A new approach for obtaining rapid uniformity in rice (Oryza sativa L.) via a 3x x 2x cross

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Abstract

A triploid (2n = 3x = 36) rice plant was obtained by screening a twin seedling population in which each seed germinated to two or three sprouts that were then crossed with diploid plants. One diploid plant was chosen among the various F₁ progenies and developed into an F₂ population via self-pollination. Compared with the control variety Shanyou 63, this F₂ population had a stable agronomical performance in field trials, as confirmed by the F-test. The stability of the F₂ population was further substantiated by molecular analysis with simple sequence repeat markers. Specifically, of 160 markers assayed, 37 (covering all 12 chromosomes) were polymorphic between the parental lines. Testing the F₁ hybrid individually with these markers showed that each PCR product had only a single band instead of two bands from each parent. The bands were identical to either maternal (23 markers) or paternal (eight markers) bands or distinct from both parents (six markers). The amplified bands of all 60 randomly selected F₂ plants were uniform and identical to those of the F₁ hybrid. These results suggest that the F₁ plant is a non-segregating hybrid and that a stable F₂ population was obtained. This novel system provides an efficient means for shortening the cycle of hybrid rice seed production.

Key words: F-test, polyploidy, rice, SSR marker, stability.

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Since the discovery that tetraploid plants can be regenerated from callus tissue on cut stems of diploid Solanum nigrum (Winkler, 1916) polyploidy has been recognized as a common phenomenon in nature and an important factor in the evolution of plant genomes. Polyploidy occurs in many taxa and is particularly widespread in flowering plants. At least half of the known angiosperm species have experienced polyploidy in their evolutionary history (Hieter and Griffiths, 1999; Echardt, 2001; Wu et al., 2001). Polyploidy often results in considerable genomic changes such as chromosomal rearrangements, gene loss and changes in DNA methylation (reviewed by Adams, 2007).

Compared to their diploid and haploid counterparts, polyploid organisms often express specific characteristics such as larger cell and body sizes (Sugiyama, 2005) and a propensity to develop apomixis (Naumova et al., 1999). Studies in rice have identified stable lines in an early generation from the progeny of 3x x 2x or 4x x 2x crosses (Wu et al., 1999; Xing et al., 2000). Wang et al. (1999) also reported that loss of heterozygosity (LOH) from 2x x 2x crosses led to stable panicle rows in F₂ progeny and subsequently proposed a mechanism of “assortment mitosis” (Wang et al., 2001) that was supported by cytological evidence (Wang et al., 2006).

In this study, we screened another triploid x diploid cross that differs from the crosses reported by Wu et al. (1999) and obtained a diploid F₁ plant that generated a stable F₂ population. This system will be helpful in providing new insights into the potential application of polyploidy and should allow the development of an efficient breeding system to greatly shorten the breeding cycle.

Individuals of the triploid plant DB43, originally derived from a twin seedling population, served as the maternal parent. A diploid japonica-type cultivar, ZD2, served as the paternal parent. The 25 F₁ seeds from a DB43/ZD2 cross were obtained by direct hybridization followed by embryo rescue. Five plants among the F₁ seedlings were cytologically confirmed to be diploid (Xing and Zhou, 2000). Self-pollinated F₂ seeds were collected to generate five F₂ populations in the following year. Only one of these five...
populations appeared to be phenotypically uniform in the field.

To verify the phenotypic uniformity of the F2 population, five major morphological traits (plant height, panicle length, number of productive tillers, seed-setting rate and 1000-grain weight) were investigated and compared with the very widespread Shanyou 63 as the control variety by using the F-test (Table 1). The F value (sd1/sd2) for each trait was < 1.0, indicating that the F2 population was stable for these agronomical traits under the field conditions used.

Microsatellite markers were used to assess the relationship between parents and the F1 hybrid and to test the stability of the F2 population. PCR was done with the following assay mixture in 25 μL: 40 ng of template DNA, 200 μM each of the four dNTPs, 2.5 μL of 10x buffer, 1 unit of DNA Taq polymerase, 2 mM MgCl2 and 0.25 μM of each of the two primers. The PCR amplifications were done in a Perkin Elmer 9600 GeneAmp PCR System with the following conditions: 94 °C for 7 min, followed by 35 cycles of 94 °C for 1 min, 55 °C for 1 min and 72 °C for 2 min, and a final extension at 72 °C for 10 min. The amplification products were separated by electrophoresis in 3% (w/v) agarose gels followed by staining with ethidium bromide and examination under UV light.

One hundred and sixty simple sequence repeat (SSR) markers were used to screen for polymorphisms in the parental lines: 37 of these markers covering all 12 rice chromosomes were polymorphic (Figure 1). More importantly, when these polymorphic markers were used to amplify the F1 DNA template individually, each F1 product showed only a single band instead of the expected two bands that were supposed to be identical to those from the two parents. Comparison of the PCR patterns of the parents with those of the F1 hybrid plant allowed the polymorphic SSR markers to be classified into three groups: Group 1 included 23 SSR markers for which the size of the band amplified from F1 was identical to that of the maternal band. *Group 2 markers for which the size of the amplified F1 band was identical to that of paternal band. *Group 3 markers for which the size of the amplified F1 band was distinct from that of both parents.

Table 1 - Comparison of stability for major agronomical traits between the F2 population and the control variety Shanyou 63 using the F-test.

| Traits               | F2 population | Control (Shanyou 63) | F value |
|----------------------|---------------|----------------------|---------|
| Plant height (cm)    | 125.9 ± 3.48  | 121.1 ± 6.32         | 0.55    |
| Panicle length (cm)  | 24.7 ± 1.14   | 25.2 ± 1.53          | 0.75    |
| Tiller number        | 8.6 ± 2.54    | 8.1 ± 2.9            | 0.87    |
| Seed-setting rate (%)| 60.7 ± 7.10   | 78.6 ± 10.1          | 0.70    |
| 1000-grain weight (g)| 24.1 ± 1.03   | 28.4 ± 1.11          | 0.93    |

The values are the mean ± SD.

Figure 1 - Chromosomal distribution of 37 polymorphic SSR markers among rice parental lines. The approximate positions of the markers and centromeres are based on the available genetic linkage maps for rice (Akagi et al., 1996; Chen et al., 1997; Temnykh et al., 2000, 2001). The superscripts indicate three different groups and the dots indicate the positions of centromeres. *Group 1 markers for which the size of the amplified F1 band was identical to that of maternal band. *Group 2 markers for which the size of the amplified F1 band was identical to that of paternal band. *Group 3 markers for which the size of the amplified F1 band was distinct from that of both parents.

Figure 2 - Pattern of PCR amplification for parental plants, F1 hybrid and F2 population. (A) RM224 marker in group 1: the size of the amplified F1 band was the same as that of the maternal plant. (B) RM241 marker in group 2: the size of the amplified F1 band was the same as that of the paternal plant. (C) RM168 marker in group 3: the size of the amplified F1 band was different from that of both parental plants. (D) Non-segregating amplified bands from F2 plants that were identical to amplified F1 bands, as assessed by using the markers RM234, RM241 and RM253 from groups 1, 2 and 3, respectively.
groups and 30 F$_2$ samples were chosen to illustrate this uniformity (Figure 2D).

Six markers had completely different PCR patterns with F$_1$ DNA template from those of their parents. This phenomenon has also been observed in wheat (Liu et al., 1998), although the mechanism of allele loss following hybridization remains unclear.

Various studies have shown that polyploidy can lead to immediate, extensive changes at the genomic levels, resulting in differential gene silencing or gene loss (reviewed by Udall and Wendel, 2006). Josefsson et al. (2006) showed that maternal imprinting of PHERES1(PHE1), the gene of type I MADS-box, and paternal imprinting of MEDEA(MEA), the gene encodes a polycomb group (PcG) protein, appeared to be lost in hybrids between tetraploid Arabidopsis thaliana and diploid Arabidopsis arenosa. This phenomenon, known as early generation stability, has previously been reported in rice from apomixis (Chen, 1992), although not all studies have confirmed this (Shi et al., 1996). The results of our experiment cannot be explained by apomixis because the markers tested in non-segregating diploid progeny were of mixed paternal and maternal origins. The most probable explanation in this case was recombination followed by chromosomal elimination in mitotic cells of the F$_1$ hybrid.

Our results indicate that the F$_2$ population was non-segregating and should theoretically be stable in subsequent generations. This unusual phenomenon, which differs from the findings previously reported by Wang et al. (1999), should prove useful for breeding restorer lines of hybrid rice (Zhou et al., 2007).

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