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Virological and serological analysis of a recent Middle East respiratory syndrome coronavirus infection case on a triple combination antiviral regimen

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1. Introduction

An upsurge of Middle East respiratory syndrome coronavirus (MERS-CoV) infection has been recently described in countries of the Arabian Peninsula resulting in exported cases from these countries to the European Union [1]. Cases of MERS-CoV infection are associated with a high case fatality rate since there is no available treatment. There is a scarcity of data on specific therapeutic interventions for the disease. Published reports propose the use of known antivirals based on extrapolation of data from: (i) the severe acute respiratory syndrome (SARS) epidemic that was also associated with the circulation of a novel coronavirus; (ii) in vitro data; (iii) animal experimental infections and therapy data; and (iv) limited clinical data for actual MERS-CoV infections [2–4]. However, no clear-cut recommended therapeutic regimen exists and the evidence for grading such interventions is generally low, with the exception of the use of convalescent serum that based on biological effects is given the highest grade [5]. Moreover, little is known about the viral kinetics of MERS-CoV-associated infection, especially when a specific antiviral or other therapeutic intervention is attempted.

A case of MERS-CoV has recently been described in Greece in a traveller who had extensive contact with the healthcare environment in Jeddah (Saudi Arabia) [6]. Here we describe molecular, serological and phylogenetic analyses of this case as well as evidence for a second case that was a close contact of the first patient. Furthermore, we provide evidence of the kinetics and the pattern of viral excretion in biological specimens obtained from the first Greek case while the patient was on a triple antiviral regimen.

2. Methods

2.1. Case investigation

A preliminary report of the first imported, laboratory-confirmed MERS-CoV case in Greece has been described elsewhere [6]. A full description of the course of illness and treatment regimen in

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relation to kinetics of virus shedding and immune response was prepared by review of the patient records. In the course of the outbreak investigation, 40 of 75 patient’s contacts, including the patient’s wife, provided an oropharyngeal sample for PCR testing 1 week after contact with the positive case; 5 additional contacts were included in the serology examination group. All were submitted to personal clinical monitoring for fever and upper respiratory infection symptoms and were advised to call the Hellenic Centre for Disease Control and Prevention (CDC) command centre immediately in such an instance. In addition, all were offered the chance to provide serum samples on a voluntary basis for specific anti-MERS antibody testing at baseline (same time as the oropharyngeal PCR testing) and 3 weeks after exposure.

2.2. Laboratory evaluation

During the patient’s stay in the intensive care unit (ICU), samples from the oropharynx, trachea, urine and faeces were tested for diagnostic evaluation and to monitor viral shedding. A real-time reverse transcription PCR (RT-PCR) method based on amplification of the upstream Envelope gene (upE), the nuleocapsid (N) gene and the open reading frame (ORF) 1a of the virus was used for detection of MERS-CoV according to previously described methodology [7,8].

Immunoglobulin G (IgG) and IgM antibody titres in serum samples were determined using an anti-MERS-CoV Indirect Immunofluorescence Assay (Euroimmun AG, Lübeck, Germany). Confirmation of the serological findings was performed with a virus neutralisation assay as described previously [9].

Samples from the patient’s upper respiratory tract underwent conventional or molecular testing for the presence of other respiratory pathogens: thus, cultures applied for bacterial testing, whilst real-time RT-PCR was performed for several respiratory viruses including influenza A and B viruses, respiratory syncytial virus (RSV), parainfluenza, adenovirus, enterovirus, bocavirus and human metapneumovirus (hMPV) (M.W.S r-gene; bioMérieux, Marcy-l’Étoile, France). Specific urine antigen testing of urine samples was utilised for Legionella pneumophila and Streptococcus pneumoniae (BinaxNOW®; Alere, Orlando, FL). A stool culture was performed due to a history of possible typhoid fever, diagnosed by treating physicians in Saudi Arabia [6].

2.3. Phylogenetic analysis

Nucleotide sequences of 3-kb concatenated sequences of representative MERS-CoVs were analysed and a phylogenetic tree was constructed by the PhyML method as described previously [10].

3. Results

3.1. Case description

A 69-year-old patient of Greek origin who was a permanent resident of Jeddah presented to a tertiary care centre a few hours after arriving in Athens (Greece) on 17 April 2014. His chief complaints included fever since 8 April 2014 and diarrhoea since 10 April 2014. The most likely source of exposure was the hospital environment in Jeddah. The patient had no known co-morbidities. At the time of initial evaluation, a fever of 38.3 °C was noted together with low oxygen saturation (92%), although the patient exhibited minimal respiratory symptoms. A chest radiograph depicted bilateral lung infiltrates consistent with viral pneumonia. The patient was immediately placed under isolation because of suspicion of MERS-CoV infection, and an antimicrobial regimen targeting community-acquired pneumonia was initiated.

On 18 April 2014, MERS-CoV infection was confirmed by means of viral RNA detection in a pharyngeal swab at the Department of Microbiology, University of Athens Medical School (Athens, Greece).

After laboratory confirmation of MERS-CoV, the patient was transferred to a specialised respiratory disease unit in the ‘Sotiria’ Chest Diseases Hospital of Athens where he was treated in a negative pressure regular room in isolation until 20 April 2014 when, due to deterioration of his respiratory function and development of acute respiratory disease syndrome (ARDS), he was intubated, ventilated and transferred to a negative pressure room in the ICU of the same hospital. An empirical antiviral regimen was initiated on Day 13 of illness consisting of oral (p.o.) lopinavir/ritonavir (400/100 mg twice daily), pegylated interferon (180 µg subcutaneously once per week for 12 days) and ribavirin (2000 mg p.o. loading dose, followed by 1200 mg p.o. every 8 h for 8 days) based on available evidence [3–5,11,12] (Fig. 1).

The patient remained intubated exhibiting hypoxia and occasionally hypercapnia while breathing inspired oxygen in the range of 0.45–0.60. He remained febrile with a plateau temperature of >39 °C and a maximum value of 40.5 °C on Day 18 of illness. Fever started subsiding below 38 °C on Day 22. Acute kidney injury was diagnosed on Day 16 of illness and rapidly progressed to non-oliguric renal failure that reverted to RFLE injury level (i.e. two-fold increase in the serum creatinine, or glomerular filtration rate decrease by 50%, or urine output <0.5 mL/kg/h for 12 h) on Day 21. The patient’s diarrhoea resolved gradually starting on Day 13 and he developed constipation thereafter with normalisation of his bowel movements and gastrointestinal function on Day 19. Owing to development of jaundice and hyperbilirubinaemia attributed to ribavirin [13], the drug was discontinued on Day 20. During the course of his hospitalisation, the patient was diagnosed with adenocarcinoma of the colon and eventually died from septic shock 2 months and 19 days after the initial diagnosis.

3.2. Testing for other pathogens

 Cultures and antigen detection were negative for L. pneumophila and S. pneumoniae. Virological testing was negative for the presence of any other respiratory virus. No relevant enteric pathogens were identified as a cause of the patient’s diarrhoea.

3.3. MERS-CoV testing and shedding

RNA was detected in several consecutive patient samples from different sites that included faecal material and serum (Fig. 1). Shedding of MERS-CoV in the respiratory secretions of the patient was noted until the fourth week of illness, whereas viraemia was last detected 15 days after onset of illness and 2 days after initiation of the triple combination antiviral regimen. Consecutive urine testing did not reveal the presence of MERS-CoV RNA (Fig. 1).

3.4. Serological testing for MERS-CoV and new possible case

Serological testing showed a peak IgG titre during the third week of illness, whilst during the fourth and fifth weeks IgG titres were substantially declining. IgM titres were persistently elevated during the whole survey period (Day 13 until Day 34 of illness) (Fig. 1). Viral neutralisation assays performed at Erasmus Medical Center (Rotterdam, The Netherlands) confirmed the immunofluorescence testing results.

Initial and follow-up serological testing were performed on serum samples from 45 patient’s contacts. Seroconversion was revealed in one of them who developed an IgG titre of 1/500 and an IgM titre of 1/100 at 21 days after making contact with the patient. This was a 63-year-old man with a past medical history of coronary artery heart disease and diabetes. The presence of specific
MERS-CoV antibodies was confirmed by the virus neutralisation assay.

3.5. Case investigation of the new case

Case investigation disclosed that the person who seroconverted had close contact with the first MERS-CoV case diagnosed in Greece on 18 April 2014. He had close contact with the index case for ca. 3 h and drove the index patient to the hospital on 17 April 2014. He had been identified as a close contact during the contact tracing conducted by the Hellenic CDC at the time and he was submitted to the following testing according to the protocol used for the investigation of close contacts by the Hellenic CDC: (i) nasopharyngeal testing (by PCR) on 23 April 2014 (negative); (ii) serology testing on 23 April 2014 (negative); and (iii) serology testing on 8 May 2014 (positive). The patient reported developing only fever (up to 38.5°C) from 4 to 7 May 2014 without other symptoms from any other system. No nasopharyngeal PCR testing was performed at the time since he was outside the incubation period of 14 days. He has been well since then and during the time that he was symptomatic he only had contact with his family members (four persons). The initial patient’s wife had a brief episode of fever on 30 May 2014. Oropharyngeal PCR testing was negative for MERS-CoV, and all contacts remained seronegative on repeat testing.

3.6. Phylogenetic results

Partial genomic sequencing [14] revealed the close phylogenetic relationship with clinical MERS-CoV strains associated with severe respiratory infection from patients in Jeddah (Fig. 2).

4. Discussion

In this report, we further characterised serological and virological parameters of the first MERS-CoV case in Greece. Rising titres of IgG were demonstrated in sequential serum samples, with the peak titres approximately 3 weeks into the course of the disease. This is in accordance with serological testing guidance from the World Health Organization (WHO) recommending baseline testing from initial contact with an affected case and repeated serological testing on Day 21 [15]. On the other hand, IgM titres of the patient remained constantly elevated above the threshold of detection, albeit at a lower level than IgG antibodies, for a prolonged period of >1 month of follow-up. Thus, isolated use of IgM testing without concomitant IgG determination appears not to be sufficient to reveal a recent infection. It should be noted, however, that in the absence of detailed studies, use of serological testing for MERS-CoV detection in humans needs to be further evaluated.

Prolonged shedding of the virus was noted from the respiratory tract of the patient. This finding is consistent with data regarding the SARS coronavirus. In a report dealing with patients affected by SARS, prolonged shedding of the virus was noted in stool (up to 126 days) and respiratory specimens (up to 52 days) [16]. Data regarding the length of MERS-CoV excretion from different body sites are scarce [17]. Excretion of the virus probably depends on the amplitude of replication in different body sites, the underlying immune status and co-morbidities, and appropriate antiviral therapy. The non-detectable viral RNA in serum by Day 3 after initiation of the antiviral treatment could be explained either by viral clearance in an otherwise immunocompetent person or by effectiveness of the instituted antiviral regimen. Literature on appropriate antiviral intervention for MERS-CoV is very limited and currently no evidence-based therapy exists. The regimen chosen was based on the best available literature as well as evidence from animal and patient data that have been described elsewhere [2–4]. The role of interferon therapy for MERS-CoV infection needs to be further elucidated. An attenuated interferon-β (IFN-β) response has been described as a result of MERS-CoV infection [18] and extensive use of interferon-based regimens alone or in combination with ribavirin has been described for SARS [4]. However, interferons appear to have a better antiviral effect on MERS-CoV compared with SARS-CoV in vitro experiments [19]. In vitro, IFN-β appears to exhibit the best anti-MERS-CoV effect [20]. Interferon activity has been enhanced by the addition of ribavirin in in vitro experiments [21]. Furthermore, this combination has shown promising clinical and radiological effects in Rhesus macaques experimentally infected with MERS-CoV [12]. Thus, the clinical team elected to use this combination despite the fact that a prestigious public health agency ranks ribavirin use as not supported by high-quality evidence [5]. In a more recent update published by the same public health agency, the use of interferons and lopinavir is ranked under the recommendation of benefit is likely to exceed risk, whereas the combination of interferon and ribavirin is ranked as data are inadequate for assessment [22]. The frequent side effects of ribavirin limit its use in combination regimens for actual MERS-CoV-infected patients, as was the experience with the current patient where liver toxicity, although not definitively associated, was mainly attributed to this medication. The renal function deterioration of the patient described here was considered multifactorial and probably also a complication of the virus infection [23]. The possibility that drug toxicity might have contributed in the renal dysfunction could not be excluded, however. No drug levels were measured since the...
patient was under continuous renal replacement therapy at that time and drug levels would be unreliable. In the actual clinical human setting, the combination of ribavirin and interferon has been tried, with no successful outcome reported among any of the MERS-CoV-infected recipients [3]. Nevertheless, the group studied consisted of severely ill patients who received this combination quite late in the course of their disease [3]. Lastly, we added the protease inhibitors lopinavir/ritonavir based on experience accumulated from the SARS epidemic where the addition of this agent to ribavirin improved the outcome of infection [24].

Expanding the knowledge regarding viral kinetics and the pattern of shedding especially in association with specific therapeutic interventions has important implications for infection control in the healthcare environment, especially as it relates to potential transmission to other patients and healthcare workers [25].

Phylogenetic analysis of the Greek MERS-CoV strain showed close similarity with circulating patient viral strains from the recent Jeddah outbreak as well as with a strain isolated from a dromedary camel in Qatar. This is in accordance with previous genetic studies that have shown identical viral strains between infected humans and dromedary camels and generated the hypothesis that dromedary camels are among the reservoirs of the virus in nature [10,26]. Also, the presence of MERS-CoV-specific antibodies in camels across a wide geographic area in Africa and the Arabian Peninsula signifies the possibility for zoonotic transmission between camels and humans [27–29]. The potential for transmission across different individual strains should be further explored. In this patient, the most likely source of exposure was the hospital environment in an endemic area, as his wife was hospitalised in a local hospital in Jeddah [6]. It appears that healthcare-associated outbreaks are playing a pivotal role in the evolution of the MERS-CoV epidemic in the recent upsurge [30].

5. Conclusions

In conclusion, we describe the genetic stability of the MERS-CoV in a strain from the recent Jeddah outbreak. Although reassuring, this finding should not limit the level of awareness regarding the increased number of cases in the Arabian Peninsula reported recently and the potential evolution and more efficient transmission of the virus. A WHO committee recently concluded that the conditions for a Public Health Emergency of International Concern have not yet been met. Nevertheless, important gaps in current knowledge about MERS-CoV exist. More investigations to clarify the natural reservoir and modes of transmission are necessary. Persistence of virus shedding in patients’ secretions and the effect of immune status and antiviral therapy together with the implementation of appropriate infection control measures are of paramount importance in limiting further spread of this potentially lethal virus.

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