CASE REPORT

A Japanese case of chronic lymphocytic leukemia with t (1;6)

Kayo Harada1, Kazuhiko Ikeda1*, Hayato Matsumoto1, Miki Furukawa1, Hiroshi Takahashi1, Hiroshi Ohkawara1, Hideyoshi Noji1, Kazuhiro Tasaki2, Masafumi Abe2, Kazuei Ogawa1 and Yasuchika Takeishi1

Abstract

Chronic lymphocytic leukemia (CLL) rarely exhibits an aggressive clinical course and its patients often have chromosomal deletions or additions. Furthermore, reciprocal translocations are barely observed in CLL. There have only been a few reports of CLL with t(1;6), and here we report the first Asian case of CLL with reciprocal translocation t(1;6). Since our case and previously reported CLL patients with t(1;6) consistently showed aggressive clinical course, t(1;6) may define a distinct type of CLL.

Keywords: CLL, T(1;6), Aggressive clinical course

Introduction

Chronic lymphocytic leukemia (CLL) is a mature B-cell neoplasm characterized by increased abnormal non-large CD5+CD20+CD23+ B cells in peripheral blood, bone marrow, and lymphoid organs [1,2]. CLL usually exhibits an indolent clinical course, but sometimes shows aggressive clinical course, including Richter’s transformation with poor outcome. Poor prognosis in CLL is associated with expression of CD38 or zeta-chain associated protein-70 (ZAP-70), mutations and/or expression of tumor protein p53 (TP53), and abnormal cytogenetics [3]. The majority of patients with CLL have chromosomal deletions or additions, but chromosomal translocations are rare, and if present, they are usually associated with the loci of immunoglobulin (IG) genes, 14q32, 2p13, or 22q11 [4]. Also, information on the roles of these translocations in clinical outcome or pathogenesis is extremely limited in CLL.

There are only a few reports of translocation t(1;6) involving chromosome 6p25 ~ 23 in patients with CLL [5-9]. This type of translocation was observed in approximately 0.5% of patients with CLL in the Belgian databases [5]. These cases showed an aggressive clinical course. Furthermore, when treated with purine analogues, patients often develop Richter’s transformation. It has been suggested that a regulator of B-cell differentiation, interferon regulatory factor 4 (IRF4) gene on 6p25, correlates with the pathogenesis of CLL cases with t (1;6) [5].

Here, we report the first Asian case of CLL with translocation t(1;6) where CLL cells expressed IRF4. The patient showed an aggressive clinical course and was successfully treated with conventional chemotherapy without purine analogue.

Case report

A 56-year-old Japanese male, who had taken medical laboratory examinations every half-year, was first found to have a slightly elevated peripheral white blood cell count (12 x 10^9/L) in July 2010, while showing no other abnormal findings on blood examination and systemic positron emission tomography using 18 F-fluorodeoxyglucose. However, only 2 months later he noticed a systemic swelling of lymph nodes, which further rapidly enlarged, and he developed a high body temperature and night sweat. He had a supraclavicular lymph node biopsy in November 2010, and was considered to have advanced B-cell lymphoma because CD20+ lymphoid cells proliferated. Based on this finding, he was referred to our department for the purpose of chemotherapy. On admission, he had mild splenomegaly and leukocytosis (17 x 10^9/L), and his platelet count and hemoglobin concentration were normal. Abnormal lymphoid cells were also present in the bone marrow (Figure 1). The
proportion of prolymphocytes and lymphocytes in the bone marrow were 2.8% and 67.2%, respectively. He immediately received chemotherapy consisting of cyclophosphamide, vincristine, doxorubicin, dexamethasone, and rituximab (R-hyper-CVAD) [10,11], resulting in complete remission. After finishing this initial treatment, it turned out that the majority of proliferated lymphoid cells in the samples taken before treatment were positive for CD5, CD23, CD38, B-cell lymphoma (BCL)-2, as well as CD20, and negative for CD3, CD10, CD56, Cyclin D1, Cyclin D2, TP53, and terminal deoxynucleotidyl transferase in immunohistochemistry and/or flow cytometry. Both IRF-4 and ZAP-70, but not TP53, were strongly expressed in the lymphoid cells (Figure 1), and approximately 10% of the cells were positive for Ki67. We then reconfirmed that there were few large cells in each sample of peripheral blood, bone marrow, and the biopsied lymph node. Based on these morphological findings and the immune phenotype, he was diagnosed with CLL without the Richter’s transformation. According to the diagnosis of CLL, clinical stage before the initial treatment was reestablished as the Binet B and Rai II. Cytogenetic analysis with the G-banding method without the addition of mitogen revealed a chromosomal translocation, 46,XY,t(1;6)(p34.1;p23) in 10 of 20 analyzed metaphase cells (Figure 2). No other abnormality was found with chromosomal analysis with G-banding, and del11q was not detected in FISH analysis. He received a total of 6 courses of R-hyper-CVAD and high-dose chemotherapy with ranimustine, cytarabine, etoposide, and melphalan, followed by autologous peripheral blood stem cell transplantation (auto-PBSCT). Since then, his CLL has not relapsed for 20 months.

Discussion
Chromosomal translocations are rare but usually involve the IG genes in CLL [4]. However, our patient showed the reciprocal chromosomal translocation t (1;6)(p34.1; p23), which is not associated with the IG genes. So far, there have been a few reports on t (1;6) in CLL, including a study on 8 patients with the t(1;6)(p36 ~ 33; p25 ~ 23) found in the Belgian Cytogenetic Institutes database [5]. The 8 patients showed an advanced and progressive clinical course, with three of them being complicated with Richter’s transformation. In addition, most of these patients required conventional chemotherapy shortly after diagnosis. Our patient also showed rapid progression of the disease after the sudden onset. CLL cases with t (1;6)(p35;p25) in the complex karyotype have also been reported [7,8]. Thus, CLL with t (1;6) may be of a distinct type. On the other hand, there have been more CLL cases with t(1;6) who had different breakpoints such as 1q25, 1q21, 6p11 and 6p12 [9], but the features of these cases remain unknown.

The CLL cells in our case significantly expressed IRF4 in immunohistochemistry. CLL cells are known to frequently express IRF4 [12], which was also implicated in the pathogenesis of CLL with t(1;6) [5]. However, IRF4 expression in the patients with t(1;6) has not been shown in the previous reports. Overexpression of IRF4
mRNA is rarely associated with a somatic mutation in the IRF4 gene [13], but unfortunately, genetic analysis for IRF4 was not performed in our case. Whether chromosomal rearrangement of t(1;6) is inducible to express IRF4 as well as the somatic point mutation in the IRF4 gene remains unclear.

Our case also showed robust expression of ZAP-70 (Figure 1), which is associated with poor prognosis and aggressive clinical course in CLL. The immunoglobulin variable heavy chain (IgVH) gene was not investigated in our case, as it is known that the expression of ZAP-70 is strongly correlated with the unmutated IgVH gene [14]. Interestingly, almost all CLL patients with t(1;6) exhibited the unmutated IgH gene [5]. As for other predictors of poor prognosis, CD38, but not TP53 or Ki67, was significantly expressed in our case.

The most common treatment for CLL is chemotherapy with a purine analogue, although high-dose chemotherapy with auto-PBSCT may be another option if a patient responds to conventional chemotherapy [15]. In the previous report of CLL with t(1;6) by Michaux et al. [5], 2 of 8 patients also underwent high-dose chemotherapy with auto-PBSCT. Notably, both of them achieved complete remission and their disease did not relapse during observation. In contrast, of the 8 CLL patients with t(1;6), patients who received purine analogues were especially complicated with Richter’s transformation. Proceeding from these findings and good response to the initial treatment, our patient received a total of 6 courses of R-hyper-CVAD followed by high-dose chemotherapy with auto-PBSCT. However, owing to the limitation in the number of patients, appropriate treatment for CLL with t(1;6) remains to be determined.

In conclusion, we report the presence of CLL with t(1;6) in Asians as well as Europeans, suggesting that t(1;6) may define a distinct type of CLL. We should anticipate chromosomal abnormalities such as t(1;6) in CLL as the prognosis may be affected.

Consent
Written informed consent was obtained from the patient for publication of this Case report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

Competing interests
The authors declare no conflict of interest.

Authors’ contributions
KH, KI, KO, YT: wrote the manuscript, treated patients. HM, MF, HT, HO, HN: treated patients, analyzed data. KT and MA: performed histopathological analysis. All authors read and approved the final manuscript.

Author details
1Department of Cardiology and Hematology, Fukushima Medical University, 1 Hikariga-oka, Fukushima, Fukushima 960-1295, Japan. 2Department of Pathology and Diagnostic Pathology, Fukushima Medical University, Fukushima, Japan.

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References
1. Byrd JC, Stilgenbauer S, Flinn IW: Chronic lymphocytic leukemia. Hematology Am Soc Hematol Educ Program 2004, 2004:163–183.
2. Swerdlow SH, Campo E, Harris NL, et al: WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues. 4th edition. Lyon: IARC Press; 2008.
3. Dighiero G, Hamblin TJ: Chronic lymphocytic leukaemia. Lancet 2008, 371:1017–1029.
4. Willis TG, Dyer MJ: The role of immunoglobulin translocations in the pathogenesis of B-cell malignancies. Blood 2000, 96:808–822.
5. Michaux L, Wlodarska I, Rack K, Stul M, Criel A, Maerevoet M, Marchal S, Demuynder H, Mineur P, Kargar Samani K, Van Hoof A, Ferrant A, Marynen P, Hagenbeijer A: Translocation t(1;6)(p35.3;p25.2); a new recurrent aberration in “unmutated” B-CLL. Leukemia 2005, 19:77–82.
6. Van Den Neste E, Robin V, Francart J, Hagenbeijer A, Stul M, Vandenberghhe P, Delannoy A, Sonet A, Denex V, Costantini S, Ferrant A, Robert A, Michaux L: Chromosomal translocations independently predict treatment failure,
treatment-free survival and overall survival in B-cell chronic lymphocytic leukemia patients treated with cladribine. Leukemia 2007, 21:1715–1722.

7. Schultz RA, Deloukina M, Gaal K, Bedell V, Smith DD, Forman SJ, McDaniell LD, Ballif BC, Shaffer LG, Slovak ML. Evaluation of chronic lymphocytic leukemia by BAC-based microarray analysis. Mol Cyogenet 2011, 4:4.

8. Rodrigues Pereira Velloso ED, Borri D, Alonso Ratis C, Fleury Perin G, Hamerschlag N, Bacal NS, Silveira PAA, Bozera AMPS, Pasqualini DC. Chronic lymphocytic leukaemia/Small lymphocytic lymphoma (CLL/SLL) associated with translocation t(1;6)(p35;p25) as part of complex karyotype. Atlas Genet Cyogenet Oncol Hemaotol 2011, 15:467–469.

9. Mayr C, Speicher MR, Kofler DM, Buhlmann R, Strehl J, Busch R, Hallek M, Wendtner CM. Chromosomal translocations are associated with poor prognosis in chronic lymphocytic leukemia. Blood 2006, 107:742–751.

10. Romaguera JE, Fayad L, Rodriguez MA, Broglio KR, Hagemeister FB, Pro B, McLaughlin P, Younes A, Samaniego F, Goy A, Saris AH, Dang NH, Wang M, Brasley V, Medeiros LJ, Katz RL, Gagneja H, Samuels BI, Smith TL, Cabanillas FF. High rate of durable remissions after treatment of newly diagnosed aggressive mantle-cell lymphoma with rituximab plus hyper-CVAD alternating with rituximab plus high-dose methotrexate and cytarabine. J Clin Oncol 2005, 23:7013–7023.

11. Thomas DA, Faderl S, O'Brien S, Bueso-Ramos C, Cortes J, Garcia-Manero G, Giles FJ, Verstovsek S, Wierda WG, Pierce SA, Shan J, Brandt M, Hagemeister FB, Keating MJ, Cabanillas F, Kantarjian H. Chemoimmunotherapy with hyper-CVAD plus rituximab for the treatment of adult Burkitt and Burkitt-type lymphoma or acute lymphoblastic leukemia. Cancer 2006, 106:1569–1580.

12. Chang CC, Lorek J, Sabath DE, Li Y, Chitambar CR, Logan B, Kampalath B, Cleveland RP. Expression of MUM1/IRF4 correlates with clinical outcome in patients with B-cell chronic lymphocytic leukemia. Blood 2002, 100:4671–4675.

13. Havelange V, Pekarsky Y, Nakamura T, PALAMARCHUK A, Alder H, Rassenti L, Kipps T, Croce CM. IRF4 mutations in chronic lymphocytic leukemia. Blood 2011, 118:2827–2829.

14. Rassenti LZ, Huynh L, Toy TL, Chen L, Keating MJ, Gribben JG, Neuberg DS, Flinn IW, Rai KR, Byrd JC, Kay NE, Greaves A, Weiss A, Kipps TJ. ZAP-70 compared with immunoglobulin heavy-chain gene mutation status as a predictor of disease progression in chronic lymphocytic leukemia. N Engl J Med 2004, 351:893–901.

15. Sutton L, Chevet S, Tounilhac O, Diviné M, Leblond V, Corront B, Leprêtre S, Eghbali H, Van Den Neste E, Michallet M, Maloisel F, Bouabdallah K, Decaudin D, Benhouc C, Brice P, Gonzalez H, Chaparo E, Radford-Weiss I, Leporrier N, Maloum K, Nguyen-Khac F, Davi F, Lejeune J, Merle-Béal H, Leporrier M. Société Française de Greffe de Moelle et de Thérapie Cellulaire (SFGRM-TC) and Groupe Français d’étude de la Leucémie Lymphoïde Chronique (GFLLC): Autologous stem cell transplantation as a first-line treatment strategy for chronic lymphocytic leukemia: a multicenter randomized controlled trial. Blood 2011, 117:6109–6119.