Peach palm plantlet growth in different culture media in a temporary immersion system

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ABSTRACT: Peach palm is a domesticated palm commercially important for the production of fruits and hearts of palm. Somatic embryogenesis, an effective technique for mass propagation, was successfully established for this species. Furthermore, a temporary immersion system improved plant regeneration. However, production can be further improved by understanding the peach palm’s growth dynamic and modifications of culture media. The aims of this study were to evaluate the growth of plantlets cultured in different culture media in a temporary immersion system and to correlate the results with nutrient uptake during the growth period. Somatic embryo-derived young plantlets approximately 1 cm in length were cultivated for 12 weeks in a twin flask system containing MS, Y3 or N6 salts, Morel and Wetmore vitamins and 3% sucrose, with a monthly medium refreshment. Growth was measured and mineral analysis of the plantlets was carried out after 12 weeks of culture. The Y3 and MS salts were the most appropriate for the plant growth. Number of roots was 52.52% higher and the root size was 40.42% between the N6 and MS medium and the root number in Y3 medium was 37.74% greater than in MS medium, which is important for post acclimatization survival. K and Na are important elements for peach palm. N is not required at such a high concentration as in Murashige and Skoog formulation. The Chu (N6) medium did not generate high quality plantlets, possibly due to the absence of some micronutrients, like Mo, Cu and Co.

Key words: Arecaceae, Bactris gasipaes, culture medium, somatic embryogenesis, twin flask.

INTRODUCTION

Peach palm (Bactris gasipaes Kunth) is a domesticated palm species of tropical America that generates several products of commercial interest, especially hearts of palm. Cultivation of this species increased in recent years in Brazil due to its rapid growth, good heart of palm quality and greater market acceptance (SCHROTH et al., 2002; CLEMENT, 2008; GRAEFE et al., 2013).

Somatic embryogenesis is a suitable technique for palm species as it allows large-scale production of selected genotypes. The protocol can be automated, resulting in mass propagation at a lower
cost than traditional in vitro methods (ETIENNE et al., 2006). The in vitro culture of peach palm was already established by somatic embryogenesis but some bottlenecks showed that the process needs to be improved (STEINMACHER et al. 2007a; STEINMACHER et al. 2007b; STEINMACHER et al. 2007c; MACIEL et al., 2010).

The temporary immersion system enhances the quality of in vitro regeneration protocols and enables larger scale production. In a liquid medium, the contact of the plant tissue with the medium facilitates the uptake of nutrients, as well as diluting possible growth inhibitors exuded by the explants (GUPTA; TIMMIS, 2005; ADELBERG et al., 2010).

A protocol using a temporary immersion system for Bactris gasipaes plantlet regeneration was developed by Steinmacher et al. (2011) and this system resulted into more vigorous and taller plantlets when compared with those from a semi-solid culture medium.

The macronutrient composition of the culture medium plays an important role in the success of micropropagation (RUŽIĆ et al., 2004). The nutritional balance of a culture medium can be determinant for in vitro productivity and a deficiency in some elements can cause physiological and morphological alterations in the plants (REED et al., 2013). Furthermore, the nutrient requirement for induction or maturation of somatic embryos is quite different from that for conversion and plantlet development (MEHROTRA et al., 2007).

The aim of this study was to test different culture media and evaluate the growth and mineral uptake of peach palm plantlets grown in an immersion culture system.

MATERIALS AND METHODS

Somatic embryogenesis induction, maturation and conversion

Peach palm seeds of spineless plants from Porto Velho, Rondônia, Brazil, were used for somatic embryogenesis induction. The procedure was based on Steinmacher et al. (2011). The endocarp was removed and the kernels were surface-sterilized under aseptic conditions, by five min immersion in 70% ethanol, followed by a 30 min immersion in a 10% sodium hypochlorite solution plus one drop of Tween 20® in each 100 mL (0.1% v/v). Thereafter, the kernels were rinsed three times in sterile distilled water. Zygotic embryos were aseptically removed from the kernels and cultured in Petri dishes containing 30 ml of somatic embryogenesis induction medium. This medium was composed of MS salts (MURASHIGE & SKOOG, 1962), Morel vitamins (MOREL & WETMORE, 1951), 3% sucrose, 0.05% glutamine, 10 µM Picloram, 1 µM silver nitrate and 0.25% Gelzan (Sigma-Aldrich®). Cultures were maintained in the dark until somatic embryos appeared and then transferred to a multiplication medium, the same as above, supplemented with 0.1% glutamine, and subcultured in a fresh medium every 45 days.

For maturation, clusters of somatic embryos were transferred to a medium composed of MS salts, Morel vitamins, 3% sucrose, 0.1% glutamine, 0.15% activated charcoal and 0.25% Gelzan (Sigma-Aldrich®) and maintained at 25±2 °C under cool-white fluorescent tubes with a photosynthetic photon flux density (PPFD) of 20 µmol m⁻² s⁻¹ and a 16 h photoperiod for four weeks. Then the somatic embryos were transferred to the conversion medium, the same as for maturation, without glutamine and with 0.7% agar (Himedia®).

Temporary immersion system (TIS) culture

Plantlets > 1.0 cm in height were selected for TIS experiments (Figure 1B), based on STEINMACHER et al. (2011). Briefly, the TIS system consisted of twin bottles, with 20 plantlets in a 5 L plastic bottle and 300 ml of culture medium in another 5 L plastic bottle (Figure 1A). Every 6 h the medium was air-pumped into the plant compartment for 3 min. The air was sterilized through an autoclavable 0.2-µm filter. Three different culture media were tested (Table 1): MS salts (MURASHIGE & SKOOG, 1962), Y3 salts (EEUWENS, 1976) or N6 salts (CHU et al., 1975), with Morel vitamins and 3% sucrose. A fresh medium was placed in the plastic bottle every 4 weeks. All media had their pH adjusted to 5.8 prior to autoclaving at 120 °C for 20 min. The cultures were placed in a growth room, under the conditions of light and temperature described above.

Morphological analysis

After 12 weeks, the plantlets were subjected to analysis (Figure 1C). All completely formed leaves and all roots were counted and the three largest leaves and roots were measured. The material was dried in an oven at 35 °C with air circulation to constant weight and the dry mass (DM) was weighed.

Mineral analysis

The DM of the plantlets was pulverized in a mortar grinder and 0.3 g were used for extraction in a Block Digester with nitric acid and hydrogen peroxide (SARRUGE & HAAG, 1974). After digestion, P, K, Mg, Mn, Cu, Fe and Zn were quantified in an Optical Emission Spectrometer (Varian 720-ES). For N,
0.15 g were weighed and quantification done in an Elemental Analyzer (Vario EL III).

**Statistical procedure**

The experiment consisted in a completely randomized design, with five repetitions comprising one bottle with 20 plantlets each, totaling 100 plantlets per culture medium. The data of morphological and mineral analysis were submitted to ANOVA and the means compared by Tukey’s test at 5% significance, using Assistat software (SILVA & AZEVEDO, 2016).

**RESULTS AND DISCUSSION**

After 12 weeks of culture, results showed that Y3 and MS media gave better results than N6 medium for almost all the parameters analyzed (Table 2). Plantlets cultured in MS and Y3 media were visually more vigorous and had less haustorial tissue formation (Figure 2 A, B) than the plantlets from N6 medium, which were also more oxidized (data not shown). The DM of plantlets cultured in Y3 medium was higher than those in N6 medium (152%), due to a lower root and leaf formation in N6 medium (Table 2 and Figure 2 C).

Results of the peach palm temporary immersion culture system showed that Y3 and MS media gave better results than N6 medium for leaf numbers and length, and for root numbers (Table 2). MS medium is widely used in in vitro culture and is a high salt medium in comparison to many other formulations, with high levels of N and of some micronutrients, particularly B and Mn (COHEN, 1995). However, the MS mineral formulation is not optimal for all plant species.

The DM value was 57.1% higher between the MS and N6 medium and 60.4% higher between the Y3 and MS medium (Table 2). This was an interesting result, considering that Y3 medium has...
half the total N concentration of MS medium. This may be due to the presence of a higher content of Na in the Y3 medium that can enhance nitrate uptake by roots, as it was shown in Beta vulgaris var. cicla L. (KABURAGI et al., 2014). Moreover, the lack of Na decreased the leaf size of peach palm seedlings growing in a nutrient solution (FERNANDES et al., 2013). This element is reported as being important for palm species, as it increases growth and productivity (FERNANDES et al., 2002).

This response could be related to the Na effect in increasing soluble carbohydrates concentration in the cell, which in turn favors cell expansion in leaves and stimulates enzymatic activity in roots (BROADLEY et al., 2012b).

The Y3 medium was first developed for Cocos nucifera (EEUWENS, 1976) and offers good results for other monocots. This medium has high levels of K, Na and Cl. The root number and length were higher in Y3 medium than in the other

Table 1 - Mineral composition of the three culture media used in a temporary immersion system.

| Macronutrients | MS | Y3 | N6 |
|----------------|----|----|----|
| NO₃⁻           | 39.406 | 19.979 | 27.991 |
| NH₄⁺           | 20.6138 | 10.001 | 4.058 |
| K⁺             | 18.793 | 39.993 | 27.991 |
| Mg²⁺           | 1.5 | 1.002 | 0.366 |
| PO₄³⁻          | 1.25 | 1.743 | 2.939 |
| Ca²⁺           | 2.99 | 2.99 | 1.1 |
| SO₄²⁻          | 1.711 | 1.168 | 0.2 |

| Micronutrients | MS | Y3 | N6 |
|----------------|----|----|----|
| Na⁺            | 0.202 | 1.814 | 0.002 |
| Cl⁻            | 5.9 | 30.01 | 2.51 |
| F              | 0.005 | 0.05 | 0.0048 |
| BO₃⁻           | 0.1002 | 0.050 | 0.0258 |
| Mn²⁺           | 0.132 | 0.132 | 0.012 |
| Zn²⁺           | 0.03 | 0.03 | 0.0052 |
| MoO₄²⁻         | 0.0011 | 0.0011 | 0 |
| Cu²⁺           | 0.0001 | 0.0006 | 0 |
| Co²⁺           | 0.0001 | 0.0010 | 0 |
| Fe²⁺           | 0.01 | 0.01 | 0.01 |
| NH₄⁺/NO₃⁻      | 0.52 | 0.50 | 0.14 |
| Total N        | 60 | 30 | 32 |

Table 2 - Growth parameters of Bactris gasipaes Kunth plantlets after 12 weeks of culture in three different media in a twin flask temporary immersion system.

| Parameters/Medium | Culture media |
|-------------------|---------------|
| Leaf number (¹)   | MS (²) N6 (²) |
| Root number (¹)   | Y3 (²) N6 (²) |
| Leaf length (cm)  | 2.86 a 1.51 b |
| Root length (cm)  | 0.66 a 0.47 a |
| Dry mass (g)      | 0.1326 ab 0.0844 b |

(¹) Leaf or root mean number per plantlet. (²) Means compared by Tukey’s test at 5% significance.
media (Table 2), which may be related to a higher concentration of K in this medium (Table 1). K plays a role in maintaining cell turgor and in protein synthesis and its presence is related to increased root growth in palm species (MEURER, 2006). In the oil palm (*E. guineensis* var. *Dura*), Muniran et al. (2008) observed a higher rooting rate when the plantlets were cultured in *y3* medium than in *MS* and *N6* media.

Considering the mineral analysis, there were significant differences among the main nutrient contents of the plantlets cultured in the different culture media (Table 3). The N, Mg, Mn, Zn and Ca contents were higher in the plantlets cultured in MS medium than in N6 media. N and Zn content were also higher in MS cultured plantlets than in Y3 plants. The K content was higher in plantlets cultured in Y3 media.

Figure 2 - *Bactris gasipaes* Kunth plantlets obtained from somatic embryogenesis and grown in a temporary immersion system. (a). MS salts (b). Y3 salts. (c). N6 salts. Arrow head: haustorium. Bar = 114 mm.
and N6 media and plantlets cultured in N6 medium had a higher P content. Only Fe content was similar in all treatments (Table 3). The presence of a higher chloride concentration in y3 medium may also have favored higher plantlet growth in this medium, even with a lower N concentration than in MS medium (Table 1). Palm species require a high chloride concentration and the lack of this element is related with vigor lost and premature senescence. Plant chloride deficiency can cause root growth decrease, related to root cell extension and decrease of root ion absorption (BROADLEY et al., 2012a).

Plantlets cultured in N6 medium had the lowest growth rates (Table 2). In addition to containing the lowest total N content, N6 medium has lower Ca, B, Mn, Na, Cl and Zn concentrations when compared with MS and Y3 media, besides the lack of Cu, Co and Mo (Table 1). Matos et al. (2013) showed the importance of these three elements for peach palm culture in a mineral solution. Moreover, it was the lack of Cu and B that influenced plantlet growth most negatively, especially leaf growth, and provoked chlorosis (MATOS et al., 2013). B deficiency reduces absorbed light energy utilization during photosynthesis, in addition to inhibiting apical meristem growth and decreasing leaf expansion and root elongation. Cu is a component of several enzymes related to the photosynthetic process and N metabolism and its deficiency lowers the photosynthetic rate and the level of carbohydrates as well as causing problems in cell wall lignification (BROADLEY et al., 2012a).

Mg and Zn were in lower concentrations in peach palm plantlets cultured in Y3 and N6 media, which could be related to problems in plantlet development (Table 3). The lack of Mg and Zn in peach palm plants growing in nutrient solution caused growth decrease and chlorosis, and was associated with the DM reduction (MATOS et al., 2013). Mg is present in several important enzymes of protein synthesis, lignin biosynthesis and in the photosynthetic process (BROADLEY et al., 2012b). Zn is also present in enzymes of several classes, acting in DNA and protein synthesis, carbohydrate metabolism, auxin synthesis and cell membrane maintenance (BROADLEY et al., 2012b).

After N, K and P were the most important nutrients for supporting peach palm growth in field conditions, followed by Ca and Mg (DEENIK et al. 2000); although, a balance between the nutrients must be established. The phosphate concentration in MS medium is inadequate for several cultures and its excess may reduce growth, possibly because it precipitates Ca and some micronutrients or decreases its absorption by the plant (GEORGE & DE KLERK, 2008). In our study, a higher P concentration could be observed in plantlets cultured in N6 medium but their growth was not improved when compared to MS and Y6 media (Table 3).

**CONCLUSION**

The salts of MS and Y3 culture media were the most suitable for peach palm growing in a temporary immersion system. K is an important element for peach palm in vitro culture. Y3 medium is recommended because it contains higher concentrations of that. Although the results obtained for plantlets

### Table 3 - Mineral analysis of *Bactris gasipaes* Kunth plantlets cultured for 12 weeks in three different media in a twin flask temporary immersion system.

| Element (1) | MS (2) | Y3 (2) | N6 (2) |
|-------------|--------|--------|--------|
| N           | 66.690 a | 41.460 c | 56.480 b |
| K           | 41.217 b | 59.379 a | 57.03 a |
| P           | 4.726 b  | 5.701 b  | 7.722 a |
| Ca          | 2.561 a  | 1.931 ab | 1.601 b |
| Mg          | 1.177 a  | 0.910 ab | 0.806 b |
| Fe          | 0.353 a  | 0.337 a  | 0.464 a |
| Mn          | 0.089 a  | 0.0645 ab| 0.046 b |
| Zn          | 0.107 a  | 0.079 b  | 0.059 b |

(1) Values in g kg⁻¹ dry material. (2) Means compared by Tukey’s test at 5% significance.
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DECLARATION OF CONFLICT OF INTERESTS

The authors declare no conflict of interest. The funding sponsors had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, and in the decision to publish the results.

AUTHORS’ CONTRIBUTIONS

All authors contributed equally for the conception and writing of the manuscript. All authors critically revised the manuscript and approved the final version.

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