Double Negative (DN) B cells: A connecting bridge between rheumatic diseases and COVID-19?

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ABSTRACT

Double Negative (DN) B cells constitute a B cell population that lacks expression of immunoglobulin D and CD27 memory marker. These cells expand in elderly healthy individuals, but also accumulate prematurely in autoimmune and infectious diseases. COVID-19 is a pandemic infectious disease caused by SARS-CoV-2, a coronavirus that was first observed in Wuhan, China in December 2019. In its more severe cases, COVID-19 causes severe pneumonia and acute respiratory syndrome with high morbidity and mortality. Recent studies have revealed that the extrafollicular DN2 B cell subset, previously described in lupus patients, does also expand in severe and/or critical groups of COVID-19 patients. These DN2 cells correlate with disease severity and laboratory parameters of inflammation. However, their exact role and function in COVID-19 require to be further investigated.

In this review, we highlight the DN immune responses in both rheumatic diseases and COVID-19, and we point out the importance of clarifying DN’s role in the immunopathology of the aforementioned infection, as it could probably enable better management of rheumatic diseases during the pandemic. Of note, the symptomatology of COVID-19, as well as the potential outcome of death, have given rise to a worldwide concern and scare of exposition to SARS-CoV-2, especially among the rheumatological patients who believe to be at higher risk due to their immunological background and the immunosuppressive therapies. Nevertheless, there is no convincing evidence so far that these patients are truly at higher risk than others.

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SARS-CoV-2: Severe acute respiratory syndrome coronavirus 2
SCD: Scleroderma
SLE: Systemic lupus erythematosus

INTRODUCTION
Coronavirus disease 2019 (COVID-19) is an infectious disease caused by severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2), a single stranded RNA virus that was first observed in Wuhan, China in December 2019. Since then, the disease has emerged as a pandemic.1 In most individuals, the infection is considered as asymptomatic or paucisymptomatic, while in some cases it causes severe pneumonia, myocarditis, acute kidney injuries, hepatitis, acute respiratory distress syndrome (ARDS), multiorgan failure and even death.1 The most severe cases of COVID-19 are characterised by a cytokine storm which induces hyperinflammation, and lymphopenia, which affects both the innate and adaptive immune responses.2 Current evidence suggests that the hyperinflammatory syndrome in COVID-19 patients results from a dysregulated host innate immune response.3,4 Furthermore, the composition of the surviving T lymphocytes, as significant reductions in peripheral T cells have been reported in the severe cases of COVID-19,5 seems to be important for the establishment of the hyperinflammatory state.3,4 Having taken into account these facts, most studies so far have focused on the innate immune system responses and the phenotypic and/or molecular T-cell alterations.

On the other hand, the knowledge regarding the immunology of B cells in COVID-19 is limited. However, a few studies have turned their attention to humoral immune responses, and thus investigated the role and function of B cells in the infection.6-9 As in many other infectious diseases such as HIV, hepatitis B and malaria,9,10,11 alterations of the B cell compartment have also been reported in COVID-19.6-9,12 Interestingly, analyses of B cell populations revealed that extrafollicular double negative (DN) B cells, a characteristic of active systemic lupus erythematosus (SLE),13 are a prominent feature of severe COVID-19 as well.7,12,14,15 Moreover, the frequencies of these DN cells correlate with disease severity profiles and some clinical and laboratory parameters of the disease.6,8,14 Their exact role and function though in COVID-19 is not fully understood.

In this review, we discuss the overlapping DN immune responses in rheumatic diseases and severe COVID-19, and we point out the importance of investigating the role and the function of DN B cells in the immunopathology of the aforementioned infection, as it could probably enable better management of rheumatological patients during the pandemic.

THE POPULATION OF DOUBLE-NEGATIVE (DN) B CELLS
Double negative (DN) B cells, also known as atypical memory B cells, constitute a B cell population that lacks expression of immunoglobulin D and CD27 memory marker.16 The population is expanded in the elderly,16 but also represents a notable component of the B cell compartment in patients with autoimmune and/or chronic infectious diseases.17-20 In general, DN B cells seem to serve different functions in the context of different conditions. For instance, in cases of HIV and malaria, these cells have been characterised as having an exhausted phenotype,19,20 while in cases of SLE they are poised to become antibody-secreting cells (ASCs) in response to TLR7 ligands and the cytokines IFN-γ and IL-21,13 and are considered to be the direct source of ASCs and serum autoantibodies during disease’s flares.13,21-23 Such a differing and condition-dependent functionality of DN B cells, probably can be justified by the heterogeneity of the DN population.

Heterogeneity among DN B cells: subsets and DN relationship to other B cells arising in autoimmunity and infections
Various names have been used to describe CD19+IgD-CD27- cells, such as age-associated B cells (ABCs), atypical memory B cells, CD11c+ B cells and of course double negative B cells.16,18,20,21 Truth is that all these terms referred to B cell populations that share a number of features and are enriched in the peripheral blood of both elderly healthy individuals and people with chronic infectious diseases or inflammatory conditions, such as autoimmune disorders, obesity and cancer.17-20,24-26 A recent study however, based on transcriptomic and IgH repertoire analyses of B cells, questions the equivalence of all these populations.27 Here, we shall discuss the DN B cell population and its subsets, as well as the DN relationship to other B cells arising in autoimmunity and infections.

The various DN B cell subsets
In 2018, Jenks and colleagues from the Sanz group were the first to divide DN B cells into two subgroups, based on the expression of follicular homing marker CXCR5.13 Up to this time, the term DN B cells had been used to describe all B cell populations that lack expression of IgD and CD27. However, according to Jenks and his colleagues, the CXCR5+ subgroup, now known as DN1, is the DN sub-population that expands in elderly healthy individuals,16 while the CXCR5− subgroup constitutes the extrafollicular DN2 subset that is more marked in active SLE.15,17 Furthermore, transcription factor T-bet, whose expression in B cells is considered to be of high importance for the induction of autoimmunity and/or the control of chronic infections,8,28,29 is expressed (now obviously) in DN2 subset only.13
Of note, the frequencies of DN2 B cells in rheumatoid arthritis (RA), primary Sjögren’s syndrome (pSS) and systemic sclerosis (SSC) patients were not as much elevated as in SLE patients.13 Thus, the subset was presumed to be SLE specific. However, subsequently published data has implicated DN2 in the pathogenesis of common variable immunodeficiency (CVID) as well.30 Moreover, despite the fact that no classification into DN1 or DN2 has been achieved in incidents involving obese people, the DN B cells in obese individuals are believed to be predominantly DN2, as they secrete anti-self IgG antibodies.24,25

In addition, a cytoplasmic FOXO1 DN population - termed cytoFOX DN B cells - was identified in SLE via deep immunophenotyping.31 FOXO1, which is a transcription factor involved in B cell development,32 translocates from the nucleus to the cytoplasm in response to BCR ligation and presumably in an AKT-dependent manner, and gets inactivated.33 The cytoplasmic localization of the transcription factor seems to serve as a biomarker of SLE progression.31 Considering the fact that highest frequencies of cytoFOX DN B cells have been observed in African-American female SLE patients, made us believe -up to this day- that these cells belong to the DN2 subset,34 as DN2 cells predominate in African-American SLE patients (especially females).13 However, we still cannot rule out the possibility that cytoFOX DN cells are equivalent to another “new” DN subset in SLE. Taking into account all these data, we believe that DN B cells that have been reported in inflammatory conditions other than SLE and obesity, such as multiple sclerosis (MS),18 display a strong possibility to belong to the DN2 subset. However, we note that no confirmatory evidence exists thus far regarding this issue, as functional, transcriptional, phenotypic and other types of analyses need to be done. For instance, our knowledge regarding the presence of DN B cells in early-stage non-small cell lung cancer (NSCLC) is extremely limited at this time.29

As far as infectious diseases are concerned, DN B cells seem to play an inhibitory role in the humoral responses.19,20,35,36 Actually, these cells express a variety of inhibitor receptors, such as FcRL4, and exhibit many features of exhaustion.19,20,35,36 In contrast to DN2 B cells, which are very active in SLE13,21-23 Moreover, the absence of FcRL4 in SLE DN B cells,13,17 further suggests that DN B cells in infectious diseases differ from DN2 cells. However, once again, no confirmatory evidence exists regarding this issue. Thus, as long as we cannot classify DN B cells in chronic infections into either DN1 or DN2 cells, we prefer to refer to them as atypical or tissue-based memory B cells.

We have already mentioned that, apart from SLE cases, DN2 cells have also been observed in severe and/or critical cases of COVID-19.14,15 and the frequencies of these cells correlate with the severity of the disease, as well as with some clinical and laboratory parameters, such as CRP, IL-6, and ferritin levels.6,8,14 Intriguingly, T-bet expression in DN2 cells from COVID-19 patients, was reduced as compared with healthy individuals,8 suggesting that these cells are probably not a major driver of inflammation in this disease but is more likely to play an inhibitory role in the regulation of T cell responses. We have to note though that the two cohorts analysed for T-bet expression, consisted of three COVID-19 patients and four healthy controls, respectively.6 We believe that a new study including more individuals from both two cohorts needs to be carried out, in order for these findings to be further strengthened.

Even more interestingly, a previously unreported DN B cell subset was detected in COVID-19 patients. This novel subset, now known as DN3, is defined by the absence of CD21 and CD11c, and seems to be associated with extrafollicular immune responses, along with DN2 cells. Of note, in comparison to DN2, DN3 cells display a broader significant correlation pattern with laboratory features that have been related to critical COVID-19.3 In general though, the subset is considered as entirely new and not yet characterised.

The main features and functions of DN B cells in rheumatic diseases and COVID-19 shall be described, in more detail, in next sections of this review. Nevertheless, a table summarizing the topics discussed above is provided here (Table 1).

| Table 1 | | |
| --- | --- | --- |
| **DN relationship to age-associated B cells** (ABCs) | **ABCs** are a CD11c+T-bet+ B cell subset that expands continuously with age in healthy individuals.37 In cases of autoimmune and/or infectious diseases though, these cells accumulate prematurely and contribute to auto-antibody or anti-pathogen antibody production, respectively.42,47 Due to overlapping phenotypic markers, as well as similar activation requirements and functionality, ABCs and DN2 B cells are considered by many -including us- as relative populations.13,21,34,42,43 However, a recent study by Maul et al., in which a comparative transcriptomic analysis was performed, suggests that ABCs are distinct from other CD11c+ B cell populations, such as DN2 cells, as they display an elevated expression in multiple cytokines and chemokines, not detected as increased in the other CD11c− subsets.27 Although we do not call into question the results of this study, we believe that the discrimination of ABCs from other CD11c+ populations came up from the use of a strict immunophenotype for the ABCs. More specifically, Maul et al. used the term “ABCs” to refer to murine spleen cells, defined as B220+CD19−CD11b+, that may express CD11c, while in the literature, consensus has not yet been reached as to which phenotypic markers should be used to properly define these cells.24 The heterogeneity of the markers used in different publications for | | |
the immunophenotyping of ABCs had previously been summarised in a review article by Phalke and Marrack.\(^42\)

In that review, it becomes apparent that some scientific groups use the term “ABCs” to refer to human circulating B cells or even mouse circulating B cells that differ from the B220\(^+\)CD19\(^+\)CD11b\(^+\) murine spleen cells, but display a similar immunophenotype to DN2 cells (as defined in Table 1 of this review article).\(^42\) Taking into consideration all the above, we still believe that ABCs and DN2 cells are related to each other, especially when we treat the first as non-murine cells only, but also as human cells,\(^18,21,38\) that express CD11c and T-bet, and lack expression of CD21.\(^34\)

### EXTRAFOLLICULAR B CELL RESPONSES IN RHEUMATIC DISEASES AND COVID-19

Germinal centres (GCs) are sites within the secondary lymphoid organs, in which somatically mutated high-affinity memory B cells and long-lived plasma cells are generated,\(^44\) thus sustaining immune protection and rapid recall responses against re-encountered antigens. In this process, apart from the proliferating antigen-specific B cells, the participation of follicular helper T cells and specialised follicular dendritic cells is also required.\(^44\) Although the GC reactions have remained the focus of B cell research, extrafollicular B cell responses - development of antibodies outside of the B cell follicles, in the absence of GCs and independently of follicular T cells - are nowadays of great interest to many scientists.\(^45,46\)

Traditionally, the extrafollicular B cell responses, in contrast to GC reactions, are thought to produce antibodies with low somatic mutations and develop short-lived immunological memory, in a T cell - independent manner.\(^47,48\) Subsequent findings though, revealed that peripheral helper T cells (or pre-GC follicular T cells) may contribute, under persistent antigen stimulation combined with Type I IFN signalling and innate-receptor sensing,\(^13\) to the induction of extrafollicular responses in patients with rheumatic diseases and/or infections.\(^7,23,49,50\) In these cases, B cells produced are class-switch recombination competent and capable of sustaining long-lived immunological responses.\(^7,23,48\) However, a key question which remains unanswered yet is whether these B cells which lead to exaggerated extrafollicular responses, such as DN2 cells reported in SLE and COVID-19,\(^7,13\) are drivers of disease pathogenesis or just a consequence of the strong inflammation that occurs. The GC and extrafollicular pathways (both T-independent and T-dependent) are presented in Figure 1.

### DN2 B CELLS IN RHEUMATIC DISEASES AND COVID-19

DN2 B cells are a unique double-negative population (IgD\(^-\)CD27\(^-\)) that displays extrafollicular characteristics, such as the lack of CXCR5 and CD62L expression.\(^13\) These cells are developmentally related to activated naive B cells. However, they are considered as a distinct B cell population due to their unique transcriptomic profile and the reported increased rates of class-switch recombination.\(^7,23,48\) DN2 cells are hyper-responsive to TLR7 and, according to transcriptomic analyses, they seem to express a variety of cytokines and cytokine receptors, transcription factors such as T-bet, signalling factors and others.\(^13\) In severe cases of SLE, particularly in African-American female patients characterised by lupus nephritis and high

#### Table 1. Discrete double-negative (DN) B cell subsets.

| Nomenclature | Phenotype | Prominence in (condition) | Properties | References |
|--------------|-----------|---------------------------|------------|------------|
| DN1          | CD19\(^+\)IgD CD27 CD21\(^+\) CD11c\(^-\) T-bet\(^+\) CXCR5\(^+\) | Elderly healthy individuals (aging) | Memory precursors | [16,17] |
| DN2          | CD19\(^+\)IgD CD27 CD21\(^+\) CD11c\(^+\) T-bet\(^-\) CXCR5\(^+\) | SLE (autoimmunity) | Extralfollicular ASC precursors | [17] |
| DN3          | CD19\(^+\)IgD CD27 CD21\(^+\) CD11c\(^-\) [T-bet\(^{low}\)] | COVID-19 | ?? | [7,8] |
| atMEM/tbMEM  | CD19\(^+\)IgD CD27\(^-\) CD21 FcRL4\(^+\) | Chronic infections | Exhausted, mucosal resident B cells | [35,36] |

DN1, DN2 and DN3: first, second and third subset of DN B cells, respectively; atMEM/tbMEM: atypical/tissue-based memory B cells; SLE: Systemic Lupus Erythematosus; ASC: antibody-secreting cell.
titres of autoantibodies, DN2 cells expand dramatically and lead to the generation of autoreactive ASCs. In fact, the DN2 subset often becomes the predominant B cell population in these patients, with the highest frequencies of them being observed in the youngest individuals.13

In order for DN2 to be activated and differentiate into ASCs, TLR7 cooperates with IL-21 and IFNγ receptors. The synergistic triggering of these receptors is known to govern T-bet and CD11c expression in activated B cells.51 The elevated expression of T-bet in DN2 cells (and also ABCs) has been associated with development of autoimmunity and more specifically with autoreactive IgG production, enhanced antigen presentation to T cells and formation of spontaneous GCs.13,38,39,52,53 It is also important to mention that, in SLE mice models, conditional T-bet targeting in B cells leads to general improvement of health status,54 thus indicating that T-bet+ B cells are a pathogenetic population that may be targeted in clinical practice for therapeutic interventions.34 Moreover, considering the fact that the frequencies of these cells correlate with disease severity, we believe that they may also serve as prognostic and/or diagnostic markers of SLE.34

The fact that DN2 expansion is pronounced in African-American female SLE patients, indicates that genetic burden, as well as the sex of individuals, are crucial factors for the generation of these DN cells. Of note, TLR7 gene, which is of high importance for DN2 activation,13 is a chromosome X-linked gene, thus explaining (at least in part) the sex-bias that accompanies SLE immunopathogenesis.55,56 It is also important to mention that, in SLE mice models, conditional T-bet targeting in B cells leads to general improvement of health status,54 thus indicating that T-bet+ B cells are a pathogenetic population that may be targeted in clinical practice for therapeutic interventions.34 Moreover, considering the fact that the frequencies of these cells correlate with disease severity, we believe that they may also serve as prognostic and/or diagnostic markers of SLE.34

DN2 B cells in COVID-19

The function of DN B cells, both in health and disease, but mainly related to infectious diseases, is not yet fully elucidated and thorough investigation is required in order to understand the importance of DN percentage alterations that occur during pathological conditions. In case of COVID-19, our knowledge regarding DN B cells is extremely limited, as the disease was first observed almost two years ago.

Accumulating evidence, arising from a few studies that focused on B cell populations in COVID-19, suggest that DN2 B cells, along with the newly discovered DN3 subset, expand in severe and/or critical cases of the disease,7,12,14,15 while on the other hand, DN1 cells exhibit significant reductions.7,8 According to longitudinal progression studies though, the frequencies of these cells and the DN1/DN2 ratio are switching back to their initial levels (meaning the frequencies and ratio reported in healthy individuals), as the resolution of the disease approaches.6

In hospitalised COVID-19 patients, DN2 cells are associated with high neutralising antibody titres and elevated
plasma concentrations of inflammatory biomarkers, such as CRP, CXCL10, and IL-6. However, in general, it is not clear whether these cells are playing an active role in disease pathogenesis, or whether they just serve as biomarkers of the strong inflammation that occurs. The fact that T-bet expression in DN2 B cells from COVID-19 patients was found to be reduced, as compared to healthy individuals, indicates that the scenario of an active pathogenetic role for these cells in the aforementioned disease is not the most plausible. Nevertheless, some direct pathogenetic effects of DN2 B cells in COVID-19 could probably include extrafollicular B cell cytokine productions, enhanced antigen presentation to T cells and of course (auto)antibody production.

Although we focus on DN2 cells here, as they are the DN subset that is also (and mainly) marked in rheumatic diseases, we find it inevitable not to make a few comments regarding DN3 cells. The fraction of DN3 cells seems to be far from the most altered in COVID-19, since their frequency was seen to significantly increase as disease severity increased. Moreover, in contrast to other DN subsets, DN3 showed significant negative correlation with ventilator parameters, such as SpO2 and PaO2/FIO2, thus indicating that in hypoxic environments, the development program tilts toward DN3 cells. Interestingly once again, in comparison to the other two DN subsets, DN3 showed more correlation with several laboratory parameters including leukocytes, neutrophils, markers of increased cellular turnover such as CPK and LDH, and others related to critical COVID-19. Of note, DN3 cells were related to predominant extrafollicular B cell responses, as they are considered as newly recruited extrafollicular B cells with low somatic mutations and low affinity maturation.

MANAGEMENT OF RHEUMATIC DISEASES DURING THE COVID-19 PANDEMIC: POTENTIAL EFFECTS OF DN B CELLS

The symptomatology of COVID-19, as well as the potential outcome of death, have given rise to a worldwide concern and scare of exposition to SARS-CoV-2, especially among the rheumatological patients who believe to be at higher risk due to their immunological background and the immunosuppressive therapies. Nevertheless, contrary to these patients’ thoughts, it is strongly recommended by their caring physicians that all their therapies should be continued, as there is no convincing evidence so far that patients with rheumatic diseases are truly at higher risk than others.

We strongly believe that the clarification of DN B cells’ role and function, especially for DN2 cells as they are an overlapping population in rheumatic diseases and COVID-19, can lead to better management of rheumatological patients during the pandemic. Of course, there are some major issues that -in our opinion- require investigation, in order to make good use of our knowledge regarding DN B cells and thus manage the rheumatic disease patients more wisely.

The first one of these major issues refers to the activation pathway of COVID-19 and more specifically to all the phases of the infection, from the moment SARS-CoV-2 invades the host to the moment cytokine release syndrome occurs, as a result of inefficacious viral response and host inflammatory response. The synthesis of pro-inflammatory cytokines promotes the recruitment of macrophages and neutrophils into the lung alveoli, thus inducing a hyper-inflammation. We mentioned before that we cannot say for sure whether DN2 cells play an active role in these processes or whether they are just a consequence of the inflammation. However, we find it interesting that TLR7, as well as IL-6, IFNα, IFNγ, TNFα, and other DN related cytokines or chemokines are involved in the pathway. Of note, TLR7 hyper-responsiveness is considered as crucial in the promotion of pathology mediated by IL-21, IL6 and IFNα. Also, the elevated soluble mediators in severe COVID-19, such as IL-6, CXCL10 and TNFα, may suppress GC reactions, and thus promote extrafollicular pathways.

The second major issue that requires investigation refers to the impact of anti-rheumatic therapies, such as biologics, on the immunological memory achieved by COVID-19 vaccination or even resolved infection. Rituximab, an anti-CD20 chimeric monoclonal antibody used in battle against RA, as well as in management of lymphoma, significantly increases the risk of HBV-reactivation in resolved patients and of viraemic flares in chronically infected patients with low viral loads. Of course, even so, long-lived ASCs which lack CD20 expression, are thought to be preserved and thus maintain serum antibody responses in the absence of memory B cells. However, we cannot guarantee that under these circumstances ASCs can be reliably formed following HBV infection. In a similar way to HBV infection, we wonder whether, and to what extent, B cell depletion via pharmacological agents, such as Rituximab, Belimumab and others, affects the immunological memory achieved by COVID-19 vaccination or infection. In addition to this, there is also something else that is of high importance. DN2 cells expand in severe and/or critical cases of COVID-19, in contrast to more mild cases. However, we do not know whether a COVID-19 vaccination, which we do not consider as a severe or critical COVID-19 case, induces DN2 expansion. Of note, an increase in circulating DN B cells has been observed after vaccination against influenza virus in healthy subjects. Finding the answers to these questions, probably could enable better management of rheumatological patients during the pandemic, as determining the most proper vaccination and medication time points for the patients, would bring the most effective therapeutic benefits.

The third major issue, which is the last we want to
discuss about, but not the least important, refers to pharmacological agents, approved for anti-rheumatic therapies and now also registered in clinical trials to treat COVID-19. Interestingly, some of this medication, such as tocilizumab, an IL-6 receptor antagonist, is known to affect DN B cell percentages in RA,73 while others such as baricitinib, a JAK1 and JAK2 inhibitor,74 can target the IL-12-STAT4 axis which is the most potent at inducing both IFNγ and IL-21 by human CD4+ T cells, the cytokines promoting the differentiation of DN B cells in human SLE.75 Taking into account this evidence, we believe that targeting DN B cells (especially DN2 and DN3) may contribute to the management of rheumatic diseases during COVID-19 era, and even become a successful therapeutic approach for the infection.

CONFLICT OF INTEREST
The authors declare no conflict of interest.

REFERENCES
1. Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel Coronavirus from patients with pneumonia in China. 2019. N Engl J Med 2020;382(9):727-33.
2. Vabret N, Britton GJ, Gruber C, Hegde S, Kim J, Kuksin M, et al. Immunopathology of hyperinflammation in emerging lessons from hepatitis B and COVID-19. Immunol Rev 2018;311:35-9.
3. Tipton CM, Fucole CF, Darce J, Chica A, Ichikawa T, Gregoretti I, et al. Diversity, cellular origin and autoantigenicity of antibody-secreting cell population expansions in acute systemic lupus erythematosus. Nat Med 2015;16(7):755-65.
4. Frasca D, Diaz A, Romero M, Blomberg BB. Phenotypic and functional characterization of Double Negative B cells in the blood of individuals with obesity. Front Immunol 2021;12:616650.
5. Frasca D, Diaz A, Romero M, Thaller S, Blomberg BB. Metabolic requirements of human pro-inflammatory B cells in aging and obesity. PLoS One 2019;14(7):e0219545.
6. Wilder NH, Abadie P, Schulte S, Brauneck F, Kohsar M, Lütgehörmann M, et al. B cell analysis in SARS-CoV-2 versus malaria: Increased frequencies of plasmablasts and atypical memory B cells in COVID-19. J Leukoc Biol 2021;109(1):77-90.
7. Woodruff MC, Ramonell RP, Nguyen DC, Cashman KS, Saini AS, et al. B cell receptor signaling and effector function. Elife [Internet]. 2015;4. Available from: http://dx.doi.org/10.7554/eLife.07218
8. Wang S, Wang, J, Kumar V, Karnell JL, Naiman B, Gross PS, et al. IL-21 drives expansion and plasma cell differentiation of autoreactive CD1+CD19+ B cells in SLE. Nat Commun 2018;9(1):1758.
9. Zumaquero E, Stone SL, Scharrer CD, Jerks SA, Nellore A, Mousseau B, et al. IFNγ induces epigenetic programming of human T-bet+ B cells and promotes TLR7/8 and IL-21 induced differentiation. E life 2019;8:e41641.
10. Portugal S, Tipton CM, Sohn H, Kone Y, Wang J, Li S, et al. Malaria-associated atypical memory B cells exhibit markedly reduced B cell receptor signaling and function. Elife [Internet]. 2021;10. Available from: http://dx.doi.org/10.7554/eLife.59237
11. Burton AR, Maini MK. Human antiviral B cell responses: A novel perspective. Am J Pathol 2021;191(1):4-17.
12. Oliviero B, Varchetta S, Mele D, Mantovani S, Cerino A, Perotti CG, et al. Cutting Edge: B Cell-Intrinsic T-bet Expression Is Required To Control Chronic Viral Infection. J Immunol 2016;197(4):1017-22.
13. Wei C, Anolić J, Cappio-Pepe A, Zheng B, Pugh-Bernard A, Brooks J, et al. A new population of cells lacking expression of CD27 represents a notable component of the B cell memory compartment in systemic lupus erythematosus. J Immunol 2007;178(10):6624-33.
14. Claeys N, Fraussen J, Vanheusden M, Hellings N, Stinissen P, Van Wijmeersch B, et al. Age-associated B cells with proinflammatory characteristics are expanded in a proportion of multiple sclerosis patients. J Immunol 2016;197(12):4576-83.
15. Mörk S, Ho J, Malaspina A, Wang W, DiPoto AC, O’Shea MA, et al. Evidence for HIV-associated B cell exhaustion in a dysfunctional memory B cell compartment in HIV-infected viremic individuals. J Exp Med 2008;205(8):1797-805.
16. Tipton CM, Sander S, Chu VT, Yasuda T, Franklin A, Graf R, et al. Failure of B Cell Tolerance in COVID. Front Immunol 2020;11:10. Available from: http://dx.doi.org/10.3389/fimmu.2020.569458
17. Mauel JW, Catalina MD, Kumar V, Bachali P, Grammer AC, Wang S, et al. Transcriptome and IgH repertoire analyses show that CD11c+ B cells are a distinct population with similarity to B cells arising in autoimmunity and infection. Front Immunol [Internet] 2021;12. Available from: http://dx.doi.org/10.3389/fimmu.2021.649458
18. Rubtsov AV, Marrack P, Rubtsova K. T-bet Expressing B cells – novel target for autoimmune therapies? Cell Immunol 2017;321:35-9.
19. Palanichamy A, et al. Extrafollicular B cell responses correlate with affinity-matured B cell population expansions in acute systemic lupus erythematosus. Nat Med 2015;16(7):755-65.
20. Sander S, Chu VT, Yasuda T, Franklin A, Graf R, Cakado DP, et al. A double-negative (IgD-CD27-) B cell population expansions in acute systemic lupus erythematosus. MedRxiv [Internet]. 2020; Available from: http://dx.doi.org/10.1101/2020.04.29.20083717
21. Farris AD, Guthridge JM. Overlapping B cell pathways in severe COVID-19 and lupus. Nat Immunol 2020;21(12):1478-80.
22. Colonna-Romano G, Bulati M, Aquino A, Pellicano M, Vitello S, Lio D, et al. A double-negative (IgD-CD27-) B cell population is increased in the peripheral blood of elderly people. Mech Ageing Dev 2009;130(10):681-90.
23. Wei C, Anolić J, Capias A, Zheng B, Pugh-Bernard A, Brooks J, et al. A new population of cells lacking expression of CD27 represents a notable component of the B cell memory compartment in systemic lupus erythematosus. J Immunol 2007;178(10):6624-33.
24. Claeys N, Fraussen J, Vanheusden M, Hellings N, Stinissen P, Van Wijmeersch B, et al. Age-associated B cells with proinflammatory characteristics are expanded in a proportion of multiple sclerosis patients. J Immunol 2016;197(12):4576-83.
25. Mörk S, Ho J, Malaspina A, Wang W, DiPoto AC, O’Shea MA, et al. Evidence for HIV-associated B cell exhaustion in a dysfunctional memory B cell compartment in HIV-infected viremic individuals. J Exp Med 2008;205(8):1797-805.
26. Tipton CM, Sander S, Chu VT, Yasuda T, Franklin A, Graf R, et al. Failure of B Cell Tolerance in COVID. Front Immunol 2020;11:10. Available from: http://dx.doi.org/10.3389/fimmu.2020.569458
27. Mauel JW, Catalina MD, Kumar V, Bachali P, Grammer AC, Wang S, et al. Transcriptome and IgH repertoire analyses show that CD11c+ B cells are a distinct population with similarity to B cells arising in autoimmunity and infection. Front Immunol [Internet] 2021;12. Available from: http://dx.doi.org/10.3389/fimmu.2021.649458
28. Rubtsov AV, Marrack P, RubtsovaK. T-bet Expressing B cells – nov- el target for autoimmune therapies? Cell Immunol 2017;321:35-9.
29. Barnett BE, Staupe RP, Odorizzi PM, Palko O, Tomov VT, Mahan AE, et al. Cutting Edge: B Cell-Intrinsic T-bet Expression Is Required To Control Chronic Viral Infection. J Immunol 2016;197(4):1017-22.
30. Richardson CT, Slack MA, Dhillon G, Marcus CZ, Barnard J, Palaricharanya A, et al. Failure of B Cell Tolerance in COVID. Front Immunol 2019;10:289.
31. Hitzo Ahye MK, Golding A. Cytoplasmic FOXP1 identifies a novel disease-activity associated B cell phenotype in SLE. Lupus Sci Med 2018;5(1):e000296.
32. Sander S, Chu VT, Yasuda T, Franklin A, Graf R, Cakado DP, et al. P13 Kinase and FOXO1 Transcription Factor Activity Differentially Control B Cells in the Germinal Center Light and Dark Zones. Immunity 2015;43(1):107-16.
33. Luo W, Weisel F, Shlomchik MJ. B Cell Receptor and CD40 Signaling Are Rewired for Synergistic Induction of the c-Myc Transcription Factor in Germinal Center B Cells. Immunity 2018;48(2):313-326.e5.
DN B CELLS IN RHEUMATIC DISEASES AND COVID-19

34. Sachinidis A, Xanthopoulos K, Garyfallos A. Age-associated B cells (ABCs) in the prognosis, diagnosis, and therapy of Systemic Lupus Erythematosus (SLE). Mediterr J Rheumatol 2020;31(3):311-8.

35. Rinaldi S, Palliikkuth S, George VK, de Armas LR, Pahuja R, Sanchez CM, et al. Paradoxical aging in HIV: immune senescence of B Cells is most prominent in young age. Aging (Albany NY) 2017;9(4):1307-25.

36. Blavatov J, Butler IS, Roentgen S, Mwacharo J, Pierias SK, Bejon P, et al. Chemokine exposure to Plasmodium falciparum is associated with phenotypic evidence of B and T cell exhaustion. J Immunol 2013;190(3):1038-47.

37. Hao Y, O’Neill P, Naradikian MS, Scholz JL, Cancro MP. A B-cell subset uniquely responsive to innate stimuli accumulates in aged mice. Blood 2011;118(8):1294-304.

38. Rubtsov AV, Rubtsova K, Fischer A, Meeein RT, Gillis JZ, Kappler JW, et al. Toll-like receptor 7 (TLR7)-driven accumulation of a novel CD11c+ B-cell population is important for the development of autoimmunity. Blood 2011;118(5):1305-15.

39. Rubtsov AV, Marrack P. T-box transcription factor T-bet, a key player in a unique type of B-cell activation essential for effective viral clearance. Proc Natl Acad Sci U S A. 2013;110(34):E3216–E3224.

40. Rubtsov AV, Rubtsova K, Halemamo K, Li SX, Kappler JW, Santiago ML et al. T Cell Production of IFNγ in Response to TLR7/IL-12 Stimulates Optimal B Cell Responses to Viruses. PLoS ONE 2016;11(11):e0166322.

41. Phakee S, Marrack P. Age (autoimmunity) associated B cells (ABCs) and their relatives. Curr Opin Immunol 2018;55:24-30.

42. Cancro MP. Age-associated B cells. Annu Rev Immunol 2020;38(1):315–40.

43. Meslin L, Ersching J, Victorla GD. Germline center B cell dynamics. Immunity 2016;45(5):471-82.

44. Allman D, Wilmore JR, Gaudette BT. The continuing story of T-cell immunopathology. J Clin Invest 2017;127(4):1392-404.

45. Elsner RA, Shlomchik MJ. Germinal center and extrafollicular B cell differentiation through type I interferon and interleukin 6. Immunity 2016;11(1):e0166322.

46. Illingworth J, Butler NS, Roetynck S, Mwacharo J, Pierce SK, Bejon P, et al. Cutting edge: IL-4, IL-21, and IFN-γ interact to govern adaptive lymphocyte responses to Plasmodium falciparum in naïve C57BL/6 mice. J Immunol 2007;178(10):6200-7.

47. Cohen MD, Keystone E. Rituximab for rheumatoid arthritis. Nat Rev Rheumatol 2015;11:606338.

48. Ly A, Liao Y, Pietrzak H, Ioannidis LJ, Sidwell T, Glory R, et al. Transcription factor T-bet in B cells modulates germline center polarization and antibody affinity maturation in response to malaria. Cell Rep 2019;29(8):2257-2269.e6.

49. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

50. Channappanavar R, Perlman S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. Semin Immunopathol 2017;39(5):529–39.

51. Kaneko N, Kuo H-H, Boucau J, Farmer JR, Allard-Charrand H, Mahajan VS, et al. Loss of Bcl-6 expressing T follicular helper cells and germinal centers in COVID-19. Cell 2020;183(1):143-157.e13.

52. Cohen MD, Keystone E. Rituximab for rheumatoid arthritis. Rhetumatol Ther 2015;22(9):111-111.

53. Boedigheimer MJ, Martin DA, Amoura Z, Sangiamento deciphering the role of B cells in rheumatoid arthritis. Rheumatology 2017;56(1):60-70.

54. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

55. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

56. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

57. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

58. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

59. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

60. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

61. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

62. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

63. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

64. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

65. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

66. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

67. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

68. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

69. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.

70. Bobrov A, Bobrova N, Mikhailova G, et al. B cell alterations during BAFF inhibition with belimumab in rheumatoid arthritis. EBioMedicine 2019;40:517-527.