Use of a linearization approximation facilitating stochastic model building

Elin M. Svensson · Mats O. Karlsson

Abstract The objective of this work was to facilitate the development of nonlinear mixed effects models by establishing a diagnostic method for evaluation of stochastic model components. The random effects investigated were between subject, between occasion and residual variability. The method was based on a first-order conditional estimates linear approximation and evaluated on three real datasets with previously developed population pharmacokinetic models. The results were assessed based on the agreement in difference in objective function value between a basic model and extended models for the standard nonlinear and linearized approach respectively. The linearization was found to accurately identify significant extensions of the model’s stochastic components with notably decreased runtimes as compared to the standard nonlinear analysis. The observed gain in runtimes varied between four to more than 50-fold and the largest gains were seen for models with originally long runtimes. This method may be especially useful as a screening tool to detect correlations between random effects since it substantially quickens the estimation of large variance-covariance blocks. To expedite the application of this diagnostic tool, the linearization procedure has been automated and implemented in the software package PsN.

Keywords Linearization · Random effects · Nonlinear mixed effects models · Pharmacometrics · Diagnostics

Introduction

Population pharmacokinetics (PK) and pharmacodynamics (PD) models, i.e. pharmacometrics, play an increasingly important role in pharmaceutical sciences and drug development [1]. Nonlinear mixed effects (NLME) models are frequently used to describe population PK/PD and are composed of a structural component (fixed effects) and a stochastic component (random effects). Fixed effects describe the typical parameter values in a population and the effects of covariates. Random effects handle unexplained variability and can be further divided into variability assigned to parameters and residual variability (RV) assigned to the observations. Parameters can vary on multiple levels: between subjects (BSV), between occasions (BOV), between studies, etc.

For NLME, especially when used in simulations, the stochastic components of the model are crucial but the development procedure can be laborious. Despite a vast increase in available computational power during the past decade, the time for parameter estimation can still be a limiting factor since the complexity of the used models and the amount of fitted data typically are increasing likewise. For highly nonlinear models numerical instability of parameter estimation with common gradient based, local methods may cause further problems. Individual parameter estimates, the empirical Bayes estimates (EBEs), can be used as a diagnostic tool to aid the development procedure. However, this practice has notable shortcomings when data are uninformative on the individual level and shrinkage towards the population mean occurs. Shrinkage can cause diagnostics based on EBEs to mask, falsely induce or distort the shape of random effects relationships [2, 3]. The objective of this work was to develop and assess a fast and stable method which is not sensitive to shrinkage for diagnostics of BSV, BOV and RV model.
components. The here proposed method, henceforth called ‘linearization’, is based on a first-order conditional estimation (FOCE) linear approximation which has previously successfully been applied for testing of covariates [4]. The linearization was evaluated in the modeling software NONMEM with a variety of different models and datasets. The results are presented as comparisons between the linearization and the corresponding nonlinear model, both in terms of estimation performance and runtimes.

Methods

Population models and linearization

NLME models commonly used in population PK/PD can be formally represented by

\[ y_{ij} = f(\vec{p}_i, \vec{x}_{ij}) + h_{ij} \]  

(1)

where \( y_{ij} \) is the data point of the \( i \)th individual’s \( j \)th observation, \( f \) is a model that relates the vector of individual parameters \( \vec{p}_i \) and the vector of independent variables \( \vec{x}_{ij} \) (for example dose and time) to the observations and \( h_{ij} \) is a model for the residual error. The individual parameters can be described as

\[ \vec{p}_i = \vec{p}'(\vec{\theta}, \vec{\eta}_i, \vec{g}(\vec{z}_i, \vec{\theta} / \vec{g})) \]  

(2)

where \( \vec{p} \) is the vector of models relating the typical parameter values in the population \( \vec{\theta} \), the parameter-specific variability \( \vec{\eta}_i \) and the vector of covariate functions \( \vec{g} \), including covariate observations \( \vec{z}_i \) and typical population values for respectively covariate-parameter relation \( \vec{g}_k \). The parameter-specific variability can have multiple levels simultaneously and is usually assumed to be normally distributed with mean 0 and variance–covariance matrix \( \Omega \). BSV and BOV are frequently modeled by assuming an exponential relationship to obtain log-normal parameter distributions. Under this assumption, parameter \( k \) in individual \( i \) with \( L \) levels of parameter-specific variability can be described by the following model

\[ p_{ki} = \theta_k \times g_k(\vec{z}_i, \vec{\theta} / \vec{g}) \times e^{(\sum_{u=1}^{L} \eta_{iku})} \]  

(3)

The residual error model is commonly modeled as a function of \( h_{ij} \) which is assumed to be normally distributed with mean 0 and variance–covariance matrix \( \Sigma \). Common forms of \( h_{ij} \) are additive error \( (h_{ij} = e_{ij}) \), proportional error \( (h_{ij} = f(\vec{p}_i, \vec{x}_{ij}) \times e_{ij}) \) or a combined error with both an additive and a proportional element \( (h_{ij} = f(\vec{p}_i, \vec{x}_{ij}) \times e_{ij} + \delta_{ij}) \). Examples of extensions and more flexible forms have been described in the following references [5–8].

Model parameters can be obtained by maximum likelihood estimation where the estimated parameter values maximizes the likelihood of the observed data [9]. Maximizing the likelihood is equivalent to minimizing the twice negative log-likelihood (objective function value, OFV). Evaluation of the function for the log-likelihood of NLME models require integration steps that are computationally demanding and a simpler linear approximation of the model can be evaluated instead. We here assess the possibility of utilizing partial derivatives and EBEs from evaluation of a nonlinear base model to assess the improvement in model fit by additional random effects through estimation of extensions implemented on the linear approximation of the base model. The linear approximation consists of first-order Taylor expansions initially around \( \vec{\eta}_i = 0 \) (Eq. 4) and subsequently around \( \vec{\eta}_i = \vec{\eta}_i \) where \( \vec{\eta}_i \) represents the EBE’s of \( \vec{c}_i \) (Eq. 5, the linearized model).

\[ y_{ij} \approx f(\vec{p}_i, \vec{x}_{ij}) + \sum_{v=1}^{s} \frac{\partial h_{ij}}{\partial c_{ijv}} \Delta c_{ijv} + e_{ij} \]  

(4)

\[ y_{ij} \approx f(\vec{p}_i, \vec{x}_{ij}) \bigg|_{Q_0} + \sum_{l=1}^{m} \frac{\partial f}{\partial \eta_{l}} \bigg|_{Q_0} (\eta_{li} - \hat{\eta}_l) + \sum_{v=1}^{s} \frac{\partial h_{ij}}{\partial c_{ijv}} \bigg|_{Q_0} e_{ijv}^* \]  

\[ + \sum_{v=1}^{s} \sum_{l=1}^{m} \frac{\partial}{\partial \eta_{l}} \left( \frac{\partial h_{ij}}{\partial c_{ijv}} \right) \bigg|_{Q_0} (\eta_{li} - \hat{\eta}_l) \]  

(5)

where \( Q_0 = (\vec{c}_{ij} = 0, \vec{\eta}_i = \vec{\eta}_i) \), \( m \) is the number of element in \( \vec{\eta}_i \) and \( \tau \) is the number of elements in \( \vec{c}_{ij} \). The partial derivatives and EBE’s are obtained by evaluating the nonlinear base model with the first order conditional estimation method with interaction (FOCE-I) in NONMEM (see section Software). If the error model is simply additive FOCE without interaction can be used. The FOCE and FOCE-I methods also utilize Taylor expansions and are described in NONMEM users guide-part VII [10]. The parameters marked with asterisks in Eqs. 4 and 5 are the unknown parameters of the linearized model that remain to be estimated. A comprehensive example code is provided as supplementary material. Since the iterative calculation of fixed effects parameters and partial derivatives are time consuming steps, especially for models including large variance–covariance matrices, the estimation of a beforehand linearized extended model (where iterative calculations only are needed for the random effects) is magnitudes faster than estimation of the nonlinear extended model.

Software

NONMEM versions 7.2 and 7.3 beta (ICON Development Solutions, Hanover, MD, USA) [10] with the estimation method FOCE-I were used for the analysis. To overcome
sensitivity to local minima when estimating individual parameters ($\eta$-values) the new NONMEM option MCETA was utilized. The default initial value for all $\eta$ is zero. MCETA allows the user to define a number of vectors containing random samples of initial $\eta$-values that should be tested in addition to zero, whichever supplies the lowest OFV will be used as initial value in the estimation. The numbers in each vector of initial $\eta$-values are randomly drawn from the normal distribution with mean zero and variance–covariance matrix $\Omega$. With a large enough number of initial $\eta$-values tested, the probability of the estimation to end up in a local minimum can be decreased.

Models were executed through PsN, using Piranha for documentation and creation of run records [11–13].

Evaluated model extensions

A comprehensive overview of the work flow for comparing standard nonlinear and linearized models is given in Fig. 1. The evaluated RV models were extensions to an additive, a proportional or a combined base error model. The evaluated extensions were:

- BSV of the residual error [14]
- A power relation with the individual model predictions ($F$), implemented as $h_{ij} = h_{ij(base)} \times F^{\delta_i}$
- Autocorrelation, i.e. serial correlation between the error of observations consecutive in time [5]
- Time dependent residual error, implemented as a step function [5]

The NONMEM code of the RV extensions is provided in the supplementary material. For extensions of the BOV and/or the correlation structure, a base model already including some BSV terms was used while for extensions of the BSV structure the base model only included RV. To this model additional parameter-specific variability were added and/or the structure of the variance–covariance block changed. To enable estimation of the linearized model with additional variability parameters, the partial derivatives of the model with respect to the new parameters must be known. The partial derivatives can be obtained in NONMEM by including the code for the additional variability parameters already in the nonlinear base model but fixed to an arbitrary small value (fixing it to zero would result in that no derivatives are calculated).

All extended models were estimated with FOCE-I both in the linearized form and as standard nonlinear models. The results were evaluated by comparing the difference in OFV ($\Delta$OFV) between the extended model and the base model for the two approaches respectively. The runtimes for the estimation step were extracted from the result files. The runtime comparisons should be interpreted in terms of magnitude and not as exact figures, since all estimations were carried out at a cluster with many nodes, which have somewhat different capacities and are randomly assigned.
Datasets

Three real data examples with previously developed models with diverse residual error structures were used to illustrate the methodology.

1. Moxonidine [14]: The data were obtained from a phase II multicenter study of oral moxonidine in patients with congestive heart failure and contained 1,022 observations from 74 patients. The structural model describing the data was a one-compartment model with first-order absorption and an additive residual error model.

2. Pefloxacin [15, 16]: The data were obtained from critically ill patients receiving 1 h intravenous infusions of pefloxacin and contained 337 observations from 74 patients. The structural model was a one-compartment model and a proportional residual error model.

3. Ethambutol [17]: The data were obtained from two prospective studies in tuberculosis patients receiving a standard treatment regimen including ethambutol. A total of 1869 observations from 189 patients were included in the dataset. The structural model was a one-compartment model with first-order absorption through one transit compartment and a combined residual error model.

Results

The agreement between the ΔOFV of the nonlinear and the linearized extended RV models were found to be good (Fig. 2a). Also for extended correlations structures the agreement was good (Fig. 2d). For extended BSV and BOV models the agreement was acceptable (Fig. 2b, c). The deviations between the nonlinear and linearized model were the largest for instances where the value of one or several fixed effect parameter(s) in the nonlinear model were notably changed by the extension. The extended linearized models do not allow a change of the fixed effects since those values are incorporated in the predictions and derivatives obtained from the base model. The linearized analysis identified the same extended models to be significant improvements as the conventional nonlinear analysis.

![Fig. 2](image-url) Difference in OFV between base and various extended RV (a), IIV (b), IOV (c) and covariance (d) models (described in methods), after estimation with the nonlinear vs. the linearized approach for the moxonidine (black triangles), pefloxacin (grey squares) and ethambutol (open circles) data examples.
in all cases except in four, resulting in an accuracy of 96% (88 of 92). In the four deviating cases the ΔOFVs were very close to the significance level (following Chi square distribution, \( \alpha = 0.05 \)).

The total runtimes for the linearized models using the three example datasets were markedly shorter compared to the corresponding nonlinear models (Table 1). When the linearization of the base model is successful, the OFV value of the linearized version and that of the corresponding nonlinear model estimated with FOCE-I should be very similar. This was the case for all evaluated models with an additive residual error model. However, for models with proportional or combined residual error models, a number of cases were discovered where the OFV values differed greatly and a lower OFV value was obtained for the nonlinear base model. Closer evaluation of the results revealed that this deviation between OFV values was caused by certain subjects for whom the individual contributions to the OFV (iOFV) were notably higher, whereas for the majority of subjects the iOFV values of the two methods matched well. For the deviating subjects the estimation of individual \( \eta \)-values had failed in the linearized model due to issues with local minima. Transformation to render the error additive in the transformed space or utilization of the MCETA option resolved the problem for all evaluated cases.

### Discussion

A novel diagnostic method for evaluation of random effects was successfully developed and evaluated. The linearization was found to accurately identify significant extensions of models’ stochastic components with notably decreased runtimes as compared to standard nonlinear analysis. The three examples used had relatively short runtimes also in their nonlinear form. When the linearization was applied to more complex models, an even more substantial decrease in runtimes (>50×) was observed. For a PD-model describing the effect of docetaxel on neutrophil counts (extension of [18]) utilizing the full random effects approach (FREM [19]) the runtime was decreased from 3 h and 50 to 5 min (2.2%). For a PK model of bedaquiline plus two metabolites including a large covariance block (extension of [20]) the runtime was reduced from 15 h and 52 to 6 min (0.6%). For a PK-model of rifabutin plus metabolite using a dataset combining 14 clinical studies (unpublished) for which the runtime of the base nonlinear model was several day, the estimation of 6 additional BSV parameters and extending the variance–covariance matrix from a diagonal to a full \( 12 \times 12 \) block structure took only 18 and 89 min, respectively.

When applied to models with a proportional or a combined residual error structure the linearization was found to be sensitive to local minima when assigning the individual \( \eta \)-values. This happens due to the potential shape distortion of the individual EBE likelihood profiles that can be caused by the interaction term (including the partial derivative with respect to both \( \varepsilon \) and \( \eta \) of the residual error model) in the linearized model. The interaction term will always be zero for additive residual error models which explains why the problem was never observed in this case. Care must be taken to ensure that the OFV of the linearized base model agrees with the corresponding nonlinear model. If a deviation is detected, simple work-arounds can preclude potential deviations. Transformations can render the error model additive in the transformed space, for example log-transformation for model with proportional error structure. Another solution is the MCETA option, available in NONMEM from version 7.3.0 [21], which decreases the risk of issues with local minima by supplying multiple initial estimates for the individual \( \eta \)-values. With MCETA values of between 10 and 1,000 all evaluated linearized models were able to obtain the same OFV value as the corresponding nonlinear model. However, run times were somewhat increase by use of the MCETA option. Yet another potential solution could be to input the EBE’s from the nonlinear base model as initial estimates for individual \( \eta \)-values in the linearized model. This is possible with the ETAS option, also available in NONMEM from version 7.3.0. The ETAS option should be faster than the MCETA option since it only tests one set of initial estimates instead of several, but it may be less reliable. In cases when the shape of the likelihood profiles contain multiple local minima, as have been observed for linearized models with proportional or combined error structures, the risk of terminating in local minima is substantial even with initial estimates close to the optimal values and therefore the use of the MCETA option could be safer.

To simplify the use of this novel methodology, an option called ‘linearize’ has been implemented in PsN (available from version 3.7.0). The procedure requires the provision of a nonlinear model, optimizes the parameters and outputs predictions and derivatives in a new dataset. This dataset

---

**Table 1** Comparison of total runtimes for nonlinear and linearized models using the three example datasets

| Data          | Total runtime nonlinear (s) | Total runtime linearized (s) | Fraction time required linearized (%) |
|--------------|-----------------------------|-----------------------------|-------------------------------------|
| Moxonidine   | 735.1                       | 106.5                       | 14.5                                |
| Pefloxacin   | 47.78                       | 6.96                        | 14.6                                |
| Ethambutol   | 103082                      | 25983\(^a\)                 | 25.2                                |

\(^a\) Executed with MCETA = 1,000
constitutes the input for an automatically created linearized version of the same model. The linearized model can serve as starting point for evaluation of manually coded extensions of the random effects. Model building could potentially be made even more efficient by automated testing of a library of RV models, comparable to the stepwise covariate search method (the SCM) already implemented in PsN. Since the values of fixed effects are not estimated in the linearized model the method should be viewed as a diagnostic tool. It is recommended to re-estimate with standard nonlinear methods once the best extended model is identified.

In conclusion, the successful use of a linear approximation method for fast diagnosis of a broad range of extended random effects models was demonstrated. The method may be especially valuable as a screening tool to detect correlations between random effects since estimating large variance–covariance blocks often is a computationally demanding and time-consuming process but can be carried out magnitudes faster with the linearization.

Acknowledgments We thank Joakim Nyberg for valuable help with understanding of the impact of the linearization on the likelihood function, Kajsa Harling for implementation in PsN and Ronald Niebecker and Hwi-yeol Yun for examples of the use of the linearization with more complex models. The research leading to these results has received support from the Innovative Medicines Initiative Joint Undertaking under Grant Agreement No. 115156, resources of which are composed of financial contributions from the European Union’s Seventh Framework Programme (FP7/2007-2013) and EFPIA companies’ in kind contribution. The DDMoRe project is also supported by financial contribution from the European Union’s Seventh Framework Programme (FP7/2007-2013) and EFPIA companies’ in kind contribution. The DDMoRe project is also supported by financial contribution from the European Union’s Seventh Framework Programme (FP7/2007-2013) and EFPIA companies’ in kind contribution.

Open Access This article is distributed under the terms of the Creative Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided the original author(s) and the source are credited.

References
1. Gobburu JV (2010) Pharmacometrics 2020. J Clin Pharmacol 50:151S–157S. doi:10.1177/0091270010376977
2. Savic RM, Karlsson MO (2009) Importance of shrinkage in empirical bayes estimates for diagnostics: problems and solutions. AAPS J 11:558–569. doi:10.1208/s12248-009-9133-0
3. Karlsson MO, Savic RM (2007) Diagnosing model diagnostics. Clin Pharmacol Ther 82:17–20. doi:10.1038/sj.clpt.6100241
4. Khandelwal A, Harling K, Jonsson EN, Hooker AC, Karlsson MO (2011) A fast method for testing covariates in population PK/PD Models. AAPS J 13:464–472. doi:10.1208/s12248-011-0289-2
5. Karlsson MO, Beal SL, Sheiner LB (1995) Three new residual error models for population PK/PD analyses. J Pharmacokin Biopharm 23:651–672
6. Dosne A-G, Keizer RJ, Bergstrand M, Karlsson MO (2012) A strategy for residual error modeling incorporating both scedasticity of variance and distribution shape. PAGE 21 Abstr 2527. www.page-meeting.org/?abstract=2527. Accessed 10 March 2014
7. Frame B, Miller R, Hutmacher MM (2009) Joint modeling of dizziness, drowsiness, and dropout associated with pregabalin and placebo treatment of generalized anxiety disorder. J Pharmacokinet Pharmacodyn 36:565–584. doi:10.1007/s10928-009-9137-5
8. Silber HE, Kjellsson MC, Karlsson MO (2009) The impact of misspecification of residual error or correlation structure on the type I error rate for covariate inclusion. J Pharmacokinet Pharmacodyn 36:81–99. doi:10.1007/s10928-009-9112-1
9. Bauer RJ, Güzy S, Ng Κ (2007) A survey of population analysis methods and software for complex pharmacokinetic and pharmacodynamic models with examples. AAPS J 9:E60–E83. doi:10.1208/aapsj0901007
10. Beal S, Sheiner LB, Boeckmann A, Bauer RJ (2010) NONMEM User’s Guides. (1989-2010). Icon Development Solutions, Ellicott City
11. Lindbom L, Phlgren P, Jonsson EN (2005) PsN-Toolkit—a collection of computer intensive statistical methods for non-linear mixed effect modeling using NONMEM. Comput Methods Progr Biomed 79:241–257. doi:10.1016/j.cmpb.2005.04.005
12. Keizer RJ, Karlsson MO, Hooker A (2013) Modeling and Simulation Workbench for NONMEM: Tutorial on Piranha, PsN, and Xpose. CPT: Pharmacomet & Syst Pharmacol 2:e50. doi:10.1038/psp.2013.24
13. Keizer RJ, van Benten M, Beijnen JH, Schellens JH, Huijtema AD (2011) Pirana and PCluster: a modeling environment and cluster infrastructure for NONMEM. Comput Methods Progr Biomed 101:72–79. doi:10.1016/j.cmpb.2010.04.018
14. Karlsson MO, Jonsson EN, Wiltsie CG, Wade JR (1998) Assumption testing in population pharmacokinetic models: illustrated with an analysis of moxifloxacin data from congestive heart failure patients. J Pharmacokinet Biopharm 26:207–246
15. Wahlby U, Thomson AH, Milligan PA, Karlsson MO (2004) Models for time-varying covariates in population pharmacokinetic-pharmacodynamic analysis. Br J Clin Pharmacol 58:367–377. doi:10.1111/j.1365-2125.2004.02170.x
16. Karlsson MO, Sheiner LB (1993) The importance of modeling interoccasion variability in population pharmacokinetic analyses. J Pharmacokinet Biopharm 21:735–750
17. Jonsson S, Davide A, Wilkins J, Van der Walt JS, Simonsson US, Karlsson MO, Smith P, McIlver H (2011) Population pharmacokinetics of ethambutol in South African tuberculosis patients. Antimicrob Agents Chemother 55:4230–4237. doi:10.1128/AAC.00274-11
18. Kloft C, Wallin J, Henningsson A, Chatelut E, Karlsson MO (2006) Population pharmacokinetic-pharmacodynamic model for neutropenia with patient subgroup identification: comparison across anticancer drugs. Clin Cancer Res: Off J Am Assoc Cancer Res 12:5481–5490. doi:10.1186/1078-0432.CCR-06-0815
19. Karlsson MO (2012) A full model approach based on the covariance matrix of parameters and covariates. PAGE 21 Abstr 2455. www.page-meeting.org/?abstract=2455. Accessed 10 March 2014
20. Svensson EM, Aweeka F, Park JG, Marzan F, Dooley KE, Karlsson MO (2013) Model-based estimates of the effects of efavirenz on bedaquiline pharmacokinetics and suggested dose adjustments for patients coinfected with HIV and tuberculosis. Antimicrob Agents Chemother 57:2780–2787. doi:10.1128/AAC.00191-13
21. Bauer RJ (2013) NONMEM users guide introduction to NONMEM 7.3.0. ICON Development Solutions, Hanover