A 65-year-old woman presented to the emergency department with respiratory distress, congestive heart failure and atrial fibrillation thought to be secondary to pneumonia and myocardial infarction. Intubation was performed, and she was admitted to the intensive care unit. A nasogastric feeding tube was placed and appropriate therapy initiated. The patient had numerous comorbidities, including end-stage renal disease, diabetes mellitus, stroke, myocardial infarction, peripheral vascular disease, hypertension and emphysema, and she was receiving hemodialysis. An examination of her head performed on admission did not reveal any abnormalities.

Three days later, numerous maggots (6–7 mm long) were observed in the patient’s nasal cavities (Figure 1) and at the left medial canthus (Figure 2). During this period, an adult fly (a green blow fly or a green bottle fly) was seen in the intensive care unit. The patient was in a windowless private room that had an open door. The intensive care unit had no windows to the outdoors; thus, it was assumed that the fly had entered the intensive care unit from another part of the hospital.

A magnetic resonance imaging scan of the head revealed slight thickening of the nasal mucosa. An ophthalmologist manually removed about 20 larvae from the canthus. None of the larvae had invaded the lacrimal duct. Using saline, an otolaryngologist irrigated about 100 larvae from the nasopharynx. There was no evidence of deeper tissue penetration. Cefotaxime was administered for 5 days as prophylaxis against bacterial complications. The patient recovered fully and was later discharged.

The larvae that were removed from the patient’s canthus and nasopharynx were identified by 2 of us (B.S. and B.G.) as second instar *Lucilia illustris* (green blow fly) of the family Calliphoridae. *Lucilia* species are primarily scavengers of vertebrate carrion (e.g., exposed meat and fish), and several species are important agents of myiasis in livestock and humans. Facultative myiasis occurs when flies lay their eggs in decaying tissue or in malodorous open wounds or orifices, including the nasopharynx and eye structures, as occurred in our patient.

Female blow flies can lay up to several hundred eggs in a mass. The eggs and larvae usually develop on ephemeral substrates, and they have evolved rapid life cycles, the speed of which is dependent on temperature.
The larvae molt 3 times, and after feeding for 1–10 days, they pupate in the surrounding media. Based on the incubation and development times of a related species, second instar larvae should take 2 days to develop; thus, in our patient’s case the infestation was nosocomially acquired.

Nosocomial myiasis is rare. Predisposing risk factors include debilitation, blood or odors of decomposition, inattention to proper nursing care and summer season. Altered consciousness or hypoesthesia may prevent detection of a fly, and paralysis may prevent the patient from fending it off. Finally, comorbidities that compromise the vascular supply, such as diabetes, peripheral vascular disease and coronary artery disease, have been found in a high proportion of cases. Our patient had all of these comorbidities. Although she had no apparent predisposing eye or nasal wounds, unvisualized bleeding around the nasogastric tube and halitosis from underlying pneumonia may have contributed to the infestation.

Larvae in the Calliphoridae family can invade local tissues with minimal damage to deeper structures. However, numerous complications may occur with larvae of certain species, including erosion of bones and teeth, cellulitis, bacteremia and death. Treatment involves manual removal of the larvae and thorough irrigation of the affected tissues with saline or a dilute antimicrobial solution (e.g., hydrogen peroxide). Systemic antiparasitics are generally not required.

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