Fine Mapping of the Gene Controlling the Fruit Skin Hairiness of *Prunus persica* and Its Uses for MAS in Progenies

Zhenhua Lu †, Lei Pan †, Bin Wei, Liang Niu, Guochao Cui, Luwei Wang, Wenfang Zeng and Zhiqiang Wang *

National Peach and Grape Improvement Center, Key Laboratory of Fruit Breeding Technology of Ministry of Agriculture, Zhengzhou Fruit Research Institute, Chinese Academy of Agricultural Sciences, Zhengzhou 450009, China; luzhenhua@caas.cn (Z.L.); panlei@caas.cn (L.P.); weibinbin2016@163.com (B.W.); niuliang@caas.cn (L.N.); cuiquochao@caas.cn (G.C.); 15839552412@163.com (L.W.); zengwenfang@caas.cn (W.Z.)

* Correspondence: wangzhiqiang@caas.cn
† These authors contributed equally to this manuscript.

Abstract: The fruit skin pubescence of *Prunus persica* is an economically important characteristic and comprises the classification criteria. The mapping and identification of a complete linkage marker to the fruit skin trichome trait locus of peach fruit are critical for the molecular marker-assisted selection for peach/nectarine. In this study, the BC$_1$ population was constructed from the parents “Zhongyou No. 4”, the recurrent parent, and “Baihuashanbitao”, the non-recurrent parent. Based on the 38 BC$_1$ individuals’ phenotypes and their genotyping using next-generation sequencing, the G (Glabrous skin) locus of the gene was first identified between 14.099 and 16.721 Mb on chromosome 5. Using other individuals of this population, the gene was fine-mapped in the range of 481 kb with SNP markers. Based on the resequencing data of other cultivars (lines), the candidate SNP in the gene Prupe.5G196400 was obtained. Subsequently, the SNP marker was designed and applied to natural and hybrid peach populations. Via genotyping analysis, we confirmed co-segregation between the peach/nectarine phenotype, which was used in the identification of peach or nectarine with 100% accuracy.

Keywords: peach; fine mapping; fruit skin hairiness

1. Introduction

Peach (*Prunus persica* L. Batsch) is one of the most important deciduous fruit trees in China, accounting for 826,325 ha of cultivation area in 2018. It is in high demand because of its flavor and high amount of nutrients [1]. As one of its important phenotypic characteristics, skin hairiness or non-hairiness (fuzziness) is considered standard for peach classification [2,3]. Before 1990, hairy peach varieties were popular and widely cultivated in China [4]. In recent years, the production and commercialization of nectarine varieties in China has increased, and some new varieties have been released. Nectarine, characterized by the absence of fruit trichomes, have a bright appearance and low allergenicity [5,6], creating high demand for them. Peaches and nectarines are native to China, with a long cultivation history [7]. However, the production and commercialization of nectarine varieties in China only started recently. Since the 1980s, breeders in China have used pollen from the USA and Italy to cross the hybrids “Jingyu” and “Qiuyu”, which has resulted in the release of several varieties [8].

To locate and identify the genes controlling the fruit skin hairiness of *Prunus persica*, markers with which to finely map the peach/nectarine locus based on the hybrid population have been developed. The RAPD marker OPZ-03 was first screened [9], and the genetic distance between OPZ-03 and the G/g locus was 23.4 cM. Since then, many Chinese researchers have been developing several markers, such as OPP20-2200, SCP20-2258, UDP96-018, and CPSCT030, which have genetic distances to the G/g locus as short as 4.5 cM. However, the genotyping results for these markers in natural peach varieties...
are not satisfactory, and the related candidate genes have not been screened. The genetic characteristics of nectarine were first reported by Blake et al. in 1932; they are controlled by the G locus and are recessive to normal peach [10]. The G (nectarine) locus was first mapped at the proximal end (bottom) of linkage group 5 [11] and is located at the distal part of linkage group 5, spanning a region from 15,126,681 to 16,315,341 (1.189 Mb), with reference to the peach genome [2,12–14]. An indel in candidate gene PpeMYB25, controlling the peaches’ skin hair, has been identified. The insertion of the LTR transcription factor in the third exon of PpeMYB25 in the MYB gene family led to the appearance of a recessive hairless phenotype [2]. Recently, Cao et al. [15] associated the trait at position 17,576,893 Mb on Chromosome 3, using GWAS, and found the candidate gene ppa010316m, accounting for 80% of the explained variation.

The cultivation of peaches, as a perennial fruit tree, using traditional breeding methods is impeded by the long juvenile stage, significantly restricting the development of the peach industry [16,17]. In this study, we used the BC₁ population from the peach cultivar “Zhongyou No. 4” to finely map the gene controlling skin hairiness (G) and to find molecular markers linked to the trait, with the aim of establishing a reliable SNP maker for peach/nectarine identification. Our results provide technical support for parental selection and early identification of hybrid progenies in peach/nectarine breeding.

2. Results

2.1. Genotyping with NGS

The peach or nectarine characteristics in the BC₁ population were distinguished in the field based on fruit skin appearance. For further observation, we used a stereomicroscope to observe the differences in the fruit skin between peaches and nectarines. At 30× magnification, the peach cultivars showed white filiform growth, which was absent in the nectarine cultivars. In the progenies, the phenotype was segregated into hairy and non-hairy (Figure 1). The phenotypes of nectarines were labeled “a” and those of peaches were labeled “h”. The parents and 38 BC₁ populations were sequenced with 70× and 10× depths, respectively. The peach genome (version 2.0) was used to select the reference genome for genotyping 38 BC₁ individuals, with scanning of the whole peach genome (chromosomes 1–8). We found a region with the same genotypes and phenotypes on chromosome 5 (Figure 2; Supplementary Data S1), with a complete linkage region with peaches/nectarines (Figure 3; Supplementary Data S2).

![Figure 1](image1.png)

Figure 1. Comparing the differences between hairy fruit skin (left) and non-hairy skin (right) observed using a zoom-stereo microscope at a magnification of 30×.

2.2. Mapping and Fine-Mapping of Hairiness/Non-Hairiness Based on Introgression Lines

To acquire closely linked SNP makers, seven SNP primer pairs (Table 1) were designed in the range of Pp05 14.099–16.721 Mb, based on the deep sequencing data for two parents. We used the 129 individuals from the Baihuashanbitao × (Zhongyou No. 4 and Baihuashanbitao) BC₁ segregating population to finely map the locus. Both SNPs that were homozygous in “Zhongyou No. 4” and heterozygous in “Baihuashanbitao” were
selected based on the parental resequencing data. Based on the SNP markers using Sanger sequencing, we identified 10 recombinants on both sides of the desired locus. Of these recombinants, the target region was narrowed down to the 481 kb region on chromosome 5, which was delimited to an interval between SNP-15760886 and SNP-16242580 by using these recombinants in the BC1 population. We obtained two SNP markers that were completely linked to the peach/nectarine phenotypes, namely, Pp05-SNP-15858687 and Pp05-SNP-15959172 (Figure 4).

Figure 2. Mapping of the locus based on the genotyping and phenotype of 38 BC1 introgression individuals and two parents. Red indicates the introgression fragment from “Baihuahsanbitao”, blue indicates the fragments from “Zhongyou No. 4”. The letter “a” indicates homozygous nectarines; “h” indicates heterozygous peaches.

Figure 3. Complete linkage region with fruit skin hairiness/non-hairiness. Red indicates the introgression fragment from “Baihuahsanbitao”, blue indicates the fragments from “Zhongyou No. 4”. The letter “a” indicates homozygous nectarines; “h” indicates heterozygous peaches.
2.3. Candidate Gene Confirmation Based on the Resequencing Data of Two Parents and Other Cultivars (Lines)

The reference peach genome database (version 2.0) contained 85 known transcripts within 481 kb of the fine-localization region. To acquire the candidate gene, we analyzed the correlation between the genotypes and phenotypes of the parents and 38 BC1 individuals. An SNP mutation site (G-A) within the Prupe.5G196400 CDS region was identified based on resequencing data for two parents (Supplementary Figure S1). To validate the candidate gene, the SNP variation in ORF of Prupe.5G196400 for 38 BC1 population individuals and for other 16 cultivars (lines) was further verified (Table 2). The results show that the phenotype and genotype matching rate of the site in both parents and the BC1 population was 100% (Supplementary Figure S2). In the finely mapped region, 86 genes were contained (Supplementary Data S3) and we determined Prupe.5G196400, with four exons and three introns, encoding a protein of unknown function, as a confident molecular marker to distinguish the peach/nectarine traits.

2.4. MAS of Hairiness/Non-Hairiness in Peach Breeding

To confirm the feasibility of the identification of peach and nectarine varieties with the SNP maker in the coding region of Prupe.5G196400, 122 cultivars or lines were tested. Based on the Sanger sequencing results, there were three genotypes of SNPs (G/G, T/G, and T/T) in 122 cultivars (lines) (Figure 5). Among them, G/G and T/G represented homozygous and heterozygous genotypes of peach, respectively, and the T/T represented the homozygous genotypes of nectarines. The genotypes of SNP-Prupe.5G196400 in the 122 peach germplasm resources are shown in Supplementary Data S4. The coincidence rate of phenotypes and genotypes was 100%; thus, SNP-Prupe.5G196400 was completely linked to the G locus and can be used in molecular marker-assisted selection of target varieties.

In addition, three populations were used to identify the phenotype of the offspring, using the SNP marker. The result indicates that the SNP (T/T, G/T, or G/G) markers can distinguish the phenotypes of the individuals with 100% accuracy (Table 3).

---

### Table 1. Primers used in fine mapping for peach skin hairiness.

| Physical Position | Primer Sequence (5’-3’) | Product Size (bp) | SNP Type |
|------------------|------------------------|------------------|----------|
| Pp05-SNP-14562747 | GGATTAGTGGAGTGGAACGC  | AAGCATCCAGCCAAAACACC | 710 | G/T |
| Pp05-SNP-15464780 | GAAGAGCAAACTTGGGACCTC | TTTTGAACCACTTGGGGAAGC | 763 | C/T |
| Pp05-SNP-15760886 | TGACGATTTGAGAGATTGATCG | TTTTTGTGGGAAGAGGAAGG | 778 | C/T |
| Pp05-SNP-15858687 | ACAGCGTTCGGCTATGAACC | TTTCTTGGGAGTTTTGTGTGC | 866 | A/G |
| Pp05-SNP-15959172 | CTCTTACGACCAAGAACCAACC | GTTGGTGGAGTGGGAGAAGC | 815 | C/T |
| Pp05-SNP-16242580 | ACTGGTGGTTTGTTGGTTGG | TTTCCATACATGTCTTAAAGGTTC | 633 | C/G |
| Pp05-SNP-15858687 | ACAGCGTTCGGCTATGAACC | TTTCTTGGGAGTTTTGTGTGC | 866 | A/G |
| Pp05-SNP-14562747 | GGATTAGTGGAGTGGAACGC  | AAGCATCCAGCCAAAACACC | 710 | G/T |

---

**Figure 4.** Genotype and phenotype of ten recombinants and two parents using 129 BC1 Individuals for fine-mapping.
Table 2. Genotyping results for 16 cultivars (lines) regarding skin hairiness.

| Cultivars (Lines)     | Phenotype  | Genotype |
|-----------------------|------------|----------|
| Zhongyou No. 4        | Non-hairy  | T/T      |
| Baihuashanbitao      | Hairy      | G/G      |
| 10–7                 | Hairy      | T/G      |
| 96–51                | Hairy      | T/G      |
| Bairuuyu             | Hairy      | G/G      |
| ludong-2-04          | Non-hairy  | T/T      |
| P7-12-03             | Non-hairy  | T/T      |
| Shanza-02            | Hairy      | T/G      |
| Shuipingzhi          | Hairy      | T/G      |
| Weni2                | Hairy      | G/G      |
| yb144                | Hairy      | T/G      |
| Zhongpan No. 01      | Hairy      | T/G      |
| Zhongtao No. 05      | Hairy      | T/G      |
| Zhongyou No. 08      | Non-hairy  | T/T      |
| Zhongyou No. 13      | Non-hairy  | T/T      |
| Zhongyou No. 20      | Non-hairy  | T/T      |

Figure 5. Three types of SNPs (T/T, T/G, and G/G) linked to peaches’ skin hairiness/non-hairiness in 122 cultivars (lines), based on Sanger sequencing results.

Table 3. The SNP marker was used for MAS in three segregation populations to predict the phenotype with 100% accuracy.

| Segregation Populations | Individuals | Hairy | Non-Hairy | Accuracy |
|-------------------------|-------------|-------|-----------|----------|
| Zhongyou No. 8          | Beijingduan (hairy) | 30    | 13        | 17       | 100%     |
| 09-bei8-25 (hairy)      | Zhongtaobaiyu (hairy) | 196   | 196       | 0        | 100%     |
| Zhongtao No. 5 (hairy)  | Zhongyou No. 15 (non-hairy) | 15    | 0         | 15       | 100%     |
3. Discussion

In China, peaches have been cultivated for more than 3000 years [18]. Because of their long juvenile period, it is important to determine tightly linked molecular markers to predict some of the economic traits, especially fruit traits, with the aim of establishing molecular marker-assisted selection in breeding [19]. With the development of next-generation sequencing, genes controlling the economic traits of fruit trees have been discovered. Some of these traits are fruit skin “anthocyanin defect” in grape [20] and the branch angle (column and weeping type) [21,22], internode length (dwarfing) [23], flesh texture [24], flesh color [25–27], and fruit shape (peaches or flat peaches) [28] in peach. In some plants, trichomes are widely distributed in the stem, leaves, fruits, and seeds; they originate from epidermal cells and play a role in protection against biotic and abiotic stressors, with a long evolutionary history, and they protect plants against insect, fungal, and excessive light damage [29–32]. For instance, the tomato mutant with hairless leaves shows decreased resistance to pest insects [33]. Although in peaches, skin hairiness enhances resistances to biotic and abiotic stress, hairy peaches have greater allergenic potential than nectarines [34].

Recently, several genes related to the development of fruit pubescence have been identified and cloned. For example, Payne et al. found that mutations of TTG1 and GL1, two genes that play a key role in the growth of Arabidopsis thaliana hairs, affect the growth of hairs [35]. Li et al. [36] identified Os05g19000 as a candidate gene for the hairiness of the glabrous leaf and hull mutants in rice. Skin hairiness or non-hairiness are commercial characteristics for peach fruit classification. In 2014, Vendramin et al. further narrowed the location range of the key loci of peach/nectarine traits to 1.1 cM and obtained the candidate gene PpeMYB25; the authors observed that the insertion of the 7 kb LTR reversal loci in exon 3 of PpeMYB25 gene was related to the recessive characteristic of nectarines [2], which is in line with our findings. However, no deletion of large fragments in PpeMYB25 locus was observed. An SNP in ORF of Prupe.5G196400 was found, which is tightly linked to the trait and could be effectively used to distinguish peach and nectarine varieties in germplasm resources.

In this study, we used next-generation sequencing and SNP markers to perform the fine mapping of the gene controlling peach/nectarine traits and obtained the candidate gene Prupe.5G196400. Although our gene has not yet been functionally verified, through our 122 peach varieties (lines) and four populations, this marker can distinguish the peach/nectarine characteristics of peaches with 100% accuracy. For this reason, we consider it as a potential candidate gene. Our results provide a foundation for the selection of early follow-up parents, variety identification, early phenotype prediction of progeny representative type, and variety selection in peach breeding.

4. Materials and Methods

4.1. Plant Material

The BC₁ lines generated from “Zhongyou No. 4”, the recurrent parent, and “Baihuashanbitao”, the non-recurrent parent, were generated in 2012. “Zhongyou No. 4” is a yellow-flesh and early-ripening nectarine cultivar bred by the Zhengzhou Fruit Research Institute, CAAS, whereas “Baihuashanbitao” is an ornamental peach from P. davidiana with female abortion, bred by the Beijing Forestry University.

The parents and 38 BC₁ individuals were phenotyped as hairy and non-hairy and subsequently genotyped using NGS for mapping. Within the mapped region, SNP markers were designed based on parental sequencing data and tested in the other 129 BC₁ individuals for fine mapping. To validate the candidate gene, resequencing data for 16 individuals were used, and 122 cultivars or lines were tested to confirm the linkage between genotypes and phenotypes.
4.2. DNA Extraction

For the recurrent (Zhongyou No. 4) and non-recurrent (Baihuashanbitao) parents and for the 38 BC\textsubscript{1} individuals, genomic DNA was extracted using the CTAB method, with slight modifications [37]. In the DNA precipitation step, pure ethanol was added, and the precipitations were picked with a toothpick to acquire high-quality DNA for next-generation sequencing. For the other BC\textsubscript{1} individuals and 122 cultivars (lines), genomic DNA was isolated with the simplified CTAB method, with one step of chloroform: isoamyl alcohol (v/v, 24:1) to remove potential contaminants.

4.3. Phenotyping

The fruit skin traits (hairiness/non-hairiness) of BC\textsubscript{1} individuals and the 122 cultivars for mapping and fine mapping were evaluated in the breeding pool (Xinxiang, Henan) by visualization. In addition, the skin was scanned at 30×, using a digital microscope VHX-7000 (Keyence, Osaka City, Japan).

4.4. NGS Genotyping and Data Analysis

We used 1.5 µg DNA per sample for DNA resequencing. Sequencing libraries were constructed using the Truseq Nano DNA HT sample preparation kit (Illumina, San Diego, CA, USA) following the manufacturer’s instruction; index codes were added to each sample. Briefly, the DNA sample was sheared by sonication into 350 bp fragments and purified. Each sample was then sequenced using HiSeq 2500 (Illumina, CA, USA), and 150 bp paired-end reads were generated. The peach genome (version 2.0) was used as the reference genome [12].

We applied the BWA (Burrows–Wheeler Aligner) [38] to align the clean reads of each sample against the reference genome (settings: mem -t 4 -k 32 -M -R). Alignment files were converted to BAM files using the SAMtools software 1.6 [39] (settings: -bS -t). Potential PCR duplications were removed using the SAMtools command “rmdup”. If multiple read pairs had identical external coordinates, only the pair with the highest mapping quality remained.

4.5. Introgression Line for Mapping

The homozygous SNPs, genotype AA and BB, in two parents were extracted, respectively, from the vcf files for SNPs. Based on the NGS data in the Excel file, a bin map was drawn to identify the introgression fragments from “Baihuashanbitao”. For the mapping analysis, the BC\textsubscript{1} individuals were divided into two phenotypic classes and then their graphical genotypes were compared in Excel file [40]. We identified chromosomes 1 to 8 to locate the candidate region, in which the genotypes were coincident with the phenotypes.

4.6. Primer Design, PCR Amplification, and SNP Genotyping

Based on the resequencing data of parents, the SNPs in the candidate region were selected. We used the Integrative Genomics Viewer 2.3 software [41] to read the BAM file to confirm the SNPs and to design the primers for fine mapping. Primer 3 (http://bioinfo.ut.ee/primer3-0.4.0/ accessed on 18 June 2020) was used to design the primers [42]. For SNP genotyping, the amplified fragment length was approximately 700 bp for Sanger sequencing, which was performed in Sangon Biotech (Shanghai, China); the SNPs were analyzed using the ContigExpress software (2000).

4.7. Completely Linked Markers in the Fine-Mapping Region

For the fine-mapped region, the genes were listed referencing the annotation in the GFF file of the peach genome (version 2.0) [43]. Variations in the genes within the fine-mapped region were identified in the BAM files of the two parents in accordance with phenotypes and genotypes, observed in the resequencing data for 16 cultivars, and confirmed in 122 cultivars (lines).
5. Conclusions

Using 35 BC1 individuals from “Zhongyou No. 4” as the recurrent parent and “Baihuashanbitao” as the non-recurrent parent, the G locus was mapped between 14.099 and 16.721 Mb on chromosome 5 and fine-mapped in the range of 481 kb with SNP markers, using the other 129 BC1 individuals of this population. A candidate SNP in the gene Prupe. 5G196400 was obtained based on the resequencing data of other cultivars (lines). The SNP marker was able to identify the phenotypes in 122 cultivars (lines). By genotyping analysis, the SNP was co-segregated with the fruit skin fuzzy phenotype and used for identification of peaches or nectarines in three segregation populations with 100% accuracy.

Supplementary Materials: The following are available online at https://www.mdpi.com/article/10.3390/plants10071433/s1. Figure S1: The IGV snapshot of SNP mutation in gene Prupe.5G196400 based on resequencing data of two parents, Figure S2: The IGV snapshot of SNP for 16 cultivars (lines) in candidate gene Prupe.5G196400, Data S1: Complete linkage region with fruit skin hairiness/non-hairiness based on the genotype and phenotype of 35 BC1 introgression individuals and two parents, Data S2: The genotype of 129 BC1 individuals, ten recombinants and two parents used for fine mapping. The fruit skin was labelled with hairiness and non-hairiness, Data S3: The genes listed in the mapped region, containing 86 genes, Data S4: The SNP marker was used to validate the genotype and phenotype in 122 cultivars (lines).

Author Contributions: Conceptualization, Z.L. and Z.W.; methodology, Z.L. and L.P.; validation, L.P., L.W. and B.W.; resources, W.Z., and L.N.; data curation, G.C.; writing—original draft preparation, Z.L. and B.W.; writing—review and editing, Z.L. and B.W. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the National Natural and Science Foundation of China (NSFC: 31870669, 32072532) and the Agricultural Science and Technology Innovation Program (CAAS-ASTIP-2019-ZFR1).

Conflicts of Interest: The authors declare no conflict of interest.

References
1. FAO. FAOSTAT-FAO’s Online Statistical Database; FAO: Rome, Italy, 2018.
2. Staff, T.P.O. A unique mutation in a myb gene cosegregates with the nectarine phenotype in peach. PLoS ONE 2014, 9, e90574.
3. Crelle, M.A.; Werner, D.J. Characterizing the Novel Fruit Surface Morphology of ‘Marina’ Peach Using Scanning Electron Microscopy. J. Am. Soc. Hortic. Sci. 1996, 121, 198–203. [CrossRef]
4. Janick, J.; Moore, J. Peach In: Advances in Fruit Breeding; Lafayette, W., Ed.; Purdue University: West Lafayette, IN, USA, 1975.
5. Alessandro, B.; Mara, V.; Filippo, D.F.; Angelo, R.; Carla, G.; Guido, M.; Gabriella, P.; Pietro, T. Different expression of Pp-LTP1 and accumulation of Pru p 3 in fruits of two Prunus persica L. Batsch Genotypes 2006, 171, 106–113.
6. Carnés, J.; Fernandez-Caldas, E.; Gallego, M.T.; Ferre, A.; Cuesta-Herranz, J. Pru p 3 (LTP) content in peach extracts. Allergy 2002, 57, 1071–1075. [CrossRef] [PubMed]
7. Yoon, J.; Liu, D.; Song, W.; Liu, W.; Zhang, A.; Li, S. Genetic diversity and ecogeographical phylogenetic relationships among peach and nectarine cultivars based on simple sequence repeat (SSR) markers. J. Am. Soc. Hortic. Sci. 2006, 131, 513–521. [CrossRef]
8. Ma, R.; Yu, M.; Tang, X.; Guo, H.; Zhou, J.; Zhao, M. Advances in nectarine breeding. J. Fruit Sci. 2000, 17, 214–219.
9. Chaparro, J.X.; Werner, D.J.; O’Malley, D.; Sederoff, R.R. Targeted mapping and linkage analysis of morphological isozyme, and RAPD markers in peach. Theor. Appl. Genet. 1994, 87, 805–813. [CrossRef] [PubMed]
10. Blake, M.A. The JH Hale peach as a parent in peach crosses. Proc. Natl. Acad. Sci. USA 1932, 29, 131–136.
11. Dirlewanger, E.; Pronier, V.; Parvery, C.; Rothan, C.; Guye, A.; Monet, R. Genetic linkage map of peach [Prunus persica (L.) Batsch] using morphological and molecular markers. Theor. Appl. Genet. 1998, 97, 888–895. [CrossRef]
12. Verde, I.; Abbott, A.G.; Scalabrino, S.; Jung, S.; Shu, S.; Marroni, F.; Zhebentyayeva, T.; Dettori, M.T.; Grimwood, J.; Cattonaro, F. The high-quality draft genome of peach (Prunus persica) identifies unique patterns of genetic diversity, domestication and genome evolution. Nat. Genet. 2013, 45, 487–494. [CrossRef]
13. Dirlewanger, E.; Cosson, P.; Boudrehi, K.; Renaud, C.; Capdeville, G.; Tazuzin, Y.; Laigret, F.; Moing, A. Development of a second-generation genetic linkage map for peach [Prunus persica (L.) Batsch] and characterization of morphological traits affecting flower and fruit. Tree Genet. Genomes 2006, 3, 1–13. [CrossRef]
14. Dantec, L.; Cardinet, G.; Bonet, J.; Fouché, M.; Boudrehi, K.; Monfort, A.; Poessel, J.L.; Moing, A.; Dirlewanger, E. Development and mapping of peach candidate genes involved in fruit quality and their transferability and potential use in other Rosaceae species. Tree Genet. Genomes 2010, 6, 995–1012. [CrossRef]
15. Cao, K.; Zhou, Z.; Wang, Q.; Guo, J.; Zhao, P.; Zhu, G.; Fang, W.; Chen, C.; Wang, X.; Wang, X. Genome-wide association study of 12 agronomic traits in peach. Nat. Commun. 2016, 7, 13246. [CrossRef] [PubMed]

16. Kale, S.M.; Pardeshi, V.C.; Kadool, N.Y.; Ghorpade, P.B.; Jana, M.M.; Gupta, V.S. Development of genomic simple sequence repeat markers for linseed using next-generation sequencing technology. Mol. Breed. 2012, 30, 597–606. [CrossRef]

17. Aranazana, M.J.; Ila, E.; Howad, W.; Aris, P. A first insight into peach [Prunus persica (L.) Batsch] SNP variability. Tree Genet. Genomes 2012, 8, 1359–1369. [CrossRef]

18. Li, H.; Keightly, D.N. The domestication of plants in China: Ecogeographical considerations. In The Origins of Chinese Civilization; University of California Press: Berkeley, CA, USA, 1983; Volume 1, pp. 21–64.

19. Aranazana, M.J.; Abbassi, E.K.; Howad, W.; Aris, P. Genetic variation, population structure and linkage disequilibrium in peach commercial varieties. BMC Genet. 2010, 11, 69. [CrossRef] [PubMed]

20. Kobayashi, S. Retrotransposon-Induced Mutations in Grape Skin Color. Science 2004, 304, 982. [CrossRef]

21. Hollender, C.A.; Pascal, T.; Tabb, A.; Hadiarto, T.; Dardick, C. Loss of a highly conserved sterile alpha motif domain gene (weep) results in pendulous branch growth in peach trees. Proc. Natl. Acad. Sci. USA 2018, 115, 201704515. [CrossRef]

22. Dardick, C.; Callahan, A.; Horn, R.; Ruiz, K.B.; Zhebentyayeva, T.; Hollender, C.; Whitaker, M.; Abbott, A.; Scorza, R. PpeTAC1 promotes the horizontal growth of branches in peach trees and is a member of a functionally conserved gene family found in diverse plants species. Plant J. 2013, 75, 618–630. [CrossRef]

23. Brandi, F.; Bar, E.; Mourgues, F.; Horvath, G.; Turcsi, E.; Giuliano, G.; Liverani, A.; Tartarini, S.; Lewinsohn, E.; Rosati, C. Study of ‘Redhaven’ peach and its white-fleshed mutant suggests a key role of CCD4 carotenoid dioxygenase in carotenoid and norisoprenoid volatile metabolism. BMC Plant Biol. 2011, 11, 24. [CrossRef]

24. Pan, L.; Zeng, W.; Niu, L.; Lu, Z.; Wang, X.; Liu, H.; Cui, G.; Zhu, Y.; Chu, J.; Li, W. PpYUC11, a strong candidate gene for the down stream of a ppofp1 gene is responsible for flat fruit shape in peach. Plant Mol. Biol. Rep. 2013, 31, 1166–1175. [CrossRef]

25. Falchi, R.; Vendramin, E.; Zanon, L.; Scalabrin, S.; Cipriani, G.; Verde, I.; Vizzotto, G.; Morgante, M. Three distinct mutational mechanisms acting on a single gene underpin the origin of yellow flesh in peach. Plant J. 2013, 76, 175–187. [CrossRef] [PubMed]

26. Zhou, H.; Ma, R.; Gao, L.; Zhang, J.; Zhang, A.; Zhang, X.; Ren, F.; Zhang, W.; Liao, L.; Yang, Q. A 1.7-mb chromosomal inversion downstream of a ppoF1 gene is responsible for flat fruit shape in peach. Plant Biotechnol. J. 2020, 19, 192–205. [CrossRef] [PubMed]

27. Fang, W.; Chen, C.; Wang, X.; Wang, X. Genome-wide association study of agronomic traits in peach. J. Exp. Bot. 2010, 61, 1053–1064. [CrossRef] [PubMed]

28. Oh, G.D.; Shim, E.J.; Jun, S.J.; Park, Y.D. Development of SNP Molecular Markers Related to Seed-hair Characteristic Based on EST Sequences in Carrot. Korean J. Hortic. Sci. Technol. 2013, 31, 80–88. [CrossRef]

29. Balcke, G.U.; Bennewitz, S.; Bergau, N.; Athmer, B.; Henning, A.; Majovsky, P.; Jiménez-Gómez, J.M.; Hoehenwarter, W.; Tissier, A. Multi-Omics of Tomato Glandular Trichomes Reveals Distinct Features of Central Carbon Metabolism Supporting High Productivity of Specialized Metabolites. Plant Cell 2017, 29, 960–983. [CrossRef]

30. Schilmiller, A.L.; Miner, D.P.; Larson, M.; McDowell, E.; Gang, D.; Wilkerson, C.; Last, R.L. Studies of a Biochemical Factory: Tomato Trichome Deep Expressed Sequence Tag Sequencing and Proteomics. Plant Physiol. 2010, 153, 1212–1223. [CrossRef]

31. Martin, C.; Glover, B.J. Functional aspects of cell patterning in aerial epidermis. Curr. Opin. Plant Biol. 2007, 10, 70–82. [CrossRef]

32. Jin, J.; Gao, L.; Zhao, L.; Gao, Z.; Li, X.; Xie, H.; Ni, J.; Gan, K.; Wu, S.; Ye, Z. Selection of Pru p 3 hypoallergenic peach and nectarine varieties. Allergy 2020, 75, 1256–1260. [CrossRef] [PubMed]

33. Payne, C.T.; Zhang, F.; Lloyd, A.M. GL3 encodes a bHLH protein that regulates trichome development in arabidopsis through interaction with GL1 and TTG1. Genetics 2000, 156, 1349–1362. [CrossRef] [PubMed]

34. Li, W.; Wu, J.; Weng, S.; Zhang, D.; Zhang, Y.; Shi, C. Characterization and fine mapping of the glabrous leaf and hull mutants (gl1) in rice (Oryza sativa L.). Plant Cell Rep. 2010, 29, 617–627. [CrossRef]

35. Doyle, J.J.; Doyle, J.L. A rapid DNA isolation procedure for small quantities of fresh leaf tissue. Phytochem. Bull. 1987, 19, 11–15.

36. Robinson, J.T. Integrative genomics viewer. Nat. Biotechnol. 2011, 29, 24–26. [CrossRef]
42. Koressaar, T.; Remm, M. Enhancements and modifications of primer design program Primer3. *Bioinformatics* **2007**, *23*, 1289–1291. [CrossRef]

43. VerdeI, G.; Jenkins, J.; Dondini, L.; Micali, S.; Pagliarani, G.; Vendramin, E.; Paris, R.; Aramini, V.; Gazza, L.; Rossini, L. The Peach v2.0 release: High-resolution linkage mapping and deep resequencing improve chromosome-scale assembly and contiguity. *BMC Genom.* **2017**, *18*, 225. [CrossRef]