Comparative study between liver biopsy and non-invasive biomarkers in assessment of hepatic fibrosis in children with chronic liver diseases

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Abstract

Background: Liver biopsy is the gold standard for detecting the degree of liver fibrosis; however, invasiveness constitutes its main limiting factor in clinical application, so we aimed to evaluate the non-invasive biomarker formulas (APRI and FIB-4) and their modified forms by BMI z-score (M-APRI, M-FIB-4, and B-AST) compared to liver biopsy in the assessment of liver fibrosis in children with chronic liver diseases. Two hundred children aged 6.3 ± 3.8 years (98 males, 102 females) with chronic liver diseases underwent liver biopsy. The stage of fibrosis was assessed according to the METAVIR system for all children, and the following non-invasive biomarker formulas were calculated: APRI, modified APRI (M-APRI: BMI z-score × APRI), Fibrosis-4 index (FIB-4), modified FIB-4 (M-FIB-4: BMI z-score × FIB-4), and B-AST (BMI z-score × AST). The best cutoff value was calculated to detect early fibrosis (F1–F2) from advanced liver fibrosis (F3–F4).

Results: There were positive correlations between all studied non-invasive biomarker models (APRI, FIB-4, M-APRI, M-FIB-4, B-AST) and fibrosis score as an increase in fibrosis score was associated with an increase in mean ± SD of all studied biomarker formulas. The best cutoff values of non-invasive biomarker models in the diagnosis of early fibrosis (F1–F2) were APRI > 0.96, M-APRI > 0.16, FIB-4 > 0.019, M-FIB-4 > 0.005, and B-AST > −8 with an area under the curve above 0.7 each, while the best cutoff values of non-invasive biomarker models (APRI, M-APRI, FIB-4, M-FIB-4, and B-AST) in the diagnosis of advanced liver fibrosis (F3–F4) were >1.96, >2.2, >0.045, and >0.015, >92.1, respectively, with an area under the curve above 0.8 each.

Conclusion: APRI, M-APRI, FIB-4, M-FIB-4, and B-AST are good non-invasive alternatives to liver biopsy in the detection of liver fibrosis in children with chronic liver diseases of different etiologies especially those that include BMI z-scores in their formulas.

Keywords: Chronic liver diseases, Children, Liver biopsy, Fibrosis, Non-invasive biomarkers

Background

The degree and progression of liver fibrosis are major determents of the outcome of liver disease in children. Liver biopsy is still the gold standard for diagnosis, grading, and staging of liver fibrosis [1]. However, liver biopsy is restricted being an invasive technique that might result in patient hazards such as pain, bleeding, and biliary system injury, along with variable accessibility, high cost, sampling mistakes, and inaccurate due to inconsistency of pathologic interpretations [2]. These limitations promote the development of less invasive methods to assess liver fibrosis especially in children [3].

Hepatic fibrosis scores and serum biomarkers can serve as non-invasive, reproducible, and sensitive
screening tools to predict fibrosis. Fibrosis scores such as aspartate aminotransferase/platelet ratio index (APRI) and fibrosis index based on four factors (Fibrosis-4 index, FIB-4) were developed based on the progression of liver pathology to cirrhosis and derived from the Apricot database in patients with chronic hepatitis C virus (HCV) infection [4]. Many of serum fibrosis models have been used to evaluate liver fibrosis and cirrhosis in adults with chronic hepatitis C. However, limited data on their accuracy in pediatric patients is available, and none of them has been fully validated in children [5]. Therefore, in this study, we aimed at the evaluation of non-invasive serum biomarker fibrosis models (APRI and FIB-4) and their modified forms by BMI z-score (M-APRI, M-FIB-4, and B-AST), compared to liver biopsy to assess liver fibrosis in children with chronic liver diseases.

Methods

Subjects
This was a 5-year observational cross-sectional study performed on 200 children suffering from various chronic liver diseases (a long-standing irreversible change in the hepatic structure that may end in complications like cirrhosis leading to premature death) [6], and their diagnosis was based on clinical, laboratory, and histopathological examination. Patients were recruited from the outpatient pediatric hepatology clinic between May 2015 and May 2019. Any patient with chronic liver disease of different etiologies (chronic viral hepatitis B or/and C, cholestatic liver disease, autoimmune hepatitis, and metabolic liver diseases) who underwent liver biopsy was included in the study. Patients were excluded if they had chronic liver disease with other co-morbidity as renal or C.V.S. or C.N.S. affection. This study was approved by the Ethical Committee of Faculty of Medicine, in accordance with “The Code of Ethics of the World Medical Association” (Declaration of Helsinki). Informed consent was obtained from parents prior to enrollment in the study after being fully informed about the study and its procedures.

Methods

- All enrolled children were subjected to the following:
  - **Full history taking and thorough clinical examination**: with emphasis on clinical presentations (jaundice, abdominal pain and distension, melena and bleeding manifestations as hematuria, epistaxis, or bleeding gums), abdominal examination, and anthropometric measurements including weight, height, and BMI. Patients’ weights were measured in kilograms (to the nearest 100 g) using an electronic digital scale which accuracy was periodically verified using reference weights, while their heights were measured in centimeters (to the nearest millimeter). Children were measured on scales with height gauges, and the subject stood with back against the gauge and feet on the weighing platform. A software program that combines the raw data of the variables (age, sex, height, and weight) to compute a nutritional status index such as z-score, weight-for-height, weight-for-age, and height-for-age was used [7]. Patients’ weights and heights were recorded on the day of the biomarker examination. BMI z-scores were calculated using the World Health Organization (WHO) Child Growth Standards and Growth Reference data with the WHO anthropometric calculator, AnthroPlus v.1.0.4.

**Laboratory investigations**: 9 ml venous blood was drawn by aseptic venipuncture using a disposable sterile syringe. Blood was divided in different vacutainer tubes as follows: 3 ml on K3-EDTA for CBC (CELL-DYN Emerald Hematology Analyzer, Abbott, IL, USA); 3 ml on sodium-citrate for prothrombin time (PT), INR, and activated partial thromboplastin time (∼aPTT) (STA Compact Max, STAGO, France); and 3 ml on the plain tube that was allowed to clot at room temperature for 30 min then serum was separated by centrifugation on 1262 g for 10 min for biochemical and serological testing including:

- Liver function tests: aspartate aminotransferase (AST), alanine aminotransferase (ALT), alkaline phosphatase (ALP), gamma-glutamyl transpeptidase (GGT), total and direct bilirubin, serum protein, and albumin (Biosystem A15 Auto-analyzer, BioSystem, Spain)
- Hepatitis B surface antigen (HBsAg) and anti-HCV antibody (HCV-Ab) by enzyme-linked immunosorbent assay (ELISA) were done and confirmation of HCV and HBV infection was done by using PCR (Biokit, Spain)
- Serum immunoglobulin G (IgG) by radial immunodiffusion (RID) (Binding Site, Birmingham, UK)
- Qualitative determination of anti-nuclear antibodies (ANA), anti-smooth muscle antibodies (ASMA), liver-kidney microsome antibodies (LKM-1), and anti-mitochondrial antibodies (AMA) by indirect immunofluorescence (IIF) (Nova Lite, INOVA diagnostics, USA)
- Work-up for Wilson’s disease (serum ceruloplasmin, 24-h urinary copper before and after penicillamine) and Keyser-Fleischer ring
Liver biopsy: Ultrasound-guided sample of living tissue of the liver was done for all patients either for diagnosis (when the above investigations were not enough for diagnosis) and/or detection of the degree of fibrosis using Menghini aspiration needle (Hepafix Luer Lock Braun Melsungen AG, Melsungen, Germany). A good enough core that contains at least five portal tracts was obtained. Biopsy specimens were fixed in formalin and deeply set within paraffin. Five-micrometer-length sections were cut and stained with hematoxylin and eosin to assess the histological activity of liver disease using the METAVIR scoring system [8] which comprises of five stages: F0 (no fibrosis), F1 (minimal fibrosis, portal fibrosis without septa), F2 (moderate fibrosis, portal fibrosis with few septa), F3 (severe fibrosis, septal fibrosis with many septa but no cirrhosis), and F4 (cirrhosis). Sections were stained with Mason-Trichrome to assess the amount of iron in the liver. Prussian blue stain to show iron deposition, and periodic acid-chief stain to exclude alpha 1 anti-trypsin deficiency. Slides were blindly evaluated by two pathologists.

Non-invasive serum biomarker models for fibrosis evaluation: were calculated according to the published analytic recommendations [9, 10] as follows:

- Aspartate transaminase (AST)-to-platelet ratio index (APRI):
  \[ \text{APRI} = \left( \frac{\text{AST (IU/L)}}{\text{AST ULN (IU/L)}} \right) / \text{platelet count (10}^9/\text{L}) \times 100 \]
- Fibrosis-4 index (FIB-4):
  \[ \text{FIB-4} = \left[ \frac{\text{age (years)} \times \text{AST (IU/L)}}{\text{platelet count (10}^9/\text{L}) \times \sqrt{\text{ALT (IU/L)}}} \right] \]
- The modified forms by BMI z-score:
  \[ \text{M-APRI} = \text{APRI} \times \text{BMI z-score} \]
  \[ \text{M-FIB-4} = \text{FIB-4} \times \text{BMI z-score} \]
  \[ \text{B-AST} = \text{B-AST (IU/L)} \times \text{BMI z-score} \]

The upper limits of normal (ULN) for ALT and AST were established as 40 IU/L, and the age used in formula and laboratory investigations was at the time of liver biopsy.

Statistical methods
The collected data were tabulated and analyzed using SPSS software, ver. 16 (SPSS Inc., Chicago, IL, USA) and MedCalc statistical software, ver. 17.2 (MedCalc Software, Mariakerke, Belgium). Categorical data were presented as numbers and percentages while quantitative data were expressed as mean ± standard deviation, median, interquartile range (IQR), and range. Quantitative data were tested for normality using the Kolmogorov-Smirnov test, assuming normality at \( P > 0.05 \). Differences between groups were tested using the Kruskal-Wallis (KW) test for non-parametric variables; when significant, it was followed by post hoc multiple comparisons using the Bonferroni test to detect the significant pairs. Spearman’s correlation coefficient (rho) was used to assess the correlation between non-parametric variables. ROC curve analysis was used to detect cutoff values for the studied markers with optimum sensitivity and specificity in the prediction of early fibrosis and advanced fibrosis. The accepted level of significance in this work was stated at 0.05 (\( P < 0.05 \) was considered significant).

Results
This study was carried out on 200 children suffering from chronic liver disease of different etiologies; they were 98 male and 102 female, aged 6.3 ± 3.8 years. Forty percent of them had positive consanguinity and 30% had a family history of chronic liver disease. Among the studied patients, 25% were diagnosed with metabolic liver diseases (MLD) [glycogen storage disease 17%, Niemann-pick disease 2%, Wilson disease 6%], 44% were diagnosed with chronic hepatitis [autoimmune hepatitis 15%, chronic hepatitis of unknown etiology 13%, steatohepatitis 8%, congenital hepatic fibrosis 8%], 15% were diagnosed with viral hepatitis [HCV 10%, HBV 5%], and 16% had cholestasis disorders (progressive familial intra-hepatic cholestasis 5%, Alagille syndrome 6%, alpha 1 anti-trypsin deficiency 1%, Caroli’s disease 2%, Dubin-Johnson syndrome 2%). The most clinical presentations of the studied hepatic patients were abdominal distension (60%), jaundice (44%), pallor (21%), hepatomegaly (65%), and splenomegaly (16%). Anthropometric measurements are shown in Table 1.

The baseline data of liver function tests in the studied hepatic patients at the time of liver biopsy were found to be elevated above their upper limit of normal (ULN) (Table 1).

Histopathological evaluation of liver biopsy specimens revealed that the majority of the studied patients (81%) showed mild disease activity. Regarding the degree of fibrosis, 41% had minimal fibrosis (F1), 19% had moderate fibrosis (F2), and severe fibrosis (F3) was present in 9% of patients (glycogen storage disease 5 patients, Wilson disease 4 patients, autoimmune hepatitis 5 patients, chronic hepatitis of unknown etiology 3 patients, alpha 1 anti-trypsin deficiency 1 patient), while cirrhosis (F4) was observed in 9% of cases (congenital hepatic fibrosis 8 patients, chronic hepatitis of unknown etiology 4 patients, autoimmune hepatitis 6 patients) (Table 2).

The median, interquartile range (IQR) of the five non-invasive serum biomarkers in our study group is shown in Table 3.

The studied non-invasive serum biomarkers (APRI, FIB-4, M-APRI, M-FIB-4, B-AST) showed significant positive correlations with HAI and fibrosis stage. The
Table 1: Clinical and laboratory characteristics of the study group at the time of liver biopsy

| Variable          | Hepatic patients (N = 200) |
|-------------------|-----------------------------|
| Age (years)       | 6.3 ± 3.8 (2–15)            |
| Sex               | Male                        |
|                   | Female                      |
| BMI (kg/m²)       | 19.3 ± 7.3 (13.0–50.7)      |
| Height (cm)       | 95.4 ± 32.4 (70.0–149.0)    |
| Weight (kg)       | 18.2 ± 5.9 (9.0–40.0)       |
| Sex               | 98 (49%)                    |
|                  | 102 (51%)                   |
| HAI score         | 0.12 ± 2.9 (<48.6)          |
| ALT (IU/L)        | 142.8 ± 198.8 (20–1080)     |
| AST (IU/L)        | 155.7 ± 205.8 (18–1170)     |
| D. bilirubin (mg/dL) | 4.5 ± 5.03 (0.2–21.0)    |
| T. bilirubin (mg/dL) | 2.6 ± 3.1 (0.1–13.0)    |
| ALP (IU/L)        | 520.6 ± 493 (80–1192)       |
| GGT (IU/L)        | 73.7 ± 119.5 (41–668)       |
| Hb (g/dL)         | 9.6 ± 1.7 (8.5–11.0)        |
| PLTs (10³/μL)     | 246.5 ± 137.9 (150.0–435.0) |

Data expressed as mean ± SD (range). BMI: body mass index, AST: aspartate aminotransferase, ALT: alanine transaminase, T. bilirubin: total bilirubin, D. bilirubin: direct bilirubin, ALP: alkaline phosphatase, GGT: gamma-glutamyl transferase, Hb: hemoglobin, PLTs: platelets.

increase in the fibrosis score was associated with an increase in mean ± SD of all five non-invasive biomarkers as the APRI score increases from 0.48 ± 0.51 in F0 to 8.11 ± 5.29 in F4, the FIB-4 score increases from 0.008 ± 0.010 in F0 to 0.145 ± 0.078 in F4, the M-APRI score increases from −2.03 ± 2.88 in F0 to 3.90 ± 1.79 in F4, M-FIB-4 increases from −0.012 ± 0.037 in F0 to 0.56 ± 0.071 in F4, and B-AST increases from −213.4 ± 369.8 in F0 to 93.60 ± 59.1 in F4.

Moreover, APRI showed a significant positive correlation with liver enzymes (ALT, ALP, and GGT) and serum bilirubin (total and direct) levels, while M-APRI showed a significant positive correlation with age of patient and serum total bilirubin. The FIB-4 index was found to be positively correlated with serum ALP level. There was a significant positive correlation between B-AST and total bilirubin levels (Table 4).

In order to assess the performance of the studied non-invasive biomarkers in the detection of early fibrosis (F1–F2) and detection of advanced liver fibrosis (F3–F4), it was found that the best cutoff for APRI in the detection of early fibrosis (F1–F2) was > 0.96 with AUC 0.745, sensitivity 46.4%, and specificity 91.9%, and in the detection of advanced liver fibrosis (F3–F4), the best cutoff point was >1.96, with AUC 0.849, sensitivity 89.9%, and specificity 84.5%, while for M-APRI, the best cutoff for detection of early fibrosis (F1–F2) was > 0.16 with AUC 0.831, sensitivity 61.3%, and specificity 96.5%, and for the detection of advanced liver fibrosis (F2–F4), the cutoff point was >2.2 with AUC 0.958, sensitivity 87.8%, and specificity 96.6%. In FIB-4, early fibrosis (F1–F2) could be detected at a cutoff point >0.019 with AUC 0.750, sensitivity 54.8%, and specificity 94.5%, while advanced liver fibrosis (F3–F4) could be detected at a cutoff point >0.045 with AUC 0.853, sensitivity 78.8%, and specificity 85.6%. Using M-FIB-4, at a cutoff value >0.004, early fibrosis (F1–F2) could be detected with AUC 0.759, sensitivity 49.7%, and specificity 96.5%, while at a cutoff value > 0.015 with AUC 0.896, sensitivity 89.8%, and specificity 82.4%, advanced liver fibrosis (F3–F4) could be detected. The values of B-AST for detection of early fibrosis (F1–F2) was >8 with AUC 0.758, sensitivity 74.2%, and specificity 75.7%, while in the detection of advanced liver fibrosis (F3–F4), the cutoff point was > 92.1, with AUC 0.955, sensitivity 89.8%, and specificity 91.2% (Tables 5 and 6, Figs. 1 and 2).

Discussion

Liver biopsy is an approved procedure that is indicated to establish a diagnosis, assess the prognosis, and

Table 2: Histopathological evaluation of the liver biopsy specimens among the study group according to METAVIR scoring system

| Variable                   | Hepatic patients (N = 200) | % (100%) |
|----------------------------|-----------------------------|----------|
| HAI                        |                            | 12.0%    |
| A0 = no activity           | 24                          |          |
| A1 = mild activity         | 162                         | 81.0%    |
| A2 = moderate activity     | 8                           | 4.0%     |
| A3 = severe activity       | 6                           | 3.0%     |
| No fibrosis                | F0 44                       | 22.0%    |
| Minimal fibrosis           | F1 82                       | 41.0%    |
| Moderate fibrosis          | F2 38                       | 19.0%    |
| Severe fibrosis            | F3 18                       | 9.0%     |
| Cirrhosis                  | F4 18                       | 9.0%     |

HAI: histological activity index

Table 3: Non-invasive biomarkers in the study group

| Non-invasive biomarkers | Median | Interquartile range (IQR) |
|------------------------|--------|---------------------------|
| APRI                   | 0.6    | 0.24–1.7                  |
| FIB-4                  | 0.01   | 0.04–0.05                 |
| M-APRI                 | 0.16   | −0.64–0.65                |
| M-FIB-4                | 0.001  | −0.004–0.015              |
| B-AST                  | 15.65  | −84.4–64.5                |

APRI: AST-platelet ratio index, FIB-4: Fibrosis-4 index, M-APRI: modified APRI, M-FIB-4: modified FIB-4, B-AST: AST in relation to BMI.
Table 4 Correlation between the non-invasive biomarkers and lab investigations and histopathological evaluation in the study group

|                      | APRI          | M-APRI        | FIB-4         | M-FIB-4       | B-AST         |
|----------------------|---------------|---------------|---------------|---------------|---------------|
|                      | Rho | P   | Rho | P   | Rho | P   | Rho | P   | Rho | P   |
| Age (years)          |    0.079       | 0.27          | 0.321         | 0.036         | –              | –              | –              | –              | 0.147       | 0.18          |
| ALT (IU/L)           |    0.649       | <0.001        | 0.018         | 0.85          | –              | –              | –              | –              | 0.018       | 0.85          |
| T. bilirubin (mg/dL) |    0.297       | 0.003         | 0.235         | 0.018         | 0.156          | 0.12           | 0.189          | 0.059          | 0.223       | 0.026         |
| D. bilirubin (mg/dL) |    0.314       | 0.002         | 0.186         | 0.67          | 0.173          | 0.08           | 0.167          | 0.098          | 0.160       | 0.11          |
| ALP (IU/L)           |    0.340       | 0.001         | 0.130         | 0.19          | 0.223          | 0.034          | 0.102          | 0.31           | 0.125       | 0.21          |
| GGT (IU/L)           |    0.253       | 0.011         | 0.107         | 0.29          | 0.179          | 0.074          | –0.063         | 0.63           | 0.08        | 0.37          |
| Hb (g/dL)            |   –0.007       | 0.94          | –0.076        | 0.45          | –0.055         | 0.62           | –0.135         | 0.18           | –0.136      | 0.18          |
| HAI                   |    0.326       | 0.001         | 0.418         | <0.001        | 0.316          | 0.002          | 0.343          | <0.001         | 0.327       | 0.0013        |
| Fibrosis stage       |    0.568       | <0.001        | 0.629         | <0.001        | 0.473          | <0.001         | 0.537          | <0.001         | 0.473       | <0.001        |

BMI body mass index, ALT alanine aminotransferase, T. bilirubin total bilirubin, D. bilirubin direct bilirubin, ALP alkaline phosphatase, GGT gamma-glutamyl transferase, Hb hemoglobin, HAI histological activity index

monitor therapy protocols [11]. Although the associated complications are infrequent, finding new alternative non-invasive methods for the evaluation of liver disease is of great interest [12]. In the current work, the majority of the studied 200 children with chronic liver disease had mild, moderate, and severe disease activity that were 81%, 4%, and 3%, respectively, while minimal, moderate, and severe fibrosis were 41%, 19%, and 18%, respectively. This comes in consistent with Behairy et al., who found that most of the studied HCV patients had minimal disease activity (80%) and no/mild fibrosis (72%); on the other hand, they found that the majority of the AIH group had mild to moderate activity (70%) and moderate to severe fibrosis (95%) and all Wilson disease group had mild to moderate activity (100%) and moderate to severe fibrosis (100%) [13] and Pokorska-Spiewak et al., who reported that most of the cases showed minimal to mild fibrosis [5]. However, Dhole et al. confirmed that advanced fibrosis appeared in 23% of cases [14]. Fibrosis staging in chronic liver disease is crucial in determining the prognosis, selecting patients fit for anti-fibrotic treatment, and monitoring treatment outcomes [15].

The median of the non-invasive biomarkers was APRI 0.6, FIB-4 index 0.01, M-APRI 0.16, M-FIB-4 0.001, and B-AST 15.65. There were significant positive correlations between all of them and fibrosis score and HAI (P < 0.05 each), with a consistent increase in fibrosis score with the increase in mean ± SD of the studied five non-invasive biomarkers. This comes in line with Abd El-Ghaffar et al., who reported a highly significant positive correlation between APRI score and stage of fibrosis as assessed by the METAVIR scoring system (r = 0.53 and P-value = 0.000). APRI score mean in patients was 0.71 ± 0.48 and increased from 0.3 ± 0.45 in fibrosis stage <2 to 0.71 ± 0.96 in fibrosis stage ≥2 [16]. Also, de Ledinghen et al. studied APRI in comparison to Fibrosure and Fibroscan and found that APRI correlated with stages of fibrosis (rb = 0.32, P = 0.03) [17].

The aspartate aminotransferase (AST) to platelet ratio index (APRI) is a simple biomarker that was developed based on the progression of liver pathology and includes standard-of-care tests that may reflect hepatocellular damage and early development of portal hypertension [10]. Furthermore, Leung et al. found that APRI performed better than FIB-4 in predicting fibrosis studied in children with cystic fibrosis liver disease. APRI may hold great promise for earlier detection of fibrosis or clinically silent liver disease in order to decrease further complications [18]. Similarly, Pokorska-Spiewak et al. found the mean ± SD of the five biomarkers were APRI score 0.48 ± 0.26, FIB-4 0.22 ± 0.13, M-APRI 0.28 ± 0.69, M-FIB-4 0.09 ± 0.28, and B-AST 31.71 ± 69.87 and confirmed a significant positive association between the fibrosis

Table 5 Performance of the studied non-invasive markers in the detection of early (F1–F2) in the studied group

|                      | Cutoff value | Sens% | Spec% | PPV% | NPV% | AUC  | 95% CI  |
|----------------------|--------------|-------|-------|------|------|------|--------|
| APRI                 | > 0.96       | 46.4% | 91.9% | 94.7%| 32.2%| 0.745| 0.71–0.82 |
| M-APRI               | >0.16        | 61.3% | 96.5% | 97.9%| 40.4%| 0.831| 0.63–0.88 |
| FIB-4                | >0.019       | 54.8% | 94.5% | 97.7%| 36.9%| 0.750| 0.55–0.85 |
| M-FIB-4              | >0.005       | 49.7% | 96.5% | 97.4%| 34.4%| 0.759| 0.64–0.79 |
| B-AST                | >–8          | 74.2% | 75.7% | 90.5%| 43.2%| 0.758| 0.66–0.85 |

Sens sensitivity, Spec specificity, PPV positive predictive value, NPV negative predictive value, AUC area under the curve, CI confidence interval
stages and M-APRI and B-AST scores with a trend toward such an association with APRI and M-FIB-4 [5].

Elhenawy et al. stated that APRI and FIB-4 were significantly correlated with fibrosis in BA (P = 0.007) and were significantly higher in those with advanced fibrosis (Russo F4 and F5; P = 0.007), and they stated that these non-invasive serological markers, which are derived from simple routine laboratory tests, may be of help in predicting advanced fibrosis and in long-term follow-up of infants with BA and minimize the need for repeated follow-up liver biopsies [19]. Many studies had reported a positive correlation between APRI score and degree of liver fibrosis [14, 17, 20].

Values of fibrosis biomarker models increase with the progression of fibrosis stages because they are dependent on ALT, AST, and platelet count, with ALT and AST increase and platelet count decrease with increasing of fibrosis. Various factors induce decreased platelet count such as secondary to decreased thrombopoietin production by hepatocytes and/or sequestration and destruction of platelets in the spleen when liver fibrosis advances and portal hypertension develops with age. However, with ongoing liver injury, AST release from mitochondria is increased and hepatic fibrosis decreases its clearance [21].

While assessing the performance of studied non-invasive biomarkers, we found that the best cutoff for APRI in the detection of early fibrosis (F1–F2) was >0.96 with an AUC of 0.745, while in advanced liver fibrosis (F3–F4), it was >1.96, with AUC 0.849. This comes in accordance with Kim et al. who reported that APRI AUC for F ≥ 3 and F = 4 were 0.92 and 0.91, respectively. Distinct optimal cutoff values of APRI for F ≥ 3 and F = 4 were obtained (1.01 and 1.41, respectively) [20]. Also, de Ledinghen et al. found an AUC of 0.73 for predicting cirrhosis in children with various chronic liver

| Performance of the studied non-invasive biomarkers in the detection of advanced liver fibrosis (F3–F4) in the studied group |
|----------------------------------|-----------|-----------|-----------|-----------|---------|-----------|
| Cutoff value | Sens% | Spec% | PPV% | NPV% | AUC | 95% CI |
| APRI > 1.96 | 89.9% | 84.5% | 34.8% | 98.7% | 0.849 | 0.76–0.92 |
| M-APRI > 2.2 | 87.8% | 96.6% | 72.7% | 98.9% | 0.958 | 0.92–0.98 |
| FIB-4 > 0.045 | 78.8% | 85.6% | 33.3% | 97.5% | 0.853 | 0.77–0.91 |
| M-FIB-4 > 0.015 | 89.8% | 82.4% | 32% | 98.7% | 0.896 | 0.84–0.97 |
| B-AST > 92.1 | 89.8% | 91.2% | 47.1% | 98.8% | 0.955 | 0.875–0.97 |

Sens sensitivity, Spec specificity, PPV positive predictive value, NPV negative predictive value, AUC area under the curve, CI confidence interval

Fig. 1 Diagnostic performance (ROC curves) of the studied non-invasive biomarkers in the detection of early fibrosis (F1–F2)
In addition, Grieve et al. using a cutoff value of 1.22 [AUC 0.83] showed a sensitivity of 75% and a specificity of 84% for macroscopic cirrhosis [22], and McGoogan et al. found that the APRI was moderately useful in predicting significant fibrosis where it could be a substitute for liver biopsy with AUC of 0.71 [23].

Moreover, in a meta-analysis that included 40 studies, it was shown that APRI score > 1.0 had 76% sensitivity and 72% specificity for predicting cirrhosis. Additionally, APRI scores > 0.7 had a 77% sensitivity and 72% specificity for predicting significant hepatic fibrosis [24].

Regarding the performance of FIB-4, the cutoff point to detect early fibrosis (F1–F2) was > 0.019 with AUC 0.750, and to detect advanced fibrosis (F3–F4), it was > 0.045 with AUC 0.853. These results agree with Elhenawy et al. reports which showed the AUC of FIB-4 was 0.0098 with AUR 0.644, 61.9% sensitivity, and 61.9% specificity to discriminating advanced fibrosis [19]. More evidences came from Pokorska-Spiewak et al. study, where FIB-4 at cutoff point 0.18, with AUR 0.708, 85.7% sensitivity, and 93.7% NPV, helped in detecting any stage of fibrosis; the cutoff point was 0.09 with AUC 0.586 [5]. On the other hand, Chen et al. reported that FIB-4 failed to correlate with the fibrosis stage. This may be due to the small number of patients in Chen’s study (n = 24) [25].

In our study, during the assessment of the performance of BMI z-score modified models, we found M-APRI, M-FIB-4, and B-AST at cutoff points >0.16, >0.005, and >–8, respectively with corresponding AUC 0.831, 0.759, and 0.758, could detect early fibrosis (F1–F2), while in detecting advanced fibrosis (F3–F4), the cutoff points of M-APRI, M-FIB-4, and B-AST were >2.2, >0.015, and >92.1 with AUC 0.958, 0.896, and 0.955, respectively.

These results were in agreement with Pokorska-Spiewak et al., who reported the cutoff values of M-APRI, M-FIB-4, and B-AST in the detection of any stage of fibrosis were 0.08, 0.36, and 0.56 with corresponding AUC 0.597, 0.568, and 0.614, respectively, while those for detection of moderate to severe fibrosis were 0.577, 0.179, and 92.82 with corresponding AUC 0.842, 0.823, and 0.848, respectively [5]. Also, Pokorska-Spiewak et al. suggest that liver fibrosis in children with CHC is positively associated with the BMI z-score. In both univariate and multivariate analyses, the BMI z-score was found to be an independent predictor of fibrosis among 42 pediatric patients with CHC (P = 0.03) [12].

We had found that the BMI z-scores of modified non-invasive biomarker models (M-APRI, M-FIB-4, and B-AST) had better diagnostic values in detecting early fibrosis (F1–F2) with AUC exceeded 0.7. Therefore, we supported the idea of incorporating the BMI z-score in the well-established APRI and FIB-4 non-invasive biomarker formulas to improve their performance in predicting advanced fibrosis. In addition, we confirmed the
B-AST as a new non-invasive simple biomarker that can easily detect liver fibrosis using very simple parameters.

**Conclusion**

APRI, M-APRI, FIB-4, M-FIB-4, and B-AST are good non-invasive alternatives to liver biopsy in the detection of liver fibrosis and its extent in children with chronic liver diseases of different etiologies especially those that include BMI z-scores in their formulas.

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**Authors’ contributions**

1. OB contributed to the design and implementation of the research, aided in choosing the patients and helped shape the research, supervised the findings of this study, discussed the results, and read and approved the final manuscript.
2. OE contributed to the design and implementation of the research, aided in choosing the patients, performed the laboratory work and helped shape the research, supervised the findings of this study, discussed the results, and read and approved the final manuscript. 3. NS contributed to the design and implementation of the research, aided in choosing the patients and helped shape the research, supervised the findings of this study, discussed the results, read and approved the final manuscript.

"All authors have read and approved the manuscript."

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**Availability of data and materials**

All data generated or analyzed during this study are included in this published article.

**Declarations**

**Ethics approval and consent to participate**

The current study was approved by the Medical Research Ethical Committee of the Faculty of Medicine, Benha University. All subjects were informed about the procedures and the aim of the study, and informed written consent was obtained from the parents or caregivers of enrolled children. The committee’s reference number is not available.

**Consent for publication**

Not applicable

**Competing interests**

The authors declare that they have no competing interests.

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**References**

1. Hashmi SS, Bhatti AH, Malik MI, Rana A, Nair H, Dar SF. et al (2017). Spectrum of histopathological diagnosis in pediatric patients with liver disorders in Pakistan. J Pak Med Assoc; 67(2):266-267.
2. Gaballah MA, Elsain HH, Shaheen EN (2015) Enhanced liver fibrosis test can replace liver biopsy in detection of liver fibrosis in chronic hepatitis C patients. International Journal of Advanced Research 3(4):145–150
3. Ruzman L, Mikolasevic I, Baraba Dekanic K, Milic S, Palcevski G (2018) Advances in diagnosis of chronic liver diseases in pediatric patients. World J Pediatr. 14(6):541–547. https://doi.org/10.1007/s12519-018-0197-8
4. Leung DH (2017). Hepatic fibrosis scores and serum biomarkers in pediatric hepatology. Clinical Liver Disease; 9(5):125-130. DOI: https://doi.org/10.1016/cld.634.
5. Pokorska-Śpiewak M, Kowalik-Miko B, Anisiewska M, Magdalena P, Magdalena M (2017) Novel serum biomarkers modified by the body mass index z-score for the detection of liver fibrosis and steatosis in children with chronic hepatitis C. BMC Infectious Diseases 17(1):361–368. https://doi.org/10.1186/s12879-017-2462-1
6. Vergani GM, Vergani D (2002). Autoimmune liver disease. In Symposium on hepatology. Ind J Paediatr. 69(1):93–98. https://doi.org/10.1007/BF02723785
7. WHO Child Growth Standards based on length/height, weight and age (2006). Acta Paediatrica; 450:76-85.
8. Bedossa P, Poynard T. (1996). An algorithm for the grading of activity in chronic hepatitis C. The METAVIR Cooperative Study Group. HEPATOLOGY; 24: 289–293. [PubMed: 8690394]. 2. DOI: https://doi.org/10.1002/hep.510240201
9. Sterling RK, Lissen E, Clumeck N, Soia R, Correa MC, Montanier J, S. Sukowski M, Tomiani FJ, Dieterich DT, Thomas DL, Messinger D, Nelson M, APRICOT Clinical Investigators (2006) Development of a simple noninvasive index to predict significant fibrosis in patients with HIV/HCV coinfection. Hepatology 43(6):1317–1325. DOI:https://doi.org/10.1002/hep.21178
10. Wai CT, Greenson JK, Fontana RJ, Kalbflesch JD, Marrero JA, Conjeevaram HS, Lok AS (2003) A simple noninvasive index can predict both significant fibrosis and cirrhosis in patients with chronic hepatitis C. Hepatology 38(2): 518–526. https://doi.org/10.1001/jhep.2003.50346
11. Sparzec Z (2005) Complications after percutaneous liver biopsy in diffuse hepatopathies. Rom J gastroenterol; 14(4):379–384.
12. Pokorska-Śpiewak M, Kowalik-Mikolajewska B, Anisiewska M, Pluta M, Marczyńska M (2015) Is liver biopsy still needed in children with chronic viral hepatitis? World J Gastroenterol. 21(42):12141–12149. https://doi.org/10.3748/wjg.v21.i42.12141
13. Behairy B, Sira MM, Zalata KR, Salama E, Abdallah MA (2016) Transient elastography compared to liver biopsy and morphometry for predicting fibrosis in pediatric chronic liver disease; does etiology matter? World Journal of gastroenterology 22(16):4249–4258. https://doi.org/10.3748/wjg.v22.i16.4238
14. Chole SD, Kher AS, Ghildiyal RG, Tamber MP (2015) Chronic liver diseases in children: clinical profile and histology. Journal of Clinical and Diagnostic Research 9(7):4–7
15. Stauber RE, Lackner C (2007) Noninvasive diagnosis of hepatic fibrosis in chronic hepatitis C. World Journal of Gastroenterology 13(32):4287–4294. https://doi.org/10.3748/wjg.v13.i32.4287
16. Abdel Ghaflar T, Youssef A, Zalata K, Alshaikhawzy M, Mowafy M, Wanis A et al (2016) Non-invasive assessment of liver fibrosis in Egyptian children with chronic liver diseases. Curr Pediatr Res 20(1):57–63
17. de Ledijring V, Le Bail B, Rebouissoux L, Fournier C, Foucher J, Mette V et al (2007) Liver stiffness measurement in children using FibroScan: feasibility study and comparison with Fibrotest, aspartate transaminase to platelets ratio index, and liver biopsy. J Pediatr Gastroenterol Nutr 45(4):443–450. https://doi.org/10.1097/MPG.0b013e31812e656f
18. Leung DH, Khan M, Minard CG, Guffey D, Ramm GA, Clouston AD, Miller G, Lewindon PJ, Shepherd RW, Ramm GA (2015) Aspartate aminotransferase to platelet ratio and fibrosis-4 as biomarkers in biopsy-validated pediatric cystic fibrosis liver disease. Hepatology 62(5):1576–1583. https://doi.org/10.1002/hep.28016
19. Ehretawya IA, Oghener HS, Sultan MM, Sita MM (2017) Utility of noninvasive serum biomarkers of liver fibrosis in infants with biliary atresia. Adv Res Gastroenterol Hepatol 3(2):555607.
20. Kim SY, Seok JY, Han SJ, Koh H (2010) Assessment of liver fibrosis and cirrhosis by aspartate amino transferase-to-platelet ratio index in children with chronic hepatitis C. The METAVIR Cooperative Study Group. Hepatology, 24: 150–158. https://doi.org/10.1002/hep.510240201
21. Elhenawy IA, Ghanem HS, Sultan MM, Sira MM (2017) Utility of noninvasive serum biomarkers of liver fibrosis in infants with biliary atresia. Adv Res Gastroenterol Hepatol 3(2):555607.
22. Kim SY, Seok JY, Han SJ, Koh H (2010) Assessment of liver fibrosis and cirrhosis by aspartate amino transferase-to-platelet ratio index in children with chronic hepatitis C. Adv Res Gastroenterol Hepatol 3(2):555607.
23. Kim SY, Seok JY, Han SJ, Koh H (2010) Assessment of liver fibrosis and cirrhosis by aspartate amino transferase-to-platelet ratio index in children with biliary atresia. J Pediatr Surg 48(4):789–795. https://doi.org/10.1016/j.jpedsurg.2012.10.010
23. McGoogan KE, Smith PB, Choi SS, Berman W, Jhaveri R (2010) Performance of the AST-to-platelet ratio index as a noninvasive marker of fibrosis in pediatric patients with chronic viral hepatitis. Journal of pediatric gastroenterology and nutrition 50(3):344–346. https://doi.org/10.1097/MPG.0b013e3181aed725

24. Lin ZH, Xin YN, Dong QJ, Wang Q, Jiang XL, Zhan SH, Sun Y, Xuan SY (2011) Performance of the aspartate aminotransferase-to-platelet ratio index for the staging of hepatitis C-related fibrosis: an updated meta-analysis. Hepatology 53(3):726–736. https://doi.org/10.1002/hep.24105

25. Chen S, Liao B, Zhong Z, Zheng Y, Liu B, Shan Q, Xie X, Zhou L (2016) Supersonic shearwave elastography in the assessment of liver fibrosis for postoperative patients with biliary atresia. Sci Rep 6(1). https://doi.org/10.1038/srep31057

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