Membranoproliferative glomerulonephritis

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Abstract Membranoproliferative glomerulonephritis is an uncommon kidney disorder characterized by mesangial cell proliferation and structural changes in glomerular capillary walls. It can be subdivided into idiopathic and secondary forms, which are differentially diagnosed by a review of clinical features, laboratory data, and renal histopathology. Three types—I, II, and III—have been defined by pathologic features. All three types are associated with hypocomplementemia, but they manifest somewhat different mechanisms of complement activation. Type II, also known as “dense deposit disease”, is associated with the presence of C3-nephritic factor. Membranoproliferative glomerulonephritis primarily affects children and young adults, with patients presenting with nephrotic or nephritic syndrome or with asymptomatic renal disease. This type of glomerulonephritis often progresses slowly to end-stage renal disease, and it tends to recur after renal transplantation, especially type II. The efficacy of various forms of treatment remains controversial; however, long-term steroid treatment seems to be effective in children with nephrotic-range proteinuria. Improvement in renal outcomes largely relies on the evaluation of more selective agents in carefully controlled studies.

Keywords Cryoglobulinemia · Dense deposit disease · Hypocomplementemia mesangiocapillary · Nephritic factor

Introduction

The term membranoproliferative glomerulonephritis (MPGN) is often employed to denote a general pattern of glomerular injury seen in a variety of disease processes that share a common pathogenetic mechanism, rather than to describe a single disease entity [1]. It encompasses forms of unknown cause (idiopathic) and forms associated with systemic and infectious disorders (Table 1). Mixed essential cryoglobulinemia associated with chronic hepatitis C virus (HCV) infection has now been implicated in a substantial number of cases of what were previously thought to be idiopathic MPGN [2].

Morphologically, MPGN is characterized by diffuse mesangial cell proliferation and the thickening of capillary walls due to subendothelial extension of the mesangium; hence, the alternative term “mesangiocapillary glomerulonephritis”. There are at least two (possibly three) distinct morphologic types of MPGN, and all three are related to complement activation (Fig. 1). Type I (classical MPGN), the most common variant, is characterized by the presence of subendothelial deposits of immune complexes associated with activation of the classical complement pathway. Type II MPGN (dense deposit disease), is a unique variant characterized by the presence of additional intra-membranous dense deposits. This type is known to be associated with a serum immunoglobulin, C3 nephritic factor (C3NeF), that stabilizes C3 convertase (C3bBb), thereby resulting in persistent activation of the alternative complement pathway. Type III MPGN, which some pathologists consider as a morphologic variant of type I, is characterized by the presence of additional subepithelial deposits.

Idiopathic MPGN is one of the least common types of glomerulonephritis, accounting for approximately 4 and 7% of primary renal causes of nephrotic syndrome in children.
The incidence of MPGN varies in different parts of the world, but has shown a decline in most developed countries. Interestingly, in Turkey and Nigeria, MPGN has been reported as the most common histopathologic subtype in children with nephrotic syndrome who underwent renal biopsy [4, 5]. All types of MPGN typically have a slowly progressive clinical course. Nonetheless, only 2.8% of end-stage renal disease (ESRD) in children on dialysis and 3.3% of ESRD in pediatric renal transplant recipients are caused by MPGN [6].

**Table 1** Classification of membranoproliferative glomerulonephritis

| Membranoproliferative glomerulonephritis | Associated with systemic and infectious disorders |
|-----------------------------------------|-----------------------------------------------|
| Idiopathic                              |                                               |
| Type I                                  | Viral: Hepatitis B and C, human immunodeficiency virus |
| Type II (dense deposit disease)          | Bacterial: shunt nephritis, visceral abscesses, infective endocarditis |
| Type III                                | Protozoal: quartan malaria, schistosomiasis, leprosy |
| Secondary                               | Other: mycoplasma, mycobacteria                |
| Infectious diseases                     |                                               |
| Systemic immune complex diseases        | Mixed cryoglobulinemia                        |
|                                        | Systemic lupus erythematosus                  |
|                                        | Scleroderma                                   |
|                                        | Sjögren’s syndrome                            |
|                                        | Hereditary deficiencies of complement components |
| Neoplasms                               | Hypocomplementemic urticarial vasculitis       |
| Chronic liver disease                   |                                               |
|                                        | Leukemias and lymphomas                       |
|                                        | Carcinomas                                    |
|                                        | Light-chain disease and plasma cell dyscrasias |
| Miscellaneous                           |                                               |
|                                        | Partial lipodystrophy                         |
|                                        | α1-Antitrypsin deficiency                     |
|                                        | Cystic fibrosis                               |
|                                        | Drugs (e.g. heroin, α-interferon)             |
|                                        | Sarcoidosis                                   |
|                                        | Sickle cell disease                           |
|                                        | Hemolytic uremic syndrome                     |
|                                        | Transplant glomerulopathy                     |

Fig. 1 Schematic depiction of the complement system highlighting the different pathways of complement activation in the three types of membranoproliferative glomerulonephritis (MPGN). NeFt Nephritic factor of the terminal pathway, P properdin.
Pathogenesis

The pathogenesis of MPGN is not yet clearly understood. It is believed that type I MPGN results from chronic antigenemia and the generation of nephritogenic immune complexes that preferentially localize to the subendothelial spaces. The precise nature of the putative antigen(s) in most patients with type I MPGN is unknown; however, a specific pathogenic antigen can sometimes be demonstrated in the glomerular lesions [7]. Recent studies have demonstrated the contribution of innate immunity to both the generation of antibodies that are deposited as immune complexes and to the local inflammatory responses directed at the glomerular immune deposits [8, 9]. The immune complexes activate the complement system via the classical pathway, leading to the generation of chemotactic factors (C3a, C5a) that mediate the accumulation of platelets and leukocytes and of terminal components (C5b-9) that directly induce cell injury. Leukocytes release oxidants and proteases that mediate capillary wall damage and cause proteinuria and a fall of glomerular filtration rate. Cytokines and growth factors released by both exogenous and endogenous glomerular cells lead to mesangial proliferation and matrix expansion [10].

The pathophysiologic basis for type II MPGN seems to be the uncontrolled systemic activation of the alternative pathway of the complement cascade [11, 12]. In most patients, loss of complement regulation is caused by the C3 nephritic factor (C3NeF), an immunoglobulin (Ig) autoantibody that binds and prevents the inactivation of C3 convertase (C3bBb) of the alternative pathway, thereby resulting in the perpetual breakdown of C3. A further cause of type II MPGN is due to mutations in the complement regulatory protein, factor H, or to autoantibodies that impede factor H function, highlighting the role of deregulated alternative complement pathway activity in type II MPGN [12].

Type II MPGN may occur in association with two other conditions, either separately or together: acquired partial lipodystrophy (APD) and macular degeneration. The abnormal activation of the alternative pathway of the complement system is the common link to these seemingly disparate diseases [13]. Acquired partial lipodystrophy is associated with the presence of circulating C3NeF, which can cause a complement-mediated lysis of adipocytes that in turn produce high concentrations of factor D, also called adipin. Factor D cleaves factor B, activating the alternative complement pathway. By analogy, C3NeF may cause damage to glomerular cells that produce the complement. Nonetheless, C3NeF can occur in apparently healthy individuals and in patients with other types of glomerular diseases. In addition, C3NeF does not always correlate with the occurrence or progression of type II MPGN, suggesting the role of other factors [12].

Complement perturbation in type III MPGN is thought to be related to a slow-acting nephritogenic factor that stabilizes a properdin dependent C5-convertase, (Cb3) 2BbP, activating the terminal pathway; hence, the term nephritogenic factor of the terminal pathway (NeFt) [14]. This nephritogenic factor has not been reported in healthy subjects, unlike C3NeF. In addition, the deposits observed in renal biopsies of patients with type III MPGN are closely associated with the circulating nephritogenic factor-stabilized convertase and with hypocomplementemia, suggesting that NeFt is fundamental to the pathogenesis of type III MPGN [15].

The mechanism of renal injury in HCV-associated cryoglobulinemia remains elusive. An estimated 50–60% of patients with chronic HCV infection develop type II cryoglobulins, which are composed of a complex of an IgM kappa monoclonal antibody with rheumatoid factor activity directed against a polyclonal anti-HCV IgG. However, only a minority (10–20%) of such patients with detectible cryoglobulinemia have clinical manifestations of cryoglobulinemic MPGN [2]. It is unclear why some cryoglobulins are more pathogenic than others, or why cryoglobulins deposit in the kidneys. Recognition of the components of cryoprecipitates, which contain HCV core protein, by circulating leukocytes and intrinsic glomerular cells leads to the production of inflammatory mediators that characterize the glomerular injury of cryoglobulinemic MPGN.

Clinical manifestations

Membranoproliferative glomerulonephritis primarily affects children and young adults, with no sex predilection. In children, MPGN is frequently idiopathic, whereas in adults, MPGN is commonly associated with cryoglobulinemia and HCV infection. Patients with MPGN may present in one of four ways, as follows:

1. nephrotic syndrome (40–70%);
2. acute nephritic syndrome (20–30%);
3. asymptomatic proteinuria and hematuria detected on routine urinalysis (20–30%);
4. recurrent episodes of gross hematuria (10–20%).

A respiratory tract infection may precede the diagnosis in half of the patients. Hypertension is found at presentation in one third of patients but occurs more frequently with progressive disease. Renal dysfunction occurs in >50% of patients. The incidence of anemia out of proportion to the degree of renal failure is common and is thought to be related to complement-mediated lysis of red blood cells. Compared with adults, children
are more likely to have hematuria at onset and less likely to have renal insufficiency and hypertension [16].

The different types of MPGN cannot be distinguished based on clinical presentation. However, the presence of associated extra-renal manifestations, such as drusen or APD, suggests type II MPGN. In contrast to drusen that form in age-related macular degeneration, drusen in individuals with MPGN II occur at an early age and often are detectable in the second decade of life. Similarly, the loss of subcutaneous fat in the upper half of the body in APD usually precedes the onset of kidney disease by several years.

Hypocomplementemia is a characteristic feature of all types of MPGN, which is essentially the only cause of idiopathic nephrotic syndrome that is associated with hypocomplementemia. In type I MPGN and cryoglobulinemic MPGN, the classical pathway is preferentially activated (low or normal C3, low C4 and CH50), while in type II MPGN, the alternative pathway is activated (low C3, normal C4 and low CH50), and in type III MPGN, the complement profile typically reveals evidence of the activation of both the alternative and terminal pathways (low C3, normal C4 and low C5 through C9) [17]. The serological detection of nephritic factors is quite useful in distinguishing among the types of MPGN; however, apart from C3NeF, which is positive in over 80% of patients with type II MPGN and even more frequent in children, assays for other nephritic factors are performed primarily in research laboratories and are not widely available. The clinical and laboratory evaluation of patients with suspected MPGN is summarized in Table 2.

The natural history of MPGN is characterized by spontaneous fluctuations of the severity of the clinical picture, with very few cases of spontaneous, complete remission. Children tend to have a more acute presentation and a slower decline in renal function than adults. Approximately 40% of patients progress to ESRD within 10 years of diagnosis. Features suggestive of an adverse outcome include the nephrotic syndrome, renal dysfunction at onset, and persistent hypertension. Type II MPGN is associated with a worse prognosis, as is the presence of chronic interstitial damage on renal biopsy [18].

Membranoproliferative glomerulonephritis may recur in renal transplant recipients with a frequency of 20–30% for type I and 80–90% for type II. The North American Pediatric Renal Transplant Cooperative Study investigated the impact of type II MPGN recurrence in a large population of pediatric renal transplant patients [19]. The study indicated that recurrence had a significant negative impact on renal allograft function and survival, contributing to graft failure in 15% of recipients by 5 years post-transplantation. No clear predictors of disease recurrence were identified in the above-mentioned study.

Histopathology

Type I MPGN

The glomeruli often show a global increase in mesangial cellularity and matrix with diffuse endocapillary proliferation. Infiltrating mononuclear leukocytes and neutrophils also contribute to the glomerular hypercellularity. As a consequence of these changes, the glomerular tufts appear hyperlobulated (Fig. 2b). Another distinctive feature of MPGN is the diffuse thickening and duplication of the glomerular basement membranes (GBM) that can be seen with Periodic acid–Schiff and silver stains. This appearance, also known as tram-tracking, splitting, or reduplication of the GBM, results from the presence of subendothelial deposits and the so-called “mesangial interposition” whereby mesangial cells, infiltrating mononuclear cells, or even portions of endothelial cells interpose themselves between the endothelium and basement membrane, with new, inner GBM-like material being laid down (Fig. 2b). Deposits can occasionally be demonstrated in the subendothelial location by trichrome staining. The proportion of cells to matrix varies with the duration of the disease. In advanced cases, lobular centers may become completely sclerosed, simulating the Kimmelstiel–Wilson nodules of diabetes. Glomerular crescents are present in approximately 10% of cases and may occur in both idiopathic and secondary forms [20]. The presence of eosinophilic hyaline globules in capillary lumina suggests mixed cryoglobulinemia and should be carefully sought. The interstitium, tubules, and vessels show nonspecific changes that correlate with the severity of chronic damage. Vasculitis affecting small and medium-sized renal arteries is strongly associated with underlying cryoglobulinemic MPGN [21].

The most common immunofluorescence finding is strongly positive staining for C3 and, less frequently, for IgG and IgM, in a fine to course granular pattern along the glomerular capillaries. This corresponds to the prominent subendothelial deposits seen by electron microscopy. Early complement components (C1q or C4) and properdin are also frequently present. Occasionally the C3 deposits are predominantly mesangial.

Electron microscopy (Fig. 3a) reveals an expanded mesangium which contains cells, matrix, and often electron-dense deposits. The subendothelial space is widened and filled with mesangial cells and matrix continuous with those in the centriflobular mesangial area and with granular electron-dense deposits. The formation of a “new basement membrane” is demonstrated under the endothelial cell. There is an effacement of podocyte foot processes and occasional subepithelial small electron-dense deposits. The detection of microtubular structures suggests cryoglobulinemia.
Type II MPGN

The glomerular changes in type II MPGN can be quite variable. Glomerular hypercellularity and lobulation, as in type I MPGN, is seen in only 25% of all cases [22]. Thus, the alternative term “dense deposit disease” is probably more accurate than the designation type II MPGN as the former emphasizes the pathognomonic feature of type II MPGN, which is the presence of elongated, brightly eosinophilic, variably refractile deposits within the GBM. The deposits may be continuous and ribbon-like or periodically interrupted, thereby having the appearance of a “string of sausages” (Fig. 2c). On silver-stained specimens, these intra-membranous deposits appear to be light brown and surrounded by darker lines forming a double-contour. Similar deposits may be seen in the tubular basement membrane and Bowman’s capsule. Crescents may be present.

Specimens studied by immunofluorescence microscopy show staining for C3, usually in a linear or granular pattern along the glomerular capillaries, and in the mesangium. Immunoglobulin is not detected, indicating that the dense deposits are not classic antigen–antibody immune complexes. However, segmental IgM or less often IgG and rarely IgA have been reported. Early complement components are usually absent.

The key feature observed on electron microscopy (Fig. 3b) is a layer of highly electron-dense material within the lamina densa of the glomerular capillary basement membrane that splits into two layers. Granular deposits of similar dense material are usually present in the mesangium. Similar deposits are also seen in some tubular basement membranes and Bowman’s capsule.

Type III MPGN

This subtype of MPGN has some features in common with those seen in type I MPGN, such as double contours of the GBM and subendothelial deposits, together with widespread subepithelial deposits similar to those seen in membranous glomerulonephritis. However, hypercellularity is often less prominent than in type I MPGN [23]. Immunostaining reveals a heavy granular pattern along the glomerular capillary walls and mesangium for C3 and sometimes IgG and IgM. Ultrastructural studies may reveal multilayering of the basement membrane material and complex disruption of the lamina densa by large dense deposits.
The efficacy of the various therapeutic regimens tried in MPGN is difficult to assess because of the small patient numbers and short-term nature of published controlled trials; the larger trials carried out to date have been uncontrolled [24]. Moreover, most trials include patients with all three types of MPGN in different proportions, making the analysis of treatment results more difficult. Treatment strategies for idiopathic MPGN are controversial and have included corticosteroids, immunosuppressives, antiplatelet regimens, plasma exchange and biologic agents.

Corticosteroids

Both retrospective and prospective studies have demonstrated the beneficial effect of alternate-day steroid therapy on renal survival in pediatric patients with MPGN. Of these, a prospective, multicenter, randomized trial was conducted by the International Study of Kidney Disease in children (ISKDC) [25]. Eighty children, predominantly with type I MPGN were randomized to receive either 40 mg/m² of prednisone every other day or placebo for a mean duration of treatment of 41 months. Criteria for admission included heavy proteinuria and a glomerular filtration rate (GFR) ≥ 70 ml/min/1.73 m². Treatment failure (30% increase in serum creatinine) occurred less frequently (40%) in patients treated with prednisone than in those who received placebo (55%), and 61% of the treatment group versus 12% of the placebo group had stable renal function at the end of the study. The use of such a protracted course of steroid medication is not without adverse effects, particularly on bone growth. Non-nephrotic children may forego steroid treatment without compromising long-term kidney function [26]. While corticosteroid therapy seems to be effective in children, there is no convincing published evidence that steroids are effective in modifying disease activity or disease progression in adults with idiopathic MPGN.

Cyclophosphamide

In one study [27], 19 pediatric and adult patients with MPGN were treated with an intensive and prolonged regimen of
pulse methylprednisolone plus oral prednisone and cyclophosphamide. Of the 19 patients, 15 achieved complete remission and three achieved a partial remission. Based on these results, the researchers concluded that cyclophosphamide is effective in inducing remission and halting the progression of MPGN to ESRD. There are as yet no studies to suggest whether the addition of cyclophosphamide to a prednisolone therapeutic regimen provides any added advantage. Nonetheless, cyclophosphamide is generally recommended in patients with rapidly progressive renal failure or a recent deterioration in renal function, especially those with crescents on histopathology.

Mycophenolate mofetil

Mycophenolate mofetil (MMF) is an anti-proliferative agent that is being increasingly used for treating patients with various forms of immune-mediated renal disease. In idiopathic MPGN, preliminary studies suggest that the combination of MMF and corticosteroid can, in the short term, reduce proteinuria and may preserve renal function [28, 29].

Cyclosporine

The efficacy of cyclosporine was tested in a recent trial involving 18 patients with refractory MPGN who also received small doses of prednisolone (0.15 mg/kg/day). Long-term reductions in proteinuria with preservation of renal function was observed in 17 of the patients, suggesting that cyclosporine can be considered in corticosteroid-resistant primary MPGN [30].

Plasma exchange

Plasma exchange has not been studied in a controlled manner. Success has been reported for the treatment of type II MPGN recurring after transplantation [13]; however, there are no long-term data on graft survival. There is some evidence that patients with Factor H deficiency may benefit from plasma exchange [31, 32].

Antiplatelet therapy

The rationale for antiplatelet therapy is that platelet consumption is increased in MPGN and that platelets may play a role in glomerular injury. The combination of aspirin (975 mg/day) and dipyridamole (275 mg/day) was found to have useful effects on renal function in a randomized double-blind placebo-controlled trial in 40 patients with idiopathic MPGN, including children [33]. However, these same authors retracted this conclusion in a subsequent study [34]. Another study reported a clinically significant reduction in proteinuria in patients treated with aspirin (500 mg/day) and dipyridamole (75 mg/day), but very little change in the GFR [35].

Possible future therapeutic strategies

Anti-inflammatory agents, such as corticosteroids, may have useful effects in treating MPGN, but their non-specific nature and adverse effects mean that a high price is paid for any beneficial effect on the renal lesion. More selective forms of anti-inflammatory therapy are becoming available [36], and these are aimed at individual pro-inflammatory mediators, such as tumor necrosis factor (which can be neutralized with specific antibodies) or interleukin (IL)-1 (the effect of which can be abrogated using soluble IL-1 receptor antagonist). Therapies which inhibit complement activation by both the classical and alternative pathways, such as eculizumab (an anti-C5 antibody designed to decrease C5a-mediated glomerular damage), should also be considered. Another attractive strategy which merits study is to reduce C3NeF using
Fig. 4 Treatment options of idiopathic MPGN depending on the level of proteinuria and renal dysfunction. ARB Angiotensin receptor blocking agent, ACEI angiotensin-converting enzyme inhibitor, MMF mycophenolate mofetil

Fig. 5 Management of hepatitis C-associated cryoglobulinemic MPGN. GFR Glomerular filtration rate, * as described in the opposite box under proteinuria <3g/d
rituximab (a chimeric IgG1 monoclonal antibody that specifically targets the CD20 surface antigen expressed on B lymphocytes). The improved understanding of the pathogenesis of MPGN is likely to result in the identification of future agents that will modify the intense cellular proliferation and immune deposition seen in this glomerular disease.

In summary, the therapeutic options in idiopathic MPGN depend on the level of proteinuria and kidney failure, as shown in Fig. 4. Management of patients with hepatitis C-associated cryoglobulinemic MPGN is also dictated by the severity of proteinuria and renal dysfunction [37], necessitating two distinct approaches, as shown in Fig. 5. In all patients, the empiric use of angiotensin-converting enzyme inhibitors or angiotensin II receptor antagonists, as drugs of first choice to treat hypertension and decrease proteinuria, is a common practice that may retard the progression of renal disease. Treatment of recurrent MPGN in renal transplant is uncertain, but plasma exchange may be useful in patients whose renal function deteriorates rapidly [13], whereas rituximab seems to be an effective treatment in the presence of cryoglobulinemia [2, 37]. Lastly, improvement in renal outcomes for patients with MPGN largely relies on the evaluation of more selective agents in future studies.

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Questions:

(Answers appear following the question list)

1. Which of the following statements regarding the prevalence of MPGN is true?
   a) MPGN is more common in Europe and North America
   b) MPGN usually affects children and young adults with a male to female ratio of 3:1
   c) MPGN accounts for approximately 5% of the primary renal causes of nephrotic syndrome in children and adults
   d) MPGN is a common cause of ESRD in children

2. Which of the following statements regarding C3NeF is correct?
   a) C3NeF is an IgM autoantibody
   b) C3NeF can be found in healthy subjects
   c) C3NeF prevents the inactivation of C3 convertase (C3bBb) by factor I
   d) C3NeF is a surrogate marker of disease progression

3. Which of the following histological features is the most characteristic of MPGN?
   a) Duplication of the glomerular basement membrane
   b) Crescent formation
   c) Effacement of the podocyte foot process
   d) Positive staining for C4 along the glomerular capillaries

4. Which of the following statements regarding the recurrence of MPGN in kidney transplantation is true?
   a) More common in type I MPGN
   b) Is usually benign
   c) Nephrotic-range proteinuria is a risk factor for recurrence
   d) Can morphologically resemble transplant glomerulopathy

5. Which of the following regarding the treatment of idiopathic MPGN is true?
   a) The rarity of MPGN makes the randomized controlled study design easy to implement
   b) There is a good evidence that the combination of aspirin and dipyridamole slows the progression of the disease
   c) Steroid therapy has been proven effective for children but not for adults
   d) Cyclosporine is generally recommended in patients with rapidly progressive renal failure

References

1. Kher V, Gulati S (2005) Mesangiocapillary glomerulonephritis. In: Davidson AM (ed) Oxford textbook of clinical nephrology, 3rd edn. Oxford University Press, Oxford, pp 523–545
2. Alpers CE, Smith KD (2008) Cryoglobulinemia and renal disease. Curr Opin Nephrol Hypertens 17:243–249
3. Orth SR, Ritz E (1998) The nephrotic syndrome. N Engl J Med 338:1202–1211
4. Ozkaya N, Cakar N, Ekim M, Kara N, Akkök N, Yalçıntayla F (2004) Primary nephrotic syndrome during childhood in Turkey. Pediatr Int 46:436–438
5. Asinobi AO, Gbadegesin RA, Adeyemo AA, Akang EE, Arowolo FA, Abiola OA, Osnusi K (1999) The predominance of membranoproliferative glomerulonephritis in childhood nephrotic syndrome in Ibadan, Nigeria. West Afr J Med 18:203–206
6. Warady BA, Hébert D, Sullivan EK, Alexander SR, Tejani A (1997) Renal transplantation, chronic dialysis, and chronic renal insufficiency in children and adolescents. The 1995 annual report of the North American pediatric renal transplantation cooperative study. Pediatr Nephrol 11:49–64
7. Kai H, Shimizu Y, Hagiwara M, Yoh K, Hirayama K, Yamagata K, Ohba S, Nagata M, Koyama A (2006) Post-MRSA infection glomerulonephritis with marked Staphylococcus aureus cell envelope antigen deposition in glomeruli. J Nephrol 19:215–219
8. Walker PD (2007) Dense deposit disease: new insights. Curr Opin Nephrol Hypertens 16:204–212
25. Tarshish P, Bernstein J, Tobin JN, Edelmann CM Jr (1992) Treatment of mesangiocapillary glomerulonephritis with alternate-day prednisone—a report of the International Study of Kidney Disease in Children. Pediatr Nephrol 6:123–130

26. Somers M, Kertesz S, Rosen S, Herrin J, Colvin R, Palacios de Carreña N, Kim M (1995) Non-nephrotic children with membranoproliferative glomerulonephritis: are steroids indicated? Pediatr Nephrol 9:140–144

27. Faedda R, Satta A, Tanda F, Pirisi M, Bartoli E (1994) Immunosuppressive treatment of membranoproliferative glomerulonephritis. Nephron 67:59–65

28. De S, Al-Nabhani D, Thorner P, Catran D, Piscione TD, Licht C (2009) Remission of resistant MPGN type I with mycophenolate mofetil and steroids. Pediatr Nephrol 24:597–600

29. Jones G, Juszczyk M, Kingdon E, Harboe M, Sweny P, Burns A (2004) Treatment of idiopathic membranoproliferative glomerulonephritis with mycophenolate mofetil and steroids. Nephrol Dial Transplant 19:3160–3164

30. Bagheri N, Nemati E, Rahbar K, Nobakht A, Einollahi B, Taheri S (2008) Cyclosporine in the treatment of membranoproliferative glomerulonephritis. Arch Iran Med 11:26–29

31. Licht C, Heinen S, Jöszti M, Löschtmann I, Saunders RE, Perkins SJ, Waldherr R, Skerka C, Kirschfink M, Hoppe B, Zipfel PF (2006) Deletion of Lys224 in regulatory domain 4 of Factor H reveals a novel pathomechanism for dense deposit disease (MPGN II). Kidney Int 70:42–50

32. Jansen JH, Högåsen K, Harboe M, Hovig T (1998) In situ complement activation in porcine membranoproliferative glomerulonephritis type II. Kidney Int 53:331–349

33. Donadio JV Jr, Anderson CF, Mitchell JC 3rd, Holley KE, Ilstrup DM, Fuster V, Chesebro JH (1984) Membranoproliferative glomerulonephritis. A prospective clinical trial of platelet-inhibitor therapy. N Engl J Med 310:1421–1426

34. Donadio JV Jr, Orford KP (1989) Reassessment of treatment results in membranoproliferative glomerulonephritis, with emphasis on life-table analysis. Am J Kidney Dis 14:445–451

35. Zäuner I, Böhler J, Braun N, Grupp C, Heering P, Schollmeyer P (1994) Effect of aspirin and dipyridamole on proteinuria in idiopathic membranoproliferative glomerulonephritis: a multicentre prospective clinical trial. Collaborative Glomerulonephritis Therapy Study Group (CGTS). Nephrol Dial Transplant 9:619–622

36. Smith RJ, Alexander J, Barlow PN, Botto M, Cassavant TL, Cook HT, de Córdoba SR, Hageman GS, Jokiranta TS, Kimberling WJ, Lambis JD, Lanning LD, Levisodiots V, Licht C, Lutz HU, Meri S, Pickering MC, Quigg RJ, Raps AL, Salant DJ, Sethi S, Thurman JM, Tully HF, Tully SP, van der Vlag J, Walker PD, Würzner R, Zipfel PF (2007) Dense deposit disease focus group, new therapy options in dense deposit disease. Kidney Int 71:936–942

37. Fabrizi F, Lunghi G, Messa P, Martin P (2008) Therapy of hepatitis C virus-associated glomerulonephritis: current approaches. J Nephrol 21:813–825

Answers:

1) C
2) B
3) A
4) D
5) C