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Barr, I., Rynehart, C., Whitney, P., & Druce, J. (2020). SARS-CoV-2 does not replicate in embryonated hen’s eggs or in MDCK cell lines. Eurosurveillance, 25(25).

https://doi.org/10.2807/1560-7917.ES.2020.25.25.2001122

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Rapid communication

SARS-CoV-2 does not replicate in embryonated hen’s eggs or in MDCK cell lines

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The advent of COVID-19, has posed a risk that human respiratory samples containing human influenza viruses may also contain SARS-CoV-2. This potential risk may lead to SARS-CoV-2 contaminating conventional influenza vaccine production platforms as respiratory samples are used to directly inoculate embryonated hen’s eggs and continuous cell lines that are used to isolate and produce influenza vaccines. We investigated the ability of these substrates to propagate SARS-CoV-2 and found that neither could support SARS-CoV-2 replication.

The majority of the global production of influenza vaccine is currently performed in embryonated hen’s eggs [1] with a growing proportion in mammalian cell lines such as Madin–Darby canine kidney cells (MDCK) [2]. Both of these methods derive their initial viruses from clinical respiratory samples of patients with influenza-like illness (ILI), which are inoculated directly into these substrates [3]. A small proportion of these influenza-positive samples may now be co-infected with severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) and influenza virus, as reported in a recent study by Kim et al. 2020 [4]. They found that 0.9% of samples positive for SARS-CoV-2, from individuals in Northern California, were also positive for influenza virus (with another 5.2% of SARS-CoV-2 cases co-infected with respiratory syncytial virus (RSV), and 15.5% with another respiratory pathogen) in a period towards the end of the influenza season (3–25 March 2020) in the United States (US) [4]. During the peak of future influenza seasons, the rate of SARS-CoV-2/ influenza virus co-infections may increase if, as might be expected, SARS-CoV-2 co-circulates with influenza, thereby enhancing the risk of contamination of influenza vaccine seed stocks. Careful screening using sensitive molecular-based methods should reduce the possibility of this occurring, however, there still exists a theoretical risk of viable virus being present, which may compromise candidate influenza vaccine viruses and increase the risk to workers in the influenza vaccine industry and laboratory scientists, who routinely work under a biosafety level (BSL)2 containment and not the higher levels (BSL3 or BSL4) required to amplify SARS-CoV-2 safely.

We investigated if SARS-CoV-2 could be propagated in embryonated hen’s eggs or the most used mammalian cell lines that are currently used for propagating influenza viruses (in both the diagnostic laboratory and in vaccine production), MDCK cells and variants of this cell line.

Inoculating and passaging SARS-CoV-2 and influenza viruses on influenza virus substrates

Briefly we inoculated two different levels (100 median tissue culture infectious dose (TCID₅₀) and 1,000 TCID₅₀) of Vero (African green monkey kidney cells; ATCC CCL-81) cell-grown SARS-CoV-2 in 100 µL of phosphate buffered saline (PBS) containing 1.45 mg/mL of neomycin sulphate (Pharmacia & Upjohn, Peapack, New Jersey, US) and 0.25 mg/mL of polymyxin B sulphate (Xellia Pharmaceuticals, Copenhagen, Denmark) into the amniotic cavity or the allantoic cavity of three 15-days-old or three 11-days-old, embryonated hen’s eggs (vaccine certified) respectively and incubated them for 3 days at 35 °C. Fluids were harvested from the eggs and pooled according to their inoculum dose and site, then re-passaged a further two times by re-inoculating 100 µL of a 1:2 dilution of fluid in PBS and antibiotics into the same sites into three eggs/site/inoculum dose and harvesting fluids (but not pooling) after 3 days at 35°C. This process mimics influenza
For attempted propagation in mammalian cells, $5 \times 10^4$, $5 \times 10^3$ or $5 \times 10^2$ TCID$_{50}$ inoculums of SARS-CoV-2 derived from infection of Vero cells were applied in Dulbecco’s Modified Eagle Minimum Essential media (DMEM) to T25 flasks (Corning cell culture flasks, surface area 25 cm$^2$, canted neck, cap (vented); Sigma-Aldrich, Sydney, Australia) containing confluent cell monolayers of either regular MDCK cells (ATCC CCL-34), MDCK-SIAT-1 [5], MDCK-TMPRSS2 [6], MDCK-hCK [7] or Vero cells and after 60 min the inoculum was removed and media containing 4 µg/mL trypsin (Sigma-Aldrich) was added and cells were incubated at 37°C for 6 days (as previously described [8]). Subsequently 100 µL samples from this first passage were inoculated onto fresh cell monolayers in T25 flasks as detailed above and incubated at 35°C for a further 6 days. As a control, another set of flasks containing these same cell lines were separately inoculated with human influenza virus isolates (A/Victoria/13/2020 (H1N1pdm09) or B/Victoria/2110/2019 or B/Victoria/2113/2019 (both B/Victoria lineage viruses) for a single passage.

### Table 1
Quantification of SARS-CoV-2$^a$ and influenza$^b$ viruses’ RNA after passages into embryonated hen’s eggs and Vero cells

| Virus Route of inoculation | SARS-CoV-2 TCID$_{50}$ (Ct value) | Passage 1 | Passage 2 | Passage 3 |
|---------------------------|------------------------------------|-----------|-----------|-----------|
| Amniotic                  |                                    |           |           |           |
| E1 100 (28)               |                                    |           |           |           |
| E2 100 (28)               |                                    |           |           |           |
| E3 100 (28)               |                                    |           |           |           |
| E1 1,000 (25)             |                                    |           |           |           |
| E2 1,000 (25)             |                                    |           |           |           |
| E3 1,000 (25)             |                                    |           |           |           |
| Allantoic                 |                                    |           |           |           |
| E1 100 (28)               |                                    |           |           |           |
| E2 100 (28)               |                                    |           |           |           |
| E3 100 (28)               |                                    |           |           |           |
| E1 1,000 (25)             |                                    |           |           |           |
| E2 1,000 (25)             |                                    |           |           |           |
| E3 1,000 (25)             |                                    |           |           |           |
| Vero cells                |                                    |           |           |           |
| VC 100 (28)               |                                    | 17        | NA        | NA        |
| VC 1,000 (25)             |                                    | 15        | NA        | NA        |
| Influenza Allantoic       |                                    |           |           |           |
| • B/Victoria/2110/2019 E  | NA (26)$^d$                        | 11        | NA        | NA        |
| • B/Victoria/2113/2019 E  | NA (27)$^d$                        | 10        | NA        | NA        |

Ct: cycle threshold; E: egg; NA: not applicable/not done; ND: not detected; SARS-CoV-2: severe acute respiratory syndrome coronavirus 2; TCID$_{50}$: median tissue culture infectious dose; VC: Vero cell.

$^a$ Quantification of SARS-CoV-2 RNA after each passage in eggs and Vero cells was assessed with a real-time RT-PCR assay targeting the RNA-dependent RNA polymerase (RdRp) gene.

$^b$ Quantification of influenza virus RNA after one egg passage was assessed with a previously described real-time RT-PCR assay [9].

$^c$ At passage 1, E1, E2, E3 harvests were pooled and then inoculated into three separate eggs at passage 2 then respectively carried onto passage 3.

$^d$ Allantoic isolate initially diluted $10^{-6}$
Assessing SARS-CoV-2 and influenza viruses’ propagation on influenza virus substrates

Samples from the egg-passaged and mammalian cell-passaged SARS-CoV-2 samples were examined for propagation of SARS-CoV-2, visually inspected for embryo viability (eggs), for cytopathogenic effects – CPE (cells), and by real-time RT-PCR (both cells and eggs) targeting the RNA-dependent RNA polymerase (RdRp) gene of SARS-CoV-2 with cycle thresholds (Ct) rounded up or down to the nearest integer. Briefly, for cell samples, 200 µL were subjected to RNA extraction using the QIAmp 96 Virus QIAcube HT kit (Qiagen, Hilden, Germany) and the RNA was eluted in 60 µL, while for egg samples, RNA was extracted from 140 µL with QIAamp mini-Viral RNA columns (Qiagen) and eluted in 60 µL. Reverse transcription and real-time PCR were performed as previously described [8] using an Applied Biosystems Fast Real Time PCR machine (Applied Biosystems, Foster City, California, US). Influenza RNA was detected as previously described [9].

Ethical statement

All experiments with SARS-CoV-2 were conducted in BSL-3 facilities and the use of embryonated hen’s eggs was approved by the University of Melbourne Animal Ethics Committee (2015136.1).

Effects of inoculating SARS-CoV-2 and influenza viruses on hen’s eggs and Vero cells

Table 1 shows the results from the egg-inoculations. The initial inoculum of SARS-CoV-2 had Ct values of 28 for 100 TCID<sub>50</sub> and 25 for 1,000 TCID<sub>50</sub>. Following the first amniotic passage into three individual eggs, the pooled amniotic fluids had a Ct of 38 for the 100 TCID<sub>50</sub> inoculum and 36 for the 1,000 TCID<sub>50</sub> inoculum, and allantoically inoculated eggs gave Cts of 37 and 33 respectively for allantoic fluids. These Ct values continued to increase to undetectable levels (Ct > 40) upon further passaging for both sites of inoculation. In contrast when the 100 TCID<sub>50</sub> and the 1,000 TCID<sub>50</sub> were inoculated into Vero cells after 3 days at 37 °C they gave reduced Ct values of 17 and 15 respectively, indicating a high level of replication had occurred. No effect on embryo viability was detected following inoculation with SARS-CoV-2. Eggs inoculated separately with two influenza B viruses (initial influenza B Cts of 26 and 27) both replicated to high levels as indicated by the highly reduced influenza B Cts of 11 (for B/Victoria/2110/2019) and 10 (for B/Victoria/2113/2019), after a single
passage (Table 1). Eggs inoculated with buffer alone had no detectable SARS-CoV-2 RdRp signal.

**Effects of inoculating SARS-CoV-2 and influenza viruses on MDCK cell lines and Vero cells**

The results testing SARS-CoV-2 propagation in mammalian cell lines are shown in Table 2. At the first passage there was residual SARS-CoV-2 detected in most MDCK cell lines (Ct values ranged from not detected down to 25, with no CPE) while the Vero cells showed lower Cts of 15, 15, 14 in T25 culture flasks (and extensive CPE) when inoculated with $5 \times 10^4$, $5 \times 10^3$, $5 \times 10^2$ TCID$_{50}$ respectively, 6 days after inoculation, indicating high levels of SARS-CoV-2 replication. When a 100µL sample of each MDCK cell line from passage 1 was applied to new cells (of the same type), this second passage showed no CPE or decrease in Ct values, producing signal levels equivalent to background (Cts: 37–39), indicating that no SARS-CoV-2 replication had occurred. In contrast when 100µL from passage 1 in Vero cells (Day 6; originally inoculated with $5 \times 10^2$ TCID$_{50}$ SARS-CoV-2) was applied to fresh Vero cells, this resulted in a Ct of 12, again indicating high levels of replication at this second passage (Table 2).

As a control, MDCK cell lines inoculated separately with either an influenza A or B virus (initial influenza A and B Cts of 25 and 29 respectively) both replicated to high levels as indicated by the highly reduced Cts (influenza B Cts of 25 and 29 respectively) both replicated to high levels at this second passage (Table 2).

Effects of inoculating SARS-CoV-2 and influenza viruses on MDCK cell lines and Vero cells

The studies described above give us confidence that even if a clinical sample, containing both human influenza and SARS-CoV-2, was inoculated into substrates used to prepare seeds for influenza vaccine production (embryonated chicken eggs or MDCK-based cell lines), SARS-CoV-2 would be unlikely to be propagated and would be undetectable after a small number of passages. This finding will reassure influenza vaccine production staff and laboratory scientists who might be concerned about potential exposure to SARS-CoV-2 and also suggests that loss of potentially important influenza candidate vaccine viruses or final vaccine lots due to SARS-CoV-2 contamination is unlikely.

**Conclusions**

The authors would like to thank Rajeev Rudraraju and Francesca Mordant for provision of SARS-CoV-2 virus stock and Vero cells. The Melbourne WHO Collaborating Centre for Research and Reference on Influenza is supported by the Australian Government Department of Health.

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**Conflict of interest**

None declared.

**Authors’ contributions**

IGB conceived the study and wrote the paper. CR, PW and JD conducted the study and edited the manuscript.

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