In Vivo Antimalarial Test of Artocarpin and in vitro Antimalarial Test of Artonin M Isolated from Artocarpus

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Abstract: The derivative of flavonoid compounds, artocarpin (1) and artonin M (2), were isolated from the root wood of Artocarpus altilis and from the root bark of A. kemando, respectively. Both plants originated from Lampung, Indonesia. The structure of the two compounds has been carefully determined by physical method and spectroscopy techniques of UV, IR, and NMR. The in vivo antimalarial test of artocarpin showed very good Plasmodium activity in female mice, with ED₅₀ value of 34.88 mg/kg body weight (kgBW), whereas the in vitro antimalarial test of artonin M showed very strong activity with IC₅₀ of 0.3 μg/mL (5.967 x 10⁻⁷ M).

Keywords: antimalarial, artocarpin, artonin M, A. altilis, A. kemando

1. Introduction

The antimalarial researches on flavanoid compounds isolated from artocarpus plants have been carried out and reported previously [1–4]. The compounds isolated from these plants were very interesting to explore as an antimalarial drug. Artonin E, cycloartobiloxanthone, artocarpin, cycloartocarpin isolated from A. altilis have been found to exhibit cytotoxic activity against Plasmodium falciparum. All of these active compounds are prenylated on C3, and the antimalarial activity test previously performed on these compounds were usually in vitro activity test [1–4].

A. kemando Miq. is one of endemic plants that grows in Indonesia. From this plant, some prenylated flavanoids have been isolated, and these include artomandin,artoindonesianin C, artonol B, and artochamin A [5]. All of these compounds have been found to show cytotoxic activity against KB cell cancer (human oral epidermoid carcinoma); thus, this plant is known as a source of promising anticancer drug [6]. Furthermore, artonin E, artonin O, artobiloxanthone, and cycloartobiloxanthone have also been found active as anticancer [5–7]. However, the antimalarial activity test of the compounds isolated from A. kemando is not available yet.

The previous results of in vitro antimalarial activity test from flavanoid compounds obtained from Artocarpus indicated that many flavanoids were active and considered as antimalaria [1–4], but no reports have been found for the in vivo antimalarial test. Thus, in this paper, we reported the in vivo antimalarial test of artocarpin isolated from A. altilis and in vitro antimalarial activity of artonin M isolated from A. kemando Miq.

2. Materials and methods

2.1. Plant materials

The root woods of A. altilis were collected from Banjar Negara village, Tanggamus, Lampung, and the root bark of the Pudau plant (Artocarpus kemando Miq.) was obtained from Karang Anyar Hamlet, Klaten Village, Penengahan District, South Lampung Regency, Lampung Province, Indonesia. They were identified at Herbarium Bogoriense, Research Center for Biology, Indonesian Institute of Sciences, Bogor, Indonesia, and a voucher specimen of each plant has been deposited at the herbarium.

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2.2. General Experimental Procedures

Thin layer chromatography (TLC) analysis was carried out on pre-coated Si-gel plates (Merck Kieselgel 60 F254), and the UV lamp of Spectroline, ENF-240 C/F model was used to see the spot in TLC. VLC was carried out using Merck Si-gel 60. Melting points were determined on a Fisher Jhons micro-melting point apparatus and were uncorrected. UV-Vis and IR spectra were measured with Beckman DU-7000 and Varian 2000 FTIR spectrophotometers, respectively. $^1$H-NMR spectrum was recorded with a JEOL ECA 500 spectrometer, operating at 500.00 MHz and $^{13}$C-NMR operating at 125 MHz.

2.3. Isolation and Purification of the Compounds

2.6 kg of root wood A. altilis was mashed and was macerated using 16 L of methanol solvent, for 3x24 hours. The maceration results were then filtered and concentrated, obtained 83.46 grams of the extract. The maceration extract was fractionated using Vacuum Liquid Chromatography (VLC), received 2 stages of VLC using Merck 60 Silica Gel adsorbent (35-70 Mesh) and amylose, and eluted with a mixture of ethyl acetate/n-hexane which gradually increased its polarity. Fractionation results produced 4 main fractions (A–D), fraction B 0.9 g, C 1.93 g, and D 13.8 g.

The fractions B and C were VLC and subsequently in CC repeatedly using the same adsorbent and eluent, compound (1), as much as 0.63 grams, and exhibited melting point 185°C – 186.8°C. TLC with an artocarpine standard using three systems of ethyl acetate / n-hexane eluent 2: 8 (Rf 0.14), acetone / dichloromethane 1: 9 (Rf 0.51), and acetone / n-hexane 3: 7 (Rf 0.37), and one stain was obtained with the same RF.

2.0 kg fine powder of root bark was macerated with a methanol solvent for 1x24 hours with three repetitions. The results of maceration were evaporated using a rotary evaporator and obtained 149.8 grams of extract. The extract obtained was then fractionated by the VLC method, with adsorbent silica gel and eluent n-hexane and ethyl acetate with variations in polarity increase. The results of the fractionation obtained seven main fractions, A – G. Fraction C received 16.7 grams in VLC further with the same adsorbent and eluent, and after that, the column chromatography was repeated using silica gel adsorbents and variations in n-hexane and ethyl eluents. Acetate and n-hexane and acetone obtained yellow crystals (compound 2) weighing 8.8 mg, with a melting point 248 – 251 °C. In TLC compounds (2) with 3 eluent systems, namely n-hexane: acetone: dichloromethane (DCM) (2: 1: 1 (c), acetone: n-hexane: (3: 7) (b), EtOAc: n -hexane (2: 8), one stain was obtained.

Artocarpin (1) (Figure 1): yellow needle crystal, melting point 185°C – 186.8°C; UV-Vis $\lambda_{\text{max}}$. (MeOH), nm (log e): 279 (4.56) and 324 (4.11); $\lambda_{\text{max}}$. (MeOH + NaOH): 278 and 362; the $\lambda_{\text{max}}$. change was not observed in MeOH + AlCl$_3$ or in MeOH + NaOAc. IR $\nu_{\text{max}}$. (KBr) cm$^{-1}$: 3392, 3380, 2960, 2930, 1647, 1620, 1483, 1451, 1362, 1207, 1153, 1099, 978, 850, and 809. $^1$H-NMR (in acetone-D$_6$, 500 MHz) $\delta$ (ppm): 6.56 (1H, s, H-8), 3.12 (2H, d, J = 7.1 Hz, H-9); 5.12 (1H, m, H-10); 1.43 (3H, s, H-12), 1.57 (3H, d, J = 1.3 Hz, H=13), 6.6 (1H, d, J = 16.2 Hz, H-14), 6.7 (1H, dd, J = 7.1 and 17.2 Hz, H-15), 2.41 (1H, m, H-16), 1.07 (6H, d, J = 6.5 Hz, H-17 and 18), 7.2 (1H, d, J = 8.45 Hz, H-6'), 6.51 (1H, dd, J = 2 and 8.4 Hz, H-5'), 6.57 (1H, d, J = 2 Hz, H-3'), 3.96 (3H, -OCH$_3$, s). $^{13}$C-NMR (in acetone-D6, 125 MHz): $\delta$ (ppm): 163.9 (C-2), 122.0 (C-3), 183.4 (C-4), 105.6(C-4a), 159.9 (C-5), 109.8 (C-6), 163.9 (C-7), 90.7 (C-8), 159.9 (C-8a), 112.9 (C-1'), 157.5 (C-2'), 103.8 (C-3'), 161.6 (C-4'), 108.1 (C-5'), 132.4 (C-6'), 24.7 (C-9), 122.6 (C-10), 132.2 (C-11), 25.9 (C-12), 17.7 (C-13), 117.1 (C-14), 142.3 (C-15), 34.1 (C-16), 23.2 (C-17 and C-18), and 56.7 (-OCH$_3$). The $^1$H and $^{13}$C NMR spectra of this compound are shown in Figures 2 and 3, while the HMQC and HMBD spectra of 1 is in Figures 4 and 5, and the important correlation of 1 is in Figure 6.

Artocarpin (2) (Figure 1): yellow needle crystal, melting point 248 – 251 °C. UV-Vis $\lambda_{\text{max}}$. (MeOH) nm (log e): 203 (4.66, H); 290 (4.48); 348 (4.19); and 385 (4.27); $\lambda_{\text{max}}$. (MeOH + NaOH): 212, 266, 388, and 467. IR $\nu_{\text{max}}$. (KBr) cm$^{-1}$: 3568, 3427, 2974, 2924, 1653, 1598, 1550, 1473,1355, 1271, and 1151. $^1$H-NMR (in methanol-D4, CD$_3$OD, 500 MHz) $\delta$ (ppm): 1.31 (3H, s, H-12); 1.45 (6H, s, H-22
and H-23); 1.67 (3H, d, J = 1.2 Hz, H-17); 1.70 (3H, d, J = 1.4 Hz, H-18); 1.95 (2H, m, H-14); 2.2 (2H, m, H-13); 2.40 (1H, t, J = 15.2 Hz, H-9), 3.10 (1H, dd, J = 7 and 15.1 Hz, H-9); 3.41 (1H, dd, J = 7 and 15.2 Hz, H-10); 5.2 (1H, t, J = 1.5 and 7 Hz, H-15); 5.67 (1H, d, J = 10 Hz, H-20); 6.23 (1H, d, J = 1 Hz, H-3'); 6.40 (1H, s, H-8); and 6.65 (1H, d, J = 10 Hz, H-19).

![Chemical structure of compounds 1 and 2](image)

**Figure 1.** Chemical structure of compounds 1 and 2

![NMR spectrum](image)

**Figure 2.** $^1$H NMR spectrum of artocarpin (1)
Figure 3. $^{13}$C NMR spectrum of artocarpin (1)

Figure 4. HMQC spectrum of 1
2.4. Antimalarial activity

The in vitro antimalarial assay was performed following the procedures available in the literature and have been used previously in some of our publications [8-10]. However, the in vivo antimalarial test was carried out based on the procedure used published methods available in the literatures [11] and have also been used in our previous work [4].

3. Results and discussions
3.1. In vivo antimalarial test of artocarpin (1)

The growth of Plasmodium was inhibited upon administering the compound 1, where the higher dosage of compound 1 was used, in addition to the smaller parasitemia average and the higher percentage inhibition average. The dosage that resulted in the highest inhibition of Plasmodium was 100 mg/kgBW, where it resulted in inhibition of 73.66 ± 0.12%. In the smallest dosage used, the inhibition of Plasmodium was still observed, although it was very low, with a percentage of 12.60 ±
0.31%. The probit calculation resulted the effective dose 50 (ED₅₀) of 34.88 mg/kgBW (Table 1). The parasitemia level and inhibition percentage of artocarpin is in Figure 7. According to Munoz et al. [12], the in vivo antimalarial activity was categorized as follows: ED₅₀ ≤ 100 mg/kgBW/day is categorized as excellent, ED₅₀ 101-250 mg/kgBW/day categorized as good, ED₅₀ 251-500 mg/kgBW/day as medium, and ED₅₀ > 500 mg/kgBW/day as inactive. Based on these criteria, compound 1 is categorized as excellent drug; therefore, it will be excellent to be developed as an antimalarial drug in the near future.

![Figure 7. The parasitemia level and inhibition percentage of isolated artocarpin (I)](image)

### Table 1. Parasitemia value, inhibition percentage, ED₅₀, ED₉₀ and ED₉₉ of isolated artocarpin (I)

| Dosage (mg/KgBW) | Parasitemia (%) | Inhibition Percentage (%) | ED₅₀ (mg/KgBW) | ED₉₀ (mg/KgBW) | ED₉₉ (mg/KgBW) |
|------------------|-----------------|---------------------------|----------------|----------------|----------------|
| 100.00           | 0.69%           | 73.66%                    |                |                |                |
| 50.00            | 1.18%           | 55.14%                    |                |                |                |
| 25.00            | 1.38%           | 47.49%                    | 34.88          | 286.15         | 1591.13        |
| 12.50            | 1.91%           | 27.28%                    |                |                |                |
| 6.25             | 2.29%           | 12.64%                    |                |                |                |

In the in vivo antimalarial activity test, some factors that demonstrate an effect on the result of the test exist. The individual factor of mice really affects the growth of *P. Berghei*, which also gives the varied picture of parasitemia. This condition normally affects the growth of *P. berghei* in the body of mice. The factor of Plasmodium also affects the number of parasitemia that occur in the mice. The host factor influencing the mice is the body resistance to eliminate Plasmodium from each mouse, while the factor of *P. berghei* that does not sincronize in the mice and is normally only 10% of *P. berghei* that was inoculated and can grow [13].

The presence of schizogony erythrocytic cycle on *P. berghei* inside the visceral organ, making the ring stage and trophozoit, are mostly taken as innoculum. The character of *P. berghei* also affects the
speed of its growth inside the host [14]. Thus, these conditions perhaps is the cause of the parasitemia value being relatively low. The parasitemia negative control was only 2.63 ± 0.01 %.

The mechanism of the compound tested as antimalarial was perhaps by inhibition mechanism of the heme polymerization. It has been found that the terpenoid compound will bind to a heme electronic system, and the hydroxyl group will be bound to heme iron [15]. The other mechanism in killing Plasmodium is probably available but which mechanism used is not known exactly.

2.2. In vitro antimalarial of Artonin M (2)

The results of in vitro antimalaria of 2 showed that this compound exhibits very high activity against Plasmodium, giving an IC50 value of 0.3 µg/mL (5.976 x 10⁻⁷ M), and this is comparable with the control positive chloroquine with an IC50 value of 3 x 10⁻⁷ M. This value is stronger than the antimalarial drug from the organotin (IV) compound, which has been reported lately [9,10,16] and may be better than other synthetic compounds having antimalarial activity which have been reported by others [17-20]. The data of activity test can be seen in Figure 8 and Table 2, while the data for the chloroquine are in Figure 9.

**Figure 8. In vitro antimalarial assay of artonin M**

**Figure 9.** The result of in vitro antimalarial activity of chloroquine
Each molecule structure of 1 and 2 exhibits two hydroxyl groups at the meta position on B ring. The hydroxyl group at C-7 on A ring on these two compounds is not on a free state, and they exhibit one uncyclinazed isoprenyl group on the C ring at compound 1 and on furanodihydrobenzosanton at compound 2. The structure of these two compounds is similar for the position of two hydroxyl groups on B ring and the presence of prenyl or geranyl on C ring, thus making these compounds as active as antimalaria.

4. Conclusions

The result of in vivo antimalarial assay of artocarpin ED$_{50}$ value obtained was 34.88 mg/kgBW. This value indicated that artocarpin demonstrates very strong antiplasmodial activity. The result of in vitro antimalarial activity of artonin M resulted in strong activity with an IC$_{50}$ value of 0.3 μg/mL. This IC$_{50}$ value also is an indication that artonin M has the potential to be developed as an antimalarial drug. The in vivo antimalarial of artocarpin and in vitro antimalarial of artonin M from Artocarpus plant are the first report of antimalarial activity assay. Based on the result reported here, the conclusion exists that A. altulis and A. kemando plants are one of the main sources to obtain the potential compound as antimalarial, which will be very useful for the replacement of some commercial drugs available in the market and caused the antimalarial resistency.

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