Cytogenetic analysis of three species of Pseudacteon (Diptera, Phoridae) parasitoids of the fire ants using standard and molecular techniques

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Abstract

Pseudacteon flies, parasitoids of worker ants, are being intensively studied as potentially effective agents in the biological control of the invasive pest fire ant genus Solenopsis (Hymenoptera: Formicidae). This is the first attempt to describe the karyotype of P. curvatus Borgmeier, P. nocens Borgmeier and P. tricuspis Borgmeier. The three species possess 2n = 6; chromosomes I and II were metacentric in the three species, but chromosome pair III was subtelocentric in P. curvatus and P. tricuspis, and telocentric in P. nocens. All three species possess a C positive band in chromosome II, lack C positive heterochromatin on chromosome I, and are mostly differentiated with respect to chromosome III. P. curvatus and P. tricuspis possess a C positive band, but at different locations, whereas this band is absent in P. nocens. Heterochromatic bands are neither AT nor GC rich as revealed by fluorescent banding. In situ hybridization with an 18S rDNA probe revealed a signal on chromosome II in a similar location to the C positive band in the three species. The apparent lack of morphologically distinct sex chromosomes is consistent with proposals of environmental sex determination in the genus. Small differences detected in chromosome length and morphology suggests that chromosomes have been highly conserved during the evolutionary radiation of Pseudacteon. Possible mechanisms of karyotype evolution in the three species are suggested.

Key words: Pseudacteon, karyotype evolution, C-banding, fluorescent banding, FISH.

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Introduction

The Phoridae, one of the most diverse families within the order Diptera, includes more than 26,000 min species, varying in life style from scavengers and predators to parasitoids (Disney, 1994; Gilbert and Jervis, 1998). The phorid genus Pseudacteon Coquillet are parasitoids, most of the species parasitizing Solenopsis Westwood fire-ants (Hymenoptera: Formicidae) (Disney, 1994). Pseudacteon flies are solitary parasitoids (Disney, 1994). The females inject a single egg into each worker ant, the third larval instar of which migrating into and later pupating within the host’s cephalic capsule (Porter et al., 1995; Porter, 1998).

Solenopsis has a cosmopolitan distribution (Disney, 1994; Patrock et al., 2009). In South America, 22 Pseudacteon species have been reported as parasitizing fire-ants of the Solenopsis saevissima complex (Porter and Pesquero, 2001; Brown et al., 2003; Folgarait et al., 2005b; Calcaterra, 2007; Kronforst et al., 2007). Currently, these flies are used as biological control agents against Solenopsis invicta Buren and S. richteri Forel fire-ants, native to Argentina (Mescher et al., 2003; Ross et al., 2008) and exotic pests in the United States (Porter et al., 2004; Thead et al., 2005; Morrison and Porter, 2006).

Despite numerous studies on the life-history traits of the genus Pseudacteon, including apparent environmental sex-determination in some species, cytogenetic studies are lacking. Dipteran species generally possess low diploid chromosome numbers, these ranging from 2n = 4 to 20, with modal numbers at 6 and 8 (due to several families of “lower” Diptera i.e. SO Nematocera) and 12 (with many species of the SO Brachycera and belonging to calyptrate and acalyprate families) (White, 1973). Almost all species of Sarcophagidae and Calliphoridae (SO Brachycera, superfamily Oestroidea) have 2n = 12 with a pair of sex chromosomes XY (Parise-Maltempi and Avancini, 2000; 2001). In the family Drosophilidae as a whole (SO Brachycera, superfamily Acalyptrata) the primitive chromosome number appears to be 2n = 12 with five pairs of
long acrocentric chromosomes (including the sex chromosome pair XY) and a small dot-like pair. Reduction in chromosome number has occurred many times independently by fusion of these elements (White, 1973). Many derived sex chromosome systems have been described and the evolutionary mechanisms of sex chromosome differentiation has been analyzed in Drosophila species (Steinemann and Steinemann, 2005). In Muscidae (SO Brachycera, superfamily Muscoidea) most species also possess 2n = 12 with XY sex chromosomes in the male. However, muscid species have been described with 2n = 10 chromosomes and lacking differentiated sex chromosomes (Parise-Maltempi and Avancini, 2001). In Phoridae (SO Brachycera, superfamily Phoroidea), Megaselia scalaris Loew and M. spiracularis Schmitz, the only 2 species that have been studied cytogenetically, possess the diploid number 2n = 6, with 2 metacentric and 1 telocentric chromosome pairs. They do not possess heteromorphic sex chromosomes and sex is determined by a single Maleness factor that can be located in any of the three linkage groups (Mainx, 1964; Traut and Willhoeft, 1990).

In order to evaluate whether the well-marked differences among P. curvatus Borgmeier, P. nocens Borgmeier and P. tricuspis Borgmeier in size, morphology, behavior and phylogenetic relations are also reflected in their karyotypes, our aim was to describe the karyotype of these species by using standard and molecular cytogenetic techniques. Descriptive values of the karyotype, heterochromatin content and distribution, and the location of nucleolus organizer regions (NORs) were analyzed. A comparative analysis of cytogenetic results also provided information on the possible karyotype evolution within the genus. Furthermore, we analyzed the presence of differentiated sex chromosomes, since it has been suggested, in the case of other species in the genus Pseudacteon, that sex determination is related to host size, and thus could be environmentally determined (Morrison et al., 1999).

Material and Methods

Fly rearing

Experiments were carried out between August, 2006 and November, 2007. Pseudacteon species were reared in colonies of S. invicta ants collected near Mercedes, Corrientes province (29° 47' S, 58° 03' W). Adult males and females of P. curvatus were captured in the Reserva Ecológica Costanera Sur (RECS), Buenos Aires province (34° 37' S, 58° 22' W), those of P. tricuspis near San Javier, Santa Fe province (30° 58' S, 59° 94' W), and those of P. nocens from Mercedes, Corrientes province (29° 11' S, 58° 5' W), all in Argentina.

Slide preparation

Different oviposition assays were performed in the lab following the same methodology used by Chirino et al. (2009). The “attacked” ants were placed in a rearing room at 28 ± 1 °C with a 12:12 (light: darkness) photoperiod and 80 ± 10% RH.

In Diptera, cytogenetic analyses are mainly carried out with third instar larvae or early pupae (Traut and Willhoeft, 1990; Cevallos and Nation, 2004). Since development in Pseudacteon takes place within its host (Porter et al., 1995), it is difficult to obtain larvae at the appropriate developmental stages for cytogenetic studies. Thus we dissected pupae of 3-5 days, which lacked differentiated tissues, but showed high mitotic indices.

Since the females of P. tricuspis and P. nocens emerge from larger ant-heads than males, parasitized worker heads were measured and developed pupae were classified as either potential females or males. In contrast, both sexes of P. curvatus develop in similar sized hosts, so they could not be classified by sex (Morrison et al., 1997; Folgarait et al., 2006).

Pseudacteon pupae were dissected in a saline solution, swollen in a hypotonic solution (0.075 M KCl) for 10-20 min, and fixed for 15-30 min in freshly prepared Carnoy fixative (ethanol: chloroform: acetic acid, 6:3:1). Slides were prepared according to Traut (1976).

Chromosome morphology

Suitable cells at the mitotic prometaphase were selected and the mean descriptive values of the karyotype were calculated using information obtained from at least 3 cells per slide for each individual of each species analyzed (20.78 ± 9.76 cells). Each slide represented different individual pupae. Mitotic prometaphases were obtained from P. tricuspis pupae (17 males and 24 females); from P. nocens pupae (27 males and 21 females), and 35 specimens of P. curvatus. Slides were stained with the fluorescent dye DAPI (Rebagliati et al., 2003) for morphological studies. The nomenclature of Levan et al. (1964) was used to describe chromosome morphology.

Banding techniques

Heterochromatin content and distribution was analyzed by means of the C-banding technique and sequential DAPI and CMA3 banding. C-bands were performed according to Sumner (1972) with slight modifications. Slides were hydrolyzed in HCl 0.2N at room temperature for 15 min, briefly washed in distilled water and then incubated at room temperature for 35 min in a 5% Ba(OH)2 solution. Slides were thoroughly washed in tap water for 1 min and incubated for 1 h in 2XSSC at 60 °C. After washing in distilled water, the slides were stained with DAPI (Rebagliati et al., 2003), since band resolution improves when pretreated slides are stained with DAPI instead of Giemsa. Fluorescent staining with GC specific chromomycin A3 (CMA3) and AT specific 4′6- diamidino-2- phenylindole (DAPI) was carried out according to Rebagliati et al. (2003).
For location of NRs, Ag-NOR staining was applied according to Howell and Black (1980). Fluorescent in situ hybridization (FISH) with 18S rDNA was also undertaken.

Fluorescence in situ hybridization

Unlabelled 18S rDNA probes were generated by PCR, using primers described by Fuková et al. (2005) and sequenced using an Automated Sequencer ABI 3100 (DNA Sequencer, PE, Applied Biosystems), thereby confirming their being a section of 18S rDNA of *Ceratitis capitata* (Weidemann) (Diptera: Tephritidae). PCR was done in a Mastercycler® Gradient Eppendorf thermal cycler (Eppendorf AG, Hamburg, Germany). Reactions were carried out with template genomic DNA extracted from *C. capitata* by standard procedure according to Baruffi et al. (1995). The PCR product showed a single band of about 900 bp on a 1% agarose gel. The band was recovered from the gel and purified by using Wizard SV gel and PCR – clean up (Promega). The rDNA probe was labeled by nick translation with biotin 14-dUTP (BioNick Labeling System, Invitrogen Life Technologies Inc., Buenos Aires, Argentina). FISH with a biotinylated probe was carried out as described by Sahara et al. (1999) with several modifications as described by Fuková et al. (2005). Hybridization signals were detected with streptavidin-Cy3 conjugate (Sigma, Saint Louis, USA). The preparations were counterstained with 0.5 μg/mL DAPI in PBS, 1% Triton X-100, and mounted in anti-fade (Vectashield Mounting Medium, Vector Laboratories, Inc., Burlingame, USA).

Photographs

Preparations were observed in a Leica DMLB epifluorescence microscope. Black-and-white images of chromosomes were recorded with a CCD camera (Leica DFC350 FX, Leica IM50 Version 4.0, Leica Microsystems Imaging Solutions Ltd. Cambridge, UK) separately for each fluorescent dye. Images were pseudocolored (light blue for DAPI and red for Cy3), and, when necessary, superimposed with the aid of Adobe Photoshop version 6.0.

Statistical analysis

The following chromosome measurements were performed with MicroMeasure for Windows, version 3.3: centromeric index CI (length of short arm as a percentage of the whole chromosome), arm ratio r (relationship between the long and the short arms of each chromosome), relative length % (length of a chromosome as a percentage of the total chromosome length), and total chromosome length – TCL (the sum of the lengths of all the chromosomes of the complement). Comparisons of data within and between species were undertaken by analyses of variance (ANOVA), adjusted by the Bonferroni method. Statistical comparisons were done using the STATISTIX Program (Analytic Software 1998, Tallahassee, FL).

Results

Analysis of mitotic chromosomes of the pupae of *Pseudacteon tricuspis*, *P. curvatus* and *P. nocens* revealed a 2n = 6 diploid number (Figure 1 a-b). Chromosome pairs I and II were metacentric in the three species, but chromosome pair III was subtelocentric in *P. tricuspis* and *P. curvatus*, and telocentric in *P. nocens* (Table 1). Neither *P. tricuspis* nor *P. nocens* possessed a distinguishable heteromorphic pair in potential males or females (*t* = 0.43, df = 39, *p* = 0.67 for *P. tricuspis* and *t* = 0.07, df = 46, *p* = 0.95 for *P. nocens*, 2-tailed *t*-Test). The three species did differ in total chromosome length (TCL) (*F*2,99 = 2.38, *p* = 0.0453) which was higher in *P. curvatus*, lower in *P. nocens* and intermediate in *P. tricuspis* (Table 2). Chromosome I in *P. nocens* was relatively larger than in *P. tricuspis*.

![Figure 1](image-url) - Cells in the mitotic metaphase (a) and anaphase (b) of *P. curvatus* stained with DAPI. Fluorescent banding with DAPI (c,d,e) and CMA3 (d,f) in *P. curvatus* (c-d) and *P. tricuspis* (e-f). Bar = 10 μm.
and *P. curvatus* (*F*$_{2,123}$ = 8.68, *p* = 0.0004). Chromosome II in *P. nocens* was also larger than in *P. curvatus* (*F*$_{2,123}$ = 5.86, *p* = 0.0039), whereas in *P. tricuspis* it was of intermediate length. Finally, chromosome III was smaller in *P. nocens* than in *P. tricuspis* and *P. curvatus* (*F*$_{2,123}$ = 29.14, *p* < 0.0001) (Table 2).

Different banding techniques were applied in order to reveal heterochromatin and analyze its location and composition. DAPI and CMA$_3$ sequential fluorescent banding did not reveal any regions either in AT rich (DAPI bright) or GC rich (CMA$_3$ bright) for any chromosome pair in any of the three species (Figure 1 c-f). After C-banding treatment, slides were stained with DAPI, allowing a better analysis of the C-heterochromatin pattern (Figure 2). C positive heterochromatin was scarce in all species, with a small C positive (C+) band in the long arm (q) of chromosome II and a completely C negative (C-) chromosome I (Figure 2a-f). “Somatic pairing” was evident from prophase up to metaphase (Figure 2a, d-i), but was lost during anaphase (Figure 2b-c). The three species varied as to the C-banding pattern in chromosome III. Thus, in *P. curvatus* a C+ band present in the short arm (p) extended over 81% of its length (Figure 2a-c). In *P. tricuspis*, a C+ band observed in the long (q) arm near the centromere corresponded to approximately 4% of its length (Figure 2d-e). Finally, the telocentric pair in *P. nocens* was completely C- (Figure 2f).

Silver impregnation revealed only one nucleolus in interphase nuclei and two signals in early prophase cells in the three species. However, no positive results were obtained in metaphase chromosomes (Figure 3). *In situ* hybridization with the 18S rDNA probe on metaphase cells showed a single hybridization signal in chromosome II in the three species at a location similar to that of the C+ band (Figure 2g-i; Figure 4). This suggests that in the three *Pseudacteon* species a single cluster of rRNA genes is located in the long arm (q) of chromosome II near the centromere.

### Discussion

The *Pseudacteon* species analyzed presented 2n = 6, with chromosomes I and II being metacentric and chromosome III subtelocentric in *P. curvatus* and *P. tricuspis* but telocentric in *P. nocens*. Total chromosome length (TCL) varied among *Pseudacteon* species, with *P. curvatus* ≥ *P. tricuspis* ≥ *P. nocens*, probably representing variations in DNA content that do not affect chromosome morphology. A similar situation has been reported in other dipteran fami-

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### Table 1 - Comparison of the somatic complement of the *Pseudacteon* species analyzed. %: relative length, r: arm ratio, CI: centromeric index, N: number of slides analyzed.

| *Pseudacteon curvatus* | Average length (µm) | % | r     | CI | Classification | N |
|------------------------|---------------------|---|--------|----|----------------|---|
| Chromosomes            |                     |   |        |    |                |   |
| I                      | 47.11 ± 17.53a      | 19.96a | 1.14 ± 0.10 | 46.87 | m              |   |
| II                     | 42.19 ± 14.89a      | 17.75b  | 1.15 ± 0.09 | 46.63 | m              | 35|
| III                    | 30.01 ± 12.14b      | 12.29c  | 3.99 ± 0.89 | 21.18 | st             |   |

| *Pseudacteon tricuspis* | Average length (µm) | % | r     | CI | Classification | N |
|-------------------------|---------------------|---|--------|----|----------------|---|
| Chromosomes             |                     |   |        |    |                |   |
| I                       | 44.69 ± 11.14a      | 19.88a  | 1.20 ± 0.17 | 45.78 | m              |   |
| II                      | 40.43 ± 9.26a       | 18.07b  | 1.18 ± 0.16 | 46.20 | m              | 41|
| III                     | 27.20 ± 7.77b       | 12.05c  | 5.20 ± 0.74 | 14.53 | st             |   |

| *Pseudacteon nocens*   | Average length (µm) | % | r     | CI | Classification | N |
|------------------------|---------------------|---|--------|----|----------------|---|
| Chromosomes            |                     |   |        |    |                |   |
| I                      | 41.21 ± 10.12a      | 20.76a  | 1.14 ± 0.08 | 46.94 | m              |   |
| II                     | 36.64 ± 8.88b       | 18.48b  | 1.13 ± 0.07 | 46.98 | m              | 48|
| III                    | 21.53 ± 5.56c       | 10.75c  | 4.91 ± 0.60 | 12.20 | t              |   |

1The comparisons of chromosomal lengths and the relative lengths (%) were made by means of one-way ANOVA. Different letters indicate significant differences (p < 0.05) adjusted by Bonferroni.

### Table 2 - Comparisons of total chromosomal lengths (TCL) and relative lengths (%) among the *Pseudacteon* species analyzed.

| Species      | TCL (µm) | %   |
|--------------|----------|-----|
|              | I  | II  | III |
| *P. curvatus*| 225.31 ± 70.74 A | 19.96 B | 17.75 B | 12.29 A |
| *P. tricuspis*| 220.11 ± 54.69 AB | 19.88 B | 18.07 AB | 12.05 A |
| *P. nocens*  | 198.45 ± 46.01 B  | 20.76 A | 18.48 A | 10.75 B  |

1The comparisons of the total chromosomal lengths (TCL) and the relative lengths (%) were analyzed by means of one-way ANOVA. Different letters indicate significant differences (p < 0.05) adjusted by Bonferroni.
lies (Rafael and Tadei, 1998; Parise-Maltempi and Avancini, 2001; Selivon et al., 2005).

Heterochromatin content was very scarce in these Pseudacteon species. Chromosome I was completely C-, with a single C+ band situated at the location of the 18S rDNA hybridization signal on chromosome II. Major differences were observed in chromosome III, with a C+ band in the p-arm of the subtelocentric pair in P. curvatus and the q-arm in P. tricuspis, although this was absent in the telocentric pair of P. nocens (Figure 4). The divergence between the former two species could be explained by pericentric inversion in chromosome III during their evolutionary history (Figure 5). Its absence in P. nocens suggests that it could have diverged before the differentiation of P. tricuspis and P. curvatus. Furthermore, the smaller size of chromosome III in P. nocens, as compared to the other two species, could be explained by the absence of a heterochromatin block. Although it would be necessary to analyze other Pseudacteon species, we propose here a possible hypothesis on karyotype evolution within the genus. We propose that after the divergence of P. nocens, and through the acquisition of heterochromatin, there was an increase in DNA across the genome of the ancestor of the other two species, but principally in chromosome III. In spite of the fragmented and unresolved phylogenies of some Pseudacteon species (Kronforst et al., 2007; Calcaterra et al., 2008), several conclusions support our hypothesis. The recent common ancestor of P. nocens is different from the most recent common ancestor of P. curvatus and P. tricuspis. The possession of a single Maleness factor M that triggers off male development (Mainx, 1964; Willhoffet and Traut, 1990; Traut and Wollert, 1998; Traut et al., 1999). Through molecular cytogenetic techniques and molecular markers it has been established that the chromo-

![Figure 2](image1.png)

Figure 2 - Somatic mitosis of Pseudacteon curvatus (a-c, g), P. tricuspis (d-e, h) and P. nocens (f,i) after C banding (a-f) and in situ hybridization with a 18S rDNA probe (g-i). At the top right-hand corner the names of species are indicated as PC = P. curvatus, PT = P. tricuspis, and PN = P. nocens. Arrows (a-f) indicate blocks of heterochromatin and head arrows (g-i), rDNA sites. Bar = 10 μm.

![Figure 3](image2.png)

Figure 3 - Interphase nuclei of P. curvatus (a) and P. tricuspis (b), and the early prophase (c) in which one nucleolus was observed after silver nitrate NOR banding. d) Metaphase of P. tricuspis. No Ag-NOR band was detected on the condensed chromosomes. Bar = 10 μm.
somes X and Y in *Megaselia* are in a very early state of molecular differentiation (Traut, 1994).

In most of the studied *Pseudacteon* species, the sex of the offspring seems to be facultatively determined by the size of the host, with females emerging from the larger ones (Morrison *et al.*, 1999; Folgarait *et al.*, 2005a, 2006). From an evolutionary point of view, this makes sense since 1) fire-ant workers are highly variable in size (Tschinkel *et al.*, 2003), 2) female flies have a higher fitness if they emerge from larger hosts, exhibiting a higher efficiency in development and larger sizes (Chirino, Gilbert, Folgarait, unpublished), and 3) females emerge from a narrower range of ant-sizes than males (Folgarait *et al.*, 2005a, 2006; Chirino, Gilbert, Folgarait, unpublished). However, in two small species, *P. culpellatus* Borgmeier (Folgarait *et al.*, 2002) and *P. curvatus* (Chirino *et al.*, 2009), females and males develop in similar-sized hosts, and emerged adults are not sexually dimorphic in size, so that the above mentioned pattern is difficult to detect. The number of different species of *Pseudacteon* that develop on *S. invicta* exceeds the diversity observed in other ants (Disney, 1994). This diversity could be influenced by *Solenopsis* size polymorphism since there is a correlation between the sizes of both parasitoid and host (Morrison *et al.*, 1997). Different *Pseudacteon* species oviposit at different times during the day, at different locations (close to mounds or the foraging trails of ants) and during different seasons. *P. nocens* in particular is crepuscular and abundant in shady conditions, but *P. curvatus* and *P. tricuspis* are more abundant at midday and in sunny conditions (Folgarait *et al.*, 2007a; 2007b). Furthermore, *P. curvatus* mainly attacks small workers (Chirino *et al.*, 2009), *P. tricuspis* larger ants (Morrison *et al.*, 1997), and *P. nocens*, medium-sized ones. Finally, *P. tricuspis* females exhibit competitive, aggressive and territorial behaviors, which are not observed in either *P. curvatus* or *P. nocens*. In this work we have shown that the well-known differences among *P. curvatus*, *P. nocens* and *P. tricuspis* as regards size, morphology, behavior and phylogenetic relations are also reflected in their karyotypes.

The exact mechanism of sex determination in *Pseudacteon* flies is perplexing (Porter, 1998). Environmental sex determination (ESD) has been suggested as a possible mechanism for the observed pattern (Morrison *et al.*, 1999). Typically, taxa having non-differentiated sex chromosomes usually show labile sex-determination, and sex can be determined by environmental stimuli (Solari, 1994). Our demonstration of non-differentiated sex chromosomes in *Pseudacteon* phorids is thus consistent with the earlier suggestion of environmental sex determination in these parasitoid flies, and should encourage further investigation of how sex is determined in this genus.

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Internet Resources

MicroMeasure Web site, http://www.colostate.edu/Depts/Biology/MicroMeasure, for MicroMeasure for Windows, version 3.3.

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