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p600 Plays Essential Roles in Fetal Development

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Abstract

p600 is a multifunctional protein implicated in cytoskeletal organization, integrin-mediated survival signaling, calcium-calcmodulin signaling and the N-end rule pathway of ubiquitin-proteasome-mediated proteolysis. While push, the Drosophila counterpart of p600, is dispensable for development up to adult stage, the role of p600 has not been studied during mouse development. Here we generated p600 knockout mice to investigate the in vivo functions of p600. Interestingly, we found that homozygous deletion of p600 results in lethality between embryonic days 11.5 and 13.5 with severe defects in both embryo and placenta. Since p600 is required for placental development, we performed conditional disruption of p600, which deletes selectively p600 in the embryo but not in the placenta. The conditional mutant embryos survive longer than knockout embryos but ultimately die before embryonic day 14.5. The mutant embryos display severe cardiac problems characterized by ventricular septal defects and thin ventricular walls. These anomalies are associated with reduced activation of FAK and decreased expression of MEF2, which is regulated by FAK and plays a crucial role in cardiac development. Moreover, we observed pleiotropic defects in the liver and brain. In sum, our study sheds light on the essential roles of p600 in fetal development.

Introduction

p600, also known as UBR4, is a 600 kDa cellular protein that is ubiquitously expressed and plays various roles depending on cell type [1,2]. One obvious structural motif of p600 is the UBR box of the N-end rule pathway ubiquitin ligases. The UBR box is responsible for recognition of the N-terminus of their substrate proteins that are produced by protein processing and/or modifications in response to environmental conditions [3–6]. Additionally, p600 has a non-canonical calcmodulin-binding domain and binds to calcmodulin in a calcium-dependent manner [1]. Moreover, p600 associates with the cytoskeleton to impact cell morphology and intracellular transport [1,7].

Regarding its biological functions, p600 is involved in activation of integrin-mediated survival signaling pathways in adherent cells. Suppression of p600 expression by short hairpin RNA (shRNA) abrogates formation of integrin-mediated ruffled membranes and cellular polarity. These phenotypes in the knockdown cells are associated with reduced activation of focal adhesion kinase (FAK), which plays a role in integrin-mediated survival signaling pathways [1]. p600 functions not only in anchorage-dependent growth but also in anchorage-independent growth. Suppression of p600 expression prevents anchorage-dependent growth in various cancer cells including osteosarcoma, cervical cancers, and gastric cancers [8–10]. Moreover, knockdown of p600 suppresses growth of gastric cancer cells in SCID mice [10]. Although the exact molecular mechanisms whereby p600 contributes to cancer growth still remain unclear, p600 has been shown to be a direct target for viral oncoproteins, namely, human and bovine papillomavirus E7 [9,11]. Importantly, experiments with papillomavirus E7 mutants demonstrated a relation between p600-binding activity and transforming activity [9,9]. Therefore, formation of the E7-p600 complex in transformed cells could be crucial for anchorage-independent growth presumably by inhibiting apoptosis. Likewise, association of p600 with cellular factors may be disorganized in nonvirus-mediated cancers, although such factors have not been identified yet.

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p600 functions not only in proliferating cells but also in non-dividing cells. p600 plays an essential role in neurite outgrowth and neuronal migration during brain development through regulation of microtubule stability and transport of endoplasmic reticulum transport [7]. Neurological functions are also reported for the Drosophila counterpart of p600, termed push. Mutation in push does not cause developmental effects but triggers defects in synaptic transmission at neuromuscular junctions in adult flies [13].

In Arabidopsis, the p600 counterpart BIG plays a role in polar-dependent intercellular transport of auxin [14], a plant hormone essential for development. Specific distribution of auxin via polar transport is crucial for the regulation of auxin action and thus mutations in BIG cause developmental defects. As a mechanism, polar localization of auxin efflux carrier PIN proteins to a particular face of the plasma membrane enables the polar auxin transport [15,16]. PIN proteins dynamically cycle between the plasma membrane and endosomes. Auxin prevents the internalization step of PIN proteins in a BIG-dependent manner, resulting accumulation of PIN proteins at the plasma membrane [17]. Despite the roles assigned to p600’s counterparts, the physiological functions of p600 in mammal remain poorly understood. To gain insights into the in vivo roles of p600 we generated p600 knockout mice. Here we demonstrate that the embryonic p600 plays essential roles during fetal development.

**Figure 1. Disruption of p600 results in growth retardation and lethality during embryonic development.** (A) Targeting strategy of p600 locus. The exon encoding the initiating methionine codon of p600 allele was replaced with a Neo cassette by homologous recombination in ES cell lines. The ES cell lines were microinjected into mouse blastocysts to generate chimeric mice. These chimera were bred to obtain offspring that are heterozygous for ‘Neo allele’. p600<sup>Neo/+</sup> mice were then crossed with Ella-Cre transgenic mouse [19] to delete the loxP flanked region, yielding ‘KO allele’. The locations of PCR amplicons used for genotyping are as indicated. Restriction enzyme digestion of genomic DNA with BamHI and SpeI produces the 2.5 and 4.3 kbp fragments from WT and Neo alleles, respectively, that hybridize with the probe for Southern blotting. The regions for PCR genotypings are indicated. (B) The genotypes of living embryos produced by inter-crossing of p600<sup>+/+</sup> mice. The numbers of embryos with indicated genotypes from days E9.5 to E12.5 are shown. Significances of the frequency of survival p600<sup>−/−</sup> embryos and expected frequency of 25%, according to Mendelian distribution of the genotypes were calculated by χ<sup>2</sup> test. The p-values of χ<sup>2</sup> test are shown. ** and *** show p-value <0.01 and <0.001, respectively. (C) The appearances of typical embryos of p600<sup>−/−</sup> and p600<sup>+/+</sup> littermates from days E9.5 to E12.5. Scale bars indicate 1 mm.

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Materials and Methods

Generation of p600 Deficient Mice

Isolation of recombinant ES cell clones and blastocysts injection were performed at The Dana-Farber Transgenic Core Facility. Briefly, ES cells were electroporated with the purified NotI-linearized targeting vector constructed from pKOII [18] (see Figure 1A). Germline transmission was obtained by mating male chimeras with C57BL/6 females.

EIIa promoter-driven Cre transgenic mice [B6.FVB-Tg(EIIa-cre)C5379Lmgd/J], Sox2 promoter-driven Cre transgenic mice [STOCK Tg(Sox2-cre)1Amc/J], and ß-actin promoter-driven flippase transgenic mice [B6SJL-Tg(ACTFLPe)9205Dym/J] were purchased from Jackson Laboratory. Backcross was done with C57BL/6 stain and all the straight and conditional knockout mice were congenic with the C57BL/6 genetic background.

Ethics Statement

All animal studies were approved by the Dana-Farber Institutional Animal Care and Use Committee (IACUC) (Animal Protocol Numbers 03-038 and 05-003) and were performed in strict accordance with IACUC guidelines.

Southern Blot Analysis for Genotyping

Genomic DNA isolated from embryos were digested with BamHI and SpeI (Figure 1A), fractionated by agarose gel electrophoresis, and transferred onto nitrocellulose membrane. The membrane was hybridized with the 32P-dCTP-labeled 880 bp PstI and EcoRI fragment purified from the targeting plasmid (Figure 1A).

Figure 2. Placental abnormalities in p600−/− animals. (A) Placental labyrinth defects in p600−/− animals. H&E staining images of the labyrinth layer of p600 knockout and control littermate at E12.5 (the area below the dashed lines) are shown. The labyrinth areas of p600−/− placenta are thinner than those of wild type littermate as shown by arrows. Scale bars indicate 500 μm. (B) High magnification images of the labyrinth layer. Dilated blood vessels (arrows) are observed in p600−/− placenta. Scale bars indicate 50 μm. (C) Immunofluorescence staining of blood vessels in labyrinth areas with anti-laminin antibody. The blood vessels in labyrinth are dilated and sparse in p600 KO animals at E12.5. Scale bars indicate 100 μm.

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p600 Knockout Mice

PCR Analysis for Genotyping

PCR-based genotyping analysis was performed using genomic DNA isolated from embryos. The following primers were employed for PCR amplification. For detection of 'WT allele' (amplicon length of 158 bp), a set of primers, 5'-TGCTTGATGGCCTAGTATGTG-3' and 5'-CTTATTTCGCAGCAGCTTTTCTT-3', was used. For detection of 'KO allele' (amplicon length of 242 bp), a set of primers, 5'-AAAGACTGTGGATGGCTAC-3' and 5'-GAGAAGGACTCGCATCCTGAG-3', was used. In the conditional knockout experiments, for detection of 'floxed allele' (amplicon length of 269 bp) and 'WT allele' (amplicon length of 235 bp), a set of primers, 5'-CACCCAGACCGAGGTCTTAC-3' and 5'-CAGCAAGCJAAGGACAGTC-3', was used. For detection of Sox2-Cre transgene (amplicon length of 102 bp), a set of primers, 5'-GGGTCCTGGGACGATACTATC-3' and 5'-GTGAAAACGCACTTGTGACCTT-3', was used. As an internal control for PCR reactions, interleukin-2 gene was coamplified (amplicon length of 324 bp) with a set of primers, 5'-CTAGGCGCCA-GAATTGAAGATCT-3' and 5'-GTAGGTGGAAATTCCATGCATCC-3'.

Western Blot Analysis

For Western blot analysis, protein lysates were prepared from embryos by extracting with RIPA buffer. 10 µg of lysates were fractionated by SDS-PAGE and transferred to a Nitrocellulose membrane. The membranes were probed with anti-p600 [1] and -actin (I-19; Santa Cruz Biotechnology) antibodies and detected with horseradish peroxidase–conjugated secondary antibodies using ECL detection system.

Histology and Immunohistochemistry

Tissues were fixed in Bouin's solution or 4% paraformaldehyde in 0.1 M PBS for 24 hr, passed through graded alcohols, and embedded in paraffin. After deparaffinization, some sections were stained with Hematoxylin & Eosin (H&E). The surface areas were calculated using WinROOF software (Mitani Corporation).

For immunohistochemistry, paraformaldehyde-fixed samples were incubated with the primary antibodies after deparaffinization and heat-induced antigen retrieval. For fluorescent immunohistochemistry, samples were incubated with secondary antibody conjugated with Alexa Fluor 568 (Life Technologies). For colorimetric immunohistochemistry, samples were incubated with secondary antibody conjugated with polymeric horseradish peroxidase (Vector Laboratories) and visualized with 3,3′- Diaminobenzidine (DAB) chromogen (Dako). The antibodies employed were as follows: rabbit anti-laminin antibody (L9393, Sigma-Aldrich), rabbit anti-Ki-67 (VP-RM04, Vector Laboratories), rabbit anti-desmin (sc-17785, Santa Cruz Biotechnology), and rabbit anti-MEF2 (sc-17785, Santa Cruz Biotechnology), mouse anti-phospho-FAK (9664, Cell Signaling Technology), mouse anti-MAP2 (sc-7457, Santa Cruz Biotechnology), mouse anti-MAP2 (sc-7457, Santa Cruz Biotechnology), and rabbit anti-cleaved caspase-3 (Asp175) #9664, Cell Signaling Technology). TUNEL assays were performed by using ApoTag Peroxidase In Situ Apoptosis Detection Kit (EMD Millipore) according to the manufacturer’s protocol.

Results

p600 is Required for Embryonic Development

To investigate the role of p600 in mice, we generated floxed mutant mice, which possess loxP sites flanking the exon encoding the initiating methionine (Figure 1A). To delete the floxed alleles the mutant mice was crossed with Elsa-Cre transgenic mice, which
To delete the Neo cassette inserted between frt (flippase the mice containing heterozygous ‘Neo allele’ (Figures 1A and 3A). sites flanking the exon-encoding the initiating methionine, from (IC), separating the left and right ventricles. On the other hand, indicate 500 p600 KO embryos developed in a population with Mendelian inheritance. Moreover, appearances and sizes of p600 KO embryos were similar to those of wild type or heterozygous littersmates (Figure 1B). However, at E10.5, a large proportion of living p600 KO embryos were clearly smaller in size than wild type or heterozygous littersmates. After E11.5, the total proportion of surviving p600 KO embryos was significantly less than the predicted Mendelian ratio. This indicates that some of the knockout embryos start to die around E11.5 (Figure 1C). No living p600 KO embryos were detected at E13.5, indicating that p600 KO embryos die between E11.5 and E13.5 (Figure 1B). These results indicate that p600 is essential for embryonic development.

p600 Knockout Impairs Placental Development with Thin Labyrinth Layer

The formation of a proper placenta is critical for fetal development. In normal embryos at days from E8.5 to E10.5, branching morphogenesis of the labyrinth occurs to form dense villi, the site of exchange of nutrients. The development of placenta continues afterward and reaches maturity at E14.5. The mature placenta consists of three layers: the labyrinth, the spongio-trophoblast, and the maternal decidua [20]. Histological analysis of p600 KO placenta showed that the labyrinth layer is thinner than that of the wild-type placenta (Figure 2A). Moreover, the labyrinth layer of the KO placenta contains abnormally dilated maternal blood vessels full of serum and with monocytes margined along the endothelial inner lining of the blood vessels (Figure 2B).

During normal gestation, placental blood vessels play crucial roles by exchanging nutrients, oxygen, and wastes between embryo and mother [20]. In this perspective, we next examined the architecture of blood vessels in knockout animals by immunohistochemical staining with laminin antibody. Fluorescent immunohistochemistry showed that p600 KO placenta contains dilated vessels with disorganized architecture in the labyrinth layer (Figure 2C). Taken together, we conclude that p600 is required for the formation of the placenta.

Sox2-Cre-mediated p600 Conditional Knockout Embryos Survive Longer than the Straight Knockout Embryos

These placental defects preclude to a clear definition of the embryonic roles of p600. To eliminate secondary influences of placental abnormalities we generated conditional knockout of p600 using Sox2 promoter-driven Cre transgene [Tg (Sox2-Cre)], which disrupts p600 alleles in embryos but not in placenta [21]. We first produced p600 floxed mutant mice, which possess loxP sites flanking the exon-encoding the initiating methionine, from the mice containing heterozygous ‘Neo allele’ (Figures 1A and 3A). To delete the Neo cassette inserted between frt (flippase recognition target) sites, the heterozygous mice were crossed with β-actin promoter driven-flippase transgenic mice [22] (Figure 3A). Accordingly, we obtained mice containing the p600 ‘floxed allele’. The resulting mice were crossed with Tg (Sox2-Cre) mice [21] to delete the fragment containing the initiating methionine flanked by loxP sites in embryos (Figure 3A).

First, p600lox/lox mice were consequently crossed with Tg (Sox2-Cre) mice to obtain p600Cre/− offspring with heterozygous Sox2-Cre allele (p600lox/+ ; Sox2-Cre). Then to produce p600−/− conditional knockout (p600 cKO) mice, male p600lox/− ; Sox2-Cre mice were crossed with female p600lox/+ ; Sox2−/− mice. We analyzed the genotype and gross morphology of living embryos from E12.5 to E14.5 (Figures 3B and 3C, and S2). In the straight KO embryos generated by inter-crossing of p600lox/− parents, only ~1% of total living embryos were p600−/− at E12.5 and none of them were p600−/− at E13.5 (Figure 1B). In contrast, in the conditional knockout strain, 15% of total living embryos were p600−/− at E12.5 and 6% were p600−/− at E13.5 (Figure 3B). These results indicate that the survival period of p600−/− embryos is prolonged by the embryo-specific knockout. However, no living p600 cKO embryos were observed after E14.5. While gross morphology of p600 cKO embryos at E12.5 revealed mild growth retardation, the bulges sticking up from the middle of the head were observed in the mutant animals. This phenotype became more obvious at later stage such as E13.5 (Figure 3C), thereby suggesting an important role for p600 during brain development (see below and Discussion).

We next verified whether placental abnormalities found in the straight p600 knockout are spared in the conditional knockout animals. As expected, no structural abnormalities in the labyrinth layer were observed in p600 cKO animals (Figure S3A). Blood vessels of the labyrinth layer were unaffected in p600 cKO animals as well (Figure S3B). Taken together, these data indicate that p600 is intrinsically crucial for the fetal development.

Embryonic p600 is Essential for the Cardiac Development

Histological analysis of p600 cKO embryos revealed serious defects in cardiac development. In normal fetal heart, the interventricular septum developed from the inner surface of the ventricles is fused with the inferior atroventricular endocardial cushions, separating left and right ventricles at E12.5 (Figure 4A, left) [23,24]. In the cKO heart, development of the interventricular septum and the inferior atroventricular endocardial cushions is immature, forming a large canal between left and right ventricles (Figure 4A, right). Although the volume of ventricles increased from E12.5, at E13.5, the formation of the ventricular septum remains immature in p600 cKO (Figure 4B). Moreover, the cKO heart has a thinner ventricular wall when compared to the control heart (Figures 4C and 4D). From these observations, we conclude that loss of p600 during embryogenesis is associated with defects in heart development. This may eventually lead to gross growth retardation and embryonic death.

To gain insights into the molecular mechanisms underlying developmental defects in the p600 cKO heart we first analyzed proliferation of cardiomyocytes by immunohistochemistry with
Figure 5. Proliferation defects in conditional p600 knockout heart. Transverse sections of p600<sup>+/−</sup> (p600<sup>+/−</sup>/Sax2-Cre<sup>+</sup>) (A) and p600 cKO (B) heart at days E13.5 were stained with anti-Ki-67 antibody. After immunohistological staining, the samples were counterstained with hematoxylin. Magnified images of the ventricular wall (top) and the interventricular septum (bottom), the region indicated by rectangles, are shown in the right panels. Scales bars in the left and right panels indicate 500 and 200 μm, respectively. (C) The percentage of Ki-67 negative cells in the ventricular wall and the interventricular septum are shown. The p-values for Student’s t-test are shown in the graph. ** and *** show p-value <0.01 and <0.001, respectively.

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Figure 6. Defects in FAK activation in conditional p600 knockout heart. Transverse sections of p600+/− (p600+/−/Sox2-Cre+) (left) and p600 cKO (right) heart at days E13.5 were stained with anti-desmin (A), anti-FAK (B), anti-phospho-FAK (Tyr397) (C), and anti-MEF2 (D) antibodies. In panels C and D, the position of the inferior atrioventricular endocardial cushions (IC) is labeled. Scale bars indicate 200 μm.
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anti-Ki-67 antibody (Figures 5). Ki-67, a cellular marker for proliferation, is expressed during all cell cycle stages in proliferating cells but not in quiescent or terminally differentiated cells [25]. The ratio of Ki-67-negative cells to the total cells was significantly increased in the cKO heart when compared to that in the control heart (Figure 5C). This result suggests abnormal proliferation of cardiac cells in \( p600 \) cKO embryos.

We have attempted to detect apoptotic cells by TUNEL assay as well as immunohistochemistry with the antibody specific to the cleaved caspase-3. Interestingly, apoptotic cells were barely detected in the cKO and control heart (data not shown). These data are consistent with the H&E data, showing that there is no detectable dead cardiomyocytes in the cKO and control heart (Figure 4). Taken together, these data indicate that developmental defects in the \( p600 \) cKO embryos could be due to deceleration of cell proliferation rather than acceleration of cell death.

To examine whether cardiomyocytes differentiate properly we immunostained the sections with antibody against desmin, a subunit of intermediate filaments specific to muscle tissues. The stained images indicate that the cKO heart express desmin at the level comparable to the control heart (Figure 6A).

FAK is a non-receptor protein tyrosine kinase involved in integrin-mediated signal transduction. FAK is activated by the establishment of integrin-extracellular matrix interactions and growth hormones. Activation of FAK results in autophosphorylation at Tyr397, serving as a binding site for the Src homology 2 (SH2) domains of Src-family tyrosine kinases and the p85 subunit of phosphatidylinositol 3-kinase (PI3K) [26]. By knocking down \( p600 \) expression with shRNA, we previously demonstrated that \( p600 \) is involved in activation of FAK in cultured fibroblasts [1]. Moreover, conditional knockout of \( FAK \) in the embryonic heart has been shown to cause the similar heart defects, i.e., ventricular septal defects and thin ventricular walls [27]. Accordingly, we examined FAK activation in the \( p600 \) cKO heart. No obvious difference was observed in FAK expression between the cKO and control heart (Figure 6B). In contrast, the overall level of phosphorylated FAK at Tyr397 in the cKO heart was significantly lower than that in the control heart (Figure 6C). Taken together, these results indicate defective FAK activation in the heart of cKO embryos.

We next determined the levels of myocyte enhancer factor 2 (MEF2), which is under regulation of FAK [27] (Figure 6D). Like phosphorylated FAK, expression of MEF2 was also reduced in the cKO heart. Interestingly, both phosphorylated FAK and MEF2 accumulated at the surface cells of the inferior atrioventricular endocardial cushions in the cKO heart (Figures 6C and 6D). Taken together, we conclude that \( p600 \) may play a role in the activation of FAK signaling pathways during cardiac development.

Embryonic \( p600 \) is Required for Development of the Liver

In addition to the cardiac defects, pleiotropic defects were observed in \( p600 \) cKO embryos. At E12.5, cKO liver displays reduced cellularity and increased empty space (or the hepatic sinusoids) (Figure 7A, right), indicative of less dense packing of parenchymal hepatocytes, when compared to the control liver (Figure 7A, left). At E13.5, in the control liver, parenchymal hepatocytes become denser and form the structural units called hepatic lobules (Figure 7B, left). On the other hand, blood vessels including sinusoids are very dilated in the mutant liver (Figure 7B, right). These results indicate that \( p600 \) plays an essential role in liver development.
Embryonic p600 is Required for Development of the Brain

Consistent with the abnormal gross morphology of the head observed in p600 cKO embryos (Figure 3C), analysis of brain serial cross sections revealed significant defects. p600 cKO embryos have considerably smaller brain with markedly small ventricular spaces and cephalic mesenchyme tissues (Figure 8). Furthermore, cKO brain appears to be compressed especially in the posterior portion. Interestingly, although there is no significant difference in the area of the neural tubes between the cKO and control brain, the area of ventricles and cephalic mesenchyme tissues is significantly reduced in the cKO brain (Figure 8C). Especially, the fourth ventricle is mostly closed through all cross sections. Moreover, severe cell death with cavitation was observed in the lateral (Figure 8B) and medial (data not shown) ganglionic eminences, which localize below the cortex of forebrain. Thus, p600 is important for brain development.

Discussion

In summary, we demonstrated that p600 plays essential roles in embryonic development and formation of placenta. Moreover, by generating conditional knockout mice in which p600 is deleted
selectively in embryos but not in placenta, we showed that embryonic p600 is critical for the development of fetal organs. The embryonic lethal phenotype was totally unexpected for us since push, the Drosophila counterpart to p600, is not essential for development to adult flies [13,20]. Three independent push mutants have been characterized; two ethyl methanesulfonate-induced mutants, which possess nonsense mutations at the N-terminal region of push and a transposon-induced mutant, which has the P-element insertion at just upstream of the translation start site of push [29]. Importantly, Drosophila embryos harboring these push mutations develop into adult flies and are viable despite abnormalities in behavior and synaptic transmission at the neuromuscular junction [13,28,29]. Mouse p600 and Drosophila push have similar protein sizes (5,180 and 5,322 amino acids, respectively) and share significant sequence similarities in the broad regions, which cover >95% of the entire proteins. Nevertheless, our data demonstrate that p600 plays specific and essential roles during development in mammal.

First, we demonstrated that conditional knockout of p600 in the epiblast cells causes embryonic lethal phenotype associated with cardiac developmental problems, including ventricular septal defects and thin ventricular walls (Figure 4). Immunohistological analysis indicates that the p600 knockout cardiomyocytes have defects in activation of FAK and expression of MEF2, which is regulated by FAK [27] (Figure 6). Importantly, MEF2 has been shown to play a crucial role in myogenesis and cardiac development [30]. Consistent with our results, conditional knockout of FAK with Cre element under the control of the myosin light chain 2a promoter (MLC2a-Cre), which deletes FAK specifically in embryonic heart, results in ventricular septal defects and thin ventricular walls. Thus, failure in FAK activation could be one of the mechanisms underlying cardiac developmental defects in p600 cKO.

In support of our findings, the relevance of p600 to congenital heart defects (CHD), which are structural problems with the heart present at birth, has been suggested [31]. Ventricular septal defect (VSD), atrial septal defects (ASD), and tetralogy of Fallot (TOF) are known to be the most common CHD. To identify genes responsible for CHD, RNA arbitrarily primed-PCR fingerprinting was performed using RNAs isolated from three types of CHD and non-CHD hearts. From these screenings, p600 (hCalo) was identified as a highly expressed gene in the VSD samples, suggesting a role of p600 in the ventricular septum formation in human development.

As p600 is important for proliferation cardiomyocytes (Figure 5), it could also be critical for the proliferation of neural progenitors in the developing brain. In addition, suppression of p600 expression in the fetal brain by in utero electroporation of p600 RNA interference revealed a key role for p600 in neuronal migration in the developing neocortex [7]. Thus, defects in neurogenesis and neural migration could be among the causes underlying the microcephalic phenotype of p600 cKO embryos. At this stage, we cannot exclude the possibility that apoptosis, autophagy and/or necrosis may also contribute to the brain phenotype. Further in-depth studies are underway to unravel the multiple roles of p600 in the developing brain.

In the course of this work, the Kwon lab has independently developed UBR1/p600 knockout mice that, like our mice, also show an embryonic lethal phenotype [32]. However, their knockout animals display developmental defects at significantly earlier stage (E9.0) by pleiotropic abnormalities, including defects in the yolk sac, precluding a clear analysis of organ development at later stages. Although it is uncertain why death occurred at different stages in the two independent knockouts, a possibility would reside in the different knockout strategies employed. While we deleted a p600 region containing the first exon (Figure 1A), Tasaki et al. replaced a region containing exons 36 through 42, which encodes the UBR box, with a cassette encoding the internal ribosome entry site (IRES), tau-lacZ, and poly-A so that the lacZ reporter is expressed from the p600 transcription start site [32]. In their strategy, the p600 N-terminal region (exons 1 through 35) and lacZ is transcribed as a bicistronic mRNA. Accordingly, it would be possible that the truncated p600 protein, which is potentially expressed in the knockout animals, has inhibitory functions in the fetal development. In sum, our work with the embryo-specific knockout animals presents the great and unique advantage to understand roles of p600 in fetal development. Further studies will address the molecular mechanisms whereby p600 contributes to fetal development.

Supporting Information

Figure S1 Genotype and protein analyses of p600 straight knockout embryos. (A) Genotype analysis of the knockout embryos by Southern blotting. Genomic DNAs isolated from embryos were digested with BamHI and SpeI and hybridized with the 32P-labeled probe (Figure 1A). The probe hybridizes with 2.5 and 4.3 kbp fragments in WT and KO alleles, respectively. The identified genotypes of p600 are indicated on the top. The positions of DNA molecular weight markers are indicated on the left. (B) Genotype analysis of the knockout embryos by PCR. Genotypes of p600 were determined by genomic PCR using primer sets, which amplify WT (top) and KO alleles (bottom) (see Materials and Methods and Figure 1A). (C) Detection of p600 protein in the knockout embryos. Crude protein extracts were prepared from a day E10.5 embryo and performed Western blotting with anti-p600 (top) and anti-actin (bottom) antibodies. The genotypes of embryos are shown on the top. The positions of protein molecular weight markers are indicated on the right.

Figure S2 Genotyping of p600 conditional knockout embryos. The genotypes of p600 cKO embryos were determined by genomic PCR with primer sets which amplify WT and floxed alleles (top), KO allele (middle), and Sox2-Cre transgene as described in Material and Methods. Determined genotypes are indicated on the top.

Figure S3 Structural abnormalities of placentas found in the straight p600 KO are recovered in p600 cKO animals. (A) H&E staining sections of days E12.5 and E13.5 placentas isolated from p600 cKO and p600 cKO/Sox2-Cre animals. Scale bars indicate 50 m. (B) Immunofluorescence staining of blood vessels in labyrinth areas with anti-laminin antibody. There results indicate that there is no significant difference between lymphatics isolated from p600 cKO and p600 cKO/+ animals. Scale bars indicate 100 m.

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Author Contributions
Conceived and designed the experiments: YN, PS, KI, TN, KA, YG.
Performed the experiments: TN, KI, SM, AMR, QY, YN.
Analyzed the data: Conceived and designed the experiments: YN, PS, KI, TN, KA, YG.

References

1. Nakatani Y, Konishi H, Vassilev A, Kurooka H, Ishiguro K, et al. (2005) p600, a unique protein required for membrane morphogenesis and cell survival. Proc Natl Acad Sci U S A 102: 15093–15098.
2. Tasaki T, Mulder LC, Iwamatsu A, Lee MJ, Davydov IV, et al. (2005) A family of mammalian E3 ubiquitin ligases that contain the UBR box motif and recognize N-degrons. Mol Cell Biol 25: 7120–7136.
3. Bailey-Serres J, Fukao T, Gibbs D, Holdsworth MJ, Lee SC, et al. (2012) Making sense of low oxygen sensing. Trends Plant Sci 17: 129–138.
4. Saadfarzan R, Mastrolo A (2013) Plant oxygen sensing is mediated by the N-end rule pathway: a milestone in plant anaerobiology. Plant Cell 23: 4173–4183.
5. Sriram SM, Kim BY, Kwon YT (2011) The N-end rule pathway: emerging functions and molecular principles of substrate recognition. Nat Rev Mol Cell Biol 12: 735–747.
6. Sriram SM, Park KS, Kwon YT (2012) The N-end rule pathway. Annu Rev Biochem 81: 261–289.
7. Shim SY, Wang J, Azada N, Neumayer G, Tran HC, et al. (2008) Protein 600 is a microtubule/endoplasmic reticulum-associated protein in CNS neurons. J Neurosci 28: 3604–3614.
8. DeMasi J, Huh KW, Nakatani Y, Munger K, Howley PM (2005) Bovine papillomavirus E7 transformation function correlates with cellular p600 protein binding. Proc Natl Acad Sci U S A 102: 11496–11491.
9. Huh KW, DeMasi J, Ogawa H, Nakatani Y, Howley PM, et al. (2005) Association of the human papillomavirus type 16 E7 oncoprotein with the 600-kDa retinoblastoma protein-associated factor, p600. Proc Natl Acad Sci U S A 102: 11492–11497.
10. Sakai H, Ohuchida K, Mizumoto K, Cui L, Nakata K, et al. (2011) Inhibition of p600 expression suppresses both invasiveness and anoikis resistance of gastric cancer. Ann Surg Oncol 18: 2057–2065.
11. Black BL, Olson EN (1998) Transcriptional control of muscle development by myocyte enhancer factor-2 (MEF2) proteins. Annu Rev Cell Dev Biol 14: 167–192.
12. Webb S, Brown NA, Anderson RH (1998) Formation of the atrioventricular septal structures in the normal mouse. Circ Res 82: 645–656.
13. Rakic P, Willing K (1990) Development of structures and transport functions of mammalian E3 ubiquitin ligases that contain the UBR box motif and recognize N-degrons. Mol Cell Biol 25: 7120–7136.
14. Paciorek T, Zazimalova E, Ruthardt N, Petrasek J, Stierhof YB, et al. (2005) Auxin inhibits endocytosis and promotes its own efflux from cells. Nature 435: 1251–1256.
15. Bardeesy N, Sinha M, Herr AF, Signoretti S, Hathaway NA, et al. (2002) Loss of the Lkb1 tumour suppressor provokes intestinal polyposis but resistance to transformation. Nature 419: 162–167.
16. Watson ED, Cross JC (2005) Development of structures and transport functions in the mouse placenta. Physiology (Bethesda, Md) 20: 180–193.
17. Hacali S, Lewis P, Pevny L, McMahon AP (2002) Efficient gene modulation in mouse embryonic fibroblasts using a Nso2Cre transgenic mouse strain. Mech Dev 119 Suppl 1: S97–S101.
18. Rodriguez CI, Buchholz F, Galloway J, Sequeria R, Kasper J, et al. (2000) High-efficiency deletor mouse show that FLPs is an alternative to Cre-loxP. Nat Genet 25: 139–140.
19. Harvey RP (2002) Patterning the vertebrate heart. Nat Rev Genet 3: 544–556.
20. Sekelsky JJ, McKim KS, Messina L, French RL, Hurley WD, et al. (1999) Identification of novel Drosophila meiotic genes recovered in a P-element screen. Genetics 152: 529–542.
21. Hayashi S, Lewis P, Pevny L, McMahon AP (2002) Efficient gene modulation in mouse embryonic fibroblasts using a Nso2Cre transgenic mouse strain. Proc Natl Acad Sci U S A 103: 6638–6643.
22. Yager J, Richards S, Hekmat-Scafe DS, Hurdl RD, Sundaresan V, et al. (2001) Control of Drosophila perineural glial growth by interacting neurotransmitter-mediated signaling pathways. Proc Natl Acad Sci U S A 98: 10445–10450.
23. Sekelsky JJ, McKim KS, Messina L, French RL, Hurley WD, et al. (1999) Identification of novel Drosophila meiotic genes recovered in a P-element screen. Genetics 152: 529–542.
24. Black BL, Olson EN (1998) Transcriptional control of muscle development by myocyte enhancer factor-2 (MEF2) proteins. Annu Rev Cell Dev Biol 14: 167–192.
25. Sun G, Yuen Chan S, Yuan Y, Wang Chan K, Qiu G, et al. (2002) Isolation of a microtubule/endoplasmic reticulum-associated protein in CNS neurons. Annu Rev Biochem 81: 261–289.