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Associations Between *IGF1*, *IGFBP2* and *TGFβ3* Genes Polymorphisms and Growth Performance of Broiler Chicken Lines

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Simple Summary: The main goal of breeding programs for broiler chickens is to increase growth rate and breast and thigh muscles weight. The candidate gene approach is a powerful technique for genetically improving performance traits in chickens. We studied the associations of the single nucleotide polymorphisms (SNPs) in the *IGF1*, *IGFBP2*, and *TGFβ3* genes with performance traits in the Hubbard F15 and Cobb E chicken lines. Based on our results, it can be concluded that the *TGFβ3* gene could be used as a candidate gene marker for chicken growth traits.

Abstract: Marker-assisted selection based on fast and accurate molecular analysis of individual genes is considered an acceptable tool in the speed-up of the genetic improvement of production performance in chickens. The objective of this study was to detect the single nucleotide polymorphisms (SNPs) in the *IGF1*, *IGFBP2*, and *TGFβ3* genes, and to investigate their associations with growth performance (body weight (BW) and average daily gain (ADG) at 14, 21, 28, 35 and 42 days of age) and carcass traits in broilers. Performance (carcass) data (weight before slaughter; weights of the trunk, giblets, abdominal fat, breast muscle and thigh muscle; slaughter value and slaughter percentage), as well as blood samples for DNA extraction and SNP analysis, were obtained from 97 chickens belonging to two different lines (Hubbard F15 and Cobb E) equally divided between the two sexes. The genotypes were detected using polymerase chain reaction- restriction fragment length polymorphism (PCR-RFLP) methods with specific primers and restrictase for each gene. The statistical analysis discovered significant associations (*p* < 0.05) between the *TGFβ3* SNP and the following parameters: BW at

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Association analysis of BWs (at 21, 28 and 35 days) and SNPs was always significant for codominant, dominant and overdominant genetic models, showing a possible path for genomic selection in these chicken lines. Slaughter value was significant for codominant, recessive and overdominant patterns, whereas other carcass traits were not influenced by SNPs. Based on the results of this study, we suggested that the $\text{TGF}\beta_3$ gene could be used as a candidate gene marker for chicken growth traits in the Hubbard F15 and Cobb E population selection programs, whereas for carcass traits further investigation is needed.

**Keywords:** chicken; SNP; $\text{IGF}_1$; $\text{IGFBP}_2$; $\text{TGF}\beta_3$; Hubbard F15; Cobb E; growth; meat

## 1. Introduction

With the growth of the human population [1], the total amount of meat consumed increases at a global level worldwide [2]. Meat consumption rose worldwide from 23.08 kg per person per year in 1961 to 43.22 kg per person per year in 2013 [2]. Chicken meat is one of the most consumed types of meat worldwide [3].

Growth performance and carcass traits are the most important economic traits in broiler chicken production, and are controlled by a number of genes [4]. Growth is a complex process that involves the regulated coordination of a wide range of neuroendocrine pathways [5]. Therefore, it is very difficult to achieve rapid genetic improvement in these traits using only traditional selection methods. The growing knowledge of the structure and function of the chicken genome can be beneficial, and can lead to the recognition of causal genes and the development of new selectable molecular markers.

Although the *Gallus gallus* (chicken) genome was first sequenced as early as 2004 [6], it still required further improvements [7–9]. The newest version of the chicken genome assembly (Gallus_gallus-5.0; GCA_000002315.3), built from combined long single molecule sequencing technology, finished bacterial artificial chromosomes (BACs) and improved physical maps, was presented in 2017 [10]. Since the methodological approach has improved, the originally reported size of the chicken genome has increased from 1.05 [6] to 1.23 Gb, which has contributed to the increased number of genes observed [10]. Initial assemblies have been found insufficient for the more complete discovery of allelic contributions to complex traits [10], leading to ongoing efforts to improve the quality of the chicken reference genome [8,11].

However, the genetic improvement of polygenic traits, including growth performance and meat production, can be accomplished by marker-assisted selection that is more accurate in estimating the animal’s genetic value [12]. The molecular markers linked to quantitative trait loci (QTLs) are not affected by environmental conditions. Therefore, they could increase the speed and effectiveness of animal breeding progress. As soon as the relationship between a DNA polymorphism and an important trait is revealed, the DNA marker may be used [13]. The candidate gene approach has become a powerful technique for the genetic improvement in chicken breeding programs, and can result in increased efficiency in detecting the required production performance traits [4].

The main objectives of the strategy in commercial broiler breeding programs include increasing growth rate and breast muscle weight, reducing abdominal fat content, improved feed efficiency and increased fitness. The relationships between these individual production traits are very complex and some of them are very difficult to measure. Therefore, the use of molecular marker-assisted selection (MMAS) is necessary. In case that the favorable allele is rare, a larger positive impact can be expected [14].

The purpose of the present study was to identify polymorphisms and evaluate the association between polymorphisms in three studied genes—$\text{IGF}_1$ (insulin-like growth factor 1), $\text{IGFBP}_2$ (insulin-like growth factor binding protein 2) and $\text{TGF}\beta_3$ (transforming growth factor $\beta$)—with growth performance and meat production in chickens from two broiler lines: Hubbard F15 and Cobb E. The biological functions and interdependence of these genes are shown in Figure 1.
Figure 1. Signaling cascade of insulin-like growth factor 1 (IGF-1) and its potential impacts in metabolism, its interactions with transforming growth factor β3 (TGF-β3) and the biological functions of the IGF1, IGFBP2 and TGFβ3 genes. IGF-1 bioavailability is modulated by IGF binding proteins (IGFBPs) [15]. IGF-1 action is mediated by its binding to its receptor [16], the type 1 insulin-like growth factor receptor (IGF-1R). IGF-1R is a heterotetramer composed of two extracellular α subunits and two transmembrane β subunits, as shown in Figure 1. α subunits are cysteine-rich regions, whereas β subunits possess a tyrosine kinase domain, which constitutes the signal transduction mechanism [16]. Tyrosine phosphorylation activates a signaling cascade [17]. IGF-1 has autocrine, paracrine [18,19] and endocrine effects [18]. IGF-1 binds to its receptor (IGF-1R) in the cell membrane, resulting in autophosphorylation and the recruitment of the adaptor proteins–insulin receptor substrate IRS-1, IRS-2, and the proto-oncogene tyrosine-protein kinase (SRC) homology and collagen protein (SHC). The serine/threonine kinase (AKT) is activated by the 3-phosphoinositide-dependent protein kinase-1 (PDK1) and by the mammalian target of rapamycin (mTOR)-containing complex mTOR-C2, leading to the phosphorylation at threonine 308 and serine 473, respectively. Activated AKT regulates downstream signaling molecules such as tuberous sclerosis protein 1/2 (TSC-1/2), which inhibit mTOR-C1 complex and regulate the ribosomal protein S6 kinase 1/2 (S6K-1/2) and eukaryotic translation initiation factor 4E (eIF4E)-binding protein 1 (4E-BP1) phosphorylation, FOXO transcription factors, glycogen synthase kinase-3β (GSK-3β), p27, BCL-2 antagonist of cell death (BAD), and BCL-2. These substances are involved in some cellular processes such as protein synthesis, glucose metabolism and cell survival. SHC activation induces the activation of the RAS/mitogen-activated protein (MAP) kinase pathway, resulting in enhanced cell proliferation [15]. Activation of IRS induces the activation of intracellular RAF/MEK/ERK/RAS and PI3K signaling pathway. The first mentioned pathway mediates mitosis, and the second one mediates metabolism and cell growth effect through AKT [20]. After the ligand (IGF-1) binds to its receptor (IGF-1R), PI3K is activated, cell proliferation is promoted by activating the mitogen-activated protein kinase (MAPK) cascade, and apoptosis is blocked by inducing the phosphorylation and the inhibition of proapoptotic proteins such as BAD [21]. The protein IGFBP-2 encoded by the gene of the same name is able to control the biological actions of IGFs [22] and TGFβ [23] in vivo via the endocrine, autocrine or paracrine pathways. The protein TGFβ-3 encoded by the TGFβ3 gene controls the growth, proliferation and differentiation of cells, cell motility and apoptosis. TGFβ-3 plays an essential role in the development of skeletal muscles. It also can suppress the formation of tumors [24]. Adapted from [15,25–27] based on other works: [21,28–53].
The insulin-like growth factor 1 gene (IGF1) has been identified as a biological candidate gene for growth, body composition, metabolic and skeletal characteristics, and is also a positional candidate gene for growth and fat deposition in chicken [54]. This gene is involved in growth of various tissues such as muscle and bones [55]. The chicken IGF1 gene was mapped to 165.95 cM on chromosome 1 (GGA1). In a broiler-layer F2 population used to map body weight (BW) QTL by a genome scan, a QTL affecting BW at 6 weeks of age has been detected at 160 cM (confidence interval (CI) 114 to 180 cM) on chicken GGA1 [56]. In the same F2 cross, a QTL at 150 cM (CI from 100 to 182 cM) on GGA1 affecting abdominal fat weight (AFW) has been ascertained [57].

The bioavailability of the insulin-like growth factors (IGFs) is regulated by a family of structurally conserved insulin-like growth factor binding proteins (IGF-binding proteins; IGFBPs) [58–60]. IGFBPs selectively bind to IGF-1 and IGF-2 proteins but do not bind to insulin [61]. More than 99% of IGF molecules circulate in blood serum as complexes with these specific and high affinity-binding proteins. Although IGF-binding protein 3 (IGFBP-3) is a main component and binds over 75% of the circulating IGF [62], IGFBP-2 is sensitive to dietary protein level, and may play a substantial role in the modulation of the growth-promoting effect of circulating IGF-1 by creating the IGF-1-IGFBP-2 complex in chickens [63]. IGFBP-2 is the predominant IGFBP in serum for different species [64]. IGFs, IGFBPs and IGFBP proteases are the major regulators of somatic growth and cell proliferation [65]. IGFBP-2 controls the biological actions of IGFs [22] and TGFs [23] in vivo via endocrine [22,23], autocrine [23] or paracrine mechanisms [23], and affects the growth and development of animals [66]. IGFBP-2 might indirectly affect adipocyte differentiation by controlling IGF [67] and TGF-β biological actions in fat tissue [68].

The transforming growth factor β (TGF-β) belongs to a large family of multifunctional growth factors [69], with important regulatory roles in embryonic and adult development [70], such as morphogenesis, development and differentiation [69]. Polypeptide growth factors of TGF-β family regulate a number of cellular processes such as cell proliferation, differentiation, migration, adhesion and apoptosis [70]. TGF-β plays a key role in maintaining both bone and articular cartilage homeostasis [71].

2. Materials and Methods

2.1. Experimental Population—Animals

The chicken hatching eggs were produced and the experiment was performed in the testing station of broilers (fattening test No. 1148) at the state-owned enterprise International Testing of Poultry, Ustrasice (Czech Republic). After hatching, 50 chickens from each of the two broiler lines Hubbard F15 and Cobb E were stalled in windowless air-conditioned hall with deep bedding and controlled light mode (Table 1). Stocking density was 6.1 chicks/m². The hall was disinfected with Virkon before the chickens were stored. The chickens were watered by automatic dropper drinking basins and fed with three feed mixtures, differently for particular period of fattening, from tube feeders ad libitum. Hypermangan solution was applied to the water in the first days of age. The composition (contents of main nutrients) in individual complete feed mixtures BR1, BR2 and BR3 for fattening broiler chickens up to the 10th, 35th and 42nd day of age, respectively, are shown in Table 2.

Table 1. Light regime in the chicken house.

| Day  | The Proportion of Light and Darkness Within 24 h |
|------|--------------------------------------------------|
| Light (h) | Dark (h) |
| 1–7 | 23 | 1 |
| 8–37 | 18 | 6 |
| 38 | 20 | 4 |
| 39 | 22 | 2 |
| 40–42 | 23 | 1 |
### Table 2. The content of nutrients in feeding mixtures for broilers (BR1, BR2, BR3) in different periods of the experiment.

| Nutrient          | Feeding Mixture * | 1st–10th Day | 11th–35th Day | 36th–42nd Day |
|-------------------|-------------------|--------------|---------------|---------------|
| Crude protein (%) | BR1               | 22.01        | 19.86         | 18.50         |
|                   | BR2               | 22.01        | 19.86         | 18.50         |
|                   | BR3               | 22.01        | 19.86         | 18.50         |
| Fat (%)           | BR1               | 3.85         | 5.76          | 7.64          |
|                   | BR2               | 3.85         | 5.76          | 7.64          |
|                   | BR3               | 3.85         | 5.76          | 7.64          |
| Lysine (%)        | BR1               | 1.16         | 1.22          | 1.07          |
|                   | BR2               | 1.16         | 1.22          | 1.07          |
|                   | BR3               | 1.16         | 1.22          | 1.07          |
| Methionine (%)    | BR1               | 0.56         | 0.56          | 0.51          |
|                   | BR2               | 0.56         | 0.56          | 0.51          |
|                   | BR3               | 0.56         | 0.56          | 0.51          |
| Calcium (%)       | BR1               | 0.91         | 0.84          | 0.81          |
|                   | BR2               | 0.91         | 0.84          | 0.81          |
|                   | BR3               | 0.91         | 0.84          | 0.81          |
| Phosphorus (%)    | BR1               | 0.60         | 0.54          | 0.47          |
|                   | BR2               | 0.60         | 0.54          | 0.47          |
|                   | BR3               | 0.60         | 0.54          | 0.47          |
| Vitamin A (IU/kg) | BR1               | 15,300       | 12,300        | 10,300        |
|                   | BR2               | 15,300       | 12,300        | 10,300        |
|                   | BR3               | 15,300       | 12,300        | 10,300        |
| Vitamin D3 (IU/kg)| BR1               | 5,000        | 5,000         | 5,000         |
|                   | BR2               | 5,000        | 5,000         | 5,000         |
|                   | BR3               | 5,000        | 5,000         | 5,000         |
| Metabolizable energy (MJ/kg) | BR1 | 12.64    | 13.07        | 13.59        |
|                   | BR2               | 12.64        | 13.07        | 13.59        |
|                   | BR3               | 12.64        | 13.07        | 13.59        |

* The feeding mixtures were produced in ZZN Pelhrimov, a.s., according to given recipes.

2.2. Phenotypic Data

Body weight (BW) was measured at 14, 21, 28, 35 days and before slaughter at 42 days of age. The mortality during experiment was 3% (sudden death syndrome). Chickens were slaughtered at 42 days of age and the slaughter analysis was performed. The carcass traits, such as weight of trunk, giblets, abdominal fat, breast muscle (with and without skin) and thigh muscle (with and without skin), as well as slaughter value and slaughter percentage were investigated. The slaughter value was calculated as the ratio between the weight of the carcass (trunk weight) and the weight at 42 days of age before slaughter, and the slaughter percentage was calculated as the ratio between the sum of weight of the trunk and giblets and the weight at 42 days.

2.3. SNP Genotyping

2.3.1. DNA Extraction

Genomic DNA for genotyping assays was extracted from whole blood samples, which were collected from 97 chickens at 42 days of age before slaughtering. Blood was taken from vena ulnaris into 1.5 mL EDTA-treated microtubes. For extraction of genomic DNA, chelex 100 was used, and the concentration and purity of genomic DNA were verified by spectrophotometer Shimadzu BioSpec-nano (Shimadzu Corporation, Kyoto, Japan).

2.3.2. Optimization of PCR-RFLP Assay

Polymerase chain reaction (PCR) was performed for all assays in total volume 25 µL mixture containing 1 µL genomic DNA, 10 pmol of each primer and 12.5 µL PPP Master Mix (Top-Bio, s.r.o., Vestec, Czech Republic). Sequences of sets of primer pairs for all three gene polymorphisms used in PCR assays are shown in Table 3. The primers for the IGF1 genotyping were designed according to Moody et al. [72]. The thermal profile included pre-denaturation at 94 °C for 2 min followed by 30 cycles 94 °C for 30 s, 67 °C for 30 s and 72 °C for 50 s, with a final extension of 72 °C for 7 min. Thermocycler BIOER Life ECO (Hangzhou Bioer Technology Co., Ltd., Bin An Rd, Hi-tech (Binjiang) District, Hangzhou, China) was used for DNA amplification. SNP of IGF1 gene was detected after digesting PCR product with Hinf1 restriction endonuclease (Fermentas, Vilnius, Lithuania) at 37 °C overnight. For detection of IGFBP2 genotypes PCR amplification was done using primer set by Li et al. [73]. Amplification was performed under following conditions: pre-denaturation at 94 °C for 2 min followed by 30 cycles 94 °C for 30 s, 54 °C for 30 s and 72 °C for 30 s, with a final extension of 72 °C for 7 min. PCR products were digested (restriction fragment length polymorphism–RFLP) with Eco72I restriction endonuclease (New England Biolabs, Ipswich, MA, USA) at 37 °C overnight. The PCR primers designed by Li et al. [69] were applied for TGFβ3. The PCR reaction conditions were the same as for IGFBP2, except the annealing temperature which was 58 °C. Gene fragments
were subjected to digestion by BsmI restriction enzyme (New England Biolabs, Ipswich, MA, USA) at 37 °C overnight.

Table 3. Primers used in polymerase chain reaction (PCR) assay.

| Gene             | Primer Sequence                  | Product Length (bp) | Restriction Enzyme | Restriction Enzyme Production Size (bp) * |
|------------------|----------------------------------|--------------------|-------------------|----------------------------------------|
| IGF1 SNP/A>C     | forward a 5’-CATGGCGGCTACTCCTG-3’ | 813                | HinfI             | AA: 378+244+191                        |
|                  | reverse a 5’-TCAGAAGGACCTACTG-3’  |                    |                   | AC: 378+244+191                        |
|                  |                                   |                    |                   | CC: 622+191                            |
| IGFBP2 intron 2 C1032T | forward b 5’-GTCCCAAGATAAACCTTG-3’ | 367                | Eco72I            | AA: 367                                |
|                  | reverse b 5’-GGTGCAAGGGGTCTG-3’   |                    |                   | AB: 367+265+102                        |
|                  |                                   |                    |                   | BB: 265+102                            |
| TGFβ3 A>C SNP     | forward c 5’-TCAGGGCAGGAGGTG-3’  | 294                | BsmI              | AA: 145+75+74                          |
|                  | reverse c 5’-GCCACTGGCAGATTCT-3’  |                    |                   | AB: 145+125+75+74                      |
|                  |                                   |                    |                   | BB: 125+75+74                          |

* Moody et al. (2003) [72], Zhou et al. (2005) [54]. b Li et al. (2006) [73]. c Li et al. (2003) [69]. All possibilities of fragments.

2.3.3. Electrophoresis

The PCR products were visualized by 2% and restriction patterns by 3% agarose gel electrophoresis and ethidium bromide staining. The ENDURO™ 250 V power supply (Labnet International, Inc., New York, USA) and the HU13 midi horizontal gel electrophoresis unit (Scie-Plas Ltd., Cambourne, Cambridge, UK) were used for DNA electrophoresis. Syngene™ Ingenius 3 Manual Gel Documentation System (Syngene) was used for photo-documentation.

2.4. Statistical Analysis

Associations of three different polymorphisms of IGF1, IGFBP2 and TGFβ3 genes with growth characteristics and carcass data in 97 chicken belonging to two different lines (Hubbard F15 and Cobb E), equally divided between two sexes, were studied. Genotypes were tested for Hardy-Weinberg equilibrium (HWE) using a chi-square ($\chi^2$) test in R/SNPassoc Package (R Development Core Team). Whole-Genome association analyses were performed assuming five different genetic models (inheritance patterns) using R/SNPassoc Package (R Development Core Team): codominant, dominant, recessive, overdominant and log-additive effect. The level of significance was tested at the nominal 5% significance level after correcting for the number of tests performed (Bonferroni correction). Phenotypes were represented by the carcass data collected on poultry, whereas line and sex were included in the model as fixed effects. Hardy-Weinberg equilibrium (HWE) was calculated and tested using $\chi^2$ test at the 0.05 level of statistical significance.

Another statistical analysis was performed using box and forest plots. Average slopes of growth curve and total integrals for the weight sum of the trunk, giblets, abdominal fat, breast and thigh muscles at 42 days of age (at the slaughter of chickens) in both chicken lines, for both sexes and for all genotypes observed, were evaluated using the laboratory information system Qinslab (Prevention Medicals, Studenka, Czech Republic).

3. Results

The IGF1/HinfI PCR-RFLP analysis of 97 DNA samples obtained from chicken belonging to broiler lines Hubbard F15 and Cobb E showed only two from three genotypes, namely AA (378 + 244 + 191 bp), and AC (622 + 378 + 244 + 191 bp), as shown in Figure 2. The AA homozygotes (73.20%) predominated over heterozygotes (26.80%) Table 4, Figure 3. No CC homozygous individuals were detected on either of the two broiler lines. Correspondingly, the frequency of allele A is much higher (86.60%) than allele C (13.40%) in the investigated chicken population, as is evident from Table 4.

In contrast, in IGFBP2/Eco72I polymorphism, all three genotypes (AA, AB, and BB) were found, however, BB (265 + 102 bp) homozygotes showed very low frequency (4.12%). The most represented genotype was AB (367 + 265 + 102 bp) with a frequency of 56.70%.
Also, in TGFβ3/BsI SNP, all three genotypes were detected, with the highest observed genotypic frequency in heterozygotes (42.27%) followed by 36.08% and 21.65% in AA (145 + 75 + 74 bp) and BB (125 + 75 + 74 + 20 bp), respectively.

Only for IGFBP2 frequencies in total population, Hardy-Weinberg equilibrium (HWE) was identified: \( p < 0.01 \).

**Figure 2.** The restriction fragment length polymorphism (RFLP) patterns for IGF1 (AA: 378 + 244 + 191 bp; AC: 622 + 378 + 244 + 191 bp; CC—it was not detected), IGFBP2 (AA: 367 bp; AB: 367 + 265 + 102 bp; BB: 265 + 102 bp) and TGFβ3 (AA: 145 + 75 + 74 bp, AB: 145 + 125 + 75 + 74 bp; BB: 125 + 75 + 74 + 20 bp). Agarose 2%, 120 V, 60 min, Tris-borate-ethylenediaminetetraacetic acid (EDTA) (TBE) buffer. M—marker.

**Table 4.** Genotype and allele frequencies of IGF1, IGFBP2 and TGFβ3 genes in the chicken population.

| Broiler Line | Gene | Genotype Frequencies a | Allele Frequencies a | \( \chi^2 \) Test HWE b (p-Value) |
|--------------|------|------------------------|---------------------|---------------------------------|
| Hubbard F15  | IGF1 | AA: 28 (56.00) AC: 22 (44.00) CC: nf | A: 78 (78.00) C: 22 (22.00) | 0.0908 |
|              | IGFBP2 | AA: 20 (40.00) AB: 27 (54.00) BB: 3 (6.00) | A: 67 (67.00) B: 33 (33.00) | 0.1998 |
|              | TGFβ3 | AA: 15 (30.00) AB: 20 (40.00) BB: 15 (30.00) | A: 50 (50.00) B: 50 (50.00) | 0.1639 |
| Cobb E       | IGF1 | AA: 43 (91.49) AC: 4 (8.51) CC: nf | A: 90 (95.74) C: 4 (4.26) | 1.00 |
|              | IGFBP2 | AA: 18 (38.30) AB: 28 (59.57) BB: 1 (2.13) | A: 64 (68.09) B: 30 (31.91) | 0.0172* |
|              | TGFβ3 | AA: 20 (42.55) AB: 21 (44.68) BB: 6 (12.77) | A: 61 (64.89) B: 33 (35.11) | 1.00 |
| Total        | IGF1 | AA: 71 (73.20) AC: 26 (26.80) CC: 0 (0) | A: 168 (86.60) C: 26 (13.40) | 0.2066 |
|              | IGFBP2 | AA: 38 (39.18) AB: 55 (56.70) BB: 4 (4.12) | A: 131 (67.53) B: 63 (32.47) | 0.0050* |
|              | TGFβ3 | AA: 35 (36.08) AB: 41 (42.27) BB: 21 (21.65) | A: 111 (57.22) B: 83 (42.78) | 0.2125 |

\( a \) The numbers in brackets are percentage frequencies (relative frequencies). \( b \) HWE—Hardy-Weinberg equilibrium; \( * \) statistically significant \( p < 0.05 \); nf—not found.
Tables 5–7 show average values of growth performance and carcass traits in both chicken lines, according to individual genotypes. The highest average BW at 42 days was achieved in the Cobb E line, with the AC genotype of IGF1 (2967.50 g), BB genotype of IGFBP2 (3170.00 g) and the AB genotype of TGFβ3 (3104.29 g). The highest average breast muscle (without skin) was found in a Cobb E chicken with an AC genotype of IGF1 (629.25 g), a BB genotype of IGFBP2 (753.00 g) and an AB genotype of TGFβ3 (653.62 g). The Cobb E chicken with an AC genotype of IGF1 (52.50 g) and a BB genotype of IGFBP2 (63.00 g) also had the highest average abdominal fat weight (AFW); whereas for TGFβ3, the highest AFW was found in the AA genotype (54.95 g). The highest average thigh muscle (with skin) were measured in a Cobb E line chicken with an AC genotype of IGF1 (519.75 g), a BB genotype of IGFBP2 (552.00 g) and an AB genotype of TGFβ3 (538.72 g). On the contrary, the lowest average values of BW at 42 days and AFW showed in the Hubbard F15 line chicken with an AA genotype of IGF1 (2585.00 and 35.32 g, respectively), a BB genotype of IGFBP2 (2413.33 g and 30.33 g, respectively) and an AA genotype of TGFβ3 (2541.33 g and 31.53 g). The lowest breast muscle (without skin) showed the Hubbard F15 line chicken with an AA genotype of IGF1 (501.75 g), an AA genotype of IGFBP2 (498.75 g) and an AA genotype of TGFβ3 (494.33 g). The lowest thigh muscle (with skin) was observed in a Hubbard F15 line chicken with an AA genotype of IGF1 (470.68 g) and a BB genotype of IGFBP2 (445.00 g) and in a Cobb E line chicken with a BB genotype of TGFβ3 (458.67 g).

| Parameter                  | Hubbard F15 |                       |                        |                       |                       |
|----------------------------|-------------|-----------------------|-----------------------|-----------------------|-----------------------|
|                            | Mean ± SD   | CI                    | Mean ± SD             | CI                    | Mean ± SD             |
| BW at 14 days *            | 421.07 ± 32 | 433, 408              | 428.18 ± 31           | 442, 414              | -                     |
|                            | 33          | 39                    | 33                    | 39                    | -                     |
| BW at 21 days *            | 840.00 ± 69 | 866, 813              | 865.00 ± 67           | 894, 835              | -                     |
|                            | 33          | 39                    | 33                    | 39                    | -                     |
| BW at 28 days *            | 1393.21 ± 127 | 1446, 1340         | 1436.82 ± 114         | 1487, 1386            | -                     |
|                            | 33          | 39                    | 33                    | 39                    | -                     |
| BW at 35 days *            | 1913.21 ± 183 | 1984, 1842          | 1947.27 ± 161         | 2018, 1876            | -                     |
|                            | 33          | 39                    | 33                    | 39                    | -                     |
| BW at 42 days before slaughter * | 2585.00 ± 298 | 2700, 2469          | 2588.64 ± 265         | 2706, 2471            | -                     |
|                            | 33          | 39                    | 33                    | 39                    | -                     |
### Table 5. Cont.

| Parameter | Hubbard F15 |  |  |  |  | Cobb E |  |  |  |  |
|-----------|-------------|---|---|---|---|---------|---|---|---|---|
|           | Genotype | Genotype | Genotype | Genotype | Genotype | Genotype | Genotype | Genotype | Genotype | Genotype |
|           | AA (n = 20) | AB (n = 22) | BB (n = 22) | AA (n = 20) | AB (n = 22) | BB (n = 22) | AA (n = 20) | AB (n = 22) | BB (n = 22) | AA (n = 20) | AB (n = 22) | BB (n = 22) |
| Mean ± SD/CI |  |  |  |  |  |  |  |  |  |  |  |

- **Trunk weight** *: 1798.00 ± 190 1871.724 1901.1754 - 1989.60 ± 224 2058.192 2520.106 -
- **Giblets weight** *: 156.79 ± 18 164.55 ± 14 168.155 - 164.19 ± 16 174.163 199.132 -
- **Abdominal fat weight** *: 35.32 ± 0.9 35.68 ± 13 41.30 - 51.47 ± 12 55.47 80.25 -
- **Breast muscle weight with skin** *: 542.18 ± 53 558.00 ± 64 586.529 - 668.81 ± 84 694.643 853.515 -
- **Breast muscle weight without skin** *: 501.75 ± 50 517.09 ± 61 544.499 - 615.98 ± 83 641.590 712.530 -
- **Thigh muscle weight with skin** *: 470.68 ± 61 475.14 ± 55 499.450 - 499.68 ± 60 519.584 669.371 -
- **Thigh muscle weight without skin** *: 421.39 ± 55 426.77 ± 53 450.402 - 441.88 ± 60 460.423 608.320 -
- **Slaughter value** **: 69.66 - 70.22 - 68.14 - 69.42 -
- **Slaughter percentage***: 75.74 76.48 - 73.94 75.01 -

BW–average body weight; SD–standard deviation; CI–confidence interval (95%); * (g); ** slaughter value = weight of trunk/BW at 42 days * 100 (%); *** slaughter percentage = weight of trunk + weight of giblets/BW at 42 days * 100 (%).

### Table 6. The average growth performance and carcass traits in the chicken population according to IGFBP2 genotypes.

| Parameter | Hubbard F15 |  |  |  |  | Cobb E |  |  |  |  |
|-----------|-------------|---|---|---|---|---------|---|---|---|---|
|           | Genotype | Genotype | Genotype | Genotype | Genotype | Genotype | Genotype | Genotype | Genotype | Genotype |
|           | AA (20) | AB (27) | BB (3) | AA (17) | AB (27) | BB (1) | AA (20) | AB (27) | BB (1) | AA (20) | AB (27) | BB (1) |
| Mean ± SD/CI |  |  |  |  |  |  |  |  |  |  |  |  |

- **BW at 14 days** *: 419.50 ± 33 427.78 ± 32 423.33 ± 15 461.385 489.449 492.458 474.814 ± 39 492.458 460.00
- **BW at 21 days** *: 839.50 ± 72 864.07 ± 66 890.838 965.655 976.879 973.839 927.695 ± 95 932.22 ± 103 1010.00
- **BW at 28 days** *: 1392.50 ± 126 1436.67 ± 128 1487.1985 1552.1101 1630.1441 1628.1482 1535.29 ± 184 1554.81 ± 184 1750.00
- **BW at 35 days** *: 1894.00 ± 179 1967.78 ± 164 2032.1902 2110.1489 2216.1972 2199.1956 2096.47 ± 202 2097.41 ± 256 2300.00
- **BW at 42 days before slaughter** *: 2578.50 ± 320 2611.85 ± 257 2713.2510 3009.1818 3069.2764 3036.2789 2891.76 ± 344 2912.59 ± 311 3170.00
- **Trunk weight** *: 1791.20 ± 206 1827.41 ± 178 1898.1757 1958.06 ± 226 2074.1842 2086.1906 1996.22 ± 228 2251.00
- **Giblets weight** *: 159.45 ± 19 168.151 165.154 149.33 ± 18 194.105 177.156 166.53 ± 20 177.156 194.00
- **Abdominal fat weight** *: 35.35 ± 11 40.32 30.33 ± 18 75.14 ± 14 57.44 56.46 50.59 ± 13 51.26 ± 12 63.00
- **Breast muscle weight with skin** *: 539.65 ± 64 578.533 552.00 ± 33 634.470 689.613 712.641 651.24 ± 74 767.11 ± 90 809.00
- **Breast muscle weight without skin** *: 498.75 ± 60 515.59 ± 53 600.420 598.82 ± 72 636.562 658.587 622.37 ± 89 672.37 ± 89 753.00
- **Thigh muscle weight with skin** *: 471.80 ± 63 476.33 ± 56 450.5753 445.05 ± 52 488.06 ± 65 503.33 ± 61 454.56 ± 62 527.479 552.00
- **Thigh muscle weight without skin** *: 421.25 ± 59 426.74 ± 52 400.33 ± 49 522.278 464.399 470.421 431.24 ± 63 445.56 ± 62 487.00
- **Slaughter value** **: 69.57 69.99 71.36 67.77 68.51 71.01
- **Slaughter percentage***: 75.78 76.12 77.54 73.53 74.31 77.13

BW–average body weight; SD–standard deviation; CI–confidence interval (95%); * (g); ** slaughter value = weight of trunk/BW at 42 days * 100 (%); *** slaughter percentage = weight of trunk + weight of giblets/BW at 42 days * 100 (%).
which was not significant for the dominant inheritance pattern. Animals 2020 10, 800

Table 7. The average growth performance and carcass traits in the chicken population (according to TGFβ3 genotypes).

| Parameter | Hubbard F15 | Cobb E |
|-----------|-------------|--------|
| Genotype | Genotype |
| AA (15) | AB (20) | BB (15) | AA (20) | AB (21) | BB (6) | Mean ± SD; CI |
| BW at 14 days * | 420 ± 40 | 433.00 ± 24 | 416.00 ± 30 | 462.00 ± 38 | 484.76 ± 41 | 463.33 ± 34 |
| BW at 21 days * | 842.67 ± 72 | 866.00 ± 65 | 839.33 ± 72 | 888.50 ± 93 | 980.00 ± 91 | 923.33 ± 50 |
| BW at 28 days * | 1399.33 ± 122 | 1420.50 ± 117 | 1414.67 ± 154 | 1453.00 ± 167 | 1664.76 ± 149 | 1536.67 ± 125 |
| BW at 35 days * | 1884.67 ± 171 | 1938.00 ± 165 | 1958.67 ± 187 | 1986.00 ± 240 | 2241.91 ± 190 | 2046.67 ± 176 |
| BW at 42 days before slaughter * | 2541.33 ± 255 | 2597.00 ± 302 | 2618.00 ± 292 | 2783.50 ± 312 | 3104.29 ± 270 | 2780.00 ± 208 |
| trunk weight * | 1773.27 ± 175 | 1825.40 ± 209 | 1814.73 ± 177 | 1896.95 ± 223 | 2129.19 ± 181 | 1857.00 ± 136 |
| gullets weight * | 159.00 ± 15 | 167.33 ± 19 | 159.60 ± 15 | 161.45 ± 19 | 177.43 ± 16 | 163.33 ± 14 |
| abdominal fat weight * | 31.53 ± 7 | 37.05 ± 13 | 37.33 ± 10 | 54.95 ± 14 | 48.90 ± 9 | 49.50 ± 15 |
| breast muscle with skin * | 533.67 ± 99 | 555.15 ± 68 | 556.60 ± 41 | 648.30 ± 86 | 710.48 ± 72 | 601.83 ± 51 |
| breast muscle without skin * | 494.33 ± 54 | 515.55 ± 66 | 513.27 ± 39 | 598.60 ± 86 | 653.62 ± 73 | 551.00 ± 52 |
| thigh muscle with skin * | 499.00 ± 63 | 476.75 ± 63 | 470.80 ± 49 | 475.65 ± 61 | 538.72 ± 51 | 458.67 ± 30 |
| thigh muscle without skin * | 504.43 | 506.44 | 498.44 | 504.44 | 562.51 | 490.42 |
| slaughter value ** | 69.80 | 70.34 | 69.44 | 68.26 | 68.62 | 66.93 |
| slaughter percentage *** | 76.08 | 76.43 | 75.57 | 74.08 | 74.34 | 72.81 |

Table parameters: BW—average body weight; SD—standard deviation; CI—confidence interval (95%); * (g); ** slaughter value = weight of trunk/BW at 42 days * 100 (%); *** slaughter percentage = weight of trunk + weight of gullets/BW at 42 days * 100 (%).

By means of statistical software analysis, the relationships between SNPs and individual traits were identified. The fixed effects included in the “whole” model were sex and line, and the p-values obtained were adjusted by the number of tests using Bonferroni correction.

Only the TGFβ3 SNP (Table 8) resulted in statistical significance for the following parameters: body weight at 21, 28 and 35 days, trunk weight and slaughter value. The p values were significant for codominant, dominant and overdominant genetic models, with the exception of the slaughter value, which was not significant for the dominant inheritance pattern.

Figure 4 shows average slopes of growth curve (14–42 days of age) in individual genotypes of both chicken line and sexes.

Figure 5 represents total integrals for the sum of the trunk, gullets, abdominal fat, breast and thigh muscles at 42 days of age in all genotypes observed.
Table 8. Results of statistical analysis for testing association between TGFβ3 polymorphism and growth performance and carcass traits in the chicken population.

| Parameter               | Codominant | Dominant | Recessive | Overdominant | Log-Additive |
|-------------------------|------------|----------|-----------|--------------|--------------|
| BW at 14 days           | ns         | ns       | ns        | ns           | ns           |
| BW at 21 days           | -0.013     | 0.014    | 0.606     | 0.005        | 0.199        |
| BW at 28 days           | -0.004     | 0.002    | 0.847     | 0.004        | 0.035        |
| BW at 35 days           | -0.004     | 0.001    | 0.560     | 0.008        | 0.016        |
| BW at 42 days before slaughter | ns         | ns       | ns        | ns           | ns           |
| trunk weight            | -0.010     | 0.013    | 0.535     | 0.003        | 0.215        |
| giblets weight          | ns         | ns       | ns        | ns           | ns           |
| abdominal fat weight    | ns         | ns       | ns        | ns           | ns           |
| breast muscle with skin | ns         | ns       | ns        | ns           | ns           |
| breast muscle without skin | ns       | ns       | ns        | ns           | ns           |
| thigh muscle with skin  | ns         | ns       | ns        | ns           | ns           |
| thigh muscle without skin | ns       | ns       | ns        | ns           | ns           |
| slaughter value         | -0.015     | 0.646    | 0.015     | 0.014        | 0.306        |
| slaughter percentage    | ns         | ns       | ns        | ns           | ns           |

BW–body weight; ns–no significant SNP after Bonferroni correction. Statistical significances at significance level 0.05 are highlighted in bold.

Figure 4. Forest plot of body weight (g) of chicken (at 14–42 days of age) of both lines in individual genotypes. (A) Males; (B) females; (C) total. Statistical characteristics of individual variants: average (n), median, standard deviation (SD), p-value (compared to average value), minimum (min) and maximum (max) value. Symbol H is Hubbard F15; C is Cobb E. The line segments represent confidence interval–CI (95%).
**Figure 5.** The weight sum of the trunk, giblets, abdominal fat, breast muscle with and without skin and thigh muscle with and without skin. IGF1 gene with genotype AA and AC (A), IGFBP2 with genotype AA and AB (B), TGFβ3 with genotype AA, AB and BB (C). The pink symbol—median, the grey symbol—mean, square indicates the weight (number of samples). A comparison of individual genotypes (D). Statistical characteristics of individual variants: number (n), mean and median. The line segments represent confidence interval–CI (95%); m—males, f—females.

### 4. Discussion

The study of candidate genes is one of the primary methods to determine whether specific genes are related to economically important traits in farm animals [69]. We performed genotyping of the SNP of three genes linked to consumer-priced characteristics in chicken meat.

One of the main hormones required to normal growth process and muscle development is insulin-like growth factor 1 (IGF-1) [74]. The chicken IGF1 gene consists of four exons and three introns, spanning more than 50 kb on chromosome 1 [75].

IGF1 encodes the same-name protein (IGF-1), which has a similar molecular structure to insulin [76] and induces insulin-like metabolic effects in muscle and adipose tissues [65]. This protein plays an important role in the proliferation, differentiation and metabolism of myogenic cell lines in chickens [76]. IGF-1 is one of the three ligands (insulin, IGF-1, IGF-2) belonging to the IGF system, which also includes three cell surface binding receptors (InsR, IGF-1R, IGF-2R), and insulin-like growth factor binding proteins (IGF binding proteins, IGFBPs) and IGFBP protease [77]. In addition, the IGF-1 protein is a potent mitogen and an essential stimulus for the differentiation of adipocytes [78]. The production and secretion of IGF-1 is affected by age, nutritional status, and several hormones [79]. The predominant source of IGF-1 is the liver and some other tissues, including muscle, brain and kidney [80].

IGF-1 binds to the type 1 insulin-like growth factor receptor (IGF-1R), which plays a critical role in signaling cell survival and proliferation [21]. However, IGF-1 can also bind, albeit with lower affinity, to the insulin receptor [16], regulating some metabolic functions [25].

Insulin-like growth factors (IGFs) provide essential signals for the control of embryonic, as well as postnatal development in vertebrates [81]. In addition to the growth hormone (GH), IGF-1 is one of the two main hormones required to support normal growth in chicken. Optimal growth requires a “set-point” concentration of both IGF-1 and triiodothyronine (T3) in blood circulation. Pituitary GH plays a role in controlling the circulating levels of both IGF-1 and T3 [74]. IGFs stimulate hepatic glycogen, increase DNA synthesis and promote tissue growth in chicken [82]. The highest level of IGF1 mRNA expression was detected in the chicken liver. High levels of IGF1 mRNA (10%–30% of the value in the liver) were expressed in spleen, lung and brain of chickens. IGF1 mRNA expression was
also observed in other extrahepatic tissues such as the kidney, heart, intestine, thymus and muscle of chickens, but these expression levels were less than 4% of that in the liver [83].

The abundant expression of IGF1 gene was detected in the liver of normal chicken, but no IGF1 mRNA expression was found in that organ of dwarf chicken [83]. The expression of hepatic IGF1 mRNA level and circulating IGF-1 concentration were significantly higher in chicken with a high growth rate, compared to the line with low growth rate, supporting the hypothesis of its stimulatory effect during post-hatching growth of chickens as stated by Beccavin et al. [84]. The liver is the main site of IGF-1 production during post-hatch growing stages of chicken as described by Kita et al. [85].

IGF-1 is significantly altered by the genotype, suggesting a pivotal role in the control of growth rate in broiler chickens [84].

The SNPs within the chicken IGF1 promoter were reported by numerous previous studies [54,86–92] but according to the author’s best knowledge, no research work on gene constitution of IGF1 SNP in the Hubbard F15 and Cobb E lines has been reported until now.

Genotype frequency analysis indicated that the AA genotype (73.20%) was of higher frequency than the AC (26.80%) and CC (0%) genotypes in both chicken lines, which is consistent with another study [92]. For the other two genes (IGFBP2, TGFβ3), the predominance of heterozygous (AB) genotypes was detected. Interestingly, in Hubbard F15, a distribution of both allele in TGFβ3 was identical (Table 4).

With regards to the genotype frequencies of IGF1/Hinf1 gene polymorphism, from the three known restriction patterns, only two genotypes were detected: AA and AC, with an almost three times higher prevalence of AA homozygotes over heterozygotes. The CC homozygous genotype was not found in either chicken line, which is consistent with the finding of Moe et al. [90], which reported an absence of the CC genotype in two commercial broiler strains (Chunky and Cobb). The noticeable predominance of allele A (86.60%) over allele C (13.40%) (Table 4) observed in our study is in conspicuous accordance with previous studies. As Moe et al. [90] have shown, allele C occurs especially in native chickens, for example in nine Japanese native chicken breeds (Chabo, Ehime-jidori, Gifu-jidori, Koeyoshi, Koshamo, Mikawa, Satsuma-dori, Engie and Tokuchijidori), the frequency of this allele to be 1.0. Our finding of low incidence of C allele in broiler chicken also corresponds with the results of genotyping performed by Anh et al. [4], who observed the CC genotype of IGF1 gene with very low frequencies (0.13 to 0.15) in all populations of crossbreds from commercial parent stock broilers with four Thai synthetic chicken lines (the Kaen Thong, Khai Mook Esarn, Soi Nin, and Soi Pet). The IGF1 SNP gene constitution of these four Thai synthetic chicken lines was studied by Promwatee et al. [92], who found that the AA genotype had a considerably lower frequency than the AC and CC genotypes in all chicken lines except Soi Noi, in which the AA and CC genotypes were similar. With the exception of Soi Nin, there was the predominance of the C allele—the frequency of the A allele was lower than that of C in all lines except Soi Noi, where both allelic frequencies were the same. This indicates that allele C is evidently typical for native breeds. A higher frequency of allele A than that of allele C in commercial broiler stocks compared to native chicken can be explained as a result of selection effect on growth traits [90].

It can be concluded that the incidence of a higher A allele frequency over C allele in the IGF1 locus observed in our study could be a result of a long-term selection strategy applied in the populations of chosen broiler lines that are the subject of this study.

Various studies reported associations between IGF1 polymorphism and growth traits in chickens. Zhou et al. [54] and Amills et al. [86] reported that polymorphism of the IGF1 gene in the promoter and 5′-untranslated region (5′-UTR) was directly associated with chicken growth rate. Bian et al. [89] found that haplotypes based on three IGF1 polymorphisms (c.*366A > C, c.528G > A and c.*1024C > T—in 5′-flanking, exon 3 and 3′-flanking regions of IGF1) were associated with BW traits.

In our study, AC genotype of IGF1 evinced the highest average BW at 42 days in both chicken lines. This genotype also corresponded with a higher average AFW, breast muscle weight (with or without skin), thigh muscle (with or without skin), slaughter value and slaughter percentage in both lines. On the contrary, the AA genotype of IGF1 was associated with the average lowest BW at 42 days,
trunk weight, AFW, breast and thigh muscles, slaughter value and slaughter percentage in both lines. However, no significant difference was identified.

These results are inconsistent with the study of Zhou et al. [54], which observed that broiler line with fragment sizes of 378, 244 and 191 bp (AA genotype) showed greater improvement of marketable BW. Additionally, in Thai native chickens, the AA genotype resulted in a higher BW compared to the AC and CC genotypes [93]. Promwatee et al. [92] found, in two synthetic lines (Khai Mook Esarn, Soi Pet), the association between the AA genotype and BW at 8 and 12 weeks of age and average daily gain (ADG) at 0–12 and 0–14 weeks. On the contrary, in the Soi Nin synthetic line, BW at 8 and 12 weeks and ADG at 0–12 weeks were associated with the AC genotype. In the fourth synthetic line (Kaen Thong), no significant association was found [92]. In Thai native chickens (Chee), the IGF1 gene was significantly associated with BW at 12 and 16 weeks of age, and ADG during 0–12 and 0–16 weeks of age [94].

IGFBP-2 binds to insulin-like growth factors [64]. IGFBP-2 is the predominant binding protein produced during adipogenesis of white preadipocytes [95]. IGFBP-2 is secreted by white adipocytes and contributes to the prevention of diet-induced obesity [96]. The circulating IGFBP-2 level was significantly and negatively correlated with fasting plasma glucose, triglycerides, low-density lipoprotein (LDL) cholesterol, IGF-1, IGF-2 and insulin C-peptide [97].

IGFBP-2 regulates a broad spectrum of physiological processes involved in growth, development, and differentiation [73]. Both inhibitory and stimulatory effects of IGFBP-2 on cell proliferation have been reported [98]. IGFBP-2 plays an important role in growth and fat metabolism [64]. IGFBP-2 is the predominant IGF binding protein produced during adipogenesis, and is known to increase the insulin-stimulated glucose uptake in myotubes [99]. IGFBP-2 stimulates glucose uptake in a phosphatidylinositol-3-OH kinase (PI3K)-dependent manner. Adipocytes treated with insulin and IGF-1 for 30 min exhibited a significant ($p < 0.001$) increase in PI3K phosphorylation when compared with the control cells. Similarly, IGFBP-2 induced a significant increase in PI3K phosphorylation in 3T3-L1 adipocytes treated for either 30 min ($p < 0.01$) or 24 h ($p < 0.001$). Similarly, IGFBP-2 induced a noticeable increase in AKT phosphorylation in 3T3-L1 adipocytes treated for either 30 min ($p < 0.05$) or 24 h ($p < 0.01$) [99]. IGF-1 significantly ($p < 0.001$) increased, whereas insulin failed to induce ($p > 0.05$) AMP-activated protein kinase (AMPK) phosphorylation in 3T3-L1 adipocytes. Similarly, the treatment of adipocytes with IGFBP-2 for either 30 min or 24 h induced a significant ($p < 0.001$) increase in AMPK phosphorylation [99].

Among the seven IGFBPs, IGFBP-2 is the main binding protein secreted by differentiating white preadipocytes, indicating a potential role in the development of obesity. Overexpression of IGFBP-2 was associated with decreased susceptibility to obesity and improved insulin sensitivity [78]. IGFBP-2 expression was associated with fat mass percentage ($p < 0.02$). It was demonstrated that IGFBP-2 is expressed by subcutaneous abdominal adipocytes of obese individuals and that the expression elevated with increasing adiposity and reducing insulin sensitivity [100].

The main functions of IGFBPs are: (1) acting as carrier proteins for circulating IGF-1 and controller of its flow from the vascular space to tissues; (2) increasing IGF-1 half-life and regulating its metabolic clearance [101]; (3) modulating the interaction between IGF-1 and its receptor, and thus indirectly controlling IGF-1 biological activity [102]; (4) modulating IGF-1 in target tissues, inhibiting or activating its specific actions: cell proliferation, differentiation, survival and migration [62,103–105]; and (5) providing a specific localization pool of IGF-1, because IGFBPs can associate with cell membranes or the extracellular matrix (ECM) [106]. Moreover, some IGFBPs can possess some biological effects outside the IGF-1 signaling pathways, such as apoptosis induction and proliferation/inhibition in some tumors [105].

The IGFBP2 gene has a total length of 32 kb and it is composed by four exons, 2.0 kb (rat) and 1.6 kb (human) mRNAs are generated, and the mature protein is approximately 31 kDa and 36 kDa in rats and humans, respectively [107]. The chicken IGFBP2 gene spans to more than 38 kb on chromosome 7 (GGA7), consists of four exons, and presents similar organization compared with rats and humans.
The chicken IGFBP2 gene is expressed in a majority of tissues, such as liver, muscle, kidney, heart, ovary, brain, intestine and other tissues [108]. IGFBP2 gene expression was downregulated in the visceral white adipose tissue of mice, and its circulating levels were reduced in obese mice [96]. Eckstein et al. [109] reported that IGFBP-2 level negatively affected bone size and mineral content in mice, suggesting it was an important regulator of bone biology in vivo.

As for another gene necessary for growth and development processes, the analysis of IGFBP2/Eco72I gene polymorphism in the present study showed all three known genotypes (AA, AB, and BB), with an obvious predominance of the heterozygous genotype (56.70%). However, BB (265 + 102 bp) homozygotes showed very low frequency (4.12%). The most represented genotype AB (367 + 265 + 102 bp) had a similar frequency to the heterozygotes (53.21%) detected by Li et al. [73], who found almost identical frequencies of both homozygotes (AA 22.96%, BB 23.83%).

The predominance of A allele over B allele in IGFBP2 locus in our study may be, similarly to IGF1 locus, a long-term selection strategy employed in these chicken populations.

The study of Li et al. [73] indicated that chicken IGFBP2 gene intron 2 C1032T (accession number AY 326194) polymorphism was associated with growth and body composition traits in an F2 population. Moreover, the IGFBP2 gene was found to be highly expressed in abdominal fat [73]. QTL for fat deposition was mapped between the marker brackets LEI0064 and ROS0019 (75 kb to 27 Mb) on GGA7 in the chicken linkage map [57], which covers the chicken IGFBP2 gene (23 to 24 Mb). In Thai native chickens (Chee), the IGFBP2 gene was significantly associated with body weight at 4 weeks of age, ADG during 0–4 weeks of age and breast width at 16 weeks of age [94].

An excessive abdominal fat in chickens is undesirable and is therefore sought to be reduced, in order to improve the quality of the final product. The IGFBP-2 could inhibit the biological actions of IGF in vivo via endocrine or paracrine mechanisms [22] and indirectly control adipocyte differentiation by regulating the actions of IGF [67]. The structure and function of the IGFBP2 gene has been analyzed in detail, however, the association of this gene with growth features in chickens has been little studied [66].

In our study, heterozygous genotype AB of IGFBP2 resulted—in both chicken lines—in a higher average BW at 42 days, trunk weight, AFW, breast and thigh muscles, slaughter value and slaughter percentage compared with AA genotype (Table 6). On average, chickens with the IGFBP2-BB genotype grew slower and simultaneously deposited less fat in the body. These differences, however, were not statistically significant. The lowest breast muscle (without skin) was observed in the Hubbard F15 line chicken with an AA genotype of IGFBP2 (498.75 g).

The findings of higher BW and AFW in heterozygotes in our research are not consistent with the findings of the study of Li et al. [73], which found that F2 chicken homozygous for the B allele (IGFBP2-BB) had a higher AFW than birds of the other two genotypes.

The results point to the potential identification of IGFBP2 as a candidate gene for altering the growth rate and abdominal fat [73]. Reduced growth was associated with increased hepatic IGFBP2 mRNA expression and elevated serum IGFBP-2 levels [22], further suggesting IGFBP-2 as a negative growth regulator in vivo [73].

TGF-βs are represented in birds and mammals by three isoforms of secreted cytokines TGF-β1, TGF-β2 and TGF-β3 [70]. The research of Li et al. [69] supported the broad effects of TGFβ genes on the growth and development of chickens. Recently, eight from 17 polymorphic sites of the TGFβ3 gene [53 (T → C), 1653 (C → T), 1755 (A → G), 3343 (C → T), 3540 (C → T), 4786 (C → T), 7263 (C → T) and 7471 (G → A)] have been significantly related to reproduction traits, indicating these polymorphic sites as potential assistant selection markers for improvement of reproductive capacity of Liboyaoshan chicken [110].

The TGFβ3 gene could be a marker for genetically improving duration fertility in hens. In the recent study performed by Gu et al. [111], four SNPs were identified in intron 1 of TGFβ3, and were significantly associated with the duration of fertility in hens (p < 0.05). In addition, they identified multi-copy and copy number variants (CNVs) in chicken TGFβ3, and later determined significant
associations between TGFβ3 CNVs and duration fertility in hens. Specifically, the TGFβ3 copy number exhibited a significant positive correlation with its expression (p < 0.05).

A significant association between the TGFβ3-βsrI polymorphism and mortality between 14 and 42 days in broiler chickens was reported by Ye et al. [112].

In addition, significant effects of TGFβ3-βsrI polymorphism on the cecum content Salmonella enteritidis bacterial load were found [113], which could have been of great importance, especially in commercial broiler chicken farms. A moderate association (p < 0.17) was found between the TGFβ3-βsrI sire allele and antibody response to the S. enteritidis vaccine [113]. Polymorphism in the restriction site of TGFβ3-βsrI was associated with S. enteritidis burden. The heterozygote AC had the highest S. enteritidis burden in the cecum, spleen and liver compared with the other two genotypes (p < 0.01). The GC genotype of TGFβ3 showed the lowest bacterial burden for Village Chickens, whereas in Red Junglefowl, the A/A genotype exhibited the lowest S. enteritidis colonization [114]. Salmonella enterica serovar Enteritidis infection is a common concern in poultry production for its negative effects on growth, as well as food safety for humans [114].

In the study of Li et al. [69], the TGFβ3 polymorphism in broilers crossed with Leghorn was associated with traits of growth and body composition, such as BW, ADG, breast muscle weight, abdominal fat and spleen weight. In our study, for the TGFβ3 gene, AB genotype was the most common in both chicken lines. The allele A was identified as a dominant allele in Cobb E (64.89%), whereas in Hubbard F15, the frequency of both alleles was identical. This finding is different from another study analyzing TGFβ3 genotypes in breeder hens [115], where the allele B was a dominant allele at TGFβ3 locus, due to it having the highest frequency (0.81).

For the TGFβ3 gene, different tendencies were observed in the association of the A and B alleles with the traits observed within both chicken lines. In Cobb E, the AB genotype of TGFβ3 resulted in the highest average BW at 42 days (3104.29 g). The highest AFW had Cobb E chicken with AA genotype of TGFβ3 (54.95 g). On the contrary, in Hubbard F15, the highest average BW at 42 days and AFW were observed in chickens with the BB genotype (2618.00 g, 37.33 g, respectively).

The highest average breast muscle (without skin) was found in the AB genotype of TGFβ3 in both lines (in Cobb E chicken: 653.62 g and in Hubbard F15: 515.55 g). The highest average thigh muscle (with skin) was also found in the AB genotype of TGFβ3 in both lines, with the highest average value in Cobb E (Cobb E: 538.72 g, Hubbard F15: 476.75 g).

On the contrary, the lowest average value of BW at 42 days was observed in the AA genotype of TGFβ3 in both lines (in the Hubbard F15 line chicken with the AA genotype: 2541.33 g and in Cobb E in individuals with the AA genotype: 2778.50 g). The lowest AFW was found in the chicken with the AA genotype in the Hubbard F15 line (31.53 g), and in Cobb E in birds with the heterozygous genotype AB (48.90 g). The lowest breast muscle (without skin) was in the Hubbard F15 line chicken with the AA genotype of TGFβ3 (494.33 g) and in Cobb E in the birds with the BB genotype (551.00 g). The lowest thigh muscle (with skin) was observed in the Hubbard F15 line chicken with the AA genotype (469.00 g), and in the Cobb E line chicken with the BB genotype of TGFβ3 (458.67 g).

Association analysis showed that BsrI genotypes of TGFβ3 are related to some performance traits (Table 8). The statistical analysis revealed a significant association of TGFβ3 with BW at 21, 28 and 35 days and trunk weight in the codominant (negative value), dominant and overdominant (positive values) genetic model, and with slaughter value in codominant (negative value), recessive and overdominant (positive values) genetic model.

The average slopes of the growth curve (14–42 days of age) constructed according to the line, genotype and sex (Figure 4) was confirmed to be a result of linear growth, as well as lineage and sex differences in body weight. Constructing the graphs of total integrals for the weight sum of the trunk, giblets, abdominal fat, breast and thigh muscles at 42 days of age (at the slaughter of chickens) in both chicken lines (with a separate evaluation of both sexes) and all genotypes observed (Figure 5) showed interesting tendencies. However, no statistically significant dependence was observed, despite apparent differences.
5. Conclusions

The presented study demonstrated that the point mutation can affect chicken growth, and confirmed some significant associations between SNP and growth traits. Based on these findings, it can be concluded that the TGFβ3 gene could be applied as a candidate gene marker for chicken growth traits in the Hubbard F15 and Cobb E broiler line population selection program. However, further association analysis will be required to clarify the effects of this marker on growth and production traits in the broiler chicken population.

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Abbreviations

ADG average daily gain
AFW abdominal fat weight
AKT serine/threonine kinase
AMP adenosine monophosphate
AMPK AMP-activated protein kinase
BACs bacterial artificial chromosomes
BAD BCL-2 antagonist of cell death
Bcl-2 antiapoptotic B-cell lymphoma/leukemia-2
bp base pairs
BW body weight
CI confidence interval
CNVs copy number variants
cM centimorgan
DNA deoxyribonucleic acid
ECM extracellular matrix
EDTA ethylenediaminetetraacetic acid
ERK extracellular signal-regulated kinase
FOXO Forkhead box, class O; subfamily of Forkhead transcription factors
Gb giga base pairs
GGA1, GGA7 Gallus gallus autosome 1, Gallus gallus autosome 7
GH growth hormone
GSK-3β glycogen synthase kinase-3β
HWE Hardy-Weinberg equilibrium
IGF insulin-like growth factor
IGF-1 insulin-like growth factor 1
IGF1 insulin-like growth factor 1 gene
IGF-1R the type 1 insulin-like growth factor receptor
IGF-2R the type 2 insulin-like growth factor receptor
IGFBP, IGFBP-2, IGFBP-3, IGFBP2, InsR, IRS, IU, kb, LDL, MAP, MAPK, MEK, MMAS, mRNA, mTOR, mTOR-C1, mTOR-C2, p27, PI3K, PCR, PDK1, QTL, RAF, RAS, RFLP, S6K-1/2, SD, SHC, SMADs, SRC, TGFβ, TGFβ3, TSC-1/2, 4E-BP1

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