Effects of Onion Extracts on Growth Performance, Carcass Characteristics and Blood Profiles of White Mini Broilers

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ABSTRACT: This experiment was carried out to investigate effects of onion extract on growth performance, meat quality and blood profiles of White mini broilers. Total of 600 one-d-old male White mini broiler chicks were divided into four groups and fed control diets (non-medicated commercial diet or antibiotics medicated) or experimental diets (non-medicated diets containing 0.3% or 0.5% onion extract) for 5 wks. The final body weight (BW) and weight gain of the group fed non-medicated control diet were lower than those of medicated control group (p<0.01). The chicks fed diet with 0.3% or 0.5% onion extract showed a similar BW to that of medicated control group. The relative weight of various organs, such as liver, spleen, bursa of Fabricius, abdominal fat, and the activities of serum enzymes were not affected by dietary treatments. There were no significant differences in meat color among groups. Whereas, groups fed diets containing onion extract had slightly lower cooking loss and higher shear force value, but not significantly. The concentrations of serum free cholesterol and triacylglycerol in groups fed diet containing onion extract were significantly decreased compared with those of controls (p<0.01). In conclusion, the onion extracts exerted a growth-promoting effect when added in White mini broiler diets, reflecting potential alternative substances to replace antibiotics. (Key Words: Onion Extract, Growth Performance, Serum Cholesterol, Blood Profiles, White Mini Broilers)

INTRODUCTION

The sub-therapeutic uses of antibiotics to enhance growth and prevent the infectious intestinal diseases have led to a problem of drug residues in final animal products and emerge of new antibiotic-resistance bacteria (Frankic et al., 2009). In Korea, the routine use of antibiotics in poultry diets have been banned and thus, some endeavors are made to develop new in-feed antibiotics substitues for reducing and treating infectious diseases in poultry industry. The herb and botanicals are increasingly being used in animal feeds, in place of antibiotics, as possible alternative means to prevent infectious diseases and modulate the immune responses (Wenk, 2003).

Onion (Allium cepa L.) which belongs to the family Liliaceae is extensively used as food and the common medicinal plants. Onion bulbs have numerous organic sulphur compounds, flavonoids and phenolic acids with proven antibacterial, antioxidant and hypolipidemic efficacy (Melvin Joe et al., 2009; Srinivasan et al., 2004). The serum cholesterol was significantly decreased by dietary dehydrated onion in experimentally hypercholesterolemic rats (Vidyavati et al., 2010). Goodarzi et al. (2013) reported that the beneficial influence of onion extract on the growth performance in meat-type broiler chickens.

The White mini broiler (WMB) is a local mixed breed produced by crossbreeding between meat-type male breeder and egg-type hens. The meat from WMB has unique texture with not too tender and is mainly used as raw meat for retort products, called Samgyetang. The production of WMB has been continuously increased, still having a small market, because consumer's preference for Samgyetang is growing in this region. The WMB show relatively slow-growth with poor feed efficiency than fast-growing broilers. They were capable of a feed efficiency of 50 g of body weight (BW) gain per 100 g feed consumption with about 900 g BW at 31-d old (Choo et al., 2014). For maximal growth and optimal feed conversion ratio, the commercial...
broiler diets with antibiotic-based growth promoters has been fed to WMB until the use of antibiotics was prohibited. The objectives of this study were to determine the dietary effects of onion extracts as in-feed antibiotics substitutions on growth performance, carcass characteristics and blood lipid profiles of WMB.

MATERIALS AND METHODS

Diets, animals and management

Fresh onion bulbs (Allium cepa L.) were purchased from local market (Emart Inc., Seoul, Korea). They were peeled and grated by mixer (HR 2160, Arteal Manufacturing Ltd., Huiyang, China) without water addition. Leuconostoc citreum SK1984 was inoculated at final level of 1% into onion juice containing medium and incubated at 30°C for 24 h. The molasses of 6.5 g/L and yeast extract of 10.0 g/L were provided as media ingredients as previously suggested (Chang et al., 2010). The obtained onion extract was sprayed and well mixed with non-medicated control diet.

On the day of hatch, White mini broiler chicks were received from a local hatchery (Fineth hatchery Co. Ltd., Chungju, Korea). They were weighed individually and randomly assigned into four experimental treatments with five replicates of 30 chicks. A total of 600 chicks were housed on rice husks in floor pens with 23/1 light/dark cycle throughout the experimental period and fed one of four diets (antibiotic medicated, non-medicated, non-medicating diets with 0.3% or 0.5% onion extract) for 35 d of rearing period. The non-medicating diet were purchased from a commercial feed manufacture (Seoulfeed Co. Ltd., Inchon, Korea) (Table 1), and mixed with avilamycin (10 ppm) or different doses of onion extract. The chicks were allowed to have free access to diet and water. The chicks were initially reared at 33°C; the room temperature was gradually decreased by 4°C weekly until a final temperature of 23°C was reached. The experimental diets were freshly added everyday and feed intake of each replicate was recorded weekly. The BW on a pen basis was recorded weekly and feed efficiency was also calculated. All animal care procedures were approved by Institutional Animal Care and Use Committee in Konkak University.

Sample preparations and measurements

At the end of the experimental period, 10 chicks per each treatment were selected and weighed individually. The blood was taken from jugular veins and analyzed for the activities of glutamic-oxaloacetic transaminase (GOT) and glutamic-pyruvic transaminase (GPT). The activities of GOT and GPT in serum were measured according to the colorimetric method as previous described (An et al., 2007). The liver, spleen, bursa of Fabricius, abdominal fat, breast and leg were immediately removed and weighed. The data were expressed as grams of organ per 100 g BW. The chicks selected were sacrificed by neck cut, scalded with hot water (55°C for 2 min), and removed the feathers mechanically. Carcasses were eviscerated manually and portioned into commercial cuts such as neck, breast, leg and wing. The breasts were immediately chilled for 30 min in ice water and transferred to icebox and then prepared separately for further analyses.

The contents of each lipid fraction in the liver and serum were separated by thin layer chromatography (TLC) on silica gel chromarods (Chromarod-S III, Mitsubishi Kagaku Iatron, Inc., Tokyo, Japan), an exclusive media in the same manner of normal phase TLC, using hexane:diethylether:formic acid (85:15:0.15, v/v) as development solvents, and quantified by IATRO SCAN (TH-10 TLC/FID analyzer, Iatron, Ltd., Tokyo, Japan) as previously described (An et al., 1997).

The instrumental color of fresh breast meat, including lightness (L*), redness (a*) and yellowness (b*), were measured using a chromameter (CR 210, Minolta, Tokyo, Japan). The pH values of breast meats were estimated in

Table 1. Formula and chemical compositions of the experimental diet

| Items | Ingredients, % |
|-------|----------------|
|       | Yellow corn    | 47.29 |
|       | Wheat          | 8.00  |
|       | Soybean meal   | 31.60 |
|       | Corn gluten meal | 3.87 |
|       | Tallow         | 5.5   |
|       | Vit. and Min. mixture | 0.20 |
|       | DL-methionine (98.5%) | 0.11 |
|       | Dicalcium phosphate | 1.90 |
|       | Limestone      | 1.13  |
|       | Choline-Cl (50%) | 0.08 |
|       | Salt           | 0.32  |
|       | Total          | 100.00|

| Calculated composition | TMEn (kcal/kg) | 3.150 |
|                       | CP (%)         | 21.50 |
|                       | Ca (%)         | 1.00  |
|                       | Available P (%) | 0.45 |
|                       | Lysine (%)     | 1.10  |
|                       | Methionine+cystine (%) | 0.85 |

1TMEn, nitrogen-corrected true metabolizable energy; CP, crude protein.

Vit. and Min. mixture provided the following nutrients per kg of diet: vitamin A, 40,000 IU; vitamin D₃, 8,000 IU; vitamin E, 10 IU; vitamin K₃, 4.0 mg; vitamin B₁, 4.0 mg; vitamin B₂, 12.0 mg; vitamin B₆, 6.0 mg; vitamin B₁₂, 0.02 mg; niacin, 60.0 mg; pantothenic acid, 20 mg; folic acid, 2.0 mg; biotin, 0.02 mg; Fe, 30.0 mg; Zn, 25.0 mg; Mn, 20.0 mg; Cu, 5.0 mg; Se, 0.1 mg.
Table 2. Dietary effects of onion extract on growth performance and carcass yield of White mini broilers\(^1\)

| Items                      | (-)Control | (+)Control | Onion extract 0.3% | SEM | p-value |
|----------------------------|------------|------------|---------------------|-----|---------|
| Initial BW (g/bird)        | 39.1       | 39.1       | 39.1                | 0.3 | NS      |
| Final BW (g/bird)          | 862.3\(^a\) | 890.6\(^b\) | 882.0\(^m\)        | 3.78| 0.01    |
| Feed intake (g/d/bird)     | 49.8       | 49.6       | 50.4                | 0.51| NS      |
| BW gain (g/d/bird)         | 24.2\(^a\) | 25.1\(^a\) | 24.8\(^m\)         | 0.13| 0.01    |
| Gain:feed                  | 0.49       | 0.51       | 0.49                | 0.02| NS      |
| Carcass yield (%)          | 66.2       | 67.32      | 66.3                | 0.47| NS      |

% for carcass

Right breast muscle | 9.7       | 9.7       | 9.6                | 0.15| NS      |
Right leg           | 7.6       | 8.1       | 7.6                | 0.15| NS      |
Right thigh         | 8.0       | 8.2       | 8.4                | 0.16| NS      |

SEM, standard error of the mean; BW, body weight; NS, not significant.
\(^1\) (-)Control, basal diet; (+)Control, basal diet+10 ppm avilamycin.
\(^a,b\) Means in the same row with no common superscript differ significantly (p<0.05).

triplicate with a pH meter (340 Mettler-Toledo, Greifensee, Switzerland). Briefly, 1 g of breast meat was cut into small pieces and homogenized with 9 mL of distilled water for 1 min in an Ultra-Turrax (Model No. T25, Janken and Kunkel, Staufen, Germany). To determine the cooking loss, 60 g of breast meat was boiled individually in polyethylene bags immersed in 80°C water bath for 30 min and cooled at room temperature for 30 min. The cooking loss was calculated from the weights of uncooked and cooked meats. The shear force values were determined with a Warner-Bratzler shear attachment on a texture analyzer (Instron Corporation, Canton, MA, USA) with the following operating conditions: load cell 50 kg and cross-head speed, 180 mm/min. Each core sample was sheared once across the center of the core, perpendicular to the muscle fiber. The values were the mean of the maximum forces required to shear each set of core samples.

Statistical analysis

Each pen was considered as the experimental unit for performance parameter. The experimental unit was put on each bird for other measurements. All data were statistically analyzed using analysis of variance (SAS Institute, 2002), and significant differences of obtained means were determined using Duncan’s multiple range tests at the level of p<0.05 (Duncan, 1955).

Table 3. Dietary effects of onion extract on carcass characteristics of White mini broilers\(^1\)

| Items                     | (-)Control | (+)Control | Onion extract 0.3% | SEM | p-value |
|---------------------------|------------|------------|---------------------|-----|---------|
| Liver (g/100 g BW)        | 2.24       | 2.32       | 2.08                | 0.06| NS      |
| Spleen (g/100 g BW)       | 0.12       | 0.16       | 0.15                | 0.01| NS      |
| Bursa of Fabricius (g/100 g) | 0.29     | 0.32       | 0.29                | 0.02| NS      |
| Abdominal fat (g/100 g)   | 1.41       | 1.53       | 1.39                | 0.09| NS      |

SEM, standard error of the mean; BW, body weight; NS, not significant.
\(^1\) (-)Control, basal diet; (+)Control, basal diet+10 ppm avilamycin.

RESULTS AND DISCUSSION

Growth performance and carcass characteristics of White mini broilers fed the experimental diets are presented (Table 2 and 3). The final BW and daily BW gain in the groups fed a non-medicated control diet were significantly lower than those of medicated control group (p<0.01). The growth performance in chicks fed diet with 0.3% or 0.5% onion extract was slightly lower than that of medicated control group, but not significantly. There were no significant difference in feed intake and feed efficiency among groups.

The herbal products and spicy have been used in animal feed as digestion stimulants and growth promoters (Frankic et al., 2009). Jo et al. (2009) reported that the favorable influence of garlic extract on feed efficiency of broiler chickens. The broiler chickens fed diet with 3% onion bulbs had a greater BW gain (Goodarzi et al., 2014). Aji et al. (2011) also reported that the BW gain in chicks fed diets with garlic and onion was higher than control. It has been appeared that inclusion of onion into broiler diets did not have adverse effects on feed intake (Goodarzi et al., 2013). In present study, there was no significant effect on feed intake during starter and grower phases with slight increase in BW gain.

The carcass yield and the relative weights of edible parts were not affected by dietary treatments as shown in

- An et al. (2015) Asian Australas. J. Anim. Sci. 28:247-251
### Table 4. Dietary effects of onion extract on the enzyme activities and lipid fractions of serum and liver of White mini broilers

| Items                               | (-)Control | (+)Control | Onion extract | SEM | p-value |
|--------------------------------------|------------|------------|---------------|-----|---------|
|                                     |            |            | 0.3%          | 0.5%|         |
| Cholesterol ester (mg/g)             | 93.12      | 92.10      | 89.77         | 88.33| 2.50    | NS      |
| Free cholesterol (mg/g)              | 58.34      | 53.38<sup>a</sup> | 49.54<sup>b</sup> | 48.74<sup>b</sup>| 1.67   | 0.01    |
| Triacylglycerol (mg/g)               | 403.32<sup>a</sup> | 347.68<sup>b</sup> | 319.04<sup>bc</sup> | 280.67<sup>c</sup>| 16.09  | 0.01    |
| Phospholipid (mg/g)                  | 425.92     | 419.27     | 399.14        | 387.84| 24.89   | NS      |

Serum lipid fractions
- Cholesterol ester (mg/g)
- Free cholesterol (mg/g)
- Triacylglycerol (mg/g)
- Phospholipid (mg/g)

**NS**: not significant. 
<sup><sup>a</sup>Means in the same row with no common superscript differ significantly (p<0.05).

### Table 5. Dietary effects of onion extract on physicochemical properties of breast muscles of White mini broilers

| Items                  | (-)Control | (+)Control | Onion extract | SEM | p-value |
|------------------------|------------|------------|---------------|-----|---------|
|                        |            |            | 0.3%          | 0.5%|         |
| pH                     | 6.42       | 6.41       | 6.43          | 6.39| 0.06    | NS      |
| Right breast muscle    |            |            |               |     |         |
| L*                     | 48.58      | 48.06      | 47.66         | 49.11| 1.06   | NS      |
| a*                     | 2.44       | 2.37       | 2.46          | 2.43| 0.31    | NS      |
| b*                     | 7.99       | 8.06       | 9.46          | 7.96| 0.55    | NS      |
| Cooking loss (%)       | 22.30      | 21.44      | 20.04         | 21.19| 1.00   | NS      |
| Shear force (kgf)      | 2.63       | 2.54       | 2.89          | 2.94| 0.22    | NS      |

**SEM**: standard error of the mean; **NS**: not significant. 
<sup><sup>a</sup>b</sup>Means in the same row with no common superscript differ significantly (p<0.05).

<sup><sup>1</sup>(-):Control, basal diet; (+):Control, basal diet + 10 ppm avilamycin.</sup>
extract affecting pH, color and cooking loss of breast meats in WMB (Table 5). The shear force values in chicks fed diets containing onion extract tend to be higher as compared with those of medicated or non-medicated controls, but not significantly. The texture of meat as well as the visual appearance are the main attributes that attract consumers to purchase quality poultry meats (Yang and Jiang, 2005). Jang et al. (2011) reported that dietary quercetin and methoxylated quercetin extracted from onion did not influence the pH and meat color in chicken thigh meats after slaughter. It is assumed that the dietary onion extract will not affect the physico-chemical properties and sensory characteristics of broiler meats, while there are scanty studies on the effect of onions on the meat quality.

In conclusion, these results suggested that onion extract may be able to use as substitute for in-feed antibiotics because the growth performance in chicks fed diet with onion extracts was comparable with medicated control. The onion extract could have beneficial effects in reducing the levels of blood free cholesterol and triacylglycerol of White mini broilers. Further research is needed to describe the effects of dietary onion extract on the metabolism and tissue distribution of cholesterol and triacylglycerol in meat-type chickens.

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REFERENCES

Adamu, I., P. K. Joseph, and K. Y. Augusti. 1982. Hypocholesterolemic action of onion and garlic unsaturated oils in sucrose fed rats over two month period. Experientia 38:899-901.

Aji, S. B., K. Ignatius, A. Y. Ado, J. B. Nuhu, A. Abdulkarim, U. Aliyu, M. B. Gambo, M. A. Ibrahim, H. Abubakar, M. M. Bukar, H. M. Imam, and P. T. Numan. 2011. Effect of feeding onion (Allium cepa) and garlic (Allium sativum) on some performance characteristics of broiler chickens. Res. J. Poult. Sci. 4:22-27.

An, B. K., H. J. Im, and C. W. Kang. 2007. Nutritional values of red pepper seed oil meal and effects of its supplementation on performances and physiological responses of broiler chicks. Asian Australas. J. Anim. Sci. 20:971-975.

An, B. K., H. Nishiyama, K. Tanaka, S. Ohtani T. Iwata, K. Tsutsumi, and M. Kasai. 1997. Dietary safflower phospholipid reduces liver lipids in laying hens. Poult. Sci. 76:689-695.

Baksh, R. and M. I. D. Chughtai. 1985. Comparative study of onion and garlic on serum cholesterol, liver cholesterol, prothrombin time and faecal sterol excretion in male albino rats. J. Chem. Soc. (Pakistan) 7:285-288.

Chang, W. K., S. B. Cho, D. W. Kim, S. S. Lee, and S. K. Kim. 2010. Cell growth and antioxidant activity on onion juice fermentation by using Lactobacillus plantarum as animal probiotics. J. Life Sci. 20:1729-1737.

Choo, Y. K., H. J. Kwon, S. T. Oh, J. S. Um, B. G. Kim, C. W. Kang, S. K. Lee, and B. K. An. 2014. Comparison of growth performance, carcass characteristics and meat quality of Korean local chickens and silky fowl. Asian Australas. J. Anim. Sci. 27:398-405.

Diaz, G. J., I. P. Roldan, and A. Cortez. 2003. Intoxication of Crotalaria pallida seeds to growing broiler chicks. Vet. Hum. Toxicol. 45:187-189.

Duncan, D. B. 1955. Multiple range and multiple F test. Biometrics 11:1-42.

El-Demerdash, F. M., M. I. Yousef, and N. I. El-Naga. 2005. Biochemical study on the hypoglycemic effects of onion and garlic in alloxan-induced diabetic rats. Food Chem. Toxicol. 43:57-63.

Frankic, T., M. Voljc, J. Salobir, and V. Rezar. 2009. Use of herbs and spices and their extracts in animal nutrition. Acta Agric. Slovenica 94:95-102.

Goodarzi, M., M. Landy, and S. Nanekarani. 2013. Effect of onion (Allium cepa L.) as an antibiotic growth promoter substitution on performance, immune responses and serum biochemical parameters in broiler chicks. Health 5:1210-1215.

Goodarzi, M., S. Nanekarani, and N. Landy. 2014. Effect of dietary supplementation with onion (Allium cepa L.) on performance, carcass traits and intestinal microflora composition in broiler chickens. Asian Pac. J. Trop. Dis. 4 (Suppl. 1):S297-S301.

Jang, A., J. A. Ham, D. W. Kim, H. S. Chae, D. W. Kim, S. H. Kim, K. H. Seol, M. W. Oh, and D. H. Kim. 2011. Effect of quercetin and methoxylated quercetin on chicken thigh meat quality during cold storage. Korean J. Poult. Sci. 38:265-273.

Jo, J. K., S. Y. Yoon, J. S. Kim, Y. W. Kim, K. Yun, I. K. Kwon, and B. J. Chae. 2009. Effect of garlic extract supplementation on growth performance, nutrient digestibility, carcass characteristics and meat composition in broilers. Korean J. Poult. Sci. 36:287-292.

Lumeij, J. T. 1997. Avian clinical biochemistry. In: Clinical Biochemistry of Domestic Animals (Eds. J. J. Kanebo, J. W. Harvey, and M. L. Bruss). 5th, Academic Press, CA, USA. pp. 857-883.

Melvin Joe, M., J. Jayochitra, and M. Vijayapriya. 2009. Antimicrobial activity of some common spices against certain human pathogens. J. Med. Plants Res. 3:1134-1136.

SAS. 2002. SAS User's guide. Statistics, Version 8. e., SAS Institute Inc., Cary, NC, USA.

Sklan, D., Y. N. Bernera, and H. D. Rabinowitch. 1992. The effect of dietary onion and garlic on hepatic lipid concentrations and activity of antioxidative enzymes in chicks. J. Nutr. Biochem. 3:322-325.

Srinivasan, K., K. Sambaiah, and N. Chandrasekhar. 2004. Spices as beneficial hypolipidemic food adjuncts: A review. Food Rev. Int. 20:187-220.

Vidyavati, H. G., H. Manjunatha, J. Hemavathy, and K. Srinivasan. 2010. Hypolipidemic and antioxidant efficacy of dehydrated onion in experimental rats. J. Food Sci. Technol. 47:55-60.

Wenk, C. 2003. Herbs and botanicals as feed additives in monogastric animals. Asian Australas. J. Anim. Sci. 16:282-289.

Yang, N. and R. S. Jiang. 2005. Recent advances in breeding for quality chickens. World's Poult. Sci. J. 61:373-381.