Transcriptomic analysis reveals moderate modulation of macrophage migration inhibitory factor superfamily genes in alcohol use disorders

MARIA CRISTINA PETRALIA1, EMANUELA MAZZON2, KATIA MANGANO2, PAOLO FAGONE1, ROBERTO DI MARCO3, LUCA FAZOLONE1, MARIA SOFIA BASILE1, FERDINANDO NICOLETTI1 and EUGENIO CAVALLI2

1Department of Biomedical and Biotechnological Sciences, University of Catania, I-95123 Catania; 2IRCCS (Scientific Institute for Research, Hospitalization and Healthcare) Centro Neurolesi ‘Bonino-Pulejo’, I-98124 Messina; 3Department of Medicine and Health Sciences ‘Vincenzo Tiberio’, University of Molise, I-86100 Campobasso, Italy

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Abstract. Alcohol use disorder (AUD) is a primary, chronic and relapsing disease of brain reward, motivation and memory, which is associated with several comorbidities, including major depression and post-traumatic stress disorder. It has been revealed that Ibudilast (IBUD), a dual inhibitor of phosphodiesterase-4 and -10 and of macrophage migration inhibitory factor (MIF), exerts beneficial effects on AUD in rodent models and human patients. Therefore, IBUD has attracted increasing interest, with research focusing on the elucidation of the pathogenic role of MIF and its homologue, D-dopachrome tautomerase (DDT), in the pathogenesis and maintenance of AUD. By using DNA microarray analysis, the current study performed a transcriptomic expression analysis of MIF, DDT and their co-receptors, including CD74, C-X-C chemokine receptor (CXCR)2, CXCR4 and CXCR7 in patients with AUD. The results revealed that the transcriptomic levels of MIF, DDT and their receptors were superimposable in the prefrontal cortex of rodents and patients with AUD and human patients. Furthermore, peripheral blood cells from heavy drinkers exhibited a moderate increase in MIF and DDT levels, both at the baseline and following exposure to alcohol-associated cues, based on individual situations that included alcohol-related stimuli resulting in subsequent alcohol use (buying alcohol and being at a bar, watching others drink alcohol). Considering the overlapping effects of MIF and DDT, the inverse Fisher’s $\chi^2$ test was performed on unadjusted P-values to evaluate the combined effect of MIF and DDT. The results revealed a significant increase in these cytokines in heavy drinkers compared with controls (moderate drinkers). To the best of our knowledge, the present study demonstrated for the first time that MIF and DDT expression was upregulated in the blood of patients with AUD. These results therefore warrant further study to evaluate the role of MIF and DDT in the development and maintenance of AUD, to evaluate their use as biomarkers to predict the psychotherapeutic and pharmacological response of patients with AUD and for use as therapeutic targets.
Inflammation and altered innate immune responses are hallmarks of alcohol-induced organ damage, affecting the liver, cardiovascular system, and brain, and immune abnormalities induced by alcohol exposure have been well documented both in rodent models and humans with AUD (9). In rodent models, it has been shown that increased systemic and brain production of proinflammatory cytokines such as TNFα, IL-1β and MCP-1 and neuroinflammation occurs when 10-days ethanol administration is followed by endotoxin challenge. Alcohol exposure significantly augments the production of proinflammatory cytokines induced by LPS, with a long-lasting persistence in the brain. Furthermore, the brains of mice exposed to 10-days alcohol administration exhibited signs of microglia activation (10). In agreement with these data, it has been shown that in vivo administration of alcohol to adult male Wistar rats (1.0 g/kg) once daily for 14 days increased the concentrations of the proinflammatory cytokines (IL-1β) and TNF-α in two primary brain regions, such as the hippocampus and frontal cortex, that are implicated in mood regulation (11). In humans, an increase in serum proinflammatory cytokines including TNFα, IL-1β occurs upon prolonged alcohol exposure (12) and the monocytes isolated from the blood of alcoholics produce greater amounts of TNFα, both spontaneously and in response to endotoxin challenge (13). Chronic alcohol consumption elevates the levels of circulating TNFα, IL-1β and IL-6 both in the woman and in their fetuses (14). It is also known that AUD patients exhibit abnormal profile of circulating cytokines, including prototypical Th1, interferon-γ (IFN-γ) (15) and Th-2 cytokines (IL-4, IL-13, IL-10) (16,17), as well as IL-6 and TNF-α, IL-7 and GM-CSF (18). In lung, cytokine production is disrupted by ethanol, exacerbating respiratory distress syndrome with upregulated expression of the Th3 and primarily anti-inflammatory cytokine, transforming growth factor-β (TGF-β) (19,20). In general, AUD patients, and in particular those with MDD, seem to exhibit higher levels of proinflammatory cytokines, IL-6, IL-7, TNF-α, GM-CSF, and IFN-γ, than controls (18,21). One recent study has shown that high-risk drinkers showed significantly higher mean values of the proinflammatory cytokine IL-6 than abstainers (22). TNF-α and IL-6 serum levels have been considered as possible neurobiological markers of alcohol consumption in AUD patients (23).

It has been clarified that chronic ethanol intake stimulates the immune system though activation of the NLRP3/ASC inflammasome. In turn, this leads to activation of caspase-1 and increase of IL-1β in the cerebellum. Accordingly, selective blockade of the IL-1/IL-1R signaling suppresses both the activation of the inflammasome and of neuroinflammation that are provoked by alcohol (24). In addition, the altered immune response evoked from alcohol abuse may in turn be implicated in the generation and maintenance of the mechanisms of alcohol addiction and regulation of drinking. For example, the activation of Toll-like receptor (TLR) 3 increases voluntary alcohol intake in C57BL/6J male (25). The potential clinical relevance of the interaction between the effects of AUD on the immune system and the possible role of this latter in regulating drinking has received further in vivo validation with the observation that TLR3-TIR-domain-containing adapter-inducing interferon-β (TRIF) pathway are activated during chronic ethanol consumption. The IKKe/TBK1 inhibitor Amleroxanx down-regulates TRIF-dependent pathway in the brain and reduces ethanol consumption, suggesting that the TRIF-dependent pathway is implicated in regulation of drinking (26). Hence, the NFkB pathway that lies downstream the activation of TLR signaling may also play a non canonical non-immune role in mediating complex behaviors, including processes of learning and memory, stress responses, anhedonia and drug reward (27). Along this line of research, in a translational study entailing both mice and humans, Coleman and coworkers generated strong proof of evidence in humans that HMGB1 and IL-1β hetero-complexes formed in vivo may contribute to the pathology of alcoholism. The Authors found that HMGB1/IL-1β complexes are increased in post-mortem human alcoholic hippocampus (28). These data have led to the hypothesis of alcohol-induced neuroimmune gene induction (29).

Taken as a whole, all these data strongly support the hypothesis that AUD can be characterized by generalized immune system dysfunction, caused by chronic alcohol exposure and supports the concept that alcoholism may represent a systemic proinflammatory condition (30). These data have attracted attention on the possible interaction between immuno-inflammatory events that are induced by AUD and the possible role of immuno-inflammatory events in the initiation and maintenance of AUD.

Along this line of research, a recent study has shown that the dual inhibitor of phosphodiesterase-4 and -10 and of the pleiotropic cytokine macrophage migration inhibitory factor (MIF), Ibudilast (IBUD), exerts beneficial effects both in animal models of alcohol dependence (31) and in humans with AUD (32). On the basis of these data, we have presently studied eventual differences in transcriptomic levels of MIF and its homologue DDT along with their receptors CD74, CXCR2, CXCR4 and CXCR7 in the brain and peripheral blood of patients with AUD. The transcriptomic analysis of the present study demonstrates a trend to increased expression of MIF, DDT and CD74 in AUD patients. However, when expression levels of MIF and DDT were merged the expression of this superfamily was significantly higher than in healthy controls. These data indicate that dysregulation of MIF family members may occur during AUD and that specific tailored treatments with antagonists of MIF superfamily for AUD is worthy of further investigation.
Materials and methods

Microarray datasets selection and analysis
Analysis of prefrontal cortex transcriptomic data. In order to investigate the expression levels of MIF and related genes in alcohol-related disorder, we interrogated the publicly-available whole-genome transcriptomic microarray datasets, GSE123114, GSE53808 and GSE49376, obtained from the Gene Expression Omnibus (GEO; https://www.ncbi.nlm.nih.gov/geo/) database. GSE123114 included whole genome transcriptomic data from an animal model of chronic alcohol self-administration (33). Briefly, for the generation of this dataset, mice were given free access to three bottles containing 5 and 10% of ethanol and water for 16 weeks. On the last two weeks, both ethanol solutions were adulterated with the averse substance, quinine. Following this procedure, mice were classified into two groups, based on the ethanol intake and preference: ‘Light Drinkers’ (preference for water at all experimental stages); and ‘Inflexible Drinkers’ (preference for ethanol at all experimental stages, even after addition of quinine to the ethanol solution). At the end of the experimental period, total RNA was extracted from the Prefrontal cortex and the Affymetrix Mouse Genome 430 2.0 Array was used for the generation of the dataset. Details on the GSE123114 dataset can be obtained from the relative publication (33).

GSE53808 was used for the analysis of gene expression patterns in the prefrontal cortex of three alcoholics without hepatic encephalopathy and three neurologically normal controls (34). Brain tissues were obtained from the New South Wales Tissue Research Centre (NSW TRC), part-funded by the National Institute on Alcohol Abuse and Alcoholism (R24AA012725) (35). The Affymetrix Human Genome U219 Array was used for the generation of the dataset. For the analysis, technical replicates were averaged. Detailed description of the GSE53808 can be obtained from (35).

The GSE49376 dataset included whole-genome transcriptomic levels of the prefrontal cortex from 46 European Australians, obtained from the NSW TRC. Among the 46 subjects, 23 (16 males and 7 females) had alcohol use disorders, divided in alcohol-dependence (n=9) and alcohol abuse (n=14). The male and female samples did not significantly differ in age, postmortem intervals, brain pH and alcohol daily use. All subjects were not affected by illicit drug abuse and dependence or major psychotic disorders. The clinical information of all the patients are available in (36).

Analysis of blood transcriptomic data. GSE59206 included whole-genome blood transcriptomic data from two groups of subjects (n=11/group): Heavy drinkers (HD, defined as regular alcohol use over the past year of at least 8 standard drinks/week for men and at least 5 standard drinks/week for women), and moderate drinkers (MD, defined as up to 7 standard drinks/week for women and 14 standard drinks/week for men), as defined by the NIAAA. All subjects were between 21-50 years-old, had no history of any drug dependence (including alcohol dependence), and did not meet criteria for Axis I DSM-IV psychiatric diagnoses. Exclusion criteria included dependence on psychoactive substances; current or past use of opiates; current use of psychoactive drugs; neurological disorders; neurological, cardiovascular, endocrine, renal, liver, thyroid diseases; subjects on medications for any medical condition; oral contraceptives; and pregnant and lactating women. Blood was collected at baseline and 1 h following exposure to an alcohol-related cue. Alcohol-related cue scripts were based on individual situations that included alcohol-related stimuli and resulted in subsequent alcohol use (e.g. buying alcohol and being at a bar, watching others drink alcohol). Microarray analysis was performed on blood cells using the Illumina HumanHT-12 V4.0 expression beadchip (37).

A summary of all the microarray datasets used in this study is presented in Table I.

Statistical analysis. Expression data are presented as mean ± standard deviation (SD). The limma (Linear Models for Microarray Analysis) R package was used for identifying differentially expressed genes. An adjusted P-value < 0.05 and a log(fold change)>1 was considered as threshold of significance.

Fishier’s Inverse $\chi^2$ test was used as integrative analysis for MIF and DDT modulation in blood. This methods computes a combined statistic from the P-values obtained from the individual genes, as follows:

$$s=-2\log\sum_{i=0}^{n}(pi)$$

which follows a $\chi^2$ distribution with 2n degrees of freedom under the null hypothesis. Graphs were constructed using the GraphPad Prism 8 software.

Results

Analysis of brain MIF, DDT and receptors in murine and human alcohol abuse. Analysis of the transcriptomic levels of MIF, DDT and CD74 and the coreceptors, CXCR2, CXCR4 and CXCR7, in a murine model of alcohol dependence (GSE123114), revealed no significant modulation in Inflexible Drinkers as compared to Light Drinkers (Fig. 1A). In a similar manner, as determined in the GSE53808 dataset, in the prefrontal cortex, no significant differences between healthy control subjects and alcoholics were observed for MIF, DDT and their receptors (Fig. 1B). Similarly, as shown on Fig. 1C, alcohol abuse and dependence was not associated to significant alterations in the RNA levels of MIF, DDT and CD74 in the prefrontal cortex, as compared to healthy subjects (GSE49376). The expression levels of the coreceptors could not be evaluated in the latter dataset, as they were below the threshold of detection.

Analysis of blood MIF, DDT and receptors in heavy drinkers. The GSE59206 dataset was interrogated to evaluate the levels of MIF, DDT and their receptors/coreceptors in subjects at risk for alcohol dependence. As shown in Fig. 2A and B, heavy drinkers showed a moderate trend of increase in the levels of MIF and DDT, both at baseline and upon exposure to alcohol-related cues. Interestingly, alcohol-related cues did not, in the short term (1 h), determine a modulation in the levels of these two genes. Although the statistical significance was not reached, and in consideration of the overlapping effects of MIF and DDT (38), an inverse Fisher’s chi square test was performed on unadjusted P-values to evaluate whether the combined effect of MIF and DDT modulation could be statistically significant.
As expected, a combined p value of 0.042 was obtained at baseline and 0.016 at 1 h post-exposure (Fig. 2C and D). No significant modulation was observed for the receptor CD74 and the coreceptors, CXCR2, CXCR4 and CXCR7 (Fig. 2E-H).
Discussion

Increasing experimental and clinical evidence indicate that immunoinflammatory events are induced by alcohol exposure. It has been shown that microglia, the innate immune cells of the brain, and neurons respond to alcohol through receptors and molecules of the innate immune system, including, but not limited to, proinflammatory cytokines and their receptors (39,40). The pathogenic relevance of immunoinflammatory events to the pathogenesis and maintenance of AUD has gained translational relevance with the demonstration of the beneficial effect obtained with minocycline in rodent models of AUD. However, clinical studies with this drug in human patients failed to confirm these preclinical findings (41). Nonetheless, the multiple data supporting a role for neuroinflammation in the pathogenesis of AUD have warranted further studies aimed at identifying other cells, molecules and receptors of the immune system that may play a role in the pathogenesis of the disorder and may eventually represent suitable therapeutic target.

MIF is a protein that has been discovered at the end of the 1960s, and acquired its name from its ability to inhibit the migration of macrophages. The receptor of MIF is composed of the cell surface receptor CD74, that is associated with CD44, and of additional 3 non-cognate ligand for C-X-C chemokine receptor type 2 (CXCR2), type 4 (CXCR4) and type 7 (CXCR7) (38). A second member of the MIF family has also been identified in 2011, named D-dopachrome tautomerase (D-DT), that exhibits multiple synergisms with MIF [reviewed in (38)].

MIF, and more recently DDT, have been implicated in the pathogenesis of several pathologies, including immunoinflammatory and autoimmune diseases (38,42-44) and cancer (45-47). As anticipated above, that MIF may represent an important pathogenic molecule in the pathogenesis of AUD emerges from recent studies indicating that the dual inhibitor of phosphodiesterase -4 and -10 and MIF, named IBUD, has beneficial effects of AUD both in rodent models and human patients (31,32). In particular, this trial consisted of a randomized, crossover, double-blind, placebo-controlled laboratory study of IBUD in non treatment-seeking individuals that had suffered with mild-to-severe AUD, within the last month. The trial evaluated the safety, tolerability, and initial human laboratory efficacy of IBUD (50 mg b.i.d.) on primary measures of subjective response to alcohol, as well as secondary measures of cue- and stress-induced changes in craving and mood. IBUD was well tolerated but it failed to significantly influence primary measures. However, it significantly improved mood on the secondary measures of stress exposure and reduced tonic levels of craving. Furthermore, exploratory analyses indicated that IBUD preferentially influenced the stimulant and mood altering effects of alcohol as compared to placebo in those patients exhibiting the most severe depressive symptoms (39).

However, since IBUD is a dual inhibitor of PDE 4-10 and MIF, it is difficult to understand whether the effects in AUD is due to the combined synergistic pharmacological properties of IBUD or whether either of them accounts for by the beneficial effects of IBUD. It is therefore important to better identify which of the two pharmacological properties may be eventually relevant for the efficacy of drug in this condition. In addition, since it has recently been demonstrated that the MIF superfamily is composed from at least 2 different
homologues, such as MIF and DDT, that most often, but not always, act synergistically. It is also important to shed light on the possible combined action that MIF, DDT and their receptors, play in development and maintenance of AUD. For this reason, we carried out a transcriptomic study that evaluated the expression of MIF, DDT and their receptors in the brains of AUD patients and in peripheral blood cells of people at risk of developing AUD, under basal conditions and upon exposure to alcohol-related cues. In addition, we also evaluated the transcriptomic analysis of MIF, DDT and their receptors in the brains of a mouse model of alcoholism. This analysis was carried out by DNA microarray analysis that represents a useful in silico toll for the better understanding of pathogenic pathways and the possible prediction of new diagnostic therapeutic approaches in several clinical settings, including immunoinflammatory and autoimmune diseases and fibrotic diseases (45,46,48-59). Our analysis first demonstrated that no differences in the expression of MIF, DDT and their receptors can be found in brains of both rodent models of AUD and patients with this condition. However, when peripheral blood of heavy drinkers was studied, it was possible to observe a consistent, though not statistically significant, trend, of increased expression of MIF and DDT in the blood of heavy drinkers as compared to healthy controls, both at baseline and upon exposure to alcohol-related cues. Interestingly, knowing the overlapping effects of MIF and DDT in several biological responses (38) we performed the inverse Fisher's $\chi^2$ test on unadjusted $p$ values and found out that the combined effect of MIF and DDT modulation was statistically significant as compared to controls. Along with the trend of augmented expression of the CD74 receptors in the peripheral blood of heavy drinkers as compared to controls these data seem to indicate that a hyperactivation of the MIF superfamily may also occur in AUD patients. This finding is of particular relevance for its possible translation, as it has recently been demonstrated that dual specific antagonists of MIF and DDT, such as 4-iodo-6-phenylpyrimidine (4-IPP), have been developed (60). Hence, this new class of drugs could be worthy being studied in rodent models of AUD in a head to head comparison with single MIF inhibitors and eventually IBUD. This is of further particular relevance as MIF has been implicated in the pathogenesis of other forms of dependence and addiction that could benefit from specific dual inhibitors of MIF and DDT. Although our data are the first to demonstrate that a moderate upregulated transcriptomic expression of the MIF-DDT axis occurs in the blood of people at risk of developing AUD, other studies are necessary to fully understand the exact role of MIF and DDT in development and maintenance of AUD. In particular, it will be important to prove whether selected groups of AUD patients can be identified on the basis of their MIF-DDT secretory capacity and whether those with highest expression profile are eventually those that may respond better to tailored anti-MIF-DDT therapies. Studies of genetic polymorphism of MIF and DDT in AUD patients may be warranted. It will also be important to determine on larger number of patients whether in a manner similar to progressive multiple sclerosis, MIF and DDT are more abundantly produced in males than in females (61), as this may further facilitate identification of individual potentially responders to anti-MIF-DDT. In addition, it will be relevant to measure circulating levels of MIF and DDT in AUD patients at different time points during their development and maintenance of addiction, as our transcriptomic analysis do not take into account the amount of MIF and DDT that can be secreted from resident cells, such as epithelial and endothelial cells that are not represented in peripheral blood samples. Nonetheless, in spite of the needs of these further studies, our present analysis provides in vivo evidence for the upregulated synthesis of MIF and DDT in AUD and highlights the possibility that these molecules may offer novel important diagnostic and therapeutic opportunities in prevention and treatment of AUD along with psychotherapeutic approaches.

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Availability of data and materials

The datasets analyzed in the current study are available from the Gene Expression Omnibus website (GEO; https://www.ncbi.nlm.nih.gov/geo/) with accession nos. GSE123114, GSE53808, GSE49376 and GSE59206.

Authors’ contributions

MCP, EM, FN and EC conceived and designed the present study. KM, PF, RDM and LF analyzed the data and prepared the figures. MCP, KM, MSB and EC interpreted the data and wrote the manuscript. EM, RDM and FN critically revised the manuscript. All authors read and approved the final version of the manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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