THE EFFECT OF YEAST (SACCHAROMYCES CEREVISAE), GARLIC (ALLIUM SATIVUM) AND THEIR COMBINATION AS FEED ADDITIVES IN FINISHING DIETS ON THE PERFORMANCE, RUMINAL FERMENTATION, AND IMMUNE STATUS OF LAMBS

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SUMMARY

Four groups of male lambs (seven animals/group) were used to study the effect of natural (garlic powder; Allium sativum) and biological (dry yeast; Saccharomyces cerevisiae) additives and their combination in finishing diets as compared to control diet on feed utilization and animal performance. Animals were 8 months of age and 35.8kg ± 0.41 as average body weight. Animals were fed a basal diet including concentrate feed mixture (CFM) at level of 70% of total requirement and berseem hay (BH) was offered ad lib. The experimental diets were: 1) a basal diet without additive (control), (C); 2) a basal diet supplemented with 6g dry yeast (2.44x10¹¹ cfu/g)/head/day, (Y); 3) a basal diet supplemented with 40g garlic powder/head/day, (G); and 4) a basal diet supplemented with 3g dry yeast plus 20g garlic powder/head/day, (YG). The results revealed that all feed additive treatments showed higher (P<0.05) digestibility values of DM, OM, CP, CF and NFE, than non-additive diet (C). The highest (P<0.05) values were observed for animals fed G diet; however, C group showed the lowest (P<0.05) digestibility values. The G diet showed the highest (P<0.05) value of TDN% (73.56%) and C showed the lowest one (69.20%). However, the DCP% was not affected (P<0.05) by additives and its values ranged between 11.81 and 12.27%. Animals fed enriched diets (Y, G and YG) showed higher ADG (180, 184 and 186 g/d, respectively) compared to control group (160g/d). Additives have no significant effect on feed intake either in the form of CFM or the roughage and consequently the total feed intake. All feed additives and their combination significantly (P<0.05) enhanced, with the same extent, the feed efficiency indicators of the enriched diets compared with control one. Yeast/garlic combined addition revealed the highest daily profit percentage relative to control (42%) followed by garlic alone (31%) treatments. Energy utilization was significantly different (P<0.05) between the test groups where, the G group showed the highest values, but C group was the lowest values. When the combined additive (YG) was supplemented, N balance exhibited 15.2% increase above the control group. The concentration of blood immunoglobulins (IgA) and IgG differed (P <0.05) among groups being their concentration were enhanced by the respective additives. It could be concluded that using feed additive such as dry yeast (6 gm/h/d) or garlic powder (40 gm/h/d) or their combination (3gm plus 20 gm, respectively) in finishing diets of lamb tended to increase digestibility coefficients for most of nutrients, increasing nutritive value as TDN and appeared to increase the daily gain as well as enhanced the immune status of animals.

Keywords: Yeast, garlic, lambs, performance, in vitro, fermentation and blood.

INTRODUCTION

The ruminant livestock industry plays a major role in the production of both of meat and milk as a key source of protein for human consumption. Sheep worldwide are mostly owned by poor rural families who lack modern management skills, and thus have poor feeding and housing practices with insufficient adoption of technologies which are important to improve productivity. Various dietary additives are widely used in ruminant diets modulate rumen metabolism, which ultimately improves nutrient use and animal performance. Enhancing feed quality and utilization by using certain feed additives may be considered a partial vertical solution to the problem of negative feed balance of total digestible nutrients.
(TDN) and digestible crude protein (DCP) which stated for a long period in the animal production sector of Egypt. There is need to save about 3.4 million tons of TDN over the total actual amount produced (9.6 million tons) to cover that required (13.0 million tons) as reported by Alnaimy et al. (2017).

Many workers applied chemical substances (antibiotics and hormones) as growth promoters in animal feeding to enhance the growth rate and to provide significant economic income. Using Herbs, spices, have received greater attention as potential alternatives to antibiotic growth promotants, since they are considered as natural products (Abd El-Latif et al., 2019). There are numerous investigations that focused on utilization of natural plant as feed additives in animals (Frankic et al., 2009). The addition of herbal additives to animals feed can also help stimulate the immune response (Khosravi et al., 2010) and improve digestibility of the feedstock, thereby enhancing quality of locally available feed source which in turn, helps to increase the production of sheep in our country. Among the well-known herbal additives include garlic which is small herbal plant of 30 to 50 cm height and belongs to the Amaryllidaceae family. Garlic (Allium sativum) has anti-microbial, anti-oxidant, and anti-hypertensive properties and has been used as a flavor in the animal nutrition industry (Rivlin, 2001 and Sivam, 2001).

There is no study focused on the impacts of yeast and garlic powder together on the rumen fermentation patterns and nutrient digestibility. It was hypothesized that the combination of both products may have an additive effect on stimulating fermentation and digestion of plant cells in the rumen. So the main research objective of this study is to examine the effect of yeast or garlic powder and combination of both as additives on lamb performance. The specific objectives of this study were to evaluate growth response of finishing lambs as well as economic efficiency; to evaluate the apparent digestibility of nutrients and rumen fermentation in lambs fed diets supplemented with yeast and/or garlic powder and to assess the impacts of feed additives on the immune status of animals.

MATERIALS AND METHODS

The farm experiment and the lab work were carried out at Animal Production Department, Faculty of Agriculture, Menoufia University, and the part of the fermentation study was accomplished at Maryout Research Station, Desert Research Center, Ministry of Agriculture.

Growth trial:

A complete randomized block design was followed in a growth trial for twenty-four male lambs which were divided into four groups (seven lambs/ group). Each group was housed in three pens and distributed as follows: 2, 2, and 3 lambs/pen.

Animals were 8 months of age with an average body weight of 35.8 kg (± 0.41 SE). Animals were weighted weekly before morning feeding. Animals were fed a basal diet (control) including concentrate feed mixture (CFM) at level of 70% of total requirement of growth as recommended by NRC (1985) and berseem hay (BH) was offered ad lib. The control ration was offered to the first group without additives, while the other three experimental groups received the control ration supplemented with dry yeast and/or garlic powder. The investigational diets were: 1) a basal diet without additive (control), (C); 2) a basal diet supplemented with 6 g dry yeast (2.44x10^11 cfu/g/head/day, (Y); 3) a basal diet supplemented with 40 g garlic powder/head/day, (G), and 4) a basal diet supplemented with 3 g dry yeast plus 20 g garlic powder/head/day, (YG). The concentrate feed mixture was formulated as follows: yellow corn 50%, cotton seed meal 25.0%, wheat bran 22.0%, limestone 1.6%, common salt 1.0%, mineral and vitamin mixture 0.4%. The amount of CFM was offered daily in two portions at 09:00 AM and 17:00 PM; however, water was available for animals all times throughout the experiment.

Immunological blood parameters for lambs:

Blood samples were collected in two tubes before feeding via the jugular vein from each lamb. The first tube to separate the blood plasma so it contains ethylene tetra acetic acid (EDTA) to prevent blood clotting to the white blood cell (WBC’s) count (Kolmer et al., 1951). The other tube for separating blood serum so it was without anti-coagulant and centrifuged 2h after collection at 3500 rpm for 20 minutes, then analyzed for immunoglobulin A (IgA) and immunoglobulin G (IgG) using enzyme-linked immunosorbent assay method (Thomas, 1998).
Economic indicators:

Market prices in 2019 were used to calculate the economic indicators expressed in Egyptian pounds (L.E.). The prices were assigned as follows; Berseem hay L.E. 2900/ton; concentrate feed mixture L.E. 4500/ton; dry yeast L.E. 22/kg; garlic powder L.E. 35/kg and live body weight L.E. 60/kg.

Digestibility, nitrogen balance, and water metabolism trials:

Twelve adult rams (50.41kg±0.61) were placed in individual metabolic cages (1.6 m x 0.53 m) and offered the same previous four rations (3 rams/ rations) for two weeks as an adaptation period followed by a week-long collection period. Water was offered twice daily, and water intake was recorded. Daily excreted feces from each animal were collected. Exactly 20% of the weight-based samples were taken and dried at 60 °C for 72 hours. Urine was collected daily in plastic jars, acidified (using 100 ml of 4N H₂SO₄), measured and 10% of the volume was sampled for nitrogen determination. Feed and fecal samples were ground through a 1 mm sieve on a Wiley mill grinder and sub samples were taken for each animal for subsequent analysis. Feed and fecal samples were analyzed for dry matter (DM), crude protein (CP), crude fiber (CF), ether extract (EE) and ash. Ruminal fluid were collected via cannula in individual metabolic cages (1.6 m x 0.53 m) and 

Rumen fermentation trials:

Four adult rumen-cannulated Barki rams with an average body weight of 47.90 ±1.7 kg were arranged in Latin square design for four periods (15days each) to study the effect of experimental diets on the deferential protozoal count and other fermentation criteria. Ruminal contents were sampled at zero, 3, 6 and 12h post-morning feeding to record the pH and determine concentration of NH₃-N, and volatile fatty acid (VFA). Ruminal pH was immediately measured using a digital pH meter (WPA CD70). Samples of rumen fluid were analyzed for ammonia-nitrogen (NH₃-N) according to Preston (1995). Total volatile fatty acids (TVFAs) were determined using steam distillation method described in Warner (1964).

Apart from the collected sample at zero time were immediately flittered through one layer of gauze, then fixed and stained with 4 times the discrete lag time prior to gas production.

The in vitro gas production technique was used to estimate in vitro degradation of both DM and OM (IVDMD & IVDMD), as well as methane production for experimental regimes (as total mixed rations). Ruminal content (50:50 v/v) collected via cannula was squeezed through four-layered cheesecloth and the filtrate liquid was incubated in a water bath at 39 °C saturated with CO₂ until further incubation. The incubation medium was prepared according to Menke et al. (1979) description. Each 200 mg DM sample was incubated in 100-mL glass serum bottles where 30 mL of the incubation medium was added. The samples were incubated in triplicate and the cumulated gas production was monitored 3, 6, 12, 24, and 48h post incubation. Three bottles containing rumen juice and artificial saliva without sample were used as blank to correct for gas production values released from rumen contents.

Data on gas production were adapted for to the following model from France et al. (2000): A = b × [1 – e⁻ʳ⁻¹], Where: A is the volume of gas production at time t; b is the asymptotic gas production (based on mL/200 mg DM); k is the rate of gas production per hour from the slowly fermentable feed fraction b; and L is the discrete lag time prior to gas production.

The partitioning factor (PF) was calculated as the ratio between the true digested DM (mg) to the volume of gas (mL in 24 h).

IVDMD and IVDMD were determined after terminal of each incubation time by recovery of the undigested fraction. The true degradability of DM and OM (TDMD & TDOM) was determined at 24h after reflux residual content with a 50 mL neutral detergent solution for 3h at 105°C. After recording the final gas volume at 24h of incubation, 4 ml NaOH (10 M) was injected in each bottle to measure methane volume as described by Demeyer et al. (1988).

The microbial protein (MCP) was calculated in accordance with Czerkawaski (1986), where 19.3 g microbial nitrogen produced per kg TDOM.
Calculations and statistical analysis:

Growth energy (GE) and digestible energy (DE) for the tested rations were calculated by equations reported by Nehring and Haenlien (1973) as follows:

\[
\text{GE (M cal/kg DM)} = 5.72 \times \text{CP} + 9.50 \times \text{EE} + 4.79 \times \text{CF} + 4.03 \times \text{NFE}
\]

\[
\text{DE (M cal/kg DM)} = 5.72 \times \text{DCP} + 9.05 \times \text{DEE} + 4.8 \times \text{DCF} + 4.06 \times \text{DNFE}
\]

Metabolizable energy (ME) was calculated as DE X 0.82 (NRC, 1985). The net energy for maintenance (NEm) or growth (NEg) was computed as outlined by Rattray et al. (1973).

\[
\text{NEm} = 0.79 \times \text{ME} - 0.4
\]

\[
\text{NEg} = 0.58 \times \text{ME} - 0.52
\]

Data of growth and digestibility trials were analyzed using Statistical Analytical System (SAS, 2002), Version 9.3.1, according to the General Linear Model as a completely randomized design with animals as block. The model of statistics was the following: \(Y_{ij} = \mu + T_i + e_{ij}\)

Where: \(Y_{ij}\) = the observation; \(\mu\) = Overall mean; \(T_i\) = the fixed effect of the treatments; \(e_{ij}\) = Random error component assumed to be normally distributed.

The statistical model associated with a Latin square design for the fermentation trials is:

\[
Y_{ijk} = \mu + \alpha_i + T_j + \beta_k + \epsilon_{ijk}
\]

Where \(\mu\) is the baseline mean, \(\alpha_i\) is the block effect associated with row \(i\), \(\beta_k\) is the block effect associated with column \(k\), \(T_j\) is the \(j\)th treatment effect, and \(\epsilon_{ijk}\) is a random error.

The Duncan multiple range test was performed (Duncan, 1955) to detect significant differences among means.

RESULTS AND DISCUSSION

Chemical composition of feeds and experimental feed additives:

The proximate composition of the bakery's yeast biomass (Saccharomyces cerevisiae) presented in Table (1) is characterized by high levels of protein (45.74%) and ash (6.79%). The CF content was low being value 2.51%. Total carbohydrates (45.72%) represent nearly half of the biomass. The present results agree with those recently reported by Elaref et al. (2020), which were 91.83, 82.58, 46.13, 3.85 and 39.24% respectively for DM, OM, CP, EE and NFE. The present values of CP, CF, EE% and Ash were higher than those found by Ibrahim and El Naggar (2018) who reported 40.82, 1.10, 1.33 and 2.98% for the same items in dry yeast, but our findings were lower in case of DM (87.05%), OM (97.02%) and NFE (53.77%). Compared with the present results of dry yeast contents, Abu E-Kassim et al. (2018) reported a lower DM, OM, CP and NFE values (92.58, 86.78, 14.10 and 50.22%, respectively) but cited higher values of CF, EE and Ash (19.74, 2.71 and 5.80%, respectively).

| Item | Experimental feed | Feed additive | Experimental ration |
|------|-------------------|---------------|---------------------|
| Moisture, % | CFM | BH | Dry yeast | Garlic powder | C | Y | G | YG |
| 13.03 | 13.98 | 13.08 | 5.99 | 13.30 | 13.30 | 13.13 | 13.22 |
| DM, % | 86.97 | 86.02 | 86.92 | 94.01 | 86.70 | 86.70 | 86.87 | 86.78 |
| CP | 13.05 | 12.53 | 45.74 | 20.44 | 12.90 | 13.04 | 13.13 | 13.09 |
| CF | 11.21 | 28.40 | 2.51 | 1.66 | 16.03 | 16.06 | 15.65 | 15.96 |
| EE | 3.86 | 3.53 | 1.75 | 1.38 | 3.77 | 3.76 | 3.71 | 3.73 |
| NFE | 63.95 | 37.84 | 43.21 | 72.59 | 56.62 | 56.48 | 57.18 | 56.67 |
| TC | 75.16 | 66.24 | 45.72 | 74.25 | 72.66 | 72.54 | 72.83 | 72.63 |
| Ash | 7.93 | 17.70 | 6.79 | 3.93 | 10.67 | 10.65 | 10.33 | 10.55 |

1CFM: Concentrate feed mixture; BH: Berseem hay.
2C: control diet CFM plus BH; Y: control diet enriched with yeast (6gm/h/d), G: control diet enriched with garlic powder (40gm/h/d), YG: control diet enriched with yeast (3gm/h/d) plus garlic powder (20gm/h/d).
3TC: Total carbohydrates = OM-(CP+EE).
The chemical composition of garlic powder (Local market product) is shown in Table (1). The results are similar to the earlier results of Otnuola et al. (2010), where moisture, CP, EE, CF, total carbohydrates and ash contents in garlic sample were 4.55, 15.33, 0.72, 2.10, 73.22 and 4.08% respectively on dry basis (95.45%). Petropoulos, et al. (2018) studied 14 Greek garlic genotypes, and they detected significant differences in nutrients contents. The range values on DM basis were calculated to be 31.67-42.64% DM, 3.59-5.73% Ash, 10.83-22.85% CP, 0.28-1.10% Fat, 70.93-84.50% carbohydrate, 94.27-96.41% OM and 3821-3901 Kcal/kg DM as an energy content. Garlic is relatively high in CP (20.44%), which is close to that declared by Sahli et al. (2018) which found a value of 22.9% and 18.8%, respectively. However, Abu El-Kassim et al. (2018) reported a lower CP value (14.10%). Garlic contained appreciable amounts of carbohydrates and protein and these results emphasize that it can be classified as carbohydrate and protein-rich spice (Abayomi et al., 2018). Furthermore, the high CP content of garlic was due to the presence of active metabolites such as allicin, ajoene and capsaicin as reported earlier by Dashak et al. (2001). The herein values from this study were higher than those reported for moisture and CP at 4.88 and 17.35% respectively by Nwinuka et al. (2005) and similar to the values of 73.03, 0.68, 4.06% reported for carbohydrate, ether extract and ash contents, respectively by the same authors. The result is in disagreement with the reports from Mariam and Devi (2016) who stated that, garlic contains 3.91% moisture, 19.75% CP, 0.49% EE, 1.73% CF, 66.36% carbohydrate and 3.39% ash on dry matter basis. Observed differences in chemical composition compared to literature may be related to genetic varieties and possibly to substrate analyzed mainly for cell wall. Furthermore, Petropoulos et al. (2018), indicated that apart from the genotype, both of growing conditions and cultivating practices also have a significant impact on the feeding value of garlic bulbs. In addition, plant density, fertilizer application rate and soil type have been reported to significantly affect the protein content of garlic bulbs (Diriba-Shiferaw et al., 2014).

Animal performance and economic indicators:

The ADG values revealed significant (P<0.05) differences across experimental groups (Table 2), where animal groups fed enrichment diets (Y, G and YG) showed higher values compared to control group (160 g/d) but on the same time the treated groups were comparable between each other. Relative to control group, the feed additives improved ADG by 12.5, 15.0 and 16.25% for Y, G and YG, respectively. Higher ADG in animals fed yeast supplemented diets (Y and YG) can be attributed to improved propiono-genesis process via yeast (Kawas et al., 2007). Malekkhahi et al. (2015) reported no effect of yeast culture supplementation on ADG or FCR in growing lambs. Hassan and Mohammed (2014) concluded that the addition of S. cerevisiae to the high concentrate diet improved the digestibility of CP and CF as well as ADG (143.7 g/d) in Awassi male lambs. Numerous studies (Garg et al., 2009; Milewski, 2009) have reported an increasing gain due to the addition of S. cerevisiae to sheep diet. Ahmed and Salah (2002) estimated a higher increase (13.8 and 30.2%) as a result of the addition of S. cerevisiae to sheep at a rate of 4 and 8 g/day, respectively, compared to the control diet. Maamouri et al. (2014) recorded 145 g/day and 223 g/day as the daily weight gain for respective lambs of (C) group and (Y) one. Payandeh and Kafilzadeh (2007) found that finishing lambs received diet supplemented with S. cerevisiae had a significantly higher ADG (209 vs. 177 g day\(^{-1}\)) but without positive effect on feed conversion ratio. These results are in harmony with those obtained on West African Dwarf goats fed diets supplanted with garlic powder (Ikyume et al., 2017) or sheep supplemented with yeast (Zeid et al., 2011). However, the current results contrast with that those of Tatar et al. (2008) who reported that garlic did not have a significantly effect on the growth rate although an improvement in growth was observed. Additionally, Bampidis et al. (2005) reported that weight gain was not significantly affected by dietary garlic pulp and husk supplementation in growing lambs compared to control group. Garlic powder supplementation in Ikyume et al. (2017) study had no significant effect on the feed conversion ratio (FCR) of West WAD goats. This result is consistent with that found by Strickland et al. (2009) where the inclusion of raw garlic in the diet of Merino lambs aged 6 months reduced the FCR.

Data concerning feed intake based on DM basis (Table 2), indicates that, the feed additive have no effect on feed intake either in the form of CFM or the roughage and consequently the total feed intake. Feed intake (g/head/d) expressed as TDNI was affected by the experimental feed additives, where animal’s group fed G diet showed the highest value of TDNI but the C group showed the lowest value and the other two groups (Y and YG) were similar. On the other hand, DCPI expressed as g/head/d (Table 2) was not affected (P<0.05) by treatments although G group showed higher value compared with the other three groups. The lack of effect of Yeast and/or garlic powder on DMI in this study may be attributed to the high proportion of concentrates (high energy intake) in the diet. These results agreed with Hassan and Mohammed (2014) and Malekkhahi et al. (2015) who found no significant impact on DM intake as a result of S. cerevisiae supplementation. However, on contrary to current results, a positive influence of SC on DM intake in growing animals was observed by Lascano et al. (2009).
Data of feed utilization efficiency expressed as DM/Gain, TDN/Gain or DCP/Gain are present in Table (2). The finding provides evidence that all feed additives and their combination significantly enhanced the daily feed conversion ratio value, also Zhong et al. (2019) found that lambs fed basal diet without or with 50g garlic powder to 84d resulted in no significant change in feed conversion ratio. The best feed efficiency obtained by YG additive may be attributed to the beneficial effects of yeast plus garlic. Where yeast provided stimulator factors and essential nutrients specially protein, energy, minerals and vitamins that better utilized by sheep (Zaki, 2016). These factors and essential nutrients resulted in some change in the digestive function that led to increasing the availability and utilization of nutrients in the rumen and could have a significant impact on the feed utilization and growth rate. Moreover, garlic has improved the use of energy and nitrogen from the diet. Briefly, it could be concluded that, yeast and/or garlic powder improved DM/Gain by 10.29, 11.17 and 14.05%, TDN/Gain by 6.87, 5.75 and 9.74% and DCP/Gain by 8.41, 8.41 and 11.21%. This result confirms that a low growth rate lead to a high ratio of feed to live weight gain. These results are consistent with those found by Maamouri et al. (2014) for yeast and Ghosh et al. (2010) for garlic powder. However, Hassan and Mohammed (2014) found that addition of S. cerevisiae (5 g/ head/d) in lamb diets had no effect on FCR value, also Zhong et al. (2019) found that lambs fed basal diet without or with 50g garlic powder per kg diet for 84d resulted in no significant change in feed conversion ratio.

The economic indicators were calculated for the animal groups fed the experimental finishing diets under the present study (Table 2). Its logic matter to find supplemented diets revealed higher daily feed cost compared to control one. All treatment groups revealed higher daily gain and consequently resulted in higher price. From these results, it could be concluded that adding yeast and yeast plus garlic powder to rations of lambs were more effective in increasing the daily profit percentage relative to control, being values 36 and 29%, respectively. However, garlic alone revealed a negative value mainly because of its high price so the use of garlic additive in finishing diet of growing lambs is restricted by a low price case.

Table (2): Changes in body weight, consumption criteria and economic indicators for lambs fed with enriched diets.

| Item                   | Experimental diet | SEM  | P value |
|------------------------|-------------------|------|---------|
|                        | C                 | Y    | G       | YG     |
| Body weight            |                   |      |         |        |
| IBW, kg                | 35.7              | 36.5 | 35.2    | 36.0   | 1.26  | 0.779 |
| FBW, kg                | 44.5              | 46.4 | 45.3    | 46.2   | 0.88  | 0.610 |
| ADG, g                 | 160^b             | 180^a| 184^a   | 186^a  | 3.23  | <0.001|
| Advantage, %           |                   | 12.5 | 15.0    | 16.25  | -     | -     |
| Feed & nutrients intake (g DM/head/d) |       |      |         |        |
| CFM                    | 1042              | 1045 | 1075    | 1035   | 10.69 | 0.207 |
| B. Hay                 | 405               | 415  | 402     | 410    | 12.17 | 0.884 |
| Total DMI              | 1447              | 1460 | 1477    | 1445   | 17.82 | 0.569 |
| TDN                    | 1001^b            | 1050^ab| 1086^a | 1050^ab| 20.24 | 0.051 |
| DCP                    | 171               | 177  | 181     | 177    | 3.78  | 0.292 |
| Feed utilization efficiency (g/g) |       |      |         |        |
| DM/Gain                | 9.04^a            | 8.11^b| 8.03^b  | 7.77^b | 0.19  | <0.001|
| TDN/Gain               | 6.26^a            | 5.83^ab| 5.90^ab| 5.65^b | 0.16  | 0.039 |
| DCP/Gain               | 1.07^a            | 0.98^b| 0.98^b  | 0.95^b | 0.03  | 0.028 |
| Economic indicators^1  |                   |      |         |        |
| Price of daily gain, LE| 9.60              | 10.80| 11.04   | 11.16  | -     | -     |
| Daily CFM cost, LE     | 5.39              | 5.54 | 6.96    | 6.13   | -     | -     |
| Daily hay cost, LE     | 1.37              | 1.40 | 1.36    | 1.38   | -     | -     |
| Total daily feed cost, LE| 6.76             | 6.94 | 8.32    | 7.51   | -     | -     |
| Daily Profit, LE       | 2.84              | 3.86 | 2.72    | 3.65   | -     | -     |
| Relative daily profit, %| 100              | 136  | 96      | 129    | -     | -     |
| Improvement, %          | 0.0               | 36.0 | -4.0    | 29.0   | -     | -     |

^1 Price of year 2019; CFM 4500 LE/T, BH 2900 LE/T, Yeast, 22 LE/Kg, Garlic 35 LE/Kg, LBW 60 LE/ Kg.
Economic indicators were calculated based on as fed basis including feed additives.

a and b means at the same raw with different superscript letters are significantly (P< 0.05) different.
**Immunological blood indicators:**

The data presented in Table (3) summarized the effect of the experimental finishing diets on immunological blood indicators in lambs. An increase in the ratio of neutrophils to lymphocytes (N/L ratio) as an immune parameter proposed as a marker of chronic stressful situations in farm animals (Trevisi and Bertoni, 2009). The N/L ratio in the present study (Table 3) showed progressively higher values with group C (0.24), which is indicative of stress. A higher N/L ratio may also indicate a health issue for the animal, reflecting a weakened immune system and often as a result, an unhealthy animal (Hyun-Sun et al., 2009). The lower N/L ratio observed in the respective feed additive groups probably reflects an identical humoral immune response in lambs. Data in Table (3) revealed that immunoglobulins (IgA) and IgG differed (P <0.05) among groups being their concentration were enhanced by the respective additives. These results were consistent with those obtained by El-Shereef (2019).

**Table (3): Immunological blood indicators in sheep fed yeast and/or garlic supplementation.**

| Item                  | Experimental diet | SEM | P Value |
|-----------------------|-------------------|-----|---------|
|                       | C                 | Y   | G       | SEM     | P Value |
| Neutrophils (N), (x10³/µl) | 17.63a            | 6.50b | 6.93c | 7.10b | 1.22 | <0.001 |
| Lymphocytes (L), (x10³/µl) | 72.85b            | 87.10b | 79.25d | 89.70b | 4.00 | 0.004 |
| N/L ratio             | 0.24              | 0.07 | 0.09   | 0.08  | -    | -     |
| IgA, (IU/l)           | 5.33b             | 8.33b | 8.10a | 8.67a | 0.69 | 0.032 |
| IgG, (IU/l)           | 7.43b             | 12.90b | 12.33b | 14.67a | 0.94 | 0.003 |

SEM: standard error of the mean; P value: probability value
a, b and c means at the same raw with different superscript letters are significantly (P< 0.05) different.

**Energy and nitrogen utilization:**

Data of gross energy (GE), digestible energy (DE), metabolizable energy (ME) and net energy for maintenance (NEm) and for growth (NEg) of lambs are presented in Table (4). It could be noticed that the GE was not significantly (P> 0.05) different among groups, being values ranged between 5.99 and 6.14 Mcal/d. It might be due to that feed intake was not different among groups. However, other criteria of energy utilization (DE, ME, NEm and NEg Mcal/d) were significantly (P<0.05) different across the experimental groups, where, the G group showed the highest values but C group was the lowest values, however both Y and YG groups showed comparable values. The same trend was observed for NEg as a ratio of GE and ME. These findings may explain the enhanced effect of the tested additives on the basal diets and was in accordance with finding of (El-Meccawi et al., 2009) who stated that the energy balance of small ruminants is dependent on the quality of their diets. The TVFAs are the final products of rumen microbial fermentation and represent the major supply of ME for ruminants (Van Soest, 1982). Therefore, using garlic as a feed additive may be responsible for the improvement of energy production and carbohydrate metabolism in animals received diets supplemented with garlic. The present result of NEg was in accordance with that found by Klevenhusen et al. (2011) who found that the concentrate supplemented with 4 g diallyl di-sulphide (an important component of garlic oil) increased (P = 0.07) body energy retention to be 4.06 MJ/d as compared with un-supplemented animals (3.48 MJ/d).

The impacts of Y, G or YG additives on N intake, fecal N, urinary N and N balance are shown in Table (4). Animals fed diet supplemented with G showed slight increase (P< 0.05) in N intake (NI) and lower N voided via feces (FN), however, both Y and YG groups showed comparable (P< 0.05) values and were in between G and C groups. As DMI was similar among the experimental groups so significant differences observed in NI was not attributed to DMI but mainly to additives nitrogen intake where it was 0.44, 1.31 and 0.88 g N/d for Y, G YG groups, respectively as compared to non-additive group. No significant (P< 0.05) effects due to supplement of Y, G or their combination were observed on the urinary nitrogen (UN), total voided nitrogen (TVN) or its relative percentage to NI (UN/NI% and TVN/NI%) as compared to the control group. A lower (P<0.05) nitrogen loss in the feces of the supplemented groups especially the focus of G group, but not observed in UN, emphasis that both of G and Y were more effective for N digestion than the absorption pathways. When the combined additives (YG) were supplemented, N balance exhibited 15.2% increase above the control group, whereas Y or G supplementation had similar values and located between Y and YG groups.

In accordance, Sallam et al. (2014) reported that microbial feed additives brought about less excretion of urinary and fecal nitrogen, which led to improvement in nitrogen balance. Cole et al. (1992) showed that lambs fed YC had higher N retention than the control which confirms our findings. The higher retention of N in group Y may be explained by the optimal ruminal NH₃-N concentration that appears to

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result from increased incorporation of N into microbial protein as a consequence of stimulated microbial activity (Malekkhahi et al., 2015). The results of N balance in this study contrast with that of Mungoi et al. (2012) who reported no effect of supplementing yeast on N balance in lambs. In earlier work, Amagase (2006) found antioxidant effects for the bioactive components of Garlic could play a role in improving the use of N in sheep fed hay supplemented with garlic leaf.

Table 4: Use of energy and nitrogen in lambs fed finishing diets enriched with yeast and/or garlic powder.

| Item* | C       | Y       | G       | SEM  | P value |
|-------|---------|---------|---------|------|---------|
|       | Experimental diet |       |         |      |         |
|       | GE, Mcal/d | 5.99    | 5.99    | 0.07 | 0.439   |
|       | DE, Mcal/d  | 3.74    | 3.91    | 0.08 | 0.099   |
|       | ME, Mcal/d  | 3.07    | 3.21    | 0.07 | 0.096   |
|       | NEm, Mcal/d | 2.38    | 2.49    | 0.05 | 0.097   |
|       | NEg, Mcal/d | 1.26    | 1.34    | 0.04 | 0.105   |
|       | NEg/GE%     | 20.93   | 22.34   | 0.59 | 0.158   |
|       | NEg/ME%     | 40.87   | 41.73   | 0.39 | 0.111   |
|       | N utilization: |         |         |      |         |
|       | Total NI (TNI), g/head/d | 29.87   | 30.25   | 0.36 | 0.174   |
|       | Fecal N (FN), g/head/d | 7.84    | 7.00    | 0.34 | 0.100   |
|       | FN/NI %     | 26.25   | 23.14   | 1.00 | 0.023   |
|       | N absorbed (NA), g/head/d | 22.03   | 23.25   | 0.41 | 0.003   |
|       | NA/NI %     | 73.75   | 76.86   | 1.07 | 0.022   |
|       | Urine N (UN), g/head/d | 15.57   | 15.81   | 0.56 | 0.223   |
|       | UN/NI %     | 52.13   | 52.26   | 1.66 | 0.497   |
|       | TVN*, g     | 23.41   | 22.81   | 0.48 | 0.570   |
|       | TVN/NI %    | 78.37   | 75.40   | 1.02 | 0.237   |
|       | N retained (NR), g/head/d | 6.46    | 7.44    | 0.29 | 0.094   |
|       | NR/TNI, %   | 21.63   | 24.60   | 1.02 | 0.215   |
|       | NR/NA, %    | 29.32   | 32.00   | 1.56 | 0.614   |
|       | NR, mg/kg BW | 128.9   | 148.6   | 7.14 | 0.214   |

*GE: gross energy; DE: digestible energy; ME: metabolizable energy; NEm: net energy for maintenance; NEg: net energy for growth; TVN: Total voided N; SEM: standard error of the mean; P value: probability value a, b and c means at the same raw with different superscript letters are significantly (P< 0.05) different.

Digestibility and feeding values of diets:

Table (5) presents nutrients digestibility and feeding values of the experimental diets. All feed additive treatments resulted in higher (P<0.05) digestibility values of DM, OM, CP, CF and NFE, but not in the C diet. In general, the highest (P<0.05) values were observed in animals fed G diet. However, C group showed the lowest (P<0.05) digestibility values for the same items. The digestibility of the EE was not affected (P<0.05) by the experimental additives. This result may be due to the digestibility of fat is not affected by the presence of yeast in the gastrointestinal tract, since yeast do not hydrolyze bile acids, and fat emulsion in mixed micelles (El-Hennawy et al., 1994). The observed increment in digestibility coefficient of major nutrients of enriched feed additive diets may be attributed to its high metabolizable energy content compared to their content of control diet (Kewan et al., 2019). Otherwise, garlic powder could alter the microbial population profile, reducing the activity of Prevotella spp which is mainly responsible for protein degradation and amino acids deamination leading to improved protein digestion and metabolism (El-Sherief, 2019).

These results are consistent with those obtained by Zhong et al. (2019) which found that the digestibility of DM (p = 0.019) and CP (p = 0.007) increased by garlic powder supplementation (5% or 50/ kg DM feed) however, lipid digestibility was not affected by the same supplementation. The values were 64.21, 74.38, 72.28% DM, CP, EE for garlic vs 60.29, 68.27, 69.97% for control. Also, the present results agree with those obtained by El-Sherief (2019) who noticed that the addition of garlic powder (2% of DMI) considerably enhanced the apparent digestibility of DM, OM, CF and CP compared to control ration being values were 59.6 vs 55.9% DMD; 56.8 vs 53.7% OM; 60.9 vs 52.5% CF; 70.6 vs 65.1% CP for garlic powder treatment vs control ration, respectively.
In contrast, Ikuyem et al. (2017) found that including 0.5% garlic powder inclusion in the diet of West African Dwarf goats significantly (p<0.05) reduced the digestibility of CP compared to the control group being values were 68.70 vs 75.78%, respectively. However, digestibility of DM, OM, CF, and EE was not affected (p<0.05) by the same level of garlic powder compared to un-supplemented group. The authors suggested that garlic powder inhibits the digestibility of protein, which could be good for the animals as the protein is protected for use in the small intestine. The improvement in nutrients digestibility by yeast supplementation is compatible with the findings of Malekkhahi et al. (2015) who found that yeast supplementation increased the digestibility of CP and NDF. Higher increases in digestibility were observed in Awassi lambs fed a high concentrate diet supplemented by S. cerevisiae (Hassan and Mohammed, 2014).

Table (5): Nutrients digestibility and feeding values of diets enriched with yeast and/or garlic.

| Items | C       | Y       | G       | YG      | SEM | P value |
|-------|---------|---------|---------|---------|-----|---------|
| Nutrients digestibility (%): |         |         |         |         |     |         |
| DM    | 72.30b  | 75.03ab | 76.70a  | 76.07a  | 1.29| 0.09    |
| OM    | 73.53b  | 76.37ab | 77.98a  | 77.17ab | 1.309| 0.105   |
| CP    | 73.69b  | 75.63ab | 78.47a  | 76.89a  | 1.07| 0.022   |
| CF    | 61.34ab | 66.99ab | 67.41a  | 66.54ab | 2.069| 0.147   |
| EE    | 74.72   | 79.40   | 78.14   | 78.11   | 4.65| 0.905   |
| NFE   | 76.87ab | 80.00ab | 80.73a  | 80.15ab | 1.31| 0.185   |
| Feeding values (%): |         |         |         |         |     |         |
| TDN   | 69.20b  | 71.96ab | 73.56a  | 72.66ab | 1.32| 0.124   |
| DCP   | 11.81   | 12.14   | 12.26   | 12.27   | 0.19| 0.315   |
| NR    | 4.86    | 4.93    | 5.00    | 4.92    | 0.10| 0.829   |
| NQI   | 9.51b   | 9.86b   | 10.30a  | 10.06a  | 0.17| 0.019   |

*Standard error of the means; *Probability value; *Total digestible nutrients; *Digestible crude protein
*Nutritive ratio = (TDN-DCP)/DCP; *Nutritive quality index = (CP %) × (DMD %)/100.

Means in the same row with different superscript letters are significantly (P<0.05) different.

The enhancement of nutrients digestibility reflected on the nutritive value expressed as TDN% (Table 5), so that the same trend was observed where G diet showed the highest value (73.56%) but C showed the lowest one (69.20%). However, the percentage of DCP was unaffected by additives and its values varied between 11.81 and 12.27%.

The nutritive ratio (NR) showed insignificant (P<0.05) differences mainly due to insignificant DCP%. The nutritive quality index (NQI) indicated that, feed additives enhanced the quality of the basal diet may be owing to significance in DM digestibility.

Calculating the improvement of TDN% achieved by additives in relative to control diet (Table 5) recoded that garlic addition was superior followed by combination of yeast and garlic and then yeast in the last (6.30, 5.00, and 3.99%, respectively). However, DCP% improved by 2.79, 3.81, 3.90% as a result of adding yeast, garlic and their half combination to the control diet. These results may be explained through the increase in favorable nitrogen source for rumen microbes beside the higher available carbohydrates which may lead to more microbial fermentation so that it reduced the dietary energy sources escaping from ruminal degradation. The present results are similar to that of Bueno et al. (2013) and Zeid et al. (2011).

**Water utilization:**

Water metabolism criteria are presented in Table (6). Data demonstrates that, the experimental feed additives had no effect (p<0.05) on combined feed water, free water intake as related to metabolic body weight (g/ kgW0.82) or as related to dry matter intake (g/ g DMI), and also excreted water in feces or urine. However, total water excretion expressed as g/ kgW0.82 or g/g DMI was recorded to be the highest (P<0.05) in control group and the lowest in G group. Both of two groups Y and YG showed comparable values and were in between C and G groups.

Animal groups fed diet included garlic powder (G and YG) showed higher insensible water loss (IWL) expressed as g/kgW0.82 or as relative to TWI, TDNI, and DCPI. On the other hand, yeast group (Y) showed the lowest values of IWL or g IWL/ kg TDNI.
The respective feed additives increased metabolic water intake as compared with non-additive diet but it did not reach to be significant, although mean values of combined metabolic water intake are mainly related to TDNI of each diet (Kewan et al., 2017). Higher values of total water loss recorded for C group may be resulted as a consequence of higher water turnover rate and/or digesta flow (Aradjo et al., 2010). However, the higher (P < 0.05) insensible water loss observed in the YG group may be attributed to the inclusion of garlic powder which may cause increasing of heat increment that result from diet fermentation which may lead to increase water needed for body cooling system (Kewan et al., 2017), however yeast group (Y) showed lower insensible water loss as the yeast may have anti-oxidative stress effects for animals (Hyun-Sun et al., 2009).

Table (6): Water utilization in rams fed diets enriched with yeast and/or garlic powder.

| Item                              | Experimental diet | SEM   | P value |
|-----------------------------------|-------------------|-------|---------|
| Metabolic body weight, kgW<sub>0.82</sub> | C                 | 24.87 | 0.52    | 0.999   |
|                                  | Y                 | 24.87 | 8.96    | 0.21    | 0.993   |
|                                  | G                 | 24.92 | 8.99    | 0.21    | 0.993   |
|                                  | YG                | 24.86 | 8.98    | 0.21    | 0.993   |
| Feed combined water, g/kgW<sub>0.82</sub> | C     | 157.7 | 2.69    | 0.381   |
|                                  | Y                 | 141.8 | 2.39    | 0.122   |
|                                  | G                 | 139.6 | 2.34    | 0.122   |
|                                  | YG                | 157.7 | 2.67    | 0.122   |
| Free water intake (FWI):          |                   |       |         |
| g/kgW<sub>0.82</sub>             | C                 | 24.29 | 2.45    | 0.76    | 0.356   |
|                                  | Y                 | 25.45 | 26.45   | 0.76    | 0.356   |
|                                  | G                 | 26.45 | 25.42   | 0.76    | 0.356   |
|                                  | YG                | 25.42 | 0.76    | 0.356   |
| Metabolic water, g/kgW<sub>0.75</sub> | C     | 191.0 | 176.3   |
|                                  | Y                 | 176.3 | 192.1   |
|                                  | G                 | 175.0 | 192.1   |
| Total water intake TWI, g/kgW<sub>0.82</sub> | YG   | 176.3 | 20.18   |
| Fecal water (FW):                |                   |       |         |
| g/kgW<sub>0.82</sub>             | C                 | 18.07 | 17.22   |
|                                  | Y                 | 15.47 | 8.96    |
|                                  | G                 | 14.76 | 9.25    |
|                                  | YG                | 17.62 | 9.46    |
| % of TWI                         | C                 | 9.46  | 7.97    |
|                                  | Y                 | 8.96  | 9.25    |
|                                  | G                 | 9.25  | 9.25    |
| Urine water (UW):                |                   |       |         |
| g/kgW<sub>0.82</sub>             | C                 | 89.35 | 81.15   |
|                                  | Y                 | 86.0  | 70.61   |
|                                  | G                 | 70.61 | 86.0    |
|                                  | YG                | 103.7 | 86.0    |
| % of TWI                         | C                 | 46.78 | 40.33   |
|                                  | Y                 | 46.03 | 36.76   |
|                                  | G                 | 46.03 | 36.76   |
| Total water excretion (TWE):      |                   |       |         |
| g/kgW<sub>0.82</sub>             | C                 | 107.4 | 98.4    |
|                                  | Y                 | 98.4  | 86.0    |
|                                  | G                 | 86.0  | 88.4    |
|                                  | YG                | 88.4  | 88.4    |
| % of TWI                         | C                 | 56.23 | 49.14   |
|                                  | Y                 | 55.81 | 46.02   |
|                                  | G                 | 46.02 | 46.02   |
| g/kg DMI                         | C                 | 1.83  | 1.45    |
|                                  | Y                 | 1.64  | 1.50    |
|                                  | G                 | 1.50  | 1.50    |
| Insensible water loss (IWL):      |                   |       |         |
| g/kgW<sub>0.82</sub>             | C                 | 83.60 | 77.90   |
|                                  | Y                 | 89.00 | 103.7   |
|                                  | G                 | 89.00 | 103.7   |
|                                  | YG                | 103.7 | 103.7   |
| % of TWI                         | C                 | 43.77 | 50.86   |
|                                  | Y                 | 44.19 | 53.98   |
|                                  | G                 | 53.98 | 53.98   |
|                                  | YG                | 53.98 | 53.98   |
| g/kg TDNI                        | C                 | 2.07  | 2.04    |
|                                  | Y                 | 1.85  | 2.36    |
|                                  | G                 | 2.36  | 2.36    |
|                                  | YG                | 2.36  | 2.36    |
| g/kg DCPI                        | C                 | 12.15 | 12.15   |
|                                  | Y                 | 10.94 | 13.98   |
|                                  | G                 | 13.98 | 13.98   |
|                                  | YG                | 13.98 | 13.98   |
| g/kg NR                          | C                 | 330.4 | 335.4   |
|                                  | Y                 | 337.5 | 335.4   |
|                                  | G                 | 335.4 | 335.4   |
|                                  | YG                | 335.4 | 335.4   |

SEM: standard error of the mean, P value: probability value
a, and b means at the same raw with different superscript letter are significantly (P< 0.05) different.

In vitro rumen fermentation parameters:

Rumen pH, NH<sub>3</sub>-N and TVFA:

In vitro rumen pH, NH<sub>3</sub>-N, and TVFA concentrations at 0, 3, 6, and 12 post-feeding rams on control, Y, G and YG supplemented diets are given in Table (7). The garlic and control diets were similar (P< 0.001) in pH at zero time and also the same finding was observed for Y and YG diets. It can be observed that, pH value was higher at zero than other all incubation times or in other words, it almost declined with progressing of time from zero up to 12h for all the experimental diets with significant differences at all tested diets. Gradual decreasing of rumen pH with progressing time may be due to higher concentrate otherwise higher organic acids resulted from fermentation caused by feed additives may explain the gradually significant decreasing of rumen pH against the time detected in the current study. The present results contrast with those reported by Sahli et al. (2018) who found no changes in the in vitro fermentation of the rumen by including garlic powder. Also, the pH of rumen liquor was not affected by garlic treatment in sheep (Kongmun et al., 2010 and Abu El-Kassim et al., 2018) or dairy goats (Kholif et al., 2012). On the other hand, Yang, et al. (2004) and Gaafar, et al. (2009) found that adding yeast led to an increase in ruminal pH by decreasing the ruminal lactate concentrations through increased activity of lactate fermenting bacteria (Selenomonas ruminantium and Megasphaera elsdenii) in the rumen.
Ammonia-nitrogen concentration (Table 7) was not affected (p>0.05) by the experimental diets at zero time of feeding. Diets included garlic powder (G and YG) showed higher (P<0.001) NH₃-N concentration at early hours (3 and 6h) as compared to the other experimental diets but Y diet was the highest at the late hour (12) post-feeding. The CP content of Y, G and YG were higher than C diets owing to feed additives, so the results herein showed that supplemented diets produce higher (p<0.001) NH₃-N as compared with C diet. This may be explained by the possible difference in the degradability of CP in the rumen between supplemented and un-supplemented diets. The increased concentration of NH₃-N suggests that yeast and garlic have increased the ruminal degradable protein and hence the ability of produce higher levels of microbial protein. Carbohydrates are the most important source of energy for the transfer of NH₃-N by microorganisms; therefore, the rate of carbohydrate fermentation was highly related to the rate of rumen protein degradation to NH₃-N and then production of microbial protein (Van Soest, 1982). Previous studies showed that concentration of ammonia-N decreased significantly (P<0.05) in animals fed garlic (Abu EL-Kassim et al., 2018) or yeast (Lascano and Heinrichs, 2009) as compared to that fed control diet. This decline may be attributed to the increased incorporation of ammonia in microbial protein (Chaucheyras and Fonty, 2001), and the stimulation of microbial activity (Lascano and Heinrichs, 2009), or it can be a direct effect of yeast on the reduction of CP degradation (Eweedah, et al., 2005).

Table 7: The effect of yeast and/or garlic feed additives on in vitro rumen fermentation.

| Incubation liquor pH | Experimental diet | SEM  | P value |
|----------------------|-------------------|------|---------|
| Rumen liquor pH      |                   |      |         |
| 0                    | 6.63ᵇ             | 6.56ᵇ| <0.01   | <0.001  |
| 3h                   | 5.87ᵃ             | 5.78ᵇ| <0.01   | <0.001  |
| 6h                   | 5.65ᵃ             | 5.64ᵇ| <0.01   | 0.02    |
| 12h                  | 5.48ᵃ             | 5.46ᵇ| <0.01   | 0.019   |
| NH₃-N, mg/dL         |                   |      |         |
| 0                    | 9.80              | 9.80  | 0.55    | 0.16    |
| 3h                   | 19.63ᵇ            | 21.25ᵇ| 0.55    | <0.001  |
| 6h                   | 24.08ᵇ            | 24.07ᵇ| 0.80    | <0.001  |
| 12h                  | 23.93ᵇ            | 26.30ᵇ| 0.67    | <0.001  |
| TVFA, meq/dL         |                   |      |         |
| 0                    | 3.28              | 3.50  | 0.07    | 0.18    |
| 3h                   | 5.50ᵇ             | 5.05ᵇ| 0.16    | 0.005   |
| 6h                   | 6.28ᵇ             | 7.50ᵇ| 0.13    | 0.001   |
| 12h                  | 9.05ᵇ             | 8.03ᵇ| 0.16    | 0.008   |

SEM: standard error of the mean, P value: probability value
a, b and c means at the same raw with different superscript letters are significantly (P< 0.05) different.

Our results regarding ruminal pH and NH₃-N disagreed with those obtained by Putnam, et al. (1997), where they reported no significant effect of adding yeast on the concentration of ammonia-N or the pH of the rumen fluid. This disagreement may be attributed to differences in the level of addition and/or different SC strains used. Newbold et al. (1995) stated that certain yeast strains are effective while others are not.

Higher values of rumen ammonia concentration at 6h for YG and at 12h in Y group may be attributed to an increase in proteolysis and protein deamination by micro-organisms and increase the ruminal non ammonia nitrogen pools resulted after addition of S. cerevisiae living cells (Galip, 2006). Higher value of rumen ammonia in G group at 3h was in consistence with that found in lactating cows fed garlic oil (Yang et al., 2007) however Ikyume et al. (2017), observed reduced NH₃-N concentration during fermentation as a result of garlic supplementation.

The total VFA concentration (Table 7) was not affected (P> 0.05) by the experimental diets at zero hour post-feeding. It was noticed that the experimental additives have main effect within 6h post feeding, where combined YG increased (p< 0.01) the TVFA concentration at 3h as compared to the other experimental diets. However, separate Y or G supplementation increased TVFA at 6h as compared to both of the other two groups (C and YG). The control diet showed the highest (P< 0.01) TVFA concentration at 12h as compared to the other supplemented diets.
The previous data concerning high TVAs at 6hr for Y group matches well to those reported for sheep by Komonna (2007). They reported that the total VFA was higher in supplemented groups with YC compared to the control group. In contrast, the work of Ismaiel et al. (2010) on sheep and Gado et al. (1998) on goats revealed insignificant differences in total VFA due to yeast culture supplementation. High TVFA at 6h for G group was in accordance with that reported by Zhong et al. (2019) who found that garlic powder supplementation increased total VFA in dairy goats as well as in sheep. However, Ikyume et al. (2017) did not observe significant differences due to garlic supplementation.

**DM and OM degradability:**

As apparent digestibility is not enough to evaluate the nutritive value of ruminant feeds, therefore it is necessary to determine the ruminal kinetics of digesting dietary nutrients. In vitro degradation data for DM and OM are presented in Table (8). Respective additive showed significant effect at all incubation times except at 12h. For DM, the intercept value (a) for the different treatments representing dry mater degraded (DMD) from soluble fraction ranged from 17.50 to 20.19 and it was significantly different (P<0.05) among treatments. Where Y diet showed the highest value and G had the lowest one. In addition, all kinetic constants; dry matter degraded from the insoluble fraction (b), the potential extent of DMD (a+b), the degradation rate constant for the insoluble fraction (c); and also the effective degradability were significantly different among treatments (P<0.05). It seems that G had the highest values followed by YG, Y and then C group.

The effective degradability of DM and OM for the experimental diets is given in Table (8). Data were calculated using rumen outflow rates of 2, 4 and 8%h⁻¹. There were significant differences (p<0.01) among diets where G diet displayed the highest values as compared to other diets.

**Table (8): The effect of yeast and/or garlic feed additives on in vitro degradability of DM and OM**

| Item | Experimental diet | SE M | Experimental diet | SEM |
|------|-------------------|------|-------------------|-----|
| In Vitro DM degradability (%) | C | Y | G | YG | C | Y | G | YG |
| a₀ | 15.08ᵇ | 14.72ᵇ | 12.59ᵃ | 12.46ᵇ | 0.31 | 2.19 | 2.55 | 2.29 | 2.40 | 0.20 |
| a₁ | 36.57ᵇ | 36.22ᵇ | 39.47ᵇ | 38.04ᵇ | 0.85 | 19.60ᵇ | 13.35ᵇ | 5.92ᵇ | 17.92ᵇ | 0.77 |
| a₂ | 43.53ᵇ | 44.88ᵇ | 48.87ᵇ | 42.54ᵇ | 1.79 | 31.61ᵇ | 35.81ᵇ | 34.47ᵇ | 40.63ᵇ | 0.86 |
| a₃ | 46.03 | 46.16 | 47.16 | 45.88 | 0.42 | 41.20ᵇ | 50.07ᵃ | 47.99ᵇ | 49.55ᵇ | 0.52 |
| a₄ | 55.28ᵇ | 60.39ᵇ | 65.8ᵃ | 63.26ᵇ | 0.26 | 52.66ᵇ | 59.13ᵃ | 56.61ᵇ | 57.15ᵇ | 0.74 |
| a₅ | 66.09ᵇ | 74.27ᵇ | 74.94ᵇ | 75.61ᵇ | 0.59 | 78.67ᵇ | 83.39ᵇ | 83.06ᵇ | 78.67ᵇ | 0.41 |
| OM kinetics | | | | | | | | | |
| b₀ | 18.73ᵇ | 20.19ᵃ | 17.50ᵇ | 18.74ᵇ | 0.51 | 7.12ᵇ | 3.24ᵇ | 1.10ᵇ | 6.00ᵇ | 0.36 |
| b₁ | 43.05ᵇ | 52.79ᵇ | 54.01ᵇ | 56.33ᵇ | 1.12 | 77.01ᵇ | 80.16ᵇ | 83.98ᵇ | 78.15ᵇ | 1.02 |
| b₂ | 61.78ᵇ | 72.98ᵇ | 71.51ᵇ | 75.07ᵇ | 1.34 | 84.14 | 83.40 | 85.08 | 84.15 | 0.97 |
| b₃ | 0.117ᵇ | 0.074ᵇ | 0.114ᵇ | 0.073ᵇ | 0.01 | 0.047ᵇ | 0.066ᵇ | 0.059ᵇ | 0.065ᵇ | 0.01 |
| b₄ | 55.48ᵇ | 61.71ᵇ | 62.91ᵇ | 62.97ᵇ | 0.39 | 77.01ᵇ | 80.16ᵇ | 83.10ᵇ | 78.15ᵇ | 1.02 |
| b₅ | 50.80ᵇ | 54.42ᵇ | 56.78ᵇ | 55.15ᵇ | 0.27 | 77.01ᵇ | 80.16ᵇ | 83.98ᵇ | 78.15ᵇ | 1.03 |
| b₆ | 44.29ᵇ | 45.54ᵇ | 48.53ᵇ | 45.65ᵇ | 0.60 | 77.01ᵇ | 80.16ᵇ | 83.98ᵇ | 78.15ᵇ | 1.03 |
| True DMD₂⁴h | | | | | | | | | |
| % | 65.34ᵈ | 83.68ᵇ | 85.33ᵇ | 73.30ᵇ | 0.32 | 58.98ᵈ | 79.30ᵃ | 72.99ᵇ | 66.34ᶜ | 0.95 |
| Improve destinations | 18.20ᵈ | 38.56ᵃ | 30.27ᵇ | 19.03ᶜ | 0.05 | 11.99ᵈ | 34.11ᵃ | 28.94ᵇ | 16.07ᶜ | 0.25 |
| - | 28.08 | 26.55 | 15.25 | - | - | 34.45 | 23.75 | 12.47 | - |

**SEM:** standard error of the mean, P value: probability value. a, b and c are degradation constants. Calculated based on apparent DMD₂⁴h. Calculated based on true DMD of control diet.

*a, b and c means at the same row with different superscript letters are significantly (P<0.05) different.

True DM degradability (Table 8) was exclusively higher by G additive then Y and YG in the second category. However, the highest improvement based on apparent DMD at 24h was revealed by yeast followed by garlic powder and then YG combination. Ryan and Gray, 1989 showed that the rate of substrate fermentation increase as a result for multiplication of bacterial numbers by S. cerevisiae supplementation. Hadjiapanayiotou et al. (1997) claimed that the use of S. cerevisiae did not affect the digestibility of nutrient, whereas Plata et al. (1994) found positive in vivo or in situ responses. Garlic powder supplemented treatment (16 mg) increased in vitro true digestibility (IVTD) as compared to the control (P<0.01). However, Yang et al. (2007) observed that garlic supplementation did not affect the total digestibility’s of DM, OM, fiber and starch, while ruminal DM and OM digestibility was increased (Kongmun et al., 2010). Sahli, et al. (2018) demonstrated an increase (P <0.001) in in vitro gas production with the addition of 32 and 64 mg garlic powder. They added that, TOMD was similar for all
the doses (0, 4, 8, 16, 32 mg) except for 64 mg, where a small but significant (P < 0.001) increase was observed (77.7%).

Ikyume et al. (2018) found that garlic inclusion had no (P>0.05) influence on the measured in vitro digestibility kinetics. Where, IDMD was not statistically (P>0.05) significant, garlic powder 0.5% group had numerically higher value (68.11%) while the control had the least value of 65.23%. The non-significantly in vitro organic matter digestibility (OMD) ranged between 68.47% and 84.32%. The highest percentage OMD was observed in the garlic powder 0.5% group while the control group had the least value of 68.47%. Favorable ruminal digestion responses to yeast culture feeding in sheep include an increase DM and OM degradation (Kamel et al., 2004). The present results are within that range reported for roughage and concentrate by Mabjeesh et al. (2000) where the IVDMD of roughages ranged from 47 to 61%, the highest for grass hay and lowest for clover hay. The value of IVDMD for grains varied from 63 to 92%, the highest value recorded for corn. CP supplements also showed a wide range of IVDMD values, 55 to 91%, the lowest for cottonseed meal and the highest for fish meal. The IVDMD of whole cottonseed was low compared to other feedstuffs, averaging 38%.

All in vitro OM degradability (Table 8) at different incubation times and its kinetic values otherwise true OM degradability as well as its improvement based on either apparent OMD of the same diet or true OMD for control diet were parallel to the same criteria of DMD as found by Guney et al. (2016). It is well accepted that the significant differences observed in OMD among the experiment diets mainly resulted from feed additive effects on the same basal diet. It means that yeast, garlic powder and its combination showed higher OM fermentation along with the different period of fermentation. So, its modifications effects are still unknown, but we can expect from the other data of the present results that yeast addition mainly affect through stimulate the fibrolytic bacteria (Chaucheyras et al., 2019) or garlic supplements inhibit Archaea, which produce methane in the rumen (Kamel et al., 2004). Khattab et al. (2010) showed that dried yeast and garlic recorded higher (P< 0.05) values of IVOMD and IVOMD than control. The improvement of IVDMD and IVOMD with combinations may be attributed to one or more of the following reasons: 1) available in essential ingredients such as vitamins, enzymes and essential amino and fatty acids for microflora from yeast (El-Ashry et al., 2001), 2) improvement in the flora environment for better digestibility with yeast (Campagne et al., 2008), 3) the medicinal oils of garlic (Khater et al., 2009). It could be concluded that the present feed additive treatments were the most effective treatment in increasing dry matter and organic matter disappearances from nutritional point of view.

**Rumen protozoa and total gas and methane yield:**

The values of ruminal ciliate protozoa count as affected by the experimental treatments are illustrated in Table (9). A significant (P<0.05) difference was observed for different differential species (Entodinium sp., Epidenium sp., Diplodenum sp.) except Polyastraon sp. and total count due to experimental treatments at zero-time of feeding. The present results indicated that the highest density was recorded for Entodinium spp which is ferment cellulose and protein while the lowest densities recorded for Diplodenum spp and Polyastraon spp which is ferment cellulose, especially that Polyastraon spp can digest 50% of cellulose in the rumen (Hungate, 1966). The high count of Entodinium spp matched with lower ruminal pH in the experimental groups as it has tolerance for lower pH (Aziz et al., 2018) and can be explained that Entodinium spp. is responsible for the use of lactic acid formed in the rumen (Khaled and Baraka, 2011) and direct feed microbial that produce lactate (e.g. Lactobacillus acidophilus) maintain a tonic level of lactic acid in the rumen, which has the potential to stimulate microorganisms that utilize lactic acid (Nocek et al., 2002).

The data confirmed that Y displayed the highest (P<0.05) values for the differential species and total count at zero hour sampling time of feeding followed by C, while G had the lowest (P<0.05) values. It seems that garlic powder diet decreased (P< 0.05) the count (×10^7/mL) of Entodinium sp., Epidenium sp. and Diplodenum sp. as compared with control diet. Where, yeast supplement may have factors that encourage increasing the previous species to make record the highest (P<0.05) values for the same order of protozoa species.

The present results of C group were higher than that found in goats fed diets with 70:30% concentrate to roughage ratio (Aziz et al., 2018), this may be due to sheep was higher than goats in protozoa count as reported by Baraka (2012). With regard to protozoa number, it is clear the reduction effect on garlic powder in microbial activities. The antimicrobial properties of the aromatic plants are attributed in part to essential oils (Panghal et al., 2011). Similar results were found by Nassar et al. (2017) who clarified that the addition of garlic powder or oil in rations of Barki sheep reduces (P<0.05) the population of protozoa. Concerning yeast supplementation effect, the results were in accordance with those of Brossard et al. (2006) who stated that yeasts tended to increase the ruminal protozoal population (P< 0.1). Contrary

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to current results, Hernández et al. (2009) showed that the addition of yeast to lambs fed early and mature orchard grass altered ruminal protozoa without affecting feed intake, total tract digestion and N balance.

Table (9): Total count and differential of rumen protozoa at zero time of feeding lambs and in vitro gas yield and kinetics for diets enriched with yeast and/or garlic.

| Item                                | Experimental diet | SEM | P value |
|-------------------------------------|-------------------|-----|---------|
| Protozoal Differential Count (x 10³/mL) |                   |     |         |
| Entodenium sp.                      | C: 6.70b          | Y: 8.17b | G: 4.44d | 5.10b | 0.43 | 0.048 |
| Epipdenium sp.                      | 3.71b             | 6.13b | 2.68d | 2.99c | 0.36 | 0.044 |
| Diplodenium sp.                     | 0.66b             | 1.97b | 0.51c | 0.54c | 0.17 | 0.049 |
| Polyplastron sp.                    | 0.48              | 0.47  | 0.45  | 0.44  | 0.18 | 0.611 |
| Total (x 10³/mL)                    | 11.55b            | 16.74a | 8.08d | 9.07c | 0.44 | 0.050 |
| Total gas yield; TGY (ml/200mg DM)  |                   |     |       |       |      |       |
| TGY 3h                              | 16.13a            | 15.78a | 12.27b | 15.28a | 0.38 | <0.001 |
| TGY 6h                              | 24.53a            | 23.03a | 17.53c | 18.03c | 0.21 | <0.001 |
| TGY 12h                             | 28.03a            | 27.03b | 21.53c | 21.53c | 0.20 | <0.001 |
| TGY 24h                             | 30.53a            | 28.28b | 19.53d | 21.53c | 0.53 | <0.001 |
| TGY 48h                             | 32.2a             | 30.87b | 27.08c | 24.53d | 0.40 | <0.001 |
| PF*                                 | 1.93c             | 2.81b | 3.75a | 3.08b | 0.10 | <0.001 |
| Kinetic constants:                  |                   |     |       |       |      |       |
| A                                   | 2.78b             | 3.78b | 11.87a | 11.96a | 0.72 | <0.001 |
| B                                   | 28.41a            | 25.79a | 18.63b | 11.95c | 0.83 | <0.001 |
| a+b                                 | 31.19a            | 29.57a | 30.50a | 23.91b | 1.13 | 0.007 |
| C                                   | 0.219a            | 0.215a | 0.041c | 0.117b | 0.02 | <0.001 |
| Methane yield:                      |                   |     |       |       |      |       |
| CH₄ yield/10⁴, ml                   | 18.57             | 18.41 | 8.20  | 11.74 | 0.22 | <0.001 |
| CH₄/TGY ratio                       | 60.84             | 65.09 | 41.97 | 54.55 | 0.51 | <0.001 |
| CH₄ energy (MJ/d)                   | 5.30              | 5.30  | 2.39  | 3.34  | -    | -    |
| CH₄ energy/GEI %                    | 21.15             | 20.94 | 9.30  | 13.33 | -    | -    |
| Microbial protein synthesis:        |                   |     |       |       |      |       |
| MCP (g/kg TDOM)                     | 71.14c            | 95.66a | 88.04b | 80.02b | 0.96 | <0.001 |

*The ratio of true digestible organic matter (mg) to gas volume (milliliters in 24 h); a, b and c are gas yield constants. SEM: standard error of the mean. P value: probability value.

a, b and c means at the same raw with different superscript letters are significantly (P<0.05) different.

**Fermentation gas yield and kinetic constants:**

Gas production reflects all nutrients fermented (soluble as well as insoluble) and fractions that are not fermentable do not contribute to the gas yield (GY). The amount of gas produced is influenced by the rate of fermentation of carbohydrate, the molar proportions of the VFA and the amount of VFA produced (Dijkstra et al., 2005). Differences in the ‘a’ and ‘c’ parameters indicate different fermentation patterns, suggesting that Y is fermented more rapidly and to a greater extent. The cumulated gas production (ml/200 mg DM) for each diet the kinetics values of gas production models are given in Table (9). The total gas yield (TGY, ml/200 mg DM) increased (P<0.001) with developing fermentation time up to 48h for all experimental diets except for G that dropped GY at 24h. The diet of Yeast (Y) increased (P<0.05) the b fraction, as also shown by the C-diet, whereas it was lower (P<0.001) with the G alone and the combination YG. In contrast, the intercept fraction (a) increased (P<0.001) in G and YG diets. The later diets showed lower rate of fermentation (c) as compared with C and Y diets. The total gas yield at 24h of incubation of Y, G and YG found to be lower than those found for C diet. The present results are lower than those reported for barley (64-71 ml), wheat (60-73 ml) and corn grains (60-82 ml) by Getachew et al. (2002), these finding might be due to applying the gas production technique herein with total mixed rations. The high gas yield as well as the OMD value of the respective additive groups could be attributed to a higher fermentation process resulting by intact the feed additives with the basal diet. Potential gas production (a) was not significantly different in study of Ikyume et al. (2018) and ranged between 15.54-27.65 ml. But, the highest value 27.65 ml was observed in the control group compared to all supplemented groups that having a similar value of 15.54 ml. The constant gas production rate (c) was also observed to be non-significant (P> 0.05) across treatment groups and ranged from 1.22 and 1.73.

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ml/hr. The highest gas production rate was observed in the 1% garlic powder group (1.73 ml/h) with the least value recorded in the control group (1.22 ml/h). Incubation time (t) was not significant (P>0.05) across the treated groups and ranged from -5.96 to -0.7h. The highest incubation time was recorded in the garlic powder 0.5% group with the least time found to be in the control group (Ikyume et al., 2018).

Higher PF values (Table 9) in respective additive diets especially G reflected high ratio of OM fermented to gas production as compared with C diet. According to Geneviève et al. (2018), the quantity of gas produced in the rumen is inversely correlated with microbial yield, which means that the PF value reflects changes in microbial biomass yield. Favorable rumen digestion responses to yeast culture feeding in ruminants include increased degradation of DM and OM (Kamel et al., 2004) and stimulation of total and cellulyolitic bacterial numbers (Newbold et al., 1995). Based on mean growth, there was a tendency for S. cerevisiae supplementation to increase rumen bacteria, whereas control treatment decreased rumen bacteria (Riyanti and Evvyernie, 2016).

**Methane yield:**

Data in Table (9) obviously showed that diets contained garlic powder (G and YG) were most effective in reduction of energy lost when expressed as a ratio of methane production to gross energy intake. Ikyume et al. (2018) found that garlic inclusion had no (p>0.05) influence on all the in vitro digestibility kinetics measured except methane (CH₄) output, Gas volume (GV), CH₄: GV ratio, metabolizable energy, and short chain fatty acids. They also found that methane to gas volume ratio expressed in percentage significantly (P<0.05) decreased with increasing amount of garlic powder supplementation (0, 0.5, 1.0 and 1.5%). While the highest value of 75 % was observed in the control group, the garlic powder 1.5% group had the least value of 19.02 %. In vitro dry matter digestibility (IDMD) was not statistically (P> 0.05) significant, GP 0.5% group had numerically higher value (68.11%) while the control had the least value of 65.23%. The addition of garlic powder to the diet of rams considerably reduced production of CH₄ by 38% (Kim et al., 2018).

**Microbial protein synthesis:**

Microbial protein (MCP) synthesized in the rumen provides the majority of the protein supplied to the small intestine of ruminants, which represents 50 to 80% of total absorbable protein (Firkins et al., 2007). The microbial protein was affected by feed additives (Table 9). Where, MCP synthesis was more enhanced (P<0.001) by yeast other than treatments. However, Wanapat et al. (2008 and 2013) showed opposite results to this finding that garlic supplementation did not produce significant changes in the synthesis of microbial proteins and/or urinary purine derivatives. The multiplication of number of bacterial by yeast supplementation may increase the rate of fermentation of the substrate and the synthesis of MCP (Ryan and Gray, 1989). The addition of yeast (Y) stimulated rumen microbial growth through the use of specific soluble growth factors such as organic acids, B vitamins and provided amino acids (Waldrip and Martin, 1993). The positive effect of the addition of SC in the present study is consistent with the findings of many workers (Guedes et al., 2008). Relating garlic addition, Ikyume et al. (2018) stated that the number of bacteria increased (P> 0.05) numerically and consequently increased the microbial protein as the level of garlic powder supplementation increased. The highest count was recorded in the garlic powder 1.5% group while the least value was observed in the control group.

**CONCLUSION**

It could be concluded that using feed additive such as dry yeast (6 gm/h/d) or garlic powder (40 gm/h/d) or its combination (3gm plus 20 gm) in finishing diets of lamb tended to increase digestibility for most of nutrients, increasing nutritive value as TDN and appeared to increase the daily gain. Furthermore these feed additives have enhanced feed efficiency and improved the immune status of animals. Although the addition of garlic powder alone is restricted by a low price case, it is most effective in reducing energy loss when expressed as a ratio between methane production and gross energy intake and also proved effective in reducing methane emissions from sheep and therefore contributing to global warming.

**REFERENCES**

Abayomi, Y.; S.S. Fagburo and S.O.K. Fajemilehin (2018). Chemical composition, phytochemical and mineral profile of garlic (*Allium sativum*). Journal of Bioscience and Biotechnology Discovery, 3(5): 105-109.
Abd El-Latif, S.A.; M.A. Toson and H.A. Mehany (2019). Effect of Dietary Onion, Garlic, Red Pepper and Anise as Natural Feed Additives on Some Hematological Studies of Japanese Quail Chicks. Acta Scientific Nutritional Health, Volume 3 Issue 9.

Abu El-Kassim, M.A.; G.A. Abd El-Hafez; S.M. Mousaand E.H. Hassan (2018). Effect of Dietary Onion, Garlic and Fenugreek Seeds Powder on Feed Intake, Blood Metabolites and Rumen Fermentation in Ossimi Ewes. Assiut J. Agric. Sci., 49 (2): 38-48.

Ahmed, B.M. and M.S. Salah (2002). Effect of yeast cultures as an additive to sheep feed on performance, digestibility, nitrogen balance and rumen fermentation. Journal of King Saudi University. Agriculture Science. 14(1):1-13.

Alnaimy, A.; A. E. Gad; M.M. Mustafa; M.A.A. Atta and H.A.M. Basuony (2017). Using of citrus by-products in farm animals feeding. Open Access J Sci., 1(3):58‒67.

Amagase H.; B.L. Petesch; H. Matsuura; S. Kasuga and Y. Itakura (2006). Intake of garlic and its bioactive components. J. Nutr., 131:955-962.

AOAC. (2005). Association of Official Analytical Chemists. Official Methods of Analysis, 18th ed., Washington, D.C. Accessed 4/25/16.

Araújo, G. L. D.; T. V. Voltolini; M. L. Chizzotti; S. H. N. Turco and F. F. R. de Carvalho (2010). Water and small ruminant production. Revista Brasileira de Zootecnia, Vol.39, (Supl. especial):326-336.

Aziz, Hend, A.; M.S. Nassar; H.S. Badway and M.H. Abd Elrahman (2018). Rumen fermentations and rumen ciliate protozoa of goat kids fed diets with different concentrate: roughage ratio. Egypt. J. Nutr. and Feeds, 21(3): 667-683.

Bampidis V.A.; V. Christodoulou; E. Christaki; P. Floroupaneri and A.B. Spais (2005). Effect of dietary garlic bulb and garlic husk supplementation on performance and carcass characteristics of growing lambs. Anim. Feed. Sci. and Tech., 121:273-283.

Baraka, T. A. (2012). Comparative Studies of Rumen pH, Total Protozoa Count, Generic and Species Composition of Ciliates in Camel, Buffalo, Cattle, Sheep and Goat in Egypt. J. American Sci., 8(2): 448-462.

Brossard, L.; F. Chaucheyras-Durand; B. Michalet-Doreau and C. Martin (2006). Dose effect of live yeasts on rumen microbial communities and fermentations during butyric latent acidosis in sheep: new type of interaction. Anim. Sci., 82: 829–836.

Bueno, M.S.; M.H.T. Watanabe; J. Issakowicz and A.C.K. Sampaio (2013). Active yeast (Saccharomyces cerevisiae) supplementation improves digestibility of lamb diet J. Agric. Vet. Sci., 2(6) (Mar. - Apr. 2013), PP 21-26.

Campanile, G.; F. Zicarelli; D. Vecchio; C. Pacelli, G. Neglia; A. Balestrieri; R. Di Palo and F. Infascelli (2008). Effects of Saccharomyces cerevisiae on in vivo organic matter digestibility and milk yield in buffalo cows. Livestock Sci., 114:358-361.

Chaucheysras, D. F.; A. Ameilbonne; P. Auffret; M. Bernard; M. Mialon; L. Dunière and E. Forano (2019). Supplementation of live yeast-based feed additive in early life promotes rumen microbial colonization and fibrolytic potential in lambs. Scientific Reports, 9:1-15.

Chaucheysras, D. F. and G. Fonty (2001). Establishment of cellulolytic bacteria and development of fermentative activities in the rumen of gnotobiotically-reared lambs receiving the microbial additive Saccharomyces cerevisiae CNCM I-1077. Reprod. Nutr. Dev., 41: 57-68.

Cole, N.A., C.W. Purdy and D.P. Hutcheson (1992). Influence of yeast culture on feeder calves and lambs. J.Anim. Sci., 70: 1682-1689.

Czerkawaski, J.W. (1986). An Introduction To Rumen Studies. Pergamon Press, Oxford, New York.

Dashak, D. A.; M.L. Dawang and N.B. Lucas (2001). An assessment of the proximate composition of locally produced spices known as dadawa basso and dadawa kawla from three markets in Plateau State of Nigeria. Food Chem., 75(2), 231- 235.

Dehoryit, B. A. (1993). Laboratory Manual For Classification And Morphology Of Rumen Ciliate Protozoa. CRC Press.
Demeyer, D.; M. De Meulemeester; K. De Graeve and B.W. Gupta (1988). Effect of fungal treatment on nutritive value of straw. Med. Fac. Landbouww. Rijksuniv. Gent, 53: 1811–1819.

Dijkstra, J.; E. Kebreab; A. Bannink; J. France and S. Lopez (2005). Application of the gas production technique in feed evaluation systems for ruminants. Anim. Feed Sci. Technol., 123–124: 561–578.

Diriba-Shiferaw, G.; R. Nigussie-Dechassa; K. Woldetsadik; G. Tabor and J.I. Sharma (2014). Bulb quality of Garlic (Allium sativum L.) as influenced by the application of inorganic fertilizers. African J. Agric. Res., 9: 778–790.

Duncan, D. B. (1955). Multiple ranges and multiple F-test. Biometrics, 11: 1-42.

Elaref, M.Y., H.A.M. Hamdon; U.A. Nayel, U.A.; A.Z.M. Salem and U.Y. Anele (2020). Influence of dietary supplementation of yeast on milk composition and lactation curve behavior of Sohagi ewes, and the growth performance of their newborn lambs. Small Ruminant Research, 1, 191: 106176.

El-Ashry, M. A.; A.M. Khoif; H.A. El-Alamy; H.M. El-Sayed and T.A. El-Hamamsy (2001). Effect of dietary supplemented with medicinal herbs on nutrient digestibility and some blood metabolities of buffalo calves. Egypt. J. Nutr. and Feeds, 4:21-33.

El-Hennawy, A.A.; C.T. Tse Wong and S.A. Kocoslis (1994). Failure of Saccharomyces boulardii to hydrolyse bile acid in vitro. Micro Bios, 80:23-29.

El-Meccawi, S.; M. Kama; A. Brosh and A.A. Degen (2009). Energy intake, heat production and energy and nitrogen balances of sheep and goats fed wheat straw as a sole diet. Livestock Sci., 125: 88–91.

El-Shereef, A.A. (2019). Blood serum biochemical changes and milk fatty acids profile due to using Garlic plant as feed additives for sheep. Asian Journal of Research in Anim. Vet. Sci., 4(4): 1-8.

Elweedah, N.M.; M.K. Mohsen; M.I. Bassiouni; M.F. Ali, and M.M. Khalafalla (2005). Performance of lambs fed on rations containing soybean meal treated with formaldehyde and probiotics. 1. Feeding value, rumen fermentation and degradability. Egyp. J. Nutr. and Feeds (Special Issue), 8:361.

Firkins, J. L.; Z. Yu and M. Morrison (2007). Ruminal nitrogen metabolism: perspectives for integration of microbiology and nutrition for dairy. J. Dairy Sci. 90 (E. Suppl.):E1-E16.

France, J.; J. Dijkstra; M.S. Dhanoa; D. Lopez; A.Z.M. Salem and R. Rojo (2000). Estimating the extent of degradation of ruminant feeds from a description of their gas production profiles observed in vitro: derivation of models and other mathematical considerations. Br. J Nutr., 83: 143-150.

Frankic, T.; M. Voljc; J. Salobir and V. Rezar (2009). Use of herbs and spices and their extract in animal nutrition. Acta agriculturae Slovenica, 94 (2): 95-102.

Gaafar H.M.A.; A.M. Mohi; M.I. Basiuoni and K.F. El-Riedy (2009). Effect of concentrate to roughage ratio and baker's yeast supplementation during hot season on performance of lactating buffaloes. Slovak, J Anim. Sci., 42(4):188-195.

Gado, H. M.; H.M. Metwally and F.A.F. Salem (1998). Effect of yeast culture and monensin on the performance of Baladi goats fed ensiled potato. Animal Agric. Sci. Ain Shams Univ., Cairo, 43(2): 419 – 429.

Galip, N. (2006). Effects of dietary Saccharomyces cerevisiae live yeast culture supplementation on ruminal digestion and protozoa count in rams fed with diets with low or high ratio forage/concentrate. Revue Méd. Vét., 157, 12: 609-613.

Garg, D.D.; T. Sharma and R.K. Dhuria (2009). Evaluation groundnut straw based complete feed blocks alone and in combination with yeast in ration of sheep. Anim. Nutr. Feed Tech., 2:9.

Geneviève, Z.; K. Adama, B. Balé; C.S. Patricia; L.N. Leando; N. Vincent, T. H. Hamadou; H. Hervé, L. Helder and A.A. Luiz (2018). In vitro rumen fermentation characteristics, methane production and rumen microbial community of two major Acacia species used in sahelian region of Burkina Faso. Tropical and Subtropical Agroecosystems, 21 (2018): 357 – 366.

Getachew, G.; G.M. Crovetto; M. Fondevila; U. Krishnamoorthy; B. Singh; M. Spanghero; H. Steingass; P.H. Robinson and M.M. Kailas (2002). Laboratory variation of 24 h in vitro gas production and estimated metabolizable energy values of ruminant feeds. Anim. Feed Sci. Tech., 102: 169–180.
Ghosh, S.; R.K. Mehla; S.K. Sirohi and R Biswatjit. (2010). The effect of dietary garlic supplementation on body weight gain, feed intake, feed conversion efficiency, fecal score, fecal coliform count and feed cost in crossbred dairy calves. Tropic. Anim. Health and Prod., 42:961-968.

Guedes, C.M.; D. Goncalves and M.A.M. Rodrigues (2008). Dias A. Effects of a Saccharomyces cerevisiae yeast on ruminal fermentation and fiber degradation of maize silages in cows. Journal of Anim. Feed Sci. Tech., 145:27-40.

Gune, M.; C. Kale; D. Bolat and S. Deniz (2016). Determination of the yield characteristics and in vitro digestibility of barley forage harvested in different vegetation periods. Indian J. Anim. Res., 50 (6): 947-950.

Hadjipanayiotou, M.; I. Antoniou and A. Photiou (1997). Effects of the inclusion of yeast culture on the performance of dairy ewes and goats and the degradation of feedstuffs. Livest. Prod. Sci., 42: 129-134.

Hassan, S.A. and S.F. Mohammed (2014). Effects of Saccharomyces cerevisiae supplementation on growth rate and nutrient digestibility in Awassi lambs fed diets with different roughage to concentrate ratios. Biochem. and Biotech. Res., Vol. 2(3), pp. 37-43.

Hermández, R.; S. S. González; J.M. Pinos-Rodríguez; M.E. Ortega; A. Hernández ; G. Bueno and M. Cobos (2009). Effect of a yeast culture on nitrogen balance and digestion in lambs fed early and mature orchard grass. J. Appl. Anim. Res., 35: 53-56.

Hungate, R.E. (1966). The Rumen Protozoa. The Rumen And Its Microbes, pp. 92-147. New York: Academic Press.

Hyun-Sun, Y.; Ellis, M., Curtis, S. E. and Johnson, R. W. (2009). Environmental temperature, space allowance, and regrouping: Additive effects of multiple concurrent stressors in growing pigs. J. Swine Health and Prod., 13 (3): 131-138.

Ibrahim E.M. and El Naggar, Soad (2018). Nutrient digestibility, productive performance and some serum biochemical indicators as affected by substitution of soybean meal for inactive dry yeast in growing lambs diet. Egypt. J. Nutr. Feeds, 21 (2): 345-353.

Ikyume, T.T.; O.S. Sowande; P.A. Dele; A.O. Yusuf; S. Monday; O.K. Egunjobi and O. Fatoba (2017). Effect of varying levels of garlic (Allium sativum) powder on growth, apparent nutrient digestibility, rumen ecology, blood profile and cost analysis of feeding West African Dwarf goats. Mal. J. Anim. Sci., 20 (2): 61-74.

Ikyume, T. T.; O.S. Sowande; P.A. Dele; A.O. Yusuf; S. Monday; O.K. Egunjobi and T. Fatoba (2018). In Vitro fermentation and rumen microbial count of West African Dwarf goats fed garlic (Allium sativum) powder. Bull. Anim. Hlth. Prod. Afr., (2018), 66, 491-499

Ismaiel, A.M.; A.H. El-Far and I.I. Abou-Ganema (2010). Effect of Tonilisat and Roemin W2 supplementations on the performance of lambs. Int. J. Bio. and Life Sci., 6 (4):222-229

Kamel, H. E.M.; J. Sekine; A.M. El-Waziry and M.H.M Yacout (2004). Effect of Saccharomyces cerevisiae on the synchronization of organic matter and nitrogen degradation kinetics and microbial nitrogen synthesis in sheep fed Berseem hay (Trifolium alexandrinum). Small Ruminant Research, 52: 211–216.

Kawas J.R.; R. Garcia-Castillo; F. GarzaCazares; H. Fimbres-Durazo; E. Olivares Saenz; G. Hernandez-Vidal and C.D. Lu (2007). Effects of sodium bicarbonate and yeasts on productive performance and carcass characteristics of light – weight lambs fed finishing diets. Small Ruminant Research. 67:157-163

Kewan, K.Z.; F.A. Salem; A.Z.M. Salem; A.R. Abdou; H.M. El-Sayed; S.S. Eisa; E.A. Zaki and N.E. Odongo (2019). Nutritive utilization of Moringa oleifera tree stalks treated with fungis and yeast to replace clover hay in growing lambs. Agroforest Syst., 93:161–173.

Kewan; K.Z.; Ahlam. R. Abdou; E.A. Zaki; F.A. Salem; H.M. El-Sayed and S.S. Eisa, S.S. (2017). Using of biological treatments to enhance nutrients of Moringa stalks before utilized in ruminant feeding as a sole diet. Res. J. Anim. Vet. Sci., 9(2): 16-22.

Khale, N.F and T.A. Baraka (2011). Influence of TOMOKO® (Direct-Fed Microbials) on productive performance, selected rumen and blood constituents in Barky finishing lambs. J. Amer. Sci., 7: 564-570.
Khater, H.F.; M.Y. Ramadan and R.S. El-Madawy (2009). Lousicidal, ovi-cidal and repellent efficacy of some essential oils against lice and flies infesting water buffaloes in Egypt. Vet. Parasitol, 164: 257–266.

Khattab, H.M.; S.A.H. Abo El-Nor; S.M. Kholif; H.M. El-Sayed; O.H. Abd El-Shaifyy and M. Saada (2010). Effect of different additive sources on milk yield and composition of lactating buffaloes. Lives. Sci., 131: 8-14.

Kholif S.M.; T.A. Morsy; M.M. Abdo; O.H. Matloup and A.A. Abu El-Ella (2012). Effect of supplementing lactating goats rations with garlic, cinnamon or ginger oils on milk yield, milk composition and milk fatty acids profile. J. Life Sci., 4(1):27-34.

Khosravi, A.; F. Boldaji; B. Dastar and S. Hasani (2010). Immune response and performance of broiler chicks fed probiotics and propionic acid. Internat. J. Poult. Sci., 9: 188-191.

Kim, J.Y.; Ghassemi Nejad J.; J.Y. Park; B.H. Lee; M. Hanada; B.W. Kim and K.I. Sung (2018). In vivo evaluation of garlic (Allium sativum) supplementation to rice straw-based diet on mitigation of CH4 and CO2 emissions and blood profiles using crossbreed rams. J. Sci. Food Agric., 98: 5197–5204.

Klevenhusen F.; S. Duval; J.O. Zeitz; M. Kreuzer and C.R. Soliva (2011). Diallyl disulphide and lovastatin: effects on energy and protein utilisation in, as well as methane emission from sheep. Arch. Anim. Nutr., 65 (4): 255–266.

Kolmer, J.A.; I.H. Spouling and H.W. Robinson (1951). Approved Laboratory Techniques (5th Ed). New York: Appleton-Century Crofts. Pp: 1180.

Komonna, O.F.A. (2007). Phsiological and nutritional responses of sheep to some feed additives. Ph.D. Thesis, Fac. Agric., Minufiya University.

Kongmun P.; M. Wanapat; P. Pakdee; C. Navanukraw (2010). Effect of coconut oil and garlic powder on in vitro fermentation using gas production technique. Livestock Sci., 127: 38-44.

Lascano, G.J., and A.J. Heinrichs (2009). Rumen fermentation pattern of dairy heifers fed restricted amounts of low, medium, and high concentrate diets without and with yeast culture. Livestock Sci., 124:48–57.

Lascano G.J.; G.I. Zanton; F.X. Suarez-Mena and A.J. Heinrichs (2009). Effect of limit feeding high and low-concentrate diets with Saccharomyces cerevisiae on digestibility and on dairy heifer growth and first-lactation performance. J. Dairy Sci., 92:5100–5010.

Maamouri, O.; B. Jemmali; I. Badri; H. Selmi and H. Rouissi. (2014). Effects of yeast (Saccharomyces cerevisiae) feed supplement on growth performances in "Queue Fine de l'Ouest" lambs. J. New Sci., 8(1).

Mabjeesh, S. J.; M. Cohen and A. Arieli (2000). In Vitro Methods for Measuring the Dry Matter Digestibility of Ruminant Feedstuffs: Comparison of Methods and Inoculum Source. J. Dairy Sci., 83:2289–2294.

Macfarlane, W.V. and B. Howard (1970). Water in the physiological ecology of ruminants. In: Physiology of digestion and metabolism in the ruminant. A.T. Phillipson Editor, Oriel Press. England. pp.217-229.

Malekkahi, M.; A.M. Tahmasbi; A.A. Naserian; M. Danesh Mesgaran; J.L. Kleen and A.A. Parand, (2015). Effects of essential oils, yeast culture and malate on rumen fermentation, blood metabolites, growth performance and nutrient digestibility of Baluchi lambs fed high-concentrate diets. J. Anim. Physiol. Anim. Nutr., 99; 221–229.

Mariam, M.B. and U.C. Devi (2016). Chemical and shelf life analysis of dry garlic powder: A golden Herb. Internat. J. Agric. Food Sci. Tech. 7(1): 1-6.

Menke K.H., Raab L., Salewski A., Steingass H., Fritz D. and Schneider W. (1979). The estimation of the digestibility and metabolizable energy content of ruminant feeding stuffs from the gas production when they are incubated with rumen liquor in vitro. J. Agric. Sci., 93:217–22.

Milewski, S. (2009). Effect of yeast preparations Saccharomyces cerevisiae on meat performance traits and blood hematological indices in suckling lambs. Med Wet; 65: 51–54.

Mungoi, M.; C. Flores; R. Casals and G. Caja (2012). Effect of malate and starch source on digestibility and nutrient balance of growing- fattening lambs. Anim. Feed Sci. Tech., 22: 154–162.
Kewan et al.

Nassar, M.; A. El Shereef and S. Abo Bak. (2017). Influence of feeding garlic plant either as powder or oil on reproductive performance of ewes. GSC Biol. Pharma. Sci., 01(03):059-061.

Nehring, K. and G. F. W. Haenlein (1973). Feed evaluation and ration calculation based on net energy fat. Journal of Animal Science, 36(5): 949-964.

Newbold, C. J.; R.J. Wallace; X.B. Chen and F.M. McIntosh (1995). Different strains of Saccharomyces cerevisiae differ in their effects on ruminal bacterial numbers in-vitro and in sheep. Canadian J. Anim. Sci., 73: 1811–1818.

Nociek, J.E; W.P. Kautz; J.A.Z. Leedle and J.G. Allman (2002). Ruminal supplementation of direct-fed microbials on diurnal pH variation and in situ digestion in dairy cattle. J. Dairy Sci., 85: 429-433.

NRC (1985). National Research Council, Nutrient Requirements of Sheep, Sixth Revised Edition. National Academy Press, Washington, DC, USA.

Nwinuka, N. M.; G.O. Ibeh and G.I. Ekeke (2005). Proximate composition and levels of some toxicants in four commonly consumed spices. J. Appl. Sci. Environ. Manage., 9(1), 150-155.

Ogimoto, K. and S. Imai (1981). Atlas of rumen microbiology. Japan Scientific Soc. Press, Tokyo.

Otunola, G. A.; O.B. Oloyede; T. Adenike; T. Oladiji and A.J. Afolayan (2010). Comparative analysis of the chemical composition of three spices Allium sativum, Zingiber officinale Rosc. and Capsicum frutescens L. commonly consumed in Nigeria. Afr. J. Biotechnol., 9(1), 6921-6927.

Panghal M, V.; Kaushal and J.P. Yadav (2011). In vitro antimicrobial activity of ten medicinal plants against clinical isolates of oral cancer cases. Annals of Clinic. Microbiol. Antimicro. 2011;10: 21.

Payandeh, S. and F. Kafizadeh (2007). The effect of yeast (Saccharomyces cerevisiae) on nutritional intake, digestibility and finishing performance of lambs fed a diet based on dried molasses sugar beet-pulp. Pakistan J. Biol. Sci., 10(2): 4426-4431.

Petropoulos, S.A.; A. Fernandes; G. Ntatsi; K. Petrotos; L. Barros and I.C.F.R. Ferreira (2018). Nutritional value, chemical characterization, and bulb morphology of Greek Garlic Landraces. Molecules, 23, 319

Plata, P. F.; M.G.D. Mendoza; J.R.B. Gama and M.S. Gonzalez (1994). Effect of a yeast culture (Saccharomyces cerevisiae) on neutral detergent fiber digestion in steers fed oat straw-based diets. Anim. Feed Sci. Tech., 49:3-4: 203-210.

Rattray, P.V.; W.N. Garrett; N. Hinman; I. Garcia and J. Castillo (1973). A system for expressing the net energy requirements and net energy content of feeds for young sheep. J. Anim. Sci., 36: 115.

Rivlin, R.S. (2001). Historical perspective on the use of garlic. J. Nutr., 131: 951S–954S.

Ryan J. P. and W.R. Gray (1989). Effect of high strength yeast culture ab initio utilizes a residual source of volatile fatty acids in strained ruminal fluid from hay-fed sheep. Biochemical Society Transactions, 17 (2): 392–393.

Ryan J. P. and W.R. Gray (1989). Effect of high strength yeast culture ab initio utilizes a residual source of volatile fatty acids in strained ruminal fluid from hay-fed sheep. Biochemical Society Transactions, 17 (2): 392–393.

Sahli, F.; C. Darej and N. Moujahed (2018). Potential of white garlic powder (Allium sativum L.) to modify in vitro ruminal fermentation. South African J. Anim. Sci., 48 (No. 2): 253-260.

Sallam, S. M.A.; A.M. Allam and S.A. Najadi (2014). Comparison of two products of direct-fed microbial supplementation on the nutrient utilization and ruminal fermentation in sheep. J. Agri. Sci., 6 (3): 159-167.

SAS (2002). Statistical Analysis Systems Institute Inc., Release 8.1, Cary, NC, USA.
Sivam, G.P. (2001). Protection against Helicobacter pylori and other bacterial infections by garlic. J. Nutr., 131: 1106S–1108S.

Strickland, V.J.; G.L. Krebs and W. Potts (2009). Pumpkin kernel and garlic as alternative treatments for the control of Haemonchus contortus in sheep. Anim. Prod. Sci., 49: 139-144.

Tatara, M. R.; E. Sliwa; K. Dudek; A. Gawron and T. Piersiak (2008). Aged garlic extract and allicin improve performance and gastrointestinal tract development of piglets reared in artificial sow. Ann. Agric. Environ. Med., 15:63-69.

Thomas, L. (1998). Immunoglobulins (Ig). In: Thomas L, ed. Clinical laboratory diagnostics. Use and assessment of clinical laboratory results. Frankfurt/Main: TH-Books Verlagsgesellschaft, 667-678.

Trevisi, E and G. Bertoni (2009). Some physiological and biochemical methods for acute and chronic stress evaluation in dairy cows. Ital. J. Anim. Sci., vol. 8 (Suppl. 1): 265-286

Van Soest, P.J. (1982). Nutritional Ecology of the Ruminant, 2nd ed. Cornell University Press, New York, pp. 266, 301.

Waldrip, H.M. and S.A. Martin (1993). Effects of an Aspergillus oryzae fermentation extract and other factors on lactate utilization by the ruminal bacterium Meganphaera elsdenii. J. Anim. Sci., 71:2770.

Wanapat, M., S. Kang, P. Khejornsart and S. Wanapat (2013). Effects of plant herb combination supplementation on rumen fermentation and nutrient digestibility in beef cattle. Asian Australas. J. Anim. Sci., 26 (8): 1127-1136.

Wanapat, M., P. Khejornsart, P. Parkdee and S. Wanapat (2008). Effect of supplementation of garlic powder on rumen ecology and digestibility of nutrients in ruminants. J. Sci. Food Agric., 88:2231-2237.

Warner, A.C.I. (1964). Production of volatile fatty acids in the rumen, methods of measurement. Nutr. Abst. and Rev., 34; 339.

Yang, W. Z; C. Benchaar; B.N. Ametaj; A.V. Chaves; M.L. He and T.A. McAllister (2007). Effects of garlic and juniper berry essential oils on ruminal fermentation and on the site and extent of digestion in lactating cows. J. Dairy Sci., 90:5671–5681.

Yang, W.Z., K.A. Beauchemin; D.D. Vedres; G.R. Ghorbani; D. Colombatto and D.P. Morgavi (2004). Effects of direct-fed microbial supplementation on ruminal acidosis, digestibility, and bacterial protein synthesis in continuous culture. Anim. Feed Sci. and Tech., 114; 179–193.

Zaki, E.A. (2016). Nutritional performance of small ruminants fed by products of moringa under desert environmental conditions. Ph. D. Thesis, Institute Studies of Environmental Studies and Research, Ain Shams University, Egypt.

Zeid, A.M.M.; M.K. Mohsen; M.A. Ibrahim and A.M.E. Elkamhawy (2011). Effect of feeding rations supplemented with chamomile flowers and dried yeast on productive performance of sheep. J. Animal and Poultry Prod., Mansoura Univ., 2 (6): 167-184, 2011

Zhong, R.; H. Xiang; L. Cheng; C. Zhao; F. Wang; X. Zhao and Y. Fang (2019). Effects of feeding Garlic powder on growth performance, rumen fermentation, and the health status of lambs infected by gastrointestinal Nematodes. Animals, 9 (3) 102: 122.
تأثير الخمرة والثوم أو مخلوطهما كإضافات غذائية على الأداء وتغذية الكرش واللحوم للحامل

المقدمة

تهدف هذه الدراسة إلى تقييم تأثير الخمرة والثوم والثوم مخلوطهما على الأداء وتكوين الكرش واللحوم للحامل. أُجريت على 36 ذكرًا و36 أنثى كل منها 8 شهورًا من مختبر مزرعة الكبد الجديدة. الأشخاص الذين تم اختيارهم كانوا من منتجات ذات جودة عالية والمتنوعة. تم تقسيمهم إلى ثلاث مجموعات: مجموعة البحوث الأولى التي لم تتم تجربة ثومًا أو خمرة، مجموعة البحوث الثانية التي تم استخدام ثومًا فقط، وثالث مجموعة المختبر التي تم استخدام خمرة فقط. م schizophrenية الضغط كانت على حدود 50% مع منجمات المختبر المتنوعة في الفئة العالية للمحميات.

المتتالي، الأمثلية الفيدلية (ألفة عالية/المقدمة) ، علقت أساسية مع إضافة 6 جرام خمرة/ رأس/ يوم ، علقة أساسية مع إضافة 35.8 جرام خمرة + 30 جرام ثوم/ رأس/ يوم ، علقة أساسية مع إضافة 70% حمض الدهني الفيدل. النتائج تشير إلى أن النتينات كانت جيدة نسبيًا بالنسبة لجميع المجموعات.

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