A systems biology view of the development of new antimicrobial therapies

Introduction

Antimicrobial resistance has received an increased level of attention in the recent years due its alarming intensification and spread worldwide. Despite acknowledgment of the acceleration of antimicrobial resistance, efforts dedicated to new antibiotics discovery and development or optimization of current therapies have declined in the past two decades. Following the report on antibiotic resistance threats in the US in 2013 from the CDC and the distressing report of the WHO on the post-antibiotic era in 2014, controlling the development and spread of antibiotic resistance became a top US policy. The effect of antibiotics can now be monitored at the various cellular levels (genome, transcriptome, proteome, metabolome) and in a dynamic way (fluxome). Very exciting research on the mechanism of bacterial cell death induced by antimicrobial action is currently using systems biology approaches combining metabolomics-based strategy, network modeling, and analysis with biochemical characterizations. Pioneering work from groups like James Collins’ at MIT led to a more detailed understanding of these mechanisms.

Their work presents an integrated view of the consequences of antibiotics treatment on the cell: impacts on the central metabolism (TCA cycle, oxidative phosphorylation, etc.), the membrane potential, protein synthesis and quality control, production of reactive oxygen species (ROS), and stress responses have been uncovered. It appears that various classes of bactericidal compounds – acting on different targets – induce cell death through a shared mechanism involving the formation of hydroxyl radicals. Antibiotic treatment triggers a chain of events in the cell that leads to an increase in NADH oxidation with stimulation of the TCA cycle, overproduction of superoxide anion at the electron transport chain level that can damage Fe-S clusters, ultimately releasing ferrous iron and increasing the production of hydroxyl radicals through the Fenton reaction. It has been well established that the bacterial physiological homeostasis is a fine-tuned process with tightly controlled and regulated components that are highly interconnected. With the expansion of the use of metabolomics techniques, bacterial physiology and adaptation to stress has been investigated in more detail using integrated approaches. However, only a few studies integrate a systemic approach to elucidate the global effect of antibiotics on bacteria. It can be anticipated that such studies will flourish in the coming years.

The use of synthetic biology and in silico modeling adds a predictive aspect that allows identification of potential targets to be modulated. The establishment of the participation of ROS during antibiotic-induced bacterial cell death paved the way towards improvement strategies of established therapies. Network modeling and predictions of ROS formation upon genetic or chemical perturbations of the cell led to the identification of potential protein targets (enzymes involved in the central metabolism such as succinate dehydrogenase). Experimental validation of the synergistic effects of antibiotic treatment with chemical inhibition of the target is a promising lead towards the development of adjuvant therapies. The
predictive value of metabolic models should be more broadly used to facilitate the establishment of chemical library screening aiming at discovering compounds that enhance the killing efficacy of current antibacterial agents. This approach already led to the uncovering of potential therapeutic strategies that does not require new antibiotics discovery.12,13

A new line of reports inspired by this type of approach is increasingly emerging where antimicrobial action is monitored and perturbations in the systems are attempted to enhance the efficacy of already established therapies. Strong evidence suggests that susceptibility to antimicrobial compounds is highly correlated to the metabolic state of the cell, and particular metabolic profiles are associated with certain antibiotic resistances. It has been suggested that alteration or reversal of this resistant-prone metabolic state could improve antibiotic efficiency.14 This has proven to be particularly successful in the case of bacterial persisters. Reactivation of bacterial metabolism (switch from dormant to metabolically active bacteria) by a single metabolite restores the efficacy of the antibiotics.12,13,15 Similarly, discovery of the involvement of oxidative stress in the mechanism of action of bactericides has led to the use of ROS stimulating molecules as co-drugs to enhance the killing potential of antibiotics.12,13 Advances in systems biology that combine modelling with experimentation and advances in techniques that allow dynamic physiological processes to be followed have shifted the field of bacteriology. The combined use of antibiotics and metabolic reprogramming through metabolites is a promising approach in the development of improved therapies.

On a different front, the recent discovery of a new antibiotic compound (teixobactin) also brings some hope in the fight against bacterial resistance.16 In addition to being the first new class of antibiotics discovered in the past 30 years, this compound acts by binding to the glycolipid component of the cell wall. Other antibiotics mediate their effects through interactions with proteins; bacteria might be less efficient at developing resistance to this mode of action. More interestingly, this new compound has been discovered in cellular communities that are almost impossible to culture under traditional microbiological conditions in the laboratory. Our knowledge of the bacterial world is indeed limited by our ability to study the microorganisms in laboratory-compatible conditions. Many specimens stemming from the soil do not thrive in petri dishes and first need to be “domesticated” therefore limiting their use by scientists. It is estimated that only 1% of soil bacteria have been cultured in laboratory conditions. The development of microfluidics devices such as the iChip17 is a very exciting technological advancement allowing researchers to culture bacteria in their “native habitat”. It clearly opens new avenues in microbiological studies and offers promise in the discovery of new molecules.

Fighting antibacterial resistance may never reach a true endpoint and will remain a permanent struggle since bacteria can evolve and adapt very quickly to environmental stresses. The systems biology approach has however proven extremely efficient at understanding and deciphering the complexity of the physiological alterations occurring in bacteria during exposure to antibiotics. Discovery of new antibiotics combined with network modulation of the metabolic status of the cell could constitute a new and sustainable battle plan. These progresses should encourage the scientific community to integrate this global vision in their research.

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**Conflict of interest**

The author declares no conflict of interest.

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