CXCL-16, IL-17, and bone morphogenetic protein 2 (BMP-2) are associated with overweight and obesity conditions in middle-aged and elderly women

Silvana Mara Turbino Luz Ribeiro²,³,⁷, Laís Roquete Lopes²,³,⁷, Guilherme de Paula Costa²,⁷, Vivian Paulino Figueiredo²,⁷, Deena Shrestha²,⁷, Aline Priscila Batista², Roney Luiz de Carvalho Nicolato⁵, Fernando Luiz Pereira de Oliveira³,⁶, Juliana Assis Silva Gomes⁸ and Andre Talvani¹,²,³,⁴,⁷*

Abstract

Background: The current concept of overweight/obesity is most likely related to a combination of increased caloric intake and decreased energy expenditure. Widespread inflammation, associated with both conditions, appears to contribute to the development of some obesity-related comorbidities. Interventions that directly or indirectly target individuals at high risk of developing obesity have been largely proposed because of the increasing number of overweight/obese cases worldwide. The aim of the present study was to assess CXCL16, IL-17, and BMP-2 plasma factors in middle-aged and elderly women and relate them to an overweight or obese status. In total, 117 women were selected and grouped as eutrophic, overweight, and obese, according to anthropometric parameters. Analyses of anthropometric and circulating biochemical parameters were followed by plasma immunoassays for CXCL-16, IL-17, and BMP-2.

Results: Plasma mediators increased in all overweight and obese individuals, with the exception of BMP-2 in the elderly group, whereas CXCL16 levels were shown to differentiate overweight and obese individuals. Overweight and/or obese middle-aged and elderly individuals presented with high LDL, triglycerides, and glycemia levels. Anthropometric parameters indicating increased-cardiovascular risk were positively correlated with CXCL-16, BMP-2, and IL-17 levels in overweight and obese middle-aged and elderly individuals.

Conclusion: This study provides evidence that CXCL-16, IL-17, and BMP-2 are potential plasma indicators of inflammatory status in middle-aged and elderly women; therefore, further investigation of obesity-related comorbidities is recommended. CXCL16, in particular, could be a potential marker for middle-aged and elderly individuals transitioning from eutrophic to overweight body types, which represents an asymptomatic and dangerous condition.

Keywords: Inflammation, Elderly, Overweight, Obesity, CXCL-16, BMP-2, IL-17

* Correspondence: talvani@nupeb.ufop.br
† Equal contributors
¹Department of Biological Sciences, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil
²Post-graduation Program in Biological Sciences/NUPEB, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil
Full list of author information is available at the end of the article

© The Author(s). 2017 Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.
Background

Obesity is a worldwide epidemic and pathological condition resulting from contemporary lifestyles; it is characterized by lack of physical activity in addition to an energy-dense diet [1]. Adipose tissue was previously defined as a simple energy source used to manage energy flow in the body; however, it has taken on the integrative role of linking bioactive mediator release, homeostasis, and pathological conditions [2]. At least 24 typical inflammatory mediators such as leptin, resistin, IL-6, TNF, and chemokines, among others (e.g., C-reactive protein, haptoglobin, and amyloid A) are upregulated during obesity [3–5].

Natural ageing is currently associated with senescent immune remodeling, which occurs when innate barriers lose their abilities to restrict pathogen entry; in this condition, the internal organs become more vulnerable to metabolic disturbances resulting in a natural reduction in life expectancy [6, 7]. It is currently accepted that increased visceral omental, not abdominal subcutaneous adipose tissue, correlates the most with cardiovascular diseases, insulin resistance, and other obesity-related diseases [8, 9]. Overweight and obesity statuses often start early in child- or adulthood. These conditions become detrimental during senescence, when the human body requires balance to deal with age-associated changes. Accordingly, lifestyle and nutritional changes in infants and adults might prevent or postpone some adverse effects on the immune system and improve quality of life in elderly individuals. Both the inflammatory context and biological aging are crucial interrelated components of the understanding the association of clinical disturbances with overweight and obesity.

Based on the many potential biomarkers that have the capacity to predict obesity- and overweight-related diseases, we investigated the clinical role of three soluble mediators in human cardiac and metabolic diseases: (i) IL-17, a pro-inflammatory cytokine previously associated with obesity due to its role in the development of atherosclerosis, coronary syndromes, and glucose tolerance [10, 11], (ii) CXCL16, an interferon-gamma-regulated chemokine and scavenger receptor for oxidized low-density lipoprotein that is expressed in atherosclerotic lesions and described as a marker of acute coronary syndromes [12, 13], and finally, (iii) bone morphogenetic protein-2/BMP-2, a cytokine that promotes bone formation, but also is associated with potential adverse clinical effects including cyst-like bone formation, soft tissue swelling, and other anabolic activities in normal and osteoarthritic chondrocytes [14, 15]. Thus, the aim of the present study was to investigate the association between CXCL-16, BMP-2, and IL-17 plasma levels and overweight and obese conditions in middle-aged (35 to 55 years of age) and elderly (>60 years old) women to find possible targets to monitor early progression to obesity.

Methods

Study population

We performed this study using 46 healthy individuals (23 middle-aged and 23 elderly), 33 overweight individuals (20 middle-aged and 13 elderly), and 38 obese individuals (23 middle-aged and 15 elderly). All study participants were of the female sex and recruited at the UNIMED Inconfidentes, Ouro Preto, MG, Brazil. These women were undergoing a complete clinical examination and the following laboratory workup was performed: full blood count, free T4, TSH, glucose, cholesterol, VLDL, LDL, HDL, and triglycerides. Individuals with hypertension, diabetes, thyroid or renal disturbances, or any other cardiac or systemic diseases as well as those using steroidal drugs were excluded from this study. These conditions could prevent adequate interpretation of the association between obesity-related diseases and immune parameters. After blood collection and full clinical evaluation, all individuals were monitored weekly by health professionals (medical doctors, nutritionists, physiotherapist, and others) from UNIMED-Inconfidentes.

Definition of anthropometric measures

All study participants had body mass, triceps skinfold, and arm, waist, and hip circumference measurements collected by a trained staff member, in accordance with standardized protocols [16, 17]. Body mass index was calculated as weight divided by squared height (kg/m²). The triceps skinfold was verified on the mid-line of the posterior surface of the arm (over the triceps muscle) at the level of the mid-point between the acromiale and the radiale. The subscapular skinfold was measured just below the angle of the scapula. Waist circumference was also measured at the midpoint between the lowest rib and the iliac crest.

Arm muscle circumference was the chosen indicator for elderly individuals, as the Third National Health and Nutrition Examination Survey (NHANES III) has presented reference data for this indicator [18]. All measurements were taken in a private room and conducted by an experienced nutritionist.

Biochemical analysis

Blood samples (7 ml) were drawn using S-Monovette tubes (Sarstedt Ltda, Nümbrecht Germany) between 7:00 and 9:00 AM, from all study participants after 12–14 h of overnight fasting. All blood tests (cholesterol, VLDL, LDL, HDL, triglycerides, and glycemia) were performed using conventional enzymatic methods at the certified Clinical Analysis Laboratory from the Pharmacy...
School at Universidade Federal de Ouro Preto, MG, Brazil, on the day of blood collection.

**Immunoassays for inflammatory mediators**

Circulating levels of the inflammatory mediators, IL-17, CXCL16, and BMP-2, were detected in plasma samples that were previously stored at ~80°C. Briefly, flat-bottom 96-well microtiter plates (Nunc) were coated with 100 μl/well of appropriate monoclonal antibodies for 18 h at 4°C and then washed with PBS buffer (pH 7.4) containing 0.05% tween-20 (wash buffer). Non-specific binding sites were blocked with 200 μl/well of 1% bovine serum albumin in PBS. Plates were washed and 100 μl/well of sample was added, which was followed by incubation for 18 h at 4°C. In addition, 100 μl/well of the appropriate biotinylated detection antibody, diluted in blocking buffer containing 0.05% tween-20, was added and samples were incubated for 1 h at 24°C. Streptavidin-horseradish peroxidase was added and after subsequent incubation and washing, 100 μl/well of the chromogen substrate o-phenylenediamine (Sigma-Aldrich Brasil Ltda, SP, Brazil), diluted in 0.03 M citrate buffer containing 0.02% of H2O2 was added for 30 min, which was followed by incubation in the dark at room temperature. The reaction was stopped through the addition of 50 μl/well of 1 M H2SO4 solution and plates were read at 492 nm in a spectrophotometer (Emax_Molecular Devices, Sunnyvale, CA, USA). All samples were simultaneously measured using Peprotech ELISA kits (Ribeirão Preto, SP, Brazil) in triplicate.

**Statistical analysis**

Graph Pad InStat and Prism statistical programs were used for analysis (GraphPad, San Diego, CA, USA). In cases of normal distribution, the independent *t* test was used, whereas the non-parametric Mann-Whitney test was used for non-Gaussian sample distributions. Correlations were verified using a Spearman test and results were confirmed by performing a linear regression test. Data were considered statistically significant when *p* < 0.05.

**Results**

**Anthropometric characterization of subjects**

Our study population consisted of 63 middle-aged (mean age of 40.9 years) and 54 elderly (mean age of 68.9 years) women, categorized according anthropometric parameters as eutrophic, overweight, and obese (Table 1). All anthropometric parameters such as body mass, triceps skinfold, arm circumference, arm muscle circumference, and waist and hip measurements were elevated in overweight or obese middle-aged and elderly individuals when compared to eutrophic individuals. Body mass index similarly increased with weight gain in both age groups, but the waist-to-hip ratio only increased in obese middle-aged individuals, and not in overweight or obese elderly individuals (data not shown).

An apparent reduction in immune and physiological activity in the senescent body was observed under natural conditions and without the presence of disease. Overweight and obese middle-aged individuals presented with anthropometric parameters that were worse than those in elderly eutrophic individuals, based on all analyzed parameters, except for arm muscle circumference. In addition, overweight and obese conditions were associated with similar anthropometric parameters, in both groups of women, with a minimum age difference of 30 years.

**Cholesterol and glucose parameters in overweight and obese women**

Blood biochemical parameters are shown in Table 2. VLDL cholesterol and glycemia levels were higher in eutrophic elderly individuals than in eutrophic middle-aged individuals.

However, when anthropometric parameters were taken into consideration for this biochemical analysis, overweight and/or obese individuals had total cholesterol, triglycerides, and glucose levels significantly increased compared to the eutrophic group. Plasma levels of HDL cholesterol were significantly higher in the eutrophic group compared to overweight and obese groups among middle-aged women. In elderly women, similar results were observed in the comparison between the overweight group and the eutrophic group.

**Table 1** Baseline anthropometric parameters of the eutrophic, overweight and obesity subjects

| Parameter           | Eutrophic          | Overweight         | Obesity             |
|---------------------|--------------------|--------------------|---------------------|
| **Adult** (n = 23) / Elderly (n = 23) | Adult (n = 20) / Elderly (n = 13) | Adult (n = 23) / Elderly (n = 15) |
| Age                | 40.0 ± 1.2 / 71.5 ± 1.4 | 40.1 ± 1.5 / 64.0 ± 2.0 | 42.6 ± 1.5 / 71.4 ± 2.1 |
| Body mass (kg)     | 57.3 ± 1.6 / 59.1 ± 1.6 | 68.8 ± 1.0* / 70.7 ± 2.1* | 86.8 ± 2.6*# / 80.3 ± 1.6*A# |
| BMI (Kg/m2)        | 21.8 ± 0.4 / 23.6 ± 0.4 | 27.2 ± 0.2* / 28.26 ± 0.3* | 34.0 ± 0.6*# / 33.2 ± 0.5*# |
| TSF (mm)           | 17.0 ± 1.1 / 18.4 ± 1.4 | 25.8 ± 1.3* / 24.0 ± 1.4 | 32.2 ± 1.5*# / 28.3 ± 1.5* |
| AC (cm)            | 27.1 ± 0.5 / 29.2 ± 0.5 | 31.3 ± 0.4* / 33.2 ± 0.6* | 36.4 ± 0.7*# / 35.5 ± 0.6* |
| AMC (cm)           | 21.7 ± 0.5 / 23.4 ± 0.3 | 23.2 ± 0.4 / 25.7 ± 0.5*A | 26.0 ± 0.5*# / 26.8 ± 0.5* |
| Waist (cm)         | 82.1 ± 1.6 / 87.1 ± 1.9 | 91.5 ± 1.5* / 101.1 ± 2.2*A | 107.7 ± 2.4*# / 102.9 ± 1.7* |

Values are shown as the mean ± SEM. BMI body mass index, TSF Triceps skinfold, AC arm circumference, AMC arm muscle circumference. *p < 0.01 when eutrophic groups were compared with their respective chronological overweight (OW) or obesity (OB) groups; *p < 0.05 when adult and elderly were compared among in terms of eutrophic, OW or OB; # P < 0.01 when OW were compared with OB inside the same chronological groups.
VDLL cholesterol, and glycemia levels that were higher than those in the respective age-matched eutrophic control groups. When we compared overweight and obese statuses, obese middle-aged individuals had higher total cholesterol, VLDL, LDL, HDL, triglycerides, and glycemia levels, whereas only HDL and triglycerides were different in the obese elderly group.

Interestingly, soluble CXCL16, but not IL-17 and BMP-2, was positively correlated with glycemia in middle-aged (r = 0.734, p < 0.01) and elderly (r = 0.604, p < 0.05) individuals, as well as with VLDL cholesterol in middle-aged (r = 0.623, p < 0.05) and elderly (r = 0.546, p < 0.05) individuals.

Table 2 Biochemical parameters in eutrophic, overweight and obesity women

|                  | Eutrophic | Overweight | Obesity  |
|------------------|-----------|------------|---------|
| Adult (n = 23) / | Adult (n = 20) / | Adult (n = 23) / |           |
| Elderly (n = 23) | Elderly (n = 13) | Elderly (n = 15) |       |
| Cholesterol (mg/dL) 157.8 ± 6.5 / 211.0 ± 12.0 | 194.0 ± 9.3 / 193.8 ± 13.0 | 217.0 ± 8.4 / 195.9 ± 11.0 |
| VLDL (mg/dL) 13.4 ± 1.0 / 18.8 ± 2.1 | 17.9 ± 2.8 / 27.05 ± 1.8 | 22.0 ± 2.3 / 28.1 ± 2.9 |
| LDL (mg/dL) 125.9 ± 6.1 / 133.0 ± 11.0 | 119.9 ± 8.5 / 121.3 ± 12.1 | 143.2 ± 7.7 / 125.8 ± 11.6 |
| HDL (mg/dL) 58.3 ± 1.8 / 55.7 ± 2.3 | 53.6 ± 2.7 / 54.7 ± 3.2 | 51.5 ± 2.2 / 46.5 ± 2.1 |
| Triglycerides (mg/dL) 67.5 ± 5.1 / 86.9 ± 10.6 | 89.5 ± 14.0 / 85.6 ± 9.1 | 111.4 ± 11.5 / 115.1 ± 10.9 |
| Glycemia (mg/dL) 91.0 ± 1.9 / 103.2 ± 9.3 | 91.3 ± 1.8 / 119.6 ± 8.8 | 104.4 ± 4.7 / 117.5 ± 11.9 |

Values are shown as the mean ± SEM. VLDL very low density lipoproteins, LDL low density lipoproteins, HDL High density lipoproteins. Statistical differences are represented being that (*) p < 0.05 when the OW or OB were compared with the eutrophic individuals in the same age group; (Δ) p < 0.05 when adult and elderly were compared in the same anthropometric group, (∞) when OB is different from OW in the same age group.

Plasma indicators of overweight and obesity
Plasma levels of the inflammatory chemokine CXCL16 (Fig. 1a) were markedly elevated in overweight and obese middle-aged and elderly individuals when compared to that in eutrophic individuals. In addition, CXCL16 was capable of distinguishing overweight and obese individuals in the two age-related groups. CXCL16 production in overweight or obese middle-aged individuals was higher than that observed in 60-year old eutrophic individuals.

IL-17 (Fig. 1b) production was high in all overweight and obese individuals. However, obese elderly individuals presented with higher cytokine levels than overweight individuals of the same age group. In terms of IL-17 production, there were no differences between overweight and obese middle-aged and eutrophic elderly individuals.

BMP-2 (Fig. 1c) was higher in all overweight and obese middle-aged individuals, similar to that observed with the IL-17 pattern, but there was no difference observed between eutrophic and overweight elderly individuals. Increased production of this factor was observed in eutrophic and obese elderly individuals, suggesting natural degeneration of the bones.

Correlations among CXCL16, IL-17, and BMP-2 and anthropometric parameters
We assumed that body mass index, triceps skinfold, arm muscle circumference, and hip and waist circumferences are variables used to predict obesity-related comorbidities. The correlations among these anthropometric...
parameters and plasma CXCL16, BMP-2, and IL-17 levels are shown in Table 3. Accordingly, CXCL16 showed the highest correlation with body mass index, triceps skinfold, and waist and hip circumference in middle-aged and elderly individuals, with Pearson’s correlation coefficients ($r$) > 0.5.

Finally, IL-17 plasma levels were also positively correlated with body mass index, arm muscle circumference, and hip and waist circumferences in both age-related groups, whereas BMP-2 levels were only correlated with these anthropometric parameters in the elderly group.

**Discussion**

Results presented herein highlight the potential for CXCL16, IL-17, and BMP-2 plasma levels to be used as markers to discriminate overweight and obese individuals in both age groups. In particular, production of the chemokine CXCL16 was markedly high in the overweight and obese groups, and this correlated with anthropometric parameters that are considered comorbidity predictors.

The identification of biomarkers to predict obesity has become important research area within the basic and clinical sciences, owing to the ability of these factors to predict cardiovascular and metabolic disturbances; this allows for proper treatment and lifestyle changes in individuals [19]. Unfortunately, the underlying complexity of biological pathway interactions demands further research before any of these potential biomarkers can be accurately used for diagnostic purposes. Many cellular inflammatory mediators come from white fat tissues, and they are often overproduced because of increased adiposity. In contrast, other regulatory factors or mediators of insulin sensitivity, such as adiponectin, are found in smaller amounts in similar conditions [20]. This imbalance between adipocyte products and other inflammatory mediators, caused by overweight and obese conditions might promote obesity-linked metabolic and cardiovascular disorders [1, 9]. Therefore, investigating biomarkers in overweight individuals should be a more common procedure in research and clinic. Ordinary individuals naturally gain a small amount of weight within a certain period of time; however, continuous chronic weight gain becomes problematic.

Interestingly, CXCL16 was shown to have a strong and exclusive association with overweight and obese conditions, regardless of subject age. Elderly women with higher VLDL and glycemia typically had higher levels of circulating CXCL16; these high circulating levels have previously been associated with the development of plaque deposits, artery walls, and/or diabetes [18, 20, 21]. Anthropometric parameters such as body mass index, triceps skinfold, and waist and hip circumference were positively correlated with CXCL16 levels in middle-aged and elderly women. CXCL16 is expressed in macrophages and aortic muscle cells. Expression increases in response to inflammatory stimuli and this enhances LDL uptake in favor of foam cell formation in human atherosclerotic plaques [11, 13]. Some chemokines have been described as effective adipokines and as biomarkers of obese conditions because of their inflammatory profile, which is associated with weight gain and adipose tissue accumulation, and the fact that their receptors are overexpressed in subcutaneous and visceral adipose tissues of obese individuals [22]. Previous studies have indicated an association between CXCL16 and obesity in C57BL6 mice [23] and coronary comorbidities in humans [24–26]. By assuming that obesity predisposes individuals to coronary and metabolic diseases, the present

| Table 3 Correlation among CXCL16, IL-17 and BMP-2 and anthropometric parameters |
|---------------------------------|----------|--------|--------|--------|--------|--------|
| **ADULTS**                     | BMI      | TSF    | AMC    | WAIST  | HIP    | WHR    |
| CXCL16                         | $R = 0.798^*$ | $R = 0.552^*$ | $R = 0.119$ | $R = 0.860^*$ | $R = 0.555^*$ | $R = 0.2081$ |
| p = 0.023                      | p = 0.5521 | p = 0.1981 | p = 0.016 | p = 0.050 | p = 0.1981 |
| IL-17                          | $R = 0.188^*$ | $R = 0.354$ | $R = 0.371^*$ | $R = 0.341$ | $R = 0.556^*$ | $R = 0.221^*$ |
| p = 0.047                      | p = 0.850 | p = 0.010 | p = 0.117 | p = 0.050 | p = 0.014 |
| BMP-2                          | $R = 0.1629$ | $R = 0.0935$ | $R = 0.1778$ | $R = 0.2423$ | $R = 0.1518$ | $R = 0.1778$ |
| p = 0.1034                     | p = 0.3523 | p = 0.0752 | p = 0.0146 | p = 0.1295 | p = 0.075 |
| **ELDERLY**                    | BMI      | TSF    | AMC    | WAIST  | HIP    | WHR    |
| CXCL16                         | $R = 0.798^*$ | $R = 0.552^*$ | $R = 0.198$ | $R = 0.860^*$ | $R = 0.555^*$ | $R = 0.198$ |
| p = 0.020                      | p = 0.050 | p = 0.119 | p = 0.016 | p = 0.050 | p = 0.110 |
| IL-17                          | $R = 0.184^*$ | $R = 0.332$ | $R = 0.227^*$ | $R = 0.146$ | $R = 0.542^*$ | $R = 0.371^*$ |
| p = 0.030                      | p = 0.090 | p = 0.014 | p = 0.110 | p = 0.050 | p = 0.020 |
| BMP-2                          | $R = 0.179^*$ | $R = 0.352$ | $R = 0.187^*$ | $R = 0.242^*$ | $R = 0.251$ | $R = 0.188^*$ |
| p = 0.050                      | p = 0.090 | p = 0.050 | p = 0.014 | p = 0.129 | p = 0.050 |

* BMI: body mass index, TSF: triceps skinfold, AMC: arm muscle circumference, WHR: waist-to-hip ratio
* $^*$ positive correlation with $p < 0.05$
study included overweight and obese women. Regardless of this, high plasma levels of CXCL16 were observed in all middle-aged and elderly individuals presenting with an anthropometric status above overweight.

Overweight or obese individuals were expected to have increased adipocyte numbers, resulting from the recruitment of pluripotent stem cells to the vascular stroma of adipose tissue [27]. In our study, BMP-2 was investigated and was shown to be clearly elevated in overweight and obese middle-aged and elderly individuals. Regarding anthropometric parameters, this factor was positively correlated with body mass index, arm muscle circumference, and waist and waist-to-hip ratio in elderly women. These parameters are indicative of overweight or obese conditions, reinforcing the potential role of BMP-2 to categorize individuals according to metabolic status. BMP-2 has been shown to promote osteogenic differentiation and adipogenesis in bone marrow stromal cells [28], and it could be partially responsible for increased obesity. A recent study suggested that increased demand to store excessive energy promotes BMP-2 expression and that adipocytes contribute to this phenomenon, due to energy storage in visceral and subcutaneous adipose tissues [29]. The present study reinforces such findings, since increased BMP-2 levels were observed in overweight and obese women and were associated with anthropometric parameters that indicate adipose tissue accumulation.

Unlike BMP-2, IL-17 has emerged as a negative regulator of adipogenesis and glucose metabolism in an experimental model. This cytokine acts in a complex network of inflammatory mediators that links adipose tissue to inflammatory cells such as neutrophils and adipocytes [10, 30, 31]. Indeed, excessive production of IL-17 during the initial stage of obesity could have a different effect on maintaining a prolonged inflammatory response and on associated long-term consequences. IL-17 was remarkably high in elderly individuals, and particularly in the obese and overweight middle-aged individuals; here, higher levels of VLDL and glycemia were observed. In addition, IL-17 was also positively correlated with classical anthropometric parameters (body mass index, arm muscle circumference, and hip and waist-to-hip ratio). These parameters have been described as anthropometric markers of poor prognosis in metabolic and cardiovascular diseases [19]. They have also been shown to contribute to the release of other inflammatory cytokines such as IL-6, IFN-gamma, and TNF [10], which reinforces the potential role of IL-17 in obesity-related diseases.

Our data reinforce the fact that obese individuals (middle-aged and elderly) present with higher cholesterol fractions, triglycerides, and glycemia, when compared to those parameters in eutrophic individuals, in accordance with the previous studies [1]. However, these overweight middle-aged individuals must receive follow-up care from health professionals when considering the relevance of visceral obesity, alongside soluble biomarkers. Nonetheless, integrated intervention programs based on weight reduction are demanding, since the immune conditions (systemic inflammation) in these patients are similar to those in elderly individuals. This might indicate the presence of accelerated damage due to the production of inflammatory mediators (e.g. CXCL-16, IL-17, and BMP-2), since high levels of these circulating factors clearly represent a risk for cardiovascular disease, cancer, and metabolic disturbances [12, 19, 25]. Preventing and reducing overweight and obese conditions depends on medical interventions, in addition to individual lifestyle changes; however, further research to motivate behavioral changes is required.

Conclusion
The present study provides complementary information regarding BMP-2, IL-17, and particularly CXCL-16, and their potential to be used as soluble markers capable of differentiating overweight and obese statuses in middle-aged and elderly women. These biomarkers were shown to be correlated with anthropometric (body mass index, triceps skinfold, arm muscle circumference, and hip and waist circumference) and biochemical (VLDL and glycemia) parameters. These factors might alter the inflammation axis observed in overweight and obese individuals, as well as in related disorders such as atherosclerosis, insulin resistance, and vascular damage. However, their clinical application needs to be investigated further in prospective population studies since its application in the clinical screening of overweight could help early in the preventive management of diseases associated with obesity.

Abbreviations
BMP-2: Bone morphogenetic protein-2; CXCL-16: CXC16 ligand; HDL: High density lipoprotein; IL-17: Interleukin-17; IL-6: Interleukin-6; LDL: Low density lipoprotein; T4: Thyroxine; TNF: Tumor necrosis factor; TSH: Thyroid-stimulating hormone; VLDL: Very low density lipoprotein

Acknowledgments
The authors thank Mr. Leonardo Mesquita for the donation of the S-Monovette tubes (Sarstedt LTDA) used in the present research.

Fundings
The present study was supported by granting from Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG) and Universidade Federal de Ouro Preto (UFOP). AT, FLPO and JASG are in credit with the CNPq for the fellowship applied to the development of the research.

Availability of data and materials
Data sharing is not applicable to the present article as no datasets were generated or analyzed while the current study was still going.

Authors’ contributions
SMTLR, LR, GPC, DS, VPF and APB have helped conducting the experiments with patients and performing immunohistochemistry and anthropometric analysis; FLPO conducted the statistical analysis; RLCN was responsible for the biochemical analyses; AT has designed, supervised the project and the data analysis; JASG, FLPO and AT wrote the manuscript after the content was discussed and reviewed by all authors. All authors read and approved the final manuscript.
Competing interest
The authors declare that they have no competing interests regarding the publication.

Consent for publication
Not applicable.

Ethics approval and consent to participate
The informed consent was obtained from all participants and the herein conducted experiments were in accordance with the Declaration of Helsinki and its proposal. It was approved by the ethical clearance of the Ethics Review Board of Ouro Preto Federal University (CAAE: 0048.0.238.000-10).

Author details
1Department of Biological Sciences, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil. 2Post-graduation Program in Biological Sciences/NUPEB, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil. 3Department of Clinical Analyses Laboratory of the Pharmacy School, Federal University of Minas Gerais, Belo Horizonte, Minas Gerais, Brazil. 4Clinical Analyses Laboratory of Tropical Biomas, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil. 5Department of Statistics, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil. 6Department of the Immunobiology of Inflammation, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil. 7Department of Morphology, Federal University of Minas Gerais, Belo Horizonte, Minas Gerais, Brazil. 8Department of Morphology, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil. 9Department of Post-graduation Program in Health and Nutrition, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil. 10Department of Biological Sciences, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil. 11Clinical Analyses Laboratory of the Pharmacy School, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil. 12Department of Health Sciences, Federal University of Ouro Preto, Ouro Preto, Minas Gerais, Brazil.

Received: 19 September 2016 Accepted: 27 February 2017
Published online: 11 March 2017

References
1. Macpherson-Sánchez AE. Integrating fundamental concepts of obesity and eating disorders: implications for the obesity epidemic. Am J Public Health. 2015;105:271–85.
2. Lastra G, Sowers JR. Obesity and cardiovascular disease: role of adipose tissue, inflammation, and the renin-angiotensin-aldosterone system. Horm Mol Biol Clin Investig. 2013;15:49–57.
3. Luis DA, Sagrado MC, Conde R, Aller R, Izaola O, Romero E. Circulating adipocytokines in obese nondiabetic patients in relationship with cardiovascular risk factors, anthropometry and resting-energy expenditure. Ann Nutr Metab. 2007;51:416–20.
4. Fain JN. Release of inflammatory mediators by human adipose tissue is enhanced in obesity and primarily by the nonfat cells: a review. Mediators Inflamm. 2010;2010:1–20.
5. Jung UJ, Seo YR, Ryu R, Choi MS. Differences in metabolic biomarkers in the blood and gene expression profiles of peripheral blood mononuclear cells among normal weight, mildly obese and moderately obese subjects. Br J Nutr. 2016;115:91–11.
6. Frasca D, Landin AM, Lechner SC. Aging down-regulates the transcription factor E2A, activation-induced cytokine deaminase, and Ig class switch in human B cells. J Immunol. 2008;180:5283–90.
7. Castelo-Branco C, Soval i. The immune system and aging: a review. Gynecol Endocrinol. 2014;30:16–22.
8. Arif M, Jomayyaz FR. Inflammation as a potential link between nonalcoholic fatty liver disease and insulin resistance. J Endocrinol. 2013;218:25–36.
9. Walla JA, Thomas HE, Sutherland AP. Linking obesity with type 2 diabetes: the role of T-bet. Diabetes Metab Syndr Obes. 2014;7:331–40.
10. Tarantino G, Costantini S, Finelli C. Is serum Interleukin-17 associated with early atherosclerosis in obese patients? J Transl Med. 2014;12:1–10.
11. Reinert-Hartwall L, Honkanen J, Salo HM. TH1/TH17 plasticity is a marker of advanced β cell autoimmunity and impaired glucose tolerance in humans. J Immunol. 2015;194:68–75.
12. Lehke M, Millington SC, Letefora M. CXCL16 is a marker of inflammation, atherosclerosis, and acute coronary syndromes in humans. J Am Coll Cardiol. 2007;49:442–9.
13. Kabir SM, Lee ES, Son DS. Chemokine network during adipogenesis in 3T3-L1 cells: Differential response between growth and proinflammatory factor in preadipocytes vs. adipocytes. Adipocyte. 2014;3:97–106.
14. Huang H, Song TJ, Li X. BMP signaling pathway is required for commitment of C3H10T1/2 pluripotent stem cells to the adipocyte lineage. Proc Natl Acad Sci USA. 2009;106:12670–5.
15. Hussein KA, Choksi K, Akeel S. Bone morphogenetic protein 2: a potential new player in the pathogenesis of diabetic retinopathy. Exp Eye Res. 2014;125:79–88.
16. WHO. Obesity: preventing and managing the global epidemic: Report of a WHO consultation, Geneva, 3–5 Jun 1997. Geneva: World Health Organization; 1998. WHO/NUT/98.1.
17. Felix HC, Bradway C, Chisholm L, Pradhan R, Weech-Maldonado R. Prevalence of moderate to severe obesity among U.S. Nursing Home Residents, 2000–2010. Res Gerontol Nurs. 2015;13:1–6.
18. Kuczynski RJ, Ogden CL, Guo SS, Grummer-Strawn LM, Flegal KM, Mei Z, et al. CDC Growth charts for the United States: methods and development. Vital Health Stat. 2000;246:1–190.
19. Coffman E, Richmond-Bryant J. Multiple biomarker models for improved risk estimation of specific cardiovascular diseases related to metabolic syndrome: a cross-sectional study. Popul Health Metr. 2015;13:7–12.
20. Feng B, Zhang T, Xu H. Human adipose dynamics and metabolic health. Ann NY Acad Sci. 2013;1281:160–77.
21. Sorensen LP, Sorengaard E, Nellmann B, Christiansen JS, Gormsen LC, Nielsen S. Increased VLDL-triglyceride secretion preceds impaired control of endogenous glucose production in obese, normoglycemnic men. Diabetes. 2011;60:2257–64.
22. Huber J, Kiefer FW, Zeyda M. CC chemokine and CC chemokine receptor profiles in visceral and subcutaneous adipose tissue are altered in human obesity. J Clin Endocrinol Metab. 2008;93:3215–21.
23. Kuri E, Shi J, Martonen E, Finckenberg P, Merviaa E. Distinct effects of calorie restriction on adipose tissue cytokine and angiogenesis profiles in obese and lean mice. Nutr Metab. 2012;9:64–72.
24. Laugand LE, Asvold BO, Vatten LJ, Jansky I, Platou C, Michelsen AE, et al. Soluble CXCL16 and risk of myocardial infarction: The HUNT study in Norway. Atherosclerosis. 2016;244:188–94.
25. Mitsuoka H, Toyohara M, Kume N, Hayashida K, Jinnai T, Tanaka M, et al. Circulating solube SR-PSOX/CXCL16 as a biomarker for acute coronary syndrome-comparison with high-sensitivity C-reactive protein. J Atheroscler Thromb. 2009;16:586–93.
26. Ma A, Yang S, Wang Y, Wang X, Pan X. Increased of serum CXCL16 levels correlates well to microembolic signals in acute stroke patients with carotid artery stenosis. Clinica Chimica Acta. 2016;460:67–71.
27. Zhang J, Li L. BMP signaling and stem cell regulation. Dev Biol. 2005;284:1–11.
28. Chen D, Ji X, Harris MA. Differential roles for bone morphogenetic protein (BMP) receptor type IB and IA in differentiation and specification of mesenchymal precursor cells to osteoblast and adipocyte lineages. J Cell Biol. 1998;142:295–305.
29. Guiu-Jurado E, Unthan M, Böhrer N, Kern M, Landgraf K, Dietrich A, et al. Bone morphogenetic protein 2 (BMP2) may contribute to partition of energy storage into visceral and subcutaneous fat deposits. Obesity (Silver Spring). 2016;12:doi: 10.1002/oby.21571.
30. Züniga LA, Shen WJ, Joyce-Shaikh B. IL-17 regulates adipogenesis, glucose homeostasis, and obesity. J Immunol. 2010;185:6947–59.
31. Ahmed M, Gaffen SL. IL-17 in obesity and adipogenesis. Cytokine Growth Factor Rev. 2010;21:449–53.