Relevance and prognostic ability of Twist, Slug and tumor spread through air spaces in lung adenocarcinoma

Ao Liu1 | Xiao Sun | Jin Xu | Yunpeng Xuan | Yandong Zhao | Tong Qiu | Feng Hou | Yi Qin | Yuanyong Wang | Tong Lu | Yang Wo | Yujun Li | Xiaoming Xing | Wenjie Jiao

Abstract

Background: Tumor spread through air spaces (STAS) is a novel pathologic characteristic in lung adenocarcinomas that indicates invasive tumor behavior. We aimed to explore the relationship between Twist, Slug and STAS in lung adenocarcinoma and to investigate the potential relationship between epithelial-mesenchymal transition (EMT) and STAS.

Materials and methods: Our study retrospectively analyzed 115 patients with resected lung adenocarcinomas to evaluate the relationship between Twist, Slug and STAS in lung adenocarcinoma. STAS was diagnosed using hematoxylin-eosin (H&E) staining. Immunohistochemistry was used to evaluate the expression levels of Slug and Twist.

Results: In this study, 56 (48.7%) patients had STAS, 40 (34.8%) patients had Slug overexpression, and 28 (24.3%) patients had Twist overexpression. Patients with either STAS or Slug and Twist overexpression experienced poor recurrence-free survival (RFS) and overall survival (OS). There were significant associations between Twist overexpression, Slug overexpression and the presence of STAS. The logistic model further revealed that pathological stage, Twist overexpression and Slug overexpression were independent risk factors for STAS. A multivariate analysis that contained Twist, Slug, pathologic stage and STAS, showed that pathologic stage and STAS were independent prognostic factors for poor RFS and OS. Another multivariate model that contained Twist, Slug and pathologic stage, showed that pathologic stage, Twist overexpression and Slug overexpression were independent risk factors for poor RFS and OS. In the cohort with STAS, the multivariate analysis showed that pathologic stage and Twist overexpression were independent risk factors for poor survival. The subgroup analysis showed that patients with both Slug overexpression and Twist overexpression with STAS received a poor prognosis.

Conclusions: STAS, Slug and Twist were correlated with poor RFS and OS in resected lung adenocarcinomas. Additionally, STAS was correlated with the...
1 | INTRODUCTION

Spread through air spaces (STAS) is a new pathologic feature of tumor invasion.1 It has been reported that cancer cells beyond the edge of the tumor invaded into the alveolar spaces in the lung surrounding parenchyma shown as single cells, micropapillary patterns or solid nests.2 Previous studies have reported that STAS was significantly correlated with poor overall survival (OS) and recurrence-free survival (RFS) in patients with surgically resected lung cancer.2-6 STAS was associated with invasive characteristics, such as lymphovascular invasion, visceral pleura invasion and invasive patterns.2,3,5-10 It was reported that metastasis-associated protein I (MTA1) might provide potential information on the mechanism of STAS.11 However, further studies are still needed to identify the molecular characteristics and mechanisms underlying STAS.

The adhesion protein E-cadherin is one of the hallmarks of epithelial morphogenesis. Tumor cells cannot maintain tissue integrity through the formation of adhesion junctions since losing E-cadherin, appears as loose tumor cells, which easily escape out of the primary tumor. The decreased expression of cell adhesion molecules, such as E-cadherin, is one of the main characteristics in the activation of epithelial-mesenchymal transition (EMT). EMT is considered a major driver of tumor progression from initiation to metastasis.12,13 EMT is a highlyconserved cellular program that plays key roles in promoting tumor invasion and metastasis. In EMT, polarized epithelial cells lose their epithelial characteristics, intercellular adhesion complexes and cytoskeletal-specific architecture, and convert into a motile mesenchymal type of cell with polarity, invasive ability and metastatic capacity.12-14 E-cadherin expression is downregulated during the acquisition of metastatic potential at the later stages of epithelial tumor progression.15,16 Some molecules, including Twist and Slug, have been found to act as E-cadherin transcription factors that induce EMT.

In view of the association between the migration and invasion of cancer cells with EMT, we hypothesized that STAS regulatory molecules might be associated with EMT. Thus, we investigated the relationship between Twist and Slug with STAS in lung adenocarcinoma and aimed to investigate the potential connection between STAS and EMT.

2 | MATERIALS AND METHODS

2.1 | Patients

In this study, we reviewed patients with resected lung adenocarcinoma who were diagnosed between January 2010 and December 2010 at our institution. This retrospective study had been approved by the Institutional Review Board at the Affiliated Hospital of Qingdao University (IRB#QYFYKY 2018-10-11-2). This study followed the Helsinki Declaration. This was a retrospective study, so informed consent was not needed by the Institutional Review Board. The exclusion criteria of this study were as follows: (a) patients with multiple nodules; (b) patients with variants of adenocarcinoma, even with other components, including neuroendocrine or squamous differentiation; and (c) patients who were lost to follow-up. Patients were included when the following inclusion criteria were met: (a) patients with a single primary tumor; and (b) patients with a confirmed M0 stage. In total, we reviewed 115 patients with resected lung adenocarcinoma in this study. The end follow-up date was 30 December 2018.

2.2 | Histopathological examination of STAS

H&E-stained tumor slides selected in this study were evaluated by two experienced pathologists who were blinded to patient clinical outcomes. Any possible differences were resolved by a consensus after a discussion. Pathologic stage was based on the TNM stage (8th edition). The definition of STAS was based on the 2015 WHO classification. STAS was defined as when cancer cells were located within the alveolar space beyond the edge of the tumor in the lung surrounding the parenchyma accompanied by micropapillary patterns, solid nests and single tumor cells.2 First, the boundary of the primary tumor with a smooth surface was confirmed under low-power microscopy. Then, we identified STAS as when tumor cells or the tumor cell mass was isolated from the primary tumor and the presence of normal alveolar structures beyond the boundary of the primary tumor. STAS was distinguished and confirmed with high-power field microscopy. Moreover, to discriminate STAS from artificially detached tumor cells or pulmonary loose tumor tissue fragments arising from tumor dissection, a clear vision of a completely normal alveolar structure instead of a broken tissue structure is necessary.
2.3 Immunohistochemistry using tissue microarrays

Immunohistochemistry was carried out via two steps. Briefly, first, the paraffin sections were deparaffinized and rehydrated. Following microwave antigen retrieval, we blocked endogenous peroxidase activity by incubating slides in 0.3% H2O2 and blocked non-specific binding sites with 10% goat serum for 1 hour. Then, the sections were rinsed and incubated with the anti-Twist (ab50887, Abcam; diluted 1:100) or anti-Slug (ab27568, Abcam; diluted 1:100) antibody overnight at 4°C. Then, the Diaminobenzidine Horseradish Peroxidase Color Development Kit (Beyotime, China) was used for color development. Ultimately, the sections were counterstained in hematoxylin and mounted.

Immunohistochemical scores based on the distribution and intensity of staining were used to grade the immunoreactive level of Twist and Slug. The staining distribution indicates the percentage of positive cells in each core, with scores of 0 (0%), 1 (1%-50%), and 2 (>50%). The staining intensity was scored as 0 (no expression), 1 (mild expression), 2 (intermediate expression), and 3 (strong expression). Overall, the distribution and intensity scores were summed into a total score (0 to 5) for each patient. Immunostaining scores were dichotomized into low and high. Only a total immunohistochemical score ≥4 was regarded as a positive result.

2.4 Statistical analyses

The relationship between STAS and clinicopathologic factors was calculated by the chi-square test and Student’s t test. A logistic regression model was used to assess the association between patient characteristics and STAS. Univariate Cox regression models were preformed to calculate the prognostic factors. Furthermore, a multivariate Cox regression model was performed to analyse independent predictors. Kaplan-Meier analyses and log-rank test were performed to assess RFS and OS. Statistical analyses were performed with IBM SPSS Statistics version 25.0 (IBM Corp.). A P value less than .05 was considered statistically significant.

3 RESULTS

3.1 Patient characteristics

We examined 115 patients who underwent lobectomy in this study. Their detailed clinicopathological characteristics are summarized in Table 1. The mean age of the patients at diagnosis was 60.36 years. Among 115 patients, 27 (23.4%) had pathologic T1 stage disease (T1a stage: 4.3%, T1b stage: 13.9%, T1c stage: 5.2%), 88 (76.6%) had pathologic T2 stage disease (T2a stage: 60.9%, T2b stage: 15.7%), 35 (30.4%) lymph node metastasis (N1 stage: 7.8%, N2 stage: 22.6%), and 82 (24.3%) had the presence of visceral pleural invasion (VPI). All patients were at the M0 stage. During the follow-up, 61 (53.0%) patients experienced recurrence, while 64 (55.7%) patients died. The mean follow-up time of the analysis at the endpoint was 76.61 months.

3.2 Association between slug, twist and clinicopathological characteristics

The associations between clinicopathologic factors, the expression of Slug and Twist and the presence of STAS are summarized in Table 1. Among the 115 patients examined in this study, 75 (65.2%) had Slug overexpression and 28 (24.3%) had Twist overexpression. As shown in Table 1, correlations between the expression levels of Slug and Twist and clinicopathological characteristics are listed. There was a significant relationship between Slug overexpression and pathologic differentiation (P = .033), pathologic T stage (P = .013), and the presence of VPI (P = .018). Slug overexpression showed a potential correlative trend with lymph node metastasis (32.5% vs 40.0%). On the other hand, Twist overexpression was not significantly associated with a high pathologic stage. However, there could be potential correlations between Twist overexpression and high pathologic differentiation (20.0% vs 25.0% vs 30.0%), increased lymph node metastasis (22.5% vs 28.6%), and the presence of VPI (21.2% vs 25.6%). Regarding the overexpression of Slug and Twist, increased Slug overexpression could be associated with increased Twist overexpression (31.0% vs 46.4%, P = .137) and vice versa (20.0% vs 32.5%, P = .137).

3.3 Association between STAS and clinicopathological characteristics

Associations between clinicopathologic characteristics and the presence of STAS are shown in Table 1. STAS was observed in 56 patients (48.7%). Representative microphotographs of STAS are shown in Figure 1. There was a significant association between the presence of STAS and lymph node metastasis (P = .005) as well as pathologic N stage (P = .012), pathologic stage (P = .013), the presence of VPI (P = .012), Twist expression (P < .001) and Slug expression (P = .011).

An additional logistic regression analysis was performed to calculate the association between STAS and pathological factors. The logistic regression analysis contained pathologic stage, Twist and Slug (Table 2). The logistic model further revealed that pathologic stage (OR = 2.154, 95% CI = 1.277-3.636, P = .004), Twist overexpression (OR = 6.104, 95%
TABLE 1  Clinicopathologic characteristics according to STAS, Twist and Slug in patients with lung adenocarcinoma

| Characteristic                      | All patients (n = 115) | STAS Absence (n = 59) | STAS Presence (n = 56) | Slug expression Low (n = 75) | Slug expression High (n = 40) | Twist expression Low (n = 87) | Twist expression High (n = 28) | P Value |
|------------------------------------|------------------------|-----------------------|------------------------|-----------------------------|-------------------------------|-----------------------------|-------------------------------|---------|
| Gender                             |                        |                       |                        |                             |                               |                             |                               |         |
| Male                               | 54 (47.0)              | 27 (50.0)             | 27 (50.0)              | 36 (66.7)                   | 18 (33.3)                    | 43 (79.6)                   | 11 (20.4)                    | .792    |
| Female                             | 61 (53.0)              | 32 (52.5)             | 29 (47.5)              | 39 (63.9)                   | 22 (36.1)                    | 44 (72.1)                   | 17 (27.9)                    | .759    |
| Age                                |                        |                       |                        |                             |                               |                             |                               |         |
| ≤65                                | 82 (71.3)              | 44 (53.7)             | 38 (46.3)              | 52 (63.4)                   | 30 (36.6)                    | 60 (73.2)                   | 22 (26.8)                    | .426    |
| >65                                | 33 (28.7)              | 15 (45.5)             | 18 (54.5)              | 23 (69.7)                   | 10 (30.3)                    | 27 (81.8)                   | 6 (18.2)                     | .426    |
| Smoking history                    |                        |                       |                        |                             |                               |                             |                               |         |
| Never                              | 73 (63.5)              | 40 (54.8)             | 33 (45.2)              | 48 (65.8)                   | 25 (34.2)                    | 54 (74.0)                   | 19 (26.0)                    | .324    |
| Former/current                     | 42 (36.5)              | 19 (45.2)             | 23 (54.8)              | 27 (64.3)                   | 15 (35.7)                    | 33 (78.6)                   | 9 (21.4)                     | .324    |
| Post-operative hospitalization days|                        |                       |                        |                             |                               |                             |                               |         |
| Mean ± SD                          | 9.19 ± 2.665           | 8.81 ± 2.345          | 9.59 ± 2.934           | 8.79 ± 2.332                | 9.95 ± 3.088                 | 9.32 ± 2.764                | 8.79 ± 2.331                 | .119    |
| Range                              | 5-17                   | 5-17                  | 5-17                   | 5-17                        | 7-17                         | 5-17                        | 5-17                         | .119    |
| Tumor location                     |                        |                       |                        |                             |                               |                             |                               |         |
| RUL                                | 48 (41.7)              | 24 (50.0)             | 24 (50.0)              | 30 (62.5)                   | 18 (37.5)                    | 39 (81.2)                   | 9 (18.8)                     | .084    |
| RML                                | 8 (7.0)                | 4 (50.0)              | 4 (50.0)               | 4 (50.0)                    | 4 (50.0)                     | 4 (50.0)                    | 4 (50.0)                     | .084    |
| RLL                                | 18 (15.7)              | 6 (33.3)              | 12 (66.7)              | 13 (72.2)                   | 5 (27.8)                     | 13 (72.2)                   | 5 (27.8)                     | .361    |
| LUL                                | 25 (21.7)              | 12 (48.0)             | 13 (52.0)              | 16 (64.0)                   | 9 (36.0)                     | 19 (76.0)                   | 6 (24.0)                     | .361    |
| LLL                                | 16 (13.9)              | 13 (81.2)             | 3 (18.8)               | 12 (75.0)                   | 4 (25.0)                     | 12 (75.0)                   | 4 (25.0)                     | .361    |
| Tumor size (cm)                    |                        |                       |                        |                             |                               |                             |                               |         |
| 0-3                                | 69 (60.0)              | 33 (47.8)             | 36 (52.2)              | 48 (69.6)                   | 21 (30.4)                    | 48 (69.6)                   | 21 (30.4)                    | .361    |
| 3-5                                | 46 (40.0)              | 26 (56.5)             | 20 (43.5)              | 27 (58.7)                   | 19 (41.3)                    | 39 (84.8)                   | 7 (15.2)                     | .361    |
| Mean ± SD                          | 3.003 ± 1.237          | 3.127 ± 1.216         | 2.871 ± 1.256          | 2.989 ± 1.272               | 3.028 ± 1.184                | 3.161 ± 1.235               | 2.511 ± 1.126                | .361    |
| Pathologic differentiation         |                        |                       |                        |                             |                               |                             |                               |         |
| Low                                | 35 (30.4)              | 16 (45.7)             | 19 (54.3)              | 25 (71.4)                   | 10 (28.6)                    | 28 (80.0)                   | 7 (20.0)                     | .178    |
| Median                             | 60 (52.2)              | 29 (48.3)             | 31 (51.7)              | 33 (55.0)                   | 27 (45.0)                    | 45 (75.5)                   | 15 (25)                      | .178    |
| High                               | 20 (17.4)              | 14 (70.0)             | 6 (30.0)               | 17 (85.0)                   | 3 (15.0)                     | 14 (70.0)                   | 6 (30.0)                     | .178    |

(Continues)
### TABLE 1 (Continued)

| Characteristic                  | All patients (n = 115) | STAS                | Slug expression | Twist expression |
|---------------------------------|------------------------|---------------------|-----------------|------------------|
|                                 |                        | Absence (n = 59)    | Presence (n = 56) | Low (n = 75) | High (n = 40) | Low (n = 87) | High (n = 28) |
|                                 |                        | Number (%)          | Number (%)       | Number (%)      | Number (%)     | Number (%) | Number (%) | P Value |
| Lymph node metastasis           |                        |                     |                 |                 |                 |             |             |         |
| No                              | 80 (69.6)              | 48 (60.0)           | 32 (40.0)       | .005            | 54 (67.5)      | 26 (32.5) | .437        |         |
| Yes                             | 35 (30.4)              | 11 (31.4)           | 24 (68.6)       |                 | 21 (60.0)      | 14 (40.0) | .437        | .485 |
| Pathologic T stage              |                        |                     |                 |                 |                 |             |             |         |
| T1                              | 27 (23.5)              | 17 (63.0)           | 10 (37.0)       | .166            | 23 (85.2)      | 4 (14.8) | .013        | .827 |
| T2                              | 88 (76.5)              | 42 (47.7)           | 46 (52.3)       |                 | 52 (59.1)      | 36 (40.9) | .013        | .827 |
| Pathologic N stage              |                        |                     |                 |                 |                 |             |             |         |
| N0                              | 80 (69.6)              | 48 (60.0)           | 32 (40.0)       | .012            | 54 (67.5)      | 26 (32.5) | .701        | .686 |
| N1                              | 9 (7.8)                | 4 (44.4)            | 5 (55.6)        |                 | 5 (55.6)       | 4 (44.4) | .701        | .686 |
| N2                              | 26 (22.6)              | 7 (26.9)            | 19 (73.1)       |                 | 16 (61.5)      | 10 (38.5) | .701        | .686 |
| Pathologic stage                |                        |                     |                 |                 |                 |             |             |         |
| Stage I                         | 69 (60.0)              | 42 (60.9)           | 27 (39.1)       | .013            | 47 (68.1)      | 22 (31.9) | .722        | .231 |
| Stage II                        | 20 (17.4)              | 10 (50.0)           | 10 (50.0)       |                 | 12 (60.0)      | 8 (40.0) | .722        | .231 |
| Stage III                       | 26 (22.6)              | 7 (26.9)            | 19 (73.1)       |                 | 16 (61.5)      | 10 (38.5) | .722        | .231 |
| VPI                             |                        |                     |                 |                 |                 |             |             |         |
| Absence                         | 33 (28.7)              | 23 (69.7)           | 10 (30.3)       | .012            | 27 (81.8)      | 6 (18.2) | .018        | .619 |
| Presence                        | 82 (71.3)              | 36 (43.9)           | 46 (56.1)       |                 | 48 (58.5)      | 34 (41.5) | .018        | .619 |
| Twist expression                |                        |                     |                 |                 |                 |             |             |         |
| Low                             | 87 (75.7)              | 53 (60.9)           | 34 (39.1)       | .001            | 60 (69.0)      | 27 (31.0) | .137        | —        |
| High                            | 28 (24.3)              | 6 (21.4)            | 22 (78.6)       |                 | 15 (53.6)      | 13 (46.4) | .137        | —        |
| Slug expression                 |                        |                     |                 |                 |                 |             |             |         |
| Low                             | 75 (65.2)              | 45 (60.0)           | 30 (40.0)       | .011            | —              | —         | 60 (80.0) | .137 |
| High                            | 40 (34.8)              | 14 (35.0)           | 26 (65.0)       |                 | —              | —         | 27 (67.5) | .137 |

Abbreviations: LLL, left lower lobe; LUL, left upper lobe; RLL, right lower lobe; RML, right middle lobe; RUL, right upper lobe; SD, standard deviation; STAS, spread through air spaces; VPI, visceral pleural invasion.
CI = 2.104-17.711, \( P = .001 \) and Slug overexpression (OR = 2.471, 95% CI = 1.030-5.929, \( P = .043 \)) were independent risk factors for STAS.

### 3.4 | Survival analysis

The univariate Cox proportional hazards regression analysis was performed to assess the prognostic risk factors for RFS and OS (Table 3). Pathologic N stage (\( P < .001 \)), presence of VPI (\( P = .069 \) vs \( P = .011 \)), pathologic stage (\( P < .001 \)), presence of STAS (\( P < .001 \)), Twist overexpression (\( P = .037 \) vs
### TABLE 3  Univariate and multivariate analyses of RFS and OS in patients with lung adenocarcinoma

| Characteristic, factor | Univariate analysis | Multivariate analysis | Multivariate analysis |
|------------------------|---------------------|-----------------------|-----------------------|
|                        | RFS HR 95% CI P Value | OS HR 95% CI P Value | RFS HR 95% CI P Value |
| Clinical factors       |                     |                       |                       |
| Gender, male           | 0.888 0.537-1.467 .643 | 1.057 0.647-1.728 .824 | — — — — — — — — — — — — |
| Age, older, > 65       | 1.033 0.595-1.792 .909 | 1.261 0.748-2.126 .385 | — — — — — — — — — — — — |
| Smoking history, former/current | 1.255 0.750-2.098 .387 | 1.174 0.710-1.940 .533 | — — — — — — — — — — — — |
| Tumor location, lower lobe | 1.357 0.800-2.302 .257 | 0.884 0.512-1.525 .657 | — — — — — — — — — — — — |
| Pathologic factors     |                     |                       |                       |
| Tumor size (cm), larger (per cm) | 1.144 0.928-1.412 .208 | 1.046 0.850-1.286 .673 | — — — — — — — — — — — — |
| Pathologic differentiation | 1.334 0.925-1.925 .123 | 1.218 0.850-1.745 .282 | — — — — — — — — — — — — |
| Pathologic T stage (per stage) | 1.187 0.922-1.529 .184 | 1.263 0.976-1.635 .076 | — — — — — — — — — — — — |
| Pathologic T stage, T2 vs T1 | 1.598 0.849-3.010 .147 | 2.135 1.086-4.197 .028 | — — — — — — — — — — — — |
| Pathologic N stage (per stage) | 1.822 1.384-2.400 <.001 | 1.693 1.293-2.215 <.001 | — — — — — — — — — — — — |
| Lymph node metastasis, yes vs no | 2.884 1.727-4.815 <.001 | 2.590 1.572-4.269 <.001 | — — — — — — — — — — — — |
| Pathologic stage (per stage) | 1.376 1.179-1.606 <.001 | 1.351 1.164-1.568 <.001 | 1.783 1.313-2.421 <.001 |
| Pathologic stage, II vs I | 1.673 0.838-3.341 .145 | 1.637 0.843-3.181 .146 | 1.726 1.277-2.332 <.001 |
| Pathologic stage, III (vs I & II) | 3.049 1.776-5.233 <.001 | 2.586 1.520-4.401 <.001 | 2.016 1.494-2.720 <.001 |
| VPI, Presence           | 1.742 0.957-3.170 .069 | 2.272 1.211-4.263 <.001 | 1.897 1.408-2.556 <.001 |
| STAS, Presence          | 3.696 2.152-6.348 <.001 | 4.577 2.639-7.937 <.001 | 1.934 1.349-2.722 <.001 |
| Twist, overexpression   | 1.802 1.037-3.131 .037 | 2.002 1.178-3.404 .010 | 1.684 0.910-3.116 <.001 |
| Slug, overexpression    | 1.885 1.132-3.140 .015 | 2.155 1.314-3.532 .002 | 1.389 0.822-2.349 .220 |

Abbreviations: CI, confidence interval; HR, hazard ratio; HS, histological subtype; OS, overall survival; RFS, recurrence-free survival; STAS, spread through air spaces; VI, vascular invasion; VPI, visceral pleural invasion.
were significant prognostic risk factors for RFS and OS. Furthermore, patients with STAS experienced worse RFS or OS than those without STAS (hazard ratio (HR), 3.696; 95% confidence interval (CI), 2.152-6.348; \( P < .001 \) vs HR, 4.577; 95% CI, 2.639-7.937; \( P < .001 \)) in the univariate analysis. Moreover, patients with Slug overexpression also experienced worse RFS or OS than those without Slug overexpression (HR, 1.885; 95% CI, 1.132-3.140; \( P = .015 \) vs HR, 2.155; 95% CI, 1.314-4.352; \( P = .002 \)). Similarly, patients with Twist overexpression also experienced worse RFS or OS than those without Twist overexpression (HR, 1.802; 95% CI, 1.037-3.131; \( P = .037 \) vs HR, 2.002; 95% CI, 1.178-3.404; \( P = .010 \)).

Additional multivariate models were used to analyze independent predictors (Table 3). One multivariate analysis including pathologic stage, STAS, Twist and Slug showed that pathologic stage (HR, 1.783; 95% CI, 1.313-2.421; \( P < .001 \)) and the presence of STAS (HR, 2.638; 95% CI, 1.464-4.754; \( P = .001 \)) were independent prognostic risk factors for poor RFS. The same outcome was shown in the multivariate models of OS (pathologic stage: HR, 1.726; 95% CI, 1.277-2.332; \( P < .001 \); presence of STAS: HR, 3.475; 95% CI, 1.934-6.243; \( P < .001 \)).

Another multivariate model that contained Twist, Slug and pathologic stage was applied. Pathologic stage (HR, 2.016; 95% CI, 1.494-2.720; \( P < .001 \)), Twist overexpression (HR, 2.343; 95% CI, 1.297-4.232; \( P = .005 \)), and Slug overexpression (HR, 1.719; 95% CI, 1.029-2.872; \( P = .039 \)) were identified as independent risk factors for poor RFS. The same outcome was shown for OS (pathologic stage: HR, 1.897; 95% CI, 1.408-2.556; \( P < .001 \); Twist overexpression: HR, 2.568; 95% CI, 1.439-4.582; \( P = .001 \); Slug overexpression: HR, 1.869; 95% CI, 1.135-3.077; \( P = .014 \)).

Kaplan-Meier analysis was performed to determine survival rates according to the expression of Twist and Slug and the presence of STAS (Figure 2). Patients with STAS experienced significantly poorer RFS and OS than patients without STAS (\( P < .001 \)). The 5-year RFS rates in patients with and without STAS were 35.7% and 75.3%, respectively, and the 5-year OS rates were 50.0% and 88.1%, respectively. Similarly, patients with Twist overexpression also experienced significantly poorer RFS and OS than patients with low Twist expression (\( P = .030 \) vs \( P = .009 \)). Patients with Slug overexpression also experienced poor RFS and OS (\( P = .012 \) vs \( P = .002 \)).

### Survival analysis in the cohort according to Slug, Twist and STAS

All patients were divided into two cohorts according to STAS (Table 4). In the cohort with STAS, the multivariate analysis showed that pathologic stage (HR, 1.356; 95% CI, 1.101-1.670; \( P = .004 \)) was a significant risk factor for poor RFS. Additionally, Twist overexpression (HR, 1.737; 95% CI, 0.882-3.422; 0.110) could be acceptable as a risk factor for poor RFS. On the other hand, pathologic stage (HR, 1.519; 95% CI, 1.234-1.869; \( P < .001 \)) and Twist overexpression (HR, 2.317; 95% CI, 1.203-4.463; 0.012) were revealed as significant risk factors for poor OS.

Furthermore, all patients were divided into four groups according to Slug, Twist and STAS as follows: Group 1: STAS(−): patients with the absence of STAS regardless of the expression of Slug and Twist; Group 2: Low Slug/Low Twist&STAS(+): patients without Slug and Twist overexpression and with STAS; Group 3: High Slug/High Twist&STAS(+): patients with either Slug or Twist overexpression and with STAS; and Group 4: High Slug&High Twist&STAS(+): patients with both Slug and Twist overexpression and with STAS. Kaplan-Meier analysis was used to determine survival rates in the four cohorts (Figure 2). RFS and OS became increasingly worse from group 1 to group 4 (\( P < .001 \)).

### DISCUSSION

In this study, we examined the relationship between Twist, Slug and STAS in patients with resected stage I-III lung adenocarcinoma and aimed to investigate the potential connection between STAS and EMT. To our knowledge, this is the first study to assess the relationship between Slug, Twist and STAS in resected pathologic stage I-III lung adenocarcinoma.

Regarding the definition of STAS, STAS is a novel pathologic feature in lung cancer that includes three histomorphologic characteristics: solid nests, micropapillary patterns and single cells.\(^2\) STAS is also associated with recurrence and survival in squamous cell carcinoma,\(^18-20\) lung large cell neuroendocrine carcinoma, and lung small cell carcinoma,\(^21\) even lung pleomorphic carcinoma.\(^22\) STAS has been reported to be related with an aggressive tumor behavior, such as a larger tumor size,\(^6,18,23\) a high pathologic stage,\(^3,4,8,24,25\) lymphatic or vascular invasion,\(^2,4,6,8,10,18,23,25\) pleural invasion,\(^4,6,7,10,23,24\) tumor budding\(^19\) and increased micropapillary or solid structures in lung adenocarcinomas.\(^2,3,5-10\) We previously reported that the positive rate of STAS may be different for different invasive patterns and even lymph node metastasis.\(^10\) Thus, in this study, it was acceptable for the positive rate of STAS to be 48.7% in all patients with stage I-III lung adenocarcinomas. It was shown that STAS was significantly related to lymph node metastasis, pathologic stage and presence of VPI. Additionally, patients with STAS experienced poor RFS or OS, consistent with previous studies.

Based on current research, STAS has important significance as a reference in patient prognosis. It was reported that MTA1 might provide potential information on the mechanism of STAS.\(^11\) However, further studies are still in progress.
FIGURE 2  Kaplan-Meier survival analysis of recurrence-free survival (RFS) and overall survival (OS) according to spread through air spaces (STAS), Twist and Slug in patients with lung adenocarcinoma. A, RFS in patients with vs without STAS ($P < .001$). B, OS in patients with vs without STAS ($P < .001$). C, RFS in patients with high vs low Twist expression ($P = .030$). D, OS in patients with high vs low Twist expression ($P = .009$). E, RFS in patients with high vs low Slug expression ($P = .012$). F, OS in patients with high vs low Slug expression ($P = .002$). G, RFS in four groups of patients ($P < .001$). H, OS in four groups of patients ($P < .001$).
needed to identify the molecular characteristics and mechanisms underlying STAS. From a histomorphological perspective, tumor cells or tumor masses migrate from the primary tumor to the surrounding alveolar spaces beyond the edge of the primary tumor. The primary tumor, which is more prone to STAS, is frequently accompanied by loose tumor tissue. In addition, it has been reported that there are fewer intercellular adhesions among poorly differentiated discohesive tumor cells, including poorly differentiated squamous cell carcinomas and micropapillary adenocarcinomas.\textsuperscript{26,27} Interestingly, it has been reported that loose cells in micropapillary carcinoma most likely facilitate anchorage-independent growth and acquire resistance to apoptosis, which are advantageous for proliferation during lymphatic cancer metastasis.\textsuperscript{28} On the other hand, the presence of frequent STAS-like and decreasing E-cadherin expression in ROS1-rearranged lung cancer can predict lower disease-free survival.\textsuperscript{29} Additionally, STAS was proven to be associated with micropapillary patterns.\textsuperscript{2,3,5-10} Thus, we hypothesized that STAS could be associated with poor intercellular adhesions among tumor cells, by which the tumor cells could spread into the alveolar spaces beyond the edge of the primary tumor. Furthermore, a key molecule that maintains tissue integrity through the formation of adhesion junctions is E-cadherin. Thus, the loss of E-cadherin among tumor cells results in the degradation of cell-cell adhesions and the isolation of malignant cells from the epithelial layer, making it easy for loose tumor cells to escape from the primary tumor.

The role of EMT in promoting metastasis and cancer invasion was recently considered important. EMT is a process whereby epithelial cell layers with a loss of epithelial cell adhesion and cytoskeletal components undergo remodeling of the cytoskeleton. The main characteristics of EMT are the decrease in the expression of cell adhesion molecules such as E-cadherin, the transformation of a cytokeratin cytoskeleton into a vimentin-based cytoskeleton, and the morphological characteristics of mesenchymal cells.\textsuperscript{15,16} EMT is an important biological process for epithelial-derived malignant tumor cells to acquire migration and invasion abilities. Through EMT, epithelial cells lose cell polarity and junctions with the basement membrane and obtain higher migration and invasion, antiapoptotic and extracellular matrix degrading abilities and other mesenchymal phenotypes. Tumor cells with EMT acquire the expression of mesenchymal components and manifest a migratory phenotype. Thus, EMT can facilitate tumor cells to become loose and disseminate from the primary tumors.\textsuperscript{12,13} On the other hand, it has been reported that STAS could be related to tumor budding.\textsuperscript{19} Concerning tumor budding, it has been reported that tumor budding reflects invasive tumor behavior and is an adverse prognostic factor.\textsuperscript{30-33} Tumor budding is thought to be closely related

| Characteristic, factor | RFS | P Value | HR | 95% CI | P Value | OS | HR | 95% CI |
|------------------------|-----|---------|----|--------|---------|----|----|--------|
| Pathologic stage (per stage) | 1.286 | .012 | 1.056-1.564 | .1387 | .01 | 1.148-1.676 | .817 | 1.082-2.635 |
| Twist, overexpression | 1.274 | .451 | 0.678-2.396 | 1.461 | .01 | 0.810-2.635 | .209 | 0.900-2.607 |
| Slug, overexpression | 1.117 | .724 | 0.604-2.060 | 1.214 | .124 | 0.680-2.607 | .512 | 0.680-2.607 |

Abbreviations: CI, confidence interval; HR, hazard ratio; RFS, recurrence-free survival; STAS, spread through air spaces.
to EMT because the molecular mechanism that leads to budding may be, in part, the same as that of EMT, thereby increasing cancer cell migration and invasion. It was recently determined that the presence of tumor budding was significantly associated with the downregulation of E-cadherin and the acquisition of vimentin expression in cancer cells. In view of the existing research, STAS may be related to EMT. Furthermore, because EMT and STAS are largely restricted to the interface between the malignant epithelium and stromal elements at the invasive margin, an induction of EMT may trigger STAS.

A hallmark of EMT is the loss of E-cadherin expression, which is consistently observed at sites of EMT during cancer. E-cadherin transcription factors, such as Twist and Slug, can induce EMT. Twist is an inducer of EMT and is correlated with poor survival. Twist overexpression has been found among tumor tissues. When Twist expression is significantly inhibited, the metastatic potential of cells can be impaired. Moreover, a previous study showed that Twist overexpression could decrease OS in patients with lung cancer. On the other hand, Twist overexpression also decreased RFS. Slug is a member of the zinc finger Snail family. Slug is a significant EMT inducer that has been implicated in the progression of lung cancer. When overexpressed, Slug abrogates E-cadherin-mediated intercellular adhesion and promotes tumor cell invasion. Slug overexpression was found to be correlated with both poor disease-free survival and OS in lung cancer. In this study, Slug overexpression was significantly related to high pathologic differentiation, a high pathologic T stage, the presence of VPI, and increased lymph node metastasis. Furthermore, Twist overexpression showed a trend with high pathologic differentiation, increased lymph node metastasis, and the presence of VPI. Similar to other studies, we also found that Twist and Slug overexpression was associated with decreased RFS and OS in patients with lung adenocarcinoma according to the survival analysis. Thus, we can conclude that the overexpression of Slug and Twist could be related to invasive tumor behavior, such as STAS.

In our study, STAS was significantly associated with the overexpression of Slug and Twist. An additional logistic model further revealed that pathological stage, Twist overexpression and Slug overexpression were independent risk factors for STAS. It seems that the presence of STAS was related to Twist and Slug overexpression. The univariate Cox analysis showed that pathologic stage, STAS, Twist and Slug were significant risk factors for RFS and OS. One multivariate model that contained pathologic stage, STAS, Twist and Slug showed that the significantly independent prognostic risk factors of RFS and OS were STAS and pathologic stage. Considering the potential connection between STAS, Twist, and Slug, another multivariate model that contained Twist, Slug and pathologic stage (and not STAS) was applied. This multivariate analysis showed that pathologic stage, Twist and Slug were significant independent prognostic factors for RFS and OS. Thus, we inferred that STAS, Twist overexpression and Slug overexpression could influence each other in terms of prognosis. Furthermore, the potential relationship between STAS and the overexpression of Slug and Twist was confirmed in another manner.

To investigate the prognostic impact of Slug or Twist in patients with STAS, all patients were divided into several groups according to Slug, Twist or STAS. In the cohort with STAS, the multivariate analysis showed that pathologic stage and Twist overexpression were independent risk factors for poor survival. Therefore, Twist expression could serve as a stratification factor to evaluate prognosis among patients with the same pathologic stage and STAS. However, Slug overexpression was not identified as an independent risk factor in patients with STAS. This consequence may be related to STAS and Slug. Additionally, there was no correlation between STAS and Slug. Furthermore, we divided all patients into four groups according to Slug, Twist and STAS. RFS and OS became increasingly worse from group 1 to group 4. Patients with both Slug and Twist overexpression and STAS experienced a poor prognosis. Currently, we cannot clearly recognize the roles of Slug and Twist in the molecular mechanism of STAS, and we do not know the molecular mechanism of STAS and related signal transduction pathways. Our research shows that STAS is associated with Slug and Twist. Because Slug and Twist play an important role in the occurrence of EMT, STAS may have a potential relationship with EMT, and the induction of EMT may trigger STAS. Our study provides important reference information for further study of the molecular mechanism of STAS and related signal transduction pathways.

Furthermore, some studies have reported that Twist may be correlated with multidrug resistance, such as vincristine and paclitaxel, which are chemotherapeutic drugs. Furthermore, in lung adenocarcinoma with EGFR mutations, Slug could be correlated with resistance to gefitinib. However, we found that STAS was correlated with the overexpression of Twist and Slug. Therefore, STAS may be a novel treatment target by increasing drug responsiveness and sensitivity to improve prognosis.

Our study had several limitations. First, it was a retrospective study of a small sample size from a single institution. Selection bias was ineluctable. Second, we measured the expression levels of Slug and Twist only by immunohistochemistry because the tissue was previously embedded in paraffin. Therefore, it is necessary to further demonstrate the relationship between Twist, Slug and STAS and the molecular mechanism of STAS and related signal transduction pathways in additional multicenter prospective studies with larger samples with additional cell biology methods. Third, the related driver gene mutations and postoperative records of targeted therapy remain unknown.
REFERENCES

1. Travis WDBE, Burke AP, Marx A, Nicholson AG. *WHO Classification of Tumours of the Lung, Pleura, Thymus and Heart*. Lyon: International Agency for Research on Cancer; 2015.

2. Kadota K, Nitadori J-I, Sima CS, et al. Tumor spread through air spaces is an important pattern of invasion and impacts the frequency and location of recurrences after limited resection for small stage I lung adenocarcinomas. *J Thorac Oncol*. 2015;10(5):806-814.

3. Warth A, Muley T, Kossakowski CA, et al. Prognostic impact of the overexpression of Twist and Slug, which could potentially provide information on the mechanism of STAS.

4. Dai C, Xie H, Su H, et al. Tumor spread through air spaces affects the recurrence and overall survival in patients with lung adenocarcinoma >2 to 3 cm. *J Thorac Oncol*. 2017;12(7):1052-1060.

5. Uruha H, Fujii T, Fujimori S, Kohno T, Kishi K. Semiquantitative assessment of tumor spread through air spaces (STAS) in early-stage lung adenocarcinomas. *J Thor Oncol*. 2017;12(7):1046-1051.

6. Masai K, Sakurai H, Sukeka A, et al. Prognostic impact of margin distance and tumor spread through air spaces in limited resection for primary lung cancer. *J Thorac Oncol*. 2017;12(12):1788-1797.

7. Lee JS, Kim EK, Kim M, Shim HS. Genetic and clinicopathologic characteristics of lung adenocarcinoma with tumor spread through air spaces. *Lung Cancer*. 2018;123:121-126.

8. Morimoto J, Nakajima T, Suzuki H, et al. Impact of free tumor clusters on prognosis after resection of pulmonary adenocarcinoma. *J Thoracic Cardiovasc Surg*. 2016;152(1):64-72.e1.

9. Liu AO, Hou F, Qin YL, et al. Predictive value of a prognostic model based on pathologic features in lung invasive adenocarcinoma. *J Thorac Oncol*. 2017;12(2):223-234.

10. Liu AO, Hou F, Qin YL, et al. Predictive value of a prognostic model based on pathologic features in lung invasive adenocarcinoma. *Am J Surg Pathol*. 2018;42(2):223-234.

11. Liu Y, Chen D, Qiu X, et al. Relationship between MTA1 and spread through air space and their joint influence on prognosis of patients with stage I-III lung adenocarcinoma. *Lung Cancer*. 2018;124:211-218.

12. Kang Y, Massague J. Epithelial-mesenchymal transitions: twist in development and metastasis. *Cell*. 2004;118(3):277-279.

13. Thiery JP, Acloque H, Huang HY, Nieto MA. Epithelial-mesenchymal transitions in development and disease. *Cell*. 2009;139(5):871-890.

14. Mani SA, Guo W, Liao M-J, et al. The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell*. 2008;133(4):704-715.

15. Yang J, Mani SA, Donaher JL, et al. Twist, a master regulator of morphogenesis, plays an essential role in tumor metastasis. *Cell*. 2004;117(7):927-939.

16. Thiery JP. Epithelial-mesenchymal transitions in tumour progression. *Nat Rev Cancer*. 2002;2(6):442-454.

17. Yoshizawa A, Fukuoka J, Shimizu S, et al. Overexpression of phospho-eIF4E is associated with survival through AKT pathway in non-small cell lung cancer. *Clin Cancer Res*. 2010;16(1):240-248.

18. Lu S, Tan KS, Kadota K, et al. Spread through Air Spaces (STAS) is an independent predictor of recurrence and lung cancer-specific death in squamous cell carcinoma. *J Thoracic Oncol*. 2017;12(2):223-234.

19. Kadota K, Kushida Y, Katsuki N, et al. Tumor spread through air spaces is an independent predictor of recurrence-free survival in patients with resected lung squamous cell carcinoma. *Am J Surg Pathol*. 2017;41(8):1077-1086.

20. Yanagawa N, Shiono S, Endo M, Ogata SY. Tumor spread through air spaces is a useful predictor of recurrence and prognosis in stage I lung squamous cell carcinoma, but not in stage II and III. *Lung Cancer*. 2018;120:14-21.

21. Aly RG, Rekhtman N, Li X, et al. Spread Through Air Spaces (STAS) is prognostic in atypical carcinoid, large cell neuroendocrine carcinoma, and small cell carcinoma of the lung. *J Thoracic Oncol*. 2019;14(9):1583-1593.

22. Yokoyama S, Murakami T, Tao H, et al. Tumor spread through air spaces identifies a distinct subgroup with poor prognosis in surgically resected lung pleomorphic carcinoma. *Chest*. 2018.

23. Shiono S, Endo M, Suzuki K, Yarimizu K, Hayasaka K, Yanagawa N. Spread through air spaces is a prognostic factor in sublobar resection of non-small cell lung cancer. *Annals Thoracic Surg*. 2018.

24. Toyokawa G, Yamada Y, Tagawa T, Oda Y. Significance of spread through air spaces in early-stage lung adenocarcinomas undergoing limited resection. *Thorac Cancer*. 2018;9(10):1255-1261.

25. Toyokawa G, Yamada Y, Tagawa T, et al. Computed tomography features of resected lung adenocarcinomas with spread through air spaces. *J Thorac Cardiovasc Surg*. 2018;156(4):1670-1676.e4.

26. Bertolini G, D’Amico L, Moro M, et al. Microenvironment-modulated metastatic CD133+/CXCR4+/EpCAM+ lung cancer-initiating cells sustain tumor dissemination and correlate with poor prognosis. *Cancer Res*. 2015;75(17):3636-3649.

27. Trpiz M, McLaughlin PM, de Leij LM, Harnsen MC. Epithelial cell adhesion molecule: more than a carcinoma marker and adhesion molecule. *Am J Pathol*. 2007;171(2):386-395.

28. Kamiya K, Hayashi Y, Douguchi J, et al. Histopathological features and prognostic significance of the micropapillary pattern in lung adenocarcinoma. *Mod Pathol*. 2008;21(8):992-1001.
29. Jin Y, Sun P-L, Park SY, et al. Frequent aerogenous spread with decreased E-cadherin expression of ROS1-rearranged lung cancer predicts poor disease-free survival. *Lung Cancer*. 2015;89(3):343-349.

30. Kadota K, Miyai Y, Katsuki N, et al. A grading system combining tumor budding and nuclear diameter predicts prognosis in resected lung squamous cell carcinoma. *Am J Surg Pathol*. 2017;41(6):750-760.

31. Weichert W, Kossakowski C, Harms A, et al. Proposal of a prognostically relevant grading scheme for pulmonary squamous cell carcinoma. *Eur Respir J*. 2016;47(3):938-946.

32. Kadota K, Nitadori J-I, Woo KM, et al. Comprehensive pathological analyses in lung squamous cell carcinoma: single cell invasion, nuclear diameter, and tumor budding are independent prognostic factors for worse outcomes. *J Thorac Oncol*. 2014;9(8):1126-1139.

33. Yamaguchi Y, Ishii G, Kojima M, et al. Histopathologic features of the tumor budding in adenocarcinoma of the lung: tumor budding as an index to predict the potential aggressiveness. Journal of thoracic oncology : official publication of the International Association for the Study of. *Lung Cancer*. 2010;5(9):1361-1368.

34. Prall F. Tumour budding in colorectal carcinoma. *Histopathology*. 2007;50(1):151-162.

35. Kalluri R, Weinberg RA. The basics of epithelial-mesenchymal transition. *J Clin Invest*. 2009;119(6):1420-1428.

36. Taira T, Ishii G, Nagai K, et al. Characterization of the immunophenotype of the tumor budding and its prognostic implications in squamous cell carcinoma of the lung. *Lung Cancer*. 2012;76(3):423-430.

37. Hung JJ, Yang MH, Hsu HS, Hsu WH, Liu JS, Wu KJ. Prognostic significance of hypoxia-inducible factor-1alpha, TWIST1 and Snail expression in resectable non-small cell lung cancer. *Thorax*. 2009;64(12):1082-1089.

38. Jiang W, Pang XG, Wang Q, Shen YX, Chen XK, Xi JJ. Prognostic role of Twist, Slug, and Foxc2 expression in stage I non-small-cell lung cancer after curative resection. *Clin Lung Cancer*. 2012;13(4):280-287.

39. Yang J. Weinberg RA. Epithelial-mesenchymal transition: at the crossroads of development and tumor metastasis. *Dev Cell*. 2008;14(6):818-829.

40. Shih JY, Tsai MF, Chang TH, et al. Transcription repressor slug promotes carcinoma invasion and predicts outcome of patients with lung adenocarcinoma. *Clin Cancer Res*. 2005;11(22):8070-8078.

41. Wang S-P, Wang W-L, Chang Y-L, et al. p53 controls cancer cell invasion by inducing the MDM2-mediated degradation of Slug. *Nat Cell Biol*. 2009;11(6):694-704.

42. Shih JY, Yang PC. The EMT regulator slug and lung carcinogenesis. *Carcinogenesis*. 2011;32(9):1299-1304.

43. Chiu S-H, Wang M-L, Chou Y-T, et al. Coexpression of Oct4 and Nanog enhances malignancy in lung adenocarcinoma by inducing cancer stem cell-like properties and epithelial-mesenchymal transdifferentiation. *Cancer Res*. 2010;70(24):10433-10444.

44. Wang X, Ling MT, Guan X-Y, et al. Identification of a novel function of TWIST, a bHLH protein, in the development of acquired taxol resistance in human cancer cells. *Oncogene*. 2004;23(2):474-482.

45. Cheng GZ, Chan J, Wang Q, Zhang W, Sun CD, Wang LH. Twist transcriptionally up-regulates AKT2 in breast cancer cells leading to increased migration, invasion, and resistance to paclitaxel. *Cancer Res*. 2007;67(5):1979-1987.

46. Li Q-Q, Xu J-D, Wang W-J, et al. Twist-mediated adriamycin-induced epithelial-mesenchymal transition relates to multidrug resistance and invasive potential in breast cancer cells. *Clin Cancer Res*. 2009;15(8):2657-2665.

47. Chang T-H, Tsai M-F, Su K-Y, et al. Slug confers resistance to the epidermal growth factor receptor tyrosine kinase inhibitor. *Am J Respir Crit Care Med*. 2011;183(8):1071-1079.

How to cite this article: Liu A, Sun X, Xu J, et al. Relevance and prognostic ability of Twist, Slug and tumour spread through air spaces in lung adenocarcinoma. *Cancer Med*. 2020;9:1986–1998. [https://doi.org/10.1002/cam4.2858](https://doi.org/10.1002/cam4.2858)