Perforinopathy: a spectrum of human immune disease caused by defective perforin delivery or function

Ilia Voskoboinik1,2* and Joseph A. Trapani2,3*

1 Killer Cell Biology Laboratory, Peter MacCallum Cancer Centre, East Melbourne, VIC, Australia
2 Sir Peter MacCallum Department of Oncology, The University of Melbourne, Melbourne, VIC, Australia
3 Cancer Cell Death Laboratory, Peter MacCallum Cancer Centre, East Melbourne, VIC, Australia

*Correspondence: ilia.voskoboinik@petermac.org; joe.trapani@petermac.org

INTRODUCTION
Perforin (PRF, encoded by the PRF1 gene) is a pore-forming toxin (1, 2) stored in the secretory granules of cytotoxic T lymphocytes (CTLs) and natural killer (NK) cells (collectively known as cytotoxic lymphocytes, CLs) (3). During an immune response, CLs form an immune synapse (IS) with various types of antigen-presenting cell, and release PRF and granzyme serine proteases into the synaptic cleft (4). The extracellular milieu of the IS (high calcium and neutral pH) promotes perforin membrane binding and insertion culminating in pore formation (5, 6), an indispensable requirement for granzymes to enter the target cell cytoplasm and trigger a multitude of cell death signaling pathways (7–12).

A SPECTRUM OF CLINICAL PRESENTATIONS REFLECTING FUNCTIONAL PRF DEFICIENCY
It was first appreciated around the turn of this century that the complete loss of perforin activity results in a fatal, autosomal recessive immunoregulatory disorder of infancy (median age of onset <12 months), familial hemophagocytic lymphohistiocytosis (FHL) (13), which can be cured only by heterologous bone marrow transplantation (14). The inability to clear antigen-presenting cells by impaired CLs causes an uncontrolled activation and expansion of CD4+ and CD8+ T cells that secrete high levels of interferon-γ. This cytokine is central to disease pathogenesis, as it leads to macrophage activation and secondarily to the overproduction of pro-inflammatory cytokines by these cells. This in turn is manifested clinically as intractable fever, liver and spleen enlargement, and hemophagocytosis in the bone marrow and lymphoid organs, leading to severe anemia and leukopenia (15). It is generally thought that macrophage activation and infiltration in bone marrow leads to hemophagocytosis in FHL patients. Recently, it was shown that interferon-γ specifically potentiated phagocytosis of erythrocytes by macrophages, thus suggesting a key mechanism for anemia in these patients (16). Variations in clinical presentation stem in part from the fact that there are up to five independent genetic causes of primary FHL and related syndromes (4). Linkage analyses have identified four genetic causes that account for 80–90% of FHL, while a further locus mapping to Chr 9 is yet to be defined (17). Inactivating perforin mutations are responsible for approximately 50% of all cases (type 2 FHL, FHL2), while mutations in three other genes, UNCI3D, STX11, and STXB2 (FHL3, 4, and 5 respectively) impair or ablate the delivery of perforin to the IS (18–21). Thus, all the causes of FHL are causally linked by the failure to deliver sufficient active perforin to the IS.

Natural killer cells (innate immunity) and CTLs (adaptive immunity) were first recognized for their key role(s) in the defense against viruses; more recently these cells have also been appreciated as being critical for immune surveillance against a variety of malignancies, particularly those of hematopoietic origin (22, 23) or of multiple cancers in the same individuals (24). Both these
roles reflect the importance of perforin in initiating the apoptosis of dangerous cells, either those harboring an intracellular pathogen, or possessing the potential for uninhibited growth and spread, to the detriment of the host. Given this, it is perhaps surprising that (infantile) FHL rarely presents with overwhelming sepsis syndrome caused by viral infection (with the rare exceptions of persistent or fulminant Epstein–Barr virus infection) and almost never with malignancy. Thus, most cases of FHL are not manifested directly through the consequences of failed “target cell death” but rather, indirectly, through an apparent skewing of the immune response toward exaggerated cytokine secretion, the second major type of effector function of CTL/NK cells.

Recent work on CTL/NK cells has highlighted their regulation of various inflammatory pathways through their cross-talk with other components of the immune system (25–27); the consequent aberrations in inflammatory responses leading from failed perforin production can occur in response to known human pathogens, although a specific pathogen that triggers the onset of FHL is rarely identified. These phenomena strongly indicate that perforin and the pathways that synthesize and deliver it to the IS play a more fundamental role in immune homeostasis, centered more particularly on adaption of the neonate to the myriad “non-pathogenic” antigenic stimuli he/she will encounter after leaving the womb. In this context, we postulate it is the failure to clear “constitutive” antigen-presenting cells through perforin-dependent (largely granzyme-mediated) cell death that induces the increased secretion of interferon-γ from the killer lymphocyte, in turn provoking severe dysregulation of pro-inflammatory and chemokine cascades in bystander cells. The molecular and cellular mechanisms causally linking failed target cell apoptosis and the hypersecretion of interferon-γ are of fundamental importance to the pathophysiology of perforinopathies, and ought to be the focus of intensive research. The consequential effects on macrophages, including secondary pro-inflammatory cytokine secretion and grossly increased macrophage phagocytic activity, are the major manifestations of disease and are most marked when perforin activity is completely abolished, but less so if some residual perforin activity persists, for example due to inheritance of perforin missense mutations that are not completely inactivating. This theme is taken up again below, as we believe that the variable clinical presentations probably flow on from this fact. While this paper focuses on perforin, it has also been noted for some time that granzymes (for example, granzymes A and M) can influence inflammatory pathways (25, 27), sometimes in a perforin-dependent manner, and at other times without the need for perforin to be present. While the significance of granzyme-mediated inflammatory pathways for human health are yet to be defined and hyper- (or hypo-) inflammatory syndromes are not yet described, future research should keep this possibility in mind. This is particularly so because granzyme genes can show considerable polymorphism, both in mice (28) and humans (29).

On the basis that congenital defects in CTL/NK cells that influence the secretion of active perforin may become clinically evident at various stages of life, we would like to propose that this group of disorders be considered as perforinopathies under three subheadings: acute, subacute, and chronic, depending on the stage of disease onset (Table 1). As alluded to above, earlier presentations tend to reflect cytokine-mediated immunopathology; later in life, a patient may present more cryptically with relatively weaker inflammatory manifestations or none at all (Figure 1). We think it is quite likely that clinicians managing these less urgent clinical presentations may not have previously considered perforinopathy in the differential diagnosis, particularly in children presenting with a variety of inflammatory disorders beyond infancy.

**ACUTE PRESENTATIONS OF PERFORINOPATHY – FHL**

For the purpose of this discussion, we define acute perforinopathy as that resulting in a clinical presentation prior to the age of 24 months, with median of approximately 9 months (this may vary depending on the genetic cause of FHL). As discussed above, the pathogenesis involves a cascade of downstream events following on from the inability of NK cells and CTL to present functional perforin and, therefore, kill a cognate target cell. Affected infants typically present with “classic” FHL and meet all or most of the criteria described in DLH-2004 (15). The diagnosis is confirmed by the loss of NK cytotoxicity (which is convenient to test, as the NK cells of healthy individuals display constitutive, pathogen-independent cytotoxicity) and identification of mutations in candidate genes (PRF1, UNC13D, STX11, STXB2). Affected infants are typically very unwell, and may require admission to a high dependency or intensive care unit; given the autosomal recessive inheritance of most of these disorders, a sibling may have previously been similarly affected, while parents are almost always unaffected carriers. Patients first have their condition stabilized, and are then prepared for heterologous bone marrow transplantation, the only potentially curative therapy (14, 15).

Acute perforinopathy is caused by detrimental mutations in PRF1 or in proteins responsible for its delivery from the lumen of cytotoxic granules to the IS, UNC13D, STX11, or STXB2 (4). The molecular basis of many disease-causing PRF1 mutations has been investigated directly using recombinant expression systems (23, 30–33), and is supported by the X-ray crystal structure of perforin (1). In contrast, the biochemical bases of pathological mutations in the other three proteins, Munc13-4, Syntaxin11, and Munc18-2, remain largely unexplored due to the lack of sufficiently informative experimental systems. As a result, the loss of molecular function is predicted (34, 35), either on the basis of undetectable NK cell activity and/or severely reduced protein expression due to degradation. In the context of an early onset FHL (acute perforinopathy), the known structural and cellular defects correlate well with disease severity. Indeed, in the case of PRF1, genuinely “null” mutations are most commonly caused by nonsense or frame-shift mutations and in-frame deletions that completely abrogate function. By contrast, a careful genotype-phenotype analysis has revealed that only about 50% of missense mutations ever reported resulted in a complete loss of perforin function, either due to direct interference with a critical functional domain or unscheduled post-translational modifications such as glycosylation (30, 36). The remaining PRF1 missense mutations were more commonly associated with atypical presentations of FHL or with seemingly unrelated pathologies (immunoregulatory or otherwise) in older children, adolescents, and even adults, who did not necessarily present with FHL at all (23, 37). If
Table 1 | Possible clinical manifestations of perforinopathy.

|                          | Acute perforinopathy | Sub-acute perforinopathy | Chronic perforinopathy |
|--------------------------|----------------------|--------------------------|------------------------|
| Age at onset             | 0–2 years            | >2 years                 | Adolescents – adults   |
| Cause                    | Bi-allelic mutations in PRF1, UNC13D, STX11, STXBP2, leading to complete loss-of-function | Bi-allelic mutations in PRF1 and, putatively, in UNC13D, STX11, STXBP2, leading to partial loss-of-function | Monoallelic mutations in PRF1 [e.g., polymorphism 272C > T (Ala91Val)], and possibly, other FHL-causing genes, are contributing not causative factors |
| Onset                    | “Classical” FHL – meets all or most of the criteria described in HLH-2004 | Difficult to diagnose as does not meet minimal essential criteria of HLH-2004: e.g., inflammatory disease that responds to corticosteroid therapy and has remitting/relapsing clinical course; hematological malignancy | May include a range of conditions, including hematological malignancy, macrophage activation syndrome, lymphoproliferative disease, but not FHL |
| Diagnostic features      | Intractable fever and hepatosplenomegaly are early and prominent Investigations (including NK function assays) described in HLH-2004 will strongly suggest FHL, and disease is confirmed by DNA sequencing | Once FHL is suspected, conduct tests as described in HLH-2004. Severe reduction of NK cell function that prompts genetic analysis may indicate FHL. Test siblings for bi-allelic mutations in FHL-related genes | Mild reduction of NK function. No symptoms described in HLH-2004 are expected |
| Therapy                  | Use protocols in HLH-2004; heterologous stem cell transplantation is the only curative therapy | Initially, corticosteroid therapy When genetic cause is identified, HLH-2004 and heterologous stem cell transplantation, as the only cure. If asymptomatic siblings are carriers of bi-allelic mutations in FHL-related genes, preventative stem cell transplantation may be considered | Disease-specific therapy, ranging from corticosteroids to stem cell transplantation |

Residual perforin activity delays the clinical onset beyond the age of 12 months, we term the presentation “subacute.”

**SUB-ACUTE PRESENTATIONS OF PERFORINOPATHY**

Sub-acute perforinopathies have a wide spectrum of manifestations, all of which are all caused by a partial (“sub-total”) loss of CL cytotoxicity due to bi-allelic mutations in one of the four genes described above. Unlike the acute form of the disease, sub-acute perforinopathies may be difficult to diagnose due to their generally milder and more “patchy” clinical presentations, an intermittent clinical course, a range of ages of onset and their frequent response to non-specific immune-suppressive or immune-ablative therapies (38, 39). These patients are more likely to present with a precipitating infection or with an isolated inflammatory manifestation such as pneumonitis that can lead the treating clinician away from the underlying diagnosis. While the only curative therapy for such patients is still ultimately bone marrow transplantation, combination drug therapy may induce remissions of variable duration. These non-curative interventions may be the only available therapies that influence disease outcome, but they have frequent unwanted effects such as opportunistic infection, growth retardation, and bone marrow suppression. Alleviating symptoms that might have categorized them as FHL patients might also delay diagnosis in both the patient and their younger siblings.
siblings (38). Considering the fact that “classic” FHL is relatively rare and commonly described in infants or very young children, we believe sub-acute perforinopathies may be under-diagnosed, and thus may delay consideration of curative stem cell transfer as a definitive treatment option.

How common are sub-acute perforinopathies? The inevitable answer is “We do not know.” Due to the lack of direct functional data on Munc13-4, synt11, and Munc18-2 mutations, we will discuss perforin deficiency as a paradigm for partial loss of CL function. The first indication that bi-allelic perforin mutations may lead to atypical FHL came from the observation that individuals who inherit two mutant alleles of PRF1, one of which is the common polymorphism C272 > T, encoding the ostensibly conservative Ala91Val (A91V) substitution, presented at an older age than “classic” FHL, and with variable symptoms (13, 38, 39). The initial suggestion that the “polymorphism” may be pathogenic was initially met with skepticism, due largely to the very high frequency of the allele among Caucasians (8–17% are heterozygotes) and the rareness of early onset FHL, which was considered at that time to be the only authentic manifestation of the disease (40). However, over the next several years, additional clinical research and biochemical studies have confirmed the original prediction that the A91V substitution is a genuine mutation that impacts severely on perforin structure and function (31, 41). While a prospective cohort study has never been reported, it is nonetheless clear from many reported case studies that inheritance of A91V in the homozygous state or its co-inheritance with a genuinely null allele is very strongly associated with atypical FHL or other immunopathologies that are delayed compared with “classic” disease (sub-acute perforinopathies). In the largest epidemiological studies reported to date [involving over 2,600 healthy individuals – (42–44)], healthy A91V homozygotes have been reported only in one study (42); every other reported case of A91V homozygocity was invariably associated with a pathology. However, the statistical power of these studies linking the genotype with disease has been insufficient to achieve significance. Based on the Mendelian inheritance of perforin mutations, it is predicted that at least 1 in 600 Caucasian individuals should be homozygous for A91V. Assuming strong penetrance of an associated immunopathology in these individuals, such a perforinopathy would be almost 100 times as common as classic (“acute”) FHL. Remarkably, at this frequency, A91V-related immune deficiency would rank among the most common congenital disorders; it would be approximately four times as frequent as cystic fibrosis and at a par with Down syndrome.

Is partial perforin deficiency a common phenomenon, and is it caused only by A91V? A retrospective analysis of all reported cases of FHL associated with bi-allelic PRF1 mutations uncovered a previously unappreciated dichotomy: half of the patients who carried at least one missense mutation in PRF1 presented “acutely,” with a FHL-like syndrome before the age of 12 months, while the remaining 50% had significantly delayed FHL, or presented with other, seemingly unrelated pathologies (23, 37, 45). Most of these patients either developed hematological malignancies (usually beyond the age of 10 years, which we consider to be a “chronic” presentation) or presented with unusual or protracted viral infections (23, 46, 47). Critically, biochemical analysis of multiple missense mutations associated with atypical FHL has led to the realization that partial perforin deficiency results from perforin misfolding: all reported cases of atypical, delayed FHL had some recoverable perforin-mediated cytotoxicity in vitro (23). Structural studies recently supported these findings, as wild-type perforin was shown to be a thermo-labile protein, with a melting temperature (the temperature at which the protein denatures and starts to lose function) only slightly above 40°C (5). Even though this analysis was conducted in vitro, this finding has raised the intriguing possibility that the protracted, cytokine-induced fever of FHL might further aggravate the disease by accentuating the protein misfolding defect. Given the diversity of clinical presentation that encompasses cancer, atypical onset of viral infections, and attenuated FHL-like syndromes, it is likely that a significant proportion of sub-acute perforinopathies go undiagnosed.

Identification of missense mutations that confer partial activity for the proteins encoded by the UNC13D, STX11, and STXB2 genes may broaden the spectrum of recognized sub-acute perforinopathies. A number of non-synonymous polymorphisms in these genes have already been cataloged in single nucleotide polymorphism (SNP) databases, and future studies will determine whether any of these variations can affect the rate and/or efficiency of secretory granule exocytosis and membrane fusion, steps that ultimately determine the amount of perforin secreted into the IS and its rate of delivery.

**CHRONIC PRESENTATIONS OF PERFORINOPATHY**

We consider chronic perforinopathies to be disease states that represent as a spectrum of immune-mediated diseases associated with monoallelic mutations in FHL-related genes. The presentations typically do not have a strong resemblance to classic FHL and may include blood cancers and macrophage activation syndrome in patients with juvenile rheumatoid arthritis. Typically, age of onset will be beyond 5 years of age. In addition, some studies have reported an association of PRF1 polymorphisms and the outcome of allogeneic bone marrow transplantation. All of these associations are contentious, as studies that find no link outnumber those supporting such an association.

Once again, the most reliable and “testable” information comes from the analysis of PRF1 polymorphisms, as the functional consequence of mutations is well understood. Considering the fact that hundreds of FHL patients with known genetic defects have been diagnosed around the world since 1999, the number of their family members who are known carriers of monoallelic mutations, should exceed 1,000. This is a significant cohort of unrelated individuals, who may be potentially investigated in prospective longitudinal studies on immune surveillance of infections and cancer and for dysregulated immune homeostasis.

An association between presumed partial CL deficiency due to monoallelic A91V perforin polymorphism, and acute lymphoblastic leukemia or macrophage activation syndrome, has been demonstrated in several studies that have necessarily involved a limited number of individuals (48, 49), but no statistical difference was observed in follow up studies when larger cohorts of patients were examined (24, 43). The likely reason for this apparent discrepancy may be the relatively low statistical power of smaller studies.
Another possibility is that a partial deficiency of CL function may predispose more strongly to a specific subtype of malignancy or other pathology. For example, while the largest analysis of patents with acute lymphoblastic leukemia did not reveal an enrichment of A91V carriers compared to healthy controls, a subset of the ALL patients who also had BCR-ABL translocations were more likely to carry the A91V allele than the control group (43). Similarly, in a study that retrospectively examined patients who had been diagnosed with more than one primary malignancy during their lifetime, a significant proportion of patients diagnosed with both melanoma and B cell lymphoma were carriers of A91V or another perforin mutation that is partly inactivating (R28C), than those who had been diagnosed with either disease alone (24).

We have tried to explore the molecular basis for how a monoallelic mutation might adversely affect overall cytotoxic function in an inherited condition classically considered to be recessive. Molecular analysis has revealed that A91V has a partial dominant-negative effect on the function of wild-type perforin; partly misfolded perforin might interfere with the expression or trafficking of normal perforin through the endoplasmic reticulum/Golgi to the secretory granules (31, 50). Once wild-type and misfolded perforin molecules are “mixed” and stored together, it is easy to imagine that perforin pores with a heterogeneous composition of monomers might have disproportionately poor function. We also speculate that in heterozygous individuals, the wild-type allele may be under-expressed relative to the mutated allele, thus further diminishing overall CL function. This possibility needs to be formally excluded, but has been described for many other polymorphic genes [e.g., Ref. (51)].

Further analysis of CL function in patients with immune-mediated disease is warranted, particularly employing recent advances in single-cell microscopy technology, which may reveal CL deficiencies that would otherwise remain undetected in cell population-based experiments (6). Such technologies may validate the more common statistical approaches, where for example, the frequency of a PRF1 mutation in cancer patients is compared to that in healthy controls. In acute perforinopathy, there is a clear causal relationship between the disease phenotype and the patient’s genotype, but this is not the case in cancer (even assuming a clinician considered testing for perforin genotype, which would be unusual). Rather, most chronic perforinopathies are likely to present “indirectly.” In cancer, the PRF1 gene [and, most likely, UNC13D, STX11, and STXB2P2 (52)] would be considered to be a tumor suppressor gene, but is not the cause of malignancy per se and is certainly not the only causal factor. Rather, impaired perforin function reduces the surveillance of cancerous cells, which were generated through an unrelated genetic event. This notion raises a question of what constitutes an appropriate control in these studies. Since the hypothesis is that carriers of monoallelic mutation(s) in PRF1 or related genes predispose to immune-mediated diseases with uncertain age of onset, taking a “snapshot” of a healthy population may underestimate the impact of mutations. In contrast, it was recently shown that individual differences between the cytotoxic activity of NK cells (and cytotoxic T cells) are much smaller than previously thought (6). This opens an avenue for “personalized” analysis of CL function that may shed light on predisposition of an individual to an immune pathology, prospectively and retrospectively.

**SUMMARY**

In this discourse, we have proposed the term “perforinopathy” to denote the wide and diverse spectrum of manifestations of perforin deficiency, both temporal and clinical. Early (acute) clinical presentations are often fatal and usually represent dysregulated immune homeostasis and the resultant hypercytokinemia (“cytokine storm”) and hemophagocytosis consequent on macrophage activation. Sub-acute presentations still occur in infancy/early childhood, may typically have a more benign course that is still principally inflammatory and may accompany other pathologies such as juvenile idiopathic arthritis or virus infection. Chronic manifestations of perforin deficiency may appear as late as adulthood, and the principal cause of symptoms is the failure to clear specific dangerous cells, particularly pre-malignant cells. Hematopoietic malignancies seem to be a common outcome, and some patients may present well into adulthood with more than one primary cancer: early onset malignant melanoma and lymphoma is the commonest combination we have observed. Although most of the data supporting our proposed “classification” is centered on our knowledge of PRF1 gene mutations, we hope that this paper will stimulate further study on polymorphisms and mutations of other genes that play a role in delivering functional perforin to the target cell.

**ACKNOWLEDGMENTS**

This work was supported by Program, Project, and Fellowship grants from the National Health and Medical Research Council (NHMRC) of Australia, and a project grant from the Cancer Council Victoria. We thank Kevin Thia for technical assistance in preparation of this manuscript.

**REFERENCES**

1. Law RH, Lukoyanova N, Voskoboinik I, Caradoc-Davies TT, Baran K, Dunstone MA, et al. The structural basis for membrane binding and pore formation by lymphocyte perforin. Nature (2010) 468:447–51. doi:10.1038/nature09518
2. Baran K, Dunstone M, Chia J, Ciccone A, Browne KA, Clarke CJ, et al. The molecular basis for perforin oligomerization and transmembrane pore assembly. Immunity (2009) 30:884–95. doi:10.1016/j.immuni.2009.03.016
3. Peters PJ, Borst J, Oorschot V, Fukuda M, Kranenburg O, Tschopp J, et al. Cytotoxic T lymphocyte granules are secretory lysosomes, containing both perforin and granzymes. J Exp Med (1991) 173:1099–109. doi:10.1084/jem.173.5.1099
4. de Saint Basle G, Gaël Ménaché G, Fischer A. Molecular mechanisms of biogenesis and exocytosis of cytotoxic granules. Nat Rev Immunol (2010) 11:568–79. doi:10.1038/nri2803
5. Lopez JA, Susanto O, Jenkins MR, Lukoyanova N, Sutton VR, Law RH, et al. Perforin forms transient pores on the target cell plasma membrane to facilitate rapid access of granzymes during killer cell attack. Blood (2013) 121:2659–68. doi:10.1182/blood-2012-07-446146
6. Lopez JA, Jenkins MR, Rudd-Schmidt JA, Brennan AJ, Danze JC, Manning SL, et al. Rapid and unidirectional perforin pore delivery at the cytotoxic immune synapse. J Immunol (2013) 191:2328–34. doi:10.4049/jimmunol.1301205
7. Walsh CM, Matloubian M, Liu CC, Ueda R, Kurahara CG, Christensen JL, et al. Immune function in mice lacking the perforin gene. Proc Natl Acad Sci U S A (1994) 91:10854–8. doi:10.1073/pnas.91.23.10854
8. Su B, Bochan MR, Hanna WL, Froelich CJ, Brahmi Z. Human granzyme B is essential for DNA fragmentation of susceptible target cells. Eur J Immunol (1994) 24:2073–80. doi:10.1002/eji.1830240921
9. Shiver JW, Su L, Henkart PA. Cytotoxicity with target DNA breakdown by rat basophilic leukemia cells expressing both cytolyisin and granzyme A. Cell (1992) 71:315–22. doi:10.1016/0092-8674(92)90359-K

10. Shi L, Kraut RP, Aebersold R, Greenberg AH. A natural killer cell granule protein that induces DNA fragmentation and apoptosis. J Exp Med (1992) 175:553–66. doi:10.1084/jem.175.2.553

11. Shi L, Kam CM, Powers JC, Aebersold R, Greenberg AH. Purification of three cytotoxic lymphocyte granule serine proteases that induce apoptosis through distinct substrate and target cell interactions. J Exp Med (1992) 176:1521–9. doi:10.1084/jem.176.6.1521

12. Heusel JW, Wesselschmidt RL, Shresta S, Russell JH, Ley TJ. Cytotoxic lymphocytes require granzyme B for the rapid induction of DNA fragmentation and apoptosis in allogeneic target cells. Cell (1994) 76:977–87. doi:10.1016/0092-8674(94)90376-X

13. Stepp SE, Dufourcq-Lagelouse R, Le Deist F, Bhawon S, Certain S, Mathew PA, et al. Perforin defect in familial hemophagocytic lymphohistiocytosis. Science (1999) 286:1957–9. doi:10.1126/science.286.5446.1957

14. Janka GE. Familial and acquired hemophagocytic lymphohistiocytosis. Ann Rev Med (2012) 63:233–46. doi:10.1146/annurev-med-041610-134208

15. Henter JI, Horne A, Arico M, Egeler RM, Filipovich AH, Imashuku S, et al. HLH-2004: diagnostic and therapeutic guidelines for hemophagocytic lymphohistiocytosis. Pediatr Blood Cancer (2007) 48:124–31. doi:10.1002/pbc.21039

16. Zoller EE, Lykens JE, Terrell CE, Aliberti J, Filipovich AH, Henson PM, et al. Perforin gene defects in familial hemophagocytic lymphohistiocytosis. J Exp Med (2011) 208:1203–14. doi:10.1084/jem.20100253

17. Orilieri E, Cappellano G, Clementi R, Cometa A, Ferretti M, Cerutti E, et al. Munc13-4 is essential for cytolytic granules fusion and is mutated in a form of familial hemophagocytic lymphohistiocytosis type 5. J Immunol (2009) 182:621–4. doi:10.4049/JMI.1000430

18. Chia J, Thia KY, Brennan AJ, Little M, Williams B, Lopez JA, et al. Familial immune dysregulation due to a gain of glycosylation mutation in lymphocyte perforin. Blood (2012) 119:1713–6. doi:10.1182/blood-2011-08-374355

19. Busiello R, Adriani M, Locatelli F, Galgani M, Finigiani G, Clementi R, et al. Atypical features of familial hemophagocytic lymphohistiocytosis. Blood (2004) 103:4610–2. doi:10.1182/blood-2003-10-3551

20. Mehta PA, Davies SM, Kumar A, Devidas M, Lee S, Zamzow T, et al. Perforin polymorphism A91V and susceptibility to multiple primary cancers. J Exp Med (2005) 200:811–6. doi:10.1084/jem.20040776

21. Mehta PA, Davies SM, Kumar A, Devidas M, Lee S, Zamzow T, et al. Perforin polymorphism A91V and susceptibility to multiple primary cancers. J Exp Med (2005) 200:811–6. doi:10.1084/jem.20040776

22. McFarley D, Catrion PF, Tulley P, Dudoit Y, Samri A, Autran B, et al. A triple-mutated allele of granzyme B incapable of inducing apoptosis. Proc Natl Acad Sci U S A (2003) 100:2562–7. doi:10.1073/pnas.0334793100

23. Chia J, Thia K, Brennan AJ, Little M, Williams B, Lopez JA, et al. Familial immune dysregulation due to a gain of glycosylation mutation in lymphocyte perforin. Blood (2012) 119:1713–6. doi:10.1182/blood-2011-08-374355

24. Trapani JA, Thia KYT, Andrews M, Davis ID, Gedye C, Parente P, et al. Human perforin: a family of CD8+ T cell granule proteins. Immunol Rev (2010) 233:69–85. doi:10.1111/j.0105-2896.2010.00907.x

25. Busiello R, Adriani M, Locatelli F, Galgani M, Finigiani G, Clementi R, et al. A single amino acid change, A91V, leads to conformational changes that can impair processing to the active form of perforin. Blood (2005) 106:932–7. doi:10.1182/blood-2004-09-3713

26. Orilieri G, Cappellano G, Clementi R, Cometa A, Ferretti M, Cerutti E, et al. Variations of the perforin gene in patients with type 1 diabetes. Diabetes (2008) 57:1078–83. doi:10.2337/db07-0947

27. Mehta PA, Davies SM, Kumar A, Devidas M, Lee S, Zamzow T, et al. Perforin polymorphism A91V and susceptibility to B-precursor childhood acute lymphoblastic leukemia: a report from the Children’s Oncology Group. Leukemia (2006) 20:1539–41. doi:10.1038/sj.leu.2404299

28. Cappellano G, Orilieri E, Comi C, Chiochetti A, Bocca S, Boggio E, et al. Variations of the perforin gene in patients with multiple sclerosis. Genes Immun (2008) 9:438–44. doi:10.1038/gen.2008.35

29. Trapani JA, Dunstone MA, Baran K, Whistock JC, Trapani JA. Perforin: structure, function, and role in human immunopathology. Immunol Rev (2010) 235:53–54. doi:10.1111/j.0105-2896.2010.00896.x

30. Beatty AD, Weller C, Levy B, Vogler C, Ferguson WS, Bicknese A, et al. A young boy with late onset hemophagocytic lymphohistiocytosis with predominant neurologic disease and perforin deficiency. Pediatr Blood Cancer (2007) 50:1070–2. doi:10.1002/pbc.21438

31. Katano H, Ali MA, Patera AC, Catalafamo M, Jaffe ES, Kimura H, et al. Chronic active Epstein-Barr virus infection associated with mutations in perforin that impair its maturation. Blood (2004) 103:2442–6. doi:10.1182/blood-2003-06-2171

Frontiers in Immunology | NK Cell Biology December 2013 | Volume 4 | Article 441 | 6
48. Cannella S, Santoro A, Bruno G, Pillon M, Mussolin L, Mangili G, et al. Germline mutations of the perforin gene are a frequent occurrence in childhood anaplastic large cell lymphoma. *Cancer* (2007) 109:2566–71. doi:10.1002/cncr.22718

49. Santoro A, Cannella S, Trizzino A, Lo Nigro L, Corsello G, Arico M. A single amino acid change A91V in perforin: a novel, frequent predisposing factor to childhood acute lymphoblastic leukemia. *Haematologica* (2005) 90:697–8.

50. Brennan AJ, Chia J, Browne KA, Ciccone A, Ellis S, Lopez JA, et al. Protection from endogenous perforin: glycans and the C terminus regulate exocytic trafficking in cytotoxic lymphocytes. *Immunity* (2011) 34:879–92. doi:10.1016/j.immuni.2011.04.007

51. Gartner JJ, Parker SC, Prickett TD, Dutton-Regester K, Stitzel ML, Lin JC, et al. Whole-genome sequencing identifies a recurrent functional synonymous mutation in melanoma. *Proc Natl Acad Sci U S A* (2013) 110:13481–6. doi:10.1073/pnas.1304227110

52. Machaczka M, Klimkowska M, Chiang SC, Merths M, Muller ML, Gustafsson B, et al. Development of classical Hodgkin’s lymphoma in an adult with biallelic STXBP2 mutations. *Haematologica* (2013) 98:760–4. doi:10.3324/haematol.2012.073098

**Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 02 October 2013; paper pending published: 25 October 2013; accepted: 25 November 2013; published online: 12 December 2013.

Citation: Voskoboinik I and Trapani JA (2013) Perforinopathy: a spectrum of human immune disease caused by defective perforin delivery or function. *Front. Immunol.* 4:441. doi: 10.3389/fimmu.2013.00441

This article was submitted to NK Cell Biology, a section of the journal *Frontiers in Immunology.*

Copyright © 2013 Voskoboinik and Trapani. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.
Author/s: Voskoboinik, I; Trapani, JA

Title: Perforinopathy: a spectrum of human immune disease caused by defective perforin delivery or function

Date: 2013-01-01

Citation: Voskoboinik, I; Trapani, JA, Perforinopathy: a spectrum of human immune disease caused by defective perforin delivery or function, FRONTIERS IN IMMUNOLOGY, 2013, 4

Persistent Link: http://hdl.handle.net/11343/191363

File Description: Published version