Effect of almond consumption on the serum fatty acid profile: a dose–response study

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Abstract

Consumption of almonds has been shown to be associated with a decreased risk of CHD, which may be related to their fatty acid (FA) composition. However, the effect of almond consumption on the serum FA composition is not known. Therefore, in the present study, we investigated whether almond consumption would alter the serum FA profile and risk of CHD, as calculated using Framingham’s 10-year risk score, in a dose-dependent manner in hyperlipidaemic individuals when compared with a higher-carbohydrate control group using dietary interventions incorporating almonds. A total of twenty-seven hyperlipidaemic individuals consumed three isocaloric (mean 1770 kJ/d) supplements during three 1-month dietary phases: (1) full-dose almonds (50–100 g/d); (2) half-dose almonds with half-dose muffins; (3) full-dose muffins. Fasting blood samples were obtained at weeks 0 and 4 for the determination of FA concentrations. Almond intake (g/d) was found to be inversely associated with the estimated Framingham 10-year CHD risk score (P=0·026). In both the half-dose and full-dose almond groups, the proportions of oleic acid (OA) and MUFA in the TAG fraction (half-almond: OA P=0·003; MUFA P=0·004; full-almond: OA P<0·001; MUFA P<0·001 and in the NEFA fraction (half-almond: OA P=0·01; MUFA P=0·04; full-almond: OA P=0·12; MUFA P=0·06) increased. The estimated Framingham 10-year CHD risk score was inversely associated with the percentage change of OA (P=0·011) and MUFA (P=0·016) content in the TAG fraction. The proportions of MUFA in the TAG and NEFA fractions were positively associated with changes in HDL-cholesterol concentrations. Similarly, the estimated Framingham 10-year CHD risk score was inversely associated with the percentage change of OA (P=0·009) and MUFA content in the NEFA fraction (P=0·009). In conclusion, the results of the present study indicate that almond consumption increases OA and MUFA content in serum TAG and NEFA fractions, which are inversely associated with CHD lipid risk factors and overall estimated 10-year CHD risk.

Key words: Coronary/heart disease; Fatty acids; Nutrition; Almonds

Epidemiological1–4 and clinical studies5,6 have shown that nut consumption decreases the risk of developing CHD and improves blood lipid risk factors. Almond consumption has been shown to reduce LDL-cholesterol (LDL-C) concentrations and increase HDL-cholesterol (HDL-C) concentrations in hyperlipidaemic individuals7,8. Although almonds contain a variety of constituents that may exert cardioprotective effects through various mechanisms9, their ability to improve blood lipid profiles and reduce CHD risk may primarily be related to their healthy fatty acid (FA) profile, which is low in saturated fats (SFA) and high in monounsaturated fats (MUFA)10,11, particularly oleic acid (OA)12,13. Epidemiological studies suggest that diets with a high proportion of MUFA in the form of OA, such as the Mediterranean diet, reduce

Abbreviations: FA, fatty acids; HDL-C, HDL-cholesterol; LDL-C, LDL-cholesterol; OA, oleic acid; PREDIMED, Prevenci´ n con Dieta Mediterranea.

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CHD risk\(^{(14,15)}\). However, the effect of almond consumption on the serum FA profile and its association with CHD risk have not been investigated.

Therefore, in the present study, we investigated whether almond consumption would alter the serum FA profile and risk of CHD, as calculated using Framingham’s 10-year risk score, in a dose-dependent manner in hyperlipidaemic individuals compared with a higher-carbohydrate control group using dietary interventions incorporating almonds\(^{(7)}\). This is a secondary analysis of a trial that primarily aimed to assess the effect of almond consumption on blood lipid risk factors for CHD\(^{(7)}\).

**Methods**

The study methods have been described in detail previously\(^{(7)}\).

**Study protocol**

A total of twenty-seven subjects completed three 1-month dietary phases in a randomised cross-over manner, with each phase being separated by a washout period of minimum 2 weeks. The three dietary phases comprised a muffin phase (control) and two almond phases: full-dose almond phase and half-dose almond plus half-dose muffin phase. During all the dietary phases, the subjects followed their own self-selected, low-fat therapeutic diets into which they incorporated the supplements. The subjects were counselled on strategies to facilitate weight maintenance.

After an overnight fast (12–14 h), body weight measurements, blood samples and blood pressure measurements were obtained at the start of the study and at weeks 2 and 4 of each 4-week dietary phase. Before baseline and at week 4 of each dietary phase, 7 d diet records were obtained from the subjects. The subjects were instructed to weigh all the food consumed with the self-rating electronic food scales provided during the weeks when dietary intake was recorded.

The present study was conducted according to the guidelines laid down in the Declaration of Helsinki, and all procedures involving human subjects were approved by the Ethics Committee of the University of Toronto and St Michael’s Hospital. Written informed consent was obtained from all subjects. The present trial was registered at ClinicalTrials.gov (no. NCT00507520).

**Study subjects**

Healthy hyperlipidaemic men and postmenopausal women were recruited through newspaper advertisement and from among patients visiting the Risk Factor Modification Centre of St Michael’s Hospital, a University of Toronto teaching hospital. As has been reported previously\(^{(7)}\), sixteen of the forty-three subjects who were initially recruited withdrew during or after completing one or two dietary phases: three discontinued for reasons directly related to the study (food allergies, \(n\) 2; abdominal discomfort, \(n\) 1) and thirteen withdrew for unrelated reasons. The characteristics of these sixteen subjects were similar to those of the subjects who completed the study. The remaining twenty-seven subjects completed all the three dietary phases: fifteen men and twelve postmenopausal women (mean age 64 (SD 9) years (range: 48–86 years; four subjects aged ≥75 years); mean BMI 25·7 (SD 3·0) kg/m\(^2\) (range: 20·5–31·5 kg/m\(^2\)) and mean baseline LDL-C concentrations 4·32 (SD 0·63) mmol/l (range 2·77–5·32 mmol/l)). All subjects had elevated LDL-C concentrations during initial assessment at recruitment (>4·1 mmol/l), despite lower values being detected in some subjects at baseline, and TAG concentrations <1·0 mmol/l. None of the subjects used tobacco and had clinical or biochemical evidence of diabetes, liver disease or renal disease. Among the twenty-seven subjects, three men and five women were taking the following medications: a hypolipidaemic agent (statin) (\(n\) 2); \(\beta\)-blocking agents (\(n\) 3); angiotensin-converting enzyme inhibitors (\(n\) 3); angiotensin II receptor blockers (\(n\) 1); thiazide diuretics (\(n\) 2); levothyroxine (\(n\) 2); hormone replacement therapy (\(n\) 2). Medication use was stable for at least 2 weeks before the study. Medication dosages were kept constant throughout the study. The subjects were asked to maintain their habitual levels of physical activity throughout the study.

**Diets**

Before the start of the study, all subjects had been instructed to follow a therapeutic National Cholesterol Education Program Step 2 diet (<7% energy from saturated fats and <200 mg/d dietary cholesterol). All subjects took all the three supplements: whole raw unblanched almonds (73 (SD 3) g/d); muffins (147 (SD 6) g/d); and half portions of almonds (37 (SD 2) g/d) plus muffins (75 (SD 3) g/d), as described previously\(^{(10)}\). The amount of supplements ingested was based on the estimated daily energy requirement of the subjects\(^{(10)}\). For subjects requiring <6694 kJ/d (<1600 kcal/d), the full portions of almonds and muffins were 50 g (one package) and 100 g (two muffins), respectively (1201 kJ/d (287 kcal/d) supplement). The respective values for those requiring 6694–10 042 kJ/d (1600–2400 kcal/d) were 75 g of almonds and 150 g of muffins (three muffins; 1799 kJ/d (430 kcal/d) supplement) and for those requiring >10 042 kJ/d (>2400 kcal/d) were 100 g of almonds and 200 g of muffins (four muffins; 2402 kJ/d (574 kcal/d) supplement). During the half-almond plus half-muffin phase, the subjects consumed each supplement at half the full amount for each of the energy requirements described above. The muffins were made from whole-wheat flour with an amount of maize oil sufficient to provide the same amounts of SFA, PUFA and fibre provided by almonds. Skimmed milk powder and egg-whites were used to maintain the level of protein between the supplements, although the protein in the muffins was 46% animal origin. MUFA (kJ) in almonds balanced the starch in muffins. The macronutrient composition of the muffins as a percentage of energy was as follows: 14·7% protein; 55·3% available carbohydrate; 32·1% fat; 4·3% SFA; 7·6% MUFA; 18·9% PUFA with 4·3 g/1000 kJ (18 g/1000 kcal) cholesterol. Non-hydrogenated maize oil was the only fat used in muffin preparation to prevent the addition of trans-FA to the
supplements. Muffin supplements were provided to the subjects at biweekly intervals and were stored in a freezer until the day before use. The subjects were instructed to reduce their total food intake, especially starchy foods (breads, bagels, non-study muffins and breakfast cereals), to allow supplements to be incorporated in their diet without increasing total energy intake and to keep the background diet constant across all the three dietary phases. To minimise changes in body weight and diet composition, the subjects were given detailed dietary counselling before and at weeks 1 and 2 of each dietary phase. During the study, the subjects were asked to not consume any additional nuts or nut products or alter their intake of dietary fibre or vegetable protein foods. Compliance was assessed from 7 d diet records, a supplement checklist in which the subjects recorded the supplements consumed, and return of uneaten supplements, which were weighed and recorded.

**analyses**

Serum was analysed according to the Lipid Research Clinics protocol(17,18) for total cholesterol, TAG and LDL-C after dextran sulphate–magnesium chloride precipitation, and LDL-C concentrations were calculated(19) as described previously(20). The present study focused on serum FA concentrations and estimated 10-year CHD risk based on the Framingham equation(16). The following variables were used in the estimated 10-year CHD risk at baseline and end of each intervention period: age; sex; systolic blood pressure; total cholesterol; HDL-C; total cholesterol:HDL-C ratio; smoking status; diabetes diagnosis. Values obtained for the 10-year CHD risk were then compared with those of the ideal risk, where total cholesterol:HDL-C ratio was set at 4·44, systolic blood pressure at 120 mmHg, and smoking status as non-smoker, to obtain relative risk values.

To assess FA fractions, total lipids were extracted from the serum according to the Folch method(21), TLC plates were activated by heating at 100°C for 1 h. Total lipids were then loaded onto the plates and placed in a tank with solvents. FA fractions were separated along with authentic standards in heptane–dichloromethane–2-propanol (80:15:5, v/v). Bands corresponding to total phospholipid, NEFA, TAG and cholesterol esters were visualised under UV light, after staining with 8-anilino-1-naphthalene sulphonic acid (0·1 %, w/v). Known amounts of an unesterified heptadecanoic acid standard were added to test-tubes containing hexane and each scraped band. These were converted to FA methyl esters with 14 % boron trifluoride–methanol at 100°C for 1 h. FA methyl esters were quantified on a Varian-430 gas chromatograph (Varian, Inc.) equipped with a Varian Factor Four capillary column (VF-23ms, 30 m x 0·25 mm inner diameter x 0·25 µm film thickness; Agilent Technologies) and a flame ionisation detector. Samples were injected in splitless mode. The temperature of the injector and detector ports was set at 250°C. FA methyl esters were eluted using a temperature programme set initially at 50°C for 2 min, increased at 20°C/min and held at 170°C for 1 min and then at 3°C/min and held at 212°C for 5 min to complete the run in 28 min.

The carrier gas used was helium, set to a constant flow rate of 0·7 ml/min. Peaks were identified by the retention times of FA methyl ester standards (Nu-Chek-Prep, Inc.). FA concentrations (nmol/ml serum) were calculated by proportional comparison of GC peak areas with that of the heptadecanoic acid internal standard. Plasma SFA, MUFA and PUFA concentrations were calculated by adding the concentrations of all the respective FA with twelve to twenty-four carbon atoms 20:0 or 18:3n-3, as these FA did not get separated in the column.

**statistical analyses**

Results are expressed as means with their standard errors. The primary outcomes of the original study were total cholesterol, LDL-C, HDL-C, TAG, apoB and apoA1(7). The present study focused on FA fractions in the serum of subjects who completed the study (n 27) and estimated CHD risk using the Framingham equation(16).

Treatment effects were tested using Student’s t test for within-group analysis and one-way repeated-measures ANOVA for between-group analysis (Microsoft Excel, 2007). Pearson’s correlation analyses were conducted to test the associations between FA concentrations and CHD risk (SAS Institute Inc., version 9.3). Statistical significance was set at P<0·05 for all analyses.

**results**

**subject characteristics**

The baseline characteristics of the subjects, as well as data obtained at baseline and after the dietary interventions, have been reported previously(7). There were no differences in the effects of dietary treatments on the anthropometric or lipid profiles analysed between those who completed the study and those who did not (see Supplementary table, available online).

**table 1. Fatty acid composition of supplements used in the study**

| Fatty acids (%) | Supplements | Mean | SEM | Mean | SEM |
|----------------|-------------|------|-----|------|-----|
|                | Muffins (n 3) | Almonds (n 3) |
| 14:0           | 0·3 ± 0·1    | 0·1 ± 0·0 |
| 14:1n5         | 0·0 ± 0·0    | 0·0 ± 0·0 |
| 16:0           | 16·3 ± 0·5   | 9·6 ± 0·0 |
| 16:1n7         | 0·2 ± 0·1    | 0·6 ± 0·0 |
| 18:0           | 0·2 ± 0·1    | 1·7 ± 0·0 |
| 18:1n9         | 21·8 ± 0·8   | 64·3 ± 0·0 |
| 18:1n7         | 0·5 ± 0·0    | 1·3 ± 0·0 |
| 18:2n6         | 57·0 ± 0·1   | 23·6 ± 0·0 |
| 18:3n6         | 0·0 ± 0·0    | 0·0 ± 0·0 |
| 20:0/18:3n-3   | 1·7 ± 0·0    | 0·0 ± 0·0 |
| 20:1n9         | 0·2 ± 0·0    | 0·0 ± 0·0 |
| Total (mg/g)   | 9·4 ± 1·1    | 24·8 ± 1·1 |

* Mean values were significantly different from that of the muffin-supplemented group (P<0·05).
Table 2. Effect of dietary treatments on the proportion of fatty acids in the serum phospholipid fraction (Mean values with their standard errors)

| Fatty acids (%) | Control (n=27) | Half-dose almonds (n=27) | Full-dose almonds (n=27) |
|-----------------|----------------|-------------------------|-------------------------|
|                 | Week 0         | Week 4                  | Week 0                  | Week 4                  | Week 0                  | Week 4                  |
| 14:0            | 0.55 ± 0.03    | 0.56 ± 0.03             | 0.64 ± 0.04             | 0.56* ± 0.05            | 0.56 ± 0.03             | 0.57 ± 0.03             |
| 14:1n-5         | 0.03 ± 0.01    | 0.03 ± 0.01             | 0.19 ± 0.14             | 0.03 ± 0.01             | 0.03 ± 0.01             | 0.07 ± 0.03             |
| 16:0            | 35.57 ± 0.50   | 35.17 ± 0.51            | 35.84 ± 0.52            | 34.93 ± 0.66            | 35.04 ± 0.41            | 34.51 ± 0.45            |
| 16:1n-7         | 0.80 ± 0.06    | 0.75 ± 0.04             | 0.97 ± 0.15             | 0.72 ± 0.05             | 0.76 ± 0.04             | 0.76 ± 0.05             |
| 18:0            | 14.62 ± 0.23   | 14.71 ± 0.22            | 14.51 ± 0.25            | 14.19 ± 0.24            | 14.75 ± 0.27            | 14.39 ± 0.23            |
| 18:1n-9         | 10.28 ± 0.24   | 10.52 ± 0.22            | 10.27 ± 0.24            | 10.64* ± 0.22           | 9.96 ± 0.24             | 10.80* ± 0.20           |
| 18:2n-6         | 1.59 ± 0.05    | 1.69 ± 0.05             | 1.60 ± 0.05             | 1.64 ± 0.06             | 1.62 ± 0.05             | 1.94 ± 0.36             |
| 18:3n-6         | 0.12 ± 0.02    | 0.20 ± 0.07             | 0.15 ± 0.06             | 0.14 ± 0.06             | 0.16 ± 0.06             | 0.85 ± 0.71             |
| 20:0/18:3n-3    | 0.44 ± 0.04    | 0.47 ± 0.04             | 0.45 ± 0.04             | 0.44 ± 0.04             | 0.42 ± 0.04             | 0.39 ± 0.04             |
| 20:1n-9         | 0.14 ± 0.01    | 0.16 ± 0.01             | 0.13 ± 0.01             | 0.17* ± 0.01            | 0.13 ± 0.01             | 0.17 ± 0.02             |
| 20:3n-3†        | 0.29 ± 0.02    | 0.32* ± 0.01            | 0.30 ± 0.01             | 0.27 ± 0.02             | 0.30 ± 0.01             | 0.29 ± 0.01             |
| AA (20:4n-6)    | 9.45 ± 0.28    | 9.02 ± 0.02             | 9.32 ± 0.39             | 9.36 ± 0.37             | 9.49 ± 0.36             | 8.94 ± 0.49             |
| 22:0            | 0.04 ± 0.03    | 0.00 ± 0.00             | 0.01 ± 0.01             | 0.06 ± 0.04             | 0.03 ± 0.03             | 0.04 ± 0.03             |
| 22:1n-9         | 0.11 ± 0.06    | 0.08 ± 0.02             | 0.06 ± 0.01             | 0.05 ± 0.01             | 0.09 ± 0.04             | 0.35 ± 0.32             |
| EPA (20:5n-3)†  | 1.27 ± 0.15    | 1.22 ± 0.09             | 1.10 ± 0.10             | 1.31 ± 0.14             | 1.40 ± 0.17             | 0.96* ± 0.11             |
| 22:4n-6         | 0.23 ± 0.02    | 0.23 ± 0.02             | 0.22 ± 0.02             | 0.19* ± 0.01            | 0.35 ± 0.14             | 0.21 ± 0.03             |
| 22:5n-6         | 0.76 ± 0.10    | 0.86 ± 0.11             | 0.83 ± 0.11             | 0.89 ± 0.12             | 0.82 ± 0.10             | 0.86 ± 0.11             |
| 22:5n-3†        | 0.80 ± 0.03    | 0.80 ± 0.03             | 0.75 ± 0.04             | 0.70 ± 0.03             | 0.80 ± 0.03             | 0.68* ± 0.03             |
| DHA (22:6n-3)†  | 3.32 ± 0.19    | 3.20 ± 0.17             | 3.07 ± 0.17             | 3.20 ± 0.20             | 3.35 ± 0.19             | 3.02* ± 0.18             |
| SFA total       | 51.00 ± 0.39   | 50.68 ± 0.47            | 51.22 ± 0.43            | 49.97* ± 0.55           | 50.59 ± 0.32            | 49.71 ± 0.38             |
| MUFA total      | 13.23 ± 0.27   | 13.47 ± 0.24            | 13.39 ± 0.44            | 13.53 ± 0.25            | 12.86 ± 0.31            | 14.32 ± 0.75             |
| PUFA total      | 35.76 ± 0.40   | 35.85 ± 0.47            | 35.39 ± 0.43            | 36.50 ± 0.51            | 36.55 ± 0.43            | 36.27 ± 0.62             |
| Total (nmol/ml) | 3762.27 ± 179.20 | 3710.31 ± 172.05 | 3843.05 ± 162.56 | 3606.76* ± 177.58 | 3675.00 ± 207.72 | 3647.24 ± 166.68 |

AA, arachidonic acid.
*Mean value was significantly different from that at week 0 (P<0.05).
†Mean values were significantly different between the dietary treatment groups (P<0.05).
Serum lipid data have been published previously. In brief, serum total cholesterol concentrations were significantly lower in both the half-dose and full-dose almond groups than in the control group. Serum LDL-C concentrations were significantly lower and serum HDL-C concentrations were significantly higher in the full-dose almond group than in the control group. Serum TAG concentrations were not affected by the dietary treatments. There were no significant differences in body weight among the dietary treatment groups.

**Fatty acid profile of dietary supplements and serum fractions**

**Fatty acid composition of the dietary supplements.** FA analysis of the almond and muffin supplements revealed that almonds had significantly higher amounts of OA (18 : 1n-9) and total FA than muffins when compared on an equal amount by weight basis (Table 1). Similar findings were recorded when the supplements were analysed on a per energy basis. As has been mentioned previously, the MUFA in almonds balanced the carbohydrate in muffins.

Changes in plasma FA concentrations after the dietary treatments reflected the FA profile of the supplements, confirming subjects’ compliance with the dietary protocol. These changes were in agreement with subjects’ reported compliance with the dietary protocol, which was good, ranging from 97·8 (SEM 0·7) to 99·5 (SEM 0·6)% for all the three dietary treatments.

**Phospholipid fraction.** There was a significant increase in the concentrations of serum OA in the full-dose and half-dose almond groups (weeks 0–4) (P<0·001 and P<0·024, respectively), while there was no significant difference in the concentrations from baseline in the control group (Table 2).

The proportions of SFA in the phospholipid fraction were positively associated with the 10-year CHD risk (R = 0·70, n 80, P=0·015), while those of PUFA in the fraction were inversely associated with it (R = −0·294, n 80, P=0·008).

**TAG fraction.** There was a significant increase in the proportions of OA and MUFA in the serum TAG fraction in the half-dose (P=0·003 and P=0·004, respectively) and full-dose (P<0·001 and P<0·001, respectively) almond groups, while there was no change in the control group (Table 3; Fig. 1). The proportions of OA and MUFA in the almond groups increased significantly in a dose-dependent manner when compared with those in the control group (P<0·05).

There was a significant decrease in the proportion of palmitic acid (16 : 0) in the half-dose and full-dose almond groups when compared with that in the control group (P<0·05). The proportion of palmitic acid was positively associated with serum TAG concentrations (R = 0·47, n 80, P<0·001) and the total cholesterol:HDL-C ratio (R = 0·325, n 80, P=0·05).

| Table 3. Effect of dietary treatments on the proportions of fatty acids in the serum TAG fraction |
|---------------------------------------------------------------------------------------------|
| (Mean values with their standard errors)                                                    |

| Fatty acids (%) | Control (n 27) | Half-dose almonds (n 27) | Full-dose almonds (n 27) |
|-----------------|----------------|-------------------------|-------------------------|
|                 | Week 0         | Week 4                   | Week 0                   | Week 4                   |
|                 | Mean SEM       | Mean SEM                 | Mean SEM                 | Mean SEM                 |
| 14: 0           | 2·01 0·14      | 2·01 0·18                | 2·12 0·21                | 1·74* 0·12               |
| 14: 1n-5        | 0·23 0·03      | 0·22 0·04                | 0·29 0·05                | 0·18* 0·03               |
| 16: 0           | 27·12 0·70     | 26·41 0·71               | 27·95 0·70               | 24·39* 0·66              |
| 16: 1n-7        | 4·01 0·23      | 3·94 0·25                | 3·93 0·25                | 3·41* 0·18               |
| 18: 0           | 3·73 0·20      | 3·60 0·17                | 4·02 0·60                | 3·46 0·14                |
| 18: 1n-9        | 38·00 0·76     | 38·32 0·61               | 37·04 0·89               | 39·94* 0·75              |
| 18: 1n-7        | 3·17 0·82      | 3·38 0·97                | 3·26 0·96                | 3·94 1·11                |
| 18: 2n-6        | 16·57 0·76     | 16·74 0·81               | 16·12 0·81               | 17·50* 0·70              |
| 18: 3n-6        | 0·47 0·06      | 0·50 0·09                | 0·46 0·05                | 0·40 0·02                |
| 20: 0:18: 3n-3† | 1·28 0·08      | 1·53* 0·11               | 1·40 0·12                | 1·42 0·08                |
| 20: 1n-9        | 0·21 0·02      | 0·23 0·02                | 0·23 0·01                | 0·23 0·02                |
| 20: 3n-3        | 0·09 0·01      | 0·11 0·02                | 0·09 0·01                | 0·16 0·05                |
| AA (20: 4n-6)   | 1·26 0·06      | 1·17* 0·07               | 1·26 0·08                | 1·17 0·05                |
| 22: 0           | 0·00 0·00      | 0·00 0·00                | 0·00 0·01                | 0·01 0·01                |
| 22: 1n-9        | 0·01 0·01      | 0·01 0·01                | 0·01 0·00                | 0·03 0·02                |
| EPA (20: 5n-3)  | 0·38 0·07      | 0·50 0·21                | 0·32 0·05                | 0·40 0·08                |
| 22: 4n-6        | 0·10 0·04      | 0·09 0·03                | 0·09 0·04                | 0·10 0·04                |
| 24: 1n-9        | 0·25 0·14      | 0·26 0·13                | 0·25 0·14                | 0·36 0·18                |
| 22: 5n-6        | 0·02 0·02      | 0·00 0·00                | 0·00 0·00                | 0·00 0·00                |
| 22: 5n-3        | 0·31 0·04      | 0·35 0·03                | 0·34 0·03                | 0·36 0·03                |
| DHA (22: 6n-3)† | 0·78 0·13      | 0·63 0·08                | 0·83 0·15                | 0·82 0·09                |
| SFA total†      | 33·51 0·67     | 32·79 0·89               | 34·79 1·15               | 30·31* 0·78              |
| MUFA total†     | 45·87 0·67     | 46·36 0·74               | 45·01 0·44               | 48·09* 0·81              |
| PUFA total‡     | 20·62 0·87     | 20·85 0·90               | 20·20 0·92               | 21·60* 0·82              |
| Total (nmol/ml) | 1529·20        | 147·17                   | 1576·81                  | 121·93                   |

AA, arachidonic acid.
† Mean value was significantly different from that at week 0 (P<0·05).
‡ Mean values were significantly different between the dietary treatment groups (P<0·05).
The proportions of MUFA in the NEFA fraction were positively associated with HDL-C concentrations (R 0.329, n 80, P=0.005) and inversely associated with the 10-year CHD risk (R −0.290, n 80, P=0.009); however, this association was significant when all the MUFA, except OA, were assessed (data not shown). In contrast, the proportions of SFA and palmitic acid (16:0) were inversely associated with HDL-C concentrations (R −0.319, n 80, P=0.004; R −0.297, n 80, P=0.007, respectively), possibly related to the reduction in the proportions of palmitic acid and total SFA and increase in those of MUFA after almond supplementation. The proportions of SFA and palmitic acid were also positively associated with the 10-year CHD risk (R 0.273, n 80, P=0.014; R 0.331, n 80, P=0.003, respectively).

No significant associations were observed between the proportions of OA or PUFA in the NEFA fraction and the risk of CHD.

Overall almond intake and CHD risk

Almond intake was found to be associated with a decrease in the percentage change of the estimated Framingham 10-year CHD risk score during the study (R −0.247, n 81, P=0.026), where for every 30 g increase in almond intake, a 3.5% decrease in the 10-year CHD risk was observed (Fig. 2).

Discussion

The results of the present study indicate that an isoenergetic incorporation of almonds (approximately 10 and 20% of energy from half-dose and full-dose almonds, respectively) into a National Cholesterol Education Program Step 2 diet improves the FA profile of hyperlipidaemic adults by increasing the OA and MUFA content of serum TAG and NEFA fractions, which are inversely associated with CHD lipid risk factors and overall 10-year CHD risk.

Much of the interest in the role of MUFA in the prevention of CHD arises from the observed beneficial effects of a Mediterranean dietary pattern, which is high in olive oil, providing 14–40% of energy from MUFA. In contrast, MUFA from olive oil, almonds contain significantly high amounts of MUFA in the form of OA. The beneficial effects of almonds, particularly the MUFA component, on CHD risk observed in the present study are consistent with previous findings, including those of the Prevención con Dieta Mediterránea (PREDIMED) study. The results of a PREDIMED-based cross-over trial

Fig. 1. Effect of almond dose on the change in oleic acid (18:1n-9) (A) and MUFA (B) proportions in the serum NEFA (□) and TAG (■) fractions of hyperlipidaemic subjects. Values are means, with standard errors represented by vertical bars. * Mean value was significantly different from that in the NEFA fraction (P<0.05). a,b Mean values with unlike letters were significantly different (P<0.05).

P=0.003) and inversely associated with HDL-C concentrations (R 0.444, n 80, P<0.001).

The 10-year CHD risk was inversely associated with the proportions of OA (R −0.283, n 80, P=0.011) and MUFA (R −0.269, n 80, P=0.016) in the TAG fraction; this association was no longer significant when all the MUFA, except OA, were assessed (data not shown). The proportions of SFA were negatively associated with HDL-C concentrations (R −0.428, n 80, P=0.001) and positively associated with the 10-year CHD risk (R 0.300, n 80, P=0.007).

No significant associations were observed between MUFA proportions and 10-year CHD risk.

Cholesterol ester fraction. There were no significant differences in the proportions of serum OA or MUFA from baseline within or between the dietary treatment groups or any significant associations between FA concentrations and blood lipid profiles or CHD risk (data not shown).

NEFA fraction. The proportions of individual FA in the serum NEFA fraction are given in Table 4. There was no change in the proportion of OA (18:1n-9) in the control group, but an increase was observed in the almond groups (Fig. 1). However, this increase was only significant in the half-dose almond group (P=0.01), and the lack of a significant increase in the full-dose almond group is possibly due to high variability. Similarly, there was an increase in the proportions of serum MUFA in the half-dose and full-dose almond groups, but it was significant only in the half-dose almond group (P=0.04 and P=0.06, respectively; Fig. 2).

The proportions of MUFA in the NEFA fraction were positively associated with HDL-C concentrations (R 0.329, n 80, P=0.005) and inversely associated with the 10-year CHD risk (R −0.290, n 80, P=0.009); however, this association was significant when all the MUFA, except OA, were assessed (data not shown). In contrast, the proportions of SFA and palmitic acid (16:0) were inversely associated with HDL-C concentrations (R −0.319, n 80, P=0.004; R −0.297, n 80, P=0.007, respectively), possibly related to the reduction in the proportions of palmitic acid and total SFA and increase in those of MUFA after almond supplementation. The proportions of SFA and palmitic acid were also positively associated with the 10-year CHD risk (R 0.273, n 80, P=0.014; R 0.331, n 80, P=0.003, respectively).

No significant associations were observed between the proportions of OA or PUFA in the NEFA fraction and the risk of CHD.

Overall almond intake and CHD risk

Almond intake was found to be associated with a decrease in the percentage change of the estimated Framingham 10-year CHD risk score during the study (R −0.247, n 81, P=0.026), where for every 30 g increase in almond intake, a 3.5% decrease in the 10-year CHD risk was observed (Fig. 2).

Discussion

The results of the present study indicate that an isoenergetic incorporation of almonds (approximately 10 and 20% of energy from half-dose and full-dose almonds, respectively) into a National Cholesterol Education Program Step 2 diet improves the FA profile of hyperlipidaemic adults by increasing the OA and MUFA content of serum TAG and NEFA fractions, which are inversely associated with CHD lipid risk factors and overall 10-year CHD risk.

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Additional trials within PREDIMED have supported that dietary incorporation of almonds are associated with a reduced risk of CHD. However, this may not apply to the present study as we focused on a MUFA–saturated fats with monounsaturated fats in the diet. In a randomised, controlled, parallel study, the SFA concentrations of thirty-eight hypercholesterolaemic adults who consumed 100 g of raw almonds daily over a 4-week period were found to be lower \(^{34}\). Furthermore, increasing MUFA intake has been reported to be associated with an increase in HDL-C concentrations, as demonstrated by a randomised, controlled trial of twenty-four hypercholesterolaemic adults \(^{35}\). The improvement in blood lipid profiles has generally been associated with a reduced risk of CHD. However, this may not apply to the present study as we focused on a MUFA–carbohydrate substitution. It has also been proposed that

Table 4. Effect of dietary treatments on the proportions of fatty acids in the serum NEFA fraction

| Fatty acids (%) | Control (n 27) | Control (n 27) | Half-dose almonds (n 27) | Half-dose almonds (n 27) | Full-dose almonds (n 27) | Full-dose almonds (n 27) |
|----------------|---------------|---------------|-------------------------|-------------------------|-------------------------|-------------------------|
|                | Week 0        | Week 4        | Week 0                  | Week 4                  | Week 0                  | Week 4                  |
| 14:0           | Mean SEM      | Mean SEM      | Mean SEM                | Mean SEM                | Mean SEM                | Mean SEM                |
| 14:1n-5        | 1.70 ± 0.10   | 1.58 ± 0.13   | 1.73 ± 0.11             | 1.52 ± 0.12             | 1.64 ± 0.14             | 1.71 ± 0.16             |
| 16:0           | 0.42 ± 0.09   | 0.43 ± 0.09   | 0.39 ± 0.09             | 0.39 ± 0.09             | 0.54 ± 0.13             | 0.41 ± 0.14             |
| 16:1n-7        | 27.00 ± 0.66  | 26.58 ± 0.88  | 27.58 ± 0.53            | 25.73* ± 0.54           | 27.12 ± 0.57            | 25.90 ± 0.61            |
| 18:0           | 3.06 ± 0.21   | 2.90 ± 0.21   | 3.40 ± 0.34             | 3.97 ± 0.88             | 3.12 ± 0.26             | 3.05* ± 0.22            |
| 18:1n-9        | 33.61 ± 1.10  | 34.14 ± 0.78  | 33.36 ± 0.63            | 34.98* ± 0.74           | 33.12 ± 1.19            | 34.78 ± 0.85            |
| 18:1n-7        | 12.18 ± 0.82  | 12.11 ± 0.45  | 11.74 ± 0.45            | 11.84 ± 0.66            | 13.04 ± 0.72            | 11.55 ± 0.51            |
| 18:2n-6        | 13.08 ± 0.49  | 13.51 ± 0.49  | 13.30 ± 0.43            | 13.46 ± 0.42            | 13.38 ± 0.58            | 13.40 ± 0.42            |
| 18:3n-6        | 0.67 ± 0.48   | 0.36 ± 0.25   | 0.68 ± 0.49             | 0.56 ± 0.48             | 0.19 ± 0.16             | 1.01 ± 0.61             |
| 20:1/20:0      | 1.16 ± 0.07   | 1.27 ± 0.07   | 1.19 ± 0.07             | 1.22 ± 0.07             | 1.21 ± 0.08             | 1.11 ± 0.06             |
| 20:1n-9        | 0.16 ± 0.03   | 0.15 ± 0.03   | 0.15 ± 0.03             | 0.16 ± 0.03             | 0.14 ± 0.05             | 0.14 ± 0.03             |
| 20:3n-6        | 0.04 ± 0.02   | 0.22 ± 0.22   | 0.02 ± 0.01             | 0.21 ± 0.18             | 0.01 ± 0.02             | 0.02 ± 0.01             |
| AA (20:4n-6)   | 1.22 ± 0.17   | 1.20 ± 0.22   | 1.05 ± 0.06             | 1.35 ± 0.43             | 1.43 ± 0.38             | 1.26* ± 0.38            |
| 22:0           | 0.00 ± 0.00   | 0.00 ± 0.00   | 0.00 ± 0.00             | 0.00 ± 0.00             | 0.00 ± 0.00             | 0.00 ± 0.00             |
| 22:1n-9        | 0.26 ± 0.20   | 0.04 ± 0.02   | 0.06 ± 0.03             | 0.08 ± 0.04             | 0.07 ± 0.05             | 0.12 ± 0.07             |
| EPA (20:5n-3)  | 0.49 ± 0.29   | 0.54 ± 0.34   | 0.88 ± 0.47             | 0.37 ± 0.29             | 1.09 ± 0.10             | 0.79 ± 0.43             |
| 22:4n-6        | 0.18 ± 0.09   | 0.42 ± 0.16   | 0.36 ± 0.13             | 0.93 ± 0.14             | 0.40 ± 0.21             | 0.19 ± 0.09             |
| 24:1n-9        | 2.14 ± 0.95   | 1.90 ± 0.68   | 1.45 ± 0.64             | 0.95 ± 0.54             | 0.87 ± 0.50             | 1.67 ± 0.64             |
| 22:5n-6        | 0.00 ± 0.00   | 0.00 ± 0.00   | 0.00 ± 0.00             | 0.00 ± 0.00             | 0.00 ± 0.00             | 0.00 ± 0.00             |
| 22:5n-3        | 0.10 ± 0.07   | 0.11 ± 0.08   | 0.11 ± 0.08             | 0.04 ± 0.01             | 0.03 ± 0.01             | 0.10 ± 0.09             |
| DHA (22:6n-3)  | 0.52 ± 0.06   | 0.51 ± 0.05   | 0.51 ± 0.06             | 0.71 ± 0.26             | 0.55 ± 0.07             | 0.55 ± 0.11             |
| SFA            | 40.46 ± 1.20  | 40.89 ± 0.97  | 41.66 ± 0.83            | 39.70* ± 0.95           | 42.40 ± 1.14            | 39.72* ± 1.00           |
| MUFA           | 41.66 ± 1.11  | 41.55 ± 0.88  | 40.85 ± 0.94            | 42.60* ± 1.14           | 39.92 ± 1.31            | 42.42 ± 0.95            |
| PUFAs          | 16.88 ± 0.93  | 17.55 ± 0.76  | 17.49 ± 0.90            | 17.70 ± 0.97            | 17.68 ± 0.86            | 17.86 ± 0.98            |
| Total (nmol/ml)| 425.30 ± 35.78| 389.08 ± 21.08| 403.96 ± 25.25         | 432.03 ± 28.00         | 374.47 ± 33.86          | 417.01 ± 39.13          |

AA, arachidonic acid.

\(^{*}\) Mean value was significantly different from that at week 0 (\(P<0.05\)).

Fig 2. Regression analysis on the association between almond consumption and percentage change of the estimated 10-year CHD risk score using the Framingham equation (\(R = 0.247, n = 81, P = 0.026\)).
almonds may act endogenously to exert a cardioprotective effect. Dietary FA are rapidly incorporated into lipoprotein lipids. Enrichment of lipoprotein particles with MUFA at the expense of PUFA may enhance their resistance to oxidative stress as there are few double bonds to react with, thus potentiating the anti-atherogenic effect\textsuperscript{36–38}. Plasma phospholipid, cholesteryl ester, TAG and NEFA compositions are responsive to total dietary fat content, though NEFA and TAG are the major forms of lipids found in circulation\textsuperscript{39}.

There are several limitations to the present study. α-Linolenic acid (18 : 3\textsuperscript{ω-3}) did not get separated from arachidic acid (20 : 0) during chromatographic analysis. Almonds are low in α-linolenic acid\textsuperscript{40} and would not be expected to significantly alter serum concentrations; however, neither this nor any associations between changes in serum α-linolenic acid concentrations and CHD risk could be verified from the findings of the present study. Moreover, in the present study, almond consumption was found to increase faecal MUFA excretion (RP Bazinet and CWC Kendal, unpublished results), indicating an attenuated effect due to the inaccessibility of some of the nutrients and energy from almonds for absorption\textsuperscript{40}. This is further supported by findings from previous studies, which suggest that nuts contain less metabolisable energy than expected based on Atwater general factors and that the lipid from nuts is more poorly absorbed than that from other food sources\textsuperscript{40}.

Despite these limitations, the findings of the present study suggest that the FA profile of almonds has the potential to contribute to the total reduction of CHD risk. The cholesterol-lowering effect of almond consumption is explained, in part, by the FA profile of almonds, especially MUFA; however, there may be other bioactive components in almonds that further reduce CHD risk. In addition to unsaturated FA, almonds are a good source of vitamin E, fibre and phytochemicals (phenols, flavonoids, proanthocyanidins and phytosterols), arginine, Cu and Mg, which may beneficially influence CHD risk\textsuperscript{42–44}. Many of these nutrients may act synergistically to produce the observed favourable outcomes, although further studies are required to prove this postulation\textsuperscript{42}. Further analysis of LDL particle size may provide a better understanding of potential mechanisms by which almonds and other foods rich in MUFA influence TAG concentrations.

In conclusion, the results of the present study indicate that almond consumption favourably alters the serum FA profile by increasing the proportions of OA and total MUFA and decreasing those of SFA. These changes in the FA profile are correlated with improvements in blood lipoproteins and with a decreased 10-year CHD risk.

**Supplementary material**

To view supplementary material for this article, please visit http://dx.doi.org/10.1017/S0007114514001640

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C. W. C. K., R. P. B. and D. J. A. J. had full access to all the data in the study and were responsible for the integrity of the data and the accuracy of the data analysis. C. W. C. K. and D. J. A. J. obtained funding. C. W. C. K., R. P. B., K. G. L. and D. J. A. J. were responsible for study concept and design. S. N., C. W. C. K., R. P. B., B. B., A.-M. G. and D. J. A. J. were responsible for the acquisition and analysis of data. S. N., C. W. C. K., A.-M. G., R. P. B., K. G. L., L. S. A. A., J. L. S. and D. J. A. J. were responsible for the interpretation of data. S. N., C. W. C. K. and R. P. B. drafted the manuscript. S. N., C. W. C. K., A.-M. G., R. P. B., B. B., K. G. L., L. S. A. A. and J. L. S. provided administrative, technical and material support. C. W. C. K., R. P. B., B. B. and D. J. A. J. supervised the study. All authors critically revised the manuscript for important intellectual content and approved the manuscript submitted for publication.

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