Research Article

Performance of *Anastrepha fraterculus* (Wiedemann) (Diptera: Tephritidae) rearing on fruits of two guava varieties under forced infestation

Rendimiento de *Anastrepha fraterculus* (Wiedemann) (Díptera: Tephritidae) en frutas de dos cultivares de guayaba bajo infestación forzada

Adalton Raga¹, Ester Marques de Sousa¹, Léo Rodrigo Ferreira Louzeiro¹, Sara Braga e Silva¹ and Juliana Sanches²

¹Instituto Biológico, Secretaria da Agricultura e Abastecimento do Estado de São Paulo. Alameda dos Videiros 1097, CEP 13101-680, Campinas, SP, Brazil. adalton.raga@gmail.com
²Instituto Agronômico, Secretaria da Agricultura e Abastecimento do Estado de São Paulo. Avenida Theodureto de Almeida Camargo 1500, CEP 13075-630. Campinas. SP, Brazil.

ZooBank: urn:lsid:zoobank.org:pub:BCFD37F0-4F16-4B5F-9788-F1606497C543
https://doi.org/10.35249/rche.46.4.20.06

Abstract. *Anastrepha fraterculus* (Wiedemann) is dominant in the main guava’s crops. Guavas are an important host of *A. fraterculus* for providing a good performance for the immature and high reproductive capacity. We evaluated the performance of *A. fraterculus* in two guava varieties related to the fruit physicochemical composition under laboratory conditions. Mature-green guavas of “Tailandesa” (red pulp) and “Kumagai” (white pulp) varieties were exposed to forced infestation by *A. fraterculus* in laboratory. An additional sample of each variety was submitted to physicochemical analysis. Similar quantities of pupae per fruit (128.5 – 156.0) and pupal viability (85.9% and 87.5%) were obtained per variety, but “Kumagai” produced significantly more pupae of *A. fraterculus* per fruit mass (1000.6 pupae/kg). Pupal weight was higher in “Tailandesa” (115.3 mg/10 pupae) than “Kumagai” (82.6 mg/10 pupae). Significant positive correlations were obtained between pupae per fruit or kg versus adults per kg for both varieties. A positive correlation was observed between pupae per fruit and pupae per kg of “Kumagai” guavas. Peel and pulp firmness were inversely correlated with pupae per fruit and pupae per kg. Weight of 10 pupae of *A. fraterculus* was correlated with fruit weight of “Tailandesa” guavas. The multivariate analysis PCA showed a relationship between pupae per kg and peel firmness, pulp firmness, Ratio, soluble solids, and titratable acidity in “Kumagai” guava. This explains the higher infestation index pupae per kg and better reproductive performance of *A. fraterculus* in “Kumagai” guava.

Key words: Infestation indices, Insecta, physicochemical parameters, *Psidium guajava*.

Resumen. *Anastrepha fraterculus* (Wiedemann) es dominante en los principales cultivos de guayaba. Las guayabas son un hospedador importante de *A. fraterculus* por brindar un buen desempeño a los inmaduros y con alta capacidad reproductiva. Evaluamos el comportamiento de *A. fraterculus* en dos variedades de guayaba relacionados con la composición fisicoquímica del fruto en condiciones de laboratorio. Las guayabas maduras de color verde de los cultivares “Tailandesa” (pulpa roja) y “Kumagai” (pulpa blanca) fueron expuestas a la infestación forzada por *A. fraterculus* en el laboratorio. Una muestra adicional de cada variedad se sometió a análisis fisicoquímico. Se obtuvieron cantidades...
Raga et al.: Performance of *Anastrepha fraterculus* on fruits of guava under forced infestation.

Introduction

In fruit flies (Diptera: Tephritidae), host-marking generally involves dragging the aculeus inside the fruit and depositing a host-marking pheromone on it. Females should be selective — choosing to place eggs on hosts that tend to be associated with relatively high juvenile growth and survival (Diaz-Fleischer et al. 2000). The knowledge of larval performance under different fruit compositions should therefore shed light on the determinants of insect host ranges (Hafsi et al. 2016).

The perfect correlation between oviposition preference and performance indicate that *Bactrocera invadens* (Drew, Tsuruta & White) females maximize offspring survival (Akol et al. 2013). Females of *Anastrepha ludens* (Loew) not only produce eggs continuously to quickly respond to egg-laying opportunities, but the eggs they produce are largely fertile (Aluja et al. 2011).

Fruit flies are persistent pests in guava (*Psidium guajava* L.) crops (Gould & Raga 2002). *Anastrepha fraterculus* (Wied.) has been observed in 116 fruit hosts (Zucchi & Moraes 2008) and is found in wild and commercial guavas in Brazil (Raga et al. 2005, 2006). Fruit fly infestations begin when guavas reach 2 cm (Souza-Filho et al. 2009) and remain until harvesting.

Guava is one of the most widely grown fruits in tropical and sub-tropical regions of the world, with established markets in more than 60 countries (Mitra & Irenaeus 2018). The production area of guava in Brazil was estimated to be 21,500 ha in 2018 (IBGE 2019). Under pruning and irrigation systems, the growing region of Campinas (São Paulo) produces table guavas of both red and white pulps year-round.

Orchards have higher uniformity because trees of the same cultivar are clones produced via vegetative propagation and grafted on to rootstocks that are now commonly produced vegetatively as well (Kogan & Hilton 2009). The continuous and high-level of damage inflicted by fruit flies on guava crops is caused by both susceptible variety and continuous fructification (Raga et al. 2006). The susceptibility of commercial guava varieties likely is related to fruit volatiles for female oviposition and favourable physicochemical characteristics of the fruit for larval development.

Guava exhibit thick pulp, few seeds and stone cells, and the desirable fruit characteristics for table purposes are high sugar concentrations and typical aroma (Singh 2011). Collecting fruits from guava germplasm field, Raga et al. (2006) obtained a maximum of 49 pupae of fruit fly per fruit, where *A. fraterculus* was dominant. The larval density of tephritids varied according to the guava growing region and respective temporal sampling (Raga et al. 2005); however, little is known about the level of larvae support each fruit provides and the similes de pupas por fruto (128,5 - 156,0) y viabilidad de pupas (85,9% y 87,5%) por cultivar, pero “Kumagai” produjo significativamente más pupas de *A. fraterculus* por masa de fruto (1000,6 pupas / kg). El peso de las pupas fue mayor en “Tailandesa” (115,3 mg / 10 pupas) que en “Kumagai” (82,6 mg / 10 pupas). Se obtuvieron correlaciones positivas significativas entre pupas por fruto o kg versus adultos por kg para ambos cultivares. Se observó una correlación positiva entre pupas por fruto y pupas por kg de guayabas “Kumagai”. La firmeza de la cascara y la pulpa se correlacionó inversamente con las pupas por fruto y las pupas por kg. El peso de 10 pupas de *A. fraterculus* se correlacionó con el peso del fruto de las guayabas “Tailandesa”. El análisis multivariado PCA mostró una relación entre pupas por kg y firmeza de la cascara, firmeza de la pulpa, relación sólidos solubles y acidez titulable en la guayaba “Kumagai”. Esto explica el mayor índice de infestación de pupas por kg y el mejor desempeño reproductivo de *A. fraterculus* en la guayaba ‘Kumagai’.

Palabras clave: Índices de infestación, Insecta, parámetros fisicoquímicos, *Psidium guajava*.
consequent larval performance in different varieties. Here, we associate the performance of *A. fraterculus* rearing in two guava varieties with the physicochemical composition of the fruit under laboratory conditions.

**Material and Methods**

**Infestation Bioassay**

Mature green guavas from cultivars “Tailandesa” (red pulp) and “Kumagai” (white pulp) were collected from a commercial orchard located in Campinas, State of São Paulo, Brazil. The orchard conducts fruit bagging without application of synthetic insecticide. Fruit were immediately brought to the laboratory and washed with a neutral detergent to remove any residue. The mean fresh fruit weight (g) was measured immediately prior to infestation. *Anastrepha fraterculus* specimens tested here were sourced from colonies that have been maintained in the Instituto Biológico (Campinas) since 1993 (Raga & Sato 2011). Forty-two fruits of each variety were exposed to mature, 17-day-old females of *A. fraterculus* for 24 hours, with 10 females being exposed per fruit. The total number of guavas per variety were separated equally into two laboratory cages (1 × 1 × 1 m) during the infestation, under 14:10 L:D photoperiod.

After infestation, the guavas were separated and kept in circular plastic containers, 15 cm in diameter (1 litre), containing approximately 1.5 cm of vermiculite substrate. The containers were capped with voile and bound with an elastic. We evaluated the recovered pupae and adults approximately 22 and 38 days after infestation, respectively. Ten pupae per fruit were weighted (mg). Experiments were undertaken in a room at 21.4-26.0 °C and 46-68% RH.

**Fruit physicochemical analysis**

Ten additional fruits of each variety were collected on the same date of the infestation assay for physicochemical analysis. The following fruit parameters were analysed in the Laboratory of Physiology and Postharvest of the Instituto Agronómico.

**Colour:** determined using the Minolta BC 10 colourimeter and expressed according to the system proposed by the Commission Internacionale de L'Eclaraige (CIE) in L* a* b* (two readings per peel and pulp colour). The values were expressed in a* (from green to red), b* (from blue to yellow), luminosity (L), hue angle, which indicates the location of the colour in a diagram and is calculated using the formula: 
\[
\text{Hue} = \tan^{-1}\left(\frac{\text{b}}{\text{a}}\right)
\]
and chromaticity, which indicates the intensity of colour and is calculated using the formula: 
\[
\text{C} = (\text{a}^2 + \text{b}^2)^{1/2};
\]

**Firmness:** determined with a manual penetrometer with a tip diameter of 8 mm and a penetration of 6 mm. Two readings per fruit in the middle region (before and after peel removal) were taken. The results were expressed in Newtons (N);

**Titratable acidity (TA):** determined using 10 g of pulp ground in a blender and homogenized with 90 mL distilled water. The sample was titrated using a pH-meter with a sodium hydroxide solution (0.1N) until a pH of 8.1 was reached. Results were expressed in grams of citric acid per 100 g of sample;

**pH:** measured using a digital pH-meter with the electrode placed directly into the blended pulp;
Soluble solids (SS): estimated using a bench refractometer with a ± 0.0003 precision by placing a small sample of blended pulp on the reading prism. Results were expressed as percentages;

Ratio: the ratio between SS/TA.

Statistical analysis

We considered each fruit as one replication for the fruit fly infestation assay. The level of infestation for the two species and for the various bag colours was measured by the number of pupae and adults produced per fruit and was compared using a one-way analysis of variance (Sisvar, version 5.6) at P < 0.05 (Ferreira 2019). Pearson’s correlation (p-value < 0.05) using SAS University Edition software (Version 3.8) (SAS Enterprise Miner 13.1. SAS Institute Inc., Cary, NC.) was used to determine the interaction between the two factors. Principal component analysis (PCA) was conducted using R statistical software, version 4.0.1 (R Core Team 2019), and applied to all analytical data collected for the guavas.

Results

“Tailandesa” guavas produced similar quantities of A. fraterculus compared to “Kumagai” guavas, corresponding to 128.5 (range 0 – 382) and 156.0 (range 3 – 304) pupae per fruit (Student’s test, P < 0.05), respectively (Figs. 1, 2). However, significantly more pupae of A. fraterculus per kg of fruit were recovered from “Kumagai” guavas than from ‘Tailandesa’ guavas, corresponding to 1000.6 (range 21.1 – 1788.2) and 365.3 (range 0 – 855.5) pupae, respectively (Figs. 1, 2). No statistical differences were detected between guava varieties in terms of pupal viability (85.9% and 87.5%) (Figs. 1, 2). Pupal weight was higher in “Tailandesa” (115.3 mg/10 pupae) than in “Kumagai” (82.6 mg/10 pupae) (Figs. 1, 3).

The values of physicochemical characteristics from the two guava varieties are shown in Table 1. “Tailandesa” guavas exhibited a significantly higher fruit weight, a, hue and chroma of pulp (Student’s test, P < 0.05). L*, a*, b*, hue and chroma of skin, pH, titratable acidity (TA) and ratio (SS/TA) were statistically similar between the varieties. “Kumagai” (white pulp) fruit exhibited higher values of pulp L* and b* and peel and pulp firmness than “Tailandesa” fruit (red pulp).

We obtained a significant positive correlation between the number of pupae per fruit or kg versus the number of adults per kg for “Tailandesa” (r = 0.94, P < 0.0001; r = 0.99, P < 0.0001) and “Kumagai” (r = 0.95, P <0.0001; r = 0.94, P < 0.0001), respectively. No significant correlation was detected between pupal viability and infestation indices for “Tailandesa” guavas (Fig. 3); however, a positive correlation was observed between pupal viability with pupae per fruit (r = 0.85, P < 0.001) and pupae per kg (r = 0.80, P < 0.001) of “Kumagai” guavas (Fig. 3). Peel (r = -0.59, P = 0.0741; r = -0.68, P = 0.0308) and pulp (r = -0.71, P = 0.0225; r = -0.72, P = 0.0193) firmness were inversely correlated with pupae per fruit and pupae per kg, respectively. The weight of 10 pupae of A. fraterculus was correlated with the fruit weight of “Tailandesa” guavas (Fig. 3). No correlations were found between pupal infestation and the remaining physicochemical parameters.

Principal components analysis (PCA) was performed on the dataset of the results for the two guava varieties and 20 traits assessed. The total variability was explained by one principal component (PC). Of these, the first (PC1) accounted for 100% of the total variation (Fig. 4). PC1 was effective in separating the two guava varieties. The examination of the PC1 loadings suggests that this separation is due to pupae per fruit, pupae per kg, pupal viability, peel and pulp luminosity, peel and pulp b*, peel hue angle, peel chromaticity, peel and pulp firmness, titratable acidity, soluble solids and SS/TA ratio. The PC1 scores and
loadings suggest that the concentrations or values of these compounds were highest for white pulp and lowest for red pulp, which were located on the negative side of the PC1 axis. The opposite was observed for pH, pulp chromaticity, peel a*, pulp a*, pulp hue angle and pupal weight, which also contributed to the separation of the guava varieties in PC1, although with negative loadings. The PC1 scores and loadings suggest that pupal viability was present at a distant level in the red and white guavas, which were centred on the PC1 axis.

**Figure 1.** Infestation parameters of *Anastrepha fraterculus* in guavas “Tailandesa” (red pulp) and “Kumagai” (white pulp). Bars with different letters indicate significant differences by Student’s t-test (P < 0.05).

**Figure 2.** Pupal infestation pattern of *Anastrepha fraterculus* in guavas “Tailandesa” (red pulp) and “Kumagai” (white pulp) (n=42).
Figure 3. Pupae of *Anastrepha fraterculus* per fruit or kg as function of pupal viability and, pupal weight versus guava weight of “Tailandesa” (red pulp) and “Kumagai” (white pulp) varieties (n=42).

Figure 4. Plots of the principal component analysis of physical and physicochemical profile data for two guavas. PC1/PC2 scores (●) and loadings plot (●) accounted for 100.00% of the total variation. Samples: Red pulp (“Tailandesa” guava), White pulp (“Kumagai” guava). Trait abbreviations: pupae per fruit [PPF], pupal weight [PW], pupae per kg [PPK], pupal viability [PV], luminosity peel [LPE], a* peel [APE], b* peel [BPE], angle Hue peel [HPE], chromaticity peel [CPE], luminosity pulp [LPU], a* pulp [APU], b* pulp [BPU], angle Hue pulp [HPU], chromaticity pulp [CPU], peel firmness [PEF], pulp firmness [PUF], pH [PH], titratable acidity [TA], soluble solids [SS], ratio [RAT].
Discussion

When searching for comparative trends in the evolution of oviposition behaviour, one point of focus for any phytophagous insect is a possible linkage between the preference for host oviposition sites and the performance of larvae in the host (Diaz-Fleischer et al. 2000). Guava volatiles positively affect male sexual performance of *A. fraterculus* (Bachmann et al. 2015) and stimulate oviposition. In the present study, the forced *A. fraterculus* infestation provided similar quantities of pupae per fruit, showing adequate substrate (pulp) for larval development.

Adult emergence above 85% supports the close relationship between *A. fraterculus* and fruits of the Myrtaceae family, especially guavas (Sugayama et al. 1998). The correlation between infestation indices and pupal viability possibly is related to the available endosymbionts during the larval development of *A. fraterculus* (Selivon et al. 1996, 2002; Noman et al. 2020) and their beneficial nutritional function for immatures.

Fruit weight had an incremental effect on tephritid infestation rates of *Rhagoletis completa* Cresson (Tephritidae) in walnuts (Guillén et al. 2011) and *Bactrocera oleae* (Gmelin) in olives (Garantonakis et al. 2016). In our study, infestation differences between guava varieties when considering weight was due to “Kumagai” guava being less heavy than “Tailandesa” guava. “Kumagai” guavas produced on average 174% more pupae per kg. Higher pupal

| Parameter               | Red pulp | White pulp |
|-------------------------|----------|------------|
| Fruit weight            | 0.357a   | 0.155b     |
| L skin                  | 65.29a   | 65.93a     |
| a skin                  | -10.97a  | -12.40a    |
| b skin                  | 45.29a   | 42.71a     |
| Hue skin                | 104.55a  | 106.28a    |
| Chroma skin             | 44.04a   | 44.53a     |
| L pulp                  | 69.33b   | 83.76a     |
| a pulp                  | 20.61a   | -3.01b     |
| b pulp                  | 20.87b   | 24.27a     |
| Hue pulp                | 225.83a  | 96.69b     |
| Chroma pulp             | 29.80a   | 24.50b     |
| Peel firmness           | 60.07b   | 85.16a     |
| Pulp firmness           | 55.22b   | 74.19a     |
| pH                      | 3.94a    | 3.89a      |
| titratable acidity (TA) | 0.58a    | 0.62a      |
| soluble solid content (SS) | 7.72b  | 9.06a      |
| SS/TA (ratio)           | 13.39a   | 14.80a     |

Significant differences within a row are indicated by different letters Student’s t-test (P < 0.05).
weights of *A. fraterculus* were obtained from higher fruit weights (“Tailandesa”), although the fruit mass did not affect the pupal viability.

Rattanapun *et al.* (2009) found that ripe and fully-ripe mangoes were more suitable for larval development, with higher larval survival and shorter larval development times for *Bactrocera dorsalis* (Hendel). In our case, “Kumagai” mature guava produced on average approximately 1 pupa of *A. fraterculus* per gram of fruit, although both varieties stimulated a similar oviposition rate per fruit.

The performance of fruit species depends on the nutritional content of the host, but many tephritids prefer ripe and fully-ripe fruit for oviposition because they have adequate components for larval development and exhibit low content of secondary metabolites (Birke & Aluja 2018). In an earlier study, Raga *et al.* (2020) obtained up to 202 pupae of *A. fraterculus* per unbagged, ripe “Tailandesa” guava in the laboratory. No other fruit host species of *A. fraterculus* has shown such a high level of infestation under the same conditions.

The dimension of guava infestation range (both indices) shows the potential impacts in terms of yield losses, the spread of immature stages during the commercialisation process and quarantine risks during domestic and international trade. Added to this are the qualitative losses during pre- and post-harvesting processes (Louzeiro *et al.* 2020). The potential infestation in guavas and pupal viability of *A. fraterculus* are indicator variables of the risk of possible establishment and dispersal of pests (Heather & Halmann 2008).

An investigator might determine which of the same range of hosts supported larval development in those species (Diaz-Fleischer *et al.* 2000). Fifteen species of *Anastrepha* plus *Ceratitis capitata* (Wied.) infest *P. guajava* in Brazil (Zuchi & Moraes 2008). However, in the state of São Paulo, *A. fraterculus* is dominant (Raga *et al.* 2005), although co-infestation may occur in guavas (Raga *et al.* 2006).

Multivariate analysis of our results showed that pulp and peel firmness, soluble solids, titratable acidity, and SS/TA ratio are associated with a higher number of pupae of *A. fraterculus* per mass of “Kumagai” guavas. Oliveira *et al.* (2014) found that the highest rates of infestation of *A. fraterculus* in some guava varieties were correlated with the highest values of soluble solids and observed that the pH index had no direct relation to *A. fraterculus*’s attraction to and infestation of the guava fruits. Firmness is an important factor for *A. ludens* (Diaz-Fleischer & Aluja 2003). Mature guava is the best medium for larval development and therefore performance testing (Cunningham *et al.* 2016). This is because immediately after harvesting, the peel and pulp maintain their firmness, thus stimulating fruit fly oviposition. The same was observed for papaya infested by *C. capitata* (Joachim-Bravo *et al.* 2001) and mangoes infested by *B. dorsalis* (Rattanapun *et al.* 2009).

No significant effects on egg and pupal performance of *C. capitata* were obtained from insects reared in different citrus varieties; however, the fruits did affect larvae and pupal weights (Papachristos *et al.* 2008). The insect performance varied according to the suitable host, the region and climatic conditions (Medeiros *et al.* 2007; Bonebrake *et al.* 2010). We consider *P. guajava* as the primary host of *A. fraterculus* in southeastern Brazil, from which large populations spread to other fruit crops, such as citrus and stone fruits (Raga *et al.* 2002, 2017). Under field conditions, guavas were susceptible to *A. fraterculus* attack from when the fruits were undeveloped to when they began to ripen (Birke *et al.* 2015). Future field studies should be developed to determine the guava fruit stage preference and the percent degree of symptomatic and asymptomatic ripe fruits during the harvesting process to guarantee fruit health and quarantine safety.

**Acknowledgments**

The tested fruits were kindly supplied by Mr. Luiz Kumagai (Campinas, SP, Brazil).
Literature Cited

Akol, A.M., Masembe, C., Isabirye, B.E., Kukiriza, C.K. and Rwomushana, I. (2013) Oviposition preference and offspring performance in phytophagous fruit flies (Diptera: Tephritidae): the African invader, Bactrocera invadens. *International Research Journal of Horticulture*, 1(1): 1-14.

Aluja, M., Birke, A., Guilien, L., Diaz-Fleischer, F. and Nestel, D. (2011) Coping with an unpredicand stressful environment: the life history and metabolic response to variable food and host availability in a polyphagous tephritid fly. *Journal of Insect Physiology*, 57: 1592-1601.

Bachmann, G.E., Segura, D.F., Devescovi, F., Juárez, M.L., Ruiz, M.J., Vera, M.T., Cladera, J.L., Teal, P.E.A. and Fernández, P.C. (2015) Male sexual behavior and pheromone emission is enhanced by exposure to guava fruit volatiles in *Anastrepha fraterculus*. *PLoS One*, 10(4): e0124250. https://dx.doi.org/10.1371/journal.pone.0124250

Birke, A., Acosta, E. and Aluja, M. (2015) Limits to the host range of the highly polyphagous tephritid fruit fly *Anastrepha ludens* in its natural habitat. *Bulletin of Entomological Research*, 105: 743-753.

Birke, A. and Aluja, M. (2018) Do mothers really know best? Complexities in testing the preference-performance hypothesis in polyphagous frugivorous fruit flies. *Bulletin of Entomological Research*, 108: 674-684. https://dx.doi.org/10.1017/S0007485317001213

Bonebrake, T.C., Boggs, C.L., McNally, J.M., Ranganathan, J. and Ehrlich, P.R. (2010) Oviposition behavior and offspring performance in herbivorous insects: consequences of climatic and habitat heterogeneity. *Oikos*, 119: 927-934.

Cunningham, J.P., Carlsson, M.A., Villa, T.F., Dekker, T. and Clarke, A.R. (2016) Do fruit ripening volatiles enable resource specialism in polyphagous fruit flies? *Journal of Chemical Ecology*, 42: 931-940.

Díaz-Fleischer, F., Papaj, D.R., Prokopy, R.J., Aluja, M. and Norrbom, A.L. (2000) Evolution of fruit fly oviposition behavior. In: Aluja, M. and Norrbom, A.L. (eds). *Fruit flies (Tephritidae): phylogeny and evolution of behavior*. Boca Raton: CRC Press, pp. 811-841.

Díaz-Fleischer, F. and Aluja, M. (2003) Clutch size in frugivorous insects as a function of host firmness: the case of the tephritid fly *Anastrepha ludens*. *Ecological Entomology*, 28: 268-277.

Ferreira, D.F. (2019) Sisvar: A computer analysis system to fixed effects split plot type designs. *Revista Brasileira de Biometria*, 37(4): 529-535.

Garantonakis, N., Varikou, K., Markakis, E., Birouraki, A., Sergentani, C., Psarras, G. and Koubouris, G.C. (2016) Interaction between *Bactrocera oleae* (Diptera: Tephritidae) infestation and fruit mineral element content in *Olea europaea* (Lamiales: Oleaceae) cultivars of global interest. *Applied Entomology and Zoology*, 51: 257-265.

Gould, W.P. and Raga, A. (2002) Pests of guava. *In: Peña, J.E., Sharp, J.L. and Wysoki, M.* (eds.). *Tropical Fruit Pests and Pollinators - Biology, Economic Importance, Natural Enemies and Control*. Wallingford: CAB. p. 295-313.

Guillen, L., Aluja, M., Rull, J., Höhn, H., Schwizer, T. and Samietz, J. (2011) Influence of walnut cultivar on infestation by *Rhagoletis completa*: behavioural and management implications. *Entomologia Experimentalis et Applicata*, 140: 207-217.

Hafsi, A., Facon, B., Ravigné, V.F.C., Quilici, S., Chermiti, B. and Duyck, P. (2016) Host plant range of a fruit fly community (Diptera: Tephritidae): does fruit composition influence larval performance?. *BMC Ecology*, 16(40): 1-12. https://dx.doi.org/10.1186/s12898-016-0094-8

IBGE (2019)Produção Agrícola Municipal 2018. Rio de Janeiro: IBGE, 2019.

Heather, N.W. and Hallman, G.J. (2008) Pest management and phytosanitary trade barriers. Wallingford, UK: CABI. 257 pp.
Raga et al.: Performance of *Anastrepha fraterculus* on fruits of guava under forced infestation.

Joachim-Bravo, I.S., Fernandes, O.A., De Bortoli, S.A. and Zucoloto, F.S. (2001) Oviposition behavior of *Ceratitis capitata* Wiedemann (Diptera: Tephritidae): association between oviposition preference and larval performance in individual females. *Neotropical Entomology*, 30: 559-564.

Kogan, M. and Hilton, R.J. (2009) Conceptual Framework for Integrated Pest Management (IPM) of Tree-Fruit Pests. *In*: Aluja, M., Leskey, T.C., and Vincent, C. (eds.). *Biological tree fruit pest management*. Oxfordshire: Commonwealth Agricultural Bureau International, UK, 2009, p. 1-31.

Louzeiro, L.R.F., Souza-Filho, M.F., Raga, A. and Schmidt, F.L. (2020) Relationship between fruit fly (Diptera: Tephritidae) infestation and the physicochemical changes in fresh fruits. *African Journal of Agricultural Research*, 15: 122-133.

Medeiros, A., Oliveira, L. and Garcia, P. (2007) Suitability as medfly *Ceratitis capitata* (Diptera, Tephritidae) hosts, of seven fruit species growing on the island of São Miguel, Azores. *Arquipélago. Ciências Biológicas e Marinas*, 24: 33-40.

Mitra, S.K. and Irenaeus, K.S.T. (2018) Guava cultivars of the world. *Acta Horticulturae*, 1205: 905-910.

Noman, M.S., Liu, L., Bai, Z. and Li, Z. (2020) Tephritidae bacterial symbionts: potentials for pest management. *Bulletin of Entomological Research*, 110: 1-14.

Oliveira, F.Q., Boiça Junior, A.B., Costa, M.L.Z., Sanches, P.A.S., Walder, J.M.M. and Batista, J.L. (2014) Preference and infestation level of *Anastrepha fraterculus* (Wiedemann, 1830) on fruits of some *Psidium guajava* L. cultivars and relation to their physicochemical parameters. *Phytoparasitica*, 42: 475-483.

Papachristos, D.P., Papadopoulos, N.T. and Nanos, G.D. (2008) Survival and development of immature stages of the Mediterranean fruit fly (Diptera: Tephritidae) in citrus fruit. *Journal of Economic Entomology*, 101(3): 866-872. https://dx.doi.org/10.1093/jee/101.3.866

R Core Team (2019) R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing. Retrieved from: https://www.R-project.org

Raga, A., Prestes, D.A.O., Souza Filho, M.F., Sato, M.E., Siloto, R.C. and Zucchi, R.A. (2002) Occurrence of fruit flies in coffee varieties in the state of São Paulo, Brazil. *Boletin Sanidad Vegetal y Plagas*, 28: 519-524.

Raga, A., Machado, R.A., Souza Filho, M.F., Sato, M.E. and Siloto, R.C. (2005) Tephritoidea (Diptera) species from Myrtaceae fruits in the State of São Paulo, Brazil. *Entomotropica*, 20: 11-14.

Raga, A., Souza Filho, M.F., Prestes, D.A.O., Azevedo Filho, J.A. and Sato, M.E. (2006) Susceptibility of guava genotypes to natural infestation by *Anastrepha* spp. (Diptera: Tephritidae) in the municipality of Monte Alegre do Sul, state of São Paulo, Brazil. *Neotropical Entomology*, 35: 121-125.

Raga, A. and Sato, M.E. (2011) Toxicity of neonicotinoids to *Ceratitis capitata* and *Anastrepha fraterculus* (Diptera: Tephritidae). *Journal of Plant Protection Research*, 51: 413-419.

Raga, A., Paula, L., Souza-Filho, M.F. and Castro, J.L. (2017) Population dynamics and infestation rate of fruit flies in stone fruits in São Paulo State, Brazil. *Annual Research & Review in Biology, 14*(1): 1-11.

Raga, A., Sousa, E.M., Silva, S.B. and Louzeiro, L.R.F. (2020) Susceptibility of bagged guavas to the attack of fruit flies (Tephritidae). *Journal of Experimental Agriculture International*, 42: 82-87.

Rattanapun, W., Amornsak, W. and Clarke, A.R. (2009) *Bactrocera dorsalis* preference for and performance on two mango varieties at three stages of ripeness. *Entomologia Experimentalis et Applicata*, 131: 243-253.

Selivon, D., Morgante, J.S., Ribeiro, A.F. and Perondini, A.L.P. (1996) Extrusion of masses of yolk during embryonic development of the fruit fly *Anastrepha fraterculus*. *Invertebrate Reproduction & Development*, 29: 1-17.
Selivon, D., Perondini, A.L.P., Ribeiro, A.F., Marino, C.L., Lima, M.M.A. and Coscrato, V.E. (2002) *Wolbachia* endosymbiont in a species of the *Anastrepha fraterculus* complex (Diptera: Tephritidae). *Invertebrate Reproduction & Development, 42*(2-3): 121-127. https://dx.doi.org/10.1080/07924259.2002.9652768

Singh, S.P. (2011) Guava (*Psidium guajava* L.). pp. 213-246e. In: Elhadi, Y. (ed.). *Postharvest Biology and Technology of Tropical and Subtropical Fruits*. https://dx.doi.org/10.1533/9780857092885.213

Souza-Filho, M.F., Raga, A., Azevedo Filho, J.A., Strikis, P.C., Guimarães, J.A. and Zucchi, R.A. (2009) Diversity and seasonality of fruit flies (Diptera: Tephritidae and Lonchaeidae) and their parasitoids (Hymenoptera: Braconidae and Figitidae) in orchards of guava, loquat and peach. *Brazilian Journal of Biology, 69*: 31-40.

Sugayama, R.L., Kovaleski, A., Liedo, P. and Malavasi, A. (1998) Colonization of a new fruit crop by *Anastrepha fraterculus* (Diptera: Tephritidae) in Brazil: a demographic analysis. *Environmental Entomology, 27*(3): 642-648.

Zucchi, R.A. and Moraes, R.C.B. (2008) Fruit flies in Brazil - *Anastrepha* species their host plants and parasitoids. Retrieved from: www.lea.esalq.usp.br/anastrepha (accessed 10 August 2020)