Research Article

To Explore the Mechanism and Equivalent Molecular Group of Fuxin Mixture in Treating Heart Failure Based on Network Pharmacology

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1. Introduction

Heart failure (HF) is a closed result of most cardiovascular diseases, which may be caused by damage to ventricular filling from various structural or functional disorders of the heart [1]. In observational data from Europe, the 1-year all-cause mortality rate for HF was greater than 20 percent [2]. In search of a cheap, safe, and effective drug, we turned to traditional Chinese medicine (TCM).

FXHJ is a prescription for the treatment of heart failure. It has been shown to be effective in clinical trials, but its active ingredients and mechanism of action are not completely clear, which limits its clinical application and international promotion. In this study, we used network pharmacology to find, conclude, and summarize the mechanism of FXHJ in the treatment of heart failure. From FXHJ, we found 39 active ingredients and 47 action targets. Next, we constructed the action network and was conducted enrichment analysis. The results showed that FXHJ mainly treated heart failure by regulating the MAPK signaling pathway, PI3K/Akt signaling pathway, cAMP signaling pathway, TNF signaling pathway, toll-like receptor signaling pathway, VEGF signaling pathway, NF-kappa B signaling pathway, and the apoptotic signaling molecule BCL2. Through the research method of network pharmacology, this study summarized the preliminary experiments of the research group and revealed the probable mechanism of FXHJ in the treatment of heart failure to a certain extent, which provided some ideas for the development of new drugs.
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2. Methods

2.1. Data Collection and Processing. Chinese medicine systematic pharmacology database (TCMSP) (http://tcmspw.com/tcmsp.php) is an open and comprehensive database of Chinese medicine ingredients and action targets. In clinical treatment, TCM is often used by oral administration. Oral bioavailability (OB) and drug-likeness (DL), two ADME-related models, are the main variables affecting the absorption of drugs from the gastrointestinal tract. Therefore, we screened bioactive components under the conditions of OB ≥ 30% and DL ≥ 0.18. Next, we find the target from the list of compounds. At the same time, we used the PubChem database (https://pubchem.ncbi.nlm.nih.gov/) to standardize the names of active ingredients. In order to make the results concise and convenient, we utilized the Uniprot database (https://www.uniprot.org/) to convert the protein name of the target into the gene name. Then, we take the union of all the consequences and delete the duplicates. In order to get the disease target of heart failure, we used “heart failure” as the keyword to retrieve the target of heart failure in the Genecards database (https://www.genecards.org/). After the intersection of the target of FXHJ and the target of heart failure, we obtained the exact target of FXHJ in the treatment of heart failure.

2.2. Network Construction. We constructed the network diagram of “herb-compound-target” of FXHJ and the equivalent molecular group by Cytoscape 3.6.1. In the network diagram, “node” refers to the compound or target, and “edge” refers to the relationship between the compound and the target. Based on the analysis of the network diagram of the equivalent molecule group, we select the compound whose degree is greater than the average as the main component of the equivalent molecule group.

2.3. Enrichment Analysis. Direct targets were recorded into David 6.8 database (https://david.ncifcrf.gov/summary.jsp) to obtain KEGG signaling pathway data. We analyzed the results with P value less than 0.05 to obtain the results. According to the enrichment results and the preceding experimental results, we obtained the functional pathway of FXHJ in the treatment of heart failure. After that, we screened the effective targets and components in reverse according to the action pathway and obtained the equivalent molecular group.

2.4. Molecular Docking. AutoDock Vina_1.1.2 software was used to conduct molecular docking between major components and key targets of potential pathways so as to ensure their interaction activity. AutoDock Vina USES semiflexible molecular docking. That is, the pharmacophore is flexible while the protein remains rigid during the docking. The docking results are evaluated by semiempirical free energy function.

2.5. Specific Steps

(1) In the PubChem website (https://pubchem.ncbi.nlm.nih.gov/), download the 3d structure of the active ingredient file (“SDF” format). Open Babel is used to hydrogenate atoms in molecular structure. Select the MMFF94 field to add the charge and minimize energy. Finally, AutoDock Tools are utilized to convert the compound into a PDBQT format file.

(2) Download the crystal structure of the key targets molecule from the PDB website (http://www.rcsb.org/).

(3) AutoDock Tools were used to separate the target protein and its ligand, add hydrogen atom, calculate the electric charge, and export it to the PDBQT format file. AutoDock Tools were used to identify the size and center of the docking box. The center of the docking box was defined as the center of the protein crystal structure of the original ligand, and the size of the docking box included the key residues in the active site of the original ligand.

(4) At last, Vina was utilized to connect the active ingredients with the target protein in turn, and Affinity was extracted. PyMol was utilized to analyze and plot the results.
3. Result

3.1. FXHJ Data Collection. From the TCMSP database, we can get that DG has 2 blood active components and 67 predicted targets. FZ has 21 blood active components and 32 predicted targets. HB has 27 blood active components and 93 predicted targets. TLZ has 12 blood active components and 123 predicted targets. YHY has 23 blood active components and 140 predicted targets. ZX has 10 blood active components and 9 predicted targets. After adding up and deleting the duplicates of 6 Chinese herbs, we obtained 64 and 157 targets of blood components. We searched the Genecards database and obtained 10,961 targets for heart failure. Next, we took the intersection of TCM action targets and heart failure targets and obtained 144 targets. Meanwhile, we constructed the action network of FXHJ as shown in Figure 1.

3.2. Enrichment Analysis of Data. The enrichment analysis showed that the key target of FXHJ in the treatment of heart failure was concentrated in 113 pathways, among which 104 were with \(P\) value less than 0.05. The top 20 bits of the enrichment results are shown in Figure 2. In addition to the pathways and targets previously demonstrated by the team, the results showed that the toll-like receptor signaling pathway, VEGF signaling pathway, and NF-kappa B signaling pathway might be potential pathways for the treatment of heart failure.

3.3. Construction of Equivalent Molecular Clusters. According to the enrichment analysis results, 5 Chinese herbs, 39 compounds, and 47 action targets were reverse-screened. We constructed the action network diagram of the equivalent molecular group, as shown in Figure 3. From the analysis of the network diagram, we can find that the main components of the equivalent molecular group are 17 compounds. We ranked these compounds by degree value from large to small as quercetin, Kaempferol, Luteolin, pe-sitosterol, 2, 7-dihydrohomorsotrine, Isocorypalmine, Protopine, Stigmasterol, 8-(3-methylbut-2-enyl)-2-phenylchromen-4-one, Cavidine, Anhydroicaritin, Rutaceolanine, Phelloorpentin, Liquiritigenin, berberine, palmatine, and isorhamnatin.

3.4. Molecular Docking Results. AutoDock Vina evaluates the binding ability of small molecules to proteins, mainly by affinity. Affinity less than 0 indicates that the ligand can spontaneously bind to the receptor, and the smaller the value is, the higher the binding energy is, and the easier the active component is to bind to the receptor.

We select five key targets from toll-like receptor signaling pathway, VEGF signaling pathway, and NF-kappa B signaling pathway according to the intersection of the pathway and degree value, namely VEGFA (1VPF), TNF (2AZ5), PIK3CG (2CHZ), PTGS2 (5KIR), and MAPK1 (SNHV). The docking results are shown in Table 1.

We visualized the combination of ligand and receptor with the highest Affinity, as shown in Figure 4. From the figure, we can see that the compound enters the target protein target active site and its binding pattern.

4. Discussion

In the treatment of heart failure, how to slow or even reverse ventricular remodeling has grown up to be a hot topic in recent years. Transforming growth factor beta, angiotensin II, aldosterone and endothelin, and myocardial damage or stress produce cytokines and chemokines can induce the formation of muscle fibroblasts [10, 11]. In the early stage, we have studied the relevant pathways, including the MAPK signaling pathway, the PI3K/Akt signaling pathway, the cAMP signaling pathway, the TNF signaling pathway, and the apoptotic conduction molecule BCL2.

There are three subclasses in the MAPK signaling pathway, namely ERK, JNK, and p38MAPK. Cipolletta et al. demonstrated that the inhibition of p-ERK volume could delay cardiac hypertrophy and improve cardiac volume [12]. Our previous experiments showed that FXHJ could significantly reduce the ERK level of rat cardiac tissue compared with the model group (\(P < 0.01\)) [13].

The active form of PI3K can activate AKT, thereby promoting cell proliferation and inhibiting the apoptotic pathway. Overexpression of PI3K can cause cardiac hypertrophy. The GSK3\(\beta\) cytokine downstream of PI3K/Akt, which can be inactivated by phosphorylation, is a key negative regulator of cardiac hypertrophy [14]. Our previous experiments have shown that FXHJ can reduce the expression of PI3K and AKT, improve the expression of GSK3, and thus inhibit ventricular remodeling [15].

CAMP ACTS as excitation-coupled phosphorylation of cardiomyocyte proteins, including l-type calcium channels, sarcoplasmic reticulum ATPase 2 regulatory protein phosphor, clonidine receptor 2, phosphatase 1 inhibitors, and various contractile proteins [16]. Lin B et al. demonstrated that activation of the cAMP signaling pathway could stabilize cardiac function in rats with heart failure [17]. We studied the \(\beta1-AR-cAMP-PKA\) pathway in previous experiments and found that compared with the model group, both the FXHJ group and the captorpril group could reduce the plasma cAMP level, increase the cAMP reserve of cardiomyocytes, improve cardiac function, and delay ventricular remodeling [18].

Tumor necrosis factor \(\alpha\) (TNF-\(\alpha\)) and the apoptotic signaling molecule BCL2 play important roles in the progression of heart failure. Liu W et al.’s serum examination of patients with chronic heart failure found that, compared with the healthy control group, the serum TNF in patients with heart failure was significantly increased, while bcl-2 was significantly decreased [19]. Studies have shown that increased TNF-\(\alpha\) levels exacerbate heart failure. The optimal use of diuretics, ACE inhibitors, beta-blockers, and standard treatment with digoxin for heart failure can significantly reduce circulating TNF levels [20]. Siltanen A’s experiments showed that BCL2 gene transplantation in rats with chronic heart failure could effectively improve cardiac function,
enhance paracrine angiogenic signals, and promote cell proliferation and survival in failing myocardium [21]. Our previous animal experiments showed that FXHJ could effectively reduce the level of TNF-α in rats with heart failure, improve the expression of the BCL2 gene, and thus inhibit a variety of myocardial cell apoptosis pathways [22, 23].

In addition, through the research method of network pharmacology, we analyzed and found that the mechanism of action of FXHJ in the treatment of heart failure may also be related to the toll-like receptor signaling pathway, VEGF signaling pathway, and NF-kappa B signaling pathway. Studies have shown that 8 weeks after knocking out NF-kappa B subunit P50/NF–B1 in mice with myocardial infarction, the degree of ventricular expansion is mild, and ventricular systolic function is preserved. Therefore, the absence of the NF-kappa B subunit P50 can improve the prognosis of heart failure caused by myocardial infarction. At the same time, inhibition of the NF-kappa B signaling pathway can inhibit ventricular remodeling to a certain extent, which may be related to the reduction of preinflammatory response and the regulation of extracellular matrix [24]. Inhibiting the activation of NF-kappa B65 can prevent myocardial fibrosis and protect cardiac function by reducing the inflammatory response [25]. Vascular endothelial growth factor (VEGF) plays an important role in mediating normal cardiac function by maintaining vascular homeostasis. VEGF maintains vascular homeostasis mainly through the following mechanisms: Improve the sensitivity

Figure 1: Function network diagram of FXHJ. The red triangle represents the Chinese herbs, the blue oval represents the compound, and the green diamond represents the target.
of blood vessels to nerve response, improve the permeability of blood vessels, promote the generation and stability of new blood vessels, and recruit stem cells and promote their homing [26]. The research of Meng-Ying He showed that when the VEGF signaling pathway was inhibited, it would lead to the imbalance of vascular homeostasis, which would lead to the generation of heart failure [27]. Toll-like receptor signaling pathway mediates the occurrence of various myocardial injuries, such as myocardial remodeling and myocardial ischemia reperfusion injury [28]. Animal studies have shown that the symptoms of myocardial ischemia in mice with toll-like receptor knockout are reduced. At the same time, toll-like receptor inhibitors could reduce myocardial ischemia in myocardial ischemia model mice [29, 30].

The analysis of the equivalent molecular group shows that there is no ZX in it. This is because the active components of ZX are mainly Alisol A24-acetate and Alisol B, which have similar structures to aldosterone and its antagonists. They compete for aldosterone receptors to inhibit reabsorption in different parts of the tubules, inhibiting water reabsorption and increasing urine output [31]. This belongs to the category of diuretics, which is different from the screening scope of this study.

Some of the components in the equivalent molecular group have been studied experimentally, which can provide some reference for the further selection of the components. Moon DO et al. demonstrated that β-sitosterol promotes apoptosis by regulating the ERK and PI3K/Akt signaling pathways [32], which is not conducive to the treatment of heart failure. Animal experiments on Kaempferol have shown that Kaempferol can treat heart failure by inhibiting the MAPK signaling pathway, NF-kappa B signaling pathway, and PI3K/Akt signaling pathway, activating the VEGF signaling pathway and upregulating the expression of GSK3β [33–37]. Min Z’s experiment proved that quercetin could significantly reduce the phosphorylation of ERK and inhibit myocardial cell fibrosis [38]. Meanwhile, quercetin...
can also treat heart failure by upregulating the expression of GSK3β factor and cAMP and inhibiting the expression of the NF-kappa B signaling pathway [39–41]. Although animal studies and clinical trials have shown that Luteolin can improve myocardial fibrosis and protect cardiac function by reducing myocardial oxidative stress [42, 43], other studies have shown that Luteolin can affect angiogenesis by inhibiting the VEGF signaling pathway [44, 45]. ©fl_his suggests that more trials are needed to confirm whether Luteolin has side effects in patients with heart failure. Rutaecarpine has been shown to protect cardiomyocytes by inhibiting the NADPH oxidase-ROS pathway [46], a mechanism not identified in our study. Similarly, animal studies have shown that berberine can protect cardiac myocytes by upregulating the expression of BCL2 and activating AMPK and eNOS signaling pathways [47, 48]. Gao L’s experiment showed that Isorhamnetin could inhibit cardiac hypertrophy by blocking the pi3k-akt signaling pathway [49].

Results of molecular docking showed that the Affinity of ligand and receptor was higher. This indicated that the ligand could well bind to the receptor, thus inhibiting the expression of this pathway. As mentioned above, inhibiting these three pathways can delay the progression of heart failure.

Interestingly, it can be evident from Figure 2 that the enrichment of tumor-related pathways is relatively high. Most of the genes enriched in these pathways are related to cell proliferation, survival, angiogenesis, and inflammatory factors, mainly involving genes such as PIK3CG, MAPK1, BCL2, NOS3, EGF, etc. These genes have been linked to
Studies have shown that tumor growth is associated with increased levels of inflammatory cytokines and a significant decrease in ventricular systolic function. We know that heart failure has the same risk factors as cancer. Patients with heart failure have a higher rate of cancer than healthy control populations. We also know that modern anticancer therapies are often associated with cardiotoxicity, which exacerbates or accelerates acute or chronic heart failure. It may be that a combination of traditional Chinese medicine can reduce the risk of cancer in patients with heart failure, but more experiments and studies are needed to confirm this view.

However, there were a few shortcomings in this study. Due to limited funds, this study did not obtain the blood composition of FXHJ by mass spectrometry but selected database screening, which may cause certain errors.

Table 1: The figures in the table represent the affinity of molecular docking (kcal/mol). The red figure indicates that the compound has the highest affinity to the target.

| Compound                                      | VEGFA | TNF  | PIK3CG | PTGS2 | MAPK1 |
|-----------------------------------------------|-------|------|--------|-------|-------|
| Protopine                                     | −7.8  | −9.2 | −9.4   | −5.4  | −9.5  |
| Cavidine                                      | −7.4  | −8.4 | −8.6   | −5.1  | −9.3  |
| Rutaecarpine                                  | −7.8  | −9.8 | −9.2   | −8.9  | −9.1  |
| 8-(3-Methylbut-2-enyl)-2-phenylchromen-4-one | −7.5  | −8.7 | −9.6   | −10.7 | −9    |
| Luteolin                                      | −7.2  | −7.8 | −8.8   | −9.7  | −8.6  |
| Quercetin                                     | −7.2  | −7.2 | −7.7   | −9.7  | −8.6  |
| Isoquercetin                                   | −7.2  | −7.3 | −7.8   | −9.7  | −8.5  |
| Berberine                                     | −7.2  | −8.6 | −8.5   | −6    | −8.5  |
| Liquiritigenin                                | −7.3  | −7.6 | −8.8   | −9.3  | −8.4  |
| Kaempferol                                    | −6.9  | −6.9 | −7.5   | −9.8  | −8.3  |
| Stigmasterol                                  | −6.9  | −8.3 | −8.3   | −7.9  | −8.2  |
| Phellopterin                                  | −6.5  | −7.4 | −8     | −8.3  | −8.1  |
| Palmatine                                     | −6.2  | −8  | −7.5   | −5.2  | −7.8  |
| Isohernandiol                                 | −7    | −7.8 | −7.7   | −4.9  | −7.7  |
| Anhydroicaritin                               | −7.7  | −8  | −8.6   | 6.6   | −7.5  |
| 2,7-Dihydrohomoerysotrine                    | −6    | −7.4 | −7.1   | −2.8  | −6.3  |

Figure 4: Number 1 is the combination of proptopine and VEGFA. Number 2 is the combination of Rutaecarpine and VEGFA. Number 3 is the combination of Rutaecarpine and TNF. Number 4 is the combination of 8-(3-Methylbut-2-enyl)-2-phenylchromen-4-one and PIK3CG. Number 5 is the combination of 8-(3-Methylbut-2-enyl)-2-phenylchromen-4-one and PTGS2. Number 6 is the combination of protopine and MAPK1. Among them, the solid blue line represents a hydrogen bond, the dotted yellow line represents a salt bridge, the dotted gray line represents hydrophobic interaction, and the dotted green line represents pi-Stacking between benzene rings.
Secondly, although animal experiments and clinical trials have shown that FXHJ has a beneficial therapeutic effect, there is still a lack of experimental research on some components. Further research is needed to determine whether these components will cause side effects.

In summary, this study revealed the mechanism of FXHJ in the treatment of heart failure through the research method of network pharmacology and summarized the previous experimental results of the research group. It wishes to point out the direction for new drug development and further research. At the same time, the concept of the equivalent molecular group provides innovative ideas for the transformation and research of the modernization of TCM.

Data Availability
The data used to support the findings of this study are available from the corresponding author upon request.

Disclosure
The authors report no conflicts of interest in this work.

Conflicts of Interest
The authors declare that they have no conflicts of interest.

Authors’ Contributions
YDY drafted the manuscript, contributed to data analysis, and revised the final manuscript. YPX and YFL examined the data collection and processing process. YTX provided advice during the study and manuscript preparation. All authors read and approved the final manuscript.

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References
[1] S. A. Hunt, W. T. Abraham, M. H. Chin et al., “ACC/AHA 2005 guideline update for the diagnosis and management of chronic heart failure in the adult—summary article,” *Journal of the American College of Cardiology*, vol. 46, no. 6, pp. 1116–1143, 2005.
[2] A. P. Maggioni, U. Dahlström, G. Filippatos et al., “EURObservationalResearchResearch programme: regional differences and 1-year follow-up results of the heart failure pilot survey (ESC-HF pilot),” *European Journal of Heart Failure*, vol. 15, no. 7, pp. 808–817, 2013.
[3] X. Yitao, “Effect of Fuxin mixture on the expression of beta_1-AR mRNA in lymphocytes of patients with chronic congestive heart failure and clinical observation,” *China Journal of Traditional Chinese Medicine*, vol. 33, no. 3, pp. 533–536, 2015.
[4] A. L. Hopkins, “Network pharmacology: the next paradigm in drug discovery,” *Nature Chemical Biology*, vol. 4, no. 11, pp. 682–690, 2008.
[5] J. Fang, C. Liu, Q. Wang, P. Lin, and F. Cheng, “In silico polypharmacology of natural products,” *Brief Bioinformation*, vol. 19, no. 6, pp. 1153–1171, 2018.
[6] M. Jafari, Y. Wang, A. Amiryousefi, and J. Tang, “Unsupervised learning and multipartite network models: a promising approach for understanding traditional medicine,” *Frontiers in Pharmacology*, vol. 11, p. 1319, 2020 PMID: 32982738; PMCID: PMC7479204.
[7] Y. Wang, M. Jafari, Y. Tang, and J. Tang, “Predicting Meridian in Chinese traditional medicine using machine learning approaches,” *PLOS Computational Biology*, vol. 15, no. 11, e1007249 pages, 2019, PMID: 31765369; PMCID: PMC6876772, Article ID e1007249.
[8] M. Kibble, N. Saarinen, J. Tang, K. Wennerberg, S. Mákelä, and T. Aittokallio, “Network pharmacology applications to map the unexplored target space and therapeutic potential of natural products,” *Natural Product Reports*, vol. 32, no. 8, pp. 1249–1266, 2015.
[9] H. Yuan, Q. Ma, H. Cui et al., “How can synergism of traditional medicines benefit from network pharmacology?” *Molecules*, vol. 22, no. 7, p. 1135, 2017.
[10] Y. Sun, “Myocardial repair/remodelling following infarction: roles of local factors,” *Cardiovascular Research*, vol. 81, no. 3, pp. 482–490, 2009.
[11] M. Devalray, H. Osman, S. Kaiser, and D. Diviani, “The role of cyclic AMP signaling in cardiac fibrosis,” *Cells*, vol. 9, no. 1, p. 69, 2019.
[12] E. Cipolletta, M. R. Rusciano, A. S. Maione et al., “Targeting the CaMKII/ERK interaction in the heart prevents cardiac hypertrophy,” *Plos One*, vol. 10, no. 6, e0130477 pages, Article ID e0130477, 2015.
[13] Yi-tao Santulli, “Effects of fuxin mixture on the signal pathway of raf-mek-erk in rats with heart failure,” *Chinese Journal of Traditional Chinese Medicine*, vol. 33, no. 4, pp. 791–794, 2015.
[14] X. Yitao, “Relationship between heart failure and PI3K-Akt-GSK3β pathway and progress in traditional Chinese medicine,” *Chinese Medicine Emergancy*, vol. 25, no. 6, pp. 1066–1069, 2015.
[15] Yi-tao Xue, “Effects of fuxin decoction on the pathway of pi3k-akt-gsk3β in heart failure rats,” *Shanghai Journal of Traditional Chinese Medicine*, vol. 49, no. 8, pp. 74–78, 2015.
[16] C. C. Sucharov, S. J. Nakano, D. Slavov et al., “A PDE3A promoter polymorphism regulates cAMP-induced transcriptional activity in failing human myocardium,” *Journal of the American College of Cardiology*, vol. 73, no. 10, pp. 1173–1184, 2019.
[17] B. Lin, D. G. Feng, and J. Xu, “microRNA-665 silencing improves cardiac function in rats with heart failure through activation of the CAMP signaling pathway,” *Journal of Cellular Physiology*, vol. 234, no. 8, pp. 13169–13181, 2019.
[18] Y.-t. Xu, “Effects of fuxin mixture on the _1-ar-camp-pka pathway in rats with heart failure,” *Chinese Journal of Integrated Traditional and Western Medicine*, vol. 37, no. 4, pp. 453–457, 2017.
[19] W. Liu, L. Ru, C. Su, S. Qi, and X. Qi, “Serum levels of inflammatory cytokines and expression of BCL2 and BAX mRNA in peripheral blood mononuclear cells and in patients with chronic heart failure,” *Medical Science Monitor*, vol. 25, pp. 2633–2639, 2019.
[20] B. Bozkurt, D. L. Mann, and A. Deswal, “Biomarkers of inflammation in heart failure,” *Heart Failure Reviews*, vol. 15, no. 4, pp. 331–341, 2010.
[21] A. Siltanen, K. Kitabayashi, T. Pätilä et al., “Bel-2 improves myoblast sheet therapy in rat chronic heart failure,” *Tissue Engineering Part A*, vol. 17, no. 1-2, pp. 115–125, 2011.
Evidence-Based Complementary and Alternative Medicine

[22] X. Yitao et al., “Effects of Fuxin decoction on myocardial cytokines TNF-α and BNP in rats with heart failure,” *Journal of Integrated Traditional Chinese and Western Medicine Cardio-Cerebrovascular Disease*, vol. 11, no. 7, pp. 833-834, 2013.

[23] X. Yitao, “Effects of fuxin decoction on expression of bcl-2, a signal transduction molecule, in cardiac myocytes of rats with heart failure,” *China Journal of Traditional Chinese Medicine*, vol. 31, no. 12, pp. 2717–2720, 2013.

[24] J.-L. Jin and J.-R. Wei, “Research progress on the relationship between Notch signaling and NF-kappa B signaling pathway and ventricular remodeling after myocardial infarction,” *Chinese Journal of Circulation*, vol. 30, no. 07, pp. 718–720, 2015.

[25] J. E. Toblli, G. Cao, C. Rivas, J. F. Giani, and F. P. Dominici, “Intravenous iron sucrose reverses anemia-induced cardiac remodeling, prevents myocardial fibrosis, and improves cardiac function by attenuating oxidative/nitrosative stress and inflammation,” *International Journal of Cardiology*, vol. 212, pp. 84–91, 2016.

[26] Z. Taimeh, J. Loughran, E. J. Birks, and R. Bolli, “Vascular endothelial growth factor in heart failure,” *Nature Reviews Cardiology*, vol. 10, no. 9, pp. 519–530, 2013.

[27] M.-Y. Bolli, Z.-W. Yin, Y.-R. Zhao, H.-P. Li, W. Zheng, and J.-G. Jiang, “Mir-320a mediates adriamycin cardiac injury by targeting VEGF signaling pathway,” *Chinese Journal of Molecular Cardiology*, vol. 18, no. 05, pp. 2594–2599, 2008.

[28] E. M. Y. Moresco, D. LaVine, and B. Beutler, “Toll-like receptors,” *Current Biology*, vol. 21, no. 13, pp. R488–R493, 2011.

[29] C.-F. Liang, J. T. Liu, Y. Wang, A. Xu, and P. M. Vanhoutte, “Toll-like receptor 4 mutation protects obese mice against endothelial dysfunction by decreasing NAPDH oxidase isoforms 1 and 4,” *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 33, no. 4, pp. 777–784, 2013.

[30] Yu-J. Xu and W. Huang, “Research progress of toll-like receptor 2, 4 and myocardial ischemia-reperfusion injury,” *Advances in Cardiology*, vol. 34, no. 2, p. 240, 2013.

[31] Yu-hui Zhao, “Research progress on the chemical constituents and diuretic mechanism of fuling, fuling skin, polyphenol and alisma,” *Chinese Journal of Pharmacology and Toxicology*, vol. 28, no. 04, pp. 594–599, 2014.

[32] D.-O. Moon, K.-J. Lee, Y. H. Choi, and G.-Y. Kim, “Betaδ-Sitosterol-induced-apoptosis is mediated by the activation of ERK and the downregulation of Akt in MCA-102 murine fibrosarcoma cells,” *International Immunopharmacology*, vol. 7, no. 8, pp. 1044–1053, 2007.

[33] F. Zhang and C. Ma, “Kaempferol suppresses human gastric cancer SNU-216 cell proliferation, promotes cell autophagy, but has no influence on cell apoptosis,” *Brazilian Journal of Medical and Biological Research*, vol. 52, no. 2, Article ID e7843, 2019.

[34] Z. Zhuang, G. Ye, and B. Huang, “Kaempferol alleviates the interleukin-1β-induced inflammation in rat osteoarthritis chondrocytes via suppression of NF-κB,” *Medical Science Monitor*, vol. 23, pp. 3925–3931, 2017.

[35] H.-Y. Wang, J.-G. Zhao, Z.-G. Wei, and Y.-Q. Zhang, “The renal protection of flavonoid-rich ethanolic extract from silkworm green cocoon involves in inhibiting TNF-α-p38 MAP kinase signalling pathway in type 2 diabetic mice,” *Biomedicine & Pharmacotherapy*, vol. 118, p. 109379, 2019.

[36] L. Zhang, Z. Guo, Y. Wang, J. Geng, and S. Han, “The protective effect of kaempferol on heart via the regulation of Nrf2, NF-κβ, and PI3K/Akt/GSK-3β signaling pathways in isoproterenol-induced heart failure in diabetic rats,” *Drug Development Research*, vol. 80, no. 3, pp. 294–309, 2019.

[37] H. K. Chin, C. T. Horng, Y. S. Liu et al., “Kaempferol inhibits angiogenic ability by targeting VEGF receptor-2 and down-regulating the PI3K/AKT, MEK and ERK pathways in VEGF-stimulated human umbilical vein endothelial cells,” *Oncology Reports*, vol. 39, pp. 2351–2357, 2018.

[38] Z. Lu, L. Yangchan, W. Juqian, and Z. Changying, “Quercetin inhibition of myocardial fibrosis through regulating MAPK signaling pathway via ROS,” *Pakistan Journal of Pharmaceutical Sciences*, vol. 32, no. 3, pp. 1355–1359, 2019.

[39] A. Tripathi, M. Kumar, P. Kaur, B. Kumar, and S. S. K. Sagi, “Efficacy of Quercetin as a potent sensitizer of β2-AR in combating the impairment of fluid clearance in lungs of rats under hypoxia,” *Respiratory Physiology & Neurobiology*, vol. 273, Article ID 103334, 2020.

[40] S.-C. Cheng, W.-C. Huang, J.-H. S. Pang, Y.-H. Wu, and C.-Y. Cheng, “Quercetin inhibits the production of IL-1β-induced inflammatory cytokines and chemokines in ARPE-19 cells via the MAPK and NF-xB signaling pathways,” *International Journal of Molecular Sciences*, vol. 20, no. 12, p. 2957, 2019.

[41] R. Gupta, R. K. Shukla, A. Pandey et al., “Involvement of PKA/ DARPP-32-PP1α and β-arrestin/akt/GSK-3β signaling in cadmium-induced DA-D2 receptor-mediated motor dysfunctions: protective role of quercetin,” *Scientific Reports*, vol. 8, no. 1, p. 2528, 2018.

[42] W. Hu, T. Xu, P. Wu et al., “Luteolin improves cardiac dysfunction in heart failure rats by regulating sarcoplasmic reticulum Ca2+-ATPase 2a,” *Scientific Reports*, vol. 7, no. 1, p. 41017, 2017.

[43] G. Wang, W. Li, X. Lu, P. Bao, and X. Zhao, “Luteolin ameliorates cardiac failure in type I diabetic cardiomyopathy,” *Journal of Diabetes and Its Complications*, vol. 26, no. 4, pp. 259–265, 2012.

[44] C. Li, Q. Wang, S. Shen, X. Wei, and G. Li, “HIF-1α/VEGF signaling-mediated epithelial-mesenchymal transition and angiogenesis is critically involved in anti-metastasis effect of Luteolin in melanoma cells,” *Phytotherapy Research*, vol. 33, no. 3, pp. 798–807, 2019.

[45] M. Zang, L. Hu, B. Zhang et al., “Luteolin suppresses angiogenesis and vasculogenic mimicry formation through inhibiting Notch1-VEGF signaling in gastric cancer,” *Biochemical and Biophysical Research Communications*, vol. 490, no. 3, pp. 913–919, 2017.

[46] M.-H. Bao, W. Dai, Y.-J. Li, and C.-P. Hu, “Rutaecarpine prevents hypoxia-reoxygenation-induced myocardial cell apoptosis via inhibition of NADPH oxidases,” *Canadain Journal of Physiology and Pharmacology*, vol. 89, no. 3, pp. 177–186, 2011.

[47] Y. Ying, K. Chen, X. Dong et al., “Berberine inhibits cardiac remodeling of heart failure after myocardial infarction by reducing myocardial cell apoptosis in rats,” *Experimental and Therapeutic Medicine*, vol. 16, pp. 2499–2505, 2018.

[48] K. Chen, G. Li, F. Geng et al., “Berberine reduces ischemia/reperfusion-induced myocardial cell apoptosis via activating AMPK and PI3K-Akt signaling in diabetic rats,” *Apoptosis*, vol. 19, no. 6, pp. 946–957, 2014.

[49] L. Gao, R. Yao, Y. Liu et al., “Isorhamnetin protects against cardiac hypertrophy through blocking PI3K-AKT pathway,” *Molecular and Cellular Biochemistry*, vol. 429, no. 1-2, pp. 167–177, 2017.

[50] G. Kroemer and J. Pouyssegur, “Tumor cell metabolism: cancer’s Achilles’ heel,” *Cancer Cell*, vol. 13, no. 6, pp. 472–482, 2008.
[51] R. R. Ramjiawan, A. W. Griffioen, and D. G. Duda, "Anti-angiogenesis for cancer revisited: is there a role for combinations with immunotherapy?" *Angiogenesis*, vol. 20, no. 2, pp. 185–204, 2017.

[52] F. Balkwill and A. Mantovani, "Inflammation and cancer: back to Virchow?" *The Lancet*, vol. 357, no. 9255, pp. 539–545, 2001.

[53] M. Buoncervello, S. Maccari, B. Ascione et al., "Inflammatory cytokines associated with cancer growth induce mitochondria and cytoskeleton alterations in cardiomyocytes," *Journal of Cellular Physiology*, vol. 234, no. 11, pp. 20453–20468, 2019.

[54] A. M. Richards, "Can heart failure cause cancer?" *Nature Reviews Cardiology*, vol. 16, no. 1, pp. 7-8, 2019.

[55] A. Lena, A. J. S. Coats, and M. S. Anker, "Metabolic disorders in heart failure and cancer," *ESC Heart Failure*, vol. 5, no. 6, pp. 1092–1098, 2018.

[56] M. S. Anker, S. von Haehling, U. Landmesser, A. J. S. Coats, and S. D. Anker, "Cancer and heart failure-more than meets the eye: common risk factors and co-morbidities," *European Journal of Heart Failure*, vol. 20, no. 10, pp. 1382–1384, 2018.