Research Article

Evaluation of Nematocidal Action against Caenorhabditis elegans of Essential Oil of Flesh Fingered Citron and Its Mechanism

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Essential oils from flesh fingered citrons were obtained by mechanical pressing extraction under optimal conditions. Thirty-three components, representing 78.25% of the total oil, were identified by gas chromatography-mass spectrometry (GC-MS). In this study, we explored the toxicity of the essential oil of flesh fingered citrons and the associated regulatory mechanism using Caenorhabditis elegans as a model organism. The mortality experiment showed that the LD_{50} value is 1.48 mg/ml. Oxidative stress experiments showed that essential oils of flesh fingered citrons can reduce the antioxidation of nematodes and can shorten their lifespan. In survival rate experiments for verification, d-limonene had a nematocidal effect by destroying the waterproof protective layer on the surface of the worm. β-Pinene can interfere with the normal growth of nematodes according to behavioral experiments, and reproduction experiments showed that α-pinene has very strong nematocidal properties. c-Terpinene also has good nematocidal activity. The results demonstrated that the essential oil of flesh fingered citron has great potential to act as a new nematicide.

1. Introduction

Controlling insect pests and nematodes by synthetic nematicides is expensive and environmentally harmful. The search for naturally occurring, cheaper, and more environmentally friendly biocontrol agents is ongoing throughout the world [1]. Currently, plant chemicals are being increasingly used as effective botanical nematicides in the form of extracts and essential oils because synthetic nematicides cause many negative effects on the environment [2]. Therefore, we have explored natural plant sources for pest management agents. Due to the infrequent presence and inconsistent efficacy of synthetic pesticides, plant nematicides may be suitable alternatives to disrupt the physiological processes of target nematodes in addition to direct mortality [3, 4]. Because of their nematocidal toxicity, essential oils are considered a possible alternative [5]. These oils have been found to have significantly negative impacts on nematode survival, reproduction, development, behavior, and metabolic pathways [6, 7] and have been studied as nematicidal, larvicidal, and repellent agents [8]. Essential oils are plant-based economically important secondary metabolites [9]. Citrus essential oils are extracted from citrus peels [10] that produce characteristic odors and have been widely applied to the food (beverage, sweets, and so on), cosmetics, and pharmaceutical industries mainly because of their fragrance and flavor [11]. The essential oils of the flesh fingered citron (Citrus medica cv. sarcodactylis) are used as an expectorant to treat cough, asthma, and inflammation, which is generally produced by mechanical extraction like all citrus oils [12]. Flesh fingered citron essential oil can be divided into two fractions, with the volatile fraction (93%–96%) containing monoterpenes and sesquiterpenes, such as d-limonene, α- and β-pinene, γ-terpinene [13], and other oxygenated components (4%–7%). It is an essential oil of intermediate to low odor and flavor intensity.
Terpenoids that come from this essential oil are important nematocidal substances and have strong biological activity. Thus, this essential oil has been used in agricultural nematicide research. Different components like monoterpenoids (e.g., α-pinene) are responsible for nematocidal properties and physiological disruptions of nematodes [15, 16]. These compounds can be used as antifeedants, digestion and metabolism breakers, acetylcholinesterase inhibitors, and cell death inducers by acting as oxidizing agents, which can cause ionic leakage of cells and degeneration of mitochondria and proteins [17–22]. Limonene, a monocyclic monoterpenene, is famous for its citrus-like olfactory properties [23]. Limonene is also an important precursor for many herbal and pharmaceutical compounds such as perillyl alcohol, carvone, and menthol [24, 25]. D-Limonene is the most widely used isomer in spices, such as perillyl alcohol, carvone, and menthol [24, 25]. D-Limonene is the most widely used isomer in spices, medicines, and pesticides [26]. As a nematicide, it can break the waterproof protective layer of the worm’s body surface and has strong penetrating properties so that worms die by suffocation, reducing their survival rate.

Many studies have employed the nematode Caenorhabditis elegans as an animal model to study the effects of essential oils [27]. C. elegans is also used as an epidemiological model for stress and aging because of its small size, relatively short lifecycle, and the clear impact of genetic and environmental factors that affect the lifespan [28].

In this study, we used C. elegans as an in vivo model to determine the mortality effect of essential oils. We examined the mechanism of action of essential oils and the main individual components on C. elegans mortality and tested our hypotheses through a series of experiments on oxidative stress, lifespan, reproduction, and survival rate.

2. Materials and Methods

2.1. Plant Materials and C. elegans. Mature fruits of the flesh fingered citron were harvested in November 2017 from a local orchard in Jinhua City, Zhejiang Province, P.R. China. Wild-type Caenorhabditis elegans (N2) and food E. coli OP50 strains were obtained from the Caenorhabditis Genetics Center.

2.2. Chemicals. D-Limonene (95%), α-pinene (98%), β-pinene (≥95%), and γ-terpinene (95%) were obtained from Macklin (China). Cholesterol, calcium chloride, sodium chloride, potassium dihydrogen phosphate, disodium phosphate, magnesium sulfate, and absolute ethanol were purchased from Sinopharm Chemical Reagent Co., Ltd. (China). Agar powder was purchased from Sangon Biotech Co., Ltd. (Shanghai, China). Potassium dichromate (K₂Cr₂O₇), di-methyl sulfoxide (DMSO), and 5-fluoro-2-deoxyuridine (FUDR) were purchased from Sigma (USA). Peptone (LP0037), tryptone (LP0042), and yeast extract (LP0021) were purchased from OXOID (UK). All other chemicals and solvents used were of analytical grade.

2.3. Essential Oil Preparation. The mechanical pressing method is suitable for the extraction of easily burnt raw materials such as citrus and lemon because it is easy to maintain the structure and function of essential oils. Flesh fingered citron fruits were first screened and peeled, saturated in 3% (w/v) calcium chloride or calcium hydroxide for a specified time (4 h) to remove pectin, and then cleaned and leached with distilled water. The soaked flavedo (50 g) was weighed and turned into juice using a juice extractor. The juice was supplemented with sodium chloride (0.25%) and stirred until the sodium chloride completely dissolved. After the solution was separated into layers, the oil-water mixed liquid obtained after filtering was centrifuged, and the upper oil layer of the mixture was separated and dried with anhydrous sodium sulfate [29]. The extraction rate of essential oils was about 0.3%.

The purification was carried out using a molecular distillation method carried out by a laboratory rubbed molecular distillation device (Pope, USA). At 45°C melted in the water bath, essential oil was drawn into a feed bottle and degassed using a short-path evaporator. The distillation temperature, distillation pressure, wiping roller speed, feed flow rate, and cooling water temperature were set to 70°C, 50 Pa, 200 rpm, 4 mL/min, and 10°C, respectively. Then, the feeding was turned on, and the degassed feed liquid immediately became a very thin film and spread onto the evaporation surface. A different number of volatile components evaporated as the liquid flowed down the wall through the heating zone. More volatile components concentrated onto the closely positioned internal condensing surface and became the distillates, while the less volatile components flowed down along the cylinder and became the residues. After distillation, the distillate and residue fractions were weighed and transferred to the containers. They were stored at 4°C until analysis [30]. The purification rate of essential oils was about 85%.

2.4. GC and GC-MS Analysis. A fused silica capillary column (HP-5MS) was used for GC analysis with a Hewlett-Packard 6890 gas chromatograph (5% phenyl-methylsiloxane, 60 m × 0.25 mm, film thickness 0.25 µm, Agilent Technologies, USA). The oven temperature was set to rise from an initial temperature of 50–100°C at a rate of 10°C/min and held there for 10 min and then raised to 140°C at a rate of 3°C/min. It was kept at 140°C for 10 min and finally heated to 230°C at a rate of 2°C/min and held there for 10 min. The split injection mode with a split ratio of 1:10 was employed. Quantification of components in the essential oil was obtained based on the GC peak areas using both the internal standard method and relative response factors. GC/MS analysis was performed on a Hewlett-Packard 6890 gas chromatograph coupled with a Hewlett-Packard mass-selective detector 5973N quadrupole mass spectrometer (MS) (Agilent Technologies, USA). All GC experimental parameters were the same as described for GC-FID analysis. The MS was operated in the electron impact ionization (EI) mode with an ionization energy of 70 eV. The temperatures of the MS source and MS quadrupole were 280 and 150°C, respectively. The constituents of the essential oil were identified based on computer matching with the Wiley/NIST
library and comparisons of retention indices with those reported in the literature [31].

2.5. C. elegans Strains, Handling, and Mortality. The nematodes used in this study were Bristol wild-type N2 [32]. C. elegans was grown and assayed on NGM agar plates carrying *E. coli* OP50 at 20°C. The phytochemicals were dissolved in 0.1% DMSO. All chemicals in the NGM plate and liquid are expressed in the final concentrations.

To investigate the nematocidal effect of the flesh fingered citron essential oil, a survival assay was deployed. In this assay, the worms were treated with essential oils at varying concentrations of 0, 0.5, 1, 1.5, and 2 mg/ml. *C. elegans* mortality was determined after 24 h exposure, and LD$_{50}$ was calculated [33].

2.6. Mechanism Experiments of Essential Oil Nematicide in *C. elegans*

2.6.1. Oxidative Stress Resistance Assays for *C. elegans*. Oxidative stress assays were performed essentially as previously described [34]. Wild-type N2 *C. elegans* was synchronized to the L4 period and pretreated with 0, 0.5, 1, 1.5, and 2 mg/ml essential oil and 0.1% DMSO as a solvent control for 48 h, then exposed to K$_2$Cr$_2$O$_7$ (8 μm, 10 mmol/L), and finally cultured for 48 h. Worms were observed every 4 h, and their survival was determined by the touch-provoked movement [35]. At least three independent biological replications were performed.

2.6.2. Lifespan Assays on *C. elegans*. L4 larval *C. elegans* (wild-type N2) were transferred to 96-well plates with S-medium to inhibit progeny development in the presence of 0, 0.5, 1, 1.5, and 2 mg/ml essential oil of flesh fingered citron dissolved in 0.1% DMSO. We evaluated whether the essential oil influenced the lifespan of *C. elegans* by comparing the lifespans of untreated (control) and essential oil-treated wild-type N2 *C. elegans*. Live and dead *C. elegans* was counted every other day (from the first day of adulthood) until all *C. elegans* had died as determined by the touch-provoked movement. At least three independent biological replications were performed.

2.6.3. Survival Rate of *C. elegans*. As a verification experiment, L1 stage larvae were treated with the essential oil solution for 3 days to the L4 stage and exposed to air for half an hour, and then the survival rate was determined. We performed at least three independent biological replications.

2.6.4. Behavioral Experiments on *C. elegans*. Body bends and head swings indicate alternating muscle contractions. Synchronized L4 adults were transferred to an empty NGM board and allowed to be free for at least 1 min. The number of body flexions within 20 sec and the number of head swings with a drop of distilled water within 1 min were determined. We performed at least three independent biological replications.

2.6.5. Reproductive Assays of *C. elegans*. The effect of essential oil on fecundity was assessed using age-synchronized N2 wild-type worms [36]. The total progeny yield of each worm after exposure to essential oil (0, 0.5, 1, 1.5, 2 mg/ml) was assessed according to standard protocols. Essential oil-treated/untreated L4 larvae were transferred to fresh preparation/untreatment plates until reproduction terminated. Three days later, the number of nematodes in the two plates was determined. We performed at least three independent biological replications.

2.7. Mortality Rate Inflicted by Four Key Components in Essential Oil. The experiments were similar to those in Section 2.5. Four major components, D-limonene, γ-terpinene, α-pinene, and β-pinene (5 mM each), were used to treat *C. elegans*. At least three independent biological replications were performed using at least 100 *C. elegans* strains.

2.8. Statistical Analysis. For the oxidative stress assays and lifespan assays, we used GraphPad Prism (GraphPad Software). Survival curves resulting in $p$ values <0.05 were considered significantly different relative to untreated controls. Statistics were analyzed using SPSS Statistics Software (SPSS). Results are expressed as the mean ± standard error of the mean (SEM). Univariate analysis of variance was performed using the ANOVA algorithm, while the LSD test was used for minimally significant difference analysis. *Post hoc* tests to determine the significance and interaction between the factors of the lifespan of *C. elegans* were performed with a two-factor variation analysis (two-way ANOVA). Differences were considered significant at the $P < 0.05$, $P < 0.01$, and $P < 0.001$ levels [35].

3. Results and Discussion

3.1. GC-MS. Qualitative and quantitative analytical results of the chemical composition of essential oil of flesh fingered citron by GC/MS are listed in Table 1. Quantification of analytes was carried out by peak area normalization and expressed in terms of the percentage of the total peak area to facilitate comparison with data reported in the literature. As shown in the table, 33 compounds representing 78.25% of the total peak area were identified. The essential oil consisted mainly of monoterpene hydrocarbons, and the main ones were d-limonene (43.39%), γ-terpinene (18.57%), α-pinene (2.88%), β-pinene (2.69%), β-myrcene (1.77%), (Z)-β-ocimene (1.46%), and α-thujene (1.31%). The components of EO in the present study had a composition similar to that in the previous reports about other species of citrus fruits [37, 38].
linear for each dose concentration range (Figure 1). When the concentration range of the essential oil is 1.0–1.5 mg/ml, the equation can be obtained from the linear relationship: \( y = 46x - 18 \), so the LD\(_{50}\) value is 1.48 mg/ml.

### 3.3. Mechanism Experiments

#### 3.3.1. Essential Oil Affects Oxidative Stress Resistance of *C. elegans*.

It has been widely reported that many kinds of citrus essential oils from natural plants exhibit antioxidant, antiradical, and antimicrobial properties [34]. K\(_2\)Cr\(_2\)O\(_7\) is an oxidative stress inducer because of its heavy metal oxidant that produces an intracellular oxidation-reduction reaction with \( \text{O}_2 \) production and is commonly used to generate intracellular oxidation in *C. elegans* [39]. Pretreatment with different concentrations of the essential oil of flesh fingered citron in 0.1% dimethyl sulfoxide (DMSO) had a nematocidal effect on *C. elegans* via oxidative stress (Figure 2).

Essential oils contain a variety of compounds that may unleash different biological activities. Moreover, higher concentrations of the extract may be toxic, and the higher the concentration, the greater the toxicity, thus reducing the antioxidation effect of *C. elegans*.  

#### 3.3.2. Essential Oil of Flesh Finger Citron Affects the Lifespan of Wild-Type *C. elegans*.

The *C. elegans* model system was used to assess the effects of essential oils on aging. The increase in free radical production and the destruction of the redox balance lead to oxidative stress, causing aging and age-related diseases [40]. Senescence is well illustrated in *C. elegans* and shares homologous mechanisms with primates and humans [41–43]. The experimental results (Figure 3) show that the higher the concentration of essential oil, the more obvious the nematocidal effect, and the lower the concentration of essential oil, the longer the survival of *C. elegans*. This indicates that the essential oil of flesh fingered citron can reduce the lifespan of *C. elegans*.

#### 3.3.3. Survival Rate.

The formation of allergic oxidation products like D-limonene oxide and dD-limonene 2-hydroperoxide has been reported [44] for oxidized citrus oil (rich in d-limonene), a frequent skin sensitizer. D-Limonene can break the waterproof protective layer of the worm’s body surface and has strong penetrating properties so that worms die by suffocation. It can be seen from the graph (Figure 4) of the survival rate of *C. elegans* that the presence of essential oils of flesh fingered citron reduces the survival rate of nematodes.

#### 3.3.4. Slowed Behavioral Activity in *C. elegans*.

Terpenes are the largest group of phytochemicals resulting from secondary metabolism [45]. Some components (e.g., \( \beta \)-pinene)

### Table 1: Major components from the essential oil of fingered citron.

| No. | Constituent | Retention index | Percentage |
|-----|-------------|-----------------|------------|
| **Monoterpene hydrocarbons** |
| 1   | \( \alpha \)-Thujene | 919             | 1.31       |
| 2   | \( \alpha \)-Pinene | 931             | 2.88       |
| 3   | \( \beta \)-Pinene | 980             | 2.69       |
| 4   | \( \alpha \)-Phellandrene | 1005           | 0.26       |
| 5   | \( \delta \)-3-Carene | 1015           | 0.30       |
| 6   | \( \alpha \)-Terpinene | 1019           | 0.23       |
| 7   | D-Limonene | 1032            | 43.39      |
| 8   | \( \gamma \)-Terpinene | 1066           | 18.57      |
| 9   | Terpinolene | 1087            | 1.44       |
| 10  | \( \beta \)-Myrcene | 987             | 1.77       |
| 11  | (\( Z \))-\( \beta \)-Ocimene | 1045           | 1.46       |
| 12  | 2-Carene | 1018             | 0.31       |
| 13  | \( \beta \)-Phellandrene | 964            | 0.38       |
| **Sesquiterpene hydrocarbons** |
| 14  | \( \beta \)-Caryophyllene | 1425          | 0.19       |
| 15  | (\( S \))-\( \beta \)-Bisabolene | 1485        | 0.46       |
| 16  | \( \alpha \)-Bergamotene | 1437          | 0.33       |
| 17  | Germacrene D | 1485            | 0.032      |
| **Oxygenated monoterpenes** |
| 18  | Linalool | 1095             | 0.13       |
| 19  | trans-Limonene oxide | 1139         | 0.12       |
| 20  | Terpin-4-ol | 1183           | 0.28       |
| 21  | \( \alpha \)-Terpineol | 1196          | 0.43       |
| 22  | Neral | 1235             | 0.23       |
| 23  | Nerol | 1229             | 0.051      |
| 24  | Geranial | 1263            | 0.28       |
| 25  | Neryl acetate | 1349          | 0.039      |
| 26  | Geranyl acetate | 1368         | 0.032      |
| 27  | Geraniol | 1252            | 0.28       |
| 28  | cis-Limonene oxide | 1138         | 0.079      |
| 29  | (\( Z \))-Carveol | 1222          | 0.061      |
| 30  | Thymol | 1292             | 0.017      |
| **Oxygenated sesquiterpenes** |
| 31  | Caryophyllene oxide | 1612          | 0.13       |
| 32  | Spathulenol | 1604           | 0.061      |
| **Nonisoprenoid components** |
| 33  | Decanal | 1200             | 0.032      |
| Total |                                   |               | 78.25      |

*Relative to C\(_5\)–C\(_{25}\)*-\( \alpha \)-alkanes calculated from a nonpolar HP-5MS™ capillary column. *Based on FID peak area normalization.
are highly toxic and interfere with the normal growth of *C. elegans* and affect behavior. It can be seen from the results (Table 2) that different concentrations of essential oil have a strong inhibitory effect on the number of body bends of nematodes and reduce the number of head swings. The essential oil of flesh fingered citron therefore affects the behavior of nematodes in addition to its nematocidal action.

3.3.5. Reproduction Rate Reduced in *C. elegans*. As an essential oil component, γ-terpinene has good insecticidal activity [46] and is highly toxic. α-Pinene is a strong repellant [47], has nematocidal characteristics, and induces physiological changes in some nematodes, and through the production of oxidizing compounds, it acts as an anti-feedant, digestive and metabolic disruptor, acetylcholinesterase inhibitor, and cell death inducer, leading to the ionic leakage of cells and the degeneration of mitochondria and proteins [6]. It can be seen from the fertility rate (Figure 5) that because of reduced synthesis of DNA, the reproduction of nematodes treated by the essential oil was lowered.
3.4. Effect on Mortality of Four Key Components in the Essential Oil. The structural formulae of four components are shown in Figure 6(a). The survival rate of nematodes exposed to d-limonene was 30% at 24 h; it was 34% for γ-terpinene, 11% for α-pinene, and 9% for β-pinene (Figure 6(b)). The survival rate of nematodes exposed to

Table 2: Number of body bends and head swings.

| Essential oil/0.1% DMSO (mg/ml) | 0   | 0.5 | 1   | 1.5  | 2    |
|---------------------------------|-----|-----|-----|------|------|
| Bending bodies/20 s             | 9.0±1.63 | 8.8±0.22 | 8±0.65 | 6.6±0.24 | 6.3±0.26 |
| Swinging heads/1 min            | 157±4.32 | 147±2.94 | 139±5.35 | 126±2.94 | 110±2.87 |

Note: Data are expressed as the average value ± standard deviation (n = 3).
α-pinene was 23% and that of β-pinene was 15% at 18 h. Therefore, the components of the essential oil of flesh fingered citron have obvious nematocidal effects.

4. Conclusions

Plant essential oils and extracts directly affect nematode survival [48]. Essential oil extracted from the flesh fingered citron can be used as a plant-derived nematicide to kill C. elegans. At a dose of 2 mg/ml (about 0.2%, v/v), the mortality rate reached 88%. These concentrations are much lower than the concentrations of insecticidal and nematicidal essential oils in the literature studies [49, 50], and the LD₅₀ value was 1.48 mg/ml. Furthermore, oxidative stress experiments showed that the essential oil can shorten the survival time of nematodes by reducing the antioxidative activity of C. elegans. Essential oils can also kill C. elegans by reducing its lifespan, in addition to affecting behaviors such as body bends and head swings and lowering the reproductive rate. Finally, four isolated components had a significant mortality effect on the nematodes, indicating that these four terpenes in the essential oil are responsible for its effects. Monoterpene and the key nematicides. This article uses Monoterpenes are the main ingredients indicating that these four terpenes in the essential oil are reducing the reproductive rate. Finally, four isolated components had a significant mortality effect on the nematodes, indicating that these four terpenes in the essential oil are responsible for its effects. Monoterpene and the key nematicides. This article uses C. elegans as a biological model to kill nematodes in the realistic laboratory, which lays an experimental basis for the subsequent development of new natural pesticides. It has a very important theoretical significance and application value. This essential oil is a candidate for partial or complete replacement of current chemically synthesized pesticides to enhance sustainability.

Data Availability

The numerical data used to support the findings of this study are included within the article.

Additional Points

Highlights. (1) A kind of plant-derived nematicide is proposed. (2) The lethal mechanism of flesh fingered citron essential oil on nematodes is studied. (3) The nematocidal effect is excellent which also can replace chemically synthesized nematicides, which is sustainable and environmentally friendly.

Conflicts of Interest

The authors declare that they have no conflicts of interest.

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References

[1] P. Majumder, H. A. Mondal, and S. Das, “Insecticidal activity of Arum maculatum tuber lectin and its binding to the glycosylated insect gut receptors,” Journal of Agricultural and Food Chemistry, vol. 53, no. 17, pp. 6725–6729, 2005.

[2] D. G. Heckel, “Insecticide resistance after silent spring,” Science, vol. 337, no. 6102, pp. 1612–1614, 2012.

[3] M. B. Isman, “Botanical insecticides, deterrents, and repellents in modern agriculture and an increasingly regulated world,” Annual Review of Entomology, vol. 51, no. 1, pp. 45–66, 2006.

[4] S. Senthil-Nathan, “Physiological and biochemical effect of neem and other Meliaceae plants secondary metabolites against lepidopteran insects,” Frontiers in Physiology, vol. 4, pp. 1–17, 2013.

[5] S. Rajendran and V. Srinivujini, “Plant products as fumigants for stored-product insect control,” Journal of Stored Products Research, vol. 44, no. 2, pp. 126–135, 2008.

[6] A. K. Tripathi, V. Prajapati, and S. Kumar, “Bioactivities of l-camphene, d-camphene, and dihydrocamphene toward three stored product beetles,” Journal of Economic Entomology, vol. 96, no. 5, pp. 1594–1601, 2003.

[7] J.-H. Tak and M. B. Isman, “Metabolism of citral, the major constituent of lemongrass oil, in the cabbage looper, Tri-choplusia ni, and effects of enzyme inhibitors on toxicity and metabolism,” Pesticide Biochemistry and Physiology, vol. 133, pp. 20–25, 2016.

[8] L. Dargahi, K. Razavi-Azarkhiavi, M. Ramezani, M. R. Abaee, and J. Behravan, “Insecticidal activity of the essential oil of Thymus transcapsicus against Anopheles stephensi,” Asian Pacific Journal of Tropical Biomedicine, vol. 4, pp. S589–S591, 2014.

[9] S. Pandey, S. Tiwari, A. Kumar et al., “Antioxidant and anti-aging potential of Juniper berry (Juniperus communis L.) essential oil in Caenorhabditis elegans model system,” Industrial Crops and Products, vol. 120, pp. 113–122, 2018.

[10] Y. Li, C. Wu, T. Wu et al., “Preparation and characterization of citrus essential oils loaded in chitosan microcapsules by using different emulsifiers,” Journal of Food Engineering, vol. 217, pp. 108–114, 2018.

[11] R. Augelletti, S. Frattari, F. Gironi, and W. Guerani, “Phase equilibria and thermodynamic modeling of systems CO₂—bergamot oil and CO₂—linalyl acetate,” The Journal of Supercritical Fluids, vol. 116, pp. 1–9, 2016.

[12] D. Chouchi, D. Barth, E. Reverchon, and G. Della Porta, “Supercritical CO₂ desorption of bergamot peel oil,” Industrial & Engineering Chemistry Research, vol. 34, no. 12, pp. 4508–4513, 1995.

[13] C. Celia, E. Trapasso, M. Locatelli et al., “Anticancer activity of liposomal bergamot essential oil (BEO) on human neuroblastoma cells,” Colloids and Surfaces B: Biointerfaces, vol. 112, pp. 548–553, 2013.

[14] L. Sánchez-González, C. Pastor, M. Vargas, A. Chiralt, C. González-Martínez, and M. Cháfer, “Effect of hydroxypropylmethylecellulose and chitosan coatings with and without bergamot essential oil on quality and safety of cold stored grapes,” Postharvest Biology and Technology, vol. 60, no. 1, pp. 57–63, 2011.

[15] N. Kumrungsee, W. Pluempanupat, O. Koul, and V. Bullangpoti, “Toxicity of essential oil compounds against diamondback moth, Plutella xylostella, and their impact on detoxification enzyme activities,” Journal of Pest Science, vol. 87, no. 4, pp. 721–729, 2014.

[16] A. Dinesh-Kumar, E. Srimaan, M. Chellappandian et al., “Target and non-target response of Swietenia Mahagoni Jacq. chemical constituents against tobacco cutworm Spodoptera litura Fab. and earthworm, Eudrilus eugeniae Knib,” Chemosphere, vol. 199, pp. 35–43, 2018.

[17] S. A. M. Abdelgaleel, M. I. E. Mohamed, M. E. I. Badawy, and S. A. A. El-arami, “Fumigant and contact toxicities of
monoterpenes to *Sitophilus oryzae* (L.) and *Tribolium castaneum* (Herbst) and their inhibitory effects on acetylcholinesterase activity,” *Journal of Chemical Ecology*, vol. 35, no. 5, pp. 518–525, 2009.

[18] W. Qin, S. Huang, C. Li, S. Chen, and Z. Peng, “Biological activity of the essential oil from the leaves of *Piper sarmentosum* Roxb. (Piperaceae) and its chemical constituents on *Brontispa longissima* (Gestro) (Coleoptera: hispidae),” *Pesticide Biochemistry and Physiology*, vol. 96, no. 3, pp. 132–139, 2010.

[19] S.-W. Kim, J. Kang, and I.-K. Park, “Fumigant toxicity of Apiaceae essential oils and their constituents against *Sitophilus oryzae* and their acetylcholinesterase inhibitory activity,” *Journal of Asia-Pacific Entomology*, vol. 16, no. 4, pp. 443–448, 2013.

[20] S. Kiran and B. Prakash, “Toxicity and biochemical efficacy of chemically characterized Rosmarinus officinalis essential oil against *Sitophilus oryzae* and *Oryzaephilus surinamensis*,” *Industrial Crops and Products*, vol. 74, pp. 817–823, 2015.

[21] H. Wei, J. Liu, B. Li et al., “The toxicity and physiological effect of essential oil from *Chenopodium ambrosioides* against the diamondback moth, *Plutella xylostella* (Lepidoptera: plute-lidiae),” *Crop Protection*, vol. 76, pp. 68–74, 2015.

[22] K. Dhiyva, G. Vengateswari, M. Arunthirumani, S. Karthi, S. Senthil-Nathan, and M. S. Shivakumar, “Bioprospecting of *Prosopis juliflora* (Sw.) DC seed pod extract effect on antioxi-dant and immune system of *Spodoptera littura* (Lepidoptera: noctuidae),” *Physiological and Molecular Plant Pathology*, vol. 101, pp. 45–53, 2018.

[23] X. Cao, Y. B. Lv, J. Chen et al., “Metabolic engineering of oleaginous yeast *Yarrowia lipolytica* for limonene overpro-duction,” *Biotechnology for Biofuels*, vol. 9, p. 214, 2016.

[24] J. Alonso-Gutierrez, R. Chan, T. S. Batth et al., “Metabolic engineering of *Escherichia coli* for limonene and perillyl alcoholk production,” *Metabolic Engineering*, vol. 19, pp. 33–41, 2013.

[25] E. Jongedijk, K. Cankar, J. Ranzijn et al., “Capturing of the monoterpene olenf limonene produced in *Saccharomyces cerevisiae*,” *Yeast*, vol. 32, pp. 159–171, 2015.

[26] W. J. Wang, “Recent advances on limonene, a natural and active monoterpene,” *China Food Additives*, vol. 1, pp. 33–37, 2005.

[27] A. D. R. Nuckerayanti, I. J. Nasser, F. Speror et al., “Essential oil composition, in vivo antioxidant, and antimicrobial activities of *Pimpinella pruatjan* from West Java, Indonesia,” *The Natural Products Journal*, vol. 8, no. 1, pp. 61–69, 2018.

[28] L. Guarante and C. Kenyon, “Genetic pathways that regulate ageing in model organisms,” *Nature*, vol. 440, no. 6589, pp. 255–262, 2000.

[29] F. Yi, R. Jin, J. Sun, B. Ma, and X. Bao, “Evaluation of me-chanically-pressed essential oil from Nanfeng Mandarin (*Citrus reticulata* Blanco cv. Kinokuni) as a food preservative based on antioxidant and antibacterial activities,” *Lwt*, vol. 95, pp. 346–353, 2018.

[30] F. Yi, J. Sun, X. Bao, B. Ma, and M. Sun, “Influence of me-locular distillation on antioxidant and antimicrobial activities of rose essential oils,” *Lwt*, vol. 102, pp. 310–316, 2019.

[31] A. Farhat, A.-S. Fabiano-Tixier, M. E. Maataoui, J.-F. Maingonnat, M. Romdhane, and F. Chemat, “Microwave steam distillation for extraction of essential oil from orange peel: kinetic data, extract’s global yield and mechanism,” *Food Chemistry*, vol. 125, no. 1, pp. 255–261, 2011.

[32] C.-W. Yu, W.-H. Li, F.-L. Hsu, P.-L. Yen, S.-T. Chang, and V. H.-C. Liao, “Essential oil alloaromadendrene from mixed-type *Cinnamomum osmophloeum* leaves prolongs the lifespan in *Caenorhabditis elegans*,” *Journal of Agricultural and Food Chemistry*, vol. 62, no. 26, pp. 6159–6165, 2014.

[33] E. N. Jesser, J. O. Werdin-Gonzalez, A. P. Murray, and A. A. Ferrero, “Efficacy of essential oils to control the Indian meal moth, *Plodia interpunctella* (Hübner) (Lepidoptera: Pyralidae),” *Journal of Asia-Pacific Entomology*, vol. 20, no. 4, pp. 1122–1129, 2017.

[34] Z. Lou, J. Chen, F. Yu et al., “The antioxidant, antibacterial, antbiofilm activity of essential oil from *Citrus medica* L. var. *sarcodactylis* and its nanoemulsion,” *LWT*, vol. 80, pp. 371–377, 2017.

[35] G. J. Lithgow and G. A. Walker, “Stress resistance as a de-terminate of *C. elegans* lifespan,” *Mechanisms of Ageing and Development*, vol. 123, no. 7, pp. 765–771, 2002.

[36] A. Pant, S. K. Saikia, V. Shukla, J. Asthana, B. A. Akhoon, and R. Pandey, “Beta-caryophyllene modulates expression of stress response genes and mediates longevity in *Caeno-rhabditis elegans*,” *Experimental Gerontology*, vol. 57, pp. 81–95, 2014.

[37] L. Espina, M. Solominos, S. Lorán, P. Conchello, D. García, and R. Pagán, “Chemical composition of commercial citrus fruit essential oils and evaluation of their antimicrobial ac- tivity acting alone or in combined processes,” *Food Control*, vol. 22, no. 6, pp. 896–902, 2011.

[38] N. Tao, L. Jia, and H. Zhou, “Anti-fungal activity of *Citrus reticulata* Blanco essential oil against *Penicillium italicum* and *Penicillium digitatum*,” *Food Chemistry*, vol. 153, pp. 265–271, 2014.

[39] C.-W. Yu, C.-C. Wei, and V. H.-C. Liao, “Curcumin-mediated oxidative stress resistance in *Caenorhabditis elegans* modulated by-age-1, akt-1, pdk-1, osr-1, unc-43, sek-1, skn-1, sir-2.1, and mev-1,” *Free Radical Research*, vol. 48, no. 3, pp. 371–379, 2014.

[40] O. X. Shaw, G. Werstuck, and Y. Chen, “Oxidative stress and aging diseases,” *Oxidative Medicine and Cellular Longevity*, vol. 2014, Article ID 569146, 2 pages, 2014.

[41] C. J. Kenyon, “The genetics of ageing,” *Nature*, vol. 464, no. 7288, pp. 504–512, 2010.

[42] L. P. O’Reilly, C. J. Luke, D. H. Perlmutter, G. A. Silverman, and S. C. Pak, “*C. elegans* in high-throughput drug discovery,” *Advanced Drug Delivery Reviews*, vol. 69–70, no. 70, pp. 247–253, 2014.

[43] S. Zhu, H. Li, J. Dong et al., “Rose essential oil delayed alzheimer’s disease-like symptoms by SKN-1 pathway in *C. elegans*,” *Journal of Agricultural and Food Chemistry*, vol. 65, no. 40, pp. 8855–8865, 2017.

[44] M. Matura, A. Goossens, O. Bordalo et al., “Oxidized citrus oil (R-limonene): a frequent skin sensitizer in Europe,” *Food Chemistry*, vol. 153, pp. 265–271, 2014.

[45] M. Matura, A. Goossens, O. Bordalo et al., “Oxidized citrus oil (R-limonene): a frequent skin sensitizer in Europe,” *Food Chemistry*, vol. 153, pp. 265–271, 2014.

[46] S.-I. Kim, J.-S. Yoon, J. W. Jung, K.-B. Hong, Y.-J. Ahn, and H. W. Kwon, “Toxicity and repellency of origanum essential oil and its components against *Tribolium castaneum* (Cole-optera: tenebrionidae) adults,” *Journal of Asia-Pacific Entomo-logy*, vol. 13, no. 4, pp. 369–373, 2010.

[47] G. Gupta, U. Agarwal, H. Kaur, N. R. Kumar, and P. Gupta, “Aphidical effects of terpenoids present in *Citrus limon* on
Macrosiphum roseiformis and two generalist insect predators,”
Journal of Asia-Pacific Entomology, vol. 20, no. 4, pp. 1087–
1095, 2017.

[48] M. Shahriari, A. Zibaee, N. Sahebzadeh, and L. Shamakhi,
“Effects of α-pinene, trans-anethole, and thymol as the es-
sential oil constituents on antioxidant system and acetyl-
choline esterase of Ephesia kuehniella Zeller (Lepidoptera:
Pyralidae),” Pesticide Biochemistry and Physiology, vol. 150,
pp. 40–47, 2018.

[49] K. Polatoglu, Ö. C. Karakoc, Y. Y. Yücel et al., “Insecticidal
activity of edible Crithmum maritimum L. essential oil against
Coleopteran and Lepidopteran insects,” Industrial Crops and
Products, vol. 89, pp. 383–389, 2016.

[50] C. Maleita, I. Esteves, R. Chim et al., “Naphthoquinones from
walnut husk residues show strong nematicidal activities
against the root-knot Nematode Meloidogyne hispanica,”
ACS Sustainable Chemistry & Engineering, vol. 5, no. 4,
pp. 3390–3398, 2017.