**Toxoplasma gondii** B1 Gene Detection in Feces of Stray Cats around Seoul, Korea and Genotype Analysis of Two Laboratory-Passaged Isolates

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Abstract: The increasing prevalence of *Toxoplasma gondii* infection in the human population in the Republic of Korea (= Korea) is due to various reasons such as an increase in meat consumption. However, the importance of cats in transmitting *T. gondii* infection through oocysts to humans has seldom been assessed. A total of 300 fecal samples of stray cats captured around Seoul from June to August 2013 were examined for *T. gondii* B1 gene (indicating the presence of oocysts) using nested-PCR. Fourteen (4.7%) of 300 cats examined were positive for B1 gene. Female cats (7.5%) showed a higher prevalence than male cats (1.4%). Cats younger than 3 months (5.5%) showed a higher prevalence than cats (1.5%) older than 3 months. For laboratory passage of the positive samples, the fecal suspension (0.2 ml) of B1 gene positive cats was orally inoculated into experimental mice. Brain tissues of the mice were obtained after 40 days and examined for the presence of tissue cysts. Two isolates were successfully passaged (designated KNIH-1 and KNIH-2) and were molecularly analyzed using the SAG5D and SAG5E gene sequences. The SAG5D and SAG5E gene sequences showed high homologies with the ME49 strain (less virulent strain). The results indicated the importance of stray cats in transmitting *T. gondii* to humans in Korea, as revealed by detection of B1 gene in fecal samples. *T. gondii* isolates from cats were successfully passaged in the laboratory for the first time in Korea.

Key words: Toxoplasma gondii, stray cat, nested-PCR, prevalence, genotype

INTRODUCTION

*Toxoplasma gondii* is an intracellular protozoan parasite that can infect warm-blooded animals, including humans. Toxoplasmosis is an important clinical disease worldwide and can lead to lymphadenitis, encephalitis, retinochoroiditis, congenital infection, and stillbirth [1]. *T. gondii* can cause infection via ingestion of tissue cysts or tachyzoites in raw or undercooked meat of infected animals, or ingestion of oocysts in the water or soil contaminated with feces of infected cats [2].

The prevalence of human toxoplasmosis in the Republic of Korea (= Korea) is increasing due to various factors. Increase in meat consumption is an important reason. Also, individuals with occupations requiring soil contact in environments frequented by cats are significantly more likely to contract toxoplasmosis [2]. However, a more significant risk factor is direct contact with cats, the definitive host of *T. gondii*. Epidemiologic surveys of *T. gondii* infection in sera of stray and household cats have been reported in several areas of Korea [3-8].

*T. gondii* strains isolated from Europe and North America belong to 3 distinct clonal lineages (genotypes I, II, and III) that differ in phenotype, including the pathogenicity [9]. Dubey et al. [10] recently found the 4th clonal lineage (genotype 12) from wild life of North America. They found that 85% of different strains in North America were 1 of the 3 widespread genotypes including genotype II, genotype III, and genotype 12 [10]. With the exception of a few reports, there has been little information about *T. gondii* strains and isolates in Korea. Only 1 long-term laboratory-passaged Korean isolate (KI-1) that was originally isolated from an ocular toxoplasmosis patient is available [11]. The gene sequences of KI-1 were highly homologous with RH, and thus KI-1 has been included...
in the genotype I [12]. In addition, sequence polymorphisms and phylogenetic characteristics were studied on *T. gondii* genes from heart tissues of small mammals captured in Gyeonggi and Gangwon Provinces of Korea; they closely aligned with the genotype I [13].

To date, there have been few surveys on the prevalence of *T. gondii* in stray cats living around Seoul, Korea, particularly using fecal samples to detect B1 gene. Characterization of new geographical strains or isolates of *T. gondii* is also needed in Korea. Thus, we performed a brief survey of *T. gondii* infection in stray cats captured around Seoul, Korea by means of nested-PCR to detect B1 gene and conducted laboratory-passage of some of the isolates for determination of the genotype.

**MATERIALS AND METHODS**

**Sample collection**

Fecal samples were collected from 300 stray cats captured around Seoul (including some boundary areas of Gyeonggi Province) from June to August 2013 through Hello Earth Co., an animal welfare and education consulting company in Korea. The feces of cats were stored at 4°C until analyzed.

**Nested-PCR for *T. gondii* B1 gene**

The fecal samples (*n = 300*) were examined for the presence of *T. gondii* B1 gene using nested-PCR. The DNeasy blood and tissue kit (Qiagen, Hilden, Germany) was used for isolation of the genomic DNA of *T. gondii*. Two primer sets directed against the B1 gene were used according to the previous study [14]. The smart 2× PCR Pre-mix (Solgent Co., Daejeon, Korea) was used with the following conditions: denaturation at 94°C for 3 min, then 35 cycles at 94°C for 20 sec, 56°C for 30 sec, 72°C for 80 sec, and post-amplification at 72°C for 7 min. Automated DNA sequencing of the PCR product was performed by Solgent Co. (Seoul, Korea). Nucleotide sequences obtained from each of the 2 isolates were aligned using the Geneious v. 6.0.3 program (Geneious Co., Wellington, New Zealand).

**Fecal examination of oocysts**

To confirm the presence of *T. gondii* oocysts in the B1 gene-positive fecal samples, the sucrose flotation method was applied to concentrate the oocysts. Briefly, 2 g of feces were mixed with 10 ml sucrose solution (Sigma-Aldrich, St. Louis, Missouri, USA) with a specific gravity of 1.2, and filtered through a strainer into a 15-ml conical tube. The filtrate was then centrifuged at 1,200 rpm for 5 min at 4°C. The tube was removed from the centrifuge and completely filled with sucrose solution and a coverslip was then placed on the tube. After 10 min, oocysts of *T. gondii* were identified on the coverslip under a light microscope at 1,000× magnification.

**Bioassay of mice**

Fourteen mice were orally inoculated each with 0.2 ml of fecal suspension from 14 *T. gondii* B1 gene positive cats. The brain tissue samples were obtained from each mouse 40 days after the inoculation. A portion of the brain tissue wassquashed between a cover slip and a glass slide for microscopic detection of *T. gondii* tissue cysts.

**Genotype analysis of 2 *T. gondii* isolates by PCR-nucleotide sequencing**

PCR and nucleotide sequencing were performed on the SAG5D and SAG5E regions of *T. gondii* according to the procedures previously reported [15]. SAG5D (present in tachyzoites and encodes a polypeptide of 362 amino acids) and SAG5E (transcribed pseudogene) are novel SAG1-related genes useful for discrimination of virulent and avirulent strains of *T. gondii* [15]. The PCR product was amplified using the Cosmo Labo-pass × 2 PCR Premix kit (Cosmo Genetech, Seoul, Korea) with the following conditions: denaturation at 94°C for 3 min, then 35 cycles at 95°C for 20 sec, 56°C for 30 sec, 72°C for 80 sec, and post-amplification at 72°C for 7 min. Automated DNA sequencing of the PCR product was performed by Solgent Co. (Seoul, Korea). Nucleotide sequences obtained from each of the 2 isolates were aligned using the Geneious v. 6.0.3 program (Geneious Co., Wellington, New Zealand).

**Statistical analysis**

Differences in the prevalence of *T. gondii* infection related to gender and age were statistically evaluated using the chi-square test in Excel. The differences were considered statistically significant when *P* < 0.05.

**RESULTS**

Fourteen (4.7%) of the 300 fecal samples of stray cats were positive for *T. gondii* B1 gene by nested-PCR (Table 1). Fecal oocysts were confirmed in at least 10 of the 14 positive feces (data not shown). Female cats (7.5%, 12/161) showed a high-
er B1 gene prevalence than male cats (1.4%, 2/139). By age, cats 3 months or younger (5.5%, 13/235) showed a higher prevalence than cats older than 3 months (1.5%, 1/65). These gender- and age-related differences were statistically significant ($P<0.05$).

Fecal samples positive for B1 gene were orally inoculated into 14 experimental mice, each with 1 fecal sample. After 40 days, 2 mice were positive for brain tissue cysts (Fig. 1). Thereafter, these 2 isolates have been continuously maintained in our laboratory. They were designated as KNIH-1 and KNIH-2 isolates.

Genotyping of KNIH-1 and KNIH-2 isolates included a comparison of their SAG5D and SAG5E gene sequences with those of other strains deposited in GenBank, including RH (genotype I and highly virulent strain), ME49 (genotype II and less virulent strain), and C56 (genotype III and avirulent strain) (Table 2). The SAG5D sequences were identical between the 2 isolates (KNIH-1 and KNIH-2). The SAG5E se-

**Table 1.** Prevalence of *Toxoplasma gondii* oocysts in the feces of stray cats by gender and age groups

| Cat groups | No. examined | No. positive | Prevalence (%) |
|------------|--------------|--------------|----------------|
| Gender     |              |              |                |
| Male       | 139          | 2            | 1.4            |
| Female     | 161          | 12           | 7.5            |
| Age (month)|              |              |                |
| ≤ 3        | 235          | 13           | 5.5            |
| > 3        | 65           | 1            | 1.5            |
| Total      | 300          | 14           | 4.7            |

The feces were examined by detecting B1 gene using nested-PCR. The gender- and age- differences in the prevalence were both statistically significant ($P<0.05$).

**Fig. 1.** Tissue cysts in the brain of mice infected with *T. gondii* KNIH-1 (A) and KNIH-2 isolates (B) successfully passaged in this study. These 2 isolates showed 100% homology with the known ME49 strain based on SAG5D and SAG5E gene sequences (Scale bars=20 μm).

**Table 2.** Comparison (% identical sites) of sequence homology among the SAG5D (dark box below the diagonal) and SAG5E (empty box above the diagonal) genes with 3 known strains of *T. gondii* (RH, ME49, and C56)

| Gene  | Strain               | SAG5D Samples | RH (Genotype I) | ME49 (Genotype II) | C56 (Genotype III) |
|-------|----------------------|---------------|-----------------|--------------------|--------------------|
| SAG5D | Samples              | -             | 99              | 100                | 98.2               |
|       | RH (Genotype I)      | 95.5          | -               | 95.5               | 98.7               |
|       | ME49 (Genotype II)   | 100           | 99              | -                  | 95.3               |
|       | C56 (Genotype III)   | 95.3          | 98.4            | 98.2               | -                  |
quences of the KNIH-1 and KNIH-2 isolates had 100% homology with the *T. gondii* ME49 strain (GenBank accession no. AY190528). However, the 2 isolates showed 99.0% identity with the RH strain (no. AY299523) and 98.2% identity with the C56 strain (no. AY190530). Similarly, the SAG5E sequences of KNIH-1 and KNIH-2 were identical to each other. They showed 100% homology with the ME49 strain (no. AY363047), whereas their homologies with RH (no. AY363043) and C56 strains (AY363049) were 95.5% and 95.3%, respectively (Table 2). These results indicated that the 2 laboratory-passaged isolates are highly homologous with a less virulent ME49 strain rather than a highly virulent RH or an avirulent C56 strain.

**DISCUSSION**

Several surveys have been reported on the seroprevalence or B1 gene detection of *T. gondii* infection in stray cats in Korea [3-5,7,8]. In 1999, 13.1% (26/198) of stray cats captured from a southern area (around Jinju City) showed positive western blot patterns for *T. gondii* [3]. In 2008, of 174 stray cats captured from Gyeonggi Province, serum specific antibodies were positive in 14 cats (8.0%) by latex agglutination test, 28 cats (16.1%) by ELISA, and 23 cats (13.2%) by PCR detection of the B1 gene in blood samples [4]. However, in the same year, a strikingly high prevalence (47.2%) in blood of stray cats by nested-PCR was reported from Gyeonggi Province [5]. Subsequently, the seroprevalence of stray cats from Metropolitan Seoul was reported as 15.3% (11/72) by ELISA, with the B1 gene positive rate in blood of 30.6% (22/72) by nested PCR in 2010 [7]. In another survey, the seroprevalence of stray cats from Metropolitan Seoul was 15.8% (69/456) by ELISA, and the B1 gene positive rate in blood was 17.5% (80/456) in 2011 [8]. These data demonstrate that the seropositivity and B1 gene (in blood) positivity of *T. gondii* infection in stray cats in Korea are markedly variable, ranging from 8.0-47.2%. The reasons for this wide range of reported prevalence include different areas surveyed, different analytical methods, and different materials (blood or feces) used for analyses. However, prevalence surveys using fecal samples were seldom performed in Korea.

In our study, the positive rate of B1 gene was 4.3% (14/300) using fecal samples (but not blood), which was much lower than the seroprevalence or B1 gene prevalence reported in blood samples [3,4,8]. Although the positive rate in this study was comparatively lower than others, we could confirm that fecal samples can be used for molecular studies including B1 gene detection. The lower positivity of *T. gondii* DNA in fecal samples than in blood is interesting. The reason may be that only the oocyst stage is excreted and available in feces, whereas various other *T. gondii* stages can be present in blood. In addition, cats shed oocysts in feces only for a short period of time (a few months) after infection, although the infection can continue for a substantially longer period of time and *T. gondii* specific IgG antibodies in serum persist in cats [16]. Therefore, surveys to detect fecal oocysts in cats are less reliable than detection of B1 gene in blood or antibodies in sera, to understand the true prevalence of *T. gondii* infection. The true prevalence of *T. gondii* in stray cats in Korea seems to be at least higher than 10% and can reach 50% according to the literature.

The prevalence of *T. gondii* in cats may depend on the locality, living conditions, as well as age and gender of the cats. However, in Korea, there have been few surveys on cats from different localities. It is speculated that cats living in rural areas may have a higher prevalence of *T. gondii* than those living in cities including Seoul. Furthermore, stray cats that hunt for food have a higher prevalence of *T. gondii* than those domesticated and fed with preserved food. In Korea, household cats from Seoul revealed a remarkably lower *T. gondii* seroprevalence (2.2%, 10/437) and B1 gene prevalence (2.1%, 10/474) [6], as compared to stray cats with 8.0-17.5% and 13.2-47.2%, respectively [3-5,7,8]. Another survey of household cats reported 0% (0/40) seroprevalence by ELISA (15.3% in stray cats in the same study) and 0% B1 gene prevalence by nested PCR (30.6% in stray cats in the same study) [7].

In our study, the younger cat group (<3 months) revealed a significantly higher prevalence of *T. gondii* (5.5%) than the older cat group (>3 months) (1.5%). This contrasted to the general statement that the seroprevalence of *T. gondii* in cats increases with ages [16,17]. It may be explained by that falc oocysts are shed by infected cats only for a short period of time after exposure, and the infection can continue for a substantially longer period of time [16]. In our study, female cats (7.5%) revealed a significantly higher prevalence than male cats (1.4%). This gender difference was difficult to explain. However, no significant gender difference has previously been reported in the seroprevalence or B1 gene prevalence of *T. gondii* infection in Korea [3-8].

The study confirmed that stray cats play an important role for transmission of *T. gondii* infection in Korea. Among the 14
B1 gene positive samples, 2 were successfully passaged in the laboratory using mice. They were genetically confirmed as 100% homologous with the ME49 strain (genotype II) in the sequences of SAG5D and SAG5E [15]. The results of the present study raise public health awareness and contribute to integrated strategies of health initiatives for prevention and control of *T. gondii* infection in Korea.

**ACKNOWLEDGMENTS**

This study was supported by a research grant from The Korea National Institute of Health, Korea Centers for Disease Control and Prevention, Ministry of Health and Welfare, the Republic of Korea (no. 800-2013-E54007-00). We would like to thank the staff of Hello Earth for providing the fecal samples of stray cats for this study.

**CONFLICT OF INTEREST**

The authors declare that they have no competing interests.

**REFERENCES**

1. Lim H, Lee SE, Jung BK, Kim MK, Lee MY, Nam HW, Shin JG, Yun CH, Cho HI, Shin EH, Chai JY. Serologic survey of toxoplasmosis in Seoul and Jeju-do, and a brief review of its seroprevalence in Korea. Korean J Parasitol 2012; 50: 287-293.
2. Jones JL, Kruszon-Moran D, Wilson M, McQuillan G, Navin T, McAuley JB. *Toxoplasma gondii* infection in the United States: seroprevalence and risk factors. Am J Epidemiol 2001; 154: 357-365.
3. Sohn WM, Nam HW. Western blot analysis of stray cat sera against *Toxoplasma gondii* and the diagnostic availability of monoclonal antibodies in sandwich-ELISA. Korean J Parasitol 1999; 37: 249-256.
4. Kim HY, Kim YA, Kang S, Lee HS, Rhie HG, Ahn HJ, Nam HW, Lee SE. Prevalence of *Toxoplasma gondii* in stray cats of Gyeonggi-do, Korea. Korean J Parasitol 2008; 46: 199-201.
5. Lee JY, Lee SE, Lee EG, Song KH. Nested PCR-based detection of *Toxoplasma gondii* in German shepherd dogs and stray cats in South Korea. Res Vet Sci 2008; 85: 125-127.
6. Hong SH, Jeong YI, Kim JY, Cho SH, Lee WJ, Lee SE. Prevalence of *Toxoplasma gondii* infection in household cats in Korea and risk factors. Korean J Parasitol 2013; 51: 357-361.
7. Lee SE, Kim JY, Kim YA, Cho SH, Ahn HJ, Woo HM, Lee WJ, Nam HW. Prevalence of *Toxoplasma gondii* infection in stray and household cats in regions of Seoul, Korea. Korean J Parasitol 2010; 48: 267-270.
8. Lee SE, Kim NH, Chae HS, Cho SH, Nam HW, Lee WJ, Kim SH, Lee JH. Prevalence of *Toxoplasma gondii* infection in feral cats in Seoul, Korea. J Parasitol 2011; 97: 153-155.
9. Khan A, Dubey JP, Su C, Ajikoka JW, Rosenthal BM, Sibley LD. Genetic analyses of atypical *Toxoplasma gondii* strains reveal a fourth clonal lineage in North America. Int J Parasitol 2011; 41: 645-655.
10. Dubey JP, Velmurugan GV, Rajendran C, Yabsley MJ, Thomas NJ, Beckmen KB, Sinnett D, Ruid D, Hart J, Fair PA, McFee WE, Shearn-Bochsler V, Kwook OCH, Ferreira LR, Choudhary S, Faria EB, Zhou H, Felix TA, Su C. Genetic characterisation of *Toxoplasma gondii* in wildlife from North America revealed widespread and high prevalence of the fourth clonal type. Int J Parasitol 2011; 41: 1139-1147.
11. Chai JY, Lin A, Shin EH, Oh MD, Han ET, Nan HW, Lee SH. Laboratory passage and characterization of an isolate of *Toxoplasma gondii* from an ocular patient in Korea. Korean J Parasitol 2003; 41: 147-154.
12. Lin A, Shin EH, Kim TY, Park JH, Guk SM, Chai JY. Genetic characteristics of the Korean isolate KI-1 of *Toxoplasma gondii*. Korean J Parasitol 2005; 43: 27-32.
13. Hong SH, Lee SE, Jeong YI, Kim HC, Chong ST, Klein TA, Song JW, Gu SH, Cho SH, Lee WJ. Prevalence and molecular characterizations of *Toxoplasma gondii* and Babesia microti from small mammals captured in Gyeonggi and Gangwon Provinces, Republic of Korea. Vet Parasitol 2014; 205: 512-517.
14. Jones CD, Okhravi N, Adamson P, Tasker S, Lightman S. Comparison of PCR detection methods for B1, P30, and 18S rDNA genes of *T. gondii* in aqueous humor. Invest Ophthalmol Vis Sci 2000; 41: 634-644.
15. Tinti M, Possenti A, Cherchi S, Barca S, Spano F. Analysis of the SAG5 locus reveals a distinct genomic organisation in virulent and avirulent strains of *Toxoplasma gondii*. Int J Parasitol 2003; 33: 1605-1616.
16. Dumètre A, Dardé ML. How to detect *Toxoplasma gondii* oocysts in environmental samples? FEMS Microbiol Rev 2003; 27: 651-661.
17. Wu SM, Zhu XQ, Zhou DH, Fu BQ, Chen J, Yang JE, Song HQ, Weng YB, Ye DH. Seroprevalence of *Toxoplasma gondii* infection in household and stray cats in Lanzhou, northwest China. Parasit Vectors 2011; 4: 214.
