Phytoplankton variability in relation to some environmental factors in the eastern coast of Suez Gulf, Egypt

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Abstract Water samples were seasonally collected from 12 stations of the eastern coast of Suez Gulf during autumn of 2012 and winter, spring, and summer of 2013 in order to investigate phytoplankton community structure in relation to some physicochemical parameters. The study area harbored a diversified phytoplankton community (138 species), belonging to 67 genera. Four algal groups were represented and classified as Bacillariophyceae (90 species), Dinophyceae (28 species), Cyanophyceae (16 species), and Chlorophyceae (4 species). The results indicated a relative high occurrence of some species namely; Pleurotaenium trabecula of green algae; Chaetoceros lorenzianus, Proboscia alata var. gracillima, Pseudosolenia calcar-avis, and Pseudonitzschia pungens of diatoms; Trichodesmium erythraeum and Pseudoanabaena limnetica of cyanophytes. Most of other algal species were fairly distributed at the selected stations of the study area. The total abundance of phytoplankton was relatively low (average of 2989 unit/L) in the eastern coast of Suez Gulf, as compared its western coast and the northern part of the Red Sea. The diversity of phytoplankton species was relatively high (2.35–3.82 nats) with an annual average of 3.22 nats in the present study. The results concluded that most of eastern coast of Suez Gulf is still healthy, relatively unpolluted, and oligotrophic area, which is clearly achieved by the low values of dissolved phosphate (0.025–0.3 μM), nitrate (0.18–1.26 μM), and dissolved ammonium (0.81–5.36 μM). Even if the occurrence of potentially harmful algae species was low, the study area should be monitored continuously. The dissolved oxygen ranged between 1.77 and 8.41 mg/L and pH values between 7.6 and 8.41. The multiple regression analysis showed that the dissolved nitrate and pH values were the most effective factors that controlled the seasonal fluctuations of phytoplankton along the eastern coast of Suez Gulf during 2012–2013.

Keywords Phytoplankton · Diversity · Physicochemical parameters · Eastern Suez Gulf · Egypt

Introduction

The northern end of the Red Sea bifurcates into the Sinai Peninsula, creating the Gulf of Suez in the west and the Gulf of Aqaba to the east. The gulf is relatively shallow and formed within a relatively young but now inactive Gulf of Suez Rift basin, dating back about 28 million years. It stretches some 300 km north by northwest, terminating at the Egyptian city of Suez and the entrance to the Suez Canal. Along the mid-line of the gulf is the boundary between Africa and Asia. The length of the gulf, from its mouth at the Strait of Jubal to its head at the city of Suez, is 314 km, and it varies in width from
19 to 32 km. The Gulf of Suez is relatively shallow, with a maximum depth of about 64 m; outside its mouth, the depth drops sharply to about 1255 m.

However, the Suez Gulf is subjected to sources of pollution such as shipping activities, where transport of oil continues to play a critical role in marine pollution in the northern part of the gulf and Suez Canal. On the other hand, extensive oil production operations are taking place in the gulf, both inshore and offshore. In addition, the gulf is subjected to industrial, agricultural, and domestic sewage; thermal pollution from power, desalination plants; and tourism activities (TEAM 2000, and NPA 2003a, b, c). In fact, there is no analysis of the tourism-related literature or recent analysis of impacts. The most published topics relate to coral breakage and its management. A full account of tourism’s environmental impacts is constrained by limited tourism data (Gladstone et al. 2013). However, the western coast of the gulf is considered more polluted than the eastern coast due to urbanization resulting from the population expansion, establishment of new industries along the coast such as fertilizer and cement factories, chemicals, and organic wastes from food processing factories at Suez City, and, in addition, to more tourism activities due to the establishment of numerous touristy villages. Thus, the Gulf of Suez could be fairly considered the most polluted area in the Red Sea (TEAM 2000 and NPA 2003a, b, c).

Noticeably, all these pollutants affected the marine ecosystem, which becomes under variable pressure, causing radical changes in marine organisms, including coral reefs, invertebrates, seagrasses, seaweeds, phytoplankton, and others (TEAM 2000 and NPA 2003a, b, c).

In fact, phytoplankton communities are the basis of many marine and freshwater food webs. Their composition fluctuates depending on hydrological conditions, such as light, temperature, salinity, pH, nutrients, and turbulence (Huertas et al. 2011). Typically, diatoms dominate coastal marine communities. However, other groups of phytoplankton can dominate depending on the combination of hydrological conditions and climatic variability (Leterme et al. 2006). Changes in dominant base groups/species often propagate up the food chain, impacting on fish, marine mammals, and birds (Donnelly et al. 2007). Phytoplankton are known to exhibit rapid responses to changes in environmental conditions and are therefore commonly acknowledged as excellent bioindicators of the impact of natural and seasonal changes in coastal ecosystems (Rimet and Bouchez 2012). Their susceptibility to environmental change is usually expressed by morphological and/or behavioral changes as well as by persistent or seasonally a typical differences in abundance and distribution (Leterme et al. 2010, 2013). Where mono-or class-specific blooms are observed on an annual basis, they often vary significantly in magnitude and/or duration between years (Ji et al. 2006 and Leterme et al. 2014).

The phytoplankton community structure in the northern part of the Red Sea was investigated by several workers and revealed variable biodiversity and community structure according to different ecological conditions and different spatial and temporal scales. Nassar (1994) recorded 76 species including 50 diatoms, 18 dinoflagellates, five blue-green algae, and three species of green algae in Suez Bay of the northern part of Suez Gulf. El-Sherif and Abo El-Ezz (2000) examined the distribution of plankton at Taba, Sharm El-Sheikh, Hurgada, and Safaga at northern Red Sea, recording 41 diatom species, 53 dinoflagellates, 10 cyanophytes, and two chlorophytes. Deyab et al. (2004) recorded 200 phytoplankton species along the Suez Canal, Suez Gulf, and the northern part of the Red Sea with clear dominance of diatoms. Shams El-Din et al. (2005) identified 110 phytoplankton species belonging to seven classes on both sides of the Suez Gulf. Nassar (2007a) studied the phytoplankton dynamics in the western coast of Suez Gulf and recorded 144 species of different groups, and Nassar (2007b) conducted similar study on the phytoplankton abundance in the coastal waters of the Aqaba Gulf, recording 127 taxa. Also, Al-Najjar et al. (2007) studied the seasonal dynamics of phytoplankton in the Gulf of Aqaba. Madkour et al. (2010) reported that the spatial distribution of phytoplankton showed that Gulf of Suez differs in the dominant species and timing of abundance from both Gulf of Aqaba and the southern sites of Sinai Peninsula. Recently, a checklist of 207 phytoplankton species is detected in the Egyptian waters of the Red Sea and some surrounding habitats during the period 1990–2010 (Nassar and Khairy 2014). In fact, the available literatures on phytoplankton population dynamic in the eastern coast of the gulf are scarce, and information is lacking concerning phytoplankton in this area.

The aim of the present work is to follow up the changes that might take place in the standing crop and community structure of the phytoplankton in the coastal waters of eastern coast of the Suez Gulf in response to
changes in the physicochemical characters of water and to compare the results with the previous studies of the surrounding habitats.

Materials and methods

Description of sampling stations

Twelve stations were selected along the eastern coast of the Suez Gulf as shown in Fig. 1. These stations are subjected to different ecological conditions due to the touristic and human activities, sewage and oil effluents, and industrial and thermal effects: St. 1 is located near the Electrical Power Station of Ayon Mousa at about 500 m of the coast and is subjected to thermal water discharge. St. 2 is situated near a tourist village namely Tamara Crouze with low human and tourist activities. St. 3 is located near the beach of Ras Sedr and is subjected to high tourist and human activities especially during summer months. St. 4 is located near a new tourist village namely Daghish village, at which low human and tourist activities were observed. St. 5 is a sandy beach located north of the Hammam Pharaon and is relatively far from the human activities. St. 6 is located near the Hammam Pharaon hot springs; its water is relatively hot and characterized by high vegetations and bad odors. St. 7 (Abu-Zenima) is situated at about 2.5 km from Abu-Zenima City. St. 8 is located near from the manganese factory of Abu-Zenima and is subjected to Mn effluents of this factory. St. 9 (Abu-Redis) is located in the south of Abu-Redis City and is subjected to sewage and oil effluents discharged from the oil charging and discharging company. St. 10 (Petropil) is situated near the Petropil Oil Company and is subjected also to oil effluents. St. 11 (Al-Konaysa) is located near a fishing harbor of the Fisheries Commission and is subjected to fishing and human activities. St. 12 (Al-Tur) is situated near the eastern side of Al-Tur fishing harbor and is also subjected to fishing and human activities.

Phytoplankton estimations

Water samples were seasonally collected using Ruttiner bottles from the sub-surface waters of different 12 stations during autumn of 2012 (November) and winter (January), spring (April), and summer (August) of 2013 (Fig. 1). Cell abundance and composition of phytoplankton were estimated according to sedimentation
method (Utermöhl 1958). The species identification was carried out following Peragallo and Peragallo (1908), Ghazzawi (1939), Cupp (1943), Prescott (1962), Bourrelly (1968), Ferguson (1968), Sournia (1986), Mizuno (1990), and Al-Kandari et al. (2009). Then, the phytoplankton species are updated according to the taxonomic database sites, like algaebase.com (ab), World Register of Marine Species (WoRMS), Canadian Register of Marine Species (CaRMS), Nordic Microalgae and Aquatic Protozoa (NOD), and Integrated Taxonomic Information System (ITIS).

Physicochemical parameters

Water temperature was measured by using a simple pocket thermometer graduated to 0.1 °C. The pH value of water samples was measured in situ using a pocket pH meter model Orion 210. Dissolved oxygen was fixed in field and measured according to the modified Winkler’s method according to (Strickland and Parsons 1972), and the dissolved inorganic nutrients (NO3, NH4 and PO4) were determined spectrophotometrically, and the results were expressed in micromolar according to the methods described by APHA (2005).

Statistical analysis

The correlation matrices was applied to total phytoplankton counts, phytoplankton classes, dominant species, and the physicochemical parameters at confidence limit 95% and \( n=47 \). Multiple regression was calculated for phytoplankton during each season, using the program of STATISTICA Version 5. Similarity index between the stations of the study area, based on phytoplankton community structure, was calculated, using the program of Primer 5. The species diversity (\( H' \)) was calculated according to Shannon and Weaver (1963).

Results

Physicochemical parameters

The results of physicochemical parameters of seawater samples collected from the eastern coast of Suez Gulf during 2012–2013 are shown in Table 1.

The temperature was typical of the north part of the Suez Gulf, ranging from a minimum of 17.00 °C during winter at St. 1 and a maximum of 31.50 °C during summer at St. 7 with an annual mean value of 24.35 °C. The normal thermal cycle was clear in the study area, showing the highest temperature during summer (30.10 °C), while in winter, the lowest ones were reached (18.16 °C).

Seawater pH lied in the alkaline side during all seasons with almost the highest values during summer. The lowest value of pH was recorded during winter at St. 9 (7.60) and the highest during summer at St. 1 (8.41). Whereas, seawater dissolved oxygen (DO) varied between a minimum of 1.77 mg/L during spring at St. 3 and a maximum of 8.41 mg/L during winter at St. 9 with small seasonal differences and an annual average of 4.30 mg O2/L.

As far as nutrients are concerned, the reactive phosphate (PO4) was very low during spring and summer at all stations, whereas the maximum value was recorded during autumn at St. 9 (0.30 μM). The dissolved nitrate (NO3) in the gulf ranged between a maximum value of 1.26 μM during autumn at St. 9 and a minimum of 0.18 μM during summer at St. 3. The dissolved ammonium (NH4) varied between a minimum of 0.81 μM during spring at St. 1 and a maximum of 5.36 μM during summer at St. 9, which may be due to the effect of sewage and oil effluents. Generally, nitrate and phosphate exhibited a seasonal cycle with lower concentrations during summer, while dissolved ammonium was the most abundant source of nitrogen during summer (Table 1).

Phytoplankton

Community composition

The study area showed a discrete phytoplankton diversity (138 species), belonging to 67 genera. Four algal groups were represented in the eastern coast of Suez Gulf belonging to Bacillariophyceae (90 species), Dinophyceae (28 species), Cyanophyceae (16 species), and Chlorophyceae (4 species) (Table 2).

The diatoms were the most dominated group, forming about 67.00 % of the total counts of phytoplankton, followed by Cyanophytes that represented about 17.00 % of the total abundance. On the other hand, Dinophyceae and Chlorophyceae formed collectively about 15.50 % of the total counts of phytoplankton (Table 3).

The phytoplankton diversity displayed wide spatial variations. The station 9 was reported as the most
diversified community (93 species), while station 1 recorded the lowest diversified one (59 species) (Table 2). On the other hand, there were no distinct seasonal variations in phytoplankton diversity, where the three seasons: autumn, winter, and spring harbored closed number of species (64, 66, and 69 species), respectively, whereas the summer harbored relatively low number (53 species).

Regardless of the large number of phytoplankton species in the study area, only nine species were perennial (occurring during the four seasons). These species are *Guinardia flaccida*, *Gyrosigma attenuatum*, *Nitzschia longissima*, *Nitzschia sigma*, *Odontella obtusa*, *Synedra ulna*, *Thalassiothrix longissima*, *Phormidium* sp., and *Ceratium trichoceros*. Whereas, 21 species appeared during the three seasons and are considered as semi-perennial. The rest number of species was observed either for one or two seasons. On the other hand, there were few species restricted to one station such as *Amphora grevilleana* (St. 1); *Chaetoceros curvisetus*, *Synedra crystallina*, *Anabaena* sp. (St. 2); *Asterolampra* sp., *Paralia sulcata*, *Dinophysis* sp., and *Phalacroma* sp. (St. 4); *Fragilaria pectinalis* and *Fragilaria construens* (St. 7); *Protoperidinium***

| Table 1 | Seasonal variations of temperature (°C), pH value, DO (mg O₂/L), and the nutrients PO₄, NO₃, and NH₄ (µM) along the eastern coast of Suez Gulf during 2012–2013 |
|---------|-------------------------------------------------|
| Autumn 2012 |                                                                                   |
| Station  | 1          | 2              | 3              | 4              | 5              | 6              | 7              | 8              | 9              | 10             | 11             | 12             | Average |
| Temp     | 22.00      | 23.00          | 22.40          | 22.70          | 22.80          | 23.80          | 23.10          | 22.60          | 24.00          | 23.20          | 22.20          | 22.94          |         |
| pH       | 8.22       | 8.00           | 8.20           | 8.00           | 8.10           | 8.05           | 8.12           | 7.66           | 7.70           | 7.80           | 7.80           | –               |         |
| DO       | 3.00       | 3.63           | 2.38           | 4.25           | 3.56           | 3.30           | 3.38           | 3.11           | 7.50           | 6.50           | 4.60           | 4.32           | 4.12       |
| PO₄      | 0.122      | 0.166          | 0.09           | 0.173          | 0.147          | 0.134          | 0.144          | 0.126          | 0.30           | 0.26           | 0.186          | 0.175          | 0.17       |
| NO₃      | 0.55       | 0.70           | 0.52           | 0.71           | 0.61           | 0.50           | 0.61           | 0.40           | 1.26           | 1.10           | 0.77           | 0.73           | 0.71       |
| NH₄      | 2.66       | 1.28           | 2.32           | 1.26           | 1.16           | 1.53           | 1.51           | 1.63           | 0.84           | 1.07           | 1.10           | 1.17           | 1.46       |

| Winter 2013 |                                                                                   |
| Temp       | 17.00      | 18.40          | 17.50          | 18.00          | 18.8           | 18.2           | 19.00          | 18.6           | 17.70          | 19.00          | 18.50          | 17.30          | 18.16 |
| pH         | 8.30       | 7.90           | 8.22           | 7.90           | 8.00           | 8.10           | 8.10           | 8.20           | 7.60           | 7.85           | 7.90           | 7.90           | –       |
| DO         | 3.32       | 4.14           | 2.88           | 4.26           | 4.1            | 3.78           | 3.90           | 3.40           | 8.41           | 5.71           | 4.63           | 4.63           | –       |
| PO₄       | 0.11       | 0.14           | 0.09           | 0.14           | 0.13           | 0.13           | 0.11           | 0.28           | 0.25           | 0.17           | 0.15           | 0.15           | –       |
| NO₃       | 0.32       | 0.46           | 0.37           | 0.47           | 0.45           | 0.42           | 0.43           | 0.38           | 0.93           | 0.89           | 0.60           | 0.52           | 0.52   |
| NH₄       | 4.72       | 2.28           | 4.21           | 2.18           | 2.32           | 2.60           | 2.40           | 2.96           | 1.62           | 1.86           | 1.90           | 2.12           | 2.60   |

| Spring 2013 |                                                                                   |
| Temp       | 25.00      | 26.80          | 25.50          | 26.20          | 27.40          | 26.40          | 27.00          | 26.00          | ND             | 27.20          | 25.20          | 26.20          |         |
| pH         | 8.32       | 8.00           | 8.30           | 7.90           | 8.00           | 8.20           | 8.17           | 8.22           | 7.80           | ND             | 7.83           | 7.87           | –       |
| DO         | 1.83       | 3.43           | 1.77           | 3.72           | 3.4            | 2.74           | 3.32           | 2.37           | 5.25           | ND             | 4.75           | 4.1            | 4.425 |
| PO₄       | 0.04       | 0.08           | 0.04           | 0.08           | 0.07           | 0.06           | 0.07           | 0.05           | 0.12           | ND             | 0.11           | 0.09           | 0.10   |
| NO₃       | 0.25       | 0.50           | 0.26           | 0.53           | 0.48           | 0.4           | 0.47           | 0.34           | 0.75           | ND             | 0.68           | 0.6            | 0.64   |
| NH₄       | 0.81       | 1.52           | 2.18           | 1.26           | 1.56           | 1.71           | 1.58           | 1.87           | 2.40           | ND             | 0.84           | 1.1            | 0.97   |

| Summer 2013 |                                                                                   |
| Temp       | 28.1       | 30.6           | 28.7           | 29.4           | 31.4           | 30             | 31.5           | 31             | 29.4           | 31.2           | 31.2           | 28.5           | 30.1   |
| pH         | 8.41       | 8.3            | 8.4            | 8.3            | 8.3            | 8.35           | 8.33           | 8.4            | 8.3            | 8.12           | 8.14           | 8.26           | –       |
| DO         | 3.1        | 4.45           | 2.34           | 4.53           | 4.14           | 3.32           | 3.42           | 3.26           | 4.53           | 5.37           | 5.22           | 4.74           | 4.03   |
| PO₄       | 0.03       | 0.05           | 0.02           | 0.05           | 0.04           | 0.04           | 0.04           | 0.03           | 0.05           | 0.06           | 0.06           | 0.05           | 0.04   |
| NO₃       | 0.21       | 0.34           | 0.18           | 0.35           | 0.26           | 0.25           | 0.26           | 0.24           | 0.35           | 0.42           | 0.4            | 0.36           | 0.30   |
| NH₄       | 2.55       | 2.62           | 4.11           | 2.55           | 3.4            | 3.62           | 3.46           | 4.0            | 5.36           | 2.1            | 2.37           | 2.5            | 3.22   |

*ND not measured*
| Diatoms                              | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|-------------------------------------|---|---|---|---|---|---|---|---|---|----|----|----|
| Amphiprora alata (Ehrenberg) Kützing (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Amphiprora paludosa W. Smith (ab)    | + | + | + | + | + | + | + | + | + | + | + | + |
| Amphora grevilleana Gregory (WoRMS)  | + | + | + | + | + | + | + | + | + | + | + | + |
| Amphora lineolata Ehrenberg (ab)     | + | + | + | + | + | + | + | + | + | + | + | + |
| Amphora marina W. Smith (ab)         | + | + | + | + | + | + | + | + | + | + | + | + |
| Amphora ovalis (Kützing) Kützing (WoRMS) | + | + | + | + | + | + | + | + | + | + | + | + |
| Asterionella sp.                     | + | + | + | + | + | + | + | + | + | + | + | + |
| Asterolampra sp.                     | + | + | + | + | + | + | + | + | + | + | + | + |
| Aulacoseira granulata var angustissima (O.F.Müller) Simonsen (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Aulacoseira italica (Ehrenberg) Simonsen (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Bacillaria paradoxa J.F. Gmelin (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Campylodiscus hibernicus Ehrenberg (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Chaetoceros coarctatus* Lauder (ab)   | + | + | + | + | + | + | + | + | + | + | + | + |
| Chaetoceros curvisetus* Cleve (ab)    | + | + | + | + | + | + | + | + | + | + | + | + |
| Chaetoceros densus* (Cleve) Cleve (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Chaetoceros lorenzianus* Grunow (ab)  | + | + | + | + | + | + | + | + | + | + | + | + |
| Chaetoceros peruvianus* Brightwell (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Climacodiun biconcavum Cleve (WoRMS) | + | + | + | + | + | + | + | + | + | + | + | + |
| Climacosphenia moniliger (Ehr. (ab)  | + | + | + | + | + | + | + | + | + | + | + | + |
| Cocconeis placentula Ehrenberg (ab)  | + | + | + | + | + | + | + | + | + | + | + | + |
| Coscinodiscus centralis Ehrenberg (WoRMS) | + | + | + | + | + | + | + | + | + | + | + | + |
| Coscinodiscus granii Gough (ab)      | + | + | + | + | + | + | + | + | + | + | + | + |
| Coscinodiscus marginatus Ehrenberg (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Diaplorus radiatus Ehrenberg (WoRMS) | + | + | + | + | + | + | + | + | + | + | + | + |
| Cyclotella meneghiana* Kützing (ab)  | + | + | + | + | + | + | + | + | + | + | + | + |
| Cymbella aspera Ehrenberg Cleve (WoRMS) | + | + | + | + | + | + | + | + | + | + | + | + |
| Cymbella sp.                         | + | + | + | + | + | + | + | + | + | + | + | + |
| Cymbella ventricosa (C.Agardh) C.Agardh (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Diploneis interrupta (Kützing) Cleve (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Diploneis sp.                        | + | + | + | + | + | + | + | + | + | + | + | + |
| Fragillaria pectinalis (O.F.Müller), Lyngbye (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Fragillaria construens Ehrenberg Grunow (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Fragillaria sp.                      | + | + | + | + | + | + | + | + | + | + | + | + |
| Gramatophora marina (Lyngbye) Kützing (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Gramatophora oceanica Ehrenberg (WoRMS) | + | + | + | + | + | + | + | + | + | + | + | + |
| Guinardia flaccida (Castracane) H. Peragallo (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Gyrosigma acuminatum (Kützing) Rabenhorst (WoRMS) | + | + | + | + | + | + | + | + | + | + | + | + |
| Gyrosigma attenuatum (Kützing) Rabenhorst (ab) | + | + | + | + | + | + | + | + | + | + | + | + |
| Gyrosigma balticum (Ehrenberg) Rabenhorst (WoRMS) | + | + | + | + | + | + | + | + | + | + | + | + |
| Hemiaulus membranacea Cleve          | + | + | + | + | + | + | + | + | + | + | + | + |
| Hemiaulus sinensis Greville          | + | + | + | + | + | + | + | + | + | + | + | + |
| Lauderia annulata Cleve (WoRMS)      | + | + | + | + | + | + | + | + | + | + | + | + |
| Leptocylindrus danicus (ab)          | + | + | + | + | + | + | + | + | + | + | + | + |
| Leptocylindrus minimus Grav (WoRMS)  | + | + | + | + | + | + | + | + | + | + | + | + |
| Table 2 (continued) | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---------------------|---|---|---|---|---|---|---|---|---|----|----|----|
| Leptocylindrus sp.  | + | + | + |   | + |   |   |   |   |    |    |    |
| Licmophora abbreviata C. Agardh (ab) | + |   |   |   | + |   |   |   |   |    |    |    |
| Licmophora flabellata C. Agardh (ab) | + | + | + | ++ | + | ++ | + | ++ | + |    |    |    |
| Licmophora gracilis (Ehrenberg) Grunow (WoRMS) | + | + | + | + | + | + | + | + | + |    |    |    |
| Mastogloia sp.      | + |   |   |   |   | + |   |   |   |    |    |    |
| Melosira sp.        | + |   |   |   |   |   |   |   |   |    |    |    |
| Melosira varians C. Agardh (ab) | + |   |   |   |   | + |   |   |   |    |    |    |
| Navicula dicephala (Ehrenberg) W. Smith (ab) | + | + |   |   | + |   |   |   |   |    |    |    |
| Navicula tripunctata (O.F. Müller) Bory de Saint-Vincent (WoRMS) | + | + | + | ++ | + | ++ | + | ++ | + |    |    |    |
| Navicula placenta (Ehrenberg) Kützing (ab) | + | + | + |   | + |   |   |   |   |    |    |    |
| Navicula sp.        | + | + | + | + | + | + | + | + | + |    |    |    |
| Nitzschia acicularis (Kütz.) W.S. | + | + | + | + | + | + | + | ++ | + |    |    |    |
| Nitzschia closterium* (Ehrenberg) W. Smith (ab) | + | + | + |   |   |   |   |   |   |    |    |    |
| Nitzschia longissima* Brébisson Ralfs in Pritchard (ab) | ++ | + | + | ++ | + | ++ | + | ++ | + |    |    |    |
| Nitzschia obtusa W. Smith (ab) | + | + | + | + | + | + | + | + | + |    |    |    |
| Nitzschia pungens* var. atlantica (Grunow ex Cleve) G.R.Hasle, (ab) | + | + | +++ | ++ | + | ++ | + | +++ | + |    |    |    |
| Nitzschia sigma (Kützing) W. Smith (ab) | + | + | + | ++ | + | ++ | + | ++ | + |    |    |    |
| Nitzschia vermicularis (Kütz.) Hantzsch in Rabenh | + | + | + |   |   |   |   |   |   |    |    |    |
| Odontella aurita (Lyngbye) C.A. Agardh (ab) | + | + | + | + | + | + | + | + | + |    |    |    |
| Odontella obtusa (Kützing) Ralfs (ab) | + | + | + | ++ | + | ++ | + | ++ | + |    |    |    |
| Odontella sinensis (Greville) Grunow (WoRMS) | + | + | + | + | + | + | + | + | + |    |    |    |
| Paralia sulcata (Ehrenberg) Cleve (ab) | + |   |   |   |   |   |   |   |   |    |    |    |
| Plagiotropis lepidoptera (Gregory) Kunze (ab) | + | + | + | + | + | + | + | + | + |    |    |    |
| Pleurosigma angulatum W. Smith (ab) | + | + | + | + | + | + | + | + | + |    |    |    |
| Proboscia alata var.gracillima* (Brightwell) Sündstrom (ab) | ++ | +++ | ++++ | +++ | ++ | +++ | +++ | +++ | +++ |    |    |    |
| Proboscia alata form indica* Brightwell) Sündstrom (ab) | + | + | ++ | + |   |   |   |   |   |    |    |    |
| Pseudosolenia bergoni H. Péragallo (ab) | + | + | + | + | + | + | + | + | + |    |    |    |
| Pseudosolenia calcar avis (Schultze) Sundström (ab) | ++ | +++ | ++++ | +++ | ++ | +++ | +++ | +++ | +++ |    |    |    |
| Rhizosolenia fragilissima Bergon | + | + | + | + |   |   |   |   |   |    |    |    |
| Rhizosolenia imbricata (Cleve) Schröder WoRMS) | ++++ | + | + | + | + | + | + | + | + |    |    |    |
| Rhizosolenia stoterfóthii H. Peragallo (ab) | + | + | + | + | + | + | + | + | + |    |    |    |
| Rhizosolenia styliformis Brightwell (ab) | + | + | + | + | + | + | + | + | + |    |    |    |
| Skeletonema costatum* (Greville Cleve (ab) | + | + | + | + | + | + | + | + | + |    |    |    |
| Stephanopyxis turis (Greville & Arnott in Gregory) Ralfs in Pritchard | + | + | + | + | + | + | + | + | + |    |    |    |
| Striatella unipunctata (Lyngbye) C. Agardh (ab) | ++ | + | ++ | + | + | + | + | + | + |    |    |    |
| Surirella minuta Brébisson (WoRMS) | ++ | + | + | + | + | + | + | + | + |    |    |    |
| Surirella robusta Ehrenberg (ab) | + | + | + | + | + | + | + | + | + |    |    |    |
| Syneidra acus Kütz. | + | + | + | + | + | + | + | + | + |    |    |    |
| Syneidra crystalline (C.Agardh) Kützing (WoRMS) | + |   |   |   |   |   |   |   |   |    |    |    |
| Synedra ulna (Nitzsch) Ehrenberg (ab) | ++ | + | + | + | + | + | + | + | + |    |    |    |
| Synedra undulata (J.W.Bailey) Gregory (ab) | + | + | + | + | + | + | + | + | + |    |    |    |
| Thalassionema nitzchioides* (Grunow) | + | + | + | + | + | + | + | + | + |    |    |    |
Table 2 (continued)

| Diatoms                        | 1  | 2  | 3  | 4  | 5  | 6  | 7  | 8  | 9  | 10 | 11 | 12 |
|-------------------------------|----|----|----|----|----|----|----|----|----|----|----|----|
| Mereschkowsky (ab)            |    |    |    |    |    |    |    |    |    |    |    |    |
| Thalassiosira sp.             | +  | +  | ++ |  + | +  | +  | +  | ++ | ++ |    |    |    |
| Thalassiothrix frauenfeldii (Grunow) Grunow (WoRMS) | ++ | ++ | +  | +  | +  | ++ | +  | +  | +  |    |    |    |
| Thalassiothrix longissima Cleve & Grunow (ab) | ++ | ++ | +++ | ++ | ++ | ++ | +++ | +++ | +++ |    |    |    |
| Trachyneis aspera (Ehrenberg; Ehrenberg) Cleve | +  | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |
| Cyanophytes                   |    |    |    |    |    |    |    |    |    |    |    |    |
| Anabaena sp.                  |    |    |    |    |    |    |    |    |    |    |    |    |
| Chroococcus minutus (Kütz.) Nägeli | +  | ++ | ++ | +  | +  | +  | +  | +  | +  |    |    |    |
| Chroococcus turigidas (Kützing) Nägeli (ab) | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Coelosphaerium sp.            |    | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Lyngbya majuscula Harvey ex Gomont (ab) | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Microcystis sp.               |    |    |    |    |    |    |    |    |    |    |    |    |
| Trichodesmium erythraeum * (Ehrenberg) Geitler (ab) | +  | +  | +  | ++ | +  | ++ | +  | ++ | ++ | +++ | +++ | +++ |
| Pseudanabaena limnetica* (Lemmermann) Komárek (ab) | ++ | ++ | +++ | ++ | ++ | +++ | ++ | ++ | +++ | +++ | +++ | +++ |
| Oscillatoria simplicissima* Gomont (WoRMS) | +  | +  | +  | +  | +  | ++ | +  | ++ | ++ |    |    |    |
| Oscillatoria sp.              |    |    |    |    |    |    |    |    |    |    |    |    |
| Oscillatoria tenuis* C. Agardh (WoRMS) | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Phormidium sp.                |    |    |    |    |    |    |    |    |    |    |    |    |
| Planktothrix formosa*         |    |    |    |    |    |    |    |    |    |    |    |    |
| Spirulina major Kützing ex Gomont (WoRMS) | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Spirulina sp.                 |    |    |    |    |    |    |    |    |    |    |    |    |
| Dinoflagellates               |    |    |    |    |    |    |    |    |    |    |    |    |
| Ceratium breve (Ostenfeld & Schmidt) | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Schroder (ab)                 |    |    |    |    |    |    |    |    |    |    |    |    |
| Ceratium egyptiacum Halim (ab) |    |    |    |    |    |    |    |    |    |    |    |    |
| Ceratium extensum (Gourret) Cleve-Euler (ab) | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Ceratium furca* (Ehrenberg) Claparède & Lachmann (WoRMS) | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Ceratium fusus* (Ehrenberg) Dujardin (ab) | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Ceratium karastenii Pavillard (WoRMS) |    |    |    |    |    |    |    |    |    |    |    |    |
| Ceratium macroceros (Kofoid) Peters (ab) | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Ceratium massiliense (Gourret) E.G.Jørgensen (ab) | +  | +  | +  | +  | +  | ++ | +  | ++ | ++ | +++ | +++ | +++ |
| Ceratium trichoceros (Ehrenberg) Kofoid (WoRMS) | +  | +  | ++ | +  | +  | ++ | ++ | +++ | +++ | +++ | +++ | +++ |
| Ceratium tripus* (O.F.Müller) Nitzsch (WoRMS) | +  | +  | +  | ++ | +  | ++ | ++ | +++ | +++ | +++ | +++ | +++ |
| Dinophysis caudata* Saville-Kent (ab) | +  | +  | +  | +  | +  | +  | +  | +  | +  |    |    |    |
| Dinophysis sp.                |    |    |    |    |    |    |    |    |    |    |    |    |
| Diplopsalis lenticula         |    |    |    |    |    |    |    |    |    |    |    |    |
| Exuviaella compressa (Bailey) Knudsen & in Ostenfeld (ab) | +  | +  | +  | ++ | +  | +  | +  | ++ | ++ | +  | +  | +  |
| Goniuulax sp.                 |    |    |    |    |    |    |    |    |    |    |    |    |
| Gymnodinium sp.               |    |    |    |    |    |    |    |    |    |    |    |    |
| Oxytoxum gracile J.Schiller   |    |    |    |    |    |    |    |    |    |    |    |    |
| Phalacroma sp.                |    |    |    |    |    |    |    |    |    |    |    |    |
| Podolampas palmipes Stein     |    |    |    |    |    |    |    |    |    |    |    |    |
| Prorocentrum compressum (J.W. Bailey) Abé ex Dodge |    |    |    |    |    |    |    |    |    |    |    |    |
divergens (St. 8); Amphora ovalis, Cymbella aspera, Hemiaulus sinensis, Odontella sinensis, Rhizosolenia bergoni, Ceratium egyptiacum, Ceratium karastenii, and Prorocentrum compressum (St. 9), whereas Pediastum clathratum appeared only at St. 11 (Table 2).

Seasonal and spatial variations of phytoplankton community

The diatoms prevailed during all seasons forming numerically the highest percentage (82 %) during winter, followed by cyanophytes, which formed the highest percentage during summer (24 %). The chlorophytes were absent during winter at all stations and formed almost equal percentages during the three other seasons. The fourth group, dinoflagellates, formed numerically (5–13 %) of the total count during the study period. Like the seasonal pattern, the spatial distribution of phytoplankton revealed the dominance of diatoms at all stations (Table 3), forming numerically from 57 % (St. 10) to 80 % (St. 4). The cyanophytes and chlorophytes showed the inverse pattern, recording the highest percentage (26 %, St. 10) and the lowest (8 %, St. 4) for the former and (11 %, St. 10; 3 %, St. 4) for the later. The dinoflagellates displayed the highest contribution at St. 9 (13 %) and the lowest one at St. 5 (5 %) as shown in Fig. 2.

Standing crop

The total abundance of phytoplankton was relatively low in the present study of the eastern coast of Suez Gulf (average of 2989 unit/L) as compared with the previous studies (Table 4). The total phytoplankton showed the highest counts during autumn 2012 with an average of 3801 unit/L, followed by winter (average of 2973 unit/L), spring (average of 2713 unit/L), and summer (average of 2469 unit/L) as shown in Table 3 and Fig. 2. The peak of autumn was due to the co-dominance of C. lorenzianus (4.24 %), G. attenuatum (6.16 %), P. pungens (4.05 %), P. alata var. gracillima (4.37 %), P. calcar-avis (7.35 %), T. erythraeum (3.98 %), Pseudanabaena limnetica (3.75 %), and P. trabecula (9.41 %). However, some of these species were also observed with relative high counts during summer and spring, 2013 such as P. alata var. gracillima, P. calcar-avis, and P. trabecula. Most of other algal species were fairly distributed at the different stations in the coastal waters of eastern coast of Suez Gulf during 2012–2013.
On the spatial scale, the average total count was relatively low in all stations. The relatively high abundance was recorded at St. 9 followed by St. 10 with total counts of 4170 and 3840, respectively, whereas St. 1 and St. 3 sustained the lowest average total counts (2218 and 2123 unit/L, respectively) (Table 3). However, St. 9 sustained relatively high counts of phytoplankton during the whole period, due to the co-dominance of *P. pungens* (3.06 %), *P. alata* var. *gracillima* (3.39 %), *P. calcar-avis* (3.80 %), and *P. trabecula* (4.29 %). The species *P. trabecula* appeared also at St. 10 (9.54 %), with the contribution of *Melosira granulata* (4.77 %), *T. longissima* (3.90 %), *T. erythraeum* (5.78 %), and *P. limnetica* (3.76 %).

**Species diversity**

The total average of diversity in the eastern coast of Suez Gulf was 3.22 (Table 5). The diversity of
Fig. 2  Seasonal variations of phytoplankton abundance (unit/L) at the different stations in the eastern coast of Suez Gulf during 2012–2013
phytoplankton sustained a maximum of 3.82 during winter at St. 9, in which the highest numbers of phytoplankton species were observed (49 spp.). On the other hand, the minimum diversity of 2.35 was found during summer at St. 7, in which the lowest numbers of species were recorded (19 spp.). On the temporal scale, the winter season sustained relatively higher diversity (3.43), whereas the other three seasons sustained relatively closed diversities (Table 5). Spatially, St. 1 and 12 sustained the lowest average of diversity (2.97 and 3.03), respectively, against the highest diversity at St. 9 (3.47).

### Statistical analysis

#### Correlation matrices and multiple regressions

The statistical analysis of the data indicated that the phytoplankton abundance was positively correlated with nitrate ($r=0.66$) and dissolved oxygen ($r=0.51$) but inversely correlated with pH values ($r=-0.63$), whereas the groups and dominant species showed varied correlations with physicochemical characteristics as shown in Table 6. The multiple regression analysis indicated that the dissolved nitrate and pH values were the most effective factors that controlled the seasonal fluctuations of phytoplankton in the eastern coast of Suez Gulf during 2012–2013. The regression model was phytoplankton counts=1379.2341+0.662 NO$_3$–0.33 pH (MR=0.662, $N=47$, $p<0.1278$). The similarity index revealed four clusters (Fig. 3).

### Discussion

Coastal waters are characterized by a high degree of spatial and temporal variability of environmental parameters (Bosak et al. 2012). These ecosystems face increasing anthropogenic influences, mainly due to the increasing human population density in coastal areas, and are described as “critical transition zones” because of their position at terrestrial, freshwater, and marine interfaces (Levin et al. 2001). Therefore, in any evaluation of the ecological consequences of human activities, such as urbanization and tourism, on the functioning of coastal ecosystems, it is essential to determine the basic structural properties of phytoplankton assemblages in these marine areas (Bosak et al. 2012). The reason is that they play a central role in the structure and functioning of

### Table 4 The number of species and abundance of phytoplankton in Suez Gulf

| Algal group     | Western coast of Suez Gulf, Nassar (2000) | Western coast of Suez Gulf, Nassar (2007a) | Eastern coast of Suez Gulf (present study) |
|-----------------|-------------------------------------------|-------------------------------------------|--------------------------------------------|
| Diatoms         | 28 G spp 4252 72.53                        | 40 G spp 10958 70.30                      | 42 G spp 2019 67.54                       |
| Dinoflagellates | 9 18 45 1278 21.80                         | 11 30 957 6.13                            | 12 28 245 8.20                           |
| Cyanophytes     | 3 4 314 5.35                               | 7 12 802 5.14                            | 9 16 507 17.00                           |
| Chlorophytes    | 0 0 0 0.00                                | 10 12 2869 18.4                          | 4 4 218 7.29                            |
| Euglenophytes   | 0 0 0 0.00                                | 1 1 5 0.03                               | 0 0 0 0.00                              |
| Silicoflagellates| 1 1 17 0.29                               | 0 0 0 0.00                               | 0 0 0 0.00                              |
| Total           | 41 70 5862 100                            | 69 144 15591 100                         | 67 138 2989 100                          |

### Table 5 Seasonal variations of species diversity (nats) at the different stations in the eastern coast of Suez Gulf

| Station       | 1   | 2   | 3   | 4   | 5   | 6   | 7   | 8   | 9   | 10  | 11  | 12  | Average |
|---------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|-----|---------|
| Autumn 2012   | 2.71| 3.28| 3.02| 3.06| 3.13| 3.06| 3.09| 3.22| 3.43| 3.35| 3.44| 2.75| 3.13   |
| Winter 2013   | 3.06| 3.46| 3.28| 3.61| 3.52| 3.42| 3.61| 3.26| 3.82| 3.6 | 3.15| 3.33| 3.43   |
| Spring 2013   | 3.00| 3.43| 3.11| 3.67| 3.27| 3.37| 3.34| 3.12| 3.41| ND  | 3.48| 2.86| 3.28   |
| Summer 2013   | 3.11| 3.4  | 2.88| 3.00| 2.88| 2.93| 2.35| 3.14| 3.24| 3.36| 3.33| 3.19| 3.06   |
| Average       | 2.97| 3.39| 3.07| 3.33| 3.20| 3.19| 3.09| 3.18| 3.47| 3.43| 3.35| 3.03| 3.22   |
freshwater and marine ecosystems (Pourafrasyabi and Ramezanpour 2014). Phytoplankton populations are well known to be influenced by space-time variations in hydrochemical and physical parameters (Cloern et al. 1989), such as light, temperature, salinity, pH, nutrients, and turbulence (Leterme et al. 2006).

Variations in pH can affect algal growth in a number of ways. It can change the distribution of carbon dioxide species and carbon availability, alter the availability of trace metals and essential nutrients, and at extreme pH levels potentially cause direct physiological effects (Chen and Durbin 1994). In general, changes in pH levels in marine systems appear to correlate with changes in temperature, dissolved oxygen, and phytoplankton production. Conditions of high pH, high phytoplankton production, and low oxygen conditions are characteristic of nutrient-enriched systems and often are found in the coastal waters (Hinga 2002). However, high pH levels are commonly seen in the late afternoon of sunny summers after the consumption of the CO₂ by photosynthesis process. After sunset, the pH level may significantly be declined due to ending the photosynthesis process (Ghobrani et al. 2014).

In the present study, the highest pH value was recorded during summer and a negative correlation was found between the pH values and phytoplankton ($r=-0.63$) and dissolved oxygen ($r=-0.67$).

Oxygen concentration is an index of the balance between processes of food production and food consumption. This balance is a key descriptor of the changing status of the ecosystem. When the balance is disrupted, the oxygen concentration can fall to low levels (Kemker 2013). Accordingly, the study area is well balanced due to the recorded moderate to high values of oxygen. These values synergized with that of phytoplankton ($r=0.51$). Generally, the slight increase

| Temp | pH | DO  | PO₄ | NO₃ | NH₄ |
|------|----|-----|-----|-----|-----|
| °C   | –  | M₉  | O₂/L| μM  | –   |
| Total phytoplankton | –0.20 | –0.63 | 0.51 | 0.44 | 0.66 | –0.47 |
| Diatoms | –0.51 | –0.69 | 0.48 | 0.69 | 0.63 | –0.46 |
| Dinoflagellates | 0.01 | –0.22 | 0.52 | 0.33 | 0.34 | –0.09 |
| Cyanophytes | 0.27 | –0.18 | 0.19 | 0.14 | 0.27 | –0.18 |
| Chlorophytes | 0.38 | –0.08 | 0.02 | 0.17 | –0.22 |
| Chaetoceros lorenzianus | –0.08 | –0.35 | 0.07 | 0.42 | 0.50 | –0.51 |
| Gyrosigma attenuatum | 0.09 | –0.20 | 0.14 | 0.39 | 0.49 | –0.27 |
| Pseudosolenia calcaris-avis | 0.10 | –0.24 | –0.03 | 0.28 | 0.40 | –0.49 |
| Pleurotaenium trabecula | 0.38 | –0.02 | –0.05 | –0.03 | 0.11 | –0.16 |
| Proboscia alata var. gracillima | 0.33 | 0.27 | 0.07 | –0.10 | –0.15 | 0.23 |
| Pseudo-nitzschia pungens | –0.45 | –0.43 | 0.31 | 0.63 | 0.52 | –0.32 |
| Trichodesmium erythraeum | 0.29 | –0.15 | 0.17 | 0.14 | 0.25 | –0.17 |
| Pseudanabaena limnetica | 0.50 | 0.10 | –0.01 | –0.04 | 0.10 | –0.03 |

Bold correlations are significant at $p<0.05$ and $n=47$.

Fig. 3 Bray-Curtis of similarity of phytoplankton abundance between the different stations
in dissolved oxygen during winter and spring may be due to the increase of oxygen solubility (Nassar 1994). However, the oxygen concentration in the Red Sea is relatively low because of its high salinity and high temperature characteristic of the area (Nassar 2007a). This is also confirmed by Gab-Alla (2007) who reported that Hammam Pharaon hot springs of eastern Suez Gulf (St. 6 in the present study) was slightly acidic (pH 6.3–7.6) and hot (temperature of 25–66 °C) with low oxygen content (0.2–5.5 mg/L) and high salinity (43 %). The nutrient enrichment of coastal waters is generally the main factor driving the succession and composition of phytoplankton communities (Leterme et al. 2014). Phosphorus availability can impact primary production rates in the ocean as well as species distribution and ecosystem structure and in some marine and estuarine environments; P availability is considered the proximal macronutrient that limits primary production (Paytan 2013). According to the low values of dissolved phosphate (0.025–0.3 μM), nitrate (0.18–1.26 μM), and ammonium (0.81–5.36 μM) during 2012–2013, the eastern coast of Suez Gulf is still healthy, relatively unpolluted, and oligotrophic area. This is established with the data reported by Fahmy (2003) who concluded that nitrogen, phosphorus, and reactive silicate concentrations were generally low and allowed classifying the Egyptian Red Sea coastal water as oligotrophic to mesotrophic. This is in addition to the relatively low total abundance of phytoplankton (average of 2989 unit/L), compared with the data reported in its western coast in 1995 (average of 5862 unit/L) by Nassar (2000) and in 2006 (average 15,591 unit/L) by Nassar (2007a), as well as the data reported in 2002 by Shams El-Din et al. (2005) along the both sides of Suez Gulf (average of 6284 unit/L). However, Ghobrani et al. (2014) mentioned that oligotrophic waters are characterized by high clarity and little counts of algae. However, the relative high abundance of phytoplankton in this study was found at St. 9 followed by St. 10 with total counts of 4169 and 3840 unit/L, respectively. This may be due to their subject to fractions of petroleum hydrocarbons and sewage discharge, which could promote the phytoplankton growth as reported by Nayar et al. 2005 and Nassar et al. 2014. On the other hand, the lowest occurrence of phytoplankton was recorded at St. 1 and St. 3, with similar total counts of 2218 and 2251 unit/L, respectively. This may be due to the effect of thermal waters discharged from the cooling systems of the Electrical Power Station of Ayon Mousa near St. 1, as well as the high tourist and human activities at the beach of Ras Sudr near St. 3.

As all marine coastal areas, diatoms were the dominant group forming high percentage (67.48 %) and prevailed during the four seasons (53.03–82 %) and at all stations (57–80 %). Whereas, the cyanophytes were the second dominant group, indicating the presence of freshwater discharge in the study area. On the other hand, the contribution of three groups Cyanophyceae, Dinoflagellates, and Chlorophyceae increased during summer at high temperature (28.1–31.5 °C). Eker and Kideys (2000) suggest that there is a positive relationship between dinoflagellates and water temperature;
thus, dinoflagellates may be better adapted to the high temperatures. Most dinoflagellates are found in temperate waters, are most prevalent in summer months (Taylor 1987), and dominate the phytoplankton in warm seasons (Tait 1981). In this connection, Schabhüttl et al. (2012) reported that green algae and diatoms showed a trend to perform better at lower temperatures, while Cyanobacteria showed stronger responses with increasing temperatures in mixed communities. In the present study, temperature was negatively correlated with diatoms \((r=-0.51)\) and was positively correlated with chlorophytes \((r=0.38)\), whereas it was not a limiting factor for dinoflagellates and cyanophytes.

However, the dominant species during this study were *C. lorenzianus*, *G. attenuatum*, *P. calcari-avis* in addition to the green alga *P. trabecula*, which appeared at all stations and during all seasons, except winter, indicating the freshwater discharge. Moreover, there were dominant potentially harmful algae, and they appeared frequently and were *P. alata* var. gracillima (Özman-Say and Balkis 2012), *P. pungens* (IOC, Casteleyen et al. 2008), and the two cyanophytes *T. erythraeum* and *P. limnetica* (Ramos et al. 2005).

Whereas, other potential harmful algae appeared but less frequently or occasionally, such as *Chaetoceros* spp. (Malone 2007), *Cylindrotheca closterium*, *Cylindrus minimus*, *Skeletonema costatum* (Ismael 2014), *N. longissima*, *Odentella aurita*, *Thalassionema nitzschioïdes*, *Ceratium furca*, *C. lorenzianus*, *C. tripos* (Özaman-Say and Balkis 2012), *C. meneghiana*, *C. gracillima* (Tilstone et al. 2010), and *Prorocentrum micans* (Whilm and Doris 1966). The effect of these species is different, as water coloration and foam or mucilage production (Méndez and Gracia 2002), clogging the fish gills (Malone 2007), and foam or mucilage production (Méndez and Gracia 2002), clogging the fish gills (Malone 2007), indicating the freshwater discharge. Moreover, there are dominant potentially harmful algae, and they appeared frequently and were *P. alata* var. gracillima (Özman-Say and Balkis 2012), *P. pungens* (IOC, Casteleyen et al. 2008), and the two cyanophytes *T. erythraeum* and *P. limnetica* (Ramos et al. 2005). Where these other potential harmful algae appeared but less frequently or occasionally, such as *Chaetoceros* spp. (Malone 2007), *Cylindrotheca closterium*, *Cylindrus minimus*, *Skeletonema costatum* (Ismael 2014), *C. lorenzianus*, *C. tripos* (Özaman-Say and Balkis 2012), *C. meneghiana*, *C. gracillima* (Tilstone et al. 2010), and *Prorocentrum micans* (Whilm and Doris 1966). The effect of these species is different, as water coloration and foam or mucilage production (Méndez and Gracia 2002), clogging the fish gills (Malone 2007), indicating the freshwater discharge. Moreover, there were dominant potentially harmful algae, and they appeared frequently and were *P. alata* var. gracillima (Özman-Say and Balkis 2012), *P. pungens* (IOC, Casteleyen et al. 2008), and the two cyanophytes *T. erythraeum* and *P. limnetica* (Ramos et al. 2005). Whilm and Doris (1966) mentioned that diversity less than 1 indicates instability or heavy pollution, whereas value exceeding 3 indicates stability or clean water. Accordingly, the stations of study area can be considered stable, where all values were closed to 3 or >3, except St. 7 during summer (2.35), which may be attributed to the low number of the recorded species (19 spp.) and the dominance of only three species: *P. trabecula* (37.79 %), *P. alata* var. gracillima (11.34 %), and *P. limnetica* (6.73 %).

The similarity index based on spatial and temporal fluctuations of phytoplankton counts revealed five clusters: cluster 1 (St. 1, 3, and 12), cluster 2 (St. 10), cluster 3 (St. 4, 6, and 9), cluster 4 (St. 2, 8, 5, 7, and 11). The lowest similarity level was 41.11 % between St. 1 and St. 10, whereas the highest level was 76.91 % between St. 3 and St. 12. However, cluster 2, which includes only one station, reflects the unique ecological conditions at this station (oil effluents from Petropil Company). In contrast, the other clusters indicated that the included stations in each cluster have more or less correlated with nitrate \((r=0.49)\). The diatom species *P. calcari-avis* was positively correlated with nitrate \((r=0.40)\) and negatively correlated with ammonium \((r=-0.49)\), and *P. pungens* was negatively correlated with temperature and pH \((r=-0.45\) and \(-0.43)\), respectively, and positively correlated with phosphate and nitrate \((r=0.63\) and \(r=0.52)\), respectively. However, the cyanophyte *P. limnetica* was influenced only by temperature \((r=0.50)\). On the other hand, *P. trabecula*, *P. var. gracillima*, and *T. erythraeum* were not affected by any of these physicochemical parameters.

Marine systems are highly dynamic, with biodiversity changing at seasonal and inter-decadal timescales (Nicholas et al. 2010). The relationship between diversity and productivity has been an object of extensive research for both terrestrial and aquatic ecosystems, and the global diversity patterns observed for marine phytoplankton show a unimodal relationship with productivity using phytoplankton biomass as surrogate (Irigoien et al. 2004). High diversity leads to greater community stability and productivity and makes the system less susceptible to invasions Tilman (1999). In this trend, Friedly (2001) found that diversity was positively related to ecosystem stability, whereas unstable ecosystem will be more likely losses diversity. Meanwhile, Whilm and Doris (1966) mentioned that diversity less than 1 indicates instability or heavy pollution, whereas value exceeding 3 indicates stability or clean water. Accordingly, the stations of study area can be considered stable, where all values were closed to 3 or >3, except St. 7 during summer (2.35), which may be attributed to the low number of the recorded species (19 spp.) and the dominance of only three species: *P. trabecula* (37.79 %), *P. alata* var. gracillima (11.34 %), and *P. limnetica* (6.73 %).
similar ecological conditions, depending on the degree of similarity.

Conclusion

The study area is considered as oligotrophic and healthy, despite land-base, human, and tourism activities, since the total counts of phytoplankton was low accompanied with low nutrient concentrations and high values of diversity. But due to the appearance of potentially harmful algae species even in low counts, make the region of the eastern coast susceptible to drastic effects at flourishing of these species during favorable conditions. Thus, monitoring continuously of this area is imperative, to follow probable bloom of these species, to predict negative effects resulting from increasing land-based activities in order to protect the eastern coast of Suez Gulf from any undesirable change there.

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