Increases in iPS Transcription Factor (*Oct4*, *Sox2*, *c-Myc*, and *Klf4*)
Gene Expression after Modified Electroconvulsive Therapy

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**Objective**
Electroconvulsive therapy (ECT) is a reasonable option for intractable depression or schizophrenia, but a mechanism of action has not been established. One credible hypothesis is related to neural plasticity. Three genes (*Oct4*, *Sox2*, *c-Myc*) involved in the induction of induced pluripotent stem (iPS) cells are Wnt-target genes, which constitute a key gene group involved in neural plasticity through the TCF family. *Klf4* is the other gene among Yamanaka's four transcription factors, and increases in its expression are induced by stimulation of the canonical Wnt pathway.

**Methods**
We compared the peripheral blood gene expression of the four iPS genes (*Oct4*, *Sox2*, *c-Myc*, and *Klf4*) before and after modified ECT (specifically ECT with general anesthesia) of patients with intractable depression (n=6) or schizophrenia (n=6). Using Thymatron ten times the total bilateral electrical stimulation was evoked.

**Results**
Both assessments of the symptoms demonstrated significant improvement after mECT stimulation. Expression of all four genes was confirmed to increase after initial stimulation. The gene expression levels after treatment were significantly different from the initial gene expression in all twelve cases at the following treatment stages: at the 3rd mECT for *Oct4*; at the 6th and 10th mECT for *Sox2*; and at the 3rd, 6th and 10th mECT for *c-Myc*.

**Conclusion**
These significant differences were not present after correction for multiple testing; however, our data have the potential to explain the molecular mechanisms of mECT from a unique perspective. Further study should be conducted to clarify the pathophysiological involvement of iPS-inducing genes in ECT.

**Key Words**
iPS cells, Electroconvulsive therapy, Schizophrenia, Depression, *Oct4*, *Sox2*.

INTRODUCTION

Electroconvulsive treatment (ECT) is currently regarded to be a reasonable option for intractable psychiatric disorders. If performed correctly, ECT is one of the most effective treatments for drug-resistant depression and catatonic schizophrenia. However, the underlying molecular mechanism of ECT's therapeutic effects has not been fully elucidated, although some important findings have been reported. Briefly, in the context of animal models, electroconvulsive shock (ECS) induces the activation of N-methyl-D-aspartate (NMDA), AMPA-related (DL-α-amino-3-hydroxy-5-methylisoxazole-4-propionic acid) receptors, and hippocampal neurogenesis is subsequently increased.

Long-term potentiation (LTP), with the feature of synaptic plasticity, is thought to form the basis of memory. One well-known hippocampal LTP circuits is NMDAR-dependent, but the mechanism of LTP remains controversial. A recent finding suggested that chronic stress provoked a deficit in LTP in the rat hippocampus, although electroconvulsive stimulation recovered this LTP.

In the adult brain, as the functional activation of the NMDA receptor increases, beta-catenin (cadherin-associated protein, beta 1, 88 kDa, with the HUGO-approved official symbol of CTNNB1) forms a complex with lymphoid enhancer-binding factor-1/T cell factor (LEF1/TCF), subsequently inducing the transcription of genes related to neural plasticity. The genes...
associated with neural plasticity through the TCF-family are called Wnt-target genes.\(^{19}\) Approximately 100 genes are included among the Wnt-target genes\(^{20,22}\) and three genes \((\text{Oct}4 \text{ [POU5F1 POU class 5 homeobox 1 GenID: 5460], Sox2 [SOX2 SRY \text{ (sex determining region Y)-box 2 GenID: 6657], c-Myc [MYC v-myc avian myelocytomatisis viral oncogene homolog GenID: 4609] genes})\) involved in the induction of IPS cells are included among the Wnt-target genes.\(^{23-25}\) Regarding the LIF gene \((\text{leukemia inhibitory factor GenID: 3976})\), which is located upstream of Klf4, stimulation along the canonical Wnt pathway induced an increase in LIF expression.\(^{26}\) Therefore, it has been suspected that electric stimuli, such as ECT, promote the expression of LIF.

The aim of the current study was to reveal the mRNA expression alterations in Yamanaka’s four transcription factors before and after mECT treatment.

**METHODS**

We administered modified electroconvulsive therapy (mECT), specifically electroconvulsive therapy with general anesthesia, at the Osaka Medical College Hospital, to patients diagnosed with either depression or schizophrenia based on the DSM-IV TR\(^{27}\) and with a prescription history of two or more types of antidepressants or antipsychotics at the maximal doses for more than four weeks and with insufficient response to treatment. Six depressed patients and six schizophrenic patients were recruited for further assessment. We included only patients with improved symptoms based on the PANSS\(^{28}\) for schizophrenia or the HAM-D\(^{29}\) for depression after the treatment. This study was approved by the ethics committee of the Osaka Medical College. Written informed consent for biological assessment was provided by all of the participants after a documentary and oral explanation. This work has been conducted according to the principles expressed in the Declaration of Helsinki, and we have not obtained the consent from the surrogates of the participants in the current study. Once the patients were suspected to have a disturbance of consciousness, the patients were omitted from the study. The demographic data for the sample, including the assessed symptoms, are provided in Table 1.

![Table 1. The demographic data of the sample with depression and schizophrenia.](tablet1.png)

| Sample No. | Age | Sex | Type   |
|------------|-----|-----|--------|
| Depression |     |     |        |
| 1          | 76  | Male|        |
| 2          | 43  | Male|        |
| 3          | 75  | Male|        |
| 4          | 51  | Female|        |
| 5          | 28  | Male|        |
| 6          | 45  | Male|        |
| Schizophrenia |     |     |        |
| 7          | 57  | Female| Catatonic |
| 8          | 51  | Female| Catatonic |
| 9          | 63  | Female| Catatonic |
| 10         | 24  | Female| Hebephenic |
| 11         | 64  | Female| Catatonic |
| 12         | 62  | Male|        |

Electroconvulsive therapy was administered under general anesthesia ten times in total, at a frequency of twice per week. Using Thymatron® (Somatics LLC, Lake Bluff, IL, USA), the patients were given bilateral electrical stimulation with a pulse wave. We set the initial quantity of electricity according to the half-age method \((\text{quantity of electricity at half age}^{10})\), and the stimulation dose was increased to 1.5 times if an effective convulsive seizure was not obtained. We used 1.0 mg/kg propofol intravenously and 0.8 mg/kg suxamethonium, which is a muscle relaxant, for general anesthesia. mECT was performed during hospitalization to minimize changes in sleep habits, diet habits, smoking habits, and medication use over the course of the study. Three milliliters of peripheral blood was extracted at approximately 10:00 AM on the day before the 1st ECT \((\text{day 0})\), on the day after the 3rd ECT \((\text{day 9})\), on the day after the 6th ECT \((\text{day 19})\), and on the day after the 10th ECT \((\text{day 33})\). The samples were immediately stored at -80°C (-112°F). RNA was extracted using an mRNA Isolation Kit for Blood® (Roche Diagnostics, Tokyo, Japan), and complementary DNA was subsequently synthesized. For quantitative PCR, the gene expression of \(\text{oct}4, \text{sox}2, \text{c-myc}, \text{klf4}\) and \(\text{glyceraldehyde-3-phosphate dehydrogenase (GAPDH)}\), the last of which was used as a housekeeping gene, were analyzed using a Light Cycler® (Roche Diagnostics, Tokyo, Japan) with specific primers and probes (Table 2).

Statistical comparison was performed using a paired t-test. The repeated measures ANOVA was applied for the overall analysis, although clear statistical significance was not achieved even on the post-hoc analysis. The average value of the expression level of the mRNA of each gene one day before the 1st ECT was regarded as 1, and significance was set at <0.05. All of the statistical analyses were performed with SPSS® software (IBM, Japan).

**RESULTS**

As shown in Figures 1 (HAM-D 21 for depression) and 2 (PANSS for schizophrenia), both severity scales for clinical symptoms indicated an improvement after mECT. The scales showed significant recovery for the patients with depression, particularly for the descriptor Depressed Mood and Feeling of Guilt, while Delusions and Conceptual Disorganization were...
relatively improved for the schizophrenia patients. No obvious adverse events were observed during the course of the study.

Gene expression analysis of all of the samples revealed increases in the four tested genes over the course of the study (Figure 3), some of which were significant, as determined by uncorrected t-test (p<0.05, not corrected, after the 3rd mECT for Oct4, after the 6th and 10th mECT for Sox2, and after the 3rd, 6th and 10th mECT for c-Myc; p>0.05 after Bonferroni correction). Compared to the overall analysis of mECT, when the data for schizophrenia and depression were analyzed individually, only c-Myc gene expression was significantly different in the schizophrenic patients after mECT, as determined by uncorrected t-test (p<0.05, not corrected after the 3rd and 6th mECT; p>0.05 for multiple testing, as in the overall analysis).

**DISCUSSION**

The current results clearly validate the clinical utility of mECT for the treatment of depression and schizophrenia, based on symptomatic assessment using the HAM-D and PANSS. Additionally, the comparison of the mRNA expression of Yamanaka’s four transcription factors in the peripheral blood showed a tendency toward higher levels after treatment. Detailed assessment of the expression of each gene indicated gradual decreases in the expression of some of the genes (e.g., c-Myc and Klf4) over the course of mECT treatment. Klf4 cooperates with Oct3/4 for the maintenance of pluripotency, but not every gene work simultaneously. Therefore, the speed or timing of the expression peak during neurogenesis could be expected to differ for each of these four genes.

iPS cells show triploblastic pluripotency, similar to fertilized eggs or embryonic stem cells. Because the transcription of specific mRNA in mature cells is generally restricted by epigenetic events, such as DNA methylation in the early developing stages, the mRNA expression balance varies for each cell. Although every cell has the same DNA-sequence information in a single individual, different cells are produced because of the different transcriptional patterns in each cell type. Introducing Yamanaka’s four transcription factors into mature cells reprograms the epigenetic restriction that occurs during the early stages of development, allowing iPS cells to acquire their pluripotency. Thus, these cells can again transcribe genes that have been repressed.

It is known that BDNF expression is increased by ECT and that an epigenetic mechanism is involved in this change. Ma et al. found an increase in BDNF in the hippocampal dentate gyrus of adult wild-type mice after ECS, but this change was not observed in Gadd45b knockout mice; Gadd45b encodes a DNA demethylase. Therefore, DNA demethylation is thought
to be involved in the BDNF increase caused by ECT.

DNA methylation of the CpG island in a gene promoter region inhibits the binding of a transcription factor and thus prevents the gene's transcription. Yamanaka’s four transcription factors promote DNA demethylation on the CpG island as a part of the reprogramming. Because the assumed mechanism of ECT is based on both neurogenesis and the increase in BDNF levels, we conclude that the observed ECT-stimulated increase in the expression of Yamanaka’s four transcription factors, some of which are Wnt-target genes needed for neurogenesis, is associated with both neurogenesis and the increase in BDNF.

Our findings should be interpreted cautiously. The first limitation of this study is its small sample size. The diagnoses of schizophrenia and depression in the enrolled sample of patients were based strictly on the SCID, and strict timing of the drawing of blood was applied. Because of these restrictions, only 12 patients were analyzed. Therefore, the small sample size could have led to misleading results. The second limitation arose from the lack of an unmedicated sample group. Our sample consisted only of patients treated with a few medications (it is recommended that drugs be avoided during mECT treatment). However, some drugs were prescribed for the extraction of peripheral blood. These drugs might have affected mRNA expression. Therefore, medication-free samples should be analyzed in future studies. There have not been any analyses of how epigenetic changes in the cerebral cortex of the frontal brain and hippocampus affect mRNA expression in peripheral blood, and this lack of analyses should be considered a third limitation of this study. In depression, methylation of the BDNF gene has been suggested as a biomarker, indicating that Oct4, Sox2, c-Myc, and Klf4 could be useful as biomarkers for ECT treatment.

Although the current findings were from the analysis of mRNA derived from leukocyte, other previous groups have reported the direct analysis of leukocyte fraction changes according to the ECT. Chaturvedi et al. have reported the results, based on the hematological difference between pre-ECT and two hours after ECT stimulation, that the increased lymphocyte and granulocyte subsets, and the increased number of leukocyte soon after the ECT stimulation. In contrast, the decreased lymphocyte subset and the increased granulocyte subsets were detected at two-hours later from ECT stimulation. In addition, Kronfol et al. have reported the increment of lymphocyte natural killer cell activity (NKCA) one hour after the ECT stimulation. They have pointed out the possible mechanism of the trafficking of the accumulated NK cell at the blood vessel wall on the circulating NK cell because of the reason that the number of circulating NK cells is relatively small. The changing of number of leucocyte at the short period as early 1 to 2 hours after ECT stimulation is not mainly from the differential leucocyte but the trafficking of the accumulated NK cell on the blood vessel wall to the circulating NK cell. It is not considered that these early response is cause only by the changing of Yamanaka’s four transcription factor. In addition, the

![Figure 1. Severity scale for depression using HAM-D 21. HAM-D 21: Hamilton depression rating scale 21 items version, ECT: electroconvulsive therapy.](image1)

![Figure 2. Severity scale for schizophrenia using PANSS. PANSS: Positive and Negative Syndrome Scale, ECT: electroconvulsive therapy.](image2)

![Figure 3. Gene expression assay for four genes. HAM-D 21: Hamilton depression rating scale 21 items version, ECT: electroconvulsive therapy.](image3)
gene expression of mRNA was corrected by the house keeping gene's expression quantity, therefore the increment of leukocyte is not simply contributed by the increased expression of Yamanaka's four transcription factor, however the lack of the observation for differential subsets of pre- and post-ECT is one of the considerable limitation of the current finding.

Another limitation should be noted regarding the method of statistical analysis. By adopting the Smirnov-Grubs outlier test, we extracted 2 samples for Oct4 (sample #1 and #2), 1 sample for Sox2 (sample #7), and 2 samples for Klf4 (samples #3 and #5). There was no individual sample that did not duplicate the outlier across the gene. The Kolmogorov-Smirnov test for normality of the distribution did not reveal non-normality of our sample. We adopted the t-test to analyze the significant differences in mRNA expression between one day prior to the initial mECT session and the other days of observation (at the 3rd, 6th and 10th mECT). Significance differences were obtained when comparing the initial mRNA expression levels with the 6th mECT for Oct4 \((t(9)) = -1.95, p=0.04\), the 10th mECT for Sox2 \((t(10)) = -2.14, p=0.03\), and the 3rd, 6th and 10th mECT for c-Myc \((t(11)) = -2.22, p=0.02\) on 3rd, \((t(11)) = -2.16, p=0.03\) on 6th, \(t(11) = -1.87, p=0.04\) on 10th). However, the differences were not significant when a two-tailed t-test was applied. Another concern was the lack of Bonferroni correction in multiple testing. As described in the Results section, all of the significance differences, as indicated by the p-values from the one-tailed t-tests, disappeared on correction (number of tests: 3). The most attractive finding of the current study was that the mRNA expression of all of the selected genes increased after ECT during the observed period.

There is evidence of the correlation of cytokine or BDNF in between blood and brain, although no evidence on Yamanaka's four transcription factor has been reported till date. Y en et al.\(^{25}\) have reported the up-regulation of Sox2, Nanog, Oct4, and Lin28 at the culture of anti-tumor T cell when IL-12 plus IL-7 or IL-21 were added into the culture fluid for three days. Because of ECT, the expression changing of brain cytokine was induced. Due to the correlation of cytokine expression between blood and brain, it is suspected that the cytokine in the peripheral blood up-regulated the Yamanaka's four transcription factor in the peripheral blood. In the future, more research especially for the validation of the expression of cultural cell after ECT stimulation is warranted.

This study found that ECT reversed epigenetic changes that occur during development and resulted in clinical effects. Future progress could reveal how the Oct4, Sox2, c-Myc, and Klf4 genes are involved in the mechanism of ECT.

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