Effects of the Application of Biochar to Plant Growth and Net Primary Production in an Oak Forest

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Abstract: Few studies have evaluated the application of biochar to forest ecosystems and their responses under field conditions. We manually spread grounded biochar on the forest floor, at rates of 0 (control), 5, and 10 Mg ha⁻¹ (C0, C5 and C10, respectively), of an oak forest in central Japan to test the effects of biochar on tree growth and productivity. The relative growth rate of the diameter at breast height (dbh) of canopy oak trees (dbh > 20 cm) significantly increased in C10 compared with that of the control (C0), but not in C5, in the second to third years after application. Despite the increasing growth rate of canopy trees, foliage production (NPP_F) and woody production (NPP_W) did not respond to biochar application. Conversely, the production of reproductive organs (NPP_R, mainly oak acorns) increased in line with the biochar application rate gradients (1.04 ± 0.09 Mg ha⁻¹ yr⁻¹ in C0, 1.30 ± 0.08 Mg ha⁻¹ yr⁻¹ in C5, and 1.47 ± 0.13 Mg ha⁻¹ yr⁻¹ in C10). Since the contribution of NPP_R to total NPP was fairly small, there were no significant differences in total NPP (=NPP_W + NPP_F + NPP_R) for C5 (14.57 ± 0.20 Mg ha⁻¹ yr⁻¹) or C10 (16.11 ± 0.73 Mg ha⁻¹ yr⁻¹) compared with the control (15.07 ± 0.48 Mg ha⁻¹ yr⁻¹).

Keywords: biochar; diameter growth; field experiments; NPP; Quercus serrata; seed production

1. Introduction

Biochar is a product of the thermal degradation of organic matter, such as plant residues, in the absence of oxygen; this process is known as pyrolysis [1]. Many types of organic matter, such as trees, grasses, and crop residues, can be converted into biochar. Since biochar has a long life span, the application of biochar to soil for the purpose of storing carbon (C) has been suggested as one possible means of reducing atmospheric CO₂ concentrations [2]. Recently, biochar has been commonly applied to arable land not only for the purpose of C storage in soil, but also for soil amendment, to enhance crop productivity. Numerous studies have addressed the effects of the addition of biochar on soil properties, crop growth, and productivity [1,3–5]. Biochar soil amendment experiments have demonstrated an 11.0% increase in productivity over controls on average, with a much higher increase in crop productivity when applied to sandy and/or acidic soils [4]. On the other hand, the effects of biochar on crop growth are highly variable when viewed in individual cases, with responses ranging from increased growth to decreased growth [6] or no response [7]. Jeffery et al. [8] also revealed that biochar has no effect on crop yield in temperate latitudes, yet elicits a 25% average increase in yield in the tropics.
Sequestration of CO\textsubscript{2} through afforestation and the use of woody biomass as bioenergy have been accepted as carbon neutral methods, because a forest ecosystem naturally sequesters CO\textsubscript{2} through their biomass or in soil organic matter. Lehman [9] recently demonstrated the potential of turning bioenergy into a carbon-negative industry by using biochar in forest ecosystems. He suggested that the ability of forest ecosystems to sequester C can be taken a step further via the pyrolysis of plant biomass into biochar. Although half of the C in the litter is released into the atmosphere as CO\textsubscript{2} with pyrolysis, pyrolysis converts woody materials at a twofold higher carbon content and biochar locks rapidly decomposable C from plant biomass into a much durable form. Moreover, the storage capacity of biochar is not limited in the same way as biomass sequestration [9]. In addition to C storage in biochar, if biochar application has a positive effect on forest productivity as well as arable lands, it may be possible to strengthen the C negative effect by increasing the absorption rate of CO\textsubscript{2} from the atmosphere by trees.

There have been some reviews of the application of biochar to examine its effects on tree growth and productivity, mainly using pot experiments [10,11]. For example, Heiskanen et al. [12] measured the growth of Norway spruce seedlings after transplantation into pots with soil enhanced with biochar. Biochar was mixed into alluvial silty soil in volume proportions of 0, 15, 30, 45, and 60%. There was an unclear trend of seedling growth with the application rate gradients, although no negative effect of the addition of biochar was noted, even at the highest application rate. Lin et al. [13] conducted a mesocosm experiment of the effects of biochar application on tree seedling growth. Soil was collected at a depth of 100 cm from open land in a forest, and two 2-year-old pine trees (\textit{Pinus elliottii}) were transplanted to each mesocosm. The study revealed that chicken manure biochar provided at 2.4 kg m\textsuperscript{-2} increased tree net primary production (NPP; aboveground biomass plus litterfall) by approximately 180%. In contrast, Saruer and Coleman [14,15] showed that using biochar did not improve Douglas fir seedling growth when the seedlings were grown in containers with biochar-amended peat-based growing media in greenhouses. Nonetheless, each of these previous studies reported the effect of biochar on tree seedlings or saplings, yet few studies have been conducted using direct forest site investigation [16,17].

Until the 1960s, Japanese forests near and around human settlements were used as an energy source, such as for charcoal or firewood. These forests were later abandoned to become secondary forests, because of the rapid decline in the demand for woody energy, or were transformed into plantations. Consequently, Japan has the third-highest forest rate (68.4%) among OECD member countries, well above the world average of 31.0% [18]. Japanese forests are 41% plantations (mainly evergreen coniferous forest) and 59% natural forests, including secondary forests [19]. Secondary forests of abandoned coppice oak (\textit{Quercus serrata} Thunb. ex Murray) are widely distributed throughout the rural area of the subtropical/warm-temperate regions in central Japan. We have conducted multiple direct large-scale field manipulations to detect the effects of the application of biochar on these secondary forests. Minamino et al. [20] investigated the effect of the application of biochar on leaf litter decomposition, using a litter bag technique, on the present study site, concluding that the addition of biochar enhanced the litter decomposition. Now, we hypothesize that biochar addition will increase tree growth and plant productivity through increasing litter decomposition. This study aimed to evaluate the effects of biochar on growth rate and forest productivity, including woody parts, foliage, and reproductive organs, using a large-scale field experiment in a deciduous forest conducted over the first three years after application.

2. Materials and Methods

2.1. Study Site

A field experiment was conducted in a secondary deciduous forest in Honjo Waseda Research Park, central Japan (36°12′ N, 139°10′ E); it was previously a coppice oak forest used for charcoal production. This deciduous forest is predominated by oak (\textit{Quercus}}
serrata Thunb. ex Murray) with an area of approximately 78 ha. The primary climax vegetation around the area is dominated by subtropical/warm-temperate evergreen broad-leaved trees, such as Castanopsis sieboldii (i.e., lucidophyllous forests) [21]; however, these climax forests have been largely replaced by coppice oak forests for charcoal production. These oak forests were abandoned in the 1970s, but have since naturally regenerated into secondary oak forests with ca. 20 m canopy trees. The study site has a warm-temperate monsoon humid climate with an annual mean air temperature and precipitation of 15.0 °C and 1286.3 mm, respectively (1981–2010). The soil was originally derived from alluvion volcanic ash, classified as Alic Hapludands [22].

2.2. Experimental Design

A total of 12 experimental plots (numbered from 1 to 12, 20 m × 20 m each) were established in a randomized design across the deciduous forest. In July 2015, all stems of tree species with a height of more than 1.3 m were identified at the species level and measured to evaluate their diameter at breast height (dbh) to determine the differences in community attributes among the treatment plots before biochar application. The relative dominance of each species was calculated as the relative total basal area (RBA) of each species in each plot. The diversity index (H’) of each plot was calculated using the Shannon-Wiener formulation:

\[ H' = -\Sigma p_i \log_2 p_i \]  

where \( p_i \) is the RBA of species \( i \).

Biochar was spread manually on top of the organic layer on the forest floor at rates of 0 (control), 5, and 10 Mg ha\(^{-1}\) (plots C0, C5, and C10, respectively). Each treatment was applied to four replicate plots in November 2015. The grounded biochar (particle size <5 mm) was made from broad-leaved and coniferous wood chips pyrolyzed at 600–700 °C (Shiratori Super MOKUTAN C, Shiratori Mokuzai Kako Cooperative Society, Gifu, Japan). The proportion of C in the biochar was about 71% (i.e., 3.6 and 7.1 Mg C ha\(^{-1}\) was used as the amount of C input to plots C5 and C10, respectively). The forest floor was densely covered by an evergreen dwarf bamboo community (Pleioblastus chino) approximately 0.5 m high. The aboveground biomass of the dwarf bamboo was measured by cutting the aboveground parts (1 m\(^2\) area and 4 replications in each plot) in July 2017.

2.3. Net Primary Production (NPP)

The dbh of all tree stems greater than or equal to 5 cm was measured in November 2016 (the first year after biochar application) to estimate the NPP. A number tag was attached to each trunk at a height of 1.3 m using a stapler. The dbh of these stems was re-measured in November 2018 (the third year after biochar application) at the same position on the trunks, together with those of any recruitment stems that reached the 5 cm threshold or died during the 2 y interval. To estimate the above and belowground biomass of the trees, the common allometric equations for Japanese deciduous forest sites were used, as follows [23]:

\[ W_{\text{Top}} \approx 0.1853 \rho D^{2.491}, \quad W_{\text{Cr}} \approx 0.1074 \rho D^{2.189} \]  

where \( \rho \) is the stem wood density with bark (kg m\(^{-3}\)), \( D \) is the dbh (cm), and \( W_{\text{Top}} \) and \( W_{\text{Cr}} \) are the dry weights (kg) of the aboveground parts (stems and branches) and coarse roots, respectively. The stem wood density of Q. serrata (0.5540 ± 0.08719 kg m\(^{-3}\)) [23] was used for all tree stems in the study plot, because the mean relative biomass of Q. serrata for all plots was more than 88%.

The biometric-based NPP (Mg dry weight ha\(^{-1}\) yr\(^{-1}\)) of the deciduous forest was calculated using the following equation [24]:

\[ \text{NPP} = \text{NPP}_W + \text{NPP}_F + \text{NPP}_R \]
where \( NPP_W \) is the annual production of the woody parts of aboveground and coarse roots, \( NPP_F \) is the annual production of foliage, and \( NPP_R \) is the annual production of reproductive organs (flowers and seeds, mainly \( Q. \ serrata \) acorns). We defined \( NPP_W \) to be the net increase in the aboveground (stems and branches) and belowground (coarse roots) matter of all trees during the measuring interval, as follows:

\[
NPP_W = \sum BI_s + \sum BI_i
\]

where \( BI_s \) represents the woody increments of surviving trees in the plot and \( BI_i \) represents the woody increments of ingrowth trees that reached the minimum diameter (5 cm) for 2 y (from November 2016 to November 2018). \( BI_s \) was calculated as the difference between the estimated \( W_{Top} \) and \( W_{Cr} \) in November 2016 and November 2018 using the allometric equations. The increments of ingrowth trees (\( BI_i \)) were calculated as the difference between the estimated \( W_{Top} \) and \( W_{Cr} \) of a tree in November 2018 and the minimum measured diameter (5 cm) [25].

\( NPP_F \) is defined as the aboveground leaf litter; it was included to accommodate for the loss of new leaves produced by plants during an interval. Litter fall production was estimated using five litter traps (1 m\(^2\) in area) set in each plot. The litter traps were installed in June 2016, and litter fall was collected every month during the study period. Annual leaf litter volume was accumulated from April to the following March (e.g., from April 2017 to March 2018 for the year 2017), because almost all trees (mainly \( Q. \ serrata \)) were deciduous species, which normally open leaves in April and shed leaves in December. In 2017, we divided litter fall into foliage, woody materials (twigs and bark), and other materials (including reproductive organs). In 2018, we divided litter fall into foliage, woody materials, reproductive organs (flowers and seeds), and other materials (e.g., broken pieces of bark and insect excrement). These materials were oven-dried to a constant mass and then weighed.

\( NPP_R \) is defined as the flowers and seeds (mainly \( Q. \ serrata \) acorns) found in the litter. We included reproductive organs as other materials in 2017; thus, we could not estimate the actual \( NPP_R \). However, the litter fall of materials other than reproductive organs was relatively small in 2018; the ratio of reproductive organs/(reproductive organs + others) ranged from 81% to 94% for each month. Thus, we estimated \( NPP_R \) for the year 2017 using the ratio of reproductive organs to other materials for each month in 2018.

### 2.4. Statistical Analysis

One-way analysis of variance followed by Tukey’s test was used to test differences in community attributes prior to biochar application, and differences in forest NPP parameters after biochar application, across the three treatments. Means ± standard error for each treatment, including the four plots, are shown.

A linear mixed model (LMM) was fitted to clarify whether there was a difference in the relative growth rate of tree dbh (RGR\(_d\)) in C5 and C10 compared to the control plot (C0). We used RGR\(_d\) instead of the growth rate (annual diameter growth) to test the biochar effect on plant growth, because the growth rate of the dbh of tree stems depends on the initial diameter. The RGR\(_d\) of each tree stem was calculated as follows:

\[
RGR_d \ (cm \ cm^{-1} \ yr^{-1}) = \frac{(ln \ D_{2018} - ln \ D_{2016})}{2} \quad (5)
\]

where \( D_{2018} \) and \( D_{2016} \) are the dbh in November 2018 and November 2016, respectively. This analysis was conducted with \( Q. \ serrata \), which was the dominant tree species in each plot (more than 88% of the mean relative biomass for all plots), to eliminate species-dependent responses to biochar from the model. Moreover, we excluded trees under the canopy with a dbh of <20 cm to eliminate the shading effect for plant growth rather than the biochar effect. In the model, the RGR\(_d\) of the individual tree stem was set as the objective variable, with the normal distribution used as the error distribution, the type of treatment
was set as the explanatory variable (fixed effect), and each plot was set as a random effect, as follows:

\[
RGR_d \sim \text{Normal} (\mu, \text{variance})
\]

(6)

\[
\mu = \text{intercept} + \text{treatment [C0, C5, C10]} + \text{random effect [each plot]}
\]

(7)

where \( \mu \) indicates the expected value (=mean) of RGR\(_d\). R 3.6.1 [26] and the lmerTest package [27] were used for the analyses. Significant differences for all statistical analyses were evaluated at the level of \( p = 0.05 \).

3. Results

3.1. Community Structures before Biochar Application

Prior to biochar application, there were no significant differences in community attributes between the treatments (Table 1). Coppice oak trees naturally regenerated in the plots, and other species invaded the plots; thus, the species number in each plot ranged from two to nine, although there were no significant differences in the species number between treatments. The Shannon-Wiener diversity index ranged from 0.08 to 1.09, and \( Q. \ serrata \) mono-dominated (i.e., the relative dominance (RBA) of \( Q. \ serrata \) in each plot was >69.7%). The basal area (BA) of the plots ranged from 19.1 to 32.1 m\(^2\) ha\(^{-1}\), with no significant difference in BA across treatments. The mean aboveground tree biomass before biochar application was 178.3 Mg ha\(^{-1}\), 156.7 Mg ha\(^{-1}\), and 166.8 Mg ha\(^{-1}\), in C0, C5, and C10, respectively; there was no significant difference. The aboveground biomass of the understory dwarf bamboo ranged from 61.2 to 219.3 g m\(^{-2}\) in each plot.

Table 1. Community structures and attributes of the experimental plots in an oak (\( Q. \ serrata \)) forest prior to biochar application. All tree stems with a height ≥1.3 m were measured in each plot (20 m × 20 m).

| Biochar Application | Plot no. | No. of Species | Diversity Index | No. of Stems (plot\(^{-1}\)) | Total BA (m\(^2\) ha\(^{-1}\)) | RBA of \( Q. \ serrata \) (%) | Aboveground Biomass (Mg ha\(^{-1}\)) | Understory Biomass (g m\(^{-2}\)) |
|---------------------|----------|----------------|-----------------|-----------------------------|--------------------------------|----------------------------------|-----------------------------------|-------------------------------|
| Control (C0)        | No. 3    | 7              | 0.44            | 29                          | 20                             | 23.1                             | 90.5                             | 144.3                         |
|                     | No. 6    | 4              | 0.47            | 23                          | 18                             | 32.1                             | 85.8                             | 226.8                         |
|                     | No. 8    | 6              | 0.70            | 27                          | 16                             | 27.7                             | 80.0                             | 189.8                         |
|                     | No. 12   | 6              | 0.30            | 41                          | 20                             | 24.1                             | 94.2                             | 152.4                         |
| Average             | 6.8      | 0.48           | 30.0            | 18.5                        | 26.8                           | 87.6                             | 178.3                           | 155.9                         |
| SE                  | 0.63     | 0.08           | 3.87            | 0.96                        | 2.03                           | 3.06                             | 18.96                           | 32.1                          |
| 5 Mg ha\(^{-1}\) (C5)| No. 1    | 6              | 0.48            | 30                          | 22                             | 27.0                             | 87.8                             | 172.6                         |
|                     | No. 7    | 2              | 0.08            | 14                          | 13                             | 19.1                             | 98.5                             | 129.5                         |
|                     | No. 10   | 7              | 1.00            | 54                          | 15                             | 23.1                             | 71.4                             | 144.3                         |
|                     | No. 11   | 5              | 0.72            | 25                          | 13                             | 26.1                             | 79.6                             | 180.3                         |
| Average             | 5.0      | 0.57           | 30.8            | 15.8                        | 23.8                           | 84.3                             | 156.7                           | 141.8                         |
| SE                  | 1.08     | 0.19           | 8.44            | 2.14                        | 1.79                           | 5.78                             | 11.91                           | 19.3                          |
| 10 Mg ha\(^{-1}\) (C10)| No. 2   | 9              | 1.09            | 23                          | 12                             | 28.6                             | 69.7                             | 206.8                         |
|                     | No. 4    | 5              | 0.56            | 26                          | 13                             | 19.6                             | 86.7                             | 128.0                         |
|                     | No. 5    | 5              | 0.44            | 35                          | 15                             | 25.0                             | 87.9                             | 160.7                         |
|                     | No. 9    | 8              | 0.42            | 40                          | 20                             | 27.2                             | 91.3                             | 171.9                         |
| Average             | 6.8      | 0.63           | 31.0            | 15.0                        | 25.1                           | 83.9                             | 166.8                           | 201.8                         |
| SE                  | 1.03     | 0.16           | 3.94            | 1.78                        | 1.98                           | 4.83                             | 16.26                           | 15.8                          |

3.2. Effects of Biochar Application on Tree Growth

The annual biomass increment (BI\(_s\) and BI\(_i\)) of each stem (i.e., growth rate of trees) increased exponentially with the stem diameter (Figure 1a–c) over two years. Some trees decreased in biomass, as the decreasing diameter reflected weak and dying stems. To eliminate the size-dependent growth rate, we used the relative growth rate of dbh (RGR\(_d\)) to detect the biochar effect on plant growth. The RGR\(_d\) varied substantially for all species other than \( Q. \ serrata \) (Figure 1d–f), although the biomass increments for these species were low because of their small dbh. Table 2 shows a summary of the results of the LMM analysis for tree RGR\(_d\). The fixed effects of C5 and C10 were compared with that of the
control (C0), which was set as 0. The mean RGR_d of the canopy oak trees increased by 6.6% in C5, with no significant difference from the control plots. Conversely, the mean RGR_d was significantly greater in C10 (increased by 22%) than in the control plots.

![Graphs showing size-dependent growth rate and relative growth rate of dbh](image)

**Figure 1.** Size-dependent growth rate (annual biomass increments; (a–c)) and relative growth rate of the dbh (d–f) of all tree stems (dbh > 5 cm) in C0 (control), C5 (5 Mg ha\(^{-1}\)), and C10 (10 Mg ha\(^{-1}\)) treatments after biochar application to an oak (Q. serrata) forest.

**Table 2.** Summary of the results of a LMM to examine the difference in the relative growth rates of the dbh (RGR_d) of canopy oak trees (dbh > 20 cm) across treatments (C0, C5, and C10). The fixed effect of the control (C0) was set to zero (*).

| Coefficient          | Estimate  | Std. Error | t Value | p       |
|----------------------|-----------|------------|---------|---------|
| (Intercept)          | 1.261 × 10\(^{-2}\) | 0.182 × 10\(^{-2}\) | 13.870  | <0.001  |
| Treatment C0         | 0 *       |            |         |         |
| Treatment C5         | 0.083 × 10\(^{-2}\) | 0.272 × 10\(^{-2}\) | 0.610   | 0.543   |
| Treatment C10        | 0.280 × 10\(^{-2}\) | 0.271 × 10\(^{-2}\) | 2.066   | 0.041   |

### 3.3. Effects of Biochar Application on NPP

C5 and C10 tended to have lower and higher woody NPP (NPP\(_W\)), including coarse roots, respectively, compared with the control plots (Figure 2a), but these differences were not significant (F\(_{2,9}\) = 3.51, \(p = 0.075\)). NPP\(_F\) was nearly the same for all treatments (Figure 2b), with no significant differences across treatments (F\(_{2,9}\) = 0.95, \(p = 0.419\)). By contrast, NPP\(_R\) increased along the application rate gradients, corresponding to 1.04 Mg ha\(^{-1}\) yr\(^{-1}\), 1.30 Mg ha\(^{-1}\) yr\(^{-1}\), and 1.47 Mg ha\(^{-1}\) yr\(^{-1}\) in the control, C5, and C10, respectively (Figure 2c). NPP\(_R\) in C10 was significantly higher than the control (F\(_{2,9}\) = 4.65, \(p = 0.041\)). The total NPP in C10 was slightly higher than that of the control plots, mainly because of the higher NPP\(_R\) (Figure 2d). However, this difference was not significant across treatments (F\(_{2,9}\) = 2.29, \(p = 0.157\)), because the contribution of NPP\(_R\) to total NPP was fairly small.
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Figure 2. Effect of the application of biochar on net primary production and components in an oak (Q. serrata) forest. NPP_W (a) included woody growth of aboveground (black part) and coarse root (white part). Total NPP (d) was the sum of NPP_W (a), NPP_F (b), and NPP_R (c). Lower cases represent significant differences among biochar treatments based on one-way ANOVA followed by Tukey’s test.

4. Discussion

4.1. Effect of Biochar on Woody Tree Growth

There was a significant increase in the RGR_d for canopy oak trees of 22% in C10 compared to that of the control in our study site (Table 2). Relatively few studies have examined the effect of biochar on tree growth. Thomas and Gale [10] reviewed existing data on the growth responses of woody plants to the addition of biochar. They analyzed a total of 17 studies examining the responses of 36 woody plant species and found a consistent, strong overall pattern of a positive growth response to the addition of biochar; these responses corresponded to a 41% increase in biomass, although almost all studies were pot trials.

A few direct field experiments examined the effects of biochar application on tree growth. Omil et al. [28] applied mixed wood ash (MWA), a mixture of fly ash and charcoal generated from power plants, to two young Pinus radiata plantations with different soil textures at a rate of 4.5 Mg ha⁻¹ over three consecutive years (13.5 Mg ha⁻¹ total). They found that MWA led to increased tree growth from the second year of treatment onward. Sovu et al. [29] also applied rice husk biochar to swidden fallows in Laos (a total biochar application equivalent to 4 Mg ha⁻¹) to investigate its effects on planted tree seedling growth over four years. In the first year after planting, no significant growth responses were observed for most of the species; however, biochar boosted growth in terms of the diameter and height of saplings for all species in the fourth year. Sherman et al. [16] examined the impacts of biomass retention-level treatments with soil amendment biochar, fertilizer, and combined fertilizer and biochar treatments in two Idaho mixed-conifer forests. Fertilizer increased the basal area growth and total stem volume growth, because nitrogen was limited in the region. However, biochar had no effect on tree growth. Sarauer et al. [17] also applied biochar amendments of 0, 2.5, or 25 Mg ha⁻¹ to the soil surfaces of five types of forests in the western US. They concluded that biochar amendment increased the soil C content by 41% but did not affect tree growth. These studies suggest that tree growth response to biochar amendment is variable, but that growth of a tree species is potentially increased by the application of biochar under field conditions over the short term.

Several mechanisms promote plant growth and productivity. Biochar improves soil nutrient conditions, especially P and K availability [30], decreases Al toxicity [31], and reduces leaching nutrient losses [32]. Biochar can also improve soil water-holding capacity. Biochar-amended soils retained more water at gravity drained equilibrium (up to 15%) in Midwestern Mollisols, in Iowa [32]. Minamino et al. [20], who used our study sites, revealed that the addition of biochar at C10 enhanced litter decomposition above biochar for one year after application because of the biochar increasing the moisture content and microbial
activity and also because of the enhanced litter decomposition below biochar for two years after its application. Moreover, Tanazawa et al. [33] found that the photosynthetic parameters ($P_{\text{max}}$ and $V_{c_{\text{max}}}$) of oak saplings (ca. 10 y old) adjacent to our study site increased with the biochar application rate gradients, up to 10 Mg ha$^{-1}$ for three years. Thus, the application of biochar to the forest ecosystem changed soil properties, such as nutrient availability and water-holding capacity, soon after its addition, and could enhance the photosynthetic and growth rates of oak trees.

4.2. Effect of Biochar on Forest NPP

Even in significant increments of RGRd for canopy oak trees in C10 (Table 2), woody NPP ($\text{NPP}_{w}$) did not respond to biochar application in our study site (Figure 2a). Actual woody biomass increment (growth rate of trees) in each stem was size dependent (Figure 1a–c). Therefore, it is difficult to detect differences in woody NPP (integrated values of woody biomass increments) over the short term in natural forests with trees of various sizes compared to the even-aged artificial forests. Foliage NPP also did not respond to biochar application (Figure 2b). Foliage biomass (i.e., foliage productivity in deciduous forests) is normally a function of forest age. It recovers soon after clear cutting in coppice forests, compared with woody biomass. For example, foliage biomass in the 18 y coppice young deciduous forest (dominated by *Betula platyphylla* var. *japonica* and *Castanea crenata*) was nearly the same as in the mature deciduous forests nearby, whereas woody biomass and NPP were still quite low [34]. Thus, the saturated foliage biomass of mature forests with closed canopies could not easily respond to biochar application.

Nevertheless, there were significant increases in the production of reproductive organs (mainly *Q. serrata* acorns), amounting to 25% in C5 and 41% in C10, compared with the control (Figure 2c). In contrast to foliage production, the productivity of acorns in forest ecosystems may easily respond to nutrient conditions. The masting phenomenon, in which dominant tree species exhibit interannual variation in resource allocation toward reproduction, is known to occur in mature Japanese forests, especially for Fagaceae species that have acorns [35]. Recent studies have shown that mass flowering does not depend on the amount of stored carbohydrates in trees, and masting trees mainly use current-year photosynthates rather than stored carbohydrates [35,36]; thus, the trees require more nutrients than in a normal year. Aoyagi et al. [37] also found that tropical trees require extraordinary amounts of P and K for masting and may re-translocate stored nutrients to meet the elevated nutrient demands of masting. Biochar can improve soil properties by increasing soil pH via a liming effect [38], leading to an increased availability of K and P as soil pH increases [30]. Therefore, abrupt nutrient fluxes that are affected by the addition of biochar should increase the productivity of *Q. serrata* reproductive organs in the same manner as masting phenomena.

It is important to note that there were no negative effects of biochar on plant growth and woody production. Nakagawa et al. [39] noted that, in a tropical rain forest, general flowering had a negative effect on tree growth and aboveground biomass increment at the community level because tree growth and reproduction are subject to trade-offs in resource allocation. Therefore, masting in forest ecosystems normally reduces tree growth and woody NPP [40]. In our study site, NPP$_R$ was significantly greater in C10 than in the control plots (Figure 2c). Nevertheless, there were no differences in the foliage and woody production in C10. This suggests that biochar application might stimulate the strength of C capture in forest ecosystems through increased seed production in C10.

In our study site, the quantitative effect of the biochar application rate on plant growth and productivity is not clear. Crop productivity changes due to biochar application were not shown to be proportional to the biochar application rate (up to 20–40 Mg ha$^{-1}$), although crop productivity was saturated at biochar application rates >40 Mg ha$^{-1}$ [4]. Tanazawa et al. [33] tested the effects of the application of biochar on the photo-synthetic parameters of oak saplings in a manner similar to those used in our field experiments (5, 10, and 20 Mg ha$^{-1}$) in locations adjacent to the present study sites. They concluded that
photosynthetic parameters increased with application rate gradients up to 10 Mg ha\(^{-1}\), but with no biochar effect at 20 Mg ha\(^{-1}\). Khorram et al. [41] applied biochar, compost, or a mixture of both to an apple orchard at an application rate of 10, 25, and 10 + 25 Mg ha\(^{-1}\), respectively, in northeastern Iran. Biochar and compost were beneficial in improving soil quality, mainly by increasing soil nutrient content and decreasing soil bulk density; thus, trunk diameter and the number of apple tree shoots increased by 23%–26% by the end of the first year. Nevertheless, there were no significant changes in fruit weight or the starch pattern index, as indices of productivity. Thus, the effect of the biochar application rate on forest NPP differs depending on the components in forest ecosystems. Additional long-term studies of the response of NPP components in forest ecosystems, especially that of fine root production, and trade-offs in resource allocation for each component are necessary.

5. Conclusions

The application of biochar significantly increased the relative growth rate of canopy oak trees by 22% in the second to third years after application in C10. Responses to biochar application differed depending on the NPP components, although there was no effect on total NPP. The production of reproductive organs (mainly oak acorns) increased by 25% and 41% in C5 and C10, respectively, compared with the control, but there were no effects on foliage or woody production. Nevertheless, we noted positive effect on plant growth and no effect on woody NPP in C10 compared with the control, unlike masting in natural forest ecosystems, which are subject to trade-offs between tree growth and reproduction. This suggests that biochar application might stimulate the strength of C capture for forests in field conditions with the addition of 10 Mg ha\(^{-1}\).

Author Contributions: Conceptualization: T.O., S.Y. and H.K.; investigation: M.T., Y.T. and H.K.; data curation: M.A.; formal analysis: T.O., M.T., M.A. and Y.T.; writing—original draft preparation: T.O., M.T. and M.A.; writing—review and editing: S.Y.; funding acquisitions: S.Y. and H.K. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by JSPS (Japan Society for the Promotion of Science) KAKENHI Grant Number 15H01730 for H.K., and 10H04237 for S.Y.

Data Availability Statement: Data sharing is not applicable to this article.

Acknowledgments: We thank the members of the Laboratory for Environmental Ecology, Waseda University, for their cooperation.

Conflicts of Interest: The authors declare no conflict of interest.

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