Conserved regions of ribonucleoprotein ribonuclease MRP are involved in interactions with its substrate

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ABSTRACT

Ribonuclease (RNase) MRP is a ubiquitous and essential site-specific eukaryotic endoribonuclease involved in the metabolism of a wide range of RNA molecules. RNase MRP is a ribonucleoprotein with a large catalytic RNA moiety that is closely related to the RNA component of RNase P, and multiple proteins, most of which are shared with RNase P. Here, we report the results of an ultraviolet-cross-linking analysis of interactions between a photoreactive RNase MRP substrate and the Saccharomyces cerevisiae RNase MRP holoenzyme. The results show that the substrate interacts with phylogenetically conserved RNA elements universally found in all enzymes of the RNase P/MRP family, as well as with a phylogenetically conserved RNA region that is unique to RNase MRP, and demonstrate that four RNase MRP protein components, all shared with RNase P, interact with the substrate. Implications for the structural organization of RNase MRP and the roles of its components are discussed.

INTRODUCTION

RNase MRP is a catalytic ribonucleoprotein complex that is closely related to RNase P [a universal RNA-based enzyme responsible for the maturation of the 5'-end of tRNA and involved in some other activities (1)]. RNase MRP is an essential eukaryotic site-specific endoribonuclease (2,3) with a specificity distinct from that of RNase P. RNase MRP has been identified in practically all eukaryotes analyzed (4,5). RNase MRP is found primarily in the nucleolus (2,6–8) and—transiently—in the cytosol (9). A relatively low quantity of RNase MRP can also be found in the mitochondria, but mitochondrial RNase MRP has a distinct protein composition and specificity (10) and will not be discussed in this work.

RNase MRP is involved in the maturation of the 5.8S rRNA, cleaving the precursor molecule at a specific site (A3) within the internal transcribed spacer 1 (11–14). RNase MRP may participate in additional, earlier steps of rRNA maturation (15), but the exact nature of this activity has not been determined. RNase MRP was shown to be involved in the regulation of the cell cycle in yeast by participating in the cleavage of specific mRNAs (16–18), in the processing of U2 snRNA, as well as in the metabolism of a number of other RNAs (18–20). Defects in the activity of RNase MRP result in a variety of pleiotropic diseases in humans (21–23).

Saccharomyces cerevisiae RNase MRP [reviewed in (24–26)] contains a 340-nt-long RNA component (NME1) and 10 essential proteins, eight of which (Pop1, Pop3, Pop4, Pop5, Pop6, Pop7, Pop8 and Rpp1) are shared with RNase P (27), and two [Snm1 (28) and Rmp1 (29)] that are unique to RNase MRP. Human RNase MRP has a similar composition (5,30–33). The RNA component of RNase MRP contains a domain (Domain 1 in Figure 1) that closely resembles the catalytic (C-) domain of RNase P, sharing the major secondary structure elements and several of the conserved nucleotides that are universally found in RNase Ps throughout the three domains of life (4,33,36,39) [reviewed in (26)]; the shared elements are involved in the formation of the catalytic core in bacterial RNase P (40–42) and, by inference, in eukaryotic RNase P and RNase MRP. The structural organizations of the C-domain in eukaryotic RNase P and Domain 1 in RNase MRP seem to be similar, and the two RNA domains interact with the same set (or similar sets) of proteins that are common to RNases P and MRP, including (but possibly not limited to) proteins Pop1, Pop5, Pop6, Pop7, Pop8 and Rpp1 (30,34,35,37,43–47).

Domain 2 of the RNA component of RNase MRP (Figure 1) and its RNase P counterpart—the specificity (S-) domain—do not have apparent sequence similarities [reviewed in (26)]. However, at least two proteins that are shared by RNase P and RNase MRP (Pop1 and Pop4)

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results of a cross-linking analysis of interactions of the RNase MRP holoenzyme with its substrate, and we specify the regions of RNase MRP RNA that are interacting with the substrate, as well as the identities of the involved RNase MRP proteins.

MATERIALS AND METHODS

Saccharomyces cerevisiae strains

Saccharomyces cerevisiae strains Pop3-BTtag, Pop4-BTtag, Pop5-BTtag, Pop6-BTtag, Pop7-BTtag, Pop8-BTtag, Rpp1-BTtag and Smn1-BTtag were based on yeast strain OE1004 (MATa RMP1::TAPHIS8::TRP1 sep 1::URA3 pep4::LEU2 nuc1::LEU2 ade2-1 trp1-1 his3-11,15 can1-100 ura3-1 leu2-3,112) (49). In addition to the affinity purification tag attached to the carboxyl terminus of RNase MRP protein component Rmp1 (49), each strain contained a 7.7 kDa peptide fused to the C-terminus of one of the RNase MRP protein components (Pop3, Pop4, Pop5, Pop6, Pop7, Pop8, Rpp1 and Smn1, respectively). The peptide, a 75 amino acids segment of Propionibacterium shermanii transcarboxylase (AGKAG EGEIPAPLAGTVSKLVGEKQATKVGLAE MKMETHIA Pump4 and Pop5 as determined in (34,35)) are shown by dashed lines. The secondary structure and the nomenclature of elements are based on previously published data (36–38).

Figure 1. Secondary structure of the S. cerevisiae RNase MRP RNA (NME1). Phylogenetically conserved nucleotides (33), including the 5'GARAR-3' element (4) (where 'R' designates purines), are highlighted in black. Substrate cross-linking sites are shown by solid arrows and marked by S1, S2, S3, S4. Cross-linking sites for protein components Pop1, Pop4 and Pop5 as determined in (34,35) are shown by dashed lines. The secondary structure and the nomenclature of elements are based on previously published data (36–38).

The only Domain 2 region that demonstrates a high degree of phylogenetic conservation is a 5'-GARAR-3' (sometimes truncated to 5'-GARA-3', where R is a purine) element (4,33) (Figure 1). This element was suggested to reside on top of a short helical stem ('P8') (4,33); however, this stem was not observed in the S. cerevisiae RNase MRP holoenzyme (37).

In vitro selection experiments have identified a weak sequence consensus for single-stranded RNase MRP substrates with a U-rich region 5' to the cleavage site (positions -7 through -3) and an absolutely conserved cytosine at the position +4 (49); this pattern of RNase MRP cleavage is distinct from the pattern observed for the cleavage of single-stranded RNAs by RNase P (50).

The elements of RNase MRP RNA and protein components that are involved in interactions with RNase MRP substrates were not known. Here, we present the

Isolation of RNase MRP holoenzymes

Active RNase MRP holoenzymes were isolated from yeast strains OE1004 (49), Pop3-BTtag, Pop4-BTtag, Pop5-BTtag, Pop6-BTtag, Pop7-BTtag, Pop8-BTtag, Rpp1-BTtag and Smn1-BTtag (as aforementioned) using a tandem affinity tag fused to the carboxyl terminus of RNase MRP protein component Rmp1 as previously described (49). The affinity tag was based on the common TAP-tag (52), but the original calmodulin-binding fragment was replaced with a His6 tag to maintain the presence of magnesium during RNase MRP isolation; RNase MRP purification was performed as follows (49).

Approximately 150 g of yeast paste was washed with water, resuspended in a buffer containing 20 mM Tris-HCl, pH 7.9, 150 mM KCl, 1 mM Mg-acetate, 10% glycerol, 1 mM phenylmethanesulfonylfluoride (PMSF),

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0.1 mM ethylenediaminetetraacetic acid (EDTA) and disrupted using a BeadBeater (Biospec). Tween 20 was added to 0.1% (v/v), and the extract was clarified by centrifugation at 17 000g for 10 min (4°C) followed by 100 000g for 3 h (4°C). Three milliliters of rabbit IgG agarose (Sigma) was added to the clarified extract. After 5 h of incubation (4°C, with light agitation), the IgG agarose was washed six times with five volumes of the buffer containing 20 mM Tris–HCl, pH 7.9, 150 mM KCl, 1 mM Mg–acetate, 10% glycerol, 1 mM PMSF, 0.1 mM EDTA, 0.1% (v/v) Tween 20 (Buffer A) and resuspended in 2 ml of the same buffer. Three hundred units of tobacco etch virus (TEV) protease (Qiagen) was added to the sample. After 5 h of incubation (4°C, with light agitation), the resin was additionally washed twice with 5 ml of buffer A. The supernatant fractions were combined; the buffer was exchanged for buffer B [20 mM Na–4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES), pH 7.9, 150 mM KCl, 1 mM Mg–acetate, 10% glycerol, 1 mM PMSF, 0.1 mM EDTA and 0.1% (v/v) Tween 20] and concentrated to the final volume of 2 ml using an Amicon-Ultra 15 (100 kDa MWCO) concentrator (Millipore). Following that, 0.5 ml of Ni-NTA agarose (Qiagen) was added to the sample. After 3 h of incubation (4°C, with light agitation), the resin was washed six times with 10 ml of buffer A supplemented with 10 mM Na–imidazole (pH 7.4). Following the final wash, RNase MRP was eluted using 10 ml of a buffer containing 400 mM Na–imidazole, pH 7.4, 50 mM KCl, 1 mM Mg–acetate, 10% glycerol, 1 mM PMSF and 0.1% (v/v) Tween 20 (20 min at 4°C with light agitation). The elution buffer was exchanged for a buffer containing 20 mM Tris–HCl, pH 8.0, 150 mM KCl, 1 mM Mg–acetate, 5 mM dithiothreitol (DTT) and 0.1% (v/v) Tween 20 (buffer C), and the sample was concentrated using an Amicon-Ultra 4 (100 kDa MWCO) concentrator (Millipore). The enzyme was transferred into buffer C supplemented with 50% (v/v) glycerol and stored at −20°C.

RNase MRP isolation from yeast strain Rmp1-TAPtag (as aforementioned) was performed following a previously described TAP-based protocol (37), but the step involving affinity purification on a calmodulin resin was skipped, and the sample was concentrated and used directly after tobacco etch virus protease cleavage.

Synthesis of photoreactive RNase MRP substrate
The photoreactive (4-thiouridine-containing) RNase MRP substrate was synthesized using run-off transcription with T7 RNA polymerase and a synthetic deoxyriboboligonucleotide as the template (53). Synthesis was performed at 37°C for 5 h in a buffer containing 40 mM Tris–HCl (pH 8.1), 6 mM MgCl2, adenosine triphosphate, cytidine triphosphate, guanosine triphosphate (GTP) (1 mM each), 1 mM 4-thioUTP (TriLink Biotechnologies), 5 mM DTT, 1 mM spermidine, 50 μg/ml bovine serum albumin, 100 mM NaDNA template oligonucleotide (5’-TTTGGTTTTAAAATTTTTGGCC TATAGTGGTCTGATTA3’), 300 nM of oligonucleotide 5’-TTAATACGACCTCACT ATAGG-3’, 0.2 U/μl SUPERaseIn RNase inhibitor (Ambion) and 0.1 mg/ml T7 RNA polymerase. The resulting 4-thiouridine-containing RNase MRP substrate [5’-GGCAACAAUUUUUUA*ACACAAA-3’] (where the RNase MRP cleavage site is indicated by the asterisk, (49) and 4-thiouridines are underlined) was purified using 10% denaturing (8 M urea) polyacrylamide gels and stored at −70°C.

When a radiolabeled substrate was required, the substrate was dephosphorylated with alkaline phosphatase and 5’-end 32P-labeled with T4 polynucleotide kinase. After labeling, the substrate was gel purified again and stored at −70°C. Alternatively, internally 32P-labeled substrate 5’-GGCAACAAUUUUUUA*ACACAGAGAA-3’ (RNase MRP cleavage sites are indicated by asterisks, and 4-thiouridines are underlined) was synthesized in the presence of 32P-α-GTP. The cross-links detected using the 5’- and internally 32P-labeled substrates were identical.

Exposure of 4-thiouridine and 4-thiouridine-containing RNA to light was avoided at all steps.

Cross-linking experiments
Cross-linking of the photoreactive (4-thiouridine-containing) RNase MRP substrate to the isolated RNase MRP holoenzyme was performed in 96-well microtiter plates on ice. The RNase MRP holoenzyme (1.5 pmol) and the photoreactive substrate (1.5 or 15 pmol) were mixed in 10 μl of a buffer containing 10 mM Na–HEPES (pH 7.5), 50 mM NaCl, 10 mM MgCl2 and exposed to ultraviolet (UV) light (365 nm) generated by an 8 W handheld UV lamp kept at a 1-cm distance from the plate; the exposure time was 30 min.

Detection of cross-linking sites on RNase MRP RNA
After UV cross-linking, DTT (to 5 mM) was added to the samples, the samples were incubated with 2 mg/ml of proteinase K for 30 min at room temperature, and then RNA was extracted with phenol followed by ethanol precipitation. The locations of the cross-links were identified using reverse transcription as previously described (37). Primers RTP1AL (complementary to nucleotides 324–340), RTP1B (316–340), RTP15A (243–276), RTP25 (168–201) and RTP4 (91–126) were used. The whole length of RNase MRP RNA was analyzed, except for 3’-terminal nucleotides 314–340.

Identification of RNase MRP proteins that formed cross-links to the substrate
The photoreactive (4-thiouridine-containing) RNase MRP substrate was 32P-labeled and cross-linked to isolated RNase MRP holoenzymes as described earlier in the text. After cross-linking, 10 μl of a loading buffer [120 mM Tris–HCl (pH 6.8), 4% sodium dodecyl sulfate (SDS), 20% (v/v) glycerol, 2 M urea and 400 mM DTT] was added to the samples; the samples were incubated 5 min at 95°C and analyzed on 12% denaturing SDS–polyacrylamide gels. The radioactive bands were visualized using a PhosphoImager (Molecular Dynamics).
RESULTS

Interactions between the RNA component of the RNase MRP holoenzyme and its substrate

To identify regions of the RNA component of RNase MRP that were involved in interactions with substrates, we isolated the active RNase MRP holoenzyme from S. cerevisiae, subjected it to UV irradiation in the presence of a photoreactive RNase MRP substrate, and we located the sites of the UV-induced RNA–RNA cross-links that appeared in the presence of the photoreactive substrate (for details see ‘Materials and Methods’ section).

The design of the substrate used in cross-linking experiments was based on the results of in vitro selection of RNase MRP substrates (49), which demonstrated that a typical RNase MRP substrate has a CUC/CAC/CGC/UUC/AUC triad in the positions +2 to +4 relative to the cleavage site (with the cytosine at +4 being absolutely required for cleavage), no guanines in the positions +1 and −1 and (preferably) a U-rich stretch located 5′ to the cleavage site. The sequence of the substrate was 5′-G GCC AACAAUUUUUUA*ACACAAA-3′; the RNase MRP cleavage site is indicated by the asterisk.

UV irradiation of the RNase MRP holoenzyme in the presence of the unmodified substrate did not result in the formation of detectable enzyme–substrate cross-links (data not shown). To increase the efficiency of the cross-linking to the levels that would make a reliable detection of transient enzyme–substrate complexes possible, we substituted uridines in our substrate with 4-thiouridines. The 4-thiouridine is structurally similar to uridine, the only difference being a single atom (oxygen for sulfur) substitution that has a minimal effect on the substrate’s properties but allows for a dramatically increased efficiency of cross-linking on photoactivation (54). The substitution of uridines with 4-thiouridines did not interfere with RNase MRP cleavage (Figure 2); all uridines were substituted with 4-thiouridines during in vitro transcription in the presence of 4-thiouridine and no uridine (‘Materials and Methods’ section).

The modified substrate was exposed to UV light (365 nm) in the presence of RNase MRP (at a 1 to 1 and a 1 to 10 enzyme to substrate molar ratios), then the samples were deproteinated and analyzed by reverse transcription using primers specific to the RNA component of RNase MRP (‘Materials and Methods’ section). Typical results are shown in Figure 3; the locations of the identified cross-links are presented in Figure 1.

Interactions between protein components of the RNase MRP holoenzyme and its substrate

To identify protein components of RNase MRP that were involved in interactions with substrates, we used a 32P-radiolabeled version of the photoreactive substrate described earlier in the text. After UV-induced cross-linking between the RNase MRP holoenzyme and the radiolabeled substrate (taken at a 1 to 1 enzyme to substrate molar ratio), the samples were separated on denaturing SDS–polyacrylamide gels, and the radioactive bands were visualized (‘Materials and Methods’ section). Typical results are shown in Figure 4A.

The upper group of radioactive bands (Figure 4A) was identified as the product of cross-linking between the radiolabeled substrate (23-nt-long) and the largest protein component of RNase MRP, Pop1 (100 kDa), as well as products of cross-linking between the radiolabeled substrate and the RNase MRP RNA, NME1 (340-nt-long, 112 kDa). To differentiate between the substrate–protein complexes of interest and the substrate–RNA complexes, we treated aliquots of the samples with proteinase K and used them as controls; the bands that were not sensitive to the proteinase K treatment were interpreted as substrate–RNA complexes (Figure 4B). The remaining (proteinase K-sensitive) band was the product of cross-linking of the substrate to Pop1, as confirmed by mass spectrometric analysis.

The identities of the higher-mobility products of cross-linking (Figure 4A) could not be reliably identified by mass spectrometry analysis because of the close electrophoretic mobilities of the remaining nine RNase MRP proteins [varying in sizes from 15.5 to 32.9 kDa (29)] and because of the shifts in mobilities caused by the formation of cross-linked complexes with the RNA substrate.

To help identify the remaining RNase MRP proteins that cross-linked to the substrate, we isolated RNase MRP from additional yeast strains that had small tags fused to each of the proteins: each strain had one specific ‘elongated’ RNase MRP protein (‘Materials and Methods’ section). The addition of a tag would decrease the electrophoretic mobility of the radioactive band associated with the protein that was tagged in a particular yeast strain but should not affect other bands. A typical result of such an experiment is shown in Figure 4C, where the elongated Rpp1 protein is denoted ‘Rpp1 + Δ’. We covered all RNase MRP proteins and were able to
**DISCUSSION**

To better understand interactions of RNase MRP with its substrates, we performed cross-linking studies using a photoreactive (4-thiouridine-containing) substrate (49) and the RNase MRP holoenzyme isolated from yeast. The substitution of the uridines with 4-thiouridines in the substrate did not interfere with RNase MRP cleavage (Figure 2). The locations of the identified cross-links between the substrate and the RNase MRP RNA (NME1) are shown in Figure 1. Of the 10 RNase MRP protein components, four (Pop1, Pop4, Pop5 and Rpp1) were cross-linked to the substrate.

The identified cross-links between the substrate and the RNase MRP RNA are located in two distinct areas of the enzyme (Figure 1): in the Domain 2 near the junction with the Domain 1 (cross-links S₁, S₂), and in the phylogenetically conserved regions mCR-I, mCR-IV in the core of the Domain 1 (cross-links S₃, S₄). High-resolution structural information for the core of the Domain 1 is not yet available; however, low-resolution data (34,35,37,47), a high level of phylogenetic conservation (4,33,36,39), as well as the apparent close evolutionary relationship (4,5,55,56), all suggest that the structural organization of the core part of the Domain 1 (Figure 1) resembles that of the core of the catalytic (C-) domain (57) of bacterial RNase P [reviewed in (26)]. Several crystal structures of the C-domain of bacterial RNase P are available (40–42), and in the absence of the structure of the conserved core of the RNase MRP RNA Domain 1, the available structure of the core of the bacterial C-domain RNA can serve as its proxy.

Cross-link S₁ (Figure 1) is located in a highly conserved region near the absolutely conserved bulged uridine found in all RNases MRP/P. In bacterial RNase P, this region coordinates two metal ions that are involved in catalysis ([42] and references therein). Drawing parallels between RNase MRP and bacterial RNase P, this cross-linking site is within ~5–8 Å of the expected position of the scissile bond in the substrate (42) (Figure 5). Further, in the case of yeast RNase P, single-stranded substrates were

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**Figure 4.** UV-induced cross-linking of a radiolabeled photoreactive RNase MRP substrate to the isolated RNase MRP holoenzyme. (A) Lane 1: control (no UV irradiation); lane 2: UV-induced cross-links. The identities of the RNase MRP components that are involved in the cross-links are shown on the right; see ‘Results’ section for explanation. Fifteen per cent denaturing (SDS) polyacrylamide gel. (B) Low-mobility radioactive bands resolved on a 6% denaturing (SDS) polyacrylamide gel. Lane 1: UV-induced cross-links; lane 2: same as lane 1, but the sample was treated with proteinase K to differentiate between protein and RNA bands; lane 3: control (no UV irradiation). (C) Identification of the band corresponding to the cross-link to the protein component Rpp1. Lane 3: UV-induced cross-links for the isolated ‘wild-type’ RNase MRP holoenzyme; lane 4: UV-induced cross-links for the isolated RNase MRP holoenzyme that had an elongated version of its protein component Rpp1 (denoted ‘Rpp1+ Δ’). The single band that had changed its mobility in lane 4 was identified as the one associated with Rpp1. Lanes 1 and 2: controls (same as lanes 3, 4, respectively, but without UV irradiation). Twelve per cent denaturing (SDS) polyacrylamide gel.

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**Figure 3.** Cross-linking of a photoreactive substrate to the isolated RNase MRP holoenzyme: primer extension analysis. Lanes 1, 2, 8 and 9: sequence ladder; lanes 3 and 10: no substrate and no UV irradiation (control); lanes 4 and 11: UV irradiation in the absence of the substrate (control); lanes 5 and 12: substrate was present at a 10 to 1 substrate to enzyme molar ratio; lanes 6 and 13: UV cross-linking in the presence of the substrate (at a 1 to 1 substrate to enzyme molar ratio); lanes 7 and 14: UV irradiation in the absence of the substrate (control); lanes 5 and 12: substrate was present at a 10 to 1 substrate to enzyme molar ratio; lanes 7 and 14: UV cross-linking in the presence of the substrate (at a 10 to 1 substrate to enzyme molar ratio). The locations of secondary structure elements (Figure 1) are shown on the left. The locations of the identified substrate–RNase MRP RNA cross-links are indicated by arrows and marked S₁ to S₄ according to the labeling in Figure 1.
cross-linked to the photoreactive substrate (as aforementioned). It is not clear if Pop4 contributes to the specificity of the substrate recognition by RNase MRP or (being a basic protein) helps to bind the substrate in a non-specific fashion. It should be noted that in the related RNase P, Pop4 was shown to cross-link to poly(U) RNA, a potent inhibitor of the enzyme (50).

Pop5 and Rpp1 were also among the RNase MRP proteins cross-linked to the RNase MRP substrate. Pop5 and Rpp1 form a complex that binds to the RNase MRP Domain 1 RNA (34,35) (Figure 1) as well as to the C-domain of archaeal and eukaryotic RNases P (47,58–61). The archaeal/eukaryotic Pop5/Rpp1 complexes and the bacterial RNase P protein bind to matching parts of the respective RNA components and were proposed to play similar roles (34,60). Thus, given that the bacterial RNase P protein participates in interactions with the substrate [(42,62,63) and references therein], interactions of Pop5 and Rpp1 with the RNase MRP substrate are consistent with the overall similarity of the catalytic parts of RNases P/MRP throughout the three domains of life.

Pop1, the largest RNase MRP protein, is the fourth protein component interacting with the RNase MRP substrate. Although the role of Pop1 in RNase MRP is not yet known (64), the available low-resolution data indicate that it interacts with both of the RNase MRP RNA domains (35,47). The proximity of the substrate–RNase MRP RNA cross-link S2 to the previously identified (35) location of a Pop1 cross-link in the mCR-IV RNA region (Figure 1) can explain the Pop1-substrate cross-link; at the same time, given the large size of Pop1, it cannot be excluded that it has multiple contacts with the substrate. The large size of Pop1 and its involvement in interactions with both central and distal parts of RNases P/MRP (35,47), as well as the observed defects in RNase P/MRP assembly associated with Pop1 mutations (64), all suggested a role for Pop1 in stabilizing the overall structures of RNases P/MRP. Our cross-linking results show that Pop1 is positioned to play a role in substrate recognition as well.

Although RNase MRP is structurally and evolutionarily related to RNase P, the substrate specificities of the two enzymes differ (20,24–26,49,50,65–67). The recognition of the ‘canonical’ RNase P substrate, pre-tRNA, involves interactions between the T- and D-loops of the substrate and the specificity (S-) domain of the RNA component (40,42,48). Thus, given that pre-tRNA is not a substrate that is specifically recognized by RNase MRP, the loss of the S-domain in this enzyme seems logical. Does the RNase MRP Domain 2, which replaces the S-domain, play a role in RNase MRP substrate recognition? Our cross-linking results show that the phylogenetically conserved part of the Domain 2 (the 5’-GARAR-3’ element, Figure 1) interacts with the substrate; the determination of the exact role of this interaction in the substrate specificity of RNase MRP will require additional studies. RNase MRP cleavage absolutely requires the presence of cytosine four nucleotides 0 3’ to the cleavage site (49), whereas cleavage by RNase P does not (50). The conserved 5’-GARAR-3’ element, which is unique to RNase MRP and is found to interact with its substrate,
could potentially be involved in interaction with the conserved cytosine of the substrate. It should be noted that, given the small size of the substrate used in this study, cross-links S3, S4 (Figure 1) must be located in a proximity to the catalytic core of the enzyme (cross-links S1, S2). Thus, it is likely that the parts of the RNase MRP RNA component that form cross-links S3, S4 fold back into the Domain 1, modifying the substrate-binding interface that otherwise would be similar to that in RNase P (given the similarity between the RNase P C-domain and Domain 2 in RNase MRP). It should be noted that RNase MRP cannot cleave double-stranded substrates (49), consistent with a more limited space near the catalytic site available for the substrate compared with that in RNase P.

Given the large size of the protein part of RNase MRP, it is expected that proteins play a significant role in determining the substrate specificity of the enzyme (66). It is somewhat surprising that proteins Snm1 (28) and Rmp1 (29), which differentiate the protein part of RNase MRP from that of RNase P, are not among the proteins found to cross-link to the RNase MRP substrate. Although it is possible that Snm1 and Rmp1 play roles other than RNase MRP substrate binding, one can speculate that these proteins play roles in interacting with (unidentified) protein parts of ribonucleoprotein substrates in vivo.

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