Morphogenetic and patterning defects explain the coloboma phenotype of the eye in the Mexican cavefish

Lucie Devos¹, Florent Klee¹, Joanne Edouard², Victor Simon¹,², Laurent Legendre², Naima El Khallouki², Maryline Blin¹, Sosthène Barbachou², Frédéric Sohm² and Sylvie Rétaux¹

¹ Paris-Saclay Institute of Neuroscience, CNRS UMR 9197, Université Paris-Saclay, 91198 Gif sur Yvette, France
² UMS AMAGEN, CNRS, INRA, Université Paris-Saclay, 91198, Gif sur Yvette, France

Key words (6): Zic1; CRISPR/Cas9 knock-in; Astyanax mexicanus; live imaging, retina patterning, retinal pigmented epithelium
Abstract
The morphogenesis of the vertebrate eye consists of a complex choreography of cell
movements, tightly coupled to axial regionalization and cell type specification processes. Any
disturbance in these events can lead to developmental defects and blindness. Here we have
deciphered the sequence of defective events leading to coloboma phenotype in the
embryonic eye of the blind cavefish of the species *Astyanax mexicanus*. Using comparative
live imaging on targeted enhancer-trap $Zic1: hsp70: GFP$ reporter lines of both the normal,
river-dwelling morph and the cave morph of the species, we identified major defects in initial
optic vesicle size and optic cup invagination in cavefish. Combining these results with gene
expression analyses, we also discovered defects in axial patterning affecting mainly the
temporal retina, in optic stalk tissue specification, and in the spreading processes involving
the retinal pigmented epithelium cells. Based on these results, we propose a developmental
scenario to explain the cavefish phenotype and discuss developmental constraints to
morphological evolution. The cavefish eye appears as an outstanding natural mutant model
to study molecular and cellular processes involved in optic region morphogenesis.
Introduction

The morphogenesis of the vertebrate eye is a complex choreography of cell movements starting from a flat neural plate and leading to the formation of a spherical multi-layered structure. Owing to technological improvements, this process has been increasingly investigated in the last decade, especially on teleost models which are amenable to live imaging due to their external development and transparency (England et al., 2006; Ivanovitch et al., 2013; Kwan et al., 2012; Martinez-Morales et al., 2009; Nicolas-Perez et al., 2016; Picker et al., 2009; Rembold et al., 2006; Sidhaye and Norden, 2017). This focus on morphogenesis led to the description of cell and tissue movements during eye development in fish (reviewed in Cavodeassi, 2018) (Fig. S1).

At the end of gastrulation, the eyefield is specified in the anterior neural plate, surrounded anteriorly and laterally by the prospective telencephalon, and posteriorly by the future hypothalamus and diencephalon (Varga et al., 1999; Woo and Fraser, 1995; Woo et al., 1995). During neurulation, the first step of eye formation is the lateral evagination of the optic vesicles. Already at this step, cell behaviours are complex as some eye-fated cells behave like the nearby telencephalic cells and converge toward the midline to form the neural keel, while others lag behind and keep the eyefield wide (Ivanovitch et al., 2013; Rembold et al., 2006). The eye vesicles then elongate due to a flow of cells entering the anterior/nasal optic vesicle, in a process recently re-described as “extended evagination” (Kwan et al., 2012). Simultaneously, the optic vesicles are separated from the neural keel by the anterior-wards progression of a posterior furrow between them and the diencephalon, leaving a connection with the neural tube at the optic stalk (England et al., 2006). Cells from the medial part (inner leaflet) of the optic vesicle then migrate around the rim of the eye ventricle (the optic recess) into the lens facing neuroepithelium through a process called rim movement (Heermann et al., 2015; Kwan et al., 2012). The cells in the outer layer of the optic cup, fated to the retinal pigmented epithelium (RPE), expand and flatten to cover the back of the retina (Cechmanek and McFarlane, 2017; Heermann et al., 2015). Together with the basal constriction of lens-facing epithelial cells (Martinez-Morales et al., 2009; Nicolas-Perez et al., 2016), these movements lead to optic cup invagination. The invagination process leads to the formation of
the optic fissure at the level of the connection of the eye with the optic stalk. This fissure allows blood vessels to invade the eye and leads the way of retino-fugal axons, but needs to close to have a functional and round eye (Gestri et al., 2018). Finally, the entire eye, together with the forebrain, rotates anteriorly, bringing the optic fissure in its final ventral position. Hence, cells that are initially located in the dorsal or ventral part of the optic vesicles contribute to the nasal or temporal quadrant of the retina, respectively (Picker et al., 2009). Failure to correctly complete any of these steps can lead to vision defects; for example, failure to properly close the optic fissure is termed coloboma and can lead to congenital blindness.

During the eye morphogenetic process, three types of tissues emerge: (1) the neural retina, facing the lens and composed of various neuronal types, (2) the RPE at the back of the neural retina, with multiple functions including nurturing of photoreceptors (Strauss, 2018), and (3) the optic stalk, transiently connecting the retina to the neural tube. This thin ventral structure is invaded by the ganglionic cells axons and guides them on their way to the tectum. Optic stalk cells then differentiate into reticular astrocytes surrounding the optic nerve (Macdonald et al., 1997). All these tissues derive from the optic vesicles and have a neural origin.

Concomitantly to morphogenesis, retinal cells acquire axial positional identity. Indeed, the visual sense requires a topographic perception of the light stimuli and processing of the signal to form images. In fishes, the neuronal map of the retina is replicated onto the contralateral tectum, in a symmetrical manner so that the nasal retina projects to the posterior optic tectum, while the temporal retina projects to the anterior tectum. This retinotopy requires a precise regional identity of both the retina and the tectum for a proper matching (Rétaux and Harris, 1996; Sperry, 1963). In the retina, several transcription factors such as Vax2, FoxG1, Tbx5a, or FoxD1 present a strong regional expression, already present at optic vesicle stage, and define the future quadrants identity by controlling the expression of effector guidance molecules (French et al., 2009; Picker et al., 2009; Sakuta et al., 2006; Schulte et al., 1999). The regionalized expression of the transcription factors themselves is mainly achieved through Hedgehog and Fgf signalling from the embryonic midline, and reciprocal interactions between them (Asai-Coakwell et al., 2007; Hernandez-Bejarano et al., 2015; Kruse-Bend et al., 2012; Picker and Brand, 2005; Picker et al., 2009). Shh is involved in the regulation of the ventral and temporal fates while Fgf secreted by the anterior neural ridge and olfactory placode seem to be involved in nasal specification. Extra-ocular Bmps direct the acquisition of dorsal fates.
Astyanax mexicanus is a teleost fish that arises in two morphs: the classical river-dwelling eyed morph and the cave-dwelling blind morph. Although eyes are absent in adult cavefish, they first develop in embryos before degenerating during larval stages. The early cavefish eyes display several morphogenesis abnormalities: the optic vesicles are short (Alunni et al., 2007), the optic cup and lens are small (Hinaux et al., 2015; Hinaux et al., 2016; Yamamoto and Jeffery, 2000) and the ventral part of the optic cup is severely reduced or lacking, leaving the optic fissure wide open (coloboma phenotype) (Pottin et al., 2011; Yamamoto et al., 2004). Moreover, cavefish exhibit several modifications of morphogen expression including an expanded Shh expression at the anterior midline, a heterochrony of Fgf8 onset in the anterior neural ridge, and variations of Bmp4 and Dkk1b expression in the prechordal plate (Hinaux et al., 2016; Pottin et al., 2011; Torres-Paz et al., 2018; Yamamoto et al., 2004). These morphogen alterations trigger modifications of the cavefish eyefield and subsequent eye, as evidenced by modified expression of Lhx9/2 or Pax6 in the neural plate when compared to surface fish embryos (Pottin et al., 2011; Yamamoto et al., 2004), and they have been linked to cavefish eye development defects: overexpression of Shh in surface fish shortens its optic cups and triggers the apoptosis of the lens, while inhibition of Fgf signalling in a cavefish restores the ventral retina (Hinaux et al., 2016; Pottin et al., 2011; Yamamoto et al., 2004).

Because of these various modifications, the cavefish is a remarkable natural mutant model to study eye development, beyond the mechanisms of eye degeneration and loss. We therefore undertook to study the morphogenesis and regionalization of the cavefish eye, in comparison with surface fish, with two aims: a better understanding of the defects of the cavefish embryonic eye as well as the mechanisms of eye morphogenesis in general.

Using CRISPR/Cas9-mediated targeted enhancer trap cavefish and surface fish Zic1:hsp70:GFP lines, we performed comparative live imaging of eye morphogenesis on developing embryos. We propose that the reduction in cavefish optic vesicle size, associated with a proper extended evagination, leads to a bias towards nasal fate and an abnormal final ventral position of the lens. In addition, optic cup invagination is impaired and RPE spreading over the surface of the neural retina is delayed in cavefish. Through gene expression analyses, we show...
moreover that the cavefish eyes display modifications of their axial regionalization, with a tendency to preserve or increase nasal and dorsal fates while strongly diminishing temporal fate. Finally, the optic stalk fate is widely increased throughout the retina.
Results

Screening candidate genes to establish *Astyanax* GFP reporter lines

To perform live imaging and label regions of interest, i.e. the entire optic region of cavefish (CF) and surface fish (SF) embryos, we sought to find a reporter gene that would be expressed in the eyefield from the neural plate stage (10hpf) until at least 24 or 36hpf, when optic recess region (ORR) and retina are clearly separated (Affaticati et al., 2015). An *in situ* hybridization mini-screen for candidate genes selected from publications and the ZFIN expression database was performed. Chosen candidates were *Vax1*, *Vax2* (Takeuchi et al., 2003), *Zic1* (Hinaux et al., 2016; Maurus and Harris, 2009; Rohr et al., 1999; Tropepe et al., 2006), *Zic2a* (Sanek et al., 2009), *Rx3* (Deschet et al., 1999; Rembold et al., 2006; Stigloher et al., 2006), *Lhx2* and *Lhx9* (Pottin et al., 2011). We performed *in situ* hybridization on CF and SF embryos at 5 different stages (10, 12, 14, 24 and 36hpf) (Fig. S2A).

Among the 7 genes, 5 were expressed in the anterior neural plate at 10hpf while 2 were not: *Vax1* and *Vax2*, whose expressions were detectable from 12hpf only (Fig. S2A). Five of them were expressed at least partially in the optic vesicle per se (excluding ORR and optic stalk): *Vax2*, *Zic1*, *Rx3*, *Lhx9* and *Zic2a* (faintly). At 36hpf, only 4 of them were still expressed in the optic cup: *Zic2a* and *Zic1* (around the lens), *Lhx2* (faintly) and *Vax2* (in the ventral retina). Subtle differences between CF and SF expression patterns were observed (not shown), and only one candidate genes was consistently expressed in the eye from neural plate to 36hpf: *Zic1* (Figure 1A and Fig. S2B; see legend of Fig. S2B). Even though the *Zic1* pattern was complex and encompassed a region wider than the optic region of interest, it was chosen for transgenesis due to its early and persistent expression throughout the eye and the ORR/optic stalk regions.

Comparative expression of *Zic1* in surface fish and cavefish embryos

A closer examination of *Zic1* pattern highlighted some patterning and morphological differences between SF and CF (Figure 1A). In the anterior neural plate at 10hpf, *Zic1* expression was wide in the bilateral eyefield in SF, with a medial indentation (Figure 1A,
asterisk); in CF, Zic1 was expressed in narrower lateral bands, with a wider staining anteriorly. At 12hpf, Zic1 pattern confirmed that the CF optic vesicles were shorter than those of SF, but they also looked more “plump”. Indeed, from a dorsal view SF optic vesicles were elongated, slender and pointier while CF optic vesicles looked rounder. On lateral views, the optic vesicles showed a trapezium shape in SF and an oval shape in CF. At 36hpf the Zic1-expressing ORR was wider in cavefish. These differences in expression patterns decisively convicted us to use Zic1 as reporter gene.

Establishing Zic1:hsp70:GFP surface fish and cavefish knock-in lines

We used a targeted enhancer-trap strategy into the Zic1 locus to generate reporter lines. This option presented the advantage that the GFP reporter insertion site would be identical in CF and SF lines and avoid positional effects, which is crucial for comparative purposes. The genomic region around Zic1 was examined to find conserved elements that might point toward putative regulatory elements (Fig. 1BC). The region was large (~500 kb in zebrafish) and complex. In the zebrafish genome which has a better quality and annotation than the Astyanax genome (McGaugh et al., 2014), Zic1 and Zic4 were very close to each other in a head to head configuration, and located in the middle of a gene desert (~200kb downstream of Zic1 and ~220kb downstream of Zic4 in zebrafish, and ~275kb downstream of Zic1 and ~235kb downstream of Zic4 in Astyanax). This gene desert contained many elements conserved among fishes, also partly conserved with tetrapods and mammals (Fig. 1BC). Such a regulatory landscape suggested that the elements driving Zic1 expression are probably modular and difficult to identify, further strengthening the choice of a directed enhancer-trap approach.

Cavefish and surface fish embryos were co-injected with sgRNA2 (targeting the region between conserved non-coding elements 1 and 2), Cas9 protein and a linearized minimal promoter hsp70:GFP repair construct and were screened at 30hpf for fluorescence patterns consistent with Zic1 endogenous pattern (Fig. 1CD). Excellent Zic1 pattern recapitulation in F0 was observed at low frequency (1-2% of injected embryos), while other more partial patterns were also seen at higher frequencies. All potential founder embryos were sorted and raised until males were sexually mature (5-6 months’ post-fertilization) and could be screened by individual in vitro fertilization. We detected 3 founders for SF (out of 15 F0 males screened)
and 5 founders for CF (out of 9 F0 males screened) with various transmission rates: 4%, 7% and 30% for SF founders and 4%, 45%, 48%, 50% and 54% for CF founders, respectively. The fish were screened based on their GFP pattern, matching Zic1 (Fig. 1D). In both morphs some variations in relative intensities of fluorescence were observed, with some lines exhibiting more homogeneous levels of expression and others having an extremely strong GFP fluorescence in the telencephalon and a dimer fluorescence in the eye. We focused on the most homogeneous lines for imaging purposes. Importantly, in those lines, genomic analyses confirmed the proper insertion of the transgene at the exact targeted site (Fig. S3), and double fluorescent in situ hybridisation for Zic1 and GFP mRNAs demonstrated that the reporter fully recapitulated the endogenous Zic1 pattern at the stages of interest (Fig. 1E).

Comparing eye morphogenesis in surface fish and cavefish through live imaging

Live imaging was performed on a light-sheet microscope on both CF and SF lines from ~10.5hpf to 24-30hpf, depending on the acquisitions (Fig. 2 and Movies 1 and 2). The planes used for analysis were chosen to always cross the middle of the lens and the optic stalk (see drawings on Fig. 2), in order to follow the anterior rotation of the eye. Overall, eye morphogenesis in SF recapitulated step by step the events described in zebrafish, while in CF the morphogenetic movements were conserved but their relative timing and extent seemed different. The following analyses result from quantifications made on 4 eyes for each morph.

Evagination and elongation. The CF optic vesicles were much shorter from the beginning of evagination onwards and they spanned about half the length of the SF optic vesicles at the same stage (139µm compared to 216µm at 11.5hpf) (Fig. 2A,B and Fig. 3A,B). Elongation then proceeded at approximately the same pace as in SF until 17.5hpf (Fig. 3B). However, while the length of the eye primordium decreased between 17.5hpf and 25.5hpf in SF, due to the invagination movement, elongation continued at slower pace until 25.5hpf in CF. Hence, the increase in length from 11.5hpf (beginning of the furrow progression to separate the optic vesicles from the neural tube in both morphs) to 31.5hpf was more important in CF than in SF, because it remained flatter (Fig. 3B). Moreover, the final size of the SF optic cup was very similar to that of its early evacuating eyefield (252µm at 31.5hpf compared to 240µm at
10.5hpf) while in CF a net increase in the optic primordium size was visible (186µm at 31.5 hpf compared to 146µm at 10.5hpf) (Fig. 3A). In addition, while in SF the optic vesicles stayed closely apposed to the neural tube, in CF they first started growing away from the neural tube before getting back closer between 18.5 and 21.5hpf (Fig. 2B). Finally, throughout development, the width of the optic stalk (defined in its wide meaning as the connection between the optic vesicle/cup and the neural tube) was similar in the two morphs despite an initially smaller size in the CF due to the smaller optic vesicles (Fig. S4).

**Optic cup invagination and lens formation.** The posterior end of the optic vesicles started curling back in both CF and SF around 15.5hpf, probably due to basal constriction. The lens started being identifiable at 17.5hpf in both morphs (Fig. 2B and movies 1 and 2). At this stage, the lens was in a central position with regard to the antero-posterior extension of the optic vesicles, in both morphs (Fig. 2B and 3D). Then, in SF the invagination quickly brought closer the two edges of the optic cup, in contact with the lens (Fig. 2B and Fig. 3C). In contrast, despite initially harbouring a curvature typical of an invaginating optic cup, the edges of the CF optic cup did not undergo the same change of form and stayed “flat”, with an apparent impairment of the rim movement in the posterior optic cup (Fig. 2B, Fig. 3C and Movie 2). In fact, the CF optic vesicles continued to elongate while the lens remained static, therefore seemingly shifting the lens position anteriorly (Fig. 2B and Fig. 3D). During this period, the posterior part of the CF optic cup showed slow and reduced curling, which in some embryos led to a separation of the posterior optic vesicle from the lens. Yet the posterior (prospective dorsal) optic cup finally curved and contacted the lens (see progression, especially on the right eye from 22.5hpf to 30.5hpf, Movie 2; Fig. 2B and Fig. 3D). However, the CF optic cup remained shallower as its lens, albeit smaller, always bulged out while in SF the lens was always contained inside of the optic cup curvature.

In sum, live-imaging experiments suggested that in CF (1) the optic vesicles were reduced in size after the initial evagination, (2) elongation of the optic vesicle occurred properly, including for the extended evagination, while (3) invagination was transiently compromised. We reasoned that these impaired morphogenetic movements should impact the proper patterning and regionalization of the CF retina.
Comparing eye regionalization in surface fish and cavefish

In order to label the different quadrants of the eye, four classical markers were chosen: FoxG1 for the nasal quadrant, Tbx5a for the dorsal quadrant, FoxD1 as a temporal marker and Vax2 as a ventral marker spanning both the nasal and temporal sides of the optic fissure (Fig. 4).

The difficulty of comparing these quadrants between CF and SF resided in the difference of morphology and size of their eyes. We therefore decided to measure different parameters including angles of expression (taking as a reference the centre of the lens and the middle of the optic fissure if needed) and gene expression areas (expressed either as absolute values or as relative values normalized to eye size). Below the expression patterns of the four markers are described in a clock-wise manner starting at the optic fissure.

The nasal marker FoxG1 presented a larger angle of expression in CF (149°) than in SF (132°) due to a dorsal expansion of the expression domain (Fig. 4A-A’). In CF the dorsal marker Tbx5a started at the same angle from the optic fissure but spanned an increased angle towards the temporal retina (angle to mid-fissure: 106° for SF and 111° for CF; span: 108° for SF and 139° for CF) (Fig. 4B-B’). Reciprocally to these two increases of expression domains in the clock-wise direction, the temporal marker FoxD1 span was reduced in its dorsal part in CF (179° in SF, 141° in CF) (Fig. 4 C-C’). Finally, the ventral marker Vax2 had different features on the nasal and temporal margins of the optic fissure. Vax2 span was increased in the ventro-nasal quadrant, indicating a dorsal-wards or clock-wise expansion (66° in SF, 94° in CF) while it was unchanged in the temporal quadrant when compared to SF (Fig. 4 D-D’).

The smaller size of the CF eye was reflected by the absolute values of markers expression areas (Fig. 4E). Indeed, all of them but one (Tbx5a) were decreased in size, including nasal Vax2 and FoxG1 expression domains which were slightly but significantly reduced. Tbx5a was the only gene showing the same area of expression in CF and SF eyes, suggesting that the dorsal quadrant was proportionately increased in cavefish. The two temporal genes, FoxD1 and Vax2, exhibited a strong reduction of expression area, pointing to a temporal reduction in the cavefish eye. This conclusion was further supported when the gene expression areas were expressed in relative values normalized to eye size (Fig. 4F) or in percentages of SF labelling (Fig. 4G). Indeed, the most strongly reduced quadrant was the temporal quadrant, labelled by FoxD1 and the temporal aspect of the Vax2 domain.
Thus, in CF all markers examined presented modifications of expression in a fan-opening fashion, from nasal towards temporal, overall increasing the nasal and dorsal fates at the expense of the temporal retina. Contrarily to the usual description of the cavefish eye as ventrally-reduced, we unmasked here a temporal reduction.

**Comparing tissue and cell identity markers in surface fish and cavefish**

To assess tissue identity in the developing retina, Pax2a was used as a marker for optic stalk and optic fissure margin identity (Macdonald et al., 1997) and Bhlhe40 (Cechmanek and McFarlane, 2017) was used as a RPE identity marker.

Pax2a showed wider expression in cavefish, expanding beyond the optic fissure margins and occupying a large part of the ventral quadrants both nasally (angles, SF: 36°, CF: 67°) and temporally (angles, SF: 21°, CF: 55°) ([Fig. 5A-A”](#)). Surprisingly, in CF eyes Pax2a expression sometimes expanded throughout the retina with a lighter, although specific, expression intensity. Staining was even sometimes present in the dorsal quadrant, opposite to the optic stalk or optic fissure margins without obvious staining in between (see [Fig. 5A’]). These “dorsal” phenotypes were observed in 60% of CF embryos, but never in SF.

In 36hpf SF embryos the RPE marker Bhlhe40 was expressed all around the eye, often contacting the lens (4.7µm away) ([Fig. 5B, B’, D]), which was taken as an indicator of the correct engulfment of the retina by the migrating RPE. The edges of the optic fissure margins sometimes lacked staining but overall the expression spanned 326° around the eye ([Fig. 5B”]). Conversely, in CF, Bhlhe40 expression area was reduced, with a wider ventral gap possibly resulting from the wider optic fissure opening, and a significantly diminished covering of the retina by the RPE (289° around the eye). Bhlhe40-positive cells were also found further away from the lens (15µm), reinforcing the idea of a reduced retina covering by the RPE at this stage; at 48hpf however, the staining span was increased and no longer significantly different from the 36hpf SF (CF 48hpf: 309°; SF 36hpf: 326°) ([Fig. 5B, C, B”]). These data suggest a slower but still occurring engulfment of the back of the retina by the RPE.

The back of the retina was usually well covered with Bhlhe40-expressing RPE cells, but with a ventral gap which was well delimited by strongly contrasted, sharp edges ([Fig. S5]). We interpreted this gap as the “exit point” of the optic stalk and used it as a proxy for optic stalk
width. The *Bhlhe40* gap was wider in CF than in SF (SF: 41µm, CF: 60µm) (Fig. 5E). This was consistent with the above-reported increase in *Pax2a* expression domain and points towards an increased optic stalk size in cavefish.

Altogether, these results suggested that both the RPE engulfment/spreading movement and the optic fissure margins juxtaposition are slowed -but occur in a delayed manner- in cavefish. Below these findings are discussed altogether with regards to earlier observations of eye morphogenesis through live imaging.
Discussion

**Astyanax knock-in reporter lines obtained by CRISPR/Cas9**

Several transgenesis techniques are available in zebrafish and have already been adapted to Astyanax: a classical approach consists in cloning a promoter or regulatory element driving the expression of a reporter or effector gene under its control, flanked with either transposons or meganuclease cutting sites (Kawakami, 2005; Thermes et al., 2002). After injection with the appropriate enzyme, transposase or meganuclease, into the 1st cell of the embryo, the construct is randomly inserted one or several times in the genome. This method is technically simple and works in Astyanax (Elipot et al., 2014; Hinaux et al., 2015; Stahl et al., 2019). It has however several drawbacks: the correct identification of most if not all the regulatory sequences driving expression is necessary and often difficult; the insertion of the transgene is random and its expression depends on the insertion site; the insertion of the construct can disrupt coding or regulatory sequences.

The advent of the flexible CRISPR/Cas9 technique now allows for an “easy” RNA-mediated targeting at a precise genomic location. However, if performing a targeted knock-out by cutting DNA and relying on the imprecise non-homologous end joining (NHEJ) DNA repair mechanism to generate indels and frameshifts has proven quite efficient, more precise repair and insertions are still difficult to obtain, at least in zebrafish. The more precise methods require homology-directed repair and therefore involve homology arms flanking a repair construct. Although different ways to increase homology-directed repair efficiency have been tested (Albadri et al., 2017; Morita et al., 2017), it is still a very challenging method as NHEJ is the preferred repair mechanism in fish embryos (Hagmann et al., 1998).

The difficulty of identifying Zic1 regulatory elements with comparative genomics led us to adopt a targeted enhancer-trap strategy, using a NHEJ DNA repair mechanism-based approach to maximize integration efficiency. This option also had the advantage that the insertion site would be similar in CF and SF lines, which is crucial for comparative purposes. Enhancer traps were originally performed by random insertions in a two-step process, the first step being the selection of the expression pattern of interest, the second the identification of the region in
which the transgene was inserted. This allowed the generation of transgenic lines with various patterns of expression, reflecting the activity of one or more enhancers and regulatory elements. Here, we “addressed” the enhancer-trap construct to Zic1 downstream genomic region using CRISPR/Cas9 methodology, similarly to what was performed by Kimura and colleagues (Kimura et al., 2014). This method yielded good results as its limited efficiency was compensated by the possibility of using a pattern-based fluorescence screening in F0 embryos. Hence, we obtained an excellent ratio of founder fish in the pool of selected F0 embryos (more than 50% in cavefish). Finally, for both morphs the different Zic1:hsp70:GFP lines, although recapitulating the Zic1 pattern, showed slight variations in the relative intensities of reporter fluorescence in the telencephalon and in the eyes. The insertion method being based upon non conservative NHEJ mechanism, those variations are likely due to sequence differences from one line to the next (indels and duplications in the genomic DNA or the transgene), which may affect the nearby regulatory sequences and slightly modify transgene expression. However, those variations remain anecdotal compared to the differences observed between lines generated by traditional transgenesis techniques (such as Tol2 promoter transgenesis), validating this approach as a tool to follow gene expression in Astyanax morphotypes.

CRISPR/Cas9 has been previously reported in surface Astyanax mexicanus to generate an Oca2 null mutant and confirm the role of Oca2 in the control of pigmentation (Klaassen et al., 2018). But this is to our knowledge the first report of the CRISPR/Cas9 technology used in this emergent model species to generate identical reporter lines in the two morphotypes, and in a targeted genome edition perspective.

**Live imaging**

The choice of live imaging microscopy technique was directed by several parameters, from the intensity of the labelling to temporal-spatial resolution trade-off. Light-sheet or SPIM (single plane illumination microscopy) microscopy offered several advantages such as wide dynamic range of the camera, allowing to detect both the strong labelling of the telencephalon and the fainter labelling of the eye. The next step of this study will be to track the cells participating in the formation the optic region, i.e. eye, optic stalk, ORR. Cells in these regions
undergo fast movements during early morphogenesis, so that a good time resolution is necessary. They are also densely-packed and therefore require a good spatial resolution. Moreover, live embryos are fragile and laser power must be kept minimal to avoid bleaching of the fluorescence and photo-damage. In contrast to the point acquisition of the confocal microscope, the plane acquisition of the SPIM allowed fast imaging while retaining sufficient spatial resolution. The orthogonal illumination induced minimal photo-damage and embryos developing for more than 20 hours under the microscope recovered well and were alive with a normal head shape at 48-60hpf - even though the tail was usually twisted due to the mechanical constraint in the low-melting agarose.

Recapitulating cavefish eye development

Live imaging performed on the Zic1:hsp70:GFP transgenic lines combined to gene expression studies allowed us to reveal striking differences in eye morphogenesis, morphology and patterning between the two Astyanax morphs. Because the ventral quadrant of the eye was originally described as reduced and because the expression of the ventralizing morphogen Shh was known to be enlarged in cavefish (Yamamoto et al., 2004), we expected a global “ventralization” of the eye quadrants at the expense of the dorsal quadrant which we assumed would be reduced. However, the data revealed a quite different story and we venture to propose a developmental scenario for cavefish morphogenesis and patterning (Fig. 6).

(1) First, the shorter size of the cavefish optic vesicles (Alunni et al., 2007; Strickler et al., 2001) seems principally due to the smaller eyefield, since elongation proceeds similarly in CF and SF until 17.5hpf. Increased Hedgehog signalling in CF was shown to decrease the size of its optic vesicles (Yamamoto et al., 2004) and probably the eyefield size, which could account for the final smaller eye size. Moreover, albeit smaller, CF optic vesicles are “correctly” patterned in their future naso-temporal axis, as shown by Hernández-Bejarano and colleagues at 10/12 somite stage (~13.5hpf) using FoxG1 and FoxD1 markers (Fig. 6A) (Hernandez-Bejarano et al., 2015).

(2) Second, after the initial evagination and patterning of a small optic vesicle, morphogenesis proceeds with the “extended evagination”, whereby cells from the neural tube continue entering the optic vesicle to contribute exclusively to the ventro-nasal part of the eye (Kwan...
et al., 2012). If this step proceeds normally in CF, which seems to be the case from live imaging experiments, it could partially compensate the originally small size of the eyefield/optic vesicle, but only in the nasal part, while the temporal part would remain fully affected (Fig. 6B). This would well explain the increased angles of nasal FoxG1 and Vax2 expression in comparison to the reduced temporal FoxD1 and Vax2 territories we observed. It is also worth noting that early Hedgehog signalling can increase Vax2 expression (before early optic vesicle stage) in Xenopus (Wang et al., 2015), which could further explain Vax2 nasal expansion.

(3) Third, the lens forms correctly at the centre of the optic vesicle in both morphs, in a proper place with regard to the initial invagination of the optic cup; it is only at later stages that the lens appears more anterior (i.e., facing the presumptive ventral retina after final eye rotation) in cavefish. This apparent displacement of the lens relative to the retina is not due to a movement of the lens - which remains fixed throughout eye morphogenesis, attached to the overlying ectoderm from which it delaminates around 22hpf in Astyanax (Hinaux et al., 2017) - but rather to a persistent elongation of the optic vesicle. This suggests normal interactions between the optic vesicle and the lens to adjust their relative position and initiate optic cup invagination. Indeed, in chick, the pre-lens ectoderm is required for normal optic cup invagination while the lens placode itself is dispensable (Hyer et al., 2003). In cavefish such mechanisms could exist and correctly initiate optic cup folding. Finally, the anterior-shifted position of the lens, due to elongation of the cavefish optic vesicle without invagination, explains why the lens is ventrally displaced in the mature eye: the final anterior rotation movement brings it in a ventral position (Fig. 6C).

(4) Fourth, although the invagination of the CF optic vesicle seems to start normally between 15.5 and 19.5hpf, it only progresses very little afterwards so that the optic cup remains shallow and elongated. This timing is very reminiscent of the 2 described steps of optic cup invagination: basal constriction initiating the primary folding between 18 and 20hpf in zebrafish (18ss to 22ss), followed by the rim movement which brings the presumptive retina from the inner optic vesicle leaflet into the lens-facing epithelium by an active migration around the rims of the optic recess between 20 and 24hpf (Heermann et al., 2015; Nicolas-Perez et al., 2016; Sidhaye and Norden, 2017). In Astyanax, 18 ss corresponds to ~16.5hpf (Hinaux et al., 2011), suggesting that the initial basal constriction leading to the onset of optic cup invagination is well conserved in cavefish – this will have to be confirmed. In contrast, the
prolonged extension of the optic vesicle and the fact that the curvature of the retina remains shallow with the lens bulging out of the eye suggests that the rim movement must be impaired in cavefish. We propose that the continuous flow of cells entering the retina leads to its elongation, in the absence of an efficient rim movement. This movement does seem weaker but not absent in CF, as the posterior part of its optic cup still manages to contact the lens, but at later stages. This difference of rim movement might be due to various causes such as defects in the basal membrane or failure to establish proper focal adhesion as seen in the ojoplano medaka mutant (Martinez-Morales et al., 2009; Nicolas-Perez et al., 2016; Sidhaye and Norden, 2017). Alternatively, the active migration described by Sidhaye and Norden could be altered by extrinsic signals, as in BMP overexpression experiments where the cell flow toward the lens-facing epithelium is reduced (Heermann et al., 2015). The various morphogen modifications known in cavefish, and the fact that the ventral eye can be restored by delaying the onset of the Fgf signalling in CF to match the SF scenario (Pottin et al., 2011), support this possibility.

The “continuous extension of the optic cup with only little rim movement” hypothesis explains cavefish phenotypes. It clarifies the ventral and “floating” position of the lens in the final eye, without much contact with the edges of the retina. It also explains how grooves/folds in the retina (typically between the dorsal and temporal quadrants) are sometimes formed (Fig. S6).

(5) Fifth, Bhlhe40 expression reveals that RPE identity is maintained in the CF eye. Yet, the expansion and engulfment movement that this tissue is supposed to achieve to reach the rim of the retina and cover the whole retina is delayed compared to SF. In fact, this spreading and migration of RPE cells is concomitant with the rim movement and may contribute to it as a driving force (Cechmanek and McFarlane, 2017; Moreno-Marmol et al., 2018). This observation reinforces the idea that the rim movement is impaired in cavefish. Potentially, the RPE movement may also be involved in optic fissure closure, as suggested by the presence of a coloboma upon impairment of the rim movement by BMP4 overexpression in the optic vesicle (Heermann et al., 2015). Deficiency in this morphogenetic movement might participate in the cavefish coloboma phenotype. Of note, the transplantation of a healthy SF lens into a CF optic cup rescues the eye as a structure, i.e., prevents lens-induced degeneration, but does not rescue the coloboma phenotype (Yamamoto and Jeffery, 2000). This is in agreement with
our findings showing that the improper closure of the optic fissure is autonomous to the CF retinal tissues and results from defective morphogenetic movements.

(6) Sixth, in addition and concomitantly to impaired morphogenetic processes, morphogen signalling modifications previously reported in cavefish might contribute to the axial patterning variations observed. On the naso-temporal axis, there seems to be a slight increase in FoxG1 domain in the early optic vesicle (Hernandez-Bejarano et al., 2015), which may be explained by the earlier onset of Fgf8 expression in the cavefish anterior neural ridge (Pottin et al., 2011), but also by the increase in size of the cavefish olfactory placodes which also secrete Fgf24 (Hernandez-Bejarano et al., 2015; Hinaux et al., 2016). These Fgfs, along with Fgf3 promote retinal nasal fate (Hernandez-Bejarano et al., 2015; Picker and Brand, 2005; Picker et al., 2009), and may be responsible for the slight expansion of FoxG1 expression in cavefish, itself limited or counteracted by increased Shh signalling (Hernandez-Bejarano et al., 2015). This might contribute to the importance taken by nasal fates relative to temporal fates in cavefish. It may also account for the Tbx5a angle enlargement towards the temporal quadrant. In fact, Fgf signalling has been shown to promote dorsal identity temporally, as demonstrated by the temporal reduction of Tbx5a span and its nasal shift in Fgf8 mutant zebrafish (Ace) (Picker and Brand, 2005). Finally, besides Shh, Fgf and Bmp signalling, it is to note that the mouse Lrp6 insertional mutant displays microphthalmia and coloboma (Zhou et al., 2008), and that zebrafish treated with the Wnt-βcat pathway activator LiCl have small eyes (Shinya et al., 2000), therefore the modulations of canonical Wnt signalling observed in cavefish (Torres-Paz et al., 2018) probably also impact the development of their eyes.

(7) Pax2a expression domain is increased at the expense of the Pax6-expressing part of the eyefield at neural plate stage in CF (Yamamoto et al., 2004). Here we expand these results at later stage and show that Pax2a is no longer confined to the optic stalk and optic fissure margins but invades the ventral retina and even often, the dorsal retina. Contrarily to Vax2, Pax2a/optic stalk identity is expanded in both temporal and ventral directions. This phenotype is probably caused by the increase in Shh signalling in cavefish, as Shh injections in zebrafish embryos or Ptch1 (a negative regulator of Hedgehog pathway) loss of function in blowout mutants cause very similar phenotypes with a larger optic stalk and an invasion of the ventral retina by Pax2a expression (Lee et al., 2008). Nevertheless, the observation of cells with an optic stalk identity in the cavefish dorsal retina was surprising, as in zebrafish, Shh signalling
maintains the optic stalk-retina interface through the regulation of Vax genes (Takeuchi et al., 2003), and here we found that Vax2 was mis-expressed in a much milder manner than Pax2a. A possibility would be that these Pax2a “ectopic cells” stem from abnormal morphogenetic movements that could not be traced in our study.

**Evolutionary considerations**

In all eyeless or eye-reduced cave vertebrates examined so far, initial eye development occurs, including the evagination of optic vesicles and further morphogenetic steps (e.g., Durand, 1976; Stemmer et al., 2015; Wilkens, 2001). This represents an energetically-costly process for embryos, raising the question of why would all these different species first develop eyes which are after all fated to degeneration, and suggesting that initial eye development cannot be circumvented (Rétaux and Casane, 2013). Our results help further refine the step(s) in eye morphogenesis that are subjected to developmental constraint. Indeed, in cavefish, the (small) eyefield is correctly specified and the evagination/elongation steps, which correspond to cell movements leading to the sorting of retinal versus adjacent telencephalic, preoptic and hypothalamic cells of the neural tube, occurs properly. It is only after the segregation between these differently-fated cell populations that the cavefish eye morphogenesis starts going awry, with a defective invagination process, that will soon be followed by lens apoptosis and the subsequent progressive degeneration of the entire eye. Therefore, our data support the idea that the first steps of eye development constitute an absolute developmental constraint to morphological evolution. To the best of our knowledge, the closest to a counter-example is the medaka mutant eyeless, a temperature-sensitive rx3 mutant line in which optic vesicles evagination does not occur. However, the homozygous eyeless fish either die after hatching (Winkler et al., 2000) or, for the 1% which can be raised to adulthood in laboratory, are sterile probably due to anatomical hypothalamic or hypophysis defects (Ishikawa et al., 2001), still reinforcing the hypothesis of a strong developmental constraint.

**Conclusion**

This work proposes conclusions and hypotheses regarding defective cavefish eye morphogenesis, and interprets cavefish eye development in the frame of the current
knowledge on the topic. Further analyses including the tracking, characterization and comparison of cell movements occurring in both morphs will refine the current scenario. For example, quantifying the extent of the rim movement, the curvature of the optic cup and the amount of cells entering the optic vesicle through time would be informative. These data also pave the way for experiments aiming at understanding the defective molecular or signalling mechanisms in cavefish eye morphogenesis, using the Zic1:hsp70:GFP knock-in lines.
Methods

Animals

Laboratory stocks of *A. mexicanus* surface fish and cavefish were obtained in 2004 from the Jeffery laboratory at the University of Maryland. The surface fish were originally collected from San Solomon Spring, Texas and the cavefish are from the Pachón cave in Mexico. Surface fish are kept at 26°C and cavefish at 22°C. Natural spawns are induced after a cold shock (22°C over weekend) and a return to normal temperature for surface fish; cavefish spawns are induced by raising the temperature to 26°C. Embryos destined for *in situ* hybridization were collected after natural spawning, grown at 24°C and staged according to the developmental staging table (Hinaux et al. 2011) and fixed in 4% paraformaldehyde. After progressive dehydration in methanol, they were stored at -20°C. Embryos destined to transgenesis or live imaging were obtained by *in vitro* fertilization. Embryos were raised in an incubator until 1 month post fertilization for the surface fishes and two month post fertilization for the cavefish. They were kept at low density (15/20 per litre maximum) in embryo medium, in 1 litre plastic tanks with a soft bubbling behind the strainer. Larvae were fed from day 5 with paramecium and transitioned to artemia nauplii from day 10-15. Artemia were given twice a day except for the weekends (once a day) and carefully removed afterward to avoid polluting the medium. At least two thirds of the medium were changed every day and dead larvae removed. After one month for the surface fish and two months for the cavefish, juveniles were taken to the fish facility where they were fed dry pellets (Skretting Gemma wean 0.3) and quickly moved to bigger tanks in order to allow their fast growth.

Animals were treated according to French and European regulations of animals in research. SR’ authorization for use of animals in research is 91-116, and Paris Centre-Sud Ethic committee authorization numbers are 2012-52 and 2012-56.

*In situ* hybridization

Some cDNAs were available from our cDNA library: *Zic1* (FO290256), *Zic2a* (FO320762) and *Rx3* (FO289986), *FoxD1* (FO380710); others were already cloned in the lab: *Lhx2* (EF175737)
and Lhx9 (EF175738) (Alunni et al. 2007); obtained from other labs (Vax1: Jeffery lab, University of Maryland, (Yamamoto et al. 2004)); or cloned for the purpose of this work in pGEMT-Easy (Promega):

- FoxG1: forward primer CTGACGTTCATGGACCGAGC; reverse primer CAGTCTGCTTCTGTTGGATGT.
- Tbx5a: forward primer GCCTTCATTGCGGTCACTTC; reverse primer CCCTCGTTCCATTCAGGCAT.
- Vax2: forward primer GGGCAAAACATGCGCGTTA; reverse primer CAGTAATCCGGGTCCACTCC.
- Pax2a: forward primer AGCTGCATAACCGAGGCGA; reverse primer CTCCATTAGAGCGAGGGATTCCGA
- Bhlhe40: forward primer : GCACTTTCCCTGCGGATTTC; reverse primer : TGGAGTC TCGTTTGTCCAGC

cDNAs were amplified by PCR, and digoxigenin-labelled riboprobes were synthesized from PCR templates. Embryos were rehydrated by graded series of EtOH/PBS, then for embryos older than 24hpf, proteinase-K permeabilization at 37°C was performed for 36hpf embryos only (10 µg/ml, 15 min) followed by a post-fixation step. Riboprobes were hybridized for 16 hr at 65°C and embryos were incubated with anti-DIG-AP (Roche, dilution 1/4000) overnight at 4°C. Colorimetric detection with BCIP/NBT (Roche) was used. Mounted embryos were imaged on a Nikon Eclipse E800 microscope equipped with a Nikon DXM 1200 camera running under Nikon ACT-1 software. Brightness and contrast were adjusted using FIJI, some of the images used for illustration purpose were created from an image stack, using the extended depth of field function of Photoshop CS5. Area, distance and angle measurements were performed using FIJI (Schindelin et al., 2012; Schneider, Rasband and Eliceiri, 2012).

**In vitro fertilization (IVF) and injections**

Surface and cavefish were maintained in a room with shifted photoperiod (light: 4pm – 7am, L:D 15:11) in order to obtain spawns during the working day (Astyanax spawn at night (Simon et al., 2019)). Fish activity was monitored after induction and upon visible excitation or when first eggs were found at the bottom of the tank, fish were fished. Females were processed first.
to obtain eggs: they were quickly blotted on a moist paper towel and laid on their side in a petri dish. They were gently but firmly maintained there while their flank was gently stroked. If eggs were not released immediately, the female was put back in the tank. Once eggs were collected, a male was quickly processed similarly to females, on the lid of the petri dish to collect sperm. The sperm was then washed on the eggs with 10-20mL of tank water (conductivity ~500µS) and left for a few moments (30s to 2 min approximately), after which embryo medium was added in the petri dish. Fertilised eggs were quickly laid on a zebrafish injection dish containing agarose grooves. They were injected with a Picospritzer III (Parker Hannifin) pressure injector.

**CRISPR injections and Knock-in lines**

sgRNA were designed to target the low-conservation regions between elements 1 and 2 and between elements 3 and 4. Two sgRNA were initially designed per region and sgRNA2 was found to efficiently cut the targeted region (Fig. S7). The mix contained Cas9 protein generously provided by TACGENE and sgRNA2 with the following targeting sequence: CCCAATTCACCAGTATACGT (synthesized with AMBION T7 MEGASHORTSCRIPT™ T7 transcription kit). Concentrations were kept with a 1:1.5 Cas9 to sgRNA molar ratio and varied between 0.71µM (25ng/µL) and 5.67µM (200ng/µL) of sgRNA 2, mostly 2.84 and 1.42µM were used. The donor construct contained a HSP70 promoter used as a minimal promoter, a GFP cDNA and SV40 poly-adenylation signal, flanked by I-SceI meganuclease cutting sites. I-SceI was used to generate sticky ends and was either detached by 7 min at 96°C or injected with the construct. Concentrations of the repair construct varied between 3.33 and 10.92nM but were mostly used at 10.71nM.

**mRNA injection**

Transgenic embryos used for live imaging were injected in the cell or yolk at 1 cell stage with a H2B-mCherry fusion mRNA at a concentration of 50ng/µL.

**Imaging**
Transgenic embryos were obtained by IVF with wild-type eggs and transgenic sperm and were immediately injected with H2B-mCherry mRNA for nuclear labelling. Injected embryos were screened for GFP and mCherry fluorescence under a Leica M165C stereomicroscope around 10-11hpf, when GFP reporter fluorescence first becomes detectable.

Selected embryos were immediately mounted in a phytagel tube (SIGMA, CAS Number: 71010-52-1) molded with Phaseview Teflon mold (1.5mm of diameter) and maintained in position with 0.4% low melting point agarose (Invitrogen UltraPure™ Low Melting Point Agarose). The tube containing the embryo was placed horizontally into the chamber containing 0.04% Tricaine in embryo medium (Sigma, CAS Number: 886-86-2). The tube was rotated under the microscope so that the embryo would face the objective.

Live imaging was performed approximately from 10.5-11hpf to 24hpf every 2.5min-3min, using a Phaseview Alpha³ light sheet apparatus, coupled with an Olympus BX43 microscope and using either a 20X/NA 0.5 Leica HCX APO objective or a 20X/NA 0.5 Olympus objective. Images were acquired using QtSPIM software (Phaseview), which controlled a Hamamatsu ORCA-Flash4.0 Digital sCMOS camera.

Room temperature was maintained at 24°C by air conditioning and the chamber temperature was further controlled by a BIOEMERGENCES-made thermostat. Medium level was maintained by a home-made perfusion system and an overflow to renew the medium.

**Imaging analysis**

Images obtained were visualized with Arivis Vision4D software and re-oriented to adopt a similar optical section plane, cutting through the middle of the lens and the optic stalk at all time-steps. On one time-step per hour, measurements were performed on the re-oriented image: optic vesicle/optic cup length (at the widest), optic vesicle size increase (calculated by subtracting the length at the onset of furrow formation to the length at time t), optic stalk width, distance between the anterior optic cup and the lens, distance between the posterior optic cup and the lens, distance between the optic cup edges, position of the lens relative to anterior optic vesicle (=distance between center of the lens and anterior OV / (distance between center of the lens and anterior OV + distance between center of the lens and posterior OV)) (see schemes on Figures).
Acknowledgements

Work supported by an Equipe FRM grant (DEQ20150331745) and an UNADEV/AVIESAN grant to SR. We thank Jean-Paul Concordet and Anne De Cian (Tacgene, Sorbonne Universités, Paris) for sharing Cas9 protein; Diane Denis, Krystel Saroul, Jocelyne Gaget, the Amagen personnel and the people from the fish facility for advices and care of our Astyanax colony; Patrick Para for making custom tools for live microscopy; Adeline Boyreau, Adeline Rausch, Fanny Husson, Elena Kardash and Nadine Peyrieras (BioEmergence platform, Gif sur Yvette, France) for reagents, discussions and advices on live imaging, and for the use of the SPIM and image analysis tools; Cyprian Wozniak, Arthur Le Bris and Gaël Launay from PhaseView (Verrière-le-Buisson, France) for technical support and development of tools on the light-sheet microscope; and François Agnès for scientific discussions.
Affaticati, P., Yamamoto, K., Rizzi, B., Bureau, C., Peyrrieras, N., Pasqualini, C., Demarque, M. and Vernier, P. (2015). Identification of the optic recess region as a morphogenetic entity in the zebrafish forebrain. Sci Rep 5, 8738.

Albadri, S., Del Bene, F. and Revenu, C. (2017). Genome editing using CRISPR/Cas9-based knock-in approaches in zebrafish. Methods 121-122, 77-85.

Alunni, A., Menuet, A., Candal, E., Penigault, J. B., Jeffery, W. R. and Rétaux, S. (2007). Developmental mechanisms for retinal degeneration in the blind cavefish Astyanax mexicanus. J Comp Neurol 505, 221-33.

Asai-Coakwell, M., French, C. R., Berry, K. M., Ye, M., Koss, R., Somerville, M., Mueller, R., van Heyningen, V., Waskiewicz, A. J. and Lehmann, O. J. (2007). GDF6, a novel locus for a spectrum of ocular developmental anomalies. Am J Hum Genet 80, 306-15.

Cavodeassi, F. (2018). Dynamic Tissue Rearrangements during Vertebrate Eye Morphogenesis: Insights from Fish Models. J Dev Biol 6.

Cechmanek, P. B. and McFarlane, S. (2017). Retinal pigment epithelium expansion around the neural retina occurs in two separate phases with distinct mechanisms. Dev Dyn 246, 598-609.

Deschet, K., Bourrat, F., Ristoratore, F., Chourrout, D. and Joly, J. S. (1999). Expression of the medaka (Oryzias latipes) Ol-Rx3 paired-like gene in two diencephalic derivatives, the eye and the hypothalamus. Mech Dev 83, 179-82.

Durand, J. P. (1976). Ocular development and involution in the European cave salamander, Proteus anguinus laurenti. Biol Bull 151, 450-66.

Elipot, Y., Legendre, L., Père, S., Sohm, F. and Rétaux, S. (2014). Astyanax transgenesis and husbandry: how cavefish enters the lab. Zebrafish 11 (4), 291-299.

England, S. J., Blanchard, G. B., Mahadevan, L. and Adams, R. J. (2006). A dynamic fate map of the forebrain shows how vertebrate eyes form and explains two causes of cyclopia. Development 133, 4613-7.

French, C. R., Erickson, T., French, D. V., Pilgrim, D. B. and Waskiewicz, A. J. (2009). Gdf6a is required for the initiation of dorsal-ventral retinal patterning and lens development. Dev Biol 333, 37-47.

Gestri, G., Bazin-Lopez, N., Scholes, C. and Wilson, S. W. (2018). Cell Behaviors during Closure of the Choroid Fissure in the Developing Eye. Front Cell Neurosci 12, 42.

Gosse, N. J. and Baier, H. (2009). An essential role for Radar (Gdf6a) in inducing dorsal fate in the zebrafish retina. Proc Natl Acad Sci U S A 106, 2236-41.

Hagmann, M., Bruggmann, R., Xue, L., Georgiev, O., Schaffner, W., Rungger, D., Spaniol, P. and Gerster, T. (1998). Homologous recombination and DNA-end joining reactions in zygotes and early embryos of zebrafish (Danio rerio) and Drosophila melanogaster. Biol Chem 379, 673-81.

Heermann, S., Schutz, L., Lemke, S., Kriegstein, K. and Wittbrodt, J. (2015). Eye morphogenesis driven by epithelial flow into the optic cup facilitated by modulation of bone morphogenetic protein. Elife 4.

Hernandez-Bejarano, M., Gestri, G., Spawls, L., Nieto-Lopez, F., Picker, A., Tada, M., Brand, M., Bovolenta, P., Wilson, S. W. and Cavodeassi, F. (2015). Opposing Shh and Fgf signals initiate nasotemporal patterning of the zebrafish retina. Development 142, 3933-42.

Hinaux, H., Blin, M., Fumey, J., Legendre, L., Heuze, A., Casane, D. and Rétaux, S. (2015). Lens defects in Astyanax mexicanus Cavefish: Evolution of crystallins and a role for alphaA-crystallin. Dev Neurobiol, 505-21.
Hinaux, H., Devos, L., Bibliowicz, J., Elipot, Y., Alié, A., Blin, M. and Rétaux, S. (2016). Sensory evolution in blind cavefish is driven by early events during gastrulation and neurulation. *Development* **143**, 4521-4532.

Hinaux, H., Pottin, K., Chalhoub, H., Pere, S., Elipot, Y., Legendre, L. and Rétaux, S. (2011). A developmental staging table for Astyanax mexicanus surface fish and Pachon cavefish. *Zebrafish* **8**, 155-65.

Hinaux, H., Recher, G., Alie, A., Legendre, L., Blin, M. and Retaux, S. (2017). Lens apoptosis in the Astyanax blind cavefish is not triggered by its small size or defects in morphogenesis. *PLoS One* **12**, e0172302.

Hyer, J., Kuhlman, J., Afif, E. and Mikawa, T. (2003). Optic cup morphogenesis requires pre-lens ectoderm but not lens differentiation. *Dev Biol* **259**, 351-63.

Ishikawa, Y., Yoshimoto, M., Yamamoto, N., Ito, H., Yasuda, T., Tokunaga, F., Iigo, M., Wakamatsu, Y. and Ozato, K. (2001). Brain structures of a medaka mutant, el (eyeless), in which eye vesicles do not evaginate. *Brain Behav Evol* **58**, 173-84.

Ivanovitch, K., Cavodeassi, F. and Wilson, S. W. (2013). Precocious acquisition of neuroepithelial character in the eye field underlies the onset of eye morphogenesis. *Dev Cell* **27**, 293-305.

Kawakami, K. (2005). Transposon tools and methods in zebrafish. *Dev Dyn* **234**, 244-54.

Kimura, Y., Hisano, Y., Kawahara, A. and Higashijima, S. (2014). Efficient generation of knock-in transgenic zebrafish carrying reporter/driver genes by CRISPR/Cas9-mediated genome engineering. *Sci Rep* **4**, 6545.

Klaassen, H., Wang, Y., Adamski, K., Rohner, N. and Kowalko, J. E. (2018). CRISPR mutagenesis confirms the role of oca2 in melanin pigmentation in Astyanax mexicanus. *Dev Biol* **411**, 313-318.

Kruse-Bend, R., Rosenthal, J., Quist, T. S., Veien, E. S., Fuhrmann, S., Dorsky, R. I. and Chien, C. B. (2012). Extraocular ectoderm triggers dorsal retinal fate during optic vesicle evagination in zebrafish. *Dev Biol* **371**, 57-65.

Kwan, K. M., Otsuna, H., Kidokoro, H., Carney, K. R., Saijoh, Y. and Chien, C. B. (2012). A complex choreography of cell movements shapes the vertebrate eye. *Development* **139**, 359-72.

Lee, J., Willer, J. R., Willer, G. B., Smith, K., Gregg, R. G. and Gross, J. M. (2008). Zebrafish blowout provides genetic evidence for Patched1-mediated negative regulation of Hedgehog signaling within the proximal optic vesicle of the vertebrate eye. *Dev Biol* **319**, 10-22.

Macdonald, R., Scholes, J., Strahle, U., Brennan, C., Holder, N., Brand, M. and Wilson, S. W. (1997). The Pax protein Noi is required for commissural axon pathway formation in the rostral forebrain. *Development* **124**, 2397-408.

Martinez-Morales, J. R., Rembold, M., Greger, K., Simpson, J. C., Brown, K. E., Quiring, R., Pepperkok, R., Martin-Bermudo, M. D., Himmelbauer, H. and Wittbrodt, J. (2009). Ojoplano-mediated basal constriction is essential for optic cup morphogenesis. *Development* **136**, 2165-75.

Maurus, D. and Harris, W. A. (2009). Zic-associated holoprosencephaly: zebrafish Zic1 controls midline formation and forebrain patterning by regulating Nodal, Hedgehog, and retinoic acid signaling. *Genes Dev* **23**, 1461-73.

McGaugh, S. E., Gross, J. B., Aken, B., Blin, M., Borowsky, R., Chalopin, D., Hinaux, H., Jeffery, W. R., Keene, A., Ma, L. et al. (2014). The cavefish genome reveals candidate genes for eye loss. *Nat Commun* **5**, 5307.
Moreno-Marmol, T., Cavodeassi, F. and Bovolenta, P. (2018). Setting Eyes on the Retinal Pigment Epithelium. Front Cell Dev Biol 6, 145.

Morita, H., Taimatsu, K., Yanagi, K. and Kawahara, A. (2017). Exogenous gene integration mediated by genome editing technologies in zebrafish. Bioengineered 8, 287-295.

Nicolas-Perez, M., Kuchling, F., Letelier, J., Polvillo, R., Wittbrodt, J. and Martinez-Morales, J. R. (2016). Analysis of cellular behavior and cytoskeletal dynamics reveal a constriction mechanism driving optic cup morphogenesis. Elife 5.

Picker, A. and Brand, M. (2005). Fgf signals from a novel signaling center determine axial patterning of the prospective neural retina. Development 132, 4951-62.

Picker, A., Cavodeassi, F., Machate, A., Bernauer, S., Hans, S., Abe, G., Kawakami, K., Wilson, S. W. and Brand, M. (2009). Dynamic coupling of pattern formation and morphogenesis in the developing vertebrate retina. PLoS Biol 7, e1000214.

Pottin, K., Hinaux, H. and Rétaux, S. (2011). Restoring eye size in Astyanax mexicanus blind cavefish embryos through modulation of the Shh and Fgf8 forebrain organising centres. Development 138, 2467-76.

Rembold, M., Loosli, F., Adams, R. J. and Wittbrodt, J. (2006). Individual cell migration serves as the driving force for optic vesicle evagination. Science 313, 1130-4.

Rétaux, S. and Casane, D. (2013). Evolution of eye development in the darkness of caves: adaptation, drift, or both? Evodevo 4, 26.

Rétaux, S. and Harris, W. A. (1996). Engrailed and retinotectal topography. Trends Neurosci 19, 542-6.

Rohr, K. B., Schulte-Merker, S. and Tautz, D. (1999). Zebrafish zic1 expression in brain and somites is affected by BMP and hedgehog signalling. Mech Dev 85, 147-59.

Sakuta, H., Takahashi, H., Shintani, T., Etani, K., Aoshima, A. and Noda, M. (2006). Role of bone morphogenic protein 2 in retinal patterning and retinotectal projection. J Neurosci 26, 10868-78.

Sanek, N. A., Taylor, A. A., Nyholm, M. K. and Grinblat, Y. (2009). Zebrafish zic2a patterns the forebrain through modulation of Hedgehog-activated gene expression. Development 136, 3791-800.

Schulte, D., Furukawa, T., Peters, M. A., Kozak, C. A. and Cepko, C. L. (1999). Misexpression of the Emx-related homeobox genes cVax and mVax2 ventralizes the retina and perturbs the retinotectal map. Neuron 24, 541-53.

Shinya, M., Eschbach, C., Clark, M., Lehrach, H. and Furutani-Seiki, M. (2000). Zebrafish Dkk1, induced by the pre-MBT Wnt signaling, is secreted from the prechordal plate and patterns the anterior neural plate. Mech Dev 98, 3-17.

Sidhaye, J. and Norden, C. (2017). Concerted action of neuroepithelial basal shrinkage and active epithelial migration ensures efficient optic cup morphogenesis. Elife 6.

Simon, V., Hyacintie, C. and Retaux, S. (2019). Breeding behavior in the blind Mexican cavefish and its river-dwelling conspecific. PLoS One 14, e0212591.

Sperry, R. W. (1963). Chemoaffinity in the Orderly Growth of Nerve Fiber Patterns and Connections. Proc Natl Acad Sci U S A 50, 703-10.

Stahl, B. A., Peuss, R., McDole, B., Kenzior, A., Jaggard, J. B., Gaudenz, K., Krishnan, J., McGaugh, S. E., Duboue, E. R., Keene, A. C. et al. (2019). Stable transgenesis in Astyanax mexicanus using the Tol2 transposase system. Dev Dyn.

Stemmer, M., Schuhmacher, L. N., Foulkes, N. S., Bertolucci, C. and Wittbrodt, J. (2015). Cavefish eye loss in response to an early block in retinal differentiation progression. Development 142, 743-52.
Stigloher, C., Ninkovic, J., Laplante, M., Gelng, A., Tannhauser, B., Topp, S., Kikuta, H.,
Becker, T. S., Houart, C. and Bally-Cuif, L. (2006). Segregation of telencephalic and eye-field
identities inside the zebrafish forebrain territory is controlled by Rx3. Development 133, 2925-
35.

Strauss, O. (2018). The retinal pigmented epithelium. In "The organization of the retina and
visual system". Edited by H. Kolb, E Fernandez and R. Nelson. Webvision. University of Utah
Health Science Center, Salt Lake City.

Strickler, A. G., Yamamoto, Y. and Jeffery, W. R. (2001). Early and late changes in Pax6
expression accompany eye degeneration during cavefish development. Dev Genes Evol 211,
138-44.

Takeuchi, M., Clarke, J. D. and Wilson, S. W. (2003). Hedgehog signalling maintains the optic
stalk-retinal interface through the regulation of Vax gene activity. Development 130, 955-68.

Thermes, V., Grabher, C., Ristoratore, F., Bourrat, F., Choulika, A., Wittbrodt, J. and Joly, J.
S. (2002). I-SceI meganuclease mediates highly efficient transgenesis in fish. Mech Dev 118,
91-8.

Torres-Paz, J., Leclercq, J. and Rétaux, S. (2018). Evolution of gastrulation in Mexican cavefish:
heterochronic cell movements and maternal factors. bioRxiv Sept 2018.

Tropepe, V., Li, S., Dickinson, A., Gamse, J. T. and Sive, H. L. (2006). Identification of a BMP
inhibitor-responsive promoter module required for expression of the early neural gene zic1.
Dev Biol 289, 517-29.

Varga, Z. M., Wegner, J. and Westerfield, M. (1999). Anterior movement of ventral
diencephalic precursors separates the primordial eye field in the neural plate and requires
cyclops. Development 126, 5533-46.

Wang, X., Lupo, G., He, R., Barsacchi, G., Harris, W. A. and Liu, Y. (2015). Dorsoventral
patterning of the Xenopus eye involves differential temporal changes in the response of optic
stalk and retinal progenitors to Hh signalling. Neural Dev 10, 7.

Wilkens, H. (2001). Convergent adaptations to cave life in the Rhamdia laticauda catfish group
(Pimelodidae, Teleostei). Environmental Biology of Fishes 62, 251-261.

Winkler, S., Loosli, F., Henrich, T., Wakamatsu, Y. and Wittbrodt, J. (2000). The conditional
medaka mutation eyeless uncouples patterning and morphogenesis of the eye. Development
127, 1911-9.

Woo, K. and Fraser, S. E. (1995). Order and coherence in the fate map of the zebrafish nervous
system. Development 121, 2595-609.

Woo, K., Shih, J. and Fraser, S. E. (1995). Fate maps of the zebrafish embryo. Curr Opin Genet
Dev 5, 439-43.

Yamamoto, Y. and Jeffery, W. R. (2000). Central role for the lens in cave fish eye degeneration.
Science 289, 631-3.

Yamamoto, Y., Stock, D. W. and Jeffery, W. R. (2004). Hedgehog signalling controls eye
degeneration in blind cavefish. Nature 431, 844-7.

Zhou, C. J., Wang, Y. Z., Yamagami, T., Zhao, T., Song, L. and Wang, K. (2008). Generation of
Lrp6 conditional gene-targeting mouse line for modeling and dissecting multiple birth
defects/congenital anomalies. Dev Dyn 239, 318-26.
Figures and Legends
Figure 1: establishment of surface fish and cavefish Zic1:hsp70:GFP lines.
(A) Zic1 expression at indicated stages in SF and CF. Anterior is to the left. Dorsal views at 10, 12 and 14 hpf, lateral views at 36hpf. Asterisk: larger indentation in the CF eyefield.

(B) Zebrafish Zic1 genomic region in UCSC genome browser (2010 assembly). Green blue peaks as well as magenta and black elements correspond to high conservation, showing the complexity of the region.

(C) Close-up on Zic1. Red boxes highlight conserved elements; element 3 is not conserved in Astyanax (asterisk).

(C) sgRNA were designed to target the low-conservation regions between elements 1 and 2, and 3 and 4. SgRNA2 (pale blue) efficiently generated cuts. It was co-injected together with the Cas9 protein and the linear repair construct containing a minimal Hsp70 promoter and the GFP.

(D) Zic1-like GFP fluorescence in mosaic F0s and stable F1s, both for SF and CF.

(E) Double-fluorescent in situ hybridization at 16hpf for Zic1 (magenta) and GFP (yellow) showing that the transgene recapitulates the endogenous Zic1 pattern, both for SF and CF lines. The top panels show entire embryos and the bottom panels show close-ups on the head, including the Zic1-expressing telencephalon (t) and eye (e). Lateral views.

Scale bars=100µm.
Figure 2: Live imaging of surface fish and cavefish eye morphogenesis.

(A) Schematic drawings of the principal steps of eye morphogenesis in fish, in lateral (top) and dorsal views (bottom). Orange arrows indicate cell and tissue movements; green arrowheads show initiation of basal constriction. The grey line indicates the optical section plane used in the pictures in B, which follows an optic stalk to lens center axis and accompanies the anterior rotation illustrated by the arrows. All subsequent measures were realized on these planes.

(B) Still images of time-lapse acquisitions from 10.5hpf to 30.5hpf on SF and SF Zic1:hsp70:GFP lines (green, GFP; magenta, nuclear mCherry). Representative steps of eye morphogenesis illustrating CF/SF differences are shown. Dorsal views, anterior to the top.
Figure 3

Figure 3: Morphological quantification of eye morphogenesis.
Measures are illustrated on the diagrams on the right in each box.

(A) Optic vesicle length. The left graph shows the mean of n=4 eyes in each morph (blue, SF; red, CF); the right graph displays the trajectories of individual eyes to show the reproducibility of the results.

(B) Optic vesicle size increase.

(C) Distance between the two optic cup edges.

(D) Position of the lens relative to anterior optic vesicle, showing that the lens is progressively shifted anteriorly between 25hpf and 30hpf.
Figure 4: regionalization of the eye.

(A-D’) In situ hybridization for the indicated markers on 36hpf SF and CF embryos. Sample sizes are indicated.

(A’’-D’’) Quantification of angles of expression and position with regards to the choroid fissure. All the angles measured are represented in H. SF, blue; CF, red.

(E) Absolute area of expression, in µm².

(F) Area of expression normalized by eye size.

(G) Area of expression in CF, expressed as percentage of the expression area in SF. Horizontal line shows that the CF eye area is 64% of the SF eye.

Statistical test: Mann-Whitney; * p<0.05; ** p<0.01; *** p<0.001.
(H) Summary diagram of the expression patterns. The angles measured are represented. Expression patterns are represented in blue for SF and red for CF. The angles measured are between the middle of the choroid fissure and the borders of the expression domains.
Figure 5

Figure 5: tissue identity.

(A-B') In situ hybridization for the indicated markers on 36hpf SF and CF embryos. (C) Bhlhe40 expression in 48hpf CF. Sample sizes are indicated.

(A'') Angle between the optic fissure and the limit of Pax2a domain (see F for angles measured).

(B'') Angles of Bhlhe40 expression at 36 and 48hpf.

(D) Distance between Bhlhe40-expressing cells and the lens at 36hpf.

(E) Width of the Bhlhe40 expression gap at the back of the eye. SF, blue; CF, red.

Statistical test: Mann-Whitney; * p<0.05; ** p<0.01; *** p<0.001.

(F) Summary diagram of expression patterns. Angles measured are represented.
Combining data from live imaging and gene expression patterns, we propose that the small size of the optic vesicles, correctly patterned, explains the decrease in temporal quadrant size. Unaltered extended evagination movements bring new cells into the prospective nasal retina, partially rescuing its size (pale green). Finally, the expansion of the RPE to engulf the retina which is concomitant with the rim movement is delayed, leaving the optic stalk wide and optic fissure margins away from each other.

oe, olfactory epithelium; tel, telencephalon; hyp, hypothalamus.
Supplemental Figures and Legends
Supplemental Figure 1: eye morphogenesis in fish.

Schemes depicting the principal steps of eye morphogenesis in fish models, summarized from the available literature cited in Introduction.

- Stages and orientations are indicated.
- Orange arrows show general cell and tissue movements.
- Black arrows show the anterior-wise rotation of the eye and brain.
- Green arrows show the contribution of extended evagination.
- Pink arrowhead show cellular basal constriction.
- The blue color depicts the RPE cells, while the green color depicts retina neuroepithelium cells changing shape.
**Supplemental Figure 2: choosing a candidate gene for transgenesis.**

(A) Mini-screen of candidate genes by *in situ* hybridization at different stages of interest on surface fish embryos (anterior is to the left). Dorsal views at 10hpf; lateral views at 14hpf and
36hpf. The eyes were dissected out for Vax1 and Lhx9 (as no eye expression was detected for either of them) to allow better visibility of the inner tissue.

(B) Detailed analysis of Zic1 expression pattern at 5 different stages in surface (SF) and cavefish (CF). Anterior is to the left, at 10, 12 and 14hpf, bottom pictures are taken in dorsal view; at 24 and 36hpf, bottom picture are taken in ventral views. Arrowheads indicate an indentation in the eyefield.

**Description of expression patterns:**

Vax1 expression was detectable from 12hpf in the presumptive ORR (between the optic vesicles) and additionally in the dorsal hypothalamus (according to brain axis (Puelles & Rubenstein, 2015), closest to the ORR) and quite faintly in the ventral telencephalon.

Vax2 expression was very similar to Vax1 both in terms of onset of expression and pattern, with the addition of the ventral quadrant of the eye. Although Vax2 had a very interesting ventral pattern, we discarded it as a candidate for transgenesis for its expression onset was very late. Moreover, in Vax2 enhancer trap zebrafish line (Kawakami Laboratory), the GFP fluorescence is only visible at 18hpf (personal observation, data not shown).

Rx3 expression showed a typical eyefield expression pattern at 10hpf but progressively faded away during optic vesicle stages and was finally not expressed anymore at 24hpf. Conversely, an anterior and ventral expression in the presumptive hypothalamus was detectable from 12hpf and remained throughout the stages examined. At 36hpf, it was clear that only the dorsal half of the hypothalamus, closest to the ORR, was labelled. Due to the rapid fading of its optic vesicle expression, we did not consider Rx3 as a valid candidate.

Lhx2 and Lhx9 were both already known to be expressed in the eyefield at neural plate stages in Astyanax (Pottin et al., 2011). Lhx2 expression showed very dim expression, if any, in the optic vesicles at 12 and 14hpf but was expressed both in the prospective telencephalon and more faintly in the prospective hypothalamus. Later on at 36hpf, Lhx2 was expressed strongly in the telencephalon and the olfactory epithelia; lighter expression was also visible in the ORR, hypothalamus and sometimes eyes. Additional expression in the pineal, optic tectum and in the hindbrain was also present.

Lhx9 staining was strong in the optic vesicles at 12hpf (during evagination) and slightly lighter at 14hpf. Moreover dorsal and ventral lateral labelling at the border of the neural keel and the
optic vesicles appeared, possibly prefiguring respectively the strong telencephalic staining visible at 24 and 36 hpf and the hypothalamic cluster at the limit of the ORR already described in a previous publication (Alié et al., 2018). At these late stages, we could not detect Lhx9 expression in the eye anymore. Salt and pepper staining was visible in the olfactory epithelia; a band of expression outlining the optic tectum and lateral discrete marks in the hindbrain were present. We did not choose Lhx2 or Lhx9 because of the rapid decay of their eye expression.

At 10 hpf, Zic2a was expressed at the border of the neural plate and almost entirely surrounding the eyefield except for a medial posterior gap. Faint staining in the bilateral eyefield could also be seen on some embryos. At 12 and 14 hpf, there was a strong Zic2a expression in the telencephalon and a faint staining in the eye or distal part of the eye could often be seen. Strong staining was generally visible throughout the dorsal-most brain. At 24 hpf, Zic2a expression remained strong in the telencephalon and was also now strongly visible at the border of the eye, in the ORR or optic stalk but without reaching the midline. Faint staining in the eye remained. At 36 hpf, the expression pattern was similar, with the ORR/optic stalk staining reaching much closer to the midline. The eye expression was now more focused around the lens, probably in the CMZ. Roof plate staining persisted throughout development. Because Zic2a was never strongly expressed in the eye, we did not favour it as a candidate for transgenesis.

Zic1 was strongly expressed at 10 hpf in the neural plate border and in the anterior neural plate, at the level of the eyefield. At 12 and 14 hpf, Zic1 expression was consistently found in the optic vesicles and between them (prospective ORR and optic stalk). A strong staining was also present throughout the telencephalon. More posteriorly, the roof plate of the midbrain and hindbrain was stained. The somites were also labelled. The pattern was very similar at 24 hpf and 36 hpf with a strong telencephalic expression and a milder ORR expression (mainly laterally and posterior to the optic recess)/optic stalk and eye staining (widely around the lens). Roof plate and somites expression remained. Even though its pattern of expression was complex and encompassed a region wider than the optic region of interest, Zic1 was chosen for transgenesis due to its early and persistent expression throughout the eye and the ORR/optic stalk regions.
Supplemental Figure 3: genomic characterization of the Knock-in lines.

Knock-In insertions, based on partial sequencing. Dotted boxes indicate un-sequenced regions, leaving uncertainties. For example, in the surface fish line, there is at least a partial insertion of the repair construct, containing a truncated Hsp70 promoter and at least another insert in the same direction (but potentially several). Of note, the surrounding genomic region is very rich in T and A (GC content around 35%) with many repeats, making PCRs sometimes challenging.

The data show that for both lines the transgenes are inserted at the correct targeted site.
Supplemental Figure 4: optic stalk width.

The size of the optic stalk (in a wide meaning: the connection between the optic vesicle and the neural tube) is smaller in cavefish during early development due to the smaller size of the optic vesicles but rapidly becomes indistinguishable from the optic stalk of the surface fish.
Supplemental Figure 5: illustration of the gap of $Bhlhe40$ staining, which we interpret as the optic stalk width.

Different focuses of the same embryo, from lateral to medial on the felt eye. Arrowheads indicate the limits of the staining gap we measured. Dotted lines show the exit trajectory of the optic stalk.
Supplemental Figure 6: retinal folds in cavefish.

Illustration of the retina folds sometimes observed in cavefish: an “extreme case” on the left and a more moderate case on the right. 36hpf cavefish embryos, mounted laterally, anterior is to the left.
Supplemental Figure 7: cutting efficiency of sgRNA 2

(A) Assessment of sgRNA 2 cutting efficiency when injected with Cas9 mRNA by heteroduplex mobility assay (HMA, explained in (C)). Each column is an individual F0 embryo. Embryos with strong additional bands are labelled with a red asterisk; additional light bands can be seen in several individuals, indicating cuts and imprecise repairs. Note that the 2 heavy bands seen on many embryos are also present in some of the un-injected controls (the 6 columns on the right) indicating a polymorphism in this region in the wild-type fish (not on the sgRNA target sequence).

(B) Assessment of sgRNA 2 cutting efficiency when injected with Cas9 protein, note the strong presence of additional band compared to the 6 control embryos on the right. Embryos without any visible cuts are labelled with a blue asterisk. Additional bands are seen much more frequently and are much more important than with the Cas9 mRNA injection, probably indicating more frequent but also more precocious cut and repair events in the embryo, so that many cells share the same sequence.
(C) Principle of the heteroduplex mobility assay: in an electrophoresis, heteroduplexes are slowed down compared to homoduplexes so that they form additional bands that can be seen even if the polymorphism is only a single substitution. In short, the DNA fragments are denatured and renatured to form heteroduplexes. An electrophoresis is then performed (here with a LabChip, PerkinElmer) to detect the presence of polymorphism.

(D) Different cutting and repair events in a single injected embryo. A PCR was performed on one injected embryo (100ng/μL sgRNA2, Cas9mRNA) around the sgRNA2 target site and the product was cloned into pGEM-T Easy (Promega) and transformed into One shot TOP10 competent bacteria (Thermo Fischer). Plasmidic preparations from individual colonies were then sequenced. Various sequences were obtained, evidencing different cut and repair events in one single embryo. sgRNA2 target sequence is highlighted in yellow whenever intact. This F0 fish harbours both insertions and deletions around the cutting site of sgRNA2. A non-injected control fish sequence is included, outlined in black.