Translational regulation of *Anopheles gambiae* mRNAs in the midgut during *Plasmodium falciparum* infection

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**Abstract**

**Background:** Malaria is caused by *Plasmodium* parasites, which are transmitted via the bites of infected Anopheline mosquitoes. Midgut invasion is a major bottleneck for *Plasmodium* development inside the mosquito vectors. Malaria parasites in the midgut are surrounded by a hostile environment rich in digestive enzymes, while a rapidly responding immune system recognizes *Plasmodium* ookinetes and recruits killing factors from the midgut and surrounding tissues, dramatically reducing the population of invading ookinetes before they can successfully traverse the midgut epithelium. Understanding molecular details of the parasite-vector interactions requires precise measurement of nascent protein synthesis in the mosquito during *Plasmodium* infection. Current expression profiling primarily monitors alterations in steady-state levels of mRNA, but does not address the equally critical issue of whether the proteins encoded by the mRNAs are actually synthesized.

**Results:** In this study, we used sucrose density gradient centrifugation to isolate actively translating *Anopheles gambiae* mRNAs based upon their association with polyribosomes (polysomes). The proportion of individual gene transcripts associated with polysomes, which is determined by RNA deep sequencing, reflects mRNA translational status. This approach led to identification of 1017 mosquito transcripts that were primarily regulated at the translational level after ingestion of *Plasmodium falciparum*-infected blood. Caspar, a negative regulator of the NF-kappaB transcription factor Rel2, appears to be substantially activated at the translational levels during *Plasmodium* infection. In addition, transcripts of Dcr1, Dcr2 and Drosha, which are involved in small RNA biosynthesis, exhibited enhanced associations with polysomes after *P. falciparum* challenge. This observation suggests that mosquito microRNAs may play an important role in reactions against *Plasmodium* invasion.

**Conclusions:** We analyzed both total cellular mRNAs and mRNAs that are associated with polysomes to simultaneously monitor transcriptomes and nascent protein synthesis in the mosquito. This approach provides more accurate information regarding the rate of protein synthesis, and identifies some mosquito factors that might have gone unrecognized because expression of these proteins is regulated mainly at the translational level rather than at the transcriptional level after mosquitoes ingest a *Plasmodium*-infected blood meal.

**Keywords:** Genome-wide study, mRNA translation, Polysome, Mosquito immune reaction, Malaria parasite
Background
Malaria is the most deadly tropical parasitic disease faced by mankind, responsible for around 1.2 million deaths each year [1]. The malaria parasite *Plasmodium* must complete a complex developmental cycle in the mosquito in order to be transmitted from person to person. When a female mosquito feeds on an infected human, it takes up parasite-laden blood. *Plasmodium* gametocytes rapidly differentiate to male and female gametes, and fertilize inside the mosquito midgut to produce zygotes. The zygotes develop into motile ookinetes that invade and traverse the midgut epithelial cells [2]. In the space between the midgut epithelium and the basal lamina, the ookinetes transform into oocysts. After maturation, each oocyst ruptures and sends out thousands of sporozoites into the hemolymph. These sporozoites later migrate to the mosquito’s salivary glands and are released into the saliva during a subsequent blood meal, infecting another person and completing the parasite cycle in the mosquito [3].

Mosquitoes have developed various mechanisms to confront *Plasmodium* infection. To accomplish transmission from person to person, the malaria parasite *Plasmodium* must undergo complex developmental transitions and survive numerous attacks from the mosquito’s innate immunity system [2,4]. A variety of mosquito factors have been shown to affect the development of *Plasmodium* parasites in the mosquito [reviewed in [4,5]]. A better understanding of the cellular and molecular mechanisms that underlie vector-parasite interaction may provide critical targets and facilitate the development of new effective malaria control strategies.

The midgut represents one of the most challenging environments for the survival and development of *Plasmodium*. Microarray analyses have unravelled many transcriptional changes in the mosquito midgut in response to *Plasmodium* infection [6-8]. These informative studies detail the relative abundance of steady-state mRNA, providing the vector-borne disease community with valuable information regarding the transcriptional response to infection. However, these assays could not address whether or when the cognate proteins are actually synthesized, and whether translational regulation of mosquito mRNA takes place in response to *Plasmodium* challenge.

Polysome profiling has been previously used in other organisms to identify translational regulation in nutritional homeostasis [9], cellular stress response [9,10], embryogenesis [11], spermatogenesis [12], cancer progression and cancer chemotherapy [13,14]. This approach is based on the principle that messenger RNAs that are being actively translated usually have multiple ribosomes associated with them, forming large structures known as polysomes. In contrast, translationally inactive mRNAs generally are associated with messenger ribonucleoprotein particles or a single ribosome (collectively called “nonpolysomes”). Sucrose density gradient centrifugation is used to separate polysomes from nonpolysomes. The mRNA levels of individual genes in polysome fractions and nonpolysome fractions are determined by using a variety of different methods, and the ratios reflect mRNA translational status [15]. Here we report our systematic study of the translational status of individual mRNA species in *Anopheles gambiae* midguts after a blood meal containing *Plasmodium falciparum* gametocytes. RNA polysomal profiling indicates that transcripts of a large group of mosquito genes are enriched in polysome fractions in *P. falciparum*-infected mosquitoes, as compared with the uninfected control. The result suggests that translational regulation of gene expression in mosquito midguts has a profound impact on the anti-malaria reactions. This approach provides more molecular details in the interaction between mosquitoes and malaria parasites, and may reveal additional targets that can be exploited to reduce mosquito vector competence via genetic manipulation.

Results
Changes in mRNA association with polysomes after *P. falciparum* infection
Our hypothesis is that midgut invasion by *P. falciparum* ookinetes alters mosquito gene expression at both the transcriptional and translational levels. To test this hypothesis, we analyzed genome-wide mRNA translational status by measuring the proportion of individual mRNA species in polysome complexes.

Midguts from female *Anopheles gambiae* mosquitoes were dissected at approximately a day (22–26 hours) after ingestion of *P. falciparum*-infected blood. Mosquitoes fed on uninfected blood were used as control. A portion of the midguts were used for isolation of total cellular RNA. Extracts of the remaining midguts were fractionated over sucrose density gradients, and fifteen fractions were collected from the top of each gradient (Figure 1). Non-polysomal fractions, as well as polysomal fractions were combined, respectively, to obtain two RNA pools per gradient for three independent experiments. The first, last and the sample between non-polysomal and polysomal, were all discarded to ensure pure pools from each set. A fourth experiment was conducted where the polysomal and non-polysomal samples were not pooled, to be used later for qRT-PCR. The mRNA levels of individual mosquito genes in polysome (PS) fractions, nonpolysome (NP) fractions and unfractionated steady-state (total) RNA were determined using high throughput RNA deep sequencing. Each RNA pool generated 2.1-9.8 million raw reads (Table 1).
Signal intensities of transcripts from each PS pool were compared to those of transcripts from a matching NP pool. To quantify the translational status of individual mRNA species, we define the relative PS loading (PL) as the extent of mRNA association with polysomes:

\[
PL = \frac{\text{Expression level in PS}}{\text{Expression level in NP} + \text{Expression level in PS}}
\]

PL values were compared between the mosquitoes fed on \textit{P. falciparum}-infected or -uninfected blood. After exposure to \textit{Plasmodium} parasites, 1170 transcripts became increasingly associated with polysomes (≥ 2 fold, \( p < 0.05 \)) in mosquito midguts (Additional file 1: Table S1). In contrast, only 7 transcripts shifted more towards nonpolysomal fractions (≥ 2 fold, \( p < 0.05 \)).

To validate the alteration of polysome-mRNA association, we carried out quantitative real-time RT-PCR (qRT-PCR) analysis with PS and NP RNAs collected in the above-mentioned experiments. PL values were compared for 16 selected genes between the \textit{Plasmodium}-challenged and control mosquitoes. These genes were selected to represent various expression levels and functional categories. A decent correlation coefficient

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**Table 1 An. gambiae RNA-seq Expression Data**

|                     | Infected Non-polysomal | Infected Polysomal | Uninfected Non-polysomal | Uninfected Polysomal | Uninfected Total RNA | Infected Total RNA |
|---------------------|------------------------|--------------------|--------------------------|----------------------|----------------------|--------------------|
| Total Reads Set 1   | 7785361                | 4244042            | 4781123                  | 4766035              | 2081115              | 6150051            |
| Total Reads Set 2   | 3871849                | 3442387            | 3351084                  | 2203276              | 9837495              | 4328298            |
| Total Reads Set 3   | 6597022                | 6980908            | 8588835                  | 5980648              | 3009585              | 8407264            |
| Reads Mapped to Transcriptome Set 1 | 3527074 (45.3%) | 3423900 (80.7%) | 3608010 (75.5%) | 1049208 (22.0%) | 1483831 (71.3%) | 5144550 (83.7%) |
| Reads Mapped to Transcriptome Set 2 | 2332634 (60.2%) | 1476806 (42.9%) | 2557909 (76.3%) | 473146 (21.5%) | 7772228 (79.0%) | 2612416 (60.4%) |
| Reads Mapped to Transcriptome Set 3 | 4942339 (74.9%) | 5191081 (74.4%) | 7221237 (84.1%) | 4992710 (83.5%) | 2634845 (71.3%) | 5992586 (71.3%) |

Sequence reads are listed for three independent experiments (Sets 1–3). The six samples were multiplexed and sequenced together. Each set had roughly 30 million raw reads. Erroneous reads and reads where linkers were incomplete or missing were excluded from total number of reads. Reads mapped to transcriptome are the number (and percentage) of reads which matched back to the An. gambiae cDNA P3.4 database.
(\(R^2 = 0.8611\)) was observed between the PL values determined by RNA deep sequencing and qRT-PCR, indicating that RNA-seq analysis provides reliable PL data (Figure 2).

**Identification of mosquito genes that are primarily regulated at the translational level in response to *P. falciparum* infection**

It is conceivable that some of changes in PL values simply reflect up- or down-regulation at the transcriptional level. The central concept of translational regulation is that cellular gene expression is governed by the efficiency of translation of a given mRNA in the absence of a corresponding change in steady-state mRNA levels [16]. In order to uncover mosquito genes that are regulated by translational mechanisms after exposure to *Plasmodium* parasites, we plotted the difference in mRNA association with polysomes (\(\log_{2}PL_{\text{infected}} - \log_{2}PL_{\text{uninfected}}\)) between the *Plasmodium*-challenged and control mosquitoes against the difference in cellular transcript levels (\(\log_{2}\text{mRNA}_{\text{infected}} - \log_{2}\text{mRNA}_{\text{uninfected}}\)) for every detectable mRNA. As shown in Figure 3, the dots in rectangle I represent transcripts that show no significant difference in total cellular abundance between the two mosquito groups, but vary considerably in polysomal association, indicating that expression of those genes is up-regulated at the translational level. The transcript levels of some other mRNA species (dots in rectangle II) decrease in the *Plasmodium*-challenged mosquitoes, while the mRNA polysomal recruitment even increases in these mosquitoes, again demonstrating the translational enhancement in the infected mosquitoes.

Among the 1170 transcripts showing significantly increased association with polysomes, the steady-state mRNA levels of 160 transcripts also increased after *Plasmodium* infection (Table 2). The majority of the transcripts (905) remained at relatively constant level while the amount of mRNA for 105 genes went down in the

| Transcripts showing significantly increased polysome loading (1170 in total) | Significantly increased | Unchanged | Significantly decreased |
|---|---|---|
| Transcripts showing significantly decreased polysome loading (7 in total) | 2 | 5 | 0 |

The list includes transcripts showing enhanced polysomal association (\(\log_{2}\text{PL}_{\text{infected}} - \log_{2}\text{PL}_{\text{uninfected}}\) ≥ 1, \(p < 0.05\)) or decreased association (\(\log_{2}\text{PL}_{\text{infected}} - \log_{2}\text{PL}_{\text{uninfected}}\) ≤ −1, \(p < 0.05\)) in response to *Plasmodium* infection. Unchanged steady state mRNA levels are defined as within the cut-off \(1 \geq \log_{2}\text{mRNA}_{\text{infected}} - \log_{2}\text{mRNA}_{\text{uninfected}} \geq -1\).
Plasmodium-challenged mosquitoes. For the 7 transcripts exhibiting decreased association with polysomes, their steady-state mRNA levels either showed no significant change in total cellular abundance or moved in opposite direction. Together, we have identified 1017 transcripts in the midgut of An.gambiae that are primarily regulated at translational level after ingestion of P. falciparum-infected blood. Proteins encoded by these transcripts are predicted to have diverse molecular functions (Figure 4). Compared with the transcriptome in the midgut, overrepresented or underrepresented gene ontologies were not found in the translationally regulated genes.

Translational regulation of immune-related genes in mosquito defense reactions against invading P. falciparum

In An. gambiae, twenty-five immune-related genes displayed a significantly higher polysomal association and therefore more active translation during infection (Table 3). These genes are implicated in different pathways of mosquito innate immunity [18,19]. Five genes (APG4B, APG6, APG7A, APG8 and APG18B) are involved in autophagy. Seven genes (CLIPA6, CLIPA7, CLIPA14, CLIPB17, CLIPC3, SRPN1 and SPRN10) encode Clip-domain serine proteases and serine protease inhibitors that are presumably involved in activation cascades. Two genes (HPX8 and CuSOD2) encode enzymes that catalyze generation and detoxification of reactive oxygen species, respectively. Three genes (CACTUS, CASPAR and REL2) are components of Toll and IMD immune signalling pathways. Five genes (DCR1, DCR2, DROSHA, RM62F and SPNE) encode key proteins in the small RNA regulatory pathway.

We selected four genes from the list to perform detailed polysomal profiling analysis (Figure 5). The CLIPB17 transcript was barely associated with ribosomes in the uninfected mosquitoes, and mRNA polysome association was evident in the P. falciparum-infected mosquitoes. For CASPAR, DCR1 and DCR2, their transcripts were loaded on polysomes even in the uninfected mosquitoes. It appeared that each mRNA molecule was associated with markedly more ribosomes in the infected mosquitoes as the transcripts were shifting more toward the bottom of the sucrose gradient, suggesting that more proteins were synthesized from each mRNA template. In contrast, the housekeeping gene rps7 did not alter its mRNA distribution within the polysome profiles in the same experiment. The result demonstrated that the identified genes were regulated at the translational level in response to Plasmodium infection.

Discussion

In this study, we discovered that a plethora of mosquito genes showed no significant difference in total cellular abundance between the Plasmodium-infected and uninfected mosquitoes, but varied considerably in polysomal association. Translational regulation allows cells to respond swiftly to all types of environmental stimuli and to fine-tune protein levels in both time and space. For most of the genes, it is unknown whether the enhanced translation is directly related to defense responses against invading P. falciparum.
Some of the genes, however, have been shown previously to play pivotal roles in anti-Plasmodium immunity. When the midgut of An. gambiae was invaded by the rodent malaria parasite Plasmodium berghei, SRPN10 was transcriptionally activated and overexpressed SRPN10 proteins were detected in the parasite-invaded midgut epithelial cells [21]. However, the mRNA levels were not significantly changed when An. gambiae was infected by the human malaria parasite P. falciparum [21,22]. Our experiment suggests that more SRPN10 proteins are still synthesized in the midgut of the infected mosquitoes as a result of enhanced translation.

Rel1 and Rel2, two NF-κB-like transcription factors, are key regulators of the mosquito immune response. Rel1 plays a critical role in controlling resistance of An. gambiae to the rodent malaria parasite P. berghei, while Rel2 seems to provide more protection against the human malaria parasite P. falciparum [23,24]. Rel1 and Rel2 are controlled by the negative regulators Cactus and Caspar, respectively. Our analysis suggested that Rel2, Cactus and Caspar were all translationally upregulated in the P. falciparum-infected mosquitoes. In future studies it will be interesting to find out whether over-expression of these proteins is restricted in the parasite-invaded midgut epithelial cells or occurs in all the epithelial cells. This may help to explain why expression of Rel2 and its inhibitor are both enhanced in response to Plasmodium infection.

We found significant up-regulation at the translational level in multiple proteins (Dcr1, Dcr2, and Drosha) that are required for biogenesis of miRNA and siRNA. The siRNA pathway is essential to the survival of mosquitoes infected with Sindbis virus and West Nile virus [25,26]. The miRNA pathway has previously been implicated in

### Table 3 Translationally regulated immune-related genes in response to P. falciparum infection

| Ensembl Gene ID | ImmunoDB Identity | Polysome loading Fold change (log2) P-value | Transcript abundance Fold change (log2) P-value |
|-----------------|-------------------|--------------------------------------------|---------------------------------------------|
| AGAP005910      | APG18B            | 2.080                                      | −0.812                                      |
| AGAP008497      | APG4B             | 3.183                                      | −0.881                                      |
| AGAP003858      | APG6              | 2.031                                      | −0.269                                      |
| AGAP008637      | APG7A             | 2.953                                      | 0.169                                       |
| AGAP002685      | APG8              | 1.606                                      | −0.425                                      |
| AGAP007938      | CACTUS            | 2.229                                      | 0.041                                       |
| AGAP006473      | CASPAR            | 4.273                                      | −1.055                                      |
| AGAP000830      | CASP57            | 2.563                                      | 0.764                                       |
| AGAP011788      | CLIPA14           | 4.384                                      | −0.078                                      |
| AGAP011789      | CLIPA6            | 2.312                                      | 0.619                                       |
| AGAP011792      | CLIPA7            | 2.336                                      | −0.392                                      |
| AGAP001648      | CLIPB17           | 3.619                                      | 0.494                                       |
| AGAP004318      | CLIPC3            | 3.574                                      | 0.842                                       |
| AGAP005234      | CuSOD2            | 2.037                                      | −0.531                                      |
| AGAP002836      | DCR1              | 3.632                                      | −0.462                                      |
| AGAP012289      | DCR2              | 4.277                                      | −1.882                                      |
| AGAP008087      | DROSHA            | 5.749                                      | 0.335                                       |
| AGAP004038      | HPX8              | 3.174                                      | 0.097                                       |
| AGAP007294      | IAP1              | 3.235                                      | −1.360                                      |
| AGAP005552      | PGRPLD            | 1.942                                      | −1.146                                      |
| AGAP006747      | REL2              | 1.767                                      | −0.637                                      |
| AGAP012523      | RM62F             | 2.005                                      | −0.446                                      |
| AGAP002829      | SPNE              | 3.042                                      | −0.562                                      |
| AGAP006909      | SRPN1             | 2.017                                      | 0.630                                       |
| AGAP005246      | SRPN10            | 1.743                                      | −0.869                                      |

Among 1017 transcripts that are translationally regulated after ingestion of P. falciparum-infected blood, 35 transcripts have putative or established immune-related function (ImmunoDB identities). In several cases, alternate splice variants of the same gene were detected. Most of the transcripts in this list displayed significant translational change when insignificant change was observed in mRNA levels \([1 ≥ (\log_{2}\text{mRNA}_{\text{Infected}} - \log_{2}\text{mRNA}_{\text{Uninfected}}) ≥ −1]\). Some transcripts (i.e. DCR2) showed significant downregulation in transcription and upregulation in translation.
An. gambiae when infected with Plasmodium [27] and in Culicine mosquitoes infected with viral invaders [28]. Winter et al. found that P. berghei infection was significantly enhanced by knockdown of key components of the miRNA pathway (Dcr1, Ago1, and Drosha), suggesting a role for these proteins in the response to infection [27]. This is the first time, to our knowledge, that mosquito miRNA processing proteins have been found to be regulated at the translational level in response to P. falciparum infection in An. gambiae.

Additional proteins showing significant translational change include Spindle E (spnE), which is involved in mRNA translational states between mosquitoes that are infected by ookinetes of the wild-type Plasmodium or CTRP knockout Plasmodium mutant will reveal whether the translational regulation is triggered by ookinete invasion of the mosquito midgut. This study takes a snapshot at 22–26 h after blood feeding, and demonstrates that both transcriptional and translational regulation of mosquito genes takes place when Plasmodium parasites invade the mosquito midgut. In order to provide a comprehensive view of the mode of gene regulation, more experiments need to be conducted with midguts and some other tissues (i.e. the fat bodies) at various time points after an infected blood meal. Furthermore, it is imperative to examine translational regulation in natural vector-parasite combinations since co-evolution and natural selection has made the mosquito-Plasmodium interactions vary among different vector-parasite combinations, or even between different geographical strains of the same species. Comparing gene expression in the Plasmodium-refractory and susceptible mosquitoes taken recently from an epidemic region in Africa may lead to identification of mosquito factors that determine the ability of mosquitoes to transmit the parasites.

Conclusions

We report a previously neglected aspect of gene expression regulation in the midgut of An. gambiae mosquitoes after ingestion of a Plasmodium-infected blood meal. Compared with the mosquitoes fed on uninfected blood, a large group of genes in the infected mosquitoes exhibit a redistribution of their mRNA transcripts between monosomes (or ribonucleoprotein particles) and translationally active polysomes, without a concomitant change in the steady-state mRNA levels. This finding supports our hypothesis that the anti-malarial response of Anopheles gambiae occurs at both the transcriptional and translational level. Elucidating the profound impact on translation of mosquito mRNA will extend knowledge of the molecular interaction between malaria parasites and their mosquito hosts. Genes for which mRNA translation is affected by Plasmodium infection may have been previously deemed “irrelevant” simply because their
transcription is not altered by exposure to the parasites. Functions of genes identified in this project will be further explored in future studies to reveal new targets that can be employed to reduce mosquito vector competence.

Methods

Gametocyte production and bloodfeeding

*P. falciparum* gametocyte cultures for infecting *Anopheles gambiae* mosquitoes were generated largely based upon the protocol of Looker and Taylor-Robinson [31]. Gametocyte cultures were initiated from asexual *P. falciparum* cultures (3D7G) following daily dilution with fresh human red blood cells and medium, with a starting parasitemia typically of 3-5% and a hematocrit of 5%. Gametocytemia was monitored during the course of each culture. At 17 days after seeding, an optimal mixture of male and female stage V gametocytes were present. Gametocytes were harvested and used immediately in bloodfeeding of mosquitoes.

Female adult *An. gambiae* (G3 strain) mosquitoes were maintained on 10% sucrose at 27 °C/80% humidity. Prior to bloodfeeding, the mosquitoes (4–6 days post-eclosion) were maintained overnight on water only. Bloodfeeding was carried out using a Hemotek 5 W1 system and collagen Hemotek feeding membranes (Discovery Workshops, Lancashire, England). Mosquitoes were allowed to feed for 15 minutes. This resulted in bloodfeeding of approximately 80–90% of available *Anopheles gambiae* females. A control group of mosquitoes were fed on uninfected blood and serum by essentially the same method.

Polysomal RNA preparation

Feeding on infected and uninfected blood was scheduled 5 hours apart. Two hundred female mosquitoes were dissected for each group at 22–26 hours post blood meal (PBM) to obtain midguts. Dissections started at 22 h PBM and ended at 26 h PBM for both groups at each experiment. The samples were flash frozen in liquid nitrogen and stored at −80 °C until processed. Frozen samples were ground using a mortar and pestle, homogenized in 1 ml lysis buffer (15 mM Tris–HCl, pH 8.0, 300 mM NaCl, 5 mM MgCl2, 0.5 mM DTT, 0.1 mg/ml cycloheximide, 1 mg/ml heparin, 1% triton X-100, 0.2 U/ml RNase inhibitor) [32]. The extracts were spun 15 minutes at 14,000 rpm at 4 °C to remove cell debris and nuclei. An aliquot of the supernatant was saved for analysis of total cellular mRNA. The rest of the supernatant was directly applied to the top of a 10% sucrose gradient that was prepared as described by Arava [33], containing 20 mM Tris–HCl (pH 8), 140 mM KCl, 5 mM MgCl2, 0.5 mM DTT, 0.1 mg/ml cycloheximide, and 0.5 mg/ml heparin. Centrifugation was carried out using an SW41-Ti rotor at 116,000 x g for 3 hours at 4 °C. Following spinning, gradient fractions were collected immediately using a Density Gradient Fractionation System (Brandel, Gaithersburg, MD) with simultaneous recording of absorbance profiles at 254 nm.

Following fractionation, non-polysomal aliquots (as shown in Figure 1) were pooled together into one sample, and polysomal aliquots were pooled into a second sample. The fraction between was discarded to ensure good separation. RNA samples from three independent infection experiments were collected. For each set, we had six RNA samples: polysomal and nonpolysomal RNAs of the infected mosquitoes, polysomal and nonpolysomal RNAs of the uninfected controls, and total cellular mRNAs of the infected and uninfected mosquitoes. A fourth set of samples were maintained as individual fractions for qRT-PCR analysis of specific transcripts.

Oocyst observation

At 7–9 days after blood feeding, 10–30 females from the same cohorts that had been used earlier for RNA collection were dissected for midguts to estimate the efficiency of the *P. falciparum* infection. Midguts were placed into a well of a 24-well plate with 1 ml of 1% mercurochrome for 10 minutes, followed by multiple rinses with sterile water and 5 minutes incubation to remove residual mercurochrome. *P. falciparum* oocysts were visualized under a Zeiss fluorescent microscope. The results are shown in Additional file 3: Figure S1.

RNA Isolation and sequencing preparation

RNA was extracted from the fractionated and unfractionated samples using Trizol reagent (Invitrogen, Carlsbad, CA). Sample preparation for Illumina-based mRNA sequencing was conducted based upon the manufacturer’s protocols “mRNA Sequencing Sample Preparation Guide” and “Preparing Samples for Multiplexed Paired-End Sequencing” (Part# 1005361Rev B, and Part# 1004898Rev D, Illumina, Inc., San Diego, CA). For each set, six cDNA libraries derived from the six RNA samples were uniquely tagged with index sequences at the PCR stage of sample preparation. Six libraries were pooled together and sequenced in a single lane of a flow cell. 76-cycle single read Illumina sequencing was conducted at the Virginia Bioinformatics Institute (VBI, Blacksburg, VA).

Data analysis of the deep sequencing results

Sequencing reads were sorted into separate files according to their barcode, and linker sequences were removed. Each file was then converted into fasta format, and blasted against the annotated *An. gambiae* transcripts (CDNA-ALL.AgamP3.4.fa; from Vectorbase) with an e-value cutoff set to −20. The number of hits for each
transcript was compiled in a list with a master table generated that showed the numbers of reads for each transcript in each sample. All RNA-seq data is deposited in Gene Expression Omnibus [GEO: GSE38707].

In order to obtain an accurate picture of the change in mRNA abundance, we eliminated transcripts from further consideration when the total hits in the paired polysomal and nonpolysomal pools of either infected mosquitoes or uninfected control were below 20 reads. We reasoned that levels of these transcripts were barely above the threshold of sequencing detection, and the numbers of reads might not accurately reflect their true abundance.

RNA-seq data for the unfraccionated RNA samples (total cellular RNA) were normalized with rps-7, a commonly used internal housekeeping control gene encoding 40 S ribosomal protein S7 [34,35]. As shown in Figure 5, the distribution of rps-7 mRNA in sucrose gradients was quite similar between the P. falciparum-infected and uninfected mosquitoes. Polysomal and nonpolysomal RNA samples were each separately normalized against rps-7 in the corresponding sucrose gradient fractions.

The portion of each transcript associated with polysomes (polysome loading, PL) in each sample was assessed from the number of reads for polysomal (P) and nonpolysomal (NP) as follows: PL = P/(NP + P). Averages across the three replicate samples for each transcript were obtained, and log2 transformations of these averages were performed. To evaluate the change in translational state for every transcript, the log2 PL was compared between the P. falciparum-infected mosquitoes and the uninfected control. Unpaired two-tailed t-tests were conducted as described by Kawaguchi et al. [9]. Transcripts with p-values below 0.05 were examined further.

**Quantitative real-time RT-PCR (qRT-PCR)**

To validate the polysomal association data, we carried out qRT-PCR analysis with aliquots of the pooled polysomal and nonpolysomal samples that were used in RNA-seq analysis. Sixteen randomly selected transcripts were measured using the Invitrogen SYBR GreenER kit (Invitrogen, Carlsbad, California) according to manufacturer’s protocols, with an ABI Prism 7300 Real Time PCR System (Applied Biosystems, Carlsbad, California). The sequences of the primers used are listed in Additional file 2: Table S2.

**Additional files**

- **Additional file 1: Table S1.** Analysis of the steady-state mRNA levels and mRNA association with polysomes during Plasmodium infection.

- **Additional file 2: Table S2.** Primers used in qRT-PCR.

- **Additional file 3: Figure S1.** Plasmodium falciparum oocyst counts in Anopheles gambiae at 7–9 days post blood feeding. Midguts were examined in 56 female mosquitoes that had been fed on P. falciparum-infected blood. Each * represents one mosquito. 64.3% of the mosquitoes were infected. It is possible that the actual infection rate was higher as we possibly did not remove away some of the unfed mosquitoes. The infection rate represents a conservative estimate. The average infected midgut with oocysts had 2.22 oocysts. The image on the right panel shows oocysts that we observed in a representative mosquito.

**Competing interests**

The authors declare that they have no competing interests.

**Authors’ contributions**

EAM, ML and IZ carried out experimental procedures. EAM generated Plasmodium falciparum gametocytes from in vitro culture. EAM, ZT and IZ analyzed the RNA-seq data. EAM and IZ wrote the article. All authors read and approved the final manuscript.

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