INTRODUCTION

Freshwater ecosystems are important components of the global carbon (C) cycle (Cole et al., 2007; Tranvik et al., 2009). They have a significant effect on the atmospheric fluxes of the greenhouse gases (GHGs) carbon dioxide (CO₂) and methane (CH₄; Bastviken, Tranvik, Downing, Crill, & Enrich-Prast, 2011; Raymond et al., 2013) and also bury C in their sediments, which removes C from the active C cycle (Mendonça et al., 2017). The overall contribution to atmospheric GHG concentrations (Prairie et al., 2018) can be quantified using the CO₂-equivalent balance, which accounts for the difference in global warming potential of CO₂ (GWP = 1) and...
CH$_4$ (GWP = 34, at 100 years timescale including climate-carbon feedbacks; Myhre et al., 2013).

While several factors drive CO$_2$ emission from fresh waters (Maberly, Barker, Stott, & De Ville, 2013; Marcé et al., 2015), many fresh waters are net heterotrophic ecosystems due to import and subsequent mineralization of terrestrial organic matter. Hence, they are net sources of atmospheric CO$_2$ (Cole, Pace, Carpenter, & Kitchell, 2000; Duarte & Prairie, 2005; Pace et al., 2004; CO$_2$ emissions > 0). With increasing inorganic nutrient supply and thus productivity, net ecosystem metabolism turns to become net autotrophic (Hanson, Bade, Carpenter, & Kratz, 2003; Hanson et al., 2004), thus CO$_2$ emissions are expected to decrease (Balmer & Downing, 2011; Pacheco, Roland, & Downing, 2013; Schindler, Carpenter, Cole, Kitchell, & Pace, 1997). C burial to increase (Anderson, Bennion, & Lotter, 2014; Flanagan, Mccauley, & Wrona, 2006; Heathcote & Downing, 2012) and ecosystems can turn into CO$_2$ sinks (Balmer & Downing, 2011; CO$_2$ emissions < 0).

However, eutrophication can change the CO$_2$-equivalent balance by methanogenic microbes in the sediments transforming a fraction of the CO$_2$ fixed by autotrophic primary production into CH$_4$ (Grasset et al., 2018; West, Coloso, & Jones, 2012). Accordingly, CH$_4$ emissions have been shown to increase exponentially with freshwater productivity (Bastviken, Cole, Pace, & Tranvik, 2004; Beaulieu, DelSontro, & Downing, 2019; Davidson et al., 2018; Grasset, Abril, Guillard, Delolme, & Bornette, 2016). Eutrophication consequently has two opposite effects on the CO$_2$-equivalent fluxes, inducing both increased CO$_2$ uptake from the atmosphere, but also enhanced CH$_4$ release to the atmosphere (DelSontro, Beaulieu, & Downing, 2018). However, the effect of productivity on the overall CO$_2$-equivalent balance of freshwater ecosystems is rarely considered. The only study published to date reports a negative effect of productivity on the CO$_2$-equivalent emission of shallow-water mesocosms (Davidson et al., 2015), but did not measure CH$_4$ emission via bubbles (ebullition), which typically is a major CH$_4$ emission pathway (Bastviken et al., 2004; Davidson et al., 2018), and was further comprised of only two levels of productivity. This study could as such not answer the question of how far a shift in ecosystem productivity, which is the typical situation in natural systems (Rineau et al., 2019), may affect the CO$_2$-equivalent balance.

Autochthonous primary production in fresh waters is not only controlled by inorganic nutrient supply but also by food web structure (Carpenter et al., 2001). Animals often have indirect effects on biogeochemical processes, sometimes with a large impact on GHG emissions (Schmitz et al., 2014). For instance, increasing planktivorous fish abundance, often concomitant to eutrophication (Jeppesen, Peder Jensen, Søndergaard, Lauridsen, & Landkildehus, 2000; Moss et al., 2011), can increase primary production and thus CO$_2$ uptake by photosynthesis by reducing grazing pressure on zooplankton (Atwood et al., 2013; Cole et al., 2000; Schindler et al., 1997). On the other hand, fish may reduce CH$_4$ emission from fresh waters through top-down control of zooplankton that graze on CH$_4$ oxidizers (Devlin, Saarenheimo, Syväranta, & Jones, 2015). The overall effect of food web structure on the CO$_2$-equivalent balance of freshwater ecosystems has not yet been investigated.

Only a fraction of the CH$_4$ that is produced in anoxic sediments reaches the atmosphere, primarily due to aerobic CH$_4$ oxidation by CH$_4$-oxidizing bacteria. Between 45% and 100% of the produced CH$_4$ in lake sediments could be lost by oxidation (Bastviken, 2009), mainly during CH$_4$ transport by diffusion across the oxic-anoxic interface in the sediment or in the water column. Recent studies show that the responses to drivers such as temperature and nutrients are different for CH$_4$ production and CH$_4$ oxidation (Fuchs, Lyautey, Montuelle, & Casper, 2016; Sepulveda-Jauregui et al., 2018). This implies that the balance between CH$_4$ oxidized and CH$_4$ produced, and thus the proportion of the produced CH$_4$ that is emitted by diffusion to the atmosphere might change along environmental and climatic gradients. Accordingly, the CO$_2$-equivalent balance of fresh waters may vary in complex ways across productivity gradients in response to the combined effects of CO$_2$ fixation and mineralization, food web effects, and production as well as oxidation of CH$_4$.

To determine how the CO$_2$-equivalent balance depends on productivity, we set up a total of 20 mesocosms, two at each of the 10 nutrient levels (total phosphorus [TP] from 39 to 939 µg/L), and one at each nutrient level was stocked with fish. We hypothesized that CH$_4$ emission will increase exponentially and CO$_2$ will decrease linearly with productivity, such that the CO$_2$-equivalent emission will have a minimum along a productivity gradient. In addition, we hypothesized that the presence of fish reduces emissions of CO$_2$ and CH$_4$ due to reduction of zooplankton grazing on phytoplankton and CH$_4$ oxidizers.

## 2 | MATERIALS AND METHODS

### 2.1 | Mesocosm setup

Twenty white opaque high density polyethylene mesocosms of 2 m height and 1 m diameter were deployed in the mesotrophic hard water lake Erken (59°51’N, 18°36’E, Sweden). The mesocosms were filled on June 2017 up to 1.65 m with c. 1,200 L of Erken water filtered through a 200 µm mesh (to remove large plankton and algal colonies), and about 80 L of surface sediment sampled from Erken at 15 m depth 1 week before. TP and total nitrogen (TN) concentration were set to 10 different levels, and of the two mesocosms receiving the same nutrient addition, one mesocosm was stocked with two juvenile crucian carp (Carassius carassius) individuals, which reflects the crucian carp density of natural populations (Holopainen & Pitkänen, 1985). The diet of juvenile crucian carp consists of zooplankton and Chironomidae (Penttinen & Holopainen, 1992), and they were therefore expected to exert the hypothesized top-down control on zooplankton abundance and grazing. The experiment was run for 1 year and 3 months to allow for new detritus to deposit on the sediment and thus affect methanogenesis. Zooplankton inoculates (approximately 13 individuals L$^{-1}$), obtained from tows with a 100 µm plankton net were added to the mesocosms.
Fish were added to 10 of the mesocosms during spring and summer (July–October 2017 and May–September 2018) and were removed during winter by hand-netting in order to avoid death because of low oxygen during ice cover. The mesocosms were shaded with black plastic sheets placed on the outside of walls to limit periphyton growth. In order to allow for all autochthonous organic carbon to reach the sediment and contribute to methanogenesis, the mesocosm walls were scraped every 4 weeks during the ice-free period to detach periphyton. Primarily the fluxes measured during summer 2018 (May–September 2018) were analyzed in this study since there was a lag in the emergence of sediment methanogenesis, which was very low during the first year (2017). However, fluxes and partial pressure of CO₂ and CH₄ throughout the entirety of the experiment (July 2017 to September 2018) are presented in Figures S1–S4 in the Supplementary material.

2.2 | Nutrient gradient establishment

A gradient of TP concentration in the water column with 10 different levels was set: background TP concentration of lake Erken (no addition), 40, 60, 80, 100, 150, 200, 400, 600, and 1,000 µg/L, spanning from mesotrophic to hypereutrophic. Each target concentration was set for two mesocosms, and one mesocosm was stocked with fish during the ice-free period, while the other mesocosm was without fish. TN concentrations were calculated to obtain an N/P atomic ratio of 16, allowing other algae than N-fixing bacteria to colonize the mesocosms (TN target concentration between 0.45 and 11.29 mg/L). Monopotassium phosphate (KH₂PO₄) and ammonium nitrate (NH₄NO₃) were added to adjust to the TP and TN target concentrations on the first day, every 2 weeks during the ice-free period, and every 4 weeks during the ice-covered period. No nutrients were added in the lowest nutrient treatment to keep the background concentration. Over summer 2018, the mean TP values varied between 39 ± 35 (SD) µg/L for the no addition treatments and 939 ± 381 (SD) µg/L for the highest nutrient treatments. An extremely high TP value of 1,000 µg/L can be encountered in fresh waters and it is consequently appropriate to include it in our eutrophication gradient to cover the most extreme cases of eutrophication (DelSontro et al., 2018).

2.3 | Water analyses

Water samples were taken from each mesocosm every 2 weeks during the ice-free period, every 4 weeks otherwise, for nutrient and dissolved organic carbon (DOC) analyses. Water was collected with a 1 m long plastic tube to get integrative samples of the water column. TP concentration was measured using the ammonium molybdate spectrometric method (Swedish standard method SS-EN ISO 6878:2005, Erken Laboratory). TN and DOC concentrations were measured on GF/F filtered (effective pore size 0.7 µm, Whatman®), GE Healthcare and acidified samples (Hydrochloric acid HCl 1 M) with a Shimadzu TOC-L TNM-L analyzer. Turbidity, pH, temperature, dissolved oxygen, and Chlorophyll a (Chl a), were conjointly measured with a multiprobe (EXO2 Multiparameter Sonde, YSI) at 50, 100 and 125 cm below the water surface. In addition, dissolved oxygen was automatically measured every 10 min with oxygen probes (miniDO2T PME) at a depth of 25–30 cm below water surface.

2.4 | Zooplankton sampling

Zooplankton samples were taken from the integrated water sample used for the water analyses. Five liters of water was filtered through 55 µm and zooplankton was immediately preserved with Lugol's solution and later analyzed using an inverted microscope (Leica DM IL LED, Leica) with image analysis software (Image Pro Plus version 7.0 for Windows, Media Cybernetics Inc.). Subsamples were counted until reaching 200 individuals and zooplankton were grouped into Cladocera (Bosmina sp., Daphnia sp., Ceriodaphnia sp., Diaphanosoma sp., Polyphemus sp. and Scapholeberis sp.) and Copepoda (Cyclopoida and Calanoida). We measured the total length of up to 20 individuals of each zooplankton taxon using an image analysis software (Image Pro Plus version 7.0 for Windows, Media Cybernetics) to calculate zooplankton biomass based on published length–weight relationships (McCaulay, 1984).

2.5 | Primary producer biomass

Phytoplankton biomass was calculated from the Chl a (µg/L) measurements in the water column by assuming a C:Chl a ratio of 40 (µg/g; Lorenzen, 1968). Periphyton biomass was measured using plastic strips, made of the same material as the mesocosms, that were attached to the walls of the mesocosms and reached the full depth down to the sediment. The strips were scraped every 4 weeks during the ice-free period, and biomass was upscaled from the width of the strip (7 cm) to the full diameter of the mesocosm. Samples were dried at 50–60°C, grinded manually into a fine powder with a mortar and pestle, acidified with HCl 5% and encapsulated in tin capsules for total organic carbon and nitrogen analysis with a C/N elemental analyzer (Costech Analytical Technologies Inc.). The total primary producer biomass in the mesocosms (g C mesocosm⁻¹) was calculated by summing periphyton and phytoplankton biomass. Sediment was sometimes also visibly present on the periphyton strips and thus increased the overall estimation of periphyton C content. However, this contamination was likely to be similar among treatments and negligible for high nutrient treatments since sediment C content was low (7% wt, data not shown) in comparison to periphyton C content (algal C content is typically around 20%–50% wt).

2.6 | Net ecosystem production

Dissolved oxygen automatic measurements at 10 min intervals were used to estimate net ecosystem production (NEP in mg O₂ L⁻¹ hr⁻¹) according to Cole et al. (2000) as follows:
Air water exchange = $K_T \times (DO_{sat} - DO_t) / z$,  

with $z$ the mixing depth of the system was assumed to be the total water depth (1.65 m) as no stratification was observed in the mesocosms.

$DO_{sat}$ in mg/L was defined according to Benson and Krause Jr. (1984):

$$DO_{sat} = \exp \left( -139.34411 + 1.575701 \times 10^5 \times T^{-1} - 6.642308 \times 10^7 \times T^{-2} + 1.2438 \times 10^{10} \times T^{-3} - 8.621949 \times 10^{11} \times T^{-4} \right).$$  

with $T$ in Kelvin.

$K_T$ the gas transfer velocity of $O_2$ in m/hr was calculated from an average $K_{600}$ of 0.014 m/hr measured over summer 2016 and 2017 (details on the gas transfer velocity calculation are given in the supplementary material) as follows:

$$K_T = K_{600} \times (600/Sc)^n,$$  

with $Sc$ the Schmidt number of $O_2$ according to Wanninkhof (1992). We chose $n = 1/2$ since the water surface in the mesocosms was often not smooth.

NEP is expressed in mmol O$_2$ m$^{-2}$ day$^{-1}$ in the rest of the manuscript for comparisons with literature.

### 2.7 | CO$_2$ and CH$_4$ diffusive fluxes

CH$_4$ and CO$_2$ concentrations in the water were measured every 2 weeks during the ice-free period, and every 4 weeks during ice cover, with the headspace method (Cole & Caraco, 1998), by sampling 30 ml of surface water in each mesocosm and equilibrating 1 min with 10 ml of atmospheric air. The gas samples were then transferred to another syringe and analyzed within 24 hr with a gas chromatograph equipped with a flame ionization detector (Agilent Technologies, 7890 A GC system). CO$_2$ and CH$_4$ concentrations were calculated from their concentrations in the headspace, the volume of water, and the specific gas solubility of CO$_2$ (Weiss, 1974) and CH$_4$ (Yamamoto, Alcauskas, & Crozier, 1976). The diffusive fluxes of CO$_2$ and CH$_4$ from the water to the atmosphere were calculated according to Cole and Caraco (1998):

$$F = K_T \times (C - P_{sat} \times K_H),$$  

where $F$ is the flux in mmol m$^{-2}$ day$^{-1}$, $K_T$ is the gas transfer velocity in m/day calculated according to Equation (4), $C$ is the gas concentration in µmol/L, $P_{sat}$ is the atmospheric gas concentration in µatm and $K_H$ is Henry’s constant in mol L$^{-1}$ atm$^{-1}$.

**Daily CO$_2$ and CH$_4$ fluxes over summer 2018 were estimated from single daytime measurements in water sampled between 9 a.m. and 11 a.m. Over summer 2017, however, CO$_2$ concentration was also measured at night (between 3 a.m. and 5 a.m.) at three dates (August, September and October), and the nighttime concentrations of CO$_2$ were found to be very close to the daytime concentrations measured between 9 a.m. and 11 a.m. (for the three dates, average of the slope = 1.1 and average of the $R^2$ = .86, data not shown). In May, June and July, the nights are very short (<6 hr) and not completely dark at this latitude, which probably limits the daily variation in CO$_2$. We consequently assumed that daily variation in CO$_2$ concentration was low over summer and that a single CO$_2$ concentration measurement can be representative of its daily concentration.

### 2.8 | CH$_4$ ebullitive flux

Bubble traps consisting of a 50 ml syringe standing just below the water surface and attached to 20 cm wide inverted funnel (Davidson et al., 2018; Huttunen, Lappalainen, Saarijärvi, Väisänen, & Martikainen, 2001) were used to collect CH$_4$ bubbles. The transparent syringes were covered by an opaque light-grey cap to prevent biofilm growth on the surface of the syringe and were cleaned at each sampling. The bubble traps remained permanently in the mesocosms and were sampled every 2 weeks during the ice-free period, every 4 weeks otherwise, when gas was visibly accumulating in the bubble traps. The gas was transferred to a syringe and analyzed within 24 hr with the gas chromatograph. The ebullitive flux of CH$_4$ (in mmol m$^{-2}$ day$^{-1}$) was calculated as the amount of CH$_4$ (mmol) divided by the surface of the funnel in m$^2$ and the number of days between two consecutive measurements. The volume of the trapped gas exceeded the volume of the syringe at one occasion for mesocosm I with fish (13/06/18) and mesocosm I without fish (14/08/18; A being the lowest nutrient treatment and J the highest), resulting in an underestimation of ebullition flux for these two treatments.

### 2.9 | CO$_2$-equivalent balance

CH$_4$ diffusive as well as CH$_4$ ebullitive fluxes were converted in CO$_2$-equivalent (and reported in mg CO$_2$ m$^{-2}$ day$^{-1}$) by assuming that 1 g of CH$_4$ has 34 times the GWP of 1 g of CO$_2$ for 100 years. This number of 34 includes climate-carbon feedback (Myhre et al., 2013). The sum of ebullitive and diffusive CH$_4$ fluxes was converted to CH$_4$-equivalent balance, since it was recently estimated to contribute to only 2% of the total CO$_2$-equivalent emission of global lakes and reservoirs (DeSontro et al., 2018).
2.10 | CH₄ oxidation

Two different methods were used to quantify CH₄ oxidation, the fraction of the CH₄ produced in sediment that was oxidized in situ was calculated from δ¹³C-CH₄ measurements, and the potential CH₄ oxidation rate was calculated from aerobic incubation of the mesocosm water. Details on the potential CH₄ oxidation rates are given in the supplementary material.

The fraction of CH₄ oxidized in situ was calculated from δ¹³C of CH₄ in mesocosm surface water and δ¹³C of anaerobically produced CH₄ in a sediment incubation. δ¹³C-CH₄ in surface water samples was measured at two dates (13/08/18 and 11/09/18) in each mesocosm. CH₄ gas samples were obtained with the headspace method by equilibrating 90 ml of surface water with 20 ml of N₂ in 120 ml syringes for 2 min. The gas samples were stored in preevacuated 12 ml vials (Soda Glass Vials 819W, Labco Ltd) that were manually flushed and filled with N₂ at atmospheric pressure. Details on the anaerobic sediment incubation can be found in the supplementary material.

The fraction of CH₄ oxidized can be calculated for an open steady state system (F_{oxi open}, Equation 6) or for a closed system (F_{oxi closed}, Equation 7; Bastviken, Ejlertsson, & Tranvik, 2002):

\[
F_{oxi open} = \frac{\delta^{13}CH_{4,oxidized} - \delta^{13}CH_{4,newly formed}}{(a-1) \times 1,000},
\]

\[
\ln(1-F_{oxi closed}) = \frac{\ln(\delta^{13}CH_{4,newly formed} + 1.000)}{a-1} - \frac{\ln(\delta^{13}CH_{4,oxidized} + 1.000)}{a-1}
\]

\[\delta^{13}CH_{4,newly formed}\]

\[\delta^{13}CH_{4,oxidized}\]

\(a\), the fractionation factor, is assumed to be 1.02 (Bastviken et al., 2002). δ¹³C CH₄ oxidized is the δ¹³C of CH₄ in surface water that has undergone oxidation through the sediment and water column, and δ¹³C CH₄ newly formed is the δ¹³C of CH₄ before oxidation. Some studies use δ¹³C CH₄ measurements from gas bubbles or bottom waters to estimate δ¹³C CH₄ newly formed (Barbosa et al., 2018; Thottathil, Reis, Giorgio, & Prairie, 2018) but it is then not possible to exclude that oxidation has already occurred in the sediment or during gas transport. We consequently used δ¹³C of CH₄ produced during an anoxic sediment incubation as δ¹³C CH₄ newly formed (Zhang, Yu, Fan, Ma, & Xu, 2016) to calculate the fraction of anaerobically produced CH₄ in the sediment that is oxidized. In an open system at steady state, CH₄ production is supposed constant and CH₄ as well as the products of CH₄ oxidation can leave freely, while in closed systems, CH₄ and oxidation products accumulate (Barbosa et al., 2018; Bastviken et al., 2002). The fraction of CH₄ oxidized calculated for open systems gave values often >1 (values between 0.77 and 2.86, mean of 1.59) while it gave a mean of 0.78 and values between 0.56 and 0.95 for closed systems (Figure S5). Values often >1 were also reported in floodplains and lakes (Barbosa et al., 2018; Bastviken et al., 2002; Thottathil et al., 2018) suggesting that the assumptions for open systems might not always be valid in natural systems. We consequently chose to use the fraction of CH₄ oxidized in closed systems as a conservative measurement of CH₄ oxidation in the rest of the manuscript.

2.11 | Statistical analyses

The effects of TP concentration and fish presence on productivity (NEP) and C fluxes (total CH₄ and CO₂ emissions) and the fraction of CH₄ oxidized were tested with analysis of covariance (lm function with fish presence as a categorical variable). The relationships between C fluxes (total CH₄ and CO₂ emissions, CO₂-equivalent balance), the fraction of CH₄ oxidized, potential CH₄ oxidation rate, productivity (NEP), primary producer biomass and zooplankton biomass were tested with linear first-order regressions and polynomial models (lm function). All models were based on the averages of the C fluxes, NEP and primary producer biomass over summer 2018 for each mesocosm (n = 20) because averages were considered more robust and integrative of the whole period, and indicative of the overall treatment effects regardless of any temporal variability. Furthermore, as we did not see any consistent temporal patterns for C fluxes and NEP (Figures S3, S4 and S6), we preferred to choose the simpler models with averages rather than the more complex mixed-effect models that also show a positive effect of productivity on CH₄ emissions and a negative effect on CO₂ emissions (Table S1). All variables except the fraction of CH₄ oxidized and NEP were log-transformed (natural logarithm) before modeling to normalize distributions and decrease the effect of extreme values. Before log-transformation, a constant was added to CO₂ emissions, CO₂-equivalent balance (180 mg CO₂ m⁻² day⁻¹) and to potential CH₄ oxidation rates (1 mg L⁻¹ day⁻¹) to make all values positive. For CO₂ emissions and CO₂-equivalent balance, the minimum values were −175 and −168 mg CO₂ m⁻² day⁻¹, respectively, the addition of 180 was chosen to give the best data normalization (according to Shapiro tests and histogram plots of the data). The accuracy of the models was assessed by visualizing the residuals and the observed against predicted data. When comparing several polynomial models, the best model was chosen according to the Akaike information criterion. Thresholds of the polynomial model between CO₂-equivalent balance and NEP were determined by the “optim” function that returns parameters that minimize a function. All statistical analyses were performed with the R software (R Core Team, 2016).

3 | RESULTS

3.1 | Relationships between C fluxes, productivity and primary producer biomass

As expected, total CH₄ emissions (diffusive + ebullitive) increased with productivity while CO₂ emissions decreased (Figure 1; Table 1). The increase in total CH₄ emissions however, was less pronounced towards the highest productivity values (Figure 1; Table 1). CH₄ ebullition also increased with productivity (p = 0.004, R² = 0.37) and occurred in 11 out of the 20 mesocosms, for which it contributed in average to 20% (range 0.5%–71.9%) of total CH₄ emissions. The CO₂-equivalent balance had a U-shaped relationship with productivity.
until a threshold at high productivity (NEP = 32 mmol O₂ m⁻³ day⁻¹) after which the CO₂-equivalent balance decreased again with increasing productivity (Figure 1; Table 1). Eight out of nine mesocosms having NEP values between 5 and 18 mmol O₂ m⁻³ day⁻¹ acted as CO₂-equivalent sinks (CO₂-equivalent balance < 0) while all mesocosms with higher or lower productivity acted as CO₂-equivalent sources (Figure 1). In accordance with these observations, a polynomial model between the CO₂-equivalent balance and productivity

---

**TABLE 1** First-order and polynomial regressions between C fluxes (total CH₄ emissions, CO₂ emissions and the CO₂-equivalent balance in mg CO₂ m⁻² day⁻¹), productivity (NEP in mmol O₂ m⁻³ day⁻¹), and TP (µg/L) with presence of fish as a categorical variable

| Predictor                          | F   | Coefficient | p-value | R² (p-value of the model) |
|------------------------------------|-----|-------------|---------|---------------------------|
| Ln total CH₄ emission              |     |             |         |                           |
| NEP                               | 31  | 0.114       | <.0001  | .66 (<.0001)              |
| NEP²                              | 2   | -0.001      | NS (.1) |                           |
| Ln total CH₄ emission              |     |             |         |                           |
| Ln TP                             | 7   | 0.452       | .02     | .47 (.004)                |
| Fish                              | 8   | .01         |         |                           |
| Ln CO₂ emission + 180             |     |             |         |                           |
| NEP                               | 90  | -0.088      | <.0001  | .86 (<.0001)              |
| NEP²                              | 15  | 0.001       | .001    |                           |
| Ln CO₂ emission + 180             |     |             |         |                           |
| Ln TP                             | 3   | -0.198      | NS (.1) | .32 (.04)                 |
| fish                              | 5   | .04         |         |                           |
| Ln CO₂-eq balance + 180           |     |             |         |                           |
| NEP                               | 0.07| -0.06       | NS (.8) | .58 (.008)                |
| NEP²                              | 10  | -7.35E-04   | .006    |                           |
| NEP³                              | 6   | 2.87E-04    | .03     |                           |
| NEP⁴                              | 4   | -5.93E-06   | NS (.05)|                           |

Note: p-values are given for each predictor (continuous and categorical variables) in the model and coefficients are only given for the continuous variables. In addition, the R² and p-value are given for each model. The polynomial models test the multiple linear relationship between NEP and NEP raised at different powers (NEP², NEP³ and NEP⁴), and C fluxes. All fluxes are logged (natural logarithm) and a constant is added before log-transformation to make CO₂ emissions and CO₂-equivalent balance positive. All models are performed on averaged values over summer 2018 (n = 20, average for 9 dates for C fluxes and 117 dates for NEP). When choosing between different polynomial models, the best model was selected according to the lowest Akaike information criterion.

Abbreviations: NEP, net ecosystem production; NS, not significant.
identified two thresholds at NEP = 5 and 19 mmol O₂ m⁻³ day⁻¹ at which the mesocosms shifted from source to sink and then back again from sink to source (Figure 1; Table 1).

Productivity was strongly correlated to primary producer biomass (Figure 2; Table S2) and primary producer biomass correlated in similar ways as productivity to total CH₄ and CO₂ emissions and to the CO₂-equivalent balance (Figure S7; Table S3). In the highest productivity treatments, phytoplankton constituted most of the primary producer biomass, and in the other treatments periphyton often dominated (Figure S8).

### 3.2 Food web structure and nutrient effect on productivity and C fluxes

Both nutrient concentrations and the presence of fish had a positive effect on primary productivity, thereby regulating CO₂ and CH₄ emissions and the resulting CO₂-equivalent balance (Figure 2; Table S2). The presence of fish induced a trophic cascade and increased productivity through a decrease in zooplankton abundance (Atwood et al., 2013; Schindler et al., 1997; Schmitz et al., 2014; Figure 2; Table S2). Nutrient concentrations and the presence of fish had a positive effect on CH₄ emissions and a negative effect on CO₂ emissions (Table 1; Figure S9). The direct effect of nutrients on CO₂ emissions was however not significant probably because two mesocosms (J and H) with high nutrient concentrations and low productivity had sometimes a high respiration (Figure S9; Figure S4).

### 3.3 Relationship between CH₄ oxidation, productivity and food web structure

The fraction of CH₄ that was oxidized increased with productivity (Figure 3) and primary producer biomass, and decreased with Copepoda biomass (Figure S10; Table S4). However, when CH₄ oxidation was correlated to both fish presence and nutrient concentration, the effect of fish was not significant, suggesting that the presence of fish did not directly enhance CH₄ oxidation (Table S4).

### 4 DISCUSSION

Our study is the first to report a non-linear relationship between freshwater productivity (or primary producer biomass) and the net ecosystem CO₂-equivalent balance. This novel result means that eutrophic freshwater ecosystems have the potential to act as CO₂-equivalent sinks, but these sinks are fragile since a small increase or decrease of productivity can turn them again into a
CO₂-equivalent source. We propose a conceptual model describing the non-linear relations between productivity and C fluxes in fresh waters (Figure 4), based on our regressions with NEP (Figure 1). We identified two thresholds at NEP = 5 and 19 mmol O₂ m⁻³ day⁻¹ for which the studied freshwater ecosystems shifted from CO₂-equivalent sources to sinks and then returned to being CO₂-equivalent sources. In other fresh waters than the ones studied here, different environmental conditions can shift the thresholds. For example, higher import of dissolved inorganic carbon or higher mineralization of imported organic matter can lead to higher CO₂ emissions (Duarte & Prairie, 2005; Hanson et al., 2004) for the same level of productivity and shift the U-shaped relationship between CO₂-equivalent balance and productivity upwards. In the same way, in temperate and tropical ecosystems higher temperatures can have a positive effect on CH₄ emissions (Davidson et al., 2018; Yvon-Durocher et al., 2014) and are also likely to shift the U-shaped relationship upwards. This implies that the narrow interval of being a CO₂-equivalent sink may be even narrower, and the CO₂-equivalent sink action may disappear completely if the minimum value of CO₂-equivalent balance is positive. Accordingly, we suggest a general pattern of CO₂-equivalent balance across freshwater productivity gradients, shaped by the contrasting effects of productivity on CO₂ and CH₄ fluxes. However, the specific positions of thresholds and the magnitude of the CO₂-equivalent emissions is likely to differ among different fresh water types and climatic zones. Our findings are not directly applicable to shallow fresh waters colonized by aquatic plants because in addition to providing substrates for CH₄ production, aquatic plants have other complex effects on CH₄ fluxes that would need to be addressed (e.g., CH₄ oxidation in the rhizosphere, CH₄ transport through plant tissues; Davidson et al., 2018; Kosten et al., 2016; Schütz, Schröder, & Rennenberg, 1991). The extent to which the experimentally derived qualitative pattern depicted in Figure 4 manifests quantitatively in various types of lake ecosystems, that is, which levels of productivity represent thresholds of which levels of CO₂-equivalent balance, is therefore unknown and probably variable, and should be the subject of further studies.

Our results indicate that the positive top-down effect of planktivorous fish on productivity decreased CO₂ emissions, in accordance with several studies (Atwood et al., 2013; Cole et al., 2000; Schindler et al., 1997), and increased CH₄ emissions. For CH₄ emissions, this is in apparent contradiction with a recent study showing reduced CH₄ emissions from lakes where fish were present through top-down control of zooplankton that graze on methane oxidizers (Devlin et al., 2015). However, in the latter study the lake was highly rich in humic matter, and the fish addition may not have increased CH₄ production because primary production may have been light-limited in the dark-stained water.

In our study, even if CH₄ emissions increased with productivity, at the same time the fraction of CH₄ oxidized also increased with productivity, hence counteracting CH₄ emissions (Figure 3; Table S4). We consequently attribute the flattening of the increase in total CH₄ emissions and the decrease in CO₂-equivalent balance towards the highest productivity levels (Figure 1) to a higher proportion of CH₄ lost by oxidation in the high-NEP treatments. Several studies indeed showed a flattening or a decrease in CH₄ concentration or CH₄ diffusive emissions towards very high chlorophyll a values (i.e., Chla > 200 µg L⁻¹; Beaulieu et al., 2019; Wang, Lu, Wang, Yang, & Yin, 2006; Yan et al., 2018). However, very few observations are available for hypereutrophic systems and these patterns should be more thoroughly verified in natural systems.

The increase in the fraction of CH₄ oxidized with productivity cannot be attributed to CH₄ concentration only. Indeed, when the CH₄ concentration is limiting the rate of CH₄ oxidation, it increases linearly with CH₄ concentration (Lofton, Whalen, & Hershey, 2014; Sundh, Bastviken, & Tranvik, 2005), and the fraction of CH₄ oxidized can thus be assumed to be constant. Previous studies have underlined a negative effect of light (Shelley, Ings, Hildrew, Trimmer, & Grey, 2017) or a related positive effect of DOC (Thottathil et al., 2018) on CH₄ oxidation. In our study, the strong correlation between primary producer biomass and the fraction of CH₄ oxidized (Figure S10; Table S4), indicates that light shading by primary producers could enhance CH₄ oxidation. Furthermore, the presence of fish and the associated decrease in zooplankton (Figure 2) could release the grazing pressure on the CH₄ oxidizing bacteria (Devlin et al., 2015), thereby increasing CH₄ oxidation (Figure S10; Table S4), as we hypothesized. Fish
can also directly enhance CH4 oxidation via sediment reworking (i.e., bioturbation) and increasing O2 supply to sediment (Oliveira Junior et al., 2019) but this does not seem to be the case in our study because CH4 oxidation was not significantly correlated to the presence of fish (Table S4). The food web structure and eutrophication had consequently antagonistic effects on CH4 emissions. Primarily, and most visibly, the presence of fish and nutrient enrichment increased CH4 emissions via their positive effect on productivity. On the other hand, they also increased CH4 oxidation most likely via an increase in primary producer biomass and/or a decrease in zooplankton abundance. Importantly, our results suggest that CH4 oxidation could play an important role for the CO2-equivalent balance of freshwater ecosystems, calling for more studies on the drivers and magnitude of CH4 oxidation in natural systems.

We show for the first time that eutrophication can alter the CO2-equivalent balance of freshwater ecosystems in a non-linear way, and have a negative or a positive feedback on climate depending on the magnitude of productivity increase. In contrast, a recent study used exponential relationships between CO2, CH4 and Chl a or TP to predict a future increase of freshwater CO2-equivalent emission due to eutrophication (DelSontro et al., 2018). This difference in productivity and CO2-equivalent emission relationships may arise from spatial disconnection of measurements; while our data describe the effect of productivity on the balance of CO2 and CH4, the other study used published data from different systems to derive separate relationships for CO2 and CH4, and can therefore not reflect any combined effect within a single ecosystem (Figure 4). The experimental evidence presented here calls for studies on natural systems that investigate both the CO2 and CH4 balance over a gradient of productivity, not the least since the thresholds between a negative to a positive feedback on climate are susceptible to differ between fresh water types and climatic zones.

ACKNOWLEDGEMENTS

Funding was received from the Swedish Research Council (grant 2018-04524) and from the Knut and Alice Wallenberg Foundation (grant KAW 2013.0091) to L.J.T. Additional funding from the European Research Council under the European Union’s Seventh Framework Programme (FP7/2007-2013)/ERC grant agreement no. 336642 to S.S. is acknowledged. We thank the KAWater team for help during fieldwork and the Erken laboratory for help with laboratory analyses. We acknowledge SITES for provisioning of facilities and experimental support. SITES received funding through the Swedish Research Council (grant no. 2017-00635).

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ORCID
Charlotte Grasset ID https://orcid.org/0000-0002-3251-7974

REFERENCES

Anderson, N. J., Bennion, H., & Lotter, A. F. (2014). Lake eutrophication and its implications for organic carbon sequestration in Europe. Global Change Biology, 20(9), 2741–2751. https://doi.org/10.1111/gcb.12584

Atwood, T. B., Hammill, E., Greig, H. S., Kratina, P., Shurin, J. B., Srivastava, D. S., & Richardson, J. S. (2013). Predator-induced reduction of freshwater carbon dioxide emissions. Nature Geoscience, 6(3), 191–194. https://doi.org/10.1038/ngeo1734

Balmer, M. B., & Downing, J. A. (2011). Carbon dioxide concentrations in eutrophic lakes: Undersaturation implies atmospheric uptake. Inland Waters, 1(2), 125–132. https://doi.org/10.5268/IW-1.2.366

Barbosa, P. M., Farjalla, V. F., Melack, J. M., Amaral, J. H. F., da Silva, J. S., & Forsberg, B. R. (2018). High rates of methane oxidation in an Amazon floodplain lake. Biogeochemistry, 137(3), 351–365. https://doi.org/10.1007/s10533-018-0425-2

Bastviken, D. (2009). Methane. In G. E. Likens (Ed.), Encyclopedia of inland waters (pp. 783–805). Oxford, UK: Academic Press.

Bastviken, D., Cole, J. J., Pace, M. L., & Tranvik, L. (2004). Methane emissions from lakes: Dependence of lake characteristics, two regional assessments, and a global estimate. Global Biogeochemical Cycles, 18(4), GB4009. https://doi.org/10.1029/2004GB002238

Bastviken, D., Ejertsson, J., & Tranvik, L. (2002). Measurement of methane oxidation in lakes: A comparison of methods. Environmental Science & Technology, 36(15), 3354–3361. https://doi.org/10.1021/es010311p

Bastviken, D., Tranvik, L. J., Downing, J. A., Crill, P. M., & Enrich-Prast, A. (2011). Freshwater methane emissions offset the continental carbon sink. Science, 331(6013), 50. https://doi.org/10.1126/science.1196808

Beaulieu, J., DelSontro, T., & Downing, J. (2019). Eutrophication will increase methane emissions from lakes and impoundments during the 21st century. Nature Communications, 10. https://doi.org/10.1038/s41467-019-09100-5

Benson, B. B., & Krause Jr., D. (1984). The concentration and isotopic fractionation of oxygen dissolved in freshwater and seawater in equilibrium with the atmosphere. Limnology and Oceanography, 29(3), 620–632. https://doi.org/10.4319/lo.1984.29.3.0620

Carpenter, S. R., Cole, J. J., Hodgson, J. R., Kitchell, J. F., Pace, M. L., Bade, D., … Schindler, D. E. (2000). Trophic cascades, nutrients, and lake productivity: Whole-lake experiments. Ecological Monographs, 71(2), 163–186. https://doi.org/10.2307/2657215

Cole, J. J., & Caraco, N. F. (1998). Atmospheric exchange of carbon dioxide in a low-wind oligotrophic lake measured by the addition of SF6. Limnology and Oceanography, 43(4), 647–656.

Cole, J. J., Pace, M., Carpenter, S., & Kitchell, J. (2000). Persistence of net heterotrophy in lakes during nutrient addition and food web manipulation. Limnology and Oceanography, 45, 1718–1730. https://doi. org/10.4319/lo.2000.45.8.1718

Cole, J. J., Prairie, Y. T., Caraco, N. F., McDowell, W. H., Tranvik, L. J., Striegl, R. G., … Melack, J. (2007). Plumbing the global carbon cycle: Integrating inland waters into the terrestrial carbon budget. Ecosystems, 10(1), 172–185. https://doi.org/10.1007/s10021-006-9013-8

Davidson, T. A., Audet, J., Jeppesen, E., Landkildehus, F., Lauridsen, T. L., Sandergaard, M., & Syværenta, J. (2018). Synergy between nutrients and warming enhances methane ebullition from experimental lakes. Nature Climate Change, 8(2), 156–160. https://doi.org/10.1038/s41558-017-0063-z

Davidson, T. A., Audet, J., Svenning, J.-C., Lauridsen, T. L., Sandergaard, M., Landkildehus, F., … Jeppesen, E. (2015). Eutrophication effects on greenhouse gas fluxes from shallow-lake mesocosms override those of climate warming. Global Change Biology, 21(12), 4449–4463. https://doi.org/10.1111/gcb.13062
Schmitz, O. J., Raymond, P. A., Estes, J. A., Kurz, W. A., Holtgrieve, G. W., Ritchie, M. E., ... Wilmers, C. C. (2014). Animating the carbon cycle. *Ecosystems*, 17(2), 344–359. https://doi.org/10.1007/s10021-013-9715-7

Schütz, H., Schröder, P., & Rennenberg, H. (1991). Role of plants in regulating the methane flux to the atmosphere. In H. A. Mooney, T. D. Sharkey, & E. A. Holland (Eds.), *Trace gas emissions by plants* (pp. 29–63). San Diego, CA: Academic Press.

Sepulveda-Jauregui, A., Hoyos-Santillan, J., Martinez-Cruz, K., Walter Anthony, K. M., Casper, P., Belmonte-Izquierdo, Y., & Thalasso, F. (2018). Eutrophication exacerbates the impact of climate warming on lake methane emission. *Science of the Total Environment*, 636, 411–419. https://doi.org/10.1016/j.scitotenv.2018.04.283

Shelley, F., Ings, N., Hildrew, A. G., Trimmer, M., & Grey, J. (2017). Bringing methanotrophy in rivers out of the shadows. *Limnology and Oceanography*, 62(6), 2345–2359. https://doi.org/10.1002/lno.10569

Sundh, I., Bastviken, D., & Tranvik, L. J. (2005). Abundance, activity, and community structure of pelagic methane-oxidizing bacteria in temperate lakes. *Applied and Environmental Microbiology*, 71(11), 6746–6752. https://doi.org/10.1128/aem.71.11.6746-6752.2005

Thottathil, S., Reis, P., Giorgio, P., & Prairie, Y. (2018). The extent and regulation of summer methane oxidation in northern lakes. *Journal of Geophysical Research: Biogeosciences*, 123(10), 3216–3230. https://doi.org/10.1002/2018JG004464

Tranvik, L. J., Downing, J. A., Cotner, J. B., Loiselie, S. A., Striegl, R. G., Ballatore, T. J., ... Weyhenmeyer, G. A. (2009). Lakes and reservoirs as regulators of carbon cycling and climate. *Limnology and Oceanography*, 54(6part2), 2298–2314. https://doi.org/10.4319/lo.2009.54.6_part_2.2298

Wang, H., Lu, J., Wang, W., Yang, L., & Yin, C. (2006). Methane fluxes from the littoral zone of hypereutrophic Taihu Lake, China. *Journal of Geophysical Research: Atmospheres*, 111(D17). https://doi.org/10.1029/2005jd006864

Wanninkhof, R. (1992). Relationship between wind speed and gas exchange over the ocean. *Journal of Geophysical Research: Oceans*, 97(C5), 7373–7382. https://doi.org/10.1029/92jc00188

Weiss, R. (1974). Carbon dioxide in water and seawater: The solubility of a non-ideal gas. *Marine Chemistry*, 2, 203–215. https://doi.org/10.1016/0304-4203(74)90015-2

West, W. E., Coloso, J. J., & Jones, S. E. (2012). Effects of algal and terrestrial carbon on methane production rates and methanogen community structure in a temperate lake sediment. *Freshwater Biology*, 57(5), 949–955. https://doi.org/10.1111/j.1365-2427.2012.02755.x

Yamamoto, S., Alcauskas, J. B., & Crozier, T. E. (1976). Solubility of methane in distilled water and seawater. *Journal of Chemical & Engineering Data*, 21(1), 78–80. https://doi.org/10.1021/je60068a029

Yan, X., Xu, X., Ji, M., Zhang, Z., Wang, M., Wu, S., ... Liu, H. (2018). Cyanobacteria blooms: A neglected facilitator of CH4 production in eutrophic lakes. *Science of the Total Environment*, 651, 466–474. https://doi.org/10.1016/j.scitotenv.2018.09.197

Yvon-Durocher, G., Allen, A. P., Bastviken, D., Conrad, R., Gudasz, C., St-Pierre, A., ... del Giorgio, P. A. (2014). Methane fluxes show consistent temperature dependence across microbial to ecosystem scales. *Nature*, 507(7493), 488–491. https://doi.org/10.1038/nature13164

Zhang, G., Yu, H., Fan, X., Ma, J., & Xu, H. (2016). Carbon isotope fractionation reveals distinct process of CH4 emission from different compartments of paddy ecosystem. *Scientific Reports*, 6, 27065. https://doi.org/10.1038/srep27065