Ability of dietary factors to affect homocysteine levels in mice: a review

Christine Brütting*, Pia Hildebrand, Corinna Brandsch and Gabriele I. Stangl

Abstract
Homocysteine is associated with several diseases, and a series of dietary factors are known to modulate homocysteine levels. As mice are often used as model organisms to study the effects of dietary hyperhomocysteinemia, we collected data about concentrations of vitamin B12, vitamin B6, folate, methionine, cystine, and choline in mouse diets and the associated plasma/serum homocysteine levels. In addition, we more closely examined the composition of the control diet, the impact of the mouse strain, sex and age, and the duration of the dietary intervention on homocysteine levels. In total, 113 out of 1103 reviewed articles met the inclusion criteria. In the experimental and control diets, homocysteine levels varied from 0.1 to 280 µmol/l. We found negative correlations between dietary vitamin B12 (\(\rho = -0.125; p < 0.05\)), vitamin B6 (\(\rho = -0.191; p < 0.01\)) and folate (\(\rho = -0.395; p < 0.001\)) and circulating levels of homocysteine. In contrast, a positive correlation was observed between dietary methionine and homocysteine (methionine: \(\rho = 0.146; p < 0.05\)). No significant correlations were found for cystine or choline and homocysteine levels. In addition, there was no correlation between the duration of the experimental diets and homocysteine levels. More importantly, the data showed that homocysteine levels varied widely in mice fed control diets as well. When comparing control diets with similar nutrient concentrations (AIN-based), there were significant differences in homocysteine levels caused by the strain (ANOVA, \(p < 0.05\)) and age of the mice at baseline (\(r = 0.47; p < 0.05\)). When comparing homocysteine levels and sex, female mice tended to have higher homocysteine levels than male mice (9.3 ± 5.9 µmol/l vs. 5.8 ± 4.5 µmol/l; \(p = 0.069\)). To conclude, diets low in vitamin B12, vitamin B6, or folate and rich in methionine are similarly effective in increasing homocysteine levels. AIN recommendations for control diets are adequate with respect to the amounts of homocysteine-modulating dietary parameters. In addition, the mouse strain and the age of mice can affect the homocysteine level.

Keywords: Age, Amino acids, Diet composition, Homocysteine, Mice, Sex, Strain, B vitamins

Introduction
Homocysteine is a sulfur-containing essential amino acid. Its accumulation is associated with several diseases, including cardiovascular diseases such as stroke, cancer, Alzheimer’s disease and Parkinson’s disease [1]. Homocysteine is a component of one-carbon metabolism that is involved in the provision of methyl groups for biological methylation reactions. The enzyme S-adenosylmethionine synthetase catalyzes the synthesis of S-adenosylmethionine (SAM) through the reaction of methionine and adenosine triphosphate. SAM, an important methyl donor for methylation reactions, is converted to S-adenosylhomocysteine (SAH) after dispensing the methyl group. The formation of homocysteine from SAH is catalyzed by adenosylhomocysteinase. Homocysteine can be converted to methionine through the vitamin B12-dependent enzyme methionine synthase [2]. The acquired methyl group for remethylation comes from 5-methyltetrahydrofolate or from betaine [3]. Folate is the precursor of tetrahydrofolate [4], which is converted...
through methyl-tetrahydrofolate reductase to 5-methyltetrahydrofolate. Betaine can be formed from its precursor choline [5]. Homocysteine can also be converted to cystathionine via transsulfuration through the vitamin B\textsubscript{6}-dependent enzyme cystathionine β-synthase [6].

The metabolic steps clearly show that several nutrients are involved in the one-carbon pathway and therefore can modulate homocysteine levels: methionine, vitamin B\textsubscript{12}, B\textsubscript{6}, folate and choline (Fig. 1). Thus, any excess in methionine intake or deficiencies in vitamin B\textsubscript{12}, B\textsubscript{6}, folate and choline can contribute to an increase in homocysteine levels [7].

Mice are often used as models of induced hyperhomocysteinemia and to study the impact of homocysteine on disease development. Thus, the current review evaluates different diets regarding their efficacy in increasing homocysteine levels in mice. We particularly focused on vitamin B\textsubscript{12}, vitamin B\textsubscript{6}, folate, the sulfur-containing amino acids methionine and cystine, and choline. In addition to the experimental diets, special focus was also placed on the control diets, which were used as reference. Additionally, we reviewed the impact of mouse strains, sex, age and feeding period on plasma/serum homocysteine levels. This review may be used as a reference for planning future nutrition studies on this topic.

**Methods**

A systematic literature search was conducted using the database PubMed and the search items (vitamin B\textsubscript{12} OR cobalamin OR vitamin B\textsubscript{6} OR pyridoxine OR B vitamins OR folic acid OR folate OR folates OR homocysteine OR hyperhomocysteinemia) AND (mice OR mouse OR murine) in the title of publications. Studies were included if they met the following criteria: (I) the study was written in English and published through July 2020, (II) wild-type mice were used as the model organism, and (III) plasma or serum homocysteine levels were measured. Studies were excluded when nutrients were administered via injections, gavage or drinking water or when any kind of surgery was performed. A total of 113 studies with 305 data sets (Additional file 1: Table S1) were eligible to be included in the evaluation of this review.

The following data were extracted from each study: mouse strain, sex, age and/or body weight at baseline, duration of feeding, dietary concentrations of vitamin B\textsubscript{12}, vitamin B\textsubscript{6}, folate, the added S-containing amino acids methionine and cystine, choline and plasma or serum homocysteine levels (in the following term "plasma" is used for plasma and serum concentrations). If diet composition was not shown in the publications but was based on commercial diets, we added the manufacturer’s information on nutrient contents. If diets were termed AIN-based, we used data on the composition of the AIN-93/G and AIN-93/M diets [8]. Otherwise, corresponding authors were asked for further information (which also included information regarding strain, sex or age of the mice as well as duration of dietary intervention). Correlations between plasma homocysteine levels (means and medians) and dietary compounds, age of the mice and duration of dietary intervention were analyzed using Pearson’s correlation testing since variables are normally distributed and Spearman correlation since variables are not normally distributed. Differences between plasma homocysteine levels and sex variables were analyzed using Student’s t test, and strain differences were analyzed with Levene’s test to assess homogeneity of variances and single-factor analysis of variance (ANOVA) followed by Hochberg’s GT2 post hoc test (SPSS 2020).

**Results**

**Dietary parameters**

In 56 out of 113 studies, the composition of the experimental diets was described in detail. Experimental diets had vitamin B\textsubscript{12} concentrations varying from 0 to 81.6 µg/kg diet, vitamin B\textsubscript{6} concentrations varying from 0 to 22 mg/kg diet, folate concentrations varying from 0 to 40 mg/kg diet, methionine + cystine concentrations varying from 0 to 24.3 g/kg diet, and choline concentrations varying from 0 to 3.5 g/kg diet (Fig. 2).

In addition to the experimental diets, we also evaluated the control diets that were used as a reference, especially with regard to their potential to minimize homocysteine levels. In 85 out of 113 studies, the composition of the control diets was described in detail. Studies have shown high variations in homocysteine-relevant nutrients in control diets. The concentration of vitamin B\textsubscript{12} varied from 10 to 100 µg/kg diet, that of vitamin B\textsubscript{6} from...
5.4 to 28 mg/kg diet, that of folate from 0.5 to 16 mg/kg diet, that of methionine and cysteine from 0 to 13 g/kg diet, and that of choline from 0 to 3.5 g/kg diet (Fig. 2). Compared to nutrient recommendations for mice [8], concentrations of vitamin B12, vitamin B6, folate and S-containing amino acids in the control diets used were often markedly higher (Fig. 2). However, AIN-based control diets were administered in only 14 out of 113 studies (Table 1 and Additional file 1: Table S1).

In mice fed the experimental or control diets, circulating homocysteine levels varied from 0.1 to 280 µmol/l. Analysis of the associations between components of the experimental and control diets and plasma homocysteine levels revealed negative correlations for vitamin B12 (rho = −0.125; p < 0.05), vitamin B6 (rho = −0.191; p < 0.01) and folate (rho = −0.395; p < 0.001; Fig. 3). A positive correlation was observed between dietary methionine and plasma homocysteine levels (methionine: rho = 0.146; p < 0.05; Fig. 3). No significant correlations were found for homocysteine levels and dietary cysteine (rho = −0.076; p > 0.05) or choline (rho = 0.044; p > 0.05). The duration of the analyzed feeding experiments varied between 3 and 17 weeks. However, there was no correlation between feeding duration and plasma homocysteine level (r = −0.05; p > 0.5).

**Strain, sex, and age of mice**

When comparing the circulating homocysteine levels in mice resulting from all analyzed control diets (including AIN-based control diets), we found varying homocysteine levels ranging from 0.1 to 24.1 µmol/l (Additional file 1: Table S1). Surprisingly, homocysteine levels in mice consuming strictly AIN-based control diets also varied in a wide range (from 0.1 to 22.5 µmol/l; Table 1), indicating that parameters other than nutrients influenced homocysteine levels.

In mice that received AIN-93-based control diets (Table 1), there were differences in homocysteine levels related to the strain (p < 0.05; Fig. 4) and age of the mice at baseline (r = 0.474; p < 0.05). When comparing homocysteine levels and sex, female mice tended to have higher homocysteine levels than male mice (9.3 ± 5.9 µmol/l vs. 5.8 ± 4.5 µmol/l; p = 0.069, Table 1).

**Discussion**

The current review shows that hyperhomocysteinemia can be induced by numerous different dietary interventions, such as a reduction in vitamin B6, vitamin B12 or folate concentration and an increase in methionine concentration. Study data showed that dietary cysteine and choline had no effects on plasma homocysteine levels in
mice. In addition, there was no correlation between the duration of feeding the experimental diets and plasma homocysteine levels. When diets were fed over varying periods, in most cases, there was no difference between homocysteine levels at different time points (Additional file 1: Table S1). One study found differences after 2 weeks of feeding the experimental diets, but no differences between 2 and up to 10 weeks. Thus, dietary interventions to increase homocysteine levels appear to be rapidly effective.

The type of control diet used in these studies showed great variations (Additional file 1: Table S1). The intake of AIN-93-based diets resulted in homocysteine levels similar to those of the other control diets. Hence, higher doses of vitamin B₁₂, vitamin B₆ and folate than recommended in the AIN-93 diet [8] do not seem to further decrease homocysteine levels. However, it should be mentioned that AIN-based control diets were only used in 14 out of 113 studies (Additional file 1: Table S1).

Plasma levels of homocysteine depend on the mouse strain because the growth rate of mice and thus the nutrient requirements depend on the genetic background [21]. Older mice have higher homocysteine levels than younger mice, which is in line with homocysteine data in humans [22, 23]. An age-related reduction in renal function is attributable to this effect [24]. In addition, females tend to have higher homocysteine levels than males. It is assumed that the renal activity of cystathionine β-synthase, which catalyzes an important step in the formation of cysteine from homocysteine, is regulated by testosterone [25] and thus is commonly higher in males than in females [26].

In addition, it must be kept in mind that the different methods used for quantification of homocysteine such

---

**Table 1** Plasma homocysteine levels in mice fed AIN-93-based control diets

| Mouse strain | Sex | Age at baseline | Duration (weeks) | Plasma Hcy (µmol/l) | n  | References |
|--------------|-----|----------------|------------------|---------------------|----|------------|
| 129/5v       | f + m | 3 wks          | 6                | 0.1 ± 0.1          | 15 | [9]        |
| 129/5v       | f + m | 3 wks          | 9                | 0.1 ± 0.1          | 15 | [9]        |
| 129/5v       | f + m | 3 wks          | 9–13             | 2.0 ± 0.6          | nda| [10]       |
| CD-1         | f + m | Adult          | 9                | 2.5 ± 0.1          | nda| [11]       |
| CS7BL/6      | m     | 6 wks          | 8                | 2.5 ± 0.7          | nda| [12]       |
| CS7BL/6      | m     | 6 wks          | 4                | 2.6 ± 0.8          | nda| [12]       |
| 129/5v       | f + m | 3 wks          | 9                | 2.8 ± 0.3          | nda| [13]       |
| Swiss        | m     | 3 wks          | 5                | 3.0 ± 2.2          | nda| [14]       |
| CS7BL/6      | m     | 6 wks          | 8                | 3.3 ± 0.8          | nda| [15]       |
| CS7BL/6      | f     | 8 wks          | 9                | 3.6 ± 0.7          | nda| [16]       |
| SAMP8        | m     | 13 wks         | 4                | 4.0 ± 0.1          | nda| [17]       |
| BALB/c       | f     | 17 wks         | 2                | 5.2 ± 0.2          | nda| [18]       |
| Swiss        | m     | 3 wks          | 10               | 5.2 ± 0.6          | nda| [19]       |
| CS7BL/6      | f     | 3 wks          | 5                | 5.4 ± 1.7          | nda| [20]       |
| CS7BL/6      | f     | 6–8 wks        | 7                | 5.5 ± 5.4          | nda| [21]       |
| SAMP8        | m     | 17 wks         | 26               | 6.5 ± 1.5          | nda| [22]       |
| SAMR1        | m     | 17 wks         | 26               | 6.5 ± 1.5          | nda| [23]       |
| Swiss        | f     | 3 wks          | 10               | 7.1 ± 0.7          | nda| [24]       |
| Swiss        | f     | 3 wks          | 27               | 8.1 ± 0.8          | nda| [25]       |
| Swiss        | m     | 3 wks          | 1                | 8.7 ± 0.9          | nda| [26]       |
| Swiss        | f     | Adult          | 3                | 9.2 ± 1.7          | nda| [27]       |
| Swiss        | f     | 3 wks          | 1                | 9.7 ± 0.6          | nda| [28]       |
| CS7BL/6      | f     | 7 wks          | 2                | 16.7 ± 1.5         | nda| [29]       |
| BALB/c       | m     | 17 wks         | 52               | 18 ± 1.5           | nda| [30]       |
| BALB/c       | f     | 17 wks         | 52               | 22.5 ± 1.5         | nda| [31]       |

f, female; m, male; n, number of included mice; nda, no data available; wks, weeks

* Mean ± standard deviation

b Mean ± standard error

c Median ± interquartile range

# Read from diagrams of data from mice that received AIN-93-based control diets containing 25 µg vitamin B₁₂, 7 mg vitamin B₆, 2 mg folate, 3 g methionine + cystine and 2.5 g choline per kg diet; only studies with complete data sets about mouse strain, sex, age at baseline, and the duration of feeding (in weeks) were included.
as chromatography, immunoassays or capillary electrophoresis could have influenced the results [27]. In our review, the high-performance liquid chromatography (HPLC) was the most frequently used method to quantify plasma homocysteine (in 72 out of 113 studies, Additional file 1: Table S1).

To conclude, vitamin B₁₂, vitamin B₆, folate, and methionine are similarly effective in reducing homocysteine levels. AIN recommendations for control diets are adequate with respect to the amounts of homocysteine-modulating dietary parameters. In addition to dietary parameters, the mouse strain and the age of mice can affect homocysteine levels.

Abbreviations
ANOVA: Analysis of variance; SAH: S-adenosylhomocysteine; SAM: S-adenosylmethionine.
Supplementary Information

The online version contains supplementary material available at https://doi.org/10.1186/s12986-021-00594-9.

Additional file 1. Supplemental Table: Diet composition and mouse data.

Acknowledgements
We thank Annette Schuhmacher (Sniff), Barbara Mickelson (Envigo), Helena Sprafke (Altromin Spezialfutter GmbH & Co. KG), Lisa Wagner (Dyets, Inc.) and Melanie Hoar (LabDiet) for their support regarding diet compositions.

Authors’ contributions
ChB collected, analyzed and interpreted the data and was a major contributor in writing the manuscript. All authors read and approved the final manuscript.

Funding
Open Access funding enabled and organized by Projekt DEAL.

Availability of data and materials
All data analysed during this study are included in this published article and its supplementary information file.

Declarations

Ethics approval and consent to participate
Not applicable.

Consent for publication
Not applicable.

Competing interests
The authors declare that they have no competing interests.

Received: 9 April 2021 Accepted: 14 June 2021
Published online: 30 June 2021

References
1. Kumar A, Palfrey HA, Pathak R, Kadowitz PJ, Gettys TW, Murthy SN. The metabolism and significance of homocysteine in nutrition and health. Nutr Metab. 2017;14(1):1–12.
2. Hoffer LJ. Homocysteine remethylation and trans-sulfuration. Metabolism. 2004;53(1):140–83.
3. Selhub J. Homocysteine metabolism. Annu Rev Nutr. 1999;19(1):217–46.
4. Scott JM. Folate and vitamin B12. Proc Nutr Soc. 1999;58(2):441–8.
5. Olthof MR, Verhoef P. Effects of betaine intake on plasma homocysteine concentrations and consequences for health. Curr Drug Metab. 2005;6(1):15–22.
6. Jhee KH, Kruger WD. The role of cystathionine β-synthase in homocysteine metabolism. Antioxid Redox Sign. 2005;7(5–6):813–22.
7. Langman LJ, Cole DEC. Homocysteine. Crit Rev Clin Lab Sci. 1999;36(4):365–406.
8. Reeves PG. Components of the AIN-93 diets as improvements in the AIN-76A diet. J Nutr. 1997;127(5):838–41.
9. Fusio A, Nicolai V, Cavallaro RA, Ricceri L, Castiello GM, Cavallaro O, Corcùtta P, et al. A vitamin B9 deprivation induces hyperhomocysteinemia and brain S-adenosylhomocysteine, depletes brain S-adenosylmethionine, and enhances PS1 and BACE expression and amyloid-β deposition in mice. Mol Cell Neurosci. 2008;37(4):731–46.
10. Fusio A, Nicolai V, Ricceri L, Cavallaro RA, Isopi E, Mangia F, et al. S-adenosylmethionine reduces the progress of the Alzheimer-like features induced by β-vitamin deficiency in mice. Neurobiol Aging. 2012;33(7):1482–e1.
11. Holstein JH, Herrmann M, Schmelenbach J, Obied R, Olku J, Klein M, et al. Deficiencies of folate and vitamin B12 do not affect fracture healing in mice. Bone. 2010;47(1):151–5.
12. Yao L, Wang C, Zhang X, Peng L, Liu W, Zhang X, et al. Hyperhomocysteinemia activates the aryl hydrocarbon receptor/CD36 pathway to promote hepatic steatosis in mice. Hepatology. 2016;64(1):92–105.
13. Persichilli S, Gervasoni J, Di Napoli A, Fusio A, Nicolai V, Giarda B, et al. Plasma thiol levels in Alzheimer’s disease mice under diet-induced hyperhomocysteinemia: effect of S-adenosylmethionine and superoxide-dismutase supplementation. J Alzheimer’s Dis. 2015;44(4):1323–31.
14. da Silva VC, Fernandes L, Haseyama EJ, Agamme ALDA, Shinozawa EMG, Muniz MTC, et al. Effect of vitamin B deprivation during pregnancy and lactation on homocysteine metabolism and related metabolites in brain and plasma of mice offspring. PLoS ONE. 2014;9(4):e92683.
15. Castaño-Moreno E, Castillo V, Peñailillo R, Llanos MN, Valenzuela R, Ronco AM. Fatty acid and lipid metabolism in liver of pregnant mice and their offspring is influenced by unbalanced folates/vitamin B12 diets. Prostaglandins Leukot Essent Fatty Acids. 2020;154:102057.
16. Yao L, Cao B, Cheng Q, Cai W, Ye C, Liang J, et al. Inhibition of soluble epoxide hydrolase ameliorates hyperhomocysteinemia-induced hepatic steatosis by enhancing β-oxidation of fatty acid in mice. Am J Physiol-Gastrointest Liver Physiol. 2019;316(4):G527–38.
17. Martinez-Vega R, Partearroyo T, Vallecillo N, Varela-Moreiras G, Pajaes MA, Varela-Nieto I. Long-term omega-3 fatty acid supplementation prevents expression changes in cochlear homocysteine metabolism and ameliorates progressive hearing loss in C57BL/6J mice. J Nutr Biochem. 2015;26(12):1424–33.
18. Lv X, Wang X, Wang Y, Zhou D, Li W, Wilson JX, et al. Folic acid delays age-related cognitive decline in senescence-accelerated mouse prone 8: alleviating telomere attrition as a potential mechanism. Aging (Albany NY). 2019;11(22):10356.
19. Schwahn BC, Wang XL, Mikhail LG, Wu Q, Cohn J, Jiang H, et al. Betaine supplementation improves the atherogenic risk factor profile in a transgenic mouse model of hyperhomocysteinemia. Atherosclerosis. 2007;195(2):e100-7.
20. Huang Y, He Y, Sun X, He Y, Li Y, Sun C. Maternal high folic acid supplement promotes glucose intolerance and insulin resistance in male mouse offspring fed a high-fat diet. Int J Mol Sci. 2014;15(4):6298–313.
21. Jacoby RO, Fox JG, Davisson M. Biology and diseases of mice. Lab Anim Sci. 1999;49(4):224–31.
22. Frick B, Schroersband CA, Neuhaus G, Leibhuber F, Fuchs D. Increasing production of homocysteine and neopterin and degradation of tryptophan with age. Aging (Albany NY). 2004;6(4):675–84.
23. Powers RW, Majors AK, Lykins DL, Sims CJ, Lain KY, Roberts JM. Plasma homocysteine and malondialdehyde are correlated in an age- and gender-specific manner. Metab Clin Exp. 2002;51(11):1433–8.
24. Norlund L, Grubb A, Fex G, Leksell H, Nilsson JE, Schenck H, et al. Plasma thiols levels in Alzheimer’s disease mice under diet supplementation improves the atherogenic risk factor profile in a transgenic mouse model of hyperhomocysteinemia. Atherosclerosis. 2007;195(2):e100-7.
25. Norlund L, Grubb A, Fex G, Leksell H, Nilsson JE, Schenck H, et al. The increase of plasma homocysteine concentrations with age is partly due to the deterioration of renal function as determined by plasma cystatin C. Clin Chem Lab Med. 1998;36(3):175–8.
26. Vitvitsky V, Prudovska A, Stabler S, Zhou Y, Wang H, Lenzt SR, Banerjee R. Testosterone regulation of renal cystathionine β-synthase: implications for sex-dependent differences in plasma homocysteine levels. Am J Physiol Renal Physiol. 2007;293(2):F594-600.
27. Vitvitsky V, Dayal S, Stabler S, Zhou Y, Wang H, Lenzt SR, et al. Perturbations in homocysteine-linked redox homeostasis in a murine model for hyperhomocysteinemia. A J Physiol Regul Integr Comp Physiol. 2004;287(1):R39-46.
28. Alam SF, Kumar S, Ganguly PM. Measurement of homocysteine: a historical perspective. J Clin Biochem Nutr. 2019;65(3):171–7.
29. Pickrell L, Li D, Brown K, Mikael LG, Wang XL, Wu Q, et al. Methylene-tetrahydrofolate reductase deficiency and low dietary folate increase embryonic delay and placental abnormalities in mice. Birth Defects Res A Clin Mol Teratol. 2009;85(6):531–41.
30. Pickrell L, Brown K, Li D, Wang XL, Deng L, Wu Q, et al. High intake of folic acid disrupts embryonic development in mice. Birth Defects Res A Clin Mol Teratol. 2011;91(1):8–19.
31. Chen Z, Karpakis AC, Ackerman SL, Pogrinyi IP, Melnyk S, Lussier-Cacan S, et al. Mice deficient in methylenetetrahydrofolate reductase exhibit hyperhomocysteinemia and decreased methylation capacity, with neuropathology and aortic lipid deposition. Hum Mol Genet. 2001;10(5):433–44.
31. Devlin AM, Arning E, Bottiglieri T, Faraci FM, Rozen R, Lentz SR. Effect of Mthfr genotype on diet-induced hyperhomocysteinemia and vascular function in mice. Blood. 2004;103(7):2624–9.

32. Liu WH, Zhao YS, Gao SY, Cao J, Zhang KQ, Zou CG. Hepatocyte... 2018;68:368.

33. Hsu CC, Cheng CH, Hsu CL, Li WL, Huang SC, Huang YC. Role of vitamin B6 status on antioxidant defenses, glutathione, and related enzyme activities in mice with homocysteine-induced oxidative stress. Food Nutr Res. 2015;59(1):25702.

34. Mikael LG, Deng L, Paul M, Leo L, Lynch CM, Faraci FM, et al. Paradoxical absence of a prothrombotic phenotype in a mouse model of severe hyperhomocysteinemia. Blood. 2012;119(13):3176–83.

35. Tan H, Jiang X, Yang F, Li Z, Liao D, Trial J, et al. Hyperhomocysteinemia inhibits post-injury reendothelialization in mice. Cardiovasc Res. 2006;69(1):253–62.

36. Christensen KE, Hou W, Bahous RH, Deng L, Malysheva OV, Arning E, et al. Moderate folic acid supplementation and MTHFD1-synthetase deficiency in mice, a model for the R653Q variant, result in embryonic defects and abnormal placental development. Am J Clin Nutr. 2016;104(5):1459–69.

37. Christensen KE, Bahous RH, Hou W, Deng L, Malysheva OV, Arning E, et al. Low dietary folate interacts with MTHFD1 synthetase deficiency in mice, a model for the R653Q variant, to increase incidence of developmental delays and defects. J Nutr. 2018;148(4):501–9.

38. Zhang X, Chen S, Li L, Wang Q, Le W. Folic acid protects motor neurons against the increased homocysteine, inflammation and apoptosis in SOD1G93A transgenic mice. Neuropharmacology. 2008;54(7):1112–9.

39. Champier J, Claustrat F, Nazaret N, Montange MF, Claustrat B. Folate depletion changes gene expression of fatty acid metabolism, DNA synthesis, and circadian cycle in male mice. Nutr Res. 2012;32(2):124–32.

40. Lam TY, Seto SW, Au ALS, Poon CCW, Li RWS, Lam HY, et al. Folic acid supplementation alters the DNA methylation profile and improves insulin resistance in high-fat-diet-fed mice. J Nutr Biochem. 2018;59:76–83.

41. Vitvitsky V, Dayal S, Stabler S, Zhou Y, Wang H, Lentz SR, et al. Perturbations in homocysteine-linked redox homeostasis in a murine model for hyperhomocysteinemia. Am J Physiol-Reg I. 2004;287(1):R39–46.

42. Kamath AF, Chauhan AK, dos Santos MBE, Loos JB, Vongpatanasin W. Elevated levels of homocysteine compromise blood-brain barrier integrity in mice. Blood. 2006;107(2):591–3.

43. Devlin AM, Singh R, Bottiglieri T, Innis SM, Green TJ. Hepatic acyl-coenzyme a: cholesterol acyltransferase-2 expression is decreased in mice with hyperhomocysteinemia. J Nutr. 2010;140(2):371–7.

44. Li W, Tang R, Ma F, Ouyang S, Liu Z, Wu J. Folic acid supplementation alters the DNA methylation profile and improves insulin resistance in mice treated with testosterone and estradiol. Am J Physiol Renal Physiol. 2015;308(12):F1431–43.

45. Teng YW, Cerdena I, Zeisel SH. Homocysteine in mice with genetic betaine homocysteine S-methyltransferase deficiency is independent of dietary-folate intake. J Nutr. 2012;142(11):1964–7.

46. Bernard DJ, Pangilinan FJ, Cheng J, Molloy AM, Brody LC. Mice lacking the transcobalamin-vitamin B12 receptor, CD320, suffer from anemia and reproductive deficits when fed vitamin B12-deficient diet. Hum Mol Genet. 2018;27(20):3627–40.

47. Zhou J, Wenstuck GH, Ihotak S, Shi YY, Tedesco V, Triggari B, et al. Hyperhomocysteinemia induced by betaine supplementation does not independently cause atherosclerosis in C57BL/6 mice. FASEB J. 2008;22(7):2569–78.

48. Hamelet J, Noll C, Ripoll C, Paul JL, Nelan D, Delabar J, et al. Increased plasma homocysteine concentrations in mice with vitamin B12 deficiency in the diet lead to atherosclerosis in male mice. J Nutr Biochem. 2011;22(10):2712–20.

49. Aléssio AC, Santos CX, Debbas V, Oliveira LC, Haddad R, Annichino-Bizzaccchi JM. Evaluation of mild hyperhomocysteinemia during the development of atherosclerosis in apolipoprotein E-deficient and normal mice. Exp Mol Med. 2019;51(3):362–40.

50. Keil RP, Abler LL, Altmann HM, Wang Z, Wang P, Ricke WA, et al. Impact of a folate-enriched diet on urinary tract function in mice treated with testosterone and estradiol. Am J Physiol Renal Physiol. 2015;308(12):F1431–43.

51. Yung LW, Cerdena I, Zeisel SH. Homocysteine in mice with genetic betaine homocysteine S-methyltransferase deficiency is independent of dietary-folate intake. J Nutr. 2012;142(11):1964–7.

52. Bernard DJ, Pangilinan FJ, Cheng J, Molloy AM, Brody LC. Mice lacking the transcobalamin-vitamin B12 receptor, CD320, suffer from anemia and reproductive deficits when fed vitamin B12-deficient diet. Hum Mol Genet. 2018;27(20):3627–40.

53. Zhou J, Wenstuck GH, Ihotak S, Shi YY, Tedesco V, Triggari B, et al. Hyperhomocysteinemia induced by betaine supplementation does not independently cause atherosclerosis in C57BL/6 mice. FASEB J. 2008;22(7):2569–78.

54. Hamelet J, Noll C, Ripoll C, Paul JL, Nelan D, Delabar J, et al. Increased plasma homocysteine concentrations in mice with vitamin B12 deficiency in the diet lead to atherosclerosis in male mice. J Nutr Biochem. 2011;22(10):2712–20.
72. Ai Y, Sun Z, Peng C, Liu L, Xiao X, Li J. Homocysteine induces hepatic steatosis involving ER stress response in high methionine diet-fed mice. Nutrients. 2017;9(4):346.

73. Sim IC, Yeh HQ, Choi HS, Choi YJ, Kwak HC, Kim SK, et al. L-serine supplementation attenuates alcoholic fatty liver by enhancing homocysteine metabolism in mice and rats. J Nutr. 2015;145(2):260–7.

74. Borowczyk K, Tisończyk J, Jakubowski H. Metabolism and neurotoxicity of homocysteine thiolactone in mice: protective role of bleomycin hydrolase. Amino Acids. 2012;43(3):1339–48.

75. Berge RK, Bjørdal B, Strand E, Bohoor P, Lindquist C, Nordrehaug JE, et al. Tetracyclithiopropionic acid induces hepatic mitochondrial dysfunction and steatosis, accompanied by increased plasma homocysteine in mice. Lipids Health Dis. 2016;15(1):1–16.

76. Jacovina AT, Deora AB, Ling Q, Broekman MJ, Almeida D, Greenberg CB, et al. Homocysteine inhibits neangiogenesis in mice through blockade of annexin A2–dependent fibrolysis. J Clin Investig. 2009;119(11):3368–94.

77. Zinno P, Motta V, Guantartico B, Natella F, Roselli M, Bello C, et al. Supplementation with dairy matrices impacts on homocysteine levels and gut microbiota composition of hyperhomocysteinemic mice. Eur J Nutr. 2020;59(1):345–58.

78. Shinohara M, Ji C, Kaplowsitz N. Differences in betaine–homocysteine methyltransferase expression, endoplasmic reticulum stress response, and liver injury between alcohol-fed mice and rats. Hepatology. 2005;41(3):796–805.

79. Teng YW, Mehendiz MG, Garrow TA, Zisel SH. Deletion of betaine-homocysteine S-methyltransferase in mice perturbs choline and 1-carbon metabolism, resulting in fatty liver and hepatocellular carcinomas. J Biol Chem. 2011;286(42):36258–67.

80. Jakubowski H, Perla-Kajan J, Finnell RH, Cabrera RM, Wang H, Gupta S, et al. Genetic or nutritional disorders in homocysteine or folate metabolism increase protein N-homocysteinylation in mice. FASEB J. 2009;23(6):1217–21.

81. Tran P, Hiou-Tim F, Frosst P, Lussier-Cacan S, Bagley P, Selhub J, et al. The methylation of Fads2 and altered hepatic fatty acid and phospholipid metabolism in mice with hyperhomocysteinemia. J Biol Chem. 2007;282(51):37082–90.

82. Sudduth TL, Weekman EM, Brothers HM, Braun K, Wilcock DM. β-amyloid deposition is shifted to the vasculature and memory impairment is exacerbated when hyperhomocysteinemia is induced in APP/PS1 transgenic mice. Alzheimer's Res Ther. 2014;6(1):1–10.

83. Chen M, Zhou C, Xu H, Zhang T, Wu B. Chronopharmacological targeting of Rev-erba by puerarin alleviates hyperhomocysteinemia in mice. Biomed Pharmacother. 2020. https://doi.org/10.1016/j.biopha.2020.109936.

84. Szulistöringrum DC, Singh R, Devlin AM. Epigenetic regulation of glucocorticoid receptor expression in aorta from mice with hyperhomocysteinemia. Epigenetics. 2012;7(5):514–21.

85. Wang J, Jiang Y, Yang A, Sun W, Ma C, Ma S, et al. Hyperhomocysteinemia-induced monocyte chemoattractant protein-1 promoter DNA methylation by nuclear factor-κB in mice. Biomedicine. 2013;2(2):118–27.

86. Song YS, Rosenfeld ME. Methionine-induced hyperhomocysteinemia promotes superoxide anion generation and NF-κB activation in peritoneal macrophages of C57BL/6 mice. J Med Food. 2004;7(2):229–34.

87. Xu F, Sudo Y, Sanechika S, Yamashita J, Shimaguchi S, Honda SI, et al. Disturbed biotinoid and folate metabolism in the Qdpr-deficient mouse. FEBS Lett. 2014;588(21):3924–31.

88. Denny KJ, Kelly CF, Kumar V, Witham KL, Cabrera RM, Fennell RH, et al. Autointoxication by homocysteine-sulfated proteins in a mouse model of folate-deficiency-induced neural tube defects. Birth Defects Res Part A Clin Mol Teratol. 2016;106(3):201–7.

89. Liu Z, Choi SW, Crott JW, Keyes MW, Jang H, Smith DE, et al. Methylprednisolone, methylprednisolone mono-hemihydrate, and methotrexate enhance the expression of mouse kidney proteins involved in renal disease. Mol Genet Metabol. 2014;113(3):200–6.

90. Song YS, Rosenfeld ME. Methionine-induced hyperhomocysteinemia suppresses superoxide anion generation and NF-κB activation in peritoneal macrophages of C57BL/6 mice. J Med Food. 2004;7(2):229–34.

91. Kopp M, Morisset R, Rychlik M. Characterization and interrelations of one-carbon metabolites in tissues, erythrocytes, and plasma in mice with dietary induced folate deficiency. Nutrients. 2017;9(5):462.

92. Jiang H, Hurt KJ, Breen K, Stabler SP, Allen RH, Orlicky DJ, et al. Folate deficiency or reduced intake of folate or choline in pregnant mice results in impaired short-term memory and increased apoptosis in the hippocampus of wild-type offspring. Neuroscience. 2015;300:1–9.

93. Shaevitz LR, Picker JD, Rana J, Koldrny NH, Share B, Berger-Sweeney JE, et al. Glutamate carboxypeptidase II and folate deficiencies result in reciprocal protection against cognitive and social deficits in mice: implications for neurodevelopmental disorders. Dev Neurobiol. 2012;72(8):891–905.
111. Challet E, Dumont S, Mehdi MK, Allemann C, Bousser T, Gourmelen S, et al. Aging-like circadian disturbances in folate-deficient mice. Neurobiol Aging. 2013;34(6):1589–98.

112. Miszewski SG, Trott JF, Berryhill GE, Tat L, Green R, Borowsky AD, et al. Folate Deficiency inhibits development of the mammary gland and its associated lymphatics in fvb mice. J Nutr. 2020;150(8):2120–30.

113. Gospe SM Jr, Getzen DW, Summers PJ, Lunetta JM, Miller JW, Selhub J, et al. Behavioral and neurochemical changes in folate-deficient mice. Physiol Behav. 1995;58(5):935–41.

114. Cavallaro RA, Fuso A, Nicolai V, Scarpa S. S-adenosylmethionine prevents oxidative stress and modulates glutathione metabolism in TgCRND8 mice fed a B-vitamin deficient diet. J Alzheimer's Dis. 2010;20(4):997–1002.

115. Ernest S, Hosack A, O’Brien WE, Rosenblatt DS, Nadeau JH. Homocysteine levels in A/J and C57BL/6J mice: genetic, diet, gender, and parental effects. Physiol Genomics. 2005;21(3):404–10.

116. Zhang J, Handy DE, Wang Y, Bouchard G, Selhub J, Loscalzo J, et al. Hyperhomocysteinemia from trimethylation of hepatic phosphatidyl-lethanolamine during cholesterol cholelithogenesis in inbred mice. Hepatology. 2011;54(2):697–706.

117. Pogribny IP, Kutanzi K, Melnyk S, de Conti A, Tryndyak V, Montgomery B, et al. Strain-dependent dysregulation of one-carbon metabolism in male mice is associated with choline- and folate-deficient diet-induced liver injury. FASEB J. 2013;27(6):2233–43.

118. Knock E, Deng L, Wu Q, Lawrance AK, Wang XL, Rozen R. Strain differences in mice highlight the role of DNA damage in neoplasia induced by low dietary folate. J Nutr. 2008;138(4):653–8.

119. Jacobs RL, Stead LM, Devlin C, Tabas I, Brosnan ME, Brosnan JT, et al. Physiological regulation of phospholipid methylation alters plasma homocysteine in mice. J Biol Chem. 2005;280(31):28299–305.

120. Choumenkovitch SF, Selhub J, Bagley PJ, Maeda N, Nadeau MR, Smith DE, et al. In the cystathionine β-synthase knockout mouse, elevations in total plasma homocysteine increase tissue S-adenosylhomocysteine, but responses of S-adenosylmethionine and DNA methylation are tissue specific. J Nutr. 2002;132(8):2157–60.

121. Santti-Baron D, Aupetit J, Janel N. Plasma homocysteine levels are not increased in murine models of Alzheimer’s disease. Neurosci Res. 2005;53(4):447–9.

122. Chassé JF, Santti-Baron D, Vayssettes C, Chabili A, Aupetit J, Maeda N, et al. Altered gene expression in liver from a murine model of hyperhomocysteinemia. J Biol Chem. 2003;278(34):31504–11.

123. Robert K, Maurn N, Ledru A, Delabar J, Janel N. Hyperkeratosis in cystathionine beta synthase-deficient mice: An animal model of hyperhomocysteinemia. Anat Rec Part A. 2004;280(2):1072–6.

124. Robert K, Santti-Baron D, Chassé JF, Paly E, Aupetit J, Kamoun P, et al. The neuronal SAPK/JNK pathway is altered in a murine model of hyperhomocysteinemia. J Neurochem. 2004;89(1):33–43.

125. Eberhardt RT, Forgione MA, Cap A, Leopold JA, Rudd MA, Trolliet M, et al. Endothelial dysfunction in a murine model of mild hyperhomocyst(e)inemia. J Clin Investig. 2000;106(4):483–91.

126. Hamelet J, Demuth K, Paul JL, Delabar JM, Janel N. Hyperhomocysteinemia due to cystathionine beta synthase deficiency induces dysregulation of genes involved in hepatic lipid homeostasis in mice. J Hepatol. 2007;46(1):151–9.

127. Powers RW, Gandley RE, Lykins DL, Roberts JM. Moderate hyperhomocysteinemia decreases endothelial-dependent vasorelaxation in pregnant but not nonpregnant mice. Hypertension. 2004;44(5):527–33.

128. Ernest S, Carter M, Shao H, Hosack A, Lerner N, Colmeneres C, et al. Parallel changes in metabolite and expression profiles in crooked-tail mutant and folate-reduced wild-type mice. Hum Mol Genet. 2006;15(23):3387–93.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.