Review

Hemp Pest Spectrum and Potential Relationship between Helicoverpa zea Infestation and Hemp Production in the United States in the Face of Climate Change

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Simple Summary: Cultivation of industrial hemp Cannabis sativa in the United States is now being expanded due to the recent legalization of the crop. Multiple insect pests attack the crop. One of the common pests is the corn earworm Helicoverpa zea that causes extensive damage to the marketable parts of hemp. Changing global climate may lead to expansion of the geographic range of insect pests. Thus, growers of this crop in the United States have to face new and intense pest problems now and in the years to come. Here, we assess the potential relationship between corn earworm infestation and hemp production in the US in the face of climate change. We also provide an update on the arthropods associated with hemp cultivation across the US. Climate change can affect aspects of interactions between hemp and corn earworm. Temperature and photoperiod affect the development and diapause process in H. zea. Drought leads to a reduction in hemp growth. Overall, our assessment suggests the selection of varieties resistant to stresses from climate and insects. Host plant diversity may prevent populations of corn earworm from reaching outbreak levels. Ongoing research on effective management of H. zea on hemp is critical.

Abstract: There has been a resurgence in the cultivation of industrial hemp, Cannabis sativa L., in the United States since its recent legalization. This may facilitate increased populations of arthropods associated with the plant. Hemp pests target highly marketable parts of the plant, such as flowers, stalks, and leaves, which ultimately results in a decline in the quality. Industrial hemp can be used for several purposes including production of fiber, grain, and cannabidiol. Thus, proper management of pests is essential to achieve a substantial yield of hemp in the face of climate change. In this review, we provide updates on various arthropods associated with industrial hemp in the United States and examine the potential impact of climate change on corn earworm (CEW) Helicoverpa zea Boddie, a major hemp pest. For example, temperature and photoperiod affect the development and diapause process in CEW. Additionally, drought can lead to a reduction in hemp growth. Host plant diversity of CEW may prevent populations of the pest from reaching outbreak levels. It is suggested that hemp varieties resistant to drought, high soil salinity, cold, heat, humidity, and common pests and diseases should be selected. Ongoing research on effective management of CEW in hemp is critical.

Keywords: industrial hemp; Cannabis sativa; climate change; pests; beneficials; corn earworm

1. Introduction

Industrial hemp or hemp (Cannabis sativa L.) cultivation is assuming new geographical borders around the world [1–3]. It is of medicinal, industrial, and economic importance. It is usually cultivated for production of long and strong bast fibers, seeds, oil, and food (Figure 1) [4]. Hemp contains extremely low amounts of the psychoactive cannabinoid Δ 9-tetrahydrocannabinol (THC). Cannabis plants that contain a concentration

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of less than 0.3% THC are considered hemp; while those above this concentration are considered marijuana [5]. Here, we regard industrial hemp as hemp varieties with amounts of less than 0.3% THC.

The legality of hemp cultivation varies worldwide. A unique aspect of hemp history in the US is the ban of its cultivation in 1937 when the federal Marihuana Tax Act effectively criminalized almost all cannabis cultivation [6]. The Agricultural Act of 2014 (also called the 2014 Farm Bill) reintroduced industrial hemp production through state pilot programs [7]. In 2018, the Agriculture Improvement Act, also known as the 2018 Farm Bill, re-legalized commercial hemp production in the US [8]. These decades of prohibition of cultivation resulted in little to no research on hemp in the US. Since 2014, however, legalization of industrial hemp in the US has resulted in increased interest in the cultivation of the crop (Figure 2) [7,9]. Despite this increase, the hemp industry is still regarded as emerging; and there is a lack of established production methods around the country [10]. This has resulted in many producers modifying and experimenting with hemp production. Presently, there are efforts around North America to develop improved cultivars for production of one or more of the commodities derived from industrial hemp [4,11,12]. Some varieties are being developed for improvement of their CBD contents, fiber production, and grain content [7]. Stakeholders in the US want breeding and genetics research to produce stable and uniform cultivars and regional adaptability [13].

**Figure 1.** Flowchart of multi-purpose hemp utilization. Graphic is from Salentijn et al. [4].
Figure 2. United States hemp acreage and greenhouse area, reported 2014–2018. Graphic is from Tyler et al. [7].

Growing conditions for hemp cultivation are documented in the literature [14]; however, optimal growing conditions are expected to vary according to cultivar. An important aspect of hemp cultivation is the management of arthropod pests. As with any crop, successful cultivation of hemp can include integrated pest management strategies. Efficient management of arthropod pests on hemp starts with surveying and properly identifying its insect community. There are reports of arthropods associated with hemp globally [15] and within the US [12]. However, with the current expansion in the cultivation of hemp across several states, reports/knowledge of arthropod pests needs to be updated. Furthermore, surveys have suggested negative grower experiences with hemp production especially from first-time or inexperienced hemp growers [10]. This highlights the need to educate growers on arthropod communities and pest control on hemp.

One of the most important challenges facing agriculture worldwide is management of abiotic stressors, including increasing temperatures and prolonged periods of drought. Such climatic anomalies are expected to drive the spread of arthropods [16,17], including those associated with hemp. In addition, biotic factors play a decisive role in species spatial distributions presently and will continue under future climate change [18]. The market value of industrial hemp in the US is impacted due to several pest insect species increasingly located on the crop [12]. Some authors have previously reviewed arthropods of hemp. Mostafa and Messenger [19] reported about 272 species of insect and mite species associated with Cannabis globally. McPartland et al. [15] described about 150 species of insects and mites associated with hemp. Cranshaw et al. [12] described several arthropod pests associated with the production of hemp and the associated pest management needs in the US. Here, we review and update the arthropods (both pests and beneficials) affecting industrial hemp across the US. Furthermore, we discuss how climate change could affect one of the prevalent pests.

2. Industrial Hemp Pests

Many phytophagous insects feed on industrial hemp, though only some species attain pest status [12,15]. Cranshaw et al. [12] arranged arthropods on hemp across the United States into the following categories—pests: defoliators, sucking insects and mites.
on leaves, stem and stalk borers, sucking insects associated with flowers and seeds, chewing insects that damage flower buds and seeds, and root feeders; natural enemy species: predators, parasitoids, pathogens; and pollinators.

Some pests and beneficial arthropods reported on hemp in the US are listed in Tables 1 and 2, respectively. Some arthropods that are currently considered as neither pest nor beneficial to hemp that exist in the US are listed in Table 3.

### Table 1. A list of some pest arthropods reported on hemp in the United States.

| Family           | Common Name                  | Scientific Name (for Those Identified to Species) | Damage Type                        | Location Found | References |
|------------------|------------------------------|--------------------------------------------------|------------------------------------|----------------|------------|
| Acrididae        | Grasshopper                  | Phorodon cannabis                                | Pest                               | Field          | [20]       |
| Aeolothripidae   | Thrips                       |                                                  | Pest                               | Field          | [20,21]    |
| Aphididae        | Cannabis aphid               |                                                  | Pest                               | Field          | [20–25]    |
| Cercopidae       | Spittlebug                   | Diabrotica undecimpecta; Diabrotica v. virgifera;| Herbaceous pest                    | Field          | [20,21,26] |
| Chrysomelidae    | Leaf beetle                  |                                                  |                                    |                |            |
| Cicadellidae     | Leafhoppers, e.g., Beet leafhopper | E.g., Circulifer tenellus | Pest (some transmits beet curly top virus) | Field          | [20,21,27,28] |
| Coreidae         | Leaf-footed bug              | Ostrinia nubilalis                               | Sucking-piercing pest              | Field          | [26]       |
| Crambidae        | European corn borer          |                                                  | Pest                               | Field          | [21]       |
| Curculionidae    | Weevil                       | Solenopsis invicta                               | Herbaceous pest                    | Field          | [20,26]    |
| Elateridae       | Click beetle                 |                                                  | Pest                               | Field          | [20]       |
| Formicidae       | Fire ant                     |                                                  | Pest                               | Field          | [20,23]    |
| Meloidae         | Blister beetle               |                                                  | Herbaceous pest                    | Field          | [26]       |
| Membracidae      | Treehopper                   |                                                  | Pest                               | Field          | [20]       |
| Miridae          | Tarnished plant bug          | Lygus lineolaris                                 | Sucking-piercing pest              | Field          | [20,26]    |
| Noctuidae        | Corn earworm                 | Helicoverpa zea                                  | Primarily, laceration of reproductive branch tip | Field          | [23,26,28–32] |
| Pentatomidae     | Stink bug                    |                                                  | Sucking-piercing pest              | Field          | [20,26,28] |
| Rhopalidae       | Hibiscus scentless plant bug | Niesthrea louisianaica                           | Sucking-piercing pest              | Field          | [26]       |
| Rhyparochromidae | Seed bug, Scarabs, e.g.,     |                                                  | Pest                               | Field          | [20]       |
| Scarabaeida      | Japanese beetle, Green June beetle | E.g., Popillia japonica | Herbaceous pest                    | Field          | [20,21,26] |
| Tarsonemidae     | Broad mites                  | Polyphagotarsonenetus latus                      | Pest                               | Greenhouse     | [28]       |
| Tetranychidae    | Two-spotted spider mite      | Tetranychus urticae                              | Pest                               | Greenhouse     | [29,31]    |
| Tortricidae      | borer (adults & larvae)      | Grapholita delineana                             | Pest                               | Field          | [20,23]    |
### Table 2. A list of some beneficial arthropods reported on hemp in the United States.

| Family         | Common Name (if any)                          | Scientific Name (for Those Identified to Species) | Association Type          | Location Found | References |
|----------------|-----------------------------------------------|-------------------------------------------------|---------------------------|----------------|------------|
| Anthocoridae   | Insidious flower bug                          | *Orius insidiosus*                              | Beneficial                | Field          | [20]       |
| Anthicidae     | Ant-like beetle                                |                                                 | Beneficial                | Field          | [20]       |
| Araneae        | Spiders                                       |                                                 | Natural enemy (predator)  | Field          | [20,33]    |
| Braconidae     | Braconids                                      | *Cardiochiles*                                   | Natural enemy (parasitoid) | Field          | [33]       |
| Carabidae      | Tiger beetles                                  |                                                 | Beneficial                | Field          | [20]       |
| Chrysopidae    | Green lacewing                                 |                                                 | Natural enemy (predator)  | Field          | [20,26]    |
| Coccinellidae  | Lady beetles [Hippodamia convergens; Coleomegilla maculata; Hyperaspis lugubris; Cycloneda munda; Cycloneda sanguinea; Harmonia axyridis] | Natural enemy (predator) | Field & greenhouse | [20,24,26] |
| Dolichopodidae | Long-legged flies                              |                                                 | Beneficial                | Field          | [20]       |
| Geocoridae     | Big-eyed bug                                   | *Geocoris*                                      | Natural enemy             | Field          | [20]       |
| Hemerobiidae   | Brown lacewings                                |                                                 | Beneficial                | Field          | [20]       |
| Ichneumonidae  | Ichneumonids                                   |                                                 | Natural enemy (parasitoid) | Field          | [33]       |
| Nabidae        | Damsel bugs                                    |                                                 | Beneficial                | Field          | [20]       |
| Pentatomidae   | Spined soldier bug                             | *Podisus maculiventris*                         | Natural enemy             | Field          | [26]       |
| Reduviidae     | Assassin bug                                   |                                                 | Beneficial                | Field          | [20]       |
| Syrphidae      | Syrphid larvae                                 |                                                 | Natural enemy (predator)  | Field          | [20,33]    |
| Tachinidae     | Tachinids                                      |                                                 | Natural enemy (parasitoid) | Field          | [33]       |
| Vespidae       | Paper wasps                                    |                                                 | Natural enemy (predator)  | Field          | [33]       |
| Opiliones (spider) |                                              |                                                 | Beneficial                | Field          | [20]       |

### Table 3. A list of some arthropods considered neither pest nor beneficial reported on hemp in the United States.

| Family         | Common Name (if any)                          | Association Type | Location Found | References |
|----------------|-----------------------------------------------|------------------|----------------|------------|
| Cerambidae     | Longhorn beetle                               | Other            | Field          | [20]       |
| Cleridae       | Checkered beetles                             | Other            | Field          | [20]       |
| Gryllidae      | Cricket                                       | Other            | Field          | [20]       |
| Latridiidae    | Minute brown scavenger beetles or fungus beetle | Other            | Field          | [20]       |
| Mordellidae    | Tumbling flower beetles                       | Other            | Field          | [20]       |
| Nitidulidae    | Sap beetle                                    | Other            | Field          | [20]       |
| Pieridae       | Pierid butterfly                              | Other            | Field          | [20]       |
| Silvanidae     | Silvan flat bark beetles                      | Other            | Field          | [20]       |
| Staphylinidae  | Rove beetle                                   | Other            | Field          | [20]       |
| Tipulidae      | Crane fly                                     | Other            | Field          | [20]       |
| Caddisflies (in the order Trichoptera) | Other | Field | [20] |
| Centipede (in the class Chilopoda) | Other | Field | [20] |
| Millipede (in the class Diplopoda) | Other | Field | [20] |
| Booklice, barklice or barkflies (in the order Pscoptera) | Leaf mining fly (larvae) | Other | Field | [20] |
|--------------------------------------------------------|-------------------------|-------|-------|------|

There are some challenges to sampling in hemp, therefore future directions should include some standardization of methods. For example, there are limitations in using just a visual count in comparison to sweep-net and beat-into-alcohol methods. Other methods of collection used are pitfall traps and yellow sticky cards. A factor that could strongly influence sampling in hemp is that neighboring crops to hemp can impact insect community. Furthermore, weather pattern can impact the insect types and the population densities.

3. Corn Earworm and Hemp: Potential Effects of Climate Change

Corn earworm *Helicoverpa zea* (Boddie, 1850) (Lepidoptera: Noctuidae) is native to the Americas [15]. *Helicoverpa zea* is very common on hemp plants across the United States (Table 1). It is a polyphagous, multivoltine insect pest and has a wide range of hosts, including many vegetables, field crops, fruits, flowers, and weeds. It causes serious damage in several crops, including corn, tomato, pepper, cotton, sorghum, and lettuce [34]. Around the world, it is called by a plethora of common names, including corn earworm, cotton bollworm, tobacco budworm, tobacco fruitworm, and vetchworm [15].

*Helicoverpa zea* overwinters as a pupa within the soil; in the US, successful overwintering occurs only in the southern USA, as *H. zea* cannot successfully overwinter at areas above 39°N [15,35]. Adults emerge from overwintering pupae in spring, and can then migrate throughout most of the United States and southern Canada during the growing season [36]. Adults mate and females lay eggs in floral inflorescences. A single female can lay up to 1500 eggs in her lifetime [37,38]. Emerged larvae feed on and injure hemp bud material, causing “bud rot” [39]. In North America, *H. zea* produces one to seven generations per year, depending on the latitude (e.g., one to two generations in Ontario, seven generations in south Texas) [40–44]. Like most multivoltine insects, its development and diapause termination are expected to be driven by temperature, while its diapause initiation triggered by photoperiod [45]. *Helicoverpa zea* pupae are chill-intolerant, thus they are unable to withstand freezing and are subject to much prefreeze mortality [46]. Sex does not influence the cold response of *H. zea* pupae [46,47]. Enhanced cold hardiness is gained through diapause in *H. zea* pupae [46–48]. It has been predicted that *H. zea* would respond to climate change by altering its voltinism [49].

Over the last decades, many studies on climate trends have been carried out and results demonstrate that patterns of temperature and precipitation are rapidly shifting, affecting large parts of our planet, in both animals and plants [50–54]. Some plant and animal species may react to climate change by showing some degree of adaptation and mitigation of its effects [55,56]. It is expected that over the coming decades, many plant and animal species will be affected in all aspects of their biology [57], and that adaptation to counterbalance impacts of climate change will be a challenge. Several aspects of insect biology can be affected by warmer temperatures, including survival and reproduction [58]. Furthermore, climate change can impact aspects of plant-insect interactions including host resistance and quality [59–62]. Natural enemies of pest insects can also be impacted by climate change [63–65]. These natural enemies, including parasitoids and predators, are dependent on the resilience of their host insects in the face of climate change, thus further exacerbating the stress on them [64].

In the US, climate change is a growing threat to biodiversity and ecosystems, and their services [66]. For example, in the Mid-Atlantic region of the US, where *Cannabis* hemp is widely cultivated, it is anticipated that by mid- to late-century (i.e., 2035–2049 and 2085–2099), there will be a warmer climate with a wetter Autumn and Spring and a drier late summer season; this is expected to cause damage to plants [67]. An increasing
number of studies show a link between drought and reduced hemp growth, including stem and fiber yield e.g., [68]. Furthermore, climate change is making the western US more arid [69]. This has contributed to drier soils [70], widespread plant death [71], and more severe wildfires [72]. Hemp is a tall plant with a wide root system (at least 0.5 m deep) and it is a good candidate for soil phytoremediation as it grows fast and easily in dense stands [73]. However, several environmental conditions such as drought, flooding, heat, and salinity affect the level of hormones in plants [68,74–78]. For example, under water stress, there is reduced transport of cytokinins from the root (the site of biosynthesis) to shoots [79,80]. This reduction in cytokinin is expected to bring about a shift towards malesness [78,81]. If this occurs in Cannabis plants, then it would ultimately influence the quantity and type of insects such as pollinators on hemp plants. Furthermore, it has been predicted that the US may experience warmer winters, resulting in diminished vernalization [82,83], a process required to promote flowering in certain types of crops. It is suggested that hemp varieties resistant to drought, high soil salinity, cold, heat, humidity, and common pests and diseases should be selected [84].

The mouthpart type of phytophagous insects influences their reaction to stress-induced plant changes [85]. For instance, decreased water content, tougher foliage, elevated levels of allelochemicals and reduced nitrogen availability all reduce nutritional quality of host plant tissue for chewing insects (e.g., corn earworm) [86]. Corn earworm herbivory increases the levels of chemical defense in cotton and has caused a significant decline in the nutritional quality of the plant as a host [87]. A similar severe decline in the nutritional quality of other plants such as soybean, geranium, and clover also occurred by corn earworm herbivory [88–91]. Recent studies have demonstrated that infestations of corn earworm on C. sativa increases the levels of cannabidiol (CBD) and delta-9-tetrahydrocannabinol (THC) beyond the 0.3% legal limit [92].

Some of the most prevalent and consistent pests of hemp are also major insect pests of corn (Zea mays L.). These include the European corn borer [Ostrinia nubilalis (Hübner)] and corn earworm [Helicoverpa zea (Boddie)]. Studies have shown that O. nubilalis antennae consistently responded to at least four chemical compounds, which all co-occur in both corn and hemp [93]. This suggests that these plant volatile compounds are cues used by these insects in herbivory. More research into the role of plant volatile compounds in the mechanisms of host location by these pests on hemp plants is needed.

Though outbreaks of H. zea have occurred in some regions of the United States where C. sativa is cultivated, host-plant diversity may prevent populations of H. zea from reaching outbreak levels [94]. Host plants of the generalist H. zea larvae include corn, tomato, and cotton, which are all economically important crops in the United States. H. zea is regarded as a serious pest of these crops making effective management of H. zea necessary. In the evaluation of biological insecticides to manage H. zea in CBD hemp in Virginia, Entrust SC (Spinosad) had a significantly lower incidence of bud rot than all other treatments [95]. Furthermore, no signs of phytotoxicity were observed from any of the biopesticide treatments in the study. In a similar study, Entrust (Spinosad) resulted in significantly fewer corn earworms and less damage than untreated control [96]. Furthermore, in another study, Entrust (Spinosad) resulted in a significantly higher corn earworm mortality (95%) than any other tested biological or organic insecticide products on field-collected corn earworms tested in laboratory assays after four days [97]. In a second bioassay, Pyganic and Entrust performed significantly better than all other treatments, resulting in 100% and 97.5% respective mortality in lab-reared corn earworms [97]. A likely reason for the difference between lab-reared and wild-caught populations could be that resistance to Cry1AB Bt proteins is widespread in Virginia corn earworms [97].

Stressed plants are expected to have reduced defenses and therefore greater vulnerability to herbivores (plant stress hypothesis; [98–100]). A similar pattern might be expected with the vulnerability of stressed C. sativa plants to herbivores such as H. zea. However, the plant vigor hypothesis contradicts this. For example, Inbar et al. [101] reported that larval growth rates of H. zea were higher on tomato (Lycopersicon esculentum) exposed
to optimal growing conditions, but lower on those exposed to stress. Whether stressed or not, morphological defense mechanisms of *C. sativa* may override the extent to which chewing herbivores such as *H. zea* damage the plant. For example, *H. zea* was negatively affected by trichome density on yellow monkey flower *Mimulus guttatus* [100]. This may be similar to the response of *H. zea* to stressed *C. sativa*.

4. Conclusions

*Helicoverpa zea* is a polyphagous insect pest on hemp, *Cannabis sativa*. In North America, the pest produces one to seven generations per year, depending on the latitude. Like most multivoltine insects, its development and diapause termination are expected to be driven by temperature, while its diapause initiation by photoperiod. *Helicoverpa zea* pupae are chill-intolerant, thus subject to much prefreeze mortality. *H. zea* could respond to climate change by altering its voltinism. Furthermore, climate change can affect aspects of interactions between hemp and corn earworm, including host resistance and quality. Natural enemies of corn earworm are dependent on the resilience of their host in the face of climate change. Water stress on hemp could bring about a shift towards maleness in the plant, and this could ultimately influence the quantity and type of insects such as pollinators on hemp. Drought leads to a reduction in hemp growth. Infestations of corn earworm on hemp increases the level of THC beyond the 0.3% threshold point at which cannabinoid content is used to distinguish strains of hemp from marijuana. Plant volatile compounds could be involved in cues used by *H. zea* in herbivory. Though outbreaks of corn earworm have been experienced in some regions of the United States where hemp is cultivated, host-plant diversity may prevent populations of corn earworm from reaching outbreak levels in regions with such diversity compared with those without. Ongoing research on effective management of *H. zea* on hemp is critical. Future research should focus on understanding abiotic stress responses in hemp, corn earworm and its natural enemies in hemp. Impacts of climate change on industrial hemp production mediated through changes in populations of serious insect pests such as corn earworm need to be given more attention for planning and devising adaptation and mitigation strategies for future management programs. There are still gaps in information that need to be addressed in order to allow production of management plans for these arthropod pests. It is also necessary to encourage the conservation of beneficial insects on hemp, to use these arthropods as pest control strategies.

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