Bagaza Virus in Wild Birds, Portugal, 2021

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Bagaza virus emerged in Spain in 2010 and was not reported in other countries in Europe until 2021, when the virus was detected by molecular methods in a corn bunting and several red-legged partridges in Portugal. Sequencing revealed high similarity between the 2021 strains from Portugal and the 2010 strains from Spain.

Bagaza virus (BAGV) is a single-stranded, positive-sense RNA virus. The virus belongs to the mosquitoborne cluster of the genus Flavivirus, family Flaviviridae, which includes such other emerging pathogens as West Nile, Japanese encephalitis, dengue, Zika, and yellow fever viruses, all of which are associated with neurologic disease in animals and humans and have zoonotic potential (1). BAGV was first isolated in 1966 from a pool of Culex species mosquitoes in the Bagaza District of Central African Republic and was detected subsequently in several species of mosquitoes. The first BAGV-associated deaths in vertebrates were detected in Spain, in 2010, in red-legged partridges (Alectoris rufa) and ring-necked pheasants (Phasianus colchicus) (2) and then, in 2016, in Himalayan monal pheasants (Lophophorus impejanus) in South Africa (3).

BAGV infection causes neurologic disease in red-legged partridges, gray partridges (Perdix perdix), ring-necked pheasants, and, to a lesser degree, in common wood pigeons (Columba palumbus) (1–6). Estimated mortality rates range from 23% to 30% in naturally and experimentally infected red-legged partridges (5,7); rates are higher (up to 40%) in experimentally infected gray partridges (6) and lower rates in pheasants and columbiformes (4,7). We describe a BAGV outbreak in Portugal in autumn 2021, associated with abnormal fatalities in red-legged partridges and 1 corn bunting (Emberiza calandra).

On September 1, 2021, three red-legged partridges were found dead in Serpa, southern Portugal. From September through mid-October, 9 partridges and 1 corn bunting were found dead in the same area (Appendix Table, https://wwwnc.cdc.gov/EID/article/28/7/21-2408-App1.pdf). Local reports emerged of partridges displaying neurologic signs compatible with potential viral infection, such as disorientation and motor incoordination. Twelve of the 13 birds were necropsied. Laboratory examinations and preliminary diagnoses were conducted at the Research Institute in Hunting Resources (Ciudad Real, Spain) and at the Center for Research on Biodiversity and Genetic Resources (InBIO Laboratório Associado, Vairão, Portugal). Official diagnosis was determined at the National Institute of Agrarian and Veterinary Research, I.P. (Lisbon, Portugal). Growing feathery were collected from 30 partridges live-trapped in the same area on October 3.

Researchers conducted molecular detection by using RNA extracted from various sampling points (feather pulp, brain, heart, kidney, spleen, and intestine) and followed 2 strategies targeting different regions of the BAGV genome (nonstructural 2b, nonstructural 5 [NS5], and 3′ nontranslated region) (Appendix Table); first, a duplex quantitative reverse transcription PCR (RT-PCR) for the simultaneous and differential detection of Japanese encephalitis and Ntaya flavivirus serocomplexes (8), and second, a uniplex quantitative RT-PCR specific for the NS5 coding region of BAGV (9). The researchers used conventional nested RT-PCR for sequencing to target part of the NS5 gene (10) and an in-house RT-PCR (developed at the National Institute of Agrarian and Veterinary Research) to target part of the NS2b gene (Appendix Table).

Out of the 12 necropsied birds, 8 red-legged partridges and 1 corn bunting (75%) tested positive for BAGV, as did 4 of 30 live-captured red-legged partridges (13.3%) (Appendix Table). The 108 bp sequences obtained from duplex quantitative RT-PCR from partridge 9 and the corn bunting showed 100% similarity with the 3′ nontranslated region of the BAGV reference strain (GenBank accession no. HQ644143) detected in the 2010 outbreak in Spain (Appendix
Table). In comparing the NS5 regions, researchers found very high similarities with HQ644143 in the 110 base pair sequences obtained from 6 partridges by nested RT-PCR (99.1%) and in the 171 base pair sequences taken from 2 partridges by RT-PCR (98.8%).

Upon necropsy, all birds were in good body condition, suggesting an acute disease course. Histopathology, albeit hampered by autolysis and freezing artifacts, revealed lymphoid depletion in the spleen and severe congestion, moderate to abundant diffuse mononuclear inflammatory infiltrates, and focal necrosis in all tissues. The heart, brain, kidney, and liver were the most affected organs (Figure).

This work confirms BAGV emergence in Portugal, in autumn 2021, associated with abnormal fatalities in red-legged partridges. Active circulation of BAGV was also evidenced in the studied region, where 13.3% of live-captured red-legged partridges testing positive for BAGV, even though ecologic and demographic studies are required to determine the extent and magnitude of the outbreak. Substantial population decline in the red-legged partridge can be anticipated in this region of Portugal on the basis of the mortality rate previously estimated for this species (4,7). The fatal case in a songbird, the corn bunting, suggests that BAGV might have a broader spectrum and effect in wild bird species. This finding, combined with the small size of the analyzed sequences, suggests the need for further research to identify the vectors for BAGV in Portugal and their role in the epidemiology of the disease, and elucidate the phylogenetic relationships between the 2021 strains in Portugal and 2010 strains in Spain against known BAGV strains.

No conclusions can be made from this research regarding the origin of this infection. However, the introduction of the virus in Portugal might be linked to persistence of the disease and migration of infected wild birds from North Africa or Spain.

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**Hodgkin Lymphoma after Disseminated Mycobacterium genavense Infection, Germany**

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*Mycobacterium genavense* infection, a rare nontuberculous mycobacteria infection, occurs in heavily immunocompromised patients (i.e., those with advanced HIV disease, genetic disorders, or acquired immunologic disorders and those undergoing immunosuppressive therapy). We report a case of disseminated *M. genavense* infection preceding Hodgkin lymphoma in a patient without obvious risk factors for this infection. *Mycobacterium genavense* was first described in 1992 in HIV-positive patients with low CD4 counts and disseminated mycobacterial disease (1). Since the 2000s, additional risk factors for this bacterial infection became known (e.g., solid organ transplantation, hematopoietic stem cell transplantation, Epstein-Barr virus–associated lymphoproliferative disorder, neutralizing anti–interferon γ autoantibodies, adenosine deaminase deficiency, nuclear factor κB1 deficiency) (2,3). Clinical manifestations of *M. genavense* commonly involve blood and lymph nodes but can include the gastrointestinal tract, spleen, liver, and bone marrow; pneumonia, prostatic joint infection, endobronchial mass, and brain mass have also been described.

A previously healthy 23-year-old woman sought medical treatment at University Hospital Gießen (Gießen, Germany) for progressive cervical lymphadenopathy (Figure, panel A) and fever originating 4 months prior. A professional animal keeper, she had no history of previous infections or autoimmune disease, an remarkable family history, and no travel outside of Europe; her tattoos showed no signs of irritation. She experienced gender dysphoria and used masculinizing hormone therapy (testosterone). We
### Appendix Table. Bagaza virus–positive specimens analyzed in a study of wild birds in Portugal, 2021*

| Code   | Species | Status         | Collection date | Sex | Age class | Real time RT-PCR (RT-qPCR) | Uniplex RT-qPCR² | nested RT-PCR³ | RT-PCR⁴ |
|--------|---------|----------------|-----------------|-----|-----------|-----------------------------|-------------------|----------------|---------|
|        |         |                |                 |     |           | Duplex RT-qPCR¹             | Uniplex RT-qPCR² | nested RT-PCR³ | RT-PCR⁴ |
|        |         |                |                 |     |           | (3′NTR, 172 bp amplicon)    | (NS5, 61pb amplicon) | (NS5, 143 bp amplicon) | (NS2b, 209 bp amplicon) |
| Partridge 1 | A. rufa | Found dead    | 01/09/21        | Male | Adult | + | + | + | nt | nt | nt | +(#) | nt |
| Partridge 2 | A. rufa | Found dead    | 01/09/21        | Male | Adult | – | – | + | nt | nt | nt | +(#) | nt |
| Partridge 3 | A. rufa | Found dead    | 28/09/21        | Female | Juvenile | + | + | – | nt | nt | nt | – | nt |
| Partridge 4 | A. rufa | Found dead    | 29/09/21        | Female | Juvenile | + | + | – | nt | nt | nt | +(#) | nt |
| Corn Bunting | E. calandra | Found dead | 09/10/21        | Unknown | Unknown | – | – | – | +(#) | nt | nt | nt | nt |
| Partridge 7 | A. rufa | Found dead    | 13/10/21        | Male | Adult | nt | nt | nt | + | + | + | nt | +(#) |
| Partridge 8 | A. rufa | Found dead    | 13/10/21        | Male | Adult | nt | nt | nt | + | + | + | nt | +(#) |
| Partridge 9 | A. rufa | Found dead    | 13/10/21        | Female | Juvenile | +(#) | +(#) | + | nt | nt | nt | +(#) | nt |
| Partridge 11 | A. rufa | Found dead   | 14/10/21        | Female | Juvenile | + | + | + | nt | nt | nt | +(#) | nt |
| Partridge 14 | A. rufa | Live-trapped | 03/10/21        | Male | Adult | + | na | na | na | na | na | na | +(#) | na |
| Partridge 16 | A. rufa | Live-trapped | 03/10/21        | Male | Adult | + | na | na | na | na | na | na | – | na |
| Partridge 33 | A. rufa | Live-trapped | 03/10/21        | Male | Adult | + | na | na | na | na | na | na | – | na |
| Partridge 37 | A. rufa | Live-trapped | 03/10/21        | Male | Juvenile | + | na | na | na | na | na | na | – | na |

*+, positive; -, negative; nt, not tested; na, not applicable; # - samples for which successful sequences were obtained. 1- dRT-qPCR developed for the detection of Japanese Encephalitis virus and virus from the Ntaya serocomplex (108 nt-long sequences obtained from Partridge 9 and corn bunting showed 100% similarity with BAGV sequence HQ644143, position 10,510 to 10,680); 2- RT-qPCR developed for the specific detection of BAGV (not sequenced), 3- nested RT-PCR developed by Sánchez-Seco et al. (2005) for the detection of Flavivirus (110 nt-long sequences obtained from Partridge 1, 2, 4, 11 and 14 showed 99.1% similarity with BAGV sequence HQ644143, position 8,996 to 9,139); 4- RT-PCR, developed by the INIAV team (unpublished, 171 nt-long sequences obtained from Partridge 7 and 8 showed 98.8% similarity with BAGV sequence HQ644143). Sizes of the sequences do not include the primers’ annealing sequences. HQ644143 is the reference BAGV genome strain from the 2010 outbreak in Spain.

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