Embryonic Origin and Remodeling of the Urinary and Digestive Outlets

Chen Wang1*, JingYing Wang1,2,3†, Joseph G. Borer1, Xue Li1,2,3‡

1 Department of Urology, Boston Children’s Hospital, Boston, Massachusetts, United States of America, 2 Department of Surgery and Pathology, Harvard Medical School, Boston, Massachusetts, United States of America, 3 Harvard Stem Cell Institute, Cambridge, Massachusetts, United States of America

Abstract

Separating digestive and urinary outlets is a critical step during mammalian embryogenesis. However, the natural history of these structures is poorly studied, and little is known about their embryonic origin. Here, we show that peri-cloacal mesenchymal (PCM) progenitors are the major source of these structures. Surprisingly, PCM progenitors also contribute to perineum, a structural barrier separating the urinary and digestive tracts, suggesting a potential role of PCM progenitors in establishing independent urinary and digestive outlets. We demonstrate that Six1 and Six2 are complementarily but asymmetrically expressed in the PCM progenitors. Deletion of these genes results in decreased cell survival and proliferation, and consequently in agenesis of the perineum and severe hypoplasia of the genital tubercle. Together, these findings suggest that PCM progenitors are the unexpected source of perineum and genital tubercle, and establish a basic framework for investigating normal and abnormal development of anorectal and genitourinary structures.

Introduction

Partitioning of a hollow structure is one of the most fundamental remodeling processes during embryogenesis. For example, a single tube of cardiac outflow tract is divided into pulmonary and aortic trunks - a vital step that ensures separation of oxygen-rich and oxygen-depleted blood circulations. Cloaca, the most caudal end of the hindgut, is a common primordial structure of both digestive and urinary outlets. Developmental anomalies involving cloaca remodeling are among the most common forms of human birth defects. However, cloaca morphogenesis and remodeling of digestive and urinary outlets have received little attention and are poorly understood.

A prevailing textbook model indicates that a putative urorectal septum divides the cloaca along the dorsoventral axis. The dorsal compartment forms the digestive outlet including rectum and anus, while the ventral urogenital sinus undergoes complex transformation to form bladder, urethra as well as related reproductive organs. More than a century ago, Rathke suggested that fusion of the bilateral longitudinal folds (Rathke’s fold) led to formation of the urorectal septum [1]. In this model, two bilateral ridges fuse like a zipper moving caudally to divide the cloaca into two compartments. This concept is supported by Retterer in the 1890s [2] and recently by investigators including Hynes and Fraher [3]. However, lack of essential evidence to support tissue fusion, including localized apoptosis and/or epithelial-to-mesenchymal transition, casts serious doubt on the model [4–8]. Indeed, Tourneaux proposed an alternative interpretation, and suggested that the urorectal septum is a coronally-oriented wedge of mesenchyme, known as the Tourneux’s fold [9], which divides cloaca like a theater curtain dropping in a rostral to caudal direction. In contrast to these two urorectal septum-based models, van der Putte liked the cloaca to a “tubular structure” that is “increasingly more bent toward the surface” [5,6]. Based on this interpretation, an entirely different ventral displacement model was put forward, which suggested that a disproportionate growth of ventral relative to dorsal cloacal mesenchyme transforms instead of divides the cloaca into the urogenital and digestive compartments. It is unclear, however, how such transformation leads to the separation of the urinary and digestive tracts.

Despite the differences among these interpretations, all models suggest that a discrete population of mesenchymal progenitors is critical for dividing the cloaca. However, a paucity of molecular and cell biological studies of cloacal mesenchymal progenitors hinders our ability to reconcile the controversies of the aforementioned models. The perineum is the diamond-shape area superficial to the pelvic diaphragm and bordered by the pubic arch, ischial tuberosities and coccyx [6]. The term “perineum” is also used for the restricted area between the anus and the urethral orifice, we refer this region as the “midline epithelium of the perineum” to avoid confusion. Since the perineum is the physical barrier that separates urinary and digestive outlets, a better understanding of its embryonic origin would have an important implication in cloacal morphogenesis. According to the classic Rathke’s fold and the Tourneux’s fold models, the putative urorectal septum consisting of intra-cloacal mesenchyme (ICM) gives rise to the perineum [1,2]. The ventral displacement model,

* E-mail: sean.li@childrens.harvard.edu
† These authors contributed equally to this work.

Citation: Wang C, Wang J, Borer JG, Li X (2013) Embryonic Origin and Remodeling of the Urinary and Digestive Outlets. PLoS ONE 8(2): e55587. doi:10.1371/journal.pone.0055587

Editor: Claudia Kappen, Pennington Biomedical Research Center/LSU, United States of America

Received October 2, 2012; Accepted December 27, 2012; Published February 4, 2013

Copyright: © 2013 Wang et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work is supported by the Harvard Stem Cell Institute, National Institutes of Health/National Institute of Dental and Craniofacial Research (NIH/NIDCR) (1R01DE019823, XL) and NIH/NIDDK (National Institute of Diabetes and Digestive and Kidney Diseases) (1R01DK091645-01A1, XL). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.
on the other hand, does not explicitly address the nature of progenitors that contribute to the perineum [5]. Genetic lineage tracing studies of cloacal epithelial cells demonstrate that midline surface epithelium of the perineum has an endodermal origin [10]. This observation implies that the ICM progenitors are the source of perineum, and indirectly supports the cloacal septum-based models. However, a direct genetic fate mapping analysis of the peri-cloacal mesenchyme (PCM) progenitors instead suggests that PCM are the major source of the perineum [11]. Therefore, the central issue of embryonic origin of the perineum remains to be elucidated.

In this study, we use an inducible genetic fate-mapping approach to interrogate PCM lineages, and demonstrate that the PCM progenitors contribute directly to the perineal stromal tissue. We show for the first time the complementary and asymmetrical expression patterns, as well as their lineage distribution patterns, of Six1 and Six2 in PCM progenitors. Deletion of these two genes results in a decreased PCM progenitor cell survival and proliferation, and consequently severe genital tubercle hypoplasia and perineum agenesis. Thus, PCM is an unexpected source of perineum, which is essential for formation and remodeling of cloaca and urogenital structures. Taken together, these findings suggest that a process reminiscent to vascular occlusion results in a partitioning of cloaca, and provide a basic framework for investigating cellular and molecular mechanisms of urinary and digestive outlet development.

Results

Asymmetric and complementary expression patterns of Six1 and Six2 in PCM progenitors

Among six different members of Six1-family transcription factors, the high degree of similarity between Six1 and Six2 suggests that they may share similar function in vivo [12,13]. We have shown that Six1 is highly expressed in the PCM progenitors with a dorsal-to-ventral gradient, and that Six1 is required for normal urinary tract development [11]. To begin to characterize the potential function of Six2, we first compared its dynamic expression pattern with Six1 (Fig. 1). Six1 transcripts were detected in PCM cells as early as e10.5 (Fig. 1A). Its expression was maintained in genital mesenchyme between e11.5–e13.5 (Figs. 1B–D). At later stages (e14.5 and e15.5), Six1 expression was significantly reduced and restricted to mesenchyme adjacent to the urethral plate and became undetectable in the preputial fold at e14.5 (Figs. 1E and F).

Six1 was weakly expressed in metanephric mesenchyme (MM) but highly expressed in PCM at e10.5. On the other hand, Six2 was enriched in MM but was hardly detectable in PCM at this stage (Fig. 1G, arrow). A day later, at e11.5, both genes were highly expressed in the genital swellings (Figs. 1B and H). At later stages, Six2 was strongly expressed in mesenchymal cells surrounding the urethral plate at e13.5 but was significantly down regulated at e15.5 (Figs. 1J–L). Similar to Six1, Six2 expression was diminished in the preputial fold (Figs. 1K and L). To highlight a spatial distribution pattern of Six2 at the critical period of cloacal morphogenesis at e11.5, we performed RNA in situ hybridization experiments on serial adjacent sagittal sections. Six2 appeared to be expressed in all PCM progenitors (Fig. 1M). However, its transcripts were enriched in the ventral PCM (vPCM) and reduced in the dorsal PCM (dPCM) (Fig. 1M–P). This asymmetric expression pattern is in contrast to Six1, which is highly expressed in the dPCM [11]. In addition, Six2 was absent from the urorectal septum, which consists primarily of the ICM progenitors (Figs. 1O and P). Thus Six1 and Six2 have asymmetric, yet complementary, expression patterns in PCM progenitors, with Six1 enriched dorsally and Six2 ventrally. Both genes are absent from ICM cells.

Six2-expressing PCM progenitors contribute to urogenital tissues including the perineum

The restricted Six2 expression pattern in PCM cells provided a unique opportunity to interrogate lineage distribution patterns of PCM progenitors during development, as well as remodeling of urinary and digestive outlets. We first performed a genetic fate mapping analysis using a Six2lacZ mouse line (Fig. 2). The eGFP and Cre fusion gene (GC) replaces and fully recapitulates the endogenous Six2 gene expression pattern since the same targeting strategy were used to generate other Six2 mutant alleles, including Six2GCE allele [14]. The GC fusion protein has a constitutively-active, site-specific Cre recombinase activity that is able to turn on a LacZ reporter, R26R-lacZ (R26RloxP) [15]. Consequently, Six2-expressing progenitors and their progenies are selectively and permanently labeled by lacZ in Six2GCE; R26RloxP rats [15]. We analyzed these embryos at three developmental stages before (e11.75) and after (e13.5) cloacal septation, and during perineum formation (e15.5) (Fig. 2). Sagittal and cross sections of genital tubercles were assayed for lacZ gene activity, a surrogate of Six2 lineages. At e11.75, lacZ cells were detected in the metanephric mesenchyme, vPCM, dPCM, and to a much less extent, the urethral plate and anorectal epithelial cells. No lacZ cells were observed in the genital tubercle ectodermal epithelial cell layer (Figs. 2A and B). At e13.5 and e15.5, the majority, if not all, urogenital mesenchyme including the perineal stromal and preputial fold tissues were lacZ cells (Figs. 2C–J). Few lacZ cells at the urethral plate and anorectal epithelium were observed at e13.5 and e15.5 (Figs. 2C–J). In addition, mesenchymal cells surrounding the anal canal were all lacZ-positive (Fig. 2G and H). Thus, Six2GCE PCM progenitor cell lineages contribute to most, if not all, anogenital mesenchymal tissues.

We next sought to determine when PCM progenitors are committed to these distinct tissues. Toward this end, we used another Six2GCE mouse line, which expresses a tamoxifen-inducible eGFP and CreER (GCE) fusion protein, to map the fate of Six2-expressing PCM progenitors [14]. A single dose of tamoxifen was used to treat females pregnant with Six2GCE/+;R26RlacZ/+ double heterozygous embryos at e11.5, e13.5, e14.5 and e15.5, and these embryos were analyzed at e17.5 for lacZ reporter gene activity. Since Six2 is strongly expressed in renal progenitors (Fig. 1), we used the kidney as an indicator of efficient tamoxifen-induced Cre recombination (Figs. 3A, E, I and M). Tamoxifen treatment at e11.5 resulted in extensive lacZ cells in the kidney; as expected, progressively fewer lacZ cells were detected in kidneys that were treated with tamoxifen at later stages (Figs. 3A, E, I and M). We next analyzed the spatiotemporal distribution patterns of lacZ cells in urogenital tissues from these same embryos. Tamoxifen treatment at e11.5, a stage in which Six2 was strongly expressed in PCM but absent from ICM cells (Figs. 1M–P), resulted in abundant lacZ cells that were broadly distributed in the perineum, preputial fold and the prospective corporal body (Figs. 3B–D). Though fewer in number, a similar distribution pattern of lacZ cells was observed when tamoxifen was administered at e13.5 (Figs. 3F–H). In contrast, tamoxifen injections at later stages (e14.5 and e15.5) resulted in lacZ cells only at the distal genital tubercle region, near the urethral plate (Figs. 3J–L). lacZ cells were observed when tamoxifen was administered at e13.5 (Figs. 3F–H). No lacZ cell was detected in the perineum in these embryos. Together, results from these constitutive and inducible genetic fate-mapping analyses demonstrate that the PCM progenitors are the major source of the
perineum, and that these progenitors are committed to the fate of the perineum as early as e11.5 prior to separation of the urinary and digestive outflow tracts. 

**Six1 and Six2 have redundant functions in PCM progenitors**

Mouse mutants lacking either Six1 or Six2 die at birth due to renal agenesis defects [12,14,16–18]. All of Six2−/− (n = 12) and Six1+/−;Six2−/− (n = 18) mutants had grossly normal genital tubercle and anal structures [11] (Fig. 4A and data not shown). Only a small percentage (20%, n = 20) of Six1−/− embryos had a displacement of the urethral meatus at a ventral and proximal region of the external genitalia, resembling a hypospadias-like phenotype (Fig. 4A and data not shown) [19–21]. Loss of one allele in the Six2 in Six1−/− mutant background (Six1−/−;Six2+/−) (n = 14) increased penetrance of the hypospadias-like phenotype to 85.7% (Figs. 4A, D and E). Mutant genital tubercles were overall smaller than wild type littermate controls. In addition, the urethral meatus were displaced at the base of external genitalia (Figs. 4D and E). Loss of both genes (Six1−/−;Six2−/−, n = 3) resulted in agenesis of the perineum and severe hypoplastic external genitalia (Figs. 4A, F and G). Thus, Six1 and Six2 have redundant and essential functions in PCM progenitors during perineum and genital tubercle formation.

To better understand urogenital and anorectal defects of Six1;Six2 compound mutants, we performed histological analysis...
of serial sagittal sections from newborn male and female pups (Figs. 4H–S). Perineal stromal tissue, which separates urinary and digestive tracts, was apparent and indicated by the anogenital distance in both wild type male and female pups (Figs. 4H–S, bracket). The same tissue in the Six1−/−;Six2−/− mutant was hypoplastic, and the anogenital distance was significantly reduced (Fig. 4P and Q). Consistent with these gross defects, the mutant genital tubercles were hypoplastic. The Six1 and Six2 double null mutants exhibited a severe agenesis defect since the genital tubercle and the perineum were nearly absent (Fig. 4R and S). In

Figure 2. A genetic fate map of Six2-expressing PCM progenitors. X-gal staining (blue) of sagittal (A–D, G and H) and cross (E, F, I, J) sections from e11.75, e13.5 and e15.5 Six2GC+/+;R26RLacZ double heterozygous embryos. All sections were counterstained with eosin (pink). A, anus; PG, preputial gland; see Figure 1 for more abbreviations. doi:10.1371/journal.pone.0055587.g002

Figure 3. An inducible genetic fate map of Six2-expressing PCM progenitors. Double Six2GC+/+;R26RLacZ pregnant females were treated with a single dose of tamoxifen at e11.5, e13.5, e14.5 and e15.5, and all embryos were collected and analyzed at e17.5 with X-gal staining (blue). (A, E, I and M) kidney sections; (B–D, F–H, J–L and N–P) urogenital sections. CB, prospective corporal body; GT, genital tubercle; P, perineum; PF, preputial fold; PG, preputial gland; U, urethra. doi:10.1371/journal.pone.0055587.g003
addition, the anal canal of the double null mutants was absent, resulting in a direct exposure of rectum epithelium (Fig 4, compare asterisk in M and S). Together, these findings suggest that Six1 and Six2 are required for the development of both digestive and urinary outlets.

Survival and proliferation of PCM progenitors depend on Six1 and Six2

Because of the rarity of obtaining double null mutants, we used Six1<sup>−/−</sup>;Six2<sup>−/−</sup> compound mutants to further characterize primary defects of digestive and urinary outlets during early embryogenesis. In wild type embryos, three populations of mesenchymal cells were apparent at e11.5 along midline sagittal sections, the ventral vPCM, the dorsal dPCM and the internal sPCM. These populations are necessary for the formation of the digestive and urinary outlets. In the compound mutants, the anal canal was absent, resulting in a direct exposure of rectum epithelium. Together, these findings suggest that Six1 and Six2 are required for the development of both digestive and urinary outlets.
ICM (Fig. 5). The caudal side of the cloaca was covered by the cloacal membrane, which was a composite of endoderm and ectoderm epithelia but devoid of any mesenchyme. At this stage, the distal end of ICM was juxtapositioned but not fused with dPCM and the cloacal membrane (Fig. 5C, asterisk), the likely site of the future anal canal. This unique juxtaposition separated the urogenital sinus and rectum, thereby serving as the first sign of separation between the urinary and the digestive tract (Fig. 5C). Asymmetric growth of these mesenchymal cells was likely involved in remodeling of the urogenital sinus to form the genital tubercle and the anal canal. In Six1\(^{-/-}\);Six2\(^{-/-}\) mutants, the relative position of the cloacal mesenchyme, the cloacal membrane, and the unique juxtaposition were maintained (Fig. 5F). However, it was apparent that both the dPCM and the vPCM were hypoplastic, and that the size of the mutant genital tubercle was significantly smaller (Fig. 5D–F, and data not shown). These observations suggest that Six1 and Six2 may control the growth and/or expansion of these tissues.

Since Six1 is required for the survival of renal and cardiac progenitors [12,16,22], we first used TUNEL assays to determine if survival of the PCM progenitors depended on Six1 and Six2 (Fig. 6). Similar levels of apoptotic cells were detected in the common nephric duct in littermate controls and Six1\(^{-/-}\);Six2\(^{-/-}\) mutants (Fig. 6A and B), which therefore functioned as an internal control [23]. Apoptotic endoderm and mesenchyme were detected at the juxtaposition at e11.5, as expected [24,25], however, more apoptotic cells were present in Six1\(^{-/-}\);Six2\(^{-/-}\) mutants (Fig. 6D, asterisk). Enhanced apoptosis was also observed in the distal urethral plate epithelium in the mutants (Fig. 6D). In addition, an ectopic TUNEL signal was detected in genitalic mesenchyme at lateral sections (Fig. 6B).

To investigate the spatial distribution pattern of the apoptotic cells, we next used a vital dye LysoTracker Red to examine apoptosis pattern of whole mount embryos at e11.5 (Fig. 6E–H). Three distinct apoptotic domains - distal urethral plate, distal mesenchymal region and proximal urethral plate - were apparent with the LysoTracker Red signal (Fig. 6E). Deletions of Six1 and Six2 caused an increase of apoptosis in the urethral plate (Fig. 6F). Consistent with the TUNEL staining findings, ectopic cell death was detected in the lateral mesenchymal region in mutants (compare Fig. 6E’ to F’). This ectopic increase of cell death persisted at e12.0 (arrowhead in Fig. 6H). Surprisingly, the mutant genital tubercle underwent less cell death in the distal mesenchymal region at e11.5 and e12.0 (Figs. 6E, F and I). As illustrated in Fig. 6I, deletion of Six1 and Six2 alters both the apoptosis level and pattern, changes which may contribute to the hypoplastic defect of the genital tubercle.

We also examined the cell proliferation status of the Six1\(^{-/-}\);Six2\(^{-/-}\) mutants by staining serial embryonic sections with a p-HH3 antibody [11], a mitosis marker of cell cycle. Sections with similar overall anatomic features were selected for this study (Fig. 7). A total of 8 cryostat sections that encompassed genital tubercle were stained with the p-HH3 antibody. Using the shape of cloaca endoderm lining as a guide, proliferating cells from dPCM, vPCM and ICM were counted and compared. The total number of p-HH3 positive cells was significantly lower in Six1\(^{-/-}\);Six2\(^{-/-}\) mutants than in wild type littermate controls (Fig. 7G). Thus, the normal proliferation of PCM progenitors depends on Six1 and Six2. Collectively, these data indicate that Six1 and Six2 are required for proliferation and survival of the PCM progenitors, which may contribute directly to the observed severe hypoplastic phenotype (Fig. 4R and S).

**Six1 and Six2 coordinate expression of critical signal molecules**

Activation of the Bone morphogenetic protein (Bmp) signaling pathway inhibits genital outgrowth and induces apoptosis [26,27]. Mouse mutants without Noggin (an antagonist of Bmp signaling pathway) display hypoplastic genitalia, while Bmp receptor Ia (BmprIa) mutants have overgrowth and reduced cell death, resulting in hyperplastic genitalia [27]. Bmp7 mutants have imperforate anus and genital tubercle defects [28]. Deletion of Eya1, a transcription coactivator of Six1 and Six2, results in increased Bmp signaling [11]. To examine whether Six1 and Six2 were required for expression of candidate genes that are important for genital tubercle growth and patterning, we therefore first examined the expression pattern of Bmp4 and Bmp7 (Figs. 8A–D, 8E–H). We found that Bmp4 expression was enhanced in dorsal lateral mesenchyme (arrowhead in Fig. 8B) and ventral distal mesenchyme (arrow in Fig. 8D) of Six1\(^{-/-}\);Six2\(^{-/-}\) mutants. Bmp7

---

**Figure 5. Hypoplastic genital tubercles of Six1;Six2 compound mutants at e11.5.** Hematoxylin and eosin (H&E) staining of the serial sagittal sections of e11.5 urogenital structure. Asterisk, juxtaposition of ICM, dPCM and the cloacal membrane (CM); B, bladder; CND, common nephric duct; IPCM, lateral PCM; R, rectum; see Figure 1 for more abbreviations. doi:10.1371/journal.pone.0055587.g005
expression in the distal urethral plate epithelium (dUPE) was expanded and extended proximally toward the base of the tubercle in mutants, and its expression in genital tubercle mesenchyme was increased (Figs. 8F and H). Upregulation of Bmp4 and Bmp7 was further confirmed by real time quantitative PCR (qPCR) using microdissected genital tubercle at e11.5 (Fig. 8Q). Furthermore, the expression of Bmp4 downstream target genes Msx1 [11,27], were significantly upregulated in Six1<sup>−/−</sup>;Six2<sup>+/−</sup> mutants (Fig. 8Q), suggesting that Bmp signaling was enhanced.

The canonical Wnt/β-catenin signal pathway is critical for the normal development of urogenital structures [29,30]. Interaction between androgen and Wnt/β-catenin signal pathway promotes formation of male external genitalia [31]. Dkk1 and Dkk2 are potent inhibitors of the Wnt/β-catenin signal pathway [32–35]. In addition, Dkk1 is a downstream effector of Bmp signaling, and
together they promote apoptosis [36]. Because Six1\textsuperscript{\texttt{1\textasciicircum{-}/2\textasciicircum{-}}}\texttt{;Six2\textasciicircum{1\textasciicircum{-}/2\textasciicircum{-}}} mutants displayed increased Bmp signaling (Fig. 8A–H, Q) and apoptosis (Fig. 6), we therefore examined the expression level of Dkk1 and Dkk2. At e13.5, Dkk1 transcripts were detected in mesenchymal cells lateral to the urethral plate, and expression of Dkk1 was slightly upregulated in the Six1\textsuperscript{\texttt{1\textasciicircum{-}/2\textasciicircum{-}}}\texttt{;Six2\textasciicircum{1\textasciicircum{-}/2\textasciicircum{-}}} mutants (Figs 8M–P). Consistently, both Dkk1 and Dkk2 genes were significantly upregulated in the mutant genital tubercle at e11.5, based on a quantitative PCR analysis of micro-dissected tissues (Fig 8Q).

Six1 is required for Fibroblast growth factor (Fgf8) expression during cardiac and craniofacial development [22]. Exogenous Fgf8 promotes genital tubercle outgrowth in organ cultures [37], and its expression in the distal urethral plate depends on both Shh and Wnt/\beta-
catenin signaling pathways [29,30,38,39]. However, conditional deletion of Fgf8 has no obvious genital tubercle defect [40]. On the other hand, a mutation in murine Fgf10 results in a hypospadias-like phenotype [41]. We detected reduced expression of Fgf8 in Six1\textsuperscript{\texttt{1\textasciicircum{-}/2\textasciicircum{-}}}\texttt{;Six2\textasciicircum{1\textasciicircum{-}/2\textasciicircum{-}}} mutants at e12.5 (Figs. 8I–L), but increased expression of Fgf10 (Fig. 8Q), suggesting that downregulation of Fgf8 might be compensated by upregulation of Fgf10. Indeed, expression of dual specificity protein phosphatase 6 (Dusp6), which is downstream of the Fgf signaling pathway [11,40], was not affected (Fig. 8Q).

Taken together, these candidate gene expression analyses suggest that deletions of both Six1 and Six2 disrupt dynamic expression patterns of several critical signal molecules required for normal development of urogenital structures.

**Discussion**

Our findings uncover that PCM progenitors are the unexpected source of perineum and urogenital organs. We show for the first time that Six1 and Six2 are asymmetric and complementarily expressed in the PCM progenitors, where they are required for proliferation and survival of these progenitors. These observations are suggestive that a process reminiscent to vascular occlusion underlies the partitioning of cloaca and remodeling of urogenital structures.
Asymmetric growth of mesenchyme is the major driving force that transforms cloaca into urinary and digestive tracts (Fig. 9). Therefore, patterning of the cloacal mesoderm is a central issue of cloaca morphogenesis. Along the rostrocaudal axis, cloaca is surrounded by mesenchyme at the rostral ICM cells and lateral PCM cells but not the caudal cloacal membrane, which is devoid of mesenchyme (Fig. 9A). Thus, an intrinsic asymmetry is established because of the absence of mesenchyme in the cloacal membrane. A rapid increase in both PCM and ICM cells occludes the cloacal cavity and separates the hindgut (rectum and anal canal) and urogenital sinus (bladder and urethra). The process also pushes the cloacal duct, the remnant of cloaca, caudally towards the surface of the perineum. Consequently, independent digestive and urinary tracts are established, and the cloaca duct persists at the midline surface of the perineum epithelium.

Unlike the intrinsic asymmetry of the rostrocaudal axis, cloaca is surrounded at all sides by the PCM progenitors along the dorsoventral axis (Fig. 9B and C). It is not immediately clear how asymmetric gene expression and growth along dorsoventral axes are established. An intriguing observation is the high levels of apoptosis at the dPCM and tail gut region (Fig. 6) [24,25]. This localized cell death likely retards growth of the dPCM, thereby causing asymmetric growth along the dorsoventral axis and a ventral shift of the cloacal membrane, as proposed by van der Putte [6]. Asymmetric expression patterns of Six1 and Six2 suggest that PCM is indeed patterned along the dorsoventral axis, as Six1 is highly enriched in the dPCM [11] while Six2 is enriched in vPCM (Fig. 1M-P). Consistently, Six1-positive lineages are predominantly localized at the ventral side of the genital tubercle (Fig. 9) [11]. We have also shown that Six1 and Six2 coordinately control proliferation and survival of PCM progenitors, potentially through candidate signal molecules (Fig. 8), and that genetic deletion of Six1 and Six2 results in agenesis of the perineum and severe hypoplastic genitalia. These data suggest that patterning along the dorsoventral axis is required for completion of cloacal division, as well as outgrowth and patterning of the genital tubercle.

Shh is expressed in the cloacal endoderm and is required for all stages of genitourinary tract development [30,38,39]. Shh signals and controls cell cycle kinetics of mesenchyme [42]. It is worth noting that Six6, a homology of Six1, is directly involved in modulating cell cycle of retinal progenitor [43]. Shh is maintained in Six1 and Six2 compound mutants (data not shown) and Eya1 mutant [11], raising a possibility that Shh maybe an upstream regulator. A key future question would be to understand intrinsic and extrinsic mechanism underlying the asymmetric growth and patterning of the cloacal mesenchyme.

The proposed cloacal occlusion model is supported by the unexpected origin of the perineum discovered here and previously [10,11]. Seifert et al., reported previously that the midline epithelium of the perineum has an endodermal origin [10]. Of the various models put forth, the cloacal occlusion model best accounts for the observations of the shape (a narrow line) and asymmetric positioning (midline caudal surface) of the endoderm remnant (Fig. 9A and B). As illustrated in Figure 9A, occlusion of the cloaca results in displacement of the cloacal duct and formation of the perineum. On the other hand, the Rathke’s fold model predict that any surviving endodermal cells would be randomly distributed and embedded in the perineum stromal layer [1,2]. The Tourneux’s fold model [9], as well as the transformation model [5], would have predicted that the cloacal remnant is distribution broadly at the surface of the perineum. Both Six1 and Six2 are expressed in the PCM, but not ICM progenitors. Interestingly, Six1- and Six2- positive cell lineages contribute directly to perineum tissue. Furthermore, inducible genetic fate mapping demonstrates, for the first time, that Six2 expression in PCM progenitor contributes to the perinermal stromal tissue, as early as e11.5. While the occlusion model does not exclude the possibility that ICM progenitors might also contribute to the perineum, these observations suggest that PCM progenitors are involved in perineum formation. Indeed, Six1 and Six2 double null mutants exhibit perineum agenesis defects.

Taking together, we conclude that cloacal mesoderm progenitors are central to the separation of the urinary and digestive tracts, as well as to the outgrowth and patterning of the genital tubercle. We postulate that asymmetric growth causes narrowing of cloaca at the location marked by the cloacal membrane. The endodermal remnant is pushed to the midline epithelial surface of perineum between the anus and urogenital meatus. We now refer to the morphogenesis event of the cloaca as “cloacal occlusion”. Since the term septation has been used in many developmental contexts, and implies explicitly the existence of a septum, which is still a matter of speculation during cloacal morphogenesis, we therefore prefer the use of the cloacal occlusion to describe the mechanism of cloacal division. This model provides a basic
framework, i.e. patterning along dorsoventral and rostrocaudal axes, with which to investigate normal and abnormal development of the anorectal and genitourinary structures.

**Materials and Methods**

**Mice**

All animal studies were performed according to protocols reviewed and approved by the institutional animal care and use committee at the Children’s Hospital Boston. Six1 [12], Six2GCE/+ and Six2GCE/- [14], R26RlacZ [15] mice have previously been reported. Genotyping of the mice was performed as described. For temporal induction of Cre recombination, tamoxifen (Sigma, T5648) was dissolved in sesame oil (Sigma, S3547) and administrated by intraperitoneal (IP) injection (50 g/kg body weight).

**Histology, Immunohistochemistry and in situ hybridization**

Embryos were fixed in 4% paraformaldehyde (PFA), embedded in OCT, and frozen sections prepared at a thickness of 12 um. H&E staining were performed by standard procedures. β-galactosidase activity was detected with previously described methods and counter stained with cosin [11,12,22,43]. Whole-mount and section in situ hybridization were performed as described previously [11,22]. To label proliferating cells, e11.5-staged embryos were dissected, fixed in 4% paraformaldehyde for 2 hours at 4°C, and then sectioned at 14 um. Embryo sections were incubated with anti-pH3 (Upstate) at a 1:200 dilution, as previously described [11,22]. The number of p-HH3+ cells was averaged from 6 sections per embryo.

**TUNEL and Lysotracker Red® Staining**

Both terminal deoxynucleotidyl transferase nick end labeling (TUNEL) (Roche) and Lysotracker Red® (Invitrogen) staining were performed according to the manufacturer’s protocol and described previously [11]. Briefly, embryos were dissected and stained with 5 uM lysotracker® in PBS at 37°C for 30 min. Embryos were then washed in PBS several times prior to PFA fixation. Microdissected genital tubercles were imaged using an Olympus SZX16 fluorescence dissection microscope equipped with a DP71 digital camera. Cryostat sections were used for TUNEL assays.

**Quantitative real-time PCR analysis**

Genital tubercle tissue of e11.5 embryos was micro-dissected and snap-frozen on a dry ice/ethanol bath. RNA was purified based on manufacturer’s protocols (Qiagen RNeasy mini). cDNA was synthesized using the Stratagene Accuscript™ High Fidelity 1st strand cDNA synthesis (Agilent Technologies) using 200 ng of total RNA. Relative gene expression levels were normalized to a 2-actin internal control, and analyzed using SYBR Green Master Mix (Applied Biosystems). The following oligos were used: 2-actin F: TCG TCG ACA ACG GCT CCG GCA TGT; 2-actin R: CCA GCC AGC TGG AGA AT; Fgf8 F: GGG GAC CTA ATT GCC AAG AG; Fgf8 R: TGT ACC AAC CGC CTC GGT ACT TG; Fgf10 F: GCC ACC AAC TGC CTG TCTT CCT C; Fgf10 R: CTG ACC TCC CAA GAC CTG TTC TC; Dkk1 F: ATA TCC CAG AAG AAC CAC ACT G; Dkk1 R: CTT TCC GTG TTT GCT TGG TG; Dkk2 F: GCA TCC TCA CCC CAC AT; Dkk2 R: CAG GCA CAA CAA AAC CCA CG; Dusp6 F: CTC GTA TCA CTG GAG CCA AAA C; Dusp6 R: TCT GGA TGT AGC CCA CTG T.

**Acknowledgments**

We would like to thank Chashe Guo, Ye Sun, Gerald Mingin and Ian Teng for helpful discussions, and Melanie Pennison for comments on the manuscript.

**Author Contributions**

Conceived and designed the experiments: XL. Performed the experiments: CW JW. Analyzed the data: CW JW JGB XL. Wrote the paper: CW JW JGB XL. JGB XL.

**References**

1. Rathke H (1832) Abhandlungen zur Bildungs- und Entwicklungsgeschichte der Tiere. Leipzig.
2. Retterer E (1890) Sur l’origine et l’évolution de la région anogenitale des mammiferes. Journal of Anatomy (Paris) 26: 126–216.
3. Hynes PJ, Fraher JP (2004) The development of the male genitourinary system. I. The origin of the urogenital septum and the formation of the perineum. Br J Plast Surg 57: 27–36.
4. Penington EC, Hutson JM (2003) The absence of lateral fusion in cloacal partition. J Pediatr Surg 38: 1287–1295.
5. van der Putte SC (1986) Normal and abnormal development of the anorectum. J Pediatr Surg 21: 434–440.
6. van der Putte SC (2005) The development of the perineum in the human. A comprehensive histological study with a special reference to the role of the stromal components. Arch Anat Embryol Cell Biol 177: 1–131.
7. Paulus CN, Morerale RF, Holoka KM, Land RE, Hutchins GM (1999) Septation and differentiation of the embryonic human cloaca. J Pediatr Surg 34: 877–884.
8. Kluth D, Hillen M, Lambrecht W (1995) The principles of normal and abnormal hindgut development. J Pediatr Surg 30: 1143–1147.
9. Tournaye F (1838) Sur les premiers developpements du cloaque du tube genital et de l’anus chez l’embryon de mouton. Journal of Anatomy (Paris) 24: 503–517.
10. Seifer AW, Hafet BD, Coho MJ (2008) Cell lineage analysis demonstrates an endodermal origin of the distal urethra and perineum. Dev Biol 318: 143–152.
11. Wang C, Gargollo P, Guo C, Tang T, Mingin G, et al. (2011) Six1 and Eya1 are critical regulators of peri-cloacal mesenchymal progenitors during genitourinary tract development. Dev Biol 368: 186–194.
12. Li X, Oghi KA, Zhang J, Krones A, Bub KT, et al. (2003) Eya protein phosphatase activity regulates Six1-Dach-Eya transcriptional effects in mammalian organogenesis. Nature 426: 247–254.
13. Kawakami K, Sato S, Ozaki H, Beda K (2000) Six family genes—structure and function as transcription factors and their roles in development. Bioessays 22: 616–626.
14. Kobayashi A, Valerius MT, Magldorf JW, Carroll TJ, Selt M, et al. (2008) Six2 defines and regulates a multipotent self-renewing nephron progenitor population throughout mammalian kidney development. Cell Stem Cell 3: 169–181.
15. Soriano P (1999) Generalized lacZ expression with the ROSA26 Cre reporter strain. Nat Genet 21: 70–71.
16. Xu PX, Zheng W, Huang L, Maire P, Laclèd C, et al. (2003) Six1 is required for the early organogenesis of mammalian kidney. Development 130: 3085–3094.
17. Ozaki H, Nakamura K, Funahashi J, Ikeda K, Yamada G, et al. (2008) Six1 controls patterning of the mouse otic vesicle. Development 135: 531–562.
18. Selt M, Lagutin OV, Bowling B, Hendrix J, Cai Y, et al. (2006) Six2 is required for suppression of nephrogenesis and progenitor renewal in the developing kidney. Embo J 25: 5214–5228.
19. Baschlo AD, Cunha GR, Baskin LS (2012) Molecular mechanisms of external genitalia development. Differentiation.
20. Rodriguez E, Jr., Weiss DA, Yang JH, Menshenina J, Ferretti M, et al. (2011) A Tbx1-Six1/Eya1-Fgf8 genetic pathway controls mammalian cardiovascular and craniofacial morphogenesis. J Clin Invest 121: 1503–1515.
23. Batourina E, Tsai S, Lambert S, Sprendle P, Viana R, et al. (2005) Apoptosis induced by vitamin A signaling is crucial for connecting the ureters to the bladder. Nat Genet 37: 1082–1089.
24. Sasaki C, Yamaguchi K, Akita K (2004) Spatiotemporal distribution of apoptosis during normal cloacal development in mice. Anat Rec A Discov Mol Cell Evol Biol 279: 761–767.
25. Qi BQ, Beasley SW, Williams AK, Fizele F (2000) Apoptosis during regression of the tailgut and septation of the cloaca. J Pediatr Surg 35: 1556–1561.
26. Morgan EA, Nguyen SB, Scott V, Stadler HS (2003) Loss of Bmp7 and Fgfl signaling in Hoxa13-mutant mice causes hypospadias. Development 130: 3095–3109.
27. Suzuki K, Bachiller D, Chen YP, Kamikawa M, Ogi H, et al. (2003) Regulation of outgrowth and apoptosis for the terminal appendage: external genitalia development by concerted actions of BMP signaling [corrected]. Development 130: 6209–6220.
28. Wu X, Ferrara C, Shapiro E, Grishina I (2009) Bmp7 expression and null phenotype in the urogenital system suggest a role in re-organization of the urethral epithelium. Gene Expr Patterns 9: 224–230.
29. Lin C, Yin Y, Long F, Ma L (2008) Tissue-specific requirements of β-catenin in external genitalia development. Development 135: 2015–2025.
30. Miyagawa S, Moon A, Haraguchi R, Inoue C, Harada M, et al. (2009) Dosage-dependent hedgehog signals integrated with Wnt/β-catenin signaling regulate external genitalia formation as an appendicular program. Development 136: 3969–3978.
31. Miyagawa S, Tatoh Y, Haraguchi R, Suzuki K, Iguchi T, et al. (2009) Genetic interactions of the androgen and Wnt/β-catenin pathways for the masculinization of external genitalia. Mol Endocrinol 23: 871–880.
32. Glinka A, Wu W, Delius H, Monaghan AP, Blumenstock C, et al. (1998) Dickkopf1 is a member of a new family of secreted proteins and functions in head induction. Nature 391: 357–362.
33. Cadigan KM, Nusse R (1997) Wnt signaling: a common theme in animal development. Genes Dev 11: 3286–3305.
34. Moon RT, Kohn AD, De Ferrari GV, Kaykas A (2004) WNT and beta-catenin signalling: diseases and therapies. Nat Rev Genet 5: 691–701.
35. Mukhopadhyay M, Shrom S, Rodriguez-Esteban C, Chen L, Tsukui T, et al. (2001) Dickkopf1 is required for embryonic head induction and limb morphogenesis in the mouse. Dev Cell 1: 423–434.
36. Groteveld L, Ruther U (2002) The Wnt antagonist Dickkopf-1 is regulated by Bmp signaling and c-Jun and modulates programmed cell death. Embo J 21: 966–975.
37. Haraguchi R, Suzuki K, Murakami R, Sakai M, Kamikawa M, et al. (2000) Molecular analysis of external genitalia formation: the role of fibroblast growth factor (Fgf) genes during genital tubercle formation. Development 127: 2471–2479.
38. Lin C, Yin Y, Ventb GM, Fisher AV, Long F, et al. (2009) Temporal and spatial dissection of Shh signaling in genital tubercle development. Development 136: 3959–3967.
39. Seifert AW, Boudlin CM, Choi KS, Harfe BD, Cohn MJ (2009) Multiphasic and tissue-specific roles of sonic hedgehog in cloacal septation and external genitalia development. Development 136: 3949–3957.
40. Seifert AW, Yamaguchi T, Cohn MJ (2009) Functional and phylogenetic analysis shows that Fgf8 is a marker of genital induction in mammals but is not required for external genital development. Development 136: 2643–2651.
41. Yucel S, Liu W, Cordiero D, Donjacour A, Cunha G, et al. (2004) Anatomical studies of the fibroblast growth factor-10 mutant, Sonic Hedgehog mutant and androgen receptor mutant mouse genital tubercle. Adv Exp Med Biol 545: 123–148.
42. Seifert AW, Zheng Z, Otmerod BK, Cohn MJ (2010) Sonic hedgehog controls growth of external genitalia by regulating cell cycle kinetics. Nat Commun 1: 1–9.
43. Li X, Perissi V, Liu F, Rose DW, Rosenfeld MG (2002) Tissue-specific regulation of retinal and pituitary precursor cell proliferation. Science 297: 1180–1183.