Dissipation and Residue of Metalaxyl-M and Azoxystrobin in Scallions and Cumulative Risk Assessment of Dietary Exposure to Hepatotoxicity

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Abstract: Metalaxyl-M and azoxystrobin have been used to control various fungal diseases on scallion and other crops. In view of the adverse toxic effects of both on the mammalian liver, it is necessary to conduct a cumulative risk assessment of their dietary exposure to consumers. The residues of metalaxyl-M and azoxystrobin on scallion were determined by a quick, easy, cheap, effective, rugged, and safe method (QuEChERS) combined with high-performance liquid chromatography tandem mass spectrometry (LC-MS/MS). The half-lives were about 1.15 and 3.89 days, respectively, and the final residues after a seven-day harvest interval were <0.001–0.088 mg/kg and 0.190–4.687 mg/kg, respectively. The cumulative dietary risk quotient of the two fungicides to Chinese consumers calculated by the probability model is 13.94%~41.25%. According to the results of the contribution analysis, the risk posed by azoxystrobin is much greater than that of metalaxyl-M. Although metalaxyl-M and azoxystrobin do not pose a cumulative risk to Chinese consumers, the risk to children and adolescents is significantly higher than that to adults. This suggests that in future research, more consideration should be given to the cumulative risk of compounds to vulnerable groups.

Keywords: cumulative risk assessment; probabilistic model; metalaxyl-M; azoxystrobin; residue

1. Introduction

As a common spice, scallion is widely eaten in many parts of Asia and some Latin American countries. Scallion contains a variety of active ingredients, has a broad-spectrum antibacterial effect, and is very valuable for the development of medicine and food [1]. In addition, many studies have proved that scallion contains active ingredients to inhibit cancer and treat cardiovascular diseases [2–4]. However, with the continuous expansion of scallion production and planting areas, soil pathogenic bacteria have increased, and the occurrence and damage of scallion diseases continues to increase [5]. Rational use of chemical pesticides is needed to reduce losses. Metalaxyl-M and azoxystrobin are used as systemic fungicides to control Peronosporales of scallion.

Metalaxyl [methyl N-(2,6-dimethylphenyl)-N-(methoxyacetyl)-D-alaninate] is a phenylamide broad-spectrum fungicide which inhibits the incorporation of uridine into RNA through the interaction with RNA polymerase-1, thus it is widely used across the world to control various crops disease caused by Peronosporales [6–8]. Metalaxyl is a racemate; the product has been gradually replaced by metalaxyl-M (Figure S1A) which is composed of main bioactive enantiomeric (R-enantiomer) component and is therefore also called high-efficiency metalaxyl [9]. The original intention of replacing metalaxyl with metalaxyl-M was to reduce the amount of this pesticide application by almost half and to reduce the risk of residues [10]. The 2002 JMPR (The Joint FAO/WHO Meeting on Pesticide Residues) report stated that toxicological studies at the time found no toxicological differences between
metalaxyl and metalaxyl-M, and that the dose–response relationship was similar [11]. However, up until now, many studies have shown that metalaxyl-M may cause more serious toxicological effects on environmental organisms and mammals than racemic metalaxyl. For instance, Zhang et al. found that metalaxyl-M induced more severe inflammation, necrosis and vacuolization in mouse liver cells than racemic metalaxyl [12]. The studies by OuYang et al. [13] and Liu et al. [14] also indicate that metalaxyl-M is significantly toxic to aquatic (Tubifex tubifex) and terrestrial (Eisenia fetida) non-target animals. It is necessary to pay attention to the new risks caused by the large application of metalaxyl-M.

Azoxystrobin (Figure S1B), methyl (E)-2-[[6- (2-cyanophenoxy)-4-pyrimidinyl]oxy]-α-(methoxymethylene)benzeneacetate, is one of the new methoxy acrylate fungicides developed with natural product strobilurins as the lead compound [15]. The main mechanism of action is to inhibit mitochondrial respiration by blocking the electron transfer between cytochrome b and cytochrome c1, inhibiting both mycelial growth and spore germination [16,17]. As a typical broad-spectrum systemic fungicide, azoxystrobin has a good control effect on most diseases caused by fungi such as Ascomycetes, Basidiomycetes, Deuteromycetes and Oomycetes, and it is precisely because it can control many types of fungal diseases at the same time that azoxystrobin has achieved great success and is widely used around the world [18,19]. The JMPR report shows that the acute oral LD50 (lethal dose) in rats is >5000 mg/kg bw, and the acute dermal LD50 is > 2000 mg/kg bw, and so falls within low-toxicity pesticides. However, the toxicological evaluation also points out that azoxystrobin has certain liver toxicity to mammals [20].

Since pesticides pose potential hazards to public health, and dietary intake is the most direct and major source of population exposure to pesticides, it is necessary to monitor and evaluate pesticide residues in food in order to protect consumers [21,22]. Due to the large registered crop species and application rates of metalaxyl-M and azoxystrobin, consumers are often exposed to both due to multiple residues on the same crop and/or residues on multiple different crops, as shown by the monitoring results of pesticide residues in various places [23,24]. In the JMPR report, the toxicological descriptions of these two low-toxicity pesticides mentions their adverse effects on the mammalian liver, and that the main target organ is the liver; both can lead to different degrees of increase in mammalian liver weight [11,20]. Therefore, when assessing the potential health hazards of these two pesticide residues in food, it may be more meaningful to take into account the additive toxic effects of the two pesticides, and to assess their cumulative exposure risk.

The US Environmental Protection Agency (EPA), the European Union Food Safety Authority (EFSA), and the World Health Organization WHO/IPCS have respectively summarized the currently available methods for assessing the cumulative risk of mixtures [25–29], including the hazard index (HI), point of departure index (PODI), margin of exposure (MOE), and the relative potency factor (RPF) method, etc. Each method has its own calculation method to determine the toxicity of the dose–additive effects due to exposure, but in general the RPF method is more applicable and is the preferred method [30]. Cumulative exposure concentration obtained by these methods are compared with health reference values, such as the acceptable daily intake (ADI), the acute reference dose (ARfD), the no observed adverse effect level (NOAEL) or benchmark dose limit (BMDL) to obtain the cumulative risk.

In the present study, QuEChERS method combined with LC-MS/MS was established to determine the residues of metalaxyl-M and azoxystrobin on scallion in four regions of China. Combined with residue data from other crops, a probabilistic model was used to fit the cumulative dietary exposure of the two pesticides and to assess chronic dietary risks to consumers.

2. Materials and Methods
2.1. Reagents and Standards

Metalaxyl-M standard (purity 99.3%) and azoxystrobin standard (purity 98.7%) were purchased from Dr. Ehrenstorfer GmbH (Augsburg, Germany). Chromatographic purity
acetonitrile was purchased from Merck KGaA. Analytical-grade sodium chloride (NaCl) and anhydrous magnesium sulfate (MgSO₄) were obtained from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China) and Beijing Tongguang Fine Chemical Co. (Beijing, China), respectively. Chromatographic purity formic acid was purchased from CNW Technologies (Shanghai, China). The purification fillers of octadecylsilane (C18), primary secondary amine (PSA), and graphitized carbon black (GCB) were purchased from Agela Technologies (Tianjin, China), and a syringe filter (nylon, 0.22 µm) was purchased from ANPEL Laboratory Technologies Inc. (Shanghai, China). The 39% metalaxyl-M-azoxystrobin suspension emulsion (SE) used in the field trial tests was provided by Syngenta Investment Co., Ltd. (Shanghai, China).

To obtain a 1000 mg/L standard solution, 10 mg metalaxyl-M standard and azoxystrobin standard was accurately weighed and dissolved in acetonitrile, which was then maintained at 4 °C in the dark. A 2.5 mL amount of the standard solution was accurately weighed and diluted with acetonitrile. An appropriate amount of the standard mixed solution was accurately transferred and diluted with blank scallion matrix extract to prepare 0.0005, 0.001, 0.005, 0.01, 0.05, 0.1, 0.5, 0.75, and 1.0 mg/L matrix-matched standard solutions for sample quantification.

2.2. Extraction and Purification Process of the Samples

Firstly, 10 g (±0.01 g) homogenized scallion samples were weighed into a 50 mL centrifuge tube (the quality control and additive recovery samples were allowed to stand for 15 min after adding the standard solution). A 20 mL amount of chromatographic acetonitrile was added and shaken for 5 min in a high-throughput tissue grinder. After ultrasonic extraction for 15 min, 5 g of sodium chloride was added and then shaken by hand for 2 min. This was centrifuged at 6000 rpm for 5 min, and then the supernatant was taken and purified with a modified QuEChERS tube. After high-speed centrifugation at 10,000 rpm for 5 min, the supernatant was filtered through a 0.22 µm membrane and put into the injection bottle for LC-MS/MS analysis.

The effects of 12 different purification adsorbents on the response results were compared, including 50 mg PSA, 50 mg C18, 50 mg GCB, 25 mg PSA with 150 mg MgSO₄, 50 mg PSA with 150 mg MgSO₄, 25 mg C18 with 150 mg MgSO₄, 50 mg C18 with 150 mg MgSO₄, 25 mg GCB with 150 MgSO₄, 50 mg GCB with 150 mg MgSO₄, 50 mg PSA + 50 mg C18 with 150 mg MgSO₄, 50 mg PSA + 50 mg C18 + 50 mg GCB with 150 mg MgSO₄, and blank. The optimal adsorbent was selected by comparing the addition recovery at the level of 0.002 mg/kg.

2.3. Instrumentation

Metalaxyl-M and azoxystrobin were analyzed by high-performance liquid chromatography (Agilent 1290 Infinity II Multisampler, Agilent Technologies, City of Santa Clara, CA, USA) and a tandem triple quadrupole mass spectrometer (6470 Triple Quadrupole LC/MS System, Agilent Technologies, City of Santa Clara, CA, USA) with an electrospray ionization (ESI) source operated in positive ion mode (ESI+). An Agilent EclipsePlusC18 RRHD column (50 mm × 2.1 mm, 1.8 µm) was used for chromatographic separation at a temperature of 40 °C. The mobile phases were acetonitrile (A) and 0.1% (v/v) formic acid aqueous solution (B). The gradient elution procedure is shown in Table S1. The flow was 0.30 mL/min, and the sample injection volume was 1.5 µL. The gas temperature was set at 280 °C and the gas flow rate at 7 L/min for MS detection working conditions. The sheath gas temperature was set at 320 °C and the sheath gas flow was 11 L/min, and the capillary voltages were controlled at 4000 V under positive ion detection mode. Analytes were determined in multiple reaction monitoring (MRM) mode. For instrument control, data acquisition, and processing, MassHunter Workstation version 10.1 (Agilent Technologies, City of Santa Clara, CA, USA) was used. Under the operating conditions given above, the two compounds were quantified based on the acquisition parameters as listed in Table S2.
2.4. Method Validation

The analytical method was validated from the aspects of linearity, sensitivity, matrix effect, precision and stability [31].

The matrix effect can be expressed by the equation as follows [32]:

\[
\text{Matrix effect (ME, %)} = \frac{\text{Slope of calibration curves in matrix} - \text{slope of calibration curves in solvent}}{\text{slope of calibration curves in solvent}} \times 100\%.
\]

The different concentrations of the standard solution were prepared with blank matrix extract to eliminate the excessive matrix effect. The prepared standard solutions with different concentration gradients (0.0005, 0.001, 0.005, 0.01, 0.5, 0.75, and 1.0 mg/L) were analyzed under the above analysis conditions, and the standard curves were drawn according to the ratio of the absolute number of compounds to the peak areas to verify linearity. The accuracy and precision of the method was verified by five repeated recovery experiments at four different spiked concentrations (0.001, 0.01, 1, and 7 mg/kg).

2.5. Field Trial Tests

The open-field trials were designed in accordance with the Guideline on Pesticide Residue Trials (NY/T 788-2018) published by the Ministry of Agriculture and Rural Affairs, P. R. China. Field trials were implemented in four different scallion producing areas in China, include Changping, Beijing (116.41 E, 40.22 N, continental monsoon climate), Taian, Shandong (116.97 E, 36.10 N, temperate monsoon climate), Changsha, Hunan (113.31 E, 28.31 N, subtropical monsoon climate), and Guiyang, Guizhou (106.63 E, 26.32 N, subtropical humid and mild climate). The 39% metalaxyl-M·azoxystrobin SE (10.6% metalaxyl-M and 28.4% azoxystrobin) was sprayed at 351 g a.i./ha active ingredient, and the application was carried out three times with an interval of seven days. The treatment and control samples were sampled multiple times at different time intervals (including 0, 5, 7, 10, and 14 d) after the last application. At least 12 normally growing and healthy scallions with a total weight at least 2 kg were collected and each test for degradation was conducted in triplicate, and for final residue in duplicate. The blank control treatment was the same as the experimental treatment, and the protective line was established between each plot. All scallion samples were cut into sections after removal of soil and roots and preserved at −20 °C until use.

2.6. Cumulative Exposure Estimation

Toxicological data in the FAO JMPR assessment report for these two pesticides were selected to ensure data reliability and consistency. The NOAEL for liver toxicity of metalaxyl-M and azoxystrobin were 8 mg/kg bw per day (13-weeks study in dogs) and 18.2 mg/kg bw per day (2-year study in rats), respectively. Even though metalaxyl-M is more toxic than azoxystrobin, azoxystrobin was chosen as the index compound in order to minimize uncertainty due to the fact that the latter has more residue data and its residues are much higher than the former in many crops. The comparison of the NOAEL of the compound with the index compound gives its RPF relative to the index compound.

The cumulative residue of metalaxyl-M and azoxystrobin can be calculated with the following Equation (1) [33]:

\[
C_{\text{index},j} = \sum_{i=1}^{\text{CONS}_{i}} (C_{i,j} \times \text{RPF}_{i})
\]

where the \(C_{\text{index},j}\) is the cumulative concentration expressed as the index compound in food \(j\), \(C_{i,j}\) is the concentration of component compound \(i\) in food \(j\), and \(\text{RPF}_{i}\) is the relative potency factor of component compound \(i\) relative to the index compound.

The cumulative exposure of metalaxyl-M and azoxystrobin can be calculated with the following Equation (2) [33]:

\[
E = \sum_{j=1}^{\text{bw}} E_{j} = \sum_{j=1}^{\text{bw}} \frac{\text{CONS}_{j} \times C_{\text{index},j}}{\text{bw}}
\]
where the $E$ is the cumulative exposure of the population, $E_j$ is the cumulative exposure of the population in individual food $j$, $CONS_j$ is the food $j$ consumption of the population, and $bw$ is the body weight of the population.

2.7. Cumulative Dietary Risk Assessment on Probabilistic Method

Monte Carlo simulation was used to fit the input parameters such as pesticide residues, food consumption and body weight, so as to obtain the cumulative exposure closer to the real situation. Inspired by the probabilistic evaluation method recommended by EFSA [34], we divided the entire evaluation process into inner and outer loops. The inner loop obtains the distribution of population chronic dietary exposure risk variability by performing 2000 random Monte Carlo iterations of the input parameters. The outer loop executes the inner loop multiple times (such as 500 times) to obtain the uncertainty distribution of the iteration evaluation results. Monte Carlo sampling was completed in the @risk version 7.6.2 (Palisade, Ithaca, NY, USA) software.

The risk characterization of metalaxyl-M and azoxystrobin can be carried out using the following Equation (3) [33]:

$$R = \frac{E}{NOAEL_{index}} \times UF \times 100\%$$  \hspace{1cm} (3)

where $R$ is chronic dietary risk of the population, $E$ is the cumulative exposure of the population and comes from Equation (2), $NOAEL_{index}$ is the no observed adverse effect level of the index compound, $UF$ is the uncertainty factor. The uncertainty factor is divided into two parts, which reflect the uncertainty between species and the uncertainty within species, respectively. For metalaxyl-M and azoxystrobin, both uncertainty factors are taken as 10, that is, the total uncertainty factor is taken as 100.

3. Results and Discussion

3.1. Optimization of the Analytical Method

The existence of [M + H]$^+$ was determined by the target ion scanning method, and the detection monitoring (MRM) mode of the target compounds was optimized by multiple reactions. The chromatograms of metalaxyl-M and azoxystrobin are shown in Figure 1.

Various adsorbents including GCB, C18 and PSA are widely used to remove different impurities in the matrix, mainly for pigment removal, sugar and fatty acid removal and reverse phase extraction, respectively [35,36]. The effects of 12 different purification adsorbents on the response results were compared by 0.002 mg/kg recovery experiment and the results are shown in Figure S2.

Except for 50 mg GCB and 50 mg GCB with 150 mg MgSO$_4$$_7$, the recoveries of other adsorbent combinations including blank are mostly between 80–100%, with little difference. In contrast, the absorbent 50 mg PSA + 50 mg C18 with 150 mg MgSO$_4$$_7$ provided the best recovery results of all adsorbent combinations, and the recoveries of metalaxyl-M and azoxystrobin at the level of 0.002 mg/kg reached 97.79% ± 2.28% and 94.34% ± 7.79% respectively. Therefore, it was selected as the adsorbent combination for subsequent treatment in this study.
Figure 1. Typical LC-MS/MS chromatograms of metalaxyl-M and azoxystrobin in scallion samples: (A, B) blank sample, (C, D) matrix standard at 0.0005 mg/L, and (E, F) spiked sample at 0.001 mg/kg.

3.2. Method Validation

The regression equations of the solvent standard curves of metalaxyl-M and azoxystrobin were $y = 175194x + 4646.8$, $R^2 = 0.9919$ and $y = 505999x + 9186.5$, $R^2 = 0.9946$, respectively. The regression equations of the matrix standard curves of metalaxyl-M and azoxystrobin in scallions were $y = 71002x + 149.65$, $R^2 = 0.9996$ and $y = 237639x + 1759.7$, $R^2 = 0.9983$, respectively. The ME of these two pesticides on scallions were as high as $-59.47\%$ and $-53.04\%$, respectively. Therefore, it is necessary to use the blank scallion matrix extract to prepare the standard solution in order to eliminate the matrix effect.

As shown in Table S3, the average recovery of metalaxyl-M in scallion was $97.66\%–106.27\%$ at spiked levels of 0.001, 0.01, 1, and 7 mg/kg with RSD of 2.11–6.88\%, and the average recovery of azoxystrobin was $88.82\%–105.19\%$ with RSD of 5.30–12.64\%. The results indicate that the accuracy and precision of this method meet the requirements of pesticide residue analysis.

The limit of quantification (LOQ) was defined as the minimum spiked concentrations of target analytes in the matrix [37,38]. According to the recovery experiments, under the above analysis conditions, the LOQ of metalaxyl-M and azoxystrobin in scallion was 0.001 mg/kg. The above validation results suggest that this analytical method is reliable for the determination of the two fungicide residues in scallion.

3.3. Dissipation Behavior

Research about pesticide degradation is an important part of the risk assessment. Almost all pesticides degrade over time after application and their residues decrease, and these two fungicides are no exception (Figure 2); the dissipation process was of first-order reaction kinetics:

$$C_t = C_0 \times e^{-kt} \quad (4)$$

$$t_{1/2} = \ln 2/k \quad (5)$$

where $C_t$ (mg/kg) is the concentration of the pesticide residue at time $t$ (days), $C_0$ (mg/kg) is the initial concentration, $k$ is the degradation rate constant (day$^{-1}$), and $t_{1/2}$ (days) is the degradation half-life of the pesticide which was determined from the $k$ value.
The initial deposition of metalaxyl-M in Shandong and Guizhou were in the range of 0.055–0.080 mg/kg and 0.049–0.067 mg/kg, respectively; seven days after the last application, the degradation rate reached 96.88–98.28%, and the half-lives were 1.15 and 1.06 days, respectively, which are similar to or lower than other research results [39]. The initial deposition of azoxystrobin in these two places were in the range of 2.115–2.590 mg/kg and 1.284–1.523 mg/kg, respectively; seven days after the last application, the degradation rate reached 71.09–77.20%, and the half-lives were 2.28 and 3.89 days, which are similar to other research results [40,41]. The different dissipation rates and initial deposition may be related to the crop species, climate types and planting modes at different sites.

3.4. Final Residue Testing

As shown in Table 1, the final residues of metalaxyl-M and azoxystrobin in scallion were determined by the above analysis method. The harvest interval was 7 and 10 days; the residue of metalaxyl-M in scallion at harvest was <0.001–0.088 and <0.001–0.049 mg/kg, respectively. The residue of azoxystrobin at harvest was 0.190–4.687 mg/kg and 0.043–4.368 mg/kg, respectively. Therefore, at the pre-harvest interval (PHI) of seven days, the final residues of azoxystrobin in scallion were lower than the China maximum residue limit (MRL) 7 mg/kg, the EU MRL 10 mg/kg for Welsh onion [42] and the US MRL 7.5 mg/kg for Onion, green, subgroup 3-07B [43]. China has not yet formulated the MRL of metalaxyl-M on scallions, but the final residues were lower than the EU MRL 0.3 mg/kg for Welsh onion [44] and the US MRL 10 mg/kg for Onion, green, subgroup 3-07B [45]. These data will provide reference for metalaxyl-M-azoxystrobin SE reasonable application in scallion and the formulation of MRL of metalaxyl-M on scallion in China.
Table 1. Final residues of metalaxyl-M and azoxystrobin in scallion samples.

| Location | Harvest Interval (d) | Metalaxyl-M | Azoxystrobin |
|----------|----------------------|-------------|--------------|
|          |                      | 1  | 2  | 1  | 2  |
| Beijing  | 7                    | 0.003 | 0.003 | 0.363 | 0.190 |
|          | 10                   | 0.005 | 0.004 | 0.377 | 0.492 |
| Shandong | 7                    | 0.002 | 0.002 | 1.420 | 1.360 |
|          | 10                   | 0.003 | 0.002 | 0.071 | 0.043 |
| Guizhou  | 7                    | <0.001 | <0.001 | 0.314 | 0.326 |
|          | 10                   | <0.001 | <0.001 | 0.350 | 0.334 |
| Hunan    | 7                    | 0.082 | 0.088 | 4.687 | 4.687 |
|          | 10                   | 0.049 | 0.049 | 4.368 | 4.203 |

Supervised trial median residue (STMR) of 7 d and 10 d were 0.0025 and 0.0035 mg/kg, respectively.

Highest residue (HR) of 7 d and 10 d were 0.088 and 0.049 mg/kg, respectively.

3.5. Cumulative Dietary Risk Assessment

According to the differences between urban and rural areas, gender and age, Chinese consumers were divided into a total of 40 groups. The different body weight and food consumption data were obtained from the Report on Nutrition and Health Status of Chinese Residents (2002). Based on the residue results obtained in this study and on other pesticide residue monitoring trials in China, dietary exposures for different food categories were calculated according to the relative potency factor method. In accordance with the principle of maximizing risk, it is assumed that consumers are exposed to the food with the largest residue of each category. We classify scallions into the soy sauce food category in the risk assessment work, according to the suggestions. All residue data and food categories are shown in Table 2.

The cumulative dietary risk assessment results are shown in Tables S4 and S5. All results did not exceed 100%, including the maximum in the most extreme cases (upper 95% confidence interval of P99.9 for rural females aged 2–3 years, 90.68%). Considering the variability of residual and consumption data, the P97.5 level was selected as an appropriate assessment of chronic risk. Considering the uncertainty of iteration, the upper limit of 95% confidence interval was chosen to be an appropriate level to achieve adequate protection. Based on these results, the risk faced by Chinese consumers is 13.94–41.25%. Among these, the minimum value of 13.94% appeared in the urban male group aged 30–44, while the maximum value of 41.25% was the risk result faced by rural girls aged 2–3. At the same time, the principle of risk maximization was followed in this study as much as possible. For example, the data of the food with the largest residue in a food category was extrapolated to the whole category, which brings a certain uncertainty and makes the evaluation results more conservative. Therefore, metalaxyl-M and azoxystrobin residues do not pose a risk of cumulative exposure hazard to the liver of Chinese consumers.

In order to compare the possible differences between the risks faced by different populations, all the risk results are presented as a heat map, as shown in Figure 3. Intuitively, age led to a large difference in risk outcomes. Children and adolescents are at significantly greater risk than adults. The most important reason is that differences in body weight led to differences in tolerance to toxicity without adequately reducing compound exposure, resulting in a nearly three-fold difference in risk outcomes. The average weight of all groups in rural areas is smaller than that of urban residents of the corresponding gender and age, and the dietary structure of residents also differs between urban and rural areas; as a result, the level of risk faced by rural residents is slightly higher than that of urban residents.
Table 2. Food categories and residue data screened for cumulative risk assessment.

| Food Category        | Pesticide      | Crop    | STMR (mg/kg) | HR (mg/kg) | C_index \(^{a,b}\) (mg/kg) |
|----------------------|----------------|---------|--------------|------------|-----------------------------|
| Rice and products    | Metalaxyl-M    | Rice    | -            | 0.160      | -                           |
|                      | Azoxystrobin   |         | -            | 0.230      | 0.160                       |
| Flour and products   | Metalaxyl-M    | Wheat   | -            | 0.023      | -                           |
|                      | Azoxystrobin   |         | -            | 0.180      | 0.023                       |
| Other cereals        | Metalaxyl-M    | Corn    | -            | 0.020      | -                           |
|                      | Azoxystrobin   |         | -            | 0.020      | 0.020                       |
| Potatoes and products| Metalaxyl-M    | Potato  | -            | 0.011      | -                           |
|                      | Azoxystrobin   |         | -            | 0.039      | 0.020                       |
| Legumes and products | Metalaxyl-M    | Soybean | -            | 0.020      | -                           |
|                      | Azoxystrobin   |         | -            | 0.020      | 0.020                       |
| Dark vegetables      | Metalaxyl-M    | Tomato  | -            | 0.034      | -                           |
|                      | Azoxystrobin   |         | -            | 0.034      | 0.051                       |
| Light vegetables     | Metalaxyl-M    | Cauliflower | - | 0.109 | - |
|                      | Azoxystrobin   |         | -            | 0.454      | 0.488                       |
| Fruits               | Metalaxyl-M    | Watermelon | - | 0.010 | - |
|                      | Azoxystrobin   |         | -            | 0.010      | 0.723                       |
| Vegetable oil        | Metalaxyl-M    | Peanut  | -            | 0.050      | -                           |
|                      | Azoxystrobin   |         | -            | 0.160      | 0.050                       |
| Soy sauce            | Metalaxyl-M    | Scallion | -            | 0.0025     | -                           |
|                      | Azoxystrobin   |         | -            | 0.088      | 0.867                       |

\(^{a}\) \(C_{\text{index}}\) was the cumulative concentration expressed as the index compound. \(^{b}\) The index compound was azoxystrobin; The RPF of metalaxyl-M was 2.275.

In order to compare the differences in the contribution of different pesticides and food categories to dietary exposure risk, while avoiding the possible effects of age, region and gender, contribution analysis was carried out for 2–3 years old infants, 30–44 years old adults, and 60–69 years old elderly. The analysis results are shown in Table S6. As previously stated, azoxystrobin contributed 76.22–84.58% of the total dietary risk, which was much higher than that of metalaxyl-M despite the latter being more toxic, due to the greater residues of azoxystrobin. For food categories, dark vegetables, light vegetables and fruits are the three main sources of risk, contributing 26.64–43.05%, 24.15–34.85%, and 7.81–32.95%, respectively. In addition to this, rice and products and soy sauce contributed 5.81–8.63% and 3.29–5.35%, respectively. The contribution of different food categories varies greatly: on the one hand, due to the huge difference in intake in the dietary structure, and on the other hand, due to the residues in different crops tending to differ.

Different populations have different contribution distributions due to variety in dietary structure. By comparing the contribution of azoxystrobin in Figure 4A, it can be seen that the distribution of contributions of different pesticides across all populations is not significantly different. Mainly due to insufficient data in the assessment to distinguish pesticide residues in foods ingested by different populations, the differences in pesticide contributions are mainly affected only by different food intakes. The same is true for different food categories. Figure 4B summarizes the contribution distribution of the three main risk source food categories for different populations. Taking fruits as an example, the contribution distribution shows that children are at higher risk than adults and the elderly, urban areas are higher than rural areas, and men are slightly lower than women, which is consistent with the trend of the proportion of fruits in the dietary structure of the population. In summary, there is little difference in the contribution distribution of risks faced by different groups of people, mainly because even if there are differences in dietary structure, the residual concentration of each food category does not change, so it is not enough to have a significant impact on the contribution results.
structure, the residual concentration of each food category does not change, so it is not necessary to divide consumer groups, especially for the risks faced by vulnerable groups.

and on the other hand, due to the residues in different crops tending to differ. The residues of metalaxyl-M and azoxystrobin on scallion were determined by LC-MS/MS. The residues were in the range of 0.001–0.009 mg/kg and 0.016–0.029 mg/kg, respectively, and the final residues after a seven-day harvest interval were <0.001–0.002 mg/kg and <0.001–0.004 mg/kg, respectively. The main reason is that although the dietary structure of different age groups differs significantly, such as children and adolescents faced significantly higher risks than the adult population. In summary, there is little difference in the contribution distribution of risks faced by different groups of people, mainly because even if there are differences in dietary tolerance to toxicity caused by different body weights. Therefore, in the assessment of dietary exposure risk, it is necessary to divide consumer groups, especially for the risks faced by vulnerable groups.

Conclusions

These results of cumulative dietary risk assessment suggest that, in future research, there should be still room for improvement in current dietary exposure assessments, focusing on expanding the amount of consumption and residue data, reducing uncertainty in the assessment, and incorporating more variables. For example, the foods acceptable...
to consumers in different regions often come from different origins, so their residues are not the same, and the consumption of food categories is often different due to differences in ethnic and regional cultures. Another example is the possible impact of changes in consumption and residues caused by factors such as seasons on the risk assessment results of subchronic and acute exposure to compounds. The subdivision of food categories is another way to reduce the uncertainty of the assessment. In short, expanding the amount of data and incorporating more considerations can effectively reduce the uncertainty and ensure that the evaluation results have sufficient accuracy.

4. Conclusions

The residues of metalaxyl-M and azoxystrobin on scallion were determined by QuEChERS method combined with LC-MS/MS. The half-lives were about 1.15 and 3.89 days, respectively, and the final residues after a seven-day harvest interval were <0.001–0.088 mg/kg and 0.190–4.687 mg/kg, respectively. The cumulative exposure risk of the two pesticides to the liver was assessed by the RPF method and probabilistic model, and azoxystrobin was chosen as the index compound. The results of cumulative dietary risk quotient are 13.94%–41.25%; rural females aged 2–3 years face the greatest exposure risk. Although less toxic, azoxystrobin contributed about 80% of the dietary exposure risk, mainly due to its higher residue levels in multiple foods. From the results, it can be seen that while the cumulative dietary exposure to the hepatotoxicity of both metalaxyl-M and azoxystrobin did not exceed the threshold risk to Chinese consumers, vulnerable groups such as children and adolescents faced significantly higher risks than the adult population. The main reason is that although the dietary structure of different age groups differs according to the current statistics, it does not differ enough to make up for the difference in tolerance to toxicity caused by different body weights. Therefore, in the assessment of dietary exposure risk, it is necessary to divide consumer groups, especially for the risks faced by vulnerable groups.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/molecules27185822/s1, Figure S1: Structures of metalaxyl-M and azoxystrobin: (A) metalaxyl-M, (B) azoxystrobin. Table S1: Gradient elution procedure. Table S2: Experimental parameters and chromatographic conditions of metalaxyl-M and azoxystrobin. Figure S2: Recovery of different modified QuEChERS purification conditions for targets in scallion matrix at the level of 0.002 mg/kg (n = 5): (a) 50 mg primary secondary amine (PSA), (b) 50 mg octadecysilane (C18), (c) 50 mg graphitized carbon black (GCB), (d) 25 mg PSA with 150 mg MgSO4, (e) 50 mg PSA with 150 mg MgSO4, (f) 25 mg C18 with 150 mg MgSO4, (g) 50 mg C18 with 150 mg MgSO4, (h) 25 mg GCB with 150 mg MgSO4, (i) 50 mg GCB with 150 mg MgSO4, (j) 50 mg PSA and 50 mg C18 with 150 mg MgSO4, (k) 50 mg PSA and 50 mg C18 with 150 mg MgSO4, and (l) blank. Table S3: Recoveries of metalaxyl-M and azoxystrobin in scallions (n = 5). Table S4: Cumulative dietary risk assessment of metalaxyl-M and azoxystrobin for male. Table S5: Cumulative dietary risk assessment of metalaxyl-M and azoxystrobin for female. Table S6: Contribution distribution of dietary risks faced by different population groups.

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References

1. Li, X.; Cai, Y.; Zhou, T.T. Research Progress of Bioactivity and Chemical Composition of Allium fistulosum. China Condiment 2018, 43, 196–200.

2. Hsing, A.W.; Chokkalingam, A.P.; Gao, Y.T.; Madigan, M.P.; Deng, J.; Gridley, G.; Fraumeni, J.F., Jr. Allium vegetables and risk of prostate cancer: A population-based study. J. Natl. Cancer Inst. 2002, 94, 1648–1651. [CrossRef] [PubMed]

3. Zhou, Y.; Zhuang, W.; Hu, W.; Liu, G.; Wu, T.; Wu, X. Consumption of Large Amounts of Allium Vegetables Reduces Risk for Gastric Cancer in a Meta-analysis. Gastrenterology 2011, 141, 80–89. [CrossRef] [PubMed]

4. Sengupta, A.; Ghosh, S.; Bhattacharjee, S. Allium vegetables in cancer prevention: An overview. Asian Pac. J. Cancer Prev. APJCP 2004, 5, 237–245.

5. Zheng, H. Diagnosis and identification on the fungal disease of bulb crops in Fujian. Chin. Agric. Sci. Bull. 2007, 23, 326–329.

6. Malhat, F.M. Persistence of metalaxyl residues on tomato fruit using high performance liquid chromatography and QuEChERS methodology. Arab. J. Chem. 2017, 10, S765–S768. [CrossRef]

7. Hassal, K.A. The Biochemistry and Uses of Pesticides: Structure, Metabolism, Mode of ACTION and Use in Crop Protection; Macmillan Press: Basingstoke, UK, 1990.

8. Abass, K.; Reponen, P.; Jalonen, J.; Pelkonen, O. In vitro metabolism and interactions of the fungicide metalaxyl in human liver preparations. Environ. Toxicol. Pharmacol. 2007, 23, 39–47. [CrossRef]

9. Buser, H.R.; Müller, M.D.; Poiger, T.; Balmer, M.E. Environmental behavior of the chiral acetamide pesticide metalaxyl: Enantioselective degradation and chiral stability in soil. Environ. Sci. Technol. 2002, 36, 221–226. [CrossRef]

10. Monkiedje, A.; Spitter, M.; Bester, K. Degradation of racemic and enantiopure metalaxyl in tropical and temperate soils. Environ. Sci. Technol. 2003, 37, 707–712. [CrossRef]

11. JMPR. The Joint FAO/WHO Meeting on Pesticide Residues. JMPR Report. 2002. Available online: https://www.fao.org/fileadmin/templates/agphome/documents/Pests_Pesticides/JMPR/Reports_1991-2006/Report_2002.pdf (accessed on 7 December 2021).

12. Zhang, P.; Wang, S.; He, Y.; Xu, Y.; Shi, D.; Yang, F.; Yu, W.; Zhu, W.; He, L. Identifying Metabolic Perturbations and Toxic Effects of Rac-Metalaxyl and Metalaxyl-M in Mice Using Integrative NMR and UPLC-MS/MS Based Metabolomics. Int. J. Mol. Sci. 2019, 20, 5457. [CrossRef]

13. OuYang, M.N.; Liu, X.; Guo, H.M.; Lu, Z.H.; Zhou, D.D.; Yang, Z.H. The different toxic effects of metalaxyl and metalaxyl-M on Tubifex tubifex. Ecotoxicol. Environ. Saf. 2021, 208, 11587. [CrossRef] [PubMed]

14. Liu, T.; Zhu, L.; Han, Y.; Wang, J.; Wang, J.; Zhao, Y. The cytotoxic and genotoxic effects of metalaxy-m on earthworms (Eisenia fetida). Environ. Toxicol. Chem. 2014, 33, 2344–2350. [CrossRef] [PubMed]

15. Conversa, G.; Bonasia, A.; Lazzizera, C.; Elia, A. Pre-harvest nitrogen and azoxystrobin application enhances raw product quality and post-harvest shelf-life of baby spinach (Spinacia oleracea L.). J. Sci. Food Agric. 2014, 94, 3263–3272. [CrossRef]

16. Matthews, G. The Pesticide Manual: A World Compendium, 10th ed.; Tomlin, C.D.S., Ed.; BCPC Publications: Berkshire, UK, 2000.

17. Bartlett, D.W.; Clough, J.M.; Godwin, J.R.; Hall, A.A.; Hamer, M.; Parr-Dobrzenski, B. The streboulurin fungicides. Pest Manag. Sci. 2002, 58, 649–662. [CrossRef] [PubMed]

18. Bartett, D.W.; Clough, J.M.; Godfrey, C.R.; Godwin, J.R.; Hall, A.A.; Heaney, S.P.; Maund, S.J. Understanding the streboulurin fungicides. Pestic. Outlook 2001, 12, 143–148. [CrossRef]

19. Cao, F.; Zhu, L.; Li, H.; Yu, S.; Wang, C.; Qiu, L. Reproductive toxicity of azoxystrobin to adult zebrafish (Danio rerio). Environ. Pollut. 2016, 219, 1109–1121. [CrossRef] [PubMed]

20. JMPR. The Joint FAO/WHO Meeting on Pesticide Residues. JMPR Report AZOXYSTROBIN, No. 229. 2008. Available online: https://www.fao.org/fileadmin/templates/agphome/documents/Pests_Pesticides/JMPR/Reports_1991-2006/Report_2002.pdf (accessed on 7 December 2021).

21. Jurasek, R.; Mutel, C.L.; Stoessel, F.; Hellweg, S. Life cycle human toxicity assessment of pesticides: Comparing fruit and vegetable diets in Switzerland and the United States. Chemosphere 2009, 77, 939–945. [CrossRef]

22. Zhao, P.; Chai, Y.; Liu, R.; Yuan, L. Dissipation, Residue, and Dietary Risk Assessment of Bifenthrin, Bifenazate, and Its Metabolite Bifenazate-Diazene in Apples Based on Deterministic and Probabilistic Methods. J. Agric. Food Chem. 2018, 66, 14302–14310. [CrossRef]

23. Liu, Y.; Ge, N.; Cui, Z.; Zhang, J.; Wu, Y.P.H.X.; Zhou, L.; Yang, D.; Cao, Y. Determination of pesticide residues in 282 samples of imported fruits and vegetables from Qingdao, Shenzhen, Dalian ports in 2012–2014. Chin. J. Food Hyg. 2016, 28, 511–515.

24. Mao, Y.; Zeng, J.; Huang, L.; Wang, Y.; Ning, J.; Zhao, Y. Analysis of Chromatographic Monitoring Results of Pesticide Residues in Fruits and Vegetables in GuLin City. Bull. Agric. Sci. Technol. 2019, 50, 167–169. (In Chinese)

25. European Food Safety Authority. International Frameworks Dealing with Human Risk Assessment of Combined Exposure to Multiple Chemicals. EFSA J. 2013, 11, 3313.

26. USEPA. Framework for Cumulative Risk Assessment; Risk Assessment Forum; U.S. Environmental Protection Agency: Washington, DC, USA, 2003. Available online: https://www.epa.gov/sites/production/files/2014-11/documents/frmwrk_cum_risk_assmnt.pdf (accessed on 6 January 2022).

27. USEPA. Supplementary Guidance for Conducting Health Risk Assessment of Chemical Mixtures; EPA/630/R-00/002; U.S. Environmental Protection Agency, Risk Assessment Forum: Washington, DC, USA, 2000.
28. WHO. Assessment of Combined Exposures to Multiple Chemicals; Report of a WHO/IPCS International Workshop on Aggregate/Cumulative Risk Assessment; WHO: Geneva, Switzerland, 2009. Available online: http://www.who.int/ipcs/methods/harmonization/areas/workshopreportdocument7.pdf (accessed on 4 January 2022).

29. European Food Safety Authority. Scientific Opinion on Risk Assessment for a Selected Group of Pesticides from the Triazole Group to Test Possible Methodologies to Assess Cumulative Effects from Exposure through Food from these Pesticides on Human Health. EFSA J. 2009, 7, 1167.

30. Gallagher, S.S.; Rice, G.E.; Scarano, L.J.; Teuschler, L.K.; Bollweg, G.; Martin, L. Cumulative risk assessment lessons learned: A review of case studies and issue papers. Chemosphere 2015, 120, 697–705. [CrossRef]

31. Yang, Y.; Qin, S.; Wang, X.; Cao, J.; Li, J. Dissipation Behavior and Acute Dietary Risk Assessment of Thiamethoxam and Its Metabolite Clothianidin on Spinach. Molecules 2022, 27, 2209. [CrossRef] [PubMed]

32. Li, C.; Chen, Z.; Qin, D.; Liu, R.; Li, L.; Li, W.; He, Y.; Yuan, L. Simultaneous determination of the herbicide bixlozone and its metabolites in plant and animal samples by liquid chromatography–tandem mass spectrometry. J. Sep. Sci. 2021, 44, 822–832. [CrossRef]

33. Chen, Z.; Xu, Y.; Li, N.; Qian, Y.; Wang, Z.; Liu, Y. A national-scale cumulative exposure assessment of organophosphorous pesticides through dietary vegetable consumption in China. Food Control. 2019, 104, 34–41. [CrossRef]

34. European Food Safety Authority; Anastassiadou, M.; Choi, J.; Coja, T.; Dujardin, B.; Hart, A.; Hernandez-Jerrez, A.F.; Jarrah, S.; Lostia, A.; Machera, K.; et al. Cumulative dietary risk assessment of chronic acetylcholinesterase inhibition by residues of pesticides. EFSA J. 2021, 19, e06392.

35. Chai, Y.; Liu, R.; He, W.; Xu, F.; Chen, Z.; Li, L.; Li, W.; Yuan, L. Dissipation Behavior, Residue, and Risk Assessment of Benzothiazolinone in Apples. Int. J. Environ. Res. Public Health 2021, 18, 4478. [CrossRef]

36. Yao, S.; Zhao, Z.; Lu, W.; Dong, X.; Hu, J.; Liu, X. Evaluation of Dissipation Behavior, Residues, and Dietary Risk Assessment of Fludioxonil in Cherry via QuEChERS Using HPLC-MS/MS Technique. Molecules 2022, 26, 3344. [CrossRef]

37. Yuan, L.; Chai, Y.; Li, C.; Liu, R.; Chen, Z.; Li, L.; Li, W.; He, Y. Dissipation, residue, dietary, and ecological risk assessment of atrazine in apples, grapes, tea, and their soil. Environ. Sci. Pollut. Res. 2021, 28, 35064–35072. [CrossRef]

38. Lv, L.; Su, Y.; Dong, B.; Lu, W.; Hu, J.; Liu, X. Dissipation Residue Behaviors and Dietary Risk Assessment of Bosalid and Pyraclostrobin in Watermelon by HPLC-MS/MS. Molecules 2022, 27, 4410. [CrossRef] [PubMed]

39. He, Y.; Jiao, X.; Zhang, T.; Wang, M.; Khan, M.; Khan, M.R.; She, Y. Study on the dissipation pattern and risk assessment of metalaxyl-M in rice grains and paddy soil and water by liquid chromatography-tandem mass spectrometry. Environ. Sci. Pollut. Res. 2021, 28, 4245–4252. [CrossRef] [PubMed]

40. Wang, S.W.; Sun, H.B.; Liu, Y.P. Dissipation and residue of azoxystrobin in banana under field condition. Environ. Monit. Assess. 2013, 185, 7757–7761. [CrossRef]

41. Bian, Y.; Guo, G.; Liu, F.; Chen, X.; Wang, Z.; Hou, T. Meptyldinocap and azoxystrobin residue behaviors in different ecosystems under open field conditions and distribution on processed cucumber. J. Sci. Food Agric. 2020, 100, 648–655. [CrossRef]

42. Reg. (EU) 2021/1807. Commission Regulation (EU). Available online: https://ec.europa.eu/food/plant/pesticides/eu-pesticides-database/mrls/?event=details&pest_res_ids=22&product_ids=&v=1&search.pr (accessed on 3 March 2022).

43. Title 40. 180.507 Azoxystrobin; tolerances for residues. Code of Federal Regulations. Available online: https://www.ecfr.gov/current/title-40/chapter-I/subchapter-E/part-180/subpart-C/section-180.507 (accessed on 3 March 2022).

44. Reg. (EU) 2017/1164. Commission Regulation (EU). Available online: https://ec.europa.eu/food/plant/pesticides/eu-pesticides-database/mrls/?event=details&pest_res_ids=152&product_ids=&v=1&search.pr (accessed on 3 March 2022).

45. Title 40. 180.546 Mefenoxam; tolerances for residues. Commission Regulation (EU). Available online: https://www.ecfr.gov/current/title-40/chapter-I/subchapter-E/part-180/subpart-C/section-180.546 (accessed on 3 March 2022).