Tandem Duplication and Random Loss for mitogenome rearrangement in *Symphurus* (Teleost: Pleuronectiformes)

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Abstract

**Background:** The mitochondrial genomes (mitogenomes) of flatfishes (Pleuronectiformes) exhibit highly diversified types of large-scale gene rearrangements. We have reported that the mitogenomes of *Crossorhombus azureus* (Bothidae), *Samariscus latus* (Samaridae) and *Cynoglossus* fishes (Cynoglossidae) show different types of gene rearrangements.

**Results:** In the present study, the complete mitogenomes of two *Symphurus* species (Cynoglossidae), *Symphurus plagiusa* and *Symphurus orientalis*, were determined. The gene order in the *S. plagiusa* mitogenome is the same as that of a typical vertebrate (without any gene rearrangements). Surprisingly, large-scale gene rearrangements have occurred in *S. orientalis*. In the rearranged fragment from the control region (CR) to the WANCY tRNA cluster (tRNA cluster of tRNA-W, tRNA-A, tRNA-N, tRNA-C and tRNA-Y) in the *S. orientalis* mitogenome, tRNA-V and tRNA-M have been translocated to the 3’ end of the 16S rRNA gene, with six large intergenic spacers over 20 bp in length. In addition, an origin for light-strand replication (O_L) structure that is typically located in the WANCY region was absent in both the *S. plagiusa* and *S. orientalis* mitogenomes. It is generally recognized that a sequence in the WANCY region that encodes tRNAs forms a hairpin structure (O_L-like structure) and can act as the O_L when the typical locus is lost. Moreover, an additional O_L-like structure was identified near the control region in the *S. plagiusa* mitogenome.

**Conclusions:** The positions of the intergenic spacers and the rearranged genes of the *S. orientalis* mitogenome strongly indicate that the mechanism underlying the rearrangement of this mitogenome was Tandem Duplication and Random Loss. Additionally, two O_L-like regions substituting for the typical locus were found in the *S. plagiusa* mitogenome. We speculate that the ancestral mitogenomes of *S. plagiusa* and *S. orientalis* also had this characteristic, such that if both O_L-like structures functioned during mitochondrial replication, they could initiate duplicate replications of the light strand (L-strand), leading to duplication of the region between the two structures. We consider that this mechanism may account for the gene duplication that occurred during the gene rearrangement process in the evolution of the ancestral mitogenome to the *S. orientalis* mitogenome.

**Keywords:** Flatfish, Mitogenome, Gene rearrangement, O_L-like structure, Mitochondrial replication

Background

Vertebrate mitochondrial genomes (mitogenomes) typically contain the same 37 genes [1]. The order of these genes is generally considered conservative in most vertebrate genomes; however, gene rearrangements also have been found in many taxa, such as birds [2-4], reptiles [5,6], amphibians [7,8], and fishes [9-11]. Teleosts, with the largest number of published complete mitogenome sequences, show only a few gene rearrangement events [9-13]. In most cases, a teleostean group has only one type or a set of similar gene rearrangements [9-14]. However, the flatfish (Pleuronectiformes) mitogenomes exhibit the most diversified types of large-scale gene rearrangements. In the mitogenomes of *Cynoglossus* fishes (tongue soles, Cynoglossidae), the control region is translocated, and a tRNA (transfer ribonucleic acid) gene is inverted [10]. In contrast, no gene rearrangements have been found in soles (Soleidae), the closest family to Cynoglossidae fishes [15-17]. The mitogenome of *Crossorhombus azureus* (Bothidae) contains genomic-scale

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gene rearrangements characterized by the protein-coding gene ND6 and seven tRNA genes encoded on the light strand (L-strand; H- versus L-strands are defined by studies on AT- and GC-skewing [18,19]) that are clustered together [20]. A third type of gene rearrangement was detected in the Samariscus latus (Samaridae) mitogenome [21]. Distinct from the above-mentioned flatfishes, the gene rearrangement in this species is characterized by the duplication and translocation of the control region (CR); simultaneously, the genes located between the two CRs are divided into two clusters in which their relative gene orders have been maintained [21].

Several models have been proposed to explain gene rearrangements in animal mitogenomes. The Recombination model involves the breakage and rejoining of participating DNA strands [22]. The Tandem Duplication and Random Loss (TDRL) model posits that rearrangements of mitochondrial gene order occurred via tandem duplications of certain genes followed by random deletion of some of the duplications [23,24]. Two additional hypotheses are described in the Tandem Duplication and Non-Random Loss (TDNRL) [25] and tRNA mis-priming models [26,27]. For the gene rearrangements in flatfishes, none of the models mentioned above can provide a perfect explanation. Thus, Kong et al. [10] developed a model of inverse duplication and deletion of redundant genes to explain the gene rearrangements in tongue soles. Subsequently, Shi et al. [20] proposed the Dimer-Mitogenome and Non-Random Loss model (DMNR), which inferred the course of gene rearrangements in C. azureus. Recently, for the rearrangement events in the S. latus mitogenome, Shi et al. [21] proposed the Double Replications and Random Loss model.

Of these mechanisms proposed to explain mitochondrial gene rearrangements, the TDRL model is generally considered the most popular and important in vertebrates [9,23,24,28,29]. Generally, when decrypting gene rearrangements with the TDRL model, it is always necessary to propose multiple duplication and loss steps. It is therefore difficult to trace which steps preserved the functional genes and which DNA segments degenerated to pseudogenes or intergenic spacers. In other words, large-scale gene rearrangements cannot readily yield integrated evidence for the TDRL model [9,23,24,28,29]. San Mauro et al. [29] also indicated that the evidence for this model in the form of duplicated genes that either remain functional or have become pseudogenes in the process of being eliminated is rather limited.

In the present study, the complete mitogenomes of two flatfishes, Symphurus plagiusa and Symphurus orientalis, were sequenced. Surprisingly, the gene order of the S. plagiusa mitogenome resembles that of a typical vertebrate (un-rearranged gene order), whereas that of S. orientalis shows large-scale gene rearrangements. This is the first report of mitogenomes with a typical gene order and large-scale gene rearrangements within the same teleost genus. The characteristics of the gene order and intergenic spacers in S. orientalis provide clear evidence for the TDRL model, accounting for the gene rearrangements in the S. orientalis mitogenome.

Methods

Ethics statement

Ethical approval was not required for the present study because the examined specimens were commonly captured marine economic fishes, and all of the fish specimens were already dead when we obtained them and were sourced from commercial fisheries. Additionally, these species were not included in the IUCN list of endangered species (http://www.iucnredlist.org).

Sampling, DNA extraction, PCR and sequencing

Specimens of S. plagiusa and S. orientalis were collected from Tampa Bay, Florida (USA) and Taiwan (China), respectively. A portion of the epaxial musculature was excised from fresh specimens and immediately stored at −70°C. Total genomic DNA was extracted using the SQ Tissue DNA Kit (OMEGA) following the manufacturer’s protocol. Based on alignments and comparisons of complete mitochondrial sequences from flatfishes, dozens of primer pairs were designed for amplification of the mtDNA genomes (Additional file 1: Table S1 and Additional file 2: Table S2). More than 30 bp of overlapping fragments between tandem regions were used to ensure correct assembly and integrity of the complete sequence.

PCR (polymerase chain reaction) was performed in a 25 μl reaction volume containing 2.0 mM MgCl₂, 0.4 mM of each dNTP, 0.5 μM of each primer, 1.0 U Taq polymerase (Takara, China), 2.5 μl of 10× Taq buffer, and approximately 50 ng of DNA template. PCR cycling conditions included an initial denaturation at 95°C for 3 min, followed by 30–35 cycles of denaturation at 94°C for 45 s, annealing at 45–55°C for 45 s, and elongation at 68–72°C for 1.5–5 min. The PCR reactions were completed by a final extension at 72°C for 5 min. The PCR products were purified with the Takara Agarose Gel DNA Purification Kit (Takara, China) and used directly as templates for cycle sequencing reactions. Sequence-specific primers were further designed and used as walking primers for both strands of each fragment on an ABI 3730 DNA sequencer (Applied Biosystems, USA). The mtDNA sequences of S. plagiusa and S. orientalis have been submitted to GenBank under the accession numbers JQ639061 and KP992899, respectively.

Sequence analysis

Sequenced fragments were assembled to create complete mitochondrial genomes using CodonCode Aligner v3 and BioEdit v7 [30]. During the processing of
large fragments and walking sequences, regular manual examinations were performed to ensure reliable assembly of the genome sequence. Annotation and boundary determination of protein-coding and tRNA (Ribosomal ribonucleic acid) genes were performed using NCBI-BLAST (http://blast.ncbi.nlm.nih.gov). tRNA genes and their secondary structures were identified using tRNAscan-SE 1.21 [31], setting the cut-off values to 1 when necessary.

Results

Features of the genomes

The complete mitogenomes of S. plagiusa and S. orientalis were 17040 bp and 17498 bp in length, respectively, and contained 13 protein-coding genes, 22 tRNA genes, and 2 rRNA genes as well as one CR. Most of these genes were encoded by the heavy strand (H-strand), except ND6 and eight tRNA genes, which were encoded on the L-strand (Table 1). The 22 tRNA genes were interspersed among rRNAs and protein-coding genes, and all tRNAs can be folded into typical cloverleaf structures. The location of the CR was between the tRNA-P and tRNA-F genes, as is typical for teleosts. Compared with the CR sequences of the other flatfishes, the symbolic structures of the two Symphurus CRs were present as in other bony fishes [16,20,21,32,33]. The typical origin for L-strand replication (Oₜ), which is usually located inside the WANCY cluster (tRNA cluster of tRNA-W, tRNA-A, tRNA-N, tRNA-C and tRNA-Y), was not found in either of the two Symphurus mitogenomes at this location.

Two different mitochondrial gene orders in the genus Symphurus

The gene order of the S. plagiusa mitogeneome is the same as that of a typical vertebrate, while that of S. orientalis contains large-scale gene rearrangements. This phenomenon is rare in vertebrates and the first report in teleosts. The difference between the two mitogenomes lies between the CR and the WANCY region. The gene order of this region in the S. plagiusa mitogenome is CR-F-12S-V-16S-L1-ND1-I-Q-M-ND2-WANCY, which is the same as that of a typical vertebrate, while this region has been rearranged to CR-F-12S-16S-M-V-L1-ND1-I-Q-ND2-WANCY in the S. orientalis mitogenome.

In the rearranged fragment spanning the region from the CR to the WANCY region in the S. orientalis mitogenome, six large intergenic spacers greater than 20 bp in length remain: Gap A, between 12S and 16S (65 bp); Gap B, between 16S and tRNA-M (98 bp); Gap C, between tRNA-M and tRNA-V (100 bp); Gap D, between tRNA-V and tRNA-L1 (31 bp); Gap E, between tRNA-Q and ND2 (117 bp); and Gap F, between ND2 and tRNA-W (37 bp).

Discussion

Which mechanisms account for the gene rearrangements in S. orientalis mitogenome?

To date, approximately 1,500 complete mtDNA sequences have been determined in teleosts, and several types of gene rearrangements have been reported [17,33-36]. In the taxa showing gene rearrangements, species across the entire taxon typically show the same or similar rearrangement events [4,6,14]. That is, at the intra-family or intra-genus level, few groups include species with the typical (unchanged) gene order together with species with large-scale gene rearrangements. Nevertheless, this unique phenomenon occurs in the flatfish genus Symphurus, as described above.

Of the models that have been proposed to explain gene rearrangements in animal mitogenomes, which model most likely applies to the S. orientalis mitogenome? The recombination model is only suitable for block interchanges of small fragments, and this model is quite rare in the mitochondrial genome. As for the tRNA mis-priming [26,27] and TDNL models [25], there are no obvious corresponding model rules in the S. orientalis mitogenome.

Several species within three flatfish groups (Bothidae, Samaridae and Cynoglossidae) have been reported to possess different types of gene rearrangements. Among them, Cynoglossus fishes belong to Cynoglossidae, the same family as Symphurus. However, the rearrangements in the Cynoglossus mitogenomes are characterized by inverted tRNA genes, which were not present in the S. orientalis mitogenome. The rearrangement characteristics of the other flatfish groups also differ from those of S. orientalis.

We numbered the gene order of the S. plagiusa mitogenome (typical gene order) from CR to WANCY in the following series: CR–F (1)–12S (2)–V (3)–16S (4)–L1 (5)–ND1 (6)–I (7)–Q (8)–M (9)–ND2 (10)–W (11)–A (12)–N (13) CY. Following this scheme, the corresponding sequence of S. orientalis would be CR–F (1)–12S (2)–16S (4)–M (9)–V (3)–L1 (5)–ND1 (6)–I(7)–Q (8)–ND2 (10)–W (11)–A (12)–N (13) CY. Based on this numeric order, the genes from F to N (1–13) in S. orientalis can be divided into two gene clusters: 1–2–(4–9) and (3–5–6–7–8)–10–11–12–13, each of which retains the conserved relative gene order, from low to high. It is reasonable to assume that the two clusters were derived from a tandem-duplicated DNA fragment that spanned genes from at least V (3) to M (9) in the typical gene order. Thereafter, one of each pair of duplicated genes was randomly lost: 1–2–(3–4–5–6–7–8–9) (3–4–5–6–7–8–9)–10–11–12–13. Within the scope of our current knowledge, this rearrangement process represents the most parsimonious and reasonable hypothesis.
### Table 1 Features of the mitogenomes of Symphurus plagiusa (Left) and Symphurus orientalis (Right)

| Gene          | Position From | To | Length (bp) | Intergenic region* | Strand | Gene          | Position From | To | Length (bp) | Intergenic region | Strand |
|---------------|---------------|----|-------------|--------------------|--------|---------------|---------------|----|-------------|--------------------|--------|
| tRNA-Phe (F)  | 1             | 69 | 69          | 0                  | H      | tRNA-Phe      | 1             | 71 | 71          | 0                  | H      |
| 12S           | 70            | 1016 | 947        | 0                  | H      | 12S           | 72            | 1020 | 949        | 65                 | H      |
| tRNA-Val (V)  | 1017          | 1086 | 70          | 0                  | H      | 16S           | 1086          | 2796 | 70          | 100                | H      |
| 16S           | 1087          | 2793 | 1707        | 0                  | H      | tRNA-Met      | 2895          | 2964 | 70          | 100                | H      |
| tRNA-Leu<sup>LCA</sup> (L) | 2794          | 2866 | 73          | 0                  | H      | tRNA-Val      | 3065          | 3136 | 72          | 31                 | H      |
| ND1           | 2867          | 3841 | 975         | 2                  | H      | tRNA-Leu<sup>LCA</sup> | 3168 | 3240 | 73          | 0                  | H      |
| tRNA-Ile (I)  | 3844          | 3913 | 70          | -2                 | L      | ND1           | 3241          | 4212 | 972        | 7                  | H      |
| tRNA-Gln (Q)  | 3912          | 3982 | 71          | -1                 | L      | tRNA-Ile      | 4220          | 4289 | 70          | -2                 | H      |
| tRNA-Met (M)  | 3982          | 4050 | 69          | 124                | H      | tRNA-Gln      | 4288          | 4358 | 71          | 117                | L      |
| ND2           | 4175          | 5227 | 1053        | 42                 | H      | ND2           | 4476          | 5522 | 1047       | 37                 | H      |
| tRNA-Trp (W)  | 5270          | 5339 | 70          | 0                  | H      | tRNA-Trp      | 5560          | 5628 | 69          | 0                  | H      |
| tRNA-Ala (A)  | 5340          | 5408 | 69          | 0                  | H      | tRNA-Ala      | 5629          | 5697 | 69          | 0                  | L      |
| tRNA-Asn (N)  | 5409          | 5479 | 71          | 2                  | L      | tRNA-Asn      | 5698          | 5770 | 73          | 3                  | L      |
| tRNA-Cys (C)  | 5482          | 5546 | 65          | 3                  | L      | tRNA-Cys      | 5774          | 5839 | 66          | 4                  | L      |
| tRNA-Tyr (Y)  | 5550          | 5616 | 67          | 1                  | L      | tRNA-Tyr      | 5844          | 5910 | 67          | 10                 | L      |
| COI           | 5618          | 7168 | 1551        | 19                 | L      | COI           | 5921          | 7471 | 1551       | 0                  | H      |
| tRNA-Ser<sup>LCA</sup> (S1) | 7169          | 7239 | 71          | 19                 | L      | tRNA-Ser<sup>LCA</sup> | 7472 | 7542 | 71          | 19                 | L      |
| tRNA-Asp (D)  | 7259          | 7327 | 69          | 4                  | L      | tRNA-Asp      | 7562          | 7630 | 69          | 5                  | H      |
| COII          | 7332          | 7925 | 594         | 96                 | H      | COII          | 7636          | 8326 | 691        | 0                  | H      |
| tRNA-Lys (K)  | 8022          | 8097 | 76          | 1                  | H      | tRNA-Lys      | 8327          | 8400 | 74          | 2                  | H      |
| ATP8          | 8099          | 8266 | 168         | -10                | H      | ATP8          | 8403          | 8570 | 168        | -10                | H      |
| ATP6          | 8257          | 8940 | 684         | 2                  | H      | ATP6          | 8561          | 9244 | 684        | -1                 | H      |
| COIII         | 8943          | 9731 | 789         | 65                 | H      | COIII         | 9244          | 10029 | 786      | -1                 | H      |
| tRNA-Gly (G)  | 9797          | 9864 | 68          | 0                  | H      | tRNA-Gly      | 10029         | 10096 | 68        | 0                  | H      |
| ND1           | 9865          | 10215 | 351       | -2                 | H      | ND1           | 10097         | 10447 | 351      | -2                 | H      |
| tRNA-Arg (F)  | 10214         | 10282 | 69        | 0                  | H      | tRNA-Arg      | 10446         | 10514 | 69        | 0                  | H      |
| ND4           | 10523         | 11953 | 1381     | 0                  | H      | ND4           | 10805         | 12184 | 1380     | 1                  | H      |
| tRNA-His (H)  | 11954         | 12022 | 69        | 0                  | H      | tRNA-His      | 12186         | 12253 | 68        | 0                  | H      |
| tRNA-Ser<sup>LSC</sup> (S2) | 12023         | 12090 | 68        | 2                  | H      | tRNA-Ser<sup>LSC</sup> | 12254         | 12321 | 68        | 2                  | H      |
| tRNA-Leu<sup>LCA</sup> (L2) | 12093         | 12164 | 72        | 0                  | H      | tRNA-Leu<sup>LCA</sup> | 12324         | 12397 | 74        | 0                  | H      |
| ND5           | 12165         | 13985 | 1821     | -4                 | H      | ND5           | 12398         | 14233 | 1836     | 4                  | H      |
| ND6           | 13982         | 14500 | 519       | 0                  | L      | ND6           | 14238         | 14756 | 519       | 0                  | L      |
| tRNA-Glu (E)  | 14501         | 14569 | 69        | 2                  | L      | tRNA-Glu      | 14757         | 14825 | 69        | 3                  | L      |
| Cytb          | 14572         | 15711 | 1140     | 1                  | H      | Cytb          | 14829         | 15969 | 1141     | 0                  | H      |
| tRNA-Thr (T)  | 15713         | 15781 | 69        | -1                 | H      | tRNA-Thr      | 15970         | 16042 | 73        | -1                 | H      |
| tRNA-Pro (P)  | 15781         | 15849 | 69        | 0                  | L      | tRNA-Pro      | 16042         | 16110 | 69        | 0                  | L      |
| O<sub>1</sub>-like Seq. | 16983         | 17034 | 52        / | L      | D-loop         | 16111         | 17498 | 1388     | 0                  | H      |
| D-loop        | 15850         | 17040 | 1191     | 0                  | H      |               |               |      |            |                   |        |

*Intergenic region: non-coding bases between the feature on the same line and the line below, with a negative number indicating an overlap.

The process of gene duplication

How did the duplication occur? Moreover, was the fragment from V (3) to M (9) or a longer fragment duplicated?

Many studies have reported similar duplications. For example, Fujita et al. [37] found that several lineages of parthenogenetic lizards harbor large, tandem duplications.
which are hypothesized to represent intermediate stages in gene rearrangement. These authors suggested that the slipped-strand mispairing mechanism could have been responsible for generating the duplications in the mitogenomes of these lizards.

A special phenomenon observed in the *Symphurus* mitogenomes indicates that the underlying mechanism is different from slipped-strand mispairing and can help us to explain this duplication process. Generally, there are only two stable non-coding regions in vertebrate mitogenomes: the CR and an approximately 40 nucleotide-long segment containing the origin for L-strand replication (O₁), which is usually located inside the WANCY cluster at approximately two thirds of the genomic distance away from the CR. At O₁, the parental H-strand is displaced as a single strand by the nascent H-strand and adopts a stable hairpin structure (Figure 1A) that serves as the initiation site for L-strand DNA synthesis. We searched all of the complete mitogenome sequences of flatfishes in the GenBank database and found that most contained conserved O₁ sequences, including *Cynoglossus* fishes (Figure 1A), which are in the same family as the *Symphurus* species and also show gene rearrangements. However, neither of the two *Symphurus* fish mitogenomes contains a typical O₁ region in the WANCY cluster.

O₁ sequence loss has been reported in some other vertebrate mitogenomes, and it was suggested that a sequence in the WANCY region that encodes a tRNA forms a hairpin structure (O₁-like structure) and acts as the O₁ [38-40]. Seligmann and Labra [41] tested whether a natural absence of an O₁ is associated with a greater capacity for the formation of O₁-like structures by WANCY tRNA genes in lepidosaurian taxa. These authors concluded that WANCY tRNA genes form more O₁-like structures in the absence of a regular O₁ than in its presence.

More interestingly, in the *S. plagiusa* mitogenome, within the CR and very close to tRNA-F, we found a hairpin structure that is very similar to the typical O₁ of vertebrate mitogenomes (Table 1 and Figure 1B). This finding means that while no typical O₁ region is present in the *S. plagiusa* mitogenome, there are two regions with the potential ability to form O₁-like structures and initiate the replication of the L-strand. The existence of multiple O₁s in vertebrate mitochondria has been demonstrated in many studies [42-44]. These features of multiple replication origins most likely also appeared in the mitogenome of the common ancestor of *S. plagiusa* and *S. orientalis* and were inherited by *S. plagiusa* due to its stable mitogenome structure. In contrast, these features would have been lost in the *S. orientalis* mitogenome because of the large-scale gene rearrangements. Therefore, in the ancestral mitogenome with the aforementioned dual O₁-like structures, if only one is functioning, mitochondrial replication would occur normally, but if both structures functioned at the same time, the region between the two structures could be duplicated.

Accordingly, we speculate that the two ends of the duplicated fragment fall in the WANCY region and at the 3’ end of the CR, which both contain O₁-like sequences. The duplication process would occur when both O₁-like structures are functional. During a mitochondrial replication, after the replication fork arising from the initiation of H-strand synthesis (Figure 2A) passed the WANCY region, where the typical O₁ is located in most vertebrates, the parental H-strand in this region was exposed as a single strand, and the WANCY tRNA sequence formed an O₁-like structure (O₁-like structure 1 in Figure 2B). This structure then initiated an L-strand DNA synthesis event at this site (Nascent L-strand 1 in Figure 2B). Coincidentally, when the replication fork continued to expand and passed the CR, the other O₁-like sequence was exposed and also formed a O₁-like structure (O₁-like structure 2 in Figure 2C), which then initiated a second round of L-strand DNA synthesis (Nascent L-strand 2 in Figure 2C). If both L-strand DNA synthesis events terminated at the WANCY region, the normal termination site, the fragment between the two O₁-like sequences would be synthesized twice (Figure 2D). By a circular closure event or mitochondrial repair, the 5’ end of Nascent L-strand 1 would connect to the 3’ end of Nascent L-strand 2, while the 3’ end of Nascent L-strand 1 would connect to the 5’ end of Nascent L-strand 2 (Figure 2E). In the next round of mitochondrial
replication, the duplication would be made permanent (Figure 2F).

Speculated TDRL process
The process of tandem duplication followed by random loss discussed above is typical of the TDRL model. Therefore, we applied the TDRL model to describe the rearrangement events that altered the typical gene order (as found in *S. plagiusa*) to that observed in the *S. orientalis* mitogenome. The hypothesized intermediate steps are as follows. First, the above-mentioned double O₁-like structures initiated DNA synthesis twice during mitochondrial replication, causing tandem duplication of the genes located between the CR and the WANCY region (1–13) (Figure 3A, B) in the ancestral mitogenome (Figure 3A: typical gene order). In this case, the mitogenome would then have contained two sets of the same gene cluster (Figure 3B: 1–13 and 1’–13’). Because the mitogenome only maintains one set of functional genes, during subsequent evolutionary events, one of each of the 13 duplicated gene pairs randomly lost its function and became a pseudogene (Figure 3B; gray boxes). These pseudogenes then accumulated additional mutations to become shorter non-coding sequences or even be lost from the genome. Eventually, the existing gene order of the *S. orientalis* mitogenome was established (Figure 3C).

Intergenic spacers provide evidence supporting the model
In the speculated process, after the five duplicated genes or gene clusters lost their functions, as shown in Figure 3B, they would have degraded to form five successive pseudogene fragments or shorter intergenic spacers (Figure 3B, gray boxes). In general, intergenic spacers vanish quickly because the degradation rate of non-functional genes is high to maintain the parsimony of mitogenomes. Interestingly, the speculated intergenic spacers did not disappear, and the positions of the five degraded gene fragments occur in one-to-one correspondence with those of the five residual intergenic spacers in the *S. orientalis* mitogenome: loss A in Figure 3B between 12S and 16S, which corresponds to Gap A in Figure 3C; loss B between 16S and

Figure 2 A duplication between the CR and the \( N \) gene caused by two O₁-like structures. (A) H-strand synthesis is initiated at an origin for light-strand replication (O₁). (B) L-strand synthesis is initiated at O₁-like structure 1 when the replication fork reaches approximately two thirds of the genomic distance from the CR. (C) H-strand synthesis is terminated, and O₁-like structure 2 initiates a second round of L-strand synthesis. (D) Both L-strand synthesis events are terminated at the WANCY region. (E) Connection of the 5’ end of Nascent L-strand 1 to the 3’ end of Nascent L-strand 2 and of the 3’ end of Nascent L-strand 1 to the 5’ end of Nascent L-strand 2. (F) The genes between the CR and \( N \) are, thus, duplicated.
tRNA-M, which corresponds to Gap B; loss C between tRNA-M and tRNA-V, which corresponds to Gap C; loss D between tRNA-V and tRNA-L1, which corresponds to Gap D; and loss E between tRNA-Q and ND2, which corresponds to Gap E. The one-to-one correspondence between the loss-of-function fragments and the residual intergenic spacers offers strong evidence supporting the speculated steps of gene duplication and loss in our model (Figure 3).

Conclusions
In summary, we determined the complete mitochondrial genomes of two Symphurus fishes, S. plagiusa and S. orientalis. The gene order of the S. plagiusa mitogenome is the same as that of a typical vertebrate, while that of S. orientalis features large-scale gene rearrangements. In the rearranged fragment from the CR to WANCY in the S. orientalis mitogenome, six large intergenic spacers more than 20 bp in length remain. The positions of these intergenic spacers occur at a one-to-one correspondence with the loss-of-function fragments in our speculated gene rearrangement model based on TDRL, providing strong evidence for both our model and the TDRL.

The two Symphurus fish mitogenomes share another special characteristic: they lack the typical O2 region that is conserved in most vertebrates. In addition, in the S. plagiusa mitogenome, there are two regions with the potential ability to form O2-like structures, which means that both could initiate replication of the L-strand. Accordingly, we speculate that the ancestral mitogenome also possessed these two O2-like structures and that during an ancient mitochondrial replication event, both O2-like structures initiated DNA synthesis and induced the doubled replication of the L-strand, leading to the duplication of the region between the two structures. The findings of this study are based only on the primary sequence structures of mitogenomes. To substantiate the speculations presented herein, it will be necessary to observe replication intermediates via electron microscopy or 2D-gel analysis.

Data accessibility
DNA sequences: GenBank accessions JQ639061 and KP992899.

Additional files

Additional file 1: Table S1. The primers used for fragment amplification of the Symphurus plagiusa mitogenome.

Additional file 2: Table S2. The primers used for fragment amplification of the Symphurus orientalis mitogenome.

Abbreviations
CR: Control region; WANCY: tRNA cluster of tRNA-W, tRNA-A, tRNA-N, tRNA-C and tRNA-Y; rRNA: Ribosomal ribonucleic acid; O1: Origin for light-strand replication; tRNA: Transfer ribonucleic acid; Mitogenome: mitochondrial genome; TDRL: Tandem Duplication and Random Loss; TDNL: Tandem Duplication and
Non-random Loss; DMNR: Dermer-Mitogenome and Non-Random Loss; mtDNA: Mitochondrial DNA; PCR: Polymerase chain reaction; H-strand: Heavy strand; L-strand: Light strand.

Competing interests
The authors declare that they have no competing interests.

Authors’ contributions
WS collected the datasets, carried out a portion of the experiments, and drafted the manuscript. LG, SYW and XGM carried out a portion of the experiments. XXY directed the entire research project. All authors read and approved the final manuscript. All authors read and approved the final manuscript.

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