Gene Set Enrichment Analysis and Ingenuity Pathway Analysis to verify the impact on Wnt Signaling Pathway in Taodan Granules Treated Psoriasis

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Research Article

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Abstract

**Background:** Taodan granules (TDGs), traditional Chinese herbals, are effective in treating psoriasis. However, mechanisms of TDGs remain indistinct. The current study aims to indicate the molecular mechanisms of TDGs in treating psoriasis.

**Methods:** Primarily, transcriptional profiling was applied to identify differentially expression genes (DEGs). The following was that we used Gene Set Enrichment Analysis (GSEA) and Ingenuity Pathway Analysis (IPA) analysis for functional enrichment analysis. Eventually, RT-PCR was performed to validation.

**Results:** The results revealed that TDGs could regulated the Wnt signaling pathway to ameliorate skin lesions of imiquimod (IMQ)-induced psoriatic mouse models. IPA core network associated with Wnt signaling pathways in TDGs for psoriasis was established. Thereinto zeste homolog (EZH) 2, CTNNB1, TP63, and WD repeat domain (WDR) 5 could be considered as upstream genes in the Wnt signaling pathway.

**Conclusions:** The Wnt signaling pathway with these above upstream genes might be potential therapeutic targets of TDGs for psoriasis.

1. **Background**

Psoriasis is a common chronic and intractable skin disease, while the study of psoriatic pathogenesis, diagnosis and treatment remains the focus and difficulty of dermatology at present. With high prevalence of psoriasis, the estimate of psoriatic prevalence in adults globally ranged from 0.51–11.43%, unevenly distributed geographically, more frequently occurred in high income regions with older populations [1, 2]. Moreover, psoriasis is not only accumulated skin, accompanied by a variety of comorbidities (cardiovascular disease, hepatic disease, nephropathy, etc.) [3], fearfully affected the quality of life for patients [4], as well as caused serious economic burden with average $12,523 annual cost of all-cause health care per psoriatic patient [5].

Several studies have identified interleukin (IL)-17 and IL-23 as key drivers of psoriasis, therefore, immunotherapy targeting of IL-17 and IL-23, together with Tumor Necrosis Factor (TNF)-α through biologic therapies, recognized as the most effective therapeutic care in clinic at present for moderate to severe psoriasis [6]. However, biological preparations were shown numerous adverse reactions and contraindications with high relapse of 94.7% after 18 months of discontinuation in the treatment of psoriasis [7]. Therefore, exploitation of more effective therapeutic approaches is imperative.

Chinese herbal medicine has been widely used in the psoriatic treatment. Taodan granules (TDGs) was formulated since Chinese medicine (CM) theory, composed of *Salvia miltiorrhiza Bunge, Curcuma aeruginosa Roxb., Astragalus mongholicus Bunge, Glycyrrhiza inflata Batalin and Angelica sinensis (Oliv.) Diels, Conioselinum anthriscoides “Chuanxiong”, Prunus persica (L.) Batsch, Cyathula officinalis K.*
C. Kuan, and Smilax china L.. Our preliminary studies indicated that the improvement rate of psoriasis area and severity index (PASI) score in psoriatic patients treated with TDGs was 76.64% [8, 9]. Ru et al. [10] conducted a multicenter, double-blind randomized controlled study of TDGs in the treatment of plaque psoriasis (ClinicalTrials.gov, No. NCT03942198) to evaluate the efficacy and recurrence. TDGs proved to down-regulate the expression of IL-2, IL-4, IL-6, secretion of neuropeptide and other psoriatic phenotypes [8, 9, 11], while Tanshinone IIA, the main ingredient of Salvia miltiorrhiza Bunge, could inhibit keratinocyte (KC) proliferation, and induce cell apoptosis together with cell cycle arrest [12]. However, the regulatory mechanisms of TDGs are still unclear and need further investigations.

Recently, transcriptome sequencing has become a routine method for identifying numerous genes regulated by specific medications. Previously, we applied transcriptomic analysis for a pilot study in the mechanisms of alleviating psoriatic dermatitis using TDGs in an imiquimod (IMQ)-induced psoriasis-like mouse model, turned out that TDGs could significantly alleviate erythema, scale and thickening of typical skin lesions, along with reduce KC proliferation. Combining with RNA sequencing results and experimental verification, the effects of TDGs for psoriasis were confirmed to up-regulate metabolic signaling pathways, such as Gly-Ser-Thr axis, as well as down-regulate immune and inflammatory pathways, along with reduce Rac2 and Arhgdib concentrations [13]. Nevertheless, in view of the multi-target characteristics of CM compounds, it remains far more challenging to extract biological insights of transcriptome sequencing. Hence, we used Ingenuity Pathway Analysis (IPA) and Gene Set Enrichment Analysis (GSEA) for functional enrichment analysis to explore the possible mechanisms of TDGs. Recently, IPA is often used to analyse transcriptome data to further investigate the regulatory relationship between genes, bioinformatics application of which refers functional analyze, integration, and further understanding [14]. An IPA network was established linked by their upstream proteins in the current study. GSEA can analyse and interpret changes of the coordinate path levels in transcriptomics experiments, while algorithm of GSEA calculated according to the overall trend of actual data, can make up valuable information easily overlooked by general differential analysis, that are biological characteristics in important genes, relationships between gene regulatory networks and functions with significance of genes [15]. Therefore, the purpose of this study was to determine the complex pathway of TDGs in the treatment of psoriasis via transcriptional profiling, IPA together with GSEA, and to conduct experimental verification (Fig. 1).

2. Methods

2.1. TDG materials preparation

The TDGs were made up of nine Chinese herbs (Additional file 1), authenticated as per standard protocols by a pharmacognosist of the Yueyang Hospital of Integrated Traditional Chinese and Western Medicine, Shanghai University of Traditional Chinese Medicine. Take TDG preparation 50 ul, add 950 ul precooled 40% methanol solution and dilute it 20 times, swirl it thoroughly, and mark it as di20. Then take diluent di20 50 ul, add in 450 ul precooled 40% methanol solution and dilute it 10 times, swirl it
thoroughly, and mark it as di200. All the diluted samples were centrifuged at 16000 rpm at 4°C for 15 min. The supernatant was transferred to liquid vials and stored in a refrigerator at 4°C for later use.

Liquid Chromatograph Mass Spectrometer/Mass Spectrometer (LC-MS/MS) analysis was used for quantitative control of TDGs. LC-MS/MS analysis was conducted using Waters ACQUITY UPLC I-Class coupled with a 5500 QTRAP mass spectrometer (SCIEX), with 35°C Column temperature. The mobile phase delivered at 0.3 ml/min was a mixture of 0.1% formic acid aqueous solution together with 0.1% formic acid acetonitrile solution, as the gradient elution: 0–3 min (5%-25% B), 3-8.5 min (25%-45% B), 8.5–12 min (45%-95% B), 12–15 min (95%-98% B), 15-15.2 min (98%-5% B), 15.2–18.2 min (5% B).

2.2. Animals

Specific pathogen-free (SPF) - grade male BALB/c mice, weighted 25 ± 3 g, were offered by the Shanghai Medical Experimental Animal Center (SCXK Shanghai 2013-0016, Shanghai, China). The mice were housed in a germfree environment (temperature of 23 ± 2°C). Ethical approvals were obtained by the Ethics Committee of Yueyang Hospital affiliated to Shanghai University of Traditional Chinese Medicine (No. YYLAC-2021-107).

2.3. Model establishment and interventions

In a nutshell, mice were divided into three groups: control group; psoriatic model group (IMQ group); and psoriatic model with TDGs treated group (IMQ + TDG group) (n = 4). After adaptive feeding, the hair on the back of mice was removed (2 × 2 cm²). Establishment of psoriatic modeling was treated with 62.5 mg of 5% IMQ cream for 6 h on back skin, while mice in the control group were applied isodose petroleum jelly. Mice in IMQ + TDG group were followed by intragastric administration of 1.8 g/kg TDGs, and mice in IMQ group were conducted with intragastric administration of 1.8 g/kg 0.9% NaCl solution.

2.4. Transcriptome sequencing

On the 12th day, mice were euthanized and the back skin was extracted for Transcriptome sequencing. Illumina HiSeqTM 2500 was used for sequencing and the transcriptome analysis was performed by Shanghai OE Biotechnology Co., Ltd. Standardization disposal was performed using DESeq software (version 1.18.0) was used to standardize the gene count for each sample. The differentially expressed genes (DEGs) were screened according to the results of \(|\log_{2}\text{FoldChange}| > 1 \) and \(p\)-value < 0.05.

2.5. Gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis

After obtaining DEGs, DAVID (https://david.ncifcrf.gov/) was used to analyse the significance of GO and KEGG analysis. A \(p\)-value of < 0.05 was considered as differentially statistical significance.

2.6. IPA

DEGs with \(|\log_{2}\text{FoldChange}| > 2\) as well as \(p\)-value < 0.05, pathway analysis and the construction of protein-protein interaction (PPI) network of the expression data were executed with IPA (QIAGEN,
Redwood City, CA, USA). Results with |z-score| ≥ 2 and/or overlap p-value < 0.05 were considered as differentially statistical significance. Upstream networks were described on the IPA database.

### 2.7. GSEA

GSEA (https://www.gsea-msigdb.org/gsea/index.jsp) with probes ranked by signal-to-noise ratio and statistical significance determined by 1,000 gene set permutations was used to investigate differences among groups for exploring the potential molecular mechanisms and functions of DEGs [16, 17]. A p-value of < 0.01 and false discovery rate (FDR) with a q-value of < 0.05 were considered as differentially statistical significance.

### 2.8. RT-PCR

On day 12, the mice were euthanized with CO$_2$ inhalation. The back skin was extracted and preserved in TRIzol reagent kit. The Reverse Transcription System First Strand cDNA Synthesis Kit was used with 20.0 a reaction volume. Real-time fluorescent PCR was performed for RT-PCR. The specific experimental method was the same as the previous study [18]. The primer sequences were revealed in Additional file 2.

### 2.9. Statistical methods

SPSS 24.0 (IBM Corp., Armonk International Business Machines, New York, USA) was used for analysing the data, described as mean ± standard deviation (SD). A t-test was used to compare the two groups, while a p-value of < 0.05 was considered as statistical significance.

### 3. Results

#### 3.1. Transcriptional regulations of TDG treatment for psoriasis

**3.1.1. Transcriptional profiles analysis of DEGs following TDG treatment**

Firstly, to integrally control the quality of TDGs, LC-MS/MS analysis was used for identification and quantification of compounds in TDGs. There were 0.68 ug/ml Rutin, 14.92 ug/ml Caffeic acid, 3.56 ug/ml Cryptotanshinone, 2.77 ug/ml Tanshinone IIA, 2.95 ug/ml Formononetin, 95.87 ug/ml Liquiritin, 82.11 ug/ml Danshensu possessed the highest content in TDGs. A total ion flow chromatogram of TDGs was shown in Additional file 3. The structure of the major components was displayed in Additional file 4.

Afterwards, to further determine the effect of TDG treatment for psoriasis, the transcriptional profiles of skin lesions treated or nor treated with TDGs of psoriatic mouse models were detected and compared based on RNA sequencing [13]. The DEGs following TDG treatment were identified by RNA sequencing, and the data were stored in the SRA database (SRP292449). A total of 1233 DEGs were identified, with 539 upregulation as well as 694 downregulation (Additional file 5A). Biological characteristics of the
potential targets for TDGs were estimated by KEGG and GO analysis. KEGG analysis indicated the most upregulated gene categories (Top 3) were for the neuroactive ligand-receptor interaction, estrogen signaling pathway, biosynthesis of unsaturated fatty acids, and glycine, serine, and threonine (Gly-Ser-Thr) metabolism; and the most downregulated gene categories (Top 3) were for cytokine-cytokine receptor interaction, Staphylococcus aureus infection, osteoclast differentiation, and chemokine signaling pathway (Additional file 5B). GO analysis manifested that the most upregulated gene categories (Top 3) were for the intermediate filaments, keratin filaments, cellular components, and structural molecule activity; and the most downregulated gene categories (Top 3) were inflammatory response, immune system process, extracellular space, and innate immune response (Additional file 5C). Therefore, the above results revealed the effect of TDG treatment for psoriasis was closely related to metabolism, inflammation as well as immune regulations.

3.1.2. Upstream analysis of DEGs following TDG treatment

To further reveal the therapeutic pathway of TDG in the treatment of psoriasis, we analysed the potential upstream regulatory functions of DEGs following TDG intervention. We selected DEGs with $|\text{log}_{2}\text{FoldChange}| > 2$ and $p$-value $< 0.05$ based on the sequencing results to construct the core regulatory network in the IPA database (Fig. 2). IPA upstream regulator analysis identified 30 significant upstream regulators that were 21 downregulated (Fig. 2A) and 9 upregulated (Fig. 2B) following TDG treatment ($|z$-score| $\geq 2$ and overlap $p$-value $< 0.05$). Among them, the most remarkable inhibited transcription regulators (Top 3) were Kruppel-like factor (KLF) 4 (overlap $p$-value $= 3.11E-16$), T-cell leukemia (TCL) 1A (overlap $p$-value $= 2.04E-15$) and CCAAT/enhancer binding protein epsilon (CEBPE) (overlap $p$-value $= 1.95E-11$), while the most distinct activated transcription regulators (Top 3) were PAX1 (overlap $p$-value $= 1.43E-09$), zinc finger protein (ZFP) 36 (overlap $p$-value $= 4.98E-09$) and SRY-box transcription factor (SOX) 7 (overlap $p$-value $= 4.54E-08$). The above results indicated that these upstream transcription factors played central roles in the improvement of psoriasis following TDGs.

3.2. Transcriptional regulation of TDG treatment for psoriasis via Wnt signaling pathway

3.2.1. GSEA and IPA of critical GO following TDG treatment

Take it further comprehensively analyse the changed gene sets following TDG intervention, we wielded GSEA to critical GO. The top eight regulated gene sets incorporated intermediate filament (GO: 0005882), negative regulation of Wnt signaling pathway (GO: 0030178), collagen fibril organization (GO: 0030199), cell fate commitment (GO: 0045165), Wnt-activated receptor activity (GO: 0042813), ureteric bud development (GO: 0001657), hair follicle development (GO: 0001942), and negative regulation of canonical Wnt signaling pathway (GO: 0090090), ranked by normalized enrichment score (NES) (Fig. 3A). Given the above, it was confirmed that the therapeutic effect of TDGs on psoriasis might be closely related to the Wnt signaling pathway. Wnt signaling pathway has proved to be activated in the pathogenesis of inflammatory diseases including psoriasis vulgaris, atherosclerosis, rheumatoid arthritis,
and sepsis [19]. The differentially expressions of the core enrichment mRNA of three relevant pathways of Wnt were displayed in Fig. 3B.

For further analyse the possible regulatory relationships in the three enriched Wnt related signaling pathways, we constructed relevant IPA core network. These genes associated with Wnt signaling pathway (GO: 0030178), consisted of complex, cytokines, enzymes, group/complex, growth factor, kinase, peptidase, transcription regulator, transmembrane receptor, and others, were used for constructing the core regulatory network on the IPA database. Platelet derived growth factor (PDGF) BB, Wnt, extracellular signal-regulated kinase (ERK)1/2, and phosphoinositide 3-kinase (PI3K) were shown to have the highest correlation in the regulations of other proteins in this pathway following TDG treatment (Fig. 3C). The proteins encoded by these genes associated with Wnt-activated receptor activity (GO: 0042813) including complex, G-protein coupled receptor, group, growth factor, kinase, transmembrane receptor, together with others were integrated into IPA core network. G protein-coupled receptors (Gpcr), epidermal growth factor (EGF), as well as protein kinase B (Akt) indicated higher levels of activities (Fig. 3D). Furthermore, Genes of complex, cytokine, enzyme, G-protein coupled receptor, group, growth factor, transcription regulator, transmembrane receptor and others were consolidated into IPA core network associated with Wnt signaling pathway (GO: 0090090). Axis inhibition protein (AXIN) 1, Wnt, along with histone deacetylase (Hdac) showed significant activities in regulating other proteins (Fig. 3E).

### 3.2.2. GSEA and IPA of critical KEGG Pathways following TDG treatment

On the other hand, we also proceeded GSEA for KEGG Pathways. Briefly, ranked by normalized enrichment score, The top nine representative gene sets included basal cell carcinoma (mmu05217), hippo signaling pathway (mmu04390), melanogenesis (mmu04916), ECM-receptor interaction (mmu04512), breast cancer (mmu05224), biosynthesis of unsaturated fatty acids (mmu01040), protein digestion and absorption (mmu04974), cushing's syndrome (mmu04934), and Wnt signaling pathway (mmu04310) (Fig. 4A). The differentially expressions of the core enrichment mRNA of this pathway were displayed in Fig. 4B.

Similarly, we constructed IPA core network associated with Wnt signaling pathway (mmu04310), while genes consisted of complex, enzyme, group, kinase, ligand-dependent nuclear receptor, transcription regulator and others. In this pathway, CTNNB1, mitogen activated protein kinase (MAPK) 8, together with casein kinase (Ck) 2 played key regulatory roles in the Wnt signaling pathway following TDG treatment in psoriasis (Fig. 4C).

### 3.3. Upstream analysis of TDG treatment for psoriasis via Wnt signaling pathway and experimental validation

After the above preliminary confirmation that TDG treatment for psoriasis was closely related to the controls of Wnt signaling pathway, we further predicted the upstream transcriptional regulations. Critical genes were identified as enriched with significance using IPA database (overlap p-value < 0.05), while six
transcription regulators, consisted of enhancer of zeste homolog (EZH) 2 (overlap p-value = 1.52E-15), CTNNB1 (overlap p-value = 2.83E-12), SRY-related HMG-box transcription factor (SOX) 11 (overlap p-value = 1.26E-11), FOS (overlap p-value = 1.49E-09), tumor protein p63 (TP63) (overlap p-value = 9.28E-09), and WD repeat domain (WDR) 5 (overlap p-value = 3.13E-08), were of great importance in the upstream of TDG treatment for psoriasis via Wnt signaling pathway (Fig. 5A).

For verifying whether the six transcription regulators of upstream had significant corresponding functions in the TDG treatment for psoriasis, we conducted RT-PCR in the back skin lesions of IMQ-induced psoriatic mouse models. The results confirmed that compared with normal skin, the EZH2, CTNNB1 and WDR5 mRNA levels of psoriatic back lesions were specifically elevated, while TP63 levels was dropped. Following TDG treatment, the decline in mRNA expression levels of EZH2, CTNNB1 together with WDR5, and the rise in mRNA expression levels of as TP63 was emerged (Fig. 5B).

4. Discussion

TDGs have favorable effects on relieving psoriasis. Our previous studies have also proved that TDGs are effective for psoriatic patients [8, 9], while the in vivo results [13] confirmed that TDGs affected KC proliferation and inflammatory responses to alleviate IMQ-induced psoriatic symptoms in animal models. To integrally control the quality of TDGs, LC-MS/MS analysis was for application. The results indicated that TDGs contained Rutin, Caffeic acid, Cryptotanshinone, Tanshinone IIA, Formononetin, Liquiritin, Danshensu, and other active ingredients. Tanshinone IIA, Cryptotanshinone, and Danshensu are the three main bioactive components of *Salvia miltiorrhiza Bunge*, while all of them has proved to alleviate psoriasis [12, 20–22]. Formononetin proved to has strong anti-proliferation properties[23]. Previously, Rutin, Caffeic acid, along with Liquiritin were reported to decrease inflammation, while Liquiritin had a certain protective effect on skin injury, and inhibited angiogenesis [24–27]. However, the mechanisms of TDGs in the treatment of psoriasis remains vague, while the specific action pathways of TDGs are worth further exploration. Therefore, the current study aims to use GSEA and IPA core network to investigate the transcriptional regulation mechanism, and systematically obtain molecular functions in TDGs.

Based on transcriptome sequencing, we constructed the core regulatory network in the IPA database for DEGs following TDG application. In order to further explore the possible relationships between regulatory factors and downstream molecules, upstream analysis in IPA predicted the active or inhibited upstream regulators following TDGs. Results revealed the most remarkable inhibited transcription regulators were KLF4, TCL1A and CEBPE. KLF4, a transcription factor, regulates a diverse array of cellular processes, including development, differentiation, proliferation, and apoptosis. Compared with non-psoriatic skin, KLF4 protein levels were significantly increased in psoriatic lesions in patients [28]. Excessive KLF4 can increase the level of histone H3 acetylation in Keratin (KRT) 17 promoter region by synergistic EP300, and mediate the over-expression of KRT17 in psoriatic lesions [29]. TCL1A, a stem cell marker, is abundantly expressed in embryonic stem cells, identified as an oncogene in various hematological malignancies, beyond that also specifically expressed in proliferative solid tumors. TCL1A expression proved to be higher in colorectal cancer tissues than that in adjacent normal tissues, and significantly correlated with
tumor differentiation, TNM stage and Ki67 positive rate [30]. CEBPE, one of the CCAAT/enhancer binding proteins, plays critical role in multiple physiological and pathological processes, and is highly expressed in tumor diseases [31]. The transcriptional activities of CEBPE are regulated by interactions with other transcription factors and/or post-translational modification (such as acetylation) [32]. For another, the most distinct activated transcription regulators following TDGs were PAX1, ZFP36 and SOX7. PAX1, a pivotal tumor suppressor gene, is responsible for regulating cell differentiation and maturation. PAX1 gene methylation detection can accurately monitor cervical cancer precancerous lesions, thus has high research value [33]. ZFP36 is a type of transcription factor with finger-like domains, associated with autoimmune diseases, arthritis and other syndromes in mice and humans, and has regulatory functions on gene expression, cell differentiation, and embryo development. Mice with conditional deletion of Tristetraprolin (TTP) (encoded by the ZFP36 gene) in KCs (Zfp36fl/flK14-Cre mice) pullulated exacerbated inflammation in the IMQ-induced psoriatic models. Furthermore, the Zfp36fl/flK14-Cre mice developed progressively spontaneous pathology, including systemic inflammation, psoriatic-like lesions, and dactylitis [34]. TTP was downregulated in fibroblasts deriving from psoriatic patients, when compared to those deriving from healthy individuals, while psoriatic fibroblasts exhibited abnormal inflammasome activities. The above phenomena proved to be related to ZFP36 promoter methylation [35]. SOX7 is a member of the SOX family of transcription factors. Studies have indicated that SOX7 is a tumor suppressor gene, down-regulated in most tumors. The regulatory mechanisms may be through regulating the transcription process mediated by Wnt-β-catenin signaling pathway, inhibiting tumor proliferation, migration, and invasion [36–38]. Given all this, these upstream transcription factors played central roles in the improvement of psoriasis following TDGs.

In order to further explore and analyse the changes of the gene sets following TDG intervention, we wielded GSEA to critical GO and KEGG. GSEA results confirmed that the treatment of TDGs was closely relative with the Wnt signaling pathway. Previous studies have confirmed that Wnt signaling pathway, as a crucial pathway of proliferative signal transduction, plays a negative regulatory role in psoriasis. β-catenin, an important transcription factor of Wnt signaling pathway, is translocated in the skin granular layer of psoriasis [39]. Inhibition of the β-catenin encoding gene CTNNB1 in HaCaT cells leads to the decrease of Cyclin D1 and Axin2 expressions, thereby inhibiting IL-22-induced cell proliferation [40]. After stimulating HaCaT cells, IWP-2 (Wnt inhibitor) can inhibit cell proliferation and secretion of pro-inflammatory factors, as well as promote cell differentiation [41]. Next, the core network analysis was performed using IPA, allowed us to analyse the coordinate expression changes at pathway levels [42]. The results indicated that PDGFBB, Wnt, ERK1/2, PI3K, Gpcr, EGF, Akt, AXIN1, Hdac, CTNNB1, MAPK8, as well as Ck2 in Wnt signaling pathway were considered as critical targets of TDGs.

For exploring the upstream regulation of Wnt signaling pathway following the intervention of TDGs, we applied the upstream analysis of IPA to forecast upstream transcription factors, and carried out experimental verification. Combined with results of verification via animal experiments, critical transcription factors with prominent enrichment included EZH2, CTNNB1 and WDR5, were remarkably down-regulated by TDGs. The methyltransferase EZH2 as a valid target for psoriasis therapy, consistent with our results, overexpresses in skin lesions of psoriatic mouse models and HaCaT cells. In vivo,
GSK126, the inhibitor of EZH2, GSK126 can ameliorate the IMQ-induced psoriatic lesions. EZH2 contributes to the development of psoriasis by inhibiting the transforming growth factor-β (TGF-β)/recombinant mothers against decapentaplegic homolog (SMAD) pathway impairment of miR-125a-5p-mediated Sex comb on midleg with four malignant brain tumor domains (SFMBT) 1 inhibition [43–45]. CTNNB1 gene is located on chromosome 3p21, and the mutation of the gene exon 3 causes nuclear accumulation of β-catenin. Targeting CTNNB1 and subsequently affecting the downstream factors, CyclinD1 and Axin2, can inhibit IL-22-induced proliferation of HaCaT and HKC cells, provided diagnostic markers and novel targets for psoriatic treatment [40]. The WD40 protein family member WDR5 in the Wnt signaling pathway has several functions on tumorigenesis and development of multiple organ tumors. It has been demonstrated that overexpression of WDR5 is associated with poor prognosis in patients with esophageal squamous cell carcinoma (ESCC), while WDR5 may act as a potential novel prognostic biomarker for ESCC [46]. However, the impact on psoriasis of WDR5 has not been elucidated. At the same time, the expression of TP63 mRNA was significantly increased following TDGs by verifying. TP63 can act as transcription factors, activating or repressing expression from a variety of gene promoters, and is believed to be crucial for normal development of ectodermal derived structures such as skin and oral mucosa. It has been indicated the downregulation of TP63 mRNA in psoriatic lesions compared to both clinically normal skin from patients and matched healthy controls [47]. Gao et al. reported that the expression of TP63 in psoriatic lesions was increased after treatment, which was consistent with our verification results [48]. Notably, SOX11 and FOS of the significant upstream transcription factors enriched by IPA analysis revealed no obvious difference in mRNA levels in psoriatic lesions, normal skin, and lesions with TDGs treatment. Although differences were not statistically significant, but upstream subtle changes might lead to mutative expressions of downstream Wnt signaling pathways. In summary, we predicted and verified that TDGs alleviated psoriasis via regulating integrant upstream transcription factors in the Wnt signaling pathway.

Our data provide evidence that TDGs may improve psoriasis by regulating the Wnt signaling pathway. In the future, further experiments on this pathway can be conducted in vivo and in vitro to further explore the mechanism of TDGs.

5. Conclusion

The present study used modular pharmacology analysis to prove that TDGs regulated the Wnt signaling pathway to ameliorate skin lesions of IMQ-induced psoriatic mouse models, and identified the upstream regulators in the Wnt signaling pathway following TDG treatment.

Abbreviations

Akt: protein kinase B; AXIN: Axis inhibition protein; CEBPE: enhancer binding protein epsilon; Ck: casein kinase; CM: Chinese medicine; DEG: differentially expression genes; EGF: epidermal growth factor; ERK: extracellular signal-regulated kinase; ESCC: esophageal squamous cell carcinoma; EZH: enhancer of zeste homolog; Gly-Ser-Thr: glycine, serine, and threonine; GO: Gene ontology; Gpcr: G protein-coupled
Declarations

Ethics approval and consent to participate

The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. Experiments were performed under a project license (NO. YYLAC-2021-107) granted by the Ethics Committee of Yueyang Hospital affiliated to Shanghai University of Traditional Chinese Medicine, in compliance with the Ethics Committee of Yueyang Hospital affiliated to Shanghai University of Traditional Chinese Medicine guidelines for the care and use of animals.

Consent for publication

Not applicable.

Availability of data and materials

All data generated or analysed during this study are included in this published article [and its supplementary information files].

Competing interests

The authors declare that they have no competing interests.

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Authors' contributions

YL1 (Ying Luo), YZ, and SYY contributed equally to this study. BL and LK conceived and designed the work. YL2 (Yue Luo), XJD, YR, MX, and XYF performed experimental work, collected, and analyzed the statistical data. JKS, HPZ, and TYL interpreted results. YL1, YZ, SYY, BL, and LK drafted and critically evaluated the manuscript. All the authors read and approved the final manuscript.

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References

1. Parisi R, Iskandar IYK, Kontopantelis E, Augustin M, Griffiths CEM, Ashcroft DM, et al. National, regional, and worldwide epidemiology of psoriasis: systematic analysis and modelling study. *BMJ*. 2020;369:m1590. doi:10.1136/bmj.m1590.

2. Michalek IM, Loring B, John SM. A systematic review of worldwide epidemiology of psoriasis. *J Eur Acad Dermatol Venereol*. 2017;31(2):205–12. doi:10.1111/jdv.13854.

3. Yan D, Blauvelt A, Dey AK, Golpanian RS, Hwang ST, Mehta NN, et al. New Frontiers in Psoriatic Disease Research, Part II: Comorbidities and Targeted Therapies. *J Invest Dermatol*. 2021. doi:10.1016/j.jid.2021.02.743.

4. Lopes N, Dias LLS, Azulay-Abulafia L, Oyafuso LKM, Suarez MV, Fabricio L, et al. Humanistic and Economic Impact of Moderate to Severe Plaque Psoriasis in Brazil. *Adv Ther*. 2019;36(10):2849–65. doi:10.1007/s12325-019-01049-7.

5. Al Sawah S, Foster SA, Goldblum OM, Malatestinic WN, Zhu B, Shi N, et al. Healthcare costs in psoriasis and psoriasis sub-groups over time following psoriasis diagnosis. *J Med Econ*. 2017;20(9):982–90. doi:10.1080/13696998.2017.1345749.

6. Griffiths CEM, Armstrong AW, Gudjonsson JE, Barker J. Psoriasis. *Lancet*. 2021;397(10281):1301–15. doi:10.1016/S0140-6736(20)32549-6.

7. Chiu HY, Hui RC, Tsai TF, Chen YC, Chang Liao NF, Chen PH, et al. Predictors of time to relapse following ustekinumab withdrawal in patients with psoriasis who had responded to therapy: An
eight-year multicenter study. *J Am Acad Dermatol*. 2019. doi:10.1016/j.jaad.2019.01.035.

8. Fan B, Li B, Shen JX, Zhang Y, Tang H, Wang J. Effects of herbal medicine on cytokines in patients with psoriasis at different stages. *Chin J Dermatol Venereol*. 2006(02):70–1.

9. Fan B, Li B, Zhang Y, Shen JX, Zhao XL. Effects of Chinese herbal medicine on the secretion of Substance P and -endorphins in plasma of patients with psoriasis during progressive and inactive stages. *Shanghai J Traditional Chin Med*. 2006(10):34–5. doi:10.16305/j.1007-1334.2006.10.017.

10. Ru Y, Yan XN, Yang SQ, Gong LP, Li LE, Chen J, et al. Oral Taodan granules for mild-to-moderate psoriasis vulgaris: protocol for a randomized, double-blind, multicenter clinical trial. *Ann Transl Med*. 2019;7(18):488. doi:10.21037/atm.2019.09.05.

11. Fan B, Li X, Ze K, Xu R, Shi RF, Geng L, et al. Expression of T-helper 17 cells and signal transducers in patients with psoriasis vulgaris of blood-heat syndrome and blood-stasis syndrome. *Chin J Integr Med*. 2015;21(1):10–6. doi:10.1007/s11655-014-1769-7.

12. Li FL, Xu R, Zeng QC, Li X, Chen J, Wang YF, et al. Tanshinone IIA Inhibits Growth of Keratinocytes through Cell Cycle Arrest and Apoptosis: Underlying Treatment Mechanism of Psoriasis. *Evid Based Complement Alternat Med*. 2012;2012:927658. doi:10.1155/2012/927658.

13. Kuai L, Luo Y, Qu K, Ru Y, Luo Y, Ding X, et al. Transcriptomic Analysis of the Mechanisms for Alleviating Psoriatic Dermatitis Using Taodan Granules in an Imiquimod-Induced Psoriasis-like Mouse Model. *Front Pharmacol*. 2021;12:632414. doi:10.3389/fphar.2021.632414.

14. Kramer A, Green J, Pollard J, Jr., Tugendreich S. Causal analysis approaches in Ingenuity Pathway Analysis. *Bioinformatics*. 2014;30(4):523–30. doi:10.1093/bioinformatics/btt703.

15. Powers RK, Goodspeed A, Pielke-Lombardo H, Tan AC, Costello JC. GSEA-InContext: identifying novel and common patterns in expression experiments. *Bioinformatics*. 2018;34(13):i555-i64. doi:10.1093/bioinformatics/bty271.

16. Mootha VK, Lindgren CM, Eriksson KF, Subramanian A, Sihag S, Lehár J, et al. PGC-1alpha-responsive genes involved in oxidative phosphorylation are coordinately downregulated in human diabetes. *Nat Genet*. 2003;34(3):267–73. doi:10.1038/ng1180.

17. Subramanian A, Tamayo P, Mootha VK, Mukherjee S, Ebert BL, Gillette MA, et al. Gene set enrichment analysis: a knowledge-based approach for interpreting genome-wide expression profiles. *Proc Natl Acad Sci U S A*. 2005;102(43):15545–50. doi:10.1073/pnas.0506580102.

18. Kuai L, Zhang JT, Deng Y, Xu S, Xu XZ, Wu MF, et al. Sheng-ji Hua-yu formula promotes diabetic wound healing of re-epithelization via Activin/Follistatin regulation. *BMC Complement Altern Med*. 2018;18(1):32. doi:10.1186/s12906-017-2074-8.

19. Pashirzad M, Shafiee M, Rahmani F, Behnam-Rassouli R, Hoseinkhani F, Ryzhikov M, et al. Role of Wnt5a in the Pathogenesis of Inflammatory Diseases. *J Cell Physiol*. 2017;232(7):1611–6. doi:10.1002/jcp.25687.

20. Tang L, He S, Wang X, Liu H, Zhu Y, Feng B, et al. Cryptotanshinone reduces psoriatic epidermal hyperplasia via inhibiting the activation of STAT3. *Exp Dermatol*. 2018;27(3):268–75. doi:10.1111/exd.13511.
21. Jia J, Mo X, Liu J, Yan F, Wang N, Lin Y, et al. Mechanism of danshensu-induced inhibition of abnormal epidermal proliferation in psoriasis. *Eur J Pharmacol*. 2020;868:172881. doi:10.1016/j.ejphar.2019.172881.

22. Huang YK, Tseng KF, Tsai PH, Wang JS, Lee CY, Shen MY. IL-8 as a Potential Therapeutic Target for Periodontitis and Its Inhibition by Caffeic Acid Phenethyl Ester In Vitro. *Int J Mol Sci*. 2021;22(7). doi:10.3390/ijms22073641.

23. Ong SKL, Shanmugam MK, Fan L, Fraser SE, Arfuso F, Ahn KS, et al. Focus on Formononetin: Anticancer Potential and Molecular Targets. *Cancers (Basel)*. 2019;11(5). doi:10.3390/cancers11050611.

24. Chen X, Zong C, Gao Y, Cai R, Fang L, Lu J, et al. Curcumol exhibits anti-inflammatory properties by interfering with the JNK-mediated AP-1 pathway in lipopolysaccharide-activated RAW264.7 cells. *Eur J Pharmacol*. 2014;723:339–45. doi:10.1016/j.ejphar.2013.11.007.

25. Tian C, Liu X, Chang Y, Wang R, Yang M, Liu M. Rutin prevents inflammation induced by lipopolysaccharide in RAW 264.7 cells via conquering the TLR4-MyD88-TRAF6-NF-kappaB signalling pathway. *J Pharm Pharmacol*. 2021;73(1):110–7. doi:10.1093/jpp/rgaa015.

26. Li Y, Xia C, Yao G, Zhang X, Zhao J, Gao X, et al. Protective effects of liquiritin on UVB-induced skin damage in SD rats. *Int Immunopharmacol*. 2021;97:107614. doi:10.1016/j.intimp.2021.107614.

27. Zhai KF, Duan H, Cui CY, Cao YY, Si JL, Yang HJ, et al. Liquiritin from Glycyrrhiza uralensis Attenuating Rheumatoid Arthritis via Reducing Inflammation, Suppressing Angiogenesis, and Inhibiting MAPK Signaling Pathway. *J Agric Food Chem*. 2019;67(10):2856–64. doi:10.1021/acs.jafc.9b00185.

28. Kim KJ, Park S, Park YH, Ku SH, Cho EB, Park EJ, et al. The expression and role of kruppel-like factor 4 in psoriasis. *Ann Dermatol*. 2014;26(6):675–80. doi:10.5021/ad.2014.26.6.675.

29. Zhang S, Tan Y, Yin XQ, Luo H. [Molecular Mechanism of KLF4 Up-regulating Keratin 17 Expression in Keratinocytes]. *Sichuan Da Xue Xue Bao Yi Xue Ban*. 2020;51(6):797–802. doi:10.12182/20201160503.

30. Li H, Yan X, Liu L, Huang L, Yin M, Pan C, et al. T-cell leukemia/lymphoma-1A predicts the clinical outcome for patients with stage II/III colorectal cancer. *Biomed Pharmacother*. 2017;88:924–30. doi:10.1016/j.biopha.2017.01.128.

31. Huang Y, Lin L, Shen Z, Li Y, Cao H, Peng L, et al. CEBPG promotes esophageal squamous cell carcinoma progression by enhancing PI3K-AKT signaling. *Am J Cancer Res*. 2020;10(10):3328–44.

32. Muraoka M, Akagi T, Ueda A, Wada T, Koeffler HP, Yokota T, et al. C/EBP epsilon DeltaRS derived from a neutrophil-specific granule deficiency patient interacts with HDAC1 and its dysfunction is restored by trichostatin A. *Biochem Biophys Res Commun*. 2019;516(1):293–9. doi:10.1016/j.bbrc.2019.06.130.

33. Chang CL, Ho SC, Su YF, Juan YC, Huang CY, Chao AS, et al. DNA methylation marker for the triage of hrHPV positive women in cervical cancer screening: Real-world evidence in Taiwan. *Gynecol Oncol*. 2021;161(2):429–35. doi:10.1016/j.ygyno.2021.02.011.
34. Andrianne M, Assabban A, La C, Mogilenko D, Salle DS, Fleury S, et al. Tristetraprolin expression by keratinocytes controls local and systemic inflammation. *JCI Insight*. 2017;2(11). doi:10.1172/jci.insight.92979.

35. Bertesi M, Fantini S, Alleci C, Lotti R, Martello A, Parenti S, et al. Promoter Methylation Leads to Decreased ZFP36 Expression and Deregulated NLRP3 Inflammasome Activation in Psoriatic Fibroblasts. *Front Med (Lausanne)*. 2020;7:579383. doi:10.3389/fmed.2020.579383.

36. Wang C, Qin L, Min Z, Zhao Y, Zhu L, Zhu J, et al. SOX7 interferes with beta-catenin activity to promote neuronal apoptosis. *Eur J Neurosci*. 2015;41(11):1430–7. doi:10.1111/ejn.12910.

37. Ying K, Wang L, Long G, Lian C, Chen Z, Lin W. ACTA2-AS1 suppresses lung adenocarcinoma progression via sequestering miR-378a-3p and miR-4428 to elevate SOX7 expression. *Cell Biol Int*. 2020;44(12):2438–49. doi:10.1002/cbin.11451.

38. Hong KO, Oh KY, Yoon HJ, Swarup N, Jung M, Shin JA, et al. SOX7 blocks vasculogenic mimicry in oral squamous cell carcinoma. *J Oral Pathol Med*. 2021. doi:10.1111/jop.13176.

39. Hampton PJ, Ross OK, Reynolds NJ. Increased nuclear beta-catenin in suprabasal involved psoriatic epidermis. *Br J Dermatol*. 2007;157(6):1168–77. doi:10.1111/j.1365-2133.2007.08195.x.

40. Shen H, Zeng B, Wang C, Tang X, Wang H, Liu W, et al. MiR-330 inhibits IL-22-induced keratinocyte proliferation through targeting CTNNB1. *Biomed Pharmacother*. 2017;91:803–11. doi:10.1016/j.biopha.2017.05.005.

41. Wang W, Yu X, Wu C, Jin H. IL-36gamma inhibits differentiation and induces inflammation of keratinocyte via Wnt signaling pathway in psoriasis. *Int J Med Sci*. 2017;14(10):1002–7. doi:10.7150/ijms.20809.

42. Yang X, Xu X, Cai X, He J, Lu P, Guo Q, et al. Gene set enrichment analysis and protein-protein interaction network analysis after sciatic nerve injury. *Ann Transl Med*. 2020;8(16):988. doi:10.21037/atm-20-4958.

43. Qu S, Liu Z, Wang B. EZH2 is involved in psoriasis progression by impairing miR-125a-5p inhibition of SFMBT1 and leading to inhibition of the TGFbeta/SMAD pathway. *Ther Adv Chronic Dis*. 2021;12:2040622320987348. doi:10.1177/2040622320987348.

44. Zhang T, Yang L, Ke Y, Lei J, Shen S, Shao S, et al. EZH2-dependent epigenetic modulation of histone H3 lysine-27 contributes to psoriasis by promoting keratinocyte proliferation. *Cell Death Dis*. 2020;11(10):826. doi:10.1038/s41419-020-03028-1.

45. Muller A, Dickmanns A, Resch C, Schakel K, Hairfinger S, Dobbelstein M, et al. The CDK4/6-EZH2 pathway is a potential therapeutic target for psoriasis. *J Clin Invest*. 2020;130(11):5765–81. doi:10.1172/JCI134217.

46. Huang D, Chen X, Chen X, Qu Y, Wang Y, Yang Y, et al. WDR5 Promotes Proliferation and Correlates with Poor Prognosis in Oesophageal Squamous Cell Carcinoma. *Onco Targets Ther*. 2020;13:10525–34. doi:10.2147/OTT.S234773.

47. Gu X, Lundqvist EN, Coates PJ, Thurfjell N, Wettersand E, Nylander K. Dysregulation of TAp63 mRNA and protein levels in psoriasis. *J Invest Dermatol*. 2006;126(1):137–41. doi:10.1038/sj.jid.5700010.
48. Gao L, Dou J, Zhang B, Zeng J, Cheng Q, Lei L, et al. Ozone therapy promotes the differentiation of basal keratinocytes via increasing Tp63-mediated transcription of KRT10 to improve psoriasis. *J Cell Mol Med.* 2020;24(8):4819–29. doi:10.1111/jcmm.15160.

**Figures**

**Figure 1**

Design of study.
Figure 2

IPA upstream analysis of DEGs following TDG treatment. (A) 21 significantly downregulated upstream regulators following TDG treatment. (B) 9 significantly upregulated upstream regulators following TDG treatment.
Figure 3

GSEA analysis and IPA analysis of GO items. (A) GSEA of TDGs treated compared with disease group. (B) Heatmap of corresponding genes in three Wnt signaling pathways. (C-E) IPA network analysis of corresponding genes in three Wnt signaling pathways.

Figure 4

GSEA analysis and IPA analysis of KEGG items. (A) GSEA of TDGs treated compared with disease group. (B) Heatmap of corresponding genes in the Wnt signaling pathway. (C) IPA network analysis of corresponding genes in the Wnt signaling pathway.
Figure 5

Experiment validation of IPA upstream analysis in Wnt signaling pathway following TDG treatment. (A) IPA upstream analysis in Wnt signaling pathway following TDG treatment. (C) Experiment validation on IMQ-induced psoriatic mouse models of significant upstream Regulators in Wnt signaling pathway following TDG treatment. p-value as digits was shown. (n = 4)

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- Additionalfile1PrescriptionofTDGs.docx
- Additionalfile2PrimersequencesforRTPCR.docx
- Additionalfile3.tif
- Additionalfile4StructureofTDGs.doc
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