Video Article

Assessment of Pulmonary Capillary Blood Volume, Membrane Diffusing Capacity, and Intrapulmonary Arteriovenous Anastomoses During Exercise

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Abstract

Exercise is a stress to the pulmonary vasculature. With incremental exercise, the pulmonary diffusing capacity (DL_{CO}) must increase to meet the increased oxygen demand; otherwise, a diffusion limitation may occur. The increase in DL_{CO} with exercise is due to increased capillary blood volume (Vc) and membrane diffusing capacity (Dm). Vc and Dm increase secondary to the recruitment and distension of pulmonary capillaries, increasing the surface area for gas exchange and decreasing pulmonary vascular resistance, thereby attenuating the increase in pulmonary arterial pressure. At the same time, the recruitment of intrapulmonary arteriovenous anastomoses (IPAVA) during exercise may contribute to gas exchange impairment and/or prevent large increases in pulmonary artery pressure.

We describe two techniques to evaluate pulmonary diffusion and circulation at rest and during exercise. The first technique uses multiple-breath holds to determine Vc and Dm at rest and during exercise. Additionally, echocardiography with intravenous agitated saline contrast is used to assess IPAVAs recruitment.

Representative data showed that the DL_{CO}, Vc, and Dm increased with exercise intensity. Echocardiographic data showed no IPAVA recruitment at rest, while contrast bubbles were seen in the left ventricle with exercise, suggesting exercise-induced IPAVA recruitment.

The evaluation of pulmonary capillary blood volume, membrane diffusing capacity, and IPAVA recruitment using echocardiographic methods is useful to characterize the ability of the lung vasculature to adapt to the stress of exercise in health as well as in diseased groups, such as those with pulmonary arterial hypertension and chronic obstructive pulmonary disease.

Video Link

The video component of this article can be found at https://www.jove.com/video/54949/

Introduction

During exercise, cardiac output can increase up to six-fold above resting values¹. Given that the lungs are the only organ to receive 100% of the cardiac output, exercise presents a considerable stress to the pulmonary system. With incremental exercise, pulmonary diffusing capacity (DL_{CO}) must increase to meet the increased oxygen demand². From rest to peak exercise, DL_{CO} can increase to up to 150% of resting values without reaching an upper limit with respect to cardiac output³,⁴,⁵. The increase in diffusing capacity occurs as a result of increases in membrane diffusing capacity (Dm) and capillary blood volume (Vc), secondary to the recruitment and distension of pulmonary capillaries⁶.

Roughton and Forster (1957) developed a technique to partition Dm and Vc⁷ by modulating the fraction of inspired oxygen (F_O2) DL_{CO} breath holds to determine Vc and Dm at rest and during exercise. Additionally, echocardiography with intravenous agitated saline contrast is used to assess IPAVAs recruitment.

Representative data showed that the DL_{CO}, Vc, and Dm increased with exercise intensity. Echocardiographic data showed no IPAVA recruitment at rest, while contrast bubbles were seen in the left ventricle with exercise, suggesting exercise-induced IPAVA recruitment.

The evaluation of pulmonary capillary blood volume, membrane diffusing capacity, and IPAVA recruitment using echocardiographic methods is useful to characterize the ability of the lung vasculature to adapt to the stress of exercise in health as well as in diseased groups, such as those with pulmonary arterial hypertension and chronic obstructive pulmonary disease.
The increases in Vc and Dm during exercise are accomplished by an increase in pulmonary artery pressure, which results in the recruitment and distension of pulmonary capillaries previously hypo-perfused at rest. This results in an increase in the cross-sectional area of the pulmonary capillary network, thereby decreasing pulmonary vascular resistance and attenuating the increase in pulmonary artery pressure.

Studies using agitated saline contrast echocardiography have shown evidence of intrapulmonary arteriovenous anastomoses (IPAVA) recruitment during exercise. The significance of IPAVA recruitment is not yet clear, and while some studies suggest that they may contribute to gas exchange impairment and may serve to unload the right ventricle, the topic remains controversial. Further, while the exact mechanism of IPAVA recruitment is not known, we have found that increasing cardiac output, as well as exogenous dopamine, causes IPAVA recruitment at rest. An acutely-increasing pulmonary artery pressure or dopamine blockade does not appear to significantly affect IPAVA recruitment during exercise. There is speculation that these larger-diameter IPAVA vessels may help to protect the pulmonary capillaries from the large increases in pulmonary artery pressure by reducing pulmonary vascular resistance.

When combined with the evaluation of Vc and Dm, agitated saline contrast echocardiography is a valuable tool to examine the adaptation of the pulmonary circulation to the stress of exercise.

### Protocol

This protocol follows the guidelines of the human research ethics board at the University of Alberta and conforms to the standards set by the latest revision of the Declaration of Helsinki.

#### 1. Graded Exercise Test (VO_{2peak})

1. Obtain written, informed consent from the subject. Have the subject read and answer the questions listed on the Physical Activity Readiness Questionnaire+ (PAR-Q+) to determine their readiness for exercise.
2. Adjust the seat height of the cycle ergometer in accordance to subject preference. Place four electrocardiogram (ECG) electrodes on the back of the patient according to standard 3-lead ECG placement, with modified limb leads to measure the heart rate (HR).
3. Insert the mouthpiece into the subject's mouth to measure the exhaled gas and ventilation throughout the test using a metabolic measurement system.
   - **NOTE:** The metabolic system will measure real-time oxygen consumption (VO_{2}), carbon dioxide production (VCO_{2}), ventilation (V\text{E}), heart rate (HR), and end tidal CO_{2} (P_{ET}CO_{2}).
4. Following 2 min of collection of baseline data, instruct the subject to start cycling with an initial workload of 50 watt, to maintain a consistent cadence of ≥80 RPM. Increase the workload in 25 W steps every 2 min, until the subject reaches volitional exhaustion or requests to stop the test.

#### 2. Multiple Fraction of Inspired Oxygen (F\text{I}_{O_{2}}) Diffusing Capacity (DL_{CO}) Method

1. Calculate the workloads corresponding to 30%, 50%, 70%, and 90% of the VO_{2peak} using the peak VO_{2} obtained in the graded exercise test. At least 48 h after the graded exercise test, have the subject return to the laboratory for DLco maneuvers.
2. Do not exceed 12 DLco tests per day, as carboxyhemoglobin (COHb) build-up can occur with repeated testing. Therefore, perform testing on multiple days based on the number of exercise workloads to be conducted and the quality of the DLco data.
3. Prepare pre-breathing gases by attaching a tank of 100% O\text{2} and a tank of medical-grade air (21% O\text{2} and 79% N\text{2}) to an air blender system. Fill two 60 L non-diffusing Douglas bags, one containing 40% O\text{2}, and one containing 60% O\text{2}, using the air blender system.
4. Set up two large-bore, three-way stopcock valves that will allow for the modulation of inhaled gas mixtures. These will be referred to as the "pre-breath valves."
5. Connect the Douglas bags to the valve system using flexible, non-compressible tubing. Connect the valve system to a two-way, T-shaped non-breathing valve connected to the test gas intake assembly of the mass flow sensor of the metabolic measurement system.
6. For resting measurements, have the subject seated upright, with both feet flat on the floor. For exercise trials, ensure that the subject is in a steady state by monitoring HR using the ECG (HR ≥ 3 bpm for steady state). **NOTE:** Steady state may not be reached at 90% of the VO_{2peak}; thus, begin the measurement once the subject has reached the HR equivalent to 90% of the VO_{2peak} on the graded exercise test.
7. Collect a single drop of capillary blood via a finger prick and analyze it for hemoglobin concentration. Then, adjust all subsequent DL_{CO} for [Hb] using the following equation:
   
   \[ DL_{CO_{adj}} = DL_{CO} \times \frac{10.22 + [Hb]}{1.7 \times [Hb]} \]
8. Select an F\text{I}_{O_{2}} (21%, 40%, or 60%) at random by switching the pre-breathe valves to the desired orientation. Choose the corresponding F\text{I}_{O_{2}} DL_{CO} gas by turning the DL_{CO} gas valve selector (see Figure 1C).
9. Instruct the subject to affix the nose clips and to breathe normally into the mouthpiece for five breaths from the Douglas bag corresponding to the respective F\text{I}_{O_{2}}.
10. Instruct the subject to expire to residual volume. When the lung volume plateaus at residual volume, have the subject inhale the DL_{CO} gas mixture to total lung capacity and hold their breath for 6 s before exhaling to residual volume.
11. Monitor the methane tracing during the exhalation to ensure that the slope is horizontal, as this indicates that the CO test gas is well equilibrated in the lung. **NOTE:** Alveolar volume (V\text{A}) and breath hold time are calculated automatically and reported by the metabolic measurement system.
12. Ensure that the V\text{E} for each DLco maneuver is within 5% of previous trials. Similarly, breath hold time should be 6 ± 0.3 s. If not, repeat the maneuver.
13. Wait 4 min to allow residual carbon monoxide to wash out, and then repeat steps 2.8 - 2.11 for each remaining F\text{I}_{O_{2}} at rest.
14. At least 48 h later, repeat steps 2.9 - 2.15 during steady state at each exercise intensity (30%, 50%, 70%, and 90% of the VO$_{2\text{peak}}$) for each F$\text{I}_2$. Reduce the workload between the breath holds at 90% of the VO$_{2\text{peak}}$ workload to recover the subject.

15. Wait 2 min between DLco tests during exercise to clear alveolar CO during exercise. Do not exceed 12 DLco tests per day to avoid carboxyhemoglobin (COHb) build-up$^5$.

3. Calculating Pulmonary Capillary Blood Volume and Membrane Diffusing Capacity

1. Calculate the alveolar partial pressure of O$_2$ (P$\text{A}_2$O$_2$) using the following equation

\[ P_{\text{A}_2}\text{O}_2 = F_{\text{I}_2}\text{O}_2(P_{\text{BAR}} - P_{\text{H}_2}\text{O}) - P_a\text{CO}_2 \times \frac{(1-F_{\text{I}_2}\text{O}_2)}{\text{RER}} \]

NOTE: F$\text{I}_2$O$_2$ is the fraction of inspired O$_2$, P$_{\text{BAR}}$ is the atmospheric pressure, P$_{\text{H}_2}$O is the water vapor pressure, P$_a$CO$_2$ is the pressure of arterial CO$_2$, and RER is the respiratory exchange ratio.

2. Estimate the RER and P$_a$CO$_2$ using the measured 30-s average P$_{\text{ET}}$CO$_2$ and RER for the respective exercise intensity from the data obtained in the previous graded exercise test.

3. Calculate $\theta$CO using the following equation$^7$.

\[ \frac{1}{\theta_{\text{CO}}} = 0.0059 \times P_{\text{A}_2}\text{O}_2 + 0.73 \]

4. Graph the relationship between 1/DLco$_{\text{adj}}$ and 1/$\theta$CO for each F$\text{I}_2$O$_2$ and calculate the regression equation.

NOTE: The minimum acceptable r$^2$ value is 0.95, and DLco maneuvers should be repeated when r$^2$ values are outside of this range$^{21}$.

4. Intrapulmonary Arteriovenous Anastomosis Recruitment

1. On a separate day from the DLco data collection, insert a 20-gauge intravenous (IV) catheter into an antecubital vein and attach it to a three-way stopcock via a 6-in IV extension tube for the injection of agitated saline for contrast echocardiography$^{11,17}$. 

![Figure 2: Representative Graph of 1/DLco versus 1/$\theta$CO at Peak Exercise.](https://www.jove.com)
Figure 3: Agitated Saline Contrast Setup. An IV catheter is placed in the antecubital space and is connected to a three-way stopcock via a 6-in extension. Two 10 mL syringes are attached to the stopcock to create the contrast solution, which contains 10 mL of saline and 0.5 mL of room air. Please click here to view a larger version of this figure.

2. Connect two 10 mL syringes to the three-way stopcock. Combine 10 mL of 0.9% sterile saline with 0.5 mL of air, and forcefully agitate it through the three-way stopcock, back and forth between the two syringes, to form fine, suspended bubbles until the sonographer is ready for contrast.

3. Have an experienced sonographer or cardiologist obtain a standard apical four-chamber view of the heart. At rest, have the echocardiographer evaluate the intra-atrial septum and ventricular septum for an intra-cardiac shunt with standard echocardiographic and color Doppler imaging.
   1. If no intra-cardiac shunt is detected, instruct the subject to perform a Valsalva maneuver during the contrast injection to evaluate for a patent foramen ovale (PFO)\(^{11,17}\). Repeat the measurement during non-Valsalva.

4. Inject the contrast while the sonographer maintains the four-chamber view. Record 15 cardiac cycles following the detection of contrast in the right ventricle.

5. Repeat the contrast-enhanced imaging during steady-state exercise at 30%, 50%, and 70% of the VO\(_{2}\)\(_{\text{peak}}\). As steady state cannot be reached at 90% of the VO\(_{2}\)\(_{\text{peak}}\), begin the imaging once the target HR, identified by the HR at 90% of the VO\(_{2}\)\(_{\text{peak}}\) during the graded exercise test, is reached.
   NOTE: The time between exercise intensities depends on the clearance of contrast from both ventricles, ≥ 2 min.

6. Have an echocardiographer who is blinded to experimental conditions interpret the agitated saline contrast echocardiograms according to a previously-described scoring system\(^{17,27}\).
   NOTE: Scoring is based on the maximum number of contrast bubbles visible within the left ventricle (LV) in a single echocardiographic frame, as follows: no contrast bubbles in the LV = 0, ≤3 bubbles = 1, 4 - 12 bubbles = 2, > 12 bubbles = 3.
   NOTE: The appearance of contrast in the left ventricle after five cardiac cycles suggests an IPAVA. An intracardiac shunt is graded by the appearance of contrast in less than five cardiac cycles\(^{17} \)\(^{27}\).

Figure 4: Representative Images for IPAVA Scoring. The scale is 5 cm (solid white line). (A) Pre contrast injection. (B) IPAVA score = 0. (C) IPAVA score = 1. (D) IPAVA score = 3. Please click here to view a larger version of this figure.

Representative Results

The effect of increasing exercise intensity on oxygen consumption, diffusing capacity, pulmonary capillary blood volume, membrane diffusing capacity, and IPAVA score is shown in Table 1. VO\(_{2}\), DL\(_{CO}\), Vc, and Dm increase in response to increasing power output.

Figure 2 shows a representative calculation of Vc and Dm using the multiple F\(_{\text{O}_2}\)-DL\(_{CO}\) technique during exercise. DL\(_{CO}\) decreases with increasing F\(_{\text{O}_2}\), and this relationship is exploited to partition Vc and Dm. Calculating the inverse of the slope of 1/DL\(_{CO}\) versus 1/θ\(_{CO}\) results in the Vc, and the inverse of the y-intercept yields the value for the Dm. As expected, both the Vc and Dm increase during exercise compared to resting values.
The results show that these techniques can be used to assess the pulmonary vasculature response during exercise. The multiple-FIO2 DLCO and agitated saline contrast echocardiography method provides investigators with more insight into the contributions of pulmonary capillary and membrane recruitment to the overall diffusion capacity and could supplement traditional pulmonary function testing in the clinical setting. Failure to increase \( Vc \) or \( Dm \) during exercise would lead to a diffusion limitation and hypoxemia. For example, a low DLCO secondary to a low \( Vc \) would indicate changes to the pulmonary capillaries; similarly, a decreased \( Dm \) would indicate changes to the pulmonary membrane.

**Figure 4** shows representative tracings of four-chamber contrast echocardiographs. With increasing exercise intensity, the IPAVA score increases from 0 (i.e., no evidence of IPAVAs) at rest to 3 at the highest exercise intensity (**Table 1**). Previous work has shown that exercise increases the IPAVA score \(^{11,12,14} \), but there is no consensus as to how these IPAVAs are recruited. There is evidence that IPAVAs can be recruited pharmacologically at rest with dopamine \(^{17,28} \), as well as by increasing cardiac output with dobutamine \(^{17,28} \) and epinephrine \(^{28} \). Inotropes such as dopamine and epinephrine are of particular interest, as they increase endogenously during exercise \(^{29} \). Furthermore, there is some evidence that IPAVA recruitment may be important to exercise hemodynamics, in that the absence of IPAVAs appears to result in greater pulmonary artery pressure, decreased cardiac output, and decreased peak power output \(^{12} \). Thus, this technique may be used in studies examining individuals with pulmonary artery hypertension.

![Figure 1: Multiple FIO2 DLCO Setup.](https://www.jove.com/images/Figure%201%20-%20Multiple%20FIO2%20DLCO%20Setup.png)

**Table 1: Representative Data for One Subject at Rest and During Exercise at 30, 50, 70, and 90% of the VO2peak.** \( VO_2 \), volume of oxygen consumption relative to body mass; \( DL_{CO} \), diffusing capacity for carbon monoxide; \( Vc \), pulmonary capillary blood volume; \( Dm \), membrane diffusing capacity; IPAVA score, scoring of contrast appearance in the left ventricle after five cardiac cycles. Data modified from Tedjasaputra et al. 2016.

|            | Rest | 30% | 50% | 70% | 90% |
|------------|------|-----|-----|-----|-----|
| \( VO_2 \) (mL·kg\(^{-1}\)·min\(^{-1}\)) | 6.0  | 17.0| 28.3| 40.1| 50.9|
| \( DL_{CO} \) (mL·min\(^{-1}\)·mmHg\(^{-1}\)) | 36.6 | 45.1| 48.0| 51.5| 60.0|
| \( Vc \) (mL) | 70.8 | 92.7| 95.2| 105.0| 125.5|
| \( Dm \) (mL·min\(^{-1}\)·mmHg\(^{-1}\)) | 75.7 | 87.7| 96.9| 101.2| 115.0|
| IPAVA Score | 0    | 1   | 1   | 2   | 3   |

**Discussion**

This method enables the evaluation of the pulmonary diffusing capacity and intrapulmonary arteriovenous anastomosis recruitment during exercise.
Critical steps within the protocol

Although the DL\textsubscript{CO} breath hold is relatively simple at rest, breath holding during exercise presents a unique challenge to the subject, as it is counter-intuitive, and subjects have a high drive to breathe during exercise. Thus, a good-quality determination of Vc and Dm relies on the rapport and clear communication between the tester and the subject. The tester’s technical ability can be quantified with the variability of the alveolar volume (± 5% of previous trials) and a breath-hold time (BHT) of 6.0 ± 0.3 s.

Modifications and troubleshooting

At the conclusion of a Vc/Dm measurement, the tester should quickly graph the three DL\textsubscript{CO} maneuvers to determine the best-fit line of the data points; the DL\textsubscript{CO} measured with 21% F\textsubscript{O}2 should always be greater than that with 40%, which should be greater than that with 60%. If not, it is recommended to check if the valve switch corresponds to the correct testing gas. Similarly, check that the pre-breathing bags are filled with the correct F\textsubscript{O}2 gas corresponding to the testing gas (Figure 1B-1D). Caution should be taken when testing a participant who is a smoker, as elevated COHb levels may under estimate DLco.

For the IPAVA recruitment assessment, the position of the subject is critical to ensure high-quality image acquisition. It is possible to replace the upright cycle ergometer with a recumbent cycle ergometer to minimize the movement of the subject. However, recumbent cycle exercise will elicit a different metabolic response for a given work rate, and thus the graded exercise test should be repeated on the recumbent cycle ergometer. Scanning of the upper chest may be uncomfortable to some women; in this case, a female sonographer is recommended. Finally, the recommended exercise protocol is designed for a young, healthy individual; accordingly, the exercise protocol can be modified for a different target population.

Limitations of the technique

The principal limitations of the multiple F\textsubscript{O}2 DL\textsubscript{CO} technique are the skill of the tester and the ability of the subject to follow commands and to remain calm during the breath hold, as Valsalva or Müllerian maneuvers will affect the measurements. Secondly, the number of breath holds in one session should be limited to 12, due to an increase in CO backpressure, which may affect the Vc and Dm measurement\textsuperscript{15,30} and pose a health risk to the subject. Depending on the research design, it may be necessary to complete the testing across multiple sessions to allow for the clearance of CO and to limit participant fatigue. With good participant coaching and good technical ability, we have determined a satisfactory coefficient of variation between trials for DLco, Vc, and Dm to be 7%, 8%, and 15%, respectively.

The multiple F\textsubscript{O}2 DL\textsubscript{CO} technique assumes that the alveolar O\textsubscript{2} is the same as the capillary O\textsubscript{2}, and thus, caution should be exercised when interpreting the data in individuals with known gas exchange impairment.

Agitated saline contrast echocardiographic imaging is limited by the technical ability of the sonographer and the ability of the subject to minimize thoracic movement while exercising. It is also critical that the interpreter of the images be familiar with the scale for scoring IPAVA recruitment according to established procedures (Figure 4)\textsuperscript{12}. The significance of a positive saline contrast echocardiography during exercise remains a topic of debate\textsuperscript{15,16}, and there is some discussion that a positive agitated saline contrast in the left ventricle may be secondary to capillary distention, and not IPAVA recruitment. Ongoing work is attempting to resolve this issue.

Significance of the technique with respect to existing/alternative methods

By utilizing these physiological techniques, it is possible to assess the pulmonary vasculature during exercise in a variety of conditions, including in health, in disease, and in drug interventions. Although the quality relies with the ability of the tester, these skills are easily and quickly acquired with proper mentorship and training. The multiple F\textsubscript{O}2 DL\textsubscript{CO} method is considered the “gold standard” in the measurement of Dm and Vc\textsuperscript{31}. While these measures are not calculated clinically, the values could be used to determine the mechanisms for hypoxemia and exercise intolerance, to predict patient outcomes, and to further characterize diagnosis\textsuperscript{31,33}. Likewise, the agitated saline echocardiography technique is the most widely-used method in determining the recruitment of IPAVAs.

Future applications or directions after mastering this technique

These techniques are applicable for use in a range of experimental conditions and interventions. We demonstrate these techniques during exercise, but they can easily be modified to measure pulmonary vascular responses during a drug infusion, such as dobutamine or dopamine, inotropes known to increase cardiac output\textsuperscript{17}. Furthermore, it is possible to use these techniques in clinical populations, such as in those with heart failure\textsuperscript{34} or chronic obstructive pulmonary disease (COPD), in which the DL\textsubscript{CO} is lower compared to age-matched control subjects\textsuperscript{35}.

Disclosures

The authors declare that they have no competing financial interests.

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