Research Article

Raphanus sativus L. Extract as a Scale and Corrosion Inhibitor for Mild Steel in Tap Water

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1. Introduction

Utilization of natural water for both domestic and industrial purposes is related with scale and corrosion problems. Water containing dissolved gases such as O2 and CO2 can cause corrosion of metal equipment [1, 2], and the presence of Ca2+ and Mg2+ cations leads to scaling [3, 4], especially when water is used in heat exchange processes. As a result of corrosion, water quality deteriorates and metal equipment requires repairs and replacement more often. Scaling leads to extra fuel consumption and flow reduction. So, both the problems lead to economic losses and require the search of new efficient solutions.

The trend of today is application of “green” reagents that are environmentally friendly and biodegradable and can be obtained without any hazardous chemical synthesis [5–8]. Muryanto et al. studied malic acid as a scale inhibitor [9]. It was found that addition of malic acid in the concentration of 3–5 ppm reduces the scale mass 2 times and more. The formed crystals’ morphology is also changed. Al-Roomi et al. tested novel low molecular weight copolymers (termed as YMR-series polymers) prepared through reaction of maleic anhydride as a new generation scale inhibitor [10]. They found that scale inhibition for steel and copper was up to 98%. Peronno et al. investigated antiscaling properties of poly(acrylic acid-co-maleic acid) and polyaspartic acid [11]. Antiscaling efficiency was observed in the very low concentration of 4 mg·L−1, and the scale structure was modified in the presence of an inhibitor. One of the promising sources of such reagents is natural extracts. Recently, the application of aqueous extract of Paronychia argentea [12], Spergularia rubra, and Parietaria officinalis aqueous solutions [13] as a scale inhibitor was reported. The proposed action mechanisms include complex formation with calcium ions that keep Ca2+ in the solution and prevent formation of CaCO3; also, inhibitor molecules are believed to be adsorbed on the growing crystals thus blocking them from Ca2+ supply.

Many “green” substances were tested as corrosion inhibitors. Ibrahimi et al. in their review analysed amino acids as corrosion inhibitors [14]. The main mechanism, similar to scale inhibition, was adsorption of inhibitor molecules on the metal surface and formation of the surface film that prevent oxygen ingress. Zhang tested synergistic effect of the combination of polyaspartic acid (PASP), polyepoxysuccinic acid (PESA), polyamino polyether methylenephosphonate (PAPEMP), sodium gluconate (Glu), and Zn2+ on carbon...
2. Experimental Section

2.1. Extract Preparation. The extract was prepared by maceration of radish cake for 24 h in the ethanol. The raw material powder and ethanol were mixed in the weight ratio of 1:10. The resultant mixture was filtered with Whatman filter paper to remove solid contents.

2.2. GC-MS Extract Composition Analyses. Chemical composition of extract was analysed by GC-MS analysis (gas chromatography with a mass selective detector by mass selective integration). A Shimadzu gas chromatograph (model GC 17A) equipped with a flame ionization detector (FID) was operated under the following conditions: capillary fused silica column (CBP-5) (length, 25 m; internal diameter, 0.25 mm; film thickness, 0.22 μm) and ion source temperature of 280°C. The GC-MS was operated in the electron impact ionization mode (EI) at 70 eV. The oven temperature was programmed as follows: the initial temperature of 50°C was maintained for 2 min and then increased to 200°C at the rate of 10°C/min and held for 5 min. Afterwards, the temperature was ramped to 250°C at the rate of 25°C/min (for 15 min). Other operating conditions included the following: carrier gas, He (99.99%), inlet pressure 76 kPa, with a linear velocity of 20 cm/s; injector temperature 250°C; detector temperature 310°C; and split ratio 1:25. The relative contents of the chemical components of the extract were found with the help of the method of internal normalization of the areas of peaks without correcting the sensitivity coefficients. The percentage of each compound was determined from its peak area and the sum of the areas of all peaks detected in the total ion current (TIC) trace. The identification and quantification of the components was carried out by using commercially available databases: Nist 05 (National Institute of Standards and Technology, USA) and Mass Finder3 (Dr. Hochmut, Scientific Consulting, Germany).

2.3. Chronoamperometry. Electrochemical technique of chronoamperometry is a fast and convenient approach to study scaling processes [1, 2]. The working electrode is polarized at a constant negative potential of −1.1 V/SSCE (saturated silver chloride electrode $E = 0.2 V/NHE$). At this potential, the dissolved oxygen is reduced to $OH^−$ ions causing the alkalization of the pre-electrode layer. The carbonic equilibrium is shifted to form carbonates, and $CaCO_3$ is deposited on the electrode surface:

$$\begin{align*}
O_2 + 2H_2O + 4e = 4OH^- \\
HCO_3^- + OH^- = CO_3^{2-} + H_2O
\end{align*}$$

The current reduction means the scale is formed on the electrode surface; meanwhile, the unchanged current value means no scale is deposited. Also, the current-time dependence provides the information of scaling time $t_s$ (theoretical time value, needed to fully cover the electrode surface) and scale porosity SP (the ratio between residual and initial current values).

Chronoamperometry tests were performed in a conventional three-electrode electrochemical cell. The working electrode was a mild steel cylinder (diameter, 6 mm; length, 30 mm) rotated at 500 rpm. The electrode surface was mechanically abraded with P600 emery paper, degreased, and etched in 200 g/l HCl prior to the test. A counter electrode, platinum grid, and a saturated silver chloride electrode were as reference one. The tap water (composition is given in Table 1) of 1 l volume was kept at 25 ± 0.5°C during the test. The mass of the deposited carbonates was determined by weighing the working electrode before and after electrochemical measurements with 0.0001 g accuracy. Polarization continued for 3 h. Each test was repeated 3 times, and the data were averaged.

2.4. Thermal Scaling. Antiscalant efficiency of the extract was also tested in thermal scaling conditions. The scale was deposited on the surface of the steel rings placed on top of the Pyrex glass tube. The heating spiral was placed inside the tube and connected to the electric transformer. The electric power of the spiral was 70 W. The tube with steel rings was placed inside the glass beaker filled with tap water. The beaker was covered on the top to prevent solution evaporation. The water was replaced daily.

The scale growth rate with and without antiscalant was determined by weighing the tube every 20 hours of boiling.

The ability of the extract to form water-soluble complexes with hardiness cations was tested titrimetrically. For this purpose, water hardness was determined titrimetrically before and after daily test. A solution sample was buffered to pH 10.1, the indicator was added, and EDTA was used as the
was calculated according to the following formula:

\[ \eta = \frac{\rho_0 - \rho_1}{\rho_2 - \rho_0} \times 100\% \]  

(3)

where \( \rho_0 \) is the solution hardness before heating, \( \rho_1 \) is the solution hardness after heating without antiscalant, and \( \rho_2 \) is the solution hardness after heating with antiscalant.

### 2.5. Polarization Resistance Measurement.
Corrosion inhibition efficiency of the extract was determined with the linear polarization resistance (LPR) technique in thermal scaling conditions [20–22]. The pair of steel rings placed on the Pyrex tube was electrically connected to corrometer and worked as a corrosion probe. LPR was measured every 15 min in a galvanostatic mode: the current density of 5 A/cm² was applied to the probe, and the potential shift was measured after 30 s [23]. The initial potential difference and ohmic drop were automatically compensated, and the polarization resistance value was recalculated into steel corrosion rate with Stern’s equation using the conventional B value of 26 mV [24].

### 2.6. Scale Characterization.
The morphology of the surface after thermal scaling was studied using a SEM-106I Selmi microscope (Ukraine) operated at 20 kV. The steel rings were removed from the Pyrex tube, dried, and sent to analyses.

FT-IR spectroscopy was performed to determine the main groups in the extract and the formed surface layer. The scale from the steel rings was scraped off with a surgery blade and sent to FT-IR. Fourier transform infrared spectroscopy (FT-IR) spectra were recorded on a FT-IR spectrometer (Thermo Scientific Nicolet 6700 FT-IR spectrometer (spectral resolution 0.5 cm⁻¹)) using the KBr pellet technique. Spectra were recorded over 450–4000 cm⁻¹.

### 3. Results and Discussion

#### 3.1. GC-MS and FT-IR Analysis of Extract.
GC-MS was used to identify the organic compounds present in the radish cake extract (RCE). The GC-MS spectra of the RCE show prominent peaks (Figure 1) which are mentioned in Table 2. The chemical molecular structures of the predominant compounds identified in the studied radish cake extract are presented in Table 3. Anthocyanidins including cyanidin (6.7%) and pelargonidin (6.9%) and isothiocyanates such as 3-(methylthio)propyl isothiocyanate (ibiverin) (4.2%), 4-pentenyl isothiocyanate (8.3%), 4-methylpentyl isothiocyanate (3.1%), 3-butenyl isothiocyanate (3.9%), 4-(methylthio)-3-butenyl isothiocyanate (5.2%), 4-(methylthio)butyl isothiocyanate (erucin) (6.4%), 5-(methylthio) pentyl isothiocyanate (berteroin) (6.9%), and L-sulforaphane (1.1%) were identified in RCE. 4-Pentenyl isothiocyanate and 5-(methylthio)pentyl isothiocyanate were found in large amounts in the radish cake extract sample. Other major compounds including phenolic derivatives such as pyrogallol, vanillic and gallic acids, and eugenol were also detected together with some fatty acids in considerable amount (8.9%).

Figure 7 shows the FT-IR spectra of the RCE. The typical v(C=O) vibration of carboxylic acids (vanillic and gallic acids) is observed as a very strong IR band at 1652 cm⁻¹. The bands at 1045 and 1299 cm⁻¹ are attributed to C–O–C vibrations of glycoside content. The bands around 2900–2950 cm⁻¹ are attributed to C–H stretching including CH, CH₂, and CH₃ bending vibrations. The infrared absorption in the spectral range between 1500 and 1300 cm⁻¹ contains vibrations of fatty acids. The peaks at 2938 and 2986 cm⁻¹ can be assigned to the N=C=S symmetric and asymmetric stretches. The framework vibration of pyridine ring and stretching mode of C=N indicates the presence of isothiocyanates. The results indicate that RCE may contain mixtures of compounds such as anthocyanidins, isothiocyanates, and phenolic derivatives. These compounds could contribute to the anticorrosion properties of the extract, and they were also noted to be present in an earlier study [25–29].

#### 3.1.1. Electrochemical Scaling Test.
The normalized chronoamperometric curves obtained for the radish cake extract in concentrations of 1 and 10 mL/L are given at Figure 2. The curve for tap water is also provided for reference. The extract of 1 mL/L concentration slows the crystallization, and at 10 mL/L, the current is reduced nearly 4 times. The radish cake extract in 1 mL/L concentration increases \( t_s \) to 3600 s, but at a higher concentration of 10 mL/L, \( t_s \) is reduced to 1000 s. Also, the shape of current-time dependence in the concentration of 10 mL/L is changed. It shows fast current

| Parameter                  | Units  | Value   |
|----------------------------|--------|---------|
| pH                         |        |         |
| Dissolved oxygen           | DO     | mg/L    | 7.8–8.0 |
| Total hardness             | TH     | mmol/L  | 6.0     |
| Total alkalinity           | TA     | mmol/L  | 3.9–4.2 |
| Calcium                    | Ca²⁺   | mmol/L  | 3.9–4.1 |
| Magnesium                  | Mg²⁺   | mmol/L  | 3.0–3.1 |
| Chloride                   | Cl⁻    | mg/L    | 0.9–1.1 |
| Sulphate                   | SO₄²⁻  | mg/L    | 18–25   |
| Hydrocarbonate             | HCO₃⁻  | mmol/L  | 30–35   |
| Total dissolved solids     | TDS    | mg/L    | 3.2–3.5 |
| Total alkalinity           | TA     | mmol/L  | 3.9–4.1 |
| Total hardness             | TH     | mmol/L  | 3.9–4.2 |
| Total alkalinity           | TA     | mmol/L  | 3.9–4.1 |
| Calcium                    | Ca²⁺   | mmol/L  | 3.0–3.1 |
| Magnesium                  | Mg²⁺   | mmol/L  | 0.9–1.1 |
| Chloride                   | Cl⁻    | mg/L    | 18–25   |
| Sulphate                   | SO₄²⁻  | mg/L    | 30–35   |
| Hydrocarbonate             | HCO₃⁻  | mmol/L  | 3.2–3.5 |
| Total dissolved solids     | TDS    | mg/L    | 240–260 |
decrease during the first 1000 s, and after stabilization, the current remains unchanged. At the concentration of 1 mL/L, the current decreases slowly during the entire test.

Values of surface porosity SP, scaling time $t_s$, and area density are collected in Table 4. In 1 mL/L concentration, scale porosity values are above 6%. At 10 mL/L concentration, porosity is nearly 20% indicating weak blocking abilities of the layer. The area density values show that the extracts at 1 mL/L concentration do not reduce the mass of the scale deposited on the surface. Moreover, the mass of the scale is even higher due to longer crystallization period. At 10 mL/L concentration, the radish cake extract

| Holding time, $t$ (min) | Name of the component | Weight ratio (%) | Molecular weight |
|-------------------------|-----------------------|------------------|------------------|
| 2.91                    | 3-(Methylthio)propyl isothiocyanate (iberverin) | 4.2              | 147.01           |
| 3.02                    | Pyrogallol acids      | 2.9              | 126.03           |
| 6.96                    | 4-Pentenyl isothiocyanate | 8.3              | 127.04           |
| 7.31                    | 4-Methylpentyl isothiocyanate | 3.1              | 143.07           |
| 10.02                   | Undecane              | 1.9              | 156.18           |
| 10.41                   | Eugenol               | 2.5              | 164.08           |
| 10.62                   | Vanillic acid         | 2.8              | 168.04           |
| 10.92                   | Dodecane              | 1.0              | 170.33           |
| 11.86                   | 3-Butenyl isothiocyanate | 3.9              | 113.18           |
| 12.42                   | 4-(Methylthio)-3-butenyl isothiocyanate | 5.2              | 159.01           |
| 12.73                   | 4-(Methylthio)butyl isothiocyanate (erucin) | 6.4              | 161.03           |
| 12.99                   | Gallic acid           | 2.7              | 170.02           |
| 13.78                   | 5-(Methylthio)pentyl isothiocyanate (berteroin) | 6.9              | 175.04           |
| 14.24                   | L-sulfuraphane        | 1.1              | 177.02           |
| 15.27                   | Dextrose              | 3.7              | 180.15           |
| 15.49                   | Tridecane             | 1.9              | 184.21           |
| 16.11                   | Dodecanoic acid (lauric acid) | 0.8              | 200.31           |
| 16.33                   | Cyanidin              | 6.7              | 287.24           |
| 18.77                   | Hexadecenoic acid (palmitic acid) | 2.2              | 256.24           |
| 19.92                   | Pelargonidin          | 6.9              | 271.24           |
| 21.00                   | Octadecanoic acid     | 5.9              | 356.31           |
| 24.11                   | Methyl linoleate      | 5.1              | 294.50           |
| 25.16                   | Sophorose             | 7.1              | 342.30           |

Figure 1: GC-MS spectra of the radish cake ethanol extract.

Table 2: Composition of the radish cake extract according to GC-MS analysis.
prevents crystallization. Practically, no changes in the electrode weight were detected.

### 3.1.2. Thermal Scaling Test

The mass of the scale on the surface of mild steel is given in Figure 3. The deposition rate in tap water is 7.5 µg/(h·cm²). It remains constant during the first 130 h and then slightly decreases. In contrast to tap water, when the extract is added, the scale deposition is reduced to 1.6 µg/(h·cm²) meaning antiscaling efficiency is 78.7%. So, the RCE slows down deposition of calcium carbonate even at the temperature of 100°C.

Possible mechanism of scale inhibition could be the formation of soluble complexes with calcium ions. Such complexes prevent the deposition of calcium carbonate keeping calcium ions in the soluble form. According to GC-MS, phenolic derivatives such as pyrogallol, vanillic and gallic acids, eugenol, and polysaccharides are found in the extract [30–32]. The mechanisms for scale inhibition by the radish cake extract are associated with active functional groups (C=O, −OH, −COOH, and N=C=S groups) of the main compounds of the extract. These substances, which have hydroxyl and/or carboxyl functional groups that interact with divalent ions such as Ca²⁺, are known to form water-soluble complexes with calcium ions. In addition to this, carboxyl-terminated anthocyanidins have a larger surface area, which can chelate with more Ca²⁺. Therefore, the radish cake extract could prevent the formation of calcium carbonate via formation of water-soluble complexes.

Scale inhibition was determined from the change of hardness salt concentration in one-day run in tap water.
solution and with different RCE concentrations (Figure 4). It was found that the antiscaling efficiency according to the changes in hardness is only 52%. In contrast, the mass of the deposited scale is nearly 5 times lower when the extract is added to the solution. This means that soluble complex formation only could not explain antiscaling efficiency of the extract.

3.1.3. Corrosion Inhibition. The instantaneous corrosion rate of mild steel determined with the LPR technique in thermal scaling conditions is given in Figure 5. In tap water, the corrosion rate of mild steel right after immersion is around 0.8 mm/year and decreases with the boiling time due to the formation of the scale layer that acts as a diffusion barrier and reduces oxygen supply to the surface. In tap water with RCE, the corrosion rate decreases in the first hour and reaches the value of 0.2 mm/year meaning corrosion inhibition efficiency is 75%. So, RCE provides efficient corrosion protection on the steel surface. As scale deposition progresses, corrosion rate in both tap water and inhibited solution decreases; however, in tap water, the decrease is faster because more scale is deposited. The thicker the scale layer, the less porous it becomes; thus, the corrosion rate is lower. After 50 h of boiling, the corrosion rate in both solutions coincides at 0.2 mm/year and continues to decrease till the corrosion rate of 0.1 mm/year is reached.

3.1.4. Surface Film Formation. The analyses of chronoamperometric curves (Figure 1) suggest that in the 10 mL/L concentration, when no scale is deposited, the current is reduced due to the formation of the surface film. This film consists of extract components and acts as a barrier protecting the steel surface from the supply of both calcium carbonate and dissolved oxygen.

The SEM images of the scaled steel surface are given in Figure 6. The scale deposited from tap water has a regular structure. The crystals are 60–80 μm, and the layer is uniform. In the presence of RCE, the crystals are much smaller and the presence of the organic film is clearly observed on the surface. Figure 6(c) demonstrates the charging effect of SEM when the electrons are accumulated in the organic film. SEM analysis clearly demonstrates the formation of the organic surface film that acts as a barrier level to reduce the supply of oxygen and calcium carbonate, thus reducing scaling and corrosion.

To identify the nature of the film, the scale after deposition was analysed with FT-IR (Figure 7). Comparison of the FT-IR spectra of the radish cake extract, the scale film, and scale film after exposure in inhibited solution in the presence of RCE showed that the spectra were changed to a different extent (Table 5). As shown by curve (1), the peaks given at 873 and 712 cm\(^{-1}\) could be attributed to the vibrations of calcite of CaCO\(_3\) crystal polymorphs, the peak at 1402 cm\(^{-1}\) reflect vaterite of CaCO\(_3\) crystal polymorphs. The absorption peak at 712 cm\(^{-1}\), 870 cm\(^{-1}\), and 1402 cm\(^{-1}\) decreases in curve (2), indicating the antiscalant efficiency of the extract.

The extended peak in the range around 1200–1400 cm\(^{-1}\) indicates that phenolic compounds and isothiocyanates were adsorbed on the steel surface. This indicates that probably these compounds may be responsible for scale inhibition. The bands around 2200–2400 cm\(^{-1}\) are attributed to polymeric bond types ((C=C) stretching vibration) [25, 26, 29]. Isothiocyanates are known to be heterocumulenes with an electrophilic centre on the carbon atom. Therefore, nucleophilic addition reactions are typical for them. For many heterocumulenes, the reactions of \([2+2]\),
Table 5: Peaks from FT-IR spectra samples: RCE (a), the scale film (b), and scale film after exposure in the presence of RCE (c).

| Wavenumber (cm⁻¹) | Scale film in RCE | Scale film | Possible groups       |
|-------------------|------------------|------------|-----------------------|
| 3291              | —                | —          | O-H/N-H               |
| 2981, 2902        | —                | —          | Aliphatic, aromatic C-H or N=C=S |
| 1456, 1417, 1639  | —                | —          | C=O or C=N            |
| 1084, 1384, 1326, 1273 | —        | —          | C-O-C                 |
| —                 | 2336             | —          | C=C                   |
| —                 | 716              | 713        | Calcite               |
| —                 | 873              | 870        | Vaterite              |
| —                 | 1402             | 1400       |                       |

Figure 6: SEM images of the scaled surface at different magnification: (a) and (b), tap water; (c) and (d), tap water with 10 mL/L of the radish cake extract.

Figure 7: FT-IR spectra of crystalline calcium carbonate grown in the absence of RCE (1), presence of RCE (2), and radish cake extract (3).
cycloaddition with the formation of four-membered cycles are common. As a result, isocyanates dimerize to form uretdiones. It is possible that the formation of a partially polymerized type of film causes a high antiscalant action.

The mechanisms for scale inhibition of the radish cake extract have been associated with both formation of the protective film on the surface of steel by compounds such as isothiocyanates and saturated fatty acids and formation of water-soluble complexes of the phenolic derivatives (pyrogallol, vanillic and gallic acids, eugenol, and polysaccharides) with divalent ions Ca²⁺. The investigation of the scale and corrosion inhibition mechanism of the radish cake extract has indicated that because of multi-component composition, it provides complexation ability with calcium ions and absorption ability on the steel surface that prevent the formation of calcium scale crystals and assure the corrosion inhibition effectively [25, 26, 29].

4. Conclusions

The ethanol extract of radish cake was tested as a scale and corrosion inhibitor of mild steel in tap water. The following conclusions can be formulated:

(1) According to GC-MS and FT-IR analyses, the main components of the extract are as follows: isothiocyanates (39.1%), phenolic derivatives (10.9%), anthocyanidins (13.6%), and saturated fatty acids (8.9%).

(2) The thermal scaling test revealed that the radish cake extract in a concentration of 10 mL/L can suppress scaling 4.7 times. At the same time, the corrosion inhibition efficiency is 75%. The scaling and corrosion are both suppressed due to the formation of the film on the surface of steel. This film acts as a barrier layer preventing oxygen and calcium carbonate ingress to the surface.

(3) The presence of the film was confirmed with SEM and FT-IR analyses. The film is mainly composed of phenolic derivatives and isothiocyanates formed by extract components and their products of chemical transformation that partially polymerized.

Data Availability

The GS-MS, LPR, and chronoamperometric data used to support the findings of this study are included within the article.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

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