Research Article

Cytotoxic Constituents of the Bark of Hypericum roeperianum towards Multidrug-Resistant Cancer Cells

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1.Introduction

Cancer continues to be a global threat, appearing as the second leading cause of death globally, with estimated 9.6 million deaths representing one in six deaths and with an estimated five-year prevalence of 43.8 million people [1]. Multidrug resistance (MDR) of cancer cells is a serious concern in chemotherapy. It is responsible for many therapeutic failures and high burdens globally, in patients suffering from cancer [2, 3]. Any modern protocol for new cytotoxic drug discovery today should integrate the ability of neoplastic cells to rapidly develop resistant phenotypes. Thus, resistant cell lines should be integrated into the cell panel used for the discovery of more efficient...
substances. The present work has taken this into account and involves several models of MDR cancer cell lines such as the colon adenocarcinoma with p53 knockout phenotype, the leukemia cells with ATP-binding cassette (ABC)-transporter-overexpressing MDR-mediating-P-glycoprotein (P-gp; ABCB1/MDR1), the breast cancer bearing resistance protein (ABCG2/BCRP), and the transfectant glioblastoma multiforme harboring a mutation-activated EGFR gene (ΔEGFR). The effectiveness of natural products in the fight against cancer has been largely demonstrated [4]. Some clinically established cytotoxic drugs such as camptothecin, paclitaxel, vinblastine, or vincristine are naturally occurring compounds [4–6]. In addition, numerous botanicals and phytochemicals derived from African medicinal plants have been found active against MDR cancer cell lines [7, 8]. Some of such prominent phytochemicals include terpenoids: salvimulticol and candesalvone B methyl ester [9], epunctalone [10], and aristasiciropin B [11], phenolics: 2-acetyl-7-methoxynaphtho[2,3-b]furan-4,9quinone [12], 6α-hydroxyphaseollidin [13], licoagrochalcone A [14], 7-dihydroxy-4′-methoxy-6,8diprenylisoflavone, and 7,7′-di-O-methylchamaejasmin [15], and alkaloids: 1,3-dimethoxy-10-methylacridone [16], isotetrandrine [17], and ungeremine [18]. However, more hit compounds should be identified to increase our arsenal of cytotoxic compounds and to secure better the chances of later obtaining new clinically usable molecules. The present study was, therefore, designed to assess the cytotoxicity of botanicals and phytochemicals from the bark of Hypericum roeperianum Schimp. p. ex A. Rich (Guttiferae). The modes of action of compound 1, such as its effects on cell cycle distribution and induction of apoptosis, on caspases activation, and on the production of reactive oxygen species (ROS), were also investigated. Hypericum roeperianum is a shrub or small tree growing in the tropical part of central, eastern, and southern tropical Africa, locally used alone or in association with various plants in the treatment of female sterility [19], as antiabortifacient [20] and as antiinflamatory remedies [21]. Previous phytochemical investigations of this plant led to the isolation of a polyketide, 4-methoxy-3-(2-methylbut-3-en-2-yl)-6-phenyl-2H-pyran-2-one, xanthones: 1,5-dihydroxy-6-methoxyxanthone, 2-hydroxy-5-methoxyxanthone and 1,4,6,7-tetrahydroxyxanthone, and the dihydroxy-6-methoxyxanthone, 2-hydroxy-5-methoxyxanthone and 1,4,6,7-tetrahydroxyxanthone, and the xanthonolignoids: 8,10-dihydroxy-3-(4hydroxy-3,5-dimethoxyphenyl-2-(hydroxymethyl)-2, 3-dihydro-1,4 dioxino[2,3-c]xanthene-7-one and 8-hydroxy-10 methoxy-3-(4-hydroxy-3,5-dimethoxyphenyl-2-(hydroxymethyl)-2,3-dihydro-1,4 dioxino[2,3-c]xanthene-7-one and 8-hydroxy-10 methoxy-3-(4-hydroxy-3,5-dimethoxyphenyl-2-(hydroxymethyl)-2,3-dihydro-1,4 dioxino[2,3-c]xanthene-7-one from the bark [22] and 10 other xanthones from the roots, namely, 5-O-methyl-2-deprenylhediaxanthone B, 5-O-methylisojacareubin, 5-O-demethylpanaxonin, roeperanone, 2-hydroxyxanthone, 5-hydroxy-2-hydroxyxanthone, 1,5-dihydroxy-2-methoxyxanthone, 2-deprenyl rhedixanthone B, isojacareubin, and calycinoxanthone D [23]. The cytotoxicity of botanicals from the bark of Hypericum roeperianum is being reported for the first time.

2. Material and Methods

2.1. Chemicals. Doxorubicin (98.0% purity) from Sigma-Aldrich (Munich, Germany) was obtained from the Johannes Gutenberg University Medical Center (Mainz, Germany). Geneticin >98% (used at 800 ng/mL and 400 μg/mL) in culture media to maintain the features of MDA-MB-231-BCRP, U87MG.ΔEGFR, and HCT116 (p53+/−), respectively, was obtained from Sigma-Aldrich and stored at 72.18 mM. Hydrogen peroxide (H2O2) and valinomycin (at 1 mg/mL) were provided by Sigma-Aldrich (Taufkirchen, Germany).

2.2. Plant Material and Extraction. The bark of Hypericum roeperianum Schimp. p. ex A. Rich (Guttiferae) was collected in Bangang Wabane (South West Region of Cameroon) in October 2018. No permission was necessary for sample’s collection. The identification of the plant was carried out by Dr. Tchiengue Barthelemy at the Cameroon National Herbarium (Yaoundé) where a voucher specimen was deposited under the number 24584/SRF/Cam. Dried barks of the plant (3.0 kg) were powdered and extracted with methanol (MeOH; 3×15 L) for 72 h at room temperature to afford a crude extract (HRB; 150.0 g) after filtration with Whatman paper no.1 and evaporation in vacuum, under reduced pressure. A portion of the resulting extract (140.0 g) was, then, exhausted in ethyl acetate to yield 65.0 g of the ethyl acetate extract (EtOAc) (HRBa) and the residue (HRBb; 75 g).

2.3. Fractionation and Purification of the Bark Extract of Hypericum roeperianum. Part of the ethyl acetate extract (EtOAc; 60.0 g) was submitted to silica gel flash chromatography using hexane-EtOAc and, then, EtOAc-MeOH mixtures of increasing polarities. Eighty fractions (frs) of 500 mL each were collected as follows: hexane 100% (sub-frs 1–3), hexane-EtOAc 90:10 (sub-frs 4–12), hexane-EtOAc 80:20 (sub-frs 13–18), hexane-EtOAc 70:30 (sub-frs 19–22), hexane-EtOAc 60:40 (sub-frs 23–27), hexane-EtOAc 50:50 (sub-frs 28–37), hexane-EtOAc 40:60 (sub-frs 38–43), AcOEt 100% (sub-frs 44–52), AcOEt-MeOH 95:5 (sub-frs 53–57), AcOEt-MeOH 90:10 (sub-frs 58–62), EtOAc-MeOH 80:20 (sub-frs 63–69), and MeOH 100% (sub-frs 70–80). Based on their analytical thin-layer chromatography (TLC) profiles, these fractions were pooled into five fractions (frs) as follows: HRBa1 (Sub-frs 1–15, 8.0 g), HRBa2 (Sub-frs 16–25, 15.0 g), HRBa3 (Sub-frs 26–38, 9.5 g), HRBa4 (Sub-frs 39–68, 12.5 g), and HRBa5 (Sub-frs 69–80, 13.0 g).

Dry fraction HRBa2 (15.0 g) was dissolved in methanol affording a nonsoluble powder which was, then, filtered to give compound 1 (15 mg). The filtrate was subjected to silica gel column chromatography using hexane-AcOEt mixtures of increasing polarities as elution solvents. Sixty-five sub-fractions of 150 mL each were collected as follows: Hex 100% (1–3), Hex-AcOEt 90:10 (Sub-frs 4–9), Hex-AcOEt 80:20 (Sub-frs 10–15), Hex-AcOEt 70:30 (Sub-frs 16–19), Hex-AcOEt 60:40 (Sub-frs 20–35), Hex-AcOEt 50:50 (Sub-frs 36–42), Hex-AcOEt 40:60 (Sub-frs 43–50), AcOEt 100% (Sub-frs 51–55), AcOEt-MeOH 90:10 (Sub-frs 56–60), and MeOH 100% (Sub-frs 61–65). Sub-frs 6–9 yielded compound 2.
(15.0 mg) as a white powder. Sub-frs EC23-29 yielded compound 3 (12.0 mg) as a yellow powder.

HRBa3 (9.5 g) was subjected to silica gel column chromatography using Hex-AcOEt mixtures of increasing polarities as elution solvents. Seventy-five sub-frs of 150 mL each were collected as follows: hexane 100% (sub-frs 1–4), Hex-AcOEt 90:10 (sub-frs 5–9), Hex-AcOEt 80:20 (sub-frs 9–16), Hex-AcOEt 70:30 (sub-frs 17–23), Hex-AcOEt 60:40 (sub-frs 24–35), Hex-AcOEt 50:50 (sub-frs 36–42), Hex-AcOEt 40:60 (sub-frs 43–47), AcOEt 100% (sub-frs 48–57), AcOEt-MeOH 90:10 (sub-frs 58–63), AcOEt-MeOH 80:20 (sub-frs 64–70), and MeOH 100% (sub-frs 71–75). Sub-frs 22–25 yielded compound 4 (18.0 mg) as a green-yellowish powder. Sub-frs 29–35 yielded compound 5 as a yellow powder (15.0 mg).

HRBa4 (12.5 g) was subjected to silica gel column chromatography using CH2Cl2-MeOH mixtures of increasing polarities as elution solvents. Fifty sub-frs of 150 mL each were collected as follows: CH2Cl2 100% (sub-frs 1–4), CH2Cl2-MeOH 95:5 (sub-frs 5–13), CH2Cl2-MeOH 90:10 (sub-frs 14–25), CH2Cl2-MeOH 85:15 (sub-frs 26–35), CH2Cl2-MeOH 80:20 (sub-frs 36–42), and MeOH 100% (sub-frs 43–50). Sub-frs 6–11 yielded compound 7 (12.0 mg) as a white powder. Sub-frs 13–15 yielded compound 6 (40.0 mg) as a white powder. Sub-frs 13–15 yielded compound 7 (12.0 mg) as a yellow powder. Sub-frs 19–23 yielded compound 8 (14.0 mg) as a yellow powder.

2.4. Cell Cultures. Cell lines used in this work included drug-sensitive and drug-resistant phenotypes of earlier reported origin. They were all provided by Prof. Dr. Thomas Efferth from his cell lines collection; they have being used in cytotoxicity screening by our team for a decade [12–21]. These include two hematological cancer cell lines, namely, the drug-sensitive CCRF-CEM leukemia cell line and its multidrug-resistant P-gp-over-expressing CEM/ADR5000 cells [24–26] and nine carcinoma cell lines, namely, U87MG glioblastoma cell line and its EGFRT20 mutant U87.MGΔEGFR subline, HCT116 (p53<sup>-/-</sup>) colon cancer cell line and its knockout clone HCT116 (p53<sup>−/−</sup>), and MDA-MB-231-pcDNA3 breast cancer cell line and its BCRP-transfected multidrug-resistant MDA-MB-231-BCRP clone 23 cell line [27], as well as the normal AML12 hepatocytes, used to compare with HepG2 liver cancer cells [13].

2.5. Resazurin Reduction Assay (RRA) for Cell Growth Evaluation. The RRA was applied to evaluate the cytotoxicity of botanicals, the isolated phytochemicals (1–5, 7, and 8), and doxorubicin on the cell growth as reported earlier [18, 28]. Cells treated with various samples at different concentrations were incubated for 72 h in humidified 5% CO2 atmosphere at 37°C. Cells were further coloured with resazurin and incubated for 1–2 h; the fluorescence was further measured with an Infinite M2000 Pro™ plate reader (Tecan), as reported previously [13].

2.6. Flow Cytometric Evaluation of Cell Cycle Distribution and Apoptotic Cells. Various concentrations of botanical HRB, phytochemical 1, and doxorubicin or DMSO (solvent control) were used to treat CCRF-CEM cells (1 × 10<sup>6</sup> cells). Cells were further incubated for 24 h in humidified 5% CO2 atmosphere at 37°C and analyzed using a BD Accury C6 Flow Cytometer (BD Biosciences, Heidelberg, Germany) by measuring the propidium iodide fluorescence of individual nucleus, as described earlier [10, 11]. Experiments were conducted thrice independently with three parallel measurements.

2.7. Assessment of Apoptosis by Annexin V/PI Staining. The CCRF-CEM cells (1 × 10<sup>6</sup>; 1 ml) were also treated with HRB, compound 1 and doxorubicin for 24 h (in humidified 5% CO2 atmosphere at 37°C), and apoptosis was further assessed by flow cytometry using the fluorescein isothiocyanate-conjugated annexin V/PI assay kit (eBioscience Annexin V; Invitrogen, San Diego, USA), as previously published [10, 11]. Briefly, treated cells were centrifuged at 1200 rpm for 5 min, then washed twice with ice-cold PBS, resuspended in 500 µl binding buffer, and stained with 5 µl FITC-conjugated annexin V (10 mg/mL) and 10 µl PI (50 mg/mL). After 15 min incubation at room temperature (RT) in the dark, cells were analyzed using a BD Accury C6 Flow Cytometer (BD Biosciences). Cells stained with only annexin V were evaluated as being in early apoptosis. Cells stained with both annexin V and propidium iodide were evaluated as being in late apoptosis or in a necrotic stage.

2.8. Evaluation of Caspases Activities Using Caspase-Glo 3/7, Caspase-Glo 8, and Caspase-Glo 9. Different concentrations of HRB and compound 1 were used to treat CCRF-CEM cells for 6 h. The activities of caspases were determined using Caspase-Glo 3/7, Caspase-Glo 8, and Caspase-Glo 9 Assay kits (Promega, Mannheim, Germany) by measuring the luminescence using an Infinite M2000 Pro™ plate reader (Tecan), as reported previously [13].

2.9. Evaluation of Reactive Oxygen Species (ROS) Production. Various concentrations of HRB and triterpenoid 1 were used to treat CCRF-CEM cells (1 × 10<sup>6</sup> cells); DMSO (solvent control); or hydrogen peroxide (H2O2; positive control). After 24 h incubation in humidified 5% CO2 atmosphere at 37°C, the production of ROS was evaluated using 2′,7′-dichlorodihydrofluorescein diacetate (H2DCFH-DA) (Sigma-Aldrich) staining, as described earlier [30–32].

2.10. Statistics. Statistical analyses were performed with Graph pad prism 5 software. Representative data from three independent experiments are shown as mean value ± S.E.M. One-way Analysis Variance (ANOVA) followed by post hoc Tukey’s test was used to determine the significance of the
difference between mean values relative to the control. The \( p \) value was calculated to determine significant differences (\( p \) value < 0.05).

3. Results

3.1. Phytochemistry. The chemical structures of the isolated phytochemicals were determined by exploiting the physical, mass spectra, and NMR data, followed by direct comparison of these data with those of similar reported compounds in the literature. Compounds were identified as trichadonic acid C\(_{30}\)H\(_{48}\)O\(_{3}\) (I; white amorphous powder; \( m/z \) 456) [33], fri-delan-3-one C\(_{26}\)H\(_{26}\)O (2; white powder; m.p. 258°C; \( m/z \) 426) [33], 2-hydroxy-5-methoxyxanthone C\(_{14}\)H\(_{12}\)O\(_{4}\) (3; yellow amorphous powder; \( m/z \) 242) [34], 1,3,6,7-tetrahydroxyxanthone or norathryiol C\(_{13}\)H\(_{8}\)O\(_{6}\) (4; green-yellowish powder; m.p. 271°C; \( m/z \) 260) [35], 1,3,5,6-tetrahydroxyxanthone C\(_{13}\)H\(_{14}\)O\(_{6}\) (5; yellow powder; m.p. 136°C; \( m/z \) 260) [36], betulenic acid C\(_{23}\)H\(_{40}\)O\(_{3}\) (6; white powder; m.p. 318°C; \( m/z \) 456) [33], 3′-hydroxymethyl-2′-(4′-hydroxy-3′,5′-dimethoxyphenyl)-5′,6′,5,6′-(6,8-dihydroxyxanthone)-1′,4′-dioxane C\(_{24}\)H\(_{20}\)O\(_{10}\) (7; yellow powder; m.p. 264°C; \( m/z \) 436) [37, 38], and 3′,4′-hydroxymethyl-2′-(4′-hydroxy-3′,5′-dimethoxyphenyl)-5′,6′,5,6′-(xanthone)-1′,4′-dioxane C\(_{24}\)O\(_{10}\) (8; yellow amorphous powder; \( m/z \) 468) [37] (Figure 1). The 1D NMR spectra of these compounds are provided as Supplementary Materials.

3.2. Cytotoxicity of Phytochemicals and Doxorubicin. The cytotoxicity of crude extracts, fractions, and phytochemicals 1–5, 7, 8, and doxorubicin was investigated using RRA towards 9 cancer cell lines and normal AML12 hepatocytes (Tables 1 and 2). The degree of resistance (D.R.) of the tested samples was determined as the ratio of the IC\(_{50}\) value of the resistant cell line divided by that of the corresponding parental sensitive cell line (Tables 1 and 2). Collateral sensitivity or hypersensitivity was deduced if the D.R. was below 1 while normal sensitivity was defined as a D.R. of 1 or around 1. cross resistance was considered if the D.R. was below 1 while normal sensitivity was calculated as the ratio of the IC\(_{50}\) value in normal AML12 hepatocytes by the corresponding values in HepG2 hepatocarcinoma cells (Tables 1 and 2).

The obtained IC\(_{50}\) values ranged from 11.43 \( \mu \)g/mL (against the P-gp-overexpressing CEM/ADR5000 leukemia cells) to 26.75 \( \mu \)g/mL (against HCT116 (p53\(^{−/−}\)) colon adenocarcinoma cells) for the crude extract HRB, from 15.65 \( \mu \)g/mL (against CEM/ADR5000 leukemia cells) to 41.17 \( \mu \)g/mL (against HCT116 (p53\(^{−/−}\)) cells) for HRBa, from 13.92 \( \mu \)g/mL (against U87MG.ΔEGFR glioblastoma cells) to 33.44 \( \mu \)g/mL (against HCT116 (p53\(^{−/−}\)) cells) for HRBb, from 16.13 \( \mu \)g/mL (against U87MG glioblastoma cells) to 33.63 \( \mu \)g/mL (against HCT116 (p53\(^{−/−}\)) cells) for HRBa3, from 10.52 \( \mu \)g/mL (against U87MG.ΔEGFR cells) to 28.43 \( \mu \)g/mL (against HCT116 (p53\(^{−/−}\)) cells) for HRBa4, and from 28.30 \( \mu \)g/mL (against U87MG.ΔEGFR cells) to 69.48 \( \mu \)g/mL (against HepG2 cells) for HRBb (Table 1). Fractions HRBa1 and HRBa5 had selective activities (Table 1).

Triterpenoid 1, xanthone 5, and doxorubicin displayed cytotoxic effects towards the 9 tested cancer cell lines with IC\(_{50}\) values ranging from 14.44 \( \mu \)M (against CCRF-CEM cells) to 44.20 \( \mu \)M (against the resistant HCT116 (p53\(^{−/−}\)) cells) for I, from 38.46 \( \mu \)M (against CEM/ADR5000 cells) to 112.27 \( \mu \)M (against HCT116 (p53\(^{−/−}\)) cells) for \( p < 0.05 \) induced late apoptosis (Q2-UR) while compound 1 significantly \( p < 0.05 \) induced early apoptosis with, respectively, 14.6% annexin V (+)/PI (−) and 52.5% annexin V (+)/PI (+) cells (Figure 3).
**Figure 1**: Chemical structures of phytochemicals isolated from the bark methanol extract of *Hypericum roeperianum*. 1: Trichadonic acid; 2: fridelan-3-one; 3: 2-hydroxy-5-methoxyxanthone; 4: 1,3,6,7-tetrahydroxanthone or norathyriol; 5: 1,3,5,6-tetrahydroxyxanthone; 6: betulenic acid; and 7: 3′-hydroxymethyl-2′-(4′-hydroxy-3′,5′-dimethoxyphenyl)-5′,6′,6-(6,8-dihydroxyxanthone)-1′,4′-dioxane; 8: 3′-hydroxymethyl-2′-(4′-hydroxy-3′,5′-dimethoxyphenyl)-5′,6′,5,6-(xanthone)-1′,4′-dioxane.

**Table 1**: Recorded IC$_{50}$ values after 72 h incubation of drug-sensitive and MDR cancer cell lines with botanicals from the bark of *Hypericum roeperianum*.

| Cell lines | Samples, IC$_{50}$ values in μg/mL and degrees of resistance* or selectivity index** |
|------------|--------------------------------------------------------------------------------------|
| CCRF-CEM   | HRB | HRBa | HRBa1 | HRBa2 | HRBa3 | HRBa4 | HRBa5 | HRBb |
| CEM/ADR5000| 13.71 ± 0.26 | 15.65 ± 1.18 | 46.42 ± 1.98 | 17.87 ± 1.53 | 16.81 ± 4.02 | 16.27 ± 4.28 | 57.15 ± 3.76 | 48.76 ± 2.83 |
| MDA-MB-231-pcDNA | 11.43 ± 0.88 | 18.56 ± 1.74 | >80 | 20.18 ± 1.76 | 23.03 ± 1.22 | 13.55 ± 0.44 | >80 | 31.55 ± 2.28 |
| Degree of resistance* | 0.84 | 1.19 | >1.72 | 1.13 | 1.37 | 0.83 | >1.16 | 0.65 |
| HCT16 (p53+/−) | 19.87 ± 1.77 | 23.12 ± 1.65 | 72.23 ± 4.54 | 20.42 ± 1.71 | 28.32 ± 1.57 | 18.13 ± 2.08 | 68.91 ± 6.30 | 48.12 ± 1.99 |
| Degree of resistance | 0.92 | 1.14 | 0.87 | 0.97 | 0.86 | 0.97 | 1.08 | 0.77 |
| HCT16 (p53+/−) | 18.22 ± 0.81 | 26.42 ± 3.01 | 63.16 ± 3.30 | 19.89 ± 0.62 | 24.41 ± 1.77 | 17.66 ± 1.11 | 74.32 ± 3.92 | 37.23 ± 4.51 |
| Degree of resistance | 0.65 | 1.19 | 0.74 | 1.09 | 0.99 | 0.95 |  |
| U87MG | 12.42 ± 0.66 | 18.03 ± 1.77 | 47.84 ± 4.36 | 18.01 ± 1.19 | 16.13 ± 2.04 | 18.44 ± 2.11 | 47.89 ± 2.24 | 46.55 ± 2.43 |
| Degree of resistance | 1.26 | 1.11 | 1.25 | 0.77 | 1.16 | 0.84 | 1.15 | 0.60 |
| HepG2 | 14.36 ± 1.48 | 20.12 ± 0.81 | 59.60 ± 5.12 | 13.92 ± 2.07 | 18.72 ± 1.45 | 15.52 ± 0.88 | 55.20 ± 4.79 | 28.30 ± 1.11 |
| Degree of resistance | 1.19 | 1.12 | 1.25 | 0.77 | 1.16 | 0.84 | 1.15 | 0.60 |
| AML12 | 17.45 ± 2.05 | 29.17 ± 1.30 | >80 | 18.75 ± 1.41 | 25.88 ± 1.56 | 14.95 ± 1.34 | >80 | 69.48 ± 2.93 |
| Selectivity index** | 2.41 | >2.74 | 3.27 | 2.17 | 4.56 |  |

(*): the degree of resistance was determined as the ratio of the IC$_{50}$ value in the resistant divided by the IC$_{50}$ in the sensitive cell line; CEM/ADR5000, MDA-MB-231-BCRP, HCT16 (p53+/−), and U87MGΔEGFR were used as the corresponding resistant counterpart for CCRF-CEM, MDA-MB-231-pcDNA, HCT16 (p53+/−), and U87MG, respectively; (**): the selectivity index was determined as the ratio of the IC$_{50}$ value in the normal AML12 hepatocytes divided by the IC$_{50}$ in HepG2 hepatocarcinoma cells; in bold: significant cytotoxic effect [7, 39, 40]; nd: not determined; HRB: crude methanol extract from the bark of *Hypericum roeperianum*, HRBa: portion obtained by exhaustion of HRB with ethyl acetate; HRBa1-5: fractions from HRBa; HRBb: residual fraction obtained after exhaustion of HRB with ethyl acetate. The data for doxorubicin used as positive control in similar experimental conditions are shown in Table 2.
Table 2: Recorded IC₅₀ values following RRA for phytochemicals isolated from the bark of *Hypericum roeperianum* and reference drug, doxorubicin, towards drug-sensitive, MDR cancer cells lines and hepatocytes after 72 h incubation.

| Cell lines       | Samples, IC₅₀ values in µM, and degrees of resistance* or selectivity index** | Doxorubicin |
|------------------|--------------------------------------------------------------------------------|-------------|
|                  | 1                          | 3           | 4            | 5              | 7            | 8            |                     |
| CCRF-CEM         | 14.44 ± 0.53               | 16.80 ± 0.96 | 19.94 ± 2.12 | 38.58 ± 2.11   | 23.28 ± 1.46 | 16.31 ± 2.12 | 0.02 ± 0.00        |
| CEM/ADR5000      | 18.27 ± 1.56               | 52.95 ± 3.08 | 23.21 ± 1.66 | 38.46 ± 4.07   | 54.04 ± 4.38 | 43.47 ± 2.97 | 122.96 ± 10.94     |
| MDA-MB-231-pcDNA | 16.47 ± 0.74               | 43.80 ± 3.47 | >153.85      | 75.15 ± 4.88   | 20.73 ± 1.32 | 36.89 ± 2.73 | 0.13 ± 0.01        |
| MDA-MB-231-BCRP  | 14.95 ± 1.17               | 33.60 ± 1.99 | 20.38 ± 1.17 | 62.94 ± 5.32   | 22.16 ± 2.88 | 30.50 ± 1.88 | 0.79 ± 0.08        |
| Degree of resistance* | 1.26                      | 3.15        | 1.16         | 1.00           | 2.32         | 2.66         | 6.683.00           |
| HCT116 (p53⁴⁺)   | 17.36 ± 1.84               | 46.67 ± 3.38 | 40.17 ± 3.09 | 75.48 ± 6.10   | >91.74       | 37.79 ± 2.92 | 0.48 ± 0.06        |
| Degree of resistance | 0.91                      | 0.77        | <0.13        | 0.84           | 1.07         | 0.83         | 6.14              |
| HCT116 (p53³⁻)   | 44.20 ± 3.21               | >165.29      | >153.85      | 112.27 ± 8.49  | >91.74       | >85.47       | 1.78 ± 0.08        |
| U87MG            | 16.16 ± 1.09               | 74.44 ± 4.75 | 106.00 ± 6.74 | 61.42 ± 3.39   | 29.70 ± 1.77 | 35.50 ± 3.28 | 0.26 ± 0.03        |
| U87MGΔEGFR       | 14.69 ± 1.55               | 44.98 ± 5.22 | 30.37 ± 2.91 | 59.04 ± 6.01   | 12.72 ± 0.75 | 30.61 ± 3.14 | 0.98 ± 0.07        |
| Degree of resistance | 0.91                      | 0.60        | 0.29         | 0.96           | 0.43         | 0.86         | 3.79              |
| HepG2            | 21.68 ± 3.18               | 44.21 ± 2.65 | 32.40 ± 3.72 | 64.73 ± 5.77   | 25.19 ± 1.69 | 31.29 ± 1.19 | 4.56 ± 0.48        |
| AML12            | 47.34 ± 0.81               | >165.29      | 45.35 ± 3.52 | 150.02 ± 7.03  | 20.89 ± 1.17 | >85.47       | 52.90 ± 4.09       |
| Selectivity index** | 2.18                      | >3.74       | 1.40         | 2.32           | 0.83         | >2.73        | 11.59             |

(*) the degree of resistance was determined as the ratio of the IC₅₀ value in the resistant divided by the IC₅₀ in the sensitive cell line; CEM/ADR5000, MDA-MB-231-BCRP, HCT116 (p53⁴⁺), and U87MGΔEGFR were used as the corresponding resistant counterparts for CCRF-CEM, MDA-MB-231-pcDNA, HCT116 (p53³⁻), and U87MG, respectively; (**): the selectivity index was determined as the ratio of the IC₅₀ value in the normal AML12 hepatocytes divided by the IC₅₀ in HepG2 hepatocarcinoma cells; in bold: significant cytotoxic effect [7, 39, 40], the cytotoxicity of compound 6 (betulenic acid) on these cell lines was previous reported [41], and this compound was no more tested in this study, no IC₅₀ value was recorded at up to 100 µM with compound 2; nd: not determined; 1: trichadonic acid; 2: 2-hydroxy-5-methoxyxanthone; 4: 1,3,6,7-tetrahydroxanthone or norathryiol; 5: 1,3,5,6-tetrahydroxanthone; 7: 3′-hydroxymethyl-2′-(4′-hydroxy-3′,5′-dimethoxyphenyl)-5′,6′-dihydroxanthone)-1′,4′-dioxane; and 8: 3′-hydroxymethyl-2′-(4′-hydroxy-3′,5′-dimethoxyphenyl)-5′,6′,3′,6′-dihydroxanthone)-1′,4′-dioxane.

Figure 2: Distribution of the CCRF-CEM cells cycle after treatment with the crude extract (HRB), compound 1 (trichadonic acid), and doxorubicin. IC₅₀ values were 14.44 µM for trichadonic acid and 0.02 µM for doxorubicin.
3.4. Activation of Caspases and Production of ROS. In the presence of the tested samples, the activity of caspases in CCRF-CEM cells increased by 1.18-fold, 1.45-fold, and 1.35-fold for HRB and by 2.52-fold, 2.62-fold, and 2.23-fold for trichadonic acid (1), respectively, for caspases 3/7, 8, and 9 (Figure 4).

The production of reactive oxygen species (ROS) in CCRF-CEM cells treated with HRB, triterpenoid 1, H$_2$O$_2$ (positive control), or DMSO was analyzed, and the results are depicted in Figure 5. The crude extract HRB significantly ($p < 0.05$) induced increase of ROS production in a range of 9.89% (1/4×IC$_{50}$) to 71.92% (2×IC$_{50}$); compound 1 also significantly ($p < 0.05$) induced increase of ROS production in a range of 12.35% (7.22 µM) to 68.12% (57.76 µM). The reference compound, H$_2$O$_2$, increased the ROS levels to 94.30% at 50 µM, while ROS production in nontreated cells was 0.6%.

### 4. Discussion

Taking into account the rapid development of resistance by cancer cell lines, the use of MDR phenotypes when screening phytochemicals is an interesting approach. Collateral or normal sensitivity (D.R. below or equal to 1) of MDR cells to phytochemicals combined to their good cytotoxicity could be better criteria to select substances for clinical studies. In the present work, four MDR cells lines, CEM/ADR5000 cells, MDA-MB-231-BCRP cells, HCT116 (p53$^{-/}$) cells, and U87.MGΔEGFR cells, were used, and their susceptibilities to isolated phytochemicals were compared with those of their parental sensitive counterparts, CCRF-CEM cells, MDA-MB-231 cells, HCT116 (p53$^{+/}$) cells, and U87.MG cells, respectively (Tables 1 and 2). Interestingly, collateral sensitivity of CEM/ADR5000 cells, BCRP-expressing MDA-MB-231 cells, and HCT116 (p53$^{-/}$) cells to HRB was achieved, as well as the hypersensitivity of all resistant cell lines to fraction HRBa4 and HRBb compared to their sensitive parental cell lines (Table 1). Collateral sensitivity of BCRP-expressing MDA-MB-231 cells and U87MGΔEGFR cells to phytochemicals 1, 3–5, and 8 was observed, suggesting that they might be useful to fight drug resistance in breast cancer and glioblastoma (Table 2). This clearly indicates that these botanicals and phytochemicals can be exploited in the fight against recalcitrant cancers. The IC$_{50}$ values below 20µg/mL or below 10µM after incubation between 48 and 72h have been set for promising cytotoxic botanicals and phytochemicals, respectively [39, 40]. Importantly, IC$_{50}$ values below 20µg/mL were obtained with HRB against 8/9 tested cancer cell lines, HRBa2 and HRBb4 against 7/9 cancer cell lines, HRBa8 against 4/9 cancer cell lines, and HRBa3 against 3/9 cell lines (Table 1). It can, therefore, be confirmed that these botanicals are interesting cytotoxic agents. However, IC$_{50}$ values below the established threshold were not achieved with phytochemicals, though terpenoid 1 and xanthone 5 had cytotoxic effects towards the 9 tested cancer cell lines. However, their good selectivity indexes still suggest that they can still be good candidates to tackle cancers, especially when drug resistance is observed. To the best of our knowledge, the cytotoxicity of the crude extract and compounds 1, 2, 3, 7, and 8 on the studied cell lines is being reported, herein, for the first time. Betulinic acid (6) is a well-known cytotoxic compound and has previously been found active towards the cancer cell lines tested in the present work, with IC$_{50}$ values ranging from 7.65 µM (in CEM-ADR5000 cells) to 44.17 µM (in HepG2 cells) [41]. Although it was not further tested, herein, compound 6 can be ranked amongst the best active principles of Hypericum roeperianum. Also, the cytotoxicity of norathyriol (4) in JB6 P+ mouse skin epidermal cells was reported [42]. 1,3,5,6-Tetrahydroxyxanthone (5) had low
cytotoxic effects against K562 leukemia cells with 12.98 µg/mL (49.92 µM) [43].

Apoptosis is a programmed cell death and is also the most investigated mechanism of action of antiproliferative drugs. In this study, it was found that both HRB and compound 1 induced apoptosis in CCRF-CEM cells with cell cycle arrest in the Go/G1 phase (Figures 2 and 3). Modulation of caspases activities is one of the events observed in the apoptotic process in cancer cell lines [44], making these enzymes a target for cytotoxic drug discovery [8, 45]. However, no significant increase in the activity of initiator caspases (caspases 8 and 9) or in that of the activator caspases (caspase 3/7) was observed (Figure 4). Phytochemical 1 induced 2.52-fold, 2.62-fold, and 2.23-fold increase of the activity of caspases 3/7, 8, and 9, respectively (Figure 4), suggesting that this molecule is a caspase modulator. Botanical HRB and compound 1 were also shown to induce increase of ROS by up to 71.92% and 68.12%, respectively (2 × IC50; Figure 5); this is an indication that one of the modes of action of this triterpenoid also includes the enhancement of ROS production in cancer cells.

Regarding the structure-activity relationship, it appears that pentacyclic triterpene 1 is different from 2 by the presence of the carboxyl group (–COOH) in C-13 (Figure 1); the presence of this –COOH group significantly enhanced the cytotoxic activity of triterpene 1, with IC50 values ranging from 14.44 µM to 44.20 µM in cancer cells tested whilst no IC50 value at up to 100 µM was recorded with triterpene 2 (Table 1). Betulic acid, another pentacyclic triterpene with –COOH in C-17, previously displayed good cytotoxicity against all cancer cell lines tested in this work [41], illustrating the importance of the carboxyl function in the cytotoxicity of pentacyclic triterpenes. Concerning xanthones, though 5 was active in all tested cancer cell lines,
3-5 displayed moderate activities (Table 1). It was previously shown that additional hetrocycle in xanthone combined to prenylation improved the cytotoxicity of xanthones, with cudraxanthone 1 (additional hetrocycle combined to -C8- prenylation) displaying significant cytotoxic effects (IC50 value below 10 µM) against all cancer cell lines tested in the present study [46]. This observation is also confirmed with another prenylated xanthone bearing additional herocycle, xanthone V1 [47]. The difference between the two xanthanolignoids 7 and 8 is the presence of two hydroxyl (-OH) groups in C-6 and C-7 (Compound 8) (Figure 1). This difference seems to influence the selectivity, as compound 8 was active against HCT116 (p53+/+), with an IC50 value of 37.79 µM compared to the IC50 value above 91.74 µM obtained for compound 7 (against the same cell line (Table 1).

5. Conclusions

The present work demonstrated that Hypericum roeperianum is a source of cytotoxic compounds. Terpenoids such as trichadonic acid (1) and betulinic acid (6), xanthones (2-hydroxy-5-methoxyxanthone (3), norathylriol (4), and 1,3,5,6-tetrahydroxyxanthone (5)), and xantholignoids (3′-hydroxymethyl-2′-(4′-hydroxy-3″,5″-dimethoxyphenyl)-5′,6′,5,6-(6,8-dihydroxyxanthone)−1′,4′-dioxide (7) and 3′-hydroxymethyl-2′-(4′-hydroxy-3″,5″-dimethoxyphenyl)-5′,6′,5,6-(xanthone)-1′,4′-dioxide (8)) are amongst the active constituents of this plant. Trichadonic acid (1) induced apoptosis in CCRF-CEM leukemia cells, through caspases activation and enhancement of ROS production. The crude extract, HRB, also induced apoptosis in CCRF-CEM cells, mediated by enhancement of ROS production. These compounds can potentially be useful in the fight against recalcitrant cancers.

Abbreviations
1: Trichadonic acid
2: Fidelan-3-one
3: 2-Hydroxy-5-methoxyxanthone
4: 1,3,6,7-Tetrahydroxyxanthone or norathylriol
5: 1,3,5,6-Tetrahydroxyxanthone
6: Betulenic acid
7: 3′-Hydroxymethyl-2′-(4′-hydroxy-3″,5″-dimethoxyphenyl)-5′,6′,5,6-(6,8-dihydroxyxanthone)-1′,4′-dioxide
8: 3′-Hydroxymethyl-2′-(4′-hydroxy-3″,5″-dimethoxyphenyl)-5′,6′,5,6-(xanthone)-1′,4′-dioxide; ABC, ATP-binding cassette; BCRP, breast cancer resistance protein; DMSO, dimethylsulfoxide; D.R., resistance; EGFR, epidermal growth factor receptor; FITC, fluorescein isothiocyanate; H2O2, hydrogen peroxide; H2DCFH-DA, 2′,7′-dichlorodihydrofluorescein diacetate; HRB, methanol extract of the bark of Hypericum roeperianum; HRBa, portion obtained by exhaustion of HRB with ethyl acetate; HRBα1-5, fractions from HRBa; HRBβ, Residual fraction obtained after exhaustion of HRB with ethyl acetate; IC50, 50% inhibitory concentration; MDR, multidrug resistance; PBS, phosphate buffer saline; P-gp, P-glycoprotein; PI, propidium iodide; and ROS, reactive oxygen species.

Data Availability

All data generated or analyzed during this study are included in this published article and its supplementary information files.

Conflicts of Interest
The authors declare that they have no conflicts of interest.

Authors’ Contributions

SBT, FD, M-GGF, ATM, and VK carried out the experiments; IC recorded NMR data; FD, GTMB, and JDSM elucidated the chemical structures; ATM and VK wrote the manuscript; and all authors read and approved the final manuscript.

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Supplementary Materials

Additional File: S1: NMR spectra and main chemical shifts of the isolated compounds; S2: diagrams of annexin V/PI staining of CCRF-CEM with HRB and trichadonic acid. (Supplementary Materials)

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