Population variation in brain size of nine-spined sticklebacks (*Pungitius pungitius*) - local adaptation or environmentally induced variation?

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**Abstract**

**Background:** Most evolutionary studies on the size of brains and different parts of the brain have relied on interspecific comparisons, and have uncovered correlations between brain architecture and various ecological, behavioural and life-history traits. Yet, similar intraspecific studies are rare, despite the fact that they could better determine how selection and phenotypic plasticity influence brain architecture. We investigated the variation in brain size and structure in wild-caught nine-spined sticklebacks (*Pungitius pungitius*) from eight populations, representing marine, lake, and pond habitats, and compared them to data from a previous common garden study from a smaller number of populations.

**Results:** Brain size scaled hypo-allometrically with body size, irrespective of population origin, with a common slope of 0.5. Both absolute and relative brain size, as well as relative telencephalon, optic tectum and cerebellum size, differed significantly among the populations. Further, absolute and relative brain sizes were larger in pond than in marine populations, while the telencephalon tended to be larger in marine than in pond populations. These findings are partly incongruent with previous common garden results. A direct comparison between wild and common garden fish from the same populations revealed a habitat-specific effect: pond fish had relatively smaller brains in a controlled environment than in the wild, while marine fish were similar. All brain parts were smaller in the laboratory than in the wild, irrespective of population origin.

**Conclusion:** Our results indicate that variation among populations is large, both in terms of brain size and in the size of separate brain parts in wild nine-spined sticklebacks. However, the incongruence between the wild and common garden patterns suggests that much of the population variation found in the wild may be attributable to environmentally induced phenotypic plasticity. Given that the brain is among the most plastic organs in general, the results emphasize the view that common garden data are required to draw firm evolutionary conclusions from patterns of brain size variability in the wild.

**Background**

During the past few decades, studies on diverse taxa have demonstrated that both absolute and relative brain size, as well as absolute and relative sizes of different brain parts, are highly variable and correlate with several environmental factors [mammals e.g. [1-3], birds e.g. [4,5] and fishes e.g. [6,7]]. Most of these studies, which form the basis of our current knowledge about brain size evolution, have used correlative approaches at the interspecific level. However, several recent studies have investigated differences in brain architecture among populations of the same species [8-17]. By using inter-population comparisons, microevolutionary processes can be investigated explicitly because most populations are likely to be found in the environment that actually shaped their brains. This is an unlikely situation in the case of comparisons based on species, which might have gone through adaptive divergence after splitting from common ancestor. An additional benefit from inter-population comparisons as compared to interspecific comparisons is that the former allow adopting approaches to separate the relative roles of genetic drift and natural selection on observed differentiation [e.g. [18,19]].
Further, even fewer studies have compared brains of individuals from different populations reared under standardized settings to exclude the possible effects of phenotypic plasticity [but see: [10,12]]. This is surprising considering the fact that phenotypic plasticity in overall brain size, in addition to the size of different brain regions, has often been demonstrated [e.g. seasonality: [20,21]; spatial learning: [22,23], environmental heterogeneity: [24,25]]. Moreover, how this plasticity might influence population and species comparisons in terms of neural architecture has yet to be explored. Therefore, direct comparisons of patterns based on data collected from wild populations with those based on data from standardized common garden settings are needed to establish if any evolutionary inferences can be made from wild collected data in such a highly plastic organ as the brain.

The nine-spined stickleback (*Pungitius pungitius*) is an excellent model species for intraspecific comparative studies and exploring adaptive divergence. It occupies markedly different habitats, ranging from marine environments through large lakes to isolated ponds wherein they are often the only fish species present [e.g. [26]]. Hence, large differences can be found both in biotic (e.g. diversity of prey, competitors and predators) and abiotic (e.g. habitat structure) habitat components. These differences are expected to impose different selection pressures on complex behaviours and memory, and thus, also on the neural architecture. This is especially true in light of the high energetic costs of developing and maintaining large brains [27].

Brain size scales allometrically with body size, both on ontogenetic and on evolutionary scales [e.g. [28-32]]. The slope of the allometric relationship between brain size to body size (both variables plotted on a logarithmic scale) is higher in prenatal than adult stages in mammals [28]. Furthermore, the slope of this relationship tends to be steeper at higher taxonomic ranks [ca. 0.75 across mammalian orders; e.g. [29,30]] compared to closely related species, or in intraspecific comparisons (ca. 0.2-0.5; [31,32]). Although some intraspecific studies in brain-body size allometry exist [e.g. [33]], only very few investigations have been conducted within a single vertebrate species, perhaps due to a lack of sufficient within-species size variation among adults. Since nine-spined sticklebacks living in ponds have repeatedly evolved into giants [34,35], the species (representing tenfold body weight differences between adult individuals) also provides an excellent model to study intraspecific brain-body size allometry.

Our aim was to explore population divergence in brain size and in the size of different parts of the brain (viz. telencephalon, optic tectum, cerebellum, hypothalamus) in wild-caught nine-spined sticklebacks from different habitats, and to compare the observed patterns with previously reported common garden results [12]. We sampled fish from eight Fennoscandian populations (Figure 1) originating from three habitat types (viz. marine, lake and pond environments) to test (i) for differences in the size of the brain and different brain parts among wild populations, and (ii) whether observed differences were habitat specific. Furthermore, to establish whether data collected from the wild can be used for evolutionary inference, (iii) we tested whether data collected from the wild and common garden experiments for fish originating from the same populations are concurrent, and if not, (iv) whether observed differences are population- or habitat-specific. We expected that the higher biotic and abiotic variability of marine and lake environments as compared to pond environments have selected for relatively large brains. We also expected to find habitat-dependent differences in brain parts important in perception, learning and spatial memory, and that the stimulus-poor laboratory environment would reduce the brains of common garden fish compared to...
their wild conspecifics. We also explored the brain-body size allometric relationship in fish from different populations, expecting hypoallometry with a relatively shallow slope (<0.5).

Results

Variation in absolute brain size

Dissected brains were fixed and photographed under standardized conditions. Absolute brain size was estimated from measurements taken from digital photographs (dorsal, lateral and ventral views) by using the ellipsoid model [12,36]. General Linear Model (GLM) results revealed a significant population effect in absolute brain size ($F_{7, 112} = 153.68, P < 0.001$). Average brain sizes of marine and lake fish were similar, but smaller than those of pond fish, the latter also being highly variable (Figure 2). General Linear Mixed Model (GLMM) analyses revealed a significant habitat ($F_{1, 5} = 11.84, P = 0.018$) and non-significant population within habitat effect ($Z = 1.55, P = 0.12$). Pond fish had brains almost twice as large as marine fish (Least Squares [LS] mean ± Standard Error [SE]: marine = 18.59 ± 3.43 mm$^3$; pond = 34.22 ± 2.97 mm$^3$).

Brain size co-varied with body weight, but independently of population origin and standard length (GLM; population: $F_{7, 96} = 1.50, P = 0.18$; log body weight: $F_{1, 96} = 7.63, P = 0.007$; log standard length: $F_{7, 96} = 0.52, P = 0.47$; population × log body weight: $F_{7, 96} = 0.79, P = 0.60$; population × log standard length: $F_{7, 96} = 1.48, P = 0.18$). The log brain size - log body weight regression revealed hypoallometry (i.e. both $\beta = 0$ and $\beta = 1$ were rejected: $R^2 = 0.88, \beta = 0.50, SE[\beta] = 0.02, P < 0.001$; Figure 3), indicating that brain size increased at half the rate of body size.

Figure 2 Population variation in absolute brain size in nine-spined sticklebacks. Means ± SE are shown. For population abbreviations, see Fig. 1.

Dissected brains were fixed and photographed under standardized conditions. Absolute brain size was estimated from measurements taken from digital photographs (dorsal, lateral and ventral views) by using the ellipsoid model [12,36]. We did not consider absolute size, but corrected our estimates with body length, body weight and absolute brain size. The multivariate GLM revealed a significant population effect both at the habitat (GLMM; habitat: $F_{1, 6.89} = 10.38, P = 0.015$; log standard length: $F_{1, 57.02} = 0.27, P = 0.54$; log body weight: $F_{1, 93.48} = 28.82, P < 0.001$; population [habitat]; $Z = 0.95, P = 0.34$), and population level (GLM; population: $F_{7, 108} = 10.79, P < 0.001$; log standard length: $F_{1, 110} = 0.02, P = 0.89$; log body weight: $F_{1, 110} = 30.14, P < 0.001$). Pond (and the single lake) populations had larger relative brain sizes than marine populations (Figure 4a).

The sizes of different brain parts (telencephalon, optic tectum, cerebellum, hypothalamus) were also estimated from the digital photographs using the ellipsoid model [12,36]. Since we detected no significant population-specific relationship, only the common slope (solid line; $\beta = 0.50$) is shown. The dashed line denotes isometry. For population abbreviations, see Fig. 1.

Figure 3 Allometric relationship between brain size and body size in nine-spined sticklebacks. Since we detected no significant population-specific relationship, only the common slope (solid line; $\beta = 0.50$) is shown. The dashed line denotes isometry. For population abbreviations, see Fig. 1.

Variation in absolute brain size

To study variation in relative brain size, we corrected the absolute brain size estimates to body length and body weight. We detected significant differences in relative brain size both at the habitat (GLMM; habitat: $F_{1, 6.89} = 10.38, P = 0.015$; log standard length: $F_{1, 57.02} = 0.27, P = 0.54$; log body weight: $F_{1, 93.48} = 28.82, P < 0.001$; population [habitat]; $Z = 0.95, P = 0.34$), and population level (GLM; population: $F_{7, 108} = 10.79, P < 0.001$; log standard length: $F_{1, 110} = 0.02, P = 0.89$; log body weight: $F_{1, 110} = 30.14, P < 0.001$). Pond (and the single lake) populations had larger relative brain sizes than marine populations (Figure 4a).

Figure 4a.b.c.d Variation in relative brain size and brain part size. To study variation in relative brain size, we corrected the absolute brain size estimates to body length and body weight. We detected significant differences in relative brain size both at the habitat (GLMM; habitat: $F_{1, 6.89} = 10.38, P = 0.015$; log standard length: $F_{1, 57.02} = 0.27, P = 0.54$; log body weight: $F_{1, 93.48} = 28.82, P < 0.001$; population [habitat]; $Z = 0.95, P = 0.34$), and population level (GLM; population: $F_{7, 108} = 10.79, P < 0.001$; log standard length: $F_{1, 110} = 0.02, P = 0.89$; log body weight: $F_{1, 110} = 30.14, P < 0.001$). Pond (and the single lake) populations had larger relative brain sizes than marine populations (Figure 4a).

The sizes of different brain parts (telencephalon, optic tectum, cerebellum, hypothalamus) were also estimated from the digital photographs using the ellipsoid model [12,36]. We did not consider absolute size, but corrected our estimates with body length, body weight and absolute brain size. The multivariate GLM revealed a significant population effect on the relative sizes of different brain parts (population: Wilks’ $\lambda_{28, 380} = 0.53, P < 0.001$; log body weight: Wilks’ $\lambda_{4, 105} = 0.91, P = 0.043$; log standard length: Wilks’ $\lambda_{4, 105} = 0.95, P = 0.024$; log brain volume: Wilks’ $\lambda_{4, 105} = 0.15, P < 0.001$). Subsequent univariate tests indicated significant population differences in relative telencephalon ($F_{7, 108} = 3.53, P = 0.002$), optic tectum ($F_{7, 108} = 2.81, P = 0.010$), and cerebellum ($F_{7, 108} = 2.59, P = 0.016$) sizes, but not in hypothalamus ($F_{7, 108} = 1.47, P = 0.18$) size (Figure 4b-d). Our GLMMs revealed that neither optic tectum (habitat: $F_{1, 10.17} = 0.44, P = 0.52$; log standard length: $F_{1, 59.62} = 0.26, P = 0.61$; log body weight: $F_{1, 88.93} = 0.50, P = 0.48$; log brain volume: $F_{1, 99.99} = 262.72, P < 0.001$).
showed significant habitat-dependency in their relative size. However, in the case of telencephalon, the habitat effect approached significance (habitat: $F_{1, 9.08} = 3.95, P = 0.078$; log standard length: $F_{1, 61.39} = 5.75, P = 0.018$; log body weight: $F_{1, 113.95} = 42.12, P < 0.001$; population[habitat]: $Z = 1.05, P = 0.29$). Marine fish tended to have larger telencephalans than pond fish, while no systematic trend could be observed in the other brain parts (Figure 4b-d).

Comparison of wild and common garden brains

We compared relative brain size and relative brain part size (see above) of fish from two marine (Helsinki, Baltic Sea and Levin Navolok Bay, White Sea; Figure 1) and two pond (Bynästjärnen, Sweden and Pyöreälampi, Finland; Figure 1) populations to data from the same populations reared in a common garden experiment [12]. Fish origin (i.e. wild-caught vs. common garden) had a habitat specific effect on relative brain size (GLMM; origin: $F_{1, 110} = 70.99, P < 0.001$; habitat: $F_{1, 2.49} = 0.49$, origin × habitat: $F_{1, 113.83} = 38.36, P < 0.001$; log standard length: $F_{1, 105.99} = 5.75, P = 0.018$; log body weight: $F_{1, 113.95} = 42.12, P < 0.001$; population[habitat]: $Z = 0.84, P = 0.40$). The population-level GLM supported this result. It revealed a population-specific origin effect (origin: $F_{1, 110} = 70.41, P < 0.001$; population: $F_{3, 110} = 5.55, P = 0.003$; origin × population: $F_{3, 110} = 14.51, P < 0.001$; log standard length: $F_{1, 110} = 9.56, P = 0.003$; log body weight: $F_{1, 110} = 46.65, P < 0.001$). Relative brain size was similar for wild-caught and common garden marine fish, whereas pond fish had relatively larger brains in the wild than in the laboratory (Figure 5).

A multivariate GLM revealed significant, simple effects of population and origin on the relative size of brain parts, but no interaction between variables (origin: Wilks’ $\lambda_{4, 106} = 0.74, P < 0.001$; population: Wilks’ $\lambda_{12, 280.7} = 0.68, P < 0.001$; origin × population: Wilks’ $\lambda_{12, 280.7} = 0.91, P = 0.57$; log standard length: Wilks’ $\lambda_{4, 106} = 0.93, P = 0.12$; log body weight: Wilks’ $\lambda_{4, 106} = 0.89, P = 0.012$; log brain volume: Wilks’ $\lambda_{4, 106} = 0.107, P < 0.001$). All brain parts were affected by origin, as revealed by the subsequent univariate tests ($5.91 < F_{1, 109}$).
Wild-caught fish had relatively larger telencephalon, optic tectum, cerebellum and hypothalamus volumes than their common garden reared conspecifics (Figure 6a-d).

Discussion
We showed that there is large variation in absolute brain volume, relative brain volume and relative volume of the telencephalon, optic tectum and cerebellum across wild nine-spined stickleback populations. Brain size patterns in the wild show habitat specificity both in absolute and relative scales: pond fish have larger brains than marine fish. Further, we found a marginally significant trend in the relative telencephalon size: marine fish tend to have larger telencephala than pond fish. The isometric relationship between brain size and body size (slope = 0.5) is in accordance with a previous study on tropical fish [37]. We also found that wild-caught pond fish have larger brains than laboratory-reared pond fish, whereas no differences were observed between wild-caught and laboratory-reared marine conspecifics. The relative sizes of all brain parts were smaller in common garden than in the wild in all populations. These findings indicate that even though large brain size and brain part size variation exist in the wild, both in absolute and relative terms, patterns in nature may differ from those gathered in a standardized common garden and in some cases even in a habitat-dependent way. This strongly suggests that environmental effects on brain development can obscure and confound evolutionary inference based on purely phenotypic data collected from the wild. Hence, our results under-line the importance of not basing evolutionary inference on phenotypic patterns of brain size variation unless the environmental sources of variation have been controlled for - a point reinforced by other studies.
focussing on differentiation in morphological and life history traits [38-40].

We found large habitat-specific population variation in absolute brain size: all marine populations (and a single lake population) had similarly sized brains that were nearly twofold smaller than those of pond fish. Within the pond habitat, there was large variation in average brain size. Although most studies have investigated only relative brain size variation (correcting brain size for variation in body size), absolute brain size can also account for differences in behaviour and/or habitat use. This is evident in comparisons of closely related species [see e.g. [32]], as the brains of these species tend to be more similar than those of distant taxa. Indeed, absolute brain size variation is routinely utilized in studies of primate and human evolution [e.g. [41]]. In general, increased brain size is attributable to an increase in neuron number, and not in neuron size [32]. Further, increases in absolute brain size result in decreased proportional connectivity [32]. Obviously, larger bodies need larger brains to be controlled at a similar level [32]. Hence, it is not surprising that pond populations that have evolved to giants [34,35] have also much larger brains than smaller sized marine or lake populations.

Previous findings have demonstrated a shallower hypoallometric slope at the intraspecific level and among closely related species than across broader taxonomic groups (mammals, intraspecific: 0.2-0.4, broad interspecific: 0.66, [28]; fishes, intraspecific: 0.44, intrafamiliar: 0.5, broad interspecific: 0.66, [37]). In accordance with these results we found a hypoallometric relationship between brain and body size with a slope = 0.5. In mammals, it has been demonstrated that on a broad evolutionary time scale, there has been greater net directional selection on brain size than on body size, while the short-term differentiation in brain vs. body size in closely related mammalian taxa has resulted from directional selection acting mostly on body size with changes in brain sizes being largely correlated responses [32]. Further, Gonzalez-Voyer et al. [42] demonstrated in Tanganyikan cichlids that even strongly correlated traits, such as brain and body size, can evolve independently from each other, and that body size may be under stronger selection than brain size during adaptive radiation. In the case of the nine-spined stickleback system, habitat-dependent body size diversification has been demonstrated [34,35], and body weight differences among these recently differentiated populations can be tenfold. Hence, it seems feasible to suggest that the observed brain size divergence might have been a correlated response to body-size divergence.

To assess body-size-independent brain size trends, we also investigated brain size differences relative to body size. Similarly to results for absolute brain size, and in contrast to our expectations, pond sticklebacks had relatively larger brains than marine sticklebacks. Intraspecific variation in relative brain size and brain architecture also appears to be strongly correlated with different ecological factors and/or life history traits. For example, environmental harshness has been shown to correlate positively with the size and neuron number of the brain region linked with memory storage (hippocampus) in the black-capped chickadee, Poecile atricapillus, a food caching species for which good memory can be essential for survival [14]. Garamszegi & Eens [8] found a positive correlation between song length and repertoire size and both relative and absolute volumes of different song nuclei. By comparing two subspecies of the white-crowned sparrow, the migratory Zonotrichia leucophrys gambelii and non-migratory Z. l. nuttalli, Pravosudov et al. [43] found that migratory subspecies had larger hippocampus and more hippocampal neurons. Habitat-independent, genetically based intraspecific variation in brain architecture has also been found both in wild guppy (Poecilia reticulata) populations reared in common environments [11], and in laboratory lines of the medaka (Oryzias latipes; [10]).

Marine nine-spined sticklebacks are members of a diverse fish fauna, and as such, are faced with numerous predators and interspecific competitors. In contrast, pond fish communities are much simpler. Structural heterogeneity in the pond environment is also much lower than that found in marine environments. These environmental factors are all known to be important in shaping brain evolution. For instance, predation pressure has been shown to affect brain size evolution in Mallorcan bovids [44], diet affected brain size evolution of bats [45], both environmental complexity and social features sculpt the brain architecture of cichlid fish [36], while living in large and socially complex groups is the most accepted hypothesis for the evolution of the extremely large brain of humans [46]. Hence, we expected selective pressures stemming from predation, interspecific competition, and habitat complexity to result in relatively larger brains in marine populations. Moreover, assuming that body size divergence (pond fish > marine/lake fish [34,35]) preceded correlated brain size divergence, we also expected pond fish to have similar or smaller brains, in relative terms, than marine or lake fish. Our previous common garden experiment based on a subset of the populations used here revealed no habitat-dependence in relative brain size [12]. Therefore, the pattern found in the current study (pond fish > marine fish) is highly unlikely to be a result of selection on brain size itself. Further, while we found no habitat-dependence in the common garden setting, strong population differentiation in relative brain size in a habitat-independent
way was detected (selective force unknown; [12]). Therefore, the plasticity resulting in the habitat-dependent wild vs. common garden difference cannot be habitat-specific itself. In a controlled laboratory experiment we found that group rearing had a negative effect on brain development in pond but not in marine fish [13]. Hence, the hypothesis that the aggressive, bold and anti-social pond fish [47,48] have larger relative brain sizes due to ontogenetic phenotypic plasticity as a response to fierce intraspecific competition must be rejected. Another possible explanation for larger relative brains in pond than in marine populations can be found from differences in ontogenetic allometry: pond fish living under negligible predation can become twice as old as marine fish [34], and an ontogenetic change in body vs. brain growth might explain this pattern. However, this issue requires further investigation.

Not only absolute and relative brain size, but also the relative size of different brain parts of nine-spined sticklebacks varied in the wild. Significant population differences were found in the relative sizes of the telencephalon, optic tectum and cerebellum. Further, we found marginally significant ($P < 0.08$) habitat-specificity in the relative size of the telencephalon, with marine fish tending to develop larger telencephala than pond fish. This is in accordance with results from our previous common garden study [12]. The telencephalon is larger in monogamous species, and shows a trend towards a positive correlation with rock size in the habitats in Tanganyikan lake cichlids [36], suggesting that both social and environmental heterogeneity may select for larger telencephalon. However, quite surprisingly, generalist limnetic populations of three-spined sticklebacks (*Gasterosteus aculeatus*) that use plankton as a main food source have larger telencephala than benthic-foraging populations of the same species as measured in samples from the wild [16]. The optic tectum is relatively larger in fish that prey on fish or other fast-moving prey, and clear water fishes develop larger optic tecta than species inhabiting turbid waters [6]. Cerebellum size correlates positively with the number of sympatric species in a fish community, and hypothalamus size is larger in monogamous than polygamous cichlids [36]. However, we did not find habitat specificity in the relative size of the optic tectum or the cerebellum, neither in the present, nor in the previous common garden study [12].

There are some incongruence between the present study and our previous work [12]. Here we found habitat-specific brain size divergence and population divergence in relative optic tectum size that was not seen in the common garden study. Only the habitat-dependence of relative telencephalon size found in the common garden study could be detected in the data from wild fish. A direct comparison between common garden and wild brains from the same populations revealed a habitat-dependent effect: pond (but not marine) fish had relatively larger brains in the wild than in the common garden. Further, the relative size of all brain parts was smaller in the laboratory than in the wild, perhaps due to a stimulus-poor environment during brain development. The most plausible explanation for the differences among common garden and wild data resides on phenotypic/ontogenetic plasticity in brain architecture. The potential for plastic responses to environmental heterogeneity is very high in fish [49-51]. Neurogenesis persists long into adulthood in fish [51-53] and contributes to lifelong development of the brain. Hence, the fact that pond fish can live nearly twice as long as marine or lake fish may result in bias originating from plain ontogenetic plasticity or allometry. Furthermore, local random environmental variation may induce plasticity that could conceal genetic trends. Therefore, common garden studies seem to be of particular importance in studies of brain evolution. For instance, in this study system erroneous evolutionary conclusions could be drawn from the habitat-specificity (implying local adaptation) of relative brain size in the data from the wild given that observed differences cannot be reproduced under common garden conditions (showing that the differences are environmentally induced).

Finally, we showed that relative brain size and brain architecture are different between wild-caught and common garden sticklebacks from the same populations. The negative effect of domestication on brain size is well known both as a result of genetic adaptation and phenotypic plasticity [54,55]. In a recent paper, Burns et al. [56] demonstrated that laboratory rearing caused a significant decrease in the relative brain size of guppies (*P. reticulata*). Interestingly, we found that laboratory rearing had a negative effect on brain size in pond but not in marine nine-spined sticklebacks. The reason for this difference is unknown and warrants further investigations. We also found that all brain parts (corrected for both body and brain size) were smaller in common garden than in nature, a pattern congruent with general expectations. The reason for this can be a phenotypically plastic response to the comparatively stimulus poor laboratory environment.

**Conclusion**

In summary, we found large variation both in absolute and relative brain size, and brain architecture, among nine-spined sticklebacks in the wild. However, the patterns differed markedly from those found previously under standardized common garden settings, being most probably a result of environmental or age effects prevailing in the wild. Further, we found that the
difference between wild or common garden samples can be habitat specific. Considering the extreme plasticity of the fish brain, drawing evolutionary inference from wild-collected material alone can be challenging, and easily misleading. To understand brain size/structure variation in the wild, more intraspecific, common garden based studies, especially those that attempt to separate genetic and environmental contributions to brain development are needed.

**Methods**

**Sampling and husbandry**

Adult fish were collected from eight populations during May and June of 2007. Three habitat types were covered: marine samples came from Helsinki (Baltic Sea, Finland), Bölesviken (Baltic Sea, Sweden) and Levin Navolok (White Sea, Russia); Rytilampi (Finland), Pyöreälampi (Finland) and Bynästjärnen (Sweden) are isolated ponds, and Iso-Portontima (Finland) is a lake (Figure 1). This region started to deglaciate around 8000 years ago [e.g. [57]], thus, the populations are younger than this. Fish were collected with seine nets and minnow traps. After collection, they were moved to the aquaculture facilities of the University of Helsinki. Prior to taking brain measurements, fish were kept in standardized environment for approximately three months: temperature (14°C) and photoperiod (12 h light, 12 h dark) were held constant, and feeding (ad libitum) with bloodworms (Chironomidae sp.) was similar for all population groups. We note that the ca. three months common garden keeping (to standardize body condition, which is highly variable during spring in nature) might have caused some plastic responses induced by the artificial environment. However, this effect is highly unlikely to be profound. The experiment was conducted under license of the Animal Experiment Board in Finland, reference number: STH379A.

Sampled habitat types differed markedly, both in terms of biotic and abiotic aspects. In marine and lake habitats, nine-spined sticklebacks belong to a diverse fish community consisting of a large number of potential fish predators and interspecific competitors. Conversely, ponds lack predatory fish, and interspecific competition is absent (Ryttilampi and Bynästjärnen), or negligible (Pyöreälampi; where a few, small-sized whitefish [Coregonus lavaretus] were recently introduced). While predation by aquatic insects and cannibalism at very early stages might occur in both habitats, there are two facts indicating large differences in predation caused mortality at later-than-fry stages: pond fish (i) show marked reduction in their defensive body armour (pelvic apparatus; [58]) and (ii) have a much longer lifespan than marine fish (6-7 years vs. 3-4 years; [34]). The structural complexity of the marine and large lake environment exceeded that of the study ponds which exhibit very simple structure (viz. negligible vegetation, and only a few rocks or fallen logs at the bottom of the pond). Although we did not quantify the abundance of nine-spined sticklebacks, it was evident from catch numbers and relative effort that population densities in ponds exceed those in the marine environment.

**Brain measurements**

The entire procedure, from dissection through photography and measurement of whole brains and brain regions, followed exactly those outlined in Gonda et al. [12,13]. Fish (N = 15 per population) were euthanized with an overdose of MS 222 (tricaine methanesulfonate). Body weights were measured to the nearest 0.01 g with a digital balance and standard length (from the tip of the mouth to the end of the caudal peduncle) to the nearest 0.01 mm with digital callipers (for population variation in body weight and standard length see Additional file 1). Freshly dissected brains were fixed in 4% buffered formalin (0.1 M phosphate buffered saline) solution for 48 h. After fixation, digital photos were taken.

**Analyses**

Absolute brain size was compared among populations using a General Linear Model (GLM) with brain volume as dependent variable and population as fixed factor. To test the habitat effect directly, we also ran a General Linear Mixed Model (GLMM) with brain volume as a dependent variable, habitat type (marine vs. pond) as a fixed effect, and population, nested within habitat type,
as a random factor. Note that in this and the subsequent (see below) tests of habitat effects we excluded the single lake population and only compared three marine with four pond populations. To account for an allometric brain size - body size relationship, all metric variables were \( \log_{10} \) (hereafter log) transformed. Because a GLM with log body weight as a dependent variable revealed population dependent patterns in the log standard length - log body weight relationship (population: \( F_{7, 104} = 13.02, P < 0.001 \); standard length: \( F_{1, 104} = 152.73, P < 0.001 \); population \times standard length: \( F_{7, 104} = 14.56, P < 0.001 \)), subsequent analyses of total brain size or brain part size included both log standard length and log body weight for size correction.

To study body - brain size allometry, we performed a GLM with log brain volume as the dependent variable, population as a fixed factor, and log standard length and log body weight as covariates, including factor \( \times \) covariates. We also ran a simplified GLM with only log body weight as a covariate, but the results remained qualitatively the same (data not shown). As only log body weight was significant in the model (see Results) the slope of the log brain size - log body weight correlation was determined by linear regression.

To assess relative brain size trends, we applied two approaches. First we ran a GLMM using log brain volume as the dependent variable, with habitat fixed and population nested within the habitat (random) factor, and log body weight and log standard length as covariates. Second, to compare populations directly, we ran a GLM with log brain volume as a dependent variable, population as a fixed factor and log body weight and log standard length as covariates. To compare the relative size of different brain parts, we ran a multivariate GLM with the log brain parts as dependent variables, population, origin and their interaction as fixed factors, and log standard length, log body weight and log brain volume as covariates. Upon significant multivariate effects, the related univariate tests were considered. Because we did not find significant population \( \times \) origin interaction in the multivariate GLM (see Results), we did not address habitat \( \times \) origin effects with GLMMs.

As in all GLMs and GLMMs testing for population or habitat divergence in relative brain volume or relative brain part volume the covariates were only used for correction, we did not include factor-covariate interactions. All statistical analyses were performed with the SPSS 18.0 for Windows package (SPSS Inc., Chicago, Illinois, USA).

### Additional material

#### Additional file 1: Body size of the nine-spined sticklebacks

**Pungitius pungitius** used in this study. Standard length and body weight of the nine-spined stickleback (*Pungitius pungitius*) individuals used in the present study in the different populations. Mean \( \pm \) SD and the minimum - maximum range are presented.

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### Authors’ contributions

AG collected data for the study, dissected the brains, took photos of them, measured the sizes of the brains and the sizes of the different brain parts and participated in the design of the study and writing of the manuscript. GH performed the statistical analyses, participated in the design of the study and helped in the writing of the manuscript. JM participated in the design and coordination of the study and helped in the writing of the manuscript. All authors read and approved the final manuscript.

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References
1. Harvey PH, Clutton-Brock TH, Mace GM: Brain size and ecology in small mammals and primates. Proc Natl Acad Sci USA 1980, 77:4387-4389.
2. Clutton-Brock TH, Harvey PH: Primates, brains and ecology. J Zool 1980, 190:309-323.
3. Marino L: A comparison of encephalization between Odontocete Cetaceans and anthropoid primates. Brain Behav Evol 1998, 51:230-238.
4. Day LB, Westcott DA, Olser DH: Evolution of brower complexity and cerebellum size in bowheadbirds. Brain Behav Evol 2005, 66(6):21-72.
5. Aviles JM, Garamszegi LZ: Egg rejection and brain size among potential hosts of the common cuckoo. Ethology 2007, 113:562-572.
6. Karlen SJ, Krubitzer L: Phenotypic diversity is the cornerstone of evolution: Variation in cortical field size within Short Tailed Opossums. J Comp Neurol 2006, 499:990-999.
7. Ishikawa Y, Yoshimoto M, Yamamoto N, Ito H: Different brain morphologies from different genotypes in a single telost species, the medaka (Oryzias latipes). Brain Behav Evol 1999, 53:2-9.
8. Burns JG, Rodd H: HAtiness, brain size and predation regime affect the performance of wild guppies in a spatial memory task. Anim Behav 2008, 76:911-922.
9. Gonda A, Herczeg G, Merilä J: Adaptive brain size divergence in nine-spined sticklebacks (Pungitius pungitius). J Exp Biol 2009, 212:1721-1726.
10. Gonda A, Herczeg G, Merilä J: Habitat-dependent and -independent plastic responses to social environment in the nine-spined stickleback (Pungitius pungitius) brain. Proc R Soc B 2009, 276:2085-2092.
11. Roth TC, Pravosudov V: Hippocampal volumes and neuron numbers increase along a gradient of environmental harshness: a large-scale comparison. Proc Roy Soc B 2009, 276:401-405.
12. Kolm N, Gonzalez-Voyer A, Belin D, Winberg S: Evidence for small scale variation in the vetebrate brain: mating strategy and sex affect brain size and structure in wild brown trout (Salmo trutta). J Evol Biol 2009, 22:2524-2531.
13. Park PJ, Bell MA: Variation of telencephalon morphology of the threespine stickleback (Gasterosteus aculeatus) in relation to inferred ecology. J Evol Biol 2010, 23:1261-1277.
14. Chrissos E, Chapman LJ: Geographic variation in phenotypic plasticity in response to dissolved oxygen in an African cichlid fish. J Evol Biol 2010, 23:2091-2103.
15. Merila J, Cmoklas P: Comparison of genetic differentiation at marker loci and quantitative traits. J Evol Biol 2001, 14:892-903.
16. Lemonen T, O’Hara B, Cano JM, Merila J: Comparative studies of quantitative trait and neutral marker divergence: a meta-analysis. J Evol Biol 2008, 21:1-17.
17. Nettlethorpe F: A brain for all seasons: cyclical anatomical changes in song control nuclei of the canary brain. Science 1981, 214:1368-1370.
18. Tramontin AD, Bienowitz EA: Seasonal plasticity in adult brain. Trends Neurosci 2000, 23:251-255.
19. Patel SN, Clayton NS, Krebs JR: Spatial learning induces neurogenesis in the avian brain. Behav Brain Res 1997, 81:115-128.
20. Maguire EA, Gadian DG, Johnsrude IS, Good CD, Ashburner J, Frackowiak RSJ, Frith CD: Navigation-related structural change in the hippocampus of taxi drivers. Proc Natl Acad Sci USA 1997, 94:5986-5990.
21. Marchetti MP, Nevitt GA: Effects of hatchery rearing on brain structures of rainbow trout, Oncorhynchus mykiss. Env Biol Fish 2003, 66:9-14.
22. Khislringer RL, Nevitt GA: Early rearing environment impacts cerebellar growth in juvenile salmon. J Exp Biol 2006, 209:504-509.
23. Bånàrescu RM, Paepke HJ: In: The Freshwater Fishes of Europe. Volume SII. AULA-Verlag, Wiebelshaus, 2001.
24. Axelrod LC, Wheelie P: The expensive tissue hypothesis - The brain and digestive system in human and primate evolution. Curr Anthropol 1995, 36:199-221.
25. Lande R: Quantitative genetic analysis of multivariate evolution, applied to brain: body size allometry. Evolution 1979, 33:402-416.
26. Harvey PH, Bennett PM: Brain size, energetics, ecology, and life history patterns. Nature 1983, 306:314-315.
27. Martin RD, Harvey PH: Brain size allometry: ontogeny and phylogeny. In Size and scaling in primate biology. Edited by: Jungers WL, New York, 1985:147-173.
28. Pagel MD, Harvey PH: The taxon-level problem in the evolution of mammalian brain size: facts and artifacts. Am Nat 1988, 132:344-359.
29. Niedrger GF: Principles of Brain Evolution Sunderland: Sinauer Associates, 2005.
30. Bauchot R, Plantel R, Ridet JM: Brain-body weight relationship in Selachii. Copeia 1976, 2:305-310.
31. Herczeg G, Gonda A, Merilä J: Evolution of gigantism in nine-spined sticklebacks. Evolution 2009, 63:190-200.
32. Herczeg G, Gonda A, Merilä J: The rule inverted - female-driven gigantism in nine-spined stickleback Pungitius pungitius. J Anim Ecol 2010, 79:581-588.
33. Pollen AA, Dobberfuhl AP, Scace J, Igulu MM, Renn SCP, Shumway CA, Hofmann HA: Environmental complexity and social organisation sculpt the brain in Lake Tanganyikan cichlid fish. Brain Behav Evol 2007, 70:21-39.
34. Bauchot R, Bauchot ML, Plantel RIdet JM: Brains of Hawaiian tropical fish; brain size and evolution. Copeia 1977, 1:1977-42-46.
35. Conover DO, Schultz ET: Phylogenetic similarity and the evolutionary significance of countergradient variation. Trends: Ecol Evol 1995, 10:248-252.
36. Alho JS, Herczeg G, Soderman F, Laurila A, Jonsson KI, Merila J: Increasing melanin along a latitudinal gradient in a widespread amphibian: local adaptation, ontogenic or environmental plasticity? BMC Evol Biol 2010, 10:217.
37. Gersapp P, Teplitsky C, Alho JS, Mills JA, Merila J: Climate change and evolution: disentangling environmental and genetic responses. Mol Ecol 2008, 17:167-178.
38. Reader SM, Laland KM: Social intelligence, innovation, and enhanced brain size in primates. Proc Natl Acad Sci USA 2002, 99:4436-4441.
39. Gonzalez-Voyer A, Winberg K, Kuhn N: Distinct evolutionary patterns of brain and body size during adaptive radiation. Evolution 2009, 63:226-2274.
40. Pravosudov W, Kitayak AS, Omanaoka A: The relationship between migratory behaviour, memory and the hippocampus: an intraspecific comparison. Proc Roy Soc B 2006, 273:2641-2649.
41. Köhler M, Maya-Solá S: Reduction of brain and sense organs in the fossil insular bovid Myotragus. Brain Behav Evol 2004, 63:125-140.
42. Jones KE, MacLarnon AM: Affording larger brains: Testing hypotheses of mammalian brain evolution on bats. Am Nat 2004, 164:E20-E31.
43. Dunbar RW, Shultz S: Evolution in the social brain. Science 2007, 317:1344-1347.
44. Herczeg G, Gonda A, Merilä J: Predation mediated population divergence in complex behaviour of nine-spined stickleback (Pungitius pungitius). J Exp Biol 2009, 212:544-552.
45. Herczeg G, Gonda A, Merila J: The social cost of shoaling covaries with predation risk in nine-spined stickleback (Pungitius pungitius) populations. Anim Behav 2009, 77:775-780.
46. Brie SC, Leonard RB, Coggeshall RE: Neuronal increase in various areas of the nervous system of the guppy, Lebistes. J Comp Neurol 1980, 194:39-53.
47. Raymond PA, Easter SS Jr: Postembryonic growth of the optic tectum in goldfish. I. Location of neuronal cells and numbers of neurons produced. J Neurosci 1983, 3:1077-1091.
48. Zupanc GKH, Horschik P: Proliferation zones in the brain of adult sticklebacks. Brain Behav Evol 2010, 75:230.
49. Zupanc GKH: Adult neurogenesis and neuronal regeneration in the central nervous system of teleost fish. Brain Behav Evol 2001, 58:250-275.
50. Zupanc GKH: Neurogenesis and neuronal regeneration in the adult fish brain. J Comp Physiol A 2006, 192:649-670.
51. Kuska D: Mammalian domestication and its effect on brain structure and behavior. In Intelligence and Evolutionary Biology. Edited by: Jerison HJ, Jerison I. Springer-Verlag, Berlin, 1988:211-250.
52. Khislrin RL, Lema SC, Nevitt GA: Environmental rearing conditions produce forebrain differences in wild Chinook salmon Oncorhynchus tshawytscha. Comp Biochem Physiol A 2006, 145:145-151.
56. Burns JG, Saravanan A, Rodd FH: Rearing environment affects the brain size of guppies: lab-reared guppies have smaller brains than wild-caught guppies. Ethology 2009, 115:122-133.

57. Eronen M, Gluckert G, Hatakka L, van der Plassche O, van der Plicht J, Rantala P: Rates of Holocene isostatic uplift and relative sea-level lowering of the Baltic in SW Finland based on studies of isolation contacts. Boreas 2001, 30:17-30.

58. Herczeg G, Turtiainen M, Merila J: Morphological divergence of North-European nine-spined sticklebacks (Pungitius pungitius): signatures of parallel evolution. Biol J Linn Soc 2010, 101:413-416.

59. Rohlf FJ: 2002 tpsDig, digitize landmarks and outlines, version 1.37. Department of Ecology and Evolution, State University of New York at Stony Brook.

60. Huber R, van Staaden M, Kaufman LS, Liem KF: Microhabitat use, trophic patterns and the evolution of brain structure in African cichlids. Brain Behav Evol 1997, 50:167-182.

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