Two intertwined compartments coexisting in sporadic conventional colon adenomas

Carlos A. Rubio
Gastrointestinal Research Laboratory, Department of Pathology, Karolinska Institute and University Hospital, Stockholm, Sweden

Sporadic conventional colon adenomas are microscopically built of 2 intertwined compartments: one on top, harboring the dysplastic tissue that defines their histo-biomolecular attributes, and the other below, composed of non-dysplastic crypts with corrupted shapes (CCS). The CCS of 306 colon adenomas revealed asymmetric, haphazardly-distributed proliferating cell-domains (PC). In contrast, the PC-domains in normal controls were symmetric, being limited to the lower thirds of the crypts. In 28% out of 501 sporadic conventional adenomas, foci of p53-upregulated dysplastic tissue were found. The CCS in 30% of 108 sporadic adenomas showed p53-upregulated single cells, suggesting mounting somatic mutations. No p53-upregulated cells were found in the crypts of controls. In polypoid adenomas, the mucosa of the stalk without dysplastic tissue on top disclosed CCS with asymmetrical PC-domains and single p53-upregulated cells. The latter observations suggested that CCS had developed prior to and not after the growth of the dysplastic tissue on top. CCS were also found below colon adenomas in carcinogen-treated rats. It is concluded that the 2 intertwined histo-biological compartments of sporadic conventional colon adenomas are probably interdependent components. These findings may open new directions aimed to uncover the link between the normal colonic mucosa and the histogenesis of, conventional adenomas. (Intest Res 2021;19:12-20)

Key Words: Colon; Conventional adenomas; Evolution; Corrupted crypts; Cell proliferation

INTRODUCTION

The large intestine is built with a single layer of epithelial cells showing inward invaginations or folds called crypts. Crypts are lined with absorptive and mucus producing goblet cells, entero-endocrine cells and only in cecum and proximal right colon, with Paneth cells. The crypts replicate by symmetric fission, beginning at their base, and proceeding upwards until 2 identical, individual crypts are created. In well-oriented sections, a characteristic crypt assemblage of tightly packed crypts, parallel as “rows of test tubes” perpendicular to the surface-epithelium-muscularis cells mucosae-axis, are found. A slight variation in the configuration of the crypts and in the space between the crypts may occur, but crypt branching is rarely seen in fixed preparations.

In a previous study, we found only 3.7 (range, 2–5) single colon crypts lined with normal epithelium but exhibiting asymmetric shapes; called colon crypts with corrupted shapes (CCS), they were found amongst more than 200,000 crypts investigated in 22 colon segments, proximal or distal to surgically removed colon cancer in humans. The remnant crypts in the colon segments in the 22 patients showed crypts with normal shapes. These findings indicated that the vast majority of the normal mucosa of patients having synchronously a colon cancer, displays crypts with normal shapes and that CCS very rarely occur, thus resembling the normal mucosa in individuals without colon cancer.
**RISK FACTORS IN THE DEVELOPMENT OF COLON NEOPLASIAS**

The vast mucosal area of the human large bowel (estimated at 2 m²) is relentlessly exposed to a series of risk factors able to generate neoplastic lesions. The main risk factors for the development of neoplastic lesions in the colonic mucosa are: (1) sex (the risk is higher in males), (2) genetic differences (the risk is higher in patients with germline mutations in one of the DNA mismatch repair genes), (3) obesity (physical inactivity and obesity are strong independent determinants associated to colon cancer), (4) type 2 diabetes (hyperinsulinemia is important in the pathogenesis of colon cancer), (5) environmental carcinogens (such as polycyclic aromatic hydrocarbon when meat is heated above 180°C for long periods), (6) mutant gut microbiome, (7) breakdown of the gut macrophage-barrier (allowing the trespassing of gut microbiome into the host, destabilizing thereby host immunity in inflammatory bowel disease), and (8) life style factors (high use of alcohol and tobacco, known to encourage the development of sessile serrated adenoma/polyps).³⁹³⁵

**SPORADIC COLON ADENOMAS AND COLON CARCINOGENESIS**

Under the influence of one or more of the aforementioned risk factors, the mucosa of the large intestine may undergo hubs of histological, biochemical and molecular aberrations, called adenomas.¹⁶-¹⁹ Based on intraepithelial cellular alterations, adenomas are histologically divided into those with low-grade dysplasia and with high-grade dysplasia.²⁰ Only those colonic adenomas breaking through the muscularis mucosa are considered as invasive carcinomas. Intramucosal invasion is not to be regarded as invasive carcinoma according to the World Health Organization (WHO)³⁰ and the Vienna Classification.³¹

Triggered by the accumulation of molecular anomalies and epigenetic modification of gene expression,²²-²⁴ adenomas develop a particular propensity to progress to colon cancer. For many years, the general understanding was that the vast majority of colon cancer evolved from sporadic polypoid conventional (tubular or villous) adenomas via the polypoid conventional adenoma-carcinoma pathway.²⁵ Subsequently, serrated adenomas²⁶ and sessile serrated adenoma/polyps²⁷ were incorporated as alternative pathways of colorectal carcinogenesis. It has been estimated that about 30% of all colorectal cancers (CRCs) progress via the serrated pathway.²⁸²⁹ In a study of all polypoid conventional adenomas registered at this hospital between 1993 and 2000 (n = 3,135) it was found that 93% were polypoid conventional adenomas, and 7%, serrated adenomas and microtubular adenomas.³⁰³¹ Thus, at this hospital, polypoid conventional adenomas were also found to be the most commonly reported histologic phenotype of sporadic protruding adenomas.

In 1985 Muto et al. detected at colonoscopy, small non-polypoid mucosal lesions measuring up to 1 cm in diameter. At histological examination those non-polypoid mucosal lesions were found to be adenomas. Flat adenomas (FA), as they were called, were subsequently found associated with a more aggressive clinical behavior than their polypoid counterparts.³³ In 1997, Kudo et al.³⁴ reported non-polypoid (flat) colon neoplasms with a lateral diameter greater than 10 mm; they were called laterally-spreading tumors (LST). The biological significance of non-polypoid adenomas (i.e., FA and LST) was subsequently confirmed by an international group of Western and Japanese workers.³⁵

**CRYPT WITH IRREGULAR SHAPES ASSOCIATED WITH COLONIC NEOPLASIAS**

1. **Aberrant Crypt Foci**

There is much discussion concerning the significance of aberrant crypt foci (ACF) as precursors of colon cancer. ACF were first described by Bird in 1987. Following staining of the surface of the colonic mucosa with methylene blue in carcinogen-treated rodents, Bird found gross changes in the pit pattern consisting in groups of 5 or more colonic crypts.

In humans, Nascimbeni et al. found that ACF occurred more frequently in patients with CRC than in controls, and claimed that the phenomenon sustained their putative role as a preneoplastic marker. Using methylene-blue-spraying and magnifying colonoscopy Drew et al. identified in the human colon, ACF in the proximal colons of approximately 40% of individuals undergoing chromoendoscopy, concluding that ACF were the earlier morphologically identifiable lesion. In contrast, Fenoglio-Preiser and Noffsinger wrote: “In humans, the frequency of ACF substantially outweighs the frequency of adenomas and carcinomas in the same colons, which suggests that most ACF may be innocent bystander lesions.” Dolara et al. submitted that ACF and colon cancer risk seemed connected in Japan, but not in Europe or North America, and Ochiai et al. claimed that the specificity of ACF as tumor precur-
sors should be questioned, partly because of their predominant location in the proximal colon where tumors do not develop, and by their gradual disappearance over time. In addition, ACF frequently harbored K-Ras mutations, but rarely adenomatous polyposis coli mutations, strongly suggesting that the majority of ACF might not be true preneoplastic lesions. More recently, Quintanilla et al. concluded that rectal ACF in humans are not surrogate markers for CRC risk. Thus, 32 years after the initial report by Bird in rodents, the significance of ACF as precursors of colon cancer in humans, remains controversial.

2. Crypts with Asymmetric Shapes Beneath Colon Adenomas

In contradistinction to ACF which are detected by observing the methylene blue-stained mucosa from the top, crypts with architectural distortions, also called crypts with CCS, are identified sidewise in well-oriented histological sections. CCS in clusters were found assembled beneath the neoplastic canopy of 255 sporadic polyloid conventional adenomas. CCS were characterized by fission distortions (exhibiting 2 or multiple non-identical crypt fissions), length distortions (hyperplastic crypts being more than twice the length of a normal crypt), outline distortions (crypts with multi-lobate outlines or cystic dilatations), and/or axial-polarity distortions in relationship to the muscularis mucosae (serpentine pattern, crypts, intercommunicating horizontal crypts, inverted L-shaped or ring-shaped horizontal crypts in “Indian files”) (Fig. 1). All sporadic polyloid conventional adenomas examined in that survey revealed CCS in cluster, assembled beneath the adenomatous tissue. Nearly three-quarters of the 255 sporadic, polyloid conventional adenomas had 3 or more CCS below and 46.3% revealed ≥ 4 CCS.

In another study, 51 colonic non-polyloid tubular adenomas were investigated. Although the mean number of CCS was significantly higher in lateral spreading adenomas (≥ 10

![Fig. 1. Non-dysplastic crypts with corrupted shapes (CCS) found below the neoplastic tissue of sporadic conventional colon adenomas.](A) Low-power view showing multiple clusters of CCS with adenomatous epithelium (dark) on top (H&E, x 1). (B) For comparison: closer view of the normal colonic mucosa (H&E, x 10) portraying parallel crypts, as “rows of test tubes,” perpendicular to the surface-epithelium-muscularis cells mucosae-axis. (C-E) Detail, sporadic polyloid conventional colon adenomas portraying collections of CCS below the adenomatous epithelium (H&E, x 20). (F-I) Non-polyloid colon adenomas also demonstrating clusters of non-dysplastic crypts with CCS below the adenomatous epithelium (dark) on top (F: H&E, x 4; G: MNF116 a pan-epithelial marker, x 4; H: detail, H&E, x 20; I: detail, MNF116, x 20). (J) Colon adenoma in carcinogen-treated rat. Note CCS below the adenomatous epithelium (dark) on top (H&E, x 1).}
mm) than in small, (“flat”) adenomas (≤ 9 mm), the mean CCS per mm ratio was 0.94 (range, 0.50–2.00) in FA and 0.92 (range, 0.40–2.93) in lateral spreading adenomas. Hence, the lateral expansion of non-polypoid colonic tubular adenomas seems to have evolved as a result of increased numbers of CCS, without increasing the number of CCS per mm.44

3. Non-dysplastic Crypts with CCS Showing Redistributed Proliferating Cells and p53-Upregulated Cells

When sections from 60 sporadic polypoid conventional adenomas were immune-stained with the proliferation marker Ki-67, different abnormal cell-proliferation phenotypes were found in the CCS below the neoplastic tissue:45,46 (1) haphazardly asymmetrically-distributed single proliferating cell (PC) or PC clusters in 80%; (2) continuous PC domains in one or both slopes of the crypts in 17%; and (3) single, haphazardly distributed PCs in the remaining 3% (Fig. 2).46 In contrast, the crypts of 12 normal colon segments (controls) exhibited crypts with normal shapes having symmetrically distributed PC-domains predominantly limited to the lower third portion of the crypts. It should be mentioned that in early observations, Maskens and Deschner47 found upward shift of the proliferative zone in the normal crypts in patients with colon carcinoma. In contrast Arai and Kino48 found more recently that the proliferative zone in the crypts was normal in patients with colon cancer.

The p53 transcription factor (encoded by the human gene TP53) is a key tumor suppressor that regulates several signaling pathways involved in carcinogenesis. Given the resources required for TP53 gene sequencing, most workers have used p53 immunohistochemistry as a surrogate, assuming that p53 overexpression is connected with a mutation, and that the lack of abnormal expression is indicative of wild type p53.49 The immune-stained sections from the 60 sporadic, polypoid conventional adenomas disclosed CCS with haphazardly distributed p53-upregulated, often single cells, in 30% of the sporadic polypoid conventional adenomas (Fig. 3). None of the more than 200,000 crypts investigated in 22 colon segments, proximal or distal to surgically removed colon cancer revealed crypts

---

**Fig. 2.** Cell proliferation in normal colon crypts and in non-dysplastic crypts with corrupted shaped in colon adenomas. (A) Normal colon mucosa with proliferating cells (PCs); note PC, predominantly in the lower third of the crypt (Ki-67, batch MIB1 × 20). (B–J) Sporadic conventional colon adenomas showing non-dysplastic crypts with corrupted shapes, with relocated PC below the neoplastic tissue. (B–F) (Ki-67, batch MIB1 × 10); (G, H) (Ki-67, batch MIB1 × 20). (I, J) (Ki-67, batch MIB1 × 40).
with p53-upregulated cells.\textsuperscript{50} It should be mentioned that in previous studies of colon adenomas we found p53 upregulation in the adenomatous tissue in 28\% out of 501 adenomas.\textsuperscript{51} Thus, the apparently normal epithelium of the CCS beneath the adenomatous tissue of sporadic polypoid conventional adenomas concurred with an unprecedented relocation of the natural PC domains, and with p53-upregulated cells, suggesting thereby, possible mounting somatic mutations in the CCS of those neoplastic lesions.\textsuperscript{50}

In another investigation, this time with 48 non-polypoid adenomas, we found that 88\% of the CCS exhibited haphazardly asymmetrically-distributed PC-domains, 8\% continuous PC-domains in one or both slopes of the CCS, and 4\%, asymmetrically-haphazardly distributed single PC.\textsuperscript{46} The CCS in 29\% of the 48 non-polypoid adenomas also showed p53-upregulated often as single cells.\textsuperscript{46} Thus, the CCS accruing beneath the neoplastic tissue of non-polypoid adenomas also thrived with relocated PC-domains and p53 upregulated cells (the latter in nearly one-third of the polypoid adenomas), suggesting cell-kinetic alterations and possibly mounting somatic mutations. On the other hand, neither asymmetrically-haphazardly distributed PC-domains, nor p53 upregulated cells were found in any of the more than 200,000 normal colon crypts, including occasional, single CCS in controls.

4. Crypts with CCS in the Mucosa of the Stalk
While studying sporadic polypoid conventional adenomas we detected in the mucosa of the stalk, CCS without neoplastic tissue on top, strongly suggesting that CCS developed independently of the control of the neoplastic tissue.\textsuperscript{50} Sections immuno-stained with the proliferation marker Ki-67 and with p53 showed that the CCS in the stalk of sporadic polypoid conventional adenomas displayed haphazardly asymmetrically-distributed PC-domains and p53 upregulated cells. Those unexpected findings suggested that the CCS in the stalk were possibly undergoing somatic mutation.\textsuperscript{50}

COLON ADENOMAS WITH CORRUPTED CRYPTS IN RODENTS

1. ACF in Rodents
The pristine report of clusters of ACF in the colonic mucosa of
carcinogen-treated rodents by Bird,\textsuperscript{36} was followed by a plethora of publications in rodents, using various carcinogens. A recent search in PubMed (9/30/2019) revealed a total of 1,349 publications on ACF in mice and rats, revealing thereby the interest for the study of ACF as potential markers of CRC risk. The induction of ACF in rodents is, however, not specific for carcinogen-treated rodents exclusively: Xu et al.\textsuperscript{52} found that a high fat diet administration to C57BL mice treated with azoxymethane (AOM) promoted ACF formation concurrent with an increase of opportunistic pathogenic bacteria in the colon of C57BL/6 mice. Moreover, Zhou et al.\textsuperscript{53} found that mice fed with sodium nitrate developed ACF. On the other hand, Kristiansen\textsuperscript{54} submitted that the presence of a high number of ACF with high crypt multiplicity was predictive for tumor outcome in mice, and Ochiai et al.\textsuperscript{51} claimed that the presence of dysplastic ACF facilitated the identification of the early stages of colon carcinogenesis in AOM-treated rats. In contrast, Ghirardi et al.\textsuperscript{55} working with Fischer 344 rats, found that ACF were prevalent in the mid colon with lower frequencies in the distal colon and rectum, and rare in the proximal colon and cecum. In their study, ACF growing features and distribution were not correlated to adenoma and adenocarcinoma distribution. The authors concluded that the preneoplastic role of ACF and their function as intermediate biomarkers in colorectal carcinogenesis remained to be clarified.\textsuperscript{55} Perhaps, as with humans (see above) it may be summarized that the significance of ACF as potential markers of CRC risk in laboratory animals, remains unsettled.

2. Crypts with Normal Epithelium Albeit CCS in Carcinogen-Treated Rats
As in the human counterpart, the vast majority of the colonic crypts in untreated mice\textsuperscript{56} and rats\textsuperscript{57} displays close-packed test tube-shaped crypts with straight axes, vertical to the muscularis mucosae. Colon crypts replicate by symmetric fission starting at their base, and proceeding upwards, finally generating 2 separate identical crypts.\textsuperscript{58} In a more recent survey we found in the normal mucosa of the colon of a cohort of Sprague-Dawley rats\textsuperscript{59} and Fisher-344 rats\textsuperscript{7}—without CRC, despite protracted treatment with the colonotrophic carcinogen 1,2 dimethylhydrazine (DMH)—occasional crypts with architectural distortions (CCS). CCS in rats were regarded those with either asymmetric basal fission, asymmetric lateral sprouting/lateral fission, basal dilatations, or spatial aberrations of the normal (vertical) axis. The study of entire Swiss-roll sections from 25 male Sprague-Dawley rats treated with DMH suspended in ethylenediaminetetraacetic acid (EDTA) solution, disclosed a total of 202 CCS (38%) amongst 533 crypts with fission. In contrast, entire Swiss-roll sections from 10 EDTA-treated controls Sprague-Dawley rats exhibited only one CCS amongst the 571 crypts with fission (0.1%). In entire Swiss-roll sections from 25 male Sprague-Dawley rats treated with DMH, 4 adenomas with CCS accruing underneath were found.\textsuperscript{29} Hence, despite repeated injections with the colonotrophic carcinogen DMH, only occasional adenomas developed limited mucosal hubs, while the vast majority of the remaining colonic mucosa remained histologically unaltered.

In another study 25 male Fisher-344 rats were treated for 24 months, with pellets containing 500 ppm of the mutagenic pyrolysate compound 2-amino-6-methylpyrido imidazole (GluP-1).\textsuperscript{7} Swiss-roll sections from the entire colon showed that out of 357 crypts with fission (mean number of crypts with fission per animal: 14.2), 130 CCS were found intercalated with normal crypts (mean number of CCS per animal: 5.2; range, 2–12). Three colon adenomas included in the Swiss-roll sections of Fisher-344 rats revealed CCS accruing beneath the adenomatous tissue,\textsuperscript{7} thus resembling the CCS found beneath sporadic polypoid and non-polypoid adenomas in humans. It was advanced, that the accumulation of CCS in of carcinogen-treated rats could reflect mucosal sites from which adenomas ultimately developed, highlighting thereby the first histologic event of ‘field of cancerization’ in the colon of rats.

CONCLUSIONS

In this review, we highlight the occurrence of 2 intertwined compartments coexisting in sporadic conventional polypoid colon adenomas: one consisting of neoplastic (dysplastic) tissue on top, and the other of non-dysplastic crypts with asymmetric, irregular, CCS below. The absence of neoplastic tissue on top of the CCS found in the stalk of those adenomas, should rationally rule out the possibility that the adenomatous tissue had directly orchestrated the growth of crypts with CCS. CCS, both beneath adenomas and in the stalk, coexisted with haphazardly distributed, asymmetric PC-domains. In contrast, none of more than 200,000 normal colon crypts revealed asymmetrically-distributed PC-domains. The rearrangement of PC-domains in CCS beneath adenomas may be the result of the relocation of the cells that fuel the production of PCs, namely the stem cells.\textsuperscript{260} In addition, p53-upregulated cells were found in the CCS in nearly one-third of the polypoid adenomas suggesting somatic mutations in those crypts. CCS were
also found below adenomas in carcinogen-treated rats. The findings that crypts with disparate shapes (CCS) albeit with normal epithelium gather underneath the neoplastic tissue and in the mucosa of the stalk, might open new vistas to study the true histogenesis of conventional adenomas, the most frequent precancerous lesion in the human colon.

ADDITIONAL INFORMATION

Funding Source
The author received no financial support for the research, authorship, and/or publication of this article.

Conflict of Interest
No potential conflict of interest relevant to this article was reported.

Author Contribution
Rubio CA designed the review, performed the bibliographical research and wrote the paper.

ORCID
Rubio CA https://orcid.org/0000-0003-3879-5729

REFERENCES

1. Dahl J, Greenson JK. Colon. In: Mills SE, ed. Histology for pathologists. 3rd ed. Philadelphia: Lippincott Williams & Wilkins, 2007:627-648.
2. Baker AM, Gabbutt C, Williams MJ, et al. Crypt fusion as a homeostatic mechanism in the human colon. Gut 2019;68:1986-1993.
3. Boman BM, Fields JZ. An APC:WNT counter-current-like mechanism regulates cell division along the human colonic crypt axis: a mechanism that explains how APC mutations induce proliferative abnormalities that drive colon cancer development. Front Oncol 2013;3:244.
4. Rubio CA. The histologic structure of the large bowel mucosa and the evolution of the three pathways of colonic carcinogenesis in humans and in experimental animals. Recent Stud Dig Syst Anat 2018;3:1-12.
5. Levine DS, Haggitt RC. Normal histology of the colon. Am J Surg Pathol 1989;13:966-984.
6. Senior PV, Pritchett CJ, Sunter JP, Appleton DR, Watson AJ. Crypt regeneration in adult human colonic mucosa during prolonged organ culture. J Anat 1982;134(Pt 3):459-469.
7. Rubio CA. Are corrupted non-dysplastic colonic crypts the first histological event in experimental colonic carcinogenesis? Anticancer Res 2017;37:2265-2268.
8. Helander HF, Fändriks L. Surface area of the digestive tract: revisited. Scand J Gastroenterol 2014;49:681-689.
9. Reddy Parine N, Alanazi IO, Shaik JP, et al. TDG gene polymorphisms and their possible association with colorectal cancer: a case control study. J Oncol 2019;2019:7091815.
10. Davenport JR, Su T, Zhao Z, et al. Modifiable lifestyle factors associated with risk of sessile serrated polyps, conventional adenomas and hyperplastic polyps. Gut 2018;67:456-465.
11. Deb M, Sengupta D, Kar S, et al. Epigenetic drift towards histone modifications regulates CAV1 gene expression in colon cancer. Gene 2016;581:75-84.
12. O’Keefe SJ. Diet, microorganisms and their metabolites, and colon cancer. Nat Rev Gastroenterol Hepatol 2016;13:691-706.
13. Seidel DV, Azcárate-Peril MA, Chapkin RS, Turner ND. Shaping functional gut microbiota using dietary bioactives to reduce colon cancer risk. Semin Cancer Biol 2017;46:191-204.
14. Rubio CA, Langner C, Schmidt PT. Partial to complete abrogation of the subepithelial macrophage barrier against the gut microbiota in patients with ulcerative colitis and Crohn’s colitis. Histopathology 2018;72:580-587.
15. Bailie L, Loughrey MB, Coleman HG. Lifestyle risk factors for serrated colorectal polyps: a systematic review and meta-analysis. Gastroenterology 2017;152:92-104.
16. Nguyen HT, Duong HQ. The molecular characteristics of colorectal cancer: implications for diagnosis and therapy. Oncol Lett 2018;16:9-18.
17. Bogaert J, PreMen H. Molecular genetics of colorectal cancer. Ann Gastroenterol 2014;27:9-14.
18. Bardhan K, Liu K. Epigenetics and colorectal cancer pathogenesis. Cancers (Basel) 2013;5:676-713.
19. Bargen JA. Rectal and sigmoidal adenomas: potential carcinoma. Gastroenterology 1951;18:138-140.
20. Hamilton SR, Vogelstein B, Kudo S, et al. Carcinoma of the colon and rectum. In: Hamilton SR, Aaltonen LA, eds. World Health Organization Classification of Tumours. Pathology and genetics of tumours of the digestive system. Lyon: IARC Press, 2000:104-119.
21. Schlemper RJ, Riddell RH, Kato Y, et al. The Vienna classification of gastrointestinal epithelial neoplasia. Gut 2000;47:251-255.
22. Komor MA, Bosch LJ, Bounova G, et al. Consensus molecular subtype classification of colorectal adenomas. J Pathol 2018;246:266-276.
23. Sakai E, Fukuyo M, Ohata K, et al. Genetic and epigenetic aberrations occurring in colorectal tumors associated with serrated pathway. Int J Cancer 2016;138:1634-1644.

24. Neuville A, Nicolet C, Meyer N, et al. Histologic characteristics of non-microsatellite-instable colon adenomas correlate with distinct molecular patterns. Hum Pathol 2011;42:244-253.

25. Jackman RJ, Mayo CW. The adenoma-carcinoma sequence in cancer of the colon. Surg Gynecol Obstet 1951;93:327-330.

26. Longacre TA, Fenoglio-Preiser CM. Mixed hyperplastic adenomatous polyps/serrated adenomas: a distinct form of colorectal neoplasia. Am J Surg Pathol 1999;14:524-537.

27. Torlakovic EE, Gomez JD, Driman DK, et al. Sessile serrated adenoma (SSA) vs. traditional serrated adenoma (TSA). Am J Surg Pathol 2008;32:21-29.

28. O’Brien MJ, Gibbons D. The adenoma-carcinoma sequence in colorectal neoplasia. Surg Oncol Clin N Am 1996;5:513-530.

29. O’Brien MJ, Zhao Q, Yang S. Colorectal serrated pathway cancers and precursors. Histopathology 2015;66:49-65.

30. Rubio CA. Colorectal adenomas: time for reappraisal. Pathol Pract 2002;198:615-620.

31. Rubio CA, Kis L, Nastic D, Schmidt PT. Microtubular colorectal adenoma an aggressive histologic phenotype with propensity to evolve into invasive carcinoma. Arc J Cancer Sci 2017; 3:31-36.

32. Muto T, Kamiya J, Sawada T, et al. Small "flat adenoma" of the large bowel with special reference to its clinicopathologic features. Dis Colon Rectum 1985;28:847-851.

33. Kasumi A, Kratzer GL, Takeda M. Observations of aggressive, small, flat, and depressed colon cancer: report of three cases. Surg Endosc 1995;9:690-694.

34. Kudo S, Kashida H, Tamura S, Nakajima T. The problem of "flat" colonic adenoma. Gastrointest Endosc Clin N Am 1997;7:87-98.

35. Kudo Se, Lambert R, Allen JJ, et al. Nonpolyoid neoplastic lesions of the colorectal mucosa. Gastrointest Endosc 2008;68 (4 Suppl):S3-S47.

36. Bird RP. Observation and quantification of aberrant crypts in the murine colon treated with a colon carcinogen: preliminary findings. Cancer Lett 1987;37:147-151.

37. Nascimbeni R, Villanacci V, Mariani PP, et al. Aberrant crypt foci in the human colon: frequency and histologic patterns in patients with colorectal cancer or diverticular disease. Am J Surg Pathol 1999;23:1256-1263.

38. Drew DA, Mo A, Grady JJ, et al. Proximal aberrant crypt foci associate with synchronous neoplasia and are primed for neoplastic progression. Mol Cancer Res 2018;16:486-495.

39. Fenoglio-Preiser CM, Noffsinger A. Aberrant crypt foci: a review. Toxicol Pathol 1999;27:632-642.

40. Dolara P, Caderni G, Luceri C. Surrogate endpoint biomarkers for human colon carcinogenesis. Toxicol Lett 2000;112-113:415-420.

41. Ochiai M, Hippo Y, Izumiya M, Watanabe M, Nakagama H. Newly defined aberrant crypt foci as a marker for dysplasia in the rat colon. Cancer Sci 2014;105:943-950.

42. Quintanilla I, López-Cerón M, Jimeno M, et al. Rectal aberrant crypt foci in humans are not surrogate markers for colorectal cancer risk. Clin Transl Gastroenterol 2019;10:e00047.

43. Rubio CA, Schmidt PT. Are non-dysplastic crypts with corrupted shapes the initial recordable histological event in the development of sporadic conventional adenomas? Anticancer Res 2018;38:5315-5320.

44. Rubio CA, Schmidt PT. Crypts with corrupted shapes in non-polyoid adenomas. Anticancer Res 2019;39:833-838.

45. Rubio CA. Preliminary report: multiple clusters of proliferating cells in non-dysplastic corrupted colonic crypts underneath conventional adenomas. In Vivo 2018;32:1473-1475.

46. Rubio CA, Schmidt PT. The normal epithelium of crypts accruing below nonpolyoid adenomas thrives with relocated proliferating cell-domains and p53-up-regulated cells. Anticancer Res 2019;39:4965-4970.

47. Maskens AP, Deschner EE. Tritiated thymidine incorporation into epithelial cells of normal-appearing colorectal mucosa of cancer patients. J Natl Cancer Inst 1977;58:1221-1224.

48. Arai T, Kino I. Morphometrical and cell kinetic studies of normal human colorectal mucosa: comparison between the proximal and the distal large intestine. Acta Pathol Jpn 1989;39:725-730.

49. Gonzalez RS, Messing S, Tu X, McMahon LA, Whitney-Miller CL. Immunohistochemistry as a surrogate for molecular subtyping of gastric adenocarcinoma. Hum Pathol 2016;56:16-21.

50. Rubio CA, Schmidt PT. Disparate cell proliferation and p53 overexpression in colonic crypts with normal epithelial lining found below the neoplastic canopy of conventional adenomas. J Pathol Clin Res 2019;5:154-163.

51. Rubio CA, Rodesjö M, Duvander A, Mathies M, Garberg L, Shetye J. p53 up-regulation during colorectal carcinogenesis. Anticancer Res 2014;34:6973-6979.

52. Xu J, Galley JD, Bailey MT, Thomas-Ahner JM, Clinton SK, Olivo-Marston SE. The impact of dietary energy intake early in life on the colonic microbiota of adult mice. Sci Rep 2016;6:19083.
the induction of colonic aberrant crypts in mice fed sodium nitrite. Nutr Cancer 2016;68:105-112.
54. Kristiansen E. The role of aberrant crypt foci induced by the two heterocyclic amines 2-amino-3-methyl-imidazo[4,5-f] quinoline (IQ) and 2-amino-1-methyl-6-phenyl-imidazo[4,5-b]pyridine (PhIP) in the development of colon cancer in mice. Cancer Lett 1996;110:187-192.
55. Ghirardi M, Nascimbeni R, Villanacci V, Fontana MG, Di Betta E, Salerni B. Azoxyrnhane-induced aberrant crypt foci and colorectal tumors in F344 rats: sequential analysis of growth. Eur Surg Res 1999;31:272-280.
56. Richards TC. Early changes in the dynamics of crypt cell populations in mouse colon following administration of 1,2-di-

methylhydrazine. Cancer Res 1977;37:1680-1685.
57. Sunter JP, Wright NA, Appleton DR. Cell population kinetics in the epithelium of the colon of the male rat. Virchows Arch B Cell Pathol 1978;26:275-287.
58. Wasan HS, Park HS, Liu KC, et al. APC in the regulation of intestinal crypt fission. J Pathol 1998;185:246-255.
59. Rubio CA. Corrupted colonic crypt fission in carcinogen-treated rats. PLoS One 2017;12:e0172824.
60. Rubio CA. Putative stem cells in mucosas of the esophago-gastrointestinal tract. In: Singh SR, ed. Stem cell, regenerative medicine and cancer. New York: Nova Science Publishers Inc., 2010: 279-308.