ABSTRACT Changes in protein subcellular localization and abundance are central to biological regulation in eukaryotic cells. Quantitative measures of protein dynamics in vivo are therefore highly useful for elucidating specific regulatory pathways. Using a combinatorial approach of yeast synthetic genetic array technology, high-content screening, and machine learning classifiers, we developed an automated platform to characterize protein localization and abundance patterns from images of log phase cells from the open-reading frame—green fluorescent protein collection in the budding yeast, Saccharomyces cerevisiae. For each protein, we produced quantitative profiles of localization scores for 16 subcellular compartments at single-cell resolution to trace proteome-wide relocalization in conditions over time. We generated a collection of ~300,000 micrographs, comprising more than 20 million cells and ~9 billion quantitative measurements. The images depict the localization and abundance dynamics of more than 4000 proteins under two chemical treatments and in a selected mutant background. Here, we describe CYCLoPs (Collection of Yeast Cells Localization Patterns), a web database resource that provides a central platform for housing and analyzing our yeast proteome dynamics datasets at the single cell level. CYCLoPs version 1.0 is available at http://cyclops.ccbr.utoronto.ca. CYCLoPs will provide a valuable resource for the yeast and eukaryotic cell biology communities and will be updated as new experiments become available.

KEYWORDS
GFP subcellular localization abundance images microscopy

During the past decade, proteome-wide screens in a variety of experimental systems have begun to elucidate how protein networks are organized in eukaryotic cells (e.g., Ghaemmaghami et al. 2003; Huh et al. 2003; Newman et al. 2006; Nagaraj et al. 2012; Kulak et al. 2014). We have developed a method for integrating systematic genetics, high-throughput microscopy, image analysis and pattern classification into an automated data acquisition and analysis platform for cell biological screens in budding yeast (Chong et al. 2015). Our screening pipeline makes use of the yeast GFP (green fluorescent protein) collection, which consists of a series of haploid yeast strains in which each open-reading frame (ORF) is individually tagged, generating a full-length protein with a COOH-terminus GFP fusion, whose expression is driven by the endogenous ORF promoter (Huh et al. 2003). We worked with the set of 4144 strains from the original collection previously annotated as having a visible GFP signal and representing ~71% of the yeast proteome. We used this collection to measure the subcellular localization and abundance of yeast proteins at the single-cell level in several conditions in time courses of up to 11 hr (Chong et al. 2015).

A number of existing databases present images of yeast cells from large-scale studies. Some of these studies assess phenotypes associated with analysis of a small number of morphologic characteristics or...
markers in a collection of mutants. Databases that house this type of data include SCMD (Saito et al. 2004) and PhenOM (Jin et al. 2012). Other databases present images of a collection of GFP (or otherwise)-tagged proteins in one or a few genetic backgrounds or conditions. Examples of this type include the Yeast GFP Fusion Localization Database, YGF (Huh et al. 2003), the Yeast Protein Localization Database, YPL (Kals et al. 2005), Organelle DB (Wiwatwattana et al. 2007), the Yeast Resource Center, YRC (Riffle and Davis 2010), the Localization and Quantitation Atlas of the Yeast Proteome, LOQATE (Breker et al. 2013), and Cellbase (Dénervaud et al. 2013). Several of these databases present visually annotated protein localizations together with the images (YGF, YPL, LOQATE), two quantify protein abundance (LOQATE, Cellbase), and one assesses the probability of each cell displaying any mixture of six main spatial patterns (Cellbase); however, none of them computationally defines a localization for each GFP protein.

To enable easy access of our image compendium of subcellular localization and abundance profiles to the research community, we developed a web-accessible database called CYCLoPs (Collection of Yeast Cells and Localization Patterns) that allows retrieval and visualization of yeast cell images and permits queries of the subcellular localization and abundance profiles of the yeast proteome for each genetic or chemical perturbation in our survey. CYCLoPs currently contains a total of 330,248 images from three wild-type screens, three screens with a strain deleted for the gene encoding the conserved lysine deacetylase Rpd3, and time courses of two chemical treatments (hydroxyurea and rapamycin; Table 1). CYCLoPs differs from existing databases in a number of ways: (1) whereas other databases provide searchable localization assignments for proteins that had been assessed visually, CYCLoPs contains computationally derived quantitative localization and abundance profiles; (2) CYCLoPs provides a searchable web graphical interface for proteins with localization and/or abundance changes of interest, which reflects the proteome flux in response to varying environmental cues and genetic backgrounds; (3) the subcellular localization data hosted on CYCLoPs were determined directly from the morphologic features of the cells and accommodate the reality that many proteins localize to multiple locations; and (4) CYCLoPs provides localization and abundance profiles for individual cells screened, thus enabling analysis at the single-cell level.
the database. For each protein, the four GFP and four red fluorescent protein (RFP) micrographs, along with the corresponding overlay GFP-RFP images, are available for visualization and download through CYCLOPs as lower resolution JPEG files.

**Quantitative scoring of protein abundance**
Protein abundance was extrapolated from the mean GFP intensity. For each cell, we calculated the ratio of integrated GFP intensity measured within the area defined by the segmented cell boundary, divided by the segmented area of the cell. The mean GFP intensity ($I_g$) of the protein was taken as the arithmetic mean of these ratios. The $I_g$ measurements from our wild-type screen were highly correlated with protein abundance measurements from other techniques, namely flow cytometry, western blot analysis, and mass spectrometry (Chong et al. 2015). For each strain, protein abundance changes ($\Delta PL$) were calculated as the fold-change, i.e., $I_g$ in the presence of treatment/mutant over $I_g$ in wild type. The $I_g$ and $\Delta PL$ values for every protein in all conditions are readily searchable in CYCLOPs.

**Constructing the ensemble classifiers for quantification of subcellular localizations at single-cell level**
Previous studies have shown that combining decisions from multiple computational classifier instances—a so-called “ensemble” strategy—can improve the predictive accuracy of the classification (Gashler et al. 2008; Rokach 2010). The ensemble approach is particularly useful in boosting the performance of weak learners and has been used in recent genomic studies (Chen et al. 2011; Reboiro-Jato et al. 2013). We constructed an ensemble of classifiers—ensLOC—to accurately assign each yeast cell to one or more of 16 pre-defined subcellular localization classes based on its morphological features. Our approach allows proteins to localize to more than one compartment or to remain unclassified, based on our 16 predefined morphologic classes. For every segmented cell in the compendium, ensLOC generated a 16-element vector, where each element is an independent assessment of the cell’s membership in a localization class.

The ensLOC framework comprises several steps (Figure 1). We first segmented the cells from micrographs obtained from our wild-type screen. A total of 430 image features, including area, shape, intensity, texture, and Zernike moments (projections of image functions based on a set of orthogonal Zernike polynomials; Teague 1979) were extracted from the segmented cells. For each classifier, we used the $\chi^2$ test of independence (Liu and Setiono 1995) to select features that best discriminated the positive from the negative training instances. The filtered features were then used as input to construct the linear Support Vector Machine classifier (Platt 1998). Seventy thousand instances of cell images representative of the morphological signatures can be found in the database.

![Diagram illustrating the ensemble of 60 binary classifiers for protein localization and quantification (modified from Chong et al. 2015). Only cell images that were not filtered by the quality-control classifiers for dead cells and “ghost” objects were further classified. All cells were first classified into different cell-cycle stages using the unbudded and budded classifiers. The rest of the ensemble is organized into 20 subgroups. For some classifier groups, e.g., Cortical Patches and Cell Periphery, budded and unbudded cells were separately tested. The results from each subgroup of binary classifiers e.g., CoP, CoP-Mito, and CoP-Cyto, were consolidated with Bagging. The circles denote the percentage of 1,057,871 cells in the wild-type WT1 experiment that were assigned to each localization class, with darker green indicating a greater percentage.](image-url)
| Classifier ID | Name of Binary Classifier | No. of Positive Training Objects | No. of Negative Training Objects | Validation Using 10-fold Cross-Validation | Visual Inspection |
|---------------|--------------------------|---------------------------------|---------------------------------|------------------------------------------|------------------|
|               |                          |                                 |                                 | Recall | Precision |                      |                  |
| Quality control |                          |                                 |                                 |        |           |                      |                  |
| 1.1.1 | DEAD | 960 | 1541 | 0.986 | 0.995 |                      |                  |
| 1.1.2 | GHOST | 1840 | 2398 | 0.995 | 1 |                      |                  |
| Budded or Unbudded |                          |                                 |                                 |        |           |                      |                  |
| 2.1.1 | UNBUDDED | 1095 | 1582 | 0.997 | 0.984 |                      |                  |
| 2.1.2 | SMALLBUDDED | 434 | 733 | 0.952 | 0.948 |                      |                  |
| 2.1.3 | LARGE_MEDIUMBUDDED | 727 | 1508 | 0.985 | 0.986 |                      |                  |
| 3.1 Cytoplasm |                          |                                 |                                 |        |           |                      |                  |
| 3.1.1 | CYTOPLASM | 3493 | 4285 | 0.979 | 0.966 | ~95% |                  |
| 3.1.2 | CYTOPLASM_NOT_NUCLEAR | 2075 | 1419 | 0.915 | 0.842 |                      |                  |
| 3.2 Endosome |                          |                                 |                                 |        |           |                      |                  |
| 3.2.1 | ENDOsomE | 2245 | 4730 | 0.826 | 0.912 | <70% |                  |
| 3.2.2 | ENDOsomE_CYTOPLASM | 2245 | 3493 | 0.977 | 0.995 |                      |                  |
| 3.2.3 | ENDOsomE_NUCLEI | 2245 | 5612 | 0.995 | 0.999 |                      |                  |
| 3.2.4 | ENDOsomE_SPINDLEPOLE | 2245 | 3397 | 0.963 | 0.986 |                      |                  |
| 3.2.5 | ENDOsomE_MITOCHONDRIA | 2245 | 6315 | 0.899 | 0.967 |                      |                  |
| 3.3 ER |                          |                                 |                                 |        |           |                      |                  |
| 3.3.1 | ER | 5274 | 4259 | 0.977 | 0.919 | <80% |                  |
| 3.3.2 | ER_CYTOPLASM | 5274 | 3493 | 0.97 | 0.965 |                      |                  |
| 3.3.3 | ER_VACUOLES_VACUOLARMEMBRANE | 5274 | 3893 | 0.976 | 0.958 |                      |                  |
| 3.3.4 | ER_CELLPERIPHERY | 5274 | 4059 | 0.996 | 0.99 |                      |                  |
| 3.4 Golgi |                          |                                 |                                 |        |           |                      |                  |
| 3.4.1 | GOLGI | 1994 | 1838 | 0.964 | 0.908 | >80% |                  |
| 3.4.2 | GOLGI_MITOCHONDRIA | 1994 | 6315 | 0.809 | 0.968 |                      |                  |
| 3.4.3 | GOLGI_ENDOSOME | 1994 | 2245 | 0.919 | 0.934 |                      |                  |
| 3.4.4 | GOLGI_CYTOPLASM | 1994 | 3493 | 0.996 | 0.99 |                      |                  |
| 3.5 Mitochondria |                          |                                 |                                 |        |           |                      |                  |
| 3.5.1 | MITOCHONDRIA | 6315 | 7894 | 0.894 | 0.884 | >85% |                  |
| 3.6 Nuclear Periphery |                          |                                 |                                 |        |           |                      |                  |
| 3.6.1 | NUCLEARPERIPHERY | 2668 | 4367 | 0.94 | 0.96 | ~70% |                  |
| 3.7 Nucleus |                          |                                 |                                 |        |           |                      |                  |
| 3.7.1 | NUCLEI | 5612 | 6881 | 0.977 | 0.956 | >80% |                  |
| 3.7.2 | NUCLEINOTCYTOPLASM | 1398 | 989 | 0.99 | 0.93 | >80% |                  |
| 3.8 Nucleolus |                          |                                 |                                 |        |           |                      |                  |
| 3.8.1 | NUCLEOLUS | 3882 | 5332 | 0.926 | 0.948 | >85% |                  |
| 3.9 Peroxisome |                          |                                 |                                 |        |           |                      |                  |
| 3.9.1 | PEROXISOME | 1256 | 2099 | 0.849 | 0.922 | <70% |                  |
| 3.9.2 | PEROXISOME_GOLGI | 1256 | 1993 | 0.928 | 0.971 |                      |                  |
| 3.9.3 | PEROXISOME_SPINDLEPOLE | 1256 | 3397 | 0.965 | 0.995 |                      |                  |
| 3.9.4 | PEROXISOME_MITOCHONDRIA | 1256 | 6315 | 0.814 | 0.981 |                      |                  |
| 3.10 Vacuole/Vacuolar Membrane |                          |                                 |                                 |        |           |                      |                  |
| 3.10.1 | VACUOLE_VACUOLARMEMBRANE_COMBINED | 3893 | 3352 | 0.926 | 0.898 | >80% |                  |
| 3.10.2 | VACUOLE_VACUOLARMEMBRANE | 2224 | 1846 | 0.92 | 0.845 | >80% VAC, 65% VAC membrane |                  |
| 3.11 Cortical Patches |                          |                                 |                                 |        |           |                      |                  |
| 3.11.1 | CORTICALPATCHES_UNBUNNED | 1813 | 1279 | 0.964 | 0.877 | ~70% |                  |
| 3.11.2 | CORTICALPATCHES_UNBUNNED_CYTOPLASM | 1813 | 1661 | 0.994 | 0.996 |                      |                  |
| 3.11.3 | CORTICALPATCHES_UnBUNNED_MITOCHONDRIA | 1813 | 4440 | 0.95 | 0.984 |                      |                  |
of 16 subcellular localizations were handpicked. The distribution of these training instances is shown in Figure 2 and Table 2.

Predictions for each subcellular localization class were obtained through combining the predicted results of a set of binary classifiers (Figure 2). A binary classifier classifies elements of a given test set into only two groups. For example, a cell is assigned to the endoplasm reticulum (ER) class if it is assigned to ER in the ER-all, ER-Cytoplasm and ER-Vacuole/Vacuolar Membrane, and ER-Cell Periphery binary classifiers (Table 2). Our objective with this approach was to reduce misclassifications among subcellular localization classes with overlapping morphological patterns. In addition, an improvement of predictive accuracy was achieved through bootstrap aggregation (bagging) (Breiman 1996), specifically by plurality voting. We generated 25 bag classifiers, each from 1000 random training instances (500 positive and 500 negative samples) with replacements. Decisions for the localization assignments of a binary classification step were reconciled from

| Classifier ID | Name of Binary Classifier | No. of Positive Training Objects | No. of Negative Training Objects-Validation Using 10-fold Cross-Validation | Visual Inspection Recall |
|---------------|---------------------------|---------------------------------|------------------------------------------------------------------------|-------------------------|
| 3.11.4        | CORTICALPATCHESBUDDED     | 1345                            | 2171                      | 0.928  0.936  75%            |
| 3.11.5        | CORTICALPATCHESBUDDED_     | 1345                            | 1059                      | 0.994  0.988                |
|               | CELLPERIPHERY              |                                 |                           |                         |
| 3.11.6        | CORTICALPATCHESBUDDED_     | 1345                            | 1875                      | 0.981  0.986                |
|               | MITOCHONDRIA               |                                 |                           |                         |
| 3.11.7        | CORTICALPATCHESBUDDED_     | 1345                            | 1022                      | 0.987  0.988                |
|               | CYTOPLASM                  |                                 |                           |                         |
| 3.12 Bud      | BUD                        | 1619                            | 1691                      | 0.937  0.905  >70%          |
| 3.13 Budneck  | BUDNECK                    | 2170                            | 3095                      | 0.947  0.942  >70%          |
|               | BUDNECK_BUD                | 2170                            | 1619                      | 0.962  0.946                |
|               | BUDNECK_CELLPERIPHERY      | 2170                            | 1059                      | 1       0.994                |
|               | BUDNECK_MITOCHONDRIA       | 2170                            | 1875                      | 0.99    0.98                 |
| 3.14 Budsite  | BUDSITE                    | 453                             | 637                       | 0.982  0.961  >80%          |
|               | BUDSITE_CYTOPLASM          | 453                             | 4955                      | 0.943  0.992                |
|               | BUDSITE_CELLPERIPHERY      | 453                             | 359                       | 0.996  0.992                |
| 3.15 Cell     | CELLPERIPHERYUNBUDDED      | 2269                            | 858                       | 0.989  0.98     >95%        |
|               | CELLPERIPHERYBUDDED        | 1059                            | 1688                      | 0.981  0.991                |
| 3.16 Spindle  | Pole                       |                                  |                           |                         |
| 3.16.1        | SPINDLEPOLETWODOTFARBUDDED | 416                             | 966                       | 0.938  0.965  >70%          |
| 3.16.2        | SPINDLEPOLETWODOTFARBUDDED_ | 416                             | 2170                      | 0.913  0.997                |
|               | BUDNECK                    |                                 |                           |                         |
| 3.16.3        | SPINDLEPOLETWODOTFARBUDDED_ | 416                             | 492                       | 1       0.996                |
|               | NUclearPERIPHERY           |                                 |                           |                         |
| 3.16.4        | SPINDLEPOLETWODOTFARBUDDED_ | 416                             | 1109                      | 0.99    0.995                |
|               | NUcleolus                  |                                 |                           |                         |
| 3.16.5        | SPINDLEPOLETWODOTCLOSEBUDDED | 306                            | 1016                      | 0.905  0.97     ~80%        |
| 3.16.6        | SPINDLEPOLETWODOTCLOSEBUDDED | 306                            | 2170                      | 0.899  0.995                |
|               | BUDNECK                    |                                 |                           |                         |
| 3.16.7        | SPINDLEPOLETWODOTCLOSEBUDDED | 306                            | 1875                      | 0.974  0.996                |
|               | MITOCHONDRIA               |                                 |                           |                         |
| 3.16.8        | SPINDLEPOLETWODOTCLOSEBUDDED | 306                            | 492                       | 0.993  0.996                |
|               | NUclearPERIPHERY           |                                 |                           |                         |
| 3.16.9        | SPINDLEPOLETWODOTCLOSEBUDDED | 306                            | 1109                      | 0.98    0.988                |
|               | NUcleolus                  |                                 |                           |                         |
| 3.16.10       | SPINDLEPOLEONEDOT          | 2675                            | 3676                      | 0.974  0.983  70%          |

In total, approximately 70K handpicked cell images (objects) were used to train the classifiers. "No. of positive training objects" refers to cells which belong to the targeted class and "No. of negative training objects" refer to cells not belonging to the targeted class. For example, to construct the "DEAD" cells classifier, 960 images of dead cells were used as positive training objects and 1541 images of non-dead cells from across all 16 localization classes were used as negative training objects. The first number of the classifier ID reflects the level and therefore the sequence at which the classifier was applied. For instance, all cell images were first tested using the "DEAD" cells classifier to eliminate dead cells from further classification to the 16 localization classes, and only cells that were tested positive in the level 2 "SMALLBUDDED" and "LARGEMEDIUMBUDDED" classifiers would be further classified by the "BUDNECK" classifier. The accuracy of the classifiers was validated computationally using 10-fold cross-validation and manually using visual inspection of 500 random positive cells. Recall = True positives/(True positives + False negatives); Precision = True positives/(True positives + False positives). ER, endoplasm reticulum.
these 25 bag classifiers. Thus, in total the localization assignment for a segmented cell was determined through plurality voting of more than 1000 classifiers (25 bags × 60 binary classifiers). Because the computation was time intensive, we modified the algorithm to enable parallelized execution on a computer cluster. Both 10-fold cross-validation and visual inspection of random samples were independently conducted for each localization class to validate the accuracy of the classifier (Table 2).

We benchmarked our computationally derived localization assignments for one of our wild-type screens, WT1 (Chong et al. 2015), to visually assigned localization annotations from YGFP (Huh et al. 2003) and found 94% agreement among the set of 1097 proteins assigned to a single location by both methods (Chong et al. 2015). We also compared our computationally derived localization assignments with assignments made using other computational methods with images from YGFP (Chen et al. 2007; Huh et al. 2009). The ensLOC framework achieved greater mean accuracy (overlap proteins divided by number of proteins identified in YGFP) for proteins identified as having a single localization in both data sets. The ensLOC framework attained a mean classifier accuracy of 81%, an improvement of up to 20% across 12 subcellular localization classes defined in both methods (Figure 3). It is also worth noting that, unlike other methods, the ensLOC framework for quantifying subcellular localization does not restrict a protein to a single localization class.

**Quantitative scoring of subcellular localization using the ensLOC framework**

In our automated imaging pipeline (Chong et al. 2015), an experiment generally produced more than a million segmented cells, among which approximately 5% were of inadequate quality. These poor quality cell images were removed using a quality control step with classifiers designed to identify dead and “ghost” cells. “Ghost” objects are artifacts that get recognized in the background of an image, as a consequence of noise being recognized as signal and result in segmentation of a region containing no cells. The ensLOC framework was then applied independently to each filtered cell; that is, protein localization in each cell was predicted for up to 60 binary classifiers, where each classifier determined if a cell should be assigned positively or negatively to the class based on its morphological features. For example, the ER-Cytoplasm binary classifier determined whether a cell harbored the phenotypic signatures of ER localization class (positive) or Cytoplasm localization class (negative). To determine the subcellular localization assignment profile of a GFP-tagged protein at the single-cell level, we calculated the proportion of labeled cells that were assigned to each of the 16 subcellular localization classes. The localization profile of a protein is thus represented as a 16-element vector, where each element (“LOC-score”) reflects the proportion of “classifiable” cells (that is, assigned to at least one localization class) that are assigned to a specific localization class.

To identify changes in subcellular localization for each protein following genetic or environmental perturbation, we assessed the statistical significance of the difference between the proportion of cells with a given localization in a condition (genetic/chemical perturbation) and the proportion of cells in wild type by using a metric we designated a z-LOC score. Cutoffs for significant localization changes were determined by fitting a “background” normal Gaussian model and a uniform “outlier” density model to the z-LOC score distribution. Cutoffs were chosen such that the number of true “outliers” was optimized (Chong et al. 2015). The LOC-scores and z-LOC scores are readily searchable in CYCLoPs.

**Database system construction**

The relational database schema of CYCLoPs was developed to provide central storage and querying of different types of data generated from our systematic yeast imaging experiments. Our goal was to optimize the efficient and scalable querying of the micrographs, the LOC-score and z-LOC score profiles, and the abundance I_a and I_PL score profiles of all proteins and conditions surveyed (database schema available through the CYCLoPs online documentation). The backend of CYCLoPs features a mySQL relational database management system, which comprises more than 100 experiment-specific tables, and the front-end web interface is hosted on an Apache 2.0 web server. The web interface was developed using a combination of HTML, CGI Perl, Perl DataBase Interface, Cascading Style Sheets, Javascript and R plotting libraries.

**Database utility**

**System interfaces and visualization:** CYCLoPs is primarily accessible via a Web interface, with a focus on providing easy and efficient access to a genome-wide database of quantitative descriptors of protein dynamics, and to assist biologists in experiment planning and hypothesis generation. A number of query and visualization tools are included in CYCLoPs version 1.0, including two custom-made micrograph viewers.
Figure 4  Screen shot of sample search and result page generated by CYCLoPs is shown. A query of the Hxt2 protein produces images of yeast cells from three wild-type screens (WT1, WT2, WT3). The tables list numerical measurements of protein abundance ($I_g$), protein abundance changes ($dPL$), subcellular localization (LOC-scores), and subcellular localization changes (zLOC-scores) with localizations from the WT screens shown in red. The scores are highlighted using a color scale from white to yellow to red to allow the viewer to identify variances in a range of values with a quick glance and do not represent significance values.
Various search options are available. Search options for proteins include protein name, common name, alias, and ORF. A protein-centric search returns a general description, abundance scores, and localization scores for the best matching protein across all screens. For example, a quick search of "Hxt2" returns a report displaying: (1) the micrographs from three wild-type screens; (2) a subcellular localization table depicting the LOC-scores of Hxt2 for each of 16 localization classes (rows) across all screens (columns); (3) a protein abundance table showing the abundance Ig and changes δPL across the screens and; (4) a subcellular localization change table showing the localization changes, z-LOCs, for each localization class across all screens (Figure 4). Search results and images may be downloaded and direct visualization of each individual cell in the compendium is available through the companion Image Viewer and Cell Viewer.

**Image and cell viewer:** The Image Viewer facilitates visual inspection of pairs of micrographs. Users can toggle between 18 screens, four images per screen, and three image channels (RFP/GFP/GFP-RFP overlay). This tool is particularly useful for visual inspection of morphologic changes. For example, Figure 5 shows internalization of Hxt2, a glucose transporter, in response to rapamycin treatment. Cells in the left micrograph (from a WT screen) display morphologic patterns that define a cell-periphery localization of Hxt2, whereas most cells in the right micrograph (after 300 min of rapamycin treatment) exhibit an obvious localization to vacuole/vacuolar membrane. The Cell Viewer provides a detailed view of a specified micrograph by cropping it into individual cells (Figure 6). The position coordinates of each cell image were obtained from the cell segmentation routine in our image analysis program. The localization labels of the cell were determined using our ensemble classifiers.

**Single cell abundance measurements and localization assignments:** Both localization and abundance of a protein may vary in individual cells in a population (Chong *et al.* 2015), and analysis of single cells can give important information about cell-cycle events and stochasticity. Because all of our data were acquired at the level of the individual cell, we are able to provide a function that allows the user to download abundance and localization data for single cells. This function may be found at http://cyclops.ccbr.utoronto.ca/DOWNLOAD/Download.html. For a selected image in our compendium, this function generates a text file with the following columns:

- Object ID, X-coordinate, Y-coordinate, GFP Intensity (Ig), Localization.

**Querying top protein abundance and localization changes:** Users also may retrieve top-ranked proteins that are transported toward or away from any of the 16 subcellular compartments included in the database or that exhibit increase/decrease in protein abundance in the presence of drug treatment or gene deletion. Search results and images may be downloaded and direct visualization of each object/cell in the compendium is available through the companion Image Viewer and Cell Viewer.

We have generated a compendium of RFP/GFP micrographs and quantitative measurements of subcellular localization and abundance changes covering ~71% of the yeast proteome in response to genetic and chemical perturbations. To make this novel compendium available and useful to the research community, we have developed a web-based query system for accessing, visualizing and analyzing the data.

CYCLOPs is intended to be an active resource for quantitative genome-wide localization and abundance measurements of *S. cerevisiae* made in multiple genetic backgrounds and following different chemical treatments. Future enhancements of CYCLOPs will involve automation processes for experimental updates. In revised versions of CYCLOPs, we hope to integrate data from other external sources for on-the-fly cross-dimensional comparisons and visualization. CYCLOPs is tightly integrated with our experimental and scoring platform, and will house data from future experiments designed to test the response of the yeast proteome to a variety of chemical and environmental perturbations.

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LITERATURE CITED
Breker, M., M. Gymrek, and M. Schuldiner, 2013 A novel single-cell screening platform reveals proteome plasticity during yeast stress responses. J. Cell. Biol. 200: 839–850.
Breiman, L., 1996 Bagging predictors. Mach. Learn. 2: 123–140.
Carpenter, A. E., T. R. Jones, M. R. Lamprecht, C. Clarke, I. H. Kang et al., 2006 CellProfiler: image analysis software for identifying and quantifying cell phenotypes. Genome Biol. 7: R100.
Chen, M., L. Shi, R. Kelly, R. Perkins, H. Fang et al., 2011 Selecting a single model or combining multiple models for microarray-based classifier development--a comparative analysis based on large and diverse datasets generated from the MAQC-II project. BMC Bioinformatics 12(Suppl 10): S3.
Chen, S. C., T. Zhao, G. J. Gordon, and R. F. Murphy, 2007 Automated image analysis of protein localization in budding yeast. Bioinformatics 23: i66–i71.
Chong, Y. T., J. L. Y. Koh, H. Friesen, S. Kaluarachchi Duffy, M. J. Cox et al., 2015 Yeast proteome dynamics from single cell imaging and automated analysis. Cell (in press).
Dénervaud, N., J. Becker, R. Delgado-Gonzalo, P. Damay, A. S. Rajkumar et al., 2008 Decision tree ensemble: small heterogeneous is better than large homogeneous. The Seventh International Conference on Machine Learning and Applications, pp. 900–905.
Ghaemmaghami, S., W. K. Huh, K. Bower, R. W. Howson, A. Belle et al., 2003 Global analysis of protein expression in yeast. Nature 425: 737–741.
Huh, S., D. Lee, and R. F. Murphy, 2009 Efficient framework for automated classification of subcellular patterns in budding yeast. Cytometry A 75: 934–940.
Huh, W. K., J. V. Falvo, L. C. Gerke, A. S. Carroll, R. W. Howson et al., 2003 Global analysis of protein localization in budding yeast. Nature 425: 686–691.
Jin, K., J. Li, F. S. Vizeacoumar, Z. Li, R. Min et al., 2012 PhenoM: a database of morphological phenotypes caused by mutation of essential genes in Saccharomyces cerevisiae. Nucleic Acids Res. 40: D687–D694.
Kals, M., K. Natter, G. G. Thallinger, Z. Trajanoski, and S. D. Kohlwein, 2005 YPL.db2: the Yeast Protein Localization database, version 2.0. Yeast 22: 213–218.
Kulak, N. A., G. Pichler, I. Paron, N. Nagaraj, and M. Mann, 2014 Minimal, encapsulated proteomic-sample processing applied to copy-number estimation in eukaryotic cells. Nat. Methods 11: 319–324.
Liu, H., and R. Setiono, 1995 Chi2: Feature selection and discretization of numeric attributes. Proc IEEE 7th International Conference on Tools with Artificial Intelligence: 338–391.
Nagaraj, N., N. A. Kulak, J. Cox, N. Neuhauser, K. Mayr et al., 2012 System-wide perturbation analysis with nearly complete coverage of the yeast proteome by single-shot ultra HPLC runs on a bench top Orbitrap. Mol. Cell Proteomics 11: M111.013722.
Newman, J. R., S. Ghaemmaghami, J. Ihmels, D. K. Breslow, and M. Noble, 2006 Single-cell proteomic analysis of S. cerevisiae reveals the architecture of biological noise. Nature 441: 840–846.
Platt, J. 1998 Fast training of support vector machines using sequential minimal optimization, pp. 185–208 in Advances in Kernel Methods - Support Vector Learning, edited by B. Scholkopf, C. J. C. Burges, and A. J. Smola. MIT Press, Cambridge.
Rokach, L., 2010 Ensemble-based classifiers. Artif. Intell. Rev. 33: 1–39.
Saito, T. L., M. Ohtani, H. Sawai, F. Sano, A. Saka et al., 2004 SCMD: Saccharomyces cerevisiae Morphological Database. Nucleic Acids Res. 32: D319–D322.
Teague, M. R., 1979 Image analysis via the general theory of moments. J. Opt. Soc. Am. 70: 920–930.
Tong, A. H., M. Evangelista, A. B. Parsons, H. Xu, G. D. Bader et al., 2001 Systematic genetic analysis with ordered arrays of yeast deletion mutants. Science 294: 2364–2368.
Wiwatwattana, N., C. M. Landau, G. J. Cope, G. A. Harp, and A. Kumar, 2007 Organelle DB: an updated resource of eukaryotic protein localization and function. Nucleic Acids Res. 35: D810–D814.

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