THE FELINE CARDIOMYOPATHIES

2. Hypertrophic cardiomyopathy

Practical relevance: Hypertrophic cardiomyopathy (HCM) is the most common form of feline cardiomyopathy observed clinically and may affect up to approximately 15% of the domestic cat population, primarily as a subclinical disease. Fortunately, severe HCM, leading to heart failure or arterial thromboembolism (ATE), only occurs in a small proportion of these cats.

Patient group: Domestic cats of any age from 3 months upward, of either sex and of any breed, can be affected. A higher prevalence in male and domestic shorthair cats has been reported.

Diagnostics: Subclinical feline HCM may or may not produce a heart murmur or gallop sound. Substantial left atrial enlargement can often be identified radiographically in cats with severe HCM. Biomarkers should not be relied on solely to diagnose the disease. While severe feline HCM can usually be diagnosed via echocardiography alone, feline HCM with mild to moderate left ventricular (LV) wall thickening is a diagnosis of exclusion, which means there is no definitive test for HCM in these cats and so other disorders that can cause mild to moderate LV wall thickening (eg, hyperthyroidism, systemic hypertension, acromegaly, dehydration) need to be ruled out.

Key findings: While a genetic cause of HCM has been identified in two breeds and is suspected in another, for most cats the cause is unknown. Systolic anterior motion of the mitral valve (SAM) is the most common cause of dynamic left ventricular outflow tract obstruction (DLVOTO) and, in turn, the most common cause of a heart murmur with feline HCM. While severe DLVOTO is probably clinically significant and so should be treated, lesser degrees probably are not. Furthermore, since SAM can likely be induced in most cats with HCM, the distinction between HCM without obstruction and HCM with obstruction (HOCM) is of limited importance in cats. Diastolic dysfunction, and its consequences of abnormally increased atrial pressure leading to signs of heart failure, and sluggish atrial blood flow leading to ATE, is the primary abnormality that causes clinical signs and death in affected cats. Treatment (eg, loop diuretics) is aimed at controlling heart failure. Preventive treatment (eg, antithrombotic drugs) is aimed at reducing the risk of complications (eg, ATE).

Conclusions: Most cats with HCM show no overt clinical signs and live a normal or near-normal life despite this disease. However, a substantial minority of cats develop overt clinical signs referable to heart failure or ATE that require treatment. For most cats with clinical signs caused by HCM, the long-term prognosis is poor to grave despite therapy.

Areas of uncertainty: Genetic mutations (variants) that cause HCM have been identified in a few breeds, but, despite valiant efforts, the cause of HCM in the vast majority of cats remains unknown. No treatment currently exists that reverses or even slows the cardiomyopathic process in HCM, again despite valiant efforts. The search goes on.

Keywords: Cardiomyopathies; myocardial diseases; hypertrophic cardiomyopathy; gene mutation; systolic anterior motion; mitral valve; echocardiography

Hypertrophic cardiomyopathy (HCM) is defined as concentric left ventricular hypertrophy (LVH; thickened left ventricular [LV] wall) in the absence of another cardiac or systemic disease capable of producing the magnitude of hypertrophy evident (Figure 1). The LV chamber in diastole is normal in some cats and is small in some others. Affected cats can present with signs of heart failure or arterial thromboembolism (ATE). Some die suddenly. However, like humans with HCM, many cats never exhibit any clinical signs of cardiac disease (ie, have subclinical disease) and live a normal lifespan.
In cats, the cause of HCM is unknown except in Maine Coons and Ragdolls, where a causative mutation has been identified (see below). Consequently, HCM is a diagnosis of exclusion in most cats. Other common causes of LVH that may need to be excluded include aortic stenosis, dehydration, systemic hypertension, hyperthyroidism, and acromegaly. However, there are caveats. Systemic hypertension and hyperthyroidism do not cause severe LVH, so if a cat has severe LVH (arbitrarily defined as diastolic LV wall thickness ≥7 mm) and systemic hypertension or hyperthyroidism, it can generally be assumed that these systemic disorders are not the sole cause of the LVH. Instead, it is likely that hyperthyroidism (and probably also systemic hypertension) exacerbates the LVH seen with HCM, since it is known that successful treatment of hyperthyroidism results in a decrease in LVH.9,8 Therefore, if an older cat has severe LVH, hyperthyroidism, and systemic hypertension still need to be ruled out as complicating factors but are probably not the only disease process affecting the heart. Hyperthyroidism, acromegaly, and systemic hypertension are rare in younger cats. Aortic stenosis is rare in any cat.10

Obviously, if a cat has hyperthyroidism, it should be treated appropriately and if it has systemic hypertension it should be treated with amiodipine or telmisartan.11–13 The hope is that successful treatment will result in a reduction in LV wall thickness and in clinical improvement, if heart failure is present.

**Pathophysiology**

When the LV wall is severely thickened, myocardial blood supply is compromised.14 This results in ongoing myocyte damage and death, as evidenced by an elevation in cardiac troponin I (cTn I) in cats with HCM.15,16 Cardiomyocytes that die are replaced with fibrous tissue (replacement fibrosis), as evidenced by increased concentrations of circulating biomarkers of type I collagen.17 In humans, myocardial fibrosis is most commonly identified and quantified non-invasively using MRI and a contrast agent (gadolinium).18 The first attempt to use this modality to identify myocardial fibrosis in cats with HCM failed.19 A more recent study that utilized gadolinium to calculate extracellular volume fraction, an indirect measure of myocardial fibrosis, showed that cats with HCM had an increase in this variable, presumably due to myocardial fibrosis.20 This presumption was strengthened by the fact that this variable correlated well with echocardiographic measures of diastolic dysfunction.

While thick fibrous myocardium contracts normally in systole due to the decrease in afterload (systolic myocardial wall stress) brought about by the thick LV wall, it does not relax normally in diastole. So, a common functional abnormality seen with HCM is diastolic dysfunction, which means the LV is stiff and so for any given blood volume that flows into the LV chamber in diastole, the diastolic pressure in the LV chamber is increased.21 Since the mitral valve is open during diastole, whatever pressure is present in the LV in diastole is also

---

**ABBREVIATIONS**

- 2D = two-dimensional
- ACE = angiotensin-converting enzyme
- APC = atrial premature complex
- ATE = arterial thromboembolism
- ATP = adenosine triphosphate
- cTn I = cardiac troponin I
- DCM = dilated cardiomyopathy
- DUST = discrete upper septal thickening
- DLVOTO = dynamic left ventricular outflow tract obstruction
- ECG = electrocardiogram or electrocardiography
- GI = gastrointestinal
- HCM = hypertrophic cardiomyopathy
- HOCM = hypertrophic obstructive cardiomyopathy
- LA = left atrium or left atrial
- LA:Ao = left atrial diameter to aortic root diameter ratio
- LV = left ventricle or left ventricular
- LVH = left ventricular hypertrophy
- LVNC = left ventricular noncompaction
- LVOT = left ventricular outflow tract
- NT-proBNP = amino terminal pro-B-type natriuretic peptide
- PE = pulmonary edema
- PLE = pleural effusion
- RCM = restrictive cardiomyopathy
- RT-PCR = reverse transcriptase polymerase chain reaction
- SAM = systolic anterior motion of the mitral valve
- RR = respiratory rate
- TDI = tissue Doppler imaging
- TMT = transient myocardial thickening
- VPC = ventricular premature complex

---

**Figure 1** Gross pathologic specimen of a heart showing markedly thickened left ventricular walls and papillary muscles and an enlarged left atrium. At the upper left there is a thrombus in the left auricle (white asterisk). Another thrombus is present in the body of the left atrium on the right (black asterisk), S = interventricular septum; F = left ventricular free wall; P = base of the papillary muscles; A = body of the left atrium
present in the left atrium (LA). As a result, an increase in LV diastolic pressure causes an increase in LA pressure. An increase in LA pressure results in LA enlargement. This means in cats with HCM that have clinically significant diastolic dysfunction, the LA is enlarged. In general, the higher the pressure, the greater the enlargement. The increased LA pressure also results in increased pulmonary venous pressure, and therefore enlargement of the pulmonary veins, and in increased pulmonary capillary pressure, which causes pulmonary edema (PE).32 The veins that drain the pleura that lie on the surface of the lungs (visceral pleural veins) drain into pulmonary veins, so an increase in LA and pulmonary venous pressure is also assumed to cause pleural effusion (PLE) in cats.23,24 This is what is referred to as left heart failure (see Part 1).

While cats with the commonly classified left-sided cardiomyopathies (HCM, dilated cardiomyopathy [DCM] and restrictive cardiomyopathy [RCM]) also frequently have right heart disease, in the authors’ opinion this is usually not severe enough to cause right heart failure, which can also cause PLE in cats.25-27 Additionally, while pulmonary hypertension secondary to left heart failure could theoretically cause severe enough right heart disease to lead to right heart failure, as is well recognized in dogs, pulmonary hypertension in cats with left heart failure due to a cardiomyopathy is uncommon and, when present, is usually not severe.28

Etiology

In most humans, and in most Maine Coon, Ragdoll and probably Sphynx cats, HCM is caused by a gene mutation (variant).28,29 Most commonly in humans and in Maine Coon and Ragdoll cats (and one domestic shorthair cat), HCM is caused by a mutation in a gene that encodes for a protein that contributes to the formation of the sarcomere (the contractile element that includes myosin, actin, etc).7,21 While sarcomere gene mutations most commonly cause HCM in humans, it should be noted that they can also cause DCM or RCM, as well as left ventricular noncompaction (LVNC; see Part 3).32 In HCM, the gene mutation causes the sarcomere to contract either less than normally or more than normally.23 In cats with HCM, the sarcomere is more responsive to calcium than normal, so the myocardium is hypercontractile.33 However, the exact pathophysiologic mechanism of how this sarcomeric dysfunction causes the LV wall to grow thicker remains to be elucidated.

In Maine Coon and Ragdoll cats the cause of HCM in most cases is a mutation in the myosin binding protein C gene.29 In Maine Coon cats, the A31P (c.91G>C; p.A31P) mutation creates an abnormal protein that is incorporated into the sarcomere, where it causes sarcomeric dysfunction (acting as a poison polypeptide).34 When the A31P mutation is transferred into fruit flies (Drosophila melanogaster), numerous abnormalities are produced including downregulation of small nuclear RNAs (a class of small RNA molecules that primarily guide chemical modifications of other RNAs) and the unfolded protein response.35

Maine Coon cats heterozygous for this mutation develop subtle systolic and diastolic dysfunction but usually do not develop wall thickening and so are subclinical.36,57 Those that are homozygous for the mutation commonly develop various degrees of wall thickening (HCM), some severe enough to result in severe LA enlargement and hence left heart failure or ATE.38,39 This pattern suggests an incomplete dominance mode of inheritance.40 In humans, HCM is usually inherited in an autosomal dominant pattern but occasionally other modes of inheritance are also found.41 Prevalence of the A31P mutation in Maine Coon cats ranges from 34% to 41%.42-44 Approximately 10% of the cats with the A31P mutation are homozygous (at risk for developing clinically significant HCM) and 90% are heterozygous for the mutation. A small percentage of Maine Coon cats with HCM do not have the A31P mutation and so there must be at least one more cause in this breed. Recently, a second mutation in cardiac troponin T was identified in one Maine Coon cat with HCM.45

The natural history and pathophysiology of the R820W (c.2460C>T; p.R820W) myosin binding protein C mutation in Ragdoll cats has not been as well studied but the pattern appears to be similar to that seen in Maine Coon cats.40 The same R820W mutation seen in Ragdoll cats causes both HCM and LVNC in humans.46 Ragdoll cats that are homozygous for the R820W mutation have a thicker LV wall than those that are heterozygous for the mutation and the wall thickness of cats heterozygous for the mutation is thicker than it is in cats without the mutation.47 In the UK, 27.4% of Ragdoll cats sampled in one study had the mutation (26% heterozygous, 1.4% homozygous).42

HCM is caused by a gene mutation in most affected
Maine Coon and Ragdoll cats, and likely Sphynx cats as well.

1030 JFMS CLINICAL PRACTICE
In Sphynx cats with HCM, a mutation in the Alstrom syndrome 1 (ALMS1) gene has been identified.48 Mutations in the ALMS1 gene are associated with the development of Alstrom syndrome in humans, a multisystem familial disease that can include retinal degeneration, obesity, neurosensory deafness, type 2 diabetes, and DCM and RCM.49 The variant identified in Sphynx cats has not been identified in humans and none of the cats studied had signs of Alstrom syndrome. Of the 71 Sphynx cats with HCM examined in this study, 62 had this variant (27 heterozygotes and 35 homozygotes), so not all affected cats had the variant and no Sphynx cats without HCM were examined for this variant.48 Consequently, it has not been proven that this variant is responsible for HCM in this breed. However, this G/C mutation does change a highly conserved glycine to alanine and was predicted to be deleterious (cause disease) by three computer programs designed to analyze this probability.

In humans, in addition to the type of mutation present, the type of angiotensin-converting enzyme (ACE) gene also influences how thick the LV wall becomes in patients with myosin binding protein C mutations.50 Patients homozygous for a deletion polymorphism (D/D) have a lower circulating concentration of ACE, more severe LVH, faster disease progression and more sudden death. Similarly, Ragdoll cats homozygous for a feline ACE polymorphism have a thicker LV wall than those that either do not have the polymorphism or are heterozygous for the polymorphism.51

HCM is prevalent in several other purebred cat breeds and so is likely to be heritable in them as well. These include, but are not limited to, Bengal, American Shorthair, British Shorthair, Persian and Siberian cats.51–55 One family of domestic shorthair cats with HCM has also been described.56 While Norwegian Forest Cats have characteristics of HCM, including mild wall thickening, myocyte hypertrophy, myofiber disarray and interstitial fibrosis, they also have endomyocardial fibrosis, a form of RCM, as another component of their disease (see Part 3).57

It is suspected that other cat breeds have a genetic cause of their HCM, but intensive efforts aimed at identifying such mutations have not been successful.52,58 Consequently, and since mixed-breed cats with no family history of cardiomyopathy most commonly develop HCM, it is likely there is some other unknown cause of feline HCM.

Lymphoma is a rare cause of an increase in LV wall thickness (Figure 2).85 Chemotherapy can result in resolution of the thickening.

**Physical characteristics of cats with subclinical HCM**

Cats with subclinical HCM tend to be larger, weigh more, have a higher body condition score, have longer humeri, and have higher serum insulin and insulin-like growth factor-1 concentrations than normal cats.59,60 How or even whether this is related to the etiology of feline HCM is unknown.

---

**Transient myoccardial thickening**

Transient myocardial thickening (TMT) is a sporadically identified phenomenon characterized by an HCM phenotype (ie, thick LV) that resolves spontaneously over time (supplementary files 1–3 – see list on page 1045).51–53 The principal case series describing this entity noted LA enlargement and left heart failure at presentation, but greater awareness may reveal milder cases in the future.51 As with HCM, male domestic shortshairs are most commonly affected. If these cats are looked at as a population, the LV is not as thick and the LA not as large in cats with TMT as it is in cats with left heart failure due to conventional HCM, although substantial overlap exists. Cats with TMT and heart failure tend to be younger than cats with HCM and heart failure, although substantial overlap again exists.

The serum cTn I concentration is often elevated, sometimes dramatically so, indicating acute myocardial cell damage or death. Antecedent events are common, with the most frequent being an anesthetic event or trauma, notably thermal burn injury.52 Corticosteroid administration is a recognized antecedent event in cats being presented in heart failure, and this is true for both TMT and HCM (see Part 3).5,61,64 LV wall thickness normalizes over 2–5 months. Etiology is unknown, although speculation that the thickening is due to myocardial edema and that the edema may be secondary to myocarditis has been put forth.61 One cat with TMT that was seropositive for Bartonella henselae and whose titer and circulating cTn I concentration normalized, together with resolution of LV hypertrophy, has been reported.85

The prognosis is good to excellent if a cat with TMT survives hospitalization and the early recovery period. Recurrence appears to be rare, although systematic prospective follow-up of these cases has not been undertaken.

**For cats with TMT and heart failure, generally the LV is not as thick and the LA is not as large compared with cats with left heart failure due to HCM, although substantial overlap exists.**
Dynamic left ventricular outflow tract obstruction

Systolic anterior motion of the mitral valve (SAM) causing dynamic left ventricular outflow tract obstruction (DLVOTO) is common in cats with HCM.\(^6\) SAM is due to the hypertrophied and cranially displaced papillary muscles pulling a part of the septal (anterior) leaflet of the mitral valve into a normal or narrowed left ventricular outflow tract (LVOT) (Figures 3 and 4; and supplementary files 4–7).\(^6\) Once that portion of the leaflet is in the LVOT, blood flowing from the apex of the LV into the LVOT pushes the tip farther toward the septum, usually (but not always) to the point where at least part of the leaflet contacts the base of the interventricular septum. This results in DLVOTO, a form of subaortic stenosis that progressively worsens throughout systole (Figure 5). When the mitral valve leaflet is distorted by SAM, mitral regurgitation also occurs. This produces a characteristic bilobed jet on color flow Doppler, with simultaneous systolic turbulence in the ascending aorta and LA (supplementary file 8).

Less commonly the basilar portion of the interventricular septum, particularly when it is thicker than normal, either contributes to, or causes, DLVOTO (supplementary files 9–13). At end-systole the hypertrophied septum can contact the septal leaflet of the mitral valve (if SAM is also present) or chordae tendineae.

Uncommonly an obstruction in the middle of the LV occurs.\(^6\) This happens when the papillary muscles or a papillary muscle and an LV wall squeeze together. SAM also may affect the chordae tendineae predominantly or exclusively.\(^6\)

SAM is the usual cause of a heart murmur in a cat with HCM. The murmur is often dynamic, which means it becomes louder when the cat is excited/stressed and softer when the cat relaxes. This occurs because physical or emotional stress increases the severity of SAM and of DLVOTO, in turn increasing the velocity of flow through the dynamic outflow tract obstruction. Stress is
Cats with SAM often have a longer than normal septal mitral valve leaflet. Rarely this is a congenitally malformed mitral valve, where the SAM is the primary problem and hypertrophy is a secondary abnormality. While it has been speculated that a primary, congenital mitral valve abnormality may occur in conjunction with HCM and even predispose a cat to SAM, it seems unlikely to the authors that a cat would be born with both HCM and an abnormally formed mitral valve; it seems more plausible that the affected mitral valve leaflet remodels (grows longer) in response to being stretched by the SAM. However, there is one study in humans where the anterior mitral valve leaflet was longer in young patients with an HCM-causing mutation and no hypertrophy.

**HCM or HOCM – what is appropriate terminology?**

Cats with SAM are often said to have hypertrophic obstructive cardiomyopathy (HOCM). While it is true that some cats do and some cats do not have SAM when examined, SAM can probably be induced by increasing myocardial contractility in many, if not most, cats with HCM, just as it can in humans. Consequently, it has been recommended that the term HOCM be abandoned in humans and the general term HCM be used for all patients. Since SAM in cats is now known not to be associated with increased morbidity and mortality, unlike in humans, using the general term HCM for all feline patients is probably equally justified, if not more so.

**SAM worsens when myocardial contractility increases with stress (ie, catecholamine stimulation) and decreases when contractility is lessened with a beta blocker such as atenolol.**
Pathology

The diagnosis of feline HCM is, on occasion, reliant on a gross pathologic and possibly histopathologic examination; one example is in the context of unexplained sudden death. After death, myocardium contracts (undergoes rigor) irreversibly because energy (adenosine triphosphate [ATP]) is depleted so sarcomeres can no longer relax. Consequently, the LV walls of a gross pathology specimen from a normal cat frequently look thicker than expected. Therefore, the post-mortem diagnosis of HCM cannot rely on a cursory macroscopic examination of the LV. Instead, the LV must be measurably grossly thickened, and the heart weight must be greater than normal. Generally, a normal cat heart weighs <20 g, but a heart from a cat with HCM weighs >20 g, sometimes substantially more so (as high as 38 g).77 The normal heart weight to body weight ratio is in the range of 3–4 g/kg in cats. Those with HCM have an average ratio of 6.3 g/kg.77

While histopathology can be used to reliably provide the diagnosis of HCM in humans, this is generally not the case in cats.78 The classic findings in humans of cardiomyocyte disarray, cardiomyocyte enlargement and medial hypertrophy of the walls of small coronary arteries may be present in some, but not all, cats (Figure 6).76,78 In fact, evidence from two groups suggests that cats with HCM do not have cardiomyocyte hypertrophy or hyperplasia but instead have increased interstitium filled with fibrous tissue, macrophages or lymphocytes, small vessels and degenerate cardiomyocytes.79,81 However, a recent study by another group found cardiomyocyte disarray and fibrosis via histopathologic examination and micro-CT to

The post-mortem diagnosis of HCM cannot rely on a cursory macroscopic examination of the LV. Instead, the LV must be measurably grossly thickened, and the heart weight must be greater than normal.

Prevalence

HCM is common in domestic cats. Studies have suggested that its prevalence could be as high as 15%.74–86 In purebred cats HCM is prevalent in young cats as subclinical disease.76 In mixed-breed cats, clinically apparent HCM is more common in older animals, although young cats and even kittens are also represented.4,6–36,84 This speaks to the marked heterogeneity of HCM and, as discussed in Part 1, raises the possibility that ‘HCM’ is in fact a collection of numerous subtypes of disease, all sharing the characteristic of some form of LV thickening but each with its own genetic mutation, epigenetic triggers or unknown cause, and each with its own evolution over time, ability to respond to treatment and prognosis. Retrospective case series consistently identify a male predominance, with male to female ratios typically around 3:1.4,6,34,88,89 This may reflect a true sex difference, or the aforementioned association with larger cats (eg, perhaps some normal large cats have sometimes met the echocardiographic criteria for HCM simply by being larger).90 Alternatively, as in Maine Coon cats, the prevalence of the disease could be the same in males and females, but disease severity is often worse in males so there is a male predominance in cats with clinically apparent disease.87

In clinic populations, mixed-breed cats with HCM predominate over purebred cats with HCM, likely a reflection of the larger population of mixed-breed cats. While there are some reports of families of mixed-breed cats with HCM,64,91 for the most part the disease shows up in cats with no family history of the disease. Of course, many of those cats have no family history because only rarely is the family lineage tracked or able to be tracked in mixed-breed cats.
Natural history

With HCM, LVH and the other associated changes develop over time. In Maine Coon cats, the HCM phenotype is never present at birth. While it can develop as early as 6 months of age, more commonly it becomes apparent for the first time at 2–3 years of age. In a few Maine Coon cats, the phenotypic changes of HCM show up for the first time at 6–7 years of age. The natural history of HCM in mixed-breed cats is poorly documented. While it often appears as if such cats develop HCM at an older age, in the authors’ opinion it is more likely that they have had the disease for many years prior to presentation because HCM often is ‘silent’ (devoid of auscultatory abnormalities). Still, it is common, for example, for a cat that is >10 years old to have a heart murmur first detected at that age, and for that to lead to the echocardiographic diagnosis of HCM.

Some cats with HCM develop mild to moderate LVH and never progress to severe LVH, whereas others do. In addition, while most cats that develop LA enlargement and left heart failure have severe LVH, some do not. However, most of those with severe LVH develop LA enlargement. A recent small study found that 90% of cats with both LA enlargement and an amino terminal pro-B-type natriuretic peptide (NT-proBNP) concentration >700 pmol/l went on to develop heart failure, suffer from ATE or die suddenly within 7–60 months of diagnosis. Just 30% of cats with only LA enlargement (including mild enlargement) went on to develop one of those events, while 40% with just an elevated NT-proBNP concentration developed the same sequelae.

LA enlargement is due to the development of a higher than normal LA pressure, which, as described earlier, is caused by LV diastolic dysfunction (stiff LV). Cats that develop moderate to severe LA enlargement (left atrial diameter to aortic root diameter ratio [LA:Ao] >1.8–2.0) are at risk for developing, or being in, left heart failure (PE and/or PLE). Exactly why a cat goes from the subclinical stage to severe LA enlargement to presenting in heart failure is unknown. It is presumed that diastolic dysfunction is the primary inciting factor. Diastolic function in other species deteriorates with age, so age may be a contributing factor in cats as well. LA function is decreased in cats with HCM. This may be primary dysfunction due to atrial myocardial disease and/or may arise secondarily to atrial disease caused by the enlargement. The enlargement plus the reduced function result in blood flow stasis (reduced blood flow velocity). This contributes to thrombus formation.

Endocrine diseases such as hyperthyroidism and acromegaly can cause LVH and therefore would be expected to exacerbate the changes produced by existing HCM (see Part 3). Acute stress resulting in severe tachycardia (eg, a cat fight) can result in acute deterioration of diastolic function and so precipitate acute left heart failure (PE). Anesthesia, surgery, IV fluid therapy and possibly corticosteroid administration can tip a cat with subclinical disease over into heart failure. However, most cats with HCM that present in heart failure have no apparent exacerbating disease or precipitating event. Rarely a cat with HCM will develop myocardial failure (decreased myocardial contractility). This is often termed end-stage HCM (supplementary file 14). These cats invariably present in left heart failure. On an echocardiogram there is still evidence of LV wall thickening but with accompanying evidence of systolic dysfunction, including an increased LV end-systolic diameter and reduced LV fractional shortening. The pathophysiologic mechanism is not understood. Some cats may have distinct regional (rather than global) LV wall hypokinesis or akinesia, and this portion of the wall may be thinner than normal (atrophied). These are all characteristics of chronic myocardial infarction. It is unlikely that myocardial infarction is due to coronary atherosclerosis, like it is in humans. In the authors’ opinion, more likely it is due to coronary thromboembolic disease.

Diagnosis

The definitive diagnosis of HCM is almost always made using echocardiography, although since there are other diseases that cause LV wall thickening, it is often still a diagnosis of exclusion. Other imaging modalities, such as CT and MRI, are used in human medicine and will likely be used more frequently in veterinary medicine as conscious restraining devices become more commonly used, and as machine costs and imaging times decrease (or achieve real-time status so anesthesia is no longer required; supplementary file 15). Electrocardiography (ECG) can reveal changes in some cats with HCM but is not a reliable indicator of disease. Radiography cannot be used to distinguish HCM from the other cardiomyopathies but is valuable for identifying severe LA enlargement, primarily on ventrodorsal or dorsoventral views (left auricular bulge), PE and PLE.

Most cats with HCM that present in heart failure have no apparent exacerbating disease or precipitating event.

The definitive diagnosis of HCM is almost always made using echocardiography, but CT and MRI may start to be used more frequently in the future as conscious restraining devices become more commonly used, and as machine costs and imaging times decrease.
Echocardiography

The echocardiographic diagnosis of HCM is straightforward when the disease is severe. Marked regional or global LV wall thickening, severely enlarged papillary muscles, SAM, end-systolic cavity obliteration and moderate to severe LA enlargement are hallmark findings. In the authors’ opinion, the majority, or all, of these can be identified by most veterinarians with echocardiographic training (Figure 7 and supplementary files 16–19). Moderately severe HCM is more challenging to diagnose and so requires a more skilled operator. But even skilled operators may not agree on the diagnosis in some cases, primarily because the diagnosis relies on measurements, and both intra- and interindividual variation commonly occurs with echocardiography. Heterogeneity of LV wall thickness also makes echocardiographic assessment challenging. In its mildest form, HCM probably cannot be distinguished from normal without using specialized techniques. Some cats have a normal LA, no SAM, large papillary muscles and an LV wall thickness that is borderline (Figure 8a). These cats are placed in an equivocal category, and may or may not progress to having an overt HCM phenotype (unmistakable LV hypertrophy ± LA enlargement) over time. End-systolic cavity obliteration is common with feline HCM (Figure 8b), but can be seen in some normal cats also.
LV wall thickening

Patterns of LV wall thickening are recognized in both humans and cats with HCM. The most common phenotype in humans is asymmetric hypertrophy, with the interventricular septum being thicker than the LV free wall.\textsuperscript{114} While this form is well recognized in cats, global (symmetric) hypertrophy is more common. Wall thickening confined to the LV apex is well described in humans but rarely recognized in cats, possibly because this area is more difficult to image and because papillary muscle hypertrophy is so common.\textsuperscript{115,116} A frequent echocardiographic finding in a cat is a bulge at the base of the interventricular septum (supplementary file 20). This bulge can be an isolated finding, with no other thickened areas of the LV, or can be seen in a cat with other regions of the LV that are also too thick. In humans it has been termed discrete upper septal thickening (DUST).\textsuperscript{117} When this abnormality is identified in isolation, it is unknown if it is a type of HCM; is due to normal aging, as noted in some humans; or is idiopathic.\textsuperscript{118} Many cats with a left basilar septal bulge do not progress to having other regions of the LV become too thick, which presents a diagnostic dilemma (slowly evolving HCM vs non-HCM aging change). This bulge may cause LV outflow tract narrowing and so contribute to the formation of SAM (supplementary file 21).

While it is attractive to divide HCM into specific patterns, in reality LV wall thickening can encompass most possible patterns, from extensive and diffuse to mild and segmental. Consequently, the entire LV should be carefully examined using multiple echocardiographic views to find the thickest diastolic region(s).\textsuperscript{84-86}

Upper limits for normal LV diastolic wall thickness are commonly debated. Part of this debate revolves around the difficulty identifying normal cats within a population from which to establish normal limits. Mostly this stems from the fact that subclinical HCM is so prevalent in the general feline population.\textsuperscript{94} This makes it impossible to be confident that a control population of normal cats does not contain cats with HCM. In general, however, most agree that almost all average-sized adult cats with an LV free wall and interventricular septal thickness <5 mm have no LVH and that any value >6 mm is too thick in almost all normal-sized cats.\textsuperscript{85,119} Obviously this leaves a gray zone between 5 and 6 mm. LV wall thickness does vary with body weight (by about 1 mm from 2 to 8 kg), so it is likely that anything thinner than 5 mm in smaller cats (2–3 kg) is too thick.\textsuperscript{95,120} It is debatable whether this cut-off should be greater than 6 mm in very large (>10 kg) domestic cats.

It should be noted that measurement of LV wall thickness in cats is not an exact science and so, even though numerical results for measurements often are displayed to within tenths of a millimeter (the thickness of a sheet of paper), the authors believe one should strive to round off to the nearest 0.5 mm. Even when highly skilled operators perform repeat echocardiographic examinations on cats with the knowledge they will be compared with their colleagues, LV wall thickness still varies by up to approximately 20%.\textsuperscript{111} In a cat with regional variations in wall thickness, errors can be compounded.

As stated previously, HCM is frequently regional, which can mean only one area of the LV wall is thick or can mean one region is thicker than other thick areas.\textsuperscript{121} Because of the potential for regional heterogeneity, the LV must be examined carefully using two-dimensional (2D) echocardiography. M-mode echocardiography cannot be used to screen the LV for regions of thickening, although it might be used to measure an already identified region of thickness. This is more easily accomplished using so-called anatomical (free-positioning) M-mode echocardiography, which is available on some ultrasound machines. However, in most situations the wall thickness should be measured on a diastolic 2D image.\textsuperscript{9,84-86}

Generally, this measurement should include endocardial surfaces (eg, leading to trailing edge for the interventricular septum) and care should always be taken to exclude LV and right ventricular papillary muscles, regions of endocardial thickening, areas where false tendons insert and the pericardium.\textsuperscript{9,122}

LA size and function

Assessment of LA size is important in any cat with a left-sided cardiomyopathy, including HCM, and is within reach of most veterinarians who have received training (Figure 9).\textsuperscript{110,123} A severely enlarged LA means either the cat is already in left heart failure or the cat is at high risk of developing left heart failure, since the LA pressure is inferred to be increased (LA enlargement as an expression

The LV must be examined carefully using 2D echocardiography. M-mode echocardiography cannot be used to screen the LV for regions of thickening.

\textbf{Figure 9} Right parasternal short-axis view of the aorta (Ao), body of the left atrium (LA) and left atrial appendage (LAA). The LA and LAA are severely enlarged.
of the cardiomyopathic process, in the absence of increased atrial pressure, is not a recognized entity.\textsuperscript{124} A severely enlarged LA also places the cat at risk of blood flow stasis and, in turn, thrombus formation; this is most commonly in the left auricle and can lead to ATE.

The measurement of LA:Ao is commonly used to assess LA size.\textsuperscript{125} Although there are different methods (eg, measuring at end-systole vs end-diastole; using a right parasternal short-axis or long-axis view), most often LA size is measured when at its largest (visually or at the beginning of ventricular electrical diastole) in a right parasternal short-axis basal view. When performed in this manner, the normal LA:Ao is <1.6 and when the value is >1.8–2.0 (depending on the study; median value = 2.2) or the LA diameter is >18–19 mm, the LA is enlarged enough to put the cat at risk for being in, or developing, heart failure and/or ATE (stage B2).\textsuperscript{126,127} When viewing the LA from a right parasternal short-axis view, one should strive to have the left auricle in view to use as a landmark. If possible, pulmonary veins should be excluded from the image for a more accurate assessment.

The LA is a three-dimensional structure. When it enlarges, it usually does so globally and will look large in any view, but this is not always the case (as it is in humans), especially when it is not severely enlarged.\textsuperscript{120} For example, a minority of cats will have predominant left auricular enlargement with lesser enlargement of the body of the LA, or subjectively the LA will look larger from a left apical view than from a right parasternal view. Consequently, the LA should be examined from both right parasternal and left apical views, when possible. The size of the LA from the left apical view is a subjective assessment, comparing it with the size of the right atrium, aorta and LV chamber.

The best way to deal with the reality of the LA being a three-dimensional structure is to measure LA volume rather than diameter. This conventionally is undertaken in humans with a biplane method using orthogonal planes and utilizing the modified Simpson’s method of discs in the ultrasound software package to calculate volume.\textsuperscript{126,128} Unfortunately, obtaining orthogonal planes of the LA in the cat is difficult and has only begun to be studied. One investigation has used a simplified method utilizing a single plane from both the right parasternal long-axis four-chamber view and the left apical four-chamber view to make a rough estimate of LA volume.\textsuperscript{127} This method did not outperform linear measurements of LA size. In another study, LA ejection fraction in cats with HCM causing heart failure was significantly different from LA ejection fraction in cats with subclinical HCM.\textsuperscript{126}

Other measures of LA function are also altered in cats with heart failure due to HCM. Measures that are decreased include left auricular flow velocity and mitral A wave velocity.\textsuperscript{126,129}

It stands to reason that the pulmonary veins are also enlarged in a cat in left heart failure, since high hydrostatic venous pressures are the hallmark of heart failure and veins distend easily. Echocardiographic measures of pulmonary vein diameter can be indexed to pulmonary artery or aortic diameter. Both indices of pulmonary vein size are increased in cats in heart failure.\textsuperscript{122}

As opposed to dogs, where the LA is not known to decrease in size with diuretic administration, LA and pulmonary vein sizes are lower in both normal cats and cats in heart failure following furosemide administration (eg, median LA:Ao = 2.3 in cats in heart failure and no furosemide administration vs median LA:Ao = 1.8 in cats treated with furosemide 12 h or less before the echocardiographic examination in one study).\textsuperscript{22,130} On occasion, this can confound the diagnosis of left heart failure in a cat.

Diastolic dysfunction
Diastolic dysfunction is a characteristic feature of HCM. Diastolic function is typically assessed in cats with HCM using tissue Doppler imaging (TDI). Most commonly a pulsed wave TDI gate is placed at the lateral mitral valve annulus to measure the velocity at which it moves in early diastole (E’ wave).\textsuperscript{21} In cats with severe HCM, the E’ wave velocity is typically decreased (Figure 10).\textsuperscript{129} Global TDI analysis of diastolic function can also be assessed with higher-end echocardiographic machines. While it is interesting to assess diastolic function in cats with HCM, it often provides little additional insight for most feline patients with HCM. However, with other forms of cardiomyopathy, such as RCM, it may be diagnostic (see Part 3).

Thickened RV free wall
While the LV dominates the echocardiographic and clinical picture of HCM, the right ventricle can also be too thick. In about half the cases of feline HCM, the RV free wall is also mildly thickened.\textsuperscript{26,27}

Cardiac biomarkers
The measurement of NT-proBNP and cTn I concentrations in serum or plasma has been evaluated extensively as a means of screening for HCM in cats without heart failure (subclinical HCM) in veterinary referral hospitals and also as a means of differentiating heart failure from primary respiratory disease in cats presented for dyspnea.\textsuperscript{132–136}
In general, NT-proBNP is reasonably accurate for detecting severe HCM in a referral clinic and reasonably accurate for determining if a cat has left heart failure due to a severe cardiomyopathy when presented for dyspnea. However, it is not accurate enough to be a definitive test and so should generally be used in conjunction with other diagnostic tests, if possible. As an example, in one study approximately half the cats with subclinical HCM had a normal plasma NT-proBNP concentration (general practice population).137 Other work suggests that cats in a referral population and with more advanced stages of subclinical HCM are more likely to have a higher plasma NT-proBNP concentration.138 The quantitative NT-proBNP assay is most reliable when used to assess cats referred for a heart murmur, gallop rhythm or arrhythmia or when radiographic heart enlargement is detected.137,139 In general, a plasma NT-proBNP concentration >99 pmol/l suggests that mild to severe HCM is present, although this cut-off is less reliable in a general practice feline population.137,139 Further evaluation via echocardiography is generally indicated if this sort of elevation is identified. However, owners should be made aware that some cats with a high NT-proBNP concentration, especially male cats, will have a normal echocardiogram.140

One study examined the ability of a highly sensitive cTn I assay (ADVIA; Centaur TnI-Ultra, Siemens Healthcare Diagnostics) to distinguish normal cats from cats with HCM.141 For all cats with HCM, including those in heart failure and those with ATE, the test was 92% sensitive and 95% specific (cut-off of 0.06 ng/ml). Using the same cut-off, if only cats with subclinical HCM were evaluated the test was 88% sensitive and 95% specific. In another study, a standard cTn I assay was used to identify cats with subclinical and clinical HCM. A cut-off of 0.163 ng/ml was only 62% sensitive but 100% specific at distinguishing normal cats from cats with subclinical HCM and no LA dilation. A cut-off of 0.234 ng/ml was highly sensitive (95%) but less specific (78%) for identifying cats with heart failure due to HCM.142

The ability of these tests to identify cats in the general population, with or without auscultatory abnormalities, at risk for morbidity and mortality associated with anesthesia and surgery has not been evaluated. The authors believe these tests should not be considered comprehensive methods for ruling in or ruling out HCM nor be used as part of the chemistry panel in the general population at this time.

Figure 10 Tissue Doppler imaging (TDI) of a normal cat and a cat with severe HCM. (a) A left apical four-chamber view is used for TDI echocardiography of the lateral mitral annulus. The white bars represent the position of the pulsed wave Doppler gate. Images (b) and (c) show TDI myocardial velocity of the lateral mitral annulus in a cat with severe HCM and a normal cat, depicted with different velocity scales. The normal cat (c) had a higher heart rate (HR; 220 beats per minute [bpm] vs 115 bpm in the cat with HCM) and fusion of the early (E') and late (A') diastolic waves to form an EA' wave. Fusion of the E' and A' diastolic waves does not affect the peak velocity in normal cats. Peak diastolic velocity and systolic velocity are greatly reduced in the cat with HCM (b), indicating diastolic dysfunction. S' = systolic myocardial velocity
Screening for subclinical HCM in purebred cats

The prevalence of HCM in various purebred cat populations is increased and the disease is assumed, or shown to be, heritable in numerous cat breeds. Breeders, breeding clubs and, in some countries, government agencies are interested in screening for HCM in these purebred cat populations so that an attempt can be made to identify cats with HCM, and then remove them from the breeding pool in order to reduce the prevalence of the disease in that breed. Echocardiography is the primary mode of screening for most breeds. This approach is far from ideal for several reasons:

- In most countries there is no mandate that a cat breeder screen the cats in a cattery for HCM, so some do and some do not, and some remove affected cats from their breeding program while others do not.
- Cats with subclinical disease can have such mild HCM that it is either undetectable via an echocardiogram or is difficult to distinguish from normal.
- HCM is often a disease that develops over time and breeders primarily present for screening young cats, in which the disease phenotype (e.g., detectable hypertrophy) may not yet be present.
- Breeding cats are often only examined once, generally at a young age.
- The examination is often performed in less than ideal circumstances.
- There are no training standards for the individual performing the examination.
- There is no standard for the equipment used for the examination.

Thousands of purebred cats are screened each year, despite the fact there is no evidence that this screening technique reduces the prevalence of disease. The principle of removing affected individuals from a breeding pool is logical, and has shown benefits in dogs with mitral valve disease. Currently, however, given the limitations stated above, in the authors’ opinion the process of screening and removal of affected cats from breeding programs due to HCM is in need of standardization and improved screening methods in North America and many other areas of the world.

Maine Coon and Ragdoll cats should be screened for the specific genetic mutation found in each breed. Cats are negative (wild type), heterozygous or homozygous for their particular mutation. Breeding recommendations based on these findings are relative- ly straightforward. Cats without a mutation should be screened echocardiographically for HCM, since a small percentage will have unequivocal findings of HCM due to some unknown cause. Cats that are homozygous for a mutation should not be bred since they pass on at least one abnormal copy of the gene to every kitten. In addition, they are very likely to develop HCM. In the authors’ opinion, breeders should be discouraged from breeding cats that are heterozygous for a mutation but, if a cat has other outstanding characteristics, it can be bred once. The offspring then should be tested and only those that are free of the mutation should be bred.

The A31P and R820W mutations are not present in cats other than Maine Coons and Ragdolls, respectively, so genetic testing for these mutations in other purebred cats or in most mixed-breed cats is not warranted. Plasma concentrations of NT-proBNP and cTn I should not be used alone to screen purebred breeding cats for HCM. NT-proBNP is particularly inaccurate at diagnosing HCM in Maine Coon cats.

Maine Coon and Ragdoll cats that are homozygous for a mutation should not be bred since they pass on at least one abnormal copy of the gene to every kitten.

Arrhythmias

Numerous types of arrhythmias can be identified on a resting ECG, including atrial and ventricular premature complexes (APCs and VPCs), atrial and ventricular tachycardia, and atrial fibrillation. While the ECG is sensitive for detecting atrial fibrillation, atrial fibrillation is not an easy diagnosis to make in a cat (the ECG may not be specific for atrial fibrillation in cats). Sinus rhythm with small P waves (which may be obscured by artifact) and frequent APCs and atrial tachycardia often masquerade as atrial fibrillation in cats. The ECG is insensitive for detecting sporadic APCs and VPCs in any species. A 24-h ambulatory ECG (Holter monitor) is a more sensitive means of identifying these arrhythmias.

In one study, most cats with HCM, either subclinical or clinical, had more VPCs than normal cats. Some had tens of thousands in one 24-h period. However, some had no more than a normal cat (0–13 VPCs per 24 h), a finding that has been noted in previous work. Only cats with HCM had ventricular tachycardia. There was no correlation between plasma cTn I concentration and the number of VPCs and there was no apparent relationship between the arrhythmias and prognosis. Consequently, whether or not a drug like sotalol should be used in an attempt to prevent sudden death due to ventricular fibrillation in cats with HCM and documented ventricular tachycardia remains an open question.

Whether or not a drug like sotalol should be used in an attempt to prevent sudden death due to ventricular fibrillation in cats with HCM and documented ventricular tachycardia remains an open question.
Treatment

There is no documented reason to treat a cat with subclinical HCM that has mild to severe wall thickening and a normal to mildly enlarged LA (stage B1) if the goal is to delay the onset of heart failure. This is because there is no medication (including ACE inhibitors, beta blockers and spironolactone) that has been shown to reduce hypertrophy or slow progression of the disease, if it is destined to progress. Therefore, the best that can be done is to:
- Monitor the cat for the development of severe LA enlargement (so that antiplatelet/anticoagulant therapy can be started);
- Avoid treatments that can trigger heart failure iatrogenically (eg, injudicious fluid therapy);
- Not breed the cat if it is sexually intact;
- Monitor for the onset of left heart failure (PE and/or PLE), if the LA is moderately to severely enlarged.

The best way to do the last is to have the owner monitor the cat’s sleeping respiratory rate (RR; normal is <30 breaths/min) and to maintain a log. In general, this should only be performed in a cat with evidence of moderate to severe LA enlargement to avoid overvigilance. The owner then needs to be instructed to call a veterinarian when the sleeping RR increases, before the onset of any severe dyspnea and hopefully avoid the all-too-common weekend or evening visit to an emergency clinic.

Antiarrhythmic agents

Beta blockers (eg, atenolol) and calcium channel blockers (eg, diltiazem) have been used to treat subclinical feline HCM. The use of beta blockers is partly an extrapolation from their use in humans where they alleviate angina and exertional dyspnea, particularly in patients with SAM. There is no evidence that cats with HCM experience chest pain (angina) and they only rarely exert themselves, so extrapolation from humans in this regard is difficult to justify.

Atenolol does have effects on cardiac function in cats. In normal cats at doses of 6.25 mg or 12.5 mg, it reduces heart rate, peak myocardial velocity during systole (S'), peak myocardial velocity during early diastole (E'), LA fractional shortening and LA ejection fraction. So, it decreases systolic and diastolic LV function and LA systolic function. It does not reduce systolic blood pressure. In cats with HCM, with or without LA enlargement, it has similar effects on heart rate, and LV systolic and diastolic function.

Atenolol administration does not prolong 5-year survival in cats with subclinical HCM, nor does it reduce plasma NT-proBNP or cTn I concentrations in cats with subclinical HCM or improve activity level or quality of life. However, it does reduce heart rate and murmur intensity and NT-proBNP and cTn I are higher in cats with SAM due to HCM than those with HCM alone. However, in the authors’ opinion and based on most of the literature, there appears to be no justification for the use of a beta blocker in cats with subclinical HCM and mild to moderate DLVOTO due to SAM. The authors do, however, think it is justified in those with severe DLVOTO due to SAM, as explained below.

Diltiazem was popular for decades (1990s and 2000s) for treating cats with both subclinical and clinical HCM based on work undertaken in the 1990s. It has since fallen out of favor with most veterinary cardiologists because of perceived lack of efficacy and gastrointestinal (GI) side effects with some of the extended-release formulations, but not because new clinical trials provided evidence that overrode the original findings achieved with non-extended-release diltiazem.

DLVOTO, most commonly due to SAM, is a frequent therapeutic target. It has been treated with both atenolol and diltiazem, but atenolol is more efficacious at reducing the degree of dynamic obstruction (stenosis; pressure gradient; peak blood flow velocity via Doppler echocardiography). An increase in contractility worsens SAM so it stands to reason that a beta blocker (a negative inotropic agent) would reduce it. Disopyramide, an antiarrhythmic agent that is also a potent negative inotrope, is another agent used in human patients to reduce SAM, primarily in those in whom a beta blocker has been ineffective. It can be combined with a beta blocker. There is only one report of its successful use in a cat with severe DLVOTO; it was combined with carvedilol, which did not work on its own.

It stands to reason that SAM, and so the degree of DLVOTO, worsens when a cat is stressed (eg, in a veterinary clinic) and improves when a cat is resting/sleeping (75% of its life). Thus, SAM is likely at its most severe during an examination by a veterinarian and at its best at home. Therefore, in the authors’ opinion, it is illogical to try to lessen the degree of SAM in cats where stress is not a common event, particularly if medication administration is itself a stressful event. However, it is possible that a cat that has severe SAM either has the same degree of SAM at home or has the potential to generate the same or greater degree of SAM if maximally stressed at home. It has been shown that cats with SAM, for any given degree of LA size, have higher NT-proBNP and cTn I concentrations.

So, in the authors’ opinion, it is likely appropriate to treat a cat that has severe SAM peak velocity through the LVOT >4.5–5.0 m/s, The best way to monitor for the onset of left heart failure if a cat’s LA is moderately to severely enlarged is to have the owner monitor the cat’s sleeping RR, with normal being <30 breaths/min.
especially if that cat is likely to be maximally stressed while at home (eg, has access to the outdoors where it may get in a cat fight or be chased by a dog). Therefore, it is recommended that a cat with severe SAM be placed on atenolol (6.25–12.5 mg/cat PO q12h) if it can be administered relatively easily, consistently and without stress.8

Antiplatelet drugs and anticoagulants

A cat with subclinical HCM with severe LA enlargement is at risk for developing an intracardiac thrombus, most commonly in the left auricle (LA appendage) (Figure 1). Medication is indicated to try to prevent intracardiac thrombus formation in these cats. Clopidogrel, either alone or possibly in combination with rivaroxaban, is currently the only drug that has been shown to decrease the incidence of recurrent systemic ATE in a population of cats.170,171 While clopidogrel has only been shown to prevent ATE recurrence, one can probably assume that if a drug can delay or reduce the risk of ATE in a population of cats that has already experienced one bout of ATE, it can probably delay or reduce the risk of the first formation of a left auricular thrombus also. However, it is not 100% efficacious due to factors such as differences in bioavailability from cat to cat, polymorphisms in genes that encode for platelet proteins, and owner adherence/compliance. For example, while bioavailability of clopidogrel in cats is unknown, it is known that there is a correlation between plasma clopidogrel and clopidogrel metabolite concentrations and platelet inhibition, but that correlation is weak, meaning there are other factors that also play a role in determining platelet inhibition by clopidogrel.172 One of those factors is genetic polymorphisms (variants) in genes that encode for platelet adenosine diphosphate receptors.172 Caveats aside, clopidogrel (18.75 mg/cat PO q24h) is the current standard for trying to prevent both first-time and recurrent ATE.

Since clopidogrel is not 100% effective, the search continues for drugs that can be used on their own or in combination with clopidogrel to prevent ATE. A common combination in human medicine is clopidogrel and aspirin (25 mg/kg PO q48–72h).173 This combination is currently used by some veterinarians for treating cats at high risk for ATE. Both are antiplatelet drugs, so the combination is assumed to be more effective at inhibiting platelet function in cats than clopidogrel on its own. Anticoagulants, either alone or in combination with an antiplatelet drug such as clopidogrel, have not been formally studied, but may be as effective as, or more effective than, clopidogrel alone. These anticoagulants include the low molecular weight heparins (eg, enoxaparin [0.75–1 mg/kg SC q6–12h]) and the selective Xa inhibitors (eg, rivaroxaban [0.5–1 mg/kg PO q24h]; apixaban [0.625 mg/cat PO q12h]).174–178 Potential adverse effects primarily relate to bleeding complications. A recent study retrospectively examined use of clopidogrel in combination with rivaroxaban in cats with ATE, in cats with intracardiac thrombi and in cats with spontaneous echocardiographic contrast.173 Side effects (epistaxis, hematemesis, hematochezia or hematuria) occurred in 5/32 cats but none required hospitalization. It should also be noted that clopidogrel is bitter. Consideration should be given to placing the quartered tablet inside a small gelatin capsule or a pill pocket prior to administration.176

Loop diuretics

Cats with PE (stage C) need to be treated with a loop diuretic – either furosemide or torsemide (torasemide). Route of administration and dosage depend on the nature and severity of clinical signs at presentation, severity of radiographic changes and response to initial therapy. Cats presented for veterinary attention due to dyspnea and severe tachypnea (most will have an RR in the 70s or 80s) typically have severe PE and so need to be treated with high-dose parenteral furosemide. The initial dose for such severe cases is 3–6 mg/kg, preferably IV, but IM can be used if catheter placement is too stressful. A constant rate infusion of 0.5–1.0 mg/kg/h can also be administered.180 Once administered, the cat should be placed in an oxygen-enriched environment and left alone to reduce stress. A hiding box placed in the oxygen cage can significantly reduce stress for some cats.181

If the dyspnea has not improved and the RR has not decreased significantly within 1–2 h, the initial dose or higher should be repeated and reassessment performed (eg, is the radiographic/point-of-care diagnosis of PE correct and is the IV catheter patent and in the vein?). Assuming no errors in diagnosis or treatment, this same dose or higher should be administered until the dyspnea has improved and the RR is below 50 breaths/min. At that time, the dose and dosing frequency should be decreased, oftentimes dramatically, or the drug discontinued for several hours.

Dyspnea due to severe PLE should be treated with thoracocentesis, as outlined in Part 1. Cats that are stable or have been stabilized need to remain on a loop diuretic, usually for life. The dosage varies depending on the severity of the PE and needs to be titrated based on the sleeping RR (goal is to keep it <30 breaths/min).157 A typical furosemide dosage for a domestic cat is 1–2 mg/kg PO q8–12h but the dosage can increase up to 4 mg/kg PO q8h (12 mg/kg/day) and
sometimes even higher (as high as 16.6 mg/kg/day in one study)\(^{182}\). The initial dosage for torsemide is typically in the range of 0.1–0.3 mg/kg PO q24h.\(^{183}\)

A cat in stage D heart failure is refractory to furosemide administration, meaning clinical signs (eg, elevated sleeping RR [tachypnea], dyspnea) are persisting despite treatment. A cat that is not responding adequately to high-dose furosemide PO may not be responding because not enough drug is getting to the nephron due to poor bioavailability (poor absorption from the GI tract). Such a cat can be treated with parenteral (SC) furosemide (100% bioavailability) or switched to oral torsemide, which is more readily absorbed from the GI tract (higher bioavailability). Furosemide effect peaks 2–4 h after oral administration and is gone within 6 h (hence the brand name – LAsts SIX hours). Peak torsemide effect also occurs within 2–4 h after oral administration but its effect lasts at least 12 h.\(^{184}\) Torsemide dosages administered to cats refractory to furosemide in one study ranged from 0.4–1.6 mg/kg q24h.\(^{182}\) In another study dosages ranged from 0.1–0.75 mg/kg q24h, administered either once a day or divided into two doses.\(^{185}\) In general, when switching from furosemide to torsemide, the dose of torsemide is 1/10 to 1/20 that of furosemide.

Torsemide can also be used as first-line therapy, where the starting dosage is usually closer to 0.2 mg/kg PO q24h but can be higher (eg, 0.4 mg/kg PO q24h or 0.2 mg/kg PO q12h) in a cat with severe heart failure. The primary problem with using torsemide in cats in the USA is that the smallest commercial tablet is 5 mg. Compounding pharmacies may carry a smaller size. Smaller tablet sizes are available in other parts of the world (licensed for use in dogs).\(^{186,187}\)

**Pimobendan**

The use of pimobendan for the treatment of heart failure in cats with HCM is controversial. Pharmacokinetic and pharmacodynamic studies have shown that the serum concentration is 10 times higher in normal cats than seen in normal dogs for a comparable dose administered orally.\(^{188,189}\) Despite this markedly high serum concentration, LV myocardial function is only mildly increased for a short time when compared with dogs.\(^{99}\) This finding is corroborated by the fact that pimobendan can be safely administered to cats with HCM, even those with SAM.\(^{190,191}\) If pimobendan did have potent positive inotropic properties in cats, it would be expected to worsen SAM and probably worsen the clinical status of any cat with HCM. Instead it appears to be safe to administer to most cats with HCM and it does not worsen SAM.\(^{192,193,194}\) Consequently, if it is beneficial for treating heart failure in cats with HCM, it must be via a different mechanism. Pimobendan does have the potential ability to improve diastolic function (in dogs) and it has been suggested that it may improve systolic LA function, again for a short time (<3 h) in cats.\(^{194–196}\) Both could result in clinical and hemodynamic improvement in a cat in heart failure due to HCM.

Few controlled clinical trials have been performed looking at pimobendan in cats with HCM in heart failure. In one case-controlled, retrospective study, the drug appeared to produce benefit.\(^{197}\) However, in a more powerful prospective study it produced no benefit in cats with heart failure due to HCM.\(^{198}\) The approach the authors generally use is to add it to the treatment protocol for a cat that has persistent signs of heart failure despite maximum diuretic therapy (stage D). Pimobendan is not known to alter platelet function.\(^{199}\)

**ACE inhibitors**

ACE inhibitors are relatively weak therapeutic agents, especially when compared with loop diuretics. There is some evidence that they are beneficial in cats in heart failure but, in general, one should not rely on them to result in visible clinical improvement.\(^{200,201}\) One study found no benefit; however, it included cats that were not in heart failure and so the strength of its conclusions is limited.\(^{202}\) In general, the number of medications administered to a cat in heart failure should be minimized to reduce stress. If elimination of a drug might be beneficial in this regard, in the authors’ opinion one that might be safely removed is the ACE inhibitor.

**Spironolactone**

One small study looked at the use of spironolactone, in combination with a loop diuretic and an ACE inhibitor, and found it to be safe in cats in left heart failure. It also suggested that the drug prolongs survival.\(^{203}\)

**Diet**

While, in theory, a low sodium diet might be beneficial for a cat in heart failure, it is usually more important that the cat continues to eat. Any diet change that results in decreased food/calorie intake should be avoided. However, in a cat with persistent or recurrent signs of heart failure (eg, tachypnea, dyspnea, elevated sleeping RR), despite an appropriate level of diuretic therapy, a low sodium diet might be a beneficial adjunct to therapy. Certainly, adding sodium to the diet in any form (eg, commercial treats, canned fish, etc) should be avoided in these cats, as should high sodium diets. A list of cat foods and their sodium content is available.\(^{204}\)

**The number of medications administered to a cat in heart failure should be minimized to reduce stress. If elimination of a drug might be beneficial in this regard, in the authors’ opinion one that might be safely removed is the ACE inhibitor.**
One study has suggested that a diet restricted in starch, higher in protein than a control diet and supplemented with docosahexaenoic acid and eicosapentaenoic acid results in marginal decreases in LV wall thickness and cTn I in cats with HCM.\textsuperscript{205}

**Mavacamten**

A small molecule inhibitor of myocardial contractility (MYK-461; mavacamten) is in phase III clinical trials in humans for the treatment of HCM.\textsuperscript{206} It works by causing reversible inhibition of actin–myosin cross bridging.\textsuperscript{207} In research cats with HCM it decreases contractility, and so reduces SAM and DLVOTO.\textsuperscript{208} It also has the ability to improve diastolic function.\textsuperscript{209}

**Prognosis**

The prognosis for HCM depends on the stage of the disease. Many cats with mild to moderate HCM never progress to severe HCM and so have an excellent prognosis.\textsuperscript{5} However, if followed serially, a significant number of cats will progress to severe HCM. Most cats with severe LV wall thickening and moderate to severe LA enlargement (stage B2) that are not in heart failure will progress to heart failure or experience ATE. Cats that are at increased risk of developing heart failure more frequently have the following:

- A gallop sound or arrhythmia on physical examination;
- Moderate to severe LA enlargement;
- Decreased LA fractional shortening;
- Extreme LV hypertrophy (supplementary file 22);
- Decreased LV systolic function;
- Regional wall thinning with hypokinesis (presumed myocardial infarction);
- And/or a restrictive diastolic LV filling pattern.\textsuperscript{210-212}

Cats in heart failure due to HCM usually have a terminal disease. Most will die within months, not years. A few will live for up to 2 years. Most cats with ATE are euthanized on presentation but a few (~20%) survive. It is important to keep in mind that a self-fulfilling prophecy is possible, and owners of cats with ATE can be persuaded to euthanize if they perceive the prognosis to be hopeless. Therefore, the inevitability of future recurrence and chronic care need to be balanced against owners’ willingness to accept this and the occasional outlier cat that survives well beyond expectations.\textsuperscript{210,213}

For some perspective, one study examined time from diagnosis of subclinical HCM (stages B1 and B2) to onset of heart failure and found that approximately 7% of the cats developed heart failure within the first year, 20% within 5 years and 25% within 10 years.\textsuperscript{5} This is regardless of LA size at the time of diagnosis. Once in heart failure, approximately half were dead within 2 months. Overall, in cats with ATE, 70% were dead within a week. The last stands in contrast to the average survival of 11.5 months in the 37% of cats with ATE that survive the acute episode and the 20% of cats with ATE that lived 4 years or more in other studies.\textsuperscript{212,213}

The exceptions to ‘the most will die within months, not years’ rule include cats with TMT and those that are in heart failure due to stress, fluid administration or corticosteroid administration. These cats can stabilize after being treated for heart failure and may live for years.\textsuperscript{6} In cats that are in heart failure due to HCM and concurrent hyperthyroidism, successful treatment or control of the hyperthyroidism often makes it easier to control the heart failure.

It is tempting to think about using NT-proBNP concentration as a tool to help determine prognosis. To date there are no studies to suggest that baseline NT-proBNP at the time of diagnosis of heart failure can be used to help determine prognosis in a cat with HCM. There is one study that suggests that, as a group, cats that experience a greater decrease in NT-proBNP during hospitalization for treatment of heart failure live longer.\textsuperscript{214} However, how that translates to individual cats is unknown. That same study found that cats whose owners had difficulty administering medication fared worse. Therefore, adherence/compliance and ability to administer medications are key factors in treatment success. Pill pockets and small amounts of tuna can be helpful. One of the authors finds it easier to administer oral tablets and capsules using a hemostat.

**KEY POINTS**

- Although many cats with HCM do not have a heart murmur, the presence of a heart murmur makes it more likely that a cat has HCM compared with a cat without a murmur.
- Screening cats for subclinical HCM should be performed using echocardiography because measurement of circulating cardiac biomarkers and thoracic radiography are inaccurate, primarily due to low sensitivity.
- Impostors for HCM include TMT, dehydration, systemic hypertension, hyperthyroidism and acromegaly. These and other phenocopies (disorders that produce the appearance of LV thickening on an echocardiogram) evolve differently from HCM and should be ruled out in order to make the diagnosis of HCM, especially when the LV wall thickening is mild to moderate.
- Severe DLVOTO with HCM, commonly referred to as hypertrophic obstructive cardiomyopathy (HOCM), can be treated with a beta blocker. However, while the benefit of doing so would appear to be self-evident, it is extrapolated from human medicine and is unproven in feline HCM.
Supplementary material

Brief outlines of the supplementary files are provided below; fuller descriptions accompany the files that are available online at journals.sagepub.com/doi/suppl/10.1177/1098612X211020162.

- Files 1–3: Videos showing a right parasternal short-axis echocardiographic view of the LV over time from a cat with presumed HCM. The cat went on to have a diagnosis of TMT.
- Files 4 and 5: Videos showing a right parasternal long-axis echocardiographic view of the LV of a cat with SAM due to HCM.
- Files 6 and 7: Videos showing a left apical echocardiographic view of SAM in a cat with HCM.
- File 8: Video showing a right parasternal color flow Doppler echocardiographic view of the two turbulent jets commonly seen in a cat with SAM.
- Files 9–12: Video and still frames showing a right parasternal long-axis echocardiographic view of the basilar interventricular septum bulging into the LVOT in systole creating DLVOTO.
- File 13: Image of a continuous wave Doppler trace showing the late peaking signal due to DLVOTO.
- File 14: Video showing a right parasternal long-axis echocardiographic view of a cat with end-stage HCM.
- File 15: Video showing a real-time MRI of a heart from a cat with severe HCM. Courtesy of Kristin Lavely, DVM, PhD, DACVIM (Cardiology).
- Files 16–19: Videos showing echocardiographic findings in a cat with severe HCM, including global LVH, SAM, large and hyperechoic papillary muscles, a thick interventricular septum and LV free wall, and an enlarged LA.
- File 20: Video showing a right parasternal long-axis echocardiographic view from a cat with a bulge of the basilar interventricular septum.
- File 21: Video showing a right parasternal long-axis echocardiographic view of the heart from a cat with HCM. The interventricular septum, including the basilar part, is thicker than normal, with the thickened basilar septum narrowing the LVOT, making it easier for SAM to form.
- File 22: Video showing a right parasternal short-axis echocardiographic view of the LV of a cat with extreme hypertrophy due to HCM.

Conflict of interest

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The authors received no financial support for the research, authorship, and/or publication of this article.

Ethical approval

This work did not involve the use of animals and therefore ethical approval was not specifically required for publication in JFMS.

Informed consent

This work did not involve the use of animals (including cadavers) and therefore informed consent was not required. No animals or people are identifiable within this publication, and therefore additional informed consent for publication was not required.

References

1. Cirino AL and Ho C. Hypertrophic cardiomyopathy overview. In: Adam MP, Arding HH, Pagon RA, et al. eds. GeneReviews®. Seattle, WA: University of Washington, Seattle, 1993. Available at: http://www.ncbi.nlm.nih.gov/books/NBK176/.
2. Ommen SR, Mital S, Burke MA, et al. 2020 AHA/ACC guideline for the diagnosis and treatment of patients with hypertrophic cardiomyopathy: a report of the American College of Cardiology/American Heart Association Joint Committee on Clinical Practice Guidelines. J Am Coll Cardiol 2020; 76: e159–240.
3. Maron BJ. Clinical course and management of hypertrophic cardiomyopathy. N Engl J Med 2018; 379: 655–668.
4. Ferasin L, Sturgess CP, Cannon MJ, et al. Feline idiopathic cardiomyopathy: a retrospective study of 106 cats (1994–2001). J Feline Med Surg 2003; 5: 151–159.
5. Fox PK, Keene BW, Lamb K, et al. International collaborative study to assess cardiovascular risk and evaluate long-term health in cats with preclinical hypertrophic cardiomyopathy and apparently healthy cats: The REVEAL Study. J Vet Intern Med 2018; 32: 930–943.
6. Rush JE, Freeman LM, Fenollosa NK, et al. Population and survival characteristics of cats with hypertrophic cardiomyopathy: 260 cases (1990–1999). J Am Vet Med Assoc 2002; 220: 202–207.
7. Kittleson MD, Meurs KM and Harris SP. The genetic basis of hypertrophic cardiomyopathy in cats and humans. J Vet Cardiol 2015; 17 Suppl 1: S53–573.
8. Luque Fuentes V, Abbott J, Chetboul V, et al. ACVIM consensus statement guidelines for the classification, diagnosis, and management of cardiomyopathies in cats. J Vet Intern Med 2020; 34: 1062–1077.
9. Sangster JK, Panciera DL, Abbott JA, et al. Cardiac biomarkers in hyperthyroid cats. J Vet Intern Med 2014; 28: 465–472.
10. Schroepe DP. Prevalence of congenital heart disease in 76,301 mixed-breed dogs and 57,025 mixed-breed cats. J Vet Cardiol 2015; 17: 192–202.
11. Glaus TM, Elliott J, Herberich E, et al. Efficacy of long-term oral telmisartan treatment in cats with hypertension: results of a prospective European clinical trial. J Vet Intern Med 2019; 33: 413–422.
12. Sent U, Gössl R, Elliott J, et al. Comparison of efficacy of long-term oral treatment with telmisartan and benazepril in cats with chronic kidney disease. J Vet Intern Med 2015; 29: 1479–1487.
13. Peterson ME. Feline hyperthyroidism. Vet Clin North Am Small Anim Pract 1984; 14: 809–826.
14. Sciagrà R. Positron-emission tomography myocardial blood flow quantification in hypertrophic cardiomyopathy. Q J Nucl Med Mol Imaging 2016; 60: 354–361.
15. Connolly DJ, Cannata J, Boswood A, et al. Cardiac troponin I in cats with hypertrophic cardiomyopathy. J Feline Med Surg 2003; 5: 209–216.
REVIEWS / Feline cardiomyopathies – hypertrophic cardiomyopathy

16 Herndon WE, Kittleson MD, Sanderson K, et al. Cardiac troponin I in feline hypertrophic cardiomyopathy. J Vet Intern Med 2002; 16: 558–564.
17 Borget K, Duddia J, Luis Fuentes V, et al. Circulating concentrations of a marker of type I collagen metabolism are associated with hypertrophic cardiomyopathy mutation status in Ragdoll cats. J Small Anim Pract 2013; 56: 360–365.
18 Choudhury L, Mahmholdt H, Wagner A, et al. Myocardial scarring in asymptomatic or mildly symptomatic patients with hypertrophic cardiomyopathy. J Am Coll Cardiol 2002; 40: 2156–2164.
19 MacDonald KA, Wisser ER, Larson RF, et al. Comparison of myocardial contrast enhancement via cardiac magnetic resonance imaging in healthy cats and cats with hypertrophic cardiomyopathy. Am J Vet Res 2005; 66: 1891–1894.
20 Fries RC, Kadotani S, Keating SCJ, et al. Cardiac extracellular volume fraction in cats with preclinical hypertrophic cardiomyopathy. J Vet Intern Med 2021; 35: 812–822.
21 Schober KE and Chetboul V. Echocardiographic evaluation of left ventricular diastolic function in cats: hemodynamic determinants and pattern recognition. J Vet Cardiol 2015; 17 Suppl 1: S102–S133.
22 Patata V, Caivano D, Porciello F, et al. Pulmonary vein to pulmonary artery ratio in healthy and cardiomyopathic cats. J Vet Cardiol 2020; 27: 23–33.
23 Kay JM. Comparative morphologic features of the pulmonary vasculature in mammals. Am Rev Respir Dis 1983; 128: 553–557.
24 Moorhead PD and Cross RF. The subgross vascular anatomy of the feline lung. Am J Vet Res 1965; 26: 740–743.
25 Fossum TW, Miller MW, Rogers KS, et al. Chylothorax associated with right-sided heart failure in five cats. J Am Vet Med Assoc 1994; 204: 84–89.
26 Schober KE, Savino SI and Yildiz V. Right ventricular involvement in feline hypertrophic cardiomyopathy. J Vet Cardiol 2016; 18: 297–309.
27 Visser LC, Sloan CQ and Stern JA. Echocardiographic assessment of right ventricular size and function in cats with hypertrophic cardiomyopathy. J Vet Intern Med 2017; 31: 668–677.
28 Vezzosi T and Schober KE. Doppler-derived echocardiographic evidence of pulmonary hypertension in cats with left-sided congestive heart failure. J Vet Cardiol 2019; 23: 58–68.
29 Meurs KM, Sanchez X, David RM, et al. A cardiac myosin binding protein C mutation in the Maine Coon cat with familial hypertrophic cardiomyopathy. Hum Mol Genet 2005; 14: 3587–3593.
30 Meurs KM, Norgard MM, Ederer MM, et al. A substitution mutation in the myosin binding protein C gene in Ragdoll hypertrophic cardiomyopathy. Genomics 2007; 90: 261–264.
31 Schipper T, Van Poucke M, Sonck L, et al. A feline orthologue of the human MYH7 c.5647G>A (p.(Glu1883Lys)) variant causes hypertrophic cardiomyopathy in a domestic short-hair cat. Eur J Hum Genet 2019; 27: 1724–1730.
32 Dadson K, Hauck L and Billia F. Molecular mechanisms in cardiomyopathy. Clin Sci 2017; 131: 1375–1392.
33 Messer AE, Chan J, Daley A, et al. Investigations into the sarcomeric protein and Ca2+-regulation abnormalities underlying hypertrophic cardiomyopathy in cats (Felis catus). Front Physiol 2017; 8: 348. DOI: 10.3389/fphys.2017.00348.
34 van Dijk SJ, Bezold Kooiker K, Mazzalupo S, et al. The A31P missense mutation in cardiac myosin binding protein C alters protein structure but does not cause haploinsufficiency. Arch Biochem Biophys 2016; 601: 133–140.
35 Tallo CA, Duncan LH, Yamamoto AH, et al. Heat shock proteins and small nucleolar RNAs are dysregulated in a Drosophila model for feline hypertrophic cardiomyopathy. G3 (Bethesda) 2021; 11. DOI: 10.1093/g3journal/kka014.
36 Sampedrano C, Chetboul V, Mary J, et al. Prospective echocardiographic and tissue Doppler imaging screening of a population of Maine Coon cats tested for the A31P mutation in the myosin-binding protein C gene: a specific analysis of the heterogeneous status. J Vet Intern Med 2009; 23: 91–99.
37 Pellegrino A, Daniel AGT, Pereira GG, et al. Assessment of regional left ventricular systolic function by strain imaging echocardiography in phenotypically normal and abnormal Maine Coon cats tested for the A31P mutation in the MYBPC3 gene. Can J Vet Res 2017; 81: 137–146.
38 Longeri M, Ferrari P, Knafez P, et al. Myosin-binding protein C DNA variants in domestic cats (A31P, A74T, R820W) and their association with hypertrophic cardiomyopathy. J Vet Intern Med 2013; 27: 275–285.
39 Granström S, Godskens MT, Christiansen M, et al. Genotype-phenotype correlation between the cardiac myosin binding protein C mutation A31P and hypertrophic cardiomyopathy in a cohort of Maine Coon cats: a longitudinal study. J Vet Cardiol 2015; 17: S268–S281.
40 Borget K, Casamían-Sorrosal D, Helps C, et al. Association of the myosin binding protein C3 mutation (MYBPC3 R820W) with cardiac death in a survey of 236 Ragdoll cats. J Vet Cardiol 2014; 16: 73–80.
41 Wang Y, Wang Z, Yang Q, et al. Autosomal recessive transmission of MYBPC3 mutation results in malignant phenotype of hypertrophic cardiomyopathy. PloS One 2013; 8: e67087. DOI: 10.1371/journal.pone.0067087.
42 Casamian-Sorrosal D, Chong SK, Fonfara S, et al. Prevalence and demographics of the MYBPC3-mutations in ragdolls and Maine coons in the British Isles. J Small Anim Pract 2014; 55: 269–273.
43 Mary J, Chetboul V, Sampedrano CC, et al. Prevalence of the MYBPC3-A31P mutation in a large European feline population and association with hypertrophic cardiomyopathy in the Maine Coon breed. J Vet Cardiol 2010; 12: 155–161.
44 Fries R, Heaney AM and Meurs KM. Prevalence of the myosin-binding protein C mutation in Maine Coon cats. J Vet Intern Med 2008; 22: 893–896.
45 McNamara JW, Schuckman M, Becker RC, et al. A novel homozygous intronic variant in TNNT2 associates with feline cardiomyopathy. Front Physiol 2020; 11: 608473. DOI: 10.3389/fphys.2020.608473.
46 Ripoll Vera T, Monserrat Iglesias L, Hermida Prieto M, et al. The R820W mutation in the MYBPC3 gene, associated with hypertrophic cardiomyopathy in cats, causes hypertrophic cardiomyopathy and left ventricular non-compaction in humans. Int J Cardiol 2010; 145: 405–407.
47 Borget K, Stern J, Meurs KM, et al. The influence of clinical and genetic factors on left ventricular wall thickness in Ragdoll cats. J Vet Cardiol 2015; 17 Suppl 1: S258–S267.
48 Meurs KM, Williams BC, DeProspero D, et al. A deleterious mutation in the ALMS1 gene in a naturally occurring model of hypertrophic cardiomyopathy in the Sphynx cat. Orphanet J Rare Dis 2021; 16: 108. DOI: 10.1186/s13023-021-01740-5.
REVIEWS / Cardiomyopathies – hypertrophic cardiomyopathy

49 Brofferio A, Sachdev V, Hannoush H, et al. Characteristics of cardiomyopathy in Alström syndrome: prospective single-center data on 38 patients. Mol Genet Metab 2017; 121: 336–343.

50 Lechin M, Quiñones MA, Omran A, et al. Angiotensin-I converting enzyme genotypes and left ventricular hypertrophy in patients with hypertrophic cardiomyopathy. Circulation 1995; 92: 1808–1812.

51 Silverman SJ, Stern JA and Meurs KM. Hypertrophic cardiomyopathy in the Sphynx cat: a retrospective evaluation of clinical presentation and heritable etiology. J Feline Med Surg 2012; 14: 246–249.

52 Meurs KM, Norgard MM, Kuan M, et al. Analysis of 8 sarcomeric candidate genes for feline hypertrophic cardiomyopathy mutations in cats with hypertrophic cardiomyopathy. J Vet Intern Med 2009; 23: 840–843.

53 Trehiou-Sechi E, Tissier R, Gouni V, et al. Comparative echocardiographic and clinical features of hypertrophic cardiomyopathy in 5 breeds of cats: a retrospective analysis of 344 cases (2001–2011). J Vet Intern Med 2012; 26: 532–541.

54 Chetboul V, Petit A, Gouni V, et al. Prospective echocardiographic and tissue Doppler screening of a large Sphynx cat population: reference ranges, heart disease prevalence and genetic aspects. J Vet Cardiol 2012; 14: 497–509.

55 Granström S, Nyberg Godtensen MT, Christiansen M, et al. Prevalence of hypertrophic cardiomyopathy in a cohort of British Shorthair cats in Denmark. J Vet Intern Med 2011; 25: 866–871.

56 Baty CJ, Malarky DE, Atkins CE, et al. Natural history of hypertrophic cardiomyopathy and aortic thromboembolism in a family of domestic shorthair cats. J Vet Intern Med 2001; 15: 595–599.

57 Marz I, Wilkie LJ, Harrington N, et al. Familial cardiomyopathy in Norwegian Forest cats. J Feline Med Surg 2015; 17: 681–691.

58 Stern JA and Ueda Y. Inherited cardiomyopathies in veterinary medicine. Pflügers Arch 2019; 471: 745–753.

59 Freeman LM, Rush JE, Meurs KM, et al. Body size and metabolic differences in Maine Coon cats with and without hypertrophic cardiomyopathy. J Feline Med Surg 2013; 15: 74–80.

60 van Hoek I, Hodgkiss-Geene H, Bode EF, et al. Associations among echocardiography, cardiac biomarkers, insulin metabolism, morphology, and inflammation in cats with asymptomatic hypertrophic cardiomyopathy. J Vet Intern Med 2020; 34: 591–599.

61 Novo Matos J, Pereira N, Glaus T, et al. Transient myocardial thickening in cats associated with heart failure. J Vet Intern Med 2018; 32: 48–56.

62 Sharpe AN, Gunther-Harrington CT, Epstein SE, et al. Cats with thermal burn injuries from California wildfires show echocardiographic evidence of myocardial thickening and intracardiac thrombi. Sci Rep 2020; 10: 2648. DOI: 10.1038/s41598-020-59497-z.

63 Joseph JL, Oxford EM and Santilli RA. Transient myocardial thickening in a Bartonia henselae-positive cat. J Vet Cardiol 2018; 20: 198–203.

64 Smith SA, Tobias AH, Fine DM, et al. Corticosteroid-associated congestive heart failure in 12 cats. Int J Appl Res Vet Med 2004; 2: 159–170.

65 Brummer DG and Moise NS. Infiltrative cardiomyopathy responsive to combination chemotherapy in a cat with lymphoma. J Am Vet Med Assoc 1989; 195: 1116–1119.

66 Schober K and Todd A. Echocardiographic assessment of left ventricular geometry and the mitral valve apparatus in cats with hypertrophic cardiomyopathy. J Vet Cardiol 2010; 12: 1–16.

67 Levine RA, Vlahakes GJ, Lefebvre X, et al. Papillary-muscle displacement causes systolic anterior motion of the mitral valve. Experimental validation and insights into the mechanism of subaortic obstruction. Circulation 1995; 91: 1189–1195.

68 Sherrid MV, Balaram S, Kim B, et al. The mitral valve in obstructive hypertrophic cardiomyopathy: a test in context. J Am Coll Cardiol 2016; 67: 1846–1858.

69 MacLea HB, Boon JA and Bright JM. Doppler echocardiographic evaluation of midventricular obstruction in cats with hypertrophic cardiomyopathy. J Vet Intern Med 2013; 27: 1416–1420.

70 Blass KA, Schober KE, Li X, et al. Acute effects of ivabradine on dynamic obstruction of the left ventricular outflow tract in cats with preclinical hypertrophic cardiomyopathy. J Vet Intern Med 2014; 28: 838–846.

71 Kuipers NW and Szatmári V. Mitral valve dysplasia in a cat causing reversible left ventricular hypertrophy and dynamic outflow tract obstruction [article in Dutch]. Tijdschr Diergeneeskd 2011; 136: 326–331.

72 Maron MS, Olivotto I, Harrigan C, et al. Mitral valve abnormalities identified by cardiovascular magnetic resonance represent a primary phenotypic expression of hypertrophic cardiomyopathy. Circulation 2011; 124: 40–47.

73 Shah JS, Esteban MTT, Thaman R, et al. Prevalence of exercise-induced left ventricular outflow tract obstruction in symptomatic patients with non-obstructive hypertrophic cardiomyopathy. Heart Br Card Soc 2008; 94: 1288–1294.

74 Maron BJ and Maron MS. A discussion of contemporary nomenclature, diagnosis, imaging, and management of patients with hypertrophic cardiomyopathy. Am J Cardiol 2016; 118: 1897–1907.

75 Payne J, Luis Fuentes V, Boswood A, et al. Population characteristics and survival in 127 referred cats with hypertrophic cardiomyopathy (1997 to 2005). J Small Anim Pract 2010; 51: 540–547.

76 Fox PR, Liu SK and Maron BJ. Echocardiographic assessment of spontaneously occurring feline hypertrophic cardiomyopathy. An animal model of human disease. Circulation 1995; 92: 2645–2651.

77 Liu SK, Peterson ME and Fox PR. Hypertrophic cardiomyopathy and hyperthyroidism in the cat. J Am Vet Med Assoc 1984; 185: 52–57.

78 Hughes SE. The pathology of hypertrophic cardiomyopathy. Histopathology 2004; 44: 412–427.

79 Kitz S, Fonfara S, Hahn S, et al. Feline hypertrophic cardiomyopathy: the consequence of cardiomyocyte-initiated and macrophage-driven remodeling processes? Vet Pathol 2019; 56: 565–575.

80 Liu SK, Maron BJ and Tilley LP. Feline hypertrophic cardiomyopathy: gross anatomic and quantitative histologic features. Am J Pathol 1981; 102: 388–395.

81 Khor KH, Campbell FE, Owen H, et al. Myocardial collagen deposition and inflammatory cell infiltration in cats with preclinical hypertrophic cardiomyopathy. Vet J 2014; 203: 161–168.

82 Novo Matos J, Garcia-Canadilla P, Simcock IC, et al. Micro-computed tomography (micro-CT) for the assessment of myocardial disarray, fibrosis and ventricular mass in a feline model of hypertrophic cardiomyopathy. Sci Rep 2020; 10: 20169. DOI: 10.1038/s41598-020-76809-5.
83 Fonfara S, Kitz S, Monteith G, et al. Myocardial transcription of inflammatory and remodeling markers in cats with hypertrophic cardiomyopathy and systemic diseases associated with an inflammatory phenotype. *Rev Vet Sci* 2021; 136: 484–494.
84 Payne JR, Brodbelt DC and Luis Fuentes V. Cardiomyopathy prevalence in 780 apparently healthy cats in rehoming centres (the CatScan study). *J Vet Cardiol* 2015; 17 Suppl 1: S244–S257.
85 Wagner T, Fuentes VL, Payne JR, et al. Comparison of auscultatory and echocardiographic findings in healthy adult cats. *J Vet Cardiol* 2010; 12: 171–182.
86 Paige CF, Abbott JA, Elvinger F, et al. Prevalence of cardiomyopathy in apparently healthy cats. *J Am Vet Med Assoc* 2009; 234: 1398–1403.
87 Kittleson MD, Meurs KM, Munro MJ, et al. Familial hypertrophic cardiomyopathy in Maine Coon cats: an animal model of human disease. *Circulation* 1999; 99: 3172–3180.
88 Atkins CE, Gallo AM, Kurzman ID, et al. Risk factors, clinical signs, and survival in cats with a clinical diagnosis of idiopathic hypertrophic cardiomyopathy: 74 cases (1985–1989). *J Am Vet Med Assoc* 1992; 201: 613–618.
89 Hori Y, Iguchi M, Heishima Y, et al. Diagnostic utility of cardiac troponin I in cats with hypertrophic cardiomyopathy. *J Vet Intern Med* 2018; 32: 922–929.
90 Haggstrom J, Andersson AO, Falk T, et al. Effect of body weight on echocardiographic measurements in 19,866 pure-bred cats with or without heart disease. *J Vet Intern Med* 2016; 30: 1601–1611.
91 Cesta MF, Baty CJ, Keene BW, et al. Pathology of end-stage remodeling in a family of cats with hypertrophic cardiomyopathy. *Vet Pathol* 2005; 42: 458–467.
92 Ironside VA, Tricklebank PR and Boswood A. Risk indicators in cats with preclinical hypertrophic cardiomyopathy: a prospective cohort study. *J Feline Med Surg* 2021; 23: 149–159.
93 Hotta K, Chen B, Behnke BJ, et al. Exercise training reverses age-induced diastolic dysfunction and restores coronary microvascular function. *J Physiol* 2017; 595: 3703–3719.
94 Santoro A, Alvino F, Antonelli G, et al. Age-related diastolic function in amateur athletes. *Int J Cardiovasc Imaging* 2015; 31: 567–573.
95 Kiatsilapanan A and Surachetpong SD. Assessment of left atrial function in feline hypertrophic cardiomyopathy by using two-dimensional speckle tracking echocardiography. *BMC Vet Res* 2020; 16: 344. DOI: 10.1186/s12917-020-02557-3.
96 Tarumi T, Takebayashi S, Fujita M, et al. Pacing tachycardia exaggerates left ventricular diastolic dysfunction but not systolic function and regional asynergy or asynchrony in patients with hypertrophic cardiomyopathy. *Europace* 2010; 12: 1308–1315.
97 Bednarski RM. Anesthetic concerns for patients with cardiomyopathy. *Vet Clin North Am Small Anim Pract* 1992; 22: 460–465.
98 White AJ. End-stage hypertrophic cardiomyopathy in a cat. *Can Vet J* 2015; 56: 509–511.
99 Habib M, Hoss S and Rakowski H. Evaluation of hypertrophic cardiomyopathy: newer echo and MRI approaches. *Curr Cardioi Rep* 2019; 21: 75. DOI: 10.1007/s11886-019-1173-1.
100 MacDonald KA, Kittleson MD, Reed T, et al. Quantification of left ventricular mass using cardiac magnetic resonance imaging compared with echocardiography in domestic cats. *Vet Radiol Ultrasound* 2005; 46: 192–199.
101 van Amerom JP, Lloyd DFA, Deprez M, et al. Fetal whole-heart 4D imaging using motion-corrected multi-planar real-time MRI. *Magn Reson Med* 2019; 82: 1055–1072.
102 Vititoe KP, Fries RC, Joslyn S, et al. Detection of intra-cardiac thrombi and congestive heart failure in cats using computed tomographic angiography. *Vet Radiol Ultrasound* 2018; 59: 412–422.
103 Oliveira CR, Ranallo FN, Pijanowski GJ, et al. The VetMousetrap: a device for computed tomographic imaging of the thorax of awake cats. *Vet Radiol Ultrasound* 2011; 52: 41–52.
104 Romito G, Guglielmini C, Mazzarella MO, et al. Diagnostic and prognostic utility of surface electrocardiography in cats with left ventricular hypertrophy. *J Vet Cardiol* 2018; 20: 364–375.
105 Schwerin A, Skrodzki M and Trautvetter E. The valuation of ST segment deviations in ECG of diseased cats [article in German]. *Kleintierzpraxis* 2002; 47: 393–405.
106 Kvart K and Strömberg V. iPhone ECG: a new simple method to detect hypertrophic cardiomyopathy (HCM) in cats? [abstract] *J Vet Intern Med* 2015; 29: 1147.
107 Elsharkawy SH and Torad FA. Prominent J wave in cats with hypertrophic cardiomyopathy. *J Vet Med Sci* 2021; 83: 1093–1097.
108 Schober KE, Maerz I, Ludewig E, et al. Diagnostic accuracy of electrocardiography and thoracic radiography in the assessment of left atrial size in cats: comparison with transthoracic 2-dimensional echocardiography. *J Vet Intern Med* 2007; 21: 709–718.
109 Kittleson MD. Case 5: Case studies in small animal cardiovascular medicine. https://viper.vetmed.ucdavis.edu/public/cardio_kittleson/cases/case5/case5.htm.
110 Tse YC, Rush JE, Cunningham SM, et al. Evaluation of a training course in focused echocardiography for noncardiologist house officers. *J Vet Emerg Crit Care (San Antonio)* 2013; 23: 268–273.
111 Chetboul V, Pouchelon JL, Muller C, et al. Effects of inter- and intra-observer variability on echocardiographic measurements in awake cats. *J Vet Med Physiol Pathol Clin Med* 2003; 50: 326–331.
112 Chetboul V, Sampedro CC, Gouni V, et al. Two-dimensional color tissue Doppler imaging detects myocardial dysfunction before occurrence of hypertrophy in a young Maine Coon cat. *Vet Radiol Ultrasound* 2006; 47: 295–300.
113 Haggstrom J, Luis Fuentes V and Wess G. Screening for hypertrophic cardiomyopathy in cats. *J Vet Cardiol* 2015; 17 Suppl 1: S134–S149.
114 Parato VM, Antonecchi V, Sozzi F, et al. Echocardiographic diagnosis of the different phenotypes of hypertrophic cardiomyopathy. *Cardiovac Ultrasound* 2016; 14: 30. DOI: 10.1186/s12974-016-0072-5.
115 Ünlü S, Özden Tok Ö, Avci Demir F, et al. Differential diagnosis of apical hypertrophic cardiomyopathy and apical displacement of the papillary muscles: a multimodality imaging point of view. *Echocardiography* 2021; 38: 103–113.
116 Louie EK and Maron BJ. Apical hypertrophic cardiomyopathy: clinical and two-dimensional echocardiographic assessment. *Ann Intern Med* 1987; 106: 663–670.
117 Diaz T, Pencina MJ, Benjamin EJ, et al. Prevalence, clinical correlates, and prognosis of discrete upper septal thickening on echocardiography: the Framingham Heart Study. *Echocardiography* 2009; 26: 247–253.
118 Canepa M, Matti O, David M, et al. Prevalence, clinical correlates, and functional impact of subaortic ventricular septal bulge (from the Baltimore Longitudinal Study of Aging). *Am J Cardiol* 2014; 114: 796–802.
119 Gundler S, Tidholm A and Haggstrom J. Prevalence of myocardial hypertrophy in a population of asymptomatic Swedish Maine Coon cats. Acta Vet Scand 2008; 50: 22, DOI: 10.1186/1751-0147-50-22.
120 Schober K, Savino S and Vedat Y. Reference intervals and allometric scaling of two-dimensional echocardiographic measurements in 150 healthy cats. J Vet Med Sci 2017; 79: 1764–1771.
121 Brizard D, Amberger C, Hartnack S, et al. Phenotypes and echocardiographic characteristics of a European population of domestic shorthair cats with idiopathic hypertrophic cardiomyopathy. Schweiz Arch Tierheilkd 2009; 151: 529–538.
122 Wolf OA, Imgrund M and Wess G. Echocardiographic assessment of feline false tendons and their relationship with focal thickening of the left ventricle. J Vet Cardiol 2017; 19: 14–23.
123 Loughran KA, Rush JE, Rozanski EA, et al. The use of focused cardiac ultrasound to screen for occult heart disease in asymptomatic cats. J Vet Intern Med 2019; 33: 1892–1901.
124 Laudhittirut T, Rujivipat N, Saringkarisate K, et al. Accuracy of methods for diagnosing heart diseases in cats. Vet World 2020; 13: 872–878.
125 Abbott JA and MacLean HN. Two-dimensional echocardiographic assessment of the feline left atrium. J Vet Intern Med 2006; 20: 111–119.
126 Linney CJ, Dukes-McEwan J, Stephenson HM, et al. Left atrial size, atrial function and left ventricular diastolic function in cats with hypertrophic cardiomyopathy. J Small Anim Pract 2014; 55: 198–206.
127 Duler L, Scollan KF and LeBlanc NL. Left atrial size and volume in cats with primary cardiomyopathy with and without congestive heart failure. J Vet Cardiol 2019; 24: 36–47.
128 Khoo CW, Krishnamoorthy S, Lim HS, et al. Assessment of left atrial volume: a focus on echocardiographic methods and clinical implications. Clin Res Cardiol 2011; 100: 97–105.
129 Schober KE and Maerz I. Doppler echocardiographic assessment of left atrial appendage flow velocities in normal cats. J Vet Cardiol 2005; 7: 15–25.
130 Campbell FE and Kittleson MD. The effect of hydration status on the echocardiographic measurements of normal cats. J Vet Intern Med 2007; 21: 1008–1015.
131 MacDonald KA, Kittleson MD, Garcia-Nolen T, et al. Tissue Doppler imaging and gradient echo cardiac magnetic resonance imaging in normal cats and cats with hypertrophic cardiomyopathy. J Vet Intern Med 2006; 20: 627–634.
132 Wurtinger G, Henrich E, Hildebrandt N, et al. Assessment of a bedside test for N-terminal pro B-type natriuretic peptide (NT-proBNP) to differentiate cardiac from non-cardiac causes of pleural effusion in cats. BMC Vet Res 2017; 13: 394. DOI: 10.1186/s12917-017-1319-6.
133 Ward JL, Lisciandro GR, Ware WA, et al. Evaluation of point-of-care thoracic ultrasound and NT-proBNP for the diagnosis of congestive heart failure in cats with respiratory distress. J Vet Med Intern Med 2018; 32: 1530–1540.
134 Fox PR, Rush JE, Reynolds CA, et al. Multicenter evaluation of plasma N-terminal probrain natriuretic peptide (NT-pro BNP) as a biochemical screening test for asymptomatic (occult) cardiomyopathy in cats. J Vet Intern Med 2011; 25: 1010–1016.
135 Fox PR, Oyama MA, Reynolds C, et al. Utility of plasma N-terminal pro-brain natriuretic peptide (NT-proBNP) to distinguish between congestive heart failure and non-cardiac causes of acute dyspnea in cats. J Vet Cardiol 2009; 11 Suppl 1: S51–S61.
136 Machen MC, Oyama MA, Gordon SG, et al. Multi-centered investigation of a point-of-care NT-proBNP ELISA assay to detect moderate to severe occult (pre-clinical) feline heart disease in cats referred for cardiac evaluation. J Vet Cardiol 2014; 16: 245–255.
137 Lu T, Côté, Kuo Y, et al. Point-of-care N-terminal pro B-type natriuretic peptide assay to screen apparently healthy cats for cardiac disease in general practice. J Vet Intern Med 2021; 35: 1663–1672.
138 Wess G, Daisenberger P, Mahling M, et al. Utility of measuring plasma N-terminal pro-brain natriuretic peptide in detecting hypertrophic cardiomyopathy and differentiating grades of severity in cats: NT-proBNP and hypertrophic cardiomyopathy in cats. Vet Clin Pathol 2011; 40: 237–244.
139 Oyama MA, Boswood A, Connolly DJ, et al. Clinical usefulness of an assay for measurement of circulating N-terminal pro-B-type natriuretic peptide concentration in dogs and cats with heart disease. J Am Vet Med Assoc 2013; 243: 71–82.
140 Hanås S, Holst BS, Höglund K, et al. Effect of feline characteristics on plasma N-terminal-prohormone B-type natriuretic peptide concentration and comparison of a point-of-care test and an ELISA test. J Vet Intern Med 2020; 34: 1187–1197.
141 Hertzschi S, Roos A and Wess G. Evaluation of a sensitive cardiac troponin T assay as a screening test for the diagnosis of hypertrophic cardiomyopathy in cats. J Vet Intern Med 2019; 33: 1242–1250.
142 Birkegärd AC, Reimann MJ, Martinussen T, et al. Breeding restrictions decrease the prevalence of myxomatous mitral valve disease in Cavalier King Charles spaniels over an 8-to 10-year period. J Vet Intern Med 2016; 30: 63–68.
143 O’Donnell K, Adin D, Atkins CE, et al. Absence of known feline MYH7 and MYBPC3 variants in a diverse cohort of cats with hypertrophic cardiomyopathy. Anim Genet 2021; 52: 542–544.
144 Singh MK, Cocchiano MF and Kittleson MD. NT-proBNP measurement fails to reliably identify subclinical hypertrophic cardiomyopathy in Maine Coon cats. J Feline Med Surg 2010; 12: 942–947.
145 Hsu A, Kittleson MD and Paling A. Investigation into the use of plasma NT-proBNP concentration to screen for feline hypertrophic cardiomyopathy. J Vet Cardiol 2009; 11 Suppl 1: S563–S570.
146 Côté E, Harpster NK, Laste NJ, et al. Atrial fibrillation in cats: 50 cases (1979–2002). J Am Vet Med Assoc 2004; 225: 256–260.
147 Ware WA. Twenty-four-hour ambulatory electrocardiography in normal cats. J Vet Intern Med 1999; 13: 175–180.
148 Hanås S, Tidholm A, Egvall A, et al. Twenty-four hour Holter monitoring of unsedated healthy cats in the home environment. J Vet Cardiol 2009; 11: 17–22.
149 Hanås S, Tidholm A and Holst BS. Ambulatory electrocardiogram recordings in cats with primary asymptomatic hypertrophic cardiomyopathy. J Feline Med Surg 2017; 19: 158–164.
150 Bartoszuk U, Keene BW, Baron Toaldo M, et al. Holter monitoring demonstrates that ventricular arrhythmias are common in cats with uncompensated and compensated hypertrophic cardiomyopathy. Vet J 2019; 243: 21–25.
151 Jackson BL, Adin DB and Lehmkuhl LB. Effect of atenolol on heart rate, arrhythmias, blood pressure, and dynamic left ventricular outflow tract obstruction in cats with subclinical hypertrophic cardiomyopathy. J Vet Cardiol 2015; 17 Suppl 1: S296–S303.
152 Schober KE, Zientek J, Li X, et al. Effect of treatment with atenolol on 5-year survival in cats with preclinical (asymptomatic) hypertrophic cardiomyopathy. *J Vet Cardiol* 2013; 15: 93–104.
153 Fox PR and Schober KA. Management of asymptomatic (occult) feline cardiomyopathy: challenges and realities. *J Vet Cardiol* 2015; 17 Suppl 1: S130–S138.
154 MacDonald KA, Kittleton MD, Larson RF, et al. The effect of ramipril on left ventricular mass, myocardial fibrosis, diastolic function, and plasma neurohormones in Maine Coon cats with familial hypertrophic cardiomyopathy without heart failure. *J Vet Intern Med* 2006; 20: 1093–1105.
155 MacDonald KA, Kittleton MD, Kass PH, et al. Effect of spironolactone on diastolic function and left ventricular mass in Maine Coon cats with familial hypertrophic cardiomyopathy. *J Vet Intern Med* 2008; 22: 335–341.
156 Coleman AE, DeFrancesco TC, Griffiths EH, et al. Atenolol in cats with subclinical hypertrophic cardiomyopathy: a double-blind, placebo-controlled, randomized clinical trial of effect on quality of life, activity, and cardiac biomarkers. *J Vet Cardiol* 2020; 30: 77–91.
157 Ljungvall I, Rishniw M, Porciello F, et al. Sleeping and resting respiratory rates in healthy adult cats and cats with subclinical heart disease. *J Feline Med Surg* 2014; 16: 281–290.
158 Porciello F, Rishniw M, Ljungvall I, et al. Sleeping and resting respiratory rates in dogs and cats with medically-controlled left-sided congestive heart failure. * Vet J* 2016; 207: 164–168.
159 Ammirati E, Contr R, Coppini R, et al. Pharmacological treatment of hypertrophic cardiomyopathy: current practice and novel perspectives: pharmacological treatment of HCM. *Eur J Heart Fail* 2016; 18: 1106–1118.
160 Sugimoto K, Aoki T and Fujii Y. Effects of atenolol on left atrial and left ventricular function in healthy cats and in cats with hypertrophic cardiomyopathy. *J Vet Med Sci* 2020; 82: 546–552.
161 Riesen SC, Schober KE, Cervecen RM, et al. Comparison of the effects of ivabradine and atenolol on heart rate and echocardiographic variables of left heart function in healthy cats. *J Vet Intern Med* 2011; 25: 469–476.
162 Jung SW and Kittleton MD. The effect of atenolol on NT-proBNP and troponin in asymptomatic cats with severe left ventricular hypertrophy because of hypertrophic cardiomyopathy: a pilot study. *J Vet Intern Med* 2011; 25: 1044–1049.
163 See J, Payne JR, Novo Matos J, et al. Biomarker changes with systolic anterior motion of the mitral valve in cats with hypertrophic cardiomyopathy. *J Vet Intern Med* 2020; 34: 1718–1727.
164 Bright JM, Golden AL, Gompf RE, et al. Evaluation of the calcium-channel blocking agents diltiazem and verapamil for treatment of feline hypertrophic cardiomyopathy. *J Vet Intern Med* 1991; 5: 272–282.
165 Wall M, Calvert CA, Sanderson SL, et al. Evaluation of extended-release diltiazem once daily for cats with hypertrophic cardiomyopathy. *J Am Anim Hosp Assoc* 2005; 41: 98–103.
166 Wey AC and Kittleson MD. Comparison of the effect of intravenous diltiazem and esmolol to reduce left ventricular outflow tract velocity and heart rate in cats with hypertrophic obstructive cardiomyopathy [abstract]. *J Vet Intern Med* 2000; 14: 335.
167 Hezzell MJ and Oyama MA. Effect of atenolol therapy on echocardiographic parameters in cats with hypertrophic obstructive cardiomyopathy [abstract]. ACVIM Forum, 2012.
168 Musat D and Sherrid MV. Pathophysiology of hypertrophic cardiomyopathy determines its medical treatment. *Anadolu Kardiyol Derg* 2006; 6 Suppl 2: 9–17.
169 Hori Y, Fujimoto E, Nishikawa Y, et al. Left ventricular outflow tract pressure gradient changes after carvedilol-disopyramide cotherapy in a cat with hypertrophic obstructive cardiomyopathy. *J Vet Cardiol* 2020; 29: 40–46.
170 Hogan DF, Fox PR, Jacob K, et al. Secondary prevention of cardiogenic arterial thromboembolism in the cat: the double-blind, randomized, positive-controlled feline arterial thromboembolism; clopidogrel vs. aspirin trial (FAT CAT). *J Vet Cardiol* 2015; 17 Suppl 1: S506–S517.
171 Lo ST, Walker AL, Georges CJ, et al. Dual therapy with clopidogrel and rivaroxaban in cats with thromboembolic disease. *J Feline Med Surg*. Epub ahead of print 10 May 2021. DOI: 10.11177/1098612X211013736.
172 Li RH, Stern JA, Ho V, et al. Platelet activation and clopidogrel effects on ADP-induced platelet activation in cats with or without the A31P mutation in MYBPC3. *J Vet Intern Med* 2016; 30: 1619–1629.
173 Hogan DF. Feline cardiogenic arterial thromboembolism: prevention and therapy. *Vet Clin North Am Small Anim Pract* 2017; 47: 1065–1082.
174 Blais M-C, Bianco D, Goggs R, et al. Consensus on the rational use of antithrombotics in veterinary critical care (CURATIVE): domain 3-defining antithrombotic protocols. *J Vet Emerg Crit Care (San Antonio)* 2019; 29: 60–74.
175 Van De Wiele CM, Hogan DF, Green HW III, et al. Antithrombotic effect of enoxaparin in clinically healthy cats: a venous stasis model. *J Vet Emerg Int Med* 2010; 24: 185–191.
176 Dixon-Jimenez AC, Brainard BM, Brooks MB, et al. Pharmacokinetic and pharmacodynamic evaluation of oral rivaroxaban in healthy adult cats. *J Vet Emerg Crit Care (San Antonio)* 2016; 26: 619–629.
177 Myers JA, Wittenburg LA, Oliver CS, et al. Pharmacokinetics and pharmacodynamics of the factor Xa inhibitor apixaban after oral and intravenous administration to cats. *Am J Vet Res* 2015; 76: 732–738.
178 Weder C, Oliver C, Wittenburg L, et al. Multi-dose pharmacokinetics and pharmacodynamics of the commercially available formulation of oral apixaban in cats: a pilot study [abstract]. ACVIM Forum, 2015. https://beta.vis.com/members/cms/project/defaultadv1.asp?pxid=5397332&pid=113565&. 179 Goggs R, Blais MC, Brainard BM, et al. American College of Veterinary Emergency and Critical Care (ACVECC) Consensus on the Rational Use of Antithrombotics in Veterinary Critical Care (CURATIVE) guidelines: small animal. *J Vet Emerg Crit Care (San Antonio)* 2019; 29: 12–38.
180 Ohad DG, Segev Y, Kelmer E, et al. Constant rate infusion vs. intermittent bolus administration of IV furosemide in 100 pets with acute left-sided congestive heart failure: a retrospective study. *Vet J* 2018; 236: 70–75.
181 van der Leij WJR, Selman LDAM, Vernooij JCM, et al. The effect of a hiding box on stress levels and body weight in Dutch shelter cats; a randomized controlled trial. *PloS One* 2019; 14. DOI: 10.1371/journal.pone.0223492.
182 Giatis IZ, Nguyenba TP, Oyama MA, et al. Use of torsemide in 17 cats with advanced congestive heart failure [abstract]. ACVIM Forum, 2014. http://beta.vis.com/doc/?id=6293271.
183 Poissonnier C, Ghazal S, Passavin P, et al. Tolerance of pimobendan in cats with congestive heart failure: a retrospective study on 21 cases (2016–2019). BMC Vet Res 2020; 16: 339. DOI: 10.1186/s12917-020-02554-6.

184 Uechi M, Matsuoka M, Kuwajima E, et al. The effects of the loop diuretics furosemide and torsemide on diuresis in dogs and cats. J Vet Med Sci 2003; 65: 1057–1061.

185 McDonald R. Use of pimobendan in cats for congestive heart failure. 26th ECVIM-CA congress, 2016. https://beta.vin.com/members/cms/project/defaultadv1.asp?id=747194&pid=157406.

186 Vetoquinol. UpCard. https://www.vetoquinol.co.uk/content/upcard.

187 European Medicines Agency. Isemid epar product information. https://www.ema.europa.eu/en/documents/product-information/isemid-epar-product-information_en.pdf.

188 Yata M, McLachlan AJ, Foster DJR, et al. Pharmacokinetics and cardiovascular effects following a single oral administration of a nonaqueous pimobendan solution in healthy dogs. J Vet Pharmacol Ther 2016; 39: 45–53.

189 Hanzliková AS, Gehring R, Kukanich B, et al. Pharmacokinetics of oral pimobendan in healthy cats. J Vet Cardiol 2012; 14: 489–496.

190 Yata M, McLachlan AJ, Foster DJR, et al. Single-dose pharmacokinetics and cardiovascular effects of oral pimobendan in healthy cats. J Vet Cardiol 2016; 18: 310–325.

191 Gordon SG, Saunders AB, Roland RM, et al. Effect of oral administration of pimobendan in cats with heart failure. J Am Vet Med Assoc 2012; 241: 89–94.

192 Ward JL, Kussin EZ, Tropp MA, et al. Retrospective evaluation of the safety and tolerability of pimobendan in cats with obstructive vs nonobstructive cardiomyopathy. J Vet Intern Med 2020; 34: 2211–2222.

193 MacGregor JM, Rush JE, Laste NJ, et al. Use of pimobendan in 170 cats (2006–2010). J Vet Cardiol 2011; 13: 251–260.

194 Oldach MS, Ueda Y, Ontiveros ES, et al. Cardiac effects of a single dose of pimobendan in cats with hypertrophic cardiomyopathy; a randomized, placebo-controlled, crossover study. Front Vet Sci 2019; 6: 15. DOI: 10.3389/fvets.2019.00015.

195 Asano H, Ishizaka S, Kameyama T, et al. Disparate inotropic and lusitropic responses to pimobendan in conscious dogs with tachycardia-induced heart failure. J Cardiovasc Pharmacol 1994; 23: 268–274.

196 Baron Toaldo M, Pollesel M and Diana A. Effect of pimobendan on left atrial function: an echocardiographic pilot study in 11 healthy cats. J Vet Cardiol 2020; 28: 37–47.

197 Reina-Doreste Y, Stern JA, Keene BW, et al. Case-control study of the effects of pimobendan on survival time in cats with hypertrophic cardiomyopathy and congestive heart failure. J Am Vet Med Assoc 2014; 245: 534–539.

198 Schober KE, Rush JE, Luis Fuentes V, et al. Effects of pimobendan in cats with hypertrophic cardiomyopathy and recent congestive heart failure: results of a prospective, double-blind, randomized, nonpivotal, exploratory field study. J Vet Intern Med 2021; 35: 789–800.

199 Shipley EA, Hogan DF, Faikpui NN, et al. In vitro effect of pimobendan on platelet aggregation in dogs. Am J Vet Res 2013; 74: 403–407.

200 Rush JE, Freeman LM, Brown DJ, et al. The use of enalapril in the treatment of feline hypertrophic cardiomyopathy. J Am Anim Hosp Assoc 1998; 34: 38–41.

201 Amberger CN, Glardon O, Glaus T, et al. Effects of benazepril in the treatment of feline hypertrophic cardiomyopathy. Results of a prospective, open-label, multicenter clinical trial. J Vet Cardiol 1999; 1: 19–26.

202 King JN, Martin M, Chetboul V, et al. Evaluation of benazepril in cats with heart disease in a prospective, randomized, blinded, placebo-controlled clinical trial. J Vet Intern Med 2019; 33: 2599–2571.

203 James R, Guillett E, Garelli-Paar C, et al. The SEISICAT study: a pilot study assessing efficacy and safety of spironolactone in cats with congestive heart failure secondary to cardiomyopathy. J Vet Cardiol 2018; 20: 1–12.

204 Tufts University, Cummings School of Veterinary Medicine. Reduced sodium foods – feline. https://heartsmart.vet.tufts.edu/wp-content/uploads/low-sodium-diet-lists-nov-2019-cats.pdf.

205 van Hoek I, Hodgkiss-Geere H, Bode EF, et al. Association of diet with left ventricular wall thickness, troponin I and IGF-1 in cats with subclinical hypertrophic cardiomyopathy. J Vet Intern Med 2020; 34: 2197–2210.

206 Olivotto I, Oreziaki A, Barriales-Villa R, et al. Mavacamten for treatment of symptomatic obstructive hypertrophic cardiomyopathy (EXPLORER-HCM): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet 2020; 396: 759–769.

207 Tuohy CV, Kaul S, Song HK, et al. Hypertrophic cardiomyopathy: the future of treatment. Eur J Heart Fail 2020; 22: 226–240.

208 Stern JA, Markova S, Ueda Y, et al. A small molecule inhibitor of sarcomere contractility acutely relieves left ventricular outflow tract obstruction in feline hypertrophic cardiomyopathy. PLoS One 2016; 11: e0168407. DOI: 10.1371/journal.pone.0168407.

209 del Rio CL, Ueyama Y, Baker DC, et al. In vivo cardiac effects of mavacamten (MYK-461): evidence for negative inotropy and improved compliance. Circulation 2017; 136 Suppl 1: A20593.

210 Payne JR, Borget K, Brodbelt DC, et al. Risk factors associated with sudden death vs. congestive heart failure or arterial thromboembolism in cats with hypertrophic cardiomyopathy. J Vet Cardiol 2015; 17 Suppl 1: S318–S328.

211 Payne JR, Borget K, Connolly DJ, et al. Prognostic indicators in cats with hypertrophic cardiomyopathy. J Vet Intern Med 2013; 27: 1427–1436.

212 Locatelli C, Pradelli D, Campo G, et al. Survival and prognostic factors in cats with restrictive cardiomyopathy: a review of 90 cases. J Feline Med Surg 2018; 20: 1138–1143.

213 Laste NJ and Harpster NK. A retrospective study of 100 cases of feline distal aortic thromboembolism: 1977–1993. J Am Anim Hosp Assoc 1995; 31: 492–500.

214 Pierce KV, Rush JE, Freeman LM, et al. Association between survival time and changes in NT-proBNP in cats treated for congestive heart failure. J Vet Intern Med 2017; 31: 678–684.

Available online at jfms.com

Article reuse guidelines: sagepub.co.uk/journals-permissions

For reuse of images only, contact the corresponding author

This paper was handled and processed by the American Editorial Office (AAFP) for publication in JFMS

Accepted for publication: 2 May 2021

JFMS CLINICAL PRACTICE 1051