Introduction
Food allergy is an atopic disease, affecting children and adults around the world [1]. It develops when there is a failure on the induction or breakdown in oral tolerance. Due to the increasing incidence of food allergy, researches on preventive actions, including nutraceutical agents, are growing in the literature [2,3]. Capsaicin (8-methyl-N-vanillyl-1-6-nonenamide) is a spicy substance found in chilies (Capsicum frutescens and Capsicum annuum), with an important role in cooking. Capsaicin is a highly selective agonist for the transient receptor potential vanilloid 1 (TRPV1), a nonselective cation channel, preferably expressed by primary afferent sensory neurons of the pain pathway. This characteristic makes capsaicin potentially useful on the treatment of chronic pain [4-7]. Capsaicin creams for topical application have been commercialized to treat some inflammatory disorders. The efficacy of such an administration via has been demonstrated in studies of pain relief related to arthritis, postoperative neuralgia, diabetic neuropathy, and psoriasis [4,8]. Thus, we aimed to evaluate the effect of the topical use of capsaicin in a model of food allergy chronic characterized by a low-grade mucosa intestinal inflammation.

Material and Methods
This study was approved by the Ethical Committee for Animal Studies of Federal University of Minas Gerais (protocol CEUA/UFMG 78/2012). Female Balb-C mice aged 5 to 8 weeks with sanitary status SPF (Specific Pathogen Free) were obtained from the animal
facility of the Federal University of Minas Gerais, Brazil. The animals were kept in a controlled environment with 12 hours of light and dark cycle and temperature about 28 °C ± 2 °C, with free access to food and liquids. During the challenge period (allergen intake), animals were kept in individual cages for better control of allergen intake. Food allergy was induced according to the model described by Saldanha et al. [9]. Briefly, a solution containing 10 μg of ovalbumin (OVA) 98%, 1 mg of adjuvant (aluminum hydroxide) and 0.2 ml of 0.9% saline solution was administrated subcutaneously on the back of the animal. After 14 days, a sensitization boost containing 10 μg of OVA without adjuvant was administrated. The experimental period started 7 days after the sensitization boost, by replacing water for an OVA solution (20%). Animals were divided into the ALE group, treated topically with base cream (without capsaicin) and ALE CAPS group, which received the same base cream containing 0.075% capsaicin (v/v). Mice, without allergy induction or topic treatment were used as healthy controls.

Mice had part of their back shaved for the daily application of 100mg of the respective creams in an area of approximately 1 cm² days during 7 days, according to a previous protocol [10]. Food and liquid intakes were measured daily by the difference of the quantity offered and the rest. All animals were euthanized on the 8th experimental day and blood, and small intestine samples were withdrawn for further analyses. Blood were used for the quantification of anti OVA IgE and IgG1 antibodies concentration by ELISA kits to confirm allergy induction. The evaluation of the cellular infiltrate of the small intestine was made by the myeloperoxidase (MPO), eosinophil peroxidase (EPO) enzymes and n-acetyl glutaminidase (NAG) activities, as indirect measure of neutrophils, eosinophils, and macrophages, respectively. Histopathological analysis of the goblet cells producing mucus was performed in the small intestine by the periodic acid from Schiff (P.A.S) stain using the Image J Software (NIH, USA). PAS positive pixels were selected for the calculation of the total area mucus-producing goblet cells. The result was expressed as % of PAS positive area of intestinal mucosa/field. The data were evaluated for normal distribution by the Kolmogorov-Smirnov test. Parametric and non-parametric data were submitted to Student’s t-Test and Mann Whitney test, respectively. The results were expressed as the mean and standard error. Statistical analyzes were performed using GraphPad Prism 7.0® software (San Diego, CA, USA). The level of significance was set at p<0.05.

Results

The results of blood concentration showed that anti OVA IgE and IgG1 antibodies levels were compatible with the presence of OVA allergy. However, no difference was detected between ALE and ALE CAPS groups considering IgE or IgG1 concentrations (Figure 1). Although IgE and IgG1 levels were similar in both groups, the OVA solution intake, in turn, was higher in animals treated with topical capsaicin, showing a higher tolerance to the allergen (Figure 1). Food intake was reduced compared to healthy mice, but similar in allergic groups confirming a similar caloric intake between ALE and ALE CAPS groups. Nonetheless, ALE CAPS mice showed a better body weight evolution, with a weight loss lower than in ALE group during the experimental days (Figure 1). When the intestinal inflammatory infiltrates were analyzed, it was noted that the ALE CAPS group showed lower neutrophil and eosinophil infiltration, as suggested by the MPO and EPO activity, respectively (Figure 2). No difference was seen when the macrophage infiltration was compared (p= 0.064). ALE group exhibited hypersecretion of mucus by goblet cells. However, a smaller mucus production was observed in allergic animals treated with capsaicin when compared to allergic animals (Figure 3).

Figure 1: Evaluation of blood levels of IgE (A) and IgG1 (B), cumulative food intake (C), ovalbumin intake (D) and weight evolution (E) in ovalbumin allergic mice topically treated with capsaicin-free cream (ALE), 0.075% capsaicin cream (ALE CAPS) or in mice without allergy or capsaicin treatment (CT). The results were expressed as mean and SD. SD. N= 7/group. * Statistically significant (p <0.05).
Cite this article: Elandia A S, Bruna SL C, Lilian G Teixiera, Ester Roffê, et al. Topical Application of Capsaicin Reduces Weight Loss Allergen Aversion and Intestinal Mucosa Inflammation in A Food Allergy Experimental Model. Biomed J SciTech Res 10(5)-2018. BJSTR. MS.ID.002023. DOI: 10.26717/BJSTR.2018.10.002023.

Discussion

Our results suggest a beneficial effect of the topical application of capsaicin in reducing the manifestations of food allergy. Due to its lipophilic character, capsaicin can be absorbed from the skin, with a half-life of about 24 hours [4]. This route of administration has already been used in commercial products to help relieve a several types of minor neuralgia related to disorders such as rheumatoid arthritis or muscle sprains and strains [11, 12].

Two of the majorsigns of food allergy are weight loss and allergen aversion [13,14]. Our results show that capsaicin can reduce both since the ALE CAPS group presented higher intake of OVA solution and lower weight loss compared to ALE mice. The improvement in allergy in the capsaicin- treated group was also demonstrated by the lower recruitment of neutrophils and eosinophils to the small intestine. Neutrophils are the first cells to be recruited to the site of inflammation, being installed in the lamina propria and intestinal crypts, releasing the enzyme myeloperoxidase (MPO) among other components [15]. Eosinophils are fundamental cells in the allergic process, serving as “immunological effector sentinels”, with rapid action in inflammatory processes [13, 16]. In allergic diseases, eosinophils are activated and attracted to the intestinal mucosa, releasing the degranulation products such as toxins and free radicals,
which can trigger intestinal injury. The release of leukotrienes and cytokines by eosinophils amplifies the inflammatory response, recruiting and activating more eosinophils. The mucosa eosinophil infiltration in others intestinal disorder such as inflammatory bowel diseases is associated with unfavorable conditions such as malabsorption, weight loss and changes in mucosal architecture with decreased crypts in the colon and villous flattening [16-18].

These results suggest that topical capsaicin is acting as an anti-inflammatory agent creating a less inflamed intestinal environment. Another important feature of food allergy is the increased production of mucus by goblet cells [13]. It is known that in the late phase of food allergy, eosinophils release lipid mediators such as prostaglandin D2, which increase mucus production by intestinal goblet cells [13]. Our data suggested that the lower production of mucus by the ALE CAPS group could be the result of the lower eosinophil infiltration in this group and its lower release of lipid mediators. Our data agree with the hypothesis that topical capsaicin would act mainly in the late phase of food allergy leading to the lower infiltration of inflammatory cells. Although the anti-inflammatory action of capsaicin has been described in several in vitro studies, the mechanisms are not fully understood [4,19,20]. One of the possible mechanisms is related to the action of capsaicin as a TRPV1 ligand [21].

It has been described that for the complete activation of the TRPV1 vanilloid channel, about 10μM of capsaicin is required [22-26]. When activated, TRPV1 channels are associated with thermal hyperalgesia in the skin, visceral organs and oral cavity [21]. However, repeated exposures to capsaicin, as occurred in thermal hyperalgesia in the skin, visceral organs and oral cavity [21]. These results are in agreement with those of Lee, Shin et al. who observed lower expression of proinflammatory cytokines in obese mice topically treated with capsaicin, reinforcing the regulatory role of capsaicin on inflammation [10]. Although all parameters demonstrated improvement of food allergy symptoms with topical use of capsaicin, IgE and IgG1 concentrations were similar between groups. However, due to the 3 and 23-day half-lives of these immunoglobulins, respectively [31], the treatment time (7 days) may not have been sufficient to detect significant changes in their plasma levels. Alternatively, if capsaicin somehow interferes with IgE binding with its FcεRI receptor rather than reducing IgE production, no difference in the circulating levels of that immunoglobulin would be detected.

Conclusion
Our data suggest that the topical application of capsaicin is associated to positive effects in food allergy related to reduction of major manifestations, such as allergen aversion and body weight loss.

Acknowledgment
This study was supported by Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG), Coordenação de Aperfeiçoamento de Pessoal de Nível Superior (CAPES), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) and by Pro-reitoria de Pesquisa (PRPq) of Universidade Federal Minas Gerais. The authors thank Maria Helena Alves de Oliveira and for the support in animal facility.

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