Basic data on the hematology, serum biochemistry, urology, and organ weights of beagle dogs

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This study was conducted to provide basic data on physiological and hematological characteristics, and organ weights of beagle dogs. A total of 237 beagle dogs were used to determine differences in physiological and hematological parameters, and organ weights depending on sex and age. The respiratory rate of both sexes tended to increase as they grew older and the female heart rate was slightly higher than that of males. Male and female body weights increased rapidly to 33 weeks old followed by a gradual increase to 41-weeks-old. The relative weight of the brain was negatively correlated with body weight, whereas the weight of reproductive organs was positively correlated with body weight. The platelet count of female dogs was slightly higher than that of males. The red blood cell, hemoglobin, and hematocrit of both sexes increased non-significantly with age. In the leukocyte differential count, the neutrophils, and eosinophils of both sexes tended to increase as they grew older, whereas basophils, lymphocytes, and monocytes decreased. In the serum biochemical profiles, alkaline phosphatase was slightly higher in males than females, while the total cholesterol of female dogs at 9-months-old was higher than that of males at the same age. Other biochemical components, including alanine aminotransferase, blood urea nitrogen, creatinine, triglyceride, and total protein increased non-significantly with age in both sexes. To conclude, we observe no significant physiological or hematological differences with sex or age, although decreasing and increasing trends were detected with some parameters. These data provide valuable reference indices of the normal physiological and hematological characteristics of beagle dogs, which should prove useful in toxicological and pharmacological studies.

Key words: Physiology, hematology, serum biochemistry, age, organ weight, beagle dog

The drug development process is typically divided into three major steps: discovery, preclinical development, and clinical trials. The preclinical development of a new drug generally includes studies of efficacy, pharmacology, and experimental toxicology to define the dose, route, and frequency to be investigated in subsequent studies. Efficacy studies demonstrate that treatment with drug candidates has the desired therapeutic effect. Efficacy studies also help to identify the best drug candidates for further development [1].

The regulatory requirements of international guidelines generally demand toxicological and pharmacological studies in at least two laboratory animal species, a rodent and a nonrodent [1]. Beagle dogs are frequently used as model non-rodents for studying the efficacy and safety of newly developed drugs and chemicals, because of similarities in their anatomical morphology and physiology compared to humans [2]. In biomedical research, it is well recognized that individual animal variability can affect study results. To minimize animal variability, researchers should be provided with normal, healthy, well-adapted animals that represent a...
physiological model characterized by a narrow range of physical, behavioral, and clinical parameters [3-5]. Basic data of the test animal’s blood, urine, and blood chemical values have detected many differences depending on sex and age, and the conditions where animals are maintained [7-9]. Thus, animals may be genetically identical but environmental effects during development cannot be ignored and these differences mean that is essential to establish basic data [10]. The establishment of standard basic data is important in the biosciences, because it can reduce and refine the use of laboratory animals [11].

Baseline parameters for hematology, urine, and blood chemical values play an important role in estimating target organ effects when it is difficult to observe the changes in tissues or when they are likely to be ignored during histopathological examinations [3-5]. These basic data can provide a foundation for interpreting certain test results more accurately [10,12]. Thus, it is necessary to acquire different types of basic data for laboratory animals to facilitate the reliable interpretation of a broad range of research studies using model animals. There have been some studies to acquire basic data for laboratory animals [10,12,13] but the volume of data is not great, so many tests are conducted based on foreign basic data [13,14].

The current study aimed to provide basic data on physiological and hematological characteristics, and organ weights for beagle dogs. This study analyzed whether there were differences in physiological and hematological parameters, and the organ weights of male and female beagle dogs. Thus, basic data was generated that should prove valuable to research in fields relevant to basic medical sciences where basic data are required to evaluate tests on dogs.

**Materials and Methods**

**Animal husbandry and maintenance**

Male and female beagle dogs (5 months old) were obtained from Samtako (Beijing Marshall Biotechnology, Beijing, China) and housed in stainless net cages (80W×90L×65H cm) or a fenced room (130W×372L×125H cm) (1 dog per cage). Quarantine and acclimatization periods were 10 days. The animals were housed in a room maintained at a temperature (23±3°C) and a relative humidity (55±10%). The light and dark cycle was 12:12 h (light from 08:00 to 20:00). The temperature and humidity of the housing rooms was measured every hour and no changes were detected that could be considered to have any effect on the tests. The study evaluated 237 beagles including 148 (male 74, female 74) aged 6 months, 52 (male 27, female 25) aged 7 months, and 37 (male 20, female 17) aged 9 months. The 7-month-old and 9-month-old beagles were used as a control group in the toxicity test. The number of beagle dogs in each evaluation class is shown in Table 1.

**Food and water intakes**

Dry food for dogs (Oriental Yeast, Tokyo, Japan) was provided at a rate of 300 g per day, so no food-related factors could have affected the study. Water was sterilized and disinfected using a membrane filter with an UV ray flowing-water sterilizer, and provided ad libitum. Water examination was conducted by the Research Institute of Public Health and Environment (Daejeon, Korea) twice each year. No pollutants were detected in the water that could have influenced the evaluations conducted in this study.

**Blood collection**

Dogs were fasted overnight prior to blood collection. Blood samples were collected from the cephalic veins using a syringe with a 21- or 22-gauge needle. The blood samples were collected in CBC bottles containing EDTA-2K (Green Cross Medical Industry, Yongin, Korea) and used for hematological examination. For the serum biochemical evaluation, 3 mL blood samples were centrifuged at 3,000 rpm for 10 min within 1 h of collection. Sera were stored at −80°C in a freezer prior to analysis. All processes used in the animal testing, including blood-gathering and collection, were reviewed by

| Table 1. Number of beagle dogs in each evaluation class |
|---------------------------------|-----------|-----------|-----------|
| Evaluation parameter            | 6 months old | 7 months old | 9 months old |
|                                | Male       | Female     | Male       | Female     | Male       | Female     |
| Physical examination            | 55         | 56         | -          | -          | 21         | 18         |
| Body weight gain                | 128        | 128        | 56         | 55         | 42         | 20         |
| Organ weights                   | 13         | 13         | 10         | 10         | 15         | 15         |
| Hematology                      | 64         | 63         | 17         | 17         | 12         | 12         |
| Serum biochemistry              | 74         | 74         | 27         | 25         | 20         | 17         |
| Urinalysis                      | 9          | 9          | 9          | 8          | 6          | 5          |
the Institutional Animal Care and Use Committees before they were conducted. Blood collection was conducted after the animals adapted to their feeding conditions.

Hematology

Blood samples were analyzed within 20 min using automatic blood analyzers (T-540 Coulter Counter, Coulter Electronics, Hialeah, FL, USA; ADVIA120 Hematology System, Bayer Healthcare LLC, Tarrytown, NY, USA). The following parameters were determined: white blood cell (WBC) count, differential leukocyte count, red blood cell (RBC) count, hemoglobin concentration, hematocrit, platelet count, mean corpuscular volume, mean corpuscular hemoglobin, and mean corpuscular hemoglobin concentration.

Serum biochemistry

The following serum biochemical parameters were evaluated using an automatic analyzer (Shimadzu CL-7200, Shimadzu, Kyoto, Japan): alkaline phosphatase (ALP), alanine aminotransferase (ALT), aspartate aminotransferase (AST), total bilirubin (TB), total cholesterol (TC), triglycerides (TG), phospholipids, total protein (TP), albumin, albumin/globulin (A/G) ratio, glucose, blood urea nitrogen (BUN), creatinine, creatine phosphokinase (CPK), calcium, and inorganic phosphorus. Sodium, potassium, and chloride were measured using an automatic electrolyte analyzer (Ciba-Corning 644 Na/K/Cl Analyzer, Ciba-Corning, Medfield, MA, USA).

Statistical analysis

Data processing and statistical analysis were performed using SAS version 9.2 (SAS Institute Inc., Cary, NC, USA). Values determined for different sexes and ages were compared using two-way ANOVA (SAS Version 9.2). A one-way ANOVA test (SAS Version 9.2) was applied at each time interval to compare the reproductive organ weight within each sex. Duncan’s multiple-comparison test was used to assess the statistical significance of differences among the study groups. A level of $P<0.05$ was accepted as statistically significant.

Results

Physiological values

A significant increase in the body temperature was observed in male dogs ($P<0.001$). The mean body temperature in 6-month-old male dogs was $38.3\pm0.4^\circ C$, whereas that in 9-month-old male dogs was $39.0\pm0.3^\circ C$. In both sexes, the respiratory rate increased significantly with age ($P<0.001$), while the heart rate of females was slightly higher than that of males (Table 2).

Body weight gain and organ weights

The body weight of male and females increased rapidly up to 33 weeks ($0.18\pm0.07$ kg/week for male dogs, $0.14\pm0.06$ kg/week for female dogs), before increasing gradually up to 41 weeks ($0.14\pm0.06$ kg/week for males, $0.11\pm0.08$ kg/week for females). The body weight of males was significantly higher than that of females (Figure 1, $P<0.001$). The absolute and relative weights of the thyroid, heart, lung, liver, kidney, and adrenals increased significantly as both sexes grew older, whereas male organs weighed significantly more than those of females (Table 3, $P<0.001$). The thymus gradually decreased in weight as both sexes grew older ($P<0.001$). Brain, pituitary, and spleen weights increased significantly with age (Tables 3 and 5, $P<0.001$). In absolute and relative terms, the reproductive organ weight of males and females, including the epididymis, prostate, and uterus, increased significantly as they grew older ($P<0.05$), whereas the weight of the ovaries did not increase significantly with age (Table 4). The absolute weight of testes increased significantly from 6 to 9 months, while their relative weight increased significantly from 6 to 7 months ($P<0.05$).
The results of the hematological examination are shown in Tables 7 and 8. The platelet count of female dogs was slightly higher than that of males. The RBC, hemoglobin, and hematocrit of both sexes increased non-significantly with age. In the leukocyte differential count, the neutrophils and eosinophils of both sexes tended to increase with age, whereas basophils, lymphocytes, and monocytes decreased.

**Biochemical parameters**

The results of the serum biochemical profiles are shown in Table 9. The ALP of male beagles was slightly higher than that of females, while the TC in female dogs aged 9 months was higher than that of males at the same age. The remaining biochemical parameters, including ALT, BUN, creatinine, TG, and TP, increased non-significantly with age in both sexes.

**Urological parameters**

The amount of urine and sodium ions tended to increase as dogs grew older. However, there were large individual differences (Table 10).

**Discussion**

Beagle dogs are the most frequently used nonrodent animals in biomedical research, because of similarities in their...
Basic data on the physiology, hematology, and organ weights in beagle dog

**Table 5. Changes in the relative organ weights of male and female dogs**

| Sex  | Organ (unit) | Age (month) | 6 (♂=13, ♀=13) | 7 (♂=10, ♀=10) | 9 (♂=15, ♀=15) |
|------|--------------|-------------|----------------|----------------|----------------|
| Male | Body weight (kg) | 7.50±0.78 | 8.53±1.04 | 9.86±0.94 |
|      | Thyroid (%) | 0.071±0.008 | 0.068±0.014 | 0.080±0.015 |
|      | Heart (%) | 0.77±0.06 | 0.77±0.09 | 0.78±0.09 |
|      | Lung (%) | 1.02±0.12 | 0.81±0.11 | 0.85±0.15 |
|      | Liver (%) | 2.95±0.36 | 2.74±0.33 | 2.73±0.41 |
|      | Kidney (%) | 0.46±0.05 | 0.44±0.03 | 0.41±0.06 |
|      | Adrenals (%) | 0.104±0.020 | 0.116±0.019 | 0.110±0.020 |
|      | Thymus (%) | 0.127±0.017 | 0.080±0.013 | 0.056±0.001 |
|      | Brain (%) | 0.97±0.11 | 0.88±0.12 | 0.75±0.09 |
|      | Pituitary (%) | 0.007±0.001 | 0.007±0.001 | 0.006±0.001 |
|      | Spleen (%) | 0.28±0.04 | 0.27±0.04 | 0.28±0.04 |
| Female | Body weight (kg) | 6.80±0.70 | 7.09±0.76 | 8.74±1.25 |
|      | Thyroid (%) | 0.071±0.013 | 0.070±0.012 | 0.075±0.013 |
|      | Heart (%) | 0.79±0.07 | 0.80±0.08 | 0.79±0.13 |
|      | Lung (%) | 0.94±0.09 | 0.76±0.06 | 0.86±0.08 |
|      | Liver (%) | 2.81±0.39 | 2.68±0.45 | 2.65±0.66 |
|      | Kidney (%) | 0.43±0.03 | 0.40±0.04 | 0.40±0.06 |
|      | Adrenals (%) | 0.135±0.025 | 0.141±0.038 | 0.125±0.029 |
|      | Thymus (%) | 0.100±0.026 | 0.078±0.018 | 0.058±0.025 |
|      | Brain (%) | 1.04±0.13 | 0.93±0.08 | 0.83±0.10 |
|      | Pituitary (%) | 0.008±0.002 | 0.007±0.001 | 0.006±0.001 |
|      | Spleen (%) | 0.31±0.08 | 0.31±0.07 | 0.30±0.05 |

1Indicates that body and organ weights increased significantly with age and there were significant differences between sexes (P<0.001). 2Indicates that organ weights decreased significantly with age and there were significant differences between sexes (P<0.001). 3Indicates that organ weights increased significantly with age (P<0.02).

**Table 6. Changes in the relative reproductive organ weights of male and female dogs**

| Sex  | Organ (unit) | Age (month) | 6 (♂=13, ♀=13) | 7 (♂=10, ♀=10) | 9 (♂=15, ♀=15) |
|------|--------------|-------------|----------------|----------------|----------------|
| Male | Testes (%) | 0.58±0.30 | 1.09±0.39 | 1.36±0.31 |
|      | Epididymis (%) | 0.149±0.017 | 0.220±0.013 | 0.255±0.027 |
|      | Prostate (%) | 0.15±0.03 | 0.31±0.11 | 0.40±0.28 |
| Female | Ovaries (%) | 0.079±0.014 | 0.101±0.066 | 0.079±0.039 |
|      | Uterus (%) | 0.13±0.02 | 1.17±1.09 | 1.15±0.74 |

123Values with different superscripts in the same row were significantly different (P<0.05).

anatomical morphology and physiology to humans [2]. In biomedical research, it is well recognized that individual animal variability can affect study results. Therefore, highly defined dog models should be used in biomedical studies [3]. In clinical or non-clinical tests, the normal range of blood constituent is a standard means of diagnosis when identifying diseases or abnormalities. Thus, they are very important values, although they can vary significantly depending on the individual environment (e.g., species, sex, age, and conditions) or genetic characteristics [13,15-17]. This means it is essential to reduce errors by determining the normal range as accurately as possible to facilitate the exact diagnosis of diseases or confirming abnormalities. The current study was conducted to determine basic physiological data on the body weight, organ weight, body temperature, heart rate, and hematological, biochemical and urological parameters for male and female beagle dogs.

In the physical examination of male and female beagle dogs aged 6 and 9 months, the body temperature and respiratory rate did not differ significantly between the sexes, although the respiratory rate of both sexes tended to increase as they grew older, while the heart rate of females was slightly higher than that of males (Table 2). Ferasin et al. [18] reported that dogs aged less than 1 year appeared to have a significantly higher heart rate than older dogs, while the heart rate of healthy dogs undergoing routine clinical examination was related to their age.
The body weight of male and female dogs increased rapidly up to 33 weeks, followed by a gradual increase to 41 weeks. The body weight was significantly different between the sexes (Figure 1). Their body weight increased by 0.18 ± 0.07 kg/week in male dogs and by 0.14 ± 0.06 kg/week in female dogs. Nunamaker et al. [19] reported that the body weight increased by 19.7 g/day in a control group of dogs. The relative organ weights were similar to those reported for a control group in a previous report [20]. The relative reproductive organ weights of both sexes increased rapidly up to 7 months (Table 6). These results indicate that puberty may be reached at 7 months of age.

Table 7. Hematological parameters of male and female dogs

| Sex     | Parameter                                      | Age (month)       |
|---------|------------------------------------------------|-------------------|
|         |                                                | 6 (♂=64, ♀=63)    | 7 (♂=17, ♀=17)    | 9 (♂=12, ♀=12)    |
| Male    | White blood cell (10^3/mm³)                     | 9.29±2.14         | 9.15±1.77         | 9.01±1.27         |
|         | Red blood cell (10^6/mm³)                      | 6.63±0.53         | 6.91±0.63         | 7.29±0.94         |
|         | Hemoglobin (g/dL)                              | 14.9±1.2          | 15.5±1.5          | 16.5±2.1          |
|         | Hematocrit (%)                                 | 42.8±3.4          | 44.7±4.2          | 46.7±6.4          |
|         | Platelet (10^9/mm³)                            | 327.3±59.3        | 314.8±61.7        | 294.3±66.7        |
|         | Mean corpuscular volume (fL)                   | 64.6±2.2          | 64.7±2.4          | 64.1±2.8          |
|         | Mean corpuscular hemoglobin (pg)               | 22.4±0.6          | 22.5±0.8          | 22.6±0.7          |
|         | Mean corpuscular hemoglobin concentration (g/dL)| 34.6±1.4          | 34.7±1.3          | 35.4±1.8          |

Female

| Sex     | Parameter                                      | Age (month)       |
|---------|------------------------------------------------|-------------------|
|         |                                                | 6 (♂=64, ♀=63)    | 7 (♂=17, ♀=17)    | 9 (♂=12, ♀=12)    |
|         | White blood cell (10^3/mm³)                     | 8.43±1.47         | 8.87±1.42         | 9.18±2.66         |
|         | Red blood cell (10^6/mm³)                      | 6.79±0.52         | 6.91±0.63         | 7.21±0.58         |
|         | Hemoglobin (g/dL)                              | 15.4±1.1          | 16.3±1.2          | 16.8±1.2          |
|         | Hematocrit (%)                                 | 44.4±3.5          | 44.7±4.2          | 47.2±4.4          |
|         | Platelet (10^9/mm³)                            | 332.5±58.5        | 319.8±54.5        | 304.4±51.5        |
|         | Mean corpuscular volume (fL)                   | 65.3±2.4          | 64.7±2.4          | 65.5±4.0          |
|         | Mean corpuscular hemoglobin (pg)               | 22.5±0.7          | 22.9±0.8          | 23.3±0.8          |
|         | Mean corpuscular hemoglobin concentration (g/dL)| 34.5±1.3          | 34.7±1.3          | 35.6±1.5          |

Table 8. Differential leukocyte counts of male and female dogs

| Sex     | Parameter                                      | Age (month)       |
|---------|------------------------------------------------|-------------------|
|         |                                                | 6 (♂=64, ♀=63)    | 7 (♂=17, ♀=17)    | 9 (♂=12, ♀=12)    |
| Male    | Neutrophils (%)                                | 52.2±6.7          | 54.4±8.0          | 55.6±4.9          |
|         | Lymphocytes (%)                                | 35.1±5.9          | 34.8±7.4          | 33.4±3.1          |
|         | Eosinophils (%)                                | 5.0±2.1           | 5.1±2.1           | 5.3±2.7           |
|         | Monocytes (%)                                  | 5.2±1.3           | 5.1±0.8           | 4.5±1.1           |
|         | Basophils (%)                                  | 1.1±0.6           | 0.7±0.4           | 0.8±0.7           |
| Female  | Neutrophils (%)                                | 53.8±5.3          | 54.2±4.4          | 58.3±4.8          |
|         | Lymphocytes (%)                                | 36.3±5.0          | 35.4±4.9          | 31.3±3.8          |
|         | Eosinophils (%)                                | 3.9±1.9           | 3.9±1.9           | 5.1±3.8           |
|         | Monocytes (%)                                  | 4.2±0.9           | 4.3±1.1           | 4.0±1.1           |
|         | Basophils (%)                                  | 1.2±0.7           | 0.8±0.6           | 0.9±0.7           |

The body weight of male and female dogs increased rapidly up to 33 weeks, followed by a gradual increase to 41 weeks. The body weight was significantly different between the sexes (Figure 1). Their body weight increased by 0.18±0.07 kg/week in male dogs and by 0.14±0.06 kg/week in female dogs. Nunamaker et al. [19] reported that the body weight increased by 19.7 g/day in a control group of dogs. The relative organ weights were similar to those reported for a control group in a previous report [20]. The relative reproductive organ weights of both sexes increased rapidly up to 7 months (Table 6). These results indicate that puberty may be reached at 7 months of age.

Age-related differences were found in terms of RBC count, hemoglobin, hematocrit, neutrophils, eosinophils, basophils, lymphocytes, monocytes, ALT, BUN, creatinine, TG, and TP. Sex-related differences were found for ALP and TC (Tables 7, 8, and 9). However, these parameters were not significantly related to age or sex. These results were slightly different to those reported by Jeong et al. [12]. The number of eosinophils was higher than that reported by Jeong et al. [12], but similar to that reported by Wolford et al. [21]. The number of lymphocyte was also higher than that reported by Jeong et al. [12] and Wolford et al. [21]. This was considered to be a consequence of transportation stress, but it may also be
related to a vaccination before their final export. ALP is an enzyme found in the canalicular membrane or sinusoidal membrane, and it can be used to measure the degree of damages to these parts. However, ALP is also secreted by the bones, intestines, and placenta [22,23]. Thus, it can be increased with motion and during periods of growth and pregnancy [14]. We found a progressive reduction in serum ALP concentration as both sexes grew older. This suggests that the observed change in the ALP level is related to the turnover of bone. Uchiyama et al. [23] and Ikeuchi et al. [14] also observed a reduction in plasma ALP concentration with age in dogs. However, this biochemical blood parameter did not exceed the normal range and there was no difference between males and females. Hematological examinations, biochemical examinations of blood, and urine examinations indicate considerable differences depending on the species, source of supply, maintenance conditions of animals, and age, as well as the analytical equipment used to conduct tests [25-27].

Therefore, it is important that each research institute

| Table 9. Serum biochemistry parameters of male and female dogs |
|---------------------------------|-----------------|-----------------|-----------------|
| **Sex**  | **Item**                   | **Age (month)** |                 |
|         | ALP (IU/L)                 | 6 (♂ =74, ♀ =74) | 7 (♂ =27, ♀ =25) | 8 (♂ =20, ♀ =17) |
| Male    | 351.65±126.81              | 34.24±5.55      | 27.77±11.20     | 10.99±2.22       |
|         | 282.99±67.74               | 35.50±5.04      | 28.25±9.90      | 11.18±2.35       |
|         | 0.70±0.14                  | 0.73±0.12       | 0.81±0.14       | 0.81±0.14        |
|         | 92.44±8.69                 | 91.20±11.56     | 91.20±11.56     | 94.02±9.29       |
|         | 158.15±29.65               | 151.55±29.29    | 160.33±30.98    | 160.33±30.98     |
|         | 30.02±10.00                | 27.57±9.89      | 26.82±11.09     | 26.82±11.09      |
|         | 5.21±0.27                  | 5.32±0.34       | 5.61±0.28       | 5.61±0.28        |
|         | 2.58±0.18                  | 2.57±0.18       | 2.70±0.17       | 2.70±0.17        |
|         | 0.04±0.02                  | 0.04±0.02       | 0.06±0.03       | 0.06±0.03        |
|         | 0.99±0.13                  | 0.94±0.08       | 0.93±0.08       | 0.93±0.08        |
|         | 6.01±0.58                  | 5.73±0.59       | 5.49±0.51       | 5.49±0.51        |
|         | 4.99±0.33                  | 4.93±0.27       | 5.02±0.28       | 5.02±0.28        |
|         | 108.01±3.01                | 108.81±1.55     | 110.25±1.55     | 110.25±1.55      |
|         | 10.92±0.43                 | 10.87±0.50      | 11.02±0.39      | 11.02±0.39       |
|         | 143.41±3.07                | 143.37±2.87     | 144.50±0.71     | 144.50±0.71      |
| Female  | 317.43±98.4                | 273.79±71.68    | 205.85±61.87    | 205.85±61.87     |
|         | 34.36±7.58                 | 37.48±6.94      | 30.88±6.55      | 30.88±6.55       |
|         | 25.53±6.69                 | 29.24±8.58      | 28.25±5.09      | 28.25±5.09       |
|         | 10.79±2.13                 | 12.07±1.58      | 13.11±3.08      | 13.11±3.08       |
|         | 0.72±0.12                  | 0.76±0.13       | 0.84±0.14       | 0.84±0.14        |
|         | 97.78±10.01                | 94.87±10.88     | 95.11±9.05      | 95.11±9.05       |
|         | 155.27±30.20               | 152.45±26.95    | 168.99±26.93    | 168.99±26.93     |
|         | 26.52±7.85                 | 27.07±7.39      | 27.24±7.91      | 27.24±7.91       |
|         | 5.25±0.31                  | 5.32±0.28       | 5.62±0.37       | 5.62±0.37        |
|         | 2.67±0.18                  | 2.68±0.18       | 2.81±0.15       | 2.81±0.15        |
|         | 0.06±0.03                  | 0.08±0.04       | 0.09±0.04       | 0.09±0.04        |
|         | 1.04±0.12                  | 1.02±0.09       | 1.01±0.11       | 1.01±0.11        |
|         | 5.81±0.66                  | 5.52±0.59       | 5.03±0.46       | 5.03±0.46        |
|         | 4.76±0.27                  | 4.62±0.27       | 4.70±0.24       | 4.70±0.24        |
|         | 108.38±2.95                | 109.60±2.16     | 110.88±2.89     | 110.88±2.89      |
|         | 11.06±0.50                 | 11.05±0.63      | 11.13±0.46      | 11.13±0.46       |
|         | 144.27±2.77                | 144.20±2.31     | 143.71±3.35     | 143.71±3.35      |

ALP, alkaline phosphatase; AST, aspartate aminotransferase; ALT, alanine aminotransferase; BUN, blood urea nitrogen; TC, total cholesterol; TG, triglycerides; TP, total proteins; TB, total bilirubins; A/G, albumin/globulin. Indicates that the parameter decreased significantly with age (P<0.0001), while the parameter was significantly higher for males compared with females (P=0.03). *Indicates that the parameter increased significantly with age in both sexes (P<0.0001). Indicates that the parameter was significantly higher in males compared with females (P=0.008). *Indicates that the parameter increased significantly with age in both sexes, while the parameter was significantly higher in females compared with males (P<0.0001). 5Indicates that the parameter was significantly higher in females compared with males (P<0.0001).
maintains standard data appropriate for their facility to facilitate the accurate interpretation of test results. Test results are also influenced by various factors, so it is necessary to minimize any deviations due to the external environment by formulating an SOP covering conditions such as the period of specimen collection, environmental conditions, animal stress status, and animal feeding activity.

References

1. Steinmetz KL, Spack EG. The basics of preclinical drug development for neurodegenerative disease indications. BMC Neurol 2009; 9(Suppl 1): S2.
2. Smith D, Broadhead C, Descotes G, Fosse R, Hack R, Krauser K, Pfister R, Phillips B, Rabemampianina Y, Sanders J, Sparrow S, Stephan-Guedeliner M, Jacobsen SD. Preclinical safety evaluation using nonrodent species: an industry/welfare project to minimize dog use. ILAR J 2002; 43(Suppl 1): S39-S42.
3. Meunier LD. Selection, acclimation, training, and preparation of dogs for the research setting. ILAR J 2006; 47(4): 326-347.
4. Smith D, Combes R, Depelchin O, Jacobsen SD, Hack R, Luft J, Lamens L, von Landenberg F, Phillips B, Pfister R, Rabemampianina Y, Sparrow S, Stark C, Stephan-Gueeldiner M. Optimising the design of preliminary toxicity studies for pharmaceutical safety testing in the dog. Regul Toxicol Pharmacol 2005; 41(2): 95-101.
5. Phillips B, Smith D, Combes R, Descotes G, Durying Jacobsen S, Hack R, Kembkowski J, Krauser K, PWster R, Rabemampianina Y, Sparrow S, Stephan-Gueeldner M, von Landenberg F. An approach to minimise dog use in regulatory toxicology: production of a best practice guide to study design. Altern Lab Anim 2004; 32(5): 447-451.
6. Swallow J, Anderson D, Buckwell AC, Harris T, Hawkins P, Kirkwood J, Lomas M, Meacham S, Peter A, Prescott M, Owen S, Quest R, Sutcliffe R, Thompson K. Guidance on the study design. Altern Lab Anim 2004; 32S1: 447-451.
7. Lammens L, von Landenberg F, Rabemampianina Y, Sparrow S, Stephan-Gueeldner M, von Landenberg F. An approach to minimise dog use in regulatory toxicology: production of a best practice guide to study design. Altern Lab Anim 2004; 32(5): 447-451.
8. Burkholder WJ. Use of body condition scores in clinical assessment of the provision of optimal nutrition. J Am Vet Med Assoc 2000; 217: 650-654.
9. Fillman-Holliday D, Landi MS. Animal care best practices for regulatory testing. ILAR J 2002; 43: S49-S58.
10. Ha CS, Son HY, Kang BY, Cha SW, Park JI, Han SS. A study of hematology and serum chemistry in beagle dogs. Korean J Lab Anim Sci 1995; 11(2): 153-158.
11. Michael B, Patric A, Susanne B, Silvia C, Sandra C, Richard C, Robert C, Raffaella C, Rodger C, Chandra T, Julia F, Laura G, Marlies H, Thomas H, Sebastian H, Leonnard S, Laurie S, Horst S, William S, Raymond T, Drew W, Valerie Z. The principles of weight of evidence validation of test methods and testing strategies. Altern Lab Anim 2006; 34(6): 603-620.
12. Jeong EJ, Han SC, Cha SW, Lee HS, Ha CS, Kim CY. Hematological and blood biochemical values of laboratory beagle dogs. Lab Anim Res 2007; 23(3): 223-229.
13. Matsuzawa T, Nomura M, Unno T. Clinical pathology reference ranges of laboratory animals. Working Group II, Nonclinical Safety Evaluation Subcommittee of the Japan Pharmaceutical Manufacturers Association. J Vet Med Sci 1993; 55 (13): 351-361.
14. Ikeuchi J, Yoshizaki T, Hirata M. Plasma biochemistry values of young beagle dogs. 1991; 16: 49-59.
15. Matsuzawa T, Nomura M, Yonezawa H, Unno T. Selection of appropriate parameters, use of a quality control concept, and suitable statistical analyses for clinical pathology examination of animals in toxicity studies: results of a current survey by the Japanese Pharmaceutical Manufacturers Association. Comp Haematol Int 1995; 5: 196-200.
16. Conder JC, Laber K. A renewed look at laboratory rodent housing and management. LAR J 2007; 48(1): 29-36.
17. Sharp J, Zammit T, Aza T, Lawson D. Stress-like responses to common procedures in individually and group housed female rats. Contemp Top Lab Anim Sci 2003; 42(1): 9-18.
18. Ferasin L, Ferasin H, Little CJL. Lack of correlation between canine heart rate and body size in veterinary clinical practice. J Small Anim Pract 2010; 51: 412-418.
19. Nunemaker E, Newhall K, Thompson C, Lucas A, Owens J, Sherman JG. Safety evaluation and treatment affect of LY2190416, a CB-1 antagonist/inverse agonist in growing beagle dogs. J Vet Pharmacol Ther 2011; doi: 10.1111/j.1365-2885.2011.01280.x.
20. Shin JW, Park HK, Kwon M, Son CG. Scientific evaluation of the herbal medicine CGX in beagle dogs. Food Chem Toxicol 2010; 48: 743-749.
21. Wofford ST, Schroer RA, Gohs FX, Gallo PE, Brodeck M, Falk HB, Ruhren R. Reference range data base for serum

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Table 10. Urinary parameters of male and female dogs

| Sex     | Item                     | Age (month) | Age (month) | Age (month) |
|---------|--------------------------|-------------|-------------|-------------|
| Male    | Volume (mL)              | 185.89±74.58| 225.78±69.17| 195.50±67.15|
|         | pH                       | 7.56±0.98   | 8.11±0.93   | 8.50±1.00   |
|         | Specific gravity         | 1.04±0.01   | 1.04±0.01   | 1.04±0.01   |
|         | Sodium (mmol/day)        | 9.78±3.92   | 14.93±8.75  | 19.86±6.75  |
|         | Potassium (mmol/day)     | 33.19±17.85 | 36.88±13.52 | 41.33±9.10  |
|         | Chloride (mmol/day)      | 36.01±13.46 | 37.65±8.34  | 40.16±6.79  |
| Female  | Volume (mL)              | 198.22±81.60| 270.38±133.70| 255.50±82.73|
|         | pH                       | 7.56±1.10   | 8.63±0.73   | 9.00±0.00   |
|         | Specific gravity         | 1.04±0.01   | 1.04±0.01   | 1.02±0.02   |
|         | Sodium (mmol/day)        | 9.40±4.64   | 13.87±6.78  | 16.17±7.77  |
|         | Potassium (mmol/day)     | 36.58±20.05 | 38.89±15.42 | 40.82±10.33 |
|         | Chloride (mmol/day)      | 28.80±8.16  | 36.31±7.02  | 35.70±6.64  |
chemistry and hematology values in laboratory animals. J Toxicol Environ Health 1986; 18: 161-188.
22. Posen S, Doherty E. The measurement of serum alkaline phosphatase in clinical medicine. Adv Clin Chem 1981; 22: 163-245.
23. Amacher DE, Higgins CV, Schomaker SJ, Clay RJ. Further characterization of serum alkaline phosphatase from male and female beagle dogs. Enzyme 1989; 42: 1-7.
24. Uchiyama T, Tokoi K, Deki T. Successive changes in the blood composition of experimental normal beagle dogs associated with age. Jikken Dobutsu 1985; 34(4): 367-377.
25. Bayard SP. Another look at the statistical analysis of changes during storage of serum specimens. Health Lab Sci 1974; 26: 959-960.
26. Caisey JD, King DJ. Clinical chemical values for some common laboratory animals. Clin Chem 1980; 26: 1977-1979.
27. Kaspar LV, Norris WP. Serum chemistry values of normal dogs (beagles): associations with age, sex, and family line. Lab Anim Sci 1977; 27(6): 980-985.