ABSTRACT

Currently, the Milwaukee protocol presents healing results in human beings affected by the rabies virus. However, there are many points to clarify on the action of drugs and the immune mechanism involved in the evolution of the disease. One of the drugs used is biopterin, which is an important cofactor for nitric oxide, important for preventing vasospasm. Thus, we describe the effect of biopterin on some inflammatory factors in a rabies virus infection developed in an animal model. The immunological mediators studied in animals infected with rabies virus submitted to doses of sapropterin were Anti-RABV, IL-6, IL-2, IL-17a, INF-gamma and Anti-iNOS. It is suggested that the medication in the context of a RABV infection already installed, had the effect of modulating the inflammatory mechanisms mainly linked to the permeability of the blood-brain barrier and the migration of cytotoxic cells.

KEYWORD: Rabies. Rabies virus. Tetrahydrobiopterin. Blood-Brain Barrier. Interleukin-6.

INTRODUCTION

The Rabies virus (Rabies Lyssavirus - RABV) is an RNA virus of the family Rhadoviridae that presents tropism for the nervous system, causing encephalitis. The onset of the symptoms occurs from one to three months after exposure to saliva or fluids of an infected mammal, which may occur after a long period. The disease has three classic forms: the furious form, the paralytic form and the atypical form. The outcome of human rabies cases is, in most cases, death and the time for it to occur varies according to the support given to the patient.

The surviving rate of patients with encephalitis is low. Survivors were submitted to the Milwaukee protocol, which aims are to reach an early stabilization of the patient and control of dysautonomies resulting from neuronal and inflammatory dysfunctions generated by the virus. The surviving rate of patients with encephalitis is low. Survivors were submitted to the Milwaukee protocol, which aims are to reach an early stabilization of the patient and control of dysautonomies resulting from neuronal and inflammatory dysfunctions generated by the virus.

One of the pharmacological components of the protocol is tetrahydrobiopterin (BH4), with its functional pharmacological form, sapropterin, a cofactor in the synthesis of nitric oxide, also participating in the metabolism of phenylalanine into tyrosine and in the conversion of tyrosine to levodopa and tryptophan to 5-hydroxytryptophan. Clinically, it is indicated for phenylketonuria, which is caused by a metabolic error in the production of BH4 due to a failure of the enzyme dihydropteridin reductase, affecting the central nervous system metabolism in several ways.

BH4 is used specifically in rabies virus infection due to its reduced dosages
in the cerebrospinal fluid (CSF) of infected patients and the fact that it can, in theory, prevent cerebral spasms by regulating the production of nitric oxide. In the protocol, there is an association of BH4 with ascorbic acid to promote the recycling of dihydrobiopterin into tetrahydrobiopterin, administered together to control the disease, concomitantly with amantadine, fludrocortisone, nimodipine and ketamine, each of them with precise indications, but they are not within the scope of this study. Thus, the aim of the present study is to analyze the immunological effect of biopterin on rabies virus infection, in a murine model.

**MATERIALS AND METHODS**

**Viruses and drugs**

In this study, samples of the wild strain rabies virus from bats carrying the antigenic variant (VAg 3) were used, provided by the Section for Arboviruses and Hemorrhagic Fevers (SAARB) of the Evandro Chagas Institute (IEC), Para State, Brazil more specifically from the Rabies Laboratory. The sample was titrated to 3.2 DL50/0.02 mL using the Reed and Muench method.

The drug used was tetrahydrobiopterin (Sapropterin form) (50 mg/kg/day) orally administered by gavage, which consists of the introduction of an aluminum cannula device for mice that allows the intra-gastric inoculation of the medication. The commercial name of the drug is KUVAN® 100 mg, soluble tablets with sapropterin and dihydrochloride as an active ingredient.

**Infection experiment**

Thirty-seven female BALB-C mice (M. musculus) aged 3-4 weeks, with a maximum of 5 animals in each cage (50 x 60 cm), were used. The animals received water and feed at will, and with respect to the circadian cycle, they were placed in appropriate structures for cages to maintain the air circulation. The experimental model of rabies inoculated a suspension containing 20% of the brain of a mouse with the antigenic variant 3 (VAg 3) of RABV, in a titration of 3.2 DL50/0.02 mL, leading to a 0.03 mL intramuscular (IM) injection in the lateral region of the right hind leg.

The animals were euthanized with Xylazine and Ketamine in double doses, followed by cardiac puncture, so that a better analysis could occur by the proposed methods.

**Treatment with biopterin**

The 37 infected animals were divided into two groups; the control group (19 animals), with only infection, and the tetrahydrobiopterin group (18 animals), in which sapropterin was administered via gavage. The animals were followed for 17 days of disease progression and on the first day of manifested symptoms, the administration of the drug to all animals began. Then, the groups were paired, following the experimental case-control design of the study. The choice to start the treatment after the onset of symptoms in the first animal intended to mimic the disease in humans, and after the disease establishment, this animal was subjected to a direct immunofluorescence test to confirm the presence of the infection.

Every day gavages were carried out with the drug calculated for each animal weight, diluted in 0.9% saline. On days: 3, 5, 7, 9, 11 and 13 of drug administration, three animals from each group were euthanized to analyze the effect of the medication on the markers proposed in this study.

The following immunological markers anti-RABV, IL-6, interferon gamma, IL-2, IL-17a and Anti-iNOS were analyzed on three animals/day over the course of 13 days, and the cerebral hemisphere was collected for analysis and comparison purposes.

**Flow cytometry**

After maceration, the hemi-brains of all mice were analyzed, the technique was performed according to the manufacturer’s instructions (BDTM Cytometric Bead Array (CBA) Mouse Inflamatory- BD Biosciences/USA) in order to analyze the production of cytokines interleukin -2 (IL-2), IL-6, interferon-gamma (IFN-gamma) and IL-17A. The samples were evaluated on the flow cytometer (BD FACSCanto II), the data were processed by the FACS DIVA software and statistical analysis on the GraphPad Prism 5 software (5.0, GraphPad Software, San Diego, EUA) using the One-way ANOVA test with p <0.05.

**Immunohistochemistry**

In the immunohistochemical processing, the immunoperoxidase technique was used for immunostaining with a commercial Vector Mouse on Mouse Basic Kit (M.O.M. kit, Vector Laboratories, Burlingame, USA) in compliance with the manufacturer’s instructions with modifications. Slides containing histological sections of the organs were deparaffinized in increasing concentrations of xylol (50% and 100%) and decreasing concentrations of alcohol (100%, 95%, 80% 70% and 30%) and later washed with distilled water.

For antigenic recovery, tissue cuts were incubated at a high temperature in a pressure cooker for 30 min/125 °C.
with tris-citrate pH: 7.2, subsequently permeabilized with 0.5% Triton X 100 solution, followed by the blocking of nonspecific sites with a solution containing Mouse IgG Blocking Reagent incubated as the primary antibody for 1 h, followed by washing in PBS to continue the incubation in a protein concentrate solution for 30 min.

The incubation with the primary antibody occurred for 12 h under agitation followed by washing; for the next incubation in hydrogen peroxide diluted in water (proportion of 1:10) for 15 min and finally tissue cuts were incubated in the ABC reagent for 1 hour.

For the immunohistochemical reactions, the DAB/nickel solution was used, mounted between slide/coverslip with Entellan (Merk, Germany), analyzed in a light field optical microscope (Axiophot – Zeiss, Oberkochen, Germany) and photographed with a digital camera (AxioCam HRC – Zeiss, Oberkochen, Germany).

The reagents used to analyze the brains of euthanized animals were Anti-RABV and Anti-iNos (Figure 1). The counting of marked cells was made with a grid in ten 400x magnification fields.

**Statistical analyses**

The Student independent t test was used to compare the tetrahydrobiopterin group and the control group and the ANOVA one criterion was used within each group, and between the results obtained on days 3, 5, 7, 9, 11 and 13, and a p value <0.05 was considered significant. Data were tested to confirm that a normal distribution has taken place.

In the evolution line of the tables, the values of increase or decrease in the time line were calculated, the plus sign (+) being the increase and the minus (-) sign, the decrease.

**RESULTS**

Table 1 shows the results of the anti-RABV analysis, with a higher mean of detectable antibodies visible in

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**Figure 1** - Immunohistochemistry images of the nervous tissue of the studied mice: A) Nervous tissue marked with anti-i-NOS. The arrow points to a positively-marker cell; B) A negative control of anti-i-NOS in nervous tissue; C) Anti-RABV marking, with the arrow pointing to a positively-marked cell; D) A negative control of anti-RABV.
the experimental group, being not possible to verify the oscillation in relation to time (p=0.045). However, it is worth noting that the group submitted to the drug had significantly lower values than the control group. In Table 2, there was a comparison of IL-6 between the groups, with no statistical differences (p=0.2113). After the ninth day of medication, the animals in the group using sapropterin did not present detectable amounts of IL-6.

Table 1 - Comparison of anti-RABV immunohistochemistry analysis in the Infection Control and the Infection Experimental group, in mice, IEC, Brazil, 2020.

| Time (days) | Tetrahydrobiopterin group Mean ± SD | Control group Mean ± SD | p-value |
|-------------|-------------------------------------|-------------------------|---------|
| 3           | 0.15750 ± 0.049                     | 0.38292 ± 0.086         | 0.0086* |
| 5           | 0.41000 ± 0.103                     | 0.41000 ± 0.103         |        |
| 7           | 0.39028 ± 0.329                     | 0.34833 ± 0.085         | 0.4207  |
| 9           | 0.29114 ± 0.163                     | 0.34167 ± 0.025         | 0.3248  |
| 11          | 0.28333 ± 0.154                     | 0.35632 ± 0.044         | 0.2368  |
| 13          | 0.32398 ± 0.015                     | 0.15833 ± 0.079         | 0.0115* |

Result + 0.16648 - 0.22459
p-value 0.0453** 0.0192** 

*Student Independent t-test; **One-Way ANOVA test

Table 2 - Comparison of IL-6 Flow Cytometry analysis in the Infection Control and the Infection Experimental group in mice, IEC, Brazil, 2020.

| Time (days) | Control group Mean ± SD | Tetrahydrobiopterin group Mean ± SD | p-value |
|-------------|-------------------------|-------------------------------------|---------|
| 3           | 3.1100 ± 2.81           | 4.1200 ± 3.59                       |         |
| 9           | 3.5450 ± 5.01           | 0.0000                              |         |
| 13          | 0.4967 ± 0.86           | 0.0000                              |         |

Result** - 2.6133 - 4.1200
p-value 0.0000 

*p = 0.0401 Student Independent t-test; **One-Way ANOVA test

Table 3 - Comparison of IFN-gamma Flow Cytometry analysis in the Infection Control and the Infection Experimental group in mice, IEC, Brazil, 2020.

| Time (days) | Control group Mean ± SD | Tetrahydrobiopterin group Mean ± SD | p-value |
|-------------|-------------------------|-------------------------------------|---------|
| 3           | 4.9100 ± 1.01           | 4.7500 ± 1.17                       |         |
| 9           | 4.5300 ± 0.49           | 3.7200 ± 0.43                       |         |
| 13          | 3.6267 ± 0.07           | 4.1450 ± 0.05                       |         |

Result - 1.2833 - 0.6050
p-value 0.1529 0.3547

*p = 0.3856 Student Independent t-test; **One-Way ANOVA test

Table 4 - Comparison of IL-2 Flow Cytometry analysis in the Infection Control and the Infection Experimental group in mice, IEC, Brazil, 2020.

| Time (days) | Control group Mean ± SD | Tetrahydrobiopterin group Mean ± SD | p-value |
|-------------|-------------------------|-------------------------------------|---------|
| 3           | 11.6933 ± 1.57          | 9.9767 ± 3.61                       |         |
| 9           | 8.3150 ± 2.92           | 7.9867 ± 0.56                       |         |
| 13          | 5.4033 ± 2.12           | 9.7750 ± 1.10                       |         |

Result - 6.2900 - 0.2017
p-value 0.0398** 0.5791

*p = 0.0401 Student Independent t-test; **One-Way ANOVA test

Table 5 - Comparison of IL-17A Flow Cytometry analysis in the Infection Control and the Infection Experimental group, in mice, IEC, Brazil, 2020.

| Time (days) | Control group Mean ± SD | Tetrahydrobiopterin group Mean ± SD | p-value |
|-------------|-------------------------|-------------------------------------|---------|
| 3           | 7.5367 ± 1.06           | 8.5067 ± 1.33                       |         |
| 9           | 7.4200 ± 1.97           | 6.9133 ± 0.51                       |         |
| 13          | 7.0533 ± 0.78           | 6.5600 ± 0.41                       |         |

Result - 0.4834 - 1.9467
p-value 0.8824 0.1165

*p = 0.0146 Student Independent t-test; **One-Way ANOVA test

Table 6 presents the results of the Anti-iNOS during the days in which the animals were euthanized, and there was no difference in the quantification of antibody markers between the groups (p = 0.0066).

DISCUSSION

Human rabies is considered to be a controlled disease
Immunological impact of tetrahydrobiopterin on the central nervous system in a murine model of rabies virus infection

**Table 6 - Comparison of INOS immunohistochemistry analysis in the Infection Control and the Infection experimental group, IEC, Brazil, 2020.**

| Time (days) | Tetrahydrobiopterin group | Control group | p-value |
|------------|---------------------------|---------------|---------|
|            | Mean ± SD                 | Mean ± SD     |         |
| 3          | 0.019792 ± 0.009          | 0.06563 ± 0.023 | **0.0164*** |
| 5          | 0.120834 ± 0.044          | 0.10208 ± 0.030 | 0.2875 |
| 7          | 0.234375 ± 0.075          | 0.24306 ± 0.081 | 0.4491 |
| 9          | 0.180729 ± 0.092          | 0.35052 ± 0.128 | 0.0673 |
| 11         | 0.280209 ± 0.011          | 0.35260 ± 0.017 | 0.2875 |
| 13         | 0.272917 ± 0.019          | 0.29948 ± 0.014 | 0.4276 |
| Result     | + 0.253125               | + 0.23385     |         |
| p-value    | **0.0066**               | **0.0273**    |         |

*Student Independent t-test; **One-Way ANOVA test

Since the widespread use of vaccines and immunoglobulins, but lethal cases after the bite of contaminated non-human mammals still occur, mainly in developing countries. There are an estimated 59,000 cases of deaths annually in more than 150 countries, most of which occur in Africa and Asia, so the establishment of a treatment for the disease already installed is necessary and one of the drugs indicated in the Milwaukee Protocol currently in vigor, is sapropterin\(^3\,^4\,^6\,^7\). The main indication for tetrahydrobiopterin (BH4), and its pharmacological form sapropterin, is for the severe form of phenylketonuria, an autosomal recessive genetic disease that generates errors in the enzyme phenylalanine hydroxylase, an essential cofactor for dopamine, noradrenaline and serotonin, leading to clinically with delayed psychomotor development, microcephaly, tremors and limb uncoordination\(^8\).

In human rabies, one of the therapeutic advantages of the use of sapropterin is the low levels of BH4 s in the CSF of patients treated with the disease already installed, and in association with other drugs, the possible beneficial effects on the pathophysiology of the disease, possibly changing the outcome of the disease, that in most cases is lethal\(^9\,^{12}\).

The inflammatory process generated by the virus is the main responsible for the lethality of the disease, as stated by Hooper\(^10\). The mechanism of response to depletion of most viruses initially passes through cytotoxicity and apoptosis of infected cells that is later used by the immune system via a specific mechanism of neutralization carried out by antibodies against viral surface proteins – participating in the adaptive immunity\(^11\,^{12}\).

The activation and stimulation of cytotoxic cells, as well as the stimulation of apoptosis, instigated by several types of T lymphocytes, are extremely harmful elements in the nervous tissue, since it has little margin for plasticity and regeneration when compared to other tissues. In addition to this nerve cell destruction factor, there is also a tissue edema that generates dysfunction and dysautonomy, very common in patients infected with the rabies virus\(^15\,^{17}\).

These two mechanisms are closely related to the activity of Interleukin 6 (IL-6), which is responsible for increasing the exposure capacity of adhesion molecules on the wall of brain vessels, facilitating the migration of white cells to the CNS, in addition to promoting the entry of liquid content, rich in antibodies and other inflammatory mediators, thus initiating an exponential cycle of innate inflammatory response, which stimulates activities that, in controlled doses, are capable of generating healing in mice with installed rabies. However, in cases with high IL-6 expression, neuronal loss begins, with irreversible and lethal sequelae\(^18\,^{20}\).

In rabies strains, with high potential for IL6 expression, a high neuronal injury power is observed, due to the high expression of type I histocompatibility molecules with greater activity of CD4 cells. This element can occur through IFN-gamma (interferon gamma) mediation, which is also a stimulator of adhesion molecules on the endothelial wall of CNS cells, in addition to being a ripener of macrophages\(^21\,^{22}\).

In the present study, it is possible to observe that there is a sign of decrease in the concentration of IL-6 in the group submitted to the studied drug, IL-6 (p=0.21) and INF-gamma (p=0.38), are important actors in the control of the inflammatory response at the CNS level in response to rabies infection, since, with a greater control of the entry of cytotoxic cells, the innate response of the immune system will suffer less damage, being an appropriate time for the adaptive immunity to respond in an increasing way, without causing serious sequelae or death, which could help explaining the success of cases in humans submitted to the Milwaukee protocol. The decrease in viral concentrations was found in the experimental group, which demonstrates a greater viral clearance capacity related to the use of the drug, at least in this murine model\(^23\,^{25}\).

As stated by Luo et al\(^18\), the more immunogenic the viral strain, the greater the ability of the antibody to generate an immune response, but also generating more neuronal damage through the innate response. It is worth noting that in the case of a negative regulation of the immune response, mainly of IL-6, INF- gamma and TNF, there will be a better systemic response, so that viral strains with overexposure to IL-6 can generate more damage\(^18\,^{26}\,^{27}\).

Along with the two immunological mediators already discussed, IL17, which is produced by TCD4 cells and is closely related to cell recruitment, decreased in the experimental infection group. Interestingly, the
experimental group (p=0.014), showed less IL-2 depletion, which is extremely important mainly for the maturation of T and B lymphocytes, being responsible for the innate and adaptive immune response, respectively. The total non-depletion of IL2 is necessary for activities within the micro environment of the CNS, as it will assist in the maturation of white cells for the immediate innate response and controlling the viral dispersion within the CNS28-33.

As for the inducible NOS (iNOS), it is an element of the nitric oxide synthase (NOS) family, which acts as an inflammatory mediator, mainly for microglia and macrophages. It is an important mediator of the inflammatory process in infectious diseases, in the present study, changes in this marker were not observed in either the experimental group or in the control group (p=0.006). It can be inferred that the drug does not interfere with the activity of cells that use iNOS as an immune mediator in the CNS, which was expected, since BH4 is not an essential component in the synthesis of iNOS34,35.

Even with criticisms of the Milwaukee protocol, it is currently applied to patients with an installed viral infection, but the role of each drug is still speculated, requiring studies in experimental and clinical models, even if experimental models are difficult to manage, due to the power of viral lethality and the need for a high biosafety laboratory structure.

CONCLUSION

The present study suggests that the use of sapropterin in mice infected with the wild rabies virus has a significant impact on the permeability and integrity of the cephalic blood-brain barrier, mainly through mechanisms mediated by IL-6. However, further studies using tools such as mRNA of inflammatory mediators are needed to verify the integrity of the blood-brain barrier to deepen and elucidate the mechanism indicated by this research.

Based on the data found, it is possible to suggest that the influence of tetrahydrobiopterin on the CNS inflammatory process was affected to regulate the inflammatory process, thus generating a possible decrease in cytotoxic cells, which migrated to the CNS due to the fragility of the r hematoencephalic barrier and preserving neural tissue. More studies regarding the interaction of drugs, viral infection and the immune system, are needed to elucidate the positive impacts obtained with the use of different drugs of the Milwaukee protocol in humans.

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CONFLICT OF INTERESTS

The authors declare no conflict of interests.

ETHICAL STATEMENT

The experiment was conducted at the laboratory of the Evandro Chagas Institute with the approval of the Ethics Committee on Animal Use (CEUA) of the Evandro Chagas Institute (IEC), under the Nº 37\2017, approved on September 13, 2017.

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