Chapter from the book *Kidney Transplantation - New Perspectives*
Downloaded from: http://www.intechopen.com/books/kidney-transplantation-new-perspectives
1. Introduction

An immunosuppression [IS]-free state is the ultimate goal to achieve in solid organ transplantation [SOT] because IS-related toxicity tremendously impact on overall clinical outcome [Orlando, Hematti et al., 2010]. The IS-free state substantiates into what is referred to as clinical operational tolerance [COT], defined as the condition in which a SOT recipient retains stable graft function and lacks histological signs of rejection, after having been completely off all IS for at least 1 year. The patient in question is an immunocompetent host capable of responding to other immune challenges, including infections or transplantation of organs from third party donors [Orlando, Hematti et al., 2010; Orlando et al., 2008]. However, experimental and clinical information collected so far show that COT is extremely difficult to achieve and is organ-dependent. In fact, the majority of cases of COT reported so far relate to liver graft recipients, in reason of the well known immune-privileged status of the liver determined by factors that remain obscure.

In this chapter, we will provide evidence that the achievement of COT after RT remains exceptional and that tolerogenic strategies are not yet available for daily clinical practice. To do so, we will illustrate and discuss the most relevant cases of COT described so far in the English literature. However, by briefly touching the recent outstanding progress achieved in regenerative medicine, we will bring the attention of the reader on the fact that regenerative medicine may offer a valuable approach to generate immediate, stable and durable COT in SOT recipients.

2. Classification

Several classifications have been proposed to categorize all strategies proposed so far to induce COT in RT recipients [Fehr & Sykes, 2004; Orlando et al, 2008; Orlando, Hematti et
al, 2010]. Nevertheless, given the interest and enthusiasm generated by recent investigations in the field of stem cell biology, we believe that so-called tolerogenic strategies – namely, protocols implemented to induce COT – may be divided into two new groups, cell-based and non-cell-based, according to whether or not stem cell strains showing some immunomodulatory effect in vitro are administered in the preoperative period in addition to standard IS.

2.1 Cell-based approach

2.1.1 Metachronous bone-marrow transplant for hematologic disorders
Bone marrow transplantation [BMT], results in the total replacement of the recipient’s bone marrow by the donor’s bone marrow hematopoietic cells, a condition referred to as full chimerism [Delis et al, 2004]. If the patient receives an organ from the same bone marrow donor late after BMT, then the reconstituted white cell compartment may recognize the new organ as self; theoretically, no response will be mounted by the host immune system and the new graft will be tolerated without need for any IS. However, despite several cases of BMT plus solid organ transplants [RT included] have been reported [Butcher et al, 1999; Chiang & Lazarus, 2003; Helg et al, 1994; Jacobsen et al, 1994; Light et al, 2002; Sayegh et al, 1991; Sellers et al, 2001; Sorof et al, 1995], yet COT has been an exceptional finding. Importantly, according to what reported by all series, COT developed in patients in which the transplant was performed several years after BMT, but never where BMT and RT were performed simultaneously.

2.1.2 Perioperative infusion of HSC
Somehow, the above reported data conflicted with what described in mice by Monaco and Wood, namely that the addition of donor bone marrow to a strong lymphocyte depleting regimen may result in the long-lasting survival of skin allografts from the same donor, without any maintenance IS [Monaco et al, 1970; Monaco et al, 1976; Monaco et al, 1985]. These experimental findings represented the platform for translational investigations aimed to prove that the perioperative infusion of donor-derived bone marrow cells can prolong allograft survival [Sykes, 2009]. The group at Massachusetts General Hospital adopted such strategy in 7 adult patients with renal failure due to multiple myeloma, who received combined human leukocyte antigen [HLA]-matched kidney and bone marrow transplant after a non-myeloablative conditioning regimen consisting of cyclophosphamide, antithymocyte globulin, and pretransplant thymic irradiation [Buhler et al, 2002; Fudaba et al, 2006; Spitzer et al, 1999; Spitzer et al, 2011]. Maintenance IS consisted of cyclosporine which was discontinued as early as day 73 post-transplant. Patients were followed for a mean of 7.4 (range 4-12) years. Two of them died for relapse of the baseline disease and therapy-related acute myeloid leukemia after 7.7 and 4.2 years, respectively; noteworthy, both were IS-free at the time of death. Of the 5 patients who are still alive [4 of whom with no evidence of myeloma recurrence], 3 are IS-free, and show serum creatinine levels of 0.63, 1.14 and 1.7 mg/dL, respectively. The remaining 2 individuals are receiving either prednisone or tacrolimus for graft-versus-host disease [but no rejection!], and present serum creatinine levels of 0.9 and 1.42 mg/dL.

More recently, the same group reported the short-term results of a seminal trial in which non-cancer patients received renal grafts and bone marrow from HLA haploidentical donors following nonmyeloablative conditioning to induce renal allograft tolerance [Kawai et al,
2008; Porcheray et al, 2009]. Even in this case, cyclosporine was the only immunosuppressant used for maintenance therapy. IS could be successfully weaned in 4 out of 5 patients, whereas the remaining patient experienced acute rejection and lost his graft. Cyclosporine could be withdrawn after 294 (range 240-272) days. However, matters of concern are represented by two major findings. First, the reported most recent mean creatinine clearance (67 ml/min, range 60-75) and serum creatinine levels (1.5 mg/dl, range 1.2-1.8) levels in the 4 IS-free patients were slightly abnormal. Second, 2 of these patients have shown evidence of de novo antibodies reactive to donor antigens between 1 and 2 years after the transplant [Porcheray et al, 2009]. Overall, this series demonstrates that B-cell immunity can occur in the context of T-cell tolerance to donor antigens through unknown mechanisms, otherwise said T-cell unresponsiveness is not synonymous of tolerance of both arms of the adaptive immune system. This is consistent with information reported by Nantes. Soulillou et al. extensively studied IS-free RT recipients [Ashton-Chess et al., 2007; Baeten et al., 2006; Ballet et al., 2006; Brouard et al., 2005; Louis et al., 2006; Roussey-Kesler et al., 2006] and found that COT may occur in the presence of anti-donor class II antibodies, as well as in patients who have previously experienced acute rejection. Moreover, they showed that IS-free RT recipients are more likely to have received grafts from donors younger than 30 years of age, are low-responders to blood transfusion, may develop graft dysfunction at any time after the weaning of IS, and show an incidence of infectious diseases comparable to that in normal individuals.

The Stanford group has implemented a similar strategy in two distinct small series, where patients received a renal graft followed by the infusion of HSC from the same HLA-mismatched (x4) or -matched (x6) donors, respectively [Millan et al., 2002; Scandling et al., 2008]. IS consisted of induction with total lymphoid irradiation and rabbit anti-thymocyte globulin, followed by cyclosporine and prednisone as maintenance therapy. After steroid discontinuation on day 10, mycophenolate mofetil was introduced and administered for 1 month. Intravenous injection of HSC was repeated during the third postoperative week. Results were poor. In the first trial, only one patient could be weaned off IS, while however showing slightly elevated serum creatinine levels (1.4 mg/dL). In the second trial, one individual could be weaned off IS but renal function tests are not shown. Two further patients rejected, while the remaining 3 were still under weaning at the time of publication.

Overall, based on the reported information, the above series show the lack of safety of hematopoietic stem cell-based regimens. Despite this approach may allow minimization of overall IS, however further studies are required to improve safety and effectiveness. Importantly, most of the IS-free patients reported above do not comply with the stringent criteria for COT adopted in the present chapter.

2.1.3 Perioperative infusion of alternative types of immunomodulatory cells

Transplant-acceptance-inducing cells (TAIC) are immunoregulatory macrophages which have shown some capacity of establishing a state of alloantigen-specific partial tolerance of solid organ transplants in renal transplant recipients [Hutchinson, Brem-Exner et al., 2008; Hutchinson, Riquelme et al., 2008; Riquelme P et al., 2009]. TAIC’s presumed tolerogenic properties have been tested in two safety trials differing in the method for TAIC preparation, numbers of cells infused, induction IS, and timing of infusions. None of the 17 patients enrolled in either study could be weaned off IS, most of them rejected, 2 dropped out the study for non-immunological causes, while only 2 individuals did well yet under
tacrolimus monotherapy at the end of the study. Overall, although these trials demonstrated that the infusion of TAIC is feasible, major concerns remain regarding the efficacy and safety of such an approach. It is unclear whether or not this approach confers any benefit in the establishment of minimal IS in RT patients when compared to the protocols currently adopted, as clearly stated by authors in the discussion. Lastly, the optimal dose and timing of cell infusions, as well as the most appropriate concomitant IS regimen, is yet to be determined.

For their ability for tissue repair and immunomodulation, mesenchymal stem cells (MSCs) are currently being evaluated for a wide variety of clinical applications including the treatment of disorders characterized by a dysfunction of immune regulation, such as graft versus host disease after bone marrow transplantation and rejection after cell or organ transplantation [Crop et al., 2009; Ding et al., 2009; Hematti, 2008; Le Blanc et al., 2008; Le Blanc & Ringden, 2007]. As MSCs seem to be able to promote engraftment of allogeneic cells, tissues and organs, as well as to prevent or treat rejection in pre-clinical models, they provide an exciting opportunity for further research in the field of IS minimization and withdrawal in SOT [Hematti, 2008; Hoogduijn MJ et al., 2010].

It is worth to mention that, in addition to stem cells, the therapeutic properties of various other immune cells like regulatory T cells or dendritic cells are currently being explored for SOT purposes [Issa, Hester et al., 2010; Issa, Schiopu et al., 2010; Orlando, Hematti et al., 2010].

2.2 Non cell-based protocols

2.2.1 Non-adherence to IS

The majority of cases of COT described so far after RT is represented by patients who spontaneously decided to stop IS for non-compliance to treatment [Ashton-Chess et al., 2007; Baeten et al., 2006; Ballet et al., 2006; Brouard et al., 2005; Hussey, 1975; Louis et al., 2006; Najarian, 1975; Newell et al., 2010; Owens et al., 1975; Roussey-Kesler et al., 2006; Sagoo et al., 2010; Uehling et al., 1976; Zoller et al., 1980]. These patients have been the object of numerous investigations conducted at different times by diverse authors with methods presenting dissimilar accuracy. However, we believe that it is worth to report the investigations by Burlingham et al. In an attempt to identify patients who may be weaned off IS at no risk, he resorted to the human-to-mouse trans-vivo delayed-type hypersensitivity assay [Geissler et al., 2001; VanBuskirk et al., 2000]. By observing that 2 renal allograft recipients failed to exhibit donor-reactive delayed type hypersensitivity responses, although they frequently develop donor-reactive alloantibodies, the authors demonstrated that this pattern of immune responses is not due to an absence of allosensitization, but to the development of an immune mechanism that actively inhibits anti-donor delayed-type (i.e., cell-mediated) immune responses. In other words, authors provided evidence that COT develops following the onset of regulatory, rather than suppressing mechanisms.

Of interest is the case of spontaneous interruption of IS by a 24-year old woman who had received a deceased donor RT at the age of 13 for membranoproliferative glomerulonephritis. When the woman found out to be pregnant, she stopped all IS to protect the fetus from any possible IS-induced teratogenic effect [Fischer et al., 1996]. Interestingly, she did not resume any IS after delivery, and no immunological response was observed for 9 years after the withdrawal of IS, while the patient was showing perfectly normal renal function tests.
2.2.2 Molecule-based tolerogenic protocols

A molecule-based tolerogenic regimen was implemented by Starzl a decade ago [Starzl et al., 2003]. The working hypothesis was that the need for continual high dose IS could be avoided in most cases with the use of a strong lymphocyte-depleting regimen prior to engraftment, followed by the administration of low dose tacrolimus monotherapy. The rationale was to remove the non-specific clones of immune cells responsible for rejection before contact with foreign donor antigens occurs. Once the donor antigens are in place after implantation of the new organ, repletion of immune cells occurs, favored by the homeostatic expansion triggered by leukocyte depletion. In addition, minimization of maintenance IS was implemented to further reduce the anti-donor response with just enough treatment to prevent irreversible immune damage to the graft, but not with such heavy treatment that the donor-specific clonal exhaustion-deletion process is precluded.

Broadly reacting rabbit antithymocyte globulins were administered preoperatively, followed by tacrolimus monotherapy, to 82 adult kidney, liver, pancreas, and intestinal transplant recipients. After a mean follow-up time of 18-months, 1-year patient and graft survival rates were 95% and 82% overall, respectively, IS-related morbidity was virtually eliminated, and 48/72 surviving patients were receiving spaced doses of tacrolimus monotherapy. Noteworthy, 25/39 (64%) renal, 12/17 (70%) liver, 5/12 (42%) pancreas, as well as 6/11 (54%) intestinal transplant recipients, resulted on tacrolimus spaced doses at the time of publication. However, IS could not be weaned off in any patient. Other protocols based on a similar strategy – i.e., leukocyte depletion followed by the administration of low dose single-drug IS - have been implemented after RT [Agarwal et al., 2008; Calne et al., 2000; Calne et al., 1999; Calne et al., 1998; Ciancio & Burke, 2008; Clatworthy et al., 2009; Kirk et al., 2003; Kirk et al., 2005; Orlando, Hematti et al., 2010; Trzonkowski et al., 2006; Trzonkowski et al., 2008; Watson et al., 2005] and liver, the organ most capable of developing COT, transplantation [Eason et al., 2005]. However, none of these protocols has proved effective or safe in producing COT nor has they shown any convincing impact on overall outcomes.

2.2.3 Total lymphoid irradiation

Three cases of COT have been described in RT recipients subjected to total lymphoid irradiation, a perioperative course of antithymocyte globulin, and maintenance prednisone that was gradually weaned and eventually stopped [Strober et al., 1989; Strober et al., 2000; Strober et al., 2004]. Two patients remained IS-free for 12 years and 69 months, while the third developed severe urinary tract obstruction 10 months after the IS withdrawal, for which he was eventually retransplanted. Importantly, the complete original series comprised 28 RT recipients, 25 of whom did not develop COT.

2.2.4 COT developed after intentional IS withdrawal for lymphoproliferative disorders

Two cases of COT developing after the intentional withdrawal of IS due to the development of post transplant lymphoproliferative disorders (PTLD) have been described, mainly within a larger series [Roussey-Kesler et al., 2006]. A third case relates to a 21-year-old man who received 2 haplo-identical RTs, the first at age 11 from his mother and the second at age 15 from his father [Christensen et al., 1998]. Three years after the second RT, he developed PTLD. IS was promptly interrupted and the PTLD subsequently resolved. At the time of publication, IS had been withdrawn for 3 years with no rejection.
| Classification | Subtype       | Center                                      | Type of study                  | Claimed cases of COT | Length of follow up from the time of the withdrawal of the IS |
|----------------|---------------|---------------------------------------------|--------------------------------|-----------------------|---------------------------------------------------------------|
| Cell-based     | Previous BMT  | Boston, Brigham and Women Hospital [Sayegh et al, 1991] | Case report                   | 2*                    | 7 and 3 years                                                |
|                |               | Geneva [Helg et al, 1994]                   | Case report                   | 1                     | 2 years                                                      |
|                |               | Copenhagen [Jacobsen et al, 1994]           | Case report                   | 1                     | 1 year                                                       |
|                |               | San Francisco [Sorof et al, 1995]           | Case report                   | 1                     | 2 years                                                      |
|                |               | Milwaukee [Buchter et al, 1999]             | Case report                   | 3                     | 3, 11, 18 years                                              |
|                |               | Birmingham, US [Sellers et al, 2001]        | Case report                   | 1                     | 7                                                            |
| HSC            |               | Boston, Massachusetts General Hospital [Spitzer et al, 1999; Buhler et al, 2002; Fudaba et al, 2006; Spitzer et al, 2011] | Prospective, non comparative | 3/6                   | 12.1, 7.7, 4, 6.8 years                                      |
|                |               | Boston, Massachusetts General Hospital [Kawai et al, 2008; Porcheray et al, 2009] | Prospective, non comparative | 4/5                   | 4.6, 3.4, 2.2 and 1.2 years                                  |
|                |               | Stanford [Millan et al, 2002]              | Prospective, non comparative  | 1/4                   | 14 years                                                     |
|                |               | Stanford [Scandling et al, 2008]            | Prospective, non comparative  | 1/3                   | 28 months                                                    |
| Classification | Subtype | Center | Type of study | Claimed cases of COT | Length of follow up from the time of the withdrawal of the IS |
|----------------|---------|--------|---------------|----------------------|----------------------------------------------------------|
| Non cell-based | Non-adherence to IS | Columbus-Madison [VanBuskirk et al, 2000; Geissler et al, 2001] | Case report | 3 | 3 years |
| | | Madison [Uehling et al, 1976] | Case report | 5, but 4 rejected after few months | 5 years |
| | | Los Angeles [Owens et al, 1975] | Case report | 4, but eventually 1 rejected after 18 months | 17, 23, 52 months |
| | | Minneapolis [Najarian, 1975] | Case report | 6, but 5 rejected after few months | NA |
| | | Madison [Hussey, 1975] | Case report | 8, but 7 rejected after few months | 40 months |
| | Non cell-based | TBI | Stanford [Strober et al, 1989; Strober et al, 2000] | Prospective | 3/28, but 1 rejected after 10 months for non-immunological reasons | 144 and 69 months |
| | Pregnancy | Pregnancy | Erlangen-Munster [Fischer et al, 1996] | Case report | 1 | 9 years |
| Classification | Subtype | Center | Type of study | Claimed cases of COT | Length of follow up from the time of the withdrawal of the IS |
|----------------|---------|--------|---------------|----------------------|-------------------------------------------------------------|
| PTLD           |         | Aarhus [Christensen et al, 1998] | Case report | 1                    | >3 years                                                   |
| Molecule-based |         | Pittsburgh [Starzl et al, 2003] | Prospective, non comparative | 0/39 | - |
|                |         | Cambridge [Clatworthy et al, 2009; Watson et al, 2005; Calne et al, 2000; Calne et al, 1999; Calne et al, 1998] | Prospective, comparative, non randomized | 0/33 | - |
|                |         | Oxford [Trzonkoski et al, 2008] | Prospective, non comparative | 0/13 | - |
|                |         | Bethesda [Kirk et al, 2003] | Prospective, non comparative | 0/7 | - |
|                |         | Bethesda [Kirk et al, 2005] | Prospective, non comparative | 0/5 | - |
| Surveys        |         | Boston [Zoller et al, 1980] | Descriptive, observational | 13 | >1 year |
|                |         | Nantes [Roussey-Kesler et al, 2006] | Descriptive, investigative | 10, but 2 rejected after 7 and 13 years | 9 years [mean time, range 1-20] |
|                |         | Nantes [Braud et al, 2008; Sivozhelezov et al, 2008] | Descriptive, investigative | 8 | >1 year |
|                |         | Nantes [Bouard et al, 2007] | Descriptive, investigative | 17 | >1 year |
|                |         | Los Angeles [Owens et al, 1975] | Descriptive, observational | 24, but 22 rejected after few months | 9, 36 months |
Table 1. Synoptic view of all successful and unsuccessful cases of COT described after RT. The mismatch between the number of cases claimed by numerous authors and the real effective number of COT calculated according to the definition of COT adopted in the present manuscript is evident [116 vs. 108, respectively].

3. Immune monitoring

As illustrated above, most cases of COT have developed in individuals who have spontaneously terminated IS. When all clinical trials in which a presumed tolerogenic protocol has been implemented are taken into consideration, it is frustrating to observe that COT could be obtained in only 6 out of 248 patients, accounting for a poor, unacceptable success rate of 2.5% [Orlando, Hematti et al., 2010]. As a corollary, the tolerogenic regimens attempted so far in RT recipients are not efficient and, more importantly, lack safety. Ideally, before implementing any of such regimens, investigators should rely on parameters able to predict with high accuracy whether patients would tolerate the weaning process without any risk to reject. Unfortunately, no parameter as such is available for routine practice, not even renal biopsy. For example, Burlingham reported on a late graft rejection in a patient who received a RT 9.5 years earlier from his mother and who had been IS-free for 7 years; a gradual rise in serum creatinine level to 2.0 mg/dl prompted a biopsy that ruled out rejection, yet 10 months later severe cellular rejection arose [Burlingham et al., 2000]. Investigators have obtained promising results in the field of liver transplantation where this problem has been circumvented by the identification of so-called markers of tolerance [Martinez-Llordella et al., 2008; Martinez-Llordella et al., 2007], defined as functional and molecular correlates of immune reactivity whose purpose is to provide clinically useful information for therapeutic decision-making in view of IS withdrawal [Ashton-Chess et al., 2009]. Investigations in tolerant liver transplant recipients resulted in the discovery and validation of a tolerance-associated transcriptional patterns, consisting of several gene signatures and multiple peripheral blood lymphocyte subsets capable of identifying tolerant and non-tolerant recipients with high accuracy. As these data suggested that transcriptional profiling of peripheral blood can be employed to identify recipients who can discontinue immunosuppressive therapy at no risk for rejection, RT researchers are currently exploring a
signature of COT after RT which may allow the selection of those patients who may be more prone to develop an IS-free state with no or quasi no risk for rejection.

The group in Nantes has pioneered investigations aimed to the identification of specific biological signatures of COT [Braud et al., 2008; Brouard et al., 2007; Sivozhelezov et al., 2008]. Brouard et al. identified a set of 49 genes and differentially expressed gene transcripts using gene-expression profiling of peripheral blood from 17 tolerant RT recipients, with tolerance class prediction scores of >90% [Brouard et al., 2007]. This fingerprint is expected to identify patients who might be eligible for a progressive tapering of their immunosuppressive medications and, more importantly, those who instead need to stay on their current IS regimen. The same group has also exploited the capabilities of high throughput microarray technology to study peripheral blood specific gene expression profiles and corresponding molecular pathways associated with operational tolerance [Braud et al., 2008; Sivozhelezov et al., 2008]. Investigations revealed that tolerant patients display a set of 343 differentially expressed genes, mainly immune and defense genes, in their peripheral blood mononuclear cells (PBMC), of which 223 were also different from healthy volunteers. Using the expression pattern of these 343 genes, they were able to correctly classify >80% of the patients in a cross-validation analysis and correctly identified all of the samples over time. All together, this study identified a unique PBMC gene signature associated with human operational tolerance in kidney transplantation [Orlando, Hematti et al., 2010].

Investigations have been conducted in parallel in Europe and the United States. The European Union Indices of Tolerance Network showed that IS-free RT patients present a distinctive expansion of peripheral blood B lymphocytes and natural killer cells and differential expression of several immune-relevant genes in the absence of donor-specific antibodies [Sagoo et al., 2010]. Similar population expansion of B immune cells and selective expression of B cell-related genes in samples obtained from tolerant individuals were noted by the National Institute of Health’s Immune Tolerance Network [Newell et al., 2010].

4. The potential of regenerative medicine

The term regenerative medicine refers to that field in the health sciences which aims to replace or regenerate human cells, tissues, or organs in order to restore or establish normal function [Mason & Dunnill, 2008; Orlando, Baptista et al., 2010; Orlando et al., 2011]. The process of regenerating body parts can occur in vivo or ex vivo and may require cells, natural or artificial scaffolding materials, growth factors, or different combinations of all three elements. Instead, the term tissue engineering refers to a field which is narrower in scope and strictly limited to the production of body parts ex vivo, by seeding cells either on or into a supporting scaffold [Orlando et al., 2011].

In the last decade, investigators in the field have been able to produce and implant in patients relatively simple hollow organs like skin [Naughton et al., 1999], vessels [Hibino et al., 2010; L’Heureux N et al., 2007; McAllister et al., 2009; Shinoka et al., 2001; Shinoka et al., 2005], bladders [Atala et al., 2006], windpipes [Macchiarini et al., 2008] and urethras [Raya-Rivera et al., 2011]. Importantly, all constructs were manufactured by either combining autologous cells with scaffolding material, or through the engineering of autologous cells per se [Orlando, Baptista et al., 2010; Orlando et al., 2011]. Importantly, none of the patients did require IS at any time after implantation of the bioengineered body part. Therefore,
regenerative medicine has shown the potential to offer a valuable approach to COT. Importantly, as the above mentioned implanted body parts were bioengineered from autologous cells, recipients never required IS and therefore we can conclude that regenerative medicine offers the only possible approach to immediate, stable and durable COT. In fact, as illustrated throughout the whole chapter, all cases of COT reported so far in the English literature developed at least several weeks after the transplants and always required some IS, the exception being RT between identical twins [Orlando, Hematti et al., 2010]. Moreover, in several circumstances, patients who could be weaned off IS eventually rejected and required resumption of IS.

Bioengineering technology has been implemented to manufacture heart [Ott et al., 2008], liver [Badylak et al., 2011; Baptista et al., 2011; Uydun et al., 2010; Petersen et al., 2010] organoids from rodent organs [Orlando, Baptista et al., 2010; Orlando et al., 2011]. In extenso, rat or ferret organs were decellularized with detergents and repopulated with cells from unrelated species, humans included. In the ideal scenario, when and if this technology will be perfected, the cellular compartment will be reconstituted – even in this case – from patient’s autologous cells, whereas the supporting scaffold will be represented by an acellular porcine or non-human primate organ depleted of its native cells.

5. Conclusions

When all the above information is taken together, it is clear that, although a wealth of knowledge exists, little progress has been made in developing a sure-fire strategy towards attaining COT. Despite tolerance has been widely investigated for decades, efforts to understand the mechanisms underlying this phenomenon and how to achieve it have thus far been to no avail. In addition to the failure of all molecule-based strategies, we have learned that stem cells do exert some modulatory effect on the immune system but we do not know why and how this occurs. Therefore, we cannot predict when the opportunities for COT to develop in a patient the greatest.

As it stands, with the technology that is currently available, the withdrawal of IS after RT cannot yet be encouraged because it is neither effective nor safe and must be considered as still in an experimental phase. Efforts to identify a peripheral blood transcriptional biomarker panel associated with COT after RT are certainly laudable but, provided the safety for the withdrawal of IS is not guaranteed, any clinical implementation should be banned outside specialized cutting-edge center. The lack of safety of all tolerogenic strategies implemented so far remains their major weakness. Recent revolutionary progress in regenerative medicine has revealed the immense potential of the field pertinent to COT. In a foreseeable future, regenerative medicine will meet the two major needs of SOT, namely that of a potentially inexhaustible source of organs and COT itself [Orlando, 2011].

6. References

Agarwal, A., Shen, LY., & Kirk, AD. (2008). The role of alemtuzumab in facilitating maintenance immunosuppression minimization following solid organ transplantation. Transplant Immunology, Vol.20, No.1-2, (November 2008), pp. 6-11, ISSN 0966-3274

Ashton-Chess, J., Giralt, M., Soulillou, JP., & Brouard, S. (2009). Can immune monitoring help to minimize immunosuppression in kidney transplantation? Transplant
allograft tolerance. *Proceedings of the National Academy of Sciences of the United States of America*, Vol.104, No.39, (September 2007), pp. 15448-15453, ISSN 0027-8424

Bühler, LH., Spitzer, TR., Sykes, M., Sachs, DH., Delmonico, FL., Tolkoff-Rubin, N., Saidman, SL., Sackstein, R., McAfee, S., Dey, B., Colby, C., & Cosimi, AB. (2002). Induction of kidney allograft tolerance after transient lymphohematopoietic chimerism in patients with multiple myeloma and end-stage renal disease. *Transplantation*, Vol.74, No.10, (November 2002), pp. 1405-1409, ISSN 0041-1337

Burlingham, WJ., Jankowska-Gan, E., VanBuskirk, A., Orosz, CG., Lee, JH., & Kusaka, S. (2000). Loss of tolerance to a maternal kidney transplant is selective for HLA class II: evidence from trans-vivo DTH and alloantibody analysis. *Human Immunology*, Vol.61, No.12, (December 2000), pp. 1395-1402, ISSN 0198-8859

Butcher, JA., Hariharan, S., Adams, MB., Johnson, CP., Roza, AM., & Cohen, EP. (1999). Renal transplantation for end-stage renal disease following bone marrow transplantation: a report of six cases, with and without immunosuppression. *Clinical Transplantation*, Vol.13, No.4, (August 1999), pp. 330-335, ISSN 0902-0063

Calne, R., Moffatt, SD., Friend, PJ., Jamieson, NV., Bradley, JA., Hale, G., Firth, J., Bradley, J., Smith, KG., & Waldmann, H. (2000). Prope tolerance with induction using Campath 1H and low-dose cyclosporin monotherapy in 31 cadaveric renal allograft recipients. *Nippon Geka Gakkai Zasshi*, Vol.101, No.3, (March 2000), pp. 301-306, ISSN 0301-4894

Calne, R., Moffatt, SD., Friend, PJ., Jamieson, NV., Bradley, JA., Hale, G., Firth, J., Bradley, J., Smith, KG., & Waldmann, H. (1999). Campath 1H allows low-dose cyclosporine monotherapy in 31 cadaveric renal allograft recipients. *Transplantation*, Vol.68, No.10, (November 1999), pp.1613-1616, ISSN 0041-1337

Calne, R., Friend, P., Moffatt, S., Bradley, A., Hale, G., Firth, J., Bradley, J., Smith, K., & Waldmann H. (1998). Prope tolerance, perioperative Campath 1H, and low-dose cyclosporin monotherapy in renal allograft recipients. *Lancet*, Vol.351, No.9117, (June 1998), pp. 1701-1702, ISSN 0140-6736

Chiang, KY., & Lazarus, HM. (2003). Should we be performing more combined hematopoietic stem cell plus solid organ transplants? *Bone marrow transplantation*, Vol.31, No.8, (April 2003), pp. 633-642, ISSN 0268-3369

Christensen, LL., Grunnet, N., Rüdiger, N., Møller, B., & Birkeland, SA. (1998). Indications of immunological tolerance in kidney transplantation. *Tissue Antigens*, Vol.51, No.6, (June 1998), pp. 637-644, ISSN 0001-2815

Ciancio, G., & Burke, GW. 3rd. (2007). Alemtuzumab (Campath-1H) in kidney transplantation. *American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons*, Vol.8, No.1, (December 2007), pp.15-20, ISSN 1600-6135

Ciancio, G., Burke, GW., Moon, J., Garcia-Morales, R., Rosen, A., Esquenazi, V., Mathew, J., Jin, Y., & Miller, J. (2004). Donor bone marrow infusion in deceased and living donor renal transplantation. *Yonsei medical journal*, Vol.45, No.6, (December 2004), pp. 998-1003, ISSN 0513-5796 2004;45:998-1003.

Clatworthy, MR., Friend, PJ., Calne, RY., Rebello, PR., Hale, G., Waldmann, H., & Watson, CJ. (2009). Alemtuzumab (CAMPATH-1H) for the Treatment of Acute Rejection in Kidney Transplant Recipients: Long-Term Follow-Up. *Transplantation*, Vol.87, No.7, (April 2009), pp.1092-1095, ISSN 0041-1337
Crop, M., Baan, C., Weimar, W., & Hoogduijn, M. (2009). Potential of mesenchymal stem cells as immune therapy in solid-organ transplantation. *Transplant International: official journal of the European Society for Organ Transplantation*, Vol.22, No.4, (April 2009), pp. 365-376, ISSN 0934-0874

Delis, S., Ciancio, G., Burke, GW., Garcia-Morales, G., & Miller, J. (2004). Donor bone marrow transplantation, chimerism and tolerance. (2004). *Transplant Immunology*, Vol.13, No.2, (September-October 2004), pp. 105-115, ISSN 0966-3274

Ding, Y., Xu, D., Feng, G., Bushell, A., Muschel, RJ., & Wood, KJ. (2009). Mesenchymal stem cells prevent the rejection of fully allogenic islet grafts by the immunosuppressive activity of matrix metalloproteinase-2 and -9. *Diabetes*, Vol.58, No.8, (June 2009), pp. 1797-1806, ISSN 0012-1797

Eason, JD., Cohen, AJ., Nair, S., Alcantera, T., & Loss, GE. (2005). Tolerance: is it worth the risk? *Transplantation*, Vol.79, No.9, (May 2005), pp.1157-1159, ISSN 0041-1337

Fehr, T., & Sykes, M. (2004). Tolerance induction in clinical transplantation. *Transplant Immunology*, Vol.13, No.2, (September-October 2004), pp. 117-130, ISSN 0966-3274

Fischer, T., Schobel, H., & Barenbrock, M. (1996). Specific immune tolerance during pregnancy after renal transplantation. *European journal of obstetrics, gynecology, and reproductive biology*, Vol.70, No.2, (December 1996), pp. 217-219, ISSN 0301-2115

Fudaba, Y., Spitzer, TR., Shaffer, J., Kawai, T., Fehr, T., Delmonico, F., Preffer, F., Tolkoff-Rubin, N., Dey, BR., Saidman, SL., Kraus, A., Bonnafoix, T., McAfee, S., Power, K., Kattleman, K., Colvin, RB., Sachs, DH., Cosimi, AB., & Sykes, M. (2006). Myeloma responses and tolerance following combined kidney and nonmyeloablative marrow transplantation: in vivo and in vitro analyses. *American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons*, Vol.6, No.9, (September 2006), pp. 2121-2133, ISSN 1600-6135

Geissler, F., Jankowska-Gan, E., DeVito-Haynes, LD., Rhein, T., Kalayoglu, M., Sollinger, HW., & Burlingham, WJ. (2001). Human liver allograft acceptance and the "tolerance assay": in vitro anti-donor T cell assays show hyporeactivity to donor cells, but unlike DTH, fail to detect linked suppression. *Transplantation*, Vol.72, No.4, (August 2001), pp. 571-580, ISSN 0041-1337

Helg, C., Chapuis, B., Bolle, JF., Morel, P., Salomon, D., Roux, E., Antonioli, V., Jeannet, M., & Leski, M. (1994). Renal transplantation without immunosuppression in a host with tolerance induced by allogeneic bone marrow transplantation. *Transplantation*, Vol.58, No.12, (December 1994), pp. 1420-1422, ISSN 0041-1337

Hutchinson, JA., Brem-Exner, BG., Riquelme, P., Roelen, D., Schulze, M., Ivens, K., Grabensee, B., Witzke, O., Philipp, T., Renders, L., Humpe, A., Sotnikova, A., Matthäi, M., Heumann, A., Gövert, F., Schulte, T., Kabelitz, D., Claas, FH., Geissler, EK., Kunzendorf, U., & Fändrich, F. (2008). A cell-based approach to the minimization of immunosuppression in renal transplantation. *Transplant International: official journal of the European Society for Organ Transplantation*, Vol.21, No.8, (August 2008), pp. 742-754, ISSN 0934-0874

Hutchinson, JA., Riquelme, P., Brem-Exner, BG., Schulze, M., Matthäi, M., Renders, L., Kunzendorf, U., Geissler, EK., & Fändrich, F. (2008). Transplant acceptance-inducing cells as an immune-conditioning therapy in renal transplantation. *Transplant International: official journal of the European Society for Organ Transplantation*, Vol.21, No.8, (August 2008), pp. 728-741, ISSN 0934-0874
Hematti, P. (2008). Role of mesenchymal stromal cells in solid organ transplantation. *Transplantation reviews*, Vol.22, No.4, (October 2008), pp. 262-273, ISSN 0955-470X

Hibino, N., Mcgillicuddy, E., Matsumura, G., Ichihara, Y., Naito, Y., Breuer, C., & Shinoka, T. (2010). Late-term results of tissue-engineered vascular grafts in humans. *The journal of thoracic and cardiovascular surgery*, Vol.139, No.2, (February 2010), pp.431-436, 436.e1-2, ISSN 0022-5223

Hoogduijn, MJ., Popp, FC., Grohne, A., Crop, MJ., van Rhijn, M., Rowshani, AT., Eggenhofer, E., Renner, P., Reinders, ME., Rabelink, TJ., van der Laan, LJ., Dor, HJ., Ijzermans, JN., Genever, PG., Lange, C., Durrbach, A., Houtgraaf, JH., Christ, B., Seifert, M., Shagidulin, M., Donckier, V., Deans, R., Ringden, O., Perico, N., Remuzzi, G., Bartholomew, A., Schlitt, HJ., Weimar, W., Baan, CC., & Dahlke, MH.; MISOT Study Group. (2010). Advancement of mesenchymal stem cell therapy in solid organ transplantation (MISOT). *Transplantation*, Vol.90, No.2, (July 2010), pp. 124-126, ISSN 0934-0874

Hussey, JL. (1976). Letter: Discontinuance of immunosuppression. *Archives of surgery*, Vol.111, No.5, (May 1976), p. 614, ISSN 0272-5533

Issa, F., Hester, J., Goto, R., Nadig, SN., Goodacre, TE., & Wood, K. (2010). Ex vivo-expanded human regulatory T cells prevent the rejection of skin allografts in a humanized mouse model. *Transplantation*, Vol.90, No.12, (December 2010), pp. 1321-1327, ISSN 0934-0874

Issa, F., Schiopu, A., & Wood, KJ.(2010). Role of T cells in graft rejection and transplantation tolerance. *Expert review of clinical immunology*, Vol.6, No.1, (January 2010), pp. 155-169, ISSN 1744-666X

Jacobsen, N., Taaning, E., Ladefoged, J., Kristensen, JK., & Pedersen, FK. (1994). Tolerance to an HLA-B,DR disparate kidney allograft after bone-marrow transplantation from same donor. *Lancet*, Vol.343, No.8900, (March 1994), p. 800, ISSN 0140-6736

Kawai, T., Cosimi, AB., Spitzer, TR., Tolkoff-Rubin, N., Suthanthiran, M., Saidman, SL., Shaffer, J., Preffer, F., Ding, R., Sharma, V., Fishman, JA., Dey, B., Ko, DS., Hertl, M., Goes, NB., Wong, W., Williams, WW. Jr, Colvin, RB., Sykes, M., & Sachs DH. (2008). HLA-mismatched renal transplantation without maintenance immunosuppression. *The New England journal of medicine*, Vol.358, No.4, (January 2008), pp. 353-361, ISSN 0028-4793

Kirk, AD., Mannon, RB., Kleiner, DE., Swanson, JS., Kampen, RL., Cendales, LK., Elster, EA., Wakefield, T., Chamberlain, C., Hoffmann, SC., & Hale, DA. (2005). Results from a human renal allograft tolerance trial evaluating T-cell depletion with alemtuzumab combined with deoxyspergualin. *Transplantation*, Vol.80, No.8, (October 2005), pp. 1051-1059, ISSN 0934-0874

Kirk, AD., Hale, DA., Mannon, RB., Kleiner, DE., Hoffmann, SC., Kampen, RL., Cendales, LK., Tadaki, DK., Harlan, DM., & Swanson, SJ. (2003). Results from a human renal allograft tolerance trial evaluating the humanized CD52-specific monoclonal antibody alemtuzumab (CAMPATH-1H). *Transplantation*, Vol.76, No.1, (July 2003), pp. 120-129, ISSN 0934-0874

Le Blanc, K., Frassoni, F., Ball, L., Locatelli, F., Roelofs, H., Lewis, I., Lanino, E., Sundberg, B., Bernardo, ME., Remberger, M., Dini, G., Egeler, RM., Bacigalupo, A., Fibbe, W., & Ringdén, O.; Developmental Committee of the European Group for Blood and Marrow Transplantation. (2008). Mesenchymal stem cells for treatment of steroid-
resistant, severe, acute graft-versus-host disease: a phase II study. *Lancet*, Vol.371, No.9624, (May 2008), pp. 1579-1586, ISSN 0140-6736

Le Blanc, K., & Ringdén, O. (2007). Immunomodulation by mesenchymal stem cells and clinical experience. *Journal of internal medicine*, Vol.262, No.5, (November 2007), pp. 509-525, ISSN 0954-6820

L’Heureux, N., McAllister, TN., & de la Fuente, LM. (2007). Tissue-engineered blood vessel for adult arterial revascularization. *The New England journal of medicine*, Vol.357, No.14, (October 2007), pp. 1451-1453, ISSN 0028-4793

Light, J., Salomon, DR., Diethelm, AG., Alexander, JW., Hunsicker, L., Thistlethwaite, R., Reinsmoen, N., & Stablein, D. (2007). Bone marrow transfusions in cadaver renal allografts: pilot trials with concurrent controls. Clinical transplantation, Vol.357, No.14, (October 2007), pp.1451-1453, ISSN 0902-0063

Louis, S., Braudeau, C., Giral, M., Dupont, A., Moizant, F., Robillard, N., Moreau, A., Soulillou, JP., & Brouard, S. (2006). Contrasting CD25hiCD4+T cells/FOXP3 patterns in chronic rejection and operational drug-free tolerance. *Transplantation*, Vol.81, No.3, (February 2006), pp. 398-407, ISSN 0934-0874

Macchiarini, P., Jungebluth, P., Go, T., Asnaghi, MA., Rees, LE., Cogan, TA., Dodson, A., Martorell, J., Bellini, S., Parnigotto, PP., Dickinson, SC., Hollander, AP., Mantero, S., Conconi, MT., & Birchall, MA. (2008). Clinical transplantation of a tissue-engineered airway. *Lancet*, Vol.372, No.9655, (December 2008), pp. 2023-2030, ISSN 0140-6736

Martínez-Llordella, M., Lozano, JJ., Puig-Pey, I., Orlando, G., Tisone, G., Lerut, J., Benitez, C., Pons, JA., Parrilla, P., Ramirez, P., Bruguera, M., Rimola, A., & Sánchez-Fueyo, A. (2008). Towards a diagnostic test of operational tolerance in liver transplantation employing transcriptional profiling. *The Journal of clinical investigation*, Vol.118, No.8. (August 2008), pp.2845-2857, ISSN 0021-9738

Martínez-Llordella, M., Puig-Pey, I., Orlando, G., Tisone, G., Ramoni, M., Lerut, J., Rimola, A., Navasa, M., Margarit, C., Bilbao, I., Hernández-Fuentes, M., Soulillou, JP., & Sánchez-Fueyo, A. (2007). Multiparameter immune profiling of operational tolerance in liver transplantation. *American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons*, Vol.7, No.2, (February 2007), pp. 309-319, ISSN 1600-6135

Mason, C., & Dunnill, P. (2008). A brief definition of regenerative medicine. *Regenerative medicine*, Vol.3, No.1 (January 2008), pp. 1-5, ISSN 1746-0751

McAllister, TN., Maruszewski, M., Garrido, SA., Wystrychowski, W., Dusserre, N., Marini, A., Zagalski, K., Fiorillo, A., Avila, H., Manglano, X., Antonelli, J., Kocher, A., Zembala, M., Cierpka, L., de la Fuente, LM., & L'Heureux, N. (2009). Effectiveness of haemodialysis access with an autologous tissue-engineered vascular graft: a multicentre cohort study. *Lancet*, Vol.373, No.9673, (April 2009), pp. 1440-1446, ISSN 0140-6736

Millan, MT., Shizuru, JA., Hoffmann, P., Dejbakhsh-Jones, S., Scandling, JD., Grumet, FC., Tan, JC., Salvatierra, O., Hoppe, RT., & Strober, S. (2002). Mixed chimerism and immunosuppressive drug withdrawal after HLA-mismatched kidney and hematopoietic progenitor transplantation. *Transplantation*, Vol.73, No.9, (May 2002), pp. 1386-1391, ISSN 0934-0874
Operational Tolerance After Renal Transplantation -
Failure of Traditional Approaches versus the Potential of Regenerative Medicine

Monaco, AP., Wood, ML., Maki, T., Madras, PN., Sahyoun, AI., & Simpson, MA. (1985). Attempt to induce unresponsiveness to human renal allografts with antilymphocyte globulin and donor-specific bone marrow. Transpl Proc 1985;27:1312-1314

Monaco, AP., Clark, AW., Wood, ML., Sahyoun, AI., Codish, SD., & Brown, RW.. (1976). Possible active enhancement of a human cadaver renal allograft with antilymphocyte serum (ALS) and donor bone marrow: case report of an initial attempt. Surgery, Vol.79, No.4, (April 1976), pp. 384-392, ISSN 0039-6060

Monaco, AP., & Wood, ML. (1970). Studies on heterologous antilymphocyte serum in mice. VII. Optimal cellular antigen for induction of immunologic tolerance with antilymphocyte serum. Transplantation proceedings, Vol.2, No.4, (December 1970), pp. 489-496, ISSN 0041-1345

Najarian, JS (1975). Editorial comment. Arch Surg 1975;110:1451.

Naughton, G. (1999). The Advanced Tissue Sciences story. Scientific American, Vol.280, No.4 (April 1999), pp. 84-85, ISSN 0036-8733

Newell, KA., Asare, A., Kirk, AD., Gisler, TD., Bourcier, K., Suthanthiran, M., Burlingham, WJ., Marks, WH., Sanz, I., Lechler, RL., Hernandez-Fuentes, MP., Turka, LA., & Seyfert-Margolis, VL.; Immune Tolerance Network ST507 Study Group. (2010). Identification of a B cell signature associated with renal transplant tolerance in humans. The Journal of clinical investigation, Vol.120, No.6, (June 2010), pp. 1836-1847, ISSN 0021-9738

Orlando, G., Wood, KJ., Stratta, RJ., Yoo, J., Atala, A., & Soker, S. (n.d.) Regenerative medicine and organ transplantation: Past, present and future. Transplantation, in press, ISSN 0934-0874

Orlando, G. (2011). Transplantation as a subfield of regenerative medicine. An interview by Lauren Constable. Expert review of clinical immunology, Vol.7, No.???, (Month 2011), pp. 137-141, ISSN 1744-666X

Orlando, G., Baptista, P., Birchall, M., De Coppi, P., Farney, A., Guimaraes-Souza, NK., Opara, E., Rogers, J., Seliktar, D., Shapira-Schweitzer, K., Stratta, RJ., Atala, A., Wood, KJ., & Soker, S. (2010). Regenerative medicine as applied to solid organ transplantation: current status and future challenges. Transplant International: official journal of the European Society for Organ Transplantation, Vol.24, No.3, (March 2011), pp. 223-232, ISSN 0934-0874

Orlando, G., Hematti, P., Stratta, RJ., Burke, GW., Di Cocco, P., Pisani, F., Soker, S., & Wood, KJ. (2010). Clinical operational tolerance after renal transplantation: current status and future challenges. Annals of surgery, Vol.252, No.6, (December 2010), pp. 915-928, ISSN 0003-4932

Orlando, G., Soker, S., & Wood, K. (2009). Operational tolerance after liver transplantation. Journal of hepatology, Vol.50, No.6. (June 2009), pp. 1247-1257, ISSN 0168-8278

Ott, HC., Clippinger, B., Conrad, C., Schuetz, C., Pomerantseva, I., Ikonomou, L., Kotton, D., & Vacanti, JP. (2010). Regeneration and orthotopic transplantation of a bioartificial lung. Nature medicine, Vol.16, No.8, (August 2010), pp. 927-933, ISSN 1078-8956

Ott, HC., Matthiesen, TS., Goh, SK., Black, LD., Kren, SM., Netoff, TL., & Taylor, DA. (2008). Perfusion-decellularized matrix: using nature’s platform to engineer a bioartificial heart. Nature medicine, Vol.14, No.2, (February 2008), pp.213-221, ISSN 1078-8956
Owens, ML., Maxwell, G., Goodnight, J., & Wolcott, MW. (1975). Discontinuance of immunosuppression in renal transplant patients. *Archives of surgery*, Vol.110, No.12, (December 1975), pp.1450-1451, ISSN 0272-5533

Petersen, TH., Calle, EA., Zhao, L., Lee, EJ., Gui, L., Raredon, MB., Gavrilov, K., Yi, T., Zhuang, ZW., Breuer, C., Herzog, E., & Niklason, LE. (2010). Tissue-Engineered Lungs for in Vivo Implantation. *Science*, Vol.329, No.599, (July 2010), pp. 538-541, ISSN 0193-4511

Raya-Rivera, A., Esquiliano, DR., Yoo, JJ., Lopez-Bayghen, E., Soker, S., & Atala, A. (2011). Tissue-engineered autologous urethras for patients who need reconstruction: an observational study. *Lancet*, (n.d.) ISSN 0140-6736

Riquelme, P., Gövert, F., Geissler, EK., Fändrich, F., & Hutchinson, JA. (2009). Human transplant acceptance-inducing cells suppress mitogen-stimulated T cell proliferation. *Transplant immunology*, Vol.21, No.3, (July 2009), pp. 162-165, ISSN 0966-3274

Roussey-Kesler, G., Giral, M., Moreau, A., Subra, JF., Legendre, C., Noel, C., Pillebout, E., Brouard, S., & Soulillou, JP. (2006). Clinical operational tolerance after kidney transplantation. *American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons*, Vol.6, No.4, (April 2006), pp. 736-746, ISSN 1600-6135

Sagoo, P., Perucha, E., Sawitzki, B., Tomiuk, S., Stephens, DA., Miqueu, P., Chapman, S., Craciun, L., Sergeant, R., Brouard, S., Rovis, F., Jimenez, E., Ballow, A., Giral, M., Rebollo-Mesa, I., Le Moine, A., Braudeau, C., Hilton, R., Gerstmayer, B., Bourcier, K., Sharif, A., Krajewska, M., Lord, GM., Roberts, L., Goldman, M., Wood, K.J., Newell, K., Seyfert-Margolis, V., Warners, AN., Jansen, U., Volk, HD., Soulillou, JP., Hernandez-Fuentes, MP., & Lechler, RI. (2010). Development of a cross-platform biomarker signature to detect renal transplant tolerance in humans. *The Journal of clinical investigation*, Vol.120, No.6, (June 2010), pp. 1848-1861, ISSN 0021-9738

Sayegh, MH., Fine, NA., Smith, JL., Rennke, HG., Milford, EL., & Tilney, NL. (1991). Immunologic tolerance to renal allografts after bone marrow transplants from the same donors. *Annals of internal medicine*, Vol.114, No.11, (June 1991), pp. 954-955, ISSN 0003-4819

Scandling, JD., Busque, S., Dejbakhsh-Jones, S., Benike, C., Millan, MT., Shizuru, JA., Hoppe, RT., Lowsky, R., Engleman, EG., & Strober, S. (2008). Tolerance and chimerism after renal and hematopoietic-cell transplantation. *The New England journal of medicine*, Vol.358, No.4, (January 2008), pp. 362-368, ISSN 0028-4793

Sellers, MT., Deierhoi, MH., Curtis, JJ., Gaston, RS., Julian, BA., Lanier, DC. Jr, & Diethelm, AG. (2001). Tolerance in renal transplantation after allogeneic bone marrow transplantation-6-year follow-up. *Transplantation*, Vol.71, No.11, (June 2001), pp. 1681-1683, ISSN 0041-1337

Shinoka, T., Matsumura, G., Hibino, N., Naito, Y., Watanabe, M., Konuma, T., Sakamoto, T., Nagatsu, M., & Kurosawa, H. (2005). Midterm clinical result of tissue-engineered vascular autografts seeded with autologous bone marrow cells. *The Journal of thoracic and cardiovascular surgery*, Vol.129, No.6, (June 2005), pp. 1330-1338, ISSN 0022-5223
Shinoka, T., Imai, Y., & Ikada, Y. (2001). Transplantation of a Tissue-Engineered Pulmonary Artery. *The New England journal of medicine*, Vol.344, No.7, (February 2001), pp. 532-533, ISSN 0028-4793

Sivozhelezov, V., Braud, C., Giacomelli, L., Pechkova, E., Giral, M., Souillou, JP., Brouard, S., & Nicolini, C. (2008). Immunosuppressive drug-free operational immune tolerance in human kidney transplant recipients. Part II. Non-statistical gene microarray analysis. *Journal of cellular biochemistry*, Vol.103, No.6, (April 2008), pp. 1693-1906, ISSN 0730-2312

Sorof, JM., Koerper, MA., Portale, AA., Potter, D., DeSantes, K., & Cowan, M. (1995). Renal transplantation without chronic immunosuppression after T cell-depleted, HLA-mismatched bone marrow transplantation. *Transplantation*, Vol.59, No.11, (June 1995), pp. 1633-1635, ISSN 0041-1337

Spitzer, TR., Delmonico, F., Tolkoff-Rubin, N., McAfee, S., Sackstein, R., Saidman, S., Colby, C., Sykes, M., Sachs, DH., & Cosimi, AB. (1999). Combined histocompatibility leukocyte antigen-matched donor bone marrow and renal transplantation for multiple myeloma with end stage renal disease: the induction of allograft tolerance through mixed lymphohematopoietic chimerism. *Transplantation*, Vol.68, No.4, (August 1999), pp. 480-484, ISSN 0041-1337

Starzl, TE., Murase, N., Abu-Elmagd, K., Gray, EA., Shapiro, R., Eghtesad, B., Corry, RJ., Jordan, ML., Fontes, P., Gayowski, T., Bond, G., Scantlebury, VP., Potdar, S., Randhawa, P., Wu, T., Zeevi, A., Nalesnik, MA., Woodward, J., Marcos, A., Trucco, M., Demetris, AJ., & Fung, JJ. (2003). Tolerogenic immunosuppression for organ transplantation. *Lancet*, Vol.361, No.9368, (May 2003), pp. 1502-1510, ISSN 0140-6736

Strober, S., Lowsky, RJ., Shizuru, JA., Scandling, JD., & Millan, MT. (2004). Approaches to transplantation tolerance in humans. *Transplantation*, Vol.77, No.6, (March 2004), pp. 932-936, ISSN 0041-1337

Strober, S., Benike, C., Krishnaswamy, S., Engleman, EG., & Grumet, FC. (2000). Clinical transplantation tolerance twelve years after prospective withdrawal of immunosuppressive drugs: studies of chimerism and anti-donor reactivity. *Transplantation*, Vol.69, No.8, (April 2000), pp. 1549-1554, ISSN 0041-1337

Strober, S., Dhillon, M., Schubert, M., Holm, B., Engleman, E., Benike, C., Hoppe, R., Sibley, R., Myburgh, JA., Collins, G., & Levin, B. (1989). Acquired immune tolerance to cadaveric renal allografts. A study of three patients treated with total lymphoid irradiation. *The New England journal of medicine*, Vol.321, No.1, (July 1989), pp. 28-33, ISSN 0028-4793

Sykes, M. (2009). Hematopoietic cell transplantation for tolerance induction: animal models to clinical trials. *Transplantation*, Vol.77, No.6, (February 2009), pp. 932-936, ISSN 0041-1337

Trzonkowski, P., Zilvetti, M., Chapman, S., Wieckiewicz, J., Sutherland, A., Friend, P., & Wood, KJ. (2008). Homeostatic repopulation by CD28-CD8+ T cells in alemtuzumab-depleted kidney transplant recipients treated with reduced immunosuppression. *American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons*, Vol.8, No.2, (February 2008), pp. 338-347, ISSN 1600-6135
Uehling, DT., Hussey, JL., Weinstein, AB., Wank, R., & Bach, FH. (1976). Cessation of immunosuppression after renal transplantation. *Surgery*, Vol.79, No.3, (March 1976), pp. 278-282, ISSN 0039-6060

Uygun, BE., Soto-Gutierrez, A., Yagi, H., Izamis, ML., Guzzardi, MA., Shulman, C., Milwid, J., Kobayashi, N., Tilles, A., Berthiaume, F., Hertl, M., Nahmias, Y., Yarmush, ML., & Uygun, K. (2010). Organ reengineering through development of a transplantable recellularized liver graft using decellularized liver matrix. *Nature medicine*, Vol.16, No.7, (July 2010), pp. 814-820, ISSN 1078-8956

VanBuskirk, AM., Burlingham, WJ., Jankowska-Gan, E., Chin, T., Kusaka, S., Geissler, F., Pelletier, RP., & Orosz, CG. (2000). Human allograft acceptance is associated with immune regulation. *The Journal of clinical investigation*, Vol.106, No.1, (July 2000), pp. 145-155, ISSN 0021-9738

Watson, CJ., Bradley, JA., Friend, PJ., Firth, J., Taylor, CJ., Bradley, JR., Smith, KG., Thiru, S., Jamieson, NV., Hale, G., Waldmann, H., & Calne, R. (2005). Alemtuzumab (CAMPATH 1H) induction therapy in cadaveric kidney transplantation--efficacy and safety at five years. *American journal of transplantation: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons*, Vol.5, No.6, (June 2005), pp. 1347-1353, ISSN 1600-6135

Zoller, KM., Cho, SI., Cohen, JJ., & Harrington, JT. (1980). Cessation of immunosuppressive therapy after successful transplantation: a national survey. *Kidney international*, Vol.18, No.1, (July 1980), pp. 110-114, ISSN 0085-2538
Although many years have passed since the first successful kidney transplantation, the method, although no longer considered a medical experiment, is still perceived as controversial and, as such, it triggers many emotions and that’s why conscious educational efforts are still needed for kidney transplantation, for many people being the only chance for an active lifestyle and improved quality of life, to win common social acceptance and stop triggering negative connotations. Apart from transplantation controversies piling up over years transplantologists also have to face many other medical difficulties. The chapters selected for this book are of high level of content, and the fact that their authors come from many different countries, and sometimes even cultures, has facilitated a comprehensive and interesting approach to the problem of kidney transplantation. The authors cover a wide spectrum of transplant-related topics.

How to reference
In order to correctly reference this scholarly work, feel free to copy and paste the following:

Giuseppe Orlando, Pierpaolo Di Cocco, Lauren Corona, Tommaso Maria Manzia, Katia Clemente, Antonio Famulari and Francesco Pisani (2011). Operational Tolerance after Renal Transplantation in the Regenerative Medicine Era, Kidney Transplantation - New Perspectives, Dr Magdalena Trzcinska (Ed.), ISBN: 978-953-307-684-3, InTech, Available from: http://www.intechopen.com/books/kidney-transplantation-new-perspectives/operational-tolerance-after-renal-transplantation-in-the-regenerative-medicine-era