Ethylene-Induced Vinblastine Accumulation Is Related to Activated Expression of Downstream TIA Pathway Genes in Catharanthus roseus

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We selected different concentrations of ethephon, to stress C. roseus. We used qRT-PCR and HPLC followed by PCA to obtain comprehensive profiling of the vinblastine biosynthesis in response to ethephon. Based on our findings, the results showed that the high concentration of ethephon had a positive effect at both transcriptional and metabolite level. Meanwhile, there was a remarkable decrease of hydrogen peroxide content and a promoted peroxidase activity in leaves. The loading plot combination with correlation analysis suggested that CrPrx1 could be regarded as a positive regulator and interacts with ethylene response factor (ERF) to play a key role in vinblastine content and peroxidase (POD) activity. This study provides the foundation for a better understanding of the regulation and accumulation of vinblastine in response to ethephon.

1. Introduction

Catharanthus roseus (L.) G. Don., also called Madagascar periwinkle, is widely used in studies as a model of medical plant for many kinds of terpenoid indole alkaloids (TIAs) [1]. Because of the antineoplastic activity in the treatment of many cancers, some of them have medicinal and scientific research value, such as vinblastine [2, 3]. The early stages of vinblastine biosynthesis in C. roseus involve the formation of tryptamine from tryptophan and its condensation with secologanin to produce the central intermediate strictosidine, the common precursor for the monoterpenoid indole alkaloids, vindoline and catharanthine (Figure 1) [3]. These two monomeric alkaloids will then be enzymatically condensed to form the bisindole alkaloid vinblastine (Figure 1) [4].

Various proteins are involved in plant defense and secondary metabolic responses. Among these proteins, class III plant peroxidases (EC 1.11.1.7) are well known. As is reported, there are various abbreviations used for class III plant peroxidases such as POD, POX, and PRX [5]. Peroxidases (POD), always known as a type of antioxidant, are also involved in the biosynthesis of secondary metabolites for catalyzing production of vinblastine. Although the functions of POD are not well understood in plants, the enzyme has been recognized with medicinal properties [5]. The above statement is the case of the TIAs of Catharanthus roseus [6]. Sottomayor et al. found that peroxidase was purified to homogeneity and a channeling mechanism was proposed for the peroxidase mediated-vacuolar synthesis of α-3’,4’-anhydrovinblastine (AVLB) [4, 6]. AVLB can be converted into vinblastine, which is biosynthesized through coupling of the monomeric precursors vindoline and catharanthine [7].

The POD encoded by CrPrx1 gene is also a multifunctional enzyme that has another function of cleaning up the oxidation of small molecules including H₂O₂ in the cells. H₂O₂ is an electron-accepting substrate for a wide variety of peroxidase-dependent reactions; thus, POD is generally considered to be merely a ROS-detoxifying enzyme [8]. The breakdown of H₂O₂ accumulation by the POD reaction is highly active especially in the presence of ROS-scavenging POD substrates [9]. They have also been involved in secondary metabolism, in root elongation, and in hydrogen
2. Materials and Methods

2.1. Plant Materials and Cultivation Methods. C. roseus seedlings were germinated in perlite with distilled water until having grown out two pairs of leaves and then transferred to standalone plates with Hoagland nutrient solution. The seedlings were kept in a growth chamber at 28°C under a 16 h photoperiod.

2.2. Ethephon Treatment. Seedlings from the three-month-old plants were harvested, and 3 fully expanded leaves of seedlings were randomly selected and subjected to hydroponic treatment. For ethephon treatment, seedlings were cultivated in the Hoagland nutrient solution containing ethephon. We illustrated all the concentrations of ethephon used in our experiment as 45 μM, 60 μM, and 100 μM and the treatment lasted 4 days.

2.3. Ethylene Release Measurement. For measurement of ethylene released from grown plants, plants with or without ethephon treatment grew in a Bunsen beaker and the Bunsen beaker was sealed to air tightness with a plastic membrane. Ethylene that was released by treated plants was determined by gas chromatography (Agilent Technologies, 7890A GC Systems). One milliliter of air from each container was taken to detect the presence of ethylene.

2.4. Alkaloid Analysis. Dry leaf powder (0.3 g) was dissolved in 10 mL absolute methanol (analytical grade) for extraction of vinblastine, vindoline, and catharanthine. Low-frequency ultrasonication (250 W, 40 kHz) was used to extract the alkaloids for 20 min. The methanol extract was centrifuged at 8000 × g for 10 min, concentrated to 1 mL, and analyzed by HPLC (Jasco, VG, England) equipped with a Waters ODS C18 reversed-phase column (250 × 4.6 mm, 5 μm) and a photodiode-array detector at 220 nm. Sample injection volume was 10 μL at a flow rate of 1.5 mL min⁻¹. Samples were applied in triplicate for quantification of vinblastine. The alkaloids were quantified by using regression equation of calibration curve.

2.5. Analysis of Redox State. Endogenous \( \text{H}_2\text{O}_2 \) concentrations were determined according to Patterson et al. [28]. Hydrogen peroxide coupled with titanium sulfate generating superoxide-titanium which was a yellow precipitate. Superoxide-titanium was dissolved by sulfuric acid, and the color of the solution had a linear relation with hydrogen peroxide concentration. The absorbance was read at 415 nm using an ultraviolet-visible spectrophotometer (UV-2550, Shimadzu, Japan). \( \text{H}_2\text{O}_2 \) was determined from a calibration curve.

Accurately measured 0.5 g of \( \text{C. roseus} \) leaves for the determination of peroxidase activity (POD, EC 1.11.1.7) was homogenized to a fine powder under liquid nitrogen. Then, the enzyme was extracted from 5 mL PBS (pH 7.0) with 1.0 mM EDTA, 1.0 mM ascorbic acid, and 10 g/L PVP. The homogenate was centrifuged under \( -4°C \) and 100000 g/min for 30 min. A hundred-microliter sample solution was added to 1.8 mL hydrochloric acid buffer solution, 1.0 mL guaiacol, and 0.1 mL hydrogen peroxide. Peroxidase activities were assayed in UV-2550 (Shimadzu, Japan) at 460 nm for 3 min [29].

![Figure 1: The pathway of terpenoid indole alkaloids biosynthesis in Catharanthus roseus.](image)
SYBR Premix Ex Taq (TaKaRa, Japan). Gene-specific primers using oligo(dT) as the primer. qRT-PCR analysis using cDNA (Toyobo, Japan) according to the manufacturer's instructions, sized from total RNA (260 nm and ethidium bromide (EB) stained test agarose gel electrophoresis used to verify the quality. cDNA was synthesized from 50–100 mg samples (both leaves and roots) by TRIzol reagent and quantified by a NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies) with absorbance at 260 nm.

2.6. RNA Extraction and RT-PCR. Total RNA was extracted from 50–100 mg samples (both leaves and roots) by TRIzol reagent and quantified by a NanoDrop ND-1000 spectrophotometer (NanoDrop Technologies) with absorbance at 260 nm and ethidium bromide (EB) stained test agarose gel electrophoresis used to verify the quality. cDNA was synthesized from total RNA (2 μg) using ReverTra Ace QPCR RT Kit (Toyobo, Japan) according to the manufacturer's instructions, using oligo(dT) as the primer. qRT-PCR analysis using cDNA as template and gene-specific primers was performed using a SYBR Premix Ex Taq (TaKaRa, Japan). Gene-specific primers used are listed in Table 1 (from ExPlant Technologies BV).

PCR was performed at 94°C for 5 min and then at 94°C for 30 s, 60°C for 30 s, and 72°C for 30 s, on 80°C reading plate for 1s for 35 cycles, and remained at 72°C for 2 min. Reactions were repeated three times for each sample to ensure the reproducibility of the results. RPS9 gene was used as an internal control. After PCR reaction, a melting curve was obtained by Opticon version 3. The comparative CT (ΔΔCT) method was used to analyze the relative transcript levels for different experiment groups.

2.7. Statistical Analysis. Principal components analysis (PCA), the multivariate analysis tool, is used to reduce a set of original variables and to extract a small number of latent factors (principal components (PCs)) for analyzing relationships among the observed variables. PCA was performed to evaluate variations of gene expression in response to ethephon. Generally, there are three approaches used: Cattell scree test, Kaiser Criterion, and variance explained criteria. We used the scree plot method to assess the number of PCs to be retained. Loading plot, the two-dimensional plane formed by the two first principal components, is the most informative in PCA. The loading plots display the relationships among the detected compounds. The loading of PC1 and PC2 against each other shows the summary of the relationship among variables [30].

All experiments were conducted with three replicates. Statistical analysis was performed using PCA and one-way analysis of variance (ANOVA) followed by SPSS17.0. Differences between treatments were separated by the least significant difference (Duncan) test at a 0.05 probability level. The values are mean ± SD for three samples in each group.

3. Results

3.1. Effect of Etephon on Endogenous Ethylene Accumulation and Vinblasticine Content. We firstly verified the effect of exogenous application of ethephon, which is absorbed into plants and converted into ethylene, on endogenous ethylene release in C. roseus seedling. The results showed that endogenous ethylene release was promoted with the treatment time and concentration of ethephon (P < 0.05) (Figure 2(a)). This indicates that the increase of vinblastine accumulation may result from the high concentration of ethephon. Then, the vinblastine contents were measured after C. roseus plants were treated by different concentrations of ethephon for 6 h (as is shown in Figure 2(a), we found that this is the best treatment time for C. roseus to produce alkaloids). Increasing concentrations of ethephon and up to 100 μM ethephon concentration resulted in enhanced vinblastine content. Treatment with 100 μM ethephon induced vinblastine content about 2 times that in the control group (Figure 2(b)) (P < 0.05).

3.2. Expression of Genes and Accumulation of Alkaloids in Response to Etephon. The contents of vindoline and catharanthine in seedlings of C. roseus were also quantified. As shown in Figure 3(a), treatment with 60 μM or 100 μM ethephon increased the production of vindoline and catharanthine of C. roseus seedlings about 2-fold, compared with the control. At the transcriptional level, the expression of D4H and T16H genes followed a similar trend to that of vindoline (Figure 3(a)); expression of D4H and T16H genes was found to be 3 times more in response to high concentration of ethephon (60 μM and 100 μM), while the expression of DAT gene was 3 times more in response to low concentration of ethephon (45 μM) (P < 0.05).

As indicated by the loading plot (Figure 3(c)), principal component analysis (PCA) showed two principal components, which can explain 98.181% of the variance of the four downstream TIA pathway genes and serve as high concentrations of ethephon-dependent (D4H, T16H, and CrPrx1) and low concentrations of ethephon-dependent (DAT) variables.

3.3. Effect of Etephon on Redox State

3.3.1. Effect of Etephon on H2O2 Accumulation in C. roseus Leaf. The key function of plant PRXs (POD) is to oxidize phenolic substrates at the expense of ROS, mainly H2O2. To clearly identify the defense level of POD, the accumulation of H2O2 in C. roseus was also measured (Figure 4). Compared with the control plants, H2O2 product was inhibited by application of ethephon (P < 0.05). We observed that ethephon authentically reduced H2O2 accumulation in C. roseus leaf, and high concentration of ethephon significantly reduced H2O2 accumulation. With the treatment with 100 μM ethephon, H2O2 product was 2-fold less in

Table 1: Primers used in qRT-PCR for validation of differentially expressed genes.

|           | Sequence                        | Length |
|-----------|---------------------------------|--------|
| d4h-F     | GACTTGAACCTTCTATGCTGCTACAC      | 25     |
| d4h-R     | TCTCATCAAAAGCTCTTCAATTCG       | 23     |
| dat-F     | AATCCCTCAAGGCGTATAAAC          | 21     |
| dat-R     | ACGGATACGCAGCTTTGTGAT          | 21     |
| CrActin-F | CTTAGTCCAGGATGTCGACATGA        | 26     |
| CrActin-R | GCTGCTTGGAGCCAAGGC            | 18     |
| CrT16H-F  | GCCTCATCACCAGTCTTCAT           | 20     |
| CrT16H-R  | CCGGACATATCCTTCTTCCCA         | 20     |
| Crpx-F    | GCAACATCTCCCCAGCCACA          | 20     |
| Crpx-R    | GTTCTCCCAACACATATGAGGACC      | 23     |
| ERF-F     | CACCTCAATGGCTGCTTTT           | 20     |
| ERF-R     | TCCTGGCTGCTTCTTCTT           | 20     |
Table 2: The relative transcript level of CrPrx1 and the correlation with vinblastine and POD activity after 12 or 24 h of treatment with ethephon to C. roseus. The results shown are the means of three replicates; bars represent SE.

|       | CK      | +45 µM  | +60 µM  | +100 µM | Vinblastine | POD activity |
|-------|---------|---------|---------|---------|-------------|--------------|
| 12 h  | 1       | 2.181 ± 0.021<sup>a</sup> | 2.266 ± 0.042<sup>a</sup> | 3.249 ± 0.148<sup>b</sup> | 0.918**      | 0.940**      |
| 24 h  | 1.765 ± 0.005<sup>a</sup> | 2.354 ± 0.019<sup>b</sup> | 3.434 ± 0.040<sup>c</sup> | 6.714 ± 0.146<sup>d</sup> | 0.816**      | 0.951**      |

Different letters indicate significant differences among treatments ($P < 0.05$). **Correlation is significant at the 0.01 level.

3.3.2. Effect of Ethephon on Peroxidase Activity. In C. roseus, POD was shown to be responsible for the dimerization reaction between catharanthine and vindoline to produce α-3',4'-anhydrovinblastine, the precursor of the natural antitumor products, vinblastine and vincristine. To determine the effect of ethephon on peroxidase (POD) of C. roseus, different ethephon concentrations were added to the media (Figure 5). It is noticed that POD activity had a lasting upward trend and showed peak of maximal activity in the seedlings treated by 100 µM ethephon ($P < 0.05$).

3.4. Effect of Ethephon on CrPrx1 Transcript. The transcript level of CrPrx1 in C. roseus including control and treated by ethephon at 0 h, 12 h, and 24 h was quantified (Table 2). In comparison to vinblastine content, the exposure of seedlings to ethephon led to a positive effect on the transcription of CrPrx1 ($P < 0.05$). Responding to these concentrations of ethephon, additive transcription of CrPrx1 was observed obviously at both 12 h and 24 h and the increment was prominent at 100 µM ethephon. Moreover, the CrPrx1 transcript level at 24 h was about 2 times higher than at 12 h; particularly, maximal steady-state amounts of CrPrx1 transcripts were detected in seedlings treated by 100 µM ethephon.

3.5. The Loading Plot Combination with Correlation Analysis. We also analyzed the correlation between alkaloids and the transcript levels of genes (the downstream TIA pathway genes and ethylene responsive factor (ERF) gene) (Figure 6). As indicated by loading plot (Figure 6), there were two principal components, which can explain 84.164% of the variance of the ten factors and serve as the first principle component (DAH, vinblastine, vindoline, catharanthine, and H$_2$O$_2$) and the second principle component (DAT, T16H, CrPrx1, ERF, and POD) variables. H$_2$O$_2$ presented a significant level of negative correlation in vinblastine accumulation during ethephon treatments, while ERF presented a significant level of activity in TIA accumulation during ethephon treatments. In addition, with the loading plot combination with correlation analysis (Table 2), the results revealed that CrPrx1 transcript was significantly correlated with vinblastine content and POD activity ($P < 0.01$).

4. Discussion

From the present study, it can be concluded that vinblastine biosynthesis influenced by ethephon has a specific relationship with the downstream TIA pathway genes. The result revealed that exogenous ethephon induced a slight increase in endogenous ethylene synthesis (Figure 2(a)). A significant amount of evidence revealed that the regulator had a close
relationship with plant growth or secondary metabolism [31]. Tadiello et al. found that all plant tissues are able to produce ethylene, which has been established to modulate a number of important plant physiological activities [32]. The use of elicitors to promote secondary metabolism in plant cell and tissue cultures has become a common practice [33]. Therefore, it is not surprising that ethephon which is converted into ethylene upon metabolism by plants influenced vinblastine accumulation in *C. roseus* (Figure 2(b)). We noticed that vinblastine content was gradually increasing and exceeded the control group at 45–100 μM ethephon. The level of *CrPrx1* transcript is similar as a trend of vinblastine content of *C. roseus* treated by ethephon (Table 2). Furthermore, through analyzing the correlation between vinblastine and transcription of the downstream TIA pathway genes (Figure 6), we found that the results have significant correlation with each other. As the common precursor of all dimeric alkaloids, α,β,α'-anhydrovinblastine (AVLB) was converted into vinblastine through coupling of the monomeric precursors vindoline and catharanthine [7]. Peroxidase was proposed to be a channeling mechanism for the peroxidasemediated-vacuolar synthesis of AVLB [4, 6]. Our data also indirectly indicated that, on the molecular level, vinblastine biosynthesis had a close correlation with peroxidase.

POD is the key enzyme catalyzing vinblastine production. The data revealed that POD activity depending on *CrPrx1* transcript was assuredly promoted by ethephon. From the result of the three alkaloids and POD activity (Figures 2(b), 3, and 5), it was shown that vinblastine content was more sensitive to the activity of peroxidase; therefore, POD activity played a principal role in vinblastine synthesis. Under 60 μM ethephon treatment, vindoline and catharanthine
Figure 4: The effects of different concentrations of ethephon on H$_2$O$_2$ accumulation in Catharanthus roseus. The results shown are the means of three replicates; bars represent SE. Different letters indicate significant differences among treatments ($P < 0.05$).

Figure 5: The effects of different concentrations of ethephon on peroxidase activity in leaves of Catharanthus roseus. The results shown are the means of three replicates; bars represent SE. Different letters indicate significant differences among treatments ($P < 0.05$).

Contents were a little higher than that of 100 $\mu$M ethephon, but vinblastine content was decreased possibly leading to lower POD activity. We analyzed the correlation between POD activity and transcription of CrPrx1 (Table 2), and the results also had significant correlation. POD and CrPrx1 are small part of a complex network of factors subjected to different regulation programs in the TIA pathway and this pathway is not only a single enzyme matching the regulation [3]. Researchers enabled the observation of a clear correlation between the AVLB and CrPrx1 transcript levels, but details of the process where CrPrx1 regulated vinblastine biosynthesis are not clear. In addition, POD, dependent on CrPrx1 transcript, has defense and catalyzing vinblastine biosynthesis mechanism; thus, it plays a more complex role in the process. Our results demonstrate that ethylene regulates vinblastine accumulation via controlling CrPrx1 transcript and protein which catalyze precursor of vinblastine synthesis at the molecular level. The spatial, temporal, and inducible formation of secondary metabolites and the transcripts of corresponding biosynthetic genes are under tight regulation at different levels, in which transcriptional regulation via transcription factors has been investigated intensively [31]. Guo et al. found that activation of EIN3 (ethylene insensitive 3) increased peroxidase (POD) activity through the direct transcriptional regulation of PODs expression. Accordingly, ethylene pretreatment or EIN3 activation was able to preclude excess ROS accumulation and increased tolerance to salt stress [34]. In fact, not all regulation is at the level of the EIN3-like transcription factors, and downstream ERF and EDF transcription factors greatly increase the potential points for interaction and cross talk of ethylene signaling with other pathways [23]. Therefore, we tested the expression of ethylene responsive factor (ERF) and investigated the possible regulatory way, by which ethylene controls gene transcription. As shown in Figure 6, the ERF mainly participates in the accumulation of TIAs in response to ethephon treatment. This suggests that ERF may be the central hub of the POD and CrPrx1 and, thus, may be actively involved in accumulation of TIAs in response to ethephon.

In vitro, a range of secondary metabolites including phenols, amines, indoles, alkaloids, and sulphonates may act as reductant substrates of class III peroxidases (Prxs), at the expense of H$_2$O$_2$, which has also emerged as a pivotal molecule in the responses of a range of biotic and abiotic stresses [35]. In this work, we showed increased accumulation of vinblastine, vindoline, and catharanthine with treatment with ethephon. Meanwhile, transcription of the peroxidase (POD) encoding gene CrPrx1 is also increased by...
ethephon treatment, accompanied by increased POD activity and decreased H$_2$O$_2$ content. POD is generally considered to be merely a peroxide-detoxifying enzyme, especially and decreased H$_2$O$_2$ accumulation is the evidence to indirectly prove POD and vinblastine synthesis conditions. If this inference is established, H$_2$O$_2$ accumulation must conform to the trend of POD activity level. According to the result of H$_2$O$_2$ accumulation, H$_2$O$_2$ content was reduced by ethephon; therefore, the result consisted with the above reasoning. Some research indicated that H$_2$O$_2$ content was correlated with vinblastine accumulation [36, 37].

The role of ethylene in plant defense is controversial as it contributes to resistance in some interactions [38, 39] but promotes disease in others [40–42]. Therefore, we suppose that vinblastine may be suppressed by trace of ethephon, while being promoted by higher concentration of ethephon. It should be noted that this study does not provide enough data to understand the behavior of ethylene in these systems [5]. Ethylene interacts with other plant developmental pathways and the mechanisms must be studied in order to clarify these issues in more detail [23].

Abbreviations

| Acronym | Description |
|---------|-------------|
| AVLB:  | α-3′,4′-Anhydrovinblastine |
| CrPrx1: | Catharanthus roseus peroxidase |
| DAT:   | Acetyl-CoA: 4-O-deacetylvinindole-4-O-acetyl-transferase |
| DEPC:  | Diethylypyrocarbonate |
| D4H:   | Desacetoxyvinindole-4-hydroxylase |
| EDTA:  | Ethylenediaminetetraacetic acid |
| ERF:   | Ethylene response factor |
| MMLV:  | Moloney murine leukemia virus |
| POD:   | Peroxidase |
| RT-PCR:| Reverse transcription-polymerase chain reaction |
| TIA:   | Terpenoid indole alkaloids |
| T6H:   | Tabersonine-16-hydroxylase |
| PCA:   | Principal component analysis |
| PVP:   | Polyvinyl pyrrolidone |

Competing Interests

The authors declare that there are no competing interests regarding the publication of this paper.

Authors’ Contributions

Xiao-Rui Guo and Zhong-Hua Tang designed and supported the work. Xi Wang and Bo-Wen Chang conducted the experiments and prepared the paper. Ya-Jie Pan conducted the supplementary experiments and revised the paper. Yan-Bo Hu edited the paper and Xiao-Rui Guo and Zhong-Hua Tang reviewed and revised it. Xi Wang and Ya-Jie Pan contributed equally to this work.

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