Elevated Inflammatory Plasma Biomarkers in Patients With Fabry Disease: A Critical Link to Heart Failure With Preserved Ejection Fraction

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Background—Because systemic inflammation and endothelial dysfunction lead to heart failure with preserved ejection fraction, we characterized plasma levels of inflammatory and cardiac remodeling biomarkers in patients with Fabry disease (FD).

Methods and Results—Plasma biomarkers were studied in multicenter cohorts of patients with FD (n=68) and healthy controls (n=40). Plasma levels of the following markers of inflammation and cardiac remodeling were determined: tumor necrosis factor (TNF), TNF receptor 1 (TNFR1) and 2 (TNFR2), interleukin-6, matrix metalloprotease-2 (MMP-2), MMP-8, MMP-9, galectin-1, galectin-3, B-type natriuretic peptide (BNP), midregional pro–atrial natriuretic peptide (MR-proANP), and globotriaosylsphingosine. Clinical profile, cardiac magnetic resonance imaging, and echocardiogram were reviewed and correlated with biomarkers. Patients with FD had elevated plasma levels of BNP, MR-proANP, MMP-2, MMP-9, TNF, TNFR1, TNFR2, interleukin-6, galectin-1, globotriaosylsphingosine, and analogues. Plasma TNF levels correlated with lower BNP, MR-proANP, and MMP-2 levels. Patients with late gadolinium enhancement on cardiac magnetic resonance imaging had greater levels of BNP, MR-proANP, TNFR1, TNFR2, and MMP-2. Plasma BNP, MR-proANP, MMP-2, MMP-8, TNF, TNFR1, TNFR2, galectin-1, and galectin-3 were elevated in patients with renal dysfunction. Patients undergoing enzyme replacement therapy who have more severe disease had higher MMP-2, TNF, TNFR1, TNFR2, and globotriaosylsphingosine analogue levels.

Conclusions—Inflammatory and cardiac remodeling biomarkers are elevated in FD patients and correlate with disease progression. These features are consistent with a phenotype dominated by heart failure with preserved ejection fraction and suggest a key pathogenic role of systemic inflammation in FD. (J Am Heart Assoc. 2018;7:e009098. DOI: 10.1161/JAHA.118.009098.)

Key Words: biomarkers • Fabry disease • heart failure with preserved ejection fraction • hypertrophy • inflammation

Fabry disease (FD, Online Mendelian Inheritance in Man [OMIM] #300644) is an X-linked lysosomal storage disorder characterized by diminished or absent α-galactosidase A (EC 3.2.1.22) enzyme activity, leading to the accumulation of the glycosphingolipid globotriaosylceramide (Gb₃) in tissues. Recent neonatal screening data suggest that the actual prevalence is close to 1:3000. Because Fabry disease is X-linked, hemizygous males typically have much lower α-galactosidase A activity than heterozygous females. However, the majority of female heterozygotes develop clinically significant disease, albeit with a milder disease course than hemizygous men, likely due to X-chromosome inactivation. Cardiac manifestations of FD include left ventricular hypertrophy (LVH), diastolic dysfunction, microvascular angina, valvular abnormalities, and conduction defects, whereas proteinuria and progression to end-stage renal disease are renal complications of FD. Patients suffering from the classic phenotype of FD typically have early-onset symptoms with noticeable cardiovascular effects between 30 and 40 years of age, ultimately suffering from heart failure with preserved ejection fraction (HFpEF).
Cardiomyopathy with concentric hypertrophy and diastolic dysfunction is now the most common cause of death in patients with FD.13 FD variants are characterized by the presence of certain GLA gene mutations and are known as cardiac or renal variant phenotypes.2 Enzyme-replacement therapy (ERT) slows the progression of disease including the development of LVH.7,14 Plasma biomarkers are a rapidly growing area of research in FD and can provide prognostic value and insight into the pathophysiology of the disease.15,16 These cytokines have a significant role in cardiac disease, particularly with respect to myocardial remodeling17 and chronic heart failure.18,19 We report on the plasma levels of a variety of biomarkers in adults with FD compared with healthy controls to gain insight into the pathophysiology and burden of disease of this condition as well as its link to the HfPEF phenotype.

Methods

Ethics and Transparency Statement

All subjects gave written informed consent, and ethics approval was obtained from the University of Alberta (Edmonton, Alberta, Canada), University of Calgary (Calgary, Alberta, Canada), and QE II Health Sciences Centre (Halifax, Nova Scotia, Canada). Due to proprietary techniques used in certain portions of the data analysis section, analytic methods and study materials will not be made available to other researchers for purposes of reproducing the results or replicating the procedure. Clinicians and researchers are invited to contact the authors for the purposes of data replication or to share biomarkers samples as part of a collaborative effort.

Clinical Perspective

What Is New?

- Inflammatory biomarkers are elevated in patients with Fabry disease.
- Markers of disease progression, including kidney disease, left-ventricular hypertrophy, and myocardial fibrosis, are associated with higher levels of inflammation and markers of adverse cardiac remodeling.

What Are the Clinical Implications?

- Systemic inflammation likely plays a pathogenic role in Fabry disease and leads to a phenotype dominated by heart failure with preserved ejection fraction.
- Targeting inflammatory pathways may have a therapeutic role in Fabry disease.

Patient Population

Fabry disease patients (n=68) were recruited through Canadian metabolic clinics in Edmonton, Calgary, and Halifax. Healthy controls (n=40) with similar mean age and sex with no significant medical conditions were recruited through community outreach. Inclusion criteria for healthy controls included no history of cardiovascular disease, hypertension, diabetes mellitus, or renal disease; and no prescriptions for angiotensin-converting enzyme inhibitors, angiotensin-receptor blockers, β-blockers, digoxin, mineralocorticoid-receptor antagonists, thiazide diuretics, or loop diuretics. Enzyme-replacement therapy with either agalsidase-α (Replagal, Shire, Dublin, Ireland) or agalsidase-β (Fabrazyme, Sanofi-Genzyme, Cambridge, MA) in standard dose was given to patients who qualified under the Canadian Fabry Disease Initiative treatment guidelines.20

Baseline Analyses

Demographic information including date of birth, sex, height, and weight were collected. Clinical data including genetic mutation analysis, plasma leukocyte α-galactosidase activity, duration of treatment, serum creatinine, cardiac imaging, and Mainz Severity Score Index (MSSI)21 data were also recorded for FD patients. Estimated glomerular filtration rate was calculated using the Modification of Diet in Renal Disease equation.22 A cut-off estimated glomerular filtration rate of 60 mL/(min·1.73 m²) was used for assigning kidney disease. LVH was defined using cardiac magnetic resonance imaging (MRI), as described below.

Cardiac Imaging

Transthoracic echocardiography was performed for the Fabry disease cohort as described previously.23,24 Briefly, echocardiography was performed using standard techniques25 on commercial ultrasound equipment (M3 S Probe, Vivid 7; GE Vingmed Ultrasound AS, Horten, Norway). Chamber quantification and ejection fraction (EF) were assessed using the modified Simpson method.26 Preserved EF was defined as EF ≥50% as assessed by echocardiography. The presence of diastolic dysfunction was graded using the (peak mitral inflow during passive filling in early diastole)/(peak mitral inflow during active filling in atrial systole) ratio, peak mitral inflow velocity during passive filling in early diastole, (peak mitral inflow during passive filling in early diastole)/(mitral annular velocity during early diastole) ratio, and left atrial maximum volume index according to current guidelines,27 unless assessment was judged to be indeterminate or precluded by arrhythmia or patient factors.

Cardiac MRI was performed as previously described in a standardized fashion.24,28-30 Cardiac MRI was performed on
1.5-T Siemens Sonata or Avanto scanners (Siemens Medical Solutions, Erlangen, Germany). Typical imaging parameters using standard balanced steady-state–free precession short-axis and long-axis cines were 1.24 milliseconds echo time, 2.89 milliseconds repetition time, 51° flip angle, 360 × 270-mm field of view, 8-mm slice thickness, and 2-mm gap between short-axis slices; 10 to 14 views per segment, reconstructed to 30 phases per cardiac cycle were used. Left-ventricular mass index was measured in end-diastole and was calculated using a modified method of disks measured from steady-state free precession cines and analyzed using software (MATLAB 2010a; The MathWorks, Natick, MA) as previously described. Assessment of LVH was performed as previously defined using cutoffs of left ventricular mass index ≥ 85 g/m² in men and ≥ 81 g/m² in women to denote LVH. Papillary muscles were included as part of the myocardium for left ventricular mass calculations but excluded for volume assessment. Conventional late gadolinium enhancement (LGE) imaging was performed 7 minutes after contrast injection using a phase-sensitive inversion recovery sequence in the short-axis, 2-, 3-, and 4-chamber views to match the cine slice locations. LGE imaging was not performed in 3 patients due to advanced renal disease precluding the administration of gadolinium contrast.

Due to the specialized nature of the data acquisition and analysis, T1 cardiac resonance mapping and extracellular volume calculations were performed only in the Alberta cohort. T1 mapping used the Saturation-Recovery Single-Shot Acquisition pulse sequence as previously described. T1 mapping was performed at baseline and 15 minutes after administration of 0.15 mmol/kg of gadopentetate dimeglumine (Magnevist; Bayer Inc, Toronto, Ontario, Canada) as previously described. Endocardial and epicardial tracings were created for T1 analysis. Blood pool and myocardial T1 analysis were used based on a circular region of interest drawn in the left ventricular blood pool and a 2-mm-wide region of interest drawn over the interventricular septum, respectively. Normal left-ventricular T1 values at our site are 1167 ± 36 milliseconds (baseline) and 600 ± 38 milliseconds (postcontrast) (n = 30) in men and 1202 ± 30 milliseconds (baseline) and 539 ± 46 (postcontrast) (n = 30) in women. In each of the 18 segments the extracellular volume fraction, which is the volume in which gadolinium contrast agent is distributed, was estimated using the calculated concentrations of contrast agent in the blood and tissue.

**Sample Collection and Processing**

While patients were sitting and rested, whole blood for plasma analysis was collected into lithium-heparin and EDTA tubes and stored immediately on ice. Subsequently, plasma fractionation was completed, and the samples were stored in liquid nitrogen at the Canadian BioSample Repository (Edmonton, Alberta, Canada).

**Classical Plasma Biomarker Quantification**

Enzyme-linked immunosorbent assays were used to investigate plasma levels of tumor necrosis factor (TNF), NTF receptors (TNFR1, TNFR2), interleukin (IL)-6, matrix metalloprotease (MMP)-2, MMP-8, MMP-9, galectin-1, and galectin-3. Plasma B-type natriuretic peptide (BNP) and midregional pro–atrial natriuretic peptide (MR-proANP) levels were assessed as previously described using an Alere Triage reagent pack (Alere Inc, Ottawa, Ontario, Canada) and analyzed using an automated Dxl 800 immunoanalyzer (Beckman-Coulter, Fullerton, CA) at provincial health laboratories in the province of Alberta, Canada. Plasma α-galactosidase activity was assessed as previously described. Plasma C-reactive protein levels were measured using high-sensitivity kits at provincial health laboratories in the province of Alberta, Canada. Commercial enzyme-linked immunosorbent assays kits were used to assay plasma levels of TNF, TNFR1, TNFR2, and IL-6 (catalogue no. HSTA00D, SRT100, SRT200, and HS600B respectively; R&D Systems, Minneapolis, MN) as previously described. The described kit protocol was used for enzyme-linked immunosorbent assays for total MMP-2, MMP-8, MMP-9, galectin-1, and galectin-3 (catalogue no. MMP200, DMP800, DMP900, DGAL10, and DGAL30, respectively; R&D Systems, Minneapolis, MN). Absorbance was measured using a SpectraMax M5 Plate Reader (Molecular Devices, San Jose, CA) at 450 nm for all assays with the wavelength correction set to 540 nm for TNFR1, TNFR2, MMP-2, MMP-8, MMP-9, galectin-1, and galectin-3 and to 650 nm for TNF and IL-6. Detection rates for the enzyme-linked immunosorbent assays were 100% for all assays. The intra-assay coefficients of variation were 3.7% (n = 8), 5.2% (n = 8), 3.5% (n = 8), 3.6% (n = 8), 11.4% (n = 8), 13.4% (n = 8), 4.6% (n = 8), 7.9% (n = 8), and 2.1% (n = 8) for TNF, TNFR1, TNFR2, IL-6, MMP-2, MMP-8, MMP-9, galectin-1, and galectin-3 assays, respectively.

**Analysis of Plasma Lyso-Gb3 and Analogues**

Plasma lysoglubotriosylceramide (Lyso-Gb3) and its 6 related analogues with modified sphingosine moieties (−C2H4; H2; O; H2O; H2O2; H2O3) were analyzed in the plasma of Fabry patients with a method previously published by Boutin and Auray-Blais (Figure 1). Briefly, 100 μL of plasma was spiked with in-house–synthesized N-glycinated Lyso-Gb3 (Lyso-Gb3-Gly) as the internal standard and purified by solid-phase extraction using mixed-mode
cation-exchange cartridges (Oasis MCX, 30 mg, 60 μm; Waters Corp, Milford, MA). The samples were separated by ultraperformance liquid chromatography using an Acquity I-Class (Waters) system equipped with a BEH C18 column (2.1×50 mm, particle diameter 1.7 μm; Waters). The analysis of Lyso-Gb3 and its 6 analogues was performed by tandem mass spectrometry using the multiple reaction-monitoring mode on a Xevo TQ-S mass spectrometer (Waters). Positive electrospray was used for the ionization. Plasma total Lyso-Gb3 was reported as the sum of Lyso-Gb3 and its 6 analogues.

Statistical Analysis
Statistical analyses were carried out using IBM SPSS Statistics version 20 for Windows (SPSS Inc, Chicago, IL). Discrete variables are presented as count and/or percentage. Continuous variables with normal distributions are presented as mean±SD, whereas continuous variables with skewed distributions, including all biomarkers, are presented as median (first quartile, third quartile) unless otherwise indicated. A P-value of <0.05 was considered statistically significant. Categorical data were compared using Pearson Chi-squared tests or the Fisher Exact Test, where appropriate. Pairwise comparisons were evaluated using Mann-Whitney U Tests or Kruskal-Wallis Test with Mann-Whitney U Tests, where appropriate. Analyses of continuous covariates were performed using linear regression with the MSSI versus biomarker levels and left-atrial size versus MR-proANP. Outliers identified by visual analysis were tested for potential impacts on the regression analysis. Receiver-operator characteristic curve analysis was performed using a diagnosis of FD via α-galactosidase levels and/or genetic testing as the gold standard.

Figure 1. Examples of ion chromatograms for lysoglobotriaosylceramide (Lyso-Gb3), its 6 analogues, and Lyso-Gb3-Gly (used as the internal standard) detected in plasma from a Fabry patient. The (+H2O2) analogue has 2 structural isomers with retention times of 3.29 and 4.57 minutes. The areas of these peaks were added together for computation results. cps indicates counts per second.
Results

Clinical Characteristics

The mean age (±SD) of FD patients was 42±13 years (n=62) versus 46±12 years for the healthy controls (n=40), and there were equal numbers of women and men in the 2 cohorts (Table 1). The mean body-mass index was 25.0±2.7 kg/m² in the healthy controls versus 24.3±4.3 kg/m² for the FD cohort. Of the 68 FD patients, 41 patients (60%) had LVH by cardiac MRI criteria, and 37 (54%) were receiving ERT. Among FD patients, median plasma α-galactosidase A activity was 1.9 (0.63, 3.6) μmol/(h·g protein). By use of the MSSI,21 16 (24%) FD patients were classiﬁed as having mild disease (MSSI 0–11), 33 (50%) had moderate disease (MSSI 12–40), and 9 (13%) had severe disease (MSSI ≥41). The mean score of the cardiac subset of MSSI (maximum of 20) was 5.5±4.9. The estimated glomerular filtration rate in FD patients was 83±33 mL/(min·1.73 m²).

The cardiac phenotype of the Fabry cohort was characterized by hypertrophy, preserved EF, and diastolic dysfunction (Table 1).23,28 The high median left ventricular mass index is consistent with the relatively high prevalence of males in our FD cohort (50%). Increased postcontrast myocardial T1 values suggest increased myocardial fibrosis, and low precontrast T1 values are consistent with a diagnosis of FD.28 The mean EF

Table 1. Demographic Data and Cardiac Imaging Parameters for the FD Cohort

| Demographic and clinical information | Healthy Controls (n=40) | Fabry Disease (n=68) | Fabry Male (n=34) | Fabry Female (n=34) |
|-------------------------------------|------------------------|---------------------|-------------------|--------------------|
| **Age, y**                          | 46±12                  | 42±13               | 40±11             | 44±13              |
| **Sex (% female)**                  | 50                     | 50                  | 0                 | 100                |
| **BMI, kg/m²**                      | 25.0±2.7               | 24.3±4.3            | 24.3±3.9          | 24.4±4.8           |
| **eGFR, mL/(min·1.73 m²)**          | ...                    | 85.9±32.9           | 77.8±36.7         | 94.1±26.6          |
| **Echocardiography parameters**     |                        |                     |                   |                    |
| **LVEF, %**                         | ...                    | 62.6±8.4            | 59.9±9.9          | 66.4±3.3           |
| **End-diastolic thickness, mm**      | ...                    | 8.1±1.9             | 8.7±2.1           | 7.6±1.3            |
| **End-systolic thickness, mm**       | ...                    | 11.9±2.5            | 12.7±2.9          | 11.1±1.6           |
| **E-wave velocity, m/s**            | ...                    | 90.4±21.5           | 94.4±23.8         | 83.8±16.2          |
| **A-wave velocity, m/s**            | ...                    | 71.3±22.4           | 71.9±23.0         | 70.4±22.8          |
| **E/A ratio**                       | ...                    | 1.34±0.34           | 1.36±0.23         | 1.31±0.47          |
| **e' velocity, m/s**                | ...                    | 0.089±0.026         | 0.093±0.030       | 0.084±0.022        |
| **E/e' ratio**                      | ...                    | 11.1±4.7            | 11.8±5.8          | 10.2±2.7           |
| **LA volume index, mL/m²**          | ...                    | 29.3±9.7            | 30.9±10.4         | 27.0±8.4           |
| **Diastolic dysfunction, %**        |                        | 60                   | 56                | 69                 |
| **Grade I**                         | ...                    | 12                  | 11                | 13                 |
| **Grade II**                        | ...                    | 28                  | 33                | 19                 |
| **Grade III**                       | ...                    | 0                   | 0                 | 0                  |
| **Cardiac MRI parameters**          |                        |                     |                   |                    |
| **LVEDV, mL/m²**                    | ...                    | 81.6±16.4           | 92.1±16.0         | 71.6±9.1           |
| **LVEF, %**                         | ...                    | 30.3±8.9            | 33.8±10.1         | 27.1±6.3           |
| **LVMi, g/m²**                      | ...                    | 63.6±6.7            | 64.0±8.5          | 63.2±4.8           |
| **T₁ baseline, myo, ms (n=36)**     | ...                    | 78.5±21.6           | 91.4±8.5          | 66.4±12.7          |
| **T₁ postcontrast, myo, ms (n=33)** | ...                    | 1068±39             | 1041±36           | 1085±45            |
| **ECV, % (n=33)**                   | ...                    | 536±32              | 551±36            | 519±41             |

Demographic and imaging parameters are expressed as mean±SD. A indicates peak mitral inflow during active filling in atrial systole; BMI, body-mass index; E, peak mitral inflow during passive filling in early diastole; e’, mitral annular velocity during early diastole; ECV, extracellular volume; eGFR, estimated glomerular filtration rate (using the Modification of Diet in Renal Disease equation); FD, Fabry disease; LA, left atrial; LVEDV, left ventricular end-diastolic volume; LVEF, left ventricular ejection fraction; LVEF, left ventricular end-systolic volume; LVMi, left ventricular mass index; MRI, magnetic resonance imaging; myo, myocardium.
Table 2. List of Identified Mutations With Corresponding Phenotype and Selected Demographic and Clinical Information in FD Cohort (n=68)

| Mutation     | Phenotype       | Age, y (Sex) | ACE-I or ARB | Statin | ASA | ERT |
|--------------|-----------------|--------------|--------------|--------|-----|-----|
| A143P        | Classic ^38      | 15 (M)       | Yes          | No     | No  | No  |
| A143P        | Classic          | 19 (F)       | No           | No     | No  | No  |
| A143P        | Classic          | 26 (F)       | No           | No     | No  | No  |
| A143P        | Classic          | 28 (M)       | No           | No     | No  | No  |
| A143P        | Classic          | 29 (F)       | Yes          | Yes    | Yes | No  |
| A143P        | Classic          | 30 (M)       | No           | No     | Yes | Yes |
| A143P        | Classic          | 33 (M)       | No           | No     | Yes | Yes |
| A143P        | Classic          | 34 (F)       | No           | No     | Yes | No  |
| A143P        | Classic          | 34 (M)       | Yes          | No     | Yes | Yes |
| A143P        | Classic          | 35 (M)       | Yes          | Yes    | Yes | Yes |
| A143P        | Classic          | 35 (M)       | Yes          | Yes    | Yes | Yes |
| A143P        | Classic          | 35 (M)       | Yes          | Yes    | No  | Yes |
| A143P        | Classic          | 36 (F)       | Yes          | Yes    | Yes | Yes |
| A143P        | Classic          | 36 (F)       | No           | No     | No  | No  |
| A143P        | Classic          | 39 (F)       | No           | No     | Yes | No  |
| A143P        | Classic          | 40 (M)       | Yes          | Yes    | Yes | Yes |
| A143P        | Classic          | 45 (F)       | No           | No     | No  | No  |
| A143P        | Classic          | 46 (F)       | No           | No     | No  | No  |
| A143P        | Classic          | 47 (M)       | Yes          | No     | No  | Yes |
| A143P        | Classic          | 48 (M)       | No           | Yes    | No  | Yes |
| A143P        | Classic          | 48 (M)       | Yes          | No     | Yes | Yes |
| A143P        | Classic          | 51 (F)       | Yes          | No     | Yes | Yes |
| A143P        | Classic          | 51 (F)       | No           | Yes    | No  | No  |
| A143P        | Classic          | 51 (M)       | No           | Yes    | Yes | Yes |
| A143P        | Classic          | 52 (M)       | Yes          | Yes    | No  | Yes |
| A143P        | Classic          | 54 (F)       | Yes          | Yes    | Yes | No  |
| A143P        | Classic          | 55 (F)       | Yes          | Yes    | No  | Yes |
| A143P        | Classic          | 62 (M)       | Yes          | Yes    | No  | Yes |
| A143P        | Classic          | 66 (F)       | Yes          | No     | Yes | Yes |
| A143P        | Classic          | 68 (F)       | Yes          | Yes    | Yes | Yes |
| E338K        | Classic ^39      | 34 (F)       | Yes          | No     | Yes | No  |
| E338K        | Classic          | 36 (F)       | Yes          | No     | Yes | Yes |
| E338K        | Classic          | 62 (M)       | Yes          | Yes    | Yes | Yes |
| G261V        | Unclassified     | 36 (F)       | Yes          | Yes    | Yes | No  |
| G43V         | Classic ^30      | 35 (M)       | Yes          | Yes    | Yes | Yes |
| intron2: c.369+5G>T | Unclassified | 41 (M)       | Yes          | Yes    | Yes | No  |

Continued
via echocardiography was 63±8%, and 2 male FD patients had reduced EF. Excluding patients who were unable to be assessed for diastolic dysfunction due to arrhythmia or who were judged to be indeterminate, 60% had none, 12% had grade I (mild), 28% had grade II (moderate), and no patients had grade III (severe) diastolic dysfunction. The mean average (peak mitral inflow during passive filling in early diastole)/ (mitral annular velocity during early diastole) ratio was

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Table 2. Continued

| Mutation    | Phenotype             | Age, y (Sex) | ACE-I or ARB | Statin | ASA | ERT |
|-------------|-----------------------|--------------|--------------|--------|-----|-----|
| A143P (n=30) | Cardiac variant       | 26 (F)       | No           | No     | No  | No  |
| S345P (n=9)  | Cardiac variant       | 41 (M)       | Yes          | Yes    | Yes | No  |
| Y134S (n=4)  | Cardiac variant       | 66 (F)       | Yes          | Yes    | Yes | Yes |
| E338K (n=3)  | Cardiac variant       | 51 (F)       | Yes          | Yes    | Yes | Yes |
| N211S (n=3)  | Cardiac variant       | 35 (M)       | Yes          | Yes    | Yes | Yes |
| R112H (n=3)  | Cardiac variant       | 46 (F)       | Yes          | No     | Yes | No  |
| R227Q (n=3)  | Cardiac variant       | 55 (F)       | Yes          | No     | No  | Yes |
| Other (n=13) | Cardiac variant       | 25 (M)       | Yes          | No     | No  | No  |
| N211S Cardiac variant | 41 26 (F) | No | No | No | No |
| N211S Cardiac variant | 41 (M) | Yes | Yes | Yes | No |
| N211S Cardiac variant | 66 (F) | Yes | Yes | Yes | Yes |
| Q321E Classic | 42 51 (F) | Yes | Yes | Yes | Yes |
| Q386X Classic | 43 35 (M) | Yes | Yes | Yes | Yes |
| Q386X Classic | 46 (F) | Yes | No | Yes | No |
| R112C Classic | 42 51 (M) | No | Yes | Yes | Yes |
| R112H Renal variant | 44 25 (M) | Yes | No | No | No |
| R112H Renal variant | 48 (F) | Yes | Yes | Yes | No |
| R220X Classic | 43 29 (F) | No | No | No | No |
| R220X Classic | 55 (F) | Yes | Yes | Yes | Yes |
| R227Q Classic | 45 19 (F) | No | No | No | No |
| R227Q Classic | 40 (M) | Yes | Yes | Yes | Yes |
| R227Q Classic | 55 (F) | Yes | Yes | Yes | No |
| R227Q Classic | 52 (M) | Yes | Yes | Yes | Yes |
| S345P Classic | 45 15 (M) | No | No | No | No |
| S345P Classic | 28 (M) | Yes | No | Yes | No |
| S345P Classic | 33 (M) | Yes | Yes | Yes | Yes |
| S345P Classic | 39 (F) | Yes | Yes | Yes | No |
| S345P Classic | 45 (F) | Yes | Yes | Yes | No |
| S345P Classic | 47 (M) | Yes | Yes | Yes | Yes |
| S345P Classic | 50 (M) | Yes | Yes | Yes | Yes |
| S345P Classic | 51 (F) | Yes | Yes | Yes | No |
| S345P Classic | 54 (F) | Yes | Yes | Yes | No |
| V254Gfs Unclassified | 50 (M) | Yes | Yes | No | Yes |
| W349X Classic | 46 30 (M) | Yes | Yes | Yes | Yes |
| Y134S Classic | 43 34 (M) | Yes | No | No | Yes |
| Y134S Classic | 35 (M) | Yes | No | Yes | Yes |
| Y134S Classic | 39 (F) | Yes | No | Yes | No |
| Y134S Classic | 68 (F) | Yes | Yes | Yes | Yes |

ACE-I indicates angiotensin-converting enzyme inhibitor; ARB, angiotensin receptor blocker; ASA, acetylsalicylic acid; ERT, enzyme-replacement therapy; FD, Fabry disease.
11.1±4.7, and the mean left atrial maximum volume index was 29.3±9.7 mL/m².

**Figure 2.** Plasma levels of cardiac remodeling biomarkers and Lyso-Gb3 in cohorts of patients with FD (n=68) and healthy controls (n=40). Biomarkers BNP, MR-proANP, galectin-1, galectin-3, and Lyso-Gb3 are significantly elevated in the FD cohort relative to healthy controls. BNP indicates B-type natriuretic peptide; FD, Fabry disease; HC, healthy controls; Lyso-Gb3, lysoglobotriaosylceramide; MR-proANP, midregional pro–atrial natriuretic peptide. *P<0.05; **P<0.01; ***P<0.001.

Differences in Plasma Biomarkers Between FD and Healthy Cohorts

Plasma BNP and MR-proANP were elevated in patients with FD relative to healthy controls (P=0.006 and 0.013, respectively) (Figure 2, Table 3). Although there was no statistically significant difference between plasma MMP-8 values in the FD and healthy control cohorts (P=0.079), there were significant differences in MMP-2 and MMP-9 (P=0.017 and P<0.001, respectively) (Figure 3). Patients with FD had significantly elevated plasma levels of inflammatory markers TNF, TNFR1, and TNFR2 relative to healthy controls (P=0.008, P=0.003, and P<0.001, respectively) (Figure 3). There was no difference in C-reactive protein concentration between the FD patients and healthy control cohorts (P=0.839), but IL-6 was significantly elevated in FD patients (P=0.021). Galectin-1 was significantly elevated in the FD cohort when compared with healthy controls, whereas galectin-3 was not statistically different (P<0.001 and P=0.533, respectively) (Figure 2). TNFR2 and galectin-3 were found to have independent positive correlations with Lyso-Gb3 (P=0.020 and 0.024, respectively).

Receiver-operator characteristic curve analysis was performed for Lyso-Gb3 which performed extremely well (area under the curve=0.998) (Figure 4). As a biomarker, plasma total Lyso-Gb3 and analogues performed flawlessly for classic mutations (area under the curve=1.0 for both) but performed poorly for cardiac/renal variant mutations (area under the curve=0.41), although our sample size was low (n=6). In our cohort there was no additional utility to measuring Lyso-Gb3 analogues (area under the curve=0.994) instead of Lyso-Gb3 alone (Figure 4).

Cardiac and Renal Disease and Relationship With Plasma Biomarkers

In FD patients with LVH by cardiac MRI criteria (n=41, 60%), TNFR2, TNF, IL-6, MMP-2, and Lyso-Gb3 were significantly elevated (P=0.045, 0.025, 0.001, 0.046, and 0.002, respectively) (Figure 5). Patients with late gadolinium enhancement on cardiac MRI had greater levels of BNP, MR-proANP, TNFR1, TNFR2, and MMP-2 (P=0.001, P<0.001, P=0.014, P=0.014, and P<0.001 respectively) (Figure 6). Patients with diastolic dysfunction had elevated BNP, MR-proANP, and MMP-2 levels (P=0.002, 0.01, and 0.003, respectively) (Figure 7A). Maximal left-atrial size correlated with MR-proANP (P<0.0001) (Figure 7B). Regression analysis revealed that the disease burden assessed by the MSSI was positively associated with increased plasma MMP-9 (P=0.015), and the MSSI cardiac subset correlated with MMP-2 (P=0.003). Fabry patients with renal dysfunction (n=18, 26%) had higher levels of BNP, MR-proANP, TNF, TNFR1, TNFR2, MMP-2, MMP-8, galectin-1, and galectin-3 (P=0.001, P<0.001, P<0.001, P<0.001, P<0.001, P=0.03, P<0.001, and P=0.004, respectively) (Figure 8).

Medical Therapy and Biomarkers

Fabry patients who qualify for and receive ERT (n=37, 54%) had greater plasma levels of TNF, TNFR1, TNFR2, MMP-2,
Elevated Inflammatory Biomarkers in Fabry Disease  Yogasundaram et al

Table 3. Biomarker Data for the Fabry Disease Cohort

| Biomarkers         | Healthy Controls (n=40) | Fabry Disease (n=68) | Fabry Male (n=34) | Fabry Female (n=34) |
|--------------------|------------------------|----------------------|-------------------|---------------------|
| BNP, pg/mL         | 16.5 (11, 35.75)       | 34.5 (15, 79.25)     | 22 (10, 77.75)    | 39.5 (20, 85.5)    |
| MR-proANP, pmol/L  | 45.6 (33.0, 74.7)      | 65.6 (41.3, 119)     | 56.7 (39.5, 191)  | 77.7 (43.9, 117)   |
| TNFR1, pg/mL       | 829 (686, 939)         | 971 (696, 1726)      | 1153 (914, 1992)  | 890 (610, 1148)    |
| TNFR2, pg/mL       | 1350 (1217, 2124)      | 2730 (1507, 4471)    | 3608 (2014, 5492) | 2376 (1460, 3635)  |
| TNF, pg/mL         | 0.73 (0.49, 0.90)      | 0.90 (0.62, 1.53)    | 1.13 (0.71, 1.84) | 0.77 (0.55, 0.97)  |
| IL-6, pg/mL        | 1.05 (0.66, 1.85)      | 1.58 (0.97, 2.19)    | 1.58 (0.99, 2.18) | 1.58 (0.96, 2.21)  |
| MMP-2, ng/mL       | 204 (182, 229)         | 232 (180, 295)       | 256 (212, 348)    | 199 (174, 262)     |
| MMP-8, ng/mL       | 2.70 (0.91, 3.68)      | 2.97 (2.44, 3.36)    | 3.03 (2.45, 3.38) | 2.94 (2.09, 3.36)  |
| MMP-9, ng/mL       | 34.1 (25.8, 55.3)      | 58.7 (40.4, 78.0)    | 64.1 (41.8, 83.6) | 55.4 (40.1, 74.1)  |
| Galectin-1, ng/mL  | 16.7 (13.5, 21.7)      | 27.2 (21.0, 35.8)    | 25.5 (18.2, 38.8) | 28.9 (22.5, 35.0)  |
| Galectin-3, ng/mL  | 4.48 (3.33, 5.88)      | 4.08 (2.95, 5.85)    | 3.65 (2.81, 6.61) | 4.38 (2.95, 5.85)  |
| Lyso-Gb3, nmol/L   | 0.06 (0.25)            | 21.8 (10.3, 47.2)    | 47.1 (31.3, 70.8) | 11.2 (8.7, 18.3)   |

Biomarker data are reported as medians (25th percentile, 75th percentile). BNP indicates B-type natriuretic peptide; IL, interleukin; Lyso-Gb3, lysoglobotriaosylceramide; MMP, matrix metalloprotease; MR-proANP, midregional pro-atrial natriuretic peptide; TNF, tumor necrosis factor; TNFR, tumor necrosis factor receptor.

and Lyso-Gb3 (P=0.025, P=0.003, P<0.001, P<0.001, and P=0.001, respectively) (Figure 9). Among patients who were prescribed either an angiotensin-converting enzyme inhibitor or angiotensin receptor blocker (n=50, 74%), plasma MMP-2 and MMP-8 were elevated (P=0.027 and 0.015, respectively). Patients prescribed a statin (n=39, 57%) had significantly increased plasma levels of BNP, MR-proANP, galectin-1, and galectin-3 (P=0.007, 0.001, 0.023, and 0.001, respectively), whereas patients prescribed aspirin (n=46, 68%) had significantly elevated plasma levels of BNP, MR-proANP, and MMP-8 (P=0.008, P=0.001, and P<0.001, respectively).

Demographics and Biomarkers

Among patients with FD, TNF, TNFR1, TNFR2, MMP-2, and Lyso-Gb3 were elevated in male (n=34, 50%) relative to female patients (P=0.003, P=0.002, P=0.035, P=0.012, and P=0.001, respectively) (Figure 10). Older FD patients (n=34, 50%) over the median age (45.5 years) had higher levels of BNP, MR-proANP, MMP-2, and galectin-3, relative to younger patients with FD (P=0.002, 0.012, 0.038, and 0.011, respectively), but Lyso-Gb3 levels were lower in these older patients (P=0.012). There were no significant differences in biomarkers between overweight or obese (n=36, 53%; body-mass index ≥25.0 kg/m²) and normal–body-mass index FD patients.

Genotype and Phenotype and Plasma Biomarkers

Galectin-3 concentrations differed significantly (P=0.03) between the type of mutation, missense (n=6, 9.1%) versus nonsense (n=60, 90.9%), but no other statistically significant differences were found for other biomarkers. Plasma concentrations of total Lyso-Gb3, Lyso-Gb3, and its analogues were significantly greater in patients with classic phenotypes (n=59, 90.8%; unclassified excluded) compared with those with cardiac/renal variant phenotypes (n=6, 9.2%), but no other significant differences were noted with other biomarkers (Table 4).

Discussion

Proteomic biomarker discovery platforms have revealed several altered pathways in FD including vascular dysfunction, oxidative stress, and cytoskeletal remodeling. Proteomic biomarker discovery platforms have revealed several altered pathways in FD including vascular dysfunction, oxidative stress, and cytoskeletal remodeling.15,16 Our study used a directed approach in which inflammatory and cardiac remodeling biomarkers were analyzed from the plasma of healthy controls and FD patients. The elevation of the inflammatory markers TNF, IL-6, TNFR1, and TNFR2 in FD patients strongly implicates chronic inflammation as a major driver in the pathogenesis of FD. Mechanistically, glycolipids, including Lyso-Gb3, bind to toll-like receptor 4, activating nuclear factor κB and T lymphocytes and subsequent production of proinflammatory cytokines, leading to a chronic inflammatory state and associated vasculopathy.47-49 Both TNF and IL-6 plasma levels are elevated in chronic heart failure and correlate with a decreasing functional status of these patients as well as all-cause mortality.50-52 Furthermore, the positive correlation of inflammatory biomarkers in FD patients with higher disease burden based on MSSI scores, cardiac-specific MSSI scores,
Figure 3. Plasma levels of inflammatory biomarkers and selected matrix metalloproteases in cohorts of FD (n=68) and healthy controls (n=40). TNF, IL-6, TNFR1, and TNFR2 are significantly elevated in the FD cohort relative to healthy controls, without significant elevation in CRP. In addition, MMP-2 and MMP-9 are also significantly elevated in the FD cohort relative to healthy controls, although no difference was observed for MMP-8. CRP, C-reactive peptide; FD, Fabry disease; HC, healthy controls; IL, interleukin; MMP, matrix metalloprotease; TNF, tumor necrosis factor; TNFR, TNF receptor. *P<0.05; **P<0.01; ***P<0.001.
LVH, LGE, and renal dysfunction suggests that systemic inflammation plays a central role in the morbidity and mortality associated with FD. These findings are further supported by the relatively high prevalence of diastolic dysfunction in our cohort. Vascular dysregulation in FD may also affect the coronary arteries, leading to microvascular angina and subsequent HFpEF. These findings support the evolving paradigm of inflammation and vascular dysfunction as key pathogenic processes in HFpEF.

Fabry patients develop significant renal dysfunction, and the strong association of elevated inflammatory markers and worsened renal function observed in our cohort is especially relevant given the connection between HFpEF and renal disease. Of particular interest to FD patients, biomarker panels may, in the future, help identify HFpEF phenotypes to guide appropriate phenotype-specific therapy.

Two different receptor subtypes of TNF, TNFR1 and TNFR2, mediate signaling pathways that have opposing effects on the heart. TNFR1-mediated signaling appears to be the primary cause of deleterious effects of TNF in the heart, including increased oxidative stress and cardiomyocyte apoptosis. In contrast, TNFR2-mediated signaling appears to confer the cardioprotective benefits of TNF. The absence of effect of mutation type or phenotype suggests that TNFR1 and TNFR2 may be sensitive to a disease progression of FD that is independent of genetic makeup. The increases in both TNFR1 and TNFR2 suggest a strong systemic inflammatory component of FD. Novel chronic heart failure therapies targeting these receptors are currently being investigated and accordingly may eventually be found to play a role in the management of FD patients with HFpEF. Importantly, TNFR1 and TNFR2 were associated with late gadolinium enhancement, which represents a prehypertrophic phenotype in FD. These biomarkers may also identify prehypertrophic stages of myocardial

**Figure 4.** Receiver operating characteristic (ROC) curve demonstrating the performance of Lyso-Gb₃ and Lyso-Gb₃ with analogues for the prediction of FD (n=40 healthy controls; n=68 FD patients). Lyso-Gb₃, the gold standard, had excellent performance (AUC=0.998), and Lyso-Gb₃ with analogues had an AUC=0.994. AUC indicates area under the curve; FD, Fabry disease; Lyso-Gb₃, lysoglobotriaosylceramide.

**Figure 5.** Plasma levels of biomarkers in FD patients (n=68) with (n=41) and without (n=27) LVH via imaging criteria. TNFR2, TNF, IL-6, MMP-2, and Lyso-Gb₃ are significantly higher in FD patients with LVH than in those without LVH. FD indicates Fabry disease; IL, interleukin; LVH, left ventricular hypertrophy (left ventricular mass index ≥85 g/m² in male and ≥81 g/m² in female patients); Lyso-Gb₃, lysoglobotriaosylceramide; MMP, matrix metalloprotease; TNF, tumor necrosis factor; TNFR, TNF receptor. *P<0.05; **P<0.01.
involvement in FD patients, triggering further investigations such as cardiac MRI. Fabry disease patients with LGE and/or diastolic dysfunction had significantly higher levels of BNP and MR-proANP, suggesting the presence of significant long-term pathological cardiac remodeling and possible eventual progression to heart failure. Elevated plasma troponins I and T provide further evidence for a cardiac-specific involvement in FD and, coupled with assessment of plasma natriuretic peptides, represent important diagnostic and prognostic tools in the evaluation of the cardiomyopathy associated with FD. MMP-2 and MMP-9 are implicated in remodeling of the extracellular matrix with increased MMP-2 levels associated with the presence of HFpEF, whereas increased MMP-9 levels predict LVH and adverse extracellular matrix remodeling. Increased plasma levels of the gelatinases MMP-2 and MMP-9 in FD patients suggest that inflammation and extracellular matrix remodeling are significant components of heart disease in FD. These findings are further supported by the correlation of MMP-2 with MSSI, LVH, LGE, and diastolic dysfunction. The elevation of MMP-9 is consistent with previous work and, together with the elevated galectin-1 levels, confirms a critical role of extracellular matrix remodeling in FD. Galectin-3 correlated strongly positively with MSSI and left atrial maximum size index, which suggests that galectin-3 may be a marker of advanced disease and the development of heart failure in patients with FD.

ERT was associated with increased levels of TNF, TNFR1, TNFR2, MMP-2, and Lyso-Gb3. These findings highlight the fact that patients who qualify for and receive ERT typically have more severe disease manifestations. Early detection of FD is especially critical as progression of the disease can be slowed by ERT. Importantly, TNFR1 and TNFR2 were associated with late gadolinium enhancement, which represents a prehypertrophic phenotype in female patients with FD. These biomarkers may also identify prehypertrophic stages of myocardial involvement in FD patients, triggering further investigations such as cardiac MRI. In contrast, patients with advanced FD continue to develop major adverse clinical events despite ERT, further underlying the need for early detection and appropriate intervention. Given that dysregulated inflammation persists in some patients despite ERT, monitoring these patients after initiation of ERT using inflammatory biomarkers may provide valuable information about disease control and long-term prognosis, particularly if used in conjunction with other previously identified biomarkers.

Lyso-Gb3 is a glycosphingolipid that accumulates in FD and serves as a disease-specific biomarker to identify clinically relevant mutations. Levels of plasma Lyso-Gb3 and utility in the diagnosis of FD were similar to those reported in prior studies. Plasma Lyso-Gb3 proved to be an ideal biomarker for the diagnosis of classic FD with both sensitivity and specificity of 100%. The use of a total plasma Lyso-Gb3 level incorporating the levels of 6 analogues is novel but did not confer increased accuracy for diagnosis of FD in our cohort. Interestingly, the levels of plasma Lyso-Gb3 for the 3 patients with unclassified mutations (G261V, intron2:c.369+5G>T, and V254Gfs) were
not statistically different from those of classic mutations, suggesting that these 3 mutations may be classified as classic rather than cardiac/renal variants. In addition to its utility in diagnosis, Lyso-Gb₃ correlated strongly with left ventricular mass index, which suggests that it may be a marker of sphingolipid accumulation in the myocardium. This finding is consistent with prior work involving FD patients with cardiac-specific variants.⁷⁻³ TNF, TNFR1, TNFR2, MMP-2, and Lyso-Gb₃ were elevated in the plasma of male patients with FD compared with female patients, which is expected given that the gene for α-galactosidase A follows an X-linked inheritance pattern, and, accordingly, hemizygotes typically suffer from a more severe form of the disease.⁶⁻⁹ Older patients with FD had elevated plasma biomarkers of remodeling, including BNP, MR-proANP, and galectin-3, without higher levels of inflammatory biomarkers. This finding could have therapeutic implications in that ERT may provide limited benefit in older patients. Indeed, in many patients with advanced FD, ERT does not prevent organ failure and death.⁷⁰ Monitoring markers of cardiac remodeling and systemic markers of inflammation may confer increased sensitivity for early subclinical manifestations for disease, which may indicate the need for aggressive treatment including ERT to prevent progression to ERT-refractory FD and its major associated complications.⁷⁰ The presence of systemic inflammation in

Figure 7. A, Plasma levels of biomarkers in FD patients (n=43) with (n=17) and without diastolic dysfunction (n=26) per echocardiography. BNP, MR-proANP, and MMP-2 are significantly higher in FD patients with diastolic dysfunction than in those without diastolic dysfunction. Diastolic dysfunction could not be assessed in some patients due to arrhythmia. B, Correlation plot of MR-proANP vs maximum LA size index ($R^2$=0.44, $P<0.001$). MR-proANP and maximum LA size index are positively correlated, as increasing LA size is known to cause atrial cardiomyocytes to release ANP. BNP indicates B-type natriuretic peptide; DD, diastolic dysfunction; FD, Fabry disease; LA, left atrial; MMP, matrix metalloprotease; MR-proANP, midregional pro–atrial natriuretic peptide. *$P<0.05$; **$P<0.01$.
pediatric FD patients is of special interest because these patients may not display major cardiac or renal manifestations until significant irreversible progression of the disease has occurred. In this case, systemic inflammation may contribute to long-term morbidity and mortality before these patients are symptomatic enough to qualify for ERT. Plasma biomarkers may also be valuable in patients with Fabry polymorphisms or mild
mutations, particularly in female patients, where the diagnosis and therapy of choice may be less clear.74

Study Limitations
A limitation of this study is the sample size, primarily due to the rarity of diagnosis of FD. Although comparable to other studies involving plasma analysis in FD, the sample size may limit the generalizability of the results, particularly with respect to phenotype (classic versus cardiac/renal variant). In addition, the small sample size limits the quality of the statistical tests and may have resulted in underpowered tests. A large sample size would enable further exploration of the relationship between the clinical and imaging variables and biomarker levels. Another limitation is the multiple comparisons, but we attempted to account for these issues by only testing
biologically plausible associations. However, many of the significances are strong enough that even the most stringent corrections would still result in rejection of the null hypothesis. Another, broader limitation of the study is the lack of clinical outcomes. Given the rarity of FD, multinational registries linked to phenotypic data would be valuable. Future studies linking these biomarkers to clinical outcomes are planned, which will help to further define their role in prognostication.

Conclusions
Plasma levels of inflammatory biomarkers, cardiac remodeling biomarkers, and Lyso-Gb3 are elevated in patients with FD. Patients with more severe disease, assessed via MSSI and its cardiac subset, have higher levels of inflammatory and remodeling biomarker levels. Several inflammatory and cardiac remodeling biomarkers, as well as Lyso-Gb3, were elevated in patients with LVH, whereas cardiac remodeling biomarkers were elevated in patients with diastolic dysfunction. Markers of cardiac remodeling, extracellular matrix turnover, and inflammation are significantly elevated in patients with renal dysfunction, suggesting that multisystem disease sequelae of FD are associated with greater states of inflammation. These features are consistent with a phenotype dominated by heart disease with preserved EF and renal disease and suggest a key pathogenic role of systemic inflammation. Exciting new advances in phenotype-specific and targeted anti-inflammatory therapy have the potential to revolutionize the management of FD.

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