Removal of Iron using Isolated Bacteria (Vibrio alginolyticus) in seawater at Ship Dismantling Area

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Abstract. The ship dismantling activity can cause heavy metal pollution in the environment. One of technology to remediate is bioremediation using bioaugmentation and/or biostimulation methods. The purpose of this research was to determine the removal of iron by Vibrio alginolyticus and nutrient addition. Vibrio alginolyticus was halophilic bacteria that isolated from seawater at this area. The laboratory test was conducted using size erlenmeyer of 250 mL. The 5% (v/v) of Vibrio alginolyticus suspension was added in each reactor. After that, the nutrient with ratio of C : N : P (100 : 10 : 1) also was added in each reactor. The removal test was carried out for 14 days. Concentration of iron was analyzed using Atomic Absorption Spectrophotometers (AAS). Based on the results, the percentage of iron removal at location 1 with treatment was 94.9 ± 3.6%. It showed a higher value than compared to iron removal at control 1 which was only 50.4±1.8 %. Whereas at location 2, the percentage of iron removal with treatment reached 53.7±13.1% and it was only 16.7±13.1% at control 2. This indicated that the treatment of the addition of Vibrio alginolyticus and nutrients can improve the iron remediation process.

Keywords: Bacteria; Remediation; Contamination; Heavy Metal; Nutrient; Ship Dismantling.

1 Introduction

Activity of shipbreaking or activity of ship demolition is an activity type of ship disposal that include the breaking up of ships become some parts [1]. The results of these activities can be sold or can be used depending on the requirement. These activities were also known as ship dismantling, ship cracking, or ship recycling. Countries such as China, India, Bangladesh, and Pakistan were countries that have many ship breaking activities [1]. The activities at ship breaking such as metal material recycle, engine reuse, winches reuse, power generators reuse, air conditioners reuse, refrigerators reuse, construction materials reuse have been conducted in the past. However, the ship recycling sector should be more focused on considering the impact on environment [2].

Economic benefits can be obtained from boat demolition activities. Besides that, the environment can be affected due to this activity. The process of cutting and scraping plates or scraping the painted metal surface were some common operations during the ship breaking [3]. The pollutants released from those processes that have potential load into the intertidal area, sediments, sludge, and coast area. According to Deshpande et al. [3, 4], some of hazardous and toxic substance such as asbestos, glass wool, thermocol, oily rags, oily sludge, oily sand, polychlorinated biphenyl (PCB), polyaromatic hydrocarbons (PAH), and organotins like tributyltin (TBT) can be released during the ship dismantling activities. Based on Barua (2017) [5], heavy metals that concern associated with shipbreaking activities were lead (Pb), mercury (Hg), cadmium (Cd), iron (Fe), aluminium (Al), zinc (Zn), copper (Cu), chromium (Cr) and manganese (Mn).

Based on our earlier study, the iron pollution occurred at ship dismantling area in Madura Island both at seawater and sediment or coastal soil [6]. The concentration of Fe in seawater reached 1.03, 1.01 and 1.00 μg/mL at three sampling locations, respectively. Meanwhile, the concentration of extractable Fe in soil was 962.0, 966.05, 981.00 mg/kg at three sampling locations, respectively.

There are many technologies to remediate iron in water media. One of those technologies is bioremediation. Bioremediation is a biological technology to remediate heavy metals and organic pollutants that using naturally living organisms [7]. The principles of the bioremediation can be divided into several techniques. Those techniques are bio-filters, bio-venting, bio-sorption, composting, bio-augmentation, bio-reactor, land farming and bio-stimulation [8]. Bioaugmentation is conducted using addition with suitable bacteria to enhance the bioremediation processes. The bacteria could be from

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contaminated areas or from other places. Biostimulation is a method of bioremediation with nutrient addition to enhance the bioremediation processes. The nutrient that can be added is carbon, nitrogen, phosphate, potassium, and oxygen. The purpose of this research was to determine the removal of iron by bacteria of *Vibrio alginolyticus* and nutrient addition. Iron is an important element for *Vibrio* spp., but iron recovery is complicated by its tendency to form iron complexes that are insoluble in nature and their relationship with high-affinity iron-binding proteins in the host [9].

2 Materials and Methods

2.1 Seawater Sampling

Samples of seawater were taken at the ship dismantling activities in Tanjungjati, Madura, Indonesia. Seawater samples were taken below seawater surface at a depth of 20 cm [10]. Amount 5 L of seawater was taken at two locations ship dismantling. Figure 1 showed the two locations of sampling. Table 1 showed the parameters at the two sampling locations.

![Seawater sampling location](image)

**Table 1.** Font styles for a reference to a journal article.

| Parameter               | Location 1 | Location 2 |
|-------------------------|------------|------------|
| pH                      | 7.3        | 7.6        |
| Temperature             | < 32 °C    | < 32 °C    |
| Salinity                | 28.4 ppt   | 28.2 ppt   |
| TOC – Total Organic Carbon | 0.39 %   | 0.94 %     |
| Total N                 | 3.14 mg/L NH3-N | 1.68 mg/L NH3-N |
| Total P                 | 0.14 mg/L PO4-P | 0.32 mg/L PO4-P |

2.2 Bacteria Preparation

*Vibrio alginolyticus* was halophilic bacteria that isolated from seawater in the same area based on our earlier research [11]. These bacteria have high resistance to iron. The stock of *Vibrio alginolyticus* was inoculated on a new selective media (Thiosulfate-Citrate Bile Salts Sucrose - TCBS media, Merck, USA). The TCBS agar is a media for the selective isolation of *Vibrio spp.*

2.3 Iron Removal Test

The iron removal laboratory test was conducted using size erlenmeyer flasks of 250 mL as a reactor. The code of reactors was R1 for treatment sample location 1, R2 for treatment sample location 2, K1 for control of R1, K2 for control of R2. All tests were carried out a duplicate. The 5% (v/v) of *Vibrio alginolyticus* suspension was added in each reactor. After that, the nutrient with a ratio of C : N : P (100 : 10 : 1) also was added in each reactor. The calculation of nutrient addition was conducted based on data of TOC, Total N and Total P. The addition nutrient was NH₄Cl, H₂O pro analysis (Merck, USA) and KH₂PO₄ and K₂HPO₄ pro analysis (Merck, USA). The removal test was carried out for 14 days.

2.4 Parameter Analysis

Analysis of iron concentration from seawater samples was carried using methods based on SNI 06-6989.4 (2004) [12]. The concentration of iron was analyzed using an Atomic Absorption Spectrophotometer (AAS) instrument, Hitachi model Z-2000 Series (Japan) at Laboratory of Energy, ITS. The monitoring parameters such as pH, temperature, salinity, dissolved oxygen (DO) and Colony Form Unit (CFU) were also measured. Analysis of CFU was conducted based on Harley and Prescot Method [13].

2.5 Statistical Analysis

The experimental data of iron removal percentages were subjected to an analysis of variance (ANOVA) using SPSS Statistics for Windows version 21.0 (SPSS, Inc., Chicago, IL). Statistical significance was defined as p < 0.05.

3 Results and Discussions

Table 2 showed the results of the iron concentration from seawater samples. The pH value during the iron removal test showed that pH was in the normal pH range of 5.80 - 6.87 (Fig. 2). The pHmax of
*Vibrio alginolyticus* was regulated at around 7.6–7.4 and the pH out ranged around 6.4–9.0 [14,15]. *Vibrio alginolyticus* showed acceptable growth at pH values ranging from 4.5–5 to 10.5–11 under suitable temperature and salinity concentration [16].

Temperature during the test was 28.5 – 30.6 °C (Figure 3). According to Farid and Larsen [16], *Vibrio alginolyticus* can grow at temperature of 10–42°C. The value of DO during the test was 3.46 – 3.73 mg/L (Figure 4), this showed the process was under aerobic conditions. According to Sihag et al. [17], in stoichiometric aerobic bacteria, it needed 3.1 mg mL of oxygen to degrade 1 mg/mL of hydrocarbons without considering the total bacterial mass.

Larsen [16], *Vibrio alginolyticus* showed the most suitable growth rate in a 30 % of salinity solution (seawater salinity). Meanwhile, the growth of those bacteria decreased in a 60 % of salinity solution (higher than seawater salinity) and it showed the lowest growth occurred in the 5 % of salinity solution (freshwater).

Figure 6 showed the population of *Vibrio alginolyticus* during iron removal test. The CFU in R1 and R2 showed the higher than in control 1 and control 2. It indicated that the population of *Vibrio alginolyticus* increased after *Vibrio alginolyticus* addition on treatment reactor. However, *Vibrio alginolyticus* presence in reactor without addition of this bacteria. It estimated that *Vibrio alginolyticus* had been presence at the location due to *Vibrio alginolyticus* was as a indigenous bacteria. Based on our previous study, *Vibrio alginolyticus* were isolated at ship dismantling area that has high resistance on iron [6].

The range of salinity was 26 – 31.85 ppt or 26 – 31.85 ‰ during test (Fig. 5). According to Farid and

Based on Figure 7, the percentage of iron removal at location 1 with treatment was 94.9 ± 3.6%. It showed a higher value than compared to iron removal at control 1 which was only 50.4 ± 1.8 %. Whereas at location 2, the percentage of iron removal with treatment reached 53.7 ± 13.1 % and it was only 16.7 ± 13.1 % at control 2. This indicated that the treatment of the addition of *Vibrio alginolyticus* and addition of nutrients can improve the iron bioremediation process.
According to El-Hendawy et al. [18], *Vibrio alginolyticus* isolated from a metal industrial area has a maximum tolerance concentration of heavy metals. That maximum tolerance reached 2.5 mM of Cd, 4 mM of Cu, 2.5 mM of Pb and 3.5 mM of Zn. The ability of those bacteria to remove heavy metal from the solutions that were polluted with heavy metals were 20% for Cd, 31% for Cu, 40% for Pb and 45% for Zn. Based on our previous research, *Vibrio alginolyticus* can survive on 2,000 µg/mL of iron.

The are many processes of bioremediation that are involved in microbe-heavy metals interaction. Those processes are adsorption process, complexation process, precipitation process, oxygenation process and reductions process [7]. The heavy metals bioremediation potential occurred through microbe-heavy metals interactions [7]. According to Davies and Bennett (1983) [19], Ford and Mitchell (1992) [20], the chemical reactions between microbe and metal ions are divided into six types of processes. The first process is intracellular accumulation, second process is reaction of cell wall-associated with metals. Third process is extracellular mobilization or it also known as immobilization of metals. Fourth process is metal siderophore interactions process. Fifth process is extracellular polymer metals interaction with transformation. And the last process is volatilization of metals to air environment.

Species of *Vibrio* have developed a variety of iron transportation systems that allow bacteria to compete for this important element in each of their habitats [9]. The mechanism of iron reduction by *Vibrio alginolyticus* was predicted through using the system including the secretion system and absorption system of high-affinity iron-binding agents (siderophores) and transportation systems for iron bound to the host complex. System transporters for iron ferric and ferrous iron not complexed to siderophores were also common to *Vibrio* species for iron removal [9].

![Fig. 7. Removal percentages of iron](image)

Based on oneway ANOVA analysis, the removal of iron at treatment reactors and control reactors showed a significant difference with p < 0.05. It indicated that *Vibrio alginolyticus* and the nutrient addition can improve the removal of iron in seawater. Based on Ulitzur [21], nutrient concentration affected the *Vibrio alginolyticus* growth, so that the ratio of nutrient should be suitable for reaching the optimum growth.

4 Conclusion

The percentage of iron removal at location 1 with treatment was 94.9 ± 3.6%. It showed a higher value than compared to iron removal at control 1 which was only 50.4 ± 1.8%. Whereas at location 2, the percentage of iron removal with treatment reached 53.7 ± 13.1% and it was only 16.7 ± 13.1% at control 2. In conclusion, *Vibrio alginolyticus* was potential to be used for removal of iron in contaminated seawater.

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References

1. T. Karlis, D. Polemis, Sci. J. of Maritime Res. 30, 128-132 (2016)
2. D.M. Hirimuth. Proceedings of the International Conference on Advances in Civil and Environmental Engineering (2017)
3. P.C. Deshpande, A.K.Tilwankar, P.P. Kalbar, S.R. Asolekar. Sci. of The Total Environ. 438, 304-311 (2012)
4. P.C. Deshpande, P.P. Kalbar, A.K.Tilwankar, S.R. Asolekar. J. of Clean. Produc. 59, 251-259 (2013)
5. P. Barua. Asian profile 45, 2 (2017)
6. H.S. Titah, H. Pratikno, A. Moesriati, R.P. Islami, M.F. Imron. AASEC MATEC Web of Conferences 197, 13020 (2018)
7. S. Siddiquee, K. Rovina, S. Al Azad, L. Naher, S. Suryani, P. Chaikaew. J Microb. Biochem. Technol. 7:6 (2015)
8. J.P. Huang, C.P. Huang, A.L. In: J.P. Vernet, Heavy Metals in the Environment, Elsevier London (1991)
9. S.M. Payne, A. R. Mey, E. E. Wyckoff. Microbiol. and Molecular. Biol. Rev. 80(1), 69-90 (2016)
10. S.K. Bhasheer, S. Umavathi, D. Banupriya, M. Thangavel, Y. Thangam, Y. Int. J. Curr. Microbiol. App. Sci. 3(11), 363-369 (2014)
11. H.S. Titah, H. Pratikno, A. Moesriati, Research Report on Bioremediation (2017)
12. SNI 06-6989.4. Method for Fe analysis using AAS (2004)
13. J.P. Harley, L.M. Prescott, Laboratory exercises in microbiology 5th Edition, McGraw–Hill Companies Texas (2002)
14. T. Nakamura, H. Tokuda, T. Unemoto. Biochimica Et Biophysica Acta 776, 330-336 (1984)
15. T. Nakamura, S. Kawasaki, T. Unemoto. J. of Gen. Microbiol. 138, 1271-1276 (1992)
16. A.F. Farid, J.L. Larsen. Systematic and Applied Microbiology 68-75 (1981)
17. S. Sihag, H. Pathak, D.P. Jaroli. Int. J. of Pure Appl. Biosci. 2(3), 185-202 (2014)
18. H.H. El-Hendawy, D.A. Ali, E.H. El-Shatoury, S.M. Ghanem. Egypt. Acad. J. biol. Sci. 1(1), 23-28 (2009)
19. D.J.A. Davies, B.G. Bennett, Exposure Commitment Assessment of Environment pollutants Vol. 3 MARC (Monitoring and Assessment Research Centre) Report Number 30 (MARC Publication, London, 1983)

20. T. Ford, M. Mitchell, Microbial Transport of Toxic Metals: Environmental Microbiology, John Wiley and Sons New York (1992)

21. S. Uliizur, Microbial Ecolog. 1(3), 127-135 (1974)