Infection after transrectal ultrasound-guided prostate biopsy

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Infectious complications after transrectal ultrasound-guided prostate biopsy (TRUS-Bx) appear to be increasing, which reflects the high prevalence of antibiotic-resistant strains of Enterobacteriaceae. Identifying patients at high risk for antibiotic resistance with history taking is an important initial step. Targeted prophylaxis with a prebiopsy rectal swab culture or augmented antibiotic prophylaxis can be considered for patients at high risk of antibiotic resistance. If infectious complications are suspected, the presence of urosepsis should be evaluated and adequate antibiotic treatment should be started immediately.

Keywords: Biopsy; Infection; Prostate

INTRODUCTION

Transrectal ultrasound-guided prostate biopsy (TRUS-Bx) is one of the most commonly performed urologic procedures in the United States and Europe, with approximately one million biopsies performed annually on each continent. TRUS-Bx is a relatively safe procedure and the chances of severe complications are low, but the incidence of infectious complications has recently been rising, along with the potential for more severe complications such as sepsis [1,2]. Escherichia coli is the most common pathogen found in infections after TRUS-Bx [3-6]. Randomized controlled trials have shown that antibiotic prophylaxis is effective in preventing infectious complications following TRUS-Bx [7]. Fluoroquinolone is the most commonly used antibiotic agent for prophylaxis [7,8]. However, worldwide antibiotic resistance is rising [9-11], and as such, infectious complications after TRUS-Bx by fluoroquinolone-resistant E. coli are rising as well [9,12-14].

INCIDENCE OF INFECTIOUS COMPLICATIONS AFTER TRUS-Bx

The incidence of infectious complications after TRUS-Bx is reported to range from 0.1% to 7%; the incidence of infections requiring admission is from 0.6% to 4.1% [15]. In Korea, the reported incidence of infectious complications after TRUS-Bx is from 0.65% to 3.1% [16-18]. In another Asia-Europe multicenter study that included Korea, the reported incidence of febrile urinary tract infection (UTI) was 3.5% and the incidence of infections requiring admission was 3.1% [19]. In Japan, the reported incidence of febrile UTI was from 0.5% to 0.76% [20,21], and in Taiwan, the incidence was 5.4% before fluoroquinolone prophylaxis and 0.9% after fluoroquinolone prophylaxis [22]. A Turkish study reported an incidence of infections requiring admission of 2% [23].
PREVALENCE OF FLUOROQUINOLONE RESISTANCE IN INFECTIOUS COMPLICATIONS

In one Korean study, acute bacterial prostatitis after TRUS-Bx occurred in 1.36% of patients and the prevalence of fluoroquinolone-resistant strains was 23.8% [25]. In a Japanese study, acute bacterial prostatitis developed in 1.3% of patients and all urine and blood cultures yielded levofloxacin-resistant E. coli [26]. In another Japanese study, the rate of genitourinary tract infection was 0.76% and E. coli was the most frequently isolated strain, of which 77.8% showed levofloxacin resistance [21]. In a North American cohort, 2.77% of cases developed infection after biopsy, of which 55% had fluoroquinolone-resistant infection [6]. In a French prospective study, 0.67% of cases had acute bacterial prostatitis, of which 95% showed fluoroquinolone resistance [27]. In an Australian study that analyzed E. coli bacteremia after TRUS-Bx, 62% were fluoroquinolone-resistant [28]. Overall, the reported prevalence of fluoroquinolone resistance in infectious complications after TRUS-Bx ranges from 24% to 100%, and considering the recent trend in increasing antibiotic resistance, fluoroquinolone resistance is expected to rise rapidly. Along with the problem of fluoroquinolone resistance, one should also be wary of the emergence of extended-spectrum beta-lactamase (ESBL)-producing bacteria. One study reported an incidence of ESBL-producing bacteria of 43% in acute prostatitis after TRUS-Bx [29].

SOLUTIONS FOR FLUOROQUINOLONE RESISTANCE

1. Identifying high-risk patients with history taking

Fluoroquinolone use in the previous 3 to 6 months prior to TRUS-Bx was a common risk factor for fluoroquinolone resistance in several studies [5,33-35]. The longer the period of fluoroquinolone use, the higher the incidence of fluoroquinolone resistance [35]. Therefore, thorough history taking is of paramount importance for identifying recent fluoroquinolone usage for other conditions such as UTI, chronic prostatitis, heart valve surgery, and artificial instrument insertion surgery (Fig. 1).

2. Targeted antibiotic prophylaxis

The evidence for routine rectal swab culture before all TRUS-Bx is still indeterminate. However, in cases of a high risk of fluoroquinolone resistance, performing prebiopsy rectal swab culture to identify rectal bacterial flora would be of great assistance in preventing or treating infectious complications. Prebiopsy rectal swab culture should be done 1 to 2 weeks before TRUS-Bx. If the patient has a history of recent antibiotic use, however, prebiopsy rectal swab culture should be postponed or its results should be interpreted cautiously. Targeted antibiotic prophylaxis based on rectal
swab culture results showed a notable decrease in the incidence of infectious complications after TRUS-Bx caused by fluoroquinolone-resistant organisms as well as a decrease in the overall cost of care [31].

3. Changing prophylactic antibiotics for TRUS-Bx

Currently, fluoroquinolone is the recommended antibiotic for TRUS-Bx in US and European guidelines. It is also the most widely used antibiotic agent for antibiotic prophylaxis for TRUS-Bx in practice. However, in regions such as Korea where the rate of fluoroquinolone resistance is high, following the US and European guidelines may be less effective for antibiotic prophylaxis. If prebiopsy rectal swab culture is done, susceptible antibiotic agents should be used, and if it is not done, prophylactic antibiotic agents should be changed in patients suspected of fluoroquinolone resistance. For this, attempts have been made to add an aminoglycoside such as amikacin to fluoroquinolone or to use third-generation cephalosporins for prophylaxis. However, this may cause another problem in addition to fluoroquinolone resistance: emergence of ESBL-producing bacteria. In a Korean report, 20% of patients with infectious complications were found to have ESBL-producing bacteria [25], and in a Canadian report, the incidence of ESBL-producing bacteria was 4.6% [32]. ESBL-producing bacteria are usually resistant to most antibiotics with the exception of carbapenems (imipenem, meropenem). However, it is not recommended to use potent antibiotics such as imipenem or piperacillin/tazobactam for general prophylaxis because this may eventually cause emergence of carbapenem-resistant Enterobacteriaceae that could make us vulnerable to fatal infections.

4. Treating fluoroquinolone-resistant E. coli

Because infectious complications after TRUS-Bx could be fatal, immediate admission and implementation of antibiotics is warranted in cases suspected of sepsis. If a prebiopsy rectal swab culture is done, a susceptible antibiotic agent targeting the suspected causative bacteria should be used. If not, third-generation cephalosporins and aminoglycoside may be the optimal choice, at least in Korea [25]. Because resistance to gentamicin and tobramycin is already high in Korea, amikacin is the recommended aminoglycoside. If ESBL-producing bacteria are suspected or cephalosporins are ineffective, use of carbapenems such as imipenem or meropenem should not be delayed. Once the results of the antibiotic susceptibility test are confirmed, de-escalation therapy is recommended, which consists of switching from a broad-spectrum empiric antibiotic therapy to a narrower spectrum. After the patient is discharged, continued treatment for a sufficient period of time is necessary to cure prostatitis.

CONCLUSIONS

Fluoroquinolones have been the most common choice for prophylactic antibiotics preceding TRUS-Bx. Infectious complications after TRUS-Bx are increasing, and this appears to be due to an increasing prevalence of fluoroquinolone-resistant strains in the rectal flora. Therefore, identifying the risk for fecal carriage of fluoroquinolone-resistant strains by history taking should be the initial step in the TRUS-Bx procedure. If a risk of fluoroquinolone resistance is present, targeted antimicrobial prophylaxis using rectal swab cultures or alternative antibiotics may be recommended for prophylaxis. In patients with infectious complications after TRUS-Bx, it is essential to administer appropriate antibiotics immediately.
CONFLICTS OF INTEREST

The author has nothing to disclose.

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