GAMBOGIC ACID AS ANTICANCER AGENT: A REVIEW

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Abstract

Gambogic acid, a common traditional Chinese medicine and widely distributed throughout South China, Vietnam, Cambodia, and Thailand. It is prenylated xanthone which is the significant bioactive compound of gamboge. Gambogic acid is known as a strong apoptotic inducer in cancer cells. It has been found as strong anticancer agent against various types of cancer cells lines such as breast cancer, pancreatic, and cervical cancer. It induces apoptosis, down regulates the anti-apoptotic proteins (survivin and BCL2,) and down regulates the activities of P-glycoprotein in drug sensitive human breast MCF-7 and drug-resistant MCF-7/ADR cells. Similarly, it also exerts alteration in P13K, AKT, p21, MMP-2 &-9, and phosphorylated-AKT expressions. The current review highlights the anticancer and chemo-preventive perspectives of gambogic acid and its mechanistic role against human and animal cancers.

Key words: Gambogic acid, phytochemical, breast cancer, pancreatic cancer, glioma cancer

Introduction

Natural bioactive compounds from fruits and vegetables have been used as therapeutic agents in clinical and biological activity against various human maladies such as cancer, cardiovascular, and neurodegenerative disorders. Gambogic acid (GA), a naturally occurring compound which is derived from Garcinia hanburyi tree. It has been found to exert multiple intracellular and extracellular actions such as antimetastatic, anti-inflammatory autophagy, programmed cell death, anti-angiogenesis, and cell cycle arrest, respectively (Kashyap et al., 2016; Seo et al., 2019). Multiple molecular targets and pathways of gambogic acid have been studied such as caspase-3 apoptosis, PI3K/Akt, TNF-α, ATR-Chk1, and MET pathways (Ishaq et al., 2014). Likewise, GA
induces apoptotic cell death via mitochondrial pathway, enhancing the active oxygen concentration, suppressing the inhibiting the NF-κB, MAPK/ERK, and PI3K/AKT expressions in HT-29 cells (Lü et al., 2012; Wang et al., 2014). It also suppresses the cancer cell proliferation and cell growth in various human cancers i.e. breast, prostate, pancreatic, colon, cervical and lung etc (Kashyap et al., 2016).

**Anticancer perspectives**

**Breast cancer**

GA has been found to prevent from tumorigenesis in the MCF-7 subcutaneous xenograft tumor model through inhibiting proliferation and inducing apoptosis (Wang et al., 2018a; Sang et al., 2018). Tumor necrosis factor related apoptosis- inducing ligand (TRAIL) has the ability to induce apoptosis in cancer cells that has developed significant interest in treating different types of cancers. In breast cancer cells, gambogic acid increased sensitivity of TRAIL and increased TRAIL-induced apoptosis in cancer cells. Wang et al (2018) discovered that GA with TRAIL co-operation decreased anti-apoptotic proteins level and activated Bid (BH3 interacting-domain death agonist) which promoted cross talk between extrinsic and intrinsic apoptotic signaling, instead of enhancing the TRAIL DR4 and DR5 receptors (Wang et al., 2018b). Different researchers and investigators determined that encapsulation of GA with polyethylenimine-poly (dl-lactide-co-glycolide) caused momentous augmentation in apoptotic cell death in an in vitro study on triple-negative breast cancer, and in vivo suppression of TNBC tumor growth (Wang et al., 2017; Xu et al., 2016). In another exploration by Doddapaneni et al. 2016, they found that encapsulation of GA as PEGylated liposomal formulation in MDA-MB-231 orthotopic xenograft significantly suppressed the growth of tumor, with 50% tumor-volume, and reduced tumor weight alongside inhibition of Bcl2 expression, apoptotic markers, cyclinD1, and micro-vessel density marker-CD31. Moreover, it also increased the levels of p53 and Bax (Doddapaneni et al., 2016).

GA increased cell toxicity, apoptosis, down regulated expressions of anti-apoptotic proteins survivin and Bcl2, and downregulated expression and activities of P-glycoprotein (P-gp) in drug sensitive human breast MCF-7 and drug-resistant MCF-7/ADR cells (Wang et al., 2015b). Sensitization of doxorubicin (DOX) resistant breast cancer cells to DOX-mediated cell death and enhancement in the intra-cellular accumulation of DOX via inhibiting both P-gp expression and activity were reported after gambogic acid treatment. In addition, DOX in combination with GA
led to the production of intra-cellular reactive oxygen species and inhibition of anti-apoptotic protein surviving. In DOX-induced apoptosis, over-expression of surviving blocked the sensitizing effects of GA. Moreover, ROS-mediated activation of p38 MAPK was revealed in GA-mediated suppression of Survivin expressions (Wang et al., 2015a).

**Glioma cancer**

A group of researchers studied the anticancer role of GA against human U251 glioma cells, they found that reduction in phosphorylation of P38, AKT, and mTOR, as well as decrement in the upstream binding factor (UBF), phosphorylation of ribosomal protein precursors (Pre) and insulin-like growth factor I (IGF-1) were reported after GA treatment (Luo et al., 2020; Sang et al., 2019). In T98G glioblastoma cells, GA dose dependently showed potent anti-proliferative activity via apoptosis induction, enhancement in Bax and AIF expression, PARP and cleavage of caspase-3,-7 & -9 and down regulation of Bcl-2 expression (Thida et al., 2016). A peer of researchers, they investigated induction of up regulation of leucine-rich repeat and Ig-like domain-containing-1(LRIG1) which further exhibited downstream Akt/mTORC1 inhibition and epidermal growth factor receptor (EFGR) degradation after GA treatment in U87 glioma cells. Further, U87 cell apoptosis and growth inhibition, AMP-activated protein kinase activation mediated GA-induced LRIG1 upregulation, while AMPK inhibition by shRNA or compound C reduced GA-induced EGFR/Akt inhibition and cytotoxicity in U87 cells (He et al., 2013). Time and dose-dependent anti-cancer action of GA investigated by group of researchers against human glioma cell lines such as U251MG and U87MG showed decreased cell proliferation, induced apoptosis in cells, and inhibited colony formation. Development of monodansylcadaverine in autophagic vacuoles, up regulation of expressions of Beclin 1, LC3-II and Atg5, enhancement in punctate fluorescent signals of glioblastoma cells pre-transfected with GFP-tagged LC3 plasmid were reported (Luo et al., 2012). During an in vitro study on rat C6 glioma cells, GA dose and time dependently induced apoptotic cell death as it triggered intrinsic mitochondrial apoptotic pathways whereas during an in vivo trial, intravenous treatment of GA showed momentous reduction in tumor volumes through apoptotic induction (Qiang et al., 2008).

**Skin cancer**

In recent study reported by Li and their colleagues, they found that dose and time dependent GA in vitro study in melanoma A375, B16-F10 cells regulating the protein expressions, inhibiting the
proliferation, migration, invasive and adhesive properties, suppressing the EMT and angiogenesis processes and reducing the enzymatic activities of MMP-2 and MMP-9. Furthermore, GA also suppressed the abnormal PI3K/Akt and ERK signaling pathways (Li et al., 2019). In human malignant melanoma (MM) cells A375, GA (2.5-7.5 microg/mL) for 36 h suppressed the caspase-3 activity and Bax/ Bcl-2 ratio, respectively (Xu et al., 2009).

| Types       | Mechanisms                                                                 | References                                                                 |
|-------------|-----------------------------------------------------------------------------|-----------------------------------------------------------------------------|
| Breast cancer | Inhibited proliferation and induced apoptotic cell death                    | (Wang et al.,2018a; Sang et al., 2018).                                      |
|             | Increased the sensitivity of breast cancer cells to tumor necrosis factor    |                                                                             |
|             | (TNF)-related apoptosis-inducing ligand (TRAIL)                             |                                                                             |
|             | Enhanced TRAIL-induced apoptosis                                            |                                                                             |
|             | Caused momentous augmentation in apoptotic cell death                        |                                                                             |
|             | Suppressed TNBC tumor growth                                                | (Wang et al., 2017; Xu et al., 2016)                                        |
|             | Significantly suppressed the tumor growth, tumor volume, and caused         | (Doddapaneni et al., 2016)                                                  |
|             | reduction in tumor weight                                                   |                                                                             |
|             | Showed inhibition on expression of bcl2, apoptotic markers, surviving,     |                                                                             |
|             | cyclinD1, and microvessel density marker-CD31. Increased the levels of      |                                                                             |
|             | p53 and Bax                                                                  |                                                                             |
| Glioma cancer| Caused reduction in phosphorylation of P38, AKT, and mTOR                    | (Luo et al., 2020; Sang et al., 2019)                                        |
|             | Lowered the upstream binding factor (UBF), phosphorylation of               |                                                                             |
|             | ribosomal protein precursors (Pre) and insulin-like growth factor I (IGF-1) |                                                                             |
|             | Induced apoptosis and increased Bax and AIF expression, Enhanced           | (Thida et al., 2016)                                                        |
|             | cleavage of caspase-3, -8, -9, and PARP                                     |                                                                             |
|             | Down regulated Bcl-2 expression                                              | (He et al., 2013).                                                          |
|             | Suppressed the epidermal growth factor receptor (EFGR) degradation          |                                                                             |
|             | Inhibited downstream Akt/mTORC1                                             |                                                                             |
| Skin cancer | Suppressed the proliferation, migration, invasive and adhesive              | (Li et al., 2019)                                                           |
|             | properties                                                                  |                                                                             |
|             | Inhibited the EMT and angiogenesis processes                                |                                                                             |
|             | Lowered the enzymatic activities of MMP-2 and MMP-9                        |                                                                             |
|             | Suppressed the abnormal PI3K/Akt and ERK signaling pathways                 |                                                                             |
| Colon cancer | Induced cell proliferation, migration and invasion                          | (Zhou and Ma, 2019).                                                       |
|             | Altered expressions of AKT, p21, P13K, MMP-2 and -9, and                    |                                                                             |
|             | phosphorylated-AKT                                                          |                                                                             |
|             | Suppressed the cell viability and activated the SHH signaling                |                                                                             |
| Liver cancer | Enhanced the levels of p-AMPK                                                | (Wang et al., 2019).                                                        |
|             | Up-regulated E-cadherin linked with LKB1                                    | (Li et al., 2019; Kebebe et al., 2019; Yin et al., 2014).                   |
|             | Induced E-cadherin, and down regulated the ZEB1                             |                                                                             |
| Lung cancer  | Suppressed the antioxidant enzyme TrxR1 reduction activity                  | (Duan et al., 2014).                                                       |
|             | Enhanced intracellular ROS level, C/EBP-homologous protein, expression     | (Zhu et al., 2019).                                                        |
|             | levels of glucose-regulated protein (GRP)78                                 |                                                                             |
|             | Activated transcription factor 6 and caspase 12                              |                                                                             |
|             | Activated the inositol-requiring enzyme 1alpha and phosphorylation         |                                                                             |
|             | levels of protein kinase R-like ER kinase                                   |                                                                             |
|             | Inhibited cell growth, induced cells autophagy                              | (Ye et al., 2018).                                                         |
|             | Caused upregulation of Beclin 1, and transformation of autophagosome         |                                                                             |
|             | markers such as LC31 to LC2II                                               |                                                                             |
Lowered the expression levels of Jagged1, Jagged2, DLL1, DLL3, DLL4, PK3K and Bcl2
Enhanced expression level of active caspase3
Inhibited Akt phosphorylation, and NICD nuclear translocation
Induced the expression of Beclin-1 and LC3-II proteins

(Zhu et al., 2018)

Destructed mitochondrial membrane
Down regulated the TOP2A, ALDOA, and ATG4B
Up-regulated DUSP1, DDIT3, and DUSP5
Enhanced the efficacy rate
Inhibited cell proliferation
Down regulated the levels of nuclear factor-κB (NF-κB), BCL-2, phosphatidylinositol3-kinase (PI3K), c-myc, and phosphorylation of serine-threonine kinase (p-AKT)

(Youns et al., 2018)

(Feng et al., 2018).

(Chen et al., 2015).

Table 1: Role of Gambogic acid in different cancers

| Cancer          | Effect                                                                                      |
|-----------------|---------------------------------------------------------------------------------------------|
| Pancreatic      | Lowered the expression levels of Jagged1, Jagged2, DLL1, DLL3, DLL4, PK3K and Bcl2         |
|                 | Enhanced expression level of active caspase3                                              |
|                 | Inhibited Akt phosphorylation, and NICD nuclear translocation                             |
|                 | Induced the expression of Beclin-1 and LC3-II proteins                                     |
|                 | (Zhu et al., 2018)                                                                         |
| Blood cancer    | Destructed mitochondrial membrane                                                           |
|                 | Down regulated the TOP2A, ALDOA, and ATG4B                                                |
|                 | Up-regulated DUSP1, DDIT3, and DUSP5                                                       |
|                 | Enhanced the efficacy rate                                                                 |
|                 | Inhibited cell proliferation                                                               |
|                 | Down regulated the levels of nuclear factor-κB (NF-κB), BCL-2, phosphatidylinositol3-kinase |
|                 | (PI3K), c-myc, and phosphorylation of serine-threonine kinase (p-AKT)                     |
|                 | (Youns et al., 2018)                                                                       |

Colon cancer

Zhou and Ma recently reported anti-tumor activity of GA in humans against colon cancer cells SW620 cells at 10 µg/ml, 50 µg/ml and 100 µg/mL with 10 µg/ml 5-fluorouracil. They concluded that GA induced cell proliferation, migration and invasion of cancer cells dose dependently and it also altered expressions of AKT, p21, P13K, MMP-2 &-9, and phosphorylated-AKT (Zhou and Ma, 2019). In another animal study conducted by Wang and the colleagues, explored that GA enhanced the apoptosis in cancer cells along with suppression of cell viability and activated the sonic hedgehog (SHH) signaling (Wang et al., 2019). Different doses of GA (5,10 and 20mg/kg in saline) against HT-29 cells of BALB/c nude mice twice a week inhibited cell proliferation through apoptosis and mediated by intrinsic (mitochondrial) and extrinsic (death receptors) pathways in a dose dependent manner (Huang et al., 2015a). Nano-encapsulated GA studied by Huang and co-workers found that GA against HT-29 human colon cancer xenograft mouse models produced anti-cancer role by decreasing cell division and proliferation and inducing apoptosis (Huang et al., 2015b). GA in LOVO cells also significantly induced apoptosis and inhibited the proliferation in a dose- and time-dependent manner. It has been reported that magnetic nano-particles of GA containing Fe₃O₄ have more ability to increase transcription of cytochrome c, caspase 9, and caspase 3 genes and decreased the transcription of phosphatidylinositol 3-kinase, Akt, and Bad genes as compare to the gambogic acid without magnetic nanoparticles containing Fe₃O₄. (Fang et al., 2012;Wei et al., 2018). Combined form of GA with 5-fluorouracil (5-FU) treatment against HCT116 human CRC cells, SW480 and human intestinal epithelial cells (IECs) significantly
inhibited growth of cancer cells, apoptosis and decreased P53, survivin and TS mRNA and protein levels as well as increased P53 protein levels (Wei et al., 2017; Zhang et al., 2017). Likewise, in both 5-fluorouracil (5-FU) sensitive and 5-FU resistant colorectal cancer cells, GA suppressed the proliferation and induced apoptosis via activating JNK signaling pathway (Wen et al., 2015; Zhang et al., 2014).

Liver cancer

Liver kinase-B1 (LKB1) have tumor suppressing action by regulating the cell growth, survival, metabolism, and polarity. GA has been found to enhance the levels of p-AMPK via up-regulating E-cadherin linked with LKB1 in vivo and in vitro studies. Additionally, GA also induces E-cadherin, and down regulates the ZEB1 (Li et al., 2019; Kebebe et al., 2019; Yin et al., 2014). The TrxR1 (thioredoxin reductase (TrxR) isoenzymes,1) in nucleus or cytosol and TrxR2 in mitochondria are important mammalian selenocysteine (Sec)-containing flavor-enzymes with a C-terminal-Gly-Cys-Sec-Gly active site are frequently over-expressed in human tumor and GA reduced expression of these malignant cells which reverse tumor growth. Duan et al. in 2014 studied that GA against hepatocellular carcinoma SMMC-7721 cells targeted mammalian selenocysteine (Sec)-containing flavoenzymes residue in the antioxidant enzyme TrxR1 to inhibit its Trx-reduction activity which collapsed intracellular redox balance and leads to the accumulation of reactive oxygen species. Furthermore, over-expression of TrxR1 in cells decreased GA induced cytotoxicity while knockdown of TrxR1 sensitizes cells to GA (Duan et al., 2014). In hepato-cellular carcinoma liver cell line (HepG2) cells, GA and cadmium-tellurium quantum dots enhanced accumulation of drugs and decreased cell proliferation. Nano-composition of GA -CdTe enhanced drug action of gambogic acid molecules against HepG2 cells and induced cell cycle arrest at G2/M phase along with cell apoptosis (Xu et al., 2013). GA inhibited Huh7 and Hep3B growth in same apoptotic pathways and IC₅₀ investigated for both cell lines were 1.8 and 2.2µM, respectively (Lee and Ho, 2013).

GA has antitumor effects against hepatocellular carcinoma through multiple mechanisms such as (i) improvement in liver functions, (ii) reduction of cell lesions in hepatic tissue, (iii) suppression of expression of CD105, and (iv) prolongation of survival time, respectively (Yan et al., 2017). In hepatocellular carcinoma (HCC) tissues, gambogic acid inhibited UNC119 (uncoordinated 119 or retinal protein 4) by inhibiting cell proliferation, produced cell cycle arrest at G0 and G1 phases.
against Hep3B cells, down regulated cell cycle related proteins including cyclin A, E, D1 and cyclin-dependent kinase 2, 4, 6. GA also down-streamed UNC119 and decreased glycogen synthase kinase 3β (Gsk3β)/β-catenin signaling (Wu et al., 2016).

A study reported by Deschatrette et al. (2013) showed that gambogic acid i has been found to lower the thymidylate synthetase mRNA levels and level of thymidylate synthetase mRNA n a gastric carcinoma cell line. A contradictory relationship was observed in hepatoma cells between DHFR expression and resistance to GA with a dihydrofolate reductase (DHFR) gene amplification and cells transfected with an inducible DHFR transgene. In vitro, GA suppressed the DHFR activity, and lowered the affinity of the enzyme for dihydrofolate (Deschatrette et al., 2013). In an in vitro study human hepatocellular carcinoma (HCC) cells cytotoxicity i.e. Bel-7402 cells and HepG2 cells by GA inhibited cell proliferation, and induced apoptosis. GA might become a promising therapeutic agent for the treatment human HCC (He et al., 2012). There are multiple pathways involved in HepG2 cells through the application of GA in a concentration-dependent manner such as inhibition of viability of HepG2 cells, reduction in volume, condensation of chromatin, enhancement in apoptotic cells percentage, alteration of expression level of several apoptosis-associated proteins, Bcl-2, p53, Bax and pro-caspase-3. In addition, GA exhibited apoptosis in HepG2 cells, probably via intrinsic mitochondrial pathways (Mu et al., 2010).

A study reported by Nie and the colleagues determined that GA destroyed mitochondrial membrane potential and induced accumulation of ROS dose dependently in human hepatoma SMMC-7721 cells with the release of cytochrome c and apoptosis-inducing factor from mitochondria to cytosol, which induced apoptosis in the cells. Furthermore, it was also concluded that GA improved the level of the phosphorylation of c-jun-N-terminal protein kinase (JNK) and p38 (Nie et al., 2009). In china gamburg is developed and used as an injectable drug against cancer treatment. Wang et al. in 2009 investigated inhibition ratio and IC₅₀ values of two derivatives of gamburg (gambogic acid and gambogenic acid) against HCC and revealed its inhibitory effects on cell proliferation. Both compounds up regulated the guanine nucleotide-binding protein beta subunit 1, and cyclin-dependent kinase 4 inhibitor A whereas down regulated the stathmin 1 (STMN1) and 14-3-3 protein sigma expressions. Furthermore, overexpression of STMN1 in HCC cells decreased their sensitivity, whilst small interfering RNAs targeting STMN1 enhanced their sensitivity to GA and gambogenic acid (Wang et al., 2009).
Lung cancer

Different doses of GA as 0, 0.5, and 1.0 μmol/L were treated with non-small cell lung cancer A549 cells in a concentration-dependent manner and reduced the cell viability, enhancement intracellular ROS level, C/EBP-homologous protein, expression levels of glucose-regulated protein (GRP)78, activated transcription factor 6 and caspase 12, as well as the inositol-requiring enzyme 1 alpha and phosphorylation levels of protein kinase R-like ER kinase (Zhu et al., 2019). A study conducted by Ye and co-workers, they explored that gambogic acid in non-small cell lung cancer (NSCLC) NCI-H441 of xenografts inhibited cell growth, induced cells autophagy, caused upregulation of Beclin 1, and transformation of autophagosome markers such as LC31 to LC2II. Moreover, GA induced autophagy through an ROS-dependent pathway (Ye et al., 2018). In human non-small cell lung cancer (NSCLC) cells (A549 and SPC-A1 cells), different doses of GA at 0, 0.5, 0.75, and 1.0 μmol/l showed momentous inhibition in cell viability, enhancement in cell apoptosis, reduction in the expression levels of Jagged1, Jagged2, DLL1, DLL3, DLL4, PK3K and Bcl2, increment in expression level of active caspase-3, and inhibited Akt phosphorylation, and NICD nuclear translocation (Zhu et al., 2018). Combined form of GA with cisplatin produced a significant inhibitory effect on A549 and NCI-H460 cells. They significantly increased autophagy, and inhibited the activation of S6, mTOR and Akt. In addition, gambogic acid combined with rapamycin induced more cell death as compare to GA anti-tumor action alone (Zhao et al., 2017).

Pancreatic cancer

In a recent study, pancreatic cancer cell lines treated with gambogic acid and showed induction of the expression of Beclin-1 and LC3-II proteins and enhancement in the formation of both acidic vesicular organelles and autophagosomes, as well as autophagic flux. Moreover, increased cytotoxicity of gambogic acid was linked with inhibition of autophagy by chloroquine. In addition, promotion of ROS production and destruction of mitochondrial membrane which is linked with autophagy activation were reported after GA treatment (Wang et al., 2019).

Pancreatic cancer cells treated with GA showed growth inhibition, down-regulation of TOP2A, ALDOA, and ATG4B, and up-regulation of DUSP1, DDIT3, and DUSP5, respectively (Youns et al., 2018). Similarly, there are multiple mechanisms through gambogic acid treatment against pancreatic cancer cell lines such as suppression of the growth, and induction of cell cycle arrest at
S-phase and apoptosis. In PANC-1 and BxPC-3 cells, synergistic action of GA with gemcitabine was reported. GA activated apoptosis induced by gemcitabine via increasing expression of cleaved caspase-3 and 9, cleaved-PARP and Bax and reduced Bcl-2 expressions. Reduction in the expression of the ribonucleotide reductase subunit-M2 (RRM2) protein and mRNA were reported after GA treatment which is linked with resistance to gemcitabine via extracellular signal regulated kinase (ERK)/E2F1 signaling pathway inhibition. Furthermore, both compounds in the xenograft cancer model significantly repressed tumor growth and down regulated the p-ERK, E2F1, and RRM2 (Xia et al., 2017; Saeed et al., 2014). In Panc-1 pancreatic cancer cells, gambogic acid-loaded magnetic Fe₃O₄ nanoparticles significantly inhibited the ETS1-mediated proliferation and migration of cells, lowered the expression of ETS1, as well as it down-streamed target genes cyclin D1, u-PA and VEGF (Wang et al., 2012). In another study, encapsulation of GA with magnetic Fe₃O₄ nanoparticles improved anticancer potential via induction of apoptosis, increasing of Bax, caspase 3 and caspase 9, decreasing protein expressions of Bcl-2 (Wang et al., 2011).

**Blood cancer**

GA increased production of ROS in hematopoietic malignant cell lines (Ortiz-Sánchez et al., 2009). In acute myeloid leukemia cell lines of experimental subjects such as Jurket, HL-60 and MV4-11 cells, nano-emulsion encapsulated GA enhanced the efficacy rate in both in vivo and in vitro studies (Feng et al., 2018). Administration of gambogic acid more than 0.5 μM to human leukemia cell line K562 dose-dependently induced apoptosis, inhibited cell proliferation, and down regulated the levels of nuclear factor-κB (NF-κB), BCL-2, phosphatidylinositol3-kinase (PI3K), c-myc, and phosphorylation of serine-threonine kinase (p-AKT) (Chen et al., 2015). Cancer cells lines such as U937 and HL-60 were treated with gambogic acid and caused suppression in cell growth, promotion in differentiation, and upregulation of p21waf1/cip1 expression, respectively (Chen et al., 2014). Gambogenic acid also reported as significant anti-cancer agent against different chronic myelogenous leukemia cell lines i.e. KBM5-T315I, KBM5, and K562 of nude mice via inducing apoptosis, cell proliferation inhibition and suppression of the growth of imatinib-resistant Bcr-Abl-T315I (Shi et al., 2014). In another study reported by Li and colleagues, they found that down regulation in the expression of SRC-3 along with inhibition of Akt kinase and its down-streamed targets p70, S6 kinase 1(S6K1) and glycogen synthase kinase 3 beta (GSK3beta) were observed after gambogic acid treatment against myelogenous leukemia cell
line K562 cells in humans. In addition, these changes also affected the expressions of apoptosis (gene Bcl-2) in K562 cells (Li et al., 2009).

In another study conducted by Tao and colleagues, they explored that N-(2-ethoxyethyl) gambogamide (NG-18), a derivative of gambogenic acid in against leukemia (HL-60 cells) also effectively suppressed the culture human tumor cells proliferation, induced apoptosis whereas tumor apoptosis is linked with up-regulated pro-apoptotic Bcl-2 family member Bax, and downregulated anti-apoptotic protein Bcl-2 (Tao et al., 2007). In multidrug-resistant lymphoma Raji/DNR cells, gambogenic acid in a dose-dependent manner down-regulated expression of P-glycoprotein (Zhou et al., 2016). A study reported by Zhao and co-workers showed that gambogenic acid exhibited mechanisms such as suppression of cell growth, induction of apoptosis, caused over expression of SRC-3, suppressed cyclin D3 and Bcl-2, Bcl-6 expressions, modulated down-stream gene expression, and induced the de-acetylation of histone H3 at lysine 9 and lysine 27 in B-cells non-Hodgkin lymphoma (NHL) (Zhao et al., 2016). Likewise, a group of researchers unveiled that GA in vitro and in vivo trials has potent anticancer effect on B-cell lymphoma (DLBCL) cells such as GCB- and ABC-DLBCL cells through inducing apoptosis, and inhibiting cells growth (Shi et al., 2015). In a study against Jeko-1 human mental cells lymphoma cell apoptosis, GA inhibited cell growth as time and dose-dependently. It is involved in suppressing ratio of Bax and Bcl-2 with arresting cell cycle and decreasing mitochondrial membrane potential and activating caspase-3,-8,-9 (Xu et al., 2013).

Conclusion

Researchers conducted extensive work on natural products to explore their role as an anticancer agent in last decades. Among these products, GA has attained popularity as a promising novel antitumor agent and has ability to inhibit proliferation and induce apoptosis. It inhibits the cell viability, enhances cell apoptosis, reduces the expression levels of PK3K, DLL1, DLL3, DLL4, Jagged1, Jagged2, and Bcl2, increases the active caspase-3 level, and inhibits the Akt phosphorylation in human malignancies. Extensive work is required to further investigate its toxicity and interaction with other chemotherapeutic agents in clinical setting.

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