Analysis of ADAM17 Polymorphisms and Susceptibility to Sporadic Abdominal Aortic Aneurysm

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Key Words
ADAM17 • Abdominal aortic aneurysm • Polymorphism

Abstract
Background: Accumulating evidence suggests that the principal TNF-α converting enzyme, a disintegrin and metalloproteinase 17 (ADAM17), is involved in the development of human abdominal aortic aneurysm (AAA). However, the association between ADAM17 gene polymorphisms and AAA has not been explored. The present study was aimed to determine the association between ADAM17 promoter polymorphisms and AAA. Methods: A total of 316 patients with AAA and 306 age-matched healthy controls were enrolled in this study. Two ADAM17 promoter polymorphisms (rs12692386 and rs1524668) were determined. Real-time PCR was employed to detect the expression of ADAM17. Results: Overall, there was a significant difference in the frequency of the genotype rs12692386 between the AAA and control subjects (P=0.0096). Furthermore, men with the rs12692386 AG genotype conferred a higher risk of developing AAA (P=0.0058). Additionally, the rs12692386 mutated AG genotype of ADAM17 was significantly associated with increased ADAM17 expression (P=0.035) and TNF-α production (P=0.042) in AAA patients. In contrast, the allele frequency of rs1524668 was not statistically associated with AAA. Conclusions: Our findings indicate a positive association between the rs12692386 polymorphism of ADAM17 and AAA. This new knowledge about ADAM17 identifies a role for ADAM17 in the pathophysiology of AAA and has important clinical implications with regard to potential therapeutics.
Introduction

Human abdominal aortic aneurysm (AAA) is the most common type of aneurysm in humans with a mortality rate associated with rupture as high as 90% [1]. Multiple characteristics of AAA pathogenesis involve chronic inflammation [2], autoimmunity, increased neoangiogenesis [3], enhanced oxidative stress [4], and degradation of the extracellular matrix (ECM) [5]; however, the precise mechanisms of AAA formation and progression remain elusive [6].

Several studies have suggested that proinflammatory cytokines, especially tumor necrosis factor (TNF)-α, play a large part in the etiology of AAA [7-9]. TNF-α, a proinflammatory cytokine, is involved in the regulation of a wide spectrum of biological processes, including cell proliferation, differentiation, apoptosis, lipid metabolism, and coagulation. TNF-α levels were reported as increased in plasma and in the aorta in human AAA patients [9]. Blocking TNF-α attenuated aneurysm formation in CaCl$_2$-induced AAA in mice through the inhibition of chronic inflammation and ECM disruption in the aortic wall; this result indicates that TNF-α plays a critical role in the pathogenesis of AAA [8]. TNF-α is generated via cleavage of the 32 kDa transmembrane precursor by TNF-α-converting enzyme (TACE), also referred to as a disintegrin and metalloproteinase (ADAM)-17. TACE belongs to the ADAMs family, which is involved in a number of biological processes, such as cell fate determination, cell migration, wound healing, neurite and axon guidance, heart development, immunity, cell proliferation, angiogenesis and inflammation [10]. TACE expression was increased in aortic lesions and a higher vascular shedding activity of TACE substrates (TNF and its receptors) was measured ex vivo from aortic explants of the apoE$^{-/-}$ mice [9, 11]. These results highlight the predominant role of TNF shedding via TACE activity in the inflammatory reaction that occurs in the lesion [12]. A deficiency in TACE has been associated with reduced expression of pro-inflammatory/chemotactic cytokines and adhesion molecules, macrophage infiltration, VEGF-A expression and neoangiogenesis, MMP activation and ECM degradation [9]. In vivo evidence also demonstrated that a lack of TACE attenuated aneurysm formation in mice with AAA [13]. In summary, these lines of evidence have led us to formulate the hypothesis that ADAM17 could be of pathogenic significance in AAA.

The human ADAM17 gene is located on chromosome 2 and consists of 19 exons that span 55 kb. ADAM17 polymorphisms have been investigated in relation to Alzheimer’s disease [14], obesity [15] and Coronary artery disease [16]. Morange et al. [16] reported that the C-154A and Ser747Leu mutations of the ADAM17 gene are associated with TNF-α plasma levels and the risk of cardiovascular death, respectively. Junyent et al. [15] suggested that two ADAM17 polymorphisms, rs11684747 and rs10495563, are related to obesity and insulin resistance-related phenotypes (higher insulin and lower HDL-C concentrations). However, a genetic association between human ADAM17 polymorphisms and AAA has not yet been determined. Therefore, we carried out a case-control study to ascertain whether ADAM17 promoter polymorphisms are associated with AAA susceptibility.

Materials and Methods

Study population

We enrolled 316 cases that were randomly selected from AAA patients who were admitted to the Department of Vascular Surgery at the First Affiliated Hospital of Harbin Medical University from 2004 to 2012. Their ages ranged from 45 to 86 years, and 242 cases were male, while 74 cases were female. The AAA diameter of all patients ranged from 32 to 78 mm. AAA was diagnosed by using computed tomography of the abdomen, which can detect dilatation of the abdominal aorta. Patients with inflammatory disorders, connective tissue disease, chronic renal failure, malignant disease or familial AAAs were excluded from the study. Sixty-nine patients with a maximal aortic diameter greater than 50 mm were admitted for elective surgery. Three hundred and six sex- and age-matched control subjects from the same geographical area (Central Harbin) were recruited within the same period from among patients who were attending vascular
outpatient clinics and those who were admitted for surgery at the First Affiliated Hospital of Harbin Medical University.

DNA Isolation and Genotyping

Genomic DNA was isolated from peripheral blood using the EZ-10 Spin Column Whole Blood Genomic DNA Isolation Kit (Sangon Biotech, Shanghai, China), according to manufacturer’s instruction. The two ADAM17 promoter polymorphisms (rs12692386 and rs1524668) were selected according to the results of the ADAM17 promoter sequencing reported by Wang et al. [14]. The polymorphisms of genes were analyzed by SNaPshot Multiplex Kit (Applied Biosystems Co., Ltd., Foster City, CA, USA). The primers used in the SNaPshot were as follows: rs12692386F: GGCCTAGCCCCTCAATCCTCTT, rs12692386R: TTTTTTGGTGAACGCCACCTGCCTTC; rs1524668F: AAAAACGTTGGTACATGTTGCTG, rs1524668R: TTTTTTTTTTTTTTTTTTTTTTTTTTCTGAAATCCGATCACCATATA. SNaPshot reactions were carried out in a 10 μL final volume containing 5 μL SNaPshot Multiplex Kit (ABI), 1 μL primer mix, 2 μL water, and 2 μL templates consisting of the multiplex PCR products from the different genes. SNaPshot response procedures: (1) initial denaturation at 96°C for 1 min, (2) denaturation at 96°C for 10 seconds, (3) annealing at 52°C for 5 seconds, (4) extension at 60°C for 30 seconds, and (5) for a total of 28 cycles. Amplified samples were stored at 4°C. Extension products were purified by 1-h incubation with 1 U of shrimp alkaline phosphatase (Takara: Otsu, shiga, Japan) at 37°C and 75°C for 15 min to inactivate the enzyme. The purified products (0.5 μL) were mixed with 9 μL of Hi-Di and 0.5 μL Liz120 SIZE STANDARD. Samples were incubated at 95°C for 5 min and then loaded on an ABI 3130XL DNA sequence detector for capillary electrophoresis. The experimental results were analyzed with GeneMapper 4.0 (Applied Biosystems Co., Ltd.).

RNA extraction

A total of 50 human AAA samples were obtained from AAA patients who underwent surgical repair at the Department of Vascular Surgery of the First Affiliated Hospital of Harbin Medical University. The control aortic samples were obtained from 50 autopsy specimens from patients who died of unrelated causes at the First Affiliated Hospital of Harbin Medical University. The study was approved by the Hospital Ethics Committee, and written informed consent was obtained from all subjects.

The aortic specimens were immediately segmented in blocks of 3-4 mm, snap-frozen in liquid nitrogen and stored at −80°C for the later RNA and protein extraction. Total RNA was extracted using the RNAprep pure Blood Kit (TianGen Biotech, Beijing, China) according to the manufacturer’s instructions.

Real-time PCR

The isolated RNA was transcribed into cDNA using the cDNA Synthesis Kit RevertAid (Thermo) according to the manufacturer’s instructions. Quantitative real-time PCR was performed using the SYBR Green RT-PCR Kit (Takara). Real-time RT-PCR primers for both human GAPDH and ADAM17 coding sequences were designed using Primer Premier 5 software (Applied Biosystems). The primer sequences used were as follows: ADAM17 sense primer; CTG TGG TGC AAA AGC AGA AA; ADAM17 anti-sense primer; TGC CAA ATG CCT CAT ATT CA; GAPDH sense primer; GAA GGG CTC ATG ACC ACA GCA GTC CAT; and GAPDH anti-sense primer; TCA TTT TGG TAC CAG GAA ATG AGC TT. The ADAM17 and GAPDH mRNA levels were measured in triplicate, and the relative mRNA quantity was determined by the $2^{-\Delta\Delta Ct}$ method and normalized with respect to the housekeeping gene glyceraldehyde-3-phosphate dehydrogenase (GAPDH). The GAPDH expression used for normalization did not vary significantly among the groups. Real-time PCR was performed using a LightCycler®480 sequence detector system (Roche Applied Sciences). The following PCR conditions were used over 40 cycles: 5 min at 95°C, 5 sec at 95°C, 20 sec at 59°C, and 10 sec at 72°C. The relative expression levels for each sample were calculated based on three technical replicates. The amplification products were validated by melting curve analysis.

Enzyme-linked immunosorbent assay (ELISA)

Blood specimens for the measurement of serum cytokines concentrations were drawn in EDTA-containing tubes and centrifuged at low speed, and the plasma aliquots were stored at −20°C. Levels of TNF-α were measured in duplicate using Quantikine sandwich ELISA kits (R&D Systems, Minneapolis, MN, USA) according to the manufacturer’s instructions.
Ultrasound Measurement of Abdominal Aortic Aneurysms

AAA size was measured as the greatest diameter of the infrarenal aorta by using ultrasound. The maximum transverse and anteroposterior diameter of the infrarenal abdominal aorta was measured by an experienced vascular sonographer using a color Doppler ultrasound (GE Healthcare Technologies, Ultrasound, Milwaukee, WI, USA) with a 5 MHz transducer. An abdominal aortic aneurysm is defined as an enlargement of the aorta of at least 1.5 times its normal diameter or greater than 3 cm diameter in total. The reproducibility of aortic measurements is regularly assessed in the Vascular Surgery Department.

Statistical Analyses

Statistical analyses were performed with SPSS software, version 19.0 (IBM, Armonk, NY, USA). The clinical data are expressed as the means ± standard deviation (SD) for continuous variables and as the medians and percentage for quantitative variables; a chi-squared test and Student’s t-test were used to compare variables between the two groups. Allele frequencies were calculated from the genotypes of all of the subjects. The allele and genotype frequencies of ADAM17 between the patients and control subjects were compared using Fisher’s exact test or the Chi-squared test. Deviation of the genotype or allele frequency was assessed using Hardy-Weinberg equilibrium (HWE). The relationships between different genotypes of ADAM17 and AAA were evaluated by an ANOVA. The comparisons of ADA17 expression levels between the patients and controls were evaluated using Student’s t-test for the normally distributed data or a Mann-Whitney-U test for the non-parametric data. Statistical differences with respect to plasma TNF-α level between independent groups were calculated using the Kruskal-Wallis test. The associations of gene polymorphisms with aortic diameter were analyzed with a multiple linear regression model (SAS, version 6.12; SAS Institute Inc., Cary, NC, USA). P values less than 0.05 were considered to be statistically significant.

Results

Demographic characteristics

The demographic characteristics of all the participants in the study are summarized in Table 1. Of the 622 participants, 316 were patients with AAA and 306 were healthy controls. There were no significant differences between the patients and controls with respect to age, diabetes, or hypertension. The mean age was 70.3 years old (±6.8 years) for the AAA subjects and 69.9 years old (±7.2 years) for the control subjects. Significant gender and aortic diameter differences in the occurrence of AAA were observed. Consistent with other reports [17, 18], AAA was substantially more common among men than among women in this study. Significant differences were also found for risk factors including smoking status and dyslipidemia.

ADAM17 gene polymorphisms and the risk of AAA

The genotype and allele frequencies of the ADAM17 polymorphisms are shown in Table 2. No deviation from the Hardy-Weinberg equilibrium was observed for the polymorphisms that were examined among the genotype distributions between the AAA and control subjects (data not shown).

A comparison of genotype distributions between the AAA and control subjects using the χ2-test revealed a statistical association (P=0.0096) between the ADAM17 rs12692386 polymorphism and the risk of developing AAA. In a dominant model (AA+AG versus GG), no significant difference was detected between the AAA and control groups (P=0.11). However, in a recessive model (AA versus AG+GG), a significant difference was observed in the AAA group when compared with the controls (P=0.022). However, the rs12692386 G allele did not show a significant difference in the AAA group when compared with the controls (P=0.14). In contrast, there was no significant association between the ADAM17 rs1524668 polymorphism and the risk of developing AAA (P=0.77). Accordingly, the frequency of the rs1524668 polymorphism was not significantly different between the two groups in either the dominant (P=0.58) or recessive models (P=0.86). Additionally, there were no
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Table 1. Characteristics of AAA cases and controls. Continuous data are presented as mean ± SD, median (range) or n(%)

| Variables         | AAA (n=316) | control (n=306) | P value |
|-------------------|-------------|-----------------|---------|
| Mean age (years)  | 70.3±6.8    | 69.9±7.2        | 0.79    |
| Male/female       | 242/74      | 155/151         | < 0.001 |
| Aortic diameter (mm) | 55.1±8.9   | 21.8±5.7        | < 0.001 |
| Smokers, n (%)    | 173 (54.7)  | 104 (34.0)      | < 0.001 |
| Diabetes, n (%)   | 65 (20.6)   | 53 (17.32)      | 0.28    |
| Hypertension, n (%) | 121 (38.3) | 105 (34.3)      | 0.71    |
| Dyslipidemia n (%) | 148 (46.8) | 106 (34.6)      | < 0.001 |

Table 2. Frequencies of ADAM17 geneotypes and alleles in AAA patients and controls

| Genotypes            | AAA patients n=316(%) | Controls n=306(%) | OR (95%CI) | P value |
|----------------------|-----------------------|-------------------|------------|---------|
| rs12692386           |                       |                   |            |         |
| AA                   | 156 (49.3)            | 179 (58.6)        |            | 0.0096  |
| AG                   | 155 (49.1)            | 116 (38.0)        |            |         |
| GG                   | 5 (1.6)               | 11 (3.4)          |            |         |
| Dominant model AA/AG vs GG | 311 (98.4)   | 295 (96.4)        | 2.32 (0.80-6.76) | 0.11    |
| Recessive model AA vs AG/GG | 160 (50.6) | 127 (41.5)        | 0.69 (0.50-0.95) | 0.022   |
| A allele             | 467 (73.9)            | 474 (77.5)        | 1.00 (reference) |         |
| G allele             | 165 (26.1)            | 138 (22.5)        | 1.21 (0.94-1.57) | 0.14    |
| rs1524668            |                       |                   |            |         |
| AA                   | 241 (76.27)           | 239 (78.98)       |            | 0.77    |
| AC                   | 73 (23.10)            | 66 (20.68)        |            |         |
| CC                   | 2 (0.63)              | 1 (0.34)          |            |         |
| Dominant model AA/AC vs CC | 314 (99.37)    | 305 (99.67)       | 0.51 (0.05-5.71) | 0.58    |
| Recessive model AA vs AC/ CC | 75 (23.73) | 67 (21.90)        | 0.98 (0.77-1.24) | 0.86    |
| A allele             | 555 (87.82)           | 544 (88.89)       | 1.00 (reference) |         |
| G allele             | 77 (12.18)            | 68 (11.11)        | 1.1 (0.78-1.57) | 0.56    |

Unexpectedly, when the sample was stratified according to either gender or smoking status, a borderline association in the ADAM17 rs1524668 genotype was determined between AAA patients and the controls (P=0.045 and P=0.035, respectively). However, when the sample was stratified by either age, diabetes or hypertension, no significant differences in the genotype or allele frequencies were detected between the AAA and control groups for either of the two ADAM17 polymorphisms (rs12692386 and rs1524668) (P>0.05).

**Associations between ADAM17 gene polymorphisms and demographic characteristics**

The associations between ADAM17 gene polymorphisms and demographic characteristics are shown in Tables 3 and 4. In an analysis stratified by gender, increased risk was associated with the variant genotype rs12692386 AG in male patients (P=0.0058). No differences in the ADAM17 rs12692386 genotype and allele frequencies were observed between AAA cases and control subjects when the sample was stratified by smoking status. Unexpectedly, when the sample was stratified according to either gender or smoking status, a borderline association in the ADAM17 rs1524668 genotype was determined between AAA patients and the controls (P=0.045 and P=0.035, respectively). However, when the sample was stratified by either age, diabetes or hypertension, no significant differences in the genotype or allele frequencies were detected between the AAA and control groups for either of the two ADAM17 polymorphisms (rs12692386 and rs1524668) (P>0.05).

**Effects of ADAM17 gene polymorphisms on ADAM17 expression**

A comparative analysis was performed on the ADAM17 mRNA expression levels among 50 AAA patients' samples and 50 control samples. Consistent with a previous report [9], the mean value of the ADAM17 mRNA expression was significantly higher in the samples of AAA patients than in the controls (P=0.018) (Fig. 1).
In addition, we assessed whether there was an association between the mean values of ADAM17 mRNA levels and the ADAM17 genotype in AAA patients. The results are presented in Fig. 2. A significant increase in ADAM17 mRNA expression was observed in AAA patients who carried the mutated rs12692386 AG ADAM17 genotype (P=0.035), while in control subjects, no association was determined between the individuals with the major AA genotype and those who carried the mutated AG genotype (P>0.05) (Fig. 2A). A similar trend, although not significant, was observed in patients carrying the rs12692386 GG ADAM17 genotype (P=0.081) (Fig. 2A). With regard to the rs1524668 polymorphism, no statistically significant

### Table 3.

A comparison between the baseline characteristics of the rs12692386 genotypes and alleles in the AAA patient and control groups. $P_A$: p value of the difference in genotype between the case and control groups. $P_G$: p value of the difference in alleles between the case and control groups.

| Characteristics | rs12692386 AAA patient group | rs12692386 control group | $P_A$ | $P_G$ |
|-----------------|-------------------------------|--------------------------|-------|-------|
| Genotype n (%)  | Allele n (%)                  | Genotype n (%)            | Allele n (%)                  |       |       |
| AA              | AG                            | GG                       | A    | G     |       |       |
| Age ≥70 years   | 80(44.7)                      | 97(54.2)                 | 2(1.1)| 101(28.2)| 98(57.0)| 62(39.2)| 6(3.8)| 24(76.6)| 74(23.4)| 0.13 | 0.20 |
| <70 years       | 75(54.7)                      | 59(43.1)                 | 3(2.2)| 209(76.3)| 65(23.7)| 88(59.5)| 55(40.5)| 5(3.4)| 23(71.0)| 65(29.0)| 0.90 | 0.67 |
| Gender Male     | 116(47.9)                     | 125(53.7)                | 1(0.4)| 357(73.8)| 127(26.2)| 94(60.6)| 56(39.4)| 5(3.2)| 24(70.7)| 66(29.3)| 0.005 | 0.98 |
| Female Smoking  | 39(52.7)                      | 31(41.9)                 | 4(5.4)| 109(73.6)| 39(26.4)| 84(55.6)| 61(44.4)| 6(4.0)| 22(75.8)| 73(24.2)| 0.67 | 0.39 |
| Yes             | 78(45.1)                      | 91(52.6)                 | 4(2.3)| 247(71.4)| 99(28.6)| 61(58.7)| 39(41.3)| 4(3.8)| 16(77.4)| 47(22.6)| 0.19 | 0.14 |
| No              | 77(53.8)                      | 65(45.5)                 | 1(0.7)| 219(76.6)| 67(23.4)| 117(57.9)| 78(42.1)| 7(3.5)| 31(72.2)| 29(27.8)| 0.97 | 0.96 |
| Diabetes        | 32(49.0)                      | 30(46.2)                 | 3(4.6)| 94(72.3)| 36(27.7)| 38(56.6)| 21(43.4)| 2(3.8)| 81(76.4)| 25(23.6)| 0.085 | 0.68 |
| Hypertension    | 123(49.0)                     | 126(50.2)                | 2(0.8)| 372(74.1)| 130(25.9)| 148(58.5)| 96(41.5)| 9(3.6)| 39(77.5)| 114(22.5)| 0.97 | 0.81 |

### Table 4.

A comparison between the baseline characteristics of the rs1524668 genotypes and alleles in the AAA patient and control groups. $P_A$: p value of the difference in genotype between the case and control groups. $P_G$: p value of the difference in alleles between the case and control groups.

| Characteristics | rs1524668 AAA patient group | rs1524668 control group | $P_A$ | $P_G$ |
|-----------------|-------------------------------|--------------------------|-------|-------|
| Genotype n (%)  | Allele n (%)                  | Genotype n (%)            | Allele n (%)                  |       |       |
| AA              | AC                            | CC                       | A    | C     |       |       |
| Age ≥70 years   | 140(78.2)                     | 38(21.2)                 | 1(0.6)| 318(88.8)| 40(11.2)| 119(75.3)| 39(24.7)| 0| 27(87.7)| 39(12.3)| 0.58 | 0.37 |
| <70 years       | 102(74.5)                     | 33(24.1)                 | 2(1.4)| 237(85.6)| 37(14.5)| 114(77.0)| 33(23.2)| 1(0.7)| 26(88.2)| 35(11.8)| 0.5 | 0.84 |
| Gender Male     | 192(79.3)                     | 47(19.4)                 | 3(12.4)| 431(89.0)| 53(11.0)| 119(76.8)| 35(22.6)| 1(0.6)| 27(81.1)| 37(19.9)| 0.045 | 0.007 |
| Female Smoking  | 50(67.6)                      | 24(32.4)                 | 4(2.3)| 234(88.8)| 16(22.4)| 114(75.5)| 37(24.5)| 0| 26(87.7)| 37(12.3)| 0.57 | 0.90 |
| Yes             | 139(80.3)                     | 31(17.9)                 | 3(1.8)| 309(93.3)| 7(10.7)| 83(79.8)| 21(20.2)| 0| 187(89.9)| 21(10.1)| 0.035 | 0.21 |
| No              | 103(72.0)                     | 40(28.0)                 | 0| 246(86.0)| 40(14.0)| 150(74.3)| 51(25.2)| 1(0.5)| 35(86.9)| 53(13.1)| 0.46 | 0.28 |
| Diabetes        | 53(81.5)                      | 12(18.5)                 | 0| 118(90.8)| 12(9.2)| 41(77.4)| 12(22.6)| 0| 94(87.7)| 12(13.3)| 0.44 | 0.25 |
| Hypertension    | 109(75.3)                     | 59(23.5)                 | 3(12.2)| 437(87.1)| 65(12.9)| 192(75.9)| 60(23.7)| 1(0.4)| 44(87.7)| 62(12.3)| 0.89 | 0.79 |

In addition, we assessed whether there was an association between the mean values of ADAM17 mRNA levels and the ADAM17 genotype in AAA patients. The results are presented in Fig. 2. A significant increase in ADAM17 mRNA expression was observed in AAA patients who carried the mutated rs12692386 AG ADAM17 genotype (P=0.035), while in control subjects, no association was determined between the individuals with the major AA genotype and those who carried the mutated AG genotype (P>0.05) (Fig. 2A). A similar trend, although not significant, was observed in patients carrying the rs12692386 GG ADAM17 genotype (P=0.081) (Fig. 2A). With regard to the rs1524668 polymorphism, no statistically significant
difference in ADAM17 mRNA expression was determined between the population carrying the mutated AC or CC genotype and the population carrying the major AA genotype in either the AAA patients or control subjects (Fig. 2B).

Effects of ADAM17 gene polymorphisms on TNF-α expression
Since ADAM17 is a TNF-α-converting enzyme, another experiment was performed to determine whether the ADAM17 gene polymorphisms had an effect on TNF-α production in
controls and AAA patients. The results are shown in Fig. 3. Generally, the plasma expression levels of TNF-α in AAA patients were significantly higher than in controls (P<0.05) (Fig. 3). The AAA patients carrying the rs12692386 AG ADAM17 genotype presented with higher plasma TNF-α levels compared to the homologous AA or GG group (P=0.042) (Fig. 3A). The expression of TNF-α did not vary significantly between the population carrying the mutated AC or CC genotype and the population carrying the major AA genotype in either the AAA patients or control subjects (P>0.05) (Fig. 3B).

**Effects of ADAM17 gene polymorphisms on aortic diameter**

It is well established that the aortic diameter in AAA cases is an important predictor of rupture. In the present study, associations of the ADAM17 gene polymorphisms with AAA aortic diameter were explored, and the results are shown in Fig. 4. The mean value of the AAA aortic diameter in the AAA patients with the variant genotypes rs12692386 (AG and GG) and rs1524668 (AC and CC) was not significantly different from that of patients with the major AA genotype (P>0.05) (Fig. 4A). We also evaluated the effect of the two ADAM17 polymorphisms on aortic diameter in patients and controls stratified by gender, and found no difference in aortic diameter either in female or male patients (P>0.05) (Fig. 4B and Fig. 4C).

**Discussion**

In this hospital-based case control study, we show for the first time that the rs12692386 polymorphism in the promoter region of ADAM17 is associated with AAA, while the rs1524668 ADAM17 polymorphism is not associated with AAA. Additionally, the rs12692386 A>G polymorphism in the promoter region may affect the expression of ADAM17.

There is increasing evidence of the potential impact of members of the ADAM family on the pathogenic mechanisms that lead to AAA. ADAMS 8, 9, 10, 12, 15 and 17 were found to be expressed in both AAA and control aorta [19], and ADAM17 (TACE) was upregulated in human AAA aortic specimens and was associated with chronic inflammation, increased neoangiogenesis and ECM disruption [9]. The role that ADAM17 plays in the inflammation and...
proteolytic degradation of ECM in the vessel wall markedly contributes to the formation and rupture of an AAA [5, 6]. ADAM17 is capable of cleaving several other cell surface molecules, such as TNF-α, TNFR1 and TNFR2, as well as IL-6R and IL-1RII, which are considered to be involved in inflammatory processes of the vasculature [10]. The progression of AAA rupture can result from excessive degradation of the extracellular matrix components of the vessel wall, especially that of elastic and collagen fibers. As active metalloproteinases, ADAMs bind to various integrins and can degrade components of the ECM; ADAM17, which acts on collagen XVII as a substrate, may be involved in this process [20]. Additionally, ADAM17 has been reported to promote the transcription of MMP-2, which is thought to play a crucial role in ECM degradation [21]. Thus, increased expression of ADAM17 may facilitate the proliferation and invasive characteristics of inflammatory cells and the disruption of the ECM and therefore contribute to AAA progression [22]. The observations that ADAM17 is present in aortic aneurysm tissue and that its expression is significantly increased in aneurysms have confirmed its involvement in AAA pathogenesis. Despite these advances, the ADAM17 alleles that contribute to AAA remain undiscovered.

To date, only three genome-wide association studies (GWAS) for AAAs have been reported [23-25]. However, the reported odds ratios (ORs) for the identified risk alleles in these studies are not in agreement with the strong heritability patterns that have been observed in epidemiologic studies, which suggests that there are multiple risk loci associated with the disease and many remain unidentified [26]. Previous reports have investigated functional and nonfunctional single nucleotide polymorphisms (SNPs) in patients with aneurysmal disease [27, 28]. However, no single genetic polymorphism or defect has been identified as a common denominator for AAAs. In our case control study, we show for the first time that the rs12692386 ADAM17 polymorphism is associated with the risk of developing AAA, while the rs1524668 ADAM17 polymorphism is not associated with AAA. Carriers of the rs12692386 AG genotype showed a significantly greater prevalence of AAA than subjects with the AA or GG genotype. Our findings suggest that the rs12692386 polymorphism of ADAM17 may serve as a genetic marker for predicting the occurrence of AAA in high-risk subjects.

AAA has a complex pathophysiology whereby both environmental and genetic factors play important roles [26]. Male sex is an established contributing factor for AAA, and males form AAAs four to five times more often than females [29]. Adriana et al. reported that estrogen receptor alpha was increased in the aortic wall and correlated inversely with MMP activity and AAA formation [30]. Estrogen decreases inflammation, which suggests a protective role for circulating estrogens during AAA formation in females. In addition to estrogens, nitric oxide (NO) and reactive oxygen species are circulating factors that may be responsible for gender differences in AAA formation [31]. Being male is not only genetic but also affects environmental risk factors, such as smoking. Accumulating epidemiological evidence repeatedly confirmed that smoking is a significant risk factor for AAA [26]. The relative risk for current smokers is at least 2-fold, and the association of having ever smoked with AAA is 2.5 times greater than the association of having ever smoked with CHD [26, 32]. Continued smoking also results in increased AAA expansion, a greater risk of rupture and a worse prognosis [33]. Smokers have elevated levels of elastase activity in the arterial wall, which leads to weakening of the aorta [34]. Stolle et al. reported that cigarette mainstream smoke could enhance the proteolytic activity of MMPs, including MMP-2 and -9, induced by Ang II and accelerate both the formation and severity of AAAs in apoE-/- mice [35]. Additionally, nicotine, a principal tobacco alkaloid, was shown to play a proinflammatory role in vasculature in vivo and in vitro [36]. Together, these findings suggest that cigarette smoke may stimulate the formation and progression of AAAs. In our present study, when the rs12692386 genotype and allele frequency of ADAM17 was further stratified by age, gender, smoking status, diabetes and hypertension, an increased risk was found in the male subgroup of AAA patients when compared to control subjects. A significant association in the ADAM17 rs1524668 genotype was also determined between AAA patients and the controls in males and in the smoking subgroup. Thus, it is highly likely that environmental risk factors, gender and smoking interact with an ADAM17 genetic predisposition for the development of AAA.
In the present study, we employed real-time PCR to detect ADAM17 mRNA expression in human AAA tissues. Consistent with a previous study [9], the expression of ADAM17 in human AAA samples was significantly increased in the AAA patients in comparison to the control subjects. We also found that the individuals who carried the ADAM17 rs12692386 mutated AG genotype had higher ADAM17 mRNA expression levels compared with the population that carried the homologous AA or GG genotype. The rs12692386 A>G polymorphism in the ADAM17 promoter region could be a functional SNP. The mechanism by which the rs12692386 ADAM17 polymorphism contributes to elevated ADAM17 expression is unknown. Given that the two SNPs map to promoter regions, transcriptional enhancers and other regulatory elements in the promoter regions might play a regulatory role. Moreover, computational analysis of the rs12692386 A>G polymorphism by MAPPER indicated a potential allele-specific binding site for the upstream stimulatory factor (USF) transcription factor [37]. The rs12692386 A>G ADAM17 polymorphism could affect binding to USF and thereby regulate gene expression.

The formation and expansion of an AAA result from extracellular matrix degradation, increased proteolytic activity, and widespread inflammation. Logically, treatment with anti-inflammatory drugs could inhibit the expansion of AAAs and reduce AAA-size [7]. A platelet P2Y12 receptor inhibitor reduces vascular inflammation and thus suppresses AAA expansion in apoE-/ mice [38]. Knockout of the ADAM17 inhibitor, Tissue Inhibitor of Metalloproteinase 3 (TIMP3) in vivo exacerbates AAA expansion and inflammation in the abdominal aorta [39]. These lines of evidence indicate that inhibiting ADAM17 or inflammation may alleviate AAA expansion. Our present study did not demonstrate any significant association between aortic diameter and the ADAM17 polymorphisms, even taking the gender status into consideration, which suggests that these polymorphisms do not affect AAA growth. A previous study concerning an MMP-2 1306 C/T polymorphism also found no association between the polymorphisms and aortic diameter [40]. Our findings provide little support for ADAM17 polymorphisms contributing to AAA expansion. Nevertheless, the exact mechanism of AAA expansion remains under investigation. AAA expansion may involve multiple factors, and the positive effect associated with ADAM17 polymorphisms may be counteracted by other factors (hormonal factors, smoking, hypertension, diabetes and other risk factors).

Despite the results described above, limitations of the present study should be acknowledged. The small number of participants in this study was insufficient and therefore may have led to nonrepresentative results. Thus, this limitation may have suppressed a true relationship due to a type II statistical error. However, notably, the prevalence of AAAs in the Chinese population is comparatively lower than that in Caucasian populations [41]. The mechanism that underlies this interesting ethnic difference remains unclear. Selection bias in the patient or control populations may not be entirely excluded. Specifically, controls were not matched for environmental risk factors of aortic aneurysms because we compared the allele frequencies in the samples to those in the population from which they came. Selection of controls based upon specific phenotypes (such as smoker/non-smoker) would in itself risk introducing bias and would therefore be irrelevant. The other risk factors in the study group, such as age, gender, smoking, hypertension, diabetes or hyperlipaemia may have complicated the association between ADAM17 polymorphisms and AAA. In addition, other functional polymorphisms may influence the expression of ADAM17, and their combined effects must be studied to better predict the occurrence, severity, and outcome of AAA. The results obtained in this study require confirmation in independent, larger populations and in different ethnic backgrounds before the conclusions can be considered definitive and useful for estimating an individual’s risk for developing AAA.

In conclusion, our study shows for the first time a significant association between the ADAM17 rs12692386 polymorphism and an increased risk of AAA. ADAM17 is therefore a promising new genetic candidate that plays a role in regulating the development of AAA. However, more work is required to further understand the role of ADAM17 in the pathogenesis of AAA and to clarify its prognostic and therapeutic potential.
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Disclosure Statement

The authors have no actual or potential conflicts of interest related to this manuscript. Appropriate approval was obtained, and appropriate procedures were followed concerning human subjects.

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