CCT2 Mutations Evoke Leber Congenital Amaurosis due to Chaperone Complex Instability

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Leber congenital amaurosis (LCA) is a hereditary early-onset retinal dystrophy that is accompanied by severe macular degeneration. In this study, novel compound heterozygous mutations were identified as LCA-causative in *chaperonin-containing TCP-1, subunit 2* (CCT2), a gene that encodes the molecular chaperone protein, CCTβ. The zebrafish mutants of CCTβ are known to exhibit the eye phenotype while its mutation and association with human disease have been unknown. The CCT proteins (CCT α-θ) forms ring complex for its chaperon function. The LCA mutants of CCTβ, T400P and R516H, are biochemically instable and the affinity for the adjacent subunit, CCTγ, was affected distinctly in both mutants. The patient-derived induced pluripotent stem cells (iPSCs), carrying these CCTβ mutants, were less proliferative than the control iPSCs. Decreased proliferation under Cct2 knockdown in 661W cells was significantly rescued by wild-type CCTβ expression. However, the expression of T400P and R516H didn’t exhibit the significant effect. In mouse retina, both CCTβ and CCTγ are expressed in the retinal ganglion cells and connecting cilium of photoreceptor cells. The Cct2 knockdown decreased its major client protein, transducing β1 (Gβ1). Here we report the novel LCA mutations in CCTβ and the impact of chaperon disability by these mutations in cellular biology.

Results

Compound heterozygous mutation of CCT2 in a Chinese family with Leber Congenital Amaurosis. A whole exome sequencing (WES) analysis was conducted in a Chinese consanguineous family diagnosed with LCA based on the criteria mentioned above. The parents are second cousins (Supplemental Fig. 1A)
and their child (II-6, in Fig. 1A) was diagnosed as blind with severe retinopathy at an age of 6 months (Supplemental Fig. 1B). All of the members of this family were examined by fundus imaging and optical coherence tomography (OCT). Six members exhibited normal fundus and OCT images (Fig. 1C), while two of the six children were further diagnosed with LCA based on the presence of attenuated blood vessels and pallor fundus images (Fig. 1D). These observations were made in combination with the habitual oculodigital signs of photophobia and eye poking (Fig. 1E), clinical hallmarks of LCA. In addition, visual acuity tests performed for the affected individuals revealed no fixation of reflection and no following of movement. Except for the retinal dystrophy, both patients have hearing disorders and language barrier. But they had no polydactyly or renal cysts, the hallmarks of Bardet-Biedl syndrome (BBS) and ciliopathy.

Further genetic analysis revealed that two novel compound heterozygous mutations, c.1198A>G and c.1547G>A, were present in this CCT2, and these represented unrivalled candidates in this LCA family (Fig. 1B). These mutations were located in exon 12 and exon 15 of the CCT2 mRNA transcript, respectively.

CCT2 encodes protein, CCTβ, one of the eight subunits of the CCT machinery6,7. The hetero-octamer ring of CCTα-θ becomes a doublet to form a cavity where nascent proteins are folded into native proteins with ATP hydrolysis8–12. While various patterns of intra-ring orientations have been reported for each subunit, CCTβ is consistently reported to be a crucial anchor in combination with CCTγ for the folding of actin and tubulin proteins in the central cavity13–18. In addition to these major components of the CCT machinery, a subset of cell division control (CDC) proteins and the recently identified key factor for retinal development, Von Hippel-Lindau disease tumor suppressor protein19 are also governed by the CCT machinery20–22. Together with these retinal developmental factors, myosin (MYO7A, synonym: USH1B, the cause of Usher syndrome)23,24, transducin α (GNAT2, the cause of Achromatopsia)25, and peroxisomal targeting signal 2 receptor protein (PEX7, the cause of Refsum disease)26 are also known components of the CCT machinery27–29. Recently, protein β-sheet folding by the CCT machinery was also observed in concert with the co-factor of the undergoing protein folding30.

Phosducin-like protein (PhLP) is a cofactor of transducin β, and dominant negative expression of PhLP specifically in photoreceptor cells has been shown to induce severe retinal degeneration in mice31. Thus, the CCT machinery appears to govern retinal homeostasis based on its chaperone activity. However, while an association between this machinery and genetic retinal diseases has not been identified, it is apparent that proper protein folding by CCT chaperone machinery (referred to as CCT machinery hereafter) is indeed a crucial function for...
retinal homeostasis. Correspondingly, alterations in a gene mutation present in one of the CCT subunits has been shown to cause retinal or neural degenerative diseases due to intracellular toxicity as a result of misfolded and aggregated proteins. Here, the LCA mutations in CCT2 and CCT3 were found to involve the substitution of a threonine for a proline (T400P) and an arginine for a histidine (R516H), respectively. Family members that had either one of these mutations exhibited normal macula (Fig. 1C). In contrast, two of the children that carried both mutations exhibited retinal dystrophy and macular degeneration in fundus imaging, and OCT imaging revealed certain foveal hypoplasia (Fig. 1D).

Three-dimensional Mutant Protein Structure Prediction. To predict whether these mutations in CCT3 directly impact disease onset, molecular modeling of the hetero-octamer CCT machinery was conducted. Possible structural consequences of T400P and R516H are illustrated in Fig. 2, panels A–C and D–F, respectively. The change to a proline residue in position 400 appears to disrupt the α-helical conformation and destabilize several hydrogen bonds in the region of the C-cap of helix 14 (Fig. 2C). In addition, the introduction of a relatively bulky proline residue at the interface between the α-helices has the potential to cause outward movement of the surrounding helices, thereby reducing the ADP nucleotide (green) binding maintained by these helices. The other mutant, R516H, is located in the C-cap of α-helix 18. In the CCT3 subunit a positively charged arginine residue (R516) has a potential to stabilize α-helix 18 by forming a salt bridge with the negatively charged glutamic acid, E509 (Fig. 2F). Mutation of arginine R516 to histidine interrupts the salt bridge and results in the loosened α-helix structure. It is known that intermediate α-helices carrying these mutations are important for CCT intra-ring formation. Furthermore, computational modeling of the CCT hetero-oligomer indicated that CCT3 mutations could affect the functional activity of the CCT machinery (Fig. 2G–J).

CCT3 is decreased in LCA patient-derived T cells and iPSCs. Computational structural predictions of T400P and R516H CCT3 identified potential defects in the oligomerization of CCT proteins. Initially, it was investigated whether these defects affected intracellular levels of CCT3 in patient-derived induced pluripotent stem cells (iPSCs). Two sets of iPSCs were established, one from an LCA patient (II-6 in Fig. 1A) and one from an unaffected parent as a parental control (I-1 in Fig. 1A). As shown in Fig. 3A,B, the levels of CCT3 protein were lower in the patient-derived T cells and iPSCs compared to the corresponding parental cells (Fig. 3A,B).

Decreased mutant proteins and its degradation by proteosomal pathway. To elucidate the cause of the decrease in CCT3 levels, human wild-type CCT3, and the T400P and R516H forms of CCT3, were transiently overexpressed in HEK293T cells treated with or without the proteosomal degradation inhibitor, MG132. After cycloheximide was added to stop nascent protein synthesis, expression of T400P was found to be reduced in the soluble (supernatant) fraction, as well as in the insoluble (precipitated) fraction (Fig. 3C). The presence of T400P CCT3 in the insoluble fraction in the presence of MG132 suggests that T400P is rapidly degraded following protein synthesis. In contrast, expression of R516H CCT3 in the soluble fraction was similar to that of wild-type CCT3, while excess R516H protein was detected in the insoluble fraction without MG132 treatment. MG132 treatment further increased the amount of R516H in the insoluble fraction, thereby suggesting that these mutant proteins undergo intracellular degradation. Taken together, these results suggest that the levels of mutant proteins in the cytosol are suppressed by intracellular degradation, presumably because of their unstable structures. The observation that lower levels of mutant T400P were detected compared with mutant R516H also indicates greater instability of the T400P structure, and this may affect the cellular function of this protein.

Distinct affinity for CCTγ of T400P and R516H Mutants. Based on the observation that the intracellular dynamics of the CCT3 mutants differed from those of wild-type CCT3, liquid chromatography (LC)/mass spectrometry (MS)/MS proteomics was employed to examine whether this has any influence on formation of the CCT3-associating complex. A few T400P-specific bands were detected, yet there were no R516H-specific bands identified by silver staining. Two of the T400P-specific bands were identified by in-gel digestion and LC/MS/MS (Fig. 3D). The T400P form of CCT3 also exhibited greater affinity for HSP90 (indicated with a closed arrowhead in Fig. 3D), for one of the other major chaperone molecules, and for CCTγ (indicated with an open arrowhead in Fig. 3D), an adjacent subunit to CCT3 in the CCT hetero-octamer intra-ring structure. The association of each of the mutant proteins with HSP90 or CCTγ was confirmed with IP and Western blotting (WB) assays. Consistent with the results or LC/MS/MS, the HSP90 was precipitated only with T400P mutant (Fig. 3E). The CCTγ was precipitated with wild type CCT3 while the interaction seemed more abundant with T400P mutant (Fig. 3E). Unexpectedly, the R516H mutant precipitated less CCTγ compared to the wild type CCT3 or T400P mutant (Fig. 3E). Taken together, these results indicate that the expected from molecular modeling impact on CCT oligomerization is due to changes in affinity of the CCT3-CCTγ interface.

LCA patient-derived iPSCs are less proliferative compared to control iPSCs. During this study, we noticed that one week later from the iPSC generation, the number of patient-derived T cells reached only 63% of the number of parental T cells present (5.4 × 10⁴ cells/ml vs. 9.5 × 10⁴ cells/ml, respectively). To explore the impact of the CCT3 mutants on cellular physiology, the cell proliferations were monitored after iPSCs establishment. A significant reduction in patient-derived iPSC growth was observed, with patient-derived T400P and R516H cells being less proliferative than the parental control (CCT3/T400P) or normal control (CCT3/CCT3) cells. These results were consistent with the previously reported smaller eye phenotype observed in several zebrafish lines treated with morpholinos and mutagen-exposed zebrafish lines indicated a possibility that proliferation defects were mediated by CCT3 and CCTγ mutations. Taken together, these data suggest that proliferation activity is one of the phenotypes that characterize mutant CCT3 proteins. Indeed, the patient-derived iPSCs examined in the present
study exhibited a slower growth rate in 96-well MTS assays after 3 days in culture (Fig. 4A). Furthermore, the growth defect became obvious in long-term serum-free embryoid body cultures with the formation of spheroids (Fig. 4B).

Stable expression of LCA mutants of CCTβ are lacking the rescue effects on proliferation. To investigate the impact of the CCTβ mutants on cellular growth, endogenous levels of Cct2 were first knocked down using silencing RNA (siRNA) in the mouse photoreceptor-derived cell line, 661W (Fig. 4C). Then, stable 661W cell lines overexpressing human wild-type CCTβ, T400P CCTβ, and R516H CCTβ were generated using a lentivirus system. A mouse Cct2-specific siRNA sequence that did not exhibit off-target effects on human CCT2 was used to knockdown endogenous levels of Cct2 in the parental 661W cells, and in the three overexpression cell lines. Cell growth for each cell line was monitored for 6 d. Proliferation of the parental 661W cells was...
Figure 3. Intracellular Dynamics of CCTβ and its LCA-causing mutants. Expression levels of CCTβ protein in patient-derived T cells (A) and iPSCs (B) were analyzed by WB. Lower levels of CCTβ were detected in patient-derived T cells and iPSCs (P09) compared with the parental control iPSCs (N13) and wild-type CCTβ -carrying control iPSCs (Cont.). (C) Intracellular protein stability was monitored using the proteasomal degradation inhibitor, MG132, and protein synthesis was abolished with cycloheximide (CHX) treatment for 3 h. Cytosolic protein levels of T400P were lower than R516H and wild-type CCTβ. The insolubility of T400P
and R516H is predicted to contribute to the degradation observed. Sup., supernatant fraction; Ppt., Precipitated fraction. (D) Each FLAG-tagged transfected sample was immunoprecipitated (IP) and subjected to WB and silver staining. Two distinct binding bands were observed for the T400P variant (indicated with a black arrowhead and the white arrowhead respectively). These two bands were digested with trypsin by an in-gel method and was subsequently identified as containing HSP90 (black arrowhead) and CCT-γ (white arrowhead) by LC/MS/MS. (E) Consistent with the MS analysis data, T400P exhibited a higher affinity for HSP90 and CCT-γ in IP and WB assays. In contrast, R516H exhibited a lower affinity for CCT-γ. Thus, these CCTβ variants appear to affect CCTγ during oligomerization of the CCT-chaperonin ring. The number labels in (E) indicate the fold-change in expression compared to wild-type HSP90 or wild-type CCTβ. Bg: background from mock (empty vector transfected) control.

significantly decreased (shown as * in Fig. 4E), while the cells expressing wild-type CCTβ exhibited a significant rescue effect compared to the parental cells (* and # respectively, Student’s t-test, p < 0.05). In contrast, the T400P- and R516H-expressing 661W cells exhibited no statistically significant rescue effect, yet a slightly positive effect on cell proliferation was observed. These results indicate that the mutants were less effective in inducing cell proliferation than wild-type CCTβ. For mutants T400P and R516H, the former was significantly less effective than wild-type CCTβ, and there was no significant difference between R516H and wild-type CCTβ cells. These results are consistent with the structural predictions and degradation assay results described above thereby indicating that T400P CCTβ has a greater effect on the functional activity of the CCT machinery. These results also suggest that compound heterozygous mutations in CCT2 induce a partial, and not complete, functional defect in the CCT machinery.

CCTβ and CCTγ colocalize in photoreceptors and Retinal Ganglion Cells. While it is reasonable that the CCTβ mutations suppress cell proliferation, and this can lead to hypoplasia of the retina and the manifestation of retinal dystrophy in LCA patients, it does not explain the progression of LCA to macular degeneration. To clarify this, we next examined the tissue localization and expression of CCTβ and CCTγ, as well as their major target protein, Gβ1 (also known as transducin β in photoreceptor cells), a second messenger of phototransduction signaling. As shown in Fig. 5A, CCTβ was found to be dominantly expressed in the cells in retinal ganglion cell layer (GCL), and also expressed in photoreceptor cells. In the latter, CCTβ was diffusely localized in the cytosol, while also being localized to some dense dots just between the inner segment (IS) and the outer segment (OS). These results suggest that CCTβ localizes to the basal body or the base of the connecting cilium. CCTγ was also expressed in the RGCs, and more abundantly, in the photoreceptor cells. Of particular note is the observation that CCTγ localized just between the IS and OS, similar to CCTβ, while its localization manner was more similar to that of the connecting cilium or axoneme structure. Gβ1 protein expression was also abundant in the neural retina (Fig. 5B), and siRNA-mediated knockdown of Cct2 in 661W cells decreased the levels of Gβ1 after 4 d of treatment (Fig. 5C). In combination, these results suggest that CCTβ and CCTγ have critical roles in the retina to maintain the physiology of the photoreceptor cells and the RGCs, and they also regulate its major target, Gβ1, an important molecule in the phototransduction pathway. Previously, it was shown that a deficit of Gβ1 in photoreceptor cells that was achieved by expressing a dominant negative form of PhLP evoked retinal degeneration in a mouse model by disturbing the function of transducin and the downstream phototransduction pathway31,37,38. Thus, it appears that reduced activity of the CCT machinery can contribute to the development of macular degeneration, as well as retinal dystrophy.

Discussion
In the present study, the mutation of CCTβ is identified as a novel LCA-causative gene. Due to the clear diagnosis of LCA and the sufficient number of genome samples, the genetic analysis pointed out only one candidate gene with compound heterozygous inheritance. While it has to be noted that this CCT2 gene mutation was found in sole consanguineous family and the following reports concerning the CCT2 mutation with LCA will be anticipated. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery. The CCT chaperone machinery found to be affected by the structurally instable T400P and R516H mutant proteins that exhibited an aberrant affinity to the CCTγ machinery.
Figure 4. Patient-derived T400P/R516H-carrying cells are less proliferative, and these LCA-causing CCTβ mutants were failed to rescue the phenotype. (A) The proliferation rate for patient-derived iPSCs (P09) was significantly lower than the proliferation rate of the parental control iPSCs (N13) and the wild type CCTβ -carrying control iPSCs (Cont.) that were maintained under normal iPS culturing conditions (* and #, respectively, Student’s t-test, p < 0.05). (B) Growth rates according to spheroid size were also significantly smaller for patient-derived iPSCs grown in serum-free embryoid body cultures for 8 days compared to the controls (* and #, respectively, Student’s t-test, p < 0.05). (C) The mouse-specific knockdown of CctT2 by siRNA significantly decreased the amount of CCTβ expressed in 661W cells (**Student’s t-test, p < 0.01). (D) Proliferation of the 661W cells was largely suppressed after 3 days of Cct2 knockdown. (E) To elucidate the impact of the T400P and R516H mutants, stable 661W clones stably expressing human CCTβ, T400P, and R516H were established by lenti-virus infection. Both stable and parental 661W cell lines were transfected with Cct2-targeted siRNA, and cell proliferation was monitored for 6 days. Consistent with the former results, parental 661W cells and T400P-expressing 661W cells exhibited a marked decrease in cell proliferation following Cct2 knockdown (* and #, respectively, Student’s t-test, p < 0.05). In addition, both T400P- and R516H-expressing 661W cells exhibited no statistically significant rescue effect, yet did exhibit slightly enhanced cell proliferation.
Figure 5. Localization of CCTβ and CCTγ in RGCs and Photoreceptor cells in Mouse Retina. (A) Low magnification micrographs (top, scale bar = 20 μm) show the localization of CCTβ and CCTγ in ganglion cells and photoreceptor cells, respectively. Expression of CCTβ was more dominant in the cells of ganglion cell layer (middle micrographs, scale bar = 20 μm), while both CCTβ and CCTγ strongly localized between the IS and OS (bottom micrographs, scale bar = 5 μm). In the photoreceptor cells, CCTβ was diffusely distributed in the cytosol, and was also densely localized in punctate regions between the IS and OS, similar to the basal body dominant manner in which CCTγ localized to the basal body, thereby further connecting in a cilium manner in photoreceptor cells. (B) Expression levels of CCTβ and Gβ1 were higher in the retinal lysates than in the RPE lysates in the mouse eye. Detection of actin was used as a loading control. (C) CCTβ knockdown resulted in lower levels of Gβ1 expression after 4 days of siRNA treatment in 661W cells. GCL: ganglion cell layer; IPL: inner plexiform layer; INL: inner nuclear layer; OPL: outer plexiform layer; ONL: outer nuclear layer; IS: inner segment; OS: outer segment.
It has recently been demonstrated that CCT2 has roles in the folding of β-propeller proteins, such as Gβ31, in addition to its major target proteins, actin and tubulin. In a study of the co-chaperone protein, PhLP, use of a dominant negative form lacking the N-terminal domain demonstrated that this region is necessary for the binding of nascent Gβ31. These results also showed the importance of Gβ31 in retinal homeostasis. In a transgenic mouse model expressing a dominant negative form of PhLP1 (dnPhLP−β), deficits in Gβ31 in the retina are observed, and this is associated with severe retinal degeneration after birth due to transducin-associated phototransduction failure31,42. CCT3 has also reported to interact with chaperonin type BBSs, including BBS6, 10, and 12, and lower levels of expression for a subset of BBSs has been observed in the dnPhLP−β retina31. Taken together, these results indicate that there may be a close link between chaperonin activity and cilium transport. On the other hand, the CCT2 mutation-carrying LCA patients do not exhibit the clinical manifestations of BBS or ciliopathy in this study. This probably because the CCT2 has more broad function in development by chaperoning some critical developmental proteins while the BBS has more dominance in cilia-associating function. Though both CCTβ and CCTγ took a connecting cillum-like localization in photoreceptors, the association between CCT-BBS functional axis and human disease has to be elucidated in the future.

Artificial gene manipulation of other CCT family members, including CCT3, CCT5, and CCT7, have been also shown to evoke developmental eye defects in several zebrafish mutant lines (http://zebrafish.org/home/guide.php). In one of the well-characterized CCT3 mutant zebrafish lines, non tectal neuron (ntn), a retinal phenotype was observed that involved fewer axonal projections of the RGCs to the lateral geniculate nucleus (LGN). The retinal ganglion cell deaths was observed in the ntn mutants probably because of lacking the trophic factor from LGN. It is possible that mutations in CCT2 could lead to genetic retinal diseases based on this uncoordinated fashion of delayed development, in this case, the failure of axonal projection to the LGN. It is also a well-known fact that the migrating retinal astrocytes and the following vascular formation after birth closely associate with RGC axons44,45. Therefore, the axonal disruption of RGC could trigger the hypoplastice retinal development shown in the patients. It is an increasing fact to introducing the optic-cup and ciliarr differentiation in 3D or 2D organoid cultures from patient-derived iPSCs to evaluate the onsite genetic impacts. Recapitulate the phenotype in the optic organoid cultures will become a powerful analytic approach. Unfortunately, the patient-derived CCT2 mutation-carrying iPSCs have a few disadvantages to introduce organoid cultures and was not applicable this time. Firstly, due to the significantly less proliferative activity of patient-derived CCT2 mutation-carrying iPSCs, optimizing the total cultivation time, induction time point and endpoint validation time point to compare the phenotype of parental iPSCs were all difficult. Secondly, the CCT2 phenotype in retina is probably associating with the RGC axon projection to LGN. It has been reported that the truncated mutation of CCTγ mutant zebrafish exhibited fewer axonal projections of the RGCs to the LGN and increased number apoptotic RGCs simultaneously36. This is probably because RGC-geneulate communication is required for further RGC survival and maturation. In order to validate these hypotheses, it is critical to utilize the in vivo phenotypic model that allows us to evaluate the orchestrate development of RGC with axon projection to the LGN. Recently, the genome editing by CRISPR-Cas9 is also applicable to iPSCs to validate the mutational impacts and rescue effect. This technique will provide the onsite impacts of CCT2 mutation in retinal development and eliminate the trophic effect from LGN. Combining all these techniques and further analyses will be important to clarify the CCT2 function in detail. The forward genetics in experimental animals, in combination with genome wide sequencing in human patients, has the potential to elucidate the causes of various developmental diseases.

Gene mutations in rat CCT4 (C450Y of CCT6)46,47 and human CCT5 (H147R of CCTγ)48 have been reported to cause sensory neuropathy, although the underlying mechanisms of disease onset have not been elucidated. Recently, the effects of these two mutations on the CCT machinery were analyzed using in vitro expression of mutant homo oligomer models49,50. Both of the gene mutations evoked sensory neuropathy, albeit via two distinct mechanisms. Mutant C450Y of CCT6 was found to be structurally instable and it failed to form the CCT ring complex. In contrast, mutant H147R of CCTγ exhibited normal CCT ring complex formation, yet the activity of the CCT machinery was reduced49,50. This analysis of the biochemical features of these eukaryotic mutant CCT subunits in a homo oligomer model established in prokaryotic host cells has the disadvantage that CCT is a eukaryote-specific chaperone and is not functional in prokaryotes. Thus, it is difficult to further investigate the physiology of the hetero-oligomer complex in this model. On the other hand, in the present study, the patient iPSCs demonstrated that maintenance of CCT machinery affects cell proliferation. It is noteworthy that the mutations, T400P and R516H, in CCT2 were not embryonic lethal, but resulted in a retinal phenotype. Taken together, these results suggest that there is a distinct threshold for the impact of mutations on subunit-target interactions in specific cell types as well as the tissue specificity. Thus, disease manifestation due to CCT mutations may appear in cells/tissues where CCT subunit-governed targets have greater importance in maintaining cellular physiology. By characterizing the chemistry of the respective CCT subunits and identifying the specific targets of the CCT complex, it may be possible to further elucidate which target proteins are crucial for specific cell types, thereby providing novel insights into our understanding of retinal development and the associated pathoetiology of hereditary retinal diseases.

In conclusion, two novel compound heterozygous mutations in CCT2 were identified in association with a family affected by LCA. These mutations affect CCTβ-associated chaperone activity which has an essential role in retinal development and photoreceptor physiology.

Methods

Whole Exome Analysis. For the human sample experiments, informed consents for all individuals were obtained in written format from the parents. DNA samples from all of the family members were subjected to WES to identify the responsible gene mutations (Fig. 1A). Libraries for WES were prepared from the DNA samples using an exon capture kit (SureSelect ver. 4+ UTR, Agilent Technologies, Santa Clara, CA), according to the manufacturer’s instructions. The exons were sequenced as 100 basepair paired-end reads by an Illumina HiSeq2000.
(Illumina, San Diego, CA). The sequence data obtained were then analyzed to extract potentially causal mutations. Briefly, sequences of the whole exome were compared with a reference human genome (hs37d5), and 5,144,654 mutations were identified. Only mutations that resulted in a change in amino acid sequence were selected as candidate mutations (n = 681). The remaining common mutations were excluded. Using a pattern of inheritance with the parents, one candidate causal gene was identified (Supplementary Table 1). In the RetNet database, only a gene mutation present in the T400P specific bands were in-gel digested and processed for LC/MS/MS as previously reported52. C57BL/6J detection was used as a loading control and for the normalization of CCT

pared with M2-FLAG sepharose (Sigma) according to its manufacturer's protocol34. The IPs of wild-type, T400P, and R516H, were generated and positioned in an optimized conformation. Full atomic structures of the wild type and mutant octamers were minimized iteratively to achieve a global energy minimum structure. Finally, octamer stereochemistry was optimized and equilibrated using 1 nanosecond molecular dynamics in water.

Establishment of LCA Patient-Specific iPSCs. Prior to obtaining iPSCs from blood samples, the patients were fully informed and all procedures performed were approved by the ethics committee of the National Organization Tokyo Medical Center. All the experiments were carried out in accordance with the approved guidelines. Circulating T-cells in the blood samples were obtained by centrifugation with Ficoll reagent, and the Yamanaka 4 factors were induced by Sendai-virus infection. After transforming the iPSCs, alkaline phosphatase activity was assayed, and then the undifferentiated stem cell state was confirmed by detecting expression of Nanog, OCT3/4, TRA81, and SSEA4.

Plasmids and Biochemical Studies. The cDNA clone of human CCT2 was obtained from NITE Biological Resource Center (NBRC). The cDNA was subcloned into the BamHI site of pEF-BOS-FLAG vector51 and the mutations responsible for T400P and R316H were introduced by KOD Mutagenesis Kit (TOYOBO) respectively. The human CCT3, T400P and R516H were transfected into HEK293T cells31 with TransIt Pro Transfection Kit (Mirus) according to its manufacturer’s instructions. A mini-PROTEAN TGX Gel and TransBlot Turbo system (BioRad) were used for SDS-PAGE Western blotting (WB) according to the manufacturer’s instructions34. Actin detection was used as a loading control and for the normalization of CCT3 levels. Detected bands against CCT3 or actin were quantitated using Chemidoc XRS+ with the Image Lab software package (BioRad). Intracellular protein stability was monitored by treating transfected cells with proteasomal degradation inhibitor, MG132, and protein synthesis was abolished with cycloheximide (CHX) during 3 h. Immunoprecipitates (IPs) by FLAG-tag were pre pared with M2-FLAG sepharose (Sigma) according to its manufacturer's protocol34. The IPs of wild-type, T400P, and R516H forms of CCT3 were next subjected to SDS-PAGE and then were silver stained (Silver Quest). Then the T400P specific bands were in-gel digested and processed for LC/MS/MS as previously reported32. C57BL/6J retina was carefully removed from the posterior part of the eye and the retinal pigment epithelium (RPE) was then scraped from the remaining sclera tissue to assess the expression of CCT3 in the mouse eye. The animal experiment was carried out in accordance with the Guide for the Care and Use of Laboratory Animals (National Institutes of Health) and the Association for Research in Vision and Ophthalmology Statement for the Use of Animals in Research, and approved by the Tokyo Medical Center Experimental Animal Committee. The tissue lysate was analyzed by WB using mini-PROTEAN TGX Gel and TransBlot Turbo system (BioRad) as mentioned above.

Proliferation Assay. Cell proliferation was monitored by MTS Assays (Cell Titer, Promega). The proliferation of iPSCs that were seeded onto feeder-coated 96-well plates (1 × 10^4 cells/well) was monitored for 3 days with MTS assays. Growth rates according to spheroid size for iPSCs grown in serum-free embryoid body cultures were less than 0.05 were considered to be statistically significant.

siRNA and Stable Expression by LentiVirus. The siRNA system (Sealth RNAi siRNA, Life Technologies) for Mouse CCT2 and a control siRNA were used with RNA iMAX reagents (Life Technologies) to knockdown expression of endogenous mouse Cct2 in 661W cells, a mouse cone photoreceptor cell line. Transfection of a siRNA specific for mouse Cct2 that did not exhibit off-target effects on human CCT3 in HEK293T cell line was performed and no off-target effect was confirmed by WB (data not shown). The human wild type CCT3, T400P and R516H stably expressing 661W cells were established by lentivirus system with CSII-CMV-MCS-IRES2-Bsd vector and the packaging plasmids (RIKEN BRC).

Immunohistochemistry. Eyes from 5-week-old male C57Bl/6J mice were dissected and fixed in 4% paraformaldehyde. After sucrose replacement, the lens was removed from each eye and left retinal tissues were routinely processed to obtain 14µm-thick cryosections. The following antibodies were used for staining: anti-CCT3 (Cell Signaling Technology), anti-CCT7 (Abcam), and anti-G31 (Abcam). Briefly, the cryosections were permeabilized with 0.4% Triton X-100 in phosphate-buffered saline (PBS) for 15 min. For negative controls, a mixture of rabbit IgGs (Dako) was used instead of primary antibody. The specimens were counterstained with Hoechst33642. The LSM700 and Zen Systems (Zeiss) were used for confocal microscopy.
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Author Contributions
Y.M. conducted all of the biological assays and prepared the manuscript; Y.M. and T.I. designed the research; X.L.S., W.Z., Y.L. and W.R. diagnosed the patients; K.Y., N.M., K.I. and M.F. analyzed the whole exome sequencing; Y. Sergeev performed the protein 3D structure prediction; D.I. supplied the iPSCs; Y. Shibagaki and S.H. performed the LC/MS/MS; T.I. supervised all the experimental results and edited the manuscript.

Additional Information
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