Human papillomavirus DNA as a factor determining the survival of bladder cancer patients

A Lopez-Beltran¹, AL Escudero¹, L Vicioso², E Muñoz³ and J Carlos Carrasco⁴

¹Department of Pathology, Cordoba University Medical School and Reina Sofia University Hospital, 14071 Cordoba, Spain; ²Department of Pathology, Malaga University Medical School, 29071 Malaga, Spain; Departments of ¹Immunology and ³Urology, Cordoba University Medical School and Reina Sofia University Hospital, 14071 Cordoba, Spain.

Summary The natural history of transitional cell carcinoma (TCC) of the urinary bladder is somewhat variable, with a significant number of tumour recurrences that occasionally evolve towards an infiltrating disease. The aim of this study was to investigate the presence of human papillomavirus (HPV) DNA in 76 TCC specimens, and then correlate such findings with the overall patient survival. However, other classical prognostic clinical and pathological variables such as pathological grade and stage, koilocytosis, age and sex were also tested. HPV DNA was investigated by means of the highly sensitive polymerase chain reaction (PCR). DNA primers specific for HPV types 6, 11, 16 and 18 were used. Our results showed that 7 (9.2%) out of 76 such cases were reactive for HPV 16 DNA; one of them also reacted with HPV 6 DNA. The statistical analysis was done by the Kaplan–Meier method, Wilcoxon's generalised test for studying the differences in survival curves and Cox's regression analysis for independent prognostic factors. A significant P-value was found for pathological grade (P<0.0001) and stage (P<0.0001), HPV 16 DNA (P = 0.0418) and koilocytosis (P = 0.0140). Thus, pathological grade was the only independent factor in the bladder cancer survival. These observations may prove useful in prognostic stratification of patients with TCC of the bladder.

Keywords: human papillomavirus; polymerase chain reaction; transitional cell tumour; urinary bladder

The presence of human papillomavirus (HPV) DNA has been reported most frequently in association with cervical dysplasias which can progress to malignancies, and benign condylomata acuminata (Stoler et al., 1992; Donalson et al., 1993). Recent studies indicate that some HPVs are associated with bladder carcinoma (Del Mistro et al., 1988; Kitamura et al., 1988; Querici Della Rovere et al., 1988; Bryant et al., 1991; Anwar et al., 1992; Chetsanga et al., 1992; Lopez-Beltran et al., 1992a; Furihata et al., 1993; Chang et al., 1994; Lopez-Beltran and Muñoz, 1995). However, the exact incidence of HPV DNA involved in TCC of the bladder remains controversial (Chang et al., 1994; Lopez-Beltran and Muñoz, 1995), since the reported incidence varies between 2.5% and 62% (Anwar et al., 1992; Chetsanga et al., 1992; Lopez-Beltran et al., 1992a; Lopez-Beltran and Muñoz, 1995). Negative results have been reported by Chang et al. (1994) and Ashfaq and Vuitch (1994). The prognostic implication of HPV infection in bladder cancer survival was suggested by Lopez-Beltran et al. (1992a) and Furihata et al. (1993), using non-isotopic DNA in situ hybridisation.

The aim of the present research was to investigate HPV incidence in 76 TCC specimens, using polymerase chain reaction (PCR) analysis, and then to correlate such findings with the overall patient survival. This study also included other classic prognosticators, such as the pathological grade and stage, patient age and sex, and koilocytosis. An attempt is also made to ascertain their possible prognostic implication in bladder cancer survival.

Materials and methods

The study group consisted of 76 unselected and consecutive cases of TCC of the urinary bladder received at Reina Sofia University Hospital (Cordoba, Spain). All 76 patients underwent transurethral resection (TUR). Their mean age was 66.57 ± 1.17. Fourteen patients were female. Selected tissue specimens from all biopsies, formalin-fixed paraffin-embedded, were analysed for pathological grade and stage. All cases were followed over 5 years. Koilocytosis in TCC was evaluated following the criteria proposed by Hartveit et al. (1992).

Sample preparation for PCR

The paraffin-embedded tissues for PCR analysis were cut into 5–10 μm thin sections. To prevent contamination from one paraffin block to another, the knife and the microtome specimen holder were carefully cleaned with xylene after each specimen had been processed. To extract DNA, each section was placed into Eppendorf tubes and the paraffin removed twice with xylene and washed once with 0.5 ml of 100% ethanol to remove the solvent. The samples were then dried and resuspended in 300 μl of digestion buffer (50 mm Tris pH 8.5; 1 mM EDTA; 0.5% Tween 20 and 200 μg ml⁻¹ of proteinase K), and incubated for 6 h at 55°C. After this incubation the samples were heated at 95°C for 8 min to inactivate the proteinase K. DNA was then extracted twice with phenol–chloroform and the aqueous phase precipitated with 95% ethanol at −20°C overnight. After centrifugation at 13 000 r.p.m. for 15 min the DNA pellet was dried and dissolved in 50 μl of distilled water. The DNA was stored at −20°C until use.

PCR analysis

PCR was performed mixing 10 μl of DNA with 90 μl of a solution containing 2 mM magnesium chloride, 50 mM potassium chloride, 10 mM Tris-HCl, pH 8.3, 0.01% gelatin, 200 μM dNTP, 2.5 U of Tag DNA polymerase and 2 μM of the following primers: for HPV type 6, the primers were HPV601 and HPV602, which amplify a region of 260 bp in the E5 gene. For HPV type 11 the primers were HPV114 and HPV115, which amplify a region of 350 bp in the L1 gene. For HPV type 16 and HPV type 18, the upstream primer was H1 which is common for both virus strains, and the downstream primers were H2 and H3 respectively, which amplify a 109 bp region from the open reading frame of the E6 gene (Table 1). The primers were synthesised using an Applied Biosystem 381A DNA syntheser (Foster City, CA, USA). The reaction was performed in an automated thermocycler.
Table I Sequence of synthetic oligonucleotide primers and complementary oligonucleotide probes used for the PCR in this study

| HPV type/gene | Primer or probe | Sequence (5' to 3') | Length (bp) of amplified products |
|--------------|----------------|---------------------|---------------------------------|
| HPV 6/E5     | Primer HPV-601  | TAGGGGACTTGCTGTC    | 260                             |
|              | Primer HPV-602  | GCCATACGCGGCATGTTG  |                                 |
|              | Probe HPV-603   | CATTACGCAGGGCCGCTG  |                                 |
| HPV 11/L1    | Primer HPV-114  | GAATACCTGCGCATGTTG  | 350                             |
|              | Primer HPV-115  | CGAGCAGCTCCTGCTCG   |                                 |
|              | Probe HPV-116   | GCCTCACCACATGTAACACTG |                             |
| HPV 16/E6    | Primer H1       | ATTAGTGATGATAGACATTA | 109                            |
|              | Primer H2       | GGCCTTATGACATTACA   |                                 |
|              | Probe H4        | ATGGAAACACATTAAACATGAC |                             |
| HPV 18/E6    | Primer H1       | ATTAGTGATGATACATTA  | 109                            |
|              | Primer H3       | GTTTCTGGGCCGAGGCA   |                                 |
|              | Primer H5       | ATGGAGACACATTGGAATACATAAACACTGTTGATA | |

Table II Selected variables representative of the 76 cases of TCC included in this study

| Factors          | Categories | No. deceased (%) | No. alive (%) | P-value | Overall no. (%) |
|------------------|------------|------------------|---------------|---------|-----------------|
| No. of patients  |            | 22 (100.0)       | 54 (100.0)    | 76 (100.0) |                 |
| Age a           | Mean±s.d.  | 66.84±2.45       | 66.88±1.34    | 66.57±1.17 |                 |
| Sex             |            |                  |               |         |                 |
| Male            | 18 (81.8)  | 44 (81.4)        | 62 (81.6)     |         |
| Female          | 4 (18.1)   | 10 (18.5)        | 14 (18.4)     |         |
| Grade           |            |                  |               |         |                 |
| I               | 14 (59.2)  | 4 (15.6)         | 184 (24.7)    | P=0.0001b |
| II              | 24 (92.3)  | 3 (11.1)         | 27 (35.8)     |         |
| III             | 16 (61.5)  | 5 (18.5)         | 21 (27.0)     |         |
| Stage           |            |                  |               |         |                 |
| O               | 8 (30.0)   | 14 (53.8)        | 22 (28.9)     | P=0.0001b |
| A               | 38 (70.3)  | 24 (44.4)        | 62 (81.6)     |         |
| B               | 8 (14.8)   | 16 (29.6)        | 24 (32.0)     |         |
| C               | 4 (18.1)   |                  | 14 (18.4)     |         |
| HPV 16          |            |                  |               |         |                 |
| +               | 7 (9.2)    | 12 (16.2)        | 19 (25.0)     | P=0.0001b |
| −               | 52 (69.2)  |                  | 69 (90.7)     |         |
| Koislocytosis   |            |                  |               |         |                 |
| +               | 12 (16.1)  | 4 (5.3)          | 16 (21.0)     | P=0.0146b |
| −               | 42 (55.4)  |                  | 60 (79.0)     |         |

*Age at diagnosis (years). bSignificant P-value. Independent prognostic factor using Cox regression analysis.

Statistical analysis

The statistical analysis was undertaken using the life test procedure. Univariate analysis of cancer-corrected 5 year survival (defined as death from or with bladder cancer) was done according to the Kaplan–Meier method (Kaplan and Meier, 1958). Differences between survival curves were estimated by the Wilcoxon test (Gehan, 1962). In addition, independent prognostic factors were sought by Cox regression analysis (Cox, 1972). A P-value below 0.05 was regarded as being statistically significant.

Results

The selected variables representative of the 76 TCCs included in this study are illustrated in Table II. The PCR analysis showed positive signals for the HPV type 16 DNA in 7 (9.21%) of 76 cases investigated. Likewise, one case showed reactivity for both HPV 16 DNA and HPV 6 DNA (Figure 1). Sixteen (21.0%) of 76 cases had koilocytosis, and one of these HPV 16 DNA. Most patients with TCC (71.4%) associated with HPV DNA were of high pathological grade/stage, and died of disease within 9 to 13 months (Table III).

Similarly, the chi-squares for the Wilcoxon test showed pathological grade and stage, the presence of HPV 16-DNA and koilocytosis, to be significantly related with survival in all 76 cases. However, pathological grade was found to be an independent prognostic factor in patient survival (Cox regression analysis).
Table III  Clinicopathological characteristics of the seven cases of TCC associated with HPV DNA

| Case no. | Age | Sex | Follow-up (months) | Grade | Stage | PCR 16 | Koilocytosis |
|---------|-----|-----|-------------------|-------|-------|--------|-------------|
| 14      | 64  | M   | D/13              | III   | B     |  +     | –           |
| 16      | 64  | M   | D/10              | III   | B     |  +     | –           |
| 17      | 85  | M   | D/10              | III   | A     |        |             |
| 22      | 60  | M   | D/9               | III   | B     |  +     | –           |
| 24      | 57  | F   | NED/60            | I     | O     |        | +           |
| 28      | 57  | F   | D/13              | III   | B     |        | –           |
| 30      | 51  | M   | NED/60            | I     | A     |        | –           |

* Reactive for both HPV DNA type 16 and HPV DNA type 6. D, death; NED, no evidence of disease; M, male; F, female.

Table IV  Studies on HPV DNA of several types investigated in human urinary bladder carcinoma

| Reference          | Method/HPV type DNA studied | Prevalence no. (%) | HPV type DNA detected |
|--------------------|-----------------------------|--------------------|-----------------------|
| Kitamura et al. (1988) | SBH/HPV 16                  | 1/10 (10.0)        | HPV 16                |
| Querci et al. (1988)  | SBH/HPV 11                   | 1/1 (100.0)        | HPV 11                |
| Bryant et al. (1991)  | ISH/HPV 6/11, HPV 16/18     | 12/76 (15.7)       | HPV 16/18             |
| Lopez-Beltran et al. (1992a) | ISH/HPV 6/11.              | 9/18 (50.0)        | HPV 16/18             |
| Chetsanga et al. (1992) | PCR/HPV 6/18, 31/33/35     | 1/4 (2.5)          | HPV 6                 |
| Anwar et al. (1992)   | PCR/HPV 6, 11, 16, 18, 33   | 39/48 (81.0)       | HPV 16, 18, 33        |
| Wilczynski et al. (1993) | ISH/HPV 16, HPV 16/18    | 28/90 (31.1)       | HPV 16, 18, 33        |
| Furihata et al. (1993) | PCR/HPV 16, 18, 33/35   | 28/53 (52.8)       | HPV 16                |
| Yu et al. (1993)       | PCR/HPV                      | 2/3 (6.7)          | HPV 18                |
| Salzein et al. (1993)  | PCR                         | 0/33 (0.0)         | HPV 6, 11, 16, 18, 33 |
| Chang et al. (1994)    | PCR/HPV 6, 11, 18, 31/33, 35, 39, 40, 45, 51-59 | 1/08 (0.8) | – |
| Ashfaq et al. (1994)   | ISH/HPV 6/11, 18, 31/33    | 0/8 (0.0)          | HPV 16/18             |
| Lopez-Beltran et al. (1995) | PCR/HPV 6/11, 16/18, 33/33 | 4/76 (5.29)       | HPV 16/18             |
| Current study          | PCR/HPV 6, 11, 16, 18      | 7/76 (9.2)         | HPV 6                 |

* Patient with mild immunodeficiency.  a Squamous cell carcinoma.  b Two of eight reported cases were squamous cell carcinoma. SBH, Southern blot hybridisation; ISH, non-isotopic DNA in situ hybridisation.

Discussion

Transitional cell carcinoma of the bladder is a heterogeneous group of neoplasms that typically present a variable biological potential including high risk of recurrence and frequent evolution towards an infiltrating disease with reduced survival rates (Lopez-Beltran et al., 1994). The prognosis of bladder cancer seems to be related to pathological factors such as tumour grade and stage, although the immunohistochemistry of cell and tumour markers as well as flow cytometric analysis of abnormalities in DNA content have also been considered prognostically significant (Lopez-Beltran et al., 1992b). The purpose of this paper was to determine whether or not the finding of HPV DNA in TCC has additional prognostic value in patient survival.

HPVs are known to infect man and although most of these proliferations are benign, some may become malignant, and this malignant transformation is related to HPV type (Howley, 1991). In the genitourinary tract, HPV types 6/11 are most commonly associated with genital condylomata acuminata (Del Mistro, 1988), whereas types 16 and 18 are associated with dysplasias and carcinomas (Chang, 1990). In TCC most HPV's were reported in a small number of patients with an immunodeficient status, (Kitamura, 1988; Querci Della Rovere, 1988). Although, recently a larger series of TCCs were screened, demonstrating a variable incidence of HPV DNA which ranged from 2.5% to 62% (Anwar et al., 1992; Chetsanga et al., 1992; Lopez-Beltran et al., 1992a; Furihata et al., 1993; Lopez-Beltran and Muñoz, 1995). Negative results were reported by Chang et al. (1994) and Ashfaq and Vuitich (1994) (Table IV). In addition, HPV 16/18 DNA detected by means of non-isotopic in situ hybridisation has been related with a poor survival (Lopez-Beltran et al., 1992a; Furihata et al., 1993). Our results found a 9.2% incidence of HPV 16 DNA in TCC. Such differences could be explained by methodological reasons (Ashfaq and Vuitich, 1994). In fact, type and time of fixation have been considered important parameters for preservation of DNA (Greer et al., 1991; Karlsten et al., 1994). However, the finding presented here of a significant relationship between the detection of HPV 16 DNA and reduced patient survival, using PCR analysis confirmed previous reports on poor survival of TCC cases presenting with high-risk HPV DNA (Lopez-Beltran et al., 1992a; Furihata et al., 1993) detected by using in situ hybridisation. Taken together these results could indicate an additional prognostic value of viral infection in bladder cancer, although pathological grade is the only independent parameter in the survival of bladder cancer as showed by our results. This is in agreement with the finding that most patients with TCC (71.4%) associated with HPV 16 DNA were of high grade. Such results are of interest since pathological grade remains an important prognostic parameter in survival of patients with TCC of the urinary bladder. Finally, we found koilocytosis to be significant in patient survival, which could be related to the increasing incidence of koilocytosis concomitant with increasing pathological grade.

Acknowledgement

This research was supported by Fondo de Investigaciones Sauritarias (FIS) Grant 94/0064-01-02.
References

ANWAR K, NAIKE H, NAKAKUKI K AND INUZUKA M. (1992). High frequency of human papillomavirus infection in carcinoma of the urinary bladder. Cancer, 70, 1967–1973.

ASHFAQ R AND VUITCH F. (1994). Human papillomavirus and carcinomas of the female urethra. J. Urol. Pathol., 2, 195–201.

BRYANT P, DAVIS P AND WISON D. (1991). Detection of human papillomavirus DNA in cancer of the urinary bladder by in situ hybridization. Br. J. Urol., 68, 49–52.

CHANG F. (1990). Role of papillomavirus. J. Clin. Pathol., 43, 269–276.

CHANG F, LIPONEN P, TERVERBAUTA A, SYRJÄNEN S AND SYRJÄNEN K. (1994). Transient cell carcinoma of the bladder: failure to demonstrate human papillomavirus deoxyribonucleic acid by in situ hybridization and polymeerase chain reaction. J. Urol., 152, 1429–1433.

CHETSANGA C, MALMSTRÖM PU, GYLLENSTEN U, MORENO-LOPEZ J, DINTER Z AND PETTERSON U. (1992). Low incidence of human papillomavirus type 16 DNA in bladder tumour detected by polymeerase chain reaction. Cancer, 69, 1208–1211.

COX DR. (1972). Regression models and life-tables. J.R. Stat. Soc., 34, 187–220.

DEL MISTRO A, KOSS LG, BRAUNSTEIN J, BENNETT B, SACCAMANO G AND SIMONS KM. (1988). Condyloma acuminata of the urinary bladder. Natural history, viral typing and DNA content. Am. J. Surg. Pathol., 12, 205–215.

DONALSON YK, ARENDS MJ, DUVALL E AND BIRD CC. (1993). PCR analysis of the upstream regulatory region of human papillomavirus genes in cervical intraepithelial neoplasia and cervical carcinoma. J. Clin. Pathol., 46, 1021–1023.

FURIHATA M, INOUE K, OHTSUKI Y, HASHIMOTO H, TERAO N AND FUJITA Y. (1994). High-risk human papillomavirus infections and overexpression of p53 protein as prognostic indicators in transitional cell carcinoma of the urinary bladder. Cancer Res., 53, 4823–4827.

GEHAN E. (1962). A generalized Wilcoxon test for comparing arbitrarily single-censored samples. Biometrika, 52, 203–217.

GREER CE, LUND JK AND MANOS M. (1991). PCR amplification from paraffin-embedded tissues: Recommendations on fixatives and PCR fragments and applications. J. Natl. Cancer Inst., 1, 46–48.

HARVEIT M, MEALE BO AND THUNOLD S. (1992). Koilocytosis in neoplasia of the urinary bladder. Br. J. Urol., 69, 46–48.

HOWLEY PM. (1991). Role of human papillomavirus in human cancer. Cancer Res., 51, 5019s–5022s.

KAPLAN EL AND MEIER P. (1958). Non-parametric estimation from incomplete observation. J. Am. Stat. Assoc., 53, 457–481.

KARLSN F, KALANTARI M, CHITEMERERE M, JOHANSSON B AND HARMAR B. (1994). Modifications of human and viral deoxyribonucleic acid by formaldehyde fixation. Lab. Invest., 71, 604–611.

KITAMURA T, YOGO Y, VEKI T, MURAKAMI S AND ASO Y. (1988). The presence of human papillomavirus type 16 genome in bladder carcinoma in situ of a patient with mild immunodeficiency. Cancer Res., 48, 7207–7211.

KWOK S AND HIGUCHI R. (1989). Avoiding false positives with PCR. Nature, 339, 237–238.

LOPEZ-BELTRAN A AND MUÑOZ E. (1995). Transitional cell carcinoma of the bladder: Low incidence of human papillomavirus DNA detected by the polymeerase chain reaction and in situ hybridization. Histopathology, 26, 565–571.

LOPEZ-BELTRAN A, CARRASCO JC, REYMundo C, MORALES-JIMENEZ C, TORO-ROJAS M AND SANTAMARIA-OSSORIO M. (1992a). Bladder cancer survival and human papillomavirus infection. Immunohistochemistry and in situ hybridization. In Oncogenes and Molecular Genetics of Urological Tumours, Olsson, CA (ed.) pp. 83–89. Churchill Livingstone: Edinburgh.

LOPEZ-BELTRAN A, CROGHAN GA, CROGHAN I, HUBEN RP, METLINC AND GAETA JF. (1992b). Prognostic factors in survival of bladder cancer. Cancer, 70, 799–807.

LOPEZ-BELTRAN A, CROGHAN GA, CROGHAN I, MATILLA A AND GAETA JF. (1994). Prognostic factors in bladder cancer. A pathologic, immunohistochemical, and DNA flow-cytometric study. Am. J. Clin. Pathol., 102, 109–114.

QUERCI DELLA ROVERE G, OLIVER R, MCMANUS DJ AND CAS- TRO JE. (1988). Development of bladder tumour containing HPV type 11 DNA after renal transplantation. Br. J. Urol., 62, 36–38.

SALZSTEIN DR, ORIHUELA E, KOUCREK JN, PAYNE DA, CHAN TS AND TYRING SK. (1993). Failure of the polymeerase chain reaction (PCR) to detect human papillomavirus (HPV) in transitional cell carcinoma of the bladder. Anticancer Res., 13, 423–425.

STOLER MH, RHODES CR, WITHBECK A, WOLINSKY SM, CHOW LT AND BROKER TR. (1992). Human papillomavirus type 16 and 18 gene expression in cervical neoplasia. Hum. Pathol., 23, 117–128.

WILCZYNSKI SP, OFT M, COOK N, LIAO SY AND IFTNER T. (1993). Human papillomavirus type 6 in squamous cell carcinoma of the bladder and cervix. Hum. Pathol., 24, 96–102.

YU ST, WU MM AND LI LM. (1993). Prevalence of human papillomavirus 16 and 18 in transitional cell carcinoma of the bladder. Chin. Med. J., 106, 494–496.