Ablation of mpeg\(^+\) Macrophages Exacerbates mfrp-Related Hyperopia

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**Purpose.** Proper refractive development of the eye, termed emmetropization, is critical for focused vision and is impacted by both genetic determinants and several visual environment factors. Improper emmetropization caused by genetic variants can lead to congenital hyperopia, which is characterized by small eyes and relatively short ocular axial length. To date, variants in only four genes have been firmly associated with human hyperopia, one of which is MFRP. Zebrafish mfrp mutants also have hyperopia and, similar to reports in mice, exhibit increased macrophage recruitment to the retina. The goal of this research was to examine the effects of macrophage ablation on emmetropization and mfrp-related hyperopia.

**Methods.** We utilized a chemically inducible, cell-specific ablation system to deplete macrophages in both wild-type and mfrp mutant zebrafish. Spectral-domain optical coherence tomography was then used to measure components of the eye and determine relative refractive state. Histology, immunohistochemistry, and transmission electron microscopy were used to further study the eyes.

**Results.** Although macrophage ablation does not cause significant changes to the relative refractive state of wild-type zebrafish, macrophage ablation in mfrp mutants significantly exacerbates their hyperopic phenotype, resulting in a relative refractive error 1.3 times higher than that of non-ablated mfrp siblings.

**Conclusions.** Genetic inactivation of mfrp leads to hyperopia, as well as abnormal accumulation of macrophages in the retina. Ablation of the mpeg1-positive macrophage population exacerbates the hyperopia, suggesting that macrophages may be recruited in an effort help preserve emmetropization and ameliorate hyperopia.

Emmetropization is the precise regulation of size, morphology, and relative proportions of ocular tissues and is critical for proper refraction of light and thus clear vision. Improper emmetropization results in either myopia or hyperopia. Myopia is caused by a relative elongation of the axial length and is the more common and better studied refractive error. Hyperopia most often occurs when the axial length is too short for the focusing apparatus of the eye, resulting in light focusing behind the retina. Comparatively less is known regarding the mechanisms of hyperopia.

Although variants in many genomic regions are associated with changes in refractive error, variants in just a few genes have been associated with monogenic high hyperopia or nanophthalmos.\(^1\)\(^2\)\(^3\)\(^4\) Homozygous or compound heterozygous mutations of MFRP, which encodes membrane-type frizzled-related (MFRP) protein, are associated with microphthalmia, high hyperopia, foveoschisis, areas of retinal pigmented epithelium (RPE) atrophy, and optic disc drusen in humans.\(^1\)\(^2\)\(^5\)\(^6\)\(^7\)\(^8\)\(^9\)\(^10\)\(^11\) Evidence for the conservation of MFRP function comes from multiple animal models. First, two mouse models of spontaneous retinal degeneration, rd6 and rdX, have identified mutations in Mfrp as their cause.\(^12\)\(^13\) Although these mutations resulted in retinal degeneration, initial analysis of homozygous mutants did not find hyperopia. However, examination of the rd6 model using non-invasive imaging found that these mice indeed have slight decreases in axial length and that this effect could be rescued by gene therapy.\(^14\)\(^15\) Our lab utilized zebrafish to investigate the effects of mfrp mutation. In contrast to mice, zebrafish homozygous for mfrp mutations do not develop retinal degeneration but do recapitulate the pronounced hyperopia seen in humans homozygous for MFRP alterations.\(^16\) These findings illustrate the role of Mfrp in proper emmetropization and its functional conservation across multiple species.

Although both mouse and zebrafish models display hyperopic phenotypes in the absence of Mfrp, neither model fully recapitulates the full spectrum of human MFRP-related phenotypes, as mice appear to exhibit only small changes in eye size, failing to develop high hyperopia, and zebrafish mutants do not present photoreceptor degeneration. Intriguingly disruption of mfrp in both zebrafish and mice causes accumulation of subretinal macrophages.\(^12\)\(^16\)\(^17\)
Accumulation of retinal macrophages may also occur in human MFRP-related pathology. The presence of round yellow-white flecks has been documented in patients with MFRP-associated microphthalmia. Under fundus microscopy the subretinal macrophages present in the Mfrp mouse models also share this white fleck appearance. These observations suggest that accumulation of retinal macrophages is a unified feature of Mfrp mutations across species.

Based on the accumulation of macrophages seen in two distinct animal models of MFRP-related hyperopia, we hypothesized that those retinal macrophages, or macrophages within the eye in general, may function to regulate emmetropization. In order to test this hypothesis, we utilized established cell-specific ablation techniques in the zebrafish model. We found that, although ablation of macrophages in wild-type (WT) fish does not affect basal emmetropization, deletion of this population exacerbates the hyperopia observed in mfrp mutant fish. We also investigated changes in proliferation, cell death, and scleral collagen fiber morphology of mfrp mutant zebrafish.

Methods

Histology and Transmission Electron Microscopy

Eyes were fixed with 1.0% paraformaldehyde, 2.5% glutaraldehyde, and 3.0% sucrose in 0.06-M cacodylate buffer overnight at 4°C. Samples were then washed in cacodylate buffer and post-fixed with 1% osmium tetroxide and then dehydrated by a series of methanol washes. Larvae were infused with Epon 812 resin (Electron Microscopy Sciences, Hatfield, PA, USA) through two 15-minute acetone-trile washes followed by 1:1 acetone-trile:Epon incubation for 1 hour and 100% Epon incubation overnight. Finally, larvae were embedded in 100% Epon and hardened at 65°C for 24 hours. Then, 1-μm transverse serial sections through the length of the larvae were cut via microtome and stained with Toluidine Blue for light microscopy. Light microscopy images were taken using a NanoZoomer 2.0-HT (Hamamatsu Photonics K.K., Hamamatsu City, Japan). For transmission electron microscopy (TEM) analysis, 70-nm sections were cut, collected on hexagonal grids, and stained with uranyl acetate and lead citrate, followed by imaging on a Hitachi H-600 Transmission Electron Microscope (Hitachi, Ltd., Tokyo, Japan).

Paraffin Histology

Eyes utilized for paraffin histology were immersed in 4% paraformaldehyde overnight at 4°C and embedded in paraffin blocks for sectioning. Then, 4-μm sections were obtained and stained with hematoxylin and eosin for analysis, with serial unstained sections used for immunofluorescent staining. Unstained sections underwent de-paraffinization with xylens and an ethanol gradient prior to heated antigen retrieval in antigen retrieval solution (Dako; Agilent Technologies, Santa Clara, CA, USA). Immunofluorescent staining was then performed as follows.

Retina Flat-Mount Preparation

Retina flat-mount immunofluorescent staining was performed on dissected eyecups that were fixed overnight at 4°C in 4% paraformaldehyde. Before staining, the lens, cornea, and anterior chamber of the eye were dissected away to allow better access to the tissue. Retinae were washed in phosphate-buffered saline (PBS) to remove fixative. An identifying notch was cut into the dorsal side of the retina to maintain proper anatomical positioning. Four roughly equidistant cuts were made from the outside of the retina toward the middle in order to relieve tension and flatten the globe under a coverslip. Immunofluorescent staining was then performed as follows.

Immunofluorescence

For both paraffin sections and retina flat-mount preparations, standard immunostaining followed with 1-hour incubation in blocking solution (2% normal goat serum, 1% Triton X-100, and 1% Tween 20 in PBS). Larvae were incubated in primary antibodies overnight in blocking solution at room temperature or 4°C. Embryos were then washed three times for 1 hour in 1% Tween 20 in PBS. Antibody detection was performed using Invitrogen Alexa Fluor 488 and 568 conjugated secondary antibodies (Thermo Fisher Scientific, Waltham, MA, USA) at 1:800 dilution in blocking solution overnight at 4°C followed by washes with 1% Tween 20 in PBS. The following primary antibodies and concentrations were utilized:

- 1:200 mouse anti-4C4 (gift from Peter Hitchcock, University of Michigan)
- 1:500 rabbit anti-lymphocyte cytotoxic protein 1 (Lcp1, L-plastin; GTX124420; GeneTex, Irvine, CA, USA)
- 1:500 mouse anti-green fluorescent protein (GFP; JL-8; Takara Bio, Shiga, Japan)
- 1:500 ABfinity rabbit anti-GFP (G10362; Thermo Fisher Scientific)

Spectral Domain–Optical Coherence Tomography

Zebrafish eyes were imaged using an Envisu R2200 spectral-domain optical coherence tomography (SD-OCT) imaging system with a 12-mm telecentric lens (Biophtgen, Morrisville, NC, USA) using a Broadlighter T870 light source centered at 878.4 nm with a 186.3-nm band width (Superlum Diodes, Cork, Ireland). Axial length, lens diameter, and retinal radius were measured for populations of zebrafish at 56 days post-fertilization (dpf), as described previously. Both eyes were measured for each fish. In statistical analysis, only the right eye was utilized.

Eye and Body Length Measurement

The following zebrafish eye dimensions were measured: axial length (front of cornea to back of RPE), lens diameter (anterior surface of lens to posterior surface), and retinal radius (center of lens to the back of the RPE). Body length was measured from the tip of the head to the end of the trunk (before the caudal fin). Relative refractive error was calculated as 1 – (retinal radius/P), where idealized focal length F = lens radius × 2.324, using a coefficient extrapolated from a large population group (n = 240) plot of lens radius versus retinal radius. Relative refractive error values are unitless, with values lower than zero indicating that the eye is myopic (or that the observed distance from lens center to RPE is greater than the expected retinal radius) and values greater than zero indicating that the eye is hyperopic (or that...
the observed distance from lens center to RPE is less than the expected retinal radius). The relative refractive error is based on the following assumptions: (1) the refractive index of the fish lens is constant, and (2) the distance from the center of the lens to the RPE is equal to the focal length of the lens of WT fish.

**Metronidazole Treatment**

All fish used for macrophage ablation experiments were raised under normal conditions from 0 to 14 dpf. On 14 dpf, fish were randomly separated into control and experimental groups. Experimental groups were reared in stationary tanks with 7-mM metronidazole (MTZ; M3761-100G; Sigma-Aldrich, St. Louis, MO, USA) dissolved in fish facility water from 6:00 PM to 9:00 AM daily. From 9:00 AM to 6:00 PM, fish were returned to the circulating facility water in 3-L tanks for rearing and feeding. From 14 dpf to 21 dpf, fish in the experimental group were kept in 500 mL of MTZ for treatments, and 21-dpf and older fish were kept in 1 L of MTZ for treatments. It is important to note that fresh MTZ treatments occurred daily until the final time point. This consistent treatment was aimed at achieving consistent depletion of the targeted cell population from 14 dpf until 56 dpf. Untreated control groups were also moved to stationary tanks with equivalent volumes of water during MTZ treatments.

**Collagen Hybridizing Peptide Staining**

For collagen hybridizing peptide (CHP) staining, paraffin sections were used, and standard de-paraffinization with xylenes and an ethanol gradient were performed. Importantly, no antigen retrieval steps were taken in order to avoid denaturing or unfolding of collagenous proteins in the samples. Samples were then blocked in 5% goat serum for 20 minutes at room temperature. To stain the sample, a 20-mM CHP-Cy3 conjugate (60520; Echelon Biosciences, Salt Lake City, UT, USA) working solution diluted in PBS was used. Prior to staining, the working solution was heated at 80°C for 5 minutes and then rapidly brought to room temperature by placing it on ice for 30 seconds. The working solution was then quickly added to slides outlined with a PAP pen (Abcam, Cambridge, UK) and incubated overnight at 4°C. Counterstaining with 4’,6-diamidino-2-phenylindole (DAP) followed prior to imaging.

**Zebrafish**

All transgenic and mutant lines were generated and maintained in the ZDR genetic background originally purchased from Aquatica Biotech, Sun City Center, Florida. WT siblings or cousins were used as control groups. All animal experiments were approved by the Institutional Animal Care and Use Committee of the Medical College of Wisconsin.

**Transgenic Lines and Mutant Lines**

Tg(mpeg1:NTReYFP) is a transgenic line that utilizes an isolated 1.86-kb sequence of the promoter region of macrophage expressed gene 1 (mpeg1) identified by Ellett et al. to drive expression of an enhanced yellow fluorescent protein (eYFP)-tagged, human codon-optimized version of the *Escherichia coli* nitroreductase. This line has previously been used in conjunction with MTZ treatment for both short- and long-term ablation of macrophages.20–22

*mfrp<sup>MW78</sup>* is a mutant line that contains a 5-bp deletion in exon 8 of zebrafish *mfrp* and is predicted to change the reading frame of Mfrp protein causing premature truncation. Mfrp immunostaining on cryosections previously indicated a loss of protein detection by an Mfrp-specific antibody, supporting the notion that this mutation leads to complete loss of Mfrp function.16

**Results**

To address the role of macrophages in emmetropization, we sought to deplete the macrophage population during ocular growth and assess potential changes in axial length and refractive error. We chose zebrafish as our model organism because they display a larger shift in refractive state upon *mfrp* deletion than the *Mfrp* mutant mouse models, allowing for easier detection of phenotypic change.15,16 For efficient macrophage ablation we used an established chemical-genetic system.23 Using macrophage promoter mpeg1 we expressed bacterial nitroreductase (NTR) fused to eYFP specifically in macrophages. Fish carrying this transgene are termed mpeg1:NTR-eYFP.15 On its own, NTR expression does not harm cells; however, it converts the nontoxic prodrug MTZ into a cytotoxic metabolite that results in autonomous DNA crosslinking and subsequent cell death. This system has been effectively utilized for the ablation of multiple cell types in zebrafish.15

**Ablation of Macrophages in WT Zebrafish Did Not Significantly Impact Emmetropization**

To determine the best temporal strategy for macrophage depletion we had previously observed ocular growth of WT and *mfrp<sup>−/−</sup>* fish over time by SD-OCT.16 In both populations, we found that 15 to 84 dpf encompassed the highest growth rates in ocular anatomy, as well as overall body size.16 Prior to 31 dpf, no differences in ocular biometry or relative refractive error were seen between WT and *mfrp<sup>−/−</sup>* eyes; at 31 dpf and following, *mfrp<sup>−/−</sup>* eyes begin to show significant decreases in axial length, lens diameter, retinal radius, and body length relative to WT fish.16 By 31 dpf, WT eyes reach a stable relative refractive error measurement that continued through at least 149 dpf.16 In contrast, at 31 dpf, *mfrp<sup>−/−</sup>* eyes exhibited slightly higher relative refractive error, and this difference was accentuated at 84 dpf, when the magnitude of difference between WT and *mfrp* relative refractive error appears to level off.16 Given this information, we chose to ablate macrophages starting at 14 dpf and continuing through 56 dpf. After 14 dpf, treatment was chosen to allow for the normal development and organization of the retina, avoiding potential adverse effects of macrophage ablation on early development and focusing solely on ocular growth as it pertains to eye size and refractive state. We chose 56 dpf as the endpoint because it represents a time when WT eyes have reached an equilibrium regarding relative refractive error, but it is also a period when active emmetropization is still occurring.23

To examine the effects of mpeg1: eYFP cell ablation on emmetropization, we allowed mpeg1:NTR-eYFP<sup>−/−</sup> and their non-transgenic WT cousins to grow to 8 weeks of age, or more precisely 56 dpf, with or without MTZ treatment. MTZ was administered through daily bath applications.
Macrophages Modulate Hyperopia

FIGURE 1. Ablation of macrophages in WT zebrafish did not significantly impact emmetropization. (A) Representative SD-OCT B-scans from the center of mpeg1:NTR-eYFP+ eyes and their WT cousins with or without MTZ treatment. (B) Axial length, lens diameter, retinal radius, and body length of mpeg1:NTR-eYFP+ fish and their WT cousins with or without MTZ treatment. (C) Axial length normalized to body length and lens diameter. (D) Relative refractive error. Two-way ANOVA was used for statistical analysis for (B) to (D). P values are shown from Tukey’s multiple comparisons for post hoc analysis. Red bars indicate statistical significance likely due to mpeg1+ cell ablation.*P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001. For mpeg1:NTR-eYFP+ untreated, n = 11; for mpeg1:NTR-eYFP+ MTZ treated, n = 7; for WT cousins untreated, n = 13; for WT cousins MTZ treated, n = 9.

Macrophage Ablation in mfrp Mutants

MTZ treatment resulted in overall smaller zebrafish, as measured by decreases in axial length, lens diameter, retinal radius, and body length (Fig. 1B). This was true in both the presence and absence of mpeg1:NTR-eYFP expression, suggesting that MTZ treatment causes developmental delay or slower overall body growth independent of cell-specific ablation. After normalization to body length, no changes in axial length were detectable (Fig. 1C). We also normalized to lens diameter, and, although mpeg1:NTR-eYFP+ MTZ-treated eyes had significantly shorter ratios of axial length to lens diameter than their non-transgenic, MTZ-treated cousins, neither MTZ-treated group had significant changes relative to their untreated siblings (Fig. 1C). To more specifically assess the refractive state of these eyes, we calculated relative refractive error as described previously.9 No significant differences in relative refractive error were observed (Fig. 1D). Taken together, these results suggest that mpeg+ macrophage ablation does not significantly alter emmetropization in WT zebrafish.

Although macrophage ablation did not significantly alter the relative refractive error of WT zebrafish, the previously documented macrophage accumulation in mfrp mutants led us to hypothesize that they may still play a role in modulating the
FIGURE 2. Efficient macrophage ablation in the mfrp retina. (A–D) mpeg1:NTR-eYFP expression in WT and mfrp–/– retina flat-mount preparations with and without MTZ treatment. (A’–D’) Immune staining for 4C4 antibody marking macrophages. (A”–D”’) Merged and colorized images of A’–D’, with blue representing 4C4 and yellow representing YFP. (A”’–D”’) Higher magnification images of A”–D”. (E, F) Cell counts of mpeg1:NTR-eYFP+ cells (E) or 4C4+ cells (F) in WT and mfrp–/– retina flat-mount preparations with and without MTZ treatment. Two-way ANOVA was used for statistical analysis. Sidak’s multiple comparisons were used for post hoc analysis. **P < 0.01, ***P < 0.001, ****P < 0.0001. For WT untreated, n = 5; for WT MTZ treated, n = 2; for mfrp–/– untreated, n = 5; for mfrp–/– MTZ treated, n = 6.

pathologic hyperopic state. To evaluate this, we employed the same macrophage ablation strategy on mfrp mutant and heterozygous siblings. Again, highly efficient ablation of WT macrophages was observed by mpeg1:NTR-eYFP expression, as well as using the 4C4 antibody, which labels macrophages and microglial cells in zebrafish (Figs. 2A–2B”’). Although nearly all transgene expressing cells were depleted, some 4C4+ cells escaped ablation, likely due to incomplete overlap in expression of the 4C4 antigen and mpeg1 transgene. As expected, mfrp mutant fish showed significant increases in mpeg1:NTR-eYFP+ cells, with focal increases at the central retina (Figs. 2C–2C”). In mfrp–/–
To obtain more detailed spatial information regarding the macrophage accumulation in mfrp mutants, as well as their ablation, we performed immunofluorescent staining on histological sections. We labeled eYFP+ cells using a GFP antibody and co-stained for Lcp1 as an additional macrophage marker.25 Nuclei were stained with DAPI to identify the retinal layers. In mfrp+/−; mpeg1:NTR-eYFP+ untreated control fish, eYFP+ cells were seen sporadically across all layers of the retina. The ganglion cell layer and inner nuclear layer (INL) contained the highest frequency of these cells, but eYFP+ cells would also be found in the outer nuclear layer (ONL) and photoreceptor layer (L). (M–O) Quantification of the number of eYFP+ cells per μm2 in the ganglion cell layer (M) and inner (N) and outer (O) nuclear layer. Error bars represent standard deviations. Two-way ANOVA was used for statistical analysis for (I) to (O). P values are shown from Tukey's multiple comparisons for post hoc analysis. *P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001. For mfrp+/− untreated, n = 4; for mfrp+/− MTZ treated, n = 4; for mfrp−/− untreated, n = 4; for mfrp−/− MTZ treated, n = 5.

Fish, we saw depletion of mpeg1:NTR-eYFP-expressing cells; however, the effect was not as complete as in WT fish (Figs. 2C–2E). The 4C4 cell counts were not significantly decreased (Figs. 2C', 2D', 2F). Although the ablation of our mpeg1:NTR-eYFP-expressing cells remained efficient and significant, these results reveal that some macrophages that accumulate in mfrp−/− eyes remained.

![Figure 3](image_url)

**Figure 3.** Distribution of macrophage accumulation and ablation across mfrp+/− and mfrp−/− retinae. (A–H) Representative images of central retina sections from mfrp+/−; mpeg1:NTR-eYFP+ and mfrp−/−; mpeg1:NTR-eYFP+ MTZ treated and untreated fish. (A–H) Grayscale DAPI images at low magnification (A, C, E, G) and high magnification (B, D, F, H). (A'-H') Grayscale eYFP images at low magnification (A', C', E', G') and high magnification (B', D', F', H'). (A''-H'') Grayscale Lcp1 images at low magnification (A'', C'', E'', G'') and high magnification (B'', D'', F'', H''). Colorized merged images with DAPI are shown in blue, eYFP in yellow, and Lcp1 in magenta; images are at low magnification (A''', C''', E''', G''') and high magnification (B''', D''', F''', H''').
nuclear layer (ONL). In control fish, nearly all eYFP+ cells observed were also Lcp1+ (Figs. 3A–3B'', 3I–3K, 3M–3O). The strong endogenous fluorescence within the photoreceptor layer precluded accurate quantitation of eYFP+ cells in this layer; however, Lcp1 staining showed small numbers of macrophages present there, as well (Fig. 3I). MTZ treatment efficiently reduced or completely depleted eYFP+ cells across all layers of mfrp–/–; mpeg1:NTR-eYFP+ eyes (Figs. 3C–3D'', 3M–3O). Though seemingly reduced, sporadic Lcp1+ cells were noted across all layers of the retina (Figs. 3E–3F'''). However, statistical analysis showed a significant increase in the frequency of both eYFP+ and Lcp1+ cells only in the ONL (Figs. 3K, 3O). MTZ treatment was similarly effective in mfrp–/–; mpeg1:NTR-eYFP+ fish, as eYFP+ cells were greatly reduced or completely depleted across all layers of the retina (Fig. 3M–3O). Lcp1+ cells were also decreased in the INL and ONL (Figs. 3J, 3K). These data highlight the efficient ablation of macrophages across all layers of the eye and identify the ONL as the specific location of increased macrophage presence in mfrp–/– fish.

### Macrophage Ablation Exacerbated Hyperopia in mfrp–/– Zebrafish

We again utilized SD-OCT to image and measure the various metrics of mfrp–/–; mpeg1:NTR-eYFP+ eyes compared with the eyes of mfrp–/–; mpeg1:NTR-eYFP+ siblings with and without MTZ treatment (Fig. 4A). As in WT fish, MTZ treatment slowed overall growth in both mfrp–/– and mfrp–/– fish (Fig. 4B, body length). This effect was independent of mpeg1:NTR-eYFP expression (Supplementary Figs. S2A, S2B).

To control for the overall growth effect of MTZ treatment and possible genotype effects specific to mfrp, we MTZ treated mfrp–/–, and mfrp–/– fish lacking the mpeg1:NTR-eYFP transgene and again imaged and measured the various eye and body metrics. We found that MTZ treatment affected axial length normalized to body length in these non-transgenic animals (P = 0.0043; 2-way analysis of variance

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**Figure 4.** Macrophage ablation exacerbated hyperopia in mfrp–/– zebrafish. (A) Representative SD-OCT B-scans from the center of mfrp+/–; mpeg1:NTR-eYFP+ eyes and their mfrp–/–; mpeg1:NTR-eYFP+ siblings with or without MTZ treatment. (B) Eye size metrics of mfrp+/–; mpeg1:NTR-eYFP+ and mfrp–/–; mpeg1:NTR-eYFP+ fish with or without MTZ treatment. (C) Axial length normalized to body length and lens diameter. (D) Relative refractive error. Two-way ANOVA was used for statistical analysis. P values are shown from Tukey’s multiple comparisons for post hoc analysis. Error bars represent standard deviation. Red bars indicate statistical significance due to MTZ treatment. *P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001. For mfrp–/– untreated, n = 11; for mfrp–/– MTZ treated, n = 14; for mfrp–/– untreated, n = 15; for mfrp–/– MTZ treated, n = 9.
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**FIGURE 5.** Macrophage ablation did not significantly alter retina morphology. (A–B') Hematoxylin and eosin-stained paraffin histology of the central retina in mfrp\(^+/−\); mpeg1:NTR-eYFP\(^+\) (A, A') and mfrp\(^−/−\); mpeg1:NTR-eYFP\(^+\) (B, B') with and without MTZ treatment.

[ANOVA] (Supplementary Fig. S2C). Although this might suggest a potential nonspecific effect of MTZ treatment on relative eye size, differences were not observed for axial length normalized to lens diameter in these non-transgenic control fish (\(P = 0.3664\), 2-way ANOVA) (Supplementary Fig. S3C). Given that body length does not impact emmetropization, while lens diameter directly impacts this process, we concluded that MTZ treatment alone does not alter emmetropization in a nonspecific fashion. Further supporting this conclusion, relative refractive error was not significantly affected by MTZ treatment in these non-transgenic controls (\(P = 0.3664\), 2-way ANOVA).

As expected, mfrp\(^−/−\) fish had significantly reduced relative eye size compared with their mfrp\(^+/−\) siblings when normalized to either body length or lens diameter (\(P < 0.0001\), 2-way ANOVA) (Fig. 4C). Relative refractive error was also significantly increased in mfrp\(^−/−\) fish compared with mfrp\(^+/−\) siblings, as previously reported (Fig. 4D).\(^{16}\) These results confirm that loss of mfrp leads to hyperopia in zebrafish and reveal that mpeg\(^−/−\) cell ablation does not rescue this phenotype.

When we assessed the effect of macrophage ablation on relative eye size we found that MTZ treatment had significant effects (Fig. 4C, axial length to lens diameter; \(P < 0.0001\), 2-way ANOVA). Post hoc analysis confirmed a significant increase in relative refractive error in mfrp\(^−/−\); mpeg1:NTR-eYFP\(^+\) fish compared with their untreated mfrp\(^−/−\) siblings (Fig. 4D). Again, this was genotype specific, with mfrp\(^+/−\); mpeg1:NTR-eYFP\(^+\) siblings showing no significant changes due to MTZ treatment. These results demonstrate that macrophage ablation significantly exacerbates the hyperopic phenotype of mfrp mutant zebrafish, indicating that the accumulation of macrophages seen in mfrp-related hyperopia may be recruited to ameliorate the microphthalmia.

**Macrophage Ablation Did Not Significantly Alter Retina Morphology**

As mpeg1\(^+/−\) cell ablation affected relative eye size and refractive error, we sought to gain possible mechanistic insight into these changes by assessing overall eye and retinal morphology. We assessed the effects of macrophage ablation on overall retina morphology by histology. Hematoxylin and eosin–stained paraffin sections revealed normal retinal morphology following macrophage ablation (Figs. 5A–5B'). Neither proliferation nor cell death underlay exacerbated mfrp-related hyperopia after macrophage ablation.

**Neither Proliferation Nor Cell Death Underlay Exacerbated mfrp-Related Hyperopia After Macrophage Ablation**

We hypothesized that the exacerbated changes in relative eye size seen in mfrp mutants could be due to altered cell
proliferation and/or death. To assess changes in proliferation we performed immunohistochemistry for proliferative cell nuclear antigen (PCNA), along with DAPI to label nuclei. In mfrp+/–, mpeg1:NTR-eYFP+ eyes, PCNA+ nuclei were found frequently within both the INL and ONL, as well as in the ciliary marginal zone. In comparison, mfrp–/–, mpeg1:NTR-eYFP+ fish had PCNA+ cell frequency within the INL similar to that of their heterozygous siblings but significantly decreased frequency of PCNA+ nuclei in the ONL (Figs. 6A–6B’, 6E–6F’, 6I, 6J). No changes in the ciliary marginal zone were noted under any condition. MTZ treatment did significantly decrease the percentage of PCNA+ nuclei in the ONL of mfrp+/–, mpeg1:NTR-eYFP+ fish compared with their untreated siblings (Figs. 6C–6D’, 6J). PCNA analysis also revealed that mfrp−− had reduced numbers of proliferative cells in the ONL, although, MTZ treatment on mfrp−−, mpeg1:NTR-eYFP+ animals did not exacerbate this effect (Figs. 6G–6H’, 6J). As an alternative to changes in cell proliferation, cell death caused by mpeg+ cell ablation could lead to the decrease in relative eye size observed in mfrp mutants. To assess cell death, we performed terminal deoxynucleotidyl transferase dUTP nick end labeling (TUNEL). We found little or no TUNEL-positive cells regardless of genotype or treatment condition, suggesting that apoptosis did not cause the altered phenotype in mfrp mutant eyes (Supplementary Figs. S3A–S3C’’). Consistent with this result, DAPI staining did not reveal pyknotic nuclei for any genotype.
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Recent improvement in the detection of collagenous proteins has come in the form of CHPs, which specifically hybridize to degraded or unfolded collagen chains. The collagen triple helix structure is made of repeating glycine–X–Y repeats, where the X and the Y are most often proline and hydroxyproline. CHP contains a repeating glycine–proline–hydroxyproline amino acid triplet which gives it high binding affinity that the RPE of mfrp–/– mice following macrophage ablation, as we did not find significant changes in fibril diameter between the MTZ-treated mfrp–/–; mpeg1::NTR-eYFP+ and their untreated mfrp–/–; mpeg1::NTR-eYFP+ siblings.

Macrophage Ablation Altered Collagen Accessibility to a CHP

To assess collagen accessibility in both untreated and MTZ-treated conditions, we used a Cy3-conjugated CHP to stain mfrp–/–; mpeg1::NTR-eYFP+ and mfrp–/–; mpeg1::NTR-eYFP+ paraffin sections and counterstained with DAPI for orientation (Figs. 8A–8C). We quantified the mean pixel intensity (P = 0.001) and the posterior sclera analysis (P = 0.00204), genotype had a significant effect on the mean pixel intensity (2-way ANOVA), with mfrp–/– eyes appearing to have slight decreases in R-CHP intensity. In the whole sclera analysis, MTZ treatment also had a significant effect on mean pixel intensity (P = 0.022, 2-way ANOVA). Applying post hoc statistical analysis, we found that mfrp–/–; mpeg1::NTR-eYFP+ fish treated with MTZ exhibited significantly less R-CHP staining intensity than their untreated mfrp–/–; mpeg1::NTR-eYFP+ siblings (P = 0.0319, Sidak’s multiple comparison) (Fig. 8D). These results suggest that mfrp–/– fish have lower levels of denatured or unfolded collagens than their mfrp–/+ siblings and that macrophage ablation exacerbates this difference. This change in collagen architecture might underlie the exacerbated hyperopia seen in mfrp–/– eyes after macrophage ablation.

Discussion

We found that depletion of macrophages exacerbated mfrp–/– related hyperopia in zebrafish. Additionally, we observed that, although macrophages affect the development of mfrp–/– related hyperopia, the absence of the mpeg-1 population did not significantly alter WT emmetropization. We went on to investigate further how macrophage ablation might affect morphology and growth of retina and sclera, examining changes in retinal cell proliferation and changes in scleral collagen fibers.

We noted the nonspecific effects of MTZ treatment on the overall size of the fish and accounted for these changes in our analysis. MTZ could cause additional non-obvious effects. MTZ itself is an antibiotic that was originally used for the treatment of trichomoniasis, and it continues to be used to combat anaerobic infections. Daily antibiotic washes on zebrafish likely have a profound effect on the microbiome of the fish and could result in unintended effects. Metronidazole has also been shown to affect circadian rhythm, increasing the expression of core clock genes in the skeletal muscle of germ-free mice. However, this MTZ-based ablation technique has been used to study circadian rhythms of visual sensitivity in zebrafish, and no alterations to circadian rhythm were reported in non-transgenic MTZ treated fish. To control for potential nonspecific effects of MTZ, all non-transgenic control groups were treated exactly as the transgene-expressing experimental groups and underwent the same MTZ treatment regimen.

Notably macrophage ablation only affected emmetropization in the context of mfrp–/– hyperopia and showed no effect in WT conditions. This finding led us to hypothesize that macrophages are recruited in mfrp–/– eyes to help preserve proper refractive state. How the loss of Mfrp, a protein expressed primarily in the RPE and ciliary epithelium, results in macrophage recruitment remains an open question. Recently, zebrafish RPE were shown to express leukocyte recruitment factors among other immune-related genes following genetic ablation. Leach et al. went on to show that the macrophage and microglia cells that are recruited are also required for proper RPE regeneration, demonstrating that macrophages play a crucial role in the health of the RPE following injury. This raises the possibility that the RPE of mfrp–/– zebrafish exists in an injury-like state leading to recruitment of macrophages. Interestingly, macrophage depletion in mice via colony stimulating factor 1 receptor inhibition was associated with loss of proper RPE structure, which could be rescued following repopulation of macrophages. Studies such as these highlight crucial signaling between these two populations of cells in RPE maintenance and regeneration. Our data demonstrate that this crosstalk between RPE and macrophages also affects pathological hyperopia in the context of loss of mfrp.
Macrophages Modulate Hyperopia

Our ablation technique is limited to the cells expressing the *mpeg1*:NTR-eYFP transgene. In that context, this broad macrophage population is efficiently ablated by the action of the transgene. Still, 4C4+ macrophages remained following MTZ treatment, suggesting that non-*mpeg*1+ macrophages persist. Likewise, our histological analysis demonstrated that Lcp1+ macrophages also remained, although at reduced numbers. These findings highlight the heterogeneity of the population of monocyte-like cells in the retina, whether macrophage or microglia. It is clear that the *mpeg1*:NTR-eYFP transgene, although marking a broad population of macrophages, does not express in all retinal monocytes. Notably, both *mpeg1* positive and negative macrophages were elevated in the *mfrp* mutant retina. Future work investigating the importance of macrophage subtypes could prove useful for identifying their possible roles in emmetropization and potential additional effects on *mfrp* mutant pathogenesis. It seems likely that either the loss of Mfrp or the condition of hyperopia alters the state of macrophages in the retina or the recruitment of a separate subpopulation.

Finally, we found modulation of the collagen architecture following macrophage ablation. Recent work suggests that collagen bundle size is dynamically regulated during emmetropization, and these alterations in size can change...
their mechanical properties. Recently, alterations in scleral collagen dynamics have been partially attributed to an increased presence of scleral macrophages in a visual form-deprived mouse model of myopia. These investigators reported that macrophages appeared to be recruited to the sclera of form-deprived myopic mice by increased scleral expression of C-C motif chemokine ligand-2. The authors suggested that, upon recruitment, the macrophages are then partially responsible for the secretion of matrix metalloproteinase-2 in the sclera, resulting in extracellular matrix remodeling that contributes to the development of myopia. One possible hypothesis that stems from these data is that, as the retina grows, it exerts force on the sclera, and collagen remodeling alters scleral compliance, allowing axial elongation of the eye. In the case of mfrp mutant eyes, increased collagen bundle diameter may result in increased tensile stiffness of the sclera and prevent proper axial elongation. Indeed, we observed an upward trend in the percentage of collagen fibrils with larger diameter in mfrp−/− sclera when compared with their mfrp+/− siblings. In further support of these ideas, retinal folds are observed in mfrp−/− eyes. Perhaps the retina itself continues to expand while the sclera cannot, and this forces the retina to fold in on itself. This is relevant to the foveoschisis seen in some patients with MFRP-related microphthalmia. Furthermore, MTZ treatment resulted in a clear shift of collagen fibril diameter distribution in mfrp+/−, mpeg1:NT-eYFP+ fish, suggesting that the presence of macrophages can affect collagen fibril size. Supporting a direct role for macrophages, it has been reported that macrophages have the ability to directly secrete collagen in the context of scar formation after cardiac injury in zebrafish. Further work is required to define the precise relationship between collagen fibril size and emmetropization, as it currently remains unclear if an alteration to scleral collagen in mfrp mutants is an underlying cause of improper refractive state or simply a response to that state.

Our data suggest that, although the presence or absence of macrophages can result in altered collagen fibril diameter, alteration alone is not sufficient to modulate axial length and affect emmetropization. To further examine the collagen architecture of the sclera we utilized a fluorescently conjugated CHP to assess collagen accessibility. Based on the significant effects of both genotype and MTZ treatment in this assay, we suggest that macrophage ablation resulted in a more rigid collagen extracellular matrix in mfrp−/− sclera. Less unfolded or denatured collagens may result in increased tensile stiffness of the sclera and prevent proper axial elongation. Indeed, we observed an upward trend in the percentage of collagen fibrils with larger diameter in mfrp−/− sclera when compared with their mfrp+/− siblings. In further support of these ideas, retinal folds are observed in mfrp−/− eyes. Perhaps the retina itself continues to expand while the sclera cannot, and this forces the retina to fold in on itself. This is relevant to the foveoschisis seen in some patients with MFRP-related microphthalmia. Furthermore, MTZ treatment resulted in a clear shift of collagen fibril diameter distribution in mfrp+/−, mpeg1:NT-eYFP+ fish, suggesting that the presence of macrophages can affect collagen fibril size. Supporting a direct role for macrophages, it has been reported that macrophages have the ability to directly secrete collagen in the context of scar formation after cardiac injury in zebrafish. Further work is required to define the precise relationship between collagen fibril size and emmetropization, as it currently remains unclear if an alteration to scleral collagen in mfrp mutants is an underlying cause of improper refractive state or simply a response to that state.

Macrophages Modulate Hyperopia

**Figure 8.** Macrophage ablation alters scleral accessibility to a collagen hybridizing peptide. (A–C) Representative images of central retina sections from mfrp+/−; mpeg1:NT-eYFP+ and mfrp−/−; mpeg1:NT-eYFP+ MTZ treated and untreated fish. (A–A′) Grayscale CHP images. (B–B′) Colorized and merged images with DAPI in cyan and CHP in red. (C–C′) High-magnification images of regions of interest in the posterior sclera as indicated by white outlines in (B) to (B′). (D, E) Quantification of the mean pixel intensity of the CHP staining across full sclera (D) and at central posterior sclera (E). For whole sclera (D), both genotype (P = 0.001) and MTZ treatment (P = 0.022) had a significant effect. Two-way ANOVA was used for statistical analysis. P values are shown from Sidak’s multiple comparisons for post hoc analysis. Error bars represent standard deviations. *P < 0.05. For mfrp+/− untreated, n = 6; for mfrp−/− MTZ treated, n = 6; for mfrp+/− untreated, n = 6; for mfrp−/− MTZ treated, n = 5.

| mfrp+/−; mpeg1:NT-eYFP+ | mfrp−/−; mpeg1:NT-eYFP+ |
|-------------------------|-------------------------|
| Untreated | MTZ treated | Untreated | MTZ treated |
| CHP | CHP | CHP | CHP |
| DAPI, CHP | DAPI, CHP | DAPI, CHP | DAPI, CHP |
macrophages in pathologic hyperopia and, more specifically, mfrp-related hyperopia in zebrafish.

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