Lipid-Core Plaque Assessed by Near-Infrared Spectroscopy and Procedure Related Microvascular Injury

Hyoung-Mo Yang, MD, PhD, Myeong-Ho Yoon, MD, PhD, Hong-Seok Lim, MD, PhD, Kyoung-Woo Seo, MD, Byoung-Joo Choi, MD, So-Yeon Choi, MD, PhD, Gyo-Seung Hwang, MD, PhD, and Seung-Jea Tahk, MD, PhD

Department of Cardiology, Ajou University School of Medicine, Suwon, Korea

ABSTRACT

Background and Objectives: Microvascular damage due to distal embolization during percutaneous coronary intervention (PCI) is an important cause of periprocedural myocardial infarction. We assessed the lipid-core plaque using near-infrared spectroscopy (NIRS) and microvascular dysfunction invasively with the index of microcirculatory resistance (IMR) and evaluated their relationship.

Methods: This study is pilot retrospective observational study. We analyzed 39 patients who performed NIRS before and after PCI, while fractional flow reserve, thermo-dilution coronary flow reserve (CFR) and IMR were measured after PCI. The maximum value of lipid core burden index (LCBI) for any of the 4-mm segments at the culprit lesion (culprit LCBI_{4mm}) was calculated at the culprit lesion. We divided the patients into 2 groups using a cutoff of culprit LCBI_{4mm} ≥500.

Results: Mean pre-PCI LCBI was 333±196 and mean post-PCI IMR was 20±14 U. Post-PCI IMR was higher (15.6±7.3 vs. 42.6±17.6 U, p<0.001) and post-PCI CFR was lower (3.7±2.2 vs. 2.1±1.0, p=0.029) in the high LCBI group. Pre-PCI LCBI was positively correlated with post-PCI IMR (ρ=0.358, p=0.025) and negatively correlated with post-PCI CFR (ρ=−0.494, p=0.001). The incidence of microvascular dysfunction (IMR ≥25 U) was higher in the high LCBI group (9.4% vs. 85.7%, p<0.001). However, there were no significant differences in the incidences of creatine Kinase-MB (9.4% vs. 14.3%, p=0.563) and troponin-I elevation (12.5% vs. 14.3%, p=1.000).

Conclusions: A large lipid-core plaque at the ‘culprit’ lesion is observed higher incidence of post-PCI microvascular dysfunction after PCI. Prospective study with adequate subject numbers will be needed.

Keywords: Percutaneous coronary intervention; Coronary artery disease; Microvessel; Near-infrared spectroscopy

INTRODUCTION

Periprocedural myocardial infarction (MI) increases the risk of major adverse cardiac events including mortality. An important cause of periprocedural MI is microvascular damage due to distal embolization during percutaneous coronary intervention (PCI). Periprocedural MI
Conflict of Interest
The authors have no financial conflicts of interest.

Author Contributions
Conceptualization: Yang HM, Yoon MH, Choi SY, Tahk SJ; Data curation: Yang HM, Yoon MH, Lim HS, Seo KW, Choi BJ, Choi SY, Hwang GS, Tahk SJ; Formal analysis: Yang HM, Choi BJ; Investigation: Yang HM, Yoon MH, Seo KW; Methodology: Yang HM, Yoon MH, Lim HS, Seo KW, Choi BJ, Choi SY, Hwang GS, Tahk SJ; Supervision: Lim HS, Tahk SJ; Writing - original draft: Yang HM; Writing - review & editing: Yoon MH, Lim HS, Seo KW, Choi BJ, Choi SY, Hwang GS, Tahk SJ.

During PCI has been reported to range from 15–70% of cases, which depends on diagnostic modalities and definitions used. Plaque composition and morphology is closely related to distal embolization during PCI. Attenuated plaque on grayscale intravascular ultrasound (IVUS), thin-cap fibroatheroma (TCFA), necrotic core (NC) on virtual histology (VH)-IVUS and cap thickness on optical coherence tomography (OCT) are related to periprocedural MI.

The lipid core burden index (LCBI) measured using near-infrared spectroscopy (NIRS) can assess plaque vulnerability. Previous studies have demonstrated that pre-PCI LCBI was related with periprocedural MI indicated by cardiac enzyme elevation, and is related to poor clinical outcomes. However, there is no study on the relationship between LCBI measured with NIRS and microvascular dysfunction assessed using invasive coronary physiology. Microvascular dysfunction can be easily measured during PCI using a pressure wire. The index of microcirculatory resistance (IMR) is a pressure-derived index, which can assess microvascular dysfunction and has high reproducibility and reliability, independent of hemodynamic changes.

We aimed to evaluate the relationship between LCBI measured with NIRS and microvascular dysfunction during PCI invasively assessed using IMR.

METHODS

Study population
From February 2015 to July 2016, we enrolled 39 patients who underwent elective PCI with a drug-eluting stent (DES). This study is pilot retrospective study with an exploration study of observational finding. Inclusion criteria were silent ischemia, stable or unstable angina, and age 18–80 years. Exclusion criteria were a lesion in an infarct-related artery, left main disease, a restenosis lesion, graft vessel lesion, chronic total occlusion lesion, and a low ejection fraction (<40%). Cardiac enzyme elevation was defined as creatine kinase-MB (CK-MB) elevation ≥3 times of the upper reference limit, and cardiac troponin-I elevation was defined according to the Society for Cardiovascular Angiography and Interventions (SCAI) definition. The study protocol was approved by the Institutional Review Board (AJIRB-MED-MDB-17-235) and informed consent was obtained from the patients.

Quantitative coronary angiography
The Cardiovascular Angiography Analysis System II (Pie Medical, Maastricht, the Netherlands) was used for quantitative coronary angiography (QCA) analysis. Before and after PCI, the percentage diameter stenosis (DS), minimal luminal diameter (MLD), reference vessel diameter, and lesion length were measured and calculated.

Intravascular ultrasound and near-infrared spectroscopy
IVUS was performed after intracoronary administration of nitroglycerin, using a 3.2-Fr exchange catheter, which is a rotation and pullback device (Infrared, Burlington, Massachusetts). The catheter was advanced distally as far as possible in the target vessel followed by automatic pullback at 0.5 mm/sec. The NIRS system has been previously described. LCBI is calculated as a fraction of yellow pixels measured from the chemogram multiplied by 1,000. The maximum value of LCBI for any of the 4-mm segments at the culprit lesion (culprit LCBI\textsubscript{4mm}) is obtained and represents lipid core plaque. A previous study showed that LCBI\textsubscript{4mm} ≥ 500
before PCI was related to periprocedural MI indicated by cardiac enzyme elevation.\(^{11}\)
Accordingly, we divided the patients into 2 groups using a cutoff value of culprit LCBI\(_{4\text{mm}}\) \(\geq 500\), as high or low LCBI group. Off-line IVUS and NIRS analyses were performed by an independent physician blinded to the IMR value. All IVUS analyses were performed according to the American College of Cardiology clinical expert consensus document on standards for acquisition, measurement, and reporting of IVUS studies.\(^{13}\)

**Coronary physiology measurement**

Coronary pressure measurement was performed pre- and post-PCI using a 0.014-inch pressure wire (PressureWire, Radi Medical System; Abbott, Chicago, IL, USA). FFR was calculated by dividing the mean distal coronary pressure (\(P_d\)) by mean proximal arterial pressure, during maximal hyperemia. After successful DES implantation, IMR and coronary flow reserve (CFR) were measured simultaneously using a thermodilution technique as previously described.\(^{15,16}\) IMR was calculated as the \(P_d\) at maximal hyperemia divided by the inverse of hyperemic mean transit time (Tmn). CFR was calculated as resting Tmn divided by hyperemic Tmn. Microvascular dysfunction defined as a post-PCI IMR \(\geq 25\) U.\(^{17}\) Maximal hyperemia was induced with continuous intravenous adenosine infusion (140 \(\mu\)g/kg/min).

**Statistical analysis**

Categorical variables were presented as percentage and continuous variables were presented as mean±standard deviations. To test normal distribution, we performed Kolmogorov-Smirnov test. Continuous variables were compared using the unpaired \(t\)-test or Mann-Whitney \(U\) test and categorical variables were compared using the \(\chi^2\) test or Fisher exact test. The correlation between LCBI and physiologic parameters was assessed using Spearman correlation. All statistical analyses were performed using SPSS software (version 20.0; SPSS Inc., Chicago, IL, USA). A \(p\) value of <0.05 was considered statistically significant.

**RESULTS**

Baseline clinical characteristics are shown in Table 1. We enrolled 39 patients, and their mean age was 60±10 years. Patients with stable angina were 12 (31%), and those with unstable angina were 20 (51%). Diabetes was present in 9 (23%) patients. The most common target vessel was the left anterior descending artery (95%).

Baseline QCA and IVUS findings are summarized in Table 2. There were no differences in MLD and DS before and after PCI between the 2 groups. IVUS results showed that the low LCBI group had a larger proximal reference lumen area (12.5±4.6 vs. 9.7±1.7 \(\text{mm}^2\), \(p=0.012\)), and had a tendency for larger post-PCI minimal stent area (7.1±2.1 vs. 5.4±1.7 \(\text{mm}^2\), \(p=0.056\)). However, there was no difference in the IVUS area stenosis after PCI between 2 groups (32±13\% vs. 37±17\%, \(p=0.376\)). Mean value of culprit LCBI\(_{4\text{mm}}\) was 333±196 before PCI and 119±137 after PCI. The delta LCBI defined as the difference between culprit LCBI\(_{4\text{mm}}\) before and after PCI was higher in the high LCBI group (174±125 vs. 389±201, \(p=0.001\)).

Coronary physiology study results are shown in Table 3. Pre-PCI FFR and post-PCI FFR (0.87±0.05 vs. 0.86±0.04, \(p=0.640\)) were not different between the 2 groups. The mean value of post-PCI IMR was 20±14 U. Post-PCI IMR was higher (15.6±7.3 vs. 42.6±17.6 U, \(p<0.001\)), and post-PCI CFR was lower in the high LCBI group (3.7±2.2 vs. 2.1±1.0, \(p=0.029\)) (Fig. 1). High LCBI group had longer mean hyperemic transit time (0.20±0.10 vs. 0.57±0.25
but the Pd was not different between the 2 groups (79±14 vs. 79±18 mmHg, p=0.928) (Table 3). Pre-PCI culprit LCBI had a negative correlation with post-PCI CFR (ρ=−0.494, p=0.001) and a positive correlation with post-PCI IMR (ρ=0.358, p=0.025) (Fig. 2).

The incidence of microvascular dysfunction was more common in the high LCBI group (9.4% vs. 85.7%, p<0.001). However, there were no significant differences in the incidence of CK-MB elevation (9.4% vs. 14.3%, p=0.563) and troponin-I elevation (12.5% vs. 14.3%, p=1.000) based on the SCAI procedural MI definition.
Table 3. Coronary physiology study results

| Variables                  | Low LCBI (n=32) | High LCBI (n=7) | p value |
|----------------------------|-----------------|-----------------|---------|
| **Pre-intervention**       |                 |                 |         |
| Pd/Pa, rest               | 0.91±0.06       | 0.93±0.01       | 0.548   |
| FFR                       | 0.73±0.09       | 0.74±0.08       | 0.910   |
| **Post-intervention**      |                 |                 |         |
| Pd/Pa, rest               | 0.95±0.02       | 0.94±0.02       | 0.331   |
| FFR                       | 0.87±0.05       | 0.86±0.04       | 0.640   |
| CFR                       | 3.7±2.2         | 2.1±1.0         | 0.029   |
| Pd, hyperemia (mmHg)      | 79±14           | 79±18           | 0.928   |
| Tmn, rest (sec)           | 0.67±0.37       | 1.14±0.70       | 0.076   |
| Tmn, hyperemia (sec)      | 0.20±0.10       | 0.57±0.25       | 0.001   |
| IMR (U)                   | 15.6±7.3        | 42.6±17.6       | <0.001  |

Values are mean±standard deviation.

CFR = coronary flow reserve; IMR = index of microcirculatory resistance; FFR = fractional flow reserve; LCBI = culprit lipid-core burden index; Pa = proximal arterial pressure; Pd = distal coronary pressure; Tmn = mean transit time.

Figure 1. Comparison of physiologic parameters. The mean value of post-PCI IMR was higher (15.6±7.3 vs. 42.6±17.6 U, p<0.001) and post-PCI CFR was lower in high LCBI group (3.7±2.2 vs. 2.1±1.0, p=0.029).

CFR = coronary flow reserve; IMR = index of microcirculatory resistance; LCBI = culprit lipid-core burden index; PCI = percutaneous coronary intervention.

Figure 2. Correlation between pre-intervention LCBI and physiologic parameters.

CFR = coronary flow reserve; IMR = index of microcirculatory resistance; LCBI = culprit lipid-core burden index; PCI = percutaneous coronary intervention.
DISCUSSION

This study showed that a high lipid-rich plaque as assessed using NIRS is related to post-PCI microvascular dysfunction, and has a fair correlation with post-PCI IMR and CFR, as assessed using a pressure wire.

Periprocedural MI increases the risk of major adverse cardiac events. Distal embolization of the plaque and/or thrombus is an important cause for the development of periprocedural MI, and it leads to impairment of microvascular perfusion at the tissue level. Since plaque dissection or redistribution occurs during PCI, the vulnerable plaque is more easily redistributed and embolized to the distal vessel or to a side branch. Hence, plaque composition and morphology are closely related to distal embolization of plaque debris during PCI.

Previous studies evaluated the relationship between plaque characteristics and periprocedural MI using different invasive imaging modalities. In grayscale IVUS, an attenuated plaque was related to deterioration in the coronary blood flow after PCI. Using VH-IVUS, TCFA and high NC, increased the risk of periprocedural MI. The OCT study showed that the presence of OCT-defined TCFA could predict periprocedural MI.

NIRS is a novel imaging technique that illuminates the tissue with near-infrared light. Since the absorbance spectrum reflects the chemical makeup of the imaged tissue, it is a very effective method to detect a lipid-rich plaque. The LCBI measured with NIRS can assess plaque vulnerability, and a recent study has demonstrated that higher LCBI increases the risk of major adverse cardiac events. Every 100 units increase in culprit LCBI was associated with a 19% increase in death, non-fatal acute coronary syndrome, or unplanned revascularization during 4 years of follow-up. Goldstein et al. reported that periprocedural MI identified by cardiac enzyme elevation is more frequent in a patient with a large lipid plaque using a cutoff value of culprit LCBI ≥500 in NIRS. We divided the patients using this cutoff value as a reference. In addition, the receiver operating characteristic curve analysis using our study population showed a similar result. The best cutoff value for culprit LCBI to predict microvascular dysfunction (IMR >25 U) was >488 with a sensitivity of 67%, specificity of 97%, and accuracy of 87%. Using a cutoff value of culprit LCBI ≥500, the incidence of microvascular dysfunction after PCI is more frequent in high LCBI group. Besides the absolute value of culprit LCBI before PCI, the delta LCBI was higher in high LCBI group, which might reflect more plaque redistribution and embolization during PCI in this group.

The strength of our study is the use of IMR to assess microvascular dysfunction. Usually, periprocedural MI is evaluated using cardiac enzyme elevation after PCI. However, the reported incidence has a wide range, several different definitions, and measured cardiac enzyme at different time points. Moreover, cardiac enzyme elevation cannot specifically identify the origin, as to whether it is related with a target lesion or other vessels or a patient level. In contrast, the IMR is a reliable method for assessing coronary microvasculature at the target vessel territory. It has a significantly good correlation with true microvascular resistance, good reproducibility and lesser dependence on hemodynamic parameters including heart rate, contractility, and blood pressure.

A previous study reported the usefulness of IMR in evaluating periprocedural MI. Cuisset et al. reported that a patient treated with direct stenting had significantly lower IMR.
compared to conventional stenting. A VH-IVUS study showed that the IMR significantly increased after PCI in patients with TCFA compared with non-TCFA. In the current study as well, besides the incidence of microvascular dysfunction, the mean value of IMR was higher in high LCBI group. The difference in IMR between the high and low LCBI group results from the difference in hyperemic Tmn. There was no difference in coronary distal pressure between the 2 groups; however, hyperemic Tmn was significantly longer in the high LCBI group indicating increased microvascular resistance in this group. Moreover, the mean value of post-PCI CFR, which is another parameter reflecting microvascular dysfunction, was lower and the incidence of CFR ≤ 2 was numerically higher (57% vs. 22%, p=0.08) in the high LCBI group. Although CFR has limitations in that it is not specific for microvascular function and has a high variability with hemodynamic changes compared to IMR, our CFR result indicates that a high lipid-rich plaque was related to post-PCI microvascular dysfunction.

Although our study showed no differences in the incidence of cardiac enzyme elevation including CK-MB and cardiac troponin-1 between 2 groups, the IMR was a more accurate and sensitive for evaluating target vessel microvascular dysfunction. Several reasons can explain these discrepancies. First, IMR can increase without troponin elevation, and indicate a microvascular injury after PCI even in the absence of detectable myonecrosis. Second, since the time of cardiac enzyme measurement after PCI was different in each study, the peak value of cardiac enzyme could not reflect true myocardial damage always. Finally, our sample size was too small to be powerful enough to detect the difference. Accordingly, IMR might be a more sensitive method to detect microvascular dysfunction compared to cardiac enzyme after PCI.

Despite vulnerable plaques assessed using various imaging modalities being correlated to periprocedural MI, there was limited and debatable data for the prevention of periprocedural MI. Brilakis et al. have reported that embolic material was retrieved in 89% of the cases with lipid-rich plaques assessed with NIRS using filter devices; however, the Coronary Assessment by Near-infrared of Atherosclerotic Rupture-prone Yellow trial failed to demonstrate the efficacy of distal protection to prevent periprocedural MI in patients with lipid-rich plaques. On the other hands, another study demonstrated that pre-PCI high dose statin therapy reduced post-PCI microvascular dysfunction as assessed using IMR. Moreover, early statin therapy within 48 hours after admission in statin-naïve patients with acute MI reduced long-term clinical outcomes compared with statin initiation later. The use of high dose statin before PCI and selected use of distal protection device are helpful to prevent periprocedural MI.

There were several limitations in this study. First, the major limitation is too small number of patients, patients of high LCBI group were only 7. This is a pilot observational retrospective study, and it is a limitation to draw meaningful statistical difference. To obtain statistical significance, statistically designed prospective study is necessary. Second, we did not measure CFR and IMR before PCI. However, in cases with significant epicardial stenosis at the target vessel, the IMR cannot reflect microvascular function accurately because of the effect of epicardial resistance. Third we did not conduct IVUS volumetric analysis. Volumetric analysis is helpful to understand the mechanism of plaque redistribution and embolization.

In conclusion, a large lipid-core plaque at the ‘culprit’ lesion is observed higher incidence of post-PCI microvascular dysfunction after PCI. Prospective study with adequate subject numbers will be needed.
REFERENCES

1. Henriques JP, Zijlstra F, Ottervanger JP, et al. Incidence and clinical significance of distal embolization during primary angioplasty for acute myocardial infarction. Eur Heart J 2002;23:1112-7.
PUBMED | CROSSREF

2. Prasad A, Singh M, Lerman A, Lennon RJ, Holmes DR Jr, Rihal CS. Isolated elevation in troponin T after percutaneous coronary intervention is associated with higher long-term mortality. J Am Coll Cardiol 2006;48:1765-70.
PUBMED | CROSSREF

3. Claessen BE, Maehara A, Fahey M, Xu K, Stone GW, Mintz GS. Plaque composition by intravascular ultrasound and distal embolization after percutaneous coronary intervention. JACC Cardiovasc Imaging 2012;5:S111-8.
PUBMED | CROSSREF

4. Idris H, Lo S, Shugman IM, et al. Varying definitions for periprocedural myocardial infarction alter event rates and prognostic implications. J Am Heart Assoc 2014;3:e001086.
PUBMED | CROSSREF

5. Prati F, Pawlowski T, Gil R, et al. Stenting of culprit lesions in unstable angina leads to a marked reduction in plaque burden: a major role of plaque embolization? A serial intravascular ultrasound study. Circulation 2003;107:2320-5.
PUBMED | CROSSREF

6. Herrmann J. Peri-procedural myocardial injury: 2005 update. Eur Heart J 2005;26:2493-519.
PUBMED | CROSSREF

7. Selvanayagam JB, Porto I, Channon K, et al. Troponin elevation after percutaneous coronary intervention directly represents the extent of irreversible myocardial injury: insights from cardiovascular magnetic resonance imaging. Circulation 2005;111:1027-32.
PUBMED | CROSSREF

8. Lee SY, Mintz GS, Kim SY, et al. Attenuated plaque detected by intravascular ultrasound: clinical, angiographic, and morphologic features and post-percutaneous coronary intervention complications in patients with acute coronary syndromes. JACC Cardiovasc Inter 2009;2:65-72.
PUBMED | CROSSREF

9. Yamada R, Okura H, Kume T, et al. Target lesion thin-cap fibroatheroma defined by virtual histology intravascular ultrasound affects microvascular injury during percutaneous coronary intervention in patients with angina pectoris. Circ J 2010;74:1658-62.
PUBMED | CROSSREF

10. Lee T, Yonetsu T, Koura K, et al. Impact of coronary plaque morphology assessed by optical coherence tomography on cardiac troponin elevation in patients with elective stent implantation. Circ Cardiovasc Inter 2011;4:378-86.
PUBMED | CROSSREF

11. Goldstein JA, Maini B, Dixon SR, et al. Detection of lipid-core plaques by intracoronary near-infrared spectroscopy identifies high risk of periprocedural myocardial infarction. Circ Cardiovasc Inter 2011;4:429-37.
PUBMED | CROSSREF

12. Schuurman AS, Vroegindewey M, Kardys I, et al. Near-infrared spectroscopy-derived lipid core burden index predicts adverse cardiovascular outcome in patients with coronary artery disease during long-term follow-up. Eur Heart J 2018;39:295-302.
PUBMED | CROSSREF

13. Fearon WF, Balsam LB, Farouque HM, et al. Novel index for invasively assessing the coronary microcirculation. Circulation 2003;107:3129-32.
PUBMED | CROSSREF

14. Moussa ID, Klein LW, Shah B, et al. Consideration of a new definition of clinically relevant myocardial infarction after coronary revascularization: an expert consensus document from the Society for Cardiovascular Angiography and Interventions (SCAI). J Am Coll Cardiol 2013;62:1563-70.
PUBMED | CROSSREF

15. Mintz GS, Nissen SE, Anderson WD, et al. American College of Cardiology clinical expert consensus document on standards for acquisition, measurement and reporting of intravascular ultrasound studies (IVUS). A report of the American College of Cardiology task force on clinical expert consensus documents. J Am Coll Cardiol 2001;37:1478-92.
PUBMED | CROSSREF

16. Fearon WF, Farouque HM, Balsam LB, et al. Comparison of coronary thermodilution and Doppler velocity for assessing coronary flow reserve. Circulation 2003;108:2198-200.
PUBMED | CROSSREF
17. Kobayashi Y, Fearon WF. Invasive coronary microcirculation assessment—current status of index of microcirculatory resistance. Circ J 2014;78:1021-8.

18. Ahmed JM, Mintz GS, Weissman NJ, et al. Mechanism of lumen enlargement during intracoronary stent implantation: an intravascular ultrasound study. Circulation 2000;102:7-10.

19. Hong YI, Jeong MH, Choi YH, et al. Impact of plaque components on no-reflow phenomenon after stent deployment in patients with acute coronary syndrome: a virtual histology-intravascular ultrasound analysis. Eur Heart J 2011;32:2059-66.

20. Jaross W, Neumeister V, Lattke P, Schuh D. Determination of cholesterol in atherosclerotic plaques using near infrared diffuse reflection spectroscopy. Atherosclerosis 1999;147:327-37.

21. Neumeister V, Scheibe M, Lattke P, Jaross W. Determination of the cholesterol-collagen ratio of arterial atherosclerotic plaques using near infrared spectroscopy as a possible measure of plaque stability. Atherosclerosis 2002;165:251-7.

22. Thygesen K, Alpert JS, Jaffe AS, et al. Third universal definition of myocardial infarction. Circulation 2012;126:2020-35.

23. Ng MK, Yeung AC, Fearon WF. Invasive assessment of the coronary microcirculation: superior reproducibility and less hemodynamic dependence of index of microcirculatory resistance compared with coronary flow reserve. Circulation 2006;113:2054-61.

24. Cuisset T, Hamilos M, Melikian N, et al. Direct stenting for stable angina pectoris is associated with reduced periprocedural microcirculatory injury compared with stenting after pre-dilation. J Am Coll Cardiol 2008;51:1060-5.

25. Brilakis ES, Abdel-Karim AR, Papayannis AC, et al. Embolic protection device utilization during stenting of native coronary artery lesions with large lipid core plaques as detected by near-infrared spectroscopy. Catheter Cardiovasc Interv 2012;80:1157-62.

26. Stone GW, Maehara A, Muller JE, et al. Plaque characterization to inform the prediction and prevention of periprocedural myocardial infarction during percutaneous coronary intervention: the CANARY trial (Coronary Assessment by Near-infrared of Atherosclerotic Rupture-prone Yellow). JACC Cardiovasc Interv 2015;8:927-36.

27. Lee BK, Koo BK, Nam CW, et al. Does pre-treatment with high dose atorvastatin prevent microvascular dysfunction after percutaneous coronary intervention in patients with acute coronary syndrome? Korean Circ J 2016;46:472-80.

28. Kim MC, Ahn Y, Cho JY, et al. Benefit of early statin initiation within 48 hours after admission in statin-naive patients with acute myocardial infarction undergoing percutaneous coronary intervention. Korean Circ J 2019;49:419-33.