Short Communication:  
**Isolation and characterization of the endophytic bacteria, and their potential as maize diseases control**

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**Abstract.** Mugiastuti E, Suprayogi, Prihatininghsih N, Soesanto L. 2020. Short Communication: Isolation And Characterization Of The Endophytic Bacteria, And Their Potential As Maize Diseases Control. Biodiversitas 21: 1809-1815. Sheath blight and bacterial wilt are diseases that can reduce maize production. Biological control with the endophytic bacteria offers environmentally friendly control for these pathogens. The study aimed to isolate and characterize the endophytic bacteria morphologically and biochemically and to study their potential to control maize diseases, especially sheath blight and bacterial wilt causing pathogens. The study was conducted at the Plant Protection Laboratory, Faculty of Agriculture, Jenderal Soedirman University, from April to August 2019. The study consisted of four stages: isolation and characterization of endophytic bacteria, the antagonism test of the endophytic bacterial to *R solani*, the antagonism test of the endophytic bacteria to *Pantoea*, and the mechanical test of the endophytic bacteria as biological control agents and plant growth-promoting bacteria. Based on the research, four endophytic bacteria isolates have been successfully isolated, and characterized successfully and found have the potential to be developed as biopesticides to control maize disease, especially *R. solani* and *Pantoea* sp. *Bacillus* sp, endophytic from the root (*BK.A1; BK.A3; PP.A5*) and *Bacillus* sp. endophytic from the stem (*PPD.B2*) can suppress the growth of *R. solani* by more than 50%, have a strong antagonistic index against *Pantoea* sp (> 4), and can produce protease and lipase enzyme, and phosphate solubilization.

**Keywords:** Bacillus, fluorescent Pseudomonas, Pantoea, Rhizoctonia solani

**INTRODUCTION**

Maize is a strategic food commodity in the world. In Indonesia, the government seeks to achieve self-sufficiency in maize through increasing production of sustainable maize. However, these efforts have faced several obstacles; one of them is the presence of plant diseases such as sheath blight caused by *Rhizoctonia solani* Kuhn, and bacterial wilt caused by *Pantoea stewartii*. *R. solani* can infect up to the midrib of the cob (Djaenuddin et al. 2017), resulting in up to 100% decrease in the yield (Muis 2007). *Pantoea* sp. can attack all stages of the plant causing wilting and leaf blight, and is known as Stewart's wilt (Patak 2004; Ammar et al. 2014). The pathogens can cause 40-100% yield loss.

Over the past 3 decades, the concept of sustainable and environmentally friendly agriculture has been carried out by minimizing the use of chemicals, both synthetic fertilizers, and pesticides. In the management of pests and plant diseases, biological control is developed by applying biological agents including endophytic bacteria (Shanti and Vittal 2013). Many endophytic bacteria can pass the endodermic barrier across from the root cortex to the vascular system, and subsequently develop as endophytes in stems, leaves, tubers, and other organs (Compant et al. 2005). The use of endophytic bacteria as biological agents has an advantage compared to rhizosphere bacteria because endophytic bacteria live and survive in the plant tissue during plant development, thus protecting the plants.  

*Bacillus* sp. and fluorescent *Pseudomonas* are reported to be able to live as endophytes and are widely used as biological control agents for soil-borne and air-borne diseases. The endophytic bacteria could control plant diseases through several mechanisms including competition, hyperparasitism, producing microbial inhibiting compounds (antibiotics, lysis enzymes, other physical or chemical disorders), enhancing plant resistance, and promoting plant growth (Compant et al 2005, Pal and McSpadden-Gardener 2006; Rosenblueth and Martinez-Romero 2006).

Based on the mechanisms, the use of endophytic bacteria isolated from maize, both upland and lowland, suggested potentially alternative control for sheath blight (*R. solani*) and bacterial wilt (*Pantoea* sp). The research aimed to isolate and characterize morphologically and biochemically the endophytic bacteria as well as their potential to control pathogens that cause disease in maize especially *R. solani* and *Pantoea* sp.

**MATERIALS AND METHODS**

This research was conducted at the Laboratory of Plant Protection, Faculty of Agriculture, Jenderal Soedirman.
University, Purwokerto, Central Java, Indonesia, from April to August 2019

Isolation *Rhizoctonia solani*

*Rhizoctonia solani* was isolated from maize with sheath blight symptoms and there was sclerotium as a resistant structure from the pathogenic fungi in Banyumasan. *R. solani* isolation was carried out based on Al-Fadhal et al. 2019. Disease samples were cut 0.5 x 0.5 cm, then sterilized with NaOCl (1%) for 2 min, and rinsed with sterile water 3 times. Disease samples pieces were then dried using sterile filter papers, and transferred to Petri dishes containing PDA medium to obtain pure *R. solani* isolates.

Isolation *Pantoea* sp.

*Pantoea* sp. was isolated from diseased maize samples taken from the maize growing area in Banyumasan Regency according to Coplin et al. (2012); Aini et al. (2013) and Desi et al. (2014). Diseased leaves or stems were washed with running water, then dried with a tissue. Diseased samples were cut 1.5 x 5 cm, then sterilized with ethanol 70% and rinsed with sterile water 3 times. Furthermore, the sample was crushed with 5 ml of sterile distilled water using a sterile mortar. The bacterial suspension was streaked on nutrient agar and incubated 3-5 days. Bacterial colonies that exhibit the character of *Pantoea* sp. were yellow, shiny, slimy, flat or convex, then separated as pure cultures of *P. Stewartii* candidates. The culture was then tested by Gram Reaction (KOH test), Hugh-Leifson test, pigment production in YDC medium, oxidase test, hypersensitivity test, and pathogenicity test on maize.

Isolation and characterization of endophytic bacteria

Sampling for the isolation of endophytic bacteria was carried out in Banyumasan and Purbalingga, Central Java, Indonesia, with purposive stratified random sampling. Samples were taken from two areas of altitude, i.e., low-moderate lands (0-500 m above sea level), and highlands (> 500 m above sea level) (Nuryanto et al., 2014). In each district, 2 locations were selected for the low-medium lands, and 1 location for the highlands. Age of maize plants was 20-30 days after planting, when the number of endophytic microbial populations that can be cultured is in the highest population (Cavaglieri et al. 2009).

The endophytic bacteria were isolated from the roots and stems of healthy maize plants. Roots and stems were washed, sterilized with 70% ethanol (1 minute), 20% sodium hypochlorite (5 minutes) and Ringer's thiosulfate solution (5 minutes). Separately, the roots and stems of 10 g each were crushed with 90 ml PBS on a sterile mortar. Subsequently, samples were plated on NA and Kings B media (Cavaglieri et al. 2009). To isolate *Bacillus* sp., the suspension was heated for 10 minutes at 80 °C, before platting on NA. Bacterial isolates were further purified and characterized, such as morphological characteristics, gram properties, catalase tests, and hypersensitivity tests.

The antagonism test of endophytic bacterial to *Rhizoctonia solani*

The antagonism test of endophytic bacteria on *R. solani* was carried out using the dual culture method. The level of inhibition of antagonist is calculated using the formula (Abidin et al. 2015).

\[
I = \frac{C - T}{C} \times 100\%
\]

Where:
- \(I\): The level of inhibition of antagonist (%)
- \(C\): The radius of pathogen colonies opposite antagonist
- \(T\): The radius of the colony of pathogens towards antagonist

The antagonism test of endophytic bacterial to *Pantoea* sp.

Antagonism testing was carried out using the double-layer test method (Santiago et al. 2015). Endophytic bacteria to be tested were grown on the nutrient agar medium, incubated at 28 C for 48 hours. In the upside-down position, 1 ml of chloroform was added to the cup lid and left for 2 hours. Next, add 5 mL so that 0.6% water containing 0.5 mL of *Pantoea* sp. bacterial suspension. The culture was re-incubated for 24 hours, and there were clear zones around the antagonistic bacterial colony. The antibiotic activity was assessed based on the diameter of the clear zone compared to the diameter of the colony. Characterization of the type of antibiosis can be divided into bactericidal and bacteriostatic types based on Djamniko et al. (2007).

The mechanism test of endophytic bacteria as controlling agents biological and plant growth-promoting microbial

The testing mechanism of endophytic bacteria was carried out for bacteria that have the potential in testing the antagonism of the fungus *R. solani* and *Pantoea* sp.

Protease test

The activity of the ability of antagonistic bacteria to produce extracellular protease enzymes was tested using Skim Milk Agar (SMA) medium. Each bacterium to be tested was grown on the nutrient agar medium, incubated at 28 C for 24-48 hours. The presence of clear zones around the colony shows that positive bacteria produce protease enzymes (Abed et al. 2016). The protease activity index is assessed based on the diameter of the clear zone compared to the diameter of the colony.

\[
\text{Protease index} = \frac{(\text{clear zone diameter} - \text{colony diameter})}{\text{Colony diameter}}
\]

Lipase test

Detection of the ability of bacteria to produce the enzyme lipase was done by growing the antagonistic bacteria on a medium containing 1% Tween 80. The presence of lipase enzyme activity was demonstrated by milky white sediment around the bacterial colony, after incubating at 28 C for 4-7 days. The lipolytic index was measured using a formula Djeric et al. (2011).
Lipolytic index = \( \frac{\text{Milky white diameter-colony diameter}}{\text{Colon} \text{yan diameter}} \)

Phosphatase test

Detection of the ability of bacteria to produce the enzyme phosphatase was done by growing bacterial isolates on Pikovskaya medium. After incubating for 7 days at 28 C, the presence of a clear zone around the bacterial colony shows that the bacteria has the ability to produce the phosphatase enzyme to dissolve phosphates. The solubility index is measured using a formula (Farooq and Bano 2013)

\[
\text{Phosphatase Index} = \frac{\text{Clear zone diameter-Colony diameter}}{\text{Colon} \text{yan diameter}}
\]

RESULTS AND DISCUSSION

Isolation and characterization of endophytic bacteria

The results of the exploration, isolation, and characterization of endophytic bacteria obtained 23 isolates of endophytic bacteria, consisted of 9 isolates of the fluorescent Pseudomonas and 14 isolates of Bacillus sp. (Table 1). Fluorescent Pseudomonas colony on King’s B was round, with a flat edge, and yellowish-white, to greenish-yellow, gram-negative, rod-shaped, non-spore and fluorescent under ultraviolet light. Singh et al. 2017 reported fluorescent Pseudomonas showed light green, yellowish, creamy, circular, slimy, regular-irregular characteristics. Bacteria have short-long rod forms. The Fluorescent Pseudomonas isolates is gram-negative, which can form catalase, a positive oxidase, needed to grow aerobes. Bacillus sp. was observed with a spherical colony having cell rod-shaped, gram-positive, and endospores within cells (Table 1.). Slepecky and Hempill (2006); Amin et al. (2015) reported Bacillus sp. has the characteristics of a circular colony and punctiform (small round), variations in the entire margin and lobate, white dull, non-slimy, gram-positive, has endospores, flagellum and some are motile.

Based on its distribution, fluorescent Pseudomonas and Bacillus sp. found in all sampling locations, high or low-medium lands. This shows that fluorescent Pseudomonas and Bacillus sp. spread and can live in various altitudes, both high and low-medium land. Bacon and Hilton 2002; Ganeshan and Kumar 2005 reported P. fluorescens and Bacillus sp., are species of bacteria with a wide range of life and are very adaptive in various environments. Both types of bacteria are also found in the roots or corn stalks. Bacillus sp. and Pseudomonas sp. including a group of endophytic bacteria have a wide range of life and more isolated in maize (Ganeshan and Kumar 2005; Orole and Adejumo 2011; Costa et al. 2013)

Antagonism test between the endophytic bacteria against \( R. \) solani

Based on the results of in vitro tests (Table 2), 24 isolates of the endophytic bacteria were able to inhibit the growth of \( R. \) solani, with varying degrees of inhibition. The endophytic bacteria that have inhibition rates above 50%, i.e. fluorescent Pseudomonas BK.A1 (51%), Bacillus sp. B.K.A1 (55.39%), Bacillus sp BK.A3 (51.52%), PP.A5 (50.66%), and PPD.B2 (50.8%). The effect of the endophytic bacteria in inhibiting the growth of \( R. \) solani is inversely proportional to the dry weight mycelium. The greater the percentage of inhibition of endophytic bacteria to the growth of \( R. \) solani, the smaller the dry weight mycelium (Table 2). Endophytic bacteria can inhibit the growth of \( R. \) solani, which were shown by the inhibitory zone in the area bordering the bacterial streak (Figure 1.A).

The endophytic bacteria have anti-pathogenic properties and can produce antibiotic compounds. The ability of the endophytic bacteria to control plant pathogens occurs through the mechanism of antibiosis, competition, lysis, inducing resistance and producing growth substances. Bacteria capable of producing secondary metabolites that can inhibit growth or damage pathogens (Hastuti et al., 2014). These compounds, including alkaloids, phenols, flavonoids, glycosides, and phytoalexin (Soesanto et al., 2010). Fluorescent Pseudomonas can produce various types of antibiotics including phenazine-1-carboxylic acid, pyocyanin, pyrrolnitrin, and pyoluteorin, 2,4-diacetyl phloroglucinol (Phl). Phl is a phenolic metabolite with antibacterial and antifungal (Jain and Das 2016). Bacillus species can produce various kinds of volatile compounds and diffusible with strong inhibitory activity against plant pathogens (Lim et al., 2017).

Figure 1. Antagonism test between the endophytic bacteria against \( R. \) solani (A) and Pantoea sp. (B)
| Land          | Sampling location                         | Sample | Gram test | Catalase test | Oxidase test | Colony morphology | Colony pigment     | Fluorescence on KB medium | Cell morphology | Endospores       | Isolate                  |
|---------------|-------------------------------------------|--------|-----------|---------------|--------------|-------------------|-------------------|--------------------------|-----------------|-------------------|--------------------------|
| Highland      | Purbalingga, Pratin 7.13’33” S, 109.17’21” E, 1.190 m asl | Root   | -         | +            | +            | Round             | yellowish-white    | +                        | Medium rod      | -                 | fluorescent Pseudomonas PP.A1 |
|               |                                           | Root   | +         | +            | +            | Round             | White             | -                        | Small rod       | +                 | Bacillus sp. PP.A3    |
|               |                                           | Root   | +         | +            | +            | Round             | White             | -                        | Medium rod      | +                 | Bacillus sp. PP.A5    |
|               |                                           | Stem   | -         | +            | +            | Round             | yellowish-white    | +                        | Medium rod      | -                 | fluorescent Pseudomonas PP. B4 |
|               | Banyumas, Baturaden 7.19’1” S, 109.14’29” E, 520 m asl | Root   | -         | +            | +            | Round             | Greenish-yellow    | +                        | Medium rod      | -                 | fluorescent Pseudomonas BB.A2 |
|               |                                           | Stem   | +         | +            | +            | Round             | White             | -                        | Small rod       | +                 | Bacillus sp. BB. A3   |
|               |                                           | Stem   | +         | +            | +            | Round             | White             | -                        | Medium rod      | +                 | Bacillus sp. BB.B4    |
| Medium-Lowland| Banyumas, Sumbang 7.21’54” S, 109.17’33”E, 200 m asl | Root   | -         | +            | +            | Round             | yellowish-white    | +                        | Medium rod      | -                 | fluorescent Pseudomonas BS.A2 |
|               |                                           | Root   | +         | +            | +            | Round             | White             | -                        | Medium rod      | +                 | Bacillus sp. BS.A1    |
|               |                                           | Root   | +         | +            | +            | Round             | White             | -                        | Small rod       | +                 | Bacillus sp. BS.A3    |
|               |                                           | Stem   | -         | +            | +            | Round             | Greenish-yellow    | +                        | Small rod       | -                 | fluorescent Pseudomonas PB. A 4 |
|               | Purbalingga, Bojongsari 7.20’12” S, 109.20’22” E, 190 m asl | Stem   | +         | +            | +            | Round             | White             | -                        | Medium rod      | +                 | Bacillus sp. BS. B1   |
|               |                                           | Stem   | +         | +            | +            | Round             | White             | -                        | Medium rod      | +                 | Bacillus sp. BB. B3   |
|               | Purbalingga, Padamara 7.22’28” S, 109.13’24” E, 180 m asl | Stem   | -         | +            | +            | Round             | Greenish-yellow    | +                        | Small rod       | -                 | fluorescent Pseudomonas PPD A1 |
|               |                                           | Stem   | -         | +            | +            | Round             | yellowish white    | +                        | Medium rod      | -                 | fluorescent Pseudomonas PPD. B1 |
|               |                                           | Stem   | +         | +            | +            | Round             | yellowish-white    | +                        | Medium rod      | -                 | fluorescent Pseudomonas PPD. B5 |
|               | Banyumas, Kembaran 7.23’47” S, 109.17’9” E, 110 m asl | Root   | -         | +            | +            | Round             | yellowish-white    | +                        | Medium rod      | -                 | fluorescent Pseudomonas BK. A1 |
|               |                                           | Root   | +         | +            | +            | Round             | White             | -                        | Medium rod      | +                 | Bacillus sp. BK.A1    |
|               |                                           | Root   | +         | +            | +            | Round             | White             | -                        | Medium rod      | +                 | Bacillus sp. BK.A3    |
|               |                                           | Stem   | +         | +            | +            | Round             | White             | -                        | Medium rod      | +                 | Bacillus sp. BK.B3    |

Table 1. Isolation and characterization of endophytic bacteria.
Table 2. Inhibition of endophytic bacteria against *R. solani*.

| Isolate                        | Inhibition rate (%) | Dry weight mycelium |
|--------------------------------|---------------------|---------------------|
| Control                        | 0                   | 0.093               |
| Endophytic bacteria from the root |                     |                     |
| fluorescent *Pseudomonas* BB.A2 | 49.00               | 0.038               |
| fluorescent *Pseudomonas* BS.A 2 | 45.00               | 0.027               |
| fluorescent *Pseudomonas* BK.A1 | 51.00               | 0.017               |
| fluorescent *Pseudomonas* PPD.A1 | 10.33               | 0.059               |
| fluorescent *Pseudomonas* PP.B4 | 38.33               | 0.017               |
| fluorescent *Pseudomonas* PB.A4 | 18.00               | 0.037               |
| *Bacillus* sp.BB.A3            | 40.42               | 0.030               |
| *Bacillus* sp.BS.A1            | 48.73               | 0.016               |
| *Bacillus* sp. BS.A3           | 37.42               | 0.039               |
| *Bacillus* sp. B.K.A1          | 55.39               | 0.002               |
| *Bacillus* sp. B.K.A3          | 51.52               | 0.003               |
| *Bacillus* sp. PP.A3           | 46.65               | 0.019               |
| *Bacillus* sp.PP.A5            | 50.66               | 0.009               |
| Endophytic bacteria from the stem |                     |                     |
| fluorescent *Pseudomonas* PPD.B1 | 27.00               | 0.020               |
| fluorescent *Pseudomonas* PPD.B5 | 49.33               | 0.013               |
| fluorescent *Pseudomonas* PP.B4 | 65.67               | 0.004               |
| *Bacillus* sp. BB.B4           | 44.44               | 0.026               |
| *Bacillus* sp. BS.B1           | 49.74               | 0.012               |
| *Bacillus* sp. BK.B3           | 40.36               | 0.031               |
| *Bacillus* sp. PPD.B2          | 50.8                | 0.007               |
| *Bacillus* sp. PPD.B4          | 39.44               | 0.036               |
| *Bacillus* sp. PB.B1           | 37.29               | 0.047               |
| *Bacillus* sp. PB.B3           | 44.9                | 0.022               |

The endophytic bacterial antagonism test against *Pantoea* sp.

The results of antagonism between the endophytic bacteria and *Pantoea* sp. show varied results. The endophytic bacteria that can inhibit bacterial growth were indicated by the presence of clear zones around the endophytic bacterial colonies (Figure 1). From the nine isolate fluorescent, *Pseudomonas* were tested, only three isolates were able to inhibit the growth of the *Pantoea* sp., i.e. fluorescent *Pseudomonas* (Pf) BS.A2, Pf BK.A1 Pf PPD.B5. While the isolates Pf BB.A2, Pf PPD.A1, Pf PP.A1, Pf PPD.B1, and Pf PP.B4 are not able to inhibit the growth of pathogenic bacteria. Meanwhile, all isolate *Bacillus* sp. tested (thirteen isolates) were able to inhibit the growth of *Pantoea* sp. (Table 3).

The presence of clear zones around endophytic bacterial colonies showed the ability of endophytic bacteria to produce antibiotics to inhibit the growth of *Pantoea* sp. *P. fluorescens* P60 can produce antibiotics that inhibit the growth of pathogens (Soesanto 2011). *Pseudomonas fluorescens* is reported to produce phenazine-1-carboxylic acid (PCA) and other derivatives, 2,4 diacetyl phloroglucinol (Phl), pyrrolnitrin (PN), and pyoluteorin (Plt) (Heydari and Pessarakli 2010). Nasrun and Burhanudin (2016) mention that *P. fluorescens* produce secondary metabolites, i.e. antimicrobials, cyanide acid and 2,4 diacetyl phloroglucinol phenazine, pyrrolnitrin, pyoluteorin antibiotics.

Table 3. Inhibition of endophytic bacteria against *Pantoea* sp.

| Isolate                        | Antagonism | Antagonism index | Antagonism category* | Antagonism activity |
|--------------------------------|------------|------------------|----------------------|---------------------|
| Endophytic bacteria from the root |            |                  |                      |                     |
| fluorescent *Pseudomonas* BB.A2 | +          | 4.91             | Strong               | Bacteriostatic      |
| fluorescent *Pseudomonas* BS.A 2 | +          | 4.42             | Strong               | Bacteriostatic      |
| fluorescent *Pseudomonas* BK.A1 | +          | 0                | -                    | -                   |
| fluorescent *Pseudomonas* PPD.A1 | -          | 0                | -                    | -                   |
| fluorescent *Pseudomonas* PB.A4 | +          | 5.29             | Strong               | Bactericidal        |
| *Bacillus* sp.BB.A3            | +          | 8.17             | Strong               | Bacteriostatic      |
| *Bacillus* sp.BS.A1            | +          | 4.00             | Strong               | Bacteriostatic      |
| *Bacillus* sp. BS.A3           | +          | 5.07             | Strong               | Bactericidal        |
| *Bacillus* sp. B.K.A1          | +          | 4.01             | Strong               | Bacteriostatic      |
| *Bacillus* sp. B.K.A3          | +          | 4.91             | Strong               | Bacteriostatic      |
| *Bacillus* sp.PP.A3            | +          | 6.63             | Strong               | Bactericidal        |
| *Bacillus* sp.PP.A5            | +          | 6.56             | Strong               | Bactericidal        |
| Endophytic bacteria from the stem |            |                  |                      |                     |
| fluorescent *Pseudomonas* PPD.B1 | -          | 0                | -                    | -                   |
| fluorescent *Pseudomonas* PPD.B5 | +          | 5.86             | Strong               | Bactericidal        |
| fluorescent *Pseudomonas* PP.B4 | -          | 0                | -                    | -                   |
| *Bacillus* sp. BB.B4           | +          | 7.80             | Strong               | Bactericidal        |
| *Bacillus* sp. BS.B1           | +          | 6.22             | Strong               | Bacteriostatic      |
| *Bacillus* sp. BK.B3           | +          | 5.33             | Strong               | Bacteriostatic      |
| *Bacillus* sp. PPD.B2          | +          | 5.00             | Strong               | Bacteriostatic      |
| *Bacillus* sp. PPD.B4          | +          | 8.75             | Strong               | Bacteriostatic      |
| *Bacillus* sp. PB.B1           | +          | 1.67             | Weak                 | Bacteriostatic      |
| *Bacillus* sp. PB.B3           | +          | 5.67             | Strong               | Bactericidal        |

Note: *Based on Davis and Stout (1971)*
The level of bacteria's ability to inhibit growth can be shown by the large diameter of the clear zone. The results showed that the antagonism index ranged from 1.67-8.17. Based on this index, most endophytic bacteria have a strong antagonism (index of antagonism> 4) (Davis and Stout 1971). Furthermore, bacterial isolates that showed antagonistic activity were tested for types of antagonism based on Djatmiko (2007). Based on the type of antagonistic activity, ten isolates the endophytic bacteria were bacteriostatic and nine isolates the endophytic bacteria were bactericidal. Bacteriostatic activity, growth inhibition is temporary, it is shown that regrowth of bacteria after being transferred to a new medium, which is free from the influence of antagonistic bacteria. Bactericidal activity, inhibition is permanent. Bacteria were unable to grow even though they are transferred to new medium.

Test the mechanism of endophytic bacteria as biological control agents and plant growth-promoting microbes

The mechanism test was carried out on endophytic bacteria that have the potential to control the fungus R. solani and Pantoea sp., i.e. Bacillus sp. B.K.A1, Bacillus sp. B.K.A3, Bacillus sp. PP.A5, Bacillus sp. PPD.B2. The production of compounds related to biocontrol of pathogens and promotion of plant growth in bacterial isolates was evaluated by measuring the production of antimicrobial compounds and hydrolytic enzymes (amylases, lipases, proteases, and chitinases) and phosphate solubilization. The results of enzyme activity tests are as shown in Table 4. The four isolates Bacillus sp. tested were able to produce protease, lipase and phosphatase enzymes, with varied activity indexes. All isolates of Bacillus sp. those tested had a high index of protease and lipase enzymes (> 3) (Table 4, Figure 2). Protease and lipase enzymes, related to the ability of the endophytic bacteria as biological control agents. Based on the protease and lipase indexes, Bacillus sp. PP.A5 can produce the highest proteases and lipase enzymes compared to other isolates. The isolates that have high protein and fat hydrolysis enzymes have the potential as biological control agents because proteins and fats are constituents of pathogen cells (Mota et al 2016). Besides, the protease enzyme is thought to degrade antibiotics produced by fungal or bacterial pathogens. According to Anderson et al. (2014), the extracellular protease enzyme produced by P. fluorescens can inactivate antibiotic compounds produced by Pantoea agglomerans.

Bacillus sp. PPD.B2 has the highest phosphate solubility index. The phosphate solubilization is related to the ability of endophytic bacteria as a plant growth promoter, providing phosphates for plants. Microbes with high phosphate solubility activity are capable of producing and releasing metabolites such as organic acids that chelate cations that are bound to phosphate (especially calcium) and converting them into soluble forms. Solubilization of different forms of phosphate by microbes associated with plants, and increasing its availability for plants, will increase growth and production of the plant (Djuric et al., 2011).

In conclusion, based on research carried out, it has been successfully isolated, morphologically and biochemically characterized four the endophytic bacteria that have the potential to be developed as biopesticides to control maize disease, especially R. solani and Pantoea sp. They can suppress the growth of R. solani by more than 50%, have a

### Table 4. Test results of proteases, lipases and phosphate solubilization

| Isolate                          | Protease test Activity | Protease test Index | Lipase test Activity | Lipase test Index | Phosphate solubilization Activity | Phosphate solubilization Index |
|----------------------------------|------------------------|---------------------|----------------------|-------------------|-----------------------------------|-------------------------------|
| Endophytic bacteria from the root|                        |                     |                      |                   |                                   |                               |
| Bacillus sp. B.K.A1              | +                      | 3.75                | +                    | 3.23              | +                                 | 1.17                          |
| Bacillus sp. B.K.A3              | +                      | 3.20                | +                    | 3.73              | +                                 | 1.27                          |
| Bacillus sp. PP.A5               | +                      | 5.00                | +                    | 4.40              | +                                 | 1.46                          |
| Endophytic bacteria from the stem|                        |                     |                      |                   |                                   |                               |
| Bacillus sp.PPD.B2               | +                      | 3.00                | +                    | 3.90              | +                                 | 2.60                          |

![Figure 2. Hydrolysis enzyme activity, (A) protease, (B) lipase and (C) phosphate solubilization](image-url)
strong antagonistic index against *Pantoea* sp (> 4), and can produce protease and lipase enzymes, and phosphate solubilization.

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