TLR7 Influences Germinal Center Selection in Murine SLE

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Abstract

TLR7 enhances germinal center maturation and migration of B cells to the dark zone where proliferation and somatic hypermutation occur. Our goal was to determine how Tlr7 dose influences selection of the autoreactive B cell repertoire in NZW/BXSB. Yaa mice bearing the site-directed heavy chain transgene 3H9 that encodes for the TLR7 regulated anti-CL response. To create a physiologic setting in which autoreactive B cells compete for survival with non-autoreactive B cells, we generated bone marrow chimeras in which disease onset occurred with similar kinetics and the transferred 3H9+ female non-Yaa, male Yaa or male TLR7-/Yaa cells could be easily identified by positivity for GFP. Deletion of 3H9 B cells occurred in the bone marrow and the remaining 3H9 follicular B cells manifested a decrease in surface IgM. Although there were differences in the naïve repertoire between the chimeras it was not possible to distinguish a clear pattern of selection against lupus related autoreactivity in TLR7-/Yaa or female chimeras. By contrast, preferential expansion of 3H9+ B cells occurred in the germinal centers of male Yaa chimeras. In addition, although all chimeras preferentially selected 3H9/Vκ5 encoded B cells into the germinal center and plasma cell compartments, 3H9 male Yaa chimeras had a more diverse repertoire and positively selected the 3H9/Vκ5-48/Jκ4 pair that confers high affinity anti-cardiolipin activity. We were unable to demonstrate a consistent effect of Tlr7 dose or Yaa on somatic mutations. Our data show that TLR7 excess influences the selection, expansion and diversification of B cells in the germinal center, independent of other genes in the Yaa locus.

Introduction

Systemic lupus erythematosus (SLE) is an autoimmune disorder in which pathogenic autoantibodies directed to ubiquitous nuclear material initiate systemic inflammation. SLE patients have defective negative selection of autoreactive B cells at the immature and transitional checkpoints [1] and also fail to restrain pathogenic effector B cells arising in the germinal center (GC) [2–3]. Understanding how these defects contribute to pathogenic autoantibody production will allow therapy for SLE to be directed to the appropriate B cell developmental stage.
TLR7 is an endosomal TLR that recognizes single-stranded viral RNA and its expression in B cells is required for the generation of anti-RNA antibodies in SLE [4–5]. Haplodeficiency of TLR7 in SLE-prone mice bearing the Yaa locus also moderately decreases anti-DNA antibodies in addition to its effect on the anti-RNA response [6–7]. Engagement of TLR7 induces signaling through its adaptor MyD88 resulting in activation of the NFκB and Type 1 interferon pathways [8–9]. B cell intrinsic TLR signaling is amplified in GC B cells compared to follicular B cells, suggesting that TLRs play a role in the development of the antigen activated antibody repertoire [10–11]. TLR signaling drives B cells into the dark zone of the germinal center where they undergo clonal expansion, and differentiation to memory cells [12]. In accord with this data, mice with a B cell specific Tlr7 deficiency have impaired anti-viral responses due to decreased entry of B cells into the GC dark zones where clonal proliferation and somatic mutation occur [13]. Two recent studies have shown that in lupus models the complete absence of TLR7 compromises B cell survival and abrogates spontaneous germinal center formation and the production of anti-Sm/RNP, anti-dsDNA, anti-cardiolipin (CL) and anti-nucleosome antibodies in a B cell intrinsic manner. By contrast, deficiency of MyD88 in macrophages and dendritic cells has no effect on germinal centers [14–15].

NZW/BXSB F1 (W/B) male mice bearing the Yaa locus have a duplication of part of the X chromosome that includes the Tlr7 gene onto the Y chromosome [16–17] and therefore have a 2-fold increase in Thr7 expression. Male W/B mice spontaneously develop high titer anti-CL and anti-Sm/RNP autoantibodies that are associated with both anti-phospholipid syndrome and glomerulonephritis whereas females, with only one copy of Thr7, develop a milder disease [18]. Although there are at least 16 genes in the Yaa locus, the Thr7 duplication is the dominant genetic contributor to the Yaa phenotype [7, 19–20]. Furthermore, 4 to 8-fold overexpression of Thr7 is sufficient to induce spontaneous onset of SLE in non-autoimmune strains [19].

The purpose of our experiments was to use W/B mice bearing the site-directed anti-CL/DNA autoantibody VH transgene 3H9 [21–22] to determine how an extra Tlr7 dose influences the selection of naïve and antigen activated autoreactive B cells during the evolution of SLE. We previously showed that 3H9 male NZW/BXSB transgenic mice develop high titer anti-DNA and anti-CL antibodies and develop proteinuria whereas females have a delay in the emergence of autoantibodies and do not become proteinuric. Furthermore although both male and female W/B mice use the 3H9 transgene to encode anti-chromatin and anti-CL antibodies they have differences in selection of the GC repertoire [21]. In these experiments however, there was no competition with non-3H9 B cells and transgenic females did not develop clinical disease. We were therefore not able to control for systemic inflammation or to distinguish the effects of the Yaa locus from Thr7 duplication on autoantibody production and clinical disease. In the studies reported herein we examined B cell tolerance mechanisms in bone marrow chimeras in which we could study repertoire selection in the setting of physiologic competition with non-autoreactive B cells and evolving inflammation. Our data suggest that an increase in Thr7 dose enhances B cell proliferation, selection and diversification in the germinal center.

Methods

Mice

C57BL/6 TLR7−/− mice (from Dr Shizuo Akira, University of Osaka) and GFP+ C57BL/6 mice (Jackson Laboratories, Bar Harbor, ME) were backcrossed >13 generations to NZW mice and then crossed with 3H9 NZW mice (from Dr. Tony Marion, University of Tennessee). 3H9, GFP+NZW and 3H9.GFP+NZWTLR7−/− females were bred with BXSB males (Jackson Laboratories) to produce GFP+ male NZW/BXSB F1 progeny containing either one (W/BTLR7−/Yaa) or
two (W/B) copies of Tlr7 with or without the 3H9 transgene (S1 Table). Wild type (wt) NZW/BXSB F1 mice were made by crossing the parental strains (Jackson Labs, Bar Harbor, ME).

Bone marrow chimeras were generated by transfer of 50% 3H9.GFP+ and 50% male wild-type (wt) W/B bone marrow into totally irradiated 8–12-wk-old W/B recipients. In the experimental groups, the 3H9.GFP+ bone marrow donors were male, male TLR7-/Yaa or female and the bone marrow recipients were universally male. A group of 50% female 3H9.GFP+ and 50% female wt donors into female recipients served as non-diseased controls. Groups of control chimeras were made using GFP+ non-3H9 donors (Table 1). Chimeric mice were tested for proteinuria every 2 weeks (Multistick; Fisher, Pittsburgh, PA) and bled periodically for serologic analyses [18, 23]. Mice were sacrificed at the onset of proteinuria 12–20 wk after transplant (Table 1). Studies were approved by the IACUC of the Feinstein Institute for Medical Research (Protocols 2009–054 and 2007–038).

Quantitation of autoantibodies to CL and dsDNA
ELISpot assays of spleen cells and serial ELISA assays of serum for antibodies to CL and dsDNA were performed as previously described [18, 24–25]. Reconstituted 3H9/Vκ antibodies were expressed and cotransfected into 293T cells as previously described [21]. Supernatants were normalized to 1µg/ml and tested in serial dilution for binding to CL and dsDNA as above and to histones and chromatin as previously described. These assays were also performed after treatment of the supernatants with DNase I to remove associated nuclear material present in the cell supernatants [21].

Flow cytometry and sorting
Spleen and bone marrow B cells were analyzed as previously described [26–28]. For each stain the FITC channel was left open to enumerate GFP+ cells.

Single-cell sorting and PCR
GFP+ single cells were sorted from FO, GC and PC B cell subsets from 4–6 mice per group and cDNA preparation and PCR of the 3H9 heavy chain and associated Vκ light chain were performed as previously described [21, 29]. PCR products were sequenced by Genewiz (South Plainfield, NJ), and sequences were identified using the International ImMunoGeneTics (IMGT) database. To distinguish between Vκ5–43/45 C3 and Vκ5–45 C3 that differ by three base pairs at the 5’ end and to enumerate somatic mutations, a 5’ leader primers for Vκ5–43/45 C3 and Vκ5–48 C3 were used to generate full length light chain genes for resequencing. Somatic mutation rates were calculated using a program available on the IMGT website. Similarly, a VH82′01 leader primer was used to generate full length 3H9 heavy chain genes.

Immunohistochemistry
Spleen tips were incubated in 4% formaldehyde/10% sucrose for 2 hours before freezing in OCT. 7 micron sections were stained with antibodies to IgD and visualized as previously described [30].

Statistics
Proteinuria and survival data were analyzed using Kaplan-Meier curves and log rank test. Comparisons shown in Figs. 1A, 2 and 3A were performed using Mann-Whitney test. p values ≤ 0.05 were considered significant. Comparisons in Fig. 4 were performed using pair-wise Mann-Whitney test with a Bonferroni adjustment to account for multiple testing. In order to
Table 1. Donor strains and chimeras.

| Chimera Name | Donor 1 NZW/BXSB (GFP+) | Donor 2 NZW/ BXSB | Harvest (Days after transplant) | Recipient NZW/ BXSB |
|--------------|--------------------------|-------------------|---------------------------------|--------------------|
|              | TLR7 | Yaa | 3H9 |                |                                      |                |
| M WT         | M    | ++  | +   | -               | M                    | 86 +/- 16       | M |
| F WT         | F    | +   | -   | -               | M                    | 119 +/- 32      | M |
| M 3H9        | M    | ++  | +   | -               | M                    | 95 +/- 31       | M |
| F 3H9        | F    | +   | -   | +               | M                    | 124 +/- 32      | M |
| M 3H9TLR7-/Yaa | M    | +   | +   | +               | M                    | 95 +/- 22       | M |
| F 3H9 to F*  | F    | +   | -   | +               | F                    | 128 +/- 14      | F |

*Chimeras not proteinuric at time of euthanasia

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Fig 1. Attenuated disease in NZW/BXSB TLR7-/Yaa males. A: Titers of IgG anti-CL antibodies were lower in NZW/BXSB TLR7-/Yaa males (grey) compared with their WT littermates (black). B: Autoantibody secreting cells (ASC) were decreased in NZW/BXSB TLR7-/Yaa males (grey) compared with their WT littermates (black). B, C: NZW/BXSB TLR7-/Yaa males had a delay in onset of proteinuria (C) and an improved survival (D) compared with their WT littermates or historical male controls. * p < 0.05, † p < 0.01, ‡ p <.001.

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preserve an overall 5% significance level, a pairwise comparison was considered statistically significant if \( p < 0.025 \). For Fig. 5 the Kruskal-Wallis test was used to compare the M and F wt with their respective experimental groups. Upon finding a significant difference, Bonferroni adjusted multiple pairwise comparisons of the WT group to each of the other two gender-matched groups, were carried out using Mann-Whitney tests. A \( p \) value of \(<0.025\) was considered significant. For all male vs. female comparisons the Mann-Whitney test was used. Statistical analysis of the V\( \kappa \) repertoire data shown in Fig. 6 was performed as previously described [31]. Owing to the typically small sample sizes of single cells, the usual Pearson \( \chi^2 \) statistic is not valid for determining whether differences exist in the distributions of L chains across B cell subsets. Instead, the Fisher exact test is applicable. However, the computing time required for running the exact test on large, sparse tables is prohibitive and, moreover, the power of the test is low. Accordingly, identifying L chains of interest without using formal
inferential methods (i.e., without using $p$ values or confidence intervals) is needed. This exploratory approach is frequently used in identifying interesting genes in gene expression microarray studies [32–34]. To this end, we used a novel method for “screening” the $r \times c$ table for cell frequencies that appear to be inconsistent with the null hypothesis that the B cell subset and L chain distributions are independent. This method uses a graphical approach (scree plots) to finding the entries in the table that depart most from the null hypothesis. For additional details on the statistical methodology please refer to Lesser et al [35]. Comparisons in Fig. 7 were performed using chi-square analysis. Mutation frequencies in each subset were compared using the Mann-Whitney test.

Fig 3. Clinical outcome of the chimeras: A: Serum levels of anti-CL antibodies (mean + SEM) in the female (circles) and male (squares) chimeras. Antibody titers increased with age in all the chimeras. Levels of anti-CL antibodies were lower in the female chimeras by age 16–18 weeks. Comparisons are with male WT controls. * $p < 0.05$, † $p < 0.01$. B: Time to fixed proteinuria onset. Note the marked delay in the 3H9 F to F chimeras ($p < 0.0001$ vs. males) and the modest delay in the 3H9 F to M chimeras ($p < 0.0025$ vs. males).

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Fig 4. Deletion of 3H9 B cells occurs in the bone marrow. A. Ratio of GFP+ B cells: GFP+ B220− non-B cells in bone marrow subsets shows deletion occurring at the pre-B stage in the 3H9 chimeras (upper symbols show comparisons of pre-B cell ratios with those of WT female or male counterparts). Further deletion occurred between the pre-B and transitional (Trans) and recirculating (Recirc) B cells (lower row of symbols). n = 5–12 per group, * p < 0.025, † p < 0.01. B. Ratio of GFP+ B cells: GFP+ T cells in spleens.
Results

Phenotype of NZW/BXSB<sup>TLR7-/Yaa</sup> mice

The phenotype of male NZW/BXSB<sup>TLR7-/Yaa</sup> mice has not been reported and was compared with that of wt male littersmates and with historical male and female W/B controls. NZW/BXSB<sup>TLR7-/Yaa</sup> mice had lower titers of anti-CL antibodies than wt controls up until the age of 16 weeks (Fig. 1A). Anti-dsDNA antibody titers appeared after 12 weeks of age in the wt mice and trended lower in the NZW/BXSB<sup>TLR7-/Yaa</sup> mice after the age of 15 weeks (p = 0.07, not shown). Only low titers of anti-Sm/RNP antibodies were present by 18 weeks of age with a trend towards lower titers in NZW/BXSB<sup>TLR7-/Yaa</sup> mice (p = 0.09, not shown). Fewer antibody and autoantibody secreting B cells, particularly IgG anti-CL producing B cells were present in the spleens of NZW/BXSB<sup>TLR7-/Yaa</sup> mice harvested at 18–22 weeks of age, compared with their age matched wt littermates (Fig. 1B). These data show that TLR7 dose regulates the anti-CL response in wt NZW/BXSB males. Proteinuria onset was significantly delayed in NZW/BXSB<sup>TLR7-/Yaa</sup> mice compared with wt littermates (Fig. 1C, p = 0.011) and this was associated with a decrease in their mortality (Fig. 1D, p = 0.05). Proteinuria onset and survival were indistinguishable between wt littermates and historical male controls [18]. Survival was modestly decreased in NZW/BXSB<sup>TLR7-/Yaa</sup> mice compared with historical female controls [23] but this difference did not reach statistical significance.

The spleens of NZW/BXSB<sup>TLR7-/Yaa</sup> mice harvested between 18–22 weeks of age were smaller than those of their wt littermates and contained fewer total B cells, dendritic cells and activated B and T cells reflecting a decreased overall inflammatory burden (Fig. 2A, B). NZW/BXSB<sup>TLR7-/Yaa</sup> mice displayed a normal T1:T2 ratio and normal percentage of marginal zone B cells.
Fig 6. Repertoire analysis of 3H9-associated Vκ chains in follicular (A) germinal center (B) and plasma cell (C) subsets of chimeric mice. Genes that were in the top 10 contributors to the χ² value and constitute >2.5% of the repertoire in any of the comparisons are shown. 55–200 sequences from 3–6 mice per group were analyzed per subset. The y axis shows percent of the total repertoire for each subset. Black arrows indicate that the reconstituted 3H9/Vκ encodes an autoreactive antibody. Grey arrows indicate that the reconstituted 3H9/Vκ encodes a non-autoreactive antibody. The complete dataset is shown in S2 Table.

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cells. By contrast wt mice manifested a reversed T1:T2 ratio (p = 0.05) and had a decrease in frequency (p = 0.0007) but not the total number of marginal zone B cells. In accordance with the serologic data, NZW/BXSB TLR7-/Yaa mice had lower numbers of class switched B cells than their wt littermates (p = 0.05) and a trend towards a decrease in plasma cells (p = 0.09, Fig. 2C, D).

**Phenotype of 3H9 W/B bone marrow chimeras**

Male 3H9 (M 3H9), male 3H9 TLR7-/Yaa (M 3H9 TLR7-/Yaa), female 3H9 (F 3H9), male wt (M WT) mice, and female wt (F WT) chimeric mice (Table 1) developed IgG autoantibodies to CL by 10–13 weeks post-transplant. By 16–18 weeks, titers of anti-CL antibodies were lower in female chimeras than in M WT chimeras regardless of 3H9 status but there was no difference between M WT and either M 3H9 or M 3H9 TLR7-/Yaa chimeras (Fig. 3A). Development of high titer IgG anti-DNA antibodies was sporadic in each group, with a significant increase in titers at 14–15 weeks post-transplant only in the M WT chimeras (data not shown).

Proteinuria was markedly delayed in the F 3H9 to F chimeric mice (p < 0.0001 vs. males) and developed with a modest delay in the female to male compared to the male chimera sets regardless of 3H9 status (p = 0.0025 M vs. F). M 3H9 TLR7-/Yaa chimeras developed proteinuria with similar kinetics to the male controls (Fig. 3B, Table 1). These data, in sum, suggest that the Yaa locus confers a slight disadvantage beyond the major effect conferred by the extra copy of Tlr7 in NZW/BXSB males.

**B cell phenotype of W/B bone marrow chimeric mice**

Phenotypic analysis of spleen B cells at the time of sacrifice indicated that F 3H9 to F chimeras had a higher frequency of MZ B cells and a lower frequency of FO B cells and plasma cells than...
all the other chimeras (S1A Fig.). This is consistent with disease protection in fully female mice. The spleens of these mice were also significantly smaller than those of the other chimeras (52.3 +/- 26.8 x 10^6 cells compared with 149.4 +/- 78.7 x 10^6 cells in M 3H9 chimeras, p < 0.0001, and 126.3 +/- 55.3 x 10^6 cells in F 3H9 chimeras, p < 0.01) such that the number of FO B cells, GC B cells and plasma cells was significantly less. All the other chimeras phenotypically resembled M WT (S1B Fig. and data not shown).

As a measure of the selection pressure against transgenic B cells we assessed deletion of 3H9 B cells in each subset by calculating the ratio of % GFP+ B cells to % GFP+ non-B cells (BM non-B cells for BM and average of spleen CD4 and CD8 cells for spleens). In the absence of deletion this ratio should be 1. Ratios of % GFP+ CD4 cells to % GFP+ CD8 or CD11b+ cells were close to 1 for all chimeras (not shown) indicating no selection pressure against these cell types based on their TLR7 dose. By contrast, all the 3H9 chimeras exhibited deletion of GFP+/3H9+ transgenic B cells in the bone marrow. This was already evident at the pre-B cell stage (Fig. 4A) with further deletion occurring between the pre-B cell and transitional stages in the BM. No further deletion occurred between the transitional and recirculating or between the peripheral T1 and FO stages (Fig. 4B). Thus, central deletion is the major contributor to tolerance induction of 3H9 B cells in W/B mice. Furthermore, deletion of 3H9+ cells was more stringent in 3H9 F to F chimeras than in the male chimeras (spleen CD19+/non-B ratio 0.28 +/- 0.04 vs. 0.53 +/- 0.17 in M 3H9, p = 0.01; 0.51 +/- 0.17 in M 3H9TLR7-/Yaa, p < 0.05) with an intermediate phenotype of the F 3H9 chimeras (Fig. 4B). By contrast, GFP+ B cells were not deleted in the BM when the donors were 3H9 negative. These data were confirmed by immunohistochemical studies that showed follicular exclusion of GFP+ cells in the 3H9 chimeras but not in the WT chimeras (Fig. 4C-F).

Surface IgM staining was decreased on splenic GFP+ follicular cells from 3H9 chimeric mice compared to GFP- follicular B cells from the same mice or to non-3H9 GFP+ cells from M or F wt chimeras, suggesting that the remaining 3H9+ cells have encountered autoantigen in the periphery [36–37] (Fig. 5A). This BCR downregulation did not however prevent 3H9+ cells from entering the GCs or appearing in the plasma cell compartment in any of the chimeras. Importantly, GFP+ transgenic B cells were preferentially enriched in the GCs of M 3H9 compared to F 3H9 and M 3H9TLR7-/Yaa chimeric mice (Fig. 5B). This suggests an effect of TLR7 dose on GC selection and/or proliferation of autoreactive B cells. A similar effect was observed in the plasma cell compartment (Fig. 5C).

Single-cell PCR analysis of the transgenic 3H9 B cell light chain repertoire

To determine whether Tlr7 dose and/or the Yaa locus influenced selection of the naïve or antigen selected repertoires we performed single cell PCR of light chains associated with the 3H9 heavy chain in selected B cell subsets. We have previously established that reconstituted unmutated 3H9/Vκ combinations using Vκ1–117, 3–4, 3–12, 13–85, 9–120 and 16–104 are non-reactive with dsDNA, CL, histones or chromatin whereas Vκ1–110, 4–57, 5–43, 5–45, 5–48, 10–94 and 12–46 react with one or more of these antigens ([21] and data not shown). Analysis of the follicular light chain repertoire revealed several differences between F to F 3H9, F 3H9, M 3H9 and M 3H9TLR7-/Yaa follicular B cells. However there was no clear effect of either Yaa status or Tlr7 dose on selection against known lupus related autoreactive specificities in the follicular repertoire (Fig. 6A, S2, S3 Tables).

The repertoire of light chains associated with 3H9+/IgG GC B cells was highly restricted in all four sets of chimeric mice, with overrepresentation of Vκ5–43/45 and Vκ5–48 as previously observed in the parental 3H9 W/B strain [21] (Fig. 6B, S2, S3 Tables). Among the Vκ5 encoded
light chains, Vκ5–43/45 was over represented in the F 3H9 to F chimeras compared with all
the other chimeras (p < 0.01 vs. the male chimeras and p<0.05 vs. F 3H9 chimeras). V3–5*01
and Vκ3–12*01 were also over represented in the F 3H9 to F chimeras. The 3H9+ plasma cell
repertoire was similar to that of the GC repertoire. M 3H9 GC B cells and plasma cells exhib-
itied the most light chain diversity, with a decrease in expression of Vκ5–43/45*01 light chains
and more frequent clonal expansions of other light chains (Fig. 6C, S2, S3 Tables).

We have previously shown that the autoreactive specificity of germline encoded 3H9/Vκ5
antibodies in non-chimeric 3H9 W/B males and females is influenced by the associated Jκ. In
these studies there was preferential selection of Vκ5–43/Jκ5 and Vκ5–48/Jκ4 in 3H9 W/B
males and of Vκ5–43/Jκ2, Vκ5–43/Jκ4 and Vκ5–48/Jκ5 in 3H9 W/B females [21]. Analysis of
the antigenic specificity of these H/L combinations showed that 3H9/Vκ5–48/Jκ4 had the
highest affinity for CL and dsDNA whereas 3H9/Vκ5–43/Jκ5 bound to histone and dsDNA
(Fig. 7A–C). Some of this reactivity was attenuated after DNase treatment and Protein A col-
umn chromatography but 3H9/Vκ5–43/Jκ5 and 3H9/Vκ5–48/Jκ4 still had higher affinity for
chromatin than the other Vκ5–43/Jκ5 and Vκ5–48/Jκ4 H/L combinations [21]. Surprisingly,
the 3H9/Vκ5–43/45 Jκ5 pair was most frequent in the F 3H9 to F chimeras and least frequent
in the 3H9 male chimeras (Fig. 7D). Vκ5–43 and Vκ5–45 have only 2 amino acid differences
(S to R in FR1 and N to Y in CDR1) and can only be distinguished by full length sequencing of
the light chain. 3H9/Vκ5–45/Jκ5 binds with higher affinity to dsDNA, CL and histones than
does 3H9/Vκ5–43/Jκ5 (Fig. 7A–C) and retains specificity for dsDNA and CL after DNase 1
treatment and purification [21]. We therefore performed full length sequencing of Vκ5–43/45
light chains from the different chimeras. We found that Vκ5–45 constituted 22% of the Vκ5–
43/45 light chains in the GCs of M 3H9 chimeras compared with 12–13% in F 3H9 and M
3H9TLR7-/Yaa chimeric mice and 17% in F 3H9 to F chimeric mice; these differences were not
significantly different. Similarly, Vκ5–45 was represented equivalently in the plasma cell reperto-
aire of all chimeras.

3H9/Vκ5–48/Jκ4, that encodes for anti-CL activity accounted for 25.3% of the Vκ5 encoded
light chain repertoire in the GCs of M 3H9 chimeras compared with < 5% in the other 3H9
chimeras (Fig. 7E, p < 0.01 compared with F 3H9 chimeras and p < 0.001 compared with M
3H9TLR7-/Yaa chimeras). These data in sum show selection against Vκ5–43/Jκ5 and selection
for Vκ5–48/Jκ4 in the 3H9 male chimeras, with the opposite phenotype in the 3H9 F to F chi-
meras and an intermediate phenotype in the other chimeras. In addition the 3H9-associated
light chain repertoire in the 3H9 male chimeras exhibits more diversity than that of the
other chimeras.

Analysis of somatic mutations

We have previously shown higher affinity CL and dsDNA binding activity of 3H9/Vκ5–43 and
3H9/Vκ5–48 pairs can be acquired as a result of somatic mutations [21]. Since Vκ5–43 and
Vκ5–48 were the dominant light chains used in association with class switched 3H9 B cells in
the GCs in all the chimeras we enumerated the number of mutations in the FR regions and in
CDRs of full length Vκ5–43 sequences from M 3H9, F 3H9 3H9TLR7-/Yaa and F 3H9 to F chi-
meras and in full length Vκ5–48 light chains from M 3H9 (18) and F 3H9 (19) chimeras. Light
chain Vκ5–43 sequences from the GC B cells of F 3H9 chimeras had significantly fewer muta-
tions than those from M 3H9 chimeras (average 0.9 vs. 3.0 mutations per sequence p< 0.05—
Table 2); this difference was not however observed in the Vκ5–48 sequences (data not shown)
or in Vκ5–43 encoded sequences from plasma cells (Table 2).

Surprisingly, the Vκ5–43 encoded light chains in the 3H9 F to F GC B cells and plasma cells
had significantly more mutations than those of the other chimeras ((p < 0.001—Table 2). The
light chain CDR1 makes an important contribution to antigen binding, with arginine, asparagine and lysine having DNA and chromatin binding capacity [38–40]. For Vκ5–43 the CDR1 consists of an asparagine rich sequence QNISNN. In F 3H9 to F chimeras one or both of the two terminal asparagines was replaced in 13/18 GC sequences and 11/20 plasma cell sequences, mostly by serine or isoleucine. By contrast, replacement of these asparagines occurred in only 1/29 GC sequences from F 3H9 (p = 0.0001), 0/28 GC sequences from M 3H9 (p < 0.0001) and 2/16 GC sequences from M 3H9TLR7-/Yaa mice (p < 0.001). The two terminal asparagines in CDR1 were also conserved in 10/11 Vκ5–43 encoded light chains from anti-DNA/anti-CL hybridomas derived from male 3H9 parental mice in which there was an average of 10 mutations per sequence (p = 0.002) [21]. The heavy chain CDR2 of 3H9 has a mutation from G to R and the CDR3 has two arginine residues that contribute to the autoreactive specificity of 3H9 encoded antibodies [38–40]. Of the 110 heavy chains recovered from wells expressing Vκ5–43/45 light chains, representing all the chimeras, the CDR2 arginine was conserved in 108 and the two CDR3 arginines were conserved in 108 with the other two manifesting a conservative R to K mutation.

**Discussion**

The purpose of our experiments was to use W/B mice bearing the site directed anti-CL/anti-DNA autoantibody VH transgene 3H9 to determine how Tlr7 expression and the Yaa locus influence the selection of naïve and antigen activated autoreactive B cells during the evolution of SLE. 60% of L chains that associate with 3H9 are permissive for autoreactivity to DNA or CL [39]. Tolerance in non-autoimmune 3H9 mice is maintained by receptor editing of the light and, to a lesser extent, the heavy chains to yield a less autoreactive naive repertoire [22, 39, 41–42], by follicular exclusion of autoreactive B cells [43], and by negative selection of autoreactive B cells in the GCs [44]. By contrast, 3H9 encoded autoantibodies using a variety of light chains are detected among autoreactive IgG hybridomas from 3H9 transgenic lupus-prone MRL/lpr mice [45]. TLR7 is required for the anti-RNA response and the absence of TLR signaling significantly attenuates the anti-CL response [46]. NZW/BXSBLTR7-/Yaa mice had significantly lower anti-CL titers and a decreased number of anti-CL AFCs compared with their WT counterparts. Thus Tlr7 dose significantly influences the anti-CL response in NZW/BXSB mice. Because 3H9 does not encode for anti-RNA antibodies we focused on the anti-CL response in these studies.
The role of B cell intrinsic TLR7 in regulating class switched autoantibody production and renal inflammation has been studied in mice with B cell specific genetic deficiency of Tlr7 or its adaptor MyD88. B cell depletion of MyD88 greatly reduces titers of IgG antinuclear antibodies and the GC B cell population fails to expand with age [47–48] resulting in a decrease in renal immune complex deposition and renal damage. Similarly, in mice with Tlr7 deficient B cells anti-RNA responses are abrogated and renal damage is decreased [14]. In Unc93b1 deficient MRL/lpr mice that have no signaling through any nucleic acid recognizing TLRs, all antinuclear antibodies are completely abrogated. When both WT and Unc93b1 deficient B cells are co-transferred into B6/lpr mice autoantibodies are preferentially made by the WT cells [46].

Finally, in Tlr7 deficient Sle1 mice, the presence of spontaneous germinal centers and autoantibody production are dependent on B cell intrinsic Tlr7 (15). These data all point to an intrinsic B cell function of TLR7 in promoting autoantibody production and subsequent tissue damage. Our analysis of the effect of the effect of excess Tlr7 dose on GC selection of H/L pairs with known antigenic specificity is consistent with this concept and yielded several important mechanistic findings.

An important design aspect of this study was to provide an environment in which immature, 3H9 positive B cells harboring Yaa or expressing one or two copies of Tlr7 were subject to competition with wt non-3H9 cells for survival; this allows deletional tolerance mechanisms to be operative. In addition the transgenic B cells were exposed to an inflammatory environment as disease evolved. The GFP marker allowed us to sort single transgenic cells from each B cell subset even though they represented only a small percentage of the B cell repertoire.

We first showed that the major site for deletion of 3H9 B cells is central with little further deletion occurring in the periphery. This is in contrast to the autoreactive anti-dsDNA D42 transgenic system in which deletion of autoreactive specificities occurs predominantly at the transitional stage in the periphery [31]. Autoreactivity of the remaining 3H9 B cells appeared to persist in the periphery as manifested by downregulation of surface IgM on the mature naïve transgenic cells. Downregulation of surface BCR in non-anergic autoreactive B cells has been postulated to be a mechanism for preserving diversity of the B cell repertoire and perhaps allowing subsequent mutation and selection away from autoreactivity during antigen specific responses [37, 49].

Examination of the light chains associated with 3H9 in follicular B cells revealed several differences between the various chimeras but it was not possible to distinguish a clear pattern of selection against autoreactivity. Importantly however, the most commonly used autoreactive heavy/light chain pairs in the follicular repertoire 3H9/10–94°01 and 3H9/12–46°01/Jκ2 were excluded from the GC and plasma cell repertoires. Whether these autoreactive B cells are segregated into the follicular subset with the lowest sIgM expression will need further study.

We next demonstrated that male W/B 3H9 B cells bearing 2 copies of Tlr7 are preferentially expanded in the GC compared to W/B female or male W/B TLR7/Yaa 3H9 B cells with only 1 copy of Tlr7. To further dissect the effects of TLR7 and Yaa on GC selection of 3H9+ B cells we examined the repertoire of light chains associated with the 3H9 heavy chain in class switched GC B cells and plasma cells. We have previously shown by single cell PCR analysis and analysis of hybridomas that Vκ5–43 and Vκ5–48 light chains are overrepresented in the GC repertoire of both male and female 3H9 W/B mice but that the preferred VJ combinations differ between males and females. Higher affinity autoreactive B cells expressing Vκ 5–43/Jκ5 and Vκ 5–48/Jκ4 are preferentially selected into and expanded in the GCs of male 3H9 W/B mice compared with females. This suggests that selection of autoreactive B cells into the GC may be partially dependent on TLR7 [21]. We show here that when interclonal competition for GC entry is provided, all the chimeras select the Vκ5–43/Jκ5 encoded light chain that confers anti-chromatin activity; this specificity has been shown not to be influenced by Tlr7 dose [5]. However the
repertoire of M 3H9 chimeric mice diversifies away from Vk5–43/Jκ5 and demonstrates preferential use of Vk5–48/Jκ4 that confers anti-CL reactivity. Overall, these data support the notion that TLR7 over-expression influences the stringency of GC selection and facilitates clonal expansion of autoreactive B cells in the GC.

Somatic mutations are linked to proliferation and this occurs in the germinal center dark zone [13] that is smaller when TLR7 is absent [50]. We therefore examined the effect of TLR7 overexpression on the frequency of somatic mutation in GC B cells. We were unable to demonstrate a consistent effect of either Yaa or Tlr7 dose on the frequency of somatic mutations in Vk5 encoded light chains. It was intriguing however, that in GC B cells the rate of somatic mutations was low in 3H9 associated Vk5–43 encoded light chains from F 3H9 chimeras in which non-Yaa B cells compete with male Yaa B cells for survival and GC entry. Surprisingly, there was a high frequency of somatic mutations in the analogous GC and plasma cell populations from 3H9 F to F chimeras in which there was no competition between Yaa and non-Yaa B cells. In the latter mice, B cells encoding the autoreactive 3H9/Vk5–43/Jκ5 pair could presumably compete equally for T cell help, whereas in the former we postulate that the same cells may not compete as well for the signals that facilitate dark zone entry. Nevertheless, the total number of spleen cells, GCs and plasma cells was substantially lower in the 3H9 F to F chimeras than in the other chimeras. This most likely reflects additional effects of Tlr7 dose either on the antigen presenting ability of B cells or on the myeloid cells that facilitate T cell expansion and inflammation [47–48].

Previous studies have shown that downregulation of the BCR in autoreactive B cells does not prevent GC B cell expansion of these cells but may influence maturation to plasma cells [36]. In addition TLR7 has been shown to influence the survival of plasma cells in an anti-viral response [50]. We found a modest decrease in the ratio of GFP+ plasma cells compared with GFP+ B220+ cells in the F 3H9 and M 3H9TLR7–/Yaa compared with the M 3H9 chimeras. In addition, there were differences in the spleen plasma cell repertoire between the chimeras similar to those of the GC repertoire, with a highly restricted light chain repertoire in the F 3H9 to F controls and the most light chain diversity in the 3H9 M chimeras.

The presence of at least one Tlr7 allele has previously been shown to be necessary for an optimal GC response to viral infection and for spontaneous GC activation in lupus prone mice. Our findings in sum show that in an inflammatory microenvironment TLR7 overexpression contributes significantly to the loss of B cell tolerance in the GCs of Yaa bearing lupus prone mice where it influences selection, proliferation and diversification of the repertoire of autoreactive B cells in the GCs.

Supporting Information

S1 Fig. Percent (A) and number (B) of spleen B cell subsets in the chimeras. The 3H9 F to F chimeras have a higher percentage of marginal zone (MZ) B cells and a lower total number of follicular (FO), class switched (CS), germinal center (GC) and plasma cells (PC) than all the other chimeras. Comparisons are with male WT chimeras. n = 5–12 per group. * p < 0.05, † p < 0.01.

S1 Table. Experimental mouse strains. Generation of mouse strains used for analysis of the TLR7–/Yaa phenotype and as donors for the bone marrow chimeras.

S2 Table. Pairwise comparisons of Vk repertoires of FO, GC and PC subsets. The Table shows the number of Vk genes represented in each comparison, the number of genes
that contribute the top 50% of the statistical difference in the $\lambda^2$ analysis and the percent contribution of the most differentially expressed gene. Individual genes that contribute $>5\%$ of the statistical difference in the $\lambda^2$ analysis are shown on the right for each comparison. See methods section for a description of the statistical analysis.

(DOCX)

**S3 Table.** 3H9 associated Vκ usage of B cell subsets from chimeric mice. Vκ usage of each 3H9-expressing single cells sorted from each of the follicular, germinal center and plasma cell subsets of bone marrow chimeric mice. The number of cells expressing each Vκ is shown for each set of chimeras. See Fig. 6 for graphical representation of selected genes.

(DOCX)

**Author Contributions**

Conceived and designed the experiments: AB WH RB RS MW SA AD. Performed the experiments: AB WH RB RS MW SA. Analyzed the data: AB WH MW MA ML AD. Wrote the paper: AB MA ML AD.

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