Data Article

Safety data on in situ gelling bimatoprost loaded nanovesicular formulations

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In vitro cytotoxicity and in vivo acute and 7 days repeat-dose ocular toxicity studies, were conducted in rabbits, in accordance with the Organisation for Economic Co-operation and Development (OECD) guidelines, for bimatoprost loaded nanovesicular aqueous dispersion (BMT-NV) and its in-situ gelling subconjunctival implant (BMT-NV-IM). For details on the preparation and evaluation of BMT-NV and its BMT-NV-IM for the control of glaucoma, please refer to ‘Bimatoprost loaded nanovesicular long-acting sub-conjunctival in-situ gelling implant: In vitro and in vivo evaluation’ (Yadav et al., 2019). The in vivo ocular toxicity was performed only after confirming dermal safety, as required by OECD. Histological evaluation of various ocular tissues, following sub-conjunctival implantation with BMT-NV-IM, was done for ocular tolerance studies.

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1. Data

1.1. BMT loaded nanovesicles (BMT-NVs) and their incorporation into a gel (BMT-NV-GEL)

BMT-NVs and BMT-NV-GEL was prepared as discussed in the main article [1].

1.2. Cytotoxicity studies

All ingredients employed for preparing the BMT-NV-GEL were of biodegradable and biocompatible nature and were indicated to be safe for ocular use at the employed concentration. Poloxamer 407 (P407) and carbopol 934P have been already used in FDA approved ophthalmic formulations [2].

Both BMT-NV and BMT-NV-GEL were safe and did not show any statistically significant ($p < 0.001$) cytotoxicity (Fig. 1, Tables S1–S6 (raw data for Fig. 1, added as supplementary data)) when administered to HCLE (150 µg/ml), HCjE (1.5, 15) and R28 (1.5, and 15 µg/ml) cell lines for 24 h. Since BMT-NV-GEL is to be used as in-situ gelling ocular drops as well as subconjunctival implant so both HCLE and HCjE cell lines were used. However exposure to high concentration of 150 µg/ml in case of conjunctival cell lines HCjE showed significant cytotoxicity. It may be noted that we expect a slow release from the
subconjunctival area, and at no time is the concentration expected to reach as high as 150 mg/ml; two lower doses i.e 1.5 and 15 mg/ml show complete protection. It may also be noted that exposure of HCjE cells to 150 mg/ml of free BMT is also toxic (~10% viability; Table S4). In comparison BMT NV and BMT NV gel are 4 times less toxic (~40% viability; Table S4).

Lower concentrations were also used for R28 cell lines which showed no cytotoxicity at 1.5 and 15 μg/ml.

Marketed drops (Lumigan drops) showed significant cell death in all three cell lines at all concentrations, and this result is not surprising as it has been published by other researchers [3–6] that benzalkonium chloride (BAK), the preservative present in the marketed formulation can induce cell death in in vitro experiments.

1.3. In vivo safety studies in rabbits

OECD 405 recommends that in vivo eye irritation/corrosion test, should be conducted after the in vivo dermal safety (OECD testing guideline 404) of a product/substance is confirmed (AEI, 2002) [7].

The score for both the (i) acute dermal irritation/corrosion study (compiled in Table 1); and the (ii) acute eye irritation/corrosion (compiled in Table 2) was zero. This clearly demonstrates a non-irritant/corrosive nature of BMT-NV-GEL when applied to dermal and ocular tissues (topical instillation), and hence are concluded to be safe for ocular use.

Repeated instillation acute study was performed in view of the fact that glaucoma requires life long treatment and will need frequent instillation of developed formulation for effective control of IOP. Similar studies have been reported by us earlier [8]. The scores obtained from this study (Table 4) also prove the system to be safe for repetitive ocular use.
Similarly aggressive therapy viz. chronic repeat instillations (5 times at 5 minute interval) for a period of one week was also evaluated and BMT-NV-GEL was still found to be safe (Table 3).

1.4. Ocular tolerance evaluation

Subconjunctival injection of BMT-NV-GEL implant [8] resulted in the formation of a bleb and conjunctival hyperemia (mild and transient) in 2 out of 4 injected eyes, which was resorbed completely within 48 h after injection. All eyes appeared normal and similar to the pre-injected or uninjected eyes after 48 h. The rats tolerated the procedure well, showing no sign of distress or pain during or immediately following administration. There were no signs of any irritation, swelling or redness and infection in any of the injected eye during the study. At the end of the study period, the residual gel was no longer observed and it is assumed that the gel dissolved completely. Particular attention was directed toward the sclera and conjunctival tissue surrounding the injection site during histological examination. The tissue appeared normal and we did not detect any signs of inflammation such as accumulation of macrophages, lymphocytic infiltrate, or evidence of giant cells (Fig. 2).

![Table 1](image)

| Skin reactions | Time in h | Total count |
|----------------|-----------|-------------|
|                | 0 1 24 48 72 |            |
| Erythema       | 0 0 0 0 0 | 0/40        |
| Oedema         | 0 0 0 0 0 | 0/40        |
| Final total score | 0/80      |            |

(Scoring 0–4 was done as described in OECD guidelines 404 as per Table 1: Grading of skin reactions, page 7).

![Table 2](image)

| Tissue of the eye | Time in h | Total count |
|-------------------|-----------|-------------|
|                   | 0 1 24 48 72 |            |
| Cornea            | 0 0 0 0 0 | 0/60        |
| Iris              | 0 0 0 0 0 | 0/30        |
| Conjunctiva       | 0 0 0 0 0 | 0/45        |
| Chemosis          | 0 0 0 0 0 | 0/60        |
| Grand Total score |           | 0/195       |

(Scroing was done as defined in OECD guidelines 405 as per Table 1: Grading of ocular lesions (page: 8 of [9]).

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![Table 3](image)

| Ocular tissue | Scores of rabbit 1 | Scores of rabbit 2 | Scores of rabbit 3 | Score |
|---------------|---------------------|---------------------|---------------------|-------|
|               | 0 h 1 h 24h 48h 72h | 0 h 1 h 24h 48h 72h | 0 h 1 h 24h 48h 72h |       |
| Cornea        | 0 0 0 0 0 | 0 0 0 0 0 | 0 0 0 0 0 | 0/60  |
| Iris          | 0 0 0 0 0 | 0 0 0 0 0 | 0 0 0 0 0 | 0/30  |
| Conjunctiva   | 0 0 0 0 0 | 0 0 0 0 0 | 0 0 0 0 0 | 0/45  |
| Chemosis      | 0 0 0 0 0 | 0 0 0 0 0 | 0 0 0 0 0 | 0/60  |
| Total score   |         |         |         | 0/195 |

Scoring was done as described under Table 2.
2. Experimental design, materials and methods

2.1. Establishing safety of the developed system

2.1.1. Cytotoxicity studies

Viability of stratified HCLE (human corneal - limbal epithelial), HCjE (human conjunctival epithelial) and R28 (retinal neuronal) cells was measured by the cell proliferation assay using MTT (3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide). Cells were grown in keratinocyte

| Ocular tissue | Cornea | Iris | Conjunctiva | Chemosis | Score | Total Score |
|---------------|--------|------|-------------|----------|-------|-------------|
| rabbit 1      | 0h     | 0    | 0           | 0        | 0/104 | 0/312       |
|               | 1d     | 0    | 0           | 0        | 0     |             |
|               | 2d     | 0    | 0           | 0        | 0     |             |
|               | 3d     | 0    | 0           | 0        | 0     |             |
|               | 5d     | 0    | 0           | 0        | 0     |             |
|               | 6d     | 0    | 0           | 0        | 0     |             |
|               | 7d     | 0    | 0           | 0        | 0     |             |
| rabbit 2      | 0h     | 0    | 0           | 0        | 0/104 |             |
|               | 1d     | 0    | 0           | 0        | 0     |             |
|               | 2d     | 0    | 0           | 0        | 0     |             |
|               | 3d     | 0    | 0           | 0        | 0     |             |
|               | 5d     | 0    | 0           | 0        | 0     |             |
|               | 6d     | 0    | 0           | 0        | 0     |             |
|               | 7d     | 0    | 0           | 0        | 0     |             |
| rabbit 3      | 0h     | 0    | 0           | 0        | 0/104 |             |
|               | 1d     | 0    | 0           | 0        | 0     |             |
|               | 2d     | 0    | 0           | 0        | 0     |             |
|               | 3d     | 0    | 0           | 0        | 0     |             |
|               | 5d     | 0    | 0           | 0        | 0     |             |
|               | 6d     | 0    | 0           | 0        | 0     |             |
|               | 7d     | 0    | 0           | 0        | 0     |             |

Scoring was done as described under Table 2.

Fig. 2. Optical microscopic pictures showing histological section of (a) naïve conjunctival tissue and conjunctival tissue of eyes post-treatment (b) 2 days, (c) 1 week, (d) 1 month, and (e) 2 month indicating absence of any untoward reactions.

2. Experimental design, materials and methods

2.1. Establishing safety of the developed system

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serum-free medium (Invitrogen, Carlsbad, CA) maintained at 37 °C in 5% CO₂ and supplemented with 25 μg/ml bovine pituitary extract, 0.4 mM CaCl₂, 0.2 ng/ml epidermal growth factor (EGF) and other suitable antibiotics. For suitable stratification and differentiation, the serum-free Invitrogen medium was replaced (at confluence) with Dulbecco’s minimal essential medium (DMEM)/F12 medium, supplemented with 10% calf serum and 10 ng/ml EGF for a period of 7 days [10]. R28 cells were grown in DMEM (Invitrogen, Carlsbad, CA) supplemented with fetal bovine serum (10%), glutamine (2 mM), gentamicin (0.21 mM), non-essential amino acids and MEM vitamins (1% each).

HCLE, HCJcE and R28 cells were exposed, after stratification in DMEM/F12 medium, to various test samples viz. BMT-NV, Blank-NV, BMT-NV-GEL, Blank-NV-GEL, BMT-SOL and marketed formulation (Lumigan® 0.03% drops) for 24h. MTT assay was then used to determine cell viability [4]. Fresh MTT solution (0.5 mg/ml) was added to variously exposed cells and incubated for 2 h at 37 °C. The purple formazan complex released from the cells after lysis was then dissolved in dimethyl sulfoxide. Absorbance of these samples was read on Gen 5 plate reader (BioTek, Winooski, VT, USA) at 570 nm. The reading was corrected for background by subtracting the absorbance measured at 690 nm from that at 570 nm. The mean absorbance value of untreated cells was taken as 100% viability and all results are expressed as % cell viability compared to these cells. All experiments were repeated three times (n = 3).

2.1.2. BMT-NV-GEL in vivo safety studies as ocular drops

Safety assessment for ocular application was approved by the Institutional Animals Ethics Committee, Panjab University, Chandigarh, India (PU/IAEC/S/16/110, dated 11/7/2016) and performed as per the OECD guidelines.

2.1.3. Dermal irritation/corrosion test as per OECD guideline 404 (ADI, 2002) [7]

It is required to perform this test prior to the ocular irritation test. Only those test substances, which are safe for dermal use, are further instilled in to the rabbit eye for eye irritation test [7].

Six-month-old female albino rabbits, weighing between 1.3 and 1.7 kg, and with intact skin were used. The dorsal trunk area of rabbits was shaved using hair clippers and a depilatory cream, 1 day prior to the test. To 6-cm² gauze, 0.5 mg of BMT-NV-GEL was applied uniformly. The gauze was fixed for 4h on the shaved skin with a non-irritating tape, ensuring complete contact with the skin surface. Care was taken to apply the gauze on a site away from easy access of the animal by mouth or by limbs and to ensure that animal may not ingest the patch. The uncovered shaved area was taken as control. After 4h of application, the gauze piece was removed, and the site was examined after 1h for any signs of erythema, oedema or redness. The test was initially performed on one rabbit and only proceeded with the other two animals if no reaction was observed on the first animal.

2.1.4. Eye irritation/corrosion test as per OECD guideline 405 (AEI, 2002) [9]

Lower lid of right eye of each rabbit (n = 3) was pulled to create a space in the conjunctival sac and BMT-NV-GEL (0.1 ml) was administered either once or five X 0.1 ml instillations were made at 5 min intervals (for repeat test). Contralateral left eye was taken as the control for each animal. After instillation, each eye was examined at an interval of 1, 24, 48, and 72 h and scored for any reactions as described in the OECD guidelines (Table: 1, page: 8, reference: [9]). A chronic repeat dose study included administration of BMT-NV-GEL (0.1 ml), five X 0.1 ml instillations at 5 min intervals, every day for 7 days, to confirm safety of the formulation for long term therapy.

2.1.5. Ocular tolerance evaluation

The effect of BMT-NV-GEL implant on structure and integrity of the administered eye (left) was determined at 2 days, 1 week, 1 month and 2-month post administration. Right eye of all the treated animals was taken as control. Both the eyeballs were incised after sacrificing the animals and washed with saline. Then they were fixed in 8% w/w formalin solution and dehydrated in an alcohol gradient. After this the eyeballs were put in melted paraffin, which was then solidified to form a block. Cross-sections (<5 μm) cut from the latter, were observed microscopically (Nikon eclipse 90i, Japan), after haematoxyline and eosine (H and E) staining, for any pathological alterations [11].
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Conflict of interest

The authors declare that they have no known competing financial interests or personal relationships.

Appendix A. Supplementary data

Supplementary data to this article can be found online at https://doi.org/10.1016/j.dib.2019.104361.

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