Ameliorative Effect of the Combination of the *Cichorium intybus* Whole Extract and Melatonin on Doxorubicin-induced Hepatotoxicity in Mice

Parisa Kiani Amin 1, Siamak Mashhady Rafie 2, 3, *Corresponding author*, Saeed Hesaraki 2 and Kumarss Amini 3

1Department of Clinical Science, Faculty of Specialized Veterinary Science, Science and Research Branch, Islamic Azad University, Tehran, Iran
2Department of Pathobiology, Faculty of Specialized Veterinary Science, Science and Research Branch, Islamic Azad University, Tehran, Iran
3Department of Microbiology, School of Basic Sciences, Saveh Branch, Islamic Azad University, Saveh, Iran

Received 2021 October 03; Revised 2022 March 07; Accepted 2022 March 13.

Abstract

**Background:** Doxorubicin is anticancer that is a choice for the treatment of many malignancies. The nature of its toxic effects on the liver and other organs is the harmful character that leads to use with caution. Then, it is necessary to supplement an antioxidant with doxorubicin to reduce its side effects.

**Objectives:** *Cichorium intybus* (*C. intybus*) is a plant with hepatoprotective effects. Melatonin is an antioxidant similar to vitamins. We investigated the repairing effects of *C. intybus*-melatonin together against doxorubicin-induced hepatotoxicity.

**Methods:** Thirty balb/c mice in the weight range of 20 g to 25 g were divided into 5 equal groups of 6 animals each. The groups were as Control: normal saline; DOX: doxorubicin; Chicory: chicory whole plant extract + DOX; Melatonin; melatonin + DOX; Both Chicory-Melatonin + DOX. We assessed histopathology to define necrosis, vacuolar degeneration, and inflammation. In addition, we used immunohistochemistry to evaluate the TNF-α proving the rate of inflammation.

**Results:** The mean sum of histological grading in the control group was 0.00 in contrast to severe damage of the hepatic parenchyma grading 11.34 in sum. The mean sum grade of the other groups including Chicory, Melatonin, and Both Chicory-Melatonin were 8.17, 4.18, and 2.49, respectively. We found that the increased liver damage and TNF-α expression induced by DOX could be improved by applying therapeutic care with the coadministration of the *C. intybus* extract and Melatonin.

**Conclusions:** Chicory and Melatonin have a healing ability against doxorubicin-induced hepatic lesions.

Keywords: *Cichorium intybus*, Melatonin, Doxorubicin, Hepatotoxicity, TNF-α

1. Background

The anticancer drug doxorubicin (DOX) is mentioned to exhibit a choice treating effect on many malignancies, especially hematological ones, but the liver, blood vessels, kidneys, and other organs could be harmfully hurt by the administration of this drug (1). The most vulnerable organ to DOX damage is the liver due to its main metabolic activity. DOX is proved to induce oxygen radicals such as superoxide anions, hydrogen peroxide, and hydroxyl radicals. DOX can inactivate superoxide dismutase and reduce glutathione peroxidase. Then, the high rate of radicals of the oxygen species (ROS) production is compared to antioxidants, the high rate of tissue destruction. It is proved that the ROS derived from DOX and its metabolites in the liver can start extrinsic and intrinsic apoptotic pathways with receptor and caspase three activations respectively. Then, DOX can induce apoptosis in addition to necrosis of hepatocytes (2). The fact of its toxic effects on the liver and other organs is the harmful character that leads to use with caution. Then, it is necessary to supplement an antioxidant with doxorubicin to reduce its side effects.

2. Objectives

The melatonin from the pineal gland is described as a factor that changes and regulates the sleep-wake cycle, circulatory system, neuroendocrine, and immunity against pathogens (3). Melatonin can behave as an anti-inflammatory and antioxidant agent. Melatonin has a direct neutralizing effect on ROS. It can induce the synthesis of the other antioxidants and help to store them. Melatonin has a potent ability to react and inactivate about 10
radical species. In contrast, other antioxidants can inactivate only one radical molecule (4). The extracts of many plants have been demonstrated as a source for preventing hepatic lesions. Cichorium intybus (C. intybus), a plant with hydroxycinnamic acids and flavonoids as its dominating compounds, helps heal the hepatic injury. Administration of C. intybus can increase the number of beneficial flora in the intestine (5, 6). Administration of C. intybus root extract also improves blood flow through circulation because of its inhibitory effect on the angiotensin-convertase enzyme. It is known as a treatment for muscle aches and influenza (7, 8). Many studies have shown that C. intybus could improve the hepatic lesions induced by pathogens or drugs realizing ROS (8, 9). Administration of the C. intybus can reduce the territory effect of the ischemic reperfusion injury of myocardia. This is due to the induction of chemokine receptor type 4 and antioxidant ability (10). A study about lead acetate testis toxicity in male rats proved that C. intybus leaf extract can prevent testicular lesions (11). Tumor necrosis factor α (TNFα) is a cytokine with the ability to induce death signals in addition to other inflammatory functions. In the liver, TNF-α is important for regeneration, proliferation, and fibrosis, but can induce hepatitis in conditions such as viral hepatitis or fatty liver diseases. TNFα is a cytokine with the ability to induce death signals in addition to inflammatory functions. TNFα is significant for hepatic regeneration, proliferation, and fibrosis. It can start hepatitis in the condition such as viral hepatitis or fatty liver diseases. TNFα has two different roles in the liver; one is as a factor of cell death and the other is a hepatocyte and fibroblast proliferator (12). 20-hydroxyicosatetraenoic acid is the substance of a pro-inflammatory metabolite of the arachidonic acid. According to a study, acute DOX toxicity could increase the synthesis of the 20-hydroxyicosatetraenoic acid in the liver and kidney. DOX alters the function of P450 and Epoxide hydrolase enzymes. Then, DOX would increase inflammation (13). Some reports defined the relationship between TNFα and vast lesions of the liver or hepatotoxicity. In microbial septicemia, principally when LPS or endotoxins are released, this cytokine is excreted by Kupffer cells. TNFα is the reason for hepatocyte injury (14, 15). Almost all anticancer drugs such as cisplatin and DOX can develop hepatotoxicity. They, then, elevate related enzymes including ALT, AST, GGT, ALP, and total bilirubin. Some investigators proved that antioxidants and plant extracts could be a benefit for reducing hepatic lesions (16, 17). The histopathological examination has a powerful ability to determine the range of the toxic lesions in all organs (18).

3. Methods

3.1. Plant Materials

C. intybus (whole Plant) with intact appearance picked up in May 2021. A botanist from Biological Sciences Department identified the plants. The plants were washed and put at a 30-degree temperature to be dried. After drying, the whole plant was pulverized into a powder shape by a laboratory mill and put at -20°C until the accumulation (8).

3.2. Preparation of Extract

3.2.1. Preparation of Aqueous-alcoholic Extract

The whole plant was powdered and taken equally and extracted by maceration procedure with alcohol (95% Ethanol) and water for 7 days in an occasional shaking and stirring flask. We filtered the extract, concentrated it, and then dried it by evaporation. The percentage of alcoholic extract of the whole plant was 18% w/w. We stored the extract at -4°C (7).

3.2.2. Preparation of Dried Extracts and Standard

We macerated 100 g of the whole plant dry C. intybus in 1 liter of methanol for 24 hours, filtrated, and the prim extract was collected. In the second stage, under reflux in a water bath, we poured 1 liter of methanol on the sample residue, boiled it for 2 hours, and filtered it. The filtrate was added to the previous crude extract. Next, we added 1 liter of distilled water to the residue. We put it leftover nightly in the room and then filtered it. The filtrate was added to the previous crude extract. Another 1 liter of distilled water was added to the residue, boiled for 2 hours, and filtered. The hot water filtrate was added to the previously collected extract. This manner belongs to the hydroalcoholic extract. There we dried the hydroalcoholic extract at 60°C and froze it until usage (19).

3.3. Animal Treatments

The present study was performed on 30 balb/c mice in the weight range of 20 to 25 g. Before starting the study, to relieve stress and ensure the health of the mice, all of them were kept in cages for 14 days and fed the same diet during storage. After clinical health assessment, the studied mice were randomly divided into 5 experimental groups of 6 animals each. The groups included Control (saline group); mice were orally administered with normal saline (1 mL/kg) for 8 weeks; DOX; DOX (Cell Pharma, 50 mg per 25 mL), was injected at a dose of 15 mg/kg intraperitoneal (20). Chicory; mice were daily administered with chicory whole plant extract, 500 mg/kg (21), for 10 days followed by one dose DOX injection at 15 mg/kg intraperitoneal. Melatonin; mice were daily administered with melatonin, 10
mg/kg (22), for 10 days followed by one dose DOX injection at 15 mg/kg intraperitoneal. Both Chicory-Melatonin; mice were daily administered with both 500 mg/kg chicory and 10 mg/kg melatonin, for 10 days followed by one dose of DOX injection at 15 mg/kg intraperitoneal. Our curing and treating behavior of animals was according to Legislation for the protection of animals used for scientific purposes.

3.4. Biochemical Parameters

On day 20, we took blood samples from the cervical vein, weighted mice accurately, and then euthanized them ethically. Serum Alanine aminotransferase (ALT), Aspartate aminotransferase (AST), Gamma-glutamyl transferase (GGT), Alkaline phosphatase (ALP), and total bilirubin were measured by using an automatic biochemical analyzer (Accent 200, China). These measures are indicators of liver (hepatotoxicity) lesions (14).

3.5. Histopathological Sample Preparation and Evaluation

After each necropsy, we put the livers in 10% formalin buffer for histopathological and immunohistochemical evaluation. A histopathological score of hepatic tissues was performed as 0 = absent; 1 = low or weak; 2 = mild; 3 = moderate; and 4 = high or frequent, and the total score was the basis of judgment (23).

3.6. Immunohistochemical Sample Preparation

The anti-TNFα antibodies were used in this study to prove the degree of the inflammation. Avidin-biotin peroxidase immunohistochemical method (ABC) was used to detect the TNFα positive areas in the livers of all experimental groups. We deparaffinized the 5-µm sections exposed them to TNFα monoclonal antibody (dilution 1:100; DAKO), and then mounted them. Positive and negative controls evaluated the staining procedure. The pathologist described the lesions in a high-power field for all slides and, then, captured them by digital camera. Five high power fields were observed at random.

3.7. Statistical Analyses

First, the data obtained from the study were presented as the mean ± SD and, then, analyzed by SPSS 22, using the one-way analysis of variance (ANOVA) followed by the Bonferroni test for parametric data and the Kruskal–Wallis one-way analysis of variance test for non-parametric data. P-values less than 0.05 were considered statistically significant.

4. Results

4.1. Toxicity

All mice that received C. intybus extract or melatonin lived with normal weight and body condition during the study. In contrast to the DOX-treated mice, this drug initiated many side effects such as tangled body hair, poor body condition, loss of activity, and weakness.

4.2. Biochemical Parameters

The automatic cell counter analyzed the ALT, AST, GGT, and ALP activities in addition to total bilirubin, higher despite the non-significantly in the DOX group than in the other groups. There was no significant (P < 0.05) relationship between treatments (Table 1).

4.3. Histopathological Structure of the Liver

The liver section in mice of the control group had normal lobules composed of straight columns of hepatocytes that extend from the central vein to the periphery of the portal areas (Figure 1A). The liver of the DOX group showed severe lesions. These include micro and macrovesicular vacuolar degeneration, necrosis, and inflammation of the parenchyma. The necrosis was composed of nuclear pyknosis and more eosinophilia or loss of cytoplasm. All study groups were significantly better than the DOX group. It was observed in the treatment groups that melatonin compared to chicory improved the livers. The Co-administration of melatonin and chicory, the most ameliorating livers tissues (Figure 1A, B, C, D, and E; Table 2).

4.4. Immunohistochemistry Analysis of TNFα in the Liver

Vast TNFα positive areas of the liver in the DOX group revealed the important role of DOX anticancer in developing inflammation and hepatotoxicity. Otherwise, all groups that received melatonin were more potable of reducing lesions than that in the chicory-treated group. The Co-administration of melatonin and chicory significantly showed the best reduction in the TNFα positive areas and inflammation (Figure 2A, B, C, D, and E; Table 3).

5. Discussion

DOX is a potent inducer for the production of many free radicals in the hepatic tissues. The liver cannot eliminate it by the body-stored antioxidants (24). Hepatocytes are destroyed when DOX is metabolized by hepatic enzymes rich in mitochondria (25). The phenomena that indicated hepatic injury due to DOX induction increase ROS, reduce mitochondrial oxidation cycles, chromatin malformation and degradation, and Necrosis or Apoptosis (26).
Table 1. Effect of a DOX (15 mg/kg IP) Alone or in Combination with Drugs on Serum Biochemical Parameters in Mice

| Groups                      | AST (unit/L) | ALT (unit/L) | GGT (unit/L) | ALP (unit/L) | Total Bilirubin (mg/dL) |
|-----------------------------|--------------|--------------|--------------|--------------|-------------------------|
| Control (Saline)            |              |              |              |              |                         |
| Mean ± SD                   | 215.75 ± 100.20<sup>a</sup> | 80.50 ± 14.27<sup>a</sup> | 0.25 ± 0.50<sup>a</sup> | 225.75 ± 141.54<sup>a</sup> | 19.25 ± 8.53<sup>a</sup> |
| %CV                         | 46.44        | 17.72        | 2.0          | 62.69        | 44.31                   |
| DOX                         |              |              |              |              |                         |
| Mean ± SD                   | 221.00 ± 65.84<sup>b</sup> | 102.00 ± 76.63<sup>b</sup> | 1.17 ± 1.94<sup>b</sup> | 307.67 ± 33.90<sup>b</sup> | 15.17 ± 5.15<sup>b</sup> |
| %CV                         | 29.79        | 75.12        | 1.65         | 11.01        | 33.94                   |
| Chicory                     |              |              |              |              |                         |
| Mean ± SD                   | 150.80 ± 24.50<sup>a</sup> | 62.40 ± 6.46<sup>a</sup> | 0.40 ± 0.54<sup>a</sup> | 214.2 ± 117.69<sup>a</sup> | 32.80 ± 23.59<sup>a</sup> |
| %CV                         | 16.24        | 10.35        | 1.35         | 54.94        | 71.92                   |
| Melatonin                   |              |              |              |              |                         |
| Mean ± SD                   | 49.67 ± 17.94<sup>a</sup> | 129.33 ± 18.03<sup>a</sup> | 0.50 ± 0.54<sup>a</sup> | 184.83 ± 95.64<sup>a</sup> | 16.0 ± 3.16<sup>a</sup> |
| %CV                         | 36.11        | 13.94        | 1.08         | 51.74        | 19.75                   |
| Both (DOX + Chicory / Melatonin) |              |              |              |              |                         |
| Mean ± SD                   | 67.60 ± 11.86<sup>b</sup> | 125.67 ± 10.78<sup>a</sup> | 0.60 ± 0.54<sup>a</sup> | 254.6 ± 61.35<sup>a</sup> | 11.5 ± 1.73<sup>a</sup> |
| %CV                         | 8.57         | 8.58         | 0.9          | 24.09        | 15.04                   |

<sup>a</sup> Analysis was done by Kruskal-Wallis for AST, GGT, and total bilirubin and one-way analysis of variance test for ALT and ALP. <sup>A</sup>: significant difference from DOX. <sup>B</sup>: significant difference from A.

Figure 1. Histopathological evaluation of the livers. A: Control Group not exposed to doxorubicin. B: DOX group we treated with doxorubicin. C: Chicory. D: Melatonin, and E: Chicory + Melatonin compared to DOX group (P < 0.05); (HE × 200). Freeform: Vacuolar degeneration; Yellow arrow: necrotic hepatocyte; Chevron: Inflammation; P: Portal vein; V: Central vein.
Table 2. The Histopathological Score of Hepatic Tissues from Mice Received High Doses of DOX (15 mg/kg) \(^a\)

| Groups                     | Necrosis     | Leucocyte Infiltration | Vacular Degeneration | Congested Blood Vessels | Total Score |
|----------------------------|--------------|------------------------|----------------------|-------------------------|-------------|
| Control (Saline)           | 0.00 ± 0.00 \(^A\) | 0.00 ± 0.00 \(^A\) | 0.00 ± 0.00 \(^A\) | 0.00 ± 0.00 \(^A\) | 0.00 \(^A\) |
| DOX                        | 3.67 ± 0.51 \(^B\) | 1.83 ± 0.75 \(^B\) | 2.50 ± 1.22 \(^B\) | 2.17 ± 0.75 \(^B\) | 11.34 \(^B\) |
| Chicory                    | 2.17 ± 0.75 \(^C\) | 1.33 ± 0.5 \(^B\) | 1.33 ± 0.51 \(^C\) | 1.50 ± 0.54 \(^B\) | 8.17 \(^C\) |
| Melatonin                  | 1.67 ± 0.51 \(^C\) | 0.67 ± 0.51 \(^C\) | 1.00 ± 0.00 \(^C\) | 0.67 ± 0.51 \(^A\) | 4.18 \(^D\) |
| Both (DOX + Chicory / Melatonin) | 0.83 ± 0.75 \(^A\) | 0.33 ± 0.51 \(^D\) | 1.00 ± 0.00 \(^C\) | 0.33 ± 0.51 \(^A\) | 2.49 \(^D\) |

\(^a\) They were treated with Chicory extract and Melatonin. Scoring was performed as 0 = absent; 1 = low or weak; 2 = mild; 3 = moderate; and 4 = high or frequent, and also the total score. Analysis was done by Kruskal-Wallis one-way analysis of variance test. A: significant difference from DOX. B: significant difference from A.

Figure 2. Immunohistochemical evaluation of the livers. A: Control; B: DOX; C: Chicory; D: Melatonin, and E: Chicory + Melatonin groups (P < 0.05); (antibody against TNF-\(\alpha\) \(\times\) 200).

Table 3. Immunohistochemical Evaluation of TNF-\(\alpha\) in the Hepatic Tissues Was Based on the Percentage of the Stained Area in All Mice (Groups) Described in the Histopathology \(^a\)

| Factor    | Control (Saline) | DOX     | Chicory  | Melatonin | Both (DOX + Chicory / Melatonin) |
|-----------|------------------|---------|----------|-----------|----------------------------------|
| TNF-\(\alpha\) % | 12.26 ± 1.35 \(^A\) | 17.54 ± 0.70 \(^B\) | 17.27 ± 0.95 \(^B\) | 14.72 ± 0.66 \(^C\) | 13.59 ± 0.98 \(^A\), \(^C\) |

\(^a\) Data are mean ± SD, and analysis is carried out using one-way ANOVA. A: significant difference from DOX. B: significant difference from A. C: significant difference from A and B.

Int J Cancer Manag. 2022; 15(3):e119807.
We studied the effects of Chicory extract, Melatonin, and both of them as anti-inflammatory factors. We also assessed their protecting effects against mice liver injury induced by DOX.

Melatonin can be a powerful antioxidant and antiradical agent that limits the effects of free radicals such as hydroxyl and persulphate radicals in many organs, especially in the liver (3, 27). Melatonin is also involved in reducing the ischemia-reperfusion injury. Some researchers have revealed that the endothelial cells of small vessels may be destroyed. Following this, the repair of the hepatic lesions will not heal properly in the DOX-treated groups (28). The growth of cancers has been prevented by melatonin supplementation therapeutically, in addition to its protective ability to decrease the side effects by melatonin supplementation (29). Many studies have shown that C. intybus could improve the hepatic lesions induced by pathogens or drugs realizing ROS (8, 9). The leaf of C. intybus extract manifested a significant protecting effect against hepatotoxicity caused by CCl4. It also reduces the serum levels of the AST, ALT, ALP, and the total bilirubin in the CCl4 treated mice (30). On the other hand, melatonin in combination with DOX can lead to decrease high serum ALT and AST levels (31). According to this experiment, co-supplementation of chicory extract (500 mg/kg) and melatonin (10 mg/kg) could reduce ALT, AST, GGT, ALP, and total bilirubin more potential than each on its own.

We suppose that the melatonin administration may promote hepatic tissues more resistance to lipid oxidation. Pre-treatment with melatonin at 10 mg/kg followed by improving effects in the liver exposed to DOX could be demonstrated by decreasing the vascular degeneration. We found this microscopic feature in the chick group and then proposed that the C. intybus extract at 500 mg/kg may improve the severity of lipid oxidation in the liver. Melatonin and Chicory cooperate as an antioxidant, anticancer, and detoxifiers. They can inhibit ROS production and prevent cell injury by the reaction to radicals (32). According to our results, co-administration of chicory extract (500 mg/kg) and melatonin (10 mg/kg) cure hepatic injury significantly. Histological signs of DOX administration are sinusoidal congestion, hepatocyte vacuolar degeneration, and leukocytes infiltration (24). Following the pathologic effects induced by DOX, there is an increase in the expression of some growth factors that lead to the hyperplasia of bile ducts with infiltration of the inflammatory cells (33). In this experiment, the hepatic lesions in the DOX group were moderate lobular necrosis, severe vacuolar degeneration, interface and lobular inflammatory cells infiltration, and sinusoidal engorgement. We demonstrate minimal pathological findings in all treating groups compared to what was seen previously in the Dox group alone. There are differences between all groups except that the melatonin and both chicory and melatonin group did not have any significant difference in the total score (Table 2).

Cytokines are small signaling proteins that modulate leukocytes’ activity in immunity and aid in the movement of immune cells to regulate cell activity, differentiation, proliferation, migration, or inflammation against pathogens (34). The first powerful cytokine that stimulates other cytokines, such as IL-1β synthesis is TNF-α, which can exaggerate the inflammatory cascade (35). The most way that DOX creates lesions in the organs such as the liver is to pull out the synthesis of pro-inflammatory cytokines followed by necrosis and degeneration.

Melatonin can reduce the serum levels of TNF-α and IL-6 in the mice. It can increase the levels of IL-10 (36). According to some studies, melatonin can improve cholestatic liver lesions induced by α-naphthyl isothiocyanate and its oxidative stress. Generally, melatonin is the benefit for improving liver disease (37-39). NOD-like receptor protein 3 (NLRP3: Inflammasome) can activate the inflammatory responses in complicated conditions such as microbial influx or necrosis due to toxins. The inflammasome also stimulates the synthesis of the IL-1β and caspase-1 in some situations such as cancer and diabetes. Melatonin diminishes inflammation by inhibiting NLRP3 synthesis. It controls some regulatory molecules, such as microRNA and Wnt/β-catenin (40). According to our results, the group receiving DOX caused a significant increase in hepatic parenchyma TNF-α levels correlated to increased leukocytes and inflammatory processes (41).

Footnotes

**Authors’ Contribution:** Conception and design of the study: S. M. R; S. H; K. A.; Acquisition of data: P. A. K; K. A.; Analysis and interpretation of data: P. A. K; S. M. R; S. H; Drafting of the manuscript: S. H; P. A. K; Critical revision of the manuscript for important intellectual content: S. M. R; Statistical analysis: S. H.

**Conflict of Interests:** Authors mention that there is no conflict of interest in this study.

**Data Reproducibility:** The data presented in this study are openly available in one of the repositories or will be available on request from the corresponding author by this journal representative at any time during submission or after publication. Otherwise, all consequences of possible withdrawal or future retraction will be with the corresponding author.

**Ethical Approval:** It was not declared by the authors.

**Funding/Support:** This study was supported by a teaching and research scholarship from the Faculty of Special...
ized Veterinary Science, Science and Research Branch, Islamic Azad University, Tehran, Iran.

References

1. Hou L, Liu S, Zhao C, Fan L, Hu H, Yin S. The combination of T-2 toxin and acrylamide synergistically induces hepatotoxicity and nephrotoxicity via the activation of oxidative stress and the mitochondrial pathway. Toxicon. 2021;189:65-72. doi: 10.1016/j.toxicon.2020.11.007. [PubMed: 33272324].

2. Liu X, Qiu Y, Liu Y, Huang N, Hua C, Wang Q, et al. Citronellal ameliorates doxorubicin-induced hepatotoxicity via antioxidative stress, antiapoptosis, and proangiogenesis in rats. J Biochem Mol Toxicol. 2020;34(2). doi: 10.1002/jbt.22839.

3. Reiter R, Tang L, Garcia J, Munoz-Hoyos A. Pharmacological actions of melatonin in oxygen radical pathophysiology. Life Sci. 1997;60(25):2255-71. doi: 10.1016/s0024-3205(97)00310-1.

4. Carroll MC, Reilly M. Melatonin: Immune Stimulant, Anti-inflammatory, and Antioxidant. Intern Med. 2021;131.

5. Shin S, Park S, Lee H, Hur J. Effects of fermented chicory fiber on the improvement of intestinal function and constipation. J Korean Soc Food Sci Nutr. 2014;43(4):355-9.

6. Liu H, Ivarsson E, Dicksved J, Lundh T, Lindberg JE. Inclusion of Cichorium intybus (Linn.) root extract against carbon tetrachloride induced hepatotoxicity via antioxidative stress, and acrylamide synergistically induces hepatotoxicity and nephrotoxicity in rats. Toxicon. 2021;189:65-72. doi: 10.1016/j.toxicon.2020.11.007. [PubMed: 33272324].

7. Liu X, Qiu Y, Liu Y, Huang N, Hua C, Wang Q, et al. Citronellal ameliorates doxorubicin-induced hepatotoxicity via antioxidative stress, antiapoptosis, and proangiogenesis in rats. J Biochem Mol Toxicol. 2020;34(2). doi: 10.1002/jbt.22839.

8. Dalar A, Konczak I. Cichorium intybus from Eastern Anatolia: Phenolic composition, antioxidant and enzyme inhibitory activities. Ind Crops Prod. 2014;60:79-85. doi: 10.1016/j.indcrops.2014.05.043.

9. Nallamilli BR, Phani Kumar CS, Veer Reddy K, Lakshmi Prasanna M, Sandeep B. The effects of the Cichorium intybus Root Extract against Alcohol-Induced Hepatotoxicity in Experimental Rats. Drug Metab Dispos. 2021;50(4):692–7. doi: 10.1124/dmd.111.039123. [PubMed: 32862634]. [PubMed Central: PMC7440251].

10. Barakat BM, Ahmed HI, Bahr HI, Elbahaie AM. Protective Effect of Boswellic Acids against Doxorubicin-Induced Hepatotoxicity: Impact on Nrf2/HO-1 Defense Pathway. Oxid Med Cell Longev. 2018;2018:8296451. doi: 10.1155/2018/8296451. [PubMed: 29541348]. [PubMed Central: PMC5818967].

11. Madhu LN, Kodali M, Attaluri S, Shuai B, Melissari L, Rao X, et al. Melatonin improves brain function in a model of chronic Gulf War Illness with modulation of oxidative stress, NLRP3 inflammasomes, and BDNF-ERK-CREB pathway in the hippocampus. Redox Biol. 2021;43:101973. doi: 10.1016/j.redox.2021.101973. [PubMed: 39313884]. [PubMed Central: PMC805571].

12. Cengiz O, Baran M, Balcioglu E, Suna PA, Bilgici P, Goktepe O, et al. Use of selenium to ameliorate doxorubicin induced hepatotoxicity by targeting pro-inflammatory cytokines. Biotechnol Histochem. 2021;96(6):677-75. doi: 10.1080/10520295.2020.1760353. [PubMed: 32400214].

13. Tacar O, Siamornsak P, Dass CR. Doxorubicin: an update on anticancer molecular action, toxicity and novel drug delivery systems. J Pharm Pharaco. 2021;135:1-10. doi: 10.1128/jppj.2019.103496. [PubMed: 2042-7588]. [PubMed Central: PMC6839841].

14. Schwann J, Wolf D, Pahl A, Brune K, Papadopoulos T, van Rooijen N, et al. Improvement of Kupffer Cells for T-Cell-Dependent Liver Injury in Mice. Am J Pathol. 2000;157(5):1671-83. doi: 10.1016/s0002-9440(10)64804-3.

15. Abdel-Wahhab KG, Mannaa FA, Ashry M, Khaled DM, Hassan LK, Gomaa HF. Chenopodium quinoa ethanol extract ameliorates cyclophosphamide-induced hepatotoxicity in male rats. Comp Clin Path. 2021;30(3):267-76. doi: 10.1002/ccp.20399-2.

16. Akcam M, Artan R, Yilmaz A, Ozdem S, Gelen T, Naziroglu M. Caffeic acid phenethyl ester modulates aflatoxin B1-induced hepatotoxicity in rats. Cell Biochem Funct. 2013;31(6):707-12. doi: 10.1002/cbf.2957. [PubMed: 23400894].

17. Eidi A, Mortazavi P, Bazargan M, Zaringhalam J. Hepatoprotective activity of cinnamon ethanolic extract against CCl4-induced liver injury in rats. Evid Cln Investig. 2012;14:1-95.

18. Sakeran MI, Zidan N, Rehman H, Aziz AT, Saggou S. Abrogation by Trifolium alexandrinum root extract on hepatotoxicity induced by acetaminophen in rats. Redox Rep. 2014;19(1):26-33. doi: 10.1797/j.00032051.2014.004366. [PubMed: 24199932]. [PubMed Central: PMC6817585].

19. Sun X, Song Y, Xie Y, Han J, Chen F, Sun Y, et al. Shenlijia Attenuates Doxorubicin-Induced Chronic Heart Failure by Inhibiting Cardiac Fibrosis. Evid Based Complement Alternat Med. 2021;2021:6659676. doi: 10.1155/2021/6659676. [PubMed: 34326887]. [PubMed Central: PMC8310442].

20. Janda K, Gutowska I, Geszke-Moritz M, Jakubczyk K. The Common Cichory (Cichorium intybus L.) as a Source of Extracts with Health-Promoting Properties-A Review. Molecules. 2021:26(6). doi: 10.3390/molecules26061814. [PubMed: 33807029]. [PubMed Central: PMC8005178].

21. Madhu LN, Kodali M, Attaluri S, Shuai B, Melissari L, Rao X, et al. Melatonin improves brain function in a model of chronic Gulf War Illness with modulation of oxidative stress, NLRP3 inflammasomes, and BDNF-ERK-CREB pathway in the hippocampus. Redox Biol. 2021;43:101973. doi: 10.1016/j.redox.2021.101973. [PubMed: 39313884]. [PubMed Central: PMC805571].

22. Cengiz O, Baran M, Balcioglu E, Suna PA, Bilgici P, Goktepe O, et al. Use of selenium to ameliorate doxorubicin induced hepatotoxicity by targeting pro-inflammatory cytokines. Biotechnol Histochem. 2021;96(6):677-75. doi: 10.1080/10520295.2020.1760353. [PubMed: 32400214].

23. Tacar O, Siamornsak P, Dass CR. Doxorubicin: an update on anticancer molecular action, toxicity and novel drug delivery systems. J Pharm Pharmacol. 2021;63(2):157-70. doi: 10.1111/jppj.2042-7588.2012.01567.x. [PubMed: 23776883].

24. Pereira GC, Pereira SP, Luminii JA, Magalhaes J, Assencio A, et al. Mitochondrionopathy phenotype in doxorubicin-treated Wistar rats depends on treatment protocol and is cardiac-specific. PLoS One. 2012;7(6). e38877. doi: 10.1371/journal.pone.0038877. [PubMed: 22745682]. [PubMed Central: PMC382146].

25. Poeggeler B, Reiter RJ, Tan DX, Chen LD, Manchester LC. Melatonin, hydroxyl radical-mediated oxidative damage, and aging: a hypothesis. J Pineal Res. 1993;14(4):151-68. doi: 10.1111/j.1600-079x.1993.tb00498.x. [PubMed: 8102180].

26. Bertuglia S, Marchiafava PL, Colantuoni A. Melatonin prevents ischemia reperfusion injury in hamster cheek pouch microcirculation. Cardiovasc Res. 1996;33(6):947-52.

27. Neri B, De Leonardi V, Gemelli MT, Di Loro F, Mottola A, Ponchietti R, et al. Melatonin as biological response modifier in cancer patients. Anticancer Res. 1998;18(2):3129-32.
30. Heibatollah S, Reza NM, Izadpanah G, Sohailia S. Hepatoprotective effect of Cichorium intybus on CCl4-induced liver damage in rats. Afr J Biochem Res. 2008;2(6):141-4.

31. Mathes AM. Hepatoprotective actions of melatonin: possible mediation by melatonin receptors. World J Gastroenterol. 2010;16(48):6087-97. doi: 10.3748/wjg.v16.i48.6087. [PubMed: 21822213]. [PubMed Central: PMC3025385].

32. Kim S, Cho M. Melatonin and Polyphenol Contents in Some Edible Sprouts (Alfalfa, Chicory, Rape, Red Kale and Sunflower). Prev Nutr Food Sci. 2011;16(2):184-8. doi: 10.3746/jfn.2011.2.184.

33. Mete R, Oran M, Topcu B, Oznur M, Seber ES, Gedikbasi A, et al. Protective effects of onion (Allium cepa) extract against doxorubicin-induced hepatotoxicity in rats. Toxicol Ind Health. 2016;32(3):551-7. doi: 10.1777/0748233715040407. [PubMed: 24913056].

34. Caglayan C, Temel Y, Kandemir FM, Yildirim S, Kucukler S. Naringin protects against cyclophosphamide-induced hepatotoxicity and nephrotoxicity through modulation of oxidative stress, inflammation, apoptosis, autophagy, and DNA damage. Environ Sci Pollut Res Int. 2018;25(21):20968-84. doi: 10.1007/s11356-018-2242-5. [PubMed: 29766429].

35. Lee HS, Jung KH, Hong SW, Park IS, Lee C, Han HK, et al. Morin protects acute liver damage by carbon tetrachloride (CCl4) in rat. Arch Pharm Res. 2008;31(9):1160-5. doi: 10.1007/sl2272-008-0283-5. [PubMed: 18806959].

36. Tavares BS, Tsosura TYS, Mattera M, Santelli JO, Belardi BE, Chiba FY, et al. Effects of melatonin on insulin signaling and inflammatory pathways of rats with apical periodontitis. Int Endod J. 2021;54(6):926-40. doi: 10.1111/iej.13474. [PubMed: 3341973].

37. Zhang JI, Meng X, Li Y, Zhou Y, Xu DP, Li S, et al. Effects of Melatonin on Liver Injuries and Diseases. Int J Mol Sci. 2017;18(4). doi: 10.3390/ijms18040673. [PubMed: 28333073]. [PubMed Central: PMC5412264].

38. Chen Z, Ma X, Zhu Y, Zhao Y, Wang J, Li R, et al. Paeoniflorin ameliorates ANIT-induced cholestasis by activating Nrf2 through an PI3K/Akt-dependent pathway in rats. Phytother Res. 2015;29(1):768-75. doi: 10.1002/ptr.5431. [PubMed: 26289092].

39. Yu H, Li Y, Xu Z, Wang D, Shi S, Deng H, et al. Identification of potential biomarkers in cholestasis and the therapeutic effect of melatonin by metabolomics, multivariate data and pathway analyses. Int J Mol Med. 2018;42(5):255-26. doi: 10.3892/ijmm.2018.3859. [PubMed: 30226547]. [PubMed Central: PMC6192756].

40. Ashrafizadeh M, Najafi M, Kayyani N, Mohammadnejad R, Farkhondeh T, Samarghandian S. Anti-Inflammatory Activity of Melatonin: a Focus on the Role of NLRP3 Inflammasome. Inflammation. 2021;44(4):1-16. doi: 10.1007/s10753-021-01428-9. [PubMed: 3365308].

41. Supriya R, Tam BT, Pei XM, Lai CW, Chan LW, Yung BY, et al. Doxorubicin Induces Inflammatory Modulation and Metabolic Dysregulation in Diabetic Skeletal Muscle. Front Physiol. 2016;7:323. doi: 10.3389/fphys.2016.00323. [PubMed: 27512375]. [PubMed Central: PMC4961706].