Screening for insecticidal efficacy of two Algerian essential oils with special concern to their impact on biological parameters of *Ephestia kuehniella* (Zeller) (Lepidoptera: Pyralidae)

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**Abstract**

Chemical composition of Algerian *Mentha rotundifolia* and *Myrtus communis* essential oils, their insecticidal activities and their impact on some biological parameters of the Mediterranean flour moth *Ephestia kuehniella* were assessed. Results showed that *M. rotundifolia* essential oil contained piperitenone oxide (46.06%), d-limonene (9.10%), cis-piperitone oxide (6.81%), and endo-borneol (4.64%) as major compounds, while *M. communis* oil was rich in α-pinene (29.08%), 1,8-cineole (36.82%), α-terpineol (6.42%), geranyl acetate (4.38%), and β-linalool (4.04%). The fumigant potential and contact toxicity tests against *E. kuehniella* demonstrated the effectiveness of *M. rotundifolia* oil (LC50 = 0.54 μL/L air, LC50 = 0.004 μL/cm²) compared to *M. communis* oil (LC50 = 2.91 μL/L air, LC50 = 0.025 μL/cm²). Moreover, results revealed that all biological parameters were significantly affected (fecundity: 6 eggs/female, oviposition deterrence: 96.62%, log fertility: 0, hatching rate: 0%, copulation rate: 0% for *M. rotundifolia* oil against fecundity: 93 eggs/female, percentage of oviposition deterrence: 47.85%, log fertility: 6.7, hatching rate: 57%, copulation rate: 53.33% for *M. communis* oil). This work supports the use of botanical insecticide as active pest control agents under storage conditions.

**Keywords** *Ephestia kuehniella* · Essential oil · Round-leaved mint · Common myrtle · Copulation rate

**Introduction**

Agricultural products are often subject to biotic and abiotic damage during production and conservation. Contamination of food commodities by insect pests is an important quality control issue of concern for the food industries (Rajendran and Sriranjini 2008). About 35% on the field and 14% in storage (around 50% in total) crops are lost annually because of insect pests, which adversely affected the world food production during crop growth, harvest and storage (Jitendra et al. 2009). Among damaging insects, the Mediterranean flour moth *Ephestia kuehniella* Zeller (Lepidoptera: Pyralidae) is one of the major pests in industrial flour mills (Jacob and Cox 1977), in Algeria (Hami et al. 2005) and Tunisia (Jarraya 2003). It is a worldwide pest that causes major economic losses in the Mediterranean basin and near East regions (Al-Izzi et al. 1985). Its main habitats are flour and grout mills, corn milling plants, bakeries and any other place used for processing grains or preparing flour products (Ben Achour et al. 2008). Larvae of *E. kuehniella* reduce both quantitative and qualitative quality of the products by feeding, by their presence and by the production of feces and webbing (Johnson et al. 1997). Last years, the overuse of synthetic insecticides and fumigants such as phosphine for grain storage has resulted in a number of problems, including the development of insecticide resistance among insect pests of stored grains (Sousa et al. 2009). Currently, stored-product pest control...
strategies tend to emphasize the non-chemical aspects of pest control with the judicious use of pesticides (Titouhi et al. 2017). In this respect, plant extracts are safe, eco-friendly and more compatible with environmental components compared to synthetic pesticides and so ranked under Green pesticides category (Rahman et al. 2016).

Besides, Kim et al. (2003) showed that plant extracts are often active against a limited number of target insects, are biodegradable, and are potentially suitable for use in integrated pest management. These authors reported also that plant extracts could lead to the improvement in new classes of nontoxic insect control. Plant essential oils have been recently qualified as ecological alternatives to chemical pesticides due to their multifunctional efficacy including anti-insect pest activity (Isman and Grieneisen 2014; Mediouni Ben Jemâa 2014). Furthermore, essential oils possess important contact and fumigant toxicity (Liu and Ho 1999; Abdelgaleil et al. 2009; Kasrati et al. 2015; Quan et al. 2018), antifeedant activity (Huang et al. 1997; Ebadollahi 2013), repellent activity (Hori 2003; Prajapati et al. 2005; Nerio et al. 2009; Chang et al. 2017) as well as development and growth inhibitory activity (Waliwitiya et al. 2009; Sangha et al. 2017) against various insect species and are regarded as environmentally compatible pesticides (Cetin et al. 2004).

Their lipophilic nature facilitates their interference with basic metabolic, biochemical, physiological and behavioral functions of insects (Nishimura 2001). Commonly, essential oils with anti-insect pest ability are known to possess insecticidal and repellent effects by neurotoxic mode of action (Kostyukovsky et al. 2002). Mentha rotundifolia (L.) huds (Lamiaceae) and Myrtus communis (L.) (Myrtales: Myrtaceae) are commonly known under the vernacular names of round leaf and common myrtle. These aromatic plant species, grow wild in northern Algeria, are widely used by the population in traditional phytotherapy for their therapeutic virtues. The essential oils extracted from the leaves of Myrtus communis have shown insecticidal activity against E. kuehniella, Plodia interpunctella, Acanthoscelides obtectus (Ayvaz et al. 2010), Tribulium castaneum (Senfi et al. 2013), Callosobruchus maculatus (Khani and Farzaneh 2012), and Trogoderma granarium (Tayoub et al. 2012). Few studies have reported the insecticidal potential of M. rotundifolia essential oil on stored-product pests (El Arch et al. 2003; Brahmi et al. 2016). Nevertheless, no study has been reported before on the insecticidal activity of M. rotundifolia against E. kuehniella adults.

This work aimed to assess the fumigant and contact toxicity of M. rotundifolia and M. communis essential oils from Algeria as well as their effects on longevity, fecundity, fertility, copulation rate and hatching rate against new emerged adults of a Tunisian strain of the Mediterranean flour moth E. kuehniella.

Materials and methods

Insect rearing

A rearing colony of E. kuehniella was maintained on wheat flour and semolina under laboratory-controlled condition (temperature of 25 °C ± 1 °C, a relative humidity of 65 ± 5% and darkness). Insects were maintained in 2-L plastic storage boxes. Unsexed adults aged between 24 and 48 h were used for the bioassays.

Plant material

Myrtus communis aerial parts samples were collected during October 2017 from the Edough Massif (North-East Algeria-Annaba: 36° 55′ N, 7° 36′ E), while M. rotundifolia aerial parts were sampled during August 2018 from the locality of Berrahal (North-East of Algeria-Annaba: 36° 50′ N, 7° 27′ E).

Extraction of the essential oils

100 g of dry leaves of each plant was hydrodistilled during 90 min using a Clevenger apparatus. The crude-extracted essential oils were stored in opaque flasks in a refrigerator at 4 °C. Essential oils’ yields obtained from each plant species were evaluated according to AFNOR (1986) formula: oil% (w/w) = weight of essential oil (g)/weight of plant material (g) × 100.

Gas chromatography–mass spectrometry (GC/MS) analysis

The analysis was carried out using an Agilent 7890A gas chromatograph combined to an Agilent 5972C mass spectrometer with electron impact ionization (70 eV). The mass spectrometer was equipped with a capillary column HP-5 MS (19091S-433), length 30 m, diameter 250 μm, and 2.5 μm film thicknesses (5% phenyl methyl silicone, 95% dimethylpolysiloxane; Hewlett-Packard, CA, USA). The column temperature was automated to rise from 50 to 250 °C at a rate of 7 °C/min. The transfer line temperature was 250 °C. The flow rate of helium (carrier gas) was 1 mL/min. A sample of 2 μL was manually injected with a constant pressure of 7.65 psi using split mode (split ratio 50:1). Scan time and mass range were 1 s and 45–400 m/z, respectively. The essential oils components were identified by comparing their retention indices (RIs) relative to n-alkanes with those of authentic compounds published in the literature or available in our laboratory. Furthermore, the identification was confirmed by matching...
their mass spectra with those recorded in Wiley Registry 9th Edition/NIST 2011 edition mass spectral library. The composition of the essential oils was stated as a relative percentage of total peak area.

**Bioassays**

**Fumigant toxicity**

The insecticidal activity of *M. rotundifolia* and *M. communis* essential oils was evaluated by the fumigant test according to Titouhi et al. (2017). Ten adults of *E. kuehniella* were placed in Plexiglas bottles of 38 mL volume. The bottom surface of the screw caps was lined with Whatman n°1 paper disks (2 cm diameter with a 3 cm length fixing tab). Using a micropipette, a series of essential oils doses (without the use of any solvent) were deposited on the filter paper disks. Doses used in the essay were 0.05, 0.125, and 0.5 μL giving equivalent concentrations of 1.31, 3.28, and 13.16 μL/L air. The caps were quickly screwed tightly onto the bottles. Three repetitions were carried out for each concentration of each essential oil including a negative control without essential oil and maintained under the same conditions described for breeding. Mortality was assessed by direct observation of insects every hour until total death. Insects are considered dead when no leg or antenna movements have been observed. The LC50, LC95, and LT50 values were calculated by using Probit analysis (IBM SPSS V22.0) was used to estimate lethal concentrations LC50 and LC95 values and lethal time LT50.

**Contact toxicity**

The contact bioassay of *M. communis* and *M. rotundifolia* essential oils against *E. kuehniella* adults was done according to Mami Maazoun et al. (2017). A Whatman filter paper disks, having 9 cm of diameter, were treated with three doses 0.05, 0.125, and 0.5 μL dissolved in acetone to give concentrations equivalent to 9 × 10−4, 24 × 10−4, and 99 × 10−4 μL/cm². Acetone was used as a negative control. The solvent was allowed to evaporate completely at room temperature for 5 min. Each filter paper disk was then placed in a glass Petri dish, and 10 adults of *E. kuehniella* were placed into the center of each Petri dish. Each hour, the number of dead insects was recorded. Tests were done in triplicate. The percentage of mortality was calculated using Abbott’s correction formula (1925). Insects are considered dead when no leg or antenna movements have been observed. The LC50, LC95, and LT50 values were calculated by using Probit analysis (IBM SPSS V20.0).

**Impact on biological parameters**

The insecticidal effects of *M. communis* and *M. rotundifolia* essential oils through fumigation were studied by assessing their impact on longevity, fecundity, fertility, hatching rate, percentage of oviposition deterrence, and copulation rate of *E. kuehniella*. Five pairs of unsexed adults (0–24 h old) were exposed at the lethal concentration LC15. The longevity of males and females, the total number of eggs laid per female (fecundity), the percentage of oviposition deterrence according to Pascual Villalobos and Robledo (1998), fertility, and the percentage of egg hatching were determined. Non-exposed adults kept under the same conditions served as a control. Experiments were replicated three times. For the copulation rate study, prior to exposure, virgin females were obtained by rearing last instar larvae of *E. kuehniella* separately in plastic boxes. Female’s bursa copulatrix were dissected out under a binocular microscope in a saline solution (10% NaCl) and observed for the presence of spermatoophores as a criterion of successful mating.

**Data analysis**

For each biological parameter (longevity, percentage of oviposition deterrence, fecundity, fertility, hatching rate, and copulation rate of *E. kuehniella*), data were subjected to two-way ANOVA, with essential oils and exposure time as main fixed factors plus with essential oils * exposure time interaction term. Differences in values of each essential oil and exposure time were tested by one-way ANOVA followed by Duncan test to detect significant differences in means at 0.05 percent level. In some cases, certain data have been transformed into a common logarithm or square root in order to obtain a normal distribution of the variables. All data values represented the mean of three replications and were expressed as the mean ± standard deviation. All statistical analyses were achieved using SPSS statistical software version 22.0.

**Results**

**Essential oil composition**

Oil yields based on leaf dry matter weight were, respectively, 1.29% and 0.64% for *M. rotundifolia* and *M. communis*. GC and GC/MS analysis of *M. rotundifolia* and *M. communis* essential oils are reported in Table 1. Results indicated that *M. rotundifolia* essential oil contained 30 identified compounds constituting approximately 95.51% of the total content. The essential oil profile was characterized by piperitenone oxide (46.06%), α-limonene (9.10%), cis-piperitone...
Table 1 Essential oil composition (%) of Mentha rotundifolia and Myrtus communis leaves collected from Algeria

| No. | Compounds                | RI   | M. rotundifolia | M. communis |
|-----|--------------------------|------|-----------------|-------------|
| 1   | α-Pinene                 | 939  | 2.61            | 29.08       |
| 2   | Camphene                 | 954  | 1.69            | 1.91        |
| 3   | Sabinene                 | 976  | 0.91            | –           |
| 4   | Vinyl amyl carbinol      | 978  | 1.44            | –           |
| 5   | β-Pinene                 | 980  | 2.04            | 0.77        |
| 6   | β-Myrcene                | 991  | 1.39            | –           |
| 7   | β-Ocimene                | 995  | 0.27            | –           |
| 8   | Δ-3-Carene               | 1011 | –               | 0.64        |
| 9   | p-Cimene                 | 1023 | –               | 1.52        |
| 10  | α-Limonene               | 1028 | 9.10            | –           |
| 11  | 1,8-Cineole              | 1033 | 0.45            | 36.82       |
| 12  | γ-Terpinene              | 1053 | –               | 0.53        |
| 13  | α-Terpinolene            | 1089 | –               | 0.47        |
| 14  | β-Linalool               | 1098 | –               | 4.04        |
| 15  | Amyl vinyl carbinyl      | 1110 | 0.67            | –           |
| 16  | trans-Pinocarveol        | 1138 | –               | 1.04        |
| 17  | Camphor                  | 1143 | 0.57            | 0.46        |
| 18  | (R)-Lavandulol           | 1148 | –               | –           |
| 19  | Endo-borneol             | 1165 | 4.64            | –           |
| 20  | Terpinen-4-ol            | 1178 | 0.35            | 1.09        |
| 21  | α-Terpineol              | 1189 | 0.82            | 6.42        |
| 22  | 5-acetyl-2-hydrazino-4   | 1211 | 1.85            | –           |
|     | methylpyrimidine         |      |                 |             |
| 23  | trans-Carveol            | 1230 | –               | 0.33        |
| 24  | Geraniol                 | 1255 | –               | 1.15        |
| 25  | Linalyl acetate          | 1257 | –               | 0.51        |
| 26  | cis-Piperitone oxide     | 1261 | 6.81            | –           |
| 27  | Bornyl acetate           | 1285 | 0.52            | –           |
| 28  | m-Thymol                 | 1290 | 0.37            | –           |
| 29  | Piperitone oxide         | 1376 | 46.06           | –           |
| 30  | Geranyl acetate          | 1383 | –               | 4.38        |
| 31  | cis-Jasmone              | 1394 | 2.47            | –           |
| 32  | Methyl eugenol           | 1401 | –               | 2.59        |
| 33  | Caryophyllene            | 1420 | 3.18            | 0.42        |
| 34  | α-Humulene               | 1454 | 0.42            | –           |
| 35  | trans-β-farnesene        | 1458 | 0.87            | –           |
| 36  | α-Amorphene              | 1475 | 0.34            | –           |
| 37  | Germacrene D             | 1485 | 3.58            | –           |
| 38  | γ-Cadinene               | 1513 | 0.46            | –           |
| 39  | 1S,cis-calamenene        | 1519 | 0.50            | –           |
| 40  | Caryophyllene oxide      | 1581 | –               | 0.96        |
| 41  | Verdiflorol              | 1592 | 0.47            | –           |
| 42  | α-Cadinol                | 1643 | 0.40            | –           |
| Total|                         |      | 95.51           | 95.13       |

–, compound not detected; RI, retention index calculated on a HP-5MS capillary column (30 m × 0.25 mm × 0.25 mm)

Concerning the essential oil of M. communis, 20 compounds representing 95.13% of the total oil composition were identified. The major components were α-pinene (29.08%), 1,8-cineole (36.82%), α-terpinolene (6.42%), geranyl acetate (4.38%), and β-linalool (4.04%).

**Essential oil fumigant toxicity**

Fumigant toxicity results are presented in Fig. 1 as a percentage of mortality of E. kuehniella adult’s strains from Tunisia exposed to M. rotundifolia and M. communis essential oils collected from Algeria. Data revealed that fumigant toxicity depended significantly on plant species (\(F_{1,36} = 33.02, p < 0.01\)), concentrations (\(F_{2,36} = 20.01, p < 0.01\)), and exposure time (\(F_{1,36} = 4.82, p < 0.01\)). The mortality values increased depending on the increasing essential oil concentrations. Furthermore, M. rotundifolia essential oil was more toxic than M. communis. In fact, for the lowest concentration (1.31 μL/L air), percentage mortality of E. kuehniella after 48 h attained, respectively, 100% and 41.66% with M. rotundifolia and M. communis essential oils. For the highest concentration (13.16 μL/L air), M. rotundifolia and M. communis essential oils caused 100% mortality after, respectively, 18 and 24 h of exposure. Moreover, essential oil of M. rotundifolia caused significantly the highest adulticidal effects at all tested concentrations. Moreover, results indicated that M. rotundifolia essential oil showed 100% effectiveness at 3.28 μL/L air after only 18 h of treatment.

Probit analyses have also confirmed that E. kuehniella was more susceptible to M. rotundifolia oil (Table 2). Additionally, the LC\(_{50}\) and LC\(_{95}\) values proved that M. rotundifolia essential oil was more toxic than M. communis. The LC\(_{50}\) values reached 0.54 μL/L air for M. rotundifolia and 2.91 μL/L air for M. communis. The LC\(_{95}\) was evaluated to be, respectively, 2.11 μL/L air and 5.29 μL/L air for M. rotundifolia and M. communis essential oils. Furthermore, the LT\(_{50}\) values ranged between 1.03 and 20.76 h for M. rotundifolia and from 20.19 h to 105.09 h for M. communis (Table 3).

**Essential oil contact toxicity**

Fumigant toxicity data are illustrated in Fig. 2 as percentage of mortality of E. kuehniella adults. Statistical analysis revealed significant differences in percentage of mortality as a function of essential oils treatments (\(F_{2,12} = 7.58, p < 0.01\)). Results indicated the sensitivity of E. kuehniella adults to both essential oils.
Fig. 1 Mortality (%) of *Ephestia kuehniella* adults exposed for various periods of time and various concentrations to *Mentha rotundifolia* and *Myrtus communis* essential oils

Table 2 Fumigant toxicity of *Mentha rotundifolia* and *Myrtus communis* essential oils against *Ephestia kuehniella*

| Essential oils     | LC$_{50}^{(a,b)}$ (μL/L air) | LC$_{95}^{(a,b)}$ (μL/L air) | $\chi^2$ | Slope ± SE | Sig     | Degrees of freedom |
|--------------------|-------------------------------|-------------------------------|------|-----------|-------|-------------------|
| *M. rotundifolia*   | 0.54 (0.05–0.94)              | 2.11 (1.74–3.95)              | 0.26 | 1.05 ± 0.38 | 0.56  | 1                 |
| *M. communis*       | 2.91 (2.63–4.84)              | 5.29 (4.61–6.46)              | 0.01 | 0.67 ± 0.10 | 1.00  | 1                 |

*Units LC$_{50}$ and LC$_{95}$ = ml/l air. Applied for 24 h at 25 °C*

Table 3 LT$_{50}$ values of *Mentha rotundifolia* and *Myrtus communis* essential oils against *Ephestia kuehniella*

| Essential oils     | Concentration (μL/L air) | LT$_{50}^{(a,b)}$ (h) | $\chi^2$ | Slope ± SE | Sig     | Degrees of freedom |
|--------------------|--------------------------|-----------------------|------|-----------|-------|-------------------|
| *M. rotundifolia*   | 1.31                     | 20.76 (9.80–29.16)    | 5.87 | 0.10 ± 0.01 | 0.31  | 5                 |
|                    | 3.28                     | 5.30 (1.6–17.6)       | 3.52 | 0.37 ± 0.05 | 0.10  | 5                 |
|                    | 13.16                    | 1.03 (0.14–3.94)      | 4.04 | 0.53 ± 0.08 | 0.20  | 5                 |
| *M. communis*       | 1.31                     | 105.09 (76.42–173.91) | 5.66 | 0.013 ± 0.001 | 0.34  | 5                 |
|                    | 3.28                     | 22.41 (10.32–36.42)   | 25.12| 0.02 ± 0.002 | 0.00  | 5                 |
|                    | 13.16                    | 20.19 (17.35–22.77)   | 3.90 | 0.06 ± 0.005 | 1.00  | 5                 |

*Units LT$_{50}$ = h. Applied at 25 °C*

*95% lower and upper confidence limits are shown in parentheses*
concentrations with a high striking effect of *M. rotundifolia* essential oil. Indeed, the lowest concentration (9 × 10⁻⁴ μL/cm²) of *M. rotundifolia* oil induced 100% mortality after 72 h of exposure, while 80% mortality was achieved after 120 h of exposure to *M. communis* oil. 

Table 4 reports LC₅₀ and LC₉₅ values calculated for *M. rotundifolia* and *M. communis* essential oils against *E. kuehniella* adults. The median lethal concentration was highly dependent upon oils. Probit analysis established that *E. kuehniella* adults were more susceptible to *M. rotundifolia* oil.

Table 5 demonstrates that LT₅₀ values for *M. rotundifolia* ranged from 62 h for the lowest concentration (9 × 10⁻⁴ μL/cm²) to 26 h for the highest concentration (99 × 10⁻⁴ μL/cm²), while for *M. communis* oil LT₅₀ values were made among essential oils. Different letters are significantly different according to Duncan test at *p* ≤ 0.05.

### Table 4 Contact toxicity of *Mentha rotundifolia* and *Myrtus communis* essential oils against *Ephestia kuehniella*

| Essential oils | LC₅₀ (μL/cm²) | LC₉₅ (μL/cm²) | χ² | Slope ± SE | Sig | Degrees of freedom |
|----------------|--------------|--------------|----|-----------|-----|--------------------|
| *Mentha rotundifolia* | 0.004 (0.0028–0.0063) | 0.017 (0.012–0.027) | 6.95 | 128.74 ± 51.16 | 0.008 | 1 |
| *Myrtus communis* | 0.025 (0.016–0.041) | 0.053 (0.034–0.084) | 1.12 | 58.03 ± 57.65 | 0.289 | 1 |

*a* Units LC₅₀ and LC₉₅ = mL/L air. Applied for 24 h at 25 °C

### Table 5 LT₅₀ values of *Mentha rotundifolia* and *Myrtus communis* essential oils against *Ephestia kuehniella*

| Essential oils | Concentrations (10⁻⁴ μL/cm²) | LT₅₀ (a,b) (h) | χ² | Slope ± SE | Sig | Degrees of freedom |
|----------------|-----------------------------|----------------|----|-----------|-----|--------------------|
| *Mentha rotundifolia* | 9 | 61.73 (55.62–69.10) | 0.67 | 0.06 ± 0.014 | 0.99 | 6 |
| | 24 | 28.76 (20.13–35.67) | 7.55 | 0.05 ± 0.01 | 0.27 | 6 |
| | 99 | 26.58 (14.54–35.44) | 2.7 | 0.03 ± 0.008 | 0.84 | 6 |
| *Myrtus communis* | 9 | 85.84 (72.35–102.16) | 8.87 | 0.02 ± 0.003 | 0.18 | 6 |
| | 24 | 73.32 (64.02–86.05) | 5.43 | 0.03 ± 0.007 | 0.48 | 6 |
| | 99 | 48.45 (42.74–53.90) | 2.62 | 0.08 ± 0.01 | 0.85 | 6 |

*a* Units LT₅₀ = h. Applied at 25 °C

*b* 95% lower and upper confidence limits are shown in parentheses
varied between 86 and 48 h for the lowest and highest concentration, respectively.

**Biological parameters**

*Longevity*

Essential oils significantly reduced adults' longevity at LC$_{15}$ concentration (Fig. 3). Statistical analysis showed highly significant differences in longevity as a function of essential oils treatments for both sex [males (F$_{2,24} = 21.08$, p < 0.01), females (F$_{2,24} = 22.57$, p < 0.01)], time [males: F$_{3,24} = 16.07$, p < 0.01; females: F$_{3,24} = 23.42$, p < 0.01], and their interaction [males (F$_{6,24} = 2.85$, p < 0.01), females (F$_{6,24} = 3.57$, p < 0.05)]. Moreover, significant differences were observed in the longevity of males adults treated by *M. rotundifolia* and *M. communis* essential oils (F$_{1,22} = 5.56$, p < 0.05), whereas no significative differences were detected in the longevity of females (F$_{1,22} = 1.32$, p > 0.05).

**Fecundity, oviposition deterrence, and hatching rate**

According to the data presented in Fig. 4, the fecundity of females was highly dependent on essential oil and exposure time. In fact, the number of eggs laid increased over time, whether in the control or in *M. communis* essential oil treatment. Based on statistical analysis, there is significant differences in fecundity between control and treated females (F$_{2,6} = 412.81$, p < 0.01). The mean number of eggs bequeathed by the untreated females was 178 eggs compared to 93 eggs for the essential oil of *M. communis* and 6 eggs for *M. rotundifolia*. As it can be perceived in Fig. 4, the essential oils adversely affected the hatching rate (control: 89%, common myrtle: 57%, round-leaved mint: 0%) which decreased over time. Data showed that hatching rate declined significantly with *M. rotundifolia* essential oil treatment (F$_{1,4} = 148.23$, p < 0.01) contrary to *M. communis* treatment (F$_{1,4} = 3.73$, p > 0.05). Furthermore, results showed significant differences in percentage of oviposition deterrence as a function of essential oils treatment (F$_{2,6} = 21.02$, p < 0.01). *Mentha rotundifolia* showed the high percentage of oviposition deterrence 96.62% ± 1.87 against 47.85% ± 18.13 for *M. communis*.

**Fertility and copulation rate of *E. kuehniella* females**

Results of fertility and copulation rate of *E. kuehniella* females are illustrated in Table 6. Data showed that fertility and copulation rate significantly depended on essential
oil. Statistical analyses indicated a significant reduction in fertility between control and treated insects \((F_{2,6} = 14.90, p < 0.01)\). Significant differences were detected in the copulation rate among control and treatments \((F_{2,6} = 169, p < 0.01)\). In control, 100% of females had been fertilized by the males and had filled spermathecae. Almost 50% of the females who were treated with \(M. \text{communis}\) oil have been fertilized. Thus, the mating rate had been halved by the action of this oil. Mating rate in females treated with essential oil of \(M. \text{rotundifolia}\) was null (0%).

### Discussion

The present study showed that the essential oils of \(M. \text{rotundifolia}\) and \(M. \text{communis}\) collected from the northeast of Algeria are a promising source of phytochemicals. \(M. \text{rotundifolia}\) yielded 1.29% which is in accordance with the results reported for the same species in Tunisia 1.26%, (Riahi et al. 2013). Contrary, \(M. \text{rotundifolia}\) from other locations of Algeria have higher essential oil yields 1.6–1.8% (Brada et al. 2007) and 1.65% (Benabdallah et al. 2018) but lower than those reported in Morocco 1.54% (Derwich et al. 2009) and 4.33% (Derwich et al. 2010).

In our case, the yield of \(M. \text{communis}\) essential oil was 0.64%. This result was in agreement with the studies of Barhouchi et al. (2016) in Algeria (0.6%) and Aidi Wannes et al. (2007) in Tunisia (0.61%) but seemed to be higher than those found in Greece 0.28–0.3% (Gardeli et al. 2008) and Morocco 0.3–0.4% (Satrani et al. 2006).

In this research, GC/MS analysis revealed 30 compounds in essential oil of \(M. \text{rotundifolia}\), the major one being piperitone oxide or rotundifolone (46.06%). Preceding works pointed out that piperitone oxide was reported as main constituent of some chemotype of \(M. \text{rotundifolia}\) in diverse geographic areas (Brada et al. 2006, 2007; Benabdallah et al. 2018; Lorenzo et al. 2002; Gende et al. 2014; Bounihi 2016; Sumio 1956). However, Kokkini and Papageorgiou (1988) confirmed that piperitone oxide and menthyl acetate were also two chemotypes of Grecian populations. Moreover, Reitsema (1958) and Lawrence (2007) reported carvone as a principal constituent in \(M. \text{rotundifolia}\) oils. Similarly, El Arch et al. (2003) and Riahi et al. (2013) stated pulegone as a major compound, respectively, in Morocco and Tunisia. Other previous studies investigated that the chemical composition of \(M. \text{rotundifolia}\) revealed the presence of specific chemotypes such as trans-piperitone epoxide in Algeria (Brahmi et al. 2016), 2,4(8),6-p-menthatrien-2,3-diol and germacrene D in Cuba (Pino et al. 1999), and menthol in Morocco (Derwich et al. 2009). Many studies reported that piperitone oxide was also the main constituent of \(M. \text{suaveolens}\) (Oumzil et al. 2002; Amzouar et al. 2016; Maffei 1988; Baser et al. 1999), \(M. \text{longifolia}\) (Venskutonis 1996; Ghoulami 2001), and \(M. \text{villosa}\) (Sousa et al. 2009). In the present study, \(M. \text{communis}\) essential oil was characterized by 1,8-cineole (36.82%) and α-pinene (29.08%) as major compounds. Likewise, data showed a high level of 1,8-cineole which corroborates the findings of Bouzouita et al. (2003) and Viuda Martos et al. (2011). However, Ben Ghnaya et al. (2013) and Aidi Wannes et al. (2007) stated that two populations of Tunisian \(M. \text{communis}\) essential oils presented the pair α-pinene/1,8-cineole as major compounds with a prevalence for α-pinene. Moreover, Barhouchi et al. (2016) reported that α-pinene and 1,8-cineole represent together 88% of common myrtle essential oil from the Northeast of Algeria. Also, Messaoud et al. (2005) investigated twelve Tunisian natural populations of \(M. \text{communis}\) and detected that α-pinene and 1,8-cineole were the main volatile components. In the same way, it can be observed that Iranian, French, and Italian myrtle oil revealed the same main mixture: α-pinene, 1,8-cineole (Rasooli et al. 2002; Curini et al. 2003; Tuberoso et al. 2006). Chalchat et al. (1998) compared \(M. \text{communis}\) essential oils from different Mediterranean locations and reported that oils from Corsica and Tunisia presented a level of α-pinene above 50% and Morocco, Lebanon, Yugoslavia oils’ presented a level under 35%. In contrast, Spanish, Croatian, and Grecian common myrtle was characterized by the myrtenyl acetate chemotype (Boelens and Jimenez 1992; Jerkovic et al. 2002; Gardeli et al. 2008), whereas in the present study there was no trace of this compound. Otherwise, the ratio 1,8-cineole, myrtenyl acetate characterized the Moroccan oil (Satrani et al. 2006). Another study from Tizi Ouzou Algeria had shown that 1,8-cineole (47%) and cis-geraniol (25%) were the main volatile compounds (Djenoune et al. 2011).

As we can observe, there are several chemotypes of \(M. \text{communis}\) and \(M. \text{rotundifolia}\) essential oils over the world; this difference in the chemical composition is closely linked to several factors like adaptive metabolism of plant (Nikšić et al. 2014), geographical position, temperature, day length, nutrients, maturation stage, polyploidy (Scora 1973), air movement, rainfall (Boukhebi et al. 2011), harvest period, distillation method, and time of extraction (Rajendran and Sriranjini 2008).

### Table 6

|                  | Copulation rate (%) | Log fertility       |
|------------------|---------------------|---------------------|
| \(M. \text{rotundifolia}\) | 0.00 ± 0.00 \(^a\) | 0.00 ± 0.00 \(^a\) |
| \(M. \text{communis}\)    | 53.33 ± 6.66 \(^b\) | 4.04 ± 1.39 \(^b\) |
| Control           | 100.00 ± 0.00 \(^c\) | 6.69 ± 0.14 \(^c\) |

For each column, values followed by different letters are significantly different according to Duncan test at \(p \leq 0.01\).
Plant extracts have been traditionally used around the world to protect stored products, but the recent increased interest in essential oil was observed with the emergence of scientific evidences on their fumigant and contact insecticidal toxicity to a large range of pests (Isman 2000). In fact, essential oils represent excellent alternatives to chemicals because of their low mammalian toxicity, fast degradation, and local obtainability (Rajendran and Sriranjini 2008). Essential oils could be regarded among the new generation of chemicals that interrupt insects’ life cycle and thus seen as more target-oriented softer technique of control with less impact on the environment. The essential oils used in the current work showed a potent toxicity to *E. kuehniella* adults, especially *M. rotundifolia*. Our results demonstrated a high fumigant and contact toxicity and exceptional disruptive effects on the biological parameters of *E. kuehniella*. Brahmi et al. (2016) and El Arch et al. (2003) have reported the insecticidal activity of *M. rotundifolia* against pest beetles. Furthermore, context of Karabörkuli et al. (2011) evaluated the fumigant action of eight essential oils from Turkish aromatic plants including *M. communis* and demonstrated that this oil showed a moderate toxicity with LC50 value of 15.15 μL/L air. In another study, Ayvaz et al. (2010) testified the insecticidal activity (fumigant test) of *M. communis* (LC50 = 12.75 μL/L air). This is in contradiction with our results, since we demonstrate that myrtle oil was more efficient with a low lethal concentration (LC50 = 2.21 μL/L air). Mediouni Ben Jemâa et al. (2012) studied the impact of seasonal variations on the chemical composition and the fumigant bioassay of five *Eucalyptus* species essential oils against *E. kuehniella* and confirmed that all species were characterized by the same major compounds: 1, 8-cineole and α-pinene, and *Eucalyptus camaldulensis* was the toxic one with LC50 = 26.73 μL/L air and LT50 = 30.46 h. Also, 100% mortality of *E. kuehniella* larvae was reached by *Salvia hortensis* essential oil at 228.5 μL/L air after 12 h of exposure (Maedeh et al. 2011). Similarly, in the study of Emanjomeh et al. (2014), *Zataria multiflora* exhibited high fumigant toxicity against *E. kuehniella* adults and larvae at 0.98 μL/L air and 20.67 μL/L air LC50, respectively. Also, fumigant toxicity screening on different development stages of *E. kuehniella* showed the effectiveness of *Mentha spicata* essential oil with LC50 value of 0.5 μL/L air for adults and eggs mortality reaching 50–60% (Eliopoulos et al. 2015). In addition, Kheirkhah et al. (2015) noted that *E. kuehniella* adults were more sensitive to the fumigant effect of *Ziziphora clinopodioides* oil than larvae (LC50 = 1.39 μL, 42.17 μL/L air). Mediouni Ben Jemâa et al. (2013) conducted fumigant assays using *Laurus nobilis* essential oil from Algeria and Tunisia against *E. kuehniella* adults and recorded the superiority of the Algerian oil (LC50 = 20.77, 33.75 μL/L air). Likewise, Ecran et al. (2013) observed that *Prangos ferulacea* oil was more toxic to *E. kuehniella* adults (LC50 = 1) than eggs (LC50 = 320. 37 μL/L air). Pandir and Baş (2016) recorded that paprica, basil, papermint, and rosemary essential oils were toxic to eggs, larvae, and adults *E. kuehniella*. Ben Chaaban et al. (2019) proved the fumigant efficacy of *Mentha pulegium* against *E. kuehniella* adults (LC50 = 0.3 μL/L air).

Inhibition of oviposition using essential oils represents a pertinent criterion to control pest infestation and manage stored products (Singh and Pandey 2018). In this respect, Ulukanli et al. (2014) pointed out that essential oils extracted from cortex of *Pinus pinea* and *Pinus brutia* presented ovicidal activity on *E. kuehniella* eggs (LC50 = 343. 57, 299.9 μL/L air). According to Tunç et al. (2000), cumin and anise induced 100% mortality of *E. kuehniella* eggs. Besides, Bachrouch et al. (2010) cited that *Pistacia lentiscus* was toxic against *E. kuehniella* (LC50 = 1.84 μL/L air) and reduced adults’ longevity, fecundity, hatching rate, and copulation rate. Generally, as recorded by Delimi et al. (2013) and Taibi et al. (2018), topical bioassay of *Artemisia herba-alba* and *Origanum vulgare* oils on *E. kuehniella* chrysalis causes disorders of nymphal duration, disrupts adult reproduction, reduces the longevity of females, and therefore decreases their fecundity.

As reported in this work, strong contact and fumigant toxicities were exhibited by both Algerian essential oils. The round-leaved mint *M. rotundifolia* showed most promising results in terms of adult mortality and impact on biological parameters of the target pest *E. kuehniella*. Indeed, a complete inhibition of egg hatching and copulation rate was accomplished. Consequently, our results clearly support the application of this oil as a potential biocontrol agent that may be an appropriate alternative for achieving a safer and more sustainable management of stored commodities in Algeria. Therefore, further research including the efficacy of this oil on other stored-product pests (coleopteran species) and its impact on food quality are warranted.

**Compliance with ethical standards**

**Conflict of interest** The author declares that they have no conflict of interest.

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References

Abbott W (1925) A method of computing the effectiveness of an insecticide. J Econ Entomol 18(2):265–267

Abdelgaleil SAM, Mohamed MIE, Badawy MEI, El-Arami SAA (2009) Fumigant and contact toxicities of monoterpene to Sitophilus oryzae (L.) and Tribolium castaneum (Herbst) and their inhibitory effects on acetylcholinesterase activity. J Chem Ecol 35:518–525

AFNOR (1986) Recueil des normes Françaises « huiles essentielles». AFNOR, Paris

Aidi Wannes W, Mhamdi B, Marzouk B (2007) Essential oil composition of two Myrtus communis L. varieties grown in North Tunisia. Ital J Biochem 56(2):180–186

Al-Izzi MAJ, Al-Maliky SY, Younis MA, Jabbo NF (1985) Bionomics of Ectomyelois ceratoniae (Lepidoptera: Pyralidae) on Pomegranates in Irak, Environ Entomol 14(2):149–153

Amzouar S, Boughdad A, Maatoui A, Allam L (2016) Comparison of the chemical composition and the insecticidal activity of essential oils of Mentha suaveolens Ehrl collected from two different regions of Morocco, against Brucus rufimanus (Bohman) (Coleoptera: Chrysomelidae). Int J Innov Appl Stud 18(3):836–845

Ayvaz A, Sagdic O, Karaborklu S, Ozturk I (2010) Insecticidal activity of the essential oils from different plants against three stored-product insects. J Insect Sci 10(1):1–13

Bachrouch O, Mediouni Ben Jemâa J, Aidi Waness W, Talou T, Marzouk B, Abderrabba M (2010) Composition and insecticidal activity of essential oil from Pistacia lentiscus L. against Ectomyelois ceratoniae Zeller and Ephesia kuehniella Zeller (Lepidoptera: Pyralidae). J Stored Prod Res 46(4):242–247

Barhouchi B, Aouadi S, Abdi A (2016) Essential oil chemical composition of myrtle growing in Northeastern Algeria and estimation of its antibacterial effectiveness. Am J Biobtech Biotechnol 12(2):110–121

Baser K, Kürkçüoğlu M, Tarımcılar G, Kaynak G (1999) Essential oils of Mentha species from Northern Turkey. J Essent Oil Res 11(5):579–588

Ben Achour N, Messoudi S, Mediouni Ben Jemâa J, Ben Ammar A, Belhadj O (2008) Isolation of Bacillus thuringiensis strains active against Ceratitis capitata and Ephesia kuehniella in Tunisia. Microbiologie et hygiene alimentaire 20:35–39

Ben Chaaban S, Haouel Hamdi S, Mahjoubi K, Mediouni Ben Jemâa J (2019) Composition and insecticidal activity of essential oil from Ruta graveolens, Mentha pulegium and Ocimum basilicum against Ectomyelois ceratoniae Zeller and Ephesia kuehniella Zeller (Lepidoptera: Pyralidae). J Plant Dis Prot 126(3):237–246

Ben Ghnaya A, Chograni H, Messoud C, Boussaid M (2013) Comparative chemical composition and antibacterial activities of Myrtus communis L. Essential oils isolated from Tunisian and Algerian population. J Plant Pathol Microbiol 4(7):186

Benabdallah A, Boumendjel M, Aissi O, Rahmounne C, Boussaid M, Messaoud M (2018) Chemical composition, antioxidant activity and acetylcholinesterase inhibitory of wild Mentha species from northeastern Algeria. South Afr J Bot 116:111–139

Boelens M, Jimenez R (1992) The Chemical composition of Spanish Myrtle oils: part II. J Essent Oil Res 4(4):349–353

Boukhebti H, Chaker A, Belhadj H, Sahli F, Lauher H, Harzallah D (2011) Chemical composition and antibacterial activity of Mentha pulegium L. and Mentha spicata L. essential oils. Sch Res Libr 3(4):267–275

Bounih A (2016) Cribleage phytochimique, étude toxicologique et valorisation pharmacologique de Melissa officinalis et de Mentha rotundifolia (Lamiacée) [Thèse de doctorat]. Université Mohamed V, Faculté de médecine et de pharmacie, Rabat

Bouzouit N, Khachouri F, Hamdi M, Chaabouni M (2003) Antimicrobial activity of essential oils from Tunisian aromatic plants. Flavour Fragg J 18(5):380–383

Brada M, Bezzina M, Marlier M, Lognay G (2006) Chemical composition of the leaf oil of Mentha rotundifolia (L.) from Algeria. J Essent Oil Res 18:663–665

Brada M, Bezzina M, Marlier M, Carlier A, Lognay G (2007) Variabilité de la composition chimique des huiles essentielles de Mentha rotundifolia du Nord de l’Algérie. Base En Ligne 11(1):1–4

Brahimi F, Abdenour A, Bruno M, Silvia P, Alessandra P, Danilo F, Yalaoui-Guellal D, Elsebai MF, Madani K, Chibane M (2016) Chemical composition and in vitro antimicrobial, insecticidal and antioxidative activities of the essential oils of Mentha pulegium L. and Mentha rotundifolia L. Huds growing in Algeria. Ind Crops Prod 88:96–105

Cetin H, Erler F, Yanikoglu A (2004) Larvicidal activity of a botanical natural product AkseBio2, against Culex pipiens. Fitoterapia 75(7–8):724–728

Chalchat J, Garry R, Michet A (1998) Essential oils of Myrtle (Myrtus communis L.) of the Mediterranean littoral. J Essent Oil Res 10(6):613–617

Chang Y, Lee SH, Na JH, Chang PS, Han J (2017) Protection of grain products from Sitophilus oryzae (L.) contamination by anti-insect pest repellent sacher containing allyl mercaptan microcapsule. J Food Sci 82(11):2634–2642

Curnin M, Bianchi A, Epifano F, Brunì R, Torta L, Zambonelli A (2003) Composition and in vitro antifungal activity of essential oils of Erigeron canadensis. Chem Nat Compd 39(2):191–194

Delima A, Taibi F, Fissah A, Gherib S, Boukkari M (2013) Bioactivité des huiles essentielles de l’armoise blanche Artemessia herba alba: effet sur la reproduction et la mortalité des adultes d’un ravageur des denrées stockées Ephesia kuehniella (Lepidoptera). Afr Sci 9(3):82–90

Derwich E, Benziane Z, Boukir A, Benaabidate L (2009) GC-MS analysis of the leaf essential oil of Mentha rotundifolia, a traditional herbal medicine in Morocco. Chem Bull Politech Univ Timisora 54(2):85–88

Derwich E, Benziane Z, Boukir A (2010) Antibacterial activity and chemical composition of the leaf essential oil of Mentha rodiifolia from Morocco. Electron J Environ Agric Food Chem 9(1):19–28

Djene A, Yangiéla J, Amrouche T, Boubri S, Boussad N, Roncales P (2011) Chemical composition and antimicrobial effects of essential oils of Eucalyptus globulus, Myrtus communis and Satureja hortensis against Escherichia coli O127:H1 and Staphylococcus aureus in minced beef. Food Sci Technol Int 17(6):505–515

Ebodallahi S (2013) Essential oils isolated from Myrtaceae family as natural insecticides. Ann Rev Res Biol 3(3):148–175

Ecran F, Baş H, Koç M, Pandir D, Oztzemis S (2013) Insecticidal activity of essential oil of Prangos feraluce (Umbrelliferae) against Ephesia kuehniella (Lepidoptera: Pyralidae) and Trichogramma embryophagum (Hymenoptera: Trichogrammatidae). Turk J Agric For 37:719–725

El Arch M, Satrani B, Farah A, Bennani L, Boriky D, Fechtal M, Blaghen M, Talbi M (2003) Composition chimique et activité anticroissant et insecticide de l’huile essentielle de Mentha rotundifolia du Maroc. Acta Bot Gallica 150(3):267–274

Eliopoulos P, Hassiotis C, Andreadis S, Porichi A (2015) Fumigant toxicity of essential oils from basil and 2-pamant against two major pyralid pests of stored products. J Econ Entomol 108(2):805–810

Emamjomeh L, Imani S, Salehi K, Masoni M, Hrhamaeypour S, Larjani K (2014) Chemical composition and insecticidal activity of essential oil of Zataria multiflora Boiss (Lamiaceae) against Ephesia kuehniella (Lepidoptera: Pyralidae). Pelaga Res Libr 4(3):253–257
Gardeli C, Vassiliki P, Athanasios M, Kibouris T, Komaitis M (2008) Essential oil composition of *Pistacia lentiscus* L. and *Myrtus communis* L.: evaluation of antioxidant capacity of methanolic extracts. Food Chem 107(3):1120–1130

Gende L, Mendiara S, Fernandez N, Baren C, Lira D, Bandoni A, Fritz R, Floris I, Eguaras M (2014) Essential oils of some Mentha spp. and their relation with antimicrobial activity against *Paeni bacillus* larvae, the causative agent of American foulbrood in honey bees, by using the bio-autography technique. Bull Insectol 67(1):13–20

Ghoulami S (2001) Phytochemical study of *Mentha longifolia* of Morocco. Fitoterapia 72(5):596–598

Hami M, Taibi F, Smaghe G, Soltani Mazouni N (2005) Comparative toxicity of three eucalyptus against insecticides against the Mediterranean flour moth. Commun Agri Appl Biol Sci Gheit Univ 70(4):767–773

Hori M (2003) Repellency of essential oils against the cigarette beetle, *Lasioderma serricorne* (Fabricius) (Coleoptera: Anobiidae). Appl Entomol Zool 38(4):467–473

Huang Y, Tan JMWL, Kini RM, Ho SH (1997) Toxic and Antifeedant action of nutmeg oil against *Tribulium castaneum* (Hurbst) and *Sitophilus zeamais* Motsch. J Stored Prod Res 33(4):289–298

Ismann M (2000) Plant essential oils for pest and disease management. Crop Prot 19:603–608

Ismann MB, Grieneisen ML (2014) Botanical insecticide research: many publication, limited useful data. Trends Plant Sci 19:140–145

Jacob T, Cox P (1977) The influence of temperature and humidity on the life-cycle of *Ephedra kuehniella* Zeller (Lepidoptera: Pyralidae). J Stored Prod Res 13(3):107–118

Jaraya A (2003) Principaux nuisibles des plantes cultivées et des drenées stockées en Afrique du nord: leur biologie, leurs ennemies naturels, leurs dégâts et leur contrôleur. Maghreb Ed, Tunisia

Jerkovic I, Radonic A, Borcic I (2002) Comparative study of leaf, fruit and flower essential oils of Croatian *Myrtus communis* (L.) during a one-year vegetative cycle. J Essent Oil Res 14(4):266–270

Jitendra K, Nitin K, Kulkarni D (2009) Plant-based pesticides for control of *Helicoverpa armigera* on *Cucumis sativus*. Asian Agric Hist 13(4):327–332

Johnson J, Valero K, Hannel M (1997) Effect of low temperature storage on survival and reproduction of Indian meal moth (Lepidoptera: Pyralidae). Crop Prot 16(6):519–523

Karakořuli S, Ayvaz A, Yilmaz S, Akbulut M (2011) Chemical composition of some essential oils against *Ephestia kuehniella*. J Econ Entomol 104(4):1212–1219

Kasrati A, Alouzi Jamali C, Bekkouche K, Spooner-Heart R, Leach D, Abbad A (2015) Chemical characterization and insecticidal properties of essential oils from different wild populations of *Mentha suaveolens* subsp. *timjia* (Briq.) Harley from Morocco. Chem Biodivers 12:823–831

Khani A, Farzaneh B (2012) Chemical composition and insecticidal activity of Myrtle (*Myrtus communis* L.) essential oil against two stored-product pests. J Med Plant By-Prod 2:83–89

Kheirkhah M, Ghasemi V, Yazdi A, Rahban S (2015) Chemical composition and insecticidal activity of essential oil from *Ziziphus clinopodioides* Lam used against the Mediterranean flour moth, *Ephestia kuehniella* Zeller. J Plant Prot Res 55(3):260–265

Kim S, Roh J, Kim D, Lee H, Ahn Y (2003) Insecticidal activities of aromatic plant extracts and essential oils against *Sitophilus oryzae* and *Callosobruchus chinensis*. J Stored Prod Res 39:293–303

Kokkins S, Papageorgiou V (1988) Constituents of essential oils from *Mentha X rotundifolia* growing wild in Greece. Planta Med 54(2):166–167

Kostyukovsky M, Rafaeli A, Gileadi C, Demchenko N, Shaaya E (2002) Activation of octopaminergic receptors by essential oil constituents isolated from aromatic plants: possible mode of action against insect pests. Pest Manag Sci 58(11):1101–1106

Lawrence B (2007) Mint: the genus Mentha. Medicinal and aromatic plants-industrial profiles. CRC Press, Boca Raton

Liu ZL, Ho SH (1999) Bioactivity of the essential oil extracted from *Evodia rutaecarpa* Hook f. et Thomas against the grain storage insects, *Sitophilus zeamais* Motsch and *Tribulium castaneum* (Hurbst). J Stored Prod Res 35:317–328

Lorenzo D, Paz D, Dellacassa E, Davies P, Vila R, Canigueral S (2002) Essential oils of Mentha pulegium and Mentha rotundifolia from Uruguay, Brazil Arch Biol Technol 54(4):519–524

Maeedh M, Hamzeh I, Hossein D, Majid A, Reza R (2011) Bioactivity of essential oil from *Satureja hortensis* (Lamiaceae) against three stored-product insect species. Afr J Biotechnol 10(34):6620–6627

MaffeI M (1988) A chemotype of *Mentha longifolia* (L.) Hudson particularly rich in piperitenone oxide. Flavour Fragr J 3(1):23–26

Mami Maazoun A, Ben Hlel T, Haouel Hamdi S, Belhadj F, Mediouni Ben Jemâa J, Marzouki M (2017) Screening for insecticidal potential and acetylcholinesterase activity inhibition of *Urginea maritima* bulbs extract for the control of *Sitophilus oryzae* (L.). J Asia Pac Entomol 20(3):752–760

Mediouni Ben Jemâa J (2014) Essential oil as source of bioactive constituents for the control of insect pests of economic importance in Tunisia. Med Aromat Plants 3(2):518–525

Mediouni Ben Jemâa J, Haouel S, Bouaziz M, Khouja M (2012) Seasonal variations in chemical composition and fumigant activity of five Eucalyptus essential oils against three moth pests of stored dates in Tunisia. J Stored Prod Res 48:61–67

Mediouni Ben Jemâa J, Tersins N, Boushsh E, Taleb-Toudert K, Khouja M (2013) Fumigant control of the Mediterranean flour moth *Ephestia kuehniella* with the Nobel Laurel *Laurus nobilis* essential oils. Tunis J Plant Prot 8(1):33–44

Messoud C, Zaouali Y, Ben Salah A, Khoudja M, Boussaid M (2005) *Myrtus communis* in Tunisia: variability of the essential oil composition in natural populations. Flavour Fragr J 20(6):577–582

Nerio L, Olivero-Verbal J, Stashenko E (2009) Repellent activity of essential oils from seven aromatic plants growing against *Sitophilus zeamais* Motschulsky (Coleoptera). J Stored Prod Res 45:212–214

Nikšić H, Kovač-Bešović E, Đurić K, Korać N, Omeragić E, Muratóvić S (2014) Seasonal variation in content and chemical composition of essential oils from leaves of *Mentha longifolia* Huds. (Lamiaceae). Bull Chem Technol Bosnia Herzeg 43:29–34

Nishimura H (2001) Aroma constituents in plants and their repellent activities against mosquitoes. Aroma Res 2:257–267

Oumzil H, Ghoulami S, Rhajaoui M, Ilidrissi A, Fkih Tetouani S, Faid M, Benjouad M (2002) Antibacterial and antifungal activity of essential oils of *Mentha suaveolens* plant. Phytother Res 16(8):727–731

Pandir D, Baş H (2016) Compositional analysis and toxicity of four plant essential oils at different stages of Mediterranean flour moth, *Ephestia kuehniella* Zeller (Lepidoptera: Pyralidae). Türk Entomol Derg 40(2):185–195

Pascal Vialalobos M, Robledo A (1998) Screening for anti-insect activity in Mediterranean plants. Ind Crops Prod 8(3):183–194

Pino J, Rosado A, Fuentes V (1999) Chemical composition of the leaf oil of *Menta rotundifolia* (L.) Hudson from Cuba. J Essent Oil Res 11(2):241–242

Prajapati V, Tripathi AK, Aggarwal KK, Khanuja SPS (2005) Insecticidal, repellent and oviposition-deterrent activity of selected essential oils against *Anopheles stephensi*, *Aedes aegypti* and *Culex quinquefasciatus*. Biorec Technol 96:1749–1757

Quan M, Liu Q, Liu Z (2018) Identification of insecticidal constituents from the essential oil from the aerial parts *Stachys riederi* var *japonica*. Molecules 23(5):1–12

Rahman S, Biswas S, Barman N, Ferdous T (2016) Plant extract as selective pesticide for integrated pest management. Biotechol Res J 2(1):6–10
Rajendran S, Sriranjini V (2008) Plant products as fumigants for stored-product insect control. J Stored Prod Res 44(2):126–135
Rasooli I, Moosavi ML, Rezaee MB, Jaimand K (2002) Susceptibility of microorganisms to Myrtus communis L. essential oil and its chemical composition. J Agric Sci Technol 4:127–133
Reitsema R (1958) A biogenetic arrangement of Mint species. J Am Pharm Assoc Sci Ed 47(4):267–269
Riahi L, Elferchichi M, Ghazghazi H, Jebali J, Aouadhi C, Chogran H, Zaouali Y, Zognhami N, Milki A (2013) Phytochemistry, antioxidant and antimicrobial activities of the essential oils of Mentha rotundifolia L. in Tunisia. Ind Crops Prod 49:883–889
Sangha JS, Astatkie T, Cutler GC (2017) Ovicidal, larvicidal and behavioural effects of some plant essential oils on diamond-back moth (Lepidoptera: Plutellidae). Entomol Soc Can 149(5):639–648
Satrani B, Farah A, Talbi M (2006) Effet de la distillation fractionnée sur la composition chimique et l’activité antimicrobienne des huiles essentielles du Myrte (Myrtus communis L.) du Maroc. Acta Bot Gallica 153(2):235–242
Scora R (1973) Essential leaf oil variability in green, variegated and albino foliage of Myrtus communis. Phytochemistry 12(1):153–155
Senfi F, Safaralizadeh MH, Safavi SA, Aramidéh S (2013) Fumigant toxicity of Laurus nobilis and Myrtus communis on larvae and adults of the red flour beetle, Tribolium castaneum Herbst (Col.: Tenebrionidae). Arch Phytopathol Plant Prot. https://doi.org/10.1080/03235408.2013.812819
Singh P, Pandey A (2018) Prospective of essential oils of the Genus Mentha as biopesticides: a review. Front Plant Sci 9:1–14
Sousa P, Linard C, Azevedo Batista D, Oliveira AC, Coelho de Souza A, Leal Cardoso J (2009) Identification of flavonoid content and chemical composition of the essential oils of Moroccan herbs: Myrtle (Myrtus communis L.), Rockrose (Cistus ladanifer L.) and Montpellier cistus (Cistus monspeliensis L.). J Essent Oil Res 23(2):1–9
Venskutonis P (1996) A chemotype of Mentha longifolia L. from Lithuania rich in piperitenone oxide. J Essent Oil Res 8(1):91–95
Viuda Martos M, Sendra E, Pérez Álvarez J, Fernández López J, Amensour M, Abrini J (2011) Identification of flavonoid content and chemical composition of the essential oils of Moroccan herbs: Myrtle (Myrtus communis L.), Rockrose (Cistus ladanifer L.) and Montpellier cistus (Cistus monspeliensis L.). J Essent Oil Res 23(2):1–9
Waliwitiya R, Kennedy CJ, Lowenberger CA (2009) Larvicidal and oviposition-altering activity of monoterprenoids, trans-anethole and rosemary oil to the yellow fever mosquito Aedes aegypti (Diptera: Culicidae). Pest Manag Sci 65:241–248

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