Heterogeneous Escape from X Chromosome Inactivation Results in Sex Differences in Type I IFN Responses at the Single Human pDC Level

Graphical Abstract

Highlights
- TLR7, CYBB, RPS6KA3, BTK, and IL13RA1 can escape XCI in human pDCs
- Female pDCs with escape from XCI have higher mRNA levels of the respective genes
- pDCs with escape from XCI of TLR7 have higher IFNα/β mRNA levels

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In Brief
In human female pDCs with two X chromosomes, Hagen et al. show that TLR7 escape from X chromosome inactivation (XCI) promotes higher TLR7 mRNA and higher IFNα/β mRNA at the single-cell level. This finding highlights the contribution of X chromosomal factors to sex differences in type I IFN responses.

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Heterogeneous Escape from X Chromosome Inactivation Results in Sex Differences in Type I IFN Responses at the Single Human pDC Level

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INTRODUCTION

Immune responses differ between women and men, and type I interferon (IFN) responses following Toll-like receptor 7 (TLR7) stimulation are higher in women. The precise mechanisms driving these sex differences in immunity are unknown. To investigate possible genetic factors, we quantify escape from X chromosome inactivation (XCI) for TLR7 and four other genes (RPS6KA3, CYBB, BTK, and IL13RA1) at the single plasmacytoid dendritic cell (pDC) level. We observe escape from XCI for all investigated genes, leading to biallelic expression patterns. pDCs with biallelic gene expression have significantly higher mRNA levels of the respective genes. Unstimulated pDCs with biallelic TLR7 expression exhibit significantly higher IFNα/β mRNA levels, and IFNα exposure results in significantly increased IFNα/β protein production by pDCs. These results identify unanticipated heterogeneity in escape from XCI of several genes in pDCs and highlight the important contribution of X chromosome factors to sex differences in type I IFN responses, which might explain observed sex differences in human diseases.

SUMMARY

Immune responses differ between women and men, and type I interferon (IFN) responses following Toll-like receptor 7 (TLR7) stimulation are higher in women. The precise mechanisms driving these sex differences in immunity are unknown. To investigate possible genetic factors, we quantify escape from X chromosome inactivation (XCI) for TLR7 and four other genes (RPS6KA3, CYBB, BTK, and IL13RA1) at the single plasmacytoid dendritic cell (pDC) level. We observe escape from XCI for all investigated genes, leading to biallelic expression patterns. pDCs with biallelic gene expression have significantly higher mRNA levels of the respective genes. Unstimulated pDCs with biallelic TLR7 expression exhibit significantly higher IFNα/β mRNA levels, and IFNα exposure results in significantly increased IFNα/β protein production by pDCs. These results identify unanticipated heterogeneity in escape from XCI of several genes in pDCs and highlight the important contribution of X chromosome factors to sex differences in type I IFN responses, which might explain observed sex differences in human diseases.
The mechanisms contributing to higher IFNα production by female pDCs remain incompletely understood. Studies showed a positive regulation of IFNα production by estradiol (Seillet et al., 2012; Griesbeck et al., 2015), and further experiments suggested genetic (X chromosome linked) factors (Souyris et al., 2018) or a combination of both (Laffont et al., 2014). The location of the TLR7 gene on the X chromosome is a potential reason for genetically driven differences due to differential allosome allocation between XX females and XY males. In order to achieve dosage compensation between the sexes, one of the two X chromosomes in females is transcriptionally silenced, a process that is referred to as X chromosome inactivation (XCI) (Avner and Heard, 2001). Studies have indicated that some genes on the X chromosome can escape XCI in females, resulting in transcription of the genes from both X chromosomes (Xi) (Carrel et al., 1999; Carrel and Willard, 2005; Souyris et al., 2018). The non-coding RNA XIST is necessary for establishing XCI (Loda and Heard, 2019), and it is thought that the Xi is in most settings stably preserved over time and cell divisions (Payer and Lee, 2008; Robert Finestra and Gribnau, 2017). Yet, relatively little is known about the occurrence and maintenance of escape from XCI at the single immune cell level.

Here, we demonstrate extensive heterogeneity in escape from XCI in female pDCs. Biallelic expression of the investigated X chromosome genes resulted in higher transcript levels of the respective mRNAs. IFNα and IFNβ mRNAs were more highly transcribed in biallelic than monoallelic TLR7-expressing pDCs, and exposure to the IFNα proteins resulted in higher levels of IFNα/β protein production. Taken together, these data demonstrate the important contribution of X chromosome factors to sex differences in type I IFN responses.

RESULTS

Assessment of Escape from XCI in Different X Chromosome Genes

Previous studies have described escape from XCI in mice and humans, but the extent of escape from XCI for different X chromosome genes in pDCs and the resulting consequences for immune responses are not well understood. To investigate escape from XCI at the single pDC level, we took advantage of single nucleotide polymorphisms (SNPs) within the mature mRNA of genes encoded by the X chromosome, enabling differentiation of mRNA encoded by the two X chromosomes in heterozygous females (Berghöfer et al., 2006; Souyris et al., 2018). We selected TLR7 and four other genes (RPS6KA3, CYBB, BTK, and IL13RA1), based on their expression levels in pDCs, location on the X chromosome, and presence of a SNP in the mature mRNA with a minor allele frequency (MAF) of over 20% (Table S1; Figure 1A). After identifying healthy female individuals with heterozygosity of the respective SNPs, freshly isolated peripheral blood mononuclear cells (PBMCs) were stimulated for 2 h with the TLR7/8 agonist CL097, pDCs were subsequently sorted (Figure S1A), and cDNA of single pDCs was obtained using the
Fluidigm C1 technology (Figure 1B). pDCs expressing the respective mRNA derived from one (monoallelic) or two (biallelic) X chromosomes were identified using the Fluidigm Biomark HD platform based on detection of mRNA containing the major or minor allele nucleotide, or both, as shown for representative female individuals heterozygous for the respective SNPs in the X chromosome genes TLR7, RPS6KA3, CYBB, BTK, and IL13RA1 in Figure 2A. Female individuals homozygous for either major allele (TLR7, RPS6KA3, CYBB, and IL13RA1) or the minor allele (BTK), thus only displaying one population in the escape plot, were used as controls (Figure 2B). Any cell without detectable mRNA for the respective gene was excluded from further analysis. The percentage of pDCs with biallelic expression for the investigated genes ranged from 5.6% to 37.3%, with the broadest range in pDCs with escape from XCI observed for BTK and TLR7 (BTK: 5.6%–31.3%; TLR7: 19.0–37.3%; Figure 2C). Overall these data show that TLR7, RPS6KA3, CYBB, BTK, and IL13RA1 can escape XCI in pDCs and that escape from XCI can be assessed at the single immune cell level.

Female pDCs Present with a Heterogeneous XCI Profile

To investigate the diversity of escape from XCI within each female, we assessed the number of escape loci within single pDCs and observed that all except female 1 (F1) had at least two or more cells with escape from XCI in multiple loci (Figure 3A). We furthermore detected pDCs in F2, F3, and F4 that had escape from XCI for every loci assessed. To investigate the influence of the non-coding RNA XIST within single pDCs, cDNA of the identical single pDCs in which escape from XCI was determined was used for XIST RNA quantification. The expression of XIST RNA did not correlate with the number of escape loci per cell (r = 0.17) (Figure 3B); however, there was a
significant difference in XIST RNA expression levels comparing pDCs with at least one locus of biallelic expression compared to pDCs without biallelic expression (Figure 3C). In summary, different pDCs from the same individual had a heterogeneous profile of escape from XCI. We did not observe a correlation between the number of escape loci and XIST expression, but XIST RNA had higher expression in pDCs with biallelic expression.

**pDCs with Escape from XCI Have Significantly Higher mRNA Levels of the Respective Escaped Genes Than pDCs with Monoallelic Expression**

To determine whether biallelic expression of X chromosome genes resulted in higher mRNA expression levels of the respective genes within the individual cell, we analyzed cDNA that was used to determine mono- versus biallelic expression patterns also for quantification of mRNA levels by using the Fluidigm Biomark HD platform (Figure 1B). In all three female individuals heterozygous for the TLR7 SNP rs3853839, significantly higher TLR7 mRNA transcript levels were observed in pDCs with biallelic than those with monoallelic TLR7 expression (Figure 4; Figure S2A). Of note, monoallelic- and biallelic-TLR7-expressing pDCs from female individuals did not show any significant difference in mRNA expression of the reference gene GAPDH or the pDC genes HLA-DRA and IL3RA (CD123) (Figure S2B).

Expanding the analysis to a comparison of mRNA levels of RPS6KA3, CYBB, BTK, and IL13RA1 between monoallelic- and biallelic-expressing pDCs, higher mRNA expression levels in pDCs with escape from XCI in the respective genes were consistently observed (Figure 4). For the genes RPS6KA3, BTK, and IL13RA1, female pDCs with monoallelic expression did not have higher mRNA levels than pDCs from males, whereas significant differences in mRNA levels between pDCs from males and biallelic-expressing pDCs from females were observed. We furthermore measured the TLR7 protein levels and observed significantly higher TLR7 protein levels in female pDCs than those in male pDCs (Figure S2C), as previously described for human PBMCs (Souyris et al., 2018). Altogether, these data demonstrate that escape from XCI in TLR7, RPS6KA3, CYBB, BTK, and IL13RA1 led to higher mRNA expression levels of the respective genes in pDCs with a biallelic expression pattern.

**pDCs with Biallelic mRNA Expression of TLR7 Have Significantly Higher IFNα and IFNβ mRNA Expression**

To further examine the functional consequences of escape of TLR7 from XCI for pDC function, we assessed whether escape of TLR7 from XCI and the resulting higher TLR7 mRNA levels were associated with higher mRNA transcript levels of IFNα subtypes and IFNβ in unstimulated pDCs. We observed a significant difference in IFNα and IFNβ mRNA expression levels between female pDCs with monoallelic compared to pDCs with biallelic TLR7 expression (Figure 5A). The difference between monoallelic- and biallelic-TLR7-expressing pDCs was furthermore statistically significant for all IFNα subtypes and IFNβ mRNA expression individually (Figure S3). Following 2-h stimulation of pDCs,
significant differences between monoallelic- and biallelic-TLR7-expressing pDCs for all IFNα subtypes and IFNβ mRNA persisted; however, stimulation led to an upregulation of IFNα and IFNβ mRNA levels, in particular in monoallelic-TLR7-expressing pDCs (Figure 5B). When we compared the lowest with the highest quartile of TLR7-mRNA-expressing pDCs, unstimulated pDCs with the highest TLR7 mRNA expression encoded significantly higher levels of IFNα and IFNβ mRNAs than pDCs with the lowest TLR7 mRNA expression levels (Figure 5C), and this difference was lost after 2 h of stimulation (data not shown). This result indicates that during initial IFNα and IFNβ production, pDCs with biallelic TLR7 expression and high TLR7 mRNA levels have higher type I IFN production; however, the TLR7-independent autocrine and paracrine IFNAR feedback loops diminished these differences between low- and high-TLR7-expressing pDCs following stimulation.

Following 2-h TLR7/8 stimulation of PBMCs, the mRNA expression level for all IFNα subtypes and IFNβ was significantly higher in pDCs from females than that from males, independent of whether there was monoallelic or biallelic expression of TLR7 (Figure 5D), highlighting the important role of the initial TLR7-dependent induction of IFNα/β production for these sex differences in type I IFN responses. To investigate whether the difference in mRNA levels following 2 h of stimulation was also present at the protein level, we measured the amount of panIFNα protein that was secreted in the supernatant following 2-h TLR7/8 stimulation of human PBMCs. PBMCs of females produced significantly more IFNα protein than males (female median = 453 pg/ml; male median = 205 pg/ml) (Figure 5E), which is in line with previous studies (Berghöfer et al., 2006; Meier et al., 2009; Ziegler et al., 2017). To demonstrate that the amount of IFNα that
was secreted within 2 h of stimulation can indeed initiate an IFNAR feedback loop, we pre-treated PBMCs with IFNα2 (1 U/ml; 5 pg/ml) and IFNα4 (1 U/ml; 1.9 pg/ml) and observed that the IFNα pre-treatment led to a significant increase in IFNα and IFNβ protein production by human pDCs (Figure S4). Overall, these data demonstrate that IFNα and IFNβ mRNAs have significantly higher expression in pDCs with biallelic expression of TLR7 and that higher initial IFNα secretion by female pDCs can increase the IFNα and IFNβ protein production probably due to an auto- and paracrine IFNAR feedback loop.

**DISCUSSION**

Sex differences in immune responses have been described in infectious and autoimmune diseases (Markle and Fish, 2014), as well as in response to vaccinations (Flanagan et al., 2017).
Next to modulation of immune function by sex hormones, differences in the number of X chromosomes between women and men can contribute to this immunological sex bias (Libert et al., 2010). Previous studies have shown that 10%–25% of genes on the X chromosome can escape from XCI (Carrel et al., 1999; Carrel and Willard, 2005), with variability between different tissues and cells (Tukiainen et al., 2017). With a size of 156.04 mega base pairs (Mbps), the X chromosome is the 8th biggest chromosome (Yates et al., 2020) and contains a number of genes important for immune function and regulation (Fish, 2008), suggesting that escape from XCI in females might impact immune function. Investigating escape from XCI at the single pDC level, we show that biallelic expression of X chromosome genes occurred frequently, contributed to transcriptional heterogeneity of pDCs, and resulted in higher mRNA levels of the respective genes. IFNα and IFNβ mRNAs were more highly transcribed in female pDCs with biallelic expression of TLR7, and exposure of pDCs to IFNα protein significantly enhanced IFNα and IFNβ protein production. Taken together, these data demonstrate the important role of X chromosome factors in the regulation of the type I IFN responses and suggest that female pDCs with biallelic expression of TLR7 and stronger induction of type I IFNs are responsible for initiating sex-specific differences in IFNα and IFNβ protein production.

A number of recent studies have demonstrated escape from XCI (Wang et al., 2016) with consequences for sex-specific immune function (Souyris et al., 2018; Oghumu et al., 2019), but the mechanisms underlying escape from XCI of specific genes and their functional consequences at the single-cell level are not well understood. We investigated the expression of five X-chromosome-encoded genes in pDCs at the single-cell level, including TLR7 for which escape from XCI has been previously reported (in human B cells, pDCs, and monocytes; Souyris et al., 2018), and in four additional genes (RPS6KA3, CYBB, BTK, and IL13RA1) selected based on their expression levels in pDCs, location on the X chromosome, and presence of a SNP with a high MAF in the mature mRNA. Our data show that the five investigated genes escaped XCI in pDCs of every female that was investigated and that escape from XCI occurred in 5%–37% of pDCs. The degree of escape from XCI of these different genes varied between different pDCs, resulting in heterogeneous X chromosome gene expression within female pDC populations. In line with a previous study reporting enhanced TLR7 transcripts in biallelic–TLR7-expressing B cells and higher TLR7 protein expression in human PBMCs (Souyris et al., 2018), we observed that pDCs with escape from XCI in TLR7 had significantly higher TLR7 mRNA transcript levels than female monoallelic–TLR7-expressing pDCs, and we confirmed significantly higher TLR7 protein levels in pDCs from females than in those from males.

Investigation of the other four studied X chromosome genes consistently demonstrated higher mRNA expression in female pDCs with biallelic expression of the respective genes than in female pDCs with monoallelic expression. Available data on the regulation of escape from XCI in pDCs is very limited. One study in mice described that XIST expression varies between different immune cells and that XIST is nearly absent in murine pDCs (Syrett et al., 2019). In contrast, we detected XIST RNA in the majority of human pDCs. These dissimilarities to the study by Syrett et al. (2019) might be due to differences in the factors regulating XCI in humans and mice (Migeon, 2017). Interestingly, we observed a significant difference in XIST RNA expression between pDCs with or without biallelic expression. However, it has been suggested that the inactivation activity of XIST is limited to a short time frame during early development (Wutz and Jaenisch, 2000), and multiple studies have proposed that XIST is not required for the maintenance of the inactivation of the X chromosome (Csankovszki et al., 1999; Rack et al., 1994), indicating that additional studies are required to investigate the role of XIST in regulating escape from XCI in humans. In summary, our results show transcription from multiple genes located on the second X chromosome in women, with significant consequences for the mRNA levels within the respective cell, contributing to a transcriptional heterogeneity of individual pDCs in female individuals.

Previous studies have shown that pDCs of females produce significantly more IFNα than pDCs of males following TLR7 stimulation (Bergeöfer et al., 2006; Meier et al., 2009; Seillet et al., 2012; Griesbeck et al., 2015; Ziegler et al., 2017) and implicated these sex differences in type I IFN production in the pathogenesis of infectious and autoimmune diseases that exhibit clear differences between women and men (Farzadegan et al., 1998; Meditz et al., 2011; Sterling et al., 2001; Amur et al., 2012). Remarkably, the incidence of SLE, an autoimmune disease primarily observed in women, is also elevated in males with Klinefelter syndrome (47, XXY) (Scofield et al., 2008). Women with Turner syndrome (45, X0) appear to have a lower risk for SLE (Cooney et al., 2009), whereas the incidence for females with triple X syndrome (47, XXX) is 2.5 times higher than for women without the syndrome (46, XX) (Liu et al., 2016), suggesting an X chromosome dosage effect in the pathogenesis of this autoimmune disease (Scofield et al., 2008). SLE patients have elevated IFNα levels in their blood (Friedman et al., 1982; Kim et al., 1987) and significantly higher expression of ISGs (Crow et al., 2003), and there are case reports of human patients developing SLE following IFNα therapy, highlighting the connection between type I IFNs and SLE (Rönnblom et al., 1990; Fukuyama et al., 2000). A mouse strain with an autoimmune phenotype resembling human SLE has furthermore been shown to have an additional cluster of X-linked genes containing murine Tlr7 (Izui et al., 1994; Subramanian et al., 2006), and deletion or impairment of pDCs in vivo improved clinical symptoms of SLE (Rowland et al., 2014; Sisirak et al., 2014). These data highlight an important role of X-linked factors in autoimmune diseases that might be explained by escape of TLR7 from XCI and result in higher induction of type I IFN mRNA in these pDCs. The current SARS-CoV-2 pandemic also displays a sex bias with 1.7 times higher case mortality rates in males than in females reported in Europe (Gebhard et al., 2020). A case study with four young men (under 35) without major preexisting diseases reported severe to fatal COVID-19 progression linked to loss-of-function mutations of TLR7 (van der Made et al., 2020), highlighting the importance of functional TLR7 signaling in the control of SARS-CoV-2 infection. It was furthermore suggested that early treatment with IFNα2 during initial SARS-CoV-2 infection was associated with reduced mortality (Wang et al., 2020). Our data showing higher
IFNα and IFNβ mRNA levels in female pDCs with biallelic TLR7 expression suggest that higher type I IFN responses in females might potentially contribute to a better control of SARS-CoV-2 infection in women (Sünders and Altfeld, 2020).

Quantifying IFNα/β mRNA expression in pDCs, we observed that unstimulated pDCs with biallelic expression of TLR7 exhibit higher mRNA levels of IFNα/β. Furthermore, the highest quartile of TLR7-expressing pDCs also expressed higher levels of IFNα/β mRNA, indicating that the initial IFNα/β response importantly depends on TLR7 mRNA expression levels. These differences in IFNα/β mRNA levels between the lowest and the highest quartile of TLR7-expressing pDCs diminished following stimulation, likely due to the auto- and paracrine IFNAR signaling that is independent of TLR7. Unfortunately, current methods do not allow determination of whether an individual pDC with biallelic expression of TLR7 and higher TLR7 mRNA expression also produces more IFNα/β protein. However, these data suggest that female pDCs with biallelic and thus higher expression of TLR7 mRNA might belong to the group of pDCs described to independently initiate IFNα secretion (Wimmers et al., 2018). The observation that unstimulated pDCs harbor mRNA for type I IFNs is in line with published data describing that pDCs constitutively produce small amounts of type I IFNs as a self-priming mechanism for TLR stimulation (Kim et al., 2014). Taken together, these data suggest that pDCs as professional type I IFN producers already have a basal level of type I IFN mRNA expression, with pDCs with biallelic TLR7 expression exhibiting higher mRNA levels of IFNα/β.

In summary, the results from these studies support a model in which a subset of female pDCs with escape from XCI in TLR7 and resulting higher TLR7 mRNA levels can respond with stronger initial IFNα/β production, initiating an auto- and paracrine IFNAR feedback loop that drives higher IFNα/β production in females. Our results furthermore show the significant impact of escape from XCI at the single-cell level on transcriptional and functional heterogeneity of immune cells, which can contribute to the described differences in immune-mediated diseases between women and men.

**STAR METHODS**

Detailed methods are provided in the online version of this paper and include the following:

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**QUANTIFICATION AND STATISTICAL ANALYSIS**

**SUPPLEMENTAL INFORMATION**

Supplemental Information can be found online at https://doi.org/10.1016/j.celrep.2020.108485.

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**AUTHOR CONTRIBUTIONS**

S.H.H. and M.A. designed the study. S.H.H. and F.H. performed experiments and analyzed the data. J.H. gave critical experimental support. S.H.H. performed basic statistical tests. S.D. and H.S. performed mixed effects linear regression models and correlations with repeated measurements. L.R. provided important statistical guidance. S.M.Z. gave important intellectual input. S.H.H. and M.A. wrote the manuscript. All authors reviewed the manuscript and gave approval for publication.

**DECLARATION OF INTERESTS**

The authors declare that they have no conflict of interest.

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### STAR METHODS

#### KEY RESOURCES TABLE

| REAGENT or RESOURCE                        | SOURCE                          | IDENTIFIER                  |
|--------------------------------------------|---------------------------------|-----------------------------|
| **Antibodies**                             |                                 |                             |
| CD11c-PE/Cy7 (Bu15)                        | Biolegend                       | Cat#: 337216, RRID: AB_2129790 |
| CD123-BV711 (9F5)                          | BD                              | Cat#: 563161, RRID: AB_2738038 |
| CD123-FITC (6H6)                           | Biolegend                       | Cat#: 306014, RRID: AB_2124259 |
| CD14-APC/Cy7 (HCD14)                       | Biolegend                       | Cat#: 325620, RRID: AB_830693 |
| CD19-BUV395 (SJ25C1)                       | BD                              | Cat#: 563549, RRID: AB_2738272 |
| CD19-BUV737 (SJ25C1)                       | BD                              | Cat#: 564303, RRID: AB_2716867 |
| CD3-BUV737 (UCHT1)                         | BD                              | Cat#: 564307, RRID: AB_2744390 |
| CD56-BUV395 (NCAM16.2)                     | BD                              | Cat#: 563554, RRID: AB_2687886 |
| IFNα2b-V450 (7N4-1)                        | BD                              | Cat#: 561382, RRID: AB_10716058 |
| IFNα-PE (LT27:295)                         | Miltenyi Biotec                 | Cat#: 130-099-098, RRID: AB_671560 |
| IFNγ-FITC (MMHB-3)                         | PBL Assay Science               | Cat#: PBL-21400-3, RRID: AB_387831 |
| HLA-DR-BV605 (L243)                        | Biolegend                       | Cat#: 307640, RRID: AB_2561913 |
| TLR7-APC (4G6)                             | Novus Biologicals               | Cat#: NBP2-25274APC, RRID: N/A |
| **Biological Samples**                     |                                 |                             |
| Human blood                                | Healthy individuals from the University Medical Center Hamburg-Eppendorf | Cat#: N/A |
| **Chemicals, Peptides, and Recombinant Proteins** |                                 |                             |
| 1x DNA Suspension Buffer                    | Teknova                         | Cat#: T0221                 |
| 2x Sso Fast Eva Green Supermix with Low ROX | Bio-Rad                         | Cat#: 1725210               |
| 2x Fast Probe Master Mix                    | Biotium                         | Cat#: 31005                 |
| ACK Lysing Buffer                          | Lonza                           | Cat#: 10-548E               |
| Bioqoll-Trenløsung                         | Biochrom                       | Cat#: L6115                 |
| Brefeldin A                                | Sigma-Aldrich                  | Cat#: B7651-5MG             |
| CL097                                      | Invivogen                      | Cat#:trl-c97               |
| Dulbecco’s phosphate buffered saline (PBS)  | Sigma-Aldrich                  | Cat#: D8637                 |
| Dymethyl sulfoxide (DMSO)                   | Sigma-Aldrich                  | Cat#: D5879-100ML           |
| Ethylenediaminetetraacetic acid (EDTA) solution | Sigma-Aldrich                  | Cat#: 03690-100ML           |
| Fetal bovine serum (FBS) superior           | Biochrom                       | Cat#: S0615                 |
| Fixation Medium (Medium A)                 | ThermoFisher Scientific        | Cat#: GAS001S100            |
| Human Recombinant IFNα2a                    | Stemcell                       | Cat#: 78076.1               |

(Continued on next page)
RESOURCE AVAILABILITY

Lead Contact
Further information and requests for resources and reagents should be directed to and will be fulfilled by the Lead Contact, Marcus Altfeld (marcus.altfeld@leibniz-hpi.de)

Materials Availability
This study did not generate new unique reagents.

Data and Code Availability
This study did not generate/analyze datasets or code.

EXPERIMENTAL MODEL AND SUBJECT DETAILS

Healthy individuals were recruited at the University Medical Center Hamburg-Eppendorf. The ethical commission “Ärztekammer” Hamburg approved the study (PV4780). The age of the donors ranged from 20 – 36. A similar number of females and males were included for the experiments to study sex-specific differences and the sex of each individual donor is indicated in the figure legend. Experiments investigating the effect of escape from XCI were only performed in female individuals, due to the requirement of two X chromosomes. Informed consent was provided by each individual prior to enrollment in the study.
METHOD DETAILS

Isolation and freezing of PBMCs
Blood from healthy individuals was drawn in EDTA tubes (Sarstedt) and the processing of the blood was started within 90 minutes after venipuncture (Meier et al., 2008). PBMCs were isolated using Ficoll-Paque (Biochrom) density centrifugation. Remaining erythrocytes were lysed in 3 mL of ACK Lying Buffer (Lonza) for 3 min. Freshly isolated PBMCs were immediately used for experiments or frozen in heat inactivated FBS (Biochrom) with 10% DMSO (Sigma-Aldrich) at –80°C and then transferred the next day into liquid nitrogen storage.

Purification of pDCs
pDCs were purified from frozen PBMCs via negative selection using the Plasmacytoid Dendritic Cell Isolation Kit II, human (Miltenyi Biotec) according to manufactures instructions. For up to 25 Mio PBMCs 25% of the recommended amount of were used (e.g., 25 μL of Non-pDC Biotin-Antibody Cocktail II for up to 25 Mio PBMCs). As buffer PBS (Sigma-Aldrich) + 0.5% FBS (Biochrom) + 2mM EDTA (Sigma-Aldrich) was used.

TLR stimulation and IFNα pre-treatment
2 million freshly isolated PBMCs per ml were stimulated in GIBCO RPMI 1640 Medium (Life technologies) supplemented with 10% heat inactivated FBS (Biochrom) (R10) with 1 μg/ml of TLR7/8 agonist CL097 (Invivogen). For analysis of the supernatant the stimulation was performed for 2 h. For flow cytometric analysis the stimulation was done for 6 h in the presence of 5 μg/ml brefeldin A (Sigma-Aldrich), as previously described (Meier et al., 2009; Ziegler et al., 2017). For IFNα pre-treatment experiments PBMCs were cultured for 2 h in R10 with or without IFNα2a (Stemcell) or IFNα4b (PBL Assay Science) at the indicated concentrations prior to stimulation with CL097 (in the presence of BFA). An unstimulated, but BFA-treated control sample was included for every experiment. For subsequent mRNA analysis, the stimulation of PBMCs was performed for 2 h. For mRNA analysis pDCs were purified from frozen PBMCs and stimulated at 50 000 pDCs per ml with 1 μg/ml of CL097 for 2 h or pDCs were purified from frozen PBMCs and left unstimulated.

Cell staining, flow Cytometry and cell sorting
Prior to staining, PBMCs / isolated pDCs were washed with PBS (Sigma-Aldrich). Staining was performed for 20 minutes (RT / dark) with the indicated surface antibodies and Zombie Aqua (Biolegend). The antibodies used for staining are shown in the Key Resources Table. For mRNA analysis PBMCs / isolated pDCs were left unfixed and stored on ice until pDCs were sorted as single live CD3-CD19-CD56-CD11c-CD14-CD123+HLA-DR+ cells using a FACSAria Fusion. Gating strategies are shown in Figure S1A.

For intracellular cytokine staining and intracellular staining of TLR7, PBMCs were fixed with Medium A (Life Technologies). Intra-cellular staining was done using Medium B (Life Technologies) with the indicated antibodies for 20 minutes. IFNα2 was quantified with clone 7N4-1 (BD) which primarily detects IFNα2. PolyIFNα was quantified with clone LT27:295 (Miltenyi Biotec), which detects IFNα2 and several other IFNα subtypes (Svensson et al., 1996). Gating strategies are shown in Figure S1A (TLR7) and Figure S1B (cytokines). Frozen PBMCs were used for the staining of TLR7. For staining of TLR7 an FMO control was included for every sample and the median fluorescent intensity (MdFl) of TLR7 was normalized to the FMO control of the same donor (MdFl TLR7 / MdFl FMO). Cells were analyzed using a LSRFortessa Flow Cytometer (BD) in the core facility Fluorescence Cytometry at the HPI, and FCS data were analyzed using FlowJo version 10.

panIFNα ELISA
panIFNα concentrations were measured in duplicates in cell culture supernatant following 2 h stimulation of freshly isolated PBMCs with CL097 (1 μg/ml) (Invivogen) using Human pan IFN-alpha ELISA Kit (Stemcell) according to manufactures’ instructions. The harvested supernatant was stored at –80°C until the measurement.

Genotyping of healthy donors
Genomic DNA (gDNA) of donors was obtained using DNeasy Blood & Tissue Kit (QIAGEN) according to protocol. The genotyping was performed with the Biomark HD (Fluidigm) using the 48.48 Dynamic Array IFC for Genotyping (Fluidigm). A specific target amplification (STA) of gDNA was performed prior to SNP genotyping. STA and SNP genotyping was done according to company protocol. The primers used for preamplification and SNP genotyping are shown in Table S2 and Table S4, respectively.

Single cell gene expression analysis
A platform that consisted of the following Fluidigm devices was used for single cell analysis: C1, Juno and Biomark HD (Fluidigm).

C1
Single-Cell Preamp IFCs, 5–10 μm (Fluidigm) were used for the generation of cDNA. After cell capture, all 96 capture sites were visualized under a microscope and only capture sites containing one single cell were included into further analysis. Single Cell Lysis Solution and DNase I solution (both included in the Single Cell-to-CT qRT-PCR Kit, Invitrogen / Thermo Fisher Scientific) were mixed...
in a ratio of 10:1 before being added to the Lysis final mix. Generation of cDNA was performed following company protocol. Pre-amplification of mRNA surrounding the respective SNPs was done with the outer primers of the SNP / DG primers to create an amplicon that could be analyzed with both the SNP and DG primers (Tables S2 and S3). A negative control was included with every C1 run.

**Biomark HD**
Gene expression analysis was performed with Biomark HD using 96.96 Dynamic Array IFC for Gene Expression. SNP typing of mRNAs was performed using 192.24 Dynamic Array IFCs for SNP genotyping following company protocol but using the cDNA generated in the C1 instead of gDNA. A water control was included in every Biomark run. Primers used for real-time quantitative PCR and SNP genotyping are shown in Table S3 and Table S4 respectively.

**Data processing**
The data processing was performed as described previously (Hess et al., 2020). In short, cells lacking detectable mRNA in two out of the three reference genes (B2M, RPL13A, GAPDH) were excluded from further analysis. Since it has been previously reasoned against normalization of expression data from single cells (McDavid et al., 2013; Arsenio et al., 2014), the gene expression results were not normalized to any reference gene (McDavid et al., 2013; Arsenio et al., 2014). A limit of detection (LOD) was defined as a ct value of 24 (Livak et al., 2013; Hipp et al., 2017), subsequently all ct values higher than the LOD were set to 24. mRNA expression levels were defined as $2^{\text{LOD}-\text{ct}}$ (Livak et al., 2013; Hipp et al., 2017). Thus the value of 1 indicates a cell without detectable mRNA for the respective gene. In the Real-Time PCR analysis software (Fluidigm) the definition of the peak detection ranges of melting curves were set as the median of the temperature peak ± 1.2 °C. The median of the temperature peak of all non-failed results in the factory setting for the gene of interest for all analyzed cells was used. The remaining settings for melting curve analysis were left unchanged in the software (Peak Sensitivity: 7; Peak Ratio Threshold: 0.8). The ct value was set to the LOD for all reactions that were marked as failed under these settings. The softwares Fluidigm SNP Genotyping Analysis (Version 4.3.2) and Fluidigm Real-Time PCR Analysis (Version 4.3.1) were used for analysis.

**QUANTIFICATION AND STATISTICAL ANALYSIS**
Graphs and plots were generated with GraphPad Prism 8 (GraphPad Software). Statistical analyses were performed using GraphPad Prism 8 and R (version 3.5). Mann-Whitney test was employed for comparison between two unpaired groups. Wilcoxon matched-pairs signed rank test was used for comparison between two paired groups. For comparisons including single cells from different individuals, a mixed effects linear regression model with a random intercept was used to take into account the intra-sample correlations. Correlations for data including single cells from different donors were performed by employing the method of calculating the correlation of repeated-measurements (package rmcorr) taking into account the intra-individual associations as described previously (Bakdash and Marusich, 2017). Detailed information can be found in the respective figure legends.