Glutathione S-transferase π: a potential role in antitumor therapy

Shu-Chen Dong1,*
Huan-Huan Sha1,*
Xiao-Yue Xu1
Tian-Mu Hu2
Rui Lou1
Huizi Li1
Jian-Zhong Wu1
Chen Dan1
Jifeng Feng1

1Jiangsu Cancer Hospital and Jiangsu Institute of Cancer Research and Nanjing Medical University Affiliated Cancer Hospital, Nanjing 210009, China; 2Department of Biological Science, Purdue University, West Lafayette, IN, USA

*These authors contributed equally to this work

Abstract: Glutathione S-transferase π (GSTπ) is a Phase II metabolic enzyme that is an important facilitator of cellular detoxification. Traditional dogma asserts that GSTπ functions to catalyze glutathione (GSH)-substrate conjunction to preserve the macromolecule upon exposure to oxidative stress, thus defending cells against various toxic compounds. Over the past 20 years, abnormal GSTπ expression has been linked to the occurrence of tumor resistance to chemotherapy drugs, demonstrating that this enzyme possesses functions beyond metabolism. This revelation reveals exciting possibilities in the realm of drug discovery, as GSTπ inhibitors and its prodrugs offer a feasible strategy in designing anticancer drugs with the primary purpose of reversing tumor resistance. In connection with the authors’ current research, we provide a review on the biological function of GSTπ and current developments in GSTπ-targeting drugs, as well as the prospects of future strategies.

Keywords: tumor resistance, glutathione S-transferase π, drug treatment

Introduction

In addition to combating a variety of noxious substances from the external environment, cell-detoxification mechanisms are capable of resisting the deleterious effects of certain endogenous substances (eg, ROS, a product generated from normal cellular metabolism), in order to maintain physiological homeostasis. Drug metabolism represents an important component of cellular detoxification and involves two enzymes: Phase I and Phase II drug-metabolism enzymes. The Glutathione S-transferase π (GST) family of enzymes is a group of typical Phase II detoxification enzymes and is found in many prokaryotes and eukaryotes.1 Of these enzymes, GSTπ catalyzes the conjunction between GSH and its electrophilic substrates upon exposure to damaging free radicals. Besides metabolite detoxification, GSTπ also exhibits ligand-binding properties that initiate cellular apoptosis when triggered by cellular stress.2 Further research has also demonstrated that GSTπ is expressed abundantly in tumor cells and associated closely to carcinogenesis, tumor formation, and chemotherapy resistance.3,4 Moreover, experiments involving drug-resistant cell lines have also demonstrated increased GSTπ expression.5,6 In multidrug-resistant HL60/VCR acute myelogenous leukemia cells, GSTπ is found to be expressed at higher levels than HL60.7 GSTπ is involved in facilitating tumor resistance and suppressing apoptosis in tumor cells via two mechanisms. First, GSTπ weakens the efficacy of chemotherapy drugs by promoting their in vitro extrusion. Second, GSTπ also functions as an MAPK-pathway inhibitor to prevent tumor-cell apoptosis.

A variety of anticancer drugs based on these principles have been synthesized in efforts to improve their therapeutic indices and reverse tumor resistance. Drugs that work through the GST system include GSTπ inhibitors and their respective prodrugs.

Correspondence: Jifeng Feng; Chen Dan
Jiangsu Cancer Hospital, 42 Baiziting Street, Nanjing, Jiangsu 210009, China
Email jifeng_feng@163.com; ccece20110622@126.com
The former work by exerting high GSTπ-inhibitory activity, while the latter comprise inactive compounds designed to target tumor tissue locally by undergoing GSTπ catalysis in the tumor to release cytotoxic metabolites. Development of therapies targeting GSTπ is a major field of research. As such, we believe that there is a need for more detailed studies outlining the diverse biological functions of GSTπ, in order to assist drug discovery and unlock more exciting possibilities in the realm of tumor treatments.

Structure

GSTs consist of the following three superfamilies: cytoplasmic (cGSTs), mitochondrial (mGSTs), and microsomal (membrane-associated proteins in eicosanoid and glutathione metabolism [MAPEG]). Among these families, cGSTs are the most complex and most closely linked to the development of human diseases. The cGSTs are divided into seven subtypes according to similarities in amino-acid sequence, different structure of genes, and immunological cross-reactivity. These subtypes are α, π, μ, θ, ω, σ, and δ. Among them, GSTα is highly expressed in many normal cells. However, recent studies have shown that GSTα also takes part in promoting multidrug resistance in p53-mutated lung cancer cells. GSTμ has been found to be able to act synergistically with MRP1 to decrease the effects of vincristine treatment. GSTπ is widespread in tumor cells, and is intricately involved with cellular carcinogenesis, tumor formation, and tumor-drug resistance. Current evidence supports the role of cGSTs in facilitating multidrug resistance across different types of tumors.

GSTπ is the most frequently and extensively studied of all the GSTs. Its encoding gene is located on chromosome 11 and is composed of seven exons. In humans, GSTπ often consists of two identical dimer subunits, with each subunit consisting of 210 amino acids and two binding sites, the G-site and the H-site (Figure 1). Different G- and H-site locations in the amino-acid residue of different GSTs exert different functions. GSTπ is able specifically to bind to GSH or GSH analogs via the G-site, which catalyzes the interaction between GST amino-acid residues with GSH sulfhydryl and conventional electrophilic residues at the H-sites to promote catalytic action. Therefore, G-site modification often guides the development of specific GSTπ inhibitors.

Biological function

GSTπ in metabolite detoxification and antioxidation

The classical view holds that as a dimeric isoenzyme, GSTπ can conjugate GSH with substrate molecules in efforts to promote clearance of active ionic substances. However, tumor cells also utilize GSTπ to form a GSH–X complex between antitumor drugs and GSH, proceeding to excrete the complex out of the cell by Pgp and MRPs. Pgp, encoded by MDRI, is often found to be highly expressed in tumor cells. What is more, GSTπ and Pgp or MRPs (eg, MRP2) are synergistic in driving the development of multidrug resistance in tumor cells (Figure 2). Studies have documented high GSTπ expression in various tumor cells, such as cancers of the gastrointestinal tract, pancreas, breast, liver, lymphoma, as well as melanoma.

Recent literature has characterized GSH and other related metabolic enzymes as vital in protecting cells from ROS through oxidation and reduction (redox) mechanisms. GSH carries a cysteine residue with an active thiol group and is responsible for maintaining thiol equilibrium. Meanwhile, it can also modulate the activities of many signaling molecules and redox-sensitive transcription factors through S-glutathionylation, a form of posttranslational modification that combines cysteine residues with GSH. GSTπ serves as a general S-glutathionylase enzyme and promotes S-glutathionylation. Its enzymatic function is based on two aspects: its catalytic activity and the auto-S-glutathionylation of GSTπ by itself on Cys47 and Cys10, both of which disturb the subsequent interaction with c-Jun NH2-terminal kinase (JNK), resulting in the formation of a GSTπ multimer (Figure 3). Besides GSTπ, other members of the GSH-redox system, such as glutamate cysteine ligase, glutathione peroxidase, and glutathione reductase, also play significant roles in this process.

GSTπ in regulation of MAPK pathway

Besides metabolite detoxification, GSTπ also exhibits ligand-binding properties that allow the enzyme to interact covalently and noncovalently with compounds, resulting in inhibition of conjugation activity. GSTπ can induce cellular apoptosis in the setting of cellular stress by activating MAPK, MKK4, downstream JNK-signal components, and p38 kinase. Normal cells have low basal JNK activity to maintain optimal cellular growth conditions. However, in the presence
of oxidative or nitrosative stress, GSTπ can form homodimers that alter the reduced states of cysteine residues in its structure, resulting in JNK dissociation from the GSTπ–JNK heterocomplex and causing subsequent activation of the c-Jun protein. Ultimately, these series of reactions will activate the apoptotic pathways33,34 (Figure 4).

Further research indicates that GSTπ can influence the MAPK pathway both through JNK and TRAF2 modulation.

![Figure 2](image1.png)

**Figure 2** Involvement of GSTπ in the detoxification of exogenous and endogenous substrates.  
**Note:** In the process, Pgp or MRPs (eg, MRP2) give assistance to GSTπ to excrete the complex out of the cell.  
**Abbreviation:** GST, glutathione S-transferase.

![Figure 3](image2.png)

**Figure 3** The process of S-glutathionylation.  
**Notes:** In this process, low-pka cysteine-residue proteins are targeted upon exposure to nitrosative or oxidative stress. The cysteine residues are oxidized to produce protein sulfenic (P–OH) and sulfenic (P–OOH) acids. GSH functions as a thiol donor to facilitate GSTπ-mediated S-glutathionylation, protecting the target protein from further damage. This reaction is also influenced by enzymes involved in deglutathionylation, such as sulfiredoxin (Srx), glutaredoxin (Grx), and thioredoxin (Trx).  
**Abbreviation:** GST, glutathione S-transferase.
In-depth analysis of biological information tools has revealed that the TRAF family is strongly linked to GSTπ. Of all the TRAF members, TRAF2 is expressed most abundantly and has been subjected to intense research. Following the activation of TNFR through TNFα binding, TRAF2 is recruited to the plasma membrane, resulting in the production of ROS. ROS generation leads to oxidation of the ASK1 inhibitor thioredoxin, separating and activating ASK1 from the inactive ASK1–Trx complex. ASK1 goes on to bind to TRAF2, which in turn activates downstream signaling cascades, including the MKK3/4/6–p38 and MKK4/7–JNK signaling pathways (Figure 4). Further evidence from steady-state fluorescence analysis confirms that direct binding between TRAF2 and GSTπ also exists. In tumor cells, GSTπ can suppress JNK activity and block the interaction between TRAF2 and ASK1 to inhibit tumor-cell apoptosis. Consequently, regarding the formation of multidrug resistance in tumor cells, besides acting as a detoxification enzyme through drug excretion to decrease pharmacological efficacy, GSTπ can also act as an MAPK-pathway inhibitor to improve tumor-cell survival. GSTπ also represents a scaffold protein to unite different members across signaling pathways.

**Gene variants and polymorphisms**

The study of pharmacogenomics in recent decades has proved that genetic polymorphism of drug-metabolizing enzymes is an important mediating factor in determining individual drug responses. Genetic polymorphism is due to a single-nucleotide mutation in the genomic sequence that fundamentally changes how a person responds to chemotherapeutic drugs. Polymorphisms affecting GSTπ have been documented to exert significant modulatory effects on the biological cascade of carcinogenesis and have been discussed vigorously in the literature.
The human GSTP1 gene exists as two functionally different variants, with both variants having documented nucleotide transitions from isoleucine to valine at codon 105 (A→G) and alanine to Val at 114 (C→T). This results in four alleles of GSTP1: wild-type GSTP1*A (Ile105 + Ala114), GSTP1*B (Val105 + Ala114), GSTP1*C (Val105 + Val114), and GSTP1*D (Ile105 + Val114).49 Besides GSTP1, cytosolic GSTs demonstrate clinically significant gene polymorphism (Table 1). These changes occur in active sites, leading to a decrease in encoded protein activity, decreased excretion of foreign substances, and suboptimal catalytic efficiency. On the other hand, reducing the rate of drug excretion has the benefit of improving an individual’s sensitivity to chemotherapeutic drugs, enhancing their curative effects.50–52

There are a great number of reviews that have summarized the associations between individual GSTπ variability and the drug sensitivity of malignant tumors.53–55 However, data are scarce regarding the relationship between GSTπ and clinical response to chemotherapy. Khrunin et al showed that 104 patients with ovarian cancer with mutant-type GSTπ possessed longer progression-free survival compared to wild-type GSTπ.56 These results highlight the possibility that

### Table 1 Genetic polymorphisms of cytosolic GSTs

| Class | Chromosome | Gene | Exon count | Alleles | Nucleotide variability | Reference(s) |
|-------|------------|------|------------|---------|------------------------|---------------|
| α     | 6p12.2     | GSTA1| 7          | GSTA1*A | Wild-type              | 116           |
|       |            |      |            | GSTA1*B | Promoter point mutation|               |
|       |            |      |            | GSTA2*A | Pro110;Ser112;Lys196;Glu210 | 117, 118 |
|       |            |      |            | GSTA2*B | Pro110, Ser112, Lys196, Ala210 |               |
|       |            |      |            | GSTA2*C | Pro110;Thr112;Lys196;Glu210 |               |
|       |            |      |            | GSTA2*D | Pro110;Ser112;Asn196;Glu210 |               |
|       |            |      |            | GSTA2*E | Ser110;Ser112;Lys196;Glu210 |               |
| μ     | 1p13.3     | GSTM1| 8          | GSTM1*A | Lys173                 | 119, 120      |
|       |            |      |            | GSTM1*B | Asn173                 |               |
|       |            |      |            | GSTM1 null | Gene deletion         |               |
|       |            |      |            | GSTM1*1-2 | Duplication           |               |
|       |            |      |            | GSTM3*A | Wild-type              | 121           |
|       |            |      |            | GSTM3*B | Pro110;Ser112;Lys196;Glu210 |               |
|       |            |      |            | GSTM4*A | Wild-type              | 122, 123      |
|       |            |      |            | GSTM4*B | Pro110;Ser112;Lys196;Glu210 |               |
|       |            |      |            | GSTM4*C | Wild-type              |               |
|       |            |      |            | GSTM4*D | Wild-type              |               |
| π     | 11q13.2    | GSTP1| 7          | GSTP1*A | Ile105;Ala114          | 124–128       |
|       |            |      |            | GSTP1*B | Val105;Ala114          |               |
|       |            |      |            | GSTP1*C | Val105;Val114          |               |
|       |            |      |            | GSTP1*D | Ile105;Val114          |               |
| θ     | 22q11.23   | GSTT1| 6          | GSTT1*A | Thr104                 | 124           |
|       |            |      |            | GSTT1*B | Pro104                 | 129           |
|       |            |      |            | GSTT1*null | Gene deletion         |               |
|       |            |      |            | GSTT2*A | Met139                 | 130, 131      |
|       |            |      |            | GSTT2*B | Ile139                 |               |
| ω     | 10q25.1    | GSTO1| 7          | GSTO1*A | Ala140;Glu155          | 132, 133      |
|       |            |      |            | GSTO1*B | Ala140                 |               |
|       |            |      |            | GSTO1*C | Asp140;Glu155          |               |
|       |            |      |            | GSTO1*D | Asp140                 |               |
|       |            |      |            | GSTO2*A | Asn142                 | 134, 135      |
|       |            |      |            | GSTO2*B | Asp142                 |               |
| δ     | 14q24.3    | GSTZ1| 12         | GSTZ1*A | Lys32;Arg42;Thr82     | 136–138       |
|       |            |      |            | GSTZ1*B | Lys32;Gly42;Thr82     |               |
|       |            |      |            | GSTZ1*C | Glu32;Gly42;Thr82     |               |
|       |            |      |            | GSTZ1*D | Glu32;Gly42;Met82     |               |

**Abbreviation:** GST: Glutathione S-transferase.
genetic variation may have significant effects on susceptibility toward cancer. In future, precise genetic polymorphic screening tests may play a more central role in determining chemotherapeutic treatment regimens for patients.

**GST\(\pi\) inhibitors**

GST\(\pi\) inhibitors reverse tumor resistance by means of suppressing GST\(\pi\) activity and improving the chemotherapeutic drug sensitivity of tumor cells. Ethacrynic acid (EA) is a classic GST\(\pi\) inhibitor.\(^5^7\) However, due to its spefific pharmacological properties in targeting GST\(\pi\), the newer GST\(\pi\) inhibitors TLK117/TLK199 and NBDHEX may prove to be more promising. Antitumor agents targeting GST\(\pi\) in context are listed in Table 2.

**EA and its analogs**

EA represents the first clinical application of GST\(\pi\) inhibitors. Previously, it was widely used for decades as a diuretic in clinical research. EA works to halt GST\(\pi\) activity through a number of mechanisms. First, it is able to bind directly to substrate-binding sites of isozymes to inhibit GST\(\pi\). Second, it is able to induce the combination of \(\alpha,\beta\)-unsaturated ketones and GSH through the nucleophilic addition reaction, depleting GSH and reducing the amount of GSH available to combine with chemotherapeutic agents, thus producing an overall GST\(\pi\)-inhibitory effect by sensitizing a cell to chemotherapeutic agents.\(^5^7\) However, the clinical applications of EA have been limited, due to its diuretic properties and lack of enzyme specificity, with long-term intake possibly risking water and salt imbalance.\(^1^8\) Zhao et al attempted to modify EA using thiazole derivatives of uric acid to strengthen its GST\(\pi\)-inhibitory effects. The team demonstrated that these derivatives had higher GST\(\pi\)-inhibitory activity in comparison to unmodified EA when administered to acute myeloid leukemia parental cells (HL60).\(^6^8\) In addition, the combination of EA and GSH has also been proven to possess superior inhibitory activity over EA alone and is able functionally to inhibit many GST isoenzymes. However, this compound also possesses limited clinical viability, given its tendency toward dissociation by \(\gamma\)-glutamyltransferase.\(^5^9\) Burg et al synthesized modified peptidomimetic glutathione analogs of these EA–GSH compounds, which were hypothesized to be stabler against peptidase-mediated dissolution. Unfortunately, these analogs instead reduced its GST\(\pi\)-inhibitory activity, despite demonstrating increased resilience toward \(\gamma\)-glutamyltransferase compared to the unmodified EA–GSH compounds.\(^5^7\) Taken together, EA and its analogs still represent novel avenues of research in the search for more efficacious antitumor drugs.

**TLK117 and TLK199**

Telintra (ezatiostat hydrochloride, TER199, TLK199) is a small-peptide, glutathione-analog molecule and was developed by Telik. Upon entering the body, TLK199 undergoes esterase hydrolysis, which releases TLK117, its activated form that has anti-GST\(\pi\) activity. TLK199 is able to enhance the potency of various antineoplastic agents against various tumor cell lines. The agent is also able to inhibit MRAP1 and prevent the combination of GST\(\pi\) and JNK, resulting in high JNK production that triggers tumor-cell apoptosis.\(^6^0\)

Furthermore, clinical studies have found TLK199 to be able to promote the maturation of hematopoietic progenitor cells, influence cancer-cell death, and inhibit myeloproliferative diseases.\(^6^1\)–\(^6^3\) In 2013, TLK199 successfully passed a US Food and Drug Administration audit and was approved to treat low-to intermediate-risk myelodysplastic syndrome. Long-term observation studies have highlighted the ability of TLK199 to enhance bone-marrow maturation and cellularity.\(^6^4\)

**NBDHEX and its analogs**

NBDHEX (6-[7-nitro-2,1,3-benzoazadiazol-4-ythio] hexanol) is a recently developed compound designed as a
“mechanism-based inhibitor” that exerts potent effects on GST\(\pi\). Since its first reports by the Tor Vergata University of Rome,\textsuperscript{65} numerous preclinical studies have shown that NBDHEX exerts high GST\(\pi\)-inhibitory activity across a wide range of tumor types. Pasello et al reported that this agent effectively reversed cisplatin resistance in osteosarcoma, alluding toward potentially improved clinical outcomes when using a combination of NBDHEX and cisplatin.\textsuperscript{66} NBDHEX GST\(\pi\)-inhibitory effects have also been observed in HL60 cells and their chemotherapy-resistant phenotype HL60/DNR.\textsuperscript{67} Other cell lines that have demonstrated NBDHEX sensitivity include Ewing sarcoma,\textsuperscript{68} the human mesothelioma cell lines MPP89, MMB1, MSTO211H, and Mero48a,\textsuperscript{69} melanoma cell lines Me501 and A375,\textsuperscript{70} and the non-small-cell lung cancer cell line H69AR.\textsuperscript{70} Further research indicates that aside from inherent antimelanoma activity, NBDHEX also has the ability to enhance the function of temozolomide, with the two able to work synergistically to suppress tumor growth.\textsuperscript{71}

NBDHEX employs several methods in combating malignant cells. First, it is able to accumulate specifically in tumor cells, remaining unaffected by MRP while maintaining good cell-membrane permeability. Second, this agent can decompose the GST\(\pi\)-JNK complex and promote activation of the apoptosis pathway.\textsuperscript{42,65,67,70–72} Further research has also suggested that TRAF2 plays a key role in facilitating NBDHEX-mediated apoptosis. NBDHEX simultaneously activates JNK and TRAF2, interfering with the effect of GST\(\pi\) via two different pathways, leading to cell-cycle arrest and cell death.\textsuperscript{34} Intriguingly, researchers have provided further evidence to demonstrate that NBDHEX can also act as an autophagy inhibitor in tumor cells.\textsuperscript{40}

Despite NBDHEX’s promising anticancer activity, consideration should be given to the relatively low GST\(\pi\)-target selectivity and poor water-solubility.\textsuperscript{73} In efforts to counter this limitation, a study group designed, synthesized, and screened 40 new NBDHEX analogs.\textsuperscript{74} The group added one or two oxygen atoms on the hydroxyl chain of the NBD bone, forming two NBDHEX analogs, MC3181 and MC3165, that were able to display higher selectivity and better external activity by forming a stable \(\sigma\)-complex with the active site of GST\(\pi\). After extensive experiments, MC3181 was deemed the more promising compound of the two, due to it having a 50-fold rise in aqueous solubility and higher selectivity toward GST\(\pi\). This novel compound yielded positive results when administered to several distinct human melanoma cell lines, particularly when used in \(BRAF^V600E\)-mutation melanoma cells.\textsuperscript{75} Moreover, both intravenous and oral treatment of MC3181 in animals with different types of human melanoma xenografts resulted in astonishing curative effects and a satisfactory safety profile.\textsuperscript{75} In summary, we conclude that NBDHEX and its analogs may serve as potential treatment strategies in the management of patients with melanoma.

**Prodrugs of GST\(\pi\)**

Both conventional chemotherapeutic and targeted agents have well-established toxicity profiles, with a wide range of adverse effects, eg, bone-marrow suppression, gastrointestinal toxicity, immunosuppression, gastrointestinal toxicity, hepatotoxicity, and cardiotoxicity.\textsuperscript{76} GST\(\pi\) prodrugs appear to have a more favorable adverse-event profile, given that they are ingested as an inactive compound and undergo breakdown to release cytotoxic metabolites only in the presence of high concentrations of enzymes that occur in the proximity of tumor cells, thereby reducing collateral damage to healthy cells.\textsuperscript{2} Additionally, tumor cells generally have heightened expression of GST\(\pi\), providing ideal conditions for GST\(\pi\)-activated prodrugs. There are two primary classes of GST\(\pi\) prodrugs. The first of these are GSH or GSH derivatives, such as canfosfamide (Telcyta, TER286, TLK286), which have cytotoxic drug segments. When catalyzed by GST\(\pi\), the prodrug releases cytotoxic compounds. The other type, such as JS-K, consists of a similar structure, but without GST analogs. Upon being catalyzed by GST\(\pi\), it forms an intermediate with GSH and releases its cytotoxic drug segments. Currently, prodrugs under research comprise TLK286, purine analogs,\textsuperscript{77} sulfonamides,\textsuperscript{78} and brostallicin.

**TLK286**

L-\(\gamma\)-Glutamyl-3-(bis[bis(2-chloroethyl)amino-phosphinyl]oxy)ethylsulfonfyl-l-alanyl-2-phenyl-[2\(R\)]-glycine hydrochloride salt (TLK286) represents the most promising GST\(\pi\)-prodrug candidate. It can generate a GSH analog and a phosphorodiamidate, the latter an active, alkylating agent.\textsuperscript{81} Following activation, the former competitively inhibits molecules that stimulate drug resistance, while the nitrogen-mustard segment induces apoptotic activity by influencing the activities of MAPK, p38 kinase, JNK, MKK4, and caspase 3.\textsuperscript{53,79–81}

In vitro studies have revealed that the GSTP1-null cell lines show a different degree of resistance in response to TLK286 compared with GSTP1\(+\) cells. These differences were abrogated by cotransfecting these cells with GST\(\pi\). Similar findings were reflected in in vivo analyses with nude mice. These data support the rationale that tumors with
elevated GSTπ expression are more sensitive to the cytotoxic effects of TLK286.92

The promising mechanism of action of TLK286 and its positive preclinical data have sparked a series of clinical trials where the prodrugs have been applied alone or in combination with other standard chemotherapeutic agents. Completed Phase II and III clinical trials have indicated that the agent has a nonoverlapping toxicity profile and synergistic effects with carboplatin, paclitaxel, and anthracycline, has no cross-drug resistance and is well tolerated, with patients mostly reporting fatigue and nausea.83–88

A completed Phase I/IIA multicenter dose-ranging clinical trial that sought to assess the safety and efficacy of TLK286 found that the compound was highly efficacious. Patients who underwent TLK286 maintenance treatment experienced prolonged median survival of 16.8 months compared to the 8.8 months experienced by those who did not receive the agent.99 These clinical trials provide sound scientific evidence that supports the therapeutic efficacy of TLK286 in managing different types of malignancies.

Nitric oxide (NO) prodrugs
Another class of prodrugs are the NO prodrugs. These medications work by binding to intracellular GSTπ and undergoing GSTπ-mediated catalysis. This process releases NO molecules that go on to exert antitumor activity. NO is an ephemeral but pleiotropic molecule. It has been shown to have the capacity to affect several vital functions of the body.90 As such, this molecule has been investigated keenly for its role in carcinogenesis, tumor progression, invasion, angiogenesis, and other key biological processes.91 However, available experimental evidence suggests that NO is a double-edged sword when used to manage tumor diseases. NO itself is a source of cytotoxic molecules, and its deleterious effects are enhanced by its ability to concentrate locally around tumor cells and the tumor microenvironment.92 At modest concentrations, NO exerts a protumorigenic response that may benefit tumor growth and survival. Nevertheless, at fairly high concentrations, NO takes on antitumor-agent properties to accelerate tumor-cell death and to inhibit tumor-cell angiogenesis.93 Based on these observations, it is clear that NO has a role to play in combating tumor-cell resistance.

An example of an NO prodrug is O2-[(2,4-dinitrophenyl)1-[(4-ethoxycarbonyl)piperazin-1-yl]diazen-1-ium-1,2-diolate (JS-K), devised by Keefer et al from the National Cancer Institute.94 The antineoplastic properties of JS-K rest on two modes of action: first, it can combine with GSTπ and then be activated to release NO at a high concentration to directly kill tumor cells; second, it binds to cellular GST/GSH, depleting its intracellular content to weaken the efflux of chemotherapy drugs in tumor cells.95 Furthermore, other literature suggests that JS-K inhibits angiogenesis,96 induces cell apoptosis (a process related to PARP, caspase 8 and 9 cleavage, and cell differentiation),95,97 destroys double-stranded DNA,95,97–99 and is also able to interfere with the cell cycle and its respective signaling pathways.92,100–102 These mechanisms are highly codependent, and function in an integrated manner to exert antitumor effects.

Remarkably, flow-cytometry findings have shown that JS-K can improve the formation of acidic vesicle organelles, underscoring its ability to induce autophagy.102 Furthermore, electron-microscopy observations have indicated that JS-K induces autophagic death in cells.102 Nevertheless, JS-K was able to spare surrounding healthy mammary epithelial cells. JS-K has been shown to be effective in several types of cancers, of which leukemia and myeloma appear to be the most susceptible.92,96,100,103 In addition, it is also efficacious in the treatment of solid tumors, such as breast cancer,102,104 lung cancer,97,103,105 glioma,103,106 prostate cancer,107 kidney cancer,108 bladder cancer,109 colon cancer,110 and hepatocellular carcinoma.111 Furthermore, JS-K is tolerated well by healthy tissue. These data indicate that further investigation into JS-K as an alternative chemotherapeutic agent is much needed.102–104,109

It is worth noting that JS-K works synergistically with chemotherapy drugs, such as cytarabine,98 bortezomib,95 cisplatin, and arsenic.112 JS-K acts as a dose-sparing agent when used with typical chemotherapeutic agents and is able to alleviate the severity of adverse effects as a consequence. While JS-K has been shown to be advantageous in treating cancer, its clinical use has been hindered with reports of poor solubility. Structural modification of JS-K is able to prolong its half-life, and combining JS-K with special nanoparticles can greatly improve its solubility and stability,103,113 further improving its prospects for clinical applications. JS-K and many other NO prodrugs represent an innovative biological approach in the development of anticancer therapeutics.

Conclusion
Multidrug resistance to chemotherapy drugs is one of the main obstacles in human cancer chemotherapy and has prompted intense research into discovering novel and innovative mechanisms that can overcome this barrier.6,7,20 There is an intimate connection between abnormal GST expression and multidrug resistance, unequivocally implicating GSTπ,
a member of the GST family in tumor-drug resistance.\textsuperscript{4} GST\textpi inhibitors and prodrugs are crucial agents that can help reverse multidrug resistance in tumors and increase the therapeutic index of anticancer drugs, which collectively decreases the physical and economic burden of cancer patients. Nevertheless, while the rapid growth of research on development of medication based on GST\textpi inhibition has resulted in clinical studies on several compounds, the drugs that make it to commercial consumption are few. Realistically, fundamental issues that stand in the way of large-scale drug production include the vast number of biological GST family members, with each subtype having their own structural and functional differences, in addition to the existence of several genetic modifiers. The situation is further compounded by the presence of posttranslational modifying factors, such as kinase activities and S-glutathionylation. More in-depth research that clarifies the roles of these components of the GST-detoxification system are much needed, in order to produce compounds that have minimal side effects and high GST\textpi selectivity. While modulation of the GSH-antioxidant system has provided promising preclinical results, some of these compounds demonstrate unacceptable toxicity profiles (eg, buthionine sulfoximine). Having said that, GSH-based medication has also been successfully employed to protect against cisplatin induced nephrotoxicity.\textsuperscript{114,115} The ongoing development of chemical genomics, computer-aided drug design, and more extensive molecular and cellular biology research will serve to be extremely useful in contributing toward the preclinical and clinical development of more efficient GST\textpi-targeting drugs.

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**Disclosure**

The authors report no conflicts of interest in this work.

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