Role of sugarcane bagasse and bamboo for adsorption of hydrolysed dyes from textile effluent: An overview

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Abstract
Adsorption process is one of the best ways for removal of dyes from effluent generated from different industries. The use of absorbent manufactured from sugarcane bagasse as well as bamboo fibre bundles, an agro squander from sugar and building industries have been reviewed as an excellent replacement for stimulated carbon sorbents for the removal of dyes from wastewater. Agricultural based absorbents prepared from sugarcane bagasse and bamboo may be successfully used to remove the unused hydrolyzed dyes from an effluent of textile dyeing industry. These ready absorbents may very much be used in decolorization of the textile effluent. In this review, literature of two sorbents has been compiled. The review analyses these agricultural based materials as low-cost absorbents for the removal of hydrolyzed dyes from textile effluent. The review also draws some of the basic mechanism of dye adsorption to on to.

Keywords: Adsorption, Bamboo, Cellulose, Dyes, Lignin, Sugarcane bagasse

INTRODUCTION
There are wide varieties of dyes that are being used in the textile industries and are causing water pollution by discharge of industrial effluent. Dyes are usually present in trace quantities in treated effluents of many industries (Garg et al., 2004). There are many methods for treatment of the textile effluent. These methods include chemical methods, physical methods and biological treatments. The treatments of effluent by chemical methods include oxidative processes (Cisneros et al., 2002), ozonation (Gähr, et al., 1994) photocatalytic treatment (Namboodri and Walsh, 1996) and by use of sodium hypochlorite (Slokar and Marechal, 1997). Physical methods for effluent treatment include adsorption by biosorbents like sawdust (Sharma et al., 2009), waste orange peel (Namasiyayam et al., 1996), banana pith (Namasiyayam et al., 1998), rice husk (McKAY et al., 1985) and sugarcane bagasse (Xing and Wang, 2009). The other physical methods include ion exchange (Mishra and Tripathy, 1993), irradiation (Hosono et al., 1993), coagulation and flocculation (Zeng et al., 2014). The biological treatment includes decolorization by white-rot fungi (McMullen and Meehan, 2001). Activated carbon prepared from many substances can be used as an adsorbent. The substances that can be used to prepare activated carbon are sawdust and rice-husk (Malik, 2003), coir pith (Santhy and Selvapathy, 2006) and bamboo (Hameed et al., 2007). The manufacturing methods of these activated carbon substances again add on overall cost to cleanup methods. Therefore, cheap and eco-friendly absorbents can be used as an alternative substitution of activated carbon for removal of dyes from wastewater. The adsorption process is one of the effective methods for removal of dyes from waste effluent (Azhar et al., 2005). There are number of low cost, easily available and effective substances which can be used as absorbents for removal of various dyes from aqueous solution or industrial effluents. The use of sugarcane bagasse as an adsorbent, an agro waste from sugar industries has been studied as an alternative substitute for activated carbon for the removal of dyes from wastewater (Azhar et al., 2005; Saad et al., 2010). Sugarcane is a grass that is harvested for its sucrose content. Sugarcane bagasse is the fibrous residue remaining after sugarcane stalk has been crushed and the juice removed. The production of bagasse exceeds 100 million tons annually. Because of its fibrous nature, it has been used as a fuel, paper and pulp, structural materials, and agricultural uses (Han et al., 1983). Reported that bagasse contains carboxylic and hydroxyl group therefore it has been adopted as a cheap, attrac-
tive and effective adsorbent for removal of dyes from wastewater (Saad et al., 2010). A number of reports have been published on sugarcane bagasse acting as an adsorbent for different dyes. These reports show that different pretreatment methods can be employed in order to make bagasse an effective adsorbent. These pretreatments act by disrupting the lignocellulosic matrix (Fig. 1.) thereby reducing the amount of lignin and hemicelluloses as well as modifying the crystalline structure of cellulose to make it more susceptible for adsorption (Silverstein et al., 2007).

Sugarcane bagasse acts as an adsorbent for many dyes present in the industrial effluent. Bamboos are evergreen perennial flowering plants and have been considered as most primitive grasses which are also present in abundant form. Bamboo has similar components to bagasse and scanty of work has been reported on bamboo acting as an adsorbent for industrial effluent. So this review work is based on adsorption of dye from industrial effluent using bagasse as well as bamboo. The used fiber bundles can further be used as fuel for boilers and filling material for seats of cars, buses, trains and so many.

**Bagasse as an adsorbent:** Sugarcane is a member of Gramineae (grasses) family with scientific name of *Saccharum officianrum*. A tropical grass native to Asia, sugarcane plants have been grown for over 4000 years. Sugarcane is native to the warm temperate to tropical regions of South Asia and Melanesia, and used for sugar production. Sugarcane plant is two to six meters (six to twenty feet) tall. It has stout fibrous stalks which are rich in the sugar sucrose. Sugarcane is a C4 plant with a high rate of photosynthesis (its rates lies around 150-200 % above the average for other plants). It can be characterized by segmented stems, blade-like leaves and production by seeds. Sugarcane plant originated from New Guinea where it has been known since about 6000 BC and then spread along human migration routes. Sugarcane is common in tropical and subtropical countries throughout the world. Brazil,

![Fig. 1. Delignification process as pretreatment on biomass (Mosier et al., 2005).](image1)

- Xylose – β(1, 4) – Mannose – β(1, 4) – Glucose –
- Alpha (1, 3) – Galactose

![Fig. 2. Structure of cellulose (Saheb and Jog, 1999).](image2)

**Fig. 3.** Hemicellulose (Brienzo et al., 2016).

![Fig. 4. Main structures present in lignins of sugarcane bagasse (Silva et al., 2011).](image3)

**Table 1.** Chemical composition of sugarcane bagasse reported by various authors (Karp et al., 2013).

| Components            | Soccol et al., 2011 | Rocha et al., 2011 | Bertoti et al., 2009 |
|-----------------------|---------------------|--------------------|----------------------|
| Cellulose (%)         | 32-44               | 45.5               | 47.5-51.1            |
| Hemicellulose (%)     | 27-32               | 27                 | 26.7-28.5            |
| Lignin (%)            | 19-24               | 21.1               | 20.2-20.8            |
| Extractives (%)       | -                   | 4.6                | 0.8-3                |
| Ashes (%)             | 4.5-9               | 2.2                | other components include resin, soaps, sulphur, ash like substances |
Fig. 5. Average chemical composition of bamboo (Lee et al., 1994).

Table 2. Some important tribes and sub-tribes of bamboo (Wilson and Loomis, 1964).

| S. N. | Name of Tribe                  | Name of Sub-tribes | Number of genera and their examples | Total number of genera | Reference                |
|------|--------------------------------|--------------------|-------------------------------------|------------------------|--------------------------|
| 1    | Olyreae (Herbaceous bamboos)   | 1. Buergersiachoinea (1) Olyrea | One genus: Buergersiachoinea 20 genera: Agnesia, Alberella, Cryptochloa, Diandrotyla, Ekmanochloa, Froesichloa, Lithachne, Maclurolyra, Mniochloa, Olyra (plant), Pariana, Parianella, Parodiolyra, Firesea, Piresiella, Raddia, Raddiella, Rehia, Reitzia and Sucrera | 21 | (Shupe et al., 2007) |
| 2    | Bambuseae (Tropical-woody bamboos) | Arthrostylidiinae | 13 genera: Acinoicladum, Apoclada, Arthrostylidium, Ahrostachys, Atractantha, Aulomeinia, Colanthelia, Elytrostachys, Glaziophyton, Merostachys, Rhipidoclaum, 10 genera: Bambusa, Bonia, Dendrocalamus, Dinoochloa, Gigantochoia, Holttumochloa, Kinabaluchloa, Melocalamus, Thyrostachys, Sphaerobambos. | 91 | (Wilson and Loomis, 1964; Dransfield, 1992) |
|      | Chusqueinae                     | Bambusinae         | 2 genera: Chusquea and Neurolepis. 4 genera: Eremocaulon, Guadua, Olmeca and Otaea. |           |                          |
|      | Guaduinae                      |                    | 9 genera: Cephalostachyum, Davidsea, Leptocanna, Melocanna, Neohouzea, Ochlandra, Pseudostachyum, Schizostachyum and teinstachyum. |           |                          |
|      | Melocanninae                   |                    | 6 genera: Decarychoia, Greslania, Hickelia, Hitchcockelia, Nastus and Pernierbam. |           |                          |
|      | Nastinae                       |                    | One genus: Racemobambus. 8 genera: Chimonobambusa, Indosasa, Phyllostachys, Qionghuea, Shibataea, Semianunnaria, Sinobambusa and Temburongia. |           |                          |
|      | Racemobambodinae               |                    | 16 | (Latif, 1993) |
| 3    | Arundinariae (Temperate woody bamboos) | – | Acidosasa, Ampelocalamus, Anundinaria, Borinda, Chimonocalamus, Drepanostachyum, Fargesia, Ferrocalsamus, Indocalamus, Gaoligongshanica, Gelidocalamus, Oligostachyum, Psudosasa, Sasa, Thamnocalamus and Yushania. | 16 | (Latif, 1993) |
Philippine and China are the three largest sugarcane plantation countries in the world (Pandey et al., 2000). It offers one of the most cost-effective renewable resources among those renewable energy options that are readily available in developing countries. Among the by-products available in the cane sugar extraction process, probably the most important one is bagasse (Silverstein et al., 2007).

The main usage of sugarcane is to produce sugar, which can then be used in an infinite number of products. Sugarcane bagasse is a fibrous residue of cane stalks left over after the crushing and extraction of juice from sugarcane. Sugarcane bagasse has been utilized as a raw material for many processes and products which include electricity generation, paper production, products based on fermentation and in adsorption processes (Pandey et al., 2000). It consists of water, fibers and trace amount of soluble solids. Table 1 shows the chemical composition of sugarcane bagasse which has been reported by various authors in a range of research papers (Bertoti et al., 2009; Rocha et al., 2011; Soccol et al., 2011; Karp et al., 2013). It consists of lignin (20-30%), cellulose (40%)

### Table 3. Chemical analysis of various species of bamboo (Higuchi, 1957).

| S.N. | Species                        | Ash (%) | Ethanol-toluene extractives (%) | Lignin (%) | Cellulose (%) | Pentosan (%) |
|------|-------------------------------|---------|--------------------------------|------------|---------------|--------------|
| 1    | Phyllostachys heterocycla     | 1.3     | 4.6                            | 26.10      | 49.10         | 27.70        |
| 2    | Phyllostachys Nigra           | 2.0     | 3.4                            | 23.80      | 42.30         | 24.10        |
| 3    | Phyllostachys Reticulata      | 1.9     | 3.4                            | 25.30      | 25.30         | 26.50        |
-45 %) and hemicelluloses (30-35 %) (Peng et al., 2009). As bagasse is a waste product in sugar industry which acquires additional disposal cost, therefore, most of the mills use it in the boiler as fuel for steam production. The surplus of the bagasse is used in the industry to produce paper, ethanol and livestock feed and building materials. In addition, it can be used for production of important enzymes such as cellulase, xylanase, amylase, inulinase and lipase (Paturau, 1989).

**Cellulose in bagasse:** Cellulose occurs as the major constituent of all plant materials and forms about half to one-third of plant tissues. It is constantly replenished by photosynthesis, with estimates of annual world biosynthesis of 1011 tons. In particular, cellulose is the main constituent of higher plants, including wood, cotton, flax, hemp, jute, SCB, ramie, cereal straws, etc. (Sun et al., 2004). Cellulose is a crystalline, high molecular weight linear homopolymer of repeated units of cellobiose where two anhydrous glucose rings linked by a β-1,4 glycosidic chains. It is completed via the occurrence of 3 hydroxyl groups through different acidity/reactivity, secondary -OH at the C-2, secondary -OH at the C-3, and primary -OH at the C-6 position. Therefore, different intermolecular and intramolecular hydrogen bonds help to

![Phenylpropane, coniferyl alcohol, sinapyl alcohol](image)

**Fig. 9. Building blocks of lignin (Lee, 1996).**

![Building structures of lignin](image)

**Fig. 10. Main substructures of lignin in bamboo (Zhang et al., 2017).**

| S. N. | Age Time (years) | Position | Ash (%) | Hot-Water Solubles (%) | Alcohol-toluene soluble (%) | Lignin (%) | Holo-cellulose (%) | Alpha-cellulose (%) |
|------|------------------|----------|---------|------------------------|-----------------------------|------------|-------------------|--------------------|
| 1.   | One              | Bottom   | 1.82    | 5.83                   | 3.32                        | 21.98      | 68.92            | 46.52              |
|      |                  | Middle   | 1.94    | 5.07                   | 2.86                        | 22.11      | 70.84            | 47.30              |
|      |                  | Top      | 1.95    | 5.14                   | 3.48                        | 21.26      | 71.95            | 47.51              |
| 2.   | Three            | Bottom   | 1.30    | 6.33                   | 4.17                        | 23.21      | 68.58            | 46.21              |
|      |                  | Middle   | 1.36    | 6.91                   | 4.38                        | 23.95      | 72.69            | 46.82              |
|      |                  | Top      | 1.41    | 7.43                   | 5.21                        | 23.71      | 73.82            | 47.51              |
| 3.   | Five             | Bottom   | 1.26    | 4.89                   | 6.61                        | 22.93      | 69.94            | 46.08              |
|      |                  | Middle   | 1.30    | 5.19                   | 6.81                        | 22.97      | 72.50            | 47.65              |
|      |                  | Top      | 1.35    | 5.84                   | 7.34                        | 23.02      | 73.65            | 47.91              |
retain ribbon-like straight formation (Saheb and Jog, 1999). Cellulose possesses equally amorphous as well as crystalline arrangement. The variation in degree of crystallinity of the cellulose depends on its species. This is produced by polymerization of D-anhydroglucopyranose units through 1, 4 β-glycosidic linkage. The high degree of polymerization and linear orientation are responsible for the stiffness in the plant. The structural formula of cellulose is shown in Fig. 2.

**Hemi-cellulose in bagasse:** Larabinose, D-galactose, D-glucose, D-mannose and D-xylose as well as other components such as acetic, gluconic and ferulic acids are five common sugars of hemicelluloses. It is a linear and branched heterogeneous polymer (Fig. 3). The basic structure of this polymer is similar to cellulose but comprising shorter chain length. Pentosan and a little hexosan are also present in its structure (Klemm et al., 1999). Hemicelluloses presenting the cell wall of plant are heteropolysaccharides. On the other side, Xylans are the mainly rich of the hemicelluloses exist in the cell walls of land plants, of which they can comprise additional mass i.e. 30% of the dry weight (Sun, et al., 2004). Cellulose and hemicelluloses can be differentiated by composition of sugar parts, by existence of smaller chains, by a division of major chain molecules, and to be amorphous which insist its structure easier to hydrolyze than cellulose (Fengel and Wegener, 1989).

**Lignin in bagasse:** Lignin is a very complex aromatic molecule raised by the grouping of phenylpropane components allied in a large 3D structure. Three main phenyl propionicalcohols present as monolignils of lignin: p-coumaryl alcohol, coniferyl alcohol and sinapyl alcohol. Lignin and a few of its consequences, bent through the exclusion of lignin from biomass, may slow down the enzymes that accomplish the fermentation proce-
dure, declining biofuel acquiesce (Bottcher et al., 2017). The variation of lignin composition can also exist because of different taxa and even though among diverse tissues as well as cell-wall coatings as of the same plant. The key structures of lignin present in the sugarcane bagasse are shown in Fig. 4. (Silva et al., 2011).

The molecular weight of lignin varies from 300 to 1,40,000 depending on the source and the method of estimation (Gralen, 1946). In other words it can be concluded that it is a nonlinear polymer formed with different monomer units associated by chemically assorted and little-reactive bonds prevents the aptitude of any single enzyme to correctly identify to degrade it. Alcoholic and phenolic hydroxyl groups are more reactive groups in it. Lignin is formed by oxidative polymerization of phenyl propane units to provide bulky cross-linked molecules holding carbon-carbon and ether linkages. Sugarcane contains a prevalence of p-coumaric alcohol which facilitates to a higher concentration of p-hydroxyphenyl alcohol (Del Rio et al., 2015).

**Uses of sugarcane bagasse:** Sugarcane bagasse is a cheap waste material left in the sugar industries that incurs additional disposal cost. The excess of the bagasse is used in the industry to produce ethanol, paper, building materials and livestock feed. In addition, it can also be used to produce various important enzymes such as cellulase, xylanase, amylase, inulinase and lipase (Singh et al., 2009). Bagasse is used as a primary fuel for sugar mills. In various tropical countries, it is the major agricultural crop which is cultivated in abounded form. Sugarcane residues viz. sugarcane bagasse and its leaves have been investigated for biotechnological and non-biotechnological applications point of view. Apart from these, sugarcane bagasse and sugarcane leaves have also been surveyed for employ in lignocellulosic bioconversion, which leads to the economic utilization of residual substances in the

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Table 6. Classification of dyes (Basava Rao and Mohan Rao, 2006).

| Dye Class | Principal textile substrates | Method of application | Chemical Types present in dyes |
|-----------|-----------------------------|-----------------------|--------------------------------|
| Acid      | Nylon, wool, silk, paper, inks and leather | Usually from neutral to acidic dyebaths | Azo (including premetalised), antraquinone, triphenylmethane, azine, xanthene, nitro and nitroso |
| Basic     | Paper, polycrylonitrile, modified nylon, polyester and inks | Applied from acidic dyebaths | Cyanine, hemicyanine, diazahemicyanine, diphenylmethane, triarylmethane, azo, azine, xanthene, acridine, oxazine and anthraquinone |
| Direct    | Cotton, rayon, paper, leather and nylon | Applied from neutral or slightly alkaline baths containing additional electrolyte | Azo, phthalocyanine, stilbene and oxazine |
| Disperse  | Polyester, polyamide, acetate, acrylic and plastics | Fine aqueous dispersions often applied by high temperature/pressure or lower temperature carrier methods; dye may be padded on cloth and baked on or thermo fixed | Azo, antraquinone, styryl, nitro and benzodifuranone |
| Reactive  | Cotton, wool, silk and nylon | Reactive site on dye reacts with functional group on fiber to bind dye covalently under influence of heat and pH (alkaline) | Azo, antraquinone, phthalocyanine, formazan, oxazine and basic |
| Solvent   | Plastics, gasoline, varnishes lacquers, stains, inks, fats, oils and waxes | Dissolution in the substrate | Azo, triphenylmethane, anthraquinone and phthalocyanine |
| Sulphur   | Cotton and rayon | Aromatic substrate vatted with sodium sulphide and reoxidised to insoluble sulphur-containing products on fiber | Indeterminate structures |
| Vat       | Cotton, rayon and wool | Water-insoluble dyes solubilised by reducing with sodium hydrogensulphide, then exhausted on fiber and reoxidised | Anthraquinone (including polycyclic quinines) and indigos |
production of bioethanol as well as value-added commercial products like xylitol, enzymes, organic acids, single-cell protein, etc. (Chandel et al., 2012).

**Bamboo as an adsorbent:** Bamboo is a kind of grass with a firm, woody and empty stem. It is a recurrent evergreen, which means it grows every year and stays green year around due to a rare rhizome dependent system. The scientific name of bamboo is *Bambusoideae*. It covers 1250 species within 75 genera, most of which are relatively fast-growing, achieving maturity within five years, but flowering uncommonly. This composite material can be grown in abundance in most of the hot countries. It is known as a composite material as it consists of cellulose fibers which are surrounded by lignin matrix. Other organic components are also present in addition to cellulose and lignin. It contains about 2-6% starch, 2% deoxidized saccharide, 2-4% fat and 0.8-6% protein (Scurlock et al., 2000). The main properties of the bamboo culm are resolute because of its anatomical structure. Its culm consists of internodes as well as nodes. In the internodes the cells are axially adjusted, while the nodes afford the transversal interconnections (Lessard and Chouinard, 1980).

Bamboo is inexpensive, fast-growing, and easily available, having comparable physical and mechanical properties to wood and can be processed by existing technologies. The fast growth characteristic of bamboo is an advantage for its utilization. Its ecological functions like soil and water conservation along with erosion control are remarkable. On the behalf of Morphology of structure, bamboo species can be classified as either monopodial or sympodial ones and dissimilarities in rhizome system can be observed based on adaptations to the climatic conditions (Gratani et al., 2008). The bamboo contains three clades classified as tribes (Wilson and Loomis, 1964).

**Chemical composition of bamboo:** Bamboo fibre is composed of α-cellulose, hemi-cellulose and lignin along with some minor constituents. The average chemical composition of bamboo is presented in Fig. 5.

The chemical composition of bamboo is almost same to that of wood. Consequently the main constituents of bamboo culms are cellulose (52-60%), hemi-cellulose (20-25%) and lignin (20-30%), which is 90% of the total mass. The negligible constituents of bamboo are resins, tannins, waxes (0.5-0.7%) and some inorganic salts. Compared with wood bamboo is said to have higher alkaline extractives, ash and silica contents (Lee et al., 1994). Owing to the information that it is available in abundant form and economical, it should be used to its fullest level (Tewari, 1992). Alpha-cellulose, lignin, extractives, pentosan, ash and silica content amplified with growing age of bamboo (Yusoff, et al., 1992). Bamboo also contains other organic compounds viz. 2-6% starch, 2% deoxidized saccharide, 2-4% fat, and 0.8-6% protein, in addition to cellulose as well as lignin. It is very much durable because of the carbohydrate content of bamboo (Latif et al., 1991). Table 3 illustrates the analysis of bamboo depending on three different species (Higuchi, 1957). The properties of these bamboo species differ a lot and different species are suggested for different functions. Owing to this information, detection of bamboo species is essential before its efficient utilization.

**Cellulose in bamboo:** It contains both amorphous and crystalline structure. The degree of crystallinity of the cellulose varies from species to species. It is formed by polymerization of D-anhydro glucopyranose units through 1-4 β - glycosidic linkage and provides the stiffness to the plant due to its high degree of polymerization and linear orientation (Ha et al., 1998). The structural formula of cellulose is shown in Fig. 6. (Ha et al., 1998). The lengthy-chain cellulose polymers are connected together by hydrogen and vander Waals bonds, which cause the cellulose to be packed into microfibrils (Zhang and Lynd, 2004). By forming these hydrogen bonds, the chains tend to arrange in parallel and form a crystalline structure. Therefore, cellulose microfibrils have both highly crystalline regions (around 2/3 of the total cellulose) and less-ordered amorphous regions (Figure 1.5). More ordered or crystalline cellulose is less soluble and less degradable (Afrin et al., 2012). The degree of polymerization (DP) is defined as the number of glucose units in a cellulose molecules and cellulose possess good degree of crystallinity. Cellulose (C6H10O5)n, molecules are linear glucans having from 300,000 D to 500,000 D in molecular size. The degree of polymerization in plant/cane fibers like bamboo and bagasse is reported to be lowest (50-600) among the plant fibers depending upon the determination method used and according to one source it is 1050 of bleached bamboo (Dence, 1992).

**Hemi-cellulose in bamboo:** It is a polymer like cellulose but having shorter chain length (DP>150). It is composed of mainly pentosan and a little hexosan (Saheb and Jog, 1999). It is soluble in cold 18% caustic soda. The predominant polysaccharide in jute is composed of a backbone of β-D - xylopyranose unit carrying a terminal 4-O-methyl -α-D- gulcuronic acid residue linked through position two. Hemicellulose is a linear and pronged heterogeneous polymer finished up of five diverse sugars - Larabinose, D-galactose, D-glucose, D-mannose, and D-xylose - in addition to other components such as acetic, glucuronic, and ferulic acids (Fig. 7). The vertebral column of the chains of hemicelluloses can be a homopoly-
Dyes and pigments are measured as one of the major toxins and it is affirmed as ‘noticeable noxious waste’. When these dyes after the use, discharge into water sources, then these are not only influences their visual nature but obliterates marine life due to rich color, chemical oxygen demand (COD), high biochemical oxygen demand (BOD), total organic carbon (TOC) as well as perched solids. On the other side, unused and hydrolysed dyes may also harmful for living life forms such as dysfunction of brain, kidney, reproductive system, liver, and essential nervous structure. As a result, it is necessary to produce economical and efficient methods to solve the pollution troubles created by unused dye present in the effluent (Wei Low et al., 2012). There are many types of dyes like direct, reactive, vat, sulphur, acidic, basic, disperse (dyes for polyester), azo, diazo, anthraquinone based and metal complex dyes, generally used by textile industries. Most of these dyes are toxic, mutagenic and carcinogenic in nature and their deletion from industrial effluents before discharging into the surroundings is very much imperative (Hameed and El-Khaiary, 2008). Effluents coming from textile industries are extremely toxic in nature as they contain a large number of metal complex dyes (e.g. Cr and Co complexes) along with the traces of various acids, alkalis and salts used during wet processing of textile substances. Number of small-scale dyeing industries is facing the problem of shutting since they are not treating their effluents. Also, it is not viable for them to treat the effluent using various existing methods. Hence, it is essential that an appropriate healing method should be invented (Basava Rao and Mohan Rao, 2006). Economical sorbents are the best option for these industries. Before adopting any kind of sorbent, it is very much required to understand the difference dyes and their usages according to the application point of view. The classification dyes according their usage is shown in Table 6. Adsorption of dye: Adsorption is a trend in which gas or liquid fragments get adhered on the exterior of porous material. It is a surface phenomenon and the fluid molecule which gets adsorbed is known as adsorbate. The porous solid material on which the adsorbate gets adsorbed is called adsorbent. This method of adsorption consists of severance of a material from one stage accompanied by its accretion or concentration at the surface of another. The accurate character of the bonding depends on the particulars of the genus involved, however the adsorbed substance is normally categorized as showing physic-sorption or chemi-sorption. It has also been reported that an adsorbent is stuff, typically porous in nature along with high surface area that can absorb substances onto its surface by intermolecular forces (Chincholi et al., 2014). It has been concluded from literature survey that the sugarcane bagasse is composed around of 40% cellulose, 24% hemicellulose, and 25% lignin. On the other side, Bamboo is composed of cellulose (45-55%), hemi-cellulose (20-25%) and lignin (22-30%). In plant cells, including sugarcane and bamboo plants, a secondary wall, consisting of three layers (S1, S2 and S3) which is bounded by a thin primary wall. The secondary wall is surrounded by lignin component. Amorphous cellulose and hemicellulose have been contained by S1 and S3 layers. Out of these two, this amorphous milieu is highly responsible for attracting the hydrolysed as well as unused dye in the effluent. However, the S2 layer contains crystal-line cellulose and these crystalline areas are formed in a extremely ordered way by linear forms of hydrogen bonds which enhance the wicking prosperity of aqueous solutions in this particular cell wall. Though amorphous regions also exist in the cellulose and amorphous cellulose, hemicellulose, along with lignin are also present between the layers (S1, S2 and S3) (Fox et al., 1987). An illustration of the structure of the cell wall with its

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**Lignin in bamboo:** Lignin is a very complex molecule constructed of phenylpropane units linked in a large three-dimensional structure. Three phenyl proponionalcohols exist as monomers of lignin: p-coumaryl alcohol, coniferyl alcohol and sinapyl alcohol. Lignin is closely bound to cellulose and hemicellulose and its function is to provide rigidity and cohesion to the material cell wall, to confer water impermeability to xylem vessels, and to form a physio-chemical barrier against microbial attack (Fengel and Wegener, 1989). Lignin is mainly composed of phenylpropane or C9 units. Three different types of C9 units are present in lignin. The lignin present in bamboo is unique. The lignification process undergoes changes during the elongation of the culm, the full lignification of the bamboo culm is completed within one growing season, showing no further ageing effect. The length of fibres varies with the variation of lignin content in bamboo. The middle position has a long fibre. The outer and inner positions having short fibres possess higher lignin content. Lignin is often called the cementing agent that binds individual cells together (Campbell and Sederoff, 1996).

**Uses of bamboo:**

**Dyes as pollutants:** Dyes and pigments are involved, however the adsorbed substance is normally categorized as showing physic-sorption or chemi-sorption. It has also been reported that an adsorbent is stuff, typically porous in nature along with high surface area that can absorb substances onto its surface by intermolecular forces (Chincholi et al., 2014). It has been concluded from literature survey that the sugarcane bagasse is composed around of 40% cellulose, 24% hemicellulose, and 25% lignin. On the other side, Bamboo is composed of cellulose (45-55%), hemi-cellulose (20-25%) and lignin (22-30%). In plant cells, including sugarcane and bamboo plants, a secondary wall, consisting of three layers (S1, S2 and S3) which is bounded by a thin primary wall. The secondary wall is surrounded by lignin component. Amorphous cellulose and hemicellulose have been contained by S1 and S3 layers. Out of these two, this amorphous milieu is highly responsible for attracting the hydrolysed as well as unused dye in the effluent. However, the S2 layer contains crystal-line cellulose and these crystalline areas are formed in a extremely ordered way by linear forms of hydrogen bonds which enhance the wicking prosperity of aqueous solutions in this particular cell wall. Though amorphous regions also exist in the cellulose and amorphous cellulose, hemicellulose, along with lignin are also present between the layers (S1, S2 and S3) (Fox et al., 1987). An illustration of the structure of the cell wall with its
component organization is shown in Figure 12 (Bidlack et al., 1992).
Different pretreatments are generally given to the lignocellulosic fibers for effective adsorption. It can be observed that there is change in the cellulose, hemicelluloses or lignin structure with pretreatment methods. Alkali treatment reduces the lignin and hemicellulose content in biomass, increases the surface area, allowing penetration of water molecules to the inner layers, and breaks the bonds between hemicellulose and lignin-carbohydrate (Gratzi and Chen, 1999). Reacting biomass with dilute sulfuric acid alters the crystalline nature of the cellulose structure by expanding the surface area of the biomass, allowing water penetration into the crystalline structure. Dilute sulfuric acid treatment improves ease of solubilization of biomass and formation of glucose (Ingram and Doran, 1995). Concentrated sulfuric acid pretreatment solubilizes cellulose by breaking down the hydrogen bonds in hemicelluloses and partially degrades cellulose as well as lignin. Cellulose can be degraded to glucose by acid. The cellulose molecule is characterized by β-1,4-glucosidic linkages between sequential glucose units. There are three reactive hydroxyl groups in each glucose unit (Camacho et al., 1996). Acid can attack the β-1,4-glucosidic linkages in cellulose leading to degradation. Pretreatment is an important tool for practical cellulose conversion processes, and is the subject of this article. Pretreatment is required to alter the structure of cellulosic biomass to make cellulose more accessible to the enzymes that convert the carbohydrate polymers into fermentable sugars as represented in the schematic diagram of Figure 1.12. The goal is to break the lignin seal and disrupt the crystalline structure of cellulose (Mosier et al., 2005).

Observations drawn: The present study revealed that although some reports are available on the use of bagasse as a sorbent for the number of dyes viz. Rhodamine B (RhB), Basic Blue 9 (BB9, also known as methylene blue) etc., but yet reports concerning the use of bamboo are scanty. Natural bamboo fibers possess good potential and may be used as sorbents in textile industries. The high lignin content of the fibre is the major cause of its stiffness. The bamboo is highly renewable and may be classified as ecofriendly. Thus, keeping in view of these observations, in the present investigation, it has been suggested to design agricultural based sorbents using bagasse and extracted bamboo fibres from raw culm for textile effluent treatment.

Conclusions and future perspectives: Adsorption is an attractive and effective method for dye removal from wastewater, especially if the adsorbent is cheap and widely available. In this review, uses of bagasse and bamboo for the adsorption of unused as well as hydrolysed dyes from effluent water may be used based on a significant number of relevant published editorials. The use of bagasse and bamboo as biosorbents for removing various types of dyes from effluent water offers many attractive features such as the outstanding adsorption capacity and the fact that these materials are low-cost, non-toxic and biocompatible. There is a particular need for future studies to verify the performance of the bagasse and bamboo as low-cost adsorbents at the pilot plant scale. There is a great need for additional research concerning as to how to further process or dispose off bagasse and bamboo fibre bundles after their use for the removal of dyes.

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