Structure and Expression of Bud Dormancy-Associated MADS-Box Genes (DAM) in European Plum

Carles Quesada-Traver1, Brenda Ivette Guerrero2, María Luisa Badenes1, Javier Rodrigo2,3, Gabino Ríos1 and Alba Lloret1*

1 Centro de Citricultura y Producción Vegetal, Instituto Valenciano de Investigaciones Agrarias (IVIA), Valencia, Spain,
2 Unidad de Hortofruticultura, Centro de Investigación y Tecnología Agroalimentaria de Aragón (CITA), Zaragoza, Spain,
3 Instituto Agroalimentario de Aragón-IA2, CITA-Universidad de Zaragoza, Zaragoza, Spain

Bud dormancy in temperate perennials ensures the survival of growing meristems under the harsh environmental conditions of autumn and winter, and facilitates an optimal growth and development resumption in the spring. Although the molecular pathways controlling the dormancy process are still unclear, DORMANCY-ASSOCIATED MADS-BOX genes (DAM) have emerged as key regulators of the dormancy cycle in different species. In the present study, we have characterized the orthologs of DAM genes in European plum (Prunus domestica L.). Their expression patterns together with sequence similarities are consistent with a role of PdoDAMs in dormancy maintenance mechanisms in European plum. Furthermore, other genes related to dormancy, flowering, and stress response have been identified in order to obtain a molecular framework of these three different processes taking place within the dormant flower bud in this species. This research provides a set of candidate genes to be genetically modified in future research, in order to better understand dormancy regulation in perennial species.

Keywords: European plum (Prunus domestica), DAM, bud dormancy, stress tolerance, flowering development

INTRODUCTION

Perennial species from temperate regions have to cope with seasonal changes in temperature, photoperiod, and water availability. Bud dormancy is an important adaptive mechanism ensuring survival during the cold period and paving the way for optimal growth resumption, flowering, and fruit production. During bud dormancy, these species cease growth and activate defense mechanisms, both essential to avoid injuries caused by the harmful environmental conditions during winter (Welling and Palva, 2006; Hänninen and Tanino, 2011). In European plum (Prunus domestica L.) and other members of the Rosaceae family, growth cessation and winter dormancy are chiefly regulated by external temperature, unlike other temperate trees that are also sensitive to photoperiod control (Heide and Prestrud, 2005; Heide, 2008). Winter dormancy presents two phases: endo- and ecodormancy (Lang et al., 1987). During endodormancy (abbreviated to dormancy in this study), the meristems remain protected within the reproductive and vegetative buds without apparent growth (Cooke et al., 2012). In this phase, flower buds require exposure to a specific range of chilling temperatures for proper flowering and subsequent fruiting. Chilling fulfilment does not cause an immediate resumption of growth because exposure to higher
temperatures is also required during the phase of ecodormancy to achieve bud break and flowering. Despite the importance of distinguishing these two phases to understand the mechanisms behind dormancy, there is a lack of phenological or biological markers to detect the fulfilment of chilling (Fadón and Rodrigo, 2018). Since dormancy is highly dependent on temperature, global warming is a developing threat with potential impact on phenological transitions and fruit production. On the one hand, warmer temperatures in spring are expected to increase the risk of frosts associated with premature flowering (Luedeling et al., 2011; Guo et al., 2014; Vitasse et al., 2018; Woznicki et al., 2019). On the other hand, climatic scenarios leading to higher temperatures in winter might cause severe reductions in winter chill, becoming insufficient to fulfil chilling requirements for dormancy release and thus causing inefficient and irregular budbreak and important production losses in fruit crops (Erez, 2000; Luedeling et al., 2011; Guo et al., 2014; Legave et al., 2015). Engineering fruit crops with altered dormancy, flowering, and stress tolerance responses should enable substantial advancements in breeding time and productivity. Therefore, it is particularly important to search out the molecular factors underlying these three processes (Lloret et al., 2018).

Several studies have been focused on a group of genes, called DORMANCY-ASSOCIATED MADS-BOX (DAM) and other orthologs of SHORT VEGETATIVE PHASE (SVP) genes, which recently have emerged as potential regulators of dormancy in several species such as almond (Prunus dulcis; Prudencio et al., 2018), apple (Malus domestica; Falavigna et al., 2014; Wu et al., 2017), apricot (Prunus armeniaca; Balogh et al., 2019), Chinese cherry (Prunus pseudocerasus; Zhu et al., 2015), hybrid aspen (Populus tremula x tremuloides; Singh et al., 2018), Japanese apricot (Prunus mume; Sasaki et al., 2011), kiwifruit (Actinidia chinensis; Wu et al., 2019; Actinidia delicosa; Wu et al., 2012), leafy spurge (Euphorbia esula; Horvath et al., 2008), pear (Pyrus pyrifolia; Saito et al., 2015), and sweet cherry (Prunus avium; Rothkegel et al., 2017). These genes were firstly identified in an evergreen mutant (evg) of peach (Prunus persica) that shows a non-dormant phenotype, maintaining apical growth and persistent leaves in response to dormancy inducing conditions (Bielenberg et al., 2004). This phenotype is associated with a genomic deletion that includes four out of the six tandemly-repeated DAM genes (Bielenberg et al., 2004; Bielenberg et al., 2008). Subsequently, other functional studies have confirmed their crucial role in the dormancy process. The ectopic expression of DAM1 from leafy spurge in Arabidopsis thaliana delays bolting and flowering concomitantly with the repression of FLOWERING LOCUS T (FT) (Horvath et al., 2010). In fact, DAM genes have been proposed to directly repress FT in leafy spurge (Hao et al., 2015) and Chinese white pear (Niu et al., 2016). PpDAM1 gene has been also described to bind and up-regulate the expression of pear PpNCED3 gene, encoding a 9-cis-epoxycarotenoid dioxygenase implicated in the synthesis of the dormancy-promoting hormone abscisic acid (Tuan et al., 2017). In addition, apple plants overexpressing MdDAMb genes show delayed bud break but normal flower and fruit development (Wu et al., 2017). Finally, transgenic poplar (Populus trichocarpa) and apple constitutively expressing PmDAM6 from Japanese apricot show growth inhibition and early bud set (Sasaki et al., 2011; Yamane et al., 2019). Overall, these studies suggest that DAM genes play a crucial role in dormancy maintenance mainly by growth and hormone regulation (Liu and Sherif, 2019; Yamane et al., 2019).

Other studies have focused on discerning the molecular mechanisms that regulate DAM gene expression. Some chromatin covalent modifications and miRNA have been reported to affect DAM expression in several species (Horvath et al., 2010; Leida et al., 2012; Saito et al., 2015; Niu et al., 2016; Vimont et al., 2020), suggesting the participation of epigenetic mechanisms in DAM-dependent dormancy modulation (Ríos et al., 2014; Conde et al., 2019). A TEOSINTE BRANCHED1/CYCLOIDEA/PROLIFERATING CELL FACTOR (TCP) protein (PpTCP20) binds to a specific element and down-regulates both PpeDAM5 and PpeDAM6 in peach (Wang et al., 2020). Also, a feedback regulatory mechanism has been described involving repression of PpDAM1 by an abscisic acid response element (ABRE)-binding transcription factor (PpAREB1) in pear (Tuan et al., 2017). More recently, pear ABRE-BINDING FACTORS3 (PpyABF3) has been reported to specifically bind and activate PpyDAM2 expression, whereas PpyABF2/PpAREB1 represses PpyDAM3 by dimerising with PpyABF3 (Yang et al., 2020). Among DAM regulatory proteins, the most studied ones are C-Repeat Binding Factors (CBF), which are able to bind and activate DAM promoters by yeast one-hybrid and transient expression experiments in Japanese apricot and pear (Saito et al., 2015; Niu et al., 2016; Zhao et al., 2018b). Interestingly, CBF factors are also involved in the cold-temperature response pathway (Wisniewski et al., 2011; Wisniewski et al., 2018), revealing the close relationship between dormancy and low temperature tolerance mechanisms.

Plants reprogram their gene expression profile in order to cope with cold and desiccation stresses associated with the dormancy period. Thus, the analysis of differentially expressed genes in dormant buds has provided many candidate genes belonging to abiotic tolerance responses. The ectopic expression of the cold acclimation response gene CBF from peach improves freezing tolerance in apple (Wisniewski et al., 2011; Wisniewski et al., 2015). Several studies suggest that evg mutant of peach has lower cold tolerance due in part to a lack of a dehydrin accumulated in bark tissues during dormancy progression (Arora et al., 1992; Arora and Wisniewski, 1994; Arora et al., 1996; Artlip et al., 1997). In addition, the ectopic expression of STRESS-ASSOCIATED PROTEIN1 (PpSAP1) gene, highly expressed in dormant buds of peach, improves water retention under desiccation conditions in transgenic plum (Lloret et al., 2017a). Moreover, some genes related to sugar metabolism pathways with altered expression along bud development have been shown to participate in tolerance mechanisms. In peach, PpeS6PDH gene encoding a sorbitol-6-phosphate dehydrogenase has been postulated to synthesize the compatible solute sorbitol in order to protect dormant buds against cold and hydric stresses (Lloret et al., 2017b). In addition, galactinol synthase (GOLS) genes, involved in the synthesis of
raffinose family oligosaccharides, are up-regulated in dormant buds of chestnut and apple (Ibáñez et al., 2013; Falavigna et al., 2018). Interestingly, the overexpression of apple MdGolS2 confers tolerance to water deficit in Arabidopsis (Falavigna et al., 2018).

During winter dormancy, growth and development of flower organs ceases in order to deal with the harmful environmental conditions. In stone fruits, flower bud formation takes place in summer in the year preceding flowering and fruiting, whereas flower primordia stop growing and remain dormant inside the buds. The dormancy arrest guarantees an optimal development of gametes under more suitable conditions after dormancy release, which associates with a sharp up-regulation of genes involved in pollen microsporogenesis, among other flowering processes (Rios et al., 2013).

European plum is the fruit tree species with the highest number of cultivars in Europe (Esmenjaud and Dirlewanger, 2007). Among them, ‘Reine Claude Verte’ is the most grown cultivar for its excellent organoleptic qualities (Gharbi et al., 2014). This cultivar has been cultivated for more than 500 years in Europe, from where it has expanded worldwide (Tabuenca and Iturrioz, 1991). European plum is also a genetic model for other Prunus species by virtue of the availability of efficient procedures for genetic transformation and regeneration (Petri et al., 2018). Based on previous studies in peach and other species, this study aims at identifying and characterizing the DAM and other dormancy related genes from European plum, in order to obtain a dynamic snapshot of molecular mechanisms and factors affecting dormancy in flower buds of this species. This research will open the possibility to initiate functional studies on dormancy related genes with the use of transgenic plum plants overexpressing or down-regulating some of these genes.

RESULTS

Chilling Requirements for Dormancy Release in European Plum

The chilling requirements for dormancy release of flower buds from European plum cv. ‘Reine Claude Verte’ were estimated during two consecutive years. The annual temperature regime was very similar in both years, showing a close pattern of chilling accumulation (Supplementary Figure S1). Chilling fulfilment was achieved at similar dates in both seasons, January 25 for 2018–2019 and January 27 for 2019–2020 (Figures 1A, B). The date of chilling fulfilment allowed estimating the chilling requirements of ‘Reine Claude Verte’ by the calculation of the number of chilling hours (CH), chilling units (CU), and chilling portions (CP) accumulated until then. The range of the values obtained in the two seasons, 979–1,086 CH, 1,248–1,287 CU, and 62.8 CP (same value in both years), were considered as the chilling requirements of this cultivar (Supplementary Table S1).

From the end of autumn and during winter, all the flower buds were closed and covered by dark brown scales at phenological stage A (Baggiolini, 1952) and stage BBCH 50 (Fadón et al., 2015) (Figure 1C). At this stage, two to three

![FIGURE 1 | Estimation of breaking of dormancy and flower bud characterization in European plum cv. ‘Reine Claude Verte.’ Flower bud weight in orchard conditions (gray squares) and after 8 days in the growth chamber (black rhombi) over two seasons: 2018–2019 (A) and 2019–2020 (B). Snowflakes mark the weight increment of 30%, when chilling was fulfilled. (C) Shoot of European plum cv. ‘Reina Claude Verte’ during dormancy, showing flower (FB), and vegetative (VB) buds. (D) Longitudinal section of a flower primordium at the date of dormancy breaking. Sepals (SE), petals (PE), anthers (AN), and pistil (PI); scale bar, 300 µm.](Frontiersin.org/FPSI/1288/images/fig1.png)
flower primordia were present inside each flower bud with all the whorls (sepsals, petals, anthers, and pistil) differentiated (Figures 1D). In autumn, the average weight of flower buds was around 0.1 g, and it remained without significant variations throughout dormancy in both years (Figures 1A, B). No external phenological changes were observed until bud burst in spring several weeks after chilling fulfilment.

**Genome-Wide Identification of PdoDAM1-6 Genes From European Plum**

We used the coding sequence of PpeDAM1-6 genes from peach (Prunus persica) for a BLASTN search in the Prunus domestica v1.0 draft genome, recently uploaded to the Genome Database for Rosaceae (GDR; https://www.rosaceae.org/). We found three assembled scaffolds that exhibited high similarity to DAM locus of peach and contained at least the putative orthologues of the six PpeDAM genes and their flanking genes. The circle plot revealed strong conservation along the three scaffolds with the PpeDAM region, localized in the chromosome 1 of peach (Figure 2). However, they also presented a weaker synteny with peach chromosomes 6 and 8. In this region of the chromosome 6, we found the putative ortholog in the peach genome of Arabidopsis SHORT VEGETATIVE PHASE (SVP) systematically named Prupe.6G199000, belonging to the StMADS11 lineage of type II MIKCC MADS-box genes in which PrupeDAM1 and PrupeDAM4 like gene structures consisting of eight exons and seven introns flanked by the translation initiation and stop codons (Figure 4). Transcriptomic data suggested an alternative splicing for PdoDAM4-like and PdoDAM5-like, based on RNA-seq data and the presence of stop codons at the 3’ end. Both transcripts were confirmed by gene expression analyses during bud development although PdoDAM4.1 and PdoDAM5.2 were clearly less abundant (Supplementary Table S2). As the exons were well conserved across the three scaffolds (Supplementary Table S3), we selected PdoDAMs predicted coding sequences from scaffold104-v.1.0 and the most abundant isoform in each case for subsequent analysis and used the names PdoDAM1-6 to designate them (Supplementary Figure S2).

The deduced amino acid sequences of all identified PdoDAMs contained the highly conserved DNA binding MADS-box domain at the N-terminal end, the K-domain

![Figure 2](image-url) | Syntenic relationships between European plum scaffolds containing the DAM locus and the peach genome. European plum scaffold104-v.1.0, scaffold1884-v.1.0, and scaffold3194-v.1.0 (purple) and peach chromosomes Ppe chr1, Ppe chr6, and Ppe chr8 (yellow) are represented using ClicO FS. Scoring regions are connected by a ribbon and colored according to a grayscale that associates black color to the maximum score for each representation. DAM region in Ppe chr1 is marked with a black asterisk. European plum scaffolds are five-fold magnified in the representation for a proper visualization.
participating in protein-protein interactions and finally the variable intervening region that connects both domains (Figure 5). These observations confirmed that PdoDAM genes belong to the MIKC⁺-type of MADS-box genes, consistently with previous studies in other species (Jiménez et al., 2009; Zhao et al., 2018a).

The phylogenetic analysis of PdoDAM1-6 predicted protein sequences indicated that they form part of a differentiated group in conjunction with other Prunus sequences, which was divided into six subgroups corresponding to the six tandemly repeated DAM genes (Figure 6). In addition, DAM proteins from genera Malus and Pyrus constituted another well-defined group, that jointly with Prunus DAMs were clearly differentiated from a cluster composed by SVP-like proteins from Arabidopsis thaliana and kiwifruit (Actinidia deliciosa) and EeDAM1-2 from leafy spurge.

Expression Analysis of PdoDAM1-6 Genes

Since the best-known function of previously described DAM-like genes has been related to bud dormancy regulation, we measured PdoDAM1-6 gene expression across bud development in European plum cv. ‘Reine Claude Verte’ during the winter of 2018/2019. The six PdoDAM1-6 genes showed a general progressive decrease in mRNA level until the dormancy period was completely overcome, with specific gene particularities (Figure 7A). Despite PdoDAM1 expression did not show great changes along bud development, a slight down-regulation was observed from the first two samples, collected in autumn. On the contrary, PdoDAM2 was the European plum DAM gene with strongest gene down-regulation, occurring in early stages before dormancy release (November–December), and maintained along the whole bud development process. PdoDAM3 and PdoDAM4 presented a similar expression pattern, with transcripts levels increasing slightly from November to December and then declining during bud dormancy progression. Finally, PdoDAM5 and PdoDAM6 also showed paralleled expression profiles. They were strongly down-regulated prior to bud dormancy release, and subsequently they slightly peaked in ecodormant Cv7 sample, at the beginning of February.

When comparing these expression patterns with those of PpeDAM genes in peach cv. ‘Crimson Baby’ at five different bud dormancy samples (CB1–CB5), we found both common and specific features (Figure 7B). Whereas PpeDAM1, PpeDAM2, and PpeDAM3 showed a quite different expression profile from their European plum counterparts, PpeDAM4, PpeDAM5, and PpeDAM6 reduced their expression during bud development in a broadly similar fashion to European plum orthologs. PpeDAM4 slightly increased its expression level in the first samples, followed by a sharp down-regulation concomitantly with dormancy release. As previously described, PpeDAM5 and PpeDAM6 transcript levels decreased in precise concordance with bud dormancy release. Interestingly, plum PdoDAM4, PdoDAM5, and PdoDAM6 gene repression occurred similarly but in a slightly more advanced manner than in their peach orthologs.

Analysis of Cis-Regulatory Elements in PdoDAM1-6 Regulatory Regions

We searched in PdoDAM genes different motifs described in previous reports as regulatory elements of DAM genes from other species. Since these elements were previously identified in both, the promoter and the intronic region between the first exons of DAM genes, we focused on those regulatory genomic regions in PdoDAM genes. MADS-box transcription factors bind DNA sequences known as CArG box motif with the consensus sequence CC(A/T)₆GG or the non-canonical C(A/T)₈G. MADS-box genes, as DAM genes, have been reported to interact with these elements to regulate the expression of other genes but also to regulate themselves (Zhu and Perry, 2005; Gregis et al., 2013). The presence of CArG box sequences in all PdoDAM genes is thus consistent with self-regulation mechanisms (Figure 8). On the other hand, CBF proteins play a critical role in activation of cold responsive genes. PmuDAM6 gene was reported to be
FIGURE 4 | RNA coverage representation in DAM genes for each European plum scaffold. For each DAM gene, three coverage graphs corresponding to the different scaffolds represent number of reads. The predicted exonic structure is depicted below (orange boxes). The coverage plot was obtained by merging bam files from three RNA samples.
transcriptionally regulated by the direct binding of PmuCBFs to CRT/DRE elements. As shown in Figure 8, these elements were identified along the whole genomic regions under study, but were particularly abundant on the promoters of DAM6 genes from Japanese apricot, peach, and European plum, in a position close to active CRTs described by the literature. We also highlighted the site II motif associated with the sequence GCCCA. This element is recognized by TCP transcription factors described as down-regulators of PpeDAM5 and PpeDAM6 expression in peach (Wang et al., 2020). Site II motifs were identified in the intronic regions of all six PdoDAM genes and the promoters of PdoDAM3 and PdoDAM6, closely located to the one reported by the literature.

Expression Analysis of Other Dormancy-Related Genes

Previous genome-wide transcriptional analyses have provided numerous genes differentially expressed in flower buds of peach, which are related to three major coincident processes: bud dormancy regulation, stress tolerance and flowering development (Ríos et al., 2013; Lloret et al., 2017a, Lloret et al., 2017b). We analyzed the bud-dependent expression of the putative orthologs in European plum of some of these genes. Firstly, a TONOPLAST INTRINSIC PROTEIN (TIP)-like gene increased its expression in flower buds of European plum after dormancy release (Figure 9A), in close agreement with the behavior of its peach ortholog and its proposed function in tonoplast turgor and growth resumption (Figure 9B). We also studied the expression patterns of SORBITOL-6-PHOSPHATE DEHYDROGENASE (S6PDH)-like and STRESS ASSOCIATED PROTEIN (SAP)-like genes, which are postulated to participate in the stress tolerance response during bud dormancy in peach. Interestingly, S6PDH-like increased its expression level during dormancy progression until January 16th (CV4) and from that point it became steady and started to decrease. On the other hand, SAP-like expression was quite stable along dormancy progression and decreased after dormancy release (Figure 9A). Finally, RUPTURED POLLEN GRAIN1 (RPG1)-like and peroxidase-like genes, associated with microsporogenesis and pollen development in anthers, were up-regulated after the dormancy period, coincidently with the last phases of flower development usually activated at this stage. The expression profile of these genes was in concordance with peach bud expression patterns (Figure 9B).

DISCUSSION

The susceptibility of phenological transitions to changing climatic scenarios is a major challenge for temperate fruit crops, with a high potential impact on flowering and fruit production, as was already reported for European plum (Woznicki et al., 2019). Among these phenological transitions, bud dormancy modulates consecutively growth cessation and resumption in response to seasonal environmental conditions, and directly affects flowering timing and efficiency, which makes the understanding of this process essential for improving adaptation strategies to global temperature changes. The estimation of chilling requirements for bud dormancy release, based on bud-break forcing assays in combination with chilling quantification using one or more mathematical models, has become a key tool for the estimation of adaptability of a species or cultivar. In particular, chilling requirement of European plum cv. ‘Reine Claude Verte’ calculated in this study (979–1,086 CH) is in close agreement with previous estimations (976–1,275 CH; Tabuenca, 1967), and similar to those reported in other European plum cultivars (Fadón et al., 2020).
Bud dormancy is regulated by DAM transcription factor genes, mainly in Prunus species (Bielenberg et al., 2008; Sasaki et al., 2011; Balogh et al., 2019). The availability of the draft genome of European plum, recently uploaded to the GDR database, has allowed the identification and genomic characterization of six plum DAM genes in this study, designated PdoDAM1-6. Domain alignments and phylogenetic analysis indicated that they are closely related to the corresponding Prunus DAM sequences. PdoDAM genomic regions showed high synteny with three different regions of the peach genome. The strongest synteny corresponds to the end of chromosome 1 where PpeDAM are located. However, the second syntenic block is situated within chromosome 6, containing the putative ortholog of SVP in peach (PpeMADS57), as stated in a systematic analysis of the MADS-box family in peach (Wells et al., 2015). This observation showed the relationship between PdoDAM genes and SVP genes, as has been already reported in peach (Jiménez et al., 2009). On the other hand, the expansion of DAM genes has been proposed to be originated by serial tandem duplications before the diversification of the Prunus genus (Jiménez et al., 2009; Zhang et al., 2012), consistently also with this study. Expansion of DAM genes occurred independently in
the Maleae tribe including apple and pear, as supported by the identification of several DAM-like genes with a common genomic organization, essentially different from *Prunus* species (Wu et al., 2017). The independent origin of these duplication events and the observation of a strong purifying selection in all six *PpeDAM* genes, in spite of their sequence similarity, suggest the specialization of these genes for unique roles (Jiménez et al., 2009).

This proposed neo-functionalization of DAM genes is coherent with their different expression timing during dormancy progression. In this respect, our gene expression data in European plum matched previous studies in other related species. Our results showed an increase in *PdoDAM4* expression levels from November to December, followed by gene down regulation prolonged after dormancy release, as similarly reported in other *Prunus* species (Leida et al., 2010; Yu et al., 2020). On the other hand, peach *PpeDAM5* and *PpeDAM6* expression were inversely correlated with dormancy release, and thus were considered good predictors of the dormancy stage of buds (Li et al., 2009; Jiménez et al., 2010; Leida et al., 2010; Leida et al., 2012). European plum *PdoDAM5* and *PdoDAM6* showed a similar transcript level decrease, although it occurred earlier than in peach, using the estimated date of dormancy release as reference. Once dormancy was released, both *PdoDAM5* and *PdoDAM6* profiles reached a secondary minor peak, which was similarly reported in apricot *ParDAM5* and *ParDAM6* genes (Balogh et al., 2019). In spite of their transcriptional particularities, *PdoDAM* genes were in general

---

**FIGURE 7** | Relative expression of DAM genes in flower buds. *PdoDAM1-6* gene expression in European plum cv. ‘Reine Claude Verte’ (CV) is represented in panel (A), and *PpeDAM1-6* gene expression in peach cv. ‘Crimson Baby’ (CB) in panel (B). Timepoint codes are found in Material and Methods. The dashed bar indicates dormancy release. For each graph, an expression of one is assigned to the highest value. Each point represents data of three biological replicates accompanied by error bars representing its standard deviation. Significant differences among samples are represented by different letters (a–e), with a confidence level of 95%.
down-regulated along bud dormancy and development, according to previous studies (Sasaki et al., 2011; Zhao et al., 2018a; Yu et al., 2020).

Gene expression diversification and neo-functionalization of duplicated DAM genes could be at least partially originated in the variation of regulatory elements responding to different, but closely related environmental signals (Haake et al., 2002; Lawton-Rauh, 2003) both in promoters and in first kilobases of intronic regions (Rose, 2019). The identification of chromatin modifications related to gene activity and silencing in the large intron of DAM genes in peach (Leida et al., 2012; de la Fuente et al., 2015) recommends the search of intronic sequences in addition to promoters. The direct binding of CBF proteins to CRT/DRE elements in promoter regions has been proposed to mediate the effect of low temperature on DAM gene expression in leafy spurge (Horvath et al., 2010), Japanese pear (Saito et al., 2015; Niu et al., 2016), apple (Mimida et al., 2015; Wisniewski et al., 2015), and Japanese apricot (Zhao et al., 2018a). The distinct number and position of these CRT/DRE elements in the specific PdoDAM1-6 promoters are expected to affect their particular environmental response. Other transcription factors involved in regulation of DAM genes in other species have also cis-regulatory elements in the promoter and large intron of PdoDAMs. Among them, site II motifs are able to bind TCP factors similar to PpeTCP20, which is involved in PpeDAM5 and PpeDAM6 down-regulation when dormancy is released in peach (Wang et al., 2020). On the other hand, MADS-box factors binding to CArG box sequences are in certain cases self-regulating their own expression (Zhu and Perry, 2005; Gregis et al., 2013), and thus we cannot discard the cross-regulation of DAM genes in complex with other DAM or MADS-box proteins, as was indirectly suggested by the down-regulation of the whole DAM array of genes when four of them were deleted in the evg mutant of peach (Bielenberg et al., 2008). In addition to conventional transcription factors, also chromatin modification complexes are affecting DAM genes (Horvath et al., 2010; Leida et al., 2012; Saito et al., 2015; Vimont et al., 2020). Conserved GA repeat sequences have been found associated with regions enriched in the H3K27me3 histone modification in introns of PpeDAM genes (de la Fuente et al., 2015), other peach dormancy-regulated genes (Lloret et al., 2017b), and Arabidopsis genes (Deng et al., 2013). These GA repeats were found in a similar position and structure in PdoDAM genes, which suggests the conservation of chromatin-dependent mechanisms involved in DAM silencing responding to seasonal chilling accumulation (Ríos et al., 2014).

In addition to DAMs, other genes have been postulated to participate in dormancy regulation and other concurrent processes which are precisely orchestrated within flower buds, that is cold and hydric stress tolerance and flowering pathways (Lloret et al., 2018).

We identified putative orthologs of some of these genes in European plum. One of them encoded a TIP aquaporin involved in water permeability and transport of small molecules across the tonoplast membrane. TIP-like genes have been related to cell turgor and cell enlargement (Ludevid et al., 1992; Maurel et al., 2015). TIP-like gene was induced once dormancy was released in both peach and European plum, supporting a role in growth resumption and bud-break in ecodormant buds (Lloret et al., 2017a). Interestingly, the transport of metabolites and regulatory molecules across cell membranes has been found related to cell-to-cell communication and bud dormancy regulation in previous articles (Rinne et al., 2001; Rinne et al., 2011; Tylewicz et al., 2018). Meristem cells are isolated by plasmodesmata callose plugging during dormancy, and exposure to chilling temperatures causes the degradation of the callose and the
subsequent restoration of cell-to-cell communication. This mechanism is based on the action of 1-3-β-D-glucanases (Rinne et al., 2001) and abscisic acid (ABA) (Tylewicz et al., 2018). Three groups of genes, members of the \( \text{GLUCAN HYDROLASE 17 (GH17)} \) family, are upregulated by the chilling temperatures and biosynthesis of gibberellins. Short-term photoperiodic exposure transiently regulates the group 1 \( \text{GH17} \) genes to maintain the symplasmic paths to facilitate bud formation. Gibberellin \( \text{GA}_3 \) and long-term chilling exposure up-regulate the groups 2 and 3 \( \text{GH17} \) genes, allowing callose removal and reopening the signaling. After the required chilling accumulation, mild temperatures up-regulate growth-related genes, mediated by \( \text{GA}_4 \), leading to bud burst (Rinne et al., 2011). The tonoplast localization of TIP protein precludes its participation in these cell-to-cell communication processes, however it supports the key role of membrane permeability in dormancy release events.

On the other hand, two genes described as stress tolerance factors during peach dormancy, \( \text{S6PDH-like} \) and \( \text{SAP1-like} \) were also differentially regulated during bud dormancy in European plum. \( \text{PpeS6PDH} \) codifies a sorbitol-6-phosphate dehydrogenase involved in sorbitol synthesis, a major translocatable photosynthate in Rosaceae species that has been hypothesized to act as a compatible solute protecting dormant buds against cold and hydric stresses (Lloret et al., 2017b). Furthermore, the ectopic expression of \( \text{PpeSAPI1} \) altered leaf morphology and reduced water loss in transgenic plums (Lloret et al., 2017a). The expression profile of \( \text{S6PDH-like} \) and \( \text{SAPI1-like} \) genes suggests the conservation of their respective protective roles against the harmful environmental conditions in dormant buds of different \( \text{Prunus} \) species. Finally, we also identified genes related to flowering pathways previously reported to be differentially expressed in peach (Rios et al., 2013). A peroxidase-like and \( \text{RPG1-like} \) genes are involved in sporopollenin synthesis, a component of the outer cell wall of the pollen grain (exine) essential for pollen maturation and viability (Guan et al., 2008; Jacobowitz et al., 2019). These genes were up-regulated after flower bud dormancy release in peach and European plum in this work, as expected for genes participating in

FIGURE 9 | Relative expression of dormancy-related genes in flower buds. Bud samples from European plum cv. ‘Reine Claude Verte’ (CV) and peach cv. ‘Crimson Baby’ (CB) are respectively represented in panels (A, B). Timepoint codes are found in Material and Methods. The dashed bar indicates dormancy release. For each graph, an expression of one is assigned to the highest value. Each point represents data of three biological replicates accompanied by error bars representing its standard deviation. Significant differences among samples are represented by different letters (a–e), with a confidence level of 95%.
pollen development in the tapetum layer of anthers, since this process is finely coordinated with the dormancy period (Julian et al., 2011).

This work provides putative key elements of a molecular framework of bud dormancy regulation and other concurrent processes lying in flower buds of European plum. This species has efficient procedures for genetic transformation (Petri et al., 2018), as opposed to most woody fruit species, which are recalcitrant for genetic transformation and regeneration (Petri and Burgos, 2005). Thus, genes identified in this work are potential candidate genes for studying dormancy regulation and other related processes by RNA interference and other functional genetics procedures.

MATERIAL AND METHODS

Plant Material and Sample Collection
Six trees of European plum (Prunus domestica L.) cv. ‘Reina Claude Verte’ were selected for the experiment from a collection held at the Centro de Investigación y Tecnología Agroalimentaria, CITTA, Zaragoza, Spain, at 41°44’30” N, 0°47’00” W, and 220 m above sea level. For flower bud development expression analysis, buds were collected during autumn–winter 2018–2019 on November 28 (CV1), December 5 (CV2), December 18 (CV3), January 16 (CV4), January 23 (CV5), January 30 (CV6), February 6 (CV7), and February 13 (CV8).

Peach trees (Prunus persica L. Batsch cv. ‘Crimson Baby’) required in this study were grown at the Instituto Valenciano de Investigaciones Agrarias located in Moncada, Spain, at 39°35’20” N, 0°23’43” W, and 75 m above sea level. For expression analysis, flower buds were collected during autumn–winter 2015–2016 on November 30 (CB1), December 14 (CB2), December 21 (CB3), January 4 (CB4), and January 19 (CB5).

Determination of Breaking of Dormancy
Dormancy was experimentally determined for ‘Reina Claude Verte’ over two consecutive winters (2018–2019 and 2019–2020). To determine the date of dormancy release, five shoots (20–30 cm in length and 4–8 mm in diameter, with at least 10 flower buds) were randomly sampled every week, starting in late November until the onset of budbreak at the mid February. The shoots were placed on water-soaked florist foam and maintained in a growth chamber with a photosynthetic photon flux density (PPFD) of 70 µmol m⁻² s⁻¹ provided by cool daylight (6500 K) tubes (Osram L58W/865) under a 12-h light photoperiod at 22 ± 1°C for 8 days (Fadón et al., 2018). To determine differences in bud growth, 10 flower buds were randomly picked and weighed on the third and last day in the growth chamber. The date of dormancy release was established when the weight of the flower buds increased by at least 30% after 8 days in the chamber (Fadón and Rodrigo, 2019).

In peach, the date of dormancy release was measured as Leida et al. (2012). Briefly, 10 budsticks from three different trees were placed in a chamber with a PPFD of 27 µmol m⁻² s⁻¹ provided by a cool daylight (6500 K) tubes (Osram L58W/765 and set at 24°C 12 h:12 h light:dark cycle. Dormancy release was considered when more than 50% of buds showed at least the green tip of the sepals after 14 days.

Estimation of Chilling Requirements
Temperatures were recorded hourly at a meteorological station located in the experimental orchard over the two seasons. Chilling was quantified according to the three most commonly used temperature models to quantify chilling in fruit trees (Fadón et al., 2020). The Chilling Hours model defined “chilling hour” (CH) as 1 h at or below 7.2°C (Weinberger, 1950). The Utah model weights different ranges of temperatures based on their effect on dormancy fulfilment, proposing the use of “chilling-units” (CU) (Richardson et al., 1974). Finally, the Dynamic Model considered the effects of high and mild temperatures during winter on dormancy release and proposed the use of “chilling portions” (CP) (Fishman et al., 1987). In each model, the chilling requirements were considered as the range of values estimated in the 2 years studied.

Microscope Preparations
Ten flower buds from European plum sampled were fixed in ethanol 95% acetic acid 3:1 (v/v), and then transferred 24 h later to ethanol 75% at 4°C for conservation. For histochemical examination, flower buds were dehydrated in a tertiary butyl alcohol series (70, 85, 95, and 100% v/v), embedded in paraffin wax, sectioned at 10 mm in a Jung 2405 rotary microtome (Leica Microsystems), and placed onto glass slides previously coated with Happt’s adhesive. Prior to staining, the sections were rehydrated (three washes of 5 min in Histoclear II [CellPath], one in Histoclar II:ethanol [1:1, v/v] for 5 min, and one in an ethanol series [100, 70, and 40% v/v] for 2 min) (Fadón and Rodrigo, 2019). The samples were then stained using the potassium iodide-iodine reaction (I₂KI) for 5 min (Fadón et al., 2018). Preparations were observed under a bright field Leica DM250 microscope (Leica Microsystems). Micrographs were taken with an IDS UI-1490SE digital camera with the IDS Software Suite 4.93.0. (IDS Imaging Development Systems GmbH).

Identification of DAM Genes in European Plum Genome
Peach genomic PpDAM1-6 sequences and two adjacent genes at each side were selected to perform a BLASTN (Zhang et al., 2000) analysis, with default values, against the Prunus domestica Draft Genome Assembly v1.0 (Zhebentyayeva et al., 2019) available at the Genome Database for Rosaceae (GDR; Jung et al., 2019). Only those scaffolds in which the E-value was zero for all the queries were subjected to a synteny analysis utilizing GDR integrated tool Tripal Synteny Viewer, that used MScanX (Wang et al., 2012) with default settings and blast files resulting from BLASTP with an E-value cutoff of 1·10⁻¹⁰, a maximum alignment of 5 and maximum scores of 5. Results were reproduced using ClicO FS, a web-based service to generate circular plots (Cheong et al., 2015). The coding sequences (CDS) of PdoDAM1-6 were predicted using the web-server tool Prot2gene (http://genomics.brocku.ca/Prot2gene/), using protein sequences from PpeDAM1-6 as a guide. RNA-seq data were

Frontiers in Plant Science | www.frontiersin.org August 2020 | Volume 11 | Article 128812
used to manually curate CDS regions for PdoDAM1-6 previously predicted. Single reads were downloaded from the National Center for Biotechnology Information (NCBI) BioProject database (accession PRJNA630876), and aligned with the *Prunus domestica* Draft Genome Assembly v1.0 using STAR. Representations of both RNA-seq coverage and CDS was performed by means of the R package: Gviz (Hahne and Ivanek, 2016).

**Gene Structural Analysis of PdoDAM1-6 Genes**

Structural distribution of the previously studied genes was compared with other genomes. For every CDS obtained previously BLASTN with default values was performed against each of the different genomes used for comparison. The best hit for the first and last nucleotide position was taken as the genomic coordinates to draw the diagram. Diagram was generated using the “Gviz” R package for genomic representations mentioned above.

**Phylogenetic Analysis of Plum DAM Genes**

DAMs and DAM-related protein sequences from other species were downloaded from GeneBank TAIR and PlantTFDB. Their accession numbers and their references are shown in Supplementary Table S4. These sequences along the predicted protein sequences of PdoDAM1-6 constituted the input for performing a multiple alignment with Clustal Omega (Sievers et al., 2011). Blocks of highly homologous regions were selected from the multiple alignment file for further analysis using Gblocks (Talavera and Castresana, 2007). Phylogenetic tree was performed using MEGA 7 (Kumar et al., 2016) with Maximum Likelihood method based on Jones-Taylor-Thorton (JTT) matrix-based model (Jones et al., 1992) and allowing for invariant sites (+I) and using a discrete gamma model (+G) (Yang, 1994). The tree was tested using a bootstrap of 1,000 replicates, removing the nodes with less than 70% bootstrap confidence.

**Analysis of Cis-Elements in PdoDAM1-6 Regulatory Regions**

The cis-elements were predicted using the genomic sequences of PdoDAM1-6 genes from the promoter until the start of the second exon, by means of PlantCARE (http://bioinformatics.psb.ugent.be/webtools/plantcare/html/) (Lescot et al., 2002) and PLACE databases (http://www.dna.afrc.go.jp/PLACE/) (Higo et al., 1999).

**Expression Analysis by Real-Time Quantitative PCR (RT-qPCR)**

Total RNA was isolated from bud samples using Plant/Fungi Total RNA Purification Kit (Norgen, Thorold). Lysis buffer was supplemented with Polyvinylpyrrolidone (PVP-40) 1% (w/v) right before usage. Potential genomic DNA was removed with the RNase-free DNase I kit (Norgen Thorold). After assessing RNA integrity by gel electrophoresis, 500 ng of each sample were retrotranscribed using PrimeScript RT reagent kit (Takara Bio) in a total volume of 10 µl. Twenty-fold diluted samples were used to perform RT-qPCR; 2 µl in a total volume of 20 µl was analyzed in each well. RT-qPCR was conducted on a StepOnePlus Real-Time PCR System (Life Technologies) using SYBR premix Ex Taq (Tli RNaseH plus) (Takara Bio). Steps used for the chain reaction were: an initial incubation of 10 min at 95°C, followed by 40 recurred cycles of 15 s at 95°C and 1 min at 60°C each. Amplification specificity was assessed both by finding a unique peak in the melting curve and by amplicon size estimation using gel electrophoresis. Actin-like and AGL26-like genes were used as reference genes since they were previously described as suitable for plum expression experiments by Lloret et al. (2017a). The stability of the selected housekeeping genes was analyzed with two different software: BestKeeper (Pfaff et al., 2004) and geNorm (Vandesompele et al., 2002). BestKeeper recommended that reference gene should have a SD value <1.0 to be considered suitable for normalization and herein both genes had an SD value of 0.40. On the other hand, according to geNorm program, genes with an M-value below the threshold of 1.5 were considered stably expressed and the obtained value was M = 0.424. The geometric mean of Ct values obtained for the reference genes was used to normalize the Ct values obtained for each gene analysis in a sample dependent manner. For every amplification experiment, three independent biological samples with two technical replicates each were analyzed. In addition, a relative standard curve was built in order to obtain relative expression values that were averaged to obtain the final results. In the cases where two possible isoforms were noticed (PdoDAM4 and PdoDAM5), differential primers were designed to determine which form was more abundant during bud development. For each timepoint, RT-qPCR using the two possible combination of primers was performed in parallel, as described above, at the same PCR instrument. Efficiency (E) of each primer pair was determined using a relative standard curve and was used to correct Ct values and allow comparison between them. In every case the abundant isoform was taken as a reference and the minoritarian was calculated as a percentage of it. The primers used to amplify each gene are presented in Supplementary Table S5.

**Statistical Analysis**

Experimental values were processed using Statgraphics XVI.I package 324 (Statpoint Technologies) to evaluated the statistics significance. Klustal-Wallis test with a confidence level of 95% was used for comparison of multiple samples. Different letters mean significantly different samples.

**DATA AVAILABILITY STATEMENT**

The RNA-seq data analyzed for this study can be found in the National Center for Biotechnology Information (NCBI) BioProject database ID PRJNA630876 (http://www.ncbi.nlm.nih.gov/bioproject/630876).

**AUTHOR CONTRIBUTIONS**

CQ-T and AL performed RT-qPCR experiments and bioinformatic analyses. BG and JR performed dormancy measurements and microscopic analysis in European plum. MB, JR, GR, and AL conceived and designed the experiments.

Quesada-Traver et al. Characterization of Plum DAM Genes
This research was funded by Instituto Nacional de Investigación y Tecnología Agraria y Alimentaria (INIA)-FEDER (RFP2015-00015-00, RTA2017-00003-00, RTA2017-00011-C03-01) and the Gobierno de Aragón–European Social Fund, European Union (Grupo Consolidado A12_17R). CQ-T was funded by a fellowship co-financed by the European Social Fund and the Instituto Valenciano de Investigaciones Agrarias (IVIA). BG was supported by a fellowship of Consejo Nacional de Ciencia y Tecnología de México (CONACYT, 471839). AL was funded by a fellowship of Ministerio de Ciencia, Innovación y Universidades of Spanish Government.

**SUPPLEMENTARY MATERIAL**

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fpls.2020.01288/full#supplementary-material
Vitasse, Y., Signarbieux, C., and Fu, Y. H. (2018). Global warming leads to more...  

Weinberger, J. H. (1950). Chilling requirements of peach varieties.  

Wang, Q., Xu, G., Zhao, X., Zhang, Z., Wang, X., Liu, X., et al. (2020). Comprehensive cloning of Prunus mume dormancy associated MADS-box genes and their response in flower bud development and dormancy. Front. Plant Sci. 19. doi: 10.3389/fpls.2018.00017  

Zhou, K., Zhou, Y., Ahmad, S., Xu, Z., Li, Y., Yang, W., et al. (2018a).  

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.  

Copyright © 2020 Quesada-Traver, Guerrero, Badenes, Rodrigo, Rios and Lloret. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.