A rapid and sensitive assay for quantification of siRNA efficiency and specificity

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Submitted: October 14, 2004; Revised: December 21, 2004; Accepted: December 23, 2004.

Indexing terms: Thymosin beta(4); Thymosin beta(10).

ABSTRACT

RNA Interference has rapidly emerged as an efficient procedure for knocking down gene expression in model systems. However, cross-reactivity, whereby multiple genes may be simultaneously targeted by a single short interfering RNA (siRNA), can potentially jeopardize correct interpretation of gene function. As such, it is essential to test the specificity of a siRNA prior to a full phenotypic analysis. To this end, we have adapted a reporter-based assay harnessing the sensitivity of luciferase activity to provide a quantitative readout of relative RNAi efficacy and specificity. We have tested different siRNAs directed against Thymosin β4 (Tβ4); determined their effectiveness at silencing Tβ4 and have both excluded off-target silencing of the Tβ4 homologue Thymosin β10 (Tβ10) and demonstrated partial knockdown of Tβ10 despite significant (12/23; 52%) sequence mismatch. This assay system is applicable to any RNAi study where there is a risk of targeting homologous genes and to the monitoring of off-target effects at the genome level following microarray expression profiling.

INTRODUCTION

The ability to selectively abolish expression of a single gene and to examine the resulting cellular phenotype has enabled substantial progress in the understanding of gene function. Despite the reported problems of cross-reactivity (1) and induction of an interferon response (2), RNA interference (RNAi) has rapidly become the preferred technique for abolishing gene expression.

RNAi, which occurs endogenously in eukaryotes, is initiated by the cleavage of long dsRNA into 21-23 nucleotide siRNA, by the RNase III endonuclease, Dicer. The siRNAs are incorporated into a multisubunit RNA-induced silencing complex (RISC), which targets complementary mRNA for degradation.

The extent of target specificity offered by RNAi in mammalian cells is still highly controversial and is almost certainly sequence-dependent. Many groups report a high degree of specificity, such as the ability of siRNAs to discriminate between wild type and mutant p53, differing by a single nucleotide, in H1299 and U20S cells (3). Genome-wide microarray studies have attempted to resolve the matter, but have proven equally inconclusive (1, 4-6). Specific silencing was observed when an exogenously expressed protein, EGFP, was targeted in HEK 293 cells (5). However, a conflicting study in HeLa cells demonstrated that RNAi targeting two different genes, IGF1R and MAPK14, resulted in silencing of non-targeted genes containing as few as eleven nucleotides of identity (1). Disparity between these two data sets could be explained by the fact that EGFP is not an endogenous mammalian gene and, therefore, siRNAs directed towards IGF1R and MAPK14 would be expected to share sequence homology with a greater number of mammalian genes than a siRNA to target EGFP and carry a greater risk of off-target
silencing. Either way it appears that there are intrinsic limits on the specificity of RNAi. In a recent study examining 359 published siRNA sequences, it was found that approximately 75% have a risk of non-specific targeting (7). Furthermore, siRNAs can elicit non-specific effects at the level of protein synthesis without a corresponding effect on mRNA levels (8) in a manner analogous to that observed with micro RNAs (miRNAs) (9-10).

Clearly, so-called off-target gene silencing could represent a significant problem, resulting in RNAi adversely targeting multiple genes simultaneously. It is, therefore, essential to determine which genes, if any, share significant sequence identity with the proposed siRNA and to directly assess whether their expression is altered by the siRNA.

Our RNAi studies have been based upon the targeting of the actin binding protein Thymosin beta 4 (Tβ4). We have encountered significant difficulties, such as probe cross-reactivity and non-specific product detection, using conventional methods, in assessing the specificity of Tβ4 knockdown as against a non-specific effect on its closest relative Thymosin beta 10 (Tβ10). This is a potential problem associated with detecting expression levels following RNAi of many similarly sized, highly homologous genes and/or members of the same gene family.

In this study we present a reporter-based assay system to test both the efficacy and specificity of Tβ4 knockdown, which can be applied in general to other siRNAs. We have placed target sequences for either Tβ4 or Tβ10 downstream of a luciferase gene and following co-transfection of the relevant siRNA assessed specific knockdown via a quantitative reduction in luciferase activity.

**MATERIALS AND METHODS**

**Construction of Tβ4 shRNAs**

Tβ4 shRNA constructs were prepared by modifying a RasGAP shRNA transgene (11), kindly provided by G.Gish, which uses the human H1 RNA polymerase III to drive transcription of short hairpin RNA (shRNA) targeting the p120-Ras GTPase-activating protein (RasGAP). The RasGAP shRNA sequence was removed from the transgene by digest with Asp718 and XbaI, to leave a pcDNA3.1 (+) backbone (Invitrogen) containing the pol III promoter. The target Tβ4 shRNA sequences, underlined in Figure 1, were prepared by oligonucleotide synthesis: 5’-GTACCCCTGAGATCGAGAAATTCGATAAGTTCAAGAGCTTTATCGAATTTCTCGATCTCAGG–3’ (Tβ4 shRNA1, sense strand); 5’-CTAGATTCCCCAAAATCGATCGAGAATTTCTCGATCTCAGTTTTTGGAAAT-3’ (Tβ4 shRNA1, antisense strand); 5’-GTACCCAGAAGCAGCTGCGAATCGTAATTCAGAGATTACGATCGCAGCTTTTCTTTTTGAAAAT–3’ (Tβ4, shRNA2, sense strand); 5’-CTAGATTCCCCAAAAGAAGCAGCTGCGAATCGTAATCTCTTGAATTACGATTCGCCAGCTTGCTTCTG–3’ (Tβ4 shRNA2, antisense strand).

Oligonucleotides were annealed by heating to 90°C in the presence of 50mM NaCl and gradual cooling to room temperature. Double stranded oligonucleotides were introduced into the Asp718 and XbaI sites of pcDNA3-H1 pol III.

**Generation of β-thymosin reporter constructs**

Oligonucleotides were synthesised to include the 23nt target sequence along with 38nt of flanking sequence: 5’-CAAGGATGTCTGACAAACCCGATATGGCTGAGATCGAGAAATTCGATAAGTTCAAGAGCTTTATCGAATTTCTCGATCTCAGG–3’ (Tβ4 reporter 1, sense strand); 5’-CTTGGTTCTGTTTTCTTCAACTTCGACTTATCGAAGCTTTTCTTTTTGAAAAT–3’ (Tβ4 reporter 1, antisense strand); 5’-CAAGGATGTCGAGTAAGTGGAGCTGACGTGAAGTTCAAGAGAAAAACAGAAACGAGC–3’ (Tβ10 reporter 1, sense strand); 5’-CTTGGTCTTCTTTTCTTCAACTTCGACGTGAAGTTCAAGAGAAAAACAGAAACGAGC–3’ (Tβ10 reporter 1, antisense strand); 5’-CAAGGATGTCGAGTAAGTGGAGCTGACGTGAAGTTCAAGAGAAAAACAGAAACGAGC–3’ (Tβ10 reporter 1, sense strand); 5’-CTTGGTCTTCTTTTCTTCAACTTCGACGTGAAGTTCAAGAGAAAAACAGAAACGAGC–3’ (Tβ10 reporter 1, antisense strand).

![Fig. 1: Tβ4 and Tβ10 share a high degree of sequence homology. An alignment of Tβ4 and Tβ10 coding sequences, using ClustalW (MRC RFCGR, http://www.rfcgr.mrc.ac.uk/Registered/Option/clustal.html). Sequences selected for Tβ4 shRNA constructs, on the basis of lower homology with Tβ10, are underlined.](image-url)
GGCGATTTC CCC CAGTCCGGCTTGCTGCAAC-3' (Tß10 reporter 1, sense strand); 5'-
CAGGTTCAAAAAACAAATGGCAAGAGAAGCAA
GCTGCCCAATCAGTGAGAGCCCGCCCAATAT
C-3' (Tß4 reporter 2, sense strand); 5'-
CTTGGGATATTGGCTGCGCTCGCCTATGCAATCGCC
AGCTTGGCTTCTCGTTACATTGGGCTGGAC-3' (Tß4 reporter 2, antisense strand); 5'-
CAAGGACCCAAAGAGACCTGAAGAGATTTCCCCAC
C-3' (Tß10 reporter 2, sense strand); 5'-
CTTGGTGGGGAAATCTTCCTAGGCTTTAGGAGATTTC
ACTCCTCTTTTCCCAATTTGCTCTTGGTC-3' (Tß4 reporter 2, antisense strand). Oligonucleotides were
annealed as described above and ligated into the StyI
site of pGL2-Basic (Promega). To enable a high level of basal
expression, the SV40 promoter of pGL2-Control
(Promega) was subcloned as a Hind III/Kpn I
fragment into the pGL2-Basic polylinker, upstream of the target
sequences.

Culture and transfection of NIH-3T3 cells

All cell culture reagents were from Invitrogen. Cells were
maintained in DMEM, supplemented with 10% (w/v)
bovine calf serum, 100 units/ml penicillin and 100 µg/ml streptomycin, at 37°C in a humidified 5% CO₂/air
atmosphere. Cells were transfected with the shRNA or
co-transfected with shRNA and reporter constructs,
using Effectene (Qiagen) transfection reagent. 2.1 x 10⁶
cells were cultured overnight to 50-70% confluence and a
total of 500 ng DNA added per well. A pCMV-β-gal
plasmid was also included as a marker of transfection
efficiency. Duplicate transfections were performed in
each experiment and data pooled from four independent
experiments.

Luciferase reporter assays

Cells were harvested 48 hours post-transfection and
assayed using Promega’s Luciferase Assay System. Cells
were washed once in PBS, scraped into 400 µl of reporter
lysis buffer and lysis facilitated by two rounds of
freezing (-80°C) and thawing (37°C). 20 µl lysate were
combined with 100 µl assay reagent and luciferase
activity measured using a TD-20/20 Luminometer
(Turner Designs). To normalize for transfection
efficiency, the activity of co-transfected β-galactosidase
was also assayed. 100 µl of lysate were combined with
50 µl of lysis buffer and 150 µl of 2x β-galactosidase assay
buffer and incubated at 37°C for 20-90 minutes until the
yellow o-nitrophenyl was produced. The reaction was
terminated by the addition of 1M Na₂CO₃ and the
absorbance at 420 nm recorded.

RESULTS AND DISCUSSION

We sought to target Thymosin β4, which encodes an actin-
binding protein implicated in cell movement, using
RNAi. Tß4, the predominant mammalian isoform of the
β-thymosin family, shares a high degree of homology
with other isoforms, such as Tß10 and Tß9 (12). Tß10 is
the β-thymosin most closely related to Tß4, displaying
70.4% identity within the coding region. The coding
region for both Tß4 and Tß10 is only 135 bp and as such
there is limited potential to design a Tß4-specific siRNA
that will not cross-react with Tß10. Intronic sequences are
ordinarily avoided since these immature mRNAs will be
kept in the nucleus while RNAi mediated mRNA
degradation is believed to occur predominantly in the
cytoplasm (13). 3’ and 5’ untranslated regions tend to be
avoided since they may be bound by regulatory proteins
and thus be inaccessible to siRNAs or RISC. Further
limitations are imposed by the need to adopt established
criteria for optimal siRNA design (14).

Sense and antisense Tß4 sequences of 23 base pairs in
length, separated by a nine bp spacer, were introduced
downstream of the H1 RNA pol III promoter (11, 15),
followed by a stretch of five thymines which act to
terminate transcription. Two sequence stretches were
selected, as regions displaying minimum sequence
identity with Tß10; Tß4shRNA1 incorporated six
mismatches and Tß4shRNA2 twelve mismatches versus
Tß10, as shown in Figure 1. The mismatches lie
predominantly in the center of the shRNAs, with
additional mismatches at the 5’ end of Tß4shRNA1 and
at both extremes of Tß4shRNA2. The position of a
mismatch has been shown to be a critical factor in RNAi
selectivity, with mismatches at the 5’ end more readily
tolerated than those at the 3’ end (16). Given the number and positions of the mismatches, neither Tβ4shRNA would be predicted to cross-react with Tβ10. Constructs were evaluated by transient transfection of NIH-3T3 cells. We encountered considerable difficulty when attempting to demonstrate the specificity of Tβ4 RNAi. While northern blotting appeared to show down-regulation of Tβ4 (to 21% of the level in non-transfected cells), re-probing of the blot for Tβ10 levels revealed bands of equivalent size and signal intensities as seen for Tβ4. Due to the significant level of identity shared by these two genes, the Tβ10 probe was able to cross-react with the more abundant Tβ4 transcript, which is comparable in size. Quantitative Real Time RT-PCR proved equally uninformative; although specific products could be amplified for Tβ4 and Tβ10, the degree of Tβ4 knockdown revealed by this technique was modest, likely due to the sensitivity in detecting expression in cells which failed to transfect with the shRNA constructs (estimated to be as high as 40%).

The need for a more sensitive means of determining RNAi specificity led us to develop a reporter-based assay for RNAi, as outlined in Figure 2. Our assay is based upon the introduction of target sequences downstream of the luciferase reporter gene. The ability of a complementary shRNA to knockdown its target is then quantitatively determined by a decrease in luciferase activity. Synthetic oligonucleotides containing the 23nt target region along with 20nt of flanking sequence were designed to produce the appropriate StyI overhangs when annealed (Fig. 2). Oligonucleotides were ligated into the StyI site located between the splice donor site and poly(A) signal of the luciferase gene in pGL2-Basic (Promega), resulting in the transcription of a luciferase mRNA containing the target sequence. The non-palindromic nature of the StyI site in pGL2-Basic (CCAAGG) ensured that the oligonucleotides were ligated into the vector in the desired 5’-3’ orientation. The SV40 promoter of pGL2-Control (Promega) was introduced as a KpnI/HindIII fragment into the multiple cloning region, to enable strong basal luciferase expression.

Four reporter constructs were created, two of which contained the Tβ4 shRNA target regions and the other two contained the analogous regions of Tβ10 (see alignment, Fig. 1). Co-transfection of Tβ4 reporter 1 with the appropriate shRNA (Tβ4shRNA1) resulted in a significant knockdown of luciferase activity (by 87%, n=8, p<0.0001) compared with the pcDNA3 backbone, confirming the efficacy of the shRNA (Fig. 3A). Neither of the mismatched shRNAs (Tβ4shRNA2 nor a control RasGAP shRNA (11)) affected luciferase activity.

![Fig. 2: A reporter assay to determine the specificity of RNAi. A. Oligonucleotides were synthesized to contain the 23nt shRNA target region, 20nt of flanking sequence on each side and appropriate nucleotides to generate StyI overhangs. Complementary oligonucleotides were annealed and ligated into the StyI site of pGL2-Basic. The SV40 promoter of pGL2-Control was subcloned into the KpnI/HindIII sites for maximal expression. B. The use of the reporter assay to determine whether a gene can be specifically targeted for RNAi (i) and to confirm that a closely related gene remains unaffected (ii). The respective Tβ4 shRNA target sequences and analogous sequences of Tβ10 were introduced into reporter constructs as described above. Upon transfection into mammalian cells luciferase mRNA is transcribed containing the target sequence within the 3’ untranslated region. Translation is unaffected by the presence of the target sequence and functional luciferase is synthesized. Tβ4 shRNAs, when co-transfected and transcribed by H1 polymerase III, direct the RISC towards the complementary Tβ4 target sequence within the luciferase mRNA. Degradation of the mRNA ensues, translation does not occur and no luciferase activity is detected. A mismatched target sequence (Tβ10) in principal is not recognized by the shRNA, the mRNA is translated and luciferase activity detected.](image-url)
Conversely, activity of Tβ4 reporter 2 was selectively knocked down by Tβ4shRNA2 (by 89%, n=8, p<0.0001), but not by Tβ4shRNA1 (Fig. 3B). Activity of Tβ10 reporter 1 was unaffected by the Tβ4shRNA1 as compared to the RasGAPshRNA control (Fig. 3A) and neither, Tβ4shRNA1 nor Tβ4shRNA2 adversely affected Tβ10 reporter 2 (Fig. 3B). However, the Tβ4shRNA2 construct, which contained 12 sequence mismatches to the analogous Tβ10 region, reduced Tβ10 reporter 1 activity (by 42%, n=8, p<0.001; Fig. 3A) compared to the pcDNA3 backbone. Given the degree of sequence mismatch this was an unexpected, yet significant observation since it represents partial silencing of an undesired “target.” This in turn could potentially result in a hypomorphic cellular phenotype assuming a dose-dependent requirement for Tβ10 function.

The reporter assay, therefore, has enabled us to compare the potency of shRNAs in inducing RNAi against Tβ4 and, importantly, to test the selectivity of knockdown of each shRNA against Tβ10. This method can be applied to all RNAi-based studies in which there is risk of targeting homologous genes, particularly when selective knockdown has been difficult to prove by conventional semi-quantitative assays of gene expression.

The present study exposes a further issue pertaining to RNAi, which is that siRNAs do not have to target coding region; although this is usually the preferred approach (13, 17). Whilst our shRNAs are directed against the coding sequence of the gene of interest, the sequence has been inserted into the 3’UTR of the luciferase gene, where the shRNAs induce degradation of the luciferase mRNA. In cases where sequence homology is high throughout the coding region and off-target silencing is shown to occur, it may become necessary to target the more divergent untranslated sequence. It should be possible to determine, using reporter constructs based on those described herein, whether binding by regulatory proteins renders the sequence inaccessible to the RNAi machinery upon transfection into the cell type for phenotype analysis.

The use of a reporter gene assay offers a rapid and reliable means of quantitatively assessing the potency and specificity of RNAi. With the approach described here, the generation of constructs is straightforward, relying on the use of synthetic oligonucleotides and a single restriction site that ensures directional cloning. Assaying reporter activity requires little effort and unequivocal data can be rapidly obtained, enabling immediate progression to the next experimental stage. Not all siRNAs are equally effective in inducing RNAi and much optimization is often required. Using a reporter approach, multiple siRNAs, directed towards
the same gene, may be simultaneously assayed and quantitatively compared.

The caveats in specificity of gene silencing by RNAi means there is an absolute requirement to test the selectivity of a siRNA before embarking on phenotype analysis. Previous use of reporter assays, with respect to RNAi, has enabled the testing of general knockdown (18-20), but the method has not been applied as a test for specificity. The comparison in this study of two closely related β-thymosin genes is testament to the sensitivity of this approach for determining RNAi specificity. The observation that one of our Tβ4 shRNA constructs induced 42% knock-down of a Tβ10 reporter, despite significant mismatches in sequence, further underscores the need for an assessment of the specificity of multiple shRNAs. Despite the fact that the gene most closely related to Tβ4 was unaffected by the Tβ4shRNA1 construct, we cannot fully exclude the possibility that there are other genes in the genome which can be silenced by this construct. That said, use of this assay in conjunction with optimal siRNA design, the testing of several different siRNAs for equivalent phenotypic outcome and identification of RNAi triggers at lowest possible concentration (21) should significantly improve confidence in the specificity of gene knockdown.

ACKNOWLEDGMENTS

This work was supported by Project Grants PG/2000117 and PG/04/012/16626 from the British Heart Foundation.

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