Endocannabinoid 2-arachidonoylglycerol self-administration by Sprague-Dawley rats and stimulation of in vivo dopamine transmission in the nucleus accumbens shell

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2-Arachidonoylglycerol (2-AG) is the most potent endogenous ligand of brain cannabinoid CB1 receptors and is synthesized on demand from 2-arachidonate-containing phosphoinositides by the action of diacylglycerol lipase in response to increased intracellular calcium. Several studies indicate that the endocannabinoid (eCB) system is involved in the mechanism of reward and that diverse drugs of abuse increase brain eCB levels. In addition, eCB are self-administered (SA) by squirrel monkeys, and anandamide increases nucleus accumbens (NAc) shell dopamine (DA) in rats. To date, there is no evidence on the reinforcing effects of 2-AG and its effects on DA transmission in rodents. In order to fill this gap, we studied intravenous 2-AG SA and monitored the effect of 2-AG on extracellular DA in the NAc shell and core via microdialysis in male Sprague-Dawley rats. Rats were implanted with jugular catheters and trained to self-administer 2-AG [25 mg/kg/inf intravenously (iv)] in single daily 1 h sessions for 5 weeks under initial fixed ratio (FR) 1 schedule. The ratio was subsequently increased to FR2. Active nose poking increased from the 6th SA session (acquisition phase) but no significant increase of nose pokes was observed after FR2. When 2-AG was substituted for vehicle (25th SA session, extinction phase), rate responding as well as number of injections slowly decreased. When vehicle was replaced with 2-AG, SA behavior immediately recovered (reacquisition phase). The reinforcing effects of 2-AG in SA behavior were fully blocked by the CB1 receptor inverse agonist/antagonist rimonabant (1 mg/kg intraperitoneally, 30 min before SA session). In the microdialysis studies, we observed that 2-AG (0.1–1.0 mg/kg iv) preferentially stimulates NAc shell as compared to the NAc core. NAc shell DA increased by about 25% over basal value at the highest doses tested (0.5 and 1.0 mg/kg iv). The results obtained suggest that the eCB system, via 2-AG, plays an important role in reward.

Keywords: endocannabinoids, 2-arachidonoylglycerol, self-administration, reward, drug addiction, in vivo microdialysis, nucleus accumbens

INTRODUCTION

Endocannabinoid (eCB) signaling controls various central functions in mammals, such as nociception, feeding, energy homeostasis, mood, learning, memory, growth, development, and reward processes (1–6). The eCB system consists of cannabinoid receptors (CB1 and CB2), lipid-derived endogenous ligands [N-arachidonoylethanolamine or anandamide, 2-arachidonoylglycerol (2-AG)], and specific enzymes involved in the biosynthesis and degradation of eCB.

Many questions on eCB signaling still need to be answered. However, among the two endogenous ligands, anandamide and 2-AG, 2-AG appears to be the most potent endogenous ligand for brain cannabinoid CB1 receptors (7). 2-AG acts as a full agonist of CB1 and CB2 receptors (8–10), and is synthesized on demand from 2-arachidonate-containing phosphoinositides by the action of diacylglycerol lipase (DAGLα and DAGLβ) in response to elevations of intracellular calcium. Unlike classic neurotransmission systems (e.g., monoaminergic, cholinergic, aminoacidergic), 2-AG signaling in the nervous system takes place in a retrograde fashion. Thus, stimulation of the postsynaptic neuron induces...
the biosynthesis of 2-AG that thus diffuses to the presynaptic terminal to act on CB1 receptors; due to its lipophilic nature, no synaptic vesicles for storage of 2-AG exist (4, 11, 12). Given these characteristics, 2-AG is considered to be a neuromodulator rather than a transmitter. CB1 activation of Glu proteins inhibits neurotransmission but the net inhibitory or excitatory effect of 2-AG signaling depends on the type of neurons involved in the process (4, 13–16). 2-AG is mainly degraded by monoacylglycerol lipase (MAGL), a membrane-associated, cytoplasm-facing soluble enzyme located pre-synaptically at axon terminals along with the CB1 receptor (17–19). Furthermore, about 15% of 2-AG is hydrolyzed by ABHD6 and ABHD12, and the remaining 1–2% by fatty acid amide hydrolase (FAAH) (20). The mechanisms of 2-AG neuronal reuptake are not completely known, but putative transporters have been described (3, 21).

Drugs of abuse affect brain eCB levels and it has been proposed that the activation of the eCB system is involved in many aspects of addiction (22). Drug-induced increases in eCB synthesis play a role in drug reward, and it has been suggested that eCBs are involved in long-term synaptic plasticity of neural substrates of motivation and reward in relation to addiction (23).

In human beings, a genetic disruption of eCB clearance is associated with drug abuse (24, 25). CB1 receptor activation affects the addictive properties of opioids, cocaine, alcohol, and nicotine (26). On the other hand, CB1 KO mice show reduced ethanol and morphine self-administration (SA) and attenuated ethanol-and opiate-induced place conditioning (27–29). In rats, the CB1 receptor inverse agonist/antagonist SR-141716A (Rimonabant) reduces ethanol and opiate SA (30–32) while in mice it reverses the behavioral and neurochemical effects of cocaine (33).

Δ9-Tetrahydrocannabinol (THC), the main active component of marijuana, is known to reduce the severity of opioid withdrawal in human beings and rodents, and acute inhibition of FAAH or MAGL alleviates symptoms of precipitated and spontaneous withdrawal in opioid- and THC-dependent mice (34, 35). The eCB system is involved in the rewarding effects of THC (22), as well as ethanol and opiates (36, 37). In particular, ethanol and heroin SA increases eCB levels in the nucleus accumbens (NAc) shell, suggesting a role for NAc eCB transmission in the reinforcing effects produced by these drugs (38). Finally, eCBs are SA by squirrel monkeys, drugs that are effective reinforcers of drug-taking behavior (39, 40).

Dopamine (DA) neurotransmission is known to regulate a large number of motivated and addiction-related behaviors (41–43). Importantly, drugs of abuse of the most diverse pharmacological classes increase mesolimbic DA transmission in rats as well as in human beings preferentially or, depending on doses and conditions, selectively in the ventral striatum/NAc (42, 44–48). Accordingly, THC and the synthetic cannabinoid WIN 55,212-2 increase extracellular DA concentrations in the shell, but not in the core of the NAc, both when injected intravenously (iv) (49) and when SA by different strains of rats (50, 51). In addition, anandamide increases NAc shell DA and its effect is amplified by the FAAH inhibitor, URB597 (52). On the other hand, reduced DA transmission in the mesolimbic system is associated with spontaneous or rimonabant-precipitated THC withdrawal (53, 54).

In spite of this extensive literature, no studies are available on the reinforcing effects of 2-AG and its effects on DA transmission in rodents. In order to fill this gap, we studied intravenous 2-AG SA and monitored the effect of 2-AG on extracellular DA in the NAc shell and core via microdialysis in male Sprague-Dawley rats.

MATERIALS AND METHODS

ANIMALS

Male Sprague-Dawley rats (Harlan, Italy), weighting 250–275 g upon arrival, were housed four per cage given ad libitum access to food and water in a temperature (22°C) and humidity (60%) controlled vivarium with a 12 h light/dark cycle (on 08:00 A.M., off 08:00 P.M.). After surgery (catheter implantation), rats were individually housed in plastic cages (30 cm × 20 cm × 20 cm) given ad libitum food and water access, and in the same environmental conditions. For 7–10 days before surgery, rats were handled twice a day. SA sessions were performed during the light phase, between 9:00 a.m. and 5:00 p.m. After the experimental sessions, the rats were returned to their home cages where a daily ration of 18 g of food was made available, which maintained body weights at stable levels throughout these studies. The weight of rats at the beginning of SA studies was 300–325 g. Rats were weighed every day for the duration of the SA experiments. No significant reduction of body weight was observed. All experimental procedures met the guidelines and protocols approved by Italian (D.L. 116/92 and 152/06) and European Council directives (609/86 and 63/2010) and in compliance with the approved animal policies by the Ethical Committee for Animal Experiments (CESA, University of Cagliari) and the Italian Ministry of Health.

DRUGS

The eCB 2-AG was purchased from Tocris Cookson Ltd. (Northpoint, UK) and was dissolved in a vehicle containing 2% ethanol, 2% Tween 80, and saline and administered as an intravenous bolus of 20 µl for SA studies (12.5, 20, 50 µg/kg/infusion) or 1 ml/kg solution for microdialysis studies (0.1–1 mg/kg iv).

The CB1 receptor inverse agonist/antagonist rimonabant (SR-141716A) was obtained from Sigma (RD-Sigma, Italy) and suspended in 0.3% Tween 80 and saline. It was administered (1 mg/kg intraperitoneally, ip) 30 min prior to 2-AG SA sessions.

2-AG SOLUTIONS

2-Arachidonoylglycerol content in the solutions prepared for SA or microdialysis studies was determined by HPLC–MS/MS analysis performed on MAX-RP C18 column (150 × 4.60 mm; 4 µm). The samples (20 µL) were analyzed by ESI in positive SIM mode following the ion [M + H]⁺ 379 m/z. The HPLC conditions were as follows: flow rate: 0.4 ml/min; solvent A: 0.1% formic acid in water; solvent B: acetonitrile; and gradient: solvent B 5–100% over 10 min. The samples (20 µl) were then analyzed by a Varian 1200 triple quadrupole HPLC–MS. Mass spectral data were acquired with a scan time of 1.0 s, needle 3500 V, shield 600 V, capillary 30 V, and detector 1900 V. The source parameters were adjusted as follows: drying gas temperature 250°C, drying gas pressure 20 psi, and nebulizer pressure 45 psi.
SELF-ADMINISTRATION STUDIES
Daily SA sessions were carried out in chambers housed in soundproof boxes (Coulbourn Instruments, Allentown, NJ, USA) containing two nose-hole holes, one active and the other inactive. A yellow/green light was placed over the active hole and a red light over the inactive one as discriminative stimuli. Prior to each daily session, the jugular catheter was flushed with 0.1 ml of sterile saline and the rats were placed in the SA box.

Rats were anesthetized with Equitesin (3 ml/kg ip; chloral hydrate 2.1 g, sodium pentobarbital 0.46 g, MgSO₄ 1.06 g, propylene glycol 21.4 ml, ethanol (90%) 5.7 ml, H₂O 3 ml) and implanted in the right jugular vein with a catheter, consisting of medical-grade tubing (Silastic, Dow Corning Corporation, Michigan, USA) according to the technique previously described (50). A stable fixation in the mid-scapular region of the back was embedded by a polypropylene mesh (Evolution, BULEV; weight 48 gmq, Dipromed, Italy). During the recovery period, at least 7 days after surgery, the catheters were flushed daily with 0.1 ml of gentamicin (40 mg/ml) and with heparinized saline (heparin 250 U/ml in 0.9% sterile saline).

Ten days after recovery from surgery, 13 rats were trained to SA 2-AG (25 µg/kg/20 µl iv) in 1 h-daily sessions (5 days/week) for 13 consecutive sessions, according to a FR 1 schedule of reinforcement (FR 1, 1 nose poke; 1 injection). During the third week, when all rats had fulfilled the criterion of 85% responses in the active hole and stable responding over three sessions, the schedule of reinforcement was increased to FR 2 (2:1) (14th–24th session).

A nose poke in the active hole resulted in a 4-s infusion of 2-AG. Each 2-AG infusion was followed by a 20-s time-out period, during which further nose pokes were recorded but did not result in additional intravenous infusions. Rats were also studied in the extinction phase from the 25th session, when the 2-AG solution was substituted with vehicle (25th–32nd day of administration).

A group of six rats were also studied in reacquisition when vehicle was replaced with 2-AG (33th–40th). Another group of animals were used to study the effect of varying injection doses of 2-AG. At the end of each SA session, the catheters were flushed with 0.1 ml of heparinized saline. The responses performed by each rat on both holes for the entire 1-h daily session and the corresponding number of reinforcers received was recorded (Graphic State 2 software, Coulbourn instruments, PA, USA).

MICRODIALYSIS STUDIES
Rats were anesthetized with Equitesin (3 ml/kg ip), prepared as previously described, and placed in a stereotaxic apparatus. The skull was exposed, and a small hole was drilled on one side. The microdialysis probe at a constant rate of 1 µl/min. Dialysate samples (10 µl) were taken every 10 min and injected without purification into an HPLC apparatus equipped with a reverse-phase column (C8 3.5 µm, Waters, Milford, MA, USA) and a coulometric detector (ESA Coulochem II, Bedford, MA, USA) to quantify DA.

At the end of each SA session, the catheters were flushed with 0.1 ml of heparinized saline and the rats were placed in the SA box.

A nose poke in the active hole resulted in a 4-s infusion of 2-AG. The responses performed by each rat on both holes for the entire 1-h daily session and the corresponding number of reinforcers received was recorded (Graphic State 2 software, Coulbourn instruments, PA, USA).

At the end of the experiment, animals were sacrificed and their brains were removed and stored in formalin (8%) before histological analysis. To this end, brains were cut on a vibratome in serial coronal slices (20 µm) oriented according to Paxinos and Watson (55) to locate the placement of the microdialysis probe.

RESULTS

SELF-ADMINISTRATION STUDIES
Experiment 1: acquisition, extinction, and reacquisition of 2-AG self-administration
In this experiment, acquisition, extinction, and reacquisition of 2-AG SA were studied. Figure 1A shows that rats implanted with a jugular catheter were trained to SA 2-AG (25 µg/kg/20 µl infusion, unit dose) in a single daily 1 h session, under an initial FR (FR 1) schedule, which was then increased to FR2. Figure 1A also shows the average number of active and inactive nose pokes performed by rats trained on 2-AG SA during acquisition, extinction, and reacquisition phases. Two-way ANOVA of acquisition and extinction phases showed a significant effect of nose pokes (F₁,2₄ = 3.4667; p < 0.0001), and of sessions (F₃₁,₇₄₄ = 182.97; p < 0.0001) and a significant nose pokes × session interaction (F₃₁,₇₄₄ = 214.069; p < 0.0001). LSD post hoc tests showed significant differences between active vs inactive nose pokes from the 7th to the 29th 2-AG SA session. Two-way ANOVA of acquisition and extinction phases showed a significant effect of nose pokes (F₁,₂₄ = 3.4667; p < 0.0001) on number of nose pokes between active vs inactive nose pokes from the 7th to the 29th 2-AG SA session. Two-way ANOVA of reacquisition, applied from the period 32nd to 40th session, showed a main effect of active vs passive nose pokes (F₁,₁₀ = 1381.47; p < 0.01). LSD post hoc tests showed significant differences between active and inactive nose pokes from the 33rd to the 40th 2-AG SA session. No differences were observed in active nose poking on each Monday following the weekend abstinence compared with the last session of the preceding week. The percentage of rats that acquired 2-AG SA was 90%.

Figure 1B shows the daily intake (µg/kg) of 2-AG or vehicle during all phases of SA (left Y-axis) and the corresponding number of reinforcements obtained (right Y-axis) in each 1-h session under a FR 1 or FR 2 schedule from rats that acquired 2-AG SA.
A
![Graph A](image)

B
![Graph B](image)

C
![Graph C](image)

Figure 1 | Acquisition, extinction, and reacquisition of 2-AG self-administration (SA) behavior over consecutive session.

(A) Number of responses (nose pokes) for 2-AG SA (25 µg/kg/infusion). Results are expressed as mean ± SEM of nose pokes in the active (circle) and inactive (triangle) holes during each 1-h daily session under FR 1 and FR 2 schedule (acquisition phase: 1st–24th days, filled symbols, N = 13, extinction phase: 25th–32nd days, open symbols, N = 6; reacquisition phase: 33th–40th days, filled symbols, N = 6). *p < 0.05 vs inactive nose pokes; ANOVA followed by LSD post hoc test. (B) Daily intake and number of infusions during 2-AG SA. Data are expressed as µg/kg (left Y-axis) or number of infusions (right Y-axis) of 2-AG during each 1-h daily session (1st–24th; 33th–40th, filled squares) or vehicle (25th–32nd, open squares). (C) Number of responses during each phase of 2-AG SA. Each bar represents the mean ± SEM of the last three sessions under acquisition, extinction, and reacquisition phases of 2-AG SA. *p < 0.05 vs acq.; #p < 0.05 vs reacq. ANOVA followed by LSD post hoc test.

Experiment 2: effect of dose of 2-AG and CB1 receptor blockade on 2-AG SA

In the second experiment, the effects of varying doses of 2-AG (12.5–50 µg/kg/20 µL infusion, unit dose) and rimonabant (SR-141716A, 1 mg/kg ip, 30 min before SA session) under an FR1 schedule were studied (Figures 2A, B). Two-way ANOVA of acquisition at the dose of 25 µg/kg/infusion showed a significant effect of nose pokes (F(1,24) = 3.896; p < 0.0001) and of sessions (F(15,360) = 324.115; p < 0.0001), and a significant nose poke × session interaction (F(15,360) = 316.965; p < 0.0001). LSD post hoc tests showed significant differences between active and inactive nose pokes (7, 10, 12, 13, and 15th SA session).

Two-way ANOVA of acquisition at the dose of 12.5 µg/kg/infusion showed a main effect of active vs passive nose pokes (F(1,15) = 1.45, p < 0.0001). LSD post hoc tests showed significant differences between active and inactive nose pokes (18, 20th 2-AG SA session).

Two-way ANOVA of acquisition at the dose of 50 µg/kg/infusion showed a main effect of active vs passive nose pokes (F(1,15) = 8.25; p < 0.001). LSD post hoc tests showed significant differences between active and inactive nose pokes (29, 30, 38–40th 2-AG SA session). Two-way ANOVA of acquisition at the dose of 50 µg/kg/infusion after rimonabant injection showed a significant main effect of active vs passive nose pokes (F(1,9) = 6.22; p < 0.05) and of day (F(2,8) = 2.19; p < 0.05). LSD post hoc tests showed significant differences between active and inactive nose pokes (37–40th 2-AG SA session).

Figure 2B shows the mean number of responses during the last three sessions of 2-AG SA at the dose of 50 µg/kg/infusion (29–31st 2-AG SA session) and the effect of rimonabant pre-treatment (SR, 1 mg/kg ip, 30 min before each session) for two consecutive sessions (32nd and 33rd 2-AG SA session). One-way ANOVA showed a main effect of the different SA phases (F(2,21) = 9.91; p < 0.05). LSD post hoc tests showed significant differences between groups.

Figure 3 shows that varying the injection dose of 2-AG resulted in a classic inverted-U-shape dose–response curve. One-way ANOVA showed that 2-AG maintained significantly higher numbers of infusions per session (F(3,8) = 12.44; p < 0.01) (Figure 3A) and higher rates of responding (F(3,8) = 45.75; p < 0.001) (Figure 3B) than vehicle at the doses of 12.5, 25, and 50 µg/kg/infusion. LSD post hoc tests showed that the maximal rate of responding (0.12 ± 0.09 response/s) and the number of injections per session (7 ± 1.19 injections/session) were maintained by 25 µg/kg 2-AG/infusion. The highest 2-AG intake per session was reached at a dose of 50 µg/kg/injection (155 ± 0.40 µg/session) (Figure 3C).

Microdialysis Studies

Effect of 2-AG administration on dopamine transmission in the NAc shell and core

Basal values of DA, expressed as fmoles/10 ml sample (mean ± SEM), were NAc shell 52 ± 5 (N = 19) and NAc core 54 ± 4 (N = 12).

Figure 4 shows that 2-AG (0.1–1.0 mg/kg iv) preferentially stimulated the NAc shell with respect to the NAc core DA.
NAC shell DA increase was observed only at the higher doses tested (0.5 and 1.0 mg/kg iv) and was about 25% over basal value. One-way ANOVA of the 0.5 mg/kg dose showed a main effect of group ($F_{12,48} = 3.27; p < 0.001$). Tukey post hoc tests revealed differences at the 20 min sample with respect to basal value and to vehicle. One-way ANOVA of the 1.0 mg/kg dose showed a main effect of group ($F_{12,48} = 2.17; p < 0.05$). Tukey post hoc tests revealed differences at the 20 min sample with respect to vehicle and with respect to basal value. Three way ANOVA of DA release of the NAC shell and core implanted animals revealed a significant effect of dose ($F_{1,23} = 4.9; p < 0.001$) and a significant area × time interaction ($F_{12,276} = 4.9; p < 0.05$).

**DISCUSSION**

In the present study, we demonstrate for the first time that the eCB 2-AG is SA by Sprague-Dawley rats and stimulates DA transmission preferentially in the NAC shell as compared to the core. Hence, 2-AG possesses behavioral and DA releasing properties similar to those of food and drug rewards.

Initially, animals acquired 2-AG SA (25 µg/kg/20 µl infusion, unit dose) in single daily 1 h FR1 sessions for 5 weeks, which was then increased to FR2, under food restriction that maintained body weights at stable levels throughout these studies. Active nose pokes significantly increased over inactive nose pokes from the 6th SA session (acquisition phase), but no significant increase of nose pokes was observed under FR2. When 2-AG was replaced by vehicle (25th SA session, extinction phase), the responding rate as well as the number of injections slowly decreased. When the vehicle was replaced once again with 2-AG, SA behavior immediately recov- ers and under SR pre-treatment condition for two consecutive session (SR, 1 mg/kg ip, 30 min before each session) (white bar) and under SR pre-treatment condition for two consecutive sessions. ANOVA followed by LSD post hoc test. (B) Number of responses during 2-AG SA and effect of SR-141716A on SA. Bars represent the mean ± SEM of the last three sessions under each 2-AG SA (50 µg/kg/inf) condition (black bar) and under SR pre-treatment condition for two consecutive sessions (SR, 1 mg/kg ip, 30 min before each session) (white bar). $p < 0.05$ vs veh; $p < 0.05$ vs SR-50. ANOVA followed by LSD post hoc test.

**FIGURE 2 | 2-Arachidonoylglycerol self-administration (SA) behavior at different 2-AG doses and effect of CB1 receptor blockade on SA.**

(A) Total number of injections of 2-AG (0, 12.5, 25, and 50 µg/kg/inf), (B) overall rate of responding for 2-AG, and (C) total 2-AG intake as shown as a function of injection dose of drug 10.

| Dose (µg/kg/inf) | Total number of injections | Rate of responding | Total intake per session |
|------------------|----------------------------|-------------------|-------------------------|
| 0                | 5                          | 0.05              | 0                       |
| 12.5             | 5                          | 0.05              | 0                       |
| 25               | 5                          | 0.05              | 0                       |
| 50               | 5                          | 0.05              | 0                       |
Different doses of 2-AG showed a tendency toward a classic inverted-U-shape dose–response curve, although non-significant differences were observed between the 25 µg/kg dose and the other doses tested (Figure 3). The reinforcing effects of 2-AG in SA behavior were fully blocked by the administration of the CB1 receptor inverse agonist/antagonist SR-141716A (rimonabant, 1 mg/kg ip, 30 min before SA session) (Figure 2B). The average number of responses during the 2-AG SA session, however, was lower in the present study than the responses (either nose pokes or lever pressing) observed during SA studies using other cannabinoids or other common drugs of abuse. Thus, rats trained to SA WIN 55-212 (50, 56), nicotine (57, 58), cocaine (59, 60), or heroin (56, 61, 62) maintained higher rates of responding than what we observed for 2-AG SA. Nevertheless, our data are in agreement with a previous study showing that 2-AG is a more effective reinforcer and that its reinforcing properties are not dependent on previous SA of other drug reinforcers.

in squirrel monkeys the rate of responding maintained by 2-AG injections was similar to those observed in THC, anandamide, methanandamide, or cocaine SA studies (39, 63–65). In contrast with Justinova et al. (40), our experiments were carried out in naive animals. Thus, our findings indicate that 2-AG serves as an effective reinforcer and that its reinforcing properties are not dependent on previous SA of other drug reinforcers.

A second group of naive rats was used to estimate the neurochemical rewarding properties of 2-AG. Thus, 2-AG administered iv at 0.5 and 1.0 mg/kg doses was able to stimulate extracellular DA release preferentially in the NAc shell as compared to the core. The increase of DA in the NAc shell was about 25% over basal value, peaking at 20 min after the injection of 2-AG. DA stimulation in this area was absent after the lowest dose tested (0.1 mg/kg iv), and the DA levels were comparable to those after vehicle injections. Interestingly, the higher dose tested (1.0 mg/kg iv) elicited a biphasic DA response with a second rise in DA levels 60 min after 2-AG injection. Notably, our data show for the first time that 2-AG stimulates DA neurotransmission in the mesolimbic DA system. This feature is a common characteristic of all the drugs abused by humans and it is believed to be crucial in the activation of brain rewarding pathways (66, 67), to be an anticipatory message for rewarding events (41) and to promote incentive learning (68) specially the DA increase in the NAc shell (42). This 2-AG-dependent increase of DA in the NAc shell most likely does not depend on the direct activation of DA neurons in the VTA, but, as in the case of THC and other cannabinoid agonists, is determined by the activation of CB1 receptors located on presynaptic glutamatergic and/or GABAergic terminals onto VTA DA neurons (69–71).

Previous studies have demonstrated that DA transmission increases in a dose-dependent fashion after administration of natural (THC), synthetic (WIN-55212), or endogenous (anandamide) cannabinoids and that this stimulation of DA in the NAc shell peaks at about 50% over basal levels. The doses used in these previous studies are comparable to those used in our study (49, 50, 52, 72) but they were accompanied by higher rates of SA behavior (39, 63, 64). In the present study, we did not observe a similar effect most likely due to the chemically unstable nature of 2-AG that quickly converts to 1-AG (38, 73). This, together with MAGL activity, could be responsible for the lack of high brain levels of 2-AG. Notably, in the present study, these properties of 2-AG were carefully taken into consideration during the preparation of the solutions for iv administration; 2AG content in administered solutions was determined daily before microdialysis or SA sessions by HPLC–MS/MS analysis.

Even though the rate of responding and DA stimulation were lower for 2-AG than those observed after administration of common drugs of abuse, our study demonstrates that the eCB system stimulation, via the mesolimbic DA system (microdialysis experiments) or by 2-AG itself (SA experiments), encodes rewarding events. The present microdialysis data, however, show that passive administration as a single intravenous bolus of 2-AG increased DA levels in the NAc shell at doses of 5–10-fold higher than the 2-AG intake reached by rats during SA behavior. A possible explanation might be the sensitization of the effect of 2-AG on DA transmission in the NAc shell of animals chronically exposed to...
2-AG during the acquisition of 2-AG SA. Further experiments of SA coupled to microdialysis performed with the same experimental protocol previously published by our group (50) will serve to directly correlate the preferential increase of DA in the NAc shell to the 2-AG SA behavior observed in our experiments. On the other hand, the results obtained in 2-AG SA experiments may be also due to the food restriction regimen. Many studies show that the reduction of body weight by 20–25% dramatically enhances drug-induced SA (see (74) for review). Similarly, the suppression of natural growth used in the present study, although not associated with loss of weight, may have increased the motivation for 2-AG, suggesting that 2-AG SA may not be crucial in standard condition (e.g., ad libitum access to food). If so, the decrease of active responding for 2-AG after the administration of rimonabant could also be due to the appetite suppression induced by this drug (75).

Consistent with other studies (40, 52), the main significance of our findings is that 2-AG may pose a risk of abuse liability, but that the activation of the eCB system together with the DA system may potentiate the reinforcing effects or the abuse potential of many drugs of abuse, supporting the hypothesis that an increase in eCB tone may be involved in drug-seeking behavior (76). Recent studies show that the manipulation of eCB levels in the brain can be selectively and successfully achieved by eCB clear- ance inhibitors (2, 77–79). Altogether, the above evidence from previous work, in conjunction with our current results, suggests that the eCB system plays a role in reward and reinforcement that makes it a good candidate as a therapeutic target in behavioral disturbances of motivation and reward.

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REFERENCES

1. Pacher P, Bátkai S, Kunos G. The endocannabinoid system as an emerging target of pharmaceutical action. Pharmacol Rev (2006) 58(3):389–462. doi:10.1124/pr.58.3.2
2. Ahn K, Johnson DS, Cravatt BF. Fatty acid amide hydrolase as a potential therapeutic target for the treatment of pain and CNS disorders. Expert Opin Drug Discov (2009) 4(7):765–84. doi:10.1517/174644409018857
3. Di Marzo V. Endocannabinoids: synthesis and degradation. Rev Physiol Biochem Pharmacol (2008) 160:1–24. doi:10.1007/5708_13050
4. Blankman JL, Cravatt BF. Chemical probes of endocannabinoid metabolism. Pharmacol Rev (2013) 65(2):849–71. doi:10.1124/pr.112.006387
5. Kirkham TC, Williams CM, Fezza F, Di Marzo V. Endocannabinoids levels in rat limbic forebrain and hypothalamus in relation to fasting, feeding and satiety: stimulation of eating by 2-arachidonoyl glycerol. Br J Pharmacol (2002) 136(4):520–7. doi:10.1038/sj.bjp.0704767
6. Piomelli D. More surprises lying ahead. The endocannabinoids keep us guessing. Neuropharmacology (2014) 76(Pt B):228–34. doi:10.1016/j.neuropharm.2013.07.026
7. Sugiuira T, Kodaka T, Kondo S, Tonegawa T, Nakane S, Kishimoto S, et al. 2-Arachidonoylglycerol, a putative endogenous cannabinoid receptor ligand, induces rapid, transient elevation of intracellular free Ca2+ in neuroblastoma x glioma hybrid NG108-15 cells. Biochem Biophys Res Commun (1996) 229(1):58–64. doi:10.1016/0006-291X(96)17577
8. Gonsiorek W, Lunn C, Fan X, Narula S, Lundell D, Hipkin RW. Endocannabinoid 2-arachidonoyl glycerol is a full agonist through human type 2 cannabinoid receptor: antagonism by anandamide. Mol Pharmacol (2000) 57(5):1045–50.
9. Hillard CJ. Biochemistry and pharmacology of the endocannabinoids arachidonylthanolamide and 2-arachidonoylglycerol. Prostaglandins Other Lipid Mediat (2000) 61(2–3):183–188.
10. Sugiuira T, Kobayashi Y, Oka S, Waku K. Biosynthesis and degradation of anandamide and 2-arachidonoylglycerol and their possible physiological significance. Prostaglandins Leukot Essent Fatty Acids (2002) 66(2–3):173–92. doi:10.1515/pfel.2001.0356
11. Alger BE, Kim J. Supply and demand for endocannabinoids. Trends Neurosci (2011) 34(6):304–15. doi:10.1016/j.tins.2011.03.003
12. Howlett AC, Riege PH, Childs DR, Hampson RE, Ulloa MA, Deutsch DG. Endocannabinoid tone versus constitutive activity of cannabinoid receptors. Br J Pharmacol (2011) 163(7):1329–43. doi:10.1111/j.1368-5883.2011.01364.x
13. Lupica CR, Riegel AH, Hoffman AF. Marijuana and cannabinoid regulation of brain reward circuits. Br J Pharmacol (2004) 143(2):227–34. doi:10.1038/sj.bjp.0705931
14. Adermark L, Lovinger DM. Retrograde endocannabinoid signaling at striatal synapses requires a regulated postsynaptic release step. Proc Natl Acad Sci U S A (2007) 104(51):20564–9. doi:10.1073/pnas.0706873104
15. Howlett AC. Cannabinoid receptor signaling. Handb Exp Pharmacol (2005) 168:53–79. doi:10.1007/3-540-26571-2_2
16. Murataeva N, Straiker A, Mackie K. Parsing the players: 2-arachidonoylglycerol synthesis and degradation in the brain. Br J Pharmacol (2014) 171(7):3799–91. doi:10.1111/bph.12411
17. Dinh TP, Carpenter D, Leslie FM, Freund TF, Katona I, Sensi SL, et al. Brain monoglyceride lipase participating in endocannabinoid inactivation. Proc Natl Acad Sci U S A (2002) 99(16):10819–24. doi:10.1073/pnas.152348999
18. Gulyas AI, Cravatt BF, Bracey MH, Dinh TP, Piomelli D, Bosca F, et al. Segregations of two endocannabinoid-hydropolyzing enzymes into pre- and postsynaptic compartments in the rat hippocampus, cerebellum and amygdala. Eur J Neurosci (2004) 20(2):441–58. doi:10.1111/j.1465-9294.200403428.x
19. Long JZ, Nomura DK, Cravatt BF. Characterization of monosaccerolipid lipase inhibition reveals differences in central and peripheral endocannabinoid metabolism. Chem Biol (2009) 16(7):744–53. doi:10.1016/j.chembio.2009.05.009
20. Blankman JL, Simon GM, Cravatt BF. A comprehensive profile of brain enzyme that hydrolyzes the endocannabinoid 2-arachidonoylglycerol. Chem Biol (2007) 14(12):1347–56. doi:10.1016/j.chembiol.2007.11.006
21. Fu J, Bottegoni G, Sasso O, Bertorelli R, Rocchia W, Masetti M, et al. A catalytically silent FAAH-1 variant drives anandamide transport in neurons. Nat Neurosci (2012) 15(1):64–69. doi:10.1038/nn.2986
22. Maldonado R, Berendse FO, Oizati A, Robledo P. Neurochemical basis of cannabis addiction. Neuroscience (2011) 181:1–17. doi:10.1016/j.neuroscience.2011.02.035
23. Sulhpara N, Parsons LH. Endocannabinoid-mediated synaptic plasticity and addiction-related behavior. Neuropharmacology (2011) 61(7):1070–87. doi:10.1016/j.neuropharm.2011.05.034
24. Sipe JC, Chiang C, Gerber AL, Beutler E, Cravatt BF. A missense mutation in human fatty acid amide hydrolase associated with problem drug use. Proc Natl Acad Sci U S A (2002) 99(12):8394–9. doi:10.1073/pnas.082255799
25. Chiang KP, Gerber AL, Sipe JC, Cravatt BF. Reduced cellular expression and activity of the P129T mutant of human fatty acid amide hydrolase: evidence for a link between defects in the endocannabinoid system and problem drug use. Hum Mol Genet (2004) 13(18):2113–9. doi:10.1093/hmg/ddh216
26. Kogan NM, Mechoulam R. Cannabinoids in health and disease. Dialogues Clin Neurosci (2007) 9(4):413–30.
27. Ledent C, Valverde O, Cossu G, Petitje F, Auberit JE, Beslot F, et al. Unresponsiveness to cannabinoids and reduced addictive effects of opiates in CB1 receptor knockout mice. Science (1999) 283(5400):401–4. doi:10.1126/science.283.5400.401
28. Hungund BL, Szakall I, Adam A, Basavarajappa BS, Vadasz C. Cannabinoid CB1 receptor knockout mice exhibit markedly reduced voluntary alcohol consumption and lack alcohol-induced dopamine release in the nucleus accumbens. J Neurochem (2003) 84(4):698–704. doi:10.1046/j.1471-4159.2003.01576.x
29. Houshi H, Babovic D, Pierrefiche O, Ledent C, Daout M, Naassila M. CB1 receptor knockout mice display reduced ethanol-induced conditioned place preference and increased striatal dopamine D2 receptors. Neuropsychopharmacology (2005) 30(2):339–49. doi:10.1038/sj.1003568
Endocannabinoid system and drug addiction

30. Mereu M, Parsons LH. SR141716A reduces the reinforcing properties of heroin but not heroin-inuces increases in nucleus accumbens dopamine in rats. *Eur J Neurosci* (2003) 18(11):3145–9. doi:10.1046/j.1460-9568.2003.03961.x

31. Solinas M, Panlilio LV, Antoniou K, Pappas LA, Goldberg SR. The cannabinoid CB1 antagonist N-piperidinyl-5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methylpyrazole-3-carboxamide (SR-141716A) differentially alters the reinforcing effects of heroin under continuous reinforcement, fixed ratio, and progressive ratio. *J Pharmacol Exp Ther* (2003) 306(1):93–102. doi:10.1124/jpet.102.047028

32. Colombo G, Orru A, Lai P, Cabras C, Maccioni P, Ruiro M, et al. The cannabinoid CB1 receptor antagonist, rimonabant, as a promising pharmacotherapy for alcohol dependence: preclinical evidence. *Mol Neurobiol* (2007) 36(1):101–12. doi:10.1007/s12035-007-0017-y

33. Mereu M, Tronci V, Chun LE, Thomas AM, Green JL, Katz JL, et al. Cocaine- and drug addiction: the nucleus accumbens shell connection. *Neuropharmacology* (2006) 47(1):2048–50. doi:10.1016/j.neuropharm.2005.02.003

34. Fillenz M. In vivo neurochemical monitoring and the study of behaviour. *Neuropharmacology* (2004) 47(3):241–63. doi:10.1016/S0896-6273(02)00967-4

35. Justinova Z, Tanda G, Redhi GH, Goldberg SR. Cannabinoid substitution in heroin-abstinent rats. *Psychopharmacology* (2003) 169(2):229–40. doi:10.1007/s00213-003-1484-0

36. Justinova Z, Mangieri RA, Bortolato M, Chefer SI, Mukhin AG, Clapper JR, et al. Fatty acid amide hydrolase inhibition heightens anandamide signaling without altering endocannabinoid metabolism. *Brain Research* (2005) 1052(1–2):13–22. doi:10.1016/j.brainres.2005.01.041

37. Caillé S, Parsons LH. Cannabinoid modulation of opiate reinforcement on delta9-tetrahydrocannabinol. *Eur J Pharmacol* (1999) 37(1):137–46. doi:10.1016/S0014-2999(99)00384-2

38. Paxinos G, Watson C. *The Rat Brain in Stereotaxic Coordinates*. 4th ed. London: Academic Press (1998).

39. Fattore L, Viganò D, Fadda P, Rubino T, Fratta W, Parolino D. Bidirectional regulation of mu-opioid and CB1-cannabinoid receptor in rats self-administering heroin or WIN 55,212-2. *Eur J Neurosci* (2007) 27(5):2197–200. doi:10.1111/j.1460-9568.2007.05470.x

40. Di Chiara G. Nucleus accumbens shell and core dopamine: differential role in mesolimbic dopaminergic decline and drug addiction. *Neurotox Res* (2003) 6(1):41–52. doi:10.1023/A:1021045120872

41. Schultz W. Getting formal with dopamine and reward. *Nat Neurosci* (2000) 3(2):179–85. doi:10.1038/76220

42. Lipton MA, Young VM, O’Rourke S, McEwen BS. Stressors and drug addiction: the role of the hypothalamo-pituitary-adrenal axis. *Neurotox Res* (2007) 11(3):135–40. doi:10.1007/s00009-006-0826-3

43. Justinova Z, Mangieri RA, Bortolato M, Chefer SI, Mukhin AG, Clapper JR, et al. Fatty acid amide hydrolase inhibition heightens anandamide signaling without altering endocannabinoid metabolism. *Brain Research* (2005) 1052(1–2):13–22. doi:10.1016/j.brainres.2005.01.041

44. Di Chiara G. Brain dialysis of neurotransmitters: a commentary. *Psychopharmacology* (2003) 231(1):229–40. doi:10.1007/s00213-003-1484-0

45. Pontieri FE, Sacco MS, Salis P, Melis V, Fattore L, et al. Cannabinoid self-administration increases dopamine release in the nucleus accumbens. *Neuropsychopharmacology* (2006) 30(7):1629–32. doi:10.1038/sj.npp.1382211.e

46. Volkow ND, Wang G-J, Fowler JS, Molina PE, Logan J, Gatley SJ, et al. Dopamine in brain and of epinephrine in plasma. *Psychopharmacology* (2003) 166(3):264–70. doi:10.1007/s00213-002-1340-7

47. Fillenz M. In vivo neurochemical monitoring and the study of behaviour. *Neurosci Biobehav Rev* (2005) 29(6):949–62. doi:10.1016/j.neubiorev.2005.02.003

48. Chang L, Haning W. Insights from recent positron emission tomographic studies of drug addiction. *Curr Opin Psychiatry* (2006) 19(1):246–52. doi:10.1097/01970659.46341.2

49. Tanda G, Pontieri FE, Di Chiara G. Cannabinoid and heroin activation of mesolimbic dopamine transmission by a common mu opioid receptor mechanism. *Science* (1997) 276(5321):2048–50. doi:10.1126/science.276.5321.2048

50. Lecca D, Cacciapaglia F, Valentini V, Di Chiara G. Monitoring extracellular dopamine in the rat nucleus accumbens shell and core during acquisition and maintenance of intravenous WIN 55,212-2 self-administration. *Psychopharmacology* (2006) 188(1):63–74. doi:10.1007/s00213-006-0475-3

51. Fadda P, Scherma M, Spono MS, Salis P, Melis V, Fattore L, et al. Cannabinoid self-administration increases dopamine release in the nucleus accumbens. *Neuroreport* (2006) 17(5):1629–32. doi:10.1097/01.wnr.0000235653.40221.8e

52. Solinas M, Justinova Z, Goldberg SR, Tanda G. Anandamide administration alone and after inhibition of fatty acidamide hydrolyase (FAAH) increases dopamine levels in the nucleus accumbens shell in rats. *J Neurochem* (2006) 98(3):408–19. doi:10.1111/j.1471-4159.2006.03890.x

53. Diana M, Melis M, Muntoni AL, Gesa GL. Mesolimbic dopaminergic decline after cannabinoid withdrawal. *Proc Natl Acad Sci U S A* (1998) 95(17):10269–73. doi:10.1073/pnas.95.17.10269

54. Tanda G, Loddo P, Di Chiara G. Dependence of mesolimbic dopamine transmission on delta9-tetrahydrocannabinol. *Eur J Pharmacol* (1999) 376(1–2):23–6. doi:10.1016/S0014-2999(99)00384-2

55. Fattore L, Viganò D, Fadda P, Rubino T, Fratta W, Parolino D. Bidirectional regulation of mu-opioid and CB1-cannabinoid receptor in rats self-administering heroin or WIN 55,212-2. *Eur J Neurosci* (2007) 27(5):2197–200. doi:10.1111/j.1460-9568.2007.05470.x

56. Caillé S, Parsons LH. Cannabinoid modulation of opiate reinforcement on delta9-tetrahydrocannabinol. *Eur J Pharmacol* (1999) 37(1):137–46. doi:10.1016/S0014-2999(99)00384-2

57. Paxinos G, Watson C. *The Rat Brain in Stereotaxic Coordinates*. 4th ed. London: Academic Press (1998).

58. Fattore L, Viganò D, Fadda P, Rubino T, Fratta W, Parolino D. Bidirectional regulation of mu-opioid and CB1-cannabinoid receptor in rats self-administering heroin or WIN 55,212-2. *Eur J Neurosci* (2007) 27(5):2197–200. doi:10.1111/j.1460-9568.2007.05470.x

59. Caillé S, Parsons LH. Cannabinoid modulation of opiate reinforcement on delta9-tetrahydrocannabinol. *Eur J Pharmacol* (1999) 37(1):137–46. doi:10.1016/S0014-2999(99)00384-2

60. Di Chiara G. The “core” of the rat nucleus accumbens. *Eur J Pharmacol* (2006) 547(2):12304–8. doi:10.1016/j.ejphar.2006.03.090

61. Melis M, Fists M, Perrra S, Muntoni AL, Pillolla G, Gesa GL. Endocannabinoids mediate presynaptic inhibition of glutameric transmission in rat ventral
71. Marinelli S, Di Marzo V, Florenzano F, Fezza F, Viscomi MT, van der Stelt M, et al. N-arachidonoyl-dopamine tunes synaptic transmission onto dopaminergic neurons by activating both cannabinoid and vanilloid receptors. *Neuropharmacology* (2007) **32**(2):298–308. doi:10.1038/npp.1301118

72. De Luca MA, Solinas M, Bimpisidis Z, Goldberg SR, Di Chiara G. Cannabinoid facilitation of behavioral and biochemical hedonic taste responses. *Neuropharmacology* (2012) **63**(1):161–8. doi:10.1016/j.neuropharm.2011.10.018

73. Rouzer CA, Ghebreselasie K, Marnett LJ. Chemical stability of 2-arachidonoylglycerol under biological conditions. *Chem Phys Lipids* (2002) **119**(1–2):69–82. doi:10.1016/S0009-3084(02)00068-3

74. Carr KD. Augmentation of drug reward by chronic food restriction: behavioral evidence and underlying mechanisms. *Physiol Behav* (2002) **76**(3):353–64. doi:10.1016/S0031-9384(02)00759-X

75. McLaughlin PL, Winston K, Swezy L, Wisniecki A, Aberman I, Tardif DJ, et al. The cannabinoid CB1 antagonists SR141716A and AM 251 suppress food intake and food-reinforced behavior in a variety of tasks in rats. *Behav Pharmacol* (2003) **14**(8):583–8. doi:10.1097/00008877-200308000-00002

76. Gutierrez-Lopez MD, Llopis N, Feng S, Barrett DA, O'Shea E, Colado ML. Involvement of 2-arachidonoylglycerol in the increased consumption of and preference for ethanol by mice treated with neurototoxic doses of methamphetamine. *Br J Pharmacol* (2010) **160**(3):772–83. doi:10.1111/j.1476-5381.2010.00720.x

77. Fowler CJ. “The tools of the trade” – an overview of the pharmacology of the endocannabinoid system. *Curr Pharm Des* (2008) **14**(23):2254–65. doi:10.2174/138161208785740126

78. Petrosino S, Ligresti A, Di Marzo V. Endocannabinoid chemical biology: a tool for the development of novel therapies. *Curr Opin Chem Biol* (2009) **13**(3):309–20. doi:10.1016/j.copbu.2009.04.616

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