Design and synthesis of triazole-based peptidomimetics of a PSD-95 PDZ domain inhibitor
Bach, Anders; Pedersen, Thomas B.; Strømgaard, Kristian

Published in:
MedChemComm

DOI:
10.1039/C5MD00445D

Publication date:
2016

Document Version
Publisher's PDF, also known as Version of record

Citation for published version (APA):
Bach, A., Pedersen, T. B., & Strømgaard, K. (2016). Design and synthesis of triazole-based peptidomimetics of a PSD-95 PDZ domain inhibitor. DOI: 10.1039/C5MD00445D
Design and synthesis of triazole-based peptidomimetics of a PSD-95 PDZ domain inhibitor† †

Anders Bach, * Thomas B. Pedersen and Kristian Strengaard

PSD-95 PDZ domains are biologically important and promising drug targets. Here, we discover a triazole-based peptidomimetic, 10, by ‘click chemistry’. Compound 10 inhibits the PDZ2/GluN2B interaction with affinity similar to tripeptide SAV and better than current small-molecules. Thus, 10 represents a new class of PSD-95 PDZ inhibitors.

1. Introduction

Postsynaptic density protein-95 (PSD-95) is a scaffolding protein found in neuronal synapses. 1 PSD-95 contains three postsynaptic density protein-95/disk-large/zonula occludens-1 (PDZ) domains, which interact with the N-methyl-D-aspartate (NMDA) receptor and signalling proteins such as neuronal nitric oxide synthase (nNOS). 2 Compounds targeting PDZ1 and PDZ2 of PSD-95 have shown promising effects in animal models of ischemic stroke, 3, 10 pain, 6, 8 and neurodegenerative diseases, 9 as they can disconnect disease-related excessive NMDA receptor activity from harmful downstream mechanisms. 3, 10 PSD-95 is particularly promising as a drug target in diseases, 9 as they can disconnect disease-related excessive NMDA receptor activity from harmful downstream mechanisms. 3, 10 PSD-95 PDZ inhibitors existed. However, Vogrig et al. reported a series of indole-based compounds (Fig. 1C) able to interact with PSD-95 PDZ domains (1H-15N heteronuclear single-quantum coherence, HSQC, NMR), inhibit the PSD-95-PDZ/5-HT2A receptor interaction (pull-down) and relieve pain in hyperalgesic rats. 24 Truncation studies of peptide ligands targeting PSD-95 have shown that pentapeptides display affinities similar to longer peptides, while tetra- and tripeptides show a stepwise reduction in affinity. 15 Pentapeptides have subsequently been derivatized into dimeric inhibitors with nanomolar potency. 4, 21 Likewise, the affinity of tetrapeptides has been increased about 20-fold by N-alkylations and stability can be enhanced by replacing amides with thioamides. 15, 20 Also, cyclization and expansion of the P1 side chain of hexapeptides have led to peptidomimetic structures with increased affinities. 22, 23

Developing small-molecules targeting PDZ domains have shown to be difficult. Only few small-molecule inhibitors have been reported and affinities are in the low micromolar range in best cases. 13 Until recently, no small-molecule PSD-95 PDZ inhibitors existed. However, Vogrig et al. reported a series of indole-based compounds (Fig. 1C) able to interact with PSD-95 PDZ domains (1H-15N heteronuclear single-quantum coherence, HSQC, NMR), inhibit the PSD-95-PDZ/5-HT2A receptor interaction (pull-down) and relieve pain in hyperalgesic rats. 24 Herein, we pursue a peptidomimetic strategy introducing a rigid scaffold into a short PSD-95 peptide ligand with the potential of improving affinity, selectivity, and stability and provide novel chemotypes targeting PSD-95. We used the tripeptide Thr-Ala-Val (TAV) as starting point (Fig. 1A–C),
which binds PDZ1 and PDZ2 with medium affinity (PDZ2: \(K_i \sim 40 \, \mu M\)). TAV was identified from a peptide SAR study of the GluN2B C-terminal revealing that P\(^{1}\) Val is essential for affinity, P\(^{−1}\) Asp could be replaced with Ala, and Thr instead of Ser in P\(^{−2}\) leads to increased affinity.\(^{15}\) As TAV is small and reasonably potent it is an attractive starting point for converting peptides into more small-molecule-like compounds. Here, we prepared azide and alkyne analogues of the corresponding amino acids and dipeptides of TAV, which then served as building blocks for subsequent copper(I)-catalyzed azide–alkyne cycloaddition (CuAAC) reactions thereby producing a series of 1,2,3-triazole containing analogues of TAV.\(^{25,26}\) Triazole resembles amide in planarity, size and electronic properties and has previously been used as an amide bioisoster resulting in potent and protease stable compounds.\(^{27–30}\) Thus, here we systematically investigated triazole as a potential bioisoster in PSD-95 PDZ domain peptide inhibitors and the possibility to rigidify the peptide backbone by implementing this small and aromatic ring. This strategy led to the identification of a new triazole-containing peptidomimetic against the PDZ2 domain of PSD-95.

2. Results and discussion

2.1. Design and synthesis

In the design process we first fragmentized TAV into the amino acid and dipeptide moieties (Thr, Thr-Ala, Ala-Val and Val) and replaced the resulting amines with azide groups and carbonyls with methyl azides to generate the four azide building blocks (1–4, Fig. 2). Likewise, carbonyls of peptide fragments were replaced with acetylene to get alkyne 5 and 6. Additionally, alkynes 7–8 were made where the acetylene replaces the central part of P\(^{1}\) Ala and the important P\(^{0}\) and P\(^{−2}\) (Val and Thr) amino acids were preserved (Fig. 2). For 8, an alcohol group needed to replace the intended carbonyl in order to provide a water-stable building block. For both 7 and 8 it was envisioned that the carbonyl/alcohol could facilitate hydrogen bond interactions with the PDZ backbone. After preparing the building blocks, the strategy was to react
the four azides (1–4) with the four alkynes (5–8) applying the CuAAC reaction so that the corresponding triazol structures (9–18) contained at least one Val and carboxylic acid group intended to bind into the PDZ domain V⁰ pocket (Fig. 2). This systematic strategy allows for a thorough investigation of the idea of introducing a rigid scaffold into the TAV backbone.

The synthesis of azides (1–4) and alkynes (5–8) were carried out in satisfactorily yields generally following standard procedures (Schemes 1 and 2). Azide 1 was synthesized in one step from L-valine in a diazo transfer reaction mediated by freshly prepared trifly azide according to literature procedure. To generate azide 2, 19 (prepared by reduction of Boc- and tert-butyl protected tert-threonine) was mesylated on the alcohol group to provide intermediate 20 followed by a nucleophilic substitution reaction by treatment of sodium azide to get 21. Azide 2 was then obtained under mild deprotection conditions applying HCl in dioxane (Scheme 1). Azide 3 was synthesized by a solid-phase procedure starting from a Wang resin preloaded with Fmoc-protected L-valine (22). After deprotecting the Fmoc, (S)-2-azidopropanoic acid coupled to the free valine amine to get resin-bound intermediate 23. The desired azide 3 was obtained by cleaving the resin under standard conditions with trifluoroacetic acid (TFA) (Scheme 1). Azide 4 was synthesized by coupling 24 with Fmoc- and tert-butyl protected tert-threonine using HBTU followed by deprotection under mild acidic conditions (Scheme 1).

Next, alkyne building blocks (5–8) were synthesized. To prepare 5, the Boc- and tert-butyl protected tert-threonine was first converted to the corresponding aldehyde Boc-Thr(bu)-CHO (25). This aldehyde was then reacted with freshly in situ prepared Bestmann–Ohira reagent (dimethyl diazo-2-oxopropylphosphonate), and the resulting intermediate alkyne 26 was readily deprotected in iodotrimethylsilane to generate 5 (Scheme 2). Likewise, the synthesis of alkyne 6 started with aldehyde 27 and reaction with the Bestmann–Ohira reagent to generate 28 (Scheme 2). After deprotecting the Boc this intermediate was coupled with Boc-Thr(bu)-OH followed by another deprotection to facilitate alkyne 6. Alkyne 7 was generated by a solid-phase strategy where propionic acid was coupled to resin-bound L-valine using N-ethoxycarbonyl-2-ethoxy-1,2-dihydroquinoline (EEDQ) to get intermediate 30 followed by cleavage from the resin (Scheme 2). Alkyne 8 was synthesized by a Grignard reaction where ethynylmagnesium chloride was added to the carbonyl of aldehyde 25 followed by deprotection with TFA (Scheme 2).

The triazole-based peptidomimetic structures 9–18 were then synthesized by standard CuAAC reaction conditions using a 1:1 ratio of alkyne and azide, 0.4 eq. CuSO₄ and 2 eq. sodium ascorbate in water tert-butanol at room temperature (ESI†). For 11–18, the reactions completed within 30 minutes and compounds were obtained in reasonably to fine yields (>30%) after HPLC purification. Triazoles 9 and 10, both based on alkyne 5, needed modified conditions to give satisfactorily yields (66–67%). For 9, heating to 55 °C for 18 hours was applied, while for 10, acetonitrile (20%) replaced tert-butanol, more sodium ascorbate (6 eq.) was added together with DIPA and reaction time was prolonged to 65 hours at room temperature. Solutions of all tested compounds were prepared in deuterated PBS buffer and concentrations accurately determined by qNMR prior to assay tests.

2.2. Biological evaluation

Triazole-based compounds 9–18 were tested in a fluorescence polarization (FP) assay in a heterologus competition mode. Increasing concentrations of test substance were added to a fixed concentration of PDZ protein and fluorescent peptide probes, which were prepared by Cy5-labeling 11-mer peptides corresponding to the C-termini of the GluN2B subunit of NMDA receptors or CRIP1. We first tested 9–18 against PDZ2 of PSD-95 together with the control tripeptides TAV and the weaker binding tripeptide

---

**Scheme 1** Synthesis of azides 1–4. Reagents and conditions: (a) Tf₂O, NaNO₂, H₂O, CH₂Cl₂, 0 °C, 2 h; (b) CuSO₄·5H₂O, H₂O, CH₂OH, CH₂Cl₂, rt, overnight; (c) NaClO, Et₂N, CH₂Cl₂, 0 °C to rt, 4 h; (d) Na₂SO₄, DMF, 50 °C, 18 h; (e) 4 M HCl in dioxan, rt, 1.5 h; (f) 20% piperidine in DMF; (g) (S)-2-azidopropanoic acid, HATU, HOAt, collidine, DMF, rt, 3 h; (h) TFA/tips/H₂O (90/5/5), rt, 2 h; (i) Boc-Thr(bu)-OH, HBTU, DIPEA, DMF, rt, 12 h; (j) 4 M HCl in dioxan, rt, 1.5 h.

**Scheme 2** Synthesis of alkynes 5–8. Reagents and conditions: (a) p-TsN₂, K₂CO₃, CH₂CN, rt, 2 h; (b) Boc-Thr(bu)-CHO (25), CH₃OH, rt, overnight; (c) (CH₃)₂SiL, CH₂Cl₂, rt, 10 min; (d) Boc-Ala-CHO (27), CH₃OH, rt, overnight; (e) TFA:CH₂Cl₂ (1:1), 0 °C; (f) Boc-Thr(bu)-OH, HATU, DIPEA, DMF, rt, 3 h; (g) 20% piperidine in DMF; (h) propionic acid, EEDQ, DMF, rt, 2 h; (i) TFA:CH₂Cl₂:H₂O (47.5:47.5:5), rt, 2 h; (j) ethynylmagnesium chloride, THF, 0 °C, 1 h.
To compare the triazole compounds with current leading small-molecule PSD-95 PDZ inhibitors we synthesized Indole 1 and 2 (Fig. 1C) according to literature and tested them in the FP assay.

Among 9–18, compound 10 showed robust inhibition of the PDZ2/probe interaction with a \( K_i \) of 289 \( \mu \)M (Fig. 3). This affinity is ~8-fold lower than the TAV tripeptide (\( K_i = 38 \mu \)M) from which it was designed, but within the same affinity-range as SAV (\( K_i = 229 \mu \)M). Triazoles 11 and 13 showed some sign of inhibition (Fig. 3), but affinities were too low to reliably determine the \( K_i \) values.

Surprisingly, Indole 1 only demonstrated marginal inhibitory activity towards PDZ2, and a \( K_i \) value could not be determined due to a high bottom plateau. Indole 2 was found more potent and gave a credible inhibition curve and a \( K_i \) value of 530 \( \mu \)M towards PDZ2 (Fig. 3).

Next, we tested the active triazole 10 and reference compounds towards PDZ1 and PDZ3 of PSD-95 by FP in a similar way as for PDZ2. From previous studies it is known that PDZ1 recognizes the same peptide sequences as PDZ2 although generally with a lower affinity than PDZ2; while the selectivity profile for PDZ3 is very different from PDZ1 and PDZ2. Triazoles 11 and 13 showed some sign of inhibition (Fig. 3), but affinities were too low to reliably determine the \( K_i \) values.

Among 9–18, compound 10 showed robust inhibition of the PDZ2/probe interaction with a \( K_i \) of 289 \( \mu \)M (Fig. 3). This affinity is ~8-fold lower than the TAV tripeptide (\( K_i = 38 \mu \)M) from which it was designed, but within the same affinity-range as SAV (\( K_i = 229 \mu \)M). Triazoles 11 and 13 showed some sign of inhibition (Fig. 3), but affinities were too low to reliably determine the \( K_i \) values.

Surprisingly, Indole 1 only demonstrated marginal inhibitory activity towards PDZ2, and a \( K_i \) value could not be determined due to a high bottom plateau. Indole 2 was found more potent and gave a credible inhibition curve and a \( K_i \) value of 530 \( \mu \)M towards PDZ2 (Fig. 3).

Overall, these FP data demonstrate that the novel triazole-based peptidomimetic structure 10 binds PDZ2 of PSD-95 with a \( K_i \) value of 289 \( \mu \)M and can fully displace the PDZ2/probe interaction. The affinity to PDZ1 was much weaker (\( K_i > 500 \mu \)M) and no activity to PDZ3 was observed. Compared to tripeptides, 10 is equipotent to SAV and 8-fold weaker than TAV. On the other hand, 10 is several fold more potent than Indole 1 and about 2-fold more potent than Indole 2. Vogrig et al. demonstrated that Indole 1 inhibits the PSD-95/PDZ1/5-HT2A interaction with an IC50 of 190 \( \mu \)M in an in vitro pull-down assay. Indole 2 was not tested in this assay, as it was estimated to be a weak binder based on \(^1\)H-\(^1\)N HSQC NMR. Also, Indole 1 showed analgesic effects in a rat neuropathic pain model. However, based on our FP data these reported pharmacological effects might be mediated by other functions than a specific inhibition of PSD-95 PDZ domains.

In order to exclude any unconventional mechanism or artefactual events of the novel triazole compound we tested 10 by isothermal titration calorimetry (ITC). The ligands, TAV and 10, were titrated into PDZ2 under low c value conditions. The \( c \) value corresponds to the ratio of the receptor concentration and the dissociation constant and should normally be 10–100 to get high-quality data. However, for weak interactions this is not practical obtainable and instead experiments are run at low \( c \) values (often <1). In this alternative setup it is important that ligand concentration is several fold higher than protein concentration and that the concentration of ligand and receptor are precisely known (as stoichiometry information is lost). When properly designed the integrated heats form a hyperbolic graph from which the affinity can be reliably extracted. TAV titrated into PDZ2 resulted in substantial signal, and when integrated heats were fitted to a one-site binding model \( K_d \) was determined to 101 \( \mu \M \) (Fig. 4A). For 10, the measured heats were smaller but still resulted in a typical and reliable low \( c \) value thermogram and \( K_d \) of 598 \( \mu \M \) (Fig. 4B).

Overall, the ITC measurements confirm the FP data and corroborate that 10 binds specifically to the PDZ2 binding pocket strongly enough to produce measurable heat in ITC and displace a peptide probe in FP.

**Fig. 3** FP assay: triazoles and reference compounds tested for affinity to PDZ1 and PDZ2 of PSD-95. Cy5-GluN2B is used as probe at 50 nM; \([\text{PDZ2}] = 3.75 \mu \text{M}; [\text{PDZ1}] = 15 \mu \text{M}\). For PDZ2, the following \( K_i \) values were determined: 38 ± 9 \( \mu \text{M} \) (TAV), 229 ± 15 \( \mu \text{M} \) (SAV), 530 ± 96 \( \mu \text{M} \) (Indole 2) and 289 ± 41 \( \mu \text{M} \) (10). \( K_i \) values for Indole 1, 11 and 13 are >500 \( \mu \text{M} \). For PDZ1, the following \( K_i \) values were determined: 74 ± 8 \( \mu \text{M} \) (TAV) and 480 ± 160 \( \mu \text{M} \) (SAV). Error bars indicate SEM based on three individual measurements.
2.3. Computational evaluation

In order to explore how 10 could interact with PSD-95 PDZ2 we performed computational docking studies using Glide and a homology model of PDZ2 based on the crystal structure of PDZ3 in complex with the CRIPIT peptide (1BE9), as previously described.15,20,21 When docking penta- and tetra-peptides into this model these are positioned in the same conformation as CRIPIT in PDZ3 (see Fig. 1) and with an excellent agreement among the highest ranking docking poses. For TAV the suggested conformations also align very well with experimentally determined peptide-PDZ structures, e.g. Val is positioned in the P0 pocket and backbone interactions to PDZ are apparent (Fig. 1 and 5). The N-terminal amino acid, Thr in TAV, shows a higher degree of conformational variation among the poses, but most conformations suggest that Thr’s hydroxyl group interacts with the β-strand (βB) of PDZ while the N-terminal amine engages with His225 on the α-helix (B) as illustrated in Fig. 1 and 5. Docking poses of 10 suggest a binding mode very similar to TAV (Fig. 5). The P0 Val side chain binds in the hydrophobic P0 pocket of PDZ and the carboxylic acid of 10 interacts with the GLGF loop in all top 10 poses. Additionally, both 10 and TAV donate their P0 amide hydrogen to the PDZ backbone. Interestingly, in the top ranking pose of 10 (Fig. 5), the N3 triazole nitrogen accepts a hydrogen atom from a PDZ backbone amide bond similar to the carbonyl group of TAV’s P-2 amide illustrating the amide isosteric properties of triazole. Also, the top pose of 10 suggests that the P-2 Thr-residue binds PDZ in the same way as seen for TAV (Fig. 5). However, generally the poses of 10 agree less well in the P-2 position compared to TAV, and for example other poses of 10 suggest that its amine does not interact with PDZ2 at all. Perhaps this variation in the P-2 region reflects less tight interactions in that part of the molecule and could perhaps explain why TAV is more potent than 10. Overall, this computational analysis illustrates that 10 possesses suitable proportions and electronic properties allowing it to fit into the PDZ pocket, and that compound 10 is likely to bind PDZ2 in an similar mode as the tripeptide TAV.

3. Conclusions

In this study we have pursued a peptidomimetic strategy where we rigidify the original peptide backbone by introducing a small triazole group. The feasibility of the CuAAC reaction is exploited to make several close analogues in order to provide a thorough investigation of the strategy. The short tripeptide TAV was used as starting point for the design of triazole-based compounds. Azide and alkyne building blocks 1–8 were synthesized in satisfactorily yields and so were the corresponding triazole-based peptidomimetic structures 9–18. Compound 10 fully inhibited the PDZ2/peptide probe interaction with a Ki value of 289 μM. This affinity is lower than for TAV, but similar to the corresponding SAV tripeptide and two to several fold better than seen for the recently published indole-based small-molecule PSD-95 PDZ inhibitors (Indole 1–2). ITC studies of 10 show that its interaction with PDZ2 produces heat and gives a thermogram typical for a true, but low-affinity, binder. Finally, computational docking studies suggest that 10 binds PDZ2 in a binding mode very similar to TAV.

This study illustrates the challenges of generating non-peptide based PDZ inhibitors with reasonable potencies. When deviating from the peptide structure ligand-affinity is often lost, and as seen for the triazole-series even subtle changes dramatically affect affinity. Also, the affinities of reported indole small-molecules were found to be low in our hands. Still, based on our ‘click chemistry’ strategy we discovered compound 10, which represents a novel class of PSD-95 PDZ inhibitors. It demonstrated reliable inhibitory activity.
in our well-established FP and ITC assays and could inspire future efforts in the field of PDZ domain medicinal chemistry.

Notes and references

1 M. Sheng and C. C. Hoogenraad, Annu. Rev. Biochem., 2007, 76, 823.
2 K. S. Christopherson, B. J. Hillier, W. A. Lim and D. S. Bredt, J. Biol. Chem., 1999, 274, 27467.
3 M. Aarts, Y. Liu, L. Liu, S. Bessho, M. Arundine, J. W. Gurd, Y. T. Wang, M. W. Salter and M. Tymianski, Science, 2002, 298, 846.
4 A. Bach, B. H. Clausen, M. Møller, B. Vestergaard, C. N. Chi, A. Round, P. L. Sørensen, K. B. Nissen, J. S. Kastrup, M. Gajhede, P. Jemth, A. S. Kristensen, P. Lundstrøm, K. L. Lambertsen and K. Stromgaard, Proc. Natl. Acad. Sci. U. S. A., 2012, 109, 3317.
5 D. J. Cook, I. Teves and M. Tymianski, Nature, 2012, 483, 213.
6 F. Tao, Q. Su and R. A. Johns, Mol. Ther., 2008, 16, 1776.
7 B. W. LeBlanc, M. Iwata, A. P. Mallon, C. N. Rupasinghe, D. J. Goebel, J. Marshall, M. R. Spaller and C. Y. Saab, Neuroscience, 2010, 167, 490.
8 R. D’Mello, F. Marchand, S. Pezet, S. B. McMahon and A. H. Dickinson, Mol. Ther., 2011, 19, 1780.
9 L. M. Ittner, Y. D. Ke, F. Delerue, M. Bi, A. Gladbach, J. van Eersel, H. Wolfling, B. C. Chieng, M. J. Christie, I. A. Napier, A. Eckert, M. Staufenbiel, E. Hardeman and J. Gotz, Cell, 2010, 142, 387.
10 F. X. Soriano, M. A. Martel, S. Papadia, A. Vasin, P. Baxter, C. Rickman, J. Forder, M. Tymianski, R. Duncan, M. Aarts, P. Clarke, D. J. Wylie and G. E. Hardingham, J. Neurosci., 2008, 28, 10696.
11 M. D. Hill, R. H. Martin, D. Mikulis, J. H. Wong, F. L. Silver, K. G. Terbrugge, G. Milot, W. M. Clark, R. L. Macdonald, M. E. Kelly, M. Boulton, I. Fleetwood, C. McDougall, T. Gunnarsson, M. Chow, C. Lum, R. Dodd, J. Poublianc, T. Krings, A. M. Demchuk, M. Goyal, R. Anderson, J. Bishop, D. Garman, M. Tymianski and ENACT trial investigators, Lancet Neurol., 2012, 11, 942.
12 J. Schultz, R. R. Copley, T. Dero, C. P. Ponting and P. Bork, Nucleic Acids Res., 2000, 28, 231.
13 C. N. Chi, A. Bach, K. Stromgaard, S. Gianni and P. Jemth, BioFactors, 2012, 38, 338.
14 D. A. Doyle, A. Lee, J. Lewis, E. Kim, M. Sheng and R. MacKinnon, Cell, 1996, 85, 1067.
15 A. Bach, C. N. Chi, T. B. Olsen, S. W. Pedersen, M. U. Roder, G. F. Pang, R. P. Clausen, P. Jemth and K. Stromgaard, J. Med. Chem., 2008, 51, 6450.
16 M. A. Stiffler, J. R. Chen, V. P. Grantcharova, Y. Lei, D. Fuchs, J. E. Allen, L. A. Zaslavskaya and G. MacBeath, Science, 2007, 317, 364.
17 B. Z. Harris, F. W. Lau, N. Fujii, R. K. Guy and W. A. Lim, Biochemistry, 2003, 42, 2797.
18 S. W. Pedersen, S. B. Pedersen, L. Anker, G. Hultqvist, A. S. Kristensen, P. Jemth and K. Stromgaard, Nat. Commun., 2014, 5, 3215.
19 J. N. Eildal, G. Hultqvist, T. Balle, N. Stuhr-Hansen, S. Padrah, S. Gianni, K. Stromgaard and P. Jemth, J. Am. Chem. Soc., 2013, 135, 12998.
20 A. Bach, J. N. Eildal, N. Stuhr-Hansen, R. Deeskim, M. Gottschalk, S. W. Pedersen, A. S. Kristensen and K. Stromgaard, J. Med. Chem., 2011, 54, 1333.
21 A. Bach, C. N. Chi, G. F. Pang, L. Olsen, A. S. Kristensen, P. Jemth and K. Stromgaard, Angew. Chem., Int. Ed., 2009, 48, 9685.
22 A. Pisecchio, G. D. Salinas, T. Li, J. Marshall, M. R. Spaller and D. F. Mierke, Chem. Biol., 2004, 11, 469.
23 D. G. Udagamasonic, S. C. Sharma and M. R. Spaller, ChemBioChem, 2008, 9, 1387.
24 A. Vogrig, I. Dorr, N. Bouzidi, B. Boucherle, A. S. Wattiez, E. Cassier, G. Vallon, I. Ripoche, I. Arunbrarosa-Thomas, P. Marin, L. Nauton, V. Thery, C. Courteix, L. Y. Lian and S. Ducki, ACS Chem. Biol., 2013, 8, 2209.
25 C. W. Tornoe, C. Christensen and M. Meldal, J. Org. Chem., 2002, 67, 3057.
26 V. V. Rostovtsev, L. G. Green, V. V. Fokin and K. B. Sharpless, Angew. Chem., Int. Ed., 2002, 41, 2596.
27 Y. L. Angell and K. Burgess, Chem. Soc. Rev., 2007, 36, 1674.
28 A. Trabocchi and A. Guarina, in Peptidodinetics in Organic and Medicinal Chemistry, John Wiley & Sons, Ltd, 2014, ch. 5, pp. 99–121.
29 A. Brik, J. Alexandratos, Y. C. Lin, J. H. Elder, A. J. Olson, A. Wlodawer, D. S. Goodsell and C. H. Wong, ChemBioChem, 2005, 6, 1167.
30 I. E. Valverde, A. Bauman, C. A. Kluba, S. Vomstein, M. A. Walter and T. L. Mindt, Angew. Chem., Int. Ed., 2013, 52, 8957.
31 J. T. Lundquist and J. C. Pelletier, Org. Lett., 2001, 3, 781.
32 H. Johansson and D. S. Pedersen, Eur. J. Org. Chem., 2012, 4267.
33 A. Bach and K. Stromgaard, Synthesis, 2011, 807.
34 N. B. Kondekar and P. Kumar, Synth. Commun., 2011, 41, 1301.
35 S. Mosca, F. Wojcik and L. Hartmann, Macromol. Rapid Commun., 2011, 32, 197.
36 G. J. Roth, B. Leipold, S. G. Müller and H. J. Bestmann, Synthesis, 2004, 640.
37 G. Reginato, A. Mordini, F. Messina, A. Degi-Innocenti and G. Poli, Tetrahedron, 1996, 52, 10985.
38 P. K. Chakraverty, W. J. Greenlee, W. H. Parsons, A. A. Patchett, P. Combs, A. Roth, R. D. Busch and T. N. Mellin, J. Med. Chem., 1989, 32, 1886.
39 N. Umezawa, N. Matsumoto, S. Iwana, N. Kato and T. Higuchi, Bioorg. Med. Chem., 2010, 18, 6340.
40 T. Wiseman, S. Williston, J. F. Brandts and L. N. Lin, Anal. Biochem., 1989, 179, 131.
41 W. B. Turnbull and A. H. Daranas, J. Am. Chem. Soc., 2003, 125, 14859.