The Thioamides Methimazole and Thiourea Inhibit Growth of \textit{M. avium} Subspecies \textit{paratuberculosis} in Culture

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Abstract

\textbf{Background:} Thyrotoxicosis is conceptualized as an “autoimmune” disease with no accepted infectious etiology. There are increasingly compelling data that another “autoimmune” affliction, Crohn disease, may be caused by \textit{Mycobacterium avium} subspecies \textit{paratuberculosis} (MAP). Like \textit{M. tb}, MAP is systemic. We hypothesized that some cases of thyrotoxicosis may be initiated by a MAP infection. Because other thioamides treat tuberculosis, leprosy and \textit{M. avium} complex, we hypothesized that a mode of action of some thioamide anti-thyrotoxicosis medications may include MAP growth inhibition.

\textbf{Methods:} The effect of the thioamides, thiourea, methimazole and 6-propo-2-thiouracil (6-PTU) were studied in radiometric Bactec culture, on ten strains of three mycobacterial species (six of MAP, two of \textit{M. avium} and two of \textit{M. tb} complex). Data are presented as “cumulative growth index,” (cGI) or “percent decrease in cumulative GI” (%ΔcGI).

\textbf{Principal Findings:} Methimazole was the most effective thioamide at inhibiting MAP growth. At 128μg/ml: MAP UCF-4; 65\%ΔcGI & MAP ATCC 19698; 90\%ΔcGI. Thiourea inhibited MAP “Ben” maximally; 70\%ΔcGI. Neither methimazole nor thiourea inhibited \textit{M. avium} or \textit{M. tb} at the doses tested. 6-PTU has no inhibition on any strain studied, although a structurally analogous control, 5-PTU, was the most inhibitory thioamide tested.

\textbf{Significance:} We show inhibition of MAP growth by the thioamides, thiourea and methimazole in culture. These data are compatible with the hypothesis that these thioamides may have anti-prokaryotic in addition to their well-established eukaryotic actions in thyrotoxic individuals.

Citation: Greenstein RJ, Su L, Brown ST (2010) The Thioamides Methimazole and Thiourea Inhibit Growth of \textit{M. avium} Subspecies \textit{paratuberculosis} in Culture. PLoS ONE 5(6): e11099. doi:10.1371/journal.pone.0011099

Editor: Leonardo A. Sechi, Universita di Sassari, Italy

Received April 5, 2010; Accepted April 23, 2010; Published June 14, 2010

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Funding: The authors thank Becton-Dickinson for providing the Bactec vials. Becton-Dickinson had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. No other extramural funds were obtained. This material is the result of work supported with resources and the use of facilities at the James J. Peters VAMC Bronx NY. Other than the JJP VAMC Bronx Research and Development Committee reviewing, and approving, our submitted research protocol the James J. Peters VAMC Bronx NY had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. This study was supported by the Bronx Veterans Medical Research Foundation, Inc. at the James J. Peters VAMC Bronx NY. The Bronx Veterans Medical Research Foundation, Inc. had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: RJG has submitted provisional patents based on the data that multiple medications used in the therapy of inflammatory diseases inhibit MAP in culture. STB was a member of the National Academy of Sciences of the USA panel that issued the Report “Diagnosis and Control of Johne’s Disease.” ISBN 0-309-08611-6. RJG and STB affirm that this does not alter their adherence to all the PLoS ONE policies on sharing data and materials. LS has no potential conflict of interest.

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Introduction

Prevailing medical dogma conceptualizes thyrotoxicosis as an “autoimmune” disease, with no universally accepted causative etiology. The mechanism of the thionamide class of anti-thyroid medications is accepted to be due to direct action on the hyperactive eukaryotic thyroid tissue. [1] There are suggestions that these autoimmune concepts should be readdressed. [2]

There is an intriguing, unexplained, association between thyroid hyperactivity and other “autoimmune” diseases, notably “inflammatory” bowel disease [3] including Crohn disease (CD) [4,5] and ulcerative colitis (UC.) [6]

As with thyrotoxicosis, the etiology of CD and UC is (are) not known. \textit{Mycobacterium avium} subspecies \textit{paratuberculosis} (MAP), causes a chronic wasting diarrheal disease in cattle called Johne disease [7], that is evocative of CD. There are increasing direct [8] & see [9] for review and circumstantial data [10,11] that MAP is zoonotic.[12] Intriguingly, the presence of MAP has been documented in a patient with thyrotoxicosis. [13]

It is of considerable interest that the thioamides ethionamide and prothionamide are used in the therapy of leprosy, tuberculosis and \textit{M. avium} complex infections diseases[14,15], and thiourea isoxyl is active against \textit{M. tb}. [16] Additionally
both of the antithyrototoxic medications Methimazole and Propothiouracil inhibit \textit{M. leprae} in the mouse footpad model.[17–19]

Accordingly, we hypothesized that anti-thyrotoxicosis thioamide medications in addition to their multifold well documented eukaryotic actions [1], may have prokaryote activity in thyrotoxic individuals. Specifically, we hypothesized that these medications may interfere with the growth kinetics of MAP. The clinical responses to these anti-thyroid medications are idiosyncratic. Therefore, we further hypothesized that any MAP culture inhibition would be strain and agent specific.

We herein report on the effect on the growth kinetics on MAP, of the thioamide anti-thyroid medications methimazole and 6-propo-2-thiouricil (6-PTU) as well as thiourea, (an integral structural component of both methimazole and 6-PTU;) \textit{M. leprae} cannot be grown in culture. [20] Therefore, as experimental control mycobacteria we studied the \textit{M. avium} and the \textit{M. tuberculosis} complexes.

**Methods**

This study was approved by the Research & Development Committee at the VAMC Bronx NY (0720-06-038) and was conducted under the Institutional Radioactive Materials Permit (#31-00636-07).

![Bacterial Culture](image)

Our Bactec® 460 (Becton-Dickinson Franklin Lakes NJ) \(^14\)C radiometric culture inhibition methods have previously been published in detail. [21–26] Because of interference with the assay [25], we do not use the detergent Tween 80 (recommended to prevent mycobacterial clumping) in culture. [27] Except for the amount of test agent, every vial has the identical concentration of all constituents (including identical 3.2% concentration of the dissolving agent, DMSO.) Vials are assayed on a daily basis, quantifying the amount of \(^14\)C released as \(^14\)CO\(_2\), by the integral detector in the Bactec 460. The data are obtained as a manufacturer determined, arbitrary Growth Units (GU) of 0-999.

In this study we evaluated ten strains of mycobacteria, six of which were MAP. Four MAP strains had been isolated from humans with Crohn’s disease. “Dominic” (ATCC 43545), “Ben” (ATCC 43544) (both originally isolated by R. Chiodini [28]) and UCF 4 and ST-5 (both gifts of Saleh Naser, Burnett College of Biomedical Sciences, University of Central Florida, Orlando FL,[8]). The other two MAP strains were from ruminants with Johne disease, ATCC 19698 and 303 (gift of Michael Collins Madison WI.) The \textit{M. avium} subspecies \textit{avium} strains (hereinafter called \textit{M. avium}) were ATCC 25291 (veterinary source) and \textit{M. avium} 101 [29]. To study the \textit{M. tuberculosis} complex, we used two BioSafety level 2 strains, Bacillus Calmette Guerin (BCG) \textit{M. bovis}}
Karlson & Lessel (ATCC 19015) and an avirulent M. tb strain; ATCC 25177 (all ATCC from ATCC Rockville MD).

The agents used to treat thyrotoxicosis that we studied were: 6-Propylthiouracil (6-propyl-2-thiouracil; 6-PTU; Sigma Cat # P3755) and methimazole (1-Methyl-2-imidazolethiol, 2-Mercapto-1-methylimidazole; Sigma Cat # M8506). We additionally studied Thiourea (Sigma Cat # T8656), an integral structural component found in both 5-PTU and methimazole (See Fig 1 in [1]). As an additional control, we studied a structural analog of PTU, 5-propyl-2-thiouracil (5-PTU; Sigma Cat # P0643.) Our positive antibiotic controls was monensin [24] and the negative control was the gluteraldehyde antibiotic phthalimide.[25]

Chemical were dissolved in DMSO, aliquoted, stored at -80°C, thawed, used once and discarded. (All Sigma, St Louis, MO.) Agents were studied at concentrations ranging from 0.25 to 128 µg/ml (See Figures.)

For clarity and ease of understanding data are presented in two ways. Graphically (Figures 1–4) we present data for individual mycobacteria from a single experiment. Data are presented as the cumulative Growth Index (cGI.) The same data are then manipulated mathematically (1) and are presented (see Tables 1–6) as change in growth kinetics as the “percent change from control cGI” (Increase as “%+ΔcGI” or Inhibition; “%−ΔcGI”). Each Table has data from a single chemical agent but every mycobacterium studied. This is in contrast to the Figures where each graph is for an individual mycobacterium.

During the course of these experiments, for technical reasons, the doses tested were progressively modified. There were four ranges. In every experiment the doses ranged from “Minimal” (0.25–2 µg/ml), to “Low” (4–8 µg/ml), to “Medium” 16–32 µg/ml and “Maximal” doses (64–128 µg/ml.) See individual Figures for actual dosage tested in each separate experiment. See [22] for

Figure 2. Shown are the cumulative Growth Indices (cGI) for the two MAP strains isolated from ruminants with Johne disease. The positive control is Monensin, and the negative control is Phthalimide. Note that 5-PTU is consistently the most effective agent, and 6-PTU exhibits no inhibition. Methimazole inhibits growth. Thiourea does not inhibit MAP 303.

doi:10.1371/journal.pone.0011099.g002
calculation of %ΔcGI. In the data presented in the Tables, the “Maximal” dose is always the 128μg/ml value.

Results

As previously [24], in this study we show that all MAP strains are inhibited by Monensin (Table 1 and Figures 1 & 2.) As previously [24], Monensin does not inhibit one of our two M. avium control strains (M. avium 101: Table 1 and Figure 3.) In our initial Monensin manuscript [24], we had only studied BCG from the M. tb complex. We now additionally study a Biosafety level II non-virulent strain of M. tb. ATCC 25177. Thus, we find that Monensin is even more inhibitory against M. tb. ATCC 25177 (>98% ΔcGI @ 4μg/ml. Table 1 & Figure 4) than it is against BCG.

The negative control that we use is Phthalimide, a glutarimide antibiotic that has no mycobacterial inhibition.[25] In this study, as previously, Phthalimide has no dose dependent inhibition against any of the mycobacterial strains we study (Table 2 and Figures 1–4.)

Thiourea, an integral structural component of both 6-PTU and methimazole, causes dose dependent inhibition of all four MAP strains isolated from humans (Table 3 and Figure 1). In contrast, thiourea does not inhibit either the two bovine MAP isolates (Table 3 and Figure 2) or any of our four mycobacterial controls species M. avium (Table 3 & Figure 3) or M. tb. (Table 3 & Figure 4).

Methimazole, causes dose dependent inhibition of all MAP strains (Table 4 and Figures 1& 2.) It is most inhibitory on a bovine MAP isolate ATCC 19698 (90%ΔcGI at 128μg/ml.) Methimazole has no inhibition on our mycobacterial controls,
M. avium (Table 4 & Figure 3) or M. tb complex (Table 4 & Figure 4).

At the doses tested, the anti thyroid medication 6-PTU causes no inhibition of MAP growth against any strain tested (Table 5 & Figures 1–4). As a control we compared this to a structural analog, 5-PTU. To our surprise, the control, 5-PTU markedly inhibits every mycobacterial strain we studied. (Table 6 and Figures 1–4.)

Discussion

Our data show that both thiourea and methimazole cause strain specific, dose dependent inhibition of MAP in radiometric culture. Thiourea is more active against the MAP strains isolated from humans and less active against MAP strains isolated from ruminants. Methimazole inhibits all MAP strains studied. In contrast, neither methimazole nor thiourea has any dose dependent inhibition against our M. avium and M. tb control strains. These data could explain the lack of a consistent response to the medical therapy of clinical thyrotoxicosis.

Other thioamides have antimycobacterial activity in leprosy, tuberculosis and in M. avium complex infections. [14–16] Methimazole and 6-PTU inhibit M. leprae,[17–19] In our assay neither Methimazole nor 6-PTU inhibits M. avium subspecies avium or the M. tb complex. We conclude that the inhibition of growth by these antithyroid thioamides is specific to MAP and M. leprae[17–19], but not to mycobacteria in general.

Our data show no inhibition by 6-PTU on any of the ten mycobacterial strains we evaluated. These data have multiple possible explanations. 6-PTU is actively concentrated in vivo by both lymphocytes (by 666%) [31] and thyroid tissue. [32,33] Thus the doses achieved in vivo may well exceed the concentrations used in our culture inhibition study.

Figure 4. Shown are the cumulative Growth Indices (cGI) for the two M. tb complex strains. The positive control is Monensin, and the negative control is Phthalimide. 5-PTU is consistently the most effective agent. Note that, uniquely in this study, 6-PTU inhibits the BioSafety level 2 avirulent strain of M. tb (ATCC 25177) that we study.
doi:10.1371/journal.pone.0011099.g004
In vivo there are multiple metabolites of 6-PTU. [34,35] Some of these 6-PTU metabolites may have anti-MAP activity. This would be analogous to 5-ASA inhibiting MAP in culture whereas intact sulfasalazine, a parent molecule of 5-ASA, has no antiMAP activity in culture. [22] We have not been able to identify a commercial source of any 6-PTU metabolites [34,35] to test in our culture inhibition system. However, we were able to obtain 5-PTU a structural analog of 6-PTU. 5-PTU markedly inhibits all ten strains of mycobacteria that we studied. We are unaware of any studies that have evaluated the safety or efficacy of 5-PTU in the therapy of thyrotoxicosis. Nor does this study attempt to correlate the clinical doses given and tissue levels achieved in vivo, with antimycobacterial activity in tissue culture.

The time required to achieve a clinical response in the therapy of tuberculosis [36], leprosy [37] and IBD [38] is months. Likewise, a clinical response to thioamide anti-thyrotoxicosis medication requires months. [1,2] This tardiness is ascribed to substantial reserves of thyroid hormone, which must be depleted before a clinical response can be observed. Our data are

### Table 1. Positive Control: Monensin.

| Dose Range µg/ml | MAP       | Bovine | M. avium | M. tb complex |
|------------------|-----------|--------|----------|---------------|
|                  | Human     |        |          |               |
|                  | Dominic   | UCF4   | Ben      | 19698 303     |
| Minimal          | −27%      | −92%   | −95%     | −76% −56%     |
| Low              | −48%      | −92%   | −98%     | −95% −78%     |
| Medium           | −84%      | −92%   | −98%     | −60% −92%     |
| Maximal          | −91%      | −94%   | −99%     | −90% −93%     |

Our positive control antibiotic causes dose dependent inhibition in all but M. avium 101.

Shown are the effect of the test agent as either enhancement (%ΔGI) or inhibition of growth (− = %ΔGI) compared to control vials containing the same concentration of DMSO. See Methods for calculating %ΔGI. The dose ranges are “Minimal” = 0.25–2 µg/ml; “Low” = 4–8 µg/ml; “Medium” = 16–32 µg/ml and “Maximal” = 64–128 µg/ml. In these Tables “Maximal” is always the 128µg/ml data. For actual dose tested in individual experiments see the Abscissa on individual Figures.

doi:10.1371/journal.pone.0011099.t001

### Table 2. Negative Control: Phthalimide.

| Dose Range µg/ml | MAP       | Bovine | M. avium | M. tb complex |
|------------------|-----------|--------|----------|---------------|
|                  | Human     |        |          |               |
|                  | Dominic   | UCF4   | Ben      | 19698 303     |
| Minimal          | −6%       | 3%     | 17%      | 28% 7% 15%    |
| Low              | −4%       | −9%    | 23%      | −11% 2% 51%   |
| Medium           | 9%        | −4%    | 24%      | 7% 61% 9%     |
| Maximal          | 11%       | −9%    | 32%      | 12% 73% 83%   |

Our negative control antibiotic has no inhibition in any of the ten mycobacterial strains tested. See Legend to Table 1 for explanation.

doi:10.1371/journal.pone.0011099.t002

### Table 3. Thiourea.

| Dose Range µg/ml | MAP       | Bovine | M. avium | M. tb complex |
|------------------|-----------|--------|----------|---------------|
|                  | Human     |        |          |               |
|                  | Dominic   | UCF4   | Ben      | 19698 303     |
| Minimal          | 3%        | −7%    | −31%     | −28% −13% 27% |
| Low              | −15%      | 5%     | −35%     | −33% 1% 4%  |
| Medium           | −20%      | −1%    | −20%     | −43% 4% 12% |
| Maximal          | −41%      | −23%   | −70%     | −56% −35% −1%|

The basic thioamide molecule causes dose dependent inhibition four MAP strains isolated from humans but none of the other six strains. See Legend to Table 1 for explanation.

doi:10.1371/journal.pone.0011099.t003
compatible with an alternative hypothesis. Mycobacteria replicate very slowly. We suggest that successful treatment of a mycobacterial trigger for thyrotoxicosis would take months to manifest clinically.

Other than a single report [13], to our knowledge an association between MAP and thyrotoxicosis has not been previously reported. This may be because detecting mycobacteria is not possible in some forms of mycobacterial diseases such as tuberculoid leprosy [39] or MAP in humans. [12] We suggest that to understand human MAP infections, more insights will be gained from analogies with leprosy [10,11] than with tuberculosis.

The thioamides used to treat thyrotoxicosis have anti *M. leprae* effects in an animal model. [17–19] We conclude this prokaryotic inhibition may have therapeutic implications in thyrotoxicosis. Our data are compatible with the hypothesis that some cases of "autoimmune" thyroid disease may be instigated by a mycobacterial, specifically we posit a MAP, infection.

### Author Contributions

Conceived and designed the experiments: RJG. Performed the experiments: RJG LS. Analyzed the data: RJG LS STB. Contributed reagents/materials/analysis tools: RJG STB. Wrote the paper: RJG.

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**Table 4. Methimazole.**

| Dose Range µg/ml | MAP | Human | UCF4 | Ben | ST-5 | 19698 | 303 | 25291 | 101 | 25177 | 19015 |
|------------------|-----|--------|------|-----|------|-------|-----|-------|-----|-------|-------|
| Minimal          |     | 2%     | 1%   | 4%  | −14% | 13%   | 22% | 34%   | 1%  | −2%   | 5%    |
| Low              |     | −16%   | −3%  | −20% | −41% | 14%   | 16% | 27%   | 3%  | −24%  | 5%    |
| Medium           |     | −7%    | −30% | −28% | −52% | −27%  | 5%  | 10%   | −1% | 4%    | 10%   |
| Maximal          |     | −57%   | −65% | −47% | −57% | −90%  | −46%| 31%   | 15% | −5%   | 7%    |

Note dose dependent inhibition in five of six MAP strains, but none of the four Control strains. See Legend to Table 1 for explanation.

**Table 5. 5P-2Thiouracil.**

| Dose Range µg/ml | MAP | Human | UCF4 | Ben | ST-5 | 19698 | 303 | 25291 | 101 | 25177 | 19015 |
|------------------|-----|--------|------|-----|------|-------|-----|-------|-----|-------|-------|
| Minimal          |     | −6%    | −16% | −11% | 27%  | 44%   | 55% | 20%   | −5% | −20%  | −2%   |
| Low              |     | −8%    | 6%   | −45% | 40%  | 32%   | 25% | 4%    | −6% | −34%  | −41%  |
| Medium           |     | −14%   | −20% | −85% | 25%  | −5%   | −39%| −60%  | −50%| −97%   | −71%  |
| Maximal          |     | −89%   | −81% | −96% | −69% | −94%  | −80%| −92%  | −97%| −99%   | −92%  |

With this structural analog of 6-PTU there is dose dependent inhibition in eight of ten strains and inhibition at 128 µg/ml in the remaining two. Note that *M. avium* 101 is inhibited. See Legend to Table 1 for explanation.

**Table 6. 6P-2Thiouracil.**

| Dose Range µg/ml | MAP | Human | UCF4 | Ben | ST-5 | 19698 | 303 | 25291 | 101 | 25177 | 19015 |
|------------------|-----|--------|------|-----|------|-------|-----|-------|-----|-------|-------|
| Minimal          |     | −4%    | 25%  | 34% | 54%  | 43%   | 58% | 32%   | −5% | −1%   | 8%    |
| Low              |     | −6%    | 26%  | 32% | 62%  | 81%   | 30% | −6%   | −2% | −1%   | 13%   |
| Medium           |     | 1%     | 21%  | 31% | 116% | 88%   | 34% | 11%   | −11%| −24%  | −1%   |
| Maximal          |     | −4%    | 29%  | 33% | 112% | 26%   | 47% | 2%    | 6%  | −96%  | 3%    |

The medication used clinically to treat thyrotoxicosis causes no inhibition in any MAP strain. "Medium" for ATCC 19698 is 64 µg/ml as 32 µg/ml was contaminated. 6-PTU causes dose dependent inhibition in only *M. tb* ATCC 25177. See Legend to Table 1 for explanation.
References

1. Cooper DS (2005) Antithyroid drugs. N Engl J Med 352: 905–917.
2. Laurberg P (2006) Remission of Graves’ disease during antithyroid drug therapy. Time to reconsider the mechanism? Eur J Endocrinol 155: 703–706.
3. Bianchi GP, Marchesini G, Garatt C, Zoli M (1990) Thyroid involvement in patients with active inflammatory bowel diseases. Ital J Gastroenterol 27: 291-295.
4. Inokuchi T, Moriwaki Y, Takahashi S, Tsutsumi Z, Ka T, et al. (2005) Autoimmune thyroid disease. Graves’ disease and Hashimoto’s thyroiditis in two patients with Crohn’s disease: case reports and literature review. Intern Med 44: 303–306.
5. Shah SA, Peppercorn MA, Pallotta JA (1998) Autoimmune (Hashimoto’s) thyroiditis associated with Crohn’s disease. J Clin Gastroenterol 26: 117–120.
6. Jarnerot G, Azad Khan AK, Truelove SC (1975) The thyroid in ulcerative colitis and Crohn’s disease. II. Thyroid enlargement and hyperthyroidism in ulcerative colitis. Acta Med Scand 197: 83-87.
7. Jobe HA, Frothingham LR (1895) Eugenitheriisfall von tuberculose beim rind (A particular case of tuberculosis in a cow). Duchs Zeitschr Tiermed, Vergl Pathol 21: 438–454.
8. Naser SA, Ghobrial G, Romero G, Valentine JP (2004) Culture of Mycobacterium avium subspecies paratuberculosis from the blood of patients with Crohn’s disease. Lancet 364: 1039–1044.
9. Greenstein R, Gillis T, Scolar D, Brown S (2009) Mycobacteria: Leprosy, a clinical trial of ethionamide and prothionamide for treatment of lepromatous leprosy. J Exp Med 204: 73–78.
10. Zhang FR, Huang W, Chen SM, Sun LD, Liu H, et al. (2009) Genomewide association study of leprosy. N Engl J Med 361: 2609–2618.
11. Schurr E, Gros P (2009) A common genetic fingerprint in leprosy and Crohn’s disease? N Engl J Med 361: 2511–2513.
12. Wang F, Langley R, Gahlen G, Doort LG, Besra GS, et al. (2007) Mechanism of thiamide drug action against tuberculosis and leprosy. J Exp Med 204: 73–76.
13. Fajardo TT, Guinto RS, Cebuon RV, Abalos RM, dela Cruz EC, et al. (2006) A clinical trial of ethionamide and prothionamide for treatment of lepromatous leprosy. Am J Trop Med Hyg 74: 457–461.
14. Phetsuksiri B, Jackson M, Scherman H, McNeil M, Besra GS, et al. (2003) Unique mechanism of action of the thiourea drug isoxyl on Mycobacterium tuberculosis. J Biol Chem 278: 53123–53130.
15. Levy L, Anandan JA (1975) Further studies of the action of antithyroid drugs on Mycobacterium leprae. Proc Soc Exp Biol Med 150: 582–585.
16. Levy L, Moon N (1972) Inhibition of the multiplication of Mycobacterium leprae by methimazole. Am Rev Respir Dis 106: 917–920.
17. Levy L, Ullmann NM (1973) Inhibition of multiplication of Mycobacterium leprae by several antithyroid drugs. Am Rev Respir Dis 111: 651–655.
18. Stewart-Tull DES (1982) Mycobacterium leprae - The bacteriologist’s enigma. In: Ratledge C, Stanford J, eds. The Biology of the Mycobacteria, Volume 1: Physiology, Identification, and Classification, 1 ed. New York: Academic Press. pp 273–307.
19. Greenstein RJ, Su L, Harrounian V, Shahidi A, Brown ST (2007) On the Action of Methotrexate and 6-Mercaptopurine on M. avium Subspecies paratuberculosis. PLoS ONE 2: e161.
20. Greenstein RJ, Su L, Shahidi A, Brown ST (2007) On the Action of 5-Amino-Saliclyc Acid and Sulfapyridine on M. avium including Subspecies paratuberculosis. PLoS ONE 2: e351.
21. Rastogi N, Goh KS, Lahroose V (1992) Activity of clarithromycin compared with those of other drugs against Mycobacterium paratuberculosis and further enhancement of its extracellular and intracellular activities by ethambutol. Antimicrob Agents Chemother 36: 2843–2846.
22. Greenstein RJ, Su L, Whitlock R, Brown ST (2009) Monensin causes dose dependent inhibition of Mycobacterium avium subspecies paratuberculosis in radiometric culture. Gut Pathogens 1: 4.
23. Greenstein RJ, Su L, Brown ST (2009) On the effect of thalidomide on Mycobacterium avium subspecies paratuberculosis in culture. Int J Infect Dis 13: e254–263.
24. Greenstein RJ, Su L, Juste RA, Brown ST (2008) On the Action of Cyclosporine A, Rapamycin and Tacrolimus on M. avium including Subspecies paratuberculosis. PLoS ONE 3: e2496.
25. Damato JJ, Collins MT (1990) Growth of Mycobacterium paratuberculosis in radiometric, Middlebrook and egg-based media. Vet Microbiol 22: 31–42.
26. Chioldini RJ, Van Kruiningen HJ, Thayer Jr. WJ, Couto J (1986) Spherophagic phase of mycobacteria isolated from patients with Crohn’s disease. J Clin Microbiol 24: 357–363.
27. Bertram MA, Inhelder CB, Yadeegar S, Kolanski P, Yamada JK, et al. (1986) Confirmation of the beige mouse model for study of disseminated infection with Mycobacterium avium complex. J Infect Dis 154: 194–195.
28. Nakamura H, Noh, IJH, Itoh K, Fukata S, Miyaochi A, et al. (2007) Comparison of methimazole and propylthiouracil in patients with hyperthyroidism caused by Graves’ disease. J Clin Endocrinol Metab 92: 2157–2162.
29. Lam D, DC, Lynch R, Davis R, Lazarus JH (1971) Concentration of 35S-propylthiouracil by the thyroid gland and its relationship to anion trapping mechanism. Metabolism 20: 989-990.
30. Marchant B, Alexander WD, Robertson JW, Lazarus JH (1979) Accumulation of 2-[14C]propylthiouracil in human polymorphonuclear leukocytes. Biochem Pharmacol 28: 2299–2296.
31. Lindsay RH, ABS, Abraham HRV (1975) Enzymatic S-methylation of n-propyl-2-thiouracil and other antithyroid drugs. Biochem Pharmacol 24: 463–468.
32. Lindsay RH, Kelly K, Hill JJ (1979) Oxidative metabolites of [2-14C]propylthiouracil in rat thyroid. Enizymology 104: 1686–1697.
33. Small PM, Fujisawa PI (2001) Management of tuberculosis in the United States. N Engl J Med 345: 189–200.
34. Britton WJ, Lockwood DN (2004) Leprosy. Lancet 363: 1209–1219.
35. Probishley DK (2002) Inflammatory bowel disease. N Engl J Med 347: 417–429.
36. Ridley DS, Jopling WH (1962) A classification of leprosy for research purposes. Lepr Rev 33: 119–128.