) Glans, dorsal process | 0. Absent; 1. Present. |
| 55 (Penis) Glans, ventral process | 0. Absent; 1. Present. |
| 56 (Penis) Ventral process, presence of stem | 0. Absent; 1. Present. |
| 57 (Penis) Ventral process, apex shape | 0. As a flabellum; 1. Tapered at the apex and rolled; 2. Flattened circular; 3. Flattened quadrangular; 4. Fringed triangular; 5. Large rectangular; 6. Rectangular bifid; 7. Rectangular with pointed projections. |
| 58 (Penis) Stylus, ventral process length ratio | 0. Stylus shorter than ventral process; 1. Stylus longer than ventral process. |
| 59 (Penis) Stylus, apical lateral projections | 0. Absent; 1. Present. |
| 60 (Penis) Stylus, apex shape (DaSilva and Gnaspini 2010) | 0. Rounded; 1. With an apical back beak. |
| 61 (Penis) Stylus, trichomes on median apical region | 0. Absent; 1. Present. |
| 62 (Penis) Insertion on the glans in lateral view | 0. Ventrall; 1. Median |
| 63 (Penis) Trunk of the penis, subapical region | 0. Truncated; 1. Projected on the glans |
| 64 (Color) Carapace, presence of dry-mark | 0. Absent; 1. Present. |
Systematics of harvestman *Sadocus*

(Roeweriinae Brazilian species) is supported by seven synapomorphies, three of them exclusive: hexagonal shape of the penial ventral plate [49(1)]; penial ventral process apex tapering distally, becoming rolled [57(1)]; and penial stylus with latero-apical projections [59(1)]. *Discocyrtus catharinensis* is probably a Roeweriinae especially based on the shape of the penial ventral plate as well as the overall penial morphology. However, we refrained further taxonomic actions as Roeweriinae diversity grows further fueled by the dismemberment of *Discocyrtus*, which is currently under revision (Kury and Carvalho 2016; Carvalho and Kury 2018, 2021).

Once we settled the issue related to *S. brasiliensis*, we propose a new concept of *Sadocus*. Under the new definition, *Sadocus* is monophyletic and supported by seven synapomorphies, four of which are exclusive: leg IV twisted from the trochanter to patella [28(0)]; trochanter IV with a finger shaped, robust prodorsal apical apophysis, its basal half of ca. uniform diameter [32(3)], trochanter IV twice longer than wide [37(1)] (modified from Hara 2016); and penis ventral plate with slightly concave distal margin [47(1)]. *Sadocus* is also the best supported clade of the analysis, with a high Goodman-Bremer support (4).

So far, *Sadocus* (represented especially by *S. polyacanthus*, its type species) has often been used in cladistic analysis as outgroups (Hara et al. 2012; Hara 2016) or as an ingroup taxon of a more comprehensive analysis testing monophyly of Gonyleptidae or Gonyleptoidea (Pinto-da-Rocha et al. 2014; Benavides et al. 2021). According to the analyses based on morphological characters (Hara et al. 2012; Hara 2016), *Sadocus* is nes-
Table 2. Matrix of character states for the cladistic analysis of the *Sadocus* (Gonyleptidae: Pachylinae).

| Species                  | 0 1 0 0 0 1 2 0 1 0 1 0 | 0 1 0 0 1 1 0 1 0 1 0 0 0 0 1 0 0 0 1 0 | - 0 - 0 2 0 0 0 1 0 1 1 0 0 0 0 1 0 0 1 0 | - - - - 0 0 1 0 0 0 1 0 0 1 0 0 1 0 0 0 0 0 1 0 0 1 0 4 0 0 0 0 0 1 0 0 0 0 0 1 0 0 0 0 0 1 0 0 0 0 0 1 0 0 0 0 0 1 0 0 0 0 0 1 1 0 0 0 0 0 1 0 0 0 0 0 1 0 |
tled in a clade mainly composed of Brazilian species. However, we have to stress that the clade with *Sadocus* in Hara’s analyses is not well supported (Bremer support: 1), its sole homoplastic synapomorphy being the proventral apical armature of tibia IV as a tubercle. In the present analysis, we have a roughly similar outcome, as the clade including *Sadocus* (*Sadocus* + *Neogonyleptes*) is sister group to a clade composed of solely Brazilian species. This outcome differs considerably from Pinto-da-Rocha et al. (2014) or Benavides et al. (2021): in those analyses based on molecular data, *Sadocus* is often retrieved closely related to Chilean Pachylinae genera. Regarding this, Pinto-da-Rocha et al. (2014) indicate that *Sadocus* is in a clade with other Chilean species (*Neogonyleptes karschii* and *Tumbesia aculeata*), which in turn is sister group to Pachylinae sensu stricto. Benavides et al. (2021) also corroborates a close relationship of *Sadocus* with Chilean genera.

The sister taxon closest to *Sadocus* is also an unsettled issue, mainly because different taxa are employed in those analyses. In the present analysis, *Sadocus* sister group is the Chilean genus *Neogonyleptes*, supported by seven synapomorphies, two of them exclusive: ventral row of granules increasing in size apically on tibia IV [46(1)]; and apex of glans ventral process flattened circle shaped [57(2)]. This sister group relationship is similar to Pinto-da-Rocha et al. (2014) sampling wise. On the other hand, Benavides et al. (2021) did not include *Neogonyleptes* in their analysis, and the Chilean *Eubalta planiceps* is the sister taxon to *Sadocus*. It is interesting to note that *Sadocus* sister taxon is strongly affected by the Chilean Pachylinae sampling not belonging to Pachylinae sensu stricto.

The main goals of this study were to revise and to test the monophyly of *Sadocus*, a hundred-year-old genus, with a convoluted taxonomic history. We believe that we succeeded in those, and the present study is an important step towards the understanding the evolution of the genus. Considering all the evidence (including the taxonomic history), *Sadocus* seems to be related to the Chilean-Argentinean Pachylinae. We understand that *Sadocus* relationship within Gonyleptidae is still an unsettled issue that deserves further investigation. As a mean to tackle that, we can suggest the inclusion of more Chilean Pachylinae genera (especially those already used in previous analyses and not belonging to Pachylinae sensu stricto) and Brazilian species as well, such as DRMN (Carvalho and Kury 2018) and K92 (Kury 1992). The latter suggestion is because of the Brazilian clade closest to Chilean Pachylinae clade depicted by Pinto-da-Rocha et al. (2014) and Benavides et al. (2021).

**Taxonomic accounts**

**Gonyleptidae Sundevall, 1833**

**Pachylinae Sørensen, 1884**

**Sadocus Sørensen, 1886**

*Gonyleptes* [part]: Gervais, 1842: 2 [rdesc]; 1844: 105 [rdesc]; 1847: 576–577 [cit]; 1849: 21, 24–26 [desc, rdesc]; Butler 1873: 113–114 [cat]; 1876: 153 [desc].

*Discocyrtus* [part]: Loman, 1899: 6 [desc].
Sadocus Sørensen, 1886: 85 [desc]; 1902: 14–20 [rdesc]; Hogg 1913: 48 [cit]; Roewer 1913: 244–245 [cit, key]; 1923: 492 [cit, key]; Mello-Leitão 1923: 190 [cit]; 1926: 31 [key]; Roewer 1930: 381 [key, cit]; Mello-Leitão 1931a: 136 [cit]; 1932: 348 [rdesc]; 1935: 105 [key]; Canals 1936: 70 [cat]; Kästner 1937: 389 [desc]; Roewer 1938: 02 [cit]; Mello-Leitão 1939: 625 [cit]; B. Soares 1944a: 166 [sys]; Soares and Soares 1949: 211 [rdesc]; Ringuelet 1955a: 438; Ringuelet 1959: 409 [rdesc]; Roewer 1961: 102 [cat]; Cekalovic 1985: 14 [cat]; Kury 2003: 191 [cat]; Hará et al. 2012: 38–39, 18 [sys]; Pinto-da-Rocha et al. 2012: 61 [cit]; Pinto-da-Rocha et al. 2014: 18 [sys]; Hará 2016: 106–109 [sys]; Pérez-Schultheiss et al. 2019, 4, 12–15 [cit]; Kury et al. 2020a: 5 [cit]; Acosta 2020 [cit]; Kury et al. 2020b [cat]. (Type species Sadocus vitellinosulcatus Sørensen 1886, by monotypy).

Lycomedes Sørensen, 1902: 17 [desc]; Roewer 1913: 126–127 [rdesc, key]; Mello-Leitão 1926: 31 [key]; Muñoz-Cuevas 1973: 226–228 [cit]. (Type species Gonyleptes asperatus Gervais, 1847, by subsequent designation by Roewer 1913).

Lycomedicus Roewer, 1923: 442 (nom. nov. for Lycomedes Sørensen, 1902, rdesc); 1925: 17 [cit]; 1929: 213 [cat]; Mello-Leitão; 1931b: 84 [cit]; 1932: 216 [rdesc]; 1935: 101 [cit]; Canals 1936: 69 [cat]; Roewer 1943: 28 [cit]; Ringuelet 1959: 329 [rdesc]; Cekalovic 1985: 18 [cat]. Synonym established by Kury 2003.

Lycomedius [lapsus]: Kästner, 1937: 389 [rdesc]; Strand 1942: 397 [cit].

Carampangue Mello-Leitão, 1937: 152 [desc]; 1945: 156 [cit]; 1949: 17 [sys]; Soares and Soares 1954: 241 [rdesc, cat]; Cekalovic 1985: 16 [cat]. (Type species Carampangue ingens Mello-Leitão, 1937 by monotypy). Synonymy established by Kury 2003.

Jighas Roewer, 1943: 28 [desc]; Mello-Leitão 1949: 17 [sys]. (Type species Jighas vastus Roewer, 1943 by monotypy). Synonymy established with Carampangue by Mello-Leitão 1949.

Araucanoleptes Mello-Leitão, 1946: 4 [desc]; Soares and Soares 1949: 160 [rdesc]. (Type species Araucanoleptes exceptionalis Mello-Leitão, 1946 by monotypy). Synonymy established by Kury 2003.

Arauconoleptes [lapsus]: Cekalovic, 1985: 12 [cat].

Type species. Sadocus vitellinosulcatus Sørensen, 1886, by monotypy. Synonymized with S. polyacanthus by Sørensen (1902).

Other species included. S. asperatus (Gervais, 1847), S. dilatatus Roewer, 1913, S. funestus (Butler, 1874), S. ingens (Mello-Leitão, 1937) and S. polyacanthus (Gervais, 1847).

Diagnosis. Sadocus are large Pachylinae (dorsal scutum maximum length 5.5–13.8 mm) with paired spines on ocularium and prominent frontal hump on dorsal scutum anterior margin. Dorsal scutum shape types gamma triangular and gamma pyriform, its posterior margin concave. Dorsal scutum mid-bulge placed close to scutal groove IV (scutal groove III in S. funestus) and transversal (S. funestus, S. ingens, S. polyacanthus) or oblique (S. asperatus, S. dilatatus); free tergites II and III each with a pair of spines. Legs IV are twisted retro-laterad from the trochanter and gradually distorted along the femur and patella (except in S. funestus). Coxa IV bearing a long, large pro-dorsal apical apophysis and a short, retro-apical one (except in S. funestus, in which is lacking). Trochanter IV with a short, blunt prolateral sub-basal apophysis and a long,
robust prodorsal apical one. Penis glans turgid and dorsally projected (with antero-lateral projections), with ventral process (half stylus length) and without dorsal process. General color (in living specimens) of the body and most parts of legs and ventral area dark brown, with lighter tones at the tips of podomeres. Yellowish to reddish tone in scutal area, scutal posterior margin, free tergites, part of legs and apophysis. Green on the arthrodial membranes between the free tergites.

**Redescription. Male: Dorsum.** Anterior margin of carapace with a prominent median frontal hump (bell shaped in dorsal view). Ocularium with one pair of spines posterior to the eyes. Dorsal scutum type varying from gamma to gamma triangular and gamma pyriform, its posterior margin concave, mid-bulge slightly asymmetrical and displaced posteriorly, widest at the scutal groove IV (scutal groove III in *S. funestus*). The curvature of mid-bulge can be transversal (*S. funestus, S. ingens, and S. polyacanthus*) or oblique (*S. asperatus and S. dilatatus*). Four scutal areas (three in *S. polyacanthus*); scutal area I divided into right and left halves by a longitudinal median groove. Scutal area III with one pair of paramedian spiniform tubercles or spines. Two pairs of ozopores close to coxa II. Lateral margin of dorsal scutum with an external and internal rows of tubercles (the external row of slightly larger tubercles) (except *S. asperatus*, with granules covering most of the lateral margin of dorsal scutum and *S. ingens*, smooth or with only few granules). Posterior margin of dorsal scutum and free tergite I each with one paramedian pair of tubercles (except *S. funestus* and *S. polyacanthus*, unarmed). Free tergites II and III each with one paramedian pair of spines. **Venter.** Coxa I–IV granulate; coxa I with a median longitudinal row of granules increasing in size apically, becoming tubercles. **Chelicerae.** Isomorphic in males and females. Segment I with well-marked bulla. Segment II fixed finger and segment III toothed. **Pedipalps.** Trochanter dorsal face inflated; ventral face with one or two setiferous tubercles. Tibiae and tarsi dorsal and lateral faces with few minute granules and variable setation. **Legs.** Coxae I–III each with one prodorsal and one retro-dorsal spiniform tubercles, ventral faces granulate (except *S. polyacanthus*, coxa I with tubercles and others with setae). Coxa IV dorso-lateral face with sparsely distributed granules, ventral face entirely granulated, with one long, oblique, bifid prodorsal apical apophysis (transversal in *S. dilatatus*, uniramous in *S. funestus*), dorsal branch longest and curved ventrad and ventral branch short and blunt; and one ventro-apical retro-lateral spine. Trochanters I–III granulate. Leg IV twisted retro-laterad from the trochanter, gradually untwisting along the femur (except *S. funestus*, straight). Trochanter IV longer than wide; prolateral face with one short, conical, blunt sub-basal apophysis, and one robust, blunt dorso-apical apophysis. Femora I–IV with granules roughly organized in six longitudinal rows (prodorsal, retro-dorsal, pro- and retro-lateral, proventral and retro-ventral rows); femora I and II unarmed. Femur IV curved, with marked inner curvature on the distal half (*S. asperatus and S. ingens*) or almost straight (*S. dilatatus, S. funestus, and S. polyacanthus*). Patellae I–III granulate, unarmed; patella IV dorsal face granulate, ventral face tuberculate. Tibiae I–III granulate, unarmed (except *S. dilatatus and S. funestus*, tibia III dorsal face granulate with retro-ventral row of tubercles increasing in size apically). Tibia IV dorsal face granulate, ventral face with tubercles sparsely distributed. Metatarsi I–IV minute granulate, unarmed. Tarsus III and IV each
with ventral process, tarsal claws smooth. **Penis.** Ventral plate distal margin with slight (but conspicuous) to moderate concavity, two or three pairs of MS A, one pair of MS B or entirely absent, four or five pairs of MS C, one or two pairs of MS D, and one or two pairs of MS E. Glans sac tall, turgid, dorsally projected with antero-lateral projections, forming a sheath for the stylus. Glans without dorsal process; stylus inserted ventrally and smooth. Glans ventral process is short (half of stylus length), parallel to the stylus, apex curved ventrad, with a short semi-circular antero-lateral projection.

**Geographic distribution (Fig. 2).** Central Chile, Región XIV Los Ríos; Metropolitan Region of Santiago; Región V Valparaíso; Región VIII Bio-Bío; Región IX Araucanía; and Region X Los Lagos. There are other localities mentioned in the literature besides the material studied here for *S. polyacanthus* in Neuquén (Argentina) and Magallanes, in the extreme south of Chile, however, we did not examine any material from there. The record of *Sadocus funestus* for Ecuador (Chimborazo, Riobamba) by Roewer (1913) is certainly a mislabeling because it does not agree with the known generic distribution (Cekalovic 1985; Kury 2003). Two species are widely distributed (*S. asperatus* and *S. polyacanthus*) and three others occur mainly in coastal mountains of Central Chile (*S. funestus, S. dilatatus, and S. ingens*).

**Key to males of Sadocus species**

1. Coxa IV with a bifid prodorsal apical apophysis (Fig. 3A, C) and one retro-ventral apical apophysis (Fig. 3A) .................................................................................................................................................................2
   - Coxa IV with an unbranched prodorsal apical apophysis (Fig. 5A, C), without retro-ventral apical apophysis (Fig. 5A) .................................................................................................................................................................**S. funestus**

2. Lateral margin of the dorsal scutum with a posterior large tubercle (Figs 4A, 7A) ........................................................................................................................................3
   - Lateral margin of the dorsal scutum only with similar sized tubercles (Fig. 6C) .................................................................................................................................................................................................4

3. Dorsal scutum with four areas (Fig. 4A), femur IV with a long, retro-lateral sub-basal apophysis (Fig. 4D) .......................................................... **S. dilatatus**
   - Dorsal scutum with three areas (Fig. 7A), femur IV retro-lateral face with spines of similar size (Fig. 7B) .......................................................... **S. polyacanthus**

4. Trochanter IV dorso-apical face only with one prolateral apophysis of similar length as the podomere, strongly curved (in lateral view), pointing frontwards (Fig. 6C) ........................................................................................................................................**S. ingens**
   - Trochanter IV dorso-apical face with two apophyses, both shorter than the podomere length, femur IV slightly curved (Fig. 3B, E) .......... **S. asperatus**

**Sadocus asperatus** (Gervais, 1847)
Figures 2, 3A–E, 8A, B, 9A–C, E, 11A, B, 12A–C

**Gonyleptes asperatus** Gervais, 1847: 577 [desc]; 1849: 26, pl. 1, fig 9 [rdesc]; Butler 1873: 113 [cat]; Sørensen 1902: 17 [syst]. Transferred to Lycomedes by Sørensen
1902. (MNHN, type lost, not examined) (RPR visited the museum but the curator was unable to find the type material)

*Lycomedes asperatus* Sørensen, 1902: 17 [syst]; Roewer 1913: 127–130, fig 57 [key, rdesc].

*Lycomedicus asperatus*: Roewer, 1923: 442, fig 556 [rdesc]; 1925: 17 [cit]; 1929: 213 [cat]; Mello-Leitão 1931b: 84 [cit]; Canals 1936: 69 [cat]; Roewer 1938: 6 [cat]; Mello-Leitão 1939: 624 [cat]; Soares and Soares 1954: 270 [cat]; Ringuelet 1959: 329, fig 44 [rdesc]; H. Soares 1968: 264 [cit]; Cekalovic 1968: 8 [cat]; 1985: 18 [cat].

*Sadocus asperatus*: Kury, 2003: 191 [cat]; Kury et al. 2020b [cat].

*Sadocus (?) sub similis*: Sørensen, 1902: 17 [cit]; Roewer 1913: 127–130, fig 57 [syst].

Synonymy with *L. asperatus* established by Roewer 1913.

*Gonyleptes polyacanthoides* Gervais, 1847: 577 [desc]; Butler 1873: 114 [cat]. Synonymy with *G. sub similis* established by Butler 1873. (MNHN, type lost, material not examined) (RPR visited the Museum but the curator was unable to find the type material)

*Gonyleptes sub similis* Gervais, 1849 [desc]: 25, pl. 1, fig 8; Butler 1873: 114 [cat]; Sørensen 1902: 6 [syst], 16 [syst]. Synonymy with *Sadocus (?) sub similis* established by Sørensen 1902 (MNHN, type lost, material not examined) (RPR visited the museum but the curator was unable to find the type material)

*Gonyleptes bicornis* Gervais, 1849: 21, pl. 1, fig 4a–b [desc]; Butler 1873: 114 [cat]; Sørensen 1902: 19 [syst]. (type material depository unknown). syn. nov.

*Lycomedes bicornis*: Sørensen, 1902: 20, fig 4–4b [rdesc]; Roewer 1913: 136–137, fig 62 [rdesc].

*Lycomedicus bicornis*: Roewer, 1923: 445, fig 561 [cit]; Canals 1936: 69 [cat]; Soares and Soares 1954a: 270 [cat]; Cekalovic 1968: 7 [cat], 1985: 18 [cat].

*Sadocus bicornis*: Kury, 2003: 191 [cat]; Kury et al. 2020b [cat].

*Discocyrtus calcitrosus* Loman, 1899: 7, fig 5 [desc]; Sørensen 1902: 19 [syst]. (type ZMB 7837, ma holotype – examined by detailed photographs).

*Lycomedes calcitrosus*: Sørensen, 1902: 19 [syst]. Synonymy with *L. asperatus* established by Roewer 1913.

*Lycomedicus calcitrosus*: Moritz, 1971: 193 [cat].

**Material examined.** Chile, date and collector unknown, 1 ma (MCZ 31267). Región Metropolitana de Santiago, *Santiago*, date or collector unknown, 6 ma, 3 fe (SMF 1369); Same, *El Canelo*, 16.I.1980, collector unknown, 2 ma (AMNH); Same, 16.I.1980, collector unknown, 1 ma, 1 fe (AMNH). Región de Valparaíso, collection date or collector unknown, 8 ma, 19 fe (SMF 5382). Región de Biobío, *Provincia de Concepción*, 22.I.1985, N.I. Platnick & O.F. Francke coll., 1 ma (AMNH); Same, Estero Nonguén (-36.831501, -73.008374), 13.III.1977, T. Cekalovic coll., 1 ma (MCZ 31275); Same, Cerro Caracol, Mirador Aleman (-36.834167, -73.047778), 15.IX.1968, T. Cekalovic coll., 1 ma, 3 fe (MZSP 9965); Same, Parque Nacional Nahuelbuta (-37.800000, -73.033333), 30.XI.2003, I. Avila, S. Ocares & D. Silva coll., 1 ma (CAS 9055050); Same, Parque Nacional Nahuelbuta (-37.8043374259, -73.0344813614, date or collector unknown, 1 ma, 1 fe (CAS 9052227); Same, Parque Nacional Nahuelbuta (-37.827500, -73.009722), 9.XII.2010, F. Marques, F.
Sadocus asperatus resembles S. ingens, S. polyacanthus, and S. dilatatus by the bifid prodorsal apical apophysis on coxa IV. S. asperatus can be distinguished from the latter species by the combination of the following characters: lateral margin of dorsal scutum covered by granules; trochanter IV with a blunt retro-dorsal apical apophysis being half of the podomere length, and a rhombus retro-ventral apical tubercle; femur IV curved (in dorsal view), with a retro-lateral row of spiniform apophysis (the middle one longest).
Redescription. Male (CAS 9055035). Measurements. Dorsal scutum maximum length 6.3; dorsal scutum maximum width 7.2; prosoma maximum length 2.5; prosoma maximum width 3.2; leg femora I 3.2; II 6.5; III 5.5; IV 7.2. Dorsum (Fig. 3A). Dorsal scutum type gamma triangular. Carapace with granules sparsely distributed. Scutal areas I–IV with eight, four, two and four granules, respectively; scutal area III with one pair of paramedian spiniform tubercles; scutal area IV incompletely divided. Lateral margin of dorsal scutum mostly covered in granules (from posterior half of carapace to posterior margin of dorsal scutum). Posterior margin of dorsal scutum and free tergite I each with a pair of paramedian tubercles. Chelicerae. Segment I with basal tubercle, bulla with small setae, each finger with five teeth. Pedipalps. Coxa dorsal face smooth, ventral face with two apical tubercles. Trochanter dorsal and ventral faces smooth. Femur ventral face granulate. Patella with sparsely distributed setae. Tibial setation: prolateral lili/lili; retro-lateral lili/lili. Tarsal setation: prolateral and retro-lateral lili/lili. Legs (Fig. 3B–E). Coxa IV with one long, oblique, bifid prodorsal apical apophysis and one retro-ventral apical spine. Trochanters I and II each with one pair of prodorsal spiniform tubercles. Trochanter III with one medio-ventral
tubercle and three retro-lateral ones. Trochanter IV prodorsal and proventral faces with few granules, the prodorsal apical apophysis long (ca. half the podomere length); retro-lateral face with one basal, one central, and one apical tubercles; one retro-dorsal apical spiniform apophysis (ca. ¼ podomere length); ventral face with setiferous tubercles sparsely distributed. Femur III with one retro-basal tubercle. Femur IV sigmoid, with dorsal row of apophysis on the basal half abruptly decreasing in size apically, becoming granules; prolateral row with central–subapical tubercles; retro-lateral row of granules with spiniform apophysis (basal most and central one longer than the others, apical most oblique, curved ventrad); ventral face with two short retro-lateral sub-apical spiniform apophyses and one proapical spiniform apophysis. Patella IV ventral face mostly smooth, with one probasal, one proapical and one retro-apical large, spiniform tubercles. Tibia IV roughly with two ventral rows of granules increasing in size from central to apical becoming spines. Tarsal counts: 6, 9, 7, 8. Penis (Fig. 11A, B). Ventral plate of penis with moderate cleft on anterior margin, three pairs of MS A, one pair of MS B, four or five pairs of MS C and one pair of MS D, without MS E.

Coloration. Immersed in ethanol: carapace, trochanter–patella IV brown, tibia light brown, legs I–III, pedipalps, and chelicerae yellowish-brown. Live specimens (Fig. 9A–C, E): carapace, coxa, and trochanter black, light gray spot (inverted T shape) from ocellarium to scutal area II; femora I–III orange; patellae–tibiae I–IV brown.

Variations (n = 56). Scutal area I–IV tubercles (5 minimum, 16 maximum per site), Measurements. Dorsal scutum maximum length 6.0–8.5; dorsal scutum maximum width 6.7–11.6; prosoma maximum length 2.5–3.5; prosoma maximum width 3.2–4.8; leg femora: I 3.2–5.0; II 6.5–10.0; III 5.5–8.5; IV 7.2–11.0.

Female (CAS 9055035). Measurements. Dorsal scutum maximum length 6.3; dorsal scutum maximum width 8.1; prosoma maximum length 2.6; prosoma maximum width 3.7; leg femora: I 3.7; II 7; III 6.1; IV 7.2.

Redescription. Dorsum (Fig. 8A, B). Scutal areas I, II, and IV with three, five, and two granules, respectively. Legs. Coxa IV with one prodorsal apical apophysis and one retro-ventral apical spine shorter than on the male, trochanter IV with retro-lateral row of tubercles, the apical one longest, femur IV with pro- and retro-ventral rows of tubercles, patella–metatarsi IV unarmed. Tarsal counts: 6, 9, 7, 8. Ovipositor (Fig. 12A–C). Two main groups of lobes delimited by a constriction, ovipositors peripheral setae inserted into sockets that are a mixture of dorsal and ventral sockets, the dorsal lobe with five setae and the ventral one with six; each main group of lobes divided by a fissure.

Variations (n = 66). Tubercle variation in the scutal areas: I 2–7; II 1–8; III 2–8; IV 3–5. Free tergites I–III each with one pair of blunt or pointed spiniform tubercles. Measurements. Dorsal scutum maximum length 5.9–8.2; dorsal scutum maximum width 8.0–9.6; prosoma maximum length 2.4–3.5; prosoma maximum width 3.7–4.5; leg femora: I 3.5–4.5; II 6.7–8.7; III 5.7–7.0; IV 7.0–8.9.

Type locality. Of Gonyleptes asperatus and Gonyleptes subsimilis: CHILE. Of Disco-cyrtus calcitrosus: CHILE, Región de Los Ríos, Provincia de Valdivia, Corral.

Geographical distribution (Fig. 2). CHILE, Región de Los Ríos, Valdivia, Corral; Región Metropolitana de Santiago, Santiago; Región de Valparaíso; Región de
Figure 3. *Sadocus asperatus* (Gervais). Male (CAS 9055035) A Habitus, dorsal view B–E right leg IV, Trochanter–tibia IV B dorsal view C prolateral view D ventral view E retro-lateral view. Scale: 1 mm.
Biobío, Provincia de Concepción, Provincia de Arauco, Provincia de Nuble; Región de Araucanía, Provincia de Cautín, Provincia de Malleco; Región de Los Lagos, Provincia de Llanquihue.

**Taxonomic notes.** After examining the original description and the drawing of *Gonyleptes bicornis*, we concluded that it is of a male of *S. asperatus*. In the original description, the spines on the free tergite, the two apical apophyses on the trochanter IV and uneven spines in the inner part of the “leg” (referring to the femur IV) are mentioned. Those characters lead us to conclude that it is *S. asperatus*.

**Sadocus dilatatus** Roewer, 1913
Figures 2, 4A–E, 8C, D, 9D, 11G, H

*Sadocus dilatatus* Roewer, 1913: 249, fig 102 [desc]; 1923: 493–494, fig 620 [rdesc]; Canals 1936: 70 [cat]; Soares and Soares 1949: 211 [cat]; Cekalovic 1968: 7 [cat]; Acosta 1996: 223 [cat]; Kury 2003: 191 [cat]; Kury et al. 2020b [cat] (SMF RI, 886, ma holotype – examined).

*Lycomedicus dilatatus*: H. Soares, 1968: 264 [rdesc]; Cekalovic 1985: 18 [cat].

**Material examined.** Chile, Región de Biobío, Provincia Concepción, date or, collector unknown, 1 ma (SMF 886 – Holotype); Same, 1.XI.1964, T. Cekalovic coll., 2 ma, 1 fe (MZSP 7875); Same, Quebrada Pinares, 4.XI.1964, T. Cekalovic coll., 1 ma (MZSP 7876); Same, Reserva Nacional Nonguén (-36.878430, -72.994350), G. Giribet, G. Hormiga & A. Pérez-González coll., 11.XI.2014, 1 ma, 2 fe (MCZ 140078).

**Diagnosis.** *Sadocus dilatatus* resembles *S. polyacanthus* by the lesser-armed femur IV (compared to other species) and by the posterior large tubercle on the lateral margin of dorsal scutum. *Sadocus dilatatus* can be distinguished from the other species of the genus by the single retro-ventral central apophysis on femur IV and the very long prodorsal apical apophysis on coxa IV (ca. ⅔ of the scutum width).

**Redescription.** Male (SMF 886). **Measurements.** Dorsal scutum maximum length 7.5; dorsal scutum maximum width 10.4; prosoma maximum length 3.2; prosoma maximum width 4.2; leg femora: I 6.0; II 13.0; III 10.3; IV 11.0. **Dorsum** (Fig. 4A). Dorsal scutum type gamma triangular. Anterior margin of dorsal scutum with median frontal hump bearing six tubercles and three granules on each side. Carapace with granules sparsely distributed. Scutal areas I–IV with 13, 16, eight and six granules, respectively; scutal area III with one pair of paramedian spines; scutal area IV completely divided (from area III). Lateral margin of dorsal scutum mostly covered by granules (from the posterior half of carapace to posterior margin of dorsal scutum), with one large tubercle near scutal area IV. Posterior margin of dorsal scutum and free tergite I each with few granules on the corners. **Chelicerae.** Segment I with one seta on mesal side of the bulla, each finger with five or six teeth. **Pedipalps.** Coxa mostly
Figure 4. *Sadocus dilatatus* Roewer. Male holotype (SMF 886) A habitus, dorsal view B–E male left trochanter–tibia IV B dorsal view C prolateral view D ventral view E retro-lateral view. Scale bars: 1 mm.
smooth, with one ventro-central tubercle. Trochanter dorsal face smooth, with one retro-ventral apical tubercle and one retro-apical spine. Femur dorsal face smooth, ventral row of few granules and one retro-ventral spine. Patella smooth. Tibial setation: prolateral III/Iii/Ili; retro-lateral Ili/Ili/Ili. Tarsal setation: prolateral Iii/Iii/Ili; retro-lateral Iii/Iii/Ili. Legs (Fig. 4B–E). Coxa IV with one long, oblique, bifid prodorsal apical apophysis and one retro-ventral apical spine. Trochanters I–III granulate; trochanter III with one retro-apical spine. Trochanter IV with sparse granules, the prodorsal apical apophysis long, (ca. ⅓ of podomere length), pointing prolaterad. Femur III with proventral and retro-ventral rows of granules increasing in size apically, becoming tubercles. Femur IV with three blunt dorsal spines on basal ⅓; proventral row of pointed granules increasing in size apically becoming spines; retro-lateral row with two spines and one central apophysis and one proventral basal spine. Patella IV with ventral row of tubercles. Tibia IV with three retro-ventral apical spines (apical one the largest). Tarsal counts: 6, 11, 7, 10. Penis (Fig. 11G, H). Ventral plate of penis with attenuated cleft on anterior margin; three pairs of MS A, one pair of MS B, four pairs of MS C, one pair of MS D, and two pairs of ventral MS E.

**Coloration.** Immersed in ethanol: carapace and leg IV dark brown; legs I–III, pedipalps and chelicerae light brown. Specimen color badly preserved. Live specimens (Fig. 9D): carapace, coxa, and trochanter black, dry-mark on carapace; femora I–III, patellae–tibiae I–IV brown; border of dorsal scutum and free tergite green; free tergite yellowish.

**Variations** (n = 4). Scutal areas I–IV with 10–13, 14–16, 8–10, 5–6 granules, respectively. In smaller males, the lateral margin of dorsal scutum may bear cluster of tubercles instead of a large tubercle. Measurements. Dorsal scutum maximum length 7.0–8.1; dorsal scutum maximum width 8.5–11.5; prosoma maximum length 3.0–3.4; prosoma maximum width 4.1–4.4; leg femora: I 5.5–6.5; II 11.4–14.5; III 9.0–11.5; IV 9.7–12.0.

**Female** (MZSP – 8022). Measurements. Dorsal scutum maximum length 7.5; dorsal scutum maximum width 8.9; prosoma maximum length 3.4; prosoma maximum width 4.5; leg femora: I 5.0; II 11.4; III 9.0; IV 11.0. Dorsum (Fig. 8C, D). Lateral margin of dorsal scutum with five slightly large granules. Scutal areas I–IV with ten, twelve, eleven, and seven granules, respectively. Legs. Coxa IV with moderate prodorsal spiniform apophysis (as long as the podomere width, but shorter than male) and one retro-ventral apical spine shorter than on the male. Trochanter IV with a retro-apical apophysis. Femur III with proventral and retro-ventral row of granules increasing in size apically, becoming tubercles; femur IV with prolateral row of pointed granules on distal half and a retro-lateral row of pointed granules on basal half. Tarsal counts: 6, 11, 7, 9.

**Type locality.** Chile, Región de Biobío, Provincia Concepción.

**Geographical distribution** (Fig. 2). Chile. Región de Biobío, Provincia Concepción, Quebrada Pinares.

**Note.** The allotype MZSP 7874 was not used for the variation or in the distribution maps because it is a female, which lacks the diagnostic characters of the species (solely based on male characters).
Erratum. H. Soares (1968) incorrectly cited the collection number 7676 for the vial labelled: CHILE, Región de Biobío, Provincia Concepción, Quebrada Pinares, 4.XI.1964, T. Cekalovic coll., 1 ma. It is in fact 7876. The vial with number 7875 mentions 2 ma, when in fact, it includes 2 ma and 1 fe, and vial 7874 mentions only 1 ma, when it is in fact 1 fe.

**Sadocus funestus** (Butler, 1874)
Figures 2, 5A–E, 8E, F, 9F, 11I–J, 12D–F

*Gonyleptes funestis* Butler, 1874: 153, figs 5, 5a [desc]; Loman 1899: 6, fig 3 [cit]; Sørensen 1902: 20 [cit]. (Type material NHM, ma holotype, examined by detailed photographs).

*Discocyrtus funestus*: Loman, 1899: 6, fig 3 [syst]; Sørensen 1902: 20 [syst].

*Lycomedes funestus*: Sørensen, 1902: 20 [syst]; Roewer 1913: 127, 130, fig 58 [rdesc].

*Lycomedicus funestus*: Roewer, 1923: 443, fig 557 [rdesc]; 1929: 214 [cat]; Canals 1936: 69 [cat]; Soares and Soares 1954: 271 [cat]; Muñoz-Cuevas 1973: 232, fig 4 [cit]; Cekalovic 1968: 7 [cat]; 1985: 18 [cat]; Cokendolpher 1993: 136 [eco].

*Sadocus funestis*: Kury, 2003: 191 [cat]; Pérez-González et al. 2020: 3 [cit].

*Sadocus funestus*: Kury et al., 2020b [cat].

**Material examined.** Chile, Región de Los Ríos, Provincia de Valdivia, Corral, date or collector unknown, 1 ma (SMF 766); Same, Valdivia, date or collector unknown, 1 ma (SMF 276); Same, Curicué, 12.I.2006, Elizabeth Arias et al. coll., 1 ma, 2 fe, 3 juvs. (AMNH); Same, Oncol Park (-39.70025, -73.0), 12.I.2006, Elizabeth Arias et al. coll., 2 ma, 3 fe (AMNH); Same, 7.II.2004, T. Cekalovic coll., 2 ma (CAS 9026265); Same, 1 ma (AMNH); Same, 10.I.2006, Elizabeth Arias et al. coll., 1 ma, 2 fe (AMNH); Same, 7.II.2004, T. Cekalovic coll., 4 ma, 1 fe (AMNH); Same, (-39.712333, -73.307278), 01.I.2017, A. Anker & P.H. Martins coll., 1 ma (UFMG 22650); Same, 1 ma (UFMG 22651); Same, 2 ma (UFMG 22652); Same, 2 ma (UFMG 22653); Same, on the way to Oncol Park, 15.II.2004, T. Cekalovic coll., 1 ma (AMNH); Same, Chiguayco, 1.I.1980, collector unknown, 1 ma (AMNH); Same, Las Lajas, Las Tablas, 9.I.1989, L.S. Kimsey coll., 1 ma, 3 fe (MCZ 31262); Región de Araucanía, Villarrica (-39.03721, -72.121), 1–30.I.1965, L. Peña coll., 1 ma (MCZ 38272); ECUADOR, Canelos [doubtful record], date or collector unknown, 1 ma (SMF 777).

**Diagnosis.** *Sadocus funestus* can be distinguished from other species of the genus by the following characters: uniramous prodorsal apical apophysis on coxa IV; leg IV straight; and trochanter IV with four apophyses.

**Redescription.** **Male** (CAS 9026265). **Measurements.** Dorsal scutum maximum length 9.3; dorsal scutum maximum width 10.0; prosoma maximum length 3.9; prosoma maximum width 5.0; leg femora: I 4.5; II 8.3; III 7.0; IV 8.0. **Dorsum** (Fig. 5A). Dorsal scutum type gamma pyriform. Dorsal scutum anterior margin with nine granules, lateral margins with a row of granules and cluster of granules near scutal groove I.
Figure 5. *Sadocus funestus* Butler. Male (CAS 9026265) A habitus, dorsal view B–E male right trochanter–tibia IV B dorsal view C prolateral view D ventral view E retro-lateral view. Scale bar: 1 mm.
Ocularium with one pair of granules on anterior face. Scutal areas I–IV with six (three on each side), three, one, and six granules, respectively; scutal areas I–III each with one pair of paramedian tubercles. Scutal area IV completely divided. Lateral margin of dorsal scutum with row of granules between ozopore area and anterior part of scutal area IV. Posterior margin of dorsal scutum and free tergites I–III each with a row of granules. Chelicerae. Segment I with setae on bulla; fixed finger with four teeth, movable finger with three teeth. Pedipalps. Coxa with one ventro-apical spine. Trochanter dorsal face smooth, with one pair of gminated ventro-apical setiferous tubercles. Femur with a row of ventro-basal granules. Patella smooth. Tibial setation: prolateral Iii/Ii; retro-lateral IIiIi/IiIi. Tarsal setation: prolateral IIiIi/IiIi; retro-lateral IIIIIi/iIIiIIi. Legs (Fig. 5B–E). Coxa IV with one long, oblique, uniramous prodorsal apical apophysis, without retro-ventral apical spine. Trochanters I and III granulate; trochanter III with one retro-apical spine. Trochanter IV proapical spiniform apophysis with large base (ca. ⅓ of podomere length); additionally with two retro-dorsal apical apophyses with the ridges touching each other; retro-lateral face with two central granules and one apical spine. Femur III dorsal face granulate; ventral face with two rows of granules increasing in size apically. Femur IV with prodorsal row of large tubercles decreasing in size apically; prolateral row of tubercles; one retro-ventral row of granules; proventral row of granules increasing in size apically; one proventral apical spiniform apophysis, one retro-sub-apical spiniform apophysis, and two or three retro-ventral apical apophyses. Patella IV with five retro-ventral spines. Tibia III with ventral row of tubercles. Femur IV with retro-lateral row of spines and four ventro-apical spines. Tarsal counts: 6, 12, 7, 8. Penis (Fig. 11I, J). Ventral plate of penis with deeper (than moderate) cleft on anterior margin, three pairs of MS A, four pairs of MS C, one pair of MS D, without MS B or E.

Coloration. Immersed in ethanol: carapace, trochanter, and leg IV brown; legs I–III, pedipalps fading from brown to yellowish. Tubercles and spines of dorsal scutum and free tergites yellowish. Live specimens (Fig. 9F): carapace, coxa, and trochanter black, dry-mark on dorsal scutum; femora I–III, patellae–tibiae I–IV brown; free tergite green.

Variations (n = 21). Granules between lateral margin of carapace and ocularium varying from none to 4–19. Color of granules on carapace range from brown to yellowish. Measurements. Dorsal scutum maximum length 8.3–10.0; dorsal scutum maximum width 8.6–10.5; prosoma maximum length 3.4–4.0; prosoma maximum width 4.7–5.5; leg femora: I 4.5–5.0; II 8.3–9.4; III 7.0–7.9; IV 7.5–8.5.

Female (AMNH CHILE, Región de Los Rios, Provincia Valdivia, Parque Oncol, 10.I.2006, Elizabeth Arias et al. coll., 1 ma, 2 fe). Measurements. Dorsal scutum maximum length 8.4; dorsal scutum maximum width 10.0; prosoma maximum length 3.4; prosoma maximum width 5.0; leg femora: I 5.0; II 9.5; III 7.8; IV 9.7. Dorsum (Fig. 8E, F). Lateral margin of dorsal scutum with two rows of tubercles. Legs. Coxa IV with short prodorsal apical apophysis, seen in a ventral view. Trochanter IV prolateral and dorsal faces unarmed, with one retro-lateral spiniform apophysis. Femora III–IV with a prolateral and a retro-lateral rows of tubercles, increasing in size apically. Tarsal counts: 5, 11, 7, 8.
**Variations** (n = 8). Dorsal scutum with 10–24 granules, the armature on the free tergites varying from one paramedian pair of blunt to pointed tubercles. Measurements. Dorsal scutum maximum length 5.5–8.3; dorsal scutum maximum width 8.7–10.0; prosoma maximum length 3.4–3.5; prosoma maximum width 4.5–5.0; leg femora: I 4.8–5.0; II 9.0–9.5; III 7.5–7.8; IV 9.3–9.7.

**Geographical distribution** (Fig. 2). Chile, Región de Los Ríos, Provincia de Valdivia, Curinanco & Chiguayco. The record for Ecuador is doubtful (Kury 2003).

**Sadocus ingens** (Mello-Leitão, 1937)
Figures 2, 6A–E, 8G, H, 10A, B, 11C, D, 12G–I

*Carampangue ingens* Mello-Leitão, 1937: 152–153, figs 14–15 [desc]; B. Soares 1945: 370 [cat]; Mello-Leitão 1949: 17 [syst]; Soares and Soares 1954: 242 [cat]; Cekalovic 1968: 7 [cat]; 1985: 17 [cat] (type MNRJ 5263, 1 ma, 1 fe, syntype, previously preserved in dry collection, examined).

**Sadocus ingens**: Kury, 2003; 191 [cat]; Kury et al. 2020b [cat].

**Jighas vastus** Roewer, 1943: 28, pl. 3, fig 22 [desc]; Mello-Leitão 1949: 17 [syst]; Acosta 1996: 217 [cat]. (type SMF RII 1380/73, 2 ma, 2 fe, syntypes, examined). Synonymy established by Mello-Leitão 1949.

**Material examined.** Chile, Región de Araucanía, *Provincia de Malleco*, Contulmo Natural Monument (-38.012778, -73.187500), 12.XII.2010, F. Marques, F. Cadiz & F. Carbayo coll., 1 ma, 1 fe (MZSP 36965); Same, Purén, Salto el Rayen (-38.013528, -73.162639), 04.I.2017, A. Anker & P.H. Martins coll., 1 ma (UFMG 22654); Same, 1 ma (UFMG 22655); Same, *Provincia de Cautín*, Temuco, 1943, collector unknown, 2 ma, 2 fe (SMF 1380/73); Same, Región de Biobío, *Provincia de Arauco*, Carampangue, 1937, W. Feed coll., 1 ma 1 fe (MNRJ 5263 – Syntypes).

**Diagnosis.** *Sadocus ingens* can be distinguished from the other species of the genus by being the largest among them (and quite large among gonyleptid harvestmen); by the prodorsal apical apophysis on trochanter IV of the same length as the podomere (in other *Sadocus* species, that apophysis length is up to ½ the podomere length); lateral margin of dorsal scutum smooth posterior to scutal area II.

**Redescription. Male** (MZSP 36965). **Measurements.** Dorsal scutum maximum length 12.0; dorsal scutum maximum width 15.5; prosoma maximum length 4.5; prosoma maximum width 7.2; leg femora: I 8.7; II 17.4; III 11.7; IV 19.4. **Dorsum** (Fig. 6A). Dorsal scutum type gamma pyriform. Anterior margin of dorsal scutum with 13 granules, lateral margin of dorsal scutum with three to nine granules on carapace, seven granules behind ocularium. Scutal areas I–IV with four or five, nine, six, and four granules, respectively; scutal area III with one paramedian pair of spines. Scutal area IV completely divided by fading scutal groove IV. Lateral margin of dorsal scutum and free tergites I and II with three or four, eight and two granules, respectively. Free tergite III smooth. **Chelicerae.** Segment I with one probasal spine on bulla, one retro- and
Figure 6. *Sadocus ingens* (Mello-Leitão). Male (MZSP 36965) A habitus, dorsal view B–E male right trochanter–tibia IV B dorsal view C prolateral view D ventral view E retro-lateral view. Scale bars: 1 mm.
one prolateral pair of filiform spines; segment II, fixed finger with four teeth, movable finger with three teeth. **Pedipalps.** Coxa smooth and barely visible. Trochanter ventral face with one retro-apical tubercle and one prolateral spine. Femur dorsal face granulate, one ventro-basal tubercle, retro-ventral row of granules and a ventral row of tubercles. Patella smooth. Tibial setation: prolateral II/II/III; retro-lateral II/III. **Tarsal setation:** prolateral II/III; retro-lateral III/III. **Legs** (Fig. 6B–E). Coxa IV covered by setae, with one robust, long, bifid prodorsal apical apophysis and one short retro-ventral spiniform apophysis. Trochanters I–III granulate; trochanters I and III dorsal face smooth. Trochanter IV with few granules on ventral central and apical areas; one retro-lateral tubercle; the prodorsal apical apophysis long, curved, as long as the podomere length. Femur IV with retro-lateral row of spines (five prominent) decreasing in size apically, becoming blunt tubercles; retro-ventral row of granules with one basal pointed tubercle, few tubercles on the middle ½ and two spines on apical area; ventro-apical face with one retro-lateral spine and one pointed, prolateral tubercle. Patella IV with three or four retro-ventral spines. Tibia IV dorsal face granulate and with retro-ventral row of granules increasing in size apically, becoming spines on distal half. Tarsal counts: 8, 16, 8, 8. **Penis** (Fig. 11C, D). Ventral plate of penis with moderate cleft on anterior margin, two or three pairs of MS A, four or five pairs of MS C and one pair of MS D.

**Coloration.** Immersed in ethanol: carapace, trochanters, femora, patella IV, and tibia IV dark brown. Scutal areas II and III, free tergites, patellae and tibiae I–III orange. Live specimens (Fig. 10A, B): carapace, scutal areas and legs I–IV black; lateral margin of dorsal scutum dark brown with green pleurites; posterior margin of dorsal scutum and free tergites orange, arthrodial membranes green.

**Variations** (n = 6) – Free tergites II and III with one paramedian pair of spines which length varies from similar to slightly longer than the tergite length, its apex varying from blunt to pointed; femur IV with granules in between the retro-lateral spines. Measurements. Dorsal scutum maximum length 12.0–13.8; dorsal scutum maximum width 12.7–16.5; prosoma maximum length 4.5–5.4; prosoma maximum width 7.2–7.8; leg femora: I 8.7–10.0; II 16.8–18.5; III 11.7–15.0; IV 16.0–20.0.

**Female** (MZSP 36965). **Measurements.** Dorsal scutum maximum length 12.0; dorsal scutum maximum width 11.5; prosoma maximum length 5.0; prosoma maximum width 7.0; leg femora: I 8.4; II 16.2; III 12.0; IV 17.0. **Dorsum** (Fig. 8G, H). Scutal areas I–IV with six, six, four, and four granules, respectively. **Legs.** Coxa IV with discreet apophysis, not seen in ventral view. Tarsal counts: 8; 15; 8; 9. **Ovipositor** (Fig.12 G–I). Two main groups of lobes delimited by constriction, ovipositors peripheral setae inserted into sockets that are a mixture of dorsal and ventral sockets, left lobe with six setae and right lobe with six. Each main group of lobes divided by a fissure.

**Variations** (n = 4). Tubercle variation in the areas: I–2–8, II–4–6, III–3–6, IV–2–4. Measurements. Dorsal scutum maximum length 11.0–12.0; dorsal scutum maximum width 10.0–11.7; prosoma maximum length 4.5–5.0; prosoma maximum width 6.4–7.2; leg femora: I 7.1–8.4; II 13.8–16.2; III 11.0–12.0; IV 13.0–17.0.

**Type locality.** Chile, Región de Araucanía, Provincia de Malleco, Monumento Nacional Contulmo.
Geographical distribution (Fig. 2). CHILE. Región de Araucanía, Provincia de Cautín; Idem, Provincia de Malleco, Temuco; Región de Biobío, Provincia de Arauco.

Sadocus polyacanthus (Gervais, 1847)
Figures 2, 7A–E, 8I, J, 10C–F, 11E, F, 12J–L

Gonyleptes polyacanthus Gervais, 1847: 576 [desc]; 1849: 24, pl. 1, figs 7–7c [rdesc]; Butler 1873: 114 [cat]. (Syntypes MNHN, 1 ma, 1 fe examined by detailed photographs). Transferred to Sadocus by Sørensen 1902.

Sadocus polyacanthus: Sørensen, 1902: 14 [syst; rdesc]; Roewer 1913: 245–246, fig 101 [key, rdesc]; 1923: 493, fig 619 [rdesc]; 1930: 381 [cit]; Canals 1934: 6 [cit]; 1936: 70 [cat]; Roewer 1938: 02, 06 [cat]; Mello-Leitão 1939: 625 [cit]; Soares and Soares 1949: 212 [cat]; Ringuelet 1955b: 113 [cit]; 1957: 13, 19 [cit]; 1959: 409, figs XVIII-1, 2 [rdesc]; Cekalovic 1968: 7 [cat]; 1976: 24 [cat]; 1985: 15 [cat]; Acosta and Maury 1998: 569 [cit]; Kury 2003: 191 [cat]; Hara et al. 2012: 38–39 [syst]; Pinto-da-Rocha et al. 2012: 61 [cit]; Pinto-da-Rocha et al. 2014: 18 [syst]; Hara 2016: 106–109 [syst]; Kury et al. 2020b [cat].

Sadocus vitellinosulcatus Sørensen, 1886: 85, pl. 6, fig 7 [desc]. (Type depository unknown, fe). Synonymy with S. polyacanthus established by Sørensen 1902.

Gonyleptes platei Loman, 1899: 5, pl. 5, fig 3–3a [desc]. (Holotype ZMB 7843, 1 ma examined). Synonymy with S. polyacanthus established by Sørensen 1902.

Araucanoleptes exceptionalis Mello-Leitão, 1946: 5, fig 5 [desc]; Soares and Soares 1949: 160 [cat] (Syntypes – URMU 703, 1 ma, 3 fe examined). Transferred to Sadocus by Kury 2003. syn. nov.

Araucanoleptes exceptionalis [spelling mistake]: Cekalovic, 1968: 7 [cat]; 1985: 12 [cat].

Sadocus platei: Moritz, 1971: 207 [cat].

Sadocus conspicillatus Roewer, 1913: 251 – 253, pl. 1, fig 3 [desc]; 1923: 495 [rdesc]; Canals 1936: 70 [cat]; Soares and Soares 1949: 211 [cat]; Cekalovic 1968, 7 [cat]; 1985: 14 [cat]; Pinto-da-Rocha et al. 2012: 61 [cit]; Kury et al. 2020b [cat]. (Syntypes – SMF, 1 ma 1 fe examined). syn. nov.

Sadocus guttatus Sørensen, 1902: 15 [desc]; Roewer 1913: 248 [rdesc]; 1923: 494 [rdesc]; Canals 1936: 70 [cat]; Soares and Soares 1949: 211 [cat]; Roewer 1961: 102 [cit]; Cekalovic 1968, 7 [cat]; 1985: 14 [cat]; Kury 2003: 191 [cat]; Kury et al. 2020b [cat] (Holotype – ZMUC, ma examined). syn. nov.

Sadocus exceptionalis: Kury, 2003: 191 [cat]; Kury et al. 2020b [cat].

Material examined. CHILE, Región Metropolitana de Santiago, Santiago, date or collector unknown, 4 ma, 2 fe (SMF 1384); Región de Valparaíso, 13.II.89, collector unknown, 1 ma (ZMUC – holotype of S. guttatus); Región de Biobío, Provincia de Arauco, Parque Nacional Nahuelbuta (-37.805567, -73.03505), 30XI.2003, I. Avila, S. Ocares & D. Silva coll. (CAS 9055055); Same (-37.806433, -73.036333), date or collector unknown (CAS 9055038); Same (-37.810556, -73.0575), 13.XII.2010, F.
Marques, F. Cádiz & F. Carbayo coll., 4 ma, 4 fe (MZSP 36964); Same (-37.8275, -73.009722), 9.XII.2010, F. Marques, F. Cádiz & F. Carbayo coll., 3 ma (MZSP 36838); Same, Provincia Concepción, date or collector unknown, 2 ma (SMF 906; holotype of S. conspicillatus); Región de la Araucanía, Provincia de Valdivia, Las Ochocientas (-39.1668, -71.98755), 14.XII.2003, E. Arias & D. Silva et al. coll. (CAS 9055047); Same, Provincia de Malleco, Lonquimay, 23–24.XII.1976, collector unknown, 1 ma, 1 fe (AMNH); Same (Las Raíces), collector unknown, 1 ma (AMNH); Same, Con- tulmo Natural Monument, 23.I.1985, N.I. Platnick & O.F. Francke coll., 3 ma, 1 fe (AMNH); Same (-40.73715, -72.31062), 12.XI.2014, G. Giribet, G. Hormiga & A. Pérez-González coll., 2 ma, 3 fe (MCZ 138065); Same, Parque Nacional de Tolu- huca (-38.226111, -71.730833), 18.I.2008, R. Pinto-da-Rocha coll., 4 ma, 3 fe, 1 juv (MZSP 29076); Same, 12.XII.2010, F. Marques, F. Cádiz & F. Carbayo coll., 5 ma, 2 fe (MZSP 36966); Curacatín, Salto del Indio no Rio Cautín (-38.46125, -71.741333), 28.XII.2016, A. Anker & P.H. Martins coll., 1 ma, 1 fe (UFMG 22632); Purén, Salto el Rayen (-38.013528, -73.162639), 04.I.2017, A. Anker & P.H. Martins coll., 1 ma (UFMG 22637); Same, 1 fe (UFMG 22640); Same, 1 ma (UFMG 22641); Same, 1 ma (UFMG 22642); Same, Provincia de Cautín, Angol, Parque Nacional Nahuelbuta (-37.829000, -73.007278), 25–27.XII.2016, A. Anker & P.H. Martins coll., 1 ma, 1 fe (UFMG 22628); Same, 1 fe (UFMG 22629); Same, 1 ma (UFMG 22630); Same, 1 fe (UFMG 22631); Same, Fundo de las Selvas, 16–20.I.1981, L.E. Peña coll., 3 ma (AMNH); Same, Vulcán Villarrica, 15–29.XII.1982, A. Newton & M. Thayer coll., 1 ma (AMNH); Same, Villarrica (-39.03 -72.121), 16–31.XII.1982, L. Peña coll., 1 ma, 1 fe (MCZ 38270); Same, 1–30.I.1965, 4 ma, 3 fe (MCZ 38271); Same, 5 ma, 4 fe (MCZ 38273); Same, 4 ma (MCZ 38272); Same, 2 ma (MCZ 31240); Same, Lago Conguil- lio, Parque Nacional Conguillío (-38.647778, -71.610278), 24–25.I.2010, R. Pinto-da-Rocha, F. Cádiz L. & D. Cádiz L. coll., 8 ma, 1 fe (MZSP 36799); Same, 2 ma, 1 fe (MZSP 36803); Same, Puesco, Parque Nacional Villarica (-39.565583, -72.175694), 22.I.2010, R. Pinto-da-Rocha, F. Cádiz L. & D. Cádiz L. coll., 9 ma, 6 fe (MZSP 36802); Región de Los Lagos, Provincia de Osorno, Pucatrihue, 23.III.1963, Tha & Lep coll., (AMNH); Same, Parque Nacional Puyehue, 12–22.II.1979, L.E. Peña coll., 2 ma, 4 fe (AMNH); Same, 5–12.II.1978, collector unknown, 1 ma (AMNH); Same (-40.666389, -72.171944), 15.I.2003, S.E. Lew & K.W. Will coll., 1 ma, 1 fe (CAS 9017709); Same, 31.I.1985, N.I. Platnick & O.F. Francke coll., 2 ma, 1 fe, 1 juv (AMNH); Same, Puyehue, 5–12.II.1978, collector unknown, 1 ma (AMNH); Same (-40.73715 -72.31062), 16.XI.2014, G. Giribet, G. Hormiga & A. Pérez-González coll., 2 ma, 2 fe (MCZ 138128); Same, Puyehue, Parque Nacional Puyehue (-40.665583, -72.175694), 30–31.XII.2016, A. Anker & P.H. Martins coll., 1 ma (UFMG 22633); Same, La Picada, 15–20.I.1980, L.E. Peña coll., 1 ma (AMNH); Same, Entre Lagos, 14–17.II.1978, collector unknown, 1 ma (AMNH); Same, 14–17.II.1978, collector unknown, 1 ma (AMNH); Same, Provincia de Chiloé, Ilha Chiloé, Chepu, 2.II.1985, N.I. Platnick & O.F. Francke coll., 1 ma (AMNH); Same, 20.II.2000, T. Cekalovic coll., 2 ma (AMNH); Same, Chepu, 14.II.2002, T. Cekalovic coll., 1 ma (AMNH); Same, Parque Nacional Chiloé (-42.574417, -74.077350), 3.XII.2009, H. Wood, L. Almeida & C. Griswold coll., 1 ma (CAS 9036299); Same, Termas de Puyehue
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Diagnosis. Sadocus polyacanthus resembles S. dilatatus by the posterior apophysis on the lateral margin of dorsal scutum, and femur IV almost straight. It is distinguished from the latter by the lack of a long, single retro-central apophysis (present in S. dilatatus) on femur IV, and by the presence of prolareral and retro-lateral rows of similar sized tubercles on femur IV and coxa IV being closer to the body (instead of spread to the sides as S. dilatatus).

Redescription. Male (CAS 9055047). Measurements. Dorsal scutum maximum length 8.0; dorsal scutum maximum width 10.0; prosoma maximum length 3.3; prosoma maximum width 4.5; leg femora: I 4.9; II 9.6; III 8.2; IV 8.5. Dorsum (Fig. 7A). Dorsal scutum type gamma pyriform. Anterior margin of dorsal scutum with 15 granules, few closer to ocularium on carapace, lateral margin of carapace with a row of granules. Three scutal areas; scutal areas I–III with four, five, and eight granules, respectively. Scutal area III with one pair of spines. Lateral margin of dorsal scutum with row
Figure 7. *Sadocus polyacanthus* (Gervais). Male (CAS 9055047) A habitus, dorsal view B–E male right trochanter–tibia IV B dorsal view C prolateral view D ventral view E retro-lateral view. Scale bars: 1 mm.
of granules increasing in size from the ozopore area to scutal area III, ending in a large spiniform tubercle. Posterior margin of dorsal scutum with a row of ten tubercles. *Chelicerae*. Segment I with a pair of retro-apical setae; fixed finger with three teeth, movable finger with four teeth. *Pedipalps*. Coxa with one central ventral spine. Trochanter with one ventro-apical tubercle. Femur with one ventro-basal seta and three ventro-basal granules. Patella covered with setae. Tibial setation: prolateral IlI/IIii; retro-lateral: IiIlI. Tarsal setation: prolateral IiIlI; retro-lateral IlIi. *Legs* (Fig. 7B–E). Trochanters I–IV granulate; trochanter II with one retro-lateral tubercle; trochanter III with one retro-apical spine; trochanter IV with one dorso-central tubercle, one retro-central tubercle, one retro-apical tubercle, one retro-dorsal apical spine; prodorsal apical apophysis with a rounded apex extending towards the dorsal region. Femur III ventral face with two rows of granules increasing in size apically. Femur IV with retro-dorsal row of spines decreasing in size towards the center; retro-lateral row of sparse spines increasing in size apically; ventro-apical face with a prolateral spine and retro-lateral granule. Patella IV with retro-lateral row of spines, the apical one curved, pointing towards tibia IV. Tibia IV with two retro-ventral apical spines. Tarsal counts: 6, 11, 7, 8. *Penis* (Fig. 11E, F). Ventral plate of penis with attenuated cleft on anterior margin; three pairs of MS A, two pairs of MS B, two or three pairs of MS C, two or three pairs of MS D.

**Coloration.** Immersed in ethanol: carapace, trochanters I–IV and femur IV dark brown. Legs with a gradient from brown to caramel. Live specimens (Fig. 10C–F): carapace, patellae, and tibiae I–IV black, except the areas of dorsal scutum that can vary (yellow, orange or red). Coxa and trochanters black. Femora I–IV varying between black and orange. Posterior margin of dorsal scutum to free tergite III arthrodial membranes green.

**Variations** (*n* = 58). Even in ethanol there is a great variation of coloration, which ranges from orange to caramel and yellow. The number of granules on the anterior margin of dorsal scutum varying from few sparsely distributed to completely covered. Measurements. Dorsal scutum maximum length 5.6–9.8; dorsal scutum maximum width 6.0–11.9; prosoma maximum length 2.2–4.0; prosoma maximum width 3.0–5.4; leg femora: I 3.5–6.9; II 6.4–13.0; III 5.3–11.4; IV 6.0–11.5.

**Female** (AMNH – 11- Chile, Region de Los Lagos, Oncol Parque, 12.1.2006, Luma Arias et al. coll., Berkeley). **Measurements.** Dorsal scutum maximum length 9.3; dorsal scutum maximum width 9.4; prosoma maximum length 3.6; prosoma maximum width 5.0; leg femora: I 6.0; II 11.5; III 9.5; IV 11.5. **Dorsum** (Fig. 8I, J). Scutal areas I and II with three and five granules, respectively. **Legs.** Coxa IV granulate, with short prodorsal apical spiniform apophysis, not visible in ventral view. Trochanter IV with one retro-lateral spiniform tubercle. Tarsal counts: 6, 11, 7, 8. **Ovipositor** (Fig. 12J–L). Two main groups of lobes delimited by constriction, ovipositors peripheral setae inserted into sockets that are a mixture of dorsal and ventral sockets, left lobe with four setae and right lobe with six; each main group of lobes divided by a fissure.

**Variations** (*n* = 25). Color in dorsal scutum and femur varying between black and orange. Dorsal scutum with 10–30 granules. Measurements. Dorsal scutum maximum
Figure 8. Female specimens of Sadocus A, B S. asperatus (CAS–9055035) A dorsal view B lateral view C, D S. dilatatus (MZSP–8022) C dorsal view D lateral view E, F S. funestus (AMNH) E dorsal view F lateral view G, H S. ingens (MZSP–36965) G dorsal view H lateral view I, J S. polyacanthus (AMNH) I dorsal view J lateral view K Drawing of species inquirenda Carampangue nigronotatum extracted from Mello-Leitão 1943.
length 7.6–9.3; dorsal scutum maximum width 8.3–10.0; prosoma maximum length 3.0–3.6; prosoma maximum width 4.2–5.0; leg femora: I 4.5–6.0; II 7.0–13.0; III 7.0–11.4; IV 8.4–11.5.
Figure 10. Photographs of living specimens of Sadocus A. *S. ingens* (ma) B. *S. ingens* (fe) C. *S. polyacanthus* (ma) D. *S. polyacanthus* (fe) E. *S. polyacanthus* (ma) F. *S. polyacanthus* (fe). B, E taken by R. Pinto-da-Rocha; A, C, D, F taken by Pedro H. Martins.

**Type locality.** Of *G. polyacanthus*: Chile. Provincias del Sur. Of *S. vitellinosulcat us*: either Australasia or South America. Of *G. platei*: Chile. Región de Los Lagos. Provincia de Valdivia. Corral. Of *A. exceptionalis*: Chile. Región de Los Lagos. Osorno. Barra del Rio Bueno. Of *S. conspicillatus*: Chile. Región de Biobío. Provincia de Concepción. Concepción. Of *Sadocus guttatus*: Chile. Región de Biobío. Provincia de Arauco. Lebu.
Figure 11. Distal part of penis *Sadocus* and *Discocyrtus*, dorsal and ventral views, respectively A, B *S. asperatus*, macrosetae colorized and numbered C, D *S. ingens* E, F *S. polyacanthus* G, H *S. dilatatus* I, J *S. funestus* K, L *D. catharinensis*. Scale bar of drawings: 0.1mm.

**Geographical distribution** (Fig. 2). Chile, Región Metropolitana de Santiago; Región de Biobío; Región de Araucanía; Región de Los Lagos; Ilha Chiloé; Región de Los Ríos.

**Taxonomic notes.** After examining the holotypes of *S. conspicillatus*, *S. guttatus*, and *S. exceptionalis*, we concluded that they were males within *S. polyacanthus* size variation. The apophyses size and shape on trochanter IV and the armature of femur IV (especially the retro-dorsal and retro-lateral row of spines size pattern) of all species are the same.
Figure 12. Distal part of ovipositor of *Sadocus* A–C *S. asperatus* A upper view B villi C setae D–F *S. funestus* D upper view E lateral view F seta G–I *S. ingens* G upper view H, I setae J–L *S. polyacanthus* J upper view K lateral view L setae.
Species inquirendae

Sadocus allermayeri (Mello-Leitão, 1945)

Carampangue allermayeri Mello-Leitão, 1945: 158 [desc]; Soares and Soares 1954: 241 [cat]; Cekalovic 1968: 7 [cat]; 1985: 16 [cat] (type material MNRJ, 1 ma & 1 fe syntypes, destroyed, not examined).

Sadocus allermayeri: Kury, 2003: 191 [cat]; Kury et al. 2020b [cat].

Type locality. Chile. Región de Biobío. Concepción. Concepción.

Taxonomic notes. The type material, belonging to MNRJ, was lost in the fire that destroyed most of the arachnid collection (Kury, pers. comm.; Kury et al. 2018). The original description is poor for modern standards and it has no illustrations. However, the description allows to be diagnosed by: the presence of scutal area IV on dorsal scutum, coxa IV with one prodorsal apical bifid apophysis; trochanter IV with one retro-basal apophysis and three proapical apophyses. Sadocus asperatus, S dilatatus, and S. ingens have the scutal area IV in dorsal scutum and coxa IV with a bifid prodorsal apical apophysis; but none of them has three apical apophyses on trochanter IV. The only species with four scutal areas on the dorsal scutum and trochanter IV with three apical apophyses is S. funestus, but the prodorsal apical apophysis on coxa IV is uniramous. Assuming that the description is correct, it implies that this is a valid species that we have not yet located among the material gathered for this revision of Sadocus.

Sadocus nigronotatus (Mello-Leitão, 1943)

Figure 8K

Carampangue nigronotatum Mello-Leitão, 1943: 8, fig 7 [desc]; Soares and Soares 1954: 242 [cat]; Cekalovic 1968: 7 [cat]; 1985: 17 [cat] (type material MNRJ, 1 fe holotype lost, not examined).

Sadocus nigronotatus: Kury, 2003, 191 [cat]; Kury et al. 2020b [cat].

Type locality. Chile. Provincia de Llanquihue. Región de Los Lagos. Maullín.

Taxonomic notes. The type material, belonging to the MNRJ, is lost (for the same reason as S. allermayeri). The original description is poor for modern standards, and both description and figure are based on a female. The females of the different Sadocus species are very similar and difficult to identify unequivocally. According to the original description, S. nigronotatus resembles S. polyacanthus in the presence of a spiniform apophysis on the lateral margin of the dorsal scutum, but can be distinguished by the presence of scutal area IV.
Species removed from *Sadocus*

*Discocyrtus catharinensis* (Mello-Leitão, 1923)

Figure 11K, L

*Sadocus catharinensis* Mello-Leitão, 1923: 152 [desc]; B. Soares 1944a: 166 [syst]; 1944b: 222 [cit]; 1945: 364 [cat]; Soares and Soares 1949: 211 [cat]; H. Soares 1966: 90 [cat] (Types MNRJ 1510, syntypes examined by detailed photographs).

*Parasadocus catharinensis* Mello-Leitão, 1927: 8 [syst]; Roewer 1930: 425 [rdesc]; Mello-Leitão 1932: 329 [rdesc].

*Discocyrtus catharinensis*: Soares & Soares, 1987: 458, figs 3–6 [syst, rdesc].

*Sadocus aquifugus* Mello-Leitão, 1931a: 136, fig 8 [desc]; 1935: 106 [cit] (Types MNRJ 11390 syntypes, male genitalia examined by photographs).

*Gonyleptes pugilator* Mello-Leitão, 1932: 303, fig 163 [desc]; B. Soares 1944b: 222 [cit]; Soares and Soares 1949: 180 [cat] (Type MNRJ – lost).

*Gonyleptes acaanthopus* Mello-Leitão, 1945: 156 [cit] [nec Quoy and Gaimard 1824] – misidentification: Soares and Soares 1987.

*Lycomedicus brasiensis* Soares & Soares, 1949: 52, figs 6–8 [desc]; 1954: 271 [cat] (type material – old collection CGPC, 1 ma, 1 fe – MZSP 36165, holotype, MZSP 1029 paratype, examined). syn. nov.

*Sadocus brasiliensis*: Kury, 2003: 191 [cat]; Kury et al. 2020b [cat].

**Material examined.** Brazil, Paraná, Piraquara, Banhado, IV.1946, Godfergé coll., 1 ma holotype, 1 fe paratype (MZSP 36165); same, 1 ma paratype (MZSP 1029); Santa Catarina, Joinville, III.1947, Goffergé coll., 1 ma (MZSP 36419).

**Type locality.** *S. catharinensis* and *S. aquifugus*: BRAZIL. Santa Catarina. Joinville. Of *G. pugilator*: BRAZIL. Santa Catarina. Of *Lycomedicus brasiensis*: BRAZIL. Paraná. Piraquara: Banhado.

**Taxonomic notes.** We examined the type material of *Sadocus brasiliensis* and its external and penial morphology did not match that of other Chilean species of the genus. Based on its type locality, we examined other Brazilian Pachylinae genera and found striking similarities between *S. brasiliensis* and *D. catharinensis*. We examined detailed pictures of the type material kindly shared by Rafael N. Carvalho and additional material from the MZSP collection. Those species are the same, and we propose that *S. brasiliensis* is the junior synonym of *D. catharinensis*. Many *Discocyrtus* spp. have been transferred to revalidated genera or newly created subfamilies, such as Roewerii-inae (Carvalho and Kury 2018) or Neopachylinae (Carvalho and Kury 2021). Indeed, the penial features of *D. catharinensis* (Fig. 15K, L) resembles those of that subfamily, which is corroborated in the present analysis: *D. catharinensis* is the sister species of *R. bittencourtii*. Based on male genitalia and leg IV (see Carvalho and Kury 2018), *D. catharinensis* seems to belong to *Discocyrtanus* Roewer, 1929. However, because there is an ongoing study revising *Discocyrtus* conducted by Rafael N. Carvalho (MNRJ) and taking into account that it will have serious taxonomic consequences, we opted to propose only the synonymy of *S. brasiliensis* with *D. catharinensis*. 
**Gonyleptes horridus Kirby, 1819**

*Gonyleptes horridus* Kirby, 1819: 452, pl. 22, fig 16 [desc]; Kury 2003: 128 [cat]; Hara et al. 2012: 38–39 [syst]; Pinto-da-Rocha et al. 2012: 26, 41–45 [syst]; Pinto-da-Rocha et al. 2014: 4, 17 [syst]; Hara 2016: 106–109 [syst]. (Type holotype NHM 1863.41, ma examined by detailed photographs).

*Lycomedes calcar* Roewer, 1913: 132, fig 59 [desc]; Acosta 1996: 218 [cit] (Type holotype SMF RI, 782, ma examined). syn. nov.

*Lycomedicus calcar*: Roewer, 1923: 444, fig 558 [rdesc]; Canals 1936: 69 [cat]; Soares and Soares 1954: 271 [cat]; Cekalovic 1968: 8 [cat], 1985: 18 [cat].

**Sadocus calcar**: Kury, 2003, 191 [cat]; Kury et al. 2020b [cat].

**Taxonomic note.** The holotype of *Sadocus calcar* is in a very bad state of preservation; only part of the carapace, with the ozopores, and leg IV remain. The rest of the prosoma and all of the other legs are absent. Even in this condition, we noted that *S. calcar* lacks the large tubercles and spines on the free tergites, which are diagnostic of *Sadocus*. Furthermore, the armature of trochanter IV and the long, bifid, C-shaped dorso-basal apophysis on femur IV are strikingly similar to those of *Gonyleptes horridus*, a common species in the state of Rio de Janeiro. Therefore, we propose *S. calcar* as a junior synonym of *G. horridus*. This synonymy made us conclude that the provenance of *S. calcar* is mistaken, because *G. horridus* is endemic to the Brazilian Atlantic rainforest. It is widely known that Roewer, unfortunately, indicated wrong provenance of a few species, and this seems to be the case for this species.

**Eubalta planiceps** (Gervais, 1842)

**Remarks.** *Sadocus planiceps* (originally *Gonyleptes planiceps* Gervais, 1842) has a convoluted taxonomic history, with many previously unknown synonyms detected, which will be published elsewhere (briefly commented in Pessoa-Silva et al. 2020). We excluded it from *Sadocus* because it lacks the synapomorphies of the genus. It also lacks the diagnostic characters of the genus, such as the typical ocularium shape and type of armature, dorsal scutum shape, just to name a few. Comparing with other species of Chilean Pachyliniae, we detected striking similarities with *Eubalta meridionalis*. This synonymy did not go unnoticed by Kury et al. (2020a) in his catalogue, who also detected this in parallel with this revision. Finally, Kury et al. kindly invited us to publish this synonymy that resulted in a publication of that synonymy (Pessoa-Silva et al. 2020).

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