CASE REPORT

Diagnosis of an ectopic adrenocorticotropic hormone-secreting bronchial carcinoid by somatostatin receptor scintigraphy

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INTRODUCTION

Ectopic adrenocorticotropic hormone (ACTH) syndrome accounts for 10%–20% of the cases of ACTH-dependent Cushing’s syndrome (1). Ectopic ACTH secretion typically results from an occult, slow-growing bronchial carcinoid tumor. The diagnosis of these relatively small tumors can be difficult with conventional imaging procedures, such as computerized tomography (CT) or magnetic resonance imaging (MRI) (2). Herein, we present a case of ectopic ACTH syndrome arising from a bronchial carcinoid tumor that was diagnosed with somatostatin receptor scintigraphy (SRS).

CASE DESCRIPTION

A 25-year-old Caucasian woman was admitted for oligomenorrhea and hirsutism. A physical examination revealed central obesity, a “buffalo hump”, acne, and severe hirsutism (Ferriman-Gallwey score of 18). After overnight dexamethasone suppression testing (DST), her serum cortisol level was 14.4 μg/dL. She was hospitalized with a presumptive diagnosis of Cushing’s syndrome.

Her diurnal cortisol secretion rhythm was found to be impaired; her serum cortisol level at midnight was 27.1 μg/dL. Low-dose DST was also consistent with this diagnosis, with an unsuppressed serum cortisol level of 4.2 μg/dL. Her basal ACTH and cortisol levels were 67 pg/mL and 15.2 μg/dL, respectively.

With a refined presumptive diagnosis of ACTH-dependent Cushing’s syndrome, we performed overnight high-dose DST with 8 mg dexamethasone. Her serum cortisol level was 10 μg/dL, revealing a lack of suppression. An MRI revealed a 3-mm microadenoma of the hypophysis. Because adenomas are incidentally discovered in up to 10% of the normal population, IPSS is advised for patients with ACTH-dependent Cushing’s syndrome and a hypophyseal adenoma smaller than 6 mm, particularly when test results conflict (4).

IPSS can demonstrate the ratio of the ACTH level in the central sinus relative to the peripheral sinus. A central/peripheral ratio of 2 before the administration of corticotropin-releasing hormone and a ratio of ≥3 after its administration strongly suggests Cushing’s disease (5). However, a positive result is highly suggestive of Cushing’s disease, false-negative results might be more common than previously appreciated (2%–4%). In cases of a negative response, clinicians should perform a careful search for an ectopic source (6).

No potential conflict of interest was reported.
Table 1 - Adrenocorticotropic hormone (ACTH) levels in the inferior petrosal sinus before and after the infusion of corticotropin-releasing hormone.

| ACTH Level (pg/mL)            | Periphery | Right  | Left  |
|-------------------------------|-----------|--------|-------|
| Time before infusion          |           |        |       |
| 5 minutes                     | 141       | 163    | 151   |
| 2 minutes                     | 144       | 159    | 164   |
| Time after infusion           |           |        |       |
| 2 minutes                     | 148       | 160    | 151   |
| 5 minutes                     | 149       | 166    | 181   |
| 10 minutes                    | 160       | 175    | 172   |

Figure 1 - Computerized tomography of the bronchial carcinoid.

Figure 2 - Somatostatin receptor scintigraphy of the bronchial carcinoid.
frequently bronchial carcinoid tumors. Locating these tumors can be challenging; they are relatively small and slow-growing, and conventional imaging studies, such as CT and MRI, identify the tumor in only 50% of cases (2,4,7). The tumor can remain occult long after the diagnosis of Cushing’s syndrome (4).

Functional imaging studies, such as PET and SRS, are complementary imaging tools for revealing the presence of carcinoids (6). SRS might be superior to PET in detecting bronchial carcinoids. Although PET can detect highly active proliferative tumors, bronchial carcinoids usually have a low proliferation index and are slow growing, quite small lesions (7,8). However, the success of SRS depends only on the presence of somatostatin receptors, which have been identified on many cells of neuroendocrine origin (8). The radiolabelled somatostatin analog octreotide can bind to somatostatin receptors 2 and 5 with high affinity (8). Bronchial carcinoids express both receptors. Nevertheless, these tumors may show heterogeneity in the degree of somatostatin receptor expression (8).

The diagnostic sensitivity of SRS is approximately 25%–73% for the detection of an ectopic source of ACTH secretion (9). Ilias and colleagues reported that the sensitivity of SRS is only approximately 49%, and it is insufficient for lesions that are not present on CT or MRI (1). Zemskova and colleagues reported a 57% sensitivity and a 79% positive predictive value for SRS for detecting ectopic ACTH-producing tumors (7).

The use of a single imaging tool may not be sufficient to diagnose ectopic ACTH-secreting tumors; conventional and functional imaging studies should be used in combination when needed (2,6-9). In our case, the ectopic lesion was shown on CT but not on PET. SRS confirmed the lesion shown on CT, and the tumor was extracted. Suppression by DST revealed biochemical recruitment, and ACTH staining of the excised tissue confirmed the ectopic tumor postoperatively.

In conclusion, in cases of ectopic ACTH syndrome, the exact location of the lesion is needed for a curative approach. Conventional imaging studies may not always show the lesion. The use of somatostatin receptor scintigraphy in addition to conventional imaging may help localize lesions.

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AUTHOR CONTRIBUTIONS

Anaforoğlu I drafted and approved the final version of the manuscript. Ersoy K drafted the manuscript. Asik M and Karyagar S substantially contributed to the acquisition and interpretation of data. Algun E critically reviewed the manuscript. Each author participated sufficiently to take public responsibility for appropriate portions of the content. All authors read and approved the final version of the manuscript.

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