Research Article

Effect of Single-Dose and Short-Term Administration of Si Jun Zi Tang on the Pharmacokinetics of Gefitinib in Rats

Ying Li,1 Xiaowei Zhou,1 Ming Niu,2 Mingyu Zhang,1 Qiong Gu,1 Wei Chen,1 Bin Dong,1 Yuanyuan Zhang,1 Ruisheng Li,3 Chunyu Li,1, and Guohui Li1

1Department of Pharmacy, National Cancer Center/National Clinical Research Center for Cancer/Cancer Hospital, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing, China
2Integrative Medical Center, The Fifth Medical Center of Chinese People’s Liberation Army General Hospital, Beijing, China
3Research Center for Clinical and Translational Medicine, The Fifth Medical Center of Chinese People’s Liberation Army General Hospital, Beijing, China

Correspondence should be addressed to Chunyu Li; chunyu_li@126.com and Guohui Li; tcm_sci@126.com

Received 6 January 2021; Revised 8 July 2021; Accepted 15 July 2021; Published 27 July 2021

Academic Editor: Silvia Wein

Copyright © 2021 Ying Li et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Background. Si Jun Zi Tang (SJZ), a four-herb Chinese medicine formula that has been described for approximately one thousand years, is often prescribed for cancer patients as a complementary therapy in China. However, the mechanism by which Si Jun Zi Tang enhances the efficacy of gefitinib is unclear. Methods. We investigated how Si Jun Zi Tang affected the pharmacokinetics of gefitinib in rats. A rapid, specific, and reliable ultra-performance liquid chromatography method with mass spectrometry was established to determine the plasma concentration of gefitinib.

Results. The results showed that a single intragastrically administered dose of Si Jun Zi Tang increased the pharmacokinetic parameters of gefitinib ($C_{\text{max}}$, 3156.13 $\mu$g/L; $\text{AUC}$, 46281.5 $\mu$g/L/h) by 3 folds in rats compared with the administration of gefitinib alone ($C_{\text{max}}$, 1352.07 $\mu$g/L; $\text{AUC}$, 11823.7 $\mu$g/L/h). Si Jun Zi Tang could also alter the pharmacokinetics of gefitinib by prolonging the time to reach $C_{\text{max}}$.

Conclusions. Potential pharmacokinetic interactions between gefitinib and SJZ were evaluated, and SJZ extended $T_{\text{max}}$ and $T_{1/2}$ and increased the $C_{\text{max}}$ and $\text{AUC}$ of gefitinib. Long-term administration of gefitinib in combination with Si Jun Zi Tang would improve the efficacy of gefitinib.

1. Introduction

Gefitinib is a selective small-molecule epidermal growth factor receptor (EGFR) tyrosine kinase inhibitor (TKI) that is approved for the treatment of advanced non-small-cell lung cancer (NSCLC) with mutations of EGFR that are sensitive to gefitinib [1, 2]. Gefitinib is metabolized in the liver by cytochrome P450 (CYP) enzymes, mainly CYP3A4 [3]. Thus, it is susceptible to drug-drug interactions with any coadministered drug, and it has been known to be an inhibitor or inducer of CYP450 enzymes [3–5]. It is necessary to monitor the clinical application of gefitinib to ensure efficacy and to prevent adverse reactions.

Si Jun Zi Tang (SJZ) was first described in the “Prescription People’s Welfare Pharmacy” approximately one thousand years ago for the treatment of gastrointestinal disorders, and it can effectively treat nausea, vomiting, and diarrhoea [6]. SJZ is a traditional Chinese medicine compound, and its main ingredients are four traditional herbal medicines, namely, Panax ginseng, Atractylodes macrocephala Koidz, Poria cocos, and Nardostachys jatamansi.

Recently, the use of herbal medicines in combination with antitumour drugs has developed considerable interest [7, 8]. It plays a critical role in improving the quality of life of chemotherapy patients with malignant tumours such as lung cancer, colon cancer, and gastric cancer [9, 10]. It has been found that SJZ indeed improved the quality of life of chemotherapy patients, and it was used in the clinic in combination with chemotherapeutics for the treatment of cancer [10]. Emerging evidence has shown that SJZ can play a significant role in suppressing tumours and has a protective effect against gastrointestinal mucosa damage induced by
chemotherapy [9]. Some studies [11–16] have shown that Chinese herbal ingredients in Si Jun Zi decoction influence the CYP450 enzyme. The Liang Yang group [11] found that ginseng reduced the metabolic rate of midazolam and caffeine, which were metabolized by CYP450 enzymes.

A previous study showed that SJZ combined with gefitinib could inhibit tumour growth and cancer cell metastasis to the lungs in a mouse model of Lewis lung cancer cells [17]. Nevertheless, the mechanism by which SJZ enhances the efficacy of gefitinib is unclear. We established an ultra-performance liquid chromatography method with mass spectrometry detection (UPLC-MS/MS) to determine the gefitinib concentration in rat plasma and studied the effect of SJZ on the pharmacokinetics (PK) of gefitinib.

2. Methods

2.1. Materials and Reagents. Gefitinib was purchased from Macklin Biochemical Co., Ltd. (Shanghai, China). The reference standards of gefitinib (purity 99.8%) and propranolol hydrochloride (purity 100%, internal standard) were obtained from the National Institutes for Food and Drug Control (Beijing, China). Si Jun Zi Tang consisting of Panax ginseng C. A. Mey. (roots), Atractylodes macrocephala Koidz. (roots), Poria cocos (Schw.), Wolf (sclerotia), and Nardostachys jatamansi DC were provided by Beijing Luye Pharmaceutical Company.

HPLC-grade solvents, including methanol and acetonitrile (Thermo Fisher, MA, USA), were used in the study. HPLC-grade formic acid was purchased from Thermo Fisher (MA, USA), and the water was Wahaha purified (Shenyang, China).

2.2. Instrumentation. UPLC-MS/MS analysis was carried out on a Waters Model Xevo TQD separation system (Waters, Milford, MA, USA) supplied with a sample manager and a binary solvent manager. The instrument was also equipped with electrospray ionization mass spectrometry in multiple reaction monitoring (MRM) mode using a triple-quadrupole mass spectrometric detector (Waters, Milford, MA, USA) supplied with a sample manager and a binary solvent manager. The instrument was also equipped with electrospray ionization mass spectrometry in multiple reaction monitoring (MRM) mode using a triple-quadrupole mass spectrometric detector (Waters, Milford, MA, USA). The data were acquired and processed using the MassLynx TM Version 4.1 software (Micromass, Manchester, UK).

2.3. Preparation of Calibration Standards and QC Samples. Stock solutions of gefitinib were prepared from two independent weightings, one for the calibration standards and the other for the quality control (QC) samples. Stock solutions (1.0 mg/mL) were prepared by dissolving gefitinib in methanol. Propranolol was used as an internal standard (IS) and was dissolved in methanol to obtain a 1.0 μg/mL working solution. All solutions were kept at −20°C and were increased to room temperature before use.

The stock of gefitinib at 1.0 mg/mL was diluted with methanol to obtain a standard solution containing 10 μg/mL of gefitinib, and then the standard sample was diluted with rat blank plasma to achieve final concentrations of 2000, 1500, 1000, 500, 200, 50, 20, and 10 μg/L. Using the same method, QC samples with high, medium, and low concentrations were prepared with blank plasma at 1400, 300, and 30 μg/L.

2.4. Sample Preparation. A simple protein precipitation method was carried out to extract gefitinib from all plasma samples. The IS was diluted with acetonitrile to a final concentration of 100 μg/L. IS (450 μL) was pipetted into 150 μL of plasma, vortexed for 2 min, and centrifuged at 13000 rpm for 10 min to precipitate the protein. The supernatant was collected and evaporated to a dried residue by a speed vacuum concentrator. The dried residue was reconstituted with 100 μL of the initial mobile phase and centrifuged at 13000 rpm for 5 min. Portions of the 10 μL volume were directly injected into the UPLC-MS/MS system.

2.5. Preparation of the Test Substance. The powder obtained from gefitinib was dissolved in 0.5% sodium carboxymethyl cellulose (CMC-Na) at a concentration of 4.5 mg/mL. SJZ comprises four commonly used herbs, Panax ginseng C. A. Mey. (roots), Atractylodes macrocephala Koidz. (roots), Poria cocos (Schw.), Wolf (sclerotia), and Nardostachys jatamansi DC, (roots), in the ratio of 9:9:9:6, respectively. The quality of each herb conformed to the requirements of the standards specified by the Chinese Pharmacopoeia [18]. The dried aqueous extract of SJZ was prepared by twice reflushing the four herb powders for 30 min with 8 volumes (vs. four herb weights) of water, and then the water extracts were pooled and concentrated under reduced pressure and finally dried to a powder under a low temperature.

2.6. Pharmacokinetic Experiments in Rats. Male Sprague-Dawley rats weighing 280 ± 30 g were supplied by Beijing Vital River Laboratory Animal Technology Co., Ltd. (Licence No. SYXX 2014-0003) (Beijing, China). The animals were kept in a room at 22–24°C with 55–60% relative humidity and a light cycle (12-hour light and 12-hour dark). They had free access to standard rodent chow and clean water ad libitum. The rats were fasted for 12 hours before the experiments while water was freely supplied.

The healthy rats were divided into six groups of six rats each. The control group was treated orally with gefitinib (45 mg/kg) that was dissolved in 0.5% CMC-Na. SJZ at an equivalent dose (2.97 g/kg), three times the equivalent dose (8.91 g/kg), six times the equivalent dose (17.82 g/kg), and twelve times the equivalent dose (35.64 g/kg) was administered by gavage to rats in groups A-D, and 1 h later, the rats in groups A-D were given gefitinib at a dose of 45 mg/kg.

SJZ (8.91 g/kg) was administered by gavage to six rats daily for 12 consecutive days in group E to examine the effects of multiple doses of the herbal medicines one hour after the last administration of SJZ at day 12, and gefitinib (45 mg/kg) was administered (Figure 1).

Blood samples (0.5 mL) were collected from the fosse orbital veins and were placed into heparinized polyethylene tubes at 0.25, 0.5, 1, 1.5, 2, 3, 4, 6, 8, 12, and 24 h after dose.
The supernatant of blood samples was collected after centrifugation at 4000 rpm for 10 min and stored at −80°C until analysis. The pharmacokinetics analysis was performed by a noncompartmental approach using DAS version 2.0 (China).

2.7. Statistical Analysis. Statistical analyses were performed using SPSS software (SPSS for Windows, version 22.0, IBM Corp, Armonk, NY, USA). Noncompartmental PK parameters are presented as the mean ± standard deviation (mean ± SD) and were analysed with unpaired t-tests for comparisons between two groups or a single-factor ANOVA followed by the Dunnett tests for comparisons among multiple groups. P values less than 0.05 were considered statistically significant.

3. Results

3.1. Mass Method Development. The mass spectrometric parameters were optimized with the corresponding standard solutions. The final analysis was conducted in positive ionization mode since it gave a good response to gefitinib and IS, which was consistent with previous reports [1, 5, 12]. In the precursor ion full scan spectra, [M + H]+ fragments were the most abundant at m/z 447.3 (gefitinib) and m/z 260.0 (IS), and the products selected for quantification were at m/z 127.8 (gefitinib) and m/z 116.1 (IS) (Figure 2). MS/MS parameters, including collision energy, cone voltage, capillary voltage, desolvation temperature, ESI source temperature, collision gas flow rate, cone gas flow, and desolvation gas flow rate, were optimized to obtain the highest response of the protonated molecules of the studied compounds.

3.2. Optimization of Chromatographic Conditions. A mixture of gefitinib and IS was eluted using different mobile phases composed of mixtures of methanol, water, and formic acid. The percentage of acetonitrile in the mobile phase had a significant effect on the shape and retention time. Gradient elution was used to ensure the preferable peak shape and retention time of the two compounds. Moreover, formic acid was essential to obtain sharp peaks for gefitinib. The final analysis was thus performed with gradient elution using a mobile phase of acetonitrile and water with 0.1% (v/v) formic acid. The shortest analytical run time of the present method was 5 min, although all compounds were eluted sufficiently within 2.5 min after injection. The remaining 2.5 min was spent diminishing the memory effect from the column and stabilizing it before the next injection.

3.3. Method Validation

3.3.1. Specificity. The specificity was thoroughly assessed by screening the chromatograms of rat blank plasma spiked with gefitinib and propranolol (Figure 3). No endogenous interference was observed at the retention time of gefitinib (2.23 min) and IS (2.43 min). This result indicated that the developed method had efficient specificity under the working conditions.

3.3.2. Linearity of the Calibration Curves and the Limit of Quantification. Calibration curves were fitted by a linear weighted (1/x2) least squares regression method over the concentration range of 10–2000 μg/L for gefitinib. The linear equation was $y = 0.001x - 0.014$ with a correlation coefficient $r > 0.9995$. The deviation from the nominal concentration should be within ±20% for the lowest limit of quantification (LLOQ). Based on the criteria, the LLOQ value was set at 10 μg/L, and the measured results are presented in Table 1. Carryover was tested by injecting two processed blank matrix samples sequentially after injecting an upper limit of quantification (ULOQ) sample. Carryover should not exceed 20% of the LLOQ in the blank samples. ULOQ was set at 2000 μg/L, and to ensure the analysis of samples over ULOQ, dilution integrity was carried out by diluting an ultrahigh QC sample at ten times the highest calibration level.

3.3.3. Accuracy and Precision. The mean concentration values for the QC samples at 30, 300, and 1400 μg/L were estimated using six replicates per day and run repeated on three separate days to determine the intraday/interday accuracy and precision. The intraday and interday precisions of gefitinib were within ±15% of the nominal concentration (Table 1).

3.3.4. Recovery and Matrix Effect. Recovery was assessed by spiking plasma samples with gefitinib at 30, 300, and 1400 μg/L, along with propranolol (IS). Plasma samples were then processed, and the drugs were extracted as described in the experimental section. The extraction recovery was determined in sextuplicate by comparing the three concentrations of samples with reference solutions containing blank plasma extracts spiked with gefitinib at the same concentrations. The recoveries are presented in Table 2. Recovery values of not less than 86.64 indicated good extraction efficiency of the proposed sample treatment method. The matrix effect was assessed in sextuplicate by comparing the concentrations obtained with the three solutions at 30, 300, and 1400 μg/L in blank plasma extracts with the same solutions in acetonitrile for gefitinib. The matrix factor of not more than 9.52 (Table 2) showed that a negligible matrix effect was found for plasma.

3.3.5. Stability Study. Gefitinib stability in rat plasma was assessed at three concentrations (30, 300, and 1400 μg/L) in sextuplicate. The processed samples were left in the autosampler (10°C) for 48 h to study the autosampler stability. Short-term stability was evaluated by detecting the samples...
left at room temperature (25°C) for 12 h. The samples were stored at −20°C for 60 days for the assessment of long-term stability. Three freeze (at approximately −20°C)-thaw (room temperature) cycles were used to assess the freeze-thaw stability. The results of the stability testing are reported in Table 3 and indicated the stability of gefitinib under the above-mentioned conditions since the recoveries were within the acceptance criteria (±15%). In addition, plasma samples exceeding the upper limit of the assay were adequately diluted 10 times with blank plasma in this study, and the recovery of samples diluted 10 times was 102.43% (RSD = 4.41%).

3.4. Pharmacokinetics Studies in Rats. The parameters of gefitinib are reported in Table 4. In the control group, gefitinib was rapidly absorbed within 3 h of administration, and the elimination half-life (T1/2) was 4.79 h. A longer time to reach the maximum concentration (Tmax), T1/2, and the area under the curves (AUC) were observed in the rats that were pretreated with SJZ prior to the gefitinib dose, and a higher maximum concentration (Cmax) was reported. In addition, clearance (CL/F) was lower than that in the control group.

3.4.1. Effect of SJZ Dosage on Gefitinib and PK. The average concentration-time profiles of gefitinib in the rats receiving different doses of SJZ are shown in Figure 4. There were prolonged Tmax and T1/2 and a higher Cmax and AUC in these groups. According to this study, the Cmax (3156.13 μg/L) was significantly increased only in the group with three times the equivalent dose (8.91 g/kg) of SJZ (P = 0.007), as well as the group with six times the equivalent dose (17.82 g/kg) of SJZ (Cmax: 2888.32 μg/L, P = 0.028) compared with the control group. Tmax was prolonged to 4.5 h, and the average AUC0-t was 46281.5 μg/L/h in the group treated with three times the equivalent dose of SJZ. The CL/F of gefitinib in all groups treated with SJZ was significantly lower than that of the control (P < 0.05). Three times the equivalent dose of SJZ had the greatest influence on the PK of gefitinib. Cmax and AUC0-t of gefitinib increased with increasing SJZ dose in the range of 2.97 g/kg–8.91 g/kg, while Cmax and AUC0-t decreased when the SJZ dose exceeded 8.91 g/kg.

3.4.2. Effect of Multiple Doses of SJZ on Gefitinib and PK. The concentration-time profiles in Figure 4 show that the PK data of gefitinib after the rats were pretreated with SJZ...
(8.91 g/kg) once daily for 12 consecutive days was similar to the single dose (8.91 g/kg), while $C_{\text{max}}$ (4934.85 $\mu$g/L) and AUC$_{0-t}$ (73049.4 $\mu$g/L/h) were higher than the rats with the single-dose and the difference was statistically significant ($C_{\text{max}}$: $P = 0.029$; AUC$_{0-t}$: $P = 0.011$). The CL$_{F}$ (0.39 L/h/kg) was lower than that of the group with a single dose. A long-term administration of gefitinib in combination with SJZ increased the concentration of gefitinib in the rats.

4. Discussion

The UPLC-MS/MS method is becoming an important tool for clinical laboratories because it is a very specific, selective,
we successfully established a UPLC-MS/MS method to determine the concentration of gefitinib. We used a protein precipitation method to prepare samples rather than a solid-phase extraction [5]. This method was simple and convenient to fully precipitate the protein. The accuracy, precision,
The changes in the pharmacokinetic characteristics of gefitinib may be related to the effect of Si Jun Zi decoction on CYP450 enzymes. Gefitinib is metabolized in the liver by CYP3A4 [3]. The concurrent use of other drugs and gefitinib may augment or oppose the pharmacokinetics or pharmacodynamics, which can then increase or decrease the pharmacological or toxicological effects of either constituent. Some studies have shown that Chinese herbal ingredients in Si Jun Zi decoction influence CYP450 enzymes. A study [11] showed that ginseng reduced the CL of midazolam and caffeine, which were metabolized by CYP450 enzymes. CYP3A2 (an isozyme of CYP3A4) and gene expression levels in Sprague-Dawley rats exposed to P. ginseng extracts were evaluated by western blotting and quantitative PCR in the study [11] and compared with the control groups. P. ginseng downregulated the expression of CYP3A2 at both the protein and gene levels. Another study [12] indicated that repeated administration of Salviae Miltiorrhizae radix P. ginseng and Glycyrrhiza uralensis inhibited the expression of CYP3A4. TQ he concurrent use of other drugs and gefitinib may be related to the effect of Si Jun Zi decoction on CYP450 enzymes.

**5. Conclusions**

A simple, rapid, and sensitive UPLC-MS/MS bioanalytical method has been developed and validated for the quantification of gefitinib in rat plasma. The method was successfully applied to PK determination following the oral administration of two combinations of gefitinib and SJZ to rats. Potential pharmacokinetic interactions between gefitinib and SJZ were evaluated, and SJZ prolonged T<sub>max</sub> and T<sub>1/2</sub> and increased C<sub>max</sub> and AUC of gefitinib. Thus, SJZ may enhance the efficacy of gefitinib by changing the pharmacokinetics of gefitinib. These findings should be noted in clinical therapeutics, especially when using two concurrent drugs which may have drug-drug interactions.

**Data Availability**

The data used in the current study can be accessed by request via the corresponding author.

**Ethical Approval**

All procedures were performed in accordance with the Regulations of Experimental Animal Administration issued by the Ministry of Science and Technology of the People’s

---

**Figure 4: Average plasma concentration-time profiles of gefitinib in the control group and in rats pretreated with SJZ.**

| Time (h) | Control | A group | B group | C group | D group | E group |
|---------|---------|---------|---------|---------|---------|---------|
| 0       |         |         |         |         |         |         |
| 5       |         |         |         |         |         |         |
| 10      |         |         |         |         |         |         |
| 15      |         |         |         |         |         |         |
| 20      |         |         |         |         |         |         |
| 25      |         |         |         |         |         |         |
| 30      |         |         |         |         |         |         |

The parameters of the pharmacokinetic characteristics of gefitinib in the control group and in rats pretreated with SJZ were summarized in Table 1. The AUC increased by 175.9% in the group that was pretreated with SJZ, while the CL<sub>z</sub> decreased by 61.3%. In addition, the T<sub>max</sub> and T<sub>1/2</sub> of gefitinib were significantly prolonged in the SJZ pretreated groups.

**Data Availability**

The data used in the current study can be accessed by request via the corresponding author.

**Ethical Approval**

All procedures were performed in accordance with the Regulations of Experimental Animal Administration issued by the Ministry of Science and Technology of the People’s...
Republic of China (http://www.most.gov.cn) and were approved by the Laboratory Animal Ethics Committee of Cancer Hospital, the Chinese Academy of Medical Sciences, and the Peking Union Medical College. After the study, all rats were sacrificed by inhaling anaesthetics for euthanasia.

Disclosure

The funding source had no role in the design of this study and did not have any role during the collection, analysis, and interpretation of the data, as well as the preparation of the manuscript.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

Acknowledgments

The authors thank the Integrative Medical Center of the Fifth Medical Center of Chinese People’s Liberation Army General Hospital for instrumental support. The materials were supported by the CAMS Innovation Fund for Medical Sciences (CIFMS) (Grant nos. 2016-12 M-1-001).

References

[1] N. Zheng, C. Zhao, X.-R. He et al., "Simultaneous determination of gefitinib and its major metabolites in mouse plasma by HPLC-MS/MS and its application to a pharmacokinetics study," Journal of Chromatography B, vol. 1011, pp. 215–222, 2016.

[2] K.-Y. Weon, M. G. Kim, S. Shin et al., "Alterations of gefitinib pharmacokinetics by co-administration of herbal medications in rats," Chinese Journal of Integrative Medicine, vol. 24, no. 6, pp. 460–466, 2018.

[3] A. Chahbouni, J. C. G. den Burger, R. M. Vos, A. Sinjewel, and A. J. Wilhelm, "Simultaneous quantification of erlotinib, gefitinib, and imatinib in human plasma by liquid chromatography tandem mass spectrometry," Therapeutic Drug Monitoring, vol. 31, no. 6, pp. 683–687, 2009.

[4] H. C. Swaisland, M. Ranson, R. P. Smith et al., "Pharmacokinetic drug interactions of gefitinib with rifampicin, itraconazole and metoprolol," Clin Pharmacolinet, vol. 44, pp. 1067–1081, 2005.

[5] H. M. Maher, N. Z. Alzoman, and S. M. Shehata, "Simultaneous determination of selected tyrosine kinase inhibitors with corticosteroids and antiemetics in rat plasma by solid phase extraction and ultra-performance liquid chromatography tandem mass spectrometry: application to pharmacokinetic interaction studies," Journal of Pharmaceutical and Biomedical Analysis, vol. 124, pp. 216–227, 2016.

[6] Y. Lu, L. Li, J.-W. Zhang, X.-Q. Zhong, J.-A. Wei, and L. Han, "Total polysaccharides of the Si Jun Zi Tang enhancing anticancer of gefitinib by an integrating approach," Journal of Separation Science, vol. 33, no. 6, pp. 2167–2175, 2010.

[7] K. Chan, "Understanding interactions between Chinese medicines and pharmaceutical drugs in integrative healthcare," Chinese Journal of Integrative Medicine, vol. 21, no. 2, pp. 83–89, 2015.

[8] H. Xu and K.-j. Chen, "Herb-drug interaction: an emerging issue of integrative medicine," Chinese Journal of Integrative Medicine, vol. 16, no. 3, pp. 195–196, 2010.

[9] Z. Guan, J. Wu, C. Wang et al., "Investigation of the preventive effect of Si Jun Zi decoction on mitomycin C-induced immunotoxicity in rats by 1 H NMR and MS-based untargeted metabolomic analysis," Journal of Ethnopharmacology, vol. 210, pp. 179–191, 2018.

[10] Y. Shun, "Effects of Si Jun Zi decoction on immunologic function and quality of life in patients with rectal carcinoma after surgery," Nanjing University of Chinese Medicine, vol. 32, pp. 334–336, 2016.

[11] L. Yang, Y. Wang, H. Xu et al., "Panax ginseng inhibits metabolism of diester alkaloids by downregulating CYP3A4 enzyme activity via the pregnane X receptor," Evidence-Based Complementary and Alternative Medicine, vol. 2019, Article ID 3508658, 13 pages, 2019.

[12] Y. Zhang, L. Miao, L. Lin, C.-Y. Ren, J.-X. Liu, and Y.-M. Cui, "Repeated administration of Sailuotong, a fixed combination of Panax ginseng, Ginkgo biloba, and Crocus sativus extracts for vascular dementia, alters CYP450 activities in rats," Phytomedicine, vol. 38, pp. 125–134, 2018.

[13] C.-t. Yu, J. Chen, X. W. Teng, V. Tong, and T. K. H. Chang, "Lack of evidence for induction of CYP2B1, CYP3A23, and CYP1A2 gene expression by panax ginseng and panax quinquefolius extracts in adult rats and primary cultures of rat hepatocytes," Drug Metabolism and Disposition, vol. 33, no. 1, pp. 19–22, 2005.

[14] Y. Liu, J.-W. Zhang, W. Li et al., "Ginsenoside metabolites, rather than naturally occurring Ginsenosides, lead to inhibition of human cytochrome P450 enzymes," Toxicological Sciences, vol. 91, no. 2, pp. 356–364, 2006.

[15] G. L. Hendersonab, M. R. Harkeyab, M. E. Gershwinac, R. M. Hackmanad, J. S. Sternad, and D. M. Stresser, "Effects of Ginseng Components on c-DNA-expressed cytochrome P450 enzymes catalytic activity," Life Science, vol. 65, pp. 209–214, 1999.

[16] H. Chen, X. Zhang, Y. Feng, W. Rui, Z. Shi, and L. Wu, "Bioactive components of Glycyrrhiza uralensis mediate drug functions and properties through regulation of CYP450 enzymes," Molecular Medicine Reports, vol. 10, no. 3, pp. 1355–1362, 2014.

[17] C. Li, M. Niu, R. Wang et al., "The modulatory properties of Si Jun Zi Tang enhancing anticancer of gefitinib by an integrating approach," Biomedicine & Pharmacotherapy, vol. 111, pp. 1132–1140, 2019.

[18] National Pharmacopoeia Commission, Pharmacopoeia of the People’s Republic Of China, China Medical Science and Technology Press, Beijing, China, 2015.

[19] C.-K. Noh, J.-H. Lee, M.-S. Kim, H.-J. Maeng, and S.-J. Chung, "Simultaneous quantification of volitinib and gefitinib in rat plasma by HPLC-MS/MS for application to a pharmacokinetic study in rats," Journal of Separation Science, vol. 40, no. 19, pp. 3782–3791, 2017.