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A Systematic Review of Pharmacological Treatment Options Used to Reduce Ischemia Reperfusion Injury in Rat Liver Transplantation

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Abstract

Background

Although animal studies models are frequently used for the purpose of attenuating ischemia reperfusion injury (IRI) in liver transplantation (LT), many of pharmacological agents have not become part of clinical routine.

Methods

A search was performed using the PubMed database to identify agents, from which 58 articles containing 2700 rat LT procedures were selected. The identified pharmacological agents were categorized as follows: I - adenosine agonists, nitric oxide agonists, endothelin antagonists, and prostaglandins, II – Kupffer cell inactivator, III - complement inhibitor, IV - antioxidant, V - neutrophil inactivator, VI -anti-apoptosis agent, VII - heat shock protein and nuclear factor kappa B inducer, VIII - metabolic agent, IX - traditional Chinese medicine, and X - others. Meta-analysis using 7-day-survival rate was also performed with Mantel-Haenszel's Random effects model.

Results

The categorization revealed that the rate of donor-treated experiments in each group was highest for agents from Group II (70%) and VII (71%), whereas it was higher for agents from Group V (83%) in the recipient-treated experiments. Furthermore, 90% of the experiments with agents in Group II provided 7-day-survival benefits. The Risk Ratio (RR) of the meta-analysis was 2.43 [95% CI: 1.88-3.14] with moderate heterogeneity. However, the RR of each of the studies was too model-dependent to be used in the search for the most promising pharmacological agent.
Conclusion
With regard to hepatic IRI pathology, the categorization of agents of interest would be a first step in designing suitable multifactorial and pleiotropic approaches to develop pharmacological strategies.

Introduction
Liver transplantation (LT) has been established as an effective therapy for end-stage liver disease and a standard surgical management option for hepatocellular carcinoma [1, 2]. Despite improvements in immunosuppressive protocols and surgical techniques, graft rejection episodes, as well as primary non-function (PNF) and primary delayed graft function (PDF) are still prevalent [3]. Ischemia Reperfusion Injury (IRI) is inevitable after LT and a major risk factor for PNF and PDF [4]. Furthermore, the shortage of organs available for LT has led to the increasing use of liver grafts with extended donor criteria (EDC) that have greater susceptibility to IRI [5].

Hepatic IRI occurs via a complex pathologic network that features a combination of factors, including impairment of sinusoidal endothelial cells (SECs), activation of Kupffer cells (KCs), disturbance of microcirculation, oxidative stress, inflammation, activation of complement factors, accumulation of leukocytes, apoptosis, and necrosis [6]. Some strategies that have been applied in experimental LT models to decrease IRI include the use of ischemic preconditioning, additives in preservation solutions, gene therapy, and the application of numerous pharmacological agents [7]. From the point of clinical application, various experimental studies have focused on developing pharmacological strategies to reduce PDF and PNF with the aim of disrupting the pathways of IRI [8]. The identification of effective pharmacological agents could expand the available options for surgeons and allow for the use of liver grafts with EDC for transplantation. Unfortunately, promising agents and strategies against IRI have not become part of the clinical routine yet. Additionally, there are few systematically summarized reports which are limited in rat animal model experiments as preclinical studies.

The aim of this study is to systematically review the reported literature in which pharmacological agents against IRI have been studied using rat LT models. Additionally, the study is focused on finding pharmaceutical strategies that could be used in clinical routine as a mean of categorizing the identified studies according to the pathology of hepatic IRI.

Materials and Methods
Literature search
A systematic search of the PubMed database for literature reported in the period between January 1993 and December 2012 was performed. The search parameters were restricted to studies reported in the English language that had an available online abstract. The search command used for the review was "(rat liver transplantation) AND (preconditioning OR pharmacological OR drug OR modification) NOT (partial) NOT (small for size) NOT (ischemic preconditioning)". In addition, literature that examined the identified agents as clinical trial candidates were also assessed for future clinical application. All experimental studies to examine pharmacological agents that were effective against IRI by means of rat LT models were included. Studies were excluded if one or more of the following conditions were applicable: 1) rat models in which machine perfusion, isolated perfused liver, ex vivo treatment, ex vivo perfusion, xenograft, or partial
LT procedures were performed, 2) non-heart beating donors, brain dead models, or fatty liver models, 3) the presence of gene transfection or potentially harmful agents, and 4) a pharmacological agent that was principally used as an immunosuppressant. This systematic review was examined according to PRISMA guideline [9].

Included Studies
The database search yielded 1489 studies, of which 184 studies reported the effects of pharmacological agents on rat LT models. In the end, a total of 58 articles could be included in this review (Fig 1) [10–67].

Data extraction and outcome measures
Data on the type of rat models used in each study, the species and number of rats in the model, the type of cold preservation solution, the cold ischemia time (CIT), hepatic artery reconstruction (HAR), and donor and/or recipient treatment protocols were extracted from the articles. The 7-day survival rates were used to perform a meta-analysis. [68] Approximately 2700 rats underwent LT. All studies used syngeneic rat LT models. In thirty studies, HAR was performed. Pharmacological agents were administered as donor- and/or recipient-treated regimens; 29 studies examined the effect of donor preconditioning, 21 studies focused on recipient treatment options and 8 studies looked at a combined donor-recipient treatment option. The subsequent survival benefit was examined in 31 studies. Transaminases were detected with several methods at various timepoints after LT; thus, these parameters were not compared to assess the effects of an agent.

Categorization of pharmacological agents according to the pathology of the hepatic IRI
The pharmaceutical agents were categorized as follows: I—adenosine and nitric oxide (NO) agonists, endothelin (ET) antagonists, and prostaglandins (PGs), II—KC inactivators, III—complement inhibitors, IV—antioxidants, V—neutrophil inactivators, VI—anti-apoptosis agents, VII—heat shock protein (HSP) and nuclear factor kappa B (NF-kB) inducers, VIII—metabolic agents, IX—agents used in traditional Chinese medicine, and X—others (Table 1).

Group I agents were known to generally preserve microvascular structure and microcirculation in the liver. Treprostinil, a PGI2 analog, plays a critical role in microcirculation [10], and the selective COX-2 inhibitor, FK3311, prevents platelet aggregation and causes vasodilatation [11]. Enalapril is a ACE inhibitor that acts by inducing vasodilation via different pathways [43].

Sotraustaurin is an immunosuppressant that prevents early T-cell activation via a calcineurin-independent pathway. Sotraustaurin treatment was reported to be linked with T-cell-macrophage crosstalk [24]. FR167653 is a potent suppressant of IL-1β and TNF-α production in monocytes and has been reported to be associated with the reduced expression of TF in KCs [31]. It is for these reasons that sotraustaurin and FR167653 were categorized in Group II.

Statistical Analysis
Both the Risk Ratio (RR) and the 95% confidence Interval (CI) for the 7-day survival probability were determined using Mantel-Haenszel’s Random Effects model. The I² statistics were calculated in order to assess the heterogeneity of the studies under review. The I² values of 0%, 25%, 50% and 75% were estimated as “No”, “Low”, “Moderate” and “High” heterogeneity, respectively [69]. A two-tailed p value of less than 0.05 was deemed statistically significant. All
Fig 1. Study flow diagram included in the systematic review.

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Table 1. Characteristics of experimental studies included in the systematic review.

| Drug category (n = 58) | Author, Year | Treatment drug | Treatment | Species | Number | Solution | CI time | HAR | Survival study |
|------------------------|--------------|----------------|-----------|---------|--------|----------|---------|-----|----------------|
| **Group I Adenosine agonist, NO agonist, ET antagonist, PGs (n = 13)** | Ghenem N, 2011\(^{10}\) | Treprostinil | D | Lewis | 50 | UW | 18h | No | No |
| | Oshima K, 2009\(^{11}\) | FK3311 | R | Lewis | 71 | UW | 18h | No | Yes |
| | Huser N, 2009\(^{12}\) | FK506, Aminoguanidine | D | DA | 41 | - | 2h | Yes | No |
| | Farmer DG, 2008\(^{13}\) | Tezosenta | R | SD | 28 | UW | 24h | No | Yes |
| | Reid KM, 2007\(^{14}\) | nor-NOA | R | Lewis | 36 | UW | 18h | No | No |
| | Tsuchihashi S, 2006\(^{15}\) | FK330 | D,R | SD | 48 | UW | 30, 40, 48h | No | Yes |
| | Yangnik GP, 2002\(^{16}\) | L-arginine | R | Lewis | 48 | UW | 18h | No | No |
| | Geller DA, 2001\(^{17}\) | L-arginine | R | Lewis | - | UW | 18h | No | No |
| | Tian YH, 2000\(^{18}\) | Adenosine deaminase inhibitor | D | Lewis | 23 | UW | 44h | No | Yes |
| | Tanaka W, 2000\(^{19}\) | TAK-044 | D | Wistar | 60 | EC | 1h | No | No |
| | Liu H, 1998\(^{20}\) | Prostaglandin E1 | D | Wistar | 16 | EC | 6h | No | No |
| | Xu HS, 1994\(^{21}\) | Prostaglandin E1 | R | SD | 97 | NS | No | Yes |
| | Maeda T, 1998\(^{22}\) | cAMP, cGMP | D,R | Lewis | 112 | UW | 24h | No | Yes |
| **Group II KC inactivator (n = 10)** | Sun K, 2012\(^{23}\) | Taunine | R | SD | 64 | UW | 1h | No | Yes |
| | Schemmer P, 2005\(^{24}\) | Taunine | D | SD | 86 | HTK | 4h | Yes | Yes |
| | Kamo N, 2011\(^{25}\) | Sotraustaurin | D,R | SD | 38 | UW | 30h | Yes | Yes |
| | Chimakonda AP, 2007\(^{26}\) | Methylprednisolone (DMP) | D | SD | 45 | UW | 24h | No | Yes |
| | Liu ZJ, 2006\(^{27}\) | Glycine | D | SD | 80 | UW | 1h | No | Yes |
| | Rentsch M, 2005\(^{28}\) | Glycine | D | Lewis | 69 | UW | 24h | Yes | Yes |
| | Schemmer P, 1999\(^{29}\) | Glycine | D | Lewis | 54 | UW | 1h | Yes | Yes |
| | Urata K, 2000\(^{30}\) | Nisoldipine, Thalidomide | D | Lewis | 24 | UW | 24h | No | Yes |
| | Hashimoto K, 2000\(^{31}\) | FR167653 | R | BN | 36 | UW | 48h | No | Yes |
| | Nishizawa H, 1997\(^{32}\) | Penetrin | D | Lewis | 36 | UW | 12h | No | No |
| **Group III Complement inhibitor (n = 2)** | Zhang J, 2011\(^{33}\) | Complement with CV factor | D | Wistar | 12 | UW | 2h | No | No |
| **Group IV Antioxidant (n = 4)** | Lehmann TG, 1998\(^{34}\) | Soluble complement receptor 1 | R | Lewis | 16 | UW | 24h | Yes | No |
| | Schauer RJ, 2004\(^{35}\) | Glutathione | R | Lewis | 36 | UW | 24h | Yes | No |
| | Koeppel TA, 1996\(^{36}\) | N-acetylcysteine | D,R | Lewis | 16 | UW | 24h | Yes | No |
| | Walcher F, 1995\(^{37}\) | N-acetylcysteine | D,R | SD | 12 | UW | 20h | No | No |
| | Consenza CA, 1994\(^{38}\) | Lazaroid U74006F | D | Lewis | 30 | UW | 24h | No | Yes |

(Continued)
| Drug category (n = 58) | Author, Year | Treatment drug | Treatment | Species | Number | Solution | Cl time | HAR | Survival study |
|-----------------------|--------------|----------------|-----------|---------|--------|----------|---------|-----|----------------|
| Group V Neutrophil inactivator (n = 6) | Schen XD, 200739 | Diannexin | R | SD | 61 | UW | 24h | No | Yes |
| | Tsuchihashi S, 200640 | anti-PSGL | R | Lewis | 32 | UW | 24h | No | Yes |
| | Soejima Y, 200741 | ONO-5046 | R | Lewis | 24 | Ringer | 5h | No | No |
| | Dulkanchainun TS, 199842 | sPSGL-1 | D,R | SD | 20 | UW | 24h | No | Yes |
| | Anthuber M, 199743 | Enalapril | R | Lewis | 18 | UW | 24h | Yes | No |
| | Walcher F, 199644 | WEB2086 | R | SPRD | 26 | UW | 5h | No | No |
| Group VI Anti-apoptosis agent (n = 4) | Grutzner U, 200645 | ANP | D | Lewis | 16 | UW | 24h | Yes | No |
| | Nowak G, 200546 | UDCA | D | Lewis | 16 | UW | 4h | Yes | No |
| | Meuller TH, 200447 | DEVD-fluoromethylketone | D | Lewis | 54 | UW | 16h | Yes | Yes |
| | Natori S, 199948 | IDN-1965 | D,R | Lewis | 10 | UW | 30h | No | Yes |
| | Zeng Z, 201249 | Diazoxide | D | SD | 80 | UW | 4h | No | No |
| | Cheng MX, 201250 | NBD peptides | D | SD | 48 | UW | 18h | No | No |
| | Kaizu T, 200851 | Carbon monoxide | R | Lewis | 42 | UW | 18h | No | No |
| | Fondevila C, 200452 | Biliverdin | D,R | SD | 152 | UW | 24h | Yes | No |
| | Tsuchihashi S, 200353 | Pyrrolidine dithiocarbamate | D | Lewis | 47 | UW | 24h | No | Yes |
| | Fudaba Y, 200154 | Geranylgeranlylactone | D | BN | 46 | NS | 45min* | No | Yes |
| | Fudaba Y, 200055 | Geranylgeranlylactone | D | BN | 20 | NS | 45min* | No | Yes |
| Group VII HSP, NFκB inducer (n = 7) | Zeng Z, 201249 | Diazoxide | D | SD | 80 | UW | 4h | No | No |
| | Cheng MX, 201250 | NBD peptides | D | SD | 48 | UW | 18h | No | No |
| | Kaizu T, 200851 | Carbon monoxide | R | Lewis | 42 | UW | 18h | No | No |
| | Fondevila C, 200452 | Biliverdin | D,R | SD | 152 | UW | 24h | Yes | No |
| | Tsuchihashi S, 200353 | Pyrrolidine dithiocarbamate | D | Lewis | 47 | UW | 24h | No | Yes |
| | Fudaba Y, 200154 | Geranylgeranlylactone | D | BN | 46 | NS | 45min* | No | Yes |
| | Fudaba Y, 200055 | Geranylgeranlylactone | D | BN | 20 | NS | 45min* | No | Yes |
| Group VIII Metabolic agent (n = 2) | Ma ZW, 200756 | Fat emulsion | R | SD | 96 | Ringer | 15min | No | Yes |
| | Morimoto Y, 199657 | Insulin | D | Lewis | 28 | UW | 24h | No | No |
| Group IX Traditional Chinese medicine (n = 6) | Song S 201058 | Sinomenine | D | SD | 76 | UW | 24h | No | Yes |
| | Liang R, 200959 | Danshen | D | SD | 52 | Ringer | 1h | Yes | Yes |
| | Chen T, 201260 | Shenfu | R | SD | 96 | - | 100min | No | No |
| | Zhu WH, 200661 | Shenfu | R | SD | 30 | NS | 4h | No | No |
| | Zhu X, 200362 | Matrine | D | SD | 80 | Ringer | 5h | No | Yes |
| | Zhu XH, 200363 | Matrine | D | SD | 72 | Ringer | 5h | No | No |
| | Tarrab E, 201264 | Cyclosporin-A | D | Lewis | 17 | UW | 24h | No | No |
| | Chen LP, 201065 | Rapamycin | R | Wistar | 128 | UW | 12h | Yes | No |
| | Gao W, 199766 | Minocycline, IFNα, Fumagillin | D | Lewis | 14 | EC | 16h | Yes | Yes |
| | Terakura M, 199567 | Putrescine | R | Wistar | 16 | EC | 6h | No | No |

NO: nitric oxide, ET: endothelin, PGs: prostaglandins, KC: Kupffer cell, NFκB: nuclear factor kappa B, CV: cobra venom, INF: interferon, HAR: hepatic artery reconstruction, SD: Sprague-Dawley, BN: Brown Norway, DA: Dark Agouti, D: Donor, R: Recipient, EC: Euro–Collins solution, UW: University of Wisconsin solution, NS: normal saline, UDCA: ursodeoxycholic acid, *

*: 37°C, -: not estimated
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statistical analyses were performed using Review Manager, Version 5 (The Cochrane Collaboration, Oxford, UK).

Results

Agents that deactivated Kupffer cells and agents that induced HSP and NF-κB were mostly used for donor preconditioning, whereas the agents that prohibited neutrophil activation were administered during recipient treatment.

The number of studies focused on each type of pharmaceutical agent was: Group I—13 studies, Group II—10 studies, Group III—2 studies, Group IV—4 studies, Group V—6 studies, Group VI—4 studies, Group VII—7 studies, Group VIII—2 studies, Group IX—6 studies, Group X—4 studies in total. The number of donor-treated experiments, recipient-treated experiments and both treated experiments and the rate in each group was 5 (39%), 6 (46%), 2 (15%) in Group I, 7 (70%), 2 (20%), 1 (10%) in Group II, 1 (50%), 1 (50%), 0 (0%) in Group III, 1 (25%), 1 (25%), 2 (50%) in Group IV, 0 (0%), 5 (83%), 1 (16%) in Group V, 3 (75%), 1 (25%), 0 (0%) in Group VI, 5 (71%), 1 (14%), 1 (14%) in Group VII, 1 (50%), 1 (50%), 0 (0%) in Group VIII, 4 (67%), 2 (33%), 0 (0%) in Group IX, 2 (50%), 2 (50%), 0 (0%) in Group X, respectively (Fig 2A). The differences of the rates of donor and/or recipient were observed among the 10 groups, suggesting that the categorization might predict suitable phase of treatment options. Most notably, the rates of donor-treated experiment were highest in group II (70%) and VII (71%), whereas the rate in recipient-treated experiments was higher in category V (83%).

The agents that deactivated Kupffer cell potentially have short-term survival benefits

Of the 31 studies that examined survival benefit, only one was excluded from the subgroup analysis on the grounds that it did not use a control group.55 The number of the studies that examined survival benefit in each group is as follows: Group I—6 studies, Group II—9 studies, Group III—0 studies, Group IV—1 study, Group V—3 studies, Group VI—2 studies, Group VII—4 studies, Group VIII—1 study, Group IX—3 studies, group X—1 study (Fig 2B). The number of the studies in Group I decreased from thirteen to six. Meanwhile, the number in group II decreased only from ten to nine, giving impression that agents in Group II were more likely to offer short-term survival benefits. In the subgroup analysis, the number and rates of experimental studies in donor-treated experiments, recipient-treated experiments, and both donor and recipient-treated experiments were 1 (17%), 3 (50%), 2 (33%) in Group I, 6 (67%), 2 (22%), 1 (11%) in Group II, 0 (0%), 2 (67%), 1 (33%) in Group V, 3 (75%), 0 (14%), 1 (25%) in Group VII, and 3 (100%), 0 (0%), 0 (0%) in Group IX, respectively (Fig 2B). The rates of donor-treated experiments in group II and VII were 67% and 75%, and that of recipient-treated experiments in group V was 67%. In Group I, however, the rate of the number of donor-treated experiments decreased from 39% to 17%, suggesting that agents in Group I provide relatively less short-term survival benefits.

The meta-analysis demonstrated that the Risk Ratio was 2.43 [95% CI: 1.88–3.14] with moderate heterogeneity

The meta-analysis showed that RR was 2.43 [95% CI: 1.88–3.14] (Fig 3). However, moderate heterogeneity was observed with statistical significance ($I^2 = 48\%$, $P = 0.002$). In the subgroup analysis in which experimental conditions of 24 hours CIT with University of Wisconsin (UW) solution were used (n = 13), RR was 2.21 [95% CI: 1.77–2.75] and no heterogeneity was
observed ($I^2 = 0, P = 0.87$). In addition, if the subgroup was divide into donor- and/or recipient-treatment regimens, the RR obtained for donor-treated experiment was 2.49 [95% CI: 1.78–3.50], for the recipient-treated experiment was 2.20 [95% CI: 1.40–3.47], and for the both-treated experiment was 2.14 [1.28–3.55], respectively.

**Discussion**

This is the first systematic review and meta-analysis of the efficacy of pharmacological agents in rat LT models. The result of meta-analysis using the 7-day survival rate showed that pharmacological
agents conferred short-term survival benefits that were probably associated with the prevention of PNF and PDF. Pharmacological treatment is believed to be effective to reduce IRI in LT, because their benefits in survival after LT have been proven by experimental researches. Therefore, based on the experimental data that are available today, the identified agents should be further evaluated in human LT. Actually, among the identified agents, methylprednisolone, a pan-caspase inhibitor, recombinant P-selectin glycoprotein ligand (rPSGL-Ig), and N-acetylcysteine (NAC) have already been studied in clinical trials. The agents except NAC have short-term survival benefits that are proven by the identified experimental researches. However, none of the pharmacological agents against IRI have become part of the clinical routine.

First of all, we would consider the results of the reported clinical trials to clarify why the pharmacological agents against IRI in LT are not established as the clinical routine. One study on the effects of methylprednisolone revealed that the administration of the agent reduced the levels of cytokines in donor subjects and preserved the graft function (which was estimated by

![Fig 3. Annotated forest plot for meta-analysis of risk ratio of seven-day-survival probability.](https://repository.kulib.kyoto-u.ac.jp)

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examining the aspartate aminotransferase (AST) and alanine aminotransferase (ALT) levels [70], whereas another research group showed that methylprednisolone treatment conferred little to no survival benefits and was associated with a higher risk of biopsy-confirmed rejection [71]. Baskin-Bey, et al. reported that when a pan-caspase inhibitor was administered only to storage and flash solutions, it reduced the prevalence of graft injury. However, treating the recipient with this agent had detrimental consequences [72], even if the pan-caspase inhibitor administered in the trial under investigation was IDN-6556 and not the variant IDN-1965. RPSGL-Ig was used for recipient-treated procedures, as well as in ex vivo liver flushes [73]. In patients with a donor risk index above the accepted study average, administration of rPSGL-Ig improved serum AST levels. Weigand’s study on the effectiveness of NAC revealed that the agent inhibited the increase in glutathione S-transferase ($\alpha$GST), serum intercellular adhesion molecule (ICAM)-1, and vascular cell adhesion molecule (VCAM)-1 levels after reperfusion of the donor liver [74]. However, Hilmi, et al. reported that NAC was ineffective against reducing the risk of acute kidney injury after LT and was not beneficial in terms of liver function or subject survival [75]. None of these agents resulted in a decrease in the mortality rate, liver failure, or perioperative morbidity in clinical setting, even though some promising pharmaceuticals engendered an improvement in the secondary outcomes of AST, ALT, and some other molecules. Thus, none of these agents resulted in a decrease in the mortality rate, liver failure, or perioperative morbidity in clinical trials. From this point of view, this study revealed other promising agents that had beneficial effects against IRI in LT as shown in Table 1. However, the differences in the RR among identified studies were too model-dependent to be used to find out the most promising agent because each experiment used different cold preserve solutions and CIT with or without HAR. Considering the fact that none of these agents decrease the mortality rate in clinical setting, obtaining the RR of only 2.5 times in the meta-analysis (which could be achieved by each single agent) might be too small to achieve definitive effects against hepatic IRI. Additionally, there are relatively small differences in the observed RR among the donor- and/or recipient- treated subgroups, suggesting that it is unclear which phase is more critical for pharmacological treatment. Therefore, additional strategies will need to be investigated in order to find an action plan that will effectively overcome the complexity of IRI in the clinical setting. Since a rat liver transplantation model is technical demanding, there is only a limited number of publications in contrast to studies using IRI to mimic in part what occur after liver transplantation. A transplant model is a more clinically relevant and thus should be used to address the question whether a new agent may be beneficial to prevent livers from IRI in LT. To increase the number of the studies that can be analyzed large animal studies were included; the effects of ET receptor antagonist (TAK-004) [76], L-arginine [77] and N-acetylcysteine [78] were proved by a pig liver transplantation model as well as a rat liver transplantation model. Four agents which were not included in Table 1 were found, a selective ETA receptor antagonist (BSF208075) [79], thromboxane A2 synthase inhibitor (sodium ozagrel) [80], platelet-activating factor antagonist (E5880) [81], Cardiotrophin-1, which is a cytokine belonging to the IL-6 family [82].

Multifactorial and pleiotropic approaches have been advocated for simultaneous action on several IRI pathologies [9, 83]. However, very few studies have reported on the effectiveness of cocktail treatments as potential pharmacological strategies for clinical application [84]. From the result of this review, the agents that deactivate KCs and the agents that induce HSP and NF-κB can be used in donor preconditioning and the agents that prohibit neutrophil activation can be administered in recipient courses. Additionally, it has been determined that agents classified as KC inactivators can be administered with the aim of engendering short-term benefits after reperfusion. Thus, multifactorial and pleiotropic approaches based on the stated categorizations could be designed as a first step with the pharmacological effects in donor and/or
recipient treatment being taken into full consideration. In our manuscript, all the agents were categorized based on the findings of the evaluated publications.

Secondly, the degree of IRI is dependent on the length and method of ischemia applied to the liver as well as the background condition of the organ [85]. For example, liver steatosis is an important risk factor for IRI in the clinical setting [86]. Differences in the action mechanisms that occur in steatotic and non-steatotic livers were observed [87]. The following drugs were reportedly examined in several studies using fatty liver models that were excluded from this review: a cyclin RGD peptide, recombinant human erythropoietin, and fibronectin-α4β1 integrin [88–91]. Due to the fact that these agents were not used in non-steatotic models, they have not been included in the selected literatures of this review. Therefore, pharmacological effects of a newly designed multifactorial and pleiotropic approach could be examined using different background liver conditions.

Finally, the additional or synergic effects, in combination with the different categories of agents to be used in multifactorial and pleiotropic approaches, should be examined. It must be noted that it would be extremely difficult to anticipate and measure these effects without a biomarker, which could be integrated into the complex pathology of hepatic IRI. Several studies regarding Damage-associated Molecular Patterns (DAMPs) in hepatic IRI were recently published. DAMPs are interestingly indicators of tissue injury as well as first line responders of immunological systems in LT [85, 92], as such, they might be useful biomarkers when examining short- and/or long-term survival benefits of multifactorial and pleiotropic treatment. Biomarkers including AST and ALT should be investigated in a parallel manner in order to measure pharmacological effects and to establish multifactorial and pleiotropic approaches in experimental LT models.

In conclusion, pharmacological strategies could be effective in reducing IRI in LT. The agents identified in this study should be further evaluated in human LT. However, further development of the strategies will be needed in order to better determine the effectiveness of agents in clinical application. The categorization of agents with consideration to hepatic IRI pathology might be the first step in designing multifactorial and pleiotropic approaches in rat LT models.

Supporting Information

S1 PRISMA Checklist. Meta-analysis on Genetic Association Studies Checklist. (DOCX)

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Author Contributions

Conceived and designed the experiments: KY PH HB DS EH PS. Performed the experiments: KY PH PS. Analyzed the data: KY PH PS. Contributed reagents/materials/analysis tools: KY PH PS. Wrote the paper: KY PH PS.

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