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Sleep deprivation reduces the density of individual spine subtypes in a branch-specific fashion in CA1 neurons

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Summary
Sleep deprivation has a negative impact on hippocampus-dependent memory, which is thought to depend on cellular plasticity. We previously found that 5 h of sleep deprivation robustly decreases dendritic spine density in the CA1 area of the hippocampus in adult male mice. However, recent work by others suggests that sleep deprivation increases the density of certain spine types on specific dendritic branches. Based on these recent findings and our previous work, we conducted a more in-depth analysis of different spine types on branches 1, 2 and 5 of both apical and basal dendrites to assess whether 5 h of sleep deprivation may have previously unrecognized spine-type and branch-specific effects. This analysis shows no spine-type specific changes on branch 1 and 2 of apical dendrites after sleep deprivation. In contrast, sleep deprivation decreases the number of mushroom and branched spines on branch 5. Likewise, sleep deprivation reduces thin, mushroom and filopodia spine density on branch 5 of the basal dendrites, without affecting spines on branch 1 and 2. Our findings indicate that sleep deprivation leads to local branch-specific reduction in the density of individual spine types, and that local effects might not reflect the overall impact of sleep deprivation on CA1 structural plasticity. Moreover, our analysis underscores that focusing on a subset of dendritic branches may lead to potential misinterpretation of the overall impact of, in this case, sleep deprivation on structural plasticity.

KEYWORDS
CA1, dendritic spines, hippocampus, sleep, structural plasticity, synaptic plasticity

1 | INTRODUCTION

A lack of sleep is a common problem in our 24/7 society and has a detrimental impact on brain functioning. Work by numerous laboratories indicated that the hippocampus is one of the brain regions particularly vulnerable to sleep deprivation (reviewed in Havekes & Aton, 2020; Kreutzmann et al., 2015; Raven et al., 2018). Sleep deprivation not only impacts fundamental hippocampus-dependent processes such as memory, but can also lead to the shrinkage of hippocampal volume (Novati et al., 2011).

Biochemical analyses of adult mice hippocampi revealed that as little as 5-6 h of sleep deprivation leads to numerous changes in molecular and cellular processes (Havekes et al., 2012), including the overall reduction of spine density in the hippocampal subregions CA1 (Acosta-peña et al., 2015; Havekes et al., 2016; Wong et al., 2019) and dentate gyrus (DG) (Raven et al., 2019), but not in area CA3 (Havekes et al., 2016). Further examination of the CA1 subregion revealed that the reduction in spine density is similar across apical and basal dendrites (Havekes et al., 2016; Wong et al., 2019). However, the reduction in spine density is not uniform along dendrites, but...
varies depending on the distance from the soma and branch number (Havekes et al., 2016; Raven et al., 2019). Indeed, sleep deprivation decreases spine numbers in both apical and basal CA1 neurons at a distance of approximately 30–150 µm from the soma (Havekes et al., 2016), whereas in the DG, the reduction in spines is found closer to the soma (0–30 µm) (Raven et al., 2019). Furthermore, only specific branches (i.e., branches 3–9 of CA1 apical/basal dendrites and branches 1–4 of both blades in the DG) show significant decreases in spine numbers following sleep deprivation (Raven et al., 2019). Such branch-specific changes in response to learning and sleep have also been reported for the motor cortex (Yang et al., 2014). With respect to spine type, it should be noted that sleep deprivation leads to an overall reduction of all spine types in CA1 neurons, whereas it specifically affects thin and branched spines in the DG (Havekes et al., 2016; Raven et al., 2019). Together, these observations indicate that sleep deprivation may lead to local changes in structural plasticity that are specific for hippocampal subregion, dendritic branch and spine type.

Despite the general consensus that sleep deprivation has a negative impact on hippocampal connectivity (Acosta-peña et al., 2015; Havekes et al., 2016; Raven et al., 2019; Wong et al., 2019), other studies reported opposing findings (De Vivo et al., 2017; Gisabella et al., 2020; Spano et al., 2019). Of special interest is the work from Gisabella and colleagues, which uses comparable experimental parameters, such as the sleep deprivation method and age of animals. They observed an overall increase in density of CA1 spines of sleep-deprived mice. Interestingly, the observed increase was found specifically for thin spines, and profoundly in distal (150–300 µm) regions from the soma of apical dendrites, whereas mushroom spines were exclusively increased in basal dendrites more close to the soma (0–150 µm) (Gisabella et al., 2020).

Prompted by these, at a first glance, opposing findings by Gisabella and colleagues (Gisabella et al., 2020), we decided to conduct a further in-depth analysis of our previously gathered spine data (Havekes et al., 2016). This additional analysis examined the impact of sleep deprivation on different spine subtypes at a branch-specific level. Because Gisabella et al. (2020) restricted their study to the first two branches stemming from the main apical or basal dendrite, we determined the density of specific spine subtypes along the first and second branches of both apical and basal dendrites. We will also include the fifth branch as a representative for branches 3–9, which were all similarly affected by sleep deprivation (Havekes et al., 2016). Altogether our current study aims to elucidate the origin of the seemingly contradictory

![Diagram of hippocampus](image)

**FIGURE 1** Overview of CA1 pyramidal neurons, branch numbering and spine subtypes. (a) A schematic representation of the hippocampus including the CA1 pyramidal neurons. (b) Illustration of the assignment of branch numbers to dendritic branches (centrifugal branch ordering method). (c) Overview of the different spine subtypes analysed in this study. (d) Example of golgi-stained pyramidal dendrites with different spine subtypes. The example pictures were taken from animals of the non-sleep deprivation group (NSD). DG, dentate gyrus.
findings with respect to the impact of sleep deprivation on structural plasticity in CA1 neurons.

2 | MATERIALS AND METHODS

For this study, we re-analysed previously gathered and published data from Havekes et al. (2016) and briefly summarize the experimental procedures below. Three-month-old male C57BL/6J mice were obtained from Jackson Laboratories. Upon arrival, mice were group housed with four littermates on a 12 h:12 h light:dark cycle (lights on at 7:00 a.m.) with food and water available ad libitum. One week prior to the start of the experiment all mice were single housed. Mice were sleep deprived for 5 h at the start of the lights-on period (7:00 a.m.) using the gentle handling method (Havekes & Aton, 2020; Havekes et al., 2016; Raven et al., 2019).

After sleep deprivation, both experimental groups were immediately sacrificed and the brains were impregnated using the Rapid Golgi stain kit (FD Neurotechnologies Inc., Columbia, MD, USA), and coronal 80-μm-thick hippocampal sections were prescreened to find neurons qualified for spine analysis. We used a stereology-based software (Neurolucida, v10, Microbrightfield, VT) and a Zeiss Axioplan 2 image microscope with an Optronics MicroFire CCD (1600 × 1200) digital camera, motorized in X, Y and Z-focus for high-resolution image acquisition and digital quantification in combination with a 100x objective. Dorsal hippocampus slices, with anterior-posterior coordinates between −1.5 and −2.30, were prescreened for impregnated CA1 pyramidal neurons along the anterior/posterior axis to see if they were qualified for further analysis. Pyramidal CA1 neurons that were insufficiently impregnated or truncated were excluded for further analysis. After prescreening, five neurons per animal were selected for further spine analysis. For the visualization of the spines, we used a 100x Zeiss objective lens with immersion oil, to identify the spine subclass. Prescreening and analysis were carried out by an experimenter blind to treatment; see Havekes et al. (2016) for a more detailed description of data collection. All experiments were conducted according to National Institutes of Health guidelines for animal care and were approved by the Institutional Animal Care and Use Committee of the University of Pennsylvania.

For the more in-depth analysis of the current study, we focused on apical and basal dendrites of the CA1 pyramidal neurons (Figure 1a) and analysed the first, second and fifth branches, in which we investigated whether sleep deprivation affects the different subtypes of spines in a branch-specific fashion (Figure 1b–d). These specific dendritic branch numbers were deliberately chosen in such a way that they include branches investigated previously in our work (Havekes et al., 2016) and more recently by others (Gisabella et al., 2020). Importantly, we used the centrifugal branch ordering method to label branches for spine counting (Figure 1b). This branch counting method starts from the soma, and new branch numbers are assigned after every bifurcation (or trifaction) of the dendrite. Spines were analysed using Neurolucida. The neuronal tracing was conducted by an experimenter blind to treatment at Neurodigitech (San Diego, CA, USA). We analysed data from five neurons per animal and five to six animals per group. The chosen neurons were fully traced and the data from these five to six neurons per animal were averaged, leading to a single data point per animal. Datasets were analysed using non-paired t-tests with adjusted p-values for multiple testing according to the Benjamini-Hochberg method.

3 | RESULTS

3.1 | Sleep deprivation leads to branch-specific reduction in spines along apical dendrites

We previously reported that sleep deprivation causes an overall reduction in spine density selectively on apical branches 3–9 (Figure 2a; Havekes et al., 2016). Our new analysis on branches 1, 2 and 5 now indicates that the effects of sleep deprivation on spine density are branch specific. Particularly, after correcting the p-value for multiple testing, we found no effect of sleep deprivation on the number of spines on branch 1 (Figure 2b), neither did sleep deprivation impact the number of dendritic spines for any subtype on branch 2 (Figure 2c). In contrast, sleep deprivation caused a decrease in the number of mushroom spines (p = 0.004) and branched spines (p = 0.045) on branch 5 (Figure 2d).

3.2 | Sleep deprivation leads to branch-specific reduction in spines along basal dendrites

Overall spine densities on branches 3 to 6 were significantly reduced by sleep deprivation (Figure 2e; Havekes et al., 2016). Spine-specific analyses indicated that sleep deprivation did not alter the density of any type of spine for branches 1 and 2 (Figure 2f,g). In contrast, sleep deprivation attenuated the number of thin (p < 0.001) and mushroom spines (p = 0.011), as well as the number of filopodia (p = 0.042) on branch 5 (Figure 2h). The observed changes in spine densities in both apical and basal dendrites largely returned to control levels after 3 h of recovery sleep (no data shown).

4 | DISCUSSION

Our spine-specific analysis revealed that sleep deprivation alters the density of individual spine types in a branch-specific fashion in CA1 neurons. Even though there was an observed tendency towards an increase in the number of thin spines on apical branch 1, there was no significant increase in thin spines after correcting for multiple testing. Also on apical branch 2, sleep deprivation did not significantly alter the density of any spine type. Furthermore, sleep deprivation decreases the number of mushroom and branched
spines on the apical dendrites of branch 5 and reduces the density of thin, mushroom and filopodia spines on the basal dendrites of branch 5. These findings indicate that, depending on the branch number examined, sleep deprivation reduces the number of specific spine subtypes on apical and basal dendrites. Hence, sleep deprivation has local effects on structural plasticity that might not
reflect the overarching impact of sleep deprivation on CA1 spine density (e.g., Havekes et al., 2016).

The observed tendency towards increased thin spines exclusively along the first branch following sleep deprivation was not significant after correcting for multiple testing. This non-significant tendency is in line with the work of Gisabella et al. (2020) suggesting that sleep deprivation might locally increase the number of thin spines in CA1 neurons. It is important to note that Gisabella et al. (2020) used a different branch counting method (shaft ordering) and limited their analysis to the first two branches, leaving all other branches unanalysed. As a result, it is unclear whether the changes they reported are representative for all branches or, alternatively, are branch specific and not reflecting the overall impact of sleep deprivation. Indeed, our results suggest that local changes might not necessarily reflect the overarching negative effect of sleep deprivation on structural plasticity reported by us and others (e.g., Acosta-peña et al., 2015; Havekes & Aton, 2020; Havekes et al., 2016; Raven et al., 2019; Raven et al., 2018; Wong et al., 2019). As such, these findings together with our previous work underscore the importance of tracing all branch numbers when examining the impact of a manipulation on structural plasticity.

Our data also suggest that the discrepancy in the literature concerning the impact of sleep deprivation on spines might not be related to the method of spine visualization, but is rather dependent on the dendritic branch examined. Whereas Gisabella et al. (2020) used a genetic and viral approach to visualize their spines, the current study uses a different approach, the Golgi-Cox staining method. Although the visualization approaches differ, it seems that the results are not contradictory and are even relatively comparable if similar branches are examined. Moreover, our previous study (Havekes et al., 2016) compared the changes in spines between Golgi-stained and Dil-stained CA1 neurons, revealing similar spine alterations following sleep deprivation. Nevertheless, it should be noted that Golgi only stains a small fraction of the CA1 pyramidal neurons. However, this seems not to influence the impact of sleep deprivation on the spine changes.

Although previous studies found that a large number of molecular mechanisms are affected by sleep deprivation (e.g., Havekes et al., 2016), none of these identified molecular mechanisms can easily explain the observed branch- and spine type-specific alterations on the CA1 pyramidal neurons. Spine plasticity is determined by presynaptic input (or lack of input) and neuronal activity. A previous study by Yang et al. (2014) demonstrated that sleep causes branch-specific spine formation after a learning trial due to enhanced reactivation of neurons involved in the learning process during non-REM sleep (Yang et al., 2014). Although our Golgi staining does not provide any information about the presynaptic input, it is known that the CA1 receives input from both area CA3 and cortical regions, and that some of these brain areas are showing aberrant activity patterns after sleep deprivation (Delorme et al., 2019). Interestingly, proximal dendrites are more prone to receive inputs from closer sources such as the Schaffer collaterals of the CA3, whereas the input of distal dendrites is more related to cortical and thalamic inputs (Spruston, 2008). Moreover, CA1 innervation by interneurons can be highly branch specific, and can even target specific areas on individual dendritic branches (Bloss et al., 2016). A recent study found that the input of CA1 PV+ interneurons to pyramidal cells is altered during post-learning sleep deprivation (Ognjanovski et al., 2018). Altogether, it remains unclear how these specific and heterogeneous input pathways are affected by sleep deprivation. Future studies examining these specific inputs may shed light on the reported local changes in CA1 structural plasticity and its functional consequences.

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CONFLICTS OF INTEREST
None declared.

AUTHOR CONTRIBUTIONS
Y.G.B., R.H., P.M., T.A. and M.J.K. designed the studies. Y.B. and R.H. conducted the analyses. The manuscript was written by Y.G.B. and R.H. with input from P.M., M.J.K. and T.A.

DATA AVAILABILITY STATEMENT
The data that support the findings of this study are available from the corresponding author upon reasonable request.

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