Histopathological Osteomyelitis Evaluation Score (HOES) – an innovative approach to histopathological diagnostics and scoring of osteomyelitis

Abstract

Background: Treatment and diagnosis of osteomyelitis are still a challenging problem for surgeons, microbiologists and histopathologists. A direct microbiological detection of bacteria in tissues is still gold standard, but it is not always successful for example in chronic osteomyelitis and/or when an antibiotic treatment has already been started or in cases of low virulent bacteria. The goal of this study was to define diagnostic criteria of osteomyelitis, the inflammatory regression of osteomyelitis (“osteomyelitis score”) under specific therapy by the correlation of histopathological and microbiological and clinical standard tests.

Methods: In this retrospective analysis patients with medical history and clinically clear signs of bacterial infection and osteomyelitis underwent surgery between 01.01.2013 and 31.12.2012. Their formal consent was given. Tissue samples were taken during surgery according to defined criteria including surgical interventions. Histopathological diagnosis was carried out by conventional techniques based on defined criteria of bacterial infection in connective tissue, peri-implant membrane and bone. These results were carried out in tables by numbers representing the histopathological criteria of acute osteomyelitis (A1 to A3) as well as the chronic criteria (C1 and C2) in a semiquantitative way (scale 0 to 3). On the other hand a notational, graduated histopathological report was performed.

Preoperative clinical diagnosis, perioperative macroscopic diagnosis, histopathological and microbiological findings were correlated.

Results: Histopathological samples of 52 surgical interventions based on the preoperative diagnosis “osteomyelitis” (AOM, ECOM or COM) were included. 37 times preoperatively signs of a chronic osteomyelitis (COM), 10 times preoperatively acute osteomyelitis (AOM) was diagnosed. Another 5 patients were preoperatively diagnosed as acute exacerbated osteomyelitis (ECOM). The correlation of the histopathological infection including the inflammatory activity and microbiological detection of bacteria was 57%. The correlation between preoperative diagnosis and histopathological findings was 68%.

Conclusion: The relatively small 68% correlation between clinical preoperative and histopathological diagnosis and 57% correlation between preoperative clinical diagnosis and microbiological findings indicates:

• Clinical findings are not sufficient for the diagnosis “osteomyelitis”.
• Clinical findings are not sufficient for the differentiation between AOM, ECOM and COM.
• Histopathological analysis is the critical factor for the diagnosis (“osteomyelitis”) and differential diagnosis (AOM vs. COM).
• Histopathological analysis represents the basis for further treatment.
• HOES facilitates the classification of the histopathological findings.

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• HOES is a sufficient tool for the treating physician in order to define the further treatment.

**Keywords:** osteomyelitis, histopathology, microbiology, HOES

**Zusammenfassung**

**Grundlegende Überlegung:** Diagnose und Therapie der Osteomyelitis fordern auch heute Chirurgen, Mikrobiologen und Pathologen gleichermaßen. Der direkte mikrobiologische Nachweis des krankheitsverursachenden Erregers stellt einen „Gold Standard“ in der Diagnostik der Knocheninfektion dar. Leider gelingt der Keimnachweis nicht in allen Fällen, speziell bei chronischen Krankheitsverläufen, laufender Antibiotikatherapie oder im Falle der „low grade Infektion“. Die histopathologische Analyse ist insofern eine Condicio sine qua non. Nur anhand dieser Ergebnisse lässt sich zweifelsfrei das Vorliegen einer Osteomyelitis detektieren und eine Aussage zu ihrer Akuität machen. Ziel dieser Studie ist die Vorstellung eines standardisierten histopathologischen Scores, anhand dessen analog zum TNM-System bei Tumorerkrankungen eine valide Kartierung einer Osteomyelitis möglich ist. Weiterhin wurde die Korrelation zwischen histopathologischen Ergebnissen und der klinischen Diagnose ebenso wie dem positiven Keimnachweis überprüft.

**Methode:** In einer retrospektiven Analyse wurden die histopathologischen und mikrobiologischen Befunde von Patienten mit dem eindeutigen klinischen Symptomen einer Osteomyelitis untersucht. Alle in die Studie eingeschlossenen Patienten wurden zwischen dem 01.01.2013 und dem 31.12.2013 operiert. Sämtliche Gewebsproben wurden während der operativen Eingriffe gewonnen. Die histologischen Untersuchungen basierten auf den Standardtechniken für bakterielle Infektionen im Bindegewebe, perimplantär und im Knochen. Die Ergebnisse wurden erfasst:

1. in einer tabellarischen Form durch Zahlen, welche die Ausprägung von akuten (A1 bis A3) und chronischen (C1 und C2) Osteomyelitis-Kriterien semiquantitativ (Scala 0–3) in einer getrennten Form für akute und chronische Veränderungen darstellt (Histopathologischer Osteomyelitis-Evaluationsscore),
2. in einer schriftlichen, abgestuften Form, welche sich durch die Summation der tabellarischen Werte ergibt.

Die präoperative und die perioperative Diagnose, das histologische Ergebnis und die Mikrobiologie wurden hinsichtlich ihrer Übereinstimmung korreliert (dabei war nicht die Keimtypisierung, sondern der Keimnachweis an sich relevant).

**Ergebnisse:** 52 chirurgische Proben wurden ausgewertet. Sie alle stammten von Patienten mit der präoperativen Diagnose „Osteomyelitis“ (akute Osteomyelitis = AOM; akute Exazerbation einer chronischen Osteomyelitis = ECOM; chronische Osteomyelitis = COM). Es fanden sich: COM n=37, AOM n=10, ECOM n=5. Die Korrelation zwischen dem histopathologischen Bild inklusive der inflammatorischen Reaktion und einem positiven ErregerNachweis betrug 57%. Die Korrelation zwischen der präoperativen Diagnose und der histologischen Analyse betrug 68%.

**Schlussfolgerung:** Die relative geringe Übereinstimmung von präoperativer Diagnose, histopathologischem Ergebnis und der Mikrobiologie legt Folgendes nahe:

• Die klinische Vermutung allein ist nicht ausreichend für die Diagnose „Osteomyelitis“.
• Die klinische Vermutung allein ist nicht ausreichend zur Differenzierung zwischen AOM, ECOM und COM.
Die histopathologische Analyse ist das entscheidende Kriterium. Einerseits für die Diagnose „Osteomyelitis“ an sich und andererseits für deren Akuțität.

Die histopathologische Analyse ist die Basis für die Therapie.

HOES ist ein brauchbares Instrument zur standardisierten Kartierung der histopathologischen Ergebnisse.

Schlüsselwörter: Osteomyelitis, Histopathologie, Mikrobiologie, HOES

Introduction

Analogously to the majority of diseases, in the case of osteomyelitis also the physician performing treatment is faced with the problem of arriving at his (objective or objectifiable) diagnosis with the aid of analysis of specific, possibly pathognomonic symptoms of the (subjective) suspected existence of this entity. That is to say the syndrome (bone infection) is initially defined from the most precise assignment of individual symptoms. The diagnosis is then made with the aid of specific diagnostic procedures.

Problems of clinical diagnostics/the diagnostic dilemma

Bone infections and especially their chronic forms are often characterised by the absence of clear clinical signs. A pathognomonic (or pathognostic) symptom is lacking. Instead there is a mixture of various symptoms which, if interpreted correctly, can lead to the diagnosis of bone infection.

The extent of this problem manifests itself clearly in the fact that in 2010 Schmidt et al. developed an osteitis definition score by which the probability of the existence of osteitis/osteomyelitis can be estimated by evaluation of various symptoms occurring in the context of the disease [1].

According to this, the diagnosis of a bone infection is based on interpretation of the following factors [1]:

1. Case history (with respect to local findings and systemic risk factors)
2. Clinical findings
3. Laboratory findings (here the absolute values in particular, but also the course of specific infection parameters such as e.g. the leukocyte count or the C-reactive protein are relevant)
4. Imaging diagnostics (sonography, projection radiography, CT, MRI, nuclear medicine and hybrid methods)
5. Pathogen diagnostics
6. Cellular analysis

In practice, the items listed under 1 to 4 lead to “tentative diagnosis of osteomyelitis” (syndrome).

For years (decades) the basis of treatment of bone infection has been surgical cleansing of the infection [2], [3], [4]. In this respect the indication for surgery is deduced from items 1 to 4. In view of this circumstance, the analysis of these individual factors must necessarily result in the following:

- assessment of the acuteness of the infection
- preoperative planimetry especially with respect to the required extent of the resection of the bone and the surrounding soft tissue

The histological-microbiological analysis of the tissue samples obtained intraoperatively from the so-called “representative locations” should accordingly lead to

- objectifying of the bone infection and therefore to the diagnosis “osteomyelitis”.

On the basis of the “Schmidt 100% rule” and analogously to oncosurgery, only complete surgical removal of the infection focus leads to calming of the infection [5]. In this respect preoperative planning of the surgery as well as the intraoperative procedure, that is to say the quality of the surgical cleansing of the focus, depend critically on the

- preoperative and intraoperative diagnosis of the spread of the infection
- intraoperative surgical radicality.

Considering items 1 to 4 of the above list (syndrome), it is found that these are potentially associated with a number of possible errors which may lead to an assessment which in reality is inappropriate. Examples of these are:

- Subjective (incorrect) description of the disease history by the patient. Incorrect preliminary assessment of the disease (doctor’s notes).
- Misinterpretation of the existing clinical symptoms.
- Atypical courses and patterns in the laboratory parameters.
- Blurring in the separation of healthy from infected tissue inherent in the method of preoperative imaging.
- Intraoperative assessment on the basis of macroscopic findings is subjective and depends on the surgeon’s experience [6], [7].
- Lack of a standardised, valid, reproducible evaluation and diagnosis of the histopathological samples obtained. Instead the findings are often presented descriptively. This makes it difficult for the clinician to manage the histopathological results.
Problems of objectifiable parameters (obtained perioperatively): problems of making a diagnosis

Microbiological diagnostics

Problem 1
Infections of the skeletal system are as a rule characterised by a low pathogen count in samples. In the case of infections associated with foreign bodies (enclosed osteosyntheses) in particular, demarcation from contamination by the skin flora is of prime importance when evaluating the microbiological results [8].

Problem 2
Detection of the pathogen may take up to 14 days, depending on the quantity. If a calculated antibiotic treatment is necessary, for example, this can lead to an overdosed or incorrect treatment. There is also the danger of a shift in selection pressure and therefore an opportunistic infection. Equally, the side effects of such an incorrect or overdosed treatment should not be ignored.

Problem 3
Due to sensitive PCR-based detection methods, sometimes also only after concentration, pathogens of low virulence can be detected nowadays, the pathogenicity and clinical relevance of which may remain in doubt.

Histopathological diagnostics

The histopathological criteria for the soft tissue, peri-implant infection and for bacterial synovialitis and their differential diagnoses have been laid down [9], [10], [11]. Diagnostic criteria for osteomyelitis, however, are non-existent or only orientating [12]. The facts presented in this reference show:

a) That solely on the basis of the histopathological assignment
   • the correct (that is to say objectifiable) diagnosis
   and
   • a conclusion regarding the extent of the infection can be made.

It follows from this that the histopathological evaluation is a “conditio sine qua non” for making the diagnosis and deciding on the treatment (“must have”).

b) That the microbiological analysis is helpful but is associated with an unacceptably high rate of falsely negative results (up to 30% negative pathogen detection with otherwise proven osteomyelitis).

It can accordingly be deduced that the quality of the histopathological analysis and its standardised, reproducible and valid presentation is preferably the basis for the above state of affairs. At the present time there is no such system or such score specifically for osteomyelitis. The HOES presented in this study pursues this approach.

Histopathological diagnostic criteria for bacterial infections in bradytrophic tissue

Diagnostics: Tissue reaction patterns caused by the pathogen

Histopathological diagnostics of bacterial infections is generally to be regarded as diagnostics which supplement microbiology. These primarily are performed by evaluating the tissue reaction pattern caused by the pathogen (so-called pathological infection substrate) and are therefore an indirect form of infection diagnostics [12], [13], [14], [15], [16]. These diagnostics are based on evaluation of the leukocyte infiltration pattern and the tissue changes in connective tissue and bone tissue. Since focal infiltration patterns, especially small granulocyte accumulations, are not necessarily caused by an infection, this finding should also be evaluated in a clinical and microbiological context.

Neutrophilic granulocyte detection

Neutrophilic granulocyte detection by means of HE staining, PAS reaction, chloroacetate esterase staining and immunohistochemical CD15 detection stand at the centre of histopathological infection diagnostics (detection of the pathological infection substrate) of acute infectious non-specific infections in particular. The identification and quantification of immunohistochemically detected neutrophilic granulocytes (CD15) are to be regarded as valid methods of bacterial infection diagnostics [13], [14], [15], [16]. Enzyme histochemical staining in particular is subject to qualitative variations, although also depending on the staining procedures and the decalcification time, so that an automated, standardised immunohistochemical detection of the specific antigen CD15 should be given preference for neutrophilic granulocyte detection [17]. An immunohistochemical detection of CD68 for detection of epitheloid cells and macrophages may be necessary for investigations involving granulomatous
osteomyelitis (e.g.: brucellosis, mycobacterial infections, mycoses, parasitoses).

**Direct pathogen typing**

Direct pathogen typing by enzyme histochemical staining is possible to only a limited extent in orthopaedic pathology in particular. Exceptions exist here in specific infection, fungal infection, mycobacterial infection, mycoses and parasitoses (e.g.: osseous echinococcosis). The advantage of histopathological infection diagnostics lies in a timely evaluation of tissue samples, especially in the case of infections with a so-called minimal pathogen quantity (“low-grade infections”). Falsely positive results (“contamination of the tissue sample”) can be largely ruled out histopathologically since the evaluation of the reaction pattern which is a manifestation and consequence of the so-called pathogen-host reaction takes place.

**Immunohistochemistry and PCR-based methods**

Immunohistochemistry and PCR-based methods complete the repertoire of methods of histopathological infection diagnostics [18]. These methods allow direct identification of the pathogen. The sensitivity of PCR-based tissue analyses is adversely affected by fixing in formalin, native material being associated with a higher sensitivity and specificity. Microbiological infection diagnostics is consequently always obligatory.

**Histopathological osteomyelitis diagnostics**

**Patterns of acute osteomyelitis**

1. **Osseous changes**: Osteonecroses: Bone trabeculae with visually empty osteocyte cavities are detectable as a criterion for necrotic bone tissue especially with EDTA decalcification. The bone trabeculae have irregular contours and are fragmented. They may be fractured and completely necrotic (so-called bone sequester). There are intramedullary granulocyte infiltrates and fibrin exudates. In bone tissue with a haemopoietic function (e.g.: axial skeleton) there is a reduced or complete lack of haemopoiesis.

2. **Soft tissue changes**: Soft tissue necroses: Criteria for soft tissue necroses are apoptoses, a tissue eosinophilia, fibrin exudations and a confining texture of the tissue.

3. **Inflammatory infiltrate pattern**: Neutrophilic granulocyte infiltrate: Diffuse and grouped deposits (so-called microabscesses, ≥5 granulocytes) of segmented neutrophilic granulocytes in the usually highly oedematous medullary spaces. The neutrophilic granulocytes are PAS cytoplasmic, coarsely granular positive and display a plumped, pyknotic chromatin texture. (Granulocyte apoptosis with pathogen phagocytosis and NETosis). Immunohistochemically there is a specific intensive, coarsely granular, predominantly cytoplasmic CD15 positivity. Osteoclasts are also detectable alongside neutrophilic granulocytes on the irregular trabecular surface.

**Patterns of chronic osteomyelitis**

1. **Osseous changes**: Bone neogenesis: Spongy osseous tissue with reactive network bone neogenesis (POL detection of irregularly running fibrils), the bone surface is bordered by osteoblasts. Medullary space fibrosis with ectatic sinus. The medullary space tissue shows fibrosing with granulation tissue formation. The infiltrate consists of macrophages, lymphocytes, plasma cells and a few neutrophilic granulocytes.

2. **Soft tissue changes**: There is fibrosing with granulation tissue formation, the infiltrate consists of macrophages, lymphocytes, plasma cells and a few neutrophilic granulocytes.

3. **Inflammatory infiltrate pattern**: Lymphocyte/macrophage/plasma cell infiltrate: In the highly fibrosed medullary spaces there is a lymphocyte- and macrophage-rich, sometimes also plasma cell-rich, sometimes focal, sometimes inflammatory infiltration with a few neutrophilic granulocytes.

**Structure of the Histopathological Osteomyelitis Evaluation Score (HOES) for standardised recording and documentation of findings**

The HOES is presented in two forms:

1. In a tabular form by numerical values representing the degree of acute (A1 to A3) and chronic (C1 and C2) osteomyelitis criteria semiquantitatively (scale 0–3) in a separate form for acute and chronic changes.

2. In a written, graduated form showing the summation of the tabular values.

**HOES: Tabular form**

Graduated semiquantitative (0, 1, 2 and 3) and an additive evaluation of the criteria for the acute (A1 to A3) and chronic osteomyelitis (C1 and C2) results in the HOES. The evaluation is performed in a semiquantitative mode, and therefore in a graduated, semiquantitative and reproducible manner usual for diagnostic histopathological scores. The semiquantitative evaluation mode comprises: non-existent, mild, moderate and severe. In numbers: non-existent = 0, mild = 1, moderate = 2, severe = 3. Mild, moderate and severe are based on a semiquantitative evaluation of area, since evaluations in a three-part step are readily reproducible: non-existent = 0. Mild (= 1) one third of the section area shows these changes. Moderate (= 2) two thirds of the section area show these changes. Severe (= 3) three thirds of the section area, that is to say the entire section area, show these changes.
HOES human osteomyelitis score (tabular form):

- A1 Osseonecrosis 0/1/2/3 (...)
- A2 Soft tissue necrosis 0/1/2/3 (...)
- A3 Granulocyte infiltrate 0/1/2/3 (...)
- C1 Bone neogenesis/fibrosis 0/1/2/3 (...)
- C2 Lymphocyte/macrophage infiltrate 0/1/2/3 (...)

HOES: Written graduated form

This semiquantitative and numerical evaluation of the criteria for acute and chronic osteomyelitis results in the HOES in the written form (Figure 1).

- I Signs of an acute osteomyelitis
- II Signs of a chronically florid (that is to say active) osteomyelitis
- III Signs of a chronic osteomyelitis
- IV Signs of a subsided (calmed) osteomyelitis
- V No indication of osteomyelitis

Assignment of HOES score values and the written form of evaluation

- Sum of A1 to A3: ≥4 = signs of an acute osteomyelitis
- Sum of A1 to A3 and C1 to C2: ≥6 = signs of a chronically florid (that is to say active) osteomyelitis
- Sum of C1 to C2: ≥4 = signs of a chronic osteomyelitis
- Sum of C1 to C2: ≤4 = signs of a subsided (calmed) osteomyelitis
- Sum of C1 to C2: ≤1 = no indication of osteomyelitis

Aim of the study

a. To define criteria for osteomyelitis and the spectrum of osteomyelitis and to establish them as a graduated evaluation mode (HOES: Histopathological Osteomyelitis Evaluation Score).

b. To analyse the correlation between the preoperative estimation of the syndrome osteomyelitis and the histopathological diagnosis according to the HOES.

c. To analyse the correlation between the histopathological diagnosis according to the HOES and the microbiological processing.

Material and method

In a study over a period of 12 months (01.01.2013 to 31.12.2013) the histopathological intraoperative findings of interventions with the preoperative diagnosis “acute or chronic osteomyelitis” were analysed according to the above HOES criteria.

For this retrospective analysis the formal agreement of the patients was obtained and the Helsinki criteria were adhered to. The evaluation was encoded and anonymous. The inclusion criterion was high suspicion from clinical, laboratory chemistry and imaging results of osteomyelitis originating either from the bone itself or from a joint infection.

In the first step the result of the histopathological analysis was compared with the preoperative diagnosis and the agreement was correlated. In addition, agreement with the microbiological pathogen detection was correlated (Table 1).

Histopathological processing of the tissue samples

The cutting of tissue samples for diagnostics, the processing of the samples (tissue sample processing, cutting of resected tissue, the decalcifying, histochemical and immunohistochemical methods) and archiving of the samples are performed in a certified and accredited framework (quality standard according to DIN EN ISO/IEC 17020).

Cutting of osseous tissue samples, decalcification, enzyme histochemistry and histochemistry

Cutting of osseous tissue samples

Soft tissue fraction and osseous tissue fraction: After fixing in buffered formalin (4%), tissue samples were cut representatively. Osseous tissue is separated in principle from adhering soft tissue and embedded separately, so that a soft tissue fraction (1.1) and an osseous tissue fraction (1.2) result.

Osseous tissue fraction:
- Fragmented osseous tissue samples
  - Fragmented osseous tissue samples with a diameter of less than 3 cm are additionally cut with a bone knife, fragmented mechanically and embedded completely (up to about 4 capsules).
  - Continuous osseous tissue samples (resected osseous tissue)

Continuous osseous tissue samples in the sense of resected osseous tissue with a diameter of more than 3 cm are lamellated with a bone saw (bone lamella width about one cm) and embedded according to the investigation (up to about 9 capsules). The different consistencies of corticalis and spongiosa are taken into account in the decalcification times. In the case of labelled resected osseous tissue, a so-called orientated processing is carried out with the fractions: resected tissue edges (proximal, distal), osseous focal findings with subspecification (ventral, dorsal, medial, lateral), macroscopically normal osseous tissue apart from the focal findings and the sectionsurface (ventral, dorsal, medial, lateral). This extended mode of processing of resected osseous tissue includes macroscopic staining of relevant section surfaces by four different stains (yellow, black, green and blue). Imaging findings (e.g.: native x-ray, MRI) are helpful for precise orientation and so-called orientated processing or may be necessary in the event of complex resected osseous tissue.
Figure 1: Histopathological presentation of features of the HOES score by graduation (I–IV): I. Signs of an acute osteitis/osteomyelitis, II. Signs of a chronically florid, that is to say active osteitis/osteomyelitis, III. Signs of a chronic osteitis/osteomyelitis, IV. Signs of a subsided/calmed osteitis/osteomyelitis.

All images in the same magnification (original magnification about 150 x) in standard HE stain.
Table 1: Correlation of preoperative diagnosis, intraoperative diagnosis (based on the macroscopic findings) and HOES (1 = no signs of infection, 2 = acute infection, 3 = chronically florid infection, 4 = chronic infection, 5 = calmed infection)

Abbreviations: AOM = acute osteomyelitis; ECOM = exacerbated chronic osteomyelitis; COM = chronic osteomyelitis

| intraop. diagnosis | HOES  | pathogens intraop. | clin. correl. | mibi correl. |
|--------------------|-------|--------------------|---------------|--------------|
| COM acetabulum      | chronic | 4                  | 0             | 1            | 0            |
| COM OSG            | chronic florid | 3                  | 0             | 1            | 0            |
| COM femur          | infection calmed | 5                  | 0             | 1            | 1            |
| ECOM tarsus        | acute infection | 2                  | 1             | 1            | 1            |
| COM calcaneus      | infection calmed | 3                  | 0             | 0            | 0            |
| COM tibia, pseud. infection | chronic florid | 3                  | 0             | 1            | 0            |
| ECOM               | chronic florid | 4                  | 1             | 0            | 1            |
| AOM pelvis         | chronic florid | 4                  | 1             | 0            | 1            |
| COM talus          | chronic florid | 3                  | 1             | 1            | 1            |
| COM tibia          | chronic      | 4                  | 0             | 1            | 0            |
| COM tibia          | acute infection | 2                  | 1             | 1            | 1            |
| COM                | infection calmed | 5                  | 0             | 1            | 1            |
| AOM pelvis         | chronic florid | 3                  | 1             | 1            | 1            |
| AOM femur          | acute infection | 4                  | 1             | 0            | 1            |
| COM tibia          | chronic florid | 2                  | 1             | 0            | 1            |
| COM tibia          | infection calmed | 5                  | 0             | 1            | 1            |
| AOM femur          | acute infection | 5                  | 1             | 0            | 0            |
| AOM femur          | acute infection | 5                  | 1             | 0            | 0            |
| AOM pelvis         | acute infection | 2                  | 0             | 1            | 0            |
| AOM                | acute infection | 2                  | 1             | 1            | 1            |
| COM calcaneus      | chronic florid | 4                  | 1             | 0            | 1            |
| COM knee           | chronic florid | 5                  | 1             | 0            | 0            |
| COM                | chronic florid | 4                  | 0             | 0            | 0            |
| ECOM tibia         | chronic florid | 4                  | 1             | 0            | 1            |
| COM knee           | infection calmed | 5                  | 0             | 1            | 1            |
| COM tibia          | infection calmed | 5                  | 1             | 0            | 0            |
| COM OSG            | chronic florid | 4                  | 1             | 0            | 1            |
| COM tibia          | infection calmed | 5                  | 0             | 1            | 1            |
| COM OSG            | chronic      | 4                  | 1             | 1            | 1            |
| ECOM femur         | acute infection | 2                  | 1             | 1            | 1            |
| COM                | infection calmed | 5                  | 1             | 1            | 0            |
| ECOM               | acute infection | 5                  | 1             | 0            | 0            |
| COM tibia          | infection calmed | 5                  | 0             | 1            | 1            |
| COM femur          | infection calmed | 5                  | 1             | 1            | 0            |
| COM tibia          | chronic      | 4                  | 0             | 1            | 0            |
| COM                | chronic      | 4                  | 0             | 1            | 0            |
| COM tibia          | infection calmed | 4                  | 0             | 0            | 0            |
| COM tibia          | chronic      | 4                  | 1             | 1            | 1            |
| ECOM sternum       | acute infection | 2                  | 1             | 1            | 1            |
| COM tibia          | chronic florid | 3                  | 1             | 1            | 1            |
| AOM OSG            | acute infection | 2                  | 1             | 1            | 1            |
| COM pelvis         | chronic florid | 3                  | 0             | 1            | 0            |
| COM tibia          | chronic florid | 5                  | 1             | 0            | 0            |
| AOM tibia          | acute infection | 2                  | 1             | 1            | 1            |
| COM OSG            | acute infection | 2                  | 1             | 1            | 1            |
| COM tibia          | infection calmed | 5                  | 1             | 1            | 0            |
| COM OSG            | infection calmed | 5                  | 1             | 1            | 0            |
| AOM femur          | acute infection | 2                  | 1             | 1            | 1            |
| COM tibia          | chron. florid | 3                  | 1             | 1            | 1            |
| COM femur          | infection calmed | 5                  | 0             | 1            | 1            |
| COM calcaneus      | chronic florid | 3                  | 0             | 1            | 0            |
Decalcification

The decalcification was carried out by means of acid (hydrochloric acid) and/or with the chelating agent EDTA. The ratio of decalcification liquid volume to tissue sample volume was about 1:20. The reaction temperature was room temperature over an incubation time of about 1 to 2 days. The decalcification by means of EDTA was carried out in the same ratio of volumes, but over a longer incubation time (up to about 3–7 days), the consistency of the samples being checked daily.

Enzyme histochemistry

The microtomised sections with a section thickness of 1–3 µm were stained with haematoxylin and eosin (HE), and a periodic acid-Schiff (PAS) staining was additionally performed.

Immunohistochemistry

The immunohistochemical CD15, CD68 staining is carried out in a fully automated staining system (BenchmarkXT, IHC Slide Stainer of the Roche brand, Ventana Medical Solutions). The sections were first deparaffinised with xylene and an ethanol series. Cell conditioning was then first carried out using Cell-Conditioning 1 (CC1) at 95 °C for 8 minutes, followed by a mild cell conditioning for 30 minutes. The anti-CD15 antibody (clone MMA, Roche, Basle, Switzerland) – a monoclonal murine antibody (ready-to-use, according to Roche undiluted) – was used as the primary antibody for identification of neutrophilic granulocytes. The sections were incubated with the antibody for 32 minutes. DAB (3,3-diaminobenzidine; DAKO Denmark) was used as the chromogen for the reaction with the peroxidase. The endogenous peroxidase was blocked by prior addition of $H_2O_2$. So-called negative controls were produced by omitting the primary antibody.

Evaluation

The evaluation of the findings was based on the following assumptions and criteria:

1. The preoperative tentative diagnosis “osteomyelitis” (syndrome) ALWAYS exists. It is the prerequisite for establishing the indication for operation.
2. The intraoperative assessment results from the diagnosis documented in the operation report (if the indication is correctly established, it coincides with the preoperative tentative diagnosis).
3. The diagnoses from 1 and 2 are checked with the aid of the histopathological analysis. This is carried out by the HOES. The following accordingly emerges:
   - The histopathological analysis does NOT confirm the clinical assessment
   - The histopathological analysis confirms the clinical assessment
4. The histopathological evaluation is correlated with the microbiological.
5. A correlation of the “pathogen/histological osteomyelitis” group is additionally conducted according to the following criteria:
   - Acute osteomyelitis
   - Chronic osteomyelitis

An acute osteomyelitis was diagnosed in 10 cases, an acutely exacerbated chronic osteomyelitis in 5 cases and a chronic osteomyelitis in 37 cases. Pathogens were detected in the samples taken in 32 cases.

In the clinical diagnosis “acute osteomyelitis”, pathogens were detected in 9 cases (90%).

In the clinical diagnosis “acutely exacerbated chronic osteomyelitis”, this happened in 5 out of 5 cases (100%).

In the clinical diagnosis “chronic osteomyelitis”, the ratio of positive to negative detection of pathogens was 19 to 18 (51.4% to 48.6%).

No significant correlation was to be found between the HOES and the microbiological findings in the diagnosis "chronic osteomyelitis" (Table 2).

The correlation between the HOES stages and the detection of pathogens can be seen from Table 3.

The correlation between the preoperative diagnosis and a detection of pathogens was 57%.

The correlation between the preoperative diagnosis and histopathological analysis of the samples taken was 68%, i.e. the preoperative tentative diagnosis could be confirmed by the histopathological diagnosis in 68% of the cases.

The correlation between the preoperative diagnosis, the HOES and the microbiological result is shown in Table 4.

Discussion

Histopathological infection diagnostics is of considerable diagnostic value. For example, histopathology is a diagnostic constituent in peri-implant infection diagnostics [19]. Quantitative criteria for bacterial low-grade and high-grade infection are defined here [11], [13], [14], [16], [17].

The situation is the opposite for osteomyelitis diagnostics: Only orientating [12] or no diagnostic criteria have hitherto existed here, in particular no classification criteria exist for bacterial acute and chronic osteomyelitis. The aim of this work is therefore to develop a diagnostic score which reflects the disease of osteomyelitis in a precise manner by defined criteria and gives a reliable, reproducible diagnostic score.

There is good agreement between the microbiological infection diagnostics and the HOES score especially in the acute diseases, but in the evaluation of “signs of a calmed, subsided osteomyelitis” there is no good agreement with the microbiological diagnosis. This is possibly to be attributed to a sampling error or to the microbiol-
The definition of the HOES is the basis for future studies concerned specifically with treatment of bone infections which correlates with the HOES stages. The following questions will need to be answered in these:

**Table 2: Correlation between the preoperative diagnosis “chronic osteomyelitis”, the HOES and the microbiological result**

| Microbiological result | Detection of pathogens | No detection of pathogens |
|------------------------|------------------------|---------------------------|
| **HOES (clinical diagnosis COM)** |                         |                           |
| (2) acute infection    | n=4                    | 4                         | 5                         |
| (3) chron. florid      | n=8                    | 3                         | 7                         |
| (4) chronic            | n=13                   | 4                         | 7                         |
| (5) infection calmed   | n=12                   | 7                         | 7                         |

**Table 3: Correlation between HOES and detection of pathogens**

| Microbiological result | Detection of pathogens | No detection of pathogens |
|------------------------|------------------------|---------------------------|
| **HOES**               |                         |                           |
| (2) acute infection    | n=12                   | 2                         | 10                        |
| (3) chron. florid      | n=9                    | 5                         | 4                         |
| (4) chronic            | n=14                   | 6                         | 8                         |
| (5) infection calmed   | n=17                   | 7                         | 10                        |

**Table 4: Correlation between the HOES, the preoperative diagnosis and the microbiological result**

| Preoperative diagnosis | AOM | ECOM | COM | Detection of pathogens | No detection of pathogens |
|------------------------|-----|------|-----|------------------------|---------------------------|
| **HOES**               |     |      |     |                        |                           |
| (2) acute infection    | n=12| 5    | 3   | 4                      | 10                        | 2                         |
| (3) chron. florid      | n=9 | 1    | 8   | 4                      | 5                         |
| (4) chronic            | n=14| 2    | 10  | 8                      | 6                         |
| (5) infection calmed   | n=17| 2    | 14  | 10                     | 7                         |

gical and histological tissue samples not being taken analogously. Although the main task of the HOES score consists of evaluation of bacterial, non-specific infections and the evaluation of floridity and chronicity, histopathological infection diagnostics always allows specific osseous infections to be ruled out and in particular also differential diagnosis of chronic osteomyelitis from neoplastic diseases.

In the case of bone segment resected tissue, a further function of HOES can be seen in the evaluation of the absence of inflammation in the resected tissue margin status. In an analogous manner to bone tumour resection (R status), a conclusion can be drawn regarding the complete removal of osteomyelitis. Further studies will demonstrate whether this R0/R1 concept can also be applied clinically to osteomyelitis.

Overall, it is to be said that histopathological criteria for reproducible osteomyelitis diagnostics have been laid down for the first time by the HOES score. Correct diagnosis of musculoskeletal infections is always difficult and is based on analysis of widely differing factors [1].

These problems also become clear in our study. Preoperative diagnosis based on case history, clinical findings, laboratory tests etc. coincide only inadequately with the detection of pathogens, and in particular with the evaluation of histopathological findings. It is therefore not suitable as the basis for deciding on the subsequent procedure.

The known fact that detection of pathogens is often not to be achieved where osteomyelitis exists clinically also becomes clear in this study, especially for the preoperative diagnosis “chronic osteomyelitis” [9]. The distribution here between positive and negative microbiological results was almost 50/50 (51.4% to 48.6%). The situation is different for the acute diseases (AOM, ECOM). In the study described, a pathogen could be detected in a total of 14 out of 15 cases. These results illustrate that the disease present can be successfully assigned unambiguously only via precise histopathological investigation [10].

For the clinician, however, interpretation of the available findings is “a complete mystery” here. The HOES described here now makes a classification of the available findings into 5 groups possible (see above). By this classification of the osteomyelitis the histomorphological findings of the disease present become less blurred and can be better applied to the clinical circumstances. The decision about the subsequent procedure will therefore be made easier for the clinician in future.
At what stage is (further) surgical revision (not) necessary (specifically with regard to the “individual revision concept”)?

What influence does microbiological detection of pathogens have here? Is antibiotic treatment necessary?

Is the HOES stage (1) = no infection or (5) = infection cooled a conditio sine qua non for implantation of osteosynthesis material?

What influence does microbiological detection of pathogens have here? Is antibiotic treatment necessary?

Only when these questions can be clearly answered, treatment of bone infections can be standardised and gold standards and SOPs defined.

Notes

Competing interests

The authors declare that they have no competing interests.

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