Role and regulation of autophagy in heat stress responses of tomato plants

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INTRODUCTION

Autophagy is a highly conserved intracellular degradation system in eukaryotes for removal and recycling of cytoplasmic components including damaged proteins and organelles (Klionsky, 2005). Central to autophagy is the formation of autophagosomes resulting from the dynamic membrane reorganization. In yeast, more than 30 autophagy-related (ATG) genes have been identified and their products often form functional groups that cooperate to perform the physiologically continuous but mechanistically distinct processes of autophagy including the induction of autophagy, autophagosome nucleation, elongation, maturation, and fusion with vacuoles (He and Klionsky, 2009). Autophagy is active at very low levels but is highly inducible in responses to stress and extracellular cues (He and Klionsky, 2009). In yeast and animal systems, the serine/threonine protein kinase TOR (target of rapamycin) functions as a central inhibitor of autophagosome formation. In yeast, inhibition of TOR leads to activation of ATG1, which can then bind ATG13 and ATG17 with increased affinities to promote assembly of the ATG1-ATG13-ATG17 scaffold and initiation of autophagosome formation through recruitment of multiple ATG proteins (He and Klionsky, 2009). The rapid increase in the autophagic flux during the early minutes or hours of exposure to stress conditions is mostly mediated by post-translational modifications of the core machinery of autophagy (He and Klionsky, 2009). A delayed and protracted autophagic response, however, also relies on activation of specific transcription programs involving stress-responsive transcription factors.

As sessile organisms, plants are constantly exposed to a wide spectrum of stress conditions such as high temperature, which causes protein misfolding. Misfolded proteins are highly toxic and must be efficiently removed to reduce cellular proteotoxic stress if restoration of native conformations is unsuccessful. Although selective autophagy is known to function in protein quality control by targeting degradation of misfolded and potentially toxic proteins, its role and regulation in heat stress responses have not been analyzed in crop plants. In the present study, we found that heat stress induced expression of autophagy-related (ATG) genes and accumulation of autophagosomes in tomato plants. Virus-induced gene silencing (VIGS) of tomato ATG5 and ATG7 genes resulted in increased sensitivity of tomato plants to heat stress based on both increased development of heat stress symptoms and compromised photosynthetic parameters of heat-stressed leaf tissues. Silencing of tomato homologs for the selective autophagy receptor NBR1, which targets ubiquitinated protein aggregates, also compromised tomato heat tolerance. To better understand the regulation of heat-induced autophagy, we found that silencing of tomato ATG5, ATG7, or NBR1 compromised heat-induced expression of not only the targeted genes but also other autophagy-related genes. Furthermore, we identified two tomato genes encoding proteins highly homologous to Arabidopsis WRKY33 transcription factor, which has been previously shown to interact physically with an autophagy protein. Silencing of tomato WRKY33 genes compromised tomato heat tolerance and reduced heat-induced ATG gene expression and autophagosome accumulation. Based on these results, we propose that heat-induced autophagy in tomato is subject to cooperative regulation by both WRKY33 and ATG proteins and plays a critical role in tomato heat tolerance, mostly likely through selective removal of heat-induced protein aggregates.

Keywords: autophagy, heat tolerance, tomato, WRKY33, NBR1, ATG5, ATG7

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As in other organisms, formation of autophagosomes and expression of \(\text{ATG}\) genes are induced by a variety of stresses and environmental cues in plants. TOR is also a negative regulator of autophagy in plants (Liu and Bassham, 2010). Furthermore, a NADPH oxidase inhibitor blocks autophagy induction upon nutrient starvation and salt stress, but not during osmotic stress (Liu and Bassham, 2010). Thus, ROS may mediate induction of autophagy during some, but not all stress conditions. There is, however, little information available about the transcriptional regulation of plant autophagy-associated genes under stress conditions.

In the present study, we analyze the role and regulation of autophagy in heat stress tolerance of tomato plants (\textit{Solanum lycopersicum}). Heat is an important abiotic stress condition that can cause misfolding and denaturation of proteins. As an important protein quality control mechanism, autophagy could play a critical role in removal of those misfolded/denatured and potentially highly toxic proteins or protein aggregates that fail to be reestablished for normal protein conformations (Kraft et al., 2010; Johansen and Lamark, 2011; Shaid et al., 2013). Tomato is one of the most important vegetable plants closely related to many commercially important plants including potato, eggplant, peppers, tobacco, and petunias and an important model plant because it has a number of interesting features such as fleshy fruits not shared by other model plants (e.g., Arabidopsis and rice) and has many recognized wild species. The tomato genome has been sequenced (Consortium, 2012) and its genes can be functionally analyzed through a number of complementary approaches including virus-induced gene silencing (VIGS) (Liu et al., 2002). From the sequenced tomato genome, we have identified tomato homologs for \text{ATG}5 and \text{ATG}7, two important autophagy-related proteins that have been subjected to functional analysis in Arabidopsis (Yoshimoto, 2010; Lai et al., 2011b; Zhou et al., 2013). We have also identified tomato homologs for \text{NBR}1, a plant selective autophagy receptor (Svenning et al., 2011; Zhou et al., 2013, 2014c), and \text{WRKY}33, a transcription factor that physically interacts with \text{ATG}18a in Arabidopsis (Lai et al., 2011b). Using a variety of molecular approaches including VIGS, we analyzed the roles of these genes in heat-induced autophagy and heat stress tolerance. These experiments not only support the critical role of autophagy in plant heat tolerance but also provide new important insights into the regulation of autophagy during plant stress responses.

**MATERIALS AND METHODS**

**PLANT MATERIALS AND GROWTH CONDITIONS**

Tomato (\textit{Solanum lycopersicum} L. cv. Ailsa Craig) seeds were germinated in a growth medium filled with a mixture of peat and vermiculite (7:3, v/v) in trays in a growth chamber. When the first true leaf was fully expanded, seedlings were transplanted into plastic pots containing the same medium. The growth conditions were as follows: light/dark cycle, 22/20°C, and photosynthetic photon flux density (PPFD), 600 \(\mu\)mol m\(^{-2}\) s\(^{-1}\).

**QUANTITATIVE RT-PCR (qRT-PCR)**

Total RNA was isolated from tomato leaves using Trizol reagent (Sangon Co., Shanghai, China), according to the manufacture’s recommendations. Genomic DNA was removed with the RNaseasy Mini Kit (Qiagen Co., Hilden, Germany). 1 \(\mu\)g RNA was reverse-transcribed using the ReverTra Ace qPCR RT Kit (Toyobo Co., Osaka, Japan), following the manufacturer’s instructions. Gene-specific RT-PCR primers were designed based on their cDNA sequences (Supplemental Table S1).

The quantitative real-time PCR was performed using the iCycler QTm real-time PCR detection system (Bio-Rad Co., Hercules, CA, USA). Each reaction (25 \(\mu\)L) consisted 12.5 \(\mu\)L of SYBR Green PCR Master Mix (Takara Co., Chiga, Japan), 1 \(\mu\)L of diluted cDNA and 0.1 \(\mu\)mol forward and reserve primers. The PCR cycling conditions and the calculation of relative gene expression were as previously described. The tomato \text{ACTIN} gene was used as internal control as previously described (Zhou et al., 2014a).

**VIRUS-INDUCED GENE SILENCING (VIGS)**

The tobacco rattle virus (TRV) VIGS constructs for silencing of tomato \text{ATG}5, \text{ATG}7, \text{NBR}1a, \text{NBR}1b, \text{WRKY}33a, and \text{WRKY}33b genes were generated by PCR amplification using gene-specific primers (Supplemental Table S2), digested with appropriate restriction enzymes and ligated into the same sites of pTRV2. The resulting plasmids were transformed into \textit{Agrobacterium tumefaciens} GV3101. \textit{Agrobacterium}-mediated virus infection was performed as previously described (Ekengren et al., 2003). Plants were then kept at 22/20°C under 150 \(\mu\)mol m\(^{-2}\) s\(^{-1}\) PPFD for 30 days before they were used for the experiments (Kandoth et al., 2007). Leaflets in the terminal of the fifth fully expanded leaves, which showed 20–30% transcript levels of control plants, were used. Each replicate had 12 plants.

**ASSESSMENT OF HEAT TOLERANCE**

To evaluate the role of autophagy in heat stress responses of tomato plants, tomato plants at the five-leaf stage were transferred to a growth chamber for heat stress treatment (45°C, 400 \(\mu\)mol m\(^{-2}\) s\(^{-1}\) PPFD, 8 h).

For determination of electrolyte leakage (EL) caused by high temperature, the leaflets in the terminal of the fifth fully expanded leaves were measured after heat stress as previous described (Huang et al., 2010). Chlorophyll fluorescence was measured using an Imaging-PAM Chlorophyll Fluorometer equipped with a computer-operated PAM-control unit (IMAG-MAXI; Heinz Walz, Effeltrich, Germany). The plants were maintained in the dark for more than 30 min before the measurements were performed. The intensities of the actinic light and saturating light were 280 and 2500 \(\mu\)mol mol\(^{-1}\) s\(^{-1}\) PPFD, respectively. The maximum quantum yield of PSII (Fv/Fm) was measured and calculated as previous described (Zhou et al., 2014a). Three replicates for each treatment were used with 12 plants for each replicate.

The light-saturated CO\(_2\) assimilation (\text{Asat}), stomatal conductance (\text{Gs}), and intracellular CO\(_2\) concentration (\text{Ci}) were determined in the silenced and pTRV plants with an infrared gas analyzer-based potable photosynthesis system (LI-6400; Li-COR, Lincoln, NE, USA). The air temperature, relative humidity, CO\(_2\) concentration, and PPFD were maintained at 22°C, 85%, 380 \(\mu\)mol mol\(^{-1}\), and 1000 \(\mu\)mol m\(^{-2}\) s\(^{-1}\), respectively.
**SEPARATION AND MEASUREMENTS OF TOTAL AND INSOLUBLE PROTEINS**

Tomato leaves were collected before and after heat treatment, ground in liquid nitrogen and homogenized in a detergent containing extraction buffer (100 mMTris/HCl, pH 8.0, 10 mM NaCl, 1 mM EDTA, 1% Triton X-100, 0.2% β-mercaptoethanol). Soluble and detergent-resistant insoluble proteins were separated through low-speed centrifugation and measured as previously described (Zhou et al., 2013).

**VISUALIZATION OF INDUCTION OF AUTOPHAGY**

For visualization of autophagosomes, tomato leaves were vacuum-infiltrated with 1 μM of the fluorescence dye LysoTracker Green DND-26 (Cell Signaling Technology, Danvers, MA, USA) or 500 μM of monodansylcadaverine (MDC) (Sigma-Aldrich, St. Louis, MO, USA). Fluorescence was visualized using a Zeiss LSM510 UVMeta laser scanning confocal microscope (Zeiss Co., Munchen, Germany) and images were superimposed using Zeiss LSM510 software.

**RESULTS**

**IDENTIFICATION OF TOMATO ATG5, ATG7, AND NBR1 GENES**

To analyze the role of autophagy in tomato heat tolerance, we chose first to focus on tomato ATG5 and ATG7 as potential targets for gene silencing as their products are required for the core process of autophagy and mutants of their Arabidopsis homologs, which are single-copy genes, have been widely used for functional analysis of autophagy (Yoshimoto, 2010; Lai et al., 2011b; Zhou et al., 2013). From the sequenced tomato genome, we identified two tomato ATG5 genes, ATG5a (Sl02g038380) and ATG5b (Sl06g041400). Based on its genomic and full-length cDNA sequences, ATG5a has an intron-exon structure similar to that of Arabidopsis ATG5 with eight exons and seven introns (Supplemental Figure 1A). ATG5a encodes a protein containing 369 amino acid residues and sharing approximately 60% sequence identity with Arabidopsis ATG5. The genomic sequence of tomato ATG5b is 97% identical to the coding sequence of tomato ATG5a (Supplemental Figure 1B). The lack of introns in tomato ATG5b indicates that the gene was copied from mRNA of tomato ATG5a and incorporated into the tomato genome. Further sequence analysis revealed multiple mutations in tomato ATG5b, including a G to A transition at nucleotide position 246 that changes a highly conserved ubiquitin-associated (UBA) domain and a WxI ATG8-interacting motif at their respective C-terminus (Supplemental Figure 3).

**HEAT INDUCTION OF AUTOPHAGY-RELATED GENES IN TOMATO**

To determine the role of autophagy in tomato heat tolerance, we firstly analyzed the effect of heat stress on expression of tomato ATG5, ATG7, and NBR1 genes. Tomato seedlings were placed in the 22 and 45°C chambers, and the ATG and NBR1 gene transcripts were analyzed by qRT-PCR using total RNA isolated from the leaflets of the fifth fully expanded leaves. As shown in Figure 1, the levels of tomato ATG5, ATG7 and NBR1 transcripts remained unchanged at 22°C throughout the 8-h period of the experiments. In contrast, transcript levels of the autophagy genes were elevated after 2–4 h of heat stress (45°C) and displayed large increases after 6 h of heat stress (Figure 1). After 6-h heat stress, the transcript levels of the ATG and NBR1 genes started to decline and reduced to basal levels by 8 h at 45°C, when the leaves of seedlings started to show symptoms of dehydration (Figure 1). Thus, heat stress induced the expression of autophagy-related genes.

**SILENCING OF TOMATO ATG5, ATG7, AND NBR1 GENES**

To determine directly the role of autophagy in tomato responses to heat stress, we used VIGS to assess the impact of downregulated expression of tomato ATG5, ATG7, NBR1a, and NBR1b on tomato heat tolerance. Gene-specific DNA fragments were cloned into the pTRV vector and Agrobacterium cells harboring the VIGS vectors were infiltrated into tomato cotyledons. We used qRT-PCR to compare the transcript levels for tomato ATG5, ATG7, NBR1a, and NBR1b in tomato plants infiltrated with the pTRV empty vector or infiltrated with the pTRV-ATG5, pTRV-ATG7, pTRV-NBR1a, or pTRV-NBR1b silencing vector. As shown in Figure 2, the basal transcript levels of ATG5, ATG7, NBR1a, and NBR1b were unchanged in tomato plants after infiltration with the pTRV empty vector but decreased 70–80% in the leaves of plants after infiltration with their respective silencing vectors (Figure 2). No significant alteration in growth or development was observed upon silencing of the ATG5, ATG7, NBR1a, or NBR1b gene. Abiotic stress including high temperature induces both ATG gene expression and formation of autophagosomes. To further assess the effect of silencing of tomato ATG5 and ATG7 on heat-induced autophagy, we used LysoTracker Green dye as a probe to detect autolysosome-like structures. The LysoTracker dye has been widely used as a probe for detecting autophagic activity in a variety of organisms including plants (Otegui et al., 2013).
FIGURE 1 | Induction of tomato ATG5, ATG7, NBR1, and WRKY33 genes by heat stress. Six weeks-old tomato plants were placed in the 22 and 45°C growth chambers and total RNA was isolated from leaf samples collected at indicated times. Transcript levels were determined using real-time qRT-PCR. Error bars indicate SE (n = 3).

We also used MDC as a probe for detection of autophagic activity in tomato leaves. MDC is an autofluorescent dye that stains autophagosomes in mammals and plants (Biederbick et al., 1995; Munafo and Colombo, 2001; Contento et al., 2005; Liu and Bassham, 2010). Under the normal temperature (22°C), again, we observed low numbers of punctate fluorescent signals in both control (pTRV) and ATG5- or ATG7-silenced plants (Figure 4). After treatment with dithiothreitol (DTT), a known inducer of autophagy (Liu et al., 2012), the numbers of punctate fluorescent signals increased by more than 8 fold in control plants infiltrated with the pTRV empty vector (Figure 4). After 6-h heat treatment, the numbers of punctate fluorescent signals also increased by about 6 fold in control plants infiltrated with the pTRV empty vector (Figure 4). In the plants infiltrated with the pTRV-ATG5 or pTRV-ATG7
silencing vector, the numbers of the punctate fluorescence signals after DTT or heat stress were substantially reduced when compared to those in DTT- or heat-treated control plants (Figure 4). These observations confirmed that heat-induced autophagy was partially blocked by silencing of the tomato ATG5 and ATG7 genes.

**COMPROMISED HEAT TOLERANCE OF AUTOPHAGY-SILENCED TOMATO PLANTS**

For comparison of heat tolerance of pTRV, pTRV-ATG5, pTRV-ATG7, pTRV-NBR1a, and pTRV-NBR1b plants, they were placed in a 45°C growth chamber for 8 h and then moved to room temperature for 3-day recovery. For heat–treated pTRV control plants, only patches of old leaves displayed symptoms of dehydration while a majority of the leaves remained green and viable after recovery (Figure 5A). On the other hand, a majority of fully expanded leaves from the pTRV-ATG5, pTRV-ATG7, pTRV-NBR1a, and pTRV-NBR1b plants exhibited wilting after the recovery (Figure 5A). The more severe symptoms in autophagy-suppressed tomato plants after heat stress were confirmed by increased EL in the silenced plants relative to that in the pTRV control plants (Figure 5B).

Heat has a harmful effect on various biology processes including photosynthesis. To further investigate responses of ATG5-, ATG7-, and NBR1-silenced tomato plants to heat stress, we compared these tomato plants with the pTRV control tomato plants for the effects of heat stress on the maximum quantum yield of photosystem II (PSII) (Fv/Fm) of leaves immediately after heat treatment and light-saturated CO2 assimilation rate (Asat) following 1-day recovery after heat stress. As shown in Figure 6, after 8 h at 45°C, Fv/Fm values for silencing plants were 22–37% lower than those of control pTRV plants. Likewise, the Asat values for the silenced plants were 35–68% lower than those of

![Figure 2](image2.png)

**FIGURE 2 | TRV-mediated silencing of tomato ATG5, ATG7, NBR1, and WRKY33 genes.** Transcript levels for each silenced gene in tomato plants infiltrated with Agrobacterium cells harboring the empty pTVR vector or the corresponding silencing vector were determined by qRT-PCR analysis using total RNA isolated from the terminal leaflets of the fifth leaves of Agrobacterium-infiltrated tomato plants. Error bars indicate SE (n = 3). According to Duncan’s multiple range test (P = 0.05), means of lesion areas do not differ significantly if they are indicated with the same letter.

![Figure 3](image3.png)

**FIGURE 3 | Detection of autophagic activity using LysoTracker Green.** Comparison of tomato plants infiltrated with Agrobacterium cells harboring the empty pTVR vector or a silencing vector for an indicated gene in terms of LysoTracker Green fluorescence signals. The plants were placed into the 22 and 45°C growth chambers and analyzed after 6-h treatment. Numbers of punctate LysoTracker Green fluorescence spots per 10,000 μm² section from the cells in the central areas of the terminal leaflets of the fifth leaves were indicated below the images. Means and SE were calculated from three experiments. According to Duncan’s multiple range test (P = 0.05), means of lesion areas do not differ significantly if they are indicated with the same letter.
pTRV control plants when assayed after 1-day recovery following heat stress (Figure 7). In addition, ATG5-, ATG7-, and NBR1-silenced tomato plants had reduced stomatal conductance (Gs) and intracellular CO2 concentration (Ci) compared to the unsilenced pTRV control plants after heat stress (Figure 7). Thus, photosynthetic efficiency and capacity were more compromised by heat stress in the autophagy-suppressed tomato plants than in the control plants.

INCREASED ACCUMULATION OF INSOLUBLE PROTEINS IN AUTOPHAGY-SILENCED PLANTS

Heat stress causes protein misfolding and denaturation, which can result in formation of protein aggregates and proteotoxic
FIGURE 6 | The effect of heat stress on the efficiency of PSII photochemistry. Fv/Fm images (A) and values (B) of the terminal leaflets of the fifth leaves were determined immediately after 8 h at 22 or 45°C heat treatment. The color code in the images ranged from 0 (black) to 1.0 (purple). Means and SE were calculated from average Fv/Fm values determined from three experiments with 10 leaves per experiment for each type of plants. According to Duncan's multiple range test ($P = 0.05$), means of lesion areas do not differ significantly if they are indicated with the same letter.

stress. To analyze the role of autophagy in protection against heat-induced proteotoxic stress, we investigated the accumulation of insoluble, detergent-resistant proteins in the pTRV, pTRV-ATG5, pTRV-ATG7, pTRV-NBR1a, and pTRV-NBR1b plants after 8 h heat treatment. Total proteins were first isolated and insoluble proteins were separated by low speed centrifugation. As shown in Figure 8, the percentages of insoluble to total proteins were similar in all plants when they were grown at 22°C. After 8-h heat stress, insoluble proteins as percentages to total proteins increased only by 40% in unsilenced pTRV control plants but increased by 138–154% in the pTRV-ATG5, pTRV-ATG7, pTRV-NBR1a, and pTRV-NBR1b plants (Figure 8). By the end the heat stress, the levels of insoluble proteins in the ATG5-, ATG7-, and NBR1-silenced tomato plants were more than two times higher than those in the unsilenced control plants (Figure 8).

FIGURE 7 | The effect of heat stress on the capacity of photosynthesis. Light-saturated CO2 assimilation rate (Asat) (A), stomatal conductance ($G_s$) (B) and intracellular CO2 concentration ($C_i$) (C) were determined following 1-day recovery after heat stress. Means and SE were calculated from average values determined from three experiments with 10 leaves per experiment for each type of plants. According to Duncan's multiple range test ($P = 0.05$), means of lesion areas do not differ significantly if they are indicated with the same letter.

IDENTIFICATION AND FUNCTIONAL ANALYSIS OF TOMATO WRKY33 IN HEAT TOLERANCE

Arabidopsis WRKY33 is a transcription factor important for plant resistance to necrotrophic fungal pathogens and for plant
heat tolerance (Zheng et al., 2006; Li et al., 2011). Arabidopsis WRKY33 interacts with Arabidopsis ATG18a, a critical component of autophagy, and plays a positive role in pathogen-induced ATG18a expression and autophagosome formation (Lai et al., 2011a). These results suggest that the critical role of WRKY33 in plant responses to biotic and abiotic stresses may be at least in part mediated through its positive regulation of pathogen/stress-induced autophagy. To investigate whether tomato contains similar WRKY transcription factor(s) with a critical role in heat tolerance and regulation of stress-induced autophagy, we searched the sequenced tomato genomes and identified two close WRKY33 homologs, WRKY33a (Sl09g014990) and WRKY33b (Sl06g066370). As shown in Supplemental Figure 4, Arabidopsis WRKY33, tomato WRKY33a and WRKY33b all belong to Group I WRKY transcription factors containing two WRKY domains with highly conserved amino acid sequences. High sequence similarities are also found in the N-terminal domains including the highly conserved SP clusters as putative MAPK phosphorylation sites and the intervening sequences between the two WRKY domains (Supplemental Figure 4). Furthermore, both tomato WRKY33a and WRKY33b contain a segment of about 100 amino acid residues on the C-terminal side of the second WRKY domain with substantial sequence homology with Arabidopsis WRKY33, which are absent in other related Group I WRKY transcription factors such as Arabidopsis WRKY25 and WRKY26 (Supplemental Figure 4) (Lai et al., 2011a).

To analyze heat-induced expression of tomato WRKY33 genes, we analyzed their transcripts in the tomato seedlings grown at 22 or 45°C. As shown in Figure 1, the transcript levels of both tomato WRKY33a and WRKY33b remained low throughout the 8-h period of the experiments at 22°C. At 45°C, however, the transcript levels of tomato WRKY33a and WRKY33b were elevated with similar kinetics (Figure 1). Transcripts levels for both genes displayed substantial increases after 4-h exposure to 45°C and peaked after 6-h heat stress (Figure 1). Like those of other analyzed autophagy-related genes, the transcript levels for both WRKY33a and WRKY33b declined after 6-h heat exposure and approached those of control plants by 8-h heat exposure (Figure 1).

To investigate directly the roles of tomato WRKY33 genes, we used VIGS technology to assess the impact of their downregulated expression on tomato heat tolerance. Tomato WRKY33-specific DNA fragments were cloned into the pTRV vector and Agrobacterium cells harboring the VIGS vectors were infiltrated into tomato leaves. As shown in Figure 2, basal expression of WRKY33a or WRKY33b was observed in the tomato plants infiltrated with the pTRV empty vector. By contrast, infiltration with either pTRV-SlWRKY33a or pTRV-SlWRKY33b silencing vector resulted in approximately 5–7-fold reduction in the transcript levels for both tomato WRKY33a and WRKY33b (Figure 2). The cross silencing likely resulted from the high sequence homology between the two genes, which share more than 75% nucleotide sequence identity. The tomato plants silenced for WRKY33a and WRKY33b were normal in growth and development and displayed no detectable morphological phenotype.

We analyzed the impact of silencing of WRKY33 genes on tomato heat tolerance. Both control and silenced plants were placed in a 45°C growth chamber for 8 h and then moved to room temperature for 3-day recovery. Unlike heat–treated pTRV control plants, which had only some patches of old leaves that displayed symptoms of dehydration, a majority of leaves from the pTRV-WRKY33a and pTRV-WRKY33b plants exhibited extensive wilting or even bleaching after the recovery (Figure 5). Thus, silencing tomato WRKY33 genes caused increased sensitivity to heat stress. Assays of EL, the maximum quantum yield of PSII (Fv/Fm), light-saturated CO2 assimilation rate (Asat), stomatal conductance (Gs), and intracellular CO2 concentration (Ci) confirmed that silencing of tomato WRKY33a and WRKY33b compromised tomato heat tolerance (Figures 5–7). Furthermore, silencing of tomato WRKY33a and WRKY33b led to increased accumulation of insoluble proteins under heat stress (Figure 8).

REGULATION OF HEAT-INDUCED AUTOPHAGY BY WRKY33 AND AUTOPHAGY PROTEINS

To investigate whether the critical role of tomato WRKY33 genes in heat tolerance is associated with their positive roles in regulation of heat-induced autophagy, we analyzed whether silencing the WRKY33 genes in tomato compromised heat-induced autophagosome formation. As shown in Figure 3, while there was a more than 10 fold increase in the numbers of punctate green fluorescent signals after 6-h heat stress in the pTRV control plants, there was only 3–5-fold increase in the pTRV-WRKY33a and pTRV-WRKY33b plants. As a result of combined reduction of both basal and induced autophagosome formation, the levels of autophagosomes in the WRKY33-silenced plants were only about

FIGURE 8 | Accumulation of insoluble protein aggregates under heat stress. Leaf tissues from tomato plants infiltrated with Agrobacterium cells harboring the empty pTRV vector or the silencing pTRV-ATG5, pTRV-ATG7, pTRV-NBR1, or pTRV-WRKY33 vector collected at indicated hours (h) under 45°C for preparation of total, soluble and insoluble proteins as described in Materials and Methods. Total proteins in the starting homogenates and insoluble proteins in the last pellets were determined the percentages of insoluble proteins to total proteins were calculated.
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25–30% of those in the control plants after 6-h heat stress. Thus, tomato WRKY33 proteins play a positive role in heat-induced autophagosome formation.

We also analyzed the mutual regulation among the silencing tomato ATG5, ATG7, NBR1, and WRKY33 genes during plant responses to heat stress. For this purpose, we analyzed the transcript levels of ATG5, ATG7, NBR1a, NBR1b, WRKY33a, and WRKY33b in the silencing plants after 6-h heat stress. In pTRV control plants, as expected, the transcript levels for ATG5, ATG7, NBR1a, NBR1b, WRKY33a, and WRKY33b were elevated under heat stress. However, induction of these genes was all reduced not only in the plants harboring their respective silencing vectors but also in the plants harboring silencing vectors for the other genes (Figure 9). Induction of three heat shock proteins (HSP17.6, HSP20, and HSP100) in ATG5-, ATG7-, NBR1-, or WRKY33-silencing plants or pTRV plants was almost as strong as that in the pTRV control plants after 6-h heat stress (Figure 10). However, induction of a tomato HSP40 gene was significantly compromised by silencing of the autophagy-related or WRKY33 genes (Figure 10). These results indicated that ATG and WRKY33 proteins have a positive role in heat-induced expression of autophagy-related genes.

DISCUSSION
In the present study, we analyzed the role of autophagy in responses to heat stress in tomato, an important horticultural crop. Both the expression of ATG5 and ATG7 genes and formation of autophagosomes were induced in heat-stressed tomato plants leaf samples collected after 6-h heat stress for determination of transcript levels of indicated genes by qRT-PCR. Error bars indicate SE (n = 3). According to Duncan’s multiple range test (P = 0.05), means of lesion areas do not differ significantly if they are indicated with the same letter.
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FIGURE 10 | Regulation of heat-induced expression of tomato HSP genes. Tomato plants infiltrated with Agrobacterium cells harboring the empty pTVR vector or the silencing pTRV-ATG5, pTRV-ATG7, pTRV-NBR1, or pTRV-WRKY33 vector were placed in a 45°C growth chamber and total RNA was isolated from leaf samples collected after 6-h heat stress for determination of transcript levels of indicated genes by qRT-PCR. Error bars indicate SE (n = 3). According to Duncan’s multiple range test (P = 0.05), means of lesion areas do not differ significantly if they are indicated with the same letter.

(Figures 1, 3, 4). The heat tolerance of autophagy-suppressed tomato plants due to silencing of ATG5 and ATG7 genes was compromised based on their increased morphological symptoms associated with enhanced defects in the efficiency and capacity of photosynthesis after heat stress (Figures 5–7). These results indicate that autophagy plays an important role in tomato heat tolerance.

Heat stress causes protein misfolding and denaturation. Misfolded/denatured proteins are highly toxic due to nonspecific binding to a variety of cellular constituent and, therefore, must be efficiently removed to prevent proteotoxic stresses (Hightower, 1991). Extensive studies in yeast and animal organisms have revealed that misfolded proteins are recognized by the protein quality control system, ubiquitinated by chaperone-dependent E3 ubiquitin ligases such as the C-terminus of Hsc70-interacting protein (CHIP) and subjected to degradation by the ubiquitin proteasome system (UPS) (Kraft et al., 2010; Shaid et al., 2013). Very recently we have conducted comprehensive genetic analysis of Arabidopsis CHIP E3 ubiquitin ligase and discovered its critical role in plant responses to a spectrum of abiotic stresses including heat stress (Zhou et al., 2014c). Previously, it has also been reported that Arabidopsis CHIP E3 ubiquitin ligase and Hsc70-4 mediate plastid-destination precursor degradation through UPS when the import of the precursors are blocked in a plastid-import mutant (Lee et al., 2009). In the present study, we demonstrated that silencing of tomato genes encoding NBR1a and NBR1b, two close homologs of mammalian ubiquitin-binding autophagy receptors P62 and NBR1, also compromised tomato heat tolerance (Figures 5–7). Thus, NBR1-mediated selective autophagy is critical in tomato heat tolerance most likely through its activity in removing heat-induced misfolded proteins. For degradation by UPS, proteins must be unfolded to enter the narrow central cavity of its barrel-shaped 20S proteolytic core since the steric conditions of a folded protein would not be able to pass through the entrance channel. Under heat stress, misfolded, or denatured proteins may form protein aggregates that are difficult to dissociate or unfold. These protein aggregates are likely to be targeted by NBR1-mediated selective autophagy if they fail to be processed by UPS. Consistent with this interpretation, compromised heat tolerance of NBR1-silenced tomato plants was associated with increased accumulation of protein aggregates under heat stress (Figure 8). These results provide further support that UPS and NBR1-mediated selective autophagy function in cooperation in the removal of misfolded proteins for protection against proteotoxic stress under adverse environmental and physiological conditions (Zhou et al., 2014c).

Heat stress induced both LysoTracker or MDC-stained autolysosome-like structures (Figures 3, 4) and expression of ATG and NBR1 genes (Figure 1). Interestingly, silencing of an ATG or NBR1 gene in tomato plants led to down regulation of not only the silenced gene but also other sequence-unrelated autophagy genes (Figure 9). Since ATG5, ATG7, and NBR1
proteins are not known to be regulators of gene transcription, their effect on the expression of other genes is probably indirect and most likely related to induced autophagy under heat stress. It is possible that induced autophagy under heat and perhaps other stress conditions as well has a positive role in the upregulation of autophagy genes. Consistent with the potential signaling role of autophagy, silencing of Arabidopsis TOR gene leads to not only constitutive formation of autophagosomes but also induced expression of some ATG genes (Liu and Bassham, 2010). We have also previously observed that in autophagy-deficient mutants, induction of jasmonate-regulated PDF1.2 gene by Botrytis infection was compromised (Lai et al., 2011b). Likewise, in the ATG5- or ATG7-silenced tomato mutants, induction of a gene encoding a HSP40 was significantly reduced (Figure 10), indicating that induced autophagy, perhaps through formation and turnover of autophagosomes, has a positive role in up-regulation of not only autophagy genes but also other genes associated with defense and stress responses. We have previously shown that in Arabidopsis atg5 and atg7 mutants, the protein levels of NBR1 increased greatly under heat stress (Zhou et al., 2013). Other studies have also showed that some of the autophagy-related proteins and NBR1 are themselves autophagy substrates (Svenning et al., 2011) that undergo rapid turnover and must be replenished through increased synthesis for sustained autophagy. On the other hand, when autophagy is suppressed or blocked as in the ATG5- and ATG7-silenced plants, there would be no need for increased transcription of the ATG or NBR1 genes since their degradation by autophagy is inhibited.

Arabidopsis WRKY33 transcription factor plays a critical role in plant resistance to necrotrophic fungal pathogens and in plant tolerance to heat stress (Zheng et al., 2006; Li et al., 2011). Autophagy is induced by necrotrophic pathogens, heat, and salt stresses and plays an important role in plant responses to these biotic and abiotic stresses as well. We have previously shown that in the Arabidopsis wrky33 mutants, increased formation of autophagosomes was observed in Botrytis-infected lesion areas but not in the areas surrounding the lesions found in wild-type plants (Lai et al., 2011b). In addition, induction of ATG18a was normal at 1 day post Botrytis infection (dpi) but was severely compromised at 2, 3, and 4 dpi in wrky33 (Lai et al., 2011b). Thus, WRKY33 is dispensable for early induction of autophagy but necessary for sustained induction of autophagy in Botrytis-infected plants. Likewise, silencing of tomato WRKY33 compromised heat-induced autophagy gene expression and reduced autophagosome formation (Figures 3, 8). Thus, it is likely that the critical role of WRKY33 in plant disease resistance and stress tolerance is, at least in part, mediated by its critical role in induction of autophagy.

In Arabidopsis, WRKY33 is subjected to dual-level regulation by the mitogen protein kinase 3 and 6 (MPK3/6) cascade (Mao et al., 2011). Upon pathogen infection, WRKY33 is phosphorylated by the pathogen/stress-induced MPK3/MPK6 and phosphorylation of WRKY33 is likely to promote the transcription activity of WRKY33, which can bind to the W boxes in its promoter and turning on its own expression (Mao et al., 2011). Expression of Arabidopsis WRKY33 is also induced by abiotic stress conditions and by paraquat, which generates ROS in exposed plant cells (Zheng et al., 2006). In plants, autophagy is also induced by a variety of stresses, including nutrient deprivation, drought, salt stress, ROS, and pathogen infection (Liu and Bassham, 2010; Lai et al., 2011b; Zhou et al., 2013). It has also been shown that NADPH oxidase inhibitors block autophagy induction by nutrient starvation and salt stress, indicating that ROS may also function as a signal in induction of autophagy by some environmental stresses (Liu and Bassham, 2010). It is tempting to speculate that multiple stress-initiated pathways may converge to the activation and induction of WRKY33 for induction of some autophagy- and other stress-related genes. Although autophagy is highly conserved in eukaryotic organisms, WRKY transcription factors are mostly plant-specific (Zhang and Wang, 2005). A critical role of WRKY33 in the regulation of plant autophagy genes would strongly indicate that the regulatory mechanisms of autophagy in plants have diverged from those in other eukaryotic organisms.
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