Dual Role of Superoxide Dismutase 2 Induced in Activated Microglia

OXIDATIVE STRESS TOLERANCE AND CONVERGENCE OF INFLAMMATORY RESPONSES*

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Microglia are activated quickly in response to external pathogens or cell debris and clear these substances via the inflammatory response. However, excessive activation of microglia can be harmful to host cells due to the increased production of reactive oxygen species and proinflammatory cytokines. Superoxide dismutase 2 (SOD2) is reportedly induced under various inflammatory conditions in the central nervous system. We herein demonstrated that activated microglia strongly express SOD2 and examined the role of SOD2, focusing on regulation of the microglial activity and the susceptibility of microglia to oxidative stress. When rat primary microglia were treated with LPS, poly(I:C), peptidoglycan, or CpG oligodeoxynucleotide, respectively, the mRNA and protein levels of SOD2 largely increased. However, an increased expression of SOD2 was not detected in the primary neurons or astrocytes, indicating that SOD2 is specifically induced in microglia under inflammatory conditions. The activated microglia showed high tolerance to oxidative stress, whereas SOD2 knockdown conferred vulnerability to oxidative stress. Interestingly, the production of proinflammatory cytokines was increased in the activated microglia treated with SOD2 siRNA compared with that observed in the control siRNA-treated cells. Pretreatment with NADPH oxidase inhibitors, diphenylene iodonium and apocynin, decreased in not only reactive oxygen species generation but also the proinflammatory cytokine expression. Notably, SOD2 knockdown largely potentiated the nuclear factor κB activity in the activated microglia. Taken together, increased SOD2 conferred tolerance to oxidative stress in the microglia and decreased proinflammatory cytokine production by attenuating the nuclear factor κB activity. Therefore, SOD2 might regulate neuroinflammation by controlling the microglial activities.

Microglia constitute the primary immune cells of the central nervous system (CNS) and are activated quickly in response to external pathogens or cell debris, after which they act by releasing inflammatory factors and/or engulfing foreign bodies to mediate the inflammatory response. However, excessive activation of microglia may be harmful to host cells; for example, microglia can promote the development of various neuronal diseases by producing large amounts of inflammatory molecules, such as tumor necrosis factor-α (TNFα), interleukin-1β (IL-1β), and reactive oxygen species (ROS).2 Indeed, microglia with abnormal activity reportedly induce neuroinflammation and are implicated in the pathogenesis of Parkinson disease, Alzheimer disease, brain ischemia-reperfusion injury, trauma, epilepsy, depression, and schizophrenia (1–5). Therefore, understanding the mechanisms that control the microglial activity is critical, not only for comprehending the physiology of microglia but also for developing new therapeutic approaches to treating neuroinflammation and/or neurodegenerative diseases.

The main source of ROS in microglia is NADPH oxidase (6). NADPH oxidase is a multisubunit enzyme complex that transfers electrons to molecular oxygen from NADPH or, to a lesser extent, NADH (7), resulting in the formation of superoxide anions as the primary product. Earlier investigations have demonstrated the relevant role of NADPH oxidase in the acute inflammatory response induced by neurophilis, with evidence showing its participation in respiratory...
Regulation of Microglial Proinflammatory Responses by SOD2

bursts killing pathogens (8, 9). Mitochondria are another source of ROS inside activated microglia. Copper chloride has been shown to stimulate mitochondrial superoxide production in murine BV-2 microglial cells (10). Lipopolysaccharide (LPS), an endotoxin found in the outer membrane of Gram-negative bacteria, also elicits mitochondrial superoxide production, which is regulated by the mitochondrial chaperone glucose-regulated protein 75 (Grp75/mortalin) (11). Naik and Dixit (12) suggested that alterations in the redox environment of the plasma membrane could cause mitochondrial ROS formation. Therefore, ROS derived from NADPH oxidase are closely linked to mitochondrial ROS production.

Proinflammatory cytokines and chemokines are transcriptionally activated by several inflammatory transcription factors, including activator protein-1 (AP-1) and nuclear factor κB (NF-κB), that act downstream of pattern-recognition receptors, such as Toll-like receptors (TLRs) in microglia. Growing evidence has shown that the ROS generated within microglia can increase the expression of proinflammatory mediators, suggesting that ROS are a modulator of microglial activation. Indeed, LPS, interferon γ, and amloid β (1–42) stimulate ROS production by NADPH oxidase, especially NOX1, NOX2, and NOX4, and increases in ROS subsequently induce the expression of several proinflammatory molecules, such as TNFα, IL-1β, inducible nitric-oxide synthase, prostaglandin E2, and monocyte chemotactic protein-1 (MCP-1) (13–18). Furthermore, the antioxidant molecule α-tocopherol has been reported to decrease the levels of proinflammatory and adhesion molecules and elicit microglial ramification by scavenging ROS (19–22). Therefore, the redox balance is thought to regulate a series of neuroinflammatory processes mediated by microglia.

Superoxide dismutase (SOD) is a major antioxidant enzyme in aerobic organisms and dismutates superoxide anions to form hydrogen peroxide and molecular oxygen. SOD1 (Cu/Zn-SOD) is present in the cytoplasm, nucleus, and peroxisomes of all mammalian cells, where it scavenges superoxide. SOD2 (Mn-SOD) is localized within mitochondria and efficiently eliminates the superoxide generated from molecular oxygen in the respiratory chain. SOD2 is reportedly induced in the CNS under inflammatory conditions. Notably, the intraventricular injection of LPS has been shown to increase the SOD2 expression in the whole brains of rats (23), and treatment of mesencephalic cultures with LPS has been demonstrated to elicit SOD2 expression, probably in astrocytes (24). Moreover, there are a few reports in which mouse primary microglia or the murine microglial cell line, BV-2, showed an increased expression of SOD2 after treatment with LPS (25, 26), and inducible SOD2 is thought to attenuate oxidative injury (25, 27). However, if the redox environment regulates microglial inflammatory processes, SOD2 might also modulate the microglial activity. In this study, we demonstrated that activated microglia strongly express SOD2. We then examined the role of the SOD2 induced in microglia, focusing on the sensitivity of these cells to oxidative stress as well as regulation of the microglial activity.

Experimental Procedures

Materials—LPS from Escherichia coli 026:B6, peptidoglycan (PGN), SP600125, acetyl cytochrome c, xanthine oxidase, and diphenylene iodonium (DPI) were obtained from Sigma-Aldrich. U0126 and hydrogen peroxide were purchased from Wako Pure Chemical Industries, Ltd. (Osaka, Japan). Poly(I:C) was obtained from Tocris (Minneapolis, MN). Caffeic acid phenylethyl ester (CAPE) was purchased from Cayman Chemical (Ann Arbor, MI). Dihydroethidium (DHE) was purchased from Molecular Probes, Inc. (Eugene, OR). Apocynin was purchased from Nacalai Tesque (Kyoto, Japan). Xanthine was obtained from Merck Millipore. Single-stranded DNA molecules containing unmethylated CpG dinucleotides (CpG DNA) were synthesized by Eurofins Genomics (Huntsville, AL). WP9QY was purchased from AnaSpec, Inc. (Fremont, CA). IL-1RA was obtained from Bioworld Technology (St. Louis Park, MN). All other chemicals were obtained from Wako Pure Chemical Industries or Sigma-Aldrich and were of reagent grade.

Animals—All animal procedures were performed in accordance with the Fundamental Guidelines for Proper Conduct of Animal Experiments and Related Activities in Academic Research Institutions under the jurisdiction of the Ministry of Education, Culture, Sports, Science, and Technology (Tokyo, Japan) and the Animal Care and Use Committee of Hiroshima University (Hiroshima, Japan). Pregnant Wistar rats and male ICR mice were obtained from Kyudo (Kumamoto, Japan) and maintained in a temperature-controlled animal facility with a 12-h light/dark cycle.

Isolation and Culture of Rat Primary Microglia— Cultures of primary microglia were prepared from 1–3-day-old Wistar rats, according to our previous report (28). Briefly, the forebrain was dissociated, and the cells were plated in a poly-L-lysine-coated plastic culture flask with tissue culture medium, which consisted of Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10% fetal bovine serum (FBS) and 5 μg/ml insulin. After 9–12 days, microglia were harvested by shaking the flask and seeded at a density of $1 \times 10^5$ cells/cm$^2$. The culture medium was changed to remove non-adherent cells 30 min after seeding. The cultures of isolated microglia were uniformly immunopositive for CD11b and contained >95% microglial cells.

Total RNA Extraction and Real-time PCR— Determination of the mRNA levels was performed according to our previous report (29). The primer sequences are shown in Table 1. The amount of mRNA was normalized to that of glyceraldehyde-3-phosphate dehydrogenase (GAPDH), and the values for the treated samples were divided by those for the untreated samples to calculate the relative mRNA levels.

Immunoblotting— Microglia were collected and lysed with radioimmune precipitation assay buffer (25 mM Tris-HCl, pH 7.6, 150 mM NaCl, 1% Nonidet P-40, 1% sodium deoxycholate, and 0.1% SDS). Equal amounts of protein were loaded and separated via SDS-PAGE using 10 or 12% (w/v) polyacrylamide gels and then transferred onto polyvinylidene difluoride membranes. The blocked membranes were incubated with the following primary antibodies: anti-SOD2 (ADI-SOD-111 rabbit, Enzo Life Sciences, Farmingdale, NY) and anti-α-tubulin.
Regulation of Microglial Proinflammatory Responses by SOD2

TABLE 1
List of rat primers for real-time PCR used in this study

| Forward (5’–3’) | Reverse (5’–3’) |
|-----------------|-----------------|
| TNFα            | AGCCCTCGTGATAGCAGCTGTA | CCGGACTCCCGTTGATAGTC |
| IL-1β           | CACCATCTTCAAACGAGGAGACA | ACGGGTTCCCTGGAAGATGC |
| COX-2           | TTTTTGAGATCTCCAGAGAGATG | CACAGTGTAAGTTTCCAGGGAGAG |
| MCP-1           | TGTTCTCCAGGATGCAAGTT | CAGCCGACTTATGGAAGTC |
| MIP-2α          | CCCCCTCTGTGCTGCACACTC | CACCAACACCCCTGTACCA |
| SOD1            | TTCCAGGCAAGAACGACGATTG | TCTTCTGCTCGAACGCACCA |
| SOD2            | GCGCTGCGCAAGGAGAGATTG | ATGCCGCCCGCATGAGACTT |
| Catalase        | GCTGGTCTGGCCACACTA | AGTTGGAACACACCTGCTT |
| Gpx1            | TCCTCAGTTAGGAGAAGAAA | GCTGGAACACTTTGAGAG |
| Glutathione reductase | GTGGTTCTTCCAGCACCACCC | ATCGGAGTGAGACGCTCC |
| Thioredoxin reductase | TCCCTGCTGCACTGATCA | CGCCCGTCTGAGACAGAG |
| GAPDH           | AGGCCACCTTGCTGGACCT | CCTTGCTGCTGCTGAGACT |

(T5168 mouse, Sigma-Aldrich). Finally, the membranes were incubated in solutions of peroxide-conjugated secondary antibodies (Thermo Fisher Scientific) and then visualized using peroxide substrates (SuperSignal West Femto, Thermo Fisher Scientific). The band intensity was quantified using the ImageJ software program (National Institutes of Health, Bethesda, MD).

Culture of Rat Primary Cortical Neurons—Rat primary cortical neurons were prepared from Wistar rats on the 17th day of gestation, as reported previously with minor modifications (30). The cortex was separated, and the meninges were removed. The tissues were cut into small pieces and then dissociated using a papain dissociation system (Worthington). The purity of the neurons was assayed using microtubule-associated protein 2 (MAP2) staining, and >95% of the cultured astrocytes showed MAP2 immunoreactivity (Fig. 3D).

Culture of Rat Primary Cortical Astrocytes—Cultures of primary astrocytes were prepared from the cerebral cortex of 1–2-day-old male Wister rats (31). The cerebral hemispheres were excised, and the meninges were removed. The cerebral cortex was cut into small pieces and then dissociated using a papain dissociation system (Worthington). The resulting cell suspension was filtered through a cell strainer (40 μm, Falcon) and plated on polyethyleneimine-coated dishes at a density of 5 × 10^5 cells/ml with Neurobasal medium and B27 supplement (Gibco). After 2 days of culture, cytosine β-d-arabinofuranoside was added to inhibit glial proliferation (final concentration, 1 μM), and the medium was changed completely after 2 days to remove the cytosine β-d-arabinofuranoside. The purity of the neurons was assayed using microtubule-associated protein 2 (MAP2) staining, and >95% of cells in the culture showed MAP2 immunoreactivity (Fig. 3A).

Measurement of the ROS Levels—ROS generated inside the microglia were quantified using acetylated cytochrome c (33). Briefly, 60 μM acetylated cytochrome c was added to the culture and incubated for another 5 min in the presence of 100 μM diethyldithiocarbamate and 100 units/ml of catalase. The amount of reduced acetylated cytochrome c was determined based on the difference in absorbance between 550 and 557 nm. The total amount of superoxide anions in the culture medium was calculated using the absorption coefficient of reduced acetylated cytochrome c (ε_{550–557} = 21.0 mM⁻¹ cm⁻¹).

Measurement of Cell Viability—The degree of cell viability was determined by comparing the percentage of the lactate dehydrogenase activity in the medium and cell lysates, according to our previous report (33).

Cloning and Site-directed Mutagenesis of the Rat SOD2 Promoter Region—Rat genomic DNA was extracted from rat whole brains using the DNeasy blood and tissue kit (Qiagen, Valencia, CA). The SOD2 promoter region (1,267 bp) was amplified with Platinum Taq DNA polymerase high fidelity (Invitrogen; primers: AP-1 Se, 5’-CAGGACACAGCCACTACAGTAT-3’ (forward) and 5’-ACCGCTGCTCTCTCGTCAGAA-3’ (reverse)) and ligated into pGL4.24 to create pGL4.24-SOD2Prom, which contains a SOD2Prom-luc transcriptional fusion. Mutation of the AP-1 and NF-κB binding sites in pGL4.24-SOD2Prom was performed via site-directed mutagenesis using the QuikChange kit (Stratagene, Santa Clara, CA; primers: AP-1 Se, 5’-CAGGACACAGCCACTACAGTAT-3’ (forward) and 5’-ACCGCTGCTCTCTCGTCAGAA-3’ (reverse)); the mutants were named pGL4.24-SOD2Prom AP-1m, pGL4.24-SOD2Prom NF-κBm, and pGL4.24-SOD2Prom AP-1m NF-κBm, respectively.

 Luciferase Assay—The constructs of pGL4.24-SOD2Prom, pGL4.24-SOD2Prom AP-1m, pGL4.24-SOD2Prom NF-κBm, pGL4.24-SOD2Prom AP-1m NF-κBm, and pNL3.2.NF-κB-RE (Promega, Madison, WI) were transfected into rat primary
Regulation of Microglial Proinflammatory Responses by SOD2

microglia using Lipofectamine 2000 (Invitrogen) according to the manufacturer’s instructions. Twenty-four hours after transfection, the cells were used in the experiments. The luciferase activity was measured using the Luciferase Assay System (Promega) for the pGLO vector or the Nano-Glo Luciferase Assay System (Promega) for the pNL vector with a GloMax 20/20 Luminometer (Promega).

Evaluation of the NF-kB Binding Activity—Nuclear extracts were prepared according to our previous report with slight modifications (34). Briefly, cells were suspended in buffer A (10 mM HEPES, pH 7.8, 10 mM KCl, 2 mM MgCl$_2$, 0.1 mM EDTA, 0.5 mM DTT, and protease inhibitor mixture) and incubated on ice for 15 min. Nonidet P-40 at a final concentration of 0.6% was added to the cell suspension, which was immediately vortexed and centrifuged. The resulting white pellets were washed with buffer A, and nuclear proteins were extracted with buffer B (50 mM HEPES, pH 7.8, 50 mM KCl, 300 mM NaCl, 0.1 mM EDTA, 0.5 mM DTT, 10% (v/v) glycerol). The specific DNA binding activity of NF-kB p65 was measured using the NF-kB (p65) transcription factor assay kit (Cayman Chemical), according to the manufacturer’s instructions.

Intracerebroventricular Injection of LPS—Eight-week-old male ICR mice were initially anesthetized for surgery using a mixture of 1.5–2.0% isoflurane (160 ml/min, Wako) and oxygen. The head was skinned, and a 27-gauge injection needle was inserted into the cerebroventricular region (anterior, 0.5 mm; lateral, 1.0 mm; ventral, 5.0 mm from bregma). LPS was injected at a volume of 2 l over 2 min using a Hamilton syringe, and the needle was left in place for an additional 1 min. The mice were killed 3 h after injection. RNA was extracted from the cerebral cortex, and the cDNA was synthesized via RT-PCR, followed by real-time PCR using the specific primers listed in Table 2.

Statistical Analyses—All data are expressed as the means ± S.E. The statistical analyses were performed using a one-way analysis of variance (ANOVA), followed by Student’s t test or Dunnett’s test. The Holm or Bonferroni methods were used to correct for multiple comparisons. p values of <0.05 were considered to be statistically significant.

Results

SOD2 Up-regulation Occurs Downstream of TLRs in Microglia—When the rat primary microglia were stimulated with LPS for 6 h, the mRNA expression of proinflammatory molecules, such as TNFα, IL-1β, COX-2 (cyclooxygenase-2), MCP-1, and MIP-2α (macrophage inflammatory protein-2α), was largely increased (Fig. 1A), clearly indicating the induction of microglial activation by LPS. Interestingly, the activated microglia strongly expressed SOD2 mRNA, compared with the unstimulated cells, whereas the expression of the antioxidant enzymes, SOD1, catalase (Cat), glutathione reductase (GR), and thioredoxin reductase (TR), remained unchanged in the presence of LPS (Fig. 1B). The expression of glutathione peroxidase 1 (GPx1) slightly decreased by LPS (Fig. 1B). The time course of the changes in the SOD2 mRNA expression was almost the same as that for the proinflammatory cytokines, TNFα and IL-1β; namely, the SOD2 mRNA levels started to increase 1 h after LPS treatment, reached a peak at 6 h, and then slightly decreased (Fig. 1C). The protein levels of SOD2 also increased 1 h after LPS treatment, and a prolonged increment was observed during 24 h of incubation of the microglia with LPS (Fig. 1D).

To date, more than 10 functional TLRs have been identified in humans and rodents (35) and are expressed in a variety of cells, including microglia. We stimulated rat primary microglia with major TLR ligands other than LPS, including poly(I:C) as a ligand for TLR3, PGN as a ligand for TLR6, and synthetic CpG DNA as a ligand for TLR9. Poly(I:C), PGN, and CpG DNA increased the mRNA expression of TNFα and IL-1β in the primary microglia (Fig. 2A), indicating that the microglia were also activated by these TLR ligands. The effect of CpG DNA on microglial activation was less pronounced than that of the other TLR ligands. Remarkably, poly(I:C), PGN, and CpG DNA elicited an increased expression of SOD2 mRNA as well as SOD2 proteins (Fig. 2B). A TLR4 antagonist, TAK-242, clearly suppressed the mRNA expression of TNFα, IL-1β, and SOD2 induced by LPS (Fig. 2C), indicating that the increased expression of TNFα, IL-1β, and SOD2 was elicited downstream of TLR4. The transcriptional activation of SOD2 has been reported in response to several proinflammatory cytokines (36, 37). However, the TNF receptor antagonist WP9QY and IL-1 receptor antagonist IL-1RA did not have an effect on the SOD2 expression induced by LPS (Fig. 2D). These data indicate that the SOD2 expression is induced downstream of TLRs, which is independent of TNFα and IL-1β signaling.

We examined whether SOD2 may be induced in other cells in the CNS, such as neurons and astrocytes. In primary neurons, the expression of TNFα, IL-1β, and SOD2 was not affected by LPS treatment for 24 h (Fig. 3B). Additionally, the primary astrocytes showed an increased expression of TNFα and IL-1β mRNA at early time points; however, no changes were noted in the mRNA levels of SOD2 following treatment with LPS during the experiments (Fig. 3C). Therefore, SOD2 induction downstream of TLRs can be considered specific to microglia in the CNS.

Transcription Factors Responsible for the Inducible Expression of SOD2 in Microglia—A promoter analysis revealed that the promoter region of rat SOD2 has NF-kB and AP-1 binding sequences (Fig. 4A). Furthermore, because AP-1 and NF-kB have been reported to drive the SOD2 expression (38–41), we next examined the involvement of NF-kB and AP-1 in the SOD2 expression induced by LPS in the rat primary microglia.

*Table 2* List of mouse primers for real-time PCR used in this study

| Primer | Forward (5'-3') | Reverse (5'-3') |
|--------|----------------|----------------|
| TNFα   | AGGGCTCTCCCTTCATCACTT | CTTGGTGTGGTTCTGACAGG |
| IL-1β  | AGGGCTCTCCCTTCATCACTT | GAGCCTCTTGCTTCTTGGCT |
| MCP-1  | GCACGTAATGGCCTTGTGTTG | CTGACAGACGCTGAGGAAGG |
| MIP-2α | CTCAGCACTCAACGGTTGAGT | CGGCTACTCCAGGCTGAGA |

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FIGURE 1. Increased expression of SOD2 in microglia treated with LPS. Rat primary microglia were stimulated with 10 ng/ml LPS for 24 h. The mRNA levels of proinflammatory molecules TNFα, IL-1β, COX-2, MCP-1, and MIP-2α (A) and antioxidant enzymes SOD1, SOD2, catalase (Cat), glutathione peroxidase 1 (GPx1), glutathione reductase (GR), and thioredoxin reductase (TR) (B) were determined at 6 h using real-time PCR and are represented as -fold changes from the levels measured in untreated cells. The values represent the mean ± S.E. (error bars) of 8–14 separate experiments. The data were compared using Student’s t test. **, p < 0.01 versus the untreated cells.

C, time course of the mRNA expression of TNFα, IL-1β, and SOD2 in the LPS-stimulated microglia. The values represent the mean ± S.E. of five separate experiments. The data were analyzed using one-way ANOVA, followed by Dunnett’s test. *, p < 0.05; **, p < 0.01 versus the cells at time 0.

D, images of immunoblotting showing the protein levels of SOD2 and α-tubulin at 24 h. The results are representative of three independent experiments.

FIGURE 2. SOD2 expression downstream of TLR stimulation. A, rat primary microglia were treated with 10 ng/ml LPS, 50 μg/ml poly(I:C), 50 μg/ml PGN, or 1 μg CpG DNA for 6 h. The mRNA levels of TNFα, IL-1β, and SOD2 were evaluated using real-time PCR and are represented as -fold change from the levels measured in untreated cells. The values represent the mean ± S.E. (error bars) of eight separate experiments. The data were compared using Student’s t test with Holm’s corrections for multiple comparisons. *, p < 0.05; **, p < 0.01 versus the untreated cells. B, after 24 h of incubation of microglia with LPS, poly(I:C), PGN, or CpG DNA, the levels of SOD2 and α-tubulin proteins were measured using immunoblotting. The results are representative of three independent experiments.

C, rat primary microglia were pretreated with a TLR4 antagonist, TAK-242 (1 μM), followed by treatment with 10 ng/ml LPS for 6 h. The levels of TNFα, IL-1β, and SOD2 mRNA were measured using real-time PCR. The values represent the mean ± S.E. of five separate experiments. The data were compared using Student’s t test with Holm’s corrections for multiple comparisons. **, p < 0.01 versus the untreated cells. ##, p < 0.01 versus the LPS-treated cells.

D, rat primary microglia were pretreated with TNFα or IL-1 receptor antagonists (20 μM WP9QY or 1.5 μg/ml IL-1RA, respectively) followed by treatment with 10 ng/ml LPS for 6 h. The levels of SOD2 mRNA were measured using real-time PCR.
also the amount of SOD2 mRNA induced by LPS (Fig. 4B). Challenges with the MEK1 inhibitor, U0126, and JNK inhibitor, SP600125, also reduced the mRNA expression of TNFα, IL-1β, and SOD2 elicited by LPS (Fig. 4B). A luciferase assay using the cloned rat SOD2 promoter region demonstrated that treatment of microglia with LPS transcriptionally activated the luciferase expression, whereas pretreatment with CAPE, U0126, or SP600125 attenuated the luciferase activity induced by LPS (Fig. 4C). In addition, mutations targeting the NF-κB and AP-1 binding sequences in the SOD2 promoter significantly suppressed luciferase transactivation (Fig. 4D). These results suggest that SOD2 is transcriptionally activated via NF-κB- and AP-1-related pathways downstream of TLRs.

Involvement of Up-regulated SOD2 in the Antioxidant Defenses of Microglia—Because SOD2 is a well known enzyme that scavenges superoxide anions, the most obvious role for the SOD2 induced by LPS is to eliminate the superoxide produced by microglia and attenuate oxidative stress. Therefore, the relationship between an increased expression of SOD2 and oxidative injury in microglia was investigated using the RNAi method. Transfection of SOD2 siRNA clearly suppressed the SOD2 mRNA and protein expression elicited by LPS (Fig. 5, A–C). The intracellular ROS levels in the microglia were increased by treatment with LPS (Fig. 5D). In SOD2-knockdown microglia, ROS production was clearly potentiated in the presence of LPS (Fig. 5D). However, ROS production did not increase in the LPS-treated microglia with SOD2 knockdown to which membrane-permeable SOD (SOD-PEG) was added (Fig. 5D), indicating that the SOD2 elicited by treatment with LPS contributes to the elimination of the ROS produced by LPS stimulation. In addition, the extracellular ROS levels were also increased by treatment with LPS, and ROS production was enhanced by the knockdown of SOD2 (Fig. 5E). The NADPH oxidase inhibitors, DPI and apocynin, partially but significantly suppressed the potentiated ROS generation noted in SOD2-knockdown cells (Fig. 5E), suggesting that the SOD2 up-regulated in response to LPS eliminates the superoxide derived from NADPH oxidase.

Treatment with LPS did not injure microglia carrying control siRNA, whereas the microglia treated with SOD2 knockdown were significantly damaged by LPS, suggesting that SOD2 plays a role in the self-protection of microglia under inflammatory conditions (Fig. 6). When unstimulated microglia were treated with the oxidants, 300 μM hydrogen peroxide or 100 μM xanthine plus 30 milliunits/ml xanthine oxidase for 24 h, cell viability decreased to 41.0 and 36.1%, respectively (Fig. 6). Interestingly, treatment with hydrogen peroxide and xanthine/xanthine oxidase did not induce any reductions in cell viability in the LPS-treated microglia (Fig. 6), demonstrating higher tolerance of the activated microglia to oxidative stress. In contrast, SOD2 knockdown clearly abolished the tolerance of the microglia stimulated with LPS (Fig. 6). Therefore, the SOD2 up-regu-

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**FIGURE 3. Effects of LPS on the SOD2 expression in primary neurons and astrocytes.** A, immunocytochemical staining of rat primary neurons treated with the MAP2 antibody (left) and rat primary astrocytes treated with glial fibrillary acidic protein (GFAP) (right). B and C, rat primary neurons (B) or astrocytes (C) were treated with 10 ng/ml LPS for 24 h, and the time course of the mRNA expression of TNFα, IL-1β, and SOD2 was evaluated using real-time PCR. The values represent the mean ± S.E. of 3–5 separate experiments. The data were analyzed using one-way ANOVA, followed by Dunnett’s test. *, p < 0.05; **, p < 0.01 versus the cells at time 0.
Regulation of Microglial Proliferative Responses by SOD2

![Diagram](https://via.placeholder.com/150)

**FIGURE 4. Involvement of NF-κB and AP-1 in the SOD2 expression in activated microglia.** A, promoter region of the rat SOD2 gene. AP-1m and NF-κBm, mutated sequences of the pGL4.24-SOD2Prom. B, rat primary microglia were pretreated with 10 μM CAPE, 10 μM U0126, or 10 μM SP600125 for 20 min and then stimulated with 10 ng/ml LPS for 6 h. The mRNA levels of TNFα, IL-1β, and SOD2 were evaluated using real-time PCR and are represented as -fold changes from the levels measured in untreated cells. The values represent the mean ± S.E. (error bars) of six separate experiments. The data were analyzed using one-way ANOVA, followed by Student’s t test or Dunnett’s test. The Bonferroni method was used to correct for multiple comparisons. *, p < 0.05; **, p < 0.01 versus the untreated cells. #, p < 0.05; ##, p < 0.01 versus the LPS-treated cells. C, after transfection with pGL4.24-SOD2Prom into rat primary microglia, the cells were cultured for 24 h. The cells were pretreated with 10 μM CAPE, 10 μM U0126, or 10 μM SP600125 for 20 min and then treated with 10 ng/ml LPS and subsequently incubated for 3 h, at which time the luciferase activity was measured. The values represent the mean ± S.E. of 8–10 separate experiments. The data were analyzed using one-way ANOVA, followed by Student’s t test or Dunnett’s test. The Bonferroni method was used to correct for multiple comparisons. **, p < 0.01 versus the untreated group carrying pGL4.24-SOD2Prom; #, p < 0.05; ##, p < 0.01 versus the LPS-treated group carrying pGL4.24-SOD2Prom. D, rat primary microglia were transfected with pGL4.24-SOD2Prom or mutant pGL4.24-SOD2Prom and then cultured for 24 h. The cells were treated with 10 ng/ml LPS for 3 h, followed by measurement of the luciferase activity. The values represent the mean ± S.E. of 8–11 separate experiments. The data were analyzed using one-way ANOVA, followed by Student’s t test or Dunnett’s test. The Bonferroni method was used to correct for multiple comparisons. **, p < 0.01 versus the untreated group carrying pGL4.24-SOD2Prom; ##, p < 0.01 versus the LPS-treated group carrying pGL4.24-SOD2Prom.

ululated by LPS confers tolerance to oxidative stress on microglia in order to enhance their survival under conditions of severe oxidative stress, such as inflammation.

**Involvement of Increased SOD2 in Suppression of the Inflammatory Cytokine Expression**—Of note, we found that the mRNA expression of TNFα and IL-1β 6 h after LPS stimulation was significantly enhanced by SOD2 knockdown, and this enhancement was largely suppressed by pretreatment with the NADPH oxidase inhibitors, DPI and apocynin (Fig. 7A). Potentiation of the cytokine expression was also observed in microglia with SOD2 knockdown 12 h after the addition of LPS (Fig. 7B), indicating that the proinflammatory response is prolonged by SOD2 knockdown. The levels of TNFα and IL-1β proteins released into the culture medium by the SOD2-knockdown microglia were higher than the levels noted in the medium of the control microglia 24 h after stimulation with LPS (Fig. 7C). These data suggest that SOD2 suppresses the cytokine expression to terminate microglial proinflammatory responses.

NF-κB is responsible for the expression of several cytokines under inflammation in the CNS, and ROS are known to regulate the activity of NF-κB (42, 43). Treatment of microglia with ROS or ROS-generating reagents increased the mRNA expression of TNFα, IL-1β, and SOD2 (data not shown). Therefore, we examined the involvement of NF-κB in the potentiation of the TNFα and IL-1β expression mediated by SOD2 knockdown. Consequently, a luciferase assay showed that the NF-κB activity induced by LPS at 1.5 h; however, the activity was predominantly potentiated at the 3-h time point (Fig. 8A). Additionally, SOD2 knockdown slightly increased the NF-κB activity induced by LPS at 1.5 h; however, the activity was predominantly potentiated at the 3-h time point (Fig. 8A). Moreover, the LPS-induced binding of NF-κB to its response element was largely enhanced by SOD2 knockdown (Fig. 8B). Therefore, it is
suggested that the up-regulation of SOD2 suppresses the NF-κB activation induced by ROS, followed by the attenuation of microglial activation.

**Discussion**

In this study, the stimulation of microglial TLR4 by LPS increased the levels of proinflammatory cytokines and chemokines as well as the superoxide-scavenging enzyme, SOD2. TLR family ligands other than LPS, such as poly(I:C), PGN, and CpG DNA, also up-regulated the SOD2 expression, indicating the existence of a common mechanism among members of the TLR family for elevating the SOD2 expression in microglia. Downstream of TLR stimulation, adaptor proteins, such as MyD88 and TRIF, are recruited to the intracellular domain of TLRs, followed by the activation of kinases, including the IRAK family.
ROS generation is induced via uncoupling of the mitochondrial electron transport chain. Once ROS are generated anywhere within cells, further ROS generation from mitochondria has also been reported (10, 11). Of note, once ROS are generated anywhere within cells, further ROS generation is induced via uncoupling of the mitochondrial respiratory chain, known as ROS-induced ROS release (46). In this regard, the regulation of ROS generation by mitochondria is important for controlling the redox balance. In another case, West et al. (47) recently reported that stimulation of TLR1, -2, or -4 in macrophages elicited mitochondrial ROS production via the TRAF6-ECSIT pathway, resulting in bacterial killing. Because SOD2 is localized in mitochondria and increases in the ROS levels induced by SOD2 knockdown were partially abolished by the inhibitors of NADPH oxidase in the present study, ROS are considered to be generated in activated microglia primarily from NADPH oxidase and secondarily from ROS-induced ROS release within the mitochondria.

Cell death was not induced by hydrogen peroxide or xanthine/xanthine oxidase in the microglia treated with LPS, indicating that activated microglia show higher tolerance to oxidative stress than microglia in the resting state. Because activated microglia with SOD2 knockdown were injured by hydrogen peroxide and xanthine/xanthine oxidase, it appears that treatment with LPS up-regulates SOD2 expression, which protects cells form subsequent challenges of oxidative stress induced by hydrogen peroxide or xanthine/xanthine oxidase. Activated microglia generate massive amounts of ROS, which play an important (patho)physiological role in the removal of bacteria, whereas the oxidative stress condition elicited by microglia may potentially damage the microglia themselves. One of the major roles of the SOD2 elicited in activated microglia could be to protect the microglia from the oxidative stress that they generate as a defense against foreign organisms.

Interestingly, the expression of proinflammatory cytokines was increased in the activated microglia with SOD2 knockdown compared with the control siRNA-treated cells. Therefore, it is suggested that SOD2 in activated microglia might negatively regulate the expression of proinflammatory cytokines. In addition, treatment of microglia with LPS stimulated ROS production, and SOD2 knockdown potentiated the increases in ROS induced by LPS, whereas the inhibition of NADPH oxidase by DPI or apocynin largely suppressed ROS production in the microglia. DPI and apocynin also attenuated the LPS-induced expression of TNFα and IL-1β potentiated by the transfection of SOD2 siRNA. Therefore, the ROS levels in microglia correlate with the cytokine expression, suggesting that ROS might regulate the proinflammatory responses of microglia.

The expression of proinflammatory cytokines is tightly controlled by the transcription factor, NF-κB (35), and NF-κB is notably known to be activated by ROS (42, 43). Carboxyalkylpyrroles, the end products of lipid peroxidation, activate NF-κB via TLR2 stimulation and subsequent MyD88 recruitment (48). ROS inactivates phosphatases, after which kinases that are relatively activated, especially IKK, induce NF-κB phosphorylation and subsequent ubiquitination and degradation by proteasomes, followed by the translocation of released NF-κB into the nucleus and subsequent gene transcription (49). In this study, increased NF-κB activity was detected in the microglia treated with LPS, and this effect was enhanced by SOD2 knockdown. Therefore, SOD2 could be considered to negatively regulate the NF-κB activity by eliminating ROS. The mechanism by which microglia are activated under inflammatory conditions is well studied and involves pattern recognition receptors, such as TLRs, which recognize xenobiotics, followed by the...
activation of downstream signals (35). On the other hand, there are few reports about the physiological mechanisms that positively terminate microglial activation and neuroinflammation (e.g. the dopamine D2 receptor-β-crystallin pathway has been investigated) (50). Our findings indicate that microglia up-regulate SOD2 during activation (at the beginning of inflammation), and the increased SOD2 subsequently inactivates microglia (terminating inflammation). Specifically, SOD2 may restrict microglial activation to the minimum extent necessary (Fig. 10). Because SOD2 was transcriptionally increased by NF-κB in activated microglia and SOD2 subsequently decreased the NF-κB activity to reduce the quantity of proinflammatory cytokines, the microglial production of proinflammatory cytokines may be regulated, at least in part, by "negative feedback" involving the NF-κB-SOD2 pathway (Fig. 10). In addition, dead cells contribute to inflammation by scattering their intracellular contents. Therefore, the suppression of microglial oxidative injury by SOD2 might also be involved in attenuating neuroinflammation. Because the excess or prolonged activation of microglia is known to induce various CNS

Regulation of Microglial Proinflammatory Responses by SOD2
Regulation of Microglial Proinflammatory Responses by SOD2

SOD2 is therefore considered to regulate the immune system in the CNS by acting as a switch to control microglial activation/inactivation.

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Regulation of Microglial Proinflammatory Responses by SOD2
Regulation of Microglial Proinflammatory Responses by SOD2

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