RESEARCH ARTICLE

QUALITATIVE ASSESSMENT OF THE FATTY ACID COMPOSITION OF EDIBLE SEAWEEDS FROM COASTAL AREAS OF MANILA BAY AND ROXAS CITY, PHILIPPINES.

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Abstract

This study aimed to come up with a qualitative profile of the fatty acid composition of some of the most abundant seaweeds in the Philippines. Lipid extraction on the freeze-dried samples was done using a modified version of Bligh & Dyer. The lipid extracts were derivatized into fatty acid methyl esters using based-catalyzed transesterification via methanolic potassium hydroxide solution and analyzed using gas chromatography mass spectrometry. Results revealed the presence of the following omega-3 fatty acids: linoleic, eicosatrienoic, and eicosapentaenoic. In addition polyunsaturated omega-6 and monounsaturated C18 fatty acids (oleic and eladic acid) have also been detected. This investigation has demonstrated the potential of edible seaweed as a significant source of omega-3 and other essential fatty acids.

Introduction:

Seaweeds or marine macro-algae, have found a wide variety of applications. It is used as a constituent in pharmaceutical and personal care products; as fertilizers; as feed for livestock, as filtering agent in wastewater treatment; and as gelling agent, stabilizer, or emulsifier. Above all, the most popular use of seaweeds is as an ingredient in various exotic dishes like salads and soups. Seaweeds, widely accepted as health food, have been found to be rich in vitamins and minerals, proteins, fiber, phytochemicals, and have exhibited antibacterial and antioxidant properties (Benjama & Masniyom 2011; Boonchu et al. 2011; Brownlee et al. 2012; Dawczynski et al. 2007; Fleurence 1999; Gessler et al. 2010; Ismail and Hong 2002; Lou et al. 2010; Matanjun et al. 2009; Mohammadi 2013; Norziah and Ching 2000; Ortiz et al. 2006; Polat & Ozogul 2013; Ratana-arporn and Chirapart 2006; Shanab 2007; Seenivasan et al. 2012; Suresh et al. 2012). Studies have also shown that the relatively low lipid composition of seaweeds was found to be rich source of polyunsaturated fatty acids (PUFA) particularly that of omega-3 fatty acids which have been associated with the lowering of plasma cholesterol and triglycerides levels, reduced risk of developing cardiovascular diseases, improved immune system, etc. (Agree et al. 1997; Bonaa et al. 1990; Connor 1994; Harris 1997; Ismail 2003; Horrocks 1999; Kinsella et al., 1990; Kris-Etherton et al., 2003; Oomen et. al. 2000; Schmidt 1997; Stone 1994). The American Heart Association recommended the ingestion of fish, preferably oil, at least twice a week (Kris-Etherton et al. 2003). In line with of the above, this study aimed to come up with a qualitative profile of the fatty acid composition of the most commonly consumed seaweeds in the Philippines.
Materials and Methods:
Seaweeds were gathered in June (Batch 1) and October (Batch 2). There were two areas of collection: Manila Bay in Luzon (*Caulerpa racemosa* Forsk. J.Ag. and *Gracilaria tenuistipitata*) and Roxas City in Central Philippines (*Caulerpa racemosa* Forsk. J.Ag. and *Hynea nidulans* Setchell). The samples were washed repeatedly with tap water and three times with distilled water, freeze-dried, grounded, and stored at -20ºC.

A modified version of Bligh & Dyer method (1959) was used to extract the lipids from the freeze-dried samples. Exactly 2 grams of each sample was successively soaked in the following solvents: methanol, 1:1 methanol:chloroform, and chloroform. After each addition, the mixture is stirred for two minutes and filtered. The filtrates were collected, pooled, and transferred to a separatory funnel. An equal volume of water was added and the mixture was allowed to stand for about three hours. The denser chloroform layer was collected, dried using anhydrous sodium sulphate, and concentrated under vacuum. The crude extract was re-dissolved in 2 mL hexane.

A 2.0 mL of 2M methanolic potassium hydroxide solution was added to each sample. The test tube containing the reaction mixture was covered by a rubber stopper and mixed for 1 minute using a vortex mixer. Afterwards, the upper hexane layer was collected using a glass syringe. Extracts were filtered into its respective glass vials using a Whatman filter, and then subjected to GC-MS analysis.

GCMS model used was the Agilent Technologies 7890A GC System and 5977A MS System using two parameters. The first parameter was designed for optimizing peaks for EPA and DHA. For this method, total run time was set to 35 minutes with an injection volume of 1 μL. Initial temperature was set to 80ºC with an increase of 10ºC per minute until 300ºC was reached. The second parameter was designed to optimize peaks relating to ALA. The total run time was set to 58 mins. with an injection volume 1 μL. Initial temperature was set to 80ºC with an increase of 10ºC per minute until 300ºC was reached. Each run was done in duplicate.

Identification of Seaweeds:
The seaweeds were identified and authenticated by the Botany Division of the Philippine National Museum.

Results:
Table 1 shows the qualitative profile of the saturated fatty acid composition. Results demonstrate that C12-18 saturated fatty acids are present in all the samples regardless of the time and the place of collection. The difference is seen for C20 and above. The presence of C20 and above saturated fatty acids was noted in the green algae *C. racemosa* and very little up to none in the two red algae samples.

### Table 1: Qualitative Profile of the Saturated Fatty Acid Composition

| Common Name       | Notation | *G. tenuistipitata* | *C. racemosa* (Luzon) | *H. nidulans* | *C. racemosa* (Central Philippines) |
|-------------------|----------|---------------------|-----------------------|--------------|-------------------------------------|
| Lauric acid       | C 12:0   | +                   | +                     | +            | +                                   |
| Tridecanoic acid  | C 13:0   | +                   | +                     | +            | +                                   |
| Myristic acid     | C 14:0   | +                   | +                     | +            | +                                   |
| Pentadecanoic acid| C 15:0   | +                   | +                     | +            | +                                   |
| Palmitic acid     | C 16:0   | +                   | +                     | +            | +                                   |
| Heptadecanoic acid| C 17:0   | +                   | +                     | +            | +                                   |
| Searic acid       | C 18:0   | +                   | +                     | +            | +                                   |
| Arachidic acid    | C 20:0   |                     | +                     | +            |                                     |
| Behenic acid      | C 22:0   | +                   | +                     | +            | +                                   |
| Tricosanoic acid  | C 23:0   |                     | +                     | +            | +                                   |
| Lignoceric acid   | C 24:0   | +                   | +                     | +            | +                                   |

The monounsaturated fatty acid (MUFA) composition is summarized in Table 2. Data shows the presence of both cis and trans C18:1 Δ9, oleic and elaidic acid, respectively.
Table 2: Qualitative Profile of the Monounsaturated Fatty Acid (MUFA) Composition

| Common Name          | Notation | G. tenuistiptata Luzon | H. nidulans (Luzon) | C. racemosa (Central Philippines) |
|----------------------|----------|------------------------|---------------------|----------------------------------|
| Myristoleic acid     | C 14:1 Δ9 | +                      | +                   | +                                |
| Pentadecenoic acid   | C 15:1 Δ10 | +                      | +                   | +                                |
| Palmitoleic acid     | C 16:1 Δ9 | +                      | +                   | +                                |
| Heptadecenoic acid   | C 17:1 Δ10 | +                      | +                   | +                                |
| Oleic acid           | C 18:1 Δ9 (cis) | +                      | +                   | +                                |
| Elaidic acid         | C 18:1 Δ9 (trans) | +                      | +                   | +                                |
| Eicosanoic acid      | C 20:1 Δ11 | +                      | +                   | +                                |
| Erucic acid          | C 22:1 Δ13 | +                      | +                   | +                                |

The polyunsaturated fatty acid (PUFA) composition is shown in Table 3. Omega-3 fatty acids ALA, ETA, and EPA were demonstrated to be present in all samples but not DHA. Results also reveal presence of several omega-6 fatty acids, among them are linoleic acid, an essential fatty acid that is incorporated in biological membranes; and arachidonic acid, the precursor of eicosanoids such as prostaglandins, leukotrienes, and thromboxanes.

Table 3: Qualitative Profile of the Polyunsaturated Fatty Acid (PUFA) Composition

| Common Name            | Notation | G. tenuistiptata Luzon | H. nidulans (Luzon) | C. racemosa (Central Philippines) |
|------------------------|----------|------------------------|---------------------|----------------------------------|
| Linoleic acid          | C 18:2 Δ9,12 (Δ-6) | +                      | +                   | +                                |
| Linolenic acid (ALA)   | C 18:3 Δ9, 12, 15 (Δ-3) | +                      | +                   | +                                |
| Eicosadienoic acid     | C 20:2 Δ 11,14 (Δ-6) | +                      | +                   | +                                |
| 8, 11,14 eicosatrienoic acid | C 20:3 Δ 8,11,14 (Δ-6) | +                      | +                   | +                                |
| 11,14,17 eicosatrienoic acid (ETA) | C 20:3 Δ 11,14,17 (Δ-3) | +                      | +                   | +                                |
| Eicosatetraenoic acid (arachidonic acid) | C 20:4 Δ 5,8,11,14 (Δ-6) | +                      | +                   | +                                |
| Eicosapentaenoic acid (EPA) | C 20:5 Δ 5, 8, 11,14,17 (Δ-3) | +                      | +                   | +                                |
| Docosadienoic acid     | C 22:2 Δ 13,16 (Δ-6) | +                      | +                   | +                                |
| Docosahexaenoic acid (DHA) | C 22:6 Δ 4, 7, 10,13,16, 19 (Δ-3) | +                      | +                   | +                                |

Discussion:

Studies done by Gressler et al. (2010), Dawcynzki et al. (2007), Muralidhar et al. (2010), Ortiz et al. (2006), Ratana-arpon & Chirapart (2006), and Sanchez-Machado et al. (2016), showed similar saturated fatty acid profile. Generally, data indicated the presence of polyunsaturated C12-C18 in most of the samples with variations when it comes to C20 or higher. In this study, saturated fatty acids C20 – C24 were detected in the green algae C. racemosa. However they were not observed in G. tenuistiptata and once only in H nidulans, both classified as red algae. The presence of elaidic acid, a trans fatty acid, have also been shown in studies by J. Ortiz (2006). Oleic acid is a naturally occurring fatty acid. On the other hand elaidic, a trans fat, is implicated in the lowering of HDL and increasing LDL levels (Abbey and Nestel 1994). Erucic acid, a fatty acid found in rapeseed, mustard seed, and sunflower seed, was not detected. The presence of erucic diet is a health concern (Food and Standards Australia...
New Zealand (2003) since studies have shown that high amounts of erucic acid in the diet of laboratory rats led to the development of myocardial lipidosis and heart lesions (Chariton et al. 1975). There were some differences in the lipid composition in the June and October collection and such is consistent with studies done by Khairy & El-Shafay (2013) and Nelson et al. (2002).

Omega-3 fatty acids ALA, ETA, and EPA were found to be present in all the samples. However the presence of DHA was not detected. Dawczynski et al. (2007) and Sanchez-Machado et al. (2016) also demonstrated the presence of ALA and EPA. In the work of Dawczynski et al. (2007) docosapentaenoic acid (DPA) was observed instead of DHA in two out of the five samples. Gressler et al. (2010) demonstrated the presence of EPA in some of the samples but ALA, ETA, or DHA was not detected. On the other hand, J. Ortiz et al. (2006) and Ratana-aronp & Chirapart (2006) detected ALA, EPA, and DHA in most of the seaweed samples, while Norziah and Ching (2000) detected EPA and DHA in G. changii. Consistent with previous studies, omega-6 fatty acids were found to be also present. Linoleic acid is a component of phospholipids in biological membrane. On the other hand arachidonic acid is the precursor of eicosanoids like prostaglandins, leukotrienes, and thromboxanes. The US Dietary Guidelines in 2005 have recommended diets rich in MUFA and PUFA (US Dept of Health and Human Services, 2005). However with the potential health risks such as cardiovascular diseases associated with diets high in omega-6 fatty acids, it has been recommended that food rich in PUFA must have relatively high omega-3 as compared to that of omega-6 fatty acids (Intl. Food Information Council 2009; Simopoulos, A. 2010).

Conclusion:
This study has confirmed the presence of essential fatty acids including omega-3 fatty acids in common seaweeds used for both human and animal consumption. These edible seaweeds, with its high nutritional value, can definitely be healthy alternative to the usual meal. However like other food products, possible risks of daily consumption should be evaluated.

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