Prediction of the Risk for Essential Hypertension among Carriers of C825T Genetic Polymorphism of G Protein β3 (GNB3) Gene

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ABSTRACT
BACKGROUND: The guanine nucleotide-binding protein beta polypeptide 3 (GNB3) 825T allele encodes a product that enhances the activation of heterotrimeric G proteins, which is associated with the occurrence of the splice variant Gβ3 that could play a role in vascular reactivity and hyperpolarization of smooth muscle cells, that makes such proteins attractive candidate gene products for susceptibility to essential hypertension (EH).

OBJECTIVE: To predict the risk for EH in individuals with C825T genetic polymorphism of G protein β3 gene.

METHODS: The study consisted of 222 normotensive individuals and 216 hypertensive patients. Individuals were genotyped for C825T genetic polymorphism of G protein β3 gene rs5443 by using restriction fragment length polymorphism.

RESULTS: Frequencies of C and T alleles were 58.1% and 41.9%, respectively, in the control group compared with 47.7% and 52.3%, respectively, in the hypertensive group. The carriers of rs5443 (T) allele exhibited a significant greater risk for EH compared with the carriers of rs5443 (C) allele (odds ratio = 1.5, 95% confidence interval = 1.2–2.0).

CONCLUSION: T allele is a risk factor for EH in the Egyptian population, which may be used as a diagnostic and a therapeutic target of prophylaxis.

KEYWORDS: G protein, polymorphism, essential hypertension, RFLP, PCR, BseDI

INTRODUCTION
Both genetic and environmental factors contribute to the pathogenesis of essential hypertension (EH). EH is about twice as common in subjects who have one or two hypertensive parents, and many epidemiological studies suggest that genetic factors account for approximately 30% of the variation in blood pressure in various populations. Northwestern American studies have reported that premature onset of hypertension among first-degree relatives yielded a remarkable high risk of 3.8 times to develop hypertension.

Guanine nucleotide-binding proteins (G proteins) are signal transducers that communicate signals from many hormones, neurotransmitters, chemokines, and autocrine and paracrine factors. Ligand binding to heptahelical receptors results in the dissociation of the heterotrimeric G protein into Gα-GTP and Gβγ complexes, resulting in various cellular functions. Because of their crucial role in the function of many types of cells, genetic abnormalities in G protein subunits have the potential to be involved in the etiology of a wide range of clinical conditions. C825T genetic polymorphism is associated with obesity, depression, and cardiovascular diseases.

The entire nucleotide-binding protein beta polypeptide 3 (GNB3) gene is located in chromosome 12p13 spans 7.5 kb and is composed of 11 exons and 10 introns (Fig. 1). Its promoter lacks TATA box but harbors GC-rich regions. All G protein β subunits identified so far consist of seven WD repeats (referring to the conserved amino acids tryptophan “W” and aspartate “D”) that form a regular torus-like structure (Fig. 1). WD proteins are found in all eukaryotes but not in prokaryotes and regulate cellular division, cell-fate determination, gene transcription, transmembrane signaling, mRNA modification, and vesicle fusion.

In GNB3, polymorphism C → T (rs5443) at nucleotide number 825 in exon 10 of the β3 subunit of pertussis toxin-sensitive Gi type protein has been identified. This polymorphism induces the occurrence of a splice variant in which the nucleotides 498–620 of exon 9 are deleted. This deletion causes the loss of 41 amino acids of exon 9 along with the fourth Trp–Asp (WD) of the seven WD repeats that form a propeller structure. Such a structure change could alter the function of critical β-propeller residue that contacts the lip of the α-propeller subunit, thereby inducing a GDP exit route, thereby accounting for the
enhanced activity of GNB3. So, this splice variant is found to be a functional protein and causes enhanced activation of G protein in reconstituted systems. Therefore, a higher blood pressure would arise from increased sensitivity to vasoactive pressor hormones known to transmit their signals through Gβ3 proteins. In vivo studies confirmed this concept and demonstrated an enhanced vascular reactivity on the stimulation of coronary α1- adrenergic receptors in carriers of GNB3 825T. Likewise, neutrophils from carriers of rs5443T allele exhibit an increased chemotactic response.

Several studies have evaluated the association between GNB3 C825T polymorphism and EH. There is a lot of controversy in GNB3 C825T polymorphism in hypertension cases. For novel analysis of the GNB3 polymorphisms, samples from Egyptian subjects are analysed, whom the studies upon them are very limited. Some ethnic groups with this polymorphism have a very strong association with hypertension, whereas some ethnic groups do not. GNB3 polymorphism (C825T) was not associated with EH among Asian population as reported by Guo et al and Li et al. Another meta-analytic study claims a significant association with GNB3 polymorphism (C825T) in Caucasians, with no detected associations between GNB3 C825T and the risk for overall EH in Asian and Japanese people.

We aimed to study the risk for EH in the Egyptian population with C825T genetic polymorphism of GNB3 gene rs5443 to clarify the discrepancies between the ethnic groups that may have a therapeutic impact on these patients, on whom sufficient epidemiological studies on this subject are not available.

Materials and Methods

Subjects. The control group (group I) consisted of 222 normotensive patients (108 males and 114 females) with mean age (38.5 ± 12.4 years). The case group (group II) consisted of 216 hypertensive patients (106 males and 110 females) with mean age (38.6 ± 9.9 years). Total 438 patients’ blood samples were recruited from Ain Shams University Hospitals, from October 2011 to March 2015. All subjects gave their written, informed consent to be included in this study. The study was conducted in the Internal Medicine Department of Ain Shams University Hospitals, and approved by the Research Ethics Committee of Ain Shams University. This research was conducted in accordance with the principles of the Declaration of Helsinki.

Subjects with body mass index (BMI) ≥25 kg/m² were considered positive for obesity, as defined by the World Health Organization (WHO). Subjects were chosen to be matched in age, gender, and BMI between the case and control groups (Table 1).

Table 1. Demographic data of the studied group.

| PARAMETER | CONTROL (NO. = 222) | CASES (NO. = 216) | P |
|-----------|---------------------|-------------------|---|
| Mean age ± SD | 38.5 ± 12.4 | 38.6 ± 9.9 | 0.93 |
| Mean BMI ± SD | 26.3 ± 4.1 | 26.7 ± 4.2 | 0.28 |
| Gender | | | |
| Male | 48.6% (108) | 49.1% (106) | χ² (P) |
| Female | 51.4% (114) | 50.9% (110) | 0.008 (0.93) |
The subjects were considered hypertensive if they were currently using antihypertensive drugs or had systolic blood pressure $>140$ mm Hg or diastolic blood pressure $>95$ mm Hg.

The clinical information obtained was medical history, current medication use, lifestyle, and anthropometric and blood pressure measurements. The physicians performed a physical examination, and blood samples were collected for laboratory procedures.

**DNA extraction.** Blood samples that were collected on blood samples collection cards then DNA was extracted from the dried blood spots from the white blood cells by a salting out method, using the QIAamp® DNA Blood Mini Kits (Qiagen). Red blood cell lysis was done by using red cell lysis buffer (20 mM Tris-HCl pH 7.6), followed by centrifugation. Nuclei lysis was carried out by cell lysis buffer (10 mM Tris-HCl pH 8.0, 1 mM Ethylene-di-amine-tetra-acetic acid (EDTA) pH 8.0, and 0.1% (w/v) SDS) and 500 µL proteinase K (20 mg/mL), which was kept in a water bath at 56 °C for one hour, and then, protein was precipitated by protein precipitation solution (60 mL of 5M potassium acetate, 11.5 mL of glacial acetic acid, 11.5 mL of glacial acetic acid, and 28.5 mL of water) followed by centrifugation at low speed ($2000 \times g$) for 10 minutes. Then, the supernatant was removed and placed into a clean Eppendorf tube. Finally, DNA was precipitated by 650 µL isopropanol and inverted to mix. The Eppendorf tube was incubated at room temperature for 15 minutes, followed by centrifugation at full speed ($17,000 \times g$) to recover DNA. Then, the DNA was washed twice with 70% ethanol by centrifugation at $4000 \times g$ after each wash. The DNA pellet was dried in air, rehydrated in TE buffer (pH 7.6), and stored at −20 °C. The DNA concentration and purity were determined by spectrophotometer measurement of absorbances at 260 and 280 nm, respectively.

**Polymorphism detection.** The gene was amplified by polymerase chain reaction (PCR) on 96-well Amp PCR System 9700 Thermocycler (Applied Biosystems). Primer sequences, PCR conditions, and restriction enzyme digestion were as follows (oligonucleotides were synthesized by Promega). The forward primer was 5’-TGACCCACTTGCACCCGTGC-3’, and the reverse primer was 5’-GCAGCAGCCACGGCTG-3’ (primers accession number: >NC_000012.12). PCR was carried out in a 50-µL reaction volume containing 100 ng of genomic DNA, 0.4 mmol/L of each primer, 0.2 mmol/L dNTPs, 2 mmol/L MgCl$_2$ in 10% PCR buffer, and 1U of DNA polymerase (Promega, UK). PCR involved a first denaturation step of 95 °C for 5 minutes, which was followed by 35 cycles at 94 °C for 1 minute, at 60 °C for 45 seconds, and at 72 °C for 1 minute. The reaction was completed by a final extension step at 72 °C for seven minutes. Aliquots of 5 µL of the PCR products were digested with BseDI (MBI Fermentas). Restriction fragment length polymorphism (RFLP) products were analyzed on 2.5% agarose gel, stained with ethidium bromide, and visualized with ultraviolet transillumination. A 25 base pairs (bp) DNA marker was used, it ranged from 25 to 300 bp (Promega). The CC genotype gave two bands of 115 bp, and 152 bp, CT was expected to give three bands of 267, 152, and 115 bp, while TT gave one band of 267 bp. Figure 2.

**Statistical analysis.** Genotype and allele frequency were calculated by allele counting as described by Emery. Genotype distribution was investigated in relation to Hardy–Weinberg equilibrium that was assessed using a $\chi^2$ goodness-of-fit test for biallelic markers (available at http://ihg.gsf.de/cgi-bin/hw/hwa1.pl). Three allele groups were considered for G protein C825T polymorphism rs5443 (CC, CT, and TT). Differences in genotype prevalence and association between the case and control groups were assessed by the chi-square test.
Table 2. Frequency of different genotypes of C825T genetic polymorphism among cases and controls (http://bioinfo.iconcologia.net/SNPstats-web).

| GENOTYPE | CONTROL (N = 222) | CASES (N = 216) | χ² (P) |
|----------|------------------|----------------|--------|
| CC       | 70 (31.5%)       | 54 (25%)       | 2.3 (0.13) |
| CT       | 118 (53.2%)      | 98 (45.4%)     | 2.65 (0.01*) |
| TT       | 34 (15.3%)       | 64 (29.6%)     | 12.9 (0.00*) |

Note: *P is highly significant (≤0.01).

Table 3. Frequency and odds ratio26 of different genotypes of C825T genetic polymorphism among cases and controls: dominant model.

| GENOTYPE | CONTROL (N = 222) | CASES (N = 216) | OR 95% CI χ² (P) |
|----------|------------------|----------------|------------------|
| CC       | 70 (31.5%)       | 54 (25.0%)     | 1 #0.9–2.1 2.3 (0.13) |
| CT and TT| 152 (68.5%)      | 162 (75.0%)    | 1.4              |

Notes: *Range includes 1 that is not a significant risk factor.
Abbreviations: OR, odds ratio; 95% CI, 95% confidence interval.

Table 4. Frequency and odds ratio26 of different genotypes of C825T genetic polymorphism among cases and control: recessive model.

| GENOTYPE | CONTROL (N = 222) | CASES (N = 216) | OR 95% CI χ² (P) |
|----------|------------------|----------------|------------------|
| CC and CT| 188 (84.7%)      | 152 (70.4%)    | 1 *1.5–3.7 12.9 (0.0**)|
| TT       | 34 (15.3%)       | 64 (29.6%)     | 2.3              |

Notes: *Range above 1 is a significant risk factor. **P is highly significant (≤0.01).
Abbreviations: OR, odds ratio; 95% CI, 95% confidence interval.

Discussion

In the present study, we investigated the association between a GNB3 825T polymorphism and EH in the Egyptian population. Our study demonstrated a frequency of 47% for T allele among the Egyptian population, which was similar to that reported for Asians (45%).26,27 This frequency was higher than that reported for whites elsewhere (28.1%, 95% CI = 26–30.2).14,15,18,20 while it was lower than that reported for black people (79.2%, 95% CI = 76.5–81.9).13,30

This C825T polymorphism is located >1700 bp upstream of the alternative splice site, indicating that the effect of GNB3 825T on splice process is a complex mechanism. However, there are examples that single distant nucleotide exchange, not related to conserved splice branch, donor, and acceptor sites, can cause such alternative splicing.31

Results of examining the association between G protein beta3 subunit C825T genetic polymorphism rs5443 and blood pressure regulation supported the observation that the G protein beta3 subunit gene variant affects renal function. The pathogenesis of the C825T polymorphism relies on the fact that the 825T allele of GNB3 is related to enhanced stimulated G protein activation in cell lines from hypertensive patients.32,33

Polymorphism in several genes has been associated with blood pressure levels.34 One of these genes is G protein. In the present study, carrier of rs5443 (T) allele either in heterozygote or homozygote form (dominant model) were at risk for hypertension (OR = 1.4, 95% CI = 0.9–2.1), the risk was much higher and significant in the recessive model (OR = 2.3, 95% CI = 1.5–3.7), when the study groups were divided into two groups based on the allele distribution, carriers of T allele were at risk of 1.5 for hypertension compared with the carriers of rs5443 (C) allele (OR = 1.5, 95% CI = 1.2–2.0).

Table 5. Frequency and odds ratio26 of different alleles of C825T genetic polymorphism among cases and control.

| ALLELE | CONTROL (N = 222) | CASES (N = 216) | OR 95% CI χ² (P) |
|--------|------------------|----------------|------------------|
| C      | 258 (58.1%)      | 206 (47.7%)    | 1 *1.2–2.0 12.183 (0.0**)|
| T      | 186 (41.9%)      | 226 (52.3%)    | 1.5              |

Notes: *Range above 1 is a significant risk factor. **P is highly significant (≤0.01).
Abbreviations: OR, odds ratio; 95% CI, 95% confidence interval.
Siffert et al had shown the significant association of T allele with EH in Germans.15 After this report, many studies have investigated the association between the C825T polymorphism and hypertension,14,35,36 particularly in a study with white population, a positive association had been shown (OR = 1.44, 95% CI = 1.1–2.41 and OR = 2.3, 95% CI = 1.7–3.3).26 In a study done in the Turkish population, T allele had 2.7 times greater risk for hypertension (OR = 2.786, 95% CI = 1.114–6.967, P = 0.028). This association was still significant after adjustment for BMI (OR = 1.78, 95% CI = 1.546–2.074, P = 0.0001).37

Similar results were reported among black population. A threefold higher risk for hypertension was found among the carriers of the T variant in both heterozygotes (OR = 3.43, 95% CI = 0.94–12.4) and homozygotes (OR = 3.87, 95% CI = 1.09–13.8). Association was detected also after adjustment for age, sex, and BMI (OR = 4.41, 95% CI = 1.11–15.4).13

Many studies reported significant associations between the GNB3 C825T polymorphism rs5443 and hypertension, as well as hemodynamic phenotypes such as renal perfusion, LVH, LV diastolic filling, and coronary vasoconstriction and cardiovascular ischemic diseases.26,38–40

In the Hypertension and Ambulatory Recording Venetia Study (HARVEST Study), patients carrying the 825T allele had an increased risk for reaching the blood pressure end point (need for antihypertensive therapy) during a mean follow-up of 4.7 years.41 Investigation in an ethnic group from United Arab Emirates demonstrated a strong association of GNB3 825T allele with LVH, but not with EH, and a high prevalence of the TT genotype (27%) and T allele (55%) in patients with EH.42

Furthermore, T allele was associated with obesity, which may affect blood pressure variations.30,41 The association with obesity has been demonstrated in both white and non-white populations.27,30,41 An effect of G protein subunit on adipogenesis has been observed.13,28 Moreover, since increased BMI is associated with increased risk for hypertension, the possibility has been raised that the association of the GNB3 allele with hypertension could be from a primary association with obesity.14 The mechanism to explain T allele and obesity that increased signaling pertussis toxin sensitive G proteins, which have been shown to stimulate adipogenesis.28 Increased Gi activity could also attenuate Gs-mediated lipolysis, leading to impaired adrennergic-mediated lipolysis and obesity.14

Interaction between the 825T polymorphism, obesity status, and physical activity in predicting hypertension in African-Americans was studied by Grove et al.44 They found that homozygotes for the 825T allele who were obese and had a low activity level were 2.7 times more likely to be hypertensive, compared with nonobese, active 825T homozygotes (OR = 2.7, 95% CI = 1.19–6.17, P = 0.02). During physical activity, catecholamines bind to beta-adrenergic receptors and catalyze the mobilization of glucose and fatty acids that are needed to fuel muscle movement.45 The GNB3 C825T polymorphism creates an altered protein product that may negatively affect interaction with beta-adrenergic receptors during physical activity possibly leading to a decrease in response to catecholamines,46 and consequently, to altered energy utilization and adipose homeostasis.47

In meta-analysis studies done in 2013 and 2014, they screened 66 studies regarding hypertension and found that only the allele model showed marginal association with hypertension (OR = 1.07, 95% CI = 1.01–1.13).15,20 Li et al, found a high statistical significance (P = 0.011) for the association between hypertension and TT genotype and T allele in Chinese population, which is in concordance with the Egyptian population in our study.7

Table 6. Frequency of different genotypes of C825T genetic polymorphism rs5443 among different gender, age, and BMI.

| GENOTYPE | CC | CT | TT |
|----------|----|----|----|
| Gender  |    |    |    |
| Male     | 64 (51.6%) | 92 (42.6%) | 58 (59.2%) |
| Female   | 60 (48.4%) | 124 (57.4%) | 40 (40.8%) |
| Age      |    |    |    |
| ≤40      | 70 (56.5%) | 131 (60.6%) | 50 (51%) |
| >40      | 54 (43.5%) | 85 (39.4%) | 48 (49%) |
| BMI      |    |    |    |
| ≤25      | 75 (60.5%) | 87 (40.3%) | 27 (27.6%) |
| >25      | 49 (39.5%) | 129 (59.7%) | 71 (72.4%) |

Notes: *P is significant (≤0.05). **P is highly significant (≤0.01).

Table 7. Ranking test comparing CC, CT, and TT.

| GNB3 | MEAN RANK | χ² (P) |
|------|-----------|--------|
| Age  |           |        |
| CC   | 219.98    | 2.6 (0.27) |
| CT   | 211.55    |        |
| TT   | 236.42    |        |
| BMI Value |        |        |
| CC   | 160.41    | 39.43 (0.0**) |
| CT   | 237.39    |        |
| TT   | 254.85    |        |

Note: **P is highly significant (=0.01).
Conclusion
In our study, 825T allele was found to be a risk factor for hypertension. Identification of the candidate genes and understanding of their function hold the promise for the establishment of the specific molecular basis for EH. More studies with larger sample sizes on different populations are needed to determine whether suggested functional candidate genes play any role at all in the etiology of EH.

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Author Contributions
Conceived and designed the experiments: NSEDH. Analyzed the data: NSEDH. Wrote the first draft of the manuscript: NSEDH. Contributed to the writing of the manuscript: AAM. Agreed with manuscript results and conclusions: NSEDH, AAM, MMA. Jointly developed the structure and arguments for the paper: NSEDH, AAM. Made critical revisions and approved the final version: AAM. All the authors reviewed and approved the final manuscript.

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