Thalidomide Effects in Patients with Hereditary Hemorrhagic Telangiectasia During Therapeutic Treatment and in Fli-EGFP Transgenic Zebrafish Model

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Abstract

Background: Hereditary hemorrhagic telangiectasia (HHT) is an autosomal dominant disease characterized by recurrent epistaxis, mucocutaneous telangiectasia, and arteriovenous malformations. The efficacy of traditional treatments for HHT is very limited. The aim of this study was to investigate the therapeutic role of thalidomide in HHT patients and the effect in FLI-EGFP transgenic zebrafish model.

Methods: HHT was diagnosed according to Shovlin criteria. Five HHT patients were treated with thalidomide (100 mg/d). The Epistaxis Severity Score (ESS), telangiectasia spots, and hepatic computed tomography angiography (CTA) were used to assess the clinical efficacy of thalidomide. The Fli-EGFP zebrafish model was investigated for the effect of thalidomide on angiogenesis. Dynamic real-time polymerase chain reaction assay, ELISA and Western blotting from patient’s peripheral blood mononuclear cells and plasma were used to detect the expression of transforming growth factor beta 3 (TGF-β3) messenger RNA (mRNA) and vascular endothelial growth factor (VEGF) protein before and after 6 months of thalidomide treatment.

Results: The average ESS before and after thalidomide were 6.966 ± 3.093 and 1.799 ± 0.627, respectively (P = 0.009). The “telangiectatic spot” on the tongue almost vanished; CTA examination of case 2 indicated a smaller proximal hepatic artery and decreased or ceased hepatic artery collateral circulation. The Fli-EGFP zebrafish model manifested discontinuous vessel development and vascular occlusion (7 of 10 fishes), and the TGF-β3 mRNA expression of five patients was lower after thalidomide therapy. The plasma VEGF protein expression was down-regulated in HHT patients.

Conclusions: Thalidomide reverses telangiectasia and controls nosebleeds by down-regulating the expression of TGF-β3 and VEGF in HHT patients. It also leads to vascular remodeling in the zebrafish model.

Key words: Hereditary Hemorrhagic Telangiectasia; Thalidomide; Zebrafish

Introduction

Hereditary hemorrhagic telangiectasia (HHT) is a hereditary autosomal dominant vascular dysplasia characterized by recurrent epistaxis, mucocutaneous telangiectasia, and arteriovenous malformations. Most HHT patients present recurrent and progressive epistaxis.¹ The efficacy of traditional treatments including compression, laser therapy, embolization, electrocautery, and sclerotherapy is unsatisfactory. Recent reports suggest that vascular endothelial growth factor (VEGF) inhibitor efficiently controlled nosebleeds; however, the normal physiology was affected.²⁻⁴ Thalidomide has also been used to treat HHT patients and control epistaxis.⁵⁻⁷ Here, we report 5 HHT cases treated with thalidomide, showing that it not only controlled nose bleeds but also reversed telangiectasia.

HHT diagnosis is based on the presence of any three of the four criteria, known as the “Curaçao criteria”⁸ as (1) epistaxis, (2) telangiectasia of the lips, oral cavity, fingers, nose, or gastrointestinal tract, (3) arteriovenous malformations of the...
lungs, liver, and central nervous system, and (4) family history involving first-degree relatives. Genetic testing is not necessary. Endoglin (ENG), activin receptor-like kinase-1 (ALK1), and SMAD4 were identified as related to HHT. The degree of epistaxis was estimated by Epistaxis Severity Score (ESS), which is based on six questions including epistaxis frequency, duration, intensity, need for treatment, the presence of anemia, and the need for blood transfusion. ESS is a valuable tool for monitoring disease progression.[1]

Thalidomide was removed from market due to severe congenital defects. It was rediscovered in the treatment of erythema nodosum leprosum and recognized for its anti-angiogenic properties in cancer treatment.[9] A study by Lebrin et al. reports that treatment with thalidomide in a mouse model of HHT stimulated mural cell coverage and thus, rescued vessel wall defects. It increased platelet-derived growth factor-B expression in endothelial cells and stimulated mural cell activation.[10] Angiodyplasia of HHT is characterized by elevated serum levels of VEGF. Transforming growth factor beta (TGF-β) signaling plays an important role in HHT vascular development.[10] We explored the mechanism of thalidomide treatment in HHT and found that VEGF and TGF-β3 were down-regulated following thalidomide therapy. Analysis of Fli-EGFP transgenic zebrafish revealed abnormal vascular development following thalidomide treatment.

**METHODS**

**Study design**

HHT was diagnosed according to Curaçao criteria.[8] Five HHT patients were treated with thalidomide (100 mg/d) continuously at Second Xiangya Hospital between November 2008 and December 2012. Patients’ charts were reviewed, and ESSs were recorded initially and 6 months later, to evaluate the efficiency of epistaxis control. Imaging of telangietasia spots on tongue every 2 months and selective hepatic computed tomography angiography (CTA) was conducted every 6 months. Written informed consent was obtained from the patients and ethical approval was given by the Medical Ethics Committee of Second Xiangya Hospital.

**DNA extraction, amplification, and sequencing**

DNA was extracted from HHT patients’ blood using the QIAGEN DNA Isolation kit (QIAGEN, Germany). ALK1, ENG, and SMAD4 exon polymerase chain reaction (PCR) reactions were carried out in a total volume of 50 μL with 10 mmol/L Tris-HCl, pH 8.3, 1.5 mmol/L MgCl2, 50 mmol/L KCl, 0.2 mmol/L dNTPs, and 30 mmol/L primers. The PCR conditions included 94 cycles for 5 min, followed by 35 cycles with annealing temperatures of 60°C for 0.5 min and 72°C for 0.5 min. The reactions were terminated with a final extension step at 72°C for 5 min.

PCR products were Sanger-sequenced and analyzed on ABI Sequencer (3100) using GeneScan and Genotyper software (PE Applied Biosystems, USA). The nucleotide sequences were deposited in GenBank.

**Real-time polymerase chain reaction**

Total cellular RNA was isolated from HHT patients’ peripheral blood mononuclear cells (PBMCs) using an RNA kit. First-strand complementary DNA (cDNA) synthesis was performed employing moloney murine leukemia virus reverse transcriptase, according to the manufacturer’s protocol (Invitrogen, USA). The TGF-β3 expression was quantified by amplification of cDNA using a real-time thermocycler (Lightcycler 96, Roche, Germany) and SYBR Green I chemistry. The primer sequences were as follows: TGF-β3 (NM_003239.3) sense, 5’GGAAAACACCGAGTCGAATA3’, and antisense, 5’GCACAAAAACCTTGAGGAAGTAAC3’. Optimized amplification conditions were 0.2 μmol/L of each primer, 2.5 mmol/L MgCl2, annealing and extension at 72°C for 40 cycles. The expression levels were standardized with reference to the housekeeping gene β-actin, and the final results were normalized to β-actin. The relative quantification was conducted using the comparative Cr method and expressed as arbitrary units.[11]

**Western blotting**

Equal quantities of plasma were run on 8% sodium dodecyl sulfate-polyacrylamide gel electrophoresis and electrophoblated onto nitrocellulose membranes, exposed to blocking reagent 10% milk in phosphate buffered saline-Tween, and then incubated overnight at 4°C with rabbit anti-human VEGF (Abcam, UK) or mouse anti-human β-actin (Sigma, USA) as the primary antibody, followed by horseradish peroxidase (HRP)-conjugated secondary antibodies. The protein bands were detected using chemiluminescence substrate solution and images were obtained with a gel analyzer system. The VEGF protein was quantified by normalization to β-actin protein and expressed as arbitrary units.

**ELISA assay for vascular endothelial growth factor**

Peripheral venous blood samples from the five HHT patients were collected into a sterile tube containing ethylenediaminetetraacetic acid and centrifuged at 2000 ×g for 10 min. The plasma was stored at −70°C before and after thalidomide treatment until assayed for VEGF. The VEGF plasma concentration was determined using a DVE00 Human VEGF Quantikine ELISA kit (R and D, Wiesbaden, Germany) according to the manufacturer’s instructions. All analyses were carried out in duplicate.

**Zebrafish strain and maintenance**

Fli-EGFP-Tg transgenic zebrafish models were gifted by Hunan Normal University and maintained in aquaria according to standard procedures on a 10-h dark/14-h light cycle at 28°C.[12] Approximately, 96 hpf (hours postfertilization) zebrafish embryos were used for experiments. Before experimental operations, all zebrafishes were anesthetized with 0.02% tricaine (Tricaine Methane Sulfonate, MS-222, Sigma, USA). The zebrafish facility and all experimental procedures were approved by the Second Xiangya Hospital Animal Ethical Committee.
Thalidomide treatment

Fli-EGFP-Tg zebrafish were exposed to 100 μmol thalidomide at 4 hpf and incubated with 96 hpf. The negative control included embryos treated only with 0.1% dimethyl sulfoxide simultaneously. The zebrafish was imaged under confocal microscope.

Confocal microscopy

Ninety-six hours postfertilization embryos were embedded in low melting-point agarose and examined under a confocal microscope (Leica TCS SP5 II microscope Leica Microsystems, Germany). Zebrafish was scanned at 2 μm per section and assembled into three-dimensional images. Vessel analysis was performed with at least 10 fish in each group using the color range tool of Adobe Photoshop CS2 version 9.0.2 software program (Adobe, USA).

Statistical analysis

Data were presented as mean ± standard deviation (SD) analyzed by SPSS Statistics 17 (IBM, Chicago, USA). A standard Student’s t-test was used for statistical analysis, P < 0.05 was considered statistically significant.

Results

Clinical features of five hereditary hemorrhagic telangiectasia patients

Five HHT patients were selected from three families and diagnosed according to the “Curaçao criteria” as follows:

- Family one: The proband was a 56-year-old woman with severe epistaxis, involving bleeding from gastric mucosa telangiectasia and bleeding anemia; the gene mutation involved ALK1.
- Family two: The proband (case 2) was a 45-year-old woman with recurrent epistaxis and severe mucocutaneous telangiectasia of the tongue (Figure 1a). Her two affected sisters (case 3 and case 4) also manifested epistaxis and telangiectasia of lips and fingers. The whole exome sequencing identified ALK1 gene mutation (exon 3, +244 A > C, leading to Thr 82 to Pro variation).
- Family three: The proband (case 5) was a 51-year-old woman who presented with severe epistaxis, mucocutaneous telangiectasia of lip and oral cavity. ALK1, ENG, and Smad4 exon sequencing revealed no mutations.

Thalidomide efficiently controls nosebleeds and reverses telangiectasia

The severity of epistaxis was evaluated by ESS, including frequency, intensity, duration, need for medical attention, transfusion related to epistaxis, and anemia. In the pretreatment period, the average ESS was 6.966 (standard deviation [SD] = 3.093). Following 6 months of thalidomide therapy, the average ESS was 1.799 (SD = 0.627), which was statistically significant (P = 0.009) [Table 1].

In all cases, the “telangiectatic spot” on tongue almost disappeared or decreased in size [Figure 1b]. CTA
examination of case 2 indicated a tortuous malformation in hepatic artery and its branches [Figure 1c]. After a 6-month therapy, the proximal hepatic artery was smaller and hepatic artery collateral circulation decreased or ceased [Figure 1d].

**Thalidomide down-regulated the transforming growth factor beta 3 and vascular endothelial growth factor expression in hereditary hemorrhagic telangiectasia patients**

To explore the potential mechanism of thalidomide therapy, dynamic real-time PCR assay was used to detect the PBMCs of HHT patients before and after thalidomide treatment. The TGF-β3 messenger RNA expression of five patients was lower after thalidomide therapy. The plasma VEGF protein expression in HHT patients was down-regulated, as determined by ELISA (239 ng/ml compared with 159 ng/ml; \( P = 0.019 \)) and in three of five patients tested by Western blotting [Figure 2].

**Thalidomide leads to blood vessel remodeling in zebrafish model**

Confocal microscopy of the Fli-EGFP transgenic zebrafish model revealed discontinuous development and blood vessel occlusion of the dorsal longitudinal anastomotic vessel and intersegmental blood vessel in 7 of 10 fish in the thalidomide treatment group [Figure 3].

**DISCUSSION**

The efficacy of traditional therapies for HHT including local therapy, surgery, and drug therapy are unsatisfactory. Since 2010, a few cases were reported showing that thalidomide successfully controlled HHT-related nose bleeds.\[^7,14-16\] Most of them included case reports except for Franck Lebrin’s study exploring the mechanism of thalidomide in vessel development.\[^7\] We not only observed improved epistaxis, but telangiectasia was actually reversed after thalidomide therapy, unlike previous studies. The hepatic vessel abnormality was also partly reversed.

Mutations in ENG and ALK1 in the TGF-β signaling pathway have been reported to cause up to 85% of HHT. TGF-β family members play an important role in vascular development. The effect of TGF-β on angiogenesis is context-dependent.\[^17,18\] TGF-β was shown to promote EC proliferation and migration at low concentrations whereas high concentrations had the opposite effect.\[^17-20\] Low concentrations of TGF-β enhanced the angiogenic effects of VEGF. Of the three isoforms of TGF-β, TGF-β1 was the most effective. We detected the expression of TGF-β1, TGF-β2, and TGF-β3 before and after thalidomide therapy, and found that only TGF-β3 was regularly altered, which was probably related to limited sample size.

VEGF signaling plays a crucial role in angiogenesis, and several studies suggest a crosstalk between VEGF and ALK1/endoglin signaling in angiogenesis.\[^21\] Our previous study showed a higher VEGF expression in HHT patients. Our data showed that the VEGF expression level was down-regulated after thalidomide treatment. VEGF activation triggers multiple signaling networks that result in endothelial cell survival, mitogenesis, migration, and differentiation. Down-regulated VEGF probably explains the effect of thalidomide in reversing the vessel abnormality.\[^22\]

Fli-EGFP is an ideal model for vascular research and development involving EGFP with VEGF receptor. Our research suggests that zebrafish vessel development was affected following thalidomide intervention. Thalidomide appears to be a good choice in severe HHT-related bleeding, without any of the adverse effects associated with traditional therapies.

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**Figure 1:** Telangiectasia images of case two with hereditary hemorrhagic telangiectasia. (a) Multiple mucocutaneous telangiectasia on the tough (before thalidomide therapy); (b) 6 months after thalidomide therapy; (c) the image of hepatic CTA (before thalidomide therapy); and (d) the image of hepatic CTA for 6 months after thalidomide therapy; the hepatic proper artery proximal transverse diameter is 125 mm, which is smaller than before thalidomide therapy (130 mm). Arrows show the hepatic proper artery proximal.

**Figure 2:** Changes of transforming growth factor beta 3 messenger RNA in peripheral blood mononuclear cells before and after thalidomide treatment in patients with hereditary hemorrhagic telangiectasia. (a) Real-time polymerase chain reaction results; (b) plasma vascular endothelial growth factor protein expression Western blotting results after 6 months of thalidomide therapy in 5 hereditary hemorrhagic telangiectasia patients ("-" means no thalidomide treatment; "+" represents thalidomide treatment).
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