Comparative genomic analysis reveals ecological differentiation in the genus Carnobacterium

Iskandar, Christelle F.; Borges, Frédéric; Taminiau, Bernard; Daube, Georges; Zagorec, Monique; Remenant, Benoît; Leisner, Jørgen; Hansen, Martin Asser; Sørensen, Søren Johannes; Mangavel, Cécile; Cailliez-Grimal, Catherine; Revol-Junelles, Anne-Marie

Published in:
Frontiers in Microbiology

DOI:
10.3389/fmicb.2017.00357

Publication date:
2017

Document Version
Publisher's PDF, also known as Version of record

Citation for published version (APA):
Iskandar, C. F., Borges, F., Taminiau, B., Daube, G., Zagorec, M., Remenant, B., ... Revol-Junelles, A-M. (2017). Comparative genomic analysis reveals ecological differentiation in the genus Carnobacterium. DOI: 10.3389/fmicb.2017.00357
Comparative Genomic Analysis Reveals Ecological Differentiation in the Genus Carnobacterium

Christelle F. Iskandar1*, Frédéric Borges1*, Bernard Taminiau2, Georges Daube2, Monique Zagorec3, Benoît Remenant3†, Jørgen J. Leisner4, Martin A. Hansen5, Søren J. Sørensen5, Cécile Mangavel1, Catherine Cailliez-Grimal1 and Anne-Marie Revol-Junelles1

1 Laboratoire d’Ingénierie des Biomolécules, École Nationale Supérieure d'Agronomie et des Industries Alimentaires – Université de Lorraine, Vandoeuvre-lès-Nancy, France, 2 Laboratory of Food Microbiology, Department of Food Science, Fundamental and Applied Research for Animal and Health, University of Liège, Liège, Belgium, 3 UMR1014 SECALIM, INRA, Oniris, Nantes, France, 4 Department of Veterinary Disease Biology, Faculty of Health and Medical Sciences, University of Copenhagen, Frederiksberg, Denmark, 5 Molecular Microbial Ecology Group, University of Copenhagen, Copenhagen, Denmark

Lactic acid bacteria (LAB) differ in their ability to colonize food and animal-associated habitats: while some species are specialized and colonize a limited number of habitats, others are generalist and are able to colonize multiple animal-linked habitats. In the current study, Carnobacterium was used as a model genus to elucidate the genetic basis of these colonization differences. Analyses of 16S rRNA gene meta-barcoding data showed that C. maltaromaticum followed by C. divergens are the most prevalent species in foods derived from animals (meat, fish, dairy products), and in the gut. According to phylogenetic analyses, these two animal-adapted species belong to one of two deeply branched lineages. The second lineage contains species isolated from habitats where contact with animal is rare. Genome analyses revealed that members of the animal-adapted lineage harbor a larger secretome than members of the other lineage. The predicted cell-surface proteome is highly diversified in C. maltaromaticum and C. divergens with genes involved in adaptation to the animal milieu such as those encoding biopolymer hydrolytic enzymes, a heme uptake system, and biopolymer-binding adhesins. These species also exhibit genes for gut adaptation and respiration. In contrast, Carnobacterium species belonging to the second lineage encode a poorly diversified cell-surface proteome, lack genes for gut adaptation and are unable to respire. These results shed light on the important genomics traits required for adaptation to animal-linked habitats in generalist Carnobacterium.

Keywords: lactic acid bacteria, Carnobacterium, 16S meta-barcoding, comparative genomic analyses, ecological differentiation

INTRODUCTION

Lactic acid bacteria (LAB) include various genera and many species that have been investigated for decades because of their major role in food fermentations and their health benefit potential as probiotics. Understanding how LAB fitness can increase through evolution in these various habitats is critical for exploiting their beneficial properties. LAB are well-known for their ability
to colonize animal-derived food, i.e., meat, fish, and dairy products, and for being members of the gastrointestinal (GI) tract and the vagina microbiota (Douglas and Klaenhammer, 2010; Douillard and de Vos, 2014). LAB can be divided into specialized and generalist bacteria. Typically, the specialists that are used as starter cultures for a narrow range of fermented products are characterized by a low genetic diversity (Delorme et al., 2010). Their genomes exhibit signs of massive losses of genes involved in biosynthetic pathways (Douglas and Klaenhammer, 2010). To compensate the loss of these functions, genes encoding transporters for amino acids or carbohydrates were gained to allow growth in nutritional rich fermentation environments (Lorca et al., 2007). These genomic changes were accompanied by a specialization to food matrices, particularly exemplified in dairy strains. Other LAB also exhibit genomes characteristic of their ecological specialization. Lactobacillus iners is described to solely colonize the vaginal cavity and harbors one of the smallest LAB genomes presumably because its niche specialization allowed a substantial genome reduction (Macklaim et al., 2011; Mendes-Soares et al., 2014). Another example is the GI tract symbiont Lactobacillus reuteri, which is characterized by different lineages each one being apparently adapted to one particular vertebrate host: rodent or human (Frese et al., 2011). By contrast, some species are more generalist in their ability to colonize various environments and are therefore ubiquitous. Species from the genus Enterococcus can be found in the GI tract of animals and in a multitude of fermented foods (Santagati et al., 2012). Their adaptation to various environments is strongly linked to the presence in their genome of DNA acquired through horizontal gene transfer (HGT), resulting in a large pan genome (Santagati et al., 2012). Similarly, the genomes of some strains of the ubiquitous species Lactobacillus rhamnosus encode multiple lifestyle traits allowing them to reside in diverse habitats (Douillard et al., 2013). However, the interconnection between the ecology of these bacteria and their genomics is not fully understood and need further investigation. Importantly, the genomic traits responsible for adaptation to multiple animal-associated habitats are not clearly defined.

The LAB genera Enterococcus, Lactobacillus, Lactococcus, and Streptococcus have been intensively investigated, but lately, other genera including Carnobacterium have also drawn attention since 16S meta-barcoding studies have shown their significance in food (Chaillou et al., 2015; Duan et al., 2016; Fougy et al., 2016; Jääskeläinen et al., 2016). The genus Carnobacterium encompasses 11 species that have been isolated from cold and permanently cold fjords of the Arctic Ocean, and an oceanic trench, respectively (Lauro et al., 2007; Vogel et al., 2011; Leonard et al., 2013; Nicholson et al., 2015). Other species, including Carnobacterium maltaromaticum and C. divergens, have been found in animal-associated habitats. These two species are the most frequently isolated carnobacteria from various sources (Leisner et al., 2007) and belong to the dominant bacterial communities in meat and fish derived food (Chaillou et al., 2015; Duan et al., 2016; Fougy et al., 2016; Jääskeläinen et al., 2016). These bacteria are therefore very interesting models to investigate the genomic traits responsible for adaptation to animal-associated habitats.

In the genus Carnobacterium, the genome size ranges from 2.4 Mbp for C. inhibens subsp. gilichinskiyi WN1359 (Leonard et al., 2013), to 3.7 Mbp for C. maltaromaticum (Cailliez-Grimal et al., 2013). It has been suggested that the larger genome of the latter species is the basis of its success in colonizing various habitats (Leisner et al., 2007). However, it appears that genome size is not necessarily a predictor of the ability to occupy diverse habitats since the genome of C. divergens strains is relatively small (~2.7 Mbp), but this species is still able to colonize various habitats (Sun et al., 2015; Remenant et al., 2016).

The aim of this study was to investigate the ecological niches Carnobacterium species can occupy and to identify the genomic traits responsible for the high ecological success of some Carnobacterium species and the possible underlying adaptive mechanisms. For that purpose, we analyzed the relative abundance of Carnobacterium species using 16S rDNA metagenomic data. Subsequently, we compared the genomes of Carnobacterium strains isolated from different environments (mainly from cold aquatic habitats), and animal-associated habitats (live animal and foods).

MATERIALS AND METHODS

16S Meta-Barcoding Sequence Analysis

The data obtained from 681 samples of various ecological origins were analyzed with a focus on Carnobacterium. A database of V1–V3 16S rRNA gene sequence datasets available at the FARAH Institute (University of Liège, Belgium) was used to delineate the species composition of the Carnobacterium genus within four types of matrices: food products, animal samples, human and animal feces and environment. This database was built from 2010 to 2015 by merging the data obtained from several single projects hosted at the FARAH Institute. The datasets were produced as previously described (Rodriguez et al., 2015) by sequencing the V1–V3 16S rDNA hypervariable region with an MiSeq sequencer using v3 reagents (ILLUMINA, USA).

Sequence read processing was employed as previously described (Rodriguez et al., 2015) using the MOTHUR software package v1.35 (Schloss and Handelsman, 2003) and the UCHIME algorithm (Edgar et al., 2011) for alignment and OTU clustering (distance 0.03) and chimera detection, respectively. 16S gene sequence reference alignment and taxonomic assignation were
based upon the SILVA database (v1.15) of full-length 16S rDNA sequences.

For each sample, Operational Taxonomic Units (OTUs) belonging to the *Carnobacterium* genus were extracted. Corresponding reads were further assigned to *Carnobacterium* species using a local BLASTn algorithm vs. the SILVA database (v1.15). Reads were assigned to a defined species when identical to the best hit (four mismatches were allowed).

Statistical differences of the different species proportion inside each type of matrix were assessed with non-parametric Kruskal–Wallis test with Dunn’s post hoc tests using PRISM6 (GraphPad Software).

**Carnobacterium Genome Analysis**

The *Carnobacterium* genome sequences available at the start of this study included five *C. maltaromaticum*, one *C. inhibens* subsp. *gilichinskyi*, one *C. divergens*, and two *Carnobacterium* sp. genomes among which three were complete (*C. maltaromaticum* LMA28, *Carnobacterium* sp. 17A, and *C. inhibens* subsp. *gilichinskyi* WNI359). The strains originated from different ecological habitats, some from animal-derived food such as the *C. maltaromaticum* strains and *C. divergens* V41, and others from environmental samples (Table 1).

The genome sequences were integrated in the MicroScope platform (Vallenet et al., 2013) to perform automatic and expert annotation of the genes, as well as comparative analysis and secretome prediction by using the integrated SignalP software (Petersen et al., 2011). The gene phyloprofile tool interface was used for searching common or specific genes/regions between a query genome and other genomes or replicons chosen from the ones available in Prokaryotic Genome DataBase (PKGDB; i.e., (re)annotation of bacterial genomes) or complete proteome downloaded from the RefSeq/WGS sections.

The pan/core genome was calculated using two methods. The pan/core genome tool accessible in the comparative genomics section was used with MICFAM parameters of 50 or 80% amino acid (aa) sequence identity and 80% coverage. The Phyloprofile tool from the MicroScope platform was used with a cut-off of 70% aa identity and 80% coverage with the best Bidirectional Best Hit (BBH).

Synteny, defined as an orthologous gene set having the same local organization in species A and B, was determined as sequence similarity by BlastP BBH with at least 30% identity on 80% of the shortest sequence (minLlap 0.8) analyses and co-localization. Metabolic pathways were predicted using the Kyoto Encyclopedia of Genes and the Genomes (KEGG) resources (Kanehisa and Goto, 2000; Kanehisa et al., 2014) and the MetaCyc database (Caspi et al., 2014). A percentage of 70% minimum identity was used to detect the specific and common genes for *C. maltaromaticum*, excluding genes with 30% minimum identity with the four other *Carnobacterium* strains.

Neighbor-joining-based phylogenetic reconstruction was based on the nucleic acid sequence of 10 housekeeping genes (*dnaK, gyrA, polA, lepA, dnrA, gyrB, secA, ftsZ, recG, ielS*) and was performed using MEGA6 by using the Kimura two-parameter model, including transitions and transversions. The candidate tree was tested with 1,000 bootstrap replications (Tamura et al., 2013). The Sequence Type (MLST) was updated using e-BURST analysis from Rahman et al. (2014a). The resulting tree was rooted using the closely related species *Enterococcus faecalis* as outgroup.

The search for prophages was conducted with the PHAST Search Tool (Zhou et al., 2011).

**Data Availability**

The annotations were deposited at DDBJ/EMBL/GenBank under the following references: PRJEB9002 for *C. maltaromaticum* ML1.97, and PRJEB8756 for *C. maltaromaticum* 3.18. The annotations are publicly available for consultation in MicroScope1.

**RESULTS AND DISCUSSION**

**Prevalence of Different Carnobacterium Species in Various Ecological Niches**

Metagenomic data for genes encoding 16S rDNA from 681 samples were analyzed with a focus on the genus *Carnobacterium*. The samples were categorized as animal-derived food, animal organs, feces, and environment. Overall, the presence of representatives of the species *C. divergens*, *C. iners*, *C. inhibens*, *C. jeotgali*, *C. maltaromaticum*, *C. mobile*, *C. viridans*, and uncultured *Carnobacterium* sp was detected and their relative abundance is presented in Figure 1. The most abundant species was *C. maltaromaticum* accounting for 28–60% of *Carnobacterium* reads, followed by *C. divergens* (15–49%) and *Carnobacterium* spp. from lineages that have not yet been cultured and characterized (14–47% of *Carnobacterium* reads, depending on the sample origin). *C. inhibens*, *C. mobile*, and *C. viridans* accounted for lesser reads. *C. jeotgali* was detected only in environmental samples. *C. viridans* was detected only in food samples whereas *C. maltaromaticum* reads were observed in the samples from all origins. Other species were detected in the samples from two or three different habitats.

The highest species diversity was observed in food and organs (dog lungs, pig nympha1 nodes, pig stomach, cattle forestomach, and cattle spleen), with a maximum of eight species in food. The lowest diversity was observed in feces where only two species were recorded. In food and organs, the most abundant species were *C. maltaromaticum*, *C. divergens*, and uncultured *Carnobacterium* sp. The OTUs assigned to the species *C. maltaromaticum* accounted for 54% and 60% of the reads associated to the genus *Carnobacterium* in food and organs, respectively (Figure 1). Interestingly, in the feces only *C. maltaromaticum* and *C. divergens* sequences were detected, each accounting for about half of the reads. In contrast, *C. maltaromaticum* sequences represented 27% of the reads, while 47% of the reads were attributed to uncultured *Carnobacterium* sp. in environmental samples. *C. divergens* was not found in samples originating from the environment.

These results strongly suggest that *C. maltaromaticum* and to a lesser extend *C. divergens* are the most prevalent species of

---

1http://www.genoscope.cns.fr/agc/microscope/home/
TABLE 1 | Characteristics of Carnobacterium strains.

| Species          | Strain name | Origin and reference | Genome reference                          | Accession number         |
|------------------|-------------|----------------------|-------------------------------------------|--------------------------|
| C. divergens     | V41         | Fish viscera (Pilet et al., 1994) | Remenant et al., 2016                    | FLLU01000001 to FLLU01000032 |
| C. inhibens subsp. gilchinskyi | WN1359 | Siberian permafrost (Leonard et al., 2013) | Leonard et al., 2013                  | CP006812 to CP006817    |
| C. maltaromaticum | ATCC35586* | Diseased trout (Hui et al., 1984) | Leisner et al., 2012                      | NZ_AGNS00000000.1       |
|                  | LMA28*      | Soft ripened cheese (Millière et al., 1994) | Cailléz-Grimal et al., 2013             | HE999757.2               |
|                  | DSM20342 MXS* | Milk with malty flavor (Miller et al., 1974) |                             | NZ_JCMX00000000.1       |
|                  | 3.18*       | Pork meat product (Laursen et al., 2005) | Iskandar et al., 2016                   | PRJEB8756               |
|                  | ML.1.97*    | Fresh salmon (Laursen et al., 2005) | Iskandar et al., 2016                   | PRJEB9002               |
| Carnobacterium sp. | AT7        | Aleutian trench (Lauro et al., 2007) | Lauro et al., 2007                     | NZ_ABHH00000000.1       |
|                  | 17.4        | Cold seawater (Vogel et al., 2011) | Vogel et al., 2011                     | NC_015990.1, NC_015939.1 |

*Supplementary Figure S1.

this genus in habitats associated to animals. Although animal-associated habitats differ from each other, they also share common properties: they are nutrient-rich environments, they can induce similar stresses, they are associated with a dense and diversified microbiota and carbon is mainly available in the form of polymeric biomacromolecules. It can therefore be expected that bacteria, such as Carnobacterium species that are associated to animal habitats, share general properties and thereby highly contrast with other Carnobacterium species associated with the external environment.

Comparison of the General Genomic Features

In order to identify the adaptation factors responsible for the high ability of C. maltaromaticum and C. divergens to colonize multiple ecological niches including animal-associated habitats, a genome based analysis was conducted on nine Carnobacterium genomes (Table 1). The genome sizes of the five C. maltaromaticum strains were the largest, ranging from 3.33 to 3.87 Mbp; the other Carnobacterium genomes were smaller (2.35–2.74 Mbp), with the smallest being that of
C. inhibens subsp. gilichinskyi WN1359 (Table 2). Accordingly, the number of predicted CDS in each genome ranged from 3,368 to 3,812 for C. maltaromaticum, and from 2,268 to 2,633 for the other Carnobacterium (Table 2). C. maltaromaticum genomes harbored a lower GC% (34.4–34.5) than other genomes (35.2–35.3).

A phylogenetic tree based on 10 housekeeping genes was constructed. It shows that C. maltaromaticum and C. divergens on one hand, and Carnobacterium sp. 17.4, Carnobacterium sp. AT7, and C. inhibens subsp. gilichinskyi WN1359 on the other hand, are closely related. They form two monophyletic taxa sharing a common ancestor, as shown by the outgroup E. faecalis V583 (Figure 2).

The pan/core genome analysis of the nine Carnobacterium (cut-off of 50% aa identity and 80% coverage) showed that the core genome represents 1,130 (31%) of the predicted CDS. When comparison was restricted to C. inhibens subsp. gilichinskyi WN1359, Carnobacterium sp. 17-4, and Carnobacterium sp. AT7, the core genome increased up to 1,745–1,785 (67–73%) of the predicted CDS. Similarly, when the comparison was restricted to the 5 C. maltaromaticum strains and C. divergens V41, the core genome increased up to 1,723–1,825 (68–76%) of the predicted CDS. This result is in agreement with the phylogenetic tree showing close phylogenetic proximity of C. maltaromaticum and C. divergens on the one hand, and of C. inhibens subsp. gilichinskyi WN1359, Carnobacterium sp. 17-4, and Carnobacterium sp. AT7 on the other hand.

To be more restrictive, the pan/core genome analysis was performed with a cut-off of 70% aa identity and 80% coverage. It revealed that the strains of C. maltaromaticum share 2,665 CDS, and each of the five strains possesses between 279 and 644 specific genes showing strain-to-strain variations.

The gene repertoire of the different genomes was then compared to search for functions that differ between strains in order to identify the genomic traits that could explain the potential adaptation of C. maltaromaticum and C. divergens to animal-linked habitats.

CRISPR-cas and Prophages

Clustered Regularly Interspaced Short Palindromic Repeats (CRISPRs; Barrangou et al., 2007), and the CRISPR associated (cas) genes confer resistance to phage infection. The role and mechanism of the CRISPR-cas system in bacterial species have been extensively studied and indicate that the spacer sequences can be considered as a signature of past exposure to exogenous DNA. Among all Carnobacterium genomes, only C. divergens V41 possessed a CRISPR-cas system. More precisely, two loci were predicted in the genome of this strain. The presence of these genes suggests that C. divergens V41 may have systems acting as barriers against HGT, thereby that would limit genome expansion in this taxon.

One to four prophages and/or prophage remnants were detected in all analyzed genomes except in those of C. inhibens subsp. gilichinskyi and Carnobacterium sp. 17.4 (Table 2). Their size, ranged from 11.5 to 74.4 kbp (data not shown). More importantly, complete prophages were found only in the C. maltaromaticum (Table 2). The genomes of all Carnobacterium species but one - C. maltaromaticum- exhibit a genome of small or relatively small sizes. This suggests that the common ancestor of Carnobacterium exhibited a small size and that the ancestor of the species C. maltaromaticum experienced a massive gain of genes. Compared to C. divergens, no CRISPR-Cas systems were found in the genome of C. maltaromaticum. CRISPR-Cas provide an adaptive immunity against foreign DNA and is considered as a barrier against horizontal transfer (Barrangou et al., 2007).

The lack of such a barrier against DNA transfer could have favored the acquisition of genes in the C. maltaromaticum lineage. Consistently, complete prophages were found in the genomes of C. maltaromaticum while only remnants prophages were found in the genome of C. divergens. Similarly, Lactobacillus genomes devoid of CRISPR-Cas systems exhibited the trend of being more abundant in phage sequences (Sun et al., 2015).

LAB are mainly described as evolving by massive gene loss. The lineage C. maltaromaticum suggests that LAB may also evolve by a massive gene gain. Consistently, other ubiquitous LAB also exhibited large genomes that could be a result of similar mechanisms (Sun et al., 2015).

Secretome

Homologs of genes encoding the general secretion route (Sec-pathway) were present in all strains, while no homolog of the Twin-arginine translocation pathway (Tat-pathway) was found (data not shown). The secretome was therefore predicted by identifying genes suspected to encode signal peptide-containing proteins. The five C. maltaromaticum strains contained the largest secretome (319–375 proteins predicted to harbor a signal peptide), whereas C. divergens V41 presented an intermediate number (272 predicted proteins) compared to those predicted in Carnobacterium sp. AT7, Carnobacterium sp. 17.4, and C. inhibens subsp. gilichinskyi WN1359 with only 132–155 proteins containing a peptide signal. Compared to other LAB or genera that can share the same habitats, the sizes of secretomes of C. maltaromaticum and C. divergens are among the largest (Figure 3).

Among the COG (cluster of orthologous genes) represented in the secretome, families M (cell wall/membrane/envelope biogenesis), P (inorganic ion transport and metabolism), and R (general functional prediction only) tend to be more represented in C. maltaromaticum and C. divergens compared to the other species (Supplementary Figure S2). This suggests that extracellular functions are more diversified in C. maltaromaticum, and to a lesser extent in C. divergens, than in the other Carnobacterium. For instance, C. maltaromaticum and C. divergens exhibit between 22 and 35 genes in COG R, while the three other strains Carnobacterium sp. AT7, Carnobacterium sp. 17.4, and C. inhibens subsp. gilichinskyi WN1359, would have less than 10 genes encoding a signal peptide within this subclass.

Further, C. maltaromaticum predicted secretomes encompassed a higher number (26–30) of proteins belonging to COG family G (carbohydrate transport and metabolism) than was the case for C. divergens V41 (18) and other species (10–15). This is correlated with a higher content in genes encoding PTS transporters, between 62 and 68, in C. maltaromaticum.
Iskandar et al. Ecological Differentiation in Carnobacterium

TABLE 2 | General features of Carnobacterium genomes.

| Organisms                  | C. maltaromaticum | C. divergens | C. inhibens subsp. gilichinskyi | Carnobacterium sp. |
|----------------------------|-------------------|--------------|--------------------------------|-------------------|
|                            | LMA28             | DSM20342     | ATCC35586                      | ML.1.97           |
|                            | 3.65              | 3.877        | 3.54                           | 3.33              |
| Sequence length (Mbp)      | 3.33              | 3.57         | 3.44                           | 3.33              |
| GC content (%)             | 34.5              | 34.4         | 34.5                           | 34.4              |
| Number of plasmids         | 3                 | ND           | ND                             | ND                |
| Number of CDS             | 3.687             | 3.812        | 3.448                          | 3.368             |
| Number of COG             | 2.671             | 2.800        | 2.636                          | 2.639             |
| fCDS                      | 48                | 12           | 12                             | 49                |
| Prophage clusters          | 2*                | 2 (2)        | 1 (4)                          | 1 (5)             |
| Scaffolds                  | 1                 | 5            | 74                             | 229               |
| Contigs                    | 1                 | 5            | 74                             | 229               |
|                            | (3**)             |              |                                |                   |
|                            |                   |              |                                |                   |

* intact and ** region incomplete; ND, not determined.

FIGURE 2 | Phylogenetic tree of the Carnobacterium strains subjected to comparative genome analysis. The tree is based on the alignment of nucleic acid sequence of 10 housekeeping genes (dnaK, gyrA, polA, lepA, dnaB, gyrB, secA, ftsZ, recG, and ileS).

By contrast, the other Carnobacterium, including C. divergens, would contain less of such genes (between 27 and 43 depending on the strains). C. maltaromaticum strains, compared to other Carnobacterium species harbor a larger repertoire of PTS transporters. This characteristics is typical of ubiquitous LAB, as these transporters enable the bacteria to exploit a wide range of carbon sources (Douillard and de Vos, 2014).

More strikingly, C. maltaromaticum strains and C. divergens V41 secretomes encompassed between 116 and 155 proteins unclassified in COG families (class X Supplementary Figure S2), while this category was of minor importance (0–23) in other Carnobacterium species. The detailed analysis of those unclassified genes revealed that approximately half of them encode hypothetical proteins or conserved exported proteins of unknown function, and 30% encode secreted proteins associated with the cell wall. These results strongly suggest that C. maltaromaticum and C. divergens cell-surface structures differ significantly from those of Carnobacterium sp. AT7, Carnobacterium sp. 17.4, and C. inhibens subsp. gilichinskyi WN1359. Therefore, we focused on the comparison of the gene repertoire encoding such surface associated proteins.

Non-covalent Cell-Wall Bound Proteins

A larger set of proteins non-covalently bound to the cell wall was predicted for C. maltaromaticum and C. divergens V41 than for the other Carnobacterium. These proteins contain at least one LysM domain, and one WxL domain or SH3 domain (Table 3).

For all strains, at least one protein with an SH3 domain involved in peptidoglycan binding is predicted to be anchored to the cell wall (Table 3 and Supplementary Table S1), and between four and six LysM proteins. Two of them are conserved in all strains, three are found only in...
FIGURE 3 | Secretome size in the different Carnobacterium genomes. Other genomes belonging to lactic acid bacteria (LAB) (Lactococcus and Lactobacillus) and other Gram-positive species (Staphylococcus, Enterococcus, and Streptococcus) available at Microscope Mage were also included.

TABLE 3 | Number of sortases and surface proteins.

|                  | C. maltaromaticum | C. divergens | C. inhibens subsp. gilichinskyi | Carnobacterium sp. |
|------------------|-------------------|--------------|---------------------------------|-------------------|
|                  | LMA28             | DSM20342     | ATCC35586                        | 3.18              |
| LysM             | 6                 | 5            | 5                               | 5                 |
| WxL              | 48                | 49           | 37                              | 53                |
| SH3              | 3                 | 3            | 1                               | 2                 |
| SDP              | 35                | 35           | 28                              | 29                |
| Sortase          | 0                 | 0            | 0                               | 0                 |
| Total            | 98                | 100          | 77                              | 95                |

C. maltaromaticum and C. divergens V41, and three are variable within Carnobacterium sp. strains (Table 3 and Supplementary Table S1).

The signature of the 160–190 aa long WxL domain is characterized by two WxL motifs. WxL-containing proteins are non-covalently anchored proteins associated with the cell wall (Brinster et al., 2007). Strikingly, the number of WxL-containing proteins is comprised of between 37 and 53 and present only in C. maltaromaticum and C. divergens. Remarkably, no WxL-containing proteins were found to be encoded by the other Carnobacterium genomes. Twenty-nine genes encoding WxL proteins are common to the five C. maltaromaticum strains and C. divergens V41. Overall, 56 genes are variable within the strains (i.e., are absent in at least one strain), and three belong to the C. maltaromaticum core genome (Table 3 and Supplementary Table S1). The WxL proteins belong to the cell-surface complex (Csc) protein family. The Csc protein encoding genes are typically clustered and each cluster contains at least on copy of cscA, cscB, cscC, and cscD. Similarly, C. maltaromaticum and C. divergens contain between 13 and 17 csc clusters. Typically, CscA contains a DUF916 domain with extracellular matrix binding ability (Galloway-Peña et al., 2015) and a C-terminal transmembrane anchor, while CscB and CscC contain WxL domains, and CscD is a small LPXTG protein (Siezen et al., 2006). Similarly, in C. maltaromaticum and C. divergens, all the WxL encoding genes are either cscB or cscC homologs, and are localized in the vicinity of at least one WxL encoding gene. These cscB and cscC homologs can also be clustered with homologs of cscA and cscD. Strikingly, the clusters can encode a high number of WxL proteins, as exemplified by the cluster BN424_324-BN424_330 which is predicted to encode six WxL proteins.
Whereas C. maltaromaticum and C. divergens are predicted to produce a high diversity of non-covalently bound proteins, only a small number of such proteins were found in C. inhibens subsp. gilichinskiy WN1359, Carnobacterium sp. AT7, and Carnobacterium sp. 17. Among those, most are predicted as LysM- and SH3-containing proteins and no WxL proteins were detected (Table 3 and Supplementary Table S1).

Covalently Anchored Proteins
Sortase-dependent proteins (SDP) are covalently anchored to the cell wall, and possess an LPxTG like motif at their C-terminal end. SDP nomenclature refers to proteins attached to the peptidoglycan by the sortase family of transpeptidases (Schneewind and Missiakas, 2014). Such SDP were found in C. maltaromaticum strains and C. divergens V41 whereas only one LPxTG domain protein was detected in Carnobacterium sp. 17.4 and Carnobacterium sp. AT7, and none in C. inhibens subsp. gilichinski WN1359 (Table 3 and Supplementary Table S1).

Sortases decorate the surfaces of Gram-positive bacteria with diverse proteins that enable microbes to interact with their environment (Comfort and Clubb, 2004; Maresso and Schneewind, 2008). The five C. maltaromaticum strains and C. divergens V41 possess many putative sortase A and B genes, while Carnobacterium sp. AT7 and Carnobacterium sp. 17.4 harbor only one and C. inhibens subsp. gilichinski WN1359 possesses a pseudogene that might encode a remnant protein with similarities with sortases (Table 3 and Supplementary Table S1).

Depending on the strains, the C. maltaromaticum genomes are predicted to encode 26–35 SDP, and 26 putative SDP could be predicted in C. divergens. Among those, 13 belong to the C. maltaromaticum core genome, including seven also conserved in C. divergens. In addition, 24 C. maltaromaticum SDP-encoding genes are strain specific or shared by only some of the strains. The differences between strains resulted either from the absence of homologs or the presence of 373 predicted pseudogenes. The predicted sortase-encoding gene in the genome of Carnobacterium sp 17.4 is located next to a collagen-binding surface protein encoding gene (ABHHv1_120049, Supplementary Table S1).

Functions of the Surface Proteins
As C. maltaromaticum and, to a lesser extent, C. divergens presented a large panel of surface-exposed proteins compared to other Carnobacterium species, we searched for the putative functions of the C. maltaromaticum and C. divergens species specific proteins to illuminate the potential benefits they might provide to these two species.

Enzymes
Homologs of multidomain proteins predicted as nucleotidases/metallophosphatases as well as PrtB homologs are among the proteins conserved in C. maltaromaticum and C. divergens (Figure 4 and Supplementary Table S1). Extracellular 5’-nucleotidase domains that catalyze dephosphorylation of exogenous adenosine 5’-nucleotides to adenosine and phosphate (Bengis-Garber and Kushner, 1982) are reported as providing a key function for phosphorous regeneration in aquatic ecosystems (Ammerman and Azam, 1985).

In the dairy LAB Lactobacillus delbrueckii subsp. bulgaricus, PrtB, a cell envelope-associated protease (CEP) has been shown to degrade casein into peptides (Siezen, 1999). Peptides and aas are subsequently internalized and peptides are further hydrolyzed by intracellular peptidases into small peptides and free aa (Savijoki et al., 2006). The analysis of the genomes of C. maltaromaticum and C. divergens revealed the conserved presence of oligopeptides transporter systems OppABCDF and DtpT, as well as intracellular peptidases (Supplementary Table S1). Interestingly, homologs of the Opp system and of intracellular peptidases were also found in Carnobacterium sp. 17.4, Carnobacterium sp. AT7, and C. inhibens subsp. gilichinskiy WN1359 but no homologs of CEP and no DtpT. In the dairy environment, CEP are believed to confer a selective advantage by allowing bacteria to exploit the aa from milk casein (Kunji et al., 1996; Christensen et al., 1999; Doeven et al., 2005; Savijoki et al., 2006). All these data strongly suggest that among Carnobacterium only C. maltaromaticum and C. divergens are indeed able to exploit aa from the proteins present in their environments. The presence of PrtB would be a selective advantage for these two species in protein-rich environments such as food.

Among the SDP proteins present in several C. maltaromaticum strains we noticed a CDS predicted as harboring a glycoside hydrolase activity (BN424_641, Figure 4 and Supplementary Table S1). This multidomain protein also contain a fibronectin type III-like module of unknown function, which is usually associated with glycoside hydrolase domains (Alahuhta et al., 2010). Such enzymatic activity could allow C. maltaromaticum to degrade extracellular carbohydrate polymers. Interestingly, this protein is also predicted to contain a mucin-binding domain. Mucin is the major component of the intestinal mucus. It is tempting to speculate that the putative mucin-binding glycoside hydrolase of C. maltaromaticum would hydrolyze mucin glycan moieties as has been described for some gut bacteria (Tailford et al., 2015).

Nutrient Uptake: Heme Compounds
Two SDP, predicted as heme-binding proteins in C. maltaromaticum and C. divergens, are homologs of IsdA and IsdC (Figure 5 and Supplementary Table S1). The Isd system in Staphylococcus aureus enables to utilize different sources of heme: free heme, heme bound to free hemoglobin, and heme bound to hemoglobin interacting with haptoglobin. The Isd ABC-transport system in S. aureus is encoded by isdABCD; EFGH, IsdH, IsdB, and IsdA acting as cell wall anchored receptor proteins: IsdH is the primary haptoglobin-hemoglobin receptor, IsdB the primary hemoglobin receptor, and IsdA can bind free heme or accept heme from IsdB or IsdH. Heme is subsequently transferred from IsdA to IsdC, another cell wall protein and then to a membrane associated transporter, formed by IsdD, IsdE, and IsdF. After internalization, heme is taken up by the heme degrading monooxygenase IsdG that releases the resulting iron in the cytoplasm (Choby and Skaar, 2016). At first glance, among the possible protein candidates able to bind heme from the environment, C. maltaromaticum and C. divergens would
only have IsdA as no ortholog of IsdB and IsdH were found. This would suggest that these two Carnobacterium species would primarily be able to use free heme as iron source and not heme bound to proteins. However, the IsdA homolog in C. maltaromaticum is predicted to contain four near-iron transporter (NEAT) domains while IsdA from S. aureus contains only two. NEAT domains bind heme compounds or proteins containing heme compound (Gaudin et al., 2011; Balderas et al., 2012). It could therefore be speculated that the presence of two additional NEAT domains in the C. maltaromaticum IsdA homolog might compensate the absence of IsdB.

Carnobacterium divergens and C. maltaromaticum do not exhibit any IsdD homolog. However, we found two flhC homologs which encode an ATP-binding component of ABC transporter as well. These FlhC homologs could play the equivalent role of IsdD and thus build a functional ABC transporter with the permease encoded by the IsdF homolog. In C. divergens V41, one flhC is localized in the vicinity of the putative isd genes, while in the C. maltaromaticum strains, this homolog is localized elsewhere in the genome. In addition, all the analyzed genomes of Carnobacterium contain an isdG homolog and would be able to release iron after heme import in the cytoplasm. The isdA homolog of C. maltaromaticum ML1.97 and 3.18 appear as a pseudogene, indicating that this system is not fully functional in these two strains. Overall, these analyses suggest C. divergens V41 and several strains if not all of C. maltaromaticum would be able to use extracellular heme, for respiration (see below) and/or as an iron source.

Microbial Adhesion

The previous analysis of the genome of C. maltaromaticum LMA28 reported the presence of putative cell-surface adhesins (Rahman et al., 2014b). Comparison of Carnobacterium genomes revealed that C. maltaromaticum strains and C. divergens V41 may produce several surface proteins predicted to contain domains reminiscent of adhesion function: collagen-binding, mucBP, Leucine-Rich Repeat (LRR).

Among those, 10 LPxTG proteins are predicted to have a collagen-binding domain (Supplementary Table S1 and Figure 4). Functional domains are annotated collagen-binding protein, Cna, or bacterial adhesin. Collagen-binding proteins found in S. aureus (Elasri et al., 2002) and Listeria monocytogenes (Bierne and Cossart, 2007) are suggested to participate in the infection process. However, proteins putatively involved in adhesion to collagen and mucin have also been reported to be important for the probiotic properties of LAB as shown in Lactobacillus plantarum WCFS1 (Boekhorst et al., 2006b). It is therefore difficult to predict if the binding capacity of such proteins in Carnobacterium can be considered as beneficial or not.

Three LPxTG proteins (Figure 4 and Supplementary Table S2) contain a MucBP (mucin-binding protein) domain. MucBP domains allow adhesion to mucus material (Lučić et al., 2012). Lactobacillales and Listeria species can possess between 1 and 14 MucBP-containing proteins (Boekhorst et al., 2006a; Bierne and Cossart, 2007).
or three genes encoding the pilus subunits are organized into
(Burgain et al., 2014). In general in Gram-positive bacteria, two
matrix in the probiotic strain
*L. rhamnosus*
described as surface components promoting adhesion to dairy
to be a lineage well-adapted to the dairy environment. Pili were
Both strains belong to clonal complex CC1, which is suspected
only in one other
*C. maltaromaticum*
LMA28 (Rahman et al., 2014b), were found
strains and absent in other
*Carnobacterium*
*C. maltaromaticum*
Of these 19 proteins, two are conserved in all
*C. maltaromaticum*
LRR-containing proteins which exhibit a WxL binding domain.
Overall, 19 putative adhesins were predicted from the genomes
to intestinal mucosa and extracellular matrices of animals.
In *Enterococcus faecium*, the WxL protein SwpA and the
protein DufA, which contains a DUF916 domain and is encoded
by a gene that belong to a csc cluster, are collagen and fibronectin-
binding proteins (Galloway-Peña et al., 2015). Similarly, it can
be hypothesized that at least some WxL and DUF916 likely
encoded by *C. divergens* and *C. maltaromaticum* might exhibit
similar adhesion properties. In addition, two LRR domains
were found in some strains of *C. maltaromaticum* holding a
WxL anchorage domain (Figure 4 and Supplementary Table
S2). These domains are involved in protein–protein interactions.
They are described to be associated with domains exhibiting an
Ig-like (immunoglobulin) fold, the function of which is
thought to facilitate the presentation of the adjacent LRR domain
(Bierne and Cossart, 2007). Accordingly, near the LRR domain
of the putative *C. maltaromaticum* LMA28 surface protein
BN424_3043, an Ig-like domain was found at the C-terminal
end of the LRR region (Figure 4 and Supplementary Table
S2). In *L. monocytogenes*, internalins associated to virulence are
WxL-containing proteins. LRR-containing proteins are rather
uncommon in *Lactobacillus*.
The presence of such putative adhesins suggests that
*C. maltaromaticum* and *C. divergens* exhibit the ability to adhere
to intestinal mucosa and extracellular matrices of animals.
Overall, 19 putative adhesins were predicted from the genomes
of *C. maltaromaticum* and *C. divergens* and absent from the other
*Carnobacterium* species. All these proteins are SDP except two
LRR-containing proteins which exhibit a WxL binding domain.
Of these 19 proteins, two are conserved in all *C. maltaromaticum*
strains and absent in other *Carnobacterium*: a putative collagen-
binding SDP and a mucin-binding protein.

Genes encoding the pili proteins previously described for
*C. maltaromaticum* LMA28 (Rahman et al., 2014b), were found
only in one other *C. maltaromaticum* strain (DSM20342 MX5).
Both strains belong to clonal complex CC1, which is suspected
to be a lineage well-adapted to the dairy environment. Pili were
described as surface components promoting adhesion to dairy
matrix in the probiotic strain *L. rhamnosus* GG and thereby
they might contribute to confer an advantage in dairy products
(Burgain et al., 2014). In general in Gram-positive bacteria, two
or three genes encoding the pilus subunits are organized into
an operon, along with at least one sortase gene (Mandlik et al.,
2008; Proft and Baker, 2009). The closest homologs of such
*C. maltaromaticum* genes were found in *E. faecalis*, with also a
highly similar genetic organization (Figure 6). However, pilin
gene organization is different between *C. maltaromaticum* and
*L. rhamnosus* GG suggesting that the genetic structures of pili loci
in *C. maltaromaticum*, *E. faecalis* V583, and *L. rhamnosus* GG are
the result of different gene rearrangements.

Overall, these comparative genomic analyses suggest that all
*C. maltaromaticum* strains and *C. divergens* might have adhesive
properties and that strains might exhibit differences in this
regard.

Further, these analyses demonstrated striking differences
between the group *C. maltaromaticum/C. divergens* and the other
*Carnobacterium* spp. The secretome and more specifically the
cell-surface proteome of *C. maltaromaticum* and *C. divergens*
are large, the one of *C. maltaromaticum* being the largest
described for LAB as previously noticed by Sun et al. (2015).
Conversely the secretome of *C. inhibens* subsp. *gilichinskyi*
WN1359, *Carnobacterium* sp. AT7, and *Carnobacterium* sp.
17.4 is the smallest among LAB. The detailed analysis of the
functions provided by such large secretome supports a
prediction that the cell-surface proteome would confer the
ability to hydrolyze and to adhere to biomacromolecules as
well as to capture biomolecules (heme compounds). The cell
surface of *C. maltaromaticum* and *C. divergens* might closely
interact with animal environments and use these nutrient-rich
substrates. In addition, *C. maltaromaticum* would likely exhibit
a larger repertoire of hydrolytic enzymes and adhesins that
may enable *C. maltaromaticum* to adapt to multiple habitats.
By contrast, the other *Carnobacterium* lack such a cell-surface
proteome and would therefore be expected to be less able to
colonize animal-linked habitats. The size of the secretome is
highly variable in LAB. Interestingly, all known specialized LAB
*Streptococcus thermophilus*, *L. iners*, *L. reuteri*, *L. sanfranciscensis*,
and *L. fructivorans* are characterized by a small secretome,
suggesting that colonizing one specific niche, such as dairy,
vagina, sourdough, or the GI tract, respectively, does not require
a large secretome. Consistently, the known generalist LAB such
as *L. rhamnosus*, *L. plantarum* (Martino et al., 2016), and some

---

**FIGURE 5** Genes encoding heme uptake system, cluster organization, and synteny between *C. maltaromaticum* LMA28 and *Staphylococcus aureus* 16K.
Enterococcus spp. exhibit a large secretome. We propose that there is an intimate relationship between the secretome size and the ability of LAB to colonize diverse habitats. According to this hypothesis, the bigger a secretome, the higher the capacity to colonize multiple environments.

Respiration
Carnobacterium maltaromaticum is unable to synthesize heme and exhibits better growth efficiency in the presence of hematin, suggesting that heme is used by C. maltaromaticum to respire oxygen. Consistent with this hypothesis, C. maltaromaticum has been reported to produce cytochrome b and d types when grown aerobically with hematin (Meisel et al., 1994). The gene repertoire of C. maltaromaticum suggests the ability to produce a functional respiratory chain. Indeed, the electron donor-encoding genes, noxB and ndh, and those required for the synthesis of the electron shuttle menaquinone (yhdB, menE, menB, menD, menF, ispA, ispB, and menA) were present. Further, cydABCD, encoding the heme-dependent cytochrome quinol oxidase that performs the final electron transfer to the acceptor oxygen (Lechardeur et al., 2011), were also found. These genes were present in all C. maltaromaticum strains and in C. divergens, except in C. maltaromaticum ML.1.97 whose menF appears to be a pseudogene. Therefore, this last strain might require the presence of quinone in the environment to respire, as reported for Streptococcus agalactiae (Rezaïki et al., 2008). It seems therefore that the components of the cell wall proteome involved in extracellular heme utilization may also contribute to oxygen respiration in C. maltaromaticum and C. divergens.

By contrast, only menA, noxA/noxB, and ispB were found in Carnobacterium sp. AT7, Carnobacterium sp. 17.4, and Carnobacterium inhibens subsp. gilichinskiyi WN1359 strongly suggesting that these three bacteria do not possess any functional respiratory chain.

Adaptation to the GI Tract
Bacteria have to deal with several stresses in order to survive in the GI tract, including the immune system and bile. All strains of C. maltaromaticum and C. divergens contain genes conferring resistance to some components of the immune system. Indeed, homologs were found for mprF, dltABCD, asnH/asnB, oatA, and pgdA (Supplementary Table S2). The genes mprF and dltABCD are involved in phospholipid lysinylation and teichoic acid D-alanylation, respectively, and thus confer resistance to antimicrobial peptides of the innate immune system by protecting the cell wall. The genes oatA and pgdA confer to peptidoglycan a high resistance to lysozyme, another component of the innate immune system, by introducing O-acetylation and N-deacetylation, respectively. Importantly, no mprF, dltABCD, asnH/asnB, oatA, and pgdA homologs were found in the genomes of Carnobacterium sp. AT7, Carnobacterium sp. 17.4, and Carnobacterium inhibens subsp. gilichinskiyi WN1359.

Bile salt hydrolase encoding genes were conserved in all strains of C. maltaromaticum while none were found in other Carnobacteria including C. divergens V41 (Supplementary Table S2).

Carnobacterium maltaromaticum and C. divergens are thus highly contrasting from the three other Carnobacterium taxa by their content of genes described in Lactobacillus as key factor for survival in the GI tract of animals (Kleerebezem et al., 2010). Indeed, while C. maltaromaticum and C. divergens possess genes putatively conferring resistance against the innate immune system, almost none of these homologs were found in Carnobacterium sp. AT7, Carnobacterium sp. 17.4, and C. inhibens subsp. gilichinskiyi WN1359. This might explain why the only two species identified in feces samples are C. maltaromaticum and C. divergens. However, surprisingly, genes allowing resistance to bile were only found in C. maltaromaticum and not in C. divergens. Yet the ability to hydrolyze bile is described as a key factor for colonization of the gut (Kleerebezem et al., 2010; Seedorf et al., 2014). Indeed C. maltaromaticum LMA28, a cheese isolate that possesses such an ability has not yet been tested in a mouse model (Rahman et al., 2014b; Sun et al., 2015). However, such ability has not yet been tested in C. divergens V41. Whether these two species differ in their ability to deal with bile and the ecological consequences it has on GI tract survival deserves further investigation.

AUTHOR CONTRIBUTIONS
Performed 16S meta-barcoding: BT and GD. Performed whole genome sequencing and assembly: JL, MH, and SS. Comparative genome analyses: CI, CC-G, FB, A-MR-J, MZ, and BR. Wrote the manuscript: CI, CC-G, BT, MZ, BR, JL, CM, FB, and A-MR-J. Coordinated the study: FB.
ACKNOWLEDGMENTS

The LABGeM (CEA/IG/Genoscope & CNRS UMR8030) and the France Génomique National infrastructure (funded as part of Investissement d’avenir program managed by Agence Nationale pour la Recherche, contract ANR-10-INBS-09) are acknowledged for support within the MicroScope annotation platform. They are also thankful to Myriam Michelle, Sylvie Wolff, Arnaud Khemisti, and Camille Collin for their technical support during this study. The authors would like to thank the reviewers for their helpful and constructive comments that greatly contributed to improving the final version of the paper.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: http://journal.frontiersin.org/article/10.3389/fmicb.2017.00357/full#supplementary-material

REFERENCES

Alahuhta, M., Xu, Q., Brunecky, R., Adney, W. S., Ding, S.-Y., Himmel, M. E., et al. (2010). Structure of a fibronectin type III-like module from Clostridium thermocellum. Acta Cryst. F 66, 878–880. doi: 10.1107/S1744309110022529

Ammerman, J. W., and Azam, F. (1985). Bacterial 5-nucleotidase in aquatic ecosystems: a novel mechanism of phosphorus regeneration. Science 227, 1338–1340. doi: 10.1126/science.227.4692.1338

Balderas, M. A., Nobles, C. L., Honza, E. S., Alicci, E. R., and Mareso, A. W. (2012). Hal is a Bacillus anthracis heme acquisition protein. J. Bacteriol. 194, 5513–5521. doi: 10.1128/JB.00685-12

Barrangou, R., Fremaux, C., Deveau, H., Richards, M., Boyaval, P., Moineau, S., et al. (2007). CRISPR provides acquired resistance against viruses in prokaryotes. Science 315, 1709–1712. doi: 10.1126/science.1138140

Bengt-Garber, C., and Kushner, D. J. (1982). Role of membrane-bound 5'-nucleotidase in nucleotide uptake by the moderate halophile Vibrio costalisa. J. Bacteriol. 149, 808–815.

Bierne, H., and Cossart, P. (2007). Listeria monocytogenes surface proteins: from genome predictions to function. Microbiol. Mol. Biol. Rev. 71, 377–397. doi: 10.1128/MMBR.00039-06

Boekhorst, J., Helmer, Q., Kleerebezem, M., and Siezen, R. J. (2012). UCHIME improves sensitivity and speed of chimera detection. Bioinformatics 28, 355–361. doi: 10.1093/bioinformatics/btr636

Douillard, F. P., and de Vos, W. M. (2014). Functional genomics of lactic acid bacteria: from food to health. Microb. Cell Fact. 13:58. doi: 10.1186/1475-2859-13-58

Douillard, F. P., Ribbera, A., Kant, R., Pietila, T. E., Jarvinen, H. M., Messing, M., et al. (2013). Comparative genomic and functional analysis of 100 Lactobacillus rhamnosus strains and their comparison with strain GG. PLoS Genet. 9:e1003683. doi: 10.1371/journal.pgen.1003683

Duan, S., Hu, X., Li, M., Miao, J., Du, J., and Wu, R. (2016). Composition and metabolic activities of the bacterial community in shrimp sauce at the flavor-forming stage of fermentation as revealed by metatranscriptome and 16S rRNA gene sequencing. J. Agric. Food Chem. 64, 2591–2603. doi: 10.1021/acs.jafc.5b03826

Edgar, R. C., Haas, B. J., Clemente, J. C., Quince, C., and Knight, R. (2011). UCHIME improves sensitivity and speed of chimera detection. Bioinformatics 27, 1947–1953. doi: 10.1093/bioinformatics/btr381

Elasri, M. O., Thomas, J. R., Skinner, R. A., Blevins, J. S., Beenen, K. E., Nelson, C. L., et al. (2002). Staphylococcus aureus collagen adhesin contributes to the pathogenesis of osteomyelitis. Bone 30, 275–280. doi: 10.1016/S8756-3282(01)00632-9

Fougy, L., Desmonts, M.-H., Cooret, G., Fassal, C., Hamon, E., Hézard, B., et al. (2016). Reducing salt in raw pork sausages increases spoilage and correlates with reduced bacterial diversity. Appl. Environ. Microbiol. 82, 3928–3939. doi: 10.1128/AEM.03233-16

Frese, S. A., Benson, A. K., Tannock, G. W., Loach, D. M., Kim, J., Zhang, M., et al. (2011). The evolution of host specialization in the vertebrate gut symbiont Lactobacillus reuteri. PLOS Genet. 7:e1001314. doi: 10.1371/journal.pgen.1001314

Galloway-Peña, J. R., Liang, X., Singh, K. V., Yadav, P., Chang, C., Rosa, S. L., et al. (2015). The identification and functional characterization of WxL proteins from Enterococcus faecium reveal surface proteins involved in extracellular matrix interactions. J. Bacteriol. 197, 882–892. doi: 10.1128/JB.02288-14

Hou, S.-P., Holt, R. A., Srinangathan, N., Seidler, R. J., and Fryer, J. L. (1984). Lactobacillus piscicola, a new species from salmonid fish. Int. J. Syst. Bacteriol. 34, 393–400. doi: 10.1099/00207713-4-3-393

Iskandar, C. F., Cailliez-Grimal, C., Rahman, A., Rondags, E., Remenant, B., Zagorec, M., et al. (2016). Genes associated to lactose metabolism illustrate the high diversity of Carnobacterium maltaromaticum. Food Microbiol. 58, 79–86. doi: 10.1016/j.fm.2016.03.008
Sun, C., Fukui, H., Harra, K., Kitayama, Y., Eda, H., Yang, M., et al. (2015). Expression of Reg family genes in the gastrointestinal tract of mice treated with indomethacin. *Am. J. Physiol. Gastrointest. Liver Physiol.* 308, G736–G744. doi: 10.1152/ajpgi.00362.2014

Tailford, L. E., Crost, E. H., Kavanaugh, D., and Juge, N. (2015). Mucin glycan foraging in the human gut microbiome. *Front. Genet.* 6:81. doi: 10.3389/fgene.2015.00081

Tamura, K., Stecher, G., Peterson, D., Filipski, A., and Kumar, S. (2013). MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol. Biol. Evol.* 30, 2725–2729. doi: 10.1093/molbev/mst197

Vallenet, D., Belda, E., Calteau, A., Cruveiller, S., Engelen, S., Lajus, A., et al. (2013). MicroScope—an integrated microbial resource for the curation and comparative analysis of genomic and metabolic data. *Nucleic Acids Res.* 41, D636–D647. doi: 10.1093/nar/gks1194

Voget, S., Klippel, B., Daniel, R., and Antranikian, G. (2011). Complete genome sequence of *Carnobacterium* sp 17-4. *J. Bacteriol.* 193, 3403–3404. doi: 10.1128/JB.05113-11

Zhou, Y., Liang, Y., Lynch, K. H., Dennis, J. J., and Wishart, D. S. (2011). PHAST: a fast phage search tool. *Nucleic Acids Res.* 39, W347–W352. doi: 10.1093/nar/gkr485

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Copyright © 2017 Iskandar, Borges, Taminiau, Daube, Zagorec, Remenant, Leisner, Hansen, Sørensen, Mangavel, Cailléz-Grimal and Revol-Junelles. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.**