Parietal neurons encode information sampling based on decision uncertainty

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During natural behavior, animals actively gather information that is relevant for learning or actions; however, the mechanisms of active sampling are rarely investigated. We tested parietal neurons involved in oculomotor control in a task in which monkeys made saccades to gather visual information relevant to a subsequent action. We show that the neurons encode, before the saccade, the information gain (reduction in decision uncertainty) that the saccade was expected to bring for the following action. Sensitivity to information gain correlates with the monkeys’ efficiency at processing the information in the post-saccadic fixation, but is independent of neuronal reward sensitivity. Reward sensitivity, in turn, is unreliable across task contexts, inconsistent with the view that the cells encode economic utility. The findings suggest that parietal cells involved in oculomotor decisions show uncertainty-dependent boosts of neural gain that facilitate the implementation of active sampling policies, including the selection of relevant cues and the efficient use of the information delivered by these cues.

To survive and thrive in complex environments, animals must make decisions under uncertainty and gather information that reduces that uncertainty. In neuroscience and psychology, information accumulation is studied by presenting participants with given (experimenter-selected) sensory cues and asking them to make decisions based on those cues. In natural behavior, however, animals use active sensing strategies, whereby they endogenously decide what information to sample: which stimuli to listen, touch or look at to guide future actions. However, the neural mechanisms of these strategies are seldom investigated.

In humans and monkeys, the primary means of sampling visual information are rapid eye movements (saccades) that place the fovea on selected items in visual scenes. Cortical neurons involved in spatial attention and saccadic decisions have spatial receptive fields (RFs) and selectively encode attention-worthy stimuli and locations, but the significance of these selective responses has been intensely debated. A long-standing question, debated primarily in the lateral intraparietal area (LIP), is whether saccade-related responses encode attentional priority or the reward values of alternative actions.

We have argued that understanding this question requires the use of behavioral tasks in which participants deploy eye movements not merely to gather rewards, as has been the prevailing practice so far, but also to serve their natural role of gathering information. To this end, we trained monkeys on a new task in which they made two coordinated saccades: an initial saccade to gather information from a visual cue and a subsequent saccade to report a decision based on the information. In an initial study using this paradigm, we showed that pre-saccadic responses of LIP cells encoded the percentage validity of alternative cues—that is, the extent to which a cue, when examined during the post-saccadic fixation, will reduce the uncertainty of the subsequent action.

Here, we show that LIP neurons also encode expected information gain (IG) based on dynamic changes in decision uncertainty. The neurons had stronger pre-saccadic responses if the monkeys had ex ante decision uncertainty and expected the initial saccade to reduce that uncertainty compared with an alternative context in which the monkeys had prior knowledge of the appropriate final action and expected the saccade merely to bring redundant information. Moreover, the neural sensitivity to IG was not correlated with the sensitivity to reward gain, and reward sensitivity showed positive or negative scaling in different task contexts, inconsistent with the idea that the cells encode the economic utility. Instead, the findings support a two-stage model of attention control. A monitoring stage that seems to be implemented outside LIP allocates control based on the utility (costs and benefits) of alternative actions, and a regulatory stage, which includes the parietal cortex, implements the required control, in part through an uncertainty-dependent enhancement of neural gain that enables animals to select informative cues and efficiently process the information conveyed by these cues.

Results

Two monkeys (Macaca mulatta) performed an information-sampling task in which they made two coordinated saccades on each trial (Fig. 1a). The monkeys made their first saccade to a cue to obtain visual information (100% coherent motion signaling the correct decision alternative) and a second saccade to indicate the final decision based on the information (a saccade to one of the alternatives).

We varied the IGs of the initial saccade through blockwise manipulations of the ex ante uncertainty about the correct final action. In informative (INF) trial blocks, the monkeys started each trial with uncertainty about their final decision and could expect that the motion will resolve their uncertainty (indicating which alternative was correct on that trial; Fig. 1a, top). In contrast, in uninformative (unINF) blocks, the correct target was fixed across trials so that the monkeys could identify the correct second saccade in advance and the motion merely confirmed their prior expectations (Fig. 1a, bottom). The INF and unINF conditions were presented in alternating blocks of 50 correct trials, with the initial block type randomized across sessions (Fig. 1b).

Importantly, the monkeys had to make their first saccade to the cue in both the INF and unINF blocks, meaning that this saccade...
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Fig. 1 | Task design. a, Trial structure. All the trials had an identical structure and differed only in whether they appeared in INF or unINF blocks. Each trial started with a fixation stage when the monkeys were informed about the RS (large or small, 300 ms versus 100 ms solenoid open time) through the fixation point color and the block type (INF or unINF) through an openframe shape around the fixation point. This was followed by the onset of the trial display, containing two targets (white squares) and a cue (a patch of small stationary dots). After viewing the display for 500 ms during central fixation (delay period), the fixation point disappeared, instructing the monkey to make the first saccade to the cue. When the monkey fixated on the cue for 100 ms, the dots began to move with 100% coherence toward one of the targets (motion and second saccade, gray arrows). The monkey was free to make its final decision and received a reward if it made a second saccade (cyan arrow) to the target that had been cued by the motion. The dark frames highlighting the delay and first saccade periods indicate the epochs of interest for the neural data analysis. unINF blocks (lower two rows) had an identical trial structure but were distinguished from INF blocks by the distribution of the correct second saccade targets. In INF blocks, the correct target (in this example, up or down, indicated by the solid and broken red arrows) was randomly selected on each trial. In contrast, in unINF blocks, a single target was correct for 50 consecutive correct trials (in this example, ‘up’, indicated by the solid blue arrow). Therefore, whereas in an INF block the monkeys started the trial with decision uncertainty and resolved this uncertainty by viewing the motion, in unINF blocks, the uncertainty was resolved from the offset of the block and the motion provided redundant information. b, Example block sequence in a recording session. INF and unINF conditions differed in the probability that one of the targets was correct. In this example, the two targets were ‘up’ and ‘down’. P(up target) is the probability that the up target was correct, and P(down target) = 1 - P(up target). INF and unINF blocks were presented in alternating blocks of 50 correct trials until at least 4 blocks were completed. Small and large RRs were pseudorandomly interleaved within each block. The block type that was presented first (INF or unINF) and the target direction in the first unINF block were randomized across sessions. c, The 2×2 factorial design dissociating IG and RS. The colors indicate the convention we use throughout the paper, whereby INF blocks are shown in red and unINF blocks in blue, and saturated and pale colors indicate large and small RRs, respectively.

Fig. 2 | Behavior is sensitive to IG. Each point shows the mean and s.e.m. across all neural recording sessions (n = 87) with respect to the decision accuracy (the probability of selecting the correct final target), the VT (the time that the monkeys spent viewing the motion before their second saccade) and the completion rate (the fraction of trials in which the monkeys traversed all task states up to the second saccade).

Behavior. The monkeys had higher rates of trial completion at large relative to small RRs (Fig. 2, right; two-way ANOVA, main effect of RS: F(1,344) = 20.0, P < 10⁻⁴), showing that RS impacted their motivation. However, the RS had only a modest influence on the initial cue-directed saccade (Supplementary Fig. 1a) and, importantly, on the monkeys’ final decision, including the accuracy (Fig. 2, left; two-way ANOVA, main effect of RS on accuracy: F(1,344) = 0.6, P = 0.4; IG–RS interaction: F(1,344) = 10⁻⁴, P = 1.0) and the time to make the decision (post-saccadic motion viewing time (VT); Fig. 2, center; two-way ANOVA, main effect of RS on VT: F(1,344) = 5.5, P = 0.02; IG–RS interaction: F(1,344) = 0.69, P = 0.4; but see subsequent discussion and Supplementary Fig. 5 for individual monkey behaviors).

In contrast to the weak effects of RS, IG strongly modulated the timing and accuracy of the final decision. In unINF blocks, the monkeys spent minimal time viewing the motion and nevertheless reached very high decision accuracy (Fig. 2, center and left). In contrast, in INF blocks, VTs were significantly longer and decision accuracy levels dropped, showing that the monkeys required more time to link the motion information with the correct final action (Fig. 2; two-way ANOVA, main effect of IG on VT: F(1,344) = 388, P < 10⁻⁷; accuracy F(1,344) = 222, P < 10⁻³; all P < 10⁻² in individual monkeys). Completion rates were insensitive to IG (main effect of IG: F(1,344) = 0.8, P = 0.4; IG–RS interaction: F(1,344) = 0.7, P = 0.4), showing that the monkeys were equally motivated to complete the INF and unINF blocks. In contrast with its effects on the final decision, IG had minimal influence on the initial cue-directed saccade (Supplementary Fig. 1a), and there was no negative correlation between the first saccade latency and the post-saccadic viewing durations, ruling out that the monkeys traded off the time they spent preparing the first and second saccades (Supplementary Fig. 1b).

LIP neurons respond more in informative and low-reward trials. To understand the neural mechanisms related to informational actions, we placed the cue in the RF of an LIP cell and focused on its responses during the delay period preceding the initial cue-directed saccade (Fig. 1a, delay/first saccade; see Supplementary Fig. 2 for control geometries). The neurons had visual responses to the onset of the cue, followed by sustained delay period activity until saccade onset (Fig. 3). For many neurons, this pre-saccadic activity differed as a function of IG and RS. Some neurons, like the example cell shown in Fig. 3a, left, responded more strongly in INF blocks relative to unINF blocks but were insensitive to RS. Other cells, like the example shown in Fig. 3a, right, were sensitive to RS. Strikingly, the

had reward value independently of IG. We further manipulated the reward size (RS) for the second saccade, with large and small rewards randomly interleaved in each block and signaled by the fixation point color (Fig. 1a). This created a 2×2 factorial design that statistically dissociated RS and IG (Fig. 1c).
RS-sensitive cells were typically enhanced by smaller rewards, an effect that was opposite to the enhancement by larger rewards that is commonly observed in this area (for examples, see refs. 16-17). The average population response (n=87; n=49 in monkey M) showed both effects, with stronger firing rates (FRs) in INF blocks relative to unINF blocks (Fig. 3b, red versus blue) and on small-reward trials relative to large-reward trials (Fig. 3b, left versus right).

To quantitatively measure these modulations, we fitted the delay activity of each cell to a linear model that included IG and RS as categorical regressors, along with nuisance trial-by-trial regressors indicating the first saccade latency, velocity, endpoint accuracy, post-saccadic VT and the direction of the final saccade (Methods, equation (1)). While the sensitivity to saccade parameters was slight (Supplementary Fig. 3a), sizeable fractions of cells showed significant IG and RS modulations. A total of 38% of the cells showed significant effects of RS (Fig. 4; 29% in monkey M, 50% in monkey S). The average βRS coefficient was negative across the population (Fig. 4, upper black triangle; mean (s.e.m.) = -1.7 (0.35) spikes per second (sp s⁻¹), z = -4.6, P < 10⁻⁵ relative to 0; monkey M: z = -3.1, P = 0.0022; monkey S: z = -3.3, P = 0.0011) and in 79% of the RS-sensitive cells (teal triangle; -3.3 (0.8) sp s⁻¹, z = -3.4, P < 10⁻⁴; 85% in monkey M, 74% in monkey S), indicating that the predominant response was enhancement for a smaller RS. Significant effects of IG were found in 39% of the cells (Fig. 4, ordinate; 47% in monkey M, 29% in monkey S). The IG coefficient was positive across the population (Fig. 4, right black triangle; 2.3 (0.6) sp s⁻¹, z = 4.2, P < 10⁻⁴ relative to 0; monkey M: z = 3.7, P = 0.0002; monkey S: z = 2.0, P = 0.05), and in 85% of the significant cells (orange triangle; 5.5 (1.2) sp s⁻¹, z = 3.8, P < 10⁻⁴; 91% in monkey M, 73% in monkey S), which indicates that most cells had higher FRs for INF blocks relative to unINF blocks.

We conducted several analyses to estimate the reliability of these neural effects. In the significant cells, the average RS and IG coefficients represented a change of more than 10% relative to the average FR of the neurons (mean (s.e.m.) for RS: -11% (2.7%), IG: 13% (2.4%)) and more than 35% of the FR standard deviation (RS: -0.36 (0.08) z-score units; IG: 0.43 (0.08) z-score units). The results were replicated when using receiver operating characteristic analysis, which revealed significant discrimination across the population (Supplementary Fig. 3b; RS: mean (s.e.m.) 0.55 (0.011), z = 4.0, P < 10⁻⁵, IG: 0.55 (0.012), z = 4.0, P < 10⁻⁴) and in a sizeable fraction of cells (RS: 38%; IG: 44%). Time-resolved analysis showed that the sensitivity to IG was significant throughout the delay period (Supplementary Fig. 3c). Finally, the variance in FRs was smaller in unINF blocks relative to INF blocks, which is consistent with the lower FRs in the latter blocks, but at odds with the idea that these blocks reflected mixtures of states related to the two anticipated directions of the final saccade (Supplementary Fig. 3d,e).

**IG responses are not value effects.** We conducted several analyses to determine whether the IG modulations were explained by the reward sensitivity shown by the cells. In a first analysis, we tested whether the cells encode the expected value (EV) of the initial saccade, which is defined as the product of RS and reward rate in each type of block (Fig. 5a). The EV primarily depended on RS, as intended in the design of the task, and was larger in unINF blocks relative to
The strong positive interaction between IG and RS coefficients. In the marginal distributions, significant cells are indicated in darker shades and the arrowheads indicate the average values across the entire sample (black) and the subset of cells with significant coefficients (teal or orange). The gray vertical and horizontal lines show the null effects ($\beta_z = 0$ and $\beta_z = 0$). The broken line represents the least square regression, and the $r$ and $P$ values refer to the correlation coefficient relating the IG and RS coefficients.

INF blocks because of the higher decision accuracy in the former blocks (Fig. 5a). Thus, the EV showed significant positive effects of RS and IG, and a negative IG–RS interaction (two-way ANOVA, RS main effect: $F_{(1,344)} > 1.060, P < 10^{-10}$ for the full dataset and each monkey; IG main effect: all $F_{(1,344)} > 90, P < 10^{-15}$; IG–RS interaction: all $F_{(1,344)} > 8.5, P < 10^{-3}$). This pattern was qualitatively different from the average LIP FRs, which showed a negative effect of RS and no IG–RS interaction (Fig. 5c; two-way ANOVA, main effects of IG and RS: both $F_{(1,344)} > 39, P < 10^{-3}$; $F_{(1,344)} > 7, P < 0.01$ in each monkey individually; interaction: $F_{(1,344)} = 1.3, P = 0.3$; monkey M: $F_{(1,344)} = 1.8, P = 0.2$; monkey S: $F_{(1,344)} = 0.03, P = 0.9$).

We next examined whether the cells encode the value of information (VOI)—a higher order reward function that measures the added value of obtaining information relative to what the monkeys may expect to obtain had they acted without the information. The monkeys could have estimated the VOI based on their experience with catch trials, in which no motion was shown and the monkeys acted based on their prior expectations (10% in each block; Methods). Catch trials had high success rates of 93% in unINF blocks (96% in monkey M, 90% in monkey S), but were only at chance rates in INF blocks (49% overall; 53% in monkey M, 49% in monkey S), consistent with the different levels of ex ante uncertainty in the two types of blocks. Thus, VOI—the difference in EV between motion and no-motion (catch) trials—was very low in unINF blocks regardless of RS, but it was positive in INF blocks, particularly at higher RS (Fig. 5b; two-way ANOVA, $F_{(1,344)} > 470, P < 10^{-47}$ for main effect of IG, RS and IG–RS interactions, in the full dataset and each monkey). The strong positive interaction between IG and RS distinguishes VOI from EV (which shows a negative interaction; Fig. 5a) and from uncertainty reduction per se (which is independent of reward magnitude). Importantly, this pattern also distinguishes VOI from the LIP pre-saccadic response, in which IG and RS were encoded additively and with opposite signs (Fig. 5a).

These findings were confirmed by examination of individual cells. We reasoned that, if the IG effects were fully explained by reward sensitivity, the two signals would be highly correlated, such that neurons would only show IG sensitivity if they also had reward modulations. Contrary to this view, $\beta_{\text{IG}}$ and $\beta_{\text{RS}}$ coefficients were uncorrelated (Fig. 4; $r = -0.0051, P = 0.96$; monkey M: $r = -0.009, P = 0.95$; monkey S: $r = -0.11, P = 0.5$). To rule out that this negative result was an artifact of low statistical power, we estimated the $\beta_{\text{IG}}$ separately for small-reward trials and large-reward trials—that is, using only half the number of trials for each cell. The resulting $\beta_{\text{IG}}$ coefficients were highly correlated across small-reward trials and large-reward trials ($r = 0.76, P < 10^{-10}$; monkey M: $r = 0.79, P < 10^{-10}$; monkey S: $r = 0.51, p 0.0011$). This result shows that we could detect reliable correlations even if we used half the number of trials, and confirms that the lack of correlation between the IG and RS modulations reflects a true independence of the two modulations.

Because both EV and VOI measures showed prominent interactions between RS and IG, we repeated the individual neuron analysis using a model that included IG, RS and the IG–RS interaction (Methods, equation (2)). In contrast to the EV and VOI, the interaction coefficients in LIP FRs did not differ from 0 across the population (Fig. 5d; $\beta_{\text{INT}}$ mean (s.e.m.) = 0.44 (0.42), $z = 1.6, P = 0.11$; monkey M: 0.34 (0.63), $z = 1.4, P = 0.15$; monkey S: 0.58 (0.51), $z = 0.6, P = 0.56$) and the $\beta_{\text{IG}}$ coefficients were equivalent whether the regression model did or did not include an interaction term (Fig. 5e; $z = 1.42, P = 0.16; r = 0.94, P < 10^{-9}$).

We further evaluated these observations at the population level by fitting the population responses with the full set of 127 models that resulted from all the possible combinations of 7 regressors, including categorical indicators of task context (IG, RS and IG–RS) and average EV, VOI, decision accuracy and completion rates in individual sessions (Fig. 2). The best-fitting model was a two-parameter model that included only terms for IG and RS (Supplementary Fig. 4). Consistent with the individual-neuron results, this model produced a significantly positive effect of IG and a negative effect of RS ($\beta_{\text{IG, population}}$ (s.e.m.): combined data: 0.17 (0.016); monkey M: 0.24 (0.022); monkey S: 0.079 (0.024), all $P < 0.05$; $\beta_{\text{RS, population}} = -0.17$ (0.016); monkey M: $-0.094$ (0.02); monkey S: $-0.26$ (0.024); all $P < 0.05$). A three-parameter model with IG, RS and IG–RS as regressors produced an inferior fit and a nonsignificant interaction coefficient ($P = 0.09$). Finally, the models with the lowest Bayesian information criterion scores included the IG and RS terms, but only inconsistently included other predictors (Supplementary Fig. 4, bold). In summary, at the level of the population and individual cells, LIP FRs are best described as encoding IG and RS, rather than reward gains or behavioral indicators that covaried with context.

LIP responses to IGs correlate with post-saccadic discrimination efficacy. Since the neurons showed enhanced pre-saccadic FRs when the task required more engagement during the post-saccadic fixation, we tested whether the IG modulations were related to the efficacy of the post-saccadic motion discrimination. To this end, we plotted the accuracy of the final decision as a function of post-saccadic VT (Fig. 6a,b). Decision accuracy in unINF blocks was near the ceiling regardless of VT (Supplementary Fig. 5, blue). However, accuracy and VT in INF blocks were positively related (Fig. 6a,b), showing that, although the motion was fully coherent, the monkeys needed time to select the appropriate action. A median split of the data based on the neural $\beta_{\text{IG}}$ coefficient showed that the increase in accuracy was steeper in sessions in which the recorded neurons had stronger IG modulations (Fig. 6a, black versus gray). Note that this
effect involves a comparison across distinct groups of cells, which suggests that the relationship between $\beta_{\mathrm{RS}}$ and performance is a network effect—that is, it can be detected above and beyond the specific sample of neurons that we happened to record in a session.

Interestingly, the relationship between decision efficiency and the $\beta_{\mathrm{RS}}$ coefficient was prominent in large-reward trials but not small-reward trials (Fig. 6a inset). A two-way analysis of covariance with VT as a continuous covariate confirmed that the $\beta_{\mathrm{RS}}$ had a significant effect on accuracy above and beyond the effect of VT (Lawley–Hotelling trace $T = 12.71, P = 0.0055$), and the relationship was stronger on large-reward trials relative to small-reward trials (RS–$\beta_{\mathrm{RS}}$ interaction: $T = 7.0, P = 0.0084$), even though the VT showed no effect of RS (main effect of RS: $T = 2.8, P > 0.4$; RS–VT interaction: $T = 0.6, P > 0.3$ in the combined data and individual monkeys). In contrast to the effects of IG, performance did not differ when the data were split according to $\beta_{\mathrm{RS}}$ irrespective of whether we examined low-reward trials, high-reward trials, or both (Fig. 6b; effects of $\beta_{\mathrm{RS}}$, VT–$\beta_{\mathrm{RS}}$, and RS–$\beta_{\mathrm{RS}}$, all $T < 2.0, P > 0.1$ overall and in individual monkeys). Therefore, the efficiency of the post-saccadic discrimination is not related to the LIP reward modulations but is related to the neuronal IG sensitivity in a reward-dependent fashion.

Our earlier finding that the neurons did not encode variations in post-saccadic VT (Supplementary Fig. 2) suggests that, while LIP firing correlates with decision efficiency at a constant VT, it may not encode speed–accuracy trade-offs across the different contexts. To directly examine this hypothesis, we examined how the monkeys traded off VT and accuracy for different RSs in INF blocks. The two monkeys made different adjustments in response to RS. Monkey M slowed down and became more accurate on large-reward trials relative to small-reward trials, whereas monkey S showed the opposite pattern, making faster and less accurate responses when higher rewards were at stake (Supplementary Fig. 5). We captured this difference by calculating a combined index of speed and accuracy and taking the difference between the indices on large-reward trials versus small-reward trials (Fig. 6c; Supplementary Fig. 5). Monkey M showed a positive difference on average, which indicates that he slowed down and achieved higher accuracy on large-reward trials versus small-reward trials (Fig. 6c, thick traces; $n = 87$ cells) and individual monkeys (thin traces). The majority of neurons do not show IG–RS interactions. The distribution of interaction coefficients from the three-parameter model. Significant coefficients were found in only eight cells (black bars), of which only three were positive. The gray triangle shows the mean coefficient and the vertical line shows the abscissa $= 0$. INF coefficients are equivalent when estimated with the two-parameter and three-parameter models. Each point represents one cell ($n = 87$). The broken diagonal line is the best fit least squares linear regression.

Negative reward effects are explained by the task. Our finding that the neurons had negative reward modulations was unexpected given that previous studies have reported that the cells have higher FRs for higher rewards. This raised the possibility that we may have inadvertently recorded from a different population of cells. Two
observations strongly argue against this possibility. First, all the neurons that were tested with the two-step task were pre-screened using a memory-guided saccade (MGS) task and showed spatially tuned delay period activity, thus conforming to the functional definition of LIP cells\(^\text{13,14}\) (Supplementary Fig. 6; population: \(z = 8.0, P < 10^{-14}\) overall; \(z > 5.1, P < 10^{-7}\) in individual monkeys; \(P < 0.05\) in 93% of individual cells).

For additional confirmation, we tested a subset of the cells on a traditional one-step saccade task in which the monkeys made a single saccade to receive a large or small reward (with RSs equated to those in the two-step task; see Methods for details). Replicating the findings in the entire sample, the cells tested in this control task \((n = 56\) out of 87\) showed a significant enhancement by small RSs during the delay period of the two-step task \((\beta_{\text{RS}, \text{average}} (s.e.m.) = -1.8 (0.5) \text{ sps}^{-1}, z = -3.1, P = 0.002, n = 56)\). When tested with the one-step task, however, the same neurons had a positive reward effect, as previously shown for this area \((\text{Fig. 7a}; a 200-\text{ms window centered on saccade onset};\) mean \(\beta_{\text{RS,one-step}} = 2.7 (1.2), z = 2.2, P = 0.025\) relative to 0; \(z = 3.7, P = 0.0002\) relative to the delay period of the two-step task).

We finally tested whether the neurons showed reward enhancement for the final saccade in the two-step task. However, even though this saccade collected the final reward, the 48 cells that were tested in control geometry 2 (Supplementary Fig. 2c) showed no effect of RS in this epoch \((\text{Fig. 7b}; \text{mean (s.e.m.)} = -0.46 (0.73), z = -1.21, P = 0.22, n = 48)\) and no correlation between the RS coefficients before the first and second saccades of the two-step task \((r = -0.01, P = 0.92\) overall; monkey M: \(r = 0.09, P = 0.64\); monkey S: \(r = -0.19, P = 0.46\)\). Similar to the entire sample, these cells showed a significant enhancement with the larger rewards in the one-step task \((\text{mean (s.e.m.)} = 3.0 (1.3), z = 2.04, P = 0.04)\) and a significant difference between the one-step and two-step tasks \((z = 2.25, P = 0.02)\). Thus, the discrepancy between our results and previous investigations is explained by the behavioral context and not the neuronal population we sampled.

**IC effects do not encode uncertainty, arousal or difficulty.** To determine whether the IG modulations reflect nonspecific differences between the INF and unINF contexts, we compared the modulations related to the cue/initial saccade with those related to
IG effects cannot be explained by spatial normalization. A final hypothesis we considered is that the IG modulations arose indirectly from spatial competition between pools of LIP neurons that encode plans for the first and second saccade. According to this view, IG modulations may arise because the monkeys could plan their second saccade as early as the delay period in the unINF but not in the INF blocks. If the neurons encoded this advance saccade plan, this might have triggered competitive interactions that reduced the responses to the first saccade specifically in the unINF blocks, thus masquerading as an IG effect. Several analyses argue against this interpretation.

We reasoned that an IG effect produced by spatial competition would also be seen in the neural responses to the targets. Crucially, neurons should show an apparent IG effect in trials in which the final saccade was away from the RF, as the advance planning of the null-direction saccade would produce lower activity in the unINF compared to the INF blocks relative to the first saccade (Fig. 8a). The neurons did not show significant IG modulations in their target-related response (Fig. 8a; abscissa; $\beta_{\text{IG, target}}$ mean (s.e.m.) = $-0.31$ ($0.64), z = 0.41, P = 0.68$ relative to 0; $n = 36$) and had much larger IG coefficients in the cue-related response relative to the target-related response ($z = 3.21, P = 0.0014$; cue-related IG: $\beta_{\text{IG, cue}}$ mean (s.e.m.) = $3.2$ ($1.1), z = 2.9, P = 0.0038$ relative to 0; $n = 36$). Thus, the IG modulations were specific to the visual cue rather than indicating global changes of gain in the INF context.

**Fig. 7 | LIP neurons are enhanced by reward in a traditional one-step task.**

a. Saccade-aligned population PSTHs on the standard one-step task. The cartoon shows the task geometry, which involved a single saccade to a target inside the RF, with the expectation of a large or small reward.

b. Saccade-aligned population PSTHs in the same format as in a, but for the second saccade of the two-step task (control geometry 2; Supplementary Fig. 2c).

The targets of the final saccade. We reasoned that if the IG modulations were nonspecific effects of uncertainty, arousal or difficulty, they should be found not only for the cue but also for the target-related responses. We therefore used control geometry 1, which revealed how the neurons encoded the saccade targets during the delay period preceding the initial saccade (Supplementary Fig. 2b) and computed the IG modulations in this geometry, pooling across saccade directions to detect nonspatial effects (Fig. 8a). The neurons did not show significant IG modulations in their target-related response (Fig. 8a; abscissa; $\beta_{\text{IG, target}}$ mean (s.e.m.) = $-0.31$ ($0.64), z = 0.41, P = 0.68$ relative to 0; $n = 36$) and had much larger IG coefficients in the cue-related response relative to the target-related response ($z = 3.21, P = 0.0014$; cue-related IG: $\beta_{\text{IG, cue}}$ mean (s.e.m.) = $3.2$ ($1.1), z = 2.9, P = 0.0038$ relative to 0; $n = 36$). Thus, the IG modulations were specific to the visual cue rather than indicating global changes of gain in the INF context.

**Fig. 8 | Controls ruling out alternative explanations.** IG modulations are specific rather than global effects. Comparison of IG coefficients in response to the cue (standard geometry) and to the targets during the delay period (geometry 1; Supplementary Fig. 2b). Each point represents one cell ($n = 36$). The diagonal line is the equality line. Arrowheads show the marginal means, black if $P < 0.05$, otherwise gray. During the delay period, the IG modulates responses to the cue/first saccade (black) but not the responses to the target in geometry 1 (gray).

a. IG modulations do not depend on the relative distance between the cue and targets. The IG coefficient ($\beta_{\text{IG, cue}}$; ordinate) as a function the TRF index measuring the relative responses to the locations occupied by the cue and the target in the standard geometry (Methods, equation (3)). Larger values along the abscissa indicate cells for which the target elicited a stronger response; larger values along the ordinate indicate cells with stronger IG sensitivity. Each point represents one cell ($n = 87$), and color denotes individual monkeys. The broken diagonal line is the least square regression; the $r$ and $P$ values refer to the correlation coefficients. IG modulations do not depend on the visual hemifield locations of the cue and targets. Comparison of the $\beta_{\text{IG}}$ coefficients in trials in which the final saccade was directed to the same versus the opposite hemifield relative to the cue, for cells that had a target in each hemifield ($n = 41$). Arrowheads show marginal means.

b. IG effects are not by-products of spatial competition. Comparison of $\beta_{\text{IG}}$ in response to the cue (standard geometry) and to the target when it is opposite the RF in unINF blocks (geometry 1; Supplementary Fig. 2b). Each point represents one cell ($n = 36$). The diagonal line is the equality line. Arrowheads show the marginal means, black if $P < 0.05$, otherwise gray. Contrary to the spatial competition hypothesis, the neurons did not show IG modulations when the final saccade target was out of the RF. IG modulations do not depend on the relative distance between the cue and targets. The IG coefficient ($\beta_{\text{IG, cue}}$; ordinate) as a function the TRF index measuring the relative responses to the locations occupied by the cue and the target in the standard geometry (Methods, equation (3)). Larger values along the abscissa indicate cells for which the target elicited a stronger response; larger values along the ordinate indicate cells with stronger IG sensitivity. Each point represents one cell ($n = 87$), and color denotes individual monkeys. The broken diagonal line is the least square regression; the $r$ and $P$ values refer to the correlation coefficients. IG modulations do not depend on the visual hemifield locations of the cue and targets. Comparison of the $\beta_{\text{IG}}$ coefficients in trials in which the final saccade was directed to the same versus the opposite hemifield relative to the cue, for cells that had a target in each hemifield ($n = 41$). Arrowheads show marginal means.
representation—that is, the extent to which the cue and target locations activate the same population of cells. To test this hypothesis, we used RF mapping data from the MGS task to compute a target RF (TRF) index that ranged between 1 (indicating a cell for which the target location was entirely outside the RF) and 0 (indicating a cell for which the target location strongly encroached on the RF, eliciting responses equivalent to cue location; Methods, equation (3)). If the IG sensitivity arose from spatial competition, it should be inversely correlated with the TRF—that is, be larger for neurons whose RFs included both the cue and target locations. However, no such correlation was present in the neural response (Fig. 8c; r = 0.044, P = 0.69 overall; monkey M: r = 0.083, P = 0.57; monkey S: r = −0.12, P = 0.46).

An alternative version of this hypothesis is that the relevant distance is relative to the visual field and that competition is strongest when the cue and target locations fall within the same, relative to opposite, hemifields. To evaluate this possibility, we focused on neurons for which the cue was at a diagonal location (did not fall on the vertical or horizontal meridians) and the two saccade targets fell in opposite hemifields. If the IG sensitivity were due to visual competition, it should be stronger when the monkeys planned the second saccade to the target that occupied the same hemifield rather than the opposite hemifield relative to the cue. Contrary to this prediction, the βp values were highly correlated and statistically equivalent between the two geometries (Fig. 8d; z = 0.21, P = 0.83 between the two conditions (n = 41); monkey S: z = 0.12, P = 0.91; monkey M: z = 0.4, P = 0.69). In summary, neither the temporal nor the spatial properties of the IG modulations are consistent with a spatial interaction hypothesis.

Discussion

In contrast to laboratory tasks in which participants make saccades to gather rewards, in this experiment we focused on the neural correlates of saccades that gathered information. In a previous study using this paradigm, we showed that LIP neurons encoded expected IGs based on fixed, long-term estimates of cue validity. Here, we extend this result by showing that the cells are also sensitive to IGs based on fixed, long-term estimates of cue validity. If the IG sensitivity arose from spatial competition, it should be inversely correlated with the TRF—that is, be larger for neurons whose RFs included both the cue and target locations. However, no such correlation was present in the neural response (Fig. 8c; r = 0.044, P = 0.69 overall; monkey M: r = 0.083, P = 0.57; monkey S: r = −0.12, P = 0.46).

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Discussion

In contrast to laboratory tasks in which participants make saccades to gather rewards, in this experiment we focused on the neural correlates of saccades that gathered information. In a previous study using this paradigm, we showed that LIP neurons encoded expected IGs based on fixed, long-term estimates of cue validity. Here, we extend this result by showing that the cells are also sensitive to IGs occasioned by dynamic changes in decision uncertainty. Responses to decision uncertainty are found in frontal cortical areas3,14 and subcortical structures, including dopamine cells15–17 and have been implicated in reward expectation, confidence and risk attitudes (for example, see ref. 23). Our findings suggest that these responses also play key roles in determining the spatial and temporal properties of the IG modulations are consistent with a spatial interaction hypothesis.

Given the dual sensitivity of LIP cells to rewards and saccadic decisions, it seems natural to propose that the cells encode the economic utility of competing alternatives16,31. Our results argue against this interpretation and argue instead that the apparent encoding of economic utility may be limited to laboratory tasks in which animals make saccades directly to collect a reward. Although LIP cells had the expected reward-related enhancement in a traditional one-step saccade task, they showed no reward sensitivity for the final saccade of the two-step task and, strikingly, showed enhancement for smaller rewards before the information-sampling saccade in this task. Moreover, the neural sensitivity to IGs could not be explained by the reward value of gathering information. In striking contrast to the value hypothesis—which predicts that the neural sensitivity to IGs and reward gains should be correlated, interact multiplicatively rather than additively, and have congruent signs—the sensitivities of the neurons to reward and IG were uncorrelated, combined additively and, critically, had opposite signs (increasing as a function of IG but decreasing with reward gains).

In contrast to their inconsistent encoding of utility, the neural sensitivity to IGs correlated with the efficiency with which the monkeys used the information in the post-saccadic fixation—as measured by the decision accuracy at a given VT. This relationship differed from attentional interpretations of LIP function in two significant ways. First, contrary to the traditional view that top-down attentional feedback facilitates visual discrimination in a retinotopic fashion within a single fixation13,34, the facilitatory effect in our task was non-retinotopic and linked the neural responses to a peripheral location before the saccade with motion processing at a foveal location after the saccade. Second, in contrast with the prevailing pre-motor view of attention, whereby attention is recruited merely in relation to a saccade motor plan34, the effects we describe here link the LIP pre-saccadic responses with the cognitive demands of the post-saccadic fixation.

These findings suggest that current views based on economic utility or attentional priority are insufficient to describe the neural mechanisms of information-sampling decisions. Instead, we propose that understanding this process requires a broader framework that integrates oculomotor decisions and cognitive control mechanisms. Specifically, our results support the expected value of control (EVC) theory that postulates a separation between monitoring and regulative (implementation) aspects of control34.

According to the EVC theory, the dorsal anterior cingulate cortex (dACC) monitors the rewards and costs of alternative actions and uses this information to decide whether to engage in a task and how much effort (control) to allocate to the task. The control signal generated by the dACC then recruits more posterior ‘regulatory’ areas, including lateral frontal and parietal areas35–38, to implement the required level of effort. The dACC is proposed to call for additional effort by boosting neuromodulators—including dopamine and norepinephrine38—which may have led to the enhanced neural gain we observed in conditions of higher uncertainty. In this view, ‘attentional priority’ is a type of focused arousal; that is, a transformation of global arousal into a stimulus-specific gain increase, which enables animals to efficiently reduce uncertainty by focusing on and extracting the information delivered by relevant cues.

This idea naturally accounts for the fact that the effects of IG in our task were mere modulatory effects on the larger visual and pre-saccadic responses of the neurons. The effects we describe are comparable to previously reported attentional and reward modulations3,16–31 and, although they seem modest in size, the fact that they correlate with behavioral efficiency suggests that they are functionally significant. If our hypothesis is correct, it implies that a full understanding of the functional consequences of IG-related enhancements requires an understanding of the large-scale networks that control active sampling; these networks include the dACC and possibly also the orbitofrontal cortex and dopamine cells that seem to signal information value in tasks motivated by curiosity32,33, along with downstream readout mechanisms that translate these responses into actions34.

The interpretation of priority maps as implementing cognitive effort may also explain our present result that the cells scaled negatively with reward magnitude. In experiments that randomly interleave small and large RSs, a smaller reward is perceived as a loss and elicits prepotent ‘escape’ reactions in both behavior and neural responses known as motivational conflict35,36. Motivational conflict was also seen in the behavior of our monkeys, as the rates of trial completion were smaller for lower rewards (Fig. 2). If resolving the conflict and completing a low-reward trial requires enhanced cognitive effort, this may explain the stronger responses for smaller RSs that we observed. The need for enhanced control in conditions of conflict can parsimoniously explain other seemingly paradoxical findings in LIP cells, including the fact that the cells have enhanced responses to punishment-predicting cues that monkeys must look away from37 or to stimuli signaling visuomotor conflict such as an antisaccade38 or a change in motor effect39. Should this hypothesis be correct, our finding that responses to RS and IG are uncorrelated...
suggests that different types of effort (for example, related to uncertainty reduction, visuomotor conflict or motivational conflict) require boosting through distinct circuits, which is consistent with a recent report4. Thus a central question for future investigation concerns the relation between priority maps, value and cognitive effort in implementing active sensing strategies.

Online content
Any methods, additional references, Nature Research reporting summaries, source data, statements of code and data availability and associated accession codes are available at https://doi.org/10.1038/s41593-019-0440-1.

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Author contributions
J.G., N.D. and M.H. designed and implemented the experiment. M.H. and N.D. collected the data. M.H. analyzed the data. J.G. and M.H. wrote the paper.

Competing interests
The authors declare no competing interests.

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In addition, to test for reward sensitivity, neurons were tested in a control one-step saccade task in which the monkeys made a single saccade to a target to obtain a large or small reward. In these trials, the monkeys achieved fixation and, when the fixation point disappeared, made a saccade to a target whose shape (an upward- or downward-pointing triangle) signaled the RS. RSs were identical to those used in the main task. The task randomly interleaved free-choice trials in which the monkeys were offered both reward alternatives (with the large-reward and small-reward targets falling randomly inside or opposite the RF) and forced-choice trials in which a single target was present (either inside or opposite the RF). During the free-choice trials, the monkeys nearly always chose the large RS, which shows that they were highly sensitive to these differences in RS (97.6% in monkey M, 97.8% in monkey S). Neural responses were analyzed on interleaved forced-choice trials, in which we could obtain RF-directed saccades at both reward magnitudes. Note that this task was designed to act as a screening tool for the presence of reward modulations. Therefore, it was designed to establish the extent of tasks used in previous investigations of reward in this area (for example, see ref. 18) rather than to provide a systematic comparison with the information-sampling task.

Data analyses. Data are reported from 87 neurons recorded from two 16-year-old male monkeys (49 neurons in monkey M, 38 neurons in monkey S) that were tested in the standard geometry of the two-step task. Of these cells, 36 neurons were further tested in control geometry (1 in monkey M), 48 neurons (30 in monkey M) were tested in geometry 2, and 56 neurons (36 in monkey M) were tested with the one-step reward control task. No statistical methods were used to predetermine sample sizes, but rather trial sizes were similar to those reported in previous studies of this area. Data collection and analyses were not performed blinded to the conditions of the experiments. Error trials were not considered in the analysis of neural responses. All statistical comparisons used nonparametric tests (two-sided paired rank or signed-rank tests) unless otherwise noted. The Nature Research Reporting Summary contains summaries of statistics and data of this Methods section.

The 'trial completion' rate (Fig. 2) was the number of completed trials divided by the number of the trials in which the monkeys initiated fixation. The 'decision accuracy' was the number of rewarded trials divided by the number of completed trials. Beyond the analysis of completion rates, incomplete trials were discarded and not further analyzed. The saccade latency was measured between the fixation point offset and the saccade start, as defined based on velocity and acceleration criteria. The 'endpoint accuracy' was defined as the Euclidean distance between the center of the cue/target and the saccade landing position, the 'velocity' refers to the peak saccade velocity, and the VT was the interval between the motion onset and the start of the second saccade. Note that because the monkeys had to complete 100 ms of post-saccadic fixation before motion onset, the inter-saccade interval (that is, the latency of the second saccade) was equal to VT + 100 ms.

Delay-period FRs were measured from the raw spike trains. For each neuron, trial-by-trial FRs were fit with two regression equations as follows:

\[
FR = \beta_0 + \beta_1 \times IG + \beta_2 \times RS + \beta_3 \times RT + \beta_4 \times VEL + \beta_5 \times ACC + \beta_6 \times VT + \beta_7 \times SDIR
\]

(1)

\[
FR = \beta_0 + \beta_1 \times IG + \beta_2 \times RS + \beta_3 \times INT \times IG + \beta_4 \times RS + \beta_5 \times LAT + \beta_6 \times VEL + \beta_7 \times ACC + \beta_8 \times VT + \beta_9 \times SDIR
\]

(2)

IG is the information gain (0 for unINF, 1 for INF) and RS is the reward size (0 for small, 1 for large). The coefficients for LAT, VEL ACC capture the effects of, respectively, the RT, velocity and accuracy of the first saccade. VT is the motion viewing time and SDIR is the direction of the second saccade. SDIR was arbitrarily set to 0 or 1 for each of the two final saccades. The fits were implemented with the stepwiselm function in MATLAB v2014b. All the regressors were z-scored covering all the trials in a session. The IG and RS terms in equation (1), and the IG, RS and IG–RS terms in equation (2) were locked in the model. The remaining saccade descriptors were included according to a backwardwise procedure using \(P\) values of 0.05 and 0.10 as cut-offs, respectively, including and excluding regressors. All analyses were done on raw FRs and repeated with z-scored and mean-normalized FRs. Unless otherwise noted, regression coefficients are reported in units of sp. Exploratory analyses showed that the IG and RS effects were sustained during the delay period (Fig. 3c; Supplementary Fig. 3c) and were not sensitive to a range of window sizes spanning this period. Thus, the results are based on the FR averaged throughout the delay period (that is, 150 ms after cue onset until the saccade onset). The TRF measured the extent to which the targets encroached on the RF of a cell in the standard geometry of the two-step task. The TRF was computed for each cell as follows:

\[
TRF = \frac{FR_{\text{targ.lat}} - FR_{\text{targ.lat}}} {FR_{\text{targ.lat}} + FR_{\text{targ.lat}}}
\]

(3)
All FR values were measured during the delay period of the MGS task (300–800 ms after cue onset). FRcue_loc is the FR at the RF center (the location occupied by the cue in the standard geometry of the two-step task). FRtar_loc is the average FR at the two locations closest to the center (the same locations that were occupied by the targets in the standard geometry of the two-step task). The TRF ranges between 1 (indicating a cell that does not respond at all at the target locations) and 0 (indicating a cell that responded equivalently to the cue and target locations). Note that negative values are impossible in this index since the cue location had been pre-selected as the location that had a higher FR.

**Reporting Summary.** Further information on research design is available in the Nature Research Reporting Summary linked to this article.

**Data availability**
The data generated and analyzed for this study are available from the corresponding author upon request.

**Code availability**
The code written to analyze the data and to produce the figures for this study are available from the corresponding author upon request.

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  - Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated

*Our web collection on statistics for biologists contains articles on many of the points above.*

### Software and code

Policy information about availability of computer code

| Data collection | Monkeylogic, September 2012 edition |
|-----------------|-------------------------------------|
| Data analysis   | MATLAB R2014b, used to write custom code |

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# Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

| Sample size | Two animal subjects with a total of 87 neurons. Based on the literature, this is an average sample size for the effects we expected in LIP. No statistical methods were used to pre-determine sample sizes. |
|-------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Data exclusions | Neurons were selected based on pre-defined, standard, electrophysiological response patterns, as outlined in manuscript. Error trials were not considered in analysis of neural responses. This exclusion was pre-established to focus on trials of interest. |
| Replication | Results were replicated in two animals and in 87 independently sampled cells. |
| Randomization | The process of selecting the neurons is a random sample of LIP on a given experimental session. Experimental conditions were randomized within and across sessions. |
| Blinding | Investigators were not blinded. Blinding is irrelevant, because the results are based on statistical analyses, the exclusion criteria are separate from the phenomena being investigated, clearly reported, and standard in the field. |

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All studies must disclose on these points even when the disclosure is negative.

| Study description | Briefly describe the study type including whether data are quantitative, qualitative, or mixed-methods (e.g. qualitative cross-sectional, quantitative experimental, mixed-methods case study). |
| Research sample | State the research sample (e.g. Harvard university undergraduates, villagers in rural India) and provide relevant demographic information (e.g. age, sex) and indicate whether the sample is representative. Provide a rationale for the study sample chosen. For studies involving existing datasets, please describe the dataset and source. |
| Sampling strategy | Describe the sampling procedure (e.g. random, snowball, stratified, convenience). Describe the statistical methods that were used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient. For qualitative data, please indicate whether data saturation was considered, and what criteria were used to decide that no further sampling was needed. |
| Data collection | Provide details about the data collection procedure, including the instruments or devices used to record the data (e.g. pen and paper, computer, eye tracker, video or audio equipment) whether anyone was present besides the participant(s) and the researcher, and whether the researcher was blind to experimental condition and/or the study hypothesis during data collection. |
| Timing | Indicate the start and stop dates of data collection. If there is a gap between collection periods, state the dates for each sample cohort. |
| Data exclusions | If no data were excluded from the analyses, state so OR if data were excluded, provide the exact number of exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established. |
| Non-participation | State how many participants dropped out/declined participation and the reason(s) given OR provide response rate OR state that no participants dropped out/declined participation. |
| Randomization | If participants were not allocated into experimental groups, state so OR describe how participants were allocated to groups, and if allocation was not random, describe how covariates were controlled. |

# Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

| Study description | Briefly describe the study. For quantitative data include treatment factors and interactions, design structure (e.g. factorial, nested, hierarchical), nature and number of experimental units and replicates. |
| Research sample | Describe the research sample (e.g. a group of tagged Passer domesticus, all Stenocereus thurberi within Organ Pipe Cactus National Monument), and provide a rationale for the sample choice. When relevant, describe the organism taxa, source, sex, age range and any manipulations. State what population the sample is meant to represent when applicable. For studies involving existing datasets, describe the data and its source. |
| Sampling strategy | Note the sampling procedure. Describe the statistical methods that were used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient. |
| Data collection | Describe the data collection procedure, including who recorded the data and how. |
| Timing and spatial scale | Indicate the start and stop dates of data collection, noting the frequency and periodicity of sampling and providing a rationale for
**Timing and spatial scale**
These choices. If there is a gap between collection periods, state the dates for each sample cohort. Specify the spatial scale from which the data are taken.

**Data exclusions**
If no data were excluded from the analyses, state so OR if data were excluded, describe the exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.

**Reproducibility**
Describe the measures taken to verify the reproducibility of experimental findings. For each experiment, note whether any attempts to repeat the experiment failed OR state that all attempts to repeat the experiment were successful.

**Randomization**
Describe how samples/organisms/participants were allocated into groups. If allocation was not random, describe how covariates were controlled. If this is not relevant to your study, explain why.

**Blinding**
Describe the extent of blinding used during data acquisition and analysis. If blinding was not possible, describe why OR explain why blinding was not relevant to your study.

**Field work, collection and transport**

**Field conditions**
Describe the study conditions for field work, providing relevant parameters (e.g. temperature, rainfall).

**Location**
State the location of the sampling or experiment, providing relevant parameters (e.g. latitude and longitude, elevation, water depth).

**Access and import/export**
Describe the efforts you have made to access habitats and to collect and import/export your samples in a responsible manner and in compliance with local, national and international laws, noting any permits that were obtained (give the name of the issuing authority, the date of issue, and any identifying information).

**Disturbance**
Describe any disturbance caused by the study and how it was minimized.

**Reporting for specific materials, systems and methods**

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

### Materials & experimental systems

| n/a | Involved in the study |
|-----|-----------------------|
| ❑   | Antibodies            |
| ❑   | Eukaryotic cell lines |
| ❑   | Palaeontology         |
| ❑   | Animals and other organisms |
| ❑   | Human research participants |
| ❑   | Clinical data         |

### Methods

| n/a | Involved in the study |
|-----|-----------------------|
| ❑   | ChIP-seq              |
| ❑   | Flow cytometry        |
| ❑   | MRI-based neuroimaging |

#### Antibodies

**Antibodies used**
Describe all antibodies used in the study; as applicable, provide supplier name, catalog number, clone name, and lot number.

**Validation**
Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer’s website, relevant citations, antibody profiles in online databases, or data provided in the manuscript.

#### Eukaryotic cell lines

Policy information about [cell lines](#)

**Cell line source(s)**
State the source of each cell line used.

**Authentication**
Describe the authentication procedures for each cell line used OR declare that none of the cell lines used were authenticated.

**Mycoplasma contamination**
Confirm that all cell lines tested negative for mycoplasma contamination OR describe the results of the testing for mycoplasma contamination OR declare that the cell lines were not tested for mycoplasma contamination.

**Commonly misidentified lines**
(See [FLAC register](#))
Name any commonly misidentified cell lines used in the study and provide a rationale for their use.
**Specimen provenance**
Provide provenance information for specimens and describe permits that were obtained for the work (including the name of the issuing authority, the date of issue, and any identifying information).

**Specimen deposition**
Indicate where the specimens have been deposited to permit free access by other researchers.

**Dating methods**
If new dates are provided, describe how they were obtained (e.g. collection, storage, sample pretreatment and measurement), where they were obtained (i.e. lab name), the calibration program and the protocol for quality assurance OR state that no new dates are provided.

Tick this box to confirm that the raw and calibrated dates are available in the paper or in Supplementary Information.

**Animals and other organisms**

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

| Laboratory animals | 2 Rhesus monkeys (macaca mulatta), male, adult, age 16 |
|--------------------|-----------------------------------------------------|
| Wild animals       | Wild animals were not used in this study.            |
| Field-collected samples | Field samples were not used in this study.     |
| Ethics oversight   | All methods were approved by the Animal Care and Use Committees of Columbia University and New York State Psychiatric Institute as complying with the guidelines within the Public Health Service Guide for the Care and Use of Laboratory Animals. |

Note that full information on the approval of the study protocol must also be provided in the manuscript.

**Human research participants**

Policy information about studies involving human research participants

| Population characteristics | Describe the covariate-relevant population characteristics of the human research participants (e.g. age, gender, genotypic information, past and current diagnosis and treatment categories). If you filled out the behavioural & social sciences study design questions and have nothing to add here, write "See above." |
|-----------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Recruitment                 | Describe how participants were recruited. Outline any potential self-selection bias or other biases that may be present and how these are likely to impact results.                                                                                       |
| Ethics oversight            | Identify the organization(s) that approved the study protocol.                                                                                                                                                                                                       |

Note that full information on the approval of the study protocol must also be provided in the manuscript.

**Clinical data**

Policy information about clinical studies
All manuscripts should comply with the ICMJE guidelines for publication of clinical research and a completed CONSORT checklist must be included with all submissions.

| Clinical trial registration | Provide the trial registration number from ClinicalTrials.gov or an equivalent agency.                                                                                                           |
|----------------------------|------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Study protocol            | Note where the full trial protocol can be accessed OR if not available, explain why.                                                                                                                  |
| Data collection           | Describe the settings and locales of data collection, noting the time periods of recruitment and data collection.                                                                                             |
| Outcomes                  | Describe how you pre-defined primary and secondary outcome measures and how you assessed these measures.                                                                                                  |

**ChIP-seq**

**Data deposition**

- Confirm that both raw and final processed data have been deposited in a public database such as GEO.
- Confirm that you have deposited or provided access to graph files (e.g. BED files) for the called peaks.

**Data access links**
For "initial submission" or "Revised version" documents, provide reviewer access links. For your "Final submission" document, provide a link to the deposited data.

**Files in database submission**
Provide a list of all files available in the database submission.
Genome browser session (e.g. UCSC)

Provide a link to an anonymized genome browser session for "Initial submission" and "Revised version" documents only, to enable peer review. Write "no longer applicable" for "Final submission" documents.

Methodology

Replicates

Describe the experimental replicates, specifying number, type and replicate agreement.

Sequencing depth

Describe the sequencing depth for each experiment, providing the total number of reads, uniquely mapped reads, length of reads and whether they were paired- or single-end.

Antibodies

Describe the antibodies used for the ChIP-seq experiments; as applicable, provide supplier name, catalog number, clone name, and lot number.

Peak calling parameters

Specify the command line program and parameters used for read mapping and peak calling, including the ChIP, control and index files used.

Data quality

Describe the methods used to ensure data quality in full detail, including how many peaks are at FDR 5% and above 5-fold enrichment.

Software

Describe the software used to collect and analyze the ChIP-seq data. For custom code that has been deposited into a community repository, provide accession details.

Flow Cytometry

Plots

Confirm that:

- The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).
- The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a ‘group’ is an analysis of identical markers).
- All plots are contour plots with outliers or pseudocolor plots.
- A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation

Describe the sample preparation, detailing the biological source of the cells and any tissue processing steps used.

Instrument

Identify the instrument used for data collection, specifying make and model number.

Software

Describe the software used to collect and analyze the flow cytometry data. For custom code that has been deposited into a community repository, provide accession details.

Cell population abundance

Describe the abundance of the relevant cell populations within post-sort fractions, providing details on the purity of the samples and how it was determined.

Gating strategy

Describe the gating strategy used for all relevant experiments, specifying the preliminary FSC/SSC gates of the starting cell population, indicating where boundaries between "positive" and "negative" staining cell populations are defined.

- Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.

Magnetic resonance imaging

Experimental design

Design type

Indicate task or resting state; event-related or block design.

Design specifications

Specify the number of blocks, trials or experimental units per session and/or subject, and specify the length of each trial or block (if trials are blocked) and interval between trials.

Behavioral performance measures

State number and/or type of variables recorded (e.g. correct button press, response time) and what statistics were used to establish that the subjects were performing the task as expected (e.g. mean, range, and/or standard deviation across subjects).
### Acquisition

| Imaging type(s) | Specify: functional, structural, diffusion, perfusion. |
|-----------------|--------------------------------------------------------|
| Field strength  | Specify in Tesla                                       |
| Sequence & imaging parameters | Specify the pulse sequence type (gradient echo, spin echo, etc.), imaging type (EPI, spiral, etc.), field of view, matrix size, slice thickness, orientation and TE/TR/flip angle. |
| Area of acquisition | State whether a whole brain scan was used OR define the area of acquisition, describing how the region was determined. |
| Diffusion MRI | Used | Not used |

### Preprocessing

| Preprocessing software | Provide detail on software version and revision number and on specific parameters (model/functions, brain extraction, segmentation, smoothing kernel size, etc.). |
|------------------------|-------------------------------------------------------------------------------------------------|
| Normalization | If data were normalized/standardized, describe the approach(es): specify linear or non-linear and define image types used for transformation OR indicate that data were not normalized and explain rationale for lack of normalization. |
| Normalization template | Describe the template used for normalization/transformation, specifying subject space or group standardized space (e.g. original Talairach, MNI305, ICBM152) OR indicate that the data were not normalized. |
| Noise and artifact removal | Describe your procedure(s) for artifact and structured noise removal, specifying motion parameters, tissue signals and physiological signals (heart rate, respiration). |
| Volume censoring | Define your software and/or method and criteria for volume censoring, and state the extent of such censoring. |

### Statistical modeling & inference

| Model type and settings | Specify type (mass univariate, multivariate, RSA, predictive, etc.) and describe essential details of the model at the first and second levels (e.g. fixed, random or mixed effects; drift or auto-correlation). |
|-------------------------|------------------------------------------------------------------------------------------------|
| Effect(s) tested | Define precise effect in terms of the task or stimulus conditions instead of psychological concepts and indicate whether ANOVA or factorial designs were used. |
| Specify type of analysis: | Whole brain | ROI-based | Both |
| Statistic type for inference | Specify voxel-wise or cluster-wise and report all relevant parameters for cluster-wise methods. (See Eklund et al. 2016) |
| Correction | Describe the type of correction and how it is obtained for multiple comparisons (e.g. FWE, FDR, permutation or Monte Carlo). |

### Models & analysis

| n/a | Involved in the study |
|-----|-----------------------|
|     | Functional and/or effective connectivity |
|     | Graph analysis |
|     | Multivariate modeling or predictive analysis |

| Functional and/or effective connectivity | Report the measures of dependence used and the model details (e.g. Pearson correlation, partial correlation, mutual information). |
|------------------------------------------|-----------------------------------------------------------------|
| Graph analysis                           | Report the dependent variable and connectivity measure, specifying weighted graph or binarized graph, subject- or group-level, and the global and/or node summaries used (e.g. clustering coefficient, efficiency, etc.). |
| Multivariate modeling and predictive analysis | Specify independent variables, features extraction and dimension reduction, model, training and evaluation metrics. |