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RESEARCH ARTICLE

Loss-of-Function Mutations in Three Homoeologous PHYTOCLOCK 1 Genes in Common Wheat Are Associated with the Extra-Early Flowering Phenotype

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Abstract

Triticum aestivum L. cv ‘Chogokuwase’ is an extra-early flowering common wheat cultivar that is insensitive to photoperiod conferred by the photoperiod insensitive alleles at the Photoperiod-B1 (Ppd-B1) and Ppd-D1 loci, and does not require vernalization for flowering. This reduced vernalization requirement is likely due to the spring habitat allele Vrn-D1 at the VERNALIZATION-D1 locus. Genotypes of the Ppd-1 loci that determine photoperiod sensitivity do not fully explain the insensitivity to photoperiod seen in ‘Chogokuwase’. We detected altered expression patterns of clock and clock-output genes including Ppd-1 in ‘Chogokuwase’ that were similar to those in an einkorn wheat mutant that lacks the clock-gene homologue, wheat PHYTOCLOCK 1 (WPCL1). Presumptive loss-of-function mutations in all WPCL1 homoeologous genes were found in ‘Chogokuwase’ and ‘Geurumil’, one of the parental cultivars. Segregation analysis of the two intervarietal F2 populations revealed that all the examined F2 plants that headed as early as ‘Chogokuwase’ had the loss-of-function wpcl1 alleles at all three homoeoloci. Some F2 plants carrying the wpcl1 alleles at three homoeoloci headed later than ‘Chogokuwase’, suggesting the presence of other loci influencing heading date. Flowering repressor Vrn-2 was up-regulated in ‘Chogokuwase’ and ‘Geurumil’ that had the triple recessive wpcl1 alleles. An elevated transcript abundance of Vrn-2 could explain the observation that ‘Geurumil’ and some F2 plants carrying the three recessive wpcl1 homeoealleles headed later than ‘Chogokuwase’. In spite of the up-regulation of Vrn-2, ‘Chogokuwase’ may have headed earlier due to unidentified earliness genes. Our observations indicated that loss-of-function mutations in the clock gene wpcl1 are necessary but are not sufficient to explain the extra-early heading of ‘Chogokuwase’.

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Introduction

In temperate cereals such as barley and wheat, three factors, i.e., the vernalization requirement, photoperiod sensitivity and narrow-sense earliness (earliness per se) contribute to the timing of heading [1]. The vernalization requirement is controlled by three genes, namely, VERNALIZATION 1 (Vrn-1), Vrn-2 and Vrn-3 [2]. Vrn-1 encodes an APETALA1/FRUITFUL-like (AP1/FUL-like) MADS-box transcription factor [3–6]. Recessive alleles at all Vrn-1 homoeoloci confer a winter growth habit (vernalization insensitive), whereas one or more dominant alleles at Vrn-1 homoeoloci results in a spring growth habit (vernalization insensitive). The expression levels of Vrn-1 in leaves are associated with earliness, suggesting that Vrn-1 determines the flowering time [7]. Recently, Vrn-D4 was identified as a duplicated copy of Vrn-A1 [8]. The Vrn-2 locus contains two tandemly duplicated genes (ZCCT1 and ZCCT2) that encode a protein with a CCT (CONSTANS, CO-like, and TOC1) domain and a zinc-finger motif [9]. Non-functional mutations in the ZCCT genes result in a spring growth habit and, thus, early flowering [9–11], indicating their roles as flowering repressors. Vrn-3 encodes a RAF kinase inhibitor-like protein with similarity to Arabidopsis FLOWERING LOCUS T (FT). Transgenic wheat plants overexpressing Vrn-3 have an extra-early flowering phenotype without the need for vernalization [12–13], indicating that Vrn-3 is a strong flowering promoter. Hereafter, we denote Vrn-3 as WFT (wheat FT). Three models, although controversial, for the epistatic interaction between these vernalization genes have been proposed [13–16].

Photoperiod insensitive alleles of Ppd-1 (Ppd-1a) were identified for each homoeolocus on chromosomes 2A, 2B and 2D of common wheat, respectively [17–20]. Three alleles of Ppd-A1a and one allele of Ppd-D1a possess deletions in a shared promoter region [18–19, 21]. The cultivars carrying these Ppd-1a alleles had an increased expression level with an abnormal circadian rhythm for Ppd-1 expression. Novel mutations were found in the 5′ upstream regions of Ppd-A1 and Ppd-B1 [21]. In addition to the mutations in the promoter region, increased copy numbers of Ppd-B1 altered its expression pattern and accelerated flowering time [22–23]. Wheat plants with at least one Ppd-1a allele were associated with an increased expression level of TaFT (WFT), which correlates with the early flowering phenotypes [20]. In barley and wheat, the orthologues of circadian clock genes in Arabidopsis have been identified as candidate genes conferring the early flowering phenotype [24–28]. Recently, we showed that a wheat homologue of Arabidopsis LUX ARRHYTHMO/PHOTOCLOCK 1 (LUX/PCL1), which is located on the long arm of wheat chromosome 3A, is a candidate gene that is missing in an early flowering mutant of einkorn wheat (Triticum monococcum L.) [25]. In Arabidopsis, LUX/PCL1 constitutes the Evening Complex (EC) together with EARLY FLOWERING 3 (ELF3) and ELF4 [29]; the EC directly represses PRR9 [30]. The barley and wheat mutants of LUX/PCL1 and ELF3 homologues head earlier than the wild-type lines under both long day and short day conditions [24–28]. In the einkorn wheat mutant lacking the wheat LUX/PCL1 homologue (WPCL1), Ppd-1 and WFT were up-regulated, whereas LATE ELONGATED HYCOPOTYL (LHY) and TIMING OF CAB EXPRESSION 1 (TOC1) were down-regulated [25, 28]. These expression patterns were also observed in the Arabidopsis lux mutants [31]. These results strongly suggest that barley and wheat LUX/PCL1 function similarly to their counterparts in Arabidopsis.

Intermittent rain before harvest often causes pre-harvest sprouting and poor grain quality in wheat cultivated in East Asia [32]. To avoid these problems, early-heading wheat cultivars have been bred in many countries. Among 260 wheat cultivars examined, ‘Chogokuwase’, a super-early-heading wheat cultivar in Japan, is one of the earliest heading cultivars [33]. The heading time for ‘Chogokuwase’ was much earlier than the parental cultivars, ‘Minaminokumugi’ and ‘Gurumil’ (Table 1), suggesting that earliness genes were inherited from both parents. Seki et al. (2011, 2013) [33–34] showed that ‘Chogokuwase’ had the Ppd-D1a allele...
and there are no differences in the Ppd-1 genotypes between 'Chogokuwase' and the parental cultivars at any of the homoeoloci. This result strongly suggests that genes other than Ppd-1 contribute to the earliness of 'Chogokuwase'. Here, we conducted gene expression analyses, sequence analyses and segregation analyses to elucidate the reasons for earliness in 'Chogokuwase'. Our results indicate that loss-of-function alleles of WPCL1 homoeologues contribute to the earliness of 'Chogokuwase'. We discuss the function of WPCL1 in the photoperiod and vernalization pathways of the wheat flowering gene network.

Materials and Methods

Plant materials

'Chogokuwase' is an offspring of a cross between a Japanese cultivar 'Minaminokomugi' and a Korean cultivar 'Geurumil'. The common wheat cultivar 'Chinese Spring' (CS) was used to determine copy number variation and for sequence analyses. We used the following two F2 populations for segregation analyses: (1) the "CN population" consisted of 114 F2 plants derived from a cross between 'Chogokuwase' and 'Norin 61', and (2) the "MG population" consisted of 489 F2 plants derived from a cross between 'Minaminokomugi' and 'Geurumil'. Recombinant inbred lines (F11 generation) that were derived from the cross between cultivated einkorn wheat T. monococcum L. (KT3-5) and wild einkorn wheat T. boeoticum Boiss. (KT1-1) were used for an analysis of epistatic interactions between Vrn-2 and WPCL1 [25].

Field experiments

'Chogokuwase', 'Minaminokomugi' and 'Geurumil' were sown in the middle of October in an experimental field (36.108 N, 136.275 E) at Fukui Prefectural University, Fukui, Japan (2011/2012 and 2012/2013 seasons). Sowing dates were 19 Oct 2011 and 17 Oct 2012. The heading dates of each cultivar were recorded.

Measurement of heading time in a growth chamber

To investigate their photoperiod sensitivity and vernalization requirement, plants were cultivated in a growth chamber under long-day (16 h light and 8 h dark; LD) or short-day (10 h light and 14 h dark; SD) conditions at 20°C (light intensity ~100 μE m⁻² s⁻¹). Seedlings at 5-days after germination were subjected to a vernalization treatment (5°C) for 21 days. After the vernalization treatment, plants were grown at 20°C. Heading time was measured in at least three replicated samples as the number of days between unfolding of the third leaf and the flag leaf. Differences between samples were statistically examined by Tukey Kramer's HSD tests (P < 0.05).

Genotyping of Vrn-1 and Ppd-1 homoeoloci

We used polymerase chain reaction (PCR) primers (S1 Table) that had been shown to identify the alleles of Vrn-1 homoeoloci in previous studies [35–36]. Genotypes of the three Ppd-1

Table 1. Heading dates of 'Chogokuwase' and the parental cultivars in the field.

| Cultivar          | 2011/2012a | 2012/2013a |
|-------------------|-----------|-----------|
| 'Chogokuwase'     | 18 March  | 24 March  |
| 'Minaminokomugi' | 23 April  | 15 April  |
| 'Geurumil'        | 28 April  | 23 April  |

a Sowing dates were 19 Oct 2011 or 17 Oct 2012, respectively.

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A Self-archived copy in Kyoto University Research Information Repository
https://repository.kulib.kyoto-u.ac.jp
homoeoloci in ‘Chogokuwase’, ‘Minaminokomugi’, ‘Geurumil’ and ‘Norin 61’ were determined as described in Seki et al. (2011, 2013) [33–34]. To estimate the copy number of the Ppd-B1 loci, real-time PCR analyses were carried out using a LightCycler Nano System (Roche Diagnostics, Switzerland). A wheat CONSTANS2 gene, TaCO2 (also called TaHd-1), was used as the internal control. We used gene-specific primer sets for Ppd-B1 and TaCO2 as reported in a previous report [22]. The rate of amplification was monitored using THUNDERBIRD SYBR qPCR mix (TOYOBO, Japan) according to the manufacturer’s protocol. Amplification rates of the samples in five technical replications were converted to copy numbers by comparing the rates in common wheat cultivars ‘Cheyenne’, ‘Timstein’ and ‘CS’ that are known to possess one, three, and four copies of Ppd-B1, respectively [22].

**Quantitative reverse-transcriptase (RT)-PCR**

Expression of Vrn-1 and WFT was analyzed in non-vernalized plants, 21 days-veralnized plants grown under the LD conditions, and in 21 days-veralnized plants grown under the SD conditions. For each cultivar, seedlings were sampled from the 2-leaf stage to the 4-leaf stage. In this study, the 1-leaf stage was defined as the period from the unfolding of the first leaf to the unfolding of the second leaf. Total RNAs were extracted from leaves using ISOGEN (Nippon Gene, Japan). cDNAs were synthesized from the total RNAs using an oligo dT primer according to the protocol for the Ready-To-Go T-primed First-Strand Kit (GE Healthcare Life Sciences, USA). Real-time PCR analyses were performed in three technical replications using a LightCycler 2.0 (Roche Diagnostic, Switzerland) with the gene-specific primer sets shown in Table 2. Transcript abundance was determined relative to a SYBR Green-labelled amplification product of the wheat Actin gene (AB1819991) that is often used as an internal control for gene-expression analyses under various conditions [25, 37].

For the expression analysis of the clock and clock-output genes, plants were grown at 23°C under SD conditions (9 h light and 15 h dark). Two-week-old leaves were sampled every three hours. Total RNA was isolated using an RNeasy Plant Mini Kit (Qiagen, Germany). First-strand cDNA was synthesized from 1 μg RNA in a 20 μL reaction solution with oligo-dT primers using a ReverTra Ace -α- Kit (TOYOBO, Japan). Primers used for quantitative RT-PCR are shown in S2 Table. Relative transcript levels were determined in three technical replications using a LightCycler Nano System with FastStart Essential DNA Green Master (Roche Diagnostics, Switzerland). Quantitative RT-PCR was performed according to the manufacturer’s protocol using the wheat Actin gene as the internal control.

**Sequencing of three WPCL1 homoeologues**

Total DNA was extracted from leaves of common wheat using a DNeasy Plant Mini Kit (Qiagen, Germany). To obtain genome-specific primers, we searched for the sequences of all three WPCL1 homoeologues.

**Table 2. Alleles at the Ppd-1 and Vrn-1 loci.**

| Cultivar          | Ppd-1   | Vrn-1   | WPCL1               |
|------------------|---------|---------|---------------------|
| ‘Chogokuwase’    | Ppd-A1b/Ppd-B1b/Ppd-D1a | vrn-A1/vrn-B1/VRN-D1 | wpcl-A1/wpcl-B1/wpcl-D1 |
| ‘Minaminokomugi’ | Ppd-A1b/Ppd-B1b/Ppd-D1a | vrn-A1/vrn-B1/VRN-D1 | wpcl-A1/wpcl-B1/wpcl-D1 |
| ‘Geurumil’       | Ppd-A1b/Ppd-B1b/Ppd-D1a | vrn-A1/vrn-B1/vrn-D1 | wpcl-A1/wpcl-B1/wpcl-D1 |
| ‘Norin 61’       | Ppd-A1b/Ppd-B1b/Ppd-D1a | vrn-A1/vrn-B1/VRN-D1 | WPCL-A1/WPCL-B1/WPCL-D1 |

a Alleles designated according to Seki et al. 2011 [33] and 2013 [34]
b In the absence of dominant VRN-A1 and VRN-B1 alleles, the presence of a dominant VRN-D1 confers a spring habit to ‘Chogokuwase’ and ‘Minaminokomugi’, and a recessive vrn-D1 confers a winter habit to ‘Geurumil’.

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WPCL1 homoeologues in the chromosome-arm specific survey sequences of 3AL, 3B and 3DL [38]. First, a blastn search against the survey sequences was performed using the WPCL1 sequences of T. monococcum L. (accession number; AB773826) [25] as a query. Alignments of the survey sequences obtained from the three genomes led to the design of the following genome-specific primers to amplify the entire coding region: WPCL-A1, 5′-GCTCCAAATGGTCCAGAGGA-3′ and 5′-GGACAGTGAAGGAAATGAG-3′; WPCL-B1, 5′-TGGACAAATTAATATTGCAGACA-3′ and 5′-GATCGACACAAACACACAGCC-3′; WPCL-D1, 5′-ATCTATCCACATCGGACCAGA-3′ and 5′-CCGGAGACACACATTCACA-3′.

Thirty-three cycles of PCR were performed using KAPATaq Extra (Kapa Biosystems, USA) and the following reaction conditions: 30 s at 94°C, 30 s at 60°C, and 45 s at 72°C. Nucleotide sequences were determined with BigDye Terminator version 3.1 (Applied Biosystems, USA) using an Applied Biosystems 3730xl DNA Analyzer. Nucleotide sequences and their predicted amino acid sequences were analyzed by GENETYX-MAC version 12.00 software (Whitehead Institute for Biomedical Research, USA).

Segregation analyses

One hundred fourteen F2 plants (CN population) from a cross between ‘Chogokuwase’ and ‘Norin 61’ were used in the first segregation analysis. The plants were grown at 23°C for ten days and then vernalized at 4°C for seven weeks. After vernalization, the plants were incubated at 23°C and grown under SD conditions (8 h light and 16 h dark). In addition to the CN population, the segregation of 489 F2 plants (MG population) from a cross between ‘Minaminokomugi’ and ‘Geurumil’ was also analyzed. The heading dates of the F2 plants in the MG population were determined in the experimental field at Fukui Prefectural University, Fukui, Japan in the 2013/2014 cultivation season. For genotyping of WPCL-A1 and WPCL-D1, PCR was carried out using following primer sets: WPCL-A1, 5′-CCAGCCAACCAGGCCC-3′ and 5′-TGATGCCGGATGATGTTAC-3′; WPCL-D1, 5′-CGGCAGGTGGGATGAC-3′ and 5′-TTGATGACTGAACTGGAC-3′. For genotyping of WPCL-B1, multiplex PCR was conducted using the following sets of primers: 5′-TCCGGATTGGTGTTTCGAGG-3′ and 5′-CATGATGGTTCTTGGGAC-3′, and 5′-ACGTAGTACTCCCTTGAGG-3′ and 5′-CAACCAGACCTATGACAC-3′. Thirty-four cycles of PCR were performed using KAPATaq Extra (Kapa Biosystems, USA) and following reaction conditions: 30 s at 94°C, 30 s at 60°C, and 45 s at 72°C. To detect allelic variation, the PCR products of WPCL-A1 and WPCL-D1 were treated with RsaI and BspT107I, respectively. For genotyping of Vrn-D1, two primer sets (Intr1/D/F-Intr1/D/R3 and -Intr1/D/R4) were used according to Yan et al. (2004) [35]. The amplified PCR products were separated by electrophoresis through a 1.0% or 1.5% agarose gel and stained with ethidium bromide.

Results

Reduced vernalization requirement and photoperiod sensitivity in ‘Chogokuwase’

‘Chogokuwase’ headed more than three weeks earlier than the parental cultivars ‘Minaminokomugi’ and ‘Geurumil’ in the field (Table 1). To investigate the vernalization requirement and photoperiod sensitivity of ‘Chogokuwase’, heading dates of ‘Chogokuwase’, ‘Minaminokomugi’ and ‘Geurumil’ were recorded under LD conditions without vernalization (V0-LD), LD conditions with 21 days vernalization (V21-LD), and SD conditions with 21 days vernalization (V21-SD). In all conditions, ‘Chogokuwase’ headed earlier than ‘Geurumil’ (Fig 1, panel (a)). ‘Chogokuwase’ had a reduced vernalization requirement compared with ‘Geurumil’, and the
Fig 1. Comparison of days to heading for ‘Chogokuwase’ and its parental cultivars as measured by unfolding of the flag leaf. (a) The days to unfolding of the flag leaf without vernalization treatment under long day conditions (V0-LD), after 21-days vernalization under long days (V21-LD) and after 21-days vernalization under short days (V21-SD). The different letters over the bars indicate statistically significant differences by Tukey Kramer’s HSD test (P < 0.05). (b) Comparison of the vernalization requirement among three cultivars as indicated by the ratio of V0-LD over V21-LD. (c) Comparison of the photoperiod sensitivity as represented by the ratio of V21-SD over V21-LD for the three cultivars.

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vernalization requirement of ‘Chogokuwase’ was as low as that of ‘Minaminokomugi’ (Fig 1, panel (b)). The photoperiod sensitivity of ‘Chogokuwase’ was similar to but lower than that of ‘Minaminokomugi’ (Fig 1, panel (c)).

‘Chogokuwase’ has a spring habit allele for VRN-D1 and an increased number of Ppd-B1 copies

Vrn-1 and Ppd-1 are the major genes that determine the vernalization requirement and photoperiod sensitivity, respectively. PCR analysis revealed that ‘Chogokuwase’ and ‘Minaminokomugi’ carried a VRN-D1 spring habit allele, whereas ‘Geurumil’ had a winter habit vrn-D1 (Table 2). The alleles of Vrn-A1 and Vrn-B1 loci in these three cultivars were winter habit types. The three cultivars had a photoperiod insensitive Ppd-D1a allele and photoperiod sensitive Ppd-A1b and Ppd-B1b alleles [33–34]. The photoperiod insensitive Ppd-B1a allele is associated with a higher number (2 to 4) of copies of Ppd-B1 [22–23]. Copy number estimation revealed that ‘Chogokuwase’ and ‘Minaminokomugi’ had three Ppd-B1 copies (S1 Fig) and an additional truncated copy (data not shown) as was reported for a Nepalese cultivar by Nguyen et al. (2013) [23].

Elevated transcript levels of Vrn-1 and WFT in ‘Chogokuwase’

Since the transcript abundance of Vrn-1 and WFT are often correlated with heading time in wheat [7], we examined the expression of these genes in ‘Chogokuwase’ and its parental cultivars. Vrn-1 transcript levels were significantly higher in ‘Chogokuwase’ than in the parental cultivars at all 4L stages under V0-LD, V21-LD and V21-SD conditions (Fig 2). Levels of Vrn-1 transcripts in ‘Geurumil’ were lower than in ‘Minaminokomugi’. WFT expression was up-regulated in ‘Chogokuwase’ especially under V21-LD conditions. Under the V0-LD and V21-SD condition, however, obvious difference was not observed in WFT transcript abundance between ‘Chogokuwase’ and ‘Minaminokomugi’. Vrn-1 transcript abundance was associated with the heading time rather than that of WFT.

Expression patterns of circadian clock and clock-output genes were altered in ‘Chogokuwase’

Diurnal expression patterns of Ppd-1, two wheat CONSTANS-like genes (WCO1 and TaHd1) and WFT were examined under SD conditions by quantitative RT-PCR. The transcript accumulation levels of Ppd-A1 in the early light period were higher in ‘Chogokuwase’ and ‘Geurumil’ than in ‘Minaminokomugi’ (Fig 3). A higher expression level of Ppd-B1 was observed in ‘Chogokuwase’ in the late dark period. Unlike Ppd-A1 and Ppd-B1, Ppd-D1 expression levels fluctuated, and no obvious differences in patterns among the three cultivars were evident. In ‘Chogokuwase’, the expression level of WCO1 during the dark period was low, whereas TaHd1 was up-regulated in ‘Chogokuwase’ and ‘Geurumil’ compared with ‘Minaminokomugi’. The expression level of TaLHY, a clock gene, in ‘Chogokuwase’ and ‘Geurumil’ was significantly lower than ‘Minaminokomugi’ at the peak of its expression (beginning of the day). In ‘Chogokuwase’ and ‘Geurumil’, on the other hand, transcripts of another clock gene TaTOC1 accumulated earlier than in ‘Minaminokomugi’. This early accumulation pattern was more evident for the third clock gene GIGANTEA (TaGI).

Sequence analysis of WPCL1

The expression patterns of clock and clock-output genes in ‘Chogokuwase’ and ‘Geurumil’ are similar to those in the einkorn wheat mutant lacking WPCL1 [25]. We determined the
nucleotide sequences of WPCL-A1, WPCL-B1 and WPCL-D1 in 'Chogokuwase' and the parental cultivars. A 142 bp deletion was observed in the region spanning the 5' untranslated region (UTR) and the first exon of WPCL-B1 in 'Chogokuwase' and 'Geurumil' (yellow) under V0-LD, V21-LD and V0-SD conditions. X- and Y-axis indicate days after 21 days-vernalization and relative transcript abundances, respectively. Transcript abundance was measured relative to the abundance of Actin gene transcripts. Bars on each observation indicate standard errors.

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Fig 2. Gene expression patterns of VRN-1 and WFT in 'Chogokuwase' and its parental cultivars. Expression levels were measured in the second (2L), third (3L) and fourth (4L) leaf stages of 'Chogokuwase' (blue), 'Minaminokomugi' (red), and 'Geurumil' (yellow) under V0-LD, V21-LD and V0-SD conditions. X- and Y-axis indicate days after 21 days-vernalization and relative transcript abundances, respectively. Transcript abundance was measured relative to the abundance of Actin gene transcripts. Bars on each observation indicate standard errors.

Further, a single nucleotide polymorphism (SNP) was found in WPCL-D1 of 'Chogokuwase' and 'Geurumil' in the SHAQKYF motif, a highly conserved region in the MYB domain [27] and whose consensus sequence in plant LUX-like genes is SHLQKY(R/Q). 'Minaminokomugi' and 'CS' had the SHLQKYR motif in WPCL-D1. The SNPs altered the amino acid sequence of this motif to CHLQKYR in WPCL-D1 of 'Chogokuwase' and 'Geurumil'. 'Chogokuwase', 'Minaminokomugi' and 'Geurumil' shared another
non-synonymous (K to N) substitution within the amino acid sequences of the SHAQKYF motif in WPCL-A1. The SHAQKYF motif is highly conserved in functional MYB transcription factors, and an amino acid substitution in this motif was associated with a loss-of-function mutation in the barley LUX/PCL1 gene HvLUX1 [27]. Thus, we assumed that the wpcl-A1 and

**Fig 3.** Gene expression patterns of circadian clock and clock-output genes under SD conditions in ‘Chogokuwase’ and the parental cultivars. White and black boxes indicate light and dark periods, respectively. Two-week-old seedlings were sampled every 3 hours over 24 h. Means ± standard deviations were calculated from data in three technical repeated experiments. Relative transcript abundance was calculated using the Actin gene as an internal control.

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alleles found in 'Chogokuwase' and 'Geurumil' were loss-of-function. In fact, the expression levels of wpcl-A1, wpcl-B1 and wpcl-D1 were consistently higher in 'Chogokuwase' and 'Geurumil' than in 'Minaminokomugi' (Fig 3). This finding is similar to that reported for the loss-of-function lux/pcl1 mutants in Arabidopsis and barley where LUX/PCL1 was up-regulated due to a feedback loop [27, 30].

wpcl-D1 alleles found in 'Chogokuwase' and 'Geurumil' were loss-of-function. In fact, the expression levels of wpcl-A1, wpcl-B1 and wpcl-D1 were consistently higher in 'Chogokuwase' and 'Geurumil' than in 'Minaminokomugi' (Fig 3). This finding is similar to that reported for the loss-of-function lux/pcl1 mutants in Arabidopsis and barley where LUX/PCL1 was up-regulated due to a feedback loop [27, 30].
Plants homozygous for the wpcl allele at three homoeoloci had an early heading phenotype

Segregation analyses in two F₂ populations segregating the WPCL1 loci (CN and MG populations) were used to test the association between the early heading phenotype and genotypes at the WPCL1 homoeoloci. In the CN population derived from a cross between 'Chogokuwase' (all loss-of-function wpcl1) and 'Norin 61' (all functional WPCL1), a triple recessive individual should appear in one out of 64 plants and individuals in the population should be homozygous at all Ppd-1 and Vrn-1 homoeoloci (Table 2, S1 Fig). Of the 114 F₂ plants, only three plants carrying homozygous wpcl1 at three homoeoloci headed as early as ‘Chogokuwase’ (Fig 5). The segregation ratio fit a 1:63 ratio (χ² = 0.85, P = 0.36). The other 111 F₂ plants were homozygous for a dominant allele or heterozygous at the WPCL1 locus and headed later than the three triple recessive plants. In the MG population, only alleles at the WPCL-B1 and WPCL-D1 loci segregated because ‘Geurumil’ and ‘Minaminokomugi’ have the wpcl-a1 allele. Of the 489 F₂ plants, 26 plants carrying the homozygous wpcl1 at the WPCL-B1 and WPCL-D1 loci were found. Of the 26 wpcl1 plants, eight plants headed as early as ‘Chogokuwase’ (Fig 6, panel (a)). The segregation ratio for early (n = 8) and late (n = 18) heading fit a 1:3 ratio (χ² = 0.462, P = 0.50), but also a 3:13 ratio (χ² = 2.47, P = 0.12), which does not rule out the presence of other loci controlling heading. We found that all the early heading plants were homozygous for the triple recessive wpcl1 homoeologues, and were vernalization-insensitive.
due to presence of dominant VRN-D1 allele (Fig 6, panel; (b)). In the wpcl1 triple recessive genetic background, the effect of copy-number variation at the Ppd-B1 locus on early heading is negligible as indicated by the segregation of the truncated copy of Ppd-B1 in five of the eight early-heading plants (data not shown).
Epistatic interactions between WPCL1 and vernalization genes

Day-neutral mutations in wheat WPCL1 and barley ELF3 up-regulate Vrn-2 expression, resulting in a stronger vernalization requirement [39]. Our quantitative RT-PCR experiment demonstrated that the Vrn-2 genes (ZCCT1 and ZCCT2) were expressed in ‘Chogokuwase’, ‘Minaminokomugi’ and ‘Geurumil’ (Fig 7), although Vrn-2 is generally not expressed under SD conditions [40–41]. Transcript abundance of Vrn-2 in ‘Chogokuwase’ and ‘Geurumil’ was higher than that in ‘Minaminokomugi’ during dark period. The expression level of Vrn-1 in the vrn-D1 allele carrier ‘Geurumil’ (Table 2) was extremely low, although Vrn-2 expression was comparable to that in ‘Chogokuwase’. These results suggest that the expression levels of Vrn-1 were determined by the genotype of the Vrn-D1 locus rather than by the expression levels of Vrn-2.

The recombinant inbred lines (RILs) resulting from a cross between a T. boeoticum accession with WPCL1 and Vrn-2 and an einkorn wheat mutant with deletions at both loci (denoted as ΔWPCL1 and ΔVrn-2) [25] made it possible to test epistatic interactions between WPCL1 and Vrn-2. Of the 62 RILs with ΔWPCL1, the RILs with ΔVrn-2 headed significantly earlier than those carrying Vrn-2 (Fig 8; P < 0.001). In contrast, among the RILs carrying WPCL1, there was no significant difference in heading time between the RILs with and without Vrn-2.

Discussion

‘Chogokuwase’ and ‘Geurumil’ contain loss-of-function natural variants of the WPCL1 gene

‘Chogokuwase’ is an extra-early heading cultivar that was derived from ‘Minaminokomugi’ and ‘Geurumil’ [33] and has a reduced vernalization requirement and lower sensitivity to photoperiod (Fig 1). The reduced vernalization requirement and photoperiod sensitivity of ‘Chogokuwase’ were derived from the parental cultivars ‘Minaminokomugi’ and ‘Geurumil’,
respectively. Vrn-1 and Ppd-1 are major genes determining the vernalization requirement and photoperiod sensitivity in common wheat [6, 17]. The vernalization insensitivity of 'Chogokuwase' could be explained by the dominant allele of Vrn-D1 that was inherited from 'Minamikomugi'. In contrast, Ppd-1 cannot be a candidate gene for the reduced photoperiod sensitivity of 'Chogokuwase' because 'Minamikomugi' and 'Chogokuwase' have the same genotypes for the three Ppd-1 homoeologues (Table 2, S1 Fig). The expression patterns of clock and clock-output genes in 'Chogokuwase' and 'Geurumil' were similar to those of an einkorn wheat mutant that lacks WPCL1 [25] and were different from those in 'Minamikomugi'. Sequence analyses revealed that 'Chogokuwase' and 'Geurumil' had non-synonymous substitutions in the SHAQKYF motif of both WPCL-A1 and WPCL-D1 (Fig 4, panel (b)). The SHAQKYF amino acid motif is conserved in all LUX-family genes, and amino acid changes within the SHAQKYF motif potentially lead to loss-of-function in barley [27]. In addition, a 142 bp deletion in the 5′ UTR and the first exon, which could result in a truncated protein or a frameshift mutation, was detected in WPCL-B1 of 'Chogokuwase' and 'Geurumil'. The clock genes in 'Chogokuwase' and 'Geurumil' had arrhythmic expression patterns, whereas the WPCL1 homoeologues were up-regulated (Fig 3). The lux/pcl1 mutants in Arabidopsis, barley and einkorn wheat showed the same expression patterns of these clock genes [25, 27, 30]. Taken together, these findings strongly suggest that 'Chogokuwase' and 'Geurumil' contain loss-of-function natural variants of the WPCL1 gene. Segregation analysis revealed that the homozygosity of recessive wpcl1 alleles at all homoeologs is a prerequisite for heading as early as 'Chogokuwase' (Figs 5 and 6). Segregation of F_{2S} in the MG population suggested that wpcl1 mutations are insufficient to explain the exceptionally early heading of 'Chogokuwase'. A fraction of the segregants with the triple recessive wpcl1 headed as late as those with a functional WPCL1, indicating the presence

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Fig 8. Relationship between days to heading and genotypes of the WPCL1 and Vrn-2 loci in the recombinant inbred lines of einkorn wheat. The days to heading in 89 recombinant inbred lines derived from a cross between T. monococcum mutants and T. boeoticum were compared. The parental monococcum accession lacks WPCL1 and Vrn-2, whereas the boeoticum accession has WPCL1 and Vrn-2. Arrows indicate days to heading of the parental T. monococcum (KT3-5) and T. boeoticum (KT1-1) accessions, respectively.

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of another gene or genes that condition(s) the early heading phenotype. In the CN population, on the other hand, wpc1l mutations were sufficient to explain the earliness of ‘Chogokuwase’, suggesting that the hypothetical gene or genes is/are present in both ‘Chogokuwase’ and ‘Norin 61’ and are not segregating in the CN population. Alternatively, the number of days to heading may be unaffected by the hypothetical gene or genes in SD conditions.

**Loss-of-function mutations in WPCL1 reduce photoperiod sensitivity by up-regulating Ppd-1**

In wheat and barley, Ppd-1, the homologue of Arabidopsis PRR7 and PRR3, has been regarded as the main regulator of photoperiod sensitivity [17, 42]. The photoperiod insensitive alleles of Ppd-1 (Ppd-1a) invoke the mis-expression and/or increased expression of Ppd-1 [20]. Thus, the pattern and level of Ppd-1 expression are critical in regulating photoperiod sensitivity. The mutants of circadian clock components lux/pcl1 and elf3 in barley and wheat have increased levels of Ppd-1 expression [24–28]. In Arabidopsis, LUX and ELF3 bind to the promoter of PRR9 and repress its expression [30, 43]. Therefore, we hypothesized that WPCL1 might function as the repressor of Ppd-1 (S2 Fig) [25]. In the present study, we found up-regulated expression of Ppd-A1 and Ppd-B1 in ‘Chogokuwase’ and ‘Geurumil’ where the functional WPCL1 is absent (Fig 3). The expression pattern of Ppd-B1 in ‘Chogokuwase’ was different from that in ‘Geurumil’. This difference might be reflecting the copy number variation at the Ppd-B1 locus. Unlike Ppd-A1 and Ppd-B1, there were no significant differences in expression patterns and levels of Ppd-D1 transcripts between wpc1l mutants (‘Chogokuwase’ and ‘Geurumil’) and the cultivar with two functional WPCL1 genes (‘Minaminokomugi’). All three cultivars carried the Ppd-D1a allele, which has a 2 kb deletion in the promoter region [33]. The 2 kb deletion overlaps with the positions of mutations in Ppd-A1a and Ppd-B1a [18–19, 21]. These observations suggest the 2 kb region, which includes a highly conserved ca. 100 bp sequence where putative cis-elements and regulatory motifs were predicted at the nucleotide sequence level, plays a critical role in Ppd-1 regulation. We hypothesize that WPCL1, a member of the Myb transcription factor family, might bind to the promoter region of Ppd-1 and repress Ppd-1 expression. We will directly test this hypothesis in the future by conducting protein-DNA interaction assays such as chromatin immunoprecipitation experiments.

**WPCL1 affects the vernalization requirement through Vrn-2**

Day-neutral mutations such as Ppd-D1a and ema8 increase the expression level of Vrn-2 [39]. We confirmed that wpc1l mutations up-regulated Vrn-2 expression (Fig 7). These observations suggested that photoperiodic signals were linked with the vernalization pathway via Vrn-2 (S2 Fig). In fact, Vrn-2 has a CCT domain that is found in proteins involved in light signal transduction [9]. ‘Geurumil’ is much more sensitive to vernalization than ‘Minaminokomugi’, although there was only a slight difference in the number of days to heading when plants were grown in the field where plants were fully vernalized (Table 1, Fig 1). The stronger vernalization requirement in ‘Geurumil’ is likely due to both up-regulation of Vrn-2 and the absence of dominant VRN-D1 alleles. Supporting evidence was reported in barley that overexpression of Vrn-2 delayed heading and decreased the expression levels of HvFT1 [44]. Therefore, the observed up-regulation of Vrn-2 in wpc1l mutants could explain the delayed heading of ‘Geurumil’. We also showed that the absence of Vrn-2 loci is crucial for early heading in the wpc1l mutants (Fig 8). ‘Chogokuwase’ exhibited vernalization-insensitive and early-heading phenotypes despite the up-regulated expression of Vrn-2. ‘Chogokuwase’ had higher Vrn-1 expression that was due to the spring habit allele of VRN-D1 (Fig 7). All of the early heading F2 plants in the MG population had the dominant VRN-D1 allele (Fig 6). Our preceding experiment
demonstrated that Vrn-1 expression levels in leaves were associated with earliness [7]. Therefore, VRN-D1 might be a candidate gene that confers early heading in ‘Chogokuwase’, a hypothesis that remains to be tested. Our observation that all the plants in early heading subgroup were homo- or heterozygous VRN-D1 alleles (Fig 6, panel (b)) indicates that vernalization-insensitivity is prerequisite for the ‘Chogokuwase’-type of early heading in the genetic backgrounds of the MG population. Ppd-1 is known as a major gene controlling heading time in common wheat. Our study indicates that loss-of-function wpcl1 accelerates heading time by up-regulating all Ppd-1 homoeologues. Although wpcl1 mutations could accelerate heading time, the effect of earliness is likely attenuated by Vrn-2 up-regulation.

The timing of flowering is one of the most important traits for breeding, and early-heading cultivars are desired to avoid pre-harvest sprouting and associated poor grain quality. However, extra-early heading often reduces crop yield. In fact, ‘Chogokuwase’ is short and slender in appearance and the number of tillers is greatly reduced. In this study, we demonstrated that a loss-of-function mutation in WPCL1 may accelerate and adjust flowering time due to the combination of vernalization gene activities in common wheat.

Supporting Information

S1 Fig. Copy number variation of the Ppd-B1 sequences in seven cultivars. Relative mean values of PCR amplification levels of the Ppd-B1 copy are shown with the standard deviation. (TIF)

S2 Fig. A revised model for the interaction of flowering-time genes in wheat. The role of WPCL1, whose orthologue in Arabidopsis forms the “Evening Complex” with ELF3 [29], as a repressor of Ppd-1 was added to the model proposed by Shimada et al. 2009 [13]. In their model, the triangle of Vrn1–WFT–Vrn-2 gene interactions regulate the phase transition from vegetative to reproductive phase in wheat. WFT plays an integrative role in both the WCO1-related photoperiod pathway and the VRN1-related vernalization pathway. The expression levels of Vrn-2 were up-regulated in ‘Chogokuwase’ that is triple homozygous for the loss-of-function alleles of wpcl1, indicating that WPCL1 also repress Vrn-2 directly and/or through promotion of Ppd-1 expression (indicated by dashed lines). (TIF)

S1 Table. Primers used for genotyping of Vrn-1. (DOCX)

S2 Table. Primers used for gene expression analyses. (DOCX)

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