Polymorphisms of the NRAMP1 gene: Distribution and susceptibility to the development of pulmonary tuberculosis in the Greek population

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Summary

Background: Ample evidence suggests that host genetic factors affect human susceptibility to tuberculosis. The natural resistance–associated macrophage protein 1 (NRAMP1) gene seems to play a role in the pathophysiology of a number of intracellular infections, including mycobacteria. A case-control study was conducted in the Greek population to determine whether NRAMP1 polymorphisms affect the susceptibility to development of overt pulmonary tuberculosis.

Material/Methods: NRAMP1 polymorphisms (3'UTR, D543N, INT4) were evaluated among 142 patients with culture-positive pulmonary tuberculosis and 144 ethnically matched healthy controls having latent M. tuberculosis infection. Patients with human immunodeficiency virus infection were excluded.

Results: Out of the 3 NRAMP1 polymorphisms, a trend of increased incidence of INT4 polymorphism was found in the patients’ group compared to the control group. A lack of association was observed between the 2 groups as far as the other 2 polymorphisms (D543N, 3'UTR) are concerned. INT4-CC homozygotes were found to have a higher risk to develop pulmonary tuberculosis compared to GG homozygotes (p=0.022). An increased incidence G/TGTG/C genotype combination was found in the patients’ group as compared to controls. G/TGTG/C genotype combination was associated with a 36% higher risk of developing pulmonary tuberculosis (p=0.004) compared to the baseline expression of G/TGTG/G combination.

Conclusions: INT4-NRAMP1 polymorphism may have a role in the development of culture-positive pulmonary tuberculosis after an initial M. tuberculosis latent infection. The possible role of INT4-NRAMP1 polymorphism in the development of active pulmonary tuberculosis needs further investigation.

key words: NRAMP1/SLC11A1 gene • tuberculosis • polymorphisms • ethnic groups
BACKGROUND

Tuberculosis (TB) remains the most widespread and deadly infectious disease worldwide, and it has been recently declared as a great emergency by the World Health Organization (WHO) [1]. Unfortunately, 8.9–9.9 million new cases of TB were reported in 2008, while 1.1–1.7 million human immunodeficiency virus (HIV)-negative patients died from this disease [2]. The factors which are mainly responsible for this evolving incidence are: (i) the migration of individuals from regions with high TB incidence to those with low incidence, (ii) the increasing number of HIV-positive individuals, and (iii) the emergence of multidrug-resistant TB due to unwise use of antibiotics [1,2]. In a total population of approximately 11 million people, the incidence of TB in Greece is estimated at 18/100,000 persons and the mortality rate at 2/100,000 annually [2–4].

The spread of *Mycobacterium tuberculosis* in the majority of humans is inhibited after the development of an effective immune response. Less than 10% of infected persons will eventually develop clinical disease [3]. Interestingly, only a minority of these patients have an identifiable risk factor, such as HIV infection, advanced age, alcohol abuse, chronic corticosteroid therapy, diabetes and poor nutrition/socio-economic status [6,7]. In the remaining patients, a complex interaction of genetic and environmental factors is possibly the cause. Studies in twins strongly suggest that host genetic factors play a major role in the susceptibility to development of TB [8]. Intense efforts are therefore being made to discover any possible genetic factors that may have a role in the host defense system and explain the increased susceptibility to overt *M. tuberculosis* infection. In 1981 a gene was found in mice that played an essential role in the determination of resistance to mycobacteria and other intracellular pathogens [9]. This gene was named natural resistance-associated macrophage protein 1 (Nramp1) [9]. The human homologue of the *Nramp1* gene, designated *NRAMP1*, now known as *SLC11A1*, is located on the long arm of chromosome 2 (2q35). *NRAMP1* is considered as a strong candidate gene for human resistance to TB. *NRAMP1* protein seems to play an important role in regulation of cytoplasmic calcium ions, especially iron. Iron is an essential mycobacterial nutrient, and has a major role in generating reactive oxygen and nitrogen intermediates in macrophages; hence, it affects the intracellular growth of mycobacteria [10,11]. Although it was believed that the gene is expressed solely in reticuloendothelial cells, it was recently shown to be expressed in CD11c+ dendritic cells, and exerts its pleiotropic effects on cytokine transcription, major histocompatibility complex (MHC) class II molecule expression, and presentation of protein antigens to T lymphocytes [12].

The possible association of *NRAMP1* polymorphisms with susceptibility to development of TB has been studied in different countries. The results of these studies among different ethnic groups have been controversial. A case-control study, conducted in West Africans, showed a positive association of the gene with pulmonary TB [13]. A similar study, conducted in Koreans, detected a significant association for the 3’UTR polymorphism, but not for the INT4 locus [14]. On the other hand, studies conducted in Denmark, Mexico, Morocco, Japan, Brazil and China suggested that the gene is not associated with an increased risk for developing TB [15–17]. In a study conducted in the Chinese population, *NRAMP1* allelic polymorphisms at the INT4 and D543N loci were each significantly associated with severe forms of pulmonary TB [17]. The aim of the present study was to explore the association between *NRAMP1* polymorphisms and the susceptibility to develop active pulmonary TB after a latent *M. tuberculosis* infection, in the Greek population.

MATERIAL AND METHODS

Study population

Patients’ group

Inclusion criteria

The inclusion criteria for the patients’ group were: (i) Greek origin; (ii) all patients should be genetically unrelated; and (iii) diagnosis of pulmonary TB based on suggestive radiological findings, a microscopic examination of sputum smears from an experienced microscopist, and documented in all patients with the presence of positive sputum cultures for *M. tuberculosis* infection. Written informed consent to participate was obtained from all patients who were included.

Exclusion criteria

The exclusion criteria for the patients’ group were: (i) presence of extrapulmonary TB; (ii) patients known to have an HIV infection were not recruited (all the other patients who agreed to participate were screened for HIV antibodies and HIV-positive patients were not enrolled in the study); and (iii) presence of any immunodeficiency syndrome or systemic disease.

Control group

Inclusion criteria

The inclusion criteria for the control group were: (i) ethically matched volunteers who visited Sotiria Hospital for a routine check-up; and (ii) the presence of a latent *M. tuberculosis* infection. Latent *M. tuberculosis* infection was defined as a positive tuberculin skin and at least 2 negative sputum cultures, combined with the absence of suggestive radiological findings for pulmonary TB. Written informed consent to participate was obtained from all controls who were included.
Briefly, healthy subjects who visited Sotiria Hospital for a routine check-up were enrolled. A 5 ml blood sample was drawn from every healthy volunteer, and was handled identically to the patients’ group. In all controls a tuberculin skin test was performed. Specifically, 0.1 ml of purified protein derivative (PPD), which contained 5 tuberculin units, was injected intradermally in the volar surface of the forearm. A 27-gauge needle on a tuberculin syringe was used. The transverse width of induration in millimetres (mm) at the skin test site was measured after 48-72 hours. A positive tuberculin skin test was defined as skin transverse width ≥ 12 mm. In all of the controls enrolled, a chest-X-ray (CXR) and sputum cultures for *M. tuberculosis* were obtained in order to rule out active disease. One hundred forty-four ethnically matched volunteers with latent *M. tuberculosis* infection were included.

**Exclusion criteria**

The exclusion criteria for the control group were: (i) HIV-positive subjects; (ii) history of prior TB; (iii) evidence of current or prior TB in the CXR; and (iv) presence of any immunodeficiency syndrome or systemic disease.

**NRAMP1 genotyping**

DNA samples were extracted from human whole blood collected into tubes containing EDTA. The kit used for the extraction was PureLink™ Genomic DNA kit Catalog no.s K1820-01, K1820-02, K1821-04, from Invitrogen. The NRAMP1 polymorphisms typed were: (i) a single-nucleotide change in intron 4 (INT4) (469+14G/C); (ii) D543N, a non-conservative single-base substitution at codon 543, which changes aspartic acid to asparagine; and (iii) 3'UTR, a TGTG deletion in the 3'untralsated region (1729+55del4). The PCR primers for the INT4 polymorphism were 5’CTCTGGCTG AAGGCTCTCGG3’ and 5’TGTGCTAT CAGTGAGCCTC3’. The primers for D543N and 5’UTR were 5’GCATCT CCCCATCTGTTG3’ and 5’AAC TTCCAGCTCTATCCTG3’, respectively.

Real-time PCR studies were performed in a LightCycler® 480 Instrument, Roche. For each of the 3 polymorphisms analyzed, a LightSNip kit was designed from the constructor company, TIB-MolBiol, Germany. Every reagent vial contained all primers and probes to run 96 Lightcycler reactions. The PCR reaction mixture contained 14.4–10.4 µl PCR grade H20, 1.0 µl Reagent Mix 2.0 µl Lightcycler FastStart DNA Master Hypro 1.6 µl MgCl₂, and 1.0–5.0 µl DNA template. The LightCycler® 480 Instrument was programmed according to the manufacturer’s parameters.

**Statistical analysis**

Initially, variants at 3 polymorphisms (3’UTR, INT4, D543N) in the NRAMP1 gene were compared between tuberculosis cases and controls with Fisher’s Exact test. To assess the differences in demographic characteristics, we used chi-square (for categorical data) and t test (for continuous data). Deviations from Hardy-Weinberg Equilibrium (HWE) were assessed using the chi-square test. Subsequently, the 3 NRAMP1 polymorphisms were each independently associated with tuberculosis through logistic regression analysis after adjustment for age and sex. The SAS statistical package

| Variables | pTB patients (n=142) | Controls (n=144) | P-value* |
|-----------|----------------------|------------------|----------|
| Gender    |                      |                  |          |
| Male (%)  | 98 (69.0)            | 94 (65.3)        | NS       |
| Female (%)| 44 (31.0)            | 50 (34.7)        |          |
| Age (years) ±SD | 54.32±16.44       | 59.63±10.77      | 0.001    |
| D543N     |                      |                  |          |
| G/G (%)   | 138 (97.2)           | 139 (96.5)       |          |
| G/A (%)   | 4 (2.8)              | 5 (3.5)          |          |
| Del/del (%) | 1 (0.7)             | 1 (0.7)          |          |
| 3’UTR     |                      |                  |          |
| G/G (%)   | 84 (59.2)            | 93 (64.6)        | 0.058    |
| G/C (%)   | 48 (33.8)            | 49 (34.0)        |          |
| C/C (%)   | 10 (7.0)             | 2 (1.4)          |          |

* P-values derived from: Chi-square for gender, t-test for age and Fisher’s exact test for gene polymorphisms; NS – Not statistically significant; pTB – Pulmonary tuberculosis.

(Version 9.1, SAS Institute Inc, Cary, NC) was used to analyze the data. Significance level was set at p=0.05.

**RESULTS**

All patients who were enrolled had a culture-positive pulmonary TB, combined with suggestive radiological findings. Positive acid-fast bacilli (AFB) sputum smear for *M. tuberculosis* was observed in 108 (76%) patients; the remaining 34 (24%) patients had a negative AFB smear test. The demographic characteristics and the frequency of the 3 NRAMP1 polymorphisms (3’UTR, D543N, INT4) between patients and controls are shown in Table 1. There was no evidence for deviation from HWE regarding the polymorphisms D543N (p=0.07) and INT4 (p=0.08), whereas the 3’ UTR (p=0.058) was found to deviate significantly from HWE.

The NRAMP1 allelic frequencies for the D543N and 3’UTR variants were not found to differ significantly between patients with pulmonary TB and controls. D543N-associated G/G and G/A variants were observed in 138 and 4 samples in the patients’ group, respectively. In controls, D543N-associated G/G and G/A genotypes were observed in 139 and 5 samples, respectively. The genotype A/A was absent in both groups. In the patients’ group, the 3’UTR-associated TGTG/TGTG, TGTG/del and del/del variants were observed in 137, 4 and 1 sample, respectively. In the control group, the numbers of samples with positive 3’UTR
variants were 139, 4 and 1 sample, respectively. However, we identified an increased incidence of INT4 polymorphism in the patients’ group compared to the control group, which was close to the borderline of statistical significance (p=0.058).

Three separate logistic regression analyses were performed, after adjustment for age and sex, in order to further investigate the possible association of TB patients with the allelic frequencies of the 3 NRAMP1 polymorphisms (Table 2). No significant association was found in patients with pulmonary TB for the D543N and 3'UTR allele variants. A significant association was found between the allele variants for the INT4-NRAMP1 polymorphism. Specifically, when GG allele genotype was used as baseline value, CC homozygotes were found to be at higher risk for developing pulmonary TB (OR=1.36; 95% CI: 1.11–1.66; p=0.004) compared to the baseline expression of G/TGTG/G. An association between pulmonary TB and G/del/G, A/del/G and A/TGTG/G genotype combinations, compared to the baseline expression of G/TGTG/G, was not observed.

A more detailed analysis was performed by creating 8 categories of genotype combinations: G/TGTG/G, G/TGTG/C, G/del/G, A/del/G, A/TGTG/G, G/del/C, A/TGTG/C and A/del/C (Table 3). G/TGTG/G was the main genotype combination observed. Significant differences among the frequencies between patients and controls were found regarding G/TGTG/G and G/TGTG/C genotype combinations. Specifically, lower expression of G/TGTG/G combination (p=0.005) was found in the patients’ group compared to the control group, while higher expression of the G/TGTG/C combination (p=0.004) was found in the patients’ group compared to the controls.

From a total of 8 genotype combinations, 5 (G/TGTG/G, G/TGTG/C, G/del/G, A/del/G, A/TGTG/G) were included in a multivariate model (Table 4). The remaining 3 combinations (G/del/C, A/TGTG/C, A/del/C) were excluded because of their extremely low frequencies. G/TGTG/C was found to be associated with a 36% higher risk for developing pulmonary TB (OR=1.36; 95% CI: 1.11–1.66; p=0.004).

### Table 2. Logistic regression-derived odds ratios (ORs) and 95% confidence intervals (95% CIs) for the risk of developing pulmonary tuberculosis according to the variant of each of the three allele polymorphisms, after adjustment for gender and age.

| Variable | ORs | 95% CI | P-value |
|----------|-----|--------|---------|
| D543N    |     |        |         |
| G/G      | Baseline |       |         |
| G/A      | 1.02 | 0.22–4.70 | 0.977 |
| 3'UTR    |     |        |         |
| TGTG/TGTG | Baseline |       |         |
| TGTG/del | 1.26 | 0.25–6.37 | 0.780 |
| Del/del  | 0.57 | 0.02–18.50 | 0.754 |
| INT4     |     |        |         |
| G/G      | Baseline |       |         |

### Table 3. Frequency of NRAMP1 genotype combinations among 1136 alleles derived from the 142 patients with pulmonary tuberculosis and 1152 alleles derived from 144 controls.

| Variables | pTB patients | Controls | P-value |
|-----------|--------------|----------|---------|
| G/TGTG/G  | 842 (74.1)   | 911 (79.0) | 0.005*  |
| G/TGTG/C  | 262 (23.1)   | 209 (18.1) | 0.004** |
| G/del/G   | 10 (0.9)     | 11 (1.0)  | 0.852*  |
| A/del/G   | 6 (0.5)      | 11 (1.0)  | 0.235*  |
| A/TGTG/G  | 6 (0.5)      | 7 (0.6)   | 0.800*  |
| G/del/C   | 6 (0.5)      | 1 (0.1)   | 0.068** |
| A/TGTG/C  | 2 (0.2)      | 1 (0.1)   | 0.622** |
| A/del/C   | 2 (0.2)      | 1 (0.1)   | 0.622** |

* P-value derived from chi-square test; ** P-value derived from Fisher’s Exact test; pTB – Pulmonary tuberculosis.

### Table 4. Logistic regression-derived odds ratios (ORs) and 95% confidence intervals (95% CIs), estimating the risk of developing pulmonary tuberculosis, among the five most common NRAMP1 genotype combinations.

| Variable | ORs | 95% CI | P-value |
|----------|-----|--------|---------|
| G/TGTG/G  | Baseline |       |         |
| G/TGTG/C  | 1.36 | 1.11–1.66 | 0.004 |
| G/del/G   | 0.98 | 0.42–2.33 | 0.970 |
| A/del/G   | 0.59 | 0.22–1.60 | 0.301 |
| A/TGTG/G  | 0.93 | 0.31–2.77 | 0.893 |

### Discussion

To our knowledge this is the first study in Greece that examines the possible association of NRAMP1 polymorphisms and the susceptibility to develop culture-positive pulmonary TB. Our study demonstrated that there was not any significant difference in the incidence of NRAMP1 D543N and 3'UTR polymorphisms between patients with pulmonary TB and ethnically matched healthy controls having latent M. tuberculosis infection. An increased incidence of INT4-NRAMP1 polymorphism was found in the patients’ group, which was close to the borderline of statistical significance.

A positive association of 3'UTR variant allele and the susceptibility to develop TB was reported in a small Caucasian population [18]. However, several studies performed in European populations suggested no statistically significant association between NRAMP1 polymorphisms and the susceptibility to develop TB. This was also confirmed in the biggest meta-analysis of published trials to date [19]. In
this analysis, the odds ratios (ORs) among 3'UTR, D543N, INT4 locus and TB susceptibility, in the European population, were 1.81 (95% CI: 0.66–4.93; p=0.25), 1.88 (95% CI: 0.75–4.67; p=0.18) and 0.87 (95% CI: 0.61–1.22; p=0.41), respectively [19]. It is believed that the European origin population has a high level of "genetical" resistance to M. tuberculosis infection, due to natural selection over the last 300 years [20,21]. The lower proportion of TB cases in the European population, compared to Asian and African populations, could be another reason for the lack of association [19]. We found a lack of correlation among D543N and 3'UTR NRAMP1 polymorphisms and an increased susceptibility to develop pulmonary TB, according to results of the majority of studies in European populations.

Although our results are close to statistical significance, there is a trend for a higher incidence of INT4-NRAMP1 polymorphism in patients with pulmonary TB, compared to ethnically matched volunteers with latent M. tuberculosis infection. This possible association appears for the first time in a European population, and can be possibly attributed to the diverse genetic background among various ethnic populations of the same descent. Ethnic or racial backgrounds can certainly be responsible for some variations observed [18]. Specifically, the INT4-NRAMP1 polymorphism can be found in approximately 20% of the Chinese population, whereas the frequency in white Europeans is about 50%, with significant variation among different European populations [17,19].

The INT4-CC homozygotes variants were found to have an increased risk for developing pulmonary TB, compared to their corresponding common GG alleles. When a more detailed analysis was performed by creating 8 categories of genotype combinations, higher expression of G/TGTG/C combination (p=0.004) was found in the patient’s group as compared to the controls. This combination was associated with a 36% higher risk for developing pulmonary TB (OR=1.36; 95% CI: 1.11–1.66; p=0.004) compared to the baseline expression of G/TGTG/G. However, the limited number of patients and controls found to carry the INT4-CC genotype must be taken into consideration in the interpretation of our results. Whether the incidence of INT4-CC variant or any NRAMP1 genotype combination is correlated to the development of active disease needs further investigation.

Most of the studies exploring the possible role of NRAMP1 polymorphisms in the development of TB did not distinguish between susceptibility to infection with M. tuberculosis and susceptibility to progression to active disease, since the control groups consisted of healthy volunteers without active disease. Our study design distinguishes between susceptibility to infection with M. tuberculosis and susceptibility to progression in an active M. tuberculosis pulmonary disease. All volunteers in the control group had a latent M. tuberculosis infection without developing the disease. This fact suggests that INT4-NRAMP1 polymorphisms may have a role in the development of overt pulmonary disease after a latent M. tuberculosis infection. The main limitations of our study are the lack of data in both study groups that can be associated with a higher risk of developing pulmonary TB (alcohol abuse, chronic corticosteroid therapy, diabetes, poor nutrition/socio-economic status), and the relatively small sample size of the population enrolled.

**Conclusions**

INT4-NRAMP1 polymorphism may be associated with a higher rate of culture-positive pulmonary TB in the Greek population compared to ethnically matched healthy controls with latent M. tuberculosis infection. A lack of association was observed for the other 2 NRAMP1 polymorphisms (D543N, 3'UTR). INT4-NRAMP1 CC homozygotes were found to have a higher risk compared to GG homozygotes. An increased incidence of G/TGTG/C genotype combination was found in the patients' group compared to controls. G/TGTG/C genotype combination was associated with a 36% higher risk for developing pulmonary TB compared to the baseline expression of G/TGTG/G genotype combination. The possible role of INT4-NRAMP1 polymorphism in the development of active pulmonary TB, after an initial latent M. tuberculosis infection, needs further investigation.

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**Conflict of interest**

All authors declare no conflict of interest in relation to this study.

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