Research Article

Inducible Conditional Vascular-Specific Overexpression of Peroxisome Proliferator-Activated Receptor Beta/Delta Leads to Rapid Cardiac Hypertrophy

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Peroxisome proliferator-activated receptors are nuclear receptors which function as ligand-activated transcription factors. Among them, peroxisome proliferator-activated receptor beta/delta (PPARβ/δ) is highly expressed in the heart and thought to have cardioprotective functions due to its beneficial effects in metabolic syndrome. As we already showed that PPARβ/δ activation resulted in an enhanced cardiac angiogenesis and growth without impairment of heart function, we were interested to determine the effects of a specific activation of PPARβ/δ in the vasculature on cardiac performance under normal and in chronic ischemic heart disease conditions. We analyzed the effects of a specific PPARβ/δ overexpression in endothelial cells on the heart using an inducible conditional vascular-specific mouse model. We demonstrate that vessel-specific overexpression of PPARβ/δ induces rapid cardiac angiogenesis and growth with an increase in cardiomyocyte size. Upon myocardial infarction, vascular overexpression of PPARβ/δ, despite the enhanced cardiac vessel formation, does not protect against chronic ischemic injury. Our results suggest that the proper balance of PPARβ/δ activation in the different cardiac cell types is required to obtain beneficial effects on the outcome in chronic ischemic heart disease.

1. Introduction

Peroxisome proliferator-activated receptors (PPARs) are ligand-activated transcription factors that belong to the nuclear receptor superfamily. There are three members of the PPAR family (α, β/δ, and γ) with distinct, but overlapping spatial, temporal, and regulated expression patterns. For all PPARs, lipids are endogenous ligands and PPARs are considered as important transcriptional regulators of genes involved in lipid metabolism and cardiac energy production [1].

PPARβ/δ is the predominant subtype in the heart, and several lines of evidence suggest a cardioprotective function of PPARβ/δ. Cardiac PPARβ/δ deletion in mice resulted in cardiac dysfunction, hypertrophy, and congestive heart failure [2]. Furthermore, it has been shown that the PPARβ/δ agonist L-165041 inhibits pharmacologically induced hypertrophy of cardiomyocytes through the interaction of PPARβ/δ to NF-κB and a subsequent downregulation of NF-κB target genes [3, 4]. An in vivo study demonstrated that cardiac specific overexpression of PPARβ/δ led to increased myocardial glucose utilisation and did not alter cardiac function but tended to exert a protective effect to ischemia/reperfusion-induced myocardial injury. This was attributed to an activation of the Glut-4 promoter by PPARβ/δ and the subsequently increased cardiac glucose utilisation [5]. Finally, we recently showed that pharmacological activation of PPARβ/δ with GW0742...
or GW501516 in mice led to rapid cardiomyocyte growth with a preserved myocardial function. We demonstrated that PPARβ/δ directly activates the Calcineurin gene [6], which is known to induce cardiac growth [7, 8]. Most interestingly, we observed in our study a rapid induction of cardiac angiogenesis upon pharmacological PPARβ/δ activation, a matter which surprisingly had not been investigated before, although the correlation between cardiac growth and angiogenesis seems quite evident. PPARβ/δ expression in endothelial cells has already been reported in 1999 by Bishop-Bailey and Hla [9]. Pharmacological activation of endothelial and endothelial progenitor cells with PPARβ/δ agonists had been shown to increase the migration, proliferation, and tube formation of these cells [10, 11].

Furthermore, PPARβ/δ knockout mice exhibited a diminished blood flow and immature microvascular structures in subcutaneously induced tumors, which could be rescued by reexpression of PPARβ/δ [12]. In human pancreatic tumors, PPARβ/δ expression strongly correlated with the advanced tumor stage and increased risk of tumor recurrence and distant metastasis. PPARβ/δ has therefore been suggested to be involved in the regulation of the angiogenic switch in tumor progression [13].

PPARβ/δ is also involved in physiological angiogenesis. As we and others showed, treatment with the PPARβ/δ agonists GW0742 and GW501516 induced an exercise-like phenotype in the heart. Both agonists induced a surprisingly rapid (after 24 h) remodelling of mouse hearts [6] and skeletal muscle [14] by increasing microvessel densities.

However, until now it was not clear if either the increase of the cardiac vasculature drives the myocardial hypertrophy or the enhanced cardiac angiogenesis might be a potential indirect effect of cardiomyocyte-specific PPARβ/δ overexpression.

In our present work, we address this question through the generation of transgenic mice with an inducible conditional vascular-specific overexpression of PPARβ/δ and analyze the normal cardiac phenotype and function as well as function and histology after experimental myocardial infarction.

We show that inducible vessel-specific overexpression of PPARβ/δ results in a rapid induction of angiogenesis, cardiac hypertrophy, and impairment of cardiac function as reflected by enhanced end-diastolic and end-systolic volumes, reduced fractional shortening, and decreased ejection fractions. Additionally, we demonstrate that, after myocardial infarction, despite the higher collateral vessel formation, the animals with vascular-specific PPARβ/δ overexpression display bigger infarct lesions, higher cardiac fibrosis, and further reduced cardiac function. This points to a more careful view about the potential benefits of PPARβ/δ agonists in the treatment of cardiovascular diseases, as the proper balance between cardiomyocytic and vascular PPARβ/δ seems to be crucial for cardiac health, especially under ischemic conditions.

2. Materials and Methods

2.1. Animals. All animals were used in accord with local Home Office regulations. PPARβ/δ-flx+ve [15] and Tie2-CreERT2 [16] animals were crossed to generate Tie2-CreERT2;PPARβ/δ-flx+ve mice, further referred to as Tie2-CreERT2;PPARβ/δ. The Tie2-Cre-line was backcrossed four times onto C57BL6. Age- and sex-matched Tie2-CreERT2;PPARβ/δ animals were injected for one week intraperitoneally either with sunflower oil (vehicle) or Tamoxifen dissolved in sunflower oil in a dose of 33 mg/kg per day [17]. Tie2-CreERT2 animals injected with Tamoxifen served as an additional control. Anaesthetized mice were examined by echocardiography using the iE33 xMATRIX system with a 12 MHz transducer (Philips Healthcare, DA Best, Netherlands). Myocardial infarctions were induced by ligation of the left coronary artery (LAD) as described [18]. Briefly, anaesthetized mice were endotracheally intubated, the skin was incised on the left thorax side, the pectoralis muscles were mobilized, a thoracotomy between the third and fourth rib was performed, and the LAD permanently was closed with a 7-0 suture distal to the left auricle. This resulted in large myocardial infarctions. The thoracotomy and the skin wound were closed with 4-0 sutures and the mice remained intubated until spontaneous respiration was reestablished. Lethality of the procedure was approximately 50% independent of the genotype of the mouse.

2.2. Genotyping. The genotype of animals was identified by PCR. PCR conditions and primer sequences are available on request.

2.3. Tissue Samples, Histology, and Immunohistology. Histology and measurement of cardiomyocyte diameters were performed according to established protocols [19]. Samples from at least five different animals per group (Tie2-CreERT2;PPARβ/δ + vehicle, Tie2-CreERT2 + Tamoxifen, and Tie2-CreERT2;PPARβ/δ + Tamoxifen) were analyzed. Investigators were blinded for the genotype of the mice. Three µm paraffin sections were used for histological and immunohistological procedures.

Haematoxylin-Eosin staining was routinely performed on all tissue samples; additionally, sections were stained with Trichrome Masson and Picrosirius red. For PPARβ/δ and Pecam-1 immunohistology, after heat-mediated antigen retrieval and quenching of endogenous peroxidase activity, the antigen was detected after antibody application Pecam-1 (CD31) (1:100, rabbit polyclonal, ab28364, Abcam) or PPARβ/δ (1:100, rabbit polyclonal, ab154395, Abcam) using EnVision™ Peroxidase/DAB Detection System from Dako (Trappes, France). Sections were counterstained with Haematoxylin (Sigma). Omission of the first antibody served as a negative control. Additionally, some slides were incubated with IgG Isotype Controls (1:100, rabbit monoclonal, clone SP137, Abcam). Slides were viewed under an epifluorescence microscope (DMLB, Leica, Germany) connected to a digital camera (Spot RT Slider, Diagnostic Instruments, Scotland).

Area densities for all immunohistological stainings were determined using the ImageJ software. Vessel area density was analyzed on at least five different sections of hearts per mouse.
Tie2-CreERT2;PPARβ/δ + vehicle  Tie2-CreERT2;PPARβ/δ + Tamoxifen

(a)

PPARβ/δ rel. expression

Cardiac endothelial cells

(b)

Calcineurin rel. expression

Whole cardiac tissue

(c)

PPARβ/δ rel. expression

Pecam-1 rel. expression

eNOS rel. expression

1-week induction heart

1-week induction heart

(d)

Pecam-1 area density (%)

Figure 1: Continued.
2.4. Real-Time RT-PCR. Total RNA was isolated from hearts and cardiac endothelial cells, sorted with CD31 MicroBeads (Miltenyi Biotec) from the mouse hearts using the Trizol reagent (Invitrogen). The RNA pellet was dissolved in diethyl pyrocarbonate-treated H₂O. First-strand cDNA synthesis was performed with 0.5 µg of total RNA using oligo(dT) primers and Superscript III reverse transcriptase (Invitrogen). One µL of the reaction product was taken for real-time RT-PCR amplification (ABI Prism 7000, Applied Biosystems) using a commercial SYBR® Green kit (Eurogentec, Angers, France). Primer sequences are available on request. Expression of each gene was normalized to the respective Gapdh, Actb, and Rplp0 expression.

2.5. Statistical Analysis. Data are expressed as means ± SEM. ANOVA with Bonferroni test as post hoc test or Mann-Whitney tests was performed as indicated. A p value of less than 0.05 was considered statistically significant.

3. Results and Discussion

3.1. PPARβ/δ Vascular-Specific Overexpression Rapidly Increases Cardiac Vessel Density. Immunohistochemistry for PPARβ/δ of heart sections proved the upregulation of PPARβ/δ protein expression in the endothelium of Tie2-CreERT2;PPARβ/δ mice induced with Tamoxifen as compared to vehicle-treated Tie2-CreERT2;PPARβ/δ animals (Figure 1(a)). Quantitative RT-PCRs from cardiac endothelial cells enriched with Pecam-1/CD31 MicroBeads were performed to confirm the vascular overexpression of PPARβ/δ upon Cre-mediated recombination. Endothelial cells isolated from hearts of Tamoxifen induced Tie2-CreERT2;PPARβ animals
showed a modest upregulation of PPARβ/δ and Calcineurin expression compared to cardiac vascular cells of vehicle-treated animals (Figure 1(b)). In contrast, no significant changes in PPARβ/δ expression levels in whole heart RNA preparations could be detected (Figure 1(c)), additionally confirming specificity of vascular PPARβ/δ overexpression, as endothelial cells contribute only to around seven percent of the total cell numbers in the mouse heart [20]. An increase in cardiac vessel density became evident on the RNA (Figure 1(c)) as well as on the protein level (Figure 1(d)) already one week after Cre-mediated vascular PPARβ/δ overexpression. Additionally, increased cardiac eNOS expression confirmed the enhanced cardiac angiogenesis (Figure 1(c)). The detection of Pecam-1 protein expression by immunohistochemistry allowed determining that this upregulation of Pecam-1 was due to the formation of new microvessels (for comparison, see Figure 1(d) right photomicrograph, which depicts higher microvessel formation in the hearts of Tie2-CreERT2;PPARβ/δ animals induced with Tamoxifen as compared to vehicle-treated Tie2-CreERT2;PPARβ/δ animals on the left or Tamoxifen treated Tie2-CreERT2 animals in the middle). Determination of Pecam-1 area density indicated a doubling of Pecam-1 positive vascular structures (Figure 1(d)). This angiogenic response to transgenic overexpression of PPARβ/δ in the endothelium was also observed in the kidney and the pancreas (Figure 1(e)), indicating a general proangiogenic action of PPARβ/δ in endothelial cells. These findings are in line with previous studies, which reported a rapid enhancement of vessel density upon pharmacological PPARβ/δ activation [6,14] and the general view of PPARβ/δ as a proangiogenic factor [21].

3.2. Specific Vascular Overexpression of PPARβ/δ Induces Cardiac Hypertrophy. Already one week after induction of PPARβ/δ expression in vessels, it became evident that cardiac growth was enhanced in the animals with Cre-mediated recombination as compared to both controls, vehicle-treated Tie2-CreERT2;PPARβ/δ and Tie2-CreERT2 mice treated with Tamoxifen. Heart/body weight measurements confirmed the macroscopic observation. This growth induction became more enhanced after three weeks and remained then stable for up to two months, the latest time point studied (Figure 2). The cause of this cardiac growth was an increase in cardiomyocyte size, as determined by cardiomyocyte diameter measurements at the different time points. On average, the cardiomyocyte diameter increased about 30% compared to the respective controls (Figure 3). Vascular formation during embryonic development is crucial for organ growth; for example, the inhibition of coronary vessel formation abolishes cardiac growth [22]; however, the factors determining organ size in an adult organism are not completely understood, but some lines of evidence suggest that, during tissue repair or in response to physiological stimuli vessel formation is required for organ enlargement [23]. Some evidence that at least for the heart vascular growth indeed led to an increase in the cardiac mass under normal conditions came from a study using transgenic mice with a cardiomyocyte-specific on/off regulatable system for the secretion of the proangiogenic factor PR39. The authors concluded that myocardial hypertrophy observed after three weeks was due to the induction of angiogenesis. They suggested that increased NO production due to increased endothelial cell mass mediated the observed hypertrophy [24]. This is in accordance with our finding of enhanced eNOS expression in the hearts of mice with vascular-specific overexpression of PPARβ/δ (Figure 1(b)). However, PR39 is a macrophage derived peptide, which inhibits degradation of hypoxia inducible factor α protein, thus activating angiogenesis through the induction of VEGF and fibroblast growth factor signalling and acting on all cardiac and other cell types. It can therefore not be excluded that part of the observed effects in this study was due to actions of PR39 on other cell types of the heart compared to only endothelial cells. The fact that we could observe cardiac hypertrophy already one week after vascular-specific overexpression of PPARβ/δ is mostly due to the overexpression of PPARβ/δ in endothelial cells, which induced angiogenesis leading to hypertrophy of the cardiomyocytes. Our approach was more direct as targeting the secretion of a proangiogenic factor like PR39 by cardiomyocytes, which affects secondarily the endothelium and in the end the increase in cardiomyocyte size is solely attributed to the increased angiogenesis. However, in the mentioned study, it cannot be excluded that the forced secretion of a proangiogenic molecule by cardiomyocytes also acts on other cell types compared to only endothelial cells, including cardiomyocytes themselves. The endothelial-specific conditional induction of PPARβ/δ in our model excludes a potential interference with possibly in parallel ongoing actions in other cardiac cell types.

3.3. PPARβ/δ Vascular-Specific Overexpression Also Increases Capillary Density in the Setting of Myocardial Infarction but Fails to Ameliorate the Outcome after Chronic Ischemic Heart Disease. To investigate the effect of PPARβ/δ driven angiogenesis on myocardial function in pathological settings, the left anterior descending (LAD) coronary artery in Tie2-CreERT2;PPARβ/δ animals induced with Tamoxifen or treated with vehicle was ligated. Immunohistochemical investigation of Pecam-1 expression demonstrated a significant increase in capillary density not only in the infarct zone but also in the border zone of the infarcted area and in the remote myocardial area of the right ventricle of Tie2-CreERT2;PPARβ/δ animals induced with Tamoxifen or treated with vehicle only. This was additionally confirmed by quantification of Pecam-1 area densities (Figure 4). Heart/body weight determination demonstrated a hypertrophic effect of vascular-specific overexpression of PPARβ/δ also in the setting of chronic ischemic heart disease, due to an increase in cardiomyocyte size (Figure 5(a)). Interestingly, histological analyses revealed much bigger infarct sizes in animals with vascular-specific overexpression of PPARβ/δ as compared to controls (Figure 5(b)) and an enhanced cardiac fibrosis, as determined by Picrosirius red staining for collagen (Figure 5(c)). This is in contrast to the study using a cardiomyocyte-specific on/off regulatable system for
the secretion of the proangiogenic factor PR39 from cardiomyocytes; the secretion of PR39 reduced infarct sizes after myocardial infarction [24]. However, as stated before, this study was based on the effects of PR39, a macrophage derived proangiogenic molecule, which might act on all cardiac cell types rather than solely on endothelial cells. Our results are in agreement with clinical studies suggesting cardiac hypertrophy as a risk factor for arteriosclerosis, myocardial infarction, and heart failure [25]. This is probably due to the increased energy consumption of hypertrophic myocardium.

To test whether the angiogenesis induced cardiac hypertrophy affects cardiac function, we performed premyocardial and three-week postmyocardial infarction echocardiographic examinations in Tie2-CreERT2;PPARβ/δ animals induced with Tamoxifen and the respective controls treated with vehicle. Consistent with the observed cardiac

**Figure 2:** Rapid induction of cardiac growth by Tie2-Cre-mediated conditional PPARβ/δ overexpression. Photomicrographs of Hematoxylin-Eosin- (HE-) stained cross sections of the hearts and respective heart-to-body weight ratios (Tie2-CreERT2;PPARβ/δ + Tamoxifen, n = 7, Tie2-CreERT2;PPARβ/δ + vehicle, n = 6, and Tie2-CreERT2 + Tamoxifen, n = 6). Scale bars indicate 2 mm. Data are means ± SEM. * p < 0.05 and ** p < 0.01.
hypertrophy, mice with vascular-specific overexpression of PPARβ/δ showed an increase in left ventricular end-diastolic (LVED) and -systolic (LVES) volume. Fractional shortening and the ejection fraction were slightly reduced as compared to the respective controls (Figure 6(a)).

Three weeks after myocardial infarction, control Tie2-CreERT2;PPARβ/δ animals treated with vehicle also showed an increase in the left ventricular end-diastolic and -systolic volume as well as a reduction in the fractional shortening and ejection fraction when compared to their healthy status.
Figure 4: Increased vessel formation after myocardial infarction in the hearts of mice with vascular-specific PPARβ/δ overexpression. Pecam-1-immunostaining in mouse heart sections and quantification of Pecam-1 signal area density (Tie2-CreERT2;PPARβ/δ + Tamoxifen, n = 5, and Tie2-CreERT2;PPARβ/δ + vehicle, n = 5). Scale bars indicate 50 μm. Data are means ± SEM. **p < 0.01 and ***p < 0.001.

Animals treated with PPARβ/δ agonists showed a rapid increase of the cardiac vasculature and an enhanced cardiac growth without functional impairment [6]. It seems as if the proper balance between PPARβ/δ activation in endothelial cells and cardiomyocytes (and maybe other cardiac cell types as fibroblasts) is required to confine the attribute “cardioprotective” to PPARβ/δ. Our results indicate that the specific, unbalanced activation of PPARβ/δ only in the vasculature, despite its effects on vessel and cardiac growth, is not sufficient to protect against chronic ischemic heart disease. Nevertheless, it is possible that activation of PPARβ/δ in the vasculature might have beneficial effects in the settings of smaller infarct sizes or in slowly developing arteriosclerosis, which will be subject of future studies.
Figure 5: Increased infarct sizes and higher cardiac fibrosis in animals with vascular-specific PPARβ/δ overexpression. (a) Respective heart-to-body weight ratios and high power photomicrographs of HE-stained heart sections showing individual cardiomyocytes and quantification of cardiomyocyte diameters. Scale bars indicate 50 μm. (b) Photomicrographs of Trichrome Masson stained cross sections and quantification of the infarct sizes (Tie2-CreERT2;PPARβ/δ + Tamoxifen, n = 8, and Tie2-CreERT2;PPARβ/δ + vehicle, n = 5). Scale bars indicate 2 mm. (c) Photomicrographs of Picrosirius red stained cross sections and quantification of cardiac fibrosis. Scale bars indicate 2 mm. Data are means ± SEM. **p < 0.01 and ***p < 0.001.
Figure 6: Impaired cardiac function upon vascular-specific PPARβ/δ overexpression, which worsens after myocardial infarction. (a) Echocardiographic examination indicates increased systolic and diastolic volumes, a reduced fractional shortening, and a decreased ejection fraction in animals with vessel-specific overexpression of PPARβ/δ, which becomes more evident after myocardial infarction (b) (Tie2-CreERT2;PPARβ/δ + Tamoxifen, n = 8, and Tie2-CreERT2;PPARβ/δ + vehicle, n = 5). IVS: interventricular septum; LVID: left ventricular internal diameter; LVPW: left ventricular posterior wall. Data are means ± SEM. *p < 0.05, **p < 0.01, and ***p < 0.001.
4. Conclusions

In this study, we investigated the effects of a vascular-specific overexpression of PPARβ/δ on cardiac phenotype and function. The rapid induction of cardiac vessel formation was accompanied by an induction of cardiac growth, characterized by an increase in cardiomyocyte diameter. Upon myocardial infarction, the increased cardiac angiogenesis neither reduced infarct sizes nor improved the cardiac function. The proper balance of PPARβ/δ activation in the different cardiac cell types may be important for potential cardioprotective effects of PPARβ/δ.

Conflict of Interests

The authors declare no conflict of interests.

Acknowledgments

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