A perspective on underwater photosynthesis in submerged terrestrial wetland plants

Colmer, Timothy D.; Winkel, Anders; Pedersen, Ole

Published in:
A O B Plants

DOI:
10.1093/aobpla/plr030

Publication date:
2011

Document version
Publisher’s PDF, also known as Version of record

Citation for published version (APA):
Colmer, T. D., Winkel, A., & Pedersen, O. (2011). A perspective on underwater photosynthesis in submerged terrestrial wetland plants. A O B Plants, 2011. https://doi.org/10.1093/aobpla/plr030
A perspective on underwater photosynthesis in submerged terrestrial wetland plants

Timothy D. Colmer¹, Anders Winkel¹,² and Ole Pedersen¹,²*

¹ School of Plant Biology, The University of Western Australia, Crawley 6009, WA, Australia
² Freshwater Biological Laboratory, University of Copenhagen, Helsingørsgade 51, 3400 Hillerød, Denmark

Received: 7 October 2011; Returned for revision: 2 November 2011; Accepted: 23 November 2011; Published: 30 November 2011

Citation details: Colmer TD, Winkel A, Pedersen O. 2011. A perspective on underwater photosynthesis in submerged terrestrial wetland plants. AoB PLANTS 2011: plr030; doi:10.1093/aobpla/plr030

Abstract

Background and aims Wetland plants inhabit flood-prone areas and therefore can experience episodes of complete submergence. Submergence impedes exchange of O₂ and CO₂ between leaves and the environment, and light availability is also reduced. The present review examines limitations to underwater net photosynthesis (P₀) by terrestrial (i.e. usually emergent) wetland plants, as compared with submerged aquatic plants, with focus on leaf traits for enhanced CO₂ acquisition.

Scope Floodwaters are variable in dissolved O₂, CO₂, light and temperature, and these parameters influence underwater P₀ and the growth and survival of submerged plants. Aquatic species possess morphological and anatomical leaf traits that reduce diffusion limitations to CO₂ uptake and thus aid P₀ under water. Many aquatic plants also have carbon-concentrating mechanisms to increase CO₂ at Rubisco. Terrestrial wetland plants generally lack the numerous beneficial leaf traits possessed by aquatic plants, so submergence markedly reduces P₀. Some terrestrial species, however, produce new leaves with a thinner cuticle and higher specific leaf area, whereas others have leaves with hydrophobic surfaces so that gas films are retained when submerged; both improve CO₂ entry.

Conclusions Submergence inhibits P₀ by terrestrial wetland plants, but less so in species that produce new leaves under water or in those with leaf gas films. Leaves with a thinner cuticle, or those with gas films, have improved gas diffusion with floodwaters, so that underwater P₀ is enhanced. Underwater P₀ provides sugars and O₂ to submerged plants. Floodwaters often contain dissolved CO₂ above levels in equilibrium with air, enabling at least some P₀ by terrestrial species when submerged, although rates remain well below those in air.

Introduction

Emergent wetland plants are well adapted to waterlogged soils, but can also experience episodes of complete submergence. Complete submergence has an impact on wild species in coastal marshes and river floodplains (Armstrong et al. 1985), and many rice crops are grown in regions threatened by floods, causing submergence (Jackson and Ram 2003).

* Corresponding author’s e-mail address: opedersen@bio.ku.dk

Published by Oxford University Press. This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.
Complete submergence impedes the exchange of \( \text{O}_2 \) and \( \text{CO}_2 \) between leaves and the environment (Mommer and Visser 2005; Voesenek et al. 2006). Light availability to submerged plants also decreases, and markedly so when floodwaters are turbid (Mommer and Visser 2005; Voesenek et al. 2006). Restricted photosynthesis, but ongoing substrate consumption in respiration or fermentation, causes sugars to become depleted in submerged plants, which in turn can result in damage or even death from substrate exhaustion (Bailey-Serres and Voesenek 2008; Colmer and Voesenek 2009).

The interface between land and water is not well defined as water tables fluctuate with precipitation and evaporation, so that plants experience variable periods and depths of flooding (Sculthorpe 1967). Plants exploit niches across these dynamic flooding gradients, but functional classification of plant types lacks sharp boundaries owing to the continuum of diversity. Notwithstanding these difficulties, plants from the wettest end of the gradient have been classified into two main groups: (i) aquatic plants that primarily live completely submerged and (ii) amphibious plants that live with emergent shoots or develop water forms when submerged (Iversen 1936; cited by Sculthorpe 1967). Emergent wetland plants typically maintain a large portion of their shoots in air, but occasionally become completely submerged. To clearly distinguish these emergent plants from other wetland species with shoot portions in air (e.g. emergent amphibious plants), we refer to this functional group as ‘terrestrial wetland plants’ (present review; Colmer and Pedersen 2008; Pedersen et al. 2010). Such distinction is important as terrestrial wetland plants typically grow vigorously in waterlogged soils and/or areas with shallow standing water, with the depth limit being determined by capacity for transport of atmospheric \( \text{O}_2 \) to belowground tissues (Sorrell et al. 2000).

For submerged terrestrial plants, \( \text{O}_2 \) deficiency and escape responses via shoot elongation have been elucidated, revealing sophisticated signalling, changes in gene expression and altered metabolism during submergence (e.g. reviewed by Bailey-Serres and Voesenek 2008, 2010). The capacity for some net photosynthesis (\( P_N \)) to continue when under water enhances plant tolerance of submergence, as \( P_N \) provides \( \text{O}_2 \) for internal aeration and sugars for energy metabolism and growth (Mommer and Visser 2005).

The present review examines limitations to under-water \( P_N \) by terrestrial wetland plants and compares their functioning with aquatic plants. Our focus here on underwater \( P_N \) as related to the ecophysiology of submergence tolerance adds to the vast knowledge on root adaptations in wetland species. Roots of wetland plants typically contain large volumes of aerenchyma, often a barrier to radial \( \text{O}_2 \) loss, and the ability to tolerate tissue \( \text{O}_2 \) deficits and reduced phytotoxins in waterlogged soils (Armstrong 1979; Jackson and Armstrong 1999; Bailey-Serres and Voesenek 2008; Colmer and Voesenek 2009). Here, we show that underwater \( P_N \) by submerged terrestrial wetland plants is limited by \( \text{CO}_2 \) availability even though floodwaters commonly contain dissolved \( \text{CO}_2 \) above air equilibrium, and so leaf traits influencing underwater \( P_N \) are important for submergence tolerance.

### The submergence environment during overland floods

Flooding can occur with various combinations of chemical and physical properties in the water; \( \text{O}_2 \), \( \text{CO}_2 \), temperature, pH and light can all vary (Setter et al. 1987; Pèrez-Llorens et al. 2004). Seawater pH is well buffered as it contains \( \text{HCO}_3^- \) (2.2 mM; Millero et al. 1998) and \( \text{HCO}_3^- \) also buffers against severe depletion of dissolved \( \text{CO}_2 \). In freshwater floods, \( \text{HCO}_3^- \) and \( \text{CO}_2 \) concentrations are highly variable, but dissolved \( \text{CO}_2 \) is commonly above air equilibrium (Table 1). The high \( \text{CO}_2 \) concentrations typically result from respiration by
organisms consuming labile carbon compounds (i.e. a net heterotrophic system); in addition, some water bodies receive CO2-enriched groundwater stream flows. By contrast, in net autotrophic systems photosynthesis depletes CO2 and produces O2. So, O2 concentrations can range from severely hypoxic (net heterotrophic) to well above air equilibrium (net autotrophic).

Temperature during flooding events can also vary widely (e.g. ~6–37 °C; Hamilton et al. 1997; Hamilton et al. 1995; Richey et al. 2002; Valett et al. 2005), depending on location and season. Respiration increases at warmer temperatures, which can deplete O2, and O2 concentration is further reduced owing to lower O2 solubility in water as temperature increases. So, the imbalance between O2 demand and supply to submerged terrestrial plants can be further exacerbated as temperature increases.

Flow rates during floods have only been reported, to our knowledge, in three papers: data are available for two flash floods and one seasonal river flood, and flows ranged from 0.002 to 0.3 m s⁻¹. Flow rates affect the thickness of diffusive boundary layers (DBLs) and thereby influence gas and nutrient exchanges with submerged plants (Binzer et al. 2005; Pedersen et al. 2009). So, underwater P0 can increase with increasing flow velocity since the DBLs become thinner (Jones et al. 2000), but the response would plateau (cf. O2 supply; Binzer et al. 2005) or even decline again if flows cause excessive shoot agitation (Madsen et al. 1993a). Light regimes in floodwaters are dependent on several factors. When floodwaters contain suspended particles or dissolved coloured organic matter (e.g. tannins in Amazonian floodwaters; Parolin 2009), light availability will be reduced. Particle suspension can be highest during early stages of floods and particles often then settle; however, if particles settle on leaves these can still limit light. Waters of high nutrient availability typically support growth of microalgae, with dense populations of both biofilms and phytoplankton leading to lower light penetration to leaf surfaces (Sand-Jensen and Sondergaard 1981; Sand-Jensen and Borum 1991; Lassen et al. 1997) and consequently also shallower depth limits for plant colonization (Sand-Jensen 1990). Examples of light reductions are available for floodwaters in the rice fields of India and Thailand; the depth at which 50% light remained varied from 0.07 to 0.7 m (Setter et al. 1987; Ram et al. 1999).

How does the submerged environment experienced by terrestrial wetland plants compare with that of water bodies containing permanent aquatic vegetation? In brief, environments supporting healthy stands of submerged aquatic plants, such as the shallow sea, and bodies containing permanent aquatic vegetation? In brief, environments supporting healthy stands of submerged aquatic plants, such as the shallow sea, and areas within rivers and lakes, also share many of the above-mentioned constraints to plant growth. Light attenuation in the water column (caused by water itself, dissolved coloured organic matter, phytoplankton and other particles) determines the maximum depth of colonization by aquatic plants. Seagrasses typically grow down to ~10% of the surface light (Duarte 1991), whereas the depth penetration of plants in freshwater lakes is down to <1% and typically ~5% of the surface light (Canfield et al. 1985). The lower light compensation points for the growth of deep–colonizing freshwater plants result from these having higher shoot-to-root ratios than seagrasses. The deepest-growing freshwater plants, such as species of Ceratophyllum and Utricularia, do not produce roots at all (Cook 1990). Similar to terrestrial floodwaters, dissolved inorganic carbon (DIC) in freshwater can also vary widely (e.g. from 0.02 to 5.6 mM in British lakes; Maberly and Spence 1983). Depending on pH, the above DIC concentrations may result in dissolved CO2 levels from near or below air equilibrium (15 μM in freshwater at 20 °C) to waters in streams/rivers, ponds and lakes that are typically supersaturated (Table 1); ponds can even contain up to 2000 μM CO2 (133-fold air equilibrium). The temperature in most water bodies fluctuates significantly less than surrounding air due to the much higher specific heat capacity of water compared with air (Hutchinson 1957), but there are exceptions, such as in shallow rock pools with large diel fluctuations (Pedersen et al. 2011a). Finally, the flow velocity in aquatic environments also varies widely, as described earlier for terrestrial floods, from almost

### Table 1 Dissolved CO2 and O2 concentrations in various types of floodwaters. Medians with ranges in parentheses.

| Environment     | CO2 (µM) | O2 (µM) |
|-----------------|----------|---------|
| Terrestrial     |          |         |
| Flash flood     | (1,2)    | (n = 4) | 1040 (3 – 1953) | 150 (0°–280) |
| Seasonal flood  | (3–6)    | (n = 6) | 365 (47 – 1600) | 79 (0°–240) |
| Tidal flood     | (7,8)    | (n = 4) | 16 (3 – 49)     | 292 (188–522) |
| Aquatic         |          |         |
| Streams and rivers | (9,10) | (n = 31) | 133 (11–836) | n.a. |
| Ponds (< 1 ha)  | (11)     | (n = 7) | 59 (<1–376)    | n.a. |
| Lakes           | (11)     | (n = 17) | 45 (11–210)    | n.a. |

1.(Ram et al. 1999; Setter et al. 1987); 2.(Hamilton et al. 1997; Hamilton et al. 1995; Richey et al. 2002; Valett et al. 2005).
3.(Pérez-Llorens et al. 2004; Winkel et al. 2011); 4.(Sand-Jensen and Frost-Christensen 1998); 5.(Jonsson et al. 2003) 6.(Stoehr et al. 2011). n.a., not available.

O2 was not measured in the water surveys conducted in 9, 10 and 11.
stagnant conditions in ponds and deeper areas of lakes to very high velocities in rivers and in surf zones of the sea (2–3 m s\(^{-1}\); Vogel 1994). In fast-flowing water or in wave-zones, the strap-shaped leaves typical of some aquatic plants are highly adaptive as this morphology reduces the pressure drag (Vogel 1994).

In summary, floodwaters faced by terrestrial plants invoke some common constraints of restricted gas exchange and lower light availability, but conditions (O\(_2\), CO\(_2\), light and temperature) differ between locations and times, posing variable challenges to plant functioning during submergence. Floodwater chemical and physical properties, in addition to the well-recognized importance of seasonal timing, duration, depth and frequency of floods (e.g. Vervuren et al. 2003), will influence plant growth and survival during submergence.

### Net photosynthesis under water

Low CO\(_2\) and/or low light can restrict \(P_N\) by submerged plants (Sand-Jensen 1989). This review focuses on CO\(_2\) acquisition. Aquatic species possess leaf traits to enhance DIC supply and thus rates of underwater \(P_N\). In Table 2, we compare the leaf traits of terrestrial wetland plants with those of submerged aquatic plants. Below we (i) summarize knowledge of morphological and anatomical leaf traits, and photosynthetic pathways including carbon-concentrating mechanisms (CCMs), and (ii) compare the rates of underwater \(P_N\) by different types of aquatic and terrestrial wetland plants, as influenced by these leaf traits.

#### Leaf traits of terrestrial wetland plants and submerged aquatic plants

Leaf morphology determines boundary layer resistances to exchange of dissolved gases and ions (Madsen and Sand-Jensen 1991). Boundary layer resistance can limit the rates of CO\(_2\) uptake and thus reduce underwater \(P_N\) in submerged plants as diffusion is 10\(^4\)-fold slower in water than in air (Vogel 1994). Morphological traits (Table 2) that reduce the DBL resistance, by decreasing the distance to the ‘leading edge’ (Vogel 1994), include leaf shapes of small, dissected/lobed and/or strap-like leaves. In addition, aquatic leaves lack trichomes, thus avoiding the development of thicker boundary layers adjacent to their surfaces. Leaves of aquatic species also tend to be thin (Table 2), although there are several exceptions (e.g. isoetids; Sand-Jensen and Prahl 1982). Thin leaves have short internal diffusion path lengths, reducing the overall resistance for CO\(_2\) to reach chloroplasts (Madsen and Sand-Jensen 1991; Maberly and Madsen 2002). One example is the lamina of Najas flexilis, which is only two cell layers (Tomlinson 1982). In cases where leaves are relatively thick, CO\(_2\) is typically sourced from sediments (e.g. isoetids; Winkel and Borum 2009), and these leaves tend to be of high porosity to facilitate internal gas phase diffusion (Pedersen and Sand-Jensen 1992; Pedersen et al. 1995; Sand-Jensen et al. 2005).

### Table 2 Comparison of leaf traits influencing gas exchange and photosynthesis by terrestrial wetland plants when under water and by submerged aquatic plants

| Leaf traits | Terrestrial wetland plants | Submerged aquatic plants |
|-------------|-----------------------------|--------------------------|
| Morphology  |                             |                          |
| Leaf size   | Medium–large                | Small–medium             |
| Dissected/lobed | Rare                        | Common                   |
| Strap-shaped | Rare                        | Common                   |
| Leaf thickness\(a\) | Moderate–thick             | Thin                     |
| Surface hydrophobicity/leaf gas films\(1,2\) | Common                | Absent                   |
| Hairs/trichomes | Rare                      | Absent                   |
| Anatomy     |                             |                          |
| Stomata     | Always present              | Absent/ non-functional   |
| Cuticle     | Always present              | Absent/highly reduced    |
| Chloroplasts in epidermal cells | Only in guard cells       | Common                   |
| Aerenchyma  | Variable                    | Variable                 |
| Porosity of lamina | High in thick, low in thin, lamina | High in thick, low in thin, lamina |
| Supporting fibres | Always present             | Rare                     |
| Photosynthetic pathway/CCM\(3\) | Common                | Common                   |
| C3          | Common                      | Common                   |
| C4          | Rare                        | Rare (but uncertain)     |
| CAM         | Absent                      | Rare                     |
| HCO\(_3\)\(_2\) use | Absent                | Common                   |

\(a\)For data on SLA see Fig. 2. Other leaf features/properties can also differ between terrestrial wetland plants and submerged aquatic plants, such as: venation, lignification, stiffness, surface topography, differences between adaxial and abaxial surfaces, and in the case of some halophytic wetland species, presence of salt bladders and glands.
In addition to these morphological traits, leaves of aquatic species also have anatomical traits that further reduce diffusive resistances for CO₂ to reach chloroplasts (Table 2). Aquatic leaves lack, or have very reduced, cuticles. Diffusion across the cuticle is the main pathway of dissolved gas exchange as the leaves lack stomata, or if present, the stomata are non-functional (Pedersen and Sand-Jensen 1992). The diffusion path length to chloroplasts is also minimized by having these organelles in all epidermal cells, and in sub-epidermal cells the chloroplasts are positioned towards the exterior (Table 2).

Submerged aquatic plants also display physiological adaptations to increase the CO₂ concentration at Rubisco, the site of carboxylation (Table 2)—these are referred to as CCMs (Maberly and Madsen 2002; Raven et al. 2008). In submerged aquatic plants, CCMs include HCO₃⁻ use (Prins and Elzenga 1989), C₄ (Magnin et al. 1997), C₃–C₄ intermediates (Keeley 1999) and Crassulacean Acid Metabolism (CAM) photosynthesis (Keeley 1998). Carbon-concentrating mechanisms increase Pₐ in CO₂-limited submerged environments, and have also been suggested to diminish photorespiration (Maberly and Madsen 2002). Photorespiration results from the oxygenase activity of Rubisco and is promoted by a low CO₂:O₂ ratio (Ogren 1984), a condition common in leaves when under water (Bowes 1987). The low CO₂ availability in aquatic environments would in itself lower the CO₂:O₂ ratio. Moreover, O₂ in submerged leaves can be high as escape is slower than production in Pₐ; O₂ escape is not only hampered by DBLs but also by the relatively low O₂ solubility in water; CO₂ is 28.5-fold more soluble than O₂ at 20 °C (Baranenko et al. 1990). Reduced photorespiration in a submerged aquatic CAM plant has been recently demonstrated (Pedersen et al. 2011b), supporting the view that CCMs do reduce photorespiration in aquatic species.

By contrast with aquatic species, leaves of terrestrial wetland plants lack most of the features described above (Table 2) and so suffer from large diffusion limitations to CO₂ supply for Pₐ when under water, unless they possess leaf gas films (Raskin and Kende 1983; Colmer and Pedersen 2008; Pedersen et al. 2009) or produce submergence-acclimated leaves (Møller and Visser 2005). Below, we evaluate underwater Pₐ by leaves of terrestrial wetland plants and then consider the occurrence and functioning of leaf gas films.

**Net photosynthesis of aquatic and submerged terrestrial wetland plants; leaf traits enhance CO₂ supply**

The most comprehensive comparison of underwater Pₐ by aquatic and terrestrial wetland plants is the study by Sand-Jensen et al. (1992). Thirty-five species of four life forms (terrestrial, amphibious homophyllous, amphibious heterophyllous and aquatic species) were compared (listed in Appendix 1). Inclusion of data from other studies in the present analysis was constrained by differences in techniques and conditions used for underwater Pₐ measurements, e.g. CO₂, temperature and light (Appendix 2).

Classifications of wetland plants into functional groups are convenient, but are also imperfect as the boundaries are not sharp (see Introduction). As examples, some terrestrial wetland species produce new leaves when submerged and these can display some acclimation to the underwater environment (Møller et al. 2007). Similarly, homophyllous amphibious plants can also display some acclimation, e.g. thinner cuticles and modestly thinner leaves when formed under water (Nielsen 1993), but these changes are far more subtle than those displayed by heterophyllous amphibious plants. Not surprisingly, different authors have classified some species into different life forms. Here, our focus is on the comparison of underwater Pₐ of leaves formed (i) in air by terrestrial wetland species, (ii) under water by amphibious homophyllous species, (iii) under water by amphibious heterophyllous species and (iv) under water by aquatic species.

An additional noteworthy feature of the study by Sand-Jensen et al. (1992) was documentation of dissolved CO₂ levels in lowland stream habitats. Underwater Pₐ was measured at ambient and at elevated CO₂ concentrations, to provide rates of relevance to the field situation as well as CO₂-saturated Pₐ for aquatic leaf types. The level of elevated CO₂ used (∼800 μM, being ∼50-fold air equilibrium) would have saturated Pₐ by the aquatic leaf types. It is uncertain whether rates were CO₂ saturated for some of the terrestrial leaf types, which can require as much as 75-fold of air equilibrium CO₂ when submerged (Colmer and Pedersen 2008).

We compare the rates on the dry mass basis (Fig. 1A and B) used by Sand-Jensen et al. (1992) and also on a projected leaf surface area basis (Fig. 1C and D); conversions used specific leaf area (SLA) data in the literature (Fig. 2; Appendix 1). Data for SLA were not available for three of the aquatic and three of the terrestrial wetland species in Sand-Jensen et al. (1992), so these six were omitted from the present analysis (Appendix 1).

The overall beneficial effects of aquatic leaf traits (Table 2) for underwater Pₐ, as well as the generally poor performance of leaves of terrestrial plants, were clearly demonstrated in Sand-Jensen et al. (1992). These authors highlighted that (i) underwater Pₐ on a mass basis increased from terrestrial, then amphibious, to truly aquatic leaf types and (ii) Danish lowland...
stream waters are commonly supersaturated with CO₂, allowing even some terrestrial species to have adequate Pₐ for growth when submerged in these habitats. The higher Pₐ by aquatic leaf types per unit mass with near-ambient CO₂ concentrations (∼90–400 μM) demonstrates the higher C-return per unit of dry mass investment by these leaf types in an underwater environment as compared with terrestrial types (Fig. 1A). When external CO₂ was supplied at an elevated level of ∼800 μM (Fig. 1B), underwater Pₐ values by the aquatic and heterophyllous amphibious leaves still exceeded those of the terrestrial and homophyllous leaf types. The low rates by terrestrial leaves even with elevated CO₂ further demonstrate the large diffusion limitations for CO₂ entry that restrict underwater Pₐ.

Expression of underwater Pₐ rates on a surface area basis, the units typically used in terrestrial plant physiology (whereas in aquatic sciences, rates are typically expressed per unit dry mass), interestingly, removes differences between the terrestrial and aquatic leaf types, at both ambient and elevated CO₂ (Fig. 1C and D). The order of magnitude of higher SLA (Fig. 2) of aquatic and many amphibious leaf types clearly sets an upper

---

**Fig. 1** Underwater net photosynthesis (Pₐ) in terrestrial wetland plants, in amphibious homophyllous or heterophyllous wetland plants and in submerged aquatic plants. Net photosynthesis was measured at 15 °C and is expressed per leaf dry mass (A and B) or per projected leaf area (C and D) at ambient CO₂ levels (90–400 μM in the natural habitats; A and C) or at elevated CO₂ levels (800 μM; B and D). Species and SLA data sources are listed in Appendix 1. Our analysis focused on the study by Sand-Jensen et al. (1992) as it is the most comprehensive available; addition of other data was constrained by differences in techniques and conditions used (e.g. CO₂ and temperature; Appendix 2). Terrestrial, leaves formed in air by emergent wetland plants; homophyllous, leaves formed under water by amphibious wetland plants; heterophyllous, leaves formed under water by amphibious wetland plants; aquatic, leaves formed under water by submerged aquatic plants (cf. Sculthorpe 1967). Rates on a mass basis (A and B) were converted to an area basis (C and D) using the published SLA data (Fig. 2, Appendix 1). The box-whisker plot shows the median, 10 and 90 percentiles, minimum and maximum values, and means are shown as ‘+’; the dot in the terrestrial column indicates an outlier. Differences amongst means of the four plant groups within each panel were tested by one-way analysis of variance and Tukey’s multiple comparison tests. **P < 0.01 and ***P < 0.001. Means with the same letter within each panel do not differ significantly at the 95% confidence interval.
limit for $P_N$ on an area basis. Maximum $P_N$, however, would rarely be achieved in most aquatic environments owing to light and CO$_2$ limitations (Sand-Jensen 1989; Kirk 1994) so that the lower CO$_2$-saturated rates of $P_N$ on an area basis for aquatic leaves would be of little consequence for their life under water.

Comparisons of the rates of underwater $P_N$ by terrestrial wetland plant leaf types with those achieved by aquatic leaf types are informative with respect to performance when submerged (Fig. 1), but here we also consider how these rates under water compare against those in air. For the terrestrial wetland species in Fig. 1, we could only find data on $P_N$ in air for three (Carex elata, Ranunculus repens and Phragmites australis; Appendix 1); $P_N$ in air at ambient CO$_2$ was 12.5–17 μmol m$^{-2}$ s$^{-1}$. When submerged with CO$_2$ at levels near ambient (but well above air equilibrium in these habitats), the mean $P_N$ under water was only 9% of that in air (cf. Fig. 1C). Thus, underwater $P_N$ is greatly reduced when terrestrial wetland species become submerged.

The analyses presented above for underwater $P_N$ by leaves of terrestrial wetland plants involved experiments in which leaves growing in air were tested under water. Several terrestrial wetland species produce new leaves when submerged, and these can display some acclimation to the underwater environment (e.g. thinner cuticles and thinner leaves; Mommer et al. 2007). Acclimated leaves have decreased resistances against CO$_2$ and O$_2$ movement across the cuticle and epidermis (Mommer and Visser 2005; Mommer et al. 2007). The best example is the several-fold reduction in cuticle resistance and thus the 69-fold higher underwater $P_N$ at an external CO$_2$ concentration of 250 μM by Rumex palustris (Mommer et al. 2006). Although a study of seven terrestrial wetland species established the formation of a thinner cuticle as a common response when submerged, and demonstrated enhanced underwater gas exchange, the degree of this response was not correlated with submergence tolerance among these species (Mommer et al. 2007). These anatomical, and in some cases morphological (e.g. R. palustris leaves are also more elongated), changes in submerged leaves of terrestrial species are much more subtle than the altered leaf development displayed by amphibious heterophyllous species which produce true aquatic leaf types when under water (Nielsen 1993).

In summary, $P_N$ by terrestrial wetland plants is reduced markedly when they are submerged. Leaves of terrestrial wetland plants generally lack the numerous beneficial leaf traits for underwater $P_N$ possessed by aquatic plants, although new leaves can display some acclimation (e.g. thinner cuticles and higher SLA). In addition, as discussed in the next section, some leaves of terrestrial wetland species retain a gas film when submerged, a trait that also enhances underwater $P_N$.

**Leaf gas films enhance the net photosynthesis of submerged terrestrial wetland plants**

Many terrestrial wetland plants have water-repellent (i.e. hydrophobic) leaf surfaces, resulting in self-cleaning by water droplets as these run off leaves (Neinhuis and Barthlott 1997). Leaf water repellence has been assessed by measurement of water droplet contact angles with the surface (Adam 1963; Brewer and Smith 1997; Neinhuis and Barthlott 1997)—angles of 140° or more indicate a hydrophobic surface whereas angles of 110° or less indicate a wettable surface. Water repellence (i.e. surface hydrophobicity) is determined by the micro- and nano-structures of the surface, as well as wax crystals (Wagner et al. 2003; Bhushan and Jung 2006).

Superhydrophobic leaves retain a microlayer of gas when submerged, referred to as ‘gas envelopes’ (Setter et al. 1989) and/or ‘leaf gas films’ (Colmer and Pedersen 2008). We prefer the term ‘gas film’ because although leaves of some species retain a gas layer on both sides (i.e. enveloped in gas), others retain a gas layer on only one side due to differences in hydrophobicity between adaxial and abaxial surfaces (Colmer and Pedersen 2008).
Gas films on leaves have been observed in field situations for several terrestrial wetland species when submerged in lakes, ponds, river edges and rice fields on floodplains: rice (Setter et al. 1987); P. australis, cover of New Phytologist, Volume 177(4); Spartina anglica (Winkel et al. 2011); and own observations (A. Winkel, T. D. Colmer and O. Pedersen). Information on the persistence of gas films on leaves with time following submergence is scant; gas films remained for at least 2 weeks (i.e. evaluation was terminated at 2 weeks) on leaves of Phalaris arundinacea, P. australis and Typha latifolia (all with gas films on both sides) and Glyceria maxima (gas film on only the adaxial side) in a controlled environment (Colmer and Pedersen 2008), but for some other species gas films diminish within a few days (own unpublished data; A. Winkel, T. D. Colmer and O. Pedersen).

Gas films on submerged leaves enhance CO₂ fixation, as first demonstrated for rice (9- to 10-fold increase; Raskin and Kende 1983). The beneficial effect of leaf gas films to underwater \( P_N \) has also been shown for other terrestrial wetland species; at 50 \( \mu \)M dissolved CO₂, gas films increased underwater \( P_N \) by 1.5- to 6-fold in leaves of four wetland species (Colmer and Pedersen 2008). Data demonstrating the beneficial effect of leaf gas films on underwater \( P_N \) are shown for several species in Fig. 3. Apparent resistance to CO₂ entry, at environmentally relevant CO₂ concentrations in the submergence water, was \( \sim 5\)-fold less in leaves with gas films compared with those with gas films removed (rice and P. australis; Pedersen et al. 2009).

Leaf gas films provide an enlarged gas–water interface to promote gas exchange with the surrounding floodwater (CO₂ uptake during light periods; \( O_2 \) uptake during dark periods) (Colmer and Pedersen 2008; Pedersen et al. 2009). In addition to the enlarged gas–water interface, leaf gas films might also enable stomata to remain open when leaves are submerged. By contrast, for leaves without gas films, stomata are hypothesized to close upon submergence (Mommer and Visser 2005), so that \( CO_2 \) and \( O_2 \) must then transverse the cuticle (Mommer et al. 2004). The beneficial effect of leaf gas films on underwater \( P_N \) was not only demonstrated by the marked decreases when these were removed (Fig. 3), but also leaves with this feature had higher rates of underwater \( P_N \) than leaves from species without leaf gas films (Fig. 3). Thus, leaf gas films appear to enable rates of underwater \( P_N \) by terrestrial leaves similar to those achieved by submergence-acclimated leaves of terrestrial wetland plants (data and discussion in Colmer and Pedersen 2008). Terrestrial species possessing leaf gas films would benefit from enhanced underwater \( P_N \) during short to medium periods of submergence, depending on persistence of the films. By contrast, for species lacking leaf gas films but that produce new acclimated leaves under water, these new leaves take several days to produce so that \( P_N \) would likely be less during the initial submergence period, but continued production of acclimated leaves would benefit these species during medium to prolonged submergence.

Detailed knowledge on leaf gas films is available only for rice (one cultivar only; Pedersen et al. 2009). Measurements using \( O_2 \) microelectrode profiling determined that gas film thickness varied from <10 to 140 \( \mu \)m; positional differences mainly resulted from ridges on leaves (i.e. gas films thinner at the tops of ridges, thicker between adjacent ridges). Using a ‘buoyancy method’ to measure gas volumes on the surfaces, and within, submerged leaves, showed that tissue porosity was 19% (v/v) and the gas volume of the films was 3.8 times more than the gas within the rice leaf. Diffusive boundary layer widths adjacent to submerged leaves with gas films were surprisingly larger than those adjacent to submerged leaves without gas films, so the

\[ P_N (\mu mol O_2 m^{-2} s^{-1}) \]

Fig. 3 Underwater net photosynthesis in terrestrial wetland plants with or without leaf gas films and when gas films were removed. Measurements for six species were conducted with 50 \( \mu \)M CO₂ at 20 °C and photosynthetically active radiation (PAR) of 600 \( \mu mol m^{-2} s^{-1} \); the exceptions were Oryza sativa (30 °C; PAR 350 \( \mu mol m^{-2} s^{-1} \)) and S. anglica (15 \( \mu \)M CO₂; PAR 550 \( \mu mol m^{-2} s^{-1} \)). These reflect the higher temperatures in tropical rice fields (O. sativa) and the lower CO₂ in seawater that submerges Spartina marshes. Gas films were removed from leaf surfaces by brushing with 0.05% Triton X-100. Species lacking leaf gas films were also brushed with Triton X-100 and showed no, or only a slight, reduction in \( P_N \). Data from Colmer and Pedersen (2008), Pedersen et al. (2009) and Winkel et al. (2011).
enlarged water–gas interface provided by the gas films would have been the major mechanism that reduced resistance to gas exchange of the leaves when under water. At dissolved CO₂ concentrations of relevance to field conditions (15–180 μM; e.g. in Thailand, Setter et al. 1987; India, Ram et al. 1999), underwater Pₐ was enhanced 4- to 4.9-fold by gas films on leaves of rice (Pedersen et al. 2009). Underwater Pₐ by leaves with gas films and CO₂ at near-ambient concentrations was 22% of Pₐ in air. When gas films were removed artificially from leaves of completely submerged rice, tissue sugar levels and growth were both reduced. Thus, leaf gas films contribute to submergence tolerance of rice by enhancing CO₂ entry for underwater Pₐ.

The experiments by Pedersen et al. (2009) also elucidated that when rice leaves are in flowing water (15 mm s⁻¹; simulating low flows such as might occur across rice fields), the gas film oscillates and the transition zone between mass flow in the bulk medium and diffusion in the boundary layer was wider, and more variable, than for leaves without a gas film. Oscillations of leaf gas films in flowing water were also noted by Barthlott et al. (2010), and they reported that specialized surface hairs on the leaves of Salvinia molesta can stabilize the gas film, even in fast-flowing water (such as in streams). The leaf surface of S. molesta possesses ‘eggbeater-shaped hairs’ that are hydrophobic except for the tips, a feature that enables gas film formation and retention by ‘pinning’ the water–air interface (Barthlott et al. 2010). The presence of this feature was suggested to prevent the formation and detachment of bubbles that otherwise could occur when in fast-flowing waters (Barthlott et al. 2010). This is a very interesting leaf surface feature, although the ecophysiological significance could be debated as S. molesta is a floating plant not typically found in fast-flowing waters; the large gas volume trapped by these specialized structures on the surface of the leaves would contribute significantly to the buoyancy of this floating plant.

In addition to enhanced CO₂ uptake for photosynthesis, leaf gas films also improve O₂ uptake during darkness from floodwaters into leaves (Colmer and Pedersen 2008; Pedersen et al. 2009). Thus, leaf gas films enhance leaf O₂ status both during the daytime and during nights, with benefits also of improved internal aeration of the entire body of submerged plants. Oxygen derived from Pₐ during light periods, as well as O₂ entry from the floodwater into leaves when in darkness, moves internally via aerenchyma to roots of rice (Pedersen et al. 2009) and rhizomes and roots of S. anglica (Winkel et al. 2011).

In conclusion, our recent studies of leaf gas films (Colmer and Pedersen 2008; Pedersen et al. 2009; Winkel et al. 2011) have supported the hypothesis by Setter et al. (1989), who observed this feature on submerged rice in field situations in Thailand, that gas films provide ‘an interface between the gas and water phases for collection of CO₂ and dispersal of O₂ during the day or collection of O₂ during the night’. This mechanism is analogous to the gas layer (plastron) on some aquatic insects that provides an enlarged gas–water interface between the tracheary system and surrounding water (Thorpe and Crisp 1949; Raven 2008; Pedersen and Colmer 2012). For terrestrial wetland species, the few data available indicate that leaf gas films enable rates of underwater Pₐ similar to those achieved by submergence-acclimated leaves, in both cases being higher than in terrestrial air-formed leaves without these features (data and discussion in Colmer and Pedersen 2008).

Conclusions and future perspectives

Submergence can have adverse effects on terrestrial wetland plants because of restricted gas exchange and low light. Floodwaters are variable in dissolved O₂, CO₂, light and temperature. Few data are available on key environmental parameters in various submergence environments—yet these factors influence underwater Pₐ, plant growth and survival. Knowledge of floodwater conditions will enhance one’s understanding of plant performance during submergence and enable the design of controlled experiments that better simulate particular submergence environments.

Submergence tolerance of terrestrial wetland plants is influenced by leaf traits. Although terrestrial wetland plants generally lack the numerous beneficial leaf traits possessed by aquatic plants, the few studies available demonstrate that some terrestrial species produce new leaves with a thinner cuticle under water and others possess leaf gas films. The improved gas diffusion between leaves and floodwaters enhances underwater Pₐ and so contributes significantly to sugar and O₂ supply of submerged plants. However, studies of leaf gas film functioning are in their infancy. Our priorities are (i) quantification of the occurrence and persistence of leaf gas films amongst a wide number of wetland species, and determination of whether this trait is related to species distributions in various flood-prone wetlands (cf. analysis of shoot elongation trait; Voesenek et al. 2004) and (ii) evaluation of whether rice, or its relatives, possesses variation in leaf gas film formation and persistence, and elucidation of the underlying genetic control of this trait using the array of resources available in rice.

More broadly, there are surprisingly few studies on Pₐ by terrestrial wetland plants when emergent and when submerged. Also lacking are measurements of Pₐ with
time after submergence and de-submergence. Future studies should compare the performances of species from various habitats, using a range of appropriate bases of expression of $P_N$ rates (area, mass, chlorophyll and leaf N) to facilitate interdisciplinary comparisons by aquatic and terrestrial plant biologists.

Sources of funding
This work was funded by The Danish Council for Independent Research Natural Science grant No. 645-06-0287; The Centre for Lake Restoration, a Villum Kann Rasmussen Centre of Excellence; and Australian Research Council Project LP0882350. AoB PLANTS supported O.P.’s participation at the International Botanical Congress 2011 in Melbourne.

Contributions by the authors
A.W. compiled the literature and drafted the section ‘The submergence environment during overland floods’. T.D.C. and O.P. contributed equally to the remaining sections.

Acknowledgements
We thank Kaj Sand-Jensen for constructive criticisms on a draft of our manuscript and Dina Ronzhina, Botanic Garden of Ural, Russian Academy of Science, for supplying SLA data on two species (see Appendix 1). We thank the referees for suggestions that improved our paper.

Conflicts of interest statement
None declared.

References
Adam NK. 1963. Principles of water repellency. In: Molliet J, ed. Waterproofing and water repellency. Amsterdam: Elsevier.
Andersson TN, Lundegård B. 1999. Field horsetail (Equisetum arvense): effects of potassium under different light and nitrogen conditions. Weed Science 47: 47–54.
Armstrong W. 1979. Aeration in higher plants. Advances in Botanical Research 7: 225–332.
Armstrong W, Wright EJ, Lythe S, Gaynard TJ. 1985. Plant zonation and the effects of the spring-neap tidal cycle on soil aeration in a Humber salt marsh. Journal of Ecology 73: 323–339.
Bailey-Serres J, Voesenek LACJ. 2008. Flooding stress: acclimations and genetic diversity. Annual Review of Plant Biology 59: 313–339.
Bailey-Serres J, Voesenek LACJ. 2010. Life in the balance: a signaling network controlling survival of flooding. Current Opinion in Plant Biology 13: 489–494.
Baranenko VI, Falkovskii LN, Kirov VS, Kurnyk LN, Musienko AN, Piontkovskii AI. 1990. Solubility of oxygen and carbon dioxide in water. Atomic Energy 68: 342–346.
Barthlott W, Schimmel T, Wiersch S, Koch K, Brede M, Barczewski M, Wolheim S, Weis A, Koltenmoier A, Leder A, Bohn HF. 2010. The Salvinia paradox: superhydrophobic surfaces with hydrophilic pins for air retention under water. Advanced Materials 22: 2325–2328.
Bhushan B, Jung YC. 2006. Micro- and nanoscale characterization of hydrophobic and hydrophilic leaf surfaces. Nanotechnology 17: 2758–2772.
Binzer T, Borum J, Pedersen O. 2005. Flow velocity affects internal oxygen conditions in the seagrass Cymodocea nodosa. Aquatic Botany 83: 239–247.
Bowes G. 1987. Aquatic plant photosynthesis: strategies that enhance carbon gain. In: Crawford RMM, ed. Plant life in aquatic and amphibious habitats. Oxford: Blackwell Scientific.
Brammer H. 1990. Floods in Bangladesh: geographical background to the 1987 and 1988 floods. The Geographical Journal 156: 12–22.
Brewer CA, Smith WK. 1997. Patterns of leaf surface wetness for montane and subalpine plants. Plant, Cell and Environment 20: 1–11.
Busch J, Lösch R. 1998. Stomatal behaviour and gas exchanges of sedges (Carex sp.) under different soil moisture regimes. Physics and Chemistry of the Earth 23: 443–448.
Canfield DE, Langeland KA, Linda SB, Haler WT. 1985. Relations between water transparency and maximum depth of macrophyte colonization in lakes. Journal of Aquatic Plant Management 23: 25–28.
Colmer TD, Pedersen O. 2008. Underwater photosynthesis and respiration in leaves of submerged wetland plants: gas films improve $CO_2$ and $O_2$ exchange. New Phytologist 177: 918–926.
Colmer TD, Voesenek LACJ. 2009. Flooding tolerance: suites of plant traits in variable environments. Functional Plant Biology 36: 665–681.
Cook CDK. 1990. Aquatic plant book. The Hague: SPB Academic Publishing.
Duarte CM. 1991. Seagrass depth limits. Aquatic Botany 40: 363–377.
Flynn DFB, Sudderth EA, Bazzaz FA. 2006. Effects of aphid herbivory on biomass and leaf-level physiology of Solanum dulcamara under elevated temperature and $CO_2$. Environmental and Experimental Botany 56: 10–18.
Hamilton SK, Sippel SJ, Melack JM. 1995. Oxygen depletion and carbon dioxide and methane production in waters of the Pantanal wetland of Brazil. Biogeochemistry 30: 115–141.
Hamilton SK, Sippel SJ, Calheiros DF, Melack JM. 1997. An anoxic event and other biogeochemical effects of the Pantanal wetland on the Paraguay River. Limnology and Oceanography 42: 257–272.
Hellings SE, Gallagher JL. 1992. The effects of salinity and flooding on Phragmites australis. Journal of Applied Ecology 29: 41–49.
Hutchinson GE. 1957. A treatise on limnology: geography, physics and chemistry. New York: John Wiley & Sons, Inc.
Iversen J. 1936. Biologische Pflanzenarten als Hilfsmittel in der Vegetationsforschung. Copenhagen: Levin & Munksgaard.
Jackson MB, Armstrong W. 1999. Formation of aerenchyma and the processes of plant ventilation in relation to soil flooding and submergence. Plant Biology 1: 274–287.
Jackson MB, Ram PC. 2003. Physiological and molecular basis of susceptibility and tolerance of rice plants to complete submergence. *Annals of Botany* 91: 227–241.

Jones JJ, Eaton JW, Hardwick K. 2000. The effect of changing environmental variables in the surrounding water on the physiology of *Elodea nuttallii*. *Aquatic Botany* 66: 115–129.

Jonsson A, Karlsson J, Jansson M. 2003. Sources of carbon dioxide supersaturation in clearwater and humic lakes in Northern Sweden. *Ecosystems* 6: 224–235.

Keeley JE. 1998. CAM photosynthesis in submerged aquatic plants. *Botanical Review* 64: 121–175.

Keeley JE. 1999. Photosynthetic pathway diversity in a seasonal pool community. *Functional Ecology* 13: 106–118.

Kirk JTO. 1994. *Light and photosynthesis in aquatic ecosystems*. New York: Cambridge University Press.

Lassen C, Revsbech NP, Pedersen O. 1997. Macrophyte development and resuspension regulate the photosynthesis and production of benthic microalgae. *Hydrobiologia* 350: 1–11.

Lensen JPM, Menting FBJ, Van der Putten WH. 2003. Plant responses to simultaneous stress of waterlogging and shade: amplified or hierarchical effects? *New Phytologist* 157: 281–290.

Lynn DE, Waldren S. 2002. Physiological variation in populations of *Ranunculus repens* L. (creeping buttercup) from the temporary limestone lakes (Turloughs) in the west of Ireland. *Annals of Botany* 89: 707–714.

Maberly SC, Madsen TV. 2002. Freshwater angiosperm carbon concentrating mechanisms: processes and patterns. *Functional Plant Biology* 29: 393–405.

Maberly SC, Spence DHN. 1983. Photosynthetic inorganic carbon use by freshwater plants. *Journal of Ecology* 71: 705–724.

Madsen TV, Sand-Jensen K. 1991. Photosynthetic carbon assimilation in aquatic macrophytes. *Aquatic Botany* 41: 5–40.

Madsen TV, Enevoldsen HO, Jørgensen TB. 1993a. Effects of water velocity on photosynthesis and dark respiration in submerged stream macrophytes. *Plant, Cell and Environment* 16: 317–322.

Madsen TV, Sand-Jensen K, Beer S. 1993b. Comparison of photosynthetic performance and carboxylation capacity in a range of aquatic macrophytes of different growth forms. *Aquatic Botany* 44: 373–384.

Madsen TV, Maberly SC, Bowes G. 1996. Photosynthetic acclimation of submersed angiosperms to CO2 and HCO3-. *Aquatic Botany* 53: 15–30.

Magnin NC, Cooley BA, Reiskind JB, Bowes G. 1997. Regulation and localization of key enzymes during the induction of Kranz-less, C₄-type photosynthesis in *Hydrilla verticillata*. *Plant Physiology* 115: 1681–1689.

Meziane D, Shipley B. 2001. Direct and indirect relationships between specific leaf area, leaf nitrogen and leaf gas exchange. Effects of irradiance and nutrient supply. *Annals of Botany* 88: 915–927.

Millero FJ, Lee K, Roche M. 1998. Distribution of alkalinity in the surface waters of the major oceans. *Marine Chemistry* 60: 111–130.

Mommer L, Visser EJW. 2005. Underwater photosynthesis in flooded terrestrial plants: a matter of leaf plasticity. *Annals of Botany* 96: 581–589.

Mommer L, Pedersen O, Visser EJW. 2004. Acclimation of a terrestrial plant to submergence facilitates gas exchange under water. *Plant, Cell and Environment* 27: 1281–1287.

Mommer L, Pons TL, Visser EJW. 2006. Photosynthetic consequences of phenotypic plasticity in response to submergences: *Rumex polystachis* as a case study. *Journal of Experimental Botany* 57: 283–290.

Mommer L, Wolters-Arts M, Andersen C, Visser EJW, Pedersen O. 2007. Submergence-induced leaf acclimation in terrestrial species varying in flooding tolerance. *New Phytologist* 173: 337–345.

Neinhuis C, Barthlott W. 1997. Characterization and distribution of water-repellent, self-cleaning plant surfaces. *Annals of Botany* 79: 667–677.

Nielsen SL. 1993. A comparison of aerial and submerged photosynthesis in some Danish amphibious plants. *Aquatic Botany* 45: 27–40.

Nielsen SL, Nielsen HD. 2006. Pigments, photosynthesis and photo-inhibition in two amphibious plants: consequences of varying carbon availability. *New Phytologist* 170: 311–319.

Nielsen SL, Sand-Jensen K. 1989. Regulation of photosynthetic rates of submerged rooted macrophytes. *Oecologia* 81: 364–368.

Ogren WL. 1984. *Photosynthesis: pathways, regulation, and modification*. Annual *Review of Plant Physiology* 35: 415–442.

Parolin P. 2009. Submerged in darkness: adaptations to prolonged submergence by woody species of the Amazonian floodplains. *Annals of Botany* 103: 359–376.

Pedersen O, Colmer TD. 2012. Physical gills prevent drowning of many wetland insects, spiders and plants. *Journal of Experimental Biology*, in press.

Pedersen O, Sand-Jensen K. 1992. Adaptations of submerged *Lobelia dortmanna* to aerial life form—morphology, carbon sources and oxygen dynamics. *Oikos* 65: 89–96.

Pedersen O, Sand-Jensen K, Revsbech NP. 1995. Diel pulses of O₂ and CO₂ in sandy lake sediments inhabited by *Lobelia dortmanna*. *Ecology* 76: 1536–1545.

Pedersen O, Vos H, Colmer TD. 2006. Oxygen dynamics during submergence in the halophytic stem succulent *Halosarcia pergranulata*. *Plant, Cell and Environment* 29: 1388–1399.

Pedersen O, Rich SM, Colmer TD. 2009. Surviving floods: leaf gas films improve O₂ and CO₂ exchange, root aeration, and growth of completely submerged rice. *The Plant Journal* 58: 147–156.

Pedersen O, Malik AI, Colmer TD. 2010. Submergence tolerance in *Hordeum marinum*: dissolved CO₂ determines underwater photosynthesis and growth. *Functional Plant Biology* 37: 524–531.

Pedersen O, Pulido C, Rich SM, Colmer TD. 2011a. In situ O₂ dynamics in submerged *Isoetes australis*: varied leaf gas permeability influences underwater photosynthesis and internal O₂. *Journal of Experimental Botany* 62: 4691–4700.

Pedersen O, Rich SM, Pulido C, Cawthray GR, Colmer TD. 2011b. Crassulacean acid metabolism enhances underwater photosynthesis and diminishes photorespiration in the aquatic plant *Isoetes australis*. *New Phytologist* 190: 332–339.

Pérez-Llórens JL, Brun FG, Andrià J, Vergara JJ. 2004. Seasonal and tidal variability of environmental carbon related physicochemical variables and inorganic C acquisition in *Graziariopsis*
longissima and Enteromorpha intestinalis from Los Toruños salt marsh (Cádiz Bay, Spain). *Journal of Experimental Marine Biology and Ecology* **304**: 183–201.

Prins HBA, Elzenga JTM. 1989. Bicarbonate utilization: function and mechanism. *Aquatic Botany* **34**: 59–83.

Ram PC, Singh AK, Singh BB, Singh VK, Singh HP, Setter TL, Singh VP, Singh RK. 1999. Environmental characterization of floodwater in eastern India: relevance to submergence tolerance of lowland rice. *Experimental Agriculture* **35**: 141–152.

Raskin I, Kende H. 1983. How does deep water rice solve its aeration problem? *Plant Physiology* **72**: 447–454.

Raven JA. 2008. Not drowning but photosynthesizing: probing plant plastrons. *New Phytologist* **177**: 841–845.

Raven JA, Giordano M, Beardall J. 2008. Insights into the evolution of CCMs from comparisons with other resource acquisition and assimilation. *Physiologia Plantarum* **133**: 4–14.

Reiskind JB, Beer S, Bowes G. 1989. Photosynthesis, photorespiration and ecophysiological interactions in marine macroalgae. *Aquatic Botany* **34**: 131–152.

Richey JE, Melack JM, Aufdenkampe AK, Ballester VM, Hess LL. 2002. Outgassing from Amazonian rivers and wetlands as a large tropical source of atmospheric CO₂. *Nature* **416**: 617–620.

Sand-Jensen K. 1989. Environmental variables and their effect on photosynthesis of aquatic plant-communities. *Aquatic Botany* **34**: 5–25.

Sand-Jensen K. 1990. Epiphyte shading—its role in resulting depth distribution of submerged aquatic macrophytes. *Folia Geobotanica & Phytotaxonomica* **25**: 315–320.

Sand-Jensen K, Borum J. 1991. Interactions among phytoplankton, periphyton, and macrophytes in temperate freshwaters and estuaries. *Aquatic Botany* **41**: 137–175.

Sand-Jensen K, Frost-Christensen H. 1998. Photosynthesis of amphiobious and obligately submerged plants in CO₂-rich lowland streams. *Oecologia* **117**: 31–39.

Sand-Jensen K, Frost-Christensen H. 1999. Plant growth and photosynthesis in the transition zone between land and stream. *Aquatic Botany* **63**: 23–35.

Sand-Jensen K, Prahl C. 1982. Oxygen-exchange with the lacunae and across leaves and roots of the submerged vascular macrophyte, *Lobelia dortmanna* L. *New Phytologist* **91**: 103–120.

Sand-Jensen K, Sondergaard M. 1981. Phytoplankton and epiphyte development and their shading effect on submerged macrophytes in lakes of different nutrient status. *Internationale Revue der Gesamten Hydrobiologie* **66**: 529–552.

Sand-Jensen K, Pedersen MF, Nielsen SL. 1992. Photosynthetic use of inorganic carbon among primary and secondary water plants in streams. *Freshwater Biology* **27**: 283–293.

Sand-Jensen K, Pedersen O, Binzer T, Borum J. 2005. Contrasting oxygen dynamics in the freshwater isoetid *Lobelia dortmanna* and the marine seagrass *Zostera marina*. *Annals of Botany* **96**: 613–623.

Sculthorpe CD. 1967. *The biology of aquatic vascular plants*. London: Edward Arnold Ltd.

Setter TL, Kupkchanakul T, Kupkchanakul K, Bhekasut P, Wiengweera A, Greenway H. 1987. Concentrations of CO₂ and O₂ in floodwater and in internodal lacunae of floating rice growing at 1–2 metre water depths. *Plant, Cell and Environment* **10**: 767–776.

Setter TL, Waters I, Wallace I, Bekhasut P, Greenway H. 1989. Submergence of rice. I. Growth and photosynthetic response to CO₂ enrichment of floodwater. *Australian Journal of Plant Physiology* **16**: 251–263.

Serrrell BK, Mendelsohn IA, McKee KL, Woods RA. 2000. Ecophysiology of wetland plant roots: a modelling comparison of aeration in relation to species distribution. *Annals of Botany* **86**: 675–685.

Spence DHN, Campel RM, Chrysal J. 1973. Specific leaf areas and zonation of freshwater macrophytes. *Journal of Ecology* **61**: 317–328.

Stoehr PA, Bastrup-Sporh L, Sand-Jensen K, Stedmon C. 2011. Lake metabolism scales with lake morphometry and catchment conditions. *Aquatic Sciences—Research Across Boundaries*, doi:10.1007/s00027-011-0207-6.

Thorpe WH, Crisp DJ. 1949. Studies on plastron respiration. IV. Plastron respiration in the Coleoptera. *Journal of Experimental Biology* **26**: 221–260.

Tomlinson PB. 1982. *Anatomy of the monocotyledons. VII. Helobiales (Alismatidae) (including the seagrasses).* Oxford: Clarendon Press.

Volett HM, Baker MA, Morrice JA, Crawford CS, Molles MC, Dahm CN, Moyer DL, Thibault JR, Ellis LM. 2005. Biogeochemical and metabolic responses to the flood pulse in a semiarid floodplain. *Ecology* **86**: 220–234.

Vervuren PJA, Beurskens SMJH, Blom CWPM. 1999. Light acclimation, CO₂ response and long-term capacity of underwater photosynthesis in three terrestrial plant species. *Plant, Cell and Environment* **22**: 959–968.

Vervuren PJA, Blom CWPM, De Kroon H. 2003. Extreme flooding events on the Rhine and the survival and distribution of riparian plant species. *Journal of Ecology* **91**: 135–146.

Voesenek LACJ, Rijnders JHGM, Peeters AJM, Van de Steeg HMV, De Kroon H. 2004. Plant hormones regulate fast shoot elongation under water: from genes to communities. *Ecology* **85**: 16–27.

Voesenek LACJ, Colmer TD, Pierik R, Millenaar FF, Peeters AJM. 2006. How plants cope with complete submergence. *New Phytologist* **170**: 213–226.

Vogel S. 1994. *Life in moving fluids: the physical biology of flow.* Princeton: Princeton University Press.

Wagner P, Fürstner R, Barthlott W. 2003. Quantitative assessment to the structural basis of water repellency in natural and technical surfaces. *Journal of Experimental Botany* **54**: 1295–1303.

Wang WJ, Wang HM, Zu YG. 2009. Plant biodiversity in a larch plantation from the view point of photosynthetic nitrogen use efficiency in northeast China. *Landscape Ecology and Engineering* **5**: 147–156.

Winkel A, Borum J. 2009. Use of sediment CO₂ by submersed rooted plants. *Annals of Botany* **103**: 1015–1023.

Winkel A, Colmer TD, Pedersen O. 2011. Leaf gas films of Spartina anglica enhance rhizome and root oxygen during tidal submergence. *Plant, Cell and Environment* **34**: 2083–2092.
Appendix 1: List of wetland plant species grouped into four types: terrestrial, amphibious homophyllous, amphibious heterophyllous and submerged aquatic

These species were used as data were available on underwater net photosynthesis \( (P_N) \) and specific leaf area (SLA) (see body of table for sources of data).

| Sources of information | Underwater \( P_N \) (used in Fig. 1) | SLA (used in Figs 1 and 2) | \( P_N \) in air (used in text) |
|------------------------|---------------------------------------|---------------------------|---------------------------------|
| **Terrestrial**         |                                       |                           |                                 |
| \( n = 10 \)            |                                       |                           |                                 |
| *Equisetum palustre*    | Sand-Jensen et al. (1992)              | Andersson and Lundegård (1999) | Data not available             |
| *Phragmites australis*  | Sand-Jensen et al. (1992)              | Colmer and Pedersen (2008)  | Hallings and Gallagher (1992)   |
| *Epilobium hirsutum*    | Sand-Jensen et al. (1992)              | Nielsen (1993)             | Data not available              |
| *Carex elata*           | Sand-Jensen et al. (1992)              | Meziane and Shipley (2001)  | Busch and Lösch (1998)          |
| *Poa pratensis*         | Sand-Jensen et al. (1992)              | Meziane and Shipley (2001)  | Data not available              |
| *Chrysosplenium alterniflorum* | Sand-Jensen et al. (1992) | Wang et al. 2009 | Data not available |
| *Ranunculus repens*     | Sand-Jensen et al. (1992)              | Nielsen (1993)             | Lynn and Waldren (2002)         |
| *Solanum dulcamara*     | Sand-Jensen et al. (1992)              | Flynn et al. (2006)        | Data not available              |
| *Barbara stricta*       | Sand-Jensen et al. (1992)              | Nielsen (1993)             | Data not available              |
| *Cardamine amara*       | Sand-Jensen et al. (1992)              | Nielsen (1993)             | Data not available              |
| **Amphibious (homophyllous)** \( n = 7 \) |                                       |                           |                                 |
| *Catabrosa aquatica*    | Sand-Jensen et al. (1992)              | Nielsen (1993)             | Not considered                  |
| *Glyceria maxima*       | Sand-Jensen et al. (1992)              | Colmer and Pedersen (2008)  | Not considered                  |
| *Myosotis laxa*         | Sand-Jensen et al. (1992)              | Lønssen et al. (2003)      | Not considered                  |
| *Veronica anagallis*    | Sand-Jensen et al. (1992)              | Nielsen (1993)             | Not considered                  |
| *Veronica beccabunga*   | Sand-Jensen et al. (1992)              | Nielsen (1993)             | Not considered                  |
| *Berula erecta*         | Sand-Jensen et al. (1992)              | Nielsen (1993)             | Not considered                  |
| *Myosotis palustris*    | Sand-Jensen et al. (1992)              | Nielsen (1993)             | Not considered                  |
| **Amphibious (heterophyllous)** \( n = 5 \) |                                       |                           |                                 |
| *Callitriche cophocarpa*| Sand-Jensen et al. (1992)              | Nielsen (1993)             | Not considered                  |
| *Callitriche stagnalis* | Sand-Jensen et al. (1992)              | Tom Vindbæk Madsen,       | Not considered                  |
|                        |                                        | personal communication     |                                 |
| *Sparganium emersum*    | Sand-Jensen et al. (1992)              | Colmer and Pedersen (2008)  | Not considered                  |
| *Sparganium erectum*    | Sand-Jensen et al. (1992)              | Nielsen and Sand-Jensen (1989) | Not considered |
| *Sagittaria sagittifolia* | Sand-Jensen et al. (1992)             | Dina Ronzhina, personal communication | Not considered |
| **Aquatic** \( n = 7 \) |                                       |                           |                                 |
| *Lemna trisulca*        | Sand-Jensen et al. (1992)              | Dina Ronzhina, personal communication | Not considered |
| *Potamogeton perfoliatus* | Sand-Jensen et al. (1992)             | Spence et al. (1973)       | Not considered                  |
| *Elodea canadensis*     | Sand-Jensen et al. (1992)              | Madsen et al. (1996)       | Not considered                  |
| *Potamogeton crispus*   | Sand-Jensen et al. (1992)              | Nielsen and Sand-Jensen (1989) | Not considered |
| **Continued**           |                                       |                           |                                 |

\[ \text{AoB PLANTS} \ 2011: \ plr030; doi:10.1093/aobpla/plr030, \text{available online at} \text{www.aobplants.oxfordjournals.org} \  \text{© The Authors} 2011 \]
Appendix 2: List of several studies of underwater net photosynthesis \( (P_N) \) in terrestrial wetland plants or amphibious plants, in addition to Sand-Jensen et al. (1992) (see Appendix 1)

Species, CO\(_2\) concentrations, temperatures and light (PAR) regimes used for measurements of underwater \( P_N \) are listed. For multi-species studies of underwater \( P_N \) in submerged aquatic plants, see Sand-Jensen (1989), Reiskind et al. (1989) and Madsen et al. (1993b).

| Source | Species tested | \( \text{CO}_2 (\mu\text{M}) \) | Temperature \( (\text{°C}) \) | PAR \( (\mu\text{mol m}^{-2} \text{s}^{-1}) \) | Notes |
|--------|----------------|-----------------|-----------------|-----------------|-------|
| Nielsen (1993) | Barbarea stricta, Batrachium aquatile, Berula erecta, Callitriche cophocarpa, Cardamine amara, Catabrosa aquatile, Epilobium hirsutum, Glyceria maxima, Hydrocotyle vulgaris, Littorelia uniflora, Lobelia dortmanniana, Lotus uliginosus, Montia fontana, Myosotis palustris, Polygonum amphibium, Ranunculus repens, Sparganium emersum, Sparganium erectum, Veronica anagallis-aquatica, Veronica beccabunga | 100 \& 280 | 25 \& 24 | 600 | Also \( P_N \) rates in air, although some seem unusually high |
| Sand-Jensen and Frost-Christensen (1998) | Myosotis palustris, Sparganium emersum | 20 \& 280 | 12 \& 24 | 400 | Also effects of initial \( \text{O}_2 \) concentration and temperature on underwater \( P_N \) |
| Sand-Jensen and Frost-Christensen (1999) | Berula erecta, Menta aquatica, Myosotis palustris, Veronica anagallis-aquatica | 100 \& 700 | 15 | 350 | Also initial slope at \( P_N \) rate-limited \( \text{CO}_2 \) concentrations and \( P_{\text{max}} \) |
| Vervuren et al. (1999) | Arrhenatherum elatius, Phalaris arundinacea, Rumex crispus | 2,200 | 20 | 740 | Also \( P_N \) rates under water after 30 days of submergence |
| Nielsen and Nielsen (2006) | Lobelia cardinalis, Nesaea crassicaulis | 40 \& 1500 | 20 | 1200 | Also \( P_N \) rates in air |
| Mommer et al. (2007) | Rumex palustris | 10–10,000 | 20 | 400 | Also full \( \text{CO}_2 \) response curve and \( P_N \) rate in air |
| Pedersen et al. (2006) | Halosarcia pergranulata (syn. Tecticornia pergranulata) | 20–6800 | 20 | 1500 | Also \( P_N \) rate in air |
| Source               | Species tested                                                                 | CO₂ (µM) | Temperature (°C) | PAR (µmol m⁻² s⁻¹) | Notes                                                                 |
|---------------------|---------------------------------------------------------------------------------|----------|------------------|---------------------|----------------------------------------------------------------------|
| Colmer and Pedersen (2008) | Acorus calamus, Glyceria maxima, Phalaris arundinacea, Phragmites australis, Sparganium emersum, Typha latifolia | 50 and 500 | 20               | 600                 | Also full CO₂ response curve for Phragmites australis                |
| Pedersen et al. (2009) | Oryza sativa                                                                   | 15–2000  | 30               | 350                 | Also full CO₂ response curve under water and $P_N$ rate in air        |
| Pedersen et al. (2010) | Hordeum marinum                                                                | 18–2000  | 20               | 350                 | Also full CO₂ response curve under water and $P_N$ rate in air        |