Efficacy of Turmeric Rhizome (*Curcuma longa*) and Moringa Leaf (*Moringa oleifera*) Extract in Treatment against Fungi Associated with Maize Seeds

W. C. John¹*, T. A. Ihum², O. Olusolape¹ and N. Janfa¹

¹Federal College of Forestry, Jos, Plateau State, Nigeria.  
²Nigeria Stored Products Research Institute, Ilorin, Kwara State, Nigeria.

Authors’ contributions

This work was carried out in collaboration between all authors. Author WCJ designed the study, performed the statistical analysis, wrote the protocol, and wrote the first draft of the manuscript. Authors TAI and OO manage the analyses of the study. Author NJ managed the literature searches. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/APRJ/2018/v1i226270

This research was focused on the use of different concentration of aqueous extract of Moringa leaf and Turmeric rhizome as seed treatment agents against fungi associated with maize seeds. The study was designed using 2 x 3 factorial in completely randomised design. This research was carried out in Biology Laboratory of Federal College of Forestry, Jos from April to June 2018. Untreated maize seeds were collected from farmers in three different areas of Mazah village in Jos North LG. Maize seeds were tested for the presence of fungi by culturing on Petri dish having moist filter paper. Aqueous extract of Turmeric rhizome and Moringa leaf at 25, 50, and 75 ml concentration were used as seed treatment agent against fungi. The seeds were soaked in prepared extracts for 1 hour and then cultured on petri dish having moist filter paper, control samples were soaked in sterile water. Infested seeds were counted manually after twenty days. Results were analysed using one-way analysis of variance at 95% confidence level. *Penicillium sp.*

*Corresponding author: E-mail: wchinaka@yahoo.com;
Fusarium solani, Aspergillus niger and Aspergillus flavus were isolated from the untreated maize seed. *Fusarium solani* gave the highest percentage occurrence of 35.48. 50 ml concentration of Turmeric rhizome has an antifungal mean value of 3.67 after 20 days. The control (untreated) showed the antifungal mean value of 6.67 after 20 days. The results obtained revealed significance between the treatment and the control. The results indicate Turmeric rhizome and Moringa leaf extracts could be used in seed dressing against maize infection caused by fungi.

Keywords: Turmeric; moringa; maize; fusarium; jos

1. INTRODUCTION

Maize (*Zea mays* L.) is one of the most important cereal crops in the world and rank third next to wheat and rice [1]. From the recent past, it has shown to be good cereal crop due to its low cost of production, wide adaptability and diversified used. Maize suffers from several fungal diseases in which are seed borne in nature [2]. The incidence of seed-borne pathogen cause germination failure as well as a reduction in seedling vigour which ultimately reduces the yield. Control of the seed borne fungal pathogens by chemical fungicide is costly and hazardous for the environment. Therefore, the use of plant material is important in reducing the prevalence of the seed borne pathogens associated with the maize seed and to enhance percentage of maize seed germination and seedling vigour.

Maize is an important cereal in many developed and developing countries of the world. It is widely used for animal feed and industrial raw materials such as starches, acids and alcohols in the developed countries, whereas it serves as food in developing countries. Recently there has been interest in using maize for the production of ethanol in the United States and China, as a substitute for petroleum-based fuels [3]. In Nigeria, maize is the most popular food crops in the domestic market. It is a good substitute for other food crops like sorghum, millet, plantain, yam and cocoyam when they are in short supply. It is the main feedstuff for poultry and other livestock [4].

It is estimated that in 2012, the total world production of maize was 875,226,630 tons, with the United States, China, and Brazil harvesting 31%, 24%, and 8% of the total production of maize, respectively [5]. Despite its importance, maize has many production constraints which prevent farmers from getting maximum yield. Drought, fire, flood, poor soil fertility, high labour cost, transportation problems and poor agronomic practices are some of the production constraints of maize [6]. Pest and diseases are also formed some of the problems that characterise maize production in the world. The diseases of maize seeds include; seed rot (*Fusarium moniliforme, Fusarium oxysporum, Penicillium species*) seedling blight (*Aspergillus species, Penicilum species*) *Biperis* leaf spot (*Biperis maydis*), *Curvularia* leaf spot (*Curvularia lunada*) etc. The study aims to determine the effect of different concentration of Turmeric rhizome and Moringa leaf extracts in maize seed treatment against fungal pathogens associated with maize seeds.

2. MATERIALS AND METHODS

2.1 Study Area

The experiment was carried out in Biology Laboratory of Federal College of Forestry Jos, North Local Government Area of Plateau State. Jos is located between latitude 7-11° North and longitude 7 -8° East. Temperature ranges between 10-32°C and main annual rainfall is about 1340 mm, with an average elevation of 1200 mm above sea level.

2.2 Collection of Samples

Fresh disease-free leaves of *Moringa oleifera* and rhizomes of Tumeric (*Curcuma longa*) 100 g each were purchased from Farin gada market of Jos North Local Government area, Plateau State. 500 seeds of healthy local maize (Waxy corn (F1) colourful) previously stored were collected from farmers at three different areas (Alibah, Anagohom and Anabor) in Mazah village of Jos North Local Government Area at different locations. Maize variety Waxy corn (F1) colourful was used for the research due to its availability and farmers choice in study area [7].

2.3 Determination of Fungi Presence on Collected Maize Seed

Three pieces of 9 cm filter paper (Whatman No 1) were soaked in distilled water and place at the
bottom of the Petri dish. Untreated healthy maize seeds collected from farmers at three different areas in Mazah village were used for fungi testing. This was done using Blotter method, maize seeds collected from different locations were randomly selected and 20 seeds were placed in each Petri dish containing moist filter paper base on the number of the areas and the seeds were incubated at room temperature for 10 days. Potato Dextrose Agar (PDA) was prepared according to the manufacturer's instruction. 80 g of Gentamycin was added to the prepared 500 ml of agar PDA media to inhibit bacteria growth. Fungi developed from incubated maize seeds were inoculated in the solidified potato Dextrose agar using inoculating needle flamed over a burner flame, the solidified inoculated plates were kept at room temperature (28± 2°C) until visible growth appeared on the plates. The fungal colonies that grew from the incubated plates were further sub-culture into fresh medium until pure cultures were obtained. The percentage occurrence of the fungi isolated was determined using the following formula:

\[
\text{Percentage occurrence} = \frac{\text{Number of isolates}}{\text{Number of total isolates}} \times 100
\]  

2.4 Identification of Fungi Isolates

Macroscopic and Microscopic examination were used to determine the morphological characteristic of the fungi isolates. For macroscopic identification, colony characteristics such as appearance and change in colour were observed on the Petri plates. For microscopic examination, the sterile inoculating needle was used to pick a little portion of five-day-old isolate and placed on a sterile glass slide, the slide was stained with lactophenol cotton blue, mixed and covered with a slip. The slide was then viewed under the microscope, using x10 and x40 magnification. Shapes of the conidia and conidiophores were taken note of. These features were matched with standards described by Barnett and Hunter [8] and Booth [9] and identified by the help of an expert.

2.5 Preparation of Plant Extract (Turmeric Rhizome and Moringa Leaf)

Collected fresh plants were detached and washed first in tap water than in distilled water and air dried at room temperature. 100 g of fresh sample was chopped and then crushed in a surface sterilised mortar and pestle by adding 100 ml distilled water (1:1 w/v). The extract was filtered through two layers of muslin cloth and was used as a stock solution [10].

2.6 Determination of Extract Concentrations

The plant extracts were used at 25, 50 and 75 % concentrations, for which 25, 50 and 75 ml of stock solution was mixed with 75, 50 and 25 ml of sterilised distilled water respectively.

2.7 Seed Treatment Using Turmeric rhizome and Moringa Leaf Extracts

Twenty healthy untreated maize seeds were treated using sterilised aqueous extracts of the plant (Turmeric and Moringa) separately by dipping the maize seeds into different concentrations (25, 50 and 75 ml) of the aqueous extracts for 1 hour. For the control, twenty healthy untreated seeds dipped in sterilised distilled water for 1 hour served as the control treatment.

2.8 Antifungal Efficacy of Turmeric Rhizome and Moringa Extract

After treating the seeds, the seeds were placed in sterilised Petri dishes containing four layers of blotting paper soaked in sterilised distilled water. Each Petri dish contained 20 seeds and the Petri dishes were replicated three times. The Petri dishes were then kept in an incubation chamber at room temperature for 10 days and infected seeds were counted manually after 10, 15, 20 days after sowing [10].

2.9 Treatment and Experimental Design

The experimental layout used was 2 x 3 factorial in complete random design (CRD) whereby the three treatments were replicated three times (0, 25, 50 and 75 ml).

\[
\text{Treatment combinations} = 2 \times 4 = 8 \\
\text{Replications} = 3 \\
\text{Total plots} = 3 \times 8 = 24.
\]

2.10 Data Analysis

Data analysis was carried out using turkey pairwise comparison one way analysis of variance (ANOVA) and T-test to compare means. Means separation was determined using Duncan multiple range test. Significance difference was determined at 95% confidence.
3. RESULTS

3.1 Isolation and Identification of Fungi

A total number of sixty-two fungal species were isolated. The result obtained showed Anagohom with the highest fungi occurrence of 45.16% compared to Alibah and Anabor which has 29.03 and 25.16% respectively (Table 1). The percentage distribution results showed a significant difference at \( p = 0.05 \). The fungi isolated were identified macroscopically and microscopically as *Penicillium sp*, *Fusarium sp*, *Aspergillus niger* and *Aspergillus flavus*. *Fusarium solani* (based on the characteristics presented in Table 2) gave the highest percentage occurrence of 35.48 while the *Penicillium sp* showed the lowest percentage occurrence of 11.29. The result in Table 4 showed a significant difference in fungal percentage occurrence.

3.2 Antifungal Efficacy of Moringa Leaf Extracts

Table 5 below showed the mean antifungal effect of Moringa after 10, 15 and 20 days at a different concentration of 25, 50 and 75 ml. The control (untreated) maize seeds have the mean value of 4.33 after 10 days, 5.00 after 15 days and 6.37 after 20 day. The 25 ml concentration showed mean values of 2.33 after 10 days and 2.6 after 15 days and 3.00 after 20 days. The 75 ml concentration indicated mean values of 1.33 after 10 days, 1.7 after 15 days and 2.00 after 20 days. The result also showed that Moringa leaf extract at a higher concentration of 75 ml has the highest effect of 2.00 mean value at 20 days interval. The result obtained showed the effect of Moringa extract antifungal activities is directly proportional to the extract concentration and days of exposure.

3.3 Antifungal Efficacy of Turmeric Rhizome Extract

The table 6 below showed the means effect of Turmeric rhizome extracts at 10, 15, and 20 days of the experiment at a different concentration of 25, 50 and 75 ml. The control (untreated) maize seeds have the mean value of 4.33 after 10 day, 5.00 after 15 days, 6.67 after 20 days. The 25 ml concentration showed the mean values of 2.00 after 10 days, 2.33 after 15 days and 3.00 after 20 days. The 50 ml concentration has the mean value of 2.33 after 10 days, 3.00 after 15 days and 3.67 after 20 days. The 75 ml concentration showed a mean value of 3.67 after 10 days, 4.33 after 15 days, and 5.67 after 20 days. The result shows that Turmeric rhizome at smaller concentration is more effective in control of associated with maize seeds as it showed least mean value of 2.00, 2.33 and 3.00 of infected seeds after 10, 15 and 20 days which is followed by 50 ml concentration. The control has the highest mean value of infected maize seeds, which shows that Turmeric rhizome is effective in controlling fungal pathogen associated with maize seeds.

### Table 1. Percentage distribution of fungi species in Alibah, Anabor, Anagohom

| Area     | No of fungi occurrence | Percentage distribution |
|----------|------------------------|-------------------------|
| Alibah   | 18                     | 29.03\(^{a}\)           |
| Anabor   | 16                     | 25.81\(^{a}\)           |
| Anagohom | 28                     | 45.16\(^{b}\)           |
| SE±      | -                      | 5.64                    |
| LSD      | -                      | *                       |
| Total    | 62                     | 100                     |

\( a, b, c \) = means separation indicating level of significance; Means within a column followed by the same letters are not significantly different \( P = 0.05 \) using Duncan Multiple Range Test. * = Significant at 95% level of probability. LSD = Least significant difference.

### Table 2. Fungi Features/ Characteristics

| S/NO | FUNGI             | FEATURES/CHARACTERISTICS                                                                 |
|------|-------------------|-------------------------------------------------------------------------------------------|
| 1    | *Aspergillus flavus* | It consists of a dense of yellow-green conidiophores and conidials with a head typically radiated, and splitting into several loose columns. Conidiophores become dark yellow-green, hyaline and roughened. |
| 2    | *Penicillium species* | Consist of a dense felt of conidiophores. Appeared leathery, blue green and yellow to orange. |
| 3    | *Fusarium solani* | Cottony, whitish, letter becoming yellow, pink-red. Reverse in yellow to brownish shades. |
| 4    | *Aspergillus niger* | It consists of a compact white basal felt with a dense layer at dark brown to black conidiophores, conidial head radiate ending to split into loose columns with time. |
Table 3. Morphological view of fungi isolates

| Appearance on PDA | Photomicrograph | Probable Isolates |
|-------------------|-----------------|-------------------|
| ![Aspergillus flavus](image1) | ![Aspergillus flavus](image2) | Aspergillus flavus |
| ![Penicillum sp](image3) | ![Penicillum sp](image4) | Penicillum sp |
| ![Fusarium Solani](image5) | ![Fusarium Solani](image6) | Fusarium Solani |
| ![Aspergillus niger](image7) | ![Aspergillus niger](image8) | Aspergillus niger |

Table 4. Percentage occurrence of fungal species

| Fungi isolates      | Number of occurrences | Percentage occurrence |
|---------------------|-----------------------|-----------------------|
| Aspergillus niger   | 19                    | 30.65%                |
| Fusarium species    | 22                    | 35.48%                |
| Aspergillus flavus  | 14                    | 22.58%                |
| Penicillum species  | 7                     | 11.29%                |
| SE±                 | -                     | 5.86%                 |
| LSD                 | -                     | *                     |
| Total               | 62                    | 100                   |

*a, b, c* = means separation indicating level of significance

Means within a column followed by the same letters are not significantly different (*P* = 0.05) using Duncan Multiple Range Test.

* = Significant at 95% level of probability.

LSD = Least significant difference.
4. DISCUSSION

Seed is the most important unit of crop production and its health plays an important role in agriculture, which determines the plant population and final yield. One of the major constraints that deteriorate the seed quality is the seed-borne fungi present inside or on the surface of seeds [7]. Leaf extracts of many higher plants have been reported to possess antifungal activity under laboratory trials [10]. Farmers are faced with challenges of fungal infestation which has led to great economic losses. Botanical seed dressing agents is gradually gaining attention due to the problems associated with the use of synthetic pesticides, synthetic pesticides are toxic, not friendly to the environment and also not affordable by most farmers especially the local farmers that own small farms due to its expensive nature [11]. Plant extracts of many plant parts have to possess antifungal activities in vitro [12].

The finding of this research revealed that fungal pathogen associated with stored maize seeds include; Penicillium sp, Fusarium solani, Aspergillus niger and Aspergillus flavus, which is in line with the findings of Nirmal et al. [13] which says Maize seeds diseases are produced by many species of fungi such as Aspergillus, Fusarium, Penicillium, Claviceps and Alternaria genera that are associated with maize seeds. The result of percentage occurrence of fungi isolates showed that Fusarium species has the highest percentage occurrence of 35.48 %, the result agrees with the work conducted by Forsberg et al. [14] which state that Fusarium occurs throughout cultivation period of maize. In the present work antifungal activity of two plants extracts, Moringa leaf and Turmeric rhizome were assessed at the concentration of 25, 50 and 75 ml against fungal growth by blotter technique. Similar investigation on the antifungal activity of plant extracts against seed-borne fungi has been reported by many researchers [15-17]. Between these, Turmeric rhizome extract was found more effective in inhibition of fungal growth than Moringa leaf at a lower concentration. Antifungal activity of the extract at different concentrations are found to be effective [18,19] this could be due to the activities of essential oils, a mixture of terpenoids, aromatic phenols and many other compounds such as 1, 8-Cineole which has proved to have fungicidal properties [20].

The result obtained using Turmeric extract at 25 ml concentration showed means value of 3.00 infestation which is the least compared to 50 ml and 75 ml concentration. Turmeric at small concentration after 20 days of the experiment is more effective than the other two concentrations (50 ml and 75 ml). This result agrees with the finding of Abiamere et al. [21] which evaluated the efficacy of Moringa, Ginger and Chromolena odorata plant as a treatment agent against seed-borne fungi associated with Cowpea. The 75 ml concentration after 20 days of seed incubation has high means value infestation of 5.67, this shows that Turmeric at higher concentration is less effective in control of fungal pathogen associated with maize seed, this result is in line with the findings of Lee et al. [22] which says fungicidal activities of Turmeric has been found effective in controlling certain agricultural and animal pest or pathogens due to the presence of a variety and bioactive constituents that interfere with fungi behaviour and growth. This present study has therefore shown that plant extract of Moringa and Turmeric can be used as a

Table 5. The antifungal mean effect of moringa leaf extract

| Concentration (ml) | Number of Days |
|--------------------|----------------|
|                    | 10  | 15  | 20  |
| Control            | 4.33  | 5.00  | 6.67  |
| 25                 | 2.33  | 2.67  | 3.00  |
| 50                 | 2.00  | 2.33  | 2.67  |
| 75                 | 1.33  | 1.67  | 2.00  |
| SE±                | 0.41  | 0.58  | 0.53  |
| LSD                | *    | *    | *    |

*a, b, c = means separation indicating level of significance; Means within a column followed by the same letters are not significantly different (P = 0.05) using Duncan Multiple Range Test.

* = Significant at 95% level of probability.

LSD = Least significant difference.

Table 6. The antifungal mean effect of turmeric rhizome extracts

| Concentration (ml) | Number of Days |
|--------------------|----------------|
|                    | 10  | 15  | 20  |
| Control            | 4.33  | 5.00  | 6.67  |
| 25                 | 2.00  | 2.33  | 3.00  |
| 50                 | 2.33  | 3.00  | 3.67  |
| 75                 | 3.67  | 4.33  | 5.67  |
| SE±                | 0.71  | 0.67  | 0.71  |
| LSD                | *    | *    | *    |

*a, b and c = means separation indicating the level of significance; Means within a column followed by the same letters are not significantly different (P = 0.05) using Duncan Multiple Range Test.

* = Significant at 95% level of probability.

LSD = Least significant difference.
fungicidal seed treatment for the control of seed-borne fungi of maize seeds.

5. CONCLUSION

Based on data from the present study, Moringa leaf and Turmeric rhizome extracts at different concentrations showed a significant difference at 95% confidence level. The result showed that Turmeric (concentration 25 and 50 ml) and Moringa (concentration 75 ml) gave a mean antifungal effect of 2.00, 2.33 and 1.33 respectively. This showed the extracts can be used in seed dressing against fungal pathogens associated with maize seeds.

6. RECOMMENDATION

Further research should be carried out on these same botanicals by combining the two together to determine their synergic effect.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Abida Y, Tabassum F, Zaman S, Chhabi SB, Islam N. Biological screening of Curcuma longa L. for insecticidal and repellent potentials against Tribolium castaneum (Herbst) Adults. Univ J Zool Rajshahi Univ. 2010;28:69-71.
2. Sharma RR, Singh D, Singh R. Biological control of postharvest diseases of fruits and vegetables by microbial antagonists. A review Biology Control. 2009;50:205-221.
3. Amein T, Wright SAI, Wikstrom M, Koch E, Schmitt A, Stephan D, Jahn M, Tinivella F, Gullino ML, Forsberg G, Werner S, Jan-van-der W and Groot SPC. Evaluation of non-chemical seed treatment methods for control of Alternaria brassicicola on cabbage seeds. Journal of Plant Disease Protection. 2011;118(6):214-221.
4. Elwakil MA. Use of antioxidant hydroquinone in the control of seed-borne fungi of peanut with special reference to the production of good quality seed. Plant Pathology Journal. 2003;2:75-79.
5. FAO. FAOSTAT, Production; 2012. Available:http://faostat.fao.org/site/567/Default.aspx?PageID (Cited November 2, 2018)
6. Al-Sammarai G, Singh H, Syarhahipl M. Evaluating eco-friendly botanicals of alternatives to synthetic fungicides. Annals of Agricultural and Environmental Medicine. 2012;9:673-676.
7. Bhattacharjee R, Dey U. Overview of fungal and bacterial biopesticides to control plant pathogens and diseases. African Journal of Microbiology Research. 2014;8(7):1749-1769.
8. Barnett HL, Hinter BB. Illustration genera of imperfect fungi, 3rd edition, Burgess publing co. Minnesoth, usa. 1972;273.
9. Broth C. The genus fusarium, Survey, UK: Cmi, Kew. 1971;237-238.
10. Sigrist MS, Pinheiro JB, Azevedo Filho JA, Zucchi Mi. Genetic diversity of turmeric germplasm (curcuma longa; zingiberaceae) identified by microsatellite markers. Gen Mol Res. 2011;10:419-428.
11. Chander H, Ahuja DK, Nagender A, Berry SK. Repellency of different plant extracts and commercial formulations used as prophylactic sprays to protect bagged grain against tribolium castaneum - a field study. J Food Sci Technol Mys. 2000;37:582-585.
12. Chander H, Nagender A, Ahuja DK, Berry SK. Effect of various plant materials on the breeding of lesser grain borer (rhyzopertha dominica) in milled rice in laboratory. Journal of Food Science Technology. 2003;40:482-485.
13. Nirmal BK, Minoo D, Geetha SP, Sumathi V, Praveen K. Biotechnology of turmeric and related species. In: ravindran pn, nirmal babu k, sivaraman k (eds) turmeric: The genus curcuma. Crc Press, Boca Raton, Fl, USA. 2007;107-127.
14. Forsberg G, Johnsson I, Laerholm J. Effects of aerated steam seed treatment on cereal seed-borne diseases and crop yield. Journal of Plant Diseases and Protection. 2005;12(3):247-256.
15. Regmi R, Jha R, Simon LS, Lal AA. In vitro evaluation of some plant extracts against alternaria alternata causing leaf spot of aloe vera. Arpn Journal of Agriculture and Biological Science. 2014;9:323-325.
16. Swami CS, Alane SK. Efficacy of some botanicals against seed-borne fungi of greengram. Bioscience Discovery. 2013;4:107-110.
17. Javaid A, Samad S. Screening of allelopathic trees for their antifungal potential against Alternaria alternata strains isolated from dying-back eucalyptus.
18. Manoorkar VB, Gachande BD. Evaluation of antifungal activity of some medicinal plant extracts against some storage seed-borne fungi of groundnut. Science Research Reporter. 2014;4:67-70.

19. Saroja DGM. Antifungal effects of phytoextracts on seed-borne fungi of chickpea (Cicer arietinum L.). Journal of Progressive Agriculture. 2012;3:71-73.

20. Batish DR, Singh HP, Kohli RK, Kaur S. Eucalyptus essential oil as a natural pesticide. Forest Ecology and Management. 2008;256:2166-2174.

21. Abiamere Co, Nweke FN, Ogbadu LJ, Onyia OC, John CO. Evaluation of *moringa oleifera*, *zingiber officinale* (ginger), *chromolena odorata* plants extract as seed borne of cowpea. Journal of Pharmacy and Biological Sciences. 2014;9(6):66-70.

22. Lee HS, Shin, WK, Song C, Cho KY, Ahn YJ. Insecticidal activities of identified *Curcuma longa* rhizome against *Nilaparvata higens* (homoptera: delphacidae) and *Plutella xylostella* (lepidoptera: yponomeutidae). Journal Asia-pacific Entomolology. 2001;4:181-185.