Genetic polymorphisms as determinants of pesticide toxicity: Recent advances

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1. Introduction

In the last decades, several epidemiological and observational studies [15]. Although in recent years the environmental emission of these substances has been reduced and specific guidelines regulate their use, even today a significant fraction of the population is exposed to the risk of environmental pollutants, especially as regards direct and indirect exposure to pesticides. Case studies demonstrated that occupational and environmental exposure to several factors, including benzene, heavy metals, fibers, chemicals, and pesticides may induce detrimental effects on human health by promoting the development of a wide range of disorders, including infectious diseases, immunological diseases, reproductive and neurodegenerative diseases, hematological malignancies, and solid tumors [1–5]. Although in recent years the environmental emission of these substances has been reduced and specific guidelines regulate their use, even today a significant fraction of the population is exposed to the risk of environmental pollutants, especially as regards direct and indirect exposure to pesticides [6].

The term pesticide includes a wide and miscellaneous category of chemicals conceived to prevent and defeat weeds or pests. Although their efficacy has contributed to the wide use, occupational and environmental exposure is a threat to public health. Subjects professionally exposed to pesticides include farmers, gardeners and people working in the production, transportation and sales of these compounds. Some categories of workers (e.g. greenhouses workers) may be exposed to high concentrations of pesticides with potential health consequences [7,8].

Numerous studies confirm the hypotheses according to which exposure to pesticides can produce detrimental effects on respiratory or reproductive systems and bring to the development of chronic diseases or cancer. Mutagenic mechanisms affecting oncogenes have been better understood in vitro, although, also in humans, there is evidence of the association between exposure to pesticides and the presence of mutations in key genes responsible for cancer development and progression [9]. Still, it has been observed that the chromosome translocation t (14; 18) has a higher prevalence in farmers exposed to pesticides, with a greater risk of developing non-Hodgkin’s lymphoma (NHL) [10]. Numerous pesticides have been classified by IARC in group 2A as probably carcinogenic to humans [11]. Acute exposure symptoms are easily identified, but the link between real exposure and the development of chronic pesticides-related illness is not easy to assess [5,12–16].

Recent studies have suggested oxidative stress as one of the mechanisms for the adverse health effects of pesticides exposure; the alteration of the physiological balance brings to the excess of oxidant species, resulting in severe damage to cellular components and macromolecules, especially the DNA [17–21]. Moreover, pesticides exposure may induce the modulation of cells functionality, especially that of the immune system (e.g. macrophages, lymphocytes, etc.), leading to the onset of a pro-inflammatory microenvironment responsible for the accumulation of genetic damages and, in turn, for the accumulation of

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ABSTRACT

Occupational and environmental exposure to pesticides may induce harmful effects on human health by promoting the development of a wide range of disorders. Some of the most recently hypothesized mechanisms are oxidative stress and epigenetic modifications, however biological effects seem to be mediated mainly by the occurrence of genetic polymorphisms. The susceptibility to exposure can be evaluated by studying the most common polymorphisms of genes involved in the metabolism of organophosphorus compounds (cytochrome P450, glutathione transferase, acetyltransferases or paraoxonase 1). The aim of this article is to review recent literature data concerning the influence of genetic polymorphisms on pesticides-induced oxidative damage.
altered cells that were not correctly removed by the immune system [22,23]. Indeed, it is well known that macrophages alterations, and in particular the abnormal secretion of several secreted proteins, play a key role in tumorigenesis and autoimmune diseases [24–28].

Furthermore, recent evidence showed that pesticides are able to induce both genetic and epigenetic alterations in the exposed individuals [29,30]. In particular, it was demonstrated that pesticides are associated to a wide range of genetic polymorphism affecting key genes involved in the regulation of cell cycle, the redox status and drug metabolism [31,32]. On the other hand, emerging evidence showed that pesticides may regulate gene expression through DNA methylation and microRNAs (miRNAs) de-regulation. On this matter, many bioinformatics data have been generated. Moreover, new computational approaches have been developed to identify in silico specific miRNAs or methylation hotspots associated with the presence of disease or to be used as bio-indicators of environmental exposure to pesticides [5,30,33–42].

Other studies demonstrated that some pesticides may induce modifications in the composition of gut microbiota and that the microbiota itself may alter the response of the organism and the metabolism of pesticides by enhancing or inhibiting the activity of cytochrome enzymes [43]. Of note, gut microbiota is involved in the regulation of several physiological and pathological processes, including cancer [44–45] and neurodegenerative diseases [46]. Therefore, there is raising evidence that modifications of microbiota induced by the exposure to pesticides may represent a trigger for the development of several diseases [47].

Although oxidative stress, epigenetic modifications and gut microbiota modulation represent key pathogenic mechanisms induced by pesticides exposure, their biological effects seem to be modulated mainly by the occurrence of genetic polymorphisms. In particular, some individuals could be more susceptible to pesticide-induced oxidative stress because of the influence of genetic polymorphisms. The susceptibility to exposure can be evaluated by studying the most common polymorphisms of cytochrome P450, of glutathione transferases (including GSTM1, GSTP1, GSTT1); of acetyltransferases (NAT2) and paraoxonases (mostly PON1), which are mainly involved in the metabolism of organophosphorus compounds [48]. In the last years, several Authors focused their attention on glutathione-s-transferases (GSTs) family and paraoxonase (PON1, PON2, and PON3) family and literature data are continually being updated in this field [49–54].

Genetic heritage may increase susceptibility for the development of acute or chronic diseases, especially in subjects already exposed to other pollutants [52,55–57]. This article aims to review recent literature data concerning the influence of genetic polymorphisms on pesticides-induced oxidative damage.

2. Methods

The material for this article was collected by examining the literature available in the PubMed and Scopus database. A search was performed using in combination the following keywords: pesticides AND polymorphisms OR pesticides AND genetic AND polymorphisms. The research was based on original research work and review documents related to the topic published mainly in the last 10 years. The relevance of the subject and the eligibility of all publications detected was evaluated basing on titles and abstracts. The reference lists of selected articles were further screened for relevance. Non-English studies were not taken into consideration.

2.1. Polymorphisms

Genetic polymorphism is the presence in a population of two or more alleles in a particular locus with a frequency greater than 1%. Individual differences in response to xenobiotics may result from the presence of certain polymorphisms in genes that encode DNA repairing enzymes or phase I/II detoxification enzymes. The occurrence of some allelic variants can protect a subject exposed to the effects of xenobiotics, or on the contrary make him more vulnerable. The ability to repair DNA is closely correlated with genetic polymorphism in genes that encode DNA repair enzymes. Genetic polymorphisms could therefore be associated with different levels of risk within the general population, since the proteins encoded by the various genotypes alter the biotransformation of the substrates. The impact of genetic polymorphisms concerning the biotransformation mechanisms of OPs or other endogenous and exogenous factors is today the focus of the scientific debate, with a view to better understanding pesticide metabolism, highlighting interindividual differences in metabolizing particular pesticides. This could be the reason behind a differential toxicity to exposure than normally expected. Studies on more extensive samples, even differentiated for ethnic groups, would be useful in this sense.

2.2. Cytochrome P450s

Cytochrome P450s (CYPs) family genes encode proteins that catalyze the biotransformation of several xenobiotic. Environmental toxic chemicals and carcinogens, such as some pesticides, can be converted into bioactive compounds capable to exert their toxic or carcinogenic effects. CYPs are one of the main enzyme systems in xenobiotic metabolism, playing a key role in activation of many chemicals. On the other hand, biotransformation can lead to a complete detoxification of substances, to facilitate their excretion. Individual’s capability to metabolize toxicants can be influenced by genetic polymorphisms of CYP enzymes. The study of variant alleles in CYP enzymes may uncover susceptibility biomarker to environmental toxicity [58].

A recent study reviewed literature about correlation between exposure to organochlorines pesticides and CYP polymorphisms. The aryl hydrocarbon receptor pathway seems to be one mechanism of pathogenesis mediating induction of CYP metabolism. Variant alleles lead to altered metabolism of toxic compounds manifesting their genotoxic or carcinogenic potential [59].

In an Egyptian sample of 30 subjects exposed to OP, the association of the genetic polymorphisms of PON1 Q192R and CYP2D6 G1934A and the enzymatic activity of PON 1 and the pseudo-cholinesterase enzyme (PChE) was evaluated. The Authors found that serum PChE enzyme levels were significantly reduced in patients with chronic exposure; moreover, the CYP2D6 1934A allele was significantly increased in the same subjects. These changes were associated with symptoms such as fatigue and motor weakness, headaches, tearing, depression, paresthesia and sleep disorders [49].

Some authors determined the frequency of CYP2B6 and CYP2C19 polymorphisms in a cohort of pesticide applicators exposed to OP (chlorpyrifos), reporting a similar frequency with European and North American population. The prevalence of polymorphisms in enzymes involved in OP detoxification may facilitate the emergence of pathologies as a consequence of occupational exposure [60,61].

2.3. PON

PON family genes are located in region 21.3–22.1 (7q21.3–22.1) of the chromosome 7. The family include three proteins (PON1, PON2, PON3) exhibiting different activities. PON 1 plays a defense role against atherosclerosis due to its esterase and lipoprotein antioxidant activity. PON2 and PON 3 functions are less clear well than PON 1. They showed antioxidant and antiatherosclerotic properties like PON 1 [62].

In the liver PON1 hydrolyzes organophosphates, which have been previously activated by cytochrome P450; polymorphisms in PON1 can affect the efficacy of the response to DNA damage in individuals exposed to OP [63].

The PON1 192 Q(Gln)/R(Arg) polymorphism is responsible for a remarkable difference in the enzyme activity: the R variant has a higher paraoxonase activity, but it seems to be less efficient in protecting
against LDL oxidation compared to PON1192 Q. Hypertension, coronary artery disease, stroke and Parkinson’s disease, have been associated with R genotype, while the Q variant seems to provide better health conditions [64].

A recent study investigated whether there was a correlation between DNA methylation patterns in blood cells, prenatal exposure to pesticides and PON1 Q192R genotypic expression focusing on cardiometabolic diseases in children. The authors observed that the presence of PON1 192R allele, in exposed to pesticides was associated with a different DNA methylation profile compared to subjects with a PON1 192QQ genotype and not exposed, assuming that DNA methylation may be one of the mechanisms responsible for the risk of metabolic events in children carrying the PON1 192R allele prenatally exposed to pesticides. The main limitation of this research is the non-identification of the individual pesticides to which the subjects were exposed [65].

In a Turkish sample of 54 subjects exposed to OP (Chlorpyrifos, Trichlorfon, Parathion and Malathion) the association of the genetic polymorphism of PON1 Q192R and the enzymatic activity of PON 1 and of the enzyme acetylcholinesterase (AChE) was evaluated. The Authors found the following genotypic frequencies of the PON1 192 Q / R polymorphism: 56.5% QQ, 34.2% QR and 9.3% of RR genotypes. Subjects with the PON1 192 R (+) genotype (QR + RR genotypes) showed a higher PON1 activity than the 192 R (-) (QQ) genotype. Then the activity of the AChE enzyme was investigated, showing a significant difference only in the group exposed to OP. Thus, the Authors conclude that the enzymatic activities of PON1 and AChE are different between exposed and not exposed, increasing in relation to the genotypic profile of the exposed subject [51]. Furthermore, a study carried out on placental villous samples showed that OPs are able to modulate the gene encoding the placental butyryl-cholinesterase (BChE), this up-regulation represent an adaptative response to environmental insults [66].

Another study evaluated the correlation between exposure to organophosphorous pesticides and the presence of PON1 L55 M in the context of Parkinson’s disease (PD). High exposures have been associated with faster progression of motor, depressive and cognitive symptoms, also taking into account the distance from the site of application of the OPs. Previous study demonstrated a significant association between the L55 M polymorphism and the risk of developing PD after exposure to OP [67,68]. PON1 plays a key role in the metabolism of OPs but in detail it is not clear which is the mechanism underlying cognitive decline [69,70].

2.4. GST

Glutathione S-transferases (GSTs) are enzymes involved in phase II drug metabolism participating in detoxification of xenobiotics or endogenous compounds. GSTs have a key role in the interaction between Glutathione (GSH) and the substrate. Genetic differences in expression and activity of these enzymes are due to polymorphic alleles. These polymorphisms alter GSTs activity and consequently susceptibility for toxic compounds [71].

A case-control study investigated the role of the presence of GSTM1 and GSTT1 polymorphisms in fetal growth restriction (FGR) with regard to organochlorine pesticides exposure. Organochlorine pesticides (OCPs) and oxidative stress are reported as associated with adverse reproductive outcomes. Since the glutathione S-transferase (GST) family acts as a detoxifier of numerous toxins, including organochlorine pesticides, prenatal exposure to OCPs can alter fetal growth. The authors found that the frequency of the GSTM1- / GSTT1- (null) genotype was significantly higher in FGR cases than in controls, suggesting that exposure to pesticides in pregnant women may be a risk factor for development of “idiopathic” FGR, a risk increased by the presence of the GST polymorphism [72]. A previous study had already highlighted the risk of exposure to organochlorine pesticides in women, increasing the biomarkers of oxidative stress and the risk of preterm birth [73].

A study conducted by Gómez-Martin et al. evaluated the influence of the polymorphisms PON1, GSTM1 and GSTT1 in relation to the levels of N7-methyldeoxyguanosine (N7-MedG), used as a DNA damage marker, in 36 farmers professionally exposed to pesticides. N7-MedG levels were higher during periods of exposure to high doses of pesticides compared to the period of least exposure. Furthermore, the Authors showed that the levels of erythrocYTE acetylcholinesterase and plasma cholinesterase were significantly decreased during the period of high exposure. In conclusion, the increased exposure to pesticides increases the levels of DNA alkylation, as proof of the genotoxicity of pesticides in humans. Additionally, subjects with genotypes such as null for GSTM1 and PON1 192R allele appear to have a higher risk of genotoxic DNA damage [54].

Exposure to pesticides causes an alteration of oxidative stress biomarkers and this alteration is also affected by the genetic structure of individuals. About that, a study conducted in Brazil studied numerous polymorphisms (OGG1, XRCC1 and XRCC4, PON, GSTT1, GSTM1, GSTP1, CYP2A6, RAD51) in relation to markers of oxidative stress, remarking that agricultural operators exposed to pesticides showed an altered antioxidant enzymes activity, which in the long term can promote the onset of chronic diseases [74]. Same Authors in a more recent study highlighted the central role played by OGG1 and XRCC1 in the repair mechanisms in a sample of 121 exposed farmers compared to unexposed. The exposure concerned both pesticide mixtures and nicotine, since they are workers on tobacco plantations. Polymorphisms of PON1 and SOD2 can alter the process of metabolizing xenobiotics. Such a genetic background, in professionally exposed subjects, increases cellular damage and alters DNA repair capacity [75]. A further study conducted in Brazil underlined the importance of using PPE. In 120 pesticide-exposed blood samples of farmers Authors first, assessed the presence of GST polymorphisms comparing to 115 control samples, in relation to poisoning events after exposure. Subjects with the GSTM1 genotype showed more intoxication events after exposure to pesticides, although only 45% of them used PPE. However, this association was not statistically significant. The Authors therefore conclude by stating that although no statistical evidence was found between the presence of polymorphisms and the poisoning events caused by pesticides, an important finding emerged regarding the correct use of PPE in the prevention of such events [53].

Other studies have investigated the correlations between exposure to pesticides, presence of mutated GST alleles and specific pathologies: Siddarth et al. studied how polymorphisms of xenobiotic metabolizing enzymes increase pesticide accumulation aggravating renal dysfunction, demonstrated by a significant decrease in the estimated glomerular filtration rate [76]; a case-control study conducted by Sharma et al. suggested a possible correlation with higher risk to develop urinary bladder cancer [77]; Pinhel et al. suggested that GSTT1/GSTM1 polymorphisms could promote develop of Parkinson’s disease concurrently to pesticides exposure [78]; Fortes et al. describe GSTM1 null genotypes a risk modifier for developing cutaneous melanoma [79]; moreover an Indian study showed that GSTT1 null genotype may represent a protection factor against coronary artery disease while, on the other hand, the GSTM1 null genotype can play a key role in the development of the disease as it results in a total absence of the activity of this detoxification mechanism [80]. All these studies reach similar conclusions, or that the combination of these two elements, genetic polymorphisms and pesticides exposure, significantly modifies the risk of development of many diseases.

2.5. DNA repair gene polymorphisms

The x-ray repair cross complementing group 1 (XRCC1) protein and the 8-oxoguanine glycosylase 1 (OGG1) genes are involved in Base Excision Repair (BER) pathway, playing a key role in repairing DNA damage caused by environmental factors. Mutation in XRCC1 is directly proportional with the number of chromosomal aberrations [81]. OGG1 is located on chromosome 3p25 and its inactivation is associated with a
| Author               | Polymorphic gene | Pesticides                                      | Associated diseases                                                                 | Ref. |
|---------------------|------------------|-------------------------------------------------|--------------------------------------------------------------------------------------|------|
| Kahl et al., 2018   | PON1 / SOD2 / OGG1 / XRCC1 / XRCC4 | Pesticides mixtures (Carbamates, Organophosphates, Pyrethroids and Organochlorines) | Participants who presented a chronic health condition (diabetes, metabolic, kidney or heart diseases, and cancer), were excluded |
| Kahl et al. 2018    | PON1 / OGG1 / XRCC1 | Pesticides mixtures (Not defined)               | Participants who presented a chronic health condition (diabetes, cardiovascular and neurological disease, cancer) were not included |
| Declerck et al. 2017| PON               | Not defined                                     | Cardio-metabolic disease                                                            | [65] |
| Paul et al. 2017    | PON               | Organophosphate                                 | Parkinson’s Disease and related symptoms                                            | [69] |
| Doceia et al. 2017  | CYP               | Organochlorine                                   | Cancer/Infertility/Neurological disorders                                            | [59] |
| Fortes et al. 2016  | GSTM1 / GSTT1     | Pyrethroid                                      | Melanoma                                                                            | [79] |
| Khattab et al. 2016 | PON1 / CYP2D6     | Organophosphates                                | fatigue and motor weakness/headaches/tearing / depression/paresthesia/sleep disorders | [49] |
| Adad et al. 2015    | PON1 / OGG1 / XRCC1 / XRCC4 | Organophosphates / Organic Copper / Triazines / Biological Insecticide (Abamectin) / Abamectine / Dithiocarbamates fungicides (macrocycles, xineb) | Cancer/Oncological cancer                                                           | [85] |
| Gómez-Martín et al. 2015 | PON1 / GSTM1 / GSTT1 | Organophosphates (Chlorpyrifos, Dimethoate, Malathion, Pirimiphos-methyl, Fosmet, Fenamifos / N-methylcarbamates (methomyl, oxamyl, formetanate)/ Pyrethroids (tralomethrin, cypermethrin, acrinathrin)/Neonicotinoids (imidacloprid) / Abamectine / Dithiocarbamates fungicides (macrocycles, xineb) | Data not available                                                                       | [54] |
| Sunay et al. 2015   | PON               | Organophosphates (Chlorpyrifos, Trichlorfon, Parathion and Malathion) | Cardiovascular diseases/Diabetes/Cancer                                               | [51] |
| Godoy et al. 2014   | GSTM1 / GSTT1     | Glyphosate / 2,4-Dichlorophenoxyacetic acid / Parathion / Abamectin / Thiophanate methyl / Cyproconazole | Headache/nausea/dizziness/fainting/tremors/excessive salivation/convulsions/irritation/pain | [53] |
| Siddarth et al. 2014| GT                | Organochlorine                                   | Chronic Kidney Disease                                                              | [76] |
| Lee et al. 2013     | PON1             | Organophosphate                                 | Parkinson’s Disease                                                                 | [68] |
| Sharma et al. 2013  | GSTM1 / GSTT1     | Organochlorine                                   | Urinary Bladder Cancer                                                              | [77] |
| Pinhel et al. 2013  | GSTM1 / GSTT1     | Not defined                                      | Parkinson’s Disease                                                                 | [78] |
| Sharma et al. 2012  | GSTM1 / GSTT1     | Organochlorine                                   | Fetal growth restriction                                                             | [72] |
| Da Silva et al. 2012| OGG1 / XRCC1 / XRCC4 / PON / GSTT1 / GSTM1 / GSTP1 / CYP2A6 / RADS1 | Pesticides mixtures (Not defined) | Data not available                                                                   | [74] |
| Ellison et al. 2012 | CYP2B6 / CYP2C19  | Organophosphate                                 | Neurobehavioral deficits                                                             | [60] |
| Rohr et al. 2011    | PON1 / XRCC1 / OGG1 | Bipyridyl/Organophosphates/Copper Sulphate/Carbamates/Others | Coronary artery disease/Stroke/ Hypercholesterolemia/Parkinson’s disease/ Hypertension | [64] |
| Manthripragada et al. 2010 | PON1 | Organophosphate                                 | Parkinson’s Disease                                                                 | [67] |
| Rohr et al. 2006    | PON1 / XRCC1 / OGG1 | Not defined                                     | Parkinson disease/Reproductive disorders                                             | [84] |
reduced production of cytokines and chemokines leading to acute inflammation, conferring resistance to organ damage [82]. XRCC1 gene is located on chromosome 19q13.2, and interacting with OGG1 can increase its glycosylase activity [83]. Polymorphisms in these genes can significantly alter DNA damage fixation system.

Other authors evaluated the effect of the haplotypes of the PON1, XRCC1 and hOGG1 genes on DNA damage of lymphocytes collected from farmers. The results demonstrate effective DNA repair for people with the Gln/Gln (PON1) + Arg/Trp (XRCC1) phenotype. Furthermore, the evaluation of the PON1 and hOGG1 genes showed an effective repair for the Gln/Arg + Cys/Cys or Ser/Cys phenotypes, respectively. Both combinations of the polymorphic PON1 gene involved in the metabolism of xenobiotics and related genes encoding enzymes involved in BER-like repair can modulate DNA damage resulting from exposure to pesticides. These results indicate that metabolizing capacity should be considered along with DNA damage repairing pathways when evaluating the individual susceptibility after exposure to xenobiotics [84].

Another study conducted by Rohr et al. on lymphocytes from pesticide-exposed vineyard workers, evaluated both the separate and combined effects of polymorphisms in the PON1 (Gln192Arg), XRCC1 (Arg194Trp) and hOGG1 (Ser326Cys) genes on induction of lymphocyte damage. Authors showed significantly higher DNA damage in the exposed group when a less efficient OGG1Cys allele was present independently of the PON1 genotype. Analysis of the compared effect of XRCC1 and PON1 genotypes in the exposed group suggested that, among the poorly metabolizing PON1Gln/Gln individuals, the XRCC1 Arg/Trp genotype has a protective effect [85].

3. Conclusion

Genetic polymorphisms can alter the effects of individual exposure to xenobiotics such as pesticides. Further research on the variability of many DNA repair genes and their combinations is of priority importance to assess their actual role as determinants of pesticide toxicity. Studies about the interaction between DNA repair genes and xenobiotic metabolism genes could also be the key to better understand the pathogenesis of pesticide-induced diseases. A good knowledge of these complex mechanisms could prospectively lead to the development of screening tests that would allow large-scale detection of susceptibility to exposure to pesticides or other xenobiotics. Improvement of preventive actions aimed to protection of most vulnerable subjects in both occupational and environmental settings may be the goal for the near future (Table 1).

Conflict of interest

The authors declare no conflict of interest.

References

[1] C. Fenga, S. Gangemi, V. Di Salvatore, L. Falzone, M. Libra, Immunological effects of occupational exposure to lead, Mol. Med. Rep. 15 (2017) 3355–3360, https://doi.org/10.3892/mmr.2017.6381.
[2] A. Garozzo, L. Falzone, V. Rapisarda, A. Marconi, D. Cinà, C. Fenga, D.A. Spandidos, M. Libra, The risk of HCV infection among health-care workers and its association with extratherapeutic manifestations, Mol. Med. Rep. 15 (2017) 3336–3339, https://doi.org/10.3892/mmr.2017.6501.
[3] A. Boino, A. Crispo, P. Cerino, L. Falzone, S. Candido, A. Giudice, G. De Petro, G. Ciliberto, M. Montella, A. Budillon, S. Costantini, Environment and bladder cancer: molecular analysis by interaction networks, Oncotarget 8 (2017), https://doi.org/10.18632/oncotarget.18222.
[4] V. Rapisarda, R. Salemé, A. Marconi, L. Falzone, M. Libra, OGG1 polymorphism and cancer risk among pesticide-exposed populations: a signiﬁcant epistatic eﬀect of XRCC1 + OGG1 genotypes in non-malignant human mesothelial cells, Oncol. Lett. 12 (2016) 3363–3367, https://doi.org/10.3892/ol.2016.5001.
[5] L. Falzone, A. Marconi, C. Loreto, S. Francesca, D.A. Spandidos, M. Libra, Occupational exposure to carcinogenic benzene, pesticides and ﬁbers (Review), Mol. Med. Rep. 14 (2016) 4467–4474, https://doi.org/10.3892/mmr.2016.4571.
[6] M. Babut, G.H. Arts, A. Barra Caracciolo, M. Teodoro, F. Nicoletti, P. Fagone, Overexpression of macrophage migration inhibitory factor (MIF) in HCC: correlation with increased levels of Freb2 and Mmp9 expression, Oncotarget 8 (2017), https://doi.org/10.18632/oncotarget.18226.
[7] S. Suratman, J.W. Edwards, K. Babina, Organophosphate pesticides exposure among farmers: pathways and risk of adverse health eﬀects, Rev. Environ. Health 30 (2015) 65–79, https://doi.org/10.1016/j.renvhe.2014.07.007.
[8] K.-H. Kim, E. Kabir, S.A. Jahan, Exposure to pesticides and the associated human health eﬀects, Sci. Total Environ. 575 (2017) 525–535, https://doi.org/10.1016/j.scitotenv.2016.09.039.
[9] E. Vakonaki, V.P. Androutsopoulos, J. Liesivuori, J.-M. Kääriäinen, J. Liesivuori, A.M. Tsatsakis, D.A. Spandidos, Pesticides and oncogenic modulation, Toxicology 307 (2013) 42–45, https://doi.org/10.1016/j.tox.2013.01.008.
[10] B.M. Qaqish, O. Al-Dalahmah, Y. Al-Motassem, A. Battah, S.S. Ismail, Occupational exposure to pesticides and the associated human health eﬀects of polymorphisms in the PON1 (Gln192Arg), XRCC1 and XRCC4 genes [85].
[11] K.Z. Guyton, D. Loomis, Y. Grosse, F. El Ghissassi, L. Benbrahim-Tallaa, N. Guha, C. Socé, H. Mattjus, L. Laguna, S. Pershagen, C. Semper, J. Gona, H. Bouguessa, Toxicity of organophosphorus pesticides induces reproductive dysfunction in the offspring, Environ. Toxicol. 28 (2013) 507–515, https://doi.org/10.3892/et.2013.1928.
[12] A. Mamane, I. Baldi, J.-F. Tessier, C. Raherison, G. Bouvier, Occupational exposure to pesticides and respiratory health, Eur. Respir. Rev. 24 (2015) 306–319, https://doi.org/10.1183/16000614.000614.
[13] Y. Yu, A. Yang, J. Zhang, S. Hu, Maternal exposure to the mixture of organophosphorous pesticides induces reproductive dysfunction in the offspring, Toxicol. 230 (2015) 379–388, https://doi.org/10.1007/s11356-014-0638-y.
[14] T. Parrin, M. Requena, A.F. Hernández, R. Alarcón, Environmental exposure to pesticides and cancer risk in multiple human organ systems, Toxicol. Lett. 230 (2014) 157–165, https://doi.org/10.1016/j.toxlet.2014.02.009.
[15] V. Rapisarda, C. Ledda, S. Matera, L. Fago, G. Arrabito, L. Falzone, A. Marconi, M. Libra, C. Costa, M. Teodoro, G. Braggioni, D. Nikitovic, A. Mamane, I. Baldi, J.-F. Tessier, C. Raherison, G. Bouvier, Overexpression of macrophage migration inhibitory factor (MIF) in HCC: correlation with increased levels of Freb2 and Mmp9 expression, Oncotarget 8 (2017), https://doi.org/10.18632/oncotarget.18226.
[16] A. Mamane, I. Baldi, J.-F. Tessier, C. Raherison, G. Bouvier, Occupational exposure to pesticides and respiratory health, Eur. Respir. Rev. 24 (2015) 306–319, https://doi.org/10.1183/16000614.000614.
[17] A. Mamane, I. Baldi, J.-F. Tessier, C. Raherison, G. Bouvier, Occupational exposure to pesticides and cancer risk in multiple human organ systems, Toxicol. Lett. 230 (2014) 157–165, https://doi.org/10.1016/j.toxlet.2014.02.009.
[18] Y. Yu, A. Yang, J. Zhang, S. Hu, Maternal exposure to the mixture of organophosphorous pesticides induces reproductive dysfunction in the offspring, Environ. Toxicol. 28 (2013) 507–515, https://doi.org/10.3892/et.2013.1928.
1 variants modify the risk of Parkinson's disease due to organophosphate exposure, Environ. Int. 56 (2013) 42–47, https://doi.org/10.1016/j.envint.2013.03.004.

[69] K.C. Paul, J.S. Sinheimer, M. Cockburn, J.M. Bronstein, Y. Bordelon, B. Ritz, Organophosphate pesticides and PON1 L55M in Parkinson’s disease progression, Environ. Int. 107 (2017) 75–81, https://doi.org/10.1016/j.envint.2017.06.018.

[70] V.P. Androutsopoulos, K. Kanavouras, A.M. Tsatsakis, Role of paraoxonase 1 (PON1) in organophosphate metabolism: implications in neurodegenerative diseases, Toxicol. Appl. Pharmacol. 256 (2011) 418–424, https://doi.org/10.1016/j.taap.2011.08.009.

[71] G. Ginsberg, S. Smolenski, D. Hattis, D.O. Johns, B. Sonawane, Genetic polymorphism in glutathione transferases (GST): population distribution of GSTM1, GSTT1, and PI conjugating activity, J. Toxicol. Environ. Health Part B 12 (2009) 389–439, https://doi.org/10.1080/1093740903158375.

[72] E. Sharma, M. Mustafa, R. Pathak, K. Guleria, R.S. Ahmed, N.B. Vaid, B.D. Banerjee, A case control study of gene environmental interaction in fetal growth restriction with special reference to organochlorine pesticides, Eur. J. Obstet. Gynecol. Reprod. Biol. 161 (2012) 163–169, https://doi.org/10.1016/j.ejogrb.2012.01.008.

[73] R. Pathak, S.G. Suke, T. Ahmed, R.S. Ahmed, A. Tripathi, K. Guleria, C. Sharma, S. Mahkani, B. Banerjee, Organochlorine pesticide residue levels and oxidative stress in preterm delivery cases, Hum. Exp. Toxicol. 29 (2010) 351–358, https://doi.org/10.1177/0748233710363334.

[74] F.R. Da Silva, J. Da Silva, Mda C. Allgayer, C.F. Simon, J.F. Dias, C.E.I. dos Santos, K.C. Paul, J.S. Sinsheimer, M. Cockburn, J.M. Bronstein, Y. Bordelon, B. Ritz, G. Ginsberg, S. Smolenski, D. Hattis, K.Z. Guyton, D.O. Johns, B. Sonawane, Genetic V.P. Androutsopoulos, K. Kanavouras, A.M. Tsatsakis, Role of paraoxonase 1 (PON1) in organophosphate metabolism: implications in neurodegenerative diseases, Toxicol. Appl. Pharmacol. 256 (2011) 418–424, https://doi.org/10.1016/j.taap.2011.08.009.

[75] M. Salvador, C. Branco, N.B. Schneider, V.F.S. Kahl, D. Simon, M.R. de Souza, V.H. da Rosa, C. Nicolau, F.R. Da Silva, K. Kvítko, A. Peres, G.P. Dorneles, C.T. de Souza, J.F. Dias, J. Da Silva, Base excision repair (OGG1 and XRCC1) and metabolism (PON1) gene polymorphisms act on modulation of DNA damage and immune parameters in tobacco farmers, Mutat. Res. Toxicol. Environ. Mutagen 836 (2018) 9–18, https://doi.org/10.1016/j.mrgentox.2018.04.001.

[76] J.G. Mabley, P. Pacher, A. Deb, R. Wallace, R.H. Elder, C. Szabó, Potential role for 8-oxoguanine DNA glycosylase in regulating inflammation, FASEB J. 19 (2005) 290–292, https://doi.org/10.1096/fj.04-2278fje.

[77] S. Makhijani, B. Banerjee, Organochlorine pesticide residue levels and oxidative stress and of susceptibility, J. Hazard. Mater. 225 (2012) 81, https://doi.org/10.1016/j.jhazmat.2012.04.074.

[78] V.F.S. Kahl, F.R. da Silva, Jda S. Alves, G.F. da Silva, J. Piccinini, V.S. Dhillon, M. Fenech, M.R. de Souza, J.F. Díaz, C.T. de Souza, M. Salvador, C. Branco, F.V. Thiesen, D. Simon, J. da Silva, Role of PON1, SOD2, OGG1, XRCC1, and XRCC4 polymorphisms on modulation of DNA damage in workers occupationally exposed to pesticides, Ecotoxicol. Environ. Saf. 159 (2018) 164–171, https://doi.org/10.1016/j.ecoenv.2018.04.022.

[79] M. Siddarth, S.K. Datta, M. Mustafa, R.S. Ahmed, B.D. Banerjee, O.P. Kalra, A.K. Tripathi, Increased level of organochlorine pesticides in chronic kidney disease patients of unknown etiology: Role of GSTM1/GSTM1 polymorphism, Chemosphere 96 (2014) 174–179, https://doi.org/10.1016/j.chemosphere.2013.10.029.

[80] T. Sharma, S. Jain, A. Verma, N. Sharma, S. Gupta, V.K. Arora, B. Dev Banerjee, Gene environment interaction in urinary bladder cancer with special reference to organochlorine pesticide: a case control study, Cancer Biomark. 13 (2016) 243–251, https://doi.org/10.3233/CBM-130346.

[81] M.A. de S. Pinhel, C.L. Sado, G. dos S. Lango, M.L. Gregorio, G.S. Amorim, G.M. da S. Florim, C.M. Maure, D.P. Martins, F. de N. Oliveira, M.A. Nakazone, W.A. Tognola, D.R.S. Souza, Nullity of GSTT1/GSTM1 related to pesticides is associated with Parkinson’s disease, Arq. Neuropsiquiatr. 71 (2013) 527–532, https://doi.org/10.1590/0004-282X20130076.

[82] C. Fortes, S. Mastroeni, G. Bottà, P. Roffetta, G. Antonelli, F. Venanzetti, Glutathione S-transferase M1 null genotype, household pesticides exposure and cutaneous melanoma, Melanoma Res. 26 (2016) 625–630, https://doi.org/10.1097/CMB.0000000000000295.

[83] M.A. Bhat, G. Gandhi, Association of GSTT1 and GSTM1 gene polymorphisms with coronary artery disease in North Indian Punjabi population: a case-control study, Postgrad. Med. J. 92 (2016) 701–706, https://doi.org/10.1136/postgradmedj-2015-133836.

[84] V.F.S. Kahl, D. Simon, M.R. de Souza, V.H. da Rosa, C. Nicolau, F.R. Da Silva, K. Kvítko, A. Peres, G.P. Dorneles, C.T. de Souza, J.F. Dias, J. Da Silva, Base excision repair (OGG1 and XRCC1) and metabolism (PON1) gene polymorphisms act on modulation of DNA damage and immune parameters in tobacco farmers, Mutat. Res. Toxicol. Environ. Mutagen 836 (2018) 9–18, https://doi.org/10.1016/j.mrgentox.2018.04.001.

[85] M.A. Bhat, G. Gandhi, Association of GSTT1 and GSTM1 gene polymorphisms with coronary artery disease in North Indian Punjabi population: a case-control study, Postgrad. Med. J. 92 (2016) 701–706, https://doi.org/10.1136/postgradmedj-2015-133836.

[86] S. Makhijani, B. Banerjee, Organochlorine pesticide residue levels and oxidative stress and of susceptibility, J. Hazard. Mater. 225 (2012) 81, https://doi.org/10.1016/j.jhazmat.2012.04.074.

[87] V.F.S. Kahl, F.R. da Silva, Jda S. Alves, G.F. da Silva, J. Piccinini, V.S. Dhillon, M. Fenech, M.R. de Souza, J.F. Díaz, C.T. de Souza, M. Salvador, C. Branco, F.V. Thiesen, D. Simon, J. da Silva, Role of PON1, SOD2, OGG1, XRCC1, and XRCC4 polymorphisms on modulation of DNA damage in workers occupationally exposed to pesticides, Ecotoxicol. Environ. Saf. 159 (2018) 164–171, https://doi.org/10.1016/j.ecoenv.2018.04.022.

[88] M. Siddarth, S.K. Datta, M. Mustafa, R.S. Ahmed, B.D. Banerjee, O.P. Kalra, A.K. Tripathi, Increased level of organochlorine pesticides in chronic kidney disease patients of unknown etiology: Role of GSTM1/GSTM1 polymorphism, Chemosphere 96 (2014) 174–179, https://doi.org/10.1016/j.chemosphere.2013.10.029.

[89] T. Sharma, S. Jain, A. Verma, N. Sharma, S. Gupta, V.K. Arora, B. Dev Banerjee, Genetic environment interaction in urinary bladder cancer with special reference to organochlorine pesticide: a case control study, Cancer Biomark. 13 (2016) 243–251, https://doi.org/10.3233/CBM-130346.