Pre-clinical study of induced pluripotent stem cell-derived dopaminergic progenitor cells for Parkinson’s disease

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Induced pluripotent stem cell (iPSC)-derived dopaminergic (DA) neurons are an expected source for cell-based therapies for Parkinson’s disease (PD). The regulatory criteria for the clinical application of these therapies, however, have not been established. Here we show the results of our pre-clinical study, in which we evaluate the safety and efficacy of dopaminergic progenitors (DAPs) derived from a clinical-grade human iPSC line. We confirm the characteristics of DAPs by in vitro analyses. We also verify that the DAP population include no residual undifferentiated iPSCs or early neural stem cells and have no genetic aberration in cancer-related genes. Furthermore, in vivo studies using immunodeficient mice reveal no tumorigenicity or toxicity of the cells. When the DAPs are transplanted into the striatum of 6-OHDA-lesioned rats, the animals show behavioral improvement. Based on these results, we started a clinical trial to treat PD patients in 2018.
he discovery that the transplantation of fetal midbrain tissues can restore neurological symptoms of Parkinson’s disease (PD) patients has stimulated interest in cell therapies for the disease. Human pluripotent stem cells (PSCs), such as embryonic stem cells (ESCs) and induced PSCs (iPSCs), are sources for a stable supply of cells for the therapy. Culture protocols that induce authentic midbrain dopaminergic (DA) neurons from human PSCs have already been established by several research groups, and the DA neurons have been proven safe and effective in rodent6,7 and primate8 PD models. These results suggest that PSC-derived DA neurons can contribute to the treatment of PD. However, the use of PSCs for cell-based therapies is a new challenge that regulatory rules are not universally standardized. Indeed, each country has its own regulations and different criteria for clinical application. Therefore, to develop a global standard cell-based therapy for PD, it is important to share the results of pre-clinical studies and the outcomes of clinical trials9.

Currently, two clinical trials using human ESCs are ongoing in Australia (NCT02452723) and China (NCT03119636), and their pre-clinical studies are reported10,11. We recently started a clinical trial (JMA-IIA00384, UMIN000033564) in Japan to treat PD patients by using iPSC-derived DA progenitors (DAPs). Because in these therapies the grafted cells survive and function as DA neurons for a long time (possibly until the patient dies), they must undergo more intensive risk management compared to other clinical trials using mesenchymal stem cells12, in which the final product contained no or minimal undifferentiated iPSCs. Indeed, each country has its own regulatory agency, Pharmaceuticals and Medical Devices Agency (PMDA), about details of the pre-clinical study. In this report, we show that the DAPs have no tumorigenicty or toxicity and that they improve the abnormal behavior of 6-hydroxydopamine (6-OHDA)-lesioned rats. Thus, we confirm the safety and efficacy of iPSC-derived DAPs for clinical trials.

Results
Manufacturing iPSC-derived DAPs under good manufacturing practice (GMP) conditions. As a starting material, we used a human iPSC line (QHJI01s04) derived from the peripheral blood of a human leukocyte antigen (HLA)-homozygous healthy volunteer13,14 and established for clinical application at the Center for iPS Cell Research and Application, Kyoto University, Japan. Then, we established a clonal master cell bank, MCB003, for our project. We have preserved the iPSCs in hundreds of frozen vials as one lot. To treat a patient, we thaw one of the vials, differentiate the cells into DAPs, and transplant them without freezing. In this pre-clinical study, we used multiple vials to confirm reproducibility of the results.

We modified our culture protocol to induce DAPs from human iPSCs8,15 to meet the GMP-grade culture condition (Fig. 1a−d, Supplementary Table 1). Briefly, human iPSCs (MCB003) were seeded onto 6-well plates coated with laminin 511-E8 fragment and cultured for 12 days, and then CORIN (+) cells were isolated by fluorescence-activated cell sorting (FACS). The sorted cells were cultured in 96-well plates as cell aggregates until day 30 and then used for the transplantation. In this culture condition, all reagents including an anti-CORIN antibody were at GMP grade. A clinical-grade cell sorter with a disposable fluid tube was equipped to mitigate the risk of cross-contamination.

Characterization of iPSCs (MCB003) in vitro. In the process of inducing DAPs, quality control check points were placed at the stage of MCB003, pre- and post-FACS sorted cells, and the final product (Fig. 1e). The quality standards of MCB003 include the morphology of the colonies, the expression of pluripotent markers such as TRA-1–60, TRA-2–49, and SSEA-4, and sterility. We confirmed that almost all cells expressed pluripotent cell markers (Supplementary Table 2).

Characterization of the final product in vitro. We conducted the DAP induction process in 25 randomly chosen vials from MCB003 (one time for each vial). Before sorting on day 12, the percentages of TRA-1–60+ and CORIN+ cells were 0.0% (n = 24) and 31.4 ± 12.7% (n = 25), respectively. After sorting, the percentage of CORIN+ cells were 93.2 ± 2.1% (n = 25), confirming purity (Table 1, Supplementary Fig. 1a, b). The in-process testing for the manufacturing of DAPs is described in Supplementary Table 3.

The quality standards of the final product were determined to confirm that the cells were mainly composed of DAPs and included no undifferentiated cells. For this purpose, the expression of markers for DAPs (FOXa2 and TUJ1), iPSCs (OCT3/4 = POU5F1, LIN28, and TRA-2–49), and proliferating early neural progenitors (SOX1, PAX6, and KI67) were examined by flow cytometry (FCM) and quantitative reverse transcription PCR (RT-qPCR). Flow cytometry on day 26 revealed that the percentage of DAPs (FOXa2+TUJ1+) was 92.3 ± 4.0% (n = 25; Table 1 and Supplementary Fig. 1c, d). Immunostaining also showed that most cells expressed DAP markers such as LMX1A and FOXA2 (Fig. 1f), and a smaller population expressed mature DA markers such as NURR1 and TH (Fig. 1g).

Regarding undifferentiated cells, immunostaining showed that day-26 spheres did not contain iPSCs (OCT3/4 and NANOG; Fig. 1h) nor early neural progenitors (SOX1 and PAX6; Fig. 1i). Flow cytometry revealed that OCT3/4+TRA-2–49+ double-positive cells were not detected. The same flow cytometry analysis of DAP samples spiked with 0.1% iPSCs or SOX1+PAX6+ double-positive cells confirmed that the detection levels meet the test specification (Table 1, Supplementary Fig. 1e, f). In addition, RT-qPCR analysis showed that the expression levels of POU5F1 (OCT3/4) and LIN28 were 0.08 ± 0.15% and 0.14 ± 0.13% (n = 25 each) of those undifferentiated iPSCs, respectively (Table 1, Supplementary Fig. 1g, h).

To further confirm that there were no residual iPSCs, day-26 spheres were cultured in a condition suitable for iPSCs for 2 weeks. In this experiment, we mixed 0.01 to 1.0% of iPSCs with 2 × 10^9 dissociated cells from the day-26 spheres and found that even 0.001% was enough to form visible iPSC colonies. Importantly, day-26 spheres without mixture did not show any colony formation (Supplementary Fig. 1i). Taken together, these results indicated that the final product contained no or minimal (<0.001%) undifferentiated iPSCs.

Next, we examined if the DAPs could differentiate into functional DA neurons. To evaluate the production of dopamine, day-26 spheres were cultured until day 56 on L-ornithine/laminin/fibronectin-coated plates, and dopamine release in response to high KCl stimulation was measured by liquid chromatography with tandem mass spectrometry (LC-MS/MS) (n = 5, Fig. 1j). The amount of dopamine was 3.3 ± 2.72 pmol μg^{-1} DNA in the cultured media, 0.47 ± 0.45 pmol μg^{-1} DNA in low KCl, and 1.46 ± 0.65 pmol μg^{-1} DNA in high KCl. Upon electrophysiological analyses, we found 67% of examined neurons showed spontaneous action potentials (Fig. 1k, l) and 89% of neurons showed induced action potentials (n = 18, Fig. 1m). The resting membrane potential was −49 ± 15 mV.

Genomic, epigenetic, and single-cell analyses. The results above suggest that there is no tumorigenic component in the final
However, genomic and epigenetic changes in the cells may affect cell behavior after transplantation. To examine this possibility, genomic and epigenetic stability during the differentiation was examined, and the final products were subjected to tumorigenicity studies. We repeated DAP induction six times and compared the results of whole-genome sequencing (WGS) and whole-exome sequencing (WES) obtained from original peripheral blood cells, undifferentiated iPSCs, and differentiated cells on days 12 and 26.

We extensively investigated mutations in the 686 cancer-related genes listed in the Catalog of Somatic Mutations In Cancer (COSMIC) Census (COSMIC version 83). We also...
Fig. 1 Characterization of human iPSC-derived dopaminergic progenitors. a An overview of the pre-clinical study. b–d Representative pictures of b iPSCs, c day-12 cultured cells before sorting, and d day-26 cultured aggregate spheres. Bars = 200 μm (n = 25 independent experiments). e Scheme of the cell production process in the GMP facility. f–i Immunofluorescence images of day-26 aggregate spheres sliced. Bars = 100 μm (n = 25 independent experiments). j The results of the LC-MS/MS analysis of the dopamine release by day-56 cultured cells under high potassium stimulation. Values are the mean ± SD (n = 5 independent experiments). k Representative image of whole-cell patch-clamp recordings. The patched cell was stained with Biocytin Alexa Fluor™ 488. Bar = 100 μm. l Representative current-clamp recording of spontaneous action potentials. m Representative current-clamp recording of action potentials induced by brief current pulses. k, l n = 2 independent experiments, the total number of cells = 18. PCS primary cell stock, SCS secondary cell stock, MBM master cell bank, QC quality control.

Table 1 Characterization of the final product.

| Test items                | Test method                      | Criteria                                           | Results (n = 25)                  |
|---------------------------|----------------------------------|---------------------------------------------------|----------------------------------|
| Differentiation/undifferentiation markers | Flow cytometry (day 12)          | TRA-1–60% (pre-sort)                              | 0.0% (n = 24)                    |
|                           |                                  | CORIN >90% (post-sort)                            | 31.35 ± 12.72%                  |
|                           |                                  |                                                   | 93.15 ± 2.127%                  |
| Morphology                | Microscopic observation          |                                                   | Complied                        |
| Viability                 | Flow cytometry: Cell count       | >90%                                              | 97.48 ± 1.608%                  |
| DAP markers               | Flow cytometry: FOXA2/TUJ1       | >80%                                              | 92.29 ± 3.964%                  |
| Undifferentiated markers  | qPCR: POU5F1 and LIN28           | <1% compared to undifferentiated cells            | POU5F1: 0.08 ± 0.15%            |
|                          |                                  |                                                   | LIN28: 0.14 ± 0.13%             |<0.1%                        |
| Early neural markers      | Flow cytometry: OCT3/4/TRA-2–49/6E | <0.1%                                           | <0.1%                           |
| Sterility                 | Flow cytometry: SOX1/PAX6        |                                                   | Negative                         |
| Mycoplasma                | JP17 (membrane filter)           |                                                   | Negative                         |
| Endotoxin                 | JP17 (PCR)                       |                                                   | Negative                         |
|                           | qPCR (LAL turbidity test)        | ≤1 EU·ml⁻¹                                        | Not analyzed                     |
| Residual plasmids         | qPCR: Transfected genes          | Not detected                                      | No abnormalities detected        |
| CVN                       | SNP array                        |                                                   | (n = 6)                          |
| SVN/Indel                 | Whole-genome sequencing          |                                                   | (1) Shibata’s gene list (Supplementary Table 4) |
|                           | Whole-genome sequencing          |                                                   | (2) COSMIC census                |
|                           | Whole-exome sequencing           |                                                   | (3) HGMD database                |

LAL, limulus amebocyte lysate.

included 242 genes in Shibata’s gene list (a cancer-related gene list made by the PMDA; Supplementary Table 4) and mutations annotated with the Human Gene Mutation Database (HGMD) PRO database (2016.4). No genomic mutations were detected by WGS (Table 2). WES revealed single-nucleotide variants (SNVs) in three genes, but this discovery was not consistently observed and thought to be a false positive after considering the results of the amplicon sequencing (Supplementary Tables 5, 6). In addition, there were no residual plasmids or copy number variations (CNVs) in all samples tested.

Regarding epigenetic modifications, we examined the methylation ratio at the transcriptional start site of 73 cancer-related genes. Our final products showed lower methylation levels than those of tissues with brain malignant tumor glioblastoma multiforme and similar levels with normal cells and fetal ventral mesencephalon (Fig. 2a and Supplementary Fig. 2).

To examine the variation of the gene expressions among the six independent preparations, we performed a single-cell RT-qPCR analysis. In each cell preparation, iPSCs on day 0, CORIN+ cells on day 12, and the final products on day 26 showed similar expression patterns in t-distributed stochastic neighbor embedding (tSNE) plots (Fig. 2b). In addition, there was no difference in the expression of genes related to DA differentiation (Fig. 2c, Supplementary Data 1). These results support the reproducibility of the cell manufacturing process.

Tumorigenicity, toxicity, and biodistribution studies. The same final products used for the genomic and epigenetic analyses were subjected to tumorigenicity studies. We repeated DAP induction six times, and each DAP population was injected into the right striatum, which is the same target used in the clinical trial, of immunodeficient NOG (NOD.Cg-PrkdcsidIl2rgm1Sug/Jiljic) mice. In this study, the toxicity and biodistribution of the cells were also examined. After discussion with the PMDA, we observed the animals as long as possible, namely for a life-long period. Initially, we grafted day-30 spheres (2 × 10⁵ cells per mouse, maximum dose) into 80 (males: 40, females: 40) and 50 mice (males: 25, females: 25) as the cell product-transplanted group and saline-injected control group (Supplementary Table 7), respectively, and observed the groups until the number of surviving male or female mice was 30 (transplantation group) or 20 (control group). At 52 weeks after transplantation (59–60 weeks old), the number of male mice in the control group was 19, at which point we stopped the observation. At this time, 28 males and 27 females survived in the transplantation group, and 19 males and 22 females survived in the control group. A survival curve of the animals showed no significant difference between groups (log-rank test, p = 0.3208, Fig. 3a). No changes were attributable to the transplantation in the mice’s general conditions, body weight, food consumption, pharmacological conditions (modified Irwin test), ophthalmological findings, urinalysis, hematology and blood chemistry data, necropsy, or organ weight. Hematoxylin–eosin (H–E) staining of 5-μm-thick brain slices revealed no proliferative or malignant findings, and immunostaining for tyrosine hydroxylase showed that DA neurons had survived (Fig. 3b–e). In 23 (12 males and 11 females) of the 55 mice in the transplantation group, tubular cell clusters were found in the grafts and expressed pan-cytokeratin and...
transthyretin (TTR), suggesting that these cells have characteristics of choroid plexus epithelial cells (Supplementary Fig. 3a, b). The cell clusters were very small (<0.5 mm in diameter), not frequent (mean 1.4 clusters per graft), and not proliferating (no expression of Ki67). In addition, they showed no effect on the survival or neurite extension of surrounding DA neurons. These results verified no tumorigenicity of the final products. Toxicity and biodistribution were also investigated using the same animals (see “Methods”), no abnormal findings were observed in the toxicity study, and grafted human cells (KU80+) were found only in the brain and not in systemic organs.

**Teratoma formation assay in subcutaneous space.** We further examined the existence of residual iPSCs in the final products by injecting day-30 spheres into the subcutaneous space of NOG mice with Matrigel, which is reported to be the most sensitive method to detect teratoma formation by residual undifferentiated iPSCs. As a spike control, 0.001–10% of undifferentiated iPSCs (MCB003) were mixed into the day-30 spheres. As a positive control, another cell line of iPSCs (201B7, established by a retroviral vector with c-Myc) and HeLa cells were used. When 600,000 HeLa cells or 201B7 cells were injected, subcutaneous tumors became detectable at around 10 weeks. In contrast, there was no tumor formation for 26 weeks in the DAP group, DAP with spiked MCB003 group, or even MCB003 group (Fig. 3f). Histological analysis revealed that KU80+ human cells survived in the DAP groups. On the other hand, no KU80+ cells were detected in the MCB group, suggesting that undifferentiated MCB003 cells cannot survive in the subcutaneous space (Fig. 3g–o). Similarly, MCB003 cells did not form any tumors in the mouse striatum, while 201B7 cells formed large teratomas (Supplementary Fig. 3c). When MCB003 cells were injected into the testes of NOG mice, however, they formed typical teratomas, indicating that they were viable and pluripotent (Supplementary Fig. 3d). These results suggested that undifferentiated MCB003 cells do not remain in day-30 spheres, and that even if they did, the risk of teratoma formation is negligible.

**Efficacy study using 6-OHDA-lesioned rats and MPTP-treated monkeys.** To confirm the efficacy of clinical-grade iPSC (MCB003)-derived DAPs, day-30 spheres (4 × 10^5 cells per rat) were transplanted into the striatum of 6-OHDA-lesioned nude rats (n = 8, vehicle control, n = 6), and methamphetamine-induced rotation was evaluated every 4 weeks. In the transplanted group, rotational asymmetry was reversed to normal levels by 16 weeks, while the control group showed no change in rotational behavior (Fig. 4a, two-way analysis of variance (ANOVA), Sidak’s multiple comparison test, p value: *** = 0.0002; ****<0.0001). Immunohistochemistry showed 2835 ± 2534 TH+FOX2+ DA neurons survived and extended axons in the striatum (Fig. 4b–h).

For the clinical trial, we have developed a long-thin needle that is attachable to a stereotaxic frame. To examine the usability of the needle, MCB003-derived day-30 spheres (1.5–2.0 million cells per monkey) were transplanted into the left putamen of MPTP (1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine)-treated monkeys (n = 3). Histological analysis 6 months after the transplantation revealed that the grafted DA neurons survived without any adverse effects, including hemorrhage or inflammation (Fig. 4i–n). The results of the in vivo pre-clinical studies are summarized in Supplementary Table 8.

**Discussion**

For the clinical application of a cell-based therapy, a 2 × 2 matrix (Safety, Efficacy × Cellular and Non-cellular components; Supplementary Table 9) is recommended for quality control of the final product. In our case, the active cellular component is iPSC-derived DAPs. We confirmed that 92.3% of the total cells are FOX2+TUJ1+ DAPs on culture day 26. We also confirmed that some of the survived human cells in rat were positive for GFAP, which suggests glial progenitors were included in the grafted cells with DAPs. The DAPs became mature DA neurons that produced dopamine and improved the abnormal behaviors of 6-OHDA-lesioned rats. Two previous clinical trials for PD using PSCs have transplanted ESC-derived neural progenitor cells (NCT02452723) and ESC-derived DA neurons (NCT03119636). These pre-clinical studies are summarized in Table 3.

There are three cellular components that should not be contained in the final product. The first is undifferentiated iPSCs, and we confirmed their absence by four methods in vitro. The second is early neural stem cells. Previously, we found that SOX1+ PAX6+ cells exist in the early stage of differentiation and that these cells form rosettes and proliferate in the brain. The expression of CORIN, however, is never common with SOX1 or PAX6 expression. As a result, we can eliminate SOX1+PAX6+ cells by sorting CORIN+ cells. The third is transformed cells. The genomic instability of iPSCs and their derivatives may lead to tumorigenicity. However, the need for genomic analysis is controversial, because genomic abnormalities do not always cause tumors. In addition, there is no standard gene list for which we should investigate. In this situation, we decided to investigate the genes listed in COSMIC Census and the cancer-related gene list made by the PMDA as well as familial disease-related genes listed in a HGD PRO database. We performed WGS and WES of both iPSCs and the final products, but detected no aberrations in these genes. Epigenomic changes in iPSCs are another possible

### Table 2 Summary of the genome analysis.

| Preparation | Residual plasmid | SNV/Indel (Shibata’s gene list/COSMIC Census-listed genes) | CNV Mutations in HGMD |
|-------------|------------------|----------------------------------------------------------|------------------------|
| CTIDAP-161004 | Day 0 None | 0 0 0 None | WGS Exome |
| CTIDAP-161011 | Day 0 None | 0 0 0 None |
| CTIDAP-161018 | Day 0 None | 0 0 0 None |
| CTIDAP-161025 | Day 0 None | 0 0 0 None |
| CTIDAP-161028 | Day 0 None | 0 0 0 None |
| CTIDAP-161101 | Day 0 None | 0 NCKIPSD 0 None |

NA, not analyzed.
cause of tumorigenicity, because iPSCs are reported to have higher variability in DNA methylation\textsuperscript{22}. From our results of a methylation array, iPSCs and the final products were not hypermethylated compared with malignant brain tumor tissue and instead showed a methylation rate consistent with normal cells.

Tumorigenicity studies in vivo are also not standardized. In previous pre-clinical studies, the observation periods in nude rats and SCID (severe combined immunodeficiency) mice were 9 months (Table 3)\textsuperscript{10,11,23}. In the case of spinal cord injury, ESC-derived oligodendrocyte progenitors (two million cells) were injected into the spinal cord of SCID mice, and the histological analysis was performed 12 months later\textsuperscript{24}. In the present study, we used NOG mice, in which not only T cell and B cell activities but also NK cell activity is reduced, thus reducing the immune response.
The observation period was 52 weeks, which approximates the mouse lifespan. Because we found no proliferating cells or abnormal findings in the grafts, 52 weeks (or 12 months) seem to be long enough for tumorigenicity study. Of note, before starting the pre-clinical study, we consulted the PMDA about the study design. The PMDA requested we use NOG mice, which is the least immunogenic animal model, is sensitive to tumor development, and offers a long observation period. The PMDA also requested we evaluate at least 10 animals per group in the histological analysis. Major discussion points with the PMDA and our solutions are summarized in Supplementary Table 10.
Regarding non-cellular components, we need to use clinical- or GMP-grade materials and reagents. We have made a GMP-grade anti-CORIN antibody and carefully confirmed that its remnant on cells was negligible. Regarding donor eligibility for iPSCs, a medical interview and blood exam for contamination were conducted. For other materials, γ-ray irradiation and viral elimination were applied to meet Japanese standards for biological raw materials. In addition, following guidelines of the International Council for Harmonization of Technical Requirements for Pharmaceuticals for Human Use (ICH) Q5A, we confirmed that the clinical-grade iPSCs (MBC003) were not contaminated with pathogenic viruses (Supplementary Table 11).

Unexpectedly, choroid plexus epithelia-like cells were observed in 23 of 55 mice with transplantation of the final product. In all cases, the cell clusters were microscopic (<0.5 mm), not frequent, not proliferating, and asymptomatic. Because bone morphogenic protein (BMP) signaling is essential for developing choroid plexus tissue26, we modified our differentiation protocol to extend the period of the BMP inhibition (Supplementary Fig. 4a). Accordingly, we successfully reduced the expression of TTR, a marker for the choroidal plexus, in the final product, without any changes of other DA markers (Supplementary Fig. 4b, c). Transplantation to nude rat resulted in TTR+ cells appearing in two out of seven rats without the BMP inhibitor, but in no rats out of six rats with the BMP inhibitor (Supplementary Fig. 4d). We also confirmed that extension of the BMP inhibition can reduce the risk of inducing TTR+ epithelium-like cells in the brain.

In conclusion, the safety and efficacy of clinical-grade cell products were confirmed in this study. Based on these results, we received approval to start a clinical trial for PD from the PMDA and the institutional regulatory board of Kyoto University. The trial is ongoing.

Methods

Establishment of iPSC stock and manufacturing of DAPs. This study was approved by the ethics committee of Kyoto University Hospital and Center for iPS Cell Research and Application (CiRA), Kyoto University, Kyoto, Japan. After informed consent was obtained from an HLA-homozygous volunteer donor, an iPSC stock for clinical use was established by reprogramming the donor’s peripheral blood with episomal plasmid vectors expressing OCT3/4, SOX2, KLF4, L-MYC, LIN28, MPP3LI3D, and EBNA1 at a cell processing center (Facility for iPSC Therapy, CiRA). The peripheral blood cells were isolated by using Ficoll-Paque PLUS (GE Healthcare), and 1.2 × 10^6 mononuclear cells were cultivated with StemFit AK03 without solution C (contains basic fibroblast growth factor) media (Ajinomoto) with 30 ng·mL^-1 interleukin-6 (IL-6), 50 ng·mL^-1 stem cell factor, 10 ng·mL^-1 thrombopoietin, 20 ng·mL^-1 Flt-3 ligand, 20 ng·mL^-1 IL-3, and 10 ng·mL^-1 granulocyte colony-stimulating factor (all WAKO) in four wells of a 24-well plate (3 × 10^5 cells per well). After 7 days of cultivation, the vectors were induced in 5 × 10^5 dissociated cells by a Nucleofector 4D electroporation system (Lonza), and the cells were replated on laminin 511-E8 fragment (iMatrix, Nippon)-coated 6-well plates (1.67 × 10^5 cells per well) in the same media as the mononuclear cells. One milliliter per well of StemFit AK03 media (Ajinomoto) was added 3, 5, and 7 days after the induction, and 9 days onward StemFit AK03 media were exchanged every 3 days. After 3 weeks of cultivation, single-cell-derived colonies became visible, and we picked up 15 of them manually. Each colony was dissociated with TrypLE Select CTS (Thermo Fisher), and all the cells were transferred to an iMatrix-coated 12-well plate and defined as passage 1 (P1). The P1 cells were passaged at 1.4 × 10^5 cells cm^-2 every 8–12 days, and 30 vials of a primary cell stock (PCS) were frozen at P4. Two out of fifteen PCS clones were identified as iPSCs by ESC-like morphology, the absence of residual plasmids, confirmation of a normal karyotype, the expression of surface markers of undifferentiated iPSCs, and efficient neural differentiation. We selected the clones with the best efficiency for DA differentiation for the transplantation experiments. In order to confirm that the iPSCs were plasmid free by a longer culture, one vial of the PCS was thawed (P5) and expanded to 70 vials of secondary cell stock (SCS, P9). After thawing the SCS, we confirmed that there was no residual plasmids (tested at P9, P12, and P19), that they expressed undifferentiated markers (TRA-1–60, SSEA-4, and TRA-2–49; tested at P15), and had normal karyotypes (46, XY; tested at P12 and P13). One vial of the SCS was thawed and passed two times, and then 384 frozen vials of MCB (P12) were stored. All freezing procedures were performed by using a programmed freezer. iPSCs were dispensed at 2 × 10^5 or 5 × 10^5 cells per tube in 200 or 500 μL of Stem Cell Banker GMP grade (TAKARA Bio, CB045) and frozen to ~80 °C at ~1°C·min^-1, and stored in the gas phase of a liquid nitrogen tank.

Frozen MCB was differentiated after two passages of maintenance culture on iMatrix in StemFit AK03 medium was conducted. To manufacture the DAPs, the iPSCs were dissociated into single cells after 10-min incubation with 0.5× TrypLE select CTS (Thermo Fisher) and treated with TrypLE select CTS (1:100) on iMatrix-coated 12-well plates to a density of 5 × 10^6 cells well^-1 in differentiation media containing Glasgow’s minimum essential medium (Thermo Fisher) supplemented with 8% γ-ray-irradiated knockout serum replacement (Thermo Fisher), 0.1 mM MEM (minimum essential medium) non-essential amino acids (Thermo Fisher), 1 mM sodium pyruvate (Sigma-Aldrich), 0.1×10^−4 M dibutyryl cAMP, 5 μM Y-27632 (R&D System), 50 μM A83-01 (Sigma-Aldrich) and had normal karyotypes (46, XY; tested at P12 and P13). One vial of the SCS was thawed and passed two times, and then 384 frozen vials of MCB (P12) were stored. All freezing procedures were performed by using a programmed freezer. iPSCs were dispensed at 2 × 10^5 or 5 × 10^5 cells per tube in 200 or 500 μL of Stem Cell Banker GMP grade (TAKARA Bio, CB045) and frozen to ~80 °C at ~1°C·min^-1, and stored in the gas phase of a liquid nitrogen tank.

Flow cytometry. Flow cytometric analysis was performed using cells on differentiation days 12 and 26. On differentiation day 12, pre-sorted cells were stained by fluorescein isothiocyanate (FITC)-conjugated anti–TRA-1–60 antibody (1:5, BD #560308) for 20 min, followed by dead-cell staining by 7-aminoactinomycin D (1:100, BD #559925). On differentiation day 26, cultured spheres were dissociated into single cells by Neuron Dissociation Solutions 5 (FUJIFILM Wako Pure Chemical Corporation) and stained by the live/Dead® Fixable Cell Stain Kit (Thermo Fisher) and FITC-conjugated anti–TRA-2–49 antibody (Merck-Millipore), followed by fixation by 4% paraformaldehyde for 30 min. After fixation, the cells were permeabilized by Perm/Wash buffer (BD Biosciences #554723) for 30 min, blocked with antibodies for intracellular staining (Supplementary Table 13), and rinsed with Perm/Wash buffer. Stained samples were analyzed by Canto II or FACSARia III flow cytometer. The compensation setting of each fluorescence was...
Weeks after transplantation
Rotation min⁻¹

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0 4 8 12 16 20
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**a**

**b**

**c**

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**e**

**f**

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**j**

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**l**

**m**

**n**
adjusted by using the ABC Anti-Mouse Bead Kit and ArC Aminie Reactive Com-
ensation Kit (Thermo Fisher).

Quantitative reverse transcription with polymerase chain reaction (qRT-
PCR). The total RNA fraction was extracted by NucleoSpin RNA XS (Takara Bio) and reverse transcribed by the SuperScript III First-Strand Synthesis System (Qagen) or PrimeScript™ RT reagent Kit with gDNA Eraser (Takara Bio). Quantitative PCR was carried out with TaqMan® Gene Expression Master Mix (Thermo Fisher) according to the manufacturer’s instructions. The expression levels of the genes were normalized to GAPDH using the standard curve method. Probes and primers are as follows: GAPDH probe, forward and reverse: Hs02758991_g1 (Thermo Fisher), POUSF1 probe: CGCAGTGCCCTGTCGCTGCC, POUSF1 forward: GTGGAGAAGCACTCGGATG, POUSF1 reverse: CAATTTGGCTGGCTTCTCTCG, LIN28 probe: CCAGATGGTGCTGCTCTGTCG, LIN28 forward: CACGGTCGCGGCTACTGTG, LIN28 reverse: CTCCTCACTGGTACCATCT.

Immunostaining. For in vitro studies, spheres were fixed with 4% paraformalde-
yde and sliced at 10 μm thickness by using a cryostat (CM-1850, Leica Biosys-
tems). Double- or triple-labeled immunostaining was performed after permeabili-
zation, antigen retrieval if needed, and blocking with 0.1% Triton X-100. The stained samples were visualized by using a fluores-
cence microscope (BX-6701; Keyence) and a confocal laser microscope (Fluoview
FV1000D; Olympus). The primary and secondary antibodies are listed in Sup-
plementary Table 13.

Single-cell analysis. Single-cell complementary DNA (cDNA) preparation and
gene expression analysis were performed following the manufacturer’s protocol. A cell
suspension of iPSCs (day 0), intermediate cells after sorting (days 12 and 13),
or DAoPs (day 28) at 300 cells μl−1 were used. C1 Suspension Reagent (Fludigm)
was added to the cell suspension at a ratio of 40%. Six microliters of cell sus-
pension mix were loaded on a C1 Single-Cell AutoPrep IFC microfluidic chip
(Fludigm) designed for cells 10–17 μm in size, and the chip was then processed on a
Fluidigm C1 instrument using the ‘STA: Cell Load (1862x)’ script. Live and dead cells
were confirmed under a fluorescent microscope. Then, the microfluidic chip was
loaded onto a BioMark chip (Fluidigm) for expression profiling with a panel of qRT-
PCR assays (Biokard HD, Fluidigm). The chip was then loaded onto Biomark HD to
analyze gene expression levels. The expression data were processed and ana-
alyzed using the Singular Analysis Toolset version 3.6.2 (Fluidigm). The gene
expression profiles of the analyzed cells were visualized using tSNE. A spec-
trum of gene expressions was visualized by violin plots. The sequences of the
DNA primers are listed in Supplementary Table 14.

Electrophysiological analysis. To analyze the electrophysiological properties of
cultured neurons, 42-day cultured spheres were attached on L-ornithine/laminin/
bronectin-coated glass coverslips. After 14 days, whole-cell patch-clamp record-
ings were performed with glass microelectrodes on the surface of the spheres for the
examination. The spheres were maintained in a physiological saline solution of
the following composition: 125 mM NaCl, 2.5 mM KCl, 2 mM CaCl2, 1 mM MgCl2,
26 mM NaHCO3, 1.25 mM NaH2PO4, and 17 mM glucose. Patch pipettes were
made from borosilicate glass capillaries (1B150F-4; World Precision Instruments) and
had a resistance of 3–10 MΩ when filled with an internal solution composed of
140 mM KCl, 10 mM HEPES, and 0.2 mM EGTA (pH 7.3). To visualize the patch-
ched cells, 100 μM Biocytin Alexa Fluor® 488 was added to the internal solution.
Recordings with a voltage clamp and current clamp were made with a patch-clamp
amplifier (Axopatch 200B; Molecular Devices). The giga-seal resistances were in
the range of 10–20 GΩ. The current signals from the amplifiers were filtered at
1 kHz and stored and analyzed on a 64-bit computer (Coster Master). All
experiments performed at room temperature.

Dopamine measurement by LC-MS/MS. Spheres on differentiation day 28 were
replated onto L-ornithine/laminin/bronectin-coated 12-well plates with neural
differentiation medium and cultured for 28 days. The medium was replaced 24 h
before the measurements. On culture day 56, the cells were washed twice with low
KCl solution (20 mM HEPES-NaOH, pH 7.4, 140 mM NaCl, 4.7 mM KCl, 2.5 mM
CaCl2, 1.2 mM MgSO4, 1.2 mM KH2PO4, and 11 mM glucose) and incubated in
low KCl solution for 15 min. The medium was subsequently replaced with 1 mL
of high KCl solution (20 mM HEPES-NaOH, pH 7.4, 85 mM NaCl, 60 mM KCl,
2.5 mM CaCl2, 1.2 mM MgSO4, 1.2 mM KH2PO4, and 11 mM glucose) for 15 min.
The solution was then collected, and the concentration of dopamine was deter-
mined by LC-MS/MS (Sumika Chemical Analysis). Plated cells were harvested in
PBS and sonicated. DNA concentration of the cell lysate was measured using the
Quant-iT™ dsDNA Assay Kit (Thermo Fisher) and used for compensation of the
da concentration. Data were calculated from five independent experiments.

Extended culture on iPSC maintenance condition. Cells on culture day 26 or 30
were dissociated to single cells and replated onto Matrigel-coated 24-well plates at
2 × 105 cells per sample with StemFit AK031 iPSC maintenance media. Undifferenti-
ated iPSCs were dissociated as well, and iPSCs were spiked into 2 × 106 DAoPs at 0.001–1%.
After 14 days, the cells were stained by alkaline phosphatase using the AP
Staining Kit (System Biosciences), and the colonies were counted manually.

Genomic/epigenomic analysis and testing residual plasmas. WGS and WES
libraries were prepared from 200 and 100 ng of genomic DNA as starting
materialss following the manufacturer’s instructions, respectively. For WGS, libraries
were made with the KAPA Hyper Prep Kit (Kapa Biosystems) without PCR. For WES, adapter-
ligated libraries were prepared with the KAPA Hyper Prep Kit, and sequencing
libraries were generated with the SeqCap EZ Human Exome Enrichment v3.0 (Roche).
Cluster generation was done with the HiSeq PE Cluster Kit v4-cBot (illumina) using
illumina cBot. Sequencing was performed with the HiSeq SBS Kit v4 using HiSeq2500
and 2 × 126 PE mode. After generating FASTQ files by bcl2fastq v2.17.11.14 (illumina)
and performing adapter trimming by cutadapt 1.1097, FASTQ files were mapped to
the reference human genome (hg19) by BWA MEM90 (ver. 0.7.13) (for Genomon2,
–T 0 option was used), and duplicated reads were removed by NovoSort (Novo-
craft) (ver. 1.0.9). To call SNV/Inserts in test samples compared with donor cells, bam files
were analyzed by Genomon31 (ver. 1.0.1) with Fisher’s exact test (P < 0.001) and
Genomon242 (ver. 2.3.0) with EB call (P(Fisher) < 0.1 and P(EB call) < 0.001 for
WGS, and P(Fisher) < 0.1 and P(EB call) < 0.001 for WES), followed by adapting the
filter criteria. Mutations were annotated by ANNOVAR35 (2016Fe081) and
restricted on CDS and splicing regions. To further extract the no pathogenic
mutations, we excluded synonymous mutations and focused on mutations annotated
with HGMD44, COSMIC Cancer Gene Census36 (ver. 83), and Shibata’s gene list
(http://www.pmda.go.jp/gr/files/000152599.pdf). WGS data were utilized for calling
CNVs using VarScan246 (2.4.2) in combination with Otou’s threshold method37 and
Delly90 (0.7.3) by comparing test samples with donor cells, followed by curating the
called CNVs manually. In addition to utilizing WGS data, CNVs were called using an
SNP genotyping array. Genomic DNA (starting from 200 ng) was hybridized onto the
HumanOmniExpress24 v1.2 DNA Analysis Kit (illumina), and intensities were
scanned by Iscan (illumina) following the manufacturer’s protocol. After exporting a
final report using GenomeStudio (2011.1) (illumina), CNV and WES were extracted
using PennCNV90 (1.0.3), Mosaic Alteration Detection-MAT)90 (1.0.1), and
GWAS tools10 (1.16.1), followed by comparing test samples with original cells and curating
the called CNVs manually.

For the methylation analysis, the bisulfite conversion of 500 ng genomic DNA
was performed using the EZ DNA Methylation Kit (Zymo Research), followed
by methylation profiling using the Infinium MethylationEPIC BeadChip Kit
(illumina) according to the manufacturer’s protocols. After exporting the
methylation data by using GenomeStudio (2011.1) and normalizing the beta
values using by BMiQ27, we calculated the beta values of 73 genomic blocks at the
transcription start sites of cancer-related genes as described previously28 and
visualized them using R (https://www.R-project.org) with ggplot234. As a reference,
53 methylation datasets of normal tissues (in GSE31848) and 11 methylation
datasets of glioblastoma multiforme (downloaded from TCGA, Sentrix_ID:
9976861010) were also visualized.
| Reference          | Cellular product                                         | Criteria                                                                 | In vitro study                     | In vivo study                | Number of animals | Cells/animal       | Evaluation                  | Observation period |
|--------------------|----------------------------------------------------------|---------------------------------------------------------------------------|------------------------------------|-----------------------------|-------------------|-------------------|-----------------------------|-------------------|
| Garitaonandia et al. | Human parthenogenetic stem cell-derived neural stem cells | NESTIN = 96.2%  
MSL1 = 93.5%  
SOX2 = 97.8% | Culture in ESC condition  
FCM  
QPCR  
Immunofluorescence  
Identification analysis  
PCR | Nude rat  
N = 15  
N = 4 (control)  
N = 80  
N = 20 (control)  
N = 3 (FP)  
N = 4 (EB)  
N = 3 (control) | 2.5 × 10^6  
7.7 × 10^6 | Acute toxicity  
Tumorigenicity  
Biodistribution | 7 days  
9 months |
| Wang et al.         | Human parthenogenetic ESC-derived DA neurons            | TUJ1 = 99.8%  
TRA-1-60 = 0.1%  
SSEA4 = 0.1% | Immuno  
fluorescence  
FCM  
Electrophysiological analysis | MPTP-treated monkey  
Nude mouse  
SCID mouse | Data not shown  
Data not shown | Data not shown  
Data not shown | Tumorigenicity  
Teratoma formation  
Toxicity  
Tumorigenicity  
Biodistribution | 9 months  
12 months  
6 months |
| Present study       | Human iPSC-derived dopaminergic progenitors             | FOXA2/TUJ1 = 92.3%  
OCT4/TRA-2-49 < 0.1%  
SOX1/PAX6 < 0.1% | Culture in iPSC condition  
QPCR  
Immunofluorescence  
Identification analysis  
DA measurement  
Genome/epigenome analysis  
Single-cell analysis | NOG mouse  
(brain)  
NOG mouse  
(subcutaneous) | N = 80  
N = 50 (control) | 2 × 10^5 | Efficacy  
Tumorigenicity  
Biodistribution | 5 months  
2–6 months |
the plasmid using iPSC genome DNA and real-time PCR Thermal Cyclers (StepOnePlus; Thermo Fisher). The DNA quantity was normalized to the copy number of endogenous FBX15. The DNA genomes of more than 25,000 cells were used as the template for the PCRs based on the copy number of endogenous FBX15. The probes and primers are as follows:

- **KLF4** probe, forward and reverse: Hi0058836_m1 (Thermo Fisher), MVL1 probe, forward and reverse: H00024095_m1 (Thermo Fisher), TP53 probe: CAGCCTGGCGATCACG;
- TP53 forward: GGGCGACGCTCTTCTGAG, TP53 reverse: CCAGGATGCTGAGCTAGA, LIN28A probe: CAAAAGGAAGACAGGCTGTA, LIN28A forward: CAAAAGGAAGACAGGCTGTA, LIN28A reverse: CATGATGATCTAGATCCCTACAGTTG, SOX2 probe: ATCTCAAATGGTGCCTCT, SOX2 forward: GCCATTACGCGACACTGC, SOX2 reverse: GAATGTTGCTAGTGACGCTTGG, OCT4/3 probe: TTCAACATGGGCGGACG, OCT4/3 forward: TGCTTCATCATTGTGCCAAAGAGATT, OCT/3 reverse: CGAAGAGGCGGAATCCGGA, EBNA1 probe: TGGCAGGAGACCCCA, EBNA1 forward: ATCCAGGGCAACAGCATAAGTGAT, EBNA1 reverse: GCCAAGGCAATCTGAGGT, FBX15 probe: GAACTGAGGACTTACCAAGAGT, FBX15 forward: GCCAGAGGTCTGTCCGTTGTA, FBX15 reverse: AATGCAGGCCTAGGGTCAAA.

**Amplon sequencing.** Primers for amplon sequencing were designed with primer.py (Amel0ff) and Apet (2.0.34) ([https://jorgensonbiology.uta.edu/wayned/aper/](https://jorgensonbiology.uta.edu/wayned/aper/)). For the multiplex PCR, 10 ng of genomic DNA and 200 nM of primer (primers with KAPA HiFi HotStart ReadyMix (Roche) and 5% dimethyl sulfoxide) were mixed and PCR reaction was performed with the following condition: the initial denature was done at 95 °C for 5 min, followed by 35 cycles of denaturation at 98 °C for 20 s, annealing at 60 °C for 15 s, extension at 72 °C for 30 s, and finally extension at 72 °C for 1 min. After purifying amplon with 1.8x Agencourt AMPure XP Beads for 20 s, annealing at 60 °C for 15 s, extension at 72 °C for 30 s, and denature was done at 95 °C for 5 min, followed by 35 cycles of denature at 98 °C for 1 min, and extension at 72 °C for 1 min. After purification with AMPure XP Beads (Beckman Coulter), we made sequencing library with KAPA Hyper Prep Kit (Roche). Library was sequenced with MiSeq (Illumina) using MiSeq Reagents Kit v2 according to the manufacturer’s instruction. The sequenced reads were aligned to the reference genome by BWA MEM (Ver. 0.7.15) and alternative allele frequencies at the targets were calculated by samtools (ver. 0.1.19)[9].

**Animal studies.** All animals were cared for and handled according to the Guidelines for Animal Experiments of Kyoto University and that of Shin Nippon Biomedical Laboratories (Kagoshima, Japan), and all animal experiments were certified by an ethics committee at Kyoto University and Shin Nippon Biomedical Laboratories.

**Tumorigenicity/toxicity/biodistribution study.** Seven-eight-week-old male and female NOG mice (CLEA Japan) were used for the transplantation. Cells were transplanted stereotactically, and the coordinates of the striatum were calculated with reference to the bregma: anterior (A), +0.5 mm; lateral (L), −2.0 mm; ventral (V), −3.0 mm; and tooth bar (TB), 0 mm. Two hundred thousand cells in 0.6 μL were injected through a 24-G needle (Hamilton 7105KH) into the right striatum (from the bregma: A, +1.0 mm; L, −3.0 mm; V, −5.0 and −4.0 mm; and TB, 0 mm).

**Efficacy study.** Transplantation of human iPSC-DAPs in 6-OHDA-lesioned rats: Adult (1-year-old) male nude male mice (F144/NIH-rnu/rnu, CLEA, Japan) were used for the lesioning. 6-OHDA injection was performed into the medial forebrain bundle, and only rats that showed more than six rotations per minute were used in the cell transplantation experiments. Cell transplantation was performed with the stereotaxic injection of 4 × 10^5 cells (200,000 cells μL^-1^) by using a 22 G needle (Hamilton 7105KH) into the right striatum (from the bregma: A, +1.0 mm; L, −3.0 mm; V, −5.0 and −4.0 mm; and TB, 0 mm). Behavoural analysis: Methamphetamine-induced rotation behavior was evaluated before and every 4 weeks after the transplantation. The rotation was recorded for 90 min after the intraperitoneal injection of methamphetamine (2.5 mg kg^-1^, Sumitomo Dainippon Pharma). The rotation number was counted by the RotoRat software version 2.01 (Med Associates, Inc., Vermont, USA).

**Transplantation of hiPSC-DAP in MPTP model monkey: human iPSC-DAPs were supplied by Shin Nippon Biomedical Laboratories.** Five adult (8 month-old) male cynomolgus monkeys were supplied by Shin Nippon Biomedical Laboratories) with nickel ammonium. The rotation number was counted by the RotoRat software version 2.01 (Med Associates, Inc., Vermont, USA).

**Statistical analysis.** Statistical analyses were performed by using the GraphPad Prism 7 software (GraphPad Software). Data from the survival curve of the transplanted animals were analyzed by the log-rank test (Fig. 3a), and the behavioral data were analyzed by a two-way ANOVA with Sidak’s multiple comparisons test (Fig. 4a). P values < 0.05 were considered significant.

**Data availability.** Most of the data that support the findings of this study are available within the article and its Supplementary Information files or from the corresponding author upon reasonable request. Some of the data are not publicly available due to them containing information that could compromise donor privacy.

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**Reporting summary.** Further information on research design is available in the Nature Research Reporting Summary linked to this article.
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Acknowledgements

We thank Sumitomo Dainippon Pharma Co., Ltd. for providing MCB003, and Dr. Y. Oo (KAN Research Institute) for kindly providing an anti-CORIN and anti-NURRI antibody. We also thank Dr. Y. Maruyama (PMDA) for discussing the regulatory issues, Dr. P. Caragianis for critically reading the manuscript, and Dr. E. Yamasaki, Dr. Y. Ozaki, Ms. Y. Ashida, Ms. Y. Tanikawa, Mr. Y. Fujita, Ms. Y. Katano, Ms. R. Takachi, and Ms. Y. Ishii (CIRA, Kyoto University) for their technical assistance. This study was supported by a grant from the Network Program for Realization of Regenerative Medicine from the Japan Agency for Medical Research and Development (AMED).

Author contributions

D.D. designed the study, collected and assembled the data, performed the data analysis and interpretation, and wrote the manuscript. H.M., T.K., M.I., S.H., and M.U. collected and/or assembled the data. K.Y., N.A., M.N., and A.M. performed the data analysis and interpretation. J.T. conceived and designed the study, assembled the data, carried out the data analysis and interpretation, wrote the manuscript, and made final approval of the manuscript.

Competing interests

J.T. receives a grant for collaborative research by Sumitomo Dainippon Pharma Co., Ltd. M.I., S.H., and K.Y. are employees of Sumitomo Dainippon Pharma Co., Ltd. H.M. is an employee of Shin Nippon Biomedical Laboratories. The other authors declare no competing interests.

Additional information

Supplementary information is available for this paper at https://doi.org/10.1038/s41467-020-17165-w.

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Peer review information Nature Communications thanks the anonymous reviewer(s) for their contribution to the peer review of this paper. Peer reviewer reports are available.

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