Review Article

Volatile uptake, transport, perception, and signaling shape a plant’s nose

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Herbivore-induced plant volatiles regulate defenses in undamaged neighboring plants. Understanding the mechanisms by which plant volatiles are taken up, perceived, and translated into canonical defense signaling pathways is an important frontier of knowledge. Volatiles can enter plants through stomata and the cuticle. They are likely perceived by membrane-associated receptors as well as intracellular receptors. The latter likely involves metabolization and transport across cell membranes by volatile transporters. Translation of volatiles into defense priming and induction typically involves mitogen-activated protein kinases (MAPKs), WRKY transcription factors, and jasmonates. We propose that the broad range of molecular processes involved in volatile signaling will likely result in substantial spatiotemporal and ontogenetic variation in plant responsiveness to volatiles, with important consequences for plant–environment interactions.

Introduction

Plants produce and emit volatile organic compounds to mediate interactions with other organisms [1]. Upon insect herbivory, plants emit a complex blend of herbivore-induced plant volatiles [2,3]. These plant volatiles typically include green leaf volatiles (GLVs), terpenoids, and phenylpropanoid/benzenoid volatiles. Based on the plant species and the herbivores that trigger the emission, the volatile blends differ in composition, quantity, and timing. Herbivore-induced plant volatiles play critical roles shaping the interactions between plant–insect herbivores, directly or indirectly. They function directly by triggering or mediating defense response in plants, or by acting as toxins or repellents against herbivores. They also function indirectly by attracting the natural enemies of insect herbivores [2–4].

A particularly interesting function of herbivore-induced plant volatiles is their ability to mediate defense in the systemic undamaged tissues or neighboring undamaged plants [5–7]. Many volatiles have been shown capable of mediating interactions between plants (see a recent review for a comprehensive summary) [8]. Other than the phytohormone precursors MeJA and MeSA and the volatile hormone ethylene, GLVs are likely the most conserved volatile signals mediating plant–plant interactions. They trigger an array of defense responses in many plants, such as Arabidopsis, tomato, lima bean, and maize [9]. GLVs treatment in maize triggers the expression of defense-related genes and biosynthesis of defense-related metabolites, including many plant volatiles [10,11]. The emission of these volatiles further contributes to indirect defense [2]. In sweet potato, the homoterpene (E)-4,8-dimethyl-1,3,7-nonatriene (DMNT) can induce expression of proteinase inhibitor genes and increase herbivore resistance [12]. Volatiles such as indole and linalool typically do not induce defense directly, but they can prime several plants for stronger defense upon insect herbivory [11,13,14].

Recent years have seen substantial progress in understanding the biosynthesis, emission, and ecological function of plant volatiles. The perception of plant volatiles is now also being unraveled [1–3,15]. To fully understand how volatiles mediate plant–plant interactions, it is important to address how volatiles enter plant tissues and get recognized thereafter. A few recent reviews have summarized the latest discoveries in volatile biosynthesis, emission, and bioactivity [1,8,16]. Here, we discuss the possible paths.
Figure 1. Schematic view of possible volatile entry routes into leaf tissues
Herbivore-induced volatiles emitted from damaged plants may enter the plant tissue of undamaged neighbors through stomata and/or diffuse through the cuticle. (a) During daytime, stomata may be the main entry sites due to low resistance. (b) At night, when the stomata are closed, diffusion through the cuticle may become more important.

that volatiles may take to enter leaf tissues. We review the current evidence regarding the role of receptors/receptor complexes in volatile perception, with a focus on herbivore-induced plant volatiles. We further summarize the downstream signaling events that are triggered by herbivore-induced plant volatiles, and explore the hypothesis that spatiotemporal variation in volatile uptake, transport, perception, and signaling shape a plant’s ‘nose’, i.e. the tissues that are involved in perceiving volatiles as environmental cues. Pathogen-induced volatiles are covered in a separate paper in the same issue by Vlot and colleagues [17].

Entry of volatiles into plant leaves
To trigger defense response in the cells, volatiles need to access the plasma membrane or intracellular compartments. Volatiles may enter the leaves either through stomata or pass through the cuticular wax layer (Figure 1).

Entry through stomata
Stomata are the breathing pores of plant leaves, balancing photosynthetic carbon dioxide uptake and evaporative water loss [18]. These natural openings provide entry low resistance points for environmental agents such as microbes [19]. Recent studies show that stomata also shape plant–insect interactions by controlling volatile emission [20–22]. In maize, stomatal closure, induced by darkness or abscisic acid (ABA) treatment, constrains the emission of elicitor-triggered sesquiterpenes. Maize plants induced by elicitors in the dark show a burst of sesquiterpenes emission when light is switched on, indicating stomata as the gate for these volatiles to be released into the atmosphere [20]. Similarly, tomato and soybean leaves emit less volatiles when the stomata are partially closed by glucose oxidase, a salivary protein from the caterpillar Helicoverpa zea [21].

Given the considerations above, it is reasonable to assume that stomata also serve as low-resistance entry points for volatiles. Volatiles from attacked plants may thus enter the leaves of neighboring plants through these openings. If stomata are the main entry path for plant volatiles, several aspects need to be considered. First, as plants close their stomata at night [23], this would imply that plants are not able to perceive volatiles during nighttime, despite the fact that herbivore-induced plant volatiles such as GLVs are released as danger cues at night [24]. Another important aspect to consider is the developmental stage of stomata in leaves. In grasses, for instance, the stomatal complex is under differentiation and formation in the developmental zone in the young leaves [18]. Thus, if stomata are important for
volatile uptake, the ability of young leaves to respond to volatiles will differ from that of the old leaves, where stomata are fully developed. Experiments that investigate diurnal and developmental variation in volatile perception could thus provide first indications on the potential role of stomata in volatile perception.

**Entry through the cuticle**
The plant cuticle is the final barrier for volatiles to be released into atmosphere from nonvegetative organs [16]. In petunia flowers, the cuticle acts both as a resistance barrier and a sink for VOCs. Its thickness thus affects the dynamics of volatile emission [25]. This effect also depends on the physiochemical properties of VOCs, with volatiles having lower ambient vapor pressure facing higher resistance [16,26]. Several plants use their leaves to adsorb/take up volatiles from neighboring plants for enhanced herbivore resistance [27]. A recent study shows that plant leaf cuticular waxes can sequester exogenous volatiles [28]. Thus, it can be hypothesized that plant volatiles may pass the leaf cuticle and diffuse across the more permeable cell wall to reach the plasma membrane, effectively bypassing stomata.

Plant cuticular waxes comprise mainly very long-chain fatty acids and their derivatives. Both the wax composition and structure change greatly during leaf development. In wheat, leaf surface waxes keeps accumulating until the leaf blade finishes expanding. Meanwhile, the carbon chain length of the wax constituents increases drastically, and the wax crystals form different structure [29]. These changes likely cause different volatile permeability in young leaves and old leaves. Different physiochemical properties of volatiles will further increase the variance. So far, responses in volatile perception are either analyzed on the whole plant level or on a specific leaf. Experiments with cuticle mutants will help to uncover the role of cuticle as entry sites for volatiles relative to stomata.

**Volatile perception at the cell membrane**
Once accessing the plasma membrane, VOCs may be perceived by receptor/receptor complexes to trigger cellular response or taken up for further metabolic processing. Finding plant volatile receptors has been a long-standing question, but a breakthrough has yet to be achieved [3,15]. Ample progress has been made on identifying receptors for microbe-associated molecular patterns (MAMPs), herbivore-associated molecular patterns (HAMPs), and damage-associated patterns (DAMPs) [30,31]. These discoveries may inspire the discovery of a plant volatile receptor.

GLVs, C6 aldehydes, alcohols, and esters are enzymatically generated from membrane lipids upon disruption of membrane integrity in leaves, upon mechanical wounding or insect feeding [9]. Based on the plant origin and their ability to induce defense, these fatty acid-derived molecules can also be termed as DAMPs [32]. In *Arabidopsis*, the lectin receptor kinase LORF recognizes bacterial medium-chain 3-hydroxy fatty acid as a MAMP to trigger immunity [33]. Another *Arabidopsis* lectin receptor kinase LeckR-K.8 is critical for the defense triggered by phosphatidylincholines primarily with C16- to C18-fatty acyl chains [34]. The maize ZmFACS protein, a leucine-rich repeat (LRR) receptor kinase, mediates defense triggered by the fatty acid–amino acid conjugates (FACs) [35]. GLVs, as fatty acid derivatives, share certain biochemical properties with the molecular patterns mentioned above. Thus, they may be perceived by plasma membrane-localized receptor kinases or receptor proteins as well. Typically, these receptors need to form a protein complex with coreceptors to achieve full immune responses. These coreceptors are often from the somatic embryogenesis receptor kinase (SERKs) family [31]. Thus, screening lectin/LRR receptor kinase/receptor protein mutants and SERKs mutants for abolished or reduced GLVs response may help identifying components of the hypothesized GLVs receptor complex.

**Volatile uptake into cells**
MAMPs and DAMPs receptors are plasma membrane-localized receptors [31]. Perception of danger-related molecules can also happen inside of plant cells, such as the perception of effectors by nucleotide-binding domain leucine-rich repeat-containing (NLR) proteins [36]. It is possible that perception of some VOCs happens intracellularly. Additionally, plants take up volatiles directly for further metabolic processing [27]. In either case, these volatiles need to pass through the plasma membrane. Direct diffusion may happen for membrane lipid derived volatiles such as GLVs. This is unlikely for most other volatiles [16]. In petunia, transport of VOCs across the plasma membrane relies on an adenosine triphosphate-binding cassette (ABC) transporter [37]. Similar volatile transporters may mediate the channeling of volatiles from the extracellular into the intracellular space. Transporters or ion channels may act as receptors as well. For example, the *Arabidopsis* anion channel SLAC1 plays an important role in sensing CO₂/bicarbonate in the guard cells [38]. Within the cells, volatiles may bind specific proteins to initiate cellular
Figure 2. Schematic representation of plant volatile perception and signaling

Plant volatiles are likely perceived in three different ways. The respective responsible proteins are: (1) cell surface receptors, (2) intracellular proteins, and (3) volatile transporters or ion channels. Volatile perception leads to a cascade of defense responses, including calcium influx, MAPK activation, and WRKY transcription factor-regulated expression of defense genes. Dashed arrows indicate unclear molecular mechanisms of the signaling cascade.

responses. Recently, the transcription regulators TOPLESS-like proteins (TPLs) were found to bind the sesquiterpene caryophyllene [39]. However, it is unclear whether TPLs alone are sufficient to act as receptors to transduce caryophyllene-triggered responses. So far, hypothetical plant odorant-binding proteins (OBPs) have also been proposed to act as plant volatile receptors, based on their similarity with animal OBPs. A recent study used in silico molecular docking to prove plant OBPs can bind monoterpens [40]. The specificity and in vivo activity of OBPs remains to be uncovered.

Translation into defense signaling

Volatiles are well established to trigger defense pathways that are typically associated with MAMPs and DAMPs [8,30] (Figure 2).

Upon exposure to GLVs, Arabidopsis and tomato plants show a cytosolic calcium influx. In Arabidopsis, this transient influx happens rapidly upon GLVs exposure and peaks at 5–10 min [41]. The calcium influx kinetic is unclear in tomato likely due to the lack of a calcium reporter line [42]. Additionally, ocimene, myrcene, pinene, and DMNT can also trigger transient cytosolic calcium influx in Arabidopsis, similar as the one triggered by GLVs [41]. Calcium influx is a typical early response, following the perception of MAMPs and DAMPs. NLR activation after effector recognition also leads to calcium influx, albeit with different dynamics [36]. Collectively, these studies indicate calcium influx is a conserved immune response upon danger perception, including volatiles.

Activation of mitogen-activated protein kinases (MAPKs) is another classical early immune response upon pattern recognition [43]. Similarly, GLV exposure leads to rapid activation of MAPKs in the grass Lolium temulentum [44]. The sesquiterpene (E)-Nerolidol increases both the transcript and protein of MAPK in tea plants [45]. Indole exposure does not activate MAPKs directly but increases MAPK gene expression and activation following simulated herbivory in rice. Knocking down MPK3 and MPK6 leads to abolished or greatly reduced defense priming effect by indole [46]. MAPK cascades are also points of convergence between different signaling pathways [47]. This role may explain the
phenomenon that dual exposure of the GLV (Z)-3-hexenyl-acetate (HAC) and indole in maize generates stronger defense than HAC exposure alone [11].

Other commonly reported volatile responses include the increased expression of genes-encoding transcription factors, defense hormone biosynthesis enzymes, and defense metabolite biosynthesis enzymes [8]. Members of the WRKY transcription factor family are often induced by various biotic and abiotic stress and regulate hormone biosynthesis in turn [48,49]. In Arabidopsis, the GLV (E)-2-hexenal induces the expression of several WRKY genes, including AtWRKY6, AtWRKY40, and AtWRKY53. Knocking out these genes leads to increased expression of (E)-2-hexenal-specific responsive genes, indicating that these transcription factors as negative regulators of GLVs signaling in Arabidopsis [48]. The GLV (Z)-3-hexenol increases ZmWRKY12 transcripts in maize, but the importance of this phenomenon is unclear [50]. Similarly, CsWRKY3 expression is up-regulated by (E)-Nerolidol but its role in (E)-Nerolidol signaling is unknown [45].

Plant VOCs may also suppress defense in some cases [14,51]. Whitefly-infested tomato plants emit a unique blend of volatiles to suppress JA-dependent defense but prime SA-dependent defense. The dual role in defense is triggered by two terpenes: β-myrcene or β-caryophyllene. On the contrary, linalool, a monoterpene elicited by the Spodoptera exigua caterpillar attacked tomato plants, primes the expression of two JA-pathway proteinase inhibitor genes, PI-I and PI-II [14]. The molecular mechanisms behind these differences are unknown. Future work on how volatile-triggered defense is differentially regulated will greatly help to uncover novel volatile signaling components and pathways.

**Spatiotemporal patterns of volatile perception**

Since the discovery of 'talking trees', plant volatile-mediated plant–plant interactions have fascinated many scientists and let to substantial research efforts [5]. We now have a detailed understanding of plant volatile biosynthesis and transport [1,16]. Based on this knowledge, we can infer that plant volatile perception likely involves a number of physical structures such as stomata and cuticles as well as molecular elements such as transporters, receptors, and signal integration proteins. Given the substantial variation in the expression of these elements in different plant parts and developmental stages, we predict that plant volatile perception will not be uniform, but will show significant variation within a given plant. Certain leaves are likely to be much more sensitive to others, and could thus be viewed as a plant 'nose'. Understanding these patterns and linking them to our increased understanding of the mechanisms of plant volatile perception will be important to unravel the ecological dynamics that are elicited by volatiles and to exploit plant volatiles as crop-reprogramming signals. As comprehensively summarized in a recent review, the potential applications of plant volatiles for sustainable agricultural practices will include breeding crops with enhanced volatile emissions and inducing volatile release in a targeted manner [2]. A good understanding of the mechanisms and spatiotemporal variation in volatile perception will also facilitate the selection and breeding of plants that are sensitive to plant volatiles in the right place at the right time. Such work will help to unlock the potential of plant volatiles as crop-reprogramming agents.

**Summary**

- Plant volatiles may enter the inner space of plant leaves through stomata or cuticle. The relative importance of these entry sites is likely to vary with development and environmental conditions.
- Plant volatile perception is likely mediated by cell surface receptors, plasma membrane-localized transporters or ion channels, and intracellular proteins.
- Plant volatiles regulate canonical defense signaling pathways, with MAPKs and WRKY transcription factors playing important roles as signal integration hubs.
- Spatiotemporal variation in volatile uptake and perception elements will likely determine where and when plants respond to volatiles.

**Competing Interests**

The authors declare that there are no competing interests associated with the manuscript.
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Author Contributions
L.W. conceived the manuscript, wrote the first draft and edited the final version. M.E. conceived the manuscript, helped write the first draft and edited the final version.

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Abbreviations
ABA, abscisic acid; ABC, adenosine triphosphate-binding cassette; DAMP, damage-associated molecular pattern; DMNT, (E)-4,8-dimethyl-1,3,7-nonatriene; FAC, fatty acid–amino acid conjugate; GLV, green leaf volatile; HAC, (Z)-3-hexenyl-acetate; HAMP, herbivore-associated molecular pattern; JA, jasmonic acid; LRR, leucine-rich repeat; MAMP, microbe-associated molecular pattern; MAPK, mitogen-activated protein kinase; NLR, nucleotide-binding, leucine-rich-repeat; OBP, odorant-binding protein; SA, salicylic acid; SERK, somatic embryogenesis receptor kinase; TPL, TOPLESS-like proteins; VOC, volatile organic compound.

References
1. Dudareva, N., Klempien, A., Muhlemann, J.K. and Kaplan, I. (2013) Biosynthesis, function and metabolic engineering of plant volatile organic compounds. New Phytol. 196, 16–32, https://doi.org/10.1111/nph.12145
2. Turlings, T.C.J. and Erb, M. (2018) Tritrophic interactions mediated by herbivore-induced plant volatiles: mechanisms, ecological relevance, and application potential. Annu. Rev. Entomol. 63, 433–452, https://doi.org/10.1146/annurev-ento-020117-043507
3. Heil, M. (2014) Herbivore-induced plant volatiles: targets, perception and unanswered questions. New Phytol. 204, 297–306, https://doi.org/10.1111/nph.12977
4. Ton, J., D’Alessandro, M., Jourdie, V., Jakab, G., Karlen, D., Held, M. et al. (2007) Priming by airborne signals boosts direct and indirect resistance in maize. Plant J. 49, 16–26, https://doi.org/10.1111/j.1365-313X.2006.02935.x
5. Baldwin, I.T., Hallatsche, R., Paschold, A., von Dahl, C.C. and Preston, C.A. (2006) Volatile signaling in plant-plant interactions: “talking trees” in the genomics era. Science 311, 812–815, https://doi.org/10.1126/science.1118446
6. Moreira, X. and Abdala-Roberts, L. (2019) Specificity and context-dependency of plant-plant communication in response to insect herbivory. Curr. Opin. Insect. Sci. 32, 15–21, https://doi.org/10.1016/j.cois.2018.09.003
7. Karban, R. (2021) Plant communication. Annu. Rev. Ecol. Evol. Syst. 52, 1–24, https://doi.org/10.1146/annurev-ecolsys-010421-020045
8. Brosset, A. and Blande, J.D. (2022) Volatile-mediated plant-plant interactions: volatile organic compounds as modulators of receiver plant defense, growth, and reproduction. J. Exp. Bot. 73, 511–528, https://doi.org/10.1093/jxb/erab487
9. Amede, M., Allmann, S., Verwaeren, J., Smagghe, G., Raesada, G., Schuurink, R.C. et al. (2018) Green leaf volatile production by plants: a meta-analysis. New Phytol. 220, 666–683, https://doi.org/10.1111/nph.14671
10. Engelberth, J., Albom, H.T., Schmelz, E.A. and Tumlinson, J.H. (2004) Airborne signals prime plants against insect herbivore attack. Proc. Natl. Acad. Sci. U.S.A. 101, 1781–1785, https://doi.org/10.1073/pnas.0308037100
11. Hu, L., Ye, M. and Erb, M. (2019) Integration of two herbivore-induced plant volatiles results in synergistic effects on plant defence and resistance. Plant Cell Environ. 42, 959–971, https://doi.org/10.1111/pce.13443
12. Meents, A.K., Chen, S.-P., Reichelt, M., Lu, H.-H., Bartram, S., Yeh, K.-W. et al. (2019) Volatile DMNT systemically induces jasmonate-independent direct anti-herbivore defense in leaves of sweet potato (Ipomoea batatas) plants. Sci. Rep. 9, 17431, https://doi.org/10.1038/s41598-019-53946-0
13. Erb, M., Veyrat, N., Robert, C.A.M., Xu, H., Frey, M., Ton, J. et al. (2015) Indole is an essential herbivore-induced volatile priming signal in maize. Nat. Commun. 6, 6273, https://doi.org/10.1038/ncomms7273
14. Zhang, P.-J., Wei, J.-N., Zhao, C., Zhang, Y.-F., Li, C.-Y., Liu, S.-S. et al. (2019) Airborne host-plant manipulation by whitflies via an inducible blend of plant volatiles. Proc. Natl. Acad. Sci. U.S.A. 116, 7387–7396, https://doi.org/10.1073/pnas.1818599116
15. Loreto, F. and D’Auria, S. (2022) How do plants sense volatiles sent by other plants? Trends Plant Sci. 27, 29–38, https://doi.org/10.1016/j.tplants.2021.08.009
16. Widhalm, J.R., Jaini, R., Morgan, J.A. and Dudareva, N. (2015) Rethinking how volatiles are released from plant cells. Trends Plant Sci. 20, 545–550, https://doi.org/10.1016/j.tplants.2015.06.009
17. Eccleston, L., Brambilla, A. and Vlot, A.C. (2022) New molecules in plant defence against pathogens. Essays Biochem., https://doi.org/10.1042/ERC20210076
18. Nunes, T.D.G., Zhang, D. and Raisig, M.T. (2020) Form, development and function of grass stomata. Plant J. 101, 780–799, https://doi.org/10.1111/tpj.14552
19 Melotto, M., Zhang, L., Oblessuc, P.R. and He, S.Y. (2017) Stomatal defense a decade later. Plant Physiol. 174, 561–571, https://doi.org/10.1104/pp.16.01853

20 Seldí-Adams, I., Richter, A., Boomér, K.B., Yoshinaga, N., Degenhardt, J. and Tumlinson, J.H. (2015) Emission of herbivore elicitor-induced sesquiterpenes is regulated by stomatal aperture in maize (Zea mays) seedlings. Plant Cell Environ. 38, 23–34, https://doi.org/10.1111/pce.12347

21 Lin, P.-A., Chen, Y., Chaverra-Rodriguez, D., Heu, C.C., Zainuddin, N.B., Sidhu, J.S. et al. (2021) Silencing the alarm: an insect salivary enzyme closes plant stomata and inhibits volatile release. New Phytol. 230, 793–803, https://doi.org/10.1111/nph.17214

22 Lin, P.-A., Chen, Y., Ponce, G., Acevedo, F.E., Lynch, J.P., Anderson, C.T. et al. (2021) Stomata-mediated interactions between plants, herbivores, and the environment. Trends Plant Sci.

23 Caird, M.A., Richards, J.H. and Donovan, L.A. (2007) Nighttime stomatal transpiration and C3 and C4 plants. Plant Physiol. 143, 4–10, https://doi.org/10.1090/1082-3523-2007-092940

24 de Moraes, C.M., Mescher, M.C. and Tumlinson, J.H. (2001) Caterpillar-induced nocturnal plant volatiles repel conspecific females. Nature 410, 577–580, https://doi.org/10.1038/35069058

25 Liao, P., Ray, S., Beachon, B., Lynch, J.H., Deshpande, A., McAdam, S. et al. (2021) Cuticle thickness affects dynamics of volatile emission from petunia flowers. Nat. Chem. Biol. 17, 138–145, https://doi.org/10.1038/s41589-020-00670-w

26 Matsu, K. (2016) A portion of plant airborne communication is endorsed by uptake and metabolism of volatile organic compounds.Curr. Opin. Plant Biol. 32, 24–30, https://doi.org/10.1016/j.pbi.2016.05.005

27 Sugimoto, K., Matsu, K. and Takabayashi, J. (2016) Uptake and conversion of volatile compounds in plant–plant communication. In Deciphering Chemical Language of Plant Communication (Blande, J.D. and Glenwood, R., eds), pp. 305–316, Springer International Publishing, Cham, https://doi.org/10.1007/978-3-319-33498-1_13

28 Camacho-Coronel, X., Molina-Torres, J. and Heil, M. (2020) Sequestration of exogenous volatiles by plant cuticular waxes as a mechanism of passive associative resistance: a proof of concept. Front. Plant Sci. 11, 121, https://doi.org/10.3389/fpls.2020.00121

29 Wang, Y., Wang, J., Chai, G., Li, C., Hu, Y., Chen, X. et al. (2015) Developmental changes in composition and morphology of cuticular waxes on leaves and spikes of glossy and glaucous wheat (Triticum aestivum L.). Plant Bio 10, e0141239, https://doi.org/10.1371/journal.pone.0141239

30 Erb, M. and Reymond, P. (2019) Molecular interactions between plants and insect herbivores. Annu. Rev. Plant Biol. 70, 527–557, https://doi.org/10.1146/annurev-arplant-050718-095910

31 Albert, I., Hua, C., Nürnberg, T., Pruitt, R.N. and Zhang, L. (2020) Surface sensor systems in plant immunity. Plant Physiol. 182, 1582–1596, https://doi.org/10.1104/pp.19.01299

32 Meents, A.K. and Mithöfer, A. (2020) Plant-plant communication: is there a role for volatile damage-associated molecular patterns? Front. Plant Sci. 11, 583275, https://doi.org/10.3389/fpls.2020.583275

33 Kutschera, A., Dawid, C., Gisch, N., Schmid, C., Raasch, L., Gerster, T. et al. (2019) Bacterial medium-chain 3-hydroxy fatty acid metabolites trigger immunity in Arabidopsis plants. Science 364, 178–181, https://doi.org/10.1126/science.aau1279

34 Stahl, E., Brillat, T., Ferreira Queiroz, E., Marcourt, L., Schmiesing, A., Hilékier, O. et al. (2020) Phosphatidylcholines from Pieris brassicae eggs activate an immune response in Arabidopsis. Elife 9, e60293

35 Loreto, F. and Facchiano, A. (2021) A hypothesis on the capacity of plant odorant-binding proteins to bind volatile isoprenoids based on in silico evidences. Elife 10, e66741

36 Yuan, M., Nguo, B.P.M., Ding, P. and Xin, X.-F. (2021) PTI-ETI crosstalk: an integrative view of plant immunity. Curr. Opin. Plant Biol. 62, 102030

37 Adesefin, F., Widdrham, J.R., Beachon, B., Lettreve, F., Pieman, B., Lynch, J.H. et al. (2017) Emission of volatile organic compounds from petunia flowers is facilitated by an ABC transporter. Science 356, 1386–1388

38 Zhang, J., Wang, N., Miao, Y., Hauser, F., McCammon, J.A., Rappel, W.-J. et al. (2018) Identification of SLAC1 anion channel residues required for CO2/bicarbonate sensing and regulation of stomatal movements. Proc. Natl. Acad. Sci. U.S.A. 115, 11129–11137

39 Nagashima, A., Higaki, T., Koeduka, T., Ishigami, K., Hosokawa, S., Watanabe, H. et al. (2019) Transcriptional regulators involved in responses to volatile organic compounds in plants. J. Biol. Chem. 294, 2256–2266

40 Giordano, D., Facchiano, A., D’Auria, S. and Loreto, F. (2017) A hypothesis on the capacity of plant odorant-binding proteins to bind volatile isoprenoids based on in silico evidences. Elife 10, e66741

41 Asai, N., Nishioha, T., Takabayashi, J. and Fujuchi, T. (2009) Plant volatiles regulate the activities of Ca2+-permeable channels and promote cytoplasmic calcium transients in Arabidopsis leaf cells. Plant Signal. Behav. 4, 294–300, https://doi.org/10.4161/psb.4.4.8275

42 Zebelo, S.A., Matsu, K., Ozawa, R. and Maffei, M.E. (2012) Plasma membrane potential depolarization and cytosolic calcium flux are early events involved in tomato (Solanum lycopersicum) plant-to-plant communication. Plant Sci. 196, 93–100, https://doi.org/10.1016/j.plantsci.2012.08.006

43 Boller, T. and Felix, G. (2009) A renaissance of elicitors: perception of microbe-associated molecular patterns and danger signals by pattern-recognition receptors. Annu. Rev. Plant Biol. 60, 379–406, https://doi.org/10.1146/annurev.plant.57.032905.105346

44 Dombrowski, J.E. and Martin, R.C. (2018) Activation of MAP kinases by green leaf volatiles in grasses. BMC Res. Notes 11, 79, https://doi.org/10.1186/s13104-017-3076-9

45 Chen, S., Zhang, L., Cai, X., Li, X., Buan, L., Luo, Z. et al. (2020) (E)-Nerolidol is a volatile signal that induces defenses against insects and pathogens in tea plants. Hortic Res. 7, 52, https://doi.org/10.1016/j.sreb.4.4.8275

46 Ye, M., Glauser, G., Lou, Y., Erb, M. and Hu, L. (2019) Molecular dissection of early defense signaling underlying volatile-mediated defense regulation and herbivore resistance in rice. Plant Cell. 31, 687–698, https://doi.org/10.1105/tpc.18.00569

47 Zhang, M., Su, J., Zhang, Y., Xu, J. and Zhang, S. (2018) Conveying endogenous and exogenous signals: MAPK cascades in plant growth and defense. Curr. Opin. Plant Biol. 45, 1–10, https://doi.org/10.1016/j.pbi.2018.04.012
Mirabella, R., Rauwerda, H., Allmann, S., Scala, A., Spyropoulou, E.A., de Vries, M. et al. (2015) WRKY40 and WRKY6 act downstream of the green leaf volatile E-2-hexenal in Arabidopsis. Plant J. 83, 1082–1096, https://doi.org/10.1111/tpj.12953.

Rushton, P.J., Somssich, I.E., Ringler, P. and Shen, Q.J. (2010) WRKY transcription factors. Trends Plant Sci. 15, 247–258, https://doi.org/10.1016/j.tplants.2010.02.006.

Engelberth, J., Contreras, C.F., Dalvi, C., Li, T. and Engelberth, M. (2013) Early transcriptome analyses of Z-3-Hexenol-treated zea mays revealed distinct transcriptional networks and anti-herbivore defense potential of green leaf volatiles. PLoS ONE 8, e77465, https://doi.org/10.1371/journal.pone.0077465.

Erb, M. (2018) Volatiles as inducers and suppressors of plant defense and immunity-origins, specificity, perception and signaling. Curr. Opin. Plant Biol. 44, 117–121, https://doi.org/10.1016/j.pbi.2018.03.008.