MGMT Immunohistochemical Expression in Colorectal Carcinoma and its Correlation with Tumor Progression

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Abstract

BACKGROUND: There is an urgent need to identify predictive features and markers for progression and treatment of colorectal carcinoma (CRC).

AIM: This study aimed to assess O⁶-methylguanine DNA methyltransferase (MGMT) expression in CRC and to correlate with the clinicopathological aspects of the tumor, also to evaluate the relationship between different histopathological parameters and tumor progression.

MATERIALS AND METHODS: The study was carried on 70 colectomy using formalin fixed paraffin embedded tumor tissue not subjected to chemo-radiotherapy nor with missing data. Specimens were collected from the Department of Pathology of Kasr El-Aini Hospital, Faculty of Medicine, Cairo University, during the period between (March - 2017 and May - 2018). Immunohistochemistry was used to detect MGMT expression and clinicopathologic aspects as well as to assess tumor budding, type of desmoplastic reaction (DR), inflammatory lymphocytic milieu, pattern of invasive front and necrosis, and then correlated with MGMT expression and tumor progression, using parametric and non-parametric statistical methods.

RESULTS: MGMT loss of expression was detected in 42.9% of CRC cases. MGMT expression status was significantly correlated with tumor stage and metastatic status (p < 0.05), while it was not correlated with other clinicopathologic features, (p > 0.05). DR, tumor budding, stromal tumor infiltrating lymphocytes (TIL-S), and necrosis were significantly correlated with tumor stage and metastatic status (p < 0.05), while it was not correlated with other clinicopathologic features, (p > 0.05). Both types of TIL and Crohn’s-like lymphoid reaction showed a mutual correlation (p < 0.05).

CONCLUSION: MGMT high expression and histopathologic parameters as DR, tumor budding, inflammatory lymphocytic milieu, and necrosis could be correlated with CRC progression.

Introduction

Colorectal cancer (CRC) is the third most commonly diagnosed cancer in males and the second in females worldwide, while according to cancer mortality, CRC is the third leading cause in both males and females [1]. The development of CRC is complex, with critical role for generation of mutations and failed DNA repair in carcinogenesis. Tumor progression entails the downregulation of damage surveillance mechanisms and necessitates genetic and epigenetic instability to gain uncontrolled proliferation. Epigenetic dysregulation is a driving mechanism in cancer development and progression. This dysregulation is directly acting to modify gene expression through a methylation process of DNA, leading to oncogene activation, tumor suppressor gene silencing, chromosomal instability, and chromatin modification [2], [3], [4].

O⁶-methylguanine DNA methyltransferase (MGMT) is a DNA repair enzyme expressed by all normal human cells, it prevents DNA mutations and damage due to alkylating agents, then undergoes self-inactivation.
tumor infiltrating lymphocytes (TIL), and Crohn’s-like lymphoid reaction (CLR), tumor invasive front pattern and necrosis are from those main parameters [12], [13], [14], [15], [16], [17], [18], [19].

The aim of this study is to evaluate the expression of MGMT in CRC and to correlate this expression with different clinicopathological aspects as well as to evaluate the relationship between different histopathologic parameters in CRC with tumor progression.

Materials and Methods

Collection of specimens

The study was carried on a total of 70 colectomy specimens of patients with colorectal cancer not subjected to chemo-radiotherapy nor with missing data, tumor sections were formalin fixed and paraffin embedded. Specimens were sampled by simple random method from the Department of Pathology of Kasr El-Aini Hospital, Faculty of Medicine, Cairo University during the period between (March - 2017 and May - 2018). The study was approved by the ethical committee of Kasr El-Aini Hospital and specimens were anonymously replaced by numbers for confidentiality.

The site of the tumor was classified into colonic and rectum, while the size of the tumor was calculated as the length of the largest diameter. The tumor extension into other organs, distant metastasis if present and the lymph node status were also obtained from the diagnosis present in the pathology reports (clinical data of distant metastasis in other organs detected at time of diagnosis were also obtained from the patient’s sheet). Mucinous and signet ring cases were excluded from grading.

Processing and histopathologic examination

Paraffin blocks of the tumor were serially sectioned at 4 μm thickness. Afterward, they were stained with routine hematoxylin-eosin stain for evaluation and confirmation of the diagnosis, assessment of histologic type and grade, detection of the tumor invasion depth, lymph node status and the presence of lympho-vascular and perineural invasion, according to the recommendations of the World Health Organization, while staging was performed using TNM 8th edition system for each case [12], [13]. Histopathologic examination was performed by one pathologist (MA). All microscopic measurements were taken using Leica slide scanner SCN400 and image viewer software (Leica Microsystems).

Tumor invasive front pattern was assessed whether infiltrative with jagged border or pushing with relative smooth border. Tumor budding was assessed according to the International Tumor Budding Consensus Conference, as isolated cancer cells or a cluster of <5 neoplastic cells at the invasive front of the tumor. The invasive front of the tumor was assessed at a scanning (x10 objective) magnification for the area with maximal tumor budding. In this area, the number of tumor buds was determined in a 0.785mm² area. Tumors were classified as absent budding, low tumor budding if 1–9 tumor buds, and high tumor budding if ≥10 tumor buds were identified per 0.785 mm² area [14].

DR was histologically assessed in the reactive fibrous zone at the advancing extramural edge of the tumor and classified into three categories: Mature when fibrotic stroma did not contain keloid-like collagen or myxoid stroma and was composed of fine mature collagen fibers stratified into multiple layers. Intermediate when keloid-like collagen observed as hyalinized thick bundles of hypocellular collagen was intermingled with mature stroma. Immature when stroma show myxoid changes comprising slightly basophilic extracellular matrix. The stroma is classified according to the most immature stromal area. A microscopic field of a ×40 objective was used as a cutoff [15]. DR was not assessed in 14 cases which represented T1 and T2 cases which by definition have not an extramural invasive component.

Inflammatory lymphocytic milieu was histologically assessed as TIL with categorization into two groups: Intraepithelial (TIL-E) assessed in 5 HPF, in the invasive front and center of tumor and classified as absent, one per gland (cancer Tube) or more per gland, excluding necrotic, and apopotic areas.[16] Stromal (TIL-S) assessed in the center and invasive front of the tumor in a percentage of stromal inflammatory cells and scored in 5% increments excluding necrotic and fibrotic areas, immune infiltrate outside of the tumor and granulocytic infiltrate areas, according to International TILs Working Group into three categories: Low (0-10%), moderate (15–50%), and high (>50%) of stroma [17]. CLR density is measured as number of lymphoid aggregates per mm length of tumor front and was scored as absent, low or high CLR density using cutoff (>0.38 follicles/mm) of length [18].

Tumor necrosis was assessed in the tumor’s center excluding luminal necrotic debris and graded as “absent” (0), “focal” (<10%), “moderate” (10–30%) or “extensive” (>30%) of tumor area [19].

Immunohistochemistry (IHC)

Another unstained paraffin section on positive charged slides from each case was loaded in IHC autostainer (Ventana Benchmark XT, Roche Diagnostics) and processed for immunostaining with Mouse monoclonal MGMT antibody (Santa Cruz Biotechnology) with a dilution 1:50, using Optiview IHC detection kit. MGMT external positive control of a lymph
node was used as well as internal control of lymphocytes, plasma cells, stroma, and blood vessels. Lymph node tissue was also assayed omitting the primary antibody as negative controls. Immunohistochemical staining of MGMT was evaluated as positive only with nuclear staining in tumor cells, while cytoplasmic staining is disregarded. A semi-quantitative scoring system for both staining intensity and the percentage of positive cells was used. A score was calculated by multiplying the intensity (negative: 0, mild: 1, moderate: 2, and strong: 3) by percentage of stained cells (0: absent, 1: 1–25%, 2: >25–50%, 3: >50–75%, and 4: >75–100%). Scores of multiplication were graded as follows: (−):0, (+): 1–3, (++): 4–8, and (+++): 9–12. In addition, for statistical analysis, the (−) and (+) cases were pooled into the low-expression group, and the (++) and (+++) cases were pooled into the high-expression group [20]. Immunostaining of additional tumor section was applied to cases with variation in the intensity of staining in multiple fields in the same tumor section. IHC interpretation and score assessment were performed by two pathologists (MA and MS). MGMT expression was studied with the different previously mentioned clinicopathologic aspects of CRC in collected cases.

Statistical methods

Data were statistically described in terms of mean ± standard deviation (±SD), median and range, or frequencies (number of cases) and percentages when appropriate. Percentages were approximated taking in consideration total percentage to be 100%. Comparison of numerical variables between the study groups was done using Mann Whitney U test for independent samples to compare 2 groups. For comparing categorical data, Chi square ($\chi^2$) test was performed. Exact test was used instead when the expected frequency is <5, $p < 0.05$ was considered statistically significant flagged with an asterisk, $<0.01$; highly significant flagged with 2 asterisks and $<0.001$ is extremely significant flagged with 3 asterisks. Shapiro–Wilk test was included for normality. All statistical calculations were done using Statistical Package for the Social Science (SPSS version 20.).

Results

The study included 70 cases of CRC with age ranging between 11 and 82 years old, mean age was 51.4 years and median age was 55 years. Three patients (4.3% of cases) were younger than 18 years. Males constituted 57.1% and females 42.9% of cases. Low MGMT expression was detected in 42.9% of all cases. Mean and median age for low MGMT expression group were 51.1 and 55 years, respectively, while those of high MGMT expression group were 51.6 and 52.5 years, respectively. Mean size of the tumor was 5.7 cm (ranging between 2.5 and 11cm) with low MGMT expression detected in 46.7% of size ≥5 cm and 36% of size <5 cm, with no statistically significant difference ($p > 0.05$). Figure 1 displays low and high expression of MGMT. Age, tumor size, and MGMT expression show normal distribution according to Shapiro–Wilk normality test ($p < 0.05$), except for tumor size in the high MGMT expression group ($p = 0.013$).

Regarding tumor stage and MGMT expression, low expression was detected in 54.4% of Stage I, 25% of Stage II, 60% of Stage III, and 11.1% of Stage IV, with a statistically significant relationship ($p = 0.0146$).

Table 1 summarizes the relationship between tumor staging with MGMT and some clinico-pathologic parameters. Despite there was not a statistically significant relationship ($p > 0.05$) between MGMT expression and primary tumor extension, a direct linear correlation between increase in MGMT expression and higher primary tumor extension (T) stage was noted. There were no statistically significant relationship ($p > 0.05$) between MGMT expression with age, sex, tumor size, site, lymph node status, tumor differentiation, histologic type, lymphovascular and perineural invasion, tumor budding, pattern of invasive front, TIL, CLR, DR,
Table 1: The relationship between MGMT and clinicopathologic parameters with tumor staging

| Parameter       | No. (%) | Stage I | Stage II | Stage III | Stage IV | p-value |
|-----------------|---------|---------|----------|-----------|----------|---------|
| MGMT            |         |         |          |           |          |         |
| High expression | 40 (57.1) | 5 (45.6) | 15 (75)  | 12 (40)   | 8 (88.9) | 0.0146* |
| Low expression  | 30 (42.9) | 6 (54.4) | 5 (25)   | 18 (60)   | 1 (11.1) |         |
| Strioma         |         |         |          |           |          |         |
| Mature          | 19 (33.9) | NA      | 11 (55)  | 7 (25.9)  | 1 (11.1) | 0.0353* |
| Intermediate    | 23 (41.1) | NA      | 7 (35)   | 13 (48.2) | 3 (33.3) |         |
| Immature        | 14 (25)  | NA      | 2 (10)   | 7 (25.9)  | 5 (55.6) |         |
| Budding         |         |         |          |           |          |         |
| Absent          | 20 (28.6) | 7 (63.6) | 8 (40)   | 4 (13.3)  | 1 (11.1) | 0.0185* |
| Low             | 23 (32.8) | 3 (27.3) | 6 (30)   | 12 (40)   | 2 (22.2) |         |
| High            | 27 (38.6) | 1 (9.1)  | 6 (30)   | 14 (46.7) | 6 (66.7) |         |
| Invasive front  |         |         |          |           |          |         |
| Pushing         | 7 (10)   | 3 (27.3) | 1 (5)    | 3 (15)    | 0 (0)    | 0.157   |
| Infiltrative    | 63 (90)  | 8 (72.7) | 9 (95)   | 27 (90)   | 9 (100)  |         |
| Tumor infiltrating lymphocytes stromal | 38 (54.3) | 4 (36.4) | 17 (85)  | 13 (43.3) | 4 (44.45) | 0.0188* |
| Intermediate    | 26 (37.1) | 5 (45.4) | 1 (5)    | 16 (53.3) | 4 (44.45) |         |
| High            | 6 (8.6)  | 2 (18.2) | 2 (10)   | 1 (3.4)   | 1 (11.1) |         |
| Tumor infiltrating lymphocytes intraepithelial |         |         |          |           |          |         |
| Absent          | 18 (25.7%) | 0 (0%)  | 5 (25%)  | 10 (33.3%) | 3 (33.3%) | 0.161   |
| 1               | 44 (62.9%) | 8 (72.7%) | 14 (70%) | 18 (60%)  | 4 (44.5%) |         |
| >1              | 8 (11.4%) | 3 (27.3%) | 1 (2%)   | 6 (26.7%) | 2 (22.2%) |         |
| Crohn’s-like lymphoid reaction | 33 (47.2%) | 7 (63.6%) | 10 (50%) | 11 (36.7%) | 5 (55.6%) | 0.780   |
| Low             | 25 (35.7%) | 3 (27.3%) | 7 (30%)  | 12 (40%)  | 3 (33.3%) |         |
| High            | 17 (12.1%) | 1 (9.1%)  | 3 (15%)  | 7 (23.3%) | 1 (11.1%) |         |
| Necrosis        |         |         |          |           |          |         |
| Absent          | 17 (24.3%) | 8 (72.7%) | 1 (5%)   | 6 (20%)   | 2 (22.2%) | 0.0032**|
| Focal           | 35 (50%) | 2 (18.2%) | 12 (60%) | 15 (50%)  | 6 (66.7%) |         |
| Moderate        | 16 (22.8%) | 1 (9.1%)  | 6 (30%)  | 9 (30%)   | 0 (0%)   |         |
| Extensive       | 4 (5.7%) | 0 (0%)   | 1 (5%)   | 0 (0%)    | 1 (11.1%) |         |

MGMT: O'-methylguanine DNA methyltransferase. NA: Not Applicable.

Discussion

MGMT low expression group represented 42.9% of cases, higher than Michailidi’s and Pietrantonio’s studies; 34% and 37%, respectively, while lower than Oliver’s study 48.2% [21], [22], [23]. MGMT low expression group was more frequent in females 50% than in males 37.5%, similar to Zhang’s and Michailidi’s studies, and in contrast to Oliver’s study that demonstrated MGMT low expression group was more predominant in males than in females. Regarding the age, Michailidi’s study showed higher mean age for both low and high MGMT expression categories 69.5 and 75 years, while what Zhang and Oliver et al. found a statistical correlation with MGMT expression at the other side could be attributed to differences in the racial and environmental factors between populations as all the patients included in the current study are Egyptians. Tumor histologic type and differentiation lack statistical significance with MGMT expression, while Zhang et al. found a statistical correlation between histologic type and low MGMT expression which was more common in signet ring cell carcinoma 80%, but they couldn’t detect a significant differences representing 39.2% and 52.6%, respectively, higher than what Zhang et al. found; 26.7% in colonic cases and 32.3% in rectal cases [24]. The differences between age, gender, and tumor site at one side in relation with MGMT expression at the other side could be attributed to differences in the racial and environmental factors between populations as all the patients included in the current study are Egyptians. Tumor histologic type and differentiation lack statistical significance with MGMT expression, while Zhang et al. found a statistical correlation between histologic type and low MGMT expression which was more common in signet ring cell carcinoma 80%, but they couldn’t detect a significant differences representing 39.2% and 52.6%, respectively, higher than what Zhang et al. found.
correlation with the tumor grade in the same study. Liddell et al. found frequent loss of MGMT expression in mucinous adenocarcinoma 23% [24], [25].

Regarding the inflammatory lymphocytic milieu of tumor, neither TIL-S, TIL-E, nor CLR correlated significantly with MGMT expression. An interesting significant relationship was noted between any two elements of the studied lymphocytic triad, that is, TIL-S with TIL-E, TIL-S with CLR, and TIL-E with CLR, similar to what Jakubowska found at the invasive front of the tumor [15]. This could support that the inflammatory lymphocytic milieu are orchestrated or directed along the same course of tumor’s immune status, as evidenced by immunohistochemical studies that demonstrated mixed B- and T-cell elements in CLR and variable T-cell subsets in TIL [17]. The current study also showed that only TIL-S showed a correlation with the TNM stage grouping. Most other studies correlated high density TIL and CLR with TNM stage grouping, favorable prognosis and overall survival, with additional significant correlation between TIL-E and CLR as well as TIL-E and TIL-S. [17], [29], [30], [31]. Necrosis did not correlate significantly with MGMT expression, yet it was significantly correlated with TNM stage grouping, which was also reported by Pollheimer. This could be due to rapid tumor growth and reflecting hypoxic changes, and in turn, correlate with increase in metastatic potential and adverse prognosis [32]. To the best of our knowledge, no data were published in the literature about the relation between DR, tumor budding, invasive front pattern, inflammatory lymphocytic milieu, and necrosis with MGMT expression status.

In this study, tumor invasiveness (T) was directly correlated with increase in MGMT expression where none of T1 cases showed high expression while T4 showed the highest expression in 70% with insignificant correlation (p = 0.889) which may raise a possibility that tumors with intact MGMT expression have more capability of growth and local invasion, although this needs more number of cases to be studied to confirm this current suggestion. Besides detection of MGMT low expression in T1 and T2 could give a clue that loss of MGMT is not a late event in CRC.
invasive phase and is related to invasive properties not the tumor size of the non-invasive growth, as it is noticed also from the percentage of low expression in tumors <5 cm previously mentioned. According to lymph node status (N), low MGMT expression was detected in 43.6% of N1 and N2 cases. This result is within the range between Zhang’s and Michailidi’s studies, both without significant statistical correlation [21] [24].

According to the presence or absence of metastasis (M) and TNM stage grouping, low expression of MGMT was detected in one of the nine metastatic cases, a significant correlation between MGMT high expression and stage grouping was detected ($p = 0.0146$). Studies done by Zhang, Sartore-Bianchi and Morano reported higher percentage of their cases with low MGMT expression in metastatic cases 24.2%, 43%, and 72%, respectively, with their studies focusing on Stage IV disease, while Michailidi found loss of MGMT expression in 50% of metastatic cases in a study involving the four stages using Duke’s staging system with significant statistical correlation [19]. The discrepancy in percentages is attributed to the type of study whether random or non-random, the number of cases and the scoring system used in assessment of MGMT IHC. Even though our study showed low MGMT expression in a low percentage of the metastatic cases, which theoretically could be disappointing, this will not preclude any benefit gained from applying MGMT detection in metastatic CRC cases and the trials using alkylating chemotherapeutics, as TMZ for cases with low expression [33]. Furthermore, Mori showed that patients with methylation and loss of expression of MGMT in tissue are valuable biomarkers for detection of Stage III CRC at high risk of recurrence [34]. It is worth mentioning that our study included four cases (colonic and Stage II) that showed low MGMT expression, one of them showed perineural invasion, and one of the high-risk features. Interestingly, a clinical trial using a combination of TMZ and Irinotecan for pretreated metastatic CRC, showed that all non-responder patients were MGMT positive by IHC, while patients with MGMT negative or low had a significantly longer median progression free-survival than others [33]. In addition, Shah et al. observed concordance between MGMT gene promoter methylation status and MGMT IHC results in Glioblastoma using the same IHC clone [35], making MGMT worth for investigation.

**Limitations and recommendation of the study**

This study has several potential limitations; first, the lack of detection of MGMT by methylation assay combined with IHC which could be more confirmative to the role MGMT in the pathogenesis of colorectal carcinoma, second, the lack of MSI status as we predict that it will show correlation with MGMT expression and many of the studied histopathological parameters and so additional molecular correlation is needed especially with known prognostic genes as KRAS and BRAF. Third, the lack of 5 year survival, as to date, the cases were collected from only <3 years ago and this period will not be enough to use prognostically; therefore, further studies with larger samples for pretreated cases and long-term follow-up are required to establish prognostic significance of MGMT and the other histopathologic parameters. In addition, Loss of MGMT might be of interest in metastatic and recurrent CRC to select cases that could benefit from alkylating chemotherapeutics, as TMZ. Finally, further studies focusing on the biology of tumor microenvironment are needed to characterize its role in carcinogenesis.

**Conclusion**

MGMT high expression showed a statistically significant correlation with tumor stage grouping and can be correlated with CRC progression. MGMT Low expression was detected in 42.9% of CRC cases and in 11.1% of metastatic cases, which might be a valuable biomarker in selection of metastatic and recurrent cases to benefit from alkylating chemotherapeutics, as TMZ. Histopathologic parameters as DR, tumor budding, Inflammatory lymphocytic milieu (TIL-S, TIL-E, and CLR) and necrosis lack correlation with MGMT expression, however, were also associated with tumor progression, with a significant relationship between some parameters as DR with tumor budding and the mutual correlation between elements of the inflammatory (lymphocytic) milieu. Finally, these histopathologic parameters are advocated to be included into the routine diagnostic surgical pathology reports.
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References

1. Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;1:1-41. https://doi.org/10.3322/caac.21660
PMid:33535338
2. Ioannou M, Paraskeva E, Baxevanidou K, Simos G, Papamichali R, Papacharalambous C, et al. HIF-1α in colorectal carcinoma: Review of the literature. J BUON. 2015;20(3):680-9.
PMid:26224618
3. Jeggo PA, Pearl LH, Carr AM. DNA repair, genome stability and cancer: A historical perspective. Nat Rev Cancer. 2016;16(1):35-42. https://doi.org/10.1038/nrc.2015.4
PMid:26667949
4. Mojarad EN, Kuppen PJ, Aghdaei HA, Zali MR. The CpG island methylator phenotype (CIMP) in colorectal cancer. Gastroenterol Hepatol Bed Bench. 2013;6(3):120-8.
PMid:24834258
5. Sharma S, Salehi F, Scheithauer BW, Rotondo F, Syro LV, Kovacs K. Role of MGMT in tumor development, progression, diagnosis, treatment and prognosis. Anticancer Res. 2009;29(10):3759-68.
PMid:19846906
6. Cordeiro AT, Da Silva CM, Bartchewsky B Jr., Ribeiro ML, Martinez CA. Evaluation of the expression of the MGMT gene in normal and neoplastic tissue of patients with colorectal cancer. Rev Col Bras Cir. 2012;39(1):48-53. https://doi.org/10.1590/s0100-69912012000100010
PMid:22481706
7. Lee KE. Immunohistochemical assessment of O(6)-methylguanine-DNA methyltransferase (MGMT) and its relationship with p53 expression in endometrial cancers. J Cancer Prev. 2013;18(4):351-4. https://doi.org/10.15430/jcp.2013.18.4.351
PMid:25337565
8. Cankovic M, Nikiforova MN, Snuderl M, Adesina AM, Lindeman N, Wen PY, et al. The role of MGMT testing in clinical practice: A report of the association for molecular pathology. J Mol Diagn. 2013;15(5):539-55. https://doi.org/10.1016/j.jmoldx.2013.05.011
PMid:23871769
9. Jacinto FV, Esteller M. MGMT hypermethylation: A prognostic foe, a predictive friend. DNA Repair (Amst). 2007;6(9):1155-60. https://doi.org/10.1016/j.dnarep.2007.03.013
PMid:17488285
10. Zhang J, Stevens MF, Bradshaw TD. Temozolomide: Mechanisms of action, repair and resistance. Curr Mol Pharmacol. 2012;5(1):102-14.
PMid:22122467
11. Reifenberger G, Hentschel B, Felsberg J, Schackert G, Simon M, Schnell O, et al. Predictive impact of MGMT promoter methylation in glioblastoma of the elderly. Int J Cancer. 2012;131(6):1342-50. https://doi.org/10.1002/ijc.27385
PMid:22139906
12. Nagtegaal ID, Arends MJ, Telle S. Colorectal adenocarcinoma. In: Nagtegaal ID, Arends MJ, Odze RD, Lam AK, editors. Tumors of the Colon and Rectum WHO Classification of Tumors Editorial Board. WHO Classification of Tumors: Digestive System Tumors. 5th ed. Geneva: World Health Organization Press; 2019. p. 177-87. https://doi.org/10.1007/978-3-7091-6821-9_2
13. Jessup JM, Goldberg RM, Asare EA, Benson AB, Brierley JD, Chang GJ, et al. Colon and rectum. In: Amin MB, editor. AJCC Cancer Staging Manual. 8th ed. New York: Springer Nature; 2017. p. 251-74.
14. Lugli A, Kirsch R, Ajoka Y, Bosman F, Cathomas G, Dawson H, et al. Recommendations for reporting tumor budding in colorectal cancer based on the international tumor budding consensus conference (ITBCC) 2016. Mod Pathol. 2017;30(9):1299-311. https://doi.org/10.1038/modpathol.2017.46
PMid:28548122
15. Ueno H, Kanemitsu Y, Sekine S, Ishiguro M, Ito E, Hashiguchi Y, et al. Desmoplastic pattern at the tumor front defines poor-prognosis subtypes of colorectal cancer. Am J Surg Pathol. 2017;41(11):1506-12. https://doi.org/10.1097/pas.0000000000000946
PMid:28877064
16. Jakubowska K, Kisielewski W, Kanczuga-Koda L, Koda M, Famulski W. Stromal and intraepithelial tumor-infiltrating lymphocytes in colorectal carcinoma. Oncol Lett. 2017;14(6):6421-32. https://doi.org/10.3892/ol.2017.7013
PMid:29151905
17. Salgado R, Denkert C, Demaria S, Sirtaine N, Klauschen F, Pruner G, et al. The evaluation of tumor-infiltrating lymphocytes (TILs) in breast cancer: Recommendations by an International TILs working group 2014. Ann Oncol. 2015;26(2):259-71. https://doi.org/10.1093/annonc/mdu450
PMid:25214542
18. Väyrynen JP, Sajanti SA, Klintrup K, Mäkelä J, Herzig KH, Karttunen TJ, et al. Characteristics and significance of colorectal cancer associated lymphoid reaction. Int J Cancer. 2014;134(9):2126-35. https://doi.org/10.1002/j cured.28533
PMid:24154855
19. Richards CH, Flegg KM, Roxburgh CS, Going JJ, Mohammed Z, Horgan PG, et al. The relationships between cellular components of the peritumoural inflammatory response, clinicopathological characteristics and survival in patients with primary operable colorectal cancer. Br J Cancer. 2012;106(12):2010-5. https://doi.org/10.1038/bjc.2012.211
PMid:22596238
20. Sartore-Bianchi A, Pietrantonio F, Amatu A, Milione M, Cassingena A, Ghезzi S, et al. Digital PCR assessment of MGMT promoter methylation coupled with reduced protein expression optimises prediction of response to alkylating agents in metastatic colorectal cancer patients. Eur J Cancer. 2017;71:43-50. https://doi.org/10.1016/j.ejca.2016.10.032
PMid:27997874
21. Michailidi C, Theocharis S, Tsouroulis G, Pletska V, Koutrakis G, Patsouris E, et al. Expression and promoter methylation status of hMLH1, MGMT, APC, and CDH1 genes in patients with colon adenocarcinoma. Exp Biol Med (Maywood). 2015;240(12):1599-605. https://doi.org/10.1038/bjc.2012.211
PMid:25908636
22. Pietrantonio F, De Braud F, Milione M, Maggi C, Iacovelli R, Dotti KF, et al. Dose-dense temozolomide in patients with MGMT-silenced chemorefractory colorectal cancer. Target Oncol. 2016;11(3):337-43. https://doi.org/10.1007/s11523-015-0397-2
250
PMid:26538496

23. Oliver JA, Ortiz R, Melguizo C, Alvarez PJ, Millan JG, Prados J. Prognostic impact of MGMT promoter methylation and MGMT and CD133 expression in colorectal adenocarcinoma. BMC Cancer. 2014;14:511. https://doi.org/10.1186/1471-2407-14-511

PMid:25015560

24. Zhang L, Zeng J, Zeng Z, Wang F, Wang D, Chen C, et al. MGMT in colorectal cancer: A promising component of personalized treatment. Tumor Biol. 2016;37(8):11443-56. https://doi.org/10.1007/s13277-016-5014-1

PMid:27006309

25. Liddell C, Droy-Dupré L, Métairie S, Aïraud F, Volteau C, Bezieau S, et al. Mapping clinicopathological entities within colorectal mucinous adenocarcinomas: A hierarchical clustering approach. Mod Pathol. 2017;30(8):1177-89. https://doi.org/10.1038/modpathol.2017.18

PMid:28429715

26. Lugli A, Karamitopoulou E, Zlobec I. Tumour budding: A promising parameter in colorectal cancer. Br J Cancer. 2012;106(11):1713-7. https://doi.org/10.1038/bjc.2012.127

PMid:22531633

27. Ueno H, Shinto E, Shimazaki H, Kajiwara Y, Sueyama T, Yamamoto J, et al. Histologic categorization of desmoplastic reaction: Its relevance to the colorectal cancer microenvironment and prognosis. Ann Surg Oncol. 2015;22(5):1504-12. https://doi.org/10.1245/s10434-014-4149-9

PMid:25395146

28. Cho SJ, Kakar S. Tumor budding in colorectal carcinoma: Translating a morphologic score into clinically meaningful results. Arch Pathol Lab Med. 2018;142(8):952-7. https://doi.org/10.5858/arp1.2018-0082-ra

PMid:30040461

29. Klintrup K, Mäkinen JM, Kauppila S, Väre PO, Melkko J, Tuominen H, et al. Inflammation and prognosis in colorectal cancer. Eur J Cancer. 2005;41(17):2645-54. https://doi.org/10.1016/j.ejca.2005.07.017

PMid:16239109

30. Rozeik LS, Schmit SL, Groenson JK, Tomso LP, Rennert HS, Rennert G, et al. Tumor-infiltrating lymphocytes, Crohn’s-like lymphoid reaction and survival from colorectal cancer. J Natl Cancer Inst. 2016;108(8):djw027. https://doi.org/10.1093/jnci/djw027

PMid:27172903

31. Fuchs TL, Sioson L, Sheen A, Jafari-Nejad K, Renaud CJ, Andrici J, et al. Assessment of tumor-infiltrating lymphocytes using international TILs working group (ITWG) system is a strong predictor of overall survival in colorectal carcinoma. Am J Surg Pathol. 2020;44(4):536-44. https://doi.org/10.1097/ pas.0000000000001409

PMid:31743129

32. Pollheimer MJ, Komprat P, Lindtner RA, Harbaum L, Schlemmer A, Rehak P, et al. Tumor necrosis is a new promising prognostic factor in colorectal cancer. Hum Pathol. 2010;41(12):1749-57. https://doi.org/10.1016/j.humpath.2010.04.018

PMid:20869096

33. Morano F, Corallo S, Nger M, Barault L, Milione M, Berenato R, et al. Temozolomide and irinotecan (TEMIRI regimen) as salvage treatment of irinotecan-sensitive advanced colorectal cancer patients bearing MGMT methylation. Ann Oncol. 2018;29(8):1800-6. https://doi.org/10.1093/annonc/mdy197

PMid:29860358

34. Mori Y, Nagasaka T, Tazawa H, Umeda Y, Morikawa T, Kubota N, et al. MGMT methylation as a novel biomarker for the identification of stage III colorectal cancers at high-risk of disease recurrence following curative surgery. Gastroenterology. 2013;144(5):S-85. https://doi.org/10.1053/j.gastro.2013.03.015

PMid:23019814

35. Shah N, Lin B, Sibennaller Z, Ryken T, Lee H, Yoon JG, et al. Comprehensive analysis of MGMT promoter methylation: Correlation with MGMT expression and clinical response in GBM. PLoS One. 2011;6(1):e16146. https://doi.org/10.1371/journal.pone.0016146

PMid:21249131