Enhanced Protective Efficacy of a Chimeric Form of the Schistosomiasis Vaccine Antigen Sm-TSP-2

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Abstract

The large extracellular loop of the Schistosoma mansoni tetraspanin, Sm-TSP-2, when fused to a thioredoxin partner and formulated with Freund’s adjuvants, has been shown to be an efficacious vaccine against murine schistosomiasis. Moreover, Sm-TSP-2 is uniquely recognised by IgG1 and IgG3 from putatively resistant individuals resident in S. mansoni endemic areas in Brazil. In the present study, we expressed Sm-TSP-2 at high yield and in soluble form in E. coli without the need for a solubility enhancing fusion partner. We also expressed in E. coli a chimera called Sm-TSP-2/5B, which consisted of Sm-TSP-2 fused to the immunogenic 5B region of the hookworm aspartic protease and vaccine antigen, Na-APR-1. Sm-TSP-2 formulated with alum/CpG showed significant reductions in adult worm and liver egg burdens in two separate murine schistosomiasis challenge studies. Sm-TSP-2/5B afforded significantly greater protection than Sm-TSP-2 alone when both antigens were formulated with alum/CpG. The enhanced protection obtained with the chimeric fusion protein was associated with increased production of anti-Sm-TSP-2 antibodies and IL-4, IL-10 and IFN-γ from spleen cells of vaccinated animals. Sera from 666 individuals from Brazil who were infected with S. mansoni were screened for potentially deleterious IgE responses to Sm-TSP-2. Anti-Sm-TSP-2 IgE to this protein was not detected (also shown previously for Na-APR-1), suggesting that the chimeric antigen Sm-TSP-2/5B could be used to safely and effectively vaccinate people in areas where schistosomes and hookworms are endemic.

Introduction

Schistosomiasis ranks among the most important infectious diseases in tropical regions, resulting in a loss of between 4.5 and 92 million Disability-Adjusted Life Years (DALYs) annually and almost 300,000 deaths in sub-Saharan Africa alone [1,2,3]. High rates of post-treatment reinfection [1], the inability of periodic chemotherapy to interrupt transmission [4], the exclusive reliance on praziquantel as the only chemotherapeutic option [5,6] and the unsustainability of mass drug administration [7] has led to the development of new anti-schistosomiasis control measures, including vaccines, to complement existing initiatives [5,8,9].

Molecules lodged in the apical membrane of the schistosome tegument represent vulnerable targets for immunological attack by host antibodies due to their intimate association with the host immune system. One such family of molecules – predicted by proteomic analyses of the schistosome tegument to be accessible to host immunoglobulin [10] – is the tetraspanin integral membrane proteins. Tetraspanins contain four transmembrane domains and two extracellular loops that are predicted to interact with exogenous ligands [11,12]. Indeed, the second extracellular loop of one of these schistosome tetraspanins, Sm-TSP-2, has proven to be an effective anti-schistosomiasis vaccine, eliciting 57–64% protection in mice vaccinated with the antigen followed by challenge with S. mansoni cercariae [12]. Other schistosome tetraspanins are protective in mouse models of schistosomiasis [10], including Sm23 [13,14] and Sj-TSP-2, an S. japonicum orthologue of Sm-TSP-2 [15]. Moreover, Sm-TSP-2 was strongly recognised by IgG1 and IgG3 from putatively resistant but not from chronically infected individuals [12], further highlighting the promise of this antigen as a subunit vaccine against human schistosomiasis.

The tegument of adult and schistosomula of S. mansoni is thinner and distinctly more vacuolated compared to controls after in vitro treatment with Sm-tsp-2 double-stranded RNA (dsRNA) [16]. Moreover, injection of mice with schistosomula pre-treated with Sm-tsp-2 dsRNA resulted in the recovery of 83% fewer parasites from the mesenteries compared to controls [16], highlighting the importance of Sm-TSP-2 in proper tegument development and worm survival, and providing a potential mechanism by which the vaccine exerts its protective effect.
**Author Summary**

There are currently no vaccines available to combat helminth (worm) infections in humans. The most devastating of the diseases caused by human helminths are schistosomiasis (or bilharzia) and hookworm disease. By fusing one of the lead schistosomiasis vaccine antigens, Sm-TSP-2, with a protective fragment from one of the lead hookworm vaccine antigens, Na-APR-1, we have produced a chimeric vaccine, termed Sm-TSP-2/5B that might provide protection against two debilitating and co-endemic neglected tropical diseases. Sm-TSP-2/5B provided increased protection compared to Sm-TSP-2 alone when formulated with human approved adjuvants and tested in a mouse model of schistosomiasis. Moreover, IgE against Sm-TSP-2 or Na-APR-1 has not been detected in the blood of residents from an area in Brazil that is endemic for schistosomes and hookworms, indicating that vaccines based on these molecules would be unlikely to generate allergic reactions in recipients from developing countries.

In an earlier study, we reported the production of a chimeric form of Sm-TSP-2, consisting of Sm-TSP-2 fused to the immunodominant and neutralizing 5B region of the hookworm aspartic protease Na-APR-1, termed Sm-TSP-2/5B [17]. Hookworm infection and schistosomiasis caused by *S. mansoni* are co-endemic in much of sub-Saharan Africa and Brazil, and there is potential interest in developing a vaccine that targets both of these high prevalence and high disease burden helminths [18]. Na-APR-1/5B is a 40 amino acid fragment of the protease that contains an immunodominant alpha helix, \( \alpha_{291}Y \), which is the target epitope recognized by polyclonal and monoclonal antibodies that are capable of neutralizing the catalytic activity of Na-APR-1 [17]. Na-APR-1/5B could not be produced in soluble form, but when fused to Sm-TSP-2, it was produced in soluble form by *E. coli* and induced antibodies upon vaccination that neutralized the enzymatic activity of Na-APR-1; the chimera is currently under investigation as a hookworm vaccine. Using a mouse model of schistosomiasis, we explored the efficacy of the Sm-TSP-2/5B chimera in comparison to Sm-TSP-2 alone when both antigens are formulated with alum/CpG. Given the recent safety concerns of helminth vaccines that elicit an IgE response in individuals residing in an endemic area [19], we also assessed the recognition of Sm-TSP-2/5B by IgE from individuals chronically infected with *S. mansoni*, a crucial step in determining whether or not this antigen could be used to safely and effectively vaccinate people in areas endemic for both hookworms and schistosomes.

**Methods**

**Ethics Statement**

All work involving experimental procedures with laboratory animals was approved by the animal ethics committee of James Cook University according to the regulations of the Australian Code of Practice for the Care and Use of Animals for Scientific Purposes, 7th edition (reference EA16). All work involving human subjects research was approved by the Human Research Ethics Committees or Institute Review Boards of Instituto René Rachou-FIOCRUZ, the Brazilian National Committee for Ethics in Research (CONEP), George Washington University Medical Center, and the London School of Hygiene and Tropical Medicine. Informed written consent was obtained from all adults or the parents and guardians of all children involved in the study.

**Antigen production**

Oligonucleotide primers incorporating *Ndel* and *XhoI* restriction sites (forward primer: GGCAGATATGGAAAAACCGCAGTCAAAAAGAC; reverse primer GGGTCTAGAGTGCCGCTTGCTTACGCTGAC) and *pfu* turbo DNA polymerase (Stratagene) were used to amplify the extracellular loop 2 region (Glu107–His184) of the *S. mansoni* tetraspanin Sm-TSP-2 from the pBd/TOPO/Sm-TSP-2 plasmid [12] in our laboratory. The amplicon was then cloned into the *Ndel* and *XhoI* sites of the pET41a expression vector (Novagen), removing the GST fusion tag to allow for native N-terminal expression of the protein, but retaining the vector’s C-terminal 6×his tag to facilitate purification by Immobilised Metal Affinity Chromatography (IMAC). The ensuing plasmid was then transformed into chemically competent *E. coli* BL21-AI cells (Invitrogen). Sm-TSP-2 was expressed using the auto-induction method and media formulations established by Studier [20]. Briefly, 10 ml of minimal media supplemented with 50 μg/ml kanamycin (MDG <sub>kan</sub>) was inoculated with a single, recombiant BL21-AI colony and grown overnight at 37°C with shaking (225 rpm). The entire overnight culture was then used to seed 1.0 L of defined media supplemented with 50 μg/ml kanamycin (YM-5052-A<sub>kan</sub>) which was incubated for 24 hours at 37°C with shaking (225 rpm). Bacteria were pelleted, lysed and the resultant homogenate purified by (IMAC) as described previously [17]. Purified Sm-TSP-2 was buffer-exchanged in a dialysis bag (Pierce) with a cut-off size of 3 kDa against two changes of 50 mM sodium phosphate, pH 6.5, 10 mM NaCl (CEX buffer) (2.0 L each) at 4°C for at least 2 hours and then further purified by passing through a pre-packed 5.0 ml Hi-Trap SP-FF column (GE Healthcare) (equilibrated with 10 column volumes of CEX buffer) at a flow rate of 1.0 ml/min using an AKTA Prime UPC PPLC unit (GE Healthcare). Bound protein was purified by washing with resuspension buffers containing a rising concentration (10–500 mM) of NaCl and eluting in 5 column volumes of elution buffer (50 mM sodium phosphate, pH 6.5, 1.0 M NaCl). Sm-TSP-2 was desalted in a dialysis bag (Pierce) with a cut-off size of 3 kDa against two changes of PBS (2.0 L each) at 4°C for at least 2 hours and the final protein concentration was adjusted to 1.0 mg/ml using an Amicon Ultra-15 centrifugal concentration device (Millipore).

Sm-TSP-2/5B was produced in *E. coli* and purified as previously described [17]. The pMal-4E plasmid encoding Maltose Binding Protein (MBP) was kindly provided by Dr F. Cardoso and MBP and his tag to facilitate purification of Sm-TSP-2/5B was produced in *E. coli* and purified on amylose resin according to the manufacturer’s instructions (New England Biolabs).

**Generation of rabbit antisera**

An emulsion containing 100 μg of Sm-TSP-2 or Sm-TSP-2/5B (1.0 mg/ml) and an equal volume of Freund’s complete adjuvant was subcutaneously injected into a single New Zealand White rabbit. The same amount of antigen emulsified in an equal volume of Freund’s incomplete adjuvant was similarly administered 2 and 4 weeks later. The rabbit was bled 2 weeks later and the serum collected by centrifugation.

**Recognition of parasite-derived Sm-TSP-2 using rabbit antisera**

Freshly perfused adult *S. mansoni* were fixed in 100% methanol overnight at 4°C, embedded in Tissue-tek Optimal Cutting Temperature compound (ProSciTech) and cryostatically sectioned into 7.0 μm sections. Sections were rehydrated in PBS and blocked with PBS/0.05% Tween 20 (PBST)/1% Foetal Calf Serum (FCS) for 1 hour at RT. After washing twice (5 minutes
each) with PBST, sections were incubated with either anti-Sm-
TSP2, anti-Sm-TSP2/5B or naive rabbit sera (8.0 μl in 200 μl
PBST/1% BSA) and 5.0 μl methanolic Alexa Fluor 488-
Phalloidin (Invitrogen) for 1 hour at RT and then washed again
(3×3 minutes each). The sections were then probed with goat anti-
rabbit IgG-Cy3 [Jackson] (1:500 in PBST/1% BSA) for 1 hour at
RT. After a further 3 washes with PBST, slides were air dried
briefly and mounted with cover slips using PBS/50% Vectorshield
mounting medium with DAPI (Vector Industries) to stain nuclei.
These were examined using a Leica IM1000 DMRB inverted
fluorescence microscope.

Neutralization of hemoglobinolysis by anti-Sm-TSP-2/5B
antibodies
The inhibition of hemoglobin digestion by Na-APR-1 using
anti-Sm-TSP2/5B IgG was performed as described previously
[17]. An equal amount of anti-Sm-TSP-2 IgG was used as a
negative control.

Study population
The study was conducted in Americaninhas, a rural community
in northeast Minas Gerais state, Brazil and has been described in
detail [12]. The study design was a total population survey, with all
individuals in a 10 km² area eligible for inclusion. All participants
excluded from the study were offered a fecal exam and treated for
all helminth infections, but were not considered part of the data set
for analysis. Women who were evidently pregnant, or who tested
positive on a urine pregnancy test received treatment for all
helminth infections after the end of the pregnancy or the
termination of breast-feeding.

Parasitological survey
The parasitological survey and blood draw were performed
during April-July 2004, the results of which can be found in Table 1.
Subjects were asked to provide two fecal samples on two separate
days, which were examined qualitatively by formalin-ether
digestion. Helminth-positive samples were then examined by
Kato–Katz fecal thick smear to quantify the intensity of infection, as
eggs per gram of feces (epg). Two slides were counted from each
day’s sample, i.e. 2 – 4 slides from each individual, as some
individuals only provided one sample. Individuals who were egg-
positive by sedimentation but negative by Kato-Katz were assigned
an epg count of 3 epg, half the Kato-Katz detection limit. Hookworm was
exclusively N. americanus. Adults or children positive for gastrointestinal
nematodes were offered a single 400 mg dose of albendazole and
individuals infected with S. mansoni were treated with praziquantel. Egg-negative individuals were not treated. Treated
individuals were examined post-treatment to confirm treatment
efficacy, and offered repeat treatment(s) until egg-negative.

Determination of human IgE responses against Sm-TSP-2
Approximately 20 ml of blood was collected from 666
volunteers in siliconized tubes for separation of serum. In brief,
the level of IgE against Sm-TSP-2 was measured by indirect
ELISA using Polysoorp 96-well microtiter ELISA plates (NUNC
F96, Fisher Scientific) which were incubated overnight at 4°C with
antigen (1 μg/ml in 0.13 M PBS, pH 7.2). After washing
with PBST, the plates were blocked for 2 hours at RT with 250 μl
of PBST/3% BSA. One hundred microliters of sera (1:25 in PBST/
3% BSA) were added to the wells and incubated overnight at 4°C,
then the plates were washed with PBST and 100 μl of mouse
biotin-conjugated monoclonal anti-human IgE FC (Human
Reagent Laboratory, Baltimore, MD) (1:200 in PBST/3% BSA)
was added to the plates. Plates were incubated for 2 hours at RT
and then washed with PBST. Plates were developed by adding o-
Phenylenediamine dihydrochloride in 0.05 M phosphate-citrate
buffer (pH 5.0) plus 30% hydrogen peroxide H2O2 for 30 minutes at
RT in the dark. Fifty microliters of 2N H2SO4 was added to
stop the colorimetric reaction, which was read at a wavelength of
490 nm on a SpectraMax 340 PC (Molecular Devices) microplate
reader, SOFTmax Pro for Windows was used for the analysis and
storage of data.

Mouse immunisation and parasite challenge
Approval for the work described in this study was obtained from
the James Cook University Animal Ethics Committee. Groups of
ten female C57BL/6 mice were immunised with Sm-TSP-2, Sm-
TSP-2/5B, or the control protein MBP. Each antigen (25 μg per
dose) was formulated with an equal volume (25 μl) of a 13 mg/ml
colloidal suspension of aluminium hydroxide gel (alum) (Sigma)
and 5 μg of CpG oligonucleotide 1826 (CpG) (Invitrogen) and
injected intraperitoneally on days 0, 14 and 28. Mice were
drawn on day 42 with 120 S. mansoni cercariae by abdominal
penetration [21]. Trials were conducted twice on different dates
and with different batches of cercariae. Serum samples were
collected at day –2 (pre-immunisation), day 40 (pre-challenge)
and day 91 (necropsy) to assess antibody responses.

Necropsy and estimation of parasite burden
Mouse necropsy and worm and egg burden assessments were
performed as described previously [12]. Reductions in parasite
loads were calculated as percentages of the parasite burden in the
control group. Statistical significance was assigned a threshold of
P = 0.05 and values were determined using the student’s t test
function in Graph Pad Prism.

ELISA using pre-challenge and necropsy sera
Individual anti-Sm-TSP-2 titres (total IgG, IgG1 and IgG2a)
were determined for all trial 1 animals just prior to cercarial
challenge and at necropsy using standard ELISA techniques.
Antigen was coated on microtiter plates at 1.0 μg/ml. Sera were
serially diluted (1:1,000 to 1:16,384,000 for total IgG and IgG1
measurements and 1:1,000 to 1:256,000 for IgG2a assessment) and
100 μl was added to each well. After addition of the appropriate
horseradish peroxidase-conjugated goat antibody (Jackson), per-
odase activity was detected with tetramethyl benzidine chromo-
genic substrate and measured at 655 nm.

Cytokine ELISAs with restimulated splenocytes
Spleens were taken from all animals from trial 2, and single cell
susensions prepared by passing through a 70 μm filter (BD
Biosciences). Red blood cell lysis buffer (Sigma) was used to
remove red blood cells. Splenocyte preparations were counted,
and cultured in duplicate at 1×10⁶ cells/well in 96-well plates.
Schistosome egg antigen (SEA) and soluble adult worm antigen
preparation (SWAP) were prepared as described respectively
[18,19] and added to the cultures at 10 μg/ml and cultured at
37°C, 5% CO2 for 72 h. Levels of IL-4, IL-10, and IFN-γ in cell-
free supernatants were assessed by ELISA (OptEIA, BD
Biosciences).

Results
Antigen production
The large extracellular loop of Sm-TSP-2 (Sm-TSP-2) (molecular
weight including 6 x His tag = 10 kDa) was expressed in E. coli using
the auto-induction technique of Studier [20] instead of the more
conventional method of IPTG induction normally used to drive protein expression in T7 promoter-based, inducible systems. In addition to producing an increased biomass despite using identical seeding conditions and culture volumes, Sm-TSP-2 was produced by auto-induction and purified by IMAC to a final concentration of 100 mg/L (Fig. 1A), more than twice the yield of Sm-TSP-2 obtained by IPTG-induction (data not shown). To obtain reasonable yields of soluble chimeric Sm-TSP-2/5B (molecular weight including 6×His tag = 16.1 kDa), the protein required expression in the less reductive cytoplasmic environment of the slow-growing Rosetta-Gami strain of E. coli, in addition to being cultured at a sub-optimal growth temperature of 23°C; as a result, auto-induction of Sm-TSP-2/5B was not a feasible production method. Nevertheless, when expressed using IPTG-induction and purified by IMAC, we obtained a yield of 20 mg/L of soluble Sm-TSP-2/5B (Fig. 1B).

Anti-Sm-TSP-2/5B antibodies bind to parasite-derived Sm-TSP-2 and neutralise the activity of Na-APR-1

The localization of Sm-TSP-2 to the outer tegument of S. mansoni has previously been documented using an antibody raised to the thioredoxin fusion protein [12]. The recognition of native Sm-TSP-2 by anti-Sm-TSP-2/5B antibodies (Fig. 2A) indicated that parasite-derived Sm-TSP-2 epitopes were faithfully reproduced in the recombinant protein and were not disrupted by the addition of the 5B region of Na-APR-1 to the C-terminus of Sm-TSP-2. No reaction was observed with naive rabbit serum (Fig. 2B). Similarly, the ability of anti-Sm-TSP-2/5B IgG to bind (and inhibit) Na-APR-1 hemoglobinase activity demonstrates the preservation of 5B epitopes within the chimeric protein. No hemoglobinase inhibition of the enzyme was observed when anti-Sm-TSP-2 IgG was used in the assay (Fig. 2C).

Table 1. Cohort details of chronically infected individuals used in this study.

| Group    | N  | Age  | Necator americanus Prev.^a | Intensity of Infection (EPGb) | Schistosoma mansoni Prev.^a | Intensity of Infection (EPGb) |
|----------|----|------|----------------------------|-------------------------------|-----------------------------|-------------------------------|
|          |    | Mean | Min | Max | N (%) | Min | Max | G. Mean | 95% CI | N (%) | Min | Max | G. Mean | 95% CI |
| Females  | 309| 33.78| 1   | 96  | 248 (69)| 6   | 62,856 | 766     | (606, 969) | 170 | 55 (55)| 60 | 4074 | 93.33 | (74, 116) |
| Males    | 357| 34.30| 7   | 92  | 252 (82)| 6   | 55,554 | 692     | (559, 857) | 155 | 43 (43)| 60 | 5952 | 90.82 | (71, 114) |
| Total    | 666| 33.45| 1   | 96  | 500 (75)| 6   | 62,856 | 727     | (621, 858) | 325 | 49 (49)| 60 | 5952 | 92.12 | (78, 108) |

^a “Prev.” = prevalence of infection.
^b “EPG” = eggs per gram of feces as determined by 2 slides of Kato Katz fecal thick smear for two days.
doi:10.1371/journal.pntd.0001564.t001

Figure 1. Vaccine antigen production. (A) Sm-TSP-2 was ligated to a pET41a vector backbone in-frame with the C-terminal 6×His tag, expressed in E. coli by autoinduction and purified by IMAC followed by CEX chromatography and desalting into PBS. 1 = uninduced soluble fraction; 2 = autoinduced soluble fraction; 3 = purified protein. (B) Sm-TSP-2/5B was ligated to a pET41a vector backbone in-frame with the C-terminal 6×His tag, expressed in E. coli by IPTG induction and purified by IMAC and desalting into PBS. 1 = uninduced soluble fraction; 2 = IPTG-induced soluble fraction; 3 = purified protein.
doi:10.1371/journal.pntd.0001564.g001
Chronically infected individuals do not produce a detectable IgE response to Sm-TSP-2

Sera from 666 individuals from Minas Gerais state, Brazil – an area of high S. mansoni transmission – were assessed for the presence of an IgE response against Sm-TSP-2. No detectable levels of anti-Sm-TSP-2 IgE antibodies were observed, despite the presence of a strong IgE response to SEA in some individuals (Fig. 3).

Antibody profile following mouse immunization and parasite challenge

Mice vaccinated with alum/CpG adjuvanted Sm-TSP-2 and Sm-TSP-2/5B mounted strong Sm-TSP-2-specific IgG responses (Table 2). IgG1 responses dominated and IgG2a responses were generally weak (not shown). Pre-challenge IgG endpoint titers (four-fold serial dilutions) ranged from 256,000–1,024,000 for Sm-TSP-2 vaccinated mice and 256,000–4,096,000 for Sm-TSP-2/5B vaccinated mice. At necropsy (post-challenge), titers had waned to 64,000–256,000 for Sm-TSP-2 vaccinated mice and 64,000–1,024,000 for Sm-TSP-2/5B vaccinated mice. Mean and median anti-Sm-TSP-2 antibody titers were higher in the group vaccinated with Sm-TSP-2/5B (means 486,400 vs 1,450,667; medians 256,000 vs 1,024,000), implying that mice vaccinated with the chimera made a stronger antibody response against the Sm-TSP-2 region of the immunogen, and increased titers were not due to anti-5B antibodies. No obvious association between antibody titer and parasite burden was detected. Of the mice vaccinated with Sm-TSP-2/5B, two mice had no worms, one mouse had two worms and one mouse had four worms. All four mice had the lowest liver egg burdens and high antibody titers (≥1,024,000). However, two other mice had equally strong antibody titers but had higher parasite burdens (29 and 34 worms), precluding determination of a robust correlation between worm burdens and antibody titers.

Vaccination with Sm-TSP-2 and Sm-TSP-2/5B formulated with alum/CpG protects against parasite challenge

Sm-TSP-2/5B and Sm-TSP-2 formulated with alum/CpG protected against experimental challenge with S. mansoni. Vaccinated groups had respective decreases in worm burden of 54–58% (Sm-TSP-2/5B, \( P<0.01 \)) and 25–27% (Sm-TSP-2, \( P<0.05 \)), compared to controls over two independent trials (Fig. 4A and 4B). A comparative reduction in liver egg burdens was also observed in these groups – 48–56% (Sm-TSP-2/5B, \( P<0.01 \)) and 20–27% (Sm-TSP-2, \( P<0.05 \)), respectively (Fig. 5A).
and 5B). When the data from both trials were combined, significant decreases in worm and liver egg burdens were seen between the group vaccinated with Sm-TSP-2/5B and the group vaccinated with Sm-TSP-2 ($P < 0.01$ and $P < 0.05$, respectively). Liver egg burdens were not disproportionately reduced compared with burdens of worms, suggesting no additional effect on parasite fecundity (Table 2).

**Vaccine-induced protection against parasite challenge is associated with parasite-specific IFN-γ production**

Splenocytes from vaccinated and challenged animals were restimulated with SEA and SWAP to assess the cytokine responses to vaccination and parasite challenge. Levels of IL-4, IL-10 and IFN-γ from splenocytes were elevated in all infected animals compared to uninfected MBP-vaccinated animals when restimulated ex vivo with SEA and SWAP (Figure 6), indicating that infection-related cytokine responses were produced, although responses to SEA were generally higher. SEA and SWAP-specific IL-4 responses tended to increase in Sm-TSP-2/5B-vaccinated animals compared to control (MBP-vaccinated) infected animals, however this only reached significance with SWAP restimulation. IL-10 production in response to SWAP, but not SEA, was also increased due to Sm-TSP-2/5B vaccination. IFN-γ production in response to both SEA and SWAP were also highly significantly increased ($P < 0.01$) in response to Sm-TSP-2/5B vaccination.

**Discussion**

We have previously demonstrated that the large extracellular loop of the *S. mansoni* tegument tetraspanin, Sm-TSP-2, when linked to a thioredoxin fusion partner and formulated with Freund’s adjuvants, is an efficacious vaccine antigen, eliciting high levels of protection in a murine schistosomiasis model of infection [12]. Herein, we show that modified and chimeric forms of the Sm-TSP-2 vaccine antigen are also protective, even when formulated with a human-approved adjuvant combination, and that a schistosomiasis vaccine based on Sm-TSP-2 (or Sm-TSP-2/5B) satisfies additional selection criteria for progression into clinical trials, such as safety concerns around the potentially deleterious effects of pre-existing IgE responses in helminth endemic populations [1,19].

There is a paucity of funding - driven by the lack of a commercially viable market - available for the production of vaccines against the neglected tropical diseases, and so a vaccine antigen must be amenable to low-cost manufacture [22]. Despite attempts at optimisation of production of these two antigens being preliminary at best, both Sm-TSP-2 and Sm-TSP-2/5B have been expressed at yields that, at this initial stage, may be indicative of cost-effective up-scaling and clinical development. Indeed, Sm-TSP-2 has been recently produced in *Pichia pastoris* fermentation cultures in our laboratory at a yield of over 500 mg/L (data not shown) and efforts are currently underway to express Sm-TSP-2/5B in a similar fashion.
We recently suggested that the presence of a pre-existing human serum IgE response to a helminth vaccine antigen is a down-selection criterion [1] when considering a molecule for progression towards clinical trials because of the safety risks involved [19]. No detectable levels of Sm-TSP-2-specific IgE were found in individuals chronically infected with *S. mansoni*, despite very strong IgE responses to proteins found within SEA. This is also the case for the hookworm antigen, *N*. *APR-1* [23], the origin of the 5B domain in Sm-TSP-2/5B. Despite the absence of a detectable IgE response, previous studies have shown that humans from schistosome- and hookworm-endemic areas mount IgG1 responses to Sm-TSP-2 [12] and *N*. *APR-1* [23], indicating that both antigens are recognized by the immune system in a natural infection. What determines the isotype response (IgG vs IgE) mounted by an infected individual to a helminth antigen is multifactorial and an unresolved topic of debate [24]. What is clear, however, is the potential danger of developing a vaccine based on an antigen that is the target of a naturally acquired IgE response in the target population.

Of the two test groups, mice vaccinated with Sm-TSP-2/5B had the highest level of protection against experimental schistosomiasis. We initially hypothesized that this increased protection was due to cross-reactive epitopes within the 5B region of hookworm *N*. *APR-1* and its *S. mansoni* orthologue, *Sm-catD* [25]. However, numerous attempts to show binding of anti-Sm-TSP-2/5B to recombinant *Sm-catD* and schistosome extracts using Western blotting and immunoprecipitation coupled to tandem mass spectrometry (A. Dougall and A. Loukas, unpublished) were unsuccessful. Anti Sm-TSP-2/5B did, however, bind strongly to recombinant *Na-APR-1* and inhibited the ability of the enzyme to cleave a synthetic substrate in a previous study [17] and has likewise been shown to neutralise the hemoglobinase capacity of *N*. *APR-1* in this study; indeed, the 5B region of Sm-TSP-2/5B is highly immunogenic and was the target of a panel of

| Mouse | Total IgG titer | Total Worms | Liver EPG* |
|-------|----------------|-------------|------------|
|       | Pre-challenge  | Necropsy     |            |
| Sm-TSP-2 vaccinated group |
| 1     | 256,000        | 64,000      | 4          | 14,652     |
| 2     | 256,000        | 64,000      | 38         | 18,722     |
| 3     | 256,000        | 64,000      | 30         | 16,061     |
| 4     | 256,000        | 64,000      | 44         | 18,543     |
| 5     | 1,024,000      | 256,000     | 53         | 16,473     |
| 6     | 256,000        | 64,000      | 48         | 20,430     |
| 7     | 256,000        | 256,000     | 25         | 4,813      |
| 8     | 256,000        | 64,000      | 38         | 13,900     |
| 9     | 1,024,000      | 256,000     | 56         | 16,250     |
| 10    | 1,024,000      | 64,000      | 28         | 11,293     |
| Sm-TSP-2/5B vaccinated group |
| 1     | 256,000        | 256,000     | 32         | 11,217     |
| 2     | 1,024,000      | 256,000     | 0          | 0          |
| 3     | 256,000        | 64,000      | 38         | 17,296     |
| 4     | 256,000        | 256,000     | 51         | 16,952     |
| 5     | 1,024,000      | 256,000     | 34         | 13,523     |
| 6     | 4,096,000      | 256,000     | 29         | 9,832      |
| 7     | 4,096,000      | 1,024,000   | 4          | 3,681      |
| 9     | 1,024,000      | 256,000     | 2          | 1,700      |
| 10    | 1,024,000      | 256,000     | 0          | 0          |

* "EPG" = eggs per gram of tissue.

doi:10.1371/journal.pntd.0001564.t002

Figure 4. Worm burdens of vaccinated C57BL/6 mice. All vaccinees were necropsied on day 91, 7 weeks post-challenge. Worms were perfused from the vasculature with PBS into petri dishes and counted. Reductions and significance (*P<0.05; **P<0.01; *** P<0.001) are represented relative to the Maltose Binding Protein (MBP) control group. (A) Trial 1 total worms. (B) Trial 2 total worms.

doi:10.1371/journal.pntd.0001564.g004
IgG1 mAbs raised to recombinant Na-APR-1 [17]. Production of a chimeric antigen comprising Sm-TSP-2 and the 5B region of Sm-CatD from S. mansoni instead of Na-APR-1 is currently underway in our laboratory and may have the additional benefit of being able to induce an antibody-mediated neutralization of Sm-TSP-2 in the tegument and Sm-CatD in the gut of the intra-mammalian stages of S. mansoni. Given the absence of an obvious cross-reactive schistosome epitope for antibodies to the Na-APR-1/5B fragment, we therefore sought to confirm whether the increased protection obtained with Sm-TSP-2/5B compared to Sm-TSP-2 alone was due to the increased size and therefore increased immunogenicity of the chimera. When microtiter plates were coated with Sm-TSP-2 and probed with antisera from mice immunized with Sm-TSP-2 or Sm-TSP-2/5B, the IgG endpoint titers were higher on average for the group immunized with Sm-TSP-2/5B, implying that vaccination with the larger immunogen resulted in an increased TSP-2-specific antibody titer. We also noted that individual mice with the highest antibody titers had the fewest worms, as highlighted in Table 2. Studies are also in progress to determine whether the chimeric protein generates similar levels of protection against hookworm infection caused by Necator americanus.

Figure 5. Liver egg burdens of vaccinated C57BL/6 mice. All vaccinees were necropsied on day 91, 7 weeks post-challenge. Livers were removed, weighed, digested in 5% KOH overnight at 37 °C and the resulting eggs resuspended in 1 ml formalin. The amount of eggs in multiple 50 μl aliquots were counted and the number of eggs per gram of tissue (EPG) determined. Reductions and significance (*P<0.05; **P<0.01; ***P<0.001) are represented relative to the Maltose Binding Protein (MBP) control group. (A) Trial 1 - liver EPG. (B) Trial 2 - liver EPG. doi:10.1371/journal.pntd.0001564.g005

Figure 6. Protection against parasite challenge is associated with increased IFN-γ production. Graphs of ELISA data showing IL-4 (left axis), IL-10 (right axis) and IFN-γ (right axis) production by splenocytes taken from mice at necropsy and restimulated with either (A) schistosome soluble egg antigen (SEA) or (B) schistosome soluble adult worm antigen (SWAP). All significant differences are compared to the control MBP-vaccinated, infected group (MBP inf) by ANOVA (**P<0.01, ***P<0.001). Unless otherwise indicated, levels are not significantly different to controls. doi:10.1371/journal.pntd.0001564.g006
Restimulation of splenocytes from vaccinated and infected mice prior to necropsy showed a general increase in both Th2 (IL-4), regulatory (IL-10) and Th1 (IFN-γ) responses to parasite antigens, which was especially marked in increased IFN-γ production by mice vaccinated with Sm-TSP-2/5B compared to those vaccinated with a parasite protein MNP. This implies that every animal was effectively challenged, indicating that the recovery of very few or no parasites in some mice was not due to an unsuccessful infection but successful vaccination. These data also suggest that Th1 cytokines have a role in the protective response against schistosomiasis, a finding that has been documented in infection studies with the parasite [26,27] and vaccination experiments with recombinant vaccine candidate antigens from the tegument such as Sm29 [28] and Sm14 [29].

A caveat of using human-approved adjuvants to test vaccine antigens in the early stages of process development is that the full potential of a candidate antigen may not be realized due to the increased immunostimulatory properties of adjuvants containing mycobacteria and other toxic components, such as Freund’s adjuvants. However, the levels of protection reported herein for Sm-TSP-2/5B were similar to those reported for Freund’s formulated thioredoxin-Sm-TSP-2, and well exceed the 40% benchmark set by the WHO for progression of an antigen into clinical trials irrespective of the adjuvant used [30].

Sm-TSP-2 immunolocalizes to the surface of schistosomula [16] and adult worms [12] and has been found in the outer tegument of mature schistosomes [10] in abundance using proteomic techniques [31]. The ultrastructural morphology of adult worms and schistosomula treated in vitro with Sm-tsp-2 double-stranded RNA displayed a distinctly vaculated and thinner tegument compared to controls, suggesting that Sm-TSP-2 may play a pivotal role in tegument development in the early stages of intra-mamalian development [16]. These insights into Sm-TSP-2 function, along with the apparent importance of humoral immunity in anti-Sm-TSP-2 vaccination, lead us to hypothesize that the surface of the schistosomulum and adult fluke are potential sites of immune attack where these crucially important membranes are being opsonized by anti-Sm-TSP-2 antibodies for further attack by complement, antibody-dependent cellular mechanisms, or both. We are currently exploring the immunologic mechanisms responsible for vaccine-induced efficacy using genetically modified mice.

The Sm-TSP-2-based vaccine antigens reported in this study appear to exhibit all the early-stage characteristics of a vaccine targeting developing countries where schistosomiasis is endemic, based on their ease of production, absence of IgE reactivity, preferential recognition by resistant humans [12], essential nature of the protein for parasite survival [16] and vaccine efficacy in animal models. These features, coupled with the recent finding of a lack of polymorphism between geographical isolates of Sm-TSP-2 throughout Africa [32] provide a compelling argument for the use of Sm-TSP-2-based antigens as safe and effective anti-schistosomiasis vaccines. These additional studies also open the door to exploring more than a single helminth target with a single antigen.

Acknowledgments
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Conceived and designed the experiments: MSP DAP HJM JMB AL. Performed the experiments: MSP DAP HJM LT AMD. Analyzed the data: MSP DAP HJM JMB AL. Contributed reagents/materials/analysis tools: MSP DAP HJM JMB AMD PJH AL. Wrote the paper: MSP HJM JMB PJH AL.

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