| タイトル | Title |
| --- | --- |
| Roles of Src family kinase, Ras, and mTOR signaling in intestinal epithelial homeostasis and tumorigenesis |

| 著者 Author(s) |
| --- |
| Matozaki, Takashi / Kotani, Takenori / Murata, Yoji / Saito, Yasuyuki |

| 掲載誌・巻号・ページ Citation |
| --- |
| Cancer Science,112(1):16-21 |

| 刊行日 Issue date |
| --- |
| 2021-01 |

| 資源タイプ Resource Type |
| --- |
| Journal Article / 学術雑誌論文 |

| 版区分 Resource Version |
| --- |
| publisher |

| 権利 Rights |
| --- |
| © 2020 The Authors. Cancer Science published by John Wiley & Sons Australia, Ltd on behalf of Japanese Cancer Association. This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes. |

| DOI |
| --- |
| 10.1111/cas.14702 |

| JaLCDOI |
| --- |
|  |

| URL |
| --- |
| http://www.lib.kobe-u.ac.jp/handle_kernel/90007776 |

PDF issue: 2021-04-28
INTRODUCTION

Intestinal epithelial cells (IECs) of the small and large intestine turn over rapidly and are renewed every 3-5 days in both mouse and human. Intestinal epithelial cells are continuously regenerated from Lgr5+ intestinal stem cells (ISCs) that reside in a region of the epithelium near the base of intestinal crypts (Figure 1A). These ISCs generate proliferating progeny, known as transient amplifying (TA) cells, that eventually differentiate into various cell lineages of mature intestinal villi including absorptive enterocytes, mucin-producing goblet cells, and antimicrobial peptide-producing Paneth cells. Paneth cells in the crypt are also thought to be important for maintenance of ISCs by secreting Wnt ligands such as Wnt3. The mature enterocytes then migrate up the crypt toward the tip of the villus and eventually die as a result of their expulsion from the luminal surface of the intestinal epithelium. The elimination of older cells is thus balanced by the ongoing production of new IECs in each crypt, resulting in the rapid turnover of IECs.

The proliferative activity of crypt cells, such as TA cells and ISCs, is thought to be a major determinant of the rapid turnover rate of mature IECs. However, aberrant activation of this proliferative activity likely contributes to the development of intestinal inflammation and intestinal cancer. The roles of receptor protein tyrosine kinases and their downstream signaling molecules such as Src family kinases, Ras, and mTOR in homeostatic regulation of IEC turnover have recently been evaluated. These signaling pathways have been found to promote not only the proliferation of IECs but also the differentiation of progenitor cells into secretory cell types such as goblet cells. Of note, signaling by Src family kinases, Ras, and mTOR has been shown to oppose the Wnt-β-catenin signaling pathway and thereby to limit the number of Lgr5+ intestinal stem cells or of Paneth cells. Such cross-talk of signaling pathways is important not only for proper regulation of IEC homeostasis but for the development of intestinal tumors and potentially for anticancer therapy.

KEYWORDS
Intestinal epithelial cell, mTORC1, Ras, Src, Wnt
of ISCs as well as in the generation of Paneth cells (Figure 1B), which contribute to the nursing of ISCs, as mentioned above. Epidermal growth factor (EGF), whose receptor is a protein tyrosine kinase (PTK), is also thought to serve as a major driver of proliferation of TA cells as well as IECs. Moreover, signaling molecules such as Ras and mTOR that act downstream of the EGF receptor protein tyrosine kinase (RPTK) have been much studied with regard to their role in regulation of the proliferative activity of IECs. In this review, we will describe new insights into the functions of such signaling molecules that act downstream of RPTKs in the regulation of IEC homeostasis, including their cross-talk with Wnt-β-catenin signaling and their relation to intestinal oncogenesis and its potential treatment.

2 SRC FAMILY KINASES

Src family kinases (SFKs)—such as Src, Fyn, and Yes—are nonreceptor-type PTKs that play important roles in the regulation of cell proliferation, migration, and differentiation. They are thought to interact with RPTKs such as the EGF receptor or with activated focal adhesion kinase in response to integrin ligation, resulting in the activation of downstream Ras-MAPK and PI3K pathways (Figure 2A). Src family kinases are expressed predominantly in the crypt area of the intestinal epithelium and are localized to lipid rafts, which are thought to be enriched at the apical surface of crypt IECs. Lipid rafts thus likely recruit SFKs to the apical surface of crypt IECs. Ablation of Src specifically in IECs of mice resulted in no apparent defect under the steady-state condition, whereas that of Src, Fyn, and Yes induced apoptosis in IECs of the small intestine. However, IEC-specific ablation of Src did prevent the regeneration of crypts after the induction of DNA damage by γ-irradiation. Csk is a PTK that inhibits the activity of all SFKs by catalyzing phosphorylation of a COOH-terminal regulatory tyrosine residue. Indeed, ablation of Csk specifically in IECs of mice resulted in hyperactivation not only of Src but also of Fyn and Yes, and gave rise to intestinal and colonic tumors. Therefore, SFKs play crucial roles in the regulation of intestinal epithelial cell homeostasis and oncogenesis.
epithelial hyperplasia. \(^9\) It also increased the proliferative activity and turnover of IECs as well as the number of goblet cells in the intestine, and all of these effects were accompanied by activation of the small GTP-binding protein Ras and the transduction factor Yes-associated protein (YAP) in intestinal crypts (Figure 2B). \(^9\) Indeed, YAP had previously been shown to be expressed predominantly in crypts of the small intestine. \(^13\) Src family kinases are thus likely important not only for the proliferative activity and turnover of IECs but also for control of the differentiation and maturation of goblet cells.

In contrast with its effects on IECs, IEC-specific ablation of Csk resulted in a reduction in the number of ISCs and of Paneth cells, and it attenuated the expression of Wnt target genes in the intestine. \(^9\) Given that the Wnt-\(\beta\)-catenin signaling pathway is essential for the generation and maintenance of both ISCs and Paneth cells, \(^1,2\) SFKs could negatively regulate Wnt-\(\beta\)-catenin signaling in the intestinal epithelium. Furthermore, given that YAP inhibits the Wnt-\(\beta\)-catenin signaling pathway, \(^14-16\) SFKs likely target this pathway by upregulating YAP activity in crypt IECs (Figure 2B).

The importance of SFKs for promotion of IEC proliferation in the physiological condition suggests that aberrant activation of these kinases likely contributes to the development of colorectal cancer (CRC). Indeed, more than 80% of individuals with CRC have been found to overexpress Src in tumor tissue. \(^17\) The Apc\(^{min/+}\) mouse is an experimental model of CRC in which Wnt-\(\beta\)-catenin signaling is markedly activated and in which ablation of Src was found to result in a pronounced attenuation of tumorigenesis and increase in survival of the mutant mice, \(^6\) suggesting that Src is required for tumorigenesis in response to Apc loss. By contrast, it was also suggested that there are few activating mutations in Src in any type of cancer, although upregulation of Src protein and activity is frequently observed. \(^18\) Moreover, although many inhibitors of Src have been developed and tested clinically, they have not received much attention with regard to their potential for CRC therapy. \(^17\) Given that SFKs are thought to promote IEC proliferation in a YAP-dependent manner, \(^9,19\) YAP is another promising target for CRC therapy.

3 | RAS

The small GTP-binding protein Ras is a key signaling molecule that acts downstream of RPTKs, such as the EGF receptor, to promote the proliferation and differentiation of IECs (Figure 2A). Indeed, activation of Ras specifically in IECs has been shown to result in the development of hyperplasia throughout the intestinal epithelium of mice. \(^20-22\) The IEC-specific activation of Ras also increased the proliferative activity and turnover of IECs as well as the number of goblet cells, whereas it led to a marked reduction in the number of Paneth cells in crypts. \(^21\) However, aberrant activation of Ras alone failed to induce the development of cancerous lesions in the mouse intestine. \(^21,23\) The Ras family of proteins comprises K-Ras, N-Ras, and H-Ras, \(^24\) and the effect of IEC-specific genetic ablation of all three family members has not been determined in mice. Src homology 2-containing protein tyrosine phosphatase 2 (Shp2, also known as PTPN11) is a cytoplasmic protein tyrosine phosphatase that contains two tandem Src homology 2 domains. \(^25,26\) Given that Shp2 is required for the activation of Ras by various growth factors and cytokines, ablation of Shp2 likely results in the efficient extinction of all Ras activity. \(^25,26\) The IEC-specific ablation of Shp2 was found to result in severe enterocolitis in mice. \(^27-29\) In addition, the numbers of absorptive enterocytes and goblet cells were markedly reduced in such Shp2 mutant mice, and the development of intestinal organoids from isolated crypts of these animals was impaired. \(^27-29\) These observations thus implicate the Ras-MAPK signaling pathway in promotion both of IEC proliferation and of the differentiation of goblet cells and absorptive enterocytes (Figure 3). \(^28,29\) In contrast, ablation of Shp2 in IECs resulted in a marked increase in the number of Paneth cells as well as of Lgr5/Olfm4+ ISCs at the crypt base. \(^29\) Indeed, the expression of Wnt target genes such as Myc and Cdk4 was markedly increased in IECs by ablation of Shp2, suggesting that Ras activity downregulates Wnt signaling in these cells (Figure 3).

4 | MAMMALIAN TARGET OF RAPAMYCIN

The serine-threonine kinase mTOR forms two distinct multiprotein complexes known as mTOR complex 1 (mTORC1) and mTORC2. \(^30\) The activity of mTORC1 is controlled by various upstream signals including growth factors, nutrients, and stress. The ligation of RPTKs by growth factors such as insulin or EGF induces the activation of PI3K and the downstream serine-threonine kinase Akt. Akt activates the small GTP-binding protein Rheb through phosphorylation and consequent inhibition of tuberous sclerosis complex 2 (Tsc2), a GTPase-activating protein for Rheb, and Rheb then activates mTORC1. Such Rheb-mediated activation of mTORC1 results in the phosphorylation of ribosomal protein S6 kinase and the translational repressor protein 4E-BP1 and in consequent regulation of a variety of cellular processes such as protein synthesis, autophagy, and cell aging. \(^30\) Activation of mTORC1 by IEC-specific ablation of Tsc2 was found to promote the proliferative activity of IECs in the

![FIGURE 3 Role of Ras and mTORC1 in regulation of intestinal epithelial cell proliferation and differentiation. ISC, intestinal stem cell; TA, transient amplifying; Tsc2, tuberous sclerosis complex 2.](image-url)
small intestine and colon of mice (Figure 3). Conversely, ablation of mTOR in IECs reduced their proliferative activity. As mentioned above, SFKs and Ras, both of which function downstream of growth factor receptors, promote not only the proliferation of IECs but also the generation and differentiation of secretory cells such as goblet cells (Figures 2 and 3). In contrast, activation of mTORC1 by Tsc2 ablation did not affect the number of Muc2+ mucus-secreting cells in the intestine, whereas it disturbed the clustering of Paneth cells at the crypt base, suggesting that mTORC1 activity is not essential for differentiation of either of these cell types from their progenitors. The number of Lgr5+ ISCs and the expression of Wnt target genes in the intestine were markedly reduced by activation of mTORC1. Indeed, Tsc1/2 is required for the maintenance of ISCs in Drosophila. The activity of mTORC1 was also shown to be important for downregulation of Wnt signaling in melanocytes.

Given the importance of mTORC1 activity for the proliferation of IECs, aberrant activation of mTORC1 likely participates in intestinal tumorigenesis. Indeed, tumorigenesis driven by mutation of Apc in the mouse intestine was shown to require the activity of mTORC1. Mutation of the mTOR gene appears to be infrequent in CRC, however, with aberrant activation of the mTOR pathway being instead attributable to mutation of genes in the upstream PI3K-Akt signaling pathway. For instance, mutations in the kinase domain of the α catalytic subunit of PI3K (PIK3CA) tend to arise late in tumorigenesis and have been identified in 32% of CRC tumors. Defects of the PTEN gene, which encodes a phosphatase for phosphatidylinositol 3,4,5-trisphosphate that opposes PI3K activity, have also been identified in CRC.

5 | CROSS-TALK BETWEEN SFKs, Ras, AND mTOR SIGNALING PATHWAYS AND Wnt-β-catenin SIGNALING IN IECs

Activation of SFKs, Ras, and mTOR likely suppresses the Wnt-β-catenin signaling pathway in crypts of the intestine and thereby regulates ISCs as well as the differentiation of TA cells into mature IECs, such as absorptive enterocytes and goblet cells (Figure 4). However, the molecular mechanisms by which these signaling molecules inhibit the Wnt-β-catenin pathway remain unclear. The binding of Wnt ligands to the Frizzled-LRP5/6 receptor complex inhibits the activity of glycosgen synthase kinase 3β (Gsk3β), which mediates the phosphorylation of β-catenin at serine 37 or serine 33 and thereby triggers its degradation by the ubiquitin-proteasome system. The nonphosphorylated form of β-catenin thus accumulates, translocates to the nucleus, and acts as a transcriptional cofactor for T-cell factor (Tcf), resulting in transcription of Wnt-β-catenin target genes. Activation of the Ras-MEK-MAPK pathway suppresses Wnt target gene expression in IECs by increasing the abundance of the ~50-kDa (shorter) isoforms of Tcf4 (Tcf M/S), which are thought to be transcriptionally inactive and to inhibit activation of Wnt target genes. The kinase Akt, which is an upstream activator of mTORC1, inhibits the activity of Gsk3β by mediating its phosphorylation at serine 9. Feedback inhibition of Akt as a result of aberrant activation of mTORC1 in melanocytes was shown to reduce the level of Gsk3β phosphorylation at serine 9 and thereby to promote Wnt3β and Gsk3β activity, resulting in suppression of Wnt target gene expression. Indeed, phosphorylation of both Akt and Gsk3β was also attenuated by IEC-specific activation of mTORC1 in IECs. An increase in the activity of Gsk3β could therefore promote β-catenin degradation and consequent downregulation of Wnt target gene expression. The activation of mTORC1 has also been shown to suppress the amount of the Wnt receptor Frizzled at the cell surface, resulting in inhibition of Wnt signaling. Further investigation will be required to clarify the molecular mechanisms by which SFKs, Ras, and mTOR inhibit Wnt-β-catenin signaling in IECs.

6 | CONCLUSION

Activation or inactivation of SFKs, Ras, or mTORC1 by gene targeting in mice as well as studies of intestinal organoids have recently uncovered roles for these signaling molecules that act downstream of growth factor RPTKs or integrins in the regulation of ISC maintenance as well as of the proliferation and differentiation of IECs from their progenitors. These signaling molecules have also been implicated in the opposition of Wnt-β-catenin signaling with homeostatic regulation of normal IEC turnover, although the molecular basis for such counterregulation remains to be fully elucidated. Given that Wnt ligand such as Wnt3 is predominantly produced and secreted by Paneth cells, the gradient of Wnt concentration from the bottom to the upper region of the crypt is likely important for both maintenance of stemness of ISCs at the bottom and the promotion of TA or mature IEC proliferation at the upper region, respectively. Interestingly, inhibition of Wnt signaling promotes the activation of MAPK in the crypts, suggesting that the gradient of Wnt in the crypt likely regulates the activation...
of MAPK in the upper part of the crypt and promotion of TA cell proliferation.\textsuperscript{46} Counterregulation by the Wnt-β-catenin pathway and the growth factor RPTK-activated signaling pathways (such as Ras or PI3K signaling) could also provide a basis for the development of new treatment strategies for CRC. Simultaneous targeting of these two signaling pathways could thus result in more effective eradication of cancer derived from the intestinal epithelium (Figure 4).

ACKNOWLEDGMENTS

The work in the authors’ laboratory was supported by a Grant-in-Aid for Scientific Research (A) from the Japan Society for the Promotion of Science (JSPS) grant number (18H04032), a research grant from Mitsui Foundation, and a research grant from Yakult Bio-Science Foundation.

CONFLICT OF INTEREST

T.M. has received research funding from the Mitsui Foundation and Yakult Bio-Science Foundation. The other authors declare no conflict of interest.

ORCID

Takashi Matozaki https://orcid.org/0000-0002-4393-8416
Takenori Kotani https://orcid.org/0000-0001-8571-4210
Yoji Murata https://orcid.org/0000-0002-9576-7030
Yasuyuki Saito https://orcid.org/0000-0002-9291-1383

REFERENCES

1. Barker N. Adult intestinal stem cells: critical drivers of epithelial homeostasis and regeneration. Nat Rev Mol Cell Biol. 2014;15:19-33.
2. Clevers H. The intestinal crypt, a prototype stem cell compartment. Cell. 2013;154:274-284.
3. Sato T, van Es JH, Snippert HJ, et al. Paneth cells constitute the niche for Lgr5 stem cells in intestinal crypts. Nature. 2011;469:415-418.
4. Gregoriew F, Clevers H. Wnt signaling in the intestinal epithelium: from endoderm to cancer. Genev. Dev. 2005;19:877-890.
5. Fodde R, Smits R, Clevers H APC, signal transduction and genetic instability in colorectal cancer. Nat Rev Cancer. 2001;1:55-67.
6. Sato T, Vries RG, Snippert HJ, et al. Single Lgr5 stem cells build crypt-villus structures in vitro without a mesenchymal niche. Nature. 2009;459:262-265.
7. Okada M, Regulation of the SRC family kinases by Csk. Int J Biol Sci. 2012;8:1385-1397.
8. Cordero JB, Ridgway RA, Valeri N, et al. c-Src drives intestinal regeneration and transformation. EMBO J. 2014;33:1474-1491.
9. Imada S, Murata Y, Kotani T, et al. Role of Src family kinases in regulation of intestinal epithelial homeostasis. Mol Cell Biol. 2016;36:2811-2823.
10. Galbiati F, Razani B, Lisanti MP. Emerging themes in lipid rafts and caveolae. Cell. 2001;106:403-411.
11. Liang X, Nazarian A, Erdjument-Bromage H, Bornmann W, Tempst P, Resh MD. Heterogeneous fatty acylation of Src family kinases with polyunsaturated fatty acids regulates raft localization and signal transduction. J Biol Chem. 2001;276:30987-30994.
12. Nada S, Okada M, MacAuley A, Cooper JA, Nakagawa H. Cloning of a complementary DNA for a protein-tyrosine kinase that specifically phosphorylates a negative regulatory site of p60\textsuperscript{src}. Nature. 1991;351:69-72.
but independently of nutritional status or Notch regulation. J Cell Sci. 2013;126:3884-3892.

36. Cao J, Tyburczy ME, Moss J, Darling TN, Widlund HR, Kwiatkowski DJ. Tuberous sclerosis complex inactivation disrupts melanogenesis via mTORC1 activation. J Clin Invest. 2017;127:349-364.

37. Francipane MG, Lagasse E. mTOR pathway in colorectal cancer: an update. Oncotarget. 2014;5:49-66.

38. Prossomariti A, Piazz G, Alquati C, Ricciardello L. Are Wnt/β-Catenin and PI3K/AKT/mTORC1 distinct pathways in colorectal cancer? Cell Mol Gastroenterol Hepatol. 2020;10:491-506.

39. Faller WJ, Jackson TJ, Knight JR, et al. mTORC1-mediated translational elongation limits intestinal tumour initiation and growth. Nature. 2015;517:497-500.

40. Fujishita T, Aoki K, Lane HA, Aoki M, Taketo MM. Inhibition of the mTORC1 pathway suppresses intestinal polyp formation and reduces mortality in Apc^{−/−} mice. Proc Natl Acad Sci USA. 2008;105:13544-13549.

41. Samuels Y, Wang Z, Bardelli A, et al. High frequency of mutations of the PIK3CA gene in human cancers. Science. 2004;304:554.

42. Zhou XP, Loukola A, Salovaara R, et al. PTEN mutational spectra, expression levels, and subcellular localization in microsatellite stable and unstable colorectal cancers. Am J Pathol. 2002;161:439-447.

43. Liu C, Li Y, Semenov M, et al. Control of β-catenin phosphorylation/degradation by a dual-kinase mechanism. Cell. 2002;108:837-847.

44. Nusse R, Clevers H. Wnt/β-catenin signaling, disease, and emerging therapeutic modalities. Cell. 2017;169:985-999.

45. Cross DA, Alessi DR, Cohen P, Andjelkovich M, Hemmings BA. Inhibition of glycogen synthase kinase-3 by insulin mediated by protein kinase B. Nature. 1995;378:785-789.

46. Kabiri Z, Greicius G, Zaribafzadeh H, Hemmerich A, Counter CM, Virshup DM. Wnt signaling suppresses MAPK-driven proliferation of intestinal stem cells. J Clin Invest. 2018;128:3806-3812.

How to cite this article: Matozaki T, Kotani T, Murata Y, Saito Y. Roles of Src family kinase, Ras, and mTOR signaling in intestinal epithelial homeostasis and tumorigenesis. Cancer Sci. 2021;112:16-21. https://doi.org/10.1111/cas.14702