A Review of Inflammatory Processes of the Breast with a Focus on Diagnosis in Core Biopsy Samples

Timothy M. D’Alfonso
Paula S. Ginter · Sandra J. Shin

Department of Pathology and Laboratory Medicine, Weill Cornell Medical College, New York, NY, USA

Received: June 2, 2015
Revised: June 10, 2015
Accepted: June 11, 2015

Corresponding Author
Timothy M. D’Alfonso, MD
New York-Presbyterian Hospital/Weill Cornell Medical College, 525 East 68th Street, Starr 10G1E, New York, NY 10065, USA
Tel: +1-212-746-2700
Fax: +1-212-746-6484
E-mail: tid9007@med.cornell.edu

Inflammatory and reactive lesions of the breast are relatively uncommon among benign breast lesions, and can be the source of an abnormality on imaging. Such lesions can simulate a malignant process, based on both clinical and radiographic findings, and core biopsy is often performed to rule out malignancy. Furthermore, some inflammatory processes can mimic carcinoma or other malignancy microscopically, and vice versa. Diagnostic difficulty may arise due to the small and fragmented sample of a core biopsy. This review will focus on the pertinent clinical, radiographic, and histopathologic features of the more commonly encountered inflammatory lesions of the breast that can be characterized in a core biopsy sample. These include fat necrosis, mammary duct ectasia, granulomatous lobular mastitis, diabetic mastopathy, and abscess. The microscopic differential diagnoses for these lesions when seen in a core biopsy sample will be discussed.

Key Words: Breast; Core biopsy; Inflammatory; Mammogram

Inflammatory and reactive conditions of the breast are relatively uncommon among benign breast lesions, and may present with clinical and radiologic abnormalities akin to malignant processes. As such, core biopsy may be performed to exclude the possibility of malignancy. In most cases, the diagnosis based on microscopy is clear, but in fragmented core biopsy samples, some conditions may mimic malignancy. Conversely, some malignancies can also simulate benign inflammatory or reactive conditions.

The clinical, radiographic, and histologic features of commonly encountered inflammatory and reactive breast lesions, namely, fat necrosis, mammary duct ectasia, granulomatous lobular mastitis, diabetic mastopathy, and abscess will be reviewed (Table 1). In addition, we will discuss the histlogic features on core biopsy that distinguish the items on this differential.

FAT NECROSIS

Fat necrosis is most often seen in traumatized breast tissue, particularly in areas of prior surgery or biopsy. Radiation therapy can also lead to fat necrosis in breast and has been seen in up to 50% of patients following balloon-based brachytherapy. Clinically, fat necrosis can present as a palpable mass with or without skin retraction, or it can be asymptomatic. On mammography, oil or lipid cysts, which are often calcified, are characteristic of fat necrosis (Fig. 1A). Calcifications in fat necrosis may be clustered, pleomorphic, and linear, simulating ductal carcinoma in situ (DCIS). Fat necrosis can also appear as a stellate mass with irregular margins on mammography and ultrasound.

Microscopically, fat necrosis is characterized by infiltration of foamy histiocytes and foreign body type giant cells around necrotic fat cells and lipid vacuoles (Fig. 1B). Cysts lined by foamy histiocytes are often present (Fig. 1C). Lymphocytic infiltration, often accompanied by plasma cells, can be present to varying degrees. As fat necrosis evolves, fibrosis develops within the lesion, forming a scar and fibrous walled cysts that can calcify (Fig. 1D–F). Adjacent glandular structures in a biopsied sample may show changes indicative of radiation therapy, such as cytologic atypia, squamous metaplasia, and thickened basement membranes.

The differential diagnosis for fat necrosis includes less common types of invasive carcinoma, including the lipid-rich variant and carcinomas with “histiocytoid” morphology, such as apocrine carcinoma and invasive lobular carcinoma. A broad-spectrum cytokeratin can be used to exclude the presence of carcinoma.
Table 1. Pertinent features of inflammatory lesions of the breast

| Clinical | Etiology and associations | Imaging | Microscopic features | Differential diagnosis |
|----------|--------------------------|---------|----------------------|-----------------------|
| Fat necrosis | Mass | Nipple/skin retraction | Trauma | MMG: oil cyst; calcifications; spiculated mass | Lipid vacuoles and necrotic adipocytes surrounded by foamy histiocytes and foreign body giant cells |
| Duct ectasia | Unilateral or bilateral non-bloody nipple discharge | Nipple/skin retraction | Mass | MMG: branching calcifications; stellate mass | Dilated ducts with luminal histiocytes |
| Granulomatous lobular mastitis | Mass | Erythema | Idiopathic Corynebacterim spp. | Often suspicious for malignancy |
| Diabetic mastopathy | Palpable mass or nodularity | Diabetes mellitus (types I-II) Autoimmune dx. | Endocrine dx. (i.e., thyroid dx.) | MMG: ill-defined mass; architectural distortion |
| Breast abscess | Tender mass with redness, warmth, swelling ± Draining sinus ± Nipple discharge | Lactational/puerperal abscess: S. aureus Subareolar/ non-puerperal abscess: - SMOLD - Smoking - Trauma - Mixed flora | MMG: asymmetric mass; skin thickening US: multiloculated mass/collection | Mixed inflammatory infiltrate, mainly neutrophils |

MMG, mammography; US, ultrasound; spp., species; TB, tuberculosis; dx, disease; MRI, magnetic resonance imaging; SMOLD, squamous metaplasia of lactiferous ducts.

Some histiocytic processes, although rare, should also be included in the differential diagnosis of fat necrosis. Rosai-Dorfman disease is a histiocytic proliferation that rarely involves the breast and is characterized by large histiocytes accompanied by a lymphoplasmacytic infiltrate. Histiocytes show emperipolesis and are discordant with the pathologic diagnosis of fat necrosis. Mammary duct ectasia is a histiocytic proliferation that rarely involves the breast and is characterized by dilatation of the central ducts with associated fibrosis and chronic inflammation. Perimenopausal and postmenopausal women are most often affected. Duct ectasia presents as unilateral or bilateral non-bloody nipple discharge, nipple retraction, or a palpable mass that is typically subareolar and sometimes associated with pain. On mammography, duct ectasia appears as branching calcifications, ductal dilatation, or a stellate mass. Ultrasound shows dilated subareolar ducts or a mass-like lesion filled with echogenic material, which may not be evident on mammography. On magnetic resonance imaging (MRI), duct ectasia can present with a pattern of enhancement that can mimic DCIS.
with luminal histiocytes is seen (Fig. 2A, B). Ductal epithelium is not hyperplastic and may be flattened or completely absent. Sloughed epithelium may be seen in ductal lumens (Fig. 2C). Lipid-laden foamy histiocytes can be seen within the duct lumen and in adjacent stroma. Intraepithelial histiocytes can also be seen. “Ochrocytes” refers to histiocytes in periductal stroma that show accumulation of lipofuscin pigment, which imparts a brown color to the cells (Fig. 2D). A periductal chronic inflammatory cell infiltrate composed of lymphocytes and plasma cells is also present, particularly when leakage of duct contents into the surrounding stroma has occurred. In the later stages of duct ectasia, fibrosis of duct walls, sometimes accompanied by elastosis, is the predominant histologic feature. The fibrotic wall of the duct may calcify, and can result in calcifications within duct lumens (Fig. 2E). A small portion of a fibrotic duct, with or without accompanying inflammation, may be the only finding in a limited core biopsy sample. Cholesterol granulomas, or “cholesterolomas,” may form within ducts and rupture, spilling contents into the surrounding stroma (Fig. 2F).

The differential diagnosis for duct ectasia includes cysts associated with fibrocystic change. Cysts are seen in terminal duct lobular units, in contrast to the subareolar duct involvement seen in duct ectasia. Making this distinction is not critical, as both lesions are typically managed conservatively. Juvenile papillomatosis is an uncommon localized lesion showing a constellation of histologic findings, including ductal stasis with luminal histiocytes. Florid ductal hyperplasia, apocrine metaplasia, papillary proliferations, and sclerosis are other characteristic findings in juvenile papillomatosis. Ductal hyperplasia and papillary epithelial hyperplasia are not seen in duct ectasia. Finally, duct ectasia occur because of duct obstruction due to an intraductal mass. If an intraductal mass is suspected and not represented in the core biopsy, excisional biopsy is necessary.

When duct ectasia is diagnosed in a screening core biopsy, excisional biopsy is not necessary, provided there is radiologic-pathologic concordance. Symptomatic cases of duct ectasia are treated by excision of the involved ducts. Duct ectasia is not associated with an elevated risk for breast carcinoma.

**GRANULOMATOUS LOBULAR MASTITIS**

Granulomatous mastitis have various etiologies, including infection (bacterial, fungal, mycobacterial), sarcoidosis, and other systemic granulomatous disease. After such causes are excluded, idiopathic “granulomatous lobular mastitis” describes a condition causing chronic, destructive non-necrotizing granulomatous inflammation of lobules. The etiology of granulomatous lobular mastitis is unknown, though some cases are associated with Co-
rynebacterium infection (described below). Oral contraceptive use, smoking, and autoimmune disease do not appear to predispose women to granulomatous lobular mastitis.19

Most cases of granulomatous lobular mastitis occur in women of reproductive age (20s–40s). Most women have been pregnant at least once prior to presentation, though the condition does not usually occur during pregnancy or lactation.20-23 Patients present with a unilateral palpable mass that is often accompanied by skin or nipple retraction and pain. Symptoms may be accompanied by axillary lymphadenopathy.19,21,22 On imaging, granulomatous lobular mastitis is often suggestive of malignancy. A spiculated mass or multiple nodular masses may be seen on mammography.22,24,25 Ultrasound often shows an irregular hypoechoic mass, fluid collection, tubular structures, or parenchymal mixed echogenicity.21,22,24,26

Microscopically, granulomatous lobular mastitis is characterized by non-necrotizing granulomas concentrated in lobules. The granulomas contain epithelioid histiocytes, Langhans giant cells, and lymphoplasmacytic inflammation (Fig. 3A, B). Neutrophilic microabscesses may also be seen. Granulomatous lobular mastitis may be complicated by frank abscess formation and draining skin sinuses. Cystic vacuoles, representing dissolved lipid, are often present within the granulomas, and can be lined by neutrophils; this has been termed “cystic neutrophilic granulomatous mastitis” (Fig. 3C).17-20 In such cases, gram-positive bacilli representing Corynebacterium can be seen within the cystic vacuoles (Fig. 3D). The bacteria show “coryneform” features, such as arrangement into palisades and “V” shapes, as well as clubbing of the organisms. These bacteria are not readily identifiable on hematoxylin and eosin examination, and in most cases only rare (<10) bacteria may be present in one or two vacuoles in one core biopsy sample. In fact, gram stains and microbial cultures of these samples are often negative in these cases, in part due to the fastidious nature of these organisms.27 We recently reported on a series of twelve patients with histologically identified cystic neutrophilic granulomatous mastitis.29 All patients presented with a unilateral breast mass that was painful in six of twelve cases. Imaging was either suspicious (BI-RADS 4) or highly suggestive of malignancy (BI-RADS 5) in over half of the studied cases. Gram-positive bacilli were identified in five of twelve cases, and all microbial cultures were negative for bacterial growth. Patients showed a variable response to treatment, with time to resolution of symptoms ranging from two weeks to six months.

Granulomatous lobular mastitis is often initially encountered in a core biopsy sample, with the differential including other
causes of granulomatous lobular inflammation. Special stains should be performed to rule out infection with fungi and acid-fast bacilli as the cause of the granulomatous process. Sarcoidosis should be excluded based on clinical, imaging, and laboratory findings. Sarcoidosis involvement of the breast is uncommon, and rarely the initial site of disease detection. Sarcoid granulomas tend to show less inflammation and are not associated with abscess or microabscess formation, as in granulomatous lobular mastitis. Schaumann bodies and asteroid bodies may be identified within granulomas. In cases where cystic neutrophilic granulomatous mastitis is seen, gram stains should be performed to identify gram-positive bacilli. At our institution, when this pattern is seen on biopsy, we include a note in the pathology report stating its known association with *Corynebacterium* infection, in order to help guide treatment. Microbial cultures should be obtained in all cases of granulomatous lobular mastitis to rule out infection.

Patients with granulomatous lobular mastitis are typically treated using a combination of antibiotics and surgery. Steroids have also been shown to be effective when added to the treatment regimen or used alone.22,25,26,30,35 Despite various treatments options, patients often experience persistent and recurrent disease, with complications including draining sinuses and abscess formation. Patients often have to undergo multiple surgical procedures and courses of antibiotics.

**DIABETIC MASTOPATHY**

Diabetic mastopathy, also known as "lymphocytic mastopathy" or "sclerosing lymphocytic lobulitis" is an uncommon mass-forming lesion seen in patients with insulin-dependent (type 1) diabetes mellitus, particularly in those who have long-standing disease with microvascular complications.32-34 The characteristic histologic findings seen in diabetic mastopathy can also be seen in lesions of patients with type 2 diabetes mellitus, autoimmune diseases such as Hashimoto’s thyroiditis, and even those with no history of diabetes or autoimmune disease.34,35 The cause of diabetic mastopathy is not known. One theory is that hyperglycemia leads to stromal matrix expansion and accumulation of advanced glycosylation end products, leading ultimately to an inflammatory B-cell response.32 It has also been suggested that diabetic mastopathy develops as a result of an immunologic re-

---

**Fig. 3.** Granulomatous lobular mastitis. (A, B) Non-necrotizing granulomas are centered within lobules. Granulomas contain Langhans giant cells, and are associated with lymphocytes and plasma cells. (C) Cystic neutrophilic granulomatous mastitis showing neutrophil-lined cysts within granulomas. (D) Gram-positive coryneform bacilli are present within the cysts.
response to exogenous insulin; however, this is unlikely the sole cause, as the lesion also develops in patients who have not taken exogenous insulin.36

Diabetic mastopathy most often occurs in premenopausal women, although rare cases have been reported in men.32,34,35,37 The typical presentation is a palpable unilateral mass, though screening mammography may detect the first changes. In some instances, multiple masses or ill-defined nodules are clinically detectable. The mammographic and sonographic features of diabetic mastopathy may be suspicious for malignancy: mammography may reveal an ill-defined mass, distortion, or dense glandular breast tissue;38 ultrasound may show an irregular hypoechoic mass with posterior shadowing.38-40 MRI shows non-specific enhancement.40-42

Tomaszewski et al.32 were the first to put forth criteria for the microscopic diagnosis of diabetic mastopathy. The characteristic constellation of findings includes lymphocytic lobulitis and ductitis, lymphocytic perivascularitis, and stromal fibrosis with epithelioid fibroblasts.32 Lymphocytic infiltrates, which can be fairly dense, surround ducts, lobules, and small vessels, and may sometimes be associated with plasma cells (Fig. 4A–C). Immunohistochemical characterization of these infiltrates reveals mature B-lymphocytes with a small population of T cells.43 Germinal centers are not typically seen here. Involved lobules may be atrophic or unremarkable. The stroma in diabetic mastopathy is dense and has a keloidal appearance. Intra-stromal epithelioid fibroblasts appear as plump cells with eosinophilic cytoplasm (Fig. 4B–D). Nuclei are oval to round with vesicular nuclei. Neither significant nuclear atypia nor mitotic figures are seen. The distribution of fibroblasts within the stroma can be heterogeneous, and show a whorled or nodular growth pattern.35 These distinctive fibroblasts are not present in all cases of diabetic mastopathy. In a series by Ely et al.,35 epithelioid fibroblasts were absent in five of 19 (26%) cases, including the two cases occurring in men. However, these fibroblasts were present in all three non-diabetic patients in their study.

The differential diagnosis for diabetic mastopathy as seen in core biopsy sample depends on which components of the lesion are present in the limited sample. If dense keloidal fibrosis is the predominant finding, fibrocystic change should be considered. It would be atypical for fibrosis to occur in diabetic mastopathy without coexisting perilobular or perivascular lymphocytic in-

Fig. 4. Diabetic mastopathy. (A) Lymphoid infiltrates surround ducts, lobules, and small vessels. The stroma has a hyalinized appearance. (B, C) Plump epithelioid fibroblasts are present in the stroma (C inset, high power). Perilobular (B) and perivascular (C) chronic inflammation is seen. (D) Fibroblasts in diabetic mastopathy compared with granular cell tumor (E) and multinucleated stromal giant cells (F).
filtered, unless the sample is quite limited. The most diagnostically relevant caveat would be to avoid misclassifying the epithelioid fibroblasts as neoplastic proliferations. Further, care should be taken to exclude carcinomas with abundant eosinophilic cytoplasm, such as pleomorphic lobular carcinomas that exhibit “histiocytoid” and/or apocrine features. These carcinomas, as in diabetic mastopathy, may not be associated with desmoplastic stroma; a broad-spectrum cytokeratin stain can be performed to rule out carcinoma in such cases. Granular cell tumors are composed of cells with abundant pink granular cytoplasm and bland nuclear features (Fig. 4E). These tumors stain positive for S100 and CD68, which helps to distinguish them from diabetic mastopathy. Multinucleated stromal giant cells, a benign incidental finding in breast tissue, can be distinguished from cells of diabetic mastopathy by their multiple hyperchromatic nuclei and scant, versus abundant, cytoplasm (Fig. 4F). Multinucleated stromal giant cells are incidental microscopic findings, while diabetic mastopathy is a mass-forming proliferation. Other mass-forming lesions with lymphoid infiltrates should be considered when lymphocytic ductitis, lobulitis, and perivasculitis are present in the absence of epithelioid fibroblasts. Lymphoma in the breast tends to diffusely infiltrate the stroma, a pattern of inflammation distinct from that seen in diabetic mastopathy. Additionally, immunostaining and molecular analysis will reveal a clonal proliferation of lymphocytes in lymphoma, but not in diabetic mastopathy.

Diabetic mastopathy is a benign condition, and patients can be managed with routine mammographic surveillance. In patients who have undergone excision of the lesion, recurrences may be ipsilateral or contralateral. In a series reported by Dorokhova et al., five of 34 cases (15%) of diabetic mastopathy recurred (2 ipsilateral, 3 bilateral). Patients with diabetic mastopathy are not at increased risk for subsequent development of breast carcinoma. Further, the lymphocytes in diabetic mastopathy do not show evidence of clonality by immunoglobulin heavy chain gene rearrangement studies, and patients also do not appear to be at risk for developing lymphoma. 

ABSCESS

Breast abscesses occur most commonly during lactation, but may also occur in the subareolar breast tissue of non-lactating breasts. Abscess formation is a consequence of ductal stasis in both lactational and non-lactational breasts. Though most patients with mastitis or breast abscess are treated with empiric antibiotics, core biopsy is indicated in some cases to rule out malignancy.

Lactational abscess can develop as a complication of mastitis, which occurs in up to 10% of lactating women, most commonly during the initiation and weaning phases of breastfeeding. The most common organism isolated from these abscesses is S. aureus, identified approximately 50% of the time. Primeter, prior lactational mastitis, and improper nursing technique are risk factors for the development of mastitis and abscesses. Cracking of the nipple skin may facilitate the entry of bacteria into the ductal system. In a series reported by Dener and Inan, 22 out of 128 patients (17%) with lactational abscess or mastitis had cracked nipples. Bacteria that enter the ducts are supplied with a lactose-rich environment from milk within the ductal lumen. Patients with lactational mastitis and abscess can present with redness, swelling, tenderness, or a palpable mass. Fever may additionally be present in some cases.

Subareolar, or non-puerperal abscesses, most often develop as a consequence of squamous metaplasia of lactiferous ducts. Squamous metaplasia of ducts leads to keratin accumulation with eventual rupture and spillage of duct contents into the surrounding stroma, which leads to abscess formation. This may be complicated abscess rupture and the formation of a sinus tract. The clinical scenario of recurring subareolar abscesses and sinus formation has been referred to as Zuska’s Disease or periductal mastitis. Patients with subareolar abscesses are typically premenopausal, though older women and men may also be affected. Cigarette smoking is a strong risk factor for the development of subareolar abscesses, with approximately 70%–90% of patients reporting a history of smoking. Diabetes and obesity are other associated risk factors. Patients present with a painful subareolar or periareolar mass, with or without nipple retraction. Nipple discharge, if present, may have a pasty consistency. Most cases are unilateral, but bilateral disease can also occur. In a series of 152 patients reported by Habif et al., 40 (26%) presented with bilateral abscesses. In contrast with lactational mastitis, the bacteria isolated from non-puerperal abscesses are most commonly mixed and predominantly anaerobic. In one study, coagulase-negative staphylococci was the most common aerobic organism isolated.

Mammographic features of abscesses are non-specific and may include a mass, architectural distortion, or skin thickening. Ultrasound shows a hypoechoic mass or a multiloculated fluid collection with a thick, echogenic rim. Microscopically, an abscess shows a mixed inflammatory infiltrate composed mainly of neutrophils in breast tissue, which may additionally show lactational changes in a lactational abscess.
In abscess resolution, neutrophilic inflammation is replaced by granulation tissue and chronic inflammatory changes. In some cases, subareolar abscesses may show dilated ducts that contain squamous metaplasia; a foreign body giant cell reaction to keratin may also be seen within the abscess. The differential diagnosis for breast abscess includes duct ectasia and granulomatous lobular mastitis, both of which can be associated with abscess formation.

Lactational mastitis can be adequately treated with antibiotics, though surgical drainage may be necessary in unresponsive cases. Nursing should be continued throughout treatment, and may in fact help resolve the infection. Treatment of subareolar abscess involves a combination of antibiotics and surgery, involving excision of the abscess and adjacent lactiferous duct. Significantly lower rates of recurrence are seen when the infected duct is also excised, compared excision of abscess alone. Subareolar abscesses are often chronic and recurring, with longer time to resolution and higher rates of recurrence, when compared to lactational abscesses.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

REFERENCES

1. Clarke D, Curtis JL, Martinez A, Fajardo L, Goffinet D. Fat necrosis of the breast simulating recurrent carcinoma after primary radiotherapy in the management of early stage breast carcinoma. Cancer 1983; 52: 442-5.
2. Rivera R, Smith-Bronstein V, Villegas-Mendez S, et al. Mammographic findings after intraoperative radiotherapy of the breast. Radiol Res Pract 2012; 2012: 758371.
3. Shah C, Badiyan S, Ben Wilkinson J, et al. Treatment efficacy with accelerated partial breast irradiation (APBI): final analysis of the American Society of Breast Surgeons MammoSite(R) breast brachytherapy registry trial. Ann Surg Oncol 2013; 20: 3279-85.
4. Paryani NN, Vallow L, Magalhaes W, et al. The incidence of fat necrosis in balloon-based breast brachytherapy. J Contemp Brachytherapy 2015; 7: 29-34.
5. Aqel NM, Howard A, Collier DS. Fat necrosis of the breast: a cytological and clinical study. Breast 2001; 10: 342-5.
6. Hogge JP, Robinson RE, Magnant CM, Zuurbier RA. The mammographic spectrum of fat necrosis of the breast. Radiographics 1995; 15: 1347-56.
7. Taboada JL, Stephens TW, Krishnamurthy S, Brandt KR, Whitman GJ. The many faces of fat necrosis in the breast. AJR Am J Roentgenol 2009; 192: 815-25.
8. Bilgen IG, Ustun EE, Memis A. Fat necrosis of the breast: clinical, mammographic and sonographic features. Eur J Radiol 2001; 39: 92-9.
9. Atasoy MM, Oren NC, Ilica AT, Guvenç I, Günsel A, Mossa-Basha M. Sonography of fat necrosis of the breast: correlation with mammography and MR imaging. J Clin Ultrasound 2013; 41: 415-23.
10. Soo MS, Kornguth PJ, Hertzberg BS. Fat necrosis in the breast: sonographic features. Radiology 1998; 206: 261-9.
11. Morkowski JJ, Nguyen CV, Lin P, et al. Rosai-Dorfman disease confined to the breast. Ann Diagn Pathol 2010; 14: 81-7.
12. Guo S, Yan Q, Rohr J, Wang Y, Fan L, Wang Z. Erdheim-Chester disease involving the breast: a rare but important differential diagnosis. Hum Pathol 2015; 46: 159-64.
13. Rahal RM, de Freitas-Junior R, Paulinelli RR. Risk factors for duct ectasia. Breast 2005; 11: 262-5.
14. Thomas WG, Williamson RC, Davies JD, Webb AJ. The clinical syndrome of mammary duct ectasia. Breast J 2005; 11: 262-5.
15. Dixon JM. Periductal mastitis/duct ectasia. World J Surg 1989; 13: 715-20.
16. Sweeney DJ, Wylie EJ. Mammographic appearances of mammary duct ectasia that mimic carcinoma in a screening programme. Australas Radiol 1995; 39: 18-23.
17. An HY, Kim KS, Yu IK, Kim KW, Kim HH. Image presentation. The nipple-areolar complex: a pictorial review of common and uncommon conditions. J Ultrasound Med 2010; 29: 949-62.
18. Davies JD. Pigmented periductal cells (ochrocytes) in mammary dysplasias: their nature and significance. J Pathol 1974; 114: 205-16.
19. Oran ES¸, Gürdal SÖ, Yankol Y, et al. Management of idiopathic granulomatous mastitis diagnosed by core biopsy: a retrospective multicenter study. Breast J 2013; 19: 411-8.
20. Going JJ, Anderson TJ, Wilkinson S, Chetty U. Granulomatous lobular mastitis. J Clin Pathol 1987; 40: 535-40.
21. Ocal K, Dag A, Turkmenoglu O, Kara T, Seyit H, Konca K. Granulomatous mastitis: clinical, pathological features, and management. Breast J 2010; 16: 176-82.
22. Hovanessian Larsen LJ, Peyvandi B, Klipfel N, Grant E, Iyengar G. Granulomatous lobular mastitis: imaging, diagnosis, and treatment. AJR Am J Roentgenol 2009; 193: 574-81.
23. Pandey TS, Mackinnon JC, Bressler L, Millar A, Marcus EE, Ganschow PS. Idiopathic granulomatous mastitis: a prospective study of 49 women and treatment outcomes with steroid therapy. Breast J 2014; 20: 258-66.
24. Ozturk M, Mavili E, Kahriman G, Akcan AC, Ozurk F. Granulomatous mastitis: radiological findings. Acta Radiol 2007; 48: 150-5.
25. Al-Khawari HA, Al-Manfoohi HA, Madda JP, Kovacs A, Sheikh M,
26. Joseph KA, Luu X, Mor A. Granulomatous mastitis: a New York public hospital experience. Ann Surg Oncol 2014; 21: 4159-63.
27. Renshaw AA, Derhagopian RP, Gould EW. Cystic neutrophilic granulomatous mastitis: an underappreciated pattern strongly associated with gram-positive bacilli. Am J Clin Pathol 2011; 136: 424-7.
28. Taylor GB, Paviour SD, Musaad S, Jones WO, Holland DJ. A clinicopathological review of 34 cases of inflammatory breast disease showing an association between corynebacteria infection and granulomatous mastitis. Pathology 2003; 35: 109-19.
29. D’Alfonso T, Moo TA, Arleo E, Cheng E, Antonio L, Hoda S. Cystic neutrophilic granulomatous lobular mastitis: further clinical and pathological characterization of an under-recognized entity based on eleven cases. Mod Pathol 2015; 28 Suppl 2: 40A-41A.
30. Jorgensen MB, Nielsen DM. Diagnosis and treatment of granulomatous mastitis. Am J Med 1992; 93: 97-101.
31. Akbulut S, Yilmaz D, Bakir S. Methotrexate in the management of idiopathic granulomatous mastitis: review of 108 published cases and report of four cases. Breast J 2011; 17: 661-8.
32. Tomaszewski JE, Brooks JS, Hicks D, Livolsi VA. Diabetic mastopathy: a distinctive clinicopathologic entity. Hum Pathol 1992; 23: 780-6.
33. Camuto PM, Zetrenne E, Ponn T. Diabetic mastopathy: a report of 5 cases and a review of the literature. Arch Surg 2000; 135: 1190-3.
34. Ely KA, Tse G, Simpson JF, Clarfeld R, Page DL. Diabetic mastopathy: a clinicopathologic review. Am J Clin Pathol 2000; 113: 541-5.
35. Seidman JD, Schnaper LA, Phillips LE. Mastopathy in insulin-requiring diabetes mellitus. Hum Pathol 1994; 25: 819-24.
36. Logan WW, Hoffman NY. Diabetic fibrous breast disease. Radiology 1989; 172: 667-70.
37. Moschetta M, Telegrafo M, Triggiani V, et al. Diabetic mastopathy: a diagnostic challenge in breast sonography. J Clin Ultrasound 2015; 43: 113-7.
38. Chan CL, Ho RS, Shek TW, Kwong A. Diabetic mastopathy. Breast J 2013; 19: 533-8.
39. Dorrkhova O, Fineberg S, Koenigsberg T, Wang Y. Diabetic mastopathy, a clinicopathological correlation of 34 cases. Pathol Int 2012; 62: 660-4.
40. Tuncbilek N, Karakas HM, Okten O. Diabetic fibrous mastopathy: dynamic contrast-enhanced magnetic resonance imaging findings. Breast J 2004; 10: 359-62.