Effect of Antioxidants of Sage tea and Marjoram tea on Advanced Renal Disease

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Abstract

Chronic kidney disease (CKD) is a worldwide public health problem. Oxidative stress is the causative factor for a wide variety of diseases, including CKD. Medicinal plants used in the management of CKD are effective in renal detoxification and help the effects of dialysis treatment. This study was conducted to investigate the effect of aqueous extract of sage (Salvia Officinal) and marjoram (Origanum Majoranum) on advanced chronic kidney patients under dialysis. The experiment was carried out on sixty patients (40-50 years old), diagnosis based on detailed clinical history, clinical examination and other relevant biochemical investigations. The patients were divided into 6 groups (each group contain 10 CKD patients under treatment with (hemodialysis and regular medical treatment for 3 months) as followed: control patients were treated with regular medical treatment, other patients consumed aqueous extracts of (5g sage, 5g marjoram, 10g sage, 10g marjoram, mixture of 5g sage +5g marjoram) twice today respectively. Results illustrated that the aqueous extract of marjoram or sage are rich in antioxidants components (Phenolic acids, Flavonoids, Oxygenated monoterpenes, Diterpenoids and Triterpenes), antioxidant capacity and phenolic content. Results clarify that glomerular filtration rate (GFR) increased, while erythrocyte sedimentation rate (ESR) and C-reactive protein (CRP) significantly decreased (P<0.05) in all treated patients compared to the control group. Malondialdehyde (MDA) decreased and superoxide dismutase (SOD) increased. The mixture of aqueous extracts of marjoram (5g) and sage (5g) recorded the best drink. Therefore, this study recommends the use of the aqueous extracts of marjoram (5g) and sage (5g) in decreasing the oxidative stress and improve kidney health in hemodialysis patients.

Key words: hemodialysis, oxidative stress, antioxidants, sage, marjoram superoxidizedismutase, malondialdehyde, patients.

Introduction

The kidneys filter plasma in the glomerulus to form a protein-free ultrafiltrate. This ultrafiltrate passes through the various tubular segments where reabsorption of essential constituents and secretion of unwanted products occur (Byham-Gray et al., 2014). CKD is a worldwide public health problem characterized by either reduced glomerular filtration rate or the presence of kidney damage that leads to abnormal kidney function and higher proteinuria, blood urea nitrogen, and serum creatinine levels (Romagnaniet al., 2017).
Oxidative stress is the causative factor for a wide variety of diseases, including CKD. Antioxidants are the molecules that combat the oxidative stress developed from an imbalance between the rate of production and removal of produced oxidants (Amarasiria et al., 2020).

Medicinal plants are rich sources of bioactive compounds that have been reported to exert nephron protective mechanisms, such as antioxidant (mainly in the form of phenolic compound), anti-inflammatory, diuretic, and immunomodulation. Hence, (El-Wakfet et al., 2020) predicted that these plants are of utmost importance to protect renal function and slow the occurrence and the progression of CKD.

Among such plants, sage (Salvia officinalis L.) and marjoram (Origanum majorana L.) have been described to contain high percentage of biologically active polyphenolic compounds which are useful to human health and have effective therapeutic benefits (El-Wakfet et al., 2020). Sage is considered as one of the most popular herbs consumed widely and traditionally as an herbaceous infusion. The incorporation of sage infusion in the daily diet can provide considerable benefits, being anti-mycotic, anti-carcinogenic, antidiabetic, antimicrobial, anti-inflammatory, antioxidant, and anti-proliferative. In addition to these effects, sage infusion exhibits antiradical activity which correlates strongly with their high level of total phenolic content. Rosmarinic acid, salvianolic acid K, and luteolin-7-O-glucuronide were detected as the main phenolic compounds of sage aqueous extract (Sotiropoulou et al., 2020). Marjoram is widely utilized as a spice and for better food flavor. It is a strong remedy for coughs, respiratory infections, cardiovascular disorders, skin lesions, and digestive problems. Marjoram could also assist in managing liver and kidney diseases (El-Wakfet et al., 2020).

Aim of the study
The aim of this study was to investigate the effect of aqueous extract of sage (Salvia Officinalis) and marjoram (Origanum Majoranum) on advanced chronic kidney patients under dialysis.

Materials, Patients and Methods

Materials: Sage (Salvia officinalis) and marjoram (Origanum majorana L.) were obtained from Family Pharmacia Company for Herbs, 38 Sharkiaind. Zone, Beibis – 10th of Ramadan road.

Patients: Sixty (40-50 years old) patients suffering from chronic kidney disease selected from Al-Azhar University Hospital, New Damietta, Egypt. The patients were diagnosed after detailed clinical history, clinical examination and other relevant biochemical investigations. Patients were divided into 6 groups (each group contain 10 CKD patients under treatment with hemodialysis) and regular medical treatment as followed: control patients treated with regular medical treatment, other patients consumed aqueous extracts of (5g sage, 5g marjoram, 10g sage, 10g marjoram and mixture of 5g sage +5g marjoram) respectively twice daily for 3 months.

Methods
A. Aqueous extracts preparation:

The air parts of plants were dried (5 grams) and treated with double distilled water at 100 degrees C, for a period of 15 minutes. The samples were filtered. The resultant filtrates were made off up to 50 ml with double-distilled water. Three samples for each analyzed condiment plant were done according to (Al-Turkiet al., 2007).
B. Determination of antioxidant activity by DPPH method:

Antioxidant activity was determined in the extracts using the 2, 2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging method. This method was taken from (Buřičova and Reblova, 2008 and Lo Scalzo, 2008).

C. Determination of total phenolic compounds:

The content of total phenolics was determined spectrophotometrically at 760 nm by using Folin-Ciocalteu reagent. The results were expressed as the content of gallic acid per unit mass of the sample (Dorman et al., 2003 and Stratilet et al., 2008).

D. Biochemical Assay

Blood samples were collected at zero time and at the end of the study. The serum was separated within 2 hours after blood withdrawal, by centrifugation at 3000 rpm for 10 minutes. The serum samples were kept frozen at -20°C until determination parameters in the laboratory of Al –Azhar University Hospital in New Damietta. Hemoglobin (HGB), red blood cell (RBC), white blood cells (WBC) and platelets (Pit) count were measured using automated hematology analyzer (Sysmex, Kobe, Japan). ESR was measured using (Westergren, 1926) method. Serum creatinine was determined according to (Kaplan, 1984 and Murry, 1984). Malondialdehyde (MDA) determined according to (Satoh, 1978 ), Superoxide dismutase was determined according to the Indirect Method (Flohe and Ötting, 1984). Serum C-reactive protein (CRP) was analyzed by an automated analyzer (Olympus AU400) with a latex turbidimetric immunoassay kit (CRP-UL assay, Wako Chemicals, Neuss, Germany). The glomerular filtration rate (GFR) was estimated according to criteria of the National Kidney Foundation/ Kidney Disease Outcomes Quality Initiative, using the equation proposed by (Levey et al., 1999).

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GFR = 175 \times \text{standardized } \text{Scr}^{-1.154} \times \text{age}^{-0.203} \times 1.212 \times (\text{if black}) \times 0.742 \times (\text{if female}).
\]

E. Statistical analysis:

Data was statistically analyzed using (SPSS) software version (18). Analysis of variance (ANOVA) was used to show the significance P<0.05 among groups (SPSS, 1986).

Results and Discussion

Chemical composition of aqueous extract of marjoram and sage:

The results illustrated that the aqueous extract of marjoram and sage are rich in antioxidants components as (Phenolic acids, Flavonoids, Oxygenated monoterpenes, Diterpenoids and triterpenes). Marjoram aqueous extract are rich in Terpinen-4-ol (20.23%), followed by vanillic acid (14.44%), ursolic acid (13.09 %), α -terpineol (7.64%) and transpinene hydrate (6.63%) and other components as shown in Fig (1).The main compounds in sage aqueous extract were, rosmarinic acid (26.03%), luteolin-7-O-glucuronide (16.5%), caffeic acid (12.69%), ursolic acid (11.43%) and apigenin (6.6%) as shown in Fig (2).These results agree with that reported by (Beddows et al. 2000, 2001; Exarchou et al. 2002; Fasseas et al. 2007; Celikel and Kavas 2008; Calikoglu et al. 2009; Chrpova et al., 2010; Abdel-Massih et al., 2010;Miron et al., 2011and Bina and Rahimi, 2017) who approved the high phenolic content and high antioxidants components of sage and marjoram. They detected that rosmarinic acid is the main phenolic compounds of sage aqueous extract and terpinen-4-olis the main component in marjoram aqueous extract with varying proportions of the other mentioned ingredients.
Fig (1):
The chemical constitutions of marjoram aqueous extract.

Fig (2):

The chemical constitutions of sage aqueous extract.
Antioxidant capacity and total phenolic contents of plants aqueous extract:

Table (1) shows the strongest antioxidant capacity of the aqueous extracts of marjoram and sage. The total antioxidant capacity (DPPH) of the aqueous extracts of marjoram and sage were (42.1 and 60.6), respectively and the phenolic content values were expressed as mg equivalent of gallic acid (GAE) were (27.7 and 24.3), respectively. This result exhibited the strongest antioxidant capacity of the aqueous extracts of marjoram and sage which agreed with (Roby et al., 2013 and Dabija et al., 2018).

Table (1):

| Herbs  | Latin name      | Family  | DPPH (mg AAE/g)* | Phenols (mg GAE/g)** |
|--------|-----------------|---------|------------------|----------------------|
| Marjoram  | Origanum majorana L. | Lamiaceae | 42.1              | 27.7                 |
| Sage    | Salvia officinalis L. | Lamiaceae | 60.6              | 24.3                 |

*AAE = ascorbic acid equivalent ** GAE = gallic acid equivalent

Effect of sage and marjoram extracts on some parameters of hemodialysis patients:

Table (2) compares the results before and after using the aqueous of herbs. The control group which includes the hemodialysis patients had increase in ESR and CRP which indicate inflammation occurrence, this result accepted with (Fine, 2002) with slightly improvement in GFR. But the treated groups by the aqueous extracts of sage and marjoram had a noticeable decrease in ESR, CRP with some variability among them and a noticeable increase in GFR for all other groups. Group (6) which was treated with mixture of the aqueous extract of marjoram (5 g) and sage (5 g) recorded the best result in GFR, which denotes the best glomerular function. Group (2) which was treated with the aqueous extract of sage (5 g) recorded the best decrease in ESR unlike CRP which had the best decrease with group (5) which was treated with the aqueous extract of marjoram (10 g) that denotes the best decrease in inflammation occurrence. However, this illustrates the good effect of using the aqueous of used herbs.

Table (3) showed the good effect of herbs that improve the kidney function as shown by the level of creatinine, whereas creatinine value increased in the control comparing to all treated groups which was had decreased in creatinine. The response in group (6) which consumed the mix of aqueous extract of herbs recorded the best decrease in creatinine.
Table (2):
Effect of the Aqueous Extract of Sage and Marjoram on Glomerular Filtration Rate (GFR), Erythrocyte Sedimentation Rate (ESR) and C-reactive protein (CRP) in hemodialysis patients:

| Groups                  | Parameters                  | Before          | After          | Before          | After          | Before          | After          |
|-------------------------|-----------------------------|-----------------|----------------|-----------------|----------------|-----------------|----------------|
|                         | GFR (ml/min/1.73 m²)        |                 |                |                 |                |                 |                |
| Group 1: (control)      |                             | 7.57±0.36       | 8.48±0.32      | 38.30±4.36      | 48.70±3.54     | 8.00±1.03       | 9.30±0.79      |
| Group 2: (5g sage)      |                             | 10.62±1.03      | 12.86±0.70     | 22.30±1.91      | 12.40±2.02     | 9.00±1.04       | 7.20±0.80      |
| Group 3: (10g sage)     |                             | 9.93±0.74       | 12.85±1.08     | 44.60±7.62      | 13.60±2.74     | 9.80±0.86       | 6.85±0.58      |
| Group 4: (5g marjoram)  |                             | 9.13±0.41       | 11.26±0.66     | 33.10±3.03      | 19.00±2.38     | 7.80±0.91       | 6.82±0.53      |
| Group 5: (10g marjoram) |                             | 10.04±0.37      | 11.11±0.59     | 55.10±4.95      | 20.40±3.02     | 8.60±1.07       | 6.78±0.58      |
| Group 6: (5g marjoram + 5g sage) |   | 10.08±0.56      | 15.22±0.28     | 34.30±3.01      | 15.41±1.72     | 9.50±0.87       | 6.89±0.24      |
|                         | ESR (mm/hr)                 |                 |                |                 |                |                 |                |
| Group 1: (control)      |                             | 8.00±1.03       | 9.30±0.79      | 12.40±2.02      | 7.20±0.80      | 9.80±0.86       | 6.85±0.58      |
| Group 2: (5g sage)      |                             | 9.00±1.04       | 7.20±0.80      | 12.40±2.02      | 7.20±0.80      | 9.80±0.86       | 6.85±0.58      |
| Group 3: (10g sage)     |                             | 9.80±0.86       | 6.85±0.58      | 12.40±2.02      | 7.20±0.80      | 9.80±0.86       | 6.85±0.58      |
| Group 4: (5g marjoram)  |                             | 7.80±0.91       | 6.82±0.53      | 19.00±2.38      | 6.82±0.53      | 9.80±0.86       | 6.85±0.58      |
| Group 5: (10g marjoram) |                             | 8.60±1.07       | 6.78±0.58      | 20.40±3.02      | 6.78±0.58      | 9.80±0.86       | 6.85±0.58      |
| Group 6: (5g marjoram + 5g sage) |                   | 9.50±0.87       | 6.89±0.24      | 15.41±1.72      | 6.89±0.24      | 9.80±0.86       | 6.85±0.58      |
|                         | CRP (mg/L)                  |                 |                |                 |                |                 |                |
| Group 1: (control)      |                             | 8.00±1.03       | 9.30±0.79      | 12.40±2.02      | 7.20±0.80      | 9.80±0.86       | 6.85±0.58      |
| Group 2: (5g sage)      |                             | 9.00±1.04       | 7.20±0.80      | 12.40±2.02      | 7.20±0.80      | 9.80±0.86       | 6.85±0.58      |
| Group 3: (10g sage)     |                             | 9.80±0.86       | 6.85±0.58      | 12.40±2.02      | 7.20±0.80      | 9.80±0.86       | 6.85±0.58      |
| Group 4: (5g marjoram)  |                             | 7.80±0.91       | 6.82±0.53      | 19.00±2.38      | 6.82±0.53      | 9.80±0.86       | 6.85±0.58      |
| Group 5: (10g marjoram) |                             | 8.60±1.07       | 6.78±0.58      | 20.40±3.02      | 6.78±0.58      | 9.80±0.86       | 6.85±0.58      |
| Group 6: (5g marjoram + 5g sage) |                   | 9.50±0.87       | 6.89±0.24      | 15.41±1.72      | 6.89±0.24      | 9.80±0.86       | 6.85±0.58      |

Data expressed as Means ± SE.
Means with different superscript letters in the same column are significantly different at P<0.05.

Table (3):
Effect of the Aqueous Extract of Sage and Marjoram on creatinine on patients:

| Groups                  | Creatinine (mg/dl) |
|-------------------------|-------------------|
|                         | Before            | After             |
| Group 1: (-ve control)  | 5.74±0.64         | 8.47±0.39         |
| Group 2: (5g sage)      | 8.14±0.61         | 6.38±0.44         |
| Group 3: (10g sage)     | 7.35±0.79         | 6.23±0.76         |
| Group 4: (5g marjoram)  | 7.45±0.53         | 6.60±0.47         |
| Group 5: (10g marjoram) | 7.26±0.55         | 5.71±0.39         |
| Group 6: (5g marjoram + 5g sage) | 8.26±0.30 | 4.05±0.23 |

Data expressed as Means ± SE.
Means with different superscript letters in the same column are significantly different at P<0.05.

Table (4) shows the blood picture results in groups before and after the experiment. It shows the decrease in Hb, RBCs, WBCs but not the platelets count in the control group. All other values increased after taking the extracts compared to the control group. This result agreed with (Hakim et al., 2016). The treated groups after the end of experiment compared with the control, showed an increase in Hb, RBCs, WBCs and Plts near the normal values. The best increase in Hb was in group (2) which consumed sage (5 g) aqueous extract, followed by group (3) which consumed sage (10 g)
aqueous extract. Group (6) which consumed the mix of aqueous extract of herbs recorded the best increase in WBCs and RBCs values. The last results indicate the improvement of hemodialysis patients' cases.

### Table (4):
**Effect of the Aqueous Extract of Sage and Marjoram on Complete Blood Count in hemodialysis patients**

| Groups                  | CBC                      |
|-------------------------|--------------------------|
|                         | Hb (g/dL) | RBC (×10^{12}/L) | WBC (×10^{3}/μL) | PLT (×10^{3}/μL) |
|                         | Before    | After          | Before          | After            | Before    | After            |
| Group 1: (+ ve control) | 10.65±    | 8.56±         | 3.92±          | 3.03±             | 8.37±    | 4.16±             | 108.10± | 117.9±           |
|                         | 1.07      | 0.44           | 0.20           | 0.15              | 1.25     | 0.37              | 1.81    | 2.47             |
| Group 2: (5g sage)      | 9.29±     | 11.11         | 4.07±          | 4.45±             | 4.48±    | 5.56±             | 109.20± | 140.1±           |
|                         | 0.24      | ± 0.16        | 0.15           | 0.17              | 0.25     | 0.31              | 5.03    | 4.06             |
| Group 3: (10g sage)     | 9.41±     | 11.02±        | 3.55±          | 3.77±             | 5.20±    | 5.87±             | 132.40± | 160.6±           |
|                         | 0.18      | 0.30           | 0.12           | 0.13              | 0.31     | 0.31              | 3.27    | 5.79             |
| Group 4: (5g marjoram)  | 8.32±     | 10.09±        | 3.57±          | 3.90±             | 4.27±    | 5.92±             | 117.50± | 131.9±           |
|                         | 0.57      | 0.41           | 0.31           | 0.25              | 0.54     | 0.66              | 3.20    | 2.96             |
| Group 5: (10g marjoram) | 8.66±     | 10.16±        | 3.83±          | 4.33±             | 5.33±    | 6.24±             | 121.30± | 134.6±           |
|                         | 0.30      | 0.30           | 0.25           | 0.27              | 0.34     | 0.52              | 6.09    | 5.33             |
| Group 6: (5g marjoram + 5g sage) | 9.77±   | 10.59±        | 3.95±          | 4.95±             | 5.80±    | 6.66±             | 116.±   | 151.8±           |
|                         | 0.65      | 0.54           | 0.17           | 0.28              | 0.41     | 0.67              | 5.11    | 7.88             |

Data expressed as Means ± SE.
Means with different superscript letters in the same column are significantly different at P< 0.05.

Results in Table (5) showed a decrease in MDA and increase in SOD in all treated groups compared with the control group. The best result was recorded in group (6).

### Table (5):
**Effect of the Aqueous Extract of Sage and Marjoram on MDA on hemodialysis patients:**

| Groups                  | MDA (NMO/ml) | SOD (U/ml) |
|-------------------------|--------------|------------|
|                         | Before       | After      | Before       | After          |
| Group 1: (-ve control)  | 105.40±4.64  | 213.90±8.81| 944.60±18.74| 779.20±29.31  |
| Group 2: (5g sage)      | 405.20±10.82 | 194.50±3.29| 607.50±7.47 | 911.80±44.48  |
| Group 3: (10g sage)     | 266.10±5.66  | 175.9±9.10 | 682.80±10.35| 940.40±16.72  |
| Group 4: (5g marjoram)  | 287±13.83    | 161.6±7.82 | 554.7±19.70 | 959.6±35.35   |
| Group 5: (10g marjoram) | 264.6±10.08  | 142.7±3.17 | 724.7±14.94 | 949.4±12.11   |
| Group 6: (5g marjoram + 5g sage) | 296.6±14.02 | 119.2±3.35 | 650±20.18    | 1050.2±22.55  |
These results indicate that chronic kidney patients treated by hemodialysis suffered from increase oxidative stress which agreed with (small et al., 2012) and antioxidants improved kidney health which agreed with (El-Ashmawy et al., 2005; 2007). The aqueous extracts of sage and marjoram had antioxidant effect which agreed with (Nakatani, 2000; Novak et al., 2000; Heo et al., 2002; Kelly, 2004; Hazzit et al., 2006; Ayatollahi et al., 2009; Ahmed et al., 2009 and Hossain et al., 2010) and improved kidney function that agreed with (Halliwell and Gutteridge, 2007; Nasri and Rafieian-Kopaei, 2014) This appeared in the improvement of GFR and the decline in CRP, ESR and creatinine with an increase in Hb, WBCs, RBCs and platelets. It is worth noting that the groups that registered the improvement was group (6) which consumed the mixture of aqueous extract of sage (5g) and marjoram (5g) with the difference in the positive effect of other doses used in the experiment on patients (El-Ashmawy et al., 2005).

**Conclusion**

The study confirmed the antioxidant effect of the aqueous extract of sage and marjoram on chronic kidney patients treated with hemodialysis since it include (flavonoids, phenolic acids and terpenes) The best drink was the mix of aqueous extract of both sage (5g) + marjoram (5g). It is recommended to repeat the study on higher number of patients.

**References**

Abdel-Massih R.M., Fares R., Bazzi S., El-Chami N. and Baydoun E., (2010):
The apoptotic and anti-proliferative activity of Origanum majorana extracts on human leukemic cell line. Leuk. Res., 34(8): 1052-1056.

Ahmed L.A., Ramadan R.S. and Mohamed R.A., (2009):
Biochemical and histopathological studies on the water extracts of Marjoram and chicory herbs and their mixture in obese rats. Pak. J. Nutr., 8(10): 1581-1587.

Al-Turki A. I., El-Ziney M. G. and Abdel-Salam A. M., (2007):
Chemical and anti-bacterial characterization of aqueous extracts of oregano, marjoram, sage and licorice and their application in milk and labneh, Journal of Food, Agriculture & Environment; 6(1) : 39 - 44.

Amarasiria S. S., Attanayakeb A. P., Jayatilakab K. A. P.W. and Mudduwac L. K. B., (2020):
Nephroprotective South Asian medicinal plants in the treatment of chronic kidney disease: a review. Traditional Medicine Research; 5 (5): 389–412.

Ayatollahi A., Shojaii A., Kobarfard F., Mohammadzadeh M. and Choudhary M., (2009):
Two flavones from Salvia iberifolia. Iran J Pharm Res; 8:179–84.

Beddows C.G., Jagait C., Kelly M.J. (2000):
Preservation of α-tocopherol in sunflower oil by herbs and spices. International Journal of Food Science and Nutrition, 51: 327–339.

Beddows C.G., Jagait C., Kelly M.J. (2001):
Effect of ascorbylpalmitate on the preservation of α-tocopherol in sunflower oil, alone and with herbs and spices. Food Chemistry, 73: 255–261.

Bina F. and Rahimi, R., (2017):
Sweet Marjoram: A Review of Ethnopharmacology, Phytochemistry, and Biological Activities, Journal of Evidence-Based Complementary & Alternative Medicine; 22(1): 175-185.

Buřičova L., Reblova Z. (2008):
Czech medicinal plants as possible sources of antioxidants. Czech Journal of Food Sciences, 26: 132–138.

Byham-Gray, L.D. Burrowes J.D. and Chertow G. M., (2014):
Nutrition in Kidney Disease, Second Edition, Nutrition and Health, 3 DOI 10.1007/978-1-62703-685-6_1, © Springer Science +Business Media New York.

Calikoglu E., Kiralan M., Bayrak A. (2009):
Effect of direct applications of sage (Salvia officinalis L.) leaves on oxidative stability of sunflower oil during accelerated storage. Journal of Food Quality, 32: 566–576.

Celikel N., Kavas G. (2008):
Antimicrobial properties of some essential oils against some pathogenic microorganisms. Czech Journal of Food Sciences; 26: 174–181.

Chrpova D., Kouřimska L., Gordon M.H., Heřmanova V., Roubičkova I., Panek J. (2010):
Antioxidant activity of selected phenols and herbs used in diets for medical conditions. Czech J. Food Sci., 28: 317–325.

Dabija A., Codină G.G., Ropciuc S. and G’atlan A., (2018):
Assessment of the Antioxidant Activity and Quality Attributes of Yogurt Enhanced with Wild Herbs Extracts, Journal of Food Quality; 2018:12.

Dorman H.J.D., Pelтокeto A., Hiltunen R., Tikkanen M.J. (2003):
Characterisation of the antioxidant properties of de-odourised aqueous extracts from selected Lamiaceae herbs. Food Chemistry, 83: 255–262.

El-Ashmawy I.M., el-Nahas A.F., Salama O.M., (2005):
Protective effect of volatile oil, alcoholic and aqueous extracts of Origanum majorana on lead acetate toxicity in mice. Basic Clin Pharmacol Toxicol; 97:238-43.

El-Ashmawy I.M., Saleh A. and Salama O.M., (2007b):
Effects of marjoram volatile oil and grape seed extract on ethanol toxicity in male rats. Basic Clin Pharmacol Toxicol., 101(5): 320-327.
El-Wakf A. M., El- Habibi E. M., Ali D. A., Abd El-Ghany E. and Elmougy R., (2020):
Marjoram and sage oils protect against testicular apoptosis, suppressed Ki-67 expression and cell cycle arrest as a therapy for male infertility in the obese rats; J Food Biochem;44:e13080.

Exarchou V., Nenadis N., Tsimidou M., Gerohanassis I.P., Troganis A., Boskou D. (2002):
Antioxidant activities and phenolic composition of extracts from Greek oregano, Greek sage, and summer savory.Journal of the Agricultural and Food Chemistry; 50:5294–5299.

Fasseas M.K., Mountzouris K.C., Tarantilis P.A., Polissiou M., Zervas G. (2007):
Antioxidant activity in meat treated with oregano and sage essential oils.Food Chemistry; 106:1188–1194.

Fine A., (2002):
Relevance of C-reactive protein levels in peritoneal dialysis patients. Kidney Int 61:615-620.

Flohe L. & Otting F.,(1984):
Superoxide Dismutase Assays, Methods in Enzymology 105:93-104 (1984).

Hakim Y. A. H., Abbas A. A., Khalil A. and Ibrahim H. A. M., (2016):
The Effect of Hemodialysis on Hemoglobin Concentration, Platelets count and White Blood Cells Count in End Stage Renal Failure, International Journal of Medical Research & Health Sciences; 5, 5:22-35.

Halliwell B. and Gutteridge J., (2007):
Free Radicals in Biology and Medicine. 4th ed. Oxford: Oxford University Press.

Hazzit M., Baaliouamer A., Leonor-Faleiro M. and Graca M.M., (2006):
Composition of the essential oils of Thymus and Origanum species from Algeria and their antioxidant and antimicrobial activities. J. Agric. Food Chem., 54(17): 6314-6321.

Heo H.J., Cho H.Y., Hong B., Kim H.K., Heo T.R., Kim E.K., Kim S.K., Kim C.J. and Shin D.H. (2002):
Ursolic acid of Origanum majorana L. reduces Abeta-induced oxidative injury. Mol. Cells; 13(1): 5-11.

Hossain M.B., Brunton N.P., Barry-Ryan C., Martin-Diana A.B. and Wilkinson M., (2010a):
Characterization of phenolics composition in Lamiaceae spices by LC-ESI-MS/MS. J. Agric. Food Chem.; 58(19): 10576-1058.

Kaplan, L.A. (1984):
Clin Chem. The C.V. Mosby co.st Louis. Toronto.

Kelly W.J., (2004):
Herbal medicine handbook. Lippincott Williams Wilkins AWolters, Kluwer Co.: 289-290.

Levey A. S., Coresh J., Balk E., Kausz T., et al., (2003):

50
National Kidney Foundation guidelines for chronic kidney disease: evaluation, classification, and stratification. Ann Intern Med. 2003; 139: 137–47.

Lo Scalzo R. (2008):
Organic acids influence on DPPH· scavenging by ascorbic acid. Food Chemistry, 107: 40–43.

Miron T.L., Plaza M., Bahrim G., Ibariez E. and Herrero M., (2011):
Chemical composition of biocative pressurized extracts of Romanian aromatic plants. J. Chromatogr. A. PMID: 21163488.

Murry, R. (1984):
Clin Chem The C.V. mosby co. st Louis. Toronto. Princeton: 1088-1090.

Nakatani N., (2000):
Phenolic antioxidants from herbs and spices. Biofactors; 13(1-4): 5-11.

Nasri H. and Rafieian-Kopaei M., (2014):
Protective effects of herbal antioxidants on diabetic kidney disease. J Res Med Sci; 19:82-3.

Novak J., Bitsch C., Langbehn J., Pank F., Skoula M., Gotsiou Y. and Franz C.M., (2000):
Ratios of cis- and trans-sabinene hydrate in Origanum majorana L. and Origanum microphyllum (Bentham) Vogel. Biochem. System Ecol., 28(7): 697-704.

Roby M.H.H., Sarhan M.A., Selim KA-H., Khalel K.I., (2013):
Evaluation of antioxidant activity, total phenols and phenolic compounds in thyme (Thymus vulgaris L.), sage (Salvia officinalis L.), and marjoram (Origanum majorana L.) extracts. Ind Crops Prod; 43:827-31.

Romagnani P., Remuzzi G., Glassock R., Levin A., Jager K. J., Tonelli M., et al., (2017):
Chronic kidney disease. Nature Reviews Disease Primers; 3: 17088.

Satoh K. (1978):
Clinica,Chemica ,Acta; 90: 37.

Small D.M., Coombes J.S., Bennett N., Johnson D.W. and Gobe G.C., (2012):
Oxidative stress, anti-oxidant therapies and chronic kidney disease. Nephrology; 17:311-21.

Sotiropoulou N. S., Megremi S.F. and Tarantilis P., (2020):
Evaluation of Antioxidant Activity, Toxicity, and Phenolic Profile of Aqueous Extracts of Chamomile (Matricaria chamomilla L.) and Sage (Salvia officinalis L.) Prepared at Different Temperatures, Sci.; 10, 2270

SPSS, (1986):
"Statistical package for social science!, version 11. SPSS Inc., Il. U.S.A

Stratil P., Kučiš V., Fojtova J. (2008):
Comparison of the phenolic content and total antioxidant activity in wines as determined by spectrophotometric methods. Czech Journal of Food Sciences, 26: 242–253.

Westergren A., (1926):
The technique of the red cell sedimentation reaction. Am Rev Tuberc.; 14:94–101.
تأثير مضادات الأكسدة لشاي البردقوش وشاي المريمية على مرضى الكلى المزمنة في المرحلة المتقدمة

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الملخص العربي
مرض الكلى المزمن هو مشكلة صحية عالمية. الإجهاد التأكسدي هو العامل المسبب لمجموعة متنوعة من الأمراض، بما في ذلك مرض الكلى المزمن. النباتات الطبية المستخدمة في علاج مرض الكلى المزمن فعالة في إزالة السموم من الكلى وتقليل آثار علاج الغسيل الكلوي. تم إجراء هذا العمل لمعرفة تأثير المستخلص المائي للبردقوش والمريمية على المرحلة المتقدمة لمرضى الكلى المزمن المعالجين بالغسيل الكلوي. تم إجراء التحقيق على ستين مريضاً بينهم 40-50 سنة، مقسمة إلى 6 مجموعات (كل مجموعة تحتوي على 10 مرضى الكلى المزمن) مع علاج الغسيل الدموي وعلاج طبي منتظم لمدة 3 أشهر على النحو التالي: مجموعة ضابطة تحتوي على مرضى تم علاجهم بالعلاج الطبي المنتظم وتناول المرضى الآخرون مستخلصات مائية كالآتي: (5 جم مريمية، 5 جم بردقوش، 10 جم مريمية، 10 جم بردقوش ومزيج من 5 جم مريمية + 5 جم بردقوش) مستخلصات مائية مرتين في اليوم. تم تشخيص المرضى على أساس التاريخ الطبي المفصل والفحص الوراثي والتحليل البيوكيميائي الأخرى ذات الصلة. أوضح النتائج أن مستخلص البردقوش المائي وخلاصة المريمية غنية بمكونات مضادات الأكسدة مثل الأحماض الفينولية، الفلافونويد، أحادي التربينات المؤكسدة، الديتيربينويدات والتراتوبيرين. أوضح النتائج أن معدل الترشيح الكلي قد زاد بشكل ملحوظ، وانخفاض سرعة ترسب كرات الدم الحمراء والبروتينات المتفاقلي سبي في جميع المجموعات الفعلية مقارنة بال مجموعة الضابطة. انخفض المولونداي الدهوادس السوبر أكسيدوزونيتز كما سجل مزيج المستخلصات المائية من البردقوش (5 جم) والمريمية (5 جم) أفضل نتائج. لذلك أوصت هذه الدراسة بأن استخدام المستخلصات المائية من البردقوش (5 جم) والمريمية (5 جم) مفيدة جداً في تقليل الإجهاد التأكسدي وتحسين صحة الكلى لدى مرضى الغسيل الكلوي. هناك حاجة إلى مزيد من الدراسات لتحديد تأثير مضادات الأكسدة للأعشاب والأطعمة على أعراض مرض الكلى المزمنة.