Lysinibacillus fusiformis, which prior to 2007 was known as Bacillus fusiformis, is a Gram-positive nonmotile bacterium of the genus Lysinibacillus (1). The majority of studies on L. fusiformis are related to its pathogenicity (2, 3), and few researchers have focused on its industrial and agricultural roles. We previously isolated L. fusiformis SW-B9 from soil, which can produce high amounts of vanillin from isoeugenol (4). Vanillin is widely used in the beverage, food, pharmaceutical, and medical industries (5). Several strains have been shown to be able to convert eugenol or isoeugenol to vanillin (5, 6). However, for L. fusiformis, only the SW-B9 strain was reported for its biotransformation of isoeugenol to vanillin (4). Therefore, the complete sequencing of strain SW-B9 will not only enrich the genome sequence database of L. fusiformis but also further our understanding of the genetic, phylogenetic, and physiological properties of this strain.

The genomic DNA of strain SW-B9 was sequenced using the MiSeq system. The whole-genome shotgun run yielded 2,430,573 paired-end reads, accounting for 1,220,147,646 bases in total. De novo assembly was performed using SPAdes (7), resulting in an assembly of 22 contigs of >1,000 bp. The total size of the assembly was 4.7 Mbp, with an N50 of 997.350 kbp and a G+C content of 37%. The annotation of the genome was accomplished with the NCBI Prokaryotic Genome Automatic Annotation Pipeline (PGAAP) (8). A total of 4,400 coding DNA sequences, 70 tRNAs, and 28 rRNAs were predicted. Out of all the genes, 1,721 coding sequences (CDSs) were assigned to 178 KEGG pathways.

A comparative genome analysis was used to measure the similarities between strain SW-B9 and other strains using the reciprocal smallest distance (RSD) (9). A total of 4,040 CDSs, accounting for 92% of the coding genes in the SW-B9 strain, are shared with L. fusiformis RB-21 (GenBank accession no. JPEF00000000.1). These results indicate that most of the coding sequences in the SW-B9 strain are highly conserved within other L. fusiformis strains, although there is no vanillin biotransformation in these strains. Monoxygenases were considered to be the enzymes that convert isoeugenol to vanillin (10). To find the isoeugenol monooxygenase gene (iem), we used the iem genes reported in Pseudomonas putida IE27 (11), Pseudomonas nitroreducens Jinn (12), Neofusicoccum parvum isolate UCR-NP2 (13), Verticillium albo-aeneum VaMs.102, and Colletotrichum fioriniae P17 (14), to blast against all proteins in the SW-B9 strain. The results show that there is no similar gene in strain SW-B9. However, in the genome of strain SW-B9, we found 15 candidate monoxygenases, enabling an exploration of the mechanism of vanillin conversion in this strain. Therefore, we speculate that the pathway to produce vanillin in SW-B9 is special, and further genetic research will be worthwhile.

The genome sequence of strain SW-B9 serves as a useful resource for further investigation of the molecular basis of its potential in the biotransformation from isoeugenol to vanillin. Systematic annotations of the genome will reveal physiological differences among the various L. fusiformis strains.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession no. JRBA00000000. The version described in this article is the first version, JRBA01000000.

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