A comparison between major chemical markers of the cultivated and wild harvested *Siphonochilus aethiopicus*, African ginger, from Mpumalanga, South Africa, using Liquid Chromatography–Mass Spectrometry

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**Abstract**

*Siphonochilus aethiopicus*, known as African ginger, is indigenous to South Africa and has multiple traditional uses in health to treat human diseases. The multiple traditional uses of African ginger have exacerbated the over-harvesting of the plant species from the wild for trade on the traditional medicine market. The wild populations of African ginger have almost completely depleted from the wild and a few African ginger cultivation sites have been established in South Africa, to conserve the plant species.

The aim of the study was to compare the major chemical markers of the cultivated and wild harvested African ginger from Mpumalanga using Liquid Chromatography–Mass Spectrometry (LC-MS). The Council for Scientific and Industrial Research (CSIR) wild-harvested African ginger dated 2010 was used as a reference sample for comparison purposes. The LC-MS data generated from the ethanol extracts of the cultivated African ginger detected the presence of 4,4a,5,8a,9-tetrahydro-3,5,8a-trimethynaphtho[2,3-b]fur-an-8. This chemical marker was also detected in the wild harvested African ginger as compared to a previous study, which auto oxidised in the referenced sample over time. This study supports the efforts to conserve African ginger through cultivation for further development in commercialisation.

**Introduction**

*Siphonochilus aethiopicus* (Schweinf.) B.L. Burtt is a member of the family Zingiberaceae, as described in the “Medicinal Plants of South Africa (2nd Edition)” book of Van Wyk and co-
African ginger is reported as a source of food used as a spice in dish flavouring in Nigeria. It is not clear if the communities including traditional health practitioners use the leaves of the African ginger in traditional medicines to treat human diseases, as there is limited scientific knowledge about the chemical fingerprinting of the leaves of the plant species.

An optimised LC-MS method has been developed for the diethyl ether, ethanol, and water extracts of the wild harvested African ginger in South Africa by the study of Fouche et al., which investigated the chemical profile and biological activity of the wild harvested African ginger against traditional uses to treat asthma. The major chemical marker was detected and identified as 4,4a,5,8a,9-tetrahydro-3,5,8a-trimethylnaptho[2,3-b]furan-8-one in hexane and ethyl acetate extracts prepared from the rhizomes and roots of the wild harvested African ginger from Mpumalanga using LC-MS. The furanoterpenoids have also been shown to have anti-plasmodial activity in other studies.

In 2021, the study of Al-Tannaka isolated another class of chemical compounds known as furenoterpenoids, which have the potential to treat asthma as shown in vitro and in vivo studies. The furenoterpenoids have also been shown to have anti-plasmodial activity in other studies.

In 2014, the harvest of African ginger in South Africa was reported. The plant species bears elongated leaves, distinctive cone-shaped rhizomes, and purple flowers. The communities including traditional health practitioners use rhizomes and roots to prepare a traditional medicine, to treat a variety of human diseases which have been widely reported in the literature. For instance, the plant species are used to treat coughs, colds, and asthma, as well as menstrual pains, navel pains, toothache, rheumatism, neuralgia, sexually transmitted infections, and stomach–ache. The plant species is also traditionally used to provide children health care in rural areas which often have limited access to western healthcare facilities.

Some studies have brought to light that African ginger can be cultivated on a small scale, thus these plants have been saved in the past from the verge of extinction. Cultivation is generally one of the strategies for the conservation of medicinal plant species, thus considered an alternative to wild harvesting to alleviate pressure on wild harvesting of the medicinal plants. Moreover, the cultivation of medicinal plant species can create opportunities for communities including traditional health practitioners, who no longer have easy access to wild populations of medicinal plant species, to continue practicing their traditions and cultures. However, there is limited scientific knowledge about the acceptability of cultivated medicinal plants by traditional health practitioners and concerns about the activity or potency of cultivated medicinal plant species. Nonetheless, the traditional health practitioners in the Mpumalanga province of South Africa are practicing the cultivation of African ginger in their back yards as part of the sustainable use of the plant species in traditional practices.

**Aim of the study**

The aim of the study was to compare the chemical markers of the cultivated African ginger with wild harvested African ginger from Mpumalanga using LC-MS.

**Materials and methods**

**African ginger sample collection and pre-treatment**

About 3 kg of the wet weight of the cultivated African ginger rhizomes and roots were collected from a private farm in Mpumalanga and supplied to the CSIR through a courier in August 2021. Since African ginger is the only indigenous plant species in the Zingiberaceae family in South Africa and has such a unique and distinctive morphology, no voucher specimen was collected.

The rhizomes and roots were washed with pure running water.
tap water to remove soil particles, followed by slicing and then
drying in an oven at 60°C for three days. The final moisture of
the dried rhizomes and roots after three days of drying was
determined at 7% of water, using a moisture analyser. The
dried rhizomes and roots were milled into a grounded material
using a hammer mill through a 6mm screen. Figure 2 indicates
the sequence followed for the primary processing of African
ginger.

Extraction

The Standard Operating Procedures (SOPs) developed by
the CSIR in 2011 for the extraction of chemical compounds
from the rhizomes and roots of African ginger using ethanol were
followed, according to the method of Fouche, et al. 2011. In
short, in the first cycle of extraction, 10g of grounded African
ginger rhizomes and roots was mixed in 60ml of absolute
ethanol and stirred at room temperature for one hour with
occasional stirring and filtered. The ethanol–biomass mixture
was filtered through grade 2 Filter Papers. The second cycle
of extraction involved the filter cake or pulp extracted again
with 60ml of fresh and absolute ethanol for one hour. The
third extraction of the filter cake or pulp was left overnight in
60ml ethanol. The filter cake or pulp was finally discarded, and
the extracts were dried using a Buchi rotavapor to give a dried
ethanol extract of 0.26 g (0.086%, w/w).

This extraction process was repeated on the wild harvested
African ginger of 2010 that is still available at the CSIR research
facility and stored in a freezer at 4°C in a powdery form, to
prevent microbial contamination.

LC-MS analysis for the detection of major chemical marker

The method development for chemical marker identification
by Fouche et al. 2011 was followed. In brief, each extract (25
mg) from both the cultivated and wild harvested African
ginger rhizomes and roots extracts was dissolved in ethanol
in an ultrasonic sonicator bath for 30 minutes in the dark. The
extraction solvent was removed and filtered through a 0.2 μm
syringe filter before analysis.

A pooled sample was created by mixing equal amounts of
each extract. The blank sample was the ethanol used as a
solvent for the samples. The samples were analyzed with the
Waters Acquity UPLC coupled to a Waters G1 HDMS mass
spectrometer and ESIPos (ESI+) and ESINeg (ESI−) ionization
mode covering the mass range of 50 – 1200 Dalton (Da). The
samples were analyzed in a randomized order and each sample
was analyzed four (4) times to allow for statistical processing
of the raw data.

Results and discussion

The ethanol extracts prepared from the cultivated and wild
harvested African ginger rhizomes and roots were analyzed
for major chemical marker identification using LC-MS and are
shown in Figure 3.

Good separations of the ethanol extracts were obtained,
and the chemical marker could be detected and observed in
the cultivated African ginger extract, showing the presence
of the furanoterpenoids, signifying the chemical marker as
4,4a,5,8a-tetrahydro-3,5,8a-trimethylnaptho[2,3-b]furan-
8-one (Figure 3A). A similar chemical marker compound was
detected using the LC-MS in the study of Fouche et al. 2011 on
the wild African ginger rhizomes and roots, which was shown
to have asthmatic effects on African ginger in vitro and in vivo.

The marker compound was however not detected in 2010 in
the wild harvested African ginger rhizomes and roots (Figure
3B). This is probably due to the process of auto-oxidation
of the marker compound [23–26] since the sample has been
stored in open-air contained in cold rooms to prevent possible
microbial contamination. The auto-oxidation process implies
the stability and therapeutic effects of the formulated African
ginger formulated products over time.

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**Figure 2**: Sequential primary processing of African ginger. A = washed African ginger rhizomes and roots with clean running tap water; B = Sliced African ginger rhizomes and roots; C = Oven dried African ginger rhizomes and roots; D = Grounded African rhizomes and roots.
Conclusion

The aim of the study was to compare the chemical markers of the cultivated African ginger against the wild harvested African ginger of 2010 which was used as a reference sample in this study. The LC-MS method was developed to aid in the detection of the chemical marker from the cultivated African ginger rhizomes and roots and compared to the wild harvested African ginger rhizomes and roots. The extracts were analyzed in ESI+ and ESI- modes to detect any chemical differences. The ESI+ analysis mode is the best to detect the known marker compound and similar compounds produced by African ginger.

The results demonstrated that the African ginger rhizomes and roots obtained from cultivated land in 2021 have the same marker compound as the wild-harvested African ginger chemical marker (see the study of Fouche et al., 2011), which belongs to a class of chemical compounds known as furanoterpenoids. However, over time, there was no evidence of the major compound detected in the wild harvested African ginger, due to the auto-oxidation process. The reason is that autoxidation of the African ginger powder occurs after and before extraction. Therefore the only compounds detected using LC-MS in the wild harvested African ginger samples were the oxidised derivatives of furanoterpenoid. The cultivated African ginger sample from a farm indicated the presence of major chemical marker derivative, 4,4a,5,8a,9-tetrahydro-3,5,8a-trimethylnaphtho[2,3-b]furan-8-one, as well as some significant concentrations of the oxidised derivatives (refer to Figure 3B), which has the potential to treat asthma through in vitro and in vivo studies as reported in the study of Fouche, et al. [6].

The use of LC-MS in the detection of chemical markers highlights how similar the cultivated and wild harvested African ginger is in relation to chemical market analysis. Since African ginger has become extinct in the wild due to overharvesting, the cultivated African ginger can be considered in the development of traditional remedies for the treatment of human diseases including treating pain, alleviating the perceptions among the traditional health practitioners that the cultivated African ginger is not as effective in health care from the indigenous knowledge systems perspective. The findings of this study have shed light on an underexplored aspect of African ginger, one that is significant on the African continent, and have both relevances for commercialisation and government policy.

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