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Introduction of a Nia proteinase cleavage site between the reporter gene and HC-Pro only partially restores the biological properties of GUS- or GFP-tagged LMV

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Abstract

Lettuce mosaic virus (LMV) isolates LMV-E and LMV-0 differ in their virulence on lettuce varieties carrying the mo12 resistance gene, which reduces viral accumulation and blocks the expression of symptoms after infection with avirulent isolates such as LMV-0. Previous work had indicated that reporter genes such as GUS or GFP affect the biological properties of recombinant LMV isolates in both susceptible and resistant lettuce varieties when fused to the N-terminus of the viral protein HC-Pro. The impact of the addition of a cleavage site for the Nia proteinase between the reporter gene and HC-Pro was evaluated, in an effort to recover the full spectrum of the biological properties of parental isolates. Symptoms, accumulation, cell-to-cell and long distance movement of the recombinant viruses containing the Nia cleavage site were studied in susceptible and mo12 lettuce varieties. Both LMV-0 and LMV-E recombinant viruses recovered the behaviour of their wild-type parent in susceptible plants upon addition of the Nia cleavage site. While the recombinant LMV-E modified in this way recovered the breaking properties of its wild-type counterpart in mo12 plants, similar modification of the LMV-0 derived recombinants failed to rescue a severe inhibition in systemic accumulation in mo12 plants, despite the fact that neither cell-to-cell movement nor phloem loading or unloading seemed to be severely affected.

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Keywords: Potyvirus, LMV; GFP; GUS; Resistance gene; Lettuce

1. Introduction

Lettuce mosaic virus (LMV, genus Potyvirus) is one of the most destructive viruses in lettuce (Lactuca sativa) crops all over the world (Dinant and Lot, 1992). The LMV genome consists of a positive-strand RNA of 10,080 nucleotides (Revers et al., 1997a). The RNA is polyadenylated at its 3′ end, linked to a viral protein (VPg) at its 5′ end, and encodes a single polyprotein that is processed by three virus-encoded proteinases (Dougherty et al., 1990; Reichman et al., 1992). Two alleles of a gene named mol in lettuce are associated with resistance to LMV in a recessive manner (Bannerman et al., 1969; Ryder, 1970). We have recently discovered that mol encodes the translation initiation factor eIF4E (Nicaise et al., 2003). LMV isolates differ in their ability to infect and produce symptoms in lettuce genotypes carrying mol1 or mol2 (Pink et al., 1992a; Bos et al., 1994; Revers et al., 1997b). Depending on the virus isolate/host cultivar combination, mol1 conveys either full resistance (absence of detectable virus accumulation) or tolerance (systemic virus accumulation but failure to induce symptoms). The two LMV isolates used in this study differ in their pathogenicity, resistance-breaking and seed transmission properties. LMV-E is a resistance-breaking isolate that induces severe symptoms on susceptible and resistant cultivars, and is not seed-borne (Pink et al., 1992b). LMV-0 is a seed-borne isolate that induces relatively milder symptoms on susceptible cultivars. In plants carrying mol2, LMV-0 is generally able to mount a systemic invasion with much reduced accumulation and symptoms failing to develop. In susceptible plants, LMV-0 and LMV-E induce different types of symptoms but accumulate to similar levels (Redondo et al., 2001).
2. Material and methods

2.1. Viral cDNAs

The construction and characterisation of pLMV-E-GFP/HC and pLMV-E-GUS/HC has been described previously (German-Retana et al., 2000). Similar constructs were prepared from an infectious cDNA of LMV-0 (Redondo et al., 2001). The plasmids pLMVclvHC, pLMV-GFPclvHC, and pLMV-GUSclvHC were obtained by inserting a double-stranded oligonucleotide encoding a consensus cleavage site recognised by the Nla viral proteinase (GDEVYHQ/I) into the SmaI site of the relevant pLMV-GUSHC or pLMV-GFPHC constructs.

2.2. Plant material and inoculation

The susceptible lettuce cultivar “Trocadéro” was routinely used for virus propagation. The cDNAs of all recombinant viruses were initially inoculated into Trocadéro plants as previously described (German-Retana et al., 2000). They were back-inoculated mechanically into plants of the susceptible varieties Trocadéro, Salinas, or the early flowering line 8720 M (Ryder, 1996), kindly provided by Dr. E. Ryder (USDA Salinas), or into the mo1-carrying variety Salinas 88. Salinas and Salinas 88 are near isogenic lines differing mainly in the mo1 gene (Ryder, 1991; Nicaise et al., 2003).

2.3. Grafting of lettuce plants

Salinas or Salinas 88 rootstocks were inoculated with the relevant recombinant virus 2 weeks prior to the grafting. Healthy etiolated stems of Salinas or of Salinas 88 were then grafted onto the rootstocks using inverted saddle grafts (Kains and McQuesten, 1960). Briefly, a V-shaped notch was made in the stem of the rootstock and a corresponding V-shaped wedge was cut in the stem of the scion to be grafted. Both fresh cuts were placed together and wrapped tightly. Grafted plants were kept in “mini green-houses” to prevent dehydration of the scion during recovery from grafting. After usually 9 days, the recovered plants, as shown by the survival of the scion, were then transferred to an insect-proof green-house with standard conditions (16 h day-light, 20–30 ◦C).

2.4. Detection of GUS and GFP expression

Histochemical GUS staining was performed following a protocol slightly modified from that of Jefferson (1987). Vacuum was used to infiltrate the GUS staining solution supplemented with 0.2% (v/v) Triton X-100 into leaf fragment or roots. K4[Fe(CN)6] and K3[Fe(CN)6] at 500 mM concentrations were added to the GUS buffer in order to avoid artefacts and to localise the blue precipitate only in the cytoplasm of the cells where the GUS activity is expressed (De Block and Debrouwer, 1992). The reaction was usually performed overnight at 37 ◦C. The on-line software image analysis “Image Tool for Windows” (University of Texas Health Science Center San Antonio) was used to measure and compare the area of the infection foci on inoculated leaves.

Expression of GFP at the whole plant level was visualised using a 100 W, hand-held, long-wave UV spot-light lamp (Model B-100; UV Products, Upland, CA). At the
tissue level. GFP expression was examined by fluorescence microscopy using an instrument equipped with GFP(R)-LP/HQ-FITC-LP Filters (Eclipse 800, Nikon, Tokyo, Japan) and a fluorescence stereomicroscope (MZ FLIII, Leica Microsystems, Heerburg, Switzerland) equipped with a filter with an excitation window at 470 ± 20 nm and an arrest window at 525 ± 25 nm.

2.5. Protein analysis and Western blotting

Leaf tissue was ground in electrophoresis loading buffer [125 mM Tris-HCl pH 6.8, 10% SDS, 25% β-mercaptoethanol] using a 1:5 (w/v) grinding ratio. Following boiling for 3 min, the homogenate was centrifuged at 20,000 × g for 10 min. Proteins were separated on 10%...
polyacrylamide–SDS gels and blotted onto nitrocellulose membranes. The membranes were probed with either polyclonal antibodies raised against GFP (Invitrogen, Carlsbad, CA) or with a monoclonal antibody raised against LMV HC-Pro protein (Roudet-Tavert et al., 2002), and anti-rabbit or anti-mouse IgGs coupled with alkaline phosphatase were used as secondary antibodies. Detection was finally achieved using an NBT–BCIP colorimetric reaction.

2.6. Quantification of viral accumulation

Viral concentrations were estimated by ELISA as described by Clark and Adams (1977), and the viral RNA was detected by dot– blot hybridisation with a dig-labeled in vitro transcribed cRNA probe as described by Brault et al. (1993).

3. Results

3.1. A Nia cleavage site introduced between the reporter gene and HC-Pro is efficiently utilised

A LMV consensus cleavage site (GDEVYHQ/SG, where the slash indicates the bond cleaved by the proteinase) was determined by alignment of the cleavage sites recognised by the Nia proteinase in the LMV sequence. A synthetic DNA fragment encoding this octapeptide was introduced between the GUS or GFP reporter gene and the HC-Pro of recombinants pLMV-GUSHC and pLMV-GPHC derived from either LMV-0 or LMV-E. The four new constructs were named pLMV-x-GUSclvHC and pLMV-x-GFPclvHC, respectively, where x indicates the parental virus (Fig. 1). In these viruses, two glycine residues absent from the non-recombinant virus are predicted near the N-terminus of HC-Pro after Nia cleavage. As a control, a recombinant in which the Nia site was added, without any reporter gene was also constructed and named LMV-0-clvHC (Fig. 1).

All recombinant viruses were infectious as shown by the development of disease after inoculation by particle bombardment of the susceptible lettuce variety "Trocadéro". Western blots of proteins extracted from Trocadéro leaves 15 days post-inoculation (dpi) demonstrated that the majority of the GFP-HC-Pro fusion protein was processed, as expected (Fig. 2A). While only the 80 kDa GFP-HC-Pro fusion protein was detected in the plants infected with LMV-0-GFPHC (lane 2), in extracts of plants infected with LMV-0-GFPclvHC (lane 1), HC-Pro was mostly present in a processed form which co-migrated with the HC-Pro detected in wild-type LMV-0 infected plants (lane 3). A low amount of GFP-HC-Pro fusion protein (estimated at less than 10% of the amount of free HC-Pro) was still detectable (lane 4). These results were confirmed by probing a similar blot with a GFP-specific antiserum (Fig. 2B). Similar results were obtained at 3, 4 and 7 weeks after inoculation, for both LMV-0 and LMV-E GFP derivatives, without any significant difference in the level of accumulation of free HC-Pro (data not shown). These results showed that the introduced Nia cleavage site was efficiently utilised, allowing the processing of about 90% of the fusion protein to the expected GFP and HC-Pro cleavage products.

3.2. The effect of the insertion of a Nia cleavage site on symptoms and viral accumulation depends on the viral isolate

It has previously been shown that GFP or GUS fusion of the LMV-E HC-Pro has negative effects on symptom severity and accumulation of the virus in systemically infected leaves of a susceptible variety (German-Retana et al., 2000). Susceptible plants inoculated with a similar construct derived from LMV-0 (LMV-0-GFPHC) developed attenuated symptoms as compared to wild-type LMV-0 (Fig. 3C). Introduction of the artificial Nia cleavage site between the GFP and the HC-Pro sequences in these constructs restored symptom severity to that of the wild-type parents (Fig. 3A and C). In particular, the stunting, leaf deformation and necrosis of young leaves, typical of LMV-E and abolished in LMV-E-GFPHC, were restored in LMV-E-GFPclvHC (Fig. 3B).

The accumulation of the GFP-tagged viruses in the susceptible lettuce variety salinas was estimated by enzyme-linked immunosorbent assay (ELISA), and compared with the accumulation of wild-type LMV at 15 dpi. As shown in Fig. 4A, no significant difference in accumulation was observed between LMV-0 and LMV-0-GFPclvHC or LMV-0-GFPHC. In the plant variety Trocadéro, comparable results were obtained at 35 and 40 dpi by dot-blot hybridization experiments (data not shown). Thus, the GFP-tagging
of HC-Pro, with or without an added Na cleavage site, had no effect on LMV-0 accumulation in susceptible lettuce hosts. In contrast, and as reported previously (German-Retana et al., 2000), the accumulation of LMV-E-GFPHC was significantly impaired at 15 dpi (Fig. 4B), reaching only 28% of the accumulation level of LMV-E. Introduction of the Na cleavage site restored accumulation of LMV-E-GFPclvHC to a level comparable to that of wild-type LMV-E in the susceptible variety (Fig. 4B).
Taken together, these results indicated that the two isolates were differentially affected in their accumulation in host plants by the fusion tagging of their HC-Pro, which induced a severe reduction of the accumulation of LMV-E, but apparently no significant effect on LMV-0. They demonstrated further that introduction of the NIa cleavage site in the tagged recombinants allowed the recovery of symptom severity and accumulation levels typical of wild-type isolates.

3.3. Introduction of an NIa cleavage site modifies the in planta distribution of tagged LMV-0 in susceptible lettuce varieties

GFP expression from LMV-E-GFPHC was still observed after prolonged propagation of the virus in the same host for many months (German-Retana et al., 2000). This was also the case for GFP-tagged viruses derived from LMV-0 (data not shown). We therefore tried to compare the accumulation of the LMV-0-derived recombinants in all organs and throughout the entire plant life cycle, including flowers and roots, using macroscopic and microscopic observation.

When the GFP-tagged LMV-0 derivatives were inoculated onto source leaves of the susceptible lettuce variety Trocadéro, circular fluorescent foci were apparent on the lamina within 4 dpi (data not shown). At 7 dpi, the infection foci on the inoculated leaves had slightly increased in size, and symptoms developed on systemically infected leaves. At this stage, the GFP fluorescence pattern was fully super-imposable onto the developing vein-clearing symptoms, the fluorescence being limited to the vicinity of veins (Fig. 3D). No differences could be observed up to this stage between LMV-0-GFPHC and LMV-0-GFPclvHC (Fig. 3D). However, a slight difference in the accumulation pattern of the two viruses was noted at later times (Fig. 3E). While LMV-0-GFPHC showed a mosaic of smallish GFP patches, LMV-0-GFPclvHC tended to accumulate in larger, more homogeneous patches. Although subtle, these general patterns of GFP expression and accumulation in systematically infected leaves were reproducible in a number of independent experiments.

GFP reporter activity was detected in the roots and in all floral parts of lettuce plants (early flowering line 8720 M. Ryder, 1996) infected with the LMV-0 tagged viruses, and especially in the teguments of non-mature seeds. However, differences between LMV-0-GFPHC and LMV-0-GFPclvHC localisation were again noted: only LMV-0-GFPclvHC was able to accumulate in the neck between the style and ovary of non-mature seeds as well as in pollen grains (Fig. 3F and G). In summary, the introduction of the NIa cleavage site between the GFP and the HC-Pro of LMV-0-GFPHC slightly modified the behaviour of the tagged virus in susceptible lettuce varieties and, in particular, allowed the recovery of the ability of the virus to infect some specific reproductive tissues such as pollen.

3.4. Behaviour of LMV-0 and LMV-E derived GFP-tagged recombinants in mo12 carrying lettuce

In lettuce varieties carrying the mo12 gene such as Salinas 88, LMV-E infection results in the appearance of a systemic mosaic at 20 dpi. LMV-0 does not induce symptoms although systemic virus accumulation can be detected albeit at a much lower level than that reached by LMV-E (Revers et al., 1997b; Redondo et al., 2001). However, both LMV-0-GFPHC and LMV-E-GFPHC (or equivalently GUS-tagged recombinants) have been shown to be generally unable to mount a systemic infection of mo12-containing varieties, in contrast to their wild-type parents (German-Retana et al., 2000; Candresse et al., 2002). We therefore investigated whether the introduction of the artificial NIa cleavage site would restore the wild-type biological behaviour of the tagged viruses in resistant plants. GFP fluorescence was monitored in upper non-inoculated leaves of Salinas 88 at 20 and 40 dpi. Unlike LMV-E-GFPHC, LMV-E-GFPclvHC was fully able to move long distances and systemically accumulate in Salinas 88 as shown by the detection of fluorescence and the presence of symptoms at both observation times, the symptoms being similar to those caused by wild-type LMV-E. Analysis of the virus progeny by RT-PCR confirmed the presence of LMV-E-GFPclvHC in these plants (data not shown). By contrast, no systemic accumulation of GFP could be observed using a hand-held

![Fig. 4. Accumulation of GFP-tagged LMV-0 and LMV-E in susceptible lettuce "Salinas". The accumulation of GFP-tagged virus was determined by ELISA in three to four individual plants and normalised to the mean value (∼100%) measured similarly for the wild-type virus from which the recombinant viruses was derived. The samples were collected from developmentally equivalent non-inoculated symptomatic leaves (level 4 from the top of the plant) at 15 dpi. Experiments were repeated at least three times. The bars indicate standard deviations.](image-url)
ceptible) and Salinas 88, compared in two near isogenic lettuce varieties, Salinas (susceptible) and resistant varieties (Fig. 5A and B). The surface of the infection foci at 8 dpi was, on average, two to three times larger on the susceptible variety than on the resistant one.

In an effort to identify at which level the systemically accumulated cell-to-cell movement of the tagged virus on both varieties. No significant difference in the size of the infection foci was found at 4 dpi by histochemical staining between the susceptible and resistant varieties (Fig. 5A and B). However, analysis of the size of the infection foci of LMV-0-GFPHC or GFP-tagged recombinants in Salinas 88 rootstocks was similar to those observed on the inoculated leaves of the susceptible Salinas 88. Data obtained at the early time point (4 dpi) further indicated that the cell-to-cell movement of the recombinants was not severely affected in the resistant plants as compared to the susceptible one.

3.5. Analysis of the mechanism(s) impairing systemic accumulation of GFP and GUS-tagged LMV-0 recombinants in Salinas 88

In an effort to identify at which level the systemically accumulated cell-to-cell movement of the tagged virus on both varieties. No significant difference in the size of the infection foci was found at 4 dpi by histochemical staining between the susceptible and the resistant varieties (Fig. 5A and B). Similar results were obtained using LMV-0-GFPHC or GFP-tagged recombinants and epi-fluorescence observation of the inoculated leaves (data not shown).

However, analysis of the size of the infection foci of GUS-tagged recombinants at a later time (8 dpi) revealed differences in the spread of the virus between susceptible and resistant varieties (Fig. 5A and B). The surface of the infection foci at 8 dpi was, on average, two to three times larger on the susceptible variety than on the resistant one. These results indicated that, irrespective of the presence of an artificial Na cleavage site, GFP- and GUS-tagged LMV-0 recombinants were able to replicate efficiently and to move cell-to-cell in the inoculated leaves of the mo12 cultivar Salinas 88. Data obtained at the early time point (4 dpi) further indicated that the cell-to-cell movement of the recombinants was not severely affected in the resistant plants as compared to the susceptible one.

3.6. Long distance movement of GFP and GUS-tagged LMV-0 is restricted in Salinas 88

To investigate the phloem loading and unloading of LMV-0-GFPCvHC, grafting experiments were performed. Uninfected Salinas scions were grafted onto Salinas 88 rootstocks which had been pre-inoculated with LMV-0-GFPCvHC 21 days earlier. Controls were composed of healthy Salinas scions grafted onto infected Salinas rootstocks. Thirty days after grafting, symptoms and detection of GFP in the leaves of the Salinas scions demonstrated that in both cases LMV-0-GFPCvHC had moved upwards through the graft junction. The symptoms and fluorescence observed on the Salinas scion grafted on an infected Salinas 88 rootstock were similar to those observed on the control graft combination (Fig. 6B), while no fluorescence or symptoms were seen on the inoculated Salinas 88 rootstock itself (data not shown).

In order to investigate whether the virus was impaired in its ability to unload from the phloem of the resistant variety, grafting experiments reciprocal to those described above were performed, in which healthy Salinas 88 scions were grafted onto infected Salinas rootstocks (Fig. 6C–E). Susceptible Salinas scions grafted onto infected susceptible Salinas rootstocks were again used as a control. Thirty days after grafting, no symptoms could be seen, but GFP fluorescence was observed on the Salinas 88 scions using a hand-held UV lamp (Fig. 6D). The fluorescence was, however, much less
Fig. 6. Lettuce graft combinations and in situ localization of GUS activity in Salinas and Salinas 88 lettuce. Observation of graft combinations under normal (A, C) and UV (B, D, E) light 30 days after grafting. The white arrowhead indicates the position of the scion leaf, the yellow arrow head the position of rootstock leaf. (A) and (B): Healthy Salinas scions were grafted onto infected Salinas rootstocks infected with LMV-0-GFPclvHC. At the day of grafting, fluorescence was detected under UV light on the Salinas rootstock. Thirty days after grafting, GFP fluorescence and symptoms were present in the Salinas scion (B). (C) and (E): Healthy Salinas 88 scions were grafted onto Salinas rootstocks inoculated with LMV-0-GFPclvHC. At the day of grafting, fluorescence under UV light and symptoms were present on the Salinas rootstock. Thirty days after grafting, GFP fluorescence without symptoms was detected in the Salinas 88 scion (D) but much less than in the Salinas rootstock (E). (F): Detection of LMV-0-GUSclvHC in the roots and upper systemic leaves of Salinas and Salinas 88 at 11 dpi.

Further indications that neither phloem loading nor unloading of the recombinant viruses was fully inhibited in the resistant plants came from the analysis of the roots of lettuce plants inoculated with LMV-0-GUSclvHC or LMV-0-GFPclvHC. Table 1 summarises the results of a time-course analysis of the detection of GUS- or GFP-tagged LMV-0 in roots and upper leaves of Salinas and Salinas 88. LMV-0 tagged viruses were detected in the roots of both varieties as early as 4 dpi and at 4, 8, 11, 14, 37 and 70 days post-inoculation, contrary to the situation with upper leaves of Salinas 88 (Table 1). However, irrespective of the analysis time, the recombinant viruses seemed to accumulate systemically in the root system of the resistant variety at a
In systemic leaves, ELISA and dot–blot assays were done, to confirm the presence or absence of detectable tagged viruses.

The systemic movement to the upper non-inoculated leaves appears to be more affected than the downward movement to the root system. In general, viral invasion of the root system has been poorly studied, and in most cases, when analysed, both upward and downward systemic movements were affected. However, similar to the situation reported here with LMV-0, Guerin and Murphy (1999) showed that in the resistant pepper (*Capsicum annuum* variety Avelar, carrying the recessive gene *pvr*), downward movement of Pepper mottle virus (PepMoV) to the roots still occurred while systemic movement to upper non-inoculated leaves was completely blocked due to a block of the entry into the internal phloem. Similar to our observations, albeit in transgenic plants and with another virus, Germundsson et al. (2002) showed that the CP gene-mediated resistance to Potato mop-top virus (PMTV) was expressed differently in roots and leaves.

In a number of virus-plant interactions, a block in upward systemic movement has been linked to the inability of the wild-type viruses (Dolja et al., 1997; Masuta et al., 2000).

The results presented here show that introduction of a NIa cleavage site fully restored wild-type biological properties in LMV-E, including its ability to produce symptoms in *mol*<sup>2</sup> plants. The behaviour of similar recombinants derived from LMV-0 was only partially restored, while a control virus with similar alterations of the HC-Pro but lacking a reporter gene had full wild-type properties.

The restriction in long distance movement and in systemic accumulation of the tagged LMV-0 recombinants in *mol*<sup>2</sup> varieties raises several questions. The fact that the restriction was observed in Salinas 88 but not in the near isogenic variety Salinas strongly supports the idea that the restriction is a consequence of the presence of the *mol*<sup>2</sup> resistance gene. Although the wild-type isolate LMV-0 is usually not affected in this way by the resistance, other natural isolates such as LMV-1 and LMV-9 (Dinant and Lot, 1992; Revers et al., 1997b) appear to be similarly affected in systemic accumulation in *mol*<sup>2</sup> varieties.

4. Discussion

During the last decade, the genomes of several potyviruses have been modified to express reporter genes (GUS or GFP) inserted between the PI and HC-Pro domains, either as a fusion with HC-Pro or with a cleavage site between the reporter gene and HC-Pro, in such a way that a nearly wild-type HC-Pro is produced upon action of the NIa proteinase. The effects of such insertions on the biological properties of the recombinant viruses differ greatly depending on the potyvirus/host combination. Expression of GUS or GFP as a fusion with HC-Pro was not lethal in most plants. The behaviour of similar recombinants derived from *PPV*, including its ability to produce symptoms in *mol*<sup>2</sup> varieties raises several questions. The fact that the restriction was observed in Salinas 88 but not in the near isogenic variety Salinas strongly supports the idea that the restriction is a consequence of the presence of the *mol*<sup>2</sup> resistance gene. Although the wild-type isolate LMV-0 is usually not affected in this way by the resistance, other natural isolates such as LMV-1 and LMV-9 (Dinant and Lot, 1992; Revers et al., 1997b) appear to be similarly affected in systemic accumulation in *mol*<sup>2</sup> varieties.

The systemic movement to the upper non-inoculated leaves appears to be more affected than the downward movement to the root system. In general, viral invasion of the root system has been poorly studied, and in most cases, when analysed, both upward and downward systemic movements were affected. However, similar to the situation reported here with LMV, Guerin and Murphy (1999) showed that in the resistant pepper (*Capsicum annuum* variety Avelar, carrying the recessive gene *pvr*), downward movement of Pepper mottle virus (PepMoV) to the roots still occurred while systemic movement to upper non-inoculated leaves was completely blocked due to a block of the entry into the internal phloem. Similar to our observations, albeit in transgenic plants and with another virus, Germundsson et al. (2002) showed that the CP gene-mediated resistance to Potato mop-top virus (PMTV) was expressed differently in roots and leaves.

In a number of virus-plant interactions, a block in upward systemic movement has been linked to the inability
of the virus to enter the phloem (Rajamäki and Valkonen, 2002; Guerini and Murphy, 1999; Wang et al., 1998) or to the inability to either enter into or exit from the phloem (Hull, 2002; Nelson et al., 1993; Schaad and Carrington, 1996; Thompson and García-Arenal, 1998) and has also frequently been observed to be associated with a reduced rate of cell-to-cell spread (Wong et al., 1999). In the case reported here, the grafting experiments showed that while the upward systemic movement is severely impaired, neither phloem loading nor phloem unloading appears to be completely inhibited. The limited amount of GFP fluorescence observed in resistant Salinas 82 scions grafted onto inoculated susceptible Salinas rootstocks seems to indicate, that phloem unloading or an ensuing step are partially inhibited in the resistant variety. It could be that the erratic way in which low level systemic accumulation is observed in the resistant varieties is caused by cumulative effects at each of the steps along the process: reduced cell-to-cell movement, reduced entry into the phloem, reduced ability to exit the phloem, etc. The alternate hypothesis is the existence, in the resistant plants, of a virus-induced defence signal which moves long-distance via the plant vascular system (Oparka and Tague, 1999) and leads to the establishment of a LMV restrictive state in upper non-inoculated tissues. To be compatible with the experimental grafting results, this hypothesis would, in addition, require that the susceptible genotypes be both unable to produce the signal and unable to respond to it.

It is tempting to speculate on the reasons for the substantial differences observed when using similar tagging strategies for different potyviruses or, in the case of LMV, for different strains of the same virus. Given the involvement of HC-Pro in multiple steps of the Potyvirus cycle, including long-distance movement, seed transmission, symptom expression and inhibition of host defences (Klein et al., 1994; Cronin et al., 1995; Johansen et al., 1996; Redondo et al., 2001; Li and Ding, 2001; Kasschau and Carrington, 2001), one possibility is that the observed differences in behaviour reflect minor differences in the tagging strategies. Such differences may result in a variable number of non-viral amino acids being inserted at the N-terminus of the HC-Pro or in variations in cleavage efficiency at the introduced Nla site and thus in the efficiency of the production of the free form of HC-Pro. In the work reported here, introduction of an artificial Nla cleavage site leads to processed HC-Pro with an insertion of two Glycine residues between the first and second amino acids at the N-terminus of HC-Pro. This modification is, however, unlikely to affect the biological properties of the recombinants since LMV-0-clvHC shows a wild-type behaviour for all parameters that we analysed. Similarly, minor modification of the HC-Pro N-terminus in TEV has no marked effect on the biology of the virus, including aphid transmission (Dolja et al., 1998).

Comparison of the behaviour of LMV-0-clvHC with that of its GUS or GFP containing derivatives demonstrates that the introduction of the reporter genes is responsible, directly or indirectly, of the loss of the ability to efficiently accumulate in systemic tissues. At the moment two hypotheses can be proposed to explain this observation. One possibility would be that the presence of an insert (0.7 kbp for GFP, 1.5 kbp for GUS) and the resulting enlargement of the genome by 7–15% would "mechanically" slow down viral replication and all subsequent plant invasion processes, which in turn would give a hedge to the plant in the race between viral invasion and plant defence mechanisms such as RNAi (Kasschau and Carrington, 2001). The second possibility would be that the reporter genes fused to HC-Pro could somehow affect one or all of its biological functions (Klein et al., 1994; Cronin et al., 1995; Johansen et al., 1996; Redondo et al., 2001; Li and Ding, 2001; Kasschau and Carrington, 2001). This second hypothesis is in keeping with the observations of Masuta et al. (2000) with CIYVV and Carrington et al. (1993) with TEV which demonstrated complete maturation of the fusion proteins accompanied by complete recovery of the wild-type virus biological properties. In this scenario, the constructs containing an Nla cleavage site would still be producing only about 10% less of active HC-Pro than the wild-type virus since cleavage between the reporter protein and HC-Pro was never found to be complete. Alternatively, the remaining fused HC-Pro could have a dominant negative effect on some functions exerted by HC-Pro during the virus cycle.

The wild-type behaviour of LMV-0-clvHC could in turn be explained by either a more complete cleavage reaction in the absence of a reporter gene or by a lower detrimental effect on HC-Pro functions due to the more limited structural modifications suffered by the HC-Pro in this context (insertion of only 13 amino acids between the first and second amino acids of the HC-Pro as compared to the insertion of the same 13 amino acids plus the reporter gene in the other constructs). This idea of a limited effect of small extensions to the HC-Pro is supported by the observation that a LMV-E derived recombinant in which the N-terminus of the HC-Pro is tagged by the addition of a poly-histidine tail (total of 10 amino acids added in fusion) retains all biological properties of its wild-type parent (German–Retana, unpublished results; Plisson et al., 2003). The exploration of these hypotheses and in particular the use of site-directed mutagenesis to improve the cleavage efficiency at the synthetic Nla site could further our understanding of the mechanisms involved and possibly allow the development of tagged LMV recombinants sharing all the biological properties of their wild-type counterparts. Nevertheless, the GFP- and GUS-tagged LMV developed by us are useful tools that are already used as expression vector to analyse the interaction between LMV and its host species.

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