Assessment of Potential Property of Baobab Fruit Pulp Derived Pectin as a Pharmaceutical Excipient

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ABSTRACT
The main goal about of the current study is extraction of baobab fruit peel pulp containing pectin as a potential excipient and additional characterization for its beneficial constructive choice as a alternative potent pharmaceutical excipient. The pectin used to be subjected to phytochemical screening to conformity with phytochemical then and physicochemical characterization about its safety measures and appropriateness/suitability in conformity with to be used as a disintegrating agent. FTIR-spectroscopic analysis, DSC-thermal analysis and XRD research studies were performed with subject as baobab fruit pectin powder. Piroxicam was selected as a model drug, direct compression technique was utilised as for preparation of piroxicam tablets by followed with distinct concentrations of pectin powder derived from baobab fruit and synthetic binding agent PVP k30 as a concentration of 5% w/w. The granules prepared are be subjected to assess pre compression parameters and granules are be lubricated with talc along with magnesium stearate and fallowed by compression result into the tablets. Pharmaceutical properties of pectin powder such as Hausner ratio, carr’s compressible index and critical angle of repose studies were assessed and results were determined satisfactory. In vitro dissolution study shows that release rate dimension of drug is decreased with increase in the baobab fruit peel pectin powder percentage in the formulation. Based on the above findings concluding baobab fruit peel pectin powder has showed good binding property and the study strongly suggesting that baobab fruit peel pulp containing pectin as a potential pharmaceutical excipient.

Keywords: Baobab fruit peel pectin powder, Piroxicam, FTIR, XRD and DSC.

INTRODUCTION
For quite a long time man has instituted high quality use about materials from natural origin in the characteristic cause in the therapeutics/medicinal at pharmaceutical field. Common natural origin materials hold advantages upon synthetic substances because they have points of interest over manufactured materials since they are nontoxic and non-harmful, more affordable, freely accessible, biodegradable yet safe to consumable sources. They are utilized as a thickening, much binding, suspending, emulsifying, stabilizing agents in pharmaceutical industries and utilized as matrices frameworks for sustained release of medications.

The globally most of terrestrial plants contained a multifunctional constituent like pectin within cell mass of plants. Pectin is a structural acidic non starch hetero-poly carbohydrate and it consists of major component of D-galacturonic acid with sugar acid derived from galactose. The pectins carboxyl group of D-galacturonic acid might be esterified with methanol/acetic acid, during pectin extraction this proportion may diminished to varying degrees. The percentage of esterified groups of the pectins are expressed as on basis of degree of methylation low methoxypectins (LM-Pectins) and high methoxypectins (HM-Pectins) respectively. The less than 50% esterified pectins (LM-Pectins) forms gels like thermo reversible by interaction with bivalent Ca2+ ions particularly at pH 3.0-4.5. In case of greater than 50% esterified pectins (HM-Pectins) rapidly forms gels like thermo irreversible under acidic circumstances in the attendance of high concentration of sugar at low pH than 3.5. Pectins are most predominantly utilizing as gelling agent and additionally it also acts as thickener. In discriminate of upon hearty aspects, pectin actually remains a promising pharmaceutical excipient for oral drug delivery. In the present study, we have extracted pectin from baobab fruit pulp and verified its potentials for using as disintegrating agent in the piroxicam.

MATERIALS AND METHODS
Piroxicam is acquired from Wockhardt pharmaceutical Pvt. Ltd, Mumbai, India and pectin powder was extracted from baobab fruit pulp in the laboratory. D-Mannitol [C6H12O5], Pvpk30, Talc [Mg3SiO4(OH)], Magnesium stearate [Mg(C18H34O2)2] and Lactose are of analytical grade.

Extraction of Baobab Fruit Pulp Pectin Powder
The ripped baobab fruits were acquired from local tree. The collected fruits were painstakingly washed and shade dried for 24 hrs fallowed by further drying carried in a hot
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air oven at 60°C result in dried fruit pulp. Dried fruit pulp material was slice into small pieces and made into powdered by using home electric greater. Fruit pulp powdered was further subjected to passed through a sieve no 20. The peel powder of 200gms was once dissolved in 1000ml of distilled water and additionally added 1gm of citric acid to keep up acidic pH. To extract pectin, whole prepared solution was exposed to reflux condensation for 6 hrs at 70°C. The extracted as part of an extraction unit was a whatman cellulose thimble with 80 mm external length and 33 mm internal. Warm acid extract was taken in a cheese cloth sack to squeezed result concentrated juice and cool it up to 4°C. Pectin was once precipitated through ethanol (2 v/v): water (1 v/v) treatment followed via non-stop through stirring for 15 min then permitted to kept stand for 2hrs. Coagulated Pectin was separated through cheese fabric, washes with alcohol (95%) and squeezed. Pressed pectin was additionally dried to consistent load at 35 to 45 ºC. Hard cake of pectin was ground and passed through a sieve No.60, store with in a desiccators for further utilization 6.

Preliminary phytochemical screening of baobab fruit pulp pectin

The baobab fruit pulp pectin powder was subjected to the phytochemical analysis to determine presence of phyto constituents like tannins, carbohydrates, alkaloids, glycosides, amino acids and proteins by using standard methods 7 as shown in Table no1.

Physicochemical characterization of baobab fruit pulp pectin powder

The physicochemical characterization of baobab fruit pulp pectin powder assed by soluble in water than in other solvents, percentage loss on drying, pH and ash value etc were determined as results are shown in Table no 2.

Table 1: Phyto chemical characterization of Baobab fruit pulp pectin powder

| S.no | Name of the test carried out | Test result | Inference |
|------|-------------------------------|-------------|-----------|
| 1    | Test for alkaloids             | Mayer’s test | -         | Absent    |
|      | Wanger test                    | -           | Absent    |
|      | Hager’s test                   | -           | Absent    |
| 2    | Test for carbohydrates         | Molish test  | +         | Present   |
|      | Fehling’s test                 | +           | Present   |
|      | Benedict’s test                | +           | Present   |
| 3    | Test for glycosides            | Liebermann- Burchard’s test | - | Absent |
|      | Legal’s test                   | -           | Absent    |
| 4    | Test for tannins               | Lead acetate test | - | Absent |
|      | Ferric chloride test           | -           | Absent    |
|      | Aqueous bromine test           | -           | Absent    |
| 5    | Mucilage test                  | Ruthenium red test | - | Absent |
| 6    | Test for amino acids and proteins | Ninhydrin test | - | Absent |
|      | Milons test                    | -           | Absent    |
|      | Biuret test                    | -           | Absent    |

Note: - Negative; + Positive

Table 2: Physicochemical characterization of Baobab fruit pulp pectin powder

| Parameter        | Color       | Odour         | Nature  | Solubility                                     |
|------------------|-------------|---------------|---------|-----------------------------------------------|
| Results          | Brown colour| Characteristic| Amorphous| Soluble in water Insoluble in acetone, methanol, ether and ethanol. |

FT-IR Studies

Pure drug piroxicam, baobab fruit pulp pectin powder, prepared formulations are being studied for FTIR spectra 8 are shown in Fig no 1 and 2.

Scanning Electron Microscopy

SEM studies are performed to understanding surface morphological characters of the extracted pectin powder of baobab fruit pulp as shown in Fig no 4.

Thermal analysis [DSC studies]

The DSC analytical studies are performed to know any chemical interactions in between of pure piroxicam, pectin powder of baobab fruit pulp and prepared tablet formulations.

Formulation of Piroxicam tablets

Piroxicam was used as model drug; direct compression method was used for preparation of the piroxicam tablets.
Drug and excipients are passed through the sieve no.60 individually. Piroxicam, baobab fruit pulp pectin powder, mannitol and lactose were added and mixed uniformly. The procedure was followed with different concentrations of baobab fruit pulp pectin powder and 5 %w/w concentration of PVPk30 as synthetic binding agent. The prepared granules were subjected to evaluate pre compression parameters. Granules were lubricated with magnesium stearate, talc and were compressed into the tablets of 200 mg, using 10 station rotary tablet compression machines as showed in table no.3.

### Table 3: Formulation of piroxicam tablets using different amounts of Baobab fruit pulp pectin powder

| Materials          | F-1 | F-2 | F-3 | F-4 | F-5 | F-6 | F-7 | F-8 | F-9 |
|--------------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| Drug:piroxicam     | 20mg| 20mg| 20mg| 20mg| 20mg| 20mg| 20mg| 20mg| 20mg|
| Pvpk30             | 10  | 10  | 10  | 10  | 10  | 10  | 10  | 10  | 10  |
| Xp                 | 2.50| 3.75| 5.0 | 7.25| 8.50| 9.75| 11  | 12.25| 13.5|
| Mannitol           | 133.5| 132.25| 131| 128.75| 127.5| 126.25| 125| 123.75| 122.5|
| Lactose            | 30  | 30  | 30  | 30  | 30  | 30  | 30  | 30  | 30  |
| Mg Ste             | 2   | 2   | 2   | 2   | 2   | 2   | 2   | 2   | 2   |
| Talc               | 2   | 2   | 2   | 2   | 2   | 2   | 2   | 2   | 2   |
| Total              | 200 | 200 | 200 | 200 | 200 | 200 | 200 | 200 | 200 |

### Assessment of pre-formulation studies of Piroxicam Granules

#### Angle of repose

It was assessed by using fixed funnel method and it was calculated with the formula of, Angle of Response\(\theta\) = \(\tan^{-1}\)\left[\frac{\text{height of pile (H)}}{\text{radius of the base of pile (R)}}\right]

### Compressibility Index

The compressibility index was fast and simple method to know powder flow characteristics. The compressibility index was determined by measuring both the Tapped bulk density and untapped bulk density of a powder, it was calculated by using formula:

\[
\text{Percent compressibility Index} = \left(\frac{\text{Tapped bulk density} - \text{Un Tapped bulk density}}{\text{Tapped bulk density}}\right) \times 100
\]

#### Assessment of prepared Tablets

**Weight Variation Test**

Twenty randomly chosen tablets were individually weighed and collectively in a single panoramic balance. The percentage deviation and weight average was noted was calculated.

#### Thickness and Hardness Test

Six tablets were taken in use at random and tablet hardness was calculated by using Monsanto hardness tester. The hardness was expressed in kg / cm². The diameter and thickness also measured by using of vernier caliper.

#### Friability Test

From each batch twenty tablets were selected randomly and tested at a time. Collective weight of tablets were determined prior to test (W1g) and after to the test (W2g). The percentage was calculated by the formula:

\[
\text{Percentage Friability} = \left(\frac{W1 - W2}{W1}\right) \times 100
\]

### Drug content

Individually 5 tablets were weighed and collectively powdered. The average powder equivalent of the tablets weight was weighed and the drug was extracted in a pH 6.8 phosphate buffer, there after spectro-photometrically drug content was determined. After with a suitable dilution absorbance was measured at 334 nm by using UV spectrophotometer.

### Test for Disintegration

According to I.P. method disintegration test was performed. Chosen 6 tablets were individually positioned in glass tubes of the disintegration device. The temperature of disintegration fluid was maintained at 37°C and the time duration required for disintegration was observed and noted.

### In-vitro dissolution studies

The dissolution studies of the prepared piroxicam tablets were performed by using a pH 6.8 phosphate buffer at 37°C and in case of tablets, paddle rotation speed was maintained at 50 rpm. Samples were collected in a equal intervals of every five minutes up-to 1hour and also maintained a constant volume (900ml) of the dissolution support medium up-to 1hr by replacing fresh medium at each time intervals. All Samples were spectroscopically analyzed at 334 nm by using UV Spectrophotometer.
Stability Studies

According to the ICH guide-lines stability studies were performed on optimized formulation by storing tablets at 40°C / 75% RH for 90 days.

RESULTS AND DISCUSSION

Phytochemical characterization of baobab fruit pulp pectin powder results revealed that baobab fruit pulp pectin powder showed positive result with molish test, Fehling’s test and Barford test. This demonstrates presence of carbs and diminishing sugar in the baobab fruit pulp pectin powder. The negative outcomes were observed in case of test for tannins, alkaloids, proteins, glycosides and amino acids the results are shown in table no.1.

According to the physic-chemical properties of baobab fruit pulp pectin powder is dissolvable in H₂O and insoluble in (CH₃)₂CO, indicates that’s it should not soluble in other organic solvents also. pH of baobab fruit pulp pectin powder solution showed. Microbial studies reveal that there is no microbial growth after three days study

FT-IR spectra for pure piroxicam (in the Figure 1), baobab fruit pulp pectin powder and optimized piroxicam tablet formulation (in the Figure 2) are shown. The characteristic absorption peak of pure piroxicam was retained in the spectra of a tablet formulation. It was noticed that no marked changes in the absorption peaks of model drug in the final formulation. Therefore, there is no drug interaction with pectin powder of baobab fruit pulp and also with other excipients utilized in the study.

The studies of DSC thermal history of piroxicam shown that pointed endothermic peak at 203.7°C, this indicates crystalline character of the piroxicam. Purified baobab fruit pulp pectin powder thermogram showed that baobab fruit pulp pectin powder is in amorphous nature. Endothermic peak of the piroxicam appear to be shifted to higher melting point 210°C, in fact there is merging of piroxicam peak and mannitol peak as shown in the thermograms of formulations shown in Fig no 3. Thermal analysis studies acknowledged that there was no drug interaction with excipients utilized.
The baobab fruit pectin powder particles are unsymmetrical and surface was smooth and silky has noticed. The range of the particles sizes were approximately 50-200 μm in size was determined.

Granules, angle of repose were measured to be 27° - 35° showing good quality of the flow rate of granules was noticed. The range of bulk density in between 0.28 to 0.42 gm/cc and tapped density to be in the range of 0.34 to 0.52 gm/cc were observed. Carr’s index and Hausner ratios were founded to be in the range of 11.23 to 20.01 and 1.12 to 1.26 are shown in the table no.4. The above findings strongly indicating that results of granules compression outcome was very satisfactory.

Sizes of the particles were observed to be in the range of 8 μm to 18 μm was confirmed by using microscopic technique. pH was found to be in the range of 5.29 to 6.82.

In-vitro drug release investigations of piroxicam tablets revealed that in the case of F1 with in 45 minutes 93.12% of drug was released and where as in the case of F2 formulation 96.45% of drug within 30 minutes. Within 20 min 98.34% of drug was released in case of F3 and F4, F5 formulations shown 89.67%, 95.47% of drug released within hour. As increased percentage levels of baobab fruit pulp pectin powder in the formulations showing enhancement in the means of time to releasing of drug about 98% as observed in the formulations. Formulation F3, is advanced from plan F1, F2, F4& F5 as it delivered 98% of the medication since F1, F2 and F3 didn’t pass for the friability test and didn’t possess enough hardness results are given in the table no.6 and Fig no.5.

Table 4: Pre-compression studies of prepared formulation granules of piroxicam

| Formulation-Codes | Angle repose | Bulk density (g/ml) | Tapped density (g/ml) | Percentage of Carrs index | Hausners ratio |
|-------------------|--------------|---------------------|------------------------|----------------------------|----------------|
| F-1               | 35.13±1.032  | 0.4236±1.0026       | 0.4854±1.0018          | 12.73±1.0494               | 1.14±1.0014   |
| F-2               | 35.15±1.041  | 0.4230±1.0020       | 0.4766±1.0033          | 11.23±1.1272               | 1.12±1.0035   |
| F-3               | 29.24±1.008  | 0.4127±1.0180       | 0.4821±1.0029          | 14.36±1.7566               | 1.16±1.0000   |
| F-4               | 27.47±1.027  | 0.4227±1.0038       | 0.5231±1.0253          | 19.19±1.0565               | 1.23±1.0071   |
| F-5               | 35.12±1.019  | 0.3823±1.0032       | 0.4852±1.0044          | 20.01±1.0848               | 1.26±1.0000   |
| F-6               | 34.99±1.003  | 0.3910±1.0014       | 0.4650±1.0036          | 15.90±1.3040               | 1.16±1.0070   |
| F-7               | 33.86±1.002  | 0.2896±1.0014       | 0.3449±1.0013          | 16.04±1.3676               | 1.18±1.0424   |
| F-8               | 35.23±0.001  | 0.3100±0.0035       | 0.3655±0.0031          | 15.19±0.2969               | 1.17±0.0070   |
| F-9               | 32.61±0.001  | 0.3925±0.0026       | 0.4614±0.0028          | 14.93±0.9545               | 1.16±0.0070   |

Hardness about piroxicam prepared tablets had been founded in between 4.1 kg/cm² to 4.9 kg/cm². Friability percentage range was founded in between 0.24% to 0.75% and thickness was 3.15 mm to 3.75 mm apart from above results also observed percentage of drug content and in-vitro dissolution studies results are given in the table no.4. Hence, tablets prepared with baobab fruit pulp pectin powder as disintegrating agent provided required properties to tablets.

Table 5: Post compression parameters of piroxicam tablets

| Formulation-codes | weight Average in mg | Hardness in kg/cm² | %loss of Friability | Thickness in mm | % of Drug content | In vitro disintegration Times in Sec |
|-------------------|----------------------|--------------------|---------------------|-----------------|------------------|------------------------------------|
| F1                | 148.5                | 4.2                | 0.36                | 3.69            | 98.16            | 49                                 |
| F2                | 149.3                | 4.9                | 0.24                | 3.48            | 97.62            | 36                                 |
| F3                | 147.8                | 4.6                | 0.59                | 3.15            | 99.35            | 31                                 |
| F4                | 145.6                | 4.1                | 0.37                | 3.75            | 96.28            | 15                                 |
| F5                | 149.2                | 4.7                | 0.49                | 3.61            | 97.19            | 31                                 |
| F6                | 148.8                | 4.3                | 0.36                | 3.95            | 99.25            | 28                                 |
| F7                | 147.2                | 4.2                | 0.75                | 3.47            | 99.61            | 20                                 |
| F8                | 149.2                | 4.0                | 0.48                | 3.64            | 98.18            | 18                                 |
| F9                | 146.7                | 4.8                | 0.57                | 3.18            | 97.29            | 23                                 |
Table 6: Cumulative amount of drug released from piroxicam tablets prepared with different amounts of Baobab fruit pulp pectin powder

| Time (mins) | F-1 | F-2 | F-3 | F-4 | F-5 | F-6 | F-7 | F-8 | F-9 |
|------------|-----|-----|-----|-----|-----|-----|-----|-----|-----|
| 0          | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   | 0   |
| 5          | 25.85 | 49.54 | 35.41 | 8.45 | 25.12 | 34.54 | 37.15 | 20.45 | 20.36 |
| 10         | 43.61 | 58.43 | 68.51 | 15.46 | 39.46 | 43.12 | 48.34 | 45.46 | 45.42 |
| 15         | 56.38 | 70.25 | 81.25 | 35.15 | 46.73 | 55.15 | 57.12 | 62.58 | 63.26 |
| 20         | 75.14 | 83.54 | 98.34 | 46.78 | 55.14 | 61.24 | 73.42 | 70.51 | 79.13 |
| 30         | 86.74 | 96.45 | _   | 57.46 | 70.51 | 79.31 | 82.16 | 83.61 | 94.15 |
| 45         | 93.12 | _   | _   | 70.23 | 83.61 | 91.87 | 90.02 | 96.45 | _   |
| 60         | _   | _   | _   | 89.67 | 95.47 | _   | 96.13 | _   | _   |

The stability studies for optimized formulations were carried out at 40°C/75% RH for 90 days. Hence there were no significant modifications in the drug content and also in the physical properties during the examination time frame.

CONCLUSION

The baobab fruit pulp pectin powder exhibited good disintegrating properties for the piroxicam tablets. The augmented concentration of pectin in the tablet formulations showing minute retardation in the drug release is evidence. Hence, this study strongly proved that powder of pectin extracted from baobab fruit pulp can be utilized as a pharmaceutical drug excipient.

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