Functional analysis of microRNAs in human hepatocellular cancer stem cells

Fanyin Meng, Shannon S. Glaser, Heather Francis, Sharon DeMorrow, Yuyan Han, Jenna D. Passarini, Allison Stokes, John P. Cleary, Xiuping Liu, Julie Venter, Preetham Kumar, Sally Priester, Levi Hubble, Dustin Staloch, Jay Sharma, Chang-Gong Liu, Gianfranco Alpini

Department of Medicine and Scott & White Digestive Disease Research Center, Texas A&M Health Science Center College of Medicine and Scott & White Hospital, Temple, TX, USA
Research Division, Celprogen Inc., San Pedro, CA, USA
Research & Education, Scott & White Hospital, Temple, TX, USA
Department of Experimental Therapeutics, Division of Cancer Medicine, University of Texas M.D. Anderson Cancer Center, Houston, TX, USA
Research, Central Texas Veterans Health Care System, Temple, TX, USA

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Abstract

MicroRNAs are endogenous small non-coding RNAs that regulate gene expression and cancer development. A rare population of hepatocellular cancer stem cells (HSCs) holds the extensive proliferative and self-renewal potential necessary to form a liver tumour. We postulated that specific transcriptional factors might regulate the expression of microRNAs and subsequently modulate the expression of gene products involved in phenotypic characteristics of HSCs. We evaluated the expression of microRNA in human HSCs by microarray profiling, and defined the target genes and functional effects of two groups of microRNA regulated by IL-6 and transcriptional factor Twist. A subset of highly chemoresistant and invasive HSCs was screened with aberrant expressions of cytokine IL-6 and Twist. We demonstrated that conserved let-7 and miR-181 family members were up-regulated in HSCs by global microarray-based microRNA profiling followed by validation with real-time polymerase chain reaction. Importantly, inhibition of let-7 increases the chemosensitivity of HSCs to sorafenib and doxorubicin whereas silencing of miR-181 led to a reduction in HSCs motility and invasion. Knocking down IL-6 and Twist in HSCs significantly reduced let-7 and miR-181 expression and subsequently inhibited chemoresistance and cell invasion. We showed that let-7 directly targets SOCS-1 and caspase-3, whereas miR-181 directly targets RASSF1A, TIMP3 as well as nemo-like kinase (NLK). In conclusion, alterations of IL-6- and Twist-regulated microRNA expression in HSCs play a part in tumour spreading and responsiveness to chemotherapy. Our results define a novel regulatory mechanism of let-7/miR-181s suggesting that let-7 and miR-181 may be molecular targets for eradication of hepatocellular malignancies.

Keywords: microRNAs (miRNAs) • human hepatocellular cancer stem cells (HSCs) • Interleukin-6 (IL-6) • Twist • chemotherapy • invasion

Introduction

Cancer stem cells (CSCs) are key players in the progression of cancer growth [1]. Present chemotherapy modalities eliminate the bulk of tumour cells, but cannot eliminate a core of these CSCs that have a high capacity for renewal [2]. Identification of these cells is the first step in the development of novel and improved therapeutic modalities [3].

miRNAs are small non-coding RNAs that are well suited to suppress previously active programs and, thereby, provide robustness to stem cell differentiation and transformation [4–7]. miRNAs identify their targets by base pairing of the seed sequence of the miRNA with complementary binding within the target mRNA's 3' untranslated region (UTR). This targeting is carried out in synchronization with the RNA-induced silencing complex (RISC), and often results in either degradation or
translational inhibition of the targets [8]. Recent studies have demonstrated that the microRNA pathway is essential for controlling self-renewal, differentiation and transformation in normal and CSCs [9–12].

Hepatocellular cancer (HCC) is the fifth most common cause of cancer and the third leading cause of cancer death worldwide [13]. Majority of the cases (>80%) are not curable because they are diagnosed at advanced stages. Progress in the identification of CSCs in solid tumours including those in GI malignancies has prompted the strong belief that HCC is a stem cell disease [14]. Studies have demonstrated that inflammatory cytokine IL-6 alters miRNA expression in human cancer cells [15,16]. Expression analysis of HCC tumours showed marked activation of the IL-6 pathway, suggesting that HCC may develop from IL-6-driven CSCs with disrupted TGF-β signalling. Besides IL-6, the basic helix-loop-helix transcription factor Twist, an organizer of the epithelial mesenchymal transition (EMT), is a marker for HCC. Twist has been found to be correlated with metastasis in various cancers including human HCC [17]. Several microRNAs have been demonstrated to be the key mediators in Twist-regulated tumour metastasis [18]. However, the specific roles of microRNAs in HCC stem cell (HSC) development and progression, especially characteristic changes during cancer metastasis and chemo-resistance needs to be addressed. We postulated that specific transcriptional factors regulated miRNAs may contribute to tumour spread and chemoresistance by modulating the expression of gene products involved in phenotypic characteristics of CSCs such as cell migration, invasion and response to chemotherapy.

Material and methods

Cell lines and cultures

Human hepatocellular CSCs and parental HCC cells were provided by Celprogen Inc. (San Pedro, CA, USA). Three groups of HSCs were chosen for the studies from seven sets on the basis of self-renewal and real-time polymerase chain reaction (PCR) tests for the specific markers (Fig. 1). The selected group of cells was cultured in Human Liver Cancer Stem Cell Undifferentiated Media to keep the stem cell phenotype (HSC-SR), and in Human Liver Cancer Stem Cell Differentiated Media for 10 passages for differentiated progenitor cell phenotype (HSC-DF). Malignant HCC cells (HepG2) were obtained from the American Type Culture Collection (Manassas, VA, USA) and cultured as recommended by the supplier. Normal human liver stem cells were also obtained from Celprogen Inc.

miRNA array hybridization and analysis

RNA was extracted using Trizol reagent (Invitrogen, Carlsbad, CA, USA). Total RNA (5 μg) was reverse transcribed using biotin end-labelled random octamer oligonucleotide primers. Hybridization of biotin-labelled complementary DNA was performed using a custom miRNA microarray chip (nRNA Program at Center for Targeted Therapy, M.D. Anderson Cancer Center, USA). The data from three samples for each cell type were further analysed by the BRB-ArrayTools software from National Cancer Institute.

Real-time PCR assays for mature miRNAs

microRNA was isolated as described previously [16] and the expression of specific mature miRNAs was confirmed using real-time PCR analysis using a TaqMan Human MicroRNA Assay kit (Applied Biosystems, Foster City, CA, USA).

Northern blot analysis

The Northern blot analysis was performed using MiRNA Northern Blot Assay Kit obtained from Signosis, Inc. (Sunnyvale, CA, USA).

Flow cytometry analysis and FACS sorting Oct-4 and CD133+ cells

HCC stem cells were collected and stained with PerCP-Cy5.5 and/or phycoerythrin (PE) tagged antibodies for 30 min. The staining was analysed by flow cytometry FACSCalibur (Becton Dickinson, San Jose, CA, USA). Oct-4 and CD133+ cells were sorted by DAKO cytomation (DAKO, Carpinteria, CA, USA) [19].

In vitro proliferation, migration, invasion and sphere formation assay

Commercial available kits were used for proliferation, migration, invasion and sphere formation assay in normal and malignant hepatic cells (details in Supporting Information).

Western blotting

Protein was extracted from cultured cells or homogenized tissues, and Western blot was performed as previously described [16]. Please see the Supporting Information for more details.

Xenograft model

Eight- to ten-week-old male SCID Berge mice were obtained from Charles River Laboratories (Wilmington, MA, USA) and fed food and water ad libitum. The mice were maintained in accordance with the Institutional Animal Care and Use Committee (IACUC) procedures and guidelines. HSC-SF (1000) and parental HCC cells (5 × 10^6 cells) were suspended in 0.25 ml of extracellular matrix gel, and the mixture was injected subcutaneously into the right and left flanks. At day 10 the tumour was
Characterization of Cancer Stem Cells from human HCC tissues. (A) Real-time analysis of Oct 4, CD 133, Nestin, telomerase, SSEA-4, AFP and CEA mRNA expression in HCC stem cell under self-renewal condition (HCC-SR), under differentiation (HCC-DF) and HepG2 HCC cells. All the markers are significantly up-regulated in HCC cancer stem cells. *P < 0.05 when compared with HepG2 group. (B) FACS analysis of HCC stem cells on day 1 before cell sorting. Results are given as the percentage of Oct-4 and CD133 cells in the total population. In the histograms, the red line represents staining with Oct-4 or CD133 mAb, and the green lines represent the isotype control-matched mAb. (C) Cell lysates were obtained and immunoblotting analysis was performed for Oct-4, CD133, -fetoprotein (AFP), CK-7, CK-19 and EpiCam using specific antibodies. (D) StemTAG™ Alkaline Phosphatase assays performed at passage 4–6 HSCs and LSCs. HSC-SR and LSCs are maintained in an undifferentiated stage on specific culture dishes, as indicated by the high AP activity, thus confirming the self-renewal within these cells. (E) Formation of tumour spheres in HCC stem cells. In 1% FCS culture media 70–90% of the HSC-SR and HSC-DF became adherent after 7 days, with a minority of floating cells forming spheres composed of three to five cells. After an additional 7 days, the floating spheres were expanded to contain 15–20 HSC-SR cells, with bright appearance and sharp edge. (F) Immunocytochemistry for CD133 and AFP was performed in Normal liver stem cells (N-LSC), HepG2 cells, hepatocellular cancer stem cells growing under self-renewal conditions (HCC-SR) and differentiation conditions (HCC-DF). An increase in CD133 expression along with the enhanced expression of AFP is observed in HCC-SR and HCC-DF cells.
visible, and then the doxorubicin treatment was started and the mice received the desired concentration of the drug intraperitoneal three times per week for 14 days. At the end of the 14 days, the tumour was dissociated into single cell suspension and assayed with Almar Blue to determine percent inhibition of the drug. For analysis of chemotherapymeasured miRNA and mRNA expression in vivo, the xenografts were excised, and tissue was homogenized. An aliquot of the lysates was used for miRNA isolation as described earlier and the remainder used for mRNA expression studies.

**Statistical analysis**

Data are expressed as the mean ± S.E. from at least three separate experiments performed in triplicate, unless otherwise noted. The differences between groups were analysed using a double-sided Student’s t-test when only two groups were present and the null hypothesis was rejected at the 0.05 level unless otherwise specified. Please see Supporting Information for more detailed information of this section.

**Results**

**Purification and characterization of pluripotency and self-renewal capacity in HCC stem cells**

The HCC stem cells (HSCs) obtained from Celprogen Inc. were originally isolated from liver cancer tissues using Oct-4, CD133, Nestin, telomerase, SSEA-4, alkaline phosphatase, α-fetoprotein (AFP), carcino-embryonic antigen (CEA) as positive markers [3,14,20]. Three groups of HSCs were carefully chosen from seven sets based on the self-renewal and real-time PCR tests for the specific markers. The enhanced expression of Oct-4, CD133, Nestin, telomerase, SSEA-4, AFP and CEA were verified by real-time PCR analysis using different primer sets (Fig. 1A). HSCs were further purified by flow cytometry after two culture passages in undifferentiated medium. We directed our purification for cells expressing two of eight CSC markers provided by the company: Oct-4 and CD133. Both Oct-4 and CD133 have been shown to be expressed in liver cancer as well as embryonic stem cells. HSCs were enriched into a CD133+/Oct-4− fraction of Oct-4+ cells (Fig. 1B). To determine the self-renewal properties of HSCs, we utilized two approaches using three cell lines—HSCs, HepG2 HCC cells and normal human liver stem cells (N-LSC) that are isolated from adult human liver tissue using AFP, CD34 and alkaline phosphatase as specific markers [21]. In the first approach, we performed western blotting, all of which showed that both HSCs growing under self-renewal conditions (HSC-SR) and differentiation conditions (HSC-DF) had substantially higher levels of the representative markers of self-renewal than the HepG2, which includes CK-19 and newly discovered malignant hepatic stemness marker EpCAM [20] (Fig. 1C and F). Alkaline phosphatase assays, performed to measure self-renewal within these cells (Fig. 1D), revealed that the levels of self-renewal in HSCs are 12 ± 3 times higher than HepG2 cells, along with the significant enhanced sphere formation ability in HSCs (Fig. 1E). Furthermore, the levels of self-renewal in HSCs under differentiation conditions were less than half of that in HSCs under self-renewal conditions.

To provide other potential mechanisms to explain the CSC effect of HCC progression, we performed real-time PCR-based superarray analysis. The Human Cancer & Estrogen Receptor Signalling PCR Array as well as Cancer Pathway Finder PCR Array were chosen because it contains 168 key genes (84 × 2), representing the 10 cancer-related biological pathways. The array groups also included genes associated with cancer cells’ invasion and response to chemotherapy. We found that 7 of 168 genes were dramatically up-regulated in HSCs related to HepG2 controls, whereas we could also detect several down-regulated genes. The two most up-regulated genes are IL-6 and Twist (Fig. 2A and B). Up-regulation of those genes was also evaluated by reverse transcriptional real-time PCR, which confirmed array data (Fig. 2C). Activated Stat3 was found in HSC cells accompanied by reduced level of TIMP-3. Interestingly, mesenchymal marker N-Cadherin (CDH2) was also overexpressed in HCC stem cells (Fig. 2D). Furthermore, the human liver cancer tissue array showed the signals intensity was strong (+++) or positive (+ +) in 20 of 24 HCC tissues for IL-6 and 17 of 24 HCC tissues for Twist (Fig. S1).

**miRNAs are differentially expressed in human hepatocellular cancer stem cells**

We then compared miRNA expression between HCC CSCs and HepG2/N-LSCs. The pattern of miRNA expression in HCC CSCs was markedly different from that of either normal human liver stem cells or the HepG2 HCC cells (Fig. 3A). The expression of a group of miRNA was differentially increased in HSC cells. These include the human let-7 and miR-181 family members, all of which were expressed >4.0-fold in HSC-SR and HSC-DF CSCs compared with HepG2 HCC cells as well as normal liver stem cells (Fig. 3B). Aberrant expressions of let-7a, let-7b and miR-181a were verified by Northern blot (Fig. 3A) as well as real-time PCR analysis (Fig. 3B). Furthermore, silencing IL-6 and Twist in HSCs significantly reduced let-7a and miR-181a expression respectfully, but not vice versa (Fig. S2), implicated the specific link between IL-6 and let-7, as well as Twist and miR-181 in tumour stem cell behaviour.

**Resistance of HSCs cells to standard chemotherapy**

HCCs are commonly resistant to treatment because the malignant cells survive chemotherapy [19]. To explore the possible role of CSC cells in the development of chemoresistance, cultured HSC-SR, HSC-DF, HepG2 and N-LSC cells were treated with sorafenib and doxorubicin and the IC50 was analysed by MTS assay. Doxorubicin has been considered to be the most effective single
agent and the multi-kinase inhibitor sorafenib has recently been approved for the therapy of advanced HCC [22]. Sorafenib acts by inhibition of the RAF/MEK/ERK pathway and down-regulation of MCL-1, leading to a disruption of survival signals in HCC cells [22,23]. As shown in Figure 4A–D, IC50s of HSCs in two chemotherapeutic reagents tested are significantly higher than HepG2 HCC cells as well as normal liver stem cells, suggesting that HCC stem cells have greater chemoresistance in vitro. In addition, HSC xenograft tumours in nude mice are more resistant to doxorubicin treatment than the control HCC xenografts (Fig. 4E). Enhanced CD133 and let-7a expressions are observed in doxorubicin-treated HSC xenograft tumours when compared to controls (Fig. 4F and G), further implicating a functional link between CD133/let-7a and tumour stem cell phenotypes.

To identify further the cell subpopulations that contribute to HCC chemoresistance, we studied the chemosensitivity of Oct-4+/CD133− and Oct-4−CD133− tumour stem cell subpopulations. Oct-4−CD133− cells showed much higher expression of IL-6 than Oct-4+CD133− HSCs (Fig. 5A). Because chemotherapy-induced cell killing is generally attributed to mitotic cell death, soft agar clonogenic assays to assess the replicative competence of sorafenib- and doxorubicin-treated subpopulations were performed. Although no significant difference was observed between the two subpopulations in the absence of two reagents, Oct-4+CD133− cells isolated from HSCs were more resistant to sorafenib and doxorubicin treatment than were corresponding Oct-4−CD133− cells (Fig. 5B and C). To determine the role of IL-6 signalling in mediating chemoresistance of HCC subpopulation,
purified Oct-4<sup>-</sup>CD133<sup>+</sup> cells were infected with retroviral siRNA against IL-6 and allowed to grow in Matrigel for 1 week in the presence or absence of sorafenib and doxorubicin. As expected, transduction of the cells with interfering siRNA significantly reduced IL-6 expression (Fig. 5A) and, importantly, diminished chemoresistant colonies (Fig. 5B and C). These results indicate that IL-6 signalling was required for protection of Oct-4<sup>-</sup>CD133<sup>+</sup> HCC stem cells from chemotherapeutics-induced cytotoxicity.

**Let-7a and let-7b contributes to survival signalling by IL-6 in HSCs**

We next evaluated the effect of let-7a and let-7b inhibition on the response to chemotherapy *in vitro*. The effects of the anti-let-7a and anti-let-7b were assessed by cotransfecting with the pRL-Tk let-7a and let-7b firefly luciferase constructs that contains two let-7a and let-7b sites in the 3′-UTR of the firefly luciferase reporter. The significant increases in luciferase activity confirmed the efficacy of anti-let-7a and let-7b under the conditions used for our studies (Fig. 6A and B). Meanwhile, introduction of siRNA to IL-6 in HSC cells also up-regulates the luciferase activity, suggested IL-6 directly enhanced let-7a and let-7b expressions (Fig. 6C). Cytotoxicity in response to diverse agents such as sorafenib and doxorubicin was enhanced by pre-incubation with antisense inhibitors to let-7a and let-7b. IC<sub>50</sub>s to all two reagents have significantly reduced in response to chemotherapy in HSC cells pre-incubated with anti-let-7a when compared with a control inhibitor (Fig. 6D and E). Similarly, an increase in activated caspase-3 expression, the predicted target gene of let-7 family members, was noted in response to anti-let-7a in Western blots from sorafenib-treated HSC cells. Furthermore, anti-let-7a treatment in HCC CSCs has successfully increased SOCS1 expression, another predicted target gene of let-7a, and subsequently reduced Stat3 activity (Fig. 6F). Thus, inhibition of let-7a and let-7b increases chemotherapy-induced apoptosis in HCC CSCs, which may be involved in enhanced expression of Caspase-3 as well as SOCS1-regulated Stat3 pathway.
Oct-4<sup>+</sup>CD133<sup>+</sup> HSCs demonstrate enhanced migration and invasion in vitro

It has been suggested that tumour invasion and metastasis may be mediated by the CSC population [43,44]. We observed human HCC cells and stem cells by light microscopy to examine invasion of the Matrigel. Movements of malignant cells in Matrigel were associated with formation of peripheral ruffles. Compared to HepG2 and N-LSC cells, HSCs had more cells with membrane ruffles, and cells were more invasive in the Matrigel (Fig. 7A and B). In vitro assays were used to compare sorted subpopulations from the perspective of differences in cell migration, and invasion (Fig. 7C and D). Oct-4<sup>+</sup>CD133<sup>+</sup> cells were observed to be significantly more migratory towards serum (Fig. 6C), and significantly more invasive through ECM gel (Fig. 7D) than respective Oct-4<sup>-</sup>CD133<sup>-</sup> cell subsets (P < 0.001).

To determine the effect of Twist overexpression was because of a direct effect on tumour invasion or alternatively was because of an increase in CSCs, we tested the ability of Oct-4<sup>+</sup>CD133<sup>+</sup> cells...
Fig. 5 IL-6 is involved in HCC stem cells survival. (A) Total RNA was extracted and IL-6 expression was assessed by quantitative real-time PCR. *P < 0.05 when compared with Control siRNA group. (B, C) Oct-4<sup>−</sup>CD133<sup>−</sup> and Oct-4<sup>−</sup>CD133<sup>−</sup> HCC stem cells were seeded in 96-well plates (10,000 per well) in specific medium with 20% FBS with sorafenib (B) and doxorubicin (C, 10 µM). Cells were incubated for 7 days in a humidified incubator at 37°C, after which the total number of colonies was labelled by Calcein AM (Molecular Probes, Eugene, OR, USA) and quantified fluorometrically. Colony survival rate of Oct-4<sup>−</sup>CD133<sup>−</sup> and Oct-4<sup>−</sup>CD133<sup>−</sup> HCC stem cells were shown after 7-day exposure to sorafenib and doxorubicin (10 µM) in soft agar. The surrogate cancer stem cell assay has demonstrated that silencing IL-6 significantly reduced transformed HCC cancer stem cell survival rate.
Twist overexpression has been demonstrated to correlate with oncogenic effects of HSCs invasion through different mechanisms [24,25]. Twist transcriptionally regulated gene expression through the promoter region containing an E-Box (a promoter element known to be bound by Twist [25,26]). Interestingly, several binding E-boxes element (CATCTG) known to be essential for Twist transcriptional activity were found in the promoter regions of miR-181 (Fig. 8A). Transfection of siRNA to Twist in HSC cells significantly reduced miR-181a expression by real-time PCR binding assay (Fig. 8B). To further evaluate the contribution of miR-181 to oncogenic effects of HSCs, we assessed the impact of miR-181 inhibition on HSC cell in vitro. Anti-miR-181 inhibitions have significantly reduced miR-181 in HSCs. HSCs displayed an aggressive phenotype, which became elongated, and forming the leading edge pseudopodia. Silencing Twist in HSCs significantly reduced the percentage of cells with invasive phenotypes. Matrigel invasion assay showing the invading capability of normal and malignant liver stem cells with and without Twist silencing. Cell migration and invasion was assessed in Oct-4−CD133+ and Oct-4+CD133− cells with or without Twist silencing. Cell migration was expressed as arbitrary fluorescence units. Cell invasion was assessed in 96-well plates pre-coated with extracellular matrix. RFU is the absolute fluorescence intensity value and FI is the fluorescence intensity after normalization with the standard curve. The mean and standard error from four separate experiments are illustrated. *P < 0.05 when compare with HepG2 and N-LSC groups. *P < 0.05 when compared with siRNA controls.

**miR-181 family members are involved in Twist-dependent effects in HSCs invasion**

Twist overexpression has been demonstrated to correlate with hepatocellular carcinoma metastasis through different mechanisms [24,25]. Twist transcriptionally regulated gene expression through the promoter region containing an E-Box (a promoter element known to be bound by Twist [25,26]). Interestingly, several binding E-boxes element (CATCTG) known to be essential for Twist transcriptional activity were found in the promoter regions of miR-181 (Fig. 8A). Transfection of siRNA to Twist in HSC cells significantly reduced miR-181a expression by real-time PCR binding assay (Fig. 8B). To further evaluate the contribution of miR-181 to oncogenic effects of HSCs, we assessed the impact of miR-181 inhibition on HSC cell in vitro. Anti-miR-181 inhibitions have significantly reduced HSC cell invasion ability by 0.6 ± 0.1-fold. Moreover, the specific miRNA–protein binding assay has demonstrated that Twist–miR-181 binding activity was significantly enhanced in HSC stem cells (Fig. 8C and D). In combination, these studies define an important role for miR-181 as a mediator of the oncogenic effects of Twist on CSC invasion in human HCC.
Identification of the target proteins for let-7 and miR-181

Based on 3′-UTR sequence analysis and prediction algorithms, several proteins can be potentially targeted by let-7 family members. Interestingly, SOCS1 and Caspase-3, two well-characterized genes involved in cancer cell survival, were identified as two such putative targets of almost all the members of let-7 family using the TargetScan (http://targetscan.org/) and miRBase (http://microrna.sanger.ac.uk/) databases. SOCS1 and Caspase-3 (CASP3) are both significantly down-regulated in HCC stem cells (Fig. 1C). To verify that SOCS1 and CASP3 targets of translational regulation by let-7 in HSCs, we performed studies using luciferase reporter constructs containing the let-7 recognition sequence (Fig. 9A, top) from the 3′-UTR of SOCS1 and CASP3 inserted downstream of the luciferase gene. Transfection with let-7a and let-7b mimics decreased reporter activity in HSCs, whereas miR-181a mimic failed to show any effects. However, when these studies were repeated with reporter constructs containing random mutations in the recognition sequence, the effects of reporter deactivation by let-7a and let-7b mimics were abolished (Fig. 9A, bottom). Moreover, the increases in SOCS1 and CASP3 expressions occurred after 3 days in cells incubated with anti-let-7a inhibitor (Fig. 6F). Concomitant with enhanced CASP3 expression, there was a reduction of cell survival (Fig. 6D and E). Taken together, these findings indicate that SOCS1 and CASP3 are the biologically relevant targets of let-7a and let-7b.

Recent studies have shown that TIMP-3 and NLK may be directly targeted by miR-181 [20,27]. miR-181 is also predicted to target tumour suppressor gene RASSF1A. Thus, we postulated that alterations in miR-181 expression could represent a mechanism by which Twist modulates RASSF1A, TIMP3 and NLK expression. Studies were performed to evaluate and experimentally validate RASSF1A, TIMP3 and NLK as the bona fide targets of translational regulation by miR-181a. First, we verified that the RASSF1A, TIMP3 and NLK 3′-UTR is the targets for miR-181s using luciferase reporter constructs containing the shared recognition sequence from the 3′-UTR of RASSF1A, TIMP3 and NLK inserted downstream of the luciferase gene (Fig. 9B, top). Transfection with mimics of miR-181a significantly modulated reporter activity in HCC stem cells (Fig. 9B, bottom). Next, the studies were repeated with random mutations in the shared recognition sequence, which resulted in abolition of the reporter activation by miR-181a mimics (Fig. 9B, bottom). Thereafter, we assessed whether ectopic expression of individual miR-181a and miR-181b sequences induces down-regulation of endogenous RASSF1A, TIMP3 and NLK protein expression. Transfection of HSCs with either miR-181a or miR-181b decreased RASSF1A, TIMP3 and NLK expressions after 3 days (Fig. 8E). Meanwhile, transfection with a mimic to
miR-199a, a miRNA that is regulated in a Twist-dependent manner but which does not have complementary sequences to the RASSF1A, TIMP-3 and NLK 3' UTR, did not significantly alter the expression of RASSF1A, TIMP-3 and NLK with a relative expression of 1.06 ± 0.23-fold, 0.91 ± 0.21-fold and 1.02 ± 0.15-fold of controls, respectively. In summary, these data confirm that RASSF1A, TIMP-3 and NLK are the biologically relevant targets of modulation by let-7a and let-7b (miR-181a).
Discussion

Hepatocellular carcinoma is a complex genetic disease caused by the accumulation of mutations that lead to the dysregulation of gene expression and to uncontrolled cell proliferation [28]. miRNAs can contribute to hepatic oncogenesis, functioning as tumour suppressors (e.g. miR-122a and miR-125a), or as oncogenes (e.g. miR-21, the miR221 and miR-143). Consequently, events activating or inactivating miRNAs are now viewed as cooperating with abnormal protein-coding genes in human hepatic tumourigenesis [29]. Our comprehensive investigation of IL-6 and Twist-dependent miRNAs as both potential identification markers and therapeutic agents in HCC stem cells has ultimately filled important gaps in our knowledge of the mechanisms of CSCs in hepatocellular malignancies and may perhaps address critical barriers to progress in the treatment of highly chemoresistant and metastatic tumours in HCC patients.

Expression analysis of HCC tumours showed marked activation of IL-6 pathway, suggesting that HCC may develop from IL-6-driven CSCs with disrupted TGF-β signalling [30]. Suppression of IL-6 signalling through the generation of mouse knockouts with a positive IL-6 regulator, inter-α-trypsin inhibitor heavy chain 4, resulted in a reduction of HCC in elf mice [30], thereby demonstrating a link between two major signalling pathways in the development of cancer and suggesting a possible therapeutic target. Other studies have found overexpression of IL-6 in chemotherapy-resistant breast, lung, leukaemic and colonic cancer cells [31]. Because IL-6 act as an anti-apoptotic ligand during malignant transformation, and is overexpressed in liver CSCs, this could be the reason why liver CSCs are resistant to chemo- and radiation therapy resistance. In addition, Twist, an organizer of EMT and an HCC marker, has been found to regulate the expression of specific miRNA cluster [18,26]. The identification of alterations in transcriptional-regulated miRNA expression in hepatocellular CSCs is likely to be essential to understand the mechanisms and role of altered gene expression and their mediated 'stemness' in human malignancies.

Different from the normal pluripotent state, initiation of oncogenic cellular differentiation is associated with aberrant expression of miRNA populations [32,33]. Multiplicity (one miRNA targeting more than one gene) and cooperation (one gene targeted by several miRNAs) have been demonstrated recently in normal and tumour stemness. There is increasing evidence for autoregulatory interactions involving let-7 and miR-181 in the signalling pathways that control stem cell maintenance and differentiation. Inhibition of the let-7 family has been demonstrated to promote de-differentiation of somatic cells to induced pluripotent stem cells [10]. In ES cells, the pluripotency factor Lin-28, a target gene for let-7, acts as a specific inhibitor of let-7 nuclear and cytoplasmic processing [34–37]. Lin-28 interaction with the Let-7 precur- sor loop mediates regulated microRNA processing. Within the biogenesis pathway, Dicer was recently shown to be another let-7 target [38], a mechanism proposed to account for the regulation of more than 1000 proteins upon let-7 overexpression in cancer cells [39]. Therefore, aberrant expressions of let-7 family in CSCs may promote the malignant transformation and creation of oncogenic phenotypes. We now report that Socs1 and Caspase-3, two let-7 target genes specific to hepatocellular CSCs, encodes a negative inhibitor of the miRNA pathway. Our results show that Socs1 and Caspase-3 are the direct targets that mediate the survival of CSCs.

miR-181s are components of the molecular circuitry that controls vertebrate development. The roles of miR-181 in cellular...
The identification of specific miRNA gene targets that are involved in tumour cell stemness such as SOCS1, Caspase-3, RASSF1A, NLK, TIMP3: elucidation of the mechanisms by which they are regulated; and their functional effects will provide potential targets to reduce chemoresistance and metastasis. Knowledge of specific processes that are regulated in a miRNA manner and identification of critical targets for individual miRNA in HCC stem cells will yield useful information and novel insights into the mechanisms of tumourigenesis, chemoresistance as well as metastasis in HCC and other cancers.

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Conflict of interest

The authors confirm that there are no conflicts of interest.

Supporting information

Additional Supporting Information may be found in the online version of this article:

Fig. S1 Aberrant expression of IL-6 and Twist in human HCC tumour sections.

Fig. S2 IL-6 and Twist specifically regulated let-7 and miR-181 expressions in HCC stem cells.

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