Effect of a passive sonic irrigation system on elimination of *Enterococcus faecalis* from root canal systems of primary teeth, using different concentrations of sodium hypochlorite: An in vitro evaluation

Maryam Forghani¹,² • Elham Afshari³* • Iman Parisay¹,⁴ • Reza Garajian⁵

¹Dental Research Center, Mashhad University of Medical Sciences, Mashhad, Iran
²Department of Endodontic, Faculty of Dentistry, Mashhad University of Medical Sciences, Mashhad, Iran
³Department of Pediatric Dentistry, Faculty of Dentistry, Mashhad University of Medical Sciences, Mashhad, Iran
⁴Dental Materials Research Center, Mashhad University of Medical Sciences, Mashhad, Iran
⁵Food Quality and Safety Research Center, Jahad Daneshgahi of Khorasan Razavi, Mashhad, Iran

*Corresponding Author; E-mail: afsharie861@mums.ac.ir

Received: 6 April 2017; Accepted: 26 August 2017
J Dent Res Dent Clin Dent Prospect 2017; 11(3):177-182 | doi: 10.15171/joddd.2017.032
This article is available from: http://dentistry.tbzmed.ac.ir/joddd

© 2017 Afshari et al. This is an Open Access article published and distributed by Tabriz University of Medical Sciences under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

**Background.** This in vitro study aimed to compare the antibacterial effect of different concentrations of sodium hypochlorite on elimination of *Enterococcus faecalis* from root canal systems of primary teeth with or without a passive sonic irrigation system (EndoActivator).

**Methods.** The root canals of 120 extracted single-rooted primary incisors were prepared using the crown-down technique. The teeth were autoclaved and inoculated with *E. faecalis*. The infected samples were then randomly divided into 6 experimental groups of 15 and positive and negative control groups as follows: group 1: 0.5% sodium hypochlorite solution; group 2: 2.5% sodium hypochlorite solution; group 3: 5% sodium hypochlorite solution; group 4: 0.5% sodium hypochlorite solution + sonic activation; group 5: 2.5% sodium hypochlorite solution + sonic activation; and group 6: 5% sodium hypochlorite solution + sonic activation. Microbiological samples were collected before and after disinfection procedures and the colony-forming units were counted. Statistical analyses were performed using the two-way ANOVA and post hoc Duncan's tests in cases of significant difference.

**Results.** There were no significant differences between the groups in any of the variables (concentration of antiseptic or use of sonic irrigation system).

**Conclusion.** Use of passive sonic irrigation systems in endodontic treatment of single-rooted primary teeth is of no benefit compared to regular needle irrigation. The results of this study also recommends use of lower concentrations of sodium hypochlorite solution (0.5%) for irrigation of the root canal system rather than higher concentrations given approximately equal efficacy.

**Key words:** Sonication, primary teeth, root canal, sodium hypochlorite.
Introduction

Microorganisms play a significant role in the onset and perpetuation of pulpal and periradicular disease. Endodontic therapy is aimed at elimination of bacteria from the infected root canal and at prevention of infection. Root canal system in primary teeth seems to be more complicated than permanent teeth. This complexity is mostly attributed to the presence of ramifications and microcanals, in their root canal system. This can make the root canal treatment of primary teeth a more challenging procedure due to the fact that mechanical instrumentation alone cannot eliminate bacteria from these microcanals. New irrigation protocols, solutions and delivery systems in endodontics give us the hope to reduce root canal infections.

Of all the currently used substances, sodium hypochlorite appears to be the most ideal, as it covers more of the requirements for an endodontic irrigant than any other known compound. It is an antimicrobial agent; it also dissolves pulpal remnants, collagen and necrotic and organic tissues. NaOCl is commonly used at concentrations between 0.5% and 6%. Use of lower concentrations of sodium hypochlorite might lead to reduced antimicrobial and tissue dissolving capacity depending on the exposure time. On the other hand, higher concentrations may have adverse effects such as toxic reactions due to its penetration beyond the apex, exert detrimental effects on dentin elasticity and flexural strength, decrease dentin microhardness, and potentially kill the stem cells in the apical area. This is a bigger concern in case of primary teeth, where physiologic apical resorption leads to additional communication with the apical tissue other than through the apex, which results in a bigger chance of reaching the irrigant beyond the apex. Besides, the overflow of irrigating solution through the apical region because of possible resorption areas could damage the underlying permanent tooth. Therefore choosing the right concentration to maintain the delicate balance between effectiveness and safety is important.

Studies have shown that agitation of sodium hypochlorite has an additive effect on its capacity of tissue dissolution. Ultrasonic and sonic agitation was introduced as a means to increase the effectiveness of chemomechanical preparation. EndoActivator System (Dentsply Tulsa Dental Specialties, Tulsa, OK) is a passive sonication system designed to safely activate various intracanal irrigants and vigorously produce the hydrodynamic phenomenon. EndoActivator produces intracanal fluid agitation through acoustic streaming and cavitation to improve the flow of the irrigant into regions of the root canal system that are less accessible.

This in vitro study was designed to compare the antibacterial effect of a passive sonic irrigation system (EndoActivator) with that of conventional needle irrigation on elimination of E. faecalis from the root canal systems of primary teeth, using different concentrations of sodium hypochlorite.

Methods

The research protocol was approved by the Vice Chancellor for Research of Mashhad University of Medical Sciences (No. 910878). A total of 120 single-rooted extracted primary incisors were selected. The teeth had intact roots with no physiologic or pathologic resorption, fracture, considerable curvature or other anomalies of the roots. The extraction indications included traumatic injuries, unrestorable crowns or severely compromised supporting tissues.

The tooth surfaces were cleared of any remnant tissues, using a universal periodontal curette immediately after extraction. The teeth were then stored in 0.5% sodium hypochlorite solution for 24 hours and then stored in 0.9% normal saline solution at room temperature until the time of experiment. In the next step, the samples were decoronated on the level of CEJ using a diamond bur (Diamond Coated Twist Drill, Medium Grit, Meisinger USA) in a high-speed rotary instrument and roots of 10–12 mm length were ultimately left. Samples cut shorter than this range, fractured roots or destroyed ones were excluded at any step of the study and new samples were included.

Pulpal tissue remnants and debris were removed from the root canals using appropriate Hedstrom files. A #15 or #20 K-file was inserted into the root canal and penetration through the apex so that the file tip could be seen. Working length was determined 1 mm shorter than the file length inserted into the canal.

Root canal preparation was then performed using “passive step-back technique” as described by Torabinejad (1994). The samples were then placed in a dental ultrasonic cleaner in order to remove any remaining debris and then placed in 17% EDTA solution (Master-Dent, USA) for 10 minutes followed by 10 minutes of storage in 5.25% sodium hypochlorite solution. Finally, the samples were washed for 10 minutes using distilled water. The prepared samples were sent to a microbiology laboratory.
In the microbiology laboratory, the samples were autoclaved for 15 minutes at 121°C. After that, all the other procedures were carried out under aseptic conditions using sterilized instruments.

Isolated 24-hour colonies of pure E. faecalis cultures grown on 10% sheep blood agar + BHI (Newprov, Paraná, Brazil) plates were suspended in a sterile 0.85% NaCl solution. The suspensions of the microorganisms had the optical density of 0.5 McFarland scale (1.5×10^8 CFU mL^-1). The root canals were then infected with a suspension containing the microorganism using an insulin syringe except for a group of 5 uninfected roots as the negative control. Each sample was transferred under aseptic conditions into a micro-tube filled with sterile BHI medium, the lids were closed and the samples were incubated at 37°C for 48 hours. Each sample was transferred into normal saline solution 3 times, 30 minutes each time, to remove any remnants of culture or bacteria from the surface.

In the first step of bacterial assessment and before antibacterial treatment was performed, each root canal was filled with normal saline using a sterile insulin syringe and emptied into a micro-tube. The resultant liquid was cultured on BHIA (Blood Heart Infusion Agar) and BA (Blood Agar) medium to assess presence of E. faecalis.

The infected samples were then randomly divided into 6 experimental groups of 15 in terms of the irrigation system and concentration of the irrigant used, and a positive and a negative control group of 15 each (Table 1).

The root canals were initially irrigated for 30 seconds with two mL of sodium hypochlorite solution using an insulin syringe considering concentrations mentioned above. Groups 1 to 3 had no activation while the solution in the root canals of the three remaining groups was sonicated with EndoActivator (blue tip size 35/0.04, two mm short of the WL, at 10,000 cpm for 30 seconds). Then the root canals were again irrigated for 30 seconds with two mL of sodium hypochlorite solution the same as above and groups 1 to 3 were sonicated. The whole process of irrigation and sonication was repeated for the third time. Each sample was transferred to a micro-tube containing sodium hypochlorite solution with the concentration of irrigant used in its root canal. The negative control group which was uninfected was not treated with an antimicrobial solution. In the positive control group, the root canals were rinsed only with sterile normal saline solution and were not subjected to any disinfection procedure. In the end, all the root canals were flushed with 3 mL of sterile saline delivered in the same manner described above. The root canals were then assessed for E. faecalis, in the same manner as done in the first step and the CFU counts were reported (Figure 1). Statistical analyses were performed using two-way ANOVA and post hoc Duncan's test in cases of significant difference. A significance level of P<0.05 was adopted. The SPSS software 13.5 was used.

### Results

All the positive control samples exhibited microbial growth before and after treatment, whereas the negative control samples showed no microbial growth in either assessment. The average percentage of bacterial reduction in CFU mL^-1 within the root canal after treatment is shown in Table 2.

---

**Table 1. Experimental groups**

| Group | Irrigant                      | Activation       |
|-------|-------------------------------|------------------|
| 1     | 0.5% sodium hypochlorite solution | -                |
| 2     | 2.5% sodium hypochlorite solution | -                |
| 3     | 5% sodium hypochlorite solution | -                |
| 4     | 0.5% sodium hypochlorite solution | sonic activation |
| 5     | 2.5% sodium hypochlorite solution | sonic activation |
| 6     | 5% sodium hypochlorite solution | sonic activation |
| 7     | negative control              |                  |
| 8     | positive control              |                  |

---

Figure 1. a. Negative control, b. positive control, c. 100% bacterial reduction
There were no statistically significant differences between the groups in any of the variables (concentration of antiseptic or use of sonic irrigation system). However, there was an insignificant decrease in reduction of bacterial concentrations in groups 4 and 1 (0.5% antiseptic with and without sonication, respectively).

The means and standard deviations for the number of Enterococcus faecalis colonies for each concentration of the solution and irrigation method are presented in Table 3.

Except for groups 4 and 1 (0.5% antiseptic with and without sonic irrigation, respectively) in all the other groups (2.5% and 5% sodium hypochlorite solution), the bacterial colonies were not detectable after treatment, presenting zero concentration. As a result, there was no significant difference between bacterial count reduction in these groups.

There were no significant differences in the number of colonies with or without sonic activation between different concentrations. Use of sonic activation reduced the number of colonies using 0.5% sodium hypochlorite solution, but the difference was not statistically significant.

**Discussion**

This study compared two different irrigation protocols with different concentrations of NaOCl in relation to their abilities to eliminate Enterococcus faecalis in the root canal space of primary teeth. Root canal system in primary teeth is more complex than that in permanent teeth, which is attributed to the presence of branching and microcanals. This can make the root canal treatment of primary teeth a more challenging procedure owing to the fact that mechanical instrumentation alone cannot eliminate bacteria from these microcanals. Therefore the importance of an efficient antimicrobial irrigation is emphasized.

Enterococcus faecalis was chosen for the present study because it has repeatedly been isolated from the root canal system in failing endodontic cases. According to Cogulu et al, the most prevalent microorganisms found in the root canal system of primary teeth are Enterococcus faecalis, Porphyromonas gingivalis and Treponema denticola.

The success of sodium hypochlorite solution in illumination of Enterococcus faecalis has been substantiated by a large number of studies. The results of the present study indicated that sodium hypochlorite solution at a concentration of 0.5% or more can successfully reduce Enterococcus faecalis counts in roots of necrotic anterior primary teeth, which is consistent with the results of some previous studies.

An in vitro study by Shabahang et al concluded that irrigation with 1.3% and 2.5% NaOCl could not eliminate Enterococcus faecalis as needed. The authors suggested use of high concentrations and long exposure to NaOCl solution in order to efficiently reduce the number of bacteria. This is, however, in contrast with the results of the present study, which recognizes lower concentrations to be as effective as higher ones. Although the current study did not show complete reduction of bacteria at lower concentrations (0.5% with or without sonic activation), the difference from the higher concentration was not significant. This might be explained by the difference in the study design and methods used (such as use of bovine incisor dentin rather than human teeth, different times and intervals of exposure to the antiseptic solution and the method of exposure). There is still some debate on the use of higher concentrations of NaOCl solution regarding their rather lower safety, resulting in iatrogenic trauma, especially in pediatric patients; therefore, it is reasonable to consider the use of more diluted solutions for the same efficacy.

The effect of vibrating tip of a sonic irrigation device, apart from moving the tip up and down, will result in a large number of studies. Considering the safety related to the use of different irrigation systems, Desai et al reported significantly lower amount of apical extrusion of the irrigation solution with the use of EndoVac and EndoActivator compared to manual, ultrasonic and Rinsendo.
techniques. This is beneficial specifically in pediatric dentistry due to the larger diameter of apical foramen, resulting from physiologic resorption of roots of primary teeth or immature root canal system of young permanent teeth, which allow easier extrusion of irrigants to the periapical area.

Clinical and in vitro studies show controversial results regarding the priority of sonic irrigation systems rather than routine syringe irrigation. To the best of our knowledge, this is the first study evaluating the use of a sonic activation system in root canal irrigation of primary teeth and it was shown that activation with sonic devices would not significantly improve removal of the bacteria from the root canal systems of primary teeth. However, it should be considered that the current study was performed on single-rooted teeth. Due to the more simple canal system of single-rooted teeth, it might be much easier to achieve a thorough cleaned canal. In multiple rooted teeth with more complicated root canal morphology, especially in primary molars with more branching of root canal system, sonic activation may lead to cleaner root canals compared to syringe irrigation. Further investigations are required to determine the effects of these different irrigation protocols for predictable disinfection of infected root canals of primary teeth.

Conclusion

In spite of the limitations of the current study, our results suggest that use of passive sonic irrigation systems in endodontic treatment of single-rooted primary teeth is of no benefit compared to current routine methods. The results of this study also recommend use of lower concentrations of sodium hypochlorite solution (0.5%) for irrigation of the root canal system rather than higher concentrations regarding approximately equal effectiveness.

Acknowledgments

None

Funding

This study was funded by research center of Mashhad faculty of dentistry- Mashhad university of medical sciences

Competing interests

The authors declare no competing interests with regards to the authorship and/or publication of this article.

Ethics approval

The research protocol was approved by the Vice Chancellor for Research of Mashhad University of Medical Sciences (No. 910878).

References

1. Sen, B. H., Piskin, B., & Demirci, T. (1995). Observation of bacteria and fungi in infected root canals and dentinal tubules by SEM. Dental Traumatology, 11(1), 6–9. DOI: 10.1111/j.1600-9657.1995.tb00671.x
2. Bergmans, L., Van Cleyenbreugel, J., Wevers, M., & Lambrechts, P. (2001). A methodology for quantitative evaluation of root canal instrumentation using microcomputed tomography. International Endodontic Journal, 34(5), 390–398. DOI: 10.1046/j.1365-2591.2001.00413.x
3. Salama, F. S., Anderson, R. W., McKnight-Hanes, C., Barenie, J. T., & Myers, D. R. (1992). Anatomy of primary incisor and molar root canals. Pediatr Dent, 14(2), 117–118. DOI: http://dx.doi.org/10.1016/j.ypd.2013.04.002
4. Gaurav, V., Srivastava, N., Rana, V., & Adlakha, V. K. (2013). A study of root canal morphology of human primary incisors and molars using cone beam computed tomography: An in vitro study. Journal of Indian Society of Pedodontics and Preventive Dentistry, 31(4), 254. DOI:10.4103/0970-4388.121827
5. Wang, Y. L., Chang, H. H., Kuo, C. I., Chen, S. K., Guo, M. K., Huang, G. F., & Lin, C. P. (2013). A study on the root canal morphology of primary molars by high-resolution computed tomography. Journal of Dental Sciences, 8(3), 321–327. DOI: 10.1016/j.jds.2013.04.002
6. Sumikawa DA, Marshall GW, Gee L, Marshall SJ. Microstructure of primary tooth dentin, pediatric dentistry. 1999 Nov;21(7):439-44. DOI unavailable
7. Gu LS, Kim JR, Jing J, Choi KK, Pasheley DH, Tay FR. Review of contemporary irrigant agitation techniques and devices. Journal of Endodontics. 2009 Jun 30;35(6):791-804. DOI: 10.1016/j.joen.2009.03.010
8. Zehnder M. Root canal irrigants. J Endod 2006;32:389–98. DOI: http://dx.doi.org/10.1016/j.joen.2005.09.014
9. 9.Haapasalo M, Shen Y, Qian W, Gao Y. Irrigation in endodontics. Dental Clinics of North America. 2010 Apr 30;54(2):291-312. DOI: 10.1016/j.cden.2009.12.001.
10. Stojicic S, Zivkovic S, Qian W, Zhang H, Haapasalo M. Tissue dissolution by sodium hypochlorite: effect of concentration, temperature, agitation, and surfactant. Journal of endodontics. 2010 Sep 30;36(9):1558-62 DOI: 10.1016/j.joen.2010.06.021.
11. Gomes BP, Ferraz CC, ME V, Berber VB, Teixeira FB, Souza-Filho FJ. In vitro antimicrobial activity of several concentrations of sodium hypochlorite and chlorhexidine gluconate in the elimination of Enterococcus faecalis. International endodontic journal. 2001 Sep 1;34(6):424-8. DOI: 10.1111/j.1600-9657.2006.00161.x
12. Gernhardt CR, Eppendorf K, Kozlowski A, Brandt M. Toxicity of concentrated sodium hypochlorite used as an endodontic irrigant. International Endodontic Journal. 2004 Apr 1;37(4):272-80. DOI: 10.1111/j.0143-2885.2004.00804.x
13. Chaugule VB, Panse AM, Gawali PN. Adverse Reaction of Sodium Hypochlorite during Endodontic Treatment of Primary Teeth. International journal of clinical pediatric dentistry. 2015 May;8(2);153. DOI: 10.5005/jp-journals-10005-1304
14. Zararolli D, Saquy PC, Cruz-Filho AM. Effect of sodium hypochlorite and EDTA irrigation, individually and in alternation, on dentin microhardness at the furcation area of
mandibular molars. Brazilian dental journal. 2012;23(6):654-8. DOI:http://dx.doi.org/10.1590/S0103-64402012000600005

15. Martin DE, De Almeida JF, Henry MA, Khairng ZZ, Schmidt CE, Teixeira FB, Diogenes A. Concentration-dependent effect of sodium hypochlorite on stem cells of apical papilla survival and differentiation. Journal of endodontics. 2014 Jan 31;40(1):51-5. DOI: 10.1016/j.joen.2013.07.026.

16. Bhandari SK, Prajapati U. Root canal obturation of primary teeth: Disposable injection technique. Journal of Indian Society of Pedodontics and Preventive Dentistry. 2012 Jan 1;30(1):13. DOI:10.4103/0970-4388.95566.

17. Ruddle CJ. Endodontic disinfection: Tsunami irrigation. Saudi Endodontic Journal. 2015 Jan 1;5(1):1. DOI:10.4103/1658-5984.116273.

18. Mohammadi Z. Sodium hypochlorite in endodontics: an update review. International dental journal. 2008 Dec 1;58(6):329-41. DOI: 10.1111/j.1875-595X.2008.tb00354.x

19. Torabinejad, M. (1994). Passive step-back technique: A sequential use of ultrasonic and hand instruments. Oral surgery, oral medicine, oral pathology,77(4), 402-405. DOI: http://dx.doi.org/10.1016/0030-4220(94)90204-6

20. Cogulu D, Uzel A, Oncag O, Eronat C. PCR-based identification of selected pathogens associated with endodontic infections in deciduous and permanent teeth. Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology. 2008 Sep 30;106(3):443-9. DOI:10.1016/j.tripleo.2008.03.004

21. Berber, V. B., Gomes, B. P. F. A., Sena, N. T., Vianna, M. E., Ferraz, C. C. R., Zaia, A. A., & Souza-Filho, F. J. (2006). Efficacy of various concentrations of NaOCl and instrumentation techniques in reducing Enterococcus faecalis within root canals and dentinal tubules. International endodontic journal, 39(1), 10-17. DOI: 10.1111/j.1365-2591.2005.01038.x

22. Retamozo, B., Shahbahang, S., Johnson, N., Aprecio, R. M., & Torabinejad, M. (2010). Minimum contact time and concentration of sodium hypochlorite required to eliminate Enterococcus faecalis. Journal of Endodontics, 36(3), 520-523. DOI: 10.1016/j.joen.2009.12.005.

23. Klein, U., & Kleier, D. J. (2013). Sodium hypochlorite accident in a pediatric patient. Pediatric dentistry, 35(7), 534-538. DOI, unavailable

24. Elumalai DD, Kumar DA, Tewari DR, Mishra SK, Iftekhar H, Alam S, Andrabi M. Newer Endodontic irrigation devices: An update. IOSR Journal of Dental and Medical Sciences (IOSR JDMS). 2014;13(6):04-8. DOI: http://dx.doi.org/10.9790/0853-13650408

25. Desai, P., & Himel, V. (2009). Comparative safety of various intracanal irrigation systems. Journal of endodontics, 35(4), 545-549. DOI: 10.1016/j.joen.2009.01.011

26. Huffaker, S. K., Safavi, K., Spangberg, L. S., & Kaufman, B. (2010). Influence of a passive sonic irrigation system on the elimination of bacteria from root canal systems: a clinical study. Journal of endodontics, 36(8), 1315-1318. DOI: 10.1016/j.joen.2010.04.024.

27. Chatterjee R, Venugopal P, Jyothi KN, Jayashankar CM, Kumar SA, Kumar PS. Effect of sonic agitation, manual dynamic agitation on removal of Enterococcus faecalis biofilm. Saudi Endodontic Journal. 2015 May 1;5(2):125. DOI: 10.4103/1658-5984.155451

28. Mathew J, Emil J, Paulsian B, John B, Raja J, Mathew J. Viability and antibacterial efficacy of four root canal disinfection techniques evaluated using confocal laser scanning microscopy. Journal of Conservative Dentistry. 2014 Sep 1;17(5):444. DOI: 10.4103/0972-0707.139833

29. Ordinola-Zapata R, Bramante CM, Aprecio RM, Handy-Romans R, Jaramillo DE. Biofilm removal by 6% sodium hypochlorite activated by different irrigation techniques. International endodontic journal. 2014 Jul 1;47(7):659-66. DOI: 10.1111/iej.12202.