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To cite this version:
Lluis Fajas, Emilie Blanchet, Jean-Sébastien Annicotte. CDK4, pRB and E2F1: connected to insulin..
Cell Division, BioMed Central, 2010, 5 (1), pp.6. 10.1186/1747-1028-5-6. inserm-00663531

HAL Id: inserm-00663531
https://www.hal.inserm.fr/inserm-00663531
Submitted on 27 Jan 2012

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CDK4, pRB and E2F1: connected to insulin
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Abstract
Pancreatic β-cells are metabolic sensors involved in the control of glucose homeostasis. This particular cell type controls insulin secretion through a fine-tuned process, which deregulation have important pathological consequences, such as observed during type 2 diabetes. We recently implicated E2F1 in the control of glucose homeostasis. First we showed that E2f1\textsuperscript{-/-} mice have decreased pancreatic size, as the result of impaired postnatal pancreatic growth. We observed in this study that E2F1 was highly expressed in non-proliferating pancreatic β-cells, suggesting that E2F1, besides the control of β-cell number could have a role in pancreatic β-cell function. We demonstrate in our recent study, both in vitro and in vivo that E2F1 directly regulates the expression of Kir6.2, a key component of the K\textsubscript{ATP} channel involved in the regulation of glucose-induced insulin secretion in pancreatic β-cells. Expression of Kir6.2 is lost in pancreas of E2f1\textsuperscript{-/-} mice, resulting in insulin secretion defects in these mice. Furthermore, we demonstrated by in tissue chromatin immunoprecipitation analysis that regulation of Kir6.2 expression by E2F1 follows the same regulatory pathway that the classical E2F1 target genes, implicating the participation of CDK4 and retinoblastoma protein. Moreover, in this context, E2F1 transcriptional activity is regulated by glucose and insulin through the CDK4-dependent inactivation of the pRB protein. In summary we provide evidence that the CDK4-pRB-E2F1 regulatory pathway is involved in glucose homeostasis. In our recent study we decipher a new function for these factors in the control of insulin secretion and open up new avenues for the treatment of metabolic diseases, in particular type 2 diabetes.

Introduction
The synthesis of insulin occurs in a specific compartment of the islet of Langerhans, the pancreatic β-cells. In normal conditions, transient hyperglycemia requires increased insulin secretion, which could be obtained through several adaptations of the β-cell, i.e. hyperplasia (increase in number) or hypertrophy (increase in size) \cite{1,2}. This transient increase in β-cell proliferation to properly secrete sufficient amount of insulin is required to restore normal glucose level, yet deficit in this process could have pathological consequences. Diabetes mellitus are multifactorial disorders characterized by an autoimmune β-cell destruction (type 1) or a combination of insulin resistance in peripheral tissues and β-cell failure (type 2). The resulting hyperglycemia is responsible for several long-term complications, such as retinopathy, neuropathy, kidney diseases or cardiovascular diseases. During type 2 diabetes, the chronic hyperglycemic state will enforce insulin secretion, and thus β-cell function, finally leading to a decrease in β-cell mass. Therefore, understanding how β-cells proliferate and function is of utmost importance and could have strong implications in the development of new therapeutic strategies for type 1 and 2 diabetes. Participation of cell cycle regulators in this process can be predicted, and has been demonstrated, as described below. However, the aim of this paper is to comment our recent data demonstrating that, in addition to their role in the control of cell mass, cell cycle regulators directly regulate β-cell functions \cite{3}.

Discussion
Cell cycle, β-cell and more
E2F transcription factors are often depicted as the ultimate effectors of the G1/S transition of the cell cycle. When bound to DNA, they exist either as free E2F/DP heterodimers, or associated in larger complexes containing members of the retinoblastoma family (pRB, p107, p130) and members of the cyclin/cdk protein families. Cyclin-dependent kinases (CDK) define a family of serine/threonine protein kinases that phosphorylate a number of substrates mainly implicated in cell cycle progression and transcription. Association of E2Fs with proteins of the pRB family facilitates active repression
through recruitment of histone deacetylases [4] and lysine/arginine methyl-transferases [5]. Subsequent phosphorylation of the retinoblastoma protein by the cyclin/CDK complexes results in the release of the E2F transcription factors and activation of the transcription of genes required for progression through G1 into the S phase of the cell cycle. CDKs ultimately translate external signaling into transcriptional response, which is the final step of the regulatory cascade [6]. This makes the CDK-pRB-E2Fs potential candidates for the control of the crosstalk between proliferative stimuli and metabolic response. E2Fs functions are not restricted to the control of cell proliferation and it is now accepted that E2Fs participate in cellular processes beyond the cell cycle [7]. This has been extensively demonstrated through the characterization of E2fs-null mouse models [8] and the number of genes identified as E2F targets encoding factors involved in DNA replication, mitosis, DNA repair, apoptosis, differentiation, and development (reviewed in [9]). Of particular interest is the observation that E2Fs, pRB, and cdk4 could be involved in the regulation of a metabolic network, including the control of adipose tissue function [10,11].

Proliferation of β-cells is a key mechanism to maintain postnatal β-cell mass [12,13] and is the primary mechanism for β-cell regeneration [14-16]. It is now clear that pancreatic β-cells are able to replicate, albeit the origin of newly formed islets remains controversial. However, the mechanisms controlling β-cell replication unequivocally require cell cycle regulation. Gene inactivation studies of cell cycle regulators underscore the important role for these proteins in maintaining the β-cell population. Together with our previous observation that E2f1 -/- mice have decreased β-cell mass, due to decreased proliferation [17], other cell cycle regulators have been implicated in this process (for review, see [18]). Similar to E2f1-/-, Cyclin D1 +/-; Cyclin D2 +/- [19], Cyclin D2 +/- [15], and Cdk4 -/- [20] mice have decreased β-cell mass. Conversely, mice with gene inactivation of the cell cycle inhibitors p27 [21,22], p16INK4A [23] have increased islet mass. Interestingly, genetic rescue of a mutant Cdk4 insensitive to INK4 inhibition (CDK4R24C) in a Cdk4-null background results in normal β-cell mass, suggesting a key role for CDK4 in controlling proliferation of β-cell [24].

**E2F1 in the control of β-cell function and insulin secretion**

We previously evaluated the effects of E2F1 on glucose homeostasis. E2f1 -/- mice show an overall reduction in pancreatic size, as the result of impaired postnatal pancreatic growth [17]. E2F1 is not involved in embryonic β-cell development, since E2F1 is expressed at the final step of pancreas development, as demonstrated by in situ hybridizations. Because of the disproportionate small pancreas E2f1 -/- β-cells secrete insufficient amounts of insulin in response to a glucose load, resulting in glucose intolerance. However, despite this decreased insulin secretion, these animals were protected for the development of diabetes because of a dramatic increase in insulin sensitivity in other tissues, mainly adipose tissue [25]. In contrast, E2f1/E2f2 double KO mice are diabetic, showing exocrine pancreatic insufficiency, indicating that E2fs gene dosage and/or tissue specificity are critical for glucose homeostasis and pancreatic function [26,27]. These observations suggest however that E2F1 and E2F2 might cooperate to regulate β-cell mass and functions through cell-autonomous [3,17] and non-cell autonomous [26,28] effects. Surprisingly, we observed that E2F1 was highly expressed in non-proliferating adult pancreatic β-cells [17], suggesting that besides its impact on β-cell number, E2F1 could also play a role in pancreatic β-cell function and insulin secretion.

Pancreatic β-cell KATP channels are composed by the heteromeric association of the sulfonylurea receptor 1 (SUR1) with the K+ ATP channel inward rectifier (Kir6.2). These channels play a critical role in the regulation of glucose-induced insulin secretion by controlling membrane polarization [29,30]. The observation that Kir6.2 -/- mice display defective insulin secretion and increased insulin sensitivity is interesting [31], since it strongly resembles a particular phenotype of E2f1-/- mice [17]. Global gene expression analysis of E2f1+/+ and -/- pancreas revealed that the mRNA and protein levels of Kir6.2 were decreased. Most importantly, we demonstrated that Kir6.2 is a bona fide E2F1 target gene, as observed by chromatin immunoprecipitation performed on pancreas and isolated islet tissues. Most importantly, rescue of Kir6.2 expression in E2f1 -/- isolated islets restored glucose-stimulated insulin secretion, confirming that E2F1 exerts cell-autonomous effects on insulin secretion through its transcriptional control of Kir6.2 expression [3]. Furthermore, since Kir6.2 transcription is positively regulated by glucose, we studied the impact of glucose and glucose-stimulated insulin in the control of CDK4 activity and subsequent E2F1-dependent transcription. Our data demonstrated that glucose-stimulated insulin secretion triggers CDK4-dependent pRb phosphorylation in the β-cell, and subsequent transcriptional activation of the Kir6.2 gene by E2F1/DP-1 heterodimers. Finally, chemical invalidation of CDK4 activity, through the use of a specific inhibitor, resulted in glucose intolerance and impaired glucose-stimulated insulin secretion in mice, confirming a role for the CDK4-pRB-E2F1 pathway in insulin secretion process. Altogether, our data provide evidence that the CDK4-pRB-E2F1 pathway directly contributes to insulin secretion through the regulation of Kir6.2 expression in
pancreatic β-cells both in vitro and in vivo. Our results reveal a novel feed-back control mechanism of the insulin response.

Conclusion
We demonstrated that E2F1 transcriptional activity is controlled in pancreatic β-cells by insulin signaling secondary to glucose-induced insulin secretion [3]. How E2F1 can mediate both a proliferative (during post-natal growth or regeneration) or/and a metabolic response (during adulthood) in pancreatic β-cells in a coordinated manner is still not clear. Interestingly, insulin is an important β-cell proliferating factor, as evidenced by studies using β-cell specific inactivation of the insulin receptor (IR) in mice which resulted in a decrease in β-cell mass [32-34] or treatment of β-cell lines with IR siRNA which reduced proliferation rates of Min6 cells [35]. On the other hand, insulin has been shown to exert through an autocrine effect a feedback regulation on metabolic gene transcription, such as insulin itself (for review, [36]). Insulin signaling activation ultimately results in a specific metabolic response in β-cells. Since our recent findings indicate that the CDK4-pRB-E2F1 pathway is regulated by insulin, we can hypothesize that both the proliferative and metabolic effects of insulin on β-cells are mediated by the increase of CDK4 activity and subsequent E2F1 transcriptional activity. This would further suggest that both cell proliferation and metabolic responses are intimately linked, and regulated by the same upstream factors. In agreement with this hypothesis, caAKTΔAT transgenic mice deficient in Cdk4 display reduced β-cell proliferation, demonstrating that AKT induces b-cell proliferation in a CDK4-dependent manner [37]. Which is the pathway leading to cdk4-RB-E2F1 activation is largely unknown. One of the central elements that transduce signals from nutrient availability is the mTOR pathway. (Wullschleger et al., 2006). Strikingly, the metabolic phenotype of E2F1-/- mice is also reminiscent to the phenotype of mice carrying inactivating mutations in the mTOR substrate S6K1 (Pende et al., 2000; Ohanna et al., 2005; Aguilar et al., 2007; Um et al., 2004). Both E2F1-/- and S6K1-/- mice show impaired metabolism in pancreatic β-cells, adipose tissue, muscle, and likely in other tissues with metabolic activity. The whole transcriptional program controlled in pancreatic β-cells in adult rats after short-term glucose infusion. Diabetes 1989, 38:49-53.

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doi:10.1186/1747-1028-5-6