Can interaction specificity in the fungus-farming termite symbiosis be explained by nutritional requirements of the fungal crop?

da Costa, Rafael R.; Vreeburg, Sabine M. E.; Shik, Jonathan Z.; Aanen, Duur K.; Poulsen, Michael

Published in:
Fungal Ecology

DOI:
10.1016/j.funeco.2018.08.009

Publication date:
2019

Document version
Publisher's PDF, also known as Version of record

Document license:
CC BY

Citation for published version (APA):
da Costa, R. R., Vreeburg, S. M. E., Shik, J. Z., Aanen, D. K., & Poulsen, M. (2019). Can interaction specificity in the fungus-farming termite symbiosis be explained by nutritional requirements of the fungal crop? Fungal Ecology, 38, 54-61. https://doi.org/10.1016/j.funeco.2018.08.009
Can interaction specificity in the fungus-farming termite symbiosis be explained by nutritional requirements of the fungal crop?

Rafael R. da Costa a,a,⁎, Sabine M.E. Vreeburg b,1, Jonathan Z. Shik a,c, Duur K. Aanen b, Michael Poulsen e

a Section for Ecology and Evolution, Department of Biology, University of Copenhagen, Universitetsparken 15, 2100 Copenhagen East, Denmark
b Laboratory of Genetics, Wageningen University, Droevendaalsesteeg 1, 6708 PB, Wageningen, the Netherlands
c Section for Ecology and Evolution, Department of Biology, University of Copenhagen, Universitetsparken 15, Building 3, 2100 Copenhagen East, Denmark.
d Smithsonian Tropical Research Institute, Apartado 0843-03092, Balboa, Ancon, Republic of Panama

A R T I C L E  I N F O
Article history:
Received 27 November 2017
Received in revised form 13 July 2018
Accepted 21 August 2018
Available online 9 November 2018

Corresponding Editor: Henrik Hjarvard de Fine Licht

A B S T R A C T
Fungus-growing termites are associated with genus-specific fungal symbionts, which they acquire via horizontal transmission. Selection of specific symbionts may be explained by the provisioning of specific, optimal cultivar growth substrates by termite farmers. We tested whether differences in in vitro performance of Termitomyces cultivars from nests of three termite species on various substrates are correlated with the interaction specificity of their hosts. We performed single-factor growth assays (varying carbon sources), and a two-factor geometric framework experiment (simultaneously varying carbohydrate and protein availability). Although we did not find qualitative differences between Termitomyces strains in carbon-source use, there were quantitative differences, which we analysed using principal component analysis. This showed that growth of Termitomyces on different carbon sources was correlated with termite host genus, rather than host species, while growth on different ratios and concentrations of protein and carbohydrate was correlated with termite host species. Our findings corroborate the interaction specificity between fungus-growing termites and Termitomyces cultivars and indicate that specificity between termite hosts and fungi is reflected both nutritionally and physiologically. However, it remains to be demonstrated whether those differences contribute to selection of specific fungal cultivars by termites at the onset of colony foundation.

© 2018 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

1. Introduction

Mutualisms are widespread in nature, with cooperation between species often providing entry into ecological niches that could not support either species alone (Moya et al., 2008). Yet, the interactions between mutualists vary from short-term co-existence to irreversible obligate symbiosis. In addition, there are varying degrees of interaction specificities; i.e., possible combinations of hosts and symbionts (Aanen et al., 2007). Vertical transmission of symbionts generally leads to a high degree of interaction specificity and co-evolution, whereas horizontal transmission typically leads to less specialized associations between symbionts (Bright and Bulgheresi, 2010). Interaction specificity, however, does not only depend on transmission mode. It is often observed that (metabolic) traits of a symbiont are lost because their functions become redundant if the other partner reliably provides the resources (Visser et al., 2010; Ellers et al., 2012). Such reciprocal specialization can favour obligate symbiotic partnerships and foster co-cladogenesis of symbionts, even in the absence of vertical transmission (Aanen et al., 2007).

An intriguing obligate symbiosis is that between a monophyletic group of termites (family Termitidae, subfamily Macrotermiteinae) and basidiomycete fungi of the genus Termitomyces (Agaricomycetes, Lyophyllaceae), which originated ca. 30 million years ago in sub-Saharan Africa (Aanen et al., 2002; Aanen and Eggleton, 2005; Roberts et al., 2016). Fungus farming enables the termites to utilise food sources they cannot digest themselves, as the fungi convert recalcitrant plant substrates into edible fungal biomass and

⁎ Corresponding author. Section for Ecology and Evolution, Department of Biology, University of Copenhagen, Universitetsparken 15, 2100 Copenhagen East, Denmark.
E-mail address: Rafael.dacosta@bio.ku.dk (R.R. da Costa).

Contributed equally to this work.
accessible carbohydrates. In return, the termites shelter the fungus from unfavourable abiotic and biotic conditions (Wood and Sands, 1978; Rouland-Lefèvre and Bignell, 2002). There are about 330 described species of fungus-growing termites in 11 monophyletic genera (Aanen et al., 2002; Nobre et al., 2010), all farming Termitomyces fungal symbionts (Aanen et al., 2002; Roberts et al., 2016). Even though most termites acquire their symbionts horizontally (Fig. 1A) (de Fine Licht et al., 2006; Aanen et al., 2007; Nobre et al., 2010), there is a degree of co-cladogenesis between the termite and fungus phylogenies; groups of Termitomyces associate with specific termite clades (Fig. 1B) (Johnson et al., 1981; Sieber, 1983; Darlington, 1994; Aanen et al. 2002, 2009); yet, within these groups, large differences in interaction specificity are observed. Some are highly specific, e.g., all Macrotermes natalensis studied so far associate with members from a single biological species of Termitomyces, whereas others have more diffuse co-evolutionary relationships; for example, most species of the genus Odontotermes associate with a broad range of Termitomyces lineages, which represent multiple species (Aanen, 2006; de Fine Licht et al., 2006; Aanen et al., 2007).

To explain the observed co-cladogenesis and differences in interaction specificity between termite-fungus associations, several studies have suggested that the substrates provided by termites play a role in the selection of a suitable fungal symbiont.
Lefèvre, 2000; Nobre and Aanen, 2012). The hypothesis is that the most specific associations have evolved most complementarity in the breakdown of plant biomass. According to this hypothesis, the less specifically cultivated Termitomyces, like the ones cultivated by Odontotermes spp., are expected to have a broader potential substrate range than very specifically cultivated Termitomyces, like the symbiont of M. natalensis. In addition, different termite species and genera collect different plant substrates, which could further select for association-specific metabolic capacities in the fungal symbionts (Grassé, 1982; Dangerfeld and Schuurman, 2000; Johjima et al., 2006; Soleymanjanejad et al., 2014; da Costa et al., 2018).

Here we explore the extent to which Termitomyces performance on different substrates is correlated with interaction specificity in the termite-fungus symbiosis. First, we generate a phylogeny of nine Termitomyces isolates to compare to previous work and confirm interaction specificities (Aanen et al., 2002, 2007). Second, we test whether there are qualitative or quantitative differences in vitro cultivar growth performance on 35 carbon sources between Termitomyces symbionts of the more specific M. natalensis and two less specific Odontotermes species. Third, as growth substrates are not expected to differ one-dimensionally (i.e., for carbon source only) between termite species, we use a geometric framework approach to generate nutritional landscapes by which we can visualize in vitro cultivar growth performance upon varying carbohydrate and protein concentrations and ratios simultaneously (Lee et al., 2008; Dussutour et al., 2010; Simpson and Raubenheimer, 2012; Shik et al., 2016). Finally, we identify whether the observed growth patterns, separately for the carbon-source and the nutritional-landscape experiments, reflect the interaction specificity between the fungi and their hosts.

2. Materials and methods

2.1. Termite collections and fungal isolations

We studied Termitomyces fungi from mature colonies for which selection of the resident fungal cultivar has already taken place. Fungus comb samples were collected from nine fungus-growing termite nests in 2015 at three geographical locations in South Africa (Table 1). Samples were collected from the termite species Odontotermes sp. (Od127, Od128 and Od159), Odontotermes cf. badius (Od145, Od150 and Od151), and M. natalensis (Mn156, Mn160 and Mn162), for which the termite species had previously been determined (Otani et al., 2014; da Costa et al., 2018) (Table 1). Mature, nodule-containing parts of the fungus comb were collected, placed into plastic bags and taken to the laboratory. Termitomyces fungal nodules were picked from clean parts of the fungus comb (no visible soil particles under a binocular microscope) with a sterile needle and placed on Petri plates with Malt Yeast Agar (MYA: 20 g malt, 2 g yeast extract, 15 g agar in 1 L distilled water). Fungal growth was monitored daily to check the purity of the isolates.

2.2. Termitomyces barcoding and phylogenetic analysis

Termitomyces DNA was isolated using a cetlytrimethylammonium bromide (CTAB) extraction. For isolates from mounds Od128, Od145, Od150, Od151 and Mn162, part of the nuclear ribosomal region, including both internal transcribed spacer (ITS) regions and the 5.8S ribosomal RNA (ITS1, 5.8S and ITS2), were amplified and sequenced using ITS1 and reverse primer ITS4 (White et al., 1990). Because Termitomyces is present in termite mounds as a heterokaryon, i.e., with two separate haploid nuclei, the total DNA of each isolate can contain two different copies for each region of the genome, which was the case for at least one of the two ITS regions in the fungal isolates of mounds Od159, Mn156, Mn160 and Od127. Further, if a length mutation exists between the two different copies of a genomic region, Sanger sequencing will fail. Therefore, we used both forward primers ITS1 and ITS3 and reverse primers ITS2 and IST4 (White et al., 1990) to obtain most of the ITS sequences. We obtained both ITS regions, but not the 5.8S region for Termitomyces from mound Od159. For Mn156 and Mn160 we obtained ITS1 and part of ITS2, and for Od127, part of ITS1, and part of ITS2. Sequences have been deposited to GenBank (MG283253-MG283261) (Table 1). Sequences were aligned using the CLC Genomics Workbench v9.5.3 (https://www.qiagenbioinformatics.com); gap insertion penalty 10, gap extension penalty 1.0, alignment mode "Very accurate". An unrooted tree was obtained by cluster analysis using the UPGMA algorithm with Jukes-Cantor distance measurement, and bootstrap support was assessed using 10,000 pseudoreplicates (Fig. 1C). Using BLASTn, the ITS sequences were compared to haplotypes obtained in a study of South African Termitomyces by Aanen et al. (2007).

2.3. Single nutrient assay: performance on different carbon sources

To determine biomass production of Termitomyces strains on different carbon substrates, we used a minimal medium developed for Schizophyllum commune (Dons et al., 1979, Supplementary Table 1), supplemented with 300 mg Urea/L, where glucose was replaced by one of 35 carbon sources as described at www.fung- growth.org, without birch wood xylan and oat spelt xylan that were no longer available and with chitin added (de Vries et al., 2017). The use of this specific set allows for comparison to other studies gathered in the FUNG-GROWTH database (e.g., Benoit et al., 2015; de Vries et al., 2017). Sterilised polycarbonate membranes (Profiltra, 0.1 pore size, 76 mm diameter, Almere, The Netherlands) were placed in each Petri dish to facilitate fungal biomass collection.

Table 1

| Termite species | GenBank accession numbers for termite identification | Termitomyces Isolate code | GenBank accession numbers for Termitomyces identification | Excavation location | Excavation GPS coordinates |
|-----------------|-----------------------------------------------------|---------------------------|----------------------------------------------------------|---------------------|--------------------------|
| Odontotermes sp. | KJ4590690                                             | Od127                     | MG2832255                                               | Experimental farm   | S25 44.562 E15.391       |
| Odontotermes sp. | KJ4590691                                             | Od128                     | MG2832254                                               | Experimental farm   | S25 44.544 E15.397       |
| Odontotermes cf. badius | MF092801                                    | Od145                     | MG2832258                                               | Experimental farm   | S25 45.118 E15.525       |
| Odontotermes cf. badius | MF092802                                    | Od150                     | MG2832257                                               | Rietondale          | S25 43.666 E14.112       |
| Odontotermes cf. badius | MF092803                                    | Od151                     | MG2832256                                               | Rietondale          | S25 43.650 E14.128       |
| Odontotermes sp. | MF092804                                             | Od159                     | MG2832253                                               | Experimental farm   | S25 44.623 E15.337       |
| Macrotermes natalensis | NA                                                 | Mn156                     | MG2832261                                               | Experimental farm   | S25 44.578 E15.645       |
| Macrotermes natalensis | NA                                                 | Mn160                     | MG2832260                                               | Experimental farm   | S25 44.578 E15.645       |
| Macrotermes natalensis | NA                                                 | Mn162                     | MG2832259                                               | Mookgophong         | S24 39.693 E28.4759      |
and weighing after incubation. *Termitomyces* strains were cultured for 15 d on MYA before roughly the same amount of fungal material from each strain was harvested and added to each of six Eppendorf tubes containing 750 µL 0.6% saline. The suspensions were subsequently pooled in a 12 mL tube and vortexed. Ten µL of this hypha/spore suspension was used as the inoculum. Three inoculates were placed on one plate; three replicate plates were inoculated per fungal strain per carbon substrate (total of nine replicates per fungus, per carbon source). The plates were incubated for 21–31 d (Supplementary Table 2), after which biomass from each Petri dish was collected, separately and weighed (g wet weight). Contaminated plates were discarded (18% out of 945 plates). Plates without membranes were incubated and photographed for visual inspection (Supplementary Fig. 1). Area could, however, not be evaluated, because of the small number of replicates. Principal Component Analysis (PCA) was performed on the average biomass per strain, per carbon source (35 measurements per strain) in R v. 3.3.2 (R Core Team) using the FactoMineR package (Le et al., 2008). Contamination led to missing data points, for which values were estimated using missMDA in R (Jasse and Husson, 2016). The carbon sources that contributed the most to the PCA patterns were evaluated by their cumulative percentage of variance explained, loading values, and eigenvalues (Supplementary Tables 3 and 4).

2.4. Geometric framework assay: performance across protein:carbohydrate ratios and concentrations

*Termitomyces* cultivar biomass formation and radial growth was assessed on media with nine protein:carbohydrate (P:C) ratios (1:9, 1:6, 1:3, 1:2, 1:1, 2:1, 3:1, 6:1, and 9:1) in four different concentrations (8, 32, 56 and 80 g/L protein + carbohydrate) (Dussutour et al., 2010; Shik et al., 2016). The mixtures of carbohydrate and protein were prepared in 250 mL distilled water and 4 g [1.6% (wt:vol)] agar (Supplementary Table 5). Sterilised polycarbonate membranes (Pro.Ultra, 0.1 pore µm) were placed in each plate to facilitate fungal biomass retrieval after incubation. Prior to inoculation, *Termitomyces* strains were cultured for 20 d on PDA plates, and they were harvested and inoculated as described above, except that each plate was inoculated with only a single inoculum. After 24 d, the Petri dishes were photographed (Supplementary Fig. 2), and contaminated plates excluded (1.3% of 3127 plates). The images were used to measure growth area in ImageJ (NIH Image, v.1.50g). After photographing, fungal biomass was harvested and weighed. To analyse differences in biomass formation for different P:C ratios among fungal strains, a PCA was performed in R v. 3.3.2 (R Core Team) using the FactoMineR package (Le et al., 2008). Cumulative percentage variance, loading and eigenvalues were calculated to establish which ratios of P:C contributed the most to the separation observed in the PCA (Supplementary Tables 6 and 7).

To visualize *Termitomyces* growth, we generated P:C landscapes upon which we mapped variation in fungal area (cm²) and biomass (g wet weight) after 24 d. We did this using the “fields” package in v.2.14.0.R to generate non-parametric thin-plate splines (Lee et al., 2008; Dussutour et al., 2010), setting topological resolution of response surfaces to $\lambda = 0.001$ as a smoothing parameter (Shik et al., 2016). We tested whether colony identity influenced growth, performing GLM analyses (proc GLM, SAS V9.3) for each of the three fungal strains (i.e., fungal strains from nests of a given termite host species). Models for biomass and area included the factors protein, carbohydrate, protein x carbohydrate, protein squared, carbohydrate squared, colony identity, and interactions between colony identity and each of the protein and carbohydrate effects. Colony identity was never a significant factor (Supplementary Table 8), so we generated performance landscapes based on strain-level means. For subsequent interpretation of P:C landscapes, we used least-square regressions based on strain-level means with both linear and quadratic terms to evaluate the main and interactive effects of protein and carbohydrates on performance (Supplementary Tables 9, 10, and 11).

3. Results

3.1. *Termitomyces* barcoding and phylogeny

All fungal isolates from *M. natalensis* had identical ITS sequences, apart from the occasional SNP between the two nuclei of the heterokaryons, while there were larger differences in ITS sequence and length between fungal isolates from *Odontotermes* spp. All the sequences were compared to sequences available in Genbank ([https://www.ncbi.nlm.nih.gov/genbank/](https://www.ncbi.nlm.nih.gov/genbank/)). Sequences of two isolates from *Odontotermes* sp. (Od127 and Od128) only differed in two insertions/deletions, and they were most similar to a fungal strain previously isolated from *Odontotermes latericuonis* (Aanen et al., 2007). *Odontotermes* sp. (Od159) was most similar to an isolate from *O. cf. bodius* (Od150) and both were most similar to an isolate from *O. latericuonis* (Aanen et al., 2007). The other two *O. cf. bodius* isolates (Od145 and Od151) were most similar to fungal strains previously isolated from the same termite species (Aanen et al., 2007). The placement of the strains in the phylogeny we generated based on our ITS sequences corroborated the phylogeny of the most similar strains generated by Aanen et al. (2007).

3.2. *Termitomyces* performance across single carbon sources

Contrary to the hypothesis that the less specifically cultivated *Termitomyces* are able to grow on a broader range of substrates, we did not observe qualitative differences between *Termitomyces* strains in growth on different substrates. We did, however, find consistent quantitative differences between *Termitomyces* strains associated with *M. natalensis* and *Odontotermes* spp. The PCA showed that the *Termitomyces* strains associated with *M. natalensis* clustered together and were separated from *Odontotermes* spp. strains (Fig. 2A). Loading values of the PCA indicated that the main contributors to the separation in the first principal component were cotton seed pulp, cellulose, alfalfa meal, arabic gum and soybean hulls (negative loading values), and D-galacturonic acid, D-glucose, L-arabinose, casein and soluble starch (positive loading values) (Supplementary Table 3). The main contributors to the separation in the second component were D-glucuronic acid and lignin (negative loading values), and apple pectin and beechwood xylan (positive loading values) (Supplementary Table 3). There was no effect of incubation time on biomass formation (correlation analysis: $y = 0.0047x + 0.028$, $R^2 = 0.0043$).

Overall, *Termitomyces* strains grew best on complex substrates such as wheat bran, citrus pulp, rice bran, cottonseed pulp, alfalfa meal and soybean hulls, and on the disaccharide cellobiose (Fig. 2B; Supplementary Table 12). Sugar beet pulp was the only complex substrate with limited *Termitomyces* growth. Among the monosaccharides, D-glucose was the only one that allowed substantial biomass formation across all strains. None of the strains formed substantial biomass on lignin. Also, little biomass was formed on sucrose, even though some biomass was formed on D-glucose and...
D-fructose, the monosaccharides that build sucrose. *Odontotermes* spp. strains grew slightly better on D-glucose, D-fructose, and chitin than *M. natalensis* strains, while *M. natalensis* strains showed slightly better growth on arabic gum (Fig. 2B).

### 3.3. Cultivar performance across protein:carbohydrate landscapes

In contrast to the results obtained on biomass formation patterns on different carbon sources, the PCA of biomass on different P:C ratios and concentrations indicated clustering based on termite species (Fig. 3C). Fungal cultivar performance was maximized by specific blends of protein and carbohydrates (Supplementary Tables 13 and 14), with strains exhibiting carbohydrate-biased ‘bulls eyes’ for maximal biomass production around concentrations of 35 g/L carbohydrates and 20 g/L protein (Fig. 3A; Supplementary Tables 9, 10 and 11). The effects of carbohydrates on fungal biomass production interacted with protein availability (i.e., quadratic terms for C were significant for each of the strains; Supplementary Table 11), indicating that single nutrient assays of cultivar performance (e.g., the carbon source test) need to be interpreted with caution. Visualized on the performance landscapes, the same carbohydrate concentration (i.e., a horizontal line extending from the y axis at 32 g/L) that was associated with the highest cultivar biomass when present in slightly higher abundance relative to protein (i.e., red bullseye at 1:2 to 1:3 P:C) was also associated with low biomass production at higher protein levels (i.e., blue areas to the right of red bullseyes, with diets > 2:1 P:C) (Fig. 3A).

The P:C ratios and concentrations that maximized growth area differed visually from those for biomass formation (Fig. 3A) and statistically, as the quadratic term for P was significant in each of the radial area regressions (but not in the biomass regressions, Supplementary Table 11). Regions of maximal growth area also appeared to differ across *Termomyces* strains from different termite species (Fig. 3B). The strains from *M. natalensis* showed a P:C diet maximum for radial growth that was comparable to the maximum for biomass formation (between 1:2 and 1:3, and between 32 and 56 g/L) and a second at P:C 3:1 and 6:1 at 8 g/L (Fig. 3B). The strains from *O. cf. badius* were similar to *M. natalensis*, while *Odontotermes* spp. strains only displayed one maximum (P:C 3:1 to 9:1 at 8 g/L) (Fig. 3B).

### 4. Discussion

#### 4.1. Termite-fungus interaction specificity and its associations with *Termomyces* growth

Our phylogeny of fungi isolated from *M. natalensis* and two species of *Odontotermes* revealed patterns of interaction specificity consistent with previous work (Aanen et al., 2002, 2007), demonstrating that *M. natalensis* colonies associate with a single *Termomyces* species, while symbionts associated with *Odontotermes* spp. are genetically more variable and without one-to-one termite-to-fungus species relationships. Our in vitro assays suggest that *Termomyces* nutritional requirements could contribute to interaction specificity with termite hosts. The patterns of biomass formation on different carbon sources separated *M. natalensis* from *Odontotermes* spp. strains, but not *Odontotermes* spp. from each other (Fig. 2B), consistent with higher specificity in *M. natalensis*- *Termomyces* interactions. Yet, the in vitro growth patterns in our
two-factor geometric framework experiment, also separate the strains of the two Odontotermes species.

4.2. Termitomyces performance of different carbon sources

The termite species included in this study primarily harvest decaying wood and leaf litter (Wood and Sands, 1978), and it was thus unsurprising that all Termitomyces strains performed well on complex carbon sources (Visser et al., 2011; Poulsen et al., 2014). Our findings do, however, suggest that M. natalensis should preferentially collect leaf litter and dead wood that are rich in cellulose, while Odontotermes should preferentially collect substrates richer in mono-, di- and oligosaccharides (Fig. 2B). Substrate preference is generally poorly understood in fungus-farming termites, but variation does indeed exist: fresh leaves (e.g., Macrotermes mülleri, Macrotermes ivorensis, Macrotermes michaelseni, and Odontotermes formosanus; Grassé, 1982; Dangerfield and Schuurman, 2000; Soleymanimejadian et al., 2014), grass stalks (e.g., Macrotermes bellicosus and Macrotermes subhyalinus; Grassé, 1982), roots (e.g., M. michaelseni, Ancistrotermes sp., and Odontotermes sp.; Dangerfield and Schuurman, 2000), and mammal dung (e.g., Odontotermes sp., O. cf. badius, M. natalensis, M. michaelseni; Dangerfield et al., 1978; da Costa et al., 2018). Consistently, a recent comparison found higher expression of cellulases in M. natalensis than in Odontotermes sp. fungus combs (da Costa et al., 2018), and Johjima et al. (2006) found that pectinases and hemicellulases were expressed more than cellulases in Termitomyces associated with Macrotermes gilvus, which harvests grasses and leaves.

All Termitomyces strains formed more biomass on cellobiose than on D-glucose, the building block of cellobiose. This may appear counterintuitive, since the degraded entities that are taken up by the fungus are monosaccharides (Allaway and Jennings, 1970), but is likely because we standardized carbon source concentrations and not carbon content, and cellobiose has more total carbon (and hence higher energetic value) than D-glucose. This appears to be unproblematic for our interpretations, as there was no overall correlation between the number of carbon molecules and biomass formed for the mono-, di- and oligosaccharides for which we could estimate energetic values (y = 0.003x + 0.038, R² = 0.019). Biomass formation is thus more likely associated with metabolic adaptations to certain carbon sources, consistent with the higher expression of enzymes targeting these carbon sources (e.g., cellobiohydrolases) in fungus combs (da Costa et al., 2018). Cellobiohydrolases convert cellobiose to monosaccharides, which likely leads to increased glucose levels in guts of old workers at the final step of plant decomposition (da Costa et al., 2018), and consistent with gut bacteria putatively breaking down and assimilating simple sugars (Poulsen et al., 2014). The potential importance of nutrient co-limitation (evident from the geometric framework experiment) represents a caveat to the above interpretation, since carbohydrate-mediated cultivar growth depends on protein availability.

Termite species forage on different substrates in different geographical locations (da Costa et al., 2018), so Termitomyces growth likely varies in response to the availability of substrates in given habitats. Analyses of a Termitomyces genome from M. natalensis indicate that the cultivar should be able to decompose most complex polysaccharides (Poulsen et al., 2014). This implies that the substrates should not govern symbiont selection alone; however, variation in forage availability between habitats could affect what enzymes are expressed. Further, because multiple fungal symbiont species associate with multiple Odontotermes species, genome content and enzymatic capacity variation between symbionts could be larger and allow termite host species to utilise a wider range of substrates. Additional -omics approaches will be needed to fully characterise the standing variation and the relative roles of genome differences vs. gene expression differences between Termitomyces species.

4.3. Fungal performance across gradients of P:C blends and termite species-specific farming strategies

The geometric framework provides a powerful graphical
approach for visualizing how diverse foraging organisms from slime molds (Dussutour et al., 2010) to humans (Simpson and Raubenheimer, 2012) prioritize competing nutritional requirements when confronted with imbalanced resources. We used this approach to visualize nutritional landscapes and test for nutritional specificity differences between cultivar strains. While cultivars exhibit nearly identical nutritional P:C targets for maximum biomass production (Fig. 3A), they appear to exhibit strain-specific targets for growth area (Fig. 3B). Thus, if nutrients act as a filter among competing strains in incipient nests, they likely favour cultivars best able to exploit nutrients through rapid radial growth than those able to acquire most biomass over time. This may be driven by heightened starvation responses, with low macronutrient concentrations (especially carbohydrates) inducing extensive radial growth (cf., Boddy, 1999; Boswell et al., 2002; Heaton et al., 2010).

An important next step will be to define nutritional mixtures present in each of the resources harvested by termite colonies in nature. By overlaying these maps of nutrient availability on growth maps, the degree of overlap between the ‘realized niche’ (nutrients actually provided by termites) and the ‘fundamental niche’ (resources that maximize cultivar performance) can be evaluated (Raubenheimer, 2011). A reasonable hypothesis is that the substrates available to termite foragers in nature have low overall protein concentrations and low nutritional variability. For instance, the carbohydrate bias of optimal biomass formation may reflect that dead wood can contain less than 0.5% nitrogen (LaFage and Notting, 1978) and fungus combs contain 1.9% and 41% nitrogen and carbon, respectively (Arshad and Schnitzer, 1987). The P:C cultivar growth has additional eco-evolutionary importance, since the existence of nutritional ‘bullseyes’ implies that termites may have benefit from providing cultivars with a specific blend of nutrients (i.e., to target the fundamental niche of their cultivar). This in turn implies a basis for selection of sensory mechanisms in the termite host, and potentially among termite species growing different cultivar species, to identify and select those nutritional blends.

4.4. Termite colony foundation and forage use may affect fungal cultivar selection

The ecological dynamics by which fungus-growing termites select their symbionts during early phases of colony development remain unknown. One possibility is that the first termite workers selectively pick up Termitomyces spores from the species with which they normally associate, along with plant substrate to start the fungal comb. This would, however, require recognition capabilities in the termites (Grasse and Noriot, 1958; Sands, 1960; Aanen et al., 2002). Sieber (1983) demonstrated that provisioning of Termitomyces spores to incipient Odontotermes montanus and M. michaelseni laboratory colonies allowed for the colonization of a primordial fungus comb (comprised of soil) by Termitomyces. However, these experiments did not involve testing whether only specific fungal species were collected. Alternatively, multiple Termitomyces species may randomly be collected by the first termite workers if spores are brought in with plant forage coincidentally. In this case foraged plant material could contribute to drive the subsequent competitive selection within the nest prior to the establishment of a monoculture.

Our findings provide support to the interaction specificity between fungus-growing termites and Termitomyces and indicate that specificity between termite hosts and fungi is not only visible from phylogenetic reconstructions, but also reflected physiologically. Differences between fungal symbionts of different hosts could facilitate variation in fungal colonization success of the substrate collected by specific termites in incipient nests, potentially contributing to maintaining termite-fungus interaction specificity if physiological differences cause variable growth during competition between multiple fungal strains at the onset of fungus garden formation.

4.5. Data accessibility

Termitomyces ITS sequences have been deposited in GenBank.

Author contributions

RRDC and SMEV designed the experiments and analyses, collected colonies in the field, isolated fungal strains, carried out the growth assay on different carbon sources, and drafted the first version of the figures, tables, and manuscript; RRDC and JZS designed the P:C experiment, which was carried out by RRDC with analytical help from JZS; SMEV extracted DNA and analysed Termitomyces sequences; DKA and MP supervised SMEV and RRDC, respectively, helped design the study, and contributed with comments on analyses, figures, tables, and text. All authors contributed to writing the manuscript.

Acknowledgements

We thank Z. Wilhelm de Beer, Michael J. Wingfield, and the staff and students at the Forestry and Agricultural Biotechnology Institute, University of Pretoria, for hosting field work; Christine Beelemanns, René Benndorf, Maria Otani, Benjamin H. Conlon, Jane de Verges, Nick Bos, and Jeremy M. Thomas-Poulson for help with excavations; Mandy Runderkamp, Margo Wisselink, and Kasun Bodawatta for laboratory assistance; and Ben Auxier for critically reading of the manuscript. This study was funded by the CAPES Foundation, Ministry of Education, Brazil (grant BEX: 13240/13-7) to RRDC, The Netherlands Organisation for Scientific Research Grant (ALW open competition 824.01.002) to SMEV, a Marie Curie International Incoming Fellowship (327940) to JZS, and a Villum Kann Rasmussen Young Investigator Fellowship (VKR10101) to MP.

Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.funeco.2018.08.009.

References

Aanen, D.K., de Fine Licht, H.H., Debets, A.J.M., Kerstes, N.A.G., Hooekstra, R.F., Boomsma, J.J., 2009. High symbiont relatedness stabilizes mutualistic cooperation in fungus growing termites. Science 326, 1103–1106.
Aanen, D.K., Ros, V.I.D., de Fine Licht, H.H., Mitchell, J., de Beer, Z.W., Slippers, B., Rouland-LeFevre, C., Boomsma, J.J., 2007. Patterns of interaction specificity of fungus-growing termites and Termitomyces symbions in South Africa. BMC Evol. Biol. 7, 115.
Aanen, D.K., 2006. As you reap, so shall you sow: coupling of harvesting and inoculating stabilizes the mutualism between termites and fungi. Biol. Lett. 2, 209–212.
Aanen, D.K., Eggleton, P., 2005. Fungus-growing termites originated in African rain forest. Curr. Biol. 15, 851–855.
Aanen, D.K., Eggleton, P., Rouland-LeFevre, C., Guldberg-Frøslev, T., Rosendahl, S., Boomsma, J.J., 2002. The evolution of fungus-growing termites and their mutualistic fungal symbionts. Proc. Natl. Acad. Sci. 99, 14887–14892.
Allaway, A.E., Jennings, D.H., 1970. The influence of Cations on Glucose Uptake by Fungus Dendryphilla salina’, vol. 69. The New Physiologist, pp. 567–579.
Arshad, M.A., Schnitzer, M., 1987. The chemistry of a termite fungus comb. Plant Soil 98, 247–256.
Benoit, L., Culleton, H., Zhou, M., DiFalco, M., Aguilar-Osorio, G., Bettaglia, E., Bouzdj, O., Brouwer, C.P.J.M., El-Bushari, H.B.O., Coutinho, P.M., Gruben, B.S., Hildén, K.S., Houbraken, J., Barboza, L.A.J., Levasseur, A., Majoer, E., Mäkelä, M.R., Narang, H.M., Trejo-Agular, B., van den Brik, J., vanKuyk, P.A., Wiebenga, A., McKie, V., McClearly, B., Tsang, A., Henriassat, B., de Vries, R.P., 2015. Closely
related fungi employ diverse enzymatic strategies to degrade plant biomass. Biotechnol. Biofuels 8, 107.

Boddy, L., 1990. Saprotrophic cord-forming fungi: meeting the challenge of a heterogenous environments. Mycologia 91, 13–32.

Bowtell, G., Jacobs, H., Davindson, F.A., Gadd, G.M., Ritz, K., 2002. Functional consequences of nutrient translocation in mycelial fungi. J. Theor. Biol. 217, 459–477.

Bright, M., Bulgheresi, S., 2010. A complete complex: transmission of microbial symbionts. Nat. Rev. Microbiol. 8, 218–230.

Dangor, J.M., Schuurman, G., 2000. Foraging by fungus-growing termites (isoptera Termitidae, Macrotermiinae) in the Okavango Delta, Botswana. J. Trop. Ecol. 15, 717–731.

Dangor, J.M., McCarthy, T.S., Ellery, W.N., 1978. The mound-building termite Macrotermes michaelseni as an ecosystem engineer. J. Trop. Ecol. 14, 507–520.

Darlington, J.F.E.C., 1994. Nutrition and evolution in fungus-growing termites. In: Hunt, J.H., Nalepa, C.A. (Eds.), Nourishment and Evolution in Insect Societies. Westview Press, Boulder, pp. 105–130.

Dans, J.J.M., de Vries, O.M.H., Wessels, J.G.H., 1979. Characterization of the genome of the basidiomycete Schizophyllum commune. Biochim. Biophys. Acta 563, 100–112.

daca, R.R., Hu, H., Pilgaard, B., Vreeburg, S.M.E., Schuckel, J., Pedersen, K.S.K., Kracun, S.K., Busk, P.K., Harholt, J., Sapountzis, P., Lange, L., Aanen, D.K., Poulsen, M., 2018. Enzyme activities at different stages of plant biomass decomposition in three species of fungus-growing termites. Appl. Environ. Microbiol. 84, e01815–e01817.

de Fine Licht, H.H., Boomsma, J.J., Aanen, D.K., 2006. Presumptive horizontal symbiont transmission in the fungus-growing termite Macrotermes natalensis. Mol. Ecol. 15, 3131–3138.

de Vries, R.P., Riley, R., Wiebenga, A., Aguilar-Osorio, G., Amillis, S., Uchima, C.A., Anderlah, G., Asadollahi, M., Askin, M., Barry, K., Battaglia, E., Bayram, Ö., Benocci, T., Braus-Stromeyer, S.A., Caldana, C., Cánovas, D., Cerqueira, G.C., Chen, F., Chen, W., Choi, C., Clum, A., et al., 2017. Comparative genomics reveals high biological diversity and specific adaptations in the industrially and medically important fungal genus Aspergillus. Genome Biol. 18, 28.

Dussutour, A., Latty, T., Beekman, M., Simpson, S.J., 2010. Amoeboid organism solves complex nutritional challenges. Proc. Natl. Acad. Sci. U.S.A. 107, 4607–4611.

Ellers, J., Kiers, E.T., McDonald, R.B., Visser, A., 2012. Ecological interactions drive the colonization of Madagascar by fungus-growing termites. Proc. R. Soc. Lond. B Biol. Sci. 277, 359–365.

Ontani, S., Mikaelyan, A., Nobre, T., Hansen, L.H., Koné, N.A., Sørensen, S.J., Aanen, D.K., Boomsma, J.J., Brune, A., Poulsen, M., 2014. Identifying the core microbial community in the gut of fungus-farming termites. Mol. Ecol. 23, 4631–4644.

Poulsen, M., Hu, H., Li, C., Chen, Z., Xu, L., Otani, S., Nygaard, S., Nobre, T., Klaubauf, S., Schindler, P.M., Hauser, F., Pan, H., Yang, Z., Sonnenberg, A.S.M., de Beer, Z.W., Zhang, Y., Winglefield, M.J., Gummelmachuizen, C.J.P., de Vries, R.F., Korb, J., Aanen, D.K., Wang, J., Boomsma, J.J., Zhang, G., 2014. Complementary symbiotic contributions to plant decomposition in a fungus-farming termite. Proc. Natl. Acad. Sci. U.S.A. 111, 14509–14505.

R Core Team, 2013. R: a language and environment for statistical computing. Available at: http://www.R-project.org.

Raubenheimer, D., 2011. Toward a quantitative nutritional Ecology: the right-angled mixture triangle. Ecol. Monogr. 81, 407–427.

Roberts, E.M., Todd, C.N., Aanen, D.K., Nobre, T., Hilbert-Wolf, H.L., O’Connor, P.M., Tapanila, L., Metela, C., Stevens, N.J., 2016. Oligocene termite nests with in situ fungus gardens from the rukwa rift basin, Tanzania, support a paleogene african origin for insect agriculture. PLoS One 11, e0156847.

Roulund-Lefèvre, C., 2000. In: Abe, T., Bignell, D.E., Higashi, M. (Eds.), Termites: Evolution, Sociality, Symbiosis, Ecology. Kluwer Academic, Dordrecht, pp. 289–306.

Roulund-Lefèvre, C., Bignell, D.E., 2002. Cultivation of symbiotic fungi by termites of the subfamily Macrotermiinae. In: Joseph, Sebbach (Ed.), Symbioses: Mechanisms and Model Systems. Springer Netherlands, Dordrecht, pp. 731–756.

Sands, W.A., 1960. The initiation of fungus comb construction in laboratory colonies of Macrotermitlines guineensis (Silvestri). Insectis Sociaux 7, 251–263.

Shik, J.Z., Gomez, E.B., Kooij, P.W., Santos, J.C., Wcislo, W.T., Boomsma, J.J., 2016. Nutrition mediates the expression of cultivar–farmer conflict in a fungus-growing ant. Proc. Natl. Acad. Sci. U.S.A. 113, 10121–10126.

Sieber, R., 1983. Establishment of fungus comb in laboratory colonies of Macrotermes michaelseni and Odontotermes mutans (Isoptera, Macrotermiinae). Insectis Sociaux 30, 204–209.

Simpson, S.J., Raubenheimer, D., 2012. The Nature of Nutrition: a Unifying Frame- work from Animal Adaptation to Human Obesity. Princeton University Press, Princeton, New Jersey, p. 239.

Solyamaninejad, E., Ji, B.Z., Liu, S.W., Zhang, X.W., 2014. Foraging polyethism in Pseudoxylaria, Termitomyces Loss of lipid synthesis as an evolutionary consequence of a parasitic lifestyle. In: Shik, J.Z., Gomez, E.B., Kooij, P.W., Santos, J.C., Wcislo, W.T., Boomsma, J.J., 2016. Nutrition mediates the expression of cultivar–farmer conflict in a fungus-growing ant. Proc. Natl. Acad. Sci. U.S.A. 113, 10121–10126.

Sieber, R., 1983. Establishment of fungus comb in laboratory colonies of Macrotermes michaelseni and Odontotermes mutans (Isoptera, Macrotermiinae). Insectis Sociaux 30, 204–209.

Simpson, S.J., Raubenheimer, D., 2012. The Nature of Nutrition: a Unifying Frame- work from Animal Adaptation to Human Obesity. Princeton University Press, Princeton, New Jersey, p. 239.

Solyamaninejad, E., Ji, B.Z., Liu, S.W., Zhang, X.W., 2014. Foraging polyethism in Pseudoxylaria, Termitomyces

Shik, J.Z., Gomez, E.B., Kooij, P.W., Santos, J.C., Wcislo, W.T., Boomsma, J.J., 2016. Nutrition mediates the expression of cultivar–farmer conflict in a fungus-growing ant. Proc. Natl. Acad. Sci. U.S.A. 113, 10121–10126.

Sieber, R., 1983. Establishment of fungus comb in laboratory colonies of Macrotermes michaelseni and Odontotermes mutans (Isoptera, Macrotermiinae). Insectis Sociaux 30, 204–209.

Simpson, S.J., Raubenheimer, D., 2012. The Nature of Nutrition: a Unifying Frame- work from Animal Adaptation to Human Obesity. Princeton University Press, Princeton, New Jersey, p. 239.

Solyamaninejad, E., Ji, B.Z., Liu, S.W., Zhang, X.W., 2014. Foraging polyethism in Pseudoxylaria, Termitomyces Loss of lipid synthesis as an evolutionary consequence of a parasitic lifestyle. In: Shik, J.Z., Gomez, E.B., Kooij, P.W., Santos, J.C., Wcislo, W.T., Boomsma, J.J., 2016. Nutrition mediates the expression of cultivar–farmer conflict in a fungus-growing ant. Proc. Natl. Acad. Sci. U.S.A. 113, 10121–10126.

Sieber, R., 1983. Establishment of fungus comb in laboratory colonies of Macrotermes michaelseni and Odontotermes mutans (Isoptera, Macrotermiinae). Insectis Sociaux 30, 204–209.