Anticoccidial effects of *Tinospora rumphii* extract in naturally infected goats

Pearl Muyco Saladino¹, Elsa Amarille Gonzaga², Loveille Jun Amarille Gonzaga³

¹College of Veterinary Medicine, University of Southern Mindanao, Kabacan, Cotabato, Philippines  
²Department of Veterinary Paraclinical Sciences, College of Veterinary Medicine, University of Southern Mindanao, Kabacan, Cotabato, Philippines  
³Department of Chemistry, College of Science and Mathematics, University of Southern Mindanao, Kabacan, Cotabato, Philippines

**ABSTRACT**

**Objective:** This study aimed to determine the anticoccidial potential of *Tinospora rumphii* ethanolic stem extract, resulting in an effective reduction in oocysts per gram counts of *Eimeria* species in goats in comparison to the commercial coccidiostat toltrazuril.

**Materials and Methods:** Eighteen goats that were naturally infected with coccidia were identified as experimental animals. The experimental animals were grouped and were administered a single dose of *T. rumphii* extract at concentrations of 300, 600, 1,000, and 2,000 mg extract/kg body weight. The fecal samples were collected before treatment, during the first treatment, and every week thereafter for 28 days posttreatment. Fecal examination was carried out using a modified McMaster technique to determine the oocysts per gram of feces, and the mean efficacies of the extracts were calculated.

**Results:** Stem extracts of *T. rumphii* were able to lower the oocysts per gram count than pretreatment for all concentrations, but the efficacy, in general, was still lower than toltrazuril. A significant difference existed in the efficacy of the extracts among the treatments on day 14. Administering 1,000 mg/kg extract resulted in the highest efficacy rate (95.25%) at 14 days post-treatment and was comparable to that of toltrazuril (89.89%).

**Conclusion:** The results show that the *T. rumphii* stem extract has the potential to reduce the *Eimeria* species oocysts per gram counts of naturally infected goats.

**Introduction**

Backyard goat production in the Philippines accounts for an estimate of 3.60 million heads of goats in the inventory [1]. Goat raising is popular among rural folks since it requires simple management and low-cost production inputs compared to other livestock. Goats are vital to the economy, being kept mainly for trade, meat, and milk production. However, one of the biggest constraints in the small ruminant industry is internal parasitism. One of the principal problems in goat farming is coccidiosis, a protozoal disease caused by the *Eimeria* species. Coccidiosis among young goats is commonly manifested by diarrhea, dehydration, emaciation, weakness, loss of appetite, poor weight gain, poor feed conversion, and death, causing a detrimental effect on the animal’s performance and priming for great economic loss. Acute infection is chiefly confined to kids 2–4 weeks postweaning up to 6 months of age, due to the stress of weaning, overcrowding, and unhygienic conditions, thereby exposing the goats to a large proportion of oocysts [2].

The administration of over-the-counter anticoccidial products is the common procedure of treatment. However, its high cost has limited its effective control of parasites. Moreover, a prolonged period of synthetic drug usage evidently results in it losing its strength against resistant coccidia in goats [3]. With the advent of drug resistance, alternative veterinary interventions are continuously being carried out using natural compounds to help reduce the problem.

A potential plant that can be used is *Tinospora rumphii*, which is synonymous with *Tinospora crispa* [4]. It is a climbing shrub native to lower elevations in tropical areas. The stem of the plant is green and succulent and covered...
with a thin brown bark and surrounded by warty lenticels. A diverse group of pharmacologically active substances have been isolated from *T. rumphii* [5]. These include amino acids, glycosides, flavonoids, phenols, saponins, steroids, and terpenoids, eliciting varied bioactivities. The phytochemical profile and bioactivities of *T. rumphii* make it a promising source of a natural coccidiostat. Thus, this study aimed to determine the anticoccidial activity of *T. rumphii* stem extracts against goats naturally infected with *Eimeria* species.

**Materials and Methods**

**Ethical approval**

The experimental procedures were approved by the University of Southern Mindanao College of Veterinary Medicine Institutional Animal Care and Use Committee MC-CVM-001552.

**Plant material and extraction**

Healthy and mature stems of *T. rumphii* were selected and collected early in the morning and were validated at the University of Southern Mindanao Department of Biological Sciences. Extraction of the plant material was carried out based on the method previously described [6] with modifications and a different extracting solvent. The stems were washed thrice using tap water and then with distilled water. Stems were air-dried for 7 days prior to grinding using an electronic blender. During extraction, the powdered stems were soaked in ethanol by adding 300 ml of ethanol for every 50 gm of powder. The mixture was soaked for 48 h and filtered using Whatman filter paper. The filtrate was concentrated *in vacuo* at 40°C at 60–100 rpm to obtain the concentrated ethanolic extract and weighed to determine the concentration. The extract was stored in a clean and sterile container.

**Animals**

The procedures used in the study were in accordance with the University of Southern Mindanao College of Veterinary Medicine Institutional Animal Care and Use Committee MC-CVM-001552. The methods for research and animal handling were all in compliance with Republic Act 8485, The Animal Welfare Act.

A preliminary fecal examination using the modified McMaster method [7] was carried out to determine the number of oocysts per gram (OPG) of feces and to identify which goats were infected with the *Eimeria* strain that will be used for the study. Identification of the *Eimeria* species was based on the morphometric characteristics of the oocysts and, in some cases, based on the presence or absence of a micropyle and oocyst shape [8].

Goats with an OPG of at least 875 were considered experimental animals. A total of 18 naturally coccidia-infected goats aged 3–5 months, regardless of sex and breed, but under the same management, served as the experimental animals. The management of the goats on the farm was a confined type of rearing.

The fecal samples were collected before treatment, during the first treatment, and every week thereafter for 28 days posttreatment (dpt).

**Administration of *T. rumphii* extract**

The goats were divided into six groups (A–F) of three goats each. The experimental goats were labeled according to their treatment. The goats were weighed using a digital weighing scale to measure the live weight, which determined the volume of extract to be administered. Goats in groups A–D were given a single dose of 300, 600, 1,000, 2,000 mg extract/kg bodyweight. Those in group E were treated with a single dose of 5% toltrazuril (0.4 ml/kg bodyweight), while those in group F were untreated and served as the negative control. The goats were fasted for about 12 h to slow down gastrointestinal transit and allow better absorption of the drug administered orally [9] before the *T. rumphii* stem extract was given orally early in the morning.

**Parasitological examination**

Fecal samples were collected early in the morning during feeding time at 7, 14, 21, and 28 dpt. Three to five grams of fresh fecal samples were obtained from the rectum of goats. Fecal samples were scored using a correction factor based on a scale of 1–3 with 1, normal pellets; 1.5, soft formed but the pellets did not separate; 2, soft but there was no formation of pellets; and 3, diarrheic [10]. Fecal examination was carried out using the modified McMaster technique [7] to determine the OPG of feces.

**Statistical analysis**

Results are expressed as mean ± standard errors of the mean. One-way analysis of variance was used to assess the significant differences (*p < 0.05*) in mean efficacy among the groups. Furthermore, when a significant difference was found, *post-hoc* multiple mean concentration using the Tukey honestly significant difference test was used to test the consistency of the effects for treatments and controls.

**Results and Discussion**

The mean OPG counts of goats in the different treatment groups are shown in Table 1. In general, there was a decrease in the OPG counts in all groups between 7 and 14 dpt except in group D. It was only at 21 dpt that group D exhibited a decrease in the OPG counts similar to all of the other groups. The OPG counts continued to decrease at 28 dpt, except for the negative control. Statistical analysis
revealed that there was no significant difference in the OPG counts in goats among treatments in all periods of observation.

The efficacy of *T. rumphii* stem extracts given to the goats is shown in Table 2. The results show that at 7 dpt, group C had the highest efficacy of 95.25% among the different concentrations of *T. rumphii* extract. Statistical analysis revealed that there was a significant difference (*p* < 0.05) in the efficacy of the different concentrations of *T. rumphii* extract used at 14 dpt. Moreover, it was observed that at 28 dpt, *T. rumphii* extracts had a more consistent increase in efficacy across concentrations. However, there was no significant difference observed among the six groups. Nevertheless, the highest efficacy the *T. rumphii* extract was able to elicit was 95.25% at a concentration of 1,000 mg/kg at 7 dpt, which was comparable to that of toltrazuril, which had an 89.89% efficacy at 7 dpt.

The efficacy of the *T. rumphii* extract in goats as observed in groups A, B, and C can be attributed to the anticoccidial potential of the phytochemical constituents to decrease coccidial oocyst counts in animals [11]. Saponins, tannins, and flavonoids are compounds known to have anticoccidial activity. These phytochemical compounds are present in *T. rumphii* stems [12].

Saponins are capable of membrane polarization, causing vacuolization and may enter the oocyst wall [13]. This action is similar to that of two conventional drugs such as praziquantel and toltrazuril, which produce severe vacuolization in all intracellular development stages, particularly in the protozoal endoplasmic reticulum, thereby interfering in the division of the protozoal nucleus, damaging the wall-forming bodies in the microgametes and also disturbing the activity of the mitochondria [14]. Saponins can directly bind to ruminal protozoa and lyse the outer membrane. However, since an oocyst wall is composed of two layers [15], saponins cannot destroy the oocyst wall but may enter the wall through the micropyle cap and damage the sporocyst [16].

Tannins may enter the inner oocyst wall and interfere with energy generation by uncoupling oxidative phosphorylation and causing a loss of intracellular components, resulting in the formation of an abnormally thickened, complete oocyst wall, and zygote necrosis [17]. The mode of action of tannins is quite similar to that of didazuril, since its usage in the oocysts of both *Eimeria (E.) brunetti* and *E. maxima* resulted in the formation of an abnormally thickened, incomplete oocyst wall, and zygote necrosis [18]. Tannins also bind to glycoproteins in the cuticle of helminths and cause their death [19]. Tannins can also provide resistance through partial protective passive immunity [20].

The antioxidant properties of flavonoids may also be attributed to the anticoccidial effect of plants [21]. It was also observed that coccidia can induce host cell destruction associated with oxidative stress and lipid peroxidation [22]. The antioxidants that have the ability to neutralize reactive oxygen species (ROS) are protective due to their ROS-scavenging ability [23], reducing the severity of *Eimeria* infection by ameliorating the degree of intestinal lipid peroxidation. Therefore, it can be emphasized that plant extracts containing flavonoids can inhibit the development of *Eimeria* before oocysts are released in host feces.

Variations in the efficacy of *T. rumphii* extract can also result when the coccidiostat only suspends the development of the parasite at the first intracellular stage when the drug is given from the time of infection [24]. Thus, in the later stages of *Eimeria* species, which are developing more deeply in the intestinal tissues, the parasite becomes unaffected by the treatment given. It was shown in lambs experimentally infected with *E. crandallis* that didazuril basically affects first-generation and, to a lesser extent, the late meront stages and gamonts [25]. If the meronts were destroyed by the anticoccidial treatment roughly 2 weeks after the goats had become infected, the likelihood

### Table 1. The mean fecal oocysts per gram of naturally infected goats treated with *T. rumphii* stem extract.

| Group     | 0          | 7          | 14         | 21         | 28         |
|-----------|------------|------------|------------|------------|------------|
| A (300 mg/kg) | 5,925 ± 3,868 | 725 ± 191 | 875 ± 50  | 2,842 ± 1,109 | 650 ± 293 |
| B (600 mg/kg) | 2,342 ± 1,421 | 2,042 ± 1,702 | 292 ± 36  | 317 ± 102  | 142 ± 36  |
| C (1,000 mg/kg) | 1,683 ± 872  | 77 ± 40    | 408 ± 166  | 283 ± 8    | 158 ± 22 |
| D (2,000 mg/kg) | 12,267 ± 11,367 | 12,550 ± 12,475 | 20,542 ± 19,267 | 10,683 ± 9,859 | 3,117 ± 2,955 |
| E (toltrazuril) | 2,850 ± 1,815 | 208 ± 102  | 1,317 ± 1,068 | 1,133 ± 727 | 283 ± 186 |
| F (untreated)   | 14,658 ± 13,741 | 14,142 ± 13,917 | 7,575 ± 7,401 | 317 ± 22    | 10,508 ± 10,000 |

*p*-values

|         | 0.616* | 0.487* | 0.523* | 0.052* | 0.433* |

*: There is no significant difference at the 5% level.
of attaining a greater efficacy against coccidiosis might have occurred [26].

The most common pathogenic coccidia species found in goats include *E. arloingi*, *E. ninakohlyokimovae*, *E. caprovinia*, *E. christenseni*, *E. faurei*, and *E. gilruthi*. Infection is most commonly observed in kids, 2–4 weeks after weaning, and can occur in goats raised under semi-intensive and intensive management practices. The infection arises naturally when grazing, particularly when sporulated oocytes are ingested. Poor goat management practices also contribute to the infection, particularly when feed and water supplies are contaminated with goat feces or when contaminated hair coats are licked [2].

One activity that induces drug resistance is the use of year-round coccidiostats in susceptible hosts. When the number of oocysts to which animals are exposed becomes very low, the animal may fall below the threshold of exposure needed to stimulate a protective response. Therefore, coccidiostats will only be used at times of risk like parturition, shipment, weaning, or inclement weather. Toltrazuril resistance has been experimentally reported in a field isolate of ovine *Eimeria* species through the use of fecal oocyst reduction count as a tool to evaluate anticoccidial efficacy. Similarly, the treatment of lambs with the recommended dose of 20 mg/kg of toltrazuril (Baycox®) did not result in a significant reduction in oocyst excretion in the treated animals compared with the controls. Decreased efficacy is possibly due to the haploid stages of *Eimeria* species, which are immediately selected for resistance as a result of extensive use of a drug over time [24].

It is hypothesized that the variations in the efficacy of *T. rumphii* extract can be due to the dependence of treatment on the susceptible stages of *Eimeria* species present in goats. The goats in the treatment group may, in general, not be in the same stages of infection [27]. Thus, goats with coccidia in later stages of development and the short time of exposure of *Eimeria* species in the surface epithelium during the administration of the treatment will have lower efficacy rates. Some goats metabolize drugs so rapidly that therapeutically effective concentrations are not reached. However, further studies must be carried out regarding the influence of individual drug metabolisms on the efficacy of the extract.

**Conclusion**

The result of the study shows that with the use of the stem extract of *T. rumphii*, the mean oocyst counts remained lower than the pre-treatment OPG counts except when using 2,000 mg of extract/kg bodyweight. The extract has potential in vivo anticoccidial activity, with a similar activity to that of the commercial coccidiostat toltrazuril at 14 dpt.

**List of Abbreviations**

dpt, days posttreatment; g, gram; HSD, honestly significant difference; kg, kilogram; mg, milligram; ml, milliliter; OPG, oocysts per gram of feces; ROS, reactive oxygen species; rpm, revolutions per minute; *T. rumphii*, *Tinospora rumphii*.

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**Conflict of interest**

The authors declare no conflicts of interest.

**Authors’ contribution**

PMS, EAG, and LJAG developed the research concepts and designed the study. PMS conducted the experiments under the supervision and guidance of EAG. LJAG wrote the manuscript draft, which was revised by the coauthors.
REFERENCES

[1] Philippine Statistics Authority. Goat situation report: January to March 2021. Philippine Statistics Authority, Quezon City, Philippines, 2021.

[2] Chartier C, Parraud C. Coccidiosis due to Eimeria in sheep and goats, a review. Small Rum Res 2012; 103(1):84–92; https://doi.org/10.1016/j.smallrumres.2011.10.022

[3] Odden A, Enemark HL, Ruiz A, Robertson LJ, Eradsal C, Nes SK, et al. Controlled efficacy trial confirming toltrazuril resistance in a field isolate of ovine Eimeria spp. Parasit Vectors 2018; 11(1):394; https://doi.org/10.1186/s13071-018-2976-4

[4] Forman LL. A revision of Tinospora (Menispermaceae) in Asia to Australia and the Pacific: the Menispermaceae of Malesia and Adjacent Areas. X: Kew Bull 1981; 36(2):375–421; https://doi.org/10.2307/4113613

[5] Choudhary M, Ismail M, Ali Z, Shaari K, Lajis NH, Atta-ur-Rahman. Alkaloidal constituents of Tinospora crispa. Nat Prod Commun 2010; 5(11):1747–50. https://doi.org/10.1177/1934578X1000501109

[6] Eguale T, Tadesse D, Giday M. In vitro anthelmintic activity of crude extracts of five medicinal plants against egg-hatching and larval development of Haemonchus contortus. J Ethnopharmacol 2011; 137(1):108–13; https://doi.org/10.1016/j.jep.2010.04.063

[7] Hutchinson GW. Nematode parasites of small ruminants, camelds and cattle diagnosis with emphasis on anthelmintic efficacy and resistance testing. Australia and New Zealand Standard Diagnostic Procedures, Canberra, Australia, pp 17–18, 2009.

[8] Saravia A, Miraballes C, Riet-Correa F, Castro-Janer E. Eimeria spp. in dairy calves in Uruguay. Identification, dynamics of oocyst excretion and association with the age of calves. Vet Parasitol Reg Stud Rep 2011; 2(1):96–102; https://doi.org/10.1590/S0074-02762011000300004

[9] Lespine A, Chartier C, Hoste H, Alwiner E. Endectocides in goats: pharmacology, efficacy and use conditions in the context of anthelmintics resistance. Small Rum Res 2012; 103(1):10–7; https://doi.org/10.1016/j.smallrumres.2011.10.013

[10] Le Jambre L, Dominik S, Eady S, Henshall J, Golditz I. Adjusting worm egg counts for faecal moisture in sheep. Vet Parasitol 2007; 145(1-2):108–15; https://doi.org/10.1016/j.vetpar.2006.11.017

[11] Molan A, Liu Z, De S. Effect of pine bark (Pinus radiata) extracts on sporulation of coccidian oocysts. Folia Parasitol 2009; 56(1):1–5; https://doi.org/10.1163/157078109X454349

[12] Ramada JS, Soriano ML, Abella JC. Anthelmintic properties of panyawan (Tinospora rhumphii Boerl.) crude aqueous stem extracts against gastrointestinal helminths of naturally infected upgrading goats (Capra hircus). Anim Biomed Anim Husb 2018; 10(1):9–17.

[13] Wang G, Han J, Zhao L, Jian D, Liu Y, Liu X. Anthelmintic activity of steroid saponins from Panax polyphylla. Phytochem Int J Phytother Phytopharmacol 2010; 17(14):1102–5; https://doi.org/10.1016/j.phyomed.2010.04.012

[14] Noack S, Chapman H, Selzer P. Anticoccidial drugs of the livestock industry. Parasitol Res 2019; 118(7):2009–26; https://doi.org/10.1007/s00436-019-06343-5

[15] Bushkin G, Motari E, Carpentieri A, Dubey JP, Costello CE, Robbins PW, et al. Evidence for a structural role for acid-fast lipids in oocyst walls of Cryptosporidium, Toxoplasma, and Eimeria. mBio 2013; 4(5):e0387–13; https://doi.org/10.1128/mBio.00387-13

[16] Mai K, Sharan MA, Walker RA, Katrih M, De Souza D, McConville MJ, et al. Oocyst wall formation and composition in coccidian parasites. Mem Inst Oswaldo Cruz 2009; 104(2):281–9; https://doi.org/10.1590/S0070-42162009000200022

[17] Jones G, McAllister T, Muir A, Cheng K. Effects of Sainfoin (Onobrychis vicifolia Scop.) Condensed tannins on growth and protein synthesis by four strains of ruminal bacteria. Appl Environ Microbiol 1994; 60(4):1374–8; https://doi.org/10.1128/aeem.60.4.1374-1378.1994

[18] Philipp GV, Zaim P, Taylor MA, Dorchies P. Comparative efficacy of diclazuril (Vecoxan®) and toltrazuril (Baycox bovis®) against natural infections of Eimeria bovis and Eimeria zuernii in French calves. Vet Parasitol 2014; 206(3-4):129–37; https://doi.org/10.1016/j.vetpar.2014.10.003

[19] Mali R, Mahajan S, Mehta A. In-vitro anthelmintic activity of stem bark of Mimusops elengi Linn. Pharmacogn Mag 2007; 3(10):73–6.

[20] Fang S, Lillehoj HS, Jung SL, Lee KW, Bravo D, Lillehoj EP. Effects of dietary supplementation with phytothermics on vaccine-stimulated immunity against infection with Eimeria tenella. Vet Parasitol 2011; 181(2-4):97–105; https://doi.org/10.1016/j.vetpar.2011.05.003

[21] Nweze N, Obi vulu I. Anticoccidial effects of Ageratum conyzoides. J Ethnopharmacol 2009; 122(1):6–9; https://doi.org/10.1016/j.jep.2008.11.014

[22] Khaliq T, Mumtaz F, Zia-ur-Rahman, Javed I, Iftikhar A. Nepr nephroprotective potential of Rosa damascena Mill flowers, Cichorium intybus Linn roots and their mixtures on gentamicin-induced toxicity in albino rabbits. Pak Vet J 2015; 35(1):43–7.

[23] Wang D, Zhou L, Li W, Zhou H, Hou G. Anticoccidial effect of Piper sarmentosum extracts in experimental coccidiosis in broiler chickens. Trop Anim Health Prod 2016; 48:1071–8; https://doi.org/10.1007/s11250-016-1034-5

[24] Andrews AH. Some aspects of coccidiosis in sheep and goats. Small Rum Res 2013; 110(2-3):93–5; https://doi.org/10.1016/j.smallrumres.2012.11.011

[25] Taylor M, Catchpole J, Marshall J, Marshall R, Hoeben D. Histopathological observations on the activity of diclazuril (Vecoxan) against the endogenous stages of Eimeria crandallis in sheep. Vet Parasitol 2003; 116(4):305–14; https://doi.org/10.1016/s0304-4017(03)00256-5

[26] Gregory M, Catchpole J. Oxide coccidiosis: the pathology of Eimeria crandallis infection. Int J Parasitol 1990; 20(7):849–60; https://doi.org/10.1016/0020-7519(90)90022-f

[27] Gibbons P, Love D, Craig T, Budke C. Efficacy of treatment of elevated coccidial oocyst counts in goats using amprolium versus ponazuril. Vet Parasitol 2016; 218:1–4; https://doi.org/10.1016/j.vetpar.2015.12.020