A Potential Prognostic Marker for Recognizing VEGF-Positive Hepatocellular Carcinoma Based on Magnetic Resonance Radiomics Signature

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Objectives: The objective of our project is to explore a noninvasive radiomics model based on magnetic resonance imaging (MRI) that could recognize the expression of vascular endothelial growth factor (VEGF) in hepatocellular carcinoma before operation.

Methods: 202 patients with proven single HCC were enlisted and stochastically distributed into a training set (n = 142) and a test set (n = 60). Arterial phase, portal venous phase, balanced phase, delayed phase, and hepatobiliary phase images were used to radiomics features extraction. We retrieved 1906 radiomic features from each participant’s MRI images. The F-test was applied to choose the crucial features. A logistic regression model was adopted to generate a radiomics signature. By combining independent risk indicators from the fusion radiomics signature and clinico-radiological features, we developed a multivariable logistic regression model that could predict the VEGF status preoperatively through calculating the area under the curve (AUC).

Results: The entire group comprised 108 VEGF-positive individuals and 94 VEGF-negative patients. AUCs of 0.892 (95% confidence interval [CI]: 0.839 - 0.945) in the training dataset and 0.800 (95% CI: 0.682 - 0.918) in the test dataset were achieved by utilizing radiomics features from two phase images (8 features from the portal venous phase and 5 features from the hepatobiliary phase). Furthermore, the nomogram relying on a combined model that included the clinical factors α-fetoprotein (AFP), irregular tumor margin, and the fusion radiomics signature performed well in both the training (AUC = 0.936, 95% CI: 0.898-0.974) and test (AUC = 0.836, 95% CI: 0.728-0.944) datasets.

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Specialty section: This article was submitted to Cancer Imaging and Image-directed Interventions, a section of the journal Frontiers in Oncology

Received: 21 January 2022
Accepted: 11 March 2022
Published: 04 April 2022

Citation: Fan T, Li S, Li K, Xu J, Zhao S, Li J, Zhou X and Jiang H (2022) A Potential Prognostic Marker for Recognizing VEGF-Positive Hepatocellular Carcinoma Based on Magnetic Resonance Radiomics Signature. Front. Oncol. 12:857715. doi: 10.3389/fonc.2022.857715
VEGF therapy plays an important role in the therapeutic outcomes for advanced hepatocellular carcinoma (14). Anti-sorafenib demonstrated a marked improvement in clinical combination of atezolizumab and bevacizumab versus successful phase III trial (IMbrave150) that assessed the its surroundings can be reduced by anti-VEGF therapies (13). A Immunosuppression caused by VEGF inside the malignancy and development and progression of HCC in previous studies (12). Overexpression of VEGF has been associated with the formation of fibrin scaffolds and induce new blood vessel sprouting around growing tumors. By interacting with multiple of its receptors (VEGFR 1, VEGFR 2, VEGFR 3), VEGF induces angiogenesis (9). The VEGF family comprises six members (VEGFA-E, and placental growth factor). VEGF-A (often referred to as VEGF) is the most prototypical and the best-studied VEGF family member. The overexpression of VEGF can occur in various human tumors, including HCC (10, 11). Overexpression of VEGF has been associated with the development and progression of HCC in previous studies (12). Immunosuppression caused by VEGF inside the malignancy and its surroundings can be reduced by anti-VEGF therapies (13). A successful phase III trial (IMbrave150) that assessed the combination of atezolizumab and bevacizumab versus sorafenib demonstrated a marked improvement in clinical outcomes for advanced hepatocellular carcinoma (14). Anti-VEGF therapy plays an important role in the therapeutic drugs for HCC. The difference in VEGF expression level will affect the treatment efficacy. Previous studies revealed that higher VEGF levels were associated with more invasive disease, shorter survival times, and worse results following surgery and local treatment (15). So it is necessary to investigate the VEGF expression. From this perspective, it appears that the analysis of VEGF expression may enhance the clinical management of HCC. Hence, early diagnosis of VEGF-positive HCC is the key to choosing the best optimal treatment strategy.

Radiomics is a new domain in medicine that utilizes high-throughput quantitative image features based on images from imaging examinations that cannot be resolved by the naked eye for diagnosis and prognosis (16). These features potentially capture the intratumoral heterogeneity, which can offer information about the tumor microenvironment and the phenotype of cancer (17). The strengths of multi-parametric magnetic resonance imaging (MRI) are high temporal resolution, excellent soft tissue definition, and lack of ionizing radiation in tumor imaging (18). Some studies reported that the radiomics signature derives from contrast-enhanced MRI could be used as the imaging biomarker in the microvascular invasion and Cytokeratin19 status of HCC (19, 20). However, as far as we know, few research concentrates on the prediction of VEGF-positive HCC based on MRI radiomics signature. Thus, this project was targeted to analyze the feasibility of multi-parametric MRI radiomics models for predicting VEGF-positive HCC.

**Conclusions:** The combined model acquired from two phase (portal venous and hepatobiliary phase) pictures of gadolinium-ethoxybenzyl-diethylenetriamine-pentaacetic acid (Gd-EOB-DTPA)-enhanced MRI could be considered as a credible prognostic marker for the level of VEGF in HCC.

**Keywords:** magnetic resonance imaging, hepatocellular carcinoma, radiomics, VEGF, diagnosis

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**INTRODUCTION**

Hepatocellular carcinoma (HCCs) is the most common primary malignant tumor of the liver, with a high mortality rate worldwide (1, 2). Surgical excision is the preferable treatment for HCC, but the relapse and metastasis rates are still high. The Barcelona Clinic Liver Cancer (BCLC) staging system may help with prognosis evaluation and therapy programs (3). However, clinical outcomes can vary greatly among patients within the same BCLC stage because of its molecular heterogeneity. Therefore, the management of HCC is challenging, and understanding the tumor molecular heterogeneity of HCC is important to help develop and optimize therapeutic protocols.

A number of molecular biomarkers have been suggested as potential prognostic factors for poor outcome in HCC, including biomarkers closely associated with cell motility, adhesions, angiogenesis, apoptosis, invasion, etc. (4–7). Among the angiogenesis-related factors, the primary angiogenic factor is vascular endothelial growth factor (VEGF) (8). VEGF, also known as vascular permeability factor, was discovered in 1983 as a protein released by tumor cells. VEGF can promote the formation of fibrin scaffolds and induce new blood vessel sprouting around growing tumors. By interacting with multiple of its receptors (VEGFR 1, VEGFR 2, VEGFR 3), VEGF induces angiogenesis (9). The VEGF family comprises six members (VEGFA-E, and placental growth factor). VEGF-A (often referred to as VEGF) is the most prototypical and the best-studied VEGF family member. The overexpression of VEGF can occur in various human tumors, including HCC (10, 11). Overexpression of VEGF has been associated with the development and progression of HCC in previous studies (12). Immunosuppression caused by VEGF inside the malignancy and its surroundings can be reduced by anti-VEGF therapies (13). A successful phase III trial (IMbrave150) that assessed the combination of atezolizumab and bevacizumab versus sorafenib demonstrated a marked improvement in clinical outcomes for advanced hepatocellular carcinoma (14). Anti-VEGF therapy plays an important role in the therapeutic drugs for HCC. The difference in VEGF expression level will affect the treatment efficacy. Previous studies revealed that higher VEGF levels were associated with more invasive disease, shorter survival times, and worse results following surgery and local treatment (15). So it is necessary to investigate the VEGF expression. From this perspective, it appears that the analysis of VEGF expression may enhance the clinical management of HCC. Hence, early diagnosis of VEGF-positive HCC is the key to choosing the best optimal treatment strategy.

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**MATERIALS AND METHODS**

**Patients and MRI Protocol**

The present research met with the approval of our Hospital Review Board (approval number KY2019-217) and written informed assent was exempted for all participators. The participants with HCC were enrolled from imaging database management system between March 2017 and March 2021. Gadolinium-ethoxybenzyl-diethylenetriamine-pentaacetic acid (Gd-EOB-DTPA, Primovist, Germany)-enhanced MRI imaging was carried out for a total of 284 patients who were suspected of HCC and experienced subsequent hepatic excision. The subject entry criteria were as follows: 1) Patients had a definite diagnosis of single primary HCC by pathology; 2) spanning less than 14 days between the first MRI examination and surgery. The exclusion criteria for patients included: 1) Patients had previous treatment, including partial hepatectomy, transcatheter arterial chemoembolization (TACE), radiotherapy, chemotherapy or needle biopsy (n = 23); 2) patients with tumor emboli in the bile duct or vessel (n = 24), or having distant transfer (n = 12); 3)
Clinical and pathological data were incomplete (n = 10); 4) with poor image quality (n = 13). Finally, 202 patients (169 men and 33 women) were eligible and randomly divided into two datasets at a proportion of 7:3. Details of baseline, involving age, gender, tumor diameter, etiology of liver disease, cirrhosis, ascites, aspartate aminotransferase (AST), alanine aminotransferase (ALT), total bilirubin (TBIL), neutrophil count, lymphocyte count, neutrophil-to-lymphocyte ratio (NLR), albumin, serum α-fetoprotein (AFP), differentiation degree, were derived from medical records. Supplementary Method 1 presents the exhaustive MRI procedure. The flowchart for patient selection is demonstrated in Figure 1.

Immunohistochemical Analysis of VEGF
All patients underwent subsequent radical resection of HCC. The detailed immunohistochemical protocol is explained in Supplementary Method 2. Two observers without knowledge of the clinicopathologic parameters scored the slides independently using a previously validated scoring system. Any disagreements were arbitrated by a third rater. VEGF scoring was calculated using the immune reactive score (IRS) as depicted before (21–23): IRS = SI (staining intensity) × PP (percentage of positive cells). SI was graded as 0 (negative), 1 (weak), 2 (moderate) and 3 (strong). PP was categorized as 0 (negative), 1 (less than 5% positive cells), 2 (6-25% positive cells), 3 (26-50% positive cells), 4 (51-75% positive cells), and 5 (more than 75% positive cells). The IRS was evaluated using ten visual fields from different regions of each tumor, and the average IRS was determined. The IRS classified VEGF status into four categories: grade 0 (IRS 0), grade 1 (IRS 1-4), grade 2 (IRS 5-9), and grade 3 (IRS 10-15). VEGF (grade 0 and grade 1) was defined as VEGF-negative expression, while VEGF (grade 2 and grade 3) was marked as VEGF-positive expression (24).

Morphologic Features
Two radiologists who were both lack of the histopathologic information analyzed the MR images on a Picture Archiving and Communication System (PACS) work-station. They negotiated every divergence to make a consensus. Supplemental Table 1 demonstrated the agreements of inter-observer about morphologic MRI features. The following qualitative morphologic variables were evaluated (20): (a) The tumor margin was classified as either smooth or irregular. It was defined irregular tumor margin if the ill-defined interface with normal tissue on HBP.
images was present; (b) arterial rim enhancement, described as a ring-enhanced with central low signal areas on arterial phase; (c) radiological capsule presence or absence.

**HCC Segmentation**
All images of the HCC tumors were uploaded on the online Deepwise Research Platform (https://research.deepwise.com). Radiologist A, who was ignorant of the pathological conditions and had five years of expertise interpreting HCC imaging, delineated the HCC lesions. Radiologist B validated the region of interests (ROIs). ROIs were individually delineated on pictures from the arterial phase (AP), portal venous phase (PVP), balanced phase (BP), delayed phase (DP), and hepatobiliary phase (HBP). Twenty-two HCCs were arbitrarily chosen and re-segmented one week later by the two radiologists, in order to calculate the intra-class and inter-class correlation coefficient (ICCs).

**Clinical and Morphologic Risk Features**
We used logistic regression (univariate and multivariate) to evaluate the connection between basic information, morphologic features and VEGF expression in HCC of the training dataset. The stopping rule was a backward stepwise selection based on Akaike’s information criterion and the likelihood ratio test. In the multivariate analysis, the potential risk factors were selected as those with a p value < 0.05, and the clinical model derived from these variables was established for the training dataset.

**Radiomic Feature Analysis**
The radiomics analysis technique involved ROI (HCCs) segmentation, feature extraction, feature selection, model building, and testing. The Python package Pyradiomics (Version 3.0.1, https://github.com/Radiomics/pyradiomics) was used for feature extraction (25). The following four steps presented the technique of feature extraction and selection in detail.

1. Standardization of spacing: All MRI pictures were resampled to pixel spacing of 2.0 × 2.0 × 2.0 mm with the aid of interpolator of sitkBSpline in the Python package SimpleITK. This step can eliminate interference from various scales (26).
2. Image filtering and feature calculation: The raw MRI images were processed by a wavelet filter and a LaPlacian-Gaussian filter to strengthen the differentiation of features. The shape-based characteristics, first-order and texture features were calculated and studied in present study. Eventually, 1906 radiomics features were obtained from every MRI phase for each lesion. The Z-score was standardized for the extracted features.
3. ICC test: The reproducible features were set as the features with an intra- and inter-observer ICC greater than 0.8 (27). These robust features were applied in the following feature selection.
4. Feature selection: The linear correlation coefficient between characteristics was initially estimated, and one of them was deleted when it was ≥ 0.65. Furthermore, we applied the analysis of variance (ANOVA) F-test to each feature and label pair to conduct feature selection. Based on the F-value ranking, the top 14 features were adopted as meaningful predictors in the subsequent logistic regression.

**Model Building**
For additional radiomics signature development, logistic regression was used for radiomics characteristics determined by the F-test. For the optimal parameter selection in the training dataset, we utilized ten-fold cross-validation. The radiomics signature was created via the parameters that produced the best average area under the curve (AUC). The phases with AUCs larger than 0.700 were judged noteworthy and hence were chosen for the development of fusion radiomics signatures.

A likelihood ratio backward stepwise multivariable logistic regression model that combined clinical factors with radiomics signature was constructed to investigate whether these two variables enhanced prediction of VEGF status. Clinical variables with a significant correlation with VEGF, as well as radiomics signatures, were added into the multivariable combined model. The discriminatory ability of classifier model was analyzed by the AUCs, and the Delong test was used to compare AUCs. The Hosmer-Lemeshow test was used to determine the calibration performance between the combined model’s projected probability and the actual VEGF status.

Furthermore, we established a nomogram relying on a combined model that can simply calculate the likelihood of VEGF status. The clinical utility of the nomogram was appraised by means of decision curve analysis (28). By measuring the disparity between the true- and false-positive rate at different thresholds, net benefits were calculated (29). Figure 2 describes a diagram of a radiomics investigation.

**Statistical Analysis**
We used Student’s t-test or Mann-Whitney U test for continuous variables and Chi-Square test or Fisher’s exact test for qualitative variables in training and test datasets. The statistical analyses were carried out using IBM SPSS Statistics 22.0 and MedCalc 20 software (MedCalc Software, Belgium). Statistical significance was defined as a two-tailed P-value < 0.05.

**RESULTS**

**Clinicopathologic Characteristics and Clinical Modeling**
In Table 1, demographics, laboratory results, and morphological characteristics of both the training and tested groups are summarized and compared. 202 individuals (169 male and 33 female) with solitary HCC were split into a training group (n = 142, 76 for VEGF positive, 66 for VEGF negative; 115 male and 27 female) and a testing group (n = 60, 32 for VEGF positive, 28 for VEGF negative; 54 male and 6 female).

Across the full cohort, there were 108 (53.5%) patients who were VEGF-positive and 94 (46.5%) patients who were VEGF-negative. VEGF status was not significantly different between the training and test datasets (p = 0.980). In the training cohort,
univariate analysis revealed that serum α-fetoprotein (AFP), irregular margin, and neutrophil-to-lymphocyte ratio (NLR) were significantly associated with VEGF (P < 0.05) (Table 1). However, age, gender, tumor diameter, etiology of liver disease, cirrhosis, ascites, ALT, AST, TBIL, neutrophil count, lymphocyte count, albumin, differentiation degree, arterial rim enhancement, tumor capsule, and enhancement pattern were not statistically significant (P > 0.05). A backward stepwise multivariable logistic regression was used to examine every notable variable. Eventually, the meaningful variables for clinical model establishing were serum AFP level (odds ratio [OR] = 0.260; 95% confidence interval [CI]: 0.107 - 0.633, P = 0.003), irregular tumor margin (p = 0.004, OR = 3.004, 95% CI: 1.434 - 6.295), and NLR (p = 0.027, OR = 0.622, 95% CI: 0.409 - 0.948) (Table 2). In the training cohort, the clinical model’s AUC was 0.709 (95% CI: 0.624 - 0.794) while it was 0.725 (95% CI: 0.593 - 0.858) in the test cohort.

**Feature Selection and Radiomics Model Development**

The features generated from arterial phase, PVP, balanced phase, delayed phase, and HBP were decreased to 1349, 1252, 1850, 1816, and 1202, respectively, after feature selection through the ICC test. Those robust features were applied for further analysis. Table 3 displays the forecast results for each of the five phases. Finally, the test set’s median 10-fold cross-validation AUC for PVP, HBP, AP, BP, and DP was 0.759, 0.731, 0.640, 0.592, and 0.692, respectively. The radiomics signatures of PVP and HBP (> 0.700) were captured to investigate further. Figure S1 presents the stability of radiomics characteristics in two phases (PVP and HBP) images. After F-test modeling for the prediction of VEGF-positive, 8 robust features in the PVP and 5 robust features in the HBP were regarded as the optimal feature set. The selected features were then used to create a logistic regression classifier model. Tables S2, S3 provide more detailed information on the features. In the training cohort, the radiomics signature in the PVP obtained an AUC of 0.809 (95% CI: 0.740 - 0.878) while in the test dataset, it had an AUC of 0.759 (95% CI: 0.630 - 0.888). Likewise, in the training and test groups, the radiomics signature of HBP obtained an AUC of 0.792 (95% CI: 0.716 - 0.868) and 0.731 (95% CI: 0.597 - 0.865), respectively.

Furthermore, we propose a multiphase fusion model that combines the radiomics signatures (PVP and HBP) with a multivariable logistic regression model to study the maximum potential utilization of both radiomics signatures in diverse MRI phases. The fusion radiomics signature performed well in training and test groups, with AUCs of 0.892 (95% CI: 0.839 -
### TABLE 1 | Patient characteristics in the training and test cohorts.

| Characteristics                          | Training dataset (n = 142) | P value | Test dataset (n = 60) | P value | P value* |
|------------------------------------------|----------------------------|---------|-----------------------|---------|----------|
|                                          | VEGF(+) (n = 76)           |         | VEGF(-) (n = 66)      |         |          |
| Age, mean ± SD, years                    | 55.72 ± 8.12               | 0.244   | 56.34 ± 7.12          | 0.168   | 0.809    |
| Gender, n (%)                            | 0.847                      |         | 0.796                 | 0.119   |          |
| Male                                     | 62 (81.6)                  |         | 28 (87.5)             | 0.260   | 0.637    |
| Female                                   | 14 (18.4)                  |         | 4 (12.5)              | 0.260   | 0.637    |
| Diameter, median (IQR), mm               | 35.50 (27.50,49.75)        | 0.067   | 44.00 (30.25,55.75)   | 0.876   | 0.401    |
| Etiology of liver disease                | 0.436                      |         | 0.377                 | 0.809   |          |
| HBV positive a                           | 67 (88.2)                  | 0.001   | 28 (87.5)             | 0.260   | 0.637    |
| HCV positive b                           | 1 (1.3)                    |         | 0 (0)                 | 0.260   | 0.637    |
| None or other                            | 8 (10.5)                   |         | 4 (12.5)              | 0.260   | 0.637    |
| Cirrhosis, n (%)                         | 0.184                      |         | 0.796                 | 0.613   |          |
| Present                                  | 55 (72.4)                  |         | 26 (81.3)             | 0.830   | 0.167    |
| Absent                                   | 21 (27.6)                  |         | 6 (18.8)              | 0.830   | 0.167    |
| Ascites, n (%)                           | 0.995                      |         | 3 (10.7)              | 0.830   | 0.167    |
| Present                                  | 15 (19.7)                  |         | 4 (12.5)              | 0.830   | 0.167    |
| Absent                                   | 61 (80.3)                  |         | 28 (87.5)             | 0.830   | 0.167    |
| ALT (U/L)                                | 28.95 (16.25,45.68)        | 0.305   | 29.00 (16.25,52.50)   | 0.583   | 0.793    |
| AST (U/L)                                | 30.00 (21.50,40.00)        | 0.112   | 30.00 (20.50,51.25)   | 0.320   | 0.901    |
| TBIL (μmol/L)                            | 15.15 (11.25,20.57)        | 0.756   | 14.30 (11.90,18.40)   | 0.543   | 0.817    |
| Neutrophil count (×10^9/L)               | 1.79 ± 0.63                | 0.315   | 1.97 ± 0.74           | 0.543   | 0.277    |
| Lymphocyte count (×10^9/L)               | 3.07 (2.20,3.88)           | 0.081   | 2.41 (1.88,3.39)      | 0.251   | 0.737    |
| Albumin (g/L)                            | 37.96 ± 4.17               | 0.720   | 38.18 ± 3.17          | 0.450   | 0.769    |
| Serum AFP, n (%)                         | 0.001                      |         | 0.830                 | 0.066   |          |
| ≤400 ng/mL                               | 66 (86.8)                  |         | 62 (81.6)             | 0.352   | 0.066    |
| >400 ng/mL                               | 10 (13.2)                  |         | 14 (18.4)             | 0.352   | 0.066    |
| Differentiation degree, n (%)            | 0.166                      |         | 0.722                 | 0.208   |          |
| Well                                     | 12 (15.8)                  |         | 5 (15.6)              | 0.722   | 0.208    |
| Moderate                                 | 51 (67.1)                  |         | 19 (59.4)             | 0.722   | 0.208    |
| Poor                                     | 13 (17.1)                  |         | 8 (25.0)              | 0.722   | 0.208    |
| Morphologic MR features                  | 0.015                      |         | 0.001                 | 0.336   |          |
| Irregular margin on HBP, n (%)           |                           |         | 0.726                 | 0.245   |          |
| Absence                                  | 25 (32.9)                  | 0.309   | 5 (15.6)              | 0.726   | 0.245    |
| Presence                                 | 51 (67.1)                  | 0.309   | 27 (83.4)             | 0.726   | 0.245    |
| Arterial rim enhancement, n (%)          |                           |         | 0.821                 | 0.793   |          |
| Presence                                 | 48 (63.2)                  | 0.879   | 18 (56.3)             | 0.821   | 0.793    |
| Absence                                  | 28 (36.8)                  | 14 (43.8) | 11 (39.3)            | 0.821   | 0.793    |
| Tumor capsule, n (%)                     | 0.879                      |         | 0.821                 | 0.793   |          |
| Complete                                 | 55 (72.4)                  |         | 22 (68.8)             | 0.821   | 0.793    |
| Incomplete or absent                     | 21 (27.6)                  |         | 10 (31.3)             | 0.821   | 0.793    |
| Enhancement pattern, n (%)               | 0.678                      |         | 0.768                 | 0.713   |          |
| Arterial enhancement with washout        | 62 (81.6)                  | 0.678   | 26 (81.3)             | 0.768   | 0.713    |
| No or minimal enhancement                | 6 (7.9)                    |         | 3 (9.4)               | 0.768   | 0.713    |
| Persistent enhancement                   | 6 (7.9)                    |         | 2 (6.3)               | 0.768   | 0.713    |
| Progressive enhancement                  | 2 (2.6)                    |         | 1 (3.1)               | 0.768   | 0.713    |

**VEGF (+), VEGF-positive; VEGF (-), VEGF-negative; ALT, alanine aminotransferase; AST, aspartate aminotransferase; TBIL, total bilirubin; NLR, neutrophil-to-lymphocyte ratio; AFP, α-fetoprotein; HBP, hepatobiliary phase; SD, standard deviation; IQR, interquartile range. *Represents the comparisons of characteristics between training and test dataset. Data are mean ± SD, median (IQR) or n (%), where n is the number of participants for whom data is available. aRepresents positivity for hepatitis B serum antigen. bRepresents positivity for serum HCV antibody.**

### TABLE 2 | Univariate and multivariate assessments of variables associated with VEGF levels in clinical model.

| Characteristics                   | Univariate OR (95%CI) | P value | Multivariate OR (95%CI) | P value |
|-----------------------------------|-----------------------|---------|-------------------------|---------|
| Age                               | 1.023 (0.985 - 1.062) | 0.243   | 1.023 (0.985 - 1.062)   | 0.243   |
| Gender                            | 0.921 (0.938 - 2.131) | 0.847   | 0.921 (0.938 - 2.131)   | 0.847   |
| NLR                               | 0.625 (0.423 - 0.924) | 0.019   | 0.625 (0.423 - 0.924)   | 0.019   |
| Irregular margin on HBP           | 2.303 (1.167 - 4.547) | 0.016   | 2.303 (1.167 - 4.547)   | 0.016   |
| Serum AFP level                   | 0.265 (0.115 - 0.610) | 0.002   | 0.265 (0.115 - 0.610)   | 0.002   |

**AFP, α-fetoprotein; NLR, neutrophil-to-lymphocyte ratio; CI, confidence interval; HBP, hepatobiliary phase; OR, odds ratio.**
0.945) and 0.800 (95% CI: 0.682 - 0.918), respectively. When compared to the clinical model, the difference was significant in the training (p = 0.0003) cohort; however, the p value in the test dataset was 0.3759. We also noticed substantial variations in fusion signatures between VEGF (+) and VEGF (-) patients, and the p values of both (training and test) groups were less than 0.001.

**Combined Model Building**

The combined model performed best for VEGF prediction, combining all potential parameters including the fusion radiomics signature and major clinical characteristics (irregular tumor margin and serum AFP levels). Compared with the clinical model, the combined model represents superior performance in training [0.936(0.898 - 0.974) vs. 0.709(0.624-0.794); p < 0.0001] and test [0.836(0.728-0.944) vs. 0.725(0.593-0.858); p = 0.093] cohorts, albeit the difference was not statistically meaningful (p = 0.093) in test datasets. Furthermore, the accuracies of the combined model for forecasting the VEGF were 86.6% (training group) and 71.7% (test group) (Table 3).

Comparisons among AUCs between every two model were showed in Table S4. The stratified analysis also revealed strong results in both age and gender categories (Table S5). Figure 3 displays the three ROC curves for different models. Figure 4 depicts the created nomogram. The calibration curves are displayed in Figure 5, and there is high conformity in the two groups (training: P = 0.820; test: P = 0.508). Figure 6 depicts the findings of the decision curve analysis (DCA). The DCA for combined model indicated that when the probability of VEGF-positive was between 0.060-0.976 and 0.250-0.823 in the training and test datasets, respectively, using the nomogram to evaluate pathological VEGF-positive had added benefit compared with treating all patients as positive or negative. The coefficients and relative weights of 13 selected radiomics features of logistic regression in combined model are displayed on Table S6. A correlation heat map of the selected 13 radiomics features in the combined model is depicted on Figure S2.

**DISCUSSION**

To detect VEGF-positive HCC preoperatively, we created a predictive nomogram using MRI (Gd-EOB-DTPA-enhanced) radiomics signatures, serum AFP levels, and irregular tumor margins. The combined model revealed adequate diagnostic capacity in stratifying HCCs based on VEGF status. The fusion radiomics signature outperformed radiomics signatures derived independently in PVP or HBP, with AUCs of 0.892 in training and 0.800 in test datasets for detecting VEGF-positive HCCs. The combined model attained the greatest AUC of 0.936 in the training cohort and an AUC of 0.836 in the test cohort after adding clinical risk factors and fusion imaging biomarkers. Previous research discovered that high levels of VEGF in HCC lesions were substantially related to metastatic recurrence (30). So far as we know, though, no studies have been published that have utilized a nomogram to forecast the likelihood of VEGF positivity in HCC.

In this study, radiomic features of the three-dimensional (3D) primary tumor were utilized to predict VEGF-positive HCC. The 3D tumor volume may give more precise tumor heterogeneity information as well as full size and shape information (31, 32). Qualitative and quantitative analysis using Gd-EOB-DTPA-enhanced MRI imaging can help characterize VEGF-positive HCCs. Despite the availability of different MRI sequences, only 8 and 5 radiomics characteristics in PVP and HBP images were retrieved for the greatest prediction performance. In our single-

| Model | Training dataset (n = 142) | Test dataset (n = 60) |
|-------|----------------------------|----------------------|
|       | AUC (95% CI) | PNP | SENS | SPEC | ACC | PPV | NPV | AUC (95% CI) | PNP | SENS | SPEC | ACC | PPV | NPV |
| Clinical model | 0.709 (0.624-0.794) | 92 | 0.684 | 0.606 | 0.648 | 0.667 | 0.625 | 0.725 (0.593-0.858) | 41 | 0.656 | 0.714 | 0.683 | 0.724 | 0.645 |
| Radiomics model of different MRI phase |                     |     |      |      |     |     |     |                     |     |      |      |     |     |     |
| AP    | 0.709 (0.622-0.797) | 98 | 0.711 | 0.667 | 0.690 | 0.711 | 0.667 | 0.640 (0.497-0.782) | 37 | 0.531 | 0.714 | 0.617 | 0.680 | 0.571 |
| PVP   | 0.809 (0.740-0.878) | 99 | 0.724 | 0.667 | 0.697 | 0.714 | 0.677 | 0.759 (0.630-0.888) | 38 | 0.625 | 0.643 | 0.633 | 0.667 | 0.600 |
| BP    | 0.748 (0.666-0.830) | 96 | 0.658 | 0.697 | 0.676 | 0.714 | 0.639 | 0.592 (0.437-0.746) | 32 | 0.438 | 0.643 | 0.533 | 0.583 | 0.500 |
| DP    | 0.669 (0.578-0.757) | 87 | 0.618 | 0.606 | 0.613 | 0.644 | 0.580 | 0.692 (0.554-0.830) | 35 | 0.406 | 0.786 | 0.583 | 0.684 | 0.537 |
| HBP   | 0.792 (0.716-0.868) | 109 | 0.763 | 0.773 | 0.768 | 0.795 | 0.739 | 0.731 (0.597-0.865) | 39 | 0.500 | 0.821 | 0.650 | 0.762 | 0.590 |
| PVP + HBP | 0.892 (0.839-0.945) | 120 | 0.803 | 0.894 | 0.845 | 0.897 | 0.797 | 0.800 (0.682-0.918) | 43 | 0.594 | 0.857 | 0.717 | 0.826 | 0.649 |
| Combined model | 0.936 (0.898-0.974) | 123 | 0.855 | 0.879 | 0.866 | 0.890 | 0.841 | 0.836 (0.728-0.944) | 43 | 0.625 | 0.821 | 0.717 | 0.800 | 0.657 |

ACC: accuracy; AP, arterial phase; AUC, area under curve; BP, balanced phase; HBP, hepatobiliary phase; DP, delayed phase; SPEC, specificity; SENS, sensitivity; NPV, negative predictive value; PNP, predicted number of patients correctly classified; PPV, positive predictive value; PVP, portal venous phase; PVP + HBP, a fusion radiomics model integrating the hepatobiliary and portal venous phases.
phase study, the predictive performance of the model for the PVP or HBP was better than that for the AP. Of interest, it has been previously reported that the same two phases (PVP and HBP) pictures of Gd-EOB-DTPA-enhanced MRI may be for the diagnosis of early HCC when arterial enhancement lack (33). Their results suggest that changes in hepatocyte function may precede significant vascular changes in early hepatocellular carcinoma. It could be speculated that such a pattern may be related to the development of subsequent arterial hyperenhancement. To our knowledge, VEGF expression in hepatocyte precedes the changes in angiogenesis in the tumor microenvironment. The low diagnostic performance of the radiomics model obtained in the AP may be due to the immaturity of tumor vasculature. PVP and HBP may reflect the functional changes of hepatocytes earlier than AP, so that we can find more information related to hepatocyte function from the characteristics and then better predict VEGF expression than in AP. Furthermore, we discovered two features, “wavelet-LLH_firstorder_Range” and “square_grlm_RunEntropy” that corresponded to the biological characteristics. “Wavelet-LLH_firstorder_Range” represents the range of gray values in the ROI.
which is derived from imaging after wavelet filtering. “Square_gdlm_RunEntropy” quantifies the degree of uncertainty or randomness in the distribution of run lengths and gray levels produced from imaging after square filtering. A greater value suggests that the texture patterns are more heterogeneous. Intratumoral heterogeneity may be related to the difference between VEGF positive and VEGF negative HCC (e.g., angiogenesis, invasion, and apoptosis) (34–36).

There was no considerable difference in the clinicopathologic characteristics of VEGF-positive HCCs between the training and test populations. In existing researches (30, 37), the value of VEGF expression correlated positively with tumor size and AFP level in tumor tissue. However, in our study, there was no effect of tumor size on VEGF expression in HCCs. This may relate to the sample size and sampling strategy. Although there were no significant statistical differences in our study, VEGF-positive HCCs were more prone to having an invasive character, defined as low-differentiated and bigger tumor size (30, 38).

Preoperative serum AFP level may be used as a predictive marker in the present study. In clinical practice, preoperative serum AFP level is readily available.

Our findings indicated that an irregular margin of tumor was closely tied with VEGF expression. The irregular tumor margin of preoperative MRI imaging features could identify high-risk early recurrence patients with early-stage hepatocellular carcinoma (39). The more irregular the margin, the greater the surface region; therefore, it may demonstrate the aggressiveness of the tumor (40). The invasive edge is a notable location for tumor angiogenesis activation, which leads to tumor invasion and metastasis (41).

The combined model, which included a radiomics signature and clinical risk indicators, significantly increased performance. This showed that radiomics signatures and clinical risk factors may be used in tandem. Finally, for clinical application, we created a nomogram, which was a simple tool for calculating the likelihood of VEGF positivity. Patients may be classified as low-
risk or high-risk based on the nomogram’s predicted likelihood. A previous study revealed that bevacizumab can target VEGF isoforms and inhibit their interaction with the VEGF receptor (15). In particular, it has been recently reported that among the tyrosine kinase inhibitors (TKI), regorafenib is a drug targeting all of the VEGF receptors (VEGFR1-3) (42). The application of these targeted anti-angiogenesis drugs to patients with a high expression level of VEGF can effectively inhibit angiogenesis in patients with HCC. And therefore this proposed nomogram could help clinicians choose effective systemic treatment options for cancer management. It can not only eliminate unnecessary medical care and expenditures for low-risk individuals, but also significantly minimize patient side effects.

Certainly, some limitations of this study deserve to be mentioned. Firstly, because this was a retrospective investigation, several confounding variables may have existed. Secondly, as this was a single-centre study. Therefore, our results need to be tested by multicenter studies. Thirdly, the genetic factors associated with VEGF were not included, which might give valuable new information for VEGF prediction. Finally, it is worth noting that our study did not include multimodal radiological data. In the future, MRI, computed tomography (CT), ultrasonography (US), and positron emission tomography (PET) data may be used to investigate the status of VEGF.

CONCLUSION

HBP and PVP radiomics signatures obtained from Gd-EOB-DTPA-enhanced MRI imaging could assist in predicting the VEGF levels of HCC. In the VEGF stratification of HCC, a forecasting nomogram integrating MRI radiomics characteristics, irregular tumor margins, and serum AFP levels revealed considerably enhanced diagnostic performance.

DATA AVAILABILITY STATEMENT

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

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Conflict of Interest: Author JX is employed by Beijing Deepwise & League of PHD Technology Co., Ltd.

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