POLYMERASE CHAIN REACTION AS A DIAGNOSTIC TOOL FOR SIX SEXUALLY TRANSMITTED INFECTIONS - PRELIMINARY RESULTS

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Abstract

Background and aim. Sexually transmitted infections are a very frequent and under-diagnosed cause of illness worldwide. A high number of detection methods and a large range of specimens in which sexually transmitted infections can be determined are available at the moment. Polymerase chain reaction performed on first void urine offers the advantage of being non-invasive, self-collectable and has high sensitivity and specificity. We looked to determine the frequency of Chlamydia trachomatis, Neisseria gonorrhoeae, Trichomonas vaginalis, Mycoplasma hominis, Mycoplasma genitalium and Ureaplasma urealyticum in symptomatic and asymptomatic patients.

Methods. Six sexually transmitted infections were determined in the first void urine of 15 symptomatic and asymptomatic patients by polymerase chain reaction. We used "Epicenter MasterPure™ Complete DNA and RNA Purification Kit" for the DNA purification and "Seeplex® STD6 ACE Detection" for the DNA amplification. The results were examined in UV light.

Results. A number of 5 patients had positive results for Chlamydia trachomatis or Neisseria gonorrhoeae. Sexually transmitted infections are more frequent in men between 27 and 40 years old.

Conclusions. Polymerase chain reaction is a good diagnostic tool for sexually transmitted infections because it has a high sensitivity and specificity. Chlamydia trachomatis is the most frequent sexually transmitted infection, followed by Neisseria gonorrhoeae.

Keywords: sexually transmitted infections, urine, polymerase chain reaction.

Background and aims

Sexually transmitted infections (STI) are the most common cause of illness worldwide and they represent an important economic, therapeutic and social problem.

World Health Organization (WHO) statistics show that about 498.9 million new cases of Chlamydia trachomatis (CT), Neisseria gonorrhoea (NG), syphilis and Trichomonas vaginalis (TV) occurred worldwide in 2008 in adults between the ages of 15 and 49 [1]. The number
of patients with STIs is higher than the one reported in the
statistics because of the asymptomatic form of the infection,
the absence of partners testing, self-treatments and under-
reporting the cases.

In women, STIs can determine cervicitis, pelvic
inflammatory disease, salpingitis, endometritis, spontaneous
abortion and prematurity/low birth weight [2,3,4]. Women
have an over 3 times higher rate of contracting CT than
do men [5]. Urethritis, epididymitis and chronic prostatitis
occur in men with STIs [2,6]. In both genders, STIs can
cause infertility and they increase the susceptibility to HIV
infection [7,8,9,10]. In both sexes symptomatic infection
appears to be the exception rather than the rule, up to
70% remain asymptomatic for a varying period of time
[11,12,13].

There are a lot of laboratory methods to detect CT,
NG, TV, Ureaplasma urealyticum (UU), Mycoplasma
hominis (MH) and Mycoplasma genitalium (MG) that
range from cell culture to nucleic acid amplification tests
(NAAT) [8]. In bacterial culture only viable bacteria can be
quantified, but NAATs such as polymerase chain reaction
(PCR) can quantify both viable and nonviable bacteria
[11]. The higher sensitivity and better specificity of NAATs
are their main advantages over other tests [3,14]. PCR has
been used to analyze a variety of specimen types including
male and female urine, male urethral swabs, and female
endocervical/vaginal swabs [15]. The invasiveness of
traditional collection methods determined some individuals
not to perform STIs tests [16]. Self-collectable samples
such as first-void urine (FVU) and vaginal swabs have
the advantage of being non-invasive and may increase
the number of tested individuals and the practicability of
screening programs [6].

In men, although urethral swabs turned to be more
sensitive and specific than FVU when enzyme immunoassay
(EIA) was performed, this difference between urethral
swab and FVU was reduced with the NAAT [8].

In women, there is a strong consistency between
vaginal swabs or FVU and endocervical swabs in the
detection of STIs [17]. FVU in women has the advantage
over endocervical swabs that it can be self-collected and
does not require the presence of a doctor. Urine samples
are the preferred collection method over vaginal swab [18].
FVU is a good specimen for detecting STIs in both
genders.

PCR performed on FVU is able to detect as many
or more infected patients than urethral swabs, endocervical
swabs or semen [9]. PCR is a convenient method for
clinicians because it can test multiple causative organisms
simultaneously and in short time.

The aim of this study was to investigate the utility of
PCR in the detection of CT, NG, TV, MH, MG and UU and
to estimate the frequency of this six sexually transmitted
infections.

Patients and methods

Patients
A number of 15 symptomatic and asymptomatic
patients were included in the study, 13 men and 2 women.
In the symptomatic group were included patients who
presented to the doctor asking for medical advice for acute
or chronic symptoms, including: genital discharge,
dysuria, fever and abdominal pain. Patients who addressed
to the doctor for a screening and partners of symptomatic
patients, all having no symptoms, were included in the
asymptomatic group.

All patients were recruited multidisciplinarily
from the dermatology, urology and infection diseases
ambulatory departments in Cluj Napoca over the period
between January 2014 to August 2014. From the total of
15 patients, 12 had symptoms of STIs and 3 patients were
asymptomatic. The age of the enrolled patients was between
22 years and 71 years and all of them were sexually active.
Patients who had a history of antibiotic use in the previous 2
weeks were excluded from the study. An informed consent
was obtained from all patients enrolled in the study and
they had all answered a standard questionnaire in which
they were asked to provide their age, sexual status, marital
status, origin, socio-economic status, symptoms, history of
STIs, partner’s symptomatology and other investigations
performed for STIs in order to have a complete medical
history. The study was approved by the ethics committee
of University of Medicine and Pharmacy from Cluj Napoca.
Confidentiality was addressed on the patient information
sheet.

Collection of urine
First void urine was collected from all 15 patients.
In the morning, when patients had not urinated for at least
4 hours, 50 ml of urine had been self-collected in a sterile
plastic container. The specimens were transported to the
laboratory without added transport medium and stored at
4° C, then examined within a maximum of 7 days.

Methods
In the laboratory, 1 ml from the specimens was
centrifuged in order to obtain the sediment.

DNA purification
In order to purify the DNA, we used “Epicenter
MasterPure™ Complete DNA and RNA Purification Kit”. 150 μl
from the sample was mixed with lysis solution,
which contains Proteinase K, and then incubated at 65° C.
Then, the sample was cooled at 37° C, RNase A was added
and afterwards incubated at 37° C for 30 minutes. The mix
was placed on ice for 5 minutes and then reagent for protein
precipitation was added and vortexed vigorously in order to
eliminate the proteins, afterwards it was centrifuged with
Hettich mikro 200 R centrifuge for 10 minutes at 14000 rotation per minute (rpm). The supernatant which contained DNA was mixed with isopropanol, then the microcentrifuge tube was inverted 30-40 times and afterwards centrifuged for 10 minutes at 14000 rpm, thus DNA would depose in the pellet. DNA was rinsed twice with 75% ethanol and re-suspended in TE Buffer. At the end, it would result 35 μl buffer with DNA.

**DNA amplification**
For this part, we used “Seeplex® STD6 ACE Detection”. In an Eppendorf container the following were mixed: primer, DNA polymerase, deoxynucleoside triphosphate (dNTP), internal control and a solution to prevent contamination. This mix was placed in PCR tubes, the patients DNA sample was added together with positive and negative controls. Afterwards the mix was electrophoresed in 2% agarose gel containing ethidium bromide, with a molecular weight marker.

**Results analysis**
The detection and examination of PCR products were performed in UV light. The distance of migration is proportional with the size in agarose gel. TV has the greatest size in agarose gel, so it migrates on a small distance. It is followed by MH, MG, CT and NG. UU has the smallest size in agarose gel, so it migrates on the longest distance.

**Results**
In order to analyze the collected data, we used descriptive statistics to provide essential information related to the demographic features of the patients. Thus, 86.7% of the patients were males, with an average age of 31 years old (Tab. I). Among young men aged between 27 and 40 years old, we found positive results of our tested STIs.

| N (%) | Mean Age | Symptomatic (%) | Asymptomatic (%) |
|-------|-----------|-----------------|------------------|
| Male  | 13 (86.7%)| 31              | 11 (84.6%)       | 2 (15.4%)       |
| Female| 2 (13.3%) | 35              | 1 (50%)          | 1 (50%)         |

Frequencies were conducted to provide important results related to the number of the patients that had one of the six tested STIs, which represent the categorical variables of the present study.

Therefore, CT was found in 4 patients and NG in 2 patients. One patient had positive result for CT and also for NG (Fig. 1). MH, MG, TV and UU were not found in this 15 patients.

The 3 asymptomatic patients were negative for all these six STIs. From the 12 symptomatic patients, 5 had positive results for CT or NG and 7 were negative for the tested STIs. One woman enrolled in this study was symptomatic and 1 was asymptomatic, but both had negative results for the six STIs tested. Of the 13 men enrolled in the study, 2 were asymptomatic and had negative results and 11 were symptomatic, out of which 5 had positive results.

**Discussion**
Although other diagnostic tools, such as culture, antigen detection and serology are available for the diagnostic of STIs, molecular testing seems to be highly sensitive and specific [4,8,9,10].

This is to our knowledge the first study in our area using PCR as a diagnostic tool for STIs.

Our data is consistent with other studies reporting CT as the most prevalent bacterial STI [3,4,8,12]. In 2008, from a total of 46.8 million new cases of CT, NG, syphilis and TV reported by WHO for the European Region, 20.6 million were new cases of CT [1].

The second most common STI in our prospective study was gonorrhea, which is the second most commonly reported sexually disease in United States [19]. In the WHO statistics for the European Region, in 2008, there were 3.4 million new cases of NG [1], but in this statistics syphilis was also included, a fact that placed gonorrhea on the third place after chlamydia and syphilis.

Co-infection with CT and NG seems to be frequent, 10-30% of people infected with NG are also co-infected with CT [19,20]. In the United States, empiric treatment of CT concurrently with gonococcal treatment is recommended if diagnostic testing for CT was not performed [20]. In this study we found one patient co-infected with NG and CT, thus another advantage of PCR as a diagnostic tool for STIs would be to help the clinicians to decide whether it is appropriate to give co-treatment for chlamydial infection.

![Figure 1. Example of PCR result. NC - negative control. 6, 7, 8, 9 - samples. PC - positive control. MK - marker of molecular weight. Sample 6 - positive for CT. Sample 9 - positive for CT and NG.](image-url)
if they have a positive result for NG. As it can be observed in figure 1, there is a low positive result for NG compared with other positive results, which is due to the small amount of DNA detected in the patient’s FVU. Correlating the low positive result with the patient’s medical history, we have concluded that there is an incipient gonorrhea associated with chlamydiasis.

According to WHO global estimation, in 2008 there were 22.6 million new cases of TV in the European region [1]. We did not detect TV in our 15 cases. This can be explained by the fact that only 2 women were included in the study and trichomoniasis is more frequent in women [7].

MH, MG and UU were not found in our study, probably due to the small number of tested patients and to the fact that they affect a large number of infertile couples [9].

One of the most important limitations of the current study is the reduced number of cases which could only provide descriptive data. TV, MH, MG and UU were not detected and the asymptomatic group has also reduced number of patients. However, these are important preliminary results that provide essential insights for further extended research studies.

Conclusions

PCR performed on FVU seems to be the best choice to detect STIs because it is more sensitive and specific than previously available diagnostic tests. This method of detection uses noninvasively collected samples, such as FVU, and it has the advantage of being able to test for multiple causative organisms simultaneously.

Chlamydia trachomatis is the most frequent sexually transmitted infection, followed by Neisseria gonorrhoeae.

In the future, PCR is expected to be used as a standard diagnostic test for STIs. Because it can perform analysis on hundreds of samples in a short time, PCR is a good diagnostic tool for screening programs.

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References:

1. Global incidence and prevalence of selected curable sexually transmitted infections – 2008 [Internet]. Geneva: World Health Organization; 2012 [cited 2014 Nov 21]. Available from: http://apps.who.int/iris/bitstream/10665/75181/1/9789241503839_eng.pdf?ua=1
2. Korte JE, Baseman JB, Cagle MP, Herrera C, Piper JM, Holden AE, et al. Cervicitis and genitourinary symptoms in women culture positive for Mycoplasma genitalium. Am J Reprod Immunol. 2006;55:265-275.
3. Abusarah EA, Awwad ZM, Charvalos E, Shehabi AA. Molecular detection of potential sexually transmitted pathogens in semen and urine specimens of infertile and fertile males. Diagn Microbiol Infect Dis. 2013;77:283-6.
4. Krolov K, Frolov J, Tudoran O, Suhorutsenko J, Lehto T, Sibul H et al. Sensitive and rapid detection of Chlamydia trachomatis by recombinase polymerase amplification directly from urine samples. J Mol Diagn. 2014;16:127-135.
5. Madkan VK, Giancola AA, Sra KK, Tiryg SK. Sex differences in the transmission, prevention, and disease manifestations of sexually transmitted diseases. Arch Dermatol. 2006;142:365-370.
6. Ostergaard L. Diagnosis of urogenital Chlamydia trachomatis infection by use of DNA amplification. APMIS Suppl. 1999;89:5-36.
7. Shipitsyna E, Zolotoverkhaya E, Chen CY, Chi KH, Grigoryev A, Savicheva A, et al. Evaluation of polymerase chain reaction assays for the diagnosis of Trichomonas vaginalis infection in Russia. J Eur Acad Dermatol Venereol. 2013;27:e217-223.
8. Eley A. How to detect Chlamydia trachomatis in males? J Androl. 2011;32:15-22.
9. Gdoura R, Kchaou W, Ammar-Keskes L, Chakroun N, Sellemi A, ZnaZen A, et al. Assessment of Chlamydia trachomatis, Ureaplasma urealyticum, Ureaplasma parvum, Mycoplasma hominis, and Mycoplasma genitalium in semen and first void urine specimens of asymptomatic male partners of infertile couples. J Androl. 2008;29:198-206.
10. Domeica M, Zhurauskaya L, Savicheva A, Frigo N, Sokolovskiy E, Hallen A, et al. Guidelines for the laboratory diagnosis of trichomomiasis in East European countries. J Eur Acad Dermatol Venereol. 2010;24:1125-1134.
11. Choe HS, Lee DS, Lee SJ, Hong SH, Park DC, Lee MK, et al. Performance of Anyplex™ II multiplex real-time PCR for the diagnosis of seven sexually transmitted infections: comparison with currently available methods. Int J Infect Dis 2013;17:e1135-1140.
12. Noren L, von Krogh G, Bondesson L, Nohlgard C, Grillner L. Potential public health benefits from testing with Chlamydia trachomatis PCR technique on first void urine in men. Acta Derm Venereol. 1998;78:63-66.
13. Rose SB, Bromhead C, Lawton BA, Zhang J, Stanley J, Baker MG. Access to chlamydia testing needed for high-risk groups: patterns of testing and detection in an urban area of New Zealand. Aust N Z J Public Health. 2012;36:343-350.
14. Aguilera-Arreola MG, Gonzalez-Cardel AM, Tenorio-L. Sensitive and rapid detection of Chlamydia trachomatis, Neisseria gonorrhoeae, Mycoplasma hominis and Ureaplasma urealyticum. BMC Res Notes. 2014;7:433.
15. Kumamoto Y, Matsumoto T, Fujisawa M, Arakawa S. Detection of Chlamydia trachomatis and Neisseria gonorrhoeae in urogenital and oral specimens using the cobas® 4800, APTIMA Combo 2® TMA, and ProbeTec™ ET SDA assays. Eur J Microbiol Immunol (Bp). 2012;2:121-127.
16. O’Byrne P. Self-directed sexually transmitted infection testing: providing noninvasive sexual health services. Appl Nurs Res. 2011;24:17-21.
17. Fang J, Husman C, DeSilva L, Chang R, Peralta L. Evaluation of self-collected vaginal swab, first void urine, and endocervical swab specimens for the detection of Chlamydia trachomatis and Neisseria gonorrhoeae in adolescent females. J Pediatr Adolesc
18. Tebb KP, Paukku MH, Pai-Dhungat MR, Gyamfi AA, Shafer MAB. Home STI Testing: The adolescent female’s opinion. J Adolesc Health. 2004;35:462-467.
19. Rosen T. Gonorrhea, Mycoplasma, and vaginosis. In: Goldsmith LA, Katz SI, Gilchrest BA, Paller AS, Leffell DJ, Wolff K (eds) Fitzpatrick’s dermatology in general medicine. 8th edition. New York: Mc Graw Hill; 2012:2514-2519.
20. Stary A, Stary G. Sexually transmitted infection. In: Callen JP, Cerroni L, Heymann WR, Hruza G, Mancini AJ, Patterson JW et all (eds) Dermatology. 3rd edition. Elsevier; 2012:1379-1383.