Activity-based protein profiling reveals that secondary-carbon-centered radicals of synthetic 1,2,4-trioxolanes are predominately responsible for modification of protein targets in malaria parasites†

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Endoperoxide-containing antimalarials, such as artemisinin and the synthetic trioxolane OZ439, are prodrugs activated by heme to generate primary and secondary carbon-centered radicals. We employed activity-based protein profiling (ABPP) to show that the secondary-carbon-centered radical of 1,2,4-trioxolanes is primarily responsible for protein labeling in malaria parasites.

Malaria caused by the protozoal parasite Plasmodium falciparum and related species is a mosquito-borne and life-threatening infectious disease, which leads to nearly a half million deaths globally each year.1 The first small molecule drug used to treat malaria was quinine, extracted from the cinchona tree.2 Due to limited access to this natural source during the Second World War, chloroquine, a synthetically-derived quinoline was identified as an anti-malaria drug.3 Chloroquine and its derivatives were extensively used from the 1930s until the 1950s.4 Unfortunately, widespread resistance to chloroquine eventually developed, prompting a search for next generation anti-malaria drugs. Chinese scientists led by Youyou Tu discovered the fascinating natural product artemisinin (Qinghaosu) and showed that it possesses unparalleled efficacy for treatment of malaria including complicated malaria caused by chloroquine-resistant plasmodial strains.5 Artemisinin established the pivotal role of the endoperoxide as a unique and effective antimalarial pharmacophore.6 The disadvantages of artemisinin are its rapid clearance in vivo, limited supply and impractical de novo synthesis. These challenges stimulated the development of synthetically-accessible and longer-acting trioxolanes, such as OZ277 and OZ439.7 Mechanism of action (MOA) studies revealed that the unique endoperoxide group in artemisinin and synthetic trioxolanes was first activated by heme to generate carbon-centered radicals, which subsequently alkylate multiple proteins in malaria parasites.8 During the activation of the endoperoxide by heme, two different radicals are generated, namely, primary and secondary carbon-centered radicals, each with their own unique properties (Fig. 1A and B).9 The primary carbon-centered radical is sterically smaller while the secondary-carbon-centered radical is more stable.10 However, the relative contributions of each radical for antimalarial activity and specific protein targets have not been elucidated. Herein we determine the protein targets of each radical and further refine

![Fig. 1](https://example.com/fig1.png)

**Fig. 1** Potential reaction pathways of artemisinin (A) and a synthetic trioxolane (B) activated by heme. (a) Rearrangement; (b) 1,5-H shift; (c) cleavage. (C) The structures of probes used in this study, ZJH-1 and ZJH-2. RIPA, radical-induced protein alkylation.

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our mechanistic understanding of endoperoxide-containing anti-
malaria molecules.

To answer the above-mentioned mechanistic questions, we
used activity-based protein profiling (ABPP) to capture the
alkylated proteins of each radical. This was accomplished by
designing two chemical probes containing an alkyne group.
Alkynes are typically used in ABPP because they are sterically
small and can be selectively conjugated to a fluorophore or an
affinity tag employing bioorthogonal click-chemistry.11 Artemisinin
is not an ideal substrate for these studies since both the radicals
remain in the same scaffold (Fig. 1A); thus it is impossible to
discern the specific proteins labeled by each radical regardless
of where the alkyne group is introduced. On the other hand, the
synthetic trioxolane provides an ideal scaffold for our proposed
investigation. After heme-mediated activation followed by rear-
range ment and hydrolysis, synthetic trioxolane is transformed
into either an adamantane ketone and a primary-carbon-centered
radical, or a cyclohexane ketone and a secondary-carbon-centered
radical (Fig. 1B). The protein targets of the two radicals can be
easily traced by selecting appropriate attachment sites for the
alkyne group. Following this design principle, incorporation of
the alkyne into the cyclohexane part (ZJH-1, Fig. 1C) will capture
the protein targets of the primary-carbon-centered radical while
attachment of the alkyne group into the adamantane part (ZJH-2,
Fig. 1C) will capture the protein targets of the secondary-carbon-
centered radical.

Following this rationale, we synthesized probes ZJH-1 and
ZJH-2 (Scheme 1). Oxime 1 and 2 were synthesized via the
reaction between the corresponding ketone and methoxyamine.
The Griesebaum co-ozonolysis of 4-(methoxycarbonyl)cyclohex-
anone with oxime 2 afforded trioxolane 3. Saponification of 3
followed by EDC-mediated coupling with propargyl amine pro-
vided the probe ZJH-1. Probe ZJH-2 was synthesized in a similar
manner as described.8c The chemical structures of probes ZJH-1
and ZJH-2 were confirmed by NMR spectroscopy and high-
resolution mass spectrometry (ESI,† Fig. S1–S14).

We measured the antimalarial activity of probes ZJH-1 and
ZJH-2 against *P. falciparum* 3D7 in vitro. Synchronization of
parasite cultures with two rounds of 5% (w/v) *v*-sorbitol was
performed as previously described.12 Compound 3 was included
as a positive control. The parasite sensitivity of compound 3,
ZJH-1 and ZJH-2 was measured using SYBR Green I as an
indicator.13 The excitation and emission wavelengths of SYBR
Green I were set as 485 and 528 nm, respectively. All three
compounds could effectively inhibit the parasite growth and the
measured IC50 values were 12.67 nM, 7.32 nM and 17.36 nM for
ZJH-1, ZJH-2 and compound 3, respectively (Fig. 2). These results
indicate that incorporation of an alkyne in probes ZJH-1 and
ZJH-2 did not perturb the antimalarial activity confirming that
these probes phenocopy the parent trioxolane scaffold and are
thus suitable for ABPP.

To assess the ability of probes ZJH-1 and ZJH-2 to label
proteins in malaria parasites, different concentrations of probes
ZJH-1 and ZJH-2 ranging from 2 to 20 μM were incubated with
the mixed stage of *P. falciparum* parasite cultures for 4 hours at
37 °C. Red blood cells were removed by lysis with 0.1% saponin
and parasites were pelleted down by centrifugation. The labeled
proteome in the parasites was obtained by sonication and the
probe-modified proteins were labeled with a fluorescent dye
(rhodamine-azide conjugate) using standard copper-catalyzed
click chemistry conditions. The treated proteome was resolved
by denaturing gel electrophoresis and visualized by in-gel fluores-
cence scanning (Fig. 3A and ESI,† Fig. S15). We observed that ZJH-2
efficiently labeled proteins at the lowest probe concentration
examined (2 μM), whereas ZJH-1 displayed virtually no labeling
at this concentration and could only label a few proteins at the
highest concentration evaluated (20 μM). This result indicates
that the secondary-carbon-centered radical predominates protein
labeling of synthetic 1,2,4-trioxolanes in malaria parasites. The
 poor protein labeling of the primary-carbon-centered radical

![Scheme 1 Synthetic routes of probes ZJH-1 and ZJH-2.](image)

![Fig. 2 The viability of malaria parasites treated with 1,2,4-trioxolane-based probes. The viability of malaria parasite was measured in the presence of different concentrations of compound 3 (A), ZJH-1 (B) and ZJH-2 (C). (D) The calculated IC50 of the three compounds (mean ± standard deviation).](image)
could be ascribed to two causes. First, because of the steric hindrance, the coordination of heme to the endoperoxide likely occurs at the least hindered oxygen atom resulting in generation of the secondary-carbon-centered one. Second, the primary-carbon-centered radical is less stable than the secondary carbon-centered one.

To further support the result of gel-based imaging, we next employed gel-free quantitative proteomics to pull down and identify the corresponding protein targets of ZJH-1 and ZJH-2 in parasites. Briefly, the alkyne labeled parasite proteome was reacted with biotin-azide, followed by enrichment with streptavidin beads. Enriched beads were subjected to on-bead digestion with trypsin to release peptide fragments of bound proteins. Collected peptides were separated and detected by LC-MS/MS and the results were processed with the label-free proteomics method to identify the protein targets of ZJH-1 and ZJH-2. During data processing, we used a strict cut-off fold change of 2 and a p-value of 0.05 as the qualification criterion. Accordingly the probe ZJH-1 targeted 22 proteins at the concentration of 20 μM, while ZJH-2 targeted 197 proteins at the concentration of 2 μM (Fig. 3B, ESI,† Table S1). These results confirm that the secondary-carbon-centered radical is much more effective in protein-labeling than the primary-carbon-centered one.

In conclusion, we designed two molecular probes to investigate the protein labeling capacity of primary and secondary carbon-centered radicals generated by synthetic trioxolanes. Both gel-based and gel-free proteomics methods demonstrated that the secondary-carbon-centered radical was responsible for nearly all the protein labeling in situ. These results provide a more nuanced insight into the unique mode of action of the endoperoxide-containing antimalarials. This study may also aid the further development of potent anti-malaria drugs.

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Conflicts of interest

The authors declare that there is no conflict of interest in the publication of this article.

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