SYNTHESES OF COPPER COMPLEXES OF NICOTINOHYDROXAMIC AND ISONICOTINOHYDROXAMIC ACIDS.

**ALIYU, A. O.**; **EGWAIKHIDE, A. P.** & **GIMBA, C. E.**

Department of Chemistry
Nigerian Defence Academy, Kaduna, Nigeria.

Department of Chemistry and Centre for Biomaterials Research
University of Benin, Nigeria

Department of Chemistry
Ahmadu Bello University, Zaria Nigeria

*(Corresponding author)*

 pegwaikhide@yahoo.com

---

**ABSTRACT**

Nicotinohydroxamic acid (NHA) and isonicotinohydroxamic acid (INHA) were synthesized, characterized by electronic and spectral studies, magnetic measurements and their pKa determined spectrophotometrically as 8.68 ± 0.02 in aqueous medium of 0.1mol dm⁻³ = ionic strength. The composition of the complexes was determined by Job's plot. The ratios of Cu²⁺ to ligands under investigation were M:L. The formation constants obtained and the possible binding modes for the complexes in solid states are discussed. Spectral studies of the isolated complexes indicate tetragonally distorted octahedral geometry via (O,O) and (N,O) coordination modes. The magnetic moments obtained for the complexes are in the range 1.57-1.79B.M. Microbial sensitivity test carried out on the ligands and their isolated complexes showed no activity on the microorganisms under investigation.

**Keywords:** Nicotinohydroxamic acid, isonicotinohydroxamic acid, IR spectra, ionic strength, Job's plot, pKa, microbial sensitivity.

---

**INTRODUCTION**

hydroxamic acids have general formula RCONHOH. These acids are much weaker acids than the structurally related carboxylic acids RCOOH (Celine 2000). Hydroxamic acids are ubiquitous in nature and are associated with iron transport bacteria (Nwabueze 1996). The selectivity of the mechanism of iron transport phenomena is important since other metal ions, which may be essential or toxic to the organism are present in the environment (Kehl 1982; Raymond 1990 and Crumbliss 1991). Hydroxamic acids with one or more –CONHOH groups have been extensively studied in relation to their pharmacological, toxicological and pathological properties (Paniago & Carvalho 1988; Mclachlan et al. 1983; Fatima et al. 2002) which is related with their ability to form metal ion complexes. Medical applications of the hydroxamates which utilize their affinity for high charge density metal ion include the possible use of the metal complexes as imaging agents (Biljana et al. 2002; Hirsova & Koldovish 1969). Hydroxamic acids are constituent of antibiotics, growth factors, food additives, tumor inhibitors and cell division factors (Albrecht & Crumbliss 1981; Hartley et al. 1980; Martell et al. 1981).

With regards to the strong ability of the hydroxamic acids to form chelates, clarification of their interactions with metal ions of particular biological effect is necessary. In the present study, equilibrium and structural studies have been performed on the copper (II) complexes of nicotinohydroxamic acid and isonicotinohydroxamic acid.

**MATERIALS AND METHODS**

Ethylnicotinate and ethylisonicotinate were used as purchased without further purification. Water was doubly distilled and degassed using purified N₂. All other reagents were used as supplied. Radiometer Copenhagen Research pH meter was used for pH measurement. IR spectra were recorded on ATI Maltson Genesis series FTR™ machine as Nujol mulls in the 4000-2000cm⁻¹ spectra region. MSB AUTO magnetic susceptibility balance was used to measure room temperature magnetic susceptibility.

Nicotinohydroxamic acid was prepared by adding 2.3g sodium metal in 50cm³ to 6.9g NH₂OH. HCl dissolved in 100cm³ MeOH. The mixture was stirred to room temperature and 15.12g ethylnicotinate was added. The mixture was stirred for 40min. and another solution of 2.3g Na in MeOH was added and stirring was continued for another 10min. The mixture was filtered to remove the precipitated NaCl and the filtrate was concentrated using a rotary evaporator and left in a refrigerator to crystallize. The crystals were removed by filtration and recrystallized from EtOH with 55% yield. Similarly, isonicotinhydroxamic acid was prepared by using15.12g ethylinicotinate 2.3g Na metal in 50cm³ of MeOH respectively.

\[ \text{[Cu(NHA)]}_2\text{H}_2\text{O} \quad \text{and} \quad \text{[Cu(INHA)]}_2\text{H}_2\text{O} \]

were prepared using 0.556g NHA and INHA in 20cm³ of MeOH added to 0.5g of CuSO₄₅H₂O in cold water. The mixture was allowed to stay for 2hr to allow the precipitate to settle. A green coloured precipitate was removed by filtration, washed with small aliquots of Et₂O and dried over silica gel in a vacuum desiccator.

**Equilibrium Studies:** The pKa values for the ligands were determined spectrophotometrically as described by Albert and Sergent (1971) using boric acid and borax of ionic strength 0.1mol dm⁻³ and 0.025mol dm⁻³ buffers for NHA and INHA ligands (Aliyu et al. 2008).

Antimicrobial screening: All media and bacterial suspensions were prepared as described by Cruickshank (1965). The antimicrobial
activity of the test compounds was assayed against six bacterial strains of three Gram + ve and three Gram–ve (Aliyu et al. 2008).

RESULTS AND DISCUSSION
The high basicity of the ligands under study may be ascribed to the positive inductive effect of the bulky pyridine group attached to the functional groups of NHA and INHA respectively. The pKa values of the ligands are 8.68 ± 0.02 for NHA and 8.68 ± 0.05 for INHA. The absorption spectra of solutions containing a constant metal concentration but variable ligand molar concentrations of NHA and INHA are shown in Figs 1 and 2. While graphical matrix rank analysis of the absorbance data generated from similar solution for NHA and INHA are indicated in figures 3 and 4.
The shape of the graphs (Figs 1 and 2) and the absence of an isosbestic point are typical of systems containing only one complex specie (Hartley et al. 1980).

Several equilibrium models were tried but it was only in ML\textsuperscript{2} model that convergence was achieved. The complex composition was determined by Job’s plot as shown in figures 5 and 6.

The ratios of Cu\textsuperscript{II} to the ligards under investigation were ML\textsuperscript{2}. Table 1 gives the analytical data and some physico-chemical properties of copper (II) complexes. There observed magnetic moments at room temperature were between 1.57 and 1.79MB thus ruling out the possibility of Cu – Cu interaction in these complexes (Nicholls 1979). The range of magnetic moments is irrespective of the stereochemistry. The visible spectra of copper(II) hydroxamate complexes, ranges between 630 – 810nm as shown in Figs 7 and 8.
FIG 5: CONTINUOUS VARIATION (JOBS PLOT) METHOD Cu^{2+}/INHA SYSTEM

FIG 6: CONTINUOUS VARIATION (JOBS PLOT) METHOD Cu^{2+}/INHA SYSTEM

FIG 7: VISIBLE SPECTRUM OF [Cu (NHA)_2 ] 2H_2O COMPLEX.
These bands are assigned d→d transitions of copper (II) ions and encompasses several overlapping bands. The electronic spectra of CuII – NHA and CuII-INHA (Figs. 7 and 8) do not resemble the spectra of standard square planar copper (II) complexes but more closely agree with the spectra of established tetragonally distorted octahedral complexes. The range of standard square planar copper (II) complexes is between 714 – 500nm, (Nichol 1979; Cotton & Wilkinson 1980).

Table 2 shows the diagnostic IR band of the metal free ligands and their corresponding complexes. In the spectra of the metal complexes, the observed band in the region of 3374.43 cm⁻¹ and 3420 cm⁻¹ were assigned to ν(NH) stretching vibration in CuII – NHA and CuII – INHA respectively. The observed decrease in the frequency of this band is about 44cm⁻¹ relative to the position in the metal free ligand and this is due to the deprotonation of the nitrogen atom of the hydroxamate group thereby indicating complexation through the nitrogen atom (CuII – NHA).

But in the CuII – INHA complex, there was little or no increase in the observed frequency relative to its metal free ligand implying the absence of coordination through the nitrogen atom.

The band around 1605.40cm⁻¹ and 1561.58cm⁻¹ in the spectra of copper (II) hydroxamate complexes were assigned to the ketonic carbonyl frequencies. The decrease in the frequencies to about 54.21 cm⁻¹ and 43.43 cm⁻¹ respectively relative to the position of the metal free ligand suggests of coordination through the ketonic carbonyl oxygen of the hydroxamate group (Biljana et al. 2002; Chatteryee 1978; West 1969). The V(CN) frequencies were observed around 1112.09cm⁻¹ and 1129cm⁻¹ for CuII – NHA and CuII – INHA respectively. The observed increase in the frequencies relative to their metal free ligand is expected. Based on the IR data therefore, the following bonding modes were suggested for the copper (II) hydroxamate complexes. CuII – NHA bonding mode is (N, O) while CuII-INHA bonding is (O, O) as suggested.

FIG 8: VISIBLE SPECTRUM OF [CU(INHA)₂] 2H₂O COMPLEX

FIG 9. SUGGESTED STRUCTURE FOR THE (N, O) BONDING MODE FOR TETRAHEDRALLY DISTORTED OCTAHEDRON COORDINATED COMPLEX OF COPPER (II) HYDROXAMATE.

FIG 10; SUGGESTED STRUCTURE FOR (O, O) BONDING FOR SQUARE PLANAR COMPLEXES OF COPPER (II) HYDROXAMATE.
The microbial sensitivity tests carried out on the ligands and the copper (II) complexes show no activity on the microorganism under investigation as shown in Table 3.

**TABLE 1: ANALYTICAL DATA AND PHYSICO-CHEMICAL PROPERTIES FOR THE ISOLATED COMPLEXES (CALCULATED%)**

| Compound          | Formular weight | Melting point/Decomposition point °C | Found Metal% | μeff Bohr Magneton at 298k | λmax (nm) | Colour       | Assignment |
|-------------------|-----------------|-------------------------------------|--------------|-----------------------------|-----------|--------------|------------|
| Cu(NHA)2·2H2O    | 375.5           | 269                                 | 16.82(16.91) | 1.57                        | 800       | Light green  | d→d        |
| Cu(INHA)2·2H2O   | 375.5           | 276                                 | 16.82(16.91) | 1.79                        | 800       | Green        | d→d        |

**TABLE 2: DIAGNOSTIC IR DATA FOR THE COMPLEXES (cm⁻¹)**

| Compound         | V(NH) cm⁻¹ | ΔV(NH) cm⁻¹ | V(C = O)cm⁻¹ | ΔV(C = O) cm⁻¹ |
|------------------|------------|-------------|--------------|----------------|
| NHA              | 3418.00    | -           | 1659.61      | -              |
| Cu(NHA)2·2H2O   | 3374.43    | -44.00      | 1605.40      | -54.21         |
| INHA             | 3422.59    | -2.59       | 1561.58      | -43.43         |
| Cu(INHA)2·2H2O  | 3420.00    | -           |              |                |

Key:
NHA: Nicotinohydroxamic acid
INHA: Isonicotinohydroxamic acid

**TABLE 3: MICROBIAL SENSITIVITY TEST FOR THE LIGANDS AND THEIR COPPER (II) COMPLEXES**

| Ligands/complexes | S. aureus | S. typhium | E. coli | α-hemolytic strep | Klebsiella | Pseudomonas |
|-------------------|-----------|------------|---------|-------------------|------------|------------|
| NHA               | -         | -          | -       | -                 | -          | -          |
| Cu(NHA)2·2H2O     | -         | -          | -       | -                 | -          | -          |
| INHA              | -         | -          | -       | -                 | -          | -          |
| Cu(INHA)2·2H2O    | -         | -          | -       | -                 | -          | -          |

Key – not present

The apparent drug resistance exhibited by the four species of gram-ve and gram+ve bacterial strains tested during this study suggests these could be nosocomial (hospital) microorganisms. Genetically developed multi-drug resistance mechanisms could have arisen in these microbial strains as a result of their rampant exposure to several antibiotics, characteristic of hospital environment.

These mechanisms include non permeability of the microbial outer membrane to chemical bactericides, development of multidrug resistant pump mechanisms that expel absorbed drugs by microbes, inactivation of absorbed drugs by antimicrobials through their chemical modification, bypassing of metabolic sequence inhibited by drugs and increase in the production of metabolite target of antimicrobial agents (Prescott et al. 1999).

Albert, A. & Serjeant, E. P. 1971. The determination of ionization constant. 2nd Edition p 44. Chapman and Hall Ltd. London

Albrecht-Gary A. M. & Crumbliss A. L. 1998. Metal Ions in Biological systems. Vol. 35 Marcel Dekker, New York

Aliyu, A. O., Egwuikhide, P. A Maikaye, D. B & Gimba, C. E. 2008. Complex formation between transition metals. Pakistan Journal of scientific and industrial research, 2(1):213-219.

Biljana, N., Nikola, K., Krisimir, S., & Dranen V. T. 2002. Complex formation between transition metals and 2 – pyrrolidine- 5 – hydroxamic acid. Acta Chemistry Slovenia 49:325-535.

Celine, J. M. & Kelvin, B. N. 2000. Hydroxamic acids-ion chelators, aspirin analogues, nitric oxide donors and structurally diversed metals complexes. http://www.irishscientist.

Chatterje, B. 1978. Coordination Chemistry, Revision. Elsevier Science limited London

Cotton, F.A. & Wilkinson, G. 1980. Advance Inorganic Chemistry Comprehensive test 4th Edition, Wiley, New York

Cruickshark, R. 1965. Medical micro-biology. 2nd Edition. Church and Livingstone. U.K

Crumbliss, A. L. 1991. Handbook of microbial iron chelate. 2nd Edition G. Winkelmann CRC. Press. New York.

Fatima, N, Maqsood, Z. T. & Kazmi, S. A. 2004. Complexation of vanadium (iv) with hydroamate chelators and Their stability relation with pH. Journal of Chemical Society of Pakistan 24:49

Hartley, F. R.; Burgess, C. & Alcocook, R. M. 1980. Solution
Kehl, H. 1982. *Chemistry and Biology of hydroxamic acids*. Karger, New York.

Nicholl, D. 1979. *Complexes and first row transition elements*. Macmillan Press Ltd. London.

Martell, A. E.; Anderson, W. F. & Badman, D. G. 1981. *Development of iron Chelators for Clinical use*. Elsevier. New York

Mclachlan, D. R. C.; Faenell, B.; Gallin, H.; Karlk, S.; Eichorn, G. & DeBoni, U. 1983. *Biological Aspects of metal and metal – related diseases*. Raven Press. New York

Nwabueze, J. N. 1996. Complexes iron(III) with cyclopropane carbo-and cyclohexyl acetoacylhydroxamic acids. *Transition metal chemistry*. Vol. 21 pp 258 – 261. Thermochemical acta, Elservier London.

Paniago, E. B & Carvalho, S. 1988. Ciencia e Cufura 40 pp 629. University of Mato Grosso do Sul (Hu – UFMS)

Prescett, L. M.; Harley, J. P. & Klein D. A. 1999. *Antimicrobial Chemotherapy. Drug resistance in Microbiology* 4th Edition pp 690. WCB/MCGram Hill.

West, T. S. 1969. Complexometry with EDTA and related reagents. BDH Chemicals Ltd London.