Potential Mutations Associated With Occult Hepatitis B Virus Status

Sima Besharat 1,2; Aezam Katoonizadeh 1; Abdolvahab Moradi 2,*

1 Liver and Pancreas Transplantology Diseases Research Center, Digestive Disease Research Institute, Tehran University of Medical Sciences, Tehran, IR Iran
2 Golestan Research Center of Gastroenterology and Hepatology, Golestan University of Medical Sciences, Gorgan, IR Iran
*Corresponding Author: Abdolvahab Moradi, Golestan Research Center of Gastroenterology and Hepatology, Golestan University of Medical Sciences, Gorgan, IR Iran. Tel: +98-5123408435; +98-9807772077, Fax: +98-721369720, E-mail: abmoradi@yahoo.com

Received: October 6, 2013; Revised: January 14, 2014; Accepted: February 20, 2014

Evidence Acquisition: In the present study we provided an in-depth review of the most important new data available on different mutations in HBV genome of patients with OHBS, which may play a role in the pathogenesis of OHBS. The data were collected through reviewing the full-text articles, identified by the PubMed search, using the following keywords and their different combinations: occult hepatitis B, HBV genome, “a” determinant, HBV open reading frames, S mutations, X mutations, P mutations and C mutations.

Results: Variants within the major hydrophilic region (MHR) of the HBsAg, deletions in the pre-S1 region, codon stop in the S open reading frame (ORF), sporadic non common mutations, some mutations affecting the posttranslational production of HBV proteins in the S ORF like deletion mutations, mutations in start codon and nucleotide changes in the X ORF, deletion and point mutations in P ORF and sometimes, nucleotide substitution in the C ORF are among the assumed mutations detected to have a role in OHBS appearance.

Conclusions: Studies mostly lacked a control group and the whole-length HBV sequencing was scant with conflicting results, suggesting that OHBS is often a result of multiple mechanisms. Additional studies on full-length HBV genomes from occult and non-occult HBV cases may shed more light on the interplay between different mechanisms involved in the pathogenesis of OHBS.

Keywords: Hepatitis B; Mutation; Virus Diseases

1. Context

Hepatitis B virus (HBV) infection is a global health problem, affecting more than 2 billion people worldwide, of whom approximately 350 million suffer from HBV-induced chronic liver diseases (1, 2). Depending on the interactions between the host and the virus, the natural course of HBV infection can be highly heterogeneous (3). Chronic HBV infection is diagnosed by detection of serum hepatitis B surface antigen (HBsAg), however, sometimes HBV infection can be presented in the absence of serum HBsAg, which is known as occult HBV status (OHBS). Accordingly, OHBS is characterized by the presence of HBV DNA in the liver, in the absence of serum HBsAg, with or without detectable HBV DNA in the serum. On the basis of HBV antibody profile, OHBS may be distinguished as: seropositive-OHBS (anti-HBc and/or anti-HBs positive) and Seronegative-OHBS (anti-HBc and anti-HBs negative) (4). The clinical relevance of OHBS has not been investigated extensively; however, several studies have suggested a potential association between OHBS and increased risk of cirrhosis and hepatocellular carcinoma (HCC). In addition, it can be transmitted through liver transplantation or blood transfusion (2, 5, 6).

Implication for health policy/practice/research/medical education:
This is a review of the potential mutations related to occult HBV status. There are several studies regarding the assumed mechanisms of associated mutations without identical conclusions.

Copyright © 2014, Kowsar Corp. Published by Kowsar Corp. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.
in four different open reading frames (ORFs). There are four partially overlapping ORFs encoding seven different HBV proteins. The largest ORF is the POL ORF, which encodes polymerase proteins. The S ORF comprises the pre-S1, pre-S2 and S regions and codes for large, middle and small sized inter-membrane surface proteins. The C ORF consists of pre core and core regions and codes the capsid (core) and the hepatitis B e antigen (HBeAg) proteins. HBeAg seems to have a role in the regulation of the immune response. The last ORF is X, which encodes the X protein. The transcription regulation activity of this protein has been suggested by some investigators (Figure 1) (5, 8-14).

Mechanisms underlying OHBS are poorly understood. Several possibilities have been suggested including: A) interference of HBV replication by other viruses (like HCV in case of HCV coinfection), B) integration of HBV-DNA into host cell chromosomes, C) formation of circulating HBV-containing immune complexes, which are not detected by routine HBsAg determining tests, D) altered host immune responses leading to the maintenance of HBV infection in a latent state until transmission to another individual occurs (mostly in case of immunosuppressive therapy), E) different mutations in HBV DNA sequence which is the main focus of the current review and will be discussed in detail (Table 1) (2, 6, 15).

2. Evidence Acquisition

In the present study we provided an in-depth review of the most important new data available on different mutations in HBV genome of patients with OHBS, which may play a role in the pathogenesis of OHBS. The data were collected through the review of the full-text articles identified by PubMed search, using the following keywords and their different combinations: occult hepatitis B, HBV genome, “a” determinant, HBV open reading frames, S mutations, X mutations, P mutations and C mutations.

3. Results

3.1. Mutations in the S Region of HBV and Occult HBV Status

The S region of ORFs consists of three AUG codons coding the expression of three proteins: large (L), middle (M) and small (S). Pre-S1 domain is unique for L protein. Pre-S2 domain is the shared sequence with the M protein and the S domain is seen in all three proteins. The L and S proteins are essential for virion formation and the M could enhance the virion secretion efficiency (19, 34, 35). The S and M proteins are detected as HBsAg. HBsAg is a peptide with 226 amino acids (aa) with a single major antigenic determinant called the “a” determinant, located in the

**Table 1.** Studies Investigating Mutations Associated With Occult Hepatitis B Virus Status

| Affected Region | First Author | Year | Journal Name | Sample | Main Mutations Found to be Responsible for OHBS |
|----------------|--------------|------|--------------|--------|-----------------------------------------------|
| S region       | Hou et al. (16) | 2001 | Hepatology   | OHBS patients | G145R mutation, some positions inside and outside the “a” determinant |
|                | Ma et al. (17)  | 2012 | J Med Virol  | OHBS patients | G145R mutation, escape mutations mostly in the “a” determinant |
|                | Liu et al. (18) | 2010 | Virol J       | OHBS in blood donors | substitutions in the regions from aa 117 to 121 and aa 144 to 147, located in the MHR and mutants with single-point or multi-point G145R mutations |
-amino acid positions between 100 and 160. The dominant epitopes of HBSAg, which are the targets of neutral-
izing B cell responses, are located in the "a" determinant (aa 124-147) within the MHR. Mutations inducing a conformational change within the "a" determinant cause making a protein with significant changes in the antigenic epitope. These changes lead to the production of the undetectable HBsAg (21).

The most common and problematic MHR mutation, G145R, is an increasing event due to the global implication of vaccination programs and the pressure of antiviral therapy (21). Variants within the MHR of HBsAg were the point of interest in Hou et al. study in China. In 46 cases with OHBS, there were 32 amino acid substitutions found between positions 100-160 of the MHR. In addition to the G145R, 11 positions inside and five positions outside the "a" determinant were involved. Combined mutations were also detected in some patients. Another two patients had insertion mutations immediately before the "a" determinant (16).

Ma et al. in a study conducted China, 2012, found other eight escape mutations associated with OHBS, in addition to the G145R, located mainly at positions 120, 126, 130, 133, 134, 137, 140, 143 and 144 with a genotypic heterogeneity (17). In the aforementioned study, a comparison was performed between OHBS patients and a group of HBV carriers, which could be considered as the strength of this work. Substitutions in the regions from aa 117 to 121 and aa 144 to 147 located in the MHR of the S gene and mutants with single-point or multi-point G145R mutations were also reported in the studies from China (18, 19). One of these studies was a phylogenetic one on blood donors and no comparisons with non-OHBS patients was performed in it (19). Other single or multiple aa substitutions have been reported to be responsible in OHBS (20).

Panigrahi et al. compared the 64-160 aa of 60 HBsAg (-) samples, with the reference sequences of each genotype, in their study on 729 HBsAg negative donor samples in India. They found single or multiple aa substitutions in 95% of the OHBS cases. T125M was the most common (93.3%) aa substitution found in the MHR, mostly in subgenotype D3. Substitutions were also found at codon A128V, G71 D, L95 S, M103I, P111L, S133A, S144P, S177G, T115I, T116P, T118R, and T127A (20).

In an interesting recent study Huang et al. compared the characteristics of 61 patients with OHBS to 153 HBsAg (+) carriers with low titers of serum HBsAg (HBsAg-L group) and 54 samples with high serum HBsAg (HBsAg-H group). MHR mutations were seen significantly more frequently in OHBS cases (55.7%) compared to the HBsAg-L (34.0%) or the HBsAg-H groups (17.1%). Thirteen representative MHR mutations were observed in patients with OHBS. Four out of the 13 mutations strongly decreased the analytical sensitivity of seven commercial HBsAg immunoassays and 10 significantly impaired virion and/or S protein secretion in both HuH7 cells and mice (36). This was a significant study regarding the comparisons.

Besides, several investigations have described mutations clustering in the aforementioned key immuno-dominant regions of the HBsAg, which are able of decreasing the immune recognition of the virus, structural alteration and various mutations in genomic regulatory regions, leading to a strong reduction of HBsAg expression (4, 5, 7, 22, 24). In a functional survey by Sengupta et al. in India, the production, secretion and localization of surface proteins of HBV were studied in HepG2 cells, transfected with the wild-type and mutant pre-S1 and pre-S2/S promoters of HBV molecular clones 3131. Their results indicated that transfected cells had reduced HBV surface protein secretion and showed cytoplasmic aggregation of HBV surface proteins. It could be concluded that OHBS may be caused due to mutations in pre-S1 and pre-S2/S promoters/pre-S1 coding region, which leads to reduced secretion of HBsAg, aggregation of HBsAg in the endoplasmic reticulum and HBsAg seronegativity (22). This was one of the few studies performed on HBV molecular clones and transfected hepatic cells, which made them capable of investigating the possible mechanisms causing OHBS, more closely.

Deletions in the pre-S2 region and the resulted impaired viral packaging, has also been reported as another mechanism for OHBS. In Chen et al. (23) study, two kinds of deletions were seen covering the pre-S1 start codon and B-cell antigenic epitope in the pre-S1 protein (aa19-26), leading to a decrease in HBsAg and HBV virus particles in the serum. In another subject, a deletion was observed (nt. 3145-52), covering nearly the entire pre-S2 region. In one case, a deletion in pre-S2-promoter (nt. 3145-52) was identified, covering almost the whole pre-S2 region. It has been known that deletions overlapping this region could decrease the expression of the M protein, which reduces virion secretion. In this study cloning and sequencing the full-length genome of HBV was done only on nine healthy young Chinese patients with OHBS, who received neonatal vaccination. Although it was a powerful study due to the full-length genome sequencing, there was not any control group. Pollicino et al. investigated the lack of HBsAg production (or detection) and the inhibition of viral replication as major aspects of OHBS in their study. They studied frozen liver specimens of 17 HBV patients (13 OHBS and four HBsAg (+) patients as a control group). Cloning and sequencing of the pre-S genomic region was detected in only one case with small in-frame deletion and two more cases, out of 13 patients with OHBS, showed point mutation in pre-S2 start codon. No important mutation was found in the pre-S1 region of HBV clones from 16 patients. Large intra-individual genetic heterogeneity was observed in OHBS cases, comparable to the HBsAg (+) subjects. Therefore, the authors concluded that the viral genomic variability does not appear to play a fundamental role in inducing the OHBS and host immune system but probably epigenetic mechanisms can play critical roles (24).

In another complete genome assessment conducted in New Delhi (2004), the major observations were: frequent quasi species variation, deletion in pre-S2/S region affect-
A1762T plus G1764A core promoter mutations also cause with the X gene in the concomitant reading frame, the enhancer II. Because the basal core promoter overlaps control replication, like the basal core promoter and the enhancer II sequence of HBV DNA and created a translational stop codon which truncated the X protein by 20 amino acids from the C-terminal end. All the HBV DNAs had a precore mutation at the 83rd nucleotide, resulting in disruption of HBsAg synthesis (29). In this study, serum HBV DNA from patients with non-B non-C hepatitis (NBNC) was sequenced and compared to that of the patients with alcoholic liver disease and autoimmune hepatitis. Therefore, these results could be very interesting keeping the studied group in mind.

The start codon in the X region could also be mutated and cause OHBS. In a Japanese study, the ATG (methionine) start codon had mutated to GTG (valine) and resulted in OHBS in one case (30). Fujise sequenced the full genome of HBV in this seronegative case of OHBS, which is worth giving more attention. Nucleotide exchange of A1762T and G1764A is another important mutation, which has been suggested to be responsible in OHBS. This was reported in Pollicino's study on a group of 13 OHBS and four cases of overt HBV (the control group). They reported the double mutation of A1762T and G1764A in 4 OHBS cases and three controls. Triple mutation of these two plus G766T was only observed in two OHBS cases. Point mutations (from 1 to 4) in BCP were also reported in the mentioned groups (24). The results of Pollicino's study are valuable regarding the comparison they made between OHBS cases and overt patients with HBV from the point of potential mutations assumed to be responsible in OHBS.

### 3.2. Mutations in the X Region of HBV and Occult HBV Status

X ORF produces the X protein (HBx) and although the exact function of HBx during HBV replication is still unclear, multiple studies suggest that HBx is necessary for viral replication in vivo and in vitro (30, 37-40). Mutations in the X region can involve the regulatory elements that control replication, like the basal core promoter and the enhancer II. Because the basal core promoter overlaps with the X gene in the concomitant reading frame, the A1762T plus G1764A core promoter mutations also cause changes in the X gene at xK10M and xV13I (30, 37-40).

Deletion mutations in X region are found in OHBS patients. Fukuda et al. in a very early study on X gene mutation in Japan, showed an identical 8-nucleotide deletion mutation at the distal part of the X region in a major group of these patients (85.7%). This mutation affected the core promoter and the enhancer II sequence of HBV DNA and created a translational stop codon which truncated the X protein by 20 amino acids from the C-terminal end.

### 3.3. Mutations in P Region and Occult HBV Status

One of the regions with mutation susceptibility in HBV ORF is P region, which encodes the polymerase protein (reverse transcriptase) or the POL. The HBV genome is organized in a way that the envelope (S) gene is completely overlapped by the polymerase gene, so it is logical to assume that changes in virus encoding, associated with antiviral resistance in the polymerase, may have consequent changes on the envelope gene (27), showing a close relationship between mutations in S and P regions of HBV genome.

This region is also susceptible to deletion mutations and having a key role in HCC progression. In a full-length genome study of HBV DNA in China, 14 out of the 16 clones, constructed from 3 cases of genotype B showed deletions in the P region. These deletions were located between nt. 2067 and 2349, covering the start codon of the P region, which is believed to reduce the enzymatic activity of the wild-type protein and may be accounted for low viral loads in OHBS (31). Forty point mutations in polymerase gene were found, resulting in changes in 11 amino acids in one case of OHBS, in a study by Fang et al. conducted in...
a high endemic area for HCC, in China in 2004 (32). Therefore, mutations in this region should draw the attention to the importance of related drug resistance and hepatocarcinogenesis.

3.4. Mutations in C Region and Occult HBV Status

C ORF of HBV genome encodes core protein and HBeAg (8-10). The core shell of hepatitis B virus is a potent immune stimulator, stimulating a strong neutralizing immune response to foreign epitopes (39, 40). Mutations in this region of HBV genome have not been assumed to be responsible for OHBS, as frequently as other regions spoken above, therefore there are not as many studies done on the subject. In one study in China BCP deletion mutation was investigated in three clones from one case (nt. 1754, nt. 1751, and nt. 1754). The deletions in the BCP region covered more than one TA box. In the C region, deletions were observed in 4 subjects. Among 14 strains with deletions in the C region, 11 had deletions in all parts of both the C and P regions, in all cases with genotype B. In one case, two deletions (nt. 2001-2050 and nt. 2152-2222) covered 22% of the C region (31).

In another study Garcia-Montalvo et al. reported 24 (6.4%) cases with OHB Samong 372 Mexican blood donors. Phylogenetic analysis in this subgroup showed a substitution in the core region of nine samples, mostly located in immune dominant epitopes. There was no precore stop codon mutants in these patients (33). Truncated precore and core mutants, resulting in stop signals were found in another study in India on patients diagnosed as OHBS, who were not on anti-viral treatment (25). All mentioned studies were observational surveys on a group of patients with OHBS, looking for mutations in the specific region of HBV genome.

On the other hand, Pollicino et al. in a whole genome study on OHBS and overt HBV cases reported G1896A nucleotide mutation, resulting in a stop signal at codon 28, within the precore region, which prevents the HBeAg synthesis in eight out of 13 OHBS cases and in all 4 overt HBsAg (+) cases (control group). Missense mutations within different core antigen immunogenic epitopes were also observed in HBV isolates, in both patients with OHBS and the control group with overt HBV infection (24).

4. Conclusions

OHBS is a complex clinical entity documented worldwide. HBV sequences from these individuals demonstrate numerous mutations/deletions and alterations that can result in decreased immune recognition of the virus, impaired HBV packaging and decreased HBsAg expression. Moreover, mutations affecting post-translational protein production and treatment-associated mutations are observed in these patients. However, the aforementioned studies mostly lacked a control group. In addition, whole-length HBV sequencing data, resulting in direct comparison of mutations between the genome sequences of occult and non-occult strains, even though scant, have conflicting results suggesting that OHBS is often a result of multiple mechanisms. Additional studies on full-length HBV genomes from occult and non-occult HBV cases may shed more light on the interplay between different mechanisms involved in the pathogenesis of OHBS. Such insights are of utmost importance to develop new therapeutic strategies.

Acknowledgements

Authors tend to appreciate kind support and expertise guidance of Dr. Hossein Poustchi and Dr. AshrafMohamadkhani (DDRI).

Authors’ Contribution

Sima Besharat and Aezam Katoozizadeh contributed in study concept, design and drafting the manuscript and Abdolvahab Moradi contributed in critical revision of the manuscript for important intellectual content and study supervision.

Financial Disclosure

No financial support was taken for this review.

Funding/Support

Authors received no financial support or grant for this review.

References

1. Ocana S, Casas ML, Buhigas I, Lledo J. Diagnostic strategy for occult hepatitis B virus infection. World J Gastroenterol. 2010;16(12):1553-7.
2. de la Fuente RA, Gutiérrez ML, García-Samaniego J, Fernandez-Rodriguez C, Lledo J, Castellano G. Pathogenesis of occult chronic hepatitis B virus infection. World J Gastroenterol. 2011;17(32):3543-8.
3. Han Y, Zhao J, Ma LY, Yin JH, Chang WJ, Zhang HW, et al. Factors predicting occurrence and prognosis of hepatitis-B-virus-related hepatocellular carcinoma. World J Gastroenterol. 2011;17(38):4258-70.
4. Raimondo G, Allain P, Brunetto MR, Buendia MA, Chen DS, Colombo M, et al. Statements from the Taormina expert meeting on occult hepatitis B virus infection. J Hepatol. 2008;49(4):652-7.
5. Dandi M, Locarnini S. New insight in the pathobiology of hepatitis B virus infection. Gut. 2012;61 Suppl 1:i6-17.
6. Said ZN. An overview of occult hepatitis B virus infection. World J Gastroenterol. 2011;17(35):3927-38.
7. Raimondo G, Pollicino T, Cacciola I, Squadrito G. Occult hepatitis B virus infection. J Hepatol. 2007;46(5):650-70.
8. Dienstag JL. Hepatitis B virus infection. N Engl J Med. 2008;359(14):1486-500.
9. Nguyen DH, Lidgate I, Hu J. Hepatitis B virus-cell interactions and pathogenesis. J Cell Physiol. 2008;216(2):289-94.
10. Chang J, Lewin SR. Immuno-pathogenesis of hepatitis B virus infection. Immuno Cell Biol. 2007;85(5):16-23.
11. Martin CM, Welge JA, Shire NJ, Rouster SD, Shata MT, Sherman KE, et al. Genomic variability associated with the presence of occult hepatitis B virus in HIV co-infected individuals. J Viral Hepat. 2010;17(8):588-97.
12. Bruni R, Prosperi M, Marcantonio C, Amadori A, Villano U, Tritarelli E, et al. A computational approach to identify point muta-
tions associated with occult hepatitis B: significant mutations affect coding regions but not regulative elements of HBV. Virol J. 2018; 15:39.
13. Mohammadkhani A, Montazeri G, Poulishi H. The Importance of Hepatitis B Virus Genotype Diversification in Basal Core Promoter Region. Middle East J Dig Dis. 2013; 2(3):49–50.
14. Kramvis A, Kev M, Francois G. Hepatitis B virus genotypes. Vaccine. 2005; 23(19):2409–23.
15. Raimondo G, Caccamo G, Filomia R, Pollicino T. Occult HBV infection. Semin Immunopathol. 2013; 35(5):39–52.
16. Hou J, Wang Z, Cheng J, Lin Y, Lau GK, Sun J, et al. Prevalence of naturally occurring surface gene variants of hepatitis B virus in nonimmunized surface antigen-negative Chinese carriers. Hepatology. 2008; 47(5):1027–34.
17. Ma Q, Wang Y. Comprehensive analysis of the prevalence of hepatitis B virus escape mutations in the major hydrophilic region of surface antigen. J Med Virol. 2012; 84(4):209–206.
18. Liu Y, Li P, Li C, Zhou J, Wu C, Zhou YH. Detection of hepatitis B virus DNA among accepted blood donors in Nanjing, China. Virol J. 2010; 7:293.
19. Yuan Q, Ou SH, Chen CR, Ge SX, Pei B, Chen QR, et al. Molecular characteristics of occult hepatitis B virus DNA in southeast China. Clin Microbiol. 2010; 48(2):357–62.
20. Panigrahi R, Biswas A, Datta S, Banerjee A, Chandra PK, Mahapatra PK, et al. Antibody and anti-HBc antibody testing with detection and characterization of occult hepatitis B virus by an in-house nucleic acid testing among blood donors in Behrampur, Ganjam, Orissa in southeastern India: implications for transfusion. Virol J. 2010; 7:204.
21. Huang C-H, Yuan Q, Chen P-J, Zhang YJ, Chen CR, Zheng QB, et al. Influence of hepatitis B virus in hepatitis B virus surface protein on viral antigenicity and phenotype in occult HBV strains from blood donors. J Hepatol. 2012; 57(4):720–9.
22. Sengupta S, Rehman S, Durgapal H, Acharya SK, Panda SK. Role of surface promoter mutations in hepatitis B surface antigen production and secretion in occult hepatitis B virus infection. J Med Virol. 2007; 79(3):220–8.
23. Chen CJ, Yang HI. Natural history of chronic hepatitis B REVEaled. J Gastroenterol Hepatol. 2012; 26(4):628–38.
24. Pollicino T, Raffa G, Costantino I, Lisa A, Campello C, Squadrato G, et al. Molecular and functional analysis of occult hepatitis B virus isolates from patients with hepatocellular carcinoma. Hepatology. 2007; 45(2):277–85.
25. Chaudhuri V, Telay R, Nayak B, Acharya SK, Panda SK. Occult hepatitis B virus infection in chronic liver disease: full-length genome and analysis of mutant surface promoter. Gastroenterology. 2004; 127(5):1556–71.
26. Ito K, Qin Y, Guarnieri M, Garcia T, Kwei K, Mizokami M, et al. Immunoprevention of hepatitis B virus virion secretion by single-aminocacid substitutions in the small envelope protein and rescue by a novel glycosylation site. J Virol. 2010; 84(24):12850–61.
27. Sheldon J, Soriano V. Hepatitis B virus escape mutants induced by antiviral therapy. J Antimicrob Chemother. 2008; 62(4):766–8.
28. Hass M, Hannoun C, Kalinina T, Sommier G, Manegold C, Gunther S. Functional analysis of hepatitis B virus reactivating in hepatitis B surface antigen-negative individuals. Hepatology. 2005; 42(1):93–103.
29. Fukuda R, Ishimura N, Kushiyama Y, Moriyama N, Ishihara S, Chowdhury A, et al. Hepatitis B virus with X gene mutation is associated with the majority of serologically “silent” non-B, non-C chronic hepatitis. World J Hepatol. 2010; 2(2):56–60.
30. Chen SJ, Zhao YK, Fang Y, Xu WZ, Ma YX, Song ZW, et al. Viral deletions among healthy young Chinese adults with occult hepatitis B virus infection. Virus Res. 2012; 163(1):197–201.
31. Fang ZL, Zhaoa H, Wang b, Ge XM, Harrison TJ. Hepatitis B virus genotypes, phylogeny and occult infection in a region with a high incidence of hepatocellular carcinoma in China. World J Gastroenterol. 2004; 10(22):3264–8.
32. Garcia-Montalvo BM, Ventura-Zapata LP. Molecular and serological characterization of occult hepatitis B infection in blood donors from Mexico. Ann Hepatol. 2010; 10(2):333–41.
33. Chotiyaputta W, Lok AS. Hepatitis B virus variants. Nat Rev Gastroenterol Hepatol. 2009; 6(8):453–62.
34. Hsu CW, Yeh CT. Emergence of hepatitis B virus S gene mutants in patients experiencing hepatitis B surface antigen seroconversion after peginterferon therapy. Hepatology. 2011; 54(2):301–9.
35. Tang H, Oishi N, Kaneko S, Murakami S. Molecular functions and biological roles of hepatitis B virus X protein. Cancer Sci. 2006; 97(10):977–83.
36. Wei Y, Neuvect C, Tiollais P, Buendia MA. Molecular biology of the hepatitis B virus and role of the X gene. Pathol Biol (Paris). 2010; 58(4):267–72.
37. Sreegr C, Mason WS. Hepatitis B virus biology. Microbiol Mol Biol Rev. 2000; 64(4):51–68.
38. Murakami S. Hepatitis B virus X protein: a multifunctional viral regulator. J Gastroenterol. 2001; 36(10):653–60.
39. Rosenman AM, Borschkova O, Berriman JA, Wynnne SA, Pumpens P, Crowther RA. Structures of hepatitis B virus cores presenting a model epitope and their complexes with antibodies. J Mol Biol. 2012; 423(1):63–78.