Persistence of Nasopharyngeal Pneumococcal Vaccine Serotypes and Increase of Nonvaccine Serotypes Among Vaccinated Infants and Their Mothers 5 Years After Introduction of Pneumococcal Conjugate Vaccine 13 in The Gambia

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Background. The widespread use of pneumococcal conjugate vaccine (PCV) has brought about a dramatic decrease in pneumococci of vaccine serotypes (VTs) but nonvaccine serotypes (NVTs) have emerged.

Methods. We conducted a cross-sectional survey (CSS) among infants who received 3 doses of 13-valent PCV (PCV13) and their mothers 5 years (CSS3) after PCV13 introduction. Nasopharyngeal swab samples were collected and cultured for isolation of Streptococcus pneumoniae. Whole-genome sequencing of the nontypeable strains was performed. Data were compared with those from 2 previous surveys conducted before PCV13 introduction (CSS1) and 1 year later (CSS2).

Results. Among infants, VT carriage decreased from 33.3% (113/339) in CSS1 to 11.4% (40/351) in CSS3 ($P = .001$) while NVTs increased from 53.1% (180/339) in CSS1 to 74.4% (261/351) in CSS3 ($P < .001$). Among mothers, there was a significant decrease in VTs between CSS2 8.4% (29/347) and CSS3 5.6% (19/342) ($P = .006$). NVTs increased from 16.6% (55/331) in CSS1 to 32.2% (110/342) in CSS3 ($P < .001$). In CSS3, the most prevalent VTs were 7F in infants and 3 in mothers, and the most prevalent NVTs were serogroup 16 and nontypeables, respectively. Genomic analysis showed that VTs were more likely than NVTs to lose their ability to express the capsule.

Conclusions. Five years after PCV13 introduction, we show both direct (infants) and indirect effects (mothers) of the vaccine, while NVT replacement has occurred in both groups. Ongoing circulation of VTs warrants further study of their relevance in any consideration of a reduced dose schedule.

Keywords. pneumococcal carriage; serotypes; herd-effect; nontypeable.

Prevention of pneumococcal disease remains a major global health priority owing to the high disease burden and associated mortality, especially among children in developing countries [1]. A 7-valent vaccine (pneumococcal conjugate vaccine [PCV] 7) was licensed in 2009, but higher-valency formulations (10-valent PCV and 13-valent PCV [PCV13]) have since been licensed [2]. The World Health Organization recommends worldwide PCV introduction, particularly in countries with a mortality rate of >50/1000 live births in children <5 years old [3]. To date, PCVs have been introduced in 59 low- and middle-income countries [4].

The impact of PCVs on pneumococcal disease results from a combination of the direct and indirect effects of the vaccine [5–7]. The latter are a consequence of decreased carriage prevalence among vaccinated individuals and the resulting decrease in pneumococcal transmission within communities [8, 9]. The overall vaccine impact on both disease and carriage, however, has been dampened by an increase in serotypes not included in the vaccine formulations, that is, nonvaccine serotypes (NVTs). This phenomenon is known as serotype replacement [10, 11]. Fortunately, the lower virulence of NVTs means there is nonetheless a net reduction in pneumococcal disease after PCV introduction [12, 13]. In addition to serotype replacement, increased capsular switching (genetic recombination of the...
proteins on the bacterial capsule) in response to vaccine pressure has been reported [14].

In The Gambia, where PCV7 was introduced in 2009 and replaced by PCV13 in 2011, Mackenzie et al [15] reported an 82% reduction in vaccine serotype (VT) invasive pneumococcal disease (IPD) and a 47% increase in NVT IPD 2 years after PCV13 introduction. These data are consistent with data from 2 successive carriage surveys conducted before PCV13 introduction (2 years after PCV7 and 1 year before PCV13 introduction), which showed a decrease in PCV13 VT carriage in infants but not their mothers (ie, no herd effect). Despite the decrease in VTs, a third of all pneumococcal isolates were VTs [16]. Interestingly, an increase in the prevalence of carriage with nontypeable pneumococci, the majority of which had lost their capsule, was also observed [16].

The increase in NVT carriage and the continued presence of VTs after PCV, both in carriage and disease, are a cause for concern. To continue monitoring the direct and indirect effects of PCV13 among vaccinated infants and their unvaccinated mothers on overall VT and NVT carriage, and to explore vaccine impact on nontypeable strains, we conducted a third pneumococcal carriage survey, 5 years after PCV13 introduction.

METHODS

Study Setting

The study was carried out at 2 government health facilities that offer maternal and child health services with regular Expanded Programme on Immunisation clinics. PCV13 was introduced without a catch-up in a 3+0 schedule with doses given at 2, 3 and 4 months. The World Health Organization/United Nations Children’s Fund (UNICEF) immunization data reported >95% coverage of PCV13 dose 3 in 2016 [17].

Study Design

We conducted a cross-sectional survey (CSS) 5 years after PCV13 in 2016 (CSS3). The results were compared with historical data from 2 earlier surveys conducted before PCV13 introduction in 2011 (CSS1) and 1 year after PCV13 introduction in 2012 (CSS2) [16]. The study design, recruitment sites, entry criteria, sample collection, and laboratory methods were similar, and all 3 surveys were conducted between March and June. In all 3 surveys, we recruited healthy infants 6–12 months of age and their mothers at the Expanded Programme on Immunisation clinics. Infants were recruited only if they had received 3 doses of PCV13, ≥1 month before recruitment and were accompanied by their biological mother. We collected information on demographics and risk factors for carriage by interviewing the mothers. Nasopharyngeal swab samples were collected from the mothers and their infants. Written informed consent was obtained from the mothers. The study was approved by the joint Medical Research Council–Gambia (MRCG) Government Ethics Committee.

Nasopharyngeal Swab Sample Collection

A calcium alginate swab was passed gently down the posterior wall of the nasopharynx. The swab was removed and placed in skim milk–tryptone-glucose-glycerol (STGG) transport medium. The STGG vials were taken to MRCG laboratories, as recommended elsewhere [18, 19].

Laboratory Processing

The samples were stored at −70°C and processed as described elsewhere for CSS1 and CSS2 [16]. In brief, 200 µL of thawed, vortexed STGG medium was placed into 5 mL of Todd Hewitt broth containing 5% yeast extract, and 1 mL of rabbit serum. This mixture was vortexed and then incubated at 37°C for 4–6 hours. Subsequently, a 50-µL aliquot was inoculated onto gentamycin blood agar and incubated overnight in 5% carbon dioxide at 37°C for the selective isolation of Streptococcus pneumoniae. The morphologically distinct alpha hemolytic colonies were screened for optochin susceptibility. Serotyping was performed using latex agglutination technique [20]. As in CSS1 and CSS2, serotyping was repeated for all nontypeable isolates.

Whole-genome Sequencing for Nontypeable Pneumococcal Serotypes

Twenty-nine nontypeable isolates from CSS1 and CSS2 were previously analyzed using whole-genome sequencing at the Wellcome Sanger Institute. In the current study, we analyzed a further 23 nontypeable isolates obtained from CSS3.

The sequencing, identification of contaminants, and multilocus sequence typing were done as detailed elsewhere [16]. The phylogenetic tree was reconstructed from single-nucleotide polymorphism sites [21] using RAXML software Version 8 [22]. In silico serotype was determined using the k-mer–based serotyping method, SeroBA [23]. This identified serotypes and generated capsule loci gene assemblies. No assemblies were generated for classic nontypeable pneumococci or isolates that have lost their capsule synthesis genes. Assemblies were annotated using Prokka software (Version 2) [24]. To investigate potential reasons for the noncapsule expression, all capsule loci were compared with reference capsule loci derived from Bentley et al [25] using the Artemis comparison tool [26]. Genes showing significant difference to the reference genes were aligned and visualized using SeaView software (Version 4) to investigate for truncations and mutations that may cause loss of function.

Sample Size Calculation

We targeted 350 infants, for a sample size similar to the those in CSS1 and CSS2. This provided 90% power to detect a 50% reduction in the prevalence of PCV13 VTs and a 25% increase in NVTs compared with CSS2, that, a decrease of PCV13 VT prevalence from 18% to 9% among infants and from 8.1% to 4.1% among mothers, and an increase in PCV13 NVT prevalence from 66.3% to 82.9% among infants and from 16.1% to 20.1% among mothers [16].
Data Management and Analysis
We calculated PCV7 carriage prevalence (serotypes 4, 6B, 9V, 14, 18C, 19F, and 23F), PCV13 carriage prevalence (PCV7 serotypes +1, 3, 5, 6A, 7F, and 19A), and PCV13 NVT carriage prevalence (all other serotypes, including nontypeables). In addition, we calculated the prevalence of the 6 serotypes contained in PCV13 but not in PCV7. We used Poisson regression with robust standard errors to estimate prevalence ratios, comparing both CSS1 and CSS2 with CSS3, and to adjust for potential confounders. We tested the hypothesis that the proportion of PCV13 VTs was the same among "typeables" (ie, isolates of known serotypes) and nontypeables [16] using Fisher exact test. All analyses were done using Stata 14 software (StataCorp).

RESULTS

Characteristics of the Study Participants
We recruited 351 infants and 347 mothers in CSS3. Swab samples were obtained from a total of 1020 mothers and 1040 infants over the 3 surveys (20 mothers had twins). Across the surveys, the median age was 25 years (interquartile range, 21.0–29.3 years) for mothers and 7.9 months (6.8–9.4 months) for infants, and 51.8% of the infants were males. The demographic and epidemiological characteristics of the women and infants were similar across the surveys, except that in CSS1 mothers were less educated and infant antibiotic use was more common (Table 1).

Prevalence of Pneumococcal Carriage Before and 5 Years After PCV13 Vaccinated Infants
Overall carriage was approximately 85% over the 3 surveys (Table 2). VT pneumococcal carriage decreased from 33.3% (113 of 339 infants) in CSS1 to 11.4% (40 of 351) in CSS3 ($P < .001$), while NVT carriage increased from 53.1% (180 of 339) in CSS1 to 74.4% (261 of 351) in CSS3 ($P < .001$). VT carriage decreased further between CSS2 and CSS3 (from 18.3% to 11.4%; $P = .02$), while NVT carriage increased (from 66.9% to 74.4%; $P = .02$) (Table 2). For the 6 serotypes included in PCV13 but not in PCV7, there was a significant decrease in prevalence
between CSS1 and CSS2 (from 23.9% to 13.7%), with an additional drop to 5.4% in CSS3 ($P < .001$).

Among VTs, 19A decreased in prevalence from 8.3% in CSS1 to 0.9% in CSS3 ($P < .001$), and 19F decreased slightly, from 5.6% to 2.8% ($P = .08$). Serotype 6A decreased from 15.3% in CSS1 to 0.0% in CSS3 ($P = .01$). The most prevalent VT in CSS3 was serotype 7F (3.4%), which had not been seen in the previous surveys ($P < .001$). Serotype 21 was the second most prevalent NVT (7.1%), having increased significantly from CSS1 (3.2%; $P = .03$), and there was also a significant increase in the nontypeables, from 0.3% in CCS1 to 3.4% in CSS3 ($P = .02$).

Mothers of Vaccinated Infants

Overall pneumococcal carriage increased from 23.0% (76 of 331 mothers) in CSS1 to 37.7% (129 of 342) in CSS3 ($P < .001$) (Table 3). Although VT carriage was similar in all 3 surveys (6%), NVT carriage increased from 16.6% (55 of 331) in CSS1 to 32.2% (110 of 342) in CSS3 (Table 3).

Serotype 3 was the most prevalent VT in CSS3 (1.8%), followed by 19F (1.2%). Other VT serotypes that were prevalent in CSS1 and CSS2 decreased considerably; 19A decreased from 1.8% in CSS1 to 0.0% in CSS3 ($P = .01$), and 6A from 1.8% in CSS1 to 0% in CSS3 ($P = .01$). Several NVTs increased between CSS1 and CSS3. Notably, serotype 21 increased from 0.9% in CSS1 to 2.9% in CSS3 ($P = .07$), and nontypeables increased from 0.3% in CSS1 to 3.2% in CSS3 ($P = .02$) (Table 3).

### Genotypic Analysis of the Nontypeable Serotypes

A total of 52 phenotypically nontypeable isolates were analyzed, including 3 from CSS1 (2 from mothers and 1 from an infant), 26 from CSS2 (5 from mothers and 21 from infants), and 23 from CSS3 (11 from mothers and 12 from infants). Three isolates were *Streptococcus pseudopneumoniae*—1 from a mother

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**Table 2. Prevalence of Pneumococcal Carriage in Infants Before (CSS1) and 1 (CSS2) and 5 Years (CSS3) After Introduction of PCV13 Into the Gambian Expanded Programme on Immunisation**

| Vaccine Group | CSS1 (n = 339) | CSS2 (n = 350) | CSS3 (n = 351) | Adjusted RR* (95% CI) | P Value | Adjusted RR* (95% CI) | P Value |
|---------------|---------------|---------------|---------------|-----------------------|---------|-----------------------|---------|
| PCV13 VTs     | 33.3          | 18.3          | 11.4          | 0.64 (.44–.93)        | .02     | 0.60 (.50–.71)         | <.001   |
| PCV13 NVTs    | 53.1          | 66.9          | 74.4          | 1.12 (1.01–1.23)      | .02     | 1.19 (1.11–1.26)       | <.001   |
| PCV13 – 7 VTs | 23.9          | 13.7          | 5.4           | 0.40 (.24–.67)        | .001    | 0.23 (.14–.37)         | <.001   |
| PCV7 VTs      | 9.4           | 4.9           | 6.0           | 1.30 (.67–2.53)       | .44     | 0.67 (.31–1.00)        | .051    |
| All serotypes | 85.8          | 84.3          | 85.5          | 1.02 (.96–1.09)       | .46     | 1.00 (.97–1.04)        | .88     |

### Vaccine Group

| Vaccine Group | CSS1 (n = 339) | CSS2 (n = 350) | CSS3 (n = 351) | Adjusted RR* (95% CI) | P Value | Adjusted RR* (95% CI) | P Value |
|---------------|---------------|---------------|---------------|-----------------------|---------|-----------------------|---------|
| PCV13 VTs     | 33.3          | 18.3          | 11.4          | 0.64 (.44–.93)        | .02     | 0.60 (.50–.71)         | <.001   |
| PCV13 NVTs    | 53.1          | 66.9          | 74.4          | 1.12 (1.01–1.23)      | .02     | 1.19 (1.11–1.26)       | <.001   |
| PCV13 – 7 VTs | 23.9          | 13.7          | 5.4           | 0.40 (.24–.67)        | .001    | 0.23 (.14–.37)         | <.001   |
| PCV7 VTs      | 9.4           | 4.9           | 6.0           | 1.30 (.67–2.53)       | .44     | 0.67 (.31–1.00)        | .051    |
| All serotypes | 85.8          | 84.3          | 85.5          | 1.02 (.96–1.09)       | .46     | 1.00 (.97–1.04)        | .88     |

**Abbreviations:** CI, confidence interval; CCS, cross-sectional survey; NA, not applicable; NT, nontypeable; NVTs, nonvaccine serotypes; PCV7, 7-valent pneumococcal conjugate vaccine (PCV7); PCV13, 13-valent PCV; PCV13-7, serotypes present in PCV7 but not in PCV13; RR, risk ratio; VTs, vaccine serotypes.

*aAdjusted for health center, maternal age and education, and infant’s age, sex, and antibiotic intake within 4 weeks of the survey.
in CSS1 and 2 from infants in CSS2 —and 1 was a nonpneumococcal Streptococcus from a mother in CSS3. Fifteen isolates (28.8%) were of the classic nontypeable lineage. The others (33 isolates; 63.5%) were of an encapsulated pneumococcal lineage and had lost their ability to express a capsule through 3 different mechanisms (Figure 1 and Table 4). Eight had a standard capsular polysaccharide synthesis locus but no capsule phenotypically detected, 15 had indels in capsule synthesis genes or complete loss of capsule loci, and 10 had acquired a locus typically found in classically nontypeable isolates.

Based on combined data from CSS2 and CSS3, 8 mothers and 19 infants carried nontypeables that were nonclassic; 7 (87.5%) and 8 (42.1%) were of the VT lineage, respectively. Compared with VT carriage prevalence of 22.6% (49 of 217) in mothers and 17.5% (104 of 595) in infants, this corresponds to a 3.9-fold increase (95% confidence interval, 2.7–5.6; \( P < .001 \)) in mothers and a 2.4-fold increase (1.4–4.2; \( P = .01 \)) in infants.

**DISCUSSION**

Despite the observed decline in VT pneumococcal prevalence in mothers and infants since the introduction of PCV13 in The Gambia, VT pneumococci continue to circulate in both groups. Among infants, the decrease in PCV13 VT carriage and associated increase in NVTs resulted in no change in overall pneumococcal carriage. In contrast, the decrease in VTs in mothers was less pronounced than the increase in NVTs, resulting in a net increase in overall pneumococcal carriage. Our previous observation that the increase in nontypeable pneumococci after PCV is due mainly to serotypes that have lost their capsule was confirmed by data from CSS3.

Although the direct effect of PCV13 on circulating PCV13 VTs among vaccinated infants was observed 1 year after its introduction (CSS2) —5 years later (CSS3) VTs were still circulating in infants who had received 3 doses of the vaccine. The PCV13 VT prevalence in this study (11.4%) is similar to

**Table 3. Prevalence of Pneumococcal Carriage in Mothers Before (CSS1) and 1 Year (CSS2) and 5 Years (CSS3) After Introduction of 13-Valent PCV Into the Gambian Expanded Programme on Immunisation**

| Vaccine Group | CSS1 (n = 331) | CSS2 (n = 347) | CSS3 (n = 342) | Adjusted RR* (95% CI) | \( P \) Value | Adjusted RR* (95% CI) | \( P \) Value |
|---------------|----------------|----------------|----------------|-----------------------|---------------|-----------------------|---------------|
| PCV13 VTs     | 6.6            | 8.4            | 5.6            | 0.58 (0.33–1.02)      | \(.06\)       | 0.92 (0.67–1.27)      | \(.63\)       |
| PCV13 NVTs    | 16.6           | 16.1           | 32.2           | 2.01 (1.51–2.67)      | <.001         | 1.40 (1.20–1.63)      | <.001         |
| PCV13 – 7 VTs | 3.9            | 6.1            | 2.3            | 0.34 (0.15–0.74)      | \(.007\)      | 0.65 (0.28–1.52)      | \(.32\)       |
| PCV7 VTs      | 2.7            | 2.6            | 3.2            | 1.14 (1.46–2.79)      | \(.78\)       | 0.99 (0.40–2.48)      | \(.99\)       |
| All serotypes | 23.0           | 24.2           | 37.7           | 1.51 (1.20–1.91)      | <.001         | 1.29 (1.14–1.47)      | <.001         |
| PCV13 VTs     |                |                |                |                       |               |                       |               |
| 1             | 0.3            | 0              | 0              | NA                    | NA            | NA                    | NA            |
| 3             | 0              | 0.3            | 0.1            | 6.09 (1.74–50.38)     | \(.094\)      | NA                    | NA            |
| 4             | 0.6            | 0.6            | 0.6            | 1.01 (0.14–7.17)      | \(.99\)       | 0.98 (0.37–2.62)      | \(.97\)       |
| 5             | 0              | 0.9            | 0              | NA                    | NA            | NA                    | NA            |
| 6A            | 1.8            | 0              | 0              | NA                    | NA            | NA                    | NA            |
| 6B            | 0              | 0              | 0              | NA                    | NA            | NA                    | NA            |
| 7F            | 0              | 0              | 0.3            | NA                    | NA            | NA                    | NA            |
| 9V            | 0              | 0              | 0.3            | NA                    | NA            | NA                    | Na            |
| 14            | 0.6            | 0.3            | 0.6            | 2.03 (1.18–22.31)     | \(.56\)       | 0.98 (0.37–2.62)      | \(.97\)       |
| 18C           | 0.3            | 0.6            | 0.3            | 0.51 (0.05–5.58)      | \(.58\)       | 0.98 (0.25–3.93)      | \(.98\)       |
| 19A           | 1.8            | 3.2            | 0.3            | 0.09 (0.01–7.11)      | \(.02\)       | 0.40 (0.14–1.16)      | \(.09\)       |
| 19F           | 0.6            | 0.3            | 1.2            | 4.06 (4.68–36.19)     | \(.21\)       | 1.39 (0.60–3.24)      | \(.44\)       |
| 23F           | 0.6            | 0.9            | 0.3            | 0.34 (0.04–3.24)      | \(.35\)       | 0.70 (0.21–2.31)      | \(.55\)       |
| PCV13 NVTs    |                |                |                |                       |               |                       |               |
| 10A           | 0.6            | 0.3            | 2.3            | 8.12 (1.02–64.65)     | \(.048\)      | 8.17 (0.91–4.26)      | \(.09\)       |
| 13            | 0.6            | 0.2            | 0.6            | 0.34 (0.17–1.67)      | \(.18\)       | 0.98 (0.37–2.62)      | \(.97\)       |
| 15B           | 0.6            | 0.5            | 1.2            | 2.19 (0.82–6.33)      | \(.54\)       | 1.39 (0.60–3.24)      | \(.44\)       |
| 16            | 2.4            | 0.6            | 2.3            | 4.06 (0.87–19.09)     | \(.06\)       | 0.98 (0.61–1.60)      | \(.96\)       |
| 19C           | 1.2            | 0              | 0              | NA                    | NA            | NA                    | NA            |
| 21            | 0.9            | 0.9            | 0.9            | 2.09 (0.94–12.19)     | \(.06\)       | 2.61 (0.85–8.34)      | \(.07\)       |
| 23B           | 0              | 0.9            | 2.0            | 2.37 (0.62–9.09)      | \(.21\)       | NA                    | NA            |
| 34            | 1.2            | 1.2            | 1.8            | 1.52 (0.43–5.35)      | \(.51\)       | 1.10 (0.41–2.62)      | \(.56\)       |
| 35B           | 1.8            | 0.6            | 1.8            | 3.04 (0.62–14.99)     | \(.17\)       | 0.86 (0.36–2.22)      | \(.95\)       |
| NT            | 0.3            | 1.4            | 3.2            | 2.23 (0.78–6.36)      | \(.13\)       | 3.26 (1.17–9.06)      | \(.02\)       |

Abbreviations: CSS, cross-sectional survey; NA, not available; NT, nontypeable; NVTs, nonvaccine serotypes; PCV7, 7-valent pneumococcal conjugate vaccine (PCV); PCV13, 13-valent PCV; PCV13-7, serotypes present in PCV7 but not in PCV13; RR, risk ratio; VTs, vaccine serotypes.

*Adjusted for health center, maternal age, and maternal education.
VT carriage among Gambian newborns 2 years after routine PCV13 [27] but 2-fold higher than PCV13 VT prevalence in Greenlandic children <5 years old (5%) 3 years after PCV13 [28], Belgian children (5.4%) 5 years after PCV13 and 9 years after PCV7 [29], and Australian aboriginal children <5 years old (5.8%) 3 years after PCV13 and 10 years after PCV7 [30]. We saw a significant increase in serotype 19F after PCV13 introduction which was also among the most common VTs after PCV13 in the Belgian study [29]. We also observed an increase in serotype 7F after PCV13 introduction which was also among the most common VTs after PCV13 in the Belgian study [29]. We also observed an increase in serotype 7F prevalence in CSS3. The increase is surprising because a mathematical model of pneumococcal transmission in The Gambia predicted that PCV13 would eliminate low-transmission VTs, including 7F [31]. A similar increase in 7F was observed in IPD surveillance data after introduction of PCV13 in Australia. However, elsewhere there has been a decline in 7F after PCV13 [32].

Our results suggest a PCV13 herd effect among Gambian mothers of vaccinated infants, particularly for serotypes 6A and 19A; such an effect was not yet apparent 1 year after the introduction of PCV13 (CSS2). However, we have to interpret this decrease in VTs with caution, because the decrease between CSS1 and CSS3 was not significant, and there was an increase between CSS1 and CSS2 driven by the large increase in serotype 19A in CSS2. A cluster-randomized trial conducted in rural Gambia demonstrated a herd effect of PCV [33], as have countries that have introduced PCV7 [34] and countries that have introduced PCV13 with a booster dose [6].

We found that the prevalence of serotype 19A decreased after PCV13 introduction and was low in CSS3 in both mothers and infants. This is in contrast to data from the United Kingdom and Alaska. In the United Kingdom, among the first countries to switch from PCV7 to PCV13, IPD associated with 19A has begun to increase in recent years [35], and in Alaska, PCV13 protection against 19A is waning in older vaccinated children, possibly owing to the emergence of a new genotype. Longer surveillance in our setting is needed to better understand the trend of this serotype in carriage and disease.

In mothers, serotype 3 was the most prevalent serotype in CSS3, and cases of IPD caused by this serotype are still seen in The Gambia [15]. This serotype, which had already decreased before PCV13 introduction, shows secular trends in The
Table 4. Results of Whole-genome Sequencing of Nontypeable Isolates

| Isolate No. | Mother or Infant | S. pneumoniae, % | S. pseudopneumoniae, % | ST | Nearest ST | Capsular Locus Top Hit | Serotype-Specific Sequence Detected | Ancestral Capsular Type From Phylogeny | Conclusion | VT/NVT |
|------------|-----------------|-----------------|-----------------------|----|-----------|-----------------------|-----------------------------------|--------------------------------------|------------|--------|
| CSS1       |                 |                 |                       |    |           |                       |                                   |                                      |            |        |
| 108887     | Infant          | 78.08           | 0.71                  | 3407 | ...       | 16F                   | 16F                               | wzd gene loss and truncated wzh    | NVT        |        |
| 105098     | Mother          | 80.51           | 0.14                  | 1778 | ...       | 34                    | 34                                | Intact capsule synthesis genes; no genetic explanation | NVT        |        |
| 104550     | Mother          | 19.47           | 40.9                  | Unknown | ...       | ...                   | ...                               | S. pseudopneumoniae ...            |            |        |
| CSS2       |                 |                 |                       |    |           |                       |                                   |                                      |            |        |
| 201376     | Infant          | 80.46           | 0.25                  | 5521 | ...       | 10A                   | 10A                              | Intact capsule synthesis genes; no genetic explanation | NVT        |        |
| 207381     | Infant          | 82.09           | 0.4                   | Novel ST D | 2052 | 20       | 20                   | Long branch, inconclusive         | Truncated whaF and wzx genes      | NVT        |        |
| 206628     | Infant          | 81.38           | 0.08                  | 989  | ...       | 12F                   | 12F/A/46                          | Intact capsule synthesis genes; no genetic explanation | NVT        |        |
| 210240     | Infant          | 80.96           | 0.08                  | 989  | ...       | 12F                   | 12F/A/46                          | Intact capsule synthesis genes; no genetic explanation | NVT        |        |
| 210201     | Infant          | 83.51           | 0.08                  | 2447 | ...       | 14                    | 14                               | Truncated glycosyl transferase genes, wcoY and rlp | VT         |        |
| 201867b    | Infant          | 46.64           | 0.57                  | Novel ST B | 4040 | Classic NT | ... | ... | Capsule switch from 14 to classic NT locus | VT         |        |
| 200954     | Mother          | 81.22           | 0.25                  | Novel ST G | 975       | 15B/C                | 15B/C                            | 12-nucleotide insertion at the 5' end of the initial transferase gene, whaA | NVT        |        |
| 206379     | Infant          | 81.51           | 0.16                  | 4033 | ...       | 15B/C                | 15B/C                            | 12-nucleotide insertion at the 5' end of the initial transferase gene, whaA | NVT        |        |
| 207139     | Infant          | 76.55           | 0.57                  | 3407 | ...       | 16F                   | 16F                              | wzd gene loss and truncated wzh    | NVT        |        |
| 210092     | Infant          | 80.13           | 0.28                  | Novel ST E | 847       | 19A                  | 19A                              | 3-nucleotide deletion at the 3' end of the rmlC gene | VT         |        |
| 210344     | Infant          | 79.93           | 0.19                  | Unknown | 7661 | 19B                  | ... | 19B | Intact capsule synthesis genes; no genetic explanation | NVT        |        |
| 206438     | Infant          | 81.8            | 0.1                   | 202  | ...       | 19A                  | 19A                              | Long branch, inconclusive         | 3-nucleotide deletion at the 3' end of the rmlC gene | VT         |        |
| 208478     | Infant          | 81.95           | 0.17                  | 4033 | ...       | 19F                   | 19F/15B/C                         | Intact capsule synthesis genes; no genetic explanation | VT         |        |
| 202731     | Infant          | 81.26           | 0.08                  | Novel ST C | 6712 | 28A/F                | ... | 28A/F | 6-nucleotide insertion in the wze gene | NVT        |        |
| 201297     | Mother          | 81.95           | 0.34                  | 5734 | ...       | 6A/B/C/D              | 6A/B/C/D                         | 12-nucleotide insertion in 3' end of the pseudogene HG262 | VT         |        |
| 201794     | Infant          | 78.82           | 0.8                   | Novel ST A | 71       | Classic NT | ... | ... | Loss of cps locus ... | VT         |        |
| 201398     | Mother          | 76.17           | 0.3                   | Novel ST H | 3582 | Classic NT | ... | ... | Capsule switch to classic NT locus ... | VT         |        |
| 201843     | Infant          | 77.28           | 1.11                  | 4040 | ...       | Classic NT            | ... | 14 | Capsule switch from 14 to classic NT locus | VT         |        |
Several carriage and IPD studies have shown that PCV13 has no direct or indirect effect on serotype 3 [37, 38]. Five years after PCV13, we have observed an increase in NVTs in both vaccinated infants and their mothers. For infants,
we had already observed an increase between CSS1 and CSS2. In CSS3, we detected a significant increase in serotypes 21 and 23B. These were also the most prevalent serotypes in the post-PCV13 era in studies conducted in The Gambia [27], Italy, and Norway [39, 40]. Unexpectedly, among mothers the increase in NVTs exceeded the decrease in VTs, resulting in an overall increase in carriage. Although this large increase is difficult to explain, it probably reflects an upsurge in pneumococcal transmission rather than biased sampling, because similar field and laboratory methods were used in all 3 surveys and recruitment took place during the same time of the year to minimize any effect of season [41].

Nontypeable strains increased in prevalence over the 3 surveys in both vaccinated infants and mothers, which is consistent with the rise in nontypeable strains causing noninvasive and invasive disease after PCV13 introduction in Taiwan [42]. Further whole-genome sequencing analysis of all the nontypeables confirmed our previous hypothesis that VTs are more likely than NVTs to lose their ability to express the capsule after introduction of PCV. These “capsular switches” may be due to the selection of variants that existed in the pre-PCV era. Capsular loss is relatively common for S. pneumoniae, and increased loss due to vaccine pressure has also been reported in other studies [14].

Nontypeable pneumococci are likely to be underreported [43] because they are generally excluded from the analyses in epidemiological studies and vaccine trials, including the ongoing surveillance of the IPD in The Gambia [15]. This may be because their role in disease has been limited. For example, in South Africa over a 10-year period spanning from before to after PCV, and in the United States over a 3-year period in the post-PCV era, nontypeables rarely caused disease [44, 45]. We nonetheless advocate for the inclusion of nontypeable strains in epidemiological surveillance studies owing to their increasing importance and potential for maintaining transmission.

The 3 surveys provide valuable data on the timeliness of vaccination. It is often assumed that vaccination in Africa is substantially delayed and more in line with a 2 + 1 schedule (ie, 2 doses before age 6 months and 1 “booster dose” after 9 months). However, our data suggest that in The Gambia the schedule used is close to 3 + 0 (the median age for PCV dose 3 vaccination was 5 months). Our study therefore represents a genuine evaluation of the 3 + 0 schedule and, as such, contributes to the current debate over the relative merits of the 3 + 0 versus the 2 + 1 PCV schedule [46].

The main limitations of our study were intrinsic to the study design, in that we cannot exclude secular trends in individual serotypes because variability in the prevalence of serotypes was already described before vaccine introduction [47]. In addition, our surveys started only after PCV7 was introduced into routine immunization, and therefore our comparisons were between children vaccinated with PCV7 (CSS1) and those vaccinated with PCV13 (CSS2 and CSS3), capturing only the additional effect of PCV13 over PCV7. There may have been residual confounding, because we did not adjust for the number of siblings, and other studies have shown increased risk of carriage when infants live with other children [27, 47]. However, there was no difference in maternal age between surveys, a potential proxy for number of siblings. Moreover, though we did not collect information on household size in CSS3, there was no association between carriage and household size in CSS1 and CSS2.

In conclusion, we have shown important effects of the introduction of PCV13 into routine immunization in The Gambia, including direct and indirect effects and serotype replacement. Continued disease surveillance and additional carriage surveys are necessary to monitor the persistence of VTs, the emergence of NVTs, and the role of nontypeables in transmission and disease. An alternative vaccination schedule (not necessarily reducing the number of doses) needs to be considered to halt ongoing VT transmission.

Notes

Author contributions. B. K. and A. R. designed the original studies. E. U., C. B., P. C. H., and A. R. critically reviewed the study proposal and made important contributions throughout the study. I. C. and A. B. led the microbiological isolation of S. pneumoniae from the swab samples, and E. B. and R. G. performed the whole-genome sequencing. E. U. coordinated the field work for CSS3, did the analysis, and wrote the first draft of the manuscript. All authors contributed to writing the manuscript and approved the final version.

Acknowledgments. Special thanks to the management of the Jammeh Foundation for Peace and the Sukuta Health Centre, the field staff led by Edrissa Sabally, the junior data manager, Haddy Kanyi, the laboratory technicians, and the mothers and their infants.

Financial support. This work was supported by Medical Research Council–Gambia and the London School of Hygiene and Tropical Medicine (postdoctoral fellowship to E. U.).

Potential conflicts of interest. E. U. served as a consultant for GSK Vaccines Malaria vaccine group (2014–2017). R. G. reports a PhD stipend from Pfizer for studying PCV impact on pneumococcal carriage in the United Kingdom (received 2009–2012). All other authors report no disclosures. All other authors report no potential conflicts. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest. Conflicts that the editors consider relevant to the content of the manuscript have been disclosed.

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