Obesity-Mediated Regulation of HGF/c-Met Is Associated with Reduced Basal-Like Breast Cancer Latency in Parous Mice

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Abstract

It is widely thought that pregnancy reduces breast cancer risk, but this lacks consideration of breast cancer subtypes. While a full term pregnancy reduces risk for estrogen receptor positive (ER+) and luminal breast cancers, parity is associated with increased risk of basal-like breast cancer (BBC) subtype. Basal-like subtypes represent less than 10% of breast cancers and are highly aggressive, affecting primarily young, African American women. Our previous work demonstrated that high fat diet-induced obesity in nulliparous mice significantly blunted latency in C3(1)-TAg mice, a model of BBC, potentially through the hepatocyte growth factor (HGF)/c-Met oncogenic pathway. Experimental studies have examined parity and obesity individually, but to date, the joint effects of parity and obesity have not been studied. We investigated the role of obesity in parous mice on BBC. Parity alone dramatically blunted tumor latency compared to nulliparous controls with no effects on tumor number or growth, while obesity had only a minor role in further reducing latency. Obesity-associated metabolic mediators and hormones such as insulin, estrogen, and progesterone were not significantly regulated by obesity. Plasma IL-6 was also significantly elevated by obesity in parous mice. We have previously reported a potential role for stromal-derived hepatocyte growth factor (HGF) via its cognate receptor c-Met in the etiology of obesity-induced BBC tumor onset and in both human and murine primary coculture models of BBC-aggressiveness. Obesity-associated c-Met concentrations were 2.5-fold greater in normal mammary glands of parous mice. Taken together, our studies demonstrate that, parity in C3(1)-TAg mice dramatically reduced BBC latency compared to nulliparous mice. In parous mice, c-Met is regulated by obesity in unaffected mammary gland and is associated with tumor onset. C3(1)-TAg mice recapitulate epidemiologic findings such that parity drives increased BBC risk and potential microenvironmental alterations in c-Met signaling may play a role in etiology.

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Data Availability: The authors confirm that all data underlying the findings are fully available without restriction. All relevant data are within the paper and its Supporting information files.

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Introduction

Epidemiologic and experimental data have shown that a full term pregnancy reduces breast cancer risk [1]. However, with the advent of protein and mRNA expression profiling and the Tumor Cancer Genome Atlas (TCGA), classification of tumor subtypes with specific risks and outcomes has shed new light on breast cancer incidence [2,3]. BBC represents 5–10% of breast cancers [4]. BBCs are estrogen receptor-, progesterone receptor- and human epidermal growth factor receptor 2 (HER2)- negative, thus, often referred to as triple-negative breast cancers, and as such these cancers lack a targeted therapy [5]. Patients have poor overall survival because these tumors are highly proliferative. Tumors are diagnosed predominantly in young African-American women, particularly obese women [6–8]. While a full term pregnancy reduces risk for estrogen receptor-positive breast cancers, like the luminal subtype [1], parity is actually associated
with increased probability of developing the more aggressive basal-like breast cancer (BBC) subtype [7,9–12].

The choice of mouse model is critical in modeling breast cancer subtypes. Protective effects of pregnancy have been described in luminal subtype models wherein parity antagonized the effects of carcinogens such as dimethylbenzanthracene (DMBA) [13,14] and ionizing radiation [15]. The protective effect of pregnancy can be mimicked by exogenous administration of high levels of estrogen and progesterone [16]. However, to date there have been no such studies on the effects of parity on BBC onset in murine models. We have shown that BBC is characterized by unique epithelial-stroma interactions, which likely play a role in etiology [17–22]. Gatenby and Gillies speculate that the origin of cancer may lie not in mutations within epithelial cells, but within acquired or somatic mutations in the mesenchymal cells that control tissue structure [23]. Thus, we hypothesized that pregnancy would induce long term changes such as inflammatory and metabolic alterations in the breast microenvironment that promote BBC [19,24].

Parity is often associated with excess weight gain and retention of weight after delivery, which is especially true for African American women, who gain more weight than recommended by the Institute of Medicine and retain more of that weight postpartum [25]. Obesity is a well-known risk factor for many cancers [26], with heterogeneous effects on breast cancer risk when subtypes and/or menopausal status are taken into account. For breast cancer overall, results for which are dominated by the effects of parity and obesity on tumour latency and progression. Further mechanistic insight. Using C3(1)-TAg mice, we examined interactions with parity were investigated to gain experimental findings; however, the underlying mechanisms of obesity-induced risk remain uncertain.

Materials and Methods

Reagents

c-Met goat anti-mouse antibody (detects pro- and cleaved c-Met) was obtained from R&D Systems (Minneapolis, MN). Anti-SV40-TAg was obtained from Santa Cruz (Santa Cruz, CA). Estrogen and progesterone ELISA kits were obtained from Novatein Bio (Cambridge, MA).

C3(1)-TAg Mouse Model

Diet: C3(1)-TAg mice [34] were used to study the role of obesity and parity on BBC. Studies were performed with approval and in accordance with guidelines of the Institutional Animal Care and Use Committee at the University of North Carolina at Chapel Hill (UNC-CH, NC). Female C3(1)-TAg mice were obtained through a collaboration with the UNC Lineberger Comprehensive Cancer Center (LCCC) Mouse Phase I Unit (MPIU). Female mice at seven weeks of age were placed with male mice for breeding. Males were removed at pregnancy. Following the delivery of litters, at maternal age of approximately 10 weeks old, pups were removed immediately after birth, and the mothers were randomly assigned to diet groups. Ten percent kcal from fat diet (“10%”) or 60% kcal (“60%”) lard-based diets matched for protein, vitamins, and minerals were obtained from Research Diets Inc. (New Brunswick, NJ) after customization, as in [36]. Custom diet information can be found in Table S1.

Tumor latency, number, growth and volume. Mice were monitored for tumor development by palpating thrice weekly following initiation of diets at 10 weeks of age. Initial tumor latency was defined as age at detection of first tumor and is reported as mean ± standard deviation (Table 1). Total tumor latency was defined as latency of all tumors palpated until sacrifice and is reported as Kaplan-Meier plot. Tumor volumes were measured once weekly over 3 weeks, following detection of first tumor, using ultrasound measurements with the Visualsonics 2000 (Toronto, Canada) as in [36]. The tumor volumes were calculated using the formula: length x width² x 0.5. The percent change in volume over time was calculated: (End volume – start volume)/Start volume * 100. The number of tumors per mouse was counted at sacrifice.

Body weight and composition. Prior to starting mice on diet and weekly until sacrifice, body weight was measured in grams. Body composition including lean mass, fat mass, free water content and total water content of non-anesthetized mice was measured prior to initiating diet and monthly thereafter using the EchoMRI-100 quantitative magnetic resonance whole body composition analyzer (Echo Medical Systems, Houston, TX). Obesity is defined as greater than a 5% incremental increase in fat composition. Fat mass is presented as % fat mass over total body weight measured day of MRI.

Blood glucose. Blood glucose was measured prior to start of diet and at sacrifice following a 6 h fast using a Bayer Contour Blood Glucose Monitor (Bayer HealthCare LLC, Tarrytown, NY).

Tissue harvest. 3 weeks after detection of the first tumor, mice were sacrificed by an intraperitoneal (i.p.) injection of avertin (Fisher Scientific, Pittsburgh, PA). Follow ing euthanasia, blood was collected by cardiac puncture in a tube with 10 μl of 0.05 mM EDTA. Plasma was collected by centrifuging blood at 5000 xg for 5 min. Mammary glands without palpable or visible tumors were collected as normal unaffected gland, although atypia of ductal epithelium could be present in C3(1)-TAg mice after 8 weeks of age [34]. Portions of the tissues were placed into a cassette and formalin fixed for immunohistochemical (IHC) analysis.
nulliparous mice.

**Table 1.** Tumor latency in C3(1)-TAg nulliparous and parous mice.

|          | Nulliparous [35] | Parous | p-value |
|----------|------------------|--------|---------|
| 10%      | 18.99±0.59       | 16.50±0.38 | 0.001   |
| 60%      | 16.94±0.51       | 15.91±0.47 | >0.05   |

Initial tumor latency was defined as the age at which first tumor palpated. Mean ± standard error of mean of tumor latency of the first tumor palpated. N=15 per group.

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**Plasma hormone panel.** Plasma collected at sacrifice was used for measuring metabolically relevant hormones and inflammatory mediators (insulin, IL-6, MCP-1 and TNF-α) using the Milliplex MAP Mouse Metabolic Hormone Magnetic Bead Panel in the Luminex MAGPIX system (EMD Millipore, Billerica, MA). The homeostasis model assessment-insulin resistance (HOMAIR) was used to calculate the approximate insulin resistance using the formula (glucose (mg/dl at sacrifice)/insulin (at sacrifice)/405) as previously described [21,36]. Estrogen and progesterone plasma concentrations were measured using ELISA assays following the manufacturer’s protocol (Novatein Bio; Cambridge, MA).

**IHC of c-Met and SV40 TAg in Normal Mammary Glands and Tumors**

IHC for c-Met and SV40 T antigen (TAg) was performed in normal mammary glands and tumors using methods as described [36,38]. Stained slides were scanned into the Aperio Scanscope CS system (Aperio Technologies, Vista, CA) at a magnification of 40X. Sections were then analyzed quantitatively using the Aperio Imagescope software: membrane IHC algorithm for c-Met quantification and the positive pixel counts for diaminobenzidine (DAB) staining in the color deconvolution algorithm for TAg as in [36]. N=6 random areas from sections (n=2 per mouse) were quantified and averaged per tumor per animal (n=5 mice per diet exposure group). Images (40X) shown are representative.

**Statistical Analysis**

Raw data are available in Table S2. Data in all cases are expressed as mean ± standard error of the mean (S.E.M.). Comparison of mean was carried out using a one-way analysis of variance (ANOVA) analysis with the SPSS (version 20) software (IBM SPSS Statistic 20.0, Armonk, NY). The level of significance was set at P<0.05. Kaplan-Meier analyses were conducted using GraphPad Prism 5 software to estimate tumor latency. Log rank and chi-square tests were used to investigate differences among groups. P values<0.05 were considered statistically significant.

**Results**

**Parity and obesity reduce C3(1)-TAg basal-like tumor latency**

To determine if parity or obesity altered tumor latency in C3(1)-TAg BBC mice, age-matched parous mice were fed control (10% kcal from fat) or obesogenic (60% kcal from fat) diets. Parity significantly decreased latency in lean mice fed 10% diet compared to previous reports in nulliparous mice fed identical diets (Table 1 [36]). In lean control mice fed 10% diet, parity decreased latency by almost 2.5 weeks (Table 1, P = 0.001). However, in obese mice fed 60% diet, the effect of parity on latency was not significantly different compared to obese nulliparous mice.

Within parous mice, no significant effects were detected on median initial tumor latency (data not shown) or total tumor latency (initial (first tumor) plus subsequent tumors detected until sacrifice) (Figure 1A). The hazard ratio comparing median latencies of 18.00 weeks for 10%-fed mice to 17.43 for 60%-fed mice was 1.151 (95% CI of ratio: 0.67 to 1.98). However, obesity significantly reduced mean latency (Figure 1B, P = 0.002). There were no significant alterations in tumor number at sacrifice or tumor volume changes over three weeks from time of tumor identification until sacrifice in parous mice (data not shown).

**c-Met expression in normal mammary glands and tumors**

**correlated with tumor latency**

Significant effects of parity on latency but not tumor burden or progression in the C3(1)-TAg GEMM suggested that effects of parity were occurring early in tumorigenesis to significantly alter tumor onset. Previous work by our group has demonstrated a role for obesity-mediated HGF/c-Met signaling in normal mammary gland of nulliparous C3(1)-TAg mice [36] which was reversed by weight loss [21]. In the current study, normal mammary glands from parous mice were examined for c-Met receptor expression. c-Met expression in normal mammary gland from 60%-fed mice demonstrated primarily epithelial localization (Figure 1C). Digital quantification revealed that c-Met protein concentrations were significantly increased by 2.5-fold in obese 60% (P = 0.005)-fed mice compared with 10%-fed controls (Figure 1D). Interestingly in tumors, c-Met protein levels were significantly decreased in obese 60% (P = 0.042)-fed mice compared with 10%-fed controls (data not shown).

**Metabolic mediators, but not pregnancy hormones or TAg expression, in normal mammary glands and tumors**

**associated with decreased latency**

To determine if metabolic parameters correlated with tumor onset, body weight and composition were measured. Parous mice fed 60% diets were significantly heavier compared with 10%-fed mice (P<0.05, Figure 2A). Compared to mice on 10% diet, 60%-fed mice also exhibited greater fat mass at 4 weeks on diet (P = 0.0001), and at 8 weeks (60% P = 0.0049, Figure 2B) as measured by MRI. Glucose, insulin and HOMAIR measures did not vary by diet exposure in parous C3(1)-TAg mice (Figures 2C–E).

**Systemic inflammatory mediators** were also examined as potential contributors to tumor onset [35]. IL-6 was significantly elevated in obese mice (P = 0.03) compared to 10%-fed controls (Figure 3A). However, other chemokine and cytokines markers of systemic inflammation including MCP-1 and TNF-α were not modified by diet (Figure 3B and C). Estrogen and progesterone concentrations were similar between lean and obese mice (Figure 3D and E).

To establish that parity or diet exposure did not alter expression of the oncogenic transgene SV40 T-antigen driven by the C3
Figure 1. Obesity and parity regulated a decrease in BBC latency and elevated c-Met in normal mammary glands. A) Upon initiation of diet at 10 weeks of age, parous mice were palpated three times a week for tumor onset (first tumor detected) (N = 15). Using the Kaplan-Meier analysis the hazard ratio comparing 60% to 10% was 1.151 (95% CI of ratio: 0.67 to 1.98). Median latencies in 10% and 60%-fed mice were 18.00 and 17.43 weeks, respectively. Using a chi-square test with a degree of freedom of 1, 10% vs. 60% equaled 0.61. B) Mean latencies of total tumors palpated from initial until sacrifice are shown. N = 30 tumors in 15 mice per group. *P = 0.002. C) Representative 40X photomicrographs of the IHC analysis of membrane localized c-Met staining (arrows). D) Total c-Met protein levels in normal mammary glands were quantified. *P = 0.003. doi:10.1371/journal.pone.0111394.g001

Figure 2. Diet-induced obesity affects body weight, adiposity, and leptin but not metabolic parameters in parous mice. A) C3(1)-TAg body weight was measured weekly over the course of the study until mice were sacrificed. Diet was initiated at 10 weeks of age (week 0 on diet) in n = 15 mice. *P < 0.05 over weeks 1–6. B) Body fat content by MRI was measured monthly until sacrifice. Percent fat content over total body mass is shown. n = 15. *P < 0.0001 at 4 weeks on diet, and P = 0.0049 at 8 weeks on diets. C) Blood glucose levels were measured from tail vein blood in mice fasted 6 hours. n = 15 mice. D) Insulin was measured at sacrifice in 6 hour fasted mice. n = 12. E) Homeostasis model assessment of insulin resistance (HOMA_{IR}) was calculated from measures at sacrifice. n = 12. doi:10.1371/journal.pone.0111394.g002
promoter, which is the key driver of tumorigenesis in this model, IHC was undertaken and digitally quantified. No significant differences were observed in the SV40-TAg expression in the normal mammary glands (Figures 4A and B). In the tumors, there was in fact a significant decrease in the SV40 TAg expression levels in the 60% group compared with the 10% diet-fed mice (P = 0.044) (data not shown).

**Discussion**

While pregnancy generally protects against the development of hormone-responsive estrogen and progesterone receptor positive tumors [39], it is an established risk factor for BBC. The Carolina Breast Cancer Study, among others, has identified parity as a positively associated risk factor for the development of BBC in both pre- and postmenopausal women [8,9,26,34,40,41]. Similarly, while pre-menopausal obesity is associated with protection from the development of luminal breast cancers (the most prevalent subtype), both pre- and post-menopausal obesity are associated with increased BBC risk [7]. The convergence of pregnancy and obesity is an important public health concern in relation to BBC risk. During the reproductive years (approximately age 20–39), roughly 25% of non-Hispanic whites are obese (BMI ≥30), and prevalence of obesity is approximately twice that in non-Hispanic black Americans (53.9% obese in 2003–2004) [42]. Since, obesity is a national epidemic in the United States [6] and BBC is a triple negative subtype that has no targeted therapies [5], it is important to understand the interaction between obesity and parity in relation to BBC risk.

Pregnancies cause extensive tissue remodeling, with the epithelium filling the majority of the mammary fat pad. Stromal cells including adipocytes, fibroblasts, and immune cells are important components of the mammary gland that work in concert to regulate the gland as it changes to produce milk during lactation. After weaning, tissue remodeling called involution causes the mammary gland to return to a microenvironmental architecture similar to the virgin gland [43,44]. These developmental changes and alterations in microenvironment may represent opportunities for carcinogenesis and may further interact with
other environmental exposures, such as dietary fat exposure and the obese state.

Using C3(1)-Tag mice, a BBC GEMM that represents human BBC [4], our previous work examining nulliparous mice demonstrated that obesity lead to a significant two week reduction in latency compared to lean controls [36]. Herein, we demonstrated that in lean mice parity alone significantly shortened BBC latency compared to nulliparous controls. However, in obese mice, the effect of parity was lost because parity did not reduce latency further than obesity alone. Within parous mice, obesity did not reduce median latency of the first tumor detected. However obesity significantly reduced secondary and subsequent mean tumor latency compared to lean controls. There were no effects on total tumor burden or promotion which was similar to findings in nulliparous mice [36]. Taken together, parity-induced reductions in latency may leave little opportunity for dramatic effects of obesity-associated factors that were observed in nulliparous mice. The C3(1)-Tag parous mouse model experiences tumorigenesis at a young age (akin to BBC in humans) [8], and thus poses a challenge for studying the joint effects of parity and obesity.

Obesity leads to numerous changes in the stroma of the mammary microenvironment and other fat pads including the release of growth factors and regulation of growth factor receptors [36,38,45–48]. The HGF/c-Met axis is a pathway that is linked to both obesity and breast cancer risk [21]. c-Met activation drives cell proliferation, angiogenesis, differentiation, migration, and anti-apoptosis pathways [49–51]. In nulliparous C3(1)-Tag mice, c-Met protein levels were significantly elevated by obesity in normal mammary gland and tumors [36]. Herein, we demonstrated that obesity also significantly elevated c-Met protein levels in the normal mammary glands in parous mice. However, c-Met was not elevated by obesity in tumors with parity, although this was just a single measure on tumors isolated at sacrifice. It may be possible that obesity creates a dysfunctional microenvironment in the post-partum period wherein elevated c-Met protein levels persist in the normal mammary gland and are not down-regulated after resolution of involution, therefore creating fertile grounds for the development of HGF/c-Met-driven BBC. The role of c-Met signaling in BBC is currently under investigation. Future studies will test inhibition of c-Met signaling in the onset of post-partum obesity-driven BBC. Because BBC has unique risk factors that in many cases are traditionally thought of as breast cancer preventive factors [35], future studies should investigate the role of weight gain or loss in the post-partum period as modifiable BBC risk factors.

**Supporting Information**

**Table S1** Contains information on custom diet. (DOCX)

**Table S2** Contains raw data. (XLSX)

**Author Contributions**

Conceived and designed the experiments: SS MAT LM. Performed the experiments: SS AJF JAG KKM KMB. Analyzed the data: SS. Contributed reagents/materials/analysis tools: DBD. Contributed to the writing of the manuscript: SS MAT LM.

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