Copper, zinc and iron levels in infants and their mothers during the first year of life: a prospective study

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Abstract

Background: Essential micronutrients are important for maintenance of life. Deficiency of micronutrients is more likely to be encountered in children, and women studies are required to investigate the status of micronutrients in children and women. This study aimed to longitudinally evaluate changes in zinc, copper, and iron levels in breastfed infants and their mothers during the first year of life.

Methods: Serum and hair samples were obtained from 35 healthy breastfed infants (51 % males, 49 % females) and their mothers 2, 6, and 12 months after delivery. All of the samples were assessed using an atomic absorption spectrophotometer. Serum iron levels were determined by a Roche/Hitachi/Modular analyzer. Statistical analyses were performed using SPSS-PC (Version 21.00) software.

Results: Hair zinc (p < 0.05) and serum iron (p < 0.001) levels of infants were significantly decreased towards the end of the first year. Infants’ serum copper levels were increased towards the end of the first year. Maternal serum and hair copper levels and serum iron levels were significantly decreased towards the end of the first year. There were no significant correlations between dietary zinc, copper, iron intake, and trace element levels of infants and their mothers.

Conclusions: Infants’ hair zinc levels, maternal and infants’ hair copper levels, and infants’ and maternal serum iron levels declined towards the end of the first year. Infants need more zinc after 6 months of age. Infants’ and mothers’ daily iron intake was less than the recommended intake.

Keywords: Infant, Mother, Serum trace elements, Hair trace elements, Breastfeeding, Diet

Background

Copper (Cu), zinc (Zn), and iron (Fe) are essential micronutrients for maintenance of life. These micronutrients are involved in many complex enzyme systems functioning in various biological processes [1–6]. Deficiency of trace element nutrients is more likely to be encountered in children, and pregnant and lactating women [7, 8]. There are interactions between some trace elements. Deficiency in one trace element may impair absorption of another (e.g., Cu deficiency impairs Fe absorption). Fe and Zn interact at the level of the intestinal mucosa and Zn absorption is impaired by Fe [9, 10]. There is also a strong interaction between Zn and Cu, and they compete at the level of intestinal absorption [11]. High Zn levels in the diet can reduce the absorption of Cu, but high dietary Cu does not decrease absorption of Zn [12].

Inadequate intake of Zn is considered to be responsible for 20 % of global child mortality [13]. Children with iron deficiency anemia have high serum Cu levels and low serum Zn levels [14]. Trace element deficiencies arise from low dietary intake and develop especially when requirements are increased or body stores are depleted. Absorption of trace elements may be impaired by increased intake of dietary components, such as phytate or excessive intake of mineral supplements [11, 15]. Another possible mechanism for trace element deficiency is...
excessive excretion or use. Zinc and copper deficiency is also found in malabsorption syndromes, such as chronic diarrhea, coeliac disease, inflammatory bowel disease, ileostomy, alcoholic cirrhosis, and hemolytic anemia [16].

Zinc as a trace element has three important functional roles: catalytic, structural, and regulatory [3, 5]. Copper has an antioxidant role that protects cells from free-radical injury [3, 17]. Copper also contributes to the formation of ceruloplasmin, which has a role in iron metabolism. Copper is required to absorb and use Fe [1, 14, 17, 18]. Infants and young children in developing countries are particularly vulnerable to Fe and Zn deficiency because of increased requirements, low bioavailability, and recurrent infections [7, 18].

Copper deficiency is rare, but it has been reported in preterm infants, in infants fed with cow’s milk, and in infants recovering from malnutrition accompanied by diarrhea [6, 19–21]. Deiciency of Cu leads to anemia, neutropenia, impairment of growth, abnormalities in glucose and cholesterol metabolism, and increased rates of infection [22].

Iron is another essential trace element that functions in the synthesis of hemoglobin and myoglobin [23]. A total of 25 % of the world’s population is thought to be affected by Fe deficiency. Infants aged between 4 and 24 months, school-age children, females, adolescents, and pregnant and lactating mothers are most affected by this deficiency [24].

Serum concentrations are useful parameters to assess trace elements, but they are not sufficiently specific and sensitive to detect mild deficiency [25, 26]. Hair shaft Zn and Cu levels are useful parameters to determine the quantity of trace elements that is available to the hair follicles at the time of growth, rather than the actual time that children are sampled. Hair trace element levels have been proposed as a useful index of the long-term status of trace elements [27]. Therefore, studies are required to investigate the status of trace elements and their interactions among each other in infants. To the best of our knowledge, there are no longitudinal cohort studies that have investigated the Cu, Zn, and Fe status of breastfed infants and their mothers. There is one relevant study, but it is not a cohort study [28].

Therefore, the present study aimed to longitudinally evaluate the changes in Zn, Cu, and Fe levels of breastfed infants and their mothers after delivery during the first year of life.

Methods
This longitudinal study was conducted between December 2007 and January 2010 in two month-old infants who were attending the Well Child Unit of Istanbul Medical School, Istanbul University. Blood and hair samples were obtained from 111 infants and their mothers 2 months after delivery. Although there were 111 infants and mothers at the beginning of the study, we lost 76 participants (loss to follow-up group) because of infection, medicine use, and vitamin use, and some did not continue to visit the clinic. Blood and hair samples were collected longitudinally from 35 infants (18 males, 17 females) and their mothers 2, 6, and 12 months after delivery.

Inclusion criteria for the study were as follows: uneventful pregnancy, term delivery, and birth weight ≥ 2500 g with no apparent congenital defects. Children born at the Maternity Clinic of the University Hospital constituted the majority of the infants and the children who were followed up at the clinic. Specimens were collected by convenience sampling. Children with any proven or suspected infection at the time of collection of samples were excluded from the study. All of the samples were assessed using an atomic absorption spectrophotometer (Varian Spectra AA 200, GTA-100, Australia). Serum Fe levels were determined by a Roche/Hitachi/ Modular analyzer, japan.

A complete physical examination, including anthropometric measurements, was performed for each infant. Weight, length, and head circumference measurements were performed by two trained nurses. Z scores for length, weight, and head circumference measurements of infants were calculated with a computerized program that was developed for Turkish children [29, 30]. This study was supported by the Istanbul University Research Fund. Written consent was obtained from the parents. Approval of the Medical School Ethical Committee was obtained at the beginning of the study.

Collection of data and specimens, and laboratory procedures
A validated questionnaire that was specific for this project was developed in a pilot study to collect data on the feeding habits of infants and their mothers. All of the mothers were on Fe and folic acid supplementation during pregnancy. Daily and weekly consumption of meat, milk, eggs, and vegetables was recorded. Infants were classified as either exclusively breastfed (infants receiving only breast milk, not even water), partially breastfed, or non-breastfed. Data on daily intake of Zn, Cu, Fe, and meat in infants and their mothers were calculated using a computerized nutrient analysis program (BEBiS), which has been adapted for Turkish infants and their families. Infants’ dietary habits were evaluated only once at 12 months and mothers’ dietary habits were evaluated 2, 6, and 12 months after delivery. For evaluation of breast milk intake, the duration of each feed was used to estimate the likely volume of milk. A feed lasting 10 min or longer was assumed to be 100 ml in volume (i.e., 10 ml per min) and a proportion of this if the feed was
of shorter duration [31]. For example for a feeding lasting 5 min, the milk intake was assumed to be 50 ml.

Blood specimens were collected after a fasting period of 8 or 10 h for mothers and 4 h for infants. Trace element-free syringes, stainless steel needles, and special trace element tubes (Becton-Dickinson) were used.

The serum samples were separated after 10 min of centrifugation. Serum samples were diluted at a 1:6 ratio with bi-distilled water. Serum Zn and Cu concentrations were measured using an atomic absorption flame emission spectrophotometer (Varian AA 100, Australia) (213.9 nm and 324.8, respectively). A standard curve was established using a commercial Zn and Cu reference (Merck KGaA Darmstadt, Germany). The coefficient of variation of the measurements was always below 5%.

Hair samples of infants and their mothers were collected from the suboccipital area of the head. Divided hair samples were sequentially washed three times in hexane, analytical grade ethanol, and fresh bi-distilled water. They were dried at 75 °C in a vacuum oven overnight in polyethylene vial and weighed 20–100 mg. The hair was digested with perchloric acid and nitric acid. Digestion was performed between 65 and 75 °C [33, 34]. The ashed samples were dissolved in 1 mL of bi-distilled water and 10-µL aliquots were injected into a graphite furnace with an auto sampler. Bovine liver certified standard (SRM no. 1577c certified; National Institute of Standard and Technology) and a pooled hair sample were similarly digested in perchloric acid and nitric acid, and were used as internal standards. A standard curve was established using a commercial Zn and Cu reference (Merck KGaA). Hair Zn and Cu concentrations were determined using a Varian Spectra AA 200 atomic absorption spectrophotometer equipped with a GTA-100 [32–34]. Hair Zn and Cu levels are expressed in µmol/g, and serum levels of Zn and Cu are expressed in µmol/g. Serum and hair Zn levels lower than 10.70 µmol/L and 1.07 µmol/g, respectively, were accepted as low levels [27, 35]. Normal serum Cu levels have been reported as between 3.15 and 11.07 µmol/L in infants aged up to 6 months of age, (mean ± SD) 17.50 ± 4.10 µmol/L for infants aged from 6 months to 2 years, and 12.60-24.40 µmol/L for females [36–38]. There is no exact cut off level for hair Cu levels for infants in the literature. Serum iron levels less than 8.95 µmol/L were accepted as low [39].

Statistical analysis
Statistical analyses were performed using SPSS-PC (IBM Corp; Version 21.00) software. Pearson’s analysis for non-continuous, the Student’s t-test for continuous, and the Mann–Whitney U test for non-normal distribution variables were performed between the groups of “loss to follow-up” and “completed the study”.

Results
This study was limited to neonates who were born at the Maternity Clinic of Istanbul Medical School. At discharge from the Maternity Clinic, each mother received a pamphlet with information on the Well Child Clinic. Families had relative socio-economic and cultural homogeneity in this study. All of the families were well above the poverty line, as assessed by their ability to bring their neonate to our center. All of the parents were literate. The majority of the mothers were high school graduates. Preterm infants born before 37 gestational weeks were not followed up at the Well Child Clinic.

The majority of the parents in the study had at least 5 years of schooling. Sociodemographic characteristics of infants and their mothers are shown in Table 1. Weight, length, and head circumference Z scores of all of the infants were within normal limits (Table 2). All 35 infants (18 males, 17 females) and their mothers were followed up until the children were aged 1 year. The breastfeeding status of all infants, parity, and maternal age are shown in Table 1. Hair trace element levels were not normally distributed. Therefore, 95% confidence intervals and median levels are shown in Table 3.

With regard to sociodemographic characteristics, there were no significant differences between the groups “loss to follow-up” and “completed the study” (Table 1). There were no differences in trace elements between these groups.

Infants’ and mothers’ serum zinc levels were not significantly different during the follow-up period (Table 3). As shown in Table 3 hair Zn levels of infants were significantly lower at the ages of 6, 12 months than those at 2 months (p < 0.05; p < 0.001, respectively). Mothers’ hair zinc levels were significantly higher at 6 months after delivery compared with those at 2 and 12 months (Table 3). Three (8.50 %) infants in the 2nd month, five (14.30 %) in the 6th month, and 6 (17 %) at 1 year had serum Zn levels below 10.70 µmol/L. Among these infants, 48.60, 66 and 77.10 % had hair Zn levels below 1.07 µmol/g at 2, 6, and 12 months, respectively. Among the mothers, 14.30, 2.90, and 2.90 % had serum Zn levels below 10.70 µmol/L, and 20, 11.40, and 11.40 % had hair
## Table 1 Comparison of sociodemographic characteristics of infants and their families

| Variables                      | Loss to follow-up | Completed the study | Two-tailed significance |
|--------------------------------|-------------------|---------------------|-------------------------|
|                                | n = 76            | n = 35              |                         |
| Sex                            |                   |                     |                         |
| Male                           | 42                | 18                  | 0.14; p > 0.05          |
| Female                         | 34                | 17                  |                         |
| Maternal education             |                   |                     |                         |
| Primary school                 | 30                | 8                   | 2.95; p > 0.05          |
| Completed high school          | 25                | 15                  |                         |
| University graduate            | 21                | 12                  |                         |
| Paternal education             |                   |                     |                         |
| Primary school                 | 20                | 5                   | 3.31; p > 0.05          |
| Completed high school          | 28                | 12                  |                         |
| University graduate            | 28                | 18                  |                         |
| Maternal occupation            |                   |                     |                         |
| Employee                       | 11                | 3                   | 0.63; p > 0.05          |
| Clerk                          | 27                | 13                  |                         |
| Physician                      | 1                 | 2                   |                         |
| Housewife                      | 35                | 16                  |                         |
| Others                         | 2                 | 1                   |                         |
| Feeding status of infants at 2 months |             |                     |                         |
| Exclusively breastfed          | 63                | 31                  | 0.59; p > 0.05          |
| Breast milk + other food       | 13                | 4                   |                         |
| Feeding status of infants at 6 months |              |                     |                         |
| Exclusively breastfed          | 13                | 3                   |                         |
| Breast milk + other food       | 22                | 4                   |                         |
| Feeding status of infants at 12 months |            |                     |                         |
| Breast milk + other food       | 35                | 100                 |                         |
| Maternal age (years)           | 76                | 30.12 ± 4.95        | 1.20; p > 0.05          |
| Parity                         | 76                | 1.70 ± 0.66         | 0.5; p > 0.05           |

*Pearson χ² analysis and Student’s t-tests were performed

Physicians and others were excluded from the statistical analysis because of their small numbers

## Table 2 Z scores of the infants’ length, weight, and head circumference

| Months | 2          | 6          | 12         |
|--------|------------|------------|------------|
| Length |            |            |            |
| Mean ± SD; median 95 % CI | 0.18 ± 0.91; 0.04 (−0.13)−(0.48) | 0.36 ± 0.93,0.14 (0.052)−(0.67) | 0.56 ± 0.92,0.34 (0.26)−(0.86) |
| Weight |            |            |            |
| Mean ± SD; median 95% CI | 0.22 ± 1.07; 0.15 (−0.13)−(0.57) | 0.22 ± 0.96; 0.46 (−0.1)−(0.54) | 0.10 ± 0.94;0.12 (−0.21)−(0.41) |
| Head circumference |            |            |            |
| Mean ± SD; median 95 % CI | −0.33 ± 1.08; 0.50 (−0.68)−(0.03) | −0.35 ± 0.88;0.68 (−0.64)−(−0.06) | −0.36 ± 0.89; −0.62 (−0.65)−(−0.07) |

Values are mean ± SD, median levels, and 95 % CIs. CI confidence interval
| Months after delivery | Maternal (n = 35) | Total (n = 105) | Infants (n = 35) | Total (n = 105) |
|----------------------|------------------|-----------------|-----------------|-----------------|
|                      | 2                | 6               | 12              | 2               | 6               | 12              |
| Serum Zn (μmol/L)    |                  |                 |                 |                 |
| a                    | 15.00 ± 3.10^c   | 14.80 ± 1.50^c  | 14.10 ± 3.10^c  | 13.30 ± 1.50^d  |
|                      |                  |                 |                 | 13.30 ± 3.10^d  | 13.30 ± 2.70^d  |
| Serum Cu (μmol/L)    |                  |                 |                 |                 |
| a                    | 19.70 ± 4.70^e   | 18.70 ± 3.10^e  | 18.3 ± 3.30^e   | 14.6 ± 3.10^f   |
|                      |                  |                 |                 | 14.6 ± 3.10^f   | 14.6 ± 3.10^f   |
| Serum Fe (μmol/L)    |                  |                 |                 |                 |
| a                    | 13.10 ± 5.6^g    | 13.71 ± 5.40^g  | 12.10 ± 5.60^g  | 13 ± 3.8^g      |
|                      |                  |                 |                 | 13 ± 3.8^g      | 13 ± 3.8^g      |
| Hair Zn (μmol/g)     |                  |                 |                 |                 |
| b                    | 1.48 ± 0.67^i    | 1.84 ± 0.75^i   | 1.76 ± 0.80^i   | 1.70 ± 0.74^i   |
|                      | (median; 95 % CI | (1.50;1.28–1.72 | (1.65;1.60–2.10 | (1.54;1.50–2.00 |
| lower-upper level)   | lower-upper level) | 1.17       | 1.17          | 1.17           |
|                      | 1.02 ± 0.50^j    | 0.77 ± 0.30^j   | 0.77 ± 0.30^j   | 0.77 ± 0.30^j   |
|                      | (0.86; 0.87–1.18 | (0.79;0.67–0.86 | (0.88;0.92–1.14 | (0.88;0.92–1.14 |
|                      |                  | 1.03 ± 0.60^j   | 1.03 ± 0.60^j   | 1.03 ± 0.60^j   |
| Hair Cu (μmol/g)     |                  |                 |                 |                 |
| b                    | 0.20 ± 0.11^k    | 0.22 ± 0.11^k   | 0.17 ± 0.19^k   | 0.20 ± 0.11^k   |
|                      | (0.170;0.16–0.24 | (0.19; 0.18–0.26 | (0.15;0.11–0.19 | (0.17;0.18–0.22 |
| lower-upper level)   | lower-upper level) | 0.20 ± 0.14^l  | 0.32 ± 0.14^l   | 0.32 ± 0.14^l   |
|                      | 0.25 ± 0.13^l    | 0.34 ± 0.16^l   | 0.34 ± 0.16^l   | 0.34 ± 0.16^l   |
|                      | (0.25;0.22–0.32  | (0.25;0.28–0.39 | (0.25;0.28–0.39 | (0.25;0.28–0.39 |
|                      | 0.25 ± 0.13^l    | 0.30 ± 0.16^l   | 0.30 ± 0.16^l   | 0.30 ± 0.16^l   |
|                      | (0.25;0.27–0.33  |                  |                  |                  |

Values are mean ± SD unless stated otherwise

Repeated ANOVA and the paired sample t-test were used to compare serum Zn, Cu, and Fe levels

χ² Friedman and Wilcoxon rank tests were used for hair analysis

F2,6 = 0.66, p > 0.05; F2,12 = 2.42, p > 0.05, t6,12: 1.24, p > 0.05
F2,6 = 0.70, p > 0.05; F2,12 = 0.61, p > 0.05, t6,12: 0.09, p > 0.05
F2,6 = 2.07, p > 0.05; F2,12 = 3.65, p > 0.05, t6,12: 0.99, p > 0.05
F2,6 = 8.65, p < 0.01; F2,12 = 28.03, p < 0.001, t6,12: 1.39, p > 0.05
F2,6 = 0.47, p > 0.05; F2,12 = 0.64, p > 0.05, t6,12: 1.28, p > 0.05
F2,6 = 21.9, p < 0.001; F2,12 = 17.35, p < 0.001, t6,12: 0.36, p > 0.05
χ² Friedman = 9.77, p < 0.05; Z2,6 = 2.57, p < 0.05, Z2,12 = 2.58, p < 0.05
χ² Friedman = 19.94, p < 0.001; Z2,6 = 2.06, p < 0.05, Z2,12 = 2.53, p < 0.01, Z2,12 = 4.09, p < 0.001
χ² Friedman = 10.7, p < 0.005; Z2,6 = 2.12, p < 0.05, Z2,12 = 2.24, p < 0.05, Z2,12 = 3.00, p < 0.005
χ² Friedman = 8.56, p < 0.05; Z2,6 = 3.15, p < 0.05, Z2,12 = 23.51, p < 0.5
Zn levels below 1.07 μmol/g at 2, 6, and 12 months after delivery, respectively (Table 4).

Infants’ serum Cu levels at 12 months of age were significantly higher those at 2 and 6 months (Table 3). Infants’ serum Cu levels in the total group (n = 105) ranged between 8.70 and 26.80 μmol/L. Hair Cu levels were a minimum of 0.21 μmol/g and a maximum of 0.35 μmol/g in infants and a minimum of 0.14 μmol/g and a maximum of 0.26 μmol/g in mothers. Hair Cu levels were not normally distributed. Therefore, 95% confidence intervals and median levels are shown in Table 3. Infants’ and mothers’ hair Cu levels were significantly higher at 6 months compared with 2 and 12 months after delivery (Table 3). Maternal serum and hair Cu levels at 12 months were significantly lower than those at 2 and 6 months. Serum Cu levels of mothers in the total group (n = 105) varied between 13.4 and 24.20 μmol/L.

Serum Fe levels of the infants were significantly lower at 12 months than those at 2 and 6 months (Table 3). Maternal serum Fe levels reached a maximum level at 6 months, and then were significantly decreased at 12 months (Table 3). Among the infants, 31.40, 51.40, and 63% had serum Fe levels less than 8.95 μmol/L at 2, 6, and 12 months, respectively. Among the mothers, 23, 11.40, and 28.60% had serum Fe levels less than 8.95 μmol/L at 2, 6, and 12 months, respectively (Table 4).

The mean daily Zn, Cu, and Fe intakes of infants aged 12 months were 3.2 ± 1.2 mg, 0.79 ± 0.32 mg, and 3.71 ± 1.43 mg, respectively (Table 5). The mean daily Zn, Cu, and Fe intakes for mothers were 8.20 ± 2.80 mg, 1.38 ± 0.62 mg, and 9.15 ± 2.90 mg, respectively. In the second month, the mothers’ daily Zn, Cu, and Fe intakes were higher than those at 6 and 12 months (p < 0.005) (Table 5). Among the mothers, 7.40, 59.20, and 8.50% consumed red meat, vegetables, and fruit, respectively, every day, but 70.60% consumed meat less than 2 days a week. There were no significant relationships between dietary Zn, Cu, and Fe intake and the status of trace elements of infants and their mothers.

Significant positive and negative correlations between trace elements in mothers and infants are shown in Table 6.

Discussion

This study is one of the few longitudinal studies regarding the status of trace elements in predominantly breastfed healthy infants and their mothers. We found that hair zinc and serum iron levels of infants were significantly lower, while serum copper levels were higher at 12 months than those at 2 and 6 months. Maternal serum and hair copper levels and serum iron levels were significantly decreased in the same period. Zinc, copper, and iron are the predominant nutritional trace elements [7, 8, 40]. The regulatory mechanisms of Zn, Cu, and Fe homeostasis are different during pregnancy, lactation, and infancy [8, 18, 41, 42]. Further studies are needed to investigate this issue.

Zinc

During infancy and early childhood, hair zinc concentrations decline from high neonatal values to minimum values at approximately 2–3 years [27]. This trend in hair Zn concentrations may arise from gradual depletion of tissue Zn pools induced by rapid growth. The International Zinc Nutrition Consultative Group concluded that breast milk is a sufficient source of zinc for normal birth weight term infants until approximately 6 months of age [27, 43–45]. Changes in hair Zn concentrations of breastfed and bottle-fed infants during the first 6 months of life were measured by MacDonald et al. [46], and only the bottle-fed males had a significant decline in hair Zn concentrations. There was no decline in hair Zn concentrations in any breastfed infants [46]. In our study, infants’ hair Zn levels significantly declined from high levels at 2 months to low levels at 6 and 12 months (Table 3). Children start to lose endogenous zinc from non-intestinal sites, such as the urinary tract and skin, after 6 months of age, because infants need more Zn after 6 months of age [6, 27]. All of the infants’ eating habits were included in this evaluation. The infants’ dietary habits for Zn, Cu, and Fe were evaluated only at...
12 months using a computerized nutrient analysis program (BEBiS), which has been adapted for Turkish infants and their families.

A meta-analysis in Turkey that included 17 studies showed that the mean serum Zn level of 336 children aged between 1 and 3 months was 14.10 ± 3.70 μmol/L [47]. Another meta-analysis of 28 studies in Turkish adults (n = 4298) showed that the mean serum Zn level was 14.50 ± 2.91 μmol/L [47]. In our study, the mean serum Zn level of infants at all ages was slightly lower than that in previous results mentioned above [47]. However, the mothers’ serum Zn levels were similar to those of Turkish adult levels.

Some authors have reported that dietary maternal zinc intake during lactation is approximately 13–15 mg/day [44, 48]. The recommended intake of Zn for lactating mothers is 12–13 mg /day [49]. The mean daily dietary zinc intake of lactating mothers in our study was lower than these estimated requirements. Similar low findings have also been previously reported for lactating mothers [50, 51]. In our study, the mean daily Zn intake of infants at 12 months was close to the recommended intake (3 mg/day) for infants aged 7–12 months [6]. Nevertheless, we did not gather information on phytate-containing food intake in our study.

Copper
There are few studies on children’s hair Cu levels [52, 53]. Park et al. [52] reported that the mean hair copper level in 655 preschool children was 0.24 ± 0.16 μmol/g. Throughout the whole study period hair Cu levels changed between 0.27 and 0.33 μmol/g in infants and between 0.18 and 0.22 μmol/g in mothers. These results are similar to those in the [52–55]. Eatough et al. [54] reported that hair Cu levels slightly decreased with age. Maternal and infants’ hair Cu levels reached their maximum level at 6 months and then decreased at 1 year in our study. Salmenpera et al. [55] reported that infants’ serum Cu levels increase with age and reach adult levels by 6 months of age. In the current study, serum Cu levels in infants in the 2nd month were lower than those at 6 and 12 months. Serum Cu levels of infants in the total group varied between 8.60 and 26.70 μmol/g throughout the study period. In our study, changes in serum Cu level in infants were similar to those previously reported [5, 14, 55]. Infants’ serum Cu levels at 12 months were higher; whereas maternal serum Cu levels were lower those at 2 and 6 months. There were positive correlations between maternal serum and hair Cu levels at all time periods (Table 6). These correlations showed that factors that affect the maternal Cu status after delivery did not change during the first year. All Cu analyses were performed using an atomic absorption spectrophotometer. We believe that there was minimum measurement error in our study.

The mean daily Cu intake of all infants at 12 months of age was 0.79 ± 0.32 mg in our study. The mean daily Cu intake among all mothers at all time periods was 1.38 ± 0.62 mg. These values are close to the recommended intake for infants and mothers. The estimated safe and adequate daily Cu dietary intake recommended by the Food and Nutrition Board for adults is 1.50–3.00 mg/day [56]. The average Cu intake of children is 0.80–1.90 mg/day [11]. Children (0–0.50 years) often have a low intake of Cu (0.08–0.16 mg/day) because of low Cu levels in breast milk [11]. Despite the declining Cu levels in breast milk during lactation, serum Cu levels in infants are increased, which suggests that the Cu requirements of infants are met. Cu in breast milk appears to be well absorbed and copper levels in breast milk are independent of maternal status [11, 55–57]. Salmenpera et al. [55] showed that serum Cu levels were not correlated with daily Cu intake in infants and in mothers. In our study, there were no relationships between daily intake of Cu and serum and hair Cu levels in infants and mothers. These findings suggest that Cu status is affected by multiple factors other than dietary intake.

The serum Cu level is a good indicator of Cu deficiency. However, neither serum Zn nor Cu reflects marginal status [26]. Therefore, hair Cu levels have been used as an indicator of copper status, particularly in infants [53, 58]. There were negative correlations between

### Table 5 Daily trace element intake of mothers and infants after delivery

| Months | Mothers | Cu     | Fe     |
|--------|---------|--------|--------|
|        | Zn      |        |        |
| 2      | 9.7 ± 3.80 | F = 6.63; p < 0.005 | 1.69 ± 0.56 | F = 6.58; p < 0.005 | 10.80 ± 3.25 | F = 8.26; p < 0.005 |
| 6      | 7.4 ± 2.50 | 1.24 ± 0.59 | 8.30 ± 3.10 |
| 12     | 7.6 ± 2.20 | 1.20 ± 0.72 | 8.40 ± 2.30 |
| Total  | 8.2 ± 2.80 | 1.38 ± 0.62 | 9.15 ± 2.90 |
| Infants| 3.20 ± 1.20 | 0.79 ± 0.32 | 3.71 ± 1.43 |
|        |         |        |        |

Values are mean ± SD
One Way ANOVA test was used to compare daily Zn, Cu, and Fe intake according to months.
| Variants | M serum Zn (month 2) | M hair Zn (month 2) | M serum Cu (month 2) | M serum Cu (month 6) | M hair Cu (month 2) | M hair Cu (month 6) | I serum Fe (month 6) | I serum Fe (month 12) | I hair Zn (month 12) |
|----------|---------------------|-------------------|---------------------|---------------------|-------------------|-------------------|-------------------|-------------------|---------------------|
| M hair Zn at 2 months | *-0.48 | *0.55 | *-0.36 | **0.01 | **0.001 | **0.05 | | | |
| M hair Zn at 6 months | | | | | | | | | |
| M serum Cu at 2 months | *-0.40 | | *-0.42 | | | | | | |
| M serum Cu at 6 months | | | | | | | | | |
| M serum Cu at 12 months | | | | | | | | | |
| M serum Fe at 2 months | | | | | | | | | |
| M serum Fe at 6 months | | | | | | | | | |
| M serum Fe at 12 months | | | | | | | | | |
| M hair Cu at 2 months | *0.62 | *0.41 | | **0.001 | **0.01 | | | | |
| M hair Cu at 6 months | | | | | | | | | |
| I serum Zn at 6 months | | | | | | | | | |
| I hair Zn at 6 months | | | | | | | | | |

Pearson’s correlation analysis was used for serum.
Spearman’s correlation analysis was used for hair.
M mothers, I infants; *r value; **p < value.
maternal hair Cu and Zn levels at 2 and 6 months after delivery in our study (Table 6).

Iron

Infants, children, and women during fertile years are particularly prone to Fe deficiency. In children, the highest prevalence of Fe deficiency is found between 4 months and 3 years of age because of rapid growth and inadequate dietary intake of Fe [9]. In our study, serum Fe levels decreased with age in mothers and infants. Infection was excluded in the subjects by their history, a physical examination, and complete blood count, which were performed at each clinic visit. We did not measure C-reactive protein levels, which may be a limitation of our study. Infants’ and mothers’ daily Fe intakes were less than the recommended intake [48]. There were positive correlations between infants’ and mothers’ serum Fe levels at 6 and 12 months (Table 6). This finding suggests that dietary Fe intake should be supplemented for mothers and infants. We did not evaluate Fe deficiency anemia and Fe deficiency. We only evaluated elemental Fe status and intake in mothers and infants after delivery for up to 1 year. Although ferritin and transferrin receptor need to be determined for Fe status, we only evaluated trace element levels in the study participants.

There is antagonism among Zn, Cu, and Fe absorption from the gastrointestinal tract. Increased Fe concentrations in the intestinal lumen may block the uptake of Zn [14, 15]. Copper plays a role in Fe metabolism through ceruloplasmin [4]. Dietary Zn absorption is inhibited by Fe [15]. Infants’ hair Cu levels and maternal and infants’ hair Cu and serum Fe levels declined towards the end of the first year. Our study consisted of healthy children. We found a significant negative correlation between the infants’ serum Fe and Zn levels at 6 months (Table 6). Voskaki et al. [3] reported significant correlations between serum Zn and Cu levels in children aged 13–14 years and their mothers. We did not find such a correlation in our study group. The reason for this discrepancy between studies may be due to our small sample size and different age.

Our study is one of the few studies on trace element levels of healthy breastfed infants and their mothers. Nevertheless, our study has some limitations. The families were generally from the middle socio-economic class and were not representative for all of the country. Evaluation of 3-day diets was based on the mothers’ reports and our sample size was small. Therefore, further research is required on a larger scale with participation of families from all socio-economic classes. Additionally, dietary components, such as phytate, which affect Zn, Cu, and Fe metabolism, were not assessed. This is a confounder and could affect absorption of trace elements. We only evaluated elemental Fe status and Fe intake in mothers and infants after delivery for 1 year. Our study aim was not to investigate the mechanism of possible Fe deficiency anemia, but rather to investigate the natural course of Fe levels of breastfed infants and their mothers. However, in further studies, ferritin and transferrin receptor levels should be analyzed to understand the possible mechanisms of Fe deficiency anemia. Although levels of inflammation can affect serum Cu, Fe, and Zn concentration, even if subclinical [59], hair trace element levels are not affected by acute infection [27, 58]. We did not measure C-reactive protein levels, which may have also been a limitation of our study. However, infection was excluded in all subjects by recording the subjects’ history, performing a physical examination, and measuring the complete blood count, at each clinic visit.

Conclusions

Infants’ hair Zn levels, maternal and infants’ hair Cu levels, and infants’ and maternal serum Fe levels declined towards the end of first year. We observed a significant decline in hair Zn levels of infants at 6 and 12 months than those at 2 months. Children lose endogenous zinc from non-intestinal sites (i.e., urine and body surface) after 6 months of age. Therefore, they need more zinc from that age. Infants’ and mothers’ daily Fe intake was less than the recommended intake. There were positive correlations between infants’ and mothers’ serum Fe levels at 6 and 12 months. This finding suggested that dietary Fe intake should be supplemented for mothers and infants.

Abbreviations

Zn: Zinc; Cu: Copper; Fe: Iron.

Competing interests

The authors declare that they have no competing interests.

Authors’ contributions

GG conceived the study, participated in its design and coordination, and drafted the manuscript. TAO conceived the study, participated in its design and coordination, drafted the manuscript, helped with the collection and acquisition of data, and performed trace element analysis. MSC and ÖD participated in the design of the study and drafted the manuscript. HI performed the statistical analyses. BO performed serum iron analysis. GS helped to coordinate and draft the manuscript. All of the authors read and approved the final version of the manuscript.

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References

1. Hegazy AA, Zaher MM, Abd El-Hafez MA, Morsy AA, Saleh RA. Relation between anemia and blood levels of lead, copper, zinc and iron among children. BMC Res Notes. 2010;3:1–9.

2. ESPGHAN. Iron, minerals and trace elements. J Pediatr Gastroenterol Nutr. 2005;41:339–46.

3. Voskaki I, Anvanitidou V, Athanasopoulou H, Tzagkaraki A, Tripsianis G, Giannoulla-Karanantza A. Serum copper and zinc levels in healthy Greek children and their parents. Biol Trace Elem Res. 2010;134:136–45.

4. Schneider JM, Fuji ML, Lamp CL, Lonnerdal B, Zidenberg-Cherr S. The prevalence of low serum zinc and copper levels and dietary habits associated with serum zinc and copper in 12 to 36-month-old children from low-income families at risk for iron deficiency. J Am Diet Assoc. 2007;107:1924–9.

5. Bujor-Jannaszczyk SC, Miller M. Role of trace elements in parental nutrition support of the surgical neonate. J Pediatr Surg. 2012;47:760–71.

6. Kleinman RE. Trace elements in pediatric nutrition handbook. 5th ed. Elk Grove: American Academy of Pediatrics; 2004. p. 313–21.

7. Kodama H. Trace element deficiency in infants and children. JMAJ. 2004;47:736–81.

8. Hambidge KM, Facoq WD. Changes in plasma and hair concentrations of zinc, copper, chromium and manganese during pregnancy. Obstet Gynecol. 1974;44:666–72.

9. Lind T. Iron and zinc in infancy: Results from experimental trials in Sweden and Indonesia. Umeå University Medical Dissertations. 2004;87:1–108.

10. Abrams SA. New approaches to iron fortification: Role of bioavailability studies. Am J Clin Nutr. 2004;80:1104–5.

11. Lonnerdal B. Bioavailability of copper. Am J Clin Nutr. 1996;63:821S–9.

12. Wu X, Liu Z, Guo J, Wan C, Zhang T, Cui H, et al. Influence of dietary zinc and copper on apparent mineral retention and serum biochemical indicators in young male mink (Mustela vison). Biol Trace Elem Res. 2015;165:56–66.

13. Jacks B, Ball M, Jacks G. A first assessment of zinc intake in Niger Inland Delta, Mali. Sight and Life. 2008;2:27–32.

14. Ece A, Uyanik BS, Iscan A, Ertan P, Yiğitgül MR. Increased serum copper and decreased serum zinc levels in children with iron deficiency anemia. Biol Trace Elem Res. 1997;59:31–9.

15. Lönnnerdal B. Dietary factors influencing zinc absorption. J Nutr. 2000;130:1378–85.

16. Gibson RS. Trace element deficiencies in humans. Can Med Assoc J. 1991;145:131.

17. Jones AA, Di Silvestro RA, Coleman M, Wagner TL. Copper supplementation of adult men: effect on blood copper enzyme activities and indicators of cardiovascular disease risk. Metabolism. 1997;46:1380–3.

18. Zlotkin SH, Schauer C, Ageyi SO, Wolffson J, Tondeur MC, Asante KP, et al. Demonstrating zinc and iron bioavailability from intrinsically labeled microencapsulated ferrous fumarate and zinc glucinate supplements in young children. J Nutr. 2000;130:202–5.

19. L’Abbé MR, Friel JK. Copper status of very-low-birth-weight infants during and 12 months of infancy. Pediatr Res. 1992;32:183–8.

20. Cordano A, Baertl JM, Graham GG. Copper deficiency in infancy. Pediatrics. 1964;34:324–36.

21. Levy T, Zelharia A, Grunebaum M, Nitzan M, Steinherz R. Copper deficiency in infants fed cow’s milk. J Pediatr. 1985;106:786–8.

22. Shazia Q, Mohammad ZH, Rahman T, Shekhar HU. Correlation of oxidative stress with serum trace element levels and antioxidant enzyme status in beta thalassemia major patients: A review of the literature. Anemia. 2012;2012:279093.

23. Worwood M. Iron and other trace metals. In: Jacobs A, Worwood M, editors. Food and nutrition: a medical perspective. Philadelphia: WB Saunders Company; 2001. p. 1426–41.

24. Wu AHB. General clinical tests. In: Wu AHB, editor. Tietz clinical guide to laboratory tests. 4th ed. St. Louis:Missouri: WB Saunders Company; 2006. p. 32–1202.

25. Lehmann HP, Henry JB. SI units. In: Henry JH, editor. Clinical diagnosis and management by laboratory methods. 20th ed. Philadelphia: WB Saunders Company; 2001. p. 1426–41.

26. Hinks LJ, Clayton BE, Lloyd RS. Zinc and copper concentrations in leucocytes and erythrocytes in healthy adults and the effect of oral contraceptives. J Clin Pathol. 1983;36:1016–21.

27. Hotz C, Brown RH. Overview of zinc nutrition in assessment of the risk of zinc deficiency in populations and options for its control. Food Nutr Bull. 2004;25:1 Suppl 2:196–203.

28. McMaster D, Lappin TR, Halliday HL, Patterson CC. Serum copper and zinc levels in the preterm infant. A longitudinal study of the first year of life. Biol Neonate. 1983;44:108–13.

29. Bundak R, Furman A, Gündüz H, Darendeller F, Baş F, Neyzi O. Body mass index references for Turkish Children. Acta Pediatric. 2006;95:194–8.

30. Buyþorum. http://www.itf.istanbul.edu.tr/cocukhast/buyuyorumweb/ Index.htm. Accessed 15 Jun 2015

31. Emmett PM, North K, Noble S. Types of drinks consumed by infants at 4 and 8 months of age: a descriptive study. Public Health Nutr. 2000;3:211–7.

32. Chou PP. Zinc. In: Pesce AJ, Kaplan LA, editors. Methods in clinical chemistry. St. Louis. Missouri: The C.V. Mosby Company; 1987. p. 596–602.

33. Alcock NW. Copper. In: Pesce AJ, Kaplan LA, editors. Methods in clinical chemistry. St. Louis. Missouri: The C. V. Mosby Company; 1987. p. 527–38.

34. Chen XC, Yin TA, He JS, Ma QY, Han ZM, Li LX. Low levels of zinc in hair and blood, pica, anorexia and poor growth in Chinese preschool children. Am J Clin Nutr. 1985;42:694–700.

35. Wu AHB. General clinical tests. In: Wu AHB, editor. Tietz clinical guide to laboratory tests. 4th ed. St. Louis:Missouri: WB Saunders Company; 2006. p. 32–1202.

36. Wu AHB. General clinical tests. In: Wu AHB, editor. Tietz clinical guide to laboratory tests. 4th ed. St. Louis:Missouri: WB Saunders Company; 2006. p. 32–1202.

37. Lehmann HP, Henry JB. SI units. In: Henry JH, editor. Clinical diagnosis and management by laboratory methods. 20th ed. Philadelphia: WB Saunders Company; 2001. p. 1426–41.

38. Lin CN, Wilton A, Church BB, Bhshan S, Roberts WL, McMillin GA. Pediatric reference intervals for serum copper and zinc. Clin Chim Acta. 2012;413:612–5.

39. Tietz NW. Tables of normal values in fundamentals of clinical chemistry. Philadelphia: WB Saunders Company; 1976. p. 1207–27.

40. Moser PB, Reynolds RD, Acharya S, Howard MP, Andon MB, Lewis SA. Copper, iron, zinc and selenium dietary intake and status of lactating women and their breast-fed infants. Am J Clin Nutr. 1988;47:229–34.

41. Dijkhuizen MA, Weringa FT, West CE. Concurrent micronutrient deficiencies in lactating mothers and their infants in Indonesia. Am J Clin Nutr. 2001;73:786–91.

42. Fung EB, Ritchie LD, Woodhouse LR, Roehl R, King JC. Zinc absorption in women during pregnancy and lactation: A longitudinal study. Am J Clin Nutr. 1995;61:1030–6.

43. Kriebes NF, Westcott J. Zinc and breastfed infants: If and when is there a risk of deficiency. Adv Exp Med Biol. 2002;503:69–75.

44. Kriebes NF, Redingier CJ, Hartley S, Robertson AD, Hambidge KM. Zinc supplementation during lactation: effects on maternal status and milk zinc concentrations. Am J Clin Nutr. 1995;61:1030–6.

45. Nakamoto M, Ninh NX, Ishihata H, Yoshike N, Hirn VTT, Nhug BT, et al. Nutritional status of lactating mothers and their breast milk concentration of iron, zinc and copper in rural Vietnam. J Nutr Sci Vitaminol. 2005;51:338–45.

46. Macdonald LD, Gibson RS, Miles JE. Changes in zinc and copper concentrations of breast fed and bottle fed infants during the first six months. Acta Pediatr Scand. 1982;7:185–9.

47. Tanelli B. Zinc in Anatolian population. Ege J Med. 2005;44:1–10.

48. Dolphin AE, Goodman AH. Maternal diets, nutritional status and zinc homeostasis during lactation in a population with a low zinc intake. Am J Clin Nutr. 2002;75:99–103.

49. Gibson RS. Content and bioavailability of trace elements in vegetarian diets. Am J Clin Nutr. 1994;59:1223S–32.
52. Park HS, Shin KO, Kim JS. Assessment of reference values for hair minerals of Korean preschool children. Biol Trace Elem Res. 2007;116:119–30.
53. Dongarra G, Lombardo M, Tamburo E, Varrica D, Cibella F, Cuttita G. Concentration and reference interval of trace elements in human hair from students living in Palermo, Sicily (Italy). Environ Toxicol Pharmacol. 2011;32:27–34.
54. Eatough DJ, Christensen JJ, Bhatt RM, Hartley C. Level of selected elements in human hair. In: Brown AC, editor. The first human hair symposium. New York: Medcom Press; 1974.
55. Salmenpera L, Perheentupa J, Pakarinen P, Siimes MA. Cu nutrition in infants during prolonged exclusive breast-feeding: low intake but rising serum concentrations of Cu and ceruloplasmin. Am J Clin Nutr. 1986;43:251–7.
56. Committee on Dietary Allowances. Food and Nutrition Board. National Research Council. Recommended dietary allowances. 10th ed. Washington, DC: National Academy Press; 1989.
57. Feely RM, Eitenmiller RR, Jones JB, Barnhart H. Copper, iron and zinc contents of human milk at early stages of lactation. Am J Clin Nutr. 1983;37:443–8.
58. Valkovic V. The role of trace elements. In: Valkovic V, editor. Trace elements in human hair. New York and London: Garland STPM Press; 1977. p. 89–141.
59. Raiten DJ, Ashour FAS, Ross AC, Meydani SN, Dawson HD, Stephensen CB, et al. Inflammation and Nutritional Science for Programs/Policies and Interpretation of Research Evidence (INSPIRE). J Nutr. 2015;145:1039S–108.