Research Article

Increased TNF-α Initiates Cytoplasmic Vacuolization in Whole Blood Coculture with Dengue Virus

Rahmat Dani Satria,1,2,4 Tzu-Wen Huang,4,5 Ming-Kai Jhan,4,5 Ting-Jing Shen,4,5 Po-Chun Tseng,4,6 Yun-Ting Wang,4 Zhen-Yu Yang,4,5 Chung-Hsi Hsing,7,8 and Chiou-Feng Lin4,5,6

1International Ph.D. Program in Medicine, College of Medicine, Taipei Medical University, Taipei 110, Taiwan
2Department of Clinical Pathology and Laboratory Medicine, Faculty of Medicine, Public Health and Nursing, Universitas Gadjah Mada, Yogyakarta 55281, Indonesia
3Clinical Laboratory Installation, Dr. Sardjito Central General Hospital, Yogyakarta 55281, Indonesia
4Department of Microbiology and Immunology, School of Medicine, College of Medicine, Taipei Medical University, Taipei 110, Taiwan
5Graduate Institute of Medical Sciences, College of Medicine, Taipei Medical University, Taipei 110, Taiwan
6Core Laboratory of Immune Monitoring, Office of Research & Development, Taipei Medical University, Taipei 110, Taiwan
7Department of Medical Research, Chi Mei Medical Center, Tainan 710, Taiwan
8Department of Anesthesiology, Chi Mei Medical Center, Tainan 710, Taiwan

Correspondence should be addressed to Chung-Hsi Hsing; hsing@mail.chimei.org.tw and Chiou-Feng Lin; cflin2014@tmu.edu.tw

Received 12 November 2020; Revised 9 April 2021; Accepted 26 April 2021; Published 7 May 2021

Academic Editor: Luiz Felipe Domingues Passero

Copyright © 2021 Rahmat Dani Satria et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

During the acute febrile phase of dengue virus (DENV) infection, viremia can cause severe systemic immune responses accompanied by hematologic disorders. This study investigated the potential induction and mechanism of the cytopathic effects of DENV on peripheral blood cells ex vivo. At one day postinfection, there was viral nonstructural protein NS1 but no further virus replication measured in the whole blood culture. Notably, DENV exposure caused significant vacuolization in monocytic phagocytes. With a minor change in the complete blood cell count, except for a minor increase in neutrophils and a significant decrease in monocytes, the immune profiling assay identified several changes, particularly a significant reduction in CD14-positive monocytes as well as CD11c-positive dendritic cells. Abnormal production of TNF-α was highly associated with the induction of vacuolization. Manipulating TNF-α expression resulted in cytopathogenic effects. These results demonstrate the potential hematological damage caused by ex vivo DENV-induced TNF-α.

1. Introduction

Dengue virus (DENV) infection is one of the most critical global health problems, especially in subtropical regions. Unfortunately, DENV causes disease in 50–100 million individuals per year [1]. Dengue-infected patients have different manifestations ranging from mild acute febrile illness, dengue fever, and dengue hemorrhagic fever to severe dengue shock syndrome, leading to plasma leakage hypovolemic shock, causing death [2]. In addition to hematologic disorders, patients with severe dengue infection may display various diseases, including multiple organ dysfunction and neurological complications [3]. According to clinicopathological studies, hematologic changes, such as leukopenia and thrombocytopenia, are possibly involved in the coagulopathy and vasculopathy of dengue-infected patients.
following the acute febrile phase of infection [4]. In dengue-related hematological pathogenesis initiation, a direct viral attack and indirect host effects are generally involved [5–8].

The complex interaction between the host and viral factors makes it challenging to explain the pathogenesis of DENV infection. However, it is believed that the main factors causing disease severity are typically due to numerous host factors. Several hypotheses have been formulated to explain severe dengue causes, including genetic involvement, underlying disorders, viral load, viral virulence, and immune responses [9–12]. Antibody-dependent enhancement is assumed to be pathogenic, mainly in the secondary infection of DENV, when patients are infected with a different serotype from the previous one [13]. Also, the imbalance of cytokines in some patients, namely, the cytokine storm, was related to dengue disease severity [14, 15]. Clinical studies have shown that tumor necrosis factor-α (TNF-α) is associated with increased severity and progression of DENV infection due to exacerbated proinflammatory cytokine production leading to instability in vascular endothelial cell function [16–18]. The impact of serum TNF-α on immune cells remains undefined.

Innate immune cells, such as neutrophils and monocytes, are believed to have an essential role in dengue pathogenesis [19, 20]. There is increased neutrophil degranulation in patients with DENV infection, as indicated by an increase in the levels of interleukin-8 (IL-8), elastase, and lactoferrin [21]. Following degranulation, in the acute febrile phase of DENV infection, there is a significant reduction in neutrophil counts, namely, neutropenia, in most patients with dengue disease [22]. Moreover, DENV infection triggers neutrophil activation and degranulation during the febrile phase, associated with increased plasma levels of proinflammatory mediators, such as IL-8 and TNF-α. Upon hematomal immunity, activated neutrophils and monocytes can destroy microbes by releasing various toxic components, such as reactive oxygen species and granular enzymes [20, 21, 23]. The establishment of vacuolization is caused by the fusion process of endosomes, autophagosomes, and secretory vesicles [24]. In this study, we investigated the induction of cellular vacuolization and its possible regulation by TNF-α in an ex vivo whole blood (WB) model of DENV infection.

2. Materials and Methods

2.1. Antibodies and Reagents. The reagents and antibodies (Abs) used were as follows: recombinant human TNF-α (hTNF-α, PeproTech, Rocky Hill, NJ); crystal violet (Sigma-Aldrich Co., St. Louis, MO, USA); neutralizing antibodies against TNF-α (Abcam, Cambridge, MA); PerCP-conjugated anti-CD4 (Catalog# MA197755); PE-Cyanine 7-conjugated anti-CD8 (Catalog# 25-0086-42); PE-conjugated anti-CD11c (Catalog# 12-0116-42); APC-conjugated anti-CD14 (Catalog# 17-0149-42); eFluor 506-conjugated anti-CD19 (Catalog# 69-0199-42); APC-eFluor 780-conjugated anti-CD25 (Catalog# 47-0257-42); Super Bright 600-conjugated anti-CD56 (Catalog# 63-0566-42); Qdot 705-conjugated anti-HLA-DR (Catalog# Q22159) (Invitrogen, Thermo Fisher Scientific, Waltham, MA); Alexa Fluor 488-conjugated anti-CD16 (Catalog# 302019); and Alexa Fluor 700-conjugated anti-CD62L (Catalog# 304820) (BioLegend, San Diego, CA).

2.2. Cell Culture and Virus Culture. Baby hamster kidney- (BHK-21) 21 cells (ATCC® CCL-10™) and Aedes albopictus clone C6/36 cells (ATCC® CRL-1660™) were cultured in Dulbecco’s Modified Eagle’s Medium (DMEM, Invitrogen Life Technologies) containing 10% heat-inactivated fetal bovine serum (FBS) (Sigma-Aldrich). The DENV serotype (DENV2 PL046) was maintained in C6/36 cells. C6/36 cell monolayers were seeded in a 75 cm² tissue culture flask with DENV coculture at a multiplicity of infection (MOI) of 0.01 and incubated at 28 °C in 5% CO₂ for 5 days. The virus supernatant was concentrated and filtered with Amicon Ultra centrifugal filters (Millipore, Billerica, MA, USA) and then stored at −80 °C before use.

2.3. Human Blood Collection. The human study was performed according to guidelines established by the Taipei Medical University- (TMU-) Joint Institutional Review Board (TMU-JIRB). Informed consent from all participants, as approved by TMU-JIRB, was obtained. All five participants were volunteers with confirmed functional health status and good physical condition who were free from medication and had no current infectious disease. All samples were collected at the same time by sodium heparin BD vacutainer collection tubes (5 ml; Becton Drive Vacutainer, Franklin Lakes, USA). All blood collection tubes were gently inverted to mix additives with the blood after collection.

2.4. DENV Ex Vivo Infection. One hundred microliters of WB was seeded in a 24-well plate supplemented with 100 μl of Roswell Park Memorial Institute (RPMI) 1640 medium containing DENV (MOI = 1). For infection, the total number of WB leukocytes was calculated using a hematology analyzer as described below. The WB was incubated with the calculated plaque-forming units of DENV at 37 °C for 24 h. The culture supernatants were collected for measuring viral replication and protein expression.

2.5. DENV and Antigen Detection. For the plaque assay, BHK-21 cells were plated in a 12-well plate (2 × 10⁵ cells/well) and cultured in DMEM at 28°C in 5% CO₂. After adsorption with serially diluted culture supernatants for 1 h, the solution was replaced with fresh DMEM containing 2% FBS and 0.5% methylcellulose (Sigma-Aldrich). Five days postinfection, the medium was removed, and the cells were fixed and stained with a crystal violet solution containing 1% crystal violet, 0.64% NaCl, and 2% formalin. To calculate the viral titer, the formation of plaques was counted at each dilution. For the DENV nonstructural protein NS1 detection, NS1 Antigen Rapid Test Cassette obtained from AsiaGen (Tainan, Taiwan) was used according to the manufacturer’s instructions.

2.6. Wright-Giemsa Staining. Following DENV ex vivo infection for 24 and 48 h, a drop of WB approximately 3 mm in diameter on each sample was placed at one end of the slide and then spread across the width of the slide. All smears of
blood were air-dried and stained with Wright-Giemsa stain (Tonyar Biotech, Taipei, Taiwan). Cells were photographed and counted under an optical microscope (Olympus CX23; Olympus, Tokyo, Japan).

2.7. Complete Blood Counts (CBCs). A complete blood count (CBC) test was conducted on heparinized peripheral WB following DENV infection for 24 h by using a DxH 500 hematology analyzer (Beckton Coulter, Clare, Ireland).

2.8. Immune Profiling. Following DENV (MOI = 1) infection in 200 μl of WB ex vivo for 24 h, representative flow cytometric analysis was conducted using an Attune NxT Flow Cytometer (Thermo Fisher Scientific) and performed by no-wash no-lyse staining for specific cell surface markers (CD4, CD8, CD11c, CD14, CD16, CD19, CD25, CD56, CD62L, and HLA-DR) according to the manufacturer’s instructions (https://www.thermofisher.com/order/catalog/product/100022776#100022776).

2.9. Enzyme-Linked Immunosorbent Assay (ELISA). According to the manufacturer’s instructions, the concentration of human TNF-α in the plasma samples was determined using DuoSet ELISA Development System kits (R&D Systems, Minneapolis, MN). In brief, we coated microwells with capture Abs against TNF-α and then blocked the wells with 1% bovine serum albumin in phosphate-buffered saline (PBS). We added the tested serum, added the hTNF-α detection Abs, and then developed the signal with HRP-conjugated detection Abs against human IgG. The relative optical density was determined using a microplate reader set to 450 nm.

2.10. Statistical Analysis. Values are expressed as the mean ± standard deviation (SD). Groups were compared by using Student’s two-tailed unpaired t-test. These analyses were performed using GraphPad Prism 4 software (GraphPad Software, La Jolla, CA). Statistical significance was set at p < 0.05.

3. Results

3.1. DENV Causes Infection but Not Replication in the Blood Ex Vivo. To mimic a circulation phase of DENV infection with viremia as demonstrated previously [25, 26], we created a novel ex vivo model of WB infection without peripheral blood mononuclear cell (PBMC) isolation. DENV was inoculated in the WB culture ex vivo. At 24 h postinfection of the coculture system, as summarized in the experimental flowchart (Figure 1(a)), in addition to analyzing the viral infection and antigen expression, several approaches were conducted to evaluate the induction of cytopathology, immune cell number and population changes, and cytokine response. For detecting DENV replication and release, the supernatant of the WB culture was harvested, and a plaque assay was then performed in a standard BHK-21 cell system [27]. Simultaneously, a commercial NS1-based rapid test cassette was performed to measure viral protein secretion [28]. Compared with the positive control (DENV-containing supernatant), DENV (Figure 1(b)) was not detectable, but viral NS1 (Figure 1(c)) was significantly detected in all five tests (Cases 1-5). The results showed that DENV could cause infection but failed to release the virion in WB culture ex vivo.

3.2. DENV Coculture Causes Mononuclear Phagocytic Cell Vacuolization in the Blood Ex Vivo. After DENV coculture caused infection in the WB culture, we evaluated the morphological changes that occurred in leukocytes after viral incubation. In the DENV-WB coculture ex vivo, the suspected targets of DENV infection are the myeloid lineage’s immune cells, such as neutrophils and monocytes [20, 29, 30]. Following DENV (MOI = 1) coculture in 100 μl of WB ex vivo for 24 and 48 h, Wright-Giemsa staining showed histopathological changes significantly in mononuclear cells in all five tests (Figure 2(a)). There was an increase in the percentage of vacuolated cells after inoculation with DENV at 24 h (DENV, 82.53 ± 4.84%; mock, 24.06 ± 12.07%; p = 0.001) (Figure 2(b)). While DENV was able to cause infection in WB culture ex vivo, the results showed intracellular vacuolization in monocytes.

3.3. Complete Blood Count (CBC) Results Display a Decrease in Monocytes in the Blood Coculture with DENV Ex Vivo. To determine whether there was a change in the number of leukocytes, we next examined CBCs after DENV-WB coculture for 24 h. We found an increase in the number of neutrophils in the DENV group (DENV, 56.80 ± 7.37%; mock, 44.52 ± 9.26%) and a decrease in the number of monocytes in the DENV group (DENV, 0.908 ± 0.30%; mock, 4.37 ± 1.66%) (Figure 3). Monocytes are the most critical blood mononuclear phagocyte and one of the leading cell targets of DENV [31, 32]. The results showed a decrease in the number of monocytes caused by DENV ex vivo.

3.4. Immune Profiling Shows Cell Changes in the Blood Coculture with DENV Ex Vivo. In response to infection, alterations in blood cells are dependent on the viral load and duration of the disease [33]. Since the CBC showed a minor change in leukocytes, we evaluated blood cells by using an immune profiling approach. Multiparameter flow cytometric analysis was utilized to assess the subpopulations of immune cells, including those in the DENV-inoculated group (Figure 4(a)) and the mock control group (Figure 4(b)). In this analysis, DENV infection decreased the frequency of blood monocytes (CD14+), and dendritic cells (CD11c+) but increased total T (Th+Tc), Th (CD4+CD56), naïve and memory Tc (CD4+CD56), and total NK (CD56+ cells) (Figure 4(c)). The findings indicate that DENV decreases monocytes and dendritic cells in ex vivo conditions of infection.

3.5. The Generation of TNF-α Is Correlated with Vacuolization in Blood Coculture with DENV Ex Vivo. Based on our findings, the blood monocytes’ change was consistent with increasing numbers of cytoplasmatic vacuoles in blood cells ex vivo. It is hypothesized that the cause of vacuolization is proinflammatory cytokines secreted by blood cells [24]. Therefore, we measured TNF-α production by ELISA. We found abnormal production of TNF-α in the DENV-inoculated groups at 24 h (DENV, 263.30 ± 180.08 pg/ml)
postincubation (Figure 5(a)). Notably, we found a strong correlation between TNF-α production and the number of vacuolated cells ($r = 0.738; p = 0.015$) (Figure 5(b)). These results indicate that the occurrence of vacuolization is highly correlated with DENV-induced TNF-α.

3.6. TNF-α Determines Vacuolization in Blood Coculture with DENV Ex Vivo. To explore the mechanisms of induction of vacuolization by TNF-α, we added human TNF-α (hTNF-α, 100 ng/ml) to the WB culture ex vivo. After 24 h of treatment, Wright-Giemsa staining showed histopathological changes in mononuclear and polymorphonuclear cells in both tests (Figure 6(a)). After quantification, there was an increase in the percentage of vacuolated cells after cotreatment with hTNF-α for 24 h (TNF-α, 64.83 ± 13.77%; mock, 7.90 ± 2.83%; $p = 0.01$) (Figure 6(b)). To ensure that TNF-α caused vacuolization, we administered anti-TNF-α to DENV-WB coculture, and the results showed a large decrease in the number of vacuolated cells after cotreatment with anti-TNF-α (DENV, 82.53 ± 4.84%; DENV+ anti-TNF-α, 30.70 ± 15.26%; and mock, 24.06 ± 12.08%) (Figure 6(c)). These results demonstrated that the vacuolization is not caused by viral replication but is caused by DENV-induced TNF-α.

4. Discussion

In the acute febrile phase of dengue-infected patients, the viral load in circulation is associated with disease severity
Figure 3: CBC results of whole blood (WB) cells 24 h postincubation. Following DENV (MOI = 1) coculture in 100 µl of WB ex vivo for 24 h, the CBC test showed the percentages of specific cell populations as noted. The quantitative data are depicted as the mean ± SD obtained from five cases. *p < 0.05 and **p < 0.01, compared to the mock group. ns: not significant.

Figure 4: Continued.
However, the cytopathogenic effects of DENV in WB cells have not yet been clearly defined, particularly in well-known hematological conditions, such as leukopenia and thrombocytopenia, as found in dengue-infected patients. In this study, we created an ex vivo model of DENV infection in WB culture to examine the possible conditions and effects of viremia during the disease’s acute febrile phase. Following DENV infection, decreases in monocytes and dendritic cells following the induction of intracellular vacuolization in monocytic cells were identified in the WB culture with DENV incubation. Moreover, we demonstrated that DENV-induced TNF-α determines the cytopathogenic effect. Although the limited viral load and antiviral serum factors may affect the infectivity of DENV in our model, the findings of this study indicate the possible impacts of DENV infection on causing TNF-α production to induce intracellular vacuolization in phagocytes.

Dengue viremia is defined as the presence of DENV that can be detected in peripheral blood, including plasma and blood cells. In contrast to previous works, DENV could infect isolated human PBMCs in vitro and in vivo. However, in our ex vivo model of infection in the human WB culture, no further newly assembled DENV could be detected, suggesting a limited microenvironment for virion release, probably due to antiviral immunity induction accompanied by a decrease in host factors to support the viral life cycle. Therefore, direct contact with host cells and its viral NS1 protein may cause TNF-α production, and then the indirect DENV-induced TNF-α may cause cytopathological changes in the targeted cells. The ex vivo model could be utilized for investigating the viral effects, including DENV and NS1, particularly at the acute febrile phase of DENV infection.

Upon the initial disease onset characterized by high fever, vomiting, and viremia, the hematological examination...
blood circulation. However, our ex vivo hematological manifestation and replacement compared with immune pro monocyte level, as demonstrated by using the CBC assay and ex vivo observation, our monocyte counts [2 white blood cell counts, lower platelet counts, and higher usually shows increased hemoglobin and hematocrit, lower white blood cell counts, lower platelet counts, and higher monocyte counts [2–4, 22]. Inconsistent with the clinical observation, our ex vivo model showed a decrease in the monocyte level, as demonstrated by using the CBC assay and immune profiling analysis, suggesting possible limitations on hematological manifestation and replacement compared with blood circulation. However, our ex vivo model identified the potential effects of DENV-induced TNF-α on cellular vacuolization in monocyctic cells, indicating these cells’ physiopathological stimulation for hematological manifestations. Accordingly, further investigations are needed to clarify the effectors involved in DENV-induced vacuolization, the possible regulatory effects caused by endocytosis, phagocytosis, autophagy, and cellular changes on cell survival and death, which are all involved in dengue pathogenesis.

This study found that DENV induces massive vacuolization of human leukocyte cells ex vivo, indicating this process is associated with DENV infection. However, the cytopathic effects of the DENV are less well known. Another flavivirus, ZIKV, also induces massive cytoplasmic vacuolization in human epithelial cells, primary skin fibroblasts, and astrocytes and then causes large-scale endoplasmic reticulum (ER) rearrangements and unfolded protein response (UPR) activation, namely, pyroptosis, a form of cell death characterized by swelling of the ER and mitochondria as well as cytoplasmic vacuolization [38]. When the UPR is inadequate to maintain the ER in a steady state, autophagy and cell death programs are activated [39]. Accumulated evidence shows that autophagy plays a vital role in controlling neutrophil and monocyte function when fighting off infections, including the processes of degranulation, metabolism, and the formation of neutrophil extracellular traps [40]. It is speculated that DENV may also cause large-scale ER rearrangements followed by the generation of cytoplasmic vacuoles, which come from the ER membrane.

The CBC assay results only identified a change in monocyte counts in the ex vivo model of DENV infection. However, by using an immune profiling approach, this study also identified a partial increase in total NK (CD56+) cells and a decrease in monocytes (CD14+) and DCs (CD11c+). While the approach of immune profiling confirmed the results of CBC on monocyte expression, changes in the other identified cell populations are needed to compare with previous works [41] while NK cells are activated [42] and DCs are decreased [43] in dengue-infected patients. A decrease in monocytes may indicate the effects of DENV-induced TNF-α for cellular activation followed by cytopathological changes, including cell adhesion and cell death. Regarding their potential roles in controlling both infections, the ex vivo model of DENV infection in peripheral blood may provide a different investigative strategy for verifying the mechanisms of viral immunity and immunopathogenesis in response to DENV infection.

Cytokine storms can be present at the acute febrile phase of DENV infection and are involved in dengue diseases’ immunopathogenesis [14, 15]. Among these immune parameters, TNF-α was shown to be positively linked to dengue-associated hematological changes in thrombocytopenia and vascular dysfunction [16–18]. Targeting TNF-α has been used as an immunotherapy strategy for suppressing DENV-induced peripheral and tissue-specific inflammatory disorders and mortality [44–46]. In this study, we found an increase in TNF-α in the ex vivo model of DENV infection, which may contribute to the induction of cellular vacuolization in monocyctic cells. Because monocytes are innate immune cells that probably produce antiviral and proinflammatory cytokines, it is speculated that vacuolization of
phagocytes is an immunological process that restricts or promotes microbial pathogenesis in infected patients.

Our study has several limitations. First, this study’s ex vivo model reflects the cytopathological effects of DENV infection in circulation, particularly at the acute phase of infection during viremia. However, dissimilar with circulation, the closed system in the ex vivo model of WB culture may affect different cell populations’ expression patterns. Notably, the induction of monocyte activation may be followed by functional changes, while the full expression of suspended monocyte numbers is significantly decreased. Monocytes are the primary cells probably targeted by DENV infection and may adhere to the disk culture. The response may increase the relative expression of neutrophils and NK cells in this experimental model system. Immune profiling in clinical blood samples of dengue patients is needed for validation. Second, in the immune profiling approach, more markers are needed to analyze the classification of specific immune cell populations. Finally, to explore the possible cytopathological effects caused by DENV and dengue NS1 protein, it is essential to investigate the infectivity of DENV in WB culture and the TNF-α-producing cells, particularly in circulating immune cell populations.

5. Conclusions

In conclusion, by using an ex vivo model of DENV-WB coculture, we found that the induction of proinflammatory TNF-α expression was followed by TNF-α-regulated cellular vacuolization in monocyctic cells. Additionally, we identified the changes in immune cell subpopulations related to the immunopathogenesis of DENV infection. Monitoring cytokine response, cellular vacuolization, and immune cell changes may help the clinical diagnosis of DENV-induced systemic inflammation. While the pathogenesis of DENV infection and disease progression is complicated, the ex vivo model may provide an experimental strategy for further exploring peripheral blood immunity against DENV infection.
Data Availability
The data used to support the findings of this study are available from the corresponding authors upon request.

Conflicts of Interest
The authors declare that there is no conflict of interest.

Authors’ Contributions
R.-D.S. and M.-K.J. performed most of the experiments and interpreted the results. T.-W.H., C.-H.H., and C.-F.L. participated in the design and supervision of the projects. M.-K.J. and T.-J.S. conducted the virus experiments. T.-J.S., P.-C.T., Y.-T.W., and Z.-Y.Y. contributed to flow cytometric analysis. R.-D.S., C.-H.H., and C.-F.L. designed the concept of the project and wrote the manuscript. All authors reviewed and approved the manuscript.

Acknowledgments
This work was supported by the Ministry of Science and Technology (MOST107-2321-B-038-001, 108-2320-B-038-026, 109-2320-B-038-050, and 109-2327-B-006-010) and Chi Mei Medical Center and Taipei Medical University (105CM-TMU-03, 106CM-TMU-09, and 107CM-TMU-10), Taiwan. We thank the Core Facility Center of Taipei Medical University (TMU) for providing technical support. Technology (MOST107-2321-B-038-001, 108-2320-B-038-026, 109-2320-B-038-050, and 109-2327-B-006-010) and 109-2327-B-006-010) and 107CM-TMU-10), Taiwan. We thank the Core Facility Center of Taipei Medical University (TMU) for providing technical support. We also appreciate the Office of Research and Publication, Universitas Gadjah Mada, Indonesia, for editing the manuscript.

References
[1] I. A. Rodenhuis-Zybert, J. Wilschut, and J. M. Smit, “Dengue virus life cycle: viral and host factors modulating infectivity,” Cellular and Molecular Life Sciences, vol. 67, no. 16, pp. 2773–2786, 2010.
[2] World Health Organization and UNICEF, Handbook for Clinical Management of Dengue, p. 114, 2012.
[3] C. M. Chen, K. S. Chan, W. L. Yu et al., “The outcomes of patients with severe dengue admitted to intensive care units,” Medicine (Baltimore), vol. 95, no. 31, article e4376, 2016.
[4] J. Chaloemwong, A. Tantiworawit, T. Rattanathammethe et al., “Useful clinical features and hematological parameters for the diagnosis of dengue infection in patients with acute febrile illness: a retrospective study,” BMC Hematology, vol. 18, no. 1, p. 20, 2018.
[5] M. Kotepui, B. PhunPhuech, N. Phiwklam, and K. Uthaisar, “Differentiating between dengue fever and malaria using hematological parameters in endemic areas of Thailand,” Infectious Diseases of Poverty, vol. 6, no. 1, p. 27, 2017.
[6] M. S. Diamond, D. Edgil, T. G. Roberts, B. Lu, and E. Harris, “Infection of human cells by dengue virus is modulated by different cell types and viral strains,” Journal of Virology, vol. 74, no. 17, pp. 7814–7823, 2000.
[7] D. M. Morens, M. C. Chu, N. J. Marchette, and S. B. Halstead, “Growth of dengue type 2 virus isolates in human peripheral blood leukocytes correlates with severe and mild dengue disease,” The American Journal of Tropical Medicine and Hygiene, vol. 45, no. 5, pp. 644–651, 1991.
[8] P. Sun, K. Bauza, S. Pal et al., “Infection and activation of human peripheral blood monocytes by dengue viruses through the mechanism of antibody-dependent enhancement,” Virology, vol. 421, no. 2, pp. 245–252, 2011.
[9] X. Fang, Z. Hu, W. Shang, J. Zhu, C. Xu, and X. Rao, “Genetic polymorphisms of molecules involved in host immune response to dengue virus infection,” FEBS Immunology and Medical Microbiology, vol. 66, no. 2, pp. 134–146, 2012.
[10] R. Ben-Shachar, S. Schmidller, and K. Koelle, “Drivers of individual variation in dengue viral load dynamics,” PLoS Computational Biology, vol. 12, no. 11, article e1005194, 2016.
[11] C. Zou, C. Huang, J. Zhang et al., “Virulence difference of five type I dengue viruses and the intrinsic molecular mechanism,” PLoS Neglected Tropical Diseases, vol. 13, no. 3, article e0007202, 2019.
[12] N. Uno and T. M. Ross, “Dengue virus and the host innate immune response,” Emerging Microbes & Infections, vol. 7, no. 1, p. 167, 2018.
[13] J. Flipse, M. A. Diosa-Toro, T. E. Hoornweg, D. P. I. van de Pol, S. Urcuqui-Inchima, and J. M. Smit, “Antibody-dependent enhancement of dengue virus infection in primary human macrophages; balancing higher fusion against antiviral responses,” Scientific Reports, vol. 6, no. 1, p. 29201, 2016.
[14] A. R. K. Patro, S. Mohanty, B. K. Prusty et al., “Cytokine signature associated with disease severity in dengue,” Viruses, vol. 11, no. 1, p. 34, 2019.
[15] A. L. Rothman, “Immunity to dengue virus: a tale of original antigenic sin and tropical cytokine storms,” Nature Reviews. Immunology, vol. 11, no. 8, pp. 532–543, 2011.
[16] S. Inyoo, A. Sutthitepumrong, and S. N. Pattanakitsakul, “Synergistic effect of TNF-α and dengue virus infection on adhesion molecule reorganization in human endothelial cells,” Japanese Journal of Infectious Diseases, vol. 70, no. 2, pp. 186–191, 2017.
[17] K. I. Masood, B. Jamil, M. Islam, M. Farhan, and Z. Hasan, “Role of TNF α, IL-6 and CXCL10 in dengue disease severity,” Iranian Journal of Microbiology, vol. 10, no. 3, pp. 202–207, 2018.
[18] A. A. Meena, A. Murugesan, S. Sopnajothi et al., “Increase of plasma TNF-α is associated with decreased levels of blood platelets in clinical dengue infection,” Viral Immunology, vol. 33, no. 1, pp. 54–60, 2020.
[19] E. D. Hottz, I. M. Medeiros-de-Moraes, A. Vieira-de-Abreu et al., “Platelet activation and apoptosis modulate monocyte inflammatory responses in dengue,” Journal of Immunology, vol. 193, no. 4, pp. 1864–1872, 2014.
[20] A. Opasawatchai, P. Amornsupawat, N. Jiravecjakul et al., “Neutrophil activation and early features of NET formation are associated with dengue virus infection in human,” Frontiers in Immunology, vol. 9, p. 3007, 2019.
[21] M. Juffrie, G. M. van der Meer, C. E. Hack et al., “Inflammatory mediators in dengue virus infection in children: interleukin-8 and its relationship to neutrophil degranulation,” Infection and Immunity, vol. 68, no. 2, pp. 702–707, 2000.
[22] J. G. X. Wong, T. L. Theirin, D. C. Lye, Y. Hao, A. Wilder-Smith, and Y. S. Leo, “Severe neutropenia in dengue patients: prevalence and significance,” The American Journal of Tropical Medicine and Hygiene, vol. 90, no. 6, pp. 984–987, 2014.
[23] M. Kunder, V. Lakshmiah, and A. V. Moideen Kutty, “Plasma neutrophil elastase, α1-antitrypsin, α2-macroglobulin and
neutrophil elastase–α1-antitrypsin complex levels in patients with dengue fever,” *Indian Journal of Clinical Biochemistry*, vol. 33, no. 2, pp. 218–221, 2018.

[24] C. C. Mihalache, S. Yousefi, S. Conus, P. M. Villiger, E. M. Schneider, and H. U. Simon, “Inflammation-associated autophagy-related programmed necrotic death of human neutrophils characterized by organelle fusion events,” *Journal of Immunology*, vol. 186, no. 11, pp. 6532–6542, 2011.

[25] S. Thiemmeca, A. Songjiang, N. Punyadee, K. Kongmanas, J. Atkinson, and P. Avirutnan, “Infection of whole blood with dengue virus,” *Molecular Immunology*, vol. 102, pp. 132-133, 2018.

[26] A. P. Durbin, M. J. Vargas, K. Wanionek et al., “Phenotyping of peripheral blood mononuclear cells during acute dengue illness demonstrates infection and increased activation of monocytes in severe cases compared to classic dengue fever,” *Virology*, vol. 376, no. 2, pp. 429–435, 2008.

[27] T. T. Tsai, C. L. Chen, Y. S. Lin et al., “Microglia retard dengue virus-induced acute viral encephalitis,” *Scientific Reports*, vol. 6, no. 1, p. 27670, 2016.

[28] L. Osorio, M. Ramirez, A. Boneo, L. A. Villar, and B. Parra, “Comparison of the diagnostic accuracy of commercial NS1-based diagnostic tests for early dengue infection,” *Virology Journal*, vol. 7, no. 1, p. 361, 2010.

[29] Z. Kou, M. Quinn, H. Chen et al., “Monocytes, but not T or B cells, are the principal target cells for dengue virus (DV) infection among human peripheral blood mononuclear cells,” *Journal of Medical Virology*, vol. 80, no. 1, pp. 134–146, 2008.

[30] S. Noisakran, N. Onlamoon, P. Songprakhon, H. M. Hsiao, K. Cheokphaibulkit, and G. C. Perng, “Cells in dengue virus infection in vivo,” *Advances in Virology*, vol. 2010, Article ID 164878, 15 pages, 2010.

[31] J. A. Aguilar-Briseño, V. Upasani, B. M., Ellen et al., “TLR2 on blood monocytes senses dengue virus infection and its expression correlates with disease pathogenesis,” *Nature Communications*, vol. 11, no. 1, p. 3177, 2020.

[32] S. W. Wan, B. A. Wu-Hsieh, Y. S. Lin, W. Y. Chen, Y. Huang, and R. Anderson, “The monocyte-macrophage-mast cell axis in dengue pathogenesis,” *Journal of Biomedical Science*, vol. 25, no. 1, p. 77, 2018.

[33] M. Kwissa, H. I. Nakaya, N. Onlamoon et al., “Dengue virus infection induces expansion of a CD14+ CD16+ monocyte population that stimulates plasmablast differentiation,” *Cell Host & Microbe*, vol. 16, no. 1, pp. 115–127, 2014.

[34] H. K. Jayanthi and S. K. Tulas, “Correlation study between platelet count, leucocyte count, nonhemorrhagic complications, and duration of hospital stay in dengue fever with thrombocytopenia,” *Journal of Family Medicine and Primary Care*, vol. 5, no. 1, pp. 120–123, 2016.

[35] G. Añez, D. A. R. Heisey, C. Chancey et al., “Distribution of dengue virus types 1 and 4 in blood components from infected blood donors from Puerto Rico,” *PLoS Neglected Tropical Diseases*, vol. 10, no. 2, article e0004455, 2016.

[36] L. Jiang and Q. Sun, “The expression profile of human peripheral blood mononuclear cell miRNA is altered by antibody-dependent enhancement of infection with dengue virus serotype 3,” *Virology Journal*, vol. 15, no. 1, p. 50, 2018.

[37] Y. L. Chen, N. Abdul Ghafar, R. Karuna et al., “Activation of peripheral blood mononuclear cells by dengue virus infection depotentiates balapiravir,” *Journal of Virology*, vol. 88, no. 3, pp. 1740–1747, 2014.