The Evolutionary History of Wild, Domesticated, and Feral *Brassica oleracea* (Brassicaceae)

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Abstract

Understanding the evolutionary history of crops, including identifying wild relatives, helps to provide insight for conservation and crop breeding efforts. Cultivated *Brassica oleracea* has intrigued researchers for centuries due to its wide diversity in forms, which include cabbage, broccoli, cauliflower, kale, kohlrabi, and Brussels sprouts. Yet, the evolutionary history of this species remains understudied. With such different vegetables produced from a single species, *B. oleracea* is a model organism for understanding the power of artificial selection. Persistent challenges in the study of *B. oleracea* include conflicting hypotheses regarding domestication and the identity of the closest living wild relative. Using newly generated RNA-seq data for a diversity panel of 224 accessions, which represents 14 different *B. oleracea* crop types and nine potential wild progenitor species, we integrate phylogenetic and population genetic techniques with ecological niche modeling, archaeological, and literary evidence to examine relationships among cultivars and wild relatives to clarify the origin of this horticulturally important species. Our analyses point to the Aegean endemic *B. cretica* as the closest living relative of cultivated *B. oleracea*, supporting an origin of cultivation in the Eastern Mediterranean region. Additionally, we identify several feral lineages, suggesting that cultivated plants of this species can revert to a wild-like state with relative ease. By expanding our understanding of the evolutionary history in *B. oleracea*, these results contribute to a growing body of knowledge on crop domestication that will facilitate continued breeding efforts including adaptation to changing environmental conditions.

Key words: cabbage, domestication, crop wild relatives, Mediterranean, origin, ecological niche.

Introduction

“Greek legend has it that the cabbage sprung from where Zeus’ sweat hit the ground.”

—N.D. Mitchell (1976)

A key tenet of evolutionary and plant biology is understanding how plants respond and adapt to changes in environmental conditions, which can be better understood by leveraging genotypic diversity and investigating the connections between genotype and phenotype. Crop wild relatives...
(CWRs) provide pools of allelic diversity that at one time were shared through a common ancestor with cultivated relatives. Although Vavilov recognized the potential of CWRs in the early 1900s (Vavilov 1926), advances in genomics and genome editing techniques have enabled scientists to better realize the potential of CWRs as a source of diversity and novel traits for the improvement of cultivated populations (Prohens et al. 2017; Li et al. 2018; Fernie and Yan 2019; Khoury et al. 2020; Turner-Hissong et al. 2020). Yet these scientific advancements are hindered in that we still have not identified the CWRs of many important crop species. Although cabbage may not have exactly formed from Zeus’ sweat, its evolutionary history, including identifying the closest living wild relative and origin of domestication, is still left unclear due to taxonomic confusion and the lack of genetic and archaeological evidence.

The horticultural crop Brassica oleracea L. has played an important role in global food systems for centuries, providing a source of leaf and root vegetables, fodder, and forage (Shyam et al. 2012). When first introduced to the species, Darwin drew many parallels between his theory of natural selection and the cultivation practices that led to the varied forms of this plant (Darwin 1868). Although many people may recognize that various dog breeds are all part of the same species, they are often surprised to learn that the domesticated forms of B. oleracea, broccoli (var. italica), Brussels sprouts (var. gemmifera), cabbage (var. capitata), cauliflower (var. botrytis), kale (var. acephala), and kohlrabi (var. gongylodes) are all one species as well. The global market for B. oleracea crops was around 70.1 million metric tons, in terms of production for 2019 (The Food and Agriculture Organization; www.fao.org). Although just six major crop types comprise the majority of the U.S. market (Agricultural Marketing Service, Market News Reports; www.ams.usda.gov), outside of these six major cultivars there exists at least 12 additional cultivated crop types (supplementary table S1, Supplementary Material online). These include lesser known varieties such as Chinese white kale or Cantonese gai-lan (Mandarin jié lán 花蓝, var. alboglabra), a leafy vegetable with florets, romanesco (var. botrytis) with unique fractal patterned curds, and walking stick kale (var. longata), which grows 6–12 feet (1.8–3.7 m) in height.

Compared with other crops, surprisingly little is known about the progenitor species and origin of domesticated B. oleracea. Primary challenges in identifying the progenitor species include the number of wild species that share a single cytodeme and are interfertile with B. oleracea (2n=18 chromosomes; similar genomic organization; referred to as the “C genome”), the corresponding confusion surrounding taxonomic relationships, and conflicting evidence regarding the center of origin. Wild relatives that share the C genome with domesticated B. oleracea include Brassica bourgeaui, Brassica cretica, Brassica hilarionis, Brassica incana, Brassica insularis, Brassica macrocarpa, Brassica montana, Brassica rippestris, and Brassica villosa. Throughout the literature, many of these species have been referred to by alternative names or have multiple subspecies. For example, B. cretica is described as having either three subspecies (subsp. aega, cretica, and laconica; Snogerup et al. 1990) or only two (subsp. cretica and nivea; Gustafsson et al. 1976). The taxonomic confusion is perhaps best highlighted by Bailey (1930), who stated that “Some of these plants appear to be more confused in literature than in nature.” The progenitor species of B. oleracea is further obscured by the presence of weedy, cabbage-like plants along the coastline of western Europe (England, France, and Spain), which have also been referred to as B. sylvestris (Mitchell 1976) or B. oleracea var. sylvestris (Gladis and Hammer 2001). The role of these weedy populations in the domestication of B. oleracea is unclear, with some studies suggesting these coastal wild populations represent the progenitor species (Snogerup et al. 1990; Song et al. 1990), and others identifying these wild forms as plants that escaped cultivation (Mitchell 1976; Mitchell and Richards 1979).

Given the uncertainty surrounding wild relatives and weedy populations, researchers have proposed numerous hypotheses for the progenitor species of B. oleracea (table 1). Hypotheses range from a single domestication with a single progenitor species (Song et al. 1990; Allender et al. 2007) to multiple domestications arising from multiple progenitor species (de Candolle 1855; Neutrof 1927; Lizgunova 1959; Helm 1963; Snogerup 1980; Heaney et al. 1987; Song et al. 1988; Swarup and Brahmi 2005). Findings that point to a single origin of domestication have proposed different wild species as the progenitor (Snogerup et al. 1990; Song et al. 1990; Hodgkin 1995; Maggioni et al. 2018). For instance, Neutrof (1927) suggested that B. montana was the progenitor of cabbages and that B. rupestris was the progenitor of kohlrabi, whereas Schulz (1936) identified B. cretica as the progenitor of only cauliflower and broccoli. Helm (1963) proposed a triple origin in which a single progenitor species gave rise to cauliflower, broccoli, and sprouting broccoli, whereas kale and Brussels sprouts were derived from another unknown wild species, and that all other crop forms were derived from a third unknown wild species. Snogerup (1980) proposed that cabbages were derived from wild B. oleracea, kales were derived from both B. rupestris and B. incana, and that Chinese white kale was derived specifically from B. cretica subsp. nivea.

Due to the lack of consensus on the progenitor species, the center of origin for B. oleracea has also remained obscure. One hypothesis is that domesticated B. oleracea originated in England from weedy B. oleracea populations, with early cultivated forms brought to the Mediterranean, where selection for many of the early crop types occurred (Hodgkin 1995). Other studies point specifically to Sicily, which boasts a large diversity of wild relatives, as the center of domestication (Schiemann 1932; Lizgunova 1959). This conforms with the observations of Vavilov (1951) that plants tended to be domesticated in a finite number of global centers of diversity, which includes the Mediterranean. Most recently, linguistic and literary evidence provided support for domestication in the Eastern Mediterranean, where there is a rich history of expressions related to the usage and cultivation of B. oleracea crop types in early Greek and Latin literature (Maggioni et al. 2010, 2018).

Using newly generated RNA-seq data for a diversity panel of 224 accessions that includes 14 cultivar types and nine wild
Table 1. Wild Species Which Have Been Proposed as Progenitor Species for *Brassica oleracea* Crop Types.

| Cultivar          | Wild Relative       | Author                  |
|-------------------|---------------------|-------------------------|
| Broccoli          | *B. oleracea*       | Linnaeus                |
|                   | *B. oleracea*       | Hedrick (1919) *a*      |
|                   | *B. oleracea*       | Giles (1941) *b*        |
|                   | *B. montana*        | Hegi (1919)             |
|                   | *B. oleracea* (from Italy) | Gates (1941) |
|                   | *B. cretica*        | Gates (1953)            |
|                   | *B. oleracea* and *B. alboalabera* | Song et al. (1990) |
| Brussels sprouts  | *B. oleracea*       | Linnaeus                |
|                   | *B. oleracea* (western Europe) | Gates (1953) |
|                   | *B. oleracea* (western Europe) | Snogerup (1980) |
|                   | *B. oleracea* and *B. alboalabera* | Song et al. (1990) |
| Cabbage           | *B. oleracea*       | Linnaeus                |
|                   | *B. oleracea*       | de Candolle (1824)      |
|                   | *B. oleracea*       | Hedrick (1919) *a*      |
|                   | *B. oleracea*       | Bailey (1930)           |
|                   | *B. montana*        | Hegi (1919)             |
|                   | *B. cretica*        | Schulz (1936)           |
|                   | *B. oleracea* (from Cyprus) | Gates (1941) |
|                   | *B. cretica*        | Gates (1953)            |
|                   | *B. oleracea* and *B. alboalabera* | Song et al. (1990) |
|                   | *B. cretica*        | Tutin et al. (1964)     |
| Kale              | *B. oleracea*       | Linnaeus                |
|                   | *B. oleracea*       | Hedrick (1919) *a*      |
|                   | *B. oleracea*       | Bailey (1930)           |
|                   | *B. montana*        | Hegi (1919)             |
|                   | *B. montana*        | Netrufal (1927)         |
|                   | *B. oleracea* (western Europe) | Gates (1953) |
|                   | *B. cretica, B. incana, B. rupestris* | Snogerup (1980) |
|                   | *B. incana and B. insularis* | Hosaka et al. (1990) |
|                   | *B. oleracea* and *B. alboalabera* | Song et al. (1990) |
| Kohlrabi           | *B. oleracea*       | Linnaeus                |
|                   | *B. rupestris*      | Netrufal (1927)         |
|                   | *Unknown Mediterranean species* | Gates (1953) |
|                   | *B. oleracea* and *B. alboalabera* | Song et al. (1990) |

Note.—Specific location is included in parentheses if indicated by the author. *Brassica oleracea* sometimes referred to as *B. oleracea* var. *sylvestris*.

*a*Edited observations by Sturtevant in the late 19th century.

*b*Referring to Prof Buckman’s experiment.

Phylogenetic and Population Clustering Distinguish Wild and Feral Populations

Sampling of *B. oleracea* cultivars included eight types of kales, five types of cabbages, Brussels sprouts, broccoli, cauliflower, Romanesco (var. *botrytis*), and kohlrabi (supplementary table S2, Supplementary Material online). Together, these cultivated types accounted for 188 of the 224 total samples. The remaining 36 samples included previously identified wild relatives: putatively wild *B. oleracea, B. cretica, B. incana, B. montana, B. hilarionis, B. insularis, B. macrocarpa, B. rupestris, and B. villosa*. The phylogenetic reconstruction of all 224 samples using SNPhylo (Lee et al. 2014) recovered several well-supported clades with greater than 70% bootstrap support, although overall support was generally poor (less than 70% bootstrap support), especially along the backbone. Chinese white kale, broccoli, cauliflower, romanesco, kohlrabi, curly kale, Brussels sprouts, *B. rupestris, B. macrocarpa,* and *B. insularis* were all recovered as monophyletic. Aside from red cabbages, cabbages were also monophyletic, but with only 55% bootstrap support. Seven cultivars (collards, tronchuda kale, savoy cabbage, perpetual kale, red cabbage, and narrow cabbage) were found throughout the tree as polyphyletic assemblies. Several wild samples were recovered within the cultivar clade, including two samples of *B. cretica* (196, 199), one sample of *B. montana* (222), and all samples of putatively wild *B. oleracea* (175, 176, 177; sample names in bold text; fig. 1). We also recovered a group in the cultivar clade consisting of five samples of three wild species, *B. incana* (205, 208, 209), *B. villosa* (233), and *B. cretica* (195), labeled ‘WildC-2’ (for wild samples with the C genome). Many of these ‘wild’ samples also share most or all of their ancestry with cultivars. At K = 2, in our fastSTRUCTURE analyses (Raj et al. 2014), samples clustered as either cultivars or wild (fig. 1). We find that two samples of *B. incana* (204, 207; likely both from Crimea), which are sister to all cultivated samples, share 100% of their ancestry with cultivated types, as do two samples of *B. cretica* (196, 199), one sample of *B. montana* (222), and all three samples of putatively wild *B. oleracea* (175, 176, 177). Together with the placement in the phylogeny, these analyses indicate that these are not truly wild samples, but represent feral types, defining feral here as either exoferal (a domesticated population derived from admixture with either a divergent population, a wild conspecific, another domesticated species, or another wild species) or endoferal (a
Ancestry proportions for 4422 Mabry et al. structure in the data (doi:10.1093/molbev/msab183 MBE + K = 109 ramosa 007 palmifolia 001 palmifolia 005 palmifolia 003 palmifolia 173 longata 004 palmifolia 107 ramosa 111 ramosa 044 medullosa 103 ramosa 037 medullosa 039 medullosa 180 viridis 032 costata 165 costata 027 costata 171 costata 139 alboglabra 033 costata 026 costata 034 costata 172 longata 028 costata 025 costata 105 ramosa 129 alboglabra 145 alboglabra 137 alboglabra 130 alboglabra 141 alboglabra 138 alboglabra 149 alboglabra 143 alboglabra 136 alboglabra 135 alboglabra 151 alboglabra 131 alboglabra 155 botrytis 156 botrytis 154 botrytis 153 botrytis 061 italica 169 italica 074 italica 064 italica 072 italica 071 italica 067 italica 062 italica 069 italica 070 italica 168 italica 075 italica 065 italica 127 botrytis 060 italica 123 botrytis 117 botrytis 126 botrytis 152 botrytis 121 botrytis 167 botrytis 118 botrytis 090 gongylodes 089 gongylodes 084 gongylodes 086 gongylodes 081 gongylodes 088 gongylodes 083 gongylodes 079 gongylodes 087 gongylodes 080 gongylodes 174 palmifolia 112 ramosa 063 italica 181 viridis 186 viridis 054 sabellica 059 sabellica 056 sabellica 057 sabellica 052 sabellica 058 sabellica 051 sabellica 053 sabellica 048 sabellica 050 sabellica 114 ramosa 104 ramosa 041 medullosa 045 medullosa 046 medullosa 110 ramosa 115 ramosa 116 ramosa 038 medullosa 036 medullosa 043 medullosa 112 ramosa
population of domesticated plants that has escaped from cultivation without the aid of introgression/hybridization with wild conspecifics; Gering et al. 2019). Our newly identified WildC-2 shows mixed wild and cultivar ancestry, which was also observed for one sample of tronchuda kale (30). The marginal likelihood was maximized at \( K = 3 \), in which a cluster comprised of broccoli, cauliflower, and Chinese white kale separated from other cultivated types. At \( K = 4 \), Chinese white kale was distinct from broccoli and cauliflower. The structure in the data was best explained by \( K = 5 \), in which the clade comprised of B. insularis and *B. macrocarpa* was separated and had shared ancestry with *Brassica cretica* (198), *B. hiliaronis*, *B. montana* (224), and one sample of tronchuda kale (30). Additional \( K \) values showed similar patterns (supplementary fig. S2, Supplementary Material online).

Principal component analysis (PCA) also separated cultivars from most wild samples (supplementary fig. S3D–F, Supplementary Material online). The PC1 axis distinguishes wild species from cultivars and the PC2 axis separates WildC-2 from all other wild species (triangles with black outlines). Although one sample of *B. cretica* (198) clusters closest to cultivated types, samples of *B. incana*, which were not in WildC-2, along with one sample of *B. montana* (222), two samples of *B. cretica* (196, 199), and all three samples of *B. oleracea* (175, 176, 177) cluster with the cultivars, corroborating the phylogenetic analyses. To further investigate the clustering patterns of *B. cretica* to cultivars, we included four additional wild-collected samples of two *B. cretica* subspecies (A and B = subsp. *nivea*, C and D = subsp. *cretica*, fig. 2A and B; supplementary fig. S3A, Supplementary Material online; labeled SRA in figure legend; Kioukis et al. 2020). Adding these samples supports the results of other studies that *B. cretica*, as a species, is very diverse. Although sample C does not group with other *B. cretica* samples using the PC1 axis, the PC2, PC3, and PC4 axes show much tighter clustering among the four wild-collected samples and one of our samples of *B. cretica* (198), indicating that our *B. cretica* (198) sample is an informative representative of wild-collected *B. cretica* (fig. 2A and B; supplementary fig. S3A, Supplementary Material online).

For crop samples, estimates of inbreeding coefficients from PCAngsd (Meinser and Albrechtsen 2018) roughly matched expectations for the frequency of heterozygotes under Hardy–Weinberg equilibrium, whereas inbreeding coefficients for wild species suggest excess homozygosity (supplementary fig. S4, Supplementary Material online), possibly reflecting cultivation practices for germplasm management and the relative isolation of wild populations (i.e., small effective population size), respectively. Feral samples, those which were identified as wild taxa, but were found more closely related to cultivars than to wild taxa in our phylogeny and clustered with cultivated samples in our PCA (*B. cretica*—196, 199; *B. incana*—204, 207; *B. montana*—222; and wild *B. oleracea*—175, 176, 177), show patterns of heterozygosity that are similar to crop samples, as do the four samples of *B. cretica* from Kioukis et al. (2020). Our WildC-2 exhibited patterns of excess homozygosity more similar to other wild taxa.

Domestication Is Also Reflected in the Transcriptome

Using expression profiles (transcript abundances) of 51,438 genes for our original 224 samples, we tested if cultivars and wild samples would still cluster separately based on the transcriptome. Overall, results and clustering patterns were similar to analyses using SNPs, with the axes of PC1 and PC2 separating most wild species from cultivars (fig. 2C; supplementary fig. S3B and C, Supplementary Material online). We again found the same samples of *B. incana* (204, 207), *B. cretica* (196, 199), *B. montana* (222), and *B. oleracea* (175, 176, 177) clustering with the cultivars, but in expression analyses WildC-2 clustered with the other wild samples, rather than separately as in our SNP based PCA. Hierarchical clustering of the expression profiles recovered similar patterns with two major groups: wild and cultivated, again with WildC-2 clustering with the other wild samples (supplementary fig. S5, Supplementary Material online). Although most cultivar groups were not recovered as unique clusters, there were a few exceptions. Brussels sprouts, Chinese white kale, and curly kale all formed distinct clades, which corresponds to what we know about their growth habit. Since RNA was collected at the 7th leaf-stage, before substantial morphological differentiation occurs between cultivars, it is not too surprising that they do not cluster distinctively by cultivar. Curly kale is almost immediately visually distinguishable from other cultivars in that the first true leaves have margins which are already undulate and/or frilled, in contrast to the more lanceolate (i.e., long, widest in the middle, with tapered tips) leaves observed in most cultivars. Brussels sprouts are also easily identifiable at this early growing stage as they have short, oblone to nearly circular leaves. Although Chinese white kale leaves look more similar to the lanceolate shape of other cultivars, they grow more rapidly and plants in this group are annual instead of biennial, which may explain why these accessions cluster separately from other cultivars.

To identify modules of genes that might be driving the observed clustering patterns, we used weighted correlation network analysis (WGCNA; Langfelder and Horvath 2008). We found that 48 modules, ranging in size from 34 to 35,981 genes, provided the best fit for the data (supplementary table S3, Supplementary Material online). To assess what types of biological processes were overrepresented in these modules, we used syntenic *Arabidopsis thaliana* genes and performed a GO analysis through PANTHER v. 16.0 (Mi et al. 2021). Overlap of *B. oleracea* with *A. thaliana* genes ranged from 17% to 98.3%, perhaps indicating that some modules are more conserved, whereas others are unique to *B. oleracea*. Modules which were more conserved between the two species included genes related to herbivory defense compound production (secondary metabolite biosynthetic process, phenylpropanoid biosynthetic and metabolic processes), wound formation (suberin biosynthetic processes), and wax formation (wax biosynthetic and metabolic processes), the latter of which may be correlated to the characteristic glaucous leaves of cultivated *B. oleracea* (supplementary table S4, Supplementary Material online). Within the top five conserved modules, the transcript abundance (TPM) was significantly different among the different groups (P value ≤ 2e-16).
**FIG. 2.** Principal component analysis (PCA) of SNPs and expression profiles. (A) Genetic variation PCA of PC1 versus PC2, (B) Genetic variation PC2 versus PC3, and (C) Expression profile PCA for PC1 versus PC2 of wild and cultivar samples. Triangles, wild samples; circles, cultivars; triangles with black outlines, WildC-2 samples with species identification indicated by color. Wild-collected *B. cretica* samples from Kioukis et al. (2020) indicated by asterisks, labeled as SRA. doi:10.1093/molbev/msab183
for modules 7, 13, 31, & 34; P value = 2.19e-11 for module 30). Post hoc comparisons using Tukey’s honestly significant difference (HSD) revealed that transcript abundance in cultivars was significantly different compared with that of wild relatives across conserved modules, except for B. hilaronis, which was not recovered as significantly different from cultivars for any module. WildC-2 along with other identified feral samples had significantly different transcript abundance compared with cultivars for modules 7, 13, 31, and 34, but not for module 30. Significant differences were also found between WildC-2 and feral samples compared with wild relatives for several modules, with no obvious patterns across modules (supplementary fig. S6 and table S5, Supplementary Material online).

**Species Tree and Admixture Inference Indicate**

_Brassica cretica_ Is the Closest Living Wild Relative

Given the results of population clustering using both SNPs and expression profiles, we further interrogated the species level relationships between wild relatives and cultivar groups by resolving the backbone of the phylogeny. Using the PoMo model (Schrempf et al. 2016) as implemented in IQ-Tree (Nguyen et al. 2015) and only including samples representing monophyletic groups as determined in the sample-level phylogeny, we found strong support for _B. cretica_ as the closest living wild relative to cultivated _B. oleracea_ (fig. 3A). Notably, for our species tree analyses, we included only one sample of _B. cretica_ (198). This sample was used for species reconstruction due to its placement near other wild taxa in the sample level phylogeny and its clustering with wild-collected _B. cretica_ from Kioukis et al. (2020) in the PCA. The current distribution of _B. cretica_ occurs throughout the Eastern Mediterranean, primarily in Greece, highlighting a potential origin of domestication (fig. 3B). Another suggested wild relative, _B. incana_, is strongly supported as belonging to the cultivar clade, sister to lacinato kale. Although our sampling is limited in regard to the distribution of _B. incana_ as a whole, this result supports our other findings that _B. incana_ is not a completely wild assemblage, but that at least some populations are feral. Within cultivars, several expected relationships were recovered: collards and cabbage as sister lineages (Song et al. 1988; Farnham 1996), with Brussels sprouts sister to both; cauliflower and broccoli as sister clades (Song et al. 1988; Stansell et al. 2018), with romanesco sister to both; and Chinese white kale as sister to all other cultivars, agreeing with recent literature (Cheng et al. 2016; Stansell et al. 2018).

With the overall species relationships resolved, we aimed to tease apart the evolutionary history of the wild samples that clustered within the cultivar clade. Specifically, we asked if any of the identified feral samples were the products of admixture using TreeMix (Pickrell and Pritchard 2012). Although the tree model without any migration edges explained 87.3% of the variance in the data set, sequentially adding migration events to the tree resulted in five migrations events explaining 92% of the variation (fig. 4A and supplementary fig. S7, Supplementary Material online). Adding a single migration edge resulted in an admixture event from _B. cretica_ (198) to a clade of [Chinese white kale + tronchuda cabbage]. To further test this event, we used four-population (_f4_) tests for treelessness as implemented in TreeMix, where a significant nonzero value indicates the presence of gene flow (Reich et al. 2009; Pickrell and Pritchard 2012; fig. 4B).

Although the tree {tronchuda cabbage, kohlrabi}|{B. cretica (198), B. hilaronis] showed no significant evidence of gene flow (_f4_ = 0.0008, Z = 1.094), replacing tronchuda cabbage with Chinese white kale indicated significant gene flow from _B. cretica_ (198) to Chinese white kale (_f4_ = −0.0055, Z = −5.113). This result was further verified when adding a second migration edge, as the migration edge only included Chinese white kale, but the direction was reversed (from Chinese white kale to _B. cretica_ [198]). The second event, from kohlrabi to a presumably feral sample of _B. cretica_ (199), was supported by _f4_ tests, with the tree {kohlrabi, _B. cretica_ (196), [B. cretica (199), marrow cabbage]] indicating significant evidence of gene flow from kohlrabi to _B. cretica_ (199) (_f4_ = 0.012, Z = 10.5). This migration event is also seen phenotypically, as _B. cretica_ (199) has a swollen stem when grown to maturity. No significant evidence of gene flow was found when substituting _B. cretica_ (199) with _B. oleracea_ (175), which is not expected to be involved in the admixture event (_f4_ = 0.0023, Z = 2.68). Two admixture events provide evidence of potential exoferal origins for at least two samples, _B. oleracea_ (175) and _B. cretica_ (199). The four-population tree of {_[B. montana (222), curly kale], [B. oleracea (175), broccoli]_ indicates significant gene flow from _B. montana_ (222) to _B. oleracea_ (175) (_f4_ = 0.315, Z = 15.77), as does the tree of {tronchuda cabbage, Chinese white kale], [B. cretica (199), broccoli] for gene flow from Chinese white kale to _B. cretica_ (199) (_f4_ = −0.009, Z = −7.98). The fifth added migration edge from _B. rupestris_ to WildC-2 explains the shared ancestry recovered in the fastSTRUCTURE results. The test for treelessness with [curly kale, WildC-2], _B. rupestris, B. macrocarpa_] indicated significant admixture from _B. rupestris_ to WildC-2 (_f4_ = −0.006, Z = −6.50), but was nonsignificant when substituting WildC-2 with cauliflower (_f4_ = −0.0003, Z = −0.338). In general, these analyses highlight that the evolutionary history of _B. oleracea_ is characterized by many admixture events and lineages of exoferal origins.

**Archaeological and Literary Evidence Point to a Late-Holocene Domestication**

To further investigate the origins of domesticated _B. oleracea_, we surveyed archaeological, literary, and artistic evidence (supplementary tables S6 and S7, Supplementary Material online). The earliest reported claim of _B. oleracea_ comes from an archaeological collection from the Austrian Alps. This collection comprises three seeds dated to the Middle Bronze Age (ca. 3550–3350 years before present or BP; Schmidl and Oeggl 2005). However, the lack of illustrations and discussion of separation criteria from other _Brassica_ species makes us question the reliability of this species-level identification, as seeds of _Brassica_ species are difficult to tell apart. The only other find of similar antiquity is _B. oleracea_ seeds from the Late Bronze Age/Early Iron Age, identified by scanning electron microscopy and radiocarbon dated directly between ca. 3250–2970 BP (Kaniewski et al. 2011). These finds
are associated with destruction levels at Gibala, Tell Tweini in western Syria on the Mediterranean coast. Although most of the archaeological finds are of seeds (supplementary table S6, Supplementary Material online), there is at least one documentation of pottery residues where lipids of *Brassica* leaf waxes were identified and dated to 850–750 BP (Evershed et al. 1992, 1994). The authors attribute this to the boiling of leaves of *B. oleracea*, and given the lack of evidence for other commonly eaten *Brassica* leaves in England at this time, this would appear a likely identification.

The earliest literary references to *B. oleracea* date to Greek scholars 2500–2000 BP (supplementary table S7, Supplementary Material online). Hipponax’s writing refers to a seven-leaf cabbage in an iambic verse (West 2011), whereas Hippocrates *On the Nature of Women*, written around 2410–2320 BP, refers to the use of cabbage, or
krambe, in a few recipes (Totelin 2009). As early as 2320 BP, there is evidence for cultivar diversity. Theophrastus refers to three varieties: a curly-leaved type, a smooth-leaved type, and a wild type with a bitter taste, many branches, and many small round leaves (Yonge 1854). Pliny in his Natural History writing some 200 years later describes at least ten varieties in addition to those seen in the previous classical works (The Elder and Rackham 1950). However, whereas most scholars accept that the Greek or Latin translations of “cabbage” refer to B. oleracea, it is important to note that “cabbage” is not a Greek word and that the word “raphanos” is translated as both cabbage and B. cretica in the Greek-English Lexicon (Liddell and Scott 1940) and in Hort’s (1916) translation of Theophrastus’ Historia Plantarum. Certainly, there are differences between the subspecies of B. cretica that might be reflective of the varieties described by Theophrastus and Pliny, and which may explain the diversity we observed among B. cretica samples in our PCA results. Further, the description by Nicander (quoted by Athenaeus; Yonge 1854; p. 582) indicates that wild or perhaps feral forms of B. cretica were known in Ionia, the western coast of present-day Turkey, ca. 2150–2050 BP.

Late-Holocene Environmental Niche Modeling

Highlights Wild Relatives’ Ranges

Based on archaeological information, the oldest relatively reliable occurrence for B. oleracea cultivation is dated 3250–2970 BP in Gibala NW Syria (Kaniewski et al. 2011). To predict what would be a suitable habitat for the wild relatives during the late-Holocene, we compiled occurrence records from GBIF (www.gbif.org) and (Snogerup et al. 1990), along with environmental data, to perform environmental niche modeling using MaxEnt 3.4.1 (Phillips et al. 2017). Notably, we find that B. cretica has an expanded Eastern Mediterranean habitat suitability (fig. 3C) that includes Cyprus. Presently, only B. hilarionis is known to occur in Cyprus (fig. 3B), however modeling predicts that in the late-Holocene it would also have had an expanded habitat suitability in the surrounding mainland coastal regions (fig. 3D). Since most of these wild species are narrow island endemics (Snogerup et al. 1990), species are generally estimated to have little change from current day distributions (supplementary fig. S8 and table S8, Supplementary Material online).

Discussion

Multiple Lines of Evidence Support a Single Eastern Mediterranean Origin

Our evidence from genome-scale, multilocus data along with archeology, literature, and environmental niche modeling best support a single Eastern Mediterranean domestication origin for B. oleracea, corroborating the conclusions of Maggioni et al. (2018) based on literary sources and (Maggioni et al. 2010) using linguistics. When modeling phylogeny and population structure, two Eastern Mediterranean species, B. cretica and B. hilarionis, are found as sister species to cultivars and are assigned ancestry from all cultivar populations for values of K from 2 to 5 (fig. 1), consistent with these species being likely progenitor species of B. oleracea cultivars. In our species tree reconstructions, we find just B. cretica as sister to all cultivars, specifically sample 198, which clusters with wild-collected B. cretica samples from Kioukis et al. (2020) in our PCA (fig. 2A and B), lending further support for B. cretica as the progenitor species. This same sample of B. cretica (198) as well as our sample of B. hilarionis are recovered as fairly homozygous, therefore they would likely be good starting material for future research related to de novo domestication via selective breeding or gene editing.

Although we do recover evidence of admixture between B. cretica (198) and both wild and cultivated taxa, the placement of B. cretica (198) as the closest living wild relative does not change. However, an inferred admixture event from B. cretica (198) to B. hilarionis does result in a topological change in the placement of B. hilarionis as sister to B. montana (224; originally collected in Spain) (supplementary fig. S7, Supplementary Material online). This novel relationship has not been identified before and warrants additional study with greater taxon sampling. The second migration event involving B. cretica (198) is from Chinese white kale. This event lends further evidence of admixture with wild germplasm during the domestication process, consistent with other examples demonstrating that domestication is not a single event, but a series of events characterized by continuous gene flow between wild and cultivated populations (Beebe et al. 1997; Wang et al. 2017). Together with the phylogeographic discontinuity of wild B. oleracea samples and their Eastern Mediterranean progenitors (fig. 3B), the more distant phylogenetic placement of B. insularis, B. macrocarpa, and B. villosa (fig. 3A), and strong patterns of shared ancestry between B. incana and cultivars (fig. 1), these results lead us to support the hypothesis of domestication in the Eastern Mediterranean with B. cretica as the closest living wild relative.

The Role of Ferality in the Domestication of Brassica oleracea

Multiple lines of evidence highlight the role of wild and feral populations as pools of diversity that contributed to crop diversification during domestication (Beebe et al. 1997; Allaby 2010; Fuller et al. 2014; Wang et al. 2017). Our data support a similar phenomenon in the domestication of B. oleracea: it appears that introgression from wild or feral populations contributed to the genetic composition of particular crops, and vice versa, which is revealed by in-depth analyses of admixture using population structure and tree-based methods (figs. 1 and 4; supplementary fig. S2 and S7, Supplementary Material online). Several samples of wild relatives, including B. cretica, as well as wild B. oleracea, B. incana, B. montana, and B. villosa, are recovered as feral in all analyses.

Although we find one sample of B. cretica (198) as the closest living wild relative, we also identify two samples of B. cretica (196 and 199) as likely feral and fall within the cultivar clade (fig. 1; see supplementary fig. S9, Supplementary Material online, for photos). Interestingly, Song et al. (1988) also recovered a polyphyletic B. cretica using RFLPs. Results presented here support previous findings that B. cretica was at one point at least partially domesticated.
Snogerup et al. (1990) state that wild B. cretica was consumed as late as 1962 and, as noted in our literary results, some early references to B. oleracea in the literature could be translated as B. cretica, meaning the vast amount of described morphology in these works, which may be the result of cultivation, could now be reflected in the multiple named subspecies and described genetic diversity of modern B. cretica (Snogerup et al. 1990; Widén et al. 2002; Allender et al. 2007; Edh et al. 2007). Further, B. cretica was known to occur in Ionia (western coast of present day Turkey) ca. 2150–2050 BP and the evidence of B. cretica populations today in Lebanon, which are morphologically similar to B. cretica subsp. nivea, suggests widespread trade of these species by the earliest Mediterranean civilizations (Dixon 2006). However, these plants may have been introduced into these localities without cultivation as was proposed by Snogerup et al. (1990). Previous researchers have noted that B. cretica populations are typically found in coastal locations associated with ancient seaports, occupying their preferred ecological niche on chalk cliffs undisturbed by grazing (Mitchell 1976; Snogerup et al. 1990). We believe that these early forms of B. cretica may have played underappreciated roles in the domestication of B. oleracea crops and to fully understand the evolutionary history of B. oleracea, the demographic history and domestication story of B. cretica must be resolved.

Sources have hypothesized that wild populations of B. oleracea in England are the progenitor(s) for modern cultivars (Snogerup et al. 1990; Song et al. 1990), whereas others have proposed that these are escaped cultivars (Mitchell 1976; Mitchell and Richards 1979). Consistent with these hypotheses, we find that the three wild B. oleracea samples in our study cluster with cultivars both phylogenetically and in PCA for both SNP data and expression profiles. Although these samples are from Canada (175), Denmark (176), and Germany (177), well outside the natural distribution range for B. oleracea—notably not from England, one of the hypothesized geographic origins—we suggest that an origin in England is unlikely given the archeological and literary data. Although the oldest archaeobotanical record for B. oleracea (Middle Bronze Age; ca. 3550–3350 BP) is from Austria, we regard this evidence with caution as wild populations of B. oleracea are not presently found in Austria and the major Brassica crops in this region include B. nigra (Tutin et al. 1964) or potentially cultivated turnip (B. rapa). Additionally, there is no compelling archaeological evidence to suggest the possible cultivation of cabbages in Europe prior to the Late Iron Age (2350–2050 BP) and Roman periods (1950–1650 BP), but there is evidence for knowledge of B. oleracea in Greece during this time (Maggioni et al. 2018; supplementary tables S6 and S7, Supplementary Material online). Overall, there are no records for B. oleracea from before this period within databases relating to the Eastern Mediterranean (Reif 2014), Europe (Kroll 2001, 2005), Britain (Tomlinson and Hall 1996), the Czech Republic (Kreuz and Schäfer 2002), or within predynastic and Pharaonic Egypt (Murray 2000), despite having documentation for other Brassica species. Evidence for B. oleracea in Europe does not start appearing until ca. 1850 BP, when the appearance of seeds increased and can be attributed to the spread of crops both within and on the periphery of the Roman Empire (Van der Veen 2011). Additionally, several studies that sampled wild B. oleracea populations in the British Isles (Mitchell 1976; Mitchell and Richards 1979), South West England (Raybould et al. 1999), Atlantic coasts of western Europe (Mittell et al. 2020), and Atlantic coast of France (Maggioni et al. 2020) support that these wild B. oleracea populations are feral populations, typically with low levels of genetic diversity and some degree of isolation from other populations. Lanner-Herrera et al. (1996) sampled populations across Spain, France, and Great Britain, concluding that each population evolved independently, whereas more recently, Mittell et al. (2020) found that geographically close populations were more genetically different than distant populations. Our results provide additional evidence that feralization is commonplace for B. oleracea crops and that references to wild B. oleracea likely represent multiple, independent feralization events. Additional sampling of wild populations will enable opportunities to further investigate the relationships among these feral populations and cultivated crops.

Brassica incana, another suggested progenitor species (Snogerup 1980), is also supported as feral for the samples included in our analyses. Two of our five samples (204 and 207) are recovered as sister to all cultivars in our individual level phylogeny but are found to share 100% of their ancestry with cultivars rather than other wild taxa using fastSTRUCTURE when K = 2 (fig. 1). Further, these two samples were resolved as sister to lacinato kale in our species tree analysis, providing additional evidence that these samples represent a feral lineage, possibly of lacinato kale. This result may lend insight into why previous studies have found B. incana as sister to B. oleracea (Lázaro and Aguinagalde 1998; Mei et al. 2010; Arias and Pires 2012) and the observation by Snogerup et al. (1990) that samples of B. incana from the Crimea are more interfertile with cultivated B. oleracea than others. Although Snogerup et al. (1990) suggested that B. incana was more interfertile due to historical introgression, we do not find evidence of this for samples 204 and 207. However, the three other samples of B. incana (205, 208, 209), which belong to WildC-2, do show evidence of admixture with B. rupestris, likely explaining their clustering together both in the PCAs and phylogeny with B. cretica (195) and B. villosa (233) which also show admixture with B. rupestris (see supplementary, fig. S10, Supplementary Material online, for photos). All three B. incana were collected in Italy from two locations and therefore do not well represent the known B. incana range (fig. 3B), whereas the two other samples found in this clade. B. cretica (195) and B. villosa (233), were collected in Greece and Italy, respectively. Although all five WildC-2 samples share an introgression event from B. rupestris (figs. 1 and 4; supplementary fig. S7, Supplementary Material online), they are from different germplasm collections (IPK-gatersleben and USDA National Plant Germplasm System), ruling out the inferred migration being the result of current cultivation practices. It is possible that at least three of these samples (B. incana 205, 208, 209) are related to the wild kale of Crimea, which is posited as a B.
rupestris–incana hybrid that was transferred to the Crimea via trade (Dixon 2006). This suggests that there was early widespread cultivation of these B. rupestris–incana types (Dixon 2006) and provides a plausible explanation for why B. incana and B. rupestris are closely related in previous studies (Lannér et al. 1997; Mei et al. 2010). The other two samples in WildC-2 (B. cretica 195 and B. villosa 233), possibly represent misidentifications, which is supported by their intermediate phenotypes (i.e., B. rupestris margins with varying amounts of trichomes; supplementary fig. S10, Supplementary Material online).

The last feral identification is that of B. montana, for which we find one sample as more closely related to wild taxa (224) and one more closely related to cultivars (222). The feral sample (222) is of unknown origin, but again the literature indicates that this may not be a surprising result. Many studies have previously indicated a close relationship between B. montana and B. oleracea. For example, Panda et al. (2003) concluded that B. montana may be a subspecies of B. oleracea, whereas Lannér et al. (1997) found that B. montana and B. oleracea clustered together using chloroplast data. Furthermore, several authors have suggested that some populations of B. montana were feral B. oleracea (Paolucci 1890; Onno 1933; Snogerup et al. 1990), which may be reflected in the overlapping ranges produced by our niche modeling of these two species (supplementary fig. S8, Supplementary Material online). Therefore, in combination with results from previous studies, our results support that at least some B. montana populations are of feral origin.

Taken together, it is clear that the current taxonomy of B. oleracea and its wild relatives is confounded by gene flow between wild and cultivated populations, resulting in confusion between wild and feral lineages and obscuring the true evolutionary history of this species. Additionally, although there is much interest in crop improvement using CWRs (Meyer et al. 2012; Khoury et al. 2020), feral lineages offer another, potentially more direct route to reintroducing genetic diversity into cultivated populations, as gene flow is less likely to be impeded by barriers such as reproductive isolation (Mabry et al. 2021). These feral populations may also provide additional avenues to explore the evolutionary capacity for range expansion and phenotypic plasticity.

Postdomestication Cultivar Relationships

Although our knowledge of the spread and diversification of B. oleracea crops after domestication is confounded by both the difficulties of identifying seeds of individual crop types and frequent introgression between crop types, we can infer some patterns using the species phylogeny. Like other studies (Cheng et al. 2016; Stansell et al. 2018), we find Chinese white kale sister to all other cultivars, representing the only Asian clade of crop types (fig. 3A). Although the spread of B. oleracea to eastern Asia is still undocumented archaeologically, recent pollen analysis has provided evidence for cultivation of other Brassica species, including B. rapa, in the Yangtze valley 3250–3350 BP, likely corresponding to movement across “Silk Road” trade routes (Zhang 2009). However, this only provides identification criteria, not archaeological evidence (Yang et al. 2018). A review of Chinese historical sources concluded that B. oleracea may have been introduced to China 1450–1350 BP and had evolved into Chinese white kale in Southern China by the period of the Tang Dynasty (1350–1250 BP; Zhang 2009). Due to its position as sister to all other cultivars and as the only Asian B. oleracea crop type, as well as its annual growth habit, this taxon warrants additional study to understand its own unique domestication story.

The dispersal of B. oleracea by human translocation westward, ultimately to the Atlantic coast of Europe, appears to have established both regional feral populations and the variety of modern crop types. Archaeological evidence suggests that this process may have begun with Late Bronze Age seafaring (3000–3300 years ago), when the whole Mediterranean became linked in trade perhaps for the first time (Broodbank 2015), and continued to provide a corridor for introgression and varietal diversification through the Iron Age (up to 2000 years ago). Trade links along the Atlantic seaboard from North Africa and Iberia through Britain and Ireland are clearly indicated in archaeology (Cunliffe 2004), and are associated with the first peopling of the Canary Islands from the north, where walking stick kale is endemic. Notably, many cultivars do not form monophyletic groups in our sample level phylogeny, likely indicative of admixture between crop types. This is supported by previous findings that broccoli is paraphyletic (Song et al. 1988; Stansell et al. 2018), as well as collards (Pelc et al. 2015), and by our findings that kale types such as tronchuda kale and perpetual kale are highly polyphyletic, suggesting that the kale morphotype has been selected for multiple times independently.

In conclusion, we confirm a single Eastern Mediterranean origin for B. oleracea and find B. cretica as the closest living wild relative. We highlight several feral samples that are not reflected by the current taxonomy but likely reflect important aspects of the domestication history for B. oleracea. Moving forward, it will be important to identify, collect, study, and preserve these feral samples as pools of allelic diversity, which may play an important role in future crop improvement, for example, as a source of potential pest and pathogen resistance (Mithen et al. 1987; Mithen and Magrath 1992; Mohammed et al. 2010). In clarifying the evolutionary history of B. oleracea and its wild relatives, we hope to enable this model system for additional studies on evolutionary phenomena such as parallel selection, polyploidy, and fertility. Additionally, since many of these wild species are very narrow endemics and are valuable for both crop improvement and for nature conservation, their identification and preservation are urgent. We hope this study can serve as a steppingstone, as the work before us has, for those who, like Darwin was, are intrigued by this group of plants and wish to further its study.

Materials and Methods

Taxon Sampling

Samples from cultivars accounted for 188 of the 224 total samples with the remaining 36 samples included being previously identified wild relatives (supplementary table S2, Supplementary Material online). These include accessions...
from the United States Department of Agriculture, Agriculture Research Service (USDA-ARS) Plant Genetic Resources Unit (PGRU; 114 accessions), The Leibniz Institute of Plant Genetics and Crop Plant Research (IPK; 71 accessions), Universidad Politécnica de Madrid (UPM; four accessions), The Nordic Genetic Resource Centre (NordGen; two accessions), Gomez Campo Collection (two accessions), John Innes Center (one accession), doubled haploid lines (17 samples, some accessions sampled twice), or from the Pires’ personal collection (13 accessions). Four replicates of each accession were grown from seed in a sterile growth chamber at the University of Missouri (MU; Columbia, MO) Bond Life Sciences Center in a randomized complete block design across two independent outgrowths. At the seventh leaf stage, leaf four was collected from each plant and immediately flash-frozen in liquid nitrogen for RNA extraction. Morphotype identity was validated in mature plants by growing all accessions twice over the span of 2 years (supplementary table S2, Supplementary Material online).

Whole-genome resequencing data for an additional four samples from Kioukis et al. (2020) of two varieties of B. cretica (var. cretica and var. nivea) was downloaded from the National Center for Biotechnology Information (NCBI) Sequence Read Archive (SRA) to supplement our sampling of B. cretica. These samples are under the SRA accession as follows: A=SRR9331103, B=SRR9331104, C=SRR9331105, and D=SRR9331106. Samples of A and B are B. cretica var. nivea from mainland Greece and C and D are B. cretica var. cretica, one from the mainland (C) and one from the island of Crete (D).

RNA Isolation and Sequencing
RNA was isolated using the ThermoFisher Invitrogen PureLink RNA mini kit (Invitrogen, Carlsbad, CA) followed by TruSeq library preparation (Illumina, San Diego, CA) and sequencing on the NextSeq platform (Illumina, San Diego, CA) for 2 x 75 bp reads. Library preparation and sequencing were performed through the MU DNA Core Facility. For eight flow cells, 24 samples were multiplexed and sequenced in a single flow cell, followed by a ninth flow cell with 17 samples, and a tenth flow-cell with 16 samples.

Mapping and SNP Calling
Short reads were mapped to the B. oleracea TO1000 genome (Chinese white kale; Parkin et al. 2014; release-41) by first using the STAR v. 2.5.2 (Dobin et al. 2013) two-pass alignment to identify splice junctions, which were then used in the second pass to improve mapping (Engström et al. 2013). The TO1000 genome of Chinese white kale was chosen due to wild relatives having a more kale-like phenotype and its placement as sister to the other cultivars in recent studies (Cheng et al. 2016; Stansell et al. 2018). Mapped reads (BAM format) were then processed following the GATK v. 3.8 best practices for RNA-seq reads (McKenna et al. 2010; Van der Auwera et al. 2013; Poplin et al. 2017). To ensure that reads were mapping correctly, the GATK “SplitNTrim” function was used to split reads into exon segments and trim any overhanging reads in intron segments. In total, 7,564,168 variants were called before any filtering was performed. The resulting variants were filtered to exclude those with a Fisher strand (FS) value greater than 30 and quality depth (QD) less than 2.0. The remaining 879,865 chromosomal variants were then filtered using vcftools v. 0.1.17 (Danecek et al. 2011) to exclude sites with greater than 60% missing data (~max-missing 0.4), sites with mean sample depth values less than 5 (~min-meanDP 5), and indels (~remove-indels) resulting in a total of 103,525 SNPs. Finally, SNPs were filtered for linkage disequilibrium (LD) using using PLINK v. 1.90 with a window size of 80 kb, or about two times the estimated length for 80% LD decay (Cheng et al. 2016), a step size of 5 kb, and a variance inflation factor of 2 (~indep 80 kb 5 2; Purcell et al. 2007), for a final data set of 36,750 SNPs. The four B. cretica genome resequencing samples (Kioukis et al. 2020) were also mapped to the B. oleracea TO1000 genome (Chinese white kale; Parkin et al. 2014; release-41), using BWA (Li and Durbin 2009).

Phylogenetic and Introgression Inference
To test how the different populations are related to one another and which wild relative is most closely related to the cultivated types, we used three different phylogenetic programs: SNPhylo v. 20160204 (Lee et al. 2014) to assess individual sample relationships, IQ-Tree v. 1.6 (Nguyen et al. 2015) to test species level relationships, and TreeMix v. 1.13 (Pickrell and Pritchard 2012) to assess introgression. For SNPhylo (Lee et al. 2014), we ran analyses using an r² cutoff of 0.1 for LD, minor allele frequency ≥0.01, proportion of missing sites ≤0.4, 1,000 bootstrap replicates, and rooted with sample 238 (B. villosa). For IQ-Tree, we used the Polymorphism-aware phylogenetic Models (PoMo) software (Schrempf et al. 2016; -m GTR+P) to perform phylogenetic comparisons using population genetic data, using 1,000 bootstrap replicates via the ultrafast bootstrap approximation method (Hoang et al. 2018) and B. villosa to root the tree. For our IQ-Tree analysis, we subsampled data to include only those samples which were recovered as monophyletic in our SNPhylo tree (supplementary table S2, Supplementary Material online; samples with asterisks). To test both the topology of relationships and for gene flow between populations, we used TreeMix with the following parameters: no sample size correction (-nom), rooted with B. villosa (-root villosa), bootstrapping over blocks of 500 SNPs (-bootstrap -k 500), and to incorporate between two and ten migration events (-m). TreeMix (Pickrell and Pritchard 2012) was run with samples of B. cretica, B. incana, B. montana, and B. oleracea as individuals, but used samples found in WildC-2, cultivars, and wild relatives as populations. Four-population (f4) tests for treeness (Reich et al. 2009; Pickrell and Pritchard 2012) were used to test the support of the inferred migration edges from TreeMix (Pickrell and Pritchard 2012) via the fourpop method.

Population Structure and Variation
To test ancestry proportions and identify the likely genetic structure of described populations we used fastSTRUCTURE v. 1.0 (Raj et al. 2014). We tested K values from 2 to 8 using default convergence criteria and priors followed by the
of the native range. From the Snogerup et al. (1990) data, we omitted records that could not be georeferenced to <5 km spatial uncertainty. Populations of B. cretica in Lebanon and Israel and of B. incana in Crimea are thought to be likely early human introductions (Snogerup et al. 1990) and records from these areas were omitted. Occurrences above 1200 m altitude were also omitted, as these species rarely occur above 1,000 m and observations above these altitudes may represent anthropogenic dispersals to disturbed areas or misidentifications. To minimize sampling bias due to clustered observations (Beck et al. 2014; Boria et al. 2014), we thinned the filtered occurrences to records greater than or equal to 10 km apart using the “spThin” package in R (Aiello-Lammens et al. 2015).

Clustering Based on Expression Profiles

First, Salmon v. 1.2.1 (Patro et al. 2015) was used to acquire transcript abundances for each sample and the estimated number of reads originating from transcripts. The input for expression profile analysis was prepared using tximport (Soneson et al. 2015) with design=--plantout+cultivar type. Correction for library size (estimateSizeFactors) and variance-stabilizing transformation (vst) was performed in DESeq2 v. 1.28.1 (Love et al. 2014). To test for clustering based on expression profiles, we ran a PCA on the normalized expression values and performed clustering based on Euclidean distance using the “prcomp” and “hclust” functions, respectively, in the “stats” v. 3.6.2 package for R v. 3.6.0 (R Core Team 2018). To assess networks of genes driving differences observed in the PCA, we used WGCNA v. 1.68 (Langfelder and Horvath 2008). Following (Zhang and Horvath 2005), we found that a soft-thresholding power of nine was best as it was the lowest power that satisfied the approximate scale-free topology criterion, resulting in 48 modules of genes.

To determine biological processes which were overrepresented in the resulting modules, A. thaliana orthologs of B. oleracea were determined using both synteny and BLAST. Synteny-based annotations were extracted from table S7 in Parkin et al. (2014), whereas the BLAST annotation was performed using blastn in BLAST v. 2.10.0+- (Camacho et al. 2009). The B. oleracea CDS database was downloaded from https://plantsensembl.org/Brassica_oleracea/Info/Index (last accessed September 2020), and the A. thaliana CDS database from Araport11_genes.201606.cds.fasta from https://www.arabidopsis.org/ (last accessed September 2020). The blastn parameters were -value 1E-6 -max_target_seq 1. Genes determined using synteny were then used to perform a GO analysis through PANTHER v. 16.0 (Mi et al. 2021). ANOVAs were used to test for differences in transcript abundance among cultivars, ferals (including WildC-2), and wild relatives by using the “aov” function in R v. 3.6.0 (R Core Team 2018) followed by multiple comparisons with Tukey’s HSD using the function “glht.”

Environmental Niche Modeling

We compiled occurrence records for wild relatives from the Global Biodiversity Information Facility (GBIF, www.gbif.org) data portal and data from Snogerup et al. (1990). From the GBIF data, we omitted records that were duplicated, lacked location data and/or vouchers, were collected from the grounds of botanical gardens, and that were clearly outside

**Supplementary Material**

Supplementary data are available at Molecular Biology and Evolution online.

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**Author Contributions**

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**Data Availability**

The sequences reported in this article have been deposited in the Sequence Read Archive database (accession number PRJNA544934; https://www.ncbi.nlm.nih.gov/bioproject/PRJNA544934). The resulting transcript abundances and VCF file are deposited at DRYAD (https://doi.org/10.5061/dryad.5mkkw763). Scripts used can be found on github (https://github.com/mmabry/Brassica-oleracea-Population-and-Phylogenetics).

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