Targeting microglial population dynamics in Alzheimer’s disease: are we ready for a potential impact on immune function?

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Abbreviations: AD, Alzheimer’s disease; GWASs, genome-wide association studies; Aβ, amyloid beta; LOAD, late onset AD; EOAD, early onset AD; SNPs, single-nucleotide polymorphisms; CSF1R, colony-stimulating factor 1 receptor; ALS, amyotrophic lateral sclerosis; CSF-1, colony stimulating factor 1; IL-34, interleukin 34; TGFβ-1, transforming growth factor beta-1; AML, acute myeloid leukemia; BBB, blood brain barrier; PD, Parkinson’s disease; RA, rheumatoid arthritis; TK, tyrosine kinase

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ABSTRACT

Alzheimer’s disease (AD) is the most common form of dementia, affecting two-thirds of people with dementia in the world. To date, no disease-modifying treatments are available to stop or delay the progression of AD. This chronic neurodegenerative disease is dominated by a strong innate immune response, whereby microglia plays a central role as the main resident macrophage of the brain. Recent genome-wide association studies (GWASs) have identified single-nucleotide polymorphisms (SNPs) located in microglial genes and associated with a delayed onset of AD, highlighting the important role of these cells on the onset and/or progression of the disease. These findings have increased the interest in targeting microglia-associated neuroinflammation as a potential disease-modifying therapeutic approach for AD. In this review we provide an overview on the contribution of microglia to the pathophysiology of AD, focusing on the main regulatory pathways controlling microglial dynamics during the neuroinflammatory response, such as the colony-stimulating factor 1 receptor (CSF1R), its ligands (the colony stimulating factor 1 and interleukin 34) and the transcription factor PU.1. We also discuss the current therapeutic strategies targeting proliferation to modulate microglia-associated neuroinflammation and their potential impact on peripheral immune cell populations in the short and
long-term. Understanding the effects of immunomodulatory approaches on microglia and other immune cell types might be critical for developing specific, effective and safe therapies for neurodegenerative diseases.

1 INTRODUCTION

Alzheimer’s disease (AD) is a chronic neurodegenerative disease and the most common form of dementia in the world, contributing to 60-70% of cases. It is estimated that currently over 50 million people are affected by dementia worldwide, according to the World Health Organisation and the recent report published by Alzheimer’s Disease International (ADI) (International, 2019). The total number of people with dementia is predicted to reach 82 million by 2030 and 152 million by 2050, causing an estimated economic burden of 2 trillion US$ globally (International, 2019). AD is mostly diagnosed in people over 65 years-old, termed as late onset AD (LOAD), with around 5% of AD cases being diagnosed in individuals under the age of 65, classified as early onset AD (EOAD) (Mendes, 2012). Despite these alarming figures, no disease-modifying treatment is currently available and the cause of sporadic AD is still unclear.

Clinically, AD manifests as a gradual decline in cognitive functions including loss of memory, dyspraxia, disorientation and aphasia, accompanied by behavioral changes such as irritability, aggressiveness, anxiety and social withdrawal (Atri, 2019). Patients are usually diagnosed based on cognitive assessments, assuming that AD neuropathologic changes will be found post-mortem. However, from 10% to 30% of patients clinically diagnosed as AD do not show AD neuropathological changes at autopsy (Jack et al., 2018), suggesting that cognitive symptoms are not the ideal method to diagnose AD. According to the updated National Institute of Aging and Alzheimer’s Association Research Framework, AD should be diagnosed by the detection of biomarkers indicative of neuropathologic changes, independently of clinical symptoms (Jack et al., 2018). This characterization is possible using PET imaging and/or assessment of biomarkers present in cerebrospinal fluid (Jack et al., 2018), although these methods are not currently being used broadly for individuals with symptoms, instead limited to early-onset, rapidly progressive or atypical cases (Atri, 2019). The main features of the pathology of AD are the accumulation of extracellular amyloid-beta (Aβ) plaques and intracellular neurofibrillary tangles of hyperphosphorylated Tau, dystrophic neurites, neuronal loss and brain atrophy (Gjoneska et al., 2015). In the last decades, several hypotheses have been explored to explain the pathogenesis of AD, being the amyloid cascade hypothesis the prevailing mechanistic theory so far. This hypothesis postulates that the neurodegeneration in AD is caused by an abnormal accumulation of Aβ protein plaques in several regions of the brain, such as the pre-frontal cortex, temporal and parietal lobe and hippocampus, causing memory and cognitive impairment and eventually leading to dementia (Hardy & Higgins, 1992; Karran, Mercken, & De Strooper, 2011). Many drugs targeting this pathway have been developed and entered clinical trials in recent years. However, none of these therapies have yet been successful in preventing the development or progression of the disease. This is possibly due to the existence of alternative pathways that are disrupted in AD and not directly considered in the amyloid cascade hypothesis, which present a high therapeutic potential as alternatives or in combination with the current strategies.

Neuroinflammation associated to AD was long considered a consequence of the pathology. However, it is now well accepted that neuroinflammation is a key player in several neurodegenerative diseases, including AD. The neuroinflammatory process that takes place in these diseases is characterised by a strong activation of the innate immune system, in which microglia plays a central
role as the main resident macrophages in the brain (Simon, Obst, & Gomez-Nicola, 2019). Microglia are able to respond to harmful stimuli in the brain including Aβ proteins, acting as the main regulators of the neuroinflammatory response associated with brain disease (Gomez-Nicola & Perry, 2015). In response to damage, microglia shows an activated phenotype accompanied by an increase in their proliferation and increased expression of inflammatory markers (Olmos-Alonso et al., 2016). This activation process is critical and postulated to play a beneficial role in the acute neuroinflammatory response, resulting in the engulfment of debris and dead cells to minimize and repair the brain damage (Cai, Hussain, & Yan, 2014; Calsolaro & Edison, 2016). However, the sustained activation of microglia observed in neurodegenerative diseases leads to a chronic neuroinflammatory response and an overproduction of inflammatory mediators, such as pro-inflammatory cytokines and reactive oxygen species, which are known to cause damage and neurodegeneration (Cai et al., 2014; Calsolaro & Edison, 2016; Lyman, Lloyd, Ji, Vizaychipi, & Ma, 2014). The generated damage keeps microglia in an over-activated state, thus preventing these cells from returning to their homeostatic and beneficial functions and worsening the disease. It has been shown that TREM2 is critical in regulating the balance between the homeostatic and the disease-associated microglial states (Nichols et al., 2019), stimulating phagocytosis and suppressing cytokine production and inflammation (Guerreiro et al., 2013). Genetic studies have recently identified mutations of this receptor strongly associated with risk of AD (Guerreiro et al., 2013; Jonsson et al., 2013), supporting the idea of a causative link between inflammatory cells and neurodegeneration. It has been suggested that non-steroidal anti-inflammatory drugs have a protective role in the onset or progression of AD (Hoozemans, Veerhuis, Rozemuller, & Eikelenboom, 2011), although most clinical trials to date have failed to show this beneficial effect. However, this idea is strongly supported by recent genome-wide association studies (GWAS), which have identified new single-nucleotide polymorphisms (SNPs) in immune-related genes associated with AD risk, such as the above cited Trem2 (Efthymiou & Goate, 2017; Hansen, Hanson, & Sheng, 2018; Huang et al., 2017; Verheijen & Sleegers, 2018). Most of these SNPs encode for proteins that are mainly expressed in microglia, strongly supporting a causal involvement of microglial cells in the development and progression of AD. These findings have attracted the effort of drug discovery programs aimed at targeting microglia-associated neuroinflammation as a potential disease-modifying therapeutic approach for AD. In this review we provide an overview of the main pathways controlling microglial activation and proliferation during the neuroinflammatory response and their contribution to the pathophysiology of AD. We also summarize the current therapeutic strategies to modulate microglial-associated neuroinflammation through targeting proliferation and highlight their potential impact on other immune cell populations in the systemic compartment.

2 REGULATION OF MICROGLIAL PROLIFERATION AND NEUROINFLAMMATION IN HEALTH AND AD

In recent years, GWAS studies have identified over 25 genetic loci associated with risk of LOAD, many of them related to neuroinflammation and mainly expressed in microglial cells, such as ApoE, Spi1 and Trem2 (Corder et al., 1993; Guerreiro et al., 2013; Huang et al., 2017; Jonsson et al., 2013). These findings directly implicate microglial and immune genes as key players in the development and progression of AD (Efthymiou & Goate, 2017). The neuroinflammatory response in AD is characterized by increased number of microglia cells showing an activated phenotype (Akiyama et al., 2000; Edison et al., 2008; Heneka, Golenbock, & Latz, 2015; Olmos-Alonso et al., 2016), increased expression of pro-inflammatory cytokines and chemokines (Dickson, Lee, Mattiace, Yen, & Brosnan, 1993; Fernandez-Botran et al., 2011) and an impairment in their phagocytic activity and Aβ clearance (Cai et al., 2014; Wendt et al., 2017).
2.1 Targeting CSF1R in AD

The main system controlling the differentiation, maintenance and proliferation of microglia in both healthy and pathological conditions is the colony-stimulating factor 1 receptor (CSF1R) pathway. CSF1R is encoded by the c-fms proto-oncogene (Sherr et al., 1985) and belongs to the type III tyrosine kinase family (Pixley & Stanley, 2004). This receptor is highly expressed by myeloid cells and its activation through the phosphorylation of the tyrosine residues stimulates many downstream signaling pathways (Pixley & Stanley, 2004; Rojo, Pridans, Langlais, & Hume, 2017; Stanley & Chitu, 2014; Wang & Colonna, 2014). CSF1R genetic variants have been found by genetic screening in neuropathologically confirmed AD patients and these mutations are strongly associated to LOAD susceptibility (Sassi et al., 2018). Moreover, CSF1R upregulation and an increase in microglial proliferation have been found in post-mortem samples from patients with AD (Akiyama et al., 1994; Gomez-Nicola, Fransen, Suzzi, & Perry, 2013; Olmos-Alonso et al., 2016). Studies published by our group showed that microglial proliferation increases progressively in proximity to Aβ plaques in the APP/PS1 murine model of AD, suggesting that microglial activation and proliferation is triggered by Aβ deposition (Olmos-Alonso et al., 2016). It has also been shown that the pharmacological inhibition of the tyrosine kinase (TK) activity of CSF1R decreases microglial proliferation and impedes the degeneration of synapses, ameliorating the progression of the disease without modifying the levels of Aβ in the APP/PS1 model (Olmos-Alonso et al., 2016). Similar effects have been also shown in several experimental models of neurodegenerative disease, including prion disease (Gomez-Nicola et al., 2013) and amyotrophic lateral sclerosis (ALS) (Martinez-Muriana et al., 2016). These results are also observed after administration of a potent CSF1R inhibitor leading to partial depletion of the microglial population in the 3xTg (Dagher et al., 2015) and 5xFAD models (Sosna et al., 2018; Spangenberg et al., 2016) of AD-like pathology. Microglial depletion strategies were also tested in aged Tg2576 mice with no effect on tau pathology (Bennett et al., 2018). However, a recent study from our group has validated the inhibition of CSF1R as a disease-modifying mechanism in the P301S mouse model of tauopathy. This report demonstrates that inhibition of CSF1R reduces the expansion of the microglial population and the expression of pro-inflammatory cytokines such as IL-1β and TNFα at mRNA and protein levels (Mancuso et al., 2019). Blockade of microglial proliferation and the repolarization of these cells to a homeostatic phenotype attenuate neuronal degeneration and ameliorate tau pathology (Mancuso et al., 2019). This repolarization of the microglial inflammatory profile to a homeostatic phenotype has been also observed after the inhibition of CSF1R in the APP/PS1 model of AD (Olmos-Alonso et al., 2016) and other models of neurodegenerative diseases such as multiple sclerosis (Nissen, Thompson, West, & Tsirka, 2018) and a model of Parkinson’s disease (PD) (Neal et al., 2020). Together, these studies provide evidence that reducing the number of microglia, or depleting them, have advantageous consequences, independently of the Aβ load, demonstrating that a disease-modifying approach for AD is achievable through targeting microglia alone.

Two independent ligands can activate CSF1R with high affinity, the colony stimulating factor 1 (CSF-1) (Stanley & Heard, 1977) and interleukin 34 (IL-34) (H. Lin et al., 2008). Both ligands have been shown to promote microglial proliferation (Gomez-Nicola et al., 2013) but also show differential spatiotemporal expression patterns and have complementary biological functionality (Nandi et al., 2012; Wang et al., 2012). Mice lacking IL-34 (Il34LacZ) displayed an acute reduction of microglial cells in the brain and Langerhans cells in the skin, showing that IL-34 is crucial for the development and maintenance of these populations (Greter et al., 2012; Wang et al., 2012). However, the administration of anti-CSF-1 and anti-IL-34 antibodies during development or in postnatal ages revealed that CSF-1 is necessary for the colonization and maintenance of microglia population in the embryonic brain, whereas IL-34 is mainly required for microglial maintenance later during adult life (Easley-Neal, Foreman, Sharma, Zarrin, & Weimer, 2019). In adulthood, CSF-1 is widely expressed...
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and produced by many different mesenchymal and epithelial cell types (Dai et al., 2002; Jones & Ricardo, 2013), whereas the expression of IL-34 is more tissue-restricted, mainly produced by keratinocytes located in the epidermis and neurons in the brain (Wang & Colonna, 2014), showing minimal overlap with the expression pattern of CSF-1 (Nakamichi, Udagawa, & Takahashi, 2013; Wei et al., 2010). The role of IL-34 and CSF-1 in the maintenance of microglial cells during adulthood has been investigated in several studies during recent years. IL-34 was first shown to be required for the maintenance of microglia in the adult brain, whereas CSF-1 seemed to be mainly involved in replacing microglial cells after inflammation (Greter et al., 2012; Wang et al., 2012). However, two recently published reports have shown different effects on the microglia population after peripheral administration of specific anti-IL-34- and anti-CSF-1- monoclonal antibodies in adult mice. In the first one, Lin et al. conclude that IL-34 is crucial for the maintenance and differentiation of microglial cells in the grey matter of adult mice, whereas CSF-1 is a key player in maintaining macrophage homeostasis in several peripheral tissues such as colon and liver (W. Lin et al., 2019). However, Easley-Neal et al. show that the blockade of both molecules leads to the depletion of different microglia populations in the brain of adult mice. The anti-CSF-1 blocking antibody depleted the microglia located in the white matter more effectively, while the anti-IL-34 blocking antibody depleted the microglia in the grey matter more efficiently, phenocopying the regional expression pattern of each ligand (Easley-Neal et al., 2019). Taking together, all this evidence suggests that CSF-1 and IL-34 are required differentially during development and for the maintenance of the microglial population in the adult brain. In AD and AD-like transgenic mice, CSF-1 was shown to be upregulated and played an essential role in the proliferation of microglia occurring as a consequence of the pathological activation in disease (Murphy, Zhao, Yang, & Cordell, 2000; Vincent, Selwood, & Murphy, 2002). Regarding IL-34, Mizuno et al. showed that IL-34-treated microglia attenuate the neurotoxic effects of Aβ in neuron-microglia cocultures by promoting microglial uptake and metabolism of Aβ (Mizuno et al., 2011). The neuroprotective role of IL-34 in this system seemed to be regulated by transforming growth factor beta-1 (TGFβ-1). The inhibition of TGFβ-1 receptor results in an increased microglial proliferation driven by IL-34 and the suppression of the observed neuroprotective effect of IL-34-treated microglia. These observations suggest that TGF-β produced by these cells acts as a negative regulator of microglial proliferation, improving the neuroprotective feature of microglia (Ma et al., 2012). In the APP/PS1 model of AD, the administration of IL-34 in the brain ameliorates the impairment of associative learning (Mizuno et al., 2011). These studies provided evidence that modulation of these cytokines may also be an approach to control the microglia population in the context of neurodegenerative diseases, as an alternative method to CSF1R modulation.

2.2 Role of PU.1 in the modulation of microglial proliferation and activation

The transcription factor PU.1 is also an important player in the development, proliferation and maintenance of microglia. PU.1, encoded by the gene Spi1, belongs to the ETS-family of transcription factors, and is a master regulator of myeloid and lymphoid development and function (Dakic et al., 2005; McKercher et al., 1996; Scott, Simon, Anastasi, & Singh, 1994). This transcription factor binds to a purine-rich DNA sequence (PU.1-box) located upstream of the promoter of its targets and activates the expression of a great number of downstream genes (Pham et al., 2013). PU.1 has been shown to be necessary for the correct development and functional maintenance of the microglial population since it is continuously expressed from erythromyeloid progenitors to adult microglia (Kierdorf et al., 2013; Smith et al., 2013). In fact, PU.1-deficient mice show a complete loss of microglia and other myeloid cell types such as macrophages and monocytes, indicating that PU.1 regulates key genes involved in the differentiation and the maturation of hematopoietic cells and also microglia (Beers et al., 2006; McKercher et al., 1996). Satoh et al. identified 5,264 Spi1 target protein-coding genes in the mouse microglial cell line BV2 by chromatin immunoprecipitation (ChIP)-seq analysis, including Spi1 itself,
the transcription factors Irf8 and Runx1. Aif1 (Iba1), Csf1r and its ligands Csf-1 and Il-34. Interestingly, two-thirds (63%) of the genes that define the microglial sensome are PU.1 targets, suggesting that PU.1 plays a pivotal role in the regulation of specific microglial functions (Satoh, Asahina, Kitano, & Kino, 2014) such as cell survival, phagocytosis, antigen presentation, and morphology. Recently, a GWAS study has identified a common haplotype, rs1057233 (G), located in a previously reported AD risk locus (CELF1), which displays a reduced expression of PU.1 in human myeloid cells associated to delayed age of onset of AD (Huang et al., 2017). In fact, the alteration of PU.1 levels in mouse and human microglial cells affected the expression of many AD risk genes (Huang et al., 2017) and their phagocytic activity (Huang et al., 2017; Rustenhoven et al., 2018; Smith et al., 2013). The activation of microglia through PU.1 has been shown to be critical for the progression of Alzheimer’s disease (Gjoneska et al., 2015), emphasising the role of microglia at the onset of the disease. Similarly, the activation of microglia through PU.1 is observed in response to mutant Huntingtin aggregates present in Huntington’s disease, hypoxic-ischaemic insults and traumatic injury-induced neurodegeneration (Crotti et al., 2014; Walton et al., 2000; Zhou, Liu, Sun, Cao, & Yang, 2019). Moreover, a recent study published by Litvinchuk et al. demonstrated that PU.1 and the transcription factors Irf8 and Runx1 were significantly upregulated in FACS-isolated microglia in the PS19 mouse model of tauopathy and AD (Litvinchuk et al., 2018). Together, these findings suggest that changes in the expression level of PU.1 may be a shared feature underlying several neurological disorders and highlight its modulation as a potential mechanism to control neuroinflammation. Studies using PU.1-/- mice have shown that complete loss of function of PU.1 results in stem cell failure (Antony-Debre et al., 2017), multiple hematopoietic abnormalities and, ultimately, developmental mortality (McKercher et al., 1996), highlighting the importance of achieving partial inhibition of PU.1 in order to understand its potential roles in disease. Newly described pharmacological PU.1 inhibitors have been recently developed (Munde et al., 2014; Stephens et al., 2016) and tested in murine and human acute myeloid leukemia (AML) (xeno) transplantation models, decreasing leukemia progression without affecting normal hematopoietic differentiation (Antony-Debre et al., 2017). These small molecules disrupt the interaction of PU.1 with its binding sites next to the promoters of target genes and lead to the downregulation of PU.1 transcriptional targets, holding a high potential as tool compounds for evaluating the role of PU.1 in neurodegenerative diseases.

3 CURRENT THERAPEUTIC STRATEGIES TARGETING MICROGLIA DYNAMICS AND POTENTIAL SIDE EFFECTS ON PERIPHERAL POPULATIONS

To date, drugs available for AD are restricted to relieve its symptoms, with no treatments able to stop or delay the progression of this disease. The cognitive problems in early-to-moderate AD are treated with Acetylcholinesterase inhibitors (Donepezil, Rivastigmine and Galantamine) which block the degradation of acetylcholine and enhance cholinergic neurotransmission, deficient in AD. Additionally, patients are treated with Memantine which protects against the glutamate excitotoxicity seen in neurodegenerative disorders such as AD. Currently, there are an estimated number of 132 agents in clinical trials for the treatment of AD. 30 in phase I of development, 74 in phase II and 28 in phase III. Among these agents, 96 (73%) are disease-modifying therapies; 38 (40%) and 17 (18%) of these have amyloid and tau as the primary target, respectively (Cummings, Lee, Ritter, Sabbagah, & Zhong, 2019). However, multiple failures to stop AD using similar strategies in the past have considerably increased the interest in other targets, such as those related to neuroinflammation, with 3 agents currently in phase II and 2 agents in phase III clinical trials (Cummings et al., 2019). In addition, recent genetic evidence clearly link microglia function to AD pathogenesis, placing the spotlight on microglia as a potential target to treat AD.
Several microglial genes identified as robustly-associated with risk of LOAD are now under investigation as potential targets for drug development, such as APOE, TREM2, CD33 and CR1, amongst others (for review see (Biber et al., 2019; Hemonnot, Hua, Ulmann, & Hirbec, 2019). Despite the importance of the above cited targets and their strong link with AD pathogenesis, here we focus on those related to the modulation of the dynamics of the microglial population. Microglial cells share many functions, genes and developmental lineage with other cells of the myeloid lineage across different organs (Hoeffel & Ginhoux, 2018), which are required for the proper functioning of the immune system (Figure 1). Because these gene expression signatures are conserved, it is extremely important to evaluate the impact of anti-neuroinflammatory agents on the broader immune system. The therapeutic benefit of influencing a given cellular function in a given pathology may result in the alteration of the natural balance of the broader immune system, with unknown consequences frequently not taken into consideration. Here, we review the potential side effects of manipulating immune-related pathways on other populations of immune cells, located in different organs of the systemic compartment.

Importantly, people with dementia usually have co-morbidities ranging from two to eight health conditions (Nelis et al., 2019). It is accepted that people with dementia have an average of 4 co-morbidities, compared to an average of 2 in people without dementia of similar age (Poblador-Plou et al., 2014). A recent study across various care settings has reported that 61% of the people with AD had three or more co-morbidities (Nelis et al., 2019). Over 90% of people with dementia have at least 1 co-morbidity, with some of these being often undiagnosed (Browne, Edwards, Rhodes, Brimicome, & Payne, 2017). Some of the main co-morbidities significantly associated with dementia are cardiac arrhythmia, hypertension, congestive cerebrovascular disease, diabetes and depression (Nelis et al., 2019). A common feature of several co-morbidities is a dysfunctional immune response. For example, obesity-related metabolic disorders, which are also risk factors for AD, are associated with alterations in the inflammatory status (Nguyen, Killcross, & Jenkins, 2014; Saltiel & Olefsky, 2017). Similarly, increasing evidence in recent years has demonstrated the important role of inflammation in the pathophysiology of diabetes (Tsalamandris et al., 2019), an age-related chronic disorder highly prevalent in AD patients (Nelis et al., 2019; Newcombe et al., 2018). Two of the most prevalent conditions associated to normal aging and dementia are cardiovascular disease and hypertension, both closely related to the above cited metabolic disorders (Lopez-Candales, Hernandez Burgos, Hernandez-Suarez, & Harris, 2017; Nelis et al., 2019). Similar to those, recent studies have supported the causal role of chronic inflammation in the development of these cardiovascular conditions (Lopez-Candales et al., 2017; Ruparelia, Chai, Fisher, & Choudhury, 2017). Also, incidence of systemic infections, such as urine tract infection (UTI) and gum disease, is increased in Alzheimer’s, further accelerating the cognitive deterioration (Dominy et al., 2019; Doraiswamy, Leon, Cummings, Marin, & Neumann, 2002). Psychiatric disorders with elevated prevalence in AD, such as depression, have also been related to peripheral and central chronic inflammation, which seem to drive changes in neurotransmitters leading to depressive symptoms (Felger, 2019). On the opposite spectrum, a growing body of evidence suggests an inverse link between the incidence rates of cancer and AD, even though both are age-related disorders with significant immune involvement (Majd, Power, & Majd, 2019). Taken together, this evidence highlights the fact that the co-existence of age-related comorbidities is a crucial aspect to consider in the development of immunomodulatory therapeutic strategies for treating AD, which in turn may compromise the responsiveness and immune control of these co-morbidities.

### 3.1. Inhibiting CSF1R in AD: target validation studies

The therapeutic potential of inhibiting CSF1R has been proposed for inflammatory diseases, autoimmune disorders, bone diseases and cancer (Burns & Wilks, 2011). Targeted inhibition of CSF1R
signalling has the potential to treat a wide variety of neurodegenerative diseases associated with chronic neuroinflammation such as AD, PD, Huntington’s disease, ALS, multiple sclerosis and psychiatric disorders. CSF1R can be blocked by at least two different approaches, i) using small-molecule inhibitors targeting the TK activity of the receptor or ii) antibodies that bind the receptor and block the interaction between CSF1R and CSF-1/IL-34. The first neutralising monoclonal antibody against CSF1R, AFS98, was produced by Sudo et al. (Sudo et al., 1995) and was shown to be effective in the control of CSF1-related functions in pathology. Some examples of its effectiveness are the reduction of macrophage accumulation in atherosclerotic lesions and diabetic nephropathy, the reduction of infiltrating macrophage proliferation in renal allografts and damaged skeletal muscle (for review see Hume & MacDonald, 2012), and the local inhibition of microglial proliferation in the prion disease model ME7 (Gomez-Nicola et al., 2013). In contrast with these results, prolonged treatment with a different monoclonal anti-CSF1R antibody, M279, selectively removed tissue macrophages, including macrophages inside the tumours, but had no protective effect in several models of inflammation (MacDonald et al., 2010). This antibody is incapable of crossing the blood brain barrier (BBB), depleting microglia in the retina but not affecting the brain (Hume & MacDonald, 2012). It has also been shown that after prolonged treatment with M279 bone density and trabecular volume are increased due to the ablation of osteoclasts, preventing the reduction in bone mass observed in female mice with age. This long-term effect on bone remodelling suggests that M279 could potentially be used as a treatment for osteoporosis (Sauter et al., 2014). Importantly, a side effect of CSF1R blocking antibodies is related to the role of CSF1R in the clearance of CSF-1 from the circulation by endocytosis (Hume & MacDonald, 2012). CSF1R blockade causes a massive increase in the concentration of circulating CSF-1, and rebound monocytopenia (Hume & MacDonald, 2012). However, this effect does not occur when the TK activity of the receptor is blocked by kinase inhibitors, since this activity is not required for the internalization of CSF-1 (Hume & MacDonald, 2012). One of the most important features of kinase inhibitors, compared to antibodies, is that small molecules are able to block autocrine actions of endogenous CSF-1, which is highly expressed in some mouse inflammatory macrophages and drives the expression of inflammatory cytokines (Hume & MacDonald, 2012). One of the most selective and best characterized of the available TK inhibitors probably is GW2580. GW2580 inhibits the growth of CSF1-dependent tumour cells (Conway et al., 2005) and the recruitment of macrophages into growing tumours (Priceman et al., 2010). It has also been shown to exhibit antitumor activity in AML by blocking paracrine production of hepatocyte growth factor and other cytokines signalling from support cells (Edwards et al., 2019). GW2580 has beneficial effects, by blocking microglial proliferation, in several experimental models of multiple sclerosis (Crespo et al., 2011), prion disease (Gomez-Nicola et al., 2013), AD (Olmos-Alonso et al., 2016), ALS (Martinez-Muriana et al., 2016), spinal cord injury (Gerber et al., 2018) and PD (Neal et al., 2020). Using the APP/PS1 model of AD-like pathology, we found diminished synaptic degeneration and improved behavioural and performance and learning after chronic inhibition of CSF1R with GW2580 (Olmos-Alonso et al., 2016). A different CSF1R inhibitor with significant in vivo data available is Ki20227. This inhibitor has been shown to reduce the number of macrophages and associated pathology in models of inflammatory arthritis (Ohno et al., 2008) and encephalomyelitis (Uemura et al., 2008). However, Ki20227 reduced the numbers of Ly6G+ granulocytes, an effect that generates concerns about its specificity. There are some other TK inhibitors that block CSF1R but also have affinity for other kinases, as the orally available JNJ-28312141 (Hume & MacDonald, 2012). This inhibitor has specificity against CSF1R but also the related receptor FLT3 and has been shown to reduce macrophage numbers and limit tumour growth in several models of transplanted tumours as well as in a FLT3-dependent subset of AML (Manthey et al., 2009). Despite J&J had JNJ-28312141 in phase II clinical trials for the treatment of rheumatoid arthritis (RA), this was discontinued and replaced by JNJ-40346527. This CSF1R inhibitor has been recently shown to repolarise microglia to a homeostatic phenotype and attenuate tau-induced neurodegeneration resulting in functional improvement in the
P301S mouse model of tauopathy (Mancuso et al., 2019). Currently, JNJ-40346527 is in phase II ongoing trials for AML (NCT03557970) and in phase Ib ongoing trials for AD (NCT04121208). Recently, a novel family of inhibitors developed by Plexxicon has been described to have a potent activity over CSF1R. PLX3397 (Pexidartinib) was shown to inhibit the survival of microglia and cause a fast depletion of the population in the healthy brain (Elmore et al., 2014). PLX3397 was shown to prevent neuronal degeneration, improving cognitive functions in the 5xFAD model of AD-like pathology (Sosna et al., 2018; Spangenberg et al., 2016). Similar results were obtained using the inhibitor PLX5622 in the 3xTg AD model (Dagher et al., 2015). However, PLX3397 also causes a potent inhibition of c-kit and PDGFRβ (Patwardhan et al., 2014), which may confound the observed effects on the microglial population. The inhibition of PDGFRβ and loss of PDGFβ signalling would affect the survival of NG2 pericytes, consequently damaging the BBB and influencing neurodegeneration (Montagne, Zhao, & Zlokovic, 2017). Despite the unknown side effects of these molecules in brain disease, PLX3397 is currently in phase II ongoing trials for several types of tumours such as sarcoma and glioblastoma (NCT01790503; NCT02584647). Another small molecule in development for AD is Masitinib, a pan-kinase TK inhibitor. AB Science SA is using Masitinib in phase III trials for patients with mild to moderate AD (NCT01872598), a wide variety of tumours such as gastrointestinal stromal tumours (NCT01694277), ALS (NCT02588677; NCT03127267) and multiple sclerosis (NCT01433497), based on the activity of the compound over CSF1R or c-kit, depending on the specific disease mechanism. In summary, many approaches have been designed to target the activity of CSF1R under neuroinflammatory conditions, and in coming years the field will collect valuable clinical information about their potential efficacy in AD.

3.2. Systemic impact of CSF1R inhibition: can selectivity and safety be improved?

According to the above cited studies, blocking the expansion of the microglial population results in a significant reduction of neuronal degeneration, leading to an improvement in the disease symptoms and survival. These results provide strong evidence of the potential application of CSF1R tyrosine kinase inhibitors as a promising approach to tackle microglial proliferation in neurodegeneration. However, although many CSF1R inhibitors are progressing to clinical trials, little is known about the impact of these approaches on the innate immune system. CSF1R is expressed in many cell types of the myeloid lineage, including tissue-resident macrophages, dendritic cells and their precursors (Chitu & Stanley, 2017). Therefore, the inhibition of CSF1R would not only affect microglia, but also other tissue-resident myeloid populations, possibly causing an immunosuppressive effect.

A potential approach to block this pathway more selectively is by modulating the binding of its ligands, CSF-1 and/or IL-34, to increase tissue specificity and reduce side effects. This approach is based on the differential tissue-selectivity and functions of CSF-1 vs. IL-34, reported in the literature and discussed previously. The blockade of both ligands can be achieved by the use of specific antibodies directed against these cytokines, with beneficial effects in murine models of arthritis, colitis and ileitis (W. Lin et al., 2019). However, blockade of both ligands, separately or in combination, leads to altered macrophage homeostasis in healthy mice, reducing the numbers of macrophages in the intestine, liver, kidney, skin, bone marrow and microglia in the brain (Easley-Neal et al., 2019; W. Lin et al., 2019). In contrast to these observations, a recent study from our group shows that monocyte and macrophage populations in peripheral tissues were not affected after the selective blockade of IL-34 in healthy mice, except for the skin-resident Langerhans cells (Obst et al., 2020). However, the number of monocytes and macrophages were significantly decreased after blockade of CSF1R, in accordance with the wider expression of the receptor. Despite the microglial population was not affected after systemic administration of anti-IL-34 antibodies, due to their low brain penetrance, we observed a local
reduction of microglia proliferation after the intracerebral injection of anti-IL-34 antibodies in mice infected with prion disease, showing that IL-34 is a key driver of microglial proliferation in the context of neurodegenerative disease (Obst et al., 2020). Our results support that modulation of the microglial response via IL-34 blockade could be a potential and more selective therapeutic approach in neurodegenerative diseases (Obst et al., 2020). A similar therapeutic approach modulating the granulocyte-macroage colony stimulating factor (GM-CSF) instead of targeting its receptor is currently in a phase II clinical trial for AD (NCT01409915), which has been recently completed although no results have been published yet. Testing of this recombinant human factor, named as Saragrostim, for AD is based on published results regarding GM-CSF role in AD mouse models, in which GM-CSF seems to reduce brain amyloidosis and reverse cognitive impairment by increasing microglial density and their activation state (Boyd et al., 2010; Kiyota et al., 2018). However, some studies have reported an increased expression of GM-CSF in AD patients (Tarkowski, Wallin, Regland, Blennow, & Tarkowski, 2001) and a beneficial role of blocking this factor using an anti-GM-CSF antibody in a mouse model of AD (Manczak et al., 2009). Nevertheless, the potential side effects of this approach on other myeloid populations are unknown, supporting the idea that more studies are necessary to understand the effects of modulating these molecules in neurodegenerative diseases and their potential on-target effects on tissue resident macrophages.

The functions of CSF-1, IL-34 and CSF1R in monocyte-macrophage differentiation have been demonstrated through the study of specific genetic mutations in mice, rats and humans (Chitu & Stanley, 2017; Hume & MacDonald, 2012). Mice and rats with Csfr-1 loss-of-function mutations have deficiencies in many tissue macrophage populations and are severely osteopetrotic, due to the lack of osteoclasts (Dai et al., 2002). Pleiotropic effects including severe postnatal growth retardation, neurological and reproductive deficiencies, highlight the important trophic roles of CSF1-dependent macrophages (Wynn, Chawla, & Pollard, 2013). By contrary, IL-34 mutation is less severe, only depleting microglia and Langerhans cells, consistent with its restricted regional expression (Wang et al., 2012). CSF1R knockout mice display a severe phenotype characterised by limited survival after the weaning phase (Chitu, Gokhan, Nandi, Mehler, & Stanley, 2016). Interestingly, a recent study has shown that genomic deletion of FIRE, a highly conserved Csf1r enhancer, ablates specifically microglia and resident macrophages in some tissues such as the skin, kidney, heart and peritoneum (Rojo et al., 2019). They demonstrate that Csf1r ΔFIRE/ΔFIRE mice are healthy and fertile, not showing the severe postnatal growth retardation and developmental abnormalities observed in Csf1r−/− rodents (Rojo et al., 2019). In humans, the hypomorphic mutation in CSF1R causes hereditary diffuse leukencephalopathy with spheroids, a disease originated from the loss of myelin and the destruction of axons (Wynn et al., 2013). Homozygous mutations in CSF1R in human leads to premature death, linked to severe brain abnormalities including hydrocephaly, hypomyelination and abnormal bone growth (Oosterhof et al., 2019). Given the central role of macrophages in fighting infection (Figure 1), the long-term blockade of the CSF1R/CSF-1/IL-34 axes could compromise the response to infection. In fact, mice infected with Listeria monocytogenes and treated with antibodies against CSF-1/IL-34 were more susceptible to the bacterial infection, showing that these approaches might be immunosuppressive in the rodent Listeria model (W. Lin et al., 2019). Similar results were obtained in a model of viral encephalitis, where the inactivation of CSF1R using a tyrosine kinase inhibitor reduced circulating antigen-presenting cells in the blood leading to a higher susceptibility to lethal West Nile virus infection (Funk & Klein, 2019). This study shows the importance of CSF1R in myeloid cell responses that involve the restriction of viral replication, and the local restimulation of recruited antiviral T cells within the CNS (Funk & Klein, 2019). On the other hand, a different CSF1R TK inhibitor showed a good safety and tolerability profile after 3 months treatment in patients with RA, causing only an alteration in Kupffer cell function (Figure 1) (Genovese et al., 2015). Kupffer cells may have a role in clearing several serum enzymes, including alanine aminotransferase and aspartate aminotransferase,
which are often used as indicators of hepatic injury during medical tests and clinical trials (W. Lin et al., 2019; Radi et al., 2011). The reduction in the population of Kupffer cells after treatment with anti-CSF-1/IL-34 antibodies correlated with an increase of these enzymes in the serum of rodents and monkeys, although no histopathological evidence of liver injury was observed (W. Lin et al., 2019; Radi et al., 2011). Importantly, the detection of high liver enzyme activity, unrelated to hepatocellular injury, may compromise clinical monitoring of liver injury, an aspect to take into consideration with therapeutics that target macrophages (W. Lin et al., 2019). Bone formation and resorption is also a process influenced by CSF1-CSF1R signalling (Figure 1). CSF-1 is produced in the bone marrow by osteoblasts, binding to CSF1R located on the surface of osteoclast precursors, giving rise to the formation of osteoclasts (El-Gamal et al., 2018). Mice lacking CSF-1 are unable to generate osteoclasts, leading to low bone density and osteoporosis (El-Gamal et al., 2018). However, CSF1R inhibition would likely lead to increased bone density and abnormal bone growth due to a decrease in osteoclast numbers. This may result in the development of Paget’s disease, which is characterised by enlarged and misshapen bones. Another effect of CSF-1 deficiency in the macrophage-deficient Csf1<sup>op/op</sup>/Csf1<sup>op/op</sup> model is an insulin mass deficit due to the reduction of pancreatic β cell proliferation and abnormal islet morphology in the pancreas (Banaei-Bouchareb et al., 2004). In fact, the addition of CSF-1 to embryonic pancreas explants caused a higher differentiation of β cell and increased production of insulin (Geutksens, Otonkoski, Pulkkinen, Drexhage, & Leenen, 2005). However, macrophage ablation in the pancreas and adipose tissue after long-term anti-CSF1R treatment (Figure 1), had no effect on average size or distribution of β cells within islets of Langerhans, detected by immunostaining for insulin (Sauter et al., 2014). Despite the decrease in tissue resident macrophages in many organs after the treatment with an anti-CSF1R antibody, Sauter et al. did not observe any overt pathology in hematoxylin and eosin sections of different organs (Sauter et al., 2014). In summary, CSF1R/CSF-1/IL-34 blocking strategies have different effects on tissue-resident macrophages and other cell types of the systemic compartment, leading to a dysregulation of the tissue homeostatic functions (Figure 1). Likewise, any therapeutic approach directed against potential microglial targets, e.g. TREM2, inflammasome, among others, is expected to have a comparable impact on peripheral immune cell populations and organ function. Therefore, we need further investigation of the potential side effects of manipulating immune-related pathways to modulate the microglial population during neuroinflammation, in order to design and develop highly specific therapeutic agents.

4 CONCLUSION

Over recent years the field of study of the contribution of neuroinflammation to AD has undergone a revolution. The number and quality of preclinical studies has increased, leading to some very promising early clinical studies, using agents directed against neuroinflammatory targets. In coming years this field will finally start to collect some critical clinical data, which will allow, once and for all, to address the hypothesis that neuroinflammation is a driver of neurodegeneration in AD. These early promising studies should not distract the field from trying to find better, more refined, approaches, to overcome the anticipated significant impact over the broader immune system. In the meantime, it is crucial to start to understand the impact of targeting key neuroinflammatory pathways on the function of other tissue-resident macrophages, and the key organ functions they are responsible for. If any of the postulated anti-neuroinflammation agents succeeded to progress to longer trials or eventually to enter market, it is anticipated that the AD target population would be exposed for very prolonged periods of time to agents influencing their immune balance. Considering AD patients are often multimorbid, this would have unknown consequences over their responsiveness to infection or the control of their immune-related co-morbidities.
5  AUTHOR CONTRIBUTIONS

MM-E and DG-N designed the structure and content of the manuscript. MM-E wrote the manuscript and prepared the figures. MM-E and DG-N drafted the manuscript.

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7  REFERENCES

Akiyama, H., Barger, S., Barnum, S., Bradt, B., Bauer, J., Cole, G. M., . . . Wyss-Coray, T. (2000). Inflammation and Alzheimer's disease. Neurobiol Aging, 21(3), 383-421. doi:10.1016/s0197-4580(00)00124-x

Akiyama, H., Nishimura, T., Kondo, H., Ikeda, K., Hayashi, Y., & McGeer, P. L. (1994). Expression of the receptor for macrophage colony stimulating factor by brain microglia and its upregulation in brains of patients with Alzheimer's disease and amyotrophic lateral sclerosis. Brain Res, 639(1), 171-174. doi:10.1016/0006-8993(94)91779-5

Antony-Debre, I., Paul, A., Leite, J., Mitchell, K., Kim, H. M., Carvajal, L. A., . . . Steidl, U. (2017). Pharmacological inhibition of the transcription factor PU.1 in leukemia. Journal of Clinical Investigation, 127(12), 4297-4313. doi:10.1172/JCI92504

Atri, A. (2019). The Alzheimer's Disease Clinical Spectrum: Diagnosis and Management. Med Clin North Am, 103(2), 263-293. doi:10.1016/j.mcna.2018.10.009

Banaei-Bouchareb, L., Gouon-Evans, V., Samara-Boustani, D., Castellotti, M. C., Czernichow, P., Pollard, J. W., & Polak, M. (2004). Insulin cell mass is altered in Csf1op/Csf1op macrophage-deficient mice. J Leukoc Biol, 76(2), 359-367. doi:10.1189/jlb.1103591

Beers, D. R., Henkel, J. S., Xiao, Q., Zhao, W., Wang, J., Yen, A. A., . . . Appel, S. H. (2006). Wild-type microglia extend survival in PU.1 knockout mice with familial amyotrophic lateral sclerosis. Proc Natl Acad Sci U S A, 103(43), 16021-16026. doi:10.1073/pnas.0607423103

Bennett, R. E., Bryant, A., Hu, M., Robbins, A. B., Hopp, S. C., & Hyman, B. T. (2018). Partial reduction of microglia does not affect tau pathology in aged mice. J Neuroinflammation, 15(1), 311. doi:10.1186/s12974-018-1348-5

Biber, K., Bhattacharya, A., Campbell, B. M., Piro, J. R., Rohe, M., Staal, R. G. W., . . . Moller, T. (2019). Microglial Drug Targets in AD: Opportunities and Challenges in Drug Discovery and Development. Front Pharmacol, 10, 840. doi:10.3389/fphar.2019.00840

Boutens, L., & Stienstra, R. (2016). Adipose tissue macrophages: going off track during obesity. Diabetologia, 59(5), 879-894. doi:10.1007/s00125-016-3904-9

Boyd, T. D., Bennett, S. P., Mori, T., Governatori, N., Runfeldt, M., Norden, M., . . . Potter, H. (2010). GM-CSF upregulated in rheumatoid arthritis reverses cognitive impairment and amyloidosis in Alzheimer mice. J Alzheimers Dis, 21(2), 507-518. doi:10.3233/JAD-2010-091471
Browne, J., Edwards, D. A., Rhodes, K. M., Brimicombe, D. J., & Payne, R. A. (2017). Association of comorbidity and health service usage among patients with dementia in the UK: a population-based study. *BMJ Open, 7*(3), e012546. doi:10.1136/bmjopen-2016-012546

Burns, C. J., & Wilks, A. F. (2011). c-FMS inhibitors: a patent review. *Expert Opin Ther Pat, 21*(2), 147-165. doi:10.1517/13543776.2011.545348

Cai, Z., Hussain, M. D., & Yan, L. J. (2014). Microglia, neuroinflammation, and beta-amyloid protein in Alzheimer's disease. *Int J Neurosci, 124*(5), 307-321. doi:10.3109/00207454.2013.833510

Calsolaro, V., & Edison, P. (2016). Neuroinflammation in Alzheimer's disease: Current evidence and future directions. *Alzheimers Dement, 12*(6), 719-732. doi:10.1016/j.jalz.2016.02.010

Carlin, L. M., Stamatiades, E. G., Auffray, C., Hanna, R. N., Glover, L., Vizcay-Barrena, G., . . . Geissmann, F. (2013). Nr4a1-dependent Ly6C(low) monocytes monitor endothelial cells and orchestrate their disposal. *Cell, 153*(2), 362-375. doi:10.1016/j.cell.2013.03.010

Chitu, V., & Stanley, E. R. (2017). Regulation of Embryonic and Postnatal Development by the CSF-1 Receptor and its Ligands in the Nervous System. *Trends Neurosci, 39*(6), 378-393. doi:10.1016/j.tins.2016.03.005

Chorus, L., & Geissmann, F. (2010). Development and homeostasis of 'resident' myeloid cells: the case of the Langerhans cell. *Trends Immunol, 31*(12), 438-445. doi:10.1016/j.it.2010.09.003

Chow, A., Huggins, M., Ahmed, J., Hashimoto, D., Lucas, D., Kunisaki, Y., . . . Frenette, P. S. (2013). CD169(+) macrophages provide a niche promoting erythropoiesis under homeostasis and stress. *Nat Med, 19*(4), 429-436. doi:10.1038/nm.3057

Chow, A., Lucas, D., Hidalgo, A., Mendez-Ferrer, S., Hashimoto, D., Scheiermann, C., . . . Frenette, P. S. (2011). Bone marrow CD169+ macrophages promote the retention of hematopoietic stem and progenitor cells in the mesenchymal stem cell niche. *J Exp Med, 208*(2), 261-271. doi:10.1084/jem.20101688

Colonna, M., & Butovsky, O. (2017). Microglia Function in the Central Nervous System During Health and Neurodegeneration. *Annu Rev Immunol, 35*, 441-468. doi:10.1146/annurev-immunol-051116-052358

Conway, J. G., McDonald, B., Parham, J., Keith, B., Rusnak, D. W., Shaw, E., . . . Hutchins, J. T. (2005). Inhibition of colony-stimulating-factor-1 signaling in vivo with the orally bioavailable cFMS kinase inhibitor GW2580. *Proc Natl Acad Sci U S A, 102*(44), 16078-16083. doi:10.1073/pnas.0502000102

Corder, E. H., Saunders, A. M., Strittmatter, W. J., Schmechel, D. E., Gaskell, P. C., Small, G. W., . . . Pericak-Vance, M. A. (1993). Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science, 261*(5123), 921-923. doi:10.1126/science.8346443

Crespo, O., Kang, S. C., Daneman, R., Lindstrom, T. M., Ho, P. P., Sobel, R. A., . . . Robinson, W. H. (2011). Tyrosine kinase inhibitors ameliorate autoimmune encephalomyelitis in a mouse model of multiple sclerosis. *J Clin Immunol, 31*(6), 1010-1020. doi:10.1007/s10875-011-9579-6

Crotti, A., Benner, C., Kerman, B. E., Gosselin, D., Lagier-Tourenne, C., Zuccato, C., . . . Glass, C. K. (2014). Mutant Huntingtin promotes autonomous microglia activation via myeloid lineage-determining factors. *Nature Neuroscience, 17*(4), 513-521. doi:10.1038/nn.3668
Targeting Neuroinflammation in Alzheimer’s disease: are we ready yet?

Cummings, J., Lee, G., Ritter, A., Sabbagh, M., & Zhong, K. (2019). Alzheimer’s disease drug development pipeline: 2019. Alzheimers Dement (N Y), 5, 272-293. doi:10.1016/j.trci.2019.05.008

Dagher, N. N., Najafi, A. R., Kayala, K. M., Elmore, M. R., White, T. E., Medeiros, R., . . . Green, K. N. (2015). Colony-stimulating factor 1 receptor inhibition prevents microglial plaque association and improves cognition in 3xTg-AD mice. J Neuroinflammation, 12, 139. doi:10.1186/s12974-015-0366-9

Dai, X. M., Ryan, G. R., Hapel, A. J., Dominguez, M. G., Russell, R. G., Kapp, S., . . . Stanley, E. R. (2002). Targeted disruption of the mouse colony-stimulating factor 1 receptor gene results in osteopetrosis, mononuclear phagocyte deficiency, increased primitive progenitor cell frequencies, and reproductive defects. Blood, 99(1), 111-120. doi:10.1182/blood.v99.1.111

Dakic, A., Metcalf, D., Di Rago, L., Mifsud, S., Wu, L., & Nutt, S. L. (2005). PU.1 regulates the commitment of adult hematopoietic progenitors and restricts granulopoiesis. J Exp Med, 201(9), 1487-1502. doi:10.1084/jem.20050075

Davies, L. C., Jenkins, S. J., Allen, J. E., & Taylor, P. R. (2013). Tissue-resident macrophages. Nat Immunol, 14(10), 986-995. doi:10.1038/ni.2705

den Haan, J. M., & Kraal, G. (2012). Innate immune functions of macrophage subpopulations in the spleen. J Innate Immun, 4(5-6), 437-445. doi:10.1159/000335216

Dickson, D. W., Lee, S. C., Mattiace, L. A., Yen, S. H., & Brosnan, C. (1993). Microglia and cytokines in neurological disease, with special reference to AIDS and Alzheimer's disease. Glia, 7(1), 75-83. doi:10.1002/glia.440070113

Dominy, S. S., Lynch, C., Ermini, F., Benedyk, M., Marczyk, A., Konrady, A., . . . Potempa, J. (2019). Porphyromonas gingivalis in Alzheimer's disease brains: Evidence for disease causation and treatment with small-molecule inhibitors. Sci Adv, 5(1), eaau3333. doi:10.1126/sciadv.aau3333

Doraiswamy, P. M., Leon, J., Cummings, J. L., Marin, D., & Neumann, P. J. (2002). Prevalence and impact of medical comorbidity in Alzheimer's disease. J Gerontol A Biol Sci Med Sci, 57(3), M173-177. doi:10.1093/gerona/57.3.m173

Dranoff, G., Crawford, A. D., Sadelain, M., Ream, B., Rashid, A., Bronson, R. T., . . . et al. (1994). Involvement of granulocyte-macrophage colony-stimulating factor in pulmonary homeostasis. Science, 264(5159), 713-716. doi:10.1126/science.8171324

Easley-Neal, C., Foreman, O., Sharma, N., Zarrin, A. A., & Weimer, R. M. (2019). CSF1R Ligands IL-34 and CSF1 Are Differentially Required for Microglia Development and Maintenance in White and Gray Matter Brain Regions. Frontiers in Immunology, 10, 2199. doi:10.3389/fimmu.2019.02199

Edison, P., Archer, H. A., Gerhard, A., Hinz, R., Pavese, N., Turkheimer, F. E., . . . Brooks, D. J. (2008). Microglia, amyloid, and cognition in Alzheimer's disease: An [11C](R)PK11195-PET and [11C]PIB-PET study. Neurobiol Dis, 32(3), 412-419. doi:10.1016/j.nbd.2008.08.001

Edwards, D. K. t., Watanabe-Smith, K., Rofelty, A., Damnersawad, A., Laderas, T., Lamble, A., . . . Tyner, J. W. (2019). CSF1R inhibitors exhibit antitumor activity in acute myeloid leukemia by blocking paracrine signals from support cells. Blood, 133(6), 588-599. doi:10.1182/blood-2018-03-838946
Efthymiou, A. G., & Goate, A. M. (2017). Late onset Alzheimer's disease genetics implicates microglial pathways in disease risk. *Molecular Neurodegeneration, 12*(1), 43. doi:10.1186/s13024-017-0184-x

El-Gamal, M. I., Al-Ameen, S. K., Al-Koumi, D. M., Hamad, M. G., Jalal, N. A., & Oh, C. H. (2018). Recent Advances of Colony-Stimulating Factor-1 Receptor (CSF-1R) Kinase and Its Inhibitors. *J Med Chem, 61*(13), 5450-5466. doi:10.1021/acs.jmedchem.7b00873

Elmore, M. R., Najafi, A. R., Koike, M. A., Dagher, N. N., Spangenberg, E. E., Rice, R. A., . . . Green, K. N. (2014). Colony-stimulating factor 1 receptor signaling is necessary for microglia viability, unmasking a microglia progenitor cell in the adult brain. *Neuron, 82*(2), 380-397. doi:10.1016/j.neuron.2014.02.040

Felger, J. C. (2019). Role of Inflammation in Depression and Treatment Implications. *Handb Exp Pharmacol, 250*, 255-286. doi:10.1007/164_2018_166

Fernandez-Botran, R., Ahmed, Z., Crespo, F. A., Gatenbee, C., Gonzalez, J., Dickson, D. W., & Litvan, I. (2011). Cytokine expression and microglial activation in progressive supranuclear palsy. *Parkinsonism Relat Disord, 17*(9), 683-688. doi:10.1016/j.parkreldis.2011.06.007

Funk, K. E., & Klein, R. S. (2019). CSF1R antagonism limits local restimulation of antiviral CD8(+) T cells during viral encephalitis. *J Neuroinflammation, 16*(1), 22. doi:10.1186/s12974-019-1397-4

Ganz, T. (2012). Macrophages and systemic iron homeostasis. *J Innate Immun, 4*(5-6), 446-453. doi:10.1159/000336423

Genovese, M. C., Hsia, E., Belkowski, S. M., Chien, C., Masterson, T., Thurmond, R. L., . . . Greenspan, A. (2015). Results from a Phase IIA Parallel Group Study of JNJ-40346527, an Oral CSF-1R Inhibitor, in Patients with Active Rheumatoid Arthritis despite Disease-modifying Antirheumatic Drug Therapy. *J Rheumatol, 42*(10), 1752-1760. doi:10.3899/jrheum.141580

Gerber, Y. N., Saint-Martin, G. P., Bringuier, C. M., Bartolami, S., Goze-Bac, C., Noristani, H. N., & Perrin, F. E. (2018). CSF1R Inhibition Reduces Microglia Proliferation, Promotes Tissue Preservation and Improves Motor Recovery After Spinal Cord Injury. *Frontiers in Cellular Neuroscience, 12*, 368. doi:10.3389/fncel.2018.00368

Geutskens, S. B., Otonkoski, T., Pulkkinen, M. A., Drexhage, H. A., & Leenen, P. J. (2005). Macrophages in the murine pancreas and their involvement in fetal endocrine development in vitro. *J Leukoc Biol, 78*(4), 845-852. doi:10.1189/jlb.1004624

Gjoneska, E., Pfenning, A. R., Mathys, H., Quon, G., Kundaje, A., Tsai, L. H., & Kellis, M. (2015). Conserved epigenomic signals in mice and humans reveal immune basis of Alzheimer's disease. *Nature, 518*(7539), 365-369. doi:10.1038/nature14252

Gomez-Nicola, D., Fransen, N. L., Suzzi, S., & Perry, V. H. (2013). Regulation of microglial proliferation during chronic neurodegeneration. *J Neurosci, 33*(6), 2481-2493. doi:10.1523/JNEUROSCI.4440-12.2013

Gomez-Nicola, D., & Perry, V. H. (2015). Microglial Dynamics and Role in the Healthy and Diseased Brain: A Paradigm of Functional Plasticity. *Neuroscientist, 21*(2), 169-184. doi:10.1177/1073858414530512
Targeting Neuroinflammation in Alzheimer’s disease: are we ready yet?

Gordon, S., Pluddemann, A., & Martinez Estrada, F. (2014). Macrophage heterogeneity in tissues: phenotypic diversity and functions. *Immunological Reviews, 262*(1), 36-55. doi:10.1111/imr.12223

Greter, M., Lelios, I., Pelczar, P., Hoeffel, G., Price, J., Leboeuf, M., . . . Becher, B. (2012). Stromal-derived interleukin-34 controls the development and maintenance of langerhans cells and the maintenance of microglia. *Immuinity, 37*(6), 1050-1060. doi:10.1016/j.immuini.2012.11.001

Guerreiro, R., Wojtas, A., Bras, J., Carrasquillo, M., Rogaeva, E., Majounie, E., . . . Alzheimer Genetic Analysis, G. (2013). TREM2 variants in Alzheimer's disease. *N Engl J Med, 368*(2), 117-127. doi:10.1056/NEJMoa1211851

Hansen, D. V., Hanson, J. E., & Sheng, M. (2018). Microglia in Alzheimer's disease. *J Cell Biol, 217*(2), 459-472. doi:10.1083/jcb.201709069

Hardy, J. A., & Higgins, G. A. (1992). Alzheimer's disease: the amyloid cascade hypothesis. *Science, 256*(5054), 184-185. doi:10.1126/science.1566067

Hemonnot, A. L., Hua, J., Ulmann, L., & Hirbec, H. (2019). Microglia in Alzheimer Disease: Well-Known Targets and New Opportunities. *Front Aging Neurosci, 11*, 233. doi:10.3389/fnagi.2019.00233

Heneka, M. T., Golenbock, D. T., & Latz, E. (2015). Innate immunity in Alzheimer's disease. *Nat Immunol, 16*(3), 229-236. doi:10.1038/nii.3102

Hoozemans, J. J., Veerhuis, R., Rozemuller, J. M., & Eikelenboom, P. (2011). Soothing the inflamed brain: effect of non-steroidal anti-inflammatory drugs on Alzheimer's disease pathology. *CNS Neurol Disord Drug Targets, 10*(1), 57-67. doi:10.2174/187152711794488665

Huang, K. L., Marcora, E., Pimenova, A. A., Di Narzo, A. F., Kapoor, M., Jin, S. C., . . . Goate, A. M. (2017). A common haplotype lowers PU.1 expression in myeloid cells and delays onset of Alzheimer's disease. *Nature Neuroscience, 20*(8), 1052-1061. doi:10.1038/nn.4587

Hume, D. A., & MacDonald, K. P. (2012). Therapeutic applications of macrophage colony-stimulating factor-1 (CSF-1) and antagonists of CSF-1 receptor (CSF-1R) signaling. *Blood, 119*(8), 1810-1820. doi:10.1182/blood-2011-09-379214

International, A. s. D. (2019). World Alzheimer Report 2019: Attitudes to dementia. *Alzheimer's Disease International.*

Jack, C. R., Jr., Bennett, D. A., Blennow, K., Carrillo, M. C., Dunn, B., Haeberlein, S. B., . . . Contributors. (2018). NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimers Dement, 14*(4), 535-562. doi:10.1016/j.jalz.2018.02.018

Jones, C. V., & Ricardo, S. D. (2013). Macrophages and CSF-1: implications for development and beyond. *Organogenesis, 9*(4), 249-260. doi:10.4161/org.25676

Jonsson, T., Stefansson, H., Steinberg, S., Jonsdottir, I., Jonsson, P. V., Snaedal, J., . . . Stefansson, K. (2013). Variant of TREM2 associated with the risk of Alzheimer's disease. *N Engl J Med, 368*(2), 107-116. doi:10.1056/NEJMo1211103

Karran, E., Mercken, M., & De Strooper, B. (2011). The amyloid cascade hypothesis for Alzheimer's disease: an appraisal for the development of therapeutics. *Nat Rev Drug Discov, 10*(9), 698-712. doi:10.1038/nrd3505
Targeting Neuroinflammation in Alzheimer’s disease: are we ready yet?

Kierdorf, K., Erny, D., Goldmann, T., Sander, V., Schulz, C., Perdigueru, E. G., . . . Prinz, M. (2013). Microglia emerge from erythromyeloid precursors via Pu.1- and Irf8-dependent pathways. *Nature Neuroscience, 16*(3), 273-280. doi:10.1038/nn.3318

Kierdorf, K., Prinz, M., Geissmann, F., & Gomez Perdiguero, E. (2015). Development and function of tissue resident macrophages in mice. *Semin Immunol, 27*(6), 369-378. doi:10.1016/j.smim.2016.03.017

Kiyota, T., Machhi, J., Lu, Y., Dyavarshetty, B., Nemati, M., Yokoyama, I., . . . Gendelman, H. E. (2018). Granulocyte-macrophage colony-stimulating factor neuroprotective activities in Alzheimer's disease mice. *J Neuroimmunol, 319*, 80-92. doi:10.1016/j.jneuroim.2018.03.009

Kurotaki, D., Uede, T., & Tamura, T. (2015). Functions and development of red pulp macrophages. *Microbiol Immunol, 59*(2), 55-62. doi:10.1111/1348-0421.12228

Li, Q., & Barres, B. A. (2018). Microglia and macrophages in brain homeostasis and disease. *Nat Rev Immunol, 18*(4), 225-242. doi:10.1038/nri.2017.125

Lin, H., Lee, E., Hestir, K., Leo, C., Huang, M., Bosch, E., . . . Williams, L. T. (2008). Discovery of a cytokine and its receptor by functional screening of the extracellular proteome. *Science, 320*(5877), 807-811. doi:10.1126/science.1154370

Lin, W., Xu, D., Austin, C. D., Caplazi, P., Senger, K., Sun, Y., . . . Zarrin, A. A. (2019). Function of CSF1 and IL34 in Macrophage Homeostasis, Inflammation, and Cancer. *Frontiers in Immunology, 10*, 2019. doi:10.3389/fimmu.2019.02019

Litvinchuk, A., Wan, Y. W., Swartzlander, D. B., Chen, F., Cole, A., Propson, N. E., . . . Zheng, H. (2018). Complement C3aR Inactivation Attenuates Tau Pathology and Reverses an Immune Network Deregulated in Tauopathy Models and Alzheimer's Disease. *Neuron, 100*(6), 1337-1353 e1335. doi:10.1016/j.neuron.2018.10.031

Lopez-Candales, A., Hernandez Burgos, P. M., Hernandez-Suarez, D. F., & Harris, D. (2017). Linking Chronic Inflammation with Cardiovascular Disease: From Normal Aging to the Metabolic Syndrome. *J Nat Sci, 3*(4). Retrieved from [https://www.ncbi.nlm.nih.gov/pubmed/28670620](https://www.ncbi.nlm.nih.gov/pubmed/28670620)

Lyman, M., Lloyd, D. G., Ji, X., Vizcaychipi, M. P., & Ma, D. (2014). Neuroinflammation: the role and consequences. *Neurosci Res, 79*, 1-12. doi:10.1016/j.neures.2013.10.004

Ma, D., Doi, Y., Jin, S., Li, E., Sonobe, Y., Takeuchi, H., . . . Suzumura, A. (2012). TGF-beta induced by interleukin-34-stimulated microglia regulates microglial proliferation and attenuates oligomeric amyloid beta neurotoxicity. *Neurosci Lett, 529*(1), 86-91. doi:10.1016/j.neulet.2012.08.071

MacDonald, K. P., Palmer, J. S., Cronau, S., Seppanen, E., Olver, S., Raffelt, N. C., . . . Hume, D. A. (2010). An antibody against the colony-stimulating factor 1 receptor depletes the resident subset of monocytes and tissue- and tumor-associated macrophages but does not inhibit inflammation. *Blood, 116*(19), 3955-3963. doi:10.1182/blood-2010-02-266296

Majd, S., Power, J., & Majd, Z. (2019). Alzheimer's Disease and Cancer: Whe...
suppresses microglial activity in a transgenic mouse model of Alzheimer's disease. *Hum Mol Genet, 18*(20), 3876-3893. doi:10.1093/hmg/ddp331

Manthey, C. L., Johnson, D. L., Illig, C. R., Tuman, R. W., Zhou, Z., Baker, J. F., . . . Molloy, C. J. (2009). JNJ-28312141, a novel orally active colony-stimulating factor-1 receptor/FMS-related receptor tyrosine kinase-3 receptor tyrosine kinase inhibitor with potential utility in solid tumors, bone metastases, and acute myeloid leukemia. *Mol Cancer Ther, 8*(11), 3151-3161. doi:10.1158/1535-7163.MCT-09-0255

Martinez-Muriana, A., Mancuso, R., Francos-Quijorna, I., Olmos-Alonso, A., Osta, R., Perry, V. H., . . . Lopez-Vales, R. (2016). CSF1R blockade slows the progression of amyotrophic lateral sclerosis by reducing microgliosis and invasion of macrophages into peripheral nerves. *Sci Rep, 6*, 25663. doi:10.1038/srep25663

Maus, U. A., Koay, M. A., Delbeck, T., Mack, M., Ermert, M., Ermert, L., . . . Lohmeyer, J. (2002). Role of resident alveolar macrophages in leukocyte traffic into the alveolar air space of intact mice. *Am J Physiol Lung Cell Mol Physiol, 282*(6), L1245-L1252. doi:10.1152/ajplung.00453.2001

McKercher, S. R., Torbett, B. E., Anderson, K. L., Henkel, G. W., Vestal, D. J., Baribault, H., . . . Maki, R. A. (1996). Targeted disruption of the PU.1 gene results in multiple hematopoietic abnormalities. *EMBO J, 15*(20), 5647-5658. Retrieved from https://www.ncbi.nlm.nih.gov/pubmed/8896458

Mendez, M. F. (2012). Early-onset Alzheimer's disease: nonamnestic subtypes and type 2 AD. *Arch Med Res, 43*(8), 677-685. doi:10.1016/j.arcmed.2012.11.009

Mizuno, T., Doi, Y., Mizoguchi, H., Jin, S., Noda, M., Sonobe, Y., . . . Suzumura, A. (2011). Interleukin-34 selectively enhances the neuroprotective effects of microglia to attenuate oligomeric amyloid-beta neurotoxicity. *Am J Pathol, 179*(4), 2016-2027. doi:10.1016/j.ajpath.2011.06.011

Montagne, A., Zhao, Z., & Zlokovic, B. V. (2017). Alzheimer's disease: A matter of blood-brain barrier dysfunction? *J Exp Med, 214*(11), 3151-3169. doi:10.1084/jem.20171406

Munde, M., Wang, S., Kumar, A., Stephens, C. E., Farahat, A. A., Boykin, D. W., . . . Poon, G. M. (2014). Structure-dependent inhibition of the ETS-family transcription factor PU.1 by novel heterocyclic diamidines. *Nucleic Acids Res, 42*(2), 1379-1390. doi:10.1093/nar/gkt955

Murphy, G. M., Jr., Zhao, F., Yang, L., & Cordell, B. (2000). Expression of macrophage colony-stimulating factor receptor is increased in the AbetaPP(V717F) transgenic mouse model of Alzheimer's disease. *Am J Pathol, 157*(3), 895-904. doi:10.1016/s0002-9440(10)64603-2

Nakamichi, Y., Udagawa, N., & Takahashi, N. (2013). IL-34 and CSF-1: similarities and differences. *J Bone Miner Metab, 31*(5), 486-495. doi:10.1007/s00774-013-0476-3

Nandi, S., Gokhan, S., Dai, X. M., Wei, S., Enikolopov, G., Lin, H., . . . Stanley, E. R. (2012). The CSF-1 receptor ligands IL-34 and CSF-1 exhibit distinct developmental brain expression patterns and regulate neural progenitor cell maintenance and maturation. *Dev Biol, 367*(2), 100-113. doi:10.1016/j.ydbio.2012.03.026

NCT01409915. Study of the Safety & Efficacy of Leukine® in the Treatment of Alzheimer's Disease. *ClinicalTrials.gov Identifier: NCT01409915. University of Colorado.*

NCT01433497. Efficacy and Safety of Masitinib in the Treatment of Progressive Multiple Sclerosis. *ClinicalTrials.gov Identifier: NCT01433497. AB Science.*
Targeting Neuroinflammation in Alzheimer’s disease: are we ready yet?

NCT01694277. Masitinib in Patients With Gastrointestinal Stromal Tumour After Progression With Imatinib. ClinicalTrials.gov Identifier: NCT01694277. AB Science.

NCT01790503. A Phase 1b/2 Study of PLX3397 + Radiation Therapy + Temozolomide in Patients With Newly Diagnosed Glioblastoma. ClinicalTrials.gov Identifier: NCT04121208. Daiichi Sankyo, Inc. & Plexxikon

NCT01872598. Masitinib in Patients With Mild to Moderate Alzheimer's Disease. ClinicalTrials.gov Identifier: NCT01872598. AB Science.

NCT02584647. PLX3397 Plus Sirolimus in Unresectable Sarcoma and Malignant Peripheral Nerve Sheath Tumors (PLX3397). ClinicalTrials.gov Identifier: NCT02584647. Columbia University & Plexxikon.

NCT02588677. Masitinib in Combination With Riluzole for the Treatment of Patients Suffering From Amyotrophic Lateral Sclerosis (ALS). ClinicalTrials.gov Identifier: NCT02588677. AB Science.

NCT03127267. Efficacy and Safety of Masitinib Versus Placebo in the Treatment of ALS Patients. ClinicalTrials.gov Identifier: NCT03127267. AB Science.

NCT03557970. CSF1R Inhibitor JNJ-40346527 in Treating Participants With Relapsed or Refractory Acute Myeloid Leukemia. ClinicalTrials.gov Identifier: NCT03557970, OHSU Knight Cancer Institute.

NCT04121208. MIcroglial Colony Stimulating Factor-1 Receptor (CSF1R) in Alzheimer's Disease (MICAD). ClinicalTrials.gov Identifier: NCT04121208. University of Oxford, Janssen Pharmaceutica.

Neal, M. L., Fleming, S. M., Budge, K. M., Boyle, A. M., Kim, C., Alam, G., . . . Richardson, J. R. (2020). Pharmacological inhibition of CSF1R by GW2580 reduces microglial proliferation and is protective against neuroinflammation and dopaminergic neurodegeneration. FASEB J, 34(1), 1679-1694. doi:10.1096/fj.201900567RR

Nelis, S. M., Wu, Y. T., Matthews, F. E., Martyr, A., Quinn, C., Rippon, I., . . . Clare, L. (2019). The impact of co-morbidity on the quality of life of people with dementia: findings from the IDEAL study. Age Ageing, 48(3), 361-367. doi:10.1093/ageing/afy155

Nelson, P. J., Rees, A. J., Griffin, M. D., Hughes, J., Kurts, C., & Duffield, J. (2012). The renal mononuclear phagocytic system. J Am Soc Nephrol, 23(2), 194-203. doi:10.1681/ASN.2011070680

Newcombe, E. A., Camats-Perna, J., Silva, M. L., Valmas, N., Huat, T. J., & Medeiros, R. (2018). Inflammation: the link between comorbidities, genetics, and Alzheimer's disease. J Neuroinflammation, 15(1), 276. doi:10.1186/s12974-018-1313-3

Nguyen, J. C., Killcross, A. S., & Jenkins, T. A. (2014). Obesity and cognitive decline: role of inflammation and vascular changes. Front Neurosci, 8, 375. doi:10.3389/fnins.2014.00375

Nichols, M. R., St-Pierre, M. K., Wendeln, A. C., Makoni, N. J., Gouwens, L. K., Garrad, E. C., . . . Combs, C. K. (2019). Inflammatory mechanisms in neurodegeneration. J Neurochem, 149(5), 562-581. doi:10.1111/jnc.14674
Targeting Neuroinflammation in Alzheimer’s disease: are we ready yet?

Nissen, J. C., Thompson, K. K., West, B. L., & Tsirka, S. E. (2018). Csf1R inhibition attenuates experimental autoimmune encephalomyelitis and promotes recovery. Exp Neurol, 307, 24-36. doi:10.1016/j.expneurol.2018.05.021

Obst, J., Simon, E., Martin-Estebane, M., Pipi, E., Barkwill, L. M., Gonzalez-Rivera, Y., . . . Gomez-Nicola, D. (2020). Inhibition of IL34 unveils tissue-selectivity and is sufficient to reduce microglial proliferation in chronic neurodegeneration. doi:bioRxiv doi:https://doi.org/10.1101/2020.03.09.976118

Odegaard, J. I., Ricardo-Gonzalez, R. R., Goforth, M. H., Morel, C. R., Subramanian, V., Mukundan, L., . . . Chawla, A. (2007). Macrophage-specific PPARgamma controls alternative activation and improves insulin resistance. Nature, 447(7148), 1116-1120. doi:10.1038/nature05894

Ohno, H., Uemura, Y., Murooka, H., Takanashi, H., Tokieda, T., Ohzeki, Y., . . . Serizawa, I. (2008). The orally-active and selective c-Fms tyrosine kinase inhibitor Ki20227 inhibits disease progression in a collagen-induced arthritis mouse model. Eur J Immunol, 38(1), 283-291. doi:10.1002/eji.200737199

Odegaard, J. I., Ricardo-Gonzalez, R. R., Goforth, M. H., Morel, C. R., Subramanian, V., Mukundan, L., . . . Chawla, A. (2007). Macrophage-specific PPARgamma controls alternative activation and improves insulin resistance. Nature, 447(7148), 1116-1120. doi:10.1038/nature05894

Oosterhof, N., Chang, I. J., Karimiani, E. G., Kuil, L. E., Jensen, D. M., Daza, R., . . . Bennett, J. T. (2019). Homozygous Mutations in CSF1R Cause a Pediatric-Onset Leukoencephalopathy and Can Result in Congenital Absence of Microglia. Am J Hum Genet, 104(5), 936-947. doi:10.1016/j.ajhg.2019.03.010

Ohno, H., Uemura, Y., Murooka, H., Takanashi, H., Tokieda, T., Ohzeki, Y., . . . Serizawa, I. (2008). The orally-active and selective c-Fms tyrosine kinase inhibitor Ki20227 inhibits disease progression in a collagen-induced arthritis mouse model. Eur J Immunol, 38(1), 283-291. doi:10.1002/eji.200737199

Olmos-Alonso, A., Schetters, S. T. T., Sri, S., Askew, K., Mancuso, R., Vargas-Caballero, M., . . . Gomez-Nicola, D. (2016). Pharmacological targeting of CSF1R inhibits microglial proliferation and prevents the progression of Alzheimer's-like pathology. Brain, 139, 891-907. doi:10.1093/brain/awv379

Odegaard, J. I., Ricardo-Gonzalez, R. R., Goforth, M. H., Morel, C. R., Subramanian, V., Mukundan, L., . . . Chawla, A. (2007). Macrophage-specific PPARgamma controls alternative activation and improves insulin resistance. Nature, 447(7148), 1116-1120. doi:10.1038/nature05894

Ohno, H., Uemura, Y., Murooka, H., Takanashi, H., Tokieda, T., Ohzeki, Y., . . . Serizawa, I. (2008). The orally-active and selective c-Fms tyrosine kinase inhibitor Ki20227 inhibits disease progression in a collagen-induced arthritis mouse model. Eur J Immunol, 38(1), 283-291. doi:10.1002/eji.200737199

Olmos-Alonso, A., Schetters, S. T. T., Sri, S., Askew, K., Mancuso, R., Vargas-Caballero, M., . . . Gomez-Nicola, D. (2016). Pharmacological targeting of CSF1R inhibits microglial proliferation and prevents the progression of Alzheimer's-like pathology. Brain, 139, 891-907. doi:10.1093/brain/awv379

Oosterhof, N., Chang, I. J., Karimiani, E. G., Kuil, L. E., Jensen, D. M., Daza, R., . . . Bennett, J. T. (2019). Homozygous Mutations in CSF1R Cause a Pediatric-Onset Leukoencephalopathy and Can Result in Congenital Absence of Microglia. Am J Hum Genet, 104(5), 936-947. doi:10.1016/j.ajhg.2019.03.010

Patwardhan, P. P., Surriga, O., Beckman, M. J., de Stanchina, E., Dematteo, R. P., Tap, W. D., & Schwartz, G. K. (2014). Sustained inhibition of receptor tyrosine kinases and macrophage depletion by PLX3397 and rapamycin as a potential new approach for the treatment of MPNSTs. Clin Cancer Res, 20(12), 3146-3158. doi:10.1158/1078-0432.CCR-13-2576

Pham, T. H., Minderjahn, J., Schmidl, C., Hoffmeister, H., Schmidhofer, S., Chen, W., . . . Rehli, M. (2013). Mechanisms of in vivo binding site selection of the hematopoietic master transcription factor PU.1. Nucleic Acids Res, 41(13), 6391-6402. doi:10.1093/nar/gkt355

Pixley, F. J., & Stanley, E. R. (2004). CSF-1 regulation of the wandering macrophage: complexity in action. Trends Cell Biol, 14(11), 628-638. doi:10.1016/j.tcb.2004.09.016

Poblador-Plou, B., Calderon-Larranaga, A., Marta-Moreno, J., Hanco-Saavedra, J., Sicras-Mainar, A., Soljak, M., & Prados-Torres, A. (2014). Comorbidity of dementia: a cross-sectional study of primary care older patients. BMC Psychiatry, 14, 84. doi:10.1186/1471-244X-14-84

Priceman, S. J., Sung, J. L., Shaposhnik, Z., Burton, J. B., Torres-Collado, A. X., Moughon, D. L., . . . Wu, L. (2010). Targeting distinct tumor-infiltrating myeloid cells by inhibiting CSF-1 receptor: combating tumor evasion of antiangiogenic therapy. Blood, 115(7), 1461-1471. doi:10.1182/blood-2009-08-237412

Radi, Z. A., Koza-Taylor, P. H., Bell, R. R., Obert, L. A., Runnels, H. A., Beebe, J. S., . . . Sadis, S. (2011). Increased serum enzyme levels associated with kupffer cell reduction with no signs of hepatic or skeletal muscle injury. Am J Pathol, 179(1), 240-247. doi:10.1016/j.ajpath.2011.03.029
Targeting Neuroinflammation in Alzheimer's disease: are we ready yet?

Rojo, R., Pridans, C., Langlais, D., & Hume, D. A. (2017). Transcriptional mechanisms that control expression of the macrophage colony-stimulating factor receptor locus. *Clin Sci (Lond)*, 131(16), 2161-2182. doi:10.1042/CS20170238

Rojo, R., Raper, A., Ozdemir, D. D., Lefevre, L., Grabert, K., Wollscheid-Lengeling, E., ... Pridans, C. (2019). Deletion of a Csf1r enhancer selectively impacts CSF1R expression and development of tissue macrophage populations. *Nature Communications*, 10(1), 3215. doi:10.1038/s41467-019-11053-8

Ruparelia, N., Chai, J. T., Fisher, E. A., & Choudhury, R. P. (2017). Inflammatory processes in cardiovascular disease: a route to targeted therapies. *Nat Rev Cardiol*, 14(3), 133-144. doi:10.1038/nrcardio.2016.185

Rustenhoven, J., Smith, A. M., Smyth, L. C., Jansson, D., Scotter, E. L., Swanson, M. E. V., ... Dragunow, M. (2018). PU.1 regulates Alzheimer's disease-associated genes in primary human microglia. *Molecular Neurodegeneration*, 13(1), 44. doi:10.1186/s13024-018-0277-1

Saltiel, A. R., & Olefsky, J. M. (2017). Inflammatory mechanisms linking obesity and metabolic disease. *Journal of Clinical Investigation*, 127(1), 1-4. doi:10.1172/JCI92035

Sassi, C., Nalls, M. A., Ridge, P. G., Gibbs, J. R., Lupton, M. K., Troakes, C., ... Hardy, J. (2018). Mendelian adult-onset leukodystrophy genes in Alzheimer's disease: critical influence of CSF1R and NOTCH3. *Neurobiol Aging*, 66, 179 e117-179 e129. doi:10.1016/j.neurobiolaging.2018.01.015

Sosna, J., Philipp, S., Albay, R., 3rd, Reyes-Ruiz, J. M., Baglietto-Vargas, D., LaFerla, F. M., & Glabe, C. G. (2018). Early long-term administration of the CSF1R inhibitor PLX3397 ablates microglia and reduces accumulation of intraneuronal amyloid, neuritic plaque deposition and pre-fibrillar oligomers in 5XFAD mouse model of Alzheimer's disease. *Molecular Neurodegeneration*, 13(1), 11. doi:10.1186/s13024-018-0244-x
Spangenberg, E. E., Lee, R. J., Najafi, A. R., Rice, R. A., Elmore, M. R., Blurton-Jones, M., . . . Green, K. N. (2016). Eliminating microglia in Alzheimer’s mice prevents neuronal loss without modulating amyloid-beta pathology. *Brain, 139*(Pt 4), 1265-1281. doi:10.1093/brain/aww016

Stanley, E. R., & Chitu, V. (2014). CSF-1 receptor signaling in myeloid cells. *Cold Spring Harb Perspect Biol, 6*(6). doi:10.1101/cshperspect.a021857

Stanley, E. R., & Heard, P. M. (1977). Factors regulating macrophage production and growth. Purification and some properties of the colony stimulating factor from medium conditioned by mouse L cells. *Journal of Biological Chemistry, 252*(12), 4305-4312. Retrieved from https://www.ncbi.nlm.nih.gov/pubmed/301140

Stephens, D. C., Kim, H. M., Kumar, A., Farahat, A. A., Boykin, D. W., & Poon, G. M. (2016). Pharmacologic efficacy of PU.1 inhibition by heterocyclic dications: a mechanistic analysis. *Nucleic Acids Res, 44*(9), 4005-4013. doi:10.1093/nar/gkw229

Sudo, T., Nishikawa, S., Ogawa, M., Kataoka, H., Ohno, N., Izawa, A., . . . Nishikawa, S. (1995). Functional hierarchy of c-kit and c-fms in intramarrow production of CFU-M. *Oncogene, 11*(12), 2469-2476. Retrieved from https://www.ncbi.nlm.nih.gov/pubmed/8545103

T’Jonck, W., Guilliams, M., & Bonnardel, J. (2018). Niche signals and transcription factors involved in tissue-resident macrophage development. *Cell Immunol, 330*, 43-53. doi:10.1016/j.cellimm.2018.02.005

Tarkowski, E., Wallin, A., Regland, B., Blennow, K., & Tarkowski, A. (2001). Local and systemic GM-CSF increase in Alzheimer's disease and vascular dementia. *Acta Neurol Scand, 103*(3), 166-174. doi:10.1034/j.1600-0404.2001.103003166.x

Tsalamandris, S., Antonopoulos, A. S., Oikonomou, E., Papamikroulis, G. A., Vogiatzi, G., Papaioannou, S., . . . Tousoulis, D. (2019). The Role of Inflammation in Diabetes: Current Concepts and Future Perspectives. *Eur Cardiol, 14*(1), 50-59. doi:10.15420/ecr.2018.33.1

Uemura, Y., Ohno, H., Ohzeki, Y., Takanashi, H., Murooka, H., Kubo, K., & Serizawa, I. (2008). The selective M-CSF receptor tyrosine kinase inhibitor Ki20227 suppresses experimental autoimmune encephalomyelitis. *J Neuroimmunol, 195*(1-2), 73-80. doi:10.1016/j.jneuroim.2008.01.015

Verheijen, J., & Sleegers, K. (2018). Understanding Alzheimer Disease at the Interface between Genetics and Transcriptomics. *Trends Genet, 34*(6), 434-447. doi:10.1016/j.tig.2018.02.007

Vincent, V. A., Selwood, S. P., & Murphy, G. M., Jr. (2002). Proinflammatory effects of M-CSF and A beta in hippocampal organotypic cultures. *Neurobiol Aging, 23*(3), 349-362. doi:10.1016/s0197-4580(01)00338-4

Walton, M. R., Gibbons, H., MacGibbon, G. A., Sirimanne, E., Saura, J., Gluckman, P. D., & Dragunow, M. (2000). PU.1 expression in microglia. *J Neuroimmunol, 104*(2), 109-115. doi:10.1016/s0165-5728(99)00262-3

Wang, Y., & Colonna, M. (2014). Interkeukin-34, a cytokine crucial for the differentiation and maintenance of tissue resident macrophages and Langerhans cells. *Eur J Immunol, 44*(6), 1575-1581. doi:10.1002/eji.201344365

Wang, Y., Szretter, K. J., Vermi, W., Gilfillan, S., Rossini, C., Cella, M., . . . Colonna, M. (2012). IL-34 is a tissue-restricted ligand of CSF1R required for the development of Langerhans cells and microglia. *Nat Immunol, 13*(8), 753-760. doi:10.1038/ni.2360
Figure 1. CSF1R/CSF-1/IL-34-dependent tissue-resident macrophage key functions.

CSF1R-, CSF1- and IL-34-dependent macrophage populations perform key functions to maintain homeostasis in different organs. Microglia, the main resident macrophages in the brain, are responsible for many critical functions during development and adulthood including support of neurogenesis, synaptic formation and pruning, and phagocytosis of apoptotic neurons and debris in the extracellular space (Colonna & Butovsky, 2017; Li & Barres, 2018). In the lungs, alveolar macrophages are responsible for the clearance of inhaled pathogens and particles (Davies, Jenkins, Allen, & Taylor, 2013; Maus et al., 2002), and they also play a critical role in the maintenance of alveolar homeostasis by clearing lipoprotein-containing alveolar surfactant produced by alveolar epithelial cells (Dranoff et al., 1994; TJonck, Guilliams, & Bonnardel, 2018). Kupffer cells, the resident macrophages in the liver, are involved in many immune and homeostatic functions such as clearing gut-derived toxins and pathogens from the blood, removal of damaged erythrocytes, as well as iron, bilirubin and cholesterol metabolism (Ganz, 2012; TJonck et al., 2018). The spleen contains multiple subsets of macrophages such as red pulp macrophages, located in the red pulp of the organ. They play a vital role in the clearance of senescent red blood cells and iron recycling (Kurotaki, Uede, & Tamura, 2015; TJonck et al., 2018). Next to red pulp macrophages, the spleen also contains marginal zone macrophages which are involved in the detection of antigens present in the bloodstream (den Haan & Kraal, 2012; Kierdorf, Prinz, Geissmann, & Gomez Perdiguero, 2015). Adipose-associated macrophages, present in the pancreas and adipose tissue all over the body, fulfil different functions such as removal of dead adipocytes, regulation of adipocyte lipolysis, storage and release to the bloodstream of excessive adipocyte-released lipids, and participation in the control of insulin sensitivity (Boutens & Stienstra, 2016; Odegaard et al., 2007; TJonck et al., 2018). Macrophages in the gastrointestinal tract continuously interact with the intestinal microbiome and maintain intestinal homeostasis regulating the immune response to commensals and defending the tissue against pathogens (Davies et al., 2013; Zimon & Jung, 2013). Langerhans cells are resident macrophages in the skin, involved in tissue surveillance, and uptake and transport of antigens to the skin-draining lymph nodes (Chorro & Geissmann, 2010; Kierdorf et al., 2015; TJonck et al., 2018). Renal macrophages play several roles such as surveillance of the environment, phagocytosis of pathogens and debris present in the extracellular matrix as well as support for nephrogenesis (Nelson et al., 2012). Circulating Ly-6Clo
monocytes are the predominant macrophage subset in the blood, acting as “intravascular housekeepers” in the clearance of endothelial cell debris as well as entering other tissues for the replenishment of tissue macrophage populations (Carlin et al., 2013; Gordon, Pluddemann, & Martinez Estrada, 2014).

Finally, different types of macrophages play critical roles in the bone. Osteoclasts are large multinucleated macrophages in charge of maintaining bone homeostasis and structure by resorption of the bone matrix produced by osteoblasts (Davies et al., 2013; T'Jonck et al., 2018), whereas bone marrow macrophages support erythropoiesis and maintain hematopoietic stem cells in stem cell niches (Chow et al., 2013; Chow et al., 2011; Davies et al., 2013). Considering the shared myeloid lineage of all these macrophage populations, it is anticipated that the immune and homeostatic key functions above described are susceptible to be affected by the immunomodulatory strategies to reduce neuroinflammation.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.