INTRODUCTION

ncRNAs represent a newly recognized kind of transcripts that lacks an "open reading frame" (ORF). Yet, several researches have illustrated possible latent ORFs inside ncRNAs via genome-wide techniques, which opened arguments regarding the traditionally understood nature of ncRNAs. In the past decade, ncRNAs have been revealed to be pivotal regulators at different levels by diversified mechanisms, including translation, RNA splicing, DNA replication, gene regulation, genome defence, chromosome structure, bifunctional RNA and as a hormone.

MI RNAs, as a crucial kind of ncRNAs, are a type of small, non-coding, evolutionarily conserved, single-stranded, endogenous RNA molecule of approximately 22-25 nucleotides (nt) in length. They regulate gene expression at the post-transcriptional level via their "seed sequences," which hybridize to the 3'-untranslated region (3'-UTR), 5'-UTR and/or CDS (coding sequence) of target mRNAs and lead to degradation or translational inhibition of these target genes.

The latest progress on miR-374 and its functional implications in physiological and pathological processes

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Abstract
Non-coding RNAs (ncRNAs) have been emerging players in cell development, differentiation, proliferation and apoptosis. Based on their differences in length and structure, they are subdivided into several categories including long non-coding RNAs (lncRNAs >200nt), stable non-coding RNAs (60-300nt), microRNAs (miRs or miRNAs, 18-24nt), circular RNAs, piwi-interacting RNAs (26-31nt) and small interfering RNAs (about 21nt). Therein, miRNAs not only directly regulate gene expression through pairing of nucleotide bases between the miRNA sequence and a specific mRNA that leads to the translational repression or degradation of the target mRNA, but also indirectly affect the function of downstream genes through interactions with IncRNAs and circRNAs. The latest studies have highlighted their importance in physiological and pathological processes. MiR-374 family member are located at the X-chromosome inactivation center. In recent years, numerous researches have uncovered that miR-374 family members play an indispensable regulatory role, such as in reproductive disorders, cell growth and differentiation, calcium handling in the kidney, various cancers and epilepsy. In this review, we mainly focus on the role of miR-374 family members in multiple physiological and pathological processes. More specifically, we also summarize their promising potential as novel prognostic biomarkers and therapeutic targets from bench to bedside.

KEYWORDS
calcium handle, epilepsy, miR-374, ncRNA, tumorigenesis
mRNAs.5,6 Because miRNAs imperfectly complement their targets, they are able to interact with tens to hundreds of gene products in various signaling pathways to ultimately regulate cell functions. This also indicates the diversity of miRNAs functions.

MiR-374 family members (including A (a), B (b) and C (c)), as splicing segments of the IncRNA FTX, are located at the X-inactivation center (Xic) of chromosomes Xq13.2 in humans.6 An increasing number of studies have shown that these members participate in a great diversity of physiological and pathological processes. In this review, we mainly summarize the role of miR-374 family members, a highly conserved miRNA cluster in evolution, in different physiological and pathological processes. More importantly, we also discuss their potential function in the diagnosis and treatment of miR-374 related diseases, especially cancer.

2 | THE DELIVERY OF MIR-374 AND ITS ROLE IN DEVELOPMENT

In mammalian development, X-chromosome inactivation (XCI) is well-known for epigenetic regulation.5,7 Mounting studies have shown that female embryos will die without XCI, which corroborates the significance of this epigenetic regulatory mechanism in the development of female embryos.8,9 The transcriptional silencing of an X chromosome often corrects an imbalance of X-linked gene dosage. One cis-acting element, named Xic, controls and regulates XCI. There are several transcripts that are located in Xic and can escape XCI, including X-inactive-specific transcript (XIST), TSIX (TSIX transcript, XIST antisense RNA), JPX (JPX transcript, XIST activator), FTX (FTX transcript, XIST regulator) and miRNAs generated from the FTX sequence by cleavage;10,11 which may lead to gender disparity (Figure 1). Overwhelming data have implicated that XIST, the master regulator of X-inactivation initiation, is a single and central cis-acting regulator that coordinates imprinted XCI.12,13

In addition to XIST, FTX is also one of the most abundant transcripts at the stage of preimplantation embryo and is thought to be a positive regulator of XIST.14,15 It is a well-conserved IncRNA in evolution and includes several conserved miRNAs, such as miR-374, -545 and -421, in its introns.6 In humans, intron 1 encodes a cluster of 2 miRNAs (MIR-374A and MIR-545), which is absent in rats and mice; intron 5 encodes a cluster of 3 miRNAs (MIR-374B, C and MIR-421) (Figure 1B). In addition, the sequences of miR-374 family members are highly conserved in different mammalian species, especially in miR-374-5p (Figure 2).

In recent years, studies have shown that these miRNAs, which are located at Xic, are also imprinted and involved in mammalian development and the generation of paternal sperm. For example, Kobayashi et al.10 reported that miR-374-5p and miR-421-3p mapped adjacent to Xist, were both predominantly expressed in female blastocysts from the Xp chromosome in F1 blastocysts and were indeed imprinted, as determined by allelic expression analysis. In addition, there is another report17 that miR-374b is markedly decreased in the seminal plasma of azoospermia but is increased in the seminal plasma of asthenozoospermia, as shown by Solexa sequencing and RT-qPCR analysis. The area under the receiver operating characteristic (ROC) curve of miR-374b is 0.839 in the 73 azoospermia cases and 0.813 in the 79 asthenozoospermia cases. The authors point out this is markedly higher than for routine biochemical parameters (0.510-0.622). Their results revealed that miR-374b level in seminal plasma is a novel, non-invasive approach for diagnosing male infertility.

3 | THE ROLE OF MIR-374 IN TUMORIGENESIS

Most likely because of the advancement of preventive examinations and therapeutic interventions, epidemiological studies have provided strong support for the decrease of the mortality rate due to cancer. Nevertheless, the heterogeneous complex of cancers still remains one of the leading causes of premature death worldwide.17,18 Recent reports have verified the presence of ncRNAs, including IncRNA, miRNAs, circular RNAs and so on, in the human cancer.19,20 We will summarize the role of miR-374 family members in the development and progression of different systematic cancers in this section.

3.1 | miR-374 and digestive system carcinoma

The digestive system, as one of the eight systems of the human body, is composed of two parts: the digestive tract and digestive glands. The liver, as one of the important digestive glands, plays an indispensable role in body metabolism, including the storage of glycogen, the synthesis of plasma protein, the decomposition of red blood cells, and the detoxification of toxic substances. Meanwhile, because the liver is a fragile organ, poor protection will lead to the decline of normal function and the development of disease (such as hepatitis and hepatocellular carcinoma). In 2017, Bao et al.21 reported that miR-374 levels were significantly decreased as HBV-related liver fibrosis progressed from S0-S2 to S3-S4. In addition, they identified that miR-374 had a highly diagnostic accuracy in discriminating S0-S2 from S3-S4 using multivariate logistic regression analysis. In recent years, our research group has been unanimously committed to the study of liver disease. In terms of miRNAs, we have previously revealed that the miR-545/374a cluster is up-regulated in HBV-HCC tissue and is significantly correlated with prognosis-related clinical features, including histological grade, metastasis and tumor capsule via the observation of 66 pairs of HBV-related HCC tissue and matched non-cancerous liver tissue specimens.8

Besides observing the role of miR-374 in liver, some have also investigated its effect on the digestive tract. For example, Wu et al.22 evaluated the potential value of miR-374b as a biomarker in colorectal cancer and found that miR-374b expression was significantly decreased for CRC patients with stage II and stage III disease and may be a novel biomarker for CRC. In addition, Qu et al.23 more comprehensively analysed the function of miR-374b from colon
First, they showed that the expression of miR-374b was significantly reduced in colon cancer tissues and cell lines. Second, they investigated the effect and mechanism of miR-374b and found that its overexpression inhibited cell proliferation and invasion, while the role of its knockdown was exactly opposite. Ultimately, they verified that LRH-1 is a direct target of miR-374b via a dual-luciferase reporter assay, and then they showed that miR-374b can suppresses the Wnt signaling pathway through LRH-1 in colon cancer cells.

In recent years, there have also been some reports about the relationship between miR-374 and gastric cancer. Xie J showed that the expression of miR-374b-5p was up-regulated and conducive to gastric cancer cell invasion and metastasis via inhibiting the expression of RECK. Ji et al. presented that the up-regulation of miR-374 mediated the malignant transformation of gastric cancer-associated mesenchymal stem cells and represented a novel avenue for gastric cancer therapy with an experimental rat model. In the same year, Sierzega et al. found that miR-374 was abnormally expressed in the...
Primary tissue of gastric cancer, but its expression was not changed in the serum of GC patients.26

Taken together, miR-374 expression was increased in HCC and GC. Additionally, its up-regulation promoted the malignant transformation of tumor cells. On the contrary, its expression was decreased in CRC. Meanwhile, its down-regulation repressed colon cancer cell proliferation and invasion. So the role of miR-374 is consistent in HCC, CRC and GC. Therefore, miR-374 can be as a novel biomaker in light of its function on the malignant features of digestive system carcinoma, including proliferation, apoptosis, invasion and metastasis.

3.2 | miR-374 and other system carcinoma

At present, there are quite sporadic reports of the effect of miR-374 on tumourgenesis in other systems. For example, miR-374b is diminished in prostate cancer tissue, and it can be identified as an independent predictor of biochemical recurrence-free survival by analysing the correlations between its expression and clinical-pathological features in Chinese patients.27 In 2015, Merhautova et al28 observed the relationship between miR-374-5p and metastatic renal cell carcinoma treated with or without sunitinib by TaqMan low-density arrays. However, they did not find any connection between either of them. Liao YY have reported that chemokine (C-C motif) ligand 3 (CCL3) activated MAPK (JNK, ERK, and p38) signaling pathways to reduce miR-374b expression and promoted VEGF-A expression and angiogenesis in human osteosarcoma cells.29 In 2009, Miko et al found that miR-374 expression was up-regulated in primary small cell lung cancer, but they didn't analyze the correlations between its expression and clinical parameters.30 The latest report31 confirmed that miR-374 may also participate in the pathogenesis of pituitary gonadotroph adenomas by bioinformatics analysis. However, the author did not discuss the specific mechanism of miR-374 in this disease.

Regarding skin cancer, Ning et al32 revealed that miR-374c expression was down-regulated by next-generation sequencing and RT-qPCR assay in Merkel cell carcinoma and other cutaneous tumors compared with normal skin. In 2017, Li et al33 observed that miR-374a expression was decreased in skin tissues of human squamous cell carcinoma compared to the normal skin tissue. Meanwhile, they verified through transfection of miR-374 mimics into A431 and SCL-1 cells, that miR-374a down-regulated the P53 signaling pathway to induce cell apoptosis and inhibit proliferation, migration and invasion by targeting its downstream protein, Gadd45a.

In addition to confirming that miR-374 influence the development of tumors, there were some reports that proto-oncogene mutations themselves also affect the expression of miRNAs, including miR-374. In 2015, Garcia-Cruz et al34 described how p19 affected miRNA by miRNA microarray assays and demonstrated that a p19G12S mutant up-regulated the expression of miR-374, miR-126, miR-342, miR-330, miR-335 and let-7 in Costello syndrome cell model. Their data suggested the oncogene mutant converted itself into activating status and led to the transduction of the downstream signaling pathways and this may have a sufficiently elementary impact on miRNAs expression to promote the development of numerous cancers.

Taken together, studies on miR-374 function in cancer have been relatively rare in other system than the digestive system. Only in human osteosarcoma and squamous cell carcinoma of skin tissues, there were detailed reports about miR-374 activation, its role in several signaling pathways and its downstream targets. The rest of the research has mainly focused on analysing the dependency of the expression of miR-374 and different clinical-pathological features.

3.3 | miR-374 and chemoradiotherapy of tumor

Presently, cancer treatment mainly includes three types: surgical resection, radiotherapy and chemotherapy. In this section, we mainly discussed miR-374 and chemoradiotherapy of tumors. In 2016, Schreiber et al35 evaluated the potential role of miRNAs in a cisplatin-resistant pancreatic cancer cell line (BxPc-3-R). They found that 34 miRNAs were up-regulated and 23 miRNAs were down-regulated, and then they identified that the down-regulated miR-374b was possibly and directly involved in the acquisition of drug-resistant phenotypes in pancreatic cancer cells with a hidden
Markov model algorithm. Meanwhile, miR-374b overexpression in BxPC3-R cells recovered cisplatin sensitivity almost to the levels displaying in BxPC3 parental cells. In the same year, Baek et al. first screened the change of miRNAs with microarray analysis in mouse squamous cell carcinoma line NR-S1, X60 cells (established by irradiating NR-S1 cells with 10 Gy of X-ray radiation once every 2 weeks) and C30 cells (established by irradiating NR-S1 cells with 5 Gy of carbon ion beam radiation once every 2 weeks). They also demonstrated that miR-374c-5p and miR-196a-5p were down-regulated.

When miR-374c-5p was selectively ectopically overexpressed in the human pancreatic cancer cell lines PANC1 and MIA-PaCa-2, these cells were sensitized to carbon ion beam radiation, with no change to gamma-ray sensitivity. Later, Gong et al. demonstrated that the p53/miRNA-374b/AKT1 signaling pathway may regulate BLM-induced cell apoptosis of colorectal cancer and ultimately facilitate an improvement in the outcome of chemotheraphy in colorectal cancer (CRC). In 2018, Sun et al. verified that the expression of miR-374b-5p was significantly reduced in pancreatic cancer tissues and the decreased expression was closely associated with poor progression in patients with pancreatic cancer. Meanwhile, they used multiple human pancreatic cancer cell lines and revealed that miR-374b-5p up-regulation relieved the chemoresistance of pancreatic cancer cells to gemcitabine by targeting several antiapoptotic genes, such as BCL-2, BIRC3 and XIAP.

Therefore, miR-374 may be a novel chemosensitizer and/or radiosensitizer and will be a new potential biomarker for deciding the optimal treatment for cancer.

4 | THE ROLE OF MIR-374 IN CELL GROWTH AND DIFFERENTIATION UNDER PHYSIOLOGICAL CONDITIONS

Cell division, growth and differentiation are common growth processes in organisms. Under physiological conditions, cell division, growth and differentiation are strictly and finely regulated. However, the exact mechanisms of this process are not yet clear.

In recent years, several studies have reported that miRNA-374 contributes to differentiation and proliferation of different cells in multiple organisms. For example, Dmitriev et al. performed the miRNA expression profile with cultures of CD56+ primary myoblasts and myotubes isolated from healthy individuals by an affinity purification procedure. They have demonstrated that a total of 60 miRNAs (including miR-374) were differentially expressed during serum starvation-induced myogenic differentiation. However, they did not explore the targets of miR-374. In 2015, Ma et al. further studied the role of miR-374 and found that miR-374b specifically bound to the 3'-UTR of MRF4 to down-regulate its expression at both the mRNA and protein level, leading to the negatively regulation of the differentiation of C2C12 myoblasts. In addition, Jee et al. found that the expression of miR-374-5p was higher in the proliferative zone (PZ) than the hypertrophic zones. They also identified that primary chondrocytes treated with a PTH/PTHrP receptor agonist, PTH1-34, induced the expression of miR-374-5p. The inhibition of miR-374-5p expression decreased chondrocyte proliferation and stimulated hypertrophic differentiation. Meanwhile, Rasheed et al. detected the expression level of miR-374 in the retina and demonstrated that its expression was up-regulated from the E12 to the PN1 stage and was later down-regulated. Nevertheless, this expression pattern was not an inverse with Brn3b during retinal ganglion cell (RGC) development. Subsequently, they confirmed that miR-374 by itself cannot affect Brn3b expression, but it can work with miR-23a to synergistically regulate the expression of Brn3b, thereby affecting RGC development.

Accumulated data have shown that miR-374 regulates cell growth and differentiation not only in rodents but also in poultry. In 2013, Pan et al. showed that dexamethasone-induced miR-374a and miR-374b promoted the differentiation of primary porcine adipocytes by targeting 3'-UTR of C/EBP-β. In the same year, Su et al. found that miR-374 contributed to goat hair production both in entering growth and cessation stages by the analysis of comparative genomics combined with an expression profile assay.

5 | THE ROLE OF MIR-374 IN KIDNEY DISEASE

Under physiological ranges, extracellular Ca\(^{2+}\) regulation is principally maintained by the kidney, as well as the skeleton. A series of studies have demonstrated that Ca\(^{2+}\) sensing receptor (CaSR) and claudin (CLDN) are pivotal regulators in renal Ca\(^{2+}\) balance. Among them, CaSR monitors circulating Ca\(^{2+}\) concentrations by adjusting excretion rates in the kidney. Moreover, CaSR influences Ca\(^{2+}\) transport via alterations of the transepithelial potential and paracellular channel permeability. The family of CLDNs, as four transmembrane proteins, consist of 27 members, which form paracellularly heteromeric or homomeric channels to allow selective permeation of cations (including Ca\(^{2+}\) and Mg\(^{2+}\)) through the epithelial tight junction. Early in 2009, Hou et al. showed that CLDN16 and CLDN19 are specifically expressed in the thick ascending limb (TAL) of nephrons, where a main percentage of filtered diverant cations (including calcium ions and magnesium ions) are extracellularly reabsorbed (30%-35% Ca\(^{2+}\) and 50%-60% Mg\(^{2+}\)). Then in 2012, a study using biochemical analysis and electrophysiological recordings found that CLDN14 and CLDN16 interacted to involve in renal Ca\(^{2+}\) reabsorption. CLDN14 overexpression in kidney epithelial cells impaired paracellular positive ions permeability through the CLDN16/19 heteromeric channel. Given the importance of miRNAs, Gong et al. demonstrated that CaSR activation by extracellular Ca\(^{2+}\) induced the expression of miR-374, as a novel microRNA, in TAL cells. Then, the up-regulated miR-374 dampened the transcript stability and translation of CLDN14 in a synergistic manner.

Polymeric IgA1 deposited in the mesangial of kidney leads to IgA nephropathy (IgAN). This is one of the most common cause of glomerulonephritis all around the world. Studies showed that IgAN was associated with an increase in B cells number and the
incompletely galactosylation of O-glycans in IgA1. In 2015, Hu et al. demonstrated that miR-374b expression was higher in B cells compared with controls in IgAN patients. And miR-374 can target PTEN and Cosmc to increase cell proliferation and the abnormal glycosylation of IgA1.

6 | THE ROLE OF MIR-374 IN NERVOUS SYSTEM DISEASE

6.1 | miR-374 and epilepsy

Epilepsy is one of the common diseases of the nervous system, and its prevalence is second only to stroke. Accumulated data have shown that miRNAs are involved in various neurological diseases, such as Alzheimer’s disease (AD), ischemic tolerance and Parkinson’s disease. But, less is known about miRNAs on epilepsy. In 2014, Moon et al. found that the expression of miR-374 was significantly decreased in the MDR group versus the control group in a model of mouse pilocarpine-induced epilepsy. In 2015, Liu et al. induced a rat TLE model with pentylenetetrazol. They also analysed the dysregulated miRNAs in the hippocampus by microRNA expression profiles and found that there were four up-regulated miRNAs, including miR-374.

6.2 | miR-374 and neurodegeneration

Degenerative changes (including drug-induced and physiological) in the nervous system seriously threaten people’s health. In recent years, research on the degenerative changes of the nervous system by miRNAs has grown. Wang et al. found that PQ- and MPTP-treatment inhibited the expression of miR-374-5p (P < 0.01) in Neuro-2a cells. Yet, miR-374-5p was not associated with several biological processes including regulation of DNA dependent transcription and RNA metabolic processes in the pathogenesis of Parkinson’s disease. Moreover, Manzini reported that miR-374 levels were significantly diminished in AD compared with the control group. Furthermore, miR-374 may directly target relevant AD genes such as BACE1 to regulate the progress of AD.

Amyotrophic lateral sclerosis (ALS) is one of the most common adult onset neurodegenerative diseases with a prevalence of 6-8 per 100 000. It is a complex disease with multiple pathogenic mechanisms (including excitotoxicity, oxidative stress, protein aggregation, mitochondrial dysfunction, dysregulated endosomal trafficking, defective axonal transport, dysregulation of RNA processing, and neuroinflammation). In 2017, Waller et al. reported that miR-374b-5p was significantly decreased in patient serum over time compared with 23 sALS and 22 control subjects. And it may be a compensatory role in the degeneration of muscle in ALS and be an attempt to support muscle regeneration and restore a balance by enhancing myoblast differentiation, and thus it could be used as a biomarker to assess treatment efficacy and potentially disease prognosis.

6.3 | miR-374 and other diseases in the nervous system

Hypoxic-ischemic encephalopathy (HIE) is a disease defined as ischemic injury caused by hypoxic asphyxia during the perinatal period. Late diagnosis partially leads to high mortality (approximately 15%-20%) of this disease. Therefore, finding new biomarkers to improve the diagnostic value of neuron-specific enolase (NSE) and S100B protein is especially important. In 2017, Wang et al. reported that the expressions of miR-210 and miR-374a, considered to be two important hypoxia-associated miRNAs, were down-regulated in blood samples of HIE newborns compared with those of healthy newborns. Joint analysis of miR-210, miR-374a, S100B protein and NSE help to elevate the diagnostic value and prognostic prediction for HIE by the ROC curves assay.

7 | THE ROLE OF MIR-374 IN CARDIOVASCULAR DISEASE

Studies showed that miR-374 family members can also regulate the pathophysiological process of cardiovascular disease. Early in 2013, Ward et al. analysed miRNA profiles in different blood subcomponents, such as platelets, PBMCs and plasma, via a high throughput RT-qPCR system in patients with STEMI or NSTEMI. And they found that miR-374b-5p in PBMCs was obviously lower in patients with STEMI than in patients with NSTEMI. Reversely, miR-374b-5p in plasma was markedly higher in patients with STEMI compared with NSTEMI. These results suggest the possible involvement of miR-374b-5p in ACS subtypes. Under normal circumstances, after patients with ACS are promptly treated, the myocardial cells undergo an ischemia and reperfusion process. Studies of myocardial ischemia-reperfusion have shown that it does not only brings benefits, but also causes myocardial injury. In 2018, Zhang et al. investigated the effects of miR-374 on myocardial I/R injury in rat models. Their results demonstrated that miR-374 relative expression was evidently lowered after reperfusion in the I/R and sevoflurane plus I/R groups compared with the sham group in the myocardium of rats. Compared with the I/R group, miR-374 relative expression was significantly increased in the sevoflurane plus I/R group in the myocardium of rats. Finally, they found that miR-374 could alleviate rat myocardial I/R injury by targeting SP1 through activating the PI3K/Akt signal pathway after pretreatment with sevoflurane.

Selenium deficiency has been identified as a causative factor in different kinds of heart failure. Researchers used microarray hybridization to show that there are five up-regulated (>5-fold) miRNAs, which were miR-374, miR-16, miR-199a-5p, miR-195 and miR-30e*, and three down-regulated miRNAs, which were miR-3571, miR-675 and miR-450a*, in rats with selenium deficiency by. And they verified that miR-374 expression was the highest among these up-regulated miRNAs. In the end, they explored that the Wnt/β-catenin signalling pathway was possibly associated with cardiac dysfunction caused...
by selenium deficiency. However, they did not confirm the targeting relationship between miR-374 and Wnt/β-catenin.

VEGF is a pivotal cytokine that promotes the formation of new blood vessels. However, the VEGF/VEGFR1 signalling pathway represses cardiac hypertrophy. In contrast, the VEGF/VEGFR2 signalling pathway accelerates cardiac hypertrophy. Lee et al.70 found that miR-374 inhibited the VEGFR1 signalling pathway and activated GPCR signalling pathway by targeting the 3’-UTR of VEGFR1 and cGMP-dependent protein kinase-1 to mediated pro-hypertrophic processes.

In 2016, Licholai et al.71 found that miR-374-5p can maintain vascular integrity by contrasting the different profile of microRNA expression in aneurysmal and unaffected ascending aortic tissue acquired from the same patient. In 2012, Milenkovic72 observed that miR-374* expression increased after mutagenesis in apoE mice compared to wild-type mice; however, when supplemented with polyphenols in these mice, its expression decreased in apoE. And then they analysed that miR-374* expression presented negative correlations with AKT1.

8 | THE ROLE OF MIR-374 IN IMMUNE-RELATED DISEASE

CD56 is invariably expressed in normal natural killer cell,73 a subset of normal T cells and occasionally in T cell acute lymphoblastic leukemia (T-ALL).74 Studies have implied that CD56 is associated with a poor prognosis in lymphoid tumors, including T-ALL.74 Therefore, in 2013, Gimenes-Teixeira et al.75 showed that miR-374 and miR-221 were higher in T-ALL/CD56+ than in T-ALL/CD56- cells, with 181- and 271-fold relative expression, respectively. Without regard to the expression of CD56, the expression of miR-374 was at obviously higher levels in leukemic blasts compared with normal peripheral blood thy-mocytes and T cells.

However, in 2015, Qian et al.76 further investigated the role and mechanism of how miR-374 affects T-cell lymphoblastic lymphoma. They showed that miR-374b was markedly down-regulated in the T-ALL tissues by microRNA microarray analysis, and the down-regulated miR-374b was greatly associated with worse survival and higher relapse rates in patients with T-ALL. miR-374b overexpression restrained tumorigenicity and cell proliferation, and it accelerated cell apoptosis though targeting Wnt-16 and AKT1, which led to inhibition of AKT signal pathway.

Acute graft-versus-host disease (aGvHD) has a higher mortality rate, the most frequent and serious complication. Expression analysis77 identified miR-374-5p as significantly down-regulated and with diagnostic value by ROC analysis in aGvHD.

Moreover, Delić et al.78 verified that, after female C57BL/6 mice were infected with self-healing Plasmodium chabaudi malaria, hepatic miR-374* expression was down-regulated. In addition, Uribe et al.79 found that miR-374a-5p expression was up-regulated in porcine intestinal mucosa infected with Salmonella Typhimurium by Microarray hybridization and analysis and RT-qPCR assay.

9 | THE ROLE OF MIR-374 IN OTHER DISEASES

Bhargava et al.80 found that miR-374 affected the function of rat AT2 epithelial cells during hyperoxic stress and recovery through three possible targets, including actinin alpha 4, actinin alpha 1, and Na, K-ATPase.

In addition, some have directly observed the relationship between miR-374 and other critical molecules in different cell lines. For example, Tasharrofi et al.81 wanted to investigate whether miR-374a can inhibit Fas-induced apoptosis in human primary retinal pigment epithelial (RPE) cells by targeting Fas during oxidative conditions. Their results confirmed that miR-374a indeed prevented Fas up-regulation by binding with its 3’-UTR to enhance RPE cells survival and protect the cells against oxidative stress. Unterbruner et al.82 found that miR-374a-5p regulate not only the expression of ubiquitin ligase MID1 by binding to the 3’-UTR of the MID1 mRNA but also the mTOR signaling pathway. Therefore, given that dysregulation of MID1 expression is closely associated with multiple diseases including cancer, midline malformation syndromes and neurodegenerative diseases, miR-374a-5p could serve as a potential drug target for future therapy development.

10 | CONCLUSIONS AND PERSPECTIVES

In conclusion, as shown in Table 1, although there are some studies on the impact of miR-374 family members on various diseases, especially cancer, their expression in different pathophysologies are not the same. In addition, investigations on miR-374 family members are relatively superficial, both physiologically and pathologically, and mainly include the detection of miR-374 expression performed by microarrays or RT-qPCR assays and the analysis of correlation between miR-374 expression and cell apoptosis, invasion, metastasis and relapse, etc. There are relatively few studies on their targets. To date, the targets of miR-374 family members chiefly include: AKT, VEGF, PTEN, Wnt and Fas signalling pathways. Therefore, the mechanism of miR-374 in different cells or disease models needs further exploration and verification.

With regard to the sundry possibilities for the diagnosis and treatment of the miR-374 family members in diseases, we summarize as follows: (a) the diagnostic role of miR-374 family members. In view of the relationship between the expression of these members and multiple diseases, we can measure their expressions, as a novel biomarker, in serum and/or tissue from patients to assess the likelihood of illness and the prognosis of disease, especially in cancer. (b) The therapeutic role of miR-374 family members. In this aspect, we can overexpress or knockdown these members themselves by a variety of methods, such as mimics or inhibitor, ago- or antago-miRNAs, over-expressed or interfering vectors, transgenic or knocking gene and so on. In addition, we can also affect their roles by regulating their targets. However, we found that there were only a small number of papers revealed the targets of miR-374 family members under...
| Disease                  | Family members | Species     | Tissue and/or cell | Expression | Targets or pathway | Relationship with disease or clinical significance                                                                 | Time | Ref. |
|-------------------------|----------------|-------------|--------------------|------------|-------------------|-----------------------------------------------------------------------------------------------------------------------|------|------|
| Imprinted gene cluster  | miR-374-5p     | Mouse       | Blastocysts        | Up         | /                 | miR-374-5p were imprinted                                                                                        | 2013 | 10   |
| Male infertility        | miR-374b       | Human       | Seminal plasma     | Down/Up    | /                 | Azoospermia, Asthenozoospermia                                                                                       | 2011 | 16   |
| HBV-related liver fibrosis | miR-374       | Human       | Serum              | Down       | /                 | As a noninvasive diagnostic biomarker                                                                               | 2017 | 21   |
| HBV-related hepatocellular carcinoma | miR-374a     | Human       | HBV-HCC tissue and HCC cell lines Bel-7402, HepG2, HepG2215 | Up         | /                 | Correlated with histological grade, metastasis and capsule of HCC                                                   | 2015 | 8    |
| Colorectal cancer       | miR-374b       | Human       | Colon cancer tissues | Down       | /                 | As a biomarker of CRC                                                                                               | 2015 | 22   |
|                         | miR-374b       | Human       | Colon cancer tissues and cell lines HT29, HCT116, SW480 and SW620 | Down       | LRH-1             | Inhibited colon cancer cell proliferation and invasion                                                               | 2018 | 23   |
| Gastric cancer          | miR-374b-5p    | Human       | Gastric carcinoma cell line MGC-803, SGC-7901 and the normal human gastric epithelial cell line GES-1 | Up         | RECK              | Promoted gastric cancer cell invasion and metastasis                                                                     | 2014 | 24   |
|                         | miR-374        | Rat         | Wistar rats and primary MSCs | Up         | /                 | Malignant transformation of gastric cancer associated mesenchymal stem cells (MSC)                                  | 2017 | 25   |
|                         | miR-374a-5p    | Human       | Blood and tissue samples | Up         | /                 | Evaluation of serum microRNA biomarkers for gastric cancer                                                             | 2017 | 26   |
| Prostate cancer         | miR-374b       | Human       | Prostate cancer tissue | Down       | /                 | Correlation with clinical features of prostate patients                                                                 | 2013 | 27   |
| Renal cell carcinoma    | miR-374-5p     | Human       | Tissue samples with or without sunitinib | /          | /                 | Didn't found any connection                                                                                            | 2015 | 28   |
| Osteosarcoma            | miR-374b       | Human, mouse| Tumor tissue and osteosarcoma cell lines MG-63, U-2 OS and endothelial progenitor cell (EPC) | Down       | CCL3/MAPK/miR-374b/VEGF-A | CCL3 promoted angiogenesis by regulating miR-374b/VEGF-A axis                                                          | 2016 | 29   |
| Small cell lung cancer  | miR-374        | Human       | Tissue samples and cell lines HTB-172, HTB-184, HTB-119 | Up         | /                 |                                                                                                                      | 2009 | 30   |
| Pituitary gonadotroph adenomas | miR-374    | Rat         | Pituitary tissue    | Up         | /                 | MiR-374, -153, -145 and -33 may have regulated the DEGs.                                                             | 2018 | 31   |
| Skin cancer             | miR-374c       | Human       | Tissue samples and MCC cell line MS-1 | Down       | /                 | Induced cell apoptosis and inhibited proliferation, migration and invasion                                            | 2014 | 32   |
|                         | miR-374a       | Human       | Skin SCC samples and normal skin cells and SCC skin cell line A431 and SCL-1 | Down       | Gadd45a (downstream protein of P53 signaling pathway) | /                                                                                                                      | 2017 | 33   |
| Disease                                                                 | Family members | Species         | Tissue and/or cell                             | Expression | Targets or pathway | Relationship with disease or clinical significance                                                                 | Time   | Ref. |
|------------------------------------------------------------------------|----------------|-----------------|-----------------------------------------------|------------|-------------------|-----------------------------------------------------------------------------------------------------------------------|--------|------|
| Mutant of p19 and p21 H-Ras proteins                                   | miR-374        | Human, mouse    | HeLa cells and murine embryonic fibroblasts (MEFs) | Up         | /                 | /                                                                                                                      | 2015   | 34   |
| Cisplatin resistant                                                    | miR-374b       | Human           | Cisplatin-resistant pancreatic cancer cell line BxPC3-R | Down       | /                 | Acquisition of drug-resistant phenotype of pancreatic cancer cell                                                   | 2016   | 35   |
| Carbon ion beam radiotherapy                                           | miR-374c-5p    | Mouse, Human    | Mouse squamous cell carcinoma line NR-51, human pancreatic cancer cell lines Panc1 and MIA-PaCa-2 | Down       | /                 | Increased the sensitivity of both Panc1-1 and MIA-PaCa-2 cells to carbon ion beam irradiation                        | 2016   | 36   |
| Colorectal cancer                                                      | miR-374b       | Human           | Colorectal cancer cell lines HCT116 and HT29 | Up         | p53/miRNA-374b/AKT1 | Regulate BLM-induced cell apoptosis, and improved the outcome of chemotherapy in CRC                                 | 2017   | 37   |
| Chemotherapeutic resistance of pancreatic cancer                       | miR-374b-5p    | Human           | Pancreatic cancer cell lines BxPC-3, PANC-1, AsPC-1, SW1990, Capan-2, Capan-2, CFPAC-1 and MIA PaCa-2; pancreatic cancer tissues | Down       | Antiapoptotic proteins: BCL-2, BIRC3, XIAP | The decreased expression of miR-374-5p was associated with poor overall and progression free survival. The up-regulation of miR-374b-5p ameliorated the chemoresistance of pancreatic cancer cells to gemcitabine. | 2018   | 38   |
| Myogenic differentiation                                               | miR-374        | Human           | Primary myoblasts and immortalized myoblasts (Myo) | Up         | /                 | /                                                                                                                      | 2013   | 39   |
| C2C12 myoblasts differentiation                                        | miR-374b       | Human           | C2C12 cells                                    | Down       | MRF4              | Suppressed myoblast differentiation                                                                                   | 2015   | 40   |
| Growth plate of cartilage                                              | Mir-374-5p     | Rat             | Primary chondrocytes (PZ)                      | Up         | /                 | Promoted proliferation and inhibited hypertrophic differentiation                                                   | 2018   | 41   |
|                                                                      |                |                 | Hypertrophic chondrocytes (HZ)                  | Down       |                   | Inhibited proliferation and promoted hypertrophic differentiation                                                   |        |      |
| Retinal ganglion cell development                                      | miR-374b       | Mouse           | E14 embryos, RGC-5 cells                       | Up (E12 -PN1 stage) | Brn3b          | miR-23a alone or in combination with miR-374 could attribute to the biphasic expression pattern of Brn3b, thereby affecting the RGC development, but miR-374 by itself cannot regulate the expression of Brn3b | 2014   | 42   |
| Adipocytes differentiation                                             | miR-374a and -374b-5p | Porcine       | Primary porcine preadipocyte                   | Up         | C/EBP-β          | Promoted differentiation of primary porcine adipocytes                                                             | 2013   | 43   |
| Hair production                                                        | miR-374b       | Goat            | Longissimus dorsi, leg and skin tissue         | Up         | /                 | Pushing secondary hair follicle activity changes from catagen to telogen                                           | 2015   | 44   |

Table 1 (Continued)
| Disease                                | Family members | Species              | Tissue and/or cell                                                                 | Expression | Targets or pathway                      | Relationship with disease or clinical significance | Time  | Ref. |
|----------------------------------------|----------------|----------------------|-----------------------------------------------------------------------------------|------------|----------------------------------------|----------------------------------------------------|-------|-----|
| **Ca\(^{2+}\) homeostasis**           | miR-374b       | Human, mouse         | Wild-type and CLDN14 KO mice, primary cultures of mouse TAL cells, mouse MKTAL cells, human HEK293 cells | Up         | Ca\(^{2+}\)/CaSR/ miR-374/ CLDN14     | Renal Ca\(^{2+}\) reabsorption                     | 2012  | 50  |
| **IgA nephropathy**                    | miR-374b       | Human                | Renal tissue and CD19\(^{+}\) B cells or DAKIKI cells                             | Up         | PTEN and Cosmc                         | Increase cell proliferation and abnormal glycosylation of IgA1 | 2015  | 55  |
| **Drug resistant epilepsy**            | miR-374        | Mouse                | Brain tissue                                                                      | Down       | /                                      | /                                                  | 2014  | 59  |
| **Pilocarpine induced epilepsy**       | miR-374-3p     | Rat                  | Hippocampus tissue                                                                | Up         | /                                      | /                                                  | 2015  | 60  |
| **PQ or MPTP treatment induced dopaminergic neurodegeneration** | miR-374-5p     | Mouse                | Neuro-2a cells                                                                    | Down       | /                                      | /                                                  | 2018  | 61  |
| **Alzheimer’s disease**                | miR-374        | Human                | Tissues and cell lines (neuroblastoma SH-SYSY cells)                              | Down       | BACE1                                  | As potential biomarker to improve AD diagnosis      | 2018  | 62  |
| **Amyotrophic lateral sclerosis**      | miR-374b-5p    | Human                | Serum                                                                             | Down       | /                                      | Promote myoblast differentiation to compensate for the muscle degeneration associated with ALS | 2017  | 65  |
| **Hypoxic–ischemic encephalopathy**    | miR-374a       | Human                | Serum of umbilical cord blood                                                     | Down       | /                                      | MiR-374a could help to elevate the diagnostic value and prognostic prediction of S100B and NSE for HIE | 2017  | 66  |
| **Acute Coronary Syndrome**            | miR-374b-5p    | Human                | PBMCs                                                                             | Down       | Up                                     | STEMI as compared with NSTEMI                       | 2013  | 67  |
| **Myocardial I/R**                     | miR-374        | Rat                  | Left ventricular tissue, HEK-293T cells, cardiomyocytes                           | Down (I/R) | Up (sevoflurane + I/R)                 | Exerted protective effects by inhibiting SP1 through activating the PI3K/Akt pathway in rat models pretreated with sevoflurane | 2018  | 68  |
| **Cardiac dysfunction of selenium deficiency** | miR-374        | Rat                  | Heart tissue                                                                      | Up         | Wnt/j-catenin signaling pathway        | Mainly associated with miR-374                      | 2015  | 69  |
| **Cardiac Hypertrophy**                | miR-374-3p     | Rat                  | Neonatal rat ventricular myocytes, isolated cardiomyocytes                        | Down       | VEGFR1 and PKG-1                      | Activated cardiac hypertrophy via activation of the Ca\(^{2+}\) signaling pathway | 2017  | 70  |
| **Aneurysm**                           | miR-374a-5p    | Human                | Tissue samples of ascending aorta                                                 | Up         | /                                      | Maintained vascular integrity                       | 2016  | 71  |

(Continues)
| Disease                                                                 | Family members | Species            | Tissue and/or cell                                      | Expression | Targets or pathway          | Relationship with disease or clinical significance                                                                 | Time | Ref. |
|------------------------------------------------------------------------|----------------|--------------------|--------------------------------------------------------|------------|----------------------------|---------------------------------------------------------------------------------------------------------------------|------|------|
| Polyphenols feeding                                                     | miR-374*       | Mouse              | Livers tissue of wild-type or apoE-deficient mice      | Down       | AKT1                       | Identifi ed as being commonly modulated by these polyphenols                                                        | 2012 | 72   |
| T cell acute lymphoid leukemia                                          | miR-374        | Human              | Bone marrow samples, thymocytes and peripheral blood T-cells | Up         | /                          | /                                                                                                                   | 2013 | 75   |
| T-cell lymphoblastic lymphoma                                           | miR-374b       | Human              | T-LBL tissue samples, T-cell lines (Jurkat and SUP-T1)  | Down       | Wnt-16 and AKT1            | Associated with worse survival and higher relapse rate in patients with T-ALL                                       | 2015 | 76   |
| Acute graft-versus-host disease                                         | miR-374b-5p    | Human              | Serum                                                  | Down       | /                          | Had diagnostic value by ROC analysis                                                                                 | 2017 | 77   |
| Mice infected with self-healing P. chabaudi malaria                    | miR-374*       | Mouse              | Sera and livers                                        | Down       | /                          | /                                                                                                                   | 2011 | 78   |
| Porcine infected with Salmonella Typhimurium                           | miR-374a-5p    | Porcine            | Intestinal mucosa tissue                               | Up         | /                          | /                                                                                                                   | 2016 | 79   |
| Hyperoxic stress and recovery induced lung injury                      | miR-374        | Rat                | AT2 epithelial cells                                   | /          | Actinin alpha 4, actinin alpha 1, and Na,K-ATPase /                  | /                                                                                                                   | 2013 | 80   |
| Age-related macular degeneration                                       | miR-374a       | Human              | Primary human RPE cells                                 | /          | Fas                        | miR-374a could prevent Fas up-regulation under oxidative conditions to improve survival of human RPE cells              | 2017 | 81   |
| /                                                                      | miR-374a-5p    | Human              | HEK293T, derivative of HEK293T stably expressing HTT-exon 1 with 51 CAG-repeats | /          | E3 ubiquitin ligase MID1  | /                                                                                                                   | 2018 | 82   |

HCC, Hepatocellular Carcinoma; CRC, colorectal cancer; LRH-1, Liver receptor homolog-1; GC, gastric cancer; RECK, reversion-inducing cysteine-rich protein with Kazal motif; MSC, mesenchymal stem cells; PC, prostate cancer; EPC, Endothelial progenitor cell; SCLC, small cell lung cancer; MCC, Merkel cell carcinoma; SCC, squamous cell carcinoma; DEGs, Differentially expressed genes; BCL2, B-cell lymphoma 2; BIRC3, Baculoviral IAP Repeat Containing 3; XIAP, X-linked inhibitor of apoptosis; TAL, thick ascending limb; RGC, Retinal Ganglion Cell; IgAN, IgA nephropathy; ALS, amyotrophic lateral sclerosis; ACS, acute coronary syndrome; I/R, Ischemia-Reperfusion T-ALL, T cell acute lymphoid leukemia; T-LBL, Lymphoblastic lymphoma of T-cell lineage; IgAN, IgA nephropathy; HIE, Hypoxic–ischemic encephalopathy; aGvHD, Acute graft-versus-host disease; AMD, Age-related macular degeneration; PZ, proliferative zone; HZ, hypertrophic zones; NRVMs, Neonatal rat ventricular myocytes; RPE, retinal pigment epithelial.
different pathological and physiological conditions. This will also be a deficiency in this area.

Therefore, there is still a long way to go until they are used in disease prediction and targeted therapy. So we should accelerate the process of translation of preclinical results into clinic and make them into phase I and II trials to guide clinical diagnosis and treatment. But, this will be a large challenge and hard to pursue in the future. Indeed, the safety (activation of viral delivery systems to immune response, off-target effect of ncRNAs, competition with endogenous miRNAs and multi-targeting of miRNAs) of ncRNAs, as clinical therapeutic targets, needs to be established with certainty.

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CONFLICTS OF INTEREST

None.

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REFERENCES

1. Djebali S, Davis CA, Merkel A, et al. Landscape of transcription in human cells. Nature. 2012;489(7414):101-108.
2. Gascoigne DK, Cheetham SW, Cattenoz PB, et al. Pinstripe: a suite of programs for integrating transcriptomic and proteomic datasets identifies novel proteins and improves differentiation of protein-coding and non-coding genes. Bioinformatics. 2012;28(23):3042-3050.
3. Anastasiadou E, Jacob LS, Slack FJ. Non-coding RNA networks in cancer. Nat Rev Cancer. 2018;18(1):5-18.
4. Wang W, Bian H, Li F, et al. HBeAg induces the expression of macrophage miR-155 to accelerate liver injury via promoting production of inflammatory cytokines. Cell Mol Life Sci. 2018;75(14):2627-2641.
5. Qi J, Qiao Y, Wang P, et al. microRNA-210 negatively regulates LPS-induced production of proinflammatory cytokines by targeting NF-kB1 in murine macrophages. FEBS Lett. 2012;586(8):1201-1207.
6. Guojing L, Zhang R, Xu J, et al. Functional conservation of both CDS- and 3′-UTR-located microRNA binding sites between species. Mol Biol Evol. 2015;32(12):3276.
7. Zhou H, Rigoutsos I. miR-103a-3p targets the 5′ UTR of GPRCSA in pancreatic cells. RNA. 2014;20(9):1431-1439.
8. Zhao Q, Li T, Qi J, et al. The miR-545/374a cluster encoded in the Ftx IncRNA is overexpressed in HBV-related hepatocellular carcinoma and promotes tumorigenesis and tumor progression. PLoS ONE. 2014;9(10):e109782.
9. Fedoriw A, Mugford J, Magnuson T. Genomic imprinting and epigenetic control of development. Cold Spring Harb Perspect Biol. 2012;4(7):a008136.
10. Kobayashi S, Totoki Y, Soma M, et al. Identification of an imprinted gene cluster in the X inactivation center. PLoS ONE. 2013;8(8):e71222.
11. Yang L, Kirby JE, Sunwoo H, Lee JT. Female mice lacking Xist RNA show partial dosage compensation and survive to term. Genes Dev. 2016;30(15):1747-1760.
12. Chureau C, Prissette M, Bourdet A, et al. Comparative sequence analysis of the X-inactivation center region in mouse, human, and bovine. Genome Res. 2002;12(6):894-908.
13. Liu F, Yuan JH, Huang JF, et al. Long noncoding RNA FTX inhibits hepatocellular carcinoma proliferation and metastasis by binding MCM2 and miR-374a. Oncogene. 2016;35(41):5422-5434.
14. da Rocha ST, Heard E. Novel players in X inactivation: insights into Xist-mediated gene silencing and chromosome conformations. Nat Struct Mol Biol. 2017;24(3):197-204.
15. Chureau C, Chantalat S, Romito A, et al. Ftx is a non-coding RNA which affects Xist expression and chromatin structure within the X-inactivation center region. Hum Mol Genet. 2011;20(4):705-718.
16. Romito A, Rougeulle C. Origin and evolution of the long non-coding RNAs in the X-inactivation center. Biochimie. 2011;93(11):1935-1942.
17. Wang C, Yang C, Chen X, et al. Altered profile of seminal plasma microRNAs in the molecular diagnosis of male infertility. Clin Chem. 2011;57(12):1722-1731.
18. Schwingshackl L, Schwedhelm C, Galbete C, Hoffmann C. Adherence to Mediterranean diet and risk of cancer: an updated systematic review and meta-analysis. Nutrients. 2017;9(10):E1063.
19. Masuda T, Hayashi N, Kuroda Y, et al. MicroRNAs as biomarkers in colorectal cancer. Cancers (Basel). 2017;9(9):E124.
20. Yang Z, Xie L, Han L, et al. Circular RNAs: regulators of cancer-related signaling pathways and potential diagnostic biomarkers for human cancers. Theranostics. 2017;7(12):3106-3117.
21. Bao S, Zheng J, Li N, et al. Serum microRNA levels as a noninvasive diagnostic biomarker for the early diagnosis of hepatitis B virus-related liver fibrosis. Gut Liver. 2017;11(6):860-869.
22. Wu X, Li S, Xu X, et al. The potential value of miR-1 and miR-374b as biomarkers for colorectal cancer. Int J Clin Exp Pathol. 2015;8(3):2840-2851.
23. Qu R, Hao S, Jin X, et al. MicroRNA-374b reduces the proliferation and invasion of colon cancer cells by regulation of LRH-1/Wnt signaling. Gene. 2018;642:354-361.
24. Xie J, Tan ZH, Tang X, et al. miR-374b-5p suppresses RECK expression and promotes gastric cancer cell invasion and metastasis. World J Gastroenterol. 2014;20(46):17439-17447.
25. Ji R, Zhang X, Qian H, et al. miR-374D mediates the malignant transformation of gastric cancer associated mesenchymal stem cells in an experimental rat model. Oncol Rep. 2017;38(3):1473-1481.
26. Sierzeza M, Kaczor M, Kolodziejczyk P, et al. Evaluation of serum microRNA biomarkers for gastric cancer based on blood and tissue pools profiling: the importance of miR-21 and miR-331. Br J Cancer. 2017;117(2):266-273.
27. He HC, Han ZD, Dai QS, et al. Global analysis of the differentially expressed microRNAs of prostate cancer in Chinese patients. BMC Genom. 2013;14:757.
28. Merhautova J, Hezova R, Poprach A, et al. miR-155 and miR-484 are associated with time to progression in metastatic renal cell carcinoma treated with sunitinib. Biomed Res Int. 2015;2015:941980.
29. Liao YY, Tsai HC, Chou PY, et al. CCL3 promotes angiogenesis by dysregulation of miR-374b/VEGFA-A axis in human osteosarcoma cells. Oncotarget. 2016;7(4):4310-4325.
30. Miko E, Czimerzer Z, Csényk E, et al. Differentially expressed microRNAs in small cell lung cancer. Exp Lung Res. 2009;35(8):646-664.
31. Hou Z, Yang J, Wang G, et al. Bioinformatic analysis of gene expression profiles of pituitary gonadotroph adenomas. Oncol Lett. 2018;15(2):1655-1663.

32. Ning MS, Kim AS, Prasad N, et al. Characterization of the merkel cell carcinoma miRNA. J Cancer. 2014;2014:289548.

33. Li XJ, Li ZF, Wang JJ, et al. Effects of microRNA-374 on proliferation, migration, invasion and apoptosis of human SCC cells by targeting Gadd45a through P53 signaling pathway. Biosci Rep. 2017;37(4):BSCR20170210.

34. Garcia-Cruz R, Camats M, Calin GA, et al. Claudin-14 regulates renal calcium handling. J Am Soc Nephrol. 2015;26(3):663-676.

35. Gamba G, Friedman PA. Thick ascending limb: the Na⁺:K⁺:2Cl⁻ cotransporter. Pflugers Arch. 2015;467(4):670-675.

36. Baek SJ, Sato K, Nishida N, et al. MicroRNA miR-374, a potentially radiosensitizer for carbon ion beam radiotherapy. Oncol Rep. 2016;36(5):2946-2950.

37. Schreiber R, Mezenec R, Matyunina LV, McDonald JF. Evidence for the role of microRNA 374b in acquired cisplatin resistance in pancreatic cancer cells. Cancer Gene Ther. 2016;23(8):241-245.

38. Baek SJ, Sato K, Nishida N, et al. MicroRNA, miR-374b, directly targets Myf6 and negatively regulates C2C12 myoblasts differentiation. Biochem Biophys Res Commun. 2015;467(4):518-525.

39. Dávila P, Barat A, Polesskaya A, et al. Simultaneous miRNA and mRNA transcriptome profiling of human myoblasts reveals a novel set of myogenic differentiation-associated miRNAs and their target genes. BMC Genom. 2013;14:265.

40. Schreiber R, Mezenec R, Matyunina LV, McDonald JF. Evidence for the role of microRNA 374b in acquired cisplatin resistance in pancreatic cancer cells. Cancer Gene Ther. 2016;23(8):241-245.

41. Jee YH, Wang J, Yue S, et al. Mir-374-5p, mir-379-5p, and mir-503-5p regulate proliferation and hypertrophic differentiation of growth plate chondrocytes in male rats. Endocrinology. 2018;159(3):1469-1478.

42. Reashed VA, Sreekanth S, Dhanesh SB, et al. Developmental wave of Brn3b expression leading to RGC fate specification is synergetically maintained by miR-23a and miR-374. Dev Neurobiol. 2014;74(12):1155-1171.

43. Pan S, Zheng Y, Zhao R, Yang X. miRNA-374 regulates dexamethasone-induced differentiation of primary cultures of porcine adipocytes. Horm Metab Res. 2013;45(7):518-525.

44. Su R, Fu S, Zhang Y, et al. Comparative genomic approach reveals novel conserved microRNAs in Inner Mongolia cashmere goat skin and longitudinal dorsi. Mol Biol Rep. 2015;42(5):989-995.

45. Riccardi D, Brown EM. Physiology and pathophysiology of the calcium-sensing receptor in the kidney. Am J Physiol Renal Physiol. 2010;298(3):F485-F499.

46. Gamba G, Friedman PA. Thick ascending limb: the Na⁺:K⁺:2Cl⁻ co-transporter, NKCC2, and the calcium-sensing receptor, CaSR. Pfuigers Arch. 2009;458(1):61-76.

47. Tsukita S, Furuse M, Itoh M. Multifunctional strands in tight junctions. Nat Rev Mol Cell Biol. 2001;2(4):285-293.

48. Mineta K, Yamamoto Y, Yamazaki Y, et al. Predicted expansion of the claudin multigene family. FEBS Lett. 2011;585(4):606-612.

49. Hou J, Renigunta A, Gomes AS, et al. Claudin-16 and claudin-19 interaction is required for their assembly into tight junctions and for renal reabsorption of magnesium. Proc Natl Acad Sci USA. 2009;106(36):15350-15355.

50. Gong Y, Renigunta V, Himmelreich N, et al. Claudin-14 regulates renal Ca²⁺ transport in response to CaSR signalling via a novel microRNA pathway. EMBO J. 2012;31(8):1999-2012.

51. Gong Y, Himmelreich N, Plam A, et al. Epigenetic regulation of microRNAs controlling CLDN14 expression as a mechanism for renal calcium handling. J Am Soc Nephrol. 2015;26(3):663-676.

52. Zhang YM, Zhou XJ, Zhang H. What genetics tells us about the pathogenesis of IgA nephropathy: the role of immune factors and infection. Kidney Int. 2017;2(3):318-331.

53. Soares MF. An update on pathology of IgA nephropathy. J Bras Nefrol. 2016;38(4):435-440.

54. Magistroni R, D’Agati VD, Appel GB, Kiryluk K. New developments in the genetics, pathogenesis, and therapy of IgA nephropathy. Kidney Int. 2015;88(5):974-989.

55. Hu S, Bao H, Xu X, et al. Increased miR-374b promotes cell proliferation and the production of aberrant glycosylated IgA1 in B cells of IgA nephropathy. FEBS Lett. 2015;589(24 Pt B):4019-4025.

56. Nelson PT, Wang WX. MIR-107 is reduced in Alzheimer’s disease brain neocortex: validation study. J Alzheimers Dis. 2010;21(1):75-79.

57. Lusardi TA, Farr CD, Faulkner CL, et al. Ischemic preconditioning regulates expression of microRNAs and a predicted target, MeCP2, in mouse cortex. J Cereb Blood Flow Metab. 2010;30(4):744-756.

58. Packer AN, Xing Y, Harper SQ, et al. The bifunctional microRNA miR-9/miR-9* regulates REST and CoREST and is downregulated in Huntington’s disease. J Neurosci. 2008;28(53):14341-14346.

59. Moon J, Lee ST, Choi J, et al. Unique behavioral characteristics and microRNA signatures in a drug resistant epilepsy model. PLoS ONE. 2014;9(1):e85617.

60. Liu X, Wu Y, Huang Q, et al. Grouping pentylentetrazol-induced epileptic rats according to memory impairment and microRNA expression profiles in the hippocampus. PLoS ONE. 2015;10(5):e0126123.

61. Wang Q, Zhan Y, Ren N, et al. Paragaut and MPTP alter microRNA expression profiles, and downregulated expression of miR-17-5p contributes to PQ induced dopaminergic neurodegeneration. J Appl Toxicol. 2018;38(5):665-677.

62. Manzine PR, Pelucchi S, Horst MA, et al. microRNA 221 targets ADAM10 mRNA and is downregulated in Alzheimer’s disease. J Alzheimers Dis. 2018;61(1):113-123.

63. McDermott CJ, Shaw PJ. Diagnosis and management of motor neuron disease. BMJ. 2008;336(7645):658-662.

64. Ferraiiuolo L, Kirby J, Grierson AJ, et al. Molecular pathways of motor neuron injury in amyotrophic lateral sclerosis. Nat Rev Neurol. 2011;7(11):616-630.

65. Waller R, Goodall EF, Milo M, et al. Serum miRNAs miR-206, 143-3p and 374b-5p as potential biomarkers for amyotrophic lateral sclerosis (ALS). Neurobiol Aging. 2017;55:123-131.

66. Wang Z, Liu Y, Shao M, et al. Combined prediction of miR-210 and miR-374a for severity and prognosis of hypoxic-ischemic encephalopathy. Brain Behav. 2017;8(1):e00835.

67. Ward JA, Esa N, Pidikiti R, et al. Circulating cell and plasma microRNA profiles differ between non-ST-segment and ST-segment-elevation myocardial infarction. Fam Med Med Sci Res. 2013;2(2):108.

68. Zhang SB, Liu TJ, Pu GH, et al. MicroRNA-374 exerts protective effects by inhibiting SP1 through activating the PI3K/Akt pathway in rat models of myocardial ischemia-reperfusion after sevoflurane preconditioning. Cell Physiol Biochem. 2018;46(4):1455-1470.

69. Xing Y, Liu Z, Yang G, et al. MicroRNA expression profiles in rats with selenium deficiency and the possible role of the Wnt/β-catenin signaling pathway in cardiac dysfunction. Int J Mol Med. 2015;35(1):143-152.

70. Lee JS, Song DW, Park JH, et al. miR-374 promotes myocardial hypertrophy by negatively regulating vascular endothelial growth factor receptor-1 signaling. BMB Rep. 2017;50(4):208-213.

71. Licholai S, Blaž M, Kapelak B, Sanak M. Unbiased profile of microRNA expression in ascending aortic aneurysm tissue: a new mechanism of the action of polyphenols. PLoS ONE. 2012;7(1):e29837.
73. Farag SS, VanDeusen JB, Fehninger TA, Caligiuri MA. Biology and clinical impact of human natural killer cells. *Int J Hematol*. 2003;78(1):7-17.

74. Dalmazzo LF, Jacomo RH, Marinato AF, et al. The presence of CD56/CD16 in T-cell acute lymphoblastic leukaemia correlates with the expression of cytotoxic molecules and is associated with worse response to treatment. *Br J Haematol*. 2009;144(2):223-229.

75. Gimenes-Teixeira HL, Lucena-Araujo AR, Dos Santos GA, et al. Increased expression of miR-221 is associated with shorter overall survival in T-cell acute lymphoid leukemia. *Exp Hematol Oncol*. 2013;2(1):10.

76. Qian D, Chen K, Deng H, et al. MicroRNA-374b suppresses proliferation and promotes apoptosis in T-cell lymphoblastic lymphoma by repressing AKT1 and Wnt-16. *Clin Cancer Res*. 2015;21(21):4881-4891.

77. Crossland RE, Norden J, Juric MK, et al. Expression of serum microRNAs is altered during acute graft-versus-host disease. *Front Immunol*. 2017;8:308.

78. Delić D, Dkhil M, Al-Quraishy S, Wunderlich F. Hepatic miRNA expression reprogrammed by *Plasmodium chabaudi* malaria. *Parasitol Res*. 2011;108(5):1111-1121.

79. Uribe JH, Collado-Romero M, Zaldívar-López S, et al. Transcriptional analysis of porcine intestinal mucosa infected with *Salmonella typhimurium* revealed a massive inflammatory response and disruption of bile acid absorption in ileum. *Vet Res*. 2016;47:11.

80. Bhargava M, Dey S, Becker T, et al. Protein expression profile of rat type two alveolar epithelial cells during hyperoxic stress and recovery. *Am J Physiol Lung Cell Mol Physiol*. 2013;305(9):L604-L614.

81. Tasharrofi N, Kouhkan F, Soleimani M, et al. Survival improvement in human retinal pigment epithelial cells via Fas receptor targeting by miR-374a. *J Cell Biochem*. 2017;118(12):4854-4861.

82. Unterbruner K, Matthes F, Schilling J, et al. MicroRNAs miR-19, miR-340, miR-374 and miR-542 regulate MID1 protein expression. *PLoS ONE*. 2018;13(1):e0190437.

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