Physiologically Based Pharmacokinetic Modeling to Assess the Impact of CYP2D6-Mediated Drug-Drug Interactions on Tramadol and O-Desmethyltramadol Exposures via Allosteric and Competitive Inhibition

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Abstract
Tramadol is an opioid medication used to treat moderately severe pain. Cytochrome P450 (CYP) 2D6 inhibition could be important for tramadol, as it decreases the formation of its pharmacologically active metabolite, O-desmethyltramadol, potentially resulting in increased opioid use and misuse. The objective of this study was to evaluate the impact of allosteric and competitive CYP2D6 inhibition on tramadol and O-desmethyltramadol pharmacokinetics using quinidine and metoprolol as prototypical perpetrator drugs. A physiologically based pharmacokinetic model for tramadol and O-desmethyltramadol was developed and verified in PK-Sim version 8 and linked to respective models of quinidine and metoprolol to evaluate the impact of allosteric and competitive CYP2D6 inhibition on tramadol and O-desmethyltramadol exposure. Our results show that there is a differentiated impact of CYP2D6 inhibitors on tramadol and O-desmethyltramadol based on their mechanisms of inhibition. Following allosteric inhibition by a single dose of quinidine, the exposure of both tramadol (51% increase) and O-desmethyltramadol (52% decrease) was predicted to be significantly altered after concomitant administration of a single dose of tramadol. Following multiple-dose administration of tramadol and a single-dose or multiple-dose administration of quinidine, the inhibitory effect of quinidine was predicted to be long (∼42 hours) and to alter exposure of tramadol and O-desmethyltramadol by up to 60%, suggesting that coadministration of quinidine and tramadol should be avoided clinically. In comparison, there is no predicted significant impact of metoprolol on tramadol and O-desmethyltramadol exposure. In fact, tramadol is predicted to act as a CYP2D6 perpetrator and increase metoprolol exposure, which may necessitate the need for dose separation.

Keywords
CYP2D6-mediated drug-drug interactions, metoprolol, O-desmethyltramadol, PBPK, quinidine, tramadol

Introduction
Chronic pain is defined as pain without apparent biological value that has persisted beyond the normal healing time of 3 to 6 months.1 It is a major public health concern, affecting 1 in 5 adults in the United States.2 Among primary care appointments, 22% focus on chronic pain management.3 The American College of Occupational Environmental Medicine guidelines for the chronic use of opioids4 and the American Society of Interventional Pain Physicians5 recommend combination medication therapy including opioids, antidepressants, nonsteroidal anti-inflammatory drugs, and anticonvulsants. The volume of opioid usage in the United States increased by up to 1177% between 1997 and 2006.6 The most commonly used opioids in chronic pain management are oxycodone, hydrocodone, codeine, tramadol, morphine, hydromorphone, methadone, and fentanyl.

Tramadol accounts for approximately 33% of opioid use in chronic pain management.7 It is a centrally
acting μ-opioid receptor agonist used to treat moderately severe pain. Tramadol also acts as a serotonin-norepinephrine reuptake inhibitor. It is predominantly metabolized by different cytochrome P450 (CYP) enzymes to an active O-desmethyltramadol and an inactive N-desmethyltramadol metabolite. The remainder is excreted unchanged in urine. The biotransformation to O-desmethyltramadol is primarily mediated by CYP2D6; biotransformation to N-desmethyltramadol is mediated by CYP3A4 and CYP2B6. Both O-desmethyltramadol and N-desmethyltramadol are further converted to either N,O-didesmethyltramadol or other inactive metabolites via either the same or other CYPs or by UDP-glucuronosyltransferases 1A8 and 2B7. The O-desmethyltramadol metabolite is much more potent than its parent compound (≈200-300 times greater μ-opioid receptor–binding affinity). Although tramadol per se is responsible for inhibition of the serotonin-norepinephrine reuptake that could play a role in pain perception, it is generally considered a prodrug because O-desmethyltramadol has much greater μ-opioid receptor binding affinity and has been shown to be 6 times more effective clinically.

There is wide interindividual variability in CYP2D6 activity, due to genetic polymorphisms or phenocconversion caused by drug-drug interactions (DDIs). This is expected to affect the pharmacokinetics, efficacy, and, potentially, safety of tramadol and O-desmethyltramadol. However, recent clinical studies have reported considerable amounts of O-desmethyltramadol in the plasma of CYP2D6 poor metabolizers (PMs; with no functional CYP2D6 activity) following administration of tramadol, suggesting the involvement of other CYP enzymes in the formation of O-desmethyltramadol. Decreased CYP2D6 activity due to either allosteric inhibition or competitive inhibition could decrease formation of O-desmethyltramadol, thus compromising the overall analgesic effects and potentially increasing dosing frequency and inciting prescribing cascades in an attempt to overcome this interaction. Inhibition of CYP2D6 also increases tramadol plasma levels, which could further suppress central nervous system activity, increasing the risk for serotoninergic toxicity and reducing seizure threshold.

Allosteric inhibition and competitive inhibition are 2 very different types of enzyme-mediated DDIs. Allosteric inhibition is a noncompetitive inhibition that involves binding of the perpetrator drug to a region of the enzyme protein structure that is spatially different from the active site where the substrate binds and is transformed. Drug binding to the allosteric site can cause conformational changes of the enzyme that render the active site no longer accessible for substrate binding or make the site unable to catalyze reactions. Almost all cases of noncompetitive inhibition are considered to be caused by allosteric regulation. For this type of inhibition, the apparent Michaelis-Menten constant ($K_m$) remains unchanged, and the apparent maximum reaction velocity ($V_{max}$) decreases. Quinidine is a known allosteric inhibitor of CYP2D6. In contrast, competitive inhibition involves 2 substrates competing for the same active site. Whether CYP2D6 substrates act as perpetrator or victim drugs is a function of their relative affinity for the active site as well as their concentration in the vicinity of the enzyme. For this type of inhibition, the apparent $K_m$ increases, while the apparent $V_{max}$ remains unchanged.

Tramadol affinity for CYP2D6 is generally considered to be relatively low, while some data suggest that metoprolol would have a moderate CYP2D6 affinity. The pattern and magnitude of interaction between 2 drugs, such as tramadol and metoprolol, depends on several factors, including the baseline bioavailability, the partial metabolic clearance through the inhibited enzymatic pathway (overall contribution of a metabolic pathway to the total clearance of the victim drug), the dose of both substrates, the sequence of administration, and the time delay between the perpetrator and victim drugs (synchronization). Prevention of DDIs is an essential component of appropriate prescribing, as DDIs represent a major risk factor for adverse drug events in patients with several chronic diseases and polypharmacy. As a patient’s pharmaceutical regimen becomes more complex, the added influence of multiple substrates competing for the same CYP450 enzyme becomes more difficult to manage.

Quantitative clinical pharmacology applications, including physiologically based pharmacokinetic (PBPK) modeling and simulation, have gained popularity for DDI assessment and are now routinely employed in drug development and regulatory evaluation. In the absence of a dedicated clinical DDI studies, PBPK models have been used to predict changes in the concentrations of victim drugs, derive dosing recommendations, and inform drug labeling. PBPK models can also help rule out the potential for a significant DDI or eliminate the need to conduct a clinical pharmacology study even when the drug candidate itself is a perpetrator. This research has 2 objectives: (1) to evaluate the impact of CYP2D6-mediated DDIs via allosteric and competitive inhibitions on tramadol and O-desmethyltramadol exposure, and (2) to optimize administration times to minimize DDI risk using PBPK modeling and simulation.

**Methods**

**Software**

PBPK modeling and simulation were performed in PK-Sim version 8.0, which is part of the Open
Systems Pharmacology software package (https://www.open-systems-pharmacology.org/). WebPlotDigitizer (https://automeris.io/WebPlotDigitizer/) was used to digitize average PK profiles from the literature.

Tramadol and O-desmethyltramadol PBPK Model Development and Verification

A summary of our PBPK modeling approach is summarized in Figure 1. Tramadol and O-desmethyltramadol disposition models were developed by integrating physicochemical drug properties (partition coefficient, ionization constant, etc) with clinical PK parameters collected from published clinical studies.\textsuperscript{15–18,28–32} For tramadol, specific tissue-to-plasma partition coefficients were estimated using the Rodgers and Rowland method;\textsuperscript{33} cellular permeabilities were estimated using the PK-Sim standard algorithm. Renal clearance was set to 0.13 L/h/kg, which represents $30\%$ of the reported total tramadol clearance.\textsuperscript{34,35} The remaining nonrenal clearance was considered as hepatic elimination. The unbound intrinsic hepatic clearance ($\text{CL}_{\text{int,u}}$) was derived retrogradely by integrating reported tramadol fraction unbound in plasma and the PK-Sim inputs for liver blood flow using the well-stirred liver model. The resulting $\text{CL}_{\text{int,u}}$ was further stratified to represent the relative contribution of CYP2D6 to the biotransformation of tramadol to O-desmethyltramadol.

Briefly, we derived a human mass balance diagram for tramadol and O-desmethyltramadol using urinary excretion and systemic exposure data collected from the literature.\textsuperscript{28–32} The relative contribution of CYP2D6-mediated metabolic pathway was then translated into specific $\text{CL}_{\text{int,u}}$/CYP2D6 using intersystem extrapolation factors related to enzyme abundance and variability. The resulting tramadol fraction metabolized by CYP2D6 was further confirmed by capturing the remaining CYP2D6 activity and tramadol’s pharmacokinetic (PK) profile in PMs and NMs.\textsuperscript{15–18,36}

Conceptually, to simulate the PK of tramadol and O-desmethyltramadol in PMs, the CYP2D6 metabolic pathway was completely knocked out by setting the respective $\text{CL}_{\text{int,u}} = 0$. For O-desmethyltramadol, specific tissue-to-plasma partition coefficients were estimated using the Schmitt method,\textsuperscript{37} while cellular permeabilities were estimated using the PK-Sim standard algorithm. The renal plasma clearance of O-desmethyltramadol was set to 0.16 L/h/kg.\textsuperscript{38}

Once developed and verified, the disposition model was expanded to account for factors impacting tramadol’s oral absorption. The dissolution of tramadol immediate release formulation was modeled by a Weibull function. The dissolution time (50\% dissolved) and shape was fitted to oral PK data\textsuperscript{15} and set as 7 minutes and 0.64, which were in line with the reported dissolution profile.\textsuperscript{39} The specific intestinal permeability was optimized based on available clinical data.\textsuperscript{15} The overarching combined PBPK model for tramadol and O-desmethyltramadol was further verified by comparing their predicted area under the plasma concentration–time curve (AUC) ratios in CYP2D6 NMs (ie, individuals with functional CYP2D6) to
Table 1. Summary of Physicochemical and Pharmacokinetic Parameters for Tramadol and O-desmethyltramadol in the Development of the PBPK Model

| Parameter                           | Value          | Source |
|-------------------------------------|----------------|--------|
| Molecular weight, g/mol             | 263.4          | 35     |
| logP                                | 1.93           | Optimized \(^a\) |
| Solubility, mg/L                    | 0.75 (pH = 7)  |         |
| pKa                                 | 9.41 (base)    | 34,35  |
| Fu, %                               | 80             | 35     |
| Total CL, L/h/kg                    | 0.51 (31) \(^d\) | 35     |
| In vitro intrinsic CL-CYP2D6, μL/min/pmol recombinant enzyme | 0.35 | Estimated \(^d\) |
| In vitro intrinsic CL-CYPX, μL/min/pmol recombinant enzyme | 0.03 | Estimated \(^d\) |
| Total hepatic CL-CYP3A4 and CYP2B6, L/h/kg | 0.1 | Estimated \(^d\) |
| Renal CL, L/h/kg                    | 0.13           | Optimized \(^b\) |
| V/L kg                              | 2.6-2.9        | 9,35   |
| Elimination half-life, h            | 6.3 (1.4) \(^a\) | 9,35   |
| t\(_{max}\), h                      | 1.6 (63) \(^d\) | 35     |
| B:P                                 | 1.07           | 34     |
| F                                   | 0.75           | 9,35   |
| O-desmethyltramadol parameters      | Value          | Source |
| Molecular weight                    | 249.3          | 35     |
| logP                                | 1.15           | Optimized \(^a\) |
| Solubility, mg/mL                   | 3.53 (pH = 7)  |         |
| pKa                                 | 9.62 (base)    | 35     |
| f\(_u\), %                          | 40             | Estimated |
| Total CL, L/h/kg                    | 0.3            | Estimated |
| Total hepatic CL-CYP and UGT, L/h/kg| 0.27          | Estimated \(^d\) |
| Renal CL, L/h/kg                    | 0.16           | 38     |
| V/L kg                              | 3.2            | 65     |
| Elimination half-life, h            | 7.4 (1.4) \(^a\) | 9,35   |
| t\(_{max}\), h                      | 3.0 (51) \(^d\) | 35     |

B:P, blood to plasma concentration ratio; CL, clearance; CYP, cytochrome P450; F, bioavailability; f\(_u\), fraction unbound in plasma; logP, partition coefficient; PBPK, physiologically based pharmacokinetic; pKa, ionization constant; t\(_{max}\), time to maximum concentration; UGT, UDP-glucuronosyltransferase; V, volume of distribution.

\(^a\)Optimization: In PK-Sim, it is recommended to use logMA (membrane affinity) as the input parameter. If the membrane affinity value is not available, logP can be used instead. A reasonable variation around the logP value should be allowed since this parameter is not directly related to membrane affinity.

\(^b\)Optimization, based on reported values from T’Jollyn et al.\(^{14}\) and clinical observed data from Campanero et al.\(^{17}\)

\(^c\)Estimated from retrograded calculation (based on well-stirred liver model and in vitro–in vivo extrapolation).

\(^d\)Mean (% coefficient of variation).

\(^e\)Mean (standard deviation).

Those observed in PMs, following tramadol oral administration as reported from clinical observations.\(^{17}\)

A summary of the final drug-specific parameters is summarized in Table 1.

Quinidine and Metoprolol PBPK Model Development and Verification

The perpetrator models were developed using both in vitro and clinical data. The parameterization input for quinidine and metoprolol PBPK models, including physiochemical (molecular weight, partition coefficient, ionization constant, etc.), pharmacokinetic (volume of distribution and clearance, etc.),\(^{40–43}\) metabolic (V\(_{max}\) and K\(_m\), etc.),\(^{44,45}\) and enzyme inhibition (quinidine-allosteric, metoprolol-competitive inhibitory constant)\(^{46,47}\) parameters, are shown in Table S1-S3. The developed perpetrator models were qualified using observed clinical PK study data following administration of a single or multiple intravenous (IV) and oral dose(s) of the perpetrators (Tables S4 and S5, Figures S1 and S2).\(^{48–50}\)

**DDI Simulations for Allosteric and Competitive CYP2D6 Inhibition**

Once independently developed and verified, PBPK models for tramadol/O-desmethyltramadol and quinidine or metoprolol were combined in simulations to evaluate the impact of allosteric (quinidine) and competitive (metoprolol) inhibition on tramadol and O-desmethyltramadol PK. To this end, we evaluated different scenarios that are relevant for drug development and regulatory evaluation as well as clinical practice.

For allosteric CYP2D6 inhibition via quinidine, we evaluated 3 main scenarios. In scenario 1, a single oral dose of quinidine (400 mg) was given concomitantly with a single oral dose of tramadol (100 mg) in accordance with the current FDA Clinical Drug Interaction Studies Guidance for short half-life drugs (≥6 hours for tramadol) showing time-independent PK.\(^{51}\) In scenario 2, tramadol (100 mg, 4 times daily) was dosed to steady state before concomitant administration of a single oral dose of quinidine (400 mg) at 30 hours (T0). Both of the PK metrics AUC over the dosing interval (eg, AUC\(_{0-6}\), AUC\(_{6-12}\), etc.) were evaluated and their respective AUC ratios (AUCR) were calculated. In scenario 3, tramadol (100 mg 4 times daily) and quinidine (400 mg 4 times daily) were concomitantly administered and both dosed to steady state to order to account for interactions scenarios encountered in the clinic.

For the competitive CYP2D6 inhibition via metoprolol, a single oral dose of metoprolol tartrate (100 mg, immediate-release [IR]) was administered concomitantly with a single 100-mg oral dose of tramadol. We also evaluated delayed metoprolol administration regimens (ie, delayed by 2 and 4 hours) to determine if the interaction between metoprolol and tramadol could be overcome by dose separation. Finally, we evaluated the impact of tramadol on metoprolol pharmacokinetics, as results from a recent in vitro study suggest that tramadol has a higher affinity for CYP2D6 than metoprolol,\(^{45}\) that is, that tramadol may act as the perpetrator rather than the victim drug.
All simulations were carried out in a virtual population consisting of 100 healthy White subjects aged between 25 and 55 years (1:1 ratio of men to women), which were all CYP2D6 NMs. Individual demographics, such as weight, BMI, and organ volumes, among others, were estimated from the built-in expression database available within the PK-Sim software.

Results

PBPK Model Development and Verification

Tramadol and O-desmethyltramadol disposition pathways and their relative contributions are shown in Figure S3. Approximately 26% of the administered dose of tramadol is excreted unchanged in urine. Fifty-three per cent of a tramadol dose is transformed to O-desmethyltramadol (mainly by CYP2D6, 38% of the administered dose), while CYP3A4 and CYP2B6 contribute 21% to its metabolic clearance leading to the formation of N-desmethyltramadol, suggesting that CYP2D6 accounts for ≈51% of tramadol’s hepatic clearance. For O-desmethyltramadol, 38% is excreted unchanged in urine, while 63% undergoes further transformation by phase 1 and 2 metabolism.

Visual inspection of the plasma concentration-time curves overlapping predicted and observed PK profiles for tramadol and O-desmethyltramadol after IV and oral administration of tramadol confirms that model predictions capture the central tendency and variability in tramadol and O-desmethyltramadol systemic exposure in healthy adults (Figure 2). Model adequacy was also confirmed quantitatively as the ratios between observed and predicted PK metrics were within a 1.25-fold range (Table 2). Furthermore, the predictive performance of the model was assessed by an external model verification, where model-based predictions were compared to clinical data in NMs and PMs that were not used during model development (Table 3). The predicted PM/NM exposure ratios were 1.52 and 0.44 for tramadol and O-desmethyltramadol, respectively, which are consistent with observed ratios of 1.44 and 0.37 in clinical studies.

DDI Simulations for Allosteric and Competitive CYP2D6 Inhibition

Concomitant single-dose administration of tramadol and quinidine (scenario 1) increases tramadol AUC<sub>0-∞</sub> by 51% and decreases O-desmethyltramadol AUC<sub>0-∞</sub> by 52%. Under conditions of a single tramadol dose administration, delaying quinidine administration by 4 hours attenuated the magnitude of the predicted DDI, resulting in a 21% rather than a 51% increase in tramadol AUC and a 21% rather than 52% decrease in O-desmethyltramadol AUC. AUCRs are shown in Table 4. In scenario 2, CYP2D6 inhibition associated with a single dose of quinidine leads time-dependent changes in tramadol and O-desmethyltramadol mean plasma concentrations: we observed a 16% to 41% increase in the steady-state tramadol (AUC<sub>τ,ss</sub>) and a 9% to 51% decrease in the O-desmethyltramadol AUC<sub>τ,ss</sub> within 48 hours after quinidine administration. The magnitude of changes in tramadol and O-desmethyltramadol exposure first increases and then gradually decreases, with the maximum exposure changes taking place at approximately 12 to 18 hours after quinidine administration (Table 4). CYP2D6 inhibition following a single dose of quinidine was predicted to last ≈42 hours, at which point both tramadol and O-
Table 2. Observed (OBS) Versus Predicted (PRED) Pharmacokinetic Parameters of Tramadol and O-desmethyltramadol Following Administration of a Single Dose of 100 mg Tramadol Via Intravenous Infusion (10 Minutes) or Oral Administration

| Route of Administration | Cmax,ng/mL—Tramadol | AUC0–∞,(mg·h/L)—Tramadol | Cmax,ng/mL—O-desmethyltramadol | AUC0–∞,(mg·h/L)—O-desmethyltramadol |
|-------------------------|---------------------|--------------------------|-------------------------------|----------------------------------|
| Intravenous infusion    | OBS 1.3 (0.7)       | 126 (36)                 | 142 (49)                      | 0.56 (0.7)                       |
|                         | PRED 3.0 (1.9)      | 210 (52)                 | 230 (74)                      | 1.01 (1.1)                       |
| Oral administration     | OBS 273 (71)        | 300 (71)                 | 300 (71)                      | 0.91 (0.4)                       |
|                         | PRED 1.3 (0.7)      | 126 (36)                 | 126 (36)                      | 0.56 (0.7)                       |

AUC0–∞, area under the plasma concentration–time curve from time 0 to infinity; Cmax, maximum concentration. Mean (SD) for Cmax and AUC0–∞.

Discussion

In this study, we developed and verified a combined PBPK model for tramadol and O-desmethyltramadol in a stepwise fashion. First, we determined the relative contribution of hepatic and renal clearance pathways to the elimination of tramadol and O-desmethyltramadol. Furthermore, we reproduced the individual contribution of different metabolic enzymes to the biotransformation of tramadol into O-desmethyltramadol. The results of this analysis suggest that CYP2D6 accounts for ≈51% of tramadol’s hepatic clearance, which is in line with values reported in literature (43%-48%). The results further suggest that ≈20% of tramadol’s hepatic biotransformation to O-desmethyltramadol is mediated by an unidentified CYP450 isoform, which could explain noticeable amounts of O-desmethyltramadol reported for CYP2D6 PMs.15–18 The combined PBPK model for tramadol and O-desmethyltramadol was developed and externally verified. Overall, our combined PBPK model for tramadol and O-desmethyltramadol was able to reproduce clinical observations well, with the exception of tramadol maximum concentration following a 10-minute IV infusion, which was slightly underpredicted. This underprediction could be explained by the sparse and variable sampling around maximum concentration in the clinical data used for model verification.

In addition to the combined PBPK model for tramadol and O-desmethyltramadol, we developed and verified separate models for quinidine and metoprolol. These models were able to reproduce well clinical observations for both quinidine and quinidine and metoprolol following single- and multiple-dose administration (ratios of predicted and observed PK metrics were within 1.25 range).41,49,50 Once developed and verified, victim and perpetrator models were combined to predict the impact of allosteric (quinidine) and competitive
Table 3. Observed (OBS) Versus Predicted (PRED) $AUC_{0-\infty}$ of Tramadol and O-desmethyltramadol in CYP2D6 Poor Metabolizers (PMs) Versus CYP2D6 Normal Metabolizers (NMs)

| Group | $AUC_{0-\infty}$—Tramadol, mg h/L | $AUC_{0-\infty}$—O-desmethyltramadol, mg h/L | fm, % | $AUC_{0-\infty}$—Tramadol fm a, % | $AUC_{0-\infty}$—O-desmethyltramadol fm a, % |
|-------|----------------------------------|---------------------------------------------|-------|----------------------------------|---------------------------------------------|
| OBS   | 3.14                             | 3.14                                        | 31    | 0.35                             | 0.37                                        |
| NM    | 2.18                             | 2.18                                        | 0.95  | 0.4                              | 0.44                                        |
| PRED  | 3.80                             | 3.80                                        | 34    | 0.4                              | 0.44                                        |
| PM    | 3.80                             | 3.80                                        | 0.9   | 0.4                              | 0.44                                        |

$AUC_{0-\infty}$, area under the plasma concentration–time curve from time 0 to infinity; fm, fraction metabolized. For PMs, their CYP2D6 intrinsic clearance was set to zero in model prediction.

$AUCR = AUC(\text{PM}) / AUC(\text{EM}) = 1 / (1 - fm)$.

Table 4. $AUCR$ for Tramadol and O-desmethyltramadol

| Scenario | $AUC_{0-\infty}$—Tramadol | $AUC_{0-\infty}$—O-desmethyltramadol |
|----------|--------------------------|-------------------------------------|
| 1. w/, concomitant administration | 1.51 | 0.48 |
| 1. w/, delayed administration of quinidine by 4 h | 1.21 | 0.79 |
| 1. w/, delayed administration of quinidine by 8 h | 1.12 | 0.89 |
| 2. $AUC_{0-6h}$ | 1.29 | 0.71 |
| 2. $AUC_{6-12h}$ | 1.35 | 0.64 |
| 2. $AUC_{12-18h}$ | 1.41 | 0.69 |
| 2. $AUC_{18-24h}$ | 1.40 | 0.56 |
| 2. $AUC_{24-36h}$ | 1.36 | 0.65 |
| 2. $AUC_{36-42h}$ | 1.31 | 0.74 |
| 2. $AUC_{42-48h}$ | 1.25 | 0.82 |
| 2. $AUC_{48-54h}$ | 1.16 | 0.91 |

| Scenario | $AUC_{0-\infty}$—Tramadol | $AUC_{0-\infty}$—O-desmethyltramadol |
|----------|--------------------------|-------------------------------------|
| 3. | $AUC_{0-\infty}$—Tramadol-AUCR | $AUC_{0-\infty}$—O-desmethyltramadol-AUCR |
| 3. | 1.57 | 0.40 |

$AUC_{0-\infty}$, area under the plasma concentration–time curve from time 0 to infinity; $AUCR$, area under the plasma concentration–time curve over the dosing interval at steady state; $AUCR$, area under the plasma concentration–time curve ratio. $AUCR$ with (w/) quinidine/AUC without quinidine.

Table 5. Metoprolol $AUC_{0-\infty}$ With Versus $AUC_{0-\infty}$ Without Concomitant or Delayed Administration of a Single Dose of Tramadol (100 mg)

| Tramadol Administration | Metoprolol $AUC_{0-\infty}$, mg h/L | $AUCR$ |
|-------------------------|------------------------------------|--------|
| w/o                     | 0.50 (0.36-0.74)                   | 1.48   |
| w/, concomitant administration | 0.74 (0.52-1.19) | 1.18   |
| w/, delayed administration of tramadol by 2 h | 0.59 (0.42-0.89) | 1.18   |
| w/, delayed administration of tramadol by 4 h | 0.55 (0.39-0.83) | 1.10   |

$AUC_{0-\infty}$, area under the plasma concentration–time curve from time 0 to infinity; $AUCR$, area under the plasma concentration–time curve ratio. Median (90% prediction interval). $AUCR$, AUC with (w/) tramadol/AUC without (w/o) tramadol.

For the allosteric CYP2D6 inhibition by quinidine, we evaluated 3 scenarios. In the first scenario, we followed the current FDA Guidance for Industry on Clinical Drug Interaction Studies—Cytochrome P450 Enzyme- and Transporter-Mediated Drug Interactions. According to the guidance, an inhibitor can be administered as a single dose if (1) single and multiple doses of the inhibitor are expected to have similar effects on the enzyme of interest and (2) inhibition is not time dependent. In addition, a single dose of the substrate is acceptable if the substrate does not show time-dependent PK (eg, autoinhibition or autoinduction). Since both quinidine and tramadol meet the above-mentioned criteria, we considered concomitant or delayed administrations of single oral doses of quinidine and tramadol in our scenario 1. Results indicate that there is a significant interaction between quinidine and tramadol, which could be attenuated by dose separation (administration of quinidine after the single dose of tramadol). However, this strategy is not of clinical relevance as tramadol requires repeated administration during the day, and CYP2D6 inhibition by quinidine requires about 4 to 5 quinidine half-lives to wear off.
In the second scenario, we accounted for the need of repeated tramadol dosing for chronic pain management. Therefore, tramadol was given orally 4 times daily and dosed to steady state before a single oral dose of quinidine was given. Our results show that CYP2D6 inhibition was most pronounced between 12 and 18 hours after quinidine administration (41% increase in tramadol AUC_{τ,ss} and 51% decrease in O-desmethyltramadol AUC_{τ,ss}) and wore off as quinidine concentrations decreased. Similar results were observed when both tramadol and quinidine were concomitantly administered to steady state (scenario 3). The simulations also showed that CYP2D6 allosteric inhibition by quinidine appears to be long lasting (about 42 hours). Therefore, it cannot be overcome clinically by dose separation as shown in scenarios 2 and 3. This finding is consistent with a previous review paper showing that dose separation will not alleviate allosteric inhibition.20 Given the significant CYP2D6 inhibition and the 4-times-daily dosing regimen of both drugs, coadministration of quinidine and tramadol should be avoided clinically.

For the competitive inhibition with metoprolol, our results indicate that concomitant administration of metoprolol did not significantly affect the exposure of tramadol and O-desmethyltramadol. This was somewhat surprising, as previous drug interaction studies between metoprolol and CYP2D6 low-affinity substrates had suggested that metoprolol was a moderate affinity substrate that could act as a perpetrator drug.22,23 In contrast, recent in vitro characterization of CYP2D6 substrate affinities suggested that tramadol has greater affinity than metoprolol for CYP2D6.45 Hence, our simulations indicate that tramadol acts as perpetrator and will increase total metoprolol exposure (AUC_{0–∞}) by approximately 48%.

Metoprolol is a second-generation β-blocker with high selectivity for β1 adrenoceptors. β1 selectivity for metoprolol diminishes once plasma concentrations exceed 300 nmol/L resulting in inhibition of β2 adrenoceptors in the bronchial and vascular musculature.5 Our simulations show that following competitive CYP2D6 inhibition via tramadol, metoprolol peak plasma concentrations were elevated to 120 ng/mL (≈450 nmol/L) following repeated administration of 100 mg of metoprolol, which is sufficient to exceed the reported β1 selectivity threshold.57 It may consequently be worthwhile exploring clinical outcomes data to determine if adverse events associated with β2 adrenoceptor blockage, such as bronchoconstriction, bronchospasm, and vasoconstriction, actually occur clinically. At the same time, our simulation results indicate that delayed administration of tramadol by 2 hours significantly reduces the interaction with metoprolol resulting in a <20% increase in metoprolol AUC_{0–∞}.

It has further been suggested that the SLC22A1 organic cation transporter (OCT1) plays an important role in the disposition of tramadol active metabolite O-desmethyltramadol. For example, the polymorphic OCT1 has been shown to play an additional role in O-desmethyltramadol exposure in neonates, suggesting that OCT1 is already active early after birth, which may impact the disposition of other OCT1 substrates in this population.58 Furthermore, loss-of-function polymorphisms in OCT1 have been associated with reduced postoperative tramadol consumption, but the respective mechanism is not completely understood yet.59

The expression of OCT1 at the blood-brain barrier (BBB) is also the subject of ongoing investigation, and resultant data can be used to further refine our PBPK model as well as our understanding of observed variability in clinical response to tramadol as it becomes available. It is also widely accepted that opioid-mediated pain relief and euphoria depends on the ability of opioids to cross the BBB. Previous studies in animals have shown that tramadol rapidly penetrates the BBB.60,61 Similar results were obtained with human immortalized brain capillary endothelial cells.61 The situation is somewhat more complex for perpetrators. While the moderately lipophilic metoprolol is readily able to cross the BBB,62 quinidine has restricted access to the brain due to the ABCB1-mediated active eflux at the BBB.63 This difference in the perpetrators’ ability to cross the BBB becomes important as CYP2D6 is also expressed to various degrees in the brain.64 However, the relative contribution of local O-desmethyltramadol formation in the brain to tramadol overall efficacy and safety profile is currently not yet fully understood and warrants further investigation.

**Conclusions**

There is a differentiated impact of quinidine and metoprolol via CYP2D6 inhibition on tramadol and O-desmethyltramadol pharmacokinetics. Following allosteric inhibition by single dose of quinidine, exposure of tramadol was increased and exposure of O-desmethyltramadol was decreased. This DDI cannot be overcome clinically when repeated 4-times-daily tramadol dosing is used. In comparison, there is no significant impact of metoprolol on tramadol and O-desmethyltramadol exposure. To the contrary, we found that tramadol acts as a CYP2D6 perpetrator and increases the exposure of metoprolol, which may be mitigated by dose separation. To better understand the impact of these DDIs on drug efficacy and safety, linking the developed PBPK models to representative
pharmacodynamic end-point models consequently provides a logical next step for this research.

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Conflicts of Interest
V.M., P.D., and J.T. are employees and shareholders of Tabula Rasa HealthCare. The other authors declare no conflicts of interest.

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Data Sharing Statement
The data supporting the findings of this study may be shared on request from the corresponding author Dr. Stephan Schmidt at sschmidt@cop.ufl.edu.

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