Comparison of Antioxidant and Anti-Tyrosinase Activity between Black Soybean (Glycine max (L.) Merr.) and Daidzein

Delken Kuswanto¹, I Nyoman Ehrich Lister², ErmiGirsang³, Ali NapiahNasution⁴, Wahyu Widowati⁵

¹Master Program of Biomedical Sciences, Universitas Prima Indonesia, Medan, Indonesia,
²Faculty of Medicine, Universitas Prima Indonesia, Medan, Indonesia,
³Faculty of Public Health, Universitas Prima Indonesia, Medan, Indonesia,
⁴Department of Tropical Medicine, Faculty of Medicine, Universitas Prima Indonesia, Medan, Indonesia,
⁵Faculty of Medicine, Maranatha Christian University, Bandung, Indonesia

Corresponding E-mail: ermigirsang@unprimdn.ac.id

Abstract: Free radicals in the body will increase with excessive ultraviolet (UV) light exposure, induce oxidative stress with the formation of Reactive Oxygen Species (ROS). A visible effect on skin tissue known as photoaging, including the process of melanogenesis catalyzed by the tyrosinase enzyme, risked for pigmentation or melanoma disorder. The use of natural ingredients has been widely used by the community to prevent the aging process. Natural compounds from a plant can be a source of antioxidants and have anti-aging abilities through inhibition of the tyrosinase enzyme. Black soybean (Glycine max (L.) Merr.) are high in the isoflavone compound, one of which is daidzein. This study evaluates the antioxidant and anti-aging potential of black soybean extract with daidzein. Antioxidants using the DPPH method and anti-aging tests carried out, namely the inhibition of the tyrosinase enzyme, are very important in the aging process. Daidzein has an IC⁵₀ value of DPPH scavenging activity around 109.34±2.80 µg/mL lower than black soybean extract with 116.52±2.50 µg/mL. Results on tyrosinase enzyme inhibition activity, black soybean extract had an IC⁵₀ value of 70.71±1.83 µg/mL lower compared to daidzein with 72.65±2.81 µg/mL. In contrast to the better antioxidant activity of daidzein, the black soybean extract is more potential to inhibit the enzyme tyrosinase.

Keywords: black soybean, daidzein, antioxidant, anti-aging, anti-tyrosinase

INTRODUCTION

The aging process is a physiological process that will occur in every living thing and the skin is a body tissue that gives a visual image of aging. Skin aging is a complex process influenced by genetic, environmental and nutritional properties. (1) Excessive ultraviolet (UV) light exposure increases the contribution of free radicals known as Reactive Oxygen Species (ROS). This certainly influences the process of melanogenesis on the skin catalyzed by the
enzyme tyrosinase. This enzyme regulates skin pigmentation through the synthesis of melanin. Increased UV radiation will increase the synthesis of melanin which causes a risk of pigmentation or melanoma disorders.\(^{(2)}\)

Various types of modern medicine have been applied to prevent premature aging, one of which is cosmetic. Most cosmetics have side effects and are not safe, so natural ingredients are needed.\(^{(3)}\) Currently, natural herbal plants are getting attention to solve problems. The use of traditional material has long been used by the community as an alternative treatment, for disease prevention, healing, health recovery and improving health status. Based on WHO data, mentioning Africa, Asia and Latin America using traditional materials as a complement to their primary treatment.\(^{(4)}\)

The use of natural materials as traditional ingredients has also been carried out by Indonesia since the days of our ancestors. The tendency of the use of traditional ingredients in the world is caused by plants containing compounds that are efficacious in medicine known as phytochemical compounds, namely a group of natural compounds that can be used to maintain health and treat diseases due to their antioxidant activity.\(^{(5)}\)

In Indonesia, black soybeans are one of the plants which are the main food commodities after rice and corn.\(^{(6)}\) Black soybeans contain high carbohydrates, vitamins, minerals, and proteins.\(^{(7)}\) Besides, the advantages of black soybean plants have the most content of isoflavones (one of them is *daidzein*) which acts as an antioxidant, only produced from plants and found in legumes.\(^{(8)}\) Lately, antioxidants have become somewhat interesting in the medical world, known to have the effect of preventing premature aging (anti-aging) in fighting free radicals.\(^{(9)}\)

It is necessary to conduct a study of the activity of black soybean extract as an antioxidant and anti-tyrosinase with the *daidzein* comparison compound. This needs to be done to prove the effectiveness of the black soybeans so that black soybeans can be used as natural food to slow down the aging process.

**METHODS**

**Preparation of Black Soybean (Glycine max (L.) Merr.) Extract**

Black soybean seeds (*Glycine max (L.) Merr.*) are washed and dried. *Glycine max (L.) Merr.* were milled and extracted using 70% ethanol by maceration method for 3 days at room temperature. The filtrate was evaporated with a rotary evaporator so that a thick extract was obtained in paste form.\(^{(10)}\)

**Qualitative Phytochemical Screening Assay**

**Flavonoid Identification**

The test tube containing HCl 2N was dissolved by mixing 10 mg *Glycine max (L.) Merr.* into it. Mg/Zn is added enough and heated for 10-15 minutes. Amyl alcohol 1 ml added to the test tube has been cooled and filtered. The presence of a red/orange
color indicates the sample contains a flavonoid compound.\textsuperscript{(11)}

**Saponin Identification**

*Glycine max* (L.) Merr. (10 mg) was dissolved using ddH\textsubscript{2}O in a test tube, simmer for 5 minutes. HCl 1N is added to the solution which has been filtered and shaken strongly. The presence of foam that is still present and remained stable shows a sample containing a saponin compound.\textsuperscript{(11)}

**Phenol Identification**

*Glycine max* (L.) Merr. 10 mg dissolved in 5 ml ddH\textsubscript{2}O, added 500 µl of FeCl\textsubscript{3} 1% solution. The presence of green/red/purple/blue/black color indicated *Glycine max* (L.) Merr. contains phenol compounds.\textsuperscript{(11)}

**Tannin Identification**

HCl 2N around 2 mg was dissolved with 10 mg of *Glycine max* (L.) Merr. in a test tube, heated in a water bath for 30 minutes. Amyl alcohol 500 µl is added after the solution cooled down. Orange/red color in the amyl alcohol layer indicated a tannin compound.\textsuperscript{(11)}

**Alkaloid Identification**

*Glycine max* (L.) Merr. 10 mg dissolved in 5 ml ddH\textsubscript{2}O was evaporated in the water bath. The resulting residue is added 5 ml HCl 2N. The solution obtained is divided into 2 test tubes. The first tube is added 3 drops of HCl 2N which functions as a blank. The second tube solution was transferred as much as one drop to the drip plate, then added 3 drops of Dragendorff reagent. The orange deposits formed indicate the presence of alkaloids.\textsuperscript{(11)}

**Steroid/Triterpeneoid Identification**

The drip plate contains 10 mg *Glycine max* (L.) Merr., added glacial acetic acid until submerged, left for 10-15 minutes and added with a drop of concentrated H\textsubscript{2}SO\textsubscript{4}. The presence of a blue-green color indicated the sample contains a steroid compound, whereas if it shows purple/red/orange color, the sample contains the triterpenoid compound.\textsuperscript{(11)}

**Terpenoid Identification**

Vanillin is added enough to the drip plate containing 10 mg of *Glycine max* (L.) Merr. Addition of one drop H\textsubscript{2}SO\textsubscript{4} into it and homogenize. The presence of purple color contains the terpenoid compound.\textsuperscript{(11)}

**DPPH (2,2-diphenyl-1-picryl-hydrazyl-hydrate) Assay**

DPPH 0.077mmol 200 µl in methanol was added to each 50 µl of *Glycine max* (L.) Merr. extract and daidzein respectively on the microplate. The mixture was incubated at room temperature for 30 minutes. DPPH 250 µl for negative control. The absorbance value is measured at a wavelength of 517 nm using a microplate reader.\textsuperscript{(12)}

\[
DPPH \text{ scavenging activity (\%)} = 1 - \left( \frac{\text{sample}}{\text{control}} \right) \times 100
\]
Tyrosinase Inhibitor Assay

The mixture consisted of 20 μl *Glycine max* (L.) Merr., 20 μl Tyrosinase from Mushroom (125 U/ml) enzyme, and 140 μl potassium phosphate buffer (20 mM, pH 6.8) were incubated at room temperature for 15 minutes. The control contained 20 μl enzyme and 160 μl phosphate buffer, then added 20 μl of L-DOPA substrate (1.5 mM) and incubated again at 10 minutes. The absorbance is measured using a wavelength of 470 nm. The percentage of tyrosinase inhibitor activity is calculated using the formula:

\[
\text{scavenging activity (\%)} = \frac{\text{control} - \text{sample}}{\text{control}} \times 100
\]

RESULTS

Phytochemical Screening of *Glycine Max* (L.) Merr.

Phytochemical screening of black soybean seed extract (*Glycine max* (L.) Merr.) includes alkaloid, tannin, steroid/triterpenoid, phenol, saponin, flavonoid, and terpenoid. Some of this substrate or component can affect the ability of antioxidants, such as alkaloid and terpenoid. The phytochemical test results provide an overview of the classes of compounds contained in *Glycine max* (L.) Merr. which is saponin, tannin, triterpenoid, terpenoid, alkaloid, and can be seen in Table 1.

DPPH Scavenging Activity

The result of the antioxidant activity test using the DPPH method in various concentrations can be seen in Table 2 and Figure 1 with Tukey HSD statistical analysis. In the process of analyzing whether *Glycine max* (L.) Merr. and the daidzein compound has antioxidant activity, it can be seen as a linear regression equation to find IC50 values in Table 3.

Table 1. Phytochemical Screening of *Glycine max* (L.) Merr.

| Component        | Result |
|------------------|--------|
| Flavonoid        | -      |
| Saponin          | +      |
| Phenol           | -      |
| Tannin           | +      |
| Steroid/triterpenoid | +/    |
| Terpenoid        | +      |
| Alkaloid         | +      |

Table 2. DPPH Scavenging Activity

| Concentration (µg/ml) | Glycine max (L.) Merr. | Daidzein |
|-----------------------|------------------------|----------|
|                       | Average DPPH Scavenging (%) |
| 200                   | 65.01 ± 2.02c           | 67.87 ± 1.02e |
| 100                   | 49.85 ± 0.84b           | 50.88 ± 0.99d |
| 50                    | 34.52 ± 1.25a           | 37.88 ± 0.46c |
| 25                    | 31.59 ± 1.10a           | 30.96 ± 0.44b |
| 12.5                  | 31.97 ± 1.99a           | 29.24 ± 0.51ab |
| 6.25                  | 29.90 ± 3.07a           | 27.57 ± 1.23a |

Notes:

Data were presented as mean ± standard deviation. Different small letters in the same column are significant at P < .05 (Tukey HSD Post Hoc test).
The antioxidant activity test used DPPH scavenging activity parameters as seen in Table 2, where *Glycine max* (L.) Merr. and *daidzein* had the highest antioxidant activity at a concentration of 200 μg/ml, each at 65.01±2.02 % and 67.87±1.02 %. The lower the concentration shows the smaller the percentage of antioxidant activity of *Glycine max* (L.) Merr. and *daidzein*. Tabel 3 shows that *Glycine max* (L.) Merr. and *daidzein* with DPPH scavenging activity obtained IC50 values averaging 116.52±2.50 μg/ml and 109.34±2.80 μg/ml.

### Tyrosinase Inhibitor Activity

The test results of tyrosinase enzyme inhibition activity in various concentrations can be seen in Table 4 and Figure 2. The test results of tyrosinase inhibition activity in both samples were linear regression equations to find the IC50 value in Table 5.

| Sample      | Equation               | R²  | IC50 (μg/ml) | Average IC50 (μg/ml) |
|-------------|------------------------|-----|--------------|----------------------|
| Glycine max | Y = 0.1735x + 30.066   | 0.97| 114.89       |                      |
| 2           | Y = 0.1766x + 28.914   | 0.97| 119.40       |                      |
| 3           | Y = 0.2123x + 25.526   | 0.99| 115.28       |                      |
| Mean        | Y = 0.1875x + 28.168   | 0.98| 116.44       | 116.52 ± 2.50        |
| Daidzein    | Y = 0.2062x + 27.342   | 0.99| 109.88       |                      |
| 2           | Y = 0.2190x + 26.718   | 0.99| 106.31       |                      |
| 3           | Y = 0.2112x + 26.380   | 0.99| 111.84       | 109.34 ± 2.80        |
| Mean        | Y = 0.2121x + 26.813   | 0.99| 109.32       |                      |
| Glycine max | Y = 0.1735x + 30.066   | 0.97| 114.89       |                      |
| 2           | Y = 0.1766x + 28.914   | 0.97| 119.40       |                      |
| 3           | Y = 0.2123x + 25.526   | 0.99| 115.28       | 116.52 ± 2.50        |
| Mean        | Y = 0.1875x + 28.168   | 0.98| 116.44       |                      |
| Daidzein    | Y = 0.2062x + 27.342   | 0.99| 109.88       |                      |
| 2           | Y = 0.2190x + 26.718   | 0.99| 106.31       |                      |
| 3           | Y = 0.2112x + 26.380   | 0.99| 111.84       | 109.34 ± 2.80        |
| Mean        | Y = 0.2121x + 26.813   | 0.99| 109.32       |                      |
Table 4. Tyrosinase Inhibition Assay

| Concentration (µg/ml) | Tyrosinase Inhibition Activity (%) | Glycine max (L.) Merr. | Daidzein |
|-----------------------|------------------------------------|-----------------------|---------|
| 100                   | 60.77 ± 1.12<sup>a</sup>           | 57.47 ± 1.16<sup>c</sup> |         |
| 50                    | 42.66 ± 0.66<sup>b</sup>           | 44.36 ± 0.97<sup>d</sup> |         |
| 25                    | 31.68 ± 0.57<sup>b</sup>           | 36.94 ± 1.43<sup>c</sup> |         |
| 12.5                  | 30.57 ± 0.80<sup>b</sup>           | 32.02 ± 0.72<sup>d</sup> |         |
| 6.25                  | 27.10 ± 0.73<sup>a</sup>           | 26.20 ± 0.58<sup>a</sup> |         |
| 3.13                  | 27.48 ± 0.72<sup>a</sup>           | 24.73 ± 0.42<sup>a</sup> |         |

Notes:
Data were presented as mean ± standard deviation. Different small letters in the same column are significant at P<.05 (Tukey HSD Post Hoc test).

Based on Table 4 and Figure 2, the tyrosinase inhibition activity of Glycine max (L.) Merr. was highest at a concentration of 100 µg/ml of 60.77±1.12 %. It was different at a concentration of 50 µg/ml, 25 µg/ml, and 12.5 µg/ml where tyrosinase inhibition activity by Glycine max (L.) Merr. decreased with a percentage of 42.66±0.66 %, 31.68±0.57 %, and 30.57±0.80 %. Daidzein compound inhibition activity was higher at the same concentration of 44.36±0.97 %, 36.94±1.43 % and 32.02±0.72 % and higher than the daidzein compound at the same concentration of 57.47±1.16 %.

Table 5 shows the Glycine max (L.) Merr. and daidzein compound showed that both could inhibit the tyrosinase enzyme with IC<sub>50</sub> at 70.71±1.83 µg/ml and 72.65±2.81 µg/ml.

**DISCUSSION**

The phytochemical screening is carried out as a preliminary stage which aims to determine the class of bioactive compounds in a plant studied. The selection of the solvent and extraction method is the most important factor in conducting a phytochemical screening assay. The phytochemical screening is done qualitatively using a color reagent and see the color changes that occur. Black soybean seeds (Glycine max (L.) Merr.) is known as...
nuts with nutritional value and is rich in protein. Not identified in our research, daidzein is one of the highest bioactive compounds in black soybean seeds and acts as an antioxidant. Therefore, daidzein was used as a comparison.

DPPH method is a simple method used to determine the antioxidant activity of plant material by capturing hydrogen atoms from the component of the extract of the plant. DPPH is sensitive to light, oxygen, and pH. However, it is stable in a radical form so that it may be quite an accurate measurement of antioxidant activity.

The lower the concentration shows the smaller the percentage of antioxidant activity of Glycine max (L.) Merr. and daidzein. Antioxidant activity with DPPH scavenging on daidzein compound was higher than Glycine max (L.) Merr. at concentration of 200 μg/ml, 100 μg/ml, and 50 μg/ml.

Glycine max (L.) Merr. and daidzein compound showed that both could inhibit the tyrosinase enzyme with IC50 at 70.71±1.83 μg/ml and 72.65±2.81 μg/ml. The tyrosinase enzyme inhibition study showed anti-tyrosinase activity produced by Glycine max (L.) Merr. and daidzein compound, according to Lai et.al (2012) using black soybean sprouts extract which has a strong activity of 98% in inhibiting the tyrosinase enzyme and can be used as a lightening agent in skin cosmetic product. Another study by Sitanggang et al. also showed a positive correlation in inhibiting >50% of the tyrosinase enzyme activity by yellow soybean.

The effectiveness of a compound in biological or biochemical functions that can inhibit the oxidation process by 50% (IC50) is classified in several groups including <50 μg/ml (very strong); 50-100 μg/ml (strong); 101-150 μg/ml (medium); >150 μg/ml (weak). In this study, the IC50 value in each test is grouped in Table 6.
Table 6. The IC50 Value of *Glycine max* (L.) Merr. and Daidzein

| Sample                  | Assay                      | IC50 (µg/ml) | Activity |
|-------------------------|----------------------------|--------------|----------|
| Glycine max (L.) Merr.  | DPPH assay                 | 116.52 ± 2.50| Medium   |
| Daidzein                |                            | 109.34 ± 2.80| Medium   |
| Glycine max (L.) Merr.  | Tyrosinase inhibition assay| 70.71 ± 1.83 | Strong   |
| Daidzein                |                            | 72.65 ± 2.81 | Strong   |

CONCLUSIONS

Bioactive compounds that presence in black soybean (*Glycine max* (L.) Merr.) are saponin, tannin, triterpenoid, terpenoid, an alkaloid. *Glycine max* (L.) Merr. and *daidzein*, both have antioxidant and tyrosinase inhibition activity as anti-aging. From the result, it shows that *Glycine max* (L.) Merr. has tyrosinase inhibitor activity better than *daidzein*. Therefore *Glycine max* (L.) Merr. can be developed into anti-aging ingredients in the product.

ACKNOWLEDGEMENTS

The author would like to thank DR. Ermí Girsang, SKM, M.Kes and dr. Ali Napiah Nasution, M.K.M as a mentor, DR. dr. I Nyoman Ehrich Lister, M.Kes., AIFM as head of the study program, dr. Linda Chiuman, M.K.M as dean, DR. Chrismis Nova linda Ginting, M.Kes as chancellor, and Aretha Medika Utama for supporting this research.

REFERENCES

1. Tobin DJ. Introduction to skin aging. Journal of tissue. 2016; 26(1):1-5.
2. Lai X, Wichers HJ, Soler-Lopez M, Dijkstra BW. Structure and Function of Human Tyrosinase and Tyrosinase-Related Proteins. Chem. Eur. J. 2018; 24:50-53.
3. Siti ZR, Norkhadijah SIS, Praveena SM. Hazardous Ingredients in Cosmetics and Personal Care Products and Health Concern: A Review. Public Health Research. 2015; 5(1):1.
4. World Health Organization. Traditional Medicine Strategy. 2013.
5. Iris FFB & Sissi WG. Herbal medicine : biomolecular and clinical aspects, 2nd Ed. New York: Taylor & Francis Group, (Chapter 1). 2011.
6. Arifin AS. Kajian Morfologi Anatomi dan Agronomi antara Kedelai Sehat dengan Kedelai Terserang Cowpea Mild Mottle Virus serta Pemanfaatannya sebagai Bahan Ajar Sekolah Menengah Kejuruan. Jurnal Pendidikan Sains. 2013; 1(2):115.
7. Alghamdi et al. Comparative phytochemical profiling of different soybean (Glycine max (L.) Merr) genotypes using GC–MS. Saudi Journal of Biological Sciences. 2018; 25:15.
8. Sumardi et al. Potential of local black soybean as a source of the isoflavones *daidzein* and genistein. International Food Research Journal. 2017; 24(5):2140-2141.
9. Garg C. Khurana P, Garg M. Molecular Mechanisms of Skin Photoaging and
10. Farnakope Herbal Indonesia. Jakarta: National Health Ministry; 2013.

11. Widowati et al. Antioxidant and Anti-Aging Assays of Oryza Sativa Extracts, Vanillin and Coumaric Acid. Journal of Natural Remedies. 2016; 16(3):89-90.

12. Widowati et al. Antioxidant and Antiaging Activities of JasminumSambac Extract, and its Compounds. Journal of Reports in Pharmaceutical Sciences. 2018; 23(3):272-273.

13. Tu PTB, Tawata S. Anti-Oxidant, Anti-Aging, and Anti-Melanogenic Properties of the Essential Oils from Two Varieties of Alpiniazerumbet. Molecules. 2015; 20:16727-16728.

14. Altemimi A, Lakhssassi N, Baharlouei A, Watson DG, Lightfood DA. Phytochemicals: Extraction, Isolation, and Identification of Bioactive Compounds from Plant Extracts. Plants. 2017; 6:2-4.

15. Al-Kawaz HS, Al-Mashhady LAM. Evaluation of The Phytochemical Constituents and Oxidant-Antioxidant Status for Actinidiadeliciosa Extracts. International Journal of Pharmacy & Therapeutics. 2016;7(1):35-36.

16. Fidrianny I, Elvianna D, Ruslan K. In Vitro Antioxidant Activities in Various Beans Extracts of Five Legumes from West of Java-Indonesia Using DPPH and ABTS Methods. International Journal of Pharmacognosy and Phytochemical Research. 2016; 8(3):471-474.

17. Kladna A. Berczyński P, Kruk I, Piechowska T, Aboul-Enein HY. Studies on the antioxidant properties of some phytoestrogens. Luminescence. 2016; 31(6).

18. Lai et al. Study of Active Ingredients in Black Soybean Sprouts and Their Safety in Cosmetic Use. Molecules. 2012; 17:11672.

19. Sitanggang SSDH, Yanti, Lay BW. Effect of Purified Protein Fractions from Soybean on Tyrosinase Inhibition. Sch. Acad. J. Biosci. 2017; 5(8):594.

20. Budaraga IK, Arnim, Marlinda Y, Bulanin U. Antioxidant Properties of Liquid Smoke Production Variation of Pyrolysis Temperature Raw and Different Concentration. International Journal of PharmTech Research. 2016; 9(6):370.