Immunohistochemistry Study of Tumor Vascular Normalization and Anti-angiogenic Effects of Sunitinib Versus Bevacizumab Prior to Dose Dense Doxorubicin/cyclophosphamide Chemotherapy in HER2 Negative Breast Cancer

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Research Article

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Abstract

Purpose

Tumor angiogenesis controlled predominantly by vascular endothelial growth factor and its receptor (VEGF-VEGFR) interaction plays a key role in the growth and propagation of cancer cells. However, the newly formed network of blood vessels is disorganized and leaky. Pre-treatment with anti-angiogenic agents can "normalize" the tumor vasculature allowing effective intra-tumoral delivery of standard chemotherapy.

Immunohistochemistry (IHC) analysis was applied to investigate and compare the vascular normalization and anti-angiogenic effects of two commonly used anti-angiogenic agents, Sunitinib and Bevacizumab, administered prior to chemotherapy in HER2 negative breast cancer patients.

Methods

This prospective clinical trial enrolled 38 patients into a sunitinib cohort and 24 into a bevacizumab cohort. All received 4 cycles of doxorubicin/cyclophosphamide chemotherapy and pre-treatment with either sunitinib or bevacizumab. Tumor biopsies were obtained at baseline, after cycle 1 (C1) and cycle 4 (C4) of chemotherapy. IHC was performed to assess the tumor vascular normalization index (VNI), lymphatic vessel density (LVD), Ki67 proliferation index and expression of tumor VEGFR2.

Results

In comparison to Bevacizumab, Sunitinib led to a significant increase in VNI post C1 and C4 (p <0.001 and 0.001) along with decrease in LVD post C1 (p= 0.017). Both drugs when combined with chemotherapy resulted in significant decline in tumor proliferation after C1 and C4 (baseline vs post C4 Ki67 index p=0.006 for Sunitinib vs p=0.021 for Bevacizumab). Bevacizumab resulted in a significant decrease in VEGFR2 expression post C1 (p=0.004).

Conclusion

Sunitinib, in comparison to Bevacizumab showed a greater effect on tumor vessel modulation and lymphangiogenesis suggesting that its administration prior to chemotherapy might result in improved drug delivery.

Clinical trial registration

ClinicalTrials.gov: NCT02790580 (first posted June 6, 2016).

Introduction

As per global cancer statistics 2018, breast cancer is the most commonly diagnosed cancer in females, with an incidence rate of 46.2 per 100,000 and representing 24.2% of the total female cancer burden globally. It is also the leading cause of cancer death with a mortality rate of 15% [1]. Multiple studies have highlighted the key role of tumor vascularization in facilitating tumor growth, progression and metastasis of various solid tumors including breast cancer [2, 3]. Bevacizumab is an anti-vascular endothelial growth factor (VEGF) monoclonal antibody while Sunitinib is an orally administered small molecule receptor tyrosine kinase inhibitor that exerts its action by targeting the vascular endothelial growth factor receptors (VEGFR). Both agents have been studied in clinical trials in combination with chemotherapy in breast cancer. Disappointingly, although both anti-angiogenic agents have shown promising preclinical results, their effects in breast cancer when combined with chemotherapy have been conflicting in the clinic. This could be in part attributed to the fact that optimal dosing schedule of these drugs in combination with chemotherapy are yet to be determined [4–6].

A major pathway involved in angiogenesis is the release of VEGF from hypoxic tumor cells and its binding to the VEGFR expressed on the vascular and lymphatic endothelial cells, leading to endothelial cell proliferation and migration. However, the newly formed blood vessels in a growing tumor are dilated, leaky and poorly organised with no pericyte covering. These “immature” vessels have variable blood flow resulting in sub-optimal delivery of chemotherapeutic drugs. The careful and judicious use of anti-angiogenic drugs can “normalize” these abnormally structured blood vessels within the tumor leading to more efficient drug delivery [7]. In addition, VEGFR2 is expressed on various tumors including breast cancer and is responsible for the autocrine and paracrine effect of VEGF resulting in tumor cell survival and proliferation [8].
We hypothesize that pre-treatment rather than concurrent treatment with sunitinib or bevacizumab prior to standard chemotherapy in human epidermal growth factor receptor 2 (HER2) negative breast cancer, along with a lower dose of the anti-angiogenic agent, will improve and “normalize” the tumor vasculature making it more efficient for intra-tumoral chemotherapy drug delivery. We enrolled patients into a prospective clinical trial and obtained serial tumor biopsies at baseline, during and after chemotherapy to assess and compare the vascular normalization and anti-angiogenic effects of sunitinib versus bevacizumab. Vascular normalization, lymphatic density, tumor proliferation index and activated VEGFR2 status of tumor cells were studied in the tumor specimens.

**Patients And Methods**

**Study Population**

Patients were enrolled into a prospective, phase II open label, single arm study conducted at the National University Cancer Institute, Singapore (NCIS). Eligibility criteria included female patients aged ≥ 18 years with newly diagnosed and histologically confirmed HER2 negative breast cancer. HER2 negativity was defined as HER2 score 0 or 1+ on immunohistochemistry (IHC) or HER2 IHC 2+ but HER2 Fluorescence In Situ Hybridization (FISH) negative (HER2/CEP17 ratio <2.0 with gene copy number < 4.0 signals/ cell) [9]. Other inclusion criteria were measurable primary tumor ≥2cm, Eastern Cooperative Oncology Group (ECOG) performance 0 or 1, absolute neutrophilic count ≥ 1.5 x 10⁹ /L, platelets ≥ 100 x 10⁹ /L, serum total bilirubin ≤ 1.5 x upper limit of normal (ULN), alanine and aspartate aminotransferase ≤ 2.5 x ULN, serum creatinine ≤ 1.5 x ULN and left ventricular ejection fraction ≥ 50%. Patients with clinically detectable second primary malignancy, symptomatic brain metastasis, and known history of systemic connective tissue diseases were excluded from the study. Signed written informed consent was taken from all patients before enrolment. The clinical trial was conducted in accordance with local regulatory requirements and approved by the institutional ethics review board.

**Treatment plan and study design**

The study aimed to determine the effect of pre-treatment with low-dose anti-angiogenic agent prior to chemotherapy as a strategy to normalize tumor vasculature. Patients were enrolled into two sequential Cohorts to study two different classes of anti-angiogenic agents: Sunitinib Cohort to evaluate a small molecule tyrosine kinase inhibitor against VEGFR, and Bevacizumab Cohort to evaluate a monoclonal antibody against VEGF. All subjects received 4 cycles of dose-dense doxorubicin/cyclophosphamide (ddAC) chemotherapy every 2 weeks (doxorubicin 60mg/m² and cyclophosphamide 600mg/m²) supported by prophylactic peglgrastim 6mg administered subcutaneously 24-48 hours after each cycle of chemotherapy. Patients enrolled into the Sunitinib Cohort were pre-treated with oral sunitinib 12.5mg daily for 7 days prior to cycle 1 ddAC and for 5 days prior to cycles 2, 3 and 4 ddAC. Patients enrolled into the Bevacizumab Cohort received intravenous bevacizumab 5mg/kg, 7 days prior to each chemotherapy cycle.

For each patient, detailed history was recorded. Physical examination, radiological staging with computed tomography (CT) scan or CT-PET scan and laboratory evaluation was done prior to initiating treatment. Patients were evaluated before each new cycle of chemotherapy to monitor adverse effects from treatment and to measure tumor response. Response to treatment was assessed clinically according to the Response Evaluation Criteria in Solid Tumors (RECIST) version 1.1 criteria [10].

**Tumor Core Biopsies for Immunohistochemistry (IHC) Studies**

Tumor core biopsies were obtained from the primary breast tumor under ultrasound guidance at baseline, 2 weeks after cycle 1 ddAC but before cycle 2 ddAC (post-C1) and about 2 weeks after completion of 4 cycles of ddAC (post-C4). Tumor cores were fixed in formalin for further histological and IHC analysis.

**Immunohistochemistry Studies**

Tumor biopsies taken at each time point were processed into paraffin blocks. Hematoxylin and Eosin (H&E) staining was done to identify and assess the tumor content. Biopsies which showed tumor content ≤10% were excluded from further evaluation and staining. IHC was performed on consecutive slides using the Leica Bond Max automated platform (Leica Biosystems, Nussloch GmbH) with bond polymer refine detection kit and bond polymer refine red detection kit (DS9800 and DS9390 respectively, Leica Biosystems). Briefly, 4-micron sections from tissue blocks were taken on coated slides. These were then deparaffinized, hydrated and blocked with hydrogen peroxide. Heat induced antigen retrieval was achieved using appropriate buffer for each antibody as per optimised protocol in control tissue. Slides were incubated with primary antibody followed by secondary antibody. Staining was completed with diaminobenzidine (DAB) chromogen and haematoxylin was used as a counterstain.
To assess the tumor vascular normalization index (VNI), double sequential staining for endothelial cells and pericytes was performed using CD31 and alpha-smooth muscle actin (α-SMA) respectively. This was followed by visualization with alkaline phosphatase-based red (DS9390) and peroxidase-based diaminobenzidine polymer detection systems (DS9800), respectively. VNI was calculated as the percentage of CD31 positive cells which co-express α-SMA in relation to the total number of blood vessels in the entire biopsy. This index was used as an indicator of tumor vessel maturation [11]. D2-40 antibody was selected for marking the lymphatic vessels and analysing the lymphatic vessel density (average number of vessels positive for D2-40 in the entire biopsy). To study the pharmacodynamic effect of sunitinib and bevacizumab on tumor cells, the expression of activated VEGFR2 (phosphorylated VEGFR-2 at tyrosine phosphorylation sites 951 and 996 [Y951 and Y996]) on tumor cells was examined. VEGFR2 expression was semi-quantified by H score, which is the percentage of tumor cells staining positive multiplied by an intensity score (0: no staining, 1: weak staining, 2: moderate staining, 3: strong staining). The final score ranged from 0-300 [12]. Ki67 proliferation index of tumor was calculated as number of Ki67 positive tumor cells per 100 tumor cells. Details on the antibody clone, commercial supplier, dilution and antigen retrieval are provided in Table 1.

**Evaluation of Histological Response on Surgical Specimens**

After completing 4 cycles of ddAC chemotherapy, patients with non-metastatic cancer underwent lumpectomy or mastectomy and sentinel lymph node biopsy or axillary lymph node clearance. Scoring of histological response on the primary tumor was done using the 5-point scale Miller-Payne grading (MPG) classification which was based on comparing the tumor cellularity between baseline and post C4 biopsy. Good histological response was defined as a score of ≥3, i.e., more than 30% reduction of tumor cellularity in post treatment biopsies from baseline [13].

**Statistical analysis**

All statistical analyses were performed using SPSS software version 20.0 (IBM, Armonk, NY, USA). Categorical variables were presented as frequencies and percentages, while continuous variables as mean with standard deviation (SD). Comparison of continuous data between groups was done using the non-parametric Wilcoxon signed rank test. Categorical data was compared using Chi-square test. Independent t-test was run for comparison of means. A p value of less than 0.05 was considered statistically significant.

**Results**

A total of sixty-two subjects were recruited; 38 patients were enrolled into the Sunitinib Cohort and 24 into the Bevacizumab Cohort. The baseline demographic and tumor characteristics of the patients are summarized in Table 2. Median age of the entire cohort was 51 years (range 29-70). Majority of the patients were Chinese, had invasive ductal carcinoma, hormone receptor positive cancer, and non-metastatic disease. There were no significant differences in demographic or baseline tumor characteristics between the Sunitinib and Bevacizumab Cohorts.

**Immunohistochemistry Studies**

Out of the total 62 patients, a full set of pre-treatment, post-C1 and post-C4 tumor samples were available for IHC analysis for 29 of the 38 patients enrolled into the sunitinib cohort, and 15 of the 24 patients enrolled into the Bevacizumab cohort. An additional 5 patients enrolled into the bevacizumab cohort had pre-treatment and post-C1 samples without a post-C4 tumor sample. Reasons for incomplete tumor specimens for IHC analysis include no biopsy because of complete clinical response, tumor specimen too small, or insufficient tumor content for IHC analysis (Figure 1).

**Changes in lymphatic vessel density, vascular normalization index, Ki67 and p-VEGFR2 induced by Sunitinib versus Bevacizumab**

In the Sunitinib Cohort, there was a significant decrease in tumor lymphatic vessel density (LVD) after cycle 1 that persisted after cycle 4 chemotherapy compared to baseline (mean LVD 0.94±1.39, 0.29±0.45, 0.36±0.58 for baseline, post-C1 and post-C4; p=0.017 for baseline vs post-C1, p=0.112 for baseline vs post-C4) (Figures 2A&B). In contrast, there was a numerical increase in LVD that was not statistically significant after cycle 1 or cycle 4 chemotherapy compared to baseline observed in the Bevacizumab Cohort (Table & Figure 3).

Similarly, significant increase in Vascular normalization Index (VNI) was observed in the Sunitinib Cohort after one cycle of chemotherapy and that persisted after four cycles of chemotherapy, compared to baseline (mean VNI 51.00±21.97%, 74.91±18.93%,...
75.54±21.23% for baseline, post-C1 and post-C4; p <0.001 for baseline vs post-C1, p=0.001 for baseline vs post-C4) (Figures 2C&D). While a similar trend was observed in the Bevacizumab Cohort, the differences were not statistically significant (Table & Figure 3).

In both the Sunitinib and Bevacizumab Cohorts, there was a significant decline in Ki67 proliferation index after cycle 1 and cycle 4 chemotherapy, although the decline appears quicker and more marked in the Sunitinib Cohort (mean Ki67 in Sunitinib Cohort 14.79±23.81%, 5.52±10.39, 2.00±7.54% for baseline, post-C1 and post-C4; p=0.027 for baseline vs post-C1, p=0.006 for baseline vs post-C4; mean Ki67 in Bevacizumab Cohort 30.85±33.16, 14.55±22.96, 4.80±8.26 for baseline, post-C1 and post-C4; p=0.005 for baseline vs post-C1, p=0.021 for baseline vs post-C4) (Figure 2E&F) (Table & Figure 3).

Expression of p-VEGFR2 on tumor cells was significantly reduced after cycle one chemotherapy in the Bevacizumab Cohort for both Y951 and Y996, although there appears to be a rebound in p-VEGFR2 after cycle 4 chemotherapy (mean H score [Y951] 55.25±41.15, 32.00±38.60, 50.67±41.82 for baseline, post-C1 and post-C4; p=0.008 for baseline vs post-C1, p=0.819 for baseline vs post-C4; mean H score [Y996] 27.50±38.54, 4.75±9.79, 7.33±10.83 for baseline, post-C1 and post-C4; p=0.004 for baseline vs post-C1, p=0.124 for baseline vs post-C4) (Figure 2G&H). A similar trend was noted in the Sunitinib Cohort but the difference was not statistically significant (Table & Figure 3).

Clinical and pathological outcomes and correlation with IHC parameters

After 4 cycles of ddAC chemotherapy, 2.7% and 8.3% of patients in the Sunitinib and Bevacizumab Cohorts respectively achieved complete clinical response; 73% and 75% respectively achieved clinical partial response, while 24.3% and 16.7% achieved stable disease. No patient in either Cohort had clinical progressive disease. Patients with a complete or partial response were considered as good clinical responders while those with only stable disease as poor clinical responders. 33 patients in the Sunitinib Cohort and 17 patients in the Bevacizumab Cohort underwent surgery. No patient achieved pathological complete response. Using the MPG system classification, 66.7% and 76.5% in the Sunitinib and Bevacizumab Cohorts respectively achieved good histological response after 4 cycles of ddAC. No significant differences in clinical or histological responses were observed between the Sunitinib and Bevacizumab Cohorts (percentage good clinical responders in Sunitinib vs Bevacizumab Cohorts 75.7% vs 83.3%, p=0.57; percentage good histological responders in Sunitinib vs Bevacizumab Cohorts 66.7% vs 76.5%, p=0.47). There was a statistically significant difference in the post-C4 Ki67 index between clinical good vs poor responders in both cohorts (Sunitinib cohort, mean Ki67 index 0.74±2.07% vs 8.20±17.78% for good vs poor responders; p=0.04; Bevacizumab cohort, mean Ki67 index 3.15±4.75% vs 15.50±20.50% for good vs poor responders; p=0.044). However, there was no significant difference in the other IHC parameters (VNI, LVD, p-VEGFR2 H score) between good and poor clinical and histological responders in both the cohorts (Table 4 and 5).

Discussion

Ever since the proposal of the “vascular normalization” theory, many preclinical observations show that the addition of anti-angiogenic agents to systemic chemotherapy leads to improved and uniform drug delivery and tumor control. The concept of presence of “normalization window” has provided insight into the probable benefit of administering low dose and short course anti-angiogenic drugs with chemotherapy instead of high and prolonged dosing [14, 15]. However, limited clinical data is available to prove the same. In this study we performed an immunohistochemistry analysis to investigate and compare the effects of two anti-angiogenic drugs, sunitinib and bevacizumab, that are commonly used in clinical practice. We examined vascular normalization, lymphatic vessel density, tumor proliferation index and activated VEGFR2 status of tumor cells in both treatment groups.

We demonstrated that in HER2 negative breast cancers, pre-treatment with low dose short course sunitinib leads to statistically significant increase in vascular normalization index in comparison to bevacizumab. Though both sunitinib and bevacizumab are capable of establishing a more mature vascular network in tumor microenvironment, sunitinib showed more promising results. Sunitinib led to almost 45% increase in VNI after one cycle of treatment compared to baseline although further increase from cycle 1 to cycle 4 was minor. On the other hand, pre-treatment with bevacizumab resulted in only ~10-20% increase in VNI post cycle 1 and 4 that was not statistically significant. The lymphatic vasculature also plays an important role in tumor cell progression and metastasis. Invasion in lymphatic vessels has been found to be associated with increased risk of lymph node and distant metastasis thereby leading to poor survival in breast cancer patients [16]. In our study, sunitinib appeared to inhibit lymphangiogenesis leading to significant decline in LVD after 1 cycle of treatment. In contrast, bevacizumab pre-treatment actually led to a numerical increase in LVD, albeit not statistically significant.
We previously conducted a phase Ib/II trial in which subjects were randomized to chemotherapy with or without low dose, intermittent sunitinib. IHC evaluation on serial tumor biopsies showed evidence of increased VNI and decrease in LVD after chemotherapy in patients randomized to receive sunitinib, but not in those treated with chemotherapy alone [17]. The observations in this current study in the Sunitinib Cohort are concordant with our previous findings [17]. Somewhat surprisingly, these results were not replicated in the Bevacizumab Cohort in our current study. In order to tilt drug effects towards more vascularization normalization than anti-angiogenic, we used a sunitinib dose that was one-third full dose and administered it for only 5-7 days prior to each 2-weekly cycle of chemotherapy instead of continuously. For bevacizumab, we used half dose (5mg/kg every 2 weeks rather than 10mg/kg) and administered it 1 week before chemotherapy rather than concurrently with chemotherapy. We postulate that the bevacizumab dose administered in our trial may still be too high, thus resulting in less prominent vascularization normalization effects than sunitinib.

The slight increase in LVD observed after bevacizumab treatment may be that bevacizumab largely sequesters VEGF-A, thereby blocking VEGF-A/VEGFR-2 signaling. This may result in a compensatory increase in other VEGF ligands like VEGF-C by tumor cells which then bind to VEGFR-3 on lymphatic endothelial cells leading to lymphangiogenesis [18]. On the other hand, the prominent effects on VNI and LVD seen with sunitinib can be on account of its action on multiple tyrosine kinase receptors. Apart from VEGFR, inhibition of other signaling pathways like platelet-derived growth factor receptor (PDGFR), stem cell factor receptor (KIT), FMS-like tyrosine kinase 3 (FLT3), colony-stimulating factor 1 receptor (CSF-1R), rearranged during transfection (RET), may have resulted in the supplementary effect on tumor proliferation, angiogenesis and lymphangiogenesis [19, 20].

Breast tumors are known to produce VEGF and also express VEGFR2 on their surface. This autocrine signaling is responsible for tumor cell growth and division. VEGFR2 signaling can be inhibited by directly blocking the receptor or by interfering with the binding to its ligand VEGF. This is an anti-angiogenic effect and can be affected by both sunitinib, which blocks tyrosine phosphorylation of VEGFR, and bevacizumab which binds to and neutralizes VEGF. In a study on mouse mammary tumor model, sunitinib-treated mice showed decreased levels of tumor p-VEGFR-2 [21]. We similarly observed decreased expression of tumor cell VEGFR2 after one cycle of chemotherapy in patients pretreated with sunitinib as well as bevacizumab proving that both drugs exert anti-angiogenic effects through inactivation of the VEGFR2 receptor on tumor cells. Intriguingly, p-VEGFR-2 expression rebounded after 4 cycles of chemotherapy indicating that the action of anti-angiogenic agents in inhibiting the receptor activation on tumor cells could be of limited duration. Indeed, in an earlier phase Ib trial in breast cancer, we had observed sunitinib-induced normalization of tumor vasculature to occur as early as 24 hours; yet in another phase II randomized trial, intermittent, low dose sunitinib combined with up to 6 cycles of docetaxel did not improve response rates compared to docetaxel alone [17]. We hypothesize that while initial treatment with sunitinib does normalize tumor vasculature, repeated administration may conversely compromise normal tumor vasculature and eventually impair chemotherapy delivery. In fact, it may be possible that just a single cycle or two of sunitinib prior to starting chemotherapy may be sufficient to normalise tumour vasculature [22]. On the other hand, the group that received bevacizumab showed a significant decrease in VEGFR2 expression on tumor cells in comparison to sunitinib. It is possible that the dose of the drugs could affect this autocrine loop signaling of tumor cells; a lower than clinically approved dose of sunitinib was used in this trial, while the bevacizumab dose administered was within the clinically approved range, with the latter thus exerting greater anti-angiogenic than vasculature normalization effects. Also, it is possible that the primary action of sunitinib may have been on endothelial cells rather than tumor cells. Similar results have been reported in a study by Wedam et al, where bevacizumab was administered to locally advanced breast cancer patients (n=21) and a significant inhibitory effect on tumor cell VEGFR2 expression (in both phosphorylation sites-Y951 and Y996) was demonstrated by IHC [23]. Collectively, these findings of decreased VEGFR2 expression together with lowered proliferation index suggests that both anti-angiogenic agents cause inactivation of VEGFR2 on tumor cells thus decreasing tumor cell proliferation although bevacizumab appears to exert a stronger effect than sunitinib on tumor cell VEGFR2 at the doses administered in this trial.

Conclusion

Immunohistochemistry analysis of serial tumor biopsies from patients with HER2 negative breast cancer who received lower dose sunitinib or bevacizumab before standard chemotherapy showed modulation of vessel morphology in tumor tissue along with suppression of tumor cell proliferation. Changes in LVD, VNI, Ki67 and p-VEGFR2, were generally early and observed after one cycle of treatment but tended to plateau with additional cycles of treatments. At the doses and schedule studied in this trial, sunitinib induced tumor vasculature normalization and inhibited lymphangiogenesis more prominently than bevacizumab, while bevacizumab demonstrated more significant effects on tumor VEGFR2 than sunitinib suggesting greater anti-angiogenic activity. Sunitinib with its more prominent vasculature normalization effects led to greater and sustained decline in tumor Ki67 than bevacizumab in this study, highlighting the promise of normalizing tumor vasculature to optimize chemotherapy delivery in breast cancer. The observation that
vasculature normalization and anti-angiogenic effects plateaued or even rebounded with additional treatment cycles suggest that perhaps restricting the use of an anti-angiogenic agent to just the first one to two cycles of chemotherapy could be sufficient to exert the desired effects without the need to combine with all chemotherapy cycles. This strategy of more judicious combination of an anti-angiogenic agent with chemotherapy warrants further investigations.

Declarations

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Conflicts of interest: SGWO served on advisory boards and had speaker’s engagement with Pfizer, Novartis, Eli Lilly, AstraZeneca and Roche, and has received support to attend conference/travel from Pfizer, Novartis and AstraZeneca. AW has served on advisory board of Pfizer and Astra Zeneca. NN has received honoraria from Astra Zeneca, Janssen, Thermofisher and support to attend conferences from Astra Zeneca, Eisai. WQC has received conference sponsorship by MSD (ASCO 2021). GBC has been on advisory boards of MSD, AstraZeneca and Novartis and served as consultant to Adagene. He has received research support from BMS, MSD, Adagene and Taiho and has stock ownership of Merus and Gilead Sciences. SCL served on advisory boards and had speaker’s engagement with Pfizer, Novartis, Eli Lilly, Astra Zeneca, Roche, and ACT genomics, received research grants/ grants to support clinical trials from Eisai, Taiho, Pfizer, and Karyopharm, and has received support to attend conference/travel from Amgen, Pfizer, Novartis, Roche and ACT Genomics.

Data Availability: All data generated or analyzed during this current study are included in this published article.

Author contributions: All authors contributed to at least one of the following: study conception, design, data acquisition, analysis, and/or interpretation. The first draft of the manuscript was written by KY and supervision/critical revision of the work was done by SCL. All authors have read and approved the final manuscript.

Ethics approval: This study was performed in line with the principles of the Declaration of Helsinki. Approval was granted by the National Healthcare Group Domain Specific Ethics Review Board in 2016 (DSRB 2016/00327).

Consent to participate: Signed written informed consent was obtained from all individual participants included in the study.

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Tables

Table 1: Details of antibodies used in immunohistochemistry studies
| Antibody          | Clone    | Manufacturer | Dilution | Antigen retrieval     |
|-------------------|----------|--------------|----------|-----------------------|
| CD31              | JC70A    | Dako         | 1:100    | Citrate buffer, pH6, 20 min |
| α-SMA             | 1A4      | Dako         | 1:500    | Citrate buffer, pH6, 20 min |
| D2-40/Podoplanin  | D2-40    | Dako         | 1:100    | Citrate buffer, pH6, 20 min |
| p-VEGFR2 951      | Rabbit Polyclonal | Invitrogen | 1:50    | Citrate buffer, pH6, 20 min |
| p-VEGFR2 996      | Rabbit Polyclonal | Invitrogen | 1:100   | EDTA buffer, pH9, 20 min   |
| Ki67              | MIB-1    | Dako         | 1:100    | EDTA buffer, pH9, 20 min   |

Table 2: Baseline clinical and pathological characteristics of patients in the two treatment groups
| Characteristics/Variables          | Sunitinib (n=38) | Bevacizumab (n=24) | p value |
|-----------------------------------|-----------------|--------------------|---------|
| **Age (years)**                   |                 |                    |         |
| Median                            | 53.5            | 49.5               | 0.406   |
| Range                             | 30-69           | 29-70              |         |
| **Race**                          |                 |                    |         |
| Chinese                           | 24 (63.1)       | 16 (66.7)          | 0.679   |
| Malay                             | 5 (13.2)        | 5 (20.8)           |         |
| Indian                            | 3 (7.9)         | 1 (4.2)            |         |
| Others                            | 6 (15.8)        | 2 (8.3)            |         |
| **Histological type of tumor**    |                 |                    |         |
| Ductal                            | 33 (86.8)       | 20 (83.3)          | 0.887   |
| Lobular                           | 3 (7.9)         | 2 (8.3)            |         |
| Others                            | 2 (5.3)         | 2 (8.3)            |         |
| **Histological grade of tumor**   |                 |                    |         |
| Grade 1 (Well differentiated)     | 2 (5.3)         | 0                  | 0.183   |
| Grade 2 (Moderately differentiated)| 10 (26.3)      | 11 (45.8)          |         |
| Grade 3 (poorly differentiated)   | 26 (68.4)       | 13 (54.2)          |         |
| **Hormone receptor status**       |                 |                    |         |
| ER and/or PR Positive             | 29 (76.3)       | 18 (75)            | 0.906   |
| ER/PR Negative                    | 9 (23.7)        | 6 (25)             |         |
| **Clinical T stage of primary tumor** |             |                    |         |
| T1                                | 1 (2.6)         | 1 (2.6)            | 0.988   |
| T2                                | 21 (55.3)       | 13 (34.2)          |         |
| T3                                | 10 (26.3)       | 6 (15.8)           |         |
| T4                                | 6 (15.8)        | 4 (10.5)           |         |
| **Clinical node status**          |                 |                    |         |
| N0                                | 10 (26.3)       | 10 (41.6)          | 0.569   |
| N1                                | 22 (57.9)       | 12 (50)            |         |
| N2                                | 2 (5.3)         | 1 (4.2)            |         |
| N3                                | 4 (10.5)        | 1 (4.2)            |         |
| **Metastasis**                    |                 |                    |         |
| Present                           | 5 (13.2)        | 2 (8.3)            | 0.559   |
| Absent                            | 33 (86.8)       | 22 (91.7)          |         |

ER: Estrogen Receptor, PR: Progesterone receptor

**Table 3: Comparison of IHC parameters between the two treatment groups**
Table 4: Correlation of IHC parameters with good vs poor clinical response in the two treatment groups

| IHC Parameters | Sunitinib Mean ± SD | Sunitinib p value | Bevacizumab Mean ± SD | Bevacizumab p value | p value comparison |
|---------------|---------------------|-------------------|-----------------------|---------------------|--------------------|
| LVD (n=29)    | 0.94±               | 0.017             | 0.36±                 | 0.012              | 0.874              |
|               | 1.39                |                   | 0.58                  |                     | 0.88±              |
|               | 0.29±0.45           |                   | 0.112                 |                     | 0.528              |
| VNI (%)       | 51.00±21.97         | <0.001            | 75.54±21.23           | 0.001              | 68.55±25.97        |
|               | 74.91±18.93         |                   | 56.08±20.54           |                     | 61.12±21.23        |
| Ki67 index (%)| 14.79±23.81         | 0.027             | 2.00±7.54             | 0.006              | 4.80±8.26          |
|               | 5.52±10.39          |                   | 30.85±33.16           |                     | 0.021              |
|               | 2.00±7.54           |                   | 14.55±22.96           |                     |                    |
| p-VEGFR2 (Y951) H score | 32.07±40.30 | 0.245             | 26.03±38.54           | 0.651              | 50.67±41.82        |
|               | 23.62±34.25         |                   | 55.25±41.15           |                     | 41.82              |
|               | 32.00±38.60         |                   | 32.00±38.60           |                     |                    |
| p-VEGFR2 (Y996) H score | 7.41±10.05 | 0.351             | 11.90±22.33           | 0.500              | 7.33±10.83         |
|               | 5.17±10.81          |                   | 27.50±38.54           |                     | 0.124              |
|               | 11.90±22.33         |                   | 4.75±9.79             |                     |                    |
|               | 7.33±10.83          |                   |                       |                     |                    |

a: p value comparing the difference in means at post-C1 vs baseline
b: p value comparing the difference in means at post-C4 vs baseline
### Table 5: Correlation of IHC parameters with good vs poor histological response in the two treatment groups

| IHC Parameters at timepoints | Sunitinib (n=28) | Bevacizumab (n=20) |
|-----------------------------|-----------------|-------------------|
| \[\text{Mean} \pm \text{SD}\] | \text{Good responder (n=23)} | \text{Poor responder (n=5)} | \text{p value} | \text{Good responder (n=16)} | \text{Poor responder (n=4)} | \text{p value} |
| Baseline | | | | | | |
| LVD | 0.83±1.29 | 0.80±0.98 | 0.97 | 0.42±0.51 | 0.75±1.36 | 0.43 |
| VNI (%) | 52.88±21.57 | 40.58±25.37 | 0.27 | 53.93±19.72 | 64.65±24.59 | 0.36 |
| Ki67 index (%) | 14.00±23.63 | 21.20±28.19 | 0.55 | 27.31±33.02 | 44.00±38.73 | 0.35 |
| p-VEGFR2 (Y951) H score | 33.04±42.04 | 26.00±39.74 | 0.73 | 53.13±41.42 | 63.75±44.97 | 0.65 |
| p-VEGFR2 (Y996) H score | 8.04±10.84 | 6.00±6.51 | 0.69 | 29.38±48.53 | 20.00±16.33 | 0.67 |
| Post-C1 | | | | | | |
| LVD | 0.34±0.48 | 0.12±0.16 | 0.32 | 0.68±1.11 | 0.05±0.10 | 0.27 |
| VNI (%) | 74.61±20.18 | 71.28±9.75 | 0.72 | 66.92±23.60 | 44.57±34.65 | 0.13 |
| Ki67 index (%) | 4.70±10.13 | 10.40±12.28 | 0.28 | 10.61±19.71 | 30.00±31.62 | 0.13 |
| p-VEGFR2 (Y951) H score | 27.61±36.30 | 10.00±22.30 | 0.31 | 25.00±34.83 | 60.00±45.46 | 0.1 |
| p-VEGFR2 (Y996) H score | 5.65±11.90 | 4.00±5.47 | 0.76 | 5.94±10.68 | 0 | 0.29 |
| Post-C4 (n=15 for bevacizumab) | | | | | | |
| LVD | 0.44±0.61 | 0.08±0.09 | 0.21 | 0.64±0.94 | 2.4±3.39 | 0.1 |
| VNI (%) | 78.84±16.06 | 75.49±15.71 | 0.67 | 70.63±23.92 | 55.06±46.44 | 0.45 |
| Ki67 index (%) | 0.74±2.07 | 8.20±17.78 | \textbf{0.04} | 3.15±4.75 | 15.00±20.50 | \textbf{0.04} |
| p-VEGFR2 (Y951) H score | 27.17±41.22 | 26.00±29.66 | 0.95 | 43.08±39.45 | 100.00±14.14 | 0.71 |
| p-VEGFR2 (Y996) H score | 10.43±19.93 | 21.00±33.98 | 0.35 | 6.92±10.90 | 10.00±14.14 | 0.72 |

\(a: \) Clinical response was not available for one patient of the total 29 patients whose biopsy samples were analyzed at all time points.

\(b: \) In bevacizumab cohort, post-C4 biopsy IHC analysis was possible for 15 of total 20 patients. Of those 15 patients, 13 were good responders and 2 were poor responders.
### IHC Parameters at time points

|                | Sunitinib (n=26) | Bevacizumab (n=16) |
|----------------|------------------|--------------------|
|                | Mean ± SD        | Mean ± SD          |
|                | Good responder   | Poor responder     |          | Good responder | Poor responder |          |
|                | (n=17)           | (n=9)              | p value  | (n=12)         | (n=4)          | p value   |
| Baseline       |                  |                    |          |                |                |          |
| LVD            | 0.88±1.49        | 0.68±0.83          | 0.72     | 0.56±0.86      | 0.20±0.28      | 0.42     |
| VNI            | 56.18±19.49      | 44.25±21.42        | 0.16     | 60.24±22.91    | 48.70±10.29    | 0.35     |
| Ki67 index     | 13.67±21.64      | 21.11±30.49        | 0.47     | 31.75±30.06    | 24.00±44.05    | 0.69     |
| p-VEGFR2 (Y951) H score | 36.47±44.99 | 28.89±37.23        | 0.67     | 57.08±43.82    | 47.5±40.30     | 0.7      |
| p-VEGFR2 (Y996) H score | 8.24±9.00 | 7.78±13.28         | 0.91     | 34.17±46.16    | 12.5±8.66      | 0.37     |
| Post-C1        |                  |                    |          |                |                |          |
| LVD            | 0.41±0.54        | 0.13±0.26          | 0.16     | 0.85±1.25      | 0.10±0.20      | 0.26     |
| VNI            | 75.34±14.94      | 68.91±24.67        | 0.41     | 55.18±31.88    | 74.63±5.07     | 0.25     |
| Ki67 index     | 5.24±9.82        | 7.89±12.94         | 0.56     | 9.00±13.06     | 27.5±34.03     | 0.12     |
| p-VEGFR2 (Y951) H score | 26.18±36.72 | 25.56±35.04        | 0.96     | 31.67±37.61    | 25.00±43.58    | 0.77     |
| p-VEGFR2 (Y996) H score | 7.65±13.36 | 2.22±4.41          | 0.25     | 4.17±9.96      | 7.50±15.50     | 0.61     |
| Post-C4 b      |                  |                    |          |                |                |          |
| (n=11 for bevacizumab) |                    |                    |          |                |                |          |
| LVD            | 0.37±0.64        | 0.40±0.55          | 0.92     | 0.92±1.59      | 0.20±0.34      | 0.47     |
| VNI            | 80.66±12.64      | 74.14±22.05        | 0.34     | 71.13±29.80    | 57.07±30.51    | 0.5      |
| Ki67 index     | 2.71±9.62        | 1.33±3.27          | 0.68     | 7.5±10.35      | 0.67±0.57      | 0.29     |
| p-VEGFR2 (Y951) H score | 32.67±43.70 | 22.22±31.92        | 0.53     | 43.75±48.67    | 56.67±41.63    | 0.69     |
| p-VEGFR2 (Y996) H score | 8.53±15.48 | 22.22±32.70        | 0.15     | 7.50±11.67     | 6.67±11.54     | 0.91     |

a: Histological response could be analyzed in 26 of 29 patients in sunitinib cohort and 16 of 20 patients in bevacizumab cohort due to very little tissue remaining in block after prior sectioning for IHC analysis.

b: In bevacizumab cohort, post-C4 biopsy IHC analysis was possible for 11 of total 16 patients. Of those 11 patients, 8 were good responders and 3 were poor responders.

**Figures**
Figure 1

Reasons for incomplete tumor specimens for IHC analysis include no biopsy because of complete clinical response, tumor specimen too small, or insufficient tumor content for IHC analysis.

Figure 2

D2-40 positive lymphatic vessels
Vascular Normalization
Ki67 positive cells
pVEGFR(951) positive cells
Similarly, significant increase in Vascular normalization Index (VNI) was observed in the Sunitinib Cohort after one cycle of chemotherapy and that persisted after four cycles of chemotherapy, compared to baseline (mean VNI 51.00±21.97%, 74.91±18.93%, 75.54±21.23% for baseline, post-C1 and post-C4; p <0.001 for baseline vs post-C1, p=0.001 for baseline vs post-C4).

Figure 3

A similar trend was noted in the Sunitinib Cohort but the difference was not statistically significant.