Ruthenation of Non-stacked Guanines in DNA G-Quadruplex Structures: Enhancement of c-MYC Expression

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Abstract: Guanine quadruplexes (GQs) are compact four-stranded DNA structures that play a key role in the control of a variety of biological processes, including gene transcription. Bulky ruthenium complexes featuring a bipyridine, a terpyridine, and one exchangeable ligand ([Ru(terpy)(bpy)Cl]PF6) are able to metalate exposed guanines present in the GQ of the c-MYC promoter region that are not involved in quadruplex base pairing. qRT-PCR and western-blot experiments indicated that the complexes promote a remarkable increase in the expression of this oncogene. We also show that exchangeable thioether ligands (X = RSR', Met) allow regulation of the metalating activity of the complex with visible light.

There is great interest in the development of metal-based DNA binders that show improved selectivity and reduced toxicity relative to cis-platinum. [1] Ruthenium complexes are promising alternatives owing to their kinetic stability and rich photochemistry and redox properties. [2] In addition to non-covalent recognition of double-stranded DNA (dsDNA), several ruthenium complexes that form covalent DNA adducts, especially through reaction with the N7 of guanines, have also been described. [3] While most of these complexes bind dsDNA, the functional relevance of G-quadruplexes (GQs) [4,5] calls for the development of probes capable of targeting these structures. [6] To our knowledge, there is only one precedent for the covalent metalation of a GQ structure with a ruthenium complex, and the reaction presents low selectivity. [7]

Herein, we demonstrate that coordination complexes of the type [Ru(terpy)(bpy)X]+ (X = Cl, RSR', Met) can selectively metalate unpaired guanines present in parallel GQs, a reaction that is enhanced upon irradiation. Importantly, we have found that this selective metalation increases the expression of the oncogene c-MYC, apparently by disrupting the parallel GQ structure present in its promoter region.

Our work was conceived after learning that while most DNA-metalating ruthenium agents are cytotoxic, the complex [Ru(terpy)(bpy)Cl]+ (1) exhibits very low toxicity. [8] We reasoned that the bulky and relatively hydrophobic nature of this sort of complexes could offer excellent opportunities for the selective modification of accessible guanines, thereby promoting specific biological responses with reduced toxicity. We first studied the ability of chloro complex 1 to metalate guanosine monophosphate (GMP). Mixing complex 1 with 3 equiv of GMP in phosphate buffer (pH 7.5), led to the partial formation of the aquo complex [Ru(terpy)(bpy)H2O]+ (2; over 50% after 30 min at RT), while the guanosine remained essentially unreacted (Figure 1B, trace b). Further incubation for 2 h afforded the metalated product 3 and the aquo derivative 2 (Figure 1A), with total consumption of the starting chloride (Figure 1B, trace d).

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Figure 1. A) GMP metalation reaction with [Ru(terpy)(bpy)Cl]+, [Ru(terpy)(bpy)Cl]2+ and formation of the aquo derivative 2. B) HPLC of the reaction of 1 (250 µM) and GMP (750 µM) in 10 mM phosphate buffer pH 7.5, 100 mM NaCl: in the dark at t = 0 (trace a); after 30 min in the dark (trace b); initial mixture after 30 min of irradiation at 455 nm (trace c); and initial mixture after 2 h in the dark (trace d). The injection peak is labeled with an asterisk. C) Signal in the MS spectrum corresponding to the monoadduct 3.
Interestingly, irradiation of the initial mixture for 30 min ($\lambda = 455$ nm) led exclusively to the formation of the monoadduct 3 (approximate conversion of 80% based on the disappearance of GMP; Figure 1B, trace c and Figure S5 in the Supporting Information). Importantly, the reaction is fully orthogonal, so competitive control experiments in the presence of excess adenosine monophosphate (AMP), cytidine monophosphate (CMP), and thymidine monophosphate (TMP; Figure S6), as well as lysine or even cysteine (Figure S7), led exclusively to the formation of the GMP derivative. Moreover, experiments with double-stranded oligonucleotides (dsDNAs) presenting different arrangements of guanines revealed modest reactivity, but only with those featuring terminal guanines (Table S1, and Figures S10–S13 in the Supporting Information). This selectivity most likely stems from the bulkiness of complex 1, which cannot reach the sterically hindered nucleophilic N7 site in paired, internal guanines.9

Remarkably, irradiation of a mixture of the parallel c-MYC quadruplex $d[\text{TTGAG}_3 \text{TAG}_3 \text{TG}_3 \text{TA}_3]$ (10 $\mu M$) with 5 equiv of 1 in 10 mM phosphate buffer (pH 7.5) and 100 mM KCl led to the clean formation of a product with a mass corresponding to a monoadduct derivative (Figure 2A, trace C, peak at 22 min, MYC-[Ru], 81% conversion). We also observed reaction in the absence of light (approx. 41% conversion after 30 min at RT, Figure 2A, trace b). Importantly, MS analysis of the bovine spleen phosphodiesterase (BSP) digestion of the ruthenated oligonucleotides lead to identification of the first guanine of the sequence (5’-TTGA…, Figure 2C) as the metalation site (Figure S24)11 No reaction was observed under the same conditions with a modified c-MYC GQ containing a C instead of a G in position 3 (MYCm, $d[\text{TTCAG}_3 \text{TAG}_3 \text{TG}_3 \text{TA}_3]$; Figure S18).

The chemoselectivity of the reaction can be rationalized by considering the secondary structure of the c-MYC GQ (Figure 3), which shows that G3 is exposed to the solvent and does not participate in the formation of the G-quartets. As a consequence, it has a highly accessible N7 nucleophile that can react with the bulky ruthenium complex. Importantly, circular dichroism experiments revealed that the GQ secondary structure is disrupted in the ruthenium adduct MYC-[Ru] (Figure S3, right). Control experiments with other related parallel quadruplex such as c-KITI, which also presents non-stacked guanines, revealed a similar reactivity pattern. Mass spectrometry shows that the promoter is modified by the ruthenium complex at the expected positions (Figures S21 and S22), although curiously, in this case, CD analysis revealed that the quadruplex remained mostly intact (Figure S4).12

With the above molecular information, we explored whether Ru complex 1 could affect the expression of the oncogene c-MYC, since this oncogene is involved in many important cellular processes.13 This was analyzed by real-time quantitative reverse transcription PCR (qRT-PCR) in HeLa and Vero cells (Figure 4 and Figure S26, respectively), using porphyrin TMPyP4, a G Q binder that is known to repress the expression of c-MYC,14 as a control. As shown in Figure 4, HeLa cells treated with 1 (100 $\mu M$) in Dulbecco’s modified Eagle’s medium (DMEM) presented a modest but significant increase in the transcription of c-MYC compared to untreated cells (80% at 16 h and 200% at 48 h).15 As expected, treatment with TMPyP4 led to a 60% decrease in the cellular levels of c-MYC mRNA after 48 h.

We also analyzed the expression of c-MYC protein by western blot. In agreement with the qRT-PCR results, treatment of cells with 100 $\mu M$ of 1, led to a noticeable increase in the levels of c-MYC protein (Figure 4, averaged 40%
mRNA levels increased after irradiation and then lysed, and c-MYC was present even higher kinetic stability in contrast to the chloride ligand (Figure 5) and are kinetically stable but undergo rapid ligand exchange upon irradiation (71% conversion, Figure S17). The acetyl-methionine derivative 5 presents even higher kinetic stability than complex 4, and no traces of the metalation adducts, or even of the aquo derivative 2, were observed after several hours in the dark. However, irradiation for 30 min triggers efficient covalent metalation of the GQ (Figure S25). In contrast to the chloride complex 1, in vitro experiments with the thioether derivative 4 showed that it does not react with the c-MYC quadruplex in the dark after 30 min, but yields the desired GQ monoadduct upon irradiation (71% conversion, Figure S17). The acetyl-methionine derivative 5 presents even higher kinetic stability than complex 4, and no traces of the metalation adducts, or even of the aquo derivative 2, were observed after several hours in the dark. However, irradiation for 30 min triggers efficient covalent metalation of the GQ (Figure S25). We also analyzed the effect of complex 5 on transcription of the c-MYC gene by qRT-PCR in HeLa cells. As shown in the Figure 5 (left), c-MYC mRNA levels increased after irradiation, becoming similar to those observed after treatment with the aquo complex 2. Western-blot analysis confirmed that enhancement of the gene expression only took place in the presence of light, while the levels of the protein c-MYC did not change when the cells were kept in the dark (Figure 5 and Figure S29). As expected, both qRT-PCR and western-blot experiments showed that the aquo compound 2 is active both in the dark and under irradiation. Moreover, control experiments confirmed that the irradiation does not have any measurable effect on cell viability (Figure S28), and that the complex does not generate significant amounts of singlet oxygen upon irradiation (Figure S25).

In conclusion, we have demonstrated the selective modification of accessible guanines flanking the GQ of c-MYC with designed bulky ruthenium complexes. Importantly, the bioorthogonal metalation enhances the expression of the oncogene c-MYC. Given that an increase in c-MYC transcription has been shown to be important in several cancers, in particular for the renewal ability of cancer stem cells, our discovery might lead to interesting biological applications.

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Similar results were obtained with the Vero mammalian cell line (see Figure S26).

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