Kynurenine pathway is altered in patients with SLE and associated with severe fatigue

Karolina Åkesson,1 Susanne Pettersson,2,3 Sara Ståhl,4 Izabella Surowiec,5 Mattias Hedenström,6 Susanna Eketjäll,7 Johan Trygg,5 Per-Johan Jakobsson,1 Iva Gunnarsson,1 Elisabet Svenungsson,1 Helena Idborg1

ABSTRACT

Objective Fatigue has been reported as the most disturbing symptom in a majority of patients with SLE. Depression is common and often severe. Together these symptoms cause significant morbidity and affect patients with otherwise relatively mild disease. Tryptophan and its metabolites in the kynurenine pathway are known to be important in several psychiatric conditions, for example, depression, which are often also associated with fatigue. We therefore investigated the kynurenine pathway in patients with SLE and controls.

Methods In a cross-sectional design plasma samples from 132 well-characterised patients with SLE and 30 age-matched and gender-matched population-based controls were analysed by liquid chromatography tandem mass spectrometry to measure the levels of tryptophan and its metabolites kynurenine and quinolinic acid. Fatigue was measured with Fatigue Severity Scale and depression with Hospital Anxiety and Depression Scale. SLE disease activity was assessed with Systemic Lupus Erythematosus Disease Activity Index (SLEDAI).

Results The kynurenine/tryptophan ratio, as a measure of indoleamine 2,3-dioxygenase (IDO) activity, was increased in patients with SLE. Patients with active disease (SLEDAI ≥6) showed lower tryptophan levels compared with controls (54µM, SD=19 vs 62µM, SD=14, p=0.03), although patients with SLE overall did not differ compared with controls. Patients with SLE had higher levels of tryptophan metabolites kynurenine (966nM, SD=530) and quinolinic acid (546nM, SD=480) compared with controls (kynurenine: 712nM, SD=230, p=0.0001; quinolinic acid: 380nM, SD=150, p=0.001). Kynurenine, quinolinic acid and the kynurenine/tryptophan ratio correlated weakly with severe fatigue (r=0.34, r=0.28 and r=0.24, respectively) but not with depression.

Conclusions Metabolites in the kynurenine pathway are altered in patients with SLE compared with controls. Interestingly, fatigue correlated weakly with measures of enhanced tryptophan metabolism, while depression did not. Drugs targeting enzymes in the kynurenine pathway, for example, IDO inhibitors or niacin (B12) supplementation, which suppresses IDO activity, merit further investigation as treatments in SLE.

INTRODUCTION

Depression and cognitive dysfunction are the most common psychiatric manifestations in SLE. Depression is present in 11%-39% of the patients,1 and many patients with SLE also suffer from anxiety.2 3 Fatigue has been reported to be present in up to 90% of women with SLE4 and when given open questions, a majority reported fatigue as the most troublesome SLE-related symptom.5 6 Fatigue is sometimes also known to be associated with symptoms of depression, but the underlying mechanisms are unknown.6 7 A large fraction of patients with SLE experience fatigue in the absence of psychiatric disorders. Possible connections between fatigue and SLE disease activity have been studied but with contradictory results.8

The kynurenine pathway is known to be upregulated as part of an activated immune response,9 and recently the tryptophan metabolism via the kynurenine pathway has been highlighted as a mechanism of central fatigue.10 11 The kynurenine pathway has an established role in depression where low plasma levels of tryptophan and elevated levels of metabolites, that is, increased kynurenine/tryptophan ratio, have been detected in the circulation and in the cerebrospinal fluid (CSF).12 In the kynurenine pathway (figure 1), two enzymes, tryptophan 2,3-dioxygenase (TDO) and indoleamine 2,3-dioxygenases (IDO), are responsible for the first step where tryptophan is converted to kynurenine. Kynurenine can further be converted to picolinic acid or quinolinic acid, and quinolinic acid to nicotinic acid.13 IDO is mainly present in immune cells and known to be induced by interferon gamma (IFN-γ), interleukin (IL-2), IL-1β, prostaglandin E2, oxidative stress and LPS.14 TDO is mainly expressed in the liver and regulated by the levels of tryptophan and by glucocorticoids.15 16 Higher kynurenine/tryptophan ratios, as a measure of IDO activity, have been reported in patients with SLE.17–19

1Rheumatology Unit, Department of Medicine, Karolinska Institutet, Karolinska University Hospital, Stockholm, Sweden
2Theme Inflammation and Infection, Karolinska University Hospital, Stockholm, Sweden
3Department of Neurobiology, Care Sciences and Society, Karolinska Institutet, Stockholm, Sweden
4Personalized Healthcare and Biomarkers Laboratories, Innovative Medicines, Science for Life Laboratory, Translational Science Centre, AstraZeneca, Stockholm, Sweden
5Department of Chemistry, Computational Life Science Cluster, Umeå University, Umeå, Sweden
6Cardiovascular and Metabolic Diseases, Innovative Medicines and Early Development Biotech Unit, AstraZeneca, Integrated Cardio Metabolic Centre (ICMC), Karolinska Institutet, Huddinge, Sweden

Correspondence to Dr Helena Idborg; Helena.idborg@ki.se
In this study, we investigated levels of tryptophan and its metabolites in the kynurenine pathway in patients with SLE and in controls. Among patients with SLE, associations with fatigue, depression and disease activity were evaluated.

MATERIAL AND METHODS

Study design and cohort

EDTA plasma and urine samples from a cross-sectional SLE cohort at Karolinska University Hospital, Stockholm, Sweden, were used in this study. All patients with SLE were adults (>18 years old) and fulfilled four of the American College of Rheumatology (ACR) 1982 revised classification criteria for SLE.20 Samples were selected to be representative for the Karolinska SLE cohort as previously described,21 and in addition, samples from a pilot study were included.22 In total, 132 samples and 32 age-matched and gender-matched population controls were selected for analysis of the kynurenine pathway.

Clinical assessments

To assess anxiety and depression, the participants answered the questionnaire Hospital Anxiety and Depression Scale (HADS).23 24 HADS consists of 14 items; each item is weighted from 0 to 3, giving a possible sum ranging from 0 to 21 for the anxiety scale and depression scale, respectively.23 The participants also filled in the Fatigue Severity Scale (FSS).25 26 FSS is a nine-item scale scored from one to seven; the mean value of the answered questions results in the final score (range 1–7). The cut-off for severe fatigue have been suggested to be ≥4,27 and mean score for healthy adults varies between 2.3 (SD=0.7) and 3.0 (SD=1.08).25 28 In this study, we denote a score of 1–3 as ‘no/limited fatigue’ and a score of ≥4 as ‘severe fatigue’.
of ≥4 as ‘severe fatigue’. Disease activity at sampling was defined by Systemic Lupus Erythematosus Disease Activity Index (SLEDAI) and Systemic Lupus Activity Measures (SLAM). Active disease was defined as SLEDAI ≥6 or SLAM ≥7. The Systemic Lupus International Collaborative Clinics (SLICC)/ACR damage index was used to assess organ damage.

**Analysis of tryptophan, kynurenine and quinolinic acid by liquid chromatography tandem mass spectrometry**

An aliquot of 25 µL EDTA plasma was mixed with an internal standard solution containing 13C3, [15N3]-quinolinic acid, D3-kynurenine, D3-tryptophan and D3-kynurenic acid and filtered before injection into a HPLC system. Detection was performed using a triple quadrupole mass spectrometer targeting tryptophan, kynurenine, quinolinic acid and the corresponding internal standards as previously described. Quantification was performed using internal standard and Masslynx 4.1 software (Waters Corporation). Two controls failed in the analysis, and thus this study is based on a subcohort of 132 patients with SLE and 30 controls.

**Analysis of tryptophan and picolinic acid by gas chromatography mass spectrometry (GC-MS)**

Plasma EDTA samples from patients with SLE (n=114) and controls (n=23) from our cohort were analysed by GC-MS for metabolomics profiling. Samples were randomly selected to batches, and three out of four batches were successfully analysed. Data regarding tryptophan and picolinic acid from these 87 patients and 18 controls were used in this work.

**Analysis of tryptophan, quinolinic acid and trigonelline by nuclear magnetic resonance (NMR) spectroscopy**

Urine samples from 58 patients with SLE and 12 controls were analysed for metabolite content by NMR in another project. The samples to be analysed by NMR in that project were selected based on the results from the GC-MS analysis described above. Samples in batches 1 and 3 from the GC-MS analysis were analysed by NMR since these two batches showed the largest difference in metabolic profile when comparing SLE and control. For each sample, 250 µL urine was mixed with 250 µL phosphate buffered saline (PBS) (pH 7.4) containing 3 mM Na2, and 5.5 mM trimethylsilyl propanoic acid (for chemical shift and line-shape calibration) and then transferred to 5 mm NMR tubes. The analysis was performed on a Bruker 600MHz Avance III HD spectrometer equipped with a BBO cryoprobe with Z-gradients and a SampleJet sample changer (Bruker Biospin, Rheinstetten, Germany). The samples were kept at +6°C prior to analysis, and the temperature during the experiment was +27°C. A 1D NOESY presaturation experiment (pulse sequence noesypppr1d) with 64 scans and a relaxation delay (d1) of 4s was performed on each sample. Metabolite identification and quantification was performed using Chenomx NMR suite 8.1 (www.chenomx.com), and for each sample, the quantified metabolite concentrations were normalised to concentration of creatinine. Two samples (one control and one SLE) were excluded from further analysis due to significantly different NMR spectra as compared with rest of the samples.

**Analysis of cytokines by mesoscale**

Cytokines were analysed in EDTA plasma samples from the SLE cohort using the MSD V-PLEX Human Cytokine 30-plex kit (K15054D; Mesoscale Discovery, Gaithersburg, Maryland, USA) and cytokines, which previously have been suggested to be increased in patients with SLE, that is, tumour necrosis factor alpha (TNF-α), IFN-γ-induced protein 10 (IP-10), interleukin (IL)-16, macrophage inflammatory proteins 1 beta (MIP-1β), and IL-6 were studied in this work.

**Statistics**

Unpaired t-test with Welch correction or non-parametric Mann-Whitney U test were performed for normally and non-normally distributed data, respectively. Differences were considered significant at p<0.05. The Spearman’s rank-order correlation coefficient (r) was used to measure the strength of the linear association between two variables. No adjustment for multiple testing has been applied in this work, but we consider comparisons with p<0.001 with higher confidence.

**RESULTS**

**The kynurenine pathway is altered in patients with SLE**

Characteristics of the investigated patients with SLE and controls are reported in table 1. Comparing patients with controls, we obtained significantly higher levels of kynurenine, quinolinic acid, as well as the kynurenine/tryptophan ratio in plasma from patients with SLE, while levels of tryptophan did not differ (table 2). Higher levels of tryptophan were detected in patients with active or inactive disease, compared with active disease (SLEDAI ≥6 vs control: p=0.03; SLAM ≥7 vs control: p=0.04). Both patients with active or inactive disease, according to SLEDAI, had higher levels of kynurenine, quinolinic acid and higher kynurenine/tryptophan ratio compared with controls (table 2). Higher levels of kynurenine and kynurenine/tryptophan ratio were also seen when active disease was defined by SLAM. However, none of the metabolites correlated (r<0.2) with SLAM or SLEDAI, neither when the whole SLE group nor when active disease only was considered.

In the SLE cohort (n=132), 80 patients were treated with prednisolone. Higher levels of tryptophan were detected in patients with prednisolone treatment compared with patients without prednisolone (60 µM, SD=25 vs 52 µM, SD=19, p=0.03), although glucocorticoids are known to stimulate IDO, that is increase tryptophan consumption (figure 1). Lower levels of tryptophan were detected in patients without prednisolone compared with controls (52 µM, SD=19 vs 62 µM, SD=14, p=0.006).
difference in tryptophan between patients treated with prednisolone compared with controls. The kynurenine/tryptophan ratio was not significantly different comparing patients with prednisolone (0.018, SD=0.010, n=80) to patients without prednisolone (0.022, SD=0.019, n=52).

**Fatigue and depression**

In the SLE cohort, 100 of the 132 patients (76%) as well as 10 of the 30 controls (33%) were defined as having severe fatigue. There was a significant difference in quinolinic acid, kynurenine, tryptophan levels and kynurenine/tryptophan ratio when comparing patients with severe fatigue with controls, but no difference was observed when comparing SLE patients with no/limited fatigue with either controls or SLE patients with severe fatigue (table 2).

Fatigue showed an overall weak correlation to quinolinic acid, kynurenine and kynurenine/tryptophan ratio in the SLE cohort ($r=0.21$, $r=0.22$ and $r=0.25$, respectively), but a stronger correlation to these metabolites was observed within a selection of patients that where considered to have severe fatigue ($r=0.28$, $r=0.34$ and $r=0.25$, respectively) (table 3). Index variables with connection to depression and fatigue were investigated (SLAM headache, SLAM fatigue, SLAM cortical dysfunction, SLICC neuropsychiatric, HADS anxiety and HADS depression), but no correlation was found in relation to the metabolites in the kynurenine pathway except weak correlations between kynurenine and SLICC neuropsychiatric damage ($r=0.18$, $p=0.03$) and between tryptophan and SLAM cortical dysfunction ($r=0.20$, $p=0.02$). HADS anxiety was reported for 61 of 129 patients with SLE and 11 of 30 controls, and HADS depression was reported for 42 of 129 patients and 5 of 30 controls. Additionally, SLEDAI did not correlate with fatigue, and the metabolite levels did not correlate with fatigue among the controls.

**Correlation of cytokines to metabolites in the kynurenine pathway**

Proinflammatory cytokines associated with SLE and disease activity, that is, IP-10, TNF-α and IL-16, correlated with kynurenine, quinolinic acid and kynurenine/tryptophan ratio, but not with tryptophan, in our study (table 4). MIP-1β correlated weakly with kynurenine/tryptophan ratio ($r=0.24$) and IFN-γ correlated only with quinolinic acid levels ($r=0.29$).

**Complementary measurements of the metabolites in the kynurenine pathway**

In the GC-MS study, picolinic acid (figure 1) was detected, but patients with SLE and controls did not differ. Tryptophan was detected at lower levels in patients compared with controls in the combined metabolic profile, but the difference reached significance ($p=0.02$) only in one of the analysed batches of samples. NMR data of urine samples showed significantly ($p=0.0041$) higher levels of trigonelline (figure 1) in patients with SLE (88.1, SD=85, n=58) compared with

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**Table 1 Characteristics of patients with SLE and controls**

| Parameter                  | SLE cohort n=132 | Controls n=30 | No/limited fatigue* | Severe fatigue* | No/limited versus severe fatigue* |
|----------------------------|------------------|---------------|---------------------|-----------------|----------------------------------|
| Age†                       | 48±15 (35–57)    | 47±16 (34–57) | 43±14 (31–55)       | 49±15 (36–58)   | 0.04                             |
| SLEDAI total‡              | 4 (2–8)          | –             | 2 (2–7)             | 4 (2–8)         | 0.76                             |
| SLEDAI ≥6                  | n=50             | –             | n=9                 | n=40            |                                  |
| SLAM total‡                | 7 (5–11)         | –             | 5 (2–8)             | 7 (6–12)        | 0.001                            |
| SLAM≥7                     | n=78             | –             | n=13                | n=63            |                                  |
| FSS‡                       | 5.3 (4.0–6.3)    | 3.1 (2.0–4.3) | 2.6 (2.2–3.1)       | 5.8 (5.0–6.6)   | <0.0001                          |
| HADS anxiety‡              | 6 (3–11)         | 5 (3–10)      | 4 (2–8)             | 7 (4–11)        | 0.04                             |
| HADS depression‡           | 4 (2–8)          | 2 (0–4)       | 2 (0–5)             | 5 (3–9)         | <0.0001                          |
| Serum creatinine†          | 76±31            | 68±9          | 75±30               | 76±32           | 0.93                             |
| Treatment, n (%)           |                  |               |                     |                 |                                  |
| Prednisolone dose >0 mg    | 80 (61)          | 1             | 15 (52)             | n=63 (63)       |                                  |
| Prednisolone dose >7.5 mg§ | 32 (24)          | 0             | 6 (21%)             | n=26 (26)       |                                  |
| Hydroxychloroquine         | 23 (17)          | 0             | 4 (14)              | 19 (19)         |                                  |
| No treatment¶              | 39 (30)          | 29 (97)       | 11 (38)             | 28 (28)         |                                  |

*No/limited fatigue: SLE patients with FSS <4; severe fatigue: SLE patients with FSS=4–7.
†Mean±SD (25% quantile–75% quantile).
‡Median (25% quantile–75% quantile).
§Low disease activity (LLDAS) dose of prednisolone set to >7.5 mg as suggested by Franklyn et al.48
¶No treatment implies that patients were not treated with immunosuppressants, hydroxychloroquine or corticosteroids.
FSS, Fatigue Severity Scale; HADS, Hospital Anxiety and Depression Scale; LLDAS, lupus low disease activity state; SLEDAI, Systemic Lupus Erythematosus Disease Activity Index; SLAM, Systemic Lupus Activity Measures.
In this study, we report higher circulating levels of the tryptophan metabolite kynurenine and an increased kynurenine/tryptophan ratio in patients with SLE compared with controls. These findings confirm previous studies reporting changes in the kynurenine pathway in SLE.\textsuperscript{17–19} We found that levels of tryptophan were numerically lower in SLE, as measured by the LC-MS method, but the difference was not significant, and we can thus not confirm previous studies that reported lower levels of tryptophan in SLE compared with controls. However, using the GC-MS approach we observed lower levels of tryptophan in patients for the combined metabolic profile but did not reach significance for all individual sample batches. When the patients were stratified for disease activity, as measured by two validated indices, we observed lower levels of tryptophan in patients with active SLE compared with controls. In patients with high disease activity, dietary factors might contribute to lower levels of tryptophan. However, the kynurenine/tryptophan ratio is not affected by the diet and therefore a better measure of IDO enzyme activity.

### Table 2: Metabolites in kynurenine pathway (LC-MS/MS data) in SLE, controls and SLE subgroups

|                  | Tryptophan (μM)* | Kynurenine (nM)* | Quinolinic acid (nM)* | Kynurenine/tryptophan* | Quinolinic acid/kynurenine* |
|------------------|------------------|------------------|-----------------------|------------------------|-----------------------------|
| SLE cohort (n=132) | 56.55±23         | 965.9±530        | 545.8±480             | 0.019±0.015            | 0.57±0.30                   |
| Controls (n=30)   | 61.83±14         | 712.0±230        | 379.5±150             | 0.012±0.0048           | 0.54±0.15                   |
| SLE versus controls | P=0.10           | P=0.0001         | P=0.001               | P<0.0001               | P=0.44                      |
| SLEDAI <6 (n=82)  | 58.34±25         | 977.8±590        | 511.6±450             | 0.019±0.016            | 0.54±0.27                   |
| SLEDAI ≥6 (n=50)  | 53.60±19         | 946.3±440        | 602.0±540             | 0.020±0.012            | 0.63±0.35                   |
| Active SLEDAI† versus control | P=0.03 | P=0.003 | P=0.007 | P=0.0002 | P=0.14                      |
| Inactive SLEDAI versus control | P=0.35 | P=0.0008 | P=0.02 | P=0.0002 | P=0.93                      |
| Active versus inactive SLEDAI | P=0.22 | P=0.72 | P=0.32 | P=0.97 | P=0.14                      |
| SLAM <7 (n=54)    | 58.86±28         | 909.9±570        | 498.4±490             | 0.018±0.015            | 0.54±0.26                   |
| SLAM ≥7 (n=78)    | 54.95±19         | 1005±510         | 578.6±480             | 0.020±0.014            | 0.60±0.33                   |
| Active SLAM‡ versus control | P=0.04 | P<0.0001 | P=0.001 | P<0.0001 | P=0.28                      |
| Inactive SLAM versus control | P=0.51 | P=0.03 | P=0.10 | P=0.006 | P=0.98                      |
| Active versus inactive SLAM | P=0.37 | P=0.33 | P=0.35 | P=0.36 | P=0.32                      |
| Severe fatigue (n=100)§ | 54.17±20 | 958.7±510 | 538.6±440 | 0.020±0.014 | 0.59±0.33 |
| No/limited fatigue (n=29) | 64.17±31 | 939.1±630 | 527.9±610 | 0.017±0.016 | 0.52±0.22 |
| Severe fatigue versus control | P=0.02 | P=0.0003 | P=0.002 | P<0.0001 | P=0.32                      |
| No/limited fatigue versus control | P=0.71 | P=0.07 | P=0.20 | P=0.12 | P=0.62                      |
| Severe versus no/limited fatigue | P=0.11 | P=0.88 | P=0.93 | P=0.34 | P=0.21                      |

*Mean±SD. P values obtained using Mann-Whitney U test.
†Active disease=SLEDAI ≥6.
‡Active disease=SLAM ≥7.
§No/limited fatigue were defined as FSS <4 and severe fatigue as FSS 4–7.
FSS, Fatigue Severity Scale; SLAM, Systemic Lupus Activity Measures; SLEDAI, Systemic Lupus Erythematosus Disease Activity Index.

### Table 3: Spearman rank correlation between metabolites in the kynurenine pathway and FSS in the entire SLE cohort and in the selection of patients with severe fatigue

|                              | SLE patients (FSS=1–7) | SLE patients with severe fatigue (FSS 4–7) |
|------------------------------|-------------------------|------------------------------------------|
|                              | n=129                   | n=100                                    |
| Tryptophan                   | −0.034 (p=0.71)         | 0.10 (p=0.34)                            |
| Kynurenine                   | 0.22 (p=0.01)           | 0.34 (p=0.0006)                          |
| Quinolinic acid              | 0.21 (p=0.02)           | 0.28 (p=0.005)                           |
| Kynurenine/tryptophan        | 0.25 (p=0.005)          | 0.24 (p=0.02)                            |
| Quinolinic acid/kynurenine   | −0.0046 (p=0.96)        | −0.056 (p=0.56)                          |

Fatigue Severity Scale (FSS) data were obtained from 129 of 132 patients with SLE. Spearman's correlation coefficients (r) above |0.2| are highlighted in bold.
We report higher levels of quinolinic acid in plasma from patients with SLE compared with controls. This finding may be related to an earlier finding of increased levels of quinolinic acid in CSF but, to our knowledge, it has not previously been reported in plasma from patients with SLE. Quinolinic acid has been shown to have a wide biological activity and is able to initiate inflammation and can generate reactive oxygen species. Quinolinic acid is a de novo precursor to nicotinic acid, which is used in the synthesis of nicotinamide adenine dinucleotide, a coenzyme catalysing redox reactions in the body. In urine, a methylated form of nicotinic acid, that is, trigonelline, can be found and a change in the excretion of trigonelline can be an indicator of niacin metabolism, a process that can be altered by tryptophan metabolism and also by dietary sources. We detected higher levels of urinary trigonelline in SLE compared with controls. We suggest that high urinary trigonelline indicates enhanced activity of the kynurenine pathway and possible shunting towards quinolinic acid. The majority of the patients were on prednisolone treatment, and it is known that glucocorticoids upregulate the expression of TDO that metabolises tryptophan, and we may thus expect increased kynurenine/tryptophan ratio due to treatment. However, there was no significant difference in kynurenine/tryptophan ratio comparing patients with glucocorticoid treatment and untreated patients. IDO, the other enzyme converting tryptophan to kynurenine, can be induced by proinflammatory stimuli such as IFN-γ, TNF-α and LPS, and we observed that the kynurenine/tryptophan ratio correlated positively to TNF-α (r = 0.48). It has been suggested that IDO-dependent immunosuppressive mechanisms are activated in SLE, and interferons and proinflammatory cytokines such as TNF-α, which are associated with SLE disease activity, may cause increased IDO enzyme activity and be part of the pathogenesis of SLE. Whether the metabolites in the kynurenic pathway have specific functions or contribute to disease pathogenesis has not been evaluated in this work. In the aim to suppress IDO activity, niacin (vitamin B3) supplementation and IDO blockers might be interesting treatment perspectives in SLE.

Depression, other cognitive issues or indexes for disease activity did not show any correlation to quinolinic acid, kynurenine, tryptophan or kynurenine/tryptophan ratio; however, a weak correlation to fatigue was observed. We suggest that these results indicate that the kynurenine pathway is mainly important in SLE-related fatigue. It has previously been suggested that activation of the IDO pathway may be relatively unique to inflammation-induced depression and has also been suggested as a link between inflammatory cytokines and cancer-related fatigue. The levels of fatigue were slightly higher among our controls than the values originally reported as normal for healthy adults, but they were comparable with controls in more recent European studies. The choice of questionnaires to capture patient-reported outcomes, such as fatigue, also influences the results. However, several instruments that measure fatigue showed reliable results when exploring clinically important difference, as defined by patients. The fatigue questionnaire used in this study, FSS, is widely used and recommended in SLE. In this study, a majority of the patients reported FSS ≥ 4. Fewer patients reported high levels of anxiety or depression (HADS ≥ 7), which indicate symptoms of mental disorder.

In conclusion, we confirm previous studies that the kynurenine pathway is altered in patients with SLE compared with controls. The metabolites in the kynurenine pathway have many different functions, and an increased activity of the pathway could be involved in the pathogenesis of SLE. The increased IDO activity could be a result of proinflammatory cytokines. We demonstrate that SLE-related fatigue, as captured by FSS, correlated weakly to the tryptophan metabolites kynurenine and quinolinic acid, while depression (HADS) did not. We believe that these observations may be of future clinical relevance, and it is relevant to investigate if drugs targeting IDO are effective treatments in SLE.

**Table 4** Spearman rank correlation between metabolites in the kynurenine pathway and selected cytokines

| Metabolite | Tryptophan (μM) | Kynurenine (nM) | Quinolinic acid (nM) | Kynurenine/tryptophan |
|------------|-----------------|----------------|---------------------|----------------------|
| IP-10      | -0.09           | 0.39           | 0.43                | 0.43                 |
| TNF-α      | -0.13           | 0.45           | 0.49                | 0.48                 |
| IL-16      | -0.08           | 0.43           | 0.40                | 0.40                 |
| IL-6       | -0.12           | 0.15           | 0.07                | 0.19                 |
| MIP-1β     | -0.17           | 0.15           | 0.07                | 0.24                 |
| IFN-γ      | 0.03            | 0.17           | 0.29                | 0.12                 |

Spearman’s correlation coefficients (r) above |0.2| are highlighted in bold.

**Acknowledgements** We would like to thank the patients and controls for participating in the study. Eva Jemseby for collecting all the blood samples and coordinating nurses Sonia Möller and Birgitta Mannerstedt Fogelfors for administration of patient questionnaires.

**Contributors** Å: statistical analysis and manuscript writing. SP: FSS data collection and analysis and manuscript writing. SS: LC-MS analysis of metabolites in the kynurenine pathway. IS: GC-MS analysis and manuscript writing. MH: NMR.
analysis and manuscript writing. SE: cytokine measurements and manuscript writing. JT and P-PJ: manuscript writing/approval. IG and ES: SLE cohort responsible and manuscript writing. HI: study design, statistical analyses and manuscript writing.

Funding This study was supported by the AstraZeneca–Karolinska Institutet Joint Research Program in Translational Science, Swedish Research Council, Stockholm County Council (ALF), Swedish Heart-Lung foundation, King Gustaf Vs 80th Birthday Fund, Swedish Rheumatism Association, Swedish Society of Medicine and Karolinska Institutet’s Foundations.

Competing interests SE is employed by AstraZeneca.

Patient consent Obtained.

Ethics approval The local research ethics committee approved the study.

Provenance and peer review Not commissioned; externally peer reviewed.

Data sharing statement No additional data are available.

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REFERENCES

1. Meszaros ZS, Perl A, Faraoe SV. Psychiatric symptoms in systemic lupus erythematosus: a systematic review. J Clin Psychiatry 2012;73:993.

2. Pettersson S, Lövgren M, Eriksson LE, et al. An exploration of patient-reported symptoms in systemic lupus erythematosus and the relationship to health-related quality of life. Scand J Rheumatol 2012;41:383–90.

3. Beckerman NL. Living with lupus: a qualitative report. Soc Work Health Care 2011;50:330–43.

4. Fonseca R, Bernardes M, Tereso G, et al. Silent burdens in disease: fatigue and depression in SLE. Autoimmune Dis 2014;1:41.

5. Pettersson S, Möller S, Svenungsson E, et al. Women’s experience of SLE-related fatigue: a focus group interview study. Rheumatology 2010;49:1935–42.

6. Myint AM, Schwarz MJ, Müller N. The role of the kynurenine metabolism in major depression. J Natl Transl Med 2012;11:245–51.

7. Pettersson S, Boström C, Eriksson K, et al. Lifestyle habits and fatigue among people with systemic lupus erythematosus and matched population controls. Lupus 2015;24:955–65.

8. Tench CM, McCurrie I, White PD, et al. The prevalence and associations of fatigue in systemic lupus erythematosus. Rheumatology 2000;39:1249–54.

9. Chen Y, Guillenin GJ. Kynurenine pathway metabolites in humans: disease and healthy States. Int J Tryptophan Res 2009;2:JTR. S2097–19.

10. Yamashita M, Yamamoto T. Excessive brain tryptophan and kynurenic acid lead to abnormal social behavior in central fatigue induced by chronic sleep disorder. International Journal of Psychophysiology 2014;94:229–30.

11. Yamashita M, Yamamoto T. Tryptophan circuit in fatigue: from blood to brain and cognition. Brain Res 2017;1656:16–29.

12. Kolodziej LR, Paleolog EM, Williams RO. Kynurenic metabolism in health and disease. Amino Acids 2011;41:1173–83.

13. Henderson LM, Niacon, Ann Rev Nutr 1983;3:289–307.

14. Leonard B, Maes M. Mechanistic explanations how cell-mediated immune activation, inflammation and oxidative and nitrosative stress pathways and their sequels and concomitants play a role in the pathophysiology of unipolar depression. Neurosci Biobehav Rev 2013;36:746–85.

15. Schmidt SK, Müller A, Hesseler K, et al. Antimicrobial and immunoregulatory properties of human tryptophan 2,3-dioxygenase. Eur J Immunol 2009;39:2755–64.

16. Maes M, Leonard BE, Myint AM, et al. The new ‘5-HT’ hypothesis of depression: cell-mediated immune activation induces indoleamine 2,3-dioxygenase, which leads to lower plasma tryptophan and an increased synthesis of detrimental tryptophan catabolites (TRYCATs), both of which contribute to the onset of depression. Prog Neuropsychopharmacol Biol Psychiatry 2011;35:702–21.

17. Xiang ZY, Tang AG, Ren YP, et al. Simultaneous determination of serum tryptophan metabolites in patients with systemic lupus erythematosus by high performance liquid chromatography with fluorescence detection. Clin Chem Lab Med 2010;48:513–7.

18. Widner B, Sepp N, Kowald E, et al. Enhanced tryptophan degradation in systemic lupus erythematosus. Immunobiology 2000;201:621–30.

19. Pertovaara M, Hasan T, Raitala A, et al. Indoleamine 2,3-dioxygenase activity is increased in patients with systemic lupus erythematosus and predicts disease activation in the sunny season. Clin Exp Immunol 2007;150:274–8.

20. Tan EM, Cohen AS, Fries JF, et al. The 1982 revised criteria for the classification of systemic lupus erythematosus. Arthritis Rheum 1982;25:1277–7.

21. Surowiec I, Johansson E, Torell F, et al. Multivariate strategy for the sample selection and integration of multi-batch data in metabolomics. Metabolomics 2017;13:114.

22. Idborg H, Olynyk J, Rannar S, et al. SAT0181 Systems biology of SLE. Biochemical characterization of subgroups within SLE for improved diagnosis and treatment. Ann Rheum Dis 2013;72(Suppl 3):S323–S32.

23. Bjelland I, Dahl AA, Haug TT, et al. The validity of the Hospital Anxiety and Depression Scale. An updated literature review. J Psychosom Res 2002;52:69–77.

24. Lisspers J, Nygren A, Söderman E. Hospital Anxiety and Depression Scale (HADS): some psychometric data for a Swedish sample. Acta Psychiatr Scand 1997;96:281–6.

25. Krupp LB, LaRocca NG, Muir-Nash J, et al. The fatigue severity scale. Application to patients with multiple sclerosis and systemic lupus erythematosus. Arch Neurol 1989;46:1121–3.

26. Mattsson M, Möller B, Lundberg I, et al. Reliability and validity of the fatigue severity scale in Swedish patients with systemic lupus erythematosus. Scand J Rheumatol 2008;37:269–77.

27. Zonana-Nacach A, Rosman JM, McGWin G, et al. Systemic lupus erythematosus in three ethnic groups. VI: Factors associated with fatigue within 5 years of criteria diagnosis. LUMINA Study Group, LUPus in Minority populations: NATure vs Nurture. Lupus 2000;9:101–9.

28. Vaiko PO, Bassetti CL, Bloch KE, et al. Validation of the fatigue severity scale in a Swiss cohort. Sleep 2008;31:1601–7.

29. Bondardier C, Gladman DD, Urowitz MB, et al. Derivation of the sLED. A disease activity index for lupus patients. Arthritis & Rheumatism 1992;35:630–40.

30. Lattanzi B, Consolo A, Solari N, et al. Measures of disease activity and damage in pediatric systemic lupus erythematosus: British Isles Lupus Assessment Group (BILAG), Physician’s Global Assessment of Disease Activity (MD (SLAM), Systemic Lupus Erythematosus Disease Activity Index (SLEDAI), Physician’s Global Assessment of Disease Activity (MD Global), and Systemic Lupus International Collaborating Clinics/ American College of Rheumatology Damage Index (SLICC/ACR DI; SDI), Arthritis Care Res 2011;63(S11):S112–S17.

31. Mikdashi J, Nived O. Measuring disease activity in adults with systemic lupus erythematosus: the challenges of administrative burden and responsiveness to patient concerns in clinical research. Arthritis Res Ther 2015;17:183.

32. Aĕnehands S, Brenner P, StähI S, et al. Cerebrospinal fluid kynurenines in multiple sclerosis: relation to disease course and neurocognitive symptoms. Brain Behav Immun 2016;51:47–55.

33. Ekertal S, Idborg H, Gustafsson JT, et al. TNF-a and plasma albumin as biomarkers of disease activity in systemic lupus erythematosus. Lupus Science & Medicine 2018;0–0.

34. Elia G. Interferon-γ-inducible chemokines in systemic lupus erythematosus. Clin Ter 2015;166:116–20.

35. Jacob N, Stoh W. Cytokine disturbances in systemic lupus erythematosus. Arthritis Res Ther 2011;13:228.

36. Lee S, Kanoeh K, Sekigawa I, et al. Circulating interleukin-16 in systemic lupus erythematosus. Br J Rheumatol 1998;37:1334–7.

37. Shah D, Wanchu A, Bhatnagar A. Interaction between oxidative stress and chemokines: possible pathogenic role in systemic lupus erythematosus and rheumatoid arthritis. Immunobiology 2011;216:1010–7.

38. Vogelgesang SA, Heyes MP, West SG, et al. Quinolinic acid in patients with systemic lupus erythematosus and neuropsychiatric manifestations. J Rheumatol 1996;23:850–5.

39. Stone TW, Stoy N, Darlington LG. An expanding range of targets for kynurenine metabolites of tryptophan. Trends Pharmacol Sci 2013;34:136–43.

40. Rezzi S, Ramadran Z, Martin FP, et al. Human metabolic phenotypes link directly to specific dietary preferences in healthy individuals. J Proteome Res 2007;6:4469–77.
41. Badawy AA. Tryptophan availability for kynurenine pathway metabolism across the life span: Control mechanisms and focus on aging, exercise, diet and nutritional supplements. *Neuropharmacology* 2017;112:248–63.

42. Moffett JR, Namboodiri MA. Tryptophan and the immune response. *Immunol Cell Biol* 2003;81:247–55.

43. Brochez L, Chevolet I, Kruse V. The rationale of indoleamine 2,3-dioxygenase inhibition for cancer therapy. *Eur J Cancer* 2017;76:167–82.

44. Raison CL, Dantzer R, Kelley KW et al. CSF concentrations of brain tryptophan and kynurenines during immune stimulation with IFN-alpha: relationship to CNS immune responses and depression. *Mol Psychiatry* 2010;15:393–403.

45. Kim S, Miller BJ, Stefanek ME et al. Inflammation-induced activation of the indoleamine 2,3-dioxygenase pathway: Relevance to cancer-related fatigue. 2015:2129–36.

46. Pettersson S, Lundberg IE, Liang MH et al. Determination of the minimal clinically important difference for seven measures of fatigue in Swedish patients with systemic lupus erythematosus. *Scand J Rheumatol* 2015;44:206–10.

47. Ávila-Zubieta JA, Alarcón GS, Bischoff-Ferrari HA et al. Measurement of fatigue in systemic lupus erythematosus: a systematic review. *Arthritis Rheum* 2007;57:1348–57.

48. Franklyn K, Lau CS, Navarra SV et al. Definition and initial validation of a Lupus Low Disease Activity State (LLDAS). *Ann Rheum Dis* 2016;75:1615–21.
Correction: Kynurenine pathway is altered in patients with SLE and associated with severe fatigue

Åkesson K, Pettersson S, Ståhl S, et al. Kynurenine pathway is altered in patients with SLE and associated with severe fatigue. Lupus Science & Medicine 2018;5:e000254. doi: 10.1136/lupus-2017-000254

The authors want to alert readers to the following two errors identified in the published version.

The second sentence in the Conclusions section of the Abstract should read as:

Drugs targeting enzymes in the kynurenine pathway, for example, IDO inhibitors or niacin (B3) supplementation, which suppresses IDO activity, merit further investigation as treatments in SLE.

Again, in the Discussion section, the last sentence of the second paragraph should read as:

In the aim to suppress IDO activity, niacin (vitamin B3) supplementation and IDO blockers might be interesting treatment perspectives in SLE.

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Lupus Science & Medicine 2019;6:e000254corr1. doi:10.1136/lupus-2017-000254corr1