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Mechanisms of CFTR Functional Variants That Impair Regulated Bicarbonate Permeation and Increase Risk for Pancreatitis but Not for Cystic Fibrosis

Jessica LaRusch1, Jinsei Jung2, Ignacio J. General3, Michele D. Lewis4, Hyun Woo Park2,3,*, Randall E. Brand1, Andres Gelrud1,*, Michelle A. Anderson5, Peter A. Banks6, Darwin Conwell6,*, Christine Lawrene7, Joseph Romagnuolo7, John Baillie8,*, Samer Alkaade9, Gregory Coté10, Timothy B. Gardner11, Stephen T. Amann12, Adam Slivka1, Bimaljit Sandhu13,*, Amy Aloe1, Michelle L. Kienholz1, Dhiraj Yadav1, M. Michael Barmada14, Ivet Bahar3,*, Min Goo Lee2,*, David C. Whitcomb1,14,15,*, and the North American Pancreatitis Study Group1

Abstract

CFTR is a dynamically regulated anion channel. Intracellular WNK1-SPAK activation causes CFTR to change permeability and conductance characteristics from a chloride-preferring to bicarbonate-preferring channel through unknown mechanisms. Two severe CFTR mutations (CFTRsev*) cause complete loss of CFTR function and result in cystic fibrosis (CF), a severe genetic disorder affecting sweat glands, nasal sinuses, lungs, pancreas, liver, intestines, and male reproductive system. We hypothesize that those CFTR mutations that disrupt the WNK1-SPAK activation mechanisms cause a selective, bicarbonate defect in channel function (CFTRBD) affecting organs that utilize CFTR for bicarbonate secretion (e.g., the pancreas, nasal sinuses, vas deferens) but do not cause typical CF. To understand the structural and functional requirements of the CFTR bicarbonate-preferring channel, we (a) screened 984 well-phenotyped pancreatitis cases for candidate CFTRBD mutations from among 81 previously described CFTR variants; (b) conducted electrophysiology studies on clones of variants found in pancreatitis but not CF; (c) computationally constructed a new, complete structural model of CFTR for molecular dynamics simulation of wild-type and mutant variants; and (d) tested the newly defined CFTRBD variants for disease in non-pancreas organs utilizing CFTR for bicarbonate secretion. Nine variants (CFTR R74Q, R75Q, R117H, R170H, L967S, L997F, D1152H, S1235R, and D1270N) not associated with typical CF were associated with pancreatitis (OR 1.5, p = 0.002). Clones expressed in HEK 293T cells had normal chloride but not bicarbonate permeability and conductance characteristics from a chloride-preferring to bicarbonate-preferring channel through unknown mechanisms. From among 81 previously described CFTR variants, only two severe CFTR mutations (CFTR R74Q, R75Q) cause a complete loss of CFTR function and result in cystic fibrosis (CF). We hypothesize that those CFTR mutations that disrupt the WNK1-SPAK activation mechanisms cause a selective, bicarbonate defect in channel function (CFTRBD) affecting organs that utilize CFTR for bicarbonate secretion (e.g., the pancreas, nasal sinuses, vas deferens) but do not cause typical CF. To understand the structural and functional requirements of the CFTR bicarbonate-preferring channel, we (a) screened 984 well-phenotyped pancreatitis cases for candidate CFTRBD mutations from among 81 previously described CFTR variants; (b) conducted electrophysiology studies on clones of variants found in pancreatitis but not CF; (c) computationally constructed a new, complete structural model of CFTR for molecular dynamics simulation of wild-type and mutant variants; and (d) tested the newly defined CFTRBD variants for disease in non-pancreas organs utilizing CFTR for bicarbonate secretion. Nine variants (CFTR R74Q, R75Q, R117H, R170H, L967S, L997F, D1152H, S1235R, and D1270N) not associated with typical CF were associated with pancreatitis (OR 1.5, p = 0.002). Clones expressed in HEK 293T cells had normal chloride but not bicarbonate permeability and conductance with WNK1-SPAK activation. Molecular dynamics simulations suggest physical restriction of the CFTR channel and altered dynamic channel regulation. Comparing pancreatitis patients and controls, CFTRBD increased risk for rhinosinusitis (OR 2.3, p = 0.005) and male infertility (OR 395, p < 0.0001). WNK1-SPAK pathway-activated increases in CFTR bicarbonate permeability are altered by CFTRBD variants through multiple mechanisms. CFTRsev variants are associated with clinically significant disorders of the pancreas, sinuses, and male reproductive system.

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Introduction

The cystic fibrosis transmembrane conductance regulator (CFTR, GenBank Accession: AH006034.1) is an ATP-binding cassette (ABC) transporter-type protein localized to the apical plasma membrane of epithelial cells. It differs from other ABC transporters in that it acts as a regulated anion channel rather than a transporter [1]. When the channel is open, anions move across the membrane down their electrochemical potential gradient, resulting in fluid and electrolyte secretion or absorption.

The CFTR molecule has been intensely studied because mutations in the CFTR gene are associated with cystic fibrosis (CF, OMIM #219700), the most common life-threatening genetic disorder among populations of Northern European ancestry [2,3]. However, the clinical features of CF and CFTR-related disorders are variable, and laboratory studies of CFTR regulation, its biophysical properties and molecular mechanisms of dysfunction have been challenging due to the complexity of the regulatory mechanisms and the dynamic flexibility of various structural domains [see recent reviews [4,5]].

Cystic fibrosis is an autosomal recessive syndrome usually caused by inheriting two CFTR mutations that eliminate effective chloride conductance (CFTR<sup>del</sup>) [2,3]. Although nearly 2000 CFTR variants have been described (http://www.genet.sickkids.on.ca), the majority of CF cases are associated with the CFTR 508F-del mutation as a homozygous genotype or in combination with another severe CF-associated mutation (CFTR<sup>C</sup>/CFTR<sup>T</sup>) that together result in minimal CFTR function. Thus, most research has focused on the regulation of chloride conductance, and dynamic modeling of the first of two nucleotide-binding domains (NBD1), which normally contains F508 [4,5]. Based on numerous studies, three conformations have been described for the molecule as an anion channel: a closed state, an open state, and an open-stroke permeability state [5] [4]. However, the relative permeability/conductance ratios of chloride and bicarbonate are variable [6] and may be dynamically regulated [7], suggesting that conformational changes induced by point mutations in the channel or in the permeability pore may alter ion permeation properties of CFTR.

Diagnosis of CF is based on a combination of phenotypic features, family history, functional tests and/or the identification of mutations in the CFTR gene. CFTR variants have been reported at least twice in non-pancreatic tissues, we used the phenotyping criteria for sinusitis and male infertility for the NAPS2 cases and controls (43 of them, listed in Table 1). Second, using this panel of 81 CFTR variants (Table S1 in the Supplementary Material), we genotyped the deeply phenotyped North American Pancreatitis Study 2 (NAPS2) subjects [23] to identify candidate CFTR<sup>BD</sup> variants that were also present in our cases and controls (43 of them, listed in Table 1). Third, to determine if CFTR<sup>BD</sup> variants are associated with altered WNK1-SPAK pathway-stimulated CFTR bicarbonate permeability, we generated plasmids containing the candidate CFTR<sup>BD</sup> variants selected from the NAPS2 study and expressed them in HEK-293T cells for electrophysiological analysis. Fourth, to gain insight into the molecular mechanisms of CFTR<sup>BD</sup> dysfunction, we performed molecular dynamics (MD) simulations based on homology-modeled structures of ABC transporters and examined the structure and dynamics of the channel. Finally, to determine if CFTR<sup>BD</sup> variants are associated with disease in non-pancreatic tissues, we used the phenotyping criteria for sinusitis and male infertility for the NAPS2 cases and controls.

These studies revealed at least 9 CFTR<sup>BD</sup> variants. We found that the WNK1-SPAK pathway that enhances CFTR bicarbonate permeability/conductance compared with chloride conductance in HEK-293T cells is altered by CFTR<sup>BD</sup> variants. The examination of MD trajectories suggests at least two potential mechanisms of channel dysfunction. Phenotype-genotype studies...
Author Summary

Genetic disorders of ion channels can affect the body’s ability to function properly in many ways. CFTR, an ion channel regulating movement of chloride and bicarbonate across cell membranes, is important for absorbing and secreting fluids. If the gene responsible for the CFTR channel is mutated severely, the result is cystic fibrosis, a hereditary disorder in which the patient develops thick mucus, especially in the lungs, as well as scarring (fibrosis) in the pancreas. Cystic fibrosis also affects the sweat glands, nasal sinuses, intestines, liver, and male reproductive system. Mutations to the CFTR gene that do not cause cystic fibrosis have been considered benign. However, we discovered 9 CFTR mutations that do not cause cystic fibrosis but do cause inflammation and scarring of the pancreas (chronic pancreatitis). These mutant CFTR channels secrete chloride, which is important in the sweat glands, lungs, and intestines, but not bicarbonate, which is important in the pancreas, sinuses, and male reproductive tract. We found patients with any of these 9 mutations had chronic pancreatitis, and often sinus infections, and male infertility, but not other symptoms of cystic fibrosis. Our computer models and data will help researchers develop better drugs and help physicians treating patients with chronic pancreatitis.

in humans demonstrated that CFTRBD variants are also associated with disorders of the pancreas, sinuses, and male reproductive systems.

Results

CFTR genotyping in pancreatitis patients and controls

We genotyped 984 well-phenotyped cases of pancreatitis from NAPS2 for 81 CFTR variants, including common CF mutations and variants previously reported in at least two subjects with pancreatitis but not CF. Common tag-SNPs at the CFTR locus were previously excluded in a pancreatitis genome-wide association study (all p values $\geq 0.01$) [24], suggesting that the missing heritability and predicted dysfunction was primarily associated with multiple rare variants. SPINK1 N34S was also genotyped to determine complex risk [18]. Only SPINK1 N34S heterozygotes were used for trans-heterozygote analysis with CFTR, since homozygous SPINK1 N34S is sufficient to cause pancreatitis.

Of 43 CFTR variants identified in the NAPS2 cohort (Table 1), nine were associated with typical CF but not reported in patients with pancreatitis[25–29] of particular interest: R74Q, R75Q, R117H (CFTRBD only when m cis with IVS8-T5[30]; R117H*T5), R170H, L967S, L997F, D1152H, S1235R, and D1270N. These were either independently associated with disease, or their location on the CFTR molecule. D1270N. These were either independently associated with disease, or their location on the CFTR molecule.
Table 1. Analysis of CFTR and SPINK1 variants in cases and controls.

| CFTR variant         | %Cases | %Uctrls | OR       | p-value | %Cases w/N34S | OR w/N34S | p-value w/N34S |
|----------------------|--------|---------|----------|---------|---------------|-----------|----------------|
| CF/BD or BD/BD       | 2.5    | 0.1     | 31.9     | <0.0001 | 5.5           | 7.46      | 0.12           |
| All CF               | 8.7    | 3.3     | 2.76     | <0.0001 | 16.4          | 5.65      | <0.0001        |
| F508delCF            | 6.9    | 3.1     | 2.32     | <0.0001 | 14.5          | 5.13      | <0.0001        |
| IVS8T5+CF            | 9.9    | 8.2     | 1.24     | 0.079   | 10.9          | 1.37      | 0.47           |
| 2789+5G>A CF         | 0.3    | 0.0     |          |         |               |           |                |
| 3849+10kb>C CF       | 0.3    | 0.0     |          |         |               |           |                |
| N1303K CF            | 0.3    | 0.0     |          |         |               |           |                |
| 621T+1G>T CF         | 0.1    | 0.0     |          |         |               |           | <0.0001        |
| 2184delACF           | 0.1    | 0.0     |          |         |               |           |                |
| 3120+1G>A CF         | 0.1    | 0.0     |          |         |               |           |                |
| G551D CF             | 0.2    | 0.1     | 2.50     | 0.20    | 0.0           | 0.00      | 0.83           |
| W1282X CF            | 0.2    | 0.1     | 2.50     | 0.20    | 0.0           | 0.00      | 0.83           |
| G542X CF             | 0.2    | 0.0     |          |         |               |           |                |
| R1162X CF            | 0.1    | 0.0     |          |         |               |           |                |
| 2183AA>G CF          | 0.0    | 0.1     |          |         |               |           | 0.00           | 0.83           |
| All BD               | 14.2   | 9.8     | 1.50     | 0.002   | 25.5          | 4.63      | <0.0001        |
| R75Q BD              | 6.3    | 6.2     | 1.02     | 0.30    | 16.4          | 2.97      | 0.003          |
| S1235R BD            | 2.4    | 1.4     | 1.69     | 0.052   | 1.8           | 1.30      | 0.80           |
| R117H CF/BD          | 2.3    | 0.7     | 3.49     | 0.0007  | 5.5           | 8.74      | 0.0002         |
| L967S BD             | 1.1    | 0.2     | 6.87     | 0.002   | 1.8           | 11.17     | 0.014          |
| L997F BD             | 0.8    | 1.0     | 0.82     | 0.26    | 1.8           | 1.84      | 0.55           |
| D1152H BD            | 0.4    | 0.0     |          |         |               |           | 0.00           | 0.71           |
| D1270N BD            | 0.3    | 0.2     | 1.25     | 0.29    | 0.0           | 0.00      | 0.71           |
| R170H BD             | 0.3    | 0.0     |          |         |               |           |                |
| R74Q BD              | 0.3    | 0.1     | 3.02     | 0.17    | 1.8           | 21.15     | 0.002          |
| Other                |        |         |          |         |               |           |                |
| M470V                | 76.1   | 74.2    | 1.11     | 0.14    | 70.9          | 0.85      | 0.59           |
| T854T                | 57.3   | 57.8    | 0.98     | 0.29    | 45.5          | 0.61      | 0.071          |
| Q1463Q               | 39.6   | 39.5    | 1.01     | 0.30    | 40.0          | 1.02      | 0.94           |
| 1001+111>T*          | 13.4   | 10.9    | 1.27     | 0.016   | 14.5          | 1.40      | 0.42           |
| I25G>C               | 10.3   | 9.7     | 1.07     | 0.26    | 12.7          | 1.36      | 0.45           |
| P1290P               | 7.6    | 7.9     | 0.95     | 0.28    | 7.3           | 0.91      | 0.86           |
| 1716G>A              | 4.5    | 4.1     | 1.10     | 0.26    | 1.8           | 0.43      | 0.39           |
| R668C                | 1.0    | 1.4     | 0.72     | 0.19    | 0.0           | 0.00      | 0.38           |
| G576A                | 0.7    | 1.2     | 0.58     | 0.11    | 0.0           | 0.00      | 0.41           |
### Table 1. Cont.

| CFTR variant | %Cases w/N34S | %Uctrls | OR w/N34S | OR w/N34S | P-value w/N34S | P-value w/N34S |
|--------------|---------------|---------|-----------|-----------|----------------|----------------|
| F508C        | 0.5           | 0.3     | 1.58      | 0.21      | 0.00           | 0.00           |
| R1162L       | 0.5           | 0.3     | 1.99      | 0.17      | 0.00           | 0.00           |
| I1027T       | 0.3           | 0.4     | 0.75      | 0.27      | 0.00           | 0.00           |
| R31C         | 0.3           | 0.2     | 1.89      | 0.21      | 0.00           | 0.00           |
| I148T        | 0.3           | 0.2     | 0.85      | 0.29      | 0.00           | 0.00           |
| R297Q        | 0.1           | 0.2     | 0.63      | 0.27      | 0.00           | 0.00           |
| 0.2          | 0.1           | 0.1     | 1.26      | 0.30      | 0.00           | 0.00           |
| R74W         | 0.1           | 0.1     | 1.26      | 0.30      | 0.00           | 0.00           |
| F1052V       | 0.1           | 0.1     | 1.26      | 0.30      | 0.00           | 0.00           |
| 0.2          | 0.1           | 0.1     | 1.26      | 0.30      | 0.00           | 0.00           |
| R258G        | 0.1           | 0.1     | 1.26      | 0.30      | 0.00           | 0.00           |
| 0.1          | 0.1           | 0.1     | 1.26      | 0.30      | 0.00           | 0.00           |

Of the 81 mutations tested in the cohort, 43 were observed at least once in cases or controls. Data shown for CFTR CFTR variant alone and, in cases, with a concurrent heterozygous variant in SPINK1 N34S. *1001+11C. +11C. N'' except IVS8-T5 (1210-12T[5]). MD simulations comparing the channel diameters of the WT and mutants L997F and D1152H (Figure 2c–f) demonstrate that the channel diameter is observed to narrow down from an average value of 10.3 Å to 7.5 Å (standard deviation, σ = 0.5 Å) at the pore region, near the L997F amino acid substitution (Figure 2c), and from an average of 9.9 Å to 4.3 Å (σ = 1.1 Å) for the CFTRBD mutant D1152H (Figure 2f). Note that in contrast to the WT CFTR and L997F mutant where the structure maintains its stability, the D1152H mutation induces significant fluctuations in local conformation, which are reflected on the changes in the pore diameter at this location within the channel.

In order to determine residues that play a key role in the global dynamics of the CFTR, we performed an elastic network model (ENM) analysis. ENM analysis provides information on the mechanisms of collective movements intrinsically accessible to the structure, which usually enable structural changes relevant to function [40]. Application to CFTR highlighted the critical positioning of R74, R75, R170, L967, and R1162 at the hinge region that modulates the collective movements of the nucleotide-binding domains (NBDs) with respect to membrane-spanning domains (MSDs) (mode 1 in Figure 3). We also note that L967, L997, D1152, and R1162 act as anchors in collective mode 2. In this mode, the two NBDs are observed to move in opposite directions (see color-code diagram in Figure 3). The relative movements of the two NBDs, is known to control channel gating, hence the significance of this mode, or the alterations in mode 2 potentially caused by substitutions at the corresponding hinge site.

These two results suggest that substitutions of amino acids (or their side chains) at those particular regions could have an impact on the collective dynamics of CFTR, and interfere with concerted movements that would otherwise facilitate anion permeation. We noted that the mean-square fluctuations in our model are minimal at those mutation sites originates from the contribution of global (most collective) modes. In contrast, the CFTRBD candidate variants D1270 and S1235 are in close proximity on the surface of the NBD2 (Figure 3), and had weaker functional effects than other CFTRBD variants (Figure 1).
Association of CFTR<sup>BD</sup> variants with sinus disorders and male infertility

To examine the potential clinical relevance of CFTR<sup>BD</sup> variants, we reviewed case report forms for additional CF phenotypic features of dysfunction in the sinorespiratory and male reproductive systems, which both use CFTR for bicarbonate secretion. Association with CFTR<sup>CF</sup> alleles was used to test for CFTR-mediated chloride secretion, CFTR<sup>BD</sup> to test for selective bicarbonate-mediated secretion and, because both CFTR<sup>BD</sup> and CFTR<sup>CF</sup> cause defective bicarbonate conductance, association with either CFTR<sup>BD</sup> or CFTR<sup>CF</sup> alleles, or recessive genotypes (CFTR<sup>BD</sup>/CFTR<sup>BD</sup> or CFTR<sup>CF</sup>/CFTR<sup>CF</sup>) to assess overall risk of altered bicarbonate secretion on organ dysfunction.

The sinuses may use CFTR bicarbonate secretion, in part, for mucus hydration [41]. Sinusitis is common, with a complex genetic-anatomic risk that includes anatomy, allergies and mucus hydration [41]. Sinusitis is common, with a complex gene-environment-anatomic risk that includes anatomy, allergies and mucus hydration models [41]. Sinusitis was reported in pancreatic disease associated, and these were evaluated for specific functional studies in model cell types and focusing on a context-dependent signaling pathway. Although the CFTR<sup>BD</sup> variants were scattered throughout the genetic sequence, three-dimensional models of the protein provided insight into structural and dynamic mechanisms of dysfunction. Significant association between CFTR<sup>BD</sup> variants and symptoms of sinusitis and male infertility, but not overt lung disease as in CF, provided additional evidence of context-dependent dysfunction in humans. We believe that this type of integrated approach will be important in understanding the genetic contribution to this and other complex disorders and informing the development of therapeutics that target the molecular etiology rather than the phenotype.

CFTR bicarbonate secretion also plays a role in pH regulation in the male reproductive system [42]. Male infertility is uncommon and not dependent on recurrent infections. Self-reported male infertility over age 30 years was more common among cases (n = 17; 4.2%) than controls (n = 1; 0.4%, p = 0.03) (Table 2). We identified R75Q, R117H, and S1235R as well as the R117H, L967S, L997F, D1152H, and S1235R variants in our test panel (p = 0.02; OR 1.51; CI 1.05–2.18), but risk increased among carriers of CFTR<sup>BD</sup> (p = 0.001; OR 2.60; CI 1.43–4.60), CFTR<sup>CF</sup> (p = 0.01; OR 2.47; CI 1.18–4.91) or either CFTR<sup>BD</sup> or CFTR<sup>CF</sup> variant allele (p = 0.0001; OR 2.55; CI 1.55–4.15) (Table 2). Sinusitis was not statistically associated with recessive genotypes, possibly due to the complex nature of chronic sinusitis or requirement for an unidentified epistatic risk factor.

### Table 2. CFTR variants in subjects with chronic rhinosinusitis or male infertility (age >30 years).

| Rhinosinusitis | Yes | No | p-value | OR     | CI      |
|---------------|-----|----|---------|--------|---------|
| Controls      | 53  | 468|         |        |         |
| Cases (all)   | 151 | 798| 0.002   | 1.67   | 1.19–2.38|
| 0 CFTR<sup>BD</sup> | 111 | 649| 0.021   | 1.51   | 1.05–2.18|
| 1 CFTR<sup>CF</sup> | 14  | 50 | 0.011   | 2.47   | 1.18–4.91|
| 0 CFTR<sup>BD</sup> | 23  | 78 | 0.001   | 2.60   | 1.43–4.60|
| 1 CFTR<sup>CF</sup> | 37  | 128| 0.0001  | 2.55   | 1.55–4.15|
| CFTR<sup>BD</sup>/CFTR<sup>BD</sup> or CFTR<sup>CF</sup>/CFTR<sup>CF</sup> | 3   | 21 | 0.73    | 1.26   | NS      |

| Infertility   | Yes | No | p-value | OR     | CI      |
|---------------|-----|----|---------|--------|---------|
| Controls      | 1   | 160|         |        |         |
| Cases (all)   | 17  | 390| 0.03    | 7.14   | 1.10–300|
| 0 CFTR<sup>BD</sup> | 8   | 329| 0.28    | 3.88   | 0.5–174 |
| 1 CFTR<sup>CF</sup> | 2   | 24 | 0.051   | 13.0   | 0.65–786|
| 0 CFTR<sup>BD</sup> | 2   | 35 | 0.090   | 8.99   | 0.46–541|
| 1 CFTR<sup>CF</sup> | 4   | 59 | 0.023   | 10.7   | 1.03–536|
| CFTR<sup>BD</sup>/CFTR<sup>BD</sup> or CFTR<sup>CF</sup>/CFTR<sup>CF</sup> | 5   | 2  | 1.20E-07| 303    | 23–15783|

Top: Chronic rhinosinusitis in NAPS2 controls and cases with 0, 1, or 2 CFTR mutations. Bottom: Self-reported prevalence of male infertility among males over 30 years of age. Odds ratios were calculated comparing CFTR carrier cases in each subcategory against all controls. Because CFTR<sup>CF</sup> and CFTR<sup>BD</sup> both affect bicarbonate conductance, we calculated the association and risk associated with the presence of either variant type (shaded).

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Our integrative approach revealed a new functional class of rare CFTR variants of clinical significance in pancreatic disease. Targeted genotyping of reported and plausible CFTR variants in our cohort identified candidate variants with a high pre-test probability of being diseases associated, and these were evaluated for specific functional studies in model cell types and focusing on a context-dependent signaling pathway. Although the CFTR<sup>BD</sup> variants were scattered throughout the genetic sequence, three-dimensional models of the protein provided insight into structural and dynamic mechanisms of dysfunction. Significant association between CFTR<sup>BD</sup> variants and symptoms of sinusitis and male infertility, but not overt lung disease as in CF, provided additional evidence of context-dependent dysfunction in humans. We believe that this type of integrated approach will be important in understanding the genetic contribution to this and other complex disorders and informing the development of therapeutics that target the molecular etiology rather than the phenotype.
Figure 1. Functional characteristics of the nine CFTRBD variants. Panel a. Wild-type (WT) and variant CFTR proteins were expressed in HEK 293T cells and immunoblotted with anti-CFTR and anti-Aldolase antibodies. Replicate lanes are in small font. Band B, expected size of immature ER core-glycosylated CFTR; band C, mature complex-glycosylated CFTR. Panel b. Whole-cell Cl\textsuperscript{−} currents were measured in WT and variant CFTR-expressing HEK 293T cells, as described in Methods. Panel c. Whole-cell currents of WT-CFTR were measured in HEK 293T cells co-expressed with WNK1 and SPAK using patch pipette contained a low concentration of Cl\textsuperscript{−} (10 mM). A representative trace of reversal potential measurement is shown in the left panel. The permeability ratio \( P_{\text{HCO}_3}/P_{\text{Cl}} \) was calculated according to the Goldman-Hodgkin-Katz equation. I–V relationships at the indicated points are presented in the accompanying graph. The conductance ratio \( G_{\text{HCO}_3}/G_{\text{Cl}} \) was calculated by measuring each outward current (i.e., slope between \( E_{\text{rev}} \) and \( E_{\text{rev}}+25 \) mV). RMP, resting membrane potential. Panel d. Whole-cell currents of R170H-CFTR were measured in HEK 293T cells using the same protocol shown in panel c. Panel e. A summary of the \( P_{\text{HCO}_3}/P_{\text{Cl}} \) values obtained from WT-CFTR in the standard state (left) compared to WT-CFTR and the nine CFTRBD variants with WNK1 + SPAK activation (right, underlined). Panel f. A summary of the \( G_{\text{HCO}_3}/G_{\text{Cl}} \) values in the standard state (left) with WNK1 + SPAK activation (right). Values throughout are means ± SEM. * p<0.05, **p<0.01: difference from WT in cells co-expressed with WNK1 and SPAK.

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Evidence that CFTRBD variants are associated with pancreatitis

One of the challenges of genetic association studies is determining the effect of candidate genetic variants by statistical tests when the variant is rare or the mutation effect is uncertain. One approach is to increase study power by markedly increasing study subject numbers, but this approach is prohibitively expensive and not always feasible in rare diseases. Another approach is to evaluate the combination of statistical trends linked to studies of the functional effects of a variant in a biological system and a biologically plausible framework.

In the current study, 11 variants that were previously reported to be present in chronic pancreatitis but not CF causing [16,17,43–52] underwent functional testing. Only CFTR M470V and R1162 (not shown) did not meet criteria of altered bicarbonate permeability and/or conductance after WNK1 and SPAK activation (Figure 1e–f, discussed below). The remaining 9 CFTRBD variants were identified at least twice in pancreatitis association studies over the past decade.

Five variants (R74Q, R75Q, R170H, L967S, and R1162L) were located in the hinge region that modulates the collective movements of the NBDs with respect to the MSDs (Figure 3). R74Q was previously reported in a single chronic pancreatitis patient [53] but not in the CFTR2 database. CFTR R74Q was identified by us in two cases and no controls (p = ns) and in one case who was a SPINK1 N34S carrier (p = 0.006). R75Q is considered to be a non-CF causing mutation according to the CFTR2 mutation database [54]. CFTR R75Q was identified in 61/906 cases and 75/1214 controls (6.3 vs. 6.2%, p = ns) but was also detected in nine SPINK1 N34S/- mutation carriers (9/55, 16.4%), with strong combined effect (SPINK1 OR 3.7, SPINK1 + R75Q compound OR 12.2, p 0.002). Two of the nine trans-heterozygous cases had been previously reported[18]. R75Q was also identified in four cases with a concurrent severe CF-causing mutation and in no compound controls. R170H was first reported...
in two cases of congenital bilateral aplasia of vas deferens in England [53] but is not currently in the CFTR2 mutation database. CFTR R170H was identified in three cases and no controls (p = ns). L967S has been reported in a single case of azoospermia from the CF mutation database [53] but is not in the CFTR2 mutation database. L967S was identified in ten cases (one trans-heterozygote), two controls (OR 6.9 p = 0.004), and one N345s case carrier. R1162H is predicted to be a highly deleterious variant by SIFT and damaging by PolyPhen modeling [55] and is included in the CFTR2 mutation database and classified as a variant not causing CF. Although located in a critical portion of the CFTR molecule, the association and functional threshold for inclusion as a CFTRBD variant were not fully met.

Two variants (L997F and D1152H) appeared to reduce channel diameter. L997F is considered a mutation of varying clinical consequences for CF, with low rates of pancreatic insufficiency and retention of chloride conductance [54]. In this study L997F was identified both in the cases (0.7%) and controls (1.0%), additionally, L997F was identified in one N345s case carrier and three compound heterozygous mutation case carriers, but independent statistical association with pancreatitis was not demonstrated in this study. D1152H is a mutation of varying clinical consequence for CF and is associated with low rates of pancreatic insufficiency and retention of chloride conductance [53]. CFTR D1152H was identified in four cases and no controls (p = 0.014). Two of these cases were in compound heterozygosity with F508del.

Two variants (S1235 and D1270N) were on the surface of NBD2 (Figure 2). S1235 is a non-CF-causing mutation [54] and was identified in 2.4% of cases and 1.4% of control (p = ns), three compound heterozygous cases and one N345s case carrier. While this did not reach statistical significance in this cohort, multiple previous reports of CFTR S1235R in idiopathic pancreatitis patients [56,57] and complex functional features [27] were noted. D1270N is of varying clinical consequences for CF, with low rates of pancreatic insufficiency and retention of chloride conductance [54]. D1270N was identified both in the cases (0.3%) and controls (0.2%). Although these variants have been identified in previous studies, the effects of these rare variants on altered bicarbonate permeability and conductance appear to be weak (Figure 1 e–f) and the effect on the function of NBD2 (Figures 2–3) is unclear. However, they meet minimal criteria for the class on function grounds and contribute to the overall effect on disease risk.

The final variant (R117H) is located in an extracellular domain and has functional effects beyond the other CFTRBD variants. R117H is a complex variant that is associated with CF only when found in cis with a T5 tract in intron 8. The CFTR R117H variant was identified in 22 cases (2.3%) and 8 controls (0.7%) (p = 0.001), with only 3 cases and 1 control having the CF-associated R117H*T3 haplotype (p = ns), which links the CFTR variant R117H to pancreatitis regardless of the intron 8 T5 haplotype. R117H*T7/T9 was also identified in 9 of the 80 cases with a concurrent severe CF-causing mutation and in no CF carrier controls. The R117H variant was the only one with reduced chloride current density (Figure 1b). While the variant was associated with altered bicarbonate permeability and conductance, the mechanism is yet to be determined.

Common CFTR variants previously associated with pancreatitis but not confirmed in the current study

The common polymorphisms M470V, T854T, and Q1463Q had no significant association with pancreatitis, either individually or combined in haplotypes, in contrast to a previous report [58]. Haplotypes were determined by counting homozygous carriers of each subset (M470V, T854T, P1290P, Q1463Q and M470V, IVS-T, IVS-TG) and applying Fisher’s exact test. The IVS8 T/ TG/M470V allele was evaluated in 764 NAPS subjects and controls, and no significant associations were found, in contrast to another report [59]. The possibility that a series of complex haplotypes affect CFTR expression or exon skipping was not excluded, but no evidence of direct association was seen in the current study or our previous pancreatitis GWAS [24].

Thirty-seven of the 81 CFTR variants tested were not identified in any cases among the NAPS cohort. The remaining variants were also not significantly overrepresented alone or with SPINK1 or CF mutation carrier. H148T was seen in three cases and one control, so an effect could not be detected or excluded; the in cis deletion mutation 3199del6 was not detected in any H148T carriers. The IVS8T5 variant was identified in 9.9% of cases and 8.2% of controls, which is not individually significant. There were six N348s/T5 trans-heterozygote controls and no cases, but the combined odds ratio (OR 3.9) of the SPINK1 N348s variant with IVS8T5 was not significantly higher than N348s alone. Four additional variants were identified in only one patient and no controls: CF mutations 2104delA, 3120+1G>A, R1162X and a mutation of varying clinical consequence, G1069R.

Taken together, these genotyping and functional studies provide strong rationale for inclusion of nine variants as CFTRBD class members. Although additional variants may be added to the CFTRBD class in the future, the current study did not have the very large patient size needed to provide adequate power to detect statistically significant changes in additional rare variants. In addition, other possible mechanisms of CFTR channel dysfunction linked to altered bicarbonate conductance are possible, such as mechanisms linked to CFTR R117H.

Structural significance of CFTRBD mutants

Structure-based simulations can provide insights into molecular driving forces and thereby into the mechanisms of channel dysfunction. To better understand the location and structural effects of the nine amino acid variants conferring risk of pancreatitis and causing dysfunction of the electrophysiological response to WNK1-SPAK activation, we developed structural models of CFTR and conducted dynamic simulations. Because no crystallographic structures for the entire human CFTR are currently available, we built a homology model based on the structure of a bacterial ABC transporter (Sav1866) from Staphylococcus Aureus [35]. Several computational studies have been carried out using models of CFTR and other ABC transporters that focus on the structure and/or gating cycle of the molecule and the effect of common mutations/deletions (e.g., F508del in CFTR) [4,5,35–37,39,60–64]. Our study is, to our knowledge, the first to investigate the multiscale dynamics of CFTR by examining both the global motions of the overall protein (with ENM) and the local effects of particular CFTRBD variants (with MD). The ENM analysis highlighted the critical positioning of R74, R75, R170, L967, and R1162 at the global hinge regions (those between the NBD and MSD of transporter in mode 1, and between the two NBDs in mode 2), as evidenced by the significant suppression of residue fluctuations in their close neighborhood. Mutations at those sites would thus be expected to interfere with the functional dynamics of the channel. Our all-atom MD study, on the other hand, showed that a substantial constriction could arise in channel diameter with substitutions at residues lining the wall of the channel. In particular, the L997F and D1152H mutants showed channel pore size reductions in
their neighborhoods that would directly affect conductance properties.

**Defects in CFTR bicarbonate transport**

The fact that all of the pancreatitis-associated variants identified by genetic screening in this study resulted in defective WNK1-SPAK-activated increase in bicarbonate secretion supports the argument that this mechanism is critical for bicarbonate-secreting cells that utilize CFTR as the primary anion channel. The importance of bicarbonate conductance across CFTR at the apical membrane is magnified if chloride, but not bicarbonate, conductance across the basolateral membrane is minimal, as predicted for the pancreatic duct cell [6], since the transcellular anion conductance is responsible for fluid secretion. Under basal conditions, CFTR-mediated bicarbonate permeability is only ~20% of chloride, and the capacity for facilitating high bicarbonate flux for bicarbonate-secreting tissues is limited. Under conditions of low-intracellular chloride, the WNK1-SPAK pathway are activated, and this in turn transforms CFTR into a highly bicarbonate-permeable anion channel (Figure 1). The molecular mechanisms as to how WNK1-SPAK increases the CFTR bicarbonate permeability remain unclear. However, increasing evidence suggests that ion permeability of anion channels is not fixed and can be dynamically modulated by cellular signaling and other events [65].

The pore of anion channels is believed to have a large polarizable tunnel, where ion selectivity is basically determined by the hydration energy of ions and polarizability of the channel pore [65]. Therefore, in general, the CFTR ion channel is more permeable to large anions that are more readily dehydrated [66]. However, this cannot be applied to HCO$_3^-$, although the size of HCO$_3^-$ (equivalent radius: 2.1 or 2.43 Å) is larger than Cl$^-$ (1.81 Å), most anion channels, including CFTR, exhibit poor HCO$_3^-$ permeability because of the asymmetrical charge distribution of HCO$_3^-$ [67]. A decrease in the CFTR pore diameter, as shown in L997F, can affect the permeability of HCO$_3^-$ in many ways, such as by limiting the accessibility of large, asymmetrically charged HCO$_3^-$ to the channel pore. A second mechanism of reducing HCO$_3^-$ permeability and conductance is to inhibit the interaction between CFTR and WNK1/SPAK or to reduce the WNK1/SPAK-mediated conformational change of CFTR. The eludication of precise molecular mechanisms of each mutation will provide insights into the functional change of CFTR. The elucidation of precise molecular properties.

**Materials and Methods**

**Study cohort**

The NAPS2 cohort was ascertained, and data were collected as described previously [23]. All patients were prospectively enrolled using protocols approved by the appropriate IRBs. Physician-confirmed diagnosis of pancreatitis was required for enrollment as a case, while questions on CF, chronic sinusitis, and male infertility were included on a case report form administered by a clinical research coordinator. DNA and phenotypic data for patients with chronic and recurrent acute pancreatitis (n = 984) and healthy unrelated controls (n = 467 from the NAPS2 case-control study [23,68] plus DNA from additional healthy controls from SomaLogic Inc. (Boulder, CO) (n = 377), the Inflammatory Bowel Disease Genetics Consortium (Dr Richard Duerr, University of Pittsburgh) (n = 338) and additional University of Pittsburgh studies of pancreatitis and pancreatic cancer (Dr. David Whitcomb and Randall Brand, University of Pittsburgh) (n = 42) [24] were evaluated for a final study cohort of 984 cases and 1224 unrelated controls.

**Genotyping**

PRSS1 genotyping was done by DNA sequencing [69]. SPINK1 genotyping was done by sequencing exons 2–3 and the flanking regions in a preliminary subset of 745 NAPS2 cases, with the entire cohort (cases and controls) genotyped for p.N34S, p.S558 and c.27delC using TaqMan assays. The SPINK1 c.194+5G>A variant [70] was seen in one patient and one control; c.194+2T>C [71] was not identified in the initial sequencing and was not further genotyped.

CFTR variants for the screening panel were selected from a review of published papers and abstracts between 1998 and 2010 [16,17,43–52] and the open access CFTR mutation database based in the Hospital for Sick Children in Toronto (http://www.genet.sickkids.on.ca) and Johns Hopkins University (http://CFTR2.org).

**CFTR genotyping**

was done using a custom MassARRAY iPLEX Gold assay (Sequenom, Inc, San Diego, CA) or custom TaqMan Gene Expression Assays (Life Technologies Corporation, Carlsbad, CA) through the Genomic and Proteomic Core Laboratories at the University of Pittsburgh and verified by bidirectional DNA sequencing. All cases and controls were tested for each of the 81 selected CFTR variants (Table S1). Variants were selected in three stages: the most common CF-causing mutations in North America, variations which have been reported in pancreatitis literature at least once and a subset of variants that have been reported in CF patients but for which the biological and pathological relevance remains undetermined (Mutations of Undetermined Clinical Significance). 67 SNPs (125GtoC, 1716G>A, 1717-1G>A, 1898+1G>A, 2183AA>G, 2184delA, 2789+5G>A, 3120+1G>A, 3639delC, 3819+10kbC>T, 621+1G>A, T11+4G>A, A455E, D110H, D1152H, D1270N, D443Y, D579G, F1025V, F1047L, F508C, F508del, G1069R, G1244E, G1349D, G178R, G542X, G551D, G551S, I1131L/V, I148T, I336K/T, I3507del, I807M, IVS8T5, K1180T, L1065P, L967S, L997F, MIV, M470V, M932I, M952T, N1303K, P67L, Q1463Q, R1017Q, R1162X, R117C, R117H, R170H, R253G, R297Q, R31C, R352Q, R353X, R666C, R747W, R753Q, S1235R, S1255P, S485R, S977F, T338I, T354T, V201M, W1282X) were multiplexed into 6 wells; 14 SNPs (S492F, S945L, R74Q, R360T, R1162L, G85E, I1027T, R334P, G576A, 711+1G>T,
1001+11C>T, P1290P, 3199del6) were ascertained separately via TaqMan Gene Expression Assays, with repeat confirmation of all positive results. 3199del6 was genotyped via TaqMan on all samples that tested positive for 1140T. In addition, the intron 3 boundary was directly sequenced in 873 subjects to determine the significance of the IVS8 T/TG tract.

**Statistical analysis**

Significant differences in carrier frequencies among cases and unrelated controls were determined by chi square analysis or Fisher’s exact test, and two-tailed p-values are reported. The results of each set of experiments are presented as means ± SEM. Statistical analysis was performed using Student’s t-tests or analysis of variance followed by Tukey’s multiple comparison test as appropriate. P<0.05 was considered statistically significant.

**Cell culture and plasmids**

HEK 293T cells were maintained in Dulbecco’s modified Eagle’s medium-HG (Invitrogen, Grand Island, NY) supplemented with 10% fetal bovine serum, 100 U/mL penicillin, and 0.1 mg/ml streptomycin. The mammalian expressible plasmids for hCFTR[22], Myc-rWNK1[21] and Flag-mSPAK [20] were described previously. Plasmids were transiently transfected into cells using Lipofectamine 2000 reagents (Invitrogen, Grand Island, NY). An average transfection rate over 90% was confirmed by transfection with a plasmid expressing green fluorescence protein (pEGFP-N1). Plasmids expressing variant hCFTRs were generated using a PCR-based site-directed mutagenesis kit (Stratagene, Santa Clara, CA).

**Immunoblotting**

Immunoblotting was performed using conventional methods [20]. Briefly, cells were harvested with lysis buffer (20 mM HEPES pH 7.4, 150 mM NaCl, 5 mM EDTA, 1% Triton X-100, 1 mM NaVO4, and 1 mM β-glycerophosphate) containing a complete protease inhibitor mixture (Roche Applied Science, Mannheim, Germany). Protein samples were suspended in a sodium dodecyl sulfate buffer and separated by SDS-polyacrylamide gel electrophoresis. The separated proteins were transferred to a nitrocellulose membrane and blotted with appropriate primary and secondary antibodies, and protein bands were detected with enhanced chemiluminescence solutions. Antibodies against CFTR (M347, Millipore, Billerica, MA) and aldolase A (N-15, Santa Cruz Biotechnology, Inc., Dallas, TX) were obtained from commercial sources.

**Electrophysiology**

Voltage and current clamp experiments were performed on HEK 293T cells transiently transfected with hCFTR as previously reported with slight modifications [20]. Briefly, cells were transferred into the bath mounted on a stage with an inverted microscope (IX-71, Olympus, Osaka, Japan). The pipettes were pulled by a Sutter P-57 puller and have free-tip resistances of about 2–5 MΩ. These were connected to the head stage of a patch-clamp amplifier (Axopatch-700B, Molecular Devices, Sunnyvale, CA). Ag-AgCl reference electrodes were connected to the bath via a 1.5% agar bridge containing 3 M KCl. Liquid junction potentials were corrected for each experimental solution as described previously[20]. For the anion permeability test, individual data were corrected by measuring the offset potential shift induced by the replacement of anion solution after each experiment. The conventional whole-cell clamp was achieved by rupturing the patch membrane after forming a gigaseal. Voltage and current traces were stored and analyzed using Clampfit v.10.2 (Molecular Devices, Sunnyvale, CA). currents were sampled at 5 kHz. All data were low-pass filtered at 1 kHz.

The high-chloride pipette solution contained (mM) N-methyl-D-glucamine chloride (NMGD-Cl), 5 ethylene glycol tetraacetic acid, 1 MgCl2, 3 Mg-ATP and 10 l-2-hydroxyethyl)-1-piperazinenucleosulfonic acid (HEPES)(pH 7.2). The low-chloride pipette solution was prepared by replacing Cl− with equimolar glutamate. The stand bath solution contained (mM) 146 NMDG-Cl, 1 CaCl2, 1 MgCl2, 10 glucose and 10 HEPES (pH 7.4). The high-bicarbonate-containing bath solution was made by replacing NMDG-Cl with equimolar choline-HCO3. The bicarbonate-containing solution was continuously gassed with 95% O2:5% CO2.

In all experiments, currents generated by CFTR were confirmed by the following three characteristics: 1) activation of current by the treatment with cAMP (5 µM forskolin and 100 µM 1-methyl-3-(2,2-dimethylpropyl)-7H-purine-2,6-dione (IBMX), 2) a linear I–V relationship and 3) inhibition of current by the treatment with the CFTR inhibitor CFTRinh-172.

The current reversal potential (Erev) was measured either in current clamp mode or in voltage clamp experiments. Rether membrane potential (RMP) was recorded in zero current clamp mode. To test the current-voltage relationship during zero-current clamp recording, clamp mode was shifted to the voltage clamp mode, and the I–V curve was achieved with ramp pulses from -100 to 100 mV (250 ms, holding potential; near the RMP). All currents were corrected for capacitative currents and the I–V relationship was plotted using the values of current density (pA/pF).

The relative anion permeability was determined by the reversal potential shift (ΔErev = Erev(X) – Erev(Cl)) induced by replacing extracellular Cl− with X− anion using the Goldman-Hodgkin-Katz equation as follows: 
P_X/P_Cl = exp(ΔE_rev/(RT/ZeF)) – ([Cl−]/[X−])ho × ([Cl−]/[X−])o, where [Cl−]o is the bath concentration of Cl−; [Cl−]o, is the residual Cl− in the substituted solution; [X−]o is the concentration of substitute ion; and R, T, z and F have their conventional thermodynamic meanings. The anion outward chord conductance (GX; X is anion) between Erev and Erev +25 mV was achieved by linear plotting.

**Structural modeling**

The homology model of human CFTR (UniProt accession code: P13569) was obtained using the Swiss-Model Workspace software [73]. The most recently resolved crystal structure of the *Staphylococcus Aureus* savA1866 ABC transporter, fitted to the human CFTR (PDB code: 4A82, 2.0 Å resolution) [36] was adopted as a template, and the structural model was completed using the X-ray crystallographic structure of the NMD2 region of human CFTR (PDB code: 3GD7, resolution 2.7 Å) (see Figure S3a). This model deviates from the template structure by 1.6 Å RMSD, and in our simulations the RMSD levels off at ~3.5 Å. The MSD pore-lining residues and pore radius profile (Figure S3b–c) were consistent with those observed in a homology model constructed by Norimatsu and coworkers [37,39], which was based on an earlier structure (PDB code: 2HVD, resolution 3 Å) [35]. Using this model for WT CFTR, we generated in silico models for the mutants I997F and D1152H.

Of note, the collective modes predicted by ANM are highly robust and they are not sensitive to small structural variations (like those due to a different model).

**Molecular dynamics simulations and elastic network model analysis**

Molecular dynamics simulations were performed using the AMBER11[74] package (GPU version of the pmemd program),
with the Amber99SB[75] force field and using the TIP3P water model. The protocol consisted of an initial minimization in vacuum, using 1,500 steepest descent and 1,500 conjugate gradient steps, to remove strong steric contacts, followed by another minimization of 5,000 steepest descent and 5,000 conjugate gradient steps, in explicit solvent, followed by a production run of 50 ns. The systems were kept at a temperature of 300 K, using Langevin dynamics with a collision frequency of 2 ps⁻¹; the SHAKE algorithm was adopted to use a 2 fs time step. The stability of the system was assessed by verifying the convergence of the root mean square deviation (rmds) of its heavy atoms, after the first 5 ns of simulation.

As to the pore regions where we examined the local effects of substitutions, we allowed for the relaxation and optimization of interactions during the described protocol. The simulations, thus, gave rise to local rearrangements in the neighborhood of the mutation sites and permitted us to extract statistical data on the average pore diameter at the constriction zone and its fluctuations.

The elastic network model analysis of collective dynamics was performed using the approach reviewed earlier[40]. Collective modes of motions are evaluated by eigenvalue decomposition of the connectivity/Hessian matrix, using the Gaussian/Anisotropic network model. The shape of the mode permits us to identify regions subject to large fluctuations as well as domains undergoing anti-correlated movements (colored blue and red in the ribbon diagrams, Figure 3).

Ionic diameter

The radii of the mono-atomic chloride ion was taken from Bondi [34]. The equilibrium geometry of bicarbonate ion was optimized using ab initio quantum mechanics at DFT level, with the B3LYP/6-311+G** basis set, via the Gaussian 03 software. This resulted in a bicarbonate ion that could be fit in a minimum box of size 3.40 Å×4.86 Å×5.39 Å. This yields a van der Waals radius of 2.1 (or 2.43) Å for the bicarbonate ion, based on the two smaller dimensions (or the second largest dimension) that define the minimal cross-sectional area.

Unless specified otherwise, when we refer to the diameter of the pore, we mean the minimal diameter at the specific location of the mutation, as opposed to the distribution of diameters along the pore.

Supporting Information

Figure S1 Measurement of P_{HCO3}/P_{Cl} in cells without WNK1 and SPAK co-expression. Control experiments were performed. Whole-cell recordings were performed to measure CFTR bicarbonate permeability by replacing the bath solution with high HCO₃⁻-containing (140 mM) solution. Cells were stimulated with cAMP (5 μM forskolin and 100 μM IBMX) after establishing whole-cell configuration. The current to voltage relationship (I/V curve) was obtained by depolarizing ramp pulses from −100 to +100 mV (250 ms), and all currents were corrected for capacitative currents. Treatment with the CFTR inhibitor CFTRinh-172 (20 μM) inhibited the HCO₃⁻ currents by an average of 91.8±3.0% (WT-CFTR with WNK1 & SPAK coexpression, measured at +100 mV, n = 4) indicating that CFTR mediates most of the HCO₃⁻ currents. The permeability ratio P_{HCO3}/P_{Cl} was calculated according to the Goldman-Hodgkin-Katz equation. The conductance ratio G_{HCO3}/G_{Cl} was calculated by measuring each outward current (slope between E_{rev} and E_{rev+25} mV).

Figure S2 Representative current to voltage (I–V) plots of CFTR variants in whole-cell current measurements. Whole-cell recordings were performed to measure the CFTR HCO₃⁻ permeability and conductance by replacing the bath solution with high HCO₃⁻-containing (140 mM) solution. The pipette solution contained 10 mM Cl⁻, WNK1 and SPAK kinases were coexpressed with wild-type (WT) or variant CFTR. Cells were stimulated with cAMP (5 μM forskolin and 100 μM IBMX) after establishing whole-cell configuration. The I–V curve was obtained by depolarizing ramp pulses from −100 to +100 mV (250 ms), and all currents were corrected for capacitative currents. Treatment with the CFTR inhibitor CFTRinh-172 (20 μM) inhibited the HCO₃⁻ currents by an average of 91.8±3.0% (WT-CFTR with WNK1 & SPAK coexpression, measured at +100 mV, n = 4) indicating that CFTR mediates most of the HCO₃⁻ currents. The permeability ratio P_{HCO3}/P_{Cl} was calculated according to the Goldman-Hodgkin-Katz equation. The conductance ratio G_{HCO3}/G_{Cl} was calculated by measuring each outward current (slope between E_{rev} and E_{rev+25} mV).

Figure S3 Structural features of the present homology model and comparison with previous work. Panel a shows the superposition of the current homology model onto the X-ray structure of ABC transporter from S. aureus, which yields an RMSD of 1.6 Å. Panel b shows the position of some residues lining the pore at the MSD, located on two TM helices, 6 (red) and 12 (yellow). These residues were reported by Sansom and collaborators to be lining the MSD pore [61], based on both, a homology model and experimental cysteine scanning. Panel c shows the MSD pore radius profile—measured as the radius of the smallest circle that fits the cross sectional area at each elevation along the pore axis (perpendicular to the membrane plane), and its extension toward the cytoplasmic region. The MSD portion (indicated by the upper abscissa label) is comparable to that reported earlier by Norimatsu et al [37].

Figure S4 Contribution of the slowest modes to the square displacements of residues in CFTR. Square displacements are calculated using the ENM representation of the two subunits of the structure (residues 71–645 and 846–1445, respectively). The right panel shows a color-coded ribbon diagram where regions subject to large fluctuations are colored pink, and those maximally constrained, blue. Note that R74, R75, R170, I967, D1152 and R1162 lie in the highly constrained region.

Table S1 81 CFTR variants genotyped in pancreatitis patients. The CFTR mutations investigated in this study are reported with legacy nomenclature and relative ranking among the American College of Medical Genetics most common classic cystic fibrosis-causing mutations found in North America (CF). Those found to be associated with cases in the current cohort include AN in the Pance Disease column. *IVS8 T5 and R117H are reported but CF disease causing only when in cis with each other or IVS8 T5 with IVS8 TG12or13. Intronic mutations are reported in standard nomenclature “##+/−##N” except IVS8-T5, (1210-12T)[5].

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