Preclinical and Clinical Applications of Biomaterials in the Enhancement of Wound Healing in Oral Surgery: An Overview of the Available Reviews

Giacomo Picciolo 1,†, Matteo Peditto 1,†, Natasha Irrera 2,3, Giovanni Pallio 2✉, Domenica Altavilla 1,3, Mario Vaccaro 2, Giuseppe Picciolo 3, Alessandro Scarfone 2, Francesco Squadrito 2,3,* and Giacomo Oteri 1

1 Department of Biomedical and Dental Sciences and Morphological and Functional Imaging, University of Messina, Via C. Valeria, 98125 Messina, Italy; giacomo.picciolo@unime.it (G.P.); matteo.peditto@unime.it (M.P.); daltavilla@unime.it (D.A.); giacomo.oteri@unime.it (G.O.)
2 Department of Clinical and Experimental Medicine, University of Messina, Via C. Valeria, 98125 Messina, Italy; nirrera@unime.it (N.I.); gpallio@unime.it (G.P.); vaccaro@unime.it (M.V.); alessandro.scarfone@unime.it (A.S.)
3 SunNutraPharma, Academic Spin-Off Company of the University of Messina, Via C. Valeria, 98125 Messina, Italy; beppepicciolo24@gmail.com
* Correspondence: fsquadrito@unime.it; Tel: +39-0902213648
† These authors contribute equally to this work.

Received: 28 September 2020; Accepted: 21 October 2020; Published: 24 October 2020

Abstract: Oral surgery has undergone dramatic developments in recent years due to the use of biomaterials. The aim of the present review is to provide a general overview of the current biomaterials used in oral surgery and to comprehensively outline their impact on post-operative wound healing. A search in Medline was performed, including hand searching. Combinations of searching terms and several criteria were applied for study identification, selection, and inclusion. The literature was searched for reviews published up to July 2020. Reviews evaluating the clinical and histological effects of biomaterials on post-operative wound healing in oral surgical procedures were included. Review selection was performed by two independent reviewers. Disagreements were resolved by a third reviewer, and 41 reviews were included in the final selection. The selected papers covered a wide range of biomaterials such as stem cells, bone grafts, and growth factors. Bioengineering and biomaterials development represent one of the most promising perspectives for the future of oral surgery. In particular, stem cells and growth factors are polarizing the focus of this ever-evolving field, continuously improving standard surgical techniques, and granting access to new approaches.

Keywords: biomaterials; oral wound healing; oral surgery

1. Introduction

The birth of oral surgery is difficult to trace back. Considering exodontics as a part of this specialty, its roots may go back to the origins of dentistry itself. However, since the publication of Anselme Louis Bernard Berchillet’s “Traité des maladies et des opérations réellement chirurgicales de la bouche,” published in 1778, oral surgery has dramatically evolved and expanded together with the other surgical fields of medicine, effectively encompassing a wide range of procedures beyond exodontics, such as oral pathology, orthodontics, complex reconstructive techniques, and so on. These changes are intertwined with the continuous development of evidence-based medicine and new technologies that constantly challenge the very logic behind the surgical approach, from diagnosis to treatment. Improving oral surgery gives access to new and more complex clinical scenarios stemming from the increase in patients’
demands and age along with the increase in complications such as osteoradionecrosis, drug-related osteonecrosis, and peripheral trigeminal nerve injuries [1]. Bioengineering and the development of biomaterials have opened up one of the most promising doors to the future of oral surgery [2–6]; the ability to selectively and predictably regenerate tissue, such as bone, mucosa, and nerve tissue, using new technologies, is the key to improve the surgical outcomes of oral wound healing and simplify the current reconstructive approach to complex cases, such as the atrophic mandible or soft tissue management in periodontal patients. On the way to reach this goal, clinicians are witnessing a significant shift in the field of biomimetic biomaterials and bioactive ones, specifically engineered to promote wound healing exploiting molecules or additives with different purposes, such as targeted cellular recruitment or antimicrobial potential, or specific design elements, shifting from simple scaffolds to matrixes providing a spatial orientation for tissue growth [7]. The aim of this review is to provide a comprehensive overview of the current knowledge on biomaterials used in oral surgery and to review the available systematic reviews concerning biomaterials used in oral surgery and their impact on post-operative wound healing.

2. Methods

2.1. Development of a Protocol

The overview has been carried out in agreement with a defined protocol, that included: the identification of a focused question, search strategy, criteria for review selection and inclusion.

2.2. Defining the Focused Question

The focused question was defined as: ‘What is the clinical and histological effect of biomaterials as adjuvants of the oral surgical procedures on oral wound healing as evaluated by systematic literature reviews?’.

2.3. Search Strategy

The literature was searched for reviews published up to July 2020, using the MEDLINE database. Different combinations of search terms were developed to identify suitable studies. Papers from reference lists of the considered reviews articles were also manually scanned and screened. Search strategy is shown in Figure 1.

Figure 1. Search strategy.
2.4. Criteria for Review Selection and Inclusion

The review selection was performed by two independent reviewers. Disagreements were resolved by a third reviewer. Selection was limited to reviews that included clinical studies describing both clinical and histological effects of biomaterials on post-operative wound healing in oral surgical procedures. Reviews analyzing non-surgical oral procedures (e.g., orthodontics and conservative dentistry therapies) were discarded. A time limitation of a minimum of 6 weeks for the postoperative evaluation period was applied.

3. Results

A total of 41 articles were included in the final review. Final papers are summarized in Table 1, while excluded articles are specified in Table 2.
| Authors                | Reference | Year | Included Studies | Biomaterial Type | Wound Healing Type                          | Principal Findings                                                                                                                                                                                                 |
|------------------------|-----------|------|------------------|------------------|---------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Amghar-Maach et al.    | [8]       | 2019 | 5 (animal studies) | stem cells       | regeneration of periodontal bone            | More bone volume/better periodontal health achieved with dental pulp stem cells (DPSC) grafted as cell sheets. Addition of hepatocyte growth factor (HGF) favors DPSC differentiation. |
| Arora et al.           | [9]       | 2010 | 6 (randomized clinical trials—RCT human studies) | growth factors (PRP) | sinus augmentation procedures                | No statistically significant difference was observed in histological bone density and quality of the regenerated bone.                                                                                                  |
| Barallat et al.        | [10]      | 2014 | 34 (human studies) | bone grafts      | ridge preservation procedures               | Controversial impact of grafting on preservation. Calcium sulfate, porcine xenograft, and magnesium enriched hydroxyapatite (MHA) showed a statistically significant additional benefit when compared with healing by blood clot formation. Demineralized freeze-dried bone allograft (DFDBA) seems to be more efficient than freeze-dried bone allograft (FDBA) in terms of percentage of newly formed bone. |
| Bassetti et al.        | [11]      | 2015 | 24 (18 human and 6 animal studies) | bone grafts      | alveolar ridge splitting/expansion technique (ARST) | ARST seems to be a well-functioning one-stage alternative to prolonged two-stage horizontal grafting procedures.                                                                                                      |
| Browaeys et al.        | [12]      | 2007 | 26 (animal studies) | bone grafts      | sinus augmentation procedures               | Autogenous bone represents the gold standard for sinus augmentation procedures and shows better performances with BBM or porous hydroxyapatite. Homogenous Demineralized Freeze-Dried Bone (DFDB) provides better results than heterogenous DFDB. |
| Chambrone and Tatakis  | [13]      | 2015 | 234 (not specified) | bone grafts and EMD | periodontal regeneration                   | All root coverage (RC) procedures can offer significant drop in recession depth and clinical attachment level (CAL) improvement for Miller class I and II recession-type defects. Subepithelial connective tissue graft-based procedures offered the best results for clinical practice because of their greater percentages of mean and complete RC, together with significant increase in keratinized tissue. |
| Chan et al.            | [14]      | 2013 | 8 (human clinical trials) | bone grafts      | socket preservation procedures             | Conflicting results were found with the use of xenografts on changes in the percentage of vital bone. Partial evidence suggested that alloplasts rise the amount of vital bone formation. Higher % of connective tissue was significantly reduced with the use of bone substitutes. Significant quantities of hydroxyapatite and xenograft particles detected in the healed sockets at an average of 5.6 months after grafting. |
### Table 1. Cont.

| Authors                  | Reference   | Year | Included Studies | Biomaterial Type                  | Wound Healing Type                                    | Principal Findings                                                                                                                                                                                                 |
|--------------------------|-------------|------|------------------|-----------------------------------|-------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Corbella et al.          | [15]        | 2015 | 84 (human studies) | bone grafts | lateral sinus floor elevation procedure | Autogenous bone, bovine bone, and a mixture of tricalcium phosphate (TCP) and hydroxyapatite (HA) show predictable results.                                                                                          |
| Correia et al.           | [16]        | 2017 | 18 (11 clinical trials—CT and 7 RCT) | bone grafts and stem cells | sinus lift procedures | Only a few studies have demonstrated potential of regenerative medicine in sinus lift. After 6 months significant differences were outlined in the vital bone and percentage of residual graft, while results from the connective tissue were not. Same conclusions were described with the use of periosteum-derived cells with collagen matrix. |
| Danesh-Meyer             | [17]        | 2001 | 30 (human studies) | bone grafts and membrane | guided tissue regeneration | Barrier membranes do not show to provide adequate space to predictably support periodontal regeneration in gingival recession defects.                                                                  |
| Darby and Morris         | [18]        | 2013 | 5 (human studies) | growth factors (PDGF-BB) | periodontal regeneration | The use of rhPDGF-BB led to greater CAL gain, augmented rate of bone growth, and greater percentage bone fill related to an osseoconductive control.                                               |
| De Risi et al.           | [19]        | 2013 | 38 (human studies) | bone grafts | ridge preservation procedures | No major histomorphometrical and histological differences arose among different procedures or when compared to spontaneous healing.                                                                                             |
| Del Fabbro et al.        | [20]        | 2017 | 33 (human studies) | growth factors (APC) | socket preservation procedures | Soft tissue healing was statistically better for sockets treated with Autologous Platelet Concentrates (APC) seven days after surgical procedures. New bone was statistically greater for APC group in one study.                                         |
| Del Fabbro et al.        | [21]        | 2018 | 8 (human studies) | growth factors (plasma-rich growth factors—PRGF) | alveolar socket healing | Better and faster epithelialization was observed in the sites treated with PRGF. The measurement of the thickness of the epithelial layer resulted in a thicker layer in the sockets treated with the PRGF.                                       |
| Gharpure and Bhatavadekar| [22]        | 2017 | 26 (18 animal studies and 8 human studies) | bone grafts | bone regeneration | Histological examination of the grafted sites from several studies revealed the generation of a dentin-bone complex, where tooth-bone graft was enclosed by newly forming bone. All reports failed to show complete resorption of the graft material and its substitution by newly formed bone. |
| Giannobile and Somerman  | [23]        | 2003 | 60 (human and animal studies) | growth factors (Enamel matrix derivative—EMD, bone morphogenetic proteins—BMP) | periodontal wound healing | EMD promotes bone regeneration and CAL gain. PDGF-BB promotes periodontal regeneration at the histologic level.                                                                                                                                                  |
| Authors            | Reference | Year | Included Studies | Biomaterial Type                  | Wound Healing Type          | Principal Findings                                                                 |
|--------------------|-----------|------|------------------|-----------------------------------|----------------------------|------------------------------------------------------------------------------------|
| Gupta et al.       | [24]      | 2019 | 16 (10 animal studies and 6 human studies) | statins                          | oral wound healing          | The topical application of simvastatin and chitosan gel could be used as a novel therapeutic approach to improve healing and reduced pain in the palatal donor site following the free gingival grafts (FGG) procedure. |
| Horváth et al.     | [25]      | 2012 | 14 (human studies) | bone grafts                       | ridge preservation procedures | Conflicting evidence exists on the benefit of alveolar ridge preservation (ARP) at the histological level. ARP does not appear to stimulate de novo hard tissue formation routinely. In addition, some graft materials may interfere with healing. |
| Ioannou et al.     | [26]      | 2014 | 5 (human studies) | bone graft (bioactive glass)      | bone regeneration           | The combination of bone graft with autogenous bone chips in a 1:1 ratio is a successful treatment modality for the direct sinus augmentation, with histological results comparable to 100% autogenous bone. |
| Ivanovic et al.    | [27]      | 2014 | 45 (animal studies) | bone grafts                       | periodontal regeneration    | Among the used biomaterials, autografts, in combination with flap surgery, shown the most favorable outcomes, whereas the use of most biologic factors revealed inferior results compared to flap surgery. |
| Kao et al.         | [28]      | 2015 | 124 (human studies) | growth factors (EMD, NBM, PRP, NHA) | periodontal regeneration    | Histologic evidence of periodontal regeneration has been demonstrated when EMD is used in conjunction with nanocrystalline hydroxyapatite (NHA), autogenous bone, a bovine-derived natural bone mineral (NBM), bioactive glass, NBM + PRP, or biphasic calcium phosphate. The most part of the studies indicate no added benefits in either radiographic or clinical gains when EMD is used with the addition of graft materials. |
| MacBeth et al.     | [29]      | 2016 | 9 (human studies) for q1 and 37 (human studies) for q2 | bone grafts                       | ridge preservation procedures | Debated data are available on alterations of width of the keratinised tissue following GBR. |
| Mangano et al.     | [30]      | 2015 | 39 (21 human and 18 animal studies) | stem cells                        | sinus augmentation procedures | BMSCs + PRP composite could give better outcomes in bone volume gain and osteogenesis comparable to that achieved by particulate cancellous bone in MSA. |
| Maroulakos et al.  | [31]      | 2018 | 43 (6 human and 37 animal studies) | scaffold materials and grafts     | bone regeneration           | The immediate and long-term bone repair was considered successful for the time of observation by biochemical, histological, micro-computed tomographic or histomorphometric findings. |
| Authors               | Reference  | Year | Included Studies | Biomaterial Type | Wound Healing Type | Principal Findings                                                                                                                                 |
|----------------------|------------|------|------------------|------------------|-------------------|-----------------------------------------------------------------------------------------------------------------------------------------------------|
| Miron et al.         | [32]       | 2016 | 19 (human and animal studies) | growth factors EMD) | periodontal regeneration | The use of EMD for the management of gingival recessions utilized alone is capable of improving regeneration and enhance soft tissue height/thickness. Application of EMD during conjunction with Coronally Advanced Flap (CAF) resulted in increased formation of alveolar bone, root cementum and periodontal ligament. |
| Monje et al.         | [33]       | 2014 | 15 (human studies) | bone grafts       | maxillary augmentation | Histologic analysis revealed that allogeneic block grafts perform differently in the early stages of healing when compared to autogenous block grafts. |
| Murphy and Gunsolley | [34]       | 2003 | 89 (human studies) | bone grafts       | periodontal regeneration | Augmentation materials procedures, in addition to the physical barrier, enhance the regenerative outcome in the treatment of furcation defects treated with GTR. On the other hand, in the treatment of intrabony defects, there is no advantage to the use of augmentation materials in addition to the use of the physical barrier. |
| Tavelli et al.       | [35]       | 2020 | 63 (human studies) | growth factors (PRGF) | periodontal regeneration | There is strong evidence that recombinant human platelet-derived growth factor (rhPDGF) is efficient in the regeneration of intrabony defects when applied in combination with a bone matrix. In particular, rhPDGF benefits from the delivery with an osteoconductive scaffold matrix. Clinical and histological results confirmed that rhPDGF in combination with a scaffold was also efficient in the treatment of furcation defects. |
| Pranskunas et al.    | [36]       | 2019 | 11 (9 human and 2 animal studies) | stem cells        | socket preservation procedures | The use of bioactive osteogenic molecules or mesenchymal stem cells supports bone regeneration after tooth extraction. Histologically, no particular differences are revealed between test and control groups. |
| Authors           | Reference | Year | Included Studies | Biomaterial Type | Wound Healing Type      | Principal Findings                                                                                                                                 |
|-------------------|-----------|------|------------------|------------------|-------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------|
| Reddy et al.      | [37]      | 2015 | N.S.             | bone grafts      | periodontal regeneration| Histologic proof of periodontal regeneration after the application of a regenerative treatment for the management of maxillary distal, mesial, facial and mandibular lingual or facial Class II furcation defects has been demonstrated in several studies. Evidence of histologic periodontal regeneration in mandibular Class III defects is limited to one case report. Favorable results after a regenerative therapy for maxillary Class III furcation defects are limited to few clinical case reports. In Class I furcation defects, regenerative therapy could be useful in some clinical scenarios, although generally Class I furcation defects may be treated with non-regenerative therapies. |
| Reynolds et al.   | [38]      | 2003 | 49 (human studies)| bone grafts      | regeneration of periodontal bone defects | Demineralized freeze-dried bone allograft (DFDBA) boosts the production of a new attachment apparatus in intrabony defects, while open flap debridement (OFD) results in periodontal repair characterized principally by the formation of a long junctional epithelial attachment. Several observational studies provide a large numbers of histological evidences that autogenous and demineralized allogeneic bone grafts increase the formation of new attachment. Few data also suggest that xenogenic bone grafts can sustenance the formation of a new attachment apparatus. |
| Roca-Millan et al.| [39]      | 2019 | 15 (animal studies)| statins          | Regeneration of Periodontal bone defects | Bovine bone, autogenous bone and a mixture of tricalcium phosphate (TCP) and hydroxyapatite (HA) show predictable results.                                                                                       |
| Rocchietta et al. | [40]      | 2008 | 18 (animal and human studies) | bone grafts | bone regeneration | Only a few studies have established the potential role of regenerative medicine in sinus lift. Statistical significance differences were outlined after 6 months in the vital bone and percentage of residual graft, while outcomes from the connective tissue were not. Same conclusions were described with the use of periosteum-derived cells with collagen matrix. |
| Sculean et al.    | [41]      | 2008 | 10 (Animal Studies) | Bone Grafts      | Periodontal regeneration | Barrier membranes do not provide adequate space to predictably support periodontal regeneration in gingival recession defects.                                                                                     |
Table 1. Cont.

| Authors             | Reference | Year | Included Studies | Biomaterial Type | Wound Healing Type                  | Principal Findings                                                                 |
|---------------------|-----------|------|------------------|------------------|-------------------------------------|----------------------------------------------------------------------------------|
| Sculean et al.      | [42]      | 2015 | 58 (Human Studies) | Bone Grafts      | Periodontal regeneration            | The use of rhPDGF-BB led to greater percentage bone fill, greater CAL gain and increased rate of bone growth compared to an osseoconductive control. |
| Stähli et al.       | [43]      | 2018 | 22 (15 RCT and 7 CCT) | Growth Factors (PRP) | Bone Regeneration                    | No major histomorphometrical and histological differences occurred among different techniques or when compared to spontaneous healing. | Faster and better epithelialization was appreciated in the sites treated with PRGF. The measurement of the thickness of the epithelial layer resulted in a thicker layer in the sockets treated with the PRGF. |
| Strauss et al.      | [44]      | 2018 | 12 (RCT)         | Growth Factors (PRP) | Bone Regeneration                    | Faster and better epithelialization was appreciated in the sites treated with PRGF. The measurement of the thickness of the epithelial layer resulted in a thicker layer in the sockets treated with the PRGF. |
| Varshney et al.     | [45]      | 2020 | 10 (Human Studies) | Stem Cells        | Bone Regeneration                    | Fast tissue healing was statistically better for sockets treated with Autologous Platelet Concentrates (APCs) seven days after surgery procedures. New bone was statistically greater for APC group in one study. |
| Willenbacher et al. | [46]      | 2015 | 18 (Human Studies) | Bone Grafts       | Ridge Preservation Procedures       | Histological examination of the grafted sites from a large number of papers showed the formation of a dentin-bone complex, where tooth-bone graft was enclosed by newly formed bone. All papers failed to demonstrate a complete resorption of the graft material and its substitution by newly formed bone. |
| Yen et al.          | [47]      | 2013 | 22 (Human and Animal Studies) | Bone Grafts | Guided Tissue Regeneration (GTR)     | Despite the quality assessments is different between human and animal studies, some papers suggested that animal models and human results displayed comparable bone-filling ratios in infrabony defects treated with GTR only or with GTR + bone grafting. |
| Zhao H. and Zhao L. | [48]      | 2020 | 5 (Human Studies)  | Bone Grafts       | Socket preservation procedures      | Socket Preservation adopting deproteinized bovine bone mineral (DBBM) did not provide extra benefit regard to post-extraction new bone generation compared to natural healing. |
Table 2. Excluded articles.

| Authors                  | Reference | Year | Reason for Exclusion        |
|--------------------------|-----------|------|----------------------------|
| Castro et al.            | [49]      | 2017 | no histological results     |
| Del Fabbro et al.        | [50]      | 2014 | full text not found         |
| Fawzy El-Sayed et al.    | [51]      | 2019 | different topic             |
| Helgeland et al.         | [52]      | 2018 | different topic             |
| Granate-Marques et al.   | [53]      | 2019 | different topic and comorbidity |
| Monje et al.             | [54]      | 2016 | different topic             |
| Ragucci et al.           | [55]      | 2019 | different topic             |
| Reynolds and Aichelmann-Reidy | [56] | 2012 | no systematic review        |
| Schliephake et al.       | [57]      | 2018 | no histologic results       |

Results are discussed below and divided into specific topics: stems cells, bone grafts, and growth factors.

3.1. Stem Cells

3.1.1. Background

Stem cells can be defined as undifferentiated cells characterized by a set of unique properties. A stem cell is capable of proliferation, self-renewal, production of differentiated daughter cells, self-maintenance of their population, and regeneration of injured tissue. An additional key aspect behind stem cells behavior is the flexibility in their behavior based on environmental conditions [58,59]. Stem cells belong to two main subtypes: pluripotent (or totipotent), able to differentiate in any kind of human cell, and multipotent, that can develop into multiple cell types within their lineage. They can be successfully isolated from the inner part of the blastocyst, prior to the implantation of the embryo, together with fetal and adult tissue. Adult stem cells are generally multipotent and are found in most human tissues, as they support the active cell turnover for tissues undergoing self-renovation at different degrees, and enable tissue repair by replacing damaged or lost cells. Adult stem cells can be harvested from various tissues, such as the bone marrow and the oral cavity. In fact, it is possible to find multipotent adult stem cells in exfoliated deciduous teeth, dental pulp, and periodontal ligaments that show osteogenic and neurogenic capacities [60]. Mesenchymal stem cells obtained from the bone marrow are able to differentiate into various cell types and can respond to the medium where they are inserted to differentiate themselves in the appropriate tissues, as needed [6]. Donor area of choice is usually the iliac crest bone marrow; however, the harvesting procedure from this site may be trivial and painful, so new solutions are being looked for to bypass this kind of inconvenience. Interestingly, mandible periosteum and maxillary tuberosity have been proved as a reliable source of mesenchymal stem cells with osteogenic potential, easy to access under local anesthesia and with low to no post-operative discomfort [61].

3.1.2. Overview of Reviews

In total, six reviews were found on the topic, one considering only animal studies, three only human study, and two accounting for both animal and human studies. The review from Amghar-Maach et al. [8], focused on animal studies, assesses the efficacy of dental pulp stem cells (DPSC) in the regeneration of periodontal defects, but remarks how the biomaterial architecture is relevant to the regeneration outcome. In fact, grafting stem cells in the form of cell sheets leads to better results when compared to the injection of dissociated cells. Additionally, pairing stem cells with growth factor such as hepatocyte growth factor (HGF) may favor DPSC differentiation. Correia et al. [16] and Mangano et al. [30] discuss the impact of mesenchymal stem cells in maxillary sinus augmentation, even if paired up with other biomaterials. Mesenchymal stem cells (MSCs) show a positive impact on wound healing and bone regeneration considering vital bone and vital bone percentage, leading to better outcomes in terms of osteogenesis and bone volume gain. Socket preservation procedures, analyzed by Pranskunas et al. [36],
show an increase in the clinical and radiographical aspects of wound healing in both animal and human studies; however, no significant difference from a histological point of view was found. Considering implant-related bone regeneration procedures [45] (Varshney et al.), both adipose-derived and bone marrow-derived stem cells have been proved to improve the expected results. However, while this efficacy is particularly relevant in animal models, the treatment of large defects in humans does not always relate to a predictable outcome in terms of regeneration. Therefore, even if very promising, stem cell use in the improvement of oral wound healing is not highly predictable. A better understanding of cellular interactions in the healing phases could help overcome this flaw. The combined use of stem cells and growth factors may improve the efficacy of the regenerative approach in a significant way.

3.2. Bone Graft and Resorbable Membranes

3.2.1. Background

Bone grafts are natural or synthetic biomaterials used in the regeneration of defective bone volumes. They can be classified according to their source, microscopic architecture, form, and blood supply [62]. Considering their origin, bone grafts can be defined as:

- Autografts, obtained from the same individual that receives the graft;
- Isografts, from an individual from the same species sharing the same antigenic profile (twins);
- Allografts, harvested from an individual from the same species but with a different antigenic profile;
- Xenografts, obtained from species other than human;
- Alloplastic materials, synthetic bone graft substitutes [63,64].

Moreover, bone grafts can exhibit different properties that provide the rationale for their use in regenerative procedures:

- Osteogenesis: the graft contains living osteoblasts that contribute to new bone formation;
- Osteoinduction: the graft is able to stimulate the differentiation of osteoprogenitor cells into osteoblasts;
- Osteoconduction: the graft acts as a scaffold to sustain the development of capillaries and precursor bone cells [65].

Osteogenesis requires the presence of mesenchymal cells able to differentiate into mature osteoblasts (such in autografts). Osteoinduction usually relies on the presence of growth factors, molecules able to mediate cells recruitment, proliferation, and differentiation, and represents one of the most challenging tasks for the development of bone graft substitutes [66].

Regardless of their osteogenetic and osteoinductive properties, every bone graft has to grant a three-dimensional mechanical structure that hosts and supports cells and extra-cellular matrix [62]. The key feature to a scaffold is the porosity of its structure, since pores increase contact surface of the bone graft, favoring its degradation, and allow cell migration and proliferation [30]. Pore diameter, together with pore morphology and interconnectivity, [67] seem to affect cell behavior, favoring neoangiogenesis with a diameter greater than 300 μm, and osteoblasts migration, adhesion, and proliferation with a diameter of 200–400 μm. Current literature suggests that a porosity of more than 50% by volume and pore sizes of 200–800 μm are the most adequate feature for the development of bone tissue [62,68,69]. These grafting properties are defined as osteocondustive, as described above.

The ideal bone graft should exhibit osteogenic, osteoinductive, and osteoconductive properties while lacking antigenic, teratogenic, or carcinogenic reactions, favor neoangiogenesis, be resorbable, possess a hydrophilic nature, and have low morbidity and cost. Resorbable membranes are devices commonly paired up with bone grafting materials; among them, resorbable collagen membranes (RCMs) are the most commonly found in clinical practices [70]. RCMs are manufactured from allogeneic or xenogeneic sources to manage oral wounds such as extraction sockets, sinus-lift, and ridge augmentation procedures, and periodontal and endodontic surgeries [71–74]. They are one of the essential tools in
guided bone regeneration (GBR) techniques, enhancing wound healing through promotion platelet aggregation, clot stabilization, and fibroblast attraction [75,76]. Time of resorption varies from 2 to 32 weeks and they are biocompatible, easy to manipulate, and with low immunogenicity [77]. They are available as membranes, plugs, or pads for ease of use [70].

3.2.2. Bone Grafts Categories

Autografts

Autografts are often regarded as the ‘gold standard’ among bone grafts due to their osteogenic and osteoinductive properties. Additionally, they are almost safe from the risk of immune reaction/rejection, being harvested from the same subject that receives the graft itself. However, one of their major drawbacks is represented by the necessity of a surgical intervention to collect the graft, and this may affect a patient’s systemic health, increase morbidity, and expose the subject to the risk of chronic postoperative pain and hypersensitivity of the donor area [78]. In oral surgery, autografts are proven to not be able to counteract the volume contraction of the hard tissues of the edentulous sites [79]. Autograft can be harvested by various intra- and extra-oral donor sites. Intraoral donor sites include edentulous ridges, extraction sockets, mandibular ramus, symphysis, and maxillary tuberosity, while extraoral donor sites are tibia, iliac crest, and calvarium [80–95].

Allografts

Allografts are collected from individuals, either dead or alive, of the same species but with a different genotype, processed in order to prevent the host’s immune response and transmission of infectious diseases [89]. They are available as cortical, cancellous, or cortico-cancellous grafts, in various shapes and sizes. Allografts processing has evolved over the years, from the use of fresh frozen bone, simply frozen at −80 °C and no longer used due to the risk of disease transmission and immune response, to the demineralized freeze–dried bone allograft (DFDBA), processed to preserve the organic part that contains bone morphogenetic proteins, growth factors responsible for the graft osteoinductive properties [96].

Xenografts

Xenografts are obtained from donors of a species other than the host’s one, and mostly act as scaffolds showing osteoconductive features and slow resorption time. They can be used both alone and paired up with growth factors or other grafts to enhance their properties. Their lack of osteogenetic and osteoinductive properties is balanced by their availability and relatively low cost. Originating from non-human species, the risk for disease transmission and immunogenicity has to be accounted when using xenografts [97].

Among the many available xenografts, the main categories refer to bovine substitutes, equine substitutes, porcine substitutes, algae substitutes, and coral substitutes.

Alloplastic Materials

Alloplastic materials are biomimetic synthetic bone substitutes, characterized exclusively by osteoconductive features, with no osteoinductive or osteogenic properties. Therefore, they act as a scaffold to support cell migration, proliferation, growth and bone tissue formation [98]. Considering the many chemical and physical properties, they are considered the most heterogeneous group of materials that includes, among the many synthetic bone substitutes, calcium phosphate, calcium carbonate, calcium sulfate, bioactive glasses, and polymers.
3.2.3. Overview of Reviews

A total of twenty-five reviews were found regarding bone grafts and different surgical procedures, such as alveolar socket/ridge preservation, periodontal regeneration, atrophic jaws augmentation, sinus augmentation procedures, and alveolar ridge splitting/expansion technique (ARST).

Preservation procedures found controversial results in literature, with not always clear evidence regarding new bone formation when comparing natural healing to grafts [10,19,31,48].

Magnesium-enriched hydroxyapatite (mHA), calcium sulphate, and porcine grafts granted a better outcome when compared to natural healing, while DFDBA proved to be the most efficient allograft among all [10].

Short term bone wound healing, from 3 to 4 months from dental extraction, shows similar clinical, radiographical, and histological characteristics regardless of the use of a bone graft.

Chan et al. [14] found conflicting results concerning the percentage variation of the vital bone with the use of xenografts, ranging from –22% (decrease) to 9.8% (increase), while connective tissue formation likely decreases with the use of bone substitute. Only limited evidences support an increase in vital bone formation following the use of alloplasts. Significant amounts of hydroxyapatite and xenograft particles (15 to 36%) were found at the healing site at an average of 5.6 months after grafting, as a proof of their stability and resistance to resorption. According to Horvath et al. [25], only a limited cluster of studies report a statistically significant increase in trabecular bone formation when using bone grafts in the alveolar ridge preservation; the use of bone substitute does not prevent ridge resorption but rather delays it, due to the permanence of graft particles inside the healing sockets. From the histological point of view, conflicting evidences are found regarding the benefits of ridge preservation, with no active promotion of bone formation sustained by bone grafts, and rather peculiar histological pictures of what resembles a foreign-body reaction from the host to the bone substitute particles [19,34].

While showing an impact on the reduction in the vertical bone dimension following tooth extraction, socket grafting showed no clear evidence of bone dimensional preservation, bone formation, or keratinised tissue dimensions [29].

On the other hand, Willenbacher et al. [46] found an increase in the preserved bone quota, approximately 1.31 to 1.54 mm bucco-oral bone width and 0.91 to 1.12 mm bone height, in alveolar sockets preserved with grafting materials.

The use of alternative graft solutions, such as tooth-bone graft, demonstrated no added benefits over conventional graft materials [22].

Allogeneic bone blocks represent a good alternative to autologous bone blocks, however, histological analysis highlights differences in their behavior during the healing phases. At 6 months, no connective tissue was found and the presence of inflammatory cells was meaningfully lower when recurring to autologous bone, while in the allogeneic blocks large segments of necrotic bone with empty osteocytes lacunae and little osteoclastic activity were found, along with blood vessels invading the Haversian canals of the graft [33].

The use of bone blocks enables vertical-deficient sites to be rehabilitated with implants in animal models [40].

Advanced atrophic bone augmentation techniques, such as ARST, may benefit from the use of bone grafting to preserve buccal bone height and width [11].

Autologous bone shows the best results in sinus augmentation procedures despite its high resorption rate (40%) in animal studies. This downside can be overcome by mixing it to other bone grafts, such as porous hydroxyapatite or bioglasses [12,15,26].

Periodontal regeneration of intrabony defect using bone grafts proved to be superior in terms of regenerative outcomes when compared to simple flap surgery with no use of biomaterials in both animal and human studies, with autologous bone showing the most favorable results [27,34,47].
The use of grafts combined with membranes proved the best result in terms of periodontal regeneration [37], especially in supra-alveolar and two wall intrabony defects models in animals. Three wall intrabony defects do not benefit consistently of the use of grafts or membrane systems [41].

Regeneration of gingival defects is, however, according to Danesh-Meyer et al. [17], compromised by the wound stabilizing effect of the membrane itself, which does not provide adequate space to promote periodontal regenerational while simultaneously impeding apical migration of the gingival epithelium. Interestingly, alloplastic grafts likely support periodontal repair rather than regeneration [38] and appear to show limited amounts of periodontal regeneration when compared to the other biomaterials [42].

Overall, bone grafts and membranes represent an essential tool in the hard and soft tissue regeneration. In recent years, the efficacy of bone grafts in the socket/ridge preservation techniques has been debated and more and more controversial evidences are emerging, while technology evolves and opens up new scenarios, such as in the case of Computer-Aided Design/Computer-Aided Manufacturing (CAD/CAM) manufactured bone scaffolds. The key to an optimal wound healing relies, as always in the choice of the correct device, in a deep comprehension of their biological and mechanical properties.

3.3. Growth Factors

3.3.1. Background

Growth factors (GFs) are molecules able to regulate DNA synthesis, chemotaxis, matrix synthesis, and promote cellular growth, proliferation, and cellular differentiation, usually of proteic or steroid nature.

Among the wide category of growth factors, some have polarized the research attention over the recent years, such as:

- Bone morphogenic proteins (BMPs): cytokines able to stimulate bone cell differentiation and promote new bone formation, responsible for the osteoinductive features of bone grafts;
- Fibroblast growth factor (FGF) and vascular endothelial growth factors (VEGF): mostly stimulate neoangiogenesis;
- Platelet-derived growth factor (PDGF): known as one of the initiators of wound healing, with multiple functions ranging from chemotaxis and mitogenesis to promotion of angiogenesis, acts on both soft and hard tissues;
- Transforming growth factor beta (TGF-β) and insulin-like growth factor (IGF): regulate collagen and fibronectin synthesis through osteoblasts or fibroblasts stimulation;
- Amelogenins: extracellular matrix proteins secreted by ameloblasts that regulate hydroxyapatite crystal growth and orientation and are able to promote periodontal tissues regeneration; in the clinical practice, they are commonly found in enamel matrix derivatives (EMD) compounds, a mix of enamel matrix proteins (EMP), of which amelogenins represent circa 90% of the total protein quota [99,100];
- Statins: recently discovered to possess anti-inflammatory, antimicrobial and pro-osteogenic properties.

An important biological vehicle responsible for delivering GFs to wounded sites is represented by platelets that, in addition to their procoagulant effect, release many biomolecules like PDGF, TGF-β, VEGF, etc.

Therefore, despite the availability of recombinant GFs, the use of autologous platelet concentrates has found many applications in oral surgery. Platelet concentrates belong to four main categories:

1. Pure Platelet Rich Plasma (P-PRP) or leukocyte-poor PRP that does not contain leukocytes;
2. Leukocyte and PRP (L-PRP) products;
3. Pure Platelet Rich Fibrin (P-PRF) or leukocyte-poor PRF;
4. Leukocyte and PRF (L-PRF).

PRP is a first-generation platelet concentrate containing platelets in super-natural concentration and minimal amount of natural fibrinogen. Platelets’ α granules are responsible for the release of growth factors within 3–5 days of platelet activation, which sustain their stimulation of proliferative phase for 10 days after release. However, calcium chloride and bovine thrombin are added to reach gel consistency and these components may interfere with wound healing [101,102].

Preparation rich in growth factors (PRGF-Endoret) technology was invented as an answer to some of the limitations of PRP preparations. The clot activator, calcium chloride, leads to the formation of native thrombin. This mimicked physiological clotting process enables a more sustained release of growth factors. Moreover, this procedure reduces the risk of immunological reactions and disease transmission associated with the use of exogenous bovine thrombin [103].

PRF represents a new generation platelet concentrate, an evolution of PRP. Similar to the blood clot, it is a tetramolecular fibrin matrix that contains all the molecular and cellular elements, such as platelets, leukocytes, cytokines, and circulating stem cells, that promote healing simultaneously being more stable and homogenous. Furthermore, 20 PRF does not require addition of bovine thrombine or other substances, thus it does not share the coagulant-related drawbacks of PRP [104,105].

3.3.2. Overview of Reviews

A total of thirteen reviews were considered regarding the use of growth, both autologous or recombinant.

In sinus augmentation, PRP does not significantly affect the histological density and quality of the regenerated bone; however, early wound healing was observed [9].

According to Stähli et al. [43], while having no evidence supporting the clinical benefit of PRP in healthy patients, PRP might have a positive effect on wound healing and bone regeneration in compromised patients.

PRF, however, showed superior outcomes in bone regeneration procedures, as per ridge dimension, bone regeneration, osseointegration process, and soft tissue healing [44].

According to Darby and Morris [18], periodontal regeneration performed through the use of PDGF led to greater CAL gain of around 1mm, a greater percentage bone fill of around 40%, and an increased rate of bone growth, compared to an osteoconductive control (β-TCP), with no particular adverse effects. This consideration is backed up by the systematic review of Giannobile and Somerman [23], assessing that PDGF promotes periodontal regeneration at the histological level.

The efficacy of PDGF may be further improved, associating it with an osteoconductive scaffold matrix [35].

Similarly, EMD were found to consistently promote CAL gain and probing depth reduction when compared to flap surgery alone, and its effect is improved using it in combination with graft materials [23,28].

The efficacy of EMD is further highlighted in the treatment of gingival recession [16], improving soft tissue height and thickness; using EMD together with coronally advanced flaps in root coverage seemingly leads to periodontal regeneration with formation of root cementum, periodontal ligament, and alveolar bone [32]. Platelet concentrates showed positive effects on the healing outcome of both soft and hard tissue in the post-extraction alveolar socket, with a significant increase in the keratinized mucosa quota and in the new bone formation percentage, although this result is controversial [20,21].

Local application of statins shows an apparent osteogenic and angiogenic effect in periodontal defects models in animal studies; topical simvastatin enhances wound healing and improves patient outcomes, stimulating bone formation, promoting soft tissue healing, as well as reducing post-operative pain and inflammation [24,39].
4. Conclusions

The development of biomaterials represents one of the most promising perspectives for the future of oral surgery. In particular, stem cells and growth factors are polarizing the focus of this ever-evolving field, continuously improving standard surgical techniques, and granting access to new approaches. Bone grafts and membranes usually play a pivotal role in GBR procedures. Despite their long history as essential tools in regenerative procedures, controversial evidences are emerging regarding the socket/ridge preservation techniques, that represent the basic approach in the modern oral surgery to the post-extraction socket and edentulous ridge. Technology evolves and opens up new scenarios, such as in the case of CAD/CAM manufactured bone scaffolds.

The regenerative properties of the biomaterials used in oral surgery may be improved thanks to growth factors. Their combined use, in fact, likely enhances the healing processes and favors early wound healing after oral surgical procedures.

Author Contributions: Conceptualization, G.P. (Giacomo Picciolo) and M.P.; methodology, G.P. (Giacomo Picciolo) and M.P.; investigation, G.P. (Giacomo Picciolo) and M.P.; resources, G.O.; data curation, G.P. (Giovanni Pallio), G.P. (Giuseppe Picciolo) and A.S.; writing—original draft preparation, G.P. (Giacomo Picciolo) and M.P.; writing—review and editing, N.I., M.V. and D.A.; supervision, F.S.; project administration, F.S. and G.O.; funding acquisition, F.S. and G.O. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Conflicts of Interest: The authors declare no conflict of interest. The SunNutraPharma had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

1. Saunders, W.B. Introduction. In Current Therapy in Oral and Maxillofacial Surgery; Bagheri, S.C., Bell, R.B., Khan, H.A., Eds.; W.B. Saunders: Saint Louis, MO, USA, 2012; p. 122. ISBN 978-1-4160-2527-6.
2. AlQurashi, H.; Khurshid, Z.; Yaqin, S.A.U.; Habib, S.R.; Rokaya, D.; Zafar, M.S. Polyetherketoneketone (PEKK): An emerging biomaterial for oral implants and dental prostheses. J. Adv. Res. 2020. [CrossRef]
3. Khurshid, Z.; Zafar, M.S.; Najeeb, S.; Nejatian, T.; Sefat, F. Introduction to dental biomaterials and their advances. In Advanced Dental Biomaterials; Khurshid, Z., Najeeb, S., Zafar, M.S., Sefat, F., Eds.; Woodhead Publishing: Cambridge, UK, 2019; pp. 1–5. ISBN 978-0-08-102476-8.
4. Zafar, M.S.; Ullah, R.; Qamar, Z.; Fareed, M.A.; Amin, F.; Khurshid, Z.; Sefat, F. Properties of dental biomaterials. In Advanced Dental Biomaterials; Khurshid, Z., Najeeb, S., Zafar, M.S., Sefat, F., Eds.; Woodhead Publishing: Cambridge, UK, 2019; pp. 7–35. ISBN 978-0-08-102476-8.
5. Tanveer, W. Biomaterials for maxillofacial prosthetic rehabilitation. In Advanced Dental Biomaterials; Khurshid, Z., Najeeb, S., Zafar, M.S., Sefat, F., Eds.; Woodhead Publishing: Cambridge, UK, 2019; pp. 615–641. ISBN 978-0-08-102476-8.
6. Anil, S.; Chaliserry, E.P.; Nam, S.Y.; Venkatesan, J. Biomaterials for craniofacial tissue engineering and regenerative dentistry. In Advanced Dental Biomaterials; Khurshid, Z., Najeeb, S., Zafar, M.S., Sefat, F., Eds.; Woodhead Publishing: Cambridge, UK, 2019; pp. 643–674. ISBN 978-0-08-102476-8.
7. Ferracane, J.L.; Giannobile, W.V. Novel biomaterials and technologies for the dental, oral, and craniofacial structures. J. Dent. Res. 2014, 93, 1185–1186. [CrossRef] [PubMed]
8. Amghar-Maach, S.; Gay-Escoda, C.; Sánchez-Garcés, M. Àngeles Regeneration of periodontal bone defects with dental pulp stem cells grafting: Systematic Review. J. Clin. Exp. Dent. 2019, 11, e373–e381. [CrossRef] [PubMed]
9. Arora, N.S.; Ramanayake, T.; Ren, Y.-F.; Romanos, G.E. Platelet-rich plasma in sinus augmentation procedures: A systematic literature review: Part II. Implant. Dent. 2010, 19, 145–157. [CrossRef]
10. Barallat, L.; Ruiz-Magaz, V.; Levi, P.A.; Mareque-Bueno, S.; Galindo-Moreno, P.; Nart, J. Histomorphometric results in ridge preservation procedures comparing various graft materials in extraction sockets with Nongrafted sockets in humans: A systematic review. Implant. Dent. 2014, 23, 539–554. [CrossRef]
11. Bassetti, M.A.; Bassetti, R.G.; Bosshardt, D.D. The alveolar ridge splitting/expansion technique: A systematic review. Clin. Oral Implant. Res. 2015, 27, 310–324. [CrossRef]
12. Browaeys, H.; Bouvy, P.; De Bruyn, H. A literature review on biomaterials in sinus augmentation procedures. *Clin. Implant. Dent. Relat. Res.* 2007, 9, 166–177. [CrossRef]

13. Chambrone, L.; Tatakos, D.N. Periodontal soft tissue root coverage procedures: A systematic review from the AAP regeneration workshop. *J. Periodontol.* 2015, 86, S8–S51. [CrossRef]

14. Chan, H.-L.; Lin, G.-H.; Fu, J.-H.; Wang, H.-L. Alterations in bone quality after socket preservation with grafting materials: A systematic review. *Int. J. Oral Maxillofac. Implant.* 2013, 28, 710–720. [CrossRef]

15. Corbella, S.; Taschieri, S.; Weinstein, R.L.; Del Fabbro, M. Histomorphometric outcomes after lateral sinus floor elevation procedure: A systematic review of the literature and meta-analysis. *Clin. Oral Implant. Res.* 2015, 27, 1106–1122. [CrossRef]

16. Correia, F.; Pozza, D.H.; Gouveia, S.; Felino, A.C.; Almeida, R.F. The applications of regenerative medicine in sinus lift procedures: A systematic review. *Clin. Implant. Dent. Relat. Res.* 2017, 20, 229–242. [CrossRef] [PubMed]

17. Danesh-Meyer, M.J.; Wikesjo, U.M.E. Gingival recession defects and guided tissue regeneration: A review. *J. Periodontal Res.* 2001, 36, 341–354. [CrossRef] [PubMed]

18. Darby, I.B.; Morris, K.H. A systematic review of the use of growth factors in human periodontal regeneration. *J. Periodontol.* 2013, 84, 465–476. [CrossRef] [PubMed]

19. De Risi, V.; Clementini, M.; Vittorini, G.; Mannocci, A.; De Sanctis, M. Alveolar ridge preservation techniques: A systematic review and meta-analysis of histological and histomorphometrical data. *Clin. Oral Implant. Res.* 2013, 26, 50–68. [CrossRef]

20. Del Fabbro, M.; Bucchi, C.; Lolato, A.; Corbella, S.; Testori, T.; Taschieri, S. Healing of Postextraction sockets preserved with autologous platelet concentrates. A systematic review and meta-analysis. *J. Oral Maxillofac. Surg.* 2017, 75, 1601–1615. [CrossRef]

21. Del Fabbro, M.; Panda, S.; Taschieri, S. Adjunctive use of plasma rich in growth factors for improving alveolar socket healing: A systematic review. *J. Evid. Based Dent. Pract.* 2018, 19, 166–176. [CrossRef]

22. Gharpure, A.S.; Bhatavadekar, N.B. Clinical efficacy of tooth-bone graft: A systematic review and risk of bias analysis of randomized control trials and observational studies. *Implant. Dent.* 2018, 27, 119–134. [CrossRef]

23. Giannobile, W.V.; Somerman, M.J. Growth and Amelogenin-like factors in periodontal wound healing. *Clin. Oral Investig.* 2003, 8, 193–204. [CrossRef]

24. Gupta, S.; Del Fabbro, M.; Chang, J. The impact of simvastatin intervention on the healing of bone, soft tissue, and TMJ cartilage in dentistry: A systematic review and meta-analysis. *Int. J. Implant. Dent.* 2019, 5, 17. [CrossRef]

25. Horváth, A.; Mardas, N.; Mezzomo, L.A.; Needleman, I.; Donos, N. Alveolar ridge preservation. A systematic review. *Clin. Oral Investig.* 2012, 17, 341–363. [CrossRef]

26. Ioannou, A.; Kotsakis, G.A.; Kumar, T.; Hinrichs, J.E.; Romanos, G. Evaluation of the bone regeneration potential of bioactive glass in implant site development surgeries: A systematic review of the literature. *Clin. Oral Investig.* 2014, 19, 181–191. [CrossRef]

27. Ivanovic, A.; Nikou, G.; Miron, R.J.; Nikolidakis, D.; Sculean, A. Which biomaterials may promote periodontal regeneration in intrabony periodontal defects? A systematic review of preclinical studies. *Quintessence Int.* 2014, 45, 385–395.

28. Kao, R.T.; Nares, S.; Reynolds, M.A. Periodontal regeneration—Intrabony defects: A systematic review from the AAP regeneration workshop. *J. Periodontol.* 2015, 86, S77–S104. [CrossRef] [PubMed]

29. Macbeth, N.; Trullenque-Eriksson, A.; Donos, N.; Mardas, N. Hard and soft tissue changes following alveolar ridge preservation: A systematic review. *Clin. Oral Implant. Res.* 2016, 28, 982–1004. [CrossRef] [PubMed]

30. Mangano, F.G.; Colombo, M.; Veronesi, G.; Caprioglio, A.; Mangano, C. Mesenchymal stem cells in maxillary sinus augmentation: A systematic review with meta-analysis. *World J. Stem Cells* 2015, 7, 976–991. [CrossRef]

31. Maroulakos, M.; Kamperos, G.; Tayebi, L.; Halazonetis, D.; Ren, Y. Applications of 3D printing on craniofacial bone repair: A systematic review. *J. Dent.* 2019, 80, 1–14. [CrossRef] [PubMed]

32. Miron, R.J.; Sculean, A.; Cochrane, D.L.; Froum, S.; Zucchelli, G.; Nemcovsky, C.; Donos, N.; Lyngstadaas, S.P.; Deschner, J.; Dard, M.; et al. Twenty years of enamel matrix derivative: The past, the present and the future. *J. Clin. Periodontol.* 2016, 43, 668–683. [CrossRef] [PubMed]

33. Monje, A.; Pikos, M.A.; Chan, H.-L.; Suarez, F.; Gargallo-Albiol, J.; Hernández-Alfaro, F.; Galindo-Moreno, P.; Wang, H.-L. On the feasibility of utilizing allogeneic bone blocks for atrophic maxillary augmentation. *BioMed Res. Int.* 2014, 2014, 1–12. [CrossRef]
34. Murphy, K.G.; Gunsolley, J.C. Guided tissue regeneration for the treatment of periodontal Intrabony and furcation defects. A systematic review. *Ann. Periodontol.* 2003, 8, 266–302. [CrossRef]
35. Tavelli, L.; Ravidà, A.; Barootti, C.; Chambrone, L.; Giannobile, W.V. Recombinant human platelet-derived growth factor: A systematic review of clinical findings in oral regenerative procedures. *JDR Clin. Transl. Res.* 2020. [CrossRef]
36. Pranskunas, M.; Galindo-Moreno, P.; Padial-Molina, M. Extraction socket preservation using growth factors and stem cells: A systematic review. *J. Oral Maxillofac. Res.* 2019, 10, e7. [CrossRef] [PubMed]
37. Reddy, M.S.; Aichelmann-Reidy, M.E.; Avila-Ortiz, G.; Klokkevold, P.R.; Murphy, K.G.; Rosen, P.S.; Schallhorn, R.G.; Sculean, A.; Wang, H.-L. Periodontal regeneration—Furcation defects: A consensus report from the AAP regeneration workshop. *J. Periodontol.* 2015, 86, S131–S133. [CrossRef] [PubMed]
38. Reynolds, M.A.; Aichelmann-Reidy, M.E.; Branch-Mays, G.L.; Gunsolley, J.C. The application of statins in the regeneration of bone defects. Systematic review and meta-analysis. *Int. Endod. J.* 2013, 46, 106–116. [CrossRef] [PubMed]
39. Sculean, A.; Nikolidakis, D.; Schwarz, F. Regeneration of periodontal tissues: Combinations of barrier membranes and grafting materials—Biological foundation and preclinical evidence: A systematic review. *J. Clin. Periodontol.* 2008, 35, 203–215. [CrossRef] [PubMed]
40. Sculean, A.; Nikolidakis, D.; Spindler, M.; Ivanovic, A.; Chapple, I.L.C.; Stavropoulos, A. Biomaterials for promoting periodontal regeneration in human intrabony defects: A systematic review. *Periodontology 2000* 2015, 68, 182–216. [CrossRef]
41. Stähli, A.; Strauss, F.J.; Gruber, R. The use of platelet-rich plasma to enhance the outcomes of implant therapy: A systematic review. *Clin. Oral Implant. Res.* 2018, 29, 20–36. [CrossRef]
42. Strauss, F.J.; Stähli, A.; Gruber, R. The use of platelet-rich fibrin to enhance the outcomes of implant therapy: A systematic review. *Clin. Oral Implant. Res.* 2018, 29, 6–19. [CrossRef]
43. Willenbacher, M.; Al-Nawas, B.; Berres, M.; Kämmerer, P.W.; Schiegnitz, E. The effects of alveolar ridge preservation: A meta-analysis. *Clin. Implant. Dent. Relat. Res.* 2015, 18, 1248–1268. [CrossRef]
44. Yen, C.-C.; Tu, Y.-K.; Chen, T.-H.; Lu, H.-K. Comparison of treatment effects of guided tissue regeneration on infrabony lesions between animal and human studies: A systematic review and meta-analysis. *J. Periodontal Res.* 2013, 48, 415–424. [CrossRef]
45. Zhao, H.; Hu, J.; Zhao, L. Histological analysis of socket preservation using DBBM. A systematic review and meta-analysis. *J. Stomatol. Oral Maxillofac. Surg.* 2020. [CrossRef] [PubMed]
46. Castro, A.B.; Meschi, N.; Temmerman, A.; Pinto, N.; Lambrechts, P.; Teughels, W.; Quirynen, M. Regenerative potential of leucocyte- and platelet-rich fibrin. Part A: Intra-bony defects, furcation defects and periodontal plastic surgery. A systematic review and meta-analysis. *J. Clin. Periodontol.* 2017, 44, 67–82. [CrossRef]
47. Del Fabbro, M.; Corbella, S.; Taschieri, S.; Francetti, L.; Weinstein, R. Autologous platelet concentrate for post-extraction socket healing: A systematic review. *Eur. J. Oral Implantol.* 2014, 7, 333–344. [CrossRef]
48. Fawzy El-Sayed, K.M.; Ahmed, G.M.; Abouaouf, E.A.; Schwendicke, F. Stem/progenitor cell-mediated pulpal tissue regeneration: A systematic review and meta-analysis. *Int. Endod. J.* 2019, 52, 1573–1585. [CrossRef]
49. Helgeland, E.; Shanbhag, S.; Pedersen, T.O.; Mustafa, K.; Rosén, A. Scaffold-Based Temporomandibular Joint Tissue Regeneration in Experimental Animal Models: A Systematic Review. *Tissue Eng. Part B Rev.* 2018, 24, 300–316. [CrossRef] [PubMed]
50. Granate-Marques, A.; Polis-Yanes, C.; Seminario-Arnez, M.; Jané-Salas, E.; López-López, J. Medication-related osteonecrosis of the jaw associated with implant and regenerative treatments: Systematic review. *Med. Oral Patol. Oral Cir. Bucal.* 2019, 24, e195–e203. [CrossRef]
54. Monje, A.; Diaz, K.T.; Aranda, L.; Insua, A.; Garcia-Nogales, A.; Wang, H.L. Schneiderian Membrane Thickness and Clinical Implications for Sinus Augmentation: A Systematic Review and Meta-Regression Analyses. J. Periodontol. 2016, 87, 888–899. [CrossRef] [PubMed]

55. Ragucci, G.M.; Almazrooa, S.A.; Noonan, V.; Wang, H.L.; Hernández-Alfaro, F.; Gargallo-Albiol, J. Influence of exposing dental implants into the sinus cavity on survival and complications rate: A systematic review. Int. J. Implant. Dent. 2019, 5, 6. [CrossRef]

56. Reynolds, M.A.; Aichelmann-Reidy, M.E. Protein and peptide-based therapeutics in periodontal regeneration. J. Evid. Based Dent. Pract. 2012, 12, 118–126. [CrossRef]

57. Schliephake, H.; Sicilia, A.; Nawas, B.A.; Donos, N.; Gruber, R.; Jepsen, S.; Milinkovic, I.; Mombelli, A.; Navarro, J.M.; Quirynen, M.; et al. Drugs and diseases: Summary and consensus statements of group 1. The 5th EAO Consensus Conference 2018. Clin. Oral Implants Res. 2018, 29, 93–99. [CrossRef] [PubMed]

58. Potten, C.S. Stem cells: Attributes, cycles, spirals, pitfalls and uncertainties. Lessons for and from the crypt. Development 1990, 110, 1001–1020.

59. Loeffler, M.; Roeder, I. Tissue Stem Cells: Definition, Plasticity, Heterogeneity, Self-Organization and Models—A Conceptual Approach. Cells Tissues Organs 2002, 171, 8–26. [CrossRef] [PubMed]

60. Mosquera-Perez, R.; Fernández-Olavarri, A.; Diaz-Sanchez, R.-M.; Gutierrez-Perez, J.-L.; Serrera-Figallo, M.-Á.; Torres-Lagares, D. Stem cells and oral surgery: A systematic review. J. Clin. Exp. Dent. 2019, 11, e1181–e1189. [CrossRef] [PubMed]

61. Almazrooa, S.A.; Noonan, V.; Wang, H.L.; Hernández-Alfaro, F.; Gargallo-Albiol, J. Influence of exposing dental implants into the sinus cavity on survival and complications rate: A systematic review. Int. J. Implant. Dent. 2019, 5, 6. [CrossRef]

62. Schliephake, H.; Sicilia, A.; Nawas, B.A.; Donos, N.; Gruber, R.; Jepsen, S.; Milinkovic, I.; Mombelli, A.; Navarro, J.M.; Quirynen, M.; et al. Drugs and diseases: Summary and consensus statements of group 1. The 5th EAO Consensus Conference 2018. Clin. Oral Implants Res. 2018, 29, 93–99. [CrossRef] [PubMed]

63. Luitaud, C.; Laflamme, C.; Semlali, A.; Saidi, S.; Grenier, G.; Zakrzewski, A.; Rouabhia, M. Development of an engineering autologous palatal mucosa-like tissue for potential clinical applications. J. Biomed. Mater. Res. Part B Appl. Biomater. 2007, 83, 554–561. [CrossRef]
76. Yamada, M.; Kubo, K.; Ueno, T.; Iwasa, F.; Att, W.; Hori, N.; Ogawa, T. Alleviation of commercial collagen sponge- and membrane-induced apoptosis and dysfunction in cultured osteoblasts by an amino acid derivative. *Int. J. Oral Maxillofac. Implant.* 2010, 25, 939–946.

77. Bunyaratavej, P.; Wang, H.-L. Collagen membranes: A review. *J. Periodontol.* 2001, 72, 215–229. [CrossRef] [PubMed]

78. Chiapasco, M.; Casentini, P.; Zaniboni, M. Bone augmentation procedures in implant dentistry. *Int. J. Oral Maxillofac. Implant.* 2009, 24, 237–259.

79. Araújo, M.G.; Da Silva, J.C.C.; De Mendonça, A.F.; Lindhe, J. Ridge alterations following grafting of fresh extraction sockets in man. A randomized clinical trial. *Clin. Oral Implant. Res.* 2014, 26, 407–412. [CrossRef] [PubMed]

80. Iizuka, T.; Smolka, W.; Hallermann, W.; Mericske-Stern, R. Extensive augmentation of the alveolar ridge using autogenous calvarial split bone grafts for dental rehabilitation. *Clin. Oral Implant. Res.* 2004, 15, 607–615. [CrossRef]

81. Kökat, A.M.; Erçöçen, A.R.; Karayazgan-Saraço˘glu, B. Simultaneous implant placement in a mandibular defect reconstructed with a free fibula graft and fabrication of a fixed metal porcelain prosthesis. *J. Craniofacial Surg.* 2018, 29, 1307–1311. [CrossRef]

82. Osborn, T.M.; Helal, D.; Mehra, P. Iliac crest bone grafting for mandibular reconstruction: 10-year experience outcomes. *J. Oral Biol. Craniofacial Res.* 2017, 8, 25–29. [CrossRef]

83. Jeong, E.C.; Yoon, S.; Jung, Y.H. Radial forearm Osteocutaneous free flap for reconstruction of hard palate with alveolar defect. *J. Craniofacial Surg.* 2017, 28, e438-e440. [CrossRef]

84. Lee, Y.H.; Choi, J.H.; Hwang, K.; Choi, J.H. Rib bone graft adjusted to fit the facial asymmetry: A frame structure graft. *J. Craniofacial Surg.* 2015, 26, 2160–2162. [CrossRef]

85. Atil, F.; Kocyigit, I.D.; Suer, B.T.; Alp, Y.E.; Yazan, M.; Tekin, U.; Tuz, H.H. Clinical evaluation of the use of Tibial bone grafting in Dentoalveolar reconstructive surgery. *Med. Princ. Pract.* 2015, 24, 72–78. [CrossRef]

86. Bande, C.R.; Daware, S.; Lambade, P.; Patle, B. Reconstruction of orbital floor fractures with Autogenous bone graft application from anterior wall of maxillary sinus: A retrospective study. *J. Maxillofac. Oral Surg.* 2014, 14, 605–610. [CrossRef] [PubMed]

87. Cho, Y.-S.; Hwang, K.-G.; Park, C. Postoperative effects of anterior nasal spine bone harvesting on overall nasal shape. *Clin. Oral Implant. Res.* 2012, 24, 618–622. [CrossRef] [PubMed]

88. Güngörmüş, M.; Yavuz, M.S. The ascending ramus of the mandible as a donor site in maxillofacial bone grafting. *J. Oral Maxillofac. Surg.* 2002, 60, 1316–1318. [CrossRef] [PubMed]

89. Sabhlok, S.; Waknis, P.; Gadre, K.S. Applications of coronoid process as a bone graft in maxillofacial surgery. *J. Craniofacial Surg.* 2014, 25, 577–580. [CrossRef]

90. Peñarrocha-Oltra, D.; García-Mira, B.; Martínez, O. Localized vertical maxillary ridge preservation using bone cores and a rotated palatal flap. *Int. J. Oral Maxillofac. Implant.* 2005, 20, 131–134.

91. Velásquez, H.; Olate, S.; Diaz, C.; Navarro, P.; Borie, E.; De Moraes, M. Quantitation of mandibular symphysis bone as source of bone grafting: Description in class I and class III skeletal conditions. *J. Oral Implant.* 2017, 43, 211–217. [CrossRef]

92. Tolstunov, L. Maxillary tuberosity bone block graft: Innovative technique and case report. *J. Oral Maxillofac. Surg.* 2009, 67, 1723–1729. [CrossRef]

93. Hernández-Alfaro, F.; Pages, C.M.; García, E.; Corchero, G.; Arranz, C. Palatal core graft for alveolar reconstruction: A new donor site. *Int. J. Oral Maxillofac. Implant.* 2005, 20, 777–783.

94. Ganz, S.D. Mandibular tori as a source for onlay bone graft augmentation: A surgical procedure. *Pract. Periodontics Aesthetic Dent. PPAD* 1998, 9, 973–982.

95. Kainulainen, V.T.; Sándor, G.K.; Clokie, C.; Keller, A.; Oikarinen, K. The zygomatic bone as a potential donor site for alveolar reconstruction—A quantitative anatomic cadaver study. *Int. J. Oral Maxillofac. Surg.* 2004, 33, 786–791. [CrossRef]

96. Sheikh, Z.; Hamdan, N.; Ikeda, Y.; Grynpas, M.; Ganss, B.; Glogauer, M. Natural graft tissues and synthetic biomaterials for periodontal and alveolar bone reconstructive applications: A review. *Biomater. Res.* 2017, 21, 9. [CrossRef] [PubMed]

97. Tovar, N.; Jimbo, R.; Gangolli, R.; Perez, L.; Manne, L.; Yoo, D.-Y.; Lorenzoni, F.; Witek, L.; Coelho, P. Evaluation of bone response to various anorganic bovine bone xenografts: An experimental calvaria defect study. *Int. J. Oral Maxillofac. Surg.* 2014, 43, 251–260. [CrossRef] [PubMed]
98. Alghamdi, A.S.; Shibly, O.; Ciancio, S.G. Osseous grafting part II: Xenografts and alloplasts for periodontal regeneration—A literature review. *J. Int. Acad. Periodontol.* 2010, 12, 39–44. [PubMed]

99. Bansal, A.K.; Shetty, D.C.; Bindal, R.; Pathak, A. Amelogenin: A novel protein with diverse applications in genetic and molecular profiling. *J. Oral Maxillofac. Pathol.* 2012, 16, 395–399. [CrossRef]

100. Wyganowska-Switkowska, M.; Urbaniak, P.; Nohawica, M.; Kotwicka, M.; Jankun, J. Enamel matrix proteins exhibit growth factor activity: A review of evidence at the cellular and molecular levels. *Exp. Ther. Med.* 2015, 9, 2025–2033. [CrossRef]

101. Mohan, S.P.; Jaishangar, N.; Devy, S.; Narayanan, A.; Cherian, D.; Madhavan, S.S. Platelet-rich plasma and platelet-rich fibrin in periodontal regeneration: A review. *J. Pharm. Bioallied Sci.* 2019, 11, S126–S130. [CrossRef]

102. Kobayashi, E.; Flückiger, L.; Fujioka-Kobayashi, M.; Sawada, K.; Sculean, A.; Schaller, B.; Miron, R.J. Comparative release of growth factors from PRP, PRF, and advanced-PRF. *Clin. Oral Investig.* 2016, 20, 2353–2360. [CrossRef]

103. Giannini, S.; Cielo, A.; Bonanome, L.; Rastelli, C.; Derla, C.; Corpaci, F.; Falisi, G. Comparison between PRP, PRGF and PRF: Lights and shadows in three similar but different protocols. *Eur. Rev. Med. Pharmacol. Sci.* 2015, 19, 927–930.

104. Feigin, K.; Shope, B. Use of platelet-rich plasma and platelet-rich fibrin in dentistry and oral surgery: Introduction and review of the literature. *J. Veter Dent.* 2019, 36, 109–123. [CrossRef]

105. Albanese, A.; Licata, M.; Polizzi, B.; Campisi, G. Platelet-rich plasma (PRP) in dental and oral surgery: From the wound healing to bone regeneration. *Immun. Ageing* 2013, 10, 1–23. [CrossRef]

**Publisher’s Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.

© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).