Microfibrillar-associated protein 5 is linked with markers of obesity-related extracellular matrix remodeling and inflammation

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Objective: Microfibrillar-associated protein 5 (MFAP5) is an extracellular matrix (ECM) glycoprotein, which is colocalized with microfibrils in elastin networks. Its function in adipose tissue (AT) is not known. We have recently shown that the expression of MFAP5 is downregulated in AT along with weight reduction (WR) in persons with metabolic syndrome (MetS). The aim of this work was to study whether the change of MFAP5 mRNA expression in response to WR is associated with markers of adiposity, glucose metabolism and insulin resistance in human AT.

Design: Weight reduction intervention study in parallel study design (The Genobin study). Altogether 46 obese subjects with impaired glucose tolerance and features of MetS were randomized to a WR (n = 28) or a control group (n = 18) lasting for 33 weeks.

Measurements: Circulating glucose and insulin concentrations were measured and subcutaneous AT biopsies were performed before and after the intervention. The mRNA expression was studied by quantitative real-time PCR (QPCR).

Results: QPCR of human AT biopsy samples confirmed that MFAP5 is highly expressed in AT and its expression is decreased during WR. The mRNA expression of MFAP5 correlated positively with body mass index, and the change in MFAP5 mRNA expression during WR correlated positively with the change of body fat mass. Furthermore, the MFAP5 mRNA expression correlated negatively with circulating fasting concentrations of adiponectin and interleukin (IL)-1β and positively with leptin, insulin and IL-1Ra levels. In addition, the MFAP5 mRNA expression correlated positively with the mRNA expressions of peroxisome proliferator-activated receptor gamma, cyclin D2 and A disintegrin and metalloproteinase domain 12, genes involved in AT remodeling.

Conclusion: This study demonstrates that MFAP5 is highly expressed in human AT and is correlated with markers of insulin resistance. Furthermore, it is possible that MFAP5 is related to ECM remodeling during development of obesity.

Keywords: MFAP5; adipose tissue; insulin resistance; ECM remodeling

Introduction

Obesity is a chronic low-grade inflammatory state, which is characterized by an increase in circulating inflammatory factors partly due to changes in cytokine and adipokine production in adipose tissue (AT).¹ In addition to mature adipocytes, AT is composed of different cell types in the stromavascular fraction¹ that may have different roles in obesity-related inflammation of AT. It has been suggested that a proinflammatory state with a concomitant upregulation of inflammation-related genes might lead to changes in the expression of genes linked to the extracellular matrix (ECM) in order to accommodate the growing AT.² Weight reduction (WR) is an effective way to reverse the state of inflammation and decrease biological markers of inflammation in the circulation, and also the expression of inflammatory markers in AT.³ Genome-wide transcriptomics analysis performed from AT of the subjects with metabolic syndrome (MetS) participating in the Genobin study⁴ showed that the ECM-associated gene, microfibrillar-associated protein 5 (MFAP5),⁵ also known as MAGP2, was one of the genes, whose expression was downregulated in subcutaneous AT along with WR and improved insulin sensitivity. It was found that MFAP5 mRNA
was highly expressed in AT, although its function in human AT is not known. In this work, our aim was to investigate whether the change of MFAP5 gene expression, along with WR, is correlated with the measures of glucose metabolism and body adiposity in the Genobin study individuals and with circulating adipokines and expression of genes, which were changed after WR.

Materials and methods

Altogether 75 overweight/obese (body mass index (BMI) 28–40 kg m\(^{-2}\)) subjects aged 40–70 years were recruited into the Genobin study (NTC00621205) described in detail earlier.\(^4\) The subjects had impaired fasting glucose or impaired glucose tolerance test and by frequently sampled intravenous glucose tolerance test, according to the Minimal Model analysis. Briefly, the TRIzol method followed by purification with RNeasy Mini Kit (Invitrogen, Carlsbad, CA, USA) and Qiagen, Valencia, CA, USA) were used for extracting total RNA according to the manufacturer’s instructions. Nanodrop (Nanodrop Technologies, Wilmington, DE, USA) was used for measuring the RNA concentration and the A260/A280 ratio. RNA was reverse-transcribed into cDNA using High-capacity cDNA Reverse Transcription Kit (Applied Biosystems, Foster City, CA, USA) according to the instructions provided by the manufacturer. QPCR with TaqMan chemistry (Applied Biosystems) by using ABI Prism 7500 analyzer (Applied Biosystems) was used for the confirmation of microarray gene expression results. The analysis for the relative quantity of a specific gene, before and after the intervention in AT, was performed as described previously.\(^4\) Expression of target genes were normalized to cyclophilin A1 (PPIA) expression.

The clinical data were analyzed using the SPSS software for Windows version 14.0 (SPSS Inc., Chicago, IL, USA). Data are given as mean ± s.d., unless otherwise indicated. The normality of distributions of the variables was tested with the Kolmogorov–Smirnov test, with Lilliefors significance correction. Logarithmic transformation was used to achieve normal distribution, whenever needed (indicated in tables and/or figures). General linear model for univariate analysis was used to test the difference in fold-change values of mRNA expression between the groups. Paired samples t-test was used for comparing the baseline and endpoint measurements within the study group. Correlation analyses were carried out using Pearson’s method. Partial correlation analysis with adjustment for weight at baseline and gender was used when appropriate. The WR and control groups were combined in the correlation analysis at baseline and when studying the correlations of change values. At baseline, this was carried out because the participants who were randomized either to an intervention or a control group represents the same risk population, all having MetS. Thus, the participants were homogenous regarding the selection criteria. After the intervention, the two treatment groups were analyzed separately because of potential treatment effect. Moreover, the correlations between the change of MFAP5 mRNA expression and the changes of clinical and biochemical measures were studied when the WR and control groups were combined, but also separately, for both the groups. For the clinical and biochemical measurements, P<0.05 was considered as statistically significant.

Results

Values for body weight, fasting plasma glucose and serum insulin concentration were at baseline 92.1 ± 14.9 kg, 6.2 ± 0.4 mmol l\(^{-1}\) and 11.5 ± 5.4 pmol l\(^{-1}\), and at the end of the intervention 88.7 ± 14.0 kg, 5.9 ± 0.3 mmol l\(^{-1}\) and 10.7 ±
7.0 pmol l$^{-1}$ for the WR group, respectively. The respective values for the control group were 87.3 ± 8.3 kg, 6.4 ± 0.6 mmol l$^{-1}$ and 9.9 ± 5.0 pmol l$^{-1}$ at baseline and 87.7 ± 9.0 kg, 6.1 ± 0.6 mmol l$^{-1}$ and 10.6 ± 6.0 pmol l$^{-1}$ at the end of the intervention, respectively.

The QPCR analysis for MFAP5 expression confirmed the data obtained by microarray (Figure 1). When the comparisons were made within the groups, the mRNA expression levels of MFAP5 showed significant decrease (94.2 ± 49.0 to 81.7 ± 41.7 AU, $P = 0.017$) in the WR group and no change was seen in the control group (106.4 ± 40.4 to 109.4 ± 34.5 AU, $P = 0.636$). Similar results were also obtained by microarray showing that MFAP5 expression was significantly downregulated in the WR group ($P = 0.004$).3 Fold-change results by QPCR showed a significant reduction in MFAP5 mRNA expression in the WR group ($P = 0.028$) when compared with the control group.

Correlation analyses were performed with the combined study groups at baseline. MFAP5 mRNA expression correlated significantly at baseline with BMI (Table 1). Moreover, the expression of MFAP5 correlated significantly with fasting serum insulin concentration at baseline, when adjusted for baseline body weight (Table 1). There was no correlation between MFAP5 mRNA expression and fasting plasma glucose levels.

Interestingly, at baseline, the mRNA expression of MFAP5 correlated negatively with fasting serum adiponectin and positively with leptin concentrations. Moreover, a positive correlation was also found for fasting plasma interleukin (IL)-1Ra and a negative one for fasting plasma IL-1β concentrations, when adjusted for baseline body weight (Table 1). MFAP5 mRNA expression correlated positively with leptin gene expression (Table 1) at baseline (adjusted for baseline body weight) and with adiponectin and IL-6 expression after the WR (Table 1). However, at the expression level, there was no correlation with cytokines like transforming growth factor beta 1, tumour necrosis factor alpha, IL-1β or IL-1Ra (Table 1).

Taken into account that MFAP5 is involved in the NOTCH pathway,9 we studied the correlation of MFAP5 mRNA expression with NOTCH1 and NOTCH2 mRNA expressions. MFAP5 mRNA expression did not correlate with the expressions of these genes.

We also studied whether MFAP5 mRNA expression is correlated with expressions of genes, which are involved in modulation of AT formation. MFAP5 expression correlated positively with peroxisome proliferator-activated receptor gamma (PPARγ), cyclin D2 (CCND2) and a disintegrin and metallopeptinase domain 12 (ADAM12) mRNA expressions at baseline (adjusted for baseline body weight), as well as after the WR (Table 1), when adjusted for the corresponding body weight.

The change in MFAP5 mRNA expression level correlated significantly with the change in body fat mass ($r = 0.392$, $P = 0.009$). Furthermore, the change of MFAP5 expression level correlated significantly with the changes of ADAM12, adiponectin (ADIPOQ) and NOTCH2 expression levels ($r = 0.343$, $P = 0.026$; $r = 0.433$, $P = 0.004$; $r = 0.438$, $P = 0.006$), respectively, when adjusted for the change of body weight and the WR and control groups were combined. When the groups were analyzed separately, there was also a significant correlation in the WR group between the change of MFAP5 mRNA expression and the change of ADIPOQ mRNA expression. In the control group, there were no significant correlations.

**Discussion**

MFAP5 is associated with microfibrils in elastin networks in a number of tissues, and its function may be related to cell signaling during microfibril assembly, elastogenesis and cell survival.5,10,11 It has been shown that MFAP5 promotes angiogenesis and interacts with NOTCH1 by either activating or suppressing its activity, depending on the cell type involved.9,12 Furthermore, MFAP5 is increased in fibrotic skin of humans and in mouse models with systemic sclerosis.13 Whether its function in AT is similar to previous findings is not known.

ECM is composed of structural and multifunctional molecules such as collagen, adhesive glycoproteins and proteoglycans.14,15 In AT, ECM maintains the structural integrity of adipocytes, and has an important role in AT formation.15 Development of obesity induces changes in AT (e.g., adipocyte hypertrophy, new adipocyte formation), which are associated with remodeling of ECM proteins and angiogenesis.14,15,16 Genes related to ECM and cytoskeleton have been shown to be upregulated by high-fat diet and
Correlations of MFAP5 mRNA expression in subcutaneous adipose tissue with anthropometric, biochemical measures, and with selected genes expressed in subcutaneous adipose tissue before and after the intervention in weight reduction and control groups; adjusted for body weight

|                         | Groups combined (n = 46) | Weight reduction group (n = 28) | Control group (n = 18) |
|-------------------------|-------------------------|-------------------------------|-----------------------|
|                         | 0 week                  | 34 weeks                     | 34 weeks              |
| r*                     | P-value                 | r*                           | P-value               |
| **Anthropometric and biochemical measures** |                         |                               |                       |
| Body mass index (kg·m⁻²) | 0.369***               | 0.014                        | 0.068**               | 0.737                 | 0.128                 | 0.613 |
| Body weight (kg)        | 0.043***               | 0.785                        | -0.213*               | 0.297                 | -0.122*               | 0.641 |
| Body fat mass (kg)      | 0.278***               | 0.069                        | 0.052**               | 0.798                 | 0.114                 | 0.653 |
| Waist circumference (cm) | 0.282***               | 0.071                        | 0.431*                | 0.032                 | -0.011*               | 0.968 |
| S (μU·min⁻¹)           | -0.260***              | 0.101                        | -0.162*               | 0.439                 | -0.429*               | 0.017 |
| f5-adiponectin (μg·ml⁻¹) | -0.378***              | 0.014                        | -0.369*               | 0.063                 | -0.626*               | 0.009 |
| f5-leptin (ng·ml⁻¹)     | 0.361***               | 0.019                        | 0.155*                | 0.459                 | 0.153*                | 0.271 |
| f5-insulin (pmol·l⁻¹)   | 0.397**                | 0.010                        | 0.165*                | 0.429                 | 0.352*                | 0.181 |
| f5-glucose (mmol·l⁻¹)   | 0.132**                | 0.41                         | -0.093*               | 0.66                  | 0.358*                | 0.174 |
| f5-TNFα (pg·ml⁻¹)       | -0.162***              | 0.305                        | -0.296*               | 0.151                 | 0.028*                | 0.919 |
| f5-IL1β (pg·ml⁻¹)       | -0.403***              | 0.009                        | -0.22                | 0.29                  | -0.207*               | 0.441 |
| f5-IL1Ra (pg·ml⁻¹)      | 0.347***               | 0.024                        | 0.325*                | 0.113                 | 0.469*                | 0.067 |
| hsCRP (mg·l⁻¹)          | -0.021***              | 0.895                        | -0.106*               | 0.615                 | -0.076*               | 0.781 |

**Gene expression in adipose tissue (AU)**

|                     | 0 week                  | 34 weeks                     | 34 weeks              |
|---------------------|-------------------------|-------------------------------|-----------------------|
| r*                  | P-value                 | r*                           | P-value               |
| Leptin              | 0.338***               | 0.028                        | 0.259*                | 0.211                 | 0.357*                | 0.174 |
| Adiponectin         | 0.033***               | 0.838                        | 0.488*                | 0.013                 | 0.011*                | 0.968 |
| PPARγ              | 0.521***               | 0.001                        | 0.621*                | 0.001                 | 0.195*                | 0.469 |
| TNFα                | 0.133***               | 0.404                        | 0.018*                | 0.932                 | 0.39*                 | 0.136 |
| IL-1β               | 0.071**                | 0.659                        | 0.140*                | 0.524                 | 0.09*                 | 0.741 |
| IL-1Ra              | 0.106**                | 0.504                        | 0.054*                | 0.807                 | 0.386*                | 0.14  |
| IL-6                | 0.007***               | 0.966                        | 0.409*                | 0.042                 | -0.267*               | 0.317 |
| TGF-β1              | 0.25*                  | 0.12                         | 0.388*                | 0.067                 | 0.089*                | 0.743 |
| NOTCH1              | -0.277*                | 0.083                        | -0.191*               | 0.383                 | -0.121*               | 0.656 |
| NOTCH2              | 0.191*                 | 0.239                        | 0.324*                | 0.131                 | 0.37*                 | 0.158 |
| CCND2               | 0.523***               | -0.001                       | 0.401*                | 0.047                 | 0.392*                | 0.133 |
| ADAM22              | 0.248***               | 0.114                        | 0.261*                | 0.229                 | 0.473*                | 0.064 |
| ADAM12              | 0.584***               | <0.001                       | 0.552*                | 0.004                 | 0.579*                | 0.019 |

Abbreviations: ADAM12, A disintegrin and metalloproteinasine domain 12; ADAM22, A disintegrin and metalloproteinasine domain 22; AU, arbitrary unit; CCND2, cyclin D2; fP, fasting plasma; f5, fasting serum; hsCRP, high-sensitivity C reactive protein; IL-6, interleukin 6; IL-1Ra, interleukin 1 receptor antagonist; IL-1β, interleukin 1 beta; NOTCH1, notch1 proprotein; NOTCH2, notch2 proprotein; PPARγ, peroxisome proliferator-activated receptor gamma; S, insulin sensitivity index; TGF-β1, transforming growth factor beta 1; TNFα, tumour necrosis factor alpha. *Adjusted for body weight (kg). †Correlation analyses were carried out using Pearson’s method, no adjustments; n = 41; **n = 42; ***n = 43; ****n = 44; *****n = 44; ***n = 24; ‡n = 26; §n = 27; ¶n = 17. Values are logarithmized, when appropriate. Bold font: result statistically significant (P < 0.05).
We thank Mrs Paivi Turunen for the technical assistance.

Acknowledgements

The authors declare no conflict of interest.

Conflict of interest

No conflicts of interest to declare.

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