Article

A Continuously Tunable Full-Color Emission Nitrogen-Doped Carbon Dots and for Ultrasensitive and Highly Selective Detection of Ascorbic Acid

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Abstract: Nitrogen-doped carbon dots exhibiting excitation-dependent full-color emissions (F-NCDs) were prepared via the one-step hydrothermal method with citric acid and phenylenediamine. Specifically, the emission wavelength of the F-NCDs tuned from 452 nm to 602 nm due to the introduction of new energy levels by C=O and C=N functional groups. We exploited its stability in illumination, ionic strength, and pH, as well as its specificity, sensitivity, especially in ascorbic acid (AA) detection. F-NCDs could measure the AA concentration in the linear ranges of 0~0.1 and 0.1~1 mmol/L with the detection limit (LOD, S/N = 3) as low as 2.6 nmol/L. Additionally, we successfully detected AA in bovine serum with our F-NCDs and obtained the result within 1 min. Because of full-color emission features, we believe our F-NCDs have a great potential in fluorescent sensor detection.

Keywords: nitrogen-doped carbon dots; full-color emission; ascorbic acid; fluorescence detection

1. Introduction

Carbon dots (CDs), which are fluorescent nanoparticles with particle size less than 10 nm, have excellent optical properties and high stability. Moreover, they have several advantages, including simple synthesis, low toxicity, easy modification, good biocompatibility, and good water solubility. Therefore, they have been widely used in many fields, such as drug-loaded therapy [1], bioimaging [2–6], sensors [7], medical diagnosis [8,9] and printing inks [10]. Previous studies focused on their excitation-dependent vs. the excitation-independent features. However, most CDs show their emissions in the blue to green-light regions, which significantly restrict their biological applications. In recent years, the full-color emissions CDs (F-CDs) have been discovered, which display continuously tunable excitation-dependent full-color emission with reliable intensity. With an unaltered chemical structure, they can widely integrate with photoluminescence body and build versatile sensing systems by ignoring the matching of energy gaps. Therefore, it is of great significance to attain the F-CDs for further research and application, particularly in the biological field.
Ascorbic acid (AA) is not only one of the essential nutrients for the human body, but also a good natural antioxidant which has been used as an additive in many commercial products. As one of the essential nutrients, AA plays vital roles in metabolism, body growth, development [11–13], antibody formation, iron absorption [14], folic acid and vitamin E stabilization. By scavenging free radicals, AA can prevent cardiovascular diseases, diabetes, and cancer. Furthermore, AA deficiency even causes cold, scurvy, atherosclerosis, and oxidative damage to lipids, proteins, and DNA [15]. However, excess AA also leads to adverse reactions, including thrombosis, diarrhea, distension, urinary tract, and kidney infection. Therefore, there is an urgent demand to monitor AA concentration. Currently, various methods, including redox titration [16], high performance liquid chromatography (HPLC) [17], electrochemical [18] and fluorescence spectrophotometry analysis [19,20] have been used to detect AA concentration. Considering their excellent optical and chemical properties, CDs could also serve as an ideal fluorescent material to monitor AA concentration with low cost, easy and rapid detection, and high sensitivity and accuracy.

Herein, we present a nitrogen-doped full-color emissions carbon dots (F-NCDs) for ascorbic acid (AA) detection using citric acid and \( p \)-phenylenediamine as raw material. According to detailed structure and properties characterizations, we postulated that the C=O and C=N functional groups imported new energy levels for electronic conversion and generated the continuously tunable excitation-dependent full-color emissions. Furthermore, we successfully applied F-NCDs as a fluorescence sensor to detect ascorbic acid (AA) in serum.

2. Materials and Methods
2.1. Chemicals and Reagents
Citric acid was purchased from Tiantian Chemical Reagent Factory (Tianjin, China). \( p \)-phenylenediamine was obtained from Aladdin Reagent Co., Ltd. (Shanghai, China). Phenylalanine, arginine, glycine, threonine, aspartate, tryptophan, glutamate, and tyrosine were purchased from Tianjin Institute of Fine Chemical Industry Co., Ltd. (Tianjin, China). KCl, MgCl\(_2\), CaCl\(_2\), NaCl, MnSO\(_4\), SrCl\(_2\), CdSO\(_4\), NiSO\(_4\), and Fe\(_2\)(SO\(_4\))\(_3\) were obtained from Tianjin Kaitong Chemical Reagent Co., Ltd. (Tianjin, China). CuSO\(_4\), HgCl\(_2\), PbSO\(_4\), ZnCl\(_2\), AlCl\(_3\), HCl, NaOH, and anhydrous ethanol were acquired from Beijing Chemical Plant (Beijing, China). Quinine sulfate was acquired from Beijing Enoch Technology Co., Ltd. (Beijing, China). All chemicals were analytical pure, and the experimental water was ultrapure water. Fetal bovine serum was obtained from Zhejiang Tianhang Biotechnology Co., Ltd. (Zhejiang, China).

2.2. Synthesis of F-NCDs
F-NCDs were synthetized using a hydrothermal synthesis method. As shown in Figure 1, citric acid and \( p \)-phenylenediamine were dissolved in 10 mL ultrapure water with a mass ratio of 2.5:1. Then, the mixture was autoclaved and left to react at 200 °C for 8 h. After reaction, the sample was cooled naturally to room temperature. Later, the yellow-brown solution to be dialyzed was placed in a sealed dialysis membrane (MWCO = 500 Da) and immersed in ultrapure water for 64 h. After dialysis, solutions were frozen and light yellow carbon dot powder was obtained, named F-NCDs. Furthermore, 5 mg F-NCDs powder was dissolved in 5 mL ultrapure water to create a 1 mg/mL F-NCDs solution.
2.3. Characterization of F-NCDs

A JEM-2100F transmission electron microscope (Tokyo, Japan) was used to acquire high-resolution transmission electron microscope (HRTEM) images of F-NCDs.

The D8-FOCUS X-ray powder diffractometer (Waltham, MA, USA) was applied to measure the XRD spectrum of F-NCDs.

Additionally, the ESCALAB 250Xi X-ray photoelectron spectrometer (Waltham, MA, USA) was used to analyze the composition element and surface functional groups of F-NCDs.

The NICOLET 380 spectroscopy (Waltham, MA, USA) was employed to acquire the Fourier transform infrared (FTIR) spectrum of F-NCDs with KBr particles as the background, with a spectral resolution of 4 cm\(^{-1}\) and scanning range between 4000–500 cm\(^{-1}\).

The TU-1901 UV–vis spectrophotometer (Beijing, China) was utilized to measure the UV–vis spectra of F-NCDs. F-NCDs spectra were acquired with xenon lamps which automatically converts excitation wavelengths as a light source for testing, scan range between 200–600 nm, 2 mm spectral bandwidth, 2.0 nm variable slit, 1 nm scan interval, at medium scanning speed.

Shimadzu RF4301-PC Fluorescence Spectrometer (Kyoto, Japan) was employed to measure the fluorescence spectra of F-NCDs. All optical tests were conducted at room temperature. The conditions were set as follows: the excitation wavelength at 325 nm (\(\lambda_{\text{max}} = 325 \text{ nm}\)), scanning range between 300–600 nm, incident slit width at 5 nm, shot slit width at 5 nm, medium scan speed.

Finally, a FLUOROMAX-4 high sensitivity fluorescence spectrometer (Kyoto, Japan) was used for researching the fluorescence lifetime of F-NCDs.

2.4. Detection of AA

To detect ascorbic acid, 100 \(\mu\)L F-NCDs solution and 2 mL Tris HCl buffer (pH = 7.4) were mixed. Various concentrations of AA solution were added and then the spectra were recorded after 60 s.

3. Results and Discussion

3.1. Characterization of F-NCDs

The HRTEM image of F-NCDs showed that the particles were approximately spherical (Figure 2a), with size distribution between 0.7–3.5 nm, and the average size of 2.0 nm (Figure 2b). The XRD spectrum of F-NCDs showed that there was an obvious diffraction peak around 23° (Figure 2c), which corresponded to the amorphous structure [21,22]. FTIR spectrum of F-NCDs displayed that a typical wide absorption band emerged at
3000~3500 cm$^{-1}$ which corresponded to N–H, O–H and unsaturated C–H stretching vibrations [23–25]. The peaks at 2929 cm$^{-1}$ and 2850 cm$^{-1}$ were caused by saturated C–H bond stretching vibrations. The absorption bands at 1660~1720 cm$^{-1}$ were related to C=O and C=NH stretching vibrations. The shoulders between 1600~1400 cm$^{-1}$ were related to C=C stretching vibration, indicating the possible presence of a benzene ring structure in F-NCDs. In addition, three obvious bands at 1310 cm$^{-1}$, 1170 cm$^{-1}$ and 839 cm$^{-1}$ were correlated to O–H stretching vibration, C–N stretching vibration, and N–H deformation vibration, respectively (Figure 2).

![Figure 2. (a) HRTEM image of F-NCDs; (b) Size distribution of F-NCDs; (c) XRD spectrum of F-NCDs; (d) FTIR spectrum of F-NCDs.](image)

To reveal more detailed information about the functional groups of F-NCDs, we performed an XPS analysis. Full XPS survey spectrum showed the characteristic peaks of C1s, N1s and O1s at 285.19, 398.38 and 532.17 eV, respectively (Figure 3a). A further elemental analysis indicated the composition ratio of C, N and O were 65.95%, 7.7% and 24.55%, respectively. The C1 characteristic peaks were decomposed into three peaks at 283.58, 284.78 and 286.68 eV, which corresponded to the functional groups of C–C/C=C, C–N/C–O and C=NH/C=O, respectively (Figure 3b). The decomposition peaks of N1s were at 396.82 and 398.38 eV, which correlated with C–N and C=NH bond, respectively (Figure 3c). O1s characteristic peaks displays three peaks at 529.88 eV, 530.66 eV and 533.68 eV, which matched with C=O, C–O–C, and O–H, respectively [26–29] (Figure 3d). In conclusion, we speculated that the surface of F-NCDs might contain –OH, –COOH, –NH$_2$, benzene ring,
imine and carbonyl groups, indicating that it can be easily modified and has a good water solubility.

![Graph](image)

**Figure 3.** (a) XPS spectrum of F-NCDs; (b) C1s high-resolution XPS pattern of F-NCDs; (c) N1s high-resolution XPS pattern of F-NCDs; (d) O1s high-resolution XPS pattern of F-NCDs.

### 3.2. Optical Characteristics of F-NCDs

The UV-absorption spectrum of F-NCDs showed a strong absorption peak at 255 nm, suggesting the existence of a B absorption band \((\pi \rightarrow \pi^*)\), correlated to the presence of a benzene ring (Figure 4). Furthermore, a constant intensity of absorption was observed in the range of 300–600 nm. Fluorescent quantum yield, measured with comparative method detection [30], was 67.59% at 345 nm excitation, which using quinine sulfate (QY = 54%) in 0.1 mol/L H$_2$SO$_4$ as the reference.

We decided to examine the excitation and emission properties of F-NCDs afterwards. The F-NCDs exhibited nearly continuous excitation-dependent emission. The emission wavelengths of F-NCDs were dependent on the excitation wavelengths ranging from 305 nm to 585 nm (Figure 5a). Interestingly, the emission spectra were divided into three parts and we noticed three distinct emission peaks of F-NCDs at 452 nm, 517 nm and 602 nm (Figure 5a). When we examined them individually, we found these emission peaks corresponded to an excitation wavelength between 305–385 nm, 395–465 nm and 475–585 nm, respectively (Figure 5b–d).
Nanomaterials 2022, 12, x FOR PEER REVIEW 5 of 14

Figure 3. (a) XPS spectrum of F-NCDs; (b) C1s high-resolution XPS pattern of F-NCDs; (c) N1s high-resolution XPS pattern of F-NCDs; (d) O1s high-resolution XPS pattern of F-NCDs.

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Figure 5. (a) Fluorescence spectra of F-NCDs under various excitation wavelengths (305~585 nm); (b) Fluorescence spectra under the excitation wavelengths between 305~385 nm; (c) Fluorescence spectra under the excitation wavelengths between 395~465 nm; (d) Fluorescence spectra under the excitation wavelengths between 475~585 nm.

To further investigate the tunable change of color under the excitation-dependent fluorescence, we performed 3D fluorescent matrix scans on F-NCDs. It had a broad emission spectrum ranging from 220 to 700 nm, with the excitation wavelengths ranging from 325 to 585 nm with 20 nm increments (Figure 6a). As expected, with a red-shift of the
excitation wavelength, the emission spectrum displayed three fluorescence centers. The emission centers at 432 nm and 602 nm were obvious; however, the center at 517 nm was unnoticeable, possibly due to the low fluorescence intensity.

Figure 6. (a) Excitation-emission 3D matrix of F-NCDs; (b) CIE chromaticity coordinates of F-NCDs.

CIE color coordinates from fluorescent matrix scans of F-NCDs gradually moved from blue to cyan, green, yellow, orange, and eventually red region under different excitation wavelengths displaying from 325 to 585 nm (Figure 6b). Detailed photoluminescence properties of F-NCDs, including \( \lambda_{\text{em}} \), \( \lambda_{\text{ex}} \), \( \Delta \lambda (\lambda_{\text{em}} - \lambda_{\text{ex}}) \), fluorescence intensity, and CIE color coordinates are listed in Table 1. From the results, we observed the irregular change of the fluorescence intensity and \( \Delta \lambda \) (Table 1), suggesting the photoluminescence properties of F-NCDs contributed to multivariate surface states.

Table 1. Photoluminescence properties of F-NCDs.

| \( \lambda_{\text{ex}} / \text{nm} \) | \( \lambda_{\text{em}} / \text{nm} \) | \( \Delta \lambda / \text{nm} \) | Fluorescence Intensity | x       | y       |
|----------------|----------------|----------------|---------------------|--------|--------|
| 325           | 452            | 127            | 795                 | 0.1533 | 0.1425 |
| 345           | 455            | 110            | 642                 | 0.1533 | 0.1487 |
| 365           | 466            | 101            | 431                 | 0.1662 | 0.2165 |
| 385           | 505            | 120            | 216                 | 0.1995 | 0.3590 |
| 405           | 517            | 112            | 227                 | 0.2287 | 0.4705 |
| 425           | 520            | 95             | 171                 | 0.2370 | 0.4819 |
| 445           | 523            | 78             | 109                 | 0.2537 | 0.5324 |
| 465           | 526            | 61             | 49                  | 0.3203 | 0.5905 |
| 485           | 593            | 108            | 31                  | 0.4242 | 0.5591 |
| 505           | 598            | 93             | 46                  | 0.4880 | 0.5069 |
| 525           | 599            | 74             | 85                  | 0.5449 | 0.4537 |
| 545           | 600            | 55             | 174                 | 0.5920 | 0.4074 |
| 565           | 601            | 36             | 317                 | 0.6192 | 0.3803 |
| 585           | 602            | 17             | 600                 | 0.6584 | 0.3413 |

3.3. Full-Color Emission Mechanism of F-NCDs

Optical images of F-NCDs were obtained with excitation-dependent full-color emission and ranged from blue to red (Figure 7). Remarkably, F-NCDs demonstrated selective emission in a wide color range. From these images, we directly observed the fluorescent color changed from blue to cyan, cyan to green, green to yellow, yellow to reddish orange and finally turns red, when the excitation wavelengths ranged from 325–385 nm, 385–465 nm, 465–505 nm, 505–545 nm and 545–585 nm. This information was consistent with the fluorescent color change in the CIE color result.
Figure 7. Fluorescent images of F-NCDs at different excitation wavelengths.

Subsequently, we decided to study potential factors affecting the full-color emission characteristics of F-NCDs. Previous studies suggested that the quantum-size effect, surface states and molecular states might have influenced the emission characteristics of carbon dots [31,32]. We speculated that the full-color emission properties of F-NCDs were determined by its structure, for instance the functional groups C=O and C=N on its surface. Both functional groups produced rich structural arrangements and imported new energy levels into their electronic structures, causing more electronic conversion probabilities [33]. We illustrated the possible energy levels of F-NCDs (Figure 8). For the F-NCDs, HOMO-1 and HOMO energy levels emerged as a result of the introduction of C=O, C=N groups. Then, electron transitions could occur from the two new HOMO-1 and HOMO to the LUMO ($\pi^*$); meanwhile, the excited electrons returned to HOMO-1, or HOMO levels by radiative transition, leading to fluorescence in the green and red regions. When the electrons absorbed short-wave light, electron transitions occurred from HOMO-2, with an energy level to the LUMO ($\pi^*$) level. Then, the excited electrons returned to HOMO-2, HOMO-1, or HOMO levels by radiative transition, and, therefore, caused the broad fluorescence emission of F-NCDs from blue to red region.

Figure 8. Proposed full-color emission mechanism of F-NCDs.

3.4. Application of F-NCDs

To examine the potential application of F-NCDs, we measured its fluorescence stability and analytical performance. To achieve the best specificity and sensitivity, we examined the fluorescence emission spectrum of F-NCDs at 325 nm excitation.

3.4.1. Stability of F-NCDs

To investigate the fluorescence stability of F-NCDs, we explored the fluorescent emission peak under various conditions, including illumination, ionic strength, and pH. After 90 min of continuous UV light irradiation, the fluorescence intensity of F-NCDs remained stable, showing good fluorescence stability (Figure 9a). When we examined the impact of ionic strength (NaCl), the fluorescence intensity of F-NCDs did not show significant change, with an ionic strength up to 1.0 mol·L$^{-1}$, indicating its tolerance to ionic change (Figure 9b). Above all, we confirmed that F-NCDs had stable and excellent optical perfor-
mance even under extreme environmental conditions, indicting its significant potential for sensor application in physiological environment.

![Fluorescence stability of F-NCDs under 365 nm UV light](image1)

**Figure 9.** (a) Fluorescence stability of F-NCDs under 365 nm UV light; (b) Fluorescence stability of F-NCDs under various ionic strengths.

Since pH is a key factor in actual detection, it would be very important to examine the fluorescence stability of F-NCDs under different pH conditions. The fluorescence intensity gradually increased with an increasing pH value from 1 to 6, then greatly increased with an increasing pH from 6 to 7, and finally reached the maximum at pH 7 (Figure 10a). However, the intensity dramatically decreased when the pH value rose from 8 to 14, and it was almost completely quenched with pH value 13 and 14. In conclusion, we postulated that F-NCDs had a good fluorescence stability in an acidic and neutral environment, suggesting its utilization in vivo.

![Fluorescence intensity of F-NCDs under different pH](image2)

**Figure 10.** (a) Fluorescence intensity of F-NCDs under different pH; (b) Response time of F-NCDs with the presence of AA.

Ascorbic acid (AA) interacted with F-NCDs and could quench its fluorescence, we decided to explore its response time when AA was present. As expected, when AA was added, the fluorescence intensity decreased remarkably within 15 s and continued to decline within 45 s; however, after 45 s, the fluorescence intensity was kept constant until 90 s (Figure 10b). To ensure the stability of F-NCDs, 1 min was used as the appropriate reaction time for the following experiments when detecting AA.
3.4.2. Detection of Ascorbic Acid (AA)

To evaluate the response of F-NCDs to AA, we examined its fluorescent emission intensity under pH 7. With rising AA concentrations from 0 mM to 10 mM, the fluorescence strength of F-NCDs gradually decreased (Figure 11a). The relationship between \((F_0 - F)/F_0\) and AA concentration was calculated, where \(F\) and \(F_0\) represent the fluorescence strength of F-NCDs with and without AA, respectively (Figure 11b). In the range of 0–0.1 mM and 0.1–1 mM, it showed a perfect linear relationship, fitting linear equations \((F_0 - F)/F_0 = 1.8020[AA] + 0.0016\) \((R^2 = 0.9945)\) and \((F_0 - F)/F_0 = 0.3698[AA] + 0.1799\) \((R^2 = 0.9992)\), respectively. The lowest detection limit (LOD, \(S/N = 3\)) of AA was 26 nM, indicating its sensitivity for AA detection. Moreover, we compared our F-NCDs results with previous studies and concluded that only our F-NCDs could detect extremely low AA concentration (Table 2).

![Fluorescence emission spectra of F-NCDs in the presence of AA](image)

**Figure 11.** (a) Fluorescence emission spectra of F-NCDs in the presence of AA; (b) the relationship between fluorescence change \((F_0 - F)/F_0\) and AA concentrations.

**Table 2.** Performance comparison of different methods for AA detection.

| Materials Used                  | Method          | Method APPLIED   | linear Range (µmol/L) | LODs (µmol/L) | Ref.     |
|---------------------------------|-----------------|------------------|------------------------|---------------|---------|
| acriflavine                     | spectrofluorometric method | fluorometric       | 11,400–56,800           | 454           | [34]    |
| PAP/ZrO2NPs/CNTs/GCE            | voltammetric     | electrode         | 1–295                  | 0.35          | [35]    |
| boron doped diamond             | voltammetric     | electrode         | 18.5–370               | 5.4           | [36]    |
| CoTMPyP/Sr2Nb3O10 nanocomposite | voltammetric     | electrode         | 50–3250                | 10.6          | [37]    |
| Carbon dots                     | spectrofluorometric method | fluorometric       | 5–50                   | 3.2           | [38]    |
| Carbon dots                     | on-off-on        | fluorometric       | 0–200                  | 0.35          | [39]    |
| Carbon dots                     | on-off-on        | fluorometric       | 5–350                  | 3.11          | [40]    |
| Carbon dots                     | spectrofluorometric method | fluorometric       | 0–1000                 | 0.026         | This work |

To evaluate the specificity of F-NCDs to AA detection, we investigated their performances to various metal ions \((\text{Fe}^{3+}, \text{Cd}^{2+}, \text{Na}^+, \text{Al}^{3+}, \text{Cu}^{2+}, \text{Cr}^{3+}, \text{Mg}^{2+}, \text{Zn}^{2+}, \text{Ni}^{2+}, \text{Ca}^{2+}, \text{Sr}^{2+}, \text{Mn}^{2+}, \text{K}^+, \text{Hg}^{2+})\) and amino acids (arginine (Arg), threonine (Thr), aspartic acid (Asp), glycine (Gly), tyrosine (Tyr), glutamic acid (Glu), serine (Ser) and phenylalanine (Phe)). Compared to other ions and amino acids, only AA exhibited the greatest change in the fluorescence intensity of F-NCDs, indicating the high selectivity of F-NCDs for AA detection (Figure 12a). Furthermore, when interference substances coexisted with AA, AA
still dominated the change in F-NCDs, and the effect caused by the coexistent interference substance could be neglected, further suggesting the specificity of F-NCDs for AA detection (Figure 12b).

Figure 12. (a) Effects of different metal ions and amino acids on fluorescence intensity change of F-NCDs. AA served as a control; (b) Effects of different metal ions and amino acids on fluorescence intensity change of F-NCDs with and without AA.

3.4.3. Mechanism for Ascorbic Acid (AA) Detection

To explore how AA quenched the fluorescence of F-NCDs, we performed UV absorption and fluorescence lifetime experiments. The absorption peak intensity of F-NCDs at 255 nm increased when AA was added. However, no additional absorption peak was observed (Figure 13a). When we measured the average fluorescence lifetime of F-NCDs, the calculated average (amplitude-weighted) lifetime was reduced from 10.2 ns to 7.68 ns with the addition of AA, suggesting that the presence of AA reduced the average lifetime of F-NCDs significantly (Figure 13b). Above all, these findings revealed that the fluorescence quenching mechanism of F-NCDs/AA system was dynamic [30,41,42].

Figure 13. (a) UV-vis spectrum of AA only, F-NCDs only, and F-NCDs with AA; (b) Fluorescence decay curves of F-NCDs with or without AA.

3.4.4. Detection of Ascorbic Acid (AA) in Bovine Serum

To further investigate the feasibility and practical application of F-NCDs, we decided to detect ascorbic acid in the bovine serum. We calculated the original AA concentration...
as 42.28 µmol/L (RSD = 0.20) in animal serum, which was consistent with a previous report. Additionally, three different concentrations (20, 30, 40 µmol/L) of AA were added into the serum and the performance of our F-NCDs was evaluated. As expected, the recovery rate was as high as 99.89~101.54% (RSD < 2.1%) (Table 3). In conclusion, we proved the practicability and accuracy of our F-NCDs in detecting the concentration of AA in animal serum.

Table 3. Detected ascorbic acid (AA) concentration in animal serum.

| No. | Found (µmol/L) | Added (µmol/L) | Theoretical value(µmol/L) | Measured Value (µmol/L) | Recovery (%) | RSD (% n = 3) |
|-----|----------------|----------------|--------------------------|-------------------------|--------------|---------------|
| 1   | 42.28          | 20             | 62.28                    | 63.24                   | 101.54       | 2.1           |
| 2   | 42.28          | 30             | 72.28                    | 72.20                   | 99.89        | 0.5           |
| 3   | 42.28          | 40             | 82.28                    | 82.63                   | 100.43       | 1.1           |

4. Conclusions

In this study, we developed nitrogen-doped full-color emissions carbon dots through a one-step hydrothermal method. The surface states of F-NCDs contributed its tunable excitation-dependent full-color emissions. Our F-NCDs exhibited excellent optical properties, fluorescence intensity and stability. Additionally, based on the dynamic quenching mechanism, the F-NCDs could detect AA in serum with decent results. Compared with other sensors, our F-NCDs do not require any surface modification, and thus it could perform simple, efficient, and direct detection. Overall, our work achieved significant advances in the excitation-dependent full-color emissions of carbon dots, and we believe our F-NCDs could be applied in many areas for detection.

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References

1. Mohammadinejad, R.; Dadashzadeh, A.; Moghassemi, S.; Ashrafizadeh, M.; Dehshahri, A.; Pardakhty, A.; Hassan, H.; Sohrevardi, S.-M.; Mandegary, A. Shedding light on gene therapy: Carbon dots for the minimally invasive image-guided delivery of plasmids and noncoding RNAs. J. Adv. Res. 2019, 18, 81–93. [CrossRef] [PubMed]
2. Pandey, P.K.; Preeti, K.; Rawat, T.; Prasad, H.B. Bohidar, Multifunctional, fluorescent DNA-derived carbon dots for biomedical applications: Bioimaging, luminescent DNA hydrogels, and dopamine detection. J. Mater. Chem. B 2020, 8, 1277–1289. [CrossRef] [PubMed]
3. Sarkar, T.; Rawat, K.; Solanki, P.R.; Bohidar, H.B. Carbon dots-embedded fluorescent silica xerogel. Colloids Surf. A Physicochem. Eng. Asp. 2019, 583, 123844. [CrossRef]
4. Shi, X.; Meng, H.; Sun, Y.; Qu, L.; Lin, Y.; Li, Z.; Du, D. Far-Red to Near-Infrared Carbon Dots: Preparation and Applications in Biotechnology. Small 2019, 15, 1901507. [CrossRef]
5. Du, J.J.; Xu, N.; Fan, J.L.; Sun, W.; Peng, X.J. Carbon Dots for In Vivo Bioimaging and Theranostics. Small 2019, 15, 1805087. [CrossRef] [PubMed]
6. Bandi, R.; Devulapalli, N.P.; Dadigala, R.; Gangapuram, B.R.; Guttena, V. Facile Conversion of Toxic Cigarette Butts to N, S-Codoped Carbon Dots and Their Application in Fluorescent Film, Security Ink, Bioimaging, Sensing and Logic Gate Operation. ACS Omega 2018, 3, 13454–13466. [CrossRef] [PubMed]
7. Bakier, Y.M.; Ghali, M.; Zahra, W.K. Highly sensitive fluorescent detection of pyridine using small size carbon quantum dots derived from folic acid. J. Phys. D Appl. Phys. 2020, 53, 405103. [CrossRef]
8. Jin, C.C.; Wang, K.K.; Oppong-Gyebi, A.; Hu, J.N. Application of Nanotechnology in Cancer Diagnosis and Therapy-A Mini-Review. Int. J. Med. Sci. 2020, 17, 2964–2973. [CrossRef]
9. Li, H.W.; Mao, J.Y.; Lien, C.W.; Wang, C.K.; Lai, J.Y.; Mandal, R.P.; Chang, H.T.; Chang, L.; Ma, H.K.; Huang, C.C. Platinum ions mediate the interactions between DNA and carbon quantum dots: Diagnosis of MRSA infections. J. Mater. Chem. B 2020, 8, 3506–3512. [CrossRef]
10. Yi, Z.; Li, X.; Zhang, H.; Ji, X.; Sun, W.; Yu, Y.; Liu, Y.; Huang, J.; Sarshar, Z.; Sain, M. High quantum yield photoluminescent N-doped carbon dots for switch sensing and imaging. Talanta 2021, 222, 121663. [CrossRef]
11. Li, Y.R.; Zhu, H. Vitamin C for sepsis intervention: From redox biochemistry to clinical medicine. Mol. Cell. Biochem. 2021, 476, 4449–4460. [CrossRef] [PubMed]
12. Laustsen, A.; Van der Sluis, R.M.; Gris-Oliver, A.; Hernandez, S.S.; Cemalovic, E.; Tang, H.Q.; Pedersen, L.H.; Uldbjerg, N.; Jakobsen, M.R.; Bak, R.O. Ascorbic acid supports in vivo generation of plasmacytoid dendritic cells from circulating hematopoietic stem cells. eLife 2021, 10, e65528. [CrossRef]
13. Gao, M.; Zhao, Z.; Lv, P.; Li, Y.; Gao, J.; Zhang, M.; Zhao, B. Quantitative combination of natural anti-oxidants prevents metabolic syndrome by reducing oxidative stress. Redox Biol. 2016, 6, 206–217. [CrossRef]
14. O’Connell, P.J.; Gormally, C.; Pravda, M.; Guibault, G.G. Development of an amperometric L-ascorbic acid (Vitamin C) sensor based on electropolymerised aniline on palladium and food analysis. Anal. Chim. Acta 2001, 431, 239–247. [CrossRef]
15. Chen, Q.; Espey, M.G.; Sun, A.Y.; Pooput, C.; Kirk, K.L.; Krishna, M.C.; Khosh, D.B.; Drisko, D.J.; Levine, M. Pharmacologic doses of ascorbate act as a prooxidant and decrease growth of aggressive tumor xenografts in mice. Proc. Natl. Acad. Sci. USA 2008, 105, 11105–11109. [CrossRef] [PubMed]
16. Salkić, M.; Selimovic, A. Spectrophotometric determination of L-Ascorbic Acid in Pharmaceuticals Based on Its Oxidation by Potassium Peroxymonosulfate and Hydrogen Peroxide. Croat. Chem. Acta 2015, 88, 73–79. [CrossRef]
17. Tsao, C.S.; Salimi, S.L. Differential determination of L-ascorbic acid and D-isoascorbic acid by reversed-phase high-performance liquid chromatography with electrochemical detection. J. Chromatogr. A 1982, 245, 355–358. [CrossRef]
18. Chauhan, N.; Narang, J.; Pundir, C.S. Fabrication of multilayered carbon nanotubes/polyaniline modified Au electrode for ascorbic acid determination. Analyst 2011, 136, 1938–1945. [CrossRef] [PubMed]
19. Wu, Y.; Chen, Q.; Zhao, L.; Du, D.; Guo, N.; Ren, H.; Liu, W. Spectrofluorometric method for the determination of ascorbic acid in pharmaceutical preparation using L-tyrosine as fluorescence probe. Luminescence 2020, 35, 1092–1100. [CrossRef]
20. Zhou, X.; Xu, Q.; Wang, L.; Li, L.; Li, S.L.; Xia, K. Nitrogen dozen carbon quantum dots as one dual function sensing platform for electrochemical and fluorescent detecting ascorbic acid. J. Nanopart. Res. 2020, 22, 52–57. [CrossRef]
21. Xu, F.Z.; Tang, H.Y.; Yu, J.H.; Ge, J. A Cu2+-assisted fluorescence switch biosensor for detecting of coenzyme A employing nitrogen-doped carbon dots. Talanta 2021, 224, 121838. [CrossRef] [PubMed]
22. Qi, H.Y.; Sun, X.N.; Jing, T.; Li, J.L.; Li, J. Integration detection of mercury (II) and GSH with a fluorescent “on-off-on” switch sensor based on nitrogen, sulfur co-doped carbon dots. RSC Adv. 2012, 2, 1899–1997. [CrossRef]
23. Uriarte, D.; Vidal, E.; Canals, A.; Domini, C.E.; Garrido, M. Simple-to-use and portable device for free chlorine determination based on microwave-assisted synthesised carbon dots and smartphone images. Talanta 2021, 229, 122298. [CrossRef] [PubMed]
24. Qi, H.; Wu, X.; Zhang, H.; Tang, Y.; Gao, H.; Qian, M.; Qi, H. Synthesis of multiple-color emissive carbon dots towards white-light emission. Nanotechnology 2020, 31, 245001. [CrossRef]
25. Uriarte, D.; Domini, C.; Garrido, M. New carbon dots based on glycerol and urea and its application in the determination of tetracycline in urine samples. Talanta 2019, 201, 143–148. [CrossRef]
26. Liu, Y.; Zhou, Q.; Wu, Y.; Li, S.; Sun, Y.; Sheng, X.; Zhan, Y.; Zhao, J.; Guo, J.; Zhou, B. Sensitive detection of 2,4,6-trinitrotoluene utilizing fluorescent sensor from carbon dots and reusable magnetic core-shell nanomaterial. Talanta 2021, 233, 122498. [CrossRef]
27. Liu, Q.S.; Dong, Z.C.; Hao, A.J.; Guo, X.J.; Dong, W. Synthesis of highly fluorescent carbon dots as a dual-excitation rationmetric fluorescent probe for the fast detection of chlorogenic acid. Talanta 2021, 221, 121372. [CrossRef] [PubMed]
28. Chen, K.; Qiang, W.X.; Hu, W.P.; Lu, M.H.; Wang, Y.; Liu, X.H. On-off-on fluorescent carbon dots from waste tea: Their properties, antioxidant and selective detection of CrO42−, Fe3+, ascorbic acid and L-cysteine in real samples. Spectrochim. Acta A 2019, 213, 228–234. [CrossRef]
29. Song, N.N.; Wang, Y.Z.; Yang, X.Y.; Zong, H.L.; Chen, Y.X.; Ma, Z.; Chen, C.X. A novel electrochemical biosensor for the determination of dopamine and ascorbic acid based on graphene oxide/poly(aniline-co-thionine) nanocomposite. J. Electroanal. Chem. 2020, 873, 114352. [CrossRef]
30. Feng, S.N.; Yu, L.Y.; Yan, M.X.; Ye, J.; Huang, J.S.; Yang, X.R. Holey nitrogen-doped graphene aerogel for simultaneously electrochemical determination of ascorbic acid, dopamine and uric acid. Talanta 2021, 224, 121851. [CrossRef]
31. Zhu, S.; Zhang, J.; Tang, S.; Qiao, C.; Wang, L.; Wang, H.; Liu, X.; Li, B.; Li, Y.; Yu, W.; et al. Surface chemistry routes to modulate the photoluminescence of graphene quantum dots: From fluorescence mechanism to up-conversion bioimaging applications. *Adv. Funct. Mater.* 2012, 22, 4732–4740. [CrossRef]

32. Wang, L.; Zhu, S.J.; Wang, H.Y.; Qu, S.N.; Zhang, Y.L.; Zhang, J.H.; Chen, Q.D.; Xu, H.L.; Han, W.; Yang, B.; et al. Common origin of green luminescence in carbon nanodots and graphene quantum dots. *ACS Nano* 2014, 8, 2541–2547. [CrossRef]

33. Nie, H.; Li, M.; Li, Q.; Liang, S.; Tan, Y.; Sheng, L.; Shi, W.; Zhang, X.A. Carbon Dots with Continuously Tunable Full-Color Emission and Their Application in Ratiometric pH Sensing. *Chem. Mater.* 2014, 26, 3104–3112. [CrossRef]

34. Abd, A.L.I.; Qader, A.F.; Salih, M.I.; Aboul-Enein, H.Y. Sensitive spectrofluorometric method for the determination of ascorbic acid in pharmaceutical nutritional supplements using acriflavin as a fluorescence reagent. *Luminescence* 2019, 34, 168–174. [CrossRef]

35. Duzmen, S.; Baytak, A.K.; Aslanoglu, M. A novel voltammetric platform composed of poly (aminopyrazine), ZrO$_2$ and CNTs for rapid, sensitive and selective determination of ascorbic acid in pharmaceuticals and food samples. *Mater. Chem. Phys.* 2020, 252, 123170. [CrossRef]

36. Ribeiro, M.M.A.C.; Rocha, R.G.; Munoz, R.A.A.; Richter, E.M. A Batch Injection Analysis System With Square-wave Voltammetric Detection for Fast and Simultaneous Determination of Zinc and Ascorbic Acid. *Electroanal* 2020, 33, 90–96. [CrossRef]

37. Wu, S.; Sun, T.; Wang, H.; Fan, Z.; Li, L.; Fan, B.; Liu, L.; Ma, J.; Tong, Z. A sandwich-structured, layered CoTMPyP/5Sr$_2$Nb$_3$O$_{10}$ nanocomposite for simultaneous voltammetric determination of dopamine and ascorbic acid. *J. Electroanal. Chem.* 2020, 873, 114403. [CrossRef]

38. Meng, Y.; Jiao, Y.; Zhang, Y.; Li, Y.; Gao, Y.; Lu, W.; Liu, Y.; Shuang, S.; Dong, C. Multi-sensing function integrated nitrogen-doped fluorescent carbon dots as the platform toward multi-mode detection and bioimaging. *Talanta* 2020, 210, 120653. [CrossRef][PubMed]

39. Hu, G.; Ge, L.; Li, Y.; Mukhtar, M.; Shen, B.; Yang, D.; Li, J. Carbon dots derived from flax straw for highly sensitive and selective detections of cobalt, chromium, and ascorbic acid. *J. Colloid Interface Sci.* 2020, 579, 96–108. [CrossRef]

40. Wang, T.F.; Luo, H.; Jing, X.; Yang, J.L.; Huo, M.J.; Wang, Y. Synthesis of Fluorescent Carbon Dots and Their Application in Ascorbic Acid Detection. *Molecules* 2021, 26, 1246. [CrossRef]

41. Sonthanasamy, R.S.A.; Sulaiman, N.M.N.; Tan, L.L.; Lazim, A.M. Comprehensive spectroscopic studies of synergism between Gadong starch based carbon dots and bovine serum albumin. *Spectrochim. Acta A* 2019, 218, 85–96. [CrossRef][PubMed]

42. Alexandre, M.R.; Costa, A.I.; Berberan-Santos, M.N.; Prata, J.V. Finding Value in Wastewaters from the Cork Industry: Carbon Dots Synthesis and Fluorescence for Hemeprotein Detection. *Molecules* 2020, 25, 2320. [CrossRef][PubMed]