Treatment strategies for inherited optic neuropathies: past, present and future

Abstract
Bilateral visual loss secondary to inherited optic neuropathies is an important cause of registrable blindness among children and young adults. The two prototypical disorders seen in clinical practice are Leber hereditary optic neuropathy (LHON) and autosomal dominant optic atrophy (DOA). About 90% of LHON cases are due to one of three mitochondrial DNA (mtDNA) point mutations: m.3460G > A, m.11778G > A, and m.14484T > C, which affect critical complex I subunits of the mitochondrial respiratory chain. The majority of patients with DOA harbour pathogenic mutations within OPA1, a nuclear gene that codes for a multifunctional inner mitochondrial membrane protein. Despite their contrasting genetic basis, LHON and DOA share overlapping pathological and clinical features that serve to highlight the striking tissue-specific vulnerability of the retinal ganglion cell (RGC) layer to disturbed mitochondrial function. In addition to severe visual loss secondary to progressive optic nerve degeneration, a subgroup of patients will also develop a more aggressive syndromic phenotype marked by significant neurological deficits. The management of LHON and DOA remains largely supportive, but major advances in our understanding of the mechanisms underpinning RGC loss in these two disorders are paving the way for novel forms of treatment aimed at halting or reversing visual deterioration at different stages of the disease process. In addition to neuroprotective strategies for rescuing RGCs from irreversible cell death, innovative in vitro fertilisation techniques are providing the tantalising prospect of preventing the germline transmission of pathogenic mtDNA mutations, eradicating in so doing the risk of disease in future generations.

Introduction
Hereditary optic nerve disorders result in significant chronic visual morbidity, and the minimum prevalence of affected individuals in the population has been estimated at 1 in 10 000.1 Leber hereditary optic neuropathy (LHON) and autosomal dominant optic atrophy (DOA) are the two classical paradigms for this group of disorders, and they account for most of the inherited optic atrophy cases seen in clinical practice. LHON is caused by mitochondrial DNA (mtDNA) point mutations, whereas, in DOA, the majority of cases are due to pathogenic mutations in the OPA1 gene, which codes for an inner mitochondrial membrane protein.1,2 Strikingly, both LHON and DOA share the same characteristic pathological features with selective degeneration of the retinal ganglion cell (RGC) layer, leading to progressive optic nerve degeneration and the onset of visual symptoms.3 Rarer autosomal recessive optic atrophy genes have also been identified and a common theme is emerging, with all of them so far encoding for key components of the mitochondrial machinery.1,2 RGCs are therefore exquisitely sensitive to mitochondrial dysfunction, and the elucidation of the mechanisms involved is opening the way for therapeutic interventions targeting different stages of the disease process. In this review, recent advances in our understanding of LHON and DOA will be discussed, in addition to the practical management of this group of patients and emerging treatment options.

Mitochondrial disorders
Oxidative phosphorylation
Mitochondria are ubiquitous intracellular organelles present in all nucleated eukaryotic cells.4,5 They form a highly interconnected network extending throughout the cell’s structure and their overall morphology is
Mitochondria are unique in possessing their own genetic material in the form of a double-stranded circular DNA molecule about 16,569 base-pair long. The mitochondrial genome is therefore very compact, but it codes for structural and functional components that are indispensable for normal mitochondrial function, including two ribosomal RNAs (12S and 16S rRNA), 22 transfer RNAs (tRNAs), and 13 polypeptide subunits of the mitochondrial respiratory chain complexes (I, III, V, and V). Another remarkable feature of mitochondrial genetics is its very high copy number with several thousands of mtDNA molecules being present in metabolically active cells such as neurons. Unlike nuclear DNA, mtDNA replicates continuously in dividing as well as in non-dividing cells, and this process occurs independently of the cell cycle. As a result of this relaxed replication machinery, a cell’s total mtDNA content can be adjusted according to its bioenergetic requirements under both physiological and pathological conditions. Furthermore, the large number of mtDNA molecules present in each cell creates two possible genetic situations, referred to as homoplasmy or heteroplasmy. In the heteroplasmic state, two or more mtDNA variants are present at a specific nucleotide position, and this genetic admixture has been implicated in the variable disease expression that typifies mitochondrial disorders. Most pathogenic mtDNA mutations are heteroplasmic, and mitochondrial respiratory chain activity becomes compromised when the level of the mutant species exceeds a critical threshold (60–80%), which is both mutation and tissue specific. In terms of genetic counselling, the mitochondrial genome shows strict maternal inheritance and male mutation carriers can be reassured that transmission to their own offspring will not occur. In contrast, all the children of a homoplasmic female carrier will harbour their mother’s mtDNA mutation. The situation is rather more complex for a heteroplasmic mother as she could transmit a higher or a lower level of her mtDNA mutation to a particular offspring, which could influence the latter’s risk of becoming clinically affected. The sometimes rapid shifts in mitochondrial allele frequencies that occur between maternally related generations have been explained by a “mitochondrial bottleneck” operating in the female germline.

**Nuclear–mitochondrial interactions**

Mitochondria have limited autonomy and they rely heavily on the nuclear genome for the vast majority of proteins required for mtDNA replication, transcription, and translation. In 1988, two seminal papers were published linking for the first time specific mtDNA mutations with human disease: large-scale single mtDNA deletions in patients with mitochondrial myopathies and the m.11778G>A point mutation in families with LHON. Families segregating classical mitochondrial phenotypes in a clear-cut Mendelian pattern of inheritance were subsequently reported, and the existence of nuclear genetic defects disrupting mitochondrial proteins was widely suspected. This hypothesis was confirmed in 2001, when mutations in the nuclear genes, POLG and PEO1, were identified in families with autosomal dominant chronic progressive external ophthalmoplegia (CPEO). The pathological hallmark of all these nuclear mitochondrial disorders is mtDNA instability, which can be quantitative in nature with a reduction in mtDNA copy number (depletion) or qualitative with the accumulation of high levels of somatic mtDNA mutations (predominantly deletions) in affected tissues. MtDNA molecules are located within the matrix compartment where they are packaged within intricate physical structures known as nucleoids.
components of these fundamental replicative units have not yet been fully established, both POLG and PEO1 are closely associated with mammalian nucleoids, highlighting the critical role played by these two proteins in mitochondrial genome maintenance. Mitochondrial transcription factor A (TFAM) has also been identified as a major nucleoid-organising protein responsible for packaging mtDNA and activating mitochondrial transcription. From a diagnostic perspective, the disruption of OXPHOS that ensues from the accumulation of secondary mtDNA abnormalities can be easily detected in skeletal muscle biopsies as cytochrome c oxidase (COX)-negative fibres. When present, these various molecular and histochemical features can be extremely helpful in the investigation of patients with suspected mitochondrial disease as they provide strong clues pointing towards an underlying nuclear genetic defect.

Leber hereditary optic neuropathy

Molecular genetics

LHON (OMIM 535000) is the most common of the primary mitochondrial DNA (mtDNA) disorders, and the minimum prevalence has been estimated at 1 in 31,000 in the North of the United Kingdom (Figure 1). Comparable prevalence figures of 1 in 39,000 and 1 in 50,000 have been reported in epidemiological studies from the Netherlands and Finland, respectively. The majority of cases (90%) are due to one of three mtDNA point mutations: m.3460G>A (MTND1), m.11778G>A (MTND4), and m.14484T>C (MTND6), which affect key complex I subunits of the mitochondrial respiratory chain. The m.11778G>A mutation is by far the most common pathogenic mutation accounting for ~70% of all LHON cases worldwide. The one notable exception to this rule is the preponderance of the m.14484T>C mutation (~90%) in patients of French Canadian descent secondary to a mutational founder event. In most diagnostic laboratories, screening for the three ‘primary’ LHON mutations is carried out in the first instance in suspected cases. If this is negative and there is still a strong clinical suspicion of LHON, full mitochondrial genome sequencing can be considered. However, close liaison with an experienced laboratory is essential, not only because it remains a time consuming and expensive exercise, but interpretation of the sequencing results is not always straightforward. A number of relatively rare pathogenic mtDNA variants causing LHON have been confirmed, but others have only been reported in single patients or families and additional irrefutable proof of pathogenicity is still needed, which greatly complicates patient counselling (Table 1).

Table 1

| Disorder                                      | Minimum Prevalence |
|-----------------------------------------------|--------------------|
| Mitochondrial genetic disorders               | 1 in 5,000         |
| Leber hereditary optic neuropathy (LHON)      | 1 in 31,000        |
| Autosomal dominant optic atrophy (DOA)       | 1 in 25,000        |
| Optic neuropathy in mitochondrial cytopathies*| 1 in 10,000        |

Figure 1 The prevalence of inherited optic neuropathies in the North of the United Kingdom. *Includes patients with LHON and DOA.
Clinical features

Fundal abnormalities such as peripapillary telangiectatic vessels and transient retinal nerve fibre layer (RNFL) oedema can be observed in some asymptomatic LHON carriers.17,18 On more detailed psychophysical testing, some unaffected LHON carriers also show subclinical optic nerve impairment with reduced colour vision and contrast sensitivity, and subnormal responses on multifocal electroretinography and visual evoked potentials.19 Disease conversion in LHON is characterised by acute or subacute, painless, central visual loss in one eye, followed weeks to months later by the fellow eye.20 The median inter-eye delay is 6–8 weeks, and the second eye is invariably affected within 1 year of disease onset. Bilateral simultaneous onset occurs in a proportion of patients (~25%) with the caveat that some individuals are probably unaware that visual loss had been ongoing unnoticed in one eye before the second eye being involved. Although there are rare case reports of unilateral optic nerve involvement in LHON, it is important to exclude other underlying pathological causes in these highly atypical presentations. The peak age of onset is in the second and third decades of life, and it is unusual for LHON carriers to experience visual loss beyond the age of 50 years.20 Visual acuity worse than 6/60 is the norm at the nadir, and there is an associated dense central or centrocecal scotoma with profound dyschromatopsia. Interestingly, the pupillary light reflexes are relatively preserved in LHON due to the sparing of a special class of melanopsin-containing RGCs that seem more resistant to mitochondrial dysfunction.21

A careful dilated fundus examination can be particularly informative in the acute stage of LHON as patients classically exhibit a number of distinct abnormalities, namely, optic disc hyperaemia, peripapillary telangiectatic vessels, vascular tortuosity, and swelling of the retinal nerve fibre layer (RNFL) secondary to axoplasmic stasis (Figure 2a).1,2 It is important to stress that the posterior pole can look entirely normal in a proportion (20–40%) of affected LHON carriers and the diagnosis may be delayed until optic disc pallor becomes apparent at around 6 weeks. This group of patients are frequently labelled as having functional visual loss, at least initially. In addition to pallor of the neuroretinal rim, non-glaucomatous cupping of the optic disc is also a recognised pathological feature of longstanding LHON cases, reflecting the progressive loss of RGCs that occurs in the chronic phase of the disease process.1,2

The visual prognosis in LHON is poor and the majority of patients will remain legally blind with a significant detrimental impact on their overall quality of life.22 Spontaneous visual improvement can occur and it is most likely within the first year. Remarkably, it has also been reported several years or even decades following the initial visual loss. Visual recovery is usually heralded by the appearance of small islands of vision in the central visual field and, as the scotoma

| Mitochondrial gene | Nucleotide change |
|--------------------|-------------------|
| **Common variants** |                   |
| MTND1              | m.3460G>A         |
| MTND4              | m.11776G>A        |
| MTND6              | m.14484T>C        |
| **Rare variants**  |                   |
| MTND1              | m.3376G>A, m.3635G>A, m.3697G>A, m.3700G>A, m.3733G>A, m.4025C>T, m.4160T>C, m.4171C>A |
| MTND2              | m.4640C>A, m.5244G>A |
| MTND3              | m.10237T>C        |
| MTND4              | m.11696G>A, m.11253T>C |
| MTND4L             | m.10663T>C        |
| MTND5              | m.12811T>C, m.12848C>T, m.13637A>G, m.13730G>A |
| MTND6              | m.14325T>C, m.14568C>T, m.14459G>A, m.14729G>A, m.14482C>A, m.14482C>G, m.14495A>G, m.14498C>T, m.14568C>T, m.14596A>T |
| MTATP6             | m.9101T>C         |
| MTCO3              | m.9804G>A         |
| MTCYB              | m.14831G>A        |

*These mtDNA variants are definitely pathogenic. They have been identified in ≥2 independent LHON pedigrees and show segregation with affected disease status. The remaining putative LHON mutations have been found in singleton cases or in a single family, and additional evidence is required before pathogenicity can be irrefutably ascribed.¹
gets concurrently less dense, these fenestrations can greatly help with scanning vision and peripheral navigation.1,2 LHON carriers harbouring the m.14484T>C mutation have the best visual prognosis with a partial visual recovery rate of 37–58%, compared with 4–25% for the m.11778G>A mutation.12,23-25 Other reported positive prognostic factors for visual recovery in LHON include an earlier age of onset (<20 years), especially among young children; a subacute presentation with slow visual deterioration; and a relatively large optic disc.26,27

**LHON plus phenotypes**

An abiding mystery of LHON is the tissue-specific vulnerability of the RGC layer despite the ubiquitous presence of the pathogenic mtDNA mutation. Although visual loss is the defining feature of this mitochondrial disorder, cardiac conduction defects and neurological abnormalities such as ataxia, peripheral neuropathy, myopathy, and dystonia have been reported to be more common among LHON carriers compared with controls.1,2 However, because of the relative rarity of
Incomplete penetrance and sex bias

Not all LHON mutation carriers will experience visual loss during their lifetime. The conversion rate for male carriers is \( \sim 50\% \) compared with \( \sim 10\% \) for female carriers. LHON is therefore characterised by both marked incomplete penetrance and a striking male bias for visual loss. The secondary factors that modulate the phenotypic expression of the underlying pathogenic mtDNA mutation have been the subject of intense research over the past two decades. Although we do not yet have the full picture of what is proving to be a relatively complex disease, a number of genetic, hormonal, and environmental factors have been identified, some of which are particularly relevant to patient counselling and the development of drug treatments for LHON.

Heteroplasmy is thought to influence disease penetrance based on the observation that affected LHON carriers have mutation levels greater than 60\% in peripheral blood leukocytes.\(^{31}\) However, incomplete penetrance is still observed among heteroplasmic carriers with mutation levels exceeding this nominal threshold, and the majority of LHON families (80–90\%) harbour homoplasmic mtDNA mutations.\(^{35}\) Heteroplasmy is therefore not a major contributor to the incomplete penetrance pattern seen in LHON. Another factor that could be modulating the risk of visual loss in LHON is the mitochondrial genetic background on which the pathogenic mtDNA mutation is segregating. The mitochondrial genome is highly polymorphic, and, during human evolution, ancient mtDNA variants have gradually clustered together in specific combinations known as haplogroups.\(^{4,5}\) Mitochondrial haplogroups have a differential effect on the efficiency of the mitochondrial respiratory chain, and this could possibly influence the overall deleterious impact of the LHON mutation on RGC survival.\(^{32,33}\) In a large meta-analysis of Caucasian LHON pedigrees, haplogroup J was associated with a significantly increased risk of visual loss among m.11778G \( \rightarrow \) A and m.14484T \( \rightarrow \) C carriers, whereas m.3460G \( \rightarrow \) A carriers were more likely to become affected on a haplogroup K background.\(^{34}\) LHON carriers with the m.11778G \( \rightarrow \) A mutation had a lower risk for visual loss when the mutation arose on haplogroup H. Other haplogroup associations have been reported in families with LHON from mainland China, but none in those from South-East Asia.\(^{35,36}\) The influence of specific mtDNA polymorphisms on the risk of visual failure in LHON is therefore not entirely clear-cut, and further investigation is warranted.

The predominance of affected males in LHON cannot be explained by mitochondrial inheritance. Segregation analysis of a large number of LHON families was consistent with the existence of a recessive X-linked susceptibility gene acting in synergy with the mtDNA mutation to precipitate visual loss.\(^{37}\) Three independent linkage studies have revealed overlapping candidate regions on the X-chromosome, but the actual modifier gene has still not yet been identified.\(^{38-40}\) The existence of autosomal nuclear modifiers in LHON remains a distinct possibility, and the situation could prove even more complex if different combinations of nuclear genes are segregating in different ethnic populations.

Another obvious explanation for the striking marked male bias in LHON is a protective benefit from female sex hormones. Interestingly, treatment with 17\%oestradiol resulted in reduced reactive oxygen species (ROS) levels, increased activity of the antioxidant enzyme superoxide dismutase (SOD), and more efficient mitochondrial biogenesis in LHON cybrid cell lines.\(^{41}\) As RGC cell bodies have a high concentration of oestrogen \( \beta \) receptors, supplementation with oestrogen-like compounds represents an attractive therapeutic option. Various environmental triggers have been implicated in precipitating the onset of visual loss in LHON, including head trauma, psychological stress, exposure to industrial toxins, and drugs with mitochondrial toxic effects such as antiretrovirals and ethambutol.\(^{1,2}\) A recent multicentre study that enrolled 125 Northern European LHON pedigrees has provided important insight into the role of smoking and alcohol consumption on disease expression.\(^{42}\) Smoking was strongly associated with an increased risk of visual loss, and this detrimental effect was more marked for heavy smokers compared with...
light smokers. Various toxins present in cigarette smoke are capable of depressing mitochondrial ATP synthesis either through a direct inhibitory effect on complex I activity or by reducing arterial oxygen concentration. There was also a trend towards an increased risk of visual failure with heavy binge drinking, but this effect was not as marked as for smoking.

Disease mechanisms

The papillomacular bundle is affected early and much more severely in LHON. The preferential involvement of the RGCs within the papillomacular bundle is likely related to their relatively small calibres and limited mitochondrial energetic reserves.43 Two major mechanisms have been proposed to precipitate RGC loss in LHON—a biochemical respiratory chain defect and increased levels of reactive oxygen species (ROS).44 There are supporting lines of evidence for both these pathological pathways, and they represent obvious therapeutic targets to halt the catastrophic loss of RGCs that occurs during the acute phase of LHON. The three most common primary LHON mutations (m.3460G > A, m.11778G > A, and m.14484T > C) all affect critical complex I subunits, and impaired complex I-driven ATP synthesis has been identified using both in vitro and in vivo biochemical assays.1,3 A mouse model has recently been developed that replicates some of the key histopathological features seen in the optic nerves of affected LHON patients, namely the preferential degeneration of the smallest calibre RGC fibres, marked axonal swelling, and the accumulation of dysmorphic mitochondria within demyelinated segments.45 Increased ROS levels were observed with no reduction in ATP synthesis, strongly supporting oxidative stress as the major mechanism driving the loss of RGCs, at least in that specific mouse model.

Autosomal dominant optic atrophy

Molecular genetics

Autosomal dominant optic atrophy (DOA, OMIM 605290) is the most commonly diagnosed inherited optic neuropathy, and, in a recent epidemiological study, the minimum prevalence was estimated at 1 in 25,000 in the North of the United Kingdom (Figure 1).46 The majority of families with DOA (~ 60%) harbour pathogenic mutations in OPA1 (3q28-q29), which codes for a dynamin-related GTPase protein located within the inner mitochondrial membrane.47,48 It is a ubiquitous protein and abundant levels have been detected in RGCs, the inner ear, and the brain. OPA1 is highly polymorphic and over 200 pathogenic mutations have been identified with hotspots in the catalytic GTPase domain (exons 8–15) and the dynamin central domain (exons 16–23).49 Most pathogenic mutations result in a marked reduction in the level of the OPA1 protein, and the pathological significance of haploinsufficiency is further highlighted by those rare families with large-scale deletions spanning the entire OPA1 coding region.50

Clinical features

DOA has an insidious onset in early childhood and it typically presents with bilateral, symmetrical, central visual loss and dyschromatopsia. There is a wide variability in disease severity with visual acuities ranging from 6/6 to the detection of hand movement.51 Asymptomatic OPA1 mutation carriers with subclinical disease are frequently identified during contact tracing. If there is no definite family history, it is therefore important to examine other family members of a patient with suspected inherited optic atrophy to establish more clearly the pattern of inheritance. Visual deterioration has been reported in 19 to 67% of patients with DOA on long-term follow-up, with 13 to 46% being registered legally blind.1,2 Visual decline in DOA is usually slow, and a good proportion of patients will retain good functional vision late into their disease. However, the rate of visual loss can be highly variable both between and within families, and some patients with DOA experience a much faster rate of disease progression.51

Similar to LHON, the pathological hallmark of DOA is the preferential early loss of RGCs within the papillomacular bundle, accounting for the distinctive temporal wedge of pallor that is commonly observed at the optic nerve head (Figure 2b). However, pallor of the neuroretinal rim can be subtle, and the demonstration of pathological thinning of the retinal nerve fibre layer (RNFL), especially in the temporal quadrant, with optical coherence tomography (OCT) imaging can be particularly helpful.52 Electrophysiological testing can also be informative in these equivocal cases. The pattern visual evoked potentials in DOA is characterised by abnormally prolonged latencies and a variable decrease in magnitude. The N95 component of the pattern electroretinogram (pERG) is usually selectively depressed consistent with primary dysfunction of the RGC layer. Deep excavation of the optic disc with an enlarged cup-to-disc ratio greater than 0.5 is another typical morphological feature observed in DOA.51

It is therefore not surprising that patients with DOA are often misdiagnosed as having normal tension glaucoma, especially if the degree of optic disc pallor is mild and there is no convincing family history. Limited post mortem studies suggest the relative preservation of melanopsin-containing RGCs, and this could account for the lack of an afferent pupillary defect in patients with DOA, which is another clinical feature shared with LHON.21
DOA plus phenotypes

Progressive central visual loss is the defining feature of DOA, but with greater access to molecular genetic testing, a much broader array of clinical presentations have now been linked to pathogenic OPA1 mutations. The most common extraocular feature is sensorineural hearing loss developing in the first two decades of life and a specific OPA1 mutation in exon 14 (c.1334G > A, p.R445H) has been strongly associated with the co-occurrence of deafness and optic atrophy.53 In a large multicentre study, nearly 20% of OPA1 mutation carriers developed a more severe neuromuscular phenotype complicated by varying combinations of deafness, ataxia, myopathy, peripheral neuropathy, and CPEO.53 Interestingly, some patients have also been reported with clinical features indistinguishable from MS, hereditary spastic paraplegia (HSP), and the inherited spinocerebellar degenerations. From a mechanistic perspective, these so-called dominant optic atrophy plus (DOA +) phenotypes confirm that the pathological consequences of OPA1 mutations are not limited to RGCs, but they clearly extend to other central neuronal populations, peripheral nerves, and skeletal muscle.

Disease mechanisms

OPA1 is a multifunctional protein located within the inner mitochondrial membrane. It is a critical pro-fusion protein and pathogenic OPA1 mutations result in marked mitochondrial network fragmentation.54 This physical disruption has a knock-on effect on the stability of the mitochondrial respiratory chain complexes, resulting in increased ROS levels and a reduction in overall ATP synthesis.55,56 Pro-apoptotic cytochrome c molecules are carefully sequestered within the cristae spaces by the zipper-like action of OPA1 and mitochondria are major stores of calcium.57,58 Mitochondrial network fragmentation results in the uncontrolled release of these potent pro-apoptotic factors, which ultimately trigger irreversible cell death. Although the mechanisms still need to be fully elucidated, OPA1 mutations result in mitochondrial genome instability and the accumulation of multiple mtDNA deletions in affected tissues of some patients.59 Patients with DOA + phenotypes harbour significantly higher levels of these somatic mtDNA abnormalities suggestive of a contributory role in the development of the more severe neuromuscular complications.53 Although OPA1 regulates a whole host of interrelated cellular pathways, compromised OXPHOS and elevated ROS levels are two major disease mechanisms that overlap with LHON, raising the exciting prospect of generic neuroprotective strategies applicable to both these mitochondrial optic neuropathies.

Patient Management

General supportive measures

The sudden onset of severe visual loss in LHON causes significant distress and anxiety for patients and their families. The majority of patients are young healthy adults, and the shock of receiving a genetic confirmation of LHON is often compounded by the diagnostic delays and uncertainties that are not uncommon for rare genetic disorders. Although DOA has a less marked presentation compared with LHON, visual loss can be progressive and in a quality of life survey, half of all the patients interviewed reported high levels of anxiety and depression.50 Clinicians therefore have an important role to play in facilitating access to rehabilitative services such as low visual aids and occupational therapy. Patient groups and information websites can also provide invaluable support, especially in the immediate period after a diagnosis has been made (https://sites.google.com/site/planetleeder/lhon and http://www.lhon.org/lhon/LHON.html, accessed on 8 December 2013). In addition to visual loss, patients with LHON and DOA can also develop neurological features such as ataxia, peripheral neuropathy, and hearing impairment. Clinicians need to be aware of these ‘plus’ phenotypes to ensure their early detection, and patients in this high-risk group are best served by a multidisciplinary team to aggressively manage and minimise the functional consequences of these systemic complications.

Genetic counselling

The mitochondrial genome is maternally inherited and male LHON carriers can be reassured that their children are not at risk of inheriting their genetic defect. In contrast, female carriers will transmit the mutation to all their children, but, if the mutation is heteroplasmic, the level of the mtDNA mutation transmitted can fluctuate, sometimes markedly, as a result of the mitochondrial genetic bottleneck. Prenatal testing for heteroplasmic female LHON carriers is therefore unhelpful because the mutant load detected in amniocytes and chorionic villi may not correspond to that in other fetal cell populations, especially those destined to mature into RGCs. A frustrating aspect of the management of LHON is the current inability to accurately predict whether a familial carrier will eventually experience visual loss. Despite these limitations, individuals can be counselled based on the two major identifiable risk factors in this disorder, namely age and sex.1,2 Male carriers have about a 50% lifetime risk of visual failure compared with only 10% for female carriers, and the peak age of onset is in the second and third decades of life. As a general health measure, patients with mitochondrial disorders should be strongly
advised not to smoke and to moderate their alcohol intake. This is especially the case for unaffected LHON carriers as smoking, and to a lesser extent excessive binge drinking, has been linked with an increased risk of disease conversion.42 It also seems sensible to avoid exposure to other putative environmental triggers for visual loss in LHON, in particular industrial toxins and drugs with mitochondrial-toxic effects, for example ethambutol.1,2

DOA is an autosomal dominant Mendelian disorder, and, although genetic counselling is straightforward, providing accurate information about likely disease severity is challenging for several reasons. It is not possible to predict the rate of progression of visual loss, which can be highly variable even within families harbouring the same pathogenic OPA1 mutation.51 Furthermore, although patients harbouring missense OPA1 mutations in the GTPase domain of the protein have a statistically higher risk of developing DOA + phenotypes, not all of them will do so, and the development of neurological features only becomes manifest in mid- to late adulthood, introducing another element of uncertainty.53 OPA1 mutation carriers have a 50% chance of transmitting the pathogenic allele to each of their children, and pregnant women can request prenatal testing after having been appropriately counselled. Prenatal diagnosis is possible by analysing DNA extracted from fetal cells and the method used will depend on the mother’s personal choice and her gestational status, either amniocentesis (15–18 weeks) or chorionic villus sampling (10–12 weeks).

Treatment options—LHON

Mitochondrial cocktails

There is currently only limited evidence to support the use of any intervention in mitochondrial disorders.61 In a recent systematic review of the literature, only 35 out of 1039 publications on treatments for mitochondrial diseases included more than five patients, and most of these studies suffered from significant methodological weaknesses.62 Over the years, various combinations of vitamins (B2, B3, B12, C, E, and folic acid) and other compounds with putative mitochondrial antioxidant and bioenergetics properties have been used to treat patients with mitochondrial disease, including LHON, but none with any convincing proof of efficacy.61–63 The list of supplements that are frequently promoted on the internet includes alpha-lipoic acid, carnitine, creatine, and L-arginine.

Ubiquinone analogues

Ubiquinone is an essential carrier that ensures the efficient transfer of electrons along the mitochondrial respiratory chain. Pathogenic mtDNA mutations associated with LHON destabilise complex I resulting in a reduction in ATP synthesis and increased ROS levels. An attractive strategy to restore the flow of electrons along the mitochondrial respiratory chain is to bypass the blockage occurring at the level of complex I by enabling and maximising the direct shuttling of electrons to complex III. Coenzyme Q10 (CoQ10) is a ubiquinone analogue with a long history of use for patients with a broad range of mitochondrial disorders.61–63 Although there is a sound scientific basis based on the potential antioxidant and bioenergetic properties of CoQ10, a Cochrane review found no objective evidence of any significant patient benefit in the limited number of small studies that have been published.61 Putting these methodological issues aside, the lack of efficacy of CoQ10 could be due to its lipophilic nature, this physical property hindering its delivery to mitochondria following oral administration of the drug. There is one anecdotal report of an affected patient carrying the m.11778G>A mutation who experienced marked visual improvement after he was started on CoQ10 about 8 months following the first onset of symptoms.64 This could simply have been a chance occurrence given that spontaneous visual improvement has been noted in up to 25% of patients harbouring this mtDNA mutation.

Idebenone and EPI-743 are newer generation shorter-chain analogues of ubiquinone that are reported to be more potent compared with CoQ10, at least in vitro.28,65 The possible visual benefit of idebenone in LHON was first reported in 1992 by Mashima and colleagues who treated a 10-year-old boy harbouring the m.11778G>A with a relatively low daily dose of 90 mg/day.66 The obvious confounding factor in this case is the relatively young age of the patient, as childhood-onset LHON seems to be a distinct entity associated with a much more benign clinical course and a higher rate of spontaneous visual recovery.26 In a larger case series from the same Japanese group, 14 patients with LHON were treated with a combination of idebenone (180 mg/day), vitamin B2, and vitamin C.57 No significant difference in the proportion of eyes showing visual recovery was observed when this intervention group was retrospectively compared with 14 untreated patients. However, if it occurred, a faster rate of visual recovery was reported in the treated eyes compared with the untreated eyes. In contrast to these results, Barnils and colleagues68 found no visual benefit with the use of the same treatment cocktail in two visually affected individuals carrying the m.11778G>A mutation.

To address the true potential of idebenone as a treatment modality in LHON, a multicentre, double-blind, randomised placebo-controlled trial was initiated. RHODOS (Rescue of Hereditary Optic Disease
Outpatient Study, ClinicalTrials.gov identifier: NCT00747487) enrolled 85 patients with a confirmed primary mtDNA mutation (m.3460G > A, m.11778G > A, and m.14484T > C) and with disease duration of up to 5 years.29 The active arm received idebenone (900 mg/day), and no adverse drug reactions occurred over a treatment period of 24 weeks. This trial failed to show benefit for its pre-specified primary end point (best recovery of visual acuity at week 24), but all of the secondary end points showed a positive trend towards visual improvement in the idebenone-treated group. A post hoc analysis of the trial data also indicated that patients with discordant visual acuities (LogMAR > 0.2), and hence at highest risk of further deterioration in the least affected eye, were more likely to benefit from treatment with idebenone. In the follow-up study (RHODOS-OFU), the beneficial effect of 24 weeks of treatment with idebenone seemed to persist following discontinuation of the active medication at the end of the trial.69 In a large retrospective Italian study involving 103 patients with LHON, 44 patients with visual loss of 1 year’s duration or less were treated with idebenone and followed for at least 5 years.70 A greater proportion of patients receiving idebenone recovered vision and the most consistent prognostic factors associated with visual recovery were the early initiation of treatment and a more prolonged course of treatment.

A marketing authorisation application by Santhera Pharmaceuticals Ltd (Liestal, Switzerland) to the European Medicines Agency’s Committee for Medicinal Products for Human Use (CHMP) has recently received an unfavourable opinion and additional clinical evidence for the efficacy of idebenone in LHON has been requested by CHMP (http://www.santhera.com/index.php?docid=212&vid=&lang=en&newsdate=20130116&newsid=1671337&newslang=en, accessed on 8 December 2013). The Newcastle Mitochondrial Research Group is planning a second double-blind, randomised, placebo-controlled trial that will investigate the benefit of treatment with a higher dose of idebenone (2250 mg/day) given for a total duration of 48 weeks instead of 24 weeks as for RHODOS (Table 2). Further work is also needed to determine the appropriate duration of treatment, once initiated, and whether there is a place for the treatment of asymptomatic carriers or patients with longstanding visual loss. Although idebenone is currently not licensed for clinical use, patients with LHON frequently opt to gain access to it at their own expense from various internet sources. EPI-743 has been used in an open-labelled study of five patients with LHON who were treated within 90 days of disease conversion. Although the results need to be confirmed in a properly designed clinical trial, four out of five patients demonstrated arrest of disease progression and variable extents of visual improvement.65

**Brimonidine**

Brimonidine is a topical x2 agonist that is commonly used to lower intraocular pressure in the management of glaucoma.71 Some studies have suggested that brimonidine can exert a synergistic RGC neuroprotective effect by upregulating a number of antiapoptotic factors and by blocking glutamate excitotoxicity induced by mitochondrial oxidative stress.72 Topical brimonidine has therefore been tested as a prophylactic agent for second-eye involvement in an open-labelled study of nine patients with unilateral acute visual loss secondary to LHON.72 Brimonidine failed to prevent fellow eye involvement, and there was no evidence of any visual benefit following the onset of visual loss. Although this study was negative, raised intraocular pressure has been linked with an increased risk of visual loss in LHON and an agent with putative neuroprotective properties such as brimonidine could represent an ideal choice of treatment for unaffected LHON carriers diagnosed with glaucoma or ocular hypertension.73

**Steroids and immunosuppressants**

Patients with LHON are not infrequently treated with high-dose steroids before a molecular diagnosis has been secured to exclude the possibility of an inflammatory optic neuropathy. Steroids do not prevent the involvement of the fellow unaffected eye in LHON, and there has been no reported benefit on disease progression and the final visual outcome.63 In an in vitro experimental paradigm involving the supplementation of the culture media with hydrogen peroxide, LHON cybrids harbouring the m.11778G > A mutation showed an increased sensitivity to oxidative stress.74 This increased susceptibility to undergo apoptosis was postulated to be secondary to a toxic rise in intracellular calcium and the activation of the mitochondrial transition pore (MTP). Pre-treatment with cyclosporin A blunted the deleterious consequences of hydrogen peroxide by blocking the MTP pore, indicating a possible therapeutic pathway for LHON. The antiapoptotic effect of cyclosporine A has also been demonstrated in LHON cybrids harbouring the m.14484T > C and m.14279G > A mutations.75 On the basis of these in vitro data, a French study is currently underway recruiting patients with unilateral visual loss from LHON for treatment with cyclosporin A in an attempt to prevent the involvement of the fellow eye (Dr Dominique Bonneau, University of Angers, France, personal communication).

**Hyperbaric oxygen therapy**

There is highly anecdotal ‘internet’ evidence of patients with LHON benefiting from hyperbaric oxygen therapy.
Near-infrared light (NIR) therapy has been shown to improve mitochondrial function and cellular survival in various models of wound healing, neurodegeneration and methanol-induced retinal toxicity. However, these findings are not universally accepted and the mechanisms are not fully understood. NIR photobiomodulation is thought to increase ATP synthesis by stimulating the activity of cytochrome c oxidase (complex IV). HBOT is a controversial treatment modality that has been applied with limited success to other optic nerve disorders such as radiation-induced optic neuropathy and anterior ischaemic optic neuropathy. Given the rather slim evidence base, the theoretical toxic effects of supraphysiological levels of oxygen in LHON should be considered in the context of a dysfunctional mitochondrial respiratory chain that is already producing increased ROS levels.

Near-infrared light therapy

Near-infrared light (NIR) therapy has been shown to improve mitochondrial function and cellular survival in various models of wound healing, neurodegeneration and methanol-induced retinal toxicity. Although these findings are not universally accepted and the mechanisms are not fully understood, NIR photobiomodulation is thought to increase ATP synthesis by stimulating the activity of cytochrome c oxidase (complex IV). The application of NIR therapy to RGCs via a light-emitting diode has therefore been proposed as a possible rescue strategy for LHON. A study was initiated to investigate the visual benefit of NIR therapy in affected LHON carriers, but it has been terminated because of the inability to record reliable pERG measurements, which was the planned primary outcome measure, due to poor subject fixation.

Treatment options: DOA

Compared with LHON, the rate of RGC loss in DOA is relatively slow and the detection of a clinically meaningful benefit over the course of a 1- or 2-year treatment trial is a major methodological consideration. Long-term natural history studies are therefore urgently needed to more precisely define the visual parameters that are most sensitive at detecting a significant change in optic nerve structure and function for insidiously progressive optic nerve disorders like DOA (Table 2). Idebenone has recently been evaluated in a limited case series involving seven patients with DOA and confirmed pathogenic OPA1 mutations. A variable daily dose of idebenone was used (270–675 mg), and all the patients were treated and reviewed for at least 1 year. No adverse drug reactions were reported and some improvement in visual function was reported for five of the seven idebenone-treated patients. The results of this pilot study remain preliminary and a randomised, placebo-controlled trial with an adequate duration of follow-up will be needed to prospectively evaluate the possible benefit of using idebenone in DOA. Looking into the future, an open-labelled study of EPI-743 for patients with DOA is currently under preparation (Dr Valerio Carelli, University of Bologna, Italy, personal communication), and the potential benefit of near-infrared light in rescuing RGC loss is also being tested in a mouse model of DOA harbouring a splice site mutation (c.1065 + 5g > a) within intron 10 of the Opa1 gene (Professor Marcela Votruba, Cardiff University, UK).

Future prospects and challenges

Drug screening and development

With idebenone and EPI-743, we are witnessing the first tentative steps in our effort to try and modulate disease progression for mitochondrial optic neuropathies. Several research groups worldwide are actively pursuing the identification of novel neuroprotective agents for other, more prevalent, optic nerve disorders such as glaucoma and anterior ischaemic optic neuropathy, the results of which are likely to be highly relevant for LHON and DOA. The technological revolution of the past decade has also launched a new era of personalised medicine with powerful complementary approaches that combine in vitro screening of small molecule libraries with further in vivo validation using existing animal models. Greater collaboration with the pharmaceutical industry will be an important element of this major translational push for rare genetic disorders. More than ever, clinical academics will have an important role to

Table 2  Translational research initiatives for inherited optic neuropathies

The National Institute for Health Research (NIHR) is spearheading an ambitious translational research programme for rare genetic diseases in the United Kingdom. Inherited optic neuropathies have been selected as one of the priority disease themes. As part of this specific NIHR study, we will undertake deep phenotyping of patients with LHON and DOA to define the natural history of the visual loss and to identify the most sensitive parameters for monitoring disease progression. Importantly, this study will provide useful information on the visual outcome measures that are most likely to reveal a clinically meaningful treatment effect in future clinical trials for pipeline neuroprotective agents. Please get in touch with us if you have eligible patients who could be enrolled into this national initiative for translational research into inherited optic neuropathies.
play in taking the lead and setting the broad directions for the clinical evaluation and validation of new drugs for inherited optic neuropathies. Some obvious candidates are compounds that are thought to activate mitochondrial biogenesis, for example oestrogen-like molecules and acetyl-L-carnitine (ALCAR), which is derived from the acetylation of L-carnitine within mitochondria. ALCAR has the advantageous property of crossing the blood–brain barrier, and this molecule promotes OXPHOS by upregulating the transcript levels of ‘master’ genes involved in mitochondrial biogenesis, namely PGC-1α, PGC-1β, and TFAM. A study is currently nearing completion looking at the effect of ALCAR on neuronal conduction along the visual pathways in patients with chronic LHON and disease duration of more than 2 years (http://apps.who.int/trialsearch/trial.aspx?trialid=EUCTR2009-016982-26-IT, accessed on 8 December 2013).

**Gene therapy**

Gene replacement therapy for LHON is an attractive strategy, given the easy anatomical accessibility of the RGC layer for direct manipulation. However, the double-membrane nature of mitochondria presents a formidable series of technical challenges that need to be overcome. First and foremost, a highly efficient vector is needed to penetrate the relatively impermeable inner mitochondrial membrane and to allow a sufficient number of mitochondria to be transfected in order to achieve the desired gene replacement effect. A possible solution is to bypass the mitochondrial genome altogether by using an elegant alternative approach that relies on the nuclear allotopic expression of the gene of interest. The relevant mtDNA gene can be efficiently transfected into the nuclear compartment with an adeno-associated virus (AAV) vector after it has been reconfigured to fit the slightly different coding system operating within the nuclear genome. This hybrid nuclear-encoded protein has also been engineered with a specific targeting sequence to facilitate its efficient import into mitochondria, thereby compensating for the missing or dysfunctional mitochondrial protein. The potential of this gene therapy approach in rescuing the disease phenotype was first demonstrated in m.11778G→A LHON cybrids. The ability to rescue RGCs and improve visual function was subsequently confirmed in vivo by two independent research groups working on LHON rodent models expressing mutated ND4 (m.11778G→A) complex I subunits. These groundbreaking experiments are paving the way for more advanced studies involving primates and ultimately patients with LHON (http://www.gensight-biologics.com/, accessed on 8 December 2013). However, a note of caution is required here given the ongoing debate as to whether the imported wild-type ND4 subunit actually integrates into the native complex I to produce a stable functional unit within the inner mitochondrial membrane.

Proof-of-principle has also been demonstrated for two other forms of gene therapy based on nuclear allotopic expression. One strategy involves the transfection of the nuclear genome with the neuroprotective SOD2 gene packaged into an AAV vector. The overexpression of the superoxide dismutase ROS scavenger in m.11778G→A LHON cybrids suppressed apoptosis and resulted in increased cell survival. The consolidation of the cell’s antioxidant defences could therefore prove a complementary approach to the replacement of the mutated gene product in LHON. A radically different strategy is based on the xenotopic expression of Ndi1, an alternative NADH oxidase expressed in yeast (Saccharomyces cerevisiae) mitochondria. Ndi1 is a versatile enzyme that can bypass a malfunctioning complex I to restore downstream electron transfer while at the same time suppressing ROS overproduction. Successful rescue of optic nerve degeneration was achieved using the yeast Ndi1 gene in a rat model of LHON that was generated by the injection of rotenone-loaded microspheres into the optic layer of the rat superior colliculus.

Despite the technical difficulties of directly introducing genetic material into mitochondria, two recent studies have suggested that this could be achieved using two very different genetic engineering approaches. In one study, whole circular mtDNA molecules isolated from healthy human donors were complexed with human recombinant TFAM, and this genetic construct was able to gain direct entry into cells. When applied to m.11778G→A LHON cybrids, this novel strategy was able to restore cellular respiration and ATP synthesis by promoting mitochondrial biogenesis. In the other study, a mitochondrial targeting sequence was attached to the capsid element of an AAV vector carrying the replacement ND4 subunit gene. This additional tag allowed the direct introduction of the wild-type ND4 gene into the mitochondrial compartment, leading to the successful rescue of the pathological phenotype in both m.11778G→A LHON cybrids and a mouse model manifesting optic atrophy secondary to allotopic expression of the human ND4 mutation. The introduced ND4 gene construct is thought to remain epiosomal, which should prevent the mutagenic disruption of the endogenous genes contained within the host mitochondrial genome. Although direct mitochondrial gene delivery offers exciting therapeutic potential, these potentially groundbreaking findings...
should be viewed as preliminary until they have been independently replicated by other research groups.

Gene therapy for DOA is being contemplated for affected patients harbouring truncating OPA1 mutations that result in haploinsufficiency. The strategy being developed involves the intravitreal injection of an AAV vector to deliver the wild-type OPA1 gene to surviving RGCs in a mouse model of DOA carrying the exon 27 (c.2708-2711delTTAG) mutation. Although conceptually simpler than mitochondrial gene therapy, further methodological refinements are needed to achieve stable transfection in a sufficient proportion of RGCs to result in a clinically meaningful effect (Dr Guy Lenaers, University of Montpelier, France, personal communication). It is important to emphasise that all the gene therapy approaches being envisaged for LHON and DOA are still at an early experimental stage of development, and further evidence of their safety and efficacy is needed before the initiation of human trials can be contemplated.

**Stem cells**

The therapeutic potential of stem cells is being investigated for a wide range of genetic eye disorders, and patients with mitochondrial optic neuropathies will frequently enquire with their clinicians whether they are likely to benefit from this form of treatment. Various poorly monitored ‘stem cell institutes’ worldwide are promoting the use of non-validated experimental protocols, and patients with LHON and DOA need to be carefully advised before embarking on multiple expensive courses of treatment with the possible associated biological risks. In sharp contrast to this unregulated parallel market, there are a number of well-established research programmes that are rigorously assessing the possible application of stem cell technology for optic nerve disorders. Two main paradigms are being explored, namely the generation and transplantation of RGCs, or the use of specific stem cell populations to generate trophic factors that promote RGC survival. The technique for generating and purifying mature RGCs from embryonic stem cells or induced pluripotent stem cells is still in its infancy. Another major complicating factor is how to integrate these differentiated RGCs into the retina and then force them to make the right topographical connection. At least in the short term, it is more likely that human-derived RGCs will provide the ideal tool for drug screening and understanding the basic mechanisms regulating RGC physiology in both health and disease states.

A proportion of patients with LHON will experience partial visual recovery, mostly within 1 year of being affected, but sometimes several years after the onset of the disease. It is therefore highly likely that a subpopulation of RGCs that survived the initial massive wave of apoptosis remain in a state of suspended animation awaiting two possible fates: a final push towards apoptosis or a gradual recovery of cellular function. These surviving RGCs are being targeted by oral neuroprotective agents, but mesenchymal stem cells are also an interesting modality based on promising data from experimental glaucoma and optic neuritis studies. Autologous mesenchymal stem cells can be isolated from bone marrow isolates, and they secrete several diffusible neurotrophic factors that are thought to promote neuronal survival in adverse metabolic and excitotoxic environments. Rather encouragingly, the injection of mesenchymal stem cells was well tolerated in a small proof-of-concept study of 10 patients with secondary progressive MS. Although not definitive, there was a suggestion of structural, functional, and physiological improvement in some visual end points suggestive of a neuroprotective effect. If confirmed, a similar strategy is entirely conceivable to halt or slow down the irreversible loss of RGCs in LHON and DOA.

**Prevention of germline transmission**

In vitro fertilisation (IVF) techniques are currently being developed to prevent female carriers of childbearing age from transmitting pathogenic mtDNA mutations to the next generation. Two different IVF options have been proposed, namely pronuclear transfer and metaphase II spindle transfer, to generate the so-called ‘three-parent embryos’ devoid of mutant mtDNA molecules. The aim of both techniques is to transfer the parental nuclear genetic material (devoid of the mother’s mutant mtDNA molecules) into a donor cytoplasm containing a normal wild-type mtDNA population. In the pronuclear technique, the male and female pronuclei are transferred post fertilisation into a mitochondrial donor zygote harbouring wild-type mtDNA. The alternative approach involves the transplantation of metaphase II spindles between unfertilised oocytes followed by intracytoplasmic sperm injection fertilisation. There seems to be no significant carryover of mutant mtDNA with both IVF approaches, and further work is currently underway in higher primates. Preventing the germline transmission of pathogenic mtDNA mutations is an exciting development, but a stringent series of safety tests will first need to be fulfilled. A significant proportion of manipulated zygotes showed abnormal development and other iatrogenic genetic consequences must be considered, including an increased risk of aneuploidy and epigenetic abnormalities. The manipulation of human embryos also entails a number of important
legal, religious, and ethical ramifications that will need to be addressed. The United Kingdom has been at the forefront of this important debate and an expert panel was recently convened by the Human Fertilisation and Embryology Authority (HFEA) to review the fertilisation methods that could be used to prevent mitochondrial disease. This public consultation indicated positive support for mitochondria replacement to take place, but subject to strict safeguards and careful regulation being adhered to (https://www.gov.uk/government/news/innovative-safeguards-and-careful-regulation-being-adhered-to accessed on 8 December 2013).

Conclusion
Translational research for mitochondrial optic neuropathies is entering an exciting accelerated phase providing renewed hopes to patients and their families that the inexorable visual loss associated with this group of disorders can at least be halted. We still need a better understanding of the pathological mechanisms underpinning the preferential vulnerability of RGCs in LHON and DOA, but these more basic studies are now proceeding in parallel with major national and transnational initiatives aimed at drug development and early phase clinical trials. The establishment of patient cohorts will provide a strong base to launch properly designed randomised, placebo-controlled trials and the prospective collection of long-term natural history data will be essential to identify the best outcome measures to detect a treatment benefit. The challenges of identifying and validating treatments for relatively rare inherited optic nerve disorders remain daunting, but 25 years after the birth of mitochondrial genetic medicine, tractable solutions are finally emerging.

Conflict of interest
The authors declare no conflict of interest.

Acknowledgements
PYWM is a Medical Research Council (MRC, UK) Clinician Scientist. PYWM also receives funding from Fight for Sight (UK) and the UK National Institute of Health Research (NIHR) as part of the Rare Diseases Translational Research Collaboration. PFC is a Wellcome Trust Senior Fellow in Clinical Science and a UK NIHR Senior Investigator who also receives funding from the MRC (UK), Parkinson’s UK, and the UK NIHR Biomedical Research Centre for Ageing and Age-related disease award to the Newcastle upon Tyne Hospitals NHS Foundation Trust.

References
1 Yu-Wai-Man P, Griffiths PG, Chinnery PF. Mitochondrial optic neuropathies—Disease mechanisms and therapeutic strategies. Prog Retinal Eye Res 2011; 30(2): 81–114.
2 Fraser JA, Bioussé V, Newman NJ. The Neuro-ophthalmology of mitochondrial disease. Survey Ophthalmol 2010; 55(4): 299–334.
3 Carelli V, Ross-Cisneros FN, Sadun AA. Mitochondrial dysfunction as a cause of optic neuropathies. Prog Retinal Eye Res 2004; 23(1): 33–89.
4 DiMauro S, Schon EA. Mechanisms of disease: Mitochondrial respiratory-chain diseases. New Engl J Med 2003; 348(26): 2656–2668.
5 Schapira AHV. Mitochondrial diseases. Lancet 2012; 379(9828): 1825–1834.
6 McFarland R, Taylor RW, Turnbull DM. A neurological perspective on mitochondrial disease. Lancet Neurol 2010; 9(8): 829–840.
7 Cree LM, Samuels DC, Lopes SCdS, Rajasimha HK, Wonnapinij P, Mann JR et al. A reduction of mitochondrial DNA molecules during embryogenesis explains the rapid segregation of genotypes. Nat Genet 2008; 40(2): 249–254.
8 Holt IJ, Harding AE, Morganhughes JA. Deletions of Muscle Mitochondrial-DNA in Patients with Mitochondrial Myopathies. Nature 1988; 331(6158): 717–719.
9 Wallace DC, Singh G, Lott MT, Hodge JA, Schurr TG, Lezza AMS et al. Mitochondrial-DNA Mutation Associated with Lebers Hereditary Optic Neuropathy. Science 1988; 242(4894): 1427–1430.
10 Hudson G, Chinnery PF. Mitochondrial DNA, polymerase-gamma and human disease. Hum Mol Genet 2006; 15: R244–R252.
11 Holt IJ, He J, Mao C-C, Boyd-Kirkup JD, Martinsson P, Sembongi H et al. Mammalian mitochondrial nucleoids: organizing an independently minded genome. Mitochondrion 2007; 7(5): 311–321.
12 Man PYW, Griffiths PG, Brown DT, Howell N, Turnbull DM, Chinnery PF. The epidemiology of Leber hereditary optic neuropathy in the North East of England. Am J Hum Genet 2003; 72(2): 333–339.
13 Spruijt L, Kolbach DN, de Coo RF, Plomp AS, Bauer NJ, Smeets HJ et al. Influence of mutation type on clinical expression of Leber hereditary optic neuropathy. Am J Ophthalmol 2006; 141(4): 676–682.
14 Puomila A, Hamalainen P, Kivioja S, Savontaus M-L, Koivumaki S, Haukkanen K et al. Epidemiology and penetrance of Leber hereditary optic neuropathy in Finland. Eur J Hum Genet 2007; 15(10): 1079–1089.
15 Macmillan C, Johns TA, Fu K, Shoubridge EA. Predominance of the T14484C mutation in French-Canadian families with Leber hereditary optic neuropathy is due to a founder effect. Am J Hum Genet 2000; 66(1): 332–335.
16 Sitarz KS, Chinnery PF, Yu-Wai-Man P. Disorders of the optic nerve in mitochondrial cytopathies: new ideas on pathogenesis and therapeutic targets. Curr Neurol Neurosci Rep 2012; 12(3): 308–317.
17 Nikoskelainen E. Ophthalmological findings in leber hereditary optic neuropathy, with special reference to the mtDNA mutations (vol 103, pg 504, 1996). Ophthalmology 1996; 103(7); 998.
18 Savini G, Barboni P, Valentino ML, Montagna P, Cortelli P, De Negri AM et al. Retinal nerve fiber layer evaluation by optical coherence tomography in unaffected carriers with
Leber’s hereditary optic neuropathy mutations. *Ophtalmology* 2005; 112(1): 127–131.

Sadun AA, Salomao SR, Berezovsky A, Sadun F, Denegri AM, Quiros PA et al. Subclinical carriers and conversions in Leber hereditary optic neuropathy: a prospective psychophysical study. *Trans Am Ophthalmol Soc* 2006; 104: 51–61.

Yu-Wai-Man P, Griffiths PG, Hudson G, Chinnery PF. Inherited mitochondrial optic neuropathies. *J Med Genet* 2009; 46(3): 145–158.

La Morgia C, Ross-Cisneros FN, Sadun AA, Hannibal J, Munarini A, Mantovani V et al. Melanopsin retinal ganglion cells are resistant to neurodegeneration in mitochondrial optic neuropathies. *Brain* 2010; 133: 2426–2438.

Kirkman MA, Korsten A, Leonhardt M, Dimitriadius K, De Coo IF, Kleopstock T et al. Quality of Life in Patients with Leber Hereditary Optic Neuropathy. *Invest Ophthalmol Vis Sci* 2009; 50(7): 3112–3115.

Harding AE, Sweeney MG, Govan GG, Riordaneva P. Pedigree Analysis in Leber Hereditary Optic Neuropathy Families with a Pathogenic MtDNA Mutation. *Am J Hum Genet* 1995; 57(1): 77–86.

Macmillan C, Kirkham T, Fu K, Allinson V, Andermann E, Chitayat D et al. Pedigree analysis of French Canadian families with T14484C Leber’s hereditary optic neuropathy. *Neurology* 1998; 50(2): 417–422.

Johns DR, Heher KL, Miller NR, Smith KH. Lebers hereditary optic neuropathy—clinical manifestations of the 14484 MUTATION. *Arch Ophthalmol* 1993; 111(4): 495–498.

Barboni P, Savini G, Valentino ML, La Morgia C, Bellusci C, De Negri AM et al. Leber’s hereditary optic neuropathy with childhood onset. *Invest Ophthalmol Vis Sci* 2006; 47(12): 5303–5309.

Ramos CDVE, Bellusci C, Savini G, Carbonelli M, Berezovsky A, Tamaki C et al. Association of optic disc size with development and prognosis of Leber’s hereditary optic neuropathy. *Invest Ophthalmol Vis Sci* 2009; 50(4): 1666–1674.

Harding AE, Sweeney MG, Miller DH, Mumford CJ, Kellarwood H, Menard D et al. Occurrence of a multiple sclerosis-like illness in women who have a Lebers hereditary optic neuropathy mitochondrial-dna mutation. *Brain* 1992; 115(4): 979–989.

Palace J. Multiple sclerosis associated with Leber’s hereditary optic neuropathy. *J Neurol Sci* 2009; 286(1-2): 24–27.

Pfeffer C, Burke A, Yu-Wai-Man P, Compston DA, Chinnery PF. Clinical features of MS associated with Leber hereditary optic neuropathy mtDNA mutations. *Neurology* 2013; 81(24): 2073–2081.

Chinnery PF, Andrews RM, Turnbull DM, Howell N. Leber hereditary optic neuropathy: Does heterolysis influence the inheritance and expression of the G11778A mutation? *Am J Med Genet* 2001; 98(3): 235–243.

Gomez-Duran A, Pacheu-Grau D, Lopez-Gallardo E, Diez-Sanchez C, Montoya J, Lopez-Perez MJ et al. Unmasking the causes of multifactorial disorders: OXPHOS differences between mitochondrial haplogroups. *Hum Mol Genet* 2010; 19(17): 3343–3353.

Gomez-Duran A, Pacheu-Grau D, Martinez-Romo I, Lopez-Gallardo E, Lopez-Perez MJ, Montoya J et al. Oxidative phosphorylation differences between mitochondrial DNA haplogroups modify the risk of Leber’s hereditary optic neuropathy. *Biochim Biophys Acta* 2012; 1822(8): 1216–1222.

Hudson G, Carelli V, Spruijt L, Gerards M, Movbray C, Achilli A et al. Clinical expression of Leber hereditary optic neuropathy is affected by the mitochondrial DNA-haplogroup background. *Am J Hum Genet* 2007; 81(2): 228–233.

Ji Y, Zhang AM, Jia X, Zhang Y-P, Xiao X, Li S et al. Mitochondrial DNA Haplogroups M7b1 ‘2 and M8a affect clinical expression of Leber hereditary optic neuropathy in Chinese families with the m.11778G -> A mutation. *Am J Hum Genet* 2008; 83(6): 760–768.

Tharapan P, Chuenkongkaew WL, Luantrakool K, Sanpachudayan T, Suktittipat B, Suphavilai R et al. Mitochondrial DNA haplogroup distribution in pedigrees of Southeast Asian G11778A Leber hereditary optic neuropathy. *J Neuro-Ophthalmol* 2006; 26(4): 264–267.

Bu XD, Rotter JL. X-chromosome-linked and mitochondrial gene-control of Leber hereditary optic neuropathy—evidence from segregation analysis for dependence on X-chromosome inactivation. *Proc Natl Acad Sci USA* 1991; 88(18): 8198–8202.

Hudson G, Keers S, Man PYW, Griffiths P, Huoponen K, Savontaus ML et al. Identification of an X-chromosomal locus and haplotype modulating the phenotype of a mitochondrial DNA disorder. *Am J Hum Genet* 2005; 77(6): 1086–1091.

Shankar SP, Fingert JH, Carelli V, Valentino ML, King TM, Daiger SP et al. Evidence for a novel X-linked modifier locus for leber hereditary optic neuropathy. *Ophthalmic Genet* 2008; 29(1): 17–24.

Ji Y, Jia X, Li S, Xiao X, Guo X, Zhang Q. Evaluation of the X-linked modifier loci for Leber hereditary optic neuropathy with the G11778A mutation in Chinese. *Mol Vision* 2010; 16(47): 416–424.

Giordano C, Montopoli M, Perli E, Orlandi M, Fantin M, Ross-Cisneros FN et al. Oestrogens ameliorate mitochondrial dysfunction in Leber’s hereditary optic neuropathy. *Brain* 2011; 134: 220–234.

Kirkman MA, Yu-Wai-Man P, Korsten A, Leonhardt M, Dimitriadius K, De Coo IF et al. Gene-environment interactions in Leber hereditary optic neuropathy. *Brain* 2009; 132: 2317–2326.

Sadun AA, WIN PH, Ross-Cisneros FN, Walker SO, qCarelli V. Leber’s hereditary optic neuropathy differentially affects smaller axons in the optic nerve. *Trans Am Ophthalmol Soc* 2000; 98: 223–235.

Carelli V, Rugolo M, Sgarbi G, Chelli A, Zanna C, Baracca A et al. Bioenergetics shapes cellular death pathways in Leber’s hereditary optic neuropathy: a model of mitochondrial neurodegeneration. *Biochim Biophys Acta* 2004; 1658(1-2): 172–179.

Lin CS, Sharpley MS, Fan W, Waymire KG, Sadun AA, Carelli V et al. Mouse mtDNA mutant model of Leber hereditary optic neuropathy. *Proc Natl Acad Sci USA* 2012; 109(49): 20065–20070.

Yu-Wai-Man P, Chinnery PF. Dominant optic atrophy—Novel OPA1 mutations and revised prevalence estimates. *Ophthalmology* 2013; 120(8): 1712.

Alexander C, Votruba M, Pesch UE, Thiselton DL, Mayer S, Moore A et al. Identification of the gene responsible for dominant optic atrophy (OPA1) on chromosome 3q28. *Am J Hum Genet* 2000; 67(4): 40.

Deletrée C, Lenoars G, Griffin JM, Gigarel N, Lorenzo C, Belenguer P et al. Nuclear gene OPA1, encoding a mitochondrial dynamin-related protein, is mutated in dominant optic atrophy. *Nat Genet* 2000; 26(2): 207–210.
Ferre M, Bonneau D, Mileda D, Chevrollier A, Verny C, Dollfus H et al. Molecular screening of 980 cases of suspected hereditary optic neuropathy with a report on 77 novel OPA1 mutations. *Hum Mut* 2009; 30(7): E692–E705.

Marchbank NJ, Craig JE, Leek JP, Toohey M, Churchill AJ, Markham AF et al. Deletion of the OPA1 gene in a dominant optic atrophy family: evidence that haploinsufficiency is the cause of disease. *J Med Genet* 2002; 39(8): e47.

Yu-Wai-Man P, Griffiths PG, Burke A, Sellar PW, Clarke MP, Gnanaraj L et al. The prevalence and natural history of dominant optic atrophy due to OPA1 mutations. *Ophthalmology* 2010; 117(5): 1538–1546, 1546.e1.

Yu-Wai-Man P, Bailie M, Atawar A, Chimney PF, Griffiths PG. Pattern of retinal ganglion cell loss in dominant optic atrophy due to OPA1 mutations. *Eyes* 2011; 25(5): 597–601.

Yu-Wai-Man P, Griffiths PG, Gorman GS, Lourenco CM, Wright AF, Auer-Grumbach M et al. Multi-system neurological disease is common in patients with OPA1 mutations. *Brain* 2010; 133: 771–786.

Lenaers G, Olichon A, Deletrée C, Hamel C, Belenguer P. Functions and dysfunctions of the human dynamin OPA1. *Biochim Biophys Acta* 2004; 1657: 37.

Chevrollier A, Guillot V, Loiseau D, Gueguen N, de Crescenzo M-AP, Verny C et al. Hereditary optic neuropathies share a common mitochondrial coupling defect. *Ann Neurol* 2008; 63(6): 794–798.

Zanna C, Ghelli A, Forcelli AM, Karbowski M, Youle RJ, Schimpf S et al. OPA1 mutations associated with dominant optic atrophy impair oxidative phosphorylation and mitochondrial fusion. *Brain* 2008; 131: 352–367.

Frezza C, Cipolat S, de Brito OM, Micaroni M, Beznoussenko GV, Rudka T et al. OPA1 controls apoptotic cristae remodeling independently from mitochondrial fusion. *Cell* 2006; 126(1): 177–189.

Kushnareva YE, Gerencser AA, Bossy B, Ju WK, White KE et al. Loss of OPA1 disturbs cellular calcium homeostasis and sensitizes for excitotoxicity. *Cell Death Differ* 2013; 20(2): 373–386.

Yu-Wai-Man P, Sitzar KS, Samuels DC, Griffiths PG, Reave AK, Bindoff LA et al. OPA1 mutations cause cytochrome c oxidase deficiency due to loss of wild-type mtDNA molecules. *Hum Mol Genet* 2010; 19(15): 3043–3052.

Bailie M, Votruba M, Griffiths PG, Chimney PF, Yu-Wai-Man P. Visual and psychological morbidity among patients with autosomal dominant optic atrophy. *Acta ophthalmol* 2013; 91(5): e413–e414.

Pfeffer G, Majamaa K, Turnbull DM, Thorburn D, Chimney PF. Treatment for mitochondrial disorders. *Cochrane Database Syst Rev* 2012; 4: CD004426.

Pfeffer G, Horvath R, Klostock T, Mootha VK, Dollfus H et al. The prevalence and natural history of dominant optic atrophy due to OPA1 mutations. *Hum Mut* 2009; 30(7): E692–E705.

Sadun AA, Chiciani CE, Ross-Cisneros FN, Barboni P, Thoennes M, Shadrer WD et al. Effect of EPI-743 on the clinical course of the mitochondrial disease Leber hereditary optic neuropathy. *Arch Neurol* 2012; 69(3): 331–338.

Mashima Y, Hida Y, Oguchi Y. Remission of Lebers hereditary optic neuropathy with Idebenone. *Lancet* 1992; 340(8815): 368–369.

Mashima Y, Kigasawa K, Wakakura M, Oguchi Y. Do idebenone and vitamin therapy shorten the time to achieve visual recovery in Leber hereditary optic neuropathy? *J Neuro-Ophthalmol* 2000; 20(3): 166–170.

Barnils N, Mesa E, Munoz S, Ferrer-Artola A, Arruga J. Response to idebenone and multivitamin therapy in Leber’s hereditary optic neuropathy. *Arch de la Sociedad Espanola de Oftalmol* 2007; 82(6): 377–380.

Klostock T, Metz G, Yu-Wai-Man P, Buechner B, Gallenmueller C, Bailie M et al. Persistence of the treatment effect of idebenone in Leber’s hereditary optic neuropathy. *Brain* 2013; 136: e230.

Corelli V, La Morgia C, Valentino ML, Rizzo G, Carbonelli M, De Negri AM et al. Idebenone treatment in Leber’s hereditary optic neuropathy. *Brain* 2011; 134: e188.

Lim G, Martin KR. Current prospects in optic nerve protection and regeneration: sixth ARVO/Pfizer Ophthalmics Research Institute Conference. *Invest Ophthalmol Vis Sci* 2011; 52(8): 5941–5954.

Newman NJ, Biouss V, David R, Bhatti MT, Hamilton SR, Farris BK et al. Prophylaxis for second eye involvement in Leber hereditary optic neuropathy: An open-labeled, nonrandomized multicenter trial of topical brimonidine purite. *Am J Ophthalmol* 2005; 140(3): 407–415.

Thouin A, Griffiths PG, Hudson G, Chimney PF, Yu-Wai-Man P. Raised intraocular pressure as a potential risk factor for visual loss in Leber hereditary optic neuropathy. *PloS ONE* 2013; 8(5): e63446.

Wong A, Cortopassi G. mtDNA Mutations confer cellular sensitivity to oxidant stress that is partially rescued by calcium depletion and cyclosporin A. *Biochem Biophys Res Commun* 1997; 239(1): 139–145.

Porcelli AM, Angelini A, Ghelli A, Mariani E, Martinuzzi A, Corelli V et al. Respiratory complex I dysfunction due to mitochondrial DNA mutations shifts the voltage threshold for opening of the permeability transition pore toward resting levels. *J Biol Chem* 2009; 284(4): 2045–2052.

Malik A, Golnik K. Hyperbaric oxygen therapy in the treatment of radiation optic neuropathy. *J Neuro-Ophthalmol* 2012; 32(2): 128–131.

Desmet KD, Paz DA, Corry JJ, Eells JT, Wong-Riley MTT, Henry MM et al. Clinical and experimental applications of NIR-LED photobiomodulation. *Photomed Laser Surg* 2006; 24(2): 121–128.

Barboni P, Valentino ML, La Morgia C, Carbonelli M, Savini G, De Negri A et al. Idebenone treatment in patients with OPA1-mutant dominant optic atrophy. *Brain* 2013; 136: e231.

Davies VJ, Hollins AJ, Piechota MJ, Yip W, Davies JR, White KE et al. Opa1 deficiency in a mouse model of autosomal dominant optic atrophy impairs mitochondrial morphology, optic nerve structure and visual function. *Hum Mol Genet* 2007; 16(11): 1307–1318.

Levin LA. Axonal loss and neuroprotection in optic neuropathies. *Can J Ophthalmol* 2007; 42(3): 403–408.

Pesc V, Fragasso F, Cassano P, Lezza AM, Cantatore P, Gadala MN. Acetyl-L-carnitine supplementation to old rats partially reverses the age-related mitochondrial decay of soleus muscle by activating peroxisome proliferator-activated receptor gamma coactivator-1 alpha-dependent mitochondrial biogenesis. *Rejuven Res* 2010; 13(2-3): 148–151.
82 Pesce V, Nicassio L, Fracasso F, Musico C, Cantatore P, Gadaleta MN. Acetyl-L-carnitine activates the peroxisome proliferator-activated receptor-gamma coactivators PGC-1 alpha/PGC-1-beta-dependent signaling cascade of mitochondrial biogenesis and decreases the oxidized peroxiredoxins content in old rat liver. *Rejuvenation Res* 2012; **15**(2): 136–139.

83 Guy J. New therapies for optic neuropathies: development in experimental models. *Curr Opin Ophthalmol* 2000; **11**(6): 421–429.

84 Guy J, Qi XP, Pallotti F, Schon EA, Manfredi G, Carelli V et al. Rescue of a mitochondrial deficiency causing Leber hereditary optic neuropathy. *Ann Neurol* 2002; **52**(5): 534–542.

85 Ellouze S, Augustin S, Bouaita A, Bonnet C, Simonutti M, Forster V et al. Optimized allotopic expression of the human mitochondrial ND4 prevents blindness in a rat model of mitochondrial dysfunction. *Am J Hum Genet* 2008; **83**(3): 373–387.

86 Guy J, QI X, Koilkonda RD, Arguello T, Chou T-H, Ruggeri M et al. Efficiency and safety of AAV-mediated gene delivery of the human ND4 complex I subunit in the mouse visual system. *Invest Ophthalmol Vis Sci* 2009; **50**(9): 4205–4214.

87 Qi X, Sun L, Hauswirth WW, Lewin AS, Guy J. Use of mitochondrial antioxidant defenses for rescue of cells with a Leber hereditary optic neuropathy-causing mutation. *Arch Ophthalmol* 2007; **125**(2): 268–272.

88 See BB, Nakamur-Ogiso E, Plotte TR, Matsuno-Yagi A, Yagi T. *In vivo* complementation of complex I by the yeast Ndi1 enzyme—Possible application for treatment of Parkinson disease. *J Biol Chem* 2006; **281**(20): 14250–14255.

89 Iyer S, Bergquist K, Young K, Gnaiger E, Rao RR, Bennett JP Jr. Mitochondrial gene therapy improves respiration, biogenesis, and transcription in G11778A Leber's hereditary optic neuropathy and T8993G Leigh's syndrome cells. *Hum Gene Ther* 2012; **23**(6): 647–657.

90 Yu H, Koilkonda RD, Chou T-H, Porciatti V, Ozdemir SS, Chiodo V et al. Gene delivery to mitochondria by targeting modified adenoassociated virus suppresses Leber's hereditary optic neuropathy in a mouse model. *Proc Natl Acad Sci USA* 2012; **109**(20): E1238–E1247.

91 Yu H, Mehta A, Wang G, Hauswirth WW, Chiodo V, Boye SL et al. Next-generation sequencing of mitochondrial targeted AAV transfer of human ND4 in mice. *Mol Vision* 2013; **19**: 1482–1491.

92 Sarzi E, Angebault C, Seveno M, Guergen N, Chaix B, Bielicki G et al. The human OPA1(delTTAG) mutation induces premature age-related systemic neurodegeneration in mouse. *Brain* 2012; **135**: 3599–3613.

93 Marchetti V, Krobne TU, Friedlander DF, Friedlander M. Stemming vision loss with stem cells. *J Clin Invest* 2010; **120**(9): 3012–3021.

94 Dahlmann-Noor A, Vijay S, Jayaram H, Limb A, Khaw PT. Recent approaches and future prospects for stem cell rescue and regeneration of the retina and optic nerve. *Can J Ophthalmol* 2010; **45**(4): 333–341.

95 Johnson TV, Bull ND, Hunt DP, Marina N, Tomarev SI, Martin KR. Neuroprotective effects of intravitreal mesenchymal stem cell transplantation in experimental glaucoma. *Invest Ophthalmol Vis Sci* 2010; **51**(4): 2051–2059.

96 Connick P, Kolappan M, Crawley C, Webber DJ, Patani R, Michell AW et al. Autologous mesenchymal stem cells for the treatment of secondary progressive multiple sclerosis: an open-label phase 2a proof-of-concept study. *Lancet Neurol* 2012; **11**(2): 150–156.

97 Craven L, Elson JL, Irving L, Tuppen HA, Lister LM, Greggains GD et al. Mitochondrial DNA disease: new options for prevention. *Hum Mol Genet* 2011; **20**: R168–R174.

98 Craven L, Tuppen HA, Greggains GD, Harbottle SJ, Murphy JL, Cree LM et al. Pronuclear transfer in human embryos to prevent transmission of mitochondrial DNA disease. *Nature* 2010; **465**(7294): U82–U89.

99 Tachibana M, Sparman M, Sritanaudomchai H, Ma H, Clepper L, Woodward J et al. Mitochondrial gene replacement in primate offspring and embryonic stem cells. *Nature* 2009; **461**(7262): 367–372.

100 Tachibana M, Amato P, Sparman M, Woodward J, Sanchis DM, Ma H et al. Towards germline gene therapy of inherited mitochondrial diseases. *Nature* 2013; **493**(7434): 627–631.

This work is licensed under a Creative Commons Attribution 3.0 Unported License. To view a copy of this license, visit http://creativecommons.org/licenses/by/3.0/