A phase I/II pharmacokinetics/pharmacodynamics study of irinotecan combined with S-1 for recurrent/metastatic breast cancer in patients with selected UGT1A1 genotypes (the JBCRG-M01 study)

Hiroshi Ishiguro1, Shigehira Saji2, Shogo Nomura3, Sunao Tanaka4, Takayuki Ueno5, Masahide Onoue6, Hiroji Iwata7, Takeharu Yamanaka8, Yasutsuna Sasaki9 & Masakazu Toi4

1Department of Medical Oncology, International University of Health and Welfare Hospital, Nasushiobara, Japan
2Department of Medical Oncology, Fukushima Medical University, Fukushima, Japan
3Center for Research and Administration and Support, National Cancer Center, Kashiwa, Japan
4Department of Breast Surgery, Graduate School of Medicine, Kyoto University, Kyoto, Japan
5Department of Breast Surgery, Hyogo College of Medicine, Nishinomiya, Japan
6Department of Pharmacy, Kitano Hospital, Osaka, Japan
7Department of Breast Oncology, Aichi Cancer Center, Nagoya, Japan
8Department of Biostatistics, Yokohama City University, Yokohama, Japan
9Department of Medical Oncology, Showa University, Tokyo, Japan

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Abstract
S-1 and irinotecan combination is attractive for breast cancer refractory to anthracyclines and taxanes. Patients with advanced human epidermal growth factor receptor 2 (HER2)-negative breast cancer previously treated with anthracyclines and taxanes were eligible. Patients with brain metastases and homozygous for UGT1A1 *6 or *28 or compound heterozygous (*6/*28) were excluded. A dose-escalation design was chosen for the phase I portion (level 1: irinotecan 80 mg/m² days 1–8 and S-1 80 mg/m² days 1–14 every 3 weeks; level 2: irinotecan 100 mg/m² and S-1 80 mg/m²). Study objectives included determination of the recommended dose for phase II, response rate, progression-free survival (PFS), and safety. Pharmacokinetics and CD34+ circulating endothelial cells (CECs) as pharmacodynamics were also analyzed. Thirty-seven patients were included. One patient at each level developed dose-limiting toxicities; therefore, level 2 was the recommended dose for phase II. Diarrhea was more common in patients possessing a *6 or *28 allele compared with wild-type homozygous patients (46% and 25%). Among 29 patients treated at level 2, PFS was longer for UGT1A1 wt/*6 and wt/*28 patients than for wt/wt patients (12 vs. 8 months, P = 0.06). PFS was significantly longer in patients with a larger-than-median SN-38 area under the curve (AUC) than in those with a smaller AUC (P = 0.039). There was an association between clinical benefit and reduction in baseline CD34+ CECs by S-1 (P = 0.047). The combination of irinotecan and S-1 is effective and warrants further investigation.

Introduction
There is an increasing demand for effective regimens for patients with recurrent/metastatic HER2-negative breast cancer for whom anthracyclines and taxanes have failed. S-1 is an oral anticancer agent that contains gimeracil, oteracil, and tegafur (a prodrug of 5-FU) which has been shown to be active against breast cancer [1]. Irinotecan exhibits an antitumor effect against recurrent/metastatic breast cancer that has been pretreated with anthracyclines and taxanes [2, 3]. Because S-1 and irinotecan have different mechanisms of action to those of anthracyclines
and taxanes, and a synergistic effect of 5-FU and irinotecan has been noted [4], their combination is attractive. Furthermore, intratumoral dihydropyrimidine dehydrogenases (DPDs) are often elevated in breast cancer, and the incorporation of a DPD inhibitor, as in S-1, might be advantageous [5].

SN-38 is an active metabolite of irinotecan, that is, excreted after conjugation to form the inactive compound SN-38G with glucuronic acid by uridine diphosphate (UDP)-glucuronosyl transferase (UGT) 1A1. Variants of the gene for UGT1A1 exist, and in patients with certain polymorphisms (those who are homozygous for UGT1A1*6/*6 or UGT1A1*28/*28, or heterozygous, UGT1A1*6/*28) glucuronidation is impaired, resulting in an increase in the risk of serious adverse reactions to irinotecan [6–8].

Circulating endothelial cells (CECs) comprise two distinct populations: vascular-derived mature CECs, elicited by damage to the endothelium of the vasculature, and bone-marrow-derived circulating endothelial progenitors (CEPs), which contribute to neovascularization. Although CD34 is expressed not only in endothelial progenitor cells but also in some of the mature endothelial cells, it has been widely used to identify progenitor cells with clonogenic potential [9]. A reduction in CEPs is strongly correlated with antiangiogenic effects [10]. Because vascular endothelial growth factor-A-driven tumor angiogenesis for the formation of a functional vascular bed and the subsequent tumor growth partly depend on the mobilization of CEPs, a change in the CEP level may be a predictive marker for angiogenesis therapy [11]. In our previous studies [12, 13], the CD34+ CEC level was closely associated with the treatment response to chemotherapy, including S-1.

We therefore conducted a phase I/II study of combined therapy with irinotecan plus S-1 in patients with recurrent/metastatic breast cancer who had already received both anthracyclines and taxanes, including analyses of pharmacokinetics, pharmacogenomics, and CECs/CEPs as potential pharmacodynamic markers.

Methods

Patients

Patients were included if they had pathologically confirmed recurrent and/or metastatic human epidermal growth factor receptor 2 (HER2)-negative breast cancer, were aged 75 or younger and had previously received anthracycline and taxane chemotherapy (including adjuvant and/or neo-adjuvant use). All patients were required to have a measurable lesion according to the Response Evaluation Criteria in Solid Tumors (RECIST) guideline version 1.0 [14]. Other criteria included an Eastern Cooperative Oncology Group performance status of 0–2, expected survival of longer than 12 weeks, adequate organ function, a white blood cell count of ≥3000/mm³ or an absolute neutrophil count of ≥1500/mm³, a platelet count of ≥10 × 10⁹/mm³, a hemoglobin level of ≥9.0 g/dL, aspartate transaminase (AST)/alanine aminotransferase (ALT) levels of ≤2 × the upper normal limit, total bilirubin ≤1.5 mg/dL, a creatinine level of ≤1.2 mg/dL, estimated creatinine clearance of ≥50 mL/min, and oxygen saturation >90%. All patients provided written informed consent. Patients were excluded if they had brain metastases, diarrhea, ileus, clinically significant infection, a significant amount of fluid retention, interstitial pneumonitis, or another clinically significant condition. Eligible patients were preregistered and a test for UGT1A1 genetic polymorphism was performed. Patients who were homozygous for UGT1A1*6 or *28, or compound heterozygous (6/*28), were excluded.

Study design

This was a phase I/II, multicenter clinical trial. Phase I was a dose-escalation study using the standard 3 + 3 design. Patients received 80 (level 1) or 100 mg/m² (level 2) of intravenous irinotecan on days 1 and 8 plus 80 mg/m²/day of oral S-1 from day 1 through day 14. Cycles were repeated every 21 days until the prespecified discontinuation criteria was met. Dose-limiting toxicities were defined as ≥grade 3 nonhematological toxicities, grade 4 neutropenia, or febrile neutropenia during the first cycle. The maximum tolerated dose (MTD) was defined as the dose at which more than one patient in every three patients experienced dose-limiting toxicities. The dose one level lower than the MTD (or level 2 if no MTD was observed) was defined as the recommended dose.

The primary endpoint of phase II was response and clinical benefit rate. Secondary endpoints included safety, progression-free survival (PFS), and overall survival (OS). The study protocol was approved by the institutional review board at each participating study center before initiation and was conducted in accordance with the Ethical Guideline for Clinical Studies and Human Genome/Gene Analysis Research by the Ministry of Health, Labour and Welfare, Japan and the Declaration of Helsinki. The study was registered in the University Hospital Medical Information Network Clinical Trial Registry managed by the National University Hospital Council of Japan and operated by the University of Tokyo Hospital (UMIN000000517). All patient information forms were collected and managed by the Japan Breast Cancer Research Group (JBCRG) data center.

Study assessment

Adverse events were graded according to the National Cancer Institute Common Terminology Criteria for Adverse Events,
version 3.0. Tumor assessment was conducted every two cycles. Response was assessed based on RECIST version 1.0. Partial response (PR) and complete response (CR) were confirmed by repeated assessment at least 4 weeks apart. Stable disease (SD) was evaluated at least 8 weeks after study treatment began; if it continued beyond 24 weeks, it was defined as long SD. Clinical benefit was defined as either CR, PR, or long SD. PFS was defined as the time between treatment initiation and disease progression or death, whichever occurred first. Response duration was defined as the PFS only of patients whose disease responded. OS was defined as the time between treatment initiation and death due to any cause. PFS and OS were censored at the last day of tumor or survival assessment, respectively.

Pharmacogenomics, pharmacokinetics, and pharmacodynamics
Pharmacogenomics (PGx) analysis for UGT1A1 *6 and *28 was conducted by Bio Medical Laboratories, Inc. The pharmacokinetics (PK) of irinotecan were assessed during the first cycle for patients enrolled at Kyoto University Hospital, with additional informed consent. Blood samples were obtained prior to irinotecan infusion and at 10, 60, 120, and 240 min after the end of infusion. Plasma concentrations of irinotecan, SN-38, and SN-38G were measured by high-performance liquid chromatography at ADME & Tox. Research Institute, Sekisui Medical Co. Ltd, Ibaraki, Japan. PK parameters were calculated with WinNonlin Professional ver. 5.0.1 software (Pharsight Co., Mountain View, CA, USA) using a linear trapezoidal model. For pharmacodynamics (PD) analysis, we used CD34+ CECs as a marker for CEPs. CD34+ CECs were measured using a CellSearch system (Veridex LLC, Co. Ltd, NJ, USA). CECs were classified as 4,6-diamino-2-phenylindole (DAPI)+, CD45−, CD146+, or CD105+ and were subsequently stained with an anti-CD34 antibody and evaluated with an extra channel in the CellSearch system.

Statistical analysis
The main objective of the phase II portion of the study was to test the null hypothesis that the response rate (RR) is 10% against the alternative of 30% using a single-stage binomial study design. An RR of <10% was deemed unacceptable based on previous data from a study of S-1 [1]. At least 33 patients were required in phase II with a one-sided alpha of 0.05 and a beta of 0.2. The Kaplan–Meier method was used to estimate survival curves, and the log-rank test and univariate Cox proportional hazards model were used to evaluate the association between UGT1A1 genetic polymorphisms and survival outcomes. The Mann-Whitney U-test was used to compare CD34+ CEC counts during combination treatment. All reported P values were two-sided, P < 0.05 was considered statistically significant, and 90% confidence intervals (CIs) were calculated with the Clopper and Pearson method. Statistical analyses were performed with SAS Release 9.3 (SAS Institute Inc., Cary, NC, USA) and JMP v9.0.0 software (SAS Institute Inc., NC, USA).

Results
Patient population, clinical endpoints, and pharmacogenomics
Thirty-seven patients were enrolled in this study (13 in phase I and 24 in phase II) between November 2006 and June 2011 (the trial closed before full enrollment due to poor accrual) (Fig. 1). One patient was excluded from each phase, leaving 35 patients for analysis (12 for phase I and 23 for phase II). All patients had received two or more previous chemotherapies other than anthracyclines and taxanes and the majority had visceral metastases such as hepatic disease (Table 1).

The median number of irinotecan cycles and dose intensity were not different between patients with one of the UGT1A1 polymorphisms and homozygous wild-type patients (Table 2).

Among the 29 patients treated at level 2, RR, clinical benefit rate (CR + PR + long SD), PFS, and OS were numerically better for patients who were heterozygous for *6 or *28 compared with those for patients who were wild-type homozygous (Table 3). Figures 2A and B show Kaplan–Meyer curves by UGT1A1 genotype for PFS and OS at treatment level 2. Although not statistically significant, the median PFS (12 vs. 8 months, hazard ratio [HR] 0.47, P = 0.060, Fig. 2C) and median OS (23 vs. 17 months, HR 0.74, P = 0.56) were longer for the heterozygous patients compared with the homozygous wild-type patients, respectively.

Diarrhea was more common in patients possessing a *6 or *28 allele compared with wild-type homozygous patients (Table 4).

Pharmacokinetics and pharmacodynamics
For 16 patients, the PK of irinotecan, SN-38 and SN-38G were assessed during the first cycle on days 1 and 8 (five patients did not receive irinotecan on day 8 due to not meeting administration criteria and one did not provide samples for PK analysis on day 1 due to poor venous access). The level of the active metabolite SN-38 was constant between day 1 and day 8 of the first treatment cycle and significantly lower in the five patients who could not receive the irinotecan dose on day 8 (Fig. 3). PFS
was significantly longer in patients with a larger-than-median SN-38 AUC \( (P = 0.039) \) than in those with a smaller AUC (Fig. 2D).

Ten patients had CD34+ CEC analysis during the first cycle. There was a transient increase in CD34+ CEC levels on days 1 and 8 shortly after administering irinotecan (Fig. 4A and B). Baseline levels (just before administering irinotecan) and peak levels (within 4 h of administration) were both lower on day 8 compared with day 1 (Fig. 5A and B, \( P = 0.027, P = 0.063 \), respectively). There was an association between clinical benefit and reduction in baseline CD34+ CECs by S-1 (Fig. 5C, \( N = 10, P = 0.047 \)).

**Discussion**

This study not only determined the recommended dose and safety/efficacy profile of the combination of S-1 and irinotecan but also elucidated the PGx, PK, and PD (CD34+ CECs). The recommended dose was set at level 2 for all patients with no significant difference in the safety profiles between genetic variants. However, a higher RR and a
longer PFS were seen in patients who were heterozygous for a \textit{UGT1A1} mutation compared with homozygous wild-type patients. This was supported by the PK analysis in which patients with a larger SN-38 AUC had a significantly longer PFS. In terms of PD, association between clinical benefit and reduction in baseline CD34+ CECs by S-1 was demonstrated.

This study had several limitations. First, it was not possible to complete enrollment because of slow accrual. Despite the small sample size, however, the RR at level
was above the threshold level and associations between the efficacy endpoints and PGx/PK were demonstrated. Second, the PK and PD analyses were conducted in a limited number of patients since the sample collection and analysis was possible only at Kyoto University Hospital. Third, we could not conduct PK analysis of S-1 because it was not possible to perform such a complex sampling in an outpatient setting. Finally, S-1 may not be available in several countries, including the United States; therefore, the results may not be generalizable to these countries.

Several studies have shown associations between UGT1A1 polymorphisms and the toxicity of irinotecan, particularly neutropenia [7, 8, 15]. In this study, the combination with S-1 and a small sample size make analysis difficult. Although diarrhea is another characteristic toxicity of irinotecan, the degree of baseline constipation, a combination with S-1 and the use of 5HT3 antagonist make this analysis difficult with a limited number of patients. A few studies have indicated higher efficacy for patients with UGT1A1 polymorphisms, with small differences in the

| Dose level | Level 1 | Level 2 | Phase II (all cycles) |
|------------|---------|---------|-----------------------|
| UGT1A1 genotype | wt/wt (N = 4) | wt/mu (N = 2) | wt/wt (N = 3) | wt/mu (N = 3) | wt/wt (N = 12) | wt/mu (N = 11) |
| Diarrhea (%) | 1 (25) | 0 (0) | 0 (0) | 0 (0) | 3 (25) | 5 (46) |
| Constipation (%) | 3 (75) | 0 (0) | 0 (0) | 0 (0) | 1 (8) | 1 (9) |
| Vomiting (%) | 0 (0) | 0 (0) | 0 (0) | 1 (33) | 2 (17) | 1 (9) |
| Anorexia (%) | 0 (0) | 0 (0) | 0 (0) | 0 (0) | 3 (25) | 1 (9) |
| Fatigue (%) | 1 (25) | 0 (0) | 0 (0) | 0 (0) | 2 (17) | 0 (0) |
| Febrile neutropenia (%) | 0 (0) | 0 (0) | 0 (0) | 0 (0) | 0 (0) | 0 (0) |

wt, wild-type for UGT1A1; mu; *6 or *28 for UGT1A1.
Our PK analysis supported these polymorphisms being functionally relevant. It is possible that patients without UGT1A1 polymorphisms may require a higher dose of irinotecan.

For patients with disease refractory to anthracyclines and taxanes, single agent chemotherapy with agents such as irinotecan delivered at the MTD given every few weeks is an option for patients with low tumor burden, but PFS is relatively short and combination chemotherapy strategies are largely not successful [17]. We have previously reported that a low baseline CD34+ CEC level was associated with higher pathological CR in primary breast cancer [12]. Cytotoxic chemotherapy increases the CD34+ CEC count, possibly owing to the damage to normal vessels [12]. Chemotherapy can be more frequently administered at low doses, resulting in fewer side effects and a better quality of life; this is known as metronomic chemotherapy. Capecitabine and S-1 meet the characteristics of metronomic chemotherapy. This mode of administration suppresses the induction of endothelial cells from the bone marrow [18] and decreases CD34+ CEC levels [13]. There could be a good rationale for combining

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**Figure 5.** (A) Comparison of the baseline CD34+ CEC level before irinotecan administration between days 1 and 8. (B) Comparison of the peak CD34+ CEC level after administration of irinotecan between days 1 and 8. (C) Association between reduction in CD34+ CEC level and clinical benefit (CB).
metronomic chemotherapy with intravenous chemotherapy, such as taxanes [19] and irinotecan.

In conclusion, S-1 and irinotecan combination therapy is well tolerated with a modest RR but a promising clinical benefit rate and PFS. PGx analysis not only helps to avoid significant irinotecan-related toxicity, as previously reported, but also might identify patients who are more likely to benefit from such a treatment.

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**Conflict of Interest**

S. Saji, T. Yamanaka, and Y. Sasaki report personal fees from TAIHO PHARMACEUTICAL CO., LTD. H. Ishiguro, S. Saji, and M. Toi report grants from TAIHO PHARMACEUTICAL CO., LTD.

H. Ishiguro, S. Saji, H. Iwata, and M. Toi are board members of JBCRG. The other authors have declared no conflicts of interest.

**References**

1. Saek, T., S. Takashima, M. Sano, N. Horikoshi, S. Miura, S. Shimizu, et al. 2004. A phase II study of S-1 in patients with metastatic breast cancer—a Japanese trial by the S-1 Cooperative Study Group, Breast Cancer Working Group. Breast Cancer 11:194–202.

2. Taguchi, T., T. Tominaga, M. Ogawa, T. Ishida, K. Morimoto, and N. Ogawa. 1994. A late phase II study of CPT-11 (irinotecan) in advanced breast cancer. CPT-11 Study Group on Breast Cancer. Gan To Kagaku Ryoho 21:1017–1024.

3. Perez, E. A., D. W. Hillman, J. A. Mailliard, J. N. Ingle, J. M. Ryan, T. R. Fitch, et al. 2004. Randomized phase II study of two irinotecan schedules for patients with metastatic breast cancer refractory to an anthracycline, a taxane, or both. J. Clin. Oncol. 22:2849–2855.

4. Cao, S., and Y. M. Rustum. 2000. Synergistic antitumor activity of irinotecan in combination with 5-fluouracil in rats bearing advanced colorectal cancer: role of drug sequence and dose. Can. Res. 60:3717–3721.

5. Fukushima, M., M. Morita, K. Ikeda, and S. Nagayama. 2003. Population study of expression of thymidylate synthase and dihydropyrimidine dehydrogenase in patients with solid tumors. Int. J. Mol. Med. 12:839–844.

6. Ando, Y., H. Saka, M. Ando, T. Sawa, K. Muro, H. Ueoka, et al. 2000. Polymorphisms of UDP-glucuronosyltransferase gene and irinotecan toxicity: a pharmacogenetic analysis. Can. Res. 60:6921–6926.

7. Innocenti, F., S. D. Undevia, L. Iyer, P. Xian Chen, S. Das, M. Kocherginsky, et al. 2004. Genetic variants in the UDP-glucuronosyltransferase 1A1 gene predict the risk of severe neutropenia of irinotecan. J. Clin. Oncol. 22:1382–1388.

8. Minami, H., K. Sai, M. Saeki, Y. Saito, S. Ozawa, K. Suzuki, et al. 2007. Irinotecan pharmacokinetics/pharmacodynamics and UGT1A genetic polymorphisms in Japanese: roles of UGT1A1*6 and *28. Pharmacogenet. Genomics 17:497–504.

9. Bertolini, F., Y. Shaked, P. Mancuso, and R. S. Kerbel. 2006. The multifaceted circulating endothelial cell in cancer: towards marker and target identification. Nat. Rev. Cancer 6:835–845.

10. Shaked, Y., U. Emmenegger, S. Man, D. Cervi, F. Bertolini, Y. Ben-David, et al. 2005. Optimal biologic dose of metronomic chemotherapy regimens is associated with maximum antiangiogenic activity. Blood 106:3058–3061.

11. Lambrechts, D., H. J. Lenz, S. de Haas, P. Carmeliet, and S. J. Scherer. 2013. Markers of response for the antiangiogenic agent bevacizumab. J. Clin. Oncol. 31:1219–1230.

12. Ali, A. M., T. Ueno, S. Tanaka, M. Takada, H. Ishiguro, A. Z. Abdellah, et al. 2011. Determining circulating endothelial cells using Cell Search system during preoperative systemic chemotherapy in breast cancer patients. Eur. J. Cancer 47:2265–2272.

13. Tsuji, W., H. Ishiguro, S. Tanaka, M. Takeuchi, T. Ueno, and M. Toi. 2014. Orally administered S-1 suppresses circulating endothelial cell counts in metastatic breast cancer patients. Int. J. Clin. Oncol. 19:452–459.

14. Therasse, P., S. G. Arbuck, E. A. Eisenhauer, J. Wanders, R. S. Kaplan, L. Rubinstein, et al. 2000. New guidelines to evaluate the response to treatment in solid tumors. European Organization for Research and Treatment of Cancer, National Cancer Institute of the United States, National Cancer Institute of Canada. J. Natl Cancer Inst. 92:205–216.

15. Onoue, M., T. Terada, M. Kobayashi, T. Katsura, S. Matsumoto, K. Yanagihara, et al. 2009. UGT1A1*6 polymorphism is most predictive of severe neutropenia induced by irinotecan in Japanese cancer patients. Int. J. Clin. Oncol. 14:136–142.

16. Toffoli, G., E. Cecchin, G. Corona, A. Russo, A. Buonadonna, M. D’andrea, et al. 2006. The role of UGT1A1*28 polymorphism in the pharmacodynamics and pharmacokinetics of irinotecan in patients with
metastatic colorectal cancer. J. Clin. Oncol. 24:3061–3068.

17. Dear, R. F., K. McGeechan, M. C. Jenkins, A. Barratt, M. H. Tattersall, and N. Wilcken. 2013. Combination versus sequential single agent chemotherapy for metastatic breast cancer. Cochrane Database Syst. Rev. 12:CD008792.

18. Bertolini, F., S. Paul, P. Mancuso, S. Monestiroli, A. Gobbi, Y. Shaked, et al. 2003. Maximum tolerable dose and low-dose metronomic chemotherapy have opposite effects on the mobilization and viability of circulating endothelial progenitor cells. Cancer Res. 63:4342–4346.

19. O’Shaughnessy, J., D. Miles, S. Vukelja, V. Moiseyenko, J. P. Ayoub, G. Cervantes, et al. 2002. Superior survival with capecitabine plus docetaxel combination therapy in anthracycline-pretreated patients with advanced breast cancer: phase III trial results. J. Clin. Oncol. 20:2812–2823.