Aging is an important risk factor in the occurrence of many chronic diseases. Senescence and exhaustion of adult stem cells are considered as a hallmark of aging in organisms. In this study, a senescent human amniotic mesenchymal stem cell (hAMSC) model subjected to oxidative stress was established in vitro using hydrogen peroxide. We investigated the effects of ganoderic acid D (GA-D), a natural triterpenoid compound produced from Ganoderma lucidum, on hAMSC senescence. GA-D significantly inhibited β-galactosidase (a senescence-associated marker) formation, in a dose-dependent manner, with doses ranging from 0.1 μM to 10 μM, without inducing cytotoxic side-effects. Furthermore, GA-D markedly inhibited the generation of reactive oxygen species (ROS) and the expression of p21 and p16 proteins, relieved the cell cycle arrest, and enhanced telomerase activity in senescent hAMSCs. Furthermore, GA-D upregulated the expression of phosphorylated protein kinase R-like endoplasmic reticulum kinase (PERK), peroxidase III (PRDX3), and nuclear factor-erythroid 2-related factor (NRF2) and promoted intranuclear transfer of NRF2 in senescent cells. The PERK inhibitor GSK2656157 and/or the NRF2 inhibitor ML385 suppressed the PERK/NRF2 signaling, which was activated by GA-D. They induced a rebound for the generation of ROS and β-galactosidase-positive cells and attenuated the differentiation capacity. These findings suggest that GA-D retards hAMSC senescence through activation of the PERK/NRF2 signaling pathway and may be a promising candidate for the discovery of antiaging agents.
as observed in the premature aging diseases including Werner syndrome and Hutchinson-Gilford premature aging syndrome, as well as in the aging mouse models [11, 12]. Furthermore, aged MSCs showed spontaneous expression of embryonic factors and p53 point mutations in an age-related tumorigenesis model [13]. Interestingly, transplantation of mesodermal-derived stem cells into aging mice can prolong their life span [14], while senescent bone marrow MSCs cannot reach the injury site and lose their protection to the lungs due to weak activation, migration, and anti-immune function [15]. Hence, stem cell senescence has received increased attention in recent years in the field of antiaging and regenerative medicine.

Ganoderma lucidum (Leyss. Ex. Fr) Karst, a medicinal mushroom, also referred to as "Lingzhi," has been widely used as a famous traditional Chinese medicine to promote health and longevity for thousands of years in China and other Oriental countries. Furthermore, recent pharmacological studies have demonstrated that the extracts of G. lucidum, such as aqueous extracts and ethanol extracts, induce antiaging effects by increasing the mitochondrial antioxidant activity, scavenging free radicals, and reducing the generation of reactive oxygen species (ROS) [16–18]. Ethanol extracts of G. lucidum have delayed the progression of age-related Alzheimer’s disease by regulating the methylation level of DNA [18]. However, the antiaging bioactive ingredients in G. lucidum have not been elucidated. One study showed that several ergosterol derivatives, ganodermasides A, B, C, and D isolated from the methanol extract of spores of G. lucidum, prolonged the replicative life span of yeast by targeting an age-related gene UTH1 [16]. More than 200 distinct chemical entities with various pharmacological actions, such as antioxidation, antitumor, and antiradiation effects, have been isolated from G. lucidum [19, 20]. However, the number of confirmed antiaging ingredients is lower than the large number of known compounds that have been isolated from G. lucidum. The dysfunction of antioxidant enzymes accelerates the aging process, as they are the first line of defense for protecting biological macromolecules against oxidative stress. Although there are no reports on the antiaging activity of G. lucidum triterpenes, their antioxidant properties suggest that they may have a potential effect on the extension of life span.

Senescent cells accumulate in various aging tissues and at pathogenic sites in many chronic diseases. Notably, targeting cellular senescence is regarded as a promising approach for the delay, prevention, or alleviation of multiple age and cellular senescence-associated conditions and the fundamental aging processes [21]. The free radical theory of aging postulates that the production of intracellular ROS is the major determinant of life span. What is the link between ROS and senescence? Excessive accumulation of ROS induces oxidative damage in cells. A previous study showed that oxidative damage contributed to replicative senescence [22]. Oxidative stress triggers DNA damage, resulting in the dysregulation of cell homeostasis and aging phenotypic characteristics, eventually leading to the acceleration of the cellular senescence [23]. Additionally, H2O2-induced oxidative damage could cause the cellular and molecular changes in senescent cells. For example, both p21 and p16 proteins are elevated during the induction of premature senescence, thereby causing cell cycle arrest and loss of self-replication [22]. Thus, aging and age-related diseases are regulated by intracellular free radicals, and generation of ROS remains one of the most widely accepted causes of aging [24]. Therefore, it is an effective strategy to reduce the excessive accumulation of ROS to slow down the senescence of MSCs.

Based on the free radical theory, we developed a H2O2-induced stem cell senescent model using human amniotic MSCs (hAMSCs) with high expression of β-galactosidase, a senescence-associated marker. Dozens of natural compounds isolated from G. lucidum were screened using the H2O2-induced hAMSC senescent model. Among these compounds, ganoderic acid D (GA-D), a triterpenoid compound, dramatically alleviated stem cell senescence. Therefore, in the present study, we have reported the effect of GA-D on oxidative stress-induced stem cell senescence as well as the underlying mechanism of delayed senescence in hAMSCs.

2. Materials and Methods

2.1. Source and Identification of GA-D Compound. The GA-D compound was purchased from Baoji Chenguang Biotechnology Company, Baoji, China. The properties of GA-D, such as purity, molecular weight, and chemical structure, were identified using high-performance liquid chromatography (HPLC), liquid chromatography-mass spectrometry (LC-MS), and nuclear magnetic resonance (NMR), respectively. The HPLC-grade purity was observed to be above 98% by HPLC analysis (Figure 1(a)), and the molecular formula was determined to be C30H42O9 through ESI-MS (negative) at a mass to charge ratio (m/z) of 513.3 [M-H]− (Figure 1(b)). Consistent with its molecular formula, 30 carbon resonances were observed in the 13C-NMR spectrum, and the data of 13C- and 1H-NMR spectra (Figures 1(c) and 1(d)) were consistent with the compound ganoderic acid D identified by Kikuchi et al. [25]. Consequently, the compound was confirmed to be ganoderic acid D.

2.2. Cell Isolation, Culture, and Identification. As per previously described methods [26, 27], hAMSCs were isolated from placental amnion tissue that was collected from normal pregnant women after gaining informed consent using collagenase type II (Solarbio, Beijing, China) and deoxyribonuclease I (Solarbio, Beijing, China) enzymes. The hAMSCs were cultured in Dulbecco’s modified Eagle medium low glucose (LG-DMEM) (Gibco, New York, USA) supplemented with 10% fetal bovine serum (FBS) (Gibco, New York, USA), 1% nonessential amino acids (Gibco, New York, USA), 10 ng/mL basic fibroblast growth factor (bFGF) (Peprotech, NJ, USA), and 1% L-alanyl-L-glutamine dipeptide (L-GLutaMAX) (Gibco, New York, USA) in a humid atmosphere of 5% CO2 at 37° C. Culture medium was replaced by fresh medium every three days. When the cells reached 80% confluency, the harvested cells were passaged. Cells that belonged to passage 2 (P2) were used for further analysis in the study. Subsequently, these hAMSCs were analyzed using flow cytometry and the immunocytochemical staining method according to the protocols described in previous studies [26, 27].
(a) Figure 1: Continued.
Figure 1: Continued.
research was conducted in accordance with the Declaration of Helsinki and the guidelines of the Ethical Committee of the Affiliated Hospital of Zunyi Medical University (Zunyi China).

2.3. Cell Treatment. When P3 hAMSCs were grown to a confluency of approximately 50%, they were treated with H2O2 at different final concentrations (100 μM, 200 μM, and 400 μM) for 2h. Subsequently, the cells were washed with Dulbecco’s phosphate-buffered saline (D-PBS) to remove residual H2O2 and replaced with fresh LG-DMEM/F12 complete medium. The aging-related indicators were tested after the cells were cultured up to the specified time point. Different final concentrations (0.001 μM, 0.01 μM, 0.1 μM, 1 μM, 10 μM, and 100 μM) of GA-D were used to pretreat hAMSCs for 6h prior to the addition of H2O2. The protein kinase R- (PKR-) like endoplasmic reticulum kinase (PERK) inhibitor GSK2656157 (MCE, Shanghai, China) and nuclear factor-erythroid 2-related factor (NRF2) inhibitor ML385 (MCE, Shanghai, China) were added 1 h prior to the addition of GA-D. GA-D, GSK2656157, and ML385 were dissolved in dimethylsulfoxide (DMSO) (Solarbio, Beijing, China) at a concentration of 10 mM and diluted to the corresponding concentration with D-PBS prior to use.

2.4. Flow Cytometry Analysis. For the phenotypic characterization of hAMSCs, a BD Stemflow™ Human MSC analysis kit (Cat. No. 562245, BD Biosciences, San Diego, CA, USA) was used in the study. P3 hAMSCs in the logarithmic growth phase were harvested and labeled with different antibodies for human MSC-specific markers (CD73, CD90, and CD105) for flow cytometry analysis. Briefly, P3 hAMSCs were collected and washed twice with D-PBS containing 0.1% BSA, adjusted to a density of 1×10⁶ cells/mL, and then incubated with the corresponding antibody for 1 h in the dark. After washing again with D-PBS containing 0.1% BSA, the cell suspension was centrifuged at 1000 rpm for 5 min and the supernatant was discarded. Finally, the labeled cells were analyzed by flow cytometry (BD, Franklin Lakes, NJ, USA) using the CellQuest software after fixation with 1% paraformaldehyde.

2.5. Senescence-Associated β-Galactosidase Staining. Cell senescence was detected using a β-galactosidase staining kit.
2.7. Cell Viability Assay. Cell viability was determined using the following formula:

\[
\text{Relative proliferation rate} = \frac{X \text{ group average OD} - \text{blank control group average OD}}{\text{control group average OD} - \text{blank control group average OD}} \times 100\%.
\]

Subsequently, the absorbance was measured using an appropriate amount of D-PBS, and the cell membrane was disrupted by freezing and thawing the cells repeatedly for protein extraction. Following this, the supernatant was collected and stored at -20°C. The subsequent steps were accomplished according to the manufacturer’s instructions.

2.6. Telomerase Activity Assay. Telomerase activity in cell extracts was measured using a telomerase detection kit (Elabscience, Wuhan, China). The cells were digested and collected using trypsin. Thereafter, the cell pellet was mixed with an appropriate amount of D-PBS, and the cell membrane was disrupted by freezing and thawing the cells repeatedly for protein extraction. Following this, the supernatant was collected and stored at -20°C.

2.7. Cell Viability Assay. Cell viability was determined using the MTT assay. The cells were mixed with the MTT solution with an appropriate amount of D-PBS, and the cell membrane was disrupted by freezing and thawing the cells repeatedly for protein extraction. Subsequently, the absorbance was measured at 570 nm. Relative inhibition rate can be calculated using the following formula: relative proliferation rate (X group) = \[
1 - \frac{X \text{ group average OD} - \text{blank control group average OD}}{\text{control group average OD} - \text{blank control group average OD}} \times 100\%.
\]

Subsequently, the absorbance was measured at 450 nm. Relative proliferation rate can be calculated using the following formula:

\[
\text{Relative proliferation rate} = \frac{X \text{ group average OD} - \text{blank control group average OD}}{\text{control group average OD} - \text{blank control group average OD}} \times 100\%.
\]

2.8. Intracellular Reactive Oxygen Species (ROS) Assay. An ROS detection kit (Beyotime, Shanghai, China) was used to measure the content of intracellular ROS. Cells were treated with 2,7-dichlorodihydrofluorescein diacetate (DCFH-DA) dilution solution after washing once with D-PBS and incubated for 30 min. The level of intracellular ROS was observed using the Nikon Eclipse Ti-S inverted fluorescence microscope (Nikon, Tokyo, Japan).

2.9. Osteogenic and Chondrogenic Differentiation of hAMSCs In Vitro. Approximately 2 × 10^4 cells were seeded in 12-well plates in LG-DMEM culture medium. After incubation for 24 h, the cells were treated with GA-D and replaced with osteoblast-conditioned medium (HG-DMEM (Gibco, New York, USA) with 10% FBS, 100 mmol/L dexamethasone (Sigma, SL, USA), 50 ng/mL vitamin C (Solarbio, Beijing, China), and 10 mmol/L β-glycerophosphate (Solarbio, Beijing, China)) and chondrocyte-conditioned medium (HG-DMEM with 10 ng/L transforming growth factor- β3 (Peprotech, NJ, USA), 1 × 10^{-7} mol/L dexamethasone, and 50 mg/L vitamin C) for 7 days, as described in previous reports [26, 28]. Osteogenic differentiation was preliminarily evaluated by alkaline phosphatase staining, and chondrogenesis was assessed by toluidine blue staining.

2.10. Cell Cycle Analysis. The cell cycle was detected using a DNA content quantitation assay kit (Solarbio, Beijing, China). Cells were collected and digested using trypsin and washed with D-PBS. The cells were fixed by the addition of prechilled 70% ethanol followed by overnight incubation. Thereafter, the cells were resuspended in RNase A and incubated at 37°C for 30 min. Subsequently, propidium iodide staining solution was added and incubated at 4°C for 30 min. Cell cycle distribution was measured using flow cytometry.

2.11. Quantitative Real-Time PCR Analysis. Total RNA from hAMSCs was extracted using RNAiso Plus (Takara, Dalian, China), and 1-2 µg of the total RNA was used for cDNA synthesis with the PrimeScript™ RT reagent kit (Takara, Dalian, China). Real-time PCR was performed and monitored using the SYBR® Premix Ex Taq II (Takara, Dalian, China) quantitative real-time PCR system, and β-actin was used as the loading control. The threshold cycle (Ct) value was used to calculate the relative expression of genes. The primers for all genes tested are listed in Table 1.

2.12. Western Blotting. Total protein of hAMSCs was extracted using the radioimmunoprecipitation assay (RIPA) lysis buffer (Solarbio, Beijing, China), and nuclear protein was extracted using a nuclear protein extraction kit (Solarbio, Beijing, China). Proteins were detected through SDS-PAGE and subsequently electrotransferred to polyvinylidene fluoride membranes. The antibodies used were as follows: anti-p21 antibody (Abcam, ab80633), anti-p16 antibody (Huabio, Hangzhou, China), anti-Nrf2 antibody (Abcam, ab62352), anti-PERK antibody (CST, 3192S), anti-p-PERK antibody (CST, 3179S), and anti-peroxidase III (PRDX3) antibody (Abcam, ab128953). Then, membranes were incubated with horseradish peroxidase conjugated-secondary antibody (ProteinTech, SA00001-2) for 2 h at room temperature. Finally, the membranes were developed using enhanced chemiluminescence (ECL) hypersensitive luminescent solution (Beyotime, Shanghai, China). The intensity of the protein bands was analyzed using ImageJ software.

2.13. Statistical Analysis. All experiments were performed at least three times, and the data were expressed as the mean ± standard error of the mean (SEM). The GraphPad Prism software was used to conduct Student’s t-test and analyze the data. p < 0.05 was considered statistically significant.

3. Results

3.1. Identification of hAMSCs and Establishment of a Senescent hAMSC Model. Similar to our previous studies [28, 29], the surface molecules of MSCs were highly expressed in hAMSCs, including CD105 (88.10%), CD73 (99.84%), and CD90 (98.48%). However, the expression of cell surface molecules of hematopoietic stem cells (0.12%), including CD34, CD11b, CD19, CD45, and HLD-AR, was not observed. Additionally, hAMSCs strongly expressed vimentin (a marker protein of MSCs) but did not express cytokeratin 19 (a marker protein of epithelial cells) (Supplementary Figure 1). These results indicate that hAMSCs conform to MSC accreditation, as recommended by the International Society for Cellular Therapy [30].

To verify the effects of oxidative stress on hAMSCs, H₂O₂ was used to treat them. The formation of the senescence
marker β-galactosidase and cell viability were detected by β-galactosidase staining and MTT assay, respectively. As shown in Figure 2(a), H2O2 induced significantly premature cellular senescence of hAMSCs, as indicated by decreased cell density and increased rate of SA-β-gal-positive cells, as compared to the control group. The rate of SA-β-gal-positive cells was increased to 54.33% ± 5.15% and 58.33% ± 1.24% after treatment with 200 μM H2O2 and 400 μM H2O2, respectively. Though no significant difference was observed between the two groups (Figure 2(b)), the MTT assay showed that 400 μM H2O2 was more toxic to cells than 200 μM H2O2 (Figure 2(c)).

3.2. GA-D Inhibits the Generation of β-Galactosidase. To investigate the effects of GA-D on senescence induced by oxidative stress in hAMSCs, GA-D was used to pretreat hAMSCs prior to H2O2 treatment. The generation of β-galactosidase was suppressed in a dose-dependent manner when 0.1 μM, 1.0 μM, and 10.0 μM GA-D were employed (Figure 3(a)), but 100 μM GA-D had no significant effect against the senescent cells (Figures 3(a) and 3(b)). The data showed that the proportion of the SA-β-gal-positive cell rate was reduced from 46.92% ± 0.74% to 39.72% ± 0.17%, 28.0% ± 2.9%, and 19.58% ± 3.5%, at the respective concentrations (Figure 3(b)). The inhibitory rate against the generation of β-galactosidase of 10 μM GA-D reached 58.3%. Moreover, GA-D at various concentrations ranging from 0.001 μM to 100.0 μM was nontoxic to normal hAMSCs (Figure 3(c)). Therefore, these results suggest that 10 μM GA-D exerts positive effects on inhibition of H2O2-induced cellular senescence in hAMSCs.

3.3. GA-D Prevents H2O2-Induced Premature Senescence of hAMSCs. To confirm the preventive effect of GA-D on H2O2-induced senescence of hAMSCs, we further detected the production of intracellular ROS, cell cycle arrest, telomerase activity, and other senescence indicators. The production of intracellular ROS (Figures 4(a) and 4(b)) was obviously inhibited by GA-D at 10 μM. Additionally, H2O2 caused cell cycle arrest in the G2/M phase from 11.84% ± 0.16% to 19.64% ± 0.28%, but GA-D partially rescued this change of cell cycle from 19.64% ± 0.28% to 15.11% ± 0.20% in the G2/M phase (Figures 4(c) and 4(d)). GA-D also significantly downregulated the relative expression of p21 and p16INK4a at the translational level in senescent cells (Figures 4(e)–4(g)). Interestingly, the production of telomerase was also increased with the addition of GA-D (Figure 4(h)). These results further indicated that GA-D could prevent H2O2-induced senescence of hAMSCs.

3.4. GA-D Prevents Senescence of hAMSCs by Activating the PERK/NRF2 Signaling Pathway. In the present study, we

### Table 1: Primer sequence of target genes.

| Gene       | Sequence (5′ → 3′) | GenBank ID       | Length of product (bp) |
|------------|-------------------|------------------|------------------------|
| 14-3-3ζ    | For: CCTGCATGAAGTCTGTAACGTGAG Rev: GACCTACGGGTCCTCACAACA | NM_001135702.1 | 100                    |
| PRDX3      | For: ACAGCCGTTGCAATGGAGAG Rev: AGCTGTGGAAATTCGTTAGCTT | NM_006793.4 | 152                    |
| SIRT1      | For: TACGCTTGCAGCTAACCTTGT | NM_001142498.1 | 160                    |
| SIRT2      | For: CAGGGACAGTACATTACCTG Rev: CAGGCTGTATGTTGAGGTTG | NM_001193286.1 | 162                    |
| SIRT3      | For: GACATTGGGCTGACGTGAT Rev: ACCACATGCAGCAGAACACTTC | NM_001017524.2 | 122                    |
| SIRT4      | For: GTCTTGGCTGACTTTGACGT Rev: CCAATGGAGGCTTTGAGCA | NM_012240.2 | 79                     |
| SIRT5      | For: GCCATACGCGAGTTGAGAC Rev: CAACTCACAAGAGGTCATGG | NM_001193267.2 | 157                    |
| SIRT6      | For: CCCACGGGATCTGCGACCAT Rev: CTCGACGCTTTGTCCTG | NM_001193285.2 | 194                    |
| SIRT7      | For: AGCCAAATACCTGTCGCTC Rev: AGCACTAAGGCTCTCTT | NM_006164.5 | 190                    |
| NRF2       | For: TGGGAGGTAGTGCGACAT Rev: AGCGCCGAAAGGATGTA | NM_006793.4 | 152                    |
| β-Catenin  | For: GGGATGGTGAGGTTAAGA Rev: GCTGTTGACAGGGAGAC | NM_001904.4 | 148                    |
| ERK1       | For: CTACAGCGAGTTGCAATACAT Rev: CAGCGATGCTGGATCTCCC | NM_002746.2 | 109                    |
| ERK2       | For: TCTGGGACGATATCAGGCC Rev: CTGGTGTAAATCTGAGCTT | NM_138957.3 | 134                    |
| β-Actin    | For: TGGCACCAGCACAAAGGAA Rev: CTAAAGTCTAGTCGCCCTAGA | NM_0011101.3 | 186                    |
analyzed the transcriptional levels of some key genes that played an important role in the senescence process through qRT-PCR. The transcriptional levels of all genes analyzed were significantly altered in senescent hAMSCs. However, only NRF2 and PRDX3 were dramatically upregulated after the treatment with GA-D (Figure 5(a)), and this change was also observed at the translational level (Figure 5(b)). Furthermore, it was observed that GA-D promoted intranuclear transfer of NRF2 in senescent hAMSCs (Figure 5(c)). Generally, PERK delays cell cycle arrest by activating NRF2 [31], thereby indicating that the PERK/NRF2 signaling pathway might involve cellular senescence. Therefore, it is questionable whether there is a link between PERK and NRF2 in the antisenescence process of GA-D. We examined the protein expression levels of PERK in this study. Consistent with our prediction, the protein expression levels of phosphorylated PERK (p-PERK) were downregulated in senescent hAMSCs; however, they were upregulated after GA-D pretreatment (Figure 5(e)). Additionally, we found that GSK2656157 not only inhibited the expression of p-PERK (Figure 5(e)) but also reduced the expression of NRF2 in the nucleus (n-NRF2) and total NRF2 (t-NRF2) proteins (Figures 5(c) and 5(d)). However, only the expression of NRF2 (t-NRF2 and n-NRF2) could be suppressed in the presence of ML385 (Figures 5(c) and 5(d)), and there was no evident effect on p-PERK (Figure 5(e)). None of the treatments affected the expression of PERK protein, except for the H2O2-induced model group (Figure 5(f)). These results suggest that GA-D might activate p-PERK signaling to promote intranuclear transfer of NRF2 and ultimately prevent the senescence of hAMSCs.

3.5. Antisenescence Effect of GA-D Is Attenuated after the Inhibition of the PERK/NRF2 Signals. To verify whether GA-D delays the oxidative stress-induced senescence of

![Figure 2: H2O2 treatment induced the premature cellular senescence of hAMSCs. (a) Cell density and SA-β-gal-positive cells. Scale bar = 200 μm. (b) Statistics of SA-β-gal-positive cell rates. (c) Effects of different H2O2 concentrations on hAMSC viability. Data are presented as mean ± SEM, n = 3, **p < 0.01.](image-url)
hAMSCs through the activation of the PERK/NRF2 signals, the production of ROS and β-galactosidase was detected in hAMSCs in the presence of GSK2656157 and/or ML385. As shown in Figures 6(a)–6(d), the ROS level significantly increased after the blockage of PERK/NRF2 signals (Figures 6(a) and 6(b)), along with an increase in the generation of β-galactosidase (Figures 6(c) and 6(d)). As previously described, the senescence of MSCs led to a decrease in the ability to differentiate into osteoblasts and chondrocytes [32]. Therefore, the osteoblastic and chondrogenic differentiation capabilities of hAMSCs were preliminarily detected through alkaline phosphatase staining and toluidine blue staining, respectively. The expression of alkaline phosphatase (Figure 6(e)) and glycosaminoglycan (Figure 6(f)) was significantly decreased in senescent hAMSCs compared to the normal group, indicating that the differentiation ability of senescent hAMSCs was markedly attenuated, and this change could be restored by GA-D in senescent hAMSCs. However, the antisenescent effect of GA-D was reversed after the addition of GSK2656257 and ML385 (Figures 6(e) and 6(f)). Therefore, these results suggest that the antisenescent effect of GA-D activates PERK/NRF2 signals in senescent hAMSCs.
Figure 4: Continued.
Stem cell aging is regarded as an important driver of organism aging. Adult stem cells are essential for maintaining tissue and organ homeostasis through repair and regeneration during life. For instance, adipose-derived mesenchymal stem cells derived from old horses exhibited a typical senescence phenotype and limited regenerative capacity [33]. Aging-associated phenotypes can be reversed in vivo through the induction of stem cell rejuvenation [34]. Supporting this notion, delayed senescence of stem cells might be a promising novel strategy for ameliorating organismal aging and treating aging-related diseases. Ganoderma lucidum is a well-known traditional Chinese tonic that promotes health and longevity in East Asian countries. Recent studies have demonstrated that the extracts and polysaccharides of Ganoderma lucidum have distinct roles in delaying aging and the treatment of aging-related diseases through a multitargeted mechanism [16–18]. Polysaccharides and triterpenoids are the two major bioactive components of Ganoderma lucidum. Furthermore, Ganoderma lucidum is regarded as a cellular factory that produces a diverse set of bioactive triterpenoid compounds. So far, more than 210 triterpenoid compounds with different chemical structures have been identified [19, 20]. To our knowledge, however, the antiaging bioactivity of Ganoderma lucidum-derived triterpenoid compounds remains unknown [35]. Among all natural small-molecule compounds produced from Ganoderma lucidum, only four novel ergosterol derivatives, ganoderasides A–D, were shown to prolong the replicative life span of yeast through the regulation of the aging-related gene UTH1 [16]. In the present study, the protective effect of GA-D, a Ganoderma lucidum-derived triterpenoid compound against oxidative stress-induced stem cell senescence, was reported for the first time. Furthermore, GA-D inhibited the generation of ROS and senescence-associated markers, such as β-galactosidase, p21, and p16INK4a, and enhanced telomerase activity through the activation of the PERK/NRF2 signaling pathway.

Based on recently reported data [35, 36], antiaging activity was observed for more than 200 natural small-molecule compounds that were produced in plants and fungi. Among these natural compounds, saponin compounds with a tetracyclic triterpenoid skeleton, such as ginsenosides Rgl1, Rb1, and Rg3 from Panax ginseng CA Mey and cycloastragenol from Astragalus propinquus Schischkin, showed potent antiaging effects as did oleanolic acid from Fructus ligustri Lucidi, a triterpenoid with a pentacyclic structure. To date, ganodermic acid A, a Ganoderma lucidum-derived triterpenoid compound, was found to enhance antioxidant enzyme activity, inhibit ROS production, and increase the mitochondrial membrane potential [37]. The increased ROS and loss of the mitochondrial membrane potential were important inductive factors of cellular senescence [38], implying that Ganoderma lucidum-derived triterpenoid compounds may have antiaging potential by inhibiting oxidative stress. Consistent with these findings, GA-D also showed potent antiaging activity in hAMSCs by inhibiting oxidative stress. Therefore, natural triterpenoid compounds and their derivatives containing triterpenoid skeletons might exert strong antiaging potentials. Our findings suggest that GA-D is the first antiaging tetracyclic triterpenoid compound. However, further investigation in vivo is needed to confirm its antiaging activity.

Oxidative stress is an important factor that causes cellular senescence and body aging, and high concentrations of ROS trigger DNA damage and lead to cellular senescence through the direct or indirect regulation of aging-related signaling pathways [39]. As described in previous studies [16–18], Ganoderma lucidum could exert antiaging effects by regulating oxidative stress. In the present study, a senescent hAMSC cell model subjected to oxidative stress was established in vitro using H2O2. While GA-D can evidently decrease the ROS, β-galactosidase, p21, and p16INK4a levels and increase the content of telomerase in senescent hAMSCs, the underlying mechanism of the antisenescence effect of GA-D remains unclear. The regulatory mechanisms of stem cell senescence are extremely complex. Previous studies have shown that various signals, such as Keap1/NRF2 [40], Wnt/β-catenin [41], and ERK1/2 [42], are closely related to stem cell senescence. Sirtuins, which are metabolic sensors,
Figure 5: Continued.
are recognized as a link between the metabolic signaling and senescence. The life span-extending effects of dietary restriction involved the activation of sirtuins, and members of the sirtuin family, including SIRT1 [43], SIRT3 [44], and SIRT6 [45], could delay the senescence of stem cells by regulating oxidative stress. 14-3-3ζ, as a possible binding site of GA-D [46], could reduce the production of ROS by increasing cytoprotective target genes.

The PERK/NRF2 antioxidant signaling pathway plays an important role in oxidative stress and ER stress [54]. Oxidative and ER stresses are triggered by the accumulation of ROS in cells. Subsequently, PERK and NRF2 are activated to enhance the expression of antioxidant enzymes and detoxification enzymes that restore redox homeostasis [55]. Moreover, activation of PERK also enhances cell resistance to oxidative stress by promoting nuclear transfer of NRF2 [56]. Therefore, we speculated that the PERK signal might be activated in senescent hAMSCs after pretreatment with GA-D. Similar to the expression of NRF2 at the translational level, the subsequent detection showed that GA-D gave rise to the upregulation of PERK, especially at the phosphorylation level of PERK. Furthermore, when the specific inhibitors, including GSK2656157 for PERK signaling and ML385 for NRF2 signaling, were employed, the corresponding signaling not only was blocked but also showed mutual inhibiting effect of GA-D implicates the occurrence of UPR. PERK autophosphorylation activates the protein response (UPR), a cell protection program, is initiated to reduce the production of misfolded proteins and to finally extend the life span [58]. PERK is a key protein that mediates the occurrence of UPR.
Figure 6: Antisenescence of GA-D was attenuated after inhibition of the PERK/NRF2 signaling pathway. (a, b) Intracellular ROS levels after inhibition of PERK and NRF2 expression. Scale bar = 200 μm. (c, d) Changes of β-galactosidase expression in hAMSCs after inhibition of PERK and NRF2 expression. Scale bar = 200 μm. Data are presented as mean ± SEM, n = 3, **p < 0.01. (e) The ability of hAMSCs to differentiate into osteoblasts was analyzed by alkaline phosphatase staining. Scale bar = 200 μm. (f) The capability of hAMSCs to differentiate into chondrocytes was detected by toluidine blue staining. GA-D: ganoderic acid D. Scale bar = 200 μm.
the eukaryotic promoter, eukaryotic initiation factor 2α (eIF2α), which reduces the initiation of translation of most proteins in cells, thereby reducing the ER load. In the present study, it appears that H2O2 induces oxidative stress and ER stress in hAMSCs and finally induces cell senescence by increasing the accumulation of intracellular ROS and misfolded proteins. After GA-D pretreatment, the phosphorylation level of PERK was upregulated (Figure 5(e)), which might have mediated the occurrence of UPR to reduce ER stress. Meanwhile, p-PERK promoted the expression of NRF2 and nuclear transfer (Figures 5(c) and 5(d)) and finally triggered an increase in the transcription of downstream genes of NRF2, including PRDX3, HO-1, and quinine oxidoreductase 1 (NQO1) [53]. Consequently, hAMSC senescence was mitigated by decreased accumulation of ROS. This effect can be reversed by using specific inhibitors of PERK and/or NRF2; however, further investigation is required. Moreover, the mechanism underlying the delay in senescence of hAMSCs by GA-D remains unclear. The results of ligand-protein inverse-docking (INVDOCK) analysis suggested that GA-D could bind six isoforms of the 14-3-3 protein family, annexin A5, and aminopeptidase B [46]. Therefore, we speculate that GA-D may bind to one of these receptors for its functions; however, this needs to be further explored. Based on the above-mentioned analysis, a schematic diagram presenting the possible antisenescence mechanism of GA-D in hAMSCs can be speculated, as shown in Figure 7. To summarize, we report for the first time that natural triterpenoid compounds attenuate stem cell senescence in vitro through the activation of the PERK/NRF2 signals, and NRF2 is traceable as a key gene in the antisenescence effect of GA-D. These findings offer a new candidate agent for the prevention of aging-related diseases. However, due to the complexity of the signal regulation pathway, further studies on the signaling regulatory network of GA-D in senescent hAMSCs as well as its therapeutic effect on aging model in vivo are needed.

5. Conclusion

The present study reveals for the first time that *Ganoderma lucidum*-derived triterpenoid GA-D may exhibit potent antisenescence effects against H2O2-induced premature senescence of hAMSCs through the activation of PERK/NRF2 signals. These findings not only provide a deeper understanding of the antiaging effects of *Ganoderma* triterpenoid compounds but also provide a new theoretical basis for the development of antiaging supplements and the prevention of aging-related diseases on the basis of stem cell theory of aging.

Data Availability

The data used to support the findings of this study are all presented in the Results section and are also available from the corresponding author (jhxiao@zmu.edu.cn, or jianhuixiao@126.com).

Additional Points

**Highlights.** Ganoderic acid D retards the senescence of human amniotic mesenchymal stem cell. Ganoderic acid D exhibits potent protective effect against H2O2-induced premature senescence of hAMSCs without inducing cytotoxic side-effects. Ganoderic acid D retards hAMSC senescence through the activation of the PERK/NRF2 signaling pathway. Ganoderic acid D may act as a promising candidate for the antiaging
agent discovery. Ganoderic acid D has been reported as the first antiaging tetracyclic triterpenoid compound.

**Conflicts of Interest**

The authors confirm that there are no conflicts of interest.

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**Supplementary Materials**

Supplementary Figure 1: identification of hAMSCs. (Supplementary Materials)

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