Therapy-Induced Senescence: Opportunities to Improve Anticancer Therapy

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Abstract

Cellular senescence is an essential tumor suppressive mechanism that prevents the propagation of oncogenically activated, genetically unstable, and/or damaged cells. Induction of tumor cell senescence is also one of the underlying mechanisms by which cancer therapies exert antitumor activity. However, an increasing body of evidence from preclinical studies demonstrates that radiation and chemotherapy cause accumulation of senescent cells (SnCs) both in tumor and normal tissue. SnCs in tumors can, paradoxically, promote tumor relapse, metastasis, and resistance to therapy, in part, through expression of the senescence-associated secretory phenotype. In addition, SnCs in normal tissue can contribute to certain radiation- and chemotherapy-induced side effects. Because of its multiple roles, cellular senescence could serve as an important target in the fight against cancer. This commentary provides a summary of the discussion at the National Cancer Institute Workshop on Radiation, Senescence, and Cancer (August 10-11, 2020, National Cancer Institute, Bethesda, MD) regarding the current status of senescence research, heterogeneity of therapy-induced senescence, current status of senotherapeutics and molecular biomarkers, a concept of “one-two punch” cancer therapy (consisting of therapeutics to induce tumor cell senescence followed by selective clearance of SnCs), and its integration with personalized adaptive tumor therapy. It also identifies key knowledge gaps and outlines future directions in this emerging field to improve treatment outcomes for cancer patients.
Cells become senescent after extensive replication that causes telomere shortening (1) or from exposure to genotoxic, oncogenic, and/or oxidative stress (2). Senescent cells (SnCs) induced by different stimuli share some common characteristics including an essentially stable growth arrest, relative resistance to apoptosis, persistent DNA damage signaling, changes in heterochromatin, decreased lamin-B1 levels, and increased expression of the cyclin-dependent kinase (CDK) inhibitors, p16INK4a (p16, encoded by the INK4a/ARF locus, also known as Cdkn2a), p21Cip1/Waf1 (p21, encoded by Cdkn1a) (3) and senescence-associated β-galactosidase (SA-β-gal) (4). SnCs secrete a plethora of factors, including proinflammatory cytokines, chemokines, matrix metalloproteinases, bioactive lipids, noncoding nucleotides (miRNAs, mitochondrial DNA), vesicles, and growth factors, collectively termed the senescence-associated secretory phenotype (SASP) (5-10). SnCs can exist in a continuum of states and contribute to a variety of physiological and pathophysiological processes, including organogenesis and wound healing (11). Cellular senescence is also a critical barrier for tumorigenesis, preventing division of cells with oncogene activation and genetic instability and promoting immune clearance of these cells, in part, through the SASP (12).

Senescence occurs after treatment with radiation and/or certain chemotherapies, known as therapy-induced senescence (TIS) (13,14), through induction of DNA double-strand breaks (DSBs) (15). On one hand, senescence can contribute to antitumor effects and treatment outcomes (16); on the other, chronic accumulation of SnCs can stimulate relapse and metastasis (17). Both these effects have been linked to the SASP and clearly suggest the importance of cellular and tissue context. For example, some tumor cells may escape TIS with the acquisition of genomic changes that confer treatment resistance (18), especially under a p53-deficient environment and p21-driven genomic instability (19). Also, malignant cells reprogrammed by TIS to acquire a change in lineage and/or stemness can become self-renewing tumor-initiating cells that cause tumor relapse and promote aggressive growth (20-22). Studies demonstrated that transplanting relatively small numbers of senescent cells into young mice was sufficient to cause persistent physical dysfunction and spread cellular senescence to host tissues (23). Through the SASP, SnCs can contribute to treatment-induced side effects such as myelosuppression, fatigue, and cardiovascular dysfunction (24). Thus, for cancer treatment, inhibiting the induction of senescence is detrimental, whereas promoting posttreatment SnC clearance is beneficial, timing being critical. Therefore, targeting SnCs, with senotherapeutics, including senomorphics (small molecules that partly suppress senescence phenotypes such as the SASP without cell killing) and senolytics (small molecules that induce SnCs death), is an emerging strategy for cancer treatment (25,26).

Understanding the molecular pathways that regulate senescence in cancer can generate novel insights to guide the discovery of unique anticancer agents and molecular biomarkers. This approach may spur the development of novel “one-two punch” cancer treatments consisting of agents that induce tumor cell senescence followed by senolytics to selectively clear SnCs (27,28) in tumor and normal tissue, a strategy that has the potential to improve therapy and concurrently mitigate many treatment-related side effects (24). This commentary summarizes the research discussions on important knowledge gaps to exploit TIS (Figure 1, A) and one-two punch cancer therapy on patient outcomes (Figures 1, B and 2) at the National Cancer Institute (NCI) Workshop on Radiation, Senescence, and Cancer (August 10-11, 2020, Bethesda, MD, USA).

**Novel Cellular, Molecular, and Epigenetic Mechanisms of Senescence**

Whether senescence is a state of permanent growth arrest or reversible is controversial (29,30). Senescence is one avenue whereby tumor cells evade the direct cytotoxic impact of therapy, thereby allowing for prolonged survival in a dormant state, with the potential to recover self-renewal capacity and contribute to disease recurrence (31). Although unrepaired DSBs are a well-recognized trigger of senescence, it can also be acquired in the absence of DNA damage response (DDR) or following DDR pathway activation in the absence of DNA damage (32). Senescence can occur after treatment with inhibitors of CDK4/6 (33,34), Polo and Aurora kinases (34,35), histone deacetylases, and other epigenetic modifiers (36). Furthermore, cells may re-enter the cell cycle subsequent to a prolonged senescence arrest to produce progeny with chromosomal instability or a cancer stem cell-like phenotype (20) providing a survival advantage. Therefore, it is important to distinguish “irreversible senescence arrest” from “senescence-like arrest” (37), because the cells reentering the cell cycle after senescence-like arrest may contribute to treatment failure.

Although TIS is commonly associated with unrepaired DSBs, it may also be induced after damage to mitochondria (38-41), which in turn leads to increased production of reactive oxygen species (42) and DNA damage (43). Senescence in quiescent endothelial cells can be induced through 2 independent pathways: 1) activation of DDR and p53 and 2) dysfunction of mitochondria (41,44). In addition, other mitochondrial and cytoplasmic metabolic pathways, including glycolysis and glutaminolysis, are critical mediators of DSB repair and DNA damage checkpoint responses that regulate senescence. The hexosamine biosynthetic pathway and downstream protein O-GlcNAcylation are attractive druggable targets to modulate radiation-induced senescence (45,46). Therefore, studies on regulation of the mitochondrial and other metabolic pathways impacting chromosomal integrity and senescence are essential for the development of novel mitigators of therapy-induced toxicities.

There is now evidence for metabolic (47) and stem cell-like remodeling (20,48) among SnCs and for the presence of an immunogenic switch that renders SnCs susceptible to an adaptive T-cell attack (49). Given this dynamic nature of the senescent state and the occasional cell-cycle reentry of previously SnCs, plasticity-associated functional capabilities may become particularly relevant to selective senescence escape and tumor represenprogression.

Gopal and colleagues (50) used single-cell RNA-sequencing (RNAseq) and fluorescence reporters representing distinct transcriptional cell states to model cell-state dynamics and TIS at the single-cell level. They found that phenotypic switching during chemotherapy, influenced by a range of possible senescence scenarios, caused the persistence of quiescent tumor programs. Some of these cells subsequently reverted to more proliferative states with variable time intervals across individual tumors, resulting in the development of treatment resistance and aggressive recurrence. Understanding distinct cell states and how single-cell behaviors establish phenotypic equilbrium in cancer populations and the role that SnCs play in this process is pivotal in elucidating the critical cell fate switches that may underlie treatment resistance (workshop presentations: “Senescent Cells as both Drivers and Suppressors of Radiation-Induced Cancers,” J. Campisi, Buck Institute, Novato, CA, and “Animal Models to Study the Role of Senescence in Diseases and Cancer,” Jan van Deursen, Rochester, MN).
Nevertheless, therapeutic targeting of SnCs is a balancing act that must not affect the beneficial effects of senescence while targeting pathways of pro-tumorigenic and pathological senescence. Future studies could evaluate strategies to integrate senolytic therapies effectively into aggressive anticancer regimens to reduce late toxicities while enhancing cure.
Many months after initiating one-two punch cancer therapy with personalized adaptive tumor therapy. To therapeutically exploit and benefit from the differences in response to treatment between tumor and normal tissue for the best patient outcome, factors that should be considered for pretreatment planning include tumor molecular profiling, tumor heterogeneity, imaging, identification of target(s), metabolic status, and planned integrated biomarkers for tumor diagnosis and treatment matching (130). Similar profiling of normal tissue response to treatment may include determination of genetic susceptibility, immune status, stromal tissue subsets, the impact of the anatomical location of the tumor on normal tissue, metabolic status, and biomarkers that predict response and adverse effects. In a one-two punch therapy, punch 1 may include spatially targeted radiotherapy (eg, dose-boost to hypoxic regions), molecularly targeted drugs, and/or immune therapy to the tumor, which will induce TIS in the tumor, stroma, and bystander tissue. Thus, tumor, stroma, and bystander tissue all need to be evaluated for TIS for the second punch to be successful. Biomarker-driven TIS evaluation will be essential to optimize immune modulation, dose, and schedule of the second punch with a suitable senolytic. Along with dynamic adaptive tumor targeting (with drugs, immune modulators, and radiation), the use of different types of senolytics may be necessary to address spatial, temporal, and tissue heterogeneity among tumors and senescent cells. Repeat treatment courses (punch n) with senotherapeutics (senolytics or senomorphics) may be necessary to prevent tumor recurrence, drug resistance, plasticity, and normal tissue injury and mitigate and/or treat adverse effects months to years after completing the one-two punch therapy for optimal tissue remodeling and tissue function restoration. Dotted boxes represent current biomarkers and future opportunities to develop diagnostics or therapeutics for precision medicine in TIS. Tissues are indicated by the colors red (tumor), green (normal tissue), blue (stroma and immune related to tumor), and brown (bystander tissue). The figure was created with BioRender.com. Rx = prescription; TIS = therapy-induced senescence.

**Heterogeneity of SnCs**

Heterogeneity among SnCs is contextual, influenced by the cell type and tissue of origin, the nature of the insult causing senescence, and the elapsed time after the insult occurs. Characterization of SnC heterogeneity is fundamental to understanding its role in tumorigenesis and developing cancer treatments, such as one-two punch cancer therapy (27,28) (detailed below). Box 1 provides some important considerations regarding SnC heterogeneity.

First, heterogeneity of SnCs is reflected in their ability to employ different senescent cell anti-apoptotic pathways (SCAPs) (51,52). For example, senescent endothelial cells rely on the anti-apoptotic protein Bcl-xL for survival and thus are sensitive to Bcl-xL inhibitors, whereas senescent adipocyte progenitors are more sensitive to a pan-tyrosine kinase inhibitor, Dasatinib (52).

Second, many SnCs can escape growth arrest by a variety of mechanisms. For example, in TIS lymphoma cells, inactivation of the H3K9 histone methyltransferase, Suv39h1, or p53 can reverse their growth arrest (11,20,53). Similar observations were made in senescent melanocytes induced by RAS/BRaf oncogenes after ectopic transfection with the lysine-specific demethylase-1 and the Jumonji C domain-containing histone demethylase, JMJD2C (53). Infrequent but spontaneous escape from TIS is seen in breast, non-small cell lung, colon, and ovarian cancer cells after therapy (54). Senescence escape is often associated with high expression of CDC2 and polyploidy (55). TIS cells can reprogram to acquire stemness and become more tumorigenic and drug resistant after senescence escape (54). Thus, strategies to induce and subsequently remove residual SnCs at the right time after treatment may improve therapeutic efficacy and help mitigate treatment morbidity, thus improving patient quality of life (QOL).

Third, the SnC transcriptome is highly heterogeneous and can exert opposite effects on tumorigenesis and response to therapy. The composition and quantity of individual SASP factors secreted by SnCs can vary among cell types and depend on the stimuli (8,17,56). The comprehensive soluble SASP atlas of senescent human fibroblasts induced by radiation, RAS overexpression, or atazanavir (a HIV protease inhibitor), and radiation-induced senescent renal epithelial cells indicated that only 17 soluble SASP factors are shared among many SnCs, whereas several other factors varied depending on tissue type and insults (8). In contrast, mesenchymal stromal cells exposed to different stressors showed common senescent phenotypes characterized by 4 classes of SASP components among several phenotypes: extracellular matrix and cytoskeleton and/or cell junctions, metabolic processes, redox factors, and regulators of gene expression (57). Specific SASP factors can modulate response to therapy. The SASP factors IL-1α, IL-6, TGF-β, CXCL1, and CXCL2 secreted by oncogene-induced senescence in human...
Box 1. Important gaps and considerations in the understanding of the heterogeneity of therapy-induced senescence (TIS)

- **TIS is heterogeneous and context (e.g., tissue of origin, nature of stress, and time after insults) dependent, and therefore, characterization in a variety of contexts is important for the development of novel approaches to cancer therapeutics such as one-two punch therapy.**
- **Senescent cell (SnC) heterogeneity is also reflected in the use of different SnC anti-apoptotic pathways to resist cell death.**
- **Many TIS-induced SnCs escape from growth arrest, which can result in the acquisition of plasticity, stemness, tumorigenic, and aggressive growth phenotypes.**
- **SnC transcriptomes are also heterogenous, can exert contextually dichotomous effects on responses to therapy, and tumorigenesis.**
- **Specific senescence-associated secretory phenotype factors can modulate the responses of tumor cells and normal tissues to therapy, resulting in inhibition and/or promotion of tumorigenesis and induction of normal tissue injury.**
- **The creation of a comprehensive atlas of SnCs may accelerate the discovery and development of novel biomarkers and senotherapeutics as next-generation anticancer agents to achieve better outcomes for cancer patients.**
- **Time of administration of senotherapeutics may be an essential determinant in the efficacy and toxicity profiles of anticancer agents.**

Senotherapeutics

The advent of senotherapeutics over the last 5 years has enabled many proof-of-concept studies for age-related diseases (66,67,70,71). Although senomorphics, such as the mTOR.
inhibitor rapamycin (72), JAK1/2 inhibitor ruxolitinib (73), and BET inhibitor JQ1 (74) may also be useful, the focus has been on senolytics. Table 1 provides a summary of senotherapeutics currently under various stages of development. Over the last decade, many targets for senolytics have been discovered, and several senolytics have been tested in preclinical models. However, the translation of senolytics to the clinic has been challenging for a number of reasons. These include, but are not limited to, SnCs heterogeneity across different tissues, organs, and model systems; the selectivity of drugs to deleterious SnCs; systemic toxicities; and development of drug resistance (75).

Box 2 summarizes important pitfalls, challenges, and opportunities underlying the development of senolytics as anticancer agents for clinical use.

In mouse models, senolytics such as Navitoclax (ABT-263) selectively eliminate TIS cells and prevent or delay cancer relapse and metastasis (24,76). ABT-263, a Bcl-2/Bcl-xL inhibitor, used as an adjuvant therapy with radiation increases the survival of glioblastoma multiforme tumor-bearing mice by eliminating senescent astrocytes with a tumor-promoting role (workshop presentation: “Prevention of Glioblastoma Recurrence after Radiotherapy by Elimination of Senescent Astrocytes,” Sandeep Burma, University of Texas Health Science Center, San Antonio, TX). Similarly, DNA-replication kinase CDC7 inhibitor–induced senescent liver and lung cancer cells with TP53 mutations can be selectively cleared by mTOR inhibitors, and the combination of inhibitors of CDC7 and mTOR statistically significantly reduced the tumor burden and increased survival in liver cancer xenograft mouse models (77). Recently, Ruscetti et al. (12) demonstrated in a pancreatic ductal adenocarcinoma model, in which combination therapy–induced senescence (MEK and CDK4/6 inhibitors targeting oncogenic signaling) triggers SASP-dependent vascular remodeling, facilitating chemotherapy uptake and SASP-mediated endothelial activation, driving T-cell infiltration into the tumors, and potentiating PD-1 blockade.

Some approaches described below to reduce toxicities and increase efficacy are noteworthy. First, taking advantage of the high expression of SA-β-gal in SnCs, gemcitabine, ABT-263, and duocarmycin have been converted into promising prodrugs with galactose as a pro-moiety (78–80). Such prodrugs are preferentially activated in SnCs by conversion into the active parent drug by SA-β-gal resulting in targeting of SnCs. However, activated macrophages also express SA-β-gal (81), so such agents will not likely be fully selective against SnCs. The second is to improve selectivity by constructing proteolysis targeting chimeras (PROTACs; heterobifunctional molecules that link a ligand for a protein of interest to an E3 ligase ligand) (82). To improve the potency and reduce the severe thrombocytopenia induced by ABT-263, taking advantage of the low expression levels of E3 ligase Cereblon (CRBN) in platelets, PROTAC PZ15227, constructed by linking ABT-263 to a CRBN ligand pomalidomide, has shown improved efficacy and reduced toxicities compared with ABT-263 (82). This approach could have broad applications if SnCs-specific E3 ligases could be identified (70,82). However, although this approach may protect against thrombocytopenia, it may not be effective in protecting against the neutropenia that can be induced by ABT-263 (83). Third, the design of dendrimer-conjugated Bcl-2/XL inhibitor, AZD0466, to optimize drug-release rate has shown promise in reducing cardiovascular toxicities and improved therapeutic index in preclinical models, which allowed its progression to clinical studies (84). However, as some senolytics are repurposed anticancer agents with known on- and off-target toxicities, their dose and scheduling need to be optimized for clinical applications.

Different types of SnCs use different SCAPs to resist apoptosis. Therefore, different TIS cells may require different senolytics (67). The continued discovery of new SCAPs, senolytic targets, and senolytics, and optimization of their dose regimen is essential. However, current efforts involving structure–activity relationships and lead optimization are lagging in senolytic discovery. Several biotechnology companies are currently developing new senolytics. Many of these efforts do not directly focus on developing senolytics as anticancer agents. However, it includes the development of senolytics to target the transition between quiescence and senescence state, modulators of senescence signaling pathways and DNA repair enhancers, approaches to enhance the efficacy and reduce toxicities using PROTACs, and antibodies to promote immune clearance of SnCs as well as novel discovery platforms (85). Similarly, there are also opportunities to repurpose radiation-effect modulators (86–88) as senotherapeutics. Thus, we anticipate the development of a steady pipeline of senotherapeutics in the near future, which may be used in one-two punch cancer therapy.

One-Two Punch Cancer Therapy

To target TIS, a novel one-two punch cancer therapy approach represents an exciting area of research (27,89), which is illustrated in Figure 1. B. Cancer therapies at clinical doses, while accomplishing tumor cell killing (first punch), also induce senescence in both tumor and normal tissues (13,54). SnCs are normally cleared by immune surveillance (90). Therapy-induced SnCs are heterogeneous and dynamic, also reflected in biomarkers, cellular plasticity, expression of SASPs and SCAPs, the tissue of origin, and cell lineage (8,13,52). Selective clearance of SnCs with a senotherapeutic (second punch) in tumors can prevent tumor relapse, metastasis, and development of resistance to treatment (27,89). Similarly, selective clearance of SnCs in normal tissue in a dynamic spatiotemporal environment will prevent, treat, and mitigate therapy-induced side effects and help restore tissue homeostasis. However, the time of administration of the “second punch” therapy will be important to improve efficacy. The report on the one-two punch approach to selectively eliminate chemotherapy-induced senescent lymphoma cells in mice was achieved by using a metabolic senolytic to block glucose utilization or autophagy (47).

Genetic and pharmacological clearance of TIS cells reduces side effects and inhibits tumor relapse and metastasis (24). A combination of chemotherapy and a senolytic therapy (eg, ABT263 or cardiac glycosides) is effective in the treatment of many cancer types in mouse models (77,91–94). The combined treatment with XL413 (a potent CDC7 inhibitor) and AZD8055 (mTOR inhibitor) resulted in pronounced growth inhibition of liver cancer (77). Recently, chimeric antigen receptor T cells targeting the cell surface protein, urokinase-type plasminogen activator receptor (which is broadly present in SnCs), were found effective as a senolytic in a KrasG12D; p53−/− lung adenocarcinoma mouse model demonstrating proof-of-principle for synthetic senolytic cell-based therapy (95).

Clearance of TIS cells can improve posttreatment QOL by mitigating cancer treatment-induced acute and late effects such as radiation-induced tissue fibrosis and chemotherapy-induced neuropathy (76,96,97). Interestingly, in some immunocompetent mouse tumor models, TIS is found to be beneficial, as it can also promote immune clearance of tumor cells through...
| Drug class                                      | Agent (company)                                      | Mechanism of action                                                                 | Developmental stage                        | Reference(s)                                      |
|------------------------------------------------|------------------------------------------------------|--------------------------------------------------------------------------------------|-------------------------------------------|--------------------------------------------------|
| Natural products and derivatives               | Alvespimycin, Geldanamycin, and Tanespimycin         | HSP inhibitors                                                                       | Optimization                              | Fuhrmann-Stroissnigg et al. 2017 (125)           |
|                                               | Curcumin analog, EF24                                 | Promotes degradation of anti-apoptotic Bcl-2 proteins                                 | Discovery                                 | Li et al. 2017 (126)                             |
|                                               | Piperlongumine and analogs                           | OXR1 and other                                                                       | Discovery                                 | Liu et al. 2018 (127), Wang et al. 2016 (128), Zhang et al. 2018 (129) |
|                                               | Cardiac glycosides: Digoxin, Ouabain, and Proscillaridin A | Na⁺/K⁺ ATPase inhibitor                                                               | Drug repurposing                          | Guerrero et al. 2019 (93), Triana-Martinez et al. 2019 (94) |
|                                               | Fisetin                                               | Blocks PI3K/AKT/mTOR pathways                                                        | Clinical trials:                         | Zhu et al. 2017 (71)                             |
|                                               |                                                      | Frail elderly (NCT03675724)                                                          |                                           | Yousefzadeh et al. 2018 (130)                    |
|                                               |                                                      | Osteoarthritis (NCT04210986)                                                         |                                           |                                                  |
|                                               |                                                      | Chronic kidney disease, Diabetes mellitus, and diabetic nephropathies (NCT03325322) |                                           |                                                  |
|                                               |                                                      | Mild cognitive impairment (NCT02741804)                                               |                                           |                                                  |
|                                               |                                                      | COVID-19 (NCT04476953)                                                              |                                           |                                                  |
|                                               | Quercetinb                                            | Activates estrogen receptors and inhibits PI3 kinase                                  | Clinical trials:                         | Zhu et al. 2015 (67)                             |
|                                               |                                                      | Alzheimer’s disease (NCT04063124)                                                    |                                           |                                                  |
|                                               |                                                      | Chronic kidney disease (NCT02848131)                                                 |                                           |                                                  |
|                                               |                                                      | Hematopoietic stem cell transplant (NCT02652052)                                     |                                           |                                                  |
|                                               |                                                      | Skeletal health in older humans (NCT04313634)                                        |                                           |                                                  |
|                                               | A1155463 (Abbvie, North Chicago, IL)                 | Bcl-xL inhibitor                                                                      | Preclinical tool compound                 | Zhu et al. 2017 (71)                             |
|                                               | A1331852 (Abbvie, North Chicago, IL)                 | Bcl-xL inhibitor                                                                      | Preclinical tool compound                 | Zhu et al. 2017 (71)                             |
|                                               | Navitoclax (ABT-263) (Abbvie, North Chicago, IL)     | Bcl-2/Bcl-xL inhibitor                                                                | Preclinical                               | Zhu et al. 2015 (67), Chang et al. 2016 (97)     |
|                                               | ABT-737 (Abbvie, North Chicago, IL)                  | Bcl-2/Bcl-xL inhibitor                                                                | Preclinical                               | Yosef et al. 2016 (131)                          |
|                                               | Dasatinibb                                           | Pan receptor tyrosine kinase inhibitor                                               | Clinical trials:                         | Zhu et al. 2015 (67)                             |
|                                               |                                                      | Alzheimer’s disease (NCT04063124)                                                    |                                           |                                                  |
|                                               |                                                      | Chronic kidney disease (NCT02848131)                                                 |                                           |                                                  |
|                                               |                                                      | Hematopoietic stem cell transplant (NCT02652052)                                     |                                           |                                                  |
|                                               |                                                      | Skeletal health in older humans (NCT04313634)                                        |                                           |                                                  |
|                                               | JQ1                                                  | BET inhibitor                                                                         | Preclinical                               | Tasdemir et al. 2016 (74)                        |
|                                               | P5091 (DFCI, Boston, MA)                             | USP7 inhibitor                                                                        | Discovery                                 | He et al. 2020 (132)                             |
|                                               | Panobinostat                                         | Pan HDAC inhibitor                                                                    | Unknown                                   | Samarawera et al. 2017 (133)                     |
|                                               | Proxofimc                                            | FOXO4/P53 protein interaction inhibitor                                               | Preclinical                               | Baar et al. 2017 (134)                          |
|                                               | UBX010 (Unity Biotechnology, South San Francisco, CA) | MDM2/p53 protein interaction inhibitor                                               | Preclinical                               | Vilgelm et al. 2019 (61)                         |
|                                               |                                                      | Bcl-2/Bcl-xL inhibitor                                                                | Clinical trial terminated                 | Vilgelm et al. 2015 (62)                         |
| (continued)                                   |                                                      |                                                                                      |                                           | Jeon et al. 2017 (103)                           |
|                                               |                                                      |                                                                                      |                                           | Kirkland et al. 2020 (102)                       |
COMMENTARY

The importance of senescence in cancer therapy:

SnCs}\textsuperscript{a}

SASC factors along with clearance of SnCs (34,60,63,98). The underlying causes of the differential effects of SnCs on tumorigenesis and response to therapy have not been fully elucidated. For example, SnCs induced immediately after therapy and those that accumulate over time may have different effects regarding antitumor immunity and tumor relapse, metastasis, and drug resistance (17). In addition, the SASP itself may change over time.

The first report on the combination of dasatinib and quercetin (D+Q) as an effective senolytic therapy in a preclinical model was discovered using a hypothesis-driven, mechanism-based, and bioinformatics approach (67). The results of 2 early phase clinical trials have now been published: 1) the first-in-human open-label pilot study (NCT02874989) demonstrated the feasibility and provided initial evidence that senolytic intervention with the above combination in participants with idiopathic pulmonary fibrosis can alleviate physical dysfunction (99), and 2) subsequently, it was demonstrated that treatment with D+Q administered to subjects with diabetic kidney disease statistically significantly reduces the SnCs burden (NCT02848131) (51,100). Thus, several senolytics, including D+Q (51,67,100), fisetin (71), UBX0101 (101), and UBX1325 (102), have now progressed into clinical trials (see Table 1), and some also have progressed to phase 2 studies.

The timing of senotherapeutic administration is a key determinant of the efficacy of the one-two punch strategy, because surveillance of SnCs is executed by cytokines and chemokines; release of which is time dependent. For example, the role for surveillance of SnCs is executed by cytokines and chemokines; prominent of the efficacy of the one-two punch strategy, because (51,100). Thus, several senolytics, including D+Q (51,67,100), fisetin (71), UBX0101 (101), and UBX1325 (102), have now progressed into clinical trials (see Table 1), and some also have progressed to phase 2 studies.

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Concluding Thoughts: a Translational Perspective

Promises, Pitfalls, and Barriers for Clinical Translation of Senotherapy and One-Two Punch Cancer Therapy

Increased expression of biomarkers associated with but not specific for SnCs is detected in numerous human cancers. For example, increased expression of p16\textsuperscript{INK4a} is associated with an increased risk of tumor relapse and poor prognosis in breast cancer (104,105), and the senescence-associated gene signature in peritumoral tissue correlates with shorter recurrence-free survival in hepatocellular carcinoma (11). In contrast, in malignant pleural mesothelioma patients after chemotherapy, increased expression of p16\textsuperscript{INK4a} is associated with better survival (106). This incongruity may reflect human tumor heterogeneity in response to treatment. Therefore, the juxtaposed roles of SnCs in tumorigenesis and treatment response may be contextual; it may be dependent on tumor and tissue type of origin. In this regard, our understanding of SnC heterogeneity is limited by our inability to spatiotemporally and unambiguously detect, characterize, and monitor SnCs in patients during the course of treatment because of the lack of reliable SnC biomarkers and tools to detect them (68).

Since discovery of the first potential senescence biomarker detectable in normal human skin (4), it has become clear that there are no universal senescence-specific biomarkers, whether in normal (young or old) or malignant tissue. Hence, there has been a concerted effort to develop biomarker panels for identifying senescent cells in vivo (6,68,107,108). Among the recognized senescence, biomarkers include CDK inhibitors 2 A (p16/CDKN2A) and 1A (p21/CDKN1A), which suppress cell proliferation (8). However, recent tissue diagnostic array studies of cells expressing p16 and p21 indicated that although different organs express different levels of proteins as a function of age across the human life span, some tissues such as muscle do not appear to have either of these markers (109). Moreover, senescence biomarkers are highly variable, whether in normal or tumor tissue, and they can depend on the cell type, senescence inducer, and time after senescence induction (8). Therefore, reliable SnC biomarkers to spatially identify and longitudinally track SnCs are urgently needed (eg, noninvasive imaging) for successful translation. Of note, Dr Jesus Gil showed preliminary data from his collaboration with Dr Lars Zender at the University of Tubingen, Germany, on using positron emission tomography imaging in the brain of a glioblastoma multiforme patient to monitor TIS cells and their clearance by Digoxin (workshop presentation: “Strategies to Target Senescence,” Jesus Gil, Imperial College, London, UK).

However, a major limitation to the clinical translation of senotherapeutics is the inability of animal models (human xenografts and/or genetically engineered models) to recapitulate fully key mechanisms and predict outcomes for human patients. In this regard, patient-deriv ed xenograft or spontaneous tumor models may serve to better recapitulate tumor

Table 1. (continued)

| Drug class                      | Agent (company)                          | Mechanism of action       | Developmental stage                          | Reference(s) |
|---------------------------------|------------------------------------------|---------------------------|----------------------------------------------|--------------|
| Senescence cell-targeting       | UBX-1325 (Unity Biotechnology, South San Francisco, CA) | DNA alkylation agent      | Clinical trial: Diabetic macular edema (NCT04537884) | Guerrero et al. 2020 (79) |
| prodrugs                        | Duocarmycin galactose conjugate          | Nucleoside analog         | Discovery                                    | Cai et al. 2020 (80) |
| PROTACs                         | ARV825 (Avnias Inc, New Haven, CT)       | BET family protein degrader | Discovery                                    | Waikita et al. 2020 (135) |
|                                 | P215227 (University of FL, Gainesville, FL) | Bcl-2/2xL degrader        | Discovery                                    | He et al. 2020 (82) |

\textsuperscript{aThis is not a comprehensive list of senotherapeutics but provides some examples of classes of drugs developed as senotherapeutics.}

\textsuperscript{bUsed in combination with each other.}

\textsuperscript{cAll drugs in the Table have been reported to have anticancer effects except for Proxofin.}
Box 2. Pitfalls, challenges, and opportunities in the development of senolytics as anticancer agents for clinical use

- Discovery. Current efforts involving structure-activity relationships and lead optimization are lagging in senolytic discovery. Medicinal chemistry-based research is needed.
- Experimental models. Suitable, reliable, and efficient in vitro and in vivo models for evaluation of the safety and efficacy of senolytics are needed.
- Specificity. Further characterization of senescent cells (SnCs) is necessary to allow improved targeting of “harmful SnCs” with sparing of immune-modulating SnCs.
- Toxicities. Most senolytics developed from anticancer agent pipelines demonstrate on- and off-target toxicities.
  - For example, Bcl-xL specific inhibitors A1155463 and A1331852 can cause severe thrombocytopenia because platelets depend on Bcl-xL for survival. Similarly, Bcl-xL/Bcl-2 dual inhibitors ABT-263, ABT-737, and UBX-1325 can also cause severe neutropenia because neutrophils depend on Bcl-2 for survival. Improved selective targeting with proteolysis targeting chimeras may reduce toxicities.
  - Senolytic prodrugs rely on senescence-associated β-galactosidase for SnC selective activation. However, macrophages also express high levels of β-gal. To reduce off-target toxicities by using prodrugs, however, SnCs specific activation enzymes need to be identified.
- Time of administration. The effectiveness of senolytics may depend on the time of administration following cancer therapy. Senolytics are most effective when administered in a hit-and-run fashion reducing potential toxicities and off-target effects.
- Drug resistance. SnCs in tumors may not be in a state of “permanent growth arrest” but can subsequently acquire “stemness” and/or plasticity, leading to drug resistance and metastasis.
- Mechanisms of action. A clearer understanding of the mechanisms of action of senolytics is necessary, particularly for those natural product senolytics or their derivatives including quercetin, fisetin, piperlongumine and analogs, curcumin analog EF-24, and cardiac glycosides digoxin, ouabain, and procollaredin A.
- Intellectual property rights. Most senolytics are obtained from natural products, or derivatives of natural products, repurposed anticancer agents, or off-patent. Therefore, they are not commercially viable for pharmaceutical industries.
- Regulatory barriers. Some senolytics are botanical products or derivatives, which need to be registered as investigational drugs with the US Food and Drug Administration. Because of the heterogeneity and possible uncertainty about active constituents, the efficacy of the drugs can vary from batch to batch.
- Clinical studies.
  - Safety requirement for senolytics is relatively high as these drugs are intended to treat elderly patients or patients undergoing cancer treatments who have low tolerability to toxicity. Although multiple trials of senolytics have begun, it will take several years to complete these trials.
  - Data on senescence from clinical studies are relatively sparse.
inhibitors were also found to preserve hematopoietic stem cell function in NSCLC patients treated with etoposide and carboplatin without reducing treatment efficacy (118). Further, CDK4/6 inhibitors exhibit excellent in vivo pharmacology and tolerability in patients with metastatic triple-negative breast cancer (119). Trilaciclib is now approved to treat hormone receptor-positive and HER2-negative advanced breast cancer through induction of G1/S cell cycle arrest and senescence in tumor cells (120,121). In combination with MDM2 inhibition, preclinical studies show CDK4/6 inhibition combined with MDM2 inhibition is quite effective in inhibiting NRAS mutant melanoma tumor growth by induction of senescence, followed by immune-mediated tumor cell clearance (61). Studies with CDK4/6 inhibitors are examples of several possibilities of combining agents with differential effects and exploiting differential vulnerabilities in tumor and normal cells to improve treatment efficacy in cancers and decrease toxicities. However, tissue-specific heterogeneity in TIS is a challenge to be overcome and an opportunity to improve patient outcomes.

Discussion

Since the discovery of cellular senescence by Hayflick and Moorhead (122), senescence is now understood to be a fundamental biological process that governs a variety of pathophysiological functions and diseases including cancer. As the number of cancer survivors is growing because of ever-improving treatments, focus on treatment-induced adverse effects has become even more imperative, as is the opportunity to study and mitigate TIS to improve further the treatment and also the QOL of survivors. With SnCs emerging as an anticancer target, the NCI is developing promising strategies to prevent, slow, and reverse senescence for each drug and malignancy? Addressing such questions is vital to design appropriate clinical trials and get the full benefit from the one-two punch therapy.

By understanding TIS, we can anticipate a steady pipeline of novel biomarkers and senotherapeutics and can plan to translate senotherapeutics into next-generation anticancer agents. Furthermore, the identification of novel biomarkers should enable more personalized treatments, which could be more specific to a given patient, type of malignancy and treatment. Successful translation of senotherapeutics will ultimately involve obtaining regulatory approval from the US Food and Drug Administration. Studies of senotherapeutics present unique regulatory challenges, especially when used as combined treatment modalities with chemotherapy and radiation: different toxicity profiles and timelines are likely for the development of adverse acute and late effects between systemic therapies and radiation. Cancer therapeutic-senotherapeutic drug combinations will also present challenges in clinical trial design. Therefore, an early interaction with the US Food and Drug Administration is important if the laboratory data are compelling to consider general aspects related to the development and translation of senotherapeutics and to obtain further guidance.

Funding

This work was supported by the National Cancer Institute (P30CA060553 and R37CA222924 to MEA; R01 CA197796 and R01 CA246807 to SB; R01CA214025-01 to D Gius; R01CA239706 to D Gewertz; AG063543, ES029603, AG056278 and P01AG062413 to LJN; R01CA164492 and R01CA217182 to SJK; CA116021 to AR; R37CA233770 to AEV; R37AG03925, P01AG062413 and R33AG061456 to JK; P30CA08748 to PBR and SWL; K12 CA184746 to PBR, R01CA211963, R01CA219836, R01CA242003, and R01AG063801 to GZ and DZ; and R01CA218596 (Ewing/S Brown) and R21CA205660 (Jae Ho Kim). Further, laboratory studies done in Dr Citrin’s laboratory are supported by the intramural research program of the National Cancer Institute (grant BC10850). Dr Burma is also supported the National Aeronautics and Space Administration (NNX16AD78G). Dr Gius is also supported by the Avon Foundation for Breast Cancer Research and the Lynn Sage Cancer Research Foundation and by the Cancer Prevention and Research Institute of Texas (CPRIT) grant no. RR20012. Studies done in Dr Gil’s laboratory are supported by the Core support from MRC (MC-U120085810) and grants from Worldwide Cancer Research (18-0215) and CRUK (C15075/A28647). Studies done in Dr Kirkland’s laboratory are also supported by the Alzheimer’s Association Part the Cloud Program, Robert and Arlene Kogod, the Connor Group, Robert J. and Theresa W. Ryan, and the Noaber Foundation. Studies done in Dr O’Loghlen’s lab were funded by the BBSRC (BB/P000223/1) and Barts Charity Grant (MGU0497). Studies done in Dr Richmond’s lab was also funded by a VA MERIT Award (101BX002301) and a VA Senior Research Career Scientist Award. Dr Schmitt’s laboratory is supported by grants to C.A.S. from the Deutsche Krebshilfe (No. 110678), the BMBF e: Med program project SeneSys (No. 03IL0189A), the Deutsche Forschungsgemeinschaft DFG (GO 2688/1-1 | SCHM 1633/11-1, SCHM 1633/9-1).
Box 3. Key knowledge gaps in therapy-induced senescence (TIS) research for the discovery and development of senotherapeutics as a novel anticancer strategy and to guide the personalization of cancer treatment

- Can we use radiation/chemotherapy as a model to induce and study cellular perturbations that cause senescence and/or cancer and improve outcomes?
- Which is the critical type and level of biological damage (DNA mainly) shifting the balance toward senescence or apoptosis?
- What are the differences between TIS in normal tissue and the tumor microenvironment?
- Is induction and promotion of senescence treatment specific? Is the response uniform or heterogeneous across tumor types?
- Is the time frame of senescence occurrence after treatment dependent on treatment and tumor types?
- Can TIS be used to unravel the intersecting mechanisms of carcinogenesis and senescence?
- Will TIS be a viable approach to discover and develop novel senotherapeutics that can also serve as novel cancer therapies?
- Can senotherapeutics improve the efficacy of radiation and chemotherapy without compromising antitumor immunity?
- Is the timing of senotherapeutics following treatment important for determining the efficacy of one-two punch cancer therapeutics?
- Are there any biomarkers that can predict or monitor the effectiveness of senotherapeutics?
- How TIS effects the release of heterogeneous senescence-associated secretory phenotypes (SASPs) formed by metabolites, proteins, and vesicles and how the heterogeneous HSP regulates the tumor microenvironment?

Notes

Role of the funders: The NCI convened the workshop, “Radiation, Senescence, and Cancer” (August 2020) that informed the manuscript’s scope. The funders had no role in the writing of this commentary or the decision to submit it for publication. The manuscript was reviewed and approved for submission through the NCI Division of Cancer Diagnosis and Treatment manuscript clearance process.

Disclosures: The main organizers of the workshop, Drs. Pataje G. Prasanna, Deborah E. Citrin, and C. Norman Coleman are employees of the US government and have nothing to disclose and performed this work in the interest of general public. Drs. Jeffrey Hildesheim, Mansoor Ahmed, Sundar Venkatachalam, Gabriela Riscuta, Dan Xi, and Mitchell S. Anche are employees of the US government, and other authors, Drs. David Gius, Goronzly, O’Loghlen, and Mendonca have nothing to disclose. Drs. Guangrong Zheng and Daohong Zhou are inventors of three pending patent applications for use of Bcl-xL PROTACs as senolytic and antitumor agents and are co-founders of and have equity in Dialectic Therapeutics, which develops Bcl-xL PROTACs to treat cancer. Dr Abazeed discloses research grant and travel support from Siemens Healthcare and research grant, travel support, and honorarium from Bayer AG in subject matter unrelated to this work. Dr Jesus Gil has acted as a consultant for Unity Biotechnology, Geras Bio, Myricx Pharma, and Merck KGaA; owns equity in Unity Biotechnology and Geras Bio, and is a named inventor in Imperial College and MRC patents related to senolytic therapies. Dr. Judith Campisi, Jan van Deursen, and Daohong Zhou are co-founders and stockholders of Unity Biotechnology that develops senolytics to treat age-related diseases. Dr James Kirkland has a financial interest related to this work. Patents on senolytic drugs are held by the Mayo Clinic’s Radiation Research Program in planning the workshop.

Acknowledgements: The authors would like to acknowledge the assistance of Drs. Michael G. Espey, Jeffrey Buchsbaum, Jacek Capala, Bhadrasain Vikram, and Ceferino Obcemia of NCI’s Radiation Research Program in planning the workshop.

Disclaimer: The views and opinions expressed in this article are those of the authors and do not necessarily reflect the views and the opinions of the institutes/organizations they represent.

Data Availability

The data underlying this article are available in the article.

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