CGAP-Align: A High Performance DNA Short Read Alignment Tool

Yaoliang Chen¹, Ji Hong¹, Wanyun Cui¹, Jacques Zaneveld², Wei Wang¹, Richard Gibbs², Yanghua Xiao¹*, Rui Chen²*

1 School of Computer Science, Fudan University, Shanghai, China, 2 Human Genome Sequencing Center, Department of Molecular and Human Genetics, Baylor College of Medicine, Houston, Texas, United States of America

Abstract

Background: Next generation sequencing platforms have greatly reduced sequencing costs, leading to the production of unprecedented amounts of sequence data. BWA is one of the most popular alignment tools due to its relatively high accuracy. However, mapping reads using BWA is still the most time consuming step in sequence analysis. Increasing mapping efficiency would allow the community to better cope with ever expanding volumes of sequence data.

Results: We designed a new program, CGAP-align, that achieves a performance improvement over BWA without sacrificing recall or precision. This is accomplished through the use of Suffix Array, a novel data structure combining elements of Suffix Array and Suffix Tree. We also utilize a tighter lower bound estimation for the number of mismatches in a read, allowing for more effective pruning during inexact mapping. Evaluation of both simulated and real data suggests that CGAP-align consistently outperforms the current version of BWA and can achieve over twice its speed under certain conditions, all while obtaining nearly identical results.

Conclusion: CGAP-align is a new time efficient read alignment tool that extends and improves BWA. The increase in alignment speed will be of critical assistance to all sequence-based research and medicine. CGAP-align is freely available to the academic community at http://sourceforge.net/p/cgap-align under the GNU General Public License (GPL).

Introduction

In recent years, advances in sequencing have led to the production of unprecedented amount of sequence data. Alignment, which maps reads to the reference sequence, is one of the most computationally demanding tasks performed in typical sequence data processing. Accurate sequence alignment is critical for SNP calling [1], structural variation detection [2] and downstream analysis.

In order to efficiently and accurately map large numbers of short reads many new alignment programs have been developed. The algorithms underlying most of these tools can be classified into two major categories [3]. The first category uses hash tables to hash either read sequences, as in MAQ [1], SeqMap [4] and CloudBurst [5], or the genome reference, as in SOAPv1 [6], PASS [7], MOM [8] and ProbeMatch [9]. Although this technique can be easily parallelized, the major drawback of using hash tables is that either they must scan the whole genome, even when few reads are aligned, or they require a large amount of memory to build an index for the reference. The second category is based on string matching using a representation of prefix/suffix trie [2], including suffix tree, suffix array [10], enhanced suffix array [11] and FM-index [12]. The first three representations are used by TRELLIS [13], MUMmer [14] and Vmatch (http://www.vmatch.de). Unfortunately, these programs have poor performance on large-scale references including the human genome due to memory constraints. The FM-index is a type of substring index based on the Burrows-Wheeler transform, with some similarities to suffix array. Owing to its small memory requirement, it is utilized by numerous state of the art programs including SOAPv2 [15], Bowtie [16], Bowtie2 [17] and BWA [18].

BWA has become one of the most widely used alignment tools owing to its efficiency and accuracy. BWA possesses higher recall and precision than SOAPv2 or Bowtie. However, both Bowtie and SOAP2 are significantly faster than BWA. Therefore, it is highly desirable to improve the speed of BWA while maintaining its alignment quality. In this paper, we describe a new efficient alignment tool, CGAP-align, following the framework of BWA.
part of short read mapping in BWA, possible mismatches and gaps are enumerated during the traversal of the FM-index of the reference sequence. The time efficiency of BWA is mainly impacted by two factors. The first is the efficiency of locating a substring in the reference. The second is the ability of the program to bypass segments of the reference where the read would contain a large number of mismatches, avoiding the need to enumerate all possible sets of mismatches and gaps, through a process called pruning. Improvements to these strategies have been implemented in CGAP-align to optimize the efficiency of both the reference querying and the pruning steps.

In this report, we first introduce a novel data structure, Suffix Tarray, which speeds up the process of locating a read on the reference. Second, we present an effective pruning strategy that more accurately predicts the number of mismatches in a read prior to alignment. Pruning is significantly improved by using a set of training reads in advance to identify and study frequent patterns of nucleotides. The performance of CGAP-align was evaluated on both simulated data and several sets of real paired-end sequence data. Our results indicate that by implementing both of these improvements alignment speed is significantly increased without sacrificing recall or precision.

Methods

1.1 Suffix Tarray (STA): Improving Reference Queries

To improve the alignment speed, we utilized a data structure, Suffix Tarray (STA), to index reference sequences in CGAP-align. STA uses a new index structure that is a hybrid of trie (inspired by suffix tree) and suffix array data structures. Before we present the concept of STA, we first briefly review the two most widely-used data structures for sequence indexing: suffix array (SA) and suffix tree (ST).

1.1.1 Background: Suffix Tree (ST) & Suffix Array (SA)

Suffix tree is a data structure that encodes all of the suffixes of a given string in the form of tree, allowing quick location of substrings of suffixes of a given string in alphabetical order. The suffix tree is constructed in linear time \[O(n)\] time from the reference \(X\) (as it can be constructed in linear time from a SA) and with a small constant and can be traversed in linear time \(O(n)\) time from the start of any substring. Owing to its query efficiency, FM-index, has been recently adopted by many widely-used mapping tools, including SOAPv2 [15], Bowtie [16] and BWA [18]. However, such space efficiency comes at the cost of reduced query performance. Empirically, ST is much faster than FM-index for queries on substrings [23].

1.1.2 Overview of the Suffix Tarray (STA) Structure.

In an FM-index, for a character \(c\), \(Occ\) is a function of the SA position \(y\), which counts the number of the occurrences of \(c\) within the suffix interval \([1, y]\) of the SA. To accelerate the calculation of \(Occ\) values, the SA is divided into buckets with a fixed size. For the SA positions \(y\) that define the left bucket boundaries, the corresponding \(Occ\) values are pre-computed and stored in a table. This allows all other \(Occ\) values to be efficiently calculated by counting the occurrences of the character \(c\) from the start of their corresponding buckets instead of from the beginning of the SA.

However, retrieving a pre-computed \(Occ\) value is still much faster than retrieving an \(Occ\) value in the middle of a bucket. This observation implies that pre-computing frequent \(Occ\) values has the potential to speed up the mapping process. The basic idea of Suffix Tarray (STA) is to first find those most frequently visited \(Occ\) values and then organize them in a ST so that they can be efficiently accessed during alignment. With this novel approach, we can take advantage of the time efficiency of the ST and the space efficiency of FM-index. At a high-level, STA can be viewed as a truncated suffix tree (TST) encoding corresponding SA intervals of a FM-index at each leaf (Figure 1). As the height of the tree is bounded to a reasonably small value, we adopt trie as a light implementation of the TST.

There are two major steps to construct a STA for a reference string \(X\). In the first step, a FM-index is built for the entire reference sequence. All possible suffix strings of the reversed reference sequence \(x\) are enumerated. Instead of building the whole ST for \(x\), we truncate the ST according to the frequency values of the nodes. The frequency value of a node in ST for \(x\) is defined as the number of occurrences of the substring which it represents in \(x\). In our example, the frequency value of the node “CT” is 2 since there are 2 prefixes valued “CT” among all of the suffixes of \(x\). The nodes in trees will be discarded if their frequency values are smaller than a threshold \(\varepsilon\) which is calculated by St. We select a maximal \(\varepsilon\) such that the resulting trie has a size less than a user-specified trie size \(St\). This is accomplished by a binary-search style enumeration of the possible values of \(\varepsilon\). Initially, the possible range of \(\varepsilon\) is set to \([1, \ |X|]\). The program first tries to construct a trie using a value \(P\) in the middle of possible range, such that in the first iteration \(P\) is approximately \(|X|/2\). If, during construction, the trie size exceeds the size limit \(St\), then the maximum possible value for \(\varepsilon\) is reduced to \(P\). If, on the other hand, the trie is
constructed without exceeding St then the minimum possible value of \( e \) is increased to \( P \). This process is iterated until the possible range of \( e \) is reduced to a single value, which we assign to \( e \). At last, a truncated trie with frequency threshold \( e \) and size less than \( St \) is built. Figure 1 is an example of a STA.

In the current version of CGAP-align, each trie node takes 36 bytes. One concern when building a STA is how to choose an appropriate truncated trie size. If the size is too small, then the index will not see a significant increase in query speed since the FM-index will dominate the query process. Otherwise, memory consumption will increase. Fortunately, since the frequency values of novel nodes decreases rapidly with increasing trie size, a STA with a moderate-sized trie can perform nearly as well as a STA with a large trie. We denote ST\(_{100}\) as a full ST for truncated at depth 100 and the frequency ratio of a trie as the values of all nodes in that trie divided by the sum of frequency value of all nodes in ST\(_{100}\). According to our experiment, when indexing the human reference genome, a 300 MB sized truncated ST has a frequency ratio of 13.32% while 1 GB has 14.68%.

1.1.3 Matching Substrings to Suffix Tarray.

Algorithm 1:

**STA index construction** : \((X,b)\) //reference string \(X\) and the length of candidate string \(b\)

Calculate BWT string \(B\) for reference string \(X\);

Calculate array \(C(\cdot)\) and \(Occ(\cdot,\cdot)\) from \(B\);

Build ST for all the reversed substrings occurring in \(X\) with frequency \(\approx e\);

**Procedures**:

InexactSearch\((W,z)\)

return InexRecur\((W,|W|−1,z,1,|X|−1,ST.root)\);

InexRecur\((W, i, z, k, l, node)\)

* if \( z < D[i] \) then

* return \( \otimes \)

if \( i < 0 \) then

return \( \{[k,l]\} \)

//deletion, edit cost : 1

\( I \leftarrow \text{InexRecur}(W, i−1, z−1, k, l, node) \)

for each \( c \in \{A,C,G,T\} \) do

if node not null then

\( k \leftarrow \text{node}.k; \ l \leftarrow \text{node}.l; \)

\( \text{node} \leftarrow \text{node}.\text{child}[c]; \)

else;

\( k \leftarrow C(c) + Occ(c,k−l)+1; \)

\( l \leftarrow C(c) + Occ(c,l); \)

if \( k \leq l \) then

//insertion, edit cost : 1

\( I \leftarrow I \cup \text{InexRecur}(W, i, z−1, k, l, node) \)

if \( c = W[i] \) then

//no edit operation, edit cost : 0

\( I \leftarrow I \cup \text{InexRecur}(W, i−1, z, k, l, node) \)

else

//replace, edit cost : 1

\( I \leftarrow I \cup \text{InexRecur}(W, i−1, z−1, k, l, node) \)

return \( I \)
The procedure to query a substring from STA is given in Algorithm 1. Procedure InexactSearch([W, z]) takes substring W and maximal edit distance z as input, and returns the SA intervals of substrings that match W with no more than z differences. The algorithm framework is similar to that used in BWA, but with a different index structure. The l and k values are re-calculated based on STA. For a given node of ST, we denote node.child as its child-list, while node.k and node.l define its SA interval values. The lines with an asterisk mark the D-array pruning strategy used in BWA, which will be discussed in section 1.2.

The actual implementation of CGAP-align differs slightly from the pseudo code given in Algorithm 1. CGAP-align inherits the logic of BWA’s inexact mapping, where “gap extensions” do not add to the number of edits that have been performed (i.e. IndexRecurs will be recursively called with maximum edit distance value z). This helps to detect relatively long gaps. However, it also causes a potential issue resulting in false negative alignments in both BWA and CGAP-align. This problem is addressed in section 2.2.

1.2 Data-Conscious D-Array Calculation (DCDC): Improving Pruning

To improve the efficiency of mapping, a pruning step is implemented in BWA. A lower bound is calculated for the number of differences between \( W[0..t] \) (i.e. the t-length prefix of the read \( W \)) and any substrings of the reference sequence and stored as the \( t \)th element of array \( D \), all prior to mapping. Reads with \( D[i] \) greater than the maximum tolerated number of mismatches defined by the user are excluded from further alignment. If calculating D-array is faster than the time it would take to map the pruned part of the search trees, pruning improves mapping efficiency. The degree of improvement depends on both the speed at which the D-array can be calculated and the accuracy of the estimation of D-array. A more accurate estimation of D results in a smaller search space and more efficient mapping. Figure 2 gives an example of how D-array would work during a backward search on either FM-index or STA. D-array prunes superfluous enumerations before mapping, providing a significant boost in performance.

1.2.1 D-array: Background. BWA proposes a method to estimate the D-array by splitting \( W \) into several small segments. Let \( e(W) \) be the minimal number of the edit operations required to make \( W \) exactly align on the reference \( X \). BWA divides \( W \) into segments \( w_1 \ldots w_t \), where \( e(w_p) = 1 \) for \( 1 \leq p < t \) and \( e(w_t) = 1 \) for \( p = t \). Then \( D[i] \) is approximated as \( p - 1 \) for \( 1 \leq i < |W| \), where \( w_p \) contains the \((i+t)^{th}\) element of \( W \), or \( t \) for \( i = |W| \). For example, given a reference \( X = \{\text{AACGTATCGACG}\} \) and a read \( W = \{\text{AACGT}\} \), BWA segments the \( W \) as \( \{\text{AACG}\}/\{\text{T}\}/\{\text{G}\}/\{\text{A}\}/\{\text{CGACG}\} \) and thus produces the D-array \( \{\text{000111}\} \). The time to calculate the D-array for read \( W \) is in \( O(|W|) \) when the FM-index of the reverse of \( X \) is used. In CGAP-align, the calculation of D-array is further accelerated by using STA.

1.2.2 A Tighter Lower Bound for D-array. Identifying a \( u_p \) with \( e(u_p) > 1 \) when splitting \( W \) generates a D-array with a tighter lower bound. In the example described above, if we consider segments with an \( e(u_p) \) equal to 2, then we derive the segmentation \( \{\text{AACG}\}/\{\text{T}\}/\{\text{G}\}/\{\text{A}\}/\{\text{CGACG}\} \), corresponding to the D-array \( \{000122\} \). By allowing segments with more mismatches we derive a tighter bound for D.

However, the above tighter bound comes at the cost of longer computations. The time cost of verifying \( e(W) = k \) is exponential to the value of \( k \). Therefore, it is too expensive to calculate each \( e(w_i) \) value on the fly unless \( e(w_i) \) is strictly restricted to 1 (in this case calculation of \( e(w_i) \) is \( O(|w_i|) \)). To address this issue, we add a pre-processing step, in which we identify the frequent substrings from the training read set with a \( e(w_i) = 2 \). This approach is feasible as genomic sequences from the same species are similar among individuals. Thus, providing a training read set from each species, it is possible to generate a list of species specific frequently occurring substrings \( (w_i) \) with \( e(w_i) = 2 \).

In a pre-processing phase, an appropriate number of patterns with the greatest frequency values are identified by a depth-first visit on an FM-index built upon the training reads and then organized into an Aho-Corasick (AC) automaton [24]. The number of patterns is determined by the AC automaton size specified by the user. Given a read \( W \), an AC automaton helps to find all substrings of \( W \) (denoted as \( w \)) that match to any of the indexed patterns (denoted as \( f \)) in linear time. A description of AC automaton is given in section 1.2.3. For the sake of the efficiency, we only index patterns with \( e(f) = 2 \) into AC automaton.

1.2.3 Aho-Corasick Automaton. In general, millions of frequent patterns (FP) with \( e(f) = 2 \) are found in the pre-processing phase. We use a AC automaton \( G_T \) to organize these FPs so that when given a read \( W \), we are able to quickly query all of the FPs that are contained in \( W \) in a single scan. This allows for efficient segmentation of \( W \) according to the FPs it contains. We adopt the AC algorithm [24] in which the FPs are organized in an AC automaton. Instead of searching for occurrences of a single string \( f \) within a main text string \( W \), AC automaton supports searching for...
occurrences of a set of strings $F$ within $W$. An AC automaton makes use of the information embodied by the string set $F$ itself to determine where to begin the next matching attempt if a mismatch happens, thus bypassing re-examination of previously matched characters in $W$. There is a particular type of state called a “leaf state”, each of which corresponds to a string $f$. The transition to a leaf state $L_f$ indicates an occurrence of $f$ in $W$. When a match is detected, we reset the automaton to its initial state and continue the scanning on $W$. These steps are iteratively performed until we meet the end of $W$. A simple example is shown in Figure 3.

1.2.4 Calculating the Better D-array.

**Algorithm 2 CalculateD**($W$, $X$, $G_T$)

**Input**: A read $W$, the reference $X$ and the AC automaton $G_T$

**Output**: The $D-$ array for $W$

\[
\begin{align*}
&j \leftarrow 0; z \leftarrow 0; k \leftarrow 0 \\
&\text{for } i = 0 \text{ to } |W| - 1 \text{ do} \\
&\quad \text{if } W[j, i] \text{ is not a substring of } X \text{ then} \quad e(W(j, i)) = 1 \\
&\quad \quad z \leftarrow z + 1 \\
&\quad \quad k \leftarrow j \\
&\quad \quad j \leftarrow i + 1 \\
&\quad \text{elseif any suffix of } W[k, i] \text{ matches a patternstring } f \text{ in } G_T \text{ then} \quad e(W[k, i]) = 2 \\
&\quad \quad z \leftarrow z + 1 \\
&\quad \quad k \leftarrow i + 1 \\
&\quad \quad j \leftarrow i + 1 \\
&\quad D(i) \leftarrow z
\end{align*}
\]

Algorithm 2 shows the detailed procedure to estimate a tighter $D$-array in CGAP-align. We use a greedy strategy to iteratively find the first matched pattern string $f$ of $W$ from the AC automaton $G_T$. If no such $f$ is found, we segment $W$ by $e(W[j, i]) = 1$ instead (the first “if” condition). As described above, when we find a match in $G_T$ at position $i$, we reset the automaton, scanning matches within $W[i+1, |W|-1]$ in the following comparison.

**Results**

As described above, two major changes have been added to the current version of BWA. First, instead of FM-index we utilize STA, a novel data structure that speeds up string matching by constructing a trie on top of an FM-index. Second, we implement a new pruning method DCDC. As mapping time increases exponentially with the number of mismatches, it is highly desirable to prune reads with large numbers of mismatches. To achieve this goal, a data training process has been implemented to identify a set of frequent substrings with two mismatches from the genome. This pre-processed data allows for accurate calculation of the minimal number of mismatches between a prefix and the genome, resulting in further reduction of the search space.

Both functionalities have been implemented in C. To facilitate the usage of CGAP-align, it offers a command line interface that is almost identical to BWA and outputs SAM files (Sequence Alignment/Map format). CGAP-align is distributed under the GNU General Public License (GPL) with detailed documentation and source code freely available through Fudan University, BCM-HGSC and the Sourceforge web site. The pre-built indices for some public references (eg. Hg19) are also provided.

2.1 CGAP-align Evaluation

To evaluate the performance of CGAP-align, we have benchmarked it against BWA (version 0.5.9), SOAPv2 (version 2.20) and Bowtie2 (version 2.0.6), three of the most commonly used alignment programs. Other tools like Bowtie are not included because gapped alignments are currently not supported. All tools run on 4 threads. Both BWA and CGAP-align were evaluated on their ability to map 100 base pair long reads using either default settings or relatively loose settings that allow up to 5 edits and a gap extension of 3 base pairs (-n 5 -e 3 -l 25). The same data set is used for SOAPv2, allowing up to 5 mismatches and a gap size of 4 (-v 5 -g 4). This setting is looser than SOAPv2’s default setting, which enables it to find more alignments. For bowtie2, default parameters are used (-sensitive) since no gap settings can be adjusted. In addition, two modes of CGAP-align, the first with STA alone (denoted CGAP-align) and the second with both DCDC and STA (denoted CGAP-align*), were evaluated. STA was built based on the human reference genome hg19 with 600 MB size. All of the experiments utilized the same AC
automaton with a size of 1000 MB trained from independent WGS data for DCDC.

2.2 Evaluation on simulated data

The performance of BWA, CGAP-align, Bowtie2, and SOAPv2 on a simulated data set was examined. Wgsim, a program included in SAMtools [25], was used to generate 20 million simulated human genome shotgun 100 bp reads with a 2% error rate. As the exact position of each simulated read in the genome is known, both the sensitivity and specificity of the alignments could be precisely calculated.

As shown in Table 1, a significant speedup was achieved by CGAP-align while maintaining high mapping quality. We report both the absolute running time (Hours) and the percentage relative to BWA for all programs considered. When STA was used alone, CGAP-align was about 10% to 20% faster than BWA. Since the implementation of STA only accelerates the string matching step without affecting alignment, identical results were obtained between CGAP-align and BWA. Both programs produced mapping results with high accuracy and sensitivity. Sensitivity was measured by the true positive rate (TPR), which is defined as the overall percentage of correctly mapped reads relative to the number of input reads. In parallel, mapping specificity was measured by the positive predictive value (PPV), which indicates the percentage of correctly mapped reads relative to the number of the reads reported as mapping correctly by the algorithm. Under default settings, 97.9% of the reads were accurately mapped with a PPV of 98.7%. The effect of STA was also evaluated under loose settings. A similar speed up was observed under this condition, and identical mapping results obtained by both CGAP-align and BWA as expected. For SOAPv2 and Bowtie, lower TPRs were observed than for CGAP-align although both run faster than CGAP-align.

Surprisingly, despite the use of training data, little speedup was observed after adding DCDC. As shown in Table 1, an almost identical speed was observed for CGAP-align under default settings. We found that in this case only 5% of the reads’ D-arrays were improved by DCDC. This is because, in our simulated data set, most reads were aligned to the reference with only 1 or 2 edit operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC. On the other hand, the DCDC itself consumed more time than the operations, leaving little optimization space for DCDC.

2.3 Evaluation on Real Data

We tested CGAP-align using whole exome sequence (WES) data from two individuals [29.2 million read pairs for WES1, 31.6

Figure 3. An example AC automaton. The frequent pattern string set F is {AA, AC, AG, C, G, T}. The grey nodes represent the leaf states. The bold ‘T’ edge is connected as among all the prefixes of the 6 patterns, “T” is the longest suffix of “AT”. Given a short read $W$ = “CAT”, the state transition sequence when querying $W$ from the automaton is <$R$, $L_c$, $R$, 1, $L_T$>, which indicates the occurrence of frequent patterns ‘C’ and ‘T’. doi:10.1371/journal.pone.0061033.g003
million read pairs for WES2) and whole genome sequence (WGS) data from a separate pair of individuals (107.8 million read pairs for WGS1 and 86.3 million read pairs for WGS2). All read pairs were produced by Illumina HiSeq with a 100 bp read length. Both the consumed time (Hours) and the frequency with which reads were confidently mapped (Conf) are reported in Table 2. For both CGAP-align and BWA, we use mapping quality threshold 10 to determine confident mappings. CGAP-align, utilizing STA alone, obtained a speed up of approximately 10% to 20% compared with BWA. Surprisingly, CGAP-align*, while achieving a small speed up with default settings, shortened the running time by more than 50% with loose settings, due to the high error rates of the reads in real data. While SOAPv2 was faster than all other candidates including CGAP-align*, it found less confidently mapped reads than BWA, especially for the WGS cases. On the WGS1 data, where a significant number of mismatches between the reads and reference were observed, CGAP-align and BWA confidently mapped almost 10% more reads than SOAPv2 did. In comparison to SOAPv2, CGAP-align* further shortens the performance gap while maintaining a relatively high mapping rate. Overall, it possesses impressive mapping rates in WES while the performances when mapping WGS data still need to be improved.

Discussion

Due to the enormous number of reads generated by the next generation sequencing technologies, the efficiency of read alignment has become a critical problem. In this article, we present CGAP-align, a variant of BWA, which doubles the speed of the alignment process. While SOAPv2 is also very fast, it is unable to determine confident mappings. CGAP-align, utilizing STA alone, obtained a speed up of approximately 10% to 20% compared with BWA. Surprisingly, CGAP-align*, while achieving a small speed up with default settings, shortened the running time by more than 50% with loose settings, due to the high error rates of the reads in real data. While SOAPv2 was faster than all other candidates including CGAP-align*, it found less confidently mapped reads than BWA, especially for the WGS cases. On the WGS1 data, where a significant number of mismatches between the reads and reference were observed, CGAP-align and BWA confidently mapped almost 10% more reads than SOAPv2 did. In comparison to SOAPv2, CGAP-align* further shortens the performance gap while maintaining a relatively high mapping rate. Overall, it possesses impressive mapping rates in WES while the performances when mapping WGS data still need to be improved.

29.2 million read pairs (WES1), 31.6 million read pairs (WES2), 107.8 million read pairs (WGS1) and 86.3 million read pairs (WGS2) were mapped to the human genome. The run time (BWA in hours, CGAP-align and SOAP in percentage relative to BWA) on a 2.4 GHz Dual-Core AMD Opteron Processor 2216 HE with 4 threads running simultaneously (Hours & percentage relative to BWA), true positive rate (TPR) and positive predictive value (PPV). CGAP-align gave identical results to BWA with a shorter run time.

doi:10.1371/journal.pone.0061033.t001

doi:10.1371/journal.pone.0061033.t002

doi:10.1371/journal.pone.0061033.g004

Table 1. Evaluation on simulated data.

| Program          | Default Settings |         |         |         | Loose Settings |         |         |         |
|------------------|------------------|---------|---------|---------|----------------|---------|---------|---------|
|                  | Hours (%)        | TPR (%) | PPV (%) |         | %              | TPR (%) | PPV (%) |         |
| BWA              | 2.08             | 99.84   | 98.51   |         | 3.70           | 99.92   | 98.54   |         |
| CGAP-align       | 1.77             | 85.10   | 98.51   |         | 2.92           | 87.64   | 99.92   | 98.54   |
| CGAP-align       | 1.80             | 86.54   | 98.51   |         | 2.89           | 88.92   | 99.92   | 98.54   |
| Bowtie2          | 1.57             | 75.48   | 95.05   | 95.57   |                |         |         |         |
| SOAPv2           | 0.95             | 45.67   | 84.95   | 98.21   |                |         |         |         |

Table 2. Evaluation on real data.

| Program          | Default Settings |         |         |         | Loose Settings |         |         |         |
|------------------|------------------|---------|---------|---------|----------------|---------|---------|---------|
|                  | Hours (%)        | %       | Conf (%) | Hours (%) | %       | Conf (%) |         |         |
| BWA-WES1         | 3.350            | -       | 97.31   | 7.130    | -       | 97.39   |         |         |
| CGAP-align-WES1  | 2.887            | 86.18   | 97.31   | 6.375    | 89.41   | 97.39   |         |         |
| CGAP-align-WES1  | 2.698            | 80.54   | 97.31   | 3.813    | 53.48   | 97.33   |         |         |
| Bowtie2-WES1     | 3.177            | 93.04   | 98.43   |         | -       | -       |         |         |
| SOAPv2-WES1      | 1.076            | 32.12   | 93.85   |         | -       | -       |         |         |
| BWA-WES2         | 4.555            | -       | 96.84   | 11.158   | -       | 96.92   |         |         |
| CGAP-align-WES2  | 3.857            | 84.68   | 96.84   | 9.699    | 86.92   | 96.92   |         |         |
| CGAP-align-WES2  | 3.829            | 84.06   | 96.84   | 5.598    | 50.17   | 96.85   |         |         |
| Bowtie2-WES2     | 3.700            | 81.23   | 98.02   |         | -       | -       |         |         |
| SOAPv2-WES2      | 1.314            | 28.85   | 92.94   |         | -       | -       |         |         |
| BWA-WGS1         | 22.818           | -       | 91.84   | 71.349   | -       | 92.26   |         |         |
| CGAP-align-WGS1  | 20.817           | 91.23   | 91.84   | 66.262   | 92.87   | 92.26   |         |         |
| CGAP-align-WGS1  | 19.717           | 86.41   | 91.84   | 30.159   | 42.27   | 92.19   |         |         |
| Bowtie2-WGS1     | 8.433            | 36.96   | 83.31   |         | -       | -       |         |         |
| SOAPv2-WGS1      | 10.311           | 45.19   | 82.03   |         | -       | -       |         |         |
| BWA-WGS2         | 8.504            | -       | 93.69   | 20.244   | -       | 93.82   |         |         |
| CGAP-align-WGS2  | 7.629            | 89.71   | 93.69   | 19.218   | 94.93   | 93.82   |         |         |
| CGAP-align-WGS2  | 7.047            | 82.87   | 93.69   | 9.379    | 46.33   | 93.76   |         |         |
| Bowtie2-WGS2     | 5.283            | 61.12   | 84.45   |         | -       | -       |         |         |
| SOAPv2-WGS2      | 4.647            | 54.64   | 88.34   |         | -       | -       |         |         |

The 10 million read pairs were mapped to the human genome. We recorded the run time (BWA in hours, CGAP-align and SOAP in percentage relative to BWA) on a 2.4 GHz Dual-Core AMD Opteron Processor 2216 HE with 4 threads running simultaneously (Hours & percentage relative to BWA), true positive rate (TPR) and positive predictive value (PPV). CGAP-align gave identical results to BWA with a shorter run time.

Figure 4. The error rates of the simulated reads vs. the speed up of CGAP-align over BWA, defined as ((TBWA-TCGAP)/TCGAP). Six sets of simulated human genome shotgun 100 bp reads with error rates from 0 to 0.1 were generated. The speed up when using STA under default settings and when using both STA and DCDC under loose settings is reported.

doi:10.1371/journal.pone.0061033.g004
map as many reads as CGAP-align. CGAP-align retains its high recall and precision even when the error rate of the reads is high.

CGAP-align also outputs SAM files, allowing easy use of various downstream analysis tools.

CGAP-align requires a tunable amount of memory beyond that required by BWA, which equals to the size of the additional indices for STA and DCDC. To make DCDC work, a set of training reads are needed for CGAP-align to build the DCDC index. In practice, any FASTA file containing reads that are supposed to be mapped to the reference is qualified to be the training dataset for that reference. However, we recommend not using FASTA files larger than 2 GB to reduce memory cost and time of DCDC index construction.

It is very easy to migrate from using BWA to CGAP-align. For each reference sequence, only a single command line is needed to build the additional indices used by CGAP-align. After that, CGAP-align has an identical interface to that of BWA. In addition, CGAP-align is backwards compatible with BWA, so BWA can do read alignment using the FM-index produced by CGAP-align.

The main concern with using CGAP-align is that it occasionally misses alignments called by BWA. Figure 5 further investigates a real example of the cause of these differences. Consider CGAP-align mapping during the alignment depicted in Figure 5 under loose settings, where at most 5 edit costs are allowed. The backward search starts from right to left. When the search goes to the second mismatch, 3 edit costs have been counted. The D-array element at the next position (red number on the forth row) also has a value of 3, indicating that at least 6 edit costs are needed to map W to X according to DCDC’s D-array. As a result, this alignment is pruned by CGAP-align. However, at the same position in BWA’s case, the D-array value is 2 (telling us that at least 5 edit costs are needed), which does not trigger the pruning, leading to the discrepancy between the alignments. From the mechanism behind this example, we can see that such discrepancies are quite random and partially depend on the gap positions in the alignment. As previously noted, only a very small fraction of reads encounter this problem and the pruned alignments are typically quite marginal.

### Author Contributions
Conceived and designed the experiments: YC YX RC WW. Analyzed the data: YC YX RC. Contributed reagents/materials/analysis tools: YC JH WC. Wrote the paper: YC JZ YX RC. Provided data: RG.

### References
1. Li H, Ruan J, Durbin R (2008) Mapping short DNA sequencing reads and calling variants using mapping quality scores. Genome Res 18: 1851–1858.
2. Li H, Homer N (2010) A survey of sequence alignment algorithms for next-generation sequencing. Brief Bioinform 11: 473–483.
3. Ritz A, Bashir A, Raphael BJ (2010) Structural variation analysis with strobe reads. Bioinformatics 26:1291–1296.
4. Jiang H, Wang WH (2008) SeqMap: mapping massive amount of oligonucleotides to the genome. Bioinformatics 24: 2395–2396.
5. Schatz MC (2005) CloudBurst: highly sensitive read mapping with MapReduce. Bioinformatics 21: 1363–1369.
6. Li R, Li Y, Kristiansen K, Wang J (2008) SOAP: short oligonucleotide alignment program. Bioinformatics 24: 713–714.
7. Campana D, Albiero A, Bilardi A, Caniato E, Forcato C, Manavski S, Vitulo N, Valle G (2009) PASS: a program to align short sequences. Bioinformatics 25: 967–978.
8. Eaves HL, Gao Y (2009) MOM: maximum oligonucleotide mapping. Bioinformatics 25: 969–970.
9. Kim YJ, Teletia N, Ruotti V, Maher CA, Chinnaiyan AM, et al. (2009) ProbeMatch: a tool for aligning oligonucleotide sequences. Bioinformatics 25: 1424–1425.
10. Nahm U, Myers G (1998) Suffix arrays: a new method for on-line string searches. In Proceedings of the first annual ACM-SIAM SODA: 90: 327.
11. Abouellhoda MI, Kurtz S, Ohelundh E (2004) Replacing suffix trees with enhanced suffix arrays. Journal of Discrete Algorithms 2: 3–36.
12. Ferragina P, Manzini G (2000) Opportunistic Data Structures with Applications. Proceedings of the 41st Annual Symposium on FOCS. pp. 390–398.
13. Phoophakdee B, Zaki MJ (2007) Genome-scale disk-based suffix tree indexing. SIGMOD. ACM pp. 833–844.
14. Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, et al. (2004) Versatile and open software for comparing large genomes. Genome Biol 5: R12.
15. Li R, Yu C, Li Y, Lam TW, Yiu SM, et al. (2009) SOAP2: an improved ultrafast tool for short read alignment. Bioinformatics 25: 1966–1967.
16. Langmead B, Trapnell C, Pop M, Salzberg SL (2009) Ultrafast and memory-efficient alignment of short DNA sequences to the human genome. Genome Biol 10: R25.
17. Langmead B, Salzberg SL (2012) Fast gapped-read alignment with Bowtie 2. Nat Methods 9: 357–359.
18. Li H, Durbin R (2009) Fast and accurate read alignment with Burrows-Wheeler. Bioinformatics 25:1754–1760.
19. McCreight EM (1976) A Space-Economical Suffix Tree Construction Algorithm. Journal of the ACM 23: 262–272.
20. Tata S, Hawkins R, Patel J (2003) Practical Suffix Tree Construction. Proceedings of the 30th International Conference on VLDB. pp. 36–47.
21. Ko P, Ahura S (2003) Space efficient linear time construction of suffix arrays. In Combinatorial Pattern Matching. LNCS 2676, Springer, pp. 203–210.
22. Burrows M, Wheeler D (1994) A block sorting lossless data compression algorithm. 124, Palo Alto, CA, Digital Equipment Corporation.
23. Hon WK, Lam TW, Sung WK, Tse WL, Wong CK, et al. (2004) Practical Aspects of Compressed Suffix Arrays and FM-index. Proceedings Of The Sixth Workshop On ALENEX And The First Workshop On ANALCO. pp. 31–38.
24. Aho AV, Corasick MJ (1975) Efficient string matching: An aid to bibliographic search. Communications of the ACM 10: 333–340.
25. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, et al. (2009) The Sequence alignment/map (SAM) format and SAMtools. Bioinformatics 25: 2078–2079.