Storage Conditions and Adsorption Thermodynamic Properties for Purple Corn

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Abstract: Adsorption isotherms provide insight into the thermodynamic properties governed by food storage conditions. Adsorption isotherms of purple corn of the Canteño variety were evaluated at 18, 25, and 30 °C, for the equilibrium relative humidity (ERH) range between 0.065 and 0.95. The equilibrium moisture (Xe) was determined by the continuous weight-change method. Seven mathematical models of isotherms were modeled, using the coefficient of determination R², mean absolute error (MAE), and estimated standard error (ESE) as the convergence criterion. Thermodynamic parameters such as isosteric heat (q_a), Gibbs Free Energy (AG), differential entropy (AS), activation energy (Ea), and compliance with the isokinetic law were evaluated. It was observed that the adsorption isotherms presented cross-linking around 75% ERH and 17% Xe, suggesting adequate storage conditions at these values. The GAB and Halsey models reported better fit (R² > 97%, MAE < 10%, ESE < 0.014 and random residual dispersion). The reduction of Xe from 17 to 7%, increases q_a, from 7.7022 to 0.0165 kJ/g, while ΔG decreases considerably with the increase in Xe, presenting non-spontaneous endothermic behavior, and linear relationship with ΔS, evidencing compliance with the isokinetic theory, governed by q_a = Ea showed that more energy is required to remove water molecules from the upper layers bound to the monolayer, evaluated using C_GAB. The models predicted the storage conditions, and the thermodynamic parameters show the structural stability of the purple corn grains of the Canteño variety during storage.

Keywords: purple corn; adsorption isotherm; isosteric heat; Gibbs free energy; differential entropy; activation energy; isokinetic theory

1. Introduction

Pigments in the grains of purple corn (Zea mays L.), in addition to being used as natural colorants, are attributed biological functions as antioxidants [1,2], and are found in mainly the pericarp, aleurone, endosperm, and embryo of corn [3–6]. These compounds present nutritional interest for their contribution to human health due to their beneficial properties [7,8]. Similarly, purple corn, due to its color, is used in food, cosmetic and
pharmaceutical products [9–11], and is widely consumed in countries such as Peru, Bolivia, Ecuador, and Mexico, especially in porridge, desserts, and as a drink, due to its pleasant flavor and striking color [12–14].

However, the functional or antioxidant properties, which purple corn grains present, may be susceptible to changes, even losing their qualities due to storage conditions, such as inadequate temperature and relative humidity [4,15], and the uncontrolled combination of these can allow for the development of molds and yeasts [16–19], or at the other extreme allow weight loss, which would cause economic losses due to low humidity or the deterioration of the grain due to cracking and wear of the food surface [20,21].

The equilibrium water content (Xₑ) in food is reached when the partial vapor pressure of the material equals the vapor pressure of the air that contains it, to the ratio of the vapor pressure of the food. The ambient air is called water activity, a_w, and is a determining factor during storage [22,23].

Numerous physicochemical, semi-empirical, and empirical mathematical models have been developed that help to study the adsorption behavior of water in foods [24–26], in equilibrium with the atmosphere that contains it, at different storage temperatures, called adsorption isotherms [27–29], describing the behavior of water at the level of a monolayer (BET isotherm), multilayer (GAB isotherm) [30–32], or simply by adjusting a_w and equilibrium moisture data [33,34]. These models, such as BET and GAB, provide information on the thermodynamic behavior of the water bound to the active sites [35,36], on the surface of the food.

On the other hand, isotherms provide information on thermodynamic adsorption parameters, which are useful for the design of drying and storage equipment [37–39], thus, the isosteric heat of sorption is an indicator of the bond strength between free water and the surface of the food, and the higher this is, the greater the energy required during drying [40,41]. Another aspect to consider is the speed with which water molecules dissipate in the active sites of materials or foods, which is related to entropy [26,42]. This movement of water molecules facilitates the vaporization process, which can occur spontaneously, and can be measured using the Gibbs free energy [43,44].

There is currently a great interest in consuming foods with minimal processing, with high nutritional value, and that also provide health benefits [45], such as purple corn; however, these are susceptible to deterioration and loss of functionality during storage, which would generate economic losses in the producer and marketer, for this reason, the research aimed to study the storage conditions and thermodynamic properties of purple corn grains.

2. Materials and Methods

2.1. Samples

Grains of purple corn (Zea mays L.) of the Canteño variety, dried outdoors, were used, with an initial humidity of 11.03% dry basis (d.b.), produced in the fields of the José María Arguedas National University, Santa Rosa farm at 2804 m altitude, 13°39′05″ S and 73°26′31″ W, in the province of Andahuaylas, Peru.

2.2. Construction of Adsorption Isotherms

The construction of the adsorption isotherms was based on the static gravimetric method [46]. Nine glass jars of 200 mL with hermetic lid were conditioned, with a tripod incorporated as the support where three corn grains were placed. Previously, the flasks were loaded with saturated solutions of chemical substances with water activity values between 0.06 and 0.92 (Table 1).

The jars were placed in a Memmert model 100–800 stove at 18, 25, and 30 °C. Weighing of the corn grains was carried out every three days with precise analytical balance until the samples presented a constant weight, that is, they reached equilibrium with their atmosphere. Sodium azide at 0.25% was added to prevent microbiological growth and grain germination for water activities above 0.5.
**Table 1.** The water activity of substances for the construction of isotherms.

| Substance                  | Equation                                                                 | R²   |
|----------------------------|--------------------------------------------------------------------------|------|
| Sodium hydroxide           | \( a_w = 0.081 - 1.128 \times 10^{-3}T + 3.929 \times 10^{-5}T^2 - 5.092 \times 10^{-7}T^3 \) | 0.998 |
| Lithium chloride           | \( \ln a_w = \left( \frac{500.95}{T} \right) - 3.85 \)                      | 0.980 |
| Potassium Acetate          | \( \ln a_w = \left( \frac{861.39}{T} \right) - 4.33 \)                      | 0.970 |
| Magnesium chloride         | \( a_w = 0.365 - 2.523 \times 10^{-3}T + 5.071 \times 10^{-5}T^2 - 4.166 \times 10^{-7}T^3 \) | 0.963 |
| Magnesium Nitrate          | \( \ln a_w = \left( \frac{356.60}{T} \right) - 1.82 \)                      | 0.990 |
| Potassium iodide           | \( \ln a_w = \left( \frac{255.80}{T} \right) - 1.23 \)                      | 1.000 |
| Sodium chloride            | \( \ln a_w = \left( \frac{228.92}{T} \right) - 1.04 \)                      | 0.960 |
| Potassium chloride         | \( \ln a_w = \left( \frac{367.58}{T} \right) - 1.39 \)                      | 0.970 |
| Barium chloride            | \( a_w = 0.908 - 4.011 \times 10^{-4}T + 2.786 \times 10^{-5}T^2 - 2.037 \times 10^{-7}T^3 \) | 0.997 |

\( a_w \), \( T \) - the water activity; \( T \) is the temperature (K). Source: Labuza et al. [46].

2.3. Determination of Equilibrium Moisture

The equilibrium humidity was calculated by the difference between the mass of the sample that reaches equilibrium and the dry mass, according to equation:

\[
X_e = \frac{m_{eq} - m_s}{m_s} \tag{1}
\]

where, \( X_e \) is the equilibrium moisture on a dry basis; \( m_{eq} \) is the mass of the sample at equilibrium, g; and \( m_s \) is the mass of the dry sample, g.

2.4. Adjustment of Adsorption Isotherms

The experimental data were fitted to adsorption isotherm models (Table 2), by non-linear regression, applying the Quasi-Newton method, using Statistica 8.0 Software (Statsoft, Tulsa, OK, USA). The goodness of fit was evaluated using the fit coefficient \( R^2 \), mean absolute error (MAE) (Equation (2)) and the estimated standard error (ESE) (Equation (3)), by considering good fit when MAE < 10% and ESE lower [47–51]. Likewise, the dispersion of the residuals of \( X_e \) was taken as a convergence criterion, which evaluates the tendency of the systematic and random errors during the experimentation [51].

**Table 2.** Mathematical models of the adsorption isotherm.

| Model                  | Equation                                                                |
|------------------------|--------------------------------------------------------------------------|
| Temperature dependent  | \( x_e = \frac{x_{eBET}}{1 - \frac{x_{eBET}}{1 + \frac{x_{eBET}}{\left( \frac{m_{eq} - m_s}{m_s} \right)}}} \) (2) |
| GAB                    | \( x_e = \frac{x_{eGAB}}{1 - \frac{x_{eGAB}}{1 + \frac{x_{eGAB}}{\left( \frac{m_{eq} - m_s}{m_s} \right)}}} + C_{GAB} \) (3) |
| Oswin                  | \( x_e = A \left( \frac{a_w}{1 - a_w} \right)^B \)                     |
| Modified Henderson     | \( 1 - a_w = \exp(-kT_x^p) \)                                          |
| Chung y Pfost          | \( a_w = \exp\left( \frac{A}{RT} \exp\left(-Bx_e\right) \right) \)        |

Temperature independent
Table 2. Cont.

| Model | Equation |
|-------|----------|
| Halsey | $aw = \exp \left( -\frac{A}{Bx} \right)$ (7) |
| Henderson | $1 - aw = \exp \left( -kx^n \right)$ (8) |

where: $A$, $B$, $C_{BET}$, $k_{GAB}$, $k$, $n$, $n'$ are constants of the equations; $X_e$ is the equilibrium humidity (g water/g dry basis); $X_m$ is the humidity of the molecular monolayer (g water/g dry mass); $R$ is the universal gas constant; and, $T$ is the temperature (K).

%MAE = \frac{100}{N} \sum_{i=1}^{N} \left| \frac{Me_{i,exp} - Me_{i,pre}}{Me_{i,exp}} \right| (9)

\text{ESE} = \sqrt{\frac{\sum_{i=1}^{N} (Me_{i,exp} - Me_{i,pre})^2}{N - n}}, (10)

where, $Me_{i,exp}$ is the observed experimental equilibrium moisture content; $Me_{i,pre}$ is the predetermined moisture content in the observations; $N$ is the number of experimental observations, $n$ is the number of constants in the model.

2.5. Thermodynamic Parameters

The isosteric heat of adsorption ($q_{st}$) (or differential enthalpy) evaluates the difference between the total heat of sorption in the purple corn and the heat of vaporization of water at the system temperature \cite{52}, and can be estimated using the Clausius-Clapeyron equation (Equation (11)) \cite{53}.

The value of $q_{st}$ was obtained by plotting $\ln aw$ vs. $1/T$, at their respective humidities, where $q_{st}/R$ is the slope.

$$\frac{\partial \ln (aw)}{\partial (1/T)} |_{x_e} = -\frac{q_{st}}{R}$$ (11)

where, $aw$ is the water activity; $T$ is the absolute temperature (K); $q_{st}$ is the isosteric heat of sorption (kJ/kg); and $R$ is the universal gas constant (8.314 kJ/kmol·K) for water (0.4619 kJ/kg·K).

On the other hand, the $q_{st}$ data $X_e$, were fitted to the Tsami equation (Equation (12)) \cite{53}.

$$q_{st} = q_0 \exp \left( -\frac{X_e}{X_0} \right)$$ (12)

where, $q_0$ is the isosteric heat of sorption when the moisture content is constant; $X_e$ is the equilibrium humidity (g water/g dry sample), $q_0$ is the isosteric heat of adsorption (kJ/mol) of the first water molecule in the food and is defined as $X_e \rightarrow 0 \Rightarrow q_{st} \rightarrow q_0$; and $X_0$ is the characteristic moisture content for each product.

The differential entropy of sorption ($\Delta S$) (kJ/kg·K) was calculated using the Gibbs–Helmholtz equation (Equation (13)) \cite{54}.

$$\Delta S = \frac{q_{st} - \Delta G}{T}$$ (13)

where $\Delta G$ is the Gibbs free energy (kJ/kg), it is expressed using Equation (14).

$$\Delta G = -RT \ln(aw)$$ (14)

During the adsorption process, the variation of Gibbs free energy is related to the variation of isosteric heat and entropy, thus by replacing Equation (14) in (13), Equation (15) is obtained.

$$- \ln(aw) = \frac{q_{st}}{RT} \frac{\Delta S}{R}$$ (15)

The linear form of Equation (15), allows us to obtain the intercept and calculate $\Delta S$. 
The enthalpy–entropy compensation theory suggests the existence of a linear relationship between enthalpy and entropy according to Equation (16) [36,55,56].

\[
q_{st} = T_\beta \Delta S + \Delta G_\beta
\]  

where, \( T_\beta \) is the isokinetic temperature (K); \( \Delta G_\beta \) is the free energy (kJ/kg) at \( T_\beta \).

\( T_\beta \) is an indicator in which it is assumed that all interactions within the purple corn grains occur with the same speed [57], while the term \( +\Delta G_\beta \) represents whether the adsorption process is spontaneous or not (\( -\Delta G_\beta \)).

The validity of the compensation theory was evaluated by comparing \( T_\beta \) with the harmonic mean temperature (\( T_{hm} \)) (Equation (17)) [56,58,59], and it is valid when \( T_\beta \neq T_{hm} \), likewise if \( T_\beta > T_{hm} \), the process of sorption is governed by the isosteric heat of sorption (enthalpy of sorption), and if \( T_\beta < T_{hm} \) by the entropy [60,61].

\[
T_{hm} = \frac{n}{\sum_{i=1}^{n} 1/T_i}
\]  

where, \( n \) is the number of used temperatures.

The effect of temperature on humidity was evaluated using the Arrhenius equation (Equation (18)), for the GAB isotherm parameters.

\[
\ln(D) = \ln(D_0) - \frac{E_a}{RT}
\]  

where, \( D \) is a parameter of the GAB model, \( D_0 \) is a pre-exponential factor, and \( E_a \) is the activation energy (kJ/mol).

3. Results and Discussion

3.1. Adsorption Isotherms

An equilibrium moisture, \( X_e \), of purple corn was reached after 15 days at 18 °C, and in 12 days at 25 °C and 30 °C. The behavior of \( X_e \) at storage conditions is shown in Figure 1, and a crossover of the isotherms around \( a_w \) 0.75 is observed, due to the composition of purple corn of a higher content of carbohydrates and sugars compared to other corn varieties, this behavior is characteristic of fruits with a high sugar content [16,20,22,23,25,31,62].

![Figure 1. Adsorption isotherms adjusted with the Halsey model.](image)

Likewise, the increase in temperature would promote the availability of active sites to adsorb water on the corn grain surface, due to the effects caused by capillarity and humidity interactions [23,62,63], this being a typical behavior of a type II isotherm [27,33,64,65].

3.2. Adjustment of Adsorption Isotherms

It was observed that, at 18 °C, the GAB and Halsey models reported \( R^2 \) values of 0.967 and 0.974, MAE of 5.149% and 5.902%, and ESE 0.013 and 0.011, respectively; at
25 °C, $R^2$ values of 0.973 and 0.976, MAE 8.795% and 8.628% and ESE 0.014 and 0.012 were found, while at 30 °C, $R^2$ values were reported to be 0.984 and 0.975, MAE 8.508% and 10.412% and ESE as 0.011 and 0.013, respectively (Table 3). In the same way, both models presented random residual dispersion at the study temperatures, which indicates that the models better attenuate systematic and experimental errors due to repetitiveness, better representing the adsorption phenomenon [22,25,34,66,67].

Table 3. Model parameters for adsorption isotherms.

| Model            | Temperature dependent | Parameters          | $R^2$ | SEE  | MAE (%) | Residual Distribution |
|------------------|-----------------------|---------------------|-------|------|---------|-----------------------|
|                  |                       | $X_m$               | 0.076 | 0.967| 0.013   | 5.149                 |
|                  | GAB                   | $C_{GAB}$           | 1,502,959 |      |         | Random                |
|                  |                       | $K$                 | 0.755 |      |         |                       |
|                  |                       | $X_m$               | 0.068 | 0.973| 0.014   | 8.795                 |
|                  |                       | $C_{GAB}$           | 4,501,090 |      |         | Random                |
|                  |                       | $K$                 | 0.825 |      |         |                       |
|                  |                       | $X_m$               | 0.064 | 0.984| 0.011   | 8.508                 |
|                  |                       | $C_{GAB}$           | 1,812,258 |      |         | Random                |
|                  |                       | $K$                 | 0.842 |      |         |                       |
|                  |                       | $X_m$               | 0.028 | 0.301| 0.056   | 33.845                |
|                  |                       | $C_{BET}$           | -19.315 |      |         | Trending              |
|                  | BET                   | $X_m$               | 0.030 | 0.604| 0.049   | 26.359                |
|                  |                       | $C_{BET}$           | -20.218 |      |         | Trending              |
|                  |                       | $X_m$               | 0.029 | 0.594| 0.051   | 27.66                 |
|                  |                       | $C_{BET}$           | -21.015 |      |         | Trending              |
|                  | Oswin                 | $A$                 | 0.132 | 0.959| 0.014   | 6.657                 |
|                  |                       | $B$                 | 0.264 |      |         |                       |
|                  |                       | $A$                 | 0.127 | 0.957| 0.016   | 9.171                 |
|                  |                       | $B$                 | 0.323 |      |         | Slightly random       |
|                  |                       | $A$                 | 0.121 | 0.966| 0.015   | 9.870                 |
|                  |                       | $B$                 | 0.345 |      |         | Slightly random       |
|                  | Modified Henderson    | $k$                 | 0.336 | 0.912| 0.020   | 10.794                |
|                  |                       | $n$                 | 2.518 |      |         | Trending              |
|                  |                       | $k$                 | 0.130 | 0.903| 0.024   | 12.346                |
|                  |                       | $n$                 | 2.005 |      |         | Trending              |
|                  |                       | $k$                 | 0.095 | 0.927| 0.022   | 12.53                 |
|                  |                       | $n$                 | 1.809 |      |         | Trending              |
|                  | Chun-Pfost            | $A$                 | -24.266 | 0.948| 0.015   | 7.443                 |
|                  |                       | $B$                 | 19.759 |      |         | Random                |
|                  |                       | $A$                 | -16.300 | 0.929| 0.021   | 12.428                |
|                  |                       | $B$                 | 16.826 |      |         | Trending              |
|                  |                       | $A$                 | -13.974 | 0.940| 0.020   | 13.612                |
|                  |                       | $B$                 | 16.113 |      |         | Trending              |
|                  | Temperature independent| $A$                | 0.002 | 0.974| 0.011   | 5.902                 |
|                  |                       | $B$                 | 2.867 |      |         | Random                |
|                  |                       | $A$                 | 0.004 | 0.976| 0.012   | 8.628                 |
|                  |                       | $B$                 | 2.387 |      |         | Random                |
|                  |                       | $A$                 | 0.005 | 0.975| 0.013   | 10.412                |
|                  |                       | $B$                 | 2.276 |      |         | Random                |
On the other hand, the Oswin, Modified Henderson, Chun-Pfost, and Henderson models reported $R^2$ values > 0.90. In fact, these models are used as predictors of $X_e$ behavior at different relative humidities, generally for cereals and fruits [20,30,35,44,65].

Regarding the $C_{GAB}$ values, these were greater than unity, which indicates that the adsorption in the monolayer is fast, that is, the humidity at the monolayer level is achieved quickly during the first days and, as a consequence, the purple corn is prone to rapid attack by molds and yeasts [26,27,68].

Furthermore, $C_{GAB}$ is high because the surface of the purple corn grain is constituted by a large number of active centers, including polar groups of the $-\text{CO}$, $-\text{COO}^-$ and $-\text{NH}_3^+$, which allow it to establish a greater number of hydrogen bridge bonds.

On the other hand, the parameter $k_{GAB}$, which is related to the standard chemical potential between the molecules of the second layer and those of the pure liquid state, was observed to increase with temperature (Table 3), which suggests a decrease in humidity at low $a_w$ values.

While moisture at the monolayer $X_m$ level of the GAB model, was found to be around 7% d.b., which is a usual behavior for corn varieties [39,69], the fact that $X_m$ decreases with temperature indicates that at higher temperatures, the moisture loss is greater at the monolayer level for a defined relative humidity [67,69], due to the breaking of the intermolecular bonds of the hydrogen bridge type between the surface of the corn grain and the water available at the $X_e$ level. This suggests that temperature is a critical condition for the attack of molds and yeasts, which is a typical behavior of foods that follow type II isotherms [24,34,36,39,54,64].

### 3.3. Thermodynamic Parameters

The isosteric heat of sorption $q_{st}$ of purple corn was determined considering $a_w$ values calculated using the Halsey equation, for $X_e$ between 0.07 and 0.17. Figure 2a, presents the behavior of $q_{st}$ as a function of $X_e$, and it is observed that as the equilibrium humidity increases, the value of $q_{st}$ decreases from 7.7022 to 0.0165 kJ/g, meaning this behavior is a result of an initially high humidity at the monolayer level, requiring more energy to break the polar and hydrogen bonds on the surface of the corn grain, and as $X_e$ increases, the active sites that adsorb water are no longer available, which is usual in foods with a high carbohydrate content [39,67,69–71].

The $q_{st}$ values found are higher than those reported for this cereal [39,69] due to the coloration of the purple corn grain, which is related to the presence of phenolic compounds and sugars [2,5,15], thereby giving it a greater number of functional groups, with the capacity to establish a greater number of bonds with water, for which it would require more energy to eliminate it from the monolayer.

| Model   | Parameters | $R^2$ | SEE  | MAE (%) | Residual Distribution |
|---------|------------|-------|------|---------|------------------------|
| Henderson 18 °C | $k$ | 97.702 | 0.912 | 0.020 | 10.794 | Trending |
|           | $n$ | 2.518 |      |        |            |          |
| Henderson 25 °C | $k$ | 38.724 | 0.903 | 0.024 | 12.346 | Trending |
|           | $n$ | 2.005 |      |        |            |          |
| Henderson 30 °C | $k$ | 28.762 | 0.927 | 0.022 | 12.529 | Trending |
|           | $n$ | 1.809 |      |        |            |          |
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observed to increase with temperature (Table 3), which suggests a decrease in humidity at low temperatures, indicating that at higher temperatures, the moisture loss is greater at the monolayer level. This suggests that temperature is a critical condition for the attack of molds and yeasts, which is a typical behavior of foods that follow type II isotherms [24,34,36,39,54,64].

Figure 2. (a) Isosteric heat of sorption; (b) Gibbs free energy; (c) Differential entropy for purple corn; (d) Relationship between differential entropy and isosteric heat of sorption (**) evaluated at 5% significance.

The work required to make sorption active sites available was calculated using the Gibbs free energy, this being a thermodynamic indicator between the corn grain and the water [72]. Furthermore, it was observed that it decreases with the increase in temperature and humidity. At 7% humidity, ΔG was 493.82, 344.49, and 300.67 kJ/kg at 18, 25, and 30 °C, respectively, this significant change is observed at up to 15% humidity, for which similar values of ΔG are observed (Figure 2b). Furthermore, this behavior is characteristic of grains and cereals [35,39,41,42,73,74] because the available sites on the surface of the purple corn grain have been occupied, consistent with the crossing of the isotherms (Figure 1).

On the other hand, it was observed that ΔG > 0, suggesting an endergonic process, that is, a driving force, is required to initiate the binding of water molecules during adsorption, and that as Xc increases, the availability to form bonds is lower, thereby requiring less energy, which is characteristic of non-spontaneous processes when they reach equilibrium, as evidenced by the adsorption systems of purple corn at different relative humidities [41,54,75].

The availability of active sites depends on how fast water molecules are mobilized on the surface of the purple corn grain, and this was calculated using differential entropy (ΔS) [24,42]. It was observed that ΔS decreases from 16.51 to 0.19 kJ/kg K for the Xc interval between 7 to 15%, presenting a rapid drop up to Xc 11% (Figure 2c), this would be due to the greater availability of the active sites, and from this point, the mobility of the molecules decreases, related to ΔG [19,41,76].

Similarly, a linear relationship (R² > 0.99) was observed between q_st and ΔS (Figure 2d), that is, there is a direct relationship between the energy needed to bind free water to the food surface, and the mobility of water molecules at the monolayer level, so the isokinetic theory, or enthalpy–entropy compensation, applies to this experimentation [36,45,55,56].

The isokinetic temperature T_b was 476.53 K, while T_lun 297.0 K, which suggests a sorption process governed by q_st (T_b > T_lun) [60,61], which is usual behavior in seeds and grains [24,35,41,43], likewise, this comparison established that purple corn grains remain stable following structural modifications that could occur during water removal or drying in the range of the study temperatures [44,77].

The GAB isotherm parameters have a thermodynamic interpretation via the activation energy (E_a), which represents the necessary energy of the phenomena occurring at the level of the water monolayer of the corn grain surface.
Thus, the energy for water to be adsorbed towards the surface of the corn grain, to form the monolayer ($X_m$), and bind to the specific polar groups of corn, was 10.947 kJ/mol, for the interval from 18 to 30 °C (Table 4), on the other hand, the $C_{GAB}$ parameter is related to the difference in energy of the molecules adsorbed in the monolayer and the upper ones [27,44,56,68], whose value was 18.84 kJ/mol. Likewise, the parameter $k_{GAB}$, which refers to the chemical potential, that is, the energy necessary to form the bond between the water molecules and the active sites [68], was 6.82 kJ/mol.

### Table 4. Activation energy of the GAB isotherm parameters.

| Parameters | 18 °C  | 25 °C  | 30 °C  | $E_a$ (kJ/mol) |
|------------|--------|--------|--------|----------------|
| $X_m$      | 0.0764 | 0.0677 | 0.0640 | −10.947        |
| $C_{GAB}$  | 1,502,958.98 | 4,501,089.95 | 1,812,257.53 | 18.843        |
| $k$        | 0.7552 | 0.8252 | 0.8418 | 6.820          |

### 4. Conclusions

Adsorption isotherms presented crosslinking at around 75% RH and 17% $X_e$ at 18, 25, and 30 °C, suggesting adequate storage conditions at these values. The GAB and Halsey models reported a better fit and would allow for a description of the behavior of corn grain moisture at different equilibrium relative humidities. The reduction of $X_e$ between 17 and 7% occurs with an increase in the isosteric heat of adsorption, $q_{st}$, from 7.7022 to 0.0165 kJ/g, while the Gibbs free energy decreases considerably with the increase in $X_e$ at the study temperatures, showing a non-spontaneous endergonic behavior, and presents a positive linear relationship with the adsorption differential entropy, evidencing the compliance of the isokinetic theory, governed by $q_{st}$, which suggests the structural stability of corn grains during storage and drying. The activation energy showed that more energy is required to remove water molecules from the upper layers bound to the monolayer, evaluated using $C_{GAB}$.

### Author Contributions:

Conceptualization, D.C.-Q., B.S.R.-P. and R.F.A.-S.; methodology, D.C.-Q. and M.C.-F.; software, Y.C.-Q. and A.Z.-P.; validation, B.S.R.-P. and A.M.S.-R.; formal analysis, D.C.-Q., A.M.S.-R., R.F.A.-S., M.C.-F., Y.G.P.-M. and M.M.Z.-P.; investigation, D.C.-Q., T.A.-A., Y.C.-Q. and A.M.-Q.; writing—original draft preparation, D.C.-Q., Y.C.-Q. and B.S.R.-P.; writing—review and editing, D.C.-Q., B.S.R.-P. and Y.C.-Q.; supervision, D.C.-Q. and B.S.R.-P., funding acquisition, D.C.-Q. All authors have read and agreed to the published version of the manuscript.

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### Data Availability Statement:

The data presented in this study are available in this same article.

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### Conflicts of Interest:

The authors declare no conflict of interest.

### References

1. Lao, F.; Sigurdson, G.T.; Giusti, M.M. Health Benefits of Purple Corn (Zea mays L.) Phenolic Compounds. Compr. Rev. Food Sci. Food Saf. 2017, 16, 234–246. [CrossRef]
2. Del Pozo-Insfran, D.; Brenes, C.H.; Serna, S.O.; Talcott, S.T. Polyphenolic and antioxidant content of white and blue corn (Zea mays L.) products. Food Res. Int. 2006, 39, 696–703. [CrossRef]
3. Loarca-Piña, G.; Neri, M.; Figueroa, J.D.; Castaño, E.; Ramos, M.; Reynoso, R.; Mendoza, S. Chemical characterization, antioxidant and antimutagenic evaluations of pigmented corn. J. Food Sci. Technol. 2019, 56, 3177–3184. [CrossRef]
4. Lao, F.; Giusti, M.M. The effect of pigment matrix, temperature and amount of carrier on the yield and final color properties of spray dried purple corn (Zea mays L.) cob anthocyanin powders. Food Chem. 2017, 227, 376–382. [CrossRef]
5. Jing, P.; Giusti, M. Characterization of anthocyanin-rich waste from purple corncobs (Zea mays L.) and its application to color milk. J. Agric. Food Chem. 2005, 53, 8775–8781. [CrossRef] [PubMed]
6. De Pascual-Teresa, S.; Santos-Buelga, C.; Rivas-Gonzalo, J.C. LCMS analysis of anthocyanins from purple corn cob. J. Sci. Food Agric. 2002, 82, 1003–1006. [CrossRef]
7. Colombo, F.; Di Lorenzo, C.; Petroni, K.; Silano, M.; Pilu, R.; Falletta, E.; Biella, S.; Restani, P. Pigmented Corn Varieties as Functional Ingredients for Gluten-Free Products. *Foods* 2021, 10, 1770. [CrossRef] [PubMed]

8. Bacchetti, T.; Masciangelo, S.; Micheletti, A.; Ferretti, G. Carotenoids, Phenolic Compounds and Antioxidant Capacity of Five Local Italian Corn (*Zea Mays* L.) Kernels. *J. Nutr. Food Sci.* 2013, 3, 227. [CrossRef]

9. Guillón, P.; Eibes, G.; Lorenzo, J.M.; Pérez-Rodríguez, N.; Lú-Chau, T.A.; Guillón, B. Green sustainable process to revalorize purple corn cobs within a bioenergy frame: Co-production of bioactive extracts. *Sci. Total Environ.* 2020, 709, 136236. [CrossRef] [PubMed]

10. Lago, C.; Cassani, E.; Zanzi, C.; Landoni, M.; Trovato, R.; Pilu, R. Development and study of a maize cultivar rich in anthocyanins: Coloured polenta, a new functional food. *Plant Breed.* 2014, 133, 201–217. [CrossRef]

11. Escribano-Balión, M.T.; Santos-Buelga, C.; Rivas-Gonzalo, J.C. Anthocyanins in cereals. *J. Chromatogr. A* 2004, 1054, 129–141. [CrossRef] [PubMed]

12. Aguilar-Hernández, A.D.; Salinas-Moreno, Y.; Ramírez-Díaz, J.L.; Alemán-De la Torre, I.; Bautista-Ramírez, E.; Flores-López, H.E. Anthocyanins and color in grain and cob of peruvian purple corn grown in Jalisco, Mexico. *Rev. Mex. Cienc. Agrícolas* 2019, 10, 1071–1082. [CrossRef]

13. Cuevas-Montilla, E.; Hillebrand, S.; Antezana, A.; Winterhalter, P. Soluble and bound phenolic compounds in different Bolivian purple corn (*Zea mays* L.) Cultivars. *J. Agric. Food Chem.* 2011, 59, 7068–7074. [CrossRef] [PubMed]

14. Yang, Z.; Zhai, W. Identification and antioxidant activity of anthocyanins extracted from the seed and cob of purple corn (*Zea mays* L.). *Innovative Food Sci. Emerg. Technol.* 2011, 11, 169–176. [CrossRef]

15. Chen, C.; Weng, Y. Moisture Sorption isotherms of Oolong tea. *Food Bioprocess Technol.* 2010, 3, 226–233. [CrossRef]

16. Aghazadeh, N.; Esmaili, M.; Mohtarami, F. Prediction of Equilibrium Moisture Contents of Black Grape Seeds (*Siah Sardasht Cultivar*) at Various Temperatures and Relative Humidity: Shelf-Life Criteria. *Nutr. Food Sci. Res.* 2021, 8, 45–52.

17. Fleurat-Lessard, F. Integrated management of the risks of stored grain spoilage by seed borne fungi and contamination by storage mould mycotoxins—An update. *J. Stored Prod. Res.* 2017, 71, 22–40. [CrossRef]

18. Udomkun, P.; Argyropoulos, D.; Nagle, M.; Mahayothee, B.; Müller, J. Sorption behaviour of papayas as affected by compositional and structural alterations from osmotic pretreatment and drying. *J. Food Eng.* 2015, 157, 14–23. [CrossRef]

19. Anwar, F.; Naseer, R.; Bhanger, M.I.; Ashraf, S.; Talpur, F.N.; Aladedunye, F.A. Physico-chemical characteristics of citrus seeds and seed oils from Pakistan. *J. Am. Oil. Chem. Soc.* 2008, 85, 321–330. [CrossRef]

20. Pumacahua-Ramos, A.; Gómez, J.A.; Telis-Romero, J.; Villa-Vélez, H.A.; Lopes, J.F. Isotherms and isosteric heat of sorption of two varieties of Peruvian quinoa. *Sci. Agropecu.* 2016, 7, 409–417. [CrossRef]

21. De Oliveira, G.H.H.; Aragão, D.M.S.; De Oliveira, A.P.L.; Silva, M.G.; Gusmão, A.C.A. Modelagem e propriedades termodinâmicas na secagem de morangos. *Braz. J. Food Technol.* 2015, 18, 314–321. [CrossRef]

22. Chen, C. Validation of the Component Model for Prediction of Moisture Sorption Isotherms of Two Herbs and other Products. *Foods* 2019, 8, 191. [CrossRef] [PubMed]

23. Ramírez-Miranda, M.; Cruz y Victoria, M.T.; Vizcarra-Mendoza, M.G.; Anaya-Sosa, I. Determination of moisture sorption isotherms and their thermodynamics properties of nixtamalized maize flour. *Rev. Mex. Ing. Química* 2014, 13, 165–178.

24. Isquierdo, E.P.; Caldeira, D.S.A.; Siqueira, V.C.; Martins, E.A.S.; Quequeto, W. Fittings of adsorption isotherm models and thermodynamic properties of urunday seeds. *Eng. Agric.* 2020, 40, 374–380. [CrossRef]

25. Gonelli, A.L.D.; Corréa, P.C.; Oliveira, G.H.H.; Afonso Junior, P.C. Water sorption properties of coffee fruits, pulped and green coffee. *LWT—Food Sci. Technol.* 2013, 50, 386–391. [CrossRef]

26. Saberi, B.; Vuong, Q.V.; Chockchaisawasdee, S.; Golding, J.B.; Scarlett, C.J.; Statopoulos, C.E. Water Sorption Isotherm of Pea Starch Edible Films and Prediction Models. *Foods* 2015, 5, 1. [CrossRef] [PubMed]

27. Tallá, A. Predicting sorption isotherms and net isosteric heats of sorption of maize grains at different temperaturas. *Int. J. Food Eng.* 2014, 10, 393–401. [CrossRef]

28. Moussaoui, H.; Bahammou, Y.; Idlimam, A.; Lambarrar, A.; Abdennouri, N. Investigation of hygroscopic equilibrium and modeling sorption isotherms of the argan products: A comparative study of leaves, pulps, and fruits. *Food Bioprod. Process.* 2019, 114, 12–22. [CrossRef]

29. Saleh, R.M.; Karim, N.A.; Hensel, O.; Sturm, B. Mathematical modelling of adsorption isotherms of Malaysian variety of purple flesh sweet potato at different temperatures. *Therm. Sci. Eng. Prog.* 2018, 7, 326–330. [CrossRef]

30. Fonseca, N.N.; Resende, O.; Ferreira, J.W.N.; Silva, L.C.D.M.; Andrade, E.G.; Oliveira, L.P. Desorption isotherms of graniferous sorghum grains. *Res. Soc. Des.* 2020, 9, 1–16. [CrossRef]

31. Resende, O.; Oliveira, D.E.C.; Costa, L.M.; Ferreira, W.N. Thermodynamic properties of baru fruits (Dipteryx alata Vogel). *Eng. Agric.* 2017, 37, 739–749. [CrossRef]

32. Staudt, P.B.; Kechinski, C.P.; Tessaro, I.C.; Marczak, L.D.F.; Soares, R.D.P.; Cardozo, N.S.M. A new method for predicting sorption isotherms at different temperatures using the BET model. *J. Food Eng.* 2013, 114, 139–145. [CrossRef]

33. Zeymer, J.S.; Corrêa, P.C.; Oliveira, G.H.; Baptestini, F.M.; Campos, R.C. Mathematical modeling and hysteresis of sorption isotherms for paddy rice grains. *Eng. Agric.* 2019, 39, 524–532. [CrossRef]

34. Bustos-Vanegas, J.D.; Corrêa, P.C.; Zeymer, J.S.; Baptestini, F.M.; Campos, R.C. Moisture sorption isotherms of quinoa seeds: Thermodynamic analysis. *Eng. Agric.* 2018, 38, 941–950. [CrossRef]
35. Bessa, J.F.V.; Resende, O.; De Oliveira, D.E.C.; De Lima, R.R.; Quequeto, W.D.; Siqueira, V.C. Adsorption isotherms and thermodynamic properties of Carthamus tinctorius L. seeds. Rev. Bras. Eng. Agric. E Ambient. 2021, 25, 696–702. [CrossRef]
36. Hassini, L.; Bettaiab, E.; Desmoulires, H.; Torres, S.S.; Touil, A. Desorption isotherms and thermodynamic properties of prickly pear seeds. Ind. Crops Prod. 2015, 67, 457–465. [CrossRef]
37. Botelho, F.M.; Neto, N.J.B.; Botelho, S.C.J.; De Oliveira, G.H.H.; Hauth, M.R. Sorption isotherms of Brazil nuts. Rev. Bras. Agric. E Ambient. 2019, 23, 776–781. [CrossRef]
38. Ade, A.R.; Ajay, E.A.; Raji, A.O.; Adetayo, S.A.; Arowora, K.A. Moisture sorption isotherms of Mesquite seed (Prosopis africana). Agric. Eng. Int. CIGR J. 2016, 18, 273–281.
39. Oliveira, G.H.H.; Corrêa, F.C.; Araiô, E.F.; Valente, D.S.M.; Botelho, F.M. Desorption isotherms and thermodynamic properties of sweet corn cultivars (Zea Mays L.). Int. J. Food Sci. Technol. 2010, 45, 546–554. [CrossRef]
40. Choque-Quispe, D.; Ramos-Pacheco, B.S.; Solano-Reynoso, A.M.; Ligarda-Samanez, C.A.; Choque-Quispe, Y.; Peralta-Guevara, D.E.; Quispe-Quispe, Y. Drying and color in punamuña leaves (Satureja boliviensis). DYNA 2021, 88, 31–37. [CrossRef]
41. Zeymer, J.S.; Corrêa, P.C.; De Oliveira, G.H.H.; Baptestini, F.M. Thermodynamic properties of water desorption in lettuce seeds. Semin. Cienc. Agron. 2018, 39, 921–932. [CrossRef]
42. Moreira, R.; Chenlo, F.; Torres, M.D.; Vallejo, N. Thermodynamic analysis of experimental sorption isotherms of loquat and quince fruits. J. Food Eng. 2008, 88, 514–521. [CrossRef]
43. Campos, R.C.; Corrêa, P.C.; Zaidan, I.R.; Zaidan, U.R.; Leite, R.A. Moisture sorption isotherms of sunflower seeds: Thermodynamic analysis. Ciênc. E Agrotecnol. 2019, 43, 2. [CrossRef]
44. Wang, P.; Fu, N.; Li, D.; Wang, L. Predicting Storage Conditions for Rice Seed with Thermodynamic Analysis. Int. J. Food Eng. 2017, 13, 20170129. [CrossRef]
45. Barati, M.; Zare, D.; Zomorodian, A. Moisture sorption isotherms and thermodynamic properties of safflower seed using empirical and neural network models. Food Meas. 2016, 10, 236–246. [CrossRef]
46. Labuza, T.P. Moisture Sorption: Practical Aspects of Isotherm Measurement and Use; American Association of Cereal Chemists: St. Paul, MN, USA, 1984; ISBN 0913250341.
47. Corrêa, P.C.; Baptestini, F.M.; Vanegas, J.D.B.; Leite, R.; Botelho, F.M.; De Oliveira, G.H.H. Kinetics of water sorption of damaged bean grains: Thermodynamic properties. Rev. Bras. Eng. Agric. E Ambient. 2017, 21, 556–561. [CrossRef]
48. Silva, H.W.; Costa, L.M.; Resende, O.; Oliveira, D.E.; Soares, R.S.; Vale, L.S. Thermodynamic properties of pepper seeds—Variety “Cabacinha”. Cientifica 2016, 44, 14–22. [CrossRef]
49. Soleimani, M.; Tabil, L.; Shahedi, M.; Emani, S. Sorption isotherm of hybrid seed corn. In Proceedings of the Canadian Society for Engineering in Agricultural, Food, Environmental, and Biological Systems—CSBE, Edmonton, AB, Canada, 16–19 July 2006. [CrossRef]
50. Karunanithy, C.; Muthukumarappan, K.; Donepudi, A. Moisture Sorption Characteristics of Corn Stover and Big Bluestem. J. Renew. Energy 2013, 2013, 4. [CrossRef]
51. Resende, O.; Corrêa, C.P.; Gonell, A.L.D.; Ribeiro, D.M. Isotermas e Calor Isostático de Sorção do Feijão. Ciênc. Tecnol. Aliment. 2006, 26, 626–631. [CrossRef]
52. Tsami, E.; Desmorieux, H.; Torres, S.S.; Touil, A. Desorption isotherms and thermodynamic properties of prickly pear seeds. Ind. Crops Prod. 2015, 67, 457–465. [CrossRef]
53. Rizvi, S.S. Thermodynamic properties of foods in dehydration. In Engineering Properties of Foods; Taylor & Francis: Boca Raton, FL, USA, 2005; pp. 259–346. [CrossRef]
54. McMinn, W.A.M.; Al-Muhtaseb, A.H.; Magee, T.R.A. Enthalpy-entropy compensation in sorption phenomena of starch materials. Food Res. Int. 2005, 38, 505–510. [CrossRef]
55. Gabas, A.L.; Menegalli, F.C.; Telis-Romero, J. Water sorption enthalpy-entropy compensation based on isotherms of plum skin and pulp. J. Food Sci. 2000, 65, 680. [CrossRef]
56. Beristain, C.I.; Garcia, H.S.; Azuara, E. Enthalpyentropy compensation in food vapor adsorption. J. Food Eng. 1996, 30, 405–415. [CrossRef]
57. Madamba, P.S.; Driscoll, R.H.; Buckle, K.A. Enthalpy-entropy compensation models for sorption and browning of garlic. J. Food Eng. 1996, 28, 109–119. [CrossRef]
58. Tsami, E. Net isosteric heat of sorption in dried fruits. J. Food Eng. 1991, 14, 327–335. [CrossRef]
59. Krug, R.R.; Hunter, W.G.; Grier, R.A. Enthalpy-entropy compensation. 1. Some fundamental statistical problems associated with the analysis of van’t Hoff and Arrhenius data. J. Phys. Chem. 1976, 80, 2335–2341. [CrossRef]
60. Ryde, U.L.F. A fundamental view of enthalpy-entropy compensation. MedChemComm 2014, 5, 1324–1336. [CrossRef]
61. Leffler, J.E.; Grunwald, E. Rates and Equilibria of Organic Reactions: As Treated by Statistical, Thermodynamic and Extrathermodynamic Methods; Elsevier: Amsterdam, The Netherlands, 1963; ISBN 978-1-62198-643-0.
62. Vega, A.; Lara, E.; Lemus, R. Isotermas de adsorción en harina de maíz (Zea mays). Food Sci. Technol. 2006, 26, 4. [CrossRef]
63. Condon, J.B. Surface Area and Porosity Determinations by Physisorption: Measurements and Theory; Elsevier: Amsterdam, The Netherlands, 2006; ISBN 978-0-444-51964-1.
64. Silva, K.S.; Romero, J.T.; Mauro, M.A. Sorption isotherms and thermodynamic analysis of seed fruits used to obtain vegetable oil. Lat. Am. Appl. Res. 2015, 45, 21–26. [CrossRef]
65. Miranda, M.; Vega-Gálvez, A.; Sanders, M.; López, J.; Lemus-Mondaca, R.; Martínez, E.; Di Scala, K. Modelling the water sorption isotherms of quinoaseeds (Chenopodium quinoa Willd.) and determination of sorption heats. *Food Bioprocess Technol.* 2012, 5, 1686–1693. [CrossRef]

66. Corrêa, P.C.; Botelho, F.M.; Botelho, S.C.C.; Goneli, A.L.D. Isotermas de sorção de água de frutos de Coffea canephora. *Rev. Bras. Eng. Agríc. E Ambient.* 2014, 18, 1047–1052. [CrossRef]

67. Majd, K.M.; Karpavarfard, S.H.; Farahnaky, A.; Ansari, S. Thermodynamic properties of water sorption isotherms of grape seed. *Int. Agrophysics* 2014, 28, 63–71. [CrossRef]

68. Enrione, J.I.; Hill, S.E.; Mitchell, J.R. Sorption behavior of mixtures of glycerol and starch. *J. Agric. Food Chem.* 2007, 55, 2956–2963. [CrossRef] [PubMed]

69. Samapundo, S.; Devlieghere, F.; De Meulenaer, B.; Atukwase, A.; Lamboni, Y.; Debevere, J.M. Sorption isotherms and isosteric heats of sorption of whole yellow dent corn. *J. Food Eng.* 2007, 79, 168–175. [CrossRef]

70. Choque-Quispe, D.; Ligarda-Samanez, C.A.; Ramos-Pacheco, B.S.; Taipe-Pardo, F.; Peralta-Guevara, D.E.; Solano-Reynoso, A.M. Evaluation of sorption isotherms of grains and flour of amaranth (*Amaranthus caudatus*). *Rev. ION* 2018, 31, 67–81. [CrossRef]

71. Labuza, T.P.; Kaanane, A.; Chen, J.Y. Effect of temperature on the moisture sorption isotherm and water activity shift of two dehydrated foods. *J. Food Sci.* 1985, 50, 385–391. [CrossRef]

72. Telis, V.R.N.; Gabas, A.L.; Menegalli, F.C.; Telis-Romero, J. Water sorption thermodynamic properties applied to persimmon skin and pulp. *Thermochim. Acta* 2000, 343, 49–56. [CrossRef]

73. De Oliveira, D.E.C.; Resende, O.; Chaves, T.H.; Souza, K.A.; Smaniotto, T.A.D.S. Propriedades termodinâmicas das sementes de pinhão manso. *Biosci.* 2014, 30, 147157.

74. De Oliveira, D.E.C.; Resende, O.; Smaniotto, T.A.D.S.; De Sousa, K.A.; Campos, R.C. Propriedades termodinâmicas de grãos de milho para diferentes teores de água de equilíbrio. *Pesqui. Agropecu. Trop.* 2013, 43, 50–56. [CrossRef]

75. Nkolo, M.Y.N.; Noah, N.J.; Bardet, S. Effect of enthalpy–entropy compensation during sorption of water vapour in tropical woods: The case of Bubinga (*Guibourtia tessmannii* J. Leéonard; *G. pellegriniana* J. L.). *Thermochim. Acta* 2008, 468, 1–5. [CrossRef]

76. Ayala-Aponte, A.A. Thermodynamic properties of moisture sorption in cassava flour. *DYNA* 2016, 83, 139–145. [CrossRef]

77. Toshkov, N.; Lazarov, L.; Popova, V.; Ivanova, T.; Menkov, N. Thermodynamics of moisture sorption in tobacco (*Nicotiana tabacum* L.) seeds. *E3S Web Conf.* 2020, 207, 01019. [CrossRef]