Transmission Characteristics of Barley Yellow Striate Mosaic Virus in Its Planthopper Vector Laodelphax striatellus

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The most economically important plant viruses are specifically transmitted by phytophagous insects that significantly affect viral epidemiology. Barley yellow striate mosaic virus (BYSMV), a member of the genus Cytorhabdovirus, is transmitted by the small brown planthopper (SBPH, Laodelphax striatellus) in a persistent-propagative manner. However, the infection route of BYSMV in SBPHs is poorly understood. In this study, immunofluorescence confocal laser scanning microscopy (iCLSM) was performed to investigate the route of BYSMV in SBPHs. We unexpectedly found that BYSMV initially infected the hindgut epithelium of SBPHs, instead of the midgut epithelium initially infected by other persistent-propagative viruses. Subsequently, BYSMV disseminated to the hindgut visceral muscles and spread to other parts of alimentary canals, hemolymph, and salivary glands. Comparative analysis of gene expression on viral mRNAs and the BYSMV nucleoprotein by using different molecular detection and immunohistochemistry further demonstrated that BYSMV initially infected and replicated in the hindgut epithelial cells of SBPHs. Collectively, our study provides the first insight into that hindgut is initial infection site of BYSMV that represents a new dissemination route of persistent-propagative viruses.

Keywords: Barley yellow striate mosaic virus, cytorhabdovirus, persistent-propagative virus, Laodelphax striatellus, hindgut

INTRODUCTION

To survive in sessile plant hosts, most plant-infecting viruses rely on specific vectors for plant-to-plant transmission in nature. Although some fungi, nematodes, and various types of invertebrates serve as vectors for some plant viruses, the majority of described plant viruses are transmitted by sap-feeding hemipteran insects including aphids, leafhoppers, planthoppers, and whiteflies (Hogenhout et al., 2008). Based on acquisition and inoculation thresholds, and retention of viruses by insect vectors, transmission relationships between insect vectors and plant viruses are grouped into four types: non-persistent, semi-persistent, persistent-circulative, and persistent-propagative (Ng and Falk, 2006; Hogenhout et al., 2008; Blanc et al., 2014). The members of rhabdoviruses, orthotospoviruses, tenuiviruses, and reoviruses are transmitted by specific insect vectors in a persistent-propagative manner (Hogenhout et al., 2008). Persistent-propagative viruses
usually multiply in epithelial cells of the alimentary canal, and then pass through the gut wall to hemolymph, and finally move to salivary glands from where the viruses are introduced into healthy susceptible plant hosts (Ammar et al., 2008; Hogenhout et al., 2008). In addition, some persistent-propagative viruses, including rhabdoviruses, reoviruses, and tenuiviruses are transmitted vertically from parents to their progenies (Nault and Ammar, 1989; Falk and Tsai, 1998; Ammar et al., 2008; Wei and Li, 2016). Persistent-propagative viruses replicate and spread in their insect vectors requiring specific interaction of viral components and vector factors to overcome several major transmission barriers including midgut infection and escape barriers, salivary gland infection and escape barriers, and transovarial transmission barriers (Ammar et al., 2008; Hogenhout et al., 2008).

In competent planthopper vectors, persistent-propagative viruses must first efficiently infect the midgut epithelial cells, escape the midgut, spread to hemolymph, and finally breach the basal laminae of the salivary gland into salivary secretions that facilitate the injection of the virus into a susceptible host during insect feeding. The failure of virus transmission in incompetent insect vectors may be due to the impede of viral entrance, replication, or dissemination in insect cells or organs (Ammar, 1994; Jackson et al., 2005; Ammar et al., 2008; Whitfield et al., 2015). Midgut barriers are the first barriers and principle determinants of vector competence. Many persistent viruses are restricted to midgut epitheliums of incompetent insect vectors probably due to lack of viral receptors or some specific immune response to the viruses (de Assis et al., 2005; Ding, 2010; Donald et al., 2012; Jia et al., 2012a; Chen et al., 2015; Neelakanta and Sultana, 2016; Whitt et al., 2016). Silencing of the core component Dicer-2 of the siRNA pathway allows the efficient propagation within the midgut epithelium and transmission of Southern rice black streaked dwarf virus (SRBSDV) by its incompetent vector (Lan et al., 2015).

Plant rhabdoviruses initially fall into two genera, Nucleorhabdovirus and Cytorhabdovirus, which mature in the nucleus and cytoplasm of host cells, respectively. Besides, the Dichorhavirus and Varicosavirus genera are recently recognized as rhabdoviruses based on significant genome sequence identities with plant rhabdoviruses, even though they contain bipartite genomes/antigenomic RNA acting templates of virus replication and transcription (Jackson et al., 2005). Thus, the mRNAs and proteins accumulation of the rhabdovirus N proteins are usually detected and analyzed to represent virus infection.

In the present work, we used immunofluorescence staining and molecular analysis to investigate the temporal and spatial distribution of BYSMV within L. striatellus. We found that the hindguts of L. striatellus vectors rather than their midguts were acquisition sites for BYSMV and provided molecular evidence that BYSMV could replicate in the cytoplasm of hindgut epithelia of L. striatellus.

**MATERIALS AND METHODS**

**Insect Rearing and Virus Maintenance**

Planthopper vectors, L. striatellus, were isolated from Hebei province, China and maintained in growth chambers for nearly 4 years. The Chinese isolate of BYSMV was maintained on diseased wheat plants through serial transmission by L. striatellus as described previously (Di et al., 2014; Yan et al., 2015). Both viruliferous and healthy L. striatellus were reared separately on wheat seedlings in growth chambers with a 16 h (h) light/8 h dark and held at 25 ± 2°C during light and 20 ± 2°C during dark periods.

**Polyclonal Antibody Preparation**

The BYSMV N (GenBank: NC_028244.1) and actin genes (GenBank: KC683802.1) were amplified using specific primers of BYSMV N (5′ GGAATTCCTATGATGG AAGAAGATCATGG 3′ and 5′ CGGCTCGAGGAGAAGAGTGC TGGTCAGCAT 3′) and Actin (5′ GGAATTCACATCT GCCGAGGAGAGGG 3′ and 5′ CATGCCGACCTGGT 3′). The resulting plasmids pET-30a (+) vector. The resulting plasmids pET-30a-N and pET-30a-Actin were transformed into the Rosetta strain of Escherichia coli. BYSMV N and Actin proteins were purified from the final suspension of transformed cell treated using Ni-NTA resin (Qiagen, Hilden, Germany) as previous report (Dong et al., 2016). The purified proteins immunized rabbits, and the specific polyclonal antisera was used to purify Immunoglobulin G (lgG) using A-Sepharose affinity column (Sigma–Aldrich).

**The Acquisition Efficiency of BYSMV by L. striatellus**

Second-instar nymphs of L. striatellus (n = 50, three biological repetitions) were allowed 1, 4, 24, 36, and 48 h acquisition...
access period (AAP) on BYSMV infected wheat plants. Insects were incubated on healthy rice seedlings for a 12-day inoculation period (IP), and then examined by iCLSM (immunofluorescence confocal laser scanning microscopy). The planthoppers fed on healthy wheat plants were acted as negative controls.

**TABLE 1 | Acquisition access period (AAP) and BYSMV-positive rate of SBPHs.**

| AAP | Infectivity (No. of infected/total insects) |
|-----|------------------------------------------|
|     | Experiment 1 | Experiment 2 | Experiment 3 | Total (percentage) |
| 1 h | 1/49         | 1/50         | 2/50         | 4 (149(2.7%) |
| 4 h | 5/50         | 6/50         | 3/45         | 14/145(9.7%) |
| 24 h | 12/50       | 13/50        | 13/51        | 38/151(25.2%) |
| 36 h | 30/53       | 18/43        | 19/50        | 67/146(45.9%) |
| 48 h | 27/50       | 20/43        | 19/41        | 66/134(49.3%) |

Second-instar nymphs of SBPHs were fed on BYSMV-infected barley plants for different time acquisition access, then incubated on healthy barley seedling for 12 days. The whole alimentary canals were dissected for iCLSM test using BYSMV-N antibody conjugated directly to fluorescein-5-isothiocyanate (FITC, Sigma–Aldrich). To distinguish muscle fibers from other tissues, actin was stained with phalloidin-rhodamine.
with phalloidin–rhodamine (Invitrogen). Finally, the stained products were washed with PBS buffer and processed for Olympus immunofluorescence microscopy (Olympus FV1000). The organs dissected from healthy planthoppers were stained as negative controls. DAPI, GFP, and RFP fluorescence were visualized under 405, 488, and 543 nm, respectively. The value of gain and offset is set to 1%. The high voltage is set to 400–600. To avoid cross fluorescence effects, a line sequential scanning mode was used for image capture at a 1024 × 1024 pixel resolution.

Hemolymph Collection and Immunofluorescence Staining

Hemolymph collection and immunofluorescence staining were performed by the methods as reported previously (Wang et al., 2014; Huo et al., 2018). Briefly, SBPHs were anesthetized on ice for 30 min. The forelegs at the coxa-trochanter joint were severed by forceps. The hemolymph was disemboqued and drawn onto the tip of forceps. The collected hemolymph was quickly washed in PBS drops, where the drops were on poly (L-lysine)-coated cover glasses. The glasses were incubated at 37°C for 30 min to allow the adhesion of hemocytes. The hemolymph was fixed in 4% paraformaldehyde for 30 min at room temperature and washed in 0.01 M PBS buffer (pH 7.4). Subsequently, hemolymph was permeabilized in PBS buffer harboring 0.2% Triton X-100 for 5 min at room temperature and stained with BYSMV N-FITC antibody (1:100). Finally, stained products were washed with PBS buffer and processed for Olympus immunofluorescence microscopy (Olympus FV1000). The planthoppers fed on healthy wheat plants were used as negative controls.

Western Blotting

At 4 and 8 days after AAP, the alimentary canals of 50 planthoppers were dissected and separated into two parts, hindgut tissues and the remaining tissues (including esophagus, anterior diverticulum, midgut, and malpighian tubules). Total proteins were isolated from the two parts of alimentary canals in SDS buffer [100 mM Tris-HCl (pH 6.8), 20% glycerol, 4% SDS, 0.2% bromophenol blue, 10% β-mercaptoethanol] for western blotting, separated in SDS-PAGE gels, and transferred onto nitrocellulose membranes. Anti-N (1:3000) polyclonal antibody was used to quantify the accumulations of the BYSMV N proteins, as described previously (Yan et al., 2015). Anti-actin (1:1000) polyclonal antibody was used to quantify actin accumulations as internal loading control.

RNA Analysis

Total RNA was isolated from the hindgut tissues and the remaining tissues of the alimentary canals of 200 planthoppers. The RNA was quantified using a NanoDrop ND-1000 (Thermo Fisher Scientific). Total RNA (1.5 μg) was reverse-transcribed by using M-MLV Reverse Transcriptase (Promega) with oligo (dT) 20 primer and the produced cDNAs were used as templates for semi-quantitative RT-PCR and RT-qPCR. Expression levels of N, P, and M genes of BYSMV were examined by semi-quantitative RT-PCR with primers of N (5’GAAGATCATGGATTGGACAGAGAG-3’; 5’GCAGGAGTGAAGCCGGGAT3’), P (5’AGATGGGATCTTCGCTAGGC-3’; 5’CTCCACACCGGAGATATACC3’), M (5’GGAGGTAGACTTTGGAGAAAGGAG3’; 5’CGAAGATCCAGATGAAGCTG3’) according to the manufacturers’ instructions. The actin gene of L. striatellus served as an internal control with the specific primers (5’GCCCACCTACGAGGTTAAT3’; 5’CCATTCTCTGTTCGAA GTCCAG3’). The semi-quantitative RT-PCR was performed using the following thermal cycling profile: 94°C for 10 min, followed by 25 cycles of amplification (94°C for 30 s, 55°C for 30 s, and 72°C for 30 s), and 72°C for 10 min. The relative intensity of each lane was quantified using Quantity One software. The RT-qPCR assay was performed on a BIO-RAD CFX96™ Real-Time System (Bio-Rad) using the 1X SsoFastTM EvaGreen Supermix (Bio-Rad), according to the manufacturers’ instructions. The cycling parameters were: 95°C for 3 min, followed by 48 cycles of 95°C for 10 s and 55°C for 20 s. Actin gene of L. striatellus was used as an internal control.

Immunohistochemistry

Hindgut samples were dehydrated in ethanol series and embedded in paraffin wax. Serial sections (8 μm) were mounted on slides coated with poly-L-lysine, incubated with 10% normal donkey serum to reduce background, and incubated with the primary antibody (1:200) raised against BYSMV N protein at 4°C for 12 h. After washed three times in PBS buffer, resulting sections were incubated with a secondary antibody (1:50), donkey anti-rabbit IgG conjugated with Alexa fluor® 488 at 25°C for

| TABLE 2 | Statistics of BYSMV antigens in various organs/tissues of SBPHs. |
| Days padp | Hindgut | Midgut | Esophagus | Anterior diverticulum | Salivary glands | Total number |
| 2 days | 17 | 0 | 0 | 0 | 0 | 45 |
| 3 days | 21 | 2 | 0 | 0 | 0 | 50 |
| 5 days | 24 | 10 | 4 | 2 | 0 | 46 |
| 7 days | 24 | 23 | 17 | 10 | 7 | 50 |
| 12 days | 26 | 26 | 26 | 26 | 25 | 47 |

Second-instar nymphs of SBPHs were fed on BYSMV-infected barley plants for a 36-h acquisition access, then incubated on healthy barley seedling for 2, 3, 5, 7, and 12-day padp. The whole internal organs were dissected for iCLSM test using BYSMV-N antibody conjugated with fluorescein isothiocyanate (N-FITC).
40 min, and then processed for Olympus immunofluorescence microscopy (Olympus FV1000). Hindguts of healthy *L. striatellus* were used as negative controls.

**RESULTS**

**BYSMV Acquisition Ability of *L. striatellus***

To determine acquisition efficiency of BYSMV by its planthopper vector, we allowed the second-instar nymphs of non-viruliferous SBPH populations to feed on BYSMV-infected wheat plants for 1, 4, 24, 36, and 48 h AAP. These insects were reared on healthy wheat seedlings for a 12-day incubation period. Internal organs of about 50 insects were dissected and stained by BYSMV N protein-specific IgG directly conjugated with fluorescein isothiocyanate (N-FITC) for iCLSM. As negative controls, the internal tissues of SBPHs exposed to healthy plants did not show specific fluorescence (Figure 1A and Supplementary Figure S1). In contrast, the BYSMV-infected alimentary canals showed high intensity of GFP signal that represented accumulation of the BYSMV N protein (Supplementary Figure S1).

Results from three independent repetitions with a total number of 134 –151 insects showed that 2.7%, 9.7%, 25.2%, 45.9%, and 49.3% of SBPHs were infected by BYSMV with 1, 4, 24, 36, and 48 h AAhs, respectively (Table 1). The results indicated that *L. striatellus* could acquire BYSMV after a feeding period of 1 h, and the BYSMV acquisition rates of nymphs increased to 49.3% after 48 h feeding (Table 1).

**BYSMV Infection Starts in the Hindgut of *L. striatellus***

To determine the temporal and spatial distribution of BYSMV in its vector, we allowed second-instar nymphs of SBPH to feed
FIGURE 3 | The propagation of BYSMV in L. striatellus. (A) The virus accumulations of infected hindguts and other tissues of the alimentary canals at 4-day and 8-day padp were verified by western blotting analysis with anti-N polyclonal antiserum. The whole alimentary canals of viruliferous SBPHs served as negative controls (Mock). The actin of L. striatellus was detected as the loading controls. (B) RT-qPCR assay detecting relative accumulation of BYSMV-N mRNA in the infected hindguts and other tissues of the alimentary canals at 4-day and 8-day padp, respectively. The viral expression level of 4-day-hindgut was normalized as the relative expression level, which was set to 1. Data points are the mean value of three independent experiments. The significant difference was analyzed by Student’s t-test. **P-value < 0.01. Actin was used as an internal control. Other tissues of infected alimentary canals included esophagus, anterior diverticulum, midguts, and malpighian tubules.

TABLE 3 | Initial infection sites of plant persistent-propagative viruses.

| Genus               | Virus name                                               | Type of vector | Initial entry                  | Reference                      |
|---------------------|----------------------------------------------------------|----------------|-------------------------------|--------------------------------|
| Cytorhabdovirus     | Barley yellow striate mosaic virus (BYSMV)               | Planthopper    | Hindgut                       | –                              |
| Nucleorhabdovirus   | Maize mosaic virus (MMV)                                  | Planthopper    | Anterior diverticulum, Midgut | Ammar el and Hogenhout, 2008   |
| Tenuivirus          | Rice grassy stunt virus (RGSV)                           | Planthopper    | Midgut                        | Zheng et al., 2014             |
| Tenuivirus          | Rice stripe virus (RSV)                                  | Planthopper    | Midgut                        | Wu et al., 2014                |
| Orthotospovirus     | Tomato spotted wilt virus (TSVV)                         | Thrips         | Midgut                        | Kritzman et al., 2002          |
| Fijivirus           | Southern rice black-streaked dwarf virus (SRBSDV)        | Planthopper    | Midgut                        | Jia et al., 2012b              |
| Oryzavirus          | Rice ragged stunt virus (RRSV)                           | Planthopper    | Midgut                        | Jia et al., 2012c              |
| Phytoreovirus       | Rice dwarf virus (RDV)                                   | Leafhopper     | Filter chamber                 | Chen et al., 2011              |
| Phytoreovirus       | Rice gali dwarf virus (RGDV)                             | Leafhopper     | Filter chamber                 | Zheng et al., 2015             |

Results of three independent repetitions (n = 50) are summarized in Table 2. The micrographs illustrating key events in BYSMV dissemination route in SBPH were shown in Figure 1. At 1-day post-first access to diseased plants (padp), ingested BYSMV were labeled as punctate structures throughout the gut lumen (Figure 1B). One day later, a sign of specific labeling of BYSMV antigen was only observed in the hindgut epithelium in about 37.8% of SBPHs examined (Figures 1C,D and Table 2). At this time, no detectable virus accumulation was found in any other tissues of alimentary canal (Figure 1C). At 3-day padp, BYSMV N was detected in a high proportion of insect hindgut tissues (up to 42%), and some signals had disseminated to the hindgut visceral muscles from the initial infection sites (Figure 1E and Table 2). At 5-day padp, BYSMV subsequently spread to the longitudinal and circular muscle cells surrounding hindgut tissues in about 52% of SBPHs examined (Figure 1F and Table 2). The individual fluorescence panels of 1, 2, 3, and 5 padp were shown in Supplementary Figure S2. Simultaneously, a relative low ratio of midgut (up to 22%), esophagus (up to 8.7%), and anterior diverticulum (up to 4.3%) tissues had been infected by BYSMV (Table 2).

In the following observation, BYSMV N was extensively detected in the alimentary canals of insects, including esophagus, anterior diverticulum, midgut, malpighian tubule, and hindgut at 7-day padp (Figures 1G–I). At 12-day padp, BYSMV was detected in the hemocytes, salivary glands, as well as in other tissues of about 50% insects examined (Figures 1J–L and Table 2). The individual fluorescence panels of 7, 11, and 12 padp were shown in Supplementary Figure S3. Collectively, BYSMV initially infected the hindgut epithelium of SBPH nymphs, which is distinguishable from other persistent-propagative viruses those establish their initial infection in midgut epithelium of their vectors (Hogenhout et al., 2008; Whitfield et al., 2015; Neelakanta and Sultana, 2016; Wei and Li, 2016). To our knowledge, these observations reveal a new tissue tropism of persistent-propagative viruses in which uptake of viruses occurs in the hindgut of insect vectors.

Infection of BYSMV in Cytoplasm of SBPH Hindgut Epithelium

To determine whether the detected BYSMV antigens were from replicating BYSMV or BYSMV-bearing undigested foods, the 6–8 µm thick paraffin sections of viruliferous L. striatellus hindguts on diseased barley plants for a 36-h AAP. Then, the whole internal organs of 50 planthoppers were dissected and stained by N-FITC and the actin dye phalloidin–rhodamine (Invitrogen) for iCLSM.

on diseased barley plants for a 36-h AAP. Then, the whole internal organs of 50 planthoppers were dissected and stained by N-FITC and the actin dye phalloidin–rhodamine (Invitrogen) for iCLSM.
were processed for immunohistochemistry. Paraffin sections of healthy *L. striatellus* hindguts were used as negative control (Figure 2A). At 3-day padp, BYSMV were detected in the cytoplasm of hindgut epithelial cells (Figure 2B). At 5-day padp, BYSMV has spread to the visceral muscles surrounding the hindguts (Figure 2C), which is consistent with the observation of the whole organs described above (Figure 1). These results were further confirmed by using three-dimensional rendered confocal images (Supplementary Figure S4). Collectively, these results indicate that the observed fluorescence is due to authentic virus infections but not undigested BYSMV particles.

The Multiplication of BYSMV in the Hindgut of SBPHs

To further confirm the propagation of BYSMV in hindgut tissues, the accumulation of BYSMV N protein in hindgut tissues and the other parts of alimentary canals were detected by Western blotting analysis using the BYSMV N specific antibody. At 4-day padp, the N protein was easily detected in hindgut tissues rather than in the other parts of the alimentary canals, indicating that BYSMV established the initial infection in hindgut tissues (Figure 3A). At 8-day padp, higher amount of N proteins was accumulated in the hindgut tissues compared with those of at 4-day padp (Figure 3A). While, the N protein was detected at a relative low level in the remaining tissues at 8-day padp, indicating BYSMV begin to spread into these tissues at this time (Figure 3A).

We further used a quantitative real-time PCR assay to confirm the mRNA level of BYSMV N gene. To rule out DNA contamination, the DNase I treated RNA samples without reverse transcription (Non-RT) were directly used as templates for PCR. The result showed that no DNA bands were amplified from the RNA samples of infected hindgut tissues (Supplementary Figure S5A), indicating the contaminated DNA was treated by DNase I completely. At 4-day padp (Figure 3A), while the N protein was detected at a relative low level in the remaining tissues at 8-day padp, indicating BYSMV begin to spread into these tissues at this time (Figure 3A).

Collectively, these results suggest that BYSMV initially infected and propagated in the *L. striatellus*.

**DISCUSSION**

The plant viruses of agricultural importance are usually transmitted by plant-feeding hemipteran insects including aphids, planthoppers, leafhoppers, whiteflies. Persistent-propagative viruses have distinct tissue tropism and infection route in their insect vectors (Table 3). For example, rice dwarf virus (RDV), rice ragged stunt virus (RRSV), rice stripe virus (RSV), rice grassy stunt virus (RGSV), and tomato spotted wilt virus (TSWV) initially infected midgut epithelial cells of their insect vectors (Kritzman et al., 2002; Jia et al., 2012b,c; Wu et al., 2014; Zheng et al., 2014). Maize mosaic virus (MMV), a member of *Nucleorhabdovirus*, initially infect the midgut and anterior diverticulum of its planthopper vector (Ammar el and Hogenhout, 2008). Here, we unexpectedly discovered that the hindgut of the planthopper vector was the acquisition site for BYSMV. To the best of our knowledge, this is the first time to reveal a new tissue tropism of persistent-propagative viruses in which uptake of viruses occurs in the hindgut of insect vectors.

The microvilli and basol nalina of insect midgut is the initial barriers for viral entry and dissemination. Thus, many studies have documented that the insect midgut epithelium is a major determinant of vector competence for a variety of persistent-propagative viruses (Hogenhout et al., 2008; Whitfield et al., 2015; Neelakanta and Sultana, 2016; Wei and Li, 2016). The inability of BYSMV to infect the midgut epithelium might due to lack of viral receptor in the midgut, or the innate immune responses in the midgut epithelium that control viral accumulation and block viral transmission (de Assis et al., 2005; Ding, 2010; Donald et al., 2012; Jia et al., 2012a; Chen et al., 2015; Neelakanta and Sultana, 2016; Whitt et al., 2016). Nonetheless, the efficient infection of BYSMV entry and dissemination from the hindgut epithelium allow the *L. striatellus* to be a competent vector for BYSMV transmission.

The hindguts of aphids are initially infected by the majority of species of luteoviruses, which are transmitted by aphids in a non-propagative, circulative manner (Gildow, 1993; Garret et al., 1996; Reinbold et al., 2003; Gray et al., 2014). Unlike the luteoviruses, BYSMV is able to initially infect and efficiently propagate in the epithelial cells of the hindgut and then disseminate into the midgut muscles and other organs (Figure 3). Despite the luteoviruses unable to replicate in the vector, long feeding periods on the plants with high virus acquisition facilitate the enough amounts of viruses accumulation and transmission efficiency by the aphids (Gray et al., 1991). In addition, the non-enveloped icosahedral particles of luteoviruses can interact with specific molecules on the surface of the epithelial cells of guts and are likely to enter the epithelial cells by endocytosis (Gildow, 1993). These resulted suggested that different strategies may be employed by the viruses for entry and spread in vectors. Further experiments are needed to determine whether the pattern of the enveloped virions of BYSMV enter the epithelial cells of hindgut via a receptor-mediated endocytic fashion.

**AUTHOR CONTRIBUTIONS**

X-BW and QC conceived and designed the experiments. QC, W-YY, QG, Z-HJ, S-YL, X-DF, and D-MG performed the experiments. X-BW and QC analyzed the data and drafted the manuscript. X-BW and YW participated in experimental coordination and revision of the manuscript. X-BW and QC proofread and finalized the manuscript.
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