**Multiple Molecular Mechanisms Rescue mtDNA Disease in C. elegans**

**Graphical Abstract**

- A genetically engineered worm recapitulates the hallmarks of mtDNA disease in humans
- This worm can be exploited to generate countless additional models of mtDNA disease
- An RNAi screen identifies 25 genes that can prevent or delay mtDNA disease in worms
- IGF-1/insulin signaling, mitophagy, and UPR\textsuperscript{mt} strongly affect disease progression

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**In Brief**

Haroon et al. describe a genetically engineered *C. elegans* that carries an error-prone copy of DNA polymerase γ, the enzyme that replicates the mitochondrial genome. This worm recapitulates the major hallmarks of mitochondrial disease in humans. The authors identify multiple biological pathways that could potentially delay disease progression.
Multiple Molecular Mechanisms Rescue mtDNA Disease in C. elegans

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SUMMARY

Genetic instability of the mitochondrial genome (mtDNA) plays an important role in human aging and disease. Thus far, it has proven difficult to develop successful treatment strategies for diseases that are caused by mtDNA instability. To address this issue, we developed a model of mtDNA disease in the nematode C. elegans, an animal model that can rapidly be screened for genes and biological pathways that reduce mitochondrial pathology. These worms recapitulate all the major hallmarks of mtDNA disease in humans, including increased mtDNA instability, loss of respiration, reduced neuromuscular function, and a shortened lifespan. We found that these phenotypes could be rescued by intervening in numerous biological pathways, including IGF-1/insulin signaling, mitophagy, and the mitochondrial unfolded protein response, suggesting that it may be possible to ameliorate mtDNA disease through multiple molecular mechanisms.

INTRODUCTION

Energy animates life. Whether it’s protein synthesis, autophagy, or signal transduction, almost every biological process is driven by the consumption of energy. Most of this energy is generated by mitochondria, small tubular organelles that are frequently called the powerhouses of our cells. To optimize energy production, mitochondria carry their own genome called “mtDNA,” a small, circular molecule that encodes numerous protein products that are essential to ATP synthesis (Anderson et al., 1981). As a result, loss or mutation of mtDNA invariably affects energy production, which is particularly harmful to cells with high energy demands such as neurons and muscle fibers (Wallace, 2005). Most diseases caused by mtDNA instability are therefore characterized by some form of neuromuscular dysfunction. For example, point mutations, deletions, or overt loss of mtDNA molecules can result in premature deafness, myopathy, or severe encephalomyopathy in children (Saneto and Sedensky, 2013) and contributes to neurodegeneration (Kraytsberg et al., 2006) and muscle wasting (Wanagat et al., 2001) in aging adults. In most cases, mtDNA instability arises spontaneously, although it also can be caused by mutations in DNA polymerase γ, the enzyme that replicates the mitochondrial genome. Thus, the impact of mtDNA instability on human aging and disease is well documented. To this day though, no child has ever been cured of an mtDNA disease, nor does a treatment exist for the mtDNA component of age-related diseases. Although it is now possible to manage the symptoms of a number of pediatric mtDNA diseases, two bottlenecks have traditionally kept the scientific community from developing a more robust treatment strategy: a shortage of animal models and a lack of promising molecules to target. Here, we address these issues by establishing a model of mtDNA disease in worms that can give rise to countless additional models with the proper breeding scheme. Moreover, because worms can be easily manipulated by genetic and pharmaceutical means, we were able to use RNAi, drugs, and genetic mutants to identify multiple cellular pathways that can ameliorate the pathological consequences of mtDNA instability on organismal health, thus providing potential targets for therapeutic intervention. Together, these results suggest that multiple pathways may have evolved to prevent mtDNA disease, underlining the central role that energy plays in cellular life.

RESULTS

A Model of mtDNA Disease in C. elegans

To successfully design a therapeutic strategy for mtDNA disease, it will be essential to identify cellular mechanisms that can either increase or decrease the pathology that is caused by mtDNA instability. These experiments will reveal promising targets that can be exploited for therapeutic intervention, and may provide valuable insight into the basic biology that underlies mitochondrial function. Identifying these mechanisms requires a flexible animal model that is well suited for “discovery experiments.” The most promising model available today is the mitochondrial mutator mouse (Kujoth et al., 2005; Trifunovic et al., 2004). This mouse model carries an error-prone copy of DNA polymerase γ (PolγA Polyc, ), the enzyme that replicates the mitochondrial genome. Because of error-prone DNA replication, the PolγA Polyc mice exhibit a >100-fold increase in mtDNA instability, which is associated with numerous age-related diseases, including diabetes, atherosclerosis, cardiovascular disease, and cancer. This mouse model has proven invaluable in identifying potential therapeutic targets and has become the gold standard for mtDNA research. However, this model is far from perfect. For example, it does not recapitulate the wide range of mtDNA diseases observed in humans, nor does it have the ability to screen for potential targets. To address these issues, we developed a model of mtDNA disease in worms that can give rise to countless additional models with the proper breeding scheme. Moreover, because worms can be easily manipulated by genetic and pharmaceutical means, we were able to use RNAi, drugs, and genetic mutants to identify multiple cellular pathways that can ameliorate the pathological consequences of mtDNA instability on organismal health, thus providing potential targets for therapeutic intervention. Together, these results suggest that multiple pathways may have evolved to prevent mtDNA disease, underlining the central role that energy plays in cellular life.
Figure 1. Characterization of the polg-1(srh1) Worms

(A) The exonuclease (green) domain of DNA polymerase γ contains three highly conserved regions (red) that control the fidelity of DNA synthesis, including an aspartic acid in exonuclease domain II that is essential for the proofreading activity of polymerase γ across the tree of life. We used CRISPR/Cas9 technology to mutate this residue to alanine in the error-prone allele polg-1(srh1).

(B) The mtDNA mutation frequency of the polg-1(srh1) worms is >70-fold higher compared to WT worms.

(C) The mutation spectrum of WT and polg-1(srh1) depicted as peaks above and below the WT nucleotide according to standard electrophoretogram color coding (red, T; blue, C; black, G; green, A). The percentage of each type of mutation is listed by the peaks.

(D) The mtDNA copy number is reduced by 56% in the polg-1(srh1) worms.

(E) The basal respiration of polg-1(srh1) worms worsens with age compared to WT worms.

(F) The mitochondria of polg-1(srh1) worms display reduced reserve capacity upon FCCP treatment at 5 days of age.

(G and H) Confocal images of day 10 polg-1(srh1) and WT worms (G) do not reveal a significant difference in oxidation stress (H).

(legend continued on next page)
mutations, which results in extensive mtDNA disease (Kujoth et al., 2005; Trifunovic et al., 2004; Vermulst et al., 2007, 2008). However, logistical considerations prohibit these mice from being screened en masse to identify genes or molecular pathways that modulate their pathology. Therefore, we recreated the PolgAD257A mutation in C. elegans, a genetically tractable model organism that is amenable to high-throughput screens. Using CRISPR/Cas9 technology, we directly edited the polg-1 gene (the C. elegans homolog of PolgA) and introduced the analogous D207A mutation [polg-1(srh1)] (Figure 1A). Similar to the mutator mice, we found that the polg-1(srh1) worms display a large increase in mtDNA mutations (Vermulst et al., 2007, 2008) (Figure 1B), which was primarily driven by C:G > T:A transitions (Figure 1C). These transitions were not equally distributed over the TaqI restriction site, despite the fact that it is a palindrome, suggesting that polg-1-mediated mutagenesis has a substantial strand bias (Figure 1C). Moreover, homozygous carriers of the polg-1(srh1) allele displayed a ~50% loss in mtDNA copy number (Figure 1D), mirroring the mtDNA depletion of homozygous mutator mice. Thus, even though mtDNA is replicated differently in worms compared to mice (Lewis et al., 2015), the polg-1(srh1) worms closely mimic the genetic instability of the mutator mice and display an increased rate of mtDNA mutation and depletion, two forms of mtDNA instability that cause mtDNA disease in humans (Copeland, 2008; El-Hattab and Scaglia, 2013).

Mutator Worms Replicate Major Hallmarks of mtDNA Disease in Humans

In human patients, mtDNA instability invariably results in mitochondrial dysfunction. To determine whether the mutator worms display mitochondrial dysfunction as well, we analyzed intact mitochondrial dysfunction. To determine whether the mutator worms respire at the same rate as wild-type (WT) worms by respirometry. We found that at day 1 of adulthood, the polg-1(srh1) worms display increased rate of mtDNA mutation and depletion, worms closely mimic the genetic instability of the mutator mice and display an increased rate of mtDNA mutation and depletion, two forms of mtDNA instability that cause mtDNA disease in humans (Copeland, 2008; El-Hattab and Scaglia, 2013).

Monitor Load and Tissue Dysfunction of polg-1(srh1) Worms Worsen over Generations

The number of mutated mtDNA molecules that are transmitted from mother to child controls the severity and the type of pathology that emerges in children (Wallace, 2010). The higher this number is, the more complex the disease tends to be; however, (I–K) Neuromuscular function assessed by chemotaxis (I), thrashing (J), and a gentle touch assay (K) reveals increased dysfunction in polg-1(srh1) worms compared to WT animals.

(L) The polg-1(srh1) worms have a median lifespan of 13 days compared to 16 days for WT worms (log-rank test p < 0.01).

Data for WT and polg-1(srh1) worms are in blue and pink, respectively (in B, D–F, and H–J).

Bar graphs represent the mean ± SEM of at least three biological replicates, and the lifespan assay was performed using at least 100 worms per genotype. Unpaired t tests were performed to determine significance (‘p < 0.05, ”p < 0.01; ns, no significant difference).

Also see Figure S1 and Movies S1 and S2.
it has proven difficult to study germline transmission of mtDNA mutations in detail, because it is not possible to transform the mitochondrial genome at will. As a result, researchers are forced to rely on natural mtDNA variants to decipher the parameters that regulate mutation inheritance. We reasoned that the increased mutation rate of the polg-1(srh1) allele could greatly accelerate the process of isolating organisms with specific mtDNA mutations of interest, so that novel models of inherited mtDNA mutations can rapidly be generated. To test this hypothesis, we tracked the offspring of individual WT and polg-1(srh1) worms over 35 generations and sequenced the mtDNA of 3 siblings at generations 5, 15, 25, and 35. Over these generations, the mutator worms accrued an increasing number of detectable heteroplasmic mutations (Figures 2A, 2B, and S2), while the mtDNA of the WT worms remained pristine. Consistent with the idea that these mutations contribute to the dysfunction of the polg-1(srh1) worms, we found that the performance of the polg-1(srh1) worms in the chemotaxis (Figure 2C), gentle touch (Figure 2D) and thrashing (Figure 2E) assay decreased with increasing generation number. To prevent these generational effects from confounding our results, all studies were performed on generation-matched worms (generations 27–33).

| Generation | Mutation | Heteroplasm | Gene | Change |
|------------|----------|-------------|------|--------|
| 5          | G1006DA  | 30%         | COII | D to N |
| 25         | C2582T   | 50%         | ND1  | G to A |
| 35         | T2085    | 100%        | ND1  | early stop |

Figure 2. The Phenotype of the polg-1(srh1) Worms Worsens from Generation to Generation

(A) The outer circle represents the worm mitochondrial genome highlighting the protein-coding genes (green), rDNA (blue), AT-rich region (yellow), and all the mutations (red) that arose over 35 generations of maintaining the polg-1(srh1) mutator allele. The inner circle represents the regions that were sequenced (blue).

(B) mtDNA mutations were tracked over 35 generations in WT and polg-1(srh1) worms. At generations 5, 15, 25, and 35, ~11 kb of mtDNA of 3 progenies of a single WT and polg-1(srh1) worm were sequenced. The mutations depicted here were present in all 3 progenies, indicating that they were successfully transmitted through the germline. Three mutations, here denoted in purple, brown, and orange polg-1(srh1) were present across multiple generations and showed substantial genetic drift. No new mutations arose in the WT strain.

(C–E) Neuromuscular dysfunction was assessed by chemotaxis (C), thrashing (D), and gentle touch (E) revealed progressive dysfunction with increasing generation in the polg-1(srh1) worms.

(F) Representative electrophoretograms tracking the T2085 deletion that results in an early stop codon in the ND1 gene in both the parent with polg-1(srh1) allele and its resultant progeny where the mutator allele has been mated out. Three individuals without the polg-1(srh1) allele in the F1 generation, 10 individuals in the F2 generations, and 21 individuals in the F3 generation were isolated and tested for the transmission of T2085 heteroplasmy in a background WT for polg-1.

Bar graphs represent the mean ± SEM of at least three biological replicates. Unpaired t tests were performed to determine significance (*p < 0.05, **p < 0.01; ns, no significant difference).
However, in some cases, the rationale of our experiments required a change in generation number. If so, this change has been clearly denoted. We further found that these germline mutations both increased and decreased over successive generations, indicating substantial genetic drift. To formally demonstrate that these mutations can be isolated to study their inheritance, we decided to remove the error-prone \( \text{polg-1(srh)} \) allele from the nuclear genome. We reasoned that this strategy would prevent additional mutations from accumulating in the mitochondrial genome, and allow desirable mtDNA mutations to be fixed in a WT nuclear background. To this end we crossed the mutagenic \( \text{polg-1(srh)} \) allele out of a worm that carried a single base-pair deletion in the ND1 gene (Figures 2A, 2B, and S2E). After the removal of the \( \text{polg-1(srh)} \) allele, we selectively picked individual progeny from parents with \( \% \) heteroplasmy over several generations (Figure 2F), and were able to successfully retain this mtDNA mutation. Interestingly, we found that, three generations after breeding out the mutator allele, 33% of the worms laid inviable embryos, 33% of the worms laid progeny that became larval phase 2 dauers, and 33% of the worms laid normal embryos that developed fully. This experiment demonstrates that it is possible to exploit the \( \text{polg-1(srh)} \) worms to generate novel animal models that display gross phenotypic abnormalities, and carry highly desirable mtDNA mutations in their germline, which could fill an important void in the mitochondrial research community.

Multiple Pathways Modulate mtDNA Disease in Worms

The chemotaxis assay described above is highly quantitative and reproducible, and tests the most important clinical aspects of mtDNA disease. Therefore, we used this assay as a screening tool to identify genes that can ameliorate the pathology associated with mtDNA disease (Figure 3). To this end, we performed a candidate RNAi screening using >130 RNAi constructs that control either C. elegans lifespan, overall health, or various aspects of mitochondrial function, parameters that are likely to intersect with mtDNA disease. We considered a gene a positive hit if RNAi against that gene improved the performance of the mutator worms by >30%. This cutoff is based on the observation that the inherent variation between randomly selected \( \text{polg-1(srh)} \) worms rarely exceeded this threshold (0 out of 10 trials), indicating a <10% chance of discovering a false positive (Figure S3A). Second, RNAi against \( \text{age-1} \), which we found to be a potent modulator of mtDNA disease (35% on average) (Figures 3A–3C), improved the performance of the mutator worms by >30% in 6 out of 9 trials (Figure S3B), indicating a >60% chance of detecting a gene that increases the performance of the \( \text{polg-1(srh)} \) worms by 35% or more. Excitingly, RNAi against numerous genes decreased the pathology associated with mtDNA disease in worms. For example, manipulation of the insulin growth factor (IGF)-1/insulin signaling (IIS) pathway, mitophagy, autophagy, apoptosis, and the mitochondrial unfolded protein response (UPR\(_{\text{mt}}\)) all ameliorated the neuro-muscular defect of the \( \text{polg-1(srh)} \) worms (Figure 3D). Interestingly, reduced autophagy was previously shown to ameliorate
mitochondrial dysfunction in a model of mitochondrial disease that is caused by a nuclear mutation (Peng et al., 2015), while suppression of apoptosis activates the highly beneficial stress response in worms (Judy et al., 2013). Similarly, mitophagy and UPR\textsuperscript{mt} are implicated in various diseases that contain a response in worms (Judy et al., 2013). Similarly, mitophagy suppression of apoptosis activates the highly beneficial stress (Kenyon, 2010). Consistent with this idea, we found that it initiates a transcriptional program that underlie our observations, we picked three pathways that emerged from our screen and used drugs and genetic mutations to knockout or activate the molecular pathways they control.

The IGF-1/Insulin signaling pathway emerged as the strongest modulator of mtDNA disease, as RNAi against \textit{age-1}, \textit{par-5}, and \textit{akt-1} all rescued the mobility defect of the \textit{polg-1(srh1)} worms. In contrast, RNAi against \textit{daf-18}, which directly opposes \textit{AGE-1} (Murphy and Hu, 2013), exacerbated the phenotype of the \textit{polg-1(srh1)} worms (Table S1). At a mechanistic level, \textit{age-1}, \textit{par-5}, and \textit{akt-1} share a common goal as well: they all increase the phosphorylation of the transcription factor DAF-16, trapping it in the cytoplasm and suppressing its activity (Murphy and Hu, 2013). RNAi against \textit{age-1}, \textit{par-5}, and \textit{akt-1} releases this suppression and allows DAF-16 to translocate to the nucleus where it initiates a transcriptional program that promotes organismal health (Kenyon, 2010). Consistent with this idea, we found that RNAi against \textit{daf-16} greatly exacerbated the neuromuscular defect of the \textit{polg-1(srh1)} worms (Table S1). To verify our findings further, we introduced a hypomorph of the IGF-1/insulin receptor (\textit{daf-2(e1370)}) into the \textit{polg-1(srh1)} worms. This allele displays reduced IIS activity (Kenyon et al., 1993) and consistent with our RNAi results, we found that it rescued the neuromuscular defect of the mutator worms (Figure 4A). Conversely, a partial deletion of \textit{daf-16} (\textit{daf-16(mu86)}) exacerbated their phenotype (Figure 4B). Thus, these genetic mutations recapitulated the results of our RNAi screen. We further found that the \textit{daf-2(e1370)} allele also improved the basal respiration rate of the mutator worms (Figure 4C), indicating that reduced IIS activity improves the mobility of the worms by correcting their underlying mitochondrial dysfunction. Interestingly, the \textit{daf-2(e1370)} allele also improved mtDNA copy number in the mutator worms (Figure 4D), suggesting that it can partially rescue the etiology of the disease itself, although the mutation rate of the \textit{polg-1(srh1)} worms was unaffected (Figure 4E).

Our screen further indicated that knockdown of \textit{dct-1} and \textit{pdr-1} could rescue the mobility defect of the mutator worms. Because these genes play a pivotal role in mitophagy (Paliaras et al., 2015), these results suggest that reduced mitophagy may be beneficial for the \textit{polg-1(srh1)} worms. To verify these observations with a genetic mutant, we introduced the \textit{pdr-1(gk448)} allele into the mutator worms. \textit{Pdr-1} encodes the \textit{C. elegans} homolog of the parkin ubiquitin ligase PARK2, and the \textit{pdr-1(gk448)} allele carries a partial deletion that greatly reduces mitophagy in worms (Springer et al., 2005). Consistent with our RNAi screen, we found that the \textit{pdr-1(gk448)} allele rescued the neuromuscular defect of the \textit{polg-1(srh1)} worms (Figure 4F). Conversely, exposure of the mutator worms to urolithin A, a drug that induces mitophagy (Ryu et al., 2016), worsened their phenotype (Figure 4G).

Finally, we found that the \textit{pdr-1(gk448)} allele rescued the basal respiration rate of the mutator worms (Figure 4H), similar to the \textit{daf-2(e1370)} allele. In contrast to the \textit{daf-2(e1370)} allele though, we found that the \textit{pdr-1(gk448)} allele did not alter mtDNA copy number (Figure 4I) and paradoxically increased the mitochondrial mutation frequency by 5.5-fold (Figure 4J).

Finally, our screen indicated that RNAi against the mitochondrial protease \textit{spg-7} and several components of the electron transport chain (\textit{atp-3, nuo-2, D2030.4, and T02H6.11}) rescued the neuromuscular defect of the mutator worms. Knockdown of \textit{spg-7} or certain components of the electron transport chain are known to induce the unfolded protein response in mitochondria (Jovaisaite et al., 2014), indicating a possible involvement of this adaptive stress response in the rescue mechanism. Indeed, suppression of UPR\textsuperscript{mt} by RNAi against \textit{haf-1}, \textit{ats-1}, and \textit{ubi-5}, exacerbated the phenotype of the mutator worms (Table S1).

To further verify the protective role of UPR\textsuperscript{mt} in mtDNA disease progression, we introduced the deletion allele \textit{ats-1(tm4919)} (Pellegrino and Haynes, 2015) into the \textit{polg-1(srh1)} background. \textit{Ats-1} encodes a transcription factor required for UPR\textsuperscript{mt} activation. Interestingly, we found that homozygous carriers of the \textit{ats-1(tm4919)} allele rendered heterozygous \textit{polg-1(srh1)} worms infertile, indicating that loss of UPR\textsuperscript{mt} greatly exacerbates the phenotype of the \textit{polg-1(srh1)} worms. Accordingly, we were unable to generate homozygous \textit{polg-1(srh1)}; \textit{ats-1(tm4919)} worms for experiments. To answer the question of whether activation of this adaptive response is able to rescue mtDNA disease, we used a constitutively active \textit{ats-1} allele (\textit{ats-1(et15)}) and found that it indeed partially rescued the neuromuscular defect of the mutator worms (Figure 4K). In addition, this allele rescued the decreased reserve capacity of mitochondria (Figure 4L) and slightly increased mtDNA copy number (Figure 4M), although it did not alter the mtDNA mutation frequency (Figure 4N).

**DISCUSSION**

MtDNA instability is associated with a remarkable number of human diseases. Although it is now possible to manage, and even improve some of the symptoms of mtDNA disease, there is currently no cure for either inherited mtDNA diseases or the mtDNA component of age-related diseases. To address this problem, we generated a model of mtDNA disease in \textit{C. elegans}, which displays two forms of mtDNA instability known to cause mtDNA disease in humans: mtDNA depletion and mtDNA mutation. Both of these phenotypes are driven by a mutant DNA polymerase \textit{y}, which is a major source of mtDNA instability in patients as well. As a result, the \textit{polg-1(srh1)} worms do not model any specific mtDNA disease, but serve as a general model of mtDNA disease caused by mtDNA instability. The \textit{polg-1(srh1)} worms are especially useful because they can rapidly be screened for large numbers of genes, drugs, and small molecules that modulate mtDNA disease, which allows them to serve as a motor for discovery. Moreover, any findings can
Figure 4. Verification of the RNAi Screen with Genetic Mutants and Small Molecules

(A) The daf-2(e1370) allele rescues the neuromuscular defect of 5-day-old polg-1(srh1) worms.
(B) The daf-16(mu86) allele worsens the neuromuscular defect of 5-day-old polg-1(srh1) worms. This set of experiments was performed at generation 7 instead of generation 30, when the phenotype of the mutator worms is not significantly different from the WT worms until 7 days of age. Accordingly, the detrimental effect of the daf-16(mu86) allele is better illustrated.
(C) The daf-2(e1370) allele rescues the basal respiration rate of 10-day-old polg-1(srh1) worms.
(D and E) The daf-2(e1370) allele (D) increases mtDNA copy number by 118%, but (E) has no effect on the mutation frequency of polg-1(srh1) worms.
(F) The pdr-1(gk448) allele rescues the chemotaxis defect of 5-day-old polg-1(srh1) worms.
(G) Induction of mitopagy with 50 μM urolithin A (UA) worsens the chemotaxis phenotype of the 5-day-old polg-1(srh1) worms.
(H) The pdr-1(gk448) allele rescues the basal respiration rate of 10-day-old polg-1(srh1) worms.
(I and J) The pdr-1(gk448) allele (I) increases mtDNA copy number by 25% and (J) results in a 5.5-fold increase in the mutation frequency of polg-1(srh1) worms.
(K and L) The atfs-1(et15) allele (K) rescues the chemotaxis defect of 5-day-old polg-1(srh1) worms and (L) the reserve capacity of 5-day-old polg-1(srh1) worms.
(M and N) The atfs-1(et15) allele (M) increases mtDNA copy number by 63%, but (N) has no effect on the mutation frequency of polg-1(srh1) worms.

Bar graphs represent the mean ± SEM of at least three biological replicates. Unpaired t tests were performed to determine significance (*p < 0.05, **p < 0.01; ns, no significant difference).
immediately be tested in the mitochondrial mutator mouse to determine whether they are translatable to mammalian biology. Another useful feature of the mutator worms is that they rapidly accumulate mtDNA mutations in their germline, which allows for countless additional models of mtDNA disease to be generated. For example, after crossing the error-prone polg-1(srh1) allele out of the nuclear genome to prevent further mutation accumulation, worms with desirable germline mutations could be used to determine which parameters control the inheritance of mutated molecules. A similar approach was recently proposed for the mitochondrial mutator mice (Kauppila et al., 2016). Because mitochondrial research is greatly handicapped by a lack of inherited mutant mtDNA models, these worms would complement the mtDNA models that can be generated by restriction site targeting (Xu et al., 2008) to fill an important void in the research community.

A second problem that prevents the research community from developing a treatment for mtDNA disease is the lack of promising molecules to target. Here, we used the polg-1(srh1) worms to show that numerous molecular pathways control the severity of mtDNA disease. The most promising candidate we have identified thus far is the IGF-1/insulin signaling pathway. It has long been known that reduced IGF-1/insulin signaling has beneficial effects on the overall health of organisms (Kenyon, 2010); however, our results now suggest that for a discrete set of diseases, reduced IIS activity may also have a direct therapeutic application. Since reduced IIS activity increased mtDNA copy number, this approach may be especially useful for diseases caused by mtDNA depletion. Second, it is well known that reduced IGF-1 signaling is particularly beneficial to aging organisms (Kenyon, 2010). Since mtDNA mutations and mitochondrial dysfunction are associated with aging, these findings also provide insight into the molecular mechanisms by which reduced IGF-1 signaling prevents age-related dysfunction. We further found that reduced mitophagy could rescue the polg-1(srh1) worms from mtDNA disease, while increased mitophagy exacerbated their phenotype. This counter-intuitive observation suggests that while increased mitophagy is beneficial under normal circumstances, it can result in an adverse outcome in the context of disease. One potential explanation for this paradox could be that in the mutator worms recycled mitochondria are replaced with equally dysfunctional organelles, leading to futile cycles of mitophagy and mitochondrial biogenesis that results in further energy depletion. Interestingly, loss of PARK2 has both beneficial (reduced splenomegaly) and detrimental (substantia nigra) effects on the mutator mice (Pickrell and Youle, 2015), suggesting that the effect of reduced mitophagy on mammalian biology is highly cell-type and tissue specific. Another surprising observation was that reduced mitophagy increased the mutation burden of the mutator worms. These results suggest that mitophagy can potentially cull mutated mtDNA molecules from cells, which is consistent with the idea that mitophagy recycles dysfunctional organelles. Surprisingly though, knocking out PARK2 in the mitochondrial mutator mice did not seem to result in an increased mutation frequency. The reason for this discrepancy is currently unclear; however, an important difference between these organisms is that the mutator mice carry 10-fold more mutations than the mutator worms, which may limit the ability of mitophagy to significantly impact the mutation burden.

Finally, we found that activation of UPR mt could rescue the neuromuscular defect of the polg-1(srh1) worms. Interestingly, it was previously shown that a reduction in mtDNA copy number causes an imbalance in the number of proteins derived from the nuclear and mitochondrial genome. This imbalance causes proteotoxic stress inside mitochondria by preventing proteins from finding their natural binding partner, and UPR mt ameliorates this stress. Since the polg-1(srh1) worms suffer from mtDNA depletion, we hypothesize that a reduction in proteotoxic stress partially underlies the improved performance of the polg-1(srh1); atfs-1(et15) worms.

Taken together, these observations demonstrate that the polg-1(srh1) worms are a useful model to identify modulators of mtDNA disease. We further note that all of the modifiers we analyzed here seem to ameliorate mtDNA disease by improving mitochondrial function, suggesting that they could be beneficial for a broad range of mtDNA diseases. Moreover, since the genetic instability of the mutator worms is substantial, these modifiers are likely to be fairly powerful. It is further interesting to note that despite the fact that few targets are available today, a substantial number of genes can modulate the pathological consequences of mtDNA instability, suggesting that numerous targets may exist for therapeutic treatment of mtDNA disease. One potential conclusion from these observations is that precisely because energy is required for every biological process in our cells, multiple pathways may have evolved to manage or prevent mtDNA disease. Using the polg-1(srh1) worms, the mutator mice, and numerous other models that are still on the horizon, it now finally may be possible to identify these pathways in a reasonable period of time, which holds enormous promise for our ability to understand the molecular basis of mtDNA disease and to develop comprehensive treatment plans for patients.

**EXPERIMENTAL PROCEDURES**

**Strains and Growth Conditions**

The polg-1(srh1) worms were created using CRISPR/Cas9 technology in accordance with published protocols (Wadhams et al., 2017) and using the guide RNA sequence UGAUCGAGCCAGAUGUCGGG and the donor DNA sequence atctggagattggaatggagaataagattatgacataattgtggtgctcccGgTtgCcgTgaggcttatcaatcgataaatgggtcaaagattcgatttatggatacaatgtct. The guide RNA sequence UGAUCGAGCCAGAUGUCGGG and the donor DNA sequence atctggagattggaatggagaataagattatgacataattgtggtgctcccGgTtgCcgTgaggcttatcaatcgataaatgggtcaaagattcgatttatggatacaatgtct. Capitalized bases differ from the WT polg-1 sequence. These changes were required for changing the aspartic acid at residue 207 to alanine, as well as inhibiting CRISPR/Cas9 cleavage after integration of the donor DNA. Additionally, these changes introduced a Smal restriction site for genotyping purposes. Like in many other models of mitochondrial dysfunction, we found that the polg-1(srh1) worms displayed pronounced fertility problems when in a homozygous state. Therefore, we maintained the polg-1(srh1) allele as heterozygote animals with the balancer chromosome mnc1 that carries a pharyngeal fluorescent GFP reporter (CCG strain MT20110). Only the non-fluorescent homoygous worms were used for experiments. The daf-2(e1370), daf-16(mu86), pyd-1(kg448), and atfs-1(tm4919) alleles were obtained from CGC, which is funded by NIH Office of Research Infrastructure Programs (P40 OD010440). The atfs-1(et15) strain was created by Dr. Marc Pilon (University of Gothenburg, Sweden) and was obtained from Dr. Cole Haynes. All strains were backcrossed into the laboratory N2 WT strain at least four times before comparative experiments were carried out. Strains were maintained without starvation at 20°C for at least two generations prior to experimental use. The L4 stage of worms was counted as day 0 of adulthood for all experiments.
mtDNA Copy Number

mtDNA copy number was determined by qPCR using at least three replicates per genotype by adapting a published protocol (Polyak et al., 2012). For each replicate, 10 L4 worms were collected in 10 μL of a buffer containing 10 mM Tris-HCl, 50 mM KCl, 1.5 mM MgCl₂, 0.001% gelatin, and 5 mg/mL Proteinase K. The worms were then incubated at 65°C for 75 min, followed by a 20-min incubation at 95°C, and diluted with 10 μL of H₂O. Subsequently, 5.5 μL of each sample was used to run a TaqMan assay with a Universal Master Mix (cat. no. 444040, Thermo Fisher Scientific), where ND4 (cat. no. 4440043, assay ID: AIFAT8G) and ACT-4 (assay ID: Ce02508047_s1, Thermo Fisher Scientific) assays quantified mtDNA and nDNA content, respectively. mtDNA copy numbers were then normalized to nDNA content.

Oxygen Consumption

To measure oxygen consumption, we adapted a published protocol (Dancy et al., 2016) and used approximately 300 L4 worms grown on plates to the appropriate age and collected them in M9 media (22 mM KH₂PO₄, 42 mM Na₂HPO₄, 86 mM NaCl, and 1 mM MgSO₄). The worms were washed 3 times and aliquoted over 3–5 wells of a 24-well plate from Seaorse Biosciences containing 500 μL of M9 media. Images were taken of the wells to account for the exact number of worms deposited in each well, and the plates were analyzed with a Seahorse Biosciences XF24 Extracellular Flux Analyzer in accordance with the following program: 10 cycles of 2 min mixing, 2 min resting, and 2 min reading. This method demonstrates a linear increase in oxygen consumption rate (OCR) reading with increasing number of N2 and polg-1(srh1) worms (Figures S1A and S1B). To assess maximal respiration, we injected carbonyl cyanide-(trifluoromethoxy)phenylhydrazone (FCCP) to a final concentration 25 μM, and ten additional readings were made.

Oxidative State

For each data point, five age-matched worms with the roGFP::orp1 construct were anesthetized on a 3% agarose pad with 10 mM levamisole-HCl, and their heads were imaged by excitation at 488 and 405 nm and emission at 500–600 nm. Confocal images were analyzed using ZEN Black software where the total fluorescence emitted by the excitation of each wavelength was calculated in arbitrary units (Figure S2C). Heads of WT and generation-matched polg-1(srh1) worms (Figures S1A and S1B). To assess maximal respiration, we injected carbonyl cyanide-(trifluoromethoxy)phenylhydrazone (FCCP) to a final concentration 25 μM, and ten additional readings were made.

Random Mutation Capture and Mutation Spectrum

The original protocol (Vermulst et al., 2007) was modified from using DNA isolated from mitochondrial preps to using whole-genome preps. Briefly, we sorted 2,000 homozygous polg-1(srh1) worms and performed whole-genome preps. 10–20 μg of total DNA was digested with TaqⅠ restriction enzyme (cat. no. R0149T, New England Biolabs) for 10 hr, with 1 μL of fresh enzyme added every hour. Digested DNA was amplified by qPCR or digital droplet PCR using two primer sets to measure total mtDNA copy number and mutant mtDNA copy number. PCR reactions across TaqⅠ sites that record mutant mtDNA molecules were digested with TaqⅠ post-PCR to determine that digestion was complete, and mtDNA mutation rates were calculated based on copy number measurements and qPCR efficiency.
SUPPLEMENTAL INFORMATION

Supplemental Information includes four figures, one table, and two movies and can be found with this article online at https://doi.org/10.1016/j.celrep.2018.02.099.

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AUTHOR CONTRIBUTIONS

S.H. and M.V. conceived the project. S.H., N.G.E., and J.H.B. performed the mutation frequency analyses. S.H. and J.L.W. determined the mutation spectrum. A.L. performed the oxidative state studies. S.H., A.L., and C.F. carried out the mobility assays. S.H. generated the strains, performed the gentle touch, thrashing, and lifespan studies, and analyzed worm mtDNA over multiple generations. C.M.H., B.P.B., J.A.-F., and T.G. provided worm strains, technical expertise, and analysis tools. S.H. and M.V. wrote the manuscript. All authors contributed to and commented on this manuscript.

DECLARATION OF INTERESTS

The authors declare no competing interests.

REFERENCES

Anderson, S., Bankier, A.T., Barrell, B.G., de Brujin, M.H., Coulson, A.R., Drouin, J., Eperon, I.C., Nierlich, D.P., Roe, B.A., Sanger, F., et al. (1981). Sequence and organization of the human mitochondrial genome. Nature 290, 457–465.

Bansal, A., Zhu, L.J., Yen, K., and Tissenbaum, H.A. (2015). Uncoupling life-span and healthspan in Caenorhabditis elegans longevity mutants. Proc. Natl. Acad. Sci. USA 112, E277–E286.

Copeland, W.G. (2008). Inherited mitochondrial diseases of DNA replication. Annu. Rev. Med. 59, 131–146.

Dancy, B.M., Brockway, N., Ramadasan-Nair, R., Yang, Y., Sedeksky, M.M., and Morgan, P.G. (2016). Glutathione S-transferase mediates an ageing response to mitochondrial dysfunction. Mech. Ageing Dev. 153, 14–21.

De Henau, S., Titleman, L., Vangeheel, M., Luyckx, E., Trashin, S., Pauwels, M., Germani, F., Vlaemincx, C., Vanfleteren, J.R., Bert, W., et al. (2015). A redox signalling globin is essential for reproduction in Caenorhabditis elegans. Nat. Commun. 6, 8782.

El-Hattab, A.W., and Scaglia, F. (2013). Mitochondrial DNA depletion syndromes: review and updates of genetic basis, manifestations, and therapeutic options. Neurotherapeutics 10, 186–198.

Han, H.F., and Beckerle, M.C. (2009). The ALP-enigma protein ALP-1 functions in actin filament organization to promote muscle structural integrity in Caenorhabditis elegans. Mol. Biol. Cell 20, 2361–2370.

Haynes, C.M., Fiorese, C.J., and Lin, Y.F. (2013). Evaluating and responding to mitochondrial dysfunction: the mitochondrial unfolded-protein response and beyond. Trends Cell Biol. 23, 311–318.

Hobert, O., Moerman, D.G., Clark, K.A., Beckerle, M.C., and Ruvkun, G. (1999). A conserved LIM protein that affects muscular adherens junction integrity and mechanosensory function in Caenorhabditis elegans. J. Cell Biol. 144, 45–57.

Jova¨isaita, V., Mouchiroud, L., and Autexier, J. (2014). The mitochondrial unfolded protein response, a conserved stress response pathway with implications in health and disease. J. Exp. Biol. 217, 137–143.

Judy, M.E., Nakamura, A., Huang, A., Grant, H., McCurdy, H., Weiberth, K.F., Gao, F., Coppola, G., Kenyon, C., and Kao, A.W. (2013). A shift to osmotic stress resistance in programmed cell death mutants. PLoS Genet. 9, e1003714.

Kamath, R.S., and Ahiringer, J. (2003). Genome-wide mRNA screening in Caenorhabditis elegans. Methods 30, 313–321.

Kashyap, L., Perera, S., and Fisher, A.L. (2012). Identification of novel genes involved in saccopenia in RNAi screening in Caenorhabditis elegans. J. Gerontol. A Biol. Sci. Med. Sci. 67, 56–65.

Kauppila, J.H.K., Baines, H.L., Bratic, A., Simard, M.L., Freyer, C., Mouri¨er, A., Stamp, C., Filogrina, R., Larsson, N.G., Greaves, L.C., and Stewart, J.B. (2016). A Phenotype-driven approach to generate mouse models with pathogenic mtDNA mutations causing mitochondrial disease. Cell Rep. 16, 2980–2990.

Kenyon, C.J. (2010). The genetics of ageing. Nature 464, 504–512.

Kenyon, C., Chang, J., Gensch, E., Rudner, A., and Tabtiang, R. (1993). A C. elegans mutant that lives twice as long as wild type. Nature 366, 461–464.

Kryatskyy, B., Kudryavtseva, E., McKee, A.C., Geula, C., Kowall, N.W., and Khrapko, K. (2006). Mitochondrial DNA deletions are abundant and cause functional impairment in aged human substantia nigra neurons. Nat. Genet. 38, 518–520.

Kujoth, G.C., Hiona, A., Pugh, T.D., Someya, S., Panzer, K., Wohlgemuth, S.E., Hofer, T., Seo, A.Y., Sullivan, R., Jobling, W.A., et al. (2005). Mitochondrial DNA mutations, oxidative stress, and apoptosis in mammalian aging. Science 309, 481–484.

Lewis, S.C., Joers, P., Wilcox, S., Griffith, J.D., Jacobs, H.T., and Hyman, B.C. (2015). A rolling circle replication mechanism produces multimeric larvae of mitochondrial DNA in Caenorhabditis elegans. PLoS Genet. 11, e1004985.

Murphy, C.T., and Hu, P.J. (2013). Insulin/insulin-like growth factor signaling in C. elegans. In WormBook: The Online Review of C. elegans Biology, ed., D.M. Eisenmann, ed. (The C. elegans Research Community).https://doi.org/10.1895/wormbook.1.7.1. https://www.ncbi.nlm.nih.gov/books/NBK179230/.

Nussbaum-Krammer, C.I., Neto, M.F., Brielmann, R.M., Pedersen, J.S., and Morimoto, R.I. (2015). Investigating the spreading and toxicity of prion-like proteins using the metazoan model organism C. elegans. J. Vis. Exp. 95, 52521.

Pellegrino, M.W., and Haynes, C.M. (2015). Mitophagy and the mitochondrial unfolded protein response in neurodegeneration and bacterial infection. BMC Biol. 13, 22.

Peng, M., Ostrovsky, J., Kwon, Y.J., Polyak, E., Licata, J., Tsukikawa, M., Marty, E., Thomas, J., Felix, C.A., Xiao, R., et al. (2015). Inhibiting cytosolic translation and autophagy improves health in mitochondrial disease. Hum. Mol. Genet. 24, 4829–4847.

Pickrell, A.M., and Youle, R.J. (2015). The roles of PINK1, parkin, and mito¨phagy in programmed cell death mutants. PLoS Genet. 11, 22, 1399–1401.

Pellegrino, M.W., and Haynes, C.M. (2015). Mitophagy and the mitochondrial unfolded protein response in neurodegeneration and bacterial infection. BMC Biol. 13, 22.

Peng, M., Ostrovsky, J., Kwon, Y.J., Polyak, E., Licata, J., Tsukikawa, M., Marty, E., Thomas, J., Felix, C.A., Xiao, R., et al. (2015). Inhibiting cytosolic translation and autophagy improves health in mitochondrial disease. Hum. Mol. Genet. 24, 4829–4847.

Pickrell, A.M., and Youle, R.J. (2015). The roles of PINK1, parkin, and mitophagy in Parkinson’s disease. Neuron 85, 257–273.

Polyak, E., Zhang, Z., and Falk, M.J. (2012). Molecular profiling of mitochondrial dysfunction in Caenorhabditis elegans. Methods Mol. Biol. 837, 241–255.

Ryu, D., Mouchiroud, L., Andreux, P.A., Katsyuba, E., Moulian, N., Nicolet-Dit-Félix, A.A., Williams, E.G., Jha, P., Lo Sasso, G., Huzard, D., et al. (2016). Uracil N1 induces mitophagy and prolongs lifespan in C. elegans and increases muscle function in rodents. Nat. Med. 22, 879–888.

Saneto, R.P., and Sedensky, M.M. (2013). Mitochondrial disease in childhood: mtDNA encoded. Neurotherapeutics 10, 199–211.
Springer, W., Hoppe, T., Schmidt, E., and Baumeister, R. (2005). A Caenorhabditis elegans Parkin mutant with altered solubility couples alpha-synuclein aggregation to proteotoxic stress. Hum. Mol. Genet. 14, 3407–3423.

Trifunovic, A., Wredenberg, A., Falkenberg, M., Spelbrink, J.N., Rovio, A.T., Bruder, C.E., Bohlooly-Y, M., Gidlöf, S., Oldfors, A., Wibom, R., et al. (2004). Premature ageing in mice expressing defective mitochondrial DNA polymerase. Nature 429, 417–423.

Trifunovic, A., Hansson, A., Wredenberg, A., Rovio, A.T., Dufour, E., Khvostov, I., Spelbrink, J.N., Wibom, R., Jacobs, H.T., and Larsson, N.G. (2005). Somatic mtDNA mutations cause aging phenotypes without affecting reactive oxygen species production. Proc. Natl. Acad. Sci. USA 102, 17993–17998.

Vermulst, M., Bielas, J.H., Kujoth, G.C., Ladiges, W.C., Rabinovitch, P.S., Prolla, T.A., and Loeb, L.A. (2007). Mitochondrial point mutations do not limit the natural lifespan of mice. Nat. Genet. 39, 540–543.

Vermulst, M., Wanagat, J., Kujoth, G.C., Bielas, J.H., Rabinovitch, P.S., Prolla, T.A., and Loeb, L.A. (2008). DNA deletions and clonal mutations drive premature aging in mitochondrial mutator mice. Nat. Genet. 40, 392–394.

Waaijers, S., Portegijs, V., Kerver, J., Lemmens, B.B., Tijsterman, M., van den Heuvel, S., and Boxem, M. (2013). CRISPR/Cas9-targeted mutagenesis in Caenorhabditis elegans. Genetics 195, 1187–1191.

Wallace, D.C. (2005). A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: a dawn for evolutionary medicine. Annu. Rev. Genet. 39, 359–407.

Wallace, D.C. (2010). Mitochondrial DNA mutations in disease and aging. Environ. Mol. Mutagen. 51, 440–450.

Wanagat, J., Cao, Z., Pathare, P., and Aiken, J.M. (2001). Mitochondrial DNA deletion mutations colocalize with segmental electron transport system abnormalities, muscle fiber atrophy, fiber splitting, and oxidative damage in sarcopenia. FASEB J. 15, 322–332.

Wes, P.D., and Bargmann, C.I. (2001). C. elegans odour discrimination requires asymmetric diversity in olfactory neurons. Nature 410, 698–701.

Xu, H., DeLuca, S.Z., and O’Farrell, P.H. (2008). Manipulating the metazoan mitochondrial genome with targeted restriction enzymes. Science 321, 575–577.

Youle, R.J., and Narendra, D.P. (2011). Mechanisms of mitophagy. Nat. Rev. Mol. Cell Biol. 12, 9–14.