Synthesis, gastroprotective and acute toxicity of bio-isosteric derivative of diclofenac

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Abstract

Background and objective: Non-steroidal anti-inflammatory drugs (such as diclofenac) had been widely prescribed for the treatment of different types of pain; however, they are not devoid of adverse effects. Therefore, synthesis of new bio-isosteric analogs of diclofenac with greater COX II selectivity and less gastrointestinal side effect is demanded. This study aimed to evaluate the acute toxicity and gastroprotective activity of the bio-isosteric derivative of diclofenac against ethanol-induced gastric ulceration in rats.

Methods: 2-Cumarlanone 2 had been utilized to prepare amides 3a-e then after the bio-isosteric derivatives 4a-esynthesized from them then used to study the biological activity by testing whether high doses of the prototype compound 4c are toxic or not on albino mice and measuring the gastroprotective effect on albino rats. Rats were divided into four groups. Group 1 orally administered with Tween 20; group 2 was orally administered with 20 mg/kg esomeprazole; groups 3 and 4 received 100 and 200 mg/kg of the compound, respectively. Absolute ethanol was given orally to the groups, and rats were sacrificed after one hour.

Results: Few diarylethers 4a-e bio-isosteric to diclofenac had been prepared and fully characterized (including 1HNMR, 13CNMR, and IR spectroscopy). Serum biochemical parameters were reported to be normal. Hematological analysis of kidney and liver did not elicit any remarkable changes in the treated group compared to the control group. Thus, the 50% oral lethal dose (LD50) for the male and female mice was greater than 5 g/kg body weight. Anti-ulcer data showed a gastroprotective effect of the bio-isosteric diclofenac derivative 4c and presented the ulcer area inhibition, low stomach pH, and preserve the mucous content.

Conclusion: Many bio-isosteric derivatives of diclofenac had been prepared with good yields. The synthesized derivative 4c showed no toxicity, and the gastroprotective effect may possibly be due to the preservation of gastric wall mucus.

Keywords: NSAIDs; Diclofenac; Diarylether; Bio-isostere; Gastroprotective.
bleeding. However, it would be fatal in a few cases. The gastrointestinal irritation is because of the inactivation of the COX I, while the desired clinical efficacy is due to the COX II inactivation.\(^{10-12}\) In order to counter the side effects of NSAIDs (such as diclofenac), many prodrug approaches had been reported,\(^{13,14}\) though, bio-isosteric derivatives of diclofenac might also be useful. In the latter case, diarylether analogue 4a-e to diclofenac (diphenylamine, as shown in scheme 1) 1 could be prepared, and converting the carboxylic acid to a less acidic functionality such as an amide moiety to reduce the direct gastrointestinal irritation and indirect irritation due to greater selectivity to COX II.\(^{6}\) The current study is conducted to evaluate the acute toxicity and gastroprotective activity of the bio-isosteric derivative of diclofenac against ethanol-induced gastric ulceration in rats.

**Methods**

This study design was of an experimental type with simple randomization assignments, in which the chemical changes to the diclofenac was assumed to lead to no changes in the toxicity outcomes of albino rats and expected to possess a gastroprotective effect. The study was conducted at the College of Pharmacy and College of Medicine between April and September 2018.

**Chemistry**

The chemical reactions (as shown in scheme 1) had been carried out at the Organic and Pharmaceutical Chemistry Lab., at Hawler Medical University, College of Pharmacy. Commercially available reagents were used as received without purification. Analytical thin layer chromatography (TLC) was performed with plastic-backed TLC plates coated with silica G/\(\text{UV}_{254}\), in a variety of solvents. The plates were visualized by UV light (254 nm).\(^1\)\(^{1}\), and \(^{13}\)C NMR spectra were recorded on a BrukerUltra Shield 300 (300 MHz) spectrometer. All chemical shifts (\(\delta\)) are quoted in parts per million (ppm) relative to a calibration reference of the residual protic solvent; \(\text{CHCl}_3\) (\(\delta\)\(\text{H}\) 7.26, s) was used as the internal standard in \(^1\)H NMR spectra, and \(^{13}\)C NMR shifts were referenced using CDCl\(_3\) (\(\delta\)\(\text{C}\) 77.16, t) with broadband decoupling and the \(J\) values are measured in Hertz (Central Lab., University of Jordon). Melting points of the synthesized compounds were measured on the electrothermal melting point apparatus (Gallenkamp, UK) and are uncorrected. IR spectra were recorded on a JASCO, Japan (Pharmaceutical Chemistry Department, College of Pharmacy, Hawler Medical University/ Erbil).

**Scheme 1:** Synthesis of bio-isosteric derivatives of diclofenac 4a-e.

Reaction conditions: i) amines \(\text{R}(1.5\text{ eq.}),\) 2-cumaranone 2 (1 eq.), toluene (0.1 M), reflux, 6 hours; ii) acetamides 3a-e (1 eq.), 1-fluoro-2-nitrobenzene (FNB)\(^5\) (10 eq.), \(\text{K}_2\text{CO}_3\) (2.5 eq.), DMSO (0.1 M), 25° C, 24 hours.\(^{15}\)
1.1 General procedure for the synthesis of amides 3a-e
To a solution of 2-coumaranone (1 eq.) in toluene (0.1 M) was added the amine (1.5 eq.). The resulting solution was stirred at 110 °C for 6 hours, after which it was allowed to cool, acidified (1 M HCl), and extracted with ethyl acetate. The combined organic layers were washed with distilled water and brine, dried (MgSO₄), filtered and the solvent evaporated in vacuo to afford the pure amide directly or after flash column chromatography.

1.2 General procedure for the synthesis of bio-isosteric diarylethers 4a-e
To a solution of the prepared amides 3a-e (1 eq.) in DMSO (0.1 M) was added potassium carbonate (2.5 eq.), and the resulting solution was stirred at 25 °C for 30 minutes. 1-Fluoro-2-nitrobenzene FNB 5 (10 eq.) was added, and the reaction stirred for 24 hours at 25°C. After which, the mixture was acidified with hydrochloric acid solution (1 M). The product was extracted with ethyl acetate, and the combined organic layers were washed with distilled water, brine, dried (MgSO₄), filtered and the solvent evaporated in vacuo to afford the crude residue which was purified by flash column chromatography.

Biological study
2.1 Acute toxicity study
To determine a safe range of doses for the bio-isosteric derivatives of diclofenac, toxicity test was carried out as previously described. Compound 4b selected as the prototype as it showed the highest COX II inhibition activity. In brief, 36 mice, including 18 females and 18 males were divided into three groups categorized as the vehicle (10% Tween 20), low dose of bio-isosteric derivative of diclofenac 4c (2 g/kg) and high dose of bio-isosteric derivative of diclofenac 4c (5 g/kg). The mice were fasted for 16 h prior to the dosing (water was accessible except for the last two h). After the dosing, food was withdrawn for another one to three hours. Any signs of toxicities and mortality were documented during the next two weeks. On day 15, the animals were sacrificed, and blood serum samples were collected for hematological analysis.

2.2 Ethanol-induced gastric ulceration
Healthy male albino rats (150–180 g, 6–8 weeks old) were obtained from the Animal House Unit, College of Medicine, Hawler Medical University. The animals were allowed access to standard rat pellets and RO (reverse osmosis) water. Entirely animal experiments were conceded in agreement with the prior approval from the Ethics committee of College of Pharmacy, Hawler Medical University (Ethics No. 181112/81) Date. The preventive effect of bio-isosteric derivative of diclofenac 4c against superficial hemorrhagic mucosal lesions was investigated in the normal rats. Prior to the experiment, albino male rats were fasted for 24 hours (water was accessible except for the last two hours). Twenty four rats were divided randomly into four groups of 6 rats each and pre-treated accordingly (Table 1). The rats were sacrificed one hour later with xylazine and ketamine, and their stomachs were immediately excised. The stomach of each rat was dissected along the greater curvature, and pH-meter titration with 0.1 N NaOH was used to analyze the hydrogen ion concentration in the gastric contents expressed in mEq/l value. Then, a glass slide was applied to gently scrape the gastric mucosa of the rats followed by the weighing of the obtained mucus with a precision electronic balance.

Table 1: The experimental design and specifications of the animal study.

| Groups   | Description          | Pre-treatment          | Treatment        |
|----------|----------------------|------------------------|------------------|
| Group 1  | Ulcer control        | 10% Tween 20 (5 ml/kg)| absolute ethanol |
| Group 2  | Treatment control    | Esomeprazole (20 mg/kg)| absolute ethanol |
| Group 3  | Experimental group1  | LD (100 mg/kg)         | absolute ethanol |
| Group 4  | Experimental group2  | HD (200 mg/kg)         | absolute ethanol |
### 2.3 Macroscopic analysis of lesions

The hemorrhagic damage of the stomach was determined by the assessment of the luminal surface. The ulcer inhibition percentage (I%) of each pre-treatment was calculated using a planimeter (10 × 10 mm²) and a dissecting microscope (1.8×) where UC and UT were the ulcer area of the control and treated group, respectively. The measurement of the ulcer area was performed as previously described in detail by Abdelwahab et al.\textsuperscript{17}

\[ (I\%) = \left[ \frac{(UA_{\text{control}} - UA_{\text{treated}})}{UA_{\text{control}}} \right] \times 100. \]

### Statistical analysis

All values were reported as mean ± SEM. The statistical analysis was done using the statistical package for the social sciences (version 23). One-way ANOVA was used to compare means. A value of \( P < 0.05 \) was considered significant.

### Results

The acetamides 3a-e had been prepared from 2-cumaranone 2 and corresponding amines R with a good yield and fully characterized by different spectroscopic methods, as shown in Tables 2 and 3. After the successful preparation of the amide substrates 3a-e, their reaction with FNB 5 afforded the bio-isosteric diarylether derivatives 4a-e in their maximum percents of yield, as shown in Tables 2 and 3.

### Table 2: Physical data and isolated yields of the amides 3a-e and bio-isosteric diarylethers 4a-e.

| Entry | Yield (%) | Colour         | Melting point °C | \( R_f \) (30% EtOAc in petroleum ether) |
|-------|-----------|----------------|------------------|----------------------------------------|
| 3a    | 96        | Colorless      | 88-90            | 0.56                                   |
| 3b    | 95        | Yellow         | 139-141          | 0.69                                   |
| 3c    | 89        | Colorless      | 120-122          | 0.52                                   |
| 3d    | 98        | Yellow         | 107-109          | 0.48                                   |
| 3e    | 87        | Yellow         | 123-125          | 0.37                                   |
| 4a    | 100       | Yellow, oily substance | | 0.74                                   |
| 4b    | 100       | Yellow, oily substance | | 0.48                                   |
| 4c    | 100       | Yellow, oily substance | | 0.41                                   |
| 4d    | 100       | Yellow         | 74-76            | 0.57                                   |
| 4e    | 100       | Yellow         | oily substance   | 0.39                                   |
Table 3: Diagnostics peaks and values in IR, $^1$H NMR, and $^{13}$C NMR of the amides 3a-e and bio-isosteric diarylethers 4a-e.

| Comp. | IR (cm$^{-1}$) | $^1$H NMR (ppm) | $^{13}$C NMR (ppm) |
|-------|----------------|-----------------|---------------------|
| 3a    | 1618 (C=O stretching), 3173 (OH stretching), 1090 (C-O stretching) | 1.13 (t, 3H, $J$ = 7.5Hz), 1.29 (t, 3H, $J$ = 6.0Hz), 3.39 (q, 2H, $J$ = 6.0Hz), 3.50 (q, 2H, $J$ = 7.5Hz), 3.71 (s, 2H), 10.47 (s, 1H) | 13.05, 14.94 (CH$_3$), 37.06, 41.47, 43.67 (CH$_2$) |
|       | (C=O stretching), 3064 (O-H stretching), 1477 (C-O stretching) | 3.81 (s, 2H), 4.63 (s, 2H), 4.64 (s, 2H), 10.07 (s, 1H) | 37.18, 49.33, 51.26, (CH$_2$) |
| 3c    | 1610 (C=O stretching), 2959 (OH stretching), 1094 (C-O stretching) | 1.89 (pent, 2H, $J$ = 6.0Hz), 2.01 (pent, 2H, $J$ = 6.0Hz), 3.48 (t, 2H, $J$ = 7.5Hz), 3.68 (t, 2H, $J$ = 6.0Hz), 3.70 (s, 2H), 10.37 (s, 1H) | 24.53, 25.12, 39.03, 46.34, 47.77 (CH$_2$) |
| 3d    | 1616 (C=O stretching), 3166 (OH stretching), 1039 (C-O stretching) | 1.52-1.63 (m, 6H), 3.55 (t, 2H, $J$ = 6.0Hz), 3.61 (t, 2H, $J$ = 6.0Hz), 3.73 (s, 2H), 9.87 (s, 1H) | 26.52, 36.41, 43.45, 48.16, (CH$_2$) |
| 3e    | 1615 (C=O stretching), 2931 (OH stretching), 1092 (C-O stretching) | 3.65-3.68 (m, 8H), 3.74 (s, 2H), 9.56 (s, 1H) | 36.31, 42.59, 47.29, 66.51, 66.65 (CH$_2$) |
| 4a    | 1638 (C=O stretching), 1098 (C-O stretching), 1526, 1368 (N-O stretching) | 0.97 (t, 3H, $J$ = 7.5Hz), 1.09 (t, 3H, $J$ = 7.5Hz), 3.25-3.35 (m, 4H), 3.71 (s, 2H) | 12.74, 14.02 (CH$_3$), 34.25, 40.31, 42.33 (CH$_2$) |
| 4b    | 1644 (C=O stretching), 1079 (C-O stretching), 1523, 1349 (N-O stretching) | 3.90 (s, 2H), 4.55 (s, 2H), 4.59 (s, 2H) | 34.80, 48.65, 50.38 (CH$_2$) |
| 4c    | 1636 (C=O stretching), 1247 (C-N stretching), 1098 (C-O stretching), 1522, 1345 (N-O stretching) | 1.69-1.92 (m, 4H), 3.31 (t, 2H, $J$ = 6.0Hz), 3.43 (t, 2H, $J$ = 7.5Hz), 3.67 (s, 2H) | 24.43, 26.15, 35.89, 45.93, 46.89 (CH$_2$) |
| 4d    | 1642 (C=O stretching), 1068 (C-O stretching), 1584, 1350 (N-O stretching) | 1.48-1.57 (m, 4H), 1.65-1.70 (m, 2H), 3.52 (t, 2H, $J$ = 6.0Hz), 3.61 (t, 2H, $J$ = 6.0Hz), 3.86 (s, 2H) | 24.37, 25.45, 26.16, 34.24, 42.88, 46.99 (CH$_2$) |
| 4e    | 1642 (C=O stretching), 1068 (C-O stretching), 1584, 1350 (N-O stretching) | 3.47-352 (m, 8H), 3.71 (s, 2H) | 34.09, 42.25, 46.37, 66.64, 66.79 (CH$_2$) |
In the acute toxicity study (high doses), all mice survived and did not exhibit any sign of toxicity and abnormality at 2 and 5 g/kg dosage. For a duration of 14 days, there were no behavioral changes, and no abnormal signs were observed. As shown in Table 4 and 5, the serum biochemical parameters were reported to be normal. Hematological analysis of kidney and liver did not elicit any remarkable changes in the treated group compared to the control group. Thus, the 50% oral lethal dose (LD$_{50}$) for the male and female mice was greater than 5 g/kg body weight. Regarding the Macroscopic results of the antiulcer study; Table 6 shows that after treatment with esomeprazole (positive control), the acidity was significantly attenuated while rats pre-treated with diclofenac derivative at high dose and low dose caused slight elevation of the stomach pH however the gastric mucus content was indeed washed-out in animals pre-treated with ethanol.

**Table 4:** Effect of bio-isosteric derivative of diclofenac on serum biochemical analysis (renal function test).

| Animal groups | Urea (mg/dL) | Creatinin (mg/dL) | Uric acid (mg/dL) | Calcium (mg/dL) | Phosphorus (mg/dL) | P value |
|---------------|--------------|--------------------|-------------------|----------------|-------------------|---------|
| Vehicle       | 43.5 ± 0.4   | 0.42 ± 0.1         | 5.7 ± 0.4         | 9.95 ± 0.6     | 8.1 ± 0.4          |         |
| LD (2 g/kg)   | 44.33 ± 0.3  | 0.39 ± 0.7         | 5.6 ± 0.5         | 9.97 ± 0.7     | 8.37 ± 0.7         | 0.251   |
| HD (5 g/kg)   | 43.00 ± 0.8  | 0.44 ± 0.03        | 5.9 ± 0.5         | 10.00 ± 0.4    | 7.97 ± 0.3         | 0.432   |

Values expressed as mean ± SEM. The results did not show any significant difference between groups.

**Table 5:** Effect of bio-isosteric derivative of diclofenac on serum biochemical analysis (Liver function test).

| Animal groups | Total Bilirubin (mg/dL) | Direct Bilirubin (mg/dL) | AST (U/L) | ALT (U/L) | ALP (U/L) | P value |
|---------------|-------------------------|--------------------------|-----------|-----------|-----------|---------|
| Vehicle       | 0.05 ± 0.4              | 0.01 ± 0.9               | 183.9 ± 1.8 | 37.5 ± 0.3 | 115.5 ± 0.1 |         |
| LD (2 g/kg)   | 0.06 ± 0.5              | 0.01 ± 0.5               | 177 ± 1.2  | 40.7 ± 0.1 | 137.3 ± 0.4 | 0.371   |
| HD (5 g/kg)   | 0.05 ± 1.9              | 0.01 ± 0.1               | 189.7 ± 1.5 | 35 ± 0.4  | 106 ± 0.1  | 0.402   |

Values expressed as mean ± SEM. The results did not show any significant difference between groups. ALT: alanine aminotransferase; AST: aspartate aminotransferase; ALP: alkaline phosphatase.

**Table 6:** Antiulcer activity of bio-isosteric derivative of diclofenac against ethanol-induced gastric injury.

| Animal group | Pre-treatment (5 ml/kg) | Ulcer area (mm$^2$) | Inhibition (%) | Mucus weight (g) | pH (mEq/l) | P value |
|--------------|-------------------------|---------------------|----------------|------------------|------------|---------|
| Vehicle      | 10% Tween 20            | 220.8 ± 0.3         | ...            | 0.34 ± 0.1       | 3.7 ± 0.6  |         |
| Control      | 20 Mg/kg Esomeprazole   | 55.2 ± 0.1*         | 95.1*          | 1.16 ± 0.9*      | 6.7 ± 0.2  | <0.001  |
| Low Dose     | 100 mg/kg derivative    | 74.4 ± 0.2*         | 63.5*          | 0.8 ± 0.4*       | 5.5 ± 0.1  | <0.001  |
| High Dose    | 200 mg/kg derivative    | 129.6 ± 0.3*        | 53.8*          | 0.7 ± 0.5*       | 5.7 ± 0.5  | <0.001  |

The values are expressed as the mean ± SEM. (*) Indicates significance compared to the ulcer positive group.
The administration of ethanol to the rats induced obvious hemorrhagic lesions in the gastric walls, as shown in Figure 1, while rats treated with diclofenac derivative and esomeprazole showed fewer areas of gastric ulcer formation in comparison with the vehicle control group. It is worthy to note that the treatment helped to flatten some of the gastric mucosal folds in rat's stomachs.

**Discussion**

Ameen had focused on developing the first example of chiral version of Truce-smiles rearrangement, and the amide substrates 3a-ehad been attempted. In his work, the diarylethers 4a-e documented that had significant COX inhibition, and the compound 4c showed maximum inhibition against COX II (70%). Therefore, the amides4a-e (derivatives of diclofenac 1, shown in scheme 2) had been prepared to apply the same procedure established by Snape and Ameen. As it can be shown from Tables 2 and 3, their physical properties and diagnostic peaks imply same documented in the literature.

**Scheme 2:** The structure of diclofenac 1 and its bio-isosteric derivatives 4a-e.

**Figure 1:** Effect of bio-isosteric derivative of diclofenac on macroscopic appearance of the gastric mucosa. The esomeprazole group shows mild injuries in comparison to the ulcer control group (A). Severe injuries are observed in the gastric mucosa of the ulcer control group (B). The HD group shows mild injuries to the gastric mucosa (C), while LD group shows moderate injuries in the gastric mucosa and flattened of the mucosal folds (D).
For the synthesis of the amides of the diarylethers 4a-e, the optimized protocol (using 10 equivalents of 1-fluoro-2-nitrobenzene FNB5) had been utilized to get 100% yield, and with no rearrangement products (compounds 6 and 7, demonstrated in scheme 3), as illustrated in Tables 2 and 3. Snape and Ameen found that by increasing the number of molar equivalents of FNB5 from 2.0 to 5.0 and to 10.0 equivalents, the amount of diarylethers 4a-e also increased, at all temperatures studied (25, 60 and 100 °C), with no rearranged products 6 and 7, as shown in scheme 3. They provide the exact explanation for this occurrence that certain counter ions rendering the electron-rich enolate of diarylethers 4a-e formed during the first stage of the reaction and unable to react further with FNB 5 to rearrange and give the rearranged Truce-Smiles products such as 6 and 7. Serum biochemical parameters of acute toxicity test were normal, and the hematological analysis of kidney and liver functions parameters did not elicit any remarkable changes in the treated group compared to the control group which means, that the 50% oral lethal dose (LD₅₀) for the male and female mice was greater than 5 g/kg body weight. The presented data thus agrees with previously published results in different animal species like mice, rats, pigs. Moreover, the results of the present study showed that the bio-isosteric derivative of diclofenac 4c possessed an anti-ulcer activity against ethanol-induced hemorrhagic mucosal lesions in rats. The prototype derivative 4c increased the gastric wall mucus. Indeed, the gastroprotective effect of the chemical compounds appeared to be mediated partially through the protection of gastric mucus secretion. Alterations in gastric motility are chief focus points in the prevention of experimental gastric lesions. The results of the current study showed an increase in the flattening of the stomach mucosal folds, which suggests that the gastroprotective effect of the diclofenac derivative was due to a decrease in gastric motility. The mechanism of the anti-ulcer activity could be through increasing the cyclooxygenase enzymes that exist in two isoforms (COX I and COX II), which are the key factors for the synthesis of prostaglandins that have been shown to inhibit gastric secretion, stimulate bicarbonate secretion, and increase gastric blood volume. COX I is constitutively expressed in the gastrointestinal tract in large quantities and has been suggested to maintain mucosal integrity through continuous generation of prostaglandins that are important in the prevention and maintenance of gastric mucosal integrity and ulcer healing. However, diclofenac is a well-known non-steroidal anti-inflammatory drug (NSAID), which acts by potent inhibition of both cyclooxygenase isoenzymes, COX I and COX II. Therefore, depending on the results of this

Scheme 3: The reaction of 10 equivalents of FNB 5 with the amides 3a-e.
study, the gastroprotective effect of the bio-
isosteric derivative 4c of diclofenac might be due to its cyclooxygenase lowering activity, however, further studies need to be done to confirm that expectation.

Conclusion

Many bio-isosteric derivatives of diclofenac had been prepared with good yields, as shown in Tables 1 and 3. The extra equivalents of FNB 5 were helpful in affording the prepared products in their maximum yields (100%). The synthesized derivative 4c is not toxic up to 5g/kg, and it possesses a gastroprotective effect. An increase in the mucus results in low ulcerogenic activity of 4c, and it may involve both low direct cytotoxicity due to a decrease in an aggressive factor (acid) and protective effect against the development of gastric lesions through an increase in a protective factor (mucus).

Competing interests

The author declares no competing interests.

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