Effects of the inclusion of a mixed *Psychrotrophic* bacteria strain for sewage treatment in constructed wetland in winter seasons

Meizhen Tang, Zhengtao Li, Yuewei Yang, Junfeng Chen and Jie Jiang

College of Life Science, Qufu Normal University, Shandong, Qufu 273165, People’s Republic of China

Received: 7 January 2018
Accepted: 21 March 2018

Subject Category:
Engineering

Subject Areas:
biotechnology

Keywords:
mixed *Psychrotrophic* bacteria strain, constructed wetlands, wastewater treatment, kinetics mode

Authors for correspondence:
Meizhen Tang
e-mail: qsd_tmzh@mail.qfnu.edu.cn
Yuewei Yang
e-mail: yangyuewei@163.com

Constructed wetlands (CWs) have been used globally in wastewater treatment for years. CWs represent an efficient ecological system which is both energy-saving and low in investment for construction and operational cost. In addition, CWs also have the advantage of being easy to operate and maintain. However, the operation of CWs at northern latitudes (both mid and high) is sometimes quite demanding, due to the inhibitory effect of low temperatures that often occur in winter. To evaluate the wastewater treatment performance of a culture of mixed *Psychrotrophic* bacteria strains in an integrated vertical-flow CW, the removal rates of ammonia nitrogen (NH$_3$–N), chemical oxygen demand (COD), nitrite nitrogen (NO$_2^−$–N), nitrate nitrogen (NO$_3^−$–N) and total phosphorus (TP) were quantified at different bacterial dosages to determine the best bacterial dosage and establish kinetic degradation models of the mixed strains. The bacterial culture was made up of *Psychrobacter* TM-1, *Sphingobacterium* TM-2 and *Pseudomonas* TM-3, mixed together at a volume/volume ratio of 1 : 1 : 1 (at bacterial suspension concentrations of 4.4 × 10$^9$ ml$^{-1}$). Results showed that the organic pollutants (nitrogen and phosphorus) in the sewage could be efficiently removed by the culture of mixed *Psychrotrophic* bacteria. The optimal dosage of this mixed bacteria strain was 2.5%, and the treatment efficiency of COD, NH$_3$–N, NO$_2^−$–N, NO$_3^−$–N, total nitrogen and TP were stable at 91.8%, 91.1%, 88.0%, 93.8%, 94.8% and 95.2%, respectively, which were 1.5, 2.0, 2.1, 1.5, 2.2 and 1.3 times those of...
1. Introduction

A constructed wetland (CW) is an artificial wetland built to imitate a natural wetland ecosystem. Because of the low initial investment in construction, low operational costs, limited required maintenance, high efficiency and good compatibility of stable treatment effect and ecological landscape, CWs are widely used in processing agricultural sewage, acid mine drainage, industrial wastewater, aquaculture wastewater, landfill leachate and city highway runoff and drainage [1–8]. In recently published studies, it was found that the contaminant removal rate by CWs could reach 85% to 95% for biochemical oxygen demand (BOD), 80% for chemical oxygen demand (COD) and 92.41% for nitrogen compounds [9]. The main mechanisms of pollutant removal in constructed wetlands involve several biological processes (e.g. microbial metabolic activity and plant uptake) and physico-chemical processes (e.g. sedimentation, adsorption and precipitation at water-sediment, root-sediment and plant–water interfaces) [10]. Microbial degradation plays a dominant role in the removal of soluble/colloidal biodegradable organics in wastewater [11,12]. However, the sewage treatment efficiency of the constructed wetlands in northern China is rather low in winter. This is because microbial activity is reduced as the water temperature decreases, and (due to winter plant death) the biological activity of CWs might decrease under winter conditions. Thereby the pollutant removal efficiency and, more importantly, the efficacy of this contaminant removal technique might be affected by seasonal changes.

There is some global research on low-temperature biological strengthening techniques that can be applied to artificial wetland systems. Pei et al. [13] studied the impact of denitrification efficiency by Bacillus subtilis FY99-01 in wetland systems, and found that the microorganisms could effectively increase the removal of nitrates from the wetlands. Shao et al. studied the denitrification effects of Paenibacillus sp. XP1 dosing on cattails and reed surface flow in a CW system when temperature was between 15 and 21°C and found that Paenibacillus sp. XP1 introduction could effectively shorten the treatment time required for wastewater decontamination. Furthermore, after treatment with these bacterial cultures the concentration of ammonia (NH3–N) in effluent met the national level B standard in the CW, and compared with the reed CW, was more efficient at denitrification [14]. These results confirmed the feasibility of artificial wetland wastewater treatments at low temperatures [15]. The sewage treatment efficiency of Pseudomonas flava WD-3 in an integrated vertical-flow constructed wetland (IVCW) during winter with different dosages, P. flava WD-3 has a good processability for the wastewater treatment in the IVCW system in winter. Besides, the simplified Monod model simulated and evaluated the pollutant removal efficiency of this bacterial strain with respect to its dosages to improve water quality, and even accurately predicted the pollutant removal efficiency [16].

There are relatively few recent studies that have focused on the use of biological strengthening technology to increase the efficiency of the wastewater treatment in CWs. Owing to the effects of the natural environment, the applications of low-temperature biological reinforcement technology in CW systems still needs further research.

In this study the cold-resistant mixed bacteria cultures of Psychrobacter TM-1, Sphingobacterium TM-2 and Pseudomonas TM-3 (mixed Psychrotrophic bacteria strain), isolated from the Nansihu Lake wetlands, were used in the IVCW system. The removal rates of chemical oxygen demand (COD), ammonia nitrogen (NH3–N), nitrate nitrogen (NO3–N), nitrite nitrogen (NO2–N), total nitrogen (TN) and total phosphorus (TP) were measured to test the biotreatment performance of this mixed Psychrotrophic bacteria strain. The main objective of this paper was to develop a simplified first-order kinetics model that can be used with CW data to (1) predict COD, NH3–N, NO3–N, NO2–N, TN and TP retention in the wetland system, (2) evaluate the effects of different dosages and (3) provide a modelling tool to simulate and evaluate the pollutant removal efficiency of this mixed Psychrotrophic bacteria strain. The results of this study provide a theoretical foundation and scientific support for the use of constructed wetlands in wastewater treatment during the winter months, and may have great significance in solving the increasingly severe water pollution problem in China.
2. Material and methods

2.1. Isolation, screening and identification of the mixed Psychrotrophic bacteria strain

2.1.1. Enrichment culture, separation, screening and purification of the Psychrotrophic bacteria

Sediment samples, collected from Nansihu Lake in Shandong province, were incubated at a temperature of 2°C for 7 days in order to domesticate the microbes. Five grams of sediment was placed in each conical flask and shaken in an orbital shaker incubator (150 r.p.m.) at a temperature of (6 ± 1)°C. To allow the microbes to proliferate rapidly, we placed a few vitreous balls and 95 ml of sterile culture medium in each flask.

One millilitre of the nutrient solution was added to each test tube with 9 ml of sterilized water (1:10 dilution of the culture medium). In this way, dilution to 10\(^{-2}\), 10\(^{-3}\), 10\(^{-4}\), 10\(^{-5}\) and 10\(^{-6}\) times less than the original concentration of the culture medium could be easily obtained. Subsequently, 1 ml of the solution was taken from each flask and diluted 10\(^{-4}\), 10\(^{-5}\), 10\(^{-6}\) times less than the original concentration. These solutions were then inoculated in three kinds of isolation media (with three parallels for each kind of medium), using agar plates. After that, the plates were placed into the incubator at a temperature of 6(±1)°C and growth was recorded constantly. The nine best-growing strains were selected and repeatedly purified to acquire a single colony. Then the colonies were inoculated in a slant culture medium and reserved in the refrigerator at 4°C.

2.1.2. The screening of the mixed Psychrotrophic bacteria

After the enrichment cultivation, the nine single colonies were inoculated in the simulated wastewater while at a pH of 7.0. Inoculation quantity was controlled to within 10%. The solutions were statically cultured following 6 h of aeration, and the concentrations of COD, TP and NH\(_3\)-N were measured every 12 h. According to the removal efficiency of the pollutants, the most efficient strains (1, 2, 3, 4 and 5) were selected.

These five bacterial strains, which had high-efficient degrading capability, were inoculated in lysogeny broth (LB) culture medium at different concentrations. The growth status of the mixed flora was recorded regularly. Meanwhile, the mixed flora was inoculated in the simulated wastewater at a pH of 7.0. The inoculation quantity was controlled within 10%. The solutions were then statically cultured after 6 h of aeration at 6(±1)°C, and the concentrations of COD, TP and NH\(_3\)-N were measured every 12 h. According to the removal efficiency of the pollutants, the combination of the 1, 4 and 5 strains had the most notable effect on sewage disposal efficiency.

2.1.3. Identification of the mixed Psychrotrophic bacteria species

To identify the species of bacteria in the mixed Psychrotrophic bacteria, the 1, 4 and 5 strains were dyed, placed under the microscope and the morphological characteristics of these bacteria were recorded in table 1. A preliminary identification was carried out using physiological and biochemical characteristics of the bacterial strains.

Using a scanning electron microscope the 1, 4 and 5 strains were purified and inoculated on an LB solid culture medium. After incubation at 30°C for 24 h, the 1, 4 and 5 strains were selected, isolated and then rinsed several times with sterilized water. Firstly, the thalluses were fixed with 3% glutaraldehyde, then rinsed, and fixed with osmic acid. Finally, the bacteria were dehydrated with ethanol.

16S rDNA sequence analysis: the total DNA of strains 1, 4 and 5 was extracted. Using the following primers: 8f (5' AGAGTTTGATCCTGGCTCAG 3') 20 bp and 1492r (5' GTTACCTTGTTACGACTT 3'), PCR was performed. Following cloning and purification, the PCR product was identified by Shanghai Biological Engineering Co, Ltd [17–19].

2.2. Integrated vertical-flow constructed wetlands system and operation

Six small-scale plots were constructed in March 2012. Each plot (2 m\(^2\)) was equally divided into two chambers: a down-flow chamber and an up-flow chamber, indicating the direction of the passing water. A collecting ditch, 100 mm deep, was installed at the bottom of the chambers. Each chamber comprised three different particle-size-distribution layers: 150 mm depth of gravel (40–50 mm diameter) on the bottom, 250 mm slag (5–10 mm diameter) in the middle and 300 mm (in the down-flow chamber) or 250 mm (in the up-flow chamber) brown soil (0–4 mm diameter) on the top. All the water pipes were
made of stainless steel (75 mm diameter). For an even distribution of water, the influent pipes had holes (5 mm diameter) on the underside and were spread on the surface of the down-flow chamber. At the bottoms of the two chambers, the collecting pipes transported water coming from the down-flow chamber to the up-flow chamber. The effluent pipes were situated on the surface of the up-flow chamber (figure 1). *Typha orientalis* Presl and *Acorus calamus* were planted in the down-flow and up-flow chambers. The dosing load of the CW was 2–20 cm d⁻¹ and the organic load was 15–20 kg ha⁻¹ h⁻¹. When the system worked, the sewage was first dosed evenly on the surface of the down-flow chamber, then the sewage flowed downward vertically, finally reaching the up-flow chamber through a connecting pipe. After reaching the up-flow chamber, it flowed upward vertically, and finally was discharged from the effluent pipes. In this experiment, the CW was designed to be a discontinuous-flow system and was operated in a non-saturated state. The detention time in the system was set to be 5 days.

The influent water was taken from the sewage treatment plant in Qufu. Parameters are listed in table 2.

Preparation of bacterial suspension: the 1, 4, and 5 strains were incubated in the LB liquid culture medium until their absorbance reached 1.2. The solution was then centrifuged, then the supernatant was discarded and the bacteria thallus was diluted with a saline solution. This process was repeated

---

**Figure 1.** Schematic diagram of the IVCW system.

**Table 1.** Morphologic, physiologic and biochemical characteristics of the bacterial strains 1, 4 and 5.

| identification item               | Strain 1                      | Strain 4                      | Strain 5                      |
|-----------------------------------|------------------------------|------------------------------|------------------------------|
| colony shape                      | translucent, round           | opaque, round                 | translucent, irregular       |
| colony colour                     | milky                         | milky                         | yellow                       |
| colony state                      | moist, neat edge             | wet, irregular edges          | moist, neat edge             |
| microbial category                | bacterial                     | bacterial                     | bacterial                     |
| Gram staining                     | +                             | −                            | +                            |
| gelatin hydrolysis                | −                             | +                            | +                            |
| oxidase (V-P) assay               | −                             | −                            | +                            |
| membrane experiments              | +                             | −                            | +                            |
| oxidation and fermentation of glucose | +                         | −                             | +                            |
| oxidation and fermentation of sucrose | −                          | −                             | −                            |
| oxidation and fermentation of lactose | +                         | −                             | +                            |
| methyl red experiment             | +                             | −                            | +                            |
| citric acid experiment            | +                             | −                            | −                            |
| H₂S production experiment         | −                             | −                            | +                            |

---
Table 2. Parameters of inflow of the pH and temperature. DO, dissolved oxygen; COD, chemical oxygen demand; TSS, total suspended solids; TP, total phosphorus; NH$_3$–N, ammonium nitrogen; TN, total nitrogen.

| index | pH  | temperature (°C) | DO (mg l$^{-1}$) | COD (mg l$^{-1}$) | TSS (mg l$^{-1}$) | TP (mg l$^{-1}$) | NH$_3$–N (mg l$^{-1}$) | NO$_3^-$–N (mg l$^{-1}$) | NO$_2^-$–N (mg l$^{-1}$) | TN (mg l$^{-1}$) |
|-------|-----|------------------|------------------|------------------|------------------|------------------|----------------------|------------------------|------------------------|----------------|
| influent | 7.2 ~ 7.8 | 6 ~ 10 | 3.4 ~ 3.8 | 480 ~ 500 | 2.1 ~ 2.5 | 12.7 ~ 13.8 | 43.3 ~ 47.4 | 30.1 ~ 37.1 | 2.0 ~ 2.5 | 82.5 ~ 84.5 |
| effluent | 7.0 ~ 7.4 | 7 ~ 10 | 0.2 ~ 0.4 | — | — | — | — | — | — | — |
three times so that the nutrient of the medium could be fully removed. In the end, the absorbance of the
solution was adjusted to 1.2 with the saline water. These bacterial solutions were then mixed together at
a volume ratio of 1:1:1.

In early December 2014 (water temperature between 4 and 10°C), suspensions of mixed Psychrotrophic
bacteria ($4.4 \times 10^9$ ml$^{-1}$) were injected into the IVCW system at a 0.5–5% volume/volume ratio to the
sewage. We also included a control treatment that was not incubated with any Psychrotrophic bacteria.
The concentrations of different parameters in effluent, including COD, NH$_3$–N, NO$_3$–N, NO$_2$–N, TN
and TP, were monitored constantly for three months.

All the parameters were analysed according to standard methods. Statistical analysis was performed
using Origin8.6 and SPSS19 software.

3. Results and analysis

3.1. Identification of mixed Psychrotrophic bacteria

Identification results of morphologic, physiological and biochemical characteristics of the bacterial
strains at 6(±1)°C can be found in figure 2.

Under cold-temperature conditions (6(±1)°C), Strain 1 was composed of a Gram-positive translucent
circular ivory colony, with a moist surface, and regular edge. Strain 4 was a Gram-negative opaque round
milky colony with a moist surface, and irregular edge. Strain 5 was a Gram-positive translucent yellow
colony with a moist surface and irregular edges. Results from SEM (scanning electron microscopy) are
shown in figure 2. Strains 1, 4 and 5 were all spherical with no flagella. According to the Shanghai
Biological Engineering Co., Ltd, the specific 16S rDNA sequence of Strain 1 was 1498 bp (GenBank
acceptance number KR083014), the sequence of Strain 4 was 1489 bp (GenBank acceptance number
KR083015), and the sequence of Strain 5 is 1489 bp (GenBank acceptance number KR083016). A BLAST
search was performed on the three strains, and the phylogenetic tree was analysed and constructed by
using Clustal W and PHYLIP software. The phylogenetic tree is shown in figure 3.

According to the physiological and biochemical characteristics and the 16S rDNA of strains, strains
1, 4 and 5 could be preliminarily identified as cold Bacillus, sphingolipids of Bacillus and Pseudomonas.
Strain 1 was named Psychrobacter TM-1, and strains 4 and 5 were named Sphingobacterium TM-2 and
Pseudomonas TM-3, respectively.

3.2. Effect of simulated wastewater treatment

The degradation efficiencies of COD, NH$_3$–N and TP from the simulated sewage at 6(±1)°C by these
mixed Psychrotrophic bacteria cultures were 86.83%, 65.95% and 56.08%, respectively. Average removal
efficiencies were 1.38, 1.17 and 1.11 times higher than the single strains of P. flav a WD-3 (GenBank
acceptance number JX114950) [20]. In addition, the removal performance was found to be very stable.

3.3. The effect of sewage treatment on an artificial wetland

When the hydraulic retention time (HRT) was 5 days and the temperature of water was 6–10°C, the water
treatment efficiency in the CW with different inoculation amounts of low-temperature mixed flora was
as shown in figures 3 and 4.
Figure 3. Phylogenetic tree of the bacterial strains 1, 4 and 5. The concentration of COD, TN, NH₃⁻N, NO₂⁻N, NO₃⁻N and TP changes with time in the CW (as figure a, b, c, d, e and f).

As shown in figure 3, between 0.5% and 5.0% bacteria the purgative efficiency of sewage in wetland system increased as the dosage of the mixed Psychrotrophic bacteria was increased. Indeed, when the amount of bacteria was increased from 0.5% to 2.5%, the sewage purification efficacy of the wetland system was significantly improved. When the amount of bacteria was increased from 2.5% to 5.0%, the
purification efficacy was improved; however, the improvement was not significant. When the combined bacterium dosage was 5.0% the removal of the pollutants was improved; however, the cost was too high to make it feasible. Considering the operating costs and the efficaciousness of sewage treatment, 2.5% should be chosen as the best dosage of the mixed Psychrotrophic bacteria for the CW.

It can be seen in figure 4 that when the HRT was 5 days, the water temperature was 6–10°C and the dosage of combined bacterial liquid was 2.5%. As the hydraulic time was increased, the removal
efficiencies of COD, NH$_3$-N, NO$_2^-$-N, NO$_3^-$-N, TN and TP reached 91.8%, 91.1%, 88.0%, 93.8%, 94.8% and 95.2%, respectively, and the removal rates became 1.5, 2.0, 2.1, 1.5, 2.2 and 1.3 times higher than those of the control group. Furthermore, the average concentrations of COD, NH$_3$-N, TN and TP in the effluent were 39.56, 4.21, 4.36 and 0.64 mg l$^{-1}$, which met the first grade of Chinese national pollutant discharge standards for municipal wastewater treatment plants (GB18918-2002).

Our results showed that the removal efficiency of COD and NH$_3$-N in wetland sewage was 6.4% and 6.5% higher than expected when the removal rate of the single _P. flava_ WD-3 was 86.3% and 83.5% [20]. Other indicators of sewage treatment filtration did not show this level of improvement. Indeed, there were no significant differences among these indicators. Furthermore, the HRT was shortened by 5 days when the mixed _Psychrotrophic_ bacteria was used in the CS (5 days versus the 10 days taken when using only _P. flava_ WD-3 [20]). There this bacterial application both increased the efficacy of sewage treatment and reduced the cost of sewage purification.

Compared with one single low-temperature bacteria strain, the mixed bacteria strain better purified the sewage and shortened the HRT. In comparison, Bott & Love [21] treated aquaculture sewage with the mixed bacterial culture of _Bacillus subtilis_ and photosynthetic bacteria, and found that the mixed bacterial culture had a noticeable effect on flocculation and could display synergistic properties. They found that the removal efficiencies for COD, TP and NH$_3$-N were 62.35%, 66.78% and 52.60% after 48 h of bacterial culture had a noticeable effect on flocculation and could display synergistic properties. They added a mixture of bacteria to SBR directionally and found that the time of domestication and aeration was shortened, and the disposal ability and the effects of sequencing batch reactor (SBR) process were significantly improved, with a COD removal efficiency of greater than 92%. The wastewater purification abilities of _B. subtilis, Saccharomyces, Lactobacillus_ and mixed strains were studied by Morikawa [23], and the results showed that the mixture of flora had a synergistic effect, improving the disposal capacity of the pollutants, with NH$_3$-N and removal efficiencies reaching 93.2% and 97.8%, respectively. Therefore, a significant amount of prior research has shown that there are mutually beneficial relationships among the strains, and that mixing bacterial strains can have a synergistic effect in terms of contaminant removal efficacy. Moreover, these studies also indicated that there is potential to use these mixed bacterial strains to dispose of low-temperature sewage.

### 3.4. Wastewater treatment dynamics equation

The study of kinetics could optimize the technology and the methods associated with the biochemical disposing process. Indeed, the patterns of the microbial degradation could be predicted by establishing kinetic degradation models and simulating degradation with modern technology. The models used in the construction of the CWs were typically first-order kinetic models. This basic design equation (first-order kinetic model) has been widely applied in Australia, Europe and the USA for the prediction of bacterial treatment results. In spite of certain limitations, first-order kinetic models are still regarded as the most suitable for describing the process of degradation for pollutants in CWs. This is because calculation parameters can be worked out easily and the calculation process is simple. Jou et al. investigated the feasibility of using a CW to restore a heavily polluted creek, and estimated the reductions in biochemical oxygen demand (BOD) and nitrogenous biochemical oxygen demand (NBOD) using the first-order kinetic model in a laboratory wetland system, which provided some basis for using this model when designing the constructed wetlands [24].

As for the first-order kinetic equation applied in CWs, the main consideration was the relationship between the disposal load and the processing efficiency. The derivation of the model used here was based on the degradation of the matrix following the first-order reaction kinetics. The first-order kinetic model in a laboratory wetland system, which provided some basis for using this model when designing the constructed wetland systems, was typically first-order kinetic models. This basic design equation (first-order kinetic model) has been widely applied in Australia, Europe and the USA for the prediction of bacterial treatment results. In spite of certain limitations, first-order kinetic models are still regarded as the most suitable for describing the process of degradation for pollutants in CWs. This is because calculation parameters can be worked out easily and the calculation process is simple. Jou et al. investigated the feasibility of using a CW to restore a heavily polluted creek, and estimated the reductions in biochemical oxygen demand (BOD) and nitrogenous biochemical oxygen demand (NBOD) using the first-order kinetic model in a laboratory wetland system, which provided some basis for using this model when designing the constructed wetlands [24].

As for the first-order kinetic equation applied in CWs, the main consideration was the relationship between the disposal load and the processing efficiency. The derivation of the model used here was based on the degradation of the matrix following the first-order reaction kinetics. The first-order kinetic model for the degradation of contaminants in CW was [25, 26] as follows:

$$C_0 = C_e \exp(-k_e \cdot t) \quad (3.1)$$

$$k_e = -\frac{1}{t} \ln \left(\frac{C_0}{C_e}\right). \quad (3.2)$$

In formulae (3.1) and (3.2): $k_e$—contaminant volume removal rate constant, d$^{-1}$; $C_e$—influent concentration, mg l$^{-1}$; $C_0$—effluent concentration, mg l$^{-1}$; $t$—hydraulic retention time, d.

Based on the kinetic models listed above, the concentration of the pollutants in the effluent flow was denoted $C_e$ (hydraulic retention time, HRT $\leq$ 5 days) and the concentration of the pollutants in the influent flow was denoted $C_0$ when the dosage differed. In $C_e/C_0$ was taken as the ordinate and $t$(time)
Table 3. First-order kinetics model and the $R^2$ value of pollutants removal in the CW.

|pollution parameter| dosage (%)| First-order kinetics model| $R^2$ value| pollution parameter| dosage (%)| first-order kinetics model| $R^2$ value|
|-------------------|-----------|----------------------------|------------|-------------------|-----------|----------------------------|------------|
|COD               | 0.0       | $Y = -0.1662X - 0.0965$ | 0.9692     | NH$_3$–N         | 0.0       | $Y = -0.1439X + 0.1523$ | 0.9689     |
|                   | 0.50      | $Y = -0.2481X + 0.0216$ | 0.9501     |                   | 0.50      | $Y = -0.1716X + 0.0579$ | 0.9613     |
|                   | 1.25      | $Y = -0.3561X + 0.0204$ | 0.9874     |                   | 1.25      | $Y = -0.2442X - 0.0290$ | 0.9560     |
|                   | 2.50      | $Y = -0.5205X + 0.1297$ | 0.9895     |                   | 2.50      | $Y = -0.4361X - 0.1113$ | 0.9746     |
|                   | 5.00      | $Y = -0.5450X + 0.1331$ | 0.9952     |                   | 5.00      | $Y = -0.4449X - 0.2924$ | 0.9800     |
|NO$_3$–N         | 0.0       | $Y = -0.2022X - 0.0059$ | 0.9809     | NO$_2$–N         | 0.0       | $Y = -0.0918X - 0.0829$ | 0.9949     |
|                   | 0.50      | $Y = -0.2836X - 0.0619$ | 0.9730     |                   | 0.50      | $Y = -0.1178X - 0.1634$ | 0.9894     |
|                   | 1.25      | $Y = -0.3155X - 0.1673$ | 0.9650     |                   | 1.25      | $Y = -0.1760X - 0.1539$ | 0.9891     |
|                   | 2.50      | $Y = -0.4291X - 0.5734$ | 0.9813     |                   | 2.50      | $Y = -0.3734X - 0.0896$ | 0.9469     |
|                   | 5.00      | $Y = -0.5085X - 0.5269$ | 0.9699     |                   | 5.00      | $Y = -0.4353X - 0.0822$ | 0.9250     |
|TN                | 0.0       | $Y = -0.1259X + 0.0517$ | 0.9868     | TP               | 0.0       | $Y = -0.3009X + 0.2262$ | 0.9494     |
|                   | 0.50      | $Y = -0.2109X + 0.0552$ | 0.9823     |                   | 0.50      | $Y = -0.3331X + 0.0370$ | 0.9895     |
|                   | 1.25      | $Y = -0.2496X - 0.0630$ | 0.9826     |                   | 1.25      | $Y = -0.4280X - 0.0559$ | 0.9865     |
|                   | 2.50      | $Y = -0.5867X + 0.2602$ | 0.9387     |                   | 2.50      | $Y = -0.5638X + 0.0163$ | 0.9490     |
|                   | 5.00      | $Y = -0.6186X + 0.2274$ | 0.9186     |                   | 5.00      | $Y = -0.6153X + 0.0806$ | 0.9325     |
Table 4. The $k_v$ in the pseudo-first order reaction of pollutants removal in the CW.

| pollution parameter | dosage (%) | 0.00 | 0.50 | 1.25 | 2.50 | 5.00 |
|---------------------|------------|------|------|------|------|------|
| COD                 |            | -0.1843 | -0.2521 | -0.3732 | -0.4991 | -0.5190 |
| NH$_3$–N            |            | -0.1793 | -0.1734 | -0.2646 | -0.4413 | -0.6977 |
| NO$_2$–N            |            | -0.0692 | -0.1318 | -0.1802 | -0.2764 | -0.4979 |
| NO$_3$–N            |            | -0.1630 | -0.3106 | -0.3712 | -0.5174 | -0.3975 |
| TN                  |            | -0.0891 | -0.2065 | -0.2661 | -0.2848 | -0.3054 |
| TP                  |            | -0.0950 | -0.3289 | -0.3386 | -0.3571 | -0.5912 |

as the abscissa. The time curves of COD, NH$_3$–N, NO$_2$–N, NO$_3$–N, TN and TP are shown in figure 5. The first-order kinetic model and $R$ value of each contaminant are shown in table 3. The measurement results of COD, NH$_3$–N, NO$_2$–N, NO$_3$–N, TN and TP were substituted into equation (3.2) at HRT of 5 days, and $k_v$ was calculated as shown in table 4.

The volume degradation rate $k_v$ represents the treatment efficiency of the contaminants. Mixed Psychrotrophic bacterial flora had a noticeably high disposal rate for pollutants at a HRT of 5 days. Furthermore, the degradation rates of COD, NH$_3$–N, NO$_2$–N, NO$_3$–N, TN and TP significantly increased as the dosage of bacteria was increased. The correlation coefficients ($R$) of each $k_v$ reached 0.9816, 0.9597, 0.9473, 0.9934, 0.9267 and 0.9183. Through the analysis of treatment efficiency for the pollutants in the CW at different dosages, it is evident that the removal kinetics of contaminants in the CW was in accordance with the first-order kinetics model.

4. Conclusion

(1) The nine bacterial strains, isolated from artificial wetland sediments, were cold-resistant bacteria. Following screening, a combination of the best strains (1, 4 and 5) had the highest removal efficiency for COD, NH$_3$–N and TP in wastewater. Removal rates for this mixed Psychrotrophic bacteria strain were 86.83%, 65.95% and 56.08%, respectively. The strains 1, 4 and 5 were preliminarily identified as cold Bacillus, sphingolipids of Bacillus and Pseudomonas, and were named Psychrobacter TM-1, Sphingobacterium TM-2 and Pseudomonas TM-3.

(2) The effectiveness of the IV CW system with the mixed Psychrotrophic bacteria strain in winter conditions was demonstrated through the increased removal of COD, NH$_3$–N, NO$_2$–N, NO$_3$–N, TN and TP. At high concentrations of organic substrate, the dosage of the mixed Psychrotrophic bacteria strain and the removal rate of the contaminants were positively correlated and followed a first-order rate equation. We also evaluated the effects of different bacterial dosages and determined that the optimal dosage of this mixed Psychrotrophic bacteria strain was 2.5% for these CW systems when considering biological effect, time, cost and resource consumption.

Data accessibility. We include all the experimental data in the manuscript, and there is no electronic supplementary material.

Authors’ contributions. M.T. and Z.L. conceived the idea and designed the experiments. M.T., Z.L. Y.Y and J.C. contributed towards fabrication and characterization of materials. M.T., Z.L. Y.Y and J.C. analysed data and wrote the paper. All authors discussed the results and commented on the manuscript.

Competing interests. We declare we have no competing interests.

Funding. This research was supported by the National Natural Science Fund of China (no. 31700433 and no. 31672314).

References

1. Kivaisi AK. 2001 The potential for constructed wetlands for wastewater treatment and reuse in developing countries: a review. Ecol. Eng. 16, 545–560. (doi:10.1016/S0167-8775(00)00138-0)
2. Wood J, Fernandez G, Barker A. 2007 Efficiency of reed beds in treating dairy wastewater. Bio syst. Eng. 98, 455–469. (doi:10.1016/j.biosystemseng.2007.09.022)
3. Kynkaanniemi P, Ulen B, Torstensson G. 2013 Phosphorus retention in a newly constructed wetland receiving agricultural tile drainage water. J. Environ. Qual. 42, 596–605. (doi:10.2134/jeq2012.0266)
4. Sgroi M, Pelissari C, Rocca R, Szerino P, Garcia J, Valliasindi, F, Avila C. 2017 Removal of organic carbon, nitrogen, emerging contaminants and fluorescing organic matter in different constructed wetland configurations. Chem. Eng. J. 332, 619–627. (doi:10.1016/j.cej.2017.09.122)
5. Calheiros CSC, Quintero PVB, Silva G. 2012 Use of constructed wetland systems with Arundo and Sarcosoma for polishing high salinity tannery wastewater. J. Environ. Manage. 95, 66–71. (doi:10.1016/j.jenvman.2011.10.003)

6. Justin MZ, Zupancic M. 2009 Combined purification and reuse of landfill leachate by constructed wetland and irrigation of grass and willows. Desalination 246, 157–168. (doi:10.1016/j.desal.2008.03.049)

7. Chen J, Liu S, Yan J, Wen J, Hu Y, Zhang W. 2017 Intensive removal efficiency and mechanisms of carbon and ammonium in municipal wastewater treatment plant tail water by ozone oyster shells fix-bed bioreactor—membrane bioreactor combined system. Eco. Eng. 101, 75–83. (doi:10.1016/j.ecoleng.2016.11.029)

8. Yan Z, Chen J, Liu Y, Shao J, Shu P, Wen S. 2017 Effects of oxytetracycline on bacterial diversity in livestock wastewater. Environ. Eng. Sci. 34, 265–271. (doi:10.1089/ees.2016.0425)

9. Istenic D, Arías CA, Vollertsen J. 2012 Foreword. Environ. Sci. Heal. A. 47, 919. (doi:10.1805/j.19934529.2012.667287)

10. Stottmeister U, Wiessner A, Kuschk P, Kappelmeyer J. 2009 Combined purification and reuse of landfill leachate by constructed wetland and irrigation of grass and willows. Desalination 246, 157–168. (doi:10.1016/j.desal.2008.03.049)

11. Chen J, Zhang L, Hu Y, Huang W, Niu Z, Sun J. 2017 Bacterial community shift and improved performance induced by in situ preparing dual graphene modified bioelectrode in microbial fuel cell. Biosour. Technol. 241, 220–227. (doi:10.1016/j.biosourcetech.2017.05.123)

12. Nguyen LM. 2000 Organic matter composition, microbial biomass and microbial activity in gravel-bed constructed wetlands treating farm dairy wastewaters. Eco. Eng. 16, 199–221. (doi:10.1016/S0925-8574(00)00044-6)

13. Pei Y, Yang Z, Tian B. 2010 Nitrate removal by microbial enhancement in a riparian wetland. Biosour. Technol. 101, 5712–5718. (doi:10.1016/j.biosourcetech.2010.02.005)

14. Shao Y, Pei H, Hu W. 2013 Nitrogen removal by bioaugmentation in constructed wetlands for rural domestic wastewater in autumn. Desal. Water Treat. 51, 6624–6631. (doi:10.1080/19443994.2013.791797)

15. Templar PH, Weathers KC. 2011 Use of mixed ion exchange resin and the denitrifier method to determine isotopic values of nitrate in atmospheric deposition and canopy throughfall. Atmos. Environ. 45, 2017–2020. (doi:10.1016/j.atmosenv.2011.01.035)

16. Tang M, Zhang F, Yao S, Liu Y, Chen J. 2015 Application of Pseudomonas flavus WD-3 for sewage treatment in constructed wetland in winter. Environ. Technol. 36, 1205–1211. (doi:10.1080/21622315.2014.983183)

17. Chen J, Hu Y, Huang W, Zhang L. 2017 Enhanced electricity generation for biocathode microbial fuel cell by in situ microbial-induced reduction of graphene oxide and polarity reversion. Int. J. Hydrogen Energy. 42, 12574–12582. (doi:10.1016/j.ijhylene.2017.03.012)

18. Chen J, Hu Y, Zhang L, Huang W, Sun J. 2017 Bacterial community shift and improved performance induced by in situ preparing dual graphene modified bioelectrode in microbial fuel cell. Biosour. Technol. 238, 273–280. (doi:10.1016/j.biosourcetech.2017.04.044)

19. Chen J, Hu Y, Tan X, Zhang L, Huang W, Sun J. 2017 Enhanced performance of microbial fuel cell with in situ preparing dual graphene modified bioelectrode. Biosour. Technol. 241, 735–742. (doi:10.1016/j.biosourcetech.2017.06.020)

20. Tang M, Ding P, Xia X, Chen Y, Zheng M, Han Z, Yuan M. 2014 Application of Pseudomonas flavus WD-3 for the sewage purification in the artificial wetland. Acta Sci. Circumstantiae 34, 1955–2000. (doi:10.13671/j.hjkxxb.2014.0535)

21. Bott CB, Love NG. 2004 Implicating the glutathione-gated potassium efflux system as a cause of electrophile-induced activated sludge deflocculation. Appl. Environ. Microb. 70, 5569–5578. (doi:10.1128/AEM.70.9.5569-5578.2004)

22. Moussavi G, Heidanazad M. 2011 The performance of SBR, SCR, and MSCR for simultaneous biodegradation of high concentrations of formaldehyde and ammonia. Sep. Purr. Technol. 77, 187–195. (doi:10.1016/j.seppur.2010.11.028)

23. Monikawa M. 2016 Beneficial biofilm formation by industrial bacteria Bacillus subtilis and related species. J. Biosci. Bioeng. 101, 1–8. (doi:10.1263/jb.101.1)

24. Joss CJ, Chen SW, Kao CM, Lee CL. 2008 Assessing the efficiency of a constructed wetland using a first-order biokinetic model. Wetlands 28, 215–219. (doi:10.1672/07-60.1)

25. Kadlec RH. 2000 The inadequacy of first-order treatment wetland models. Eco. Eng. 15, 105–119. (doi:10.1016/S0925-8574(99)00039-7)

26. Gai L, Li W, Zhang Y, Wei J, Lei Y, Zhang M. 2016. Nitrogen removal in a horizontal subsurface flow constructed wetland estimated using the first-order kinetic model. Water 8, 534. (doi:10.3390/w8110534)