Prognostic impact of discrepant Ki67 and mitotic index on hormone receptor-positive, HER2-negative breast carcinoma

L Rossi1, E Laas2, P Mallon3, A Vincent-Salomon4, J-M Guinebretiere4, F Lerebours5, R Rouzier1,6, J-Y Pierga5 and F Reyal *,1,7,8

1Département de chirurgie, Institut Curie, 75005 Paris, France; 2Service de Gynécologie-Obstétrique, Hôpital Tenon, 75020 Paris, France; 3Craigavon Area Hospital Breast Unit BT63 5QQ, Portadown, Northern Ireland; 4Département de biologie des tumeurs, Institut Curie, 75005 Paris, France; 5Département d’oncologie médicale, Institut Curie, 75005 Paris, France; 6Equipe d’Accueil 7285, ‘Risk and Safety in Clinical Medicine for women and Perinatal Health’, University Versailles-Saint-Quentin, 78180 Montigny-le-Bretonneux, France; 7Département de recherche translationnelle, Equipe résidu tumoral et réponse au traitement, Institut Curie, 75005 Paris, France and 8INSERM U932 Immunity and Cancer, Institut Curie, 75005 Paris, France

Background: Inconsistencies between mitotic index (MI) and Ki67 measures have been identified in many breast tumour samples. The aim of this study was to describe the prognosis of hormone receptor-positive (HR+) HER2—tumours having discrepant MI and Ki67.

Methods: We included a cohort of breast cancer patients initially treated by surgery between 2001 and 2005 in the Institut Curie. Breast cancer-specific survival (BCSS) and disease-free survival (DFS) were analysed according to three proliferation groups: high MI/high Ki67 (MI = 3, Ki67 > 20%), low MI/low Ki67 (MI < 3, Ki67 ≤ 20%) and discrepant (high MI/low Ki67 or low MI/high Ki67).

Results: Among the 1430 patients, 19.6% had discrepant Ki67 and MI, 11.6% had high markers and 68.8% had low markers. The 5-year BCSS was 95.8%, 95% CI (0.93–0.98) in the discrepant group, 99.3%, 95% CI (0.993–0.999) in the low-proliferation group and 91.8%, 95% CI (0.88–0.96) in the high-proliferation group. In multivariate analysis, the survival of the discrepant group was lower than that of the low-proliferation group: BCSS hazard ratio (HR) = 3.01 (1.32–6.84; P = 0.008) and DFS HR = 2.07, 95% CI (1.31–3.26; P = 0.002). Among grade 2 tumours in multivariate analysis, DFS of the discrepant group was lower than that of the low MI/low Ki67 group: HR = 1.98, 95% CI (1.14–3.46), P = 0.02. Regarding BCSS, the obtained results were similar.

Conclusion: The prognosis of patients with discrepant MI and Ki67 appears intermediate between that of low MI/low Ki67 and high MI/high Ki67 groups. These markers should be jointly analysed to clarify prognosis.
Discrepant mitotic index and Ki67 in breast cancer: prognosis

or to a significant fraction of growing cells. Thus, the correlation between the number of mitosis and the proliferation rate is not strictly linear (Beresford et al, 2006).

The MI is a major prognostic factor in that patients with high MI have an increased risk of death (Stuart-Harris et al, 2008) and recurrence regardless of tumour size (Baak et al, 2008) and lymph node status (Baak et al, 2005). There is also evidence to suggest that MI predicts response to adjuvant (Andre et al, 2005; Jansen et al, 2006) and neoadjuvant (Vincent-Salomon et al, 2004; Penault-Llorca et al, 2008) chemotherapy.

The nuclear Ki67 antigen is expressed in the cell cycle phases G1, S, G2 and M, but not in G0. The level of expression of the Ki67 protein varies during the cell cycle. Rates are low in G1 and early S phase and increase to a maximum at the time of mitosis. A rapid decrease in Ki67 expression occurs in anaphase and telophase (Lopez et al, 1991). To identify cells expressing the Ki67 protein, immunohistochemistry is performed using a mouse monoclonal antibody. MIB-1 antibody has been validated more frequently (Harris et al, 2007). Immunohistochemistry can be used on tissue sections embedded in paraffin. The half-life of Ki67 is estimated to be ~60–90 min, and the duration of a cell cycle is ~24 h. Normal breast tissue and oestrogen receptor positive (ER +) tumour cells express Ki67 at low levels (<3%), whereas ER− tumours exhibit higher expression (Clarke et al, 1997). The percentage of tumour cells expressing Ki67 reflects the percentage of cells in the mitotic cycle within the tumour. Several studies support the prognostic value of Ki67 (Reyal et al, 2013). In the meta-analysis by De Azambuja et al (2007), patients with high Ki67 had worse disease-free survival (DFS; hazard ratio (HR) = 1.93 (1.74–2.14), P < 0.001) and overall survival (OS; HR = 1.95 (1.70–2.24), P < 0.001). Ki67 was also an independent prognostic factor in node-negative and node-positive patients. Stuart-Harris et al (2008) found similar results in their meta-analysis. The ability of Ki67 to predict the response to chemotherapy is controversial (Dowsett et al, 2011). Viale et al in 2008, in their analysis of 1521 patients without lymph node involvement, did not show any predictive value of Ki67 in response to adjuvant chemotherapy.

Mitotic index and Ki67 both evaluate the proliferative potential of tumours; however, we have observed in some patients’ discordance between the two markers. The aim of this study was to determine the characteristics and the prognostic of HR + HER2− breast cancers having high Ki67/low MI or low Ki67/high MI.

MATERIALS AND METHODS

Patients. This study included a cohort of patients with a first invasive breast cancer treated in the Institut Curie between January 2001 and December 2005. The inclusion criteria were HR + (ER + and/or progesterone receptor-positive (PR +)) HER2− tumours with a pathological size ranging from T1 to T3 (TNM classification) with or without lymph node involvement, no distant metastasis at diagnosis and surgery as first treatment. Patients with missing values for Ki67 or MI were excluded because they were not classifiable in a ‘proliferation group’ required for the analysis.

Data measurement. All data were collected prospectively.

Pathological characteristics. The resected specimens were fixed in AFA (alcohol–formalin–acetic acid solution), and then embedded in paraffin and cut into sections of 3–5-μm thickness. The slides were stained with haematoxylin, eosin and saffron. Tumour size was macroscopically determined on the fresh resected specimen. Pathological grade was assessed according to Elston & Ellis, 1991.

Mitotic index. The MI was assessed in the peripheral most cellular region of the tumour slides. The criteria of Van Diest et al (1992) were used to define mitotic figures. The MI was the sum of the number of mitosis counted on 10 consecutive high-power fields (each area of 0.307 mm²) using an optical microscope (objective ×40). According to Elston & Ellis’ grade (Elston and Ellis, 1991), the thresholds used were as follows: 1 point when <9 mitosis were counted, 2 points when 10–19 mitosis were counted and 3 points if mitoses were ≥20 per 10 fields. We considered that MI was low if the score was equal to 1 or 2, and high if the score was equal to 3. According to our microscope, this sets a cutoff to 19 mitosis mm⁻².

Immunohistochemical characteristics

Ki67 rate. Tissue sections were incubated with the anti-Ki67 monoclonal antibody (Clone MIB-1 M7240, Dako, Glostrup, Denmark). The revelation of the staining was performed using a peroxidase mouse IgG kit and dianaminobenzidine as chromogen. The validity of each procedure was verified with the staining of internal controls (lymphocytes, mitosis). The semiquantitative gross assessment was performed by determining at ×200 magnification the percentage of positive neoplastic nuclei within the area of highest positivity. All nuclei with homogeneous staining even with a light staining or only a nucleolar staining were interpreted as positive. A cutoff of >20% was used to define tumours with a high Ki67 rate.

**ER and PR status.** After rehydration and antigenic retrieval in citrate buffer (10 mM, pH 6.1), the tissue sections were stained for ER (clone 6F11, Novocastra, 1 out of 200) and PR (clone 1A6, Novocastra, 1 out of 200). Revelation of staining was performed using the Vectastain Elite ABC peroxidase mouse IgG kit (Vector, Burlingame, CA, USA) and dianaminobenzidine (Dako A/S) as chromogen. Positive and negative controls were included in each slide run. Cases were considered positive for ER and PR according to the standardized guidelines using a cutoff of ≥10% stained tumour nuclei (Balaton et al, 1996).

**HER2 status.** After rehydration and antigenic retrieval in citrate buffer (10 mM, pH 6.1), the tissue sections were stained for HER2 (clone CB11, Novocastra, 1 out of 1000). Revelation of staining was performed using the Vectastain Elite ABC peroxidase mouse IgG kit (Vector, Burlingame, CA, USA) and dianaminobenzidine (Dako A/S) as chromogen. Positive and negative controls were included in each slide run. The overexpression of HER2 was determined according to the GEFPICS guidelines with FISH performed in all cases of HER2 2+ result (Penault-Llorca et al, 2010).

Statistical analysis. Patients were classified into three ‘proliferation groups’: high MI/high Ki67 (MI = 3 and Ki67 > 20%), low MI/low Ki67 (MI = 1 or 2 and Ki67 ≤ 20%) and discrepant (high MI/low Ki67 or low MI/high Ki67). Data were analysed using χ²-test or Fisher exact test for qualitative variables and by Student’s t-test for quantitative variables. Breast cancer-specific survival was described using the time from diagnosis to the time of death of breast cancer. Disease-free survival was described using the time from diagnosis to the time of a recurrence (local recurrence or lymph node recurrence or distant metastasis). Survival analyses were performed using the Kaplan–Meier estimate of the survival function. Comparison between survival curves was performed using the log-rank test. Hazard ratios were estimated using the Cox proportional hazard model. Only variables with a P-value < 0.1 in univariate analyses were included in a multivariate ascending stepwise procedure using the Cox model. There was <5% missing data for all the variables. All statistical tests were reported with two-tailed P-values and 95% CIs at an alpha level of 0.05 or lower. All statistical analyses were performed using the R software (http://cran.r-project.org, The R Foundation for Statistical Computing, Vienna, Austria, version 0.98.978, 2009–2013 Rstudio Inc.) with the rms and survival packages.

www.bjcancer.com | DOI:10.1038/bjc.2015.239
Ethical consideration. The registration of patients of the Institut Curie in this cohort received a favourable agreement from the French National Committee on Computers and Liberties (CNIL, Commission nationale de l’informatique et des libertes). Patients gave informed written consent before being registered in the cohort. The study was approved by the breast cancer study group and the comity of clinical research study of the Institut Curie.

RESULTS

Patients. Among the 1487 patients with a HR+ HER2− invasive breast cancer treated by surgery first, 52 patients were excluded because they had a missing value for Ki67 or MI and five because they had missing data on the cause of death. Consequently, 1430 women treated in the Institut Curie between 2001 and 2005 were analysed (Table 1).

Proliferation groups. Proliferation groups were distributed as follows: 984 patients (68.8%) had low MI/low Ki67, 166 (11.6%) high MI/high Ki67 and 280 (19.6%) discrepant markers (227 (15.9%) low MI/high Ki67 and 53 (3.7%) high MI/low Ki67; Table 1). There was no statistical difference between the high MI/low Ki67 and the low MI/high Ki67 groups, regarding patient characteristics, tumour characteristics or administered treatments.

The median Ki67 in the high MI/low Ki67 group was 15%. This median of 15% was significantly higher than that of the low MI/low Ki67 group (median = 8%). In the low MI/high Ki67 subgroup, the median Ki67 was 30%. This median of 30% was significantly lower than that of the high MI/high Ki67 group (median = 45.8%). Mitotic index of this subgroup low MI/high Ki67 was measured equal to 1 in 46% of cases (104 patients) and to 2 in 54.2% of cases (123 patients).

In the high MI group (MI = 3), the first quartile value of Ki67 was 21.5%. The low Ki67 values of this group were not only values nearer to the threshold of 20% (Figure 1). When MI was low (equal to 1 or 2), around one quarter of the Ki67 values were >20% and were not only near the threshold of 20% but also could reach really high values.

Tumour sizes greater than 20 mm in the discrepant group were more frequent (31.5%) than in the low MI/low Ki67 group (19.5%) and less frequent than in the high MI/high Ki67 one (43.4%), P<0.001 (Table 1). Moreover, axillary lymph nodes in the discrepant group were more often metastatic (43.1%) than in the low MI/low Ki67 (28.9%) group and less often than in the high MI/high Ki67 one (47%), P<0.001. The discrepant group had grade 3 tumours in higher proportion (20.0%) than the low MI/low Ki67 group (1%) and in lower proportion than the high MI/high Ki67 group (68.1%), P<0.001 (Table 1).

BCSS and DFS. The median follow-up was 73 months. The BCSS rate at 5 years was 99.3%, 95% CI (0.993–0.999) in the low MI/low Ki67 group, 95.8%, 95% CI (0.93–0.98) in the discrepant group and 91.8%, 95% CI (0.88–0.96) in the high MI/high Ki67 group. In the univariate analysis, the discrepant group had an intermediate BCSS (Figure 2A) and DFS (Figure 2B) compared with the two other groups. The relative risks of death due to breast cancer of the discrepant and the high MI/high Ki67 groups were higher than those of the low MI/low Ki67 group, HR = 4.35, 95% CI (2.06–9.21; P = 0.0001) and HR = 9.77, 95% CI (4.80–19.89; P<0.0001), respectively (Figure 2A). Similarly, the relative risks of recurrence were higher in the discrepant and in the high MI/high Ki67 groups compared with the low MI/low Ki67 group, HR = 2.49, 95% CI (1.64–3.76; P<0.0001) and HR = 4.97, 95% CI (3.22–7.43; P<0.0001), respectively (Figure 2B).

Table 1. Characteristics of the HR+ HER2− population according to proliferation groups

| Variable/category | Low MI/low Ki67 | Discrepant | High MI/high Ki67 | P-value | All; N (%) |
|-------------------|----------------|------------|------------------|---------|-----------|
| Age (years)       |                |            |                  |         |           |
| <40               | 38             | 3.9        | 25               | 8.9     | 18        | 10.8     | <0.001 | 81 (5.7) |
| >40               | 946            | 96.1       | 255              | 91.1    | 148       | 89.2     |         | 1349 (94.3) |
| Tumour size (mm)  |                |            |                  |         |           |
| <20               | 792            | 80.5       | 192              | 68.6    | 94        | 56.6     | <0.001 | 1078 (75.4) |
| >20               | 192            | 19.5       | 88               | 31.5    | 72        | 43.4     |         | 352 (24.6) |
| Axillary lymph nodes |              |            |                  |         |           |
| Negative          | 700            | 71.1       | 159              | 56.8    | 88        | 53.0     | <0.001 | 947 (64.2) |
| Positive          | 284            | 28.9       | 121              | 43.1    | 78        | 47.0     |         | 483 (33.8) |
| Pathological type |                |            |                  |         |           |
| Ductal            | 754            | 76.6       | 238              | 85.0    | 151       | 91.0     | <0.001 | 1143 (79.9) |
| Lobular           | 146            | 14.8       | 28               | 10.0    | 11        | 6.6      |         | 185 (13.0) |
| Other             | 84             | 8.6        | 14               | 5.0     | 4         | 2.4      |         | 102 (7.1) |
| Grade             |                |            |                  |         |           |
| 1                 | 524            | 53.3       | 52               | 18.6    | 0         | 0        | <0.001 | 576 (40.3) |
| 2                 | 450            | 45.7       | 172              | 61.4    | 53        | 31.9     |         | 675 (47.2) |
| 3                 | 10             | 1.0        | 56               | 20.0    | 113       | 68.0     |         | 179 (12.5) |
| Treatment         |                |            |                  |         |           |
| None              | 267            | 27.1       | 30               | 10.7    | 8         | 4.8      | <0.001 | 305 (21.3) |
| Chemotherapy alone| 9              | 0.9        | 4                | 1.5     | 7         | 4.2      |         | 20 (1.4) |
| Hormone therapy alone | 457       | 46.4       | 114              | 40.7    | 40        | 24.1     |         | 611 (42.7) |
| Both              | 251            | 25.6       | 132              | 47.1    | 111       | 66.9     |         | 494 (34.6) |

Abbreviations: HR = hazard ratio; MI = mitotic index; N = number of patients; Other = apocrine, comedocarcinoma, mucinous carcinoma, medullary carcinoma, Paget’s disease, papillary carcinoma, tubular, undifferentiated or unknown.
In the multivariate analysis, DFS of the discrepant group still was lower than that of the low MI/low Ki67 group: HR = 2.07, 95% CI 1.31–3.26; P = 0.002 (Table 2). Regarding BCSS in the multivariate analysis, the same type of results were obtained with a HR = 3.01, 95% CI (1.32–6.84; P = 0.008) for the discrepant group compared with the low-proliferative group. The high MI/high Ki67 group also had a greater risk of dying from breast cancer when compared with the low MI/low Ki67 group: HR = 4.30, 95% CI (1.63–11.30; P = 0.003; data not shown).

As the discrepant group had intermediate BCSS and DFS, we tried to find a discriminatory factor and analysed survival according to the PR status (Prat et al, 2013). In the high MI/high Ki67 and the low MI/low Ki67 groups, the PR level was not relevant regarding BCSS (Supplementary Figure) or DFS. In the discrepant group, patients with PR ≥20% tended to have better BCSS (HR = 0.50, 95% CI (0.15–1.06), P = 0.06; Supplementary Figure) and better DFS (HR = 0.61, 95% CI (0.31–1.18), P = 0.14).

As for grade 2 tumours the decision whether or not to administer chemotherapy is still more difficult, we also analysed whether the proliferation groups had an impact on the survival in this subgroup. In grade 2 HR + HER2− tumour subgroup (N = 675), patients with discrepant MI and Ki67 had worse DFS than the low-proliferative group: HR = 2.18 (1.28–3.72; P = 0.004; Figure 3). Patients with high MI/high Ki67 had even worse DFS than the low-proliferative group: HR = 3.15 (1.57–6.32; P = 0.001; Figure 3). In the multivariate analysis, DFS of the discrepant and the high MI/high Ki67 group was still significantly lower than that of the low MI/low Ki67 group: HR = 1.98, 95% CI (1.14–3.46), P = 0.02 and HR = 2.97, 95% CI (1.42–6.20), P = 0.004, respectively (Supplementary Table). In terms of BCSS, the obtained results were similar. In the multivariate analysis, the discrepant group had a lower BCSS than the low MI/low Ki67 group: HR = 3.08, 95% CI (1.16–8.19), P = 0.02.

**DISCUSSION**

Various studies have shown that the cell proliferation markers MI and Ki67 were positively correlated (Syparatos et al, 2002; Offersen et al, 2003; Trihia et al, 2003) and were associated with breast cancer prognosis (De Azambuja et al, 2007; Stuart-Harris et al, 2008; Yerushalmi et al, 2010). However, they differ in some cases. Our study showed that HR + HER2− tumours with disproportionate MI and Ki67 had higher tumour size, higher grade and more frequent metastatic axillary node involvement than the low Ki67/low MI group. The discrepant group showed lower tumour size, lower grade and less frequent metastatic axillary lymph nodes than the high Ki67/high MI group. BCSS and DFS of the discrepant group (HR = 3.01, 95% CI (1.32–6.84), P = 0.008 and HR = 2.07, 95% CI (1.31–3.26), P = 0.002, respectively) were situated between those of the low-proliferative and high-proliferative groups. There was no significant difference regarding the tumour’s characteristics within the discrepant group between the high MI/low Ki67 and the low MI/high Ki67 tumours, except for grade.

There is no consensual cutoff value to distinguish low and high Ki67 expression reflecting low or high tumour’s proliferative potential. According to the St Gallen guidelines 2013 (Goldhirsch et al, 2013), the threshold to separate luminal A and luminal B cancers is 14%, from one study of gene-expression profiling (Cheang et al, 2009). In terms of prognosis, the threshold of 20% was suggested in these guidelines, but should be adjusted on local laboratory specificities because of the high degree of variation of Ki67 measurement. We chose the cutoff point of 20% as it was the threshold set in clinical routine in our institution to discriminate between high- and low-proliferation tumours.

One previous study focused on the prognosis of breast cancer with discrepant MI and Ki67 (Jalava et al, 2006). It included 265 invasive breast cancer cases and split them into four groups on the
basis of Ki67 (cutoff 15%) and MI (cutoff 17 mitoses mm$^{-2}$). Patients with high MI/low Ki67 had an OS approaching that of the high MI/high Ki67 group, whereas patients with low MI/high Ki67 had a better prognosis close to that of the low MI/low Ki67 group. Some care is needed while interpreting these data because of the limited number of patients. Hence, we initially analysed our data split into four groups and saw that the two discrepant groups (high MI/low Ki67 and low MI/high Ki67) were not significantly different from each other in terms of tumour characteristics and survival. There was no difference when comparing the BCSS of the low MI/high Ki67 group to the BCSS of the high MI/low Ki67 group: HR = 0.89 (0.25–3.14), $P = 0.86$ (log-rank test). There was no difference either when regarding DFS of the low MI/high Ki67 compared with that of the high MI/low Ki67 group: HR = 0.77, 95% CI (0.35–1.68), $P = 0.51$. As these two groups looked similar, we decided to analyse them together and to form a larger group of patients.

The median Ki67 in the high MI/low Ki67 group was 15%. It means that only 50% of the values of this subgroup were between 15 and 20%, close to the threshold and that a significant part of the values of Ki67 was really low. This median of 15% was significantly higher than that of the low MI/low Ki67 group (median = 8%). In the low MI/high Ki67 subgroup, the median Ki67 was 30%. This subgroup contained high values of Ki67, distant from the threshold of 20%. This median of 30% was significantly lower than that of the high MI/high Ki67 group (median = 45.8%). Moreover, MI of this subgroup low MI/high Ki67 was measured equal to 1 in 46% of cases (104 patients) and to 2 in 54.2% of cases (123 patients).

Low MI/high Ki67 could be explained by the fact that Ki67 protein reflects the proportion of cells in cycle, indicating the cells with propensity to divide. These cells may also stop their cycles or get into apoptosis and not reach mitosis phase. High MI/low Ki67 is harder to explain. MI could be overstated because of mitotic blocks for DNA repair, leading to a larger number of mitotic figures being counted. However, it includes a small number of cells and this hypothesis is insufficient to explain the 3.7% of high MI/low Ki67 tumours. Measurement errors of Ki67 cannot be excluded but the semiquantitative method used showed its equivalence to the quantitative one and its intraobserver and extraobserver reproducibility (Caly et al, 2004).

If these explanations were enough, patients with high MI/low Ki67 should have a similar prognosis to those with high MI/high Ki67 because of the underestimation of Ki67, as found by Jalava...
et al (2006). Yet, BCSS and DFS of patients with high MI/low Ki67 tumours were significantly better than those with high MI/high Ki67 and were comparable to those of low MI/high Ki67 tumours, forming the discrepant group.

Studies are more clear on the prognostic value of MI than on that of Ki67 (Offersten et al, 2003), and the interpretation of the latter is still not standardised. Some studies have shown the prognostic value of Ki67 (De Azambuja et al, 2007; Stuart-Harris et al, 2008) and it has the advantage of identifying the proliferative cells outside a mitotic stage (Vincent-Salomon et al, 2004). However, as suggested by other studies (Spryatos et al, 2002; Gudlaugsson et al, 2013), we demonstrated here that in terms of survival, it is interesting to consider the two markers together and not only independently. This particularly applies to the situations where MI is equal to 2 or when Ki67 is between 10 and 25%, in which the choice to indicate chemotherapy is not unquestionable. These results need to be confirmed in further studies with a longer follow-up, as in HR+ HER2− populations breast cancer recurrences might occur later.

It is very interesting to notice that within the HR+ HER2− grade 2 tumour subgroup the analysis of MI and Ki67 together could help to precise different prognosis. In terms of DFS (Figure 3, Supplementary Table) and BCSS (data not shown), the discrepant group had lower survival than the low MI/low Ki67 group (DFS HR = 1.98, 95% CI (1.14–3.46), P = 0.02 and BCSS HR = 3.08, 95% CI (1.16–8.19), P = 0.02). It has been shown that Ki67 could help to classify grade 2 tumours into two distinct subgroups (Aleskandarany et al, 2011); however, here we highlight that it should be considered jointly with MI to avoid misclassifications towards prognosis.

Progestosterone receptor is part of the ER signalling network and has been shown to be a major prognosis stratifier within the luminal breast cancers (Braun et al, 2013; Nishimukai et al, 2014). In our study, the analysis of BCSS (Supplementary Figure) or DFS of HR+ HER− patients did not seem to be affected by the PR status (cutoff 20% (Prat et al, 2013)); however, in the discrepant group patients with PR ≥ 20% had almost significantly better BCSS (HR = 0.50, 95% CI (0.15–1.06), log-rank P = 0.06). Further research will specify the place given to tumour characterisation techniques by multigene microarray (Albain et al, 2010) as well as other complementary prognostic markers such as the expression of Bcl-2 (B-cell lymphoma 2) anti-apoptotic protein (Hwang et al, 2012) and androgen receptor (Veronesi et al, 2014). Prognosis of HR+ HER2− tumours with inconsistent MI and Ki67 is intermediate between those of high MI/high Ki67 and low MI/low Ki67 tumours, including grade 2 tumours. This study suggests that the combined analysis of Ki67 and MI is important in accurately determining the aggressiveness of HR+ HER2− breast cancers.

Baak JPA, van Diest PJ, Janssen EAM, Gudlaugsson E, Voorhorst FJ, van der Wall E, Vermorken JB. Collaborators of Multicenter Morphometric Mammary Carcinoma Project (MMMC) (2008) Proliferation accurately identifies the high-risk patients among small, low-grade, lymph node-negative invasive breast cancers. Ann Oncol 19: 649–654.

Baak JPA, van Diest PJ, Voorhorst FJ, van der Wall E, Beex LVAM, Vermorken JB, Janssen EAM (2005) Prospective multicenter validation of the independent prognostic value of the mitotic activity index in lymph node-negative invasive breast cancers younger than 55 years. J Clin Oncol 23: 5993–6001.

Balaton AL, Coindre JM, Collin F, Ettore F, Fiche M, Jacquemier J, Le Doussal V, Mandard AM, Mathieu MC, Mignon C, Simony-Lafontaine J, Treilleux I, Verriele V, Vilain MO, Zafrañi B (1996) Recommendations for the immunohistochemical evaluation of hormone receptors on paraffin sections of breast cancer. Study Group on Hormone Receptors using Immunohistochemistry FNCLCC/AFAAQAP. National Federation of Centres to Combat Cancer/French Association for Quality Assurance in Pathology. Ann Pathol 16: 144–148.

Berestfod MJ, Wilson GD, Makris A (2006) Measuring proliferation in breast cancer: practicalities and applications. Breast Cancer Res 8: 216.

Braun L, Mietzsch F, Seibold P, Schneeweiß A, Schirrmacher P, Chang-Chau J, Peter Sinn H, Aulmann S (2013) Slintrins breast cancer subtypes defined by estrogen receptor signalling-prognostic relevance of progesterone receptor loss. Mod Pathol 26: 1161–1171.

Caly M, Genin P, Ghuzlan AA, Elie C, Fréneaux P, Klijianenjo J, Rosty C, Sigal-Zafrañi B, Vincent-Salomon A, Dousguard A, Zidane M, Sastre-Garau X (2004) Analysis of correlation between mitotic index, MIBI score and S-phase fraction as proliferation markers in invasive breast carcinoma. Methodological aspects and prognostic value in a series of 257 cases. Anticancer Res 24: 3283–3288.

Cheang MCU, Chia SK, Voduc D, Gao D, Leung S, Snider J, Watson M, Davies S, Bernard PS, Parker JS, Perou CM, Ellis MJ, Nielsen TO (2009) Ki67 index, HER2 status, and prognosis of patients with luminal B breast cancer. J Natl Cancer Inst 101: 736–750.

Clarke RB, Howell A, Potten CS, Anderson E (1997) Dissociation between steroid receptor expression and cell proliferation in the human breast. Cancer Res 57: 4987–4991.

De Azambuja E, Cardoso F, de Castro Jr G, Colozza M, Mano MS, Durveq B, Sotiriou C, Larsmont D, Piccart-Gebhart MJ, Paesmans M (2007) Ki-67 as prognostic marker in early breast cancer: a meta-analysis of published studies involving 12,155 patients. Br J Cancer 96: 1504–1513.

Dowsett M, Nielsen TO, A’Hern R, Bartlett J, Coombes RC, Cuzick J, Ellis MJ, Henry NL, Hugh JC, Lively T, McShane L, Paik S, Perlout-Llorca F, Prudkin L, Regan M, Salter J, Sotiriou C, Smith IE, Viale G, Zujewski J, Hayes DF, International Ki-67 in Breast Cancer Working Group (2011) Assessment of Ki67 in breast cancer: recommendations from the International Ki-67 in Breast Cancer working group. J Natl Cancer Inst 103: 1656–1664.

Elston CW, Ellis IO (1991) Pathological prognostic factors in breast cancer. I. The value of histological grade in breast cancer: experience from a large study with long-term follow-up. Histopathology 19: 403–410.

Goldhirsch A, Winer EP, Coates AS, Gelber RD, Piccart-Gebhart M, Thürlimann B, Senn H-J, Panel members (2013) Personalizing the treatment of women with early breast cancer: highlights of the St Gallen International Expert Consensus on the Primary Therapy of Early Breast Cancer 2013. Ann Oncol 24: 2206–2223.

Gudlaugsson E, Klos J, Skaland I, Janssen EAM, Smaalnd R, Feng W, Shao Z, Malpica A, Baak JPA (2013) Prognostic comparison of the proliferation markers (mitotic activity index, phosphohistone H3, Ki67), steroid receptors, HER2, high molecular weight cytokeratins and classical prognostic factors in T<sub>1</sub>N<sub>0</sub>M<sub>0</sub> breast cancer. Pol J Pathol 64: 1–8.

Harris L, Fritsche H, Mennel R, Norton L, Radvin P, Taube S, Somerfield MR, Anderson E, Pol J Pathol 19: 5287–5312.

Hwang K-T, Woo JW, Shin HC, Kim HS, Ahn SK, Moon H-G, Han W, Park IA, Noh D-Y (2012) Prognostic influence of BCL2 expression in breast cancer. Int J Cancer 131: E1109–E1119.

Jalava P, Kuopio T, Juntti-Pattinen L, Kotkasanto T, Kronqvist P, Collan Y (2006) Ki67 immunohistochemistry: a valuable marker in prognostication but with a risk of misclassification: proliferation subgroups formed based on Ki67 immunoreactivity and standardized mitotic index. Histopathology 48: 674–682.
Discrepant mitotic index and Ki67 in breast cancer: prognosis

Supplementary Information accompanies this paper on British Journal of Cancer website (http://www.nature.com/bjc)