Introduction

The cytology of ocular impression has numerous clinical applications. Currently, health professionals opt for invasive techniques, such as biopsies and punctures, causing discomfort to the patient. The possibility of obtaining ocular surface samples with minimal discomfort for the patient, in a non-invasive procedure, makes it the ideal method for investigating the ocular surface [1]. When the filter paper is applied to the conjunctival surface, it is possible to remove epithelial cells, still viable, from one to three layers of the conjunctiva, preserving its morphological and structural characteristics [2]. The analysis of these cells allows the evaluation of the degree of metaplasias, immunocytochemical analyzes with specific cytokeratin markers, isolation of pathogens and the obtaining of mRNA, cells for phenotyping and the diagnosis of melanose and melanoma [3]. This work aims to demonstrate an efficient cellular preservation procedure, with a simple and easily adaptable methodology in the routine of the services of ophthalmology in health units. This will allow a cytological evaluation of diseases of the ocular surface, replacing invasive techniques and reducing the implications for the patient.

Methods

Studies cases

50 samples of ocular impression cytology of the bulbar conjunctiva were obtained from volunteers, students and teachers of the Escola Superior de Tecnologia da Saúde de Lisboa. Collection and Handling of Samples

The technique of printing cytology required four fundamental methodological steps: harvest, fixation, staining and evaluation. The harvesting material includes a strip of cellulose acetate from Millipore Corporation, HAWP304F0 catalog code, with a 0.45μm pore. A paper model [4], approximately 7mm wide by 5mm high with apex was applied (Figure 1). This format allows the specimen arrangement on the filter paper to be kept constant, and posteriorly its orientation on the blade. Without the need for topical anesthesia, the apex of the filter paper is applied to the corneal surface, pressing with a swab for a period of 2 to 5 seconds. It is removed by the peeling maneuver and placed immediately in SureThin. The slides were analyzed by three independent evaluators, using an evaluation grid with the following parameters: cell size, detail and nuclear membrane, detail and cytoplasmic membrane, ratio N / C and tinctorial affinity.

Results: The filter paper with apex helped to correctly position the paper in the eye and the procedure that was applied allowed an effective collection of cells with 50-70% of the filter surface being filled without the need for topical anesthesia. The SureThin fixative presented quality in cell preservation, in addition to being more economical. The Papanicolaou technique proved to be ideal in the coloring of ocular epithelial cells. This developed methodology presented an efficiency of 80%, 9% to a maximum score 2.

Conclusions: The presented method proved to be very effective in evaluating of ocular cell samples, while simultaneously proving to be a very cheap and comfortable technique for the patient.
The “nuclear detail” presented 111 responses in 150 for score 2 (“Visualization of all nuclear details”). The “nuclear membrane” remained intact and well defined, with 114 responses for score 2 (76.0%). It was observed that the SureThin fixative was effective in the preservation, since 83.3% of the answers were obtained for score 2 (“Small nucleus with N/C ratio 1:6”). In general, the staining affinity was maintained, allowing to differentiate its cellular constituents, with 123 responses in 150 for score 2. (Figure 2 (a,b,c,d)) shows the quality of the samples collected in this study.

Discussion

Many cases in which corneal surface pathologies are present, biopsy are contraindicated [5]. The lack of literature to support the various steps of the cytological ocular printing technique, namely in the fixation step, poses a problem when practicing the technique. The method used proved to be ideal for the evaluation of cytological exams by ocular impression. Our technique differs from some authors mainly at the fixer level used. Adams et al. [5] reports the importance of fixation in this type of samples, suggesting other possible fixatives such as formaldehyde, ethanol, methanol and fixative spray. However, there are no studies in the literature that indicate the most effective fixation method. The coloring technique used presented good results, being easy to execute and economical, although it did not belong to Martinez et al. [6], which uses a modified method of Periodic Acid-Schiff, Hematoxylin and Papanicolaou. The clarification stage in xylene is emphasized by Stolz [2]. This will allow for the subsequent analysis by optical microscopy, once the filter paper becomes transparent in organic solvents, used in the stage of the diaphanization, during the staining. The use and care of alcohol-based reagents was based on quotes from Michalany [7], which recommends the exchange of ethanol and never re-use.

Conclusion

The present method proved to be effective for the evaluation of ocular surface cells, using a cost effective and comfortable technique for the patient, easily applicable in the hospital routine.

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