INTRODUCTION

Hepatitis E virus (HEV) has four genotypes. Of these four types, genotype 1 has caused epidemic outbreaks in Asia and Africa and genotype 2 is usually discovered in western Africa. Both genotype 1 and 2 are found exclusively in humans.1 On the other hand, genotypes 3 and 4 are usually isolated from sporadic hepatitis E in developed countries, and also found in swine, deer, wild boar populations.2,3 Sporadic infection in non-endemic areas such as developed countries has been known to be due to an influx from foreign countries. However, some reports of locally acquired acute viral hepatitis E in people with no history of travel to endemic regions have recently increased in non-endemic areas.4,5 Autochthonous sporadic HEV infections in a non-endemic area mostly give no clue as to their sources despite the diagnosis of acute viral hepatitis E, rendering the transmission routes undecided.

Of the viruses that cause acute viral hepatitis, HEV is known to be the only virus to have animal reservoirs. Since the discovery of HEV in swine, HEV has also been isolated from chickens, deers, mongooses, rabbits and rats, supporting zoonotic transmission and prompting its investigation.6,7 We report here a case of acute viral hepatitis E that occurred after ingestion of raw meat of a wild roe deer in the absence of contact with another hepatitis patient or travel to an endemic area.
CASE REPORT

A 43-year-old male presented with abdominal discomfort for 3 weeks and jaundice lasting 1 week. He had a past history of diabetes mellitus, which had been diagnosed 3 years prior to admission to our hospital. However, he arbitrarily stopped taking hypoglycemic agents. The patient was a heavy alcohol drinker, with the consumption of 2 to 3 bottles of Soju, distilled liquor, 4 to 5 times a week. He denied any travel outside South Korea in the preceding years. About 6-8 weeks before hospitalization, he ingested raw meat (about 300 g) of a captured wild roe deer inhabiting in Gyeongnam province with his friends, who enjoyed hunting on a regular basis.

Physical examination on admission was generally normal, except for jaundice. Mild tenderness was only noted in the epigastric area. Initial laboratory data showed white blood cell count of 4.88×10³/mm³ (polymorphonuclear neutrophils, 57.2%; lymphocytes, 36.4%; and eosinophils, 1.5%), elevated serum total bilirubin level of 12.3 mg/dL, serum aspartate aminotransferase (AST) level of 1,637 IU/L, serum alanine aminotransferase (ALT) level of 1,949 IU/L, random glucose level of 220 mg/dL and HbA1c level of 10.8%. Hepatitis B surface (HBs) antigen, immunoglobulin M (IgM) anti-hepatitis B core antigen, anti-hepatitis C virus (HCV), HCV RNA (RT PCR) were all negative with positive anti-HBs. As a result of IgM anti-hepatitis A virus (HAV) was negative with positive immunoglobulin G (IgG) anti-HAV, acute hepatitis A could be excluded. Abdominal computed tomography showed findings compatible with secondary changes in acute hepatitis, and fatty infiltration with splenomegaly, which implied concurrent alcoholic liver disease. Seven days after admission, results of IgM anti-HEV and IgG anti-HEV were both positive, the optical density value of IgM anti-HEV of 3.656 (cut-off value: 0.276) and IgG anti-HEV of 3.384 (cut-off value: 0.375), which confirmed the diagnosis of acute viral hepatitis E. IgM anti-HEV and IgG anti-HEV were measured by a commercial immunoassay (HEV IgM and HEV IgG ELISA, Genelabs Diagnostic Pte. Ltd, Singapore). The serum total bilirubin peaked at 24.3 mg/dL and rapidly decreased. The levels of AST and ALT were highest at the time of admission and then showed a rapid decrease. Twenty-three days after admission, the patient was discharged with a total bilirubin of 8.06 mg/dL, AST of 130 IU/L and ALT of 133 IU/L (Fig. 1). Two months after discharge, IgM anti-HEV and IgG anti-HEV were both still positive with the optical density value of IgM anti-HEV of 3.315 (cut-off value: 0.282) and IgG anti-HEV of 2.753 (cut-off value: 0.375). Diagnosis of hepatitis E was confirmed by the detection of both IgM and IgG anti-HEV in serial samples and by the detection of serum HEV RNA.

Detection of HEV genome in patient’s serum

Viral RNA was extracted from 140 µL of anti-HEV IgM-positive serum in phosphate-buffered saline using a QiAamp viral RNA kit (Qiagen, Valencia, CA, USA).

**Figure 1.** Patient’s clinical course with changes in ALT, TB and results of IgM / IgG anti-HEV. ALT, alanine aminotransferase; TB, total bilirubin; HEV, hepatitis E virus.

**Figure 2.** A phylogenetic tree constructed by the neighbor-joining method based on the ORF2 sequence of the Korean HEV in question, 2010-JKSH-CYY (HM769728), and 42 HEV reference strains with genotypes 1 – 4 (G1-G4). All strains were separated into four groups according to their genotype. H: isolated from humans; P: isolated from pigs. Sequence analysis was conducted using the software VectorNTI and MEGA 5.04. Bootstrap values are indicated for the major nodes as a percentage of the data obtained from 1000 resamplings.
Table 1. Comparison of the HEV isolates obtained in the present study (2010-JKSH-CYY) with 42 human and swine HEV reference strains, based on ORF2 sequence

| No. | Accession No. (Human) | Accession No. (Pig) | Genotype | Country     | Homology (%) |
|-----|----------------------|---------------------|----------|-------------|--------------|
| 1   | D11092               |                     | 1b       | China (Uighur) | 74           |
| 2   | X98292               |                     | 1c       | India       | 74           |
| 3   | M74506               |                     | 2a       | Mexico      | 78           |
| 4   | AF082843             | 3a                  | USA (Midwest) | 73           |
| 5   | AB073912             | 3b                  | Japan (Tochigi) | 76           |
| 6   | AB091394             | 3b                  | Japan (Kanagawa) | 77           |
| 7   | AF455784             | 3g                  | Kyrgyzstan | 76           |
| 8   | AY115488             | 3j                  | Canada (Ontario) | 76           |
| 9   | AB246676             | 3                   | Japan (Tokyo) | 77           |
| 10  | AF516178             | 3a                  | Korea     | 76           |
| 11  | AF516179             | 3a                  | Korea     | 76           |
| 12  | AF527942             | 3a                  | Korea     | 77           |
| 13  | AY641398             | 3a                  | Korea     | 76           |
| 14  | AY714267             | 3a                  | Korea     | 76           |
| 15  | AY714268             | 3a                  | Korea     | 76           |
| 16  | AY714269             | 3a                  | Korea     | 76           |
| 17  | AY714270             | 3a                  | Korea     | 76           |
| 18  | AY714271             | 3a                  | Korea     | 77           |
| 19  | AY714272             | 3a                  | Korea     | 76           |
| 20  | AB074915             | 3c                  | Japan (Saitama) | 83           |
| 21  | AB097812             | 4c                  | Japan (Hokkaido) | 87           |
| 22  | AB099347             | 4c                  | Japan (Tochigi) | 87           |
| 23  | AY594199             | 4d                  | China (Uighur) | 82           |
| 24  | AB197673             | 4                   | Japan     | 91           |
| 25  | AB197674             | 4                   | Japan     | 89           |
| 26  | EF077630             | 4                   | China (Changchun) | 90           |
| 27  | EU166959             | 4                   | China (Beijing) | 91           |
| 28  | FJ763142             | 4                   | Korea (Gyeonggi-do) | 98           |
| 29  | EU849134             | 4                   | Korea (Gyeonggi-do) | 97           |
| 30  | AF151963             | 4                   | China     | 93           |
| 31  | DQ445492             | 4                   | China (Changchun) | 94           |
| 32  | DQ445493             | 4                   | China (Changchun) | 94           |
| 33  | DQ445494             | 4                   | China (Changchun) | 93           |
| 34  | DQ445495             | 4                   | China (Changchun) | 94           |
| 35  | EU332143             | 4                   | China (Heilongjiang) | 94           |
| 36  | EU332144             | 4                   | China (Heilongjiang) | 94           |
| 37  | EU332145             | 4                   | China (Heilongjiang) | 94           |
| 38  | FJ172540             | 4                   | China (Heilongjiang) | 92           |
| 39  | FJ172550             | 4                   | China (Inner Mongolia) | 93           |
| 40  | AF117279             | 4                   | Taiwan   | 93           |
| 41  | AJ344180             | 4                   | China     | 92           |
| 42  | AB550624             | 4                   | China (Inner Mongolia) | 93           |
mini-kit (Qiagen, Hilden, Germany) according to the manufacturer’s instructions. Purified RNA was used to generate the ORF2 of HEV using One-Step RT-PCR with a PLATINUM Taq Kit (Takara, Shiga, Japan). Briefly, RT-PCR was performed on 5 µL of purified RNA from serum in 50 µL of 2× reaction mix with 0.2 µM each primer (forward: 5’-aggttgccgtctgcgaga-3’; reverse: 5’-acagtcgtgccgattgcg-3’). Reverse transcription was performed for 30 min at 50°C and for 2 min at 94°C followed by 40 cycles of 94°C for 30s, 55°C for 30s and 72°C for 2 min, and a final extension step of 10 min at 72°C. The second round of PCR was performed under the same conditions as the first-round PCR with 0.2 µM each of the same primers.

Phylogenetic analysis

The relationship between the sequences is shown in the dendrogram. The length of each pair of branches represents the distance between the sequences. Phylogenetic trees were constructed by the neighbor-joining method based on the ORF2 sequence. All strains were separated into four groups according to their genotypes. Sequence analysis was conducted using the software VectorNTI and MEGA 4.1 beta. Bootstrap values are indicated for the major nodes as a percentage of the data obtained from 1000 resamplings.

We identified genotype 4 human HEV from the 43-year-old Korean male patient with acute hepatitis, who had never been abroad. He was negative for serum markers of hepatitis A, B and C viruses and positive for anti-hepatitis E virus as IgM class leading to the diagnosis of hepatitis E, whose ORF2 nucleotide sequence from the patient was isolated. The identified Korean HEV strain named 2010-JKSH-CKK (HM769726) belonged to genotype 4, based on comparison with previously reported HEV strains Table 2.

| Case ref | Age (yr) /Sex | Presenting symptom | Suspicious transmission route | IgM anti-HEV (titer/cut-off) | IgG anti-HEV (titer/cut-off) | Serum HEV RNA | Stool HEV RNA | Outcome |
|----------|---------------|-------------------|-------------------------------|----------------------------|-----------------------------|---------------|---------------|---------|
| #117     | 27/M          | Fever, myalgia    | UK                            | + (1.22/0.45)               | +                           | ND            | ND            | Recovery |
| #217     | 28/F          | Fever, diarrhea   | Travel/India                  | + (1.77/0.44)               | + (4.0/0.58)                | ND            | ND            | Recovery |
| #317     | 34/F          | Jaundice          | UK                            | + (1.30/0.46)               | + (1.27/0.62)               | ND            | ND            | Recovery |
| #417     | 42/M          | Abdominal pain    | UK                            | + (1.10/0.43)               | + (3.5/0.53)                | ND            | ND            | Recovery |
| #518     | 30/M          | Fatigue           | UK                            | +                           | -                           | ND            | ND            | Recovery |
| #618     | 25/F          | Jaundice, fatigue | UK                            | +                           | -                           | ND            | ND            | Recovery |
| #718     | 24/M          | Jaundice          | Travel/India                  | +                           | -                           | ND            | ND            | Recovery |
| #818     | 29/M          | Jaundice          | UK                            | +                           | -                           | ND            | ND            | Recovery |
| #918     | 52/F          | Jaundice          | UK                            | +                           | -                           | ND            | ND            | Recovery |
| #1018    | 45/M          | Fatigue           | UK                            | +                           | -                           | ND            | ND            | Recovery |
| #1118    | 48/F          | Jaundice          | UK                            | +                           | -                           | ND            | ND            | Recovery |
| #1218    | 42/F          | Jaundice          | UK                            | +                           | -                           | ND            | ND            | Recovery |
| #1318    | 23/F          | Jaundice          | UK                            | +                           | -                           | ND            | ND            | LT       |
| #1419    | 42/F          | Fever             | UK                            | +                           | ND                           | ND            | ND            | Recovery |
| #1519    | 33/M          | Fatigue           | UK                            | +                           | ND                           | ND            | ND            | Recovery |
| #1619    | 40/F          | Fatigue           | UK                            | +                           | ND                           | ND            | ND            | Recovery |
| #178     | 51/F          | Jaundice          | UK                            | +                           | +                           | Detected genotype 4 | ND       | Recovery |
| #1822    | 54/M          | Jaundice          | Wild boar bile juice          | + (3.62/0.30)               | + (3.73/0.38)               | Detected genotype 4 | Detected genotype 4 | Recovery |
| #19†     | 43/M          | Jaundice          | Roe deer meat                 | + (3.66/0.28)               | + (3.38/0.38)               | Detected genotype 4 | ND       | Recovery |

Ref, reference; HEV, hepatitis E virus; UK, unknown; ND, not detected; LT, liver transplantation. *underlying autoimmune hepatitis. †present case.
DISCUSSION

In 1997, an animal strain of HEV was isolated from pig, characterized as swine HEV, which was revealed to be genetically associated with human HEV in the United States. Subsequently, a number of swine HEVs were isolated and proven to be closely associated genetically with human HEV. Unlike genotype 1 and 2 HEV, genotype 3 and 4 HEV are isolated from both human and animals. Recently, sporadic cases of genotype 3 and 4 hepatitis E has been reported in non endemic area, such as France, Germany, and Japan. In Germany, an autochthonous genotype 3 HEV infection was reported even in a 5-month-old female child. In Japan where raw fish and uncooked or undercooked meat are part of the traditional diet, there have been some case reports of zoonotic food-borne transmission, though domestic pig, wild boar and wild deer are the main routes for autochthonous HEV infection. For example, in 2003, hepatitis E was diagnosed in a family who had previously ingested raw meat of a wild roe deer. HEV genotype 3 was isolated from the remnant raw meat and the nucleotide sequence homology of HEV was verified between the family and the meat. Following this event, there were some published articles about zoonotic transmission of HEV in Japan.

In South Korea, there have been case reports of HEV with some reports of HEV serologically diagnosed since 2002; however, only three of them including our case could detect serum HEV RNA (Table 2). In the interpretation of HEV serological test, we should consider the possibility of false-positive result, especially in nonendemic areas. The detection of IgM anti-HEV in serial samples as well as serum or stool HEV RNA detection is helpful to confirm diagnosis. Two Korean cases of genotype 4 hepatitis E reported in 2010 and 2011 are similar to our case with a nucleotide similarity of 97 and 98%, respectively. Our case is the first human case of autochthonous genotype 4 hepatitis E from a roe deer, while most of the HEV infections previously reported were by zoonotic transmission from a domestic pig or a wild boar in South Korea. Accumulating data supports the notion that zoonotic transmission is the main route of autochthonous infection of HEV, which is an increasingly interesting issue.

Even with an increasing interest in acute hepatitis E, HEV is still regarded as an infrequent cause of acute viral hepatitis. Perhaps this is because most asymptomatic patients with newly elevated levels of aminotransferase receive supportive care without a definitive diagnosis, which can possibly underestimate the incidence of hepatitis E. Therefore, physicians should be on the alert for HEV infection in patients with acute hepatitis of unknown etiology in developed countries. Furthermore, it is important to check whether they intake the raw meat of wild animals in patients without a travel history of HEV endemic area.

Conflicts of Interest

The authors have no conflicts to disclose.

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