Regional grafting of autologous adipose tissue is effective in inducing prompt healing of indolent digital ulcers in patients with systemic sclerosis: results of a monocentric randomized controlled study

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Abstract

Background: A randomized controlled trial (RCT) was performed to confirm preliminary uncontrolled data indicating that regional adipose tissue (AT) grafting (G) is effective in inducing ischemic digital ulcer (IDU) healing in patients with systemic sclerosis (SSc).

Patients and methods: SSc patients with IDUs were randomized to be blindly treated with AT-G or a sham procedure (SP). AT-G consisted of injection, at the base of the finger with the IDU, of 0.5–1 ml AT after centrifugation of fat aspirate. The SP consisted of false liposuction and local injection of saline solution. The primary endpoint was to compare the cumulative prevalence of healed IDUs in the two groups within the following 8 weeks.

Results: AT-G and the SP were carried out in 25 and 13 patients, respectively. The two groups were comparable for age, gender, disease duration, and SSc subtypes. IDU healing was observed in 23/25 and 1/13 patients treated with AT-G and the SP, respectively (p < 0.0001). The 12 patients who received the unsuccessful SP underwent a rescue AT-G. In all of them, IDU healing was observed after 8 weeks of observation. It was noticeable that in the AT-G-treated patients a significant reduction of pain intensity (measured by visual analogue scale) was recorded after 4 and 8 weeks (p < 0.0001 in all cases). Similarly, a significant increase of capillary numbers in the affected finger was recorded by nailfold videocapillaroscopy after 4 and 8 weeks (p < 0.0001 in both cases).

Conclusion: This RCT strongly confirms that AT-G is effective in inducing IDU healing in SSc patients.

Trial registration: ClinicalTrials.gov, NCT03406988. Registered retrospectively on 25 January 2018.

Keywords: Systemic sclerosis, Digital ulcers, Autologous fat grafting, Adipose tissue stem cells

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**Introduction**

Systemic sclerosis (SSc) is an autoimmune disease characterized by a multifactorial pathological process where a central role is played by the progressive loss of the microvascular bed, with the consequent fibrotic changes in the involved organs and tissues [1]. The clinical counterpart of the impaired microcirculation in the fingers is represented by Raynaud’s phenomenon, which is present in the great majority of the patients and is often the initial manifestation of the disease [2].

The progressive derangement of the microvascular architecture can be easily observed in finger periungual areas by means of nailfold videocapillaroscopy (NVC). This simple and low-cost technique enables careful observation of the evolutionary steps of this pathological process. These steps are capillary network rarefaction, dilated or giant capillary formation, and, finally, almost complete vascular desertification [3]. The most advanced stages of capillary loss may induce the formation of digital ulcers (DUs) on the fingertips. This event, which has been observed in around half of the patients [2, 4, 5], commonly leads to a significant deterioration of the patient’s quality of life because of the often-severe disabling pain and the difficulty in performing the simplest daily living activities. The healing of DUs is often a lengthy process requiring accurate and intensive topical and systemic treatment [4]. Nevertheless, in a significant number of cases this therapeutic approach is ineffective and distal necrosis with subsequent tissue loss or phalangeal amputation may eventually occur.

Since it has been recognized that bone marrow mesenchymal stem cells (BM-MSCs) possess multiple regenerative and angiogenic properties, therapy with these cells of autologous source has been tested with regenerative and angiogenetic properties, therapy with BM-MSCs. In addition, ASCs are regarded as a more favorable source since these cells can be easily and less invasively isolated, and have shown a great rapidity of expansion [10]. Preliminary attempts at cell therapy with ASCs have been carried out with the purpose of inducing ulcer healing in peripheral vascular impairment that can be observed in some animal models and human conditions [11]. In a recent open pilot study performed by our group, it has been demonstrated that grafting (G) with autologous adipose tissue (AT), which is known to contain both ASCs and a stromal/vascular fraction (SVF) [12], was effective in inducing prompt healing of long-lasting IDUs localized in the fingertips of a small number of patients with SSc [13]. The IDU healing was accompanied by the rapid disappearance of local ischemic pain and evidence of a partial restoration of the capillary bed in the digits when assessed by NVC.

Starting from these encouraging data, and with the purpose of confirming these preliminary results, we have designed and performed a monocentric randomized controlled study. In accordance with the study protocol, patients with a typical SSc-related IDU on the fingertip were randomized to undergo a regional AT-G with autologous fat as active therapy or a ‘sham’ procedure (SP)—that simulates the active treatment—as placebo treatment. All of the patients with SSc enrolled in both arms were blind regarding the treatment received. Furthermore, during the study period all of the enrolled patients received the same systemic vasoactive and topical therapy.

**Patients and methods**

The study design is schematically represented in Fig. 1. This study is registered with ClinicalTrials.gov (NCT03406988).

**Patients**

The study was conducted from July 2017 to April 2018. Patients who were candidates for enrollment in the study were required to be over 18 years old and to meet the 2013 classification criteria of the American College of Rheumatology/European League Against Rheumatism for SSc [14]. Patients with either the limited cutaneous (lc) or the diffuse cutaneous (dc) variants of SSc were considered eligible for the study. Furthermore, all of the candidate patients had to have only one active IDU (cardinal ulcer), lasting for at least 6 weeks prior to enrolment and showing no tendency to heal despite weekly intravenous iloprost (0.5–2 ng/kg/min), oral administration of calcium-channel blockers (nifedipine 20 mg daily), local medication with meticulous attention directed toward wound care, including antimicrobial agents, keeping the DUs clean, and using a suitable dressing, and surgical removal of necrotic tissue. Previous clinical history of similar DUs did not preclude the eligibility of patients.

An IDU was defined as a painful area, at least 6 mm in diameter at its longest point, with visible depth and loss of dermis, located at the volar surface of the digit, distal to the proximal interphalangeal digital crease in a site judged compatible with a vascular etiology [15]. We excluded digital ulcers located over cutaneous calcifications or over extensor surfaces of small joints since they are generally believed to be mechanical, as a result of recurrent
microtrauma and increased skin tension—although there is emerging evidence suggesting that all DUs in SSc could be the expression of the common vascular disease [16].

Exclusion criteria from the study were the presence of severe extracutaneous manifestations, such as cardiac, lung, and renal involvement, current therapy with dual and selective endothelin inhibitors, and concomitant treatment with immune-suppressive therapies (including prednisone equivalent > 10 mg). It is known that immunosuppressor agents have a cytostatic effect which could prevent regenerative processes and promote infections complicating DUs. Furthermore, there is insufficient evidence at present to recommend immunosuppressive therapy for DUs [17].

No patient included in the trial was in current therapy with phosphodiesterase type-5 inhibitors that are not authorized in Italy for the treatment of DUs.

Patients suffering from diabetes and/or other vascular diseases and women currently pregnant or breastfeeding were also excluded from the study recruitment.

**Active therapy**

Active therapy consisted of the implantation of a small amount of autologous AT at the base of the finger where the IDU was distally located, following the procedure that was adopted in a previous open study [13]. Briefly, under local anesthesia, a total amount of 0.5–1 ml of the autologous AT was injected at the base of the affected finger, by sequential introduction of small aliquots in different directions from the injection site. This was done to spread the grafted AT as widely as possible around the finger base. “Luer lock” syringes (Artsana SpA, Grandate, Como, Italy) and Coleman 18G cannulas (Mentor-Aesthetic, Santa Barbara, CA, USA) were used for the procedure. At the moment of needle removal, a small skin incision was made at the site of injection to slightly reduce the tissue pressure due to the insertion of the additional material.

Autologous AT was previously harvested in each patient through liposuction from abdominal fat in the surgery room, under local anesthesia. Afterward, the collected AT was transferred into sterilized vials and processed at 920 x g for 3 min. The upper and lower phases, containing oil supernatant and mature adipocytes and blood and plasma residuals, respectively, were discharged. Only the intermediate layer was used for the filling procedure [13].

**Sham procedure**

To maintain the blindness of the study, a SP was administered to the patients enrolled in the placebo group to simulate the AT-G. Briefly, a false liposuction of the abdominal adipose tissue was performed followed by the injection of 0.5–1 ml of 0.9% saline solution at the base of the affected finger. The SP was carried out exactly in...
the same surgical room and following the same steps as for the active therapy.

**Basal therapy**
Weekly iloprost infusions and calcium-channel blockers that were administered to the patients before inclusion in the study were continued during the entire observation time for all of the patients enrolled in both arms of the study. The administration of analgesics to alleviate the IDU-related pain was also allowed for each patient. The amount of analgesics administered was obviously drastically reduced and stopped when pain disappeared as a consequence of IDU healing.

**Outcome parameters and follow-up of the patients**
The primary endpoint of the study was to compare the prevalence of patients in whom IDU healing was observed within 8 weeks after the AT-G and to compare this figure with that observed in the control arm of the study where the SP was performed. To verify the time of IDU healing, all of the patients treated with either AT-G or the SP were observed every week. Local medication and necrotic tissue debridement were also performed during this weekly observation when required.

Secondary endpoints to be assessed were: pain improvement (i.e., a reduction of more than 50% of the baseline pain VAS score after AT-G and the SP); and variation of the number of capillaries in the affected digits in patients who received AT-G or the SP.

To this end, a patient's self-assessment of pain severity was performed by means of a visual analogue scale (VAS) (range 0–100 mm, 100 mm indicating the most severe pain). This was done immediately before and every week after carrying out AT-G and the SP in all patients included in the study. Moreover, an NVC examination (with a 200× magnification lens, using Videocap; Scalar Co. Ltd, DS MediGroup, Milan, Italy) was performed in every affected finger immediately before and after 4 and 8 weeks for every patient enrolled and randomized to receive both types of procedure. NVC images from each patient were taken from four consecutive fields, two fields in both the right and left directions, starting from the middle of the nailfold, for a total extension of 1.2 mm and digitally stored. Although all studied patients exhibited a capillaroscopic late pattern according to Cutolo et al.'s classification [3], we decided to evaluate only the count of capillaries, since it was easier to precisely quantify this parameter than the number and extension of avascular areas. A single experienced operator, who was blinded regarding the type of procedure performed in the examined patient, counted the cumulative number of observed capillaries before and 4 and 8 weeks after both AT-G and the SP.

**Rescue AT-G therapy**
The follow-up period was extended beyond the 8 weeks when the primary and secondary endpoints were assessed up to a total of 20 weeks. It was preliminarily established that all of the patients in the control arm, in whom IDU healing was not observed after 10 weeks, had to undergo a rescue AT-G. The possibility that IDU healing could be achieved after this rescue therapy was verified by monitoring these patients every week for the following 10 weeks.

**Calculation of the patients’ samples and randomization procedure**
Starting from the results of our open study that showed 100% of IDU healing within 10 weeks in the patients treated with AT-G, and from some literature data demonstrating that, in patients suffering from distal IDUs and treated with standard vasoactive therapy, ulcer healing was observed in around 15% after 9 weeks [18], we have considered it plausible to achieve IDU healing after 8 weeks of observation in at least 80% of patients treated with AT-G and in no more than 30% of the control patients receiving the SP. To verify that the null hypothesis (i.e., the two treatments are equally effective in inducing IDU healing) could be true with a probability lower than 1%, and having preliminarily established for ethical reasons that the ratio between treated patients and control patients should be 2:1, we calculated (using Fisher’s exact test) that we had to enroll 30 patients to be treated with AT-G and 15 patients to be included in the control group.

Patients meeting the inclusion and exclusion criteria were consecutively enrolled in the study and randomized to receive AT-G or the SP following a sequence of randomization that consisted of a consecutive and repeated inclusion of one patient in the control group followed by two patients in the actively treated group until the achievement of the pre-established numbers of patients in the two groups had been reached.

**Statistical analysis**
Demographic and clinical parameters of the patients selected in the two arms of the study were compared using an unpaired t test and contingency tables for continuous and categorical variables, respectively.

Data collected before and after both therapeutic procedures were analyzed using paired t tests and Wilcoxon tests for continuous and discrete variables, respectively.

When multiple tests were performed, we corrected the level of significance for the number of tests, according to Bonferroni's method.

A Kaplan–Meier curve was built and the log-rank test was used to analyze the cumulative prevalence of healed
IDUs in the two arms of the study in each observation time during the follow-up.

Ethical rules
This study was conducted according to the Helsinki Declaration and approved by the local ethics committee. Written informed consent was obtained from all of the enrolled patients.

Results
A preplanned interim analysis of the results was performed after the enrolment of 25 and 13 patients who received AT-G and the SP, respectively. The analysis demonstrated that the primary endpoint of the study was already reached (i.e., the probability that the two therapeutic procedures had no difference in efficacy for inducing IDU healing was < 1%). Considering this figure, we decided to stop the enrolment of further patients into the study.

Table 1 summarizes the main demographic and clinical characteristics of the patients included in the study. The groups of patients randomized to undergo AT-G and the SP were comparable for age, disease duration, dcSSc/lcSSc ratio, and extension of the IDU area at the time of enrollment. Patients who received active therapy had IDUs lasting for a significantly longer time than the patients from the control arm (p < 0.01).

Twenty-three out of the 25 patients (92.0%) treated with AT-G achieved DU healing after 8 weeks. Conversely, in only 1 of the 13 patients (7.7%) who received the SP was IDU healing observed at the end of the same observation period. This difference was highly significant (p < 0.0001). The Kaplan–Meier curve representing the cumulative prevalence of IDU healing in the two differently treated groups is shown in Fig. 2. Figure 3 shows digital ulcer progressive healing of patient 2 (Fig. 3a) and patient 8 (Fig. 3b) after autologous AT-G.

The result was maintained during the following 3-month period and no new DU appeared in any of the AT-G patients, both in the treated finger and the other fingers of the same hand. This feature might have been fostered by the consequent secondary prevention with bosentan.

At the 4-week observation time, a reduction of the pain VAS of more than 50% of the baseline value was recorded in 21 out of the 25 patients treated with AT-G and in 0 out of the 13 patients of the SP group (p < 0.0001 using Fisher’s exact test). All four patients in the AT-G group in whom this level of pain improvement was not observed at the 4-week observation time reached this goal after 8 weeks. Conversely, the number of patients in the SP group with this grade of pain score reduction remained at zero even at the 8-week observation time. The variation of pain VAS within the two groups is shown in Fig. 4a, b.

In the actively treated patients, the number of capillaries in the affected digit consistently increased after 4 and 8 weeks compared to the starting time and at the 8th week in comparison with 4-week counts (p < 0.0001 in all comparisons). Conversely, capillary counts remained unchanged in patients who underwent the SP (see Fig. 5a, b). Despite the significant capillary increase in the AT-G-treated group, patients maintained the late capillary pattern, according to Cutolo et al. [3].

It is worth noting that in all of the 12 patients who initially underwent the SP and in whom this procedure was ineffective at the end of the 10th week, IDU healing was observed in the following 8 weeks after the rescue AT-G. It is also noticeable that the two patients initially treated with AT-G and in whom the IDU healing was not achieved at the 8-week observation time had a complete resolution of their IDU at the end of the 10th week (see Fig. 2).

Discussion
Fingertip IDUs are a frequent complication in patients with both lc-SSc and dc-SSc, and are considered a clear consequence of distal capillary bed desertification that can be observed in the late phase of the disorder [2–5]. The SSc-related ischemic DUs are often painful, invalidating, and may lead to a severe deterioration in quality of life. Their trend to healing is frequently very slow or null, despite the multiple local and general therapeutic measures that are commonly adopted [4].

| Table 1 Demographic and clinical features in patients with SSc treated with AT-G and SP |
|--------------------------------------|----------|----------------|
| Feature                             | AT-G (25 patients) | SP (13 patients) | Difference |
| Age (years), median (range)          | 42 (21–69)        | 37 (23–70)       | n.s.       |
| Women, n (%)                        | 23 (92)           | 13 (100)         | n.s.       |
| Disease duration (years), median (range) | 4 (1–10)  | 5 (2–12)         | n.s.       |
| Patients with dcSSc/lcSSc (n)       | 16/9              | 7/6              | n.s.       |
| IDU localization (2nd/3rd/4th/5th finger) | 10/12/2/1 | 5/4/4/0         | n.s.       |
| IDU longest diameter, mean (95% CI) | 7.5 (6.7–8.2)    | 7.3 (6.2–8.4)    | n.s.       |
| IDU duration before enrolment (weeks), mean (95% CI) | 11.4 (9.7–13.1) | 8.0 (7.1–8.8) | p < 0.001 |
| Patients who have had previous ulcers, n (%) | 15 (60) | 7 (54) | n.s.       |

AT-G adipose tissue grafting, CI confidence interval, dcSSc diffuse cutaneous systemic sclerosis, IDU ischemic digital ulcer, lcSSc limited cutaneous systemic sclerosis, n.s. not significant, SP sham procedure
This paper clearly shows that AT-G is significantly more effective than traditional vasoactive therapy alone in inducing IDU healing, restoration of the capillary bed in the treated finger, and a rapid resolution of local ischemic pain in patients with SSc. The fact that these clinical effects can be ascribed to specific actions of AT-G, and not to mechanical stimulation due to local overload of the injected material, is supported by the almost complete ineffectiveness of the SP that was blindly carried out in the control group. Furthermore, the IDUs of the patients who initially had received the ineffective SP healed within the following 8 weeks once treated with a rescue AT-G.

The results of the present RCT confirm the data we obtained in our previous open study where the same AT-G induced, after 2 months and in all of the patients, IDU healing, a significant increase in the number of fingertip capillaries, and a rapid resolution of ischemic pain [13]. Although it is likely that the local increase of the microvascular bed may be an important cause of pain reduction in the AT-G-treated patients in our study, the rapidity of this pain improvement could rather
suggest the intervention of other possible factors. It has been shown in experimental models that the ASC secretome may exert either downregulation of the inflammatory cytokines or upregulation of neural trophic factors able to modulate neuroinflammation and the related pain [19–21].

Therapeutic procedures using various cellular components that derive from AT, and potentially containing totipotent stem cells, are an emerging new therapeutic approach for different disease-related features in patients with SSc [9].

Granel et al. [22] have demonstrated that local lipofilling with the separated SVF is capable of improving Raynaud’s phenomenon, local vascular flow, and skin elasticity in the hands. In another study by our group, we observed that lipofilling with autologous AT in the perioral area improves the mouth opening capacity and at the same time induces a histologically and NVC-proven new capillary formation in the local skin area [23].

In contrast, few studies, limited to single case reports, have been performed in which SSc-related ischemic

![Fig. 4](image-url) Variations of pain intensity measured by VAS in patients treated with AT-G as active therapy (a) and in those who underwent SP as placebo treatment (b). VAS for pain measured every week in all study patients. (a) Comparisons between baseline VAS values and those recorded at 4 and 8 weeks of follow-up, and between values at 4 and 8 weeks, in patients treated with AT-G are all highly significant. (b) VAS values remained unchanged in patients who underwent SP. VAS visual analog scale, w weeks.
lesions in fingers or limbs were treated with regional infusion of purified BM-MSCs. Even if limited in number, these experimental cell therapies have uniformly shown an improvement of ischemic lesions [7–9].

Positive results obtained with cellular therapeutic procedures with different AT fractions in SSc-related vascular lesions are not completely surprising, since similar results have been reported in other ischemic conditions in both human disorders and in animal models treated with similar therapeutic modalities. A certain degree of wound improvement and healing has been observed after regional injection of isolated ASCs in the ischemic leg in a small group of patients with lower limb ulcers that were not suitable for revascularization [24, 25]. Similarly, an acceleration of excisional wound healing by locally administered ASCs has also been demonstrated in different animal models [26, 27]. Furthermore, foot ulcers have been reported to heal more frequently and rapidly, with respect to traditionally treated controls, in diabetic patients treated locally with differentially processed AT cellular fractions [24, 28].

Fig. 5 Variations in number of capillaries assessed by NVC in fingers of patients with cardinal IDU treated with AT-G as active therapy (a) and who underwent SP as placebo treatment (b). (a) Comparisons between baseline counts with those recorded at 4 and 8 weeks, and between counts recorded at 4 and 8 weeks, of follow-up in AT-G-treated patients are all highly significant. (b) Capillary counts remained unchanged in patients who underwent SP. w weeks
The reason why local implantation of AT containing adipocyte progenitors, ASCs and SVF, as well as similar procedures with BM-MSCs, may induce such positive changes in different SSc-related ischemic lesions remains almost completely speculative at the moment. A number of studies have shown that ASCs, as well as BM-MSCs, are multipotent cells potentially able to differentiate themselves into other cell types, like cells of the mesoderm (chondrogenic, osteogenic, myogenic) and endoderm (endothelial) lineages, under the conditioning of different stimulant, as demonstrated in a lot of in-vitro and in-vivo studies [12]. ASCs are also capable, when locally implanted, of producing a number of growth factors with proangiogenic and proliferation effects, such as vascular endothelial growth factor and fibroblast growth factor, that may favor local angiogenesis by exerting their paracrine actions [29]. ASCs may also directly produce or stimulate the production from other resident cell types of a number of cytokines with immune-modulating action probably capable of downregulating some pathological mechanisms in injured tissues [29]. On the other hand, damaged tissues may release various chemoattractants, like epidermal growth factor and monocyte chemoattractant protein-1, that are able to induce local migration and concentration of ASCs [12, 29] by interacting with specific receptors present on the surface of these latter cells [30]. Finally, both BM-MSCs and ASCs have shown, either in vivo or in vitro, that they better express their angiogenetic properties when exposed to hypoxic conditions [31, 32].

Since SSc is a systemic microvascular disorder where many tissue and organ systems may suffer from a reduction of capillary flow and the consequent fibrotic evolution, it could be postulated that both BM and AT may be involved in this process and the locally resident cells may be defective. This possibility could make use of stem cells of autologous origin less effective with respect to corresponding cells from a heterologous donor, when used as regenerative therapeutic elements in the treatment of SSc. Some in-vitro data suggest that isolated BM-MSCs have a senescent aspect and may be defective in their immunosuppressive properties [33]. Conversely, studies assessing the functional capacity of ASCs from SSc patients in comparison to those of healthy controls have shown that the proliferative and proangiogenic ability of these cells is not compromised, probably being preserved in the AT niche from any SSc-related pathological aggression [34].

The SVF of the medium layer of AT separated by centrifugation contains a heterogeneous population of many cells other than ASCs, including adipocyte precursors, endothelial cells and their progenitors, pericytes, hematopoietic lineage cells, and fibroblasts. At the present time, little information is available on the possible contribution given by the SVF in the regenerative effects that whole AT has shown under many different conditions [35]. Recent studies have demonstrated that pericytes possibly provide a very important contribution to tissue repair and revascularization, since these cells may work as driving stromal cells in tubuli formation during angiogenesis [36]. Therefore, it cannot be excluded that SVF, where pericytes are certainly present, might play an important role also in the healing process of IDUs in our patients. Such an assumption is further reinforced by recent data showing the higher angiogenetic potential of freshly isolated AT where the SVF is largely represented, when compared to cultured AT cells where the SVF is reduced [12]. The multiple lineage differentiation potential of ASCs, the possible support to angiogenesis given by the SVF, and the local release of a number of growth factors and cytokines, able to stimulate the angiogenesis, may all have contributed to the formation of new capillary loops, and to the rapid IDU healing observed in our study.

The present study confirms previous more preliminary and uncontrolled data suggesting that local transfer of autologous AT may be a successful option to induce healing in ischemic SSc-related fingertip IDUs that are resistant to more traditional therapeutic approaches. The AT-G procedure we have adopted in this study did not cause any adverse event and so it has been proven to be a safe treatment. Since this procedure is fairly invasive, at the present time it should be considered a second-line treatment to be carried out only in cases resistant to traditional therapy. Only the acquisition of experience in this procedure, followed by a careful analysis of its cost/efficacy balance, could change present behavior.

**Conclusions**

This study clearly shows that AT-G is significantly more effective than traditional vasoactive therapy alone in inducing IDU healing, restoration of the capillary bed in the treated finger, and a rapid resolution of local ischemic pain in patients with SSc. No adverse event was registered during the study. Altogether, our data suggest that AT-G may be considered a successful option to induce healing in ischemic SSc-related IDUs.

**Abbreviations**

ASC: Adipose-derived stromal cell; AT: Adipose tissue; BM-MSC: Bone marrow mesenchymal stem cell; dc: Diffuse cutaneous; G: Grafting; IDU: Ischemic digital ulcer; lc: Limited cutaneous; NVC: Nailfold videocapillaroscopy; SP: Sham procedure; SSc: Systemic sclerosis; SVF: Stromal/vascular fraction; VAS: Visual analogue scale

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Availability of data and materials
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors’ contributions
All authors were involved with the conception and design of the present study. NDP and CV were responsible for the design, statistical analysis and interpretation of the data. GDL was responsible for the surgical part of the study. RA, EZ, WM, and FP were involved in data acquisition. AM was involved at a later time for drafting of the additional figures, reference revision and final revision of the manuscript. All authors analyzed and interpreted data, drafted or revised the manuscript critically for important intellectual content, and approved the final version of the manuscript.

Ethics approval and consent to participate
All participants provided written informed consent prior to data collection. This study was conducted according to the Helsinki declaration and was approved by the local Ethical Committee (Comitato Etico Milano Area 2, Session 04.07.2017, Reference Number 138).

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

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References
1. Gabrielli A, Avvedimento EV, Krieg T. Sclerodermia. N Engl J Med. 2009;360:1989–2003.
2. Ferri C, Valentini G, Cozzi F, Sebastiani M, Michielisi C, La Montagna G, et al. Systemic sclerosis: demographic, clinical, and serologic features and survival in 1,012 Italian patients. Medicine (Baltimore). 2002;81:139–53.
3. Cutillo M, Pizzocori C, Secchi ME, Sulli A, Capillaroscopy. Best Pract Res Clin Rheumatol. 2006;20:1005–10.
4. Steen V, Denton CP, Pope JE, Matsuji-Cerinic M. Digital ulcers: overt vascular disease in systemic sclerosis. Rheumatology. 2009;48(Suppl 3):19–24.
5. Walker UA, Tyndall A, Czirjak L, Denton C, Farge-Bancel D, Kowal-Bielecka O, et al. Clinical risk assessment of organ manifestations systemic sclerosis: a report from the EUULAR Scleroderma Trials and Research Group database. Ann Rheum Dis. 2007;66:549–53.
6. Bronckaers A, Hilkens P, Martens W, Gervois P, Rajtazjak J, Struyt T, et al. Mesenchymal stem/stromal cells as a pharmacological and therapeutic approach to accelerate angiogenesis. Pharmacol Ther. 2014;143:181–96.
7. Christophet M, Schendel M, Foll J, Muller LP, Keyser B, Behre G. Marked improvement of severe progressive systemic sclerosis after transplantation of mesenchymal stem cells from an allogeneic haploidentical-related donor mediated by ligation of CD137L. Leukemia. 2008;22:1062–4.
8. Guiducci S, Porta F, Saccardi R, Guidi S, Ibbba-Manneschi L, Manetti M, et al. Autologous mesenchymal stem cells foster revascularization of ischemic limbs in systemic sclerosis: a case report. Ann Intern Med. 2010;153:650–4.
9. Neviati T, Ananeva L, Bykovskia M, Eremin I, Karandashev E, Rhenhnikov J, et al. Autologous mesenchymal stem cells in clinical medicine: a clinical prospective study. Cell Transplant. 2016;25:107–15.
10. Tsoyserkani NM, Christensen ML, Sheikh SP, Sorensen JA. Adipose-derived stem cells new treatment for wound healing? Ann Plast Surg. 2015;75:117–23.
11. Pikula M, Marek-Trzonkowska N, Wardowska A, Renkiewska A, Trzonkowski P. Adipose tissue-derived stem cells in clinical applications. Expert Opin Biol Ther. 2013;13:1357–70.
12. Kokai LE, Marra K, Rubin JP. Adipose stem cells: biology and clinical applications for tissue repair and regeneration. Transl Res. 2014;163:399–408.
13. Del Papa N, Di Luca G, Sambataro D, Zaccara E, Maglione W, Gabrielli A, et al. Regional implantation of autologous adipose tissue-derived cells induces a prompt healing of long-standing indolent digital ulcers in patients with systemic sclerosis. Cell Transplant. 2015;24:2297–305.
14. Van den Hoogen F, Khanna D, Frensen J, Johnson SR, Baron M, Tyndall A, et al. 2013 Classification criteria for systemic sclerosis. An American College of Rheumatology/European League Against Rheumatism Collaborative Initiative. Arthritis Rheum. 2015;65:2737–47.
15. Hachulla E, Hatan POY, Carpentier P, Agard C, Chatelo E, Jego P, Mouton H, et al. Effect of sildenafil on ischaemic digital ulcer healing in systemic sclerosis: the placebo-controlled SEDUCE study. Ann Rheum Dis. 2016;75:1009–15.
16. Hughes M, Murray A, Denton CP, Herrick AL. Should all digital ulcers be included in future clinical trials of systemic sclerosis-related digital vasculopathy? Med Hypotheses. 2018;116:101–4.
17. Hughes M, Herrick AL. Digital ulcers in systemic sclerosis. Rheumatology (Oxford). 2017;56(1):14–25.
18. Whigley FM, Wise RA, Seibold JR, McCloskey DA, Kujala G, Medsger TA Jr, et al. Intravenous iloprost infusion in patients with Raynaud phenomenon secondary to systemic sclerosis. Ann Intern Med. 1994;120:199–206.
19. Kapur SK, Katz AJ. Review of the adipose derived stem cell secretome. Biochimie. 2013;95:2222–8.
20. Sacerdote P, Niada S, Franchi S, Arrigoni E, Rossi A, Yenagi V, et al. Systemic administration of human adipose-derived stem cells reverts nociceptive hypersensitivity in an experimental model of neuropathy. Stem Cells Dev. 2013;22:1253–63.
21. Birni AT, Amodeo G, Ferreira LM, Niada S, Moschetti G, et al. Therapeutic effect of human adipose-derived stem cells and their secretome in experimental diabetic pain. Sci Rep. 2017;7(1):9904.
22. Ganel B, Daumus A, Jouve E, Harlé JR, Nguyen PS, Chabannon C, et al. Safety, tolerability and potential efficacy of injection of autologous adipose-derived stromal vascular fraction in the fingers of patients with systemic sclerosis: an open-label phase I trial. Ann Rheum Dis. 2015;74:2175–85.
23. Del Papa N, Caviggioli F, Sambataro D, Zaccara E, Di Luca G, et al. Autologous fat grafting in the treatment of fibrotic perioral changes in patients with systemic sclerosis. Cell Transplant. 2015;24:633–72.
24. Bura A, Plant-Backard V, Bourin P, Silvestre JS, Gross F, Grollou JL, et al. Phase I trial: the use of autologous cultured adipose-derived stroma/stem cells to treat patients with non-revascularizable critical limb ischemia. Cytotherapy. 2014;16:245–57.
25. Lee HC, An SG, Lee HW, Park JS, Cha KS, Hong TJ, et al. Safety and effect of adipose tissue-derived stem cell implantation in patients with critical limb ischemia: a pilot study. Circ J. 2012;76:1750–60.
26. Kim EK, Li G, Lee TJ, Hong JP. The effect of human adipose-derived stem cells on healing of ischemic wounds in a diabetic nude mouse model. Plast Reconstr Surg. 2011;128:387–94.
27. Nie C, Yang D, Xu J, Si Z, Jin X, Zhang J. Locally administered adipose-derived stem cells accelerate wound healing through differentiation and vasculogenesis. Cell Transplant. 2015;24:633–72.
28. Han SK, Kim HR, Kim WK. The treatment of diabetic foot ulcers with uncultured processed liposaprate cells: a pilot study. Wound Repair Regen. 2010;18:342–8.
29. Suga H, Glotzbach JP, Sorkin M, Longaker MT, Gurtner GC. Paracrine mechanism of angiogenesis in adipose-derived stem cell transplantation. Ann Plast Surg. 2014;72:234–31.
30. Baer PC, Schubert R, Bereiter-Hahn J, Ploessl M, Geiger H. Expression of a functional epidermal growth factor receptor on human adipose-derived mesenchymal stem cells and its signaling mechanism. Eur J Cell Biol. 2009;88:73–83.
31. Chen L, Tredget EE, Wu PY, Wu Y. Paracrine factors of mesenchymal stem cells recruit macrophages and endothelial lineage cells and enhance wound healing. PLoS One. 2008;3:e1886.
32. Yang J, Zhang H, Zhao L, Chen Y, Liu H, Zhang T. Human adipose tissue-derived stem cells protect impaired cardiomyocytes from hypoxia/reoxygenation injury through hypoxia-induced paracrine mechanism. Cell Biochem Funct. 2012;30:505–14.
33. Cipriani P, Di Benedetto P, Liakouli V, Del Papa B, Di Padova M, Di Ianni M, et al. Mesenchymal stem cells (MSCs) from scleroderma patients (SSc) preserve their immunomodulatory properties although senescent and normally induce T regulatory cells (Tregs) with a functional phenotype: implications for cellular-based therapy. Clin Exp Immunol. 2013;173:195–206.

34. Capelli C, Zaccara E, Cipriani P, Di Benedetto P, Magilione W, Andracco R, et al. Phenotypical and functional characteristics of in vitro-expanded adipose-derived mesenchymal stromal cells from patients with systemic sclerosis. Cell Transplant. 2017;26:841–54.

35. Kølle SF, Fischer-Nielsen A, Mathiasen AB, Elberg JJ, Oliveri RS, Glovinski PV, et al. Enrichment of autologous fat grafts with ex-vivo expanded adipose tissue-derived stem cells for graft survival: a randomised placebo-controlled trial. Lancet. 2013;382:1113–20.

36. Szöke K, Brinchmann J. Concise review: Therapeutic potential of adipose tissue-derived angiogenic cells. Stem Cells Transl. Med. 2012;1:658–67.