Distribution and seasonal abundance of *Biomphalaria* snails and their infection status with *Schistosoma mansoni* in and around Lake Tana, northwest Ethiopia

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*Biomphalaria* snails, namely *B. pfeifferi* and *B. sudanica*, are the principal intermediate hosts for *Schistosoma mansoni* infection in Ethiopia. Epidemiological studies of *Biomphalaria* snails and their infection status with *S. mansoni* is vital for public health planning. This study aimed to assess the spatial and seasonal abundance of *Biomphalaria* snails as well as their infection status with *S. mansoni* around Lake Tana, northwest Ethiopia. Malacological survey was conducted from January 2021 to December 2021 in ten different collection sites in and around Lake Tana. Snail collection was performed for 20 min from each collection site seasonally (four times in a year) using a standard scoop and handpicking from aquatic vegetation. All collected snails were carefully examined based on their morphological features and all live *Biomphalaria* snails were subjected to cercariae shedding experiment. Descriptive statistics were used to determine the prevalence of *S. mansoni* infection and its relationship with snail collection sites and seasons. A total of 3886 freshwater snails were collected from ten collection sites around Lake Tana. Out of the total snails collected, 1606 (41.3%; 95% CI 39.77–42.89%) were *Biomphalaria* spp. The highest (374) and the lowest numbers (98) of *Biomphalaria* snails were collected from Shinne River and Qunzela Lakeshore, respectively. Out of the 1375 live *Biomphalaria* snails, 14.4% (95% CI 12.59–16.37%) snails shed cercariae, but only 4.87% (95% CI 3.79–6.15%) were cercariae of *S. mansoni*. The infection prevalence of *S. mansoni* ranged from 10.59% at the Cherechera site to 1.49% at Gumara River. *Biomphalaria* snail infections with *S. mansoni* cercariae were observed throughout the season, the highest and the lowest infection rates being in the spring and summer seasons. Significant differences in the prevalence of *S. mansoni* infection in *Biomphalaria* snails were observed across study sites and seasons (*p* < 0.05). *Biomphalaria* snails were the most abundant freshwater snails found in nearly all of snail collection sites throughout the year. It was revealed that nearly five percent of *Biomphalaria* snails were infected with *S. mansoni* cercariae. This study highlights the importance of appropriate snail control strategies to support the ongoing prevention and control of schistosomiasis around Lake Tana.

Abbreviations

IoB  Institute of biotechnology
GPS  Global positioning system
SPSS  Statistical package for social science
MASL  Meter above sea level
NTD  Neglected tropical diseases
UNESCO  United Nations Educational, Scientific and Cultural Organization
PCR  Polymerase chain reaction

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Schistosomiasis is one of the neglected tropical diseases (NTD) that is widely distributed in Africa, South America, the Middle East and Southeast Asia. The prevalence of the disease varies among regions depending on the socio-economic level, environmental conditions, human water contact behaviour of the community as well as on the level of control strategies employed in the country. The disease is severe in Africa, particularly in sub-Saharan Africa, due to the suitability of the climatic condition and socio-economic development of the region. It is estimated that 85–95% of the global schistosomiasis are in sub-Saharan Africa with the highest prevalence among school-aged children. In Ethiopia, the prevalence of schistosomiasis could reach as high as 90% in some localities, particularly for *Schistosoma mansoni*. Although Ethiopia launched a school-based deworming program in 2015 to control schistosomiasis, the prevalence of the disease is still high in several localities.

*Schistosoma mansoni* uses freshwater snails of the genus *Biomphalaria* as an intermediate host to complete its life cycle. Malacological studies have indicated the presence of several snail groups in Ethiopia. *Biomphalaria* species, namely *B. pfeifferi* and *B. sudanica*, serve as intermediate hosts for *S. mansoni*. *B. pfeifferi* and *B. sudanica* are the principal intermediate hosts for *S. mansoni* in Ethiopia. However, limited information is available about the distribution, abundance and diversity of these snails in several endemic foci of the country. The distribution of schistosomiasis in any endemic foci is directly correlated with the distribution of snail vectors. Information regarding the distribution and abundance of *Biomphalaria* snails around Lake Tana is dated back to the beginning of the 1990s. An updated and in-depth investigation of snail intermediate hosts of *S. mansoni* is vital to designing cost-effective snail control strategies in the area.

The infection prevalence of *Biomphalaria* snails with *S. mansoni* varied from 3% to 58% in Ethiopia. Our previous review showed that about 15% of *Biomphalaria* snails of Ethiopia were positive for *S. mansoni* cercariae. However, the infection status of *Biomphalaria* snails around Lake Tana has not been investigated. A recent evidence showed a high prevalence (35%) of *S. mansoni* infection in humans in the study area despite the ongoing deworming program. In the present study, it was hypothesized that there might be a high-level of *Biomphalaria* snails infected with *S. mansoni* around Lake Tana. Knowing the infection status of freshwater snails with *S. mansoni* serves as one of the important criteria to determine the transmission dynamics of *S. mansoni* in the study area. In addition, assessment of natural snail infection with *S. mansoni* is important to elucidate the level of environmental contaminations with fecal matter from humans as well as from other non-human primates.

Epidemiological studies on the abundance, distribution, and infection status of *Biomphalaria* snails are vital for policymakers to design appropriate schistosomiasis prevention and control strategies. However, there is no recent information on the abundance and distribution of *Biomphalaria* snails and their infection status around Lake Tana. Therefore, this study aimed to investigate the spatial and seasonal abundance of *Biomphalaria* snails and their infection status with *S. mansoni* in and around Lake Tana, northwestern Ethiopia.

**Material and methods**

**Description of the study areas.** Lake Tana is located in the north-western part of Ethiopia at 12°0.00' N and 37°0.14' E. Lake Tana is the largest lake in Ethiopia and the major source of the Blue Nile River. The lake consists of more than 37 islands and peninsula and some of them serve for human habitation. Lake Tana covers an area of 3020 km² and a maximum depth of 15 m. Lake Tana is rich in biodiversity with several species of birds, fish, amphibians, macro-invertebrates, and micro-invertebrates. The lake and islands on the lake serve as homes for several species of birds including the endemic ones. Lake Tana consists of 28 known species of fish, of which 68% of them are endemic. As a result of its rich biodiversity, the United Nations Educational, Scientific and Cultural Organization (UNESCO) recognized Lake Tana as a Biosphere reserve in 2015. The present study was conducted at different sites of Lake Tana shores (Dek, Cherechera, Gorgora, Zegie and Quenza) and tributary rivers of Lake Tana (Enferanz, Gumara, Gorno, Shinnie and Robit). The selection of the collection sites was based on human habitation and the frequent human-water contact behaviour of the community.

**Operational definitions of words or phrases.** Winter is a dry season in Ethiopia that spans from December to February. Spring is span from March to May. There may be occasional rain in most parts of Ethiopia. Summer is the major rainy season in Ethiopia that spans from June to August. Autumn is the major harvesting season in Ethiopia that spans from September to November.

**Study design.** Malacological surveys were conducted from January 2021 to December 2021 to assess the distribution and seasonal abundance of *Biomphalaria* snails from the shorelines of Lake Tana (Dek, Cherechera, Gorgora, Zegie and Quenza) and its tributary rivers, namely Enferanz, Gumara, Gorno, Shinnie and Robit (Fig. 1). From each site, snails were sampled from two different points at least 200 m apart. The specific sample collection sites were selected based on the frequency of human-water contact during water fetching, washing clothes, bathing, swimming, fish processing and other domestic activities. The geographical coordinate of each sampling site was taken using a global positioning system (GPS) and it was properly recorded.

**Snail collection.** Freshwater snails were collected and examined using a standard protocol as described elsewhere. Snails were sampled using standard scoops (2 mm mesh size) and forceps from water bodies or picked with gloved hands from aquatic vegetation at the shoreline of Lake Tana as well as from the rivers that fed the lake. The snail collection sites were selected based on close proximity to human settlement and high level of open defecation. The scooping was performed for 20 min from each site between 8:00 AM and 10:00 AM on a...
seasonal basis (four times a year) by the same individual. Samplings were conducted in areas about 10 m along the shorelines of Lake Tana, selected rivers, and from an area of ca. 5m² from lake water at each sampling point. Each collected snail was kept separately in a wide-mouth glass bottle filled with water and aquatic vegetation from the same area. The snail samples were transported to the Biomedical Sciences Laboratory of the Department of Biology, Bahir Dar University. All collected snails were sorted, counted and identified in the laboratory.

Morphological identification of Biomphalaria snails. All the collected snails were carefully examined based on morphological features using standard identification keys to at least a genus level as described elsewhere. The common criteria to distinguish snail species include shell shape, shell size, nature of aperture, color and banding pattern of the shell. Once the morphological identification was completed, the snails were kept at dark for 48 h and then they were used for cercariae shedding experiment.

Testing of snails for S. mansoni infections and identification. Individual snails were carefully transferred into shedding vials that contained 10 ml of natural spring water with a neutral pH. The shedding of S. mansoni and other trematodes cercariae was induced by exposure to artificial light (60 watts) for about two hours at room temperature in the morning (10:00–12:00 AM). Each snail was observed under a dissecting microscope to determine the presence of shedding trematodes cercariae. The water in the shedding vial was carefully examined for the presence of cercariae using a dissecting microscope.

Live cercariae shedding from each snail were transferred to a microscopic slide and covered with a coverslip. The cercariae were carefully observed using a microscope with 40× magnification power and identified based on their morphological features using a standard identification key. The types and the numbers of cercariae discharged from each snail were properly recorded.

Data analysis. The data generated during the study were analysed using SPSS version 23. Descriptive statistics was used to determine the proportion of Biomphalaria snails and their infection prevalence across study sites and study seasons. Analysis of variance (ANOVA) was used to determine the differences in the abundance of Biomphalaria snails across study sites and seasons. Chi-square test was used to assess the relationship between Biomphalaria snail infection with S. mansoni and studies sites and seasons. The geographic coordinate of snail collection sites was taken using a global positioning system (GPS) from each sampling point. Mapping of the snail collection site was prepared using ArcGIS online free software. For all statistical analyses, a p-value below 0.05 was used to declare statistical significance.
Ethics approval and consent to participate. This study was conducted after obtaining ethical clearance from the Ethical Review Committee of College of Science, Bahir Dar University with Ref. No. PGRC-SVD/155/2020. The objective of the study was explained to the local administration before snail collection.

Results
The study was conducted at a latitudinal range of 11.6181°–12.3909° E and a longitudinal range of 37.0343°–37.7848°N. The collection sites were classified as periphery of Lake Tana and its tributary rivers. All snails were collected from the area where there is frequent human-water contact for various activities such as washing clothes, bathing, swimming, fetching water and fish processing (Table 1). The snail collection sites had either muddy or stony substrates with clean or turbid water. Some of the images of snail collection sites are presented in Fig. 2.

Table 1. Sampling points, GPS coordinates and other basic information of the snail collection sites.

| Study area   | Sampling point | Elevation (masl) | GPS coordinate | Human activity                                      | Nature of substrate | Nature of the water | Vegetation type       | Habitat classification |
|--------------|----------------|-----------------|----------------|---------------------------------------------------|---------------------|---------------------|-----------------------|------------------------|
| Cherechera   | a              | 1793            | 11.62069       | Swimming, bathing and fish processing             | Muddy               | Turbid water         | Floating vegetation   | Lake periphery         |
|              | b              | 1789            | 11.61805       |                                                    |                     |                     |                       |                        |
| Dek Island   | a              | 1787            | 11.88602       | Bathing, washing cloth & fish processing          | Muddy               | Turbid water         | Papyrus and other floating vegetation | Lake periphery         |
|              | b              | 1788            | 11.88646       |                                                    |                     |                     |                       |                        |
| Zegie Peninsula | a         | 1787            | 11.69279       | Swimming, bathing and washing cloth                | Rocky               | Clear               | Floating vegetation   | Lake periphery         |
|              | b              | 1782            | 11.69217       |                                                    |                     |                     |                       |                        |
| Qunzela town | a              | 1806            | 11.88451       | Bathing, fetching & washing cloth                 | Muddy               | Turbid water         | Floating vegetation   | Lake periphery         |
|              | b              | 1790            | 11.88241       |                                                    |                     |                     |                       |                        |
| Gorgora Peninsula | a        | 1802            | 12.24575       | Bathing, washing cloth & fish processing          | Rocky               | Clean               | Floating vegetation   | Lake periphery         |
|              | b              | 1790            | 12.24256       |                                                    |                     |                     |                       |                        |
| Enferanz river | a           | 1820            | 11.62090       | Fetching and washing cloth                         | Muddy               | Turbid water         | Floating vegetation   | River                  |
|              | b              | 1807            | 11.62208       |                                                    |                     |                     |                       |                        |
| Shinie river | a              | 1957            | 12.13247       | Bathing & washing cloth                            | Rocky               | Clean water          | Covered with green algae | River                  |
|              | b              | 1950            | 12.13083       |                                                    |                     |                     |                       |                        |
| Robit river  | a              | 1849            | 11.67600       | Bathing & washing cloth                            | Rocky               | Clean water          | Algae & floating vegetation | River                  |
|              | b              | 1841            | 11.67606       |                                                    |                     |                     |                       |                        |
| Gumara river | a              | 1906            | 12.39091       | Bathing, fetching & washing cloth                 | Rocky               | Turbid water         | Covered with algae    | River                  |
|              | b              | 1903            | 12.39083       |                                                    |                     |                     |                       |                        |
| Garno river  | a              | 1859            | 12.23636       | Bathing and washing cloth                          | Rocky               | Clean water          | Covered with algae    | River                  |
|              | b              | 1856            | 12.23685       |                                                    |                     |                     |                       |                        |

Figure 2. Image taken from some of the snail collection sites around Lake Tana; (a) Enferanz river (b) Cherechera site (c) Garno river (d) Qunzela port (e) Gorgora port (f) Shinne river (g) Dek Island (h) Robit river. All the images were taken by the corresponding author (TH).
The abundance of freshwater snails in Lake Tana and tributary rivers. A total of 3886 freshwater snails were collected from 20 sampling points at ten study sites during the study period. Five freshwater snail genera, namely *Biomphalaria*, *Lymnaea*, *Bulinus*, *Melanoides* and *Bellamya* snails, were recorded from the study areas (Fig. 3). The dominant snail genus observed in the study area was *Biomphalaria* (41.33%, 95% CI 39.77–42.89%) followed by *Lymnaea* (Table 2). *Lymnaea* snails were identified from all snail collection sites while *Biomphalaria* snails were observed from nine snail collection sites. *Melanoides* and *Bellamya* were observed from limited study sites, mainly from the Lake Tana periphery.
Spatial and seasonal abundance of Biomphalaria snails. Biomphalaria snails were collected from 18 sampling points around Lake Tana. A total of 1606 Biomphalaria snails were collected from the nine study sites around Lake Tana. The highest (23.29%; 95% CI 21.2–25.4%) and the lowest (6.1%; 95% CI 4.9–7.4%) number of Biomphalaria snails were collected from Shinne River and Qunzela lakeshore, respectively (Fig. 4). Similarly, the abundance of Biomphalaria snails varied across snail collection seasons. The seasonal distribution showed that the winter season had the highest Biomphalaria snail abundance, 36.43% (95% CI 34.07–38.83%), while the summer season showed the lowest abundance of Biomphalaria snail, 18.06% (95% CI 16.21–20.03%) (Fig. 5). There was a significant difference in the abundance of Biomphalaria across study sites and seasons ($p<0.05$).

Comparison of Biomphalaria snails by habitat. Freshwater snails were collected from two types of habitats: tributary rivers and Lake periphery. Biomphalaria snails were more common in rivers with a sandy and stony basement than in lakeshores (Fig. 6). There was a significant difference in the number of Biomphalaria snails between the lakeshore and riverine areas ($p<0.05$).

Infection status of Biomphalaria snails of Lake Tana and its tributary rivers. A total of 1375 live Biomphalaria snails were tested for trematode infection. Among these snails, 14.40% (95% CI 12.59–16.37%) snails shed trematode cercariae, but only 4.87% (95% CI 3.79–6.15%) were cercariae of S. mansoni (Table 3). The common trematodes observed in this study consisted of cercariae of Schistosoma mansoni, Amphistome, Echinostome, Brevifurcate apharyngeate distome and unidentified cercaria (Fig. 7). The highest S. mansoni infection was observed from the Cherechera site (10.59%) followed by Qunzela site (6.74%) while the lowest S. mansoni infection was observed from the Gumara River (1.49%). A significant difference in the infection prevalence was observed across study sites ($p=0.004$). The study was conducted in all seasons and the highest and the lowest S. mansoni cercariae were observed during the spring and summer.

Table 2. Diversity and abundance of freshwater snails of Lake Tana and its tributary rivers, 2021/22.

| Collection sites     | Freshwater snail Genus observed in the study area | Biomphalaria snails No. (%) | Bulinus snails No. (%) | Lymnaea snails No. (%) | Melanoides snails No. (%) | Bellamya snails No. (%) | Total No. (%) |
|----------------------|--------------------------------------------------|-----------------------------|------------------------|------------------------|--------------------------|-------------------------|---------------|
| Cherechera           | 194 (12.1)                                       | 87 (19.6)                  | 40 (3.0)               | 139 (70.2)            | 71 (23.0)                | 331 (13.70)             |
| Dek Island           | 173 (10.8)                                       | 119 (12.6)                 | 151 (11.4)             | 0                     | 34 (11.0)                | 477 (12.3)             |
| Zegie Peninsula      | 99 (6.2)                                         | 65 (14.6)                  | 63 (4.7)               | 37 (18.7)             | 76 (24.6)                | 340 (8.7)              |
| Qunzela town         | 98 (6.1)                                         | 68 (15.3)                  | 173 (13.0)             | 9 (4.6)               | 58 (18.8)                | 406 (10.5)             |
| Gorgora Peninsula    | 0                                                | 17 (3.8)                   | 56 (4.2)               | 0                     | 38 (12.3)                | 111 (2.8)              |
| Enferanze river      | 128 (7.9)                                        | 45 (16.1)                  | 168 (12.6)             | 12 (6.1)              | 32 (10.4)                | 385 (9.9)              |
| Shinie river         | 274 (23.3)                                       | 0                           | 240 (18.1)             | 0                     | 0                        | 641 (15.8)             |
| Robit river          | 240 (14.9)                                       | 42 (9.5)                   | 319 (24.0)             | 0                     | 0                        | 601 (15.5)             |
| Gumara river         | 162 (11.0)                                       | 1 (0.23)                   | 60 (4.5)               | 0                     | 0                        | 223 (5.7)              |
| Garmo river          | 138 (8.6)                                        | 0                           | 59 (4.4)               | 1 (0.5)               | 0                        | 198 (5.1)              |
| All sites            | 1606 (41.33)                                     | 444 (11.43)                | 1329 (34.20)           | 198 (5.10)            | 309 (7.94)               | 3886 (100)             |

Figure 4. Relative abundance of Biomphalaria snails at different collection sites.
Figure 5. Relative abundance of *Biomphalaria* snails on a seasonal basis.

Figure 6. Relative abundance of *Biomphalaria* snails at study habitat.

| Snail collection sites | Snail count | Snail examined for cercaria | *S. mansoni* cercaria | Other trematodes cercaria | Total trematodes cercaria |
|------------------------|-------------|-----------------------------|-----------------------|--------------------------|---------------------------|
|                         | Number      | Number (%)                  | Number (%)            | Number (%)               | Number (%)                |
| Cherechera             | 194         | 170 (10.59)                 | 18 (10.59)            | 23 (13.53)               | 41 (24.12)                |
| Inferanz river         | 128         | 109 (5.50)                  | 6 (5.50)              | 24 (22.02)               | 30 (27.52)                |
| Zegie Peninsula        | 99          | 84 (2.38)                   | 2 (2.38)              | 6 (7.14)                 | 8 (9.52)                  |
| Dek Island             | 173         | 143 (4.90)                  | 7 (4.90)              | 19 (13.29)               | 28 (19.58)                |
| Gumara river           | 162         | 134 (1.49)                  | 2 (1.49)              | 6 (4.48)                 | 8 (5.97)                  |
| Genaro river           | 138         | 127 (1.57)                  | 2 (1.57)              | 12 (9.45)                | 14 (11.02)                |
| Shinne river           | 374         | 313 (5.75)                  | 18 (5.75)             | 23 (7.35)                | 41 (13.10)                |
| Robit river            | 240         | 206 (2.91)                  | 6 (2.91)              | 12 (5.83)                | 18 (8.74)                 |
| Qunzela town           | 98          | 89 (6.74)                   | 6 (6.74)              | 4 (4.49)                 | 10 (11.24)                |
| Total                  | 1606        | 1375 (4.87)                 | 67 (4.87)             | 129 (9.38)               | 198 (14.40)               |

Table 3. Prevalence of *Schistosoma mansoni* and other trematodes infection among *Biomphalaria* snail species.
seasons, respectively (Fig. 8). There was a significant difference in the infection prevalence of *Biomphalaria* snails across study seasons ($p < 0.001$).

**Discussion**

Epidemiological studies about the snail intermediate host species are vital for policymakers to design appropriate schistosomiasis control strategies. The principal intermediate host for *S. mansoni* in Ethiopia is *Biomphalaria* species. Assessment of abundance, distribution and infection status of *Biomphalaria* snails contributes a lot to the prevention and control of schistosomiasis in the country. Schistosomiasis control strategies...
might not be effective without considering the snail intermediate hosts. In line with this, the present study aimed to determine the abundance, distribution and infection status of *Biomphalaria* snails with *S. mansoni* cercariae in and around Lake Tana.

The present study was conducted at lakeshores and tributary rivers of Lake Tana. This study revealed the presence of *Biomphalaria*, *Bulinus*, *Lymnaea*, *Melanoides* and *Bellamya* snails in the study sites. Among these snail genera, *Biomphalaria* snail was the predominant snail genus and it was recorded from nine study sites, which is in agreement with reports from studies conducted at Gibe River Basin, Ethiopia 14, Kenya and Tanzania 15. *Biomphalaria* snails were more common in rivers than in the lakeshores, which is in line with reports from studies conducted in Ethiopia 16, East Africa 37, South Africa 38, Nigeria 39 and Kenya 40. This shows that *Biomphalaria* snails prefer rivers and streams that have clear water with sandy and gravel substrates to lakeshores that have muddy substrates.

**Seasonal variation of *Biomphalaria* snails.** The abundance and distribution of *Biomphalaria* snails varied significantly across study seasons. *Biomphalaria* snails were dominant during winter and spring as compared with other seasons. Similar observations were reported from Egypt 41. Several studies have shown that the abundance of *Biomphalaria* snails was higher in the dry season than in wet season 42–44. In contrast to our finding, *Biomphalaria* snails were more abundant during the wet seasons than in the dry season in South Africa 45. This might be associated with the water temperature, velocity, turbidity and other environmental parameters of the study area. High rainfall, water velocity, and turbidity during the rainy season affect the natural habitats of snails in Ethiopia. As a result of these environmental conditions, the abundance of *Biomphalaria* snails may decline in the study area. This suggests that *Biomphalaria* snails may prefer stable habitat for survival.

**The spatial variation of *Biomphalaria* snails.** The abundance of *Biomphalaria* snails varied across study sites. *Biomphalaria* snails were recorded from all study sites except Gorgora. Although we attempted several times to search for *Biomphalaria* snails from Gorgora peninsula, we could not find *Biomphalaria* snails. The abundance of *Biomphalaria* snails varied from 6.1% to 23.3% in the different study sites. Spatial variations in the abundance of *Biomphalaria* snails across study sites were well documented 46–48. It is known that snail abundance varies from area to area depending on different environmental and biotic factors. In the present study, the difference in abundance of snails across sites might be associated with the nature of study sites. *Biomphalaria* snails being more abundant in rivers than in lakeshores. A study in Senegal showed that *Biomphalaria* snails preferred clean rivers and streams having stony and gravel substrates 47. The overall variation in the abundance of *Biomphalaria* snails might be associated with the nature of the water, types of aquatic vegetation, nature of water substrate, geographical locations and other environmental parameters.

**Infection status of *Biomphalaria* snails.** The current study showed that 14.4% of *Biomphalaria* snails shed different types of trematodes cercariae, which is in agreement with reports from studies conducted in Egypt 49 and Tanzania 50. In contrast to the present finding, only 4.6% of *Biomphalaria* snails were infected with trematodes around Omo Gibe River Basin in Ethiopia 14. The present study revealed that 4.87% *Biomphalaria* snails were infected with *S. mansoni*, which is in agreement with reports from studies conducted in different parts of Ethiopia 50, as well as with finding in systematic review and meta-analysis from African countries 52. In contrast, high prevalence of *Schistosoma mansoni* cercariae in *Biomphalaria* snails were reported from different parts of Ethiopia 13,20,53, Tanzania 49,54 and Nigeria 55. The proportions of *Biomphalaria* snails infected with schistosome cercariae reported from Kenya were even lower than our findings 46,48. The difference in infection status of *Biomphalaria* snails observed across studies is mainly linked to the types of diagnostic methods used. Superior detection of *S. mansoni* infection from *Biomphalaria* snails was obtained using PCR compared to cercarial shedding experiments. For example, 12% vs. 47% was reported from Tanzania 47 and 5% vs. 27% shown in review paper in African countries 52. In addition, anthropogenic activities, geographical locations, water quality, types of aquatic vegetation and other environmental factors might have contributed to the observed differences.

Significant variation of *Biomphalaria* infection with *S. mansoni* was observed across the study seasons. The highest infection rate was observed during the dry season as compared to the wet season, which is in line with reports from Tanzania 49, Sudan 27 and Nigeria 55. High levels of open-field defecation, human-water contact activities, and stable water conditions are observed during the dry seasons of the year in Ethiopia. These conditions might contribute to the long-term survival of *Biomphalaria* snails leading to high chance of infection with *S. mansoni* miracidia.

The infection status of *Biomphalaria* snails varied across study sites. In this study, it was revealed that the proportion of infected *Biomphalaria* snails was higher along the Lake periphery than in rivers. In contrast to this finding, more number of infected *Biomphalaria* snails were reported from lakeshores as compared to rivers and streams in western Kenya 49. These variations are mainly linked to the level of anthropogenic activities such as human–water contact activities and open-field defecation. High *Biomphalaria* snail infection was observed in the area where there is frequent human–water contact activities associated with washing clothes, swimming, bathing, fetching water and fish processing.

Schistosomiasis control strategies in sub-Saharan African countries including Ethiopia focus on mass-drug administration to school-aged children, with little or no emphasis on snail control. In this study, *Biomphalaria* snails were shown to be sources of *S. mansoni* infection and therefore it is an appropriate area for intervention to support the ongoing schistosomiasis prevention and control in the study area. Therefore, policymakers are advised to revisit the current schistosomiasis control and prevention strategies in Ethiopia.
Limitation of the study. Water quality and its association with snail abundance were not assessed in the present study. Water quality may have an impact on the abundance of Biomphalaria snails at different study sites as well as across seasons. The study was conducted at ten different sites and four seasons making it difficult to collect information for water quality analysis. In the present study, we used a cercarial shedding experiment which has lower sensitivity compared to PCR approaches. This might have led to the underestimation of the true prevalence of S. mansoni cercariae in Biomphalaria snails.

Conclusion
Lake Tana and its tributary rivers serve as suitable habitats for freshwater snails particularly for Biomphalaria snails. Biomphalaria species were abundant freshwater snails and they were present in varied numbers in nearly all of snail collection sites throughout the year. In this study it was revealed that nearly five percent of Biomphalaria snails were infected with S. mansoni cercariae. The prevention and control of schistosomiasis in Ethiopia totally rely on mass-drug administration without giving due consideration to snail control. This study showed that Biomphalaria snails were important sources of S. mansoni infection to humans living in the nearby snail habitats. Therefore, policymakers, regional administrators, and other stakeholders working on schistosomiasis need to incorporate appropriate snail control strategies to support the ongoing schistosomiasis prevention and control strategies.

Data availability
All data generated or analyzed during this study are included in this published article.

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References
1. McManus, D. P. et al. Schistosomiasis. Nat. Rev. Dis. Primers 4(1), 13. https://doi.org/10.1038/s41572-018-0013-8 (2018).
2. WHO. Schistosomiasis. 2022 [cited 2022 19 of March ]; Available from: https://www.who.int/news-room/fact-sheets/detail/schistosomiasis.
3. Mazigo, H. D. Participatory integrated control strategies and elimination of schistosomiasis in sub-Saharan Africa, Langet Global Health 7, 1999 https://doi.org/10.1016/S2214-109X(19)30271-2 (2019).
4. Onasanya, A., Bengtson, M., Oladejo, O., Van Engelen, J. & Diehl, J. C. Rethinking the top-down approach to schistosomiasis control and elimination in sub-saharan Africa. Front Public Health https://doi.org/10.3389/fpubh.2021.622809 (2021).
5. Worku, F. E., Danne, D., Endris, M., Tesfaye, H. & Aemero, M. Schistosoma mansoni Infection and associated determinant factors among school children in Sanja town Northwest Ethiopia: J. Parasitol. Res. https://doi.org/10.1155/2014/792536 (2014).
6. WHO Ethiopian School-Based Deworming Campaign Targets 17 Million Children. 2015. (2015).
7. Hailegebriel, T., Nibret, E., Munshea, E. & Ameha, Z. Prevalence, intensity and associated risk factors of Schistosoma mansoni infection among schoolchildren around Lake Tana, northwestern Ethiopia. PLOS NTDs 15(10), e0009861. https://doi.org/10.1371/journal.pntd.0009861 (2021).
8. Ayalew, J., Addisu, A. & Tegegne, Y. Prevalence, intensity, and associated factors of Schistosoma mansoni infection among school children in Ethiopia: J. Parasitol. Res. https://doi.org/10.1155/2020/8820220 (2020).
9. Bereket, A. et al. Epidemiology of intestinal helminthiasis among school children with emphasis on Schistosoma mansoni infection in Wolaita zone Southern Ethiopia. BMC Public Health 17(1), 587. https://doi.org/10.1186/s12889-017-4499-2 (2017).
10. Teshome, B., Hu, W., Liang, S. & Berhanu, E. Transmission of Schistosoma mansoni in Yachi areas, southwestern Ethiopia: new foci. Infect. Dis. Poverty 8(1), 1. https://doi.org/10.1186/s40249-018-0513-5 (2019).
11. Lu, X.-T. et al. Snail-borne parasitic diseases: An update on global epidemiological distribution, transmission interruption and control methods. Infect Dis. Poverty 7(1), 28. https://doi.org/10.1186/s40249-018-0414-7 (2018).
12. Erko, B., Balch, F. & Kiffe, D. The ecology of Biomphalaria sudanica in Lake Ziway. Ethiopia. Afr. J. Ecol. 44(3), 347–352. https://doi.org/10.1111/j.1365-2028.2006.00613.x (2006).
13. Abebe, G., Erko, B., Aemero, M. & Petros, B. Epidemiological study on Schistosoma mansoni infection in Sanja area, Amhara region. Ethiopia Parasites Vectors 7(1), 15. https://doi.org/10.1186/1756-3305-7-15 (2014).
14. Hailu, B. Berhanu, E., Gebre, M. & Fekadu, B. Decline of urinary schistosomiasis in Kurmkut town, western Ethiopia. BMJ Open 1(1), 0. https://doi.org/10.1136/bmjopen-2011-000324 (2011).
15. Hailu, B., Berhanu, E., Gebre, M. & Fekadu, B. Decline of urinary schistosomiasis in Kurmkut town, western Ethiopia. Ethiop. Med. J. 50(4), 331–336 (2012).
16. Mekonnen, Z., Haileselassie, H., Medhin, G., Erko, B. & Berhe, N. Schistosomiasis management and associated risk factors in Mekele city, northern Ethiopia. Ethiop. Med. J. 50(4), 331–336 (2012).
17. Mulugueta, M. et al. Human intestinal schistosomiasis in communities living near three rivers of Jimma town, southwest Ethiopia. J. Health Sci. 55(2), 1–11. https://doi.org/10.4314/jhs.v55i2.9 (2019).
18. Abebe, A., Chalachew, A., Minwuyelet, M., Goraw, G. The fish and the fisheries of Lake Tana. In: Stave K. gg, Aynalem S, editor. Ecology and Social and Ecological System Dynamics, (Springer, 2017).
19. Mekonnen, Z., Haileselassie, H., Medhin, G., Erko, B. & Berhe, N. Schistosomiasis management and associated risk factors in Mekele city, northern Ethiopia. Ethiop. Med. J. 50(4), 331–336 (2012).
20. Mulugueta, M. et al. Human intestinal schistosomiasis in communities living near three rivers of Jimma town, southwest Ethiopia. J. Health Sci. 55(2), 1–11. https://doi.org/10.4314/jhs.v55i2.9 (2019).
21. Hailegebriel, T., Nitret, E. & Munshae, A. Prevalence of Schistosoma mansoni and associated risk factors in human and Biomphalaria snails in Ethiopia: A systematic review and Meta-analysis. Acta Parasitol. https://doi.org/10.2478/s11686-021-00449-6 (2021).
22. Melese, W. Ecosystem services and tourism potential in Lake Tana peninsula: Ethiopia review. J. Tourism Hosp. 6(6) https://doi.org/10.4172/2167-0269.1000324 (2017).
23. Abebe, A., Chalachew, A., Minwuyelet, M., Goraw, G. The fish and the fisheries of Lake Tana. In: Stave K. GG, Aynalem S, editor. Ecology and Social and Ecological System Dynamics, (Springer, 2017).
24. Melese, W. Lake Tana as biosphere reserve: Review. J. Tourism Hosp. 6, 5 https://doi.org/10.4172/2167-0269.1000310 (2017).
25. WHO A practical guide to identification of African freshwater snails. Danish Biharziasis Laboratory in Collaboration with World Health Organization. 1–13 (1980).
26. Falade, M. O. & Otarigho, B. Shell morphology of three medical important tropical freshwater pulmonate snails from five sites in South-Western Nigeria. Int. J. Zool. Res. 11, 140–150. https://doi.org/10.3923/ijzr.2015.140.150 (2015).
27. Brown, D. S. Freshwater snails of Africa and their medical importance (Taylor and Francis, 1994).
28. Jordaens, K., Bruyndonckx, L., Van Goethem, J. & Backeljau, T. Morphological and anatomical differentiation of three land snails of the genus Rhyynchotrochus (Gastropoda: Camaenidae). J. Mölluscan. Stud. 75(1), 1–8. https://doi.org/10.1093/mollus/eyn055 (2008).
29. Manyangadze, T., Chimbari, M. J., Rubaba, O. & Soko, W. Spatial and seasonal distribution of Bulinus globosus and Biomphalaria pfeifferi in Ingwavuma, uMhlanyankude district, KwaZulu-Natal South Africa: Implications for schistosomiasis transmission and macro-geographical scale. Parasites Vectors 14(1), 222 (2021).
30. Tian-Bi, Y.-N.T. Molecular characterization and distribution of Schistosoma cercariae collected from naturally infected bulinid snails in northern and central Côte d’Ivoire. Parasites Vectors 12(1), 117. https://doi.org/10.1186/s13071-019-3381-3 (2019).
31. Frandsen, F. & Christensen, N. An introductory guide to the identification of cercariae from African freshwater snails with special reference to cercariae of trematode species of medical and veterinary importance. Acta Trop. 4, 181–202 (1984).
32. Fatima, M. A., Maikaje, D. B. & Umar, Y. A. Cercarial diversity in freshwater snails from selected freshwater bodies and its implication for veterinary and public health in Kaduna State Nigeria. World Acad. Sci., Eng. Technol. Int. J. Anim. Vet. Sci. 12(2), 52–58 (2018).
33. Anucherngchai, S., Tejankura, T. & Chontananarth, T. Epidemiological situation and molecular identification of cercarial stage in freshwater snails in Chao-Phraya Basin, Central Thailand. Asian Pacific J. Tropical Biomed. 6(6), 539–545. https://doi.org/10.1016/s2095-0219(v1)2020(v2020).
34. Seid, T. et al. Environmental determinants of distribution of freshwater snails and trematode infection in the Omo gibe River basin, southwest Ethiopia. Infect. Dis. Poverty 8(1), 93. https://doi.org/10.1186/s40249-019-0604-y (2019).
35. Dida, G. O. et al. Distribution and abundance of schistosomiasis and fascioliasis host snails along the mara river in Kenya and Tanzania. Infect. Ecol. Epidemiol. https://doi.org/10.3402/iee.v4.24281 (2014).
36. Ketema, D. et al. Effects of land use on intermediate host snail fauna, abundance, distribution and cercariae infection rate in Omo-Gibe river basin Ethiopia. Res. Sq.: Preprint https://doi.org/10.21203/rs.2.22097/v1(2020).
37. Magero, V. O., Kisara, S., Wade, C. M. Geographical distribution of Biomphalaria pfeifferi snails in East Africa bioRxiv Preprint. 2021:2021.11.04.662736. https://doi.org/10.1101/2021.11.04.662736.
38. De Kock, K. N., Wolmarans, C. T. & Bornman, M. Distribution and habitats of the snail Lymnaea truncatula, intermediate host of the liver fluke Fasciola hepatica in South Africa. J. S. Afr. Vet. Assoc. 74(4), 117–122 (2004).
39. Salawu, O. & Odaibo, A. Preliminary study on ecology of Bulinus jousseaumei in Schistosoma haematobium endemic rural communities of Nigeria. Afr. J. Ecol. 51(3), 441–446. https://doi.org/10.1111/aje.12054 (2012).
40. Opisa, S., Odiere, M. R., Jura, W. G. Z. O. & Karanja, D. M. S. Mwinzi PNMMalacological survey and geographical distribution of vector snails for schistosomiasis within informal settlements of Kisumu city, western Kenya. Parasites Vectors 4(1), 226. https://doi.org/10.1186/1756-3305-4-226 (2011).
41. Yousif, F., Kamel, G., el Emam, M. & Mohamed, S. Ecology of Biomphalaria alexandrina the snail vector of Schistosoma mansoni in Egypt. J. Egypt Soc. Parasitol. 23(1), 29–42 (1993).
42. Gouvras, A. N., Allan, F. & Kanoufi, K. A. Geographical distribution of Biomphalaria pfeifferi and Biomphalaria glabrata snails with reference to cercariae of trematode species of medical and veterinary importance. Acta Trop. 114(3), 203–207. https://doi.org/10.1016/j.actatropica.2011.03.007 (2011).
43. Abdulkadir, F. M., Maikaje, D. B. & Umar, Y. A. Ecology and distribution of freshwater snails in Gimbawa dam, Kaduna state Nigeria. BMC Public Health 8(1), 121–129. https://doi.org/10.3337/bjph.2011.59.2.121 (2011).
44. Odero, S.O., Ogonda, L., Munde, E.O., Shiulu, C., Cheveya, P. Distribution of biomphalaria snails in associated vegetation and schistosomiasis infection prevalence along the shores of lake Victoria, Tanzania: implications for schistosomiasis transmission and control, Parasites Vectors 10(1), 316 https://doi.org/10.1186/s13071-017-2525-z (2017).
45. Ismail, H. A. et al. Population dynamics of intermediate-host snails in the white nile river, Sudan: A year-round observational descriptive study. Korean J. Parasitol. 59(2), 121–129. https://doi.org/10.3347/kjp.2021.59.2.121 (2021).
46. Abdulkadir, F. M., Maikaje, D. B. & Umar, Y. A. Ecology and distribution of freshwater snails in Gimba wa dam, Kaduna state Nigeria. NJCRes 22(2), 98–106 (2017).
47. Rowel, C., Fred, B., Betson, M., Sousa-Figueiredo, J.C., Kabatereine, N.B., Stothard, J.R. Environmental epidemiology of intestinal schistosomiasis in Uganda: Population dynamics of Biomphalaria (Gastropoda: Planorbidae) in Lake albert and lake victoria with observations on natural infections with digenetic trematodes, Biomed. Res. Int. 2015;717261 https://doi.org/10.1155/2015/717261 (2015).
48. Siﬁy, B., Christopher, J. E. H, Cheikh Tidiane, B., Nicolas, J., Gilles, R., Jason, R. Seasonal Variations of Densities of Biomphalaria pfeifferi, the Intermediate Host of Schistosoma mansoni Parasite at the North of Senegal. In: Sajal R, Soumalia M, editors. Update on Malacology (2021).
49. Okeke, O. C. & Ubachukwu, P. O. Trematode infections of the fresh water snail Biomphalaria pfeifferi from a south-east Nigerian community with emphasis on cercarial of Schistosoma. J. Helminthol. 91, 295–301. https://doi.org/10.1093/sox22149X/16003555 (2017).
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Author contributions
T.H. was involved in the design, data collection, processing, interpretation of the findings, and drafting of the manuscript. All the images were taken by T.H. E.N. and A.M. were involved in the conception of the idea, drafting the manuscript, reviewing and editing the manuscript.

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Competing interests
The authors declare no competing interests.

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