Cell death–mediated cytokine release and its therapeutic implications

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Targeting apoptosis to treat diseases has seen tremendous success over the past decades. More recently, alternative forms of regulated cell death, including pyroptosis and necroptosis, have been described. Understanding the molecular cascades regulating both pyroptosis and necroptosis will yield even more targets to treat diseases. These lytic forms of cell death are distinct from apoptosis due to their characteristic lysis and release of cellular components that promote disease or direct a beneficial immune response. In this review, we focus on how pyroptosis and necroptosis, which release potent immune cytokines such as IL-1 and IL-18, contribute to various diseases. We also consider the important role that the executioners of these cell death pathways, GSDMD and MLKL, play in the progression of inflammatory diseases. Crosstalk between the different cell death pathways likely plays a major role physiologically. New therapeutic strategies targeting these specific molecules hold enormous potential for managing inflammatory diseases.

Introduction
Regulated cell death has been understood as a concept for decades, with apoptosis being the first well-defined process in which cells dismantle themselves in a process that is generally immunologically quiet (Kerr et al., 1972; Elmore, 2007). Apoptosis is induced homeostatically and upon exposure to a wide variety of insults, leading to the activation of initiator caspases (caspase-8, -9, -10) and effector caspases (caspase-3, -6, -7), resulting in a nonlytic cell death characterized by membrane blebbing, cell shrinkage, and chromosomal condensation (Elmore, 2007). While apoptosis facilitates the controlled degradation of intracellular proteins and organelles, pyroptosis and necroptosis lead to cell lysis and the release of a wide range of intracellular components and inflammatory cytokines. We focus in this review on the lytic forms of cell death (pyroptosis and necroptosis) and the consequences of their cytokine release, with an eye toward new ways of treating inflammatory diseases.

Unlike apoptotic cell death, in which plasma membrane integrity is maintained and intracellular components are sequestered, pyroptosis is a lytic form of cell death that, through a tightly regulated pathway, activates and releases the potent cytokines IL-1β and IL-18 (Fig. 1; Dinarello et al., 1974; Dinarello, 1998; Cookson and Brennan, 2001; Martinon et al., 2002). Pyroptosis plays a major role in the control of intracellular pathogens by destroying their replicative niche and driving immune responses through the release of matured cytokines (Man et al., 2017). Like apoptosis, pyroptosis is regulated by caspases, in particular caspase-1 and caspase-11 (in mice) or caspase-1/4/5 (in humans). Caspase-4/5/11 binds intracellular LPS directly, leading to cleavage of the substrate gasdermin D (GSDMD), the recently discovered executioner of pyroptosis (He et al., 2015; Kayagaki et al., 2015; Shi et al., 2015). Upon cleavage, the autoinhibitory C-terminal domain of GSDMD is released from the pore-forming N-terminal domain, leading to insertion of the GSDMD N-terminal domain into the plasma membrane and oligomerization with other GSDMD fragments, thereby generating a large pore 10–20 nm in diameter (Ding et al., 2016). Similar to caspase-4/5/11, caspase-1 activation by upstream sensor proteins leads to caspase-1–mediated cleavage of GSDMD. The upstream regulators of caspase-1 activation differ in their specificity to different ligands and cellular states and include the proteins NLRP3, NLRC4, AIM2, NLRP1, and pyrin, with other related proteins being suggested as additional sensors (Place and Kanneganti, 2018). A subset of these sensors (NLRP3, AIM2, and pyrin) require the adaptor protein ASC to activate caspase-1 after ligand sensing.

A single specific trigger for NLRP3 activation is not known, but many cellular insults induce NLRP3 inflammasome activation and cleavage of caspase-1, including loss of cellular ion homeostasis and plasma membrane disruption (Kanneganti et al., 2006; He et al., 2016; Jo et al., 2016). NLRC4 is activated by a set of sensor proteins including human NAIP/mouse NAIPI, NAIPI2, and NAIP5/6, which recognize the bacterial type 3 secretion system (T3SS) needle, T3SS rod, or flagellin proteins,

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respectively (Sharma and Kanneganti, 2016). The upstream NAIP proteins are also transcriptionally regulated by the transcription factor IRF8 (Karki et al., 2018). The AIM2 inflammasome is activated by cytosolic double-stranded DNA binding with AIM2, which is detected upon viral entry into the cytosol or downstream of IRF1-mediated expression of IFN-stimulated genes that lyse intracellular bacteria (Fernandes-Alnemri et al., 2009; Hornung et al., 2009; Man et al., 2015; Meunier et al., 2015). The mouse NLRP1b inflammasome is activated following proteolysis after exposure to Bacillus anthracis lethal toxin and DPP8/9 inhibitors (Chavarría-Smith and Vance, 2013; Okondo et al., 2017). Newer studies have shown that endogenous human DPP9 binds and negatively regulates NLRP1, and a mutation in this binding domain is associated with NLRP1-associated autoinflammation with arthritis and dyskeratosis (NAIAD; Zhong et al., 2018). DPP8/9 inhibitors activate murine NLRP1b by inducing proteasomal degradation of the autoinhibitory N-terminal fragment, while anthrax lethal toxin functions by induction of N-end rule–mediated proteolysis of the N-terminal fragment of NLRP1 (Okondo et al., 2018; Chui et al., 2019; de Vasconcelos et al., 2019b; Sandstrom et al., 2019). Pyrin inflammasome activation is triggered by RhoA-GTPase–modifying toxins (Xu et al., 2014). Concurrent with GSDMD cleavage, caspase-1 activation cleaves inactive pro-IL-1β and pro-IL-18 into their active forms, which are released through the GSDMD pore. Caspase-11 binding to intracellular LPS activates caspase-11 to cleave GSDMD, which drives pore formation that leads to cell lysis and downstream NLRP3 inflammasome activation through the loss of ion homeostasis. dsDNA, double-stranded DNA.

Figure 1. Overview of pyroptosis. Pyroptosis is mediated by the inflammasome sensor proteins NLRC4, NLRP1, AIM2, Pyrin, and NLRP3, leading to activation of caspase-1. The NLRP1 and NLRC4 sensor proteins do not require the adaptor protein ASC, while the remaining sensors require ASC to oligomerize with caspase-1. Caspase-1 directly cleaves GSDMD to release the autoinhibitory C-terminal domain from its pore-forming N-terminal domain and also cleaves pro-IL-1β and pro-IL-18 into their active forms, which are released through the GSDMD pore. Caspase-11 binding to intracellular LPS activates caspase-11 to cleave GSDMD, which drives pore formation that leads to cell lysis and downstream NLRP3 inflammasome activation through the loss of ion homeostasis. dsDNA, double-stranded DNA.
need processing by caspase-1, such as IL-1β, IL-33, HMGB1, ATP, and ASC complexes, are released, further contributing to the inflammatory response (Vande Walle et al., 2011; Basiorka et al., 2018; Frank and Vince, 2019). Together, the majority of these inflammasome complexes function to expel intracellular pathogens from the cell or halt viral replication in addition to releasing inflammatory cytokines, serving a critical role in host defense. Excessive or dysregulated activation of these complexes can also be detrimental to the host and is associated with inflammatory diseases, as discussed later in this review.

Similar to pyroptosis, necroptosis is a lytic form of cell death that is activated by a different pathway. In cells where caspase-8 is inhibited or missing, inflammatory signaling via TNF superfamily receptors, IFNs, TLR3, or TLR4 can lead to the phosphorylation of RIPK1, RIPK3, and ultimately mixed-lineage kinase domain-like pseudokinase (MLKL; Fig. 2; Holler et al., 2000; Cho et al., 2009; He et al., 2009; Linkermann and Green, 2014; Wang et al., 2014; Mocarski et al., 2015). Following phosphorylation, MLKL assembles into oligomers that create a channel in the plasma membrane, disrupting the ion homeostasis of the cell, similar to GSDMD-mediated lysis (Holler et al., 2000; Sun et al., 2012; Zhao et al., 2012; Linkermann and Green, 2014; Wang et al., 2014). The conditions for necroptosis are fairly strict in comparison to pyroptosis, but recent research has identified scenarios in which this pathway is engaged. Physiologically, it seems that necroptosis plays a backup role for cell death when the apoptotic pathway becomes inhibited by pathogens, mutations, changes in the regulators of apoptosis, or cancer (Pasparakis and Vandenabeele, 2015). Due to the complicated role of Fas-associated protein with death domain (FADD), caspase-8, RIPK1, RIPK3, and MLKL in regulating this form of cell death and also in regulating cytokine production, it is difficult to definitively attribute necroptosis to the progression of many diseases. New mouse models, chemical inhibitors, and assays that probe for phosphorylation of MLKL are helping to advance our understanding of the role of necroptosis in driving diseases (Pasparakis and Vandenabeele, 2015; Weinlich et al., 2017).

Crosstalk between apoptosis, necroptosis, and pyroptosis makes it difficult to determine how each pathway contributes to complex inflammatory diseases (Fig. 2). For example, apoptosis signaling through FADD and caspase-8 has been shown to link signaling between TLRs and priming of the NLRP3 inflammasome, and many of the proteins of the apoptosis pathway
also regulate necroptosis (Elmore, 2007; Gurung et al., 2014). In addition to caspase-8-mediated priming of the inflammasome, others have observed that caspase-8 can localize to caspase-1- and ASC-containing specks, facilitating inflammasome activation and downstream cytokine maturation (Gringhuis et al., 2012; Man et al., 2013; Karki et al., 2015). Treatment of colorectal cancer cells with the platinum-based, DNA-damaging chemotherapy drug lobaplatin resulted in caspase-3–dependent cleavage of GSDME, a second gasdermin family pore-forming protein (Yu et al., 2019). Infection with *Versinia spp.* has also revealed a role for caspase-8–mediated cleavage of GSDMD, suggesting this plays a previously understudied role in regulating crosstalk between apoptosis and pyroptosis (Orning et al., 2018; Sarhan et al., 2018a). While recent studies have shown that crosstalk between apoptotic signaling pathways with necroptosis and pyroptosis occurs, we focus here on the lytic forms of cell death as the final outcome that drives inflammatory pathology. In human cell lines, others have found that MLKL ion channel formation can lead to NLRP3 inflammasome activation in a cell-intrinsic manner, suggesting that this crosstalk between necroptosis and pyroptosis plays a larger role than previously appreciated (Conos et al., 2017; Gutierrez et al., 2017). Other recent studies have described key new players that regulate crosstalk between cell death pathways. Inhibition or loss of TAK1 leads to autocrine TNF-mediated RIPK1-dependent cell death through combined apoptosis, pyroptosis, and necroptosis pathways (Malireddi et al., 2018). In some viral infections, it also appears that the protein ZBP1 can act as a master regulator of all three cell death pathways, which requires further study in other contexts (Kuriakose et al., 2016). Importantly, these lytic forms of cell death also regulate the release of inflammatory cytokines that have been targeted successfully in many disease treatments. New studies examining the executioners of pyroptosis and necroptosis and their crosstalk in different contexts will shed more light on their physiological relevance.

### Monogenic inflammatory diseases

Due to the potent inflammatory signaling role of IL-1 and IL-18, the dysregulated activation and release of these cytokines contribute to many diseases (Table 1). Well-characterized gain-offunction mutations in inflammasome sensors lead to excessive production of these cytokines and cell death, underlying the pathogenesis of a number of monogenic diseases. Gain-offunction mutations in pyrin are associated with the auto-inflammatory disease familial Mediterranean fever (FMF) and drive the excessive release of IL-1β via GSDMD-mediated pyroptosis in a mouse model of FMF (Sharma et al., 2017; Kanneganti et al., 2018). While commonly treated with colchicine, recent work has shown that inhibition of IL-1 with anakinra (recombinant IL-1R antagonist), canakinumab (anti-IL-1β monoclonal antibody), or rilonacept (decoy IL-1R/IL1-RA/IgG fusion protein) can improve outcomes in colchicine-resistant FMF and that GSDMD inhibitors may also play a future role (Varan et al., 2019). In addition to the role of pyroptosis-mediated IL-1 release, a recent study showed that TNF signaling promoted the expression of pyrin in a positive feedback loop (Sharma et al., 2019). Patients with FMF also exhibit elevated serum IL-18, but whether this promotes disease and is a promising target for inhibition is unclear (Wada et al., 2018).

Autoinflammatory diseases associated with NLRP3 mutations, collectively described as cryopyrin-associated periodic syndromes (CAPS), include familial cold autoinflammatory syndrome, Muckle-Wells syndrome, and neonatal-onset multisystem inflammatory disorder/chronic infantile neurological, cutaneous, and articular syndrome (Manthiram et al., 2017). Together, these diseases are associated with systemic inflammation, neutrophilia, fever, rashes, and joint pain and are successfully treated with anakinra, rilonacept, and canakinumab and may be treated with NLRP3 or GSDMD inhibitors in the future (Lachmann et al., 2009; Gillespie et al., 2010; Coll et al., 2015; Xiao et al., 2018). In addition to FMF and CAPS, other well-characterized diseases driven by IL-1, including deficiency of IL-1R antagonist; Blau syndrome; mevalonate kinase deficiency; Majeed syndrome; and PAPA (pyogenic arthritis, pyoderma gangrenosum, and acne) syndrome, are effectively treated with IL-1 blockade, but may eventually also be treated by inhibiting GSDMD or IL-18.

Understanding the mechanism behind other monogenic diseases associated with excessive IL-1 or TNF production has led to successful treatment with neutralizing antibodies against these cytokines, summarized in Table 1. In TNF receptor-associated periodic syndrome, which is mediated by a mutation that causes hyperactivation of TNFR1, IL-1 blockade but not TNF blockade was shown to be effective in treating patients, suggesting that treatments targeting the downstream TNF-mediated cell death, possibly via inhibition of MLKL and GSDMD, may be effective alternatives (Jesus and Goldbach-Mansky, 2014;Gattorno et al., 2017). In mice, a mutation in Sharpin (Sharpin*cpdm*) causes mice to develop chronic proliferative dermatitis, which is driven by excessive TNF production, which leads to excessive epithelial cell apoptosis and inflammation that is promoted by RIPK1, RIPK3, MLKL, caspase-1/11, NLRP3, and IL-1β, suggesting that inhibition of cell death in this model may protect from similar skin disorders in humans (Gijbels et al., 1996; Kumari et al., 2014; Rickard et al., 2014; Gurung et al., 2016b). In humans, the NLRP1 P1214R mutation, adjacent to the autolysis site P1212-S1213, is associated with NAIAD, and molecular studies have found that DPP9 binding to the NLRP1 FIIND domain is disrupted by this mutation, suggesting that the binding of DPP9 and possibly DPP9-mediated degradation of the auto-inhibitory C-terminal fragment of NLRP1 is lost in this disease (Grandemange et al., 2017; Zhong et al., 2018). Loss of the executioner of pyroptotic cell death, GSDMD, in mouse models of FMF and CAPS protects mice from disease, suggesting that inhibitors against GSDMD or other steps of the pyroptosis signaling cascade may also show therapeutic potential in other inflammatory disorders.

### IL-1–mediated diseases

Mutations in key regulators of the pyroptosis pathway are known to drive autoinflammatory diseases. In addition to monogenic diseases, other complex inflammatory diseases can be successfully treated with IL-1 blockade. These inflammatory
diseases include Behcet’s disease, Sweet’s syndrome, Schnitzler’s syndrome, adult-onset Still’s disease, and juvenile idiopathic disease, summarized in Table 2. Because IL-1 is elevated in multiple inflammatory diseases, it is suspected to contribute to the progression of gout, rheumatoid arthritis, kidney disease, lung disease, cancer, and many others reviewed.

### Table 1. Monogenic autoinflammatory diseases

| Disease            | Key regulator | Treatment target                                      | Reference                                                                 |
|--------------------|---------------|-------------------------------------------------------|---------------------------------------------------------------------------|
| CAPS               | NLRP3         | IL-1β (canakinumab)                                   | Goldbach-Mansky et al., 2006; Gattorno et al., 2007;                       |
|                    | IL-1β         | IL-1β (canakinumab)                                   | Lachmann et al., 2009; Kitamura et al., 2014;                              |
|                    | GSDMD         | IL-1 decoy receptor (rilonacept)                      | Coli et al., 2015; Xiao et al., 2018                                      |
| FMF and PAAND      | Pyrin         | IL-1R (anakinra)                                      | Hashkes et al., 2012; Sharma et al., 2017, 2019; Varan et al., 2019       |
|                    | IL-1β         | IL-1β (canakinumab)                                   |                                                                          |
|                    | TNF           | IL-1β (canakinumab)                                   |                                                                          |
|                    | IL-1β         | IL-1 decoy receptor (rilonacept)                      |                                                                          |
|                    | GSDMD         | IL-1β (anakinra)                                      |                                                                          |
|                    |               | IL-1 decoy receptor (rilonacept)                      |                                                                          |
|                    |               | GSDMD?                                                |                                                                          |
| DIRA               | IL-1RA        | IL-1R (anakinra)                                      | Jesus and Goldbach-Mansky, 2014                                           |
| Blau syndrome/PGA  | NOD2          | IL-1R (anakinra)                                      | Aróstegui et al., 2007; Simonini et al., 2013                            |
|                    | IL-1β         | IL-1β (canakinumab)                                   |                                                                          |
| MKD/HIDS           | MVK           | IL-1β (anakinra)                                      | Manthiram et al., 2017                                                   |
|                    | Pyrin         | IL-1β (canakinumab)                                   |                                                                          |
| TRAPS              | TNFR          | IL-1β (canakinumab)                                   | Jesus and Goldbach-Mansky, 2014                                           |
|                    | IL-1β         | IL-1β (canakinumab)                                   |                                                                          |
|                    | GSDMD?        | IL-1β (canakinumab)                                   |                                                                          |
|                    | MLKL?         | IL-1β (canakinumab)                                   |                                                                          |
| Majeed syndrome    | LPIN2-NLRP3   | IL-1R (anakinra)                                      | Jesus and Goldbach-Mansky, 2014                                           |
|                    | IL-1β         | IL-1β (anakinra)                                      |                                                                          |
| PAPA syndrome      | PSTPIP1-Pyrin | IL-1R (anakinra)                                      | Kanameishi et al., 2017; Manthiram et al., 2017                           |
|                    | IL-1β         | IL-1R (anakinra)                                      |                                                                          |
|                    | IL-1β         | IL-1β (canakinumab)                                   |                                                                          |
|                    | TNF           | IL-1β (canakinumab)                                   |                                                                          |
| PFIT               | WDR1-Pyrin    | IL-1β (canakinumab)                                   | Standing et al., 2017                                                    |
|                    | IL-1β         | IL-1β (canakinumab)                                   |                                                                          |
|                    | GSDMD?        | IL-1β (canakinumab)                                   |                                                                          |
| AIFEC              | NLRC4         | IL-1β                                                | Romberg et al., 2017                                                     |
|                    | IL-18         | IL-1β                                                |                                                                          |
|                    | IL-17         | IL-1β                                                |                                                                          |
|                    | IFNγ?         | IL-1β                                                |                                                                          |
| NAIAD              | NLRP1         | IL-1β                                                | Grandemange et al., 2017; Zhong et al., 2018                              |
|                    | DPP9          | IL-1β                                                |                                                                          |
|                    | NLRP1?        | IL-1β                                                |                                                                          |
|                    | NLRP1?        | IL-1β                                                |                                                                          |
|                    | GSDMD?        | IL-1β                                                |                                                                          |
| NLRP12AD           | NLRP12        | IL-1R (anakinra)?                                     | Jéru et al., 2011                                                        |
|                    | IL-1β         | IL-1R (canakinumab)                                   |                                                                          |
|                    | GSDMD?        | IL-1R (canakinumab)                                   |                                                                          |
|                    | TFN           | IL-1R (canakinumab)                                   |                                                                          |
| Chronic proliferative dermatitis | Sharpin | TNF?                                                  | HogenEsch et al., 1993; Gijbels et al., 1996; Gerlach et al., 2011;       |
|                    |               | RIPK1 (Necrosisatin-1)?                               | Kumari et al., 2014; Rickard et al., 2014; Gurung et al., 2016b           |
|                    | TFN           | RIPK1 (GSK-872)?                                      |                                                                          |
|                    | RIPK3         | MLKL (necrosulfonamide)                               |                                                                          |
|                    | MLKL          | IL-1R (anakinra)?                                     |                                                                          |
| Chronic proliferative dermatitis | NLRP3      | MLKL (necrosulfonamide)                               |                                                                          |
|                    | IL-1R         | NLRP3 (MCC950)?                                       |                                                                          |
| Chronic proliferative dermatitis | IL-1β   | NLRP3 (MCC950)?                                       |                                                                          |

AIFEC, autoinflammation with infantile enterocolitis; DIRA, deficiency of IL-1R antagonist; HIDS, hyper IgD syndrome; MKD, mevalonate kinase deficiency; NLRP12AD, NLRP12-associated autoinflammatory disease; PAAND, Pyrin-associated autoinflammation with neutrophilic dermatoses; PAPA, pyogenic arthritis, pyoderma gangrenosum, and acne; PFIT, periodic fever, immunodeficiency, and thrombocytopenia; PGA, pediatric granulomatous arthritis; TRAPS, TNF receptor–associated periodic fever.

Place and Kanneganti
Inflammatory cell death and disease
The recently completed Canakinumab Antiinflammatory Thrombosis Outcomes Study (CANTOS), in which a large cohort of 10,061 patients was treated with canakinumab following myocardial infarction, found that neutralizing IL-1β–mediated inflammation reduced recurrent cardiovascular events (Ridker et al., 2017a). In atherosclerotic disease, it is thought that cell death, pyroptosis, and necroptosis lead to instability of plaques, which further suggests that blockade of these cell death pathways or their cytokine release may provide protection (Leeper, 2016; Xu et al., 2018c). Following the completion of the study, a second analysis of the CANTOS data found that canakinumab also had a significant effect on reducing lung cancer incidence and mortality, which was an unexpected finding from the trial (Ridker et al., 2017b). In the tumor microenvironment, IL-1–driven inflammation is thought to increase tumor invasiveness, metastasis, and growth (Apte et al., 2006; Gottschlich et al., 2018; Karki and Kanneganti, 2019). Together, these data provide compelling evidence that inhibiting IL-1β can have profound benefits in humans. While the protection from lung cancer was observed in patients with atherosclerosis, future studies in other patient populations will determine whether IL-1β inhibition can broadly protect from this devastating disease.

While IL-1β is often implicated in IL-1–driven diseases, the IL-1 family of proteins contains many other members including IL-1α, IL-18, IL-33, IL-36α/β/γ, IL-37, and IL-38 (Dinarello, 2018). Importantly, IL-1α and IL-1β both bind the IL-1R, but the role for IL-1α is poorly understood in most disease conditions. Treatments targeting the IL-1R or both IL-1α and IL-1β (as with IL-1R–blocking anakinra or the decoy IL-1R drug rilonacept) have shown great success, but future trials will examine whether targeting either IL-1α or IL-1β alone may provide additional benefits in specific disease models or reduce side effects of dual inhibition. An IL-1α–specific neutralizing antibody (bermekimab, MABp1) is currently in clinical trials to determine its effectiveness in multiple inflammatory diseases (Dinarello, 2018). Unlike IL-1β, IL-1α does not require caspase–1–mediated cleavage for its activation and can be released from pyroptotic or necroptotic cells. In a mouse model of neutrophilic dermatosis (via mutation in PTPN6), both RIPK1–driven up-regulation of inflammatory cytokines and IL-1α promote disease independently of inflammasome signaling and RIPK3, raising the question of whether MLKL or GSDMD may cooperate in some way to facilitate the release of IL-1α (Łukens et al., 2013). In patients with colorectal cancer, bermekimab–mediated neutralization of IL-1α appears to protect against chemotherapy-induced weight loss (Hickish et al., 2017; O’Sullivan Coyne and Burotto, 2017). In patients with TNF blockade–resistant severe hidradenitis suppurativa, bermekimab also seems to reduce symptoms (Kanii et al., 2018). Blockade of IL-1α may also protect patients from psoriasis and is being studied (Coleman et al., 2015). More studies will likely reveal an important role for IL-1α in driving many other diseases independent from inflammasome–driven production of IL-1β.

**IL-18–driven diseases**

The second key cytokine processed by caspase-1 and released by pyroptosis is IL-18, a key costimulatory cytokine of NK and T cell IFNγ responses (Dinarello, 2018). Interestingly, a subset of autoimmune inflammatory diseases associated with mutations in the pyroptosis machinery are poorly responsive to IL-1 blockade and better treated with IL-18 blockade. Excessive release of IL-18 in diseases, including Behçet’s disease, hemophagocytic histiocytosis (HLH) with macrophage activation syndrome, and NLRC4–mediated autoinflammation with infantile enterocolitis is thought to contribute to their pathology, suggesting that IL-18 blockade may be beneficial in treating these diseases (Masters, 2013; Schulert and Grom, 2015; Vitale et al., 2016; Romberg et al., 2017). Blockade of IL-18 may be beneficial in the context of colitis, inflammatory bowel disease, or cardiac disease, but conflicting findings on harmful and beneficial roles for IL-18 require more study (Zaki et al., 2010; Dinarello et al., 2013; Kanai et al., 2013; Toldo et al., 2014). IL-18 is also thought to contribute to the pathogenesis of a number of skin diseases, including psoriasis, atop dermatitis, and lupus, but more definitive studies are needed to determine whether inhibition with IL-18 binding protein (IL-18BP) will be beneficial (Lee et al., 2018). Blockade strategies include recombinant IL-18BP or neutralizing antibodies targeting IL-18. To date, it is not clear where IL-18 blockade or supplementation will be useful clinically, but ongoing clinical trials and mouse studies should reveal important contributions for this caspase-1–regulated cytokine.

**Pyroptosis–driven diseases**

While it has been understood for years that caspase-1 regulates pyroptosis and inflammasome–dependent cytokine release upon activation of specific inflammasome complexes, the recent discovery of GSDMD as the executioner of pyroptosis provides a new way of studying the relative contributions of both pyroptotic cell death and functional cytokine release. This is particularly important in light of many recent studies showing that the apoptotic, necroptotic, and pyroptotic pathways interact. Therapeutically, inhibition of GSDMD directly may prevent the release of cytokines as well as other alarmins released as a consequence of pyroptotic cell death that also contribute to disease progression, such as IL-1α, IL-33, HMGB1, or ATP. This is of particular interest in the treatment of sepsis, which in mouse models is critically dependent on caspase-11 and GSDMD, suggesting inhibitors may be able to effectively treat patients and block the release of many of the inflammatory mediators of septic shock (Kayagaki et al., 2015; Shi et al., 2015; Rathkey et al., 2018). Recent studies have identified two potential inhibitors of GSDMD that may hold therapeutic value. A GSDMD–derived peptide inhibitor, Ac-FLTD-CMK, inhibits pyroptosis by acting as an uncleavable substrate (Yang et al., 2018). Necrosulfonamide, an inhibitor that disrupts disulfide bonds in both GSDMD (Cys48) and human (but not murine) MLKL (Cys46), inhibits both pyroptosis and necroptosis, suggesting it may be useful to inhibit both cell death modalities in patients (Rathkey et al., 2018). In the brain, microglia are proposed to release ASC specks, which can seed amyloid-β plaques, suggesting that pyroptosis may promote Alzheimer’s...
| Disease                          | Key regulator | Treatment target | Reference                                      |
|---------------------------------|---------------|------------------|------------------------------------------------|
| Behcet’s disease                | IL-1, IL-18?  | IL-1R (anakinra) | Masters, 2013; Vitale et al., 2016            |
|                                 | IL-1β         | IL-1β (canakinumab) |                                               |
|                                 |               | IL-1 decoy receptor (rilonacept) |                                               |
| Sweet’s syndrome (including neutrophilic dermatoses) | PTPN6?, RIPK1, IL-1α, MLKL? | IL-1α? GSDMD? MLKL? | Nesterovitch et al., 2011; Lukens et al., 2013 |
| Schnitzler syndrome             | IL-1β         | IL-1R (anakinra) | de Koning et al., 2015                        |
|                                 | IL-18?        | IL-1β (canakinumab) |                                               |
|                                 |               | IL-1 decoy receptor (rilonacept) |                                               |
| AOSD                            | IL-1β         | IL-1R (anakinra) | Lee et al., 2015; Junge et al., 2017          |
|                                 | IL-18?        | IL-1β (canakinumab) |                                               |
|                                 |               | IL-1 decoy receptor (rilonacept) |                                               |
| JIA/soJIA                       | IL-1          | IL-1R (anakinra) | Pascual et al., 2005; Cimaz, 2016             |
|                                 | IL-1β         | IL-1β (canakinumab) |                                               |
|                                 |               | IL-1 decoy (rilonacept) |                                               |
| HLH                             | IL-1β, IL-18? | IL-1R (anakinra)? IL-18? | Schulert and Grom, 2015                        |
|                                 |               | IL-1 decoy receptor (rilonacept) |                                               |
| Osteomyelitis                   | IL-1β, NLRP3, Caspase-8, MLKL? | IL-1R (anakinra) MLKL? | Lukens et al., 2014; Gurung et al., 2016a     |
| Lung cancer                     | IL-1β         | IL-1β (canakinumab) | Ridker et al., 2017b                          |
| Cardiovascular diseases         | IL-1α?, IL-1β | IL-1β (canakinumab) | Ridker et al., 2017a                          |
|                                 |               | IL-1 decoy receptor (rilonacept) |                                               |
| Gout                            | NLRP3, MSU crystals, IL-1, GSDMD, MLKL? | IL-1R (anakinra) IL-1β (canakinumab) IL-1 decoy receptor (rilonacept) | Dayer et al., 2017 |
| Kidney diseases                 | IL-1β, IL-18, RIPK3, MLKL? GSDMD? | IL-1β? MLKL (necrosulfonamide) GSDMD? | Wu et al., 2008; Chen et al., 2018a; Sarhan et al., 2018b |
| Alzheimer’s disease             | NLRP3, ASC | NLRP3 (MCC950)? ASC? GSDMD? | Venegas et al., 2017                          |
| Ischemic brain injury           | NLRP3, NLRC4? GSDMD, RIPK1, RIPK3, MLKL? | RIPK1 (Necrostatin-1) RIPK3 (GSK-872) | Degterev et al., 2005; Chen et al., 2018b; Xu et al., 2018b; Poh et al., 2019; Zhang et al., 2019 |
| Rheumatoid arthritis            | TNF, IL-17, GSDMD, MLKL? | TNF | Yamanaka, 2015; Dayer et al., 2017            |
| Septic shock                    | Caspase-11, GSDMD, Caspase-8 | GSDMD? | Kayagaki et al., 2015; Kang et al., 2018; Mandal et al., 2018 |
| Inflammatory bowel diseases     | MLKL, NLRP3, IL-1α | NLRP3 (MCC950)? IL-1α (MABp1)? MLKL (necrosulfonamide) | Malik et al., 2016; Li et al., 2018 |

AOSD, adult-onset Still’s disease; HLH, hemophagocytic lympho-histiocytosis; JIA, juvenile idiopathic arthritis; soJIA, systemic-onset juvenile idiopathic arthritis.
Necroptosis-driven diseases

Compared with pyroptosis, where upstream caspase-1-mediated cleavage of its substrates IL-1β and IL-18 is required for their activity, necroptosis is not known to be associated with any specific processing of inflammatory mediators before cell lysis. Necroptosis is also generally considered a backup form of cell death when parts of the apoptotic pathway are inhibited by pathogen or inhibitors (Pasparakis and Vandenabeele, 2015). Interestingly, recent studies have shown that necroptosis (often determined by phospho-MLKL) plays a role in the pathogenesis of a number of diseases. Recent studies have shown that necroptosis promotes ischemic tissue damage. In kidneys, ischemia-reperfusion injury is promoted by RIPK3 and MLKL, apparently by regulating the cell death of renal proximal tubular cells, with subsequent recruitment of macrophages and NLRP3 inflammasome activation. Inhibitors targeting RIPK1, RIPK3, and MLKL appear to reduce this kidney damage, but MLKL deficiency alone does not necessarily protect from kidney reperfusion injury, suggesting there may be other interacting forms of cell death that drive injury (Linkermann et al., 2013; Newton et al., 2016; Anders, 2018; Chen et al., 2018a). In the liver, alcohol-induced damage was reduced in the absence of RIPK3 and MLKL, suggesting that necroptosis promotes disease (Roychowdhury et al., 2013; Wang et al., 2016). During spinal injury, it has also been proposed that microglia undergo necroptosis and that treatment with nercosulfonamide can reduce phospho-MLKL and pathology (Fan et al., 2015; Huang et al., 2018; Wang et al., 2018). In the mouse Sharpin−/−dm model, skin inflammation is driven by necroptosis via TNF, RIPK1, RIPK3, MLKL, and downstream inflammasome-mediated IL-1β (Kumari et al., 2014; Rickard et al., 2014; Gurung et al., 2016b). Together, these studies suggest that necroptosis can initiate and drive multiple inflammatory diseases and may be an effective target for future therapies.

Implications of cell death crosstalk

Pyroptosis and necroptosis both lead to lytic cell death after stimulation with inflammatory mediators such as pathogens, alarmins, and cytokines such as TNF. Genetic studies have shown that necroptotic signaling via RIPK3, MLKL, NLRP3, and caspase-1 leads to release of mature IL-1β independent of GSDMD (Moriwaki et al., 2015; Conos et al., 2017; Gutierrez et al., 2017). Mechanistically, this occurs through MLKL-mediated plasma membrane disruption, efflux of potassium ions, and subsequent NLRP3 inflammasome-mediated maturation of IL-1β that can be released by necroptosis. In inflamed tissues, this likely contributes significantly to the development of inflammatory diseases. Earlier studies also hinted at crosstalk between apoptotic and pyroptotic pathways, finding that caspase-8 can drive inflammasome activation (Man et al., 2013; Gurung et al., 2014, 2016a). During infections or under the condition of TAK1 inactivation, multiple cell death pathways can be engaged to prevent pathogen-mediated inhibition of cell death and facilitate effective immune responses (Kuriakose et al., 2016; Malireddi et al., 2018; Orning et al., 2018; Sarhan et al., 2018a; Xu et al., 2018a). Similarly, loss of TBK1 results in spontaneous cell death that is driven by RIPK1 (Xu et al., 2018a).

During multiple viral infections, the sensor ZBP1 coordinates parallel cell death pathways of necroptosis, apoptosis, and pyroptosis with potential to regulate cell death during other currently known diseases (Kuriakose et al., 2016; Koehler et al., 2017; Maelfait et al., 2017; Guo et al., 2018). In physiological conditions, it is not surprising that RIPK1- and RIPK3-mediated expression of inflammatory cytokines or induction of necroptosis indirectly promote further downstream apoptosis, necroptosis, or pyroptosis in other nearby cells. In cancer, for example, changes in the apoptotic death pathways likely change the form of cell death that occurs downstream of TNF or other inflammatory signals. A recent study showed that in colorectal cancer cells, lobaplatin treatment induces a unique caspase-3–mediated cleavage of GSDME, a pore-forming protein related to GSDMD, inducing a pyroptosis-like cell death (Yu et al., 2019). In endotoxic shock, recent work has shown that the combined action of caspase-11 and caspase-8, independent of RIPK1 kinase activity and RIPK3, drive endotoxin-mediated pathology in the small intestine and spleen. In addition to these key molecules, production of TNF and type I IFNs also promoted caspase-8–dependent apoptosis in the endotoxic shock model, highlighting an important role for both pyroptosis and apoptosis (Mandal et al., 2018). This highlights the importance of understanding which particular cell death pathways are involved in each unique disease scenario (Yu et al., 2019).

A key problem in treating multifactorial diseases such as chronic inflammatory bowel disease, rheumatoid arthritis, psoriasis, and many others is identifying which steps of the inflammatory process initiate and promote disease. The relative success of TNF blockade in treating many of these diseases suggests that TNF-driven inflammation and likely TNF-mediated apoptosis help to drive the disease, but some patients eventually need to discontinue TNF blockade to avoid serious infectious complications (Ko et al., 2009; Ali et al., 2013; Yamanaka, 2015; Katsanos et al., 2018). Understanding whether TNF promotes increased apoptosis, necroptosis, pyroptosis, and cytokine release to promote disease may suggest alternative treatments. In ischemic brain injury, NLRC4, NLRP3, GSDMD, RIPK1, RIPK3, and MLKL have all been proposed to promote pyroptosis and necroptosis in microglia or neuronal cells, and necrostatin-1 (an inhibitor of RIPK1) has been shown to ameliorate injury, but the role of each death pathway needs closer...
study (Degterev et al., 2005; Xu et al., 2018b; Poh et al., 2019; Zhang et al., 2019). In a model of osteomyelitis, mutations in PSTPIP2 lead to excessive TNF, osteoclastogenesis, and neutrophil recruitment that destroys bone in a caspase-8-, NLRP3-, and IL-1β-dependent manner, suggesting that crosstalk between the apoptotic, necroptotic, and pyroptotic pathways is occurring (Lukens et al., 2014; Gurung et al., 2016a). Whether the executioner molecules GSDMD or MLKL also play a role in promoting PSTPIP2-mediated bone disease remains to be seen. These cytokines and pathways also likely promote inflammation associated with rheumatoid arthritis of the joints (Jung et al., 2014; Yamanaka, 2015). Cancers are often driven by proinflammatory cytokine production, so future work may reveal interesting roles for the executioners MLKL and GSDMD in promoting tumor growth, invasiveness, and metastasis (Apte et al., 2006; Ridker et al., 2017b). Identifying the most upstream pathway in each of these diseases will help determine which treatments may be best for preventing or treating them.

Future perspectives
Better understanding of the molecular mechanisms regulating apoptosis, pyroptosis, necroptosis, and the executioner molecules of each pathway may lead to better treatment of inflammatory diseases driven by the release of inflammatory cytokines and alarmins. The extensive crosstalk that researchers have observed between each of these pathways also suggests that it will be difficult to specifically target the appropriate cell death pathway without first understanding the critical regulators of individual diseases. Indeed, when cells receive a lethal trigger, they will often set off multiple overlapping signal cascades that lead to a switch in cell death when one key molecule is inhibited, as exemplified by necroptosis. In addition to understanding which forms of cell death promote a particular disease, it will be important to understand how the release of inflammatory cytokines and alarmins also contributes. A major concern with any immunosuppressive approach targeting necroptosis or pyroptosis is that patients may become susceptible to opportunistic infections, so carefully balancing the immunosuppression with this risk is critical.

Emerging work on sublethal pyroptosis and necroptosis, where membrane repair through ESCRT negatively regulates cell lysis and cytokine release, presents an interesting new physiological relevance of these alternative pathways is unclear and requires further study. Armed with our current understanding of these pathways, novel therapeutics specifically targeting pyroptosis- and necroptosis-mediated cell death, their upstream regulators, the apoptosis pathway, and the availability of cytokine-neutralizing antibodies hold great promise to treat a wide range of inflammatory diseases.

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