GmSAL1 Hydrolyzes Inositol-1,4,5-Trisphosphate and Regulates Stomatal Closure in Detached Leaves and Ion Compartmentalization in Plant Cells

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Abstract

Inositol polyphosphatases are important regulators since they control the catabolism of phosphoinositol derivatives, which are often signaling molecules for cellular processes. Here we report on the characterization of one of their members in soybean, GmSAL1. In contrast to the substrate specificity of its Arabidopsis homologues (AtSAL1 and AtSAL2), GmSAL1 only hydrolyzes inositol-1,4,5-trisphosphate (IP₃) but not inositol-1,3,4-trisphosphate or inositol-1,4-bisphosphate. The ectopic expression of GmSAL1 in transgenic Arabidopsis thaliana led to a reduction in IP₃ signals, which was inferred from the reduction in the cytoplasmic signals of the in vivo biomarker pleckstrin homology domain–green florescent protein fusion protein and the suppression of abscisic acid-induced stomatal closure. At the cellular level, the ectopic expression of GmSAL1 in transgenic BY-2 cells enhanced vacuolar Na⁺ compartmentalization and therefore could partially alleviate salinity stress.

Introduction

Phosphoinositol derivatives play a key role in mediating cellular signals which are often related to abscisic acid (ABA) and calcium signaling pathways [1–3]. Inositol polyphosphatases are therefore potential regulators of cellular processes [1–4]. Early characterizations of inositol polyphosphatases were mainly conducted in animal systems and have successfully identified inositol polyphosphatases with different substrate specificities [4,5]. Besides phytases which act on inositol hexakishosphate (IP₆; phytate) [6,7], there are two major classes of inositol polyphosphatases identified in plants: inositol-5-phosphatases and inositol 1-phosphatases [8–10].

In Arabidopsis thaliana, a total of 15 genes were predicted to encode for inositol 5-phosphatases [11], based on the presence of two consensus domains (Domain I and Domain II) identified by aligning characterized inositol 5-phosphatases from animals, yeast and plants [4]. Despite sequence homology at the two consensus domains, the proteins encoded by these 15 Arabidopsis genes show little overall sequence similarity, suggesting a diverse group of inositol 5-phosphatases present in plants [11]. Plant inositol 5-phosphatases also exhibit different substrate specificities. For example, At5PTase1 and At5PTase2 can hydrolyze inositol-1,4,5-trisphosphate (IP₃) and inositol-1,3,4,5-tetrakisphosphate ([1,3–5]P₄) [10,11] while At5PTase3, At5PTase7, and At5PTase11 all act on phosphatidylinositol-4,5-bisphosphate (PI(4,5)P₂) and phosphatidylinositol-3,5-bisphosphate (PI(3,5)P₂) [12,13]. At the same time, At5PTase11 can also use phosphatidylinositol-3,4,5-trisphosphate (PI(3–5)P₃) as a substrate [12].

Gain-of-function experiments showed that the overexpression of At5PTase1 resulted in a reduction of the stomatal response toward light and ABA treatment, presumably due to a lowered IP₃ level [8]. The overexpression of At5PTase2 also showed decreased sensitivity toward ABA inhibitory effects on seed germination [10]. These two pieces of evidence point to the regulatory role of inositol 5-phosphatases...
in ABA signaling. On the other hand, the overexpression of  
150 m

DNA sequencing and sequence analysis

DNA sequencing was performed using the ABI PRISM

gene in  

Northern blot analysis and real time RT-PCR

Northern blot analysis was performed as previously described [21]. Antisense single-stranded digoxigenin- (Roche, Indianapolis, IN, USA)-labeled DNA probes were obtained by PCR according to the manufacturer's manual. The GmSAL1 or GFP cdNA subcloned into the pBluescript II KS (+) vector was used as the template and the primers used in the PCR were  

Gene expression under stress

Soybean seeds were germinated in vermiculite with water in a greenhouse. After one week, seedlings were transferred to hydroponic cultures with half-strength Hoagland's solution [20]. Just after the emergence of the first trifoliate, the plants were transferred to half-strength Hoagland's solution supplemented with 60mM, 125mM, and 185mM NaCl, and 10%, 14%, and 16.5% (w/v) polyethylene glycol (PEG)-6000 for 24h. Treated sample tissues were harvested and frozen in liquid nitrogen for total RNA extraction. A total of 20μg RNA for each sample was used for northern blot analysis. The osmolarity of NaCl and PEG solutions was measured by Advanced  

Materials and Methods

Cloning of GmSAL1

Soybean (Glycine max L. Merr.) plants were grown in a greenhouse. For experiments leading to the cloning of GmSAL1, the seeds were first germinated in sand irrigated with water. After the opening of the first trifoliate, the seedlings were irrigated with modified Hoagland's solution [17]. NaCl treatment was performed using 150 mM NaCl for 3 d. Total RNA samples were obtained using a modified phenol extraction protocol [18]. The first-strand cdNA was then obtained from the total RNA by reverse transcription using the Moloney murine leukemia virus-reverse transcriptase (Gibco BRL, Grand Island, NY, USA) according to the manufacturer's manual. Degenerate primers (5'TGNCANGTIGCIIGAYTAYGG3' and 5'GCRTGICCCAAITTTYTC3') (N = A/C/G/T; Y = C/T; R = A/G; I = Deoxyinosine) were designed based on the multiple alignments of the following proteins: ScHal2 from S. cerevisiae (GenBank accession number: AAR89916); AtSAL1, AtSAL2 and AHL from A. thaliana (GenBank accession number: Q42546, NP_201205 and NP_200250 respectively); and RHL from rice (GenBank accession number: Q40639). PCR using the above primer pair successfully amplified a fragment of ~600 bps, under the following conditions: 94 °C 5 min; 50 cycles of 94 °C 1 min, 54 °C 1 min and 72 °C 1 min; followed by 72 °C 5 min; in a 25 μl reaction mixture composed of 5.0 μl of the first-strand cdNA, 5.0 mM MgCl₂, 0.2 mM dNTPs, 0.8 μM of each primer, 0.5 U Taq DNA polymerase (Roche, Indianapolis, IN, USA), and 1× PCR buffer.

The DNA sequence of the full-length coding region of GmSAL1 was subsequently obtained by 5' and 3' Rapid Amplification of cdNA Ends (RACE) using the SMARTRACE cdNA amplification kit (Clontech Laboratories, K1612, Mountain View, CA, USA), according to the manufacturer's protocol. Gene-specific primers (GSPs)/nested GSPs for 5' and 3' RACE were 5'ACCACCTTCAGATTACCACCGTC3'/ 5'TGCTTTGACACCGAGTTTTTCTGC3' and 5'GTTGTATTTGGGGTTCTTGGTG3'/ 5'TGTCAAAGCACCCACGTCAGAA3'respectively. The GmSAL1 cdNA clone covering the entire coding region was amplified from the first-strand cdNA samples using the primers 5'CGCGCTGACACTAATCGTTT3' and 5'CGAGCCGACAACAAAGTTAGG3'. The DNA sequence information of GmSAL1 was deposited into GenBank (accession number: EF637045).

The in vivo functions of AtSAL1 on stress responses are still controversial. The ectopic expression of the Arabidopsis gene AtSAL1 in yeast conferred lithium tolerance, similar to the effects of overexpressing the endogenous SchHal2 gene in yeast [9]. It was an expected result since AtSAL1 and ScHal2 are homologues. However, it was subsequently reported that the overexpression of AtSAL1 in A. thaliana did not elevate NaCl tolerance [15]. In fact, AtSAL1 is a negative regulator of drought tolerance in A. thaliana, since a mutation in the AtSAL1 gene led to enhanced drought tolerance [16].

In this work, we identified the coding sequence of the SAL1 homologue in soybean, GmSAL1. We also characterized its substrate specificity and demonstrated its effects on various stress responses in plant cell through its function as an inositol polyphosphatase.
Real-time PCR was performed according to a previous report [22] using the CFX96 Touch Real-Time PCR Detection System (Bio-Rad, Hercules, CA, USA). The following primers were used: 5'-ATTGGGGTCTTGCTGGTGC-3' (forward primer for GmSAL1), 5'-TTGTGAAACCACTCCAGGC-3' (reverse primer for GmSAL1), 5'-GGCCTTGATTATCCTCGATATAG-3' (forward primer for AtUBQ10, house-keeping gene for A. thaliana samples) [23], 5'-AAAGAGAATACGAAACGGAACATG-3' (reverse primer for AtUBQ10) [23], 5'-CCCCCTCACCAAGATGTC-3' (forward primer for L25, house-keeping gene for N. tabacum samples) [24], 5'-AAGGGTGTGTTGCTCTCAATCTT-3' (reverse primer for L25) [24].

Construction of GmSAL1 transgenic A. thaliana lines

Transgenic A. thaliana ectopically expressing GmSAL1 was constructed according to a previous report [22]. The cDNA of GmSAL1 was cloned into a binary vector [25] downstream of the cauliflower mosaic virus 35S promoter. Six-week-old A. thaliana (Col-0) plants were transformed by the vacuum infiltration method [26] using the Agrobacterium tumefaciens strain GV3101 (pmp90) transformed with the GmSAL1 construct. The expression of GmSAL1 in the transformed A. thaliana was verified by real-time PCR (Figure S1 in File S1).

Construction of A. thaliana lines expressing both PH_{PLCd}\text{-GFP} and GmSAL1

Reciprocal crosses were performed between the PH_{PLCd}\text{-GFP} and the GmSAL1 transgenic lines. Five- to six-week-old plants grown on soil were used. Mature flowers were detached from the pollen donor parent. Sepals, petals, and stamens of the flower buds of the pollen recipient parent were removed with a pair of forceps. Pollens of the donor parents were transferred to the stigma of the recipient flower bud. About 2-3 weeks after artificial crossing, seeds were harvested. After a few generations of self-fertilization, double homozygous lines were screened by PCR. For the line PH_{PLCd}\text{-GFP}/GmSAL1-1, GmSAL1 was the pollen donor. For the line PH_{PLCd}\text{-GFP}/GmSAL1-2, PH_{PLCd}\text{-GFP} was the pollen donor.

Expression and purification of the GmSAL1 protein in E. coli

The coding sequence of GmSAL1 was amplified by PCR using Pfx polymerase (Invitrogen, Carlsbad, CA, USA) with the following primers: 5'CCCCACTTCTGCTTTAGGAAAGGAATTC3' and 5'CCCCCAATTTGCCAAAGAGATGACCTTTC3'. The amplified GmSAL1 cDNA was subcloned into pGEX-2T vector (GE Healthcare, Chalfont St Giles, UK) to form a fusion protein with the glutathione S-transferase (GST). The GST-GmSAL1 construct was then introduced into the E. coli strain BL21 (DE3) cells. The expression of the recombinant protein was induced by the addition of 0.1mM IPTG to the E. coli culture followed by incubation for 4h, before the cells were washed and resuspended in the lysis buffer (50mM Tris-HCl pH 7.5, 100mM NaCl and 1mM phenylmethylsulfonyl fluoride). The soluble protein fraction was obtained by sonication and subsequent centrifugation. Soluble GmSAL1 protein was affinity-purified by GST-Trap column (Amersham Biosciences, Piscataway, NJ, USA) and then dialyzed overnight in the enzyme assay buffer (25mM Tris-HCl pH 7.5, 1mM MgCl2) with 2mM dithiothreitol.

Enzyme assays and determination of the $K_m$ values

Phosphatase assays were performed according to previous reports [9,27,28] with slight modifications. A 100μl reaction mixture containing the recombinant protein and substrate in the assay buffer (25mM Tris-HCl pH7.5 and 1mM MgCl2) was incubated at 37°C for 30min and the released inorganic phosphate was quantified at 650nm using a 96-well microtiter plate reader (Tecan Group Ltd., Seestrasse, Männedorf, Switzerland). Protein concentrations were determined by the Bradford method [29]. The $K_m$ for IP3 hydrolysis was determined by measuring the rate of hydrolysis at the following substrate concentrations: 0.0125mM, 0.025mM, 0.05mM, 0.1mM, and 0.2mM. The $K_m$ of IP3 was calculated using the $K_m$ protein concentration, and molecular weight of the recombinant protein. All substrates used were from Sigma-Aldrich Co. (St Louis, MO, USA) except $I(1,3,4)P_2$ and $I(1,4)P_2$ (Echelon Biosciences Inc., Salt Lake City, UT, USA).

Relative in vivo IP3 levels in guard cells

The microscopic analysis of relative in vivo IP3 level in guard cells was according to a previous report [30]. The lower epidermal of rosette leaves of 4-week-old A. thaliana grown on soil at 22°C (16h light-8h dark cycle) was peeled off. The epidermal peels were immersed in buffer containing 50μM CaCl2, 5mM KCl, 10mM MES-Tris, (pH 6.15) for 2 h under constant light, before subjected to confocal microscopic analysis. Images were collected using Olympus FV1000(Ex: 488nm; Em: 510–525nm). The fluorescence signals were analyzed using the ImageJ program (ver. 1.37i.44p)[31].

Stomatal aperture assay

The stomatal aperture assay was performed according to a previous report [8]. Leaves of 4-week-old A. thaliana grown on soil at 22°C (16h light-8h dark cycle) were detached and incubated in a perfusion solution (50mMKCl, 10mM MES-Tris, pH 7.0) without supplements for 2h, followed by incubation in a perfusion solution with supplements for another 2h. In the control experiment, the perfusion solution was supplemented with 0.1% (v/v) MeOH (solvent of ABA). In the other two sets of experiments, perfusion solutions containing 100μM ABA with or without 5mM CaCl2 were used. The concentration of ABA employed was according to a previous report [8]. All incubations were conducted at 22°C under constant light. The differential interference contrast (DIC) images of guard cells were captured using a light microscope (Nikon Eclipse 80i). The stomatal aperture was measured using a digital ruler available in the software SPOT Advance (ver. 4.6, Diagnostic Instruments, Inc.).

Seed germination assay

The seed germination assay was performed according to a previous report [8]. A. thaliana seeds were surface-sterilized, placed on half-strength MS agar plate (1% (w/v) sucrose
supplemented with one of the following: 0.1% (v/v) MeOH alone, 2.5μM ABA in 0.1% (v/v) MeOH, or 4μM ABA in 0.1% (v/v) MeOH) and kept at 4°C in the dark for 2 d. Seeds were then allowed to germinate at 25°C under continuous light. The germination rate was calculated using 138 to 211 seeds from three independent experiments.

Establishment of transgenic tobacco BY-2 cell lines

The same recombinant construct used to transform A. thaliana was transformed into the tobacco BY-2 cells [32] using Agrobacterium (strain LB4404) by a co-cultivation method [33]. After selecting the transformants on antibiotic-containing medium, PCR screening using gene-specific primers was performed to verify the successful integration of the transgene into the genome and real-time PCR was performed to verify the expression of the transgene in the transformed cell lines. Cells were grown in a liquid MS medium [32] at room temperature in the dark with mild agitation.

Microscopic analysis of Na⁺ compartmentalization

For Na⁺ compartmentalization studies, BY-2 cells were harvested 4 d after subculture and used for all microscopic analyses. After the cells had been treated with 150mM NaCl in MS medium, they were incubated with shaking at room temperature for 1h. SodiumGreen™ indicator (S6901; Invitrogen, Carlsbad, CA, USA) was used to visualize the intracellular contents of Na⁺ [17], and the confocal images were captured using Olympus FV1000 (Exc: 488nm; Em: 510–525nm). The fluorescence signals were analyzed using the ImageJ program (ver. 1.44p) [31]. The total fluorescence intensity in the pixels was divided by the total area to obtain the average pixel fluorescence intensity. Background fluorescence intensity was measured in the same field and was subtracted. DIC images of cells were obtained by excitation with a red diode. Two replicates of each experiment were performed.

For real-time image capturing, 3-day-old cells were pre-incubated with 10μM SodiumGreen™ indicator for 30 min prior to the 200mMNaCl treatment. Images of cells were captured at 10-sec intervals after NaCl treatment for a total period of 50min, using the Bio-Rad Radiance 2100 system (Exc: 514nm, filter set HQ545/40). The first reading was taken ~20s after NaCl treatment. Cell sizes were measured by the ImageJ program (ver. 1.37) and the % change was reported. Na⁺ content was measured as the intensity of the Sodium Green™ signal per unit area using the same program. The images were collated and converted to an MPEG video and attached as a supplementary file.

Microscopic analysis of cell viability

For cell viability assays, 4-day-old BY-2 cell suspension cultures were remained untreated or treated with one of the following: 150mM NaCl in MS medium, 150mMNaCl with 1μM IP₃ in MS medium, or 13.3% (w/v) PEG-6000 in MS medium (near-isotonic to 150mM NaCl), for 24 h with shaking in an orbital shaker. After treatment, cells were stained with 0.4μg/ml Trypan blue (Sigma Aldrich Co., St Louis, MO, USA). The images of stained cells (around 150 cells in each experiment) were captured using the CCD camera attached to the light microscope (Nikon Eclipse 80i). A total of 91-247 cells were counted from 4-12 fields. Two replicates of each experiment were performed.

NaCl and PEG stress treatments on A. thaliana

Stress treatments on A. thaliana were performed as described in previous reports [34,35] with slight modifications. Ten-day-old A. thaliana seedlings grown on MS agar at 22°C (16h light-8h dark cycle) were transferred to MS agar without supplement (CK) or MS agar supplemented with 100mM NaCl, 11.1% (w/v) PEG-6000, 150mM NaCl, or 13.5% (w/v) PEG-6000 (100mM NaCl MS broth is near-isotonic to 11.1% (w/v) PEG-6000 MS broth; 150mMNaCl MS broth is near-isotonic to 13.5% (w/v) PEG-6000 MS broth). The A. thaliana seedlings were harvested 10 days after treatment. The experiments were replicated.

Determination of total chlorophyll in A. thaliana

The determination of total chlorophyll in A. thaliana was performed as described previously [34]. Leaf tissue of less than 0.02 g was immersed in 0.8 ml N, N-dimethylformamide (DMF) followed by incubation at 4°C overnight[36]. The absorbance at 603, 647 and 664 nm was recorded. The amount of total chlorophyll was calculated using a formula published previously[37].

Statistical analysis

Statistical analysis was performed using the Statistical Package for Social Sciences (version 16.0; SPSS Inc., Chicago, IL, USA).

Results

The expression of GmSAL1 was induced by NaCl but not near-isotonic PEG

We obtained the full-length coding region of GmSAL1 (the soybean homologue of AtSAL1) by PCR using degenerate primers followed by RACE. Basic Local Alignment Search Tool (BLAST) analysis showed that the overall amino acid sequence identity of GmSAL1 (GenBank accession No.: EF637045) to the closest homologues in A. thaliana, AtSAL1 (GenBank accession No.: Q42546) and AtSAL2 (GenBank accession No.: NP_201205), is 77% and 63% respectively. Multiple alignments were performed on GmSAL1, AtSAL1 and AtSAL2 (Figure 1). The consensus sequences [38] for inositol- and phosphate-binding are all conserved in GmSAL1.

Since the yeast homologue, Sch22, is known to be a salt-stress determinant [39], we tested the expression of GmSAL1 when the plants were subjected to different concentrations of NaCl (Figure 2). It was found that the levels of GmSAL1 transcripts in both leaves and roots were induced by treating the plants with NaCl (Figure 2). Since NaCl treatment consists of two stress components: ionic stress and osmotic stress, we therefore used near-isotonic PEG treatments to control for the osmotic stress. The difference in GmSAL1 expression levels between each isotonic pair of NaCl and PEG treatments showed that GmSAL1 could only be induced by ionic stress.
Figure 1. Aligning GmSAL1 with its closest homologues in A. thaliana. Multiple alignments were performed for GmSAL1, AtSAL1, and AtSAL2 (Genbank numbers EF637045, Q42546, and NP_201205, respectively), using the ClustalW program [19] in the BioEdit package (ver. 7.0.5.3). Identical amino acid residues were shaded black and similar amino acid residues were shaded grey. The conserved motifs involved in substrate- and metal-binding and nucleophilic water activation were marked as A, B and C [38].

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Due to NaCl but not osmotic stress (due to PEG) (Figure 2). PEG treatment actually led to a repression of GmSAL1 gene expression (Figure 2).

GmSAL1 hydrolyzed IP₃

The amino acid sequence alignment suggested that GmSAL1 may possess inositol polyphosphatase activities (Figure 1). We expressed and purified the GmSAL1 protein from Escherichia coli in order to determine its substrate specificity in vitro. Surprisingly, we found that GmSAL1 used IP₃ readily as the substrate but had no effect on I(1,3,4)P₃, I(1,4)P₂, inositol 1-monophosphate (I(1)P) or IP₆. The Kₘ and K₉ values were also determined (Table 1). The GmSAL1 substrate specificity is therefore completely different from that of AtSAL1, an inositol 1-phosphatase and the closest homologue of GmSAL1 in A. thaliana.

To study the physiological significance of the enzymatic activities of GmSAL1 in vivo, we employed the biosensor construct Pleckstrin homology domain–green florescent protein (PH₃PLCd-GFP). PH₃PLCd is a protein domain which binds to PI(4,5)P₂ (on the plasma membrane) and IP₃ (in the cytosol) [30]. The PH₃PLCd-GFP construct was successfully employed to indicate the cytoplasmic IP₃ levels [30].

We first generated homozygous lines of transgenic A. thaliana expressing GmSAL1 and confirmed the expression of the transgene (Figure S1 in File S1). Two independent GmSAL1 transgenic lines were crossed to an A. thaliana line expressing the PH₃PLCd-GFP construct. Homozygous transgenic lines containing both the GmSAL1 and PH₃PLCd-GFP constructs were obtained and the expression of GmSAL1 was validated (Figure S1B in File S1).

Since the expression of PH₃PLCd-GFP was reduced in the GmSAL1/PH₃PLCd-GFP double transformants (Figure S1D in File S1), instead of comparing the total GFP signals, we examined the percentage of signals localized in the cytoplasm of guard cells.

Compared to the original PH₃PLCd-GFP transgenic line, the proportion of signals in the cytoplasm was much lower in the GmSAL1/PH₃PLCd-GFP double transformants (Figure 3).

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**Table 1. Substrate specificity (Kₘ) and K₉ values of GmSAL1.**

| Substrates | Specific activity (μmol min⁻¹mg⁻¹) | Kₘ (μM) | K₉ (min⁻¹) |
|------------|-----------------------------------|--------|-----------|
| IP₃        | 1.1 ± 0.2                         | 10.8 ± 0.1 | 68 ± 13  |
| I(1,3,4)P₃ | 0                                 | -       | -         |
| I(1,4)P₂   | 0                                 | -       | -         |
| I(1)P      | 0                                 | -       | -         |
| IP₆        | 0                                 | -       | -         |

a. Enzymatic activities were measured as described [9,27,28] in 25 mM Tris-HCl pH7.5, 1 mM MgCl₂ at 37°C for 30 min. All substrates were in a final concentration of 0.5 mM except IP₃ (0.2 mM). The results were from three independent experiments, each performed in triplicates. Details for the determination of Kₘ and K₉ values were described in Materials and Methods. Numerical data represent the mean value of independent experiments±standard deviation.

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Together with the in vitro enzymatic data, this in vivo evidence supports the function of GmSAL1 to down-regulate the level of cytosolic IP$_3$.

**Ectopic expression of GmSAL1 negated the effects of ABA on stomatal closure and seed germination**

IP$_3$ plays a key role in mediating the ABA signaling in guard cells to control the stomatal aperture [8]. To test whether the ectopic expression of GmSAL1 will also affect IP$_3$-mediated stomatal closure, a stomatal aperture assay was conducted. Detached *A. thaliana* leaves were treated in a buffer containing 0.1% (v/v) MeOH, with or without 100 μM ABA. Under ABA treatment, the stomatal apertures in the wild type leaves (WT) and the empty vector-transformed control were much reduced compared to no ABA treatment (Figure 4). On the other hand, the stomatal apertures in the leaves of the GmSAL1 lines were significantly larger than those in the controls under the same ABA treatment (Figure 4).

The model of ABA-induced stomatal closure postulates that ABA increases the cytosolic IP$_3$ level in guard cells, which in turn leads to an increase in cytosolic calcium [Ca$^{2+}$]$_{cyt}$ resulting in the differential activation and inactivation of K$^+$ channels on the plasma membrane and the tonoplast [1]. The net result is the efflux of K$^+$ (and subsequently water) out of the cytosol and the vacuole, which leads to the loss of turgidity in guard cells and, consequently, stomatal closure [1,30,40]. To investigate whether the effect of GmSAL1 on stomatal aperture is Ca$^{2+}$-dependent, 5 mM Ca$^{2+}$ was also included in the medium in addition to 100 μM ABA. External Ca$^{2+}$ leads to the elevation in [Ca$^{2+}$]$_{cyt}$ and stomatal closure [41]. Our results indicated that the suppressing effect of GmSAL1 on stomatal closure under 100 μM ABA was mitigated by the addition of external Ca$^{2+}$ (Figure 4). In the medium containing 100 μM ABA and 5 mM Ca$^{2+}$, the stomatal aperture of wild type, the empty-vector line, and the GmSAL1 transgenic lines show no significant differences (Figure 4). The effect of GmSAL1 on the stomatal opening may hence be a result of its hydrolytic activities toward cellular IP$_3$.

Besides controlling the stomatal aperture, another important function of ABA in plants is the inhibition of seed germination. Germination rate is a common strategy to study ABA sensitivity [8,10]. The effects of ABA on the seed germination rate of the wild type *A. thaliana*, empty-vector transgenic control and the GmSAL1 transgenic lines were compared. Under 2.5 μM ABA and 4 μM ABA treatments, the germination rate of GmSAL1 transgenic lines was significantly higher than the controls (Figure 5) with the effects being more pronounced under 2.5 μM ABA than 4 μM ABA treatment, indicating that GmSAL1 can reduce the sensitivity of plants toward ABA.

**Ectopic expression of GmSAL1 enhanced vacuolar Na$^+$ compartmentalization in BY-2 cells under salinity stress**

Next, we examined the effects of GmSAL1 on plant cells in general. A previous study of AtSAL1 showed that its ectopic expression in yeast cells could increase salinity tolerance [9].
Figure 4. Ectopic expression of GmSAL1 in A. thaliana negated the effects of ABA on stomatal aperture. Leaves from 4-week-old A. thaliana grown on soil were used in this experiment. A: Representative DIC images of the guard cells of untransformed wild type (WT), empty-vector transgenic control (Empty vector), and two GmSAL1 transgenic lines (GmSAL1-1 and GmSAL1-2), treated with 0 μM ABA, 100 μM ABA, or 100 μM ABA + 5 mM CaCl₂, were captured using a light microscope. Scale bar = 20 μm. B: Mean stomatal aperture was measured using a digital ruler. N>168 from repeated experiments. Error bar: standard error. ** indicates a significant difference (p<0.01) between GmSAL1-1 or GmSAL1-2 and WT, based on ANOVA followed by the posthoc Tukey's test.
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Figure 5. Ectopic expression of GmSAL1 in A. thaliana negated the effects of ABA on seed germination. The germination rates of A. thaliana seeds of untransformed wild type (WT), empty-vector transgenic control (Empty vector), and GmSAL1-expressing lines, treated with 0μM (A), 2.5μM (B), or 4μM (C) ABA on half-strength MS agar plates, were determined. Results were calculated from 138-211 seeds from three independent experiments. Error bar: standard error. The differences of germination rates among WT, empty vector and GmSAL1 overexpressing lines were subjected to one-way analysis of variance (ANOVA) followed by the posthoc Tukey’s test. Under 2.5 μM ABA treatment, p<0.01 from day 4 to day 7. Under 4 μM ABA treatment, p<0.01 from day 4 to day 8.

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Since the expression of GmSAL1 is salt-inducible (Figure 2), the effect of GmSAL1 on salt tolerance in plant cells was investigated. The survival rates of GmSAL1 transgenic BY-2 cells under NaCl stress (Figure 6) and PEG-induced osmotic stress (Figure S2 in File S1) were studied. The expression of GmSAL1 in the transgenic cells was validated (Figure S1 in File S1). Trypan blue was used to stain the dead cells. NaCl (salinity stress) significantly increased the number of dead cells. The expression of GmSAL1 could alleviate NaCl-induced salinity stress (Figure 6) but not PEG-induced osmotic stress (Figure S2 in File S1). The percentage of survival in the GmSAL1 transgenic BY-2 cell lines under NaCl stress was significantly higher than in other lines (Figure 6), whereas the supplementation of 1μM IP₃ could negate the protective effects of GmSAL1 (Figure 6).

Figure 6. Ectopic expression of GmSAL1 in BY-2 cells enhanced their survival rates under NaCl stress. Four-day-old BY-2 cells grown in MS medium were used, including cells of untransformed wild type (WT), empty-vector transgenic control (Empty vector), and three independent GmSAL1 transgenic lines (A3-1, B3-1, D2-3). The survival rate was determined by Trypan blue staining. The cells were untreated, treated with 150mM NaCl or 150mM NaCl+ 1μM IP₃ in MS medium for 24 h with shaking. They were then washed with fresh MS medium and stained with 0.4μg/μl Trypan blue for 15min before microscopic analysis. A: Typical photos showing the rate of survival. The nuclei of dead cells were stained blue. Scale bar= 100μm. B: Statistical analysis. A total of 94-247 cells were counted from 4-6 fields. Error bar: standard error. ** indicates a significant difference (p<0.01) between transgenic cells and WT, based on ANOVA followed by the posthoc Tukey’s test.
We also traced the cellular compartmentalization of Na⁺ using the fluorescence dye Sodium Green™. Our results indicated that under NaCl treatment, GmSAL1 transgenic BY-2 cells exhibited enhanced vacuolar compartmentalization of Na⁺, as reflected by the higher fluorescence intensity in the vacuole when compared to the wild type BY-2 cells and empty-vector transgenic control (Figure 7). Similar to the results of the cell survival test, the effects of expressing GmSAL1 was much reduced by the supplementation of 1μM IP₃ (Figure 7).

To better visualize the changes of BY-2 cells under NaCl stress, we captured time-series images of a single cell after NaCl treatment. Two major differences were observed when comparing the wild type BY-2 cell to the GmSAL1 transgenic cell. Firstly, when NaCl was added, the size of the protoplast in the wild type cell decreased (Figure 8; Video S1), probably due to the efflux of water from the cell. Under the same treatment, the GmSAL1 transgenic cell exhibited an initial shrinkage in protoplast size followed by a recovery after about 15 min (Figure 8; Video S1). Secondly, while there was no significant elevation of Na⁺ compartmentalization into vacuoles over time in the wild type BY-2 cell under NaCl treatment, an obvious increase in vacuolar Na⁺ was observed in the GmSAL1 transgenic cell under the same conditions. The vacuolar Na⁺ was maintained at a higher level than before the NaCl treatment when the transgenic cell gradually recovered from shrinkage (Figure 8; Video S1).

**Ectopic expression of GmSAL1 did not enhance the tolerance of A. thaliana to NaCl and PEG stress**

We also investigated the effect of ectopic expression of GmSAL1 at whole plant level. Wild type (WT), transgenic empty vector (Empty vector), GmSAL1 transgenic (GmSAL1-1 and GmSAL1-2) A. thaliana were treated with NaCl and near-isotonic PEG-6000. In contrast to the protective effects of GmSAL1 on BY-2 cells, ectopic expression of GmSAL1 in A. thaliana did not confer obvious protection under salt stress and osmotic stress (Figure S3 in File S1).

**Discussion**

It is common to classify enzymes that can act on I(1,4)P₂ and I(1,3,4)P₃ as inositol 1-phosphatase and those that can act on IP₃ as inositol 5-phosphatase [5]. While GmSAL1 showed strong sequence homology to AtSAL1 and AtSAL2 which were reported to be inositol 1-phosphatases that have no activities toward IP₃, GmSAL1 employed IP₃ as the preferred substrate and is inactive toward I(1,4)P₂ and I(1,3,4)P₃ (Table 1).

There are two possible explanations for this observation. GmSAL1 may be an inositol 1-phosphatase like AtSAL1, but differs from AtSAL1 in substrate specificity. Another possibility is that GmSAL1 possesses inositol 5-phosphatase activities that act on the 5'-phosphate of IP₃. Inositol polyphosphates without a 5'-phosphate such as I(1,4)P₂ and I(1,3,4)P₃ are therefore non-substrates. Detailed sequence analysis revealed that GmSAL1 exhibits a low degree of homology to the two consensus domains found in inositol 5-phosphatases (Figure S4 in File S1). In this research, we focus on the consequence of the IP₃ hydrolytic activities exhibited by GmSAL1.

The Kₘ value of GmSAL1 toward IP₃ was found to be about 10μM, which is similar to the Kₘ value of human inositol 5-phosphatase that also acts on IP₃ [5]. This value is at least two folds higher than the cellular IP₃ level that is needed to affect K⁺ transport [42]. Therefore, GmSAL1 activity inside the cell may not be at maximum velocity under normal conditions. The IP₃ level required to induce Ca²⁺ is at the μM level [42]. While mainly located in the cytosol, IP₃ may be able to bind to receptors such as Ca²⁺ channels on the plasma membrane [30,43]. On the other hand, stress will increase the level of cytosolic IP₃ [44–46]. For instance, NaCl treatment could increase the IP₃ level up to 15 folds in A. thaliana [2]. GmSAL1 may therefore play a role in the fine adjustment of the cytosolic IP₃ concentration under stress.

The expression of GmSAL1 in its native host was responsive to NaCl (salinity stress) but not near-isotonic PEG treatment (osmotic stress) (Figure 2). This may be tied to its physiological roles. The ability of GmSAL1 to reduce the ABA-induced stomatal response (Figure 4), by lowering the IP₃ level, is apparently not a protective mechanism against long-term osmotic stress. The expression of GmSAL1 in transgenic BY-2 cells also showed no improvement in the tolerance toward PEG treatment (Figure S2 in File S1). On the other hand, GmSAL1 can help to combat salinity stress at the cellular level by enhancing the vacuolar compartmentalization of Na⁺ (Figure 6) and such an effect was not observed when IP₃ was added. It is possible that under such experimental conditions, the protective function of GmSAL1 is brought forth by reducing the IP₃ below a threshold level.

Using stomatal closure (Figure 4) and seed germination rate (Figure 5) as parameters, we showed that GmSAL1 can lower the plant’s sensitivity toward ABA treatments, presumably due to the reduction of IP₃ signals. Such effects were also observed in inositol 5-phosphatases which use IP₃ as their substrate [8,10].

The cytosolic IP₃ offers protection against water loss in planta via inducing the closure of stomata by activating the tonoplast and cell membrane K⁺ channels that remove K⁺ from the vacuole and the cytosol and inactivating K⁺ channels that increase uptake [1]. These K⁺ channels are reported to be non-specific and can also transport Na⁺ [47,48]. Consistent with this observation, it was previously reported that the addition of NaCl could lead to stomatal opening, a phenomenon that could be reversed by ABA [49].

The classical model that IP₃ is the direct signaling molecule inducing cytosolic Ca²⁺ influx in the guard cells [50] has been challenged by some recent findings. IP₆ was found to be a much more potent signalling molecule controlling Ca²⁺ influx and the effect of IP₃ on Ca²⁺ influx might be due to its conversion to IP₆ [51]. However, GmSAL1 does not use IP₆ as the substrate (Table 1) and hence the GmSAL1 effect on stomatal opening is via regulation of the cytosolic IP₃ levels.

The level of cellular IP₃ increases under stress [44–46]. If the effect of IP₃ on vacuolar cation channels also occurs in cells other than the guard cells, a higher level of IP₃ will decrease vacuolar Na⁺ compartmentalization. In contrast, the hydrolysis of IP₃ will enhance the accumulation of vacuolar Na⁺ under NaCl treatments. It was indeed what we observed using the
Figure 7. The ectopic expression of GmSAL1 in BY-2 cells increased the vacuolar compartmentalization of Na⁺ under NaCl stress. Four-day-old BY-2 cells grown in MS medium were used, including cells of the untransformed wild type (WT), empty-vector transgenic control (Empty vector), and three independent GmSAL1 transgenic lines (A3-1, B3-1, D2-3). Vacuolar Na⁺ compartmentalization was visualized with the use of Sodium Green™. Cells pre-washed with MS medium were transferred to fresh MS medium containing no supplements (untreated), MS medium supplemented with 150mM NaCl or 150mM NaCl+ 1μM IP₃ for 1h with shaking. They were then washed with fresh MS medium and stained with 5μM Sodium Green™, followed by confocal microscopic analysis. A: Typical photos showing the relative levels of vacuolar Na⁺ using the fluorescent signal of Sodium Green™ (represented by the pseudo-green color). Scale bar= 50μm. B: Statistical analysis. The relative fluorescence intensity of 17-34 cells (from 4 fields) was determined for each data point. Error bar: standard error. ** indicates a significant difference (p<0.01) between the transgenic lines and WT, based on ANOVA followed by the posthoc Tukey’s test.

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BY-2 cell model. GmSAL1 produced in the transgenic BY-2 cells hydrolyzed IP$_3$ and hence increased Na$^+$ compartmentalization in the vacuole (Figure 7), resulting in a higher survival rate for the transgenic cells under NaCl stress than for the wild type (Figure 6).

Compartmentalization of Na$^+$ in the vacuole is an effective way to protect the plant cell against salinity stress [52–54]. On one hand, the toxic Na$^+$ is removed from the cytosol. At the same time, the accumulation of Na$^+$ in the vacuole sets up an osmotic gradient to enable the plant cell to uptake water from an environment with low osmotic potential [55,56]. This is supported by our observations that severe plasmolysis occurred in the untransformed wild type BY-2 cells upon NaCl treatment and the GmSAL1 transgenic cells could accumulate Na$^+$ in the vacuole more effectively and could therefore partially restore the protoplast size, presumably through increased water intake following ion compartmentalization in the vacuole (Figure 8; Video S1). The detailed mechanism of how GmSAL1 and cytosolic IP$_3$ level regulate Na$^+$ compartmentalization into vacuole is still unclear at this point.

Since ABA is the hormone which induces stomatal closure to protect the stressed plant from water loss [57] and GmSAL1

Figure 8. Changes in the protoplast size and vacuolar Na$^+$ content under NaCl treatment in real time. GmSAL1 transgenic BY-2 cells and the untransformed wild type cells (WT) were treated with 200mM NaCl (see Materials and Methods). Differential interference contrast (DIC) images and Sodium Green™ fluorescent signals were collected by confocal microscopy (see Materials and Methods), over a period of 50min. Each data point represents the average value of 4-5 cells. Closed square: Na$^+$ content of the GmSAL1 transgenic line; closed triangle: Na$^+$ content of WT; open square: protoplast size of the GmSAL1 transgenic line; open triangle: protoplast size of the untransformed wild type (WT). Images were collated to produce the Video S1. The differences in signal intensity and cell size between WT and GmSAL1 transgenic lines were subjected to the Student’s T-test. For Sodium Green signal comparison, p<0.01 from 20 sec after NaCl treatment till the end of the experiment (50 min and 20 sec). For cell size comparison, p<0.05 from 25 min and 20 sec after NaCl treatment till the end of the experiment.
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reduces ABA-induced stomatal response, it is not surprising that the protective effect of GmSAL1 on NaCl or PEG stress was not obvious at the whole plant level (Figure S4 in File S1).

In summary, we conclude that GmSAL1 is a novel soybean SAL1 homologue that hydrolyzes IP$_3$, and plays differential roles at the whole plant level versus at the cellular level in response to salinity stress.

Supporting Information

File S1. A combined file containing one supplemental table and four supplemental figures as follows: Table S1, Osmolarity of near-isotonic solutions; Figure S1, Validation of transgene expression; Figure S2, Ectopic expression of GmSAL1 in BY-2 cells did not enhance their survival rates under PEG stress; Figure S3, Ectopic expression of GmSAL1 in A. thaliana did not enhance their tolerance toward NaCl or PEG stress; Figure S4, Multiple alignments of GmSAL1 with inositol 5-phosphatases. (DOCX)

References

1. Assmann SM (2010) Abscisic acid signal transduction in stomatal responses. In: PJ Davis. Plant hormones. New York: Springer Verlag. pp. 399-426.

2. DeWald DB, Torabinejad J, Jones CA, Shope JC, Cangelosi AR et al. (2001) Rapid accumulation of phosphatidylinositol 4,5-bisphosphate and inositol 1,4,5-trisphosphate correlates with calcium mobilization in salt-stressed Arabidopsis. Plant Physiol 126: 759-769. doi: 10.1104/pp.126.7.759. PubMed: 11420204.

3. Kim T-H, Böhmer M, Hu H, Nishimura N, Schroeder JI (2010) Guard cell signal transduction network: advances in understanding abscisic acid, CO$_2$, and Ca$^{2+}$ signaling. Annu Rev Plant Biol 61: 561-591. doi: 10.1146/annurev-plant-042809-112226. PubMed: 20192751.

4. Majerus PW, Kisseleva MV, Norris FA (1999) The role of phosphatases in inositol signaling reactions. J Biol Chem 274: 10669–10672. doi: 10.1074/jbc.274.16.10669. PubMed: 10196133.

5. Majerus PW (1992) Inositol phosphate biochemistry. Annu Rev Biochem 61: 225-250. doi: 10.1146/annurev.bi.61.070192.001301. PubMed: 1323235.

6. Kuang R, Chan KH, Yeung E, Lim BL (2009) Molecular and biochemical characterization of APAP15, a purple acid phosphatase with phytase activity, in Arabidopsis. Plant Physiol 151: 199-209. doi: 10.1104/pp.109.143180. PubMed: 19633233.

7. Lung SC, Leung A, Kuang R, Wang Y, Leung P et al. (2006) Phytase activity in tobacco Nicotiana tabacum root exudates is exhibited by a purple acid phosphatase. Phytochemistry 69: 365-373. doi: 10.1016/j.phytochem.2007.06.036. PubMed: 17897689.

8. Burnette RN, Gunasekera BM, Gillaspy GE (2003) An Arabidopsis inositol 5-phosphatase gain-of-function alters abscisic acid signaling. Plant Physiol 132: 1011-1019. doi: 10.1104/pp.010900. PubMed: 12805629.

9. Quintero FJ, García-Debals B, Rodríguez-Navarro A (1996) The SALT gene of Arabidopsis, encoding an enzyme with 3(2'),5'-bisphosphate nucleotidase and inositol polyphosphate 1-phosphatase activities, increases salt tolerance in yeast. Plant Cell B: 529-537. doi: 10.2307/3870300. PubMed: 8721754.

10. Sanchez JP, Chua NH (2001) Arabidopsis PLC1 is required for secondary responses to abscisic acid signals. Plant Cell 13: 1143-1154. doi: 10.1105/tpc.13.5.1143. PubMed: 11340167.

11. Berdy SE, Kudla J, Gruissem W, Gillaspy GE (2001) Molecular characterization of At5PTase1, an inositol phosphatase capable of terminating inositoltrisphosphate signaling. Plant Physiol 126: 801-810. doi: 10.1104/pp.126.2.801. PubMed: 11402205.

12. Ercetin ME, Gillaspy GE (2004) Molecular characterization of an Arabidopsis gene encoding a phospholipid-specific inositol polyphosphate 5-phosphatase. Plant Physiol 135: 938-946. doi: 10.1104/pp.104.040253. PubMed: 15181205.

13. Kaye Y, Golani Y, Singer Y, Lashem Y, Cohen G et al. (2011) Inositol polyphosphate 5-phosphatase7 regulates the production of reactive oxygen species and salt tolerance in Arabidopsis. Plant Physiol 157: 229-241. doi: 10.1104/pp.111.176883. PubMed: 21677096.

14. Xiong L, Lee BH, Ishisaki M, Lee H, Zhang C et al. (2001) FIERY1 encoding an inositol polyphosphate 1-phosphatase is a negative regulator of abscisic acid and stress signaling in Arabidopsis. Genes Dev 15: 1971-1984. doi: 10.1101/gad.891901. PubMed: 11485991.

15. Chen H, Zhang B, Hicks LM, Xiong L (2011) A nucleotide metabolite controls stress-responsive gene expression and plant development. PLOS ONE 6: e26661. doi: 10.1371/journal.pone.0026661. PubMed: 22028934.

16. Wilson PB, Estavillo GM, Field JK, Pomsiriwong W, Carroll AJ et al. (2009) The nucleotidease/phosphatase SAL1 is a negative regulator of drought tolerance in Arabidopsis. Plant J 58: 299-317. doi: 10.1111/j.1365-3040.2009.04790.x. PubMed: 19170934.

17. Li WYF, Wong FL, Tsai SN, Phang TH, Shao G et al. (2006) Tonoplast-located GmCLC1 and GmNHX1 from soybean enhance NaCl tolerance in transgenic bright yellow (BY-2) cells. Plant Cell Environ 29: 1122-1137. doi: 10.1111/j.1365-3040.2005.01487.x. PubMed: 17086938.

18. Ausubel FM, Brent R, Kingston RE, Moore DD, Seidman JG et al. (1995) Phenol/SDS method for plant RNA preparation. Current Protocols in Molecular Biology. New York: John Wiley & Sons, Inc.

19. Thompson JD, Higgins DG, Gibson TJ (1994) CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. nucleic Acids Res 22: 4673-4680. doi: 10.1093/nar/22.22.4673. PubMed: 7984417.

20. Hoegland DR, Ammon DI (1980) The water-culture method for growing plants without soil. Circular California Agricultural Experiment Station 347: 1-32.

21. Sambrook J, Russell DW (2001) Molecular Cloning: A Laboratory Manual. New York: Cold Spring Harbor Laboratory Press.

22. Cheung MY, Zeng NY, Tong SW, Li WYF, Yue Y et al. (2008) Constitutive expression of a rice GTPase-activating protein induces defense responses. New Phyto 179: 530-545. doi: 10.1111/j.1469-8137.2008.02473.x. PubMed: 19086295.

23. Remans T, Smeets K, Opdenakker K, Mathijsen D, Vangronsveld J et al. (2006) Normalisation of real-time RT-PCR gene expression measurements in Arabidopsis thaliana exposed to increased metal concentrations. Planta 227: 1343-1349. doi: 10.1007/s00070-007-0606-4. PubMed: 18273637.

24. Schmidt GW, Delaney SK (2010) Stable internal reference genes for normalization of real-time RT-PCR in tobacco (Nicotiana tabacum)
during development and abiotic stress. Mol Gen Genomics 283: 237-241. doi:10.1007/s00438-010-0511-1. PubMed: 2098998.

25. Brea T, Liu C, Knight TJ, Coruzzi GM (1993) Ectopic overexpression of asparagine synthase in transgenic tobacco. Plant Physiol 103: 1285-1290. PubMed: 12232020.

26. Bechet D, Thiel G (1993) In planta Agrobacterium-mediated transformation of adult; Martinez-Zapater J, Salinas J, editors. Arabidopsis: plants by vacuum infiltration, editors. Totowa: Arabidopsis Protocols: Humana Press Inc. pp. 259-266.

27. Gil-Mascarell R, López-Coronado JM, Belles JM, Serrano R, Rodríguez PL (1999) The Arabidopsis HAL2-like gene family includes a novel sodium-sensitive phosphatase. Plant J 17: 373-383. doi:10.1046/j.1365-313X.1999.00385.x. PubMed: 10205895.

28. Murphy J, Riley J (1982) A modified single solution method for the determination of phosphate in natural waters. Anal Chim Acta 27: 31-36. doi:10.1016/S0003-2670(00)88444-5.

29. Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. Anal Biochem 72: 248-254. doi:10.1016/0003-2670(76)90527-3. PubMed: 9240501.

30. Tang RH, Han S, Zheng H, Cook CW, Choi CS et al. (2007) Coupling diurnal cytosolic Ca²⁺ oscillations to the CAS-IP₃ pathway in Arabidopsis. Science 315: 1423-1426. doi:10.1126/science.1134457. PubMed: 17347443.

31. Sukumvanich P, DesMarais V, Sarmiento CV, Wang Y, Ichetovkin I et al. (2004) Cellular localization of activated N-WASP using a conformation-sensitive antibody. Cell Motil Cytoskel 59: 141-152. doi:10.1002/cm.20030. PubMed: 15362118.

32. Nagata T, Takabe I, Matsu C (1981) Delivery of tobacco mosaic virus RNA into plant protoplasts mediated by reverse-phase evaporation vesicles (liposomes). Mol Gen Genet 184: 161-165.

33. An G (1985) High efficiency transformation of cultured tobacco cells. Plant Physiol 79: 568-570. doi:10.1104/pp.79.2.568. PubMed: 7486679.

34. Cheung MY, Li MW, Yung YL, Wen QM, Lam HM (2013) The unconventional P-loop NTPTase OsYchF1 and its regulator OsGAP1 play opposite roles in salinity stress tolerance. Plant Cell Environ 35: 53-60. doi:10.1111/j.1365-3040.2011.02426.x. PubMed: 21923759.

35. Moran R, Porath D (1980) Chlorophyll determination in intact tissues using N, N-dimethylformamide. Plant Physiol 65: 478-479. doi:10.1104/pp.65.3.478. PubMed: 16664553.

36. Atack JR, Broughton HB, Pollack SJ (1995) Structure and mechanism of inositol monophosphatase. FEBS Lett 361: 1-7. doi:10.1016/0014-5793(95)00063-F. PubMed: 7890024.

37. Li WYF, Shao G, Lam HM (2008) Ectopic expression of GmPAP3 alleviates oxidative damage caused by salinity and osmotic stresses. New Phytol 178: 89-91. doi:10.1111/j.1469-8137.2007.02356.x. PubMed: 18208471.

38. Kaur N, Gupta AK (2005) Signal transduction pathways under abiotic stresses in plants. Curr Sci 28: 1771-1780.

39. Li WYF, Shao G, Lam HM (2006) Tonoplast-located GmSAL1 Regulates IP₃ signaling in plants under salinity stress. Plant Signal Behav 5: 233-238. doi:10.1081/PSB.20037468.

40. Lee SC, Luan S (2012) ABA signal transduction at the crossroad of biotic and abiotic stress responses. Plant Cell Environ 35: 53-60. doi:10.1111/j.1365-3040.2011.02426.x. PubMed: 21923759.