Viruria of Human BK Virus and John Cunningham Virus among Renal Transplant Recipients and Healthy Control in Southeast of Caspian Sea

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\textbf{Abstract}

\textbf{Background:} Members of the Polyomaviridae family, BK virus (BKV), and John Cunningham virus (JCV) are linked to polyomavirus-associated nephropathy-associated transplant rejection in immunodeficient patients. \textbf{Objective:} The aim of the study was to evaluate the prevalence of BKV and JCV in immunocompetent individuals in the north of Iran. \textbf{Methods:} Ninety-one urine samples were obtained from renal transplant recipients with a mean age of 39.78 ± 11.19 years. A healthy control group of 65 volunteers with an average age of 40.32 ± 10.7 years also contributed. After DNA extraction, positive cases were detected through PCR. Genotyping was done by alignment and phylogenetic tree construction of the VP1 region against all known JCV and BKV genotypes. \textbf{Results:} The prevalence of BKV and JCV was 15.38 and 19.78%, respectively. JCV was detected in 7.69% of the control group. The prevalence of the BKV between the case and control groups was significant ($p < 0.0001$). There was no significant association between BKV and JCV and duration of dialysis ($p > 0.05$). Overall, 62.16% of JCV cases were genotype I. Besides, genotype II was dominant within patients with BKV-positive patients. \textbf{Discussion:} The results obtained here show a relatively lower prevalence of BKV and JCV in immunocompetent renal transplant receivers and healthy control than those reported from other areas in Iran. JCV genotyping was evaluated for the first time in Iran. Genotype I for JCV and genotype II for BKV were dominant genotypes in the north of Iran.

\textbf{Introduction}

Kidney transplantation is the final treatment option for patients with ESRD. In this regard, major concerns are related to graft rejection due to immunity and a variety of viral infections. John Cunningham virus (JCV) and BK virus (BKV) are 2 important viruses within the family Polyomaviridae, which are involved in renal transplant rejection [1].
Today, it is known that ∼80% of healthy adults are seropositive for Polyomaviridae, and they may become reactive in immunodeficient conditions [2, 3]. The use of immunosuppressive drugs in a transplant recipient leads to an increased incidence of BKV and JCV diseases. Polymavirus reactivation in kidney and bone marrow transplant recipients is due to the use of immunosuppressive drugs and because of immune deficiency. Therefore, investigation of the incidence of these viruses in the transplant recipients is necessary to prevent reactivation of the virus and to prevent transplantation termination. The genomic prevalence of BKV and JCV in urine specimens of kidney recipients is 46.7 and 23.3%, respectively, and 13.3 and 24.5%, respectively, in healthy subjects [4].

BKV is a human polyomavirus first isolated from the urine of an immunocompromised renal transplant patient by Gardner et al. [5] in 1971. BKV may reactivate after renal transplantation and lead to nephropathy-associated rejection of the renal graft. In the previous work on biopsy samples from renal transplant recipients, BKV was found in 13.1% of clinical samples in Iran [6]. Samarbast-Zadeh et al. [7] have shown a 3-fold increase of BKV reactivation after 4 months of renal transplantation. BKV is divided into several serotypes. In a small population study, Motazakker et al. [8] have shown that the BKV serotype I is dominant in Iranian Turkish renal transplant recipients. In another study in Iran with more participants, it was found that serotype I of BKV with 94.11% frequency was more prevalent than serotype IV with 5.89% [9].

JCV is an opportunistic pathogen in the human population that generally infects children and can persist in the renal tissue and bone marrow [10]. The virus can lead to progressive multifocal leukoencephalopathy in patients infected with human immunodeficiency virus [11]. The prevalence of the JCV was estimated to be 38.3–40% in the healthy population [12, 13], 16.8% in patients with rheumatoid arthritis [14], and 25.9% in the central nervous system [15]. Further studies in Iran show the prevalence of JCV in renal transplant receivers is 43% [16].

Because of the clinical importance of BKV and JCV in immunocompromised patients, investigation of these viruses in renal transplant receivers is warranted. Our knowledge about the prevalence of JCV and BKV in the patients is limited in Iran, and for the first time, we report the prevalence of the viruses in the north of Iran. As a result, it was found that BKV and JCV were significantly prevalent in renal transplant recipients than healthy controls.

### Materials and Methods

#### Sample Collection

In this study, 91 urine samples were collected from renal transplant recipients admitted to the 5th Azar Hospital in Gorgan, Iran. Further 65 urine samples were also obtained from healthy controls with no history of renal transplant or usage of immunosuppressants. No pregnant women were included in the study. The samples were stored at −20°C until DNA extraction.

#### Experimental Ethics

An informed consent was obtained from the participants. This study was also approved by the Ethical Committee, Golestan University of Medical Sciences, Gorgan, Iran, with the approval code of IR.GOUMS.REC.1395.12.

#### DNA Extraction

The extraction of viral DNA from urine specimens was conducted by using a KiaSpin viral nucleic acid kit (KIAGEN, Tehran, Iran) according to the manufacturer’s protocol. The quality of extracted DNA was evaluated by the a DeNovix spectrophotometer (DeNovix, Inc., Wilmington, DE, USA). The 260/280 Optical density (OD) of 1.8–2 was considered as an ideal purity.

#### Detection of BKV and JCV

The primer that is complementary to the AgT coding sequence of the BK and JC polyomaviruses is shown in Table 1. PCR was performed in a final volume of 25 μL containing 5 μL sample, 2 μL 10× PCR buffer, 2 μL MgCl2, 1.5 μL dNTP, 10 pmol of each forward and reverse primers, and 5 units/reaction Taq DNA polymerase.

The thermocycler (Peqlab Biotechnologie GmbH, Erlangen, Germany) protocol includes 94°C for 4 min and 34 cycles consisting of 94°C for 30 s, 55°C for 40 s, and 72°C for 45 s. A final 72°C step for 5 min was also included. The PCR products were observed in 1.5% agarose gel containing SYBR Safe stain (Invitrogen, Carlsbad, CA, USA). Subsequently, all positive samples were evaluated by 2 other BK– and JC-specific primers encompassing the VPC1 gene (Table 1).

For BK, the protocol consists of 5 μL sample, 2 μL 10× PCR buffer, 5 μL MgCl2, 0.5 μL dNTP, 10 pmol of each primer, 5 units/reaction Taq polymerase in a final volume of 25 μL. The thermal condition was as follows: 95°C for 5 min, amplification was performed in 35 cycles composed of 95°C for 45 s, 59°C for 45 s, and 72°C for 60 s. A final 72°C extension step was conducted for 10 min.

The same concentrations were used for amplifying the JCV. However, thermocycler conditions include 94°C for 5 min, followed by 35 cycles consisting of 94°C for 60 s, 55°C for 60 s, and 72°C for 60 s, as well as a final 72°C step for 5 min. PCR-positive products were sent for direct sequencing (Macrogen, Seou, South Korea).

#### Genotyping

PCR products of BKV and JCV at the VP1 region were sequenced (Macrogen, Seou, South Korea). Results of sequencing were double-checked by eye and Biodeit software package [17] for quality and validity of the data. For multiple sequence alignment, MUSCLE was used in the setting of the CLC sequence viewer software tool [18]. The alignment was performed on the sequenced VP region of BKV and JCV versus all known genotypes of these vi-
A phylogenetic tree was constructed for the representation of distance-relationship of the data to other genotypes. The phylogenetic trees were constructed with the neighbor-joining algorithm within the CLC software package. Bootstrap 1000 was utilized for topological validation of the trees.

**Statistical Analysis**
Statistical analysis is performed by SPSS software version 16. The difference in the relative distribution between the groups is carried out by χ² tests, with a confidence interval of 95%. A p value <0.05 was considered as significant.

**Results**
In this study, 91 urine samples were obtained from immunosuppressed renal transplant receivers (70 male and 21 female cases) with a mean age of 39.78 ± 11.19 years. Additional sex- and age-matched control groups consisting of 65 participants were also included. The mean age of the control group (46 male and 19 female cases) was 40.32 ± 10.7 years. Table 2 shows the demographic data. The average duration of hemodialysis in the case group was 58.96 ± 50.77 (1–180) months.

As a result, the frequency of the BKV in this case-control cohort study was 15.38% (14/91), including 3 female and 11 male cases. The frequency of JCV was 19.78% (6 in female and 12 in male cases). In the control group, the genome of the BKV was not observed, while the JCV was detected in 7.69% (5/65), and all of them were males. The prevalence of BKV between the case and control groups was significant (p < 0.0001). JCV was also significantly higher in the case group (p = 0.028). There was no significant association between BKV and JCV and the age or duration of dialysis (p > 0.05).
The phylogenetic study for determination of the dominant genotype in the north of Iran has revealed that all sequenced BKV VP1 belonged to genotype II (Fig. 1). Furthermore, of 23 sequenced VP1 for JCV-positive PCR products, all were genotype I (Fig. 2).

**Discussion**

Polyomaviruses are involved in different human abnormalities, including Merkel cell carcinoma [19] and renal cell carcinoma [20]. Investigating BKV reactivation
among renal transplant recipients is of great importance as it can cause nephropathy followed by the rejection of allograft. In the present study, for the first time, the prevalence of BKV and JCV is reported in 2 case and control groups in the north of Iran. Previous works have been reported a ~29% BKV prevalence in renal transplant receivers [6–9]. On the other hand, reports on JCV in renal transplant receivers in Iran are limited to few studies. In that case, the prevalence of JCV in renal transplant receivers is almost ~29.5% [12–16]. In the immunosuppressed

Fig. 2. Phylogenetic tree demonstrating VP1 sequence of JCV. As illustrated, JCVs (those with digits left to the JCV) have a close distance to those JCVs belonging to genotype I. JCV, John Cunningham virus.
transplant receivers, the rate of JCV is almost 38% [16, 19–21].

As a result, we found 15.38% (14/91) of total patients were BKV positive, of which 3 cases were female and 11 cases were male. Meanwhile, the genome of JCV has been observed in 19.78% (18/91) of cases, including 6 female and 12 male cases. In the healthy control group, the prevalence of BKV was zero, while JCV was detected in 5 male cases. The results show a lower prevalence of BKV and JCV in renal transplant receivers in the north of Iran. Further reports of BKV and JCV prevalence in Iran are summarized in Table 3.

In previous studies, the rate of BKV prevalence is reported from 0% in the study of [20] to 62% in the study of [22]. In later research, the genome of BKV was detected in 136/220 urine samples by using the PCR technique. However, the result of the present study is consistent with that of Pakfetrat et al. [21], which was conducted in the center of Iran. Further epidemiological data are reported in [24]. This difference of detection may be attributed to the time of sampling, age of examination [24], and design of the study, for example, Pezeshgi et al. [23] have screened both BKV and JCV during 1 year after renal transplantation, and no sign of new cases was observed. The prevalence of JCV in the present study (19.78%) was lower than that reported from previous studies in Iran (31%). JCV is known to cause progressive multifocal leukoencephalopathy, which has been reported in the renal transplant population [25]. Screening of JCV is not a routine test for renal transplantation.

It was also evaluated if there is any association between BKV or JCV and age or duration of hemodialysis. Accordingly, no significant association was observed between BKV or JCV with age or duration of dialysis (p > 0.05). This finding is consistent with that in Taheri et al. [4].

According to molecular and serological studies, BKVs are divided into 4 genotypes: I, II, III, and IV. Genotype I is prevalent worldwide, while genotype IV is more prevalent in Asia, and II/III are less frequent [26]. Furthermore, 18 different genotypes of JCV are detected all around the world. Genotype Af2 is dominant in Africa and the south of Asia. Besides, genotypes SC, CY, MY, B2, B1-d, B1-b, and B1-a are frequent in Eastern Asia [27–29]. Our data clearly show that genotype II of BKV and genotype I of JCV are prevalent among renal transplant recipients in the north of Iran. There are not many studies in Iran reflecting dominant genotypes of poly-

### Table 3. Summary reports of prevalence of BKV and JCV from different parts of Iran

| Author                  | Study                  | City        | Methodology                     | No. of cases | No. of controls | Prevalence of JCV in case group (%) | Prevalence of JCV in control group (%) | Prevalence of BKV in case group (%) | Prevalence of BKV in control group (%) |
|-------------------------|------------------------|-------------|---------------------------------|--------------|----------------|------------------------------------|----------------------------------------|---------------------------------------|---------------------------------------|
| Atyabi et al. [12]      | Case-control           | Isfahan     | Traditional PCR/sequencing      | 143          | 100            | 51 (35.66)                         | 7 (7)                                  |                                      |                                      |
| Jozpanahi et al. [20]   | Cross-sectional        | Tehran      | RT-PCR                          | 50           | –              | –                                  | –                                      | 0 (0)                                 | –                                      |
| Sadeghi et al. [15]     | Cross-sectional        | Tehran      | qRT-PCR                          | 58           | 15             | 15 (25.9)                          | –                                      | –                                    | –                                     |
| Kaydani et al. [9]      | Cross-sectional        | Ahvaz       | PCR-RFLP                        | 122          | –              | –                                  | 51 (41.8)                             | –                                    |                                      |
| Pakfetrat et al. [21]   | Cross-sectional        | Shiraz      | qRT-PCR                          | 108          | –              | –                                  | 17 (15.7)                             | –                                    |                                      |
| Tajedin et al. [22]     | Cross-sectional        | Isfahan     | PCR-RFLP                        | 220          | –              | –                                  | 102 (75)                              | –                                    |                                      |
| Bozorgi et al. [13]     | Cross-sectional        | Tehran      | Traditional PCR                 | –            | 133            | 51 (38.3)                         | –                                      | –                                    | –                                      |
| Motazakker et al. [8]   | Cross-sectional        | Urmia       | PCR-RFLP                        | 120          | –              | –                                  | 12 (10)                               | –                                    |                                      |
| Pezeshgi et al. [23]    | Cross-sectional        | Tehran      | Traditional PCR                 | 31           | –              | 7 (22.5)                          | 9 (29)                                | –                                    | –                                      |
| Samarbasf Zadeh [7]     | Cross-sectional        | Ahvaz       | Double PCR and semi-nested-PCR   | 78           | –              | –                                  | 5 (6.8)                               | –                                    | –                                      |
| Ghaifari et al. [6]     | Cross-sectional        | Urmia       | Immuno-histochemistry            | 160          | –              | –                                  | 21 (13)                               | –                                    | –                                      |

JCV, John Cunningham virus; BKV, BK virus.
omaviruses. However, in one study performed by Tajeddini et al. [22] on 220 patients, genotype I (46.36%) was more frequent than genotype II (3.2%), genotype III (2.27%), and genotype IV (10%). Kaydani et al. [9] have found that genotype I (39.34%) and genotype III (2.46%) were 2 dominant genotypes among 122 patients [22]. In another study by Motazakker et al. [8] on 120 urine samples from renal transplant recipients, genotype I (10%) was prevalent. There are few studies on the prevalence of JCV among healthy and transplant recipients. Atabi et al. [16] have found JCV among patients (43%) and healthy people (7%). Further evidence of JCV infection among renal transplant recipients is from the studies of Pezeshki and Ghods [21] and Bozorgi et al. [13] on urine samples from 31 patients and 133 healthy controls, respectively. The frequency of the JCV in these studies has been 22.58 and 38.35%, respectively. Unfortunately, no evidence of JCV genotype is known in Iran, and for the first time, we report that the JCV genotype I is dominant in the north of Iran. Further investigation is warranted to develop our knowledge about the distribution of JCV and BKV in Iran.

One of the important disadvantages of the present study is that the load of BKV and JCV is not evaluated. Hence, it cannot be concluded that the positive cases are active viral infections or are resulted from quasi-species. Therefore, it is warranted that more sensitive and quantitative methods like RT-PCR be used for investigation of BKV and JCV in renal transplant recipients in the north of Iran.

**Conclusion**

The results obtained here show a relatively lower prevalence of BKV and JCV in immunocompromised renal transplant receivers and healthy controls than that reported from Iran. Additionally, it was found that genotype II of BKV and genotype I of JCV are prevalent among renal transplant recipients in the north of Iran. It was for the first time we report the JCV genotype in this country.

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**Statement of Ethics**

This study is approved by the Ethical Committee, Golestan University of Medical Sciences, Gorgan, Iran, with the approval code of IR.GOUMS.REC.1395.12.

**Conflict of Interest Statement**

Authors have nothing to declare.

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**Author Contributions**

Substantial contributions to the conception or design of the work; or the acquisition, analysis, or interpretation of data for the work: F.S., M.H., A.T., A.M., and H.R.N. Drafting the work or revising it critically for important intellectual content: F.S., A.M., and M.H. Final approval of the version to be published: A.T. and A.M. Agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved: all the authors.

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