Facile Synthesis of the Naturally Cytotoxic Triterpenoid Saponin Patrinia-Glycoside B-II and Its Conformer

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Received: 30 October 2013; in revised form: 2 December 2013 / Accepted: 3 December 2013 / Published: 10 December 2013

Abstract: The first chemical synthesis of the natural triterpenoid saponin Patrinia-glycoside B-II, namely oleanolic acid 3-O-α-L-rhamnopyranosyl-(1→2)-[β-D-glucopyranosyl-(1→3)]-α-L-arabinopyranoside, has been accomplished in a linear 11-step sequence with 9.4% overall yield. The abnormal $^1C_4$ conformation of the arabinose residue was found to occur via conformational fluctuation during preparation of the intermediates. Molecular mechanism and quantum chemistry calculations showed that Patrinia-glycoside B-II and its conformer 1 cannot interconvert under normal conditions. Preliminary structure-activity relationships studies indicated that the $^4C_1$ chair conformation of the arabinose residue in the unique $\alpha$-L-rhamnopyranosyl-(1→2)-$\alpha$-L-arabinopyranosyl disaccharide moiety is one of the chief positive factors responsible for its cytotoxic activity against tumors.

Keywords: oleanolic acid saponin; stepwise glycosylation; tumor cytotoxicity; chair conformational fluctuation

1. Introduction

Triterpenoid saponins from terrestrial plants possess enormous structural diversity and varied biological activities [1–3]. Notably, some oleanane-type triterpenoid saponins bearing the
α-L-rhamnopyranosyl-(1→2)-α-L-arabinopyranosyl disaccharide moiety at the C3-OH of the aglycone often present significant antitumor activity [4–8]. Previous research has identified this unique disaccharide moiety as a characteristic sugar sequence that can strongly induce the antitumor activity in some oleanane-type glycosides such as β-hederin [9,10].

Patrinia-glycoside B-II (PB-II, Figure 1), a typical triterpenoid saponin isolated from the seeds of *Patrinia scabiosaefolia* Fischer [11], contains the unique oligosaccharide substructure and also displays prominent inhibitory activity against many tumor cell lines [7]. To investigate the structure-activity relationships (SARs) of this type of saponin, we have developed a convenient method for the chemical preparation of Patrinia-glycoside B-II.

**Figure 1.** The structures of PB-II and its conformer 1.

2. Results and Discussion

2.1. Synthesis

The target compound consists of a trisaccharide moiety and oleanolic acid as the aglycone. With regard to the trisaccharide moiety, there is no neighboring participating group but rather a rhamnose residue connected to the C2-OH of arabinose. Because of this, the standard strategy of preparing a trisaccharide donor followed by coupling with the aglycone is not appropriate, as it would result in a mixture of α- and β-anomers. Accordingly, a stepwise glycosylation strategy was adopted in this work, as this strategy generally affords the 1,2-trans-glycoside linkage exclusively. Based on our experience, we chose isopropylidene, acetyl, and benzoyl groups as the temporary protecting groups for the hydroxyl groups of different sugars, and a benzyl group for the carboxylic acid of the oleanolic acid. In addition, perbenzoylated or peracetylated glycosyl trichloroacetimidates (SD-1 to SD-5) were used as sugar donors, which were readily prepared from L-arabinose, L-rhamnose and D-glucose according to the reported methods [12,13].

As shown in Scheme 1, the benzyl ester of oleanolic acid 2 [9] was glycosylated with perbenzoylated arabinosyl trichloroacetimidate SD-1 under trimethylsilyl trifluoromethanesulfonate (TMSOTf) catalysis to give compound 3 in excellent yield. Debenzoylation of compound 3 in a MeOH solution of NaOMe afforded saponin 4 without affecting the benzyl ester at C-28 of the aglycone. Selective protection of the 3′-OH and 4′-OH groups was successfully carried out using 2,2-dimethoxypropane (Me₂C(OMe)₂) to furnish product 5. Then, the coupling of compound 5 with perbenzoylated rhamnosyl trichloroacetimidate SD-2 under the same glycosylation conditions generated compound 6 in 79% yield. The isopropylidene moiety of compound 6 was removed with
para-toluenesulfonic acid (p-TsOH), and the 3'-OH and 4'-OH were deprotected to afford compound 7. However, products 8 and 9 were generated (in a ratio of 3:7) when we attempted to selectively protect the 4'-OH using Bu$_2$SnO/BzCl [14]. This interesting phenomenon inspired us to check the structure of the former product 7. Based on the $^1$H-NMR spectrum of compound 7, the $J_{1'-2'}$ value of the arabinose residue was adjusted to 1.3 Hz, which is much smaller than the normal value of the $\alpha$-arabinosyl conformation (usually no less than 5.0 Hz). Moreover, the chemical shift of the anomeric carbon (δ 109.3) is larger than that of the natural product (δ 104.8).

**Scheme 1.** Synthesis of compound 1.

Reagents and conditions: (a) TMSOTf, DCM, 4Å MS, r.t., 97%; (b) NaOMe, DCM-MeOH (1:2, v/v), r.t., 96%; (c) Me$_2$C(OMe)$_2$, p-TsOH, DCM, r.t., 89%; (d) TMSOTf, DCM, 4Å MS, r.t., 79%; (e) p-TsOH, DCM-MeOH (1:2, v/v), r.t., 98%; (f) Bu$_2$SnO, toluene, reflux; then BzCl, r.t., 94%; (g) BF$_3$ÆEt$_2$O, DCM, r.t.; (h) BF$_3$ÆEt$_2$O, DCM, 0 °C, 78% (for 2 steps); (i) 10% Pd-C, H$_2$, EtOAc, reflux; (j) NaOMe, DCM-MeOH (1:2, v/v), rt, 45% (for 2 steps).

Hence, we realized that the $^1$C$_4$ chair conformer of the arabinose residue was formed when compound 7 was produced. Based on our knowledge, the aglycone and perbenzoylated rhamnose (connected to the 1'-OH and 2'-OH of the arabinose residue, respectively) increase the steric bulk, resulting in the chair inversion of arabinose from the naturally stable $^4$C$_1$ conformation to the $^1$C$_4$ conformation. As expected, the 4'-OH of compound 7 is on the equatorial bond, which is favorable for generating the thermodynamically more stable compound 9 as the major species. Similar regioselectivity in 3'-OH and 4'-OH of arabinose has been reported previously [10,15–18], with the reduced activity of 3'-OH rationalized by the presence of a bulky group at 2'-OH. The polarities of
compounds 8 and 9 were so similar that it was difficult to separate them by silica gel column chromatography. Fortunately, we found that compound 8 could be slowly converted into compound 9 in the presence of boron trifluoride ethyl etherate (BF\textsubscript{3}-Et\textsubscript{2}O), via a benzoyl group migration [19]. Compound 9 was then reacted with peracetylated glucosyl donor SD-3 under the promotion of BF\textsubscript{3}-Et\textsubscript{2}O at 0 °C to give trisaccharide compound 10 in 78% yield. Finally, facile removal of the benzyl group through catalytic hydrogenation followed by debenzoylation and deacetylation in one pot yielded the conformer 1. Interestingly, this novel saponin displayed a very robust structure; the \(^1\text{C}_4\) conformation of arabinose did not convert to the \(^4\text{C}_1\) form even under reflux conditions at 150 °C for 24 h.

Considering the negative influence of the bulky group at 2\text{I}-OH on the arabinose residue, we changed the perbenzoylated rhamnosyl trichloroacetimidate SD-2 sugar donor to the much smaller SD-4. This time, compound 5 was glycosylated with SD-4 at room temperature to give disaccharide saponin 11 in 73% yield. The isopropylidene group of compound 11 was removed by \(p\)-TsOH to generate the key intermediate 12. Based on the \(^1\text{H}\)NMR spectrum of compound 12, the chemical shift (\(\delta\) 4.73) and coupling constant (\(J_{1'-2'}\)): 5.0 Hz) of the anomeric \(^1\text{H}\) atom suggested that the arabinose ring maintained the normal \(^4\text{C}_1\) conformation. Next, compound 12 was converted into a cyclic ortho ester and was then cleaved in 50% HOAc to furnish product 13 with the \(^3\text{I}-\text{OH}\) on the equatorial bond. Compound 13 was able to couple with a bulkier sugar donor, such as perbenzoylated glucosyl trichloroacetimidate SD-5, under the promotion of BF\textsubscript{3}-Et\textsubscript{2}O at a low temperature. Finally, removal of the benzyl group and all the acyl groups generated the target saponin (Scheme 2). In general, the natural Patrinia-glycoside B-II was synthesized in 11 linear steps from oleanolic acid with a 9.4% overall yield. The \(^1\text{H}\)-NMR and \(^13\text{C}\)-NMR spectra are consistent with those of the natural product. Therefore, Patrinia-glycoside B-II and its conformer were both prepared.

**Scheme 2. Synthesis of Patrinia-glycoside B-II.**

\textbf{Reagents and conditions}: (a) TMSOTf, DCM, 4Å MS, r.t., 73%; (b) \(p\)-TsOH, DCM-MeOH, r.t., 89%; (c) CH\textsubscript{2}C(OEt)\textsubscript{3}, \(p\)-TsOH, DCM, r.t.; (d) HOAc-H\textsubscript{2}O (50%), r.t., 76% (for 2 steps); (e) BF\textsubscript{3}-Et\textsubscript{2}O, DCM, 4Å MS, −78 °C; (f) Pd-C, H\textsubscript{2}, EtOAc, reflux; (g) NaOMe, DCM-MeOH, r.t., 23% (for 3 steps).
2.2. Computational Calculations

To determine the conditions in which the two compounds could interconvert, the Sybyl 6.9 software package (Tripos, St. Louis, MO, USA) was used to analyze structural conformations of compound 1. The structure of compound 1 was built with the Sketcher module, and the energy was minimized by Powell’s method using the Tripos force field. Gasteiger-Hückel charges were used for electrostatic field computations. The minimization was terminated after 1000 steps at a maximum value of the gradient at 0.05 kcal/mol⁻¹ Å⁻¹. The structure was then subjected to a simulated annealing, in which it was heated from 200 K to 1,000 K and annealed slowly to 200 K. The results showed that the $^1C_4$ and $^4C_1$ conformations are stable and unable to interconvert at 1000 K. In addition to the two-chair conformations, some boat conformations with irregular conformation are also stable conformations. Next, a 300 K to 500 K simulated annealing was used to study the structural conformations: besides the original $^1C_4$ conformation, the boat with irregular conformations also has low energy, but the $^4C_1$ conformation could not be found. This suggests that the arabinosyl conformation of compound 1 might not be able to convert to a $^4C_1$ conformation at 500 K.

Further tautomerism studies were carried out using the Amsterdam Density Functional (ADF) program [20]. The geometry optimizations for compound 1, Patrinia-glycoside B-II and the transition state were performed in a vacuum. The total energy values were obtained from the optimization output and are listed in Table 1. From the results, it can be observed that both compound 1 and Patrinia-glycoside B-II are energetically stable. Although the energy gap between them is only 0.314 kcal/mol, interconversion would be difficult given the large energy barrier of 185.79 kcal/mol (Figure 2). Further, the C₁-C₂ chemical bond needs to be broken and created, as shown by the transition state (Figure 3). Therefore, both molecular mechanism and quantum chemistry calculations showed that compound 1 and Patrinia-glycoside B-II cannot interconvert under normal conditions.

| Compound 1  | Patrinia-Glycoside B-II | Transition State |
|------------|-------------------------|-----------------|
| Energy(Kcal/mol) | $-19,986.949$ | $-19,987.263$ | $-19,801.469$ |

**Table 1.** The energy of compound 1, Patrinia-glycoside B-II and Transition State.

**Figure 2.** The energy gap between different conformers.
The transition state conformation.

2.3. Biological Evaluation

A preliminary in vitro pharmacology assay was then performed to evaluate the cytotoxicity of these saponins against eight human cancer cell lines using a standard MTT method, in which 5-fluorouracil (5-FU) was chosen as the positive control. As shown in Table 2, Patrinia-glycoside B-II showed effective inhibitory activity against all of those tumor cell lines at micromolar concentrations. Whereas compound 1 did not exhibit significantly cytotoxicity against any tumor cell lines compared to Patrinia-glycoside B-II and 5-FU. This result indicated that the $^4C_1$ chair conformation of the arabinose residue in the unique $\alpha$-L-rhamnopyranosyl-(1→2)-$\alpha$-L-arabinopyranosyl disaccharide moiety is one of the chief positive factors responsible for its cytotoxic activity against tumors.

Table 2. The IC$_{50}$$^{a,b}$ (µmol·L$^{-1}$) of 5-FU, PB-II and compound 1 on various tumor cell lines.

| Compd | HeLa | HepG2 | HT1080 | A549 | A375-S2 | K562 | HL60 | U937 |
|-------|------|-------|--------|------|---------|------|------|------|
| PB-II | 5.4 ± 0.2 | 4.2 ± 0.8 | 18.0 ± 0.5 | 27.9 ± 0.8 | 15.8 ± 0.1 | 6.2 ± 0.6 | 6.6 ± 0.3 | 5.5 ± 0.3 |
| Compound 1 | >100 | >100 | >100 | >100 | >100 | >100 | >100 | >100 |
| 5-FU | >50 | >50 | 15.2 | >50 | 25.3 ± 1.6 | 36.4 ± 3.1 | 9.6 ± 0.9 | 18.2 ± 2.0 |

$^a$ Concentration inhibiting fifty percent of cell growth for 48 h exposure period of tested samples. Data represent mean values ± standard deviation for independent experiments; $^b$ IC$_{50}$s more than 50 µM are not shown their exact values.

3. Experimental

3.1. General

Commercial reagents were used without further purification unless specialized. Solvents were dried and redistilled prior to use in the usual way. Boiling range of petroleum ether was 60–90 °C. Analytical TLC was performed with silica gel HF$_{254}$. Preparation column chromatography was performed with silica gel H. Melting points were detected with a Büchi B-540 Melting Point apparatus. Optical rotations were measured at the sodium D-line at room temperature with a Perkin-Elmer 241MC polarimeter. $^1$H and $^{13}$C-NMR spectra were recorded on an Avance AV 600 MHz spectrometer.
using Me₄Si as the internal standard. The high resolution mass spectrometry (HRMS) measurements were performed using a high resolution Fourier Transform ion cyclotron resonance (FT-ICR) instrument (Bruker, Germany) operated in electrospray ionization mode.

3.2. Benzyl Oleanolate 3-O-3,4-O-isopropylidene-α-L-arabinopyranoside (5)

The benzyl ester of oleanolic acid 2 (1.1 g, 2.01 mmol), trichloroacetimidate SD-1 (1.27 g, 2.10 mmol) and powdered 4Å molecular sieve (MS 4Å, 500 mg) were mixed in dry dichloromethane (DCM, 25 mL) and allowed to stir at r.t. for 20 min. A dry DCM solution (2 mL) of TMSOTf (0.20 mL, 0.10 mmol) was then added dropwise. The mixture was stirred for about 1 h until the starting materials were completely consumed to form a new compound. The mixture was neutralized with triethylamine (Et₃N, 0.2 mL, 1.43 mmol) and filtered through a pad of Celite. The filtrate was concentrated and purified by silica gel column chromatography (8:1, petroleum ether–ethyl acetate (EtOAc)) to afford compound 3 (1.87 g, 97%) as a white foam. To a solution of compound 3 (1.5 g, 1.50 mmol) in DCM–MeOH (1:2, v/v, 40 mL) was added a solution of NaOMe in MeOH (1.00 mol/L, 1.70 mL). The mixture was allowed to stir at r.t. for 2 h and neutralized with Dowex H⁺ resin and then filtered. The filtrate was concentrated and the residue was subjected to a silica gel column chromatography (EtOAc) to give saponin 4 (991 mg, 96%) as a white amorphous solid. To a solution of compound 4 (679 mg, 1.00 mmol) in dry acetone (10 mL) was added Me₂C(OMe)₂ (0.31 mL, 2.50 mmol) and p-TsOH (17.20 mg, 0.10 mmol). The mixture was stirred for 4 h before Et₃N (0.20 mL, 1.43 mmol) was added. The solution was concentrated and the residue was purified by silica gel column chromatography (6:1, petroleum ether–EtOAc) to afford a white foam 5 (634 mg, 89%) with Rf 0.45 (4:1, petroleum ether–EtOAc). \([\alpha]_D^{25} +45.0 (c 1.60, \text{CHCl}_3); \text{Mp} 76.8–79.2 \degree \text{C}; ^1H-NMR (600 MHz, CDCl}_3): \delta 7.34 (m, 5H, Ar-H), 5.28 (t, J = 3.0 Hz, 1H, H-12), 5.07 (dd, J = 18.7, 12.6 Hz, 2H, PhC-H₂), 4.22–4.17 (m, 3H, H-1', H-4', H-5'-1), 4.06 (dd, J = 7.8, 6.1 Hz, 1H, H-3'), 3.75 (dd, J = 13.9, 3.5 Hz, 1H, H-5'-2), 3.63 (d, J = 7.8 Hz, 1H, H-2'), 3.12 (dd, J = 11.5, 4.6 Hz, 1H, H-3), 2.91 (dd, J = 13.8, 3.3 Hz, 1H, H-18), 2.30 (br s, 1H, OH), 1.54, 1.36 (s each, 3H each, O-(CH₃)₂C-O), 1.11, 0.98, 0.92, 0.89, 0.88, 0.82, 0.60 (s each, 3H each, \(7 \times \text{Me}\)); ^13C-NMR (150 MHz, CDCl₃): \delta 177.4, 143.6, 136.3, 128.3, 127.9, 127.8, 122.4, 110.0, 104.3, 88.9, 78.1, 74.2, 73.2, 65.9, 63.0, 55.4, 47.5, 46.6, 45.8, 41.6, 41.3, 39.2, 39.0, 38.4, 36.6, 33.8, 33.1, 32.6, 32.3, 30.6, 28.2, 28.0, 27.5, 26.0, 25.8, 23.6, 23.3, 22.9, 18.1, 16.8, 16.6, 15.2; HRMS: calcd for C₃₈H₅₉O₇ (M-Bn)−: 627.4261; Found: m/z 627.4255.

3.3. Benzyl Oleanolate 3-O-2,3,4-tri-O-benzoyl-α-L-rhamnopyranosyl-(1→2)-3,4-O-isopropylidene-α-L-arabinopyranoside (6)

A mixture of compound 5 (560 mg, 0.78 mmol), trichloroacetimidate SD-2 (630 mg, 1.00 mmol) and powdered MS 4Å (300 mg) in dry DCM (10 mL) was stirred at r.t. for 20 min. A dry DCM solution of TMSOTf (0.10 mL, 0.05 mmol) was then added dropwise and the mixture was stirred for 2 h, followed by the addition of Et₃N (0.20 mL, 1.43 mmol) and filtration. The filtrate was concentrated and subjected to a silica gel column chromatography (8:1, petroleum ether–EtOAc) to furnish disaccharide saponin 6 (730 mg, 79%) as a white foam with Rf 0.60 (4:1, petroleum ether–EtOAc). \([\alpha]_D^{25} +96.7 (c 2.58, \text{CHCl}_3); \text{Mp} 122.5–124.2 \degree \text{C}; ^1H-NMR (600 MHz, CDCl₃): \delta 8.12–7.21 (m, 20H, Ar-H), 5.87
(dd, J = 10.2, 3.3 Hz, 1H, H-3''), 5.76 (brs, 1H, H-1''), 5.65 (m, 2H, H-2'', H-4''), 5.30 (brs, H-12), 5.07 (dd, J = 22.4, 12.6 Hz, 2H, PhCH₂), 4.53 (m, 1H, H-5''), 4.47 (d, J = 7.8 Hz, 1H, H-1''), 4.35 (m, 2H, H-3', H-4''), 4.17 (m, 1H, H-5'-1), 3.90 (d, J = 22.4, 12.6 Hz, 2H, PhCH₂), 3.79 (m, 1H, H-5'-2), 3.17 (dd, J = 11.3, 4.1 Hz, 1H, H-3), 2.92 (m, 1H, H-18), 1.55, 1.35 (s each, 3H each, O-(C₆H₅)₂C-O), 1.34 (d, J = 6.1 Hz, 3H, H-6''), 1.14, 0.95, 0.93, 0.92, 0.90, 0.89, 0.64 (s each, 3H each, 7 × Me); 13C-NMR (150 MHz, CDCl₃): δ 177.4, 165.7, 165.5, 165.4, 143.7, 136.4, 133.3, 133.2, 132.9, 130.0, 129.7, 129.6, 129.4, 129.3, 128.5, 128.4, 128.3, 128.2, 127.9, 122.5, 110.4, 103.5, 95.4, 89.4, 79.4, 75.5, 73.6, 72.2, 70.8, 70.1, 66.7, 66.1, 62.9, 56.0, 47.6, 46.7, 45.8, 41.7, 41.4, 39.3, 39.2, 38.7, 36.7, 33.8, 33.1, 32.7, 32.4, 30.7, 28.2, 27.8, 27.6, 26.1, 25.9, 23.6, 23.4, 23.0, 18.1, 17.5, 16.9, 16.7, 15.3; HRMS (ESI): Calcd for C₆₅H₈₁O₁₄ (M-Bn)⁻: 1085.5626; Found: m/z 1085.5619.

3.4. Benzyl Oleanolate 3-O-2,3,4-tri-O-benzoyl-α-L-rhamnopyranosyl-(1→2)-α-L-arabinopyranoside (7)

Compound 6 (705 mg, 0.60 mmol) was dissolved in a solution of DCM–MeOH (1:2, v/v, 40 mL) and then p-TsOH (78 mg, 0.45 mmol) was added. The solution was stirred at r.t. for 3 h when Et₃N (0.40 mL, 2.86 mmol) was added and the mixture was concentrated and the residue was purified by silica gel column chromatography (2:1, petroleum ether–EtOAc) to give compound 7 (666 mg, 98%) as a white amorphous solid with Rf 0.36 (2:1, petroleum ether–EtOAc); [α]₂₅D +77.3 (c 2.27, CHCl₃); Mp 143.7–147.2 °C; 1H-NMR (600 MHz, CDCl₃): δ 8.10–7.23 (m, 20H, Ar-H), 5.82 (dd, J = 10.2, 3.1 Hz, 1H, H-3''), 5.69 (m, 2H, H-2'', H-4''), 5.34 (s, 1H, H-1''), 5.29 (br s, 1H, H-12), 5.07 (dd, J = 18.7, 12.6 Hz, 2H, PhCH₂), 4.81 (d, J = 3.0 Hz, 1H, H-1'), 4.34 (m, 1H, H-5''), 4.11–3.98 (m, 3H, H-2', H-4', OH), 3.82 (m, 1H, H-5'-1), 3.67 (m, 1H, H-5'-2), 3.45 (d, J = 7.9 Hz, 1H, H-3''), 3.18 (dd, J = 11.0, 3.2 Hz, 1H, H-3), 2.91 (m, 1H, H-18), 2.52 (br s, 1H, OH), 1.34 (dd, J = 6.1 Hz, 3H, H-6''), 1.12, 1.05, 0.92, 0.89, 0.88, 0.84, 0.61 (s each, 3H each, 7 × Me); 13C-NMR (150 MHz, CDCl₃): δ 177.4, 165.7, 165.5, 143.7, 136.4, 133.3, 133.2, 132.9, 130.0, 129.7, 129.6, 129.4, 129.3, 128.5, 128.4, 128.3, 128.2, 127.9, 122.5, 102.0, 98.2, 90.3, 76.2, 71.5, 70.7, 70.7, 69.7, 67.3, 65.9, 65.5, 61.1, 55.4, 47.6, 46.7, 45.8, 41.6, 41.3, 39.2, 39.1, 38.5, 36.7, 33.8, 33.1, 32.6, 32.3, 30.6, 28.1, 27.6, 25.8, 25.7, 23.6, 23.4, 23.0, 18.2, 17.5, 16.8, 16.4, 15.3; HRMS: calcd for C₆₂H₇₇O₁₄ (M-Bn)⁻: 1045.5313; found: m/z 1045.5307.

3.5. Benzyl Oleanolate 3-O-2,3,4-tri-O-benzoyl-α-L-rhamnopyranosyl-(1→2)-3-O-benzoyl-α-L-arabinopyranoside (8) and benzyl oleanolate 3-O-2,3,4-tri-O-benzoyl-α-L-rhamnopyranosyl-(1→2)-4-O-benzoyl-α-L-arabinopyranoside (9)

Compound 7 (300 mg, 0.26 mmol) and Bu₂SnO (72 mg, 0.29 mmol) were dissolved in dry toluene (6 mL) and heated to reflux. After stirring for 1 h, the solution was cooled to r.t. followed by the dropwise addition of a toluene solution of BzCl (10% V/V, 370 μL). 4 h later, the mixture was quenched by Et₃N, then concentrated and purified by silica gel column chromatography (4:1, petroleum ether–EtOAc) to give the mixture of 8 and 9 (309 mg, 94%).
2H, PhCH$_2$), 4.76 (d, $J = 3.8$ Hz, 1H, H-1'), 4.41 (m, 1H, H-5''), 4.33 (m, 1H, H-2'), 4.29 (m, 1H, H-4'), 4.08 (m, 1H, H-5'-1), 3.71 (br d, $J = 9.6$ Hz, 1H, H-5-2), 3.18 (dd, $J = 11.5$, 3.8 Hz, 1H, H-3), 2.91 (dd, $J = 13.9$, 3.6 Hz, 1H, H-18), 1.33 (d, $J = 6.0$ Hz, 3H, H-6''), 1.11, 1.03, 0.92, 0.90, 0.87, 0.70, 0.60 (s, 3H each, 7 × Me); HRMS: calcd for C$_{69}$H$_{81}$O$_{15}$ (M-Bn)$^-$: 1149.5576; found: m/z 1149.5569.

Compound 9: $R_f$ 0.37 (4:1, petroleum ether–EtOAc); Mp 147.0–152.8 °C; 1H-NMR (600 MHz, CDCl$_3$): $\delta$ 8.12–7.25 (m, 25H, Ar-H), 5.89 (dd, $J = 10.2$, 3.3 Hz, 1H, H-3''), 5.78 (m, 1H, H-2''), 5.71 (t, $J = 10.0$ Hz, 1H, H-4''), 5.47 (s, 1H, H-1''), 5.39 (m, 1H, H-4'), 5.30 (br s, 1H, H-12), 5.07 (dd, $J = 33.1$, 12.5 Hz, 2H, PhCH$_2$), 4.82 (d, $J = 4.1$ Hz, 1H, H-1''), 4.45 (m, 1H, H-5''), 4.26 (m, 1H, H-3'), 4.15 (dd, $J = 12.1$, 6.9 Hz, 1H, H-5'-1), 4.06 (dd, $J = 4.6$, 5.6 Hz, 1H, H-2''), 3.83 (dd, $J = 12.1$, 3.4 Hz, 1H, H-5'-2), 3.21 (dd, $J = 11.7$, 4.3 Hz, 1H, H-3), 2.91 (dd, $J = 13.8$, 3.8 Hz, 1H, H-18), 1.37 (d, $J = 6.2$ Hz, 3H, H-6''), 1.13, 1.11, 0.92, 0.90, 0.90, 0.62 (s, 3H each, 7 × Me). HRMS: calcd for C$_{69}$H$_{81}$O$_{15}$ (M-Bn)$^-$: 1149.5576; found: m/z 1149.5569.

3.6. Benzyl Oleanolate 3-O-2,3,4-tri-O-benzoyl-\(\alpha-L\)-rhamnopyranosyl-(1→2)-[2,3,4,6-tetra-O-acetyl-\(\beta-D\)-glucopyranosyl-(1→3)]-3-O-benzoyl-\(\alpha-L\)-arabinopyranoside (10)

To a solution of the mixture of 8 and 9 (300 mg, 0.24 mmol) in dry DCM (8 mL) was added BF$_3$-Et$_2$O (5% V/V, 0.80 mL) and stirred at r.t. for 6 h when TLC showed complete conversion of the 8. Then, a dry DCM solution of trichloroacetimidate SD-3 (5.80 mL, 0.2 mol/L) was added dropwise at 0 °C. The mixture was allowed to stir for 1 h followed by the addition of Et$_3$N (1 mL, 7.15 mmol) and filtration. The filtrate was concentrated and subjected to a silica gel chromatography (3:1, petroleum ether–EtOAc) to furnish trisaccharide saponins 10 (197 mg, 78% in 2 steps) with $R_f$ 0.23 (3:1, petroleum ether–EtOAc). [$\alpha$]$_D^2$ +32.6 (c 1.75, CHCl$_3$); $^1$H-NMR (600 MHz, CDCl$_3$): $\delta$ 8.12–7.25 (m, 25H, Ar-H), 5.77 (dd, $J = 10.3$, 3.3 Hz, 1H, H-2''), 5.73 (m, 1H, H-3''), 5.69 (t, $J = 10.0$ Hz, 1H, H-4''), 5.39 (s, 1H, H-1''), 5.31 (s, 1H, H-1'), 5.30 (br s, 1H, H-12), 5.28 (dd, $J = 9.9$, 9.6 Hz, 1H, H-3''), 5.22 (d, $J = 3.0$ Hz, 1H, H-3'), 5.13 (dd, $J = 9.4$, 10.4 Hz, 1H, H-4''), 5.11-5.03 (m, 3H, H-2''''), 4.74 (d, $J = 8.2$ Hz, 1H, H-1'''), 4.48 (m, 1H, H-4''), 4.39 (br s, 1H, H-2''), 4.29 (dd, $J = 12.4$, 4.1 Hz, 1H, H-6''-1), 4.23 (dd, $J = 10.0$, 5.9 Hz, 1H, H-5''-1), 4.19-4.17 (m, 2H, H-5''-2), 3.81–3.78 (m, 2H, H-5''-2, H-5'''), 3.16 (dd, $J = 11.3$, 4.4 Hz, 1H, H-3), 2.91 (dd, $J = 13.7$, 3.4 Hz, 1H, H-18), 2.07, 2.01, 1.99, 1.96 (s, 3H each, 4 × CH$_3$CO), 1.35 (d, $J = 6.1$ Hz, 3H, H-6''), 1.13, 0.95, 0.92, 0.90, 0.90, 0.76, 0.61 (s, 3H each, 7 × Me). HRMS: calcd for C$_{82}$H$_{97}$O$_{24}$ (M-Bn)$^-$: 1465.6370; found: m/z 1465.6364.

3.7. Oleanolic Acid 3-O-\(\alpha-L\)-rhamnopyranosyl-(1→2)-[\(\beta-D\)-glucopyranosyl-(1→3)]-\(\alpha-L\)-arabinopyranoside (1)

A suspension of compound 10 (180 mg, 0.12 mmol) and 10% Pd/C (80 mg) in EtOAc (30 mL) was stirred under reflux and bubbled with H$_2$ (25 mL/min) for 2 h. Pd/C was removed through a pad of Celite and the filtrate was concentrated. The sticky residue was dissolved in dry DCM–MeOH (1:2, v/v, 9 mL), to which a freshly prepared NaOMe solution in MeOH (1.0 mol/L, 1.50 mL) was added. The solution was stirred at r.t. for 4 h and then neutralized with Dowex H$^+$ resin to pH 7 and filtered. The filtrate was concentrated and subjected to a silica gel column chromatography (7:3:1,
CHCl₃–MeOH–H₂O) to give a white powder 1 (46 mg, 45%, for 2 steps) with R₆ 0.25 (7:3:1, CHCl₃–MeOH–H₂O). [α]DS 35° −38.5 (c 0.52, MeOH); Mp 210.4–213.8 °C; ¹H-NMR (600 MHz, pyridine-d₅): δ 5.87 (s, 1H, H-1''), 5.51 (s, 1H, H-1'), 5.48 (br s, 1H, H-12), 4.98 (d, J = 7.8 Hz, 1H, H-1''), 4.92–4.90 (dd, J = 1.8 Hz, 1H, H-2'), 4.75–4.72 (m, 1H, H-4''), 4.70–4.68 (m, 1H, H-3'), 4.57–4.55 (m, 1H, H-2''), 4.53–4.49 (m, 3H, H-5'-1, H-3'', H-6'''-1), 4.42–4.20 (m, 6H, H-4', H-6'''-2, H-5'', H-5'-2, H-3''', H-4''''), 4.06 (m, 1H, H-2''), 3.94 (m, 1H, H-5'''), 3.29 (dd, J = 13.8, 3.8 Hz, 1H, H-18), 3.18 (dd, J = 11.7 Hz, 4.3, 1H, H-3), 1.69 (d, J = 6.2 Hz, 3H, H-6''), 1.27, 1.11, 1.00, 0.98, 0.94, 0.90, 0.80 (s, 3H each, 7 × Me); ¹³C-NMR (150 MHz, pyridine-d₅): δ 180.5, 145.1, 122.9, 109.6, 105.4, 101.5, 88.6, 87.7, 82.3, 78.8, 78.4, 75.4, 74.3, 72.9, 72.8, 71.9, 70.6, 70.4, 63.0, 56.0, 48.3, 47.0, 46.8, 42.5, 42.3, 40.0, 39.4, 38.9, 37.3, 34.6, 33.6, 33.5, 31.3, 28.7, 28.6, 26.5, 26.5, 24.1, 24.1, 24.0, 18.9, 18.9, 17.7, 17.3, 15.8; HRMS (m/z): calcd for C₄₇H₇₆NaO₁₆ (M+Na)⁺: 919.5026; found: 919.5030.

3.8. Benzyl Oleanolate 3-O-2,3,4-tri-O-acetyl-α-L-rhamnopyranosyl-(1→2)-3,4-O-isopropylidene-α-L-arabinopyranoside (11)

A suspension of compound 5 (450 mg, 0.63 mmol), trichloroacetimidate SD-4 (1.25 g, 1.50 mmol) and powdered MS 4Å (300 mg) in dry DCM (15 mL) was stirred at r.t. for 20 min and then TMSOTf (0.10 mL, 0.05 mmol) was added. The solution was stirred for 2 h before the reaction was quenched with Et₃N (1 mL, 7.15 mmol). The mixture was filtered and the filtrate was concentrated and subjected to a silica gel chromatography (8:1, petroleum ether–EtOAc) to furnish compound 11 (450 mg, 73%) as a white foam with R₆ 0.55 (4:1, petroleum ether–EtOAc). [α]DS 25° +6.8 (c 0.62, CHCl₃); ¹H-NMR (600 MHz, CDCl₃): δ 7.35–7.34 (m, 5H, Ar-H), 5.34 (brs, 1H, H-1''), 5.33-5.32 (m, 2H, H-2'', H-3''), 5.28 (brs, 1H, H-12), 5.08 (dd, J = 20.1, 12.6 Hz, 2H, PhCH₂), 4.36 (d, J = 7.2 Hz, 1H, H-1'), 4.21 (m,1H, H-3'), 4.20 (m, 1H, H-5''), 4.19 (m, 1H, H-4'), 4.17 (t, J = 7.2 Hz, 1H, H-2'), 3.77 (dd, J = 13.8, 5.4 Hz, 1H, H-5'-1), 3.75 (dd, J = 13.2, 3.6 Hz, 1H, H-5'-2), 3.09 (dd, J = 11.3, 4.2 Hz, 1H, H-3), 2.91 (dd, J = 10.2, 2.8 Hz, 1H, H-18), 2.15, 2.02, 1.97 (s, 3H each, 3 × CH₃CO), 1.53, 1.34 (s, 3H each, O-(CH₃)₂C-O), 1.20 (d, J = 6.0 Hz, 3H, H-6''), 1.12, 1.03, 0.92, 0.89, 0.89, 0.82, 0.61 (s each, 3H each, 7 × Me); ¹³C-NMR (150 MHz, CDCl₃): δ 177.6, 170.4, 170.2, 143.9, 136.6, 128.6, 128.2, 122.7, 110.6, 103.4, 95.4, 89.3, 79.4, 75.3, 73.6, 71.4, 69.8, 69.3, 66.4, 66.1, 62.8, 56.1, 47.8, 46.9, 46.1, 41.6, 39.3, 38.9, 36.9, 34.1, 33.3, 32.9, 32.6, 30.9, 29.9, 28.3, 27.9, 27.8, 26.3, 26.1, 23.8, 23.6, 21.2, 21.0, 20.9, 18.4, 17.6, 17.1, 16.6, 15.6; HRMS (m/z): calcd for C₅₈H₇₄NaO₁₅ (M+Na)⁺: 1013.5610; found: 1013.5615.

3.9. Benzyl Oleanolate 3-O-2,3,4-tri-O-acetyl-α-L-rhamnopyranosyl-(1→2)-α-L-arabinopyranoside (12)

To a solution of compound 11 (300 mg, 0.30 mmol) in a solution of DCM–MeOH (1:2, v/v, 25 mL), was added p-TsOH (104 mg, 0.60 mmol) and the mixture was stirred at room temperature. When TLC (2:1, petroleum ether-EtOAc) showed the deprotection was complete, Et₃N (0.5 mL, 3.58 mmol) was added to quench the reaction and the mixture was concentrated and purified by silica gel column chromatography (2:1, petroleum ether–EtOAc) to give compound 12 (254 mg, 89%) as a white amorphous solid with R₆ 0.36 (2:1, petroleum ether–EtOAc). [α]DS 25° +41.0 (c 0.19, CHCl₃); Mp 142.7–149.6 °C; ¹H-NMR (CDCl₃): δ 7.36–7.31 (m, 5H, Ar-H), 5.29–5.26 (m, 3H, H-3", H-4", H-6")
H-12), 5.10–5.05 (m, 3H, H-2'', PhCH₂), 5.03 (s, 1H, H-1''), 4.71 (d, J = 5.0 Hz, 1H, H-1''), 4.00 (m, 1H, H-5''), 3.93 (m, 1H, H-4''), 3.89-3.85 (m, 2H, H-2', H-3'), 3.74 (dd, J = 11.4, 4.2 Hz, 1H, H-5'-1), 3.61 (dd, J = 11.4, 4.2 Hz), 1H, H-5'-2), 3.13 (dd, J = 11.4, 4.2 Hz, 1H, H-3), 2.91 (dd, J = 13.8, 3.6 Hz, 1H, H-18), 2.15, 2.05, 2.00 (s, 3H each, 3 × CH₃CO), 1.20 (d, J = 6.0 Hz, 3H, H-6''), 1.12, 1.00, 0.99, 0.93, 0.90, 0.81, 0.60 (s, 7 × Me); 13C-NMR (CDCl₃): δ 128.6, 128.2, 128.1, 122.6, 102.0, 98.4, 76.1, 71.1, 69.9, 69.1, 67.2, 66.2, 47.8, 46.9, 46.1, 41.9, 41.6, 39.5, 39.3, 38.7, 36.9, 33.3, 32.9, 32.6, 30.9, 28.3, 27.8, 26.1, 25.9, 23.9, 23.6, 23.3, 21.1, 21.0, 18.5, 17.6, 17.1, 16.6, 15.5; HRMS (m/z): calcd for C₅₅H₈₀NaO₁₃ (M+Na)+: 973.5278; found: 973.5282.

3.10. Benzyl Oleanolate 3-O-2,3,4-tri-O-acetyl-α-L-rhamnopyranosyl-(1→2)-4-O-acetyl-α-L-arabinopyranoside (13)

A mixture of compound 12 (200 mg, 0.21 mmol), CH₃C(OEt)₃ (0.50 mL, 4.02 mmol) and p-TsOH (25 mg, 0.15 mmol) in dry DCM (12 mL) was stirred at r.t. for 1 h before the reaction was quenched with Et₃N (0.5 mL). The solution was concentrated and the residue was dissolved in 50% a.q. HOAc (10 mL) and stirred at r.t. for 2 h. Then the solvents were co-evaporated with toluene for complete removal of HOAc. The straw yellow residue was then subjected to a silica gel chromatography (2:1, petroleum ether–EtOAc) to give compound 13 (159 mg, 76%) as a white foam with Rf 0.27 (2:1, petroleum ether–EtOAc). [α]D 25 +91.0 (c 0.10, CHCl₃); 1H-NMR (600 MHz, CDCl₃): δ 7.34–7.27 (m, 5H, Ar-H), 5.32–5.31 (m, 3H, H-4', H-3'', H-2''), 5.13 (brs, 1H, H-1''), 5.11–5.04 (m, 4H, PhCH₂, H-12, H-2'), 4.09 (d, J = 6.6 Hz, 1H, H-1''), 3.98–3.95 (m, 1H, H-4''), 3.94–3.89 (m, 1H, H-3'), 3.86–3.84 (m, 1H, H-5'-1), 3.64 (dd, J = 3. 6, 12.0 Hz, 1H, H-5'-2), 3.13 (dd, J = 4.2, 1.2 Hz, 1H, H-3), 2.91(dd, 1H, J = 3.6, 10.2 Hz, H-18), 2.15, 2.13, 2.05 (s, 3H each, 3 × CH₃CO), 1.22 (d, J = 6.0 Hz, 3H, H-6''), 1.12, 1.00, 0.99, 0.93, 0.90, 0.81, 0.60 (s, 7 × Me); 13C-NMR (150 MHz, CDCl₃): δ 177.7, 170.87, 170.2, 143.9, 136.7, 129.1, 128.6, 128.2, 128.1, 122.7, 102.9, 98.2, 90.6, 77.4, 77.2, 77.0, 76.1, 71.2, 69.9, 69.1, 67.2, 66.2, 55.8, 47.8, 47.0, 46.1, 41.9, 41.6, 39.5, 39.3, 38.8, 36.9, 34.1, 33.3, 32.9, 32.6, 30.9, 28.3, 27.8, 26.1, 23.9, 23.6, 23.3, 21.1, 21.0, 20.9, 19.4, 18.5, 17.6, 17.1, 16.6; HRMS (m/z): calcd for C₅₇H₈₂NaO₁₄ (M+Na)+: 1015.5480; found: 1015.5484.

3.11. Oleanolic Acid 3-O-α-L-rhamnopyranosyl-(1→2)-β-D-glucopyranosyl-(1→3)f-α-L-arabinopyranoside (Patrinia-glycoside B-II)

To a solution of compound 13 (80 mg, 0.084 mmol) in dry DCM (8 mL) was added BF₃-Et₂O (0.016 mL, 0.13 mmol) and powdered MS 4Å (50 mg). The mixture was stirred at −78 °C for 15 min, then trichloroacetimidate SD-5 (75.0 mg, 0.101 mmol) was added and the mixture was stirred for 1 h before Et₃N (0.16 mL, 1.15 mmol) was added to quench the reaction. The mixture was then diluted with DCM (25 mL) and filtered. The filtrate was concentrated and purified by silica gel column chromatography (5:1, petroleum ether–EtOAc) to afford compound 14 (118 mg, 90%) as a white foam. A suspension of compound 14 (118 mg, 0.075 mmol) and 10% Pd/C (60 mg) in EtOAc (10 mL) was stirred under reflux and bubbled up with H₂ (25 mL/min) for 5 h. Pd/C was removed through filtration and the filtrate was concentrated to dryness. The resulted residue was dissolved in a dry solution of DCM–MeOH (1:2, v/v, 24 mL), to which a freshly prepared NaOMe solution in MeOH (1.0 mol/L, 0.30 mL) was added. The mixture was stirred at r.t. for 2 h and then neutralized with Dowex H⁺ resin
to pH 7 and filtered. The filtrate was concentrated and subjected to a silica gel column chromatography (7:3:1, CHCl3-MeOH-H2O) to give a pale yellow powder, which was recrystallized in MeOH to give the target saponin (20 mg, 23%) as a white powder. Rf 0.23 (7:3:1, CHCl3-MeOH-H2O). Mp 260.2–262.3 °C; [α]D25 −27.5 (c 0.49, MeOH); 1H-NMR (600 MHz, pyridine-d5): δ 6.20 (br s, 1H, H-1''), 5.47 (br s, 1H, H-12), 5.13 (d, 1H, J = 7.8 Hz, H-1'''), 4.87 (d, J = 5.4 Hz, 1H, H-1'), 4.77 (br s, 1H, H-2''), 4.68 (d, 1H, J = 7.8 Hz, H-2'), 4.66–4.50 (m, 4H, H-5'', H-3'', H-4', H-6''-1), 4.41–4.18 (m, 7H, H-5'-1, H-5'-2, H-6''-2, H-3', H-3'', H-4'', H-4''), 3.97-3.94 (m, 2H, H-2'', H-5''), 3.40 (d, J = 14.4 Hz, 1H, H-18), 3.18 (d, J = 10.8 Hz, 1H, H-3), 1.55 (d, J = 6.0 Hz, 3H, H-6''), 1.26, 1.18, 1.08, 0.97, 0.92, 0.87, 0.80 (s, 3H each, 7 × Me); 13C-NMR (150 MHz, pyridine-d5): δ 179.9, 144.5, 122.2, 104.5, 104.5, 101.7, 87.9, 82.0, 78.3, 78.0, 74.7, 74.5, 73.7, 72.3, 72.3, 71.2, 69.8, 67.9, 64.8, 62.3, 55.7, 47.8, 46.4, 46.2, 46.0, 41.9, 41.7, 39.5, 39.3, 38.7, 36.8, 34.0, 33.0, 33.0, 31.9, 30.7, 29.8, 29.4, 28.1, 27.8, 26.4, 28.1, 27.8, 26.4, 25.9, 23.5, 23.4, 18.4, 18.2, 17.1, 16.8, 15.3; HRMS (m/z): calcld for C47H76NaO16 (M+Na)+: 919.5026; found: 919.5030.

3.12. Computational Methods

All computational calculations were performed using the Sybyl software package and Amsterdam Density Functional (ADF) program. PBE functional and standard DZP basis set with large frozen core are used in geometry optimization and transition state search.

3.13. Cell Culture

HeLa human cervical cancer cells; HepG2 human hepatoma cells; HT1080 human fibrosarcoma cells; A549 human lung cancer cells; A375-S2 human melanoma cell; K562 human malignant myeloid cell; HL-60 human promyelocytic leukemia cell and U937 human lymphoma cell were obtained from the American Type Culture Collection (Rockville, MD, USA). The cancer cells were maintained in RPMI 1640 medium with 10% fetal bovine serum (FBS); 2% glutamine. Cultures were maintained in a humidified atmosphere incubator at 37 °C in 5% CO2.

3.14. Cell Viability Assay

The cell viability was evaluated using MTT assay. After diluting to 5 × 10^4 cells mL^-1 with the complete medium, 100 μL of the obtained cell suspension was added to each well of 96-well culture plates. The subsequent incubation was permitted at 37 °C, 5% CO2 atmosphere for 24 h before the cytotoxicity assessments. Tested samples at pre-set concentrations were added to each well, and then the cells were incubated for 48 h. The MTT solution (100 μL, 0.5 mg/mL) was added to each well, and the cells were incubated for another 4 h. The formazan crystals were dissolved in 150 μL of DMSO. Cell viability was assessed by measuring the absorbance at a 492 nm wavelength using an EMax Microplate Reader (Molecular Devices, Sunnyvale, CA, USA).

4. Conclusions

In summary, the natural cytotoxic triterpenoid saponin Patrinia-glycoside B-II and its novel conformer 1 were synthesized by a stepwise glycosylation strategy. The abnormal 1C4 conformation of
the arabinose residue was found to occur via conformational fluctuation during preparation of the intermediates. Molecular mechanism and quantum chemistry calculations showed that compound 1 and Patrinia-glycoside B-II cannot interconvert under normal conditions. Preliminary structure-activity relationships studies indicated that the $^4C_1$ chair conformation of the arabinose residue in the unique $\alpha$-L-rhamnopyranosyl-(1→2)-$\alpha$-L-arabinopyranosyl disaccharide moiety is one of the chief positive factors responsible for its cytotoxic activity against tumors.

**Supplementary Materials**

Supplementary materials can be accessed at: http://www.mdpi.com/1420-3049/18/12/15193/s1.

**Acknowledgments**

The authors gratefully acknowledge the National Natural Science Foundation of China (Grant Nos. 81273358, 21202103) for the financial support of this work.

**Conflicts of Interest**

The authors declare no conflict of interest.

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*Sample Availability*: Not available.

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