Comprehensive fatty acid fractionation profiling in preeclampsia: a case control study with multivariable analysis

Herlambang Herlambang1,2*, Anggelia Puspasari3, Citra Maharani3, Rina Nofri Enis4, Susan Tarawifa4, Amelia Dwi Fitri5, Huntari Harahap2, Asro Hayani Harahap4, Erny Kusdiyah6 and Mas Rizky Anggun Adipurna Syamsunarno7

Abstract

Background: Preeclampsia is a complication during pregnancy characterised by new-onset hypertension and proteinuria that develops after 20 weeks of gestation. Dyslipidemia in pregnancy is correlated with an increased risk of preeclampsia. However, the dynamic changes in lipid metabolic product, particularly fatty acid fraction, in preeclampsia maternal circulation, are not well understood. This study aimed to investigate fatty acid fraction in preeclampsia maternal blood compared with normotensive normal pregnancy.

Methods: A total of 34 women who developed preeclampsia and 32 women with normotensive normal pregnancy were included in our case-control study. Maternal blood samples were collected for serum fatty acid fractions analysis and other biochemical parameters. Serum fatty acid fractions included long-chain polyunsaturated fatty acid (LCPUFA), monounsaturated fatty acid (MUFA), saturated fatty acid, and total fatty acid, measured with gas chromatography-mass spectrometry (GC-MS). The mean difference of fatty acid level was analysed using parametric and non-parametric bivariate analysis based on normality distributed data, while the risk of preeclampsia based on fatty acid fraction was analysed using a logistic regression model.

Results: Women with preeclampsia have lower high-density lipoprotein (53.97 ± 12.82 mg/dL vs. 63.71 ± 15.20 mg/dL, p = 0.006), higher triglyceride (284.91 ± 97.68 mg/dL vs. 232.84 ± 73.69 mg/dL, p = 0.018) than that in the normotensive group. Higher palmitoleic acid was found in women with preeclampsia compared to normotensive normal pregnancy (422.94 ± 195.99 vs. 325.71 ± 111.03 μmol/L, p = 0.037). The binary logistic regression model showed that pregnant women who had total omega-3 levels within the reference values had a higher risk of suffering preeclampsia than those with the higher reference value (odds ratio OR (95% CI): 8.5 (1.51–48.07), p = 0.015). Pregnant women who have saturated fatty acid within reference values had a lower risk for suffering preeclampsia than those in upper reference value (OR (95% CI): 0.21 (0.52–0.88), p = 0.032).

Conclusion: Overall, palmitoleic acid was higher in women with preeclampsia. Further analysis indicated that reference omega-3 in and high saturated fatty acid serum levels are characteristics of women with preeclampsia.

Keywords: Fatty acid fraction, Omega-3, Omega-6, Palmitoleic acid, Preeclampsia, Saturated fatty acid

Introduction

Preeclampsia is a disorder that develops during pregnancy characterised by new-onset hypertension and proteinuria after 20 weeks of gestation and can rapidly
progress to severe complications, including maternal and fetal mortality [1, 2]. The mechanisms that initiate preeclampsia have many theories; nevertheless, the disruption of trophoblast invasion during placenta development leads to hypoxia, oxidative stress, inflammation, and further endothelial dysfunction is widely accepted [3, 4]. A recent study reported lipid metabolic alteration involved in preeclampsia pathogenesis. The catabolic state of fatty acid by higher lipolysis due to the rapid growth of fetal development normally occurs during the third semester of pregnancy. This catabolic state is higher in preeclampsia than in normal pregnancy, causing hyperlipidemia. This hyperlipidemia enhances oxidative stress and inflammation and leads to endothelial dysfunction [5–7]. An in vitro study reported an increased lipid droplet accumulation in endothelial cells, conditioned with maternal plasma and replicated after exposure to free fatty acid at the combined concentration, defining preeclampsia. This process enhanced cell apoptosis through a significant decrease of mitochondrial dehydrogenase [8].

Serum fatty acid consists of different fatty acid fractions, including long-chain polyunsaturated fatty acid (LCPUFA), monounsaturated fatty acid (MUFA) and saturated fatty acid (SFA), together with all of their fractions. A lipidomic analysis showed a different role of this fatty acid in the pathogenesis of the disease [9–12]. Omega-3 LCPUFA and omega-6 fraction are precursors of prostanoids and leukotrienes involved in inflammation, vasoconstriction, oxidative stress, and platelet aggregation [11, 13]. Myristic and palmitic acid, members of SFAs, are associated with the risk of hypertension in pregnancy [12]. SFAs may promote pro-coagulation [14] that is associated with placental ischemia [15], and MUFA has a protective role in cardiovascular diseases [16]. However, the information regarding comprehensive fatty acid profiles in pregnancy and preeclampsia is still limited. This study aimed to investigate fatty acid fraction in preeclampsia maternal blood compared with normotensive pregnancy.

Methods
Study design
This case-control study aimed to compare the fatty acid fraction in preeclamptic maternal blood to that of normal pregnancy. The protocol was arranged based on the Helsinki Declaration and approved by the Health Research Ethics Committee of the Faculty of Medicine, Public health and Nursing, Universitas Gadjah Mada, Dr. Sardjito Public Hospital, Yogyakarta, Indonesia (Ethical approval number: KE/FK/1189/EC/2020). All the subjects signed informed consent forms after receiving detailed information concerning the purpose of the study. Thirty-four women with preeclampsia and thirty-two women with normotensive pregnancies living in Jambi Province, Indonesia, were recruited. Preeclampsia was defined based on the American College of Obstetricians and Gynecologists (ACOG) 2013 criteria. Inclusion criteria for the preeclampsia group were a pregnant woman who had hypertension (systolic blood pressure ≥ 140 mmHg and/or diastolic blood pressure ≥ 100 mmHg) after 20 weeks of pregnancy with one or more of the following conditions: semiquantitative proteinuria > +1; acute kidney failure sign by creatinine > 1.1 g/dL; serum glutamate oxaloacetate transferase (SGOT); and/or serum glutamate pyruvate transferase (SGPT) > 40 IU/dL; thrombocyte < 100.000 cell/mm³. The inclusion criteria for the control group were antepartum normotensive healthy women with a gestational age-matched with the preeclamptic group. The exclusion criteria were twin pregnancy, the mother with signs of active clinical infection, pregnancy with assisted technology for fertilisation, history of kidney and hepatic failure. All the subjects were followed until childbirth. Dyslipidemia defined as a condition of elevated total cholesterol in blood plasma, low density lipoprotein (LDL), triglycerides, and decreased high density lipoprotein (HDL) according to the national cholesterol education program adult treatment panel (NCEP ATP) III criteria.

Blood pressure data were taken from the highest recorded blood pressure during the first came to the hospital until delivery, regardless of gestational age when the patients first came. The early-onset preeclampsia and late-onset preeclampsia was not differentiated in this study. Protein levels were taken from urine samples and measured with quantitative colourimetric methods. Creatinine serum was measured following an enzymatic-colourimetric procedure. The SGOT, SGPT serum level was measured based on the international federation of clinical chemistry (IFCC) with pyridoxal-5-phosphate activation methods. Thrombocyte count was measured with an automated hemato-analyser.

Fatty acid quantification
Fatty acid fractionation measurement used blood plasma, which was separated by centrifugation. As much as 5 mL of blood from the antecubital vein was taken from all subjects after 8–10 h of fasting. Gas chromatography-mass spectrometry (GC-MS) was used to measure fatty acid fractionation. The GC-MS was performed based on the methods described in a previous study [17]. The numeric data of the fatty acid fractionation group, based on the Mayo clinic cut off point, were set as an upper reference, reference, and below reference values.

This research quantified the comprehensive fatty acid fraction including long-chain polyunsaturated fatty acid and its fractionation including omega-3, alpha-linolenic
acid (ALA), eicosapentaenoic acid (EPA), docosahexaenoic acid (DHA), omega-6, linoleic acid (LA), gamma-linolenic acid (GLA), dihomo-gamma-linolenic acid (DGLA), and arachidonic acid (AA). We also measured the oleic acid and palmitoleic acid as the fraction of monounsaturated fatty acid. Myristic acid, palmitic acid, and stearic acid were also measured as fractionation of saturated fatty acids.

Data analysis
Numeric data was tested using the Kolmogorov-Smirnov distribution, log 10 used for data transformed. Data that were not normally distributed after being transformed were presented as median (min-max) and evaluated using the Mann-Whitney test. Normally distributed data were tested by independent t-test and presented as mean ± SD. Categorical data were analysed bivariate using chi-square or Fisher exact test based on the percentage of expected value. Multivariate analysis with a stepwise binary logistic regression model was used to predict preeclampsia based on the fatty acid fractionation profile. The data was analysed using SPSS version 26.

Results
Sixty-six pregnant women participated in this study. Baseline subject characteristics and the neonatal outcome is shown in Table 1.

Normotensive women are older, have higher BMI, and the percentage of primipara and nulliparous is higher than that of the preeclampsia group, but the difference is not statistically significant. The preeclamptic group have a higher systolic and diastolic blood pressure than normotensive women do and are statistically significant (Table 1).

The total cholesterol and LDL cholesterol were slightly different between the preeclampsia and normotensive group. Lower HDL and higher triglyceride are found in women with preeclampsia, and the difference is statistically significant (Table 1).

The control group have longer gestational time, a lower percentage of small babies for gestational age than those

### Table 1 Baseline subject characteristics

| Subject characteristic | Preeclampsia (n = 34) | Normotensive (n = 32) | P value |
|------------------------|------------------------|------------------------|---------|
| **Maternal characteristic** |                         |                         |         |
| Age, years | 29.73 ± 7.08 | 30.09 ± 7.06 | 0.838c |
| Blood Pressure | | | |
| SBP, mmHg | 170.0 (140.0–200.0) | 110.0 (100.0–130.0) | < 0.001d* |
| DBP, mmHg | 100.0 (80.0–160.0) | 70.0 (60.0–80.0) | < 0.001d* |
| Gravidarum, n | | | |
| Primigravida | 13 | 13 | 0.843a |
| Multigravida | 21 | 19 | |
| Parity, n | | | |
| Nulliparous | 11 | 13 | 0.485d |
| Multiparous | 23 | 19 | |
| BMI | 25.76 (21.93–35.93) | 25.86 (18.82–33.31) | 0.653d |
| Total cholesterol, mg/dL | 243.47 ± 44.32 | 250.91 ± 47.76 | 0.514 |
| LDL-cholesterol, mg/dL | 139.97 ± 33.48 | 143.09 ± 42.37 | 0.740 |
| HDL-cholesterol, mg/dL | 53.97 ± 12.82 | 63.71 ± 15.20 | 0.006* |
| Triglyceride, mg/dL | 284.91 ± 97.68 | 232.84 ± 73.69 | 0.018* |
| **Neonatal outcome** | | | |
| Gestational age, weeks | 37.5 (31.0–41.0) | 38.0 (28.0–41.0) | 0.155d |
| Baby birth weight, gram | 2600 (900–4500) | 3045 (1400–4690) | 0.051d |
| Baby birth for gestational age, n | | | |
| Small for gestation age | 23 | 27 | 0.113d |
| Proper for gestational age | 11 | 5 | |
| APGAR score, n | | | |
| Asphyxia, APGAR < 7 | 9 | 1 | 0.009** |
| Normal, APGAR ≥ 7 | 25 | 31 | |

* Chi-square test, †Fisher exact test, ‡t-test, §non-parametric test, *p < 0.005: statistically significant. Normally distributed data presented as mean ± standard deviation, transformed not normally distributed data presented as median (min-max). SBP Systolic blood pressure, DBP Diastolic blood pressure, BMI Body mass index, LDL Low density lipoprotein, HDL High density lipoprotein, APGAR Appearance, pulse, grimace, activity, respiration
of the preeclampsia group, but the difference is not statistically significant. The preeclamptic group have a significantly higher percentage of baby birth with asphyxia than the percentage of the control group (Table 1).

The bivariate analysis of fatty acid profiling is shown in Table 2. The group with preeclampsia has lower HDL and higher triglyceride than the normotensive group do, and both are statistically significant. The omega-6/omega-3 ratio and AA/DHA ratio was higher in the preeclamptic than that of the normotensive group, but this difference is not statistically significant.

The total LCPUFA, omega-3, and omega-6 are not significantly different between the preeclamptic and normotensive group. Omega-3 fractionation ALA, EPA, DHA and Omega-6 fractionation LA, GLA, DGLA, and AA are also not significantly different between both groups; however, the DHA is higher in the normotensive group while AA is higher in the preeclamptic group (Table 2).

Saturated fatty acid and monounsaturated fatty acid are higher in the group with preeclampsia, but this difference with that of the normotensive group is not statistically significant. Palmitoleic acid as monosaturated fatty acid fractionation is significantly higher in the preeclampsia group compared to that of normotensive women (Table 2).

A stepwise logistic regression model was created, from which the model with the most significant p-value was taken. The results show that subjects with total omega-3 blood levels within the reference value have an 8.5x higher risk of suffering preeclampsia than subjects within the upper reference value. Women who have saturated fatty acid within the reference value have as much as 21% lower risk of suffering preeclampsia than the risk of subjects within the upper reference value (Table 3).

Furthermore, we also used a stepwise regression model to predict small-for-gestational-age baby weight in the group with preeclampsia. The model with the most significant p-value was taken (Table 4).

The women who have total omega-3 blood levels within the reference value have a higher risk of having a small-for-gestational-age baby weight than those with total omega-3 within the upper reference value. A similar trend is also observed for the omega-6/3 ratio (Table 4).

**Table 2** Bivariate analysis of the fatty acid profiling

| Fatty acid fractionation | Preeclampsia (n = 34) | Normotensive (n = 32) | P-value |
|-------------------------|-----------------------|-----------------------|---------|
| LCPUFA                  | 5750.67 ± 1476.53     | 5706.87 ± 977.46      | 0.887   |
| w6/w3 ratio             | 11.36 ± 3.40          | 10.94 ± 3.12          | 0.633   |
| AA/EPA ratio            | 36.50 (9.00–134.00)   | 44.00 (10.00–116.0)   | 0.130*  |
| AA/DHA ratio            | 1.66 ± 0.387          | 1.49 ± 0.369          | 0.077   |
| Total Omega-3           | 494.64 ± 171.00       | 510.53 ± 157.47       | 0.618   |
| w3 index                | 3.00 (2.00–6.00)      | 4.00 (2.00–5.00)      | 0.159*  |
| ALA (C18:3 w3)          | 40.79 ± 17.70         | 40.15 ± 15.61         | 0.877   |
| EPA (C20:5 w3)          | 24.67 ± 23.63         | 18.34 ± 15.88         | 0.111   |
| DHA (C22:6 w3)          | 429.26 ± 145.59       | 451.87 ± 137.81       | 0.466   |
| Total Omega-6           | 5240.88 ± 1367.31     | 5196.37 ± 889.10      | 0.877   |
| LA (C18:2 w6)           | 4244.0 ± 1198.49      | 4278.84 ± 754.12      | 0.877   |
| GLA (C18.3 w6)          | 12.23 ± 7.05          | 10.50 ± 4.80          | 0.310   |
| DGLA (C20.3 w6)         | 281.97 ± 90.00        | 249.93 ± 69.32        | 0.112   |
| AA (C20.4 w6)           | 700.02 ± 250.94       | 657.03 ± 194.62       | 0.442   |
| Omega-9 (Oleic acid, C18:1 w9) | 3075.58 ± 950.43 | 2784.65 ± 795.29 | 0.184 |
| Saturated fatty acid    | 5746.79 ± 1578.68     | 5442.43 ± 1140.38     | 0.377   |
| Myristic acid, C14:0     | 151.08 ± 115.13       | 166.46 ± 76.30        | 0.188   |
| Palmitic acid, C16:0     | 4834.32 ± 1341.59     | 4504.96 ± 1003.46     | 0.261   |
| Stearic acid, C18:0      | 756.17 ± 185.33       | 774.21 ± 126.48       | 0.648   |
| Monounsaturated fatty acid | 3511.52 ± 1075.18   | 3102.25 ± 881.53      | 0.095   |
| Palmitoleic acid (C16:1 w7) | 422.94 ± 195.99    | 325.71 ± 111.03       | 0.037   |
| Oleic acid (C18:1 w9)    | 3075.58 ± 950.43     | 2784.65 ± 795.29      | 0.184   |
| Total fatty acid         | 14,974.67 ± 4052.2    | 14,275.59 ± 2883.22   | 0.420   |

* nonparametric test; otherwise t-test, *p < 0.005: statistically significant. Fatty acid fractionation in μmol/L. AA Arachidonic acid, ALA Alpha-linolenic acid, DGLA Dihomo-gamma-linolenic acid, DHA Docosahexaenoic acid, EPA Eicosapentaenoic acid, GLA Gamma-linolenic acid, LA Linoleic acid, LCPUFA long-chain polyunsaturated fatty acid; w3: omega-3; w6: omega-6
Discussion

The baseline subject characteristics of this study were not statistically different based on age, gravid status, and parity status between the preeclamptic and normotensive groups. Higher blood pressure was standard in the group with preeclampsia.

Some differences in BMI, total cholesterol, and LDL cholesterol were found between both groups. Higher plasma triglycerides and lower plasma HDL were found in women with preeclampsia. Pregnancy altered the maternal lipid metabolism; catabolic state signed by increased lipolysis occurred during the third pregnancy. Higher plasma FFA, glycerol, and triglycerides were found as lipolysis by-products and compensation for the higher energy demand due to rapid fetal development. In addition to the increased triglycerides, the triglyceride-rich HDL2 also increased [6, 18]. Lipid metabolic maladaptation in preeclampsia correlated with higher serum triglycerides, increased circulating FFA, and lower HDL than normal pregnancy, which mimics insulin resistant syndrome [19, 20].

A recent study reported the role of FFA in inflammation, oxidative stress, and cell apoptosis pathway, which is involved in the pathogenesis of preeclampsia [10]. This study quantified various fatty acid fractions such as LCP-UFA, MUFA, saturated fatty acid, and total fatty acid. The fatty acid fraction may have different impacts on the pathogenesis of preeclampsia.

Previous studies have proposed that lower omega-6 serum and higher omega-3 serum levels can decrease the risk of suffering preeclampsia [8, 11, 21]. In accordance with

Table 3Logistic regression model for fatty acid profiling used to predict preeclampsia

| Fatty acid profiling | B     | SE   | Adjusted p-value | OR (95% CI) |
|---------------------|-------|------|------------------|-------------|
| **Total Omega-3**   |       |      |                  |             |
| Upper reference value (> 500 umol/L) | ref   |      |                  |             |
| Reference value (200–500 umol/L)      | 2.141 | 0.884| 0.015*           | 8.50 (1.51–48.07) |
| Below reference value                | NS    |      |                  |             |
| **Saturated fatty acid**             |       |      |                  |             |
| Upper reference value (> 5500 umol/L) | ref   |      |                  |             |
| Reference value (2500–5500 umol/L)    | −1.540| 0.719| 0.032*           | 0.21 (0.52–0.88) |
| **Stearic acid**                    | NS    |      |                  |             |
| **EPA**                             | NS    |      |                  |             |

The model is a stepwise logistic regression model with the most significant p-value. The Hosmer and Lemeshow test was performed to analyse the goodness of fit. The p value for all models was > 0.05. B is the logistic regression model coefficient; SE the standard error; OR the odds ratio with 95% confidence interval (CI); ref. was reference genotype; p < 0.005: statistically significant.

Table 4Logistic regression model for fatty acid profiling to predict small-for-gestational-age baby weight in the preeclampsia group

| Fatty acid profiling | B     | SE   | Adjusted p-value | OR (95% CI) |
|---------------------|-------|------|------------------|-------------|
| **Total Omega-3**   |       |      |                  |             |
| Upper reference value (> 500 umol/L) | ref   |      |                  |             |
| Reference value (200–500 umol/L)      | 2.692 | 1.305| 0.039*           | 14.76 (1.14–190.37) |
| **Omega-6/3 ratio** |       |      |                  |             |
| Upper reference value (> 10.7)        | ref   |      |                  |             |
| Reference value (3.4–10.7)            | 2.718 | 1.192| 0.023*           | 15.14 (1.46–156.51) |
| **AA**                              | NS    |      |                  |             |

The model was a stepwise logistic regression model with the most significant p-value. The Hosmer and Lemeshow test was performed to analyse the goodness of fit. The p value for all models was > 0.05. B is the logistic regression model coefficient; SE the standard error; OR the odds ratio with 95% confidence interval (CI); ref. was reference genotype; p < 0.005: statistically significant.
previous studies, this research also reported that subjects who had plasma omega-3 within the reference value had a higher risk of suffering preeclampsia and small-for-gestational age baby weight than those who had plasma omega-3 within the upper reference value. Plasma level of omega-6/3 ratio in the reference value has a higher risk of having a small-for-gestational age baby in the preeclamptic group than those within our study’s upper reference value. Because this result is contrary to previous studies, the exact mechanism of this disorder is not yet clear. Hence further dietary assessments are needed.

Our study reported that the plasma level of saturated fatty acid within reference value was a protective factor for suffering preeclampsia; the mechanism of saturated fatty acid within the upper reference values in preeclampsia remains unclear. An in vitro study using culture cells reported a decreased mitochondrial dehydrogenase activity and increased apoptosis of human umbilical vein after saturated fatty acid exposure [22]. An epidemiology study also found a positive correlation between saturated fatty acids and pregnancy-induced hypertension in preeclampsia [23]. Furthermore, saturated fatty acids may promote pro-coagulation activity [14], associated with placental ischemia [15].

Our study’s main and unique finding was the higher level of palmitoleic acid in preeclamptic group than that of the normotensive group. To the best of our knowledge, no study has investigated the correlation and mechanism of palmitoleic acid in preeclampsia. Palmitoleic acid is a monounsaturated and omega-7 fatty acid derived from desaturation of palmitic or vaccenic acid, used as a de-novo lipogenesis marker. Palmitoleic acid is proposed to be the new lipokine, which influences interorgan metabolic correlation [24]. Previous studies reported higher plasma palmitoleic acid correlated with multiple cardiometabolic risks, including high blood pressure, dyslipidemia, and endothelial dysfunction. Contrary to those studies, lipokine has been reported to decrease insulin resistance and inflammation status [25]. Further research is thus needed to investigate how higher palmitoleic acid in preeclampsia mimicks adipokine resistance in many cardiovascular diseases considering that evidence shows that preeclampsia mimicks insulin resistance syndrome.

In addition, a decreased mitochondria capacity for long-chain beta-oxidation through decreased expression of long-chain 3-hydroxy acyl-CoA dehydrogenase (LCHAD) was found in a preeclampsia animal model. This metabolic shift enhanced oxidative stress [12]. Although not statistically significant, higher total free fatty acid was found in the preeclamptic group compared to that of the normotensive group.

The sample size was not too large, but it was in accordance with the sample size calculation rule. The main finding of this study enriched the knowledge on the role of fatty acids in preeclampsia. In addition, it promises clinical settings as risk screening and arrangement of nutrition patterns for preeclampsia prevention and management. However, further research with a larger sample size, multi-centre, longitudinal design was needed to reveal a more comprehensive finding.

Conclusion
Palmitoleic acid was higher in women with preeclampsia compared to that of normotensive pregnant women. Further analysis indicated omega-3 serum level as a protective factor in women with preeclampsia, while saturated fatty acid serum level as a risk factor.

Abbreviations
AA: Arachidonic acid; ALA: Alpha-linolenic acid; DGLA: Dihomo-gamma-linolenic acid, DHA: Docosahexaenoic acid; EPA: Eicosapentaenoic acid; GLA: Gamma-linolenic acid, HDL: High density lipoprotein; LA: Linoleic acid; LCPUFA: Long-chain polyunsaturated fatty acid; LDL: Low density lipoprotein; NCEP ATP: National cholesterol education program adult treatment panel; w3: Omega-3, w6: Omega-6.

Acknowledgements
This study was supported by Universitas Jambi internal research grant number 186/UN21.18/PG/SPK/2020 for Herlambang, MD, Ph.D. We thank PRODIA laboratories for the technical support provided for the metabolomic assay.

Authors’ contributions
HH, RNE and STW have made substantial contributions to the design of the work; HH, AHH, EK, and MRAAS have made the acquisition and analysis of laboratory measurement; ADF, CM, and AP interpreted the data; HH, MRAAS, CM, AP, and ADF drafted the work or substantively revised it. All the authors approved the submitted version and any substantially modified version that involves the author’s contribution to the study and have agreed to be personally accountable for the author’s contributions and to ensure that questions related to the accuracy or integrity of any part of the work, even ones in which the author was not personally involved, are appropriately investigated, resolved, and the resolution documented in the literature.

Funding
This study was supported by Universitas Jambi internal research grant number 186/UN21.18/PG/SPK/2020 for Herlambang, MD, Ph.D. The funder’s role was to fund this research, especially for sample collecting, metabolomic analysis, and publication.

Availability of data and materials
The data sets analysed during the current study are not publicly available due to confidential agreement subjects but are available from the corresponding author on reasonable request.

Declarations
Ethics approval and consent to participate
The study protocol was arranged based on the Helsinki Declaration and had approval from the Medical and Health Research Ethics Committee of the Faculty of Medicine, Public health and Nursing, Universitas Gadjah Mada, Dr. Sardjito Public Hospital, Yogyakarta, Indonesia. The ethical approval number was KE/PK/1189/EC/2020.

Consent for publication
Due to personal subject data, all subjects who participated signed informed consent forms after receiving detailed information concerning the purpose of the study.
Competing interests
We declared no conflict of interest associated with this publication, and there has been no significant financial support for this work that could have influenced its outcome.

Author details
1. Department of Obstetrics and Gynecology, Faculty of Medicine and Health Sciences, Universitas Jambi, Jambi, Indonesia.
2. Department of Physiology, Faculty of Medicine and Health Sciences, Universitas Jambi, Jambi, Indonesia.
3. Department of Medical Biology and Biochemistry, Faculty of Medicine and Health Sciences, Universitas Jambi, Jambi, Indonesia.
4. Department of Anatomy, Faculty of Medicine and Health Sciences, Universitas Jambi, Jambi, Indonesia.
5. Department of Medical Education, Faculty of Medicine and Health Sciences, Universitas Jambi, Jambi, Indonesia.
6. Department of Public Health, Faculty of Medicine and Health Sciences, Universitas Jambi, Jambi, Indonesia.
7. Department of Biomedical Sciences Faculty of Medicine Universitas Padjadjaran, Bandung, Indonesia.

Received: 20 August 2021 Accepted: 29 November 2021
Published online: 03 January 2022

References
1. Mayrink J, Costa M, Cecatti J. Preeclampsia in 2018: revisiting concepts, physiopathology, and prediction. Sci World J. 2018;2018:6268276. https://doi.org/10.1155/2018/6268276.
2. Rana S, Lemoine E, Granger JP, Karumanchi SA. Preeclampsia pathophysiology, challenges, and perspectives. Circ Res. 2019;124(7):1094–112. https://doi.org/10.1161/CIRCRESAHA.118.313276.
3. Karthikeyan VJ, Lip GY. Endothelial damage/dysfunction and hypertension in pregnancy. Front Biosci (Elite Ed). 2011;6:313–8. https://doi.org/10.2741/314.
4. Yienoglou DB, Ugrar MG, Ozcan HC, Can G, Ozkurt E, Balat O, et al. Assessment of oxidative stress markers in recurrent pregnancy loss: a prospective study. Arch Gynecol Obstet. 2013;289:1337–40. https://doi.org/10.1007/s00404-013-3113-4.
5. Ghio A, Bertolotto A, Resi V, Valpe L, Cianni GD. Triglyceride metabolism in pregnancy. Adv Clin Chem. 2011;55:133–53. https://doi.org/10.1016/b978-0-12-387042-1.00007-1.
6. Zeng Z, Liu F, Li S. Metabolic adaptations in pregnancy: a review. Ann Nutr Metab. 2017;70:539–65. https://doi.org/10.1159/000493633.
7. Enquobahrie DA, Williams MA, Butler CL, Frederick IO, Miller RS, Luthy RA. Maternal plasma lipid concentrations in early pregnancy and risk of preeclampsia. Am J Hypertens. 2004;17(7):574–81. https://doi.org/10.1016/S0148-5833(04)70066-9.
8. Wadhwani NS, Naranj AS, Mehendale SS, Waghi GN, Gupte SA, Joshi SR. Reduced maternal erythrocyte long chain polyunsaturated fatty acids exist in early pregnancy in preeclampsia. Lipids. 2016,51(1):85–94. https://doi.org/10.1007/s11775-015-2498-5.
9. Tesfa E, Nileb E, Munshea A. Maternal lipid profile and risk of preeclampsia in African pregnant women: a systematic review and meta-analysis. Plos One. 2020;15(12):e0243538. https://doi.org/10.1371/journal.pone.0243538.
10. Kaaja R. Lipid abnormalities in preeclampsia: implications for vascular health. Clin Lipidol. 2011;6(1):71–8. https://doi.org/10.2217/cip.10.82.
11. Wang Y, Walsh SW, Kay HH. Placental tissue levels of nonesterified polyunsaturated fatty acids in normal and preeclamptic pregnancies. Hypertens Pregnancy. 2005,24(3):235–45. https://doi.org/10.1080/10641950500281118.
12. Yi X, Huang Y, Zhang W, Yang C, Su W, Wu Y, et al. Association of circulating saturated fatty acids with the risk of pregnancy-induced hypertension—a nested case–control study. Hypertens Res. 2020;43(1):421–2. https://doi.org/10.1007/s41440-019-0383-7.
13. Mackay VA, Huda SS, Stewart FM, Tham K, McKenna LA, Martin I, et al. Preeclampsia is associated with compromised maternal synthesis of laong chain polyunsaturated fatty acids leading to offspring deficiency. Hypertension. 2012,60(4):1078–85. https://doi.org/10.1161/HYPERTENSIONHA.112.1979897.
14. Miller GJ. Dietary fatty acids and blood coagulation. Prostaglandins Leukot Essent Fatty Acids. 1997;57(4–5):389–94. https://doi.org/10.1016/S0952-3278(97)90416-6.
15. Dusse LM, Rios DRA, Pinheiro MB, Cooper AJ, Waleed BA. Preeclampsia: relationship between coagulation, fibrinolysis and inflammation. Clin Chim Acta. 2011;412(1–2):17–21. https://doi.org/10.1016/j.cca.2010.09.030.
16. Schwinghamml L, Hoffmann G. Monounsaturated fatty acids and risk of cardiovascular disease: synopsis of the evidence available from systematic reviews and meta-analyses. Nutrients. 2012,4(12):1989–2007. https://doi.org/10.3390/nu121239483.
17. Putri M, Hamjiyo L, Alita VR, Atik N, Syamsunarno MRA. Very long fatty acid fractionation in systemic lupus erythematosus in the active and inactive status. Int J Appl Pharm. 2020;12(3):83–6. https://doi.org/10.22159/ijap.2020.v12s3.39483.
18. Herrera E. Metabolic adaptations in pregnancy and their implications for the availability of substrates to the fetus. Eur J Clin Nutr. 2000,54(Suppl 1):S47–51. https://doi.org/10.1038/sj.ejcn.1600894.
19. Ray JG, Diamond P, Singh G, Bell CM. Brief overview of maternal triglycerides as a risk factor for preeclampsia. Br J Obstet Gynaecol. 2006;113:379–86. https://doi.org/10.1111/j.1471-0528.2006.00889.x.
20. Kaaja R, Laiuvori H, Laakso M, Tikkanen MJ, Ylikorkala O. Evidence of a state of increased insulin resistance in preeclampsia. Metabolism. 1999,48:892–6. https://doi.org/10.1016/S0026-0495(99)00922-5.
21. Jump DB. The biochemistry of n-3 polyunsaturated fatty acids. J Biol Chem. 2002,277(11):8755–8. https://doi.org/10.1074/jbc.R100062200.
22. Ding X, Yang Z, Han Y, Yu H. Fatty acid oxidation changes and the correlation with oxidative stress in different preeclampsia-like mouse models. Plos One. 2014,9(10):e109554. https://doi.org/10.1371/journal.pone.0109554.
23. Robinson NJ, Minchell LJ, Myes JE, Hubel CA, Crocker IP. A potential role for free fatty acids in the pathogenesis of preeclampsia. J Hypertens. 2009,27(6):1293–302. https://doi.org/10.1097/HJH.0b013e3282329bfe.
24. Li VL, Kim JT, Long JZ. Adipose tissue lipokine: recent progress and future direction. Diabetes. 2020,69(12):2541–8. https://doi.org/10.2337/db20-0012.
25. Frigolet ME, Gutiérrez-Aguilar R. The role of the novel lipokine palmitoleic acid in health and disease. Adv Nutr. 2017,8(1):735–815. https://doi.org/10.3945/an.115.011130.

Publisher’s Note
Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.