Spatial Epidemiology of Recently Acquired HIV Infections across Rural and Urban Areas of North Carolina

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Abstract

Transmission of HIV continues in the United States (US), despite prevention efforts aimed at education and treatment. Concurrently, drug resistance in HIV, particularly in patients being infected with HIV for the first time, poses a threat to the continued success of treatment for HIV positive individuals. In North Carolina, nearly one in five individuals with acute HIV infection (AHI) is infected with a drug-resistant strain, a phenomenon known as transmitted drug resistance (TDR). Few studies of AHI or TDR take into account both the spatial aspects of residence at time of infection and the genetic characteristics of the viruses, and questions remain about how viruses are transmitted across space and the rural-urban divide. Using AHI strains from North Carolina, we examined whether differences exist in the spatial patterns of AHI versus AHI with TDR, as well as whether the genetic characteristics of these HIV infections vary by rural-urban status and across Health Service Areas. The highest amounts of TDR were detected in persons under age 30, African Americans, and men who have sex with men (MSM) - similar to the populations where the highest numbers of AHI without TDR are observed. Nearly a quarter of patients reside in rural areas, and there are no significant differences between rural and urban residence among individuals infected with drug resistant or drug susceptible viruses. We observe similar levels of genetic distance between HIV found in rural and urban areas, indicating that viruses are shared across the rural-urban divide. Genetic differences are observed, however, across Health Service Areas, suggesting that local areas are sites of genetic differentiation in viruses being transmitted to newly infected individuals. These results indicate that future efforts to prevent HIV transmission need to be spatially targeted, focusing on local-level transmission in risky populations, in addition to statewide anti- HIV efforts.

Introduction

Despite decades of education and prevention efforts, HIV incidence remains relatively stable in the United States (US), at approximately 48,000 new infections each year [1]. However, new HIV infections are not evenly spatially distributed across the US; higher levels of new HIV infections are observed in the Southeast as compared to the rest of the country [1,2]. Additionally, certain demographic and behavioral risk groups, namely young black men who have sex with men (MSM), have high levels of new HIV acquisition [3–6].

Though incidence rates of HIV remain steady in the US, prevalence rates have risen, because people are living longer due in large part to antiretroviral (ART) therapy. Effective treatment for HIV has led to greatly decreased mortality and increased life expectancy for persons living with the virus in developed countries and resource-limited settings, worldwide [7,8]. As with other antimicrobials, widespread use of ART has led to the emergence of drug-resistant forms of the virus [9–11]. Unlike other pathogens, however, all drug resistance mutations that develop in HIV are “archived” as proviruses throughout the host genome. Thus, once resistance has developed, individual drugs or in some cases entire drug classes may not be useful again for treating the patient[12]. When individuals who are viremic with resistant HIV engage in unprotected sex or share needles, previously uninfected individuals can acquire ARV-resistant HIV, a phenomenon known as transmitted drug resistance (TDR)[13]. The TDR virus may also be passed from one individual to another in the absence of therapy. The prevalence of TDR has remained stable at approximately 10–20% of newly diagnosed individuals in North America[11,14–17] and Europe[18–21] and may be higher in acutely infected patients [22]. In the developing world, where treatment options are often limited, the prevalence of TDR appears to be increasing rapidly; 8 years since the widespread rollout of ARVs in east Africa, the prevalence of TDR is nearly 8% [23].

Despite the spatial variation in HIV infection across the US, most investigations of newly acquired HIV and evaluations of TDR versus drug-sensitive viruses in newly infected individuals typically do not take into account geographic location or utilize a
coarse spatial resolution such as county or ZIP code. With the collection of HIV genetic sequences from newly infected individuals and sociodemographic information for partner counseling and referral services, new opportunities have developed for the merger of genetics, geography and epidemiology. Spatial molecular epidemiology has the potential to illuminate patterns of HIV transmission, and of TDR variants in particular, indicating populations and places where surveillance and interventions might best be targeted [24–26]. In particular, spatial molecular epidemiology can answer questions about whether urban areas act as reservoirs for rural HIV infection, or whether circulating strains of HIV differ between rural and urban areas.

In the present study of acutely HIV-infected individuals in North Carolina (NC), we had three specific aims. First, we wished to determine the geographic and genetic distributions of new HIV cases across NC – including the subset with TDR. Second, we sought to understand whether genetic relatedness among all viruses and viruses from specific epidemiologic subgroups was associated with residential location characteristics, such as urban or rural status and health service region, as well as the geographic distance between individuals. Finally, we wanted to examine whether TDR viruses exhibited different spatial and genetic patterns than did non-TDR, drug-susceptible (DS) viruses.

**Data and Methods**

**Ethics Statement**

All participants in the Duke/UNC Acute HIV Consortium Database project provided informed consent for their de-identified sociodemographic, immunological, and virological information to be used for research purposes. The Institutional Review Boards at the University of North Carolina at Chapel Hill and the University of Iowa reviewed and approved the protocol of the study described herein.

**Study population**

All subjects included in this study were diagnosed with seronegative acute HIV infection (AHI) between 1998–2009 and enrolled in the Duke/UNC Acute HIV Consortium Database project [27]. Detailed descriptions of the case definition and database have been published previously [28–30]. In brief, subjects in the database were referred to one of the participating institutions either by community medical providers or the Screening and Tracing Active Transmission (STAT) program of the NC Department of Health and Human Services (NC-DHHS). In STAT, any individual presenting to a publicly funded HIV or sexually transmitted infection testing site who has blood drawn for HIV antibody or syphilis testing is also tested for HIV nucleic acids. HIV RNA is detectable approximately 10–14 days earlier than antibodies against HIV, allowing identification of very early infections [31]. RNA-positive, antibody-negative patients are referred for further evaluation; a majority provide consent for their data to be included in the Consortium database.

**Genotypic resistance testing**

From the earliest possible plasma sample for each subject, the pol region of the HIV-1 genome was sequenced using primers spanning all of the protease and the majority of the reverse transcriptase gene (from codons 1-100 and 38-250, respectively). GenoSure (Laboratory Corporation of America, Research Triangle Park, NC, USA) or Trugene (Siemens Healthcare Diagnostics, Tarrytown, NY, USA) primers were used for all sequencing analyses. Raw nucleotide sequence data were analyzed with the Stanford University HIV Drug Resistance Database (http://hivdb.stanford.edu) to characterize ARV-resistant viruses; relevant mutations were defined by the 2009 World Health Organization list of surveillance drug resistance mutations (SDRMs) [10].

**Phylogenetic analyses**

Sequences were aligned using Multiple Sequence Comparison by Log-Expectation (MUSCLE) [32], then edited manually using Se-Al v2.0a11 [Andrew Rambaut, University of Edinburgh]. The final data included codon positions 4-90 of protease and 38-240 of reverse transcriptase, based on numbering from an HXB2 consensus sequence (NCBI Reference Sequence NC_001802.1). A maximum likelihood (ML) tree was inferred in RAxML v7.2.0 [33], under a generalized time-reversible (GTR) model of nucleotide substitution with 1000 bootstrapped replicates. A matrix of genetic distances separating all pairs of taxa from the solved, consensus ML tree were extracted from a Newick tree file using PATRISTIC [34], with results exported as a comma-separated file for manipulation and analysis.

**Geocoding**

NC-DHHS Communicable Disease Branch personnel conducted a special query to retrieve addresses for each AHI case at the time of diagnosis. Those addresses were geocoded (assigned latitude and longitude), and this coordinate location was then matched to the 2000 Census block group (CBG) in which it was located. The centroid of the CBG was then calculated and the latitude/longitude of this anonymized point was assigned to each AHI case. Individual addresses were then deleted from the database. The CBG is the smallest unit for which demographic data collected through the Census are publicly available; each contains approximately 600–3000 individuals, and they vary in geographic size. Once the assignment to a CBG was completed, all individual address and geocoded information was deleted. This geocoding and linkage to CBG of residence at time of infection was done prior to linking to clinical and virological information from the Consortium database. Because only the CBG number and the coordinates of its centroid were recorded in the final data set, patient privacy was maintained at all times.

Cases were mapped according to their CBG of incidence and stratified by demographic and behavioral variables and drug resistance status. To investigate the potential association between residential location and genetic relatedness of viruses, cases were classified as either rural or urban according to the designations created in the 2000 Census, with 17 urbanized areas and 90 urban clusters defined in NC. Urbanized areas and urban clusters were collapsed into one “urban” category for the purposes of this study. Any case who’s CBG fell outside the bounds of an urban area were classified as rural. To further investigate potential spatial differentiation in genetic patterns of viruses, cases were assigned to the Health Service Area (HSA) of residential location at time of diagnosis. There are six HSAs in North Carolina, each of which represents a set of contiguous counties that are used in the planning of health care provision across the state.

To complement the pair-wise genetic distance matrix, a pair-wise geographic distance matrix was generated, indicating the distance in kilometers between each AHI case in the dataset.

**Statistical Methods**

Characteristics for all AHI patients, stratified by TDR status, were summarized using descriptive statistics. We assessed the relationship between demographic and behavioral characteristics and the presence of TDR using Pearson’s χ² test. Exact probability values were calculated, given the overall low numbers of observations within categories. Pearson’s χ² test was also used to
measure differences in rural/urban categorization by demographic and behavioral characteristics, as well as HSA of residence. Statistical significance was set at $P < 0.05$ for all analyses.

Scatterplots of pair-wise genetic versus geographic distances were generated, stratifying observations by TDR status, as well as HSA and rural/urban status. Pair-wise genetic distances were also stratified according to whether the pair was a rural-rural set, a rural-urban set or an urban-urban set, in order to understand differences in genetic relationships according to urbanicity. The same was done with HSA location of incidence, to explore inter-HSA and intra-HSA differences in genetic distances among viruses.

All geocoding and mapping was conducted in ArcGIS 10.0 (Esri, Redlands, CA). Chi-square tests were calculated in SAS 9.3 (SAS Institute, Inc., Cary, NC). Plotting was conducted in R using the ecodist package (R Foundation for Statistical Computing, Vienna, Austria). Median genetic values were calculated for lower triangle matrices using R and the difference in median genetic values were assessed using Wilcoxon tests for bivariate groupings and Kruskal-Wallis tests for trivariate groupings, also in R.

**Results**

We successfully geocoded 81 of 143 AHI cases identified between 1998–2009 (57%); 17 (21%) had ≥1 mutation indicating TDR. Three AHI cases without TDR and 4 cases with TDR lacked demographic and behavioral information. Among the remaining 74 cases, the median age of patients with drug-susceptible (DS) virus was 26 years (interquartile range [IQR], 21–36; Table 1), while the median age of TDR cases was 23 (IQR, 21–27). Ninety-three percent of all DS AHI cases were among men (n = 57), with a similar proportion observed among subjects with TDR (85%, n = 11). The majority of AHI cases were Black men, 52% with DS virus and 46% with TDR. Of those Black men, the majority (74% of DS and 83% of TDR) were MSM. These proportions reflect the disproportionate burden of HIV in the Southeastern US[35]– especially among young Black MSM[5].

No statistically significant differences with respect to age, sex, race/ethnicity, mode of acquisition or rural/urban residence were observed between individuals with TDR and those with DS viruses (Table 1). There was a trend toward variation in TDR status by HSA ($P = 0.08$).

| Drug Susceptible Cases (n = 61) | TDR Cases (n = 13) | Chi-square | Exact p-value |
|--------------------------------|-------------------|------------|---------------|
| **Age** | **Age** | | |
| Median (IQR) | Median (IQR) | 7.586 | 0.101 |
| **Sex** | **Sex** | 1.121 | 0.582 |
| Male | Male | 57 (93.4%) | 11 (84.6%) |
| Female | Female | 4 (6.6%) | 2 (15.4%) |
| **Race/Ethnicity** | **Race/Ethnicity** | 9.594 | 0.081 |
| White | White | 21 (34.4%) | 4 (30.8%) |
| Black | Black | 34 (55.7%) | 7 (53.8%) |
| Hispanic | Hispanic | 6 (9.9%) | 1 (7.7%) |
| Asian/Pacific Islander | Asian/Pacific Islander | 1 (7.7%) | |
| **Mode** | **Mode** | 1.107 | 0.879 |
| MSM | MSM | 42 (68.9%) | 9 (69.2%) |
| Heterosexual | Heterosexual | 14 (23%) | 4 (30.8%) |
| **Location** | **Location** | 0.574 | 0.542 |
| Urban | Urban | 47 (73.4%) | 14 (82.4%) |
| Rural | Rural | 17 (26.6%) | 3 (17.6%) |
| **HSA** | **HSA** | 9.594 | 0.081 |
| 1 | 1 | 6 (9.4%) | 0 (0%) |
| 2 | 2 | 7 (10.9%) | 6 (35.3%) |
| 3 | 3 | 5 (7.8%) | 2 (11.8%) |
| 4 | 4 | 26 (40.6%) | 7 (41.1%) |
| 5 | 5 | 10 (15.6%) | 2 (11.8%) |
| 6 | 6 | 10 (15.6%) | 0 (0%) |

Note that three drug susceptible and four drug resistant cases had no demographic and behavioral information available.

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Fifty-four individuals live in urban areas and 20 in rural areas. We observed no significant differences in the distribution of urban versus rural-dwelling patients, when categorized by race/ethnicity or mode of acquisition (MSM versus heterosexually acquired; Table 2). However, when we categorized patients by their HSA of residence, there was a statistically significant difference in rural/urban status; larger proportions lived in urban settings in all HSAs except HSA 6, which encompasses most of the northeastern portion of the state ($P = 0.008$; Figure 1B).

Maps of AHI and TDR cases reveal several noteworthy patterns. First, though the distribution of cases reflects major population centers in NC, not all cases were exclusive to urban areas (Figure 1). This is especially true for the rural and economically disadvantaged eastern portion of the state (HSA 6) and the southern coastal plain (HSA 5). Second, cases among Whites and Latinos conformed to what would be expected statistically based on subpopulation density, but Black subjects were more evenly distributed geographically, including the western mountainous region where far fewer Black North Carolinians reside (Figure 2). Cases among women seemed more concentrated in the central (Piedmont) region and the mountain west, while male cases were widespread across the entire state (Figure 3A & 3B). Finally, AHI cases among MSM were observed in both urban centers and more rural areas, widely distributed across NC. In contrast, cases of AHI among individuals reporting heterosexual sex as their HIV risk behavior were more often located in or around the central Piedmont area (Figure 3C & 3D).

Viruses from rural dwellers were more closely related to one another than they were to viruses from urban areas. Scatterplots of pairwise genetic distance versus pairwise geographic distance between cases, categorized according to rural or urban residence, indicate that viruses sampled from rural residents were separated by smaller pairwise genetic distances, even across geographic space, than viruses found in urban residents (urban-urban, median = 0.201; Figure 4A & 4B). The same was true when we examined pairings of rural viruses with urban ones; the genetic distance between pairs of rural viruses (median = 0.183) was smaller than the distance separating rural viruses from urban ones (rural-urban, median = 0.193).

In a similar analysis of DS viruses and those with TDR, clustering was observed among TDR viruses at lower geographic distances, indicating that DS viruses were more widely distributed across NC. The pattern was especially notable in the rural areas of HSA 6, where AHI cases were less prevalent (Figure 3D).

Figure 1. Spatial distribution of AHI and TDR positive census block groups (A) and Census-designated urban areas and major urban locations in North Carolina, with HSA boundaries (B).

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across the state (Figure 5A). Despite the smaller geographic range among TDR viruses, the pairwise genetic distances of TDR and DS viruses share the same general pattern over geographic space, fluctuating around 0.20.

When the various pairwise genetic distances separating TDR viruses from one another and DS viruses from one another were stratified according to residence at the time of diagnosis (rural versus urban), a different pattern emerged (Figure 5B). Among DS viruses and among TDR viruses, the range of genetic distances for urban-urban pairings (DS median = 0.201, TDR median = 0.205) was similar; Wilcoxon tests indicated no statistically significant variation between TDR and DS genetic distances among urban-urban (p = 0.37) case pairs. In contrast, genetic distances among rural-urban case pairs were significantly different for TDR versus DS viruses (median = 0.219 & 0.188 respectively, Wilcoxon p < 0.01). TDR viruses identified in rural residents were not genetically similar (median genetic distance = 0.235) and had greater genetic distance than did DS rural-rural viruses (median = 0.176). Wilcoxon tests indicated a statistically significant difference in TDR versus DS viruses in rural-rural case pairs (p = 0.008).

We then assessed the genetic relatedness between pairs of viruses from young, Black MSM, all other MSM, and heterosexuals, categorized into rural-rural, rural-urban, and urban-urban groupings (Figure 6). Young, Black MSM had narrower ranges of pairwise genetic distances than all other MSM and heterosexuals, across rural-urban categories – but young, Black MSM viruses also exhibited the highest degree of genetic distance. The median

| Race/Ethnicity   | Urban | Rural | Chi-square | Exact p-value |
|------------------|-------|-------|------------|---------------|
| White            | 19    | 6     | 4.1328     | 0.263         |
| Black            | 27    | 14    | 0.663      | 0.533         |
| Hispanic         | 7     | 0     | 0.375      | 0.738         |
| Asian            | 1     | 0     | 0.977      | 0.48          |

Table 2. Urban/rural status of cases by race/ethnicity, risk groups and HSA designations.

| Mode           | Urban | Rural | Chi-square | Exact p-value |
|----------------|-------|-------|------------|---------------|
| MSM            | 39    | 12    | 0.663      | 0.533         |
| Black MSM      | 21    | 7     | 0.375      | 0.738         |
| Young Black MSM| 20    | 5     | 0.977      | 0.48          |
| Heterosexual   | 12    | 6     | 15.299     | 0.008         |

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pairwise genetic distances for young, Black MSM were 0.215, 0.215 and 0.221 for rural-rural, rural-urban, and urban-urban categories respectively, which were higher than the values for all other MSM (0.179, 0.192 and 0.197; Kruskal-Wallis P < 0.01) and heterosexuals (0.174, 0.188, and 0.194; Kruskal-Wallis P < 0.01).

Viruses from individuals living in the same HSA had the same general distribution of pairwise genetic distances as did persons living in different HSAs, often hundreds of kilometers away from one another (Figure 7). The level of genetic dissimilarity is fairly constant across the state, although the genetic distance declined slightly as the geographic distance separating the patients increased. Plotting intra-HSA genetic variation by individual HSAs of incidence (i.e. HSA 1 versus HSA 1 viruses) revealed HSA-specific genetic patterns (Figure 8). The majority of viruses in the dataset were found in individuals residing in HSA 4 (Figure 8B & Table 1), shown in green. These viruses exhibited a narrower geographic range than did viruses in other HSAs; HSA 4 is one of the smaller HSAs in the state. Despite the smaller geographic range, there is a high level of genetic distance between viruses in HSA 4. In contrast, HSA 6 viruses are widely spaced geographically but exhibit low inter-virus genetic distance.

To explore whether these differences in genetics by HSA of residence were driven primarily by rural or urban residential location, intra-HSA genetic distances between viruses were categorized by rural-rural, rural-urban and urban-urban pairings (Figure 9). Urban-urban genetic relatedness across all HSAs varied; some viruses were closely related and other had a high degree of genetic distance. HSAs 2 and 5 had only one rural sample, so had no rural-rural pairs. Rural-rural viruses in HSA 4 have high genetic distance, as high as do rural-urban and urban-urban virus pairs. In contrast, rural-rural viruses in HSA 6 have much lower genetic distances than do rural-urban and urban-urban viruses in that region. Kruskal-Wallis tests indicated significant variation in genetic distances across rural-rural, rural-

Figure 3. Distribution of AHI and TDR cases by sex (A: men, B: women) and by mode of acquisition (C: men who have sex with men (MSM), D: heterosexual intercourse).

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Figure 4. Exploring Rural/Urban genetic variation. (A) Pair-wise genetic versus geographic distance for cases, classified as Rural-Urban, Urban-Urban or Rural-Rural based on CBG of patient at time of infection. (B) Genetic distances for all viruses, stratified by Rural/Urbun pair relationship with median value indicated by a bar. R-R indicates both cases are from rural areas, R-U indicates one rural and one urban case, and U-U indicates both cases are from urban areas.

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urban and urban-urban case pairs within HSAs 2 (p < 0.001), 4 (p < 0.001) and 6 (p < 0.001). Genetic distances across these residential designations were not significantly different in HSAs 1 (p = 0.067), 3 (p = 0.562) and 5 (p = 0.225).

**Discussion**

This is the first study of the spatial epidemiology of recently acquired HIV infection in the United States. We find some evidence for geographic restriction of DS and TDR viruses in rural versus urban areas. As in the rest of the United States, North Carolina’s HIV epidemic is principally among young, Black MSM. There were no significant differences in individual-level characteristics between rural and urban residents, suggesting that the epidemic is impacting this key risk group uniformly across the state, regardless of where these men live. Furthermore, nearly one quarter of the individuals in the sample lived in rural areas, highlighting the importance of maintaining access to HIV testing and treatment resources in less urbanized areas of the state.

The pairwise genetic distances separating viruses from rural dwellers were smaller than those separating viruses from urbanites,

Figure 5. Exploring TDR versus non-TDR genetic and geographic variation. (A) Pair-wise genetic versus geographic distance for TDR and non-TDR viruses, and (B) TDR and non-TDR genetic distances by Rural/Urbam pair relationship with median value indicated by a bar. doi:10.1371/journal.pone.0088512.g005

Figure 6. Viral pair-wise genetic distances among young, black MSM risk patients, among other MSM patients, and among heterosexual risk patients, stratified by Rural/Urbam pair relationship. Median genetic distance for each group is indicated by a bar. doi:10.1371/journal.pone.0088512.g006
and these distances decreased slightly as geographic distance between patients widened. This suggests that, to some degree, there are separate sub-epidemics occurring simultaneously in the state, with transmissions occurring with some viruses among rural residents and other, genetically unique viruses among urban dwellers. The best evidence for this phenomenon came from our analysis of viruses from each of the six HSAs across the state. In HSAs 1 (mountains), 3 (Charlotte metropolitan area), 4 (Raleigh-Durham), and 5 (Fayetteville to Wilmington), we observed pairwise genetic distances toward the lower end of the range within urban-urban pairs, while HSA 6, the rural and economically depressed Eastern region of the state, had the lowest genetic distances separating rural-rural pairs. In fact, the median distance among these rural pairs in HSA 6 was the lowest across all rural/urban categories in all HSAs, potentially indicating a tighter network of transmission in which viruses were highly genetically related. Further investigation with a larger sample from HSA 6 would allow this to be more definitively assessed.
With respect to TDR versus DS viruses, our analyses indicated that HIV isolates with primary resistance seemed to be more geographically restricted, and were more centered in urban areas. Though the presence or absence of drug resistance was not associated with any individual-level characteristics, this urban/rural dichotomy suggests that transmission of resistant HIV may be more likely in urban areas, which historically have been home to more HIV treatment and care resources. In the early days of the epidemic, many patients relocated to the areas closest to treatment centers, to facilitate frequent trips to providers for clinical care. It is therefore plausible that focally greater densities of patients with HIV treatment experience might sustain more frequent detection of TDR among newly diagnosed patients.

In understanding the potential for urban areas to act as reservoirs of infection for rural areas, several findings suggest mixing of viruses between these categories across the state and a close relationship between urban and rural HIV genetics. Similar genetic distance distributions for intra-HSA and inter-HSA case pairs suggest that there is inter-regional mixing of viruses across the state, and the fact that viruses found in individuals residing far apart from one another are genetically similar suggests that spatial distance is not a barrier to genetic similarity. Similar findings have been observed in Mississippi, where viruses clustered genetically were not clustered geographically [25]. Urban areas of North Carolina have high degrees of genetic distance at small spatial scales, typically higher than the genetic distance found in rural areas or across the rural-urban divided, as seen in Figures 6 & 9. These higher amounts of genetic differentiation could indicate either higher levels of transmission, resulting in greater amounts of genetic variation, or higher levels of introduction of new viral variants into urban areas from other sexual networks. Uniquely in the dataset, however, HSA 4 had genetic distances in rural-rural virus pairs that were higher than the levels of genetic distances in rural-urban and urban-urban virus pairs in the same regions, suggesting either that rural residents of this HSA are acquiring a variety of viruses from urban areas within the region or that there is high genetic diversity in viruses circulating in rural areas. In contrast, viruses found in HSA 6 rural resident case pairs had low amounts of genetic distance. This again highlights the possibility that rural residents in HSA 6 are all acquiring infection in the same places or from the same sexual networks, resulting in high genetic similarity.

Perhaps the most important limitation of our study is the sample size. While small numbers of observations are not uncommon in genetic studies of sensitive diseases, caution is still warranted in generalizing our results to larger populations or different geographic settings. For instance, compared to chronically infected patients entering our HIV clinical cohort based in central North Carolina, the recently infected individuals studied here are younger and more likely to be black and endorse sex with men[36]. Additionally, a variety of approaches exist for reconstruction of phylogenetic trees – and thus the estimated genetic distances between taxa may vary somewhat depending on the model of nucleotide substitution and the computational method used.

The merging of spatial analysis, phylogenetic methods and epidemiology holds potential for understanding how and why infectious diseases evolve over space and time. This capability, however, is often hampered by a lack of spatial attributes collected for places of infection. Additionally, for highly sensitive diseases such as HIV, the ability to access datasets containing such information is frequently limited because of privacy concerns.
Understanding how HIV is shared between urban and rural populations, particularly among young black MSM, the predominant group in the study, is crucial to our ability to limit new HIV infections.

**Author Contributions**

Conceived and designed the experiments: MC ME JE CBH. Performed the experiments: MC CBH. Analyzed the data: MC CBH. Contributed reagents/materials/analysis tools: JE CBH. Wrote the paper: MC ME JE CBH.

**References**

1. Centers for Disease Control (CDC) (2013) HIV surveillance report, 2011.
2. Southern HIV/AIDS Strategy Initiative (2012) SASI update: the continuing crisis in the US south.
3. Aral SO, O’Leary A, Baker C (2006) Sexually transmitted infections and HIV in the southern united states: An overview. Sex Transm Dis 33(7): S1-S3.
4. Lau Z, Lemoxy M, Hall H, Hu X, Guo X, et al. (2012) A county-level examination of the relationship between HIV and social determinants of health: 40 states, 2006-2008. The Open AIDS Journal 6: 1.
5. Lieb S, Perjean J, Thompson DR, Fallon SJ, Cooper H, et al. (2011) HIV prevalence rates among men who have sex with men in the southern united states: Population-based estimates by race/ethnicity. AIDS and Behavior 15(5): 596-606.
6. Perjean J, Song R, Hernandez A, Ziebell R, Green T, et al. (2011) Estimated HIV incidence in the united states, 2006-2009. PLoS One 6(8): e17502.
7. Harrison KMD, Song R, Zhang X (2010) Life expectancy after HIV diagnosis based on national HIV surveillance data from 23 states, united states. JAIDS J Acquired Immune Defic Syndromes 53(1): 124.
8. Collaboration HIVC, Ray M, Logan R, Sterne J, Hernandez-Diaz S, et al. (2010) The effect of combined antiretroviral therapy on the overall mortality of HIV-infected individuals. AIDS 24(12): 123.
9. Boden D, Hurley A, Zhang L, Cao Y, Guo Y, et al. (1999) HIV-1 drug resistance in newly infected individuals. JAMA: The Journal of the American Medical Association 282(12): 1135-1141.
10. Bennett DE, Camacho RJ, Otelea D, Kuritzkes DR, Fleury H, et al. (2009) Drug resistance in newly infected individuals from 40 united states cities. HIV Clinical Trials 8(1): 1-8.
11. Little SJ, Holte S, Routy JP, Daar ES, Markowitz M, et al. (2002) Antiretroviral drug resistance among patients recently infected with HIV. N Engl J Med 347(6): 385-394.
12. Panel on Antiretroviral Guidelines for Adults and Adolescents (2013) Guidelines for the use of antiretroviral agents in HIV-1 infected adults and adolescents. Department of Health and Human Services. : 1-128.
13. Soeters HM, Napravnik S, Zakharaov OM, Eron JJ, Hurt CB. (2013) Opportunities for sexual transmission of antiretroviral drug resistance among HIV-infected patients in care. AIDS 27: 000-000.
14. Grant RM, Hecht FM, Warmerdam M, Liu L, Liegler T, et al. (2002) Time trends in primary HIV-1 drug resistance among recently infected persons. JAMA: The Journal of the American Medical Association 288(2): 181-189.
15. Weinstock HS, Zaidi I, Heneine W, Bennett D, Garcia-Lerma GJ, et al. (2004) The epidemiology of antiretroviral drug resistance among drug-naive HIV-1-infected persons in 10 US cities. J Infect Dis 189(12): 2174-2180.
16. Ross L, Lai ML, Liao Q, Wine B, Rodriguez AE, et al. (2007) Prevalence of antiretroviral drug resistance and resistance-associated mutations in antiretroviral therapy-naive HIV-infected individuals from 40 united states cities. HIV Clinical Trials 8(1): 1-8.
17. Hurt CB, McCoy SL, Kurke J, Nelson J, Jerkum M, et al. (2009) Transmitted antiretroviral drug resistance among acute and recent HIV infections in north carolina, 1990 to 2007. Antivir Ther (Lond) 14(5): 673.
18. Chan P, Chyatte I, Dunn D, Evans B, Geretti A, et al. (2005) Time trends in primary resistance to HIV drugs in the united kingdom: Multicentre observational study. BMJ 331(7529): 1368-1369.
19. Masquelier B, Bhakaran K, Pillay D, Gifford R, Balestre E, et al. (2005) Prevalence of transmitted HIV-1 drug resistance and the role of resistance algorithms: Data from seroconverters in the CASCADE collaboration from 1987 to 2003. JAIDS J Acquired Immune Defic Syndromes 40(3): 505-511.
20. Wensing AM, van de Vijver, DA, Angarano G, A˚sjo¨ B, Balotta C, et al. (2005) Prevalence of drug-resistant HIV-1 variants in untreated individuals in europe: Implications for clinical management. J Infect Dis 192(6): 936-966.
21. Vercauteren J, Wensing AM, van de Vijver, David AMC, Albert J, et al. (2009) Transmission of drug-resistant HIV-1 is stabilizing in europe. J Infect Dis 200(10): 1503-1508.
22. Yanik EL, Napravnik S, Hurt CB, Dennis A, Quinlivan EB, et al. (2012) Prevalence of transmitted antiretroviral drug resistance differs between acutely and chronically HIV-infected patients. JAIDS J Acquired Immune Defic Syndromes 61(2): 258-262.
23. Gupta RK, Jordan MR, Sultan BJ, Hill A, Davis DH, et al. (2012) Global trends in antiretroviral resistance in treatment-naive individuals with HIV after rollout of antiretroviral treatment in resource-limited settings: A global collaborative study and meta-regression analysis. The Lancet.
24. Dennis AM, Hsu S, Hurt CB, Napravnik S, Sebastian J, et al. (2012) Phylogenetic insights into regional HIV transmission. AIDS 26(14): 1813-1822.
25. Oster AM, PennaMD, Ziebell X, Ziebell RA, et al. (2011) Demographic but not geographic insularity in HIV transmission among young black MSM. AIDS 25(17): 2157-2165.
26. Hurt CB, Dennis AM (2013) Putting it all together: Lessons from the jackson HIV outbreak investigation. Sex Transm Dis 40(3): 213-215.
27. McKellar MS, Cope AB, Gay CL, McGee KS, Kurucz JD, et al. (2013) Acute HIV-1 infection in the southeastern united states: A cohort study. AIDS Res Hum Retroviruses 29(1): 121-128.
28. Pichler CD, Shugas DC, Fiscus SA, Miller WC, Menezes P, et al. (2001) HIV in body fluids during primary HIV infection: Implications for pathogenesis, treatment and public health. AIDS 15(7): 837-845.
29. Pichler CD, Fiscus SA, Nguyen TQ, Foutz E, Wolf L, et al. (2005) Detection of acute infections during HIV testing in north carolina. N Engl J Med 352(18): 1873-1883.
30. Pichler CD, McPherson JT, Leone PA, Smurzynski M, Owen-O’Dowd J, et al. (2002) Real-time, universal screening for acute HIV infection in a routine HIV counseling and testing population. JAMA: The Journal of the American Medical Association 288(2): 216-221.
31. Colon MS, Shaw GM, McMichael AJ, Haynes BF (2011) Acute HIV-1 infection in the southeastern united states: Population-based estimates by race/ethnicity. AIDS and Behavior 15(3): 596-606.
32. Edgar RC (2004) MUSCLE: Multiple sequence alignment with high accuracy and high throughput. Nucleic Acids Res 32(3): 1792-1797.
33. Stamatakis A (2006) RAxML-VI-HPC: Maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. Bioinformatics 22(21): 2684-2680.
34. Fourment M, Gibbs M (2006) PATRISTIC: A program for calculating patristic distances and graphically comparing the components of genetic change. BMC Evolutionary Biology 6(1).
35. Perjean J, Tang T, Hall HI (2012) HIV diagnoses and prevalence in the southern region of the united states, 2007-2010. J Community Health : 1-13.
36. Dennis AM, Napravnik S, Senta AC, Eron JJ (2011) Late entry to HIV care among latinos compared with non-latinos in a southeastern US cohort. Clinical Infectious Diseases 53(5): 480-487.