Research Article

Smooth Muscle Progenitor Cells Preserve the Erectile Function by Reducing Corporal Smooth Muscle Cell Apoptosis after Bilateral Cavernous Nerve Crush Injury in Rats

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Radical prostatectomy causes erectile dysfunction (ED) and irreversible morphologic changes, including induction of endothelial and smooth muscle cell (SMC) apoptosis in the corpus cavernosum (CC). The injection of smooth muscle progenitor cells (SPCs) thickens the vascular intima and has demonstrated therapeutic benefit in cardiovascular disease animal. Herein, we investigated the effect of SPCs on the recovery of erectile function (EF) in rat models with bilateral cavernous nerve (CN) injury. Twenty-four male Sprague-Dawley rats were randomized into sham, vehicle only, or SPC treatment groups. Rats in the SPC treatment and vehicle groups were subjected to bilateral CN injury before intracavernosal injection. Intracavernosal injection of SPCs increased all EF parameters at day 28 after injury and simultaneously reduced apoptosis of the SMCs. Ultrastructural analysis revealed that SPCs maintained the integrity of the CC by preserving the structure of the adherens junctions. Tracking transplanted SPCs labeled with EdU showed that transplanted SPCs remained in the CC 28 days after treatment. Intracavernosal SPC injection restored EF after bilateral CN injury by reducing SMC apoptosis, which favored the maintenance of the structure of adherens junctions and regulated the stability of corporal vessels. These findings demonstrate the therapeutic potential of SPCs for treating ED in humans.

1. Background

In urology, postprostatectomy erectile dysfunction (ED) complication occurring after radical prostatectomy (RP) for earlier prostate cancer is a topic of most concern. Nerve injury may also be the main cause of erectile dysfunction after surgery [1, 2]. Epidemiology of ED after RP range is from 60% to 70%; even when nerve-sparing techniques are applied, ED rates range is still high from 30% to 87% [3]. This type of ED is most likely due to a multifactorial process, including intraoperative neurogenic and vasculogenic injury. For many years, ED after radical surgery was overcome by the implantation of a penile prosthesis [4]. Several treatment approaches are currently available for clinical treatment after radical prostatectomy. Pharmacotherapeutic options include oral treatment (phosphodiesterase 5 (PDE5) inhibitor) and intracavernosal injections (PGE-1). The PDE5 inhibitors showed benefit only in 54% of ED patients in one study [5]. Moreover, PDE5 inhibitors cannot improve the unassisted erectile function (EF) with RP [6]. There is still no progress in terms of effective agents for ED. Our previous study was mainly to find the advance methods to protect the cavernous nerve because a neurogenic component likely plays an important role in the pathogenesis of the ED [7–9]. However, in our recent study, we found that CN injury induces the most severe ED and nerve damage, which is
followed by partial spontaneous recovery of EF and re-
regeneration of the CN at day 28 after injury. Defects in the
corporal SMCs were irreversible after CN injury. This
suggests that the protection of the corpus SMCs from ap-
optosis may represent a more important treatment modality
than nerve protection in future studies using the CN crush
injury model [10]. Therefore, an alternative treatment is
highly desirable to develop an effective, clinical feasible
medication for ED patients after RP.

Several pathophysiological hypotheses have been proposed
to explain ED after CN injury, including nNOS-positive
nerve fiber decrease, vascular damage, corporal cavernosum
inflammatory, cavernosal smooth muscle hypoxia and
smooth muscle apoptosis and fibrosis [11]. Some reports
have stated that neuromodulatory treatments are possible
options, such as use of immunophilin ligands, neuro-
trophins, growth factors, and stem cell therapy, to prevent
the development of hypoxia-induced tissue damage and
fibrosis. Currently, stem cell-based therapy has been a
promising strategy for neuroprotection or tissue-protection
after bilateral CN injury [12]. It is difficult to translate to
clinical use because of adverse pharmacokinetics and be-
cause it is hard for cancer patients to use.

Bone marrow-derived progenitor cells (BMPCs), in-
cluding endothelial progenitor cells (EPCs) and smooth
muscle progenitor cells (SPCs), can be derived from the
autologous peripheral blood, and they differentiate more
easily into vascular cells during arterial remodeling than
stem cells. EPCs are capable of circulating, proliferating, and
differentiating into mature endothelial cells [13]. We have
also previously reported that EPC treatment restored EF in a
rat model of bilateral CN injury through recruitment of
EPCs toward the dorsal artery and preservation of corporal
SMCs. These findings support the therapeutic potential
of progenitor cells for treating ED in humans [14]. SPCs can
limit plaque development and promote changes in plaque
composition toward a stable phenotype in mice [15]. SPC-
released angiopoietin-1 can facilitate stabilization of endo-
thelial cell networks and may be responsible for tightly
orchestrating the complex process of neovascularization.
In addition, the proliferative ability of SPCs in vitro is greater
than that of EPCs [16]. Close cooperation between endo-
thelial cells and SMCs is important for the regulation of
vessel maturation and stability.

The effects of intracavernous injections of SPCs for the
treatment of ED after CN injury have not been explored to
date. Given that SPCs can enhance the efficiency of pro-
angiogenic cell-based therapy [17], the present study
aimed to evaluate the therapeutic potential of SPCs in-
jections into the corpora cavernosa of rats with bilateral CN
injury.

2. Methods

2.1. Experimental Animals. Twenty-five 10-week-old male
Sprague-Dawley rats (weight, 350–450 g) were used in this
study. All animals were supplied by BioLasco Taiwan Co.,
Ltd. (Taipei, Taiwan), and approved by the Fu Jen Catholic
University Animal Care and Use Committee (IACUC
approval no.: A10425). All the animal experiment methods
were conducted in accordance with the guidelines and
regulation established by Fu Jen Catholic University,
Taiwan.

2.2. Isolation and Culture of Smooth Muscle Progenitor Cells. Ten-week-old male Sprague-Dawley rats were used to isolate
the SPCs from femurs. The bone marrow was diluted with
phosphate-buffered saline (PBS). Mononuclear cells were
obtained from the bone marrow by density gradient cen-
trifugation (Histopaque-1083; Sigma–Aldrich, St. Louis,
MO, USA) at 400 ×g for 30 minutes and then at 300 ×g for 10
minutes. The cell pellet was then suspended in endothelial
basal medium-2 (EBM-2; Lonza Clonetics, Walkersville,
MD, USA) supplemented with PDGF-BB (50 ng/mL). After
2-3 days, angioblast-like cells and spindle-shaped cell out-
growths were observed. SPCs were defined by cell mor-
phology and immunofluorescence staining. To determine
the potential activity of SPCs, cells were labeled with a long-
term tracking dye using the EdU Cell Labeling Kit (Invitro-
gen) at different time points (6, 24, and 48 hours) and
concentrations (10, 50, and 100 μM) and then harvested.

2.3. Experimental Design and Surgical Procedures. Cavernous nerve crushing animal model was established by a
standardized procedure to generate a nerve injury as
described previously [7–10]. The rats were randomly
assigned to 3 groups (8 rats per group): sham, vehicle only,
and SPCs-treated groups. For the surgical procedure, the rats
were first anesthetized with an intraperitoneal injection of
sodium pentobarbital (50 mg/kg). After a lower abdomen
midline incision, major pelvic ganglions and the CNs were
observed and isolated. All rats were undergoing bilateral CN
injury except sham group. The crushing injury of the CNs
was applied using a hemostat clamp (Roboz Surgical In-
strument Co. Inc., Gaithersburg, USA) for 2 minutes before
closing the abdomen.

2.4. Measurement of Erectile Responses. The measurement of
erectile function was determined by electrical stimulation of
the CNs at 4 weeks after surgery to generate erectile re-
sponses as described previously [7–10]. The CNs were iso-
lated from prostate, and the crura of the penis were
identified. A 24 G needle containing 50 U/mL of heparin
solution was inserted into the right penile crus. The MP36
pressure transducer (Biopac Systems Inc., CA, USA) was
used to measure the intracavernous pressure (ICP) which
was recorded by BSL 3.7.3 software. Then, the CNs were
stimulated using a bipolar stainless steel electrode and the
monophasic pulses were generated by a DS3 constant
current stimulator (AutoMate Scientific Inc., CA, USA).
The stimulus parameters included a 7.5 mA amplitude, a 20 Hz
frequency, and a 0.2-millisecond pulse width of 60-second
duration. Erectile responses were expressed as intra-
cavernous pressure (ICP). The maximal ICP, the change in
ICP (ΔICP), the area under the ICP curve (AUC), and the
ratio of change in ICP and mean arterial pressure (MAP;
DICP/MAP) were assessed as the parameters of erectile function.

2.5. Immunofluorescence Staining. To determine the content of nNOS-positive nerve fiber and smooth muscle content in the cavernous nerve and penile tissue, we performed double immunofluorescent labeling for cavernous related tissue with antibodies as described previously [9, 10]. The nerve and penile tissue were fixed with formalin for 24 hours (10% formaldehyde w/v) and were subsequently embedded in paraffin. In the deparaffinized procedure, the slide was put in xylene for 10 minutes and this is repeated twice for a total of 3 treatments, followed by hydration in graded alcohols. Blocking procedures were incubated for 1 hour in 10% goat serum/2% bovine serum albumin/0.2% Triton X-100 (Sigma–Aldrich, St. Louis, MO, USA) at room temperature. The tissues were subsequently incubated overnight at 4°C with the following primary antibodies: rabbit anti-nNOS (Santa Cruz Biotechnology, Santa Cruz, CA, USA), mouse anti-neuron-specific β-III tubulin, and anti-α-smooth muscle actin (Abcam, Cambridge, UK). Then, the tissues were incubated with a 1 : 400 dilution of secondary antibody conjugated to Alexa Fluor 488 or Texas Red (Invitrogen, Carlsbad, CA, USA) for 1 hour. For further analysis of nNOS and SMC content by fluorescence microscopy, the ratios of the area of nNOS-positive cells to the area of β-III tubulin-positive cells in nerve fibers and the α-smooth muscle actin area of the corpus cavernosum (CC) were calculated at 400× and 50× magnifications, respectively. All computerized histomorphometric analyses were performed using ImageJ software (National Institutes of Health, Bethesda, MD, USA).

2.6. Statistical Analysis. Data were expressed as the mean± standard deviation. Statistical analysis for comparison of multiple treatment groups was performed with a one-way analysis of variance (ANOVA) and pairwise post hoc comparisons with the Scheffé test by using the SPSS Version 12.0 (SPSS Inc., Chicago, IL, USA) for Windows. The p < 0.05 was considered to be statistically significant.

3. Results

3.1. Cultivation and Characteristic of Smooth Muscle Progenitor Cells. Approximately 30% of the freshly isolated bone marrow mononuclear cells (BMMCs) adhered to
culture plates after 3 days of culture with PDGF-BB-containing EGM-2-MV medium (Figure 1(a)). Two weeks after isolation and differentiation, 30% of outgrowth cells displayed specific morphological characteristics of SPCs as observed through fluorescence microscopy (Figure 1(b)). Hereafter, many SPC colonies were positive with CD34 and α-SMA antigen after differentiation at 4 weeks (about 60% SPC) (Figure 1(c)) and at 5 weeks (about 90% SPC) (Figure 1(d)).

3.2. Smooth Muscle Progenitor Cells Treatment Restores Erectile Function. Figure 2 presents the intracavernosal pressure (ICP) and arterial blood pressure (BP) in the (A) sham, (B) vehicle only, and (C) smooth muscle progenitor cells (SPCs)-injected mice. The x-axis represents time in seconds, and the green bar represents one electrical stimulus of 60 seconds. The y-axis represents the ICP and BP (top and bottom panels) in the experimental animals (n = 8).

![Figure 2: The measurement of intracavernosal pressure. Recordings of the intracavernosal pressure (ICP) and arterial blood pressure (BP) in the (A) sham, (B) vehicle only, and (C) smooth muscle progenitor cells (SPCs-) injected mice. The y-axis represents the ICP and BP (top and bottom panels) in the experimental animals (n = 8).](image)

| Table 1: Intracavernous and mean arterial pressure measurements on cavernous nerve electrostimulation. |
| Groups | Maximum ICP (cm·H₂O) | ΔICP (cm·H₂O) | AUC | ΔICP/MAP | Maximum ICP/BP |
|---------|---------------------|--------------|-----|-----------|----------------|
| Sham    | 141.72 ± 12.91*     | 110.59 ± 12.79* | 4479.86 ± 948.83* | 0.68 ± 0.10* | 0.88 ± 0.12*   |
| Vehicle only | 56.19 ± 9.01        | 33.00 ± 10.12   | 1059.71 ± 413.66  | 0.19 ± 0.06  | 0.33 ± 0.05   |
| SPC     | 127.85 ± 20.89*     | 100.85 ± 25.61* | 5193.72 ± 1585.04* | 0.64 ± 0.14* | 0.81 ± 0.12*   |

SPC = smooth muscle progenitor cell; ICP = intracavernous pressure; ICP = Max ICP-Min ICP; MAP = mean arterial pressure; AUC = area under curve. * P < 0.001 versus vehicle only group.

culture plates after 3 days of culture with PDGF-BB-containing EGM-2-MV medium (Figure 1(a)). Two weeks after isolation and differentiation, 30% of outgrowth cells displayed specific morphological characteristics of SPCs as observed through fluorescence microscopy (Figure 1(b)). Thereafter, many SPC colonies were positive with CD34 and α-SMA antigen after differentiation at 4 weeks (about 60% SPC) (Figure 1(c)) and at 5 weeks (about 90% SPC) (Figure 1(d)).

was not significant. Similarly, other functional parameters of EF, such as ΔICP, area under curve (AUC), ΔICP/BP, and maximum ICP/BP, were significantly increased in the SPC-treated group (Table 1).

3.3. Intracavernous Injection of Smooth Muscle Progenitor Cells Can Prevent Atrophy of Corporal Smooth Muscle Cells. The nNOS-positive nerve fibers of the dorsal penile nerve were assessed by immunofluorescence staining. Figure 3(a) depicts the immunostaining of the dorsal penile nerves for nNOS and β-III tubulin to quantify their nNOS content. Quantitative analysis showed that the ratio of the area of expression of nNOS/β-III tubulin was dramatically reduced in the vehicle only group compared with the sham group; however, there was no significant increase in the number of nNOS-positive nerve fibers after SPC treatment (Figure 3(b)). The corporal SMC content in the corpora cavernosa was evaluated by α-smooth muscle actin staining (Figure 4(a)). The SMC content within the corpora cavernosa was significantly lower in the vehicle only group compared with the sham and the SPC treatment groups (all
These results indicate that SPC can prevent the atrophy of corporal SMCs. Intracavernous injection of smooth muscle progenitor cells maintains the structure of adherens junctions of the smooth muscle in the corpus cavernosum.

Ultrastructural analysis of the CC tissue in the vehicle only and SPC-treated groups at 28 days after injury (Figure 5) was conducted by electron microscopy. Lower-magnification TEM images revealed healthy smooth muscle of the CC near the sinusoid in the sham groups (Figure 5(A)). Healthy smooth muscle was often located along the boundary of the endothelial cells and close to the collagen tissue. Higher-magnification micrographs suggested that the ultrastructure of the adherens junctions in the corporal tissue was intact and closely placed (Figure 5(D)). However, we observed abnormal structures and apoptosis of SMCs in the vehicle only group (Figure 5(B)). In addition, loss of the adherens junctions in the SMC was also observed in the vehicle only group (Figure 5(E)). Few apoptotic cells were observed in the smooth muscle area of CC after SPC treatment (Figure 5(C)). Parts of the adherens junctions between the SMCs were observed in the SPC-treated group, but they exhibited patterns indicating partial destruction of mitochondria (Figure 5(F)).

3.4. Tracking of Smooth Muscle Progenitor Cells in the Corpus Cavernosum. To determine the potential activity of SPCs, cells were labeled with a long-term tracking dye using the EdU Cell Labeling Kit (Invitrogen) at different time points (6, 24, and 48 hours) and concentrations (10, 50, and 100 μM). The optimal time point and concentration for EdU
labeling occurred at 24 hours and at a concentration of 10 μM, respectively (Figure 6(a)). The locations of EdU-positive (stained red) cells were determined near the vascular and nerve bundle at day 3 and day 7 (Figure 6(b)) and in sinusoid of corpus cavernosum at day 28 after SPC injection in the CC (Figure 6(c)).

4. Discussion

In this study, we report the beneficial tissue-protective effects of SPC intracavernosal injection therapy on restoring EF after CN crush injury. Unfortunately, the analysis of variance in nNOS expression revealed that there were no statistically significant differences between the SPC treatment group and the vehicle only group. However, the SPC treatment group showed significantly greater recovery of EF than the vehicle only group, as indicated by the higher levels of all EF parameters evaluated (including maximum ICP, ∆ICP, AUC, and maximum ICP/BP) after treatment with SPCs (for all, *p < 0.001). The beneficial tissue-protective effects of SPCs were attributed to an increased maintenance of the structure of the adherens junctions by SMCs due to the inhibition of cell apoptosis, suggesting their involvement in cavernosal tissue remodeling.

The components of penile erection include the cavernous nerve, the endothelium, and corporal smooth muscle. For optimal EF, accurate interactions among these three components are critical [14]. Following the CN crush in animals,
nNOS expression in the penile nerves rapidly decreases and then begins to increase again over time [18]. The denervation of the penis induces significant apoptosis of corporal SMCs [19]. Corporal smooth muscle fibrosis may cause persistent ED even when other neurological functions recover after operation. In our recent study, we also found the damaged CN can lead to a partial spontaneous regeneration at 28 days after injury. Defects in the corporal SMCs were irreversible after CN crush injury [10]. Treatment with PDE5 inhibitors increases concentrations of cyclic guanosine monophosphate (cGMP); unfortunately, the composition of the corporal SMC will limit the effects of cGMP. Therefore, the main pathway of PDE5 inhibitor activity is disturbed in conditions of corporal SMC fibrosis. This may explain the low success rate of PDE5 inhibitors in treating post-RP ED. Thus, protecting the corporal SMCs from apoptosis using transplanted progenitor cells may be more important than nerve protection in patients undergoing nerve-sparing RP.

Adherens junctions are protein complexes that exist at cell-cell junctions in various cell types including epithelial, endothelial, and muscle cells, which play an essential role in intercellular tissue connections, mechanotransduction, and smooth muscle contraction [20, 21]. In this study, intracavernous injections of SPCs were able to sustain the structure of the adherens junctions of the SMCs, which play a vital role in smooth muscle contraction. In addition, we found that the smooth muscle in the contractile state was abundant around the sinusoid, which suggested that an increased presence of smooth muscle was able to maintain the contractile activity of the SMC intact and resulted in functional improvement. Rats that underwent injection of EdU-labeled SPCs for the purpose of tracking injected SPCs at 4 weeks after CN injury revealed that EdU-labeled SPCs were recruited to the damaged areas of the CC, implying that the therapeutic effect of SPCs was likely due to their mobilization and differentiation.

Currently, cell-based therapy has become a promising strategy for neuroprotection or tissue-protection after bilateral CN injury. The stem cell is capable of renewal and repair of tissue due to its capacity for division and differentiation [22]. Intracavernous injection of embryonic stem cells [23] and adult bone marrow-derived stem cells [24] has been reported to improve EF, but these studies have been limited due to clinical applicability and ethical concerns.

Figure 5: Ultrastructural analysis of the corpus cavernosum (CC) tissue. (a) Transmission electron microscopy (TEM) of the smooth muscle cell (SMC) of the CC near the sinusoid of the sham group. (b) A severe increase in extracellular matrix containing collagen was observed at vehicle only group. (c) Few apoptotic cells are observed in the smooth muscle area of CC after smooth muscle progenitor cells (SPC) treatment. (d) No apparent changes in the adherens junctions of the SMC were observed in the sham group. (e) A few fibroblasts are observed in the smooth muscle apoptotic site and loss of the adherens junctions in the SMC is shown. (f) Part of the adherens junctions between SMCs observed in the SPC group exhibits patterns of partial mitochondrial destruction (red arrow). The upper right square shows higher magnification in all groups.
Figure 6: EdU labeling of smooth muscle progenitor cells (SPCs). (a) SPCs labeled with EdU at 10, 50, and 100 μM and stained with Alexa-594 (red fluorescence) and DAPI (blue fluorescence) (200x magnification). Tracking corpus cavernosum-transplanted SPCs. (b, c) SPCs were labeled with EdU and injected into the corpus cavernosum tissue, which was then harvested at 3, 7, and 28 d and stained with Alexa-594 (red fluorescence) and DAPI (blue fluorescence). The Alexa-594 and DAPI stained images were digitally merged (scale bar = 40 μm and 100 μm).
Although adipose-derived stem cells (ADSCs) have found application in the regeneration of the CN in animal models and have been shown to be successful in the recovery of EF [25], stem cell differentiation does not easily occur during periods of repair [26]. The improvement of EF by ADSCs seems to be attributed to the paracrine action of cytokines rather than to the differentiation of ADSCs [27].

Proangiogenic cell-based therapy using autologous progenitors is a promising strategy for treating ischemic disease [28]. BMPCs, such as EPCs and SPCs, can be derived from peripheral blood and differentiate more easily into vascular cells during arterial remodeling than stem cells. EPCs are capable of circulating, proliferating, and differentiating into mature endothelial cells [13]. EPCs are required for endothelial repair and can participate in blood vessel formation [29]. The cavernous endothelium plays a crucial role in regulating the tone of the underlying smooth muscle and physiological penile erection [30]. We also reported previously that EPC treatment restored EF in a rat model of bilateral CN injury through the recruitment of EPCs toward the dorsal artery and preservation of SMCs in the CC. These findings corroborate the therapeutic potential of EPCs for treating ED in humans [14]. SPC-released angiopoietin-1 can facilitate stabilization of endothelial cell networks and may be more suitable to tightly orchestrate the complex process of neovascularization [16]. In addition, administration of SPCs can thicken the vascular intima and has demonstrated some therapeutic benefit for cardiovascular disease in animal models [31]. A tight cooperation between endothelial cells and SMCs is important to regulate vessel maturation and stability. Foubert et al. stated that SPCs may enhance the efficiency of treatment with EPCs [17]. More importantly, the proliferation ability of SPCs in vitro is higher than that of EPCs. Our study is the first report describing the use of SPCs to treat ED after CN injury and further elucidates the clinical application of SPCs in humans.

5. Conclusions

Treatment with SPCs restored EF in a rat model of bilateral CN crush injury through the recruitment of SPCs toward the sinuses in the corpora cavernosa, which sustained the structure of the adherens junctions of the SMC through a reduction of cellular apoptosis and increased stability of the corporal vessel. These findings provide support for the therapeutic potential of SPCs in treating ED in humans.

Abbreviations

| Abbreviation | Definition                      |
|--------------|--------------------------------|
| ED           | Erectile dysfunction           |
| CC           | Corpus cavernosum             |
| SMC          | Smooth muscle cell            |
| SPCs         | Smooth muscle progenitor cells |
| CN           | Cavernous nerve                |
| EPCs         | Endothelial progenitor cells   |
| BMPCs        | Bone marrow-derived progenitor cells |
| ICP          | Intracavernosal pressure       |
| BP           | Blood pressure                 |
| BMMCs        | Bone marrow mononuclear cells  |

ADSCs: Adipose-derived stem cells.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Ethical Approval

The protocol of this study was reviewed and approved by the Fu Jen Catholic University Animal Care and Use Committee (IACUC approval no.: A10425).

Disclosure

An earlier version of this study was presented as an abstract in the 2018 33rd Joint Annual Conference of Biomedical Science.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Authors’ Contributions

Yi-No Wu contributed to the conception and design, collection and/or assembly of data, data analysis and interpretation, and manuscript writing. Kuo-Chiang Chen approved the final manuscript. Chun-Hou Liao was involved in conception and design, data analysis and interpretation, and final approval of the manuscript. Chien-Liang Liu performed data analysis and interpretation. Han-Sun Chiang was responsible for conception and design and final approval of the manuscript.

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