Research Article

Agromyces arachidis sp. nov. Isolated from a Peanut (Arachis hypogaea) Crop Field

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A Gram-positive, yellowish bacterium strain AK-1T was isolated from soil sample collected from peanut (Arachis hypogaea) crop field and studied by using a polyphasic approach. The organism had morphological and chemotaxonomic properties consistent with its classification in the genus Agromyces. Phylogenetic analysis of the 16S rRNA gene sequence showed that strain AK-1T was closely related to Agromyces aurantiacus (98.6%) followed by Agromyces soli (98.3%), Agromyces tropicus (97.6%), Agromyces ulmi (97.3%), Agromyces flavus (97.2%), and Agromyces italicus (97.0%), whereas the sequence similarity values with respect to the other Agromyces species with validly published names were between 95.3 and 96.7%. However, the DNA-DNA hybridization values obtained between strain AK-1T and other related strains were well below the threshold that is required for the proposal of a novel species. The DNA G + C content of the strain is 71.8 mol%. The above data in combination with the phenotypic distinctiveness of AK-1T clearly indicate that the strain represents a novel species, for which the name Agromyces arachidis sp. nov. is proposed. The type strain is AK-1T (=MTCC 10524T =JCM 19251T).

1. Introduction

The genus Agromyces was first proposed by Gledhill and Casida Jr [1] and later on emended by Zgurskaya et al. [2]. At present, the genus Agromyces comprises 24 species with validly published names (http://www.bacterio.net/a/agromyces.html), and all these species have been isolated from different environmental sources: soils from fertile meadows, rhizosphere, and plants to rock art paintings [1–18]. In the present study, bacterial strain AK-1T, isolated from soil sample, is described and subjected to the polyphasic taxonomy. 16S rRNA gene sequence comparison revealed that the isolate is Agromyces-like organism. The aim of the present study is to determine the exact taxonomic position of the isolate.

2. Materials and Methods

Strain AK-1T was isolated from a soil sample collected from peanut (Arachis hypogaea) crop field, Srikakulam, Andhra Pradesh, India (18°14′N latitude 83°58′E longitude), by the dilution-plate technique on tryptic soy agar medium (TSA; HiMedia) and maintained as glycerol stocks at −70°C. The reference strains A. aurantiacus (MTCC 11069T), A. soli (MTCC 11074T), A. tropicus (MTCC 11075T), A. ulmi (MTCC 10783T), A. flavus (MTCC 11103T), and A. italicus (MTCC 10784T) were obtained from the Microbial Type Culture Collection and Gene Bank (MTCC), Institute of Microbial Technology, Chandigarh, India.

Colonies and cell morphologies were studied according to standard methods [19]. The Gram reaction was determined using the HiMedia Gram staining kit according to the manufacturer's instructions. Physiological tests like growth at different temperatures ranging from 10 to 55°C and NaCl concentrations (1–15%) were performed by growing the strain on TSA supplemented with different concentrations of NaCl. The pH range (5.0–12.0) and the optimum pH for growth were examined as described by Xu et al. [20] using TSB as basal medium. For anaerobiosis, the cultures
Table 1: Differential characteristics that differentiate strain AK-1\(^T\) along with the closest species AK-1\(^T\) (MTCC 10524\(^T\)), \(A.\) \textit{aurantiacus} (MTCC 1069\(^T\)), \(A.\) \textit{soli} (MTCC 11074\(^T\)), \(A.\) \textit{tropicus} (MTCC 11075\(^T\)), \(A.\) \textit{ulmi} (MTCC 10783\(^T\)), \(A.\) \textit{flavus} (MTCC 1103\(^T\)), and \(A.\) \textit{italicus} (MTCC 10784\(^T\)).

| Characteristics | AK-1\(^T\) (MTCC 10524\(^T\)) | \(A.\) \textit{aurantiacus} (MTCC 11069\(^T\)) | \(A.\) \textit{soli} (MTCC 11074\(^T\)) | \(A.\) \textit{tropicus} (MTCC 11075\(^T\)) | \(A.\) \textit{ulmi} (MTCC 10783\(^T\)) | \(A.\) \textit{flavus} (MTCC 1103\(^T\)) | \(A.\) \textit{italicus} (MTCC 10784\(^T\)) |
|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|-----------------|
| Growth at | | | | | | | |
| 37\(^\circ\)C | + | + | + | + | − | + | + |
| 42\(^\circ\)C | − | − | + | − | − | − | − |
| 2% NaCl | − | − | + | + | − | − | + |
| 5% NaCl | − | − | + | − | − | − | − |
| pH 5.0 | + | − | − | − | − | + | + |
| pH 10.0 | − | + | + | + | − | − | + |
| pH 12.0 | − | − | + | + | − | − | + |
| Starch hydrolysis | + | + | − | + | − | + | − |
| Casein hydrolysis | − | − | − | − | − | + | − |
| Urease | − | + | − | − | − | − | − |
| Catalase | − | + | + | + | − | − | + |
| Acid production from carbohydrates | | | | | | | |
| Salicin | − | + | + | + | + | − | + |
| Mannitol | − | − | − | + | + | + | − |
| Melibiose | − | − | − | + | − | − | − |
| Galactose | − | − | + | + | + | − | − |
| Arabinose | + | − | − | + | − | + | − |
| Cellohiose | + | − | + | + | + | − | + |
| Sucrose | − | + | + | + | + | − | + |
| Xylose | + | − | − | + | + | + | − |
| Galactose | − | − | + | + | + | + | + |
| Lactose | + | + | − | + | − | − | − |
| Trehalose | + | + | + | + | + | − | − |
| Mannose | − | + | + | + | + | + | + |
| Maltose | − | − | + | + | + | − | + |
| Raffinose | − | + | + | + | + | − | + |
| Sensitivity to antibiotics (\(\mu\)g/disc) | | | | | | | |
| Nitrofurantoin (300) | S | R | R | S | S | S | S |
| Norfloxacim (10) | R | S | R | R | R | S | R |
| Polymyxin B (300) | S | S | R | S | S | S | R |
| Kanamycin (30) | S | S | R | S | S | S | R |
| Colistin (10) | R | R | R | R | S | R | S |
| Methicillin (5) | R | S | R | S | R | S | S |
| Oxacillin (5) | R | S | R | S | R | S | S |
| Gentamycin (10) | S | R | S | S | S | S | S |
| Trimethoprim (5) | S | S | R | S | S | S | S |
| Oxytetracycline (30) | S | S | R | S | S | S | R |
| Cefoxitin (30) | S | S | R | S | R | S | S |
| Biochemical tests using VITEK 2GP card | | | | | | | |
| Arginine dihydrolase 1 | − | − | − | − | − | − | + |
Plates were incubated at 30°C, nitrogen (85%), carbon dioxide (10%), and hydrogen (5%). The strains were streaked on TSA plates and placed in an anaerobic jar using anaerobic gas mixture consisting of nitrogen, carbon dioxide, and hydrogen. Freeze-dried cells for other chemotaxonomic analyses were prepared following growth of the strains in tryptic soy broth for 4 days at 30°C. The peptidoglycan structure was determined by using a hydrolysate of purified cell walls, according to Schleifer and Kandler [28], with the modification that TLC on cellulose sheets (Merck 5577) was used instead of paper chromatography. Polar lipids and menaquinones were separated by single-dimensional ascending TLC as described by Schleifer and Kandler [28], with the modification that TLC on cellulose sheets (Merck 5577) was used instead of paper chromatography. Polar lipids and menaquinones were extracted and analysed by using the methods described by Minnikin et al. [29] and Kroppenstedt [30].

Genomic DNA extraction, amplification, and sequencing were performed as described previously by Mayilraj et al. [31]. The complete sequence of the 16S rRNA gene was aligned with those of representative related taxa using the EzTaxon server (http://www.eztaxon.org/) [32]. The 16S rRNA gene sequence of AK-1T and the representative of closely related species were retrieved from the EzTaxon server and aligned using MEGA version 5.0 [32]. Phylogenetic trees were constructed using the neighbour-joining as well as maximum parsimony algorithms and maximum likelihood algorithms. Bootstrap analysis was performed to assess the confidence limits of the branching. DNA-DNA hybridization was performed by the membrane filter method [33]. The G+C content of the genomic DNA was determined spectrophotometrically (Lambda 35; Perkin Elmer) using the thermal denaturation method [34].

### 3. Results and Discussion

Detailed phenotypic properties that differentiate strain AK-1T from closely related species of the genus *Agromyces* are summarized in Table 1. Most of the chemotaxonomic properties, including the fatty acid composition, were typical of members of the genus *Agromyces*. The major menaquinone detected for the strain AK-1T is MK-12 (54.13%), while MK-I1 (14.08%) and MK-I3 (31.77%) are the other minor components; major fatty acids are anteiso-C₁₅₋₀, anteiso-C₁₇₋₀, iso-C₁₅₋₀, and iso-C₁₆₋₀ (Table 2); cell wall diagnostic amino acid is 2,4-diaminobutyric acid. Major lipids are diphosphatidylglycerol (DPG), phosphatidylglycerol (PG), two unknown phospholipids, and one unknown glycolipid (Figure 2). The almost complete 16S rRNA gene sequence

### Table 1: Continued.

| Characteristics          | AK-1T (MTCC 10524ᵀ) | A. auranicus (MTCC 11069ᵀ) | A. soli (MTCC 11074ᵀ) | A. tropicus (MTCC 11075ᵀ) | A. ulmi (MTCC 10783ᵀ) | A. flavus (MTCC 10784ᵀ) | A. italicus (MTCC 11030ᵀ) |
|--------------------------|----------------------|-----------------------------|------------------------|---------------------------|------------------------|-------------------------|---------------------------|
| DNA G + C mol%           | 71.8                 | 72.8                        | 73.4                   | 72.7                       | 72.0                   | 70.9                     | 70.8                      |
| Total lipid pattern      | DPG, PG              | DPG, PG                     | DPG, PG                | DPG, PG                    | DPG, PG                | DPG, PG                  | DPG, PG                   |
| Quinone type             | MKI2, 11, 13         | MKI2, 13                    | MHI2                   | MKI2                      | MII, 11, 10            | MKI2                    | MKI2, 13                  |

All the strains were positive at pH 8.0 and 9.0, at temperatures 25°C and 30°C, and acid production from fructose; negative at 12°C, 10%, 15% NaCl, dulcitol, inositol, sorbitol, adonitol, citrate, methyl-red, Voges-Proskauer, indole, nitrate, and gelatin liquefaction. All the strains are negative for the following biochemical tests using VITEK 2-GP card: D-amylodalin, phosphatidylinositol phospholipase C, D-xylene, β-galactosidase, Ala-Phe-Pro-arylamidase, cyclodextrin, L-aspartate arylamidase, β-galactopyranosidase, α-mannosidase, phosphatase, β-glucuronidase, L-pyrrolidonyl arylamidase, D-sorbitol, urease, polymyxin B resistance, D-galactose, D-ribose, lactose, N-acetyl-D-glucosamine, D-maltose, bacitracin resistance, novobiocin resistance, growth in 6.5% NaCl, D-mannitol, D-mannose, methyl β-D-glucopyranoside, pullulan, D-raffinose, O/129 resistance (comp. vibrio.), sucrose, D-trehalose, arginine diphosphate 2 and optochin resistance. All the strains were sensitive to triple sulphas, kanamycin, sulfonamide, novobiocin, ampicillin, and rifampicin. S: sensitive; R: resistance.
Figure 1: Phylogenetic neighbour-joining tree based on 16S rRNA gene sequences (1442 bases) showing the relationship between Agromyces arachidis AK-1T and related members of the genus Agromyces. Leifsonia lichenia 2SbT (AB278552) was used as an outgroup. Bootstrap values (expressed as percentages of 1000 replications) greater than 70% are given at nodes. Filled circles indicate that corresponding nodes were also recovered in the tree generated with maximum parsimony and maximum likelihood algorithms. Bar, 0.005% sequence variation. GenBank accession numbers are given in parentheses.

Table 2: Percentage of total cellular fatty acids from strains AK-1T (MTCC10524T), A. aurantiacus (MTCC11069T), A. soli (MTCC11074T), A. tropicus (MTCC11075T), A. ulmi (MTCC10783T), A. flavus (MTCC11013T), and A. italicus (MTCC10784T).

| Type of fatty acids | AK-1T (MTCC10524T) | A. aurantiacus (MTCC11069T) | A. soli (MTCC11074T) | A. tropicus (MTCC11075T) | A. ulmi (MTCC10783T) | A. flavus (MTCC11013T) | A. italicus (MTCC10784T) |
|---------------------|---------------------|-----------------------------|----------------------|---------------------------|----------------------|-------------------------|--------------------------|
| iso-C14:0           | 0.8                 | 0.7                         | 0.6                  | 3.0                       | 2.5                  | 1.4                     | 0.6                      |
| iso-C15:0           | 9.4                 | 7.7                         | 6.1                  | 3.6                       | 15.5                 | 2.8                     | 6.3                      |
| anteiso C15:0       | 47.7                | 32.7                        | 39.5                 | 31.9                      | 58.3                 | 41.5                    | 40.2                     |
| iso-C16:0           | 11.3                | 11.4                        | 18.1                 | 32.7                      | 1.8                  | 23.6                    | 18.6                     |
| C16:0               | 0.9                 | 2.8                         | 0.6                  | 0.5                       | 2.5                  | 0.5                     | 0.7                      |
| iso-C17:0           | 3.3                 | 3.0                         | 1.8                  | 3.3                       | 3.0                  | 0.7                     | 1.7                      |
| anteiso C17:0       | 21.86               | 31.6                        | 31.2                 | 20.6                      | 1.6                  | 23.7                    | 29.2                     |
| C18:0               | tr                  | 0.6                         | tr                   | tr                        | 1.5                  | tr                      | tr                       |
| C18:2 ω6c           | ND                  | 0.7                         | ND                   | ND                        | 1.3                  | 0.7                     | tr                       |
| iso-C19:0           | ND                  | tr                          | ND                   | tr                        | ND                   | ND                      | ND                       |

Data from the present study. Fatty acids amounting to <0.5% of the total fatty acids in all strains are not shown or shown as tr: traces. ND: not detected.

of strain AK-1T (1442 bases) was determined. Phylogenetic analysis of the 16S rRNA gene sequence showed that strain AK-1T was closely related to A. aurantiacus (98.6%) followed by A. soli (98.3%), A. tropicus (97.6%), A. ulmi (97.3%), A. flavus (97.2%), and A. italicus (97.0%). The similarities with respect to the type strains of the remaining species of the genus were significantly lower (95.3–96.7%). The 16S rRNA gene sequence-based phylogenetic analysis revealed that strain AK-1T forms a separate branch within the lineage that includes A. aurantiacus, A. soli, A. tropicus, A. ulmi, and A. flavus (Figure 1); this was also evident in the phylogenetic tree constructed using maximum parsimony and maximum likelihood algorithms (shown as closed circles at the nodes in Figure 1) where the strain was recovered as a separate clade.
The DNA-DNA hybridization values for strain AK-1\textsuperscript{T} with the closely related species were less than 56.2%, which is well below the 70% threshold value recommended for the delineation of bacterial species [35]. The levels of DNA-DNA relatedness between strain AK-1\textsuperscript{T} and other *Agromyces* species were not determined, since it has been shown that organisms with more than 3% 16S rRNA gene sequence dissimilarity belong to different genomic species [36]. On the basis of the polyphasic data presented previously, strain AK-1\textsuperscript{T} should be placed in the genus *Agromyces* within a novel species, for which we propose the name *Agromyces arachidis* sp. nov.

### 3.1. Description of *Agromyces arachidis* sp. nov. *Agromyces arachidis* sp. nov. (a. r. ch. dis. N. L. n. Arachis-idis, a botanical generic name; N. L. gen. n. *arachidis*, of Arachis, isolated from a peanut (*Arachis hypogaea* crop field).

The cells are Gram-positive, strictly aerobic, nonspore forming, and occurring in straight or curved rods. Colonies are yellowish, opaque, convex, entire and 1-2 mm in diameter on tryptic soy agar medium, and capable of growing from 25°C to 37°C, with optimum for growth at 30°C and a pH range from 6.0 to 10.0; they can tolerate up to 1.0% NaCl. Strain AK-1\textsuperscript{T} shows positive reaction for hydrolysis of starch and negative for casein hydrolysis, urease production, MR-VP reaction, hydrogen sulphide production, and nitrate reduction. Acid is produced from arabinose, xylose, inulin, and lactose; it is negative for salicin, mannitol, melibiose, galactose, sucrose, rhamnose, trehalose, mannose, maltose, and raffinose. Other detailed characteristics features are mentioned in Table I. Major polar lipids are phosphatidylglycerol (PG) and diphasphatidylglycerol (DPG), two unknown phospholipids (PL), and one unknown glycolipid (GL). The major menaquinone detected for the strain AK-1\textsuperscript{T} is MK-12 (54.1%), while MK-13 (31.7%) and MK-11 (14.0%) are the other components. The predominant fatty acids are anteiso-C\textsubscript{15:0}, anteiso-C\textsubscript{17:0}, iso-C\textsubscript{15:0}, and iso-C\textsubscript{16:0}. The diagnostic diamino acid in cell wall hydrolyzate is 2,4-diaminobutyric acid. The DNA G+C content of the strain is 71.8 mol%. The type strain, AK-1\textsuperscript{T} (=MTCC 10524\textsuperscript{T} = JCM 19251\textsuperscript{T}), was isolated from a soil sample collected from peanut (*Arachis hypogaea*) crop field, Srikakulam, Andhra Pradesh, India.

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