Reference Cluster Normalization Improves Detection of Frontotemporal Lobar Degeneration by Means of FDG-PET

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Introduction

In brain imaging by means of positron emission tomography (PET) and single-photon emission computed tomography (SPECT), scaling of tracer uptake to a reference region is in most cases essential for analyses of non-quantitative data. An ideal reference region should not be affected by brain pathology and should be easy to image/analyse. The choice of the appropriate reference region is especially problematic in subjects with neurodegenerative disorders who show early metabolic and perfusion deficits [1–5]. Using statistical parametric mapping (SPM), we have recently proposed a data-driven method for normalization of [18F] fluorodeoxyglucose (FDG) uptake in cases with preclinical and manifest Alzheimer’s disease (AD) dementia [6]. As compared with traditional intensity normalization to an a priori defined reference region, the reference cluster (RC) is defined by a contrast showing areas with increased activity in patients relative to controls after global mean normalization. This RC approach proved to detect AD-related hypometabolism in a more sensitive manner than traditional ROI-based normalizations [6]. Originally developed on clinical image data, the method was soon validated...
in simulation studies and extended to perfusion studies in Parkinson’s disease (PD), another common neurodegenerative disorder [7–9].

To be applicable in clinical settings, any diagnostic test must be robust to variability in clinical presentation/assessments and methodological factors. To this end, we performed a bi-center study with a cross-validation design in frontotemporal lobar degeneration (FTLD). As compared to AD and PD, FTLD is characterized by a substantially higher heterogeneity in respect to histopathological, clinical, and imaging presentation [10–12].

FDG-PET plays a well-established role in assisting early detection and differentiation of this severe neurodegenerative disorder [13–25], after AD the second most common cause of presenile dementia [26]. Furthermore, as compared to previous single-center applications of the RC normalization in AD and PD [6,27,28], here we assess performance of the method in cross-center settings. I.e., a RC obtained at one center is applied for data normalization from another center. Results of this data-driven approach are then compared with normalization to common reference regions.

In this work, we examine the impact of intensity normalization to cerebellum (CBL), primary sensorimotor cortex (SMC), cerebral global mean (CGM) and to RC on the accuracy of FDG-PET to detect FTLD-specific metabolic deficits and to discriminate between patients with mild FTLD and healthy subjects.

Methods

Subjects

Patients were retrospectively identified from a database of subjects from the memory clinic of the Department of Psychiatry at the Technische Universität München (hereafter referred to as center 1) and from the Clinic of Cognitive Neurology at the University of Leipzig (hereafter referred to as center 2). In both centers all patients were diagnosed according to Neary diagnostic criteria of FTLD [29]. The diagnosis was based on the information from a thorough neurological and psychiatric examination, informant interview, routine blood sampling, MRI, and FDG-PET imaging. Only patients with a Mini-Mental-State Examination (MMSE) score >21 were included. For two patients at center 2 MMSE was not available. In this case, a clinical dementia rating (CDR) score of less or equal to 1 was applied as an inclusion criterion [30]. Thus, we aimed to include patients with a mild disease severity only. Exclusion criteria were evidence for lesions due to stroke, traumatic head injury, brain tumor or inflammatory diseases on structural MRI. Using these criteria, 41 patients were included at center 1 and 11 at center 2.

The control group at center 1 (n = 42) consisted of elderly individuals without relevant psychiatric or neurological symptoms; subjects were only included if they did not report subjective memory complaints and if they did not show evidence for cognitive impairment.

The control group at center 2 (n = 13) included subjects who visited the Clinic of Cognitive Neurology at the University of Leipzig with subjective cognitive complaints, which were not objectively confirmed by a comprehensive neuropsychological and clinical evaluation.

This study was carried out in accordance with the latest version of the Declaration of Helsinki after the consent procedures had been approved by the local ethics committees of the University of Leipzig and of the medical faculty at the Technische Universität München. Written informed consent was obtained from all control subjects, all patients at center 2 and from most patients at center 1. Some patients at center 1 only gave oral informed consent for participation in the study as PET examination in patients was part of routine diagnostic work-up. Such a study-specific written consent was not obtained because all the procedures are included in a diagnostic work-up for suspected neurodegenerative disorder. Irrespective of this, all patients or their proxy gave a written consent for the PET examination. Additionally the process was documented by standard hospital documentation including an indication for the diagnostic work-up (due to suspected neurodegenerative disorder). The standard written (in German) consent includes the following items: 1) Aim of the study/PET examination, 2) Contraindications to the PET examination, 3) Procedure of the PET examination, 4) Expenses and allowance, 5) Data protection, 6) Personal benefits, 7) Risks, complications and side-effects, 8) Insurance, 9) Responsible persons and consent to
Data acquisition

Table 1. Hypometabolism in patients with frontotemporal lobar degeneration compared to control subjects in center 1 (left) and 2 (right) after intensity normalization to different reference regions, in particular primary sensorimotor cortex (SMC), cerebral global mean (CGM), cerebellum (CBL), and reference cluster (RC).

| Reference Region | Center 1 | Center 2 |
|------------------|----------|----------|
|                  | Cluster extent | Peak t-value | Cluster extent | Peak t-value |
| CGM              | 36806     | 8.7       | n.s.           | n.s.        |
| SMC              | 46215     | 8.9       | 1652           | 5.0         |
| Cerebellum       | 108383    | 9.5       | 1281           | 4.3         |
| RC               | 167801    | 10.9      | 7216           | 5.1         |

The cluster extent is represented by the sum of all clusters (in voxels) which exceeded an threshold $p<0.001$ (uncorrected) at voxel level and $p<0.05$ (family-wise error corrected) at cluster level. CGM: cerebral global mean, SMC: primary sensorimotor cortex, RC: reference cluster.

n.s. not significant.
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acquired for each subject starting 30 min post injection. At center 2, data were acquired in 2D mode. A sequence of three frames (10 min each) was acquired for each subject starting 30 min post injection. Sixty-three slices were collected at both centers with an axial resolution of 5 mm full width at half maximum (FWHM) and in-plane resolution of 4.6 mm. After correction for attenuation, scatter, decay and scanner-specific dead time at both centers, images were reconstructed by filtered back-projection using a Hamm-filter (center 1: cutoff frequency 0.5 cycles/projection element; center 2: 4.9 mm FWHM). Slices obtained had a resolution of $128 \times 128$ voxels with an edge length of 2.425 mm at center 1 and 2.45 mm at center 2.

Data pre-processing

Statistical Parametric Mapping software (SPM8), http://www.fil.ion.ucl.ac.uk/spm/) implemented in Matlab 7.11 (MathWorks Inc., Sherborn, MA) was used for image processing and statistical analyses. After spatial realignment a mean image of the three frames was calculated for each subject. The image sets were spatially normalized using an in-house tracer-, scanner- and age-specific brain PET template (in MNI space) based on all available images, separately in each center. Normalized images with a voxel size of $2 \times 2 \times 2$ mm were then smoothed with a Gaussian kernel of 12 mm FWHM. Regional value extraction for subsequent intensity normalization to CBL and to SMC was performed the automatic anatomical labeling atlas [AAL, [31]]. All AAL regions belonging to cerebellar hemispheres and SMC were used as a single volume-of-interest for intensity normalization to CBL and SMC, respectively. The value of CGM was calculated using the global calculation function with default settings, as implemented in SPM [32,33].

Voxel-based analyses

To obtain a representative FTLD-specific RC, we used data from center 1, as substantially more data sets were available. By analogy with the original work [6], we computed a contrast representing relative increases in the whole patient group ($n = 41$) compared to the control sample ($n = 42$), using normalization to CGM. As we had no experience with such a contrast in FTLD, no a priori probability threshold was chosen. Instead, we started with a threshold of $p<0.05$ FWE-corrected at voxel level and increased it until most of the resultant cluster volume could be clearly assigned to one specific anatomical region. Under the threshold of $p<0.000001$ FWE-corrected at voxel level we obtained a single cluster that projected to the cerebellum. Of note, its volume was still sufficiently large, covering 4321 voxels. The cluster encompassed the medial and anterior parts of the cerebellar hemispheres as well as the cerebellar vermis (Figure 1). A small part of the cluster volume included the lower parts of the occipital cortex.

Further, we specified conventional contrasts representing relative decreases in the patient group ($n = 41$) relative to the control sample ($n = 42$). To evaluate the impact of normalization onto detection of FTLD specific hypometabolism, image intensity in these analyses was normalized to CGM, CBL, SMC, and RC as obtained above. To examine performance of normalization to RC in an independent sample, the same analyses were performed for data from center 2. Age and gender were included as nuisance variables in all analyses. A statistical threshold of $p<0.001$ uncorrected at voxel level and $p<0.05$ FWE-corrected at cluster level was applied in all analyses. Extent threshold was set at $k>100$ contiguous voxels. Detected glucose hypometabolism was compared between the normalization procedures in terms of cluster size and maximum t-value. Anatomical labeling of significant

Figure 2. Regions showing a significant decrease in glucose metabolism in frontotemporal lobar degeneration compared to control subjects in center 1 (left) and 2 (right) after intensity normalization to different reference regions, in particular primary sensorimotor cortex (SMC), cerebral global mean (CGM), cerebellum (CBL), and reference cluster (RC).

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Table 2. Hypometabolism in patients with frontotemporal degeneration after intensity normalization to different reference regions.

| Reference Region | Center 1 | Center 2 |
|------------------|----------|----------|
|                  | Cluster extent | Peak t-value | Cluster extent | Peak t-value |
| CGM              | 36806     | 8.7       | n.s.           | n.s.        |
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The cluster extent is represented by the sum of all clusters (in voxels) which exceeded an threshold $p<0.001$ (uncorrected) at voxel level and $p<0.05$ (family-wise error corrected) at cluster level. CGM: cerebral global mean, SMC: primary sensorimotor cortex, RC: reference cluster.

n.s. not significant.
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participate. As patients were studied with PET as a part of diagnostic work-up, items 4,6,8 and were not applicable. We confirm that all potential participants who declined to participate or otherwise did not participate were eligible for treatment and were not disadvantaged in any other way by not participating in the study. Demographic characteristics of participants are presented in Table 1.

Data acquisition

Scans were acquired under standard resting conditions with eyes closed at center 1 and in dimmed ambient light with eyes open at center 2, using a Siemens ECAT EXACT HR+ PET scanner (CTI, Knoxville, TN, USA).

At center 1, the acquisitions were performed in 3D mode with a total axial field of view of 15.32 cm and no interplane dead space. A sequence of three frames with a duration of 10, 5 and 5 min was

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clustering was performed using AAL [31] implemented in the WFU PickAtlas tool [34,35].

Logistic regression analyses

Accuracy for discrimination between FTLD patients and control subjects was calculated using logistic regressions with split-half cross-validation separately for each center and normalization. Thereby, the diagnostic labels were used as the dependent variable. As independent variable we used individual mean FDG-uptake values extracted after each type of intensity normalization. All mean values were extracted from the hypometabolic pattern detected after CBL normalization (Figure 2) as a kind of gold standard. Indeed, this reference region has been widely applied in FDG-PET imaging of FTLD and has been shown to be the most sensitive approach for diagnostic purposes [2]. Additionally, we extracted mean values from an overlap of hypometabolic patterns detected using all reference regions at center 1. This pattern included bilateral frontal and anterior temporal regions. To obtain accuracy distributions, the split-half cross-validation procedure was repeated 5000 times by randomly assigning patients and control subjects to training and testing datasets and calculating prediction accuracies for the data not used for training. Accuracy, sensitivity and specificity distributions obtained for FTLD patients and control subjects after normalization to each reference region were compared to each other using t-tests for independent samples and applying a significance threshold of \( p < 0.05 \) (Bonferroni corrected for multiple comparisons). Logistic regressions and t-tests comparing the obtained accuracies were implemented using functions provided by Matlab.

Demographic characteristics

Group comparisons for age were performed by conducting Student’s t-tests with a two-sided significance threshold of \( p < 0.05 \) implemented in Matlab 7.11. Group differences regarding gender were evaluated using a chi-square test for independence using the commercial software package SPSS 17.0 (http://www.spss.com/statistics/).

Results

Demographic results

The chi-square test for independence did not reveal any statistical differences in gender between the groups at center 2 \( \chi^2(2) = 2.4; p = 0.628 \). Gender distribution between FTLD patients and control subjects differed significantly at center 1 \( \chi^2(21) = 7.82; p = 0.005 \). There was a minor but significant difference in age between control subjects and FTLD patients at center 2 \( t(22) = -3.19; p = 0.004 \), which was taken generally into account in the following analyses by including age as a covariate. No significant difference in age was observed at center 1 \( t(81) = 0.98; p = 0.328 \).

Voxel-based analyses

The comparison of FTLD patients and control subjects at center 1 revealed a significantly lower relative FDG uptake in bilateral frontal, anterior temporal, cingulate cortices, caudate nucleus and thalamus for CGM and SMC (Figure 2 and Table 2). Decreased relative FDG-uptake was more pronounced, both in respect to extent and height, after intensity normalization to CBL and RC, sparing only occipital, cerebellar and vermis regions after normalization to CBL while only the vermis after normalization to RC. For data from center 2, effects were generally smaller. Left-hemispheric lower relative tracer uptake was detected in FTLD patients in inferior and middle frontal gyrus, pars triangularis, anterior superior, middle, inferior temporal gyrus and precentral gyrus after normalization to CBL. Normalization to RC additionally revealed lower relative FDG uptake in left fusiform gyrus, supramarginal gyrus, frontal inferior operculum, angular gyrus, inferior orbital gyrus, insula, Rolandic operculum, parahippocampal and inferior parietal regions. Normalization to SMC revealed lower relative tracer uptake only in left middle and superior frontal gyrus, parts of the precentral gyrus, pars triangularis and operculum. No significant cluster was detected in patients from center 2 after normalization to CGM.

For data from center 1, the opposite contrast investigating increases in glucose uptake in patients resulted in appearance of significantly higher FDG uptake in CBL, primary sensorimotor, occipital and parietal regions of patients after normalization to SMC and CGM (Figure 3). After normalization to CBL higher FDG uptake was only detected in vermis and parts of CBL. No significant clusters were observed after normalization to RC. For data from center 2, no significant clusters were observed in FTLD patients after any type of normalization.

Logistic regression

In analyses with mean uptake values extracted from the hypometabolic pattern detected using CBL as a reference region normalization to RC provided significantly higher (\( p < 0.001 \)) mean accuracies for discrimination between FTLD patients and control subjects in both cohorts (center 1: 90.4%; center 2: 97.3%) as compared to all other reference regions (Figure 4). Normalization to CBL performed second best in both centers (center 1: 86.2%; center 2: 91.9%). CGM (90.1%) was superior to SMC (86.9%) at center 2 whilst the opposite was the case at center 1 (CGM: 75%; SMC: 76.1%). The differences between all normalization procedures were highly significant (\( p < 0.001 \)). When comparing sensitivities and specificities between different normalization procedures, again all comparisons with the exception of the comparison of sensitivities for CGM and SMC...
normalizations at center 1 (p = 1.0) were highly significant (p < 0.001) showing the same order as the total accuracies. At both centers, sensitivities were significantly lower (p < 0.001) than specificities after normalization to all reference regions with the exception of normalization to RC at center 1 (p = 1.0).

Logistic regressions calculated in center 1 and center 2 based on the overlap of all clusters detected in the corresponding center revealed highly similar accuracy distributions (center 1: RC: 88.5%; CBL: 86.8%; CGM: 82.4%; SMC: 78.4%; center 2: RC: 88.7%; CBL: 78.3%; CGM: 81.8%; SMC: 79.6%) with the only difference that CGM was now superior to SMC at center 2 (Figure 4). All differences were again highly significant (p < 0.001). Similarly, the obtained sensitivities and specificities differed significantly (p < 0.001) between all normalization procedures except for the comparison of sensitivities obtained after CGM and SMC normalization (p = 1.0) and of specificities for the comparison of CBL and SMC normalization (p = 1.0), both at center 2. At both centers, all sensitivities were significantly lower than the specificities (p < 0.001).

Discussion

In the present study, we compared performance of data-driven RC normalization [6] with that of normalization to CGM, CBL, and SMC in mild FTLD. The major finding is that RC normalization allowed for a more accurate detection of early FTLD related metabolic deficits than normalization to any common reference region. Furthermore, this superior performance held its standing after cross-validation, i.e. when a representative RC cluster was applied to image data from another center that were acquired with a different protocol.

The proposed method of intensity normalization is in essence based on the assumption that CGM metabolism is significantly decreased in patients with neurodegenerative disorders. Although, to the best of our knowledge, this has not been explicitly reported in FTLD, this is rather expected, taking into account the presence of rather extensive regional metabolic deficits already at mild disease stages [20,36]. When comparing patients vs. healthy subjects, intensity normalization to CGM may lead to two unwanted effects in the patient group: underestimation of true regional hypometabolism or -perfusion and detection of apparent (but false) regional hypermetabolism or -perfusion due solely to overcorrection of global variation [37,38]. Here we show that both effects, in an extensive manner, indeed take place in FTLD, even at a mild disease stage. Moreover, under the same probability threshold, the extent of apparent hypermetabolism is substantially larger than that of true hypometabolism (data not shown). This observation suggests that so-called global normalization alone should not be applied in the comparison of FTLD patients vs. control subjects, even at mild disease stages. However, this procedure may be beneficial in detecting regions that are relatively preserved in a given neurodegenerative disease [6,9,27]. Here we demonstrate that intensity scaling to such regions increases statistical power in detecting FTLD-related regional deficits. Moreover, this gain appears substantial enough to significantly increase accuracy of discrimination between patients with mild FTLD and healthy elderly subjects.

While CGM and CBL have been most widely used for scaling of tracer uptake in FTLD (e.g., CGM: [2,19,39,40]; cerebellum:
[2, 17, 41, 42], the SMC was included, because it was shown to be a valuable reference area in AD [1]. As expected, CBL was superior to CGM, but it was also superior to SMC. This finding is in line with a quantitative FDG-PET study on FTLD [4] as well as with evidence from histopathological literature [43, 44]. The fact that our RC was also localized within the CBL further supports the view that CBL is a reference area of choice in FTLD. Our data indicate, however, that parts of the cerebellar cortex are probably also hypometabolic. Thus, the RC identifies the cerebellar subregion that is most conserved, and therefore provides the least biased normalization. This interpretation also explains why the RC performs better than employing an a priori CBL region in its entirety. In particular, we found that FDG uptake in the anterior-medial parts of the cerebellar hemispheres, as well as in the vermis, was most preserved in our patients with mild FTLD. Such a pattern might be a result of cerebellar diaschisis with a specific degeneration of cerebropontine-cerebellar pathways [45]. It should be noted, however, that FTLD is a heterogeneous group of diseases, so there might be subtype-specific patterns of preserved glucose metabolism.

In a second step, we asked if the derived RC is representative enough to improve detection of hypometabolism in an independent cohort from another center. Indeed, RC normalization was still significantly superior to other commonly used reference regions. Of note, the image data from the independent cohort were acquired using different acquisition protocols. Furthermore, as compared to control subjects at center 1, the control group at center 2 consisted of individuals with subjective cognitive complaints. Indeed, the latter situation is even closer to routine clinical practice, where a differentiation between such subjects and patients with a suspected neurodegenerative disorder might be really challenging. Nonetheless, RC normalization significantly improved discrimination between these control subjects and patients with mild FTLD. These findings suggest that the derived FTLD-specific RC is rather robust to between-center variability in data acquisition and clinical assessments. Thus, the cluster can be used in independent centers, both on a group and single subject basis.

This study has some limitations. Firstly, the sample size from the center 2 was relatively small. However, given the observation of a similar similarity of the RC approach in a second substantially bigger sample we feel confident that the results reflect a true difference between normalization procedures. However, the substantially smaller sample size in center 2 might explain the lower degree of hypometabolism detected in this FTLD cohort. Secondly, according to standards of clinical work, our PET data were acquired without arterial or venous blood sampling. As we analyzed only relative FDG-uptake, we cannot exclude the presence of true hypermetabolism in our patients with FTLD. To the best of our knowledge, however, there have been no reports describing increases in resting state regional metabolism in subjects with FTLD. Nonetheless, caution is needed when applying the RC approach in psychiatric or neurological disorders where hypermetabolism or hyperperfusion might be part of the disease process. Furthermore, accuracy values provided by RC normalization at center 1 might be overoptimistic, as the RC was obtained on the basis of the same image data. Yet, application of the same cluster to an independent cohort of subjects from center 2 resulted in a similar accuracy increase. Of note, this was also the case when normalizing image data from center 1 to a RC obtained at center 2 (data not shown). Although there was a substantial overlap between two clusters, we chose the one from center 1 to be applied in major analyses, as it is based on substantially more cases. Furthermore, we did not apply correction for partial volume effects. However, in our previous work on the same study sample from center 2 we have shown that partial volume effect correction has only little effect on the comparison of different normalization procedures [2]. Finally, and perhaps most importantly, the methodology presented in this paper is not generalized to the clinical practice in which an expert in nuclear medicine encounters with a large spectrum of conditions/disorders on a single-subject basis. This essential issue should be addressed in future studies with a different design. While we fully admit the indispensable value of visual scan reading in clinical settings, an increasing role of automated image-based classification procedures that can be applied to analyses of large-scale image databases in a user-independent manner should be recognized and noted [46, 47]. Logistic regression analyses such as those applied in the present study are a common step in such classification algorithms. However, it is important to note that the purpose of our study was not to develop or to establish a classification algorithm but rather to the examine how classification accuracy of automated classification procedures on the same data is influenced by the choice of reference region for intensity normalization.

In summary, we found that data-driven RC normalization improves detection of glucose hypometabolism in mild FTLD. Such an improvement appears substantial enough to increase accuracy of discrimination between FTLD patients and healthy subjects, also in an independent cohort. The established FTLD-specific cluster can be used for intensity normalization at independent imaging centers. The cluster as binary image is available for free download at http://www.unil.ch/webdav/site/juergen/wRe/Cls/Munich_41Pat42Kon_FWE0.000001.nii.

Author Contributions

Analysed the data: JD IY. Contributed reagents/materials/analysis tools: JD RP SF HR JDS HO ES AD AF RF AK KM OS MLS IY. Wrote the paper: JD RP SF HR JDS BD HO ES AD AF RF AK KM OS MLS IY.

References

1. Yakushev I, Landovg C, Buchholz HG, Fellgiebel A, Hammers A, et al. (2008) Choice of reference area in studies of Alzheimer's disease using positron emission tomography with fluorodeoxyglucose-F18. Psychiatry Res 164: 143–153.
2. Dukart J, Mueller K, Horstmann A, Vogt B, Frisch S, et al. (2010) Differential effects of global and cerebellar normalization on detection and differentiation of dementia in FDG-PET studies. Neuroimage 49: 1490–1495.
3. Minoshima S, Frey KA, Foster NL, Kuhl DE (1995) Preserved pontine glucose metabolism in Alzheimer disease: a reference region for functional brain image (PET) analysis. J Comput Assist Tomogr 19: 541–547.
4. Ishii K, Sasaki M, Yamaji S, Sakamoto S, Kitagaki H, et al. (1998) Paradoxical hippocampal perfusion in mild-to-moderate Alzheimer’s disease. J Nucl Med 39: 295–298.
5. Schroeter ML, Stein T, Madsowski N, Neumann J (2009) Neural correlates of Alzheimer’s disease and mild cognitive impairment: a systematic and quantitative meta-analysis involving 1531 patients. Neuroimage 47: 1196–1206.
6. Yakushev I, Hammers A, Fellgiebel A, Schmidmann I, Scheurich A, et al. (2009) SPM-based count normalization provides excellent discrimination of mild Alzheimer's disease and amnestic mild cognitive impairment from healthy aging. Neuroimage 44: 43–50.
7. Borglumard P, Cumming P, Østergaard K, Gjedde A, Rodell A, et al. (2012) Cerebral oxygen metabolism in patients with early Parkinson's disease. J Neurol Sci 313: 123–128.
8. Barfhammar P, Hansen SJ, Eggert G, Chakravarty M, Yang K, et al. (2012) Glucose metabolism in small subcortical structures in Parkinson's disease. Acta Neurol Scand 125: 303–310.
9. Borglumard P, Aanerud J, Gjedde A (2009) Data-driven intensity normalization of PET group comparison studies is superior to global mean normalization. Neuroimage 46: 981–988.
10. Ibach B, Poljsak S, Marienhaegen J, Sommer M, Manner P, et al. (2004) Contrasting metabolic impairment in frontotemporal degeneration and early onset Alzheimer’s disease. Neuroimage 23: 739–745.

11. Sampathu DM, Neumann M, Kwong LK, Ghio TT, Micsenyi M, et al. (2006) Pathological heterogeneity of frontotemporal lobar degeneration with ubiquitin-positive inclusions delineated by ubiquitin immunohistochemistry and novel monoclonal antibodies. Am J Pathol 169: 1343–1352.

12. Seelaar H, Rohrer JD, Pijnenburg YA, Fox NC, van Swieten JC (2011) Clinical, genetic and pathological heterogeneity of frontotemporal dementia: a review. J Neurol Neurosurg Psychiatry 82: 476–480.

13. Foster NL, Heidebrink JH, Clark CM, Jagust WJ, Arnold SE, et al. (2007) FDG-PET improves accuracy in distinguishing frontotemporal dementia and Alzheimer’s disease. Brain 130: 2616–2635.

14. Kipps CM, Hodges JR, Fryer TD, Nestor PJ (2009) Combined magnetic resonance imaging and positron emission tomography brain imaging in behavioural variant frontotemporal degeneration: refining the clinical phenotype. Brain 132: 2366–2376.

15. Piguet O, Hornberger M, Mioshi E, Hodges JR (2011) Behavioural-variant frontotemporal dementia: diagnosis, clinical staging, and management. Lancet Neurol 10: 162–172.

16. Schroeter ML, Neumann J (2011) Combined Imaging Markers Dissociate Alzheimer’s Disease and Frontotemporal Lobar Degeneration – An ALE Meta-Analysis. Front Aging Neurosci 3: 10.

17. Dokic J, Mueller K, Horstmann A, Barthel H, Muller HE, et al. (2011) Combined Evaluation of FDG-PET and MRI Improves Detection and Differentiation of Dementia. PLoS One 6: e18111.

18. Mosconi L, Tsui WH, Herholz K, Pupi A, Drzezga A, et al. (2006) Multicenter standardized 18F-FDG PET diagnosis of mild cognitive impairment, Alzheimer’s disease, and other dementias. J Nucl Med 49: 386–398.

19. Diehl-Schmid J, Grimmer T, Drzezga A, Bornschein S, Riemenschneider M, et al. (2007) Decline of cerebral glucose metabolism in frontotemporal dementia: a longitudinal 18F-FDG-PET-study. Neurobiol Aging 28: 42–50.

20. Diehl J, Grimmer T, Drzezga A, Riemenschneider M, Forstl H, et al. (2004) Cerebral metabolic patterns at early stages of frontotemporal dementia and semantic dementia. A PET study. Neurobiol Aging 25: 1051–1056.

21. Sadequi N, Foster NL, Wang AY, Minoshima S, Lieberman AP, et al. (2008) Automatic classification of Alzheimer’s disease vs. Frontotemporal dementia: A spatial decision tree approach with FDG-PET. 2008 IEEE International Symposium on Biomedical Imaging: From Nano to Macro, Vols 1–4: 408–411.

22. Jeong Y, Cho SS, Park JM, Kang SJ, Lee JS, et al. (2005) 18F-FDG PET findings in frontotemporal dementia: an SPM analysis of 29 patients. J Nucl Med 46: 233–239.

23. Schroeter ML, Raczk a K, Neumann J, Yves von Cranmon D (2007) Towards a nosology for frontotemporal lobar degeneration-a meta-analysis involving 267 subjects. Neuroimage 36: 497–510.

24. Schroeter ML, Raczk a K, Neumann J, von Cranmon DY (2008) Neural networks in frontotemporal dementia – a meta-analysis. Neurobiol Aging 29: 418–426. 41. Santens P, De Bleecker J, Goethals P, Strijckmans K, Lemahieu I, et al. (2001) Differential regional cerebral uptake of [18F]fluorodexyglucose in Alzheimer’s disease and frontotemporal dementia at initial diagnosis. Eur Neurol 45: 19–27.

25. Dukart J, Mueller K, Barthel H, Villinger A, Sahri O, et al. (2012) Meta-analysis based SVM classification enables accurate detection of Alzheimer’s disease across different clinical centers using FDG-PET and MRI. Psychiatry Res: 10.1016/j.psychres.2012.1004.1007.

26. Harvey RJ, Skelton-Robinson M, Rossouw MN (2003) The prevalence and causes of dementia in people under the age of 65 years. J Neurol Neurosurg Psychiatry 74: 1206–1209.

27. Berti V, Polito C, Borghammer P, Ramat S, Mosconi L, et al. (2012) Alternative normalization methods demonstrate widespread cortical hypometabolism in untreated de novo Parkinson’s disease. Q J Nucl Med Mol Imaging 56: 299–308.

28. Neary D, Snowden JS, Gustafson L, Passant U, Stuss D, et al. (1998) Frontotemporal lobar degeneration: a consensus on clinical diagnostic criteria. Neurology 51: 1546–1554.

29. Perneczky R, Wagenerf ll S, Konnossa K, Grimmer T, Diehl J, et al. (2006) Mapping scores onto stages: mini-mental state examination and clinical dementia rating. Am J Geriatr Psychiatry 14: 139–144.

30. Tzourio-Mazoyer N, Landeau B, Papathanassiou D, Crivello F, Etard O, et al. (2002) Automated anatomical labeling of activations in SPM using a macroscopic anatomical parcellation of the MNI MRI single-subject brain. Neuroimage 13: 236–248.

31. Mosconi L, Perani D, Sorbi S, Herholz K, Nacmias B, et al. (2004) MCI conversion to dementia and the APOE genotype: a prediction study with FDG-PET. Neurology 63: 2332–2340.

32. Samurakz M, Masunari I, Chen WP, Yajima K, Yamasue H, et al. (2007) Partial volume-effect corrected FDG PET and grey matter volume loss in patients with mild Alzheimer’s disease. Eur J Nucl Med Mol Imaging 34: 1658–1669.

33. Malbich JA, Laurieri PJ, Budrete JD (2004) Prenatal gyrus discrepancy in electronic versions of the Talairach atlas. Neuroimage 21: 450–455.

34. Malbich JA, Laurieri PJ, Kraft RA, Budrete JD (2003) An automated method for neuroanatomic and cytoarchitectonic atlas-based interrogation of MRI data sets. Neuroimage 19: 1233–1239.

35. Silverman DH (2004) Brain 18F-FDG PET in the diagnosis of neurodegenerative dementia: comparison with perfusion SPECT and with clinical evaluations lacking nuclear imaging. J Nucl Med 45: 594–607.

36. Aguirre GK, Zarahn E, D’Esposito M (1998) The inferential impact of global signal covariates in functional neuroimaging analyses. Neuroimage 8: 302–306.

37. Maldjian JA, Laurieri PJ, Kraft RA, Budrete JD (2003) An automated method for neuroanatomic and cytoarchitectonic atlas-based interrogation of MRI data sets. Neuroimage 19: 1233–1239.

38. Poljansky S, Ibach B, Hirschberger B, Manner P, Klinenmann H, et al. (2011) A visual [18F]FDG-PET rating scale for the differential diagnosis of frontotemporal lobar degeneration. Eur Arch Psychiatry Clin Neurosci 261: 433–446.

39. Santens P, De Bleecker J, Goethals P, Strijckmans K, Lemahieu I, et al. (2001) Differential regional cerebral uptake of [18F]fluorodeoxyglucose in Alzheimer’s disease and frontotemporal dementia. Eur Neurol 45: 19–27.

40. Rostomian AH, Madison C, Rabinovici GD, Jagust WJ (2011) Early 11C-PiB PET improves accuracy in distinguishing frontotemporal dementia and Alzheimer’s disease from normal aging. J Nucl Med 52: 173–179.

41. Neumann M, Sampathu DM, Kwong LK, Traux AC, Micsenyi MC, et al. (2006) Ubiquitinated TDP-43 in frontotemporal lobar degeneration and amyotrophic lateral sclerosis. Science 314: 130–133.

42. Mukherjee O, Pastor P, Cairns NJ, Chakraverty S, Kauwe JS, et al. (2006) HDDD2 is a familial frontotemporal lobar degeneration with ubiquitin-positive, tau-negative inclusions caused by a missense mutation in the signal peptide of progranulin. Ann Neurol 60: 314–322.

43. Tien RD, Ashdown BC (1992) Crossed cerebellar diaschisis and crossed cerebellar atrophy: correlation of MR findings, clinical symptoms, and subcortical atrophies in 26 patients. AJR Am J Roentgenol 158: 1155–1159.

44. Tzourio-Mazoyer N, Landeau B, Papathanassiou D, Crivello F, Etard O, et al. (2002) Automated anatomical labeling of activations in SPM using a macroscopic anatomical parcellation of the MNI MRI single-subject brain. Neuroimage 13: 236–248.

45. Tzourio-Mazoyer N, Landeau B, Papathanassiou D, Crivello F, Etard O, et al. (2002) Automated anatomical labeling of activations in SPM using a macroscopic anatomical parcellation of the MNI MRI single-subject brain. Neuroimage 13: 236–248.