Antigen of 49.6-kDa subunit pilus protein of Helicobacter pylori as a potential biomarker for early and rapid detection of the infection

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Abstract

Background and Aim: Helicobacter pylori infection has been identified as a major cause of peptic ulcer diseases, including gastric and duodenal ulcers, gastritis, chronic gastric carcinoma, and even gastric lymphoma [1,2]. H. pylori, also known as a zoonotic agent, is isolated from cow, sheep, and goat [3], and its antigens were detected in the milk and feces of cows [4]. This is unique because H. pylori is the only bacterium known to cause gastric carcinoma [5-7].

Infectious H. pylori was classified as carcinogen group I for gastrointestinal cancer because it has various virulent factors, such as CagA, VacA, Urease, and ammonia, that are capable of triggering carcinogenesis [5,8-10]. The pathogenic properties of these bacteria were associated with its fimbrial adhesion (pili) [11], a protein found on the bacterial cell surface that plays a role as a bacterial virulence factor [12]. In vitro studies using Western blotting analysis, hemagglutination test, adherence inhibition assays, and immunocytochemical staining revealed that the 49.6-kDa subunit pilus protein of H. pylori was considered an immunogenic protein. This study aimed to develop a serological diagnostic test using 49.6 kDa for detecting antibodies against H. pylori proteins in an early phase of the infection.

Materials and Methods: An in-house immunochromatographic test (ICT) kit was developed and used to test a panel of sera sample obtained from a randomly selected symptomatic patient, in which 40 sera were H. pylori positive and 40 sera were H. pylori negative.

Results: The results showed that ICT with 49.6 kDa as an antigen was highly sensitive and specific for detecting anti-H. pylori immunoglobulin G antibodies in human serum, with a high negative predictive value.

Conclusion: The developed test could be used to exclude H. pylori infection in symptomatic patients.

Keywords: 49.6-kDa pili protein, Helicobacter pylori, immunochromatographic test.

Introduction

Infection by Helicobacter pylori has been identified as a major cause of peptic ulcer diseases (gastric and duodenal ulcers), gastritis, chronic and gastric carcinoma, and even gastric lymphoma [1,2]. H. pylori, also known as a zoonotic agent, is isolated from cow, sheep, and goat [3], and its antigens were detected in the milk and feces of cows [4]. This is unique because H. pylori is the only bacterium known to cause gastric carcinoma [5-7].

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Further study is required to investigate the diagnostic value of this protein in the early detection of infection by this microorganism. Most diseases present high cure rates only when detected early [17]. Therefore, early diagnosis is essential to reduce morbidity and mortality. In addition, early diagnosis of infectious diseases can prevent their development into epidemics. There are several methods to detect H. pylori infection [18,19]. Detection of H. pylori infection by using serological methods is considered to be the easiest, non-invasive approach, which does not require endoscopy to diagnose the infection [20,21]. This method requires only a few drops of blood, producing results in <5 min [22]. There are numerous methods available for the detection of anti-H. pylori immunoglobulin G (IgG), immunoglobulin A, and immunoglobulin M antibodies [23], which are present
in the whole blood, serum, saliva, stool, and urine. The accuracy of the diagnostic markers varies from test to test and among sample types [24,25]. In the present study, we evaluated the performance of a new immunochromatographic test (ICT) kit using 49.6-kDa pili protein.

Materials and Methods

Ethical approval

This research was approved by the Ethical Commission for the Use of Animals in Research and Education of the Faculty of Veterinary Medicine, Udayana University, Indonesia with Ref. No. 284a/KE-PH/VII/2017.

H. pylori strains

Three H. pylori Lombok isolates were provided by the Microbiology Laboratory, Biomedical Research Unit, West Nusatenggara General Hospital, which were isolated from the gastric antral biopsies of Sasak Lombokese patients. The bacterium was first cultured using media Trypticase Soy Agar and Trypticase Soy Broth supplemented with 10% sheep blood, completed with supplement and IsoVitaleX™, and incubated at 37°C on the microaerophilic atmosphere [13].

Isolation of 49.6-kDa subunit pili protein of H. pylori

Isolation of H. pylori pili was performed by the method of Sumarno et al. [26] with a slight modification. Bacteria pili were cut by using a pili bacterial cutter, which was carried out for 30 s at the speed of 5000 rpm and repeated for five times. Subsequently, the isolation of pili fraction by centrifugation of cutting result was done at 12,000 rpm at 4°C. The supernatant containing the bacterial pili was analyzed in sodium dodecyl sulfate-polyacrylamide gel electrophoresis. Protein of 49.6-kDa subunit pili was isolated from the gel by cutting alongside close to the protein position, around 49.6 kDa. The isolated gel was sliced and inserted into the dialysis membrane soaked with phosphate-buffered saline (PBS). Subsequently, the protein of interest was electroeluted by placing the membrane in the negative electrode with current of 20 mA for 15 min. Total protein was measured using a method of detergent-compatible protein assay (Bio-Rad Laboratories Inc, USA), suspended to a concentration of 10 ng/ml, and kept at −20°C until use [13].

Sera panel from dyspepsia patients and vaccinated mice

Human sera

Eighty sera from H. pylori-positive and H. pylori-negative patients based on the culture and direct microscopic examination of mucosal gastric biopsy were used as panel sera. All sera of dyspepsia patients were provided by the Biomedical Research Unit, West Nusa Tenggara General Hospital, and stored at −20°C.

Mouse sera

One hundred healthy male balb/c mice (weight 18–22 g) were used in this study for producing animal sera. The experimental mice were grouped into 10 groups and fed with a portion of commercial food and water ad libitum. All mice were orally given culture of live H. pylori, they had been fasted one night. The H. pylori obtained from a patient with typical gastric ulcer was first cultured with Brain Heart Infusion media supplemented with 5% sheep blood, incubated at microaerophilic condition (10% CO₂, 85% N₂, and 5% O₂) at 37°C for 48 h. The cells were then washed and suspended with sterile PBS at a concentration of 109 cells/ml. The 100 balb/c mice were divided into two groups of A and B, with 50 balb/c mice each. Each animal in Group A was orally infected with 109 cells/ml 3 times every 2 days, based on the method of Marchetti et al. [27] In contrast, all animals in Group B were used as control animals that were orally given sterile PBS containing no H. pylori culture.

Blood samples were collected from both Groups A and B before they were orally infected with the bacterial culture, and 1, 2, and 3 weeks after the infection. The blood samples were collected from the tail and kept at room temperature (RT) for 2–5 h before the sera were collected and stored at −20°C until tested.

Development of IgG H. pylori ICT

A standard ICT strip typically consists of five parts: Sample (polyester) pad, conjugation (polyester) pad, nitrocellulose (NC) membrane, absorption pad, and polyvinyl chloride plastic backed card. All the pretreated parts were assembled sequentially on a plastic backing card with 2-mm overlap of each component. Antigens (49.6 kDa-sub-unit protein pili or secretary antigen [Ag]) were coated separately to serve as test line and goat anti-mouse IgG as control line [28]. Both antigens and antibodies were dispensed by XYZ dispensing system (BioDot Inc, USA) to NC membrane. The Protein A-colloidal gold (CG) conjugate was dispensed onto the polyester pad. After being dried at 37°C, the assembly was cut into a 5-mm-wide individual strip and then stored at the RT inside a sealed plastic bucket with a desiccant until used. When a serum sample is added onto the sample pad, the sample flowed from this component to the absorption pad on the membrane surface by capillary action. Once the sample reached specific positions, the antigen in the sample reacted with the conjugates (labeled with CG). The residual sample continues to move forward and be absorbed in the absorption pad. After the reaction was completed, two lines appeared on the strip: The C-line (control line to confirm whether the strip is valid) and the T-line (test line used to judge the detection results) [29].

Detection of antibodies in patient serum

The ICT cards were removed from the foil pouch and placed on a flat, dry surface. A drop (20–30 μl) of serum was applied on to the sample well. Then, two drops of the buffer were added on to the sample well. As the test began to work, purple color was seen moving across the result window at the center of the
test disk. Test results were interpreted within 20 min. The presence of two-color bands, “T” and “C,” meant that the test was positive, whereas the presence of only one band (only on “C”) was interpreted as negative. If no band or a single band only on “T” was formed after 20 min, the result was considered invalid, and the experiment was repeated [30].

**Results**

**Demographic and geographic characteristics**

We used 80 sera from dyspepsia patients undergoing an endoscopic examination at the West Nusatenggara Province General Hospital. Demographic characteristics of gastric disease patients are shown in Table-1.

The average age of the patients was 48.22 years. The representation per age group was 4 (5.0%) patients under 30 years; 60 patients (75.0%) between 30 and 50 years; and 16 patients (20.0%) >50 years. Considering the age, most of the patients were adult; hence, the diagnosis resulted in chronic *H. pylori* infection [31].

**Antigen 49.6-kDa subunit pili protein of *H. pylori***

The result of Western blotting analysis for detecting the 49.6-kDa subunit pili protein of *H. pylori* is shown in Figure-1.

**Comparison of IgG *H. pylori* detection results**

The presence of specific antibodies in mice which had been infected orally using the culture of live *H. pylori* was demonstrated using a standard ICT. Strong and specific reaction was observed when the subunit protein pili Ag reacted with the mouse sera 3 weeks after the infection. However, a negative reaction was observed when the sera were reacted with the secretory Ag (Figure-1). The prevalence of the serological reaction depended on the time after the infection; the highest prevalence of 96% was observed 3 weeks after the initial infection (Table-2).

Strong positive reaction in B (subunit protein pili Ag) was observed in sera of mice 3 weeks after the infection. No reaction was observed with the same sera in B (secretory Ag) [32].

The presence of specific antibodies against secretory Ag and subunit pili Ag in dyspepsia patients undergoing endoscopic was demonstrated using *in vitro* studies employing ICT. The subunit protein pili reacted 37 out of the 40 sera from an infected human with a specificity of 92.5% (true-positive detection) and detected only 1 out of the 40 sera (2.5%) (false-negative reaction) from the non-*H. pylori*-infected patients. Although the ICT secretory Ag reacted with sera from an infected human with a specificity of 95%, this protein also reacted with 5 out of the 40 sera (12.5%) (false negative) from the non-*H. pylori*-infected patients, suggesting that this protein was less specific than the 49.6 kDa-sub-unit pili protein of *H. pylori* (Table-3).

**Table-1**: Demographic characteristics.

| Characteristics | n (%) |
|-----------------|-------|
| Sex             |       |
| Male            | 52 (65.0) |
| Female          | 28 (35.0) |
| Age (years)     |       |
| <30             | 4 (5.0) |
| 30–50           | 60 (75.0) |
| >50             | 16 (20.0) |
| Ethnicity       |       |
| Sasak (Lomboknese) | 66 (82.5) |
| Balinese        | 12 (15.0) |
| Other           | 2 (2.5) |
| Endoscopy diagnosis |       |
| Chronic gastritis | 58 (72.5) |
| Gastric or DU   | 14 (17.5) |
| GC              | 8 (10.0) |

**Table-2**: Comparison of ICT result between subunit pili Ag and secretory Ag to detect the presence of IgG anti-*Helicobacter pylori* prepared from vaccinated mouse sera.

| Testing result | Post-infected mouse sera (n=50) |
|----------------|----------------------------------|
|                | 1 week | 2 weeks | 3 weeks |
| Secretory Ag ICT |        |         |         |
| Positive         | 0 (0)  | 2 (4)   | 39 (78) |
| Negative         | 50 (100)| 48 (96) | 11 (22) |
| Subunit pili Ag ICT |      |         |         |
| Positive         | 0 (0)  | 17 (34) | 48 (96) |
| Negative         | 50 (100)| 33 (66) | 2 (4)   |

**Table-3**: ICT result of secretory Ag and subunit pili Ag to detect the antibody of *H. pylori* in panel sera of *H. pylori*.

| Testing result | *H. pylori* (+) (n=40) | *H. pylori* (−) (n=40) |
|----------------|------------------------|------------------------|
| ICT secretory Ag |            |            |
| Positive         | 38          | 5           |
| Negative         | 2           | 35          |
| ICT subunit pili Ag |       |            |
| Positive         | 37          | 1           |
| Negative         | 3           | 39          |

*H. pylori* = Helicobacter pylori
Discussion

H. pylori infection can be diagnosed using either invasive or non-invasive approaches [33]. Among the non-invasive approaches, serological techniques are widely used because they are cost-effective, simple, and quick [22,34]. However, it is unreliable to differentiate between active and previous infections [18]. Due to this condition, it was considered essential to develop a serological test to evaluate the progression of infection, especially in conjunction with eradication therapy. Therefore, a new non-invasive diagnostic test developed based on the detection of H. pylori IgG in this study was found to be a reliable test for detecting antibodies against H. pylori proteins. Using ICT test, both the 49.6 kDa-sub-unit pili protein of H. pylori and ICT secretary Ag were found to be sensitive in detecting the presence of antibodies in vaccinated mice, from the 2nd week after vaccination. However, the 49.6 kDa-sub-unit pili protein was more sensitive than ICT secretary Ag. The first antigen demonstrated 34% and 96% of positive reaction at the 2nd and 3rd weeks after the infection. Meanwhile, the second protein showed positive reaction of 4% and 78% at the 2nd and 3rd weeks, respectively (Figure-2). The use of the prepared antigen to test sera from dyspepsia patients undergoing endoscopy using the ICT test showed that it reacted 37 out of the 40 sera from an infected human with a specificity of 92.5% and detected 1 out of the 40 sera (2.5%) from the non-H. pylori-infected patients. Although the ICT secretary Ag reacted with sera from an infected human with a specificity of 95%, this protein also reacted with 5 out of the 40 sera (12.5%) from the non-H. pylori-infected patients, suggesting that this protein was less specific than the 49.6-kDa sub-unit pili protein of H. pylori.

Conclusion

In this study, a specific 49.6-kDa sub-unit pili protein was found to be a potential biomarker for the early and specific detection of H. pylori infection. From two antigens compared in this study, both secretary Ag ICT and sub-unit pili Ag ICT recognized specific antibodies that were prepared from mice experimentally infected with live H. pylori and from dyspepsia patients undergoing endoscopy. However, the 49.6-kDa sub-unit pili protein of H. pylori was found to be more sensitive than the secretary Ag, which could detect the presence of targeted antibodies in the 2nd week after vaccination in mice. Moreover, the 49.6-kDa sub-unit pili protein demonstrated true-positive detection of 92.5% and true-negative detection of 97.5% in human sera. In contrast, the secretary Ag ICT showed true-positive and true-negative detection of 95% and 87.5%, respectively. It was concluded that a feasible serological test developed in this study, using a specific 49.6-kDa sub-unit pili protein of H. pylori, was considered to be an important antigen used in serological tests for monitoring dyspepsia patients undergoing endoscopy; therefore, it was recommended to apply this protein in serological test for the detection of H. pylori infection in humans.

Authors’ Contributions

HS: Designed and managed this research, did laboratory works, and wrote the manuscript; ZM: Collected samples and wrote the manuscript; IWMT: Did laboratory works, analyzed the data, and wrote the manuscript; KKA: Analyzed the data and wrote the manuscript; SRP: Designed the research, analyzed the data, and wrote the manuscript. All authors read and approved the final manuscript.

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Competing Interests

The authors declare that they have no competing interests.

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