**INTRODUCTION**

*Diphyllobothrium nihonkaiense* is a parasite prevalent in Korea and Japan, but rare in China, Russia, North America, Oceania, and Europe [1-7]. *Diphyllobothrium latum* and *D. nihonkaiense* are morphologically similar and difficult to distinguish. *D. nihonkaiense* has been inaccurately diagnosed as *D. latum*; however, this erroneous identification can be revised using DNA sequencing. Although the treatment is the same for both species [8], the disease severity and treatment results may vary only with limited evidence. Therefore, accumulation of data is necessary.

Many case reports have indicated salmon intake as the source of *D. latum* infection in Korea [9]; however, salmon intake is now more indicative of *D. nihonkaiense* infection. Other sea fish species from the Pacific Ocean [10] may also act as intermediate hosts for *D. nihonkaiense*. DNA analysis of suspected *D. latum* samples collected and examined in Korea indicated the presence of *D. nihonkaiense* [11]. We performed DNA analysis of 4 *Diphyllobothrium* samples from patients referred to the Parasitology Department of Hallym University, Korea between 1995 and 2014 and identified them as *D. nihonkaiense*.

**CASE DESCRIPTION**

Proglottids of *Diphyllobothrium* were obtained from 4 patients referred to the Parasitology Department of Hallym University, Chuncheon, Korea from November 1995 to February 2014 and stored in 100% ethanol. Only 1 patient in 2014 could be reached for medical history collection. The patient resided in Inje-gun, Gangwon-do, and frequently visited Sokcho-si, Gangwon-do to consume raw fish (=sashimi). The patient consumed halibut, rockfish, blowfish, flounder, filefish, mullet, sea squirt, whelk, and squid, but had no salmon intake history. The patient’s wife also consumed sashimi but not mullet because of personal preference. In November 2012, the patient experienced repeated bouts of loose stools. The patient observed a 2-m long parasite while defecating and tried to remove it, but it ruptured. The same issue occurred 1 month later. It is notable that normal stools transformed into loose stools at the point of release of the parasite. The patient received treatment with albendazole (400 mg), after which the same parasite (a part) was excreted. He was treated with albendazole (400 mg) for 3 more days but the medication had no effects. The patient continued medication for 4 more days, with repeated excretion of the parasite. The parasite excretion lasted 1 month when only 1 parasite was observed, and the excretion started to decrease thereafter.
However, in February 2014, 2–3 segments of parasites were observed during defecation. Each parasite segment was 3-4 m long, and ruptured when the patient attempted to manually remove them.

The patient visited an internist in Inje-gun with the expelled parasite. The doctor referred the patient to the Parasitology Department of Hallym University for identification of the parasite, and prescribed praziquantel (10 mg/kg in a single dose). The day after initiation of the therapy, a single dead parasite was observed in the stool. Live parasites expelled previously moved actively and were white to mild yellowish in color, whereas the dead parasite was darker yellowish in color. The proglottid but not the scolex was observed in the parasite. The patient was monitored for 3 months, with no symptoms of infection recurrence, including loose stool.

The medical histories of the other 3 patients were unavailable. Of the 4 *Diphyllobothrium* samples collected from patients, 3 were stored at the Parasitology Department, Hallym University College of Medicine in Chuncheon, Gangwon-do, and 1 at the Parasitology Department at Inje-gun, Gangwon-do. The 4 samples were sent to the Department of Parasitology, Chungbuk National University School of Medicine for DNA analysis.

A single proglottid was cut into small pieces, and total genomic DNA was extracted using a DNeasy tissue kit (Qiagen, Valencia, California, USA) according to manufacturer’s instructions. The partial mitochondrial *cox1* gene was amplified by PCR. PCR was performed in a reaction mixture containing 50 μl with 0.01 μg/μl of genomic DNA, 10× PCR buffer (20 mM Mg²⁺), 10 mM dNTP mixture, 10 pmoles of each primer, and 2.5 U/μl Taq DNA polymerase (High Fidelity PCR system, Roche, Mannheim, Germany) using a GeneAmp PCR System 9700 (Applied Biosystems, Langen, Germany).

The primers used were D1f (5′-TGTGTGGGGGCATCATAT-3′) and the corresponding sequences for the *Diphyllobothrium* species.

![Fig. 1. Aligned partial nucleotide sequences of a region of the mitochondrial cytochrome c oxidase subunit 1 gene of the DNA of adult *Diphyllobothrium* species. The sequences corresponded to 919-1,120 bp (total 203 bps) of the partial mitochondrial *cox1* gene.](image-url)
GT-3’ and D1r (5’-ATGATAAGGACAGGACCTCAGCATA-3’). The second PCR and sequencing primers used were D1f1 (5’-GTTTAGATGTAAAGACGGCTG-3’) and D1r1 (5’-GGTAGAACCGCACACACAA-3’) and yielded a 203-bp product corresponding to the position 919-1,122 bp of the cox1 gene (Fig. 1).

DNA sequencing was performed using a Big-Dye Terminator Sequencing kit (version 3.2, Applied Biosystems, Foster City, California, USA) and the reaction products were separated on electrophoresis using an automated DNA sequencer model 3739KL (Applied Biosystems). DNA sequences were assembled and aligned using Geneious 7.0.2 software (Biometer, Auckland, New Zealand). Phylogenetic analyses were performed using the neighbor-joining (NJ) method and the MEGA software (version 6). NJ analysis was performed using a distance matrix calculated with the Kimura 2 parameter method. The sequences were identified using BLAST searches and compared with sequences of D. nihonkaiense and D. latum deposited in the GenBank database. The cox1 sequences (203 bp) of specimens 1, 2, 3, and 4 showed 99.5% (202/203), 98.5% (200/203), 98.5%, and 99.5% similarities to a reference D. nihonkaiense sequence (NC 008945) and 91.6% (186/203) similarity to a reference D. latum sequence (NC 008945), respectively (Fig. 2).

**DISCUSSION**

*D. nihonkaiense* was recognized as a new species for the first time by Yamane et al. in 1986 [11], Jeon et al. [11], Park et al. [12], and Song et al. [13] have identified *D. nihonkaiense* in Korea. Before 2009, *D. latum* has been primarily identified because of its morphological resemblance with *D. nihonkaiense*, but after 2009, *D. nihonkaiense* was confirmed by DNA analysis in all cases. Our additional 4 cases support the evidence that most or all of the diphyllobothriid tapeworms in Korea are *D. nihonkaiense* but not *D. latum*. In addition, there was no DNA evidence of *D. latum* infection in Korea. In particular, patients with a salmon intake history are unlikely to be infected with *D. latum*. Therefore, during an initial morphological diagnosis of *Diphyllobothrium* infection, *D. nihonkaiense* infection should be suspected first.

A single dose of praziquantel (10 mg/kg) has been recommended for the treatment of *D. nihonkaiense* infection, similar to the dose used to treat *D. latum* infection [8]. However, in 1 case of suspected *D. nihonkaiense* infection, a single dose of praziquantel caused a continuous stomachache, which required an additional dose for recovery [14]. Therefore, 2 doses are thought to be necessary in some patients. Some authors have also used gastrograffin to treat *D. nihonkaiense* infection [15].

The limitation of this study was the small number of cases evaluated. Moreover, the medical history of 3 patients could not be obtained and evaluated. In conclusion, these 4 cases support the evidence of *D. nihonkaiense* infection in Korea. Further DNA sequencing studies are required for accumulation of evidence to prevent misidentification of *D. latum* in Korea.

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**CONFLICT OF INTEREST**

We declare that we have no conflict of interest related to this work.

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