On the interaction of an anticancer trisubstituted naphthalene diimide with G-quadruplexes of different topologies: a structural insight

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ABSTRACT

Naphthalene diimides showed significant anticancer activity in animal models, with therapeutic potential related to their ability to strongly interact with G-quadruplexes. Recently, a trifunctionalized naphthalene diimide, named NDI-5, was identified as the best analogue of a mini-library of novel naphthalene diimides for its high G-quadruplex binding affinity along with marked, selective anticancer activity, emerging as promising candidate drug for in vivo studies. Here we used NMR, dynamic light scattering, circular dichroism and fluorescence analyses to investigate the interactions of NDI-5 with G-quadruplexes featuring either parallel or hybrid topology. Interplay of different binding modes of NDI-5 to G-quadruplexes was observed for both parallel and hybrid topologies, with end-stacking always operative as the predominant binding event. While NDI-5 primarily targets the 5′-end quartet of the hybrid G-quadruplex model (m-tel24), the binding to a parallel G-quadruplex model (M2) occurs seemingly simultaneously at the 5′- and 3′-end quartets. With parallel G-quadruplex M2, NDI-5 formed stable complexes with 1:3 DNA:ligand binding stoichiometry. Conversely, when interacting with hybrid G-quadruplex m-tel24, NDI-5 showed multiple binding poses on a single G-quadruplex unit and/or formed different complexes comprising two or more G-quadruplex units. NDI-5 produced stabilizing effects on both G-quadruplexes, forming complexes with dissociation constants in the nM range.

INTRODUCTION

Cancer is the second leading cause of death worldwide (1). Considering invasiveness of tumour cells and the fact that each tumour form requires ad hoc therapeutic regimen, fighting cancer is not trivial (2). In the last decades, scientific research moved forward in cancer treatment by developing targeted therapies, featured by low-to-null toxicity, as valid and effective alternatives to conventional chemotherapies (3–6). Several antitumor targets were discovered and, among them, a strong interest arose for non-canonical nucleic acid structures known as G-quadruplexes, found at oncogene promoters and telomeres (7–13). G-quadruplexes are secondary structures of DNA and/or RNA formed by stacking of cyclic planar arrangements of four guanines called G-quartets, and stabilized by metal cations (7–13). These peculiar nucleic acid architectures play key roles in the regulation of tumour-specific genes as well as in molecular pathways involved in uncontrolled proliferation mechanisms typical of all tumour types. Thus, selectively target-

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ing G-quadruplex structures in vivo represents a very general and promising anticancer strategy (7–12,14,15).

The appealing possibility to treat common features of different cancers without impairing normal cells stimulated the synthesis of large libraries of putative selective G-quadruplex ligands. To rapidly and effectively select ‘true hits’, an affinity chromatography-based method, i.e. the G4-CPG (G-quadruplex on Controlled Pore Glass) assay, has been recently developed to identify ligands able to specifically recognize biologically relevant G-quadruplex structures (16–20). More specifically, we recently focused on a library of new multifunctionalized naphthalene diimides (NDIs) (21). NDIs have a remarkable potential as anticancer drugs because of their well-proven ability to interact with G-quartets (22–26). Moreover, their effective antitumor activity was demonstrated in several in vivo studies (27–33). By exploiting the G4-CPG assay, NDI-5 (Figure 1) was found to be the most attractive compound within the investigated series, and thus further analyzed by circular dichroism (CD) and fluorescence experiments (21). Noteworthy, biological assays underpinned NDI-5 as a promising candidate drug due to its strong and selective activity against cancer cells, showing IC₅₀ value of 79 nM on HeLa cells (21).

Encouraged by these results, we deemed it essential to undertake an in-depth study on the interaction of NDI-5 with G-quadruplexes to clarify the structural details of this strong and specific binding. Here, we reported a detailed NMR investigation on the NDI-5 interactions with G-quadruplexes of different structural topologies, complemented with dynamic light scattering (DLS), CD and fluorescence spectroscopy analyses. The DNA models of choice comprised G-rich oligonucleotides able to fold into stable and well-characterized G-quadruplex structures, i.e. M2 and m-tel24, respectively exhibiting parallel and hybrid-1 type topologies (Figure 2A and B) (34,35).

The primary sequences of the herein used model oligonucleotides, M2 d(TAGGGACGGGCGGGAGGG) and m-tel24 d[TTGGG(TTAGGG)₃A], are in line with vastly accepted consensus motif for G-quadruplex formation. M2 G-quadruplex exhibits all strands in parallel orientation, a common feature of most G-quadruplex-forming DNA segments in oncogene promoter regions (34). On the other hand, m-tel24 comprises the sequence originating from human telomeric DNA which has been only slightly modified to favour hybrid-1 G-quadruplex topology (35) and reduce the high polymorphism of telomeric DNA, thus allowing a detailed NMR investigation (36,37). Structural characterization of G-quadruplex/NDI-5 complexes was aimed at elucidating the binding of NDI-5 to both parallel and hybrid topologies, so to better guide the future rational design of optimized NDI-5 analogues in the quest for effective anticancer drugs.

MATERIALS AND METHODS

Sample preparation

Non-labelled, modified and residue-specific low-enrichment (10%) ¹⁵N- and ¹³C-labelled DNA oligonucleotides were synthesized on H-8 synthesizer (K&A LaborGeräte) with the use of standard phosphoramidite chemistry and then deprotected with aqueous ammonia. Purification and desalting of DNA oligonucleotides were performed by means of Amicon-15 centrifuge filter with 3.0 kDa MWCO. The oligonucleotide concentration was determined by measuring the absorbance at 260 nm and 90°C, using the appropriate molar extinction coefficients (38). NDI-5 was synthesized and purified as previously described (21). The NDI-5 stock solution was prepared by dissolving the solid compound in aqueous solution at 100 mM KCl, 20 mM potassium phosphate buffer (pH 7), 90%/10% H₂O/D₂O, at 20 mM concentration of the ligand for NMR studies or in pure water at 4 mM concentration of the ligand for DLS, CD and fluorescence analyses.

NMR experiments

NMR data were collected on Agilent NMR Systems 600 and 800 MHz and on Bruker AVANCE NEO 600 MHz
NMR spectrometers in the range 0–50°C. Oligonucleotide samples were prepared in 100 mM aqueous KCl, 20 mM potassium phosphate buffer (pH 7), 90%/10% H₂O/D₂O, at 0.2 mM oligonucleotide concentration per strand. In titration experiments, aliquots of the NDI-5 stock solution were directly added to the oligonucleotide solutions inside the NMR tube. NMR spectra were acquired with the use of the DPGFSE solvent suppression method. Unambiguous assignment of imino, aromatic and methyl ¹H NMR resonances was done with the aid of ¹⁵N- and ¹³C-HSQC spectra. Between sixty and twenty different gradient strengths (1–60 G·cm⁻¹) were used in DOSY NMR experiments. From translation diffusion coefficients (D) obtained from DOSY experiments, hydrodynamic dimensions for M2 and m-tel24 in the absence and presence of NDI-5 was calculated by using Stokes–Einstein equation (39):

\[ D_\alpha = \frac{k_B T}{3\pi \eta D_h} \quad \rightarrow \quad D_h = \frac{k_B T}{3\pi \eta D_\alpha} \]

where \( D_\alpha \) is the hydrodynamic diameter, \( k_B \) is the Boltzmann constant (1.38 × 10⁻²³ m²·kg·K⁻¹·s⁻²), \( T \) is the temperature (298.15 K), \( \eta \) is the fluid viscosity for 90%/10% H₂O/D₂O solution, calculated as previously described (0.917 mPa·s) (40) and \( D_h \) is the experimentally determined diffusion coefficient. NOESY spectra were acquired at mixing times between 80 and 500 ms. TOCSY spectrum was acquired at mixing time of 60 ms. ROESY spectra for NDI-5 and for M2 in presence of NDI-5 were acquired at mixing time in the range from 60 to 300 ms. For the NMR-monitored experiments evaluating the ability of NDI-5 to induce M2 G-quadruplex structuring in metal cation-free buffers, a 0.2 mM solution of M2 oligonucleotide was prepared in 90%/10% H₂O/D₂O and 6 molar equivalents of NDI-5 added to the oligonucleotide solution. DSS (4,4-dimethyl-4-silapentane-1-sulfonic acid) was used as a reference to calibrate the chemical shifts, assuming that DSS resonates at 0.0 ppm. NMR spectra were processed and analyzed with the use of VNMRJ (Varian Inc.), TopSpin 4.0.7 (Bruker) and Sparky (UCSF) softwares.

DLS experiments

DLS measurements were performed on a Zetasizer Nano ZS. M2 and m-tel24 samples were prepared at 1 mg/mL concentration respectively corresponding to 0.16 and 0.13 mM in 100 mM aqueous KCl and 20 mM potassium phosphate buffer (pH 7) and titrated with increasing amounts of NDI-5 (from 1 to 6 molar equivalents). The experiments were carried out at 25°C at a scattering angle \( \theta \) of 175° (backscatter detection). The diffusion coefficient of each population was calculated from the correlation function. The Stokes-Einstein equation was then used to evaluate the hydrodynamic diameter of free M2 and m-tel24, as well as of their complexes with NDI-5, from the related diffusion coefficients (39). Errors on hydrodynamic diameter were calculated as standard deviation of three independent measurements.

CD experiments

CD spectra were recorded in quartz cuvettes with a path length of 1 or 0.1 cm on a Jasco J-715 spectropolarimeter equipped with a Peltier-type temperature control system (model PTC-348W1). The spectra were recorded at 20°C in the range 240–800 nm with 2 s response, 200 nm/min scanning speed, 2.0 nm bandwidth, and corrected by subtraction of the background scan with buffer. All the spectra were averaged over 3 scans. Oligonucleotide samples were dissolved in a 20 mM aqueous KCl and 5 mM potassium phosphate buffer (pH 7) to obtain 2 μM solutions, then annealed by heating at 95°C for 5 min, followed by slow cooling to room temperature. CD titrations were obtained by adding increasing amounts of the ligands to M2 or m-tel24 G-quadruplexes. The ligand was added to each oligonucleotide solution until saturation of the oligonucleotide CD signals was achieved, which corresponded to 6 molar equivalents. After each ligand addition, the system was allowed equilibrating before recording the spectra. For the CD-melting experiments, the ellipticity was recorded at 265 or 290 nm, respectively for M2 and m-tel24, with a temperature scan rate of 0.5°C/min in the range 20–90°C. For the CD-monitored experiments evaluating the ability of NDI-5 to induce G-quadruplex structuring in metal cation-free buffers, 20 μM solutions of M2 or m-tel24 oligonucleotides were prepared in 10 mM Tris–HCl buffer (pH 7) and m-tel24 was titrated with increasing amounts of NDI-5 (up to 6 molar equivalents). The melting curve for the 1:6 m-tel24:NDI-5 ratio mixture was recorded at 290 nm, with a temperature scan rate of 0.5°C/min, in the range 10–90°C.

Fluorescence experiments

Fluorescence spectra were recorded at 20°C on HORIBA Jobin Yvon Inc. FluoroMax®-4 spectrophluorimeter equipped with F-3004 Sample Heater/Cooler Peltier Thermocouple Drive, by using a quartz cuvette with a 1 cm path length. NDI-5 was excited at 526 nm and emission spectra were recorded between 540 and 800 nm. Both excitation and emission slits were set at 5 nm. The experiments were performed in 20 mM aqueous KCl and 5 mM potassium phosphate buffer (pH 7). For fluorescence titration experiments, 2 μM ligands solutions were prepared in 20 mM KCl and 5 mM potassium phosphate buffer (pH 7). Increasing amounts of M2 or m-tel24 (up to 4 μM concentration) were added from 120 μM stock solutions of each DNA sample annealed in 20 mM KCl and 5 mM potassium phosphate buffer (pH 7). The obtained points were fitted with an independent and equivalent-sites model (41) using the Origin 8.0 program with the following equation:

\[
\alpha = \left( \frac{1}{\pi k_B T} \right) \left[ \left( [L]_0 + n[Qu]] + \frac{1}{k_B T} \right) \right. \\
- \left. \sqrt{\left( [L]_0 + n[Qu]] + \frac{1}{k_B T} \right) ^2 - 4[L]_0 n[Qu]] \right] 
\]

where \( \alpha \) is the molar fraction of ligand in the bound form, \( [L]_0 \) is the total ligand concentration, \([Qu]]\) is the added oligonucleotide concentration, \( n \) is the number of the equivalent and independent sites on the target DNA structure.
RESULTS AND DISCUSSION

NMR studies

NMR analysis of the G-quadruplex models and naphthalene diimide NDI-5. The interaction of NDI-5 with parallel M2 and hybrid-1 m-tel24 G-quadruplexes was studied at 100 mM KCl and 20 mM potassium phosphate buffer (pH 7.0). Under these conditions, both models gave high quality NMR spectra, with the predominant G-quadruplex conformation accounting in solution for >95% of the whole nucleotide material. Twelve narrow and well-resolved imino proton signals were observed in the $^1$H NMR spectrum of M2, consistent with the formation of a G-quadruplex with three G-quartets (Figure 3, bottom). In detail, M2 adopts a parallel G-quadruplex consisting of three stacked G-quartets, i.e. G3:G8:G12:G17, G4:G9:G13:G18 and G5:G10:G14:G19, sandwiched between the 5′- and 3′-end flanking residues (T1-A2 and T20, respectively) and connected by three propeller-type loops involving A6-C7, C11 and C15-A16 residues (Figure 2A). Notably, the M2 G-quadruplex studied herein showed identical structural features as the one analyzed in a previous study, adopting the same experimental conditions (34).

In comparison, our NMR analysis on m-tel24 was carried out at slightly higher K+ concentration than in previously published studies (100 mM KCl versus 70 mM KCl) (35). Hence, we used 2D-NOESY NMR experiments (Supplementary Figure S1) combined with $^1$H NMR analysis of oligonucleotides carrying the m-tel24 sequence containing specific thymine-to-uracil substitutions (necessary to assign the methyl groups of the thymidines in the sequence, see Supplementary Figure S2) to verify that the here used conditions did not alter the peculiar structural features of m-tel24 G-quadruplex. The obtained $^1$H NMR chemical shifts of m-tel24 G-quadruplex are reported in Supplementary Table S1. Overall, our results show that, under the used conditions, m-tel24 adopts a G-quadruplex structure with hybrid-1 type topology (Figure 2B), comprising G3:G21:G17:G9, G4:G10:G16:G22, G5:G11:G15:G23 quartets, whereby G3, G9, G15, G16 and G21 are in syn conformation along the N-glycosidic bond and the other residues exhibit anti conformation. The core of the structure is connected by one propeller and two lateral T-T-A loops, while the 5′- and 3′-ends comprise T1-T2 and A24 flanking residues, respectively (Figure 2B). In this regard, m-tel24, studied herein at 100 mM KCl, exhibits the same structural features as previously reported at 70 mM KCl (35).

Before studying the interaction of NDI-5 with both G-quadruplex models, the ligand alone was analyzed in solution by NMR. NDI-5 $^1$H NMR signals (Supplementary Figure S3) were assigned by means of 2D NMR experiments (Supplementary Figure S4). Similarly to other structurally related NDIs (24), under the studied neutral pH conditions, NDI-5 is likely to be in its dicationic form (Supplementary Figure S4A). Interestingly, among the exchangeable protons of the free ligand, i.e. 4, 9, 18 and 19, only 4 was observed (at δ 10.02 ppm, see Supplementary Figure S3). This can be attributed to the intramolecular hydrogen bond with the close inside oxygen atom, with which proton 4 can form a stable 6-membered cyclic structure (Supplementary Figure S4A). Noteworthy, the $^1$H NMR spectra of NDI-5 did not show any detectable change if recorded on freshly prepared or aged samples, thus confirming the chemical stability of NDI-5 in solution over time (Supplementary Figure S5).

NMR study of the interactions between NDI-5 and M2 G-quadruplex. The interaction between M2 G-quadruplex and NDI-5 was monitored by $^1$H NMR titration experiments. Upon addition of 0.5 molar equivalents of NDI-5, broadening of the $^1$H NMR imino signals corresponding to the free M2 G-quadruplex was observed, as well as appearance of new broad signals in the range of δ 10.67–10.78 ppm (Figure 3).

Further NDI-5 additions up to 1:2 DNA:ligand ratio resulted in sharpening of the signals in the δ region 10.67–10.78 ppm and appearance of an additional signal at δ 11.13 ppm. At 1:3 DNA:ligand ratio twelve imino signals were resolved, corresponding to G-quadruplex/NDI-5 complex, while the signals of free M2 G-quadruplex were no longer observed. The new set of twelve imino $^1$H NMR signals appearing at 1:3 DNA:ligand ratio was shifted upfield with respect to the signals of free M2 G-quadruplex, indicating the formation of defined M2/NDI-5 complexes (Figure 3). Only moderate shifts in the $^1$H NMR imino signals were observed upon increasing the DNA:ligand ratio from 1:3 to 1:6, while further addition of NDI-5 induced signal broadening again, whereby at 1:10 DNA:ligand ratio signals were hardly detectable from the baseline (Figure 3). Similar behaviour—i.e. $^1$H NMR signals broadening upon addition from 0.5 to 2 eq. of NDI-5, sharpening from 3 to 6 eq. and broadening from 7 to 10 eq.—was observed in the aromatic, anomeric and methyl regions of the oligonucleotide (Supplementary Figures S6–S8). Moreover, at 0.2 mM DNA concentration and 1:7 DNA:ligand ratio, a precipitate formed in the NMR tube. No $^3$H NMR spectral changes were observed upon monitoring the sample at 1:6 M2:NDI-5 ratio for several weeks, indicating that the complexes obtained during the titration experiments were stable over a long time (Supplementary Figure S9). Severe broadening of $^3$H NMR signals was observed for the sample at 1:6 M2:NDI-5 ratio upon lowering the temperature from
25 to 0°C (Supplementary Figure S10), consistent with the presence of different complexes at equilibrium of M2 G-quadruplex with NDI-5.

In order to get a deeper insight into the interactions between M2 G-quadruplex and NDI-5, 2D NMR experiments were performed on the samples at two different DNA/ligand ratios, i.e.: (i) 1:3, where the complete set of twelve $^1$H NMR signals corresponding to M2 G-quadruplex/NDI-5 complexes firstly resolved and (ii) 1:6, the highest ratio that could be explored in solution, considering that, at higher ligand concentration, sample precipitation occurred.

To unambiguously assign $^1$H NMR resonances for 1:3 and 1:6 M2:NDI-5 ratio samples, residue-specific $^{15}$N- and $^{13}$C-labelled M2 oligonucleotides were synthesized and analyzed by $^{15}$N- and $^{13}$C-HSQC experiments (Supplementary Figures S11-S18). $^1$H NMR chemical shifts of M2 G-quadruplex in the absence and presence of 3 or 6 molar equivalents of NDI-5 are reported in Supplementary Tables S2-S4. Addition of 3 molar equivalents of NDI-5 triggered changes in $^1$H NMR chemical shifts of M2 G-quadruplex, with the most prominently affected imino protons corresponding to G5, G10, G14 and G19 of the 3′-end quartet of M2 G-quadruplex, and to G8 and G12 of the 5′-end quartet (Figures 3 and 4). Interestingly, the same trend in $^1$H NMR chemical shift perturbations of imino protons was observed at 1:6 M2:NDI-5 ratio, i.e. the chemical shift differences observed upon addition of NDI-5 were larger, intermediate and smaller for guanines of the 3′-end quartet, of the middle quartet and of the 5′-end quartet, respectively, with the only exception of G8 and G12 (Figure 4). Notably, lower chemical shift differences observed for G3 and G17 when compared to G8 and G12 in the upper quartet could be attributed to the presence of T1 and A2 flanking residues, partly shielding G3 and G17, which in turn renders G8 and G12 more accessible to ligand binding.

Considering the aromatic, methyl and H1/H2′ sugar protons, the highest changes in $^1$H NMR chemical shifts upon addition of NDI-5 occurred at T1, A2 and T20 residues (Supplementary Figures S19 and S20). Interestingly, the chemical shift differences for aromatic and methyl protons were positive for T1 and negative in the case of A2 and T20, indicating that NDI-5 binding to M2 G-quadruplex promotes $\pi-\pi$ interactions of T1 coupled with repositioning of A2 and T20, which in turn exhibit weaker stacking to nearby G-quartets. These results are consistent with binding of NDI-5 to both the 5′- and 3′-end quartets, thus perturbing the original stacking interactions of flanking segments on the two external G-quartets. Interestingly, new resolved $^1$H NMR signals appearing upon addition of 0.5 molar equivalents of NDI-5 corresponding to the bound form of M2 G-quadruplex are those of G5, G10, G14 and G19 in the imino region (Figure 3) and T20 in the methyl region (Supplementary Figure S8). Notably, however, analysis of the $^1$H NMR signals at the low M2:NDI-5 ratio showed that binding of NDI-5 plausibly occurs to both 5′- and 3′-
end quartets of M2 G-quadruplex. Assuming a stoichiometry of binding of 1:3, the NMR data indicate that the first binding events involve stacking of the aromatic core of NDI-5 on the 5′- and 3′-end quartets (Figure 5A and B), while third binding comprises different poses including binding to outer G-quartets (Figure 5C and D) and to A6-C7 (Figure 5E) and C15-A16 loops (Figure 5F). Binding in proximity of the one-residue loop C11 and/or the loop-free groove at G3, G4, G17 and G18 may not be favoured or at least is less important if considering the perturbations in the aromatic 1H NMR chemical shifts upon titration experiments.

Interesting data were obtained by analysing increasing dilutions of 1:6 M2:NDI-5 ratio sample prepared at 0.2 mM DNA concentration. In these experiments, 1H NMR resonances progressively shifted downfield, approaching the chemical shift values, signals shape and resolution similar to that of 1:3 M2:NDI-5 ratio sample, thus further supporting formation of the 1:3 complex (Supplementary Figure S21). The NMR results show that, at higher than 1:3 DNA:ligand ratios, the species comprising M2 G-quadruplex with three bound ligand molecules represents a platform for additional binding events in which NDI-5 from bulk solution tends to interact with DNA or pre-bound ligand molecules in a ‘ligand-shell’ manner (vide infra for results of complementary methods). Upon dilution of the 1:6 system, NDI-5 molecules are progressively released from the DNA/ligand complexes, resulting in M2:NDI-5 complexes featuring three bound NDI-5 molecules (Figure 5C–F). Inspection of the differences in 1H NMR chemical shifts corresponding to G-quadruplexes:NDI-5 complexes at 1:6 versus 1:3 ratios showed the highest differences for imino protons of G3, G4, G5, G8, G9 and G10 and for aromatic and/or methyl protons of T1, A6, G9, G12 and A16 (Supplementary Figures S22 and S23). These findings suggested that the bound NDI-5 molecules released upon dilution are localized close to the 5′-end and the A6-C7 and C15-A16 loops (Figure 5G–J).

Upfield shifts of 1H NMR signals of imino and aromatic protons for all residues except T1 and T20 were observed upon increasing the DNA:NDI-5 ratio from 1:3 to 1:6. Notably, negative differences in chemical shift changes for H6 and methyl protons of T1 indicate that binding of the additional ligand molecules promotes de-stacking of T1. On the other hand, T20 was not significantly affected by further addition of NDI-5, which indicates that no additional binding occurred at the 3′-end upon increasing the M2:NDI-5 ratio from 1:3 to 1:6. Moreover, the binding of additional NDI-5 molecules caused opposite perturbations in chemical shifts at DNA:ligand ratios higher than 1:3 with respect to the trend observed on going from 1:0 to 1:3 M2:NDI-5 ratio for T1, A2, G5 and G19 residues, indicating that T1 was less involved, while A2, G5 and G19 were more involved in π–π stacking interactions at 1:6 than 1:3 M2:NDI-5 ratio. These findings are consistent with multiple and mutually dependent binding events, which differently affect M2 G-quadruplex as a function of the NDI-5 concentration, with ligand excess orchestrating the nucleotide residue re-arrangement within the overall G-quadruplex structure.

Diffusion ordered spectroscopy (DOSY) experiments were performed on M2 G-quadruplex in absence and presence of 3, 6 and 9 equivalents of NDI-5 (Supplementary Figures S24-S26). DOSY experiments at 9 molar equivalents of NDI-5 were performed at 0.1 mM DNA concentration, thus avoiding precipitation phenomena. A translation diffusion coefficient (D) of 1.5 (± 0.1) × 10−10 m2·s−1 was obtained for both the free M2 G-quadruplex and G-quadruplex/ligand samples containing 3 and 9 NDI-5 molar equivalents, and of 1.4 (± 0.1) × 10−10 m2·s−1 for the G-quadruplex/ligand sample including six NDI-5 molar equivalents. From the D values found for the M2 G-quadruplex alone, calculations based on a spherical model (39) provided a hydrodynamic diameter of 3.2 (±0.2) nm (including the hydration layer of 2 × 0.28 nm (42)), which is in full agreement with the value predicted by HYDROPRO software (43) (3.2 nm) for M2 G-quadruplex (PDB entry 5NYS) (34). Similar hydrodynamic diameter values were derived for M2 G-quadruplex in the presence of 3, 6 and 9 molar equivalents of NDI-5, i.e. 3.2 (±0.2) nm for the 1:3 and 1:9, and 3.4 (±0.2) nm for the 1:6 system. Overall, these findings indicate that the M2 G-quadruplex retains a monomeric fold even when bound to the ligand.
Figure 5. Schematic representation of the binding poses involved in the M2/NDI-5 complex formation, as inferred from NMR experiments. Free M2 and 1:1 M2/NDI-5 complex formed upon titrating M2 up to 1:0.5 M2/NDI-5 ratio are represented in (A) and (B), respectively. Putative 1:3 and 1:6 M2/NDI-5 complexes are depicted in (C)–(F) and (G)–(J), respectively.

For the NDI-5-containing systems, the absence of substantial changes in M2 G-quadruplex hydrodynamic dimensions compared to the free M2 G-quadruplex could be explained assuming that the overall G-quadruplex structure is rendered more compact and stiff upon binding, thus compensating the increase in size due to the ligand complexation.

Additionally, 2D NMR experiments, i.e. NOESY (Supplementary Figures S27 and S28) and TOCSY (Supplementary Figure S29) experiments, were performed on 1:3 and 1:6 M2:NDI-5 ratio samples in order to assign 1H NMR signals and characterize the interactions between M2 G-quadruplex and NDI-5. Higher resolution together with a higher number of NOE cross-peaks for the 1:6 M2:NDI-5 system in comparison to the 1:3 one demonstrated that saturating the complex with a ligand excess was beneficial for the spectroscopic characterization. The position of the ligand at both external G-quartets was supported by several DNA/NDI-5 intermolecular NOE cross-peaks summarized in Supplementary Table S5. Moreover, intermolecular NDI-5/NDI-5 interactions are sustained by a number of NOE cross-peaks between protons of different NDI-5 molecules (Supplementary Table S6). However, the intermolecular NDI-5/NDI-5 NOE cross-peaks were observed with very low intensities and only at mixing time of 500 ms. While broadening of 1H NMR signals precluded structural insights at low temperatures (Supplementary Figure S10), progressive heating of the 1:6 M2:NDI-5 ratio sample from 25 to 50°C allowed sharpening the 1H NMR signals, enabling more detailed analysis (Supplementary Figures S30 and S31). Notably, NDI-5/NDI-5 NOE cross-peaks resolved at 50°C (Supplementary Figure S32), thus allowing inspection of the interactions among NDI-5 molecules. This extended the details observed at 25°C, in particular the intermolecular interactions of the ligands aromatic protons and different pendant groups, altogether demonstrating equilibrium of different NDI-5/NDI-5 interacting modes. Finally, on the basis of this assignment of NDI-5 protons in the complexes with M2 G-quadruplex, differences in chemical shifts between free and bound NDI-5 were calculated and reported in Supplementary Figure S33. All NDI-5 protons were significantly perturbed upon binding ((Δδ)free-bound > 0.05 ppm) indicating that the interaction with M2 G-quadruplex affected the whole ligand molecule.

Eventually, the ability of NDI-5 to induce the formation of G-quadruplex structures starting from unfolded M2 oligonucleotide was evaluated. Interestingly, upon addition of 6 molar equivalents of NDI-5 to the oligonucleotide dissolved in a metal cation-free solution, new broad signals in the range of δ 10.56–11.10 ppm appeared (Supplementary Figure S34). This result indicates that NDI-5 induces formation of G-quadruplex structures in absence of stabilizing K+ ions, with different DNA-ligand complexes in equilibrium among them.

NMR study of the interactions of NDI-5 with modified M2 G-quadruplexes containing abasic sites in the loops. In order to investigate the role of the A6-C7 and C15-A16 loop residues in the interaction of M2 parallel G-quadruplex with NDI-5, we synthesized modified M2 oligonucleotides containing natural-to-abasic residue modifications in the first loop (M2-L1), in the third (M2-L3) or in both loops (M2-L1L3) (Supplementary Figure S35). Notably, G-quadruplexes containing natural-to-abasic residues substitutions in loop regions were expected to maintain the original parallel folding topology (44). Considering that C11, comprised in the second loop, exhibited only minor 1H NMR chemical shift perturbations upon titration experiments, and that no NOE correlations were observed between C11 and NDI-5, the M2 analogue with natural-to-abasic substitution in the second loop was not included in our study. Inspection of NMR parameters and in particular comparison of 1H NMR chemical shifts, DOSY-derived hydrodynamic diameters and NOE correlations for the M2-L1, M2-L3 and M2-L1L3 with respect to M2 demonstrated that all the modified oligonucleotides in the presence of K+...
nons adopted monomeric G-quadruplex folds with similar structural features as the parent M2. Notably, relevant \(^1\)H chemical shift differences (i.e., \(\Delta \delta > 0.05 \text{ ppm}\)) between the M2 G-quadruplex and the modified analogues were found only for imino signals of G4 and G13 and aromatic signals of G5, G8 and G9, in addition to loop sugar protons (Supplementary Figures S36–S38). However, careful comparative analysis of hydrodynamic diameters (Supplementary Table S7) and NOE connectivities (Supplementary Figure S39) for M2-L1, M2-L3 and M2-L1L3 with respect to M2 indicated that all these models shared common structural properties.

Titrations of M2-L1, M2-L3 and M2-L1L3 with NDI-5 monitored by \(^1\)H NMR analysis (Supplementary Figures S40–S48) resulted in gradual appearance of twelve well-resolved imino protons corresponding to G-quadruplex/ligand complexes in each case. The imino \(^1\)H NMR signals corresponding to these complexes were upfield shifted with respect to the signals of the free G-quadruplexes and appeared at 1:4, 1:4 and 1:3 G-quadruplex/ligand ratios for M2-L1, M2-L3 and M2-L1L3, respectively. Interestingly, narrowing of \(^1\)H NMR signals in the case of M2-L1 and M2-L3 occurred upon addition of one additional molar equivalent of NDI-5 with respect to M2-L1L3 and M2, i.e. 1:4 versus 1:3 DNA:ligand ratio, suggesting for the latter cases an interesting role of the nucleobases of the loops in the binding events. We interpret these differences in the \(^1\)H NMR spectra upon NDI-5 titration experiments considering the presence of four putative binding sites on each G-quadruplex, hereafter named \(A, B, C\) and \(D\). \(A\) and \(B\) correspond to the first NDI-5 binding sites at the 5'- and 3'-end quartets (see Figure 5B) for all the investigated M2, M2-L1, M2-L3 and M2-L1L3 G-quadruplexes. In the case of M2 as well as of M2-L1L3, binding to \(C\) (see 1:3 M2:NDI-5 complexes in Figure 5C–F) is favoured with respect to \(D\), which is not occupied by NDI-5 at 1:3 DNA:ligand ratio and may correspond to the binding site(s) where NDI-5 interacts through pre-bound NDI-5 (see 1:6 M2:NDI-5 complexes in Figure 5G–J). Regarding M2-L1 and M2-L3, the presence of abasic residues imposes similar binding affinities of NDI-5 to \(C\) and \(D\), so that binding site preference for NDI-5 is as follows: \(A = B > C = D\). Moreover, observation of \(^1\)H NMR signal sharpening at 1:4 rather than 1:3 DNA:ligand ratio in case of M2-L1 and M2-L3 is consistent with the equilibrium among species exhibiting NDI-5 bound to \(A, B, C\) and \(A, B, D\), which is intermediate on the NMR time scale at both 600 and 800 MHz. Furthermore, the fact that the M2-L1L3 G-quadruplex, containing abasic residues in two loops, shows similar NDI-5 binding profiles as the parent M2 G-quadruplex suggests that the nucleobases in the loops connecting the core of the structure do not crucially impede binding, but may orchestrate the dynamics of DNA remodelling in the sequential ligand binding.

The analysis of \(^1\)H NMR chemical shift differences for imino, aromatic, methyl and sugar protons corresponding to M2-L1, M2-L3 and M2-L1L3 in the absence or presence of 6 molar equivalents of NDI-5 revealed that changes induced by NDI-5 binding are similar as for M2 (Supplementary Figures S49–S51), consistent with NDI-5 interacting similarly with all the investigated G-quadruplexes. Moreover, DOSY experiments proved that the overall size, and thus the monomeric G-quadruplex fold was preserved for M2-L1, M2-L3 and M2-L1L3 even after NDI-5 binding (Supplementary Table S7). To get a deeper insight into the binding mode of NDI-5 towards the modified M2 oligonucleotides, 2D-NOESY spectra were recorded for M2-L1, M2-L3 and M2-L1L3 G-quadruplexes in the presence of 6 molar equivalents of NDI-5, showing similar sequential correlations as M2 (Supplementary Figure S52). Interestingly, NMR spectra of the investigated G-quadruplexes in the presence of 6 molar equivalents of NDI-5 revealed two signals at ca. \(\delta 6.8\) and 7.0 ppm for M2 (Supplementary Figure S53), whereby in the case of M2-L1, M2-L3 and M2-L1L3 only the former, only the latter or no signal was observed respectively, suggesting that these signals were diagnostic for NDI-5 interaction with the loops. Additionally, analysis of intermolecular NOE cross-peaks between NDI-5 and all the investigated G-quadruplexes (Supplementary Tables S5 and S8) revealed similarities among all the studied complexes. Moreover, almost all the contacts between different NDI-5 molecules are common to M2-L1, M2-L3 and M2-L1L3 (as well as M2) G-quadruplexes at 1:6 DNA:ligand ratio (Supplementary Table S9). Finally, analysis of \(^1\)H NMR chemical shift differences between free and bound NDI-5 in 1:6 DNA:ligand ratio samples showed that the systems comprising M2-L1, M2-L3 and M2-L1L3 had an essentially similar behaviour as M2. Additionally, the analysis of \(^1\)H NMR chemical shift perturbations upon ligand binding revealed the following trend in terms of the differences between free and bound NDI-5: M2-L1 \(\geq\) M2-L3 \(\geq\) M2-L1L3 \(\geq\) M2 (Supplementary Figure S54). The consistently higher similarity in \(^1\)H NMR chemical shifts of bound NDI-5 in case of M2-L1L3 to the parent M2 in comparison to M2-L1 and M2-L2 indicates that, albeit subtly, A6-C7 and C15-A16 loop regions modulate the ligand binding mutually. Therefore, it can be concluded that in all cases the interaction with the investigated G-quadruplexes involves the whole ligand molecules and NDI-5 is more perturbed upon binding to M2-L1 and M2-L3 than to M2-L1L3 and M2. These data provide further confirmation that the most similar complexes to those found with the unmodified M2 G-quadruplex are the ones formed with M2-L1L3 G-quadruplex.

NMR study of the interactions between NDI-5 and m-tel24 G-quadruplex. Titration of m-tel24 G-quadruplex with NDI-5—performed under the same conditions used for M2 G-quadruplex, i.e. at 100 mM KCl and 20 mM potassium phosphate buffer (pH 7.0)—showed a stepwise broadening of the signals (Figure 6 and Supplementary Figure S55–S58).

In parallel, novel \(^1\)H NMR signals of m-tel24 appeared in the imino region (Supplementary Figure S55) upon increasing NDI-5 concentration. Notably, these signals were upfield shifted with respect to the free m-tel24 G-quadruplex and assigned to m-tel24/NDI-5 complexes. The binding of NDI-5 to m-tel24 was evident from dramatic line broadening of imino \(^1\)H NMR signals even at 0.5 molar equivalents (Figure 6) with more pronounced effect observed for G3,
G4, G9, G17, G21 and G22 (Supplementary Figure S59). Notably, G3, G9, G17 and G21 compose the 5’-end quartet of m-tel24 G-quadruplex, while G4 and G22 are comprised in the middle G-quartet. Line broadening of 1H NMR signals was also evident for the aromatic protons of G17, T19, A20 and G22 and methyl groups of T1, T2, T18 and T19 (Supplementary Figures S56 and S58). Overall, these findings point to a specific binding mode of NDI-5 to m-tel24 G-quadruplex at the 5’-end, as schematically shown in Figure 7. However, even at higher NDI-5 concentration, no sharpening of 1H NMR signals was observed, denoting that complex equilibria, intermediate-to-fast on the NMR time scale, occurred. No change in 1H NMR signals could be observed upon aging the sample at 1:6 m-tel24:NDI-5 ratio for several weeks (Supplementary Figure S60). Interestingly, the number of 1H NMR signals increased upon lowering the sample temperature from 25 to 0°C (Supplementary Figure S61), while several, especially aromatic 1H NMR signals were more resolved at temperatures above 25°C (Supplementary Figures S62 and S63). Nevertheless, the spectral resolution and severity of line broadening at both lower and higher temperatures than 25°C did not warrant more detailed structural characterization. Notably, no precipitate formation was observed even at m-tel24:NDI-5 1:20 ratio. Overall, the observed behaviour suggests that NDI-5 exhibits multiple binding poses on a single m-tel24 G-quadruplex unit and/or different m-tel24:NDI-5 complexes comprising two or more G-quadruplex units could be formed. For the m-tel24:NDI-5 system, the low intensity of the 1H NMR signals in the imino as well as in the other spectral regions discouraged further, more in-depth NMR experiments.

Dynamic Light Scattering experiments

In order to measure the hydrodynamic diameter of DNA:NDI-5 complexes, in addition to the DOSY experiments described in section 3.1.2, DLS analysis was carried out. In detail, aqueous solutions of M2 and m-tel24 G-quadruplexes were titrated with increasing amounts of NDI-5 (up to 6 molar equivalents) and DLS analysis performed after equilibration of each sample (Figure 8). The volume-based particle size distribution showed the presence of a single species for both M2 and m-tel24 G-quadruplexes in the absence of ligand, with hydrodynamic diameter of 3.1 (±0.5) and 3.2 (±0.4) nm, respectively. No significant variation of hydrodynamic diameter for M2 G-quadruplex was observed upon addition of NDI-5 (Figure 8A), in full agreement with DOSY experiments. As concerns m-tel24 titration, the addition of 1 molar equivalent of ligand led to the formation of m-tel24/NDI-5 complexes with a diameter of 4.9 (±0.8) nm (Figure 8B), indicating a drastic change in size, which may correspond to species comprising two G-quadruplex units whose interaction is mediated by NDI-5 molecules. Notably, the m-tel24/NDI-5 complexes including 1 and 2 molar equivalents of NDI-5 had the same size. From 3 to 6 molar NDI-5 equivalents, species with even higher hydrodynamic diameter (average value = 7.3 nm) were observed, compatible with higher order G-quadruplex structures (Figure 8B). The results obtained for the m-tel24:NDI-5 system showed different DNA/NDI-5 complexes—comprising one, two or more G-quadruplex units—in equilibrium. Notably, these results are consistent with the above described NMR behaviour of the m-tel24:NDI-5 system. Indeed, the 1H NMR signal broadening for m-tel24:NDI-5 system observed at ratios from 1:1 onwards can be well explained considering the formation of different m-tel24:NDI-5 complexes comprising two or more G-quadruplex units which are involved in intermediate-to-fast equilibria on the NMR time scale.
Circular dichroism experiments

Circular dichroism experiments were carried out in the absence and presence of NDI-5 to evaluate the effects of this ligand on the overall conformational behaviour of M2 and m-tel24. CD spectra showed that M2 folded into a parallel G-quadruplex, featured by a maximum at 265 nm and a minimum at 243 nm, while m-tel24 folded into a hybrid G-quadruplex, featured by a maximum at 290 nm and a shoulder at 270 nm (Figure 9A and B, respectively), confirming the expected conformations for these G-quadruplex structures under the explored solution conditions. Then solutions of M2 and m-tel24 G-quadruplexes were titrated with increasing amounts of NDI-5 and CD spectra recorded after each addition. Upon titrating M2 G-quadruplex with NDI-5, reduction in the intensity of the 265 nm band was observed (Figure 9A). When m-tel24 was treated with NDI-5, a dose-dependent increase of the intensity of the 290 nm band occurred as well as a reduction of the 270 nm shoulder (Figure 9B). Thus, conformational changes were detected for both the investigated structures, proving that NDI-5 somehow affected G-quartets and/or flanking segments stacking in both M2 and m-tel24 G-quadruplexes. However, the parallel fold was preserved for M2 G-quadruplex, while, interestingly, conformational changes were induced on m-tel24 G-quadruplex by the presence of NDI-5, plausibly indicating a switch from hybrid to antiparallel topology, similarly to what previously observed for titrations on parallel c-myc and hybrid tel26 G-quadruplexes (21).

CD-melting experiments were also performed to evaluate if stabilizing or destabilizing effects on the G-quadruplexes were obtained upon incubation with NDI-5. CD melting curves of M2 and m-tel24 in the absence or presence of the ligand (1:6 DNA:NDI-5 ratio) were recorded by following the CD changes at the wavelength of intensity maximum (265 and 290 nm, respectively). The CD melting experiments (Supplementary Figure S64) showed Tm of 78 and 85 (±1) °C for M2, and Tm values of 61 and 74 (±1) °C for m-tel24, respectively in the absence or presence of 6 ligand equivalents, proving that NDI-5 was able to produce stabilizing effects on both G-quadruplexes (ΔTm = +7 and +13°C, respectively). Then, in order to evaluate the ability of NDI-5 to induce the formation of G-quadruplex structures in both M2 and m-tel24 systems starting from the unfolded oligonucleotides in a metal cation-free buffer, additional CD experiments were carried out. In detail, a solution of unstructured m-tel24 oligonucleotide in Tris–HCl buffer (pH 7) was titrated with increasing amounts of NDI-5, and CD spectra recorded after each addition (Supplementary Figure S65A). Interestingly, NDI-5 was able to induce structuring of the unfolded m-tel24 oligonucleotide, which formed an antiparallel G-quadruplex structure featured by a Tm value of 45°C (Supplementary Figures S65A and S65B, respectively). On the contrary, no data about the NDI-5 ability to induce structuring of M2 could be gained, since this system proved to fold into a stable secondary structure, i.e. a parallel G-quadruplex, even in a metal cation-free buffer (Supplementary Figure S65C).

Fluorescence experiments

To get information about the binding stoichiometry and constants for the complexes formed between NDI-5 and M2 or m-tel24 G-quadruplex, fluorescence experiments were carried out.

Fluorescence titration experiments were performed at a fixed concentration of ligand, by adding increasing amounts of each G-quadruplex. In detail, NDI-5 solutions were titrated with increasing amounts of M2 or m-tel24 solutions, and the corresponding fluorescence spectra, after each addition, were recorded (Supplementary Figures S66A and B, respectively). A significant fluorescence quenching was observed in both titration experiments, further confirming the interaction between NDI-5 and the G-quadruplex structures. The fraction of bound NDI-5 was calculated from the obtained fluorescence intensity values and reported as a function of the DNA concentration (Figure 10A and B). In the absence of suitable mathematical models for an accurate description of complex systems including multiple binding events, data were fitted with one of the most validated models for the determination of G-quadruplex/ligand binding constants, i.e. the independent and equivalent-sites model (19,41). Considering the simplicity of the above model with respect to the complexity of our G-quadruplex/ligand systems, which – for their intrinsic asymmetry – cannot offer identical binding sites, the

Figure 8. Volume-based particle size distribution for (A) M2 and (B) m-tel24 G-quadruplexes in the absence and presence of different amounts of NDI-5 (up to 6 equivalents). DLS experiments were performed at 0.16 mM (for M2) or 0.13 mM (for m-tel24) DNA, 100 mM KCl, 20 mM potassium phosphate buffer (pH 7) and 25°C. Tables report the hydrodynamic diameters (±S.D.) for the different species formed on increasing NDI-5 concentration.
Naphthalene diimides are increasingly emerging as attractive, putative anticancer drugs due to their well-proven ability to bind cancer-related G-quadruplex structures (22–26). Recently, a new trifunctionalized G-quadruplex-targeting naphthalene diimide, named NDI-5, was discovered to have a selective antitumor activity in vitro in the low nM range, showing potential as a promising lead compound to develop future candidate drugs for in vivo studies (21). Despite the wide interest in this class of compounds as G-quadruplex ligands, to the best of our knowledge no in-depth NMR study on the interactions of naphthalene diimides with G-quadruplexes in solution had been carried out thus far.

Here, we reported the first extensive NMR study on the interaction of a naphthalene diimide—the trifunctionalized NDI-5— with G-quadruplex models adopting different topologies, i.e. a parallel-type (M2) and a hybrid-type (m-tel24) structure (34,35). To complement the NMR data, dynamic light scattering, circular dichroism and fluorescence analyses were also performed. This work provided details on the binding of this dicaticionic, trifunctionalized naphthalene diimide with model G-quadruplexes of parallel and hybrid type, characterized by interplay of different binding modes. NDI-5 was found to induce hybrid-to-antiparallel conformational changes in m-tel24, in turn not significantly altering the parallel G-quadruplex structure of M2. Upon binding, NDI-5 produced marked stabilizing effects on both G-quadruplexes ($\Delta T_m \geq +7^\circ C$). From fluorescence titration experiments, apparent dissociation constants of 42 and 143 nM were determined respectively for M2/NDI-5 and m-tel24/NDI-5 complexes, indicative of the high affinity of this ligand for both G-quadruplexes, but with some preference for the parallel topology.

As concerns M2 G-quadruplex, its monomeric fold was preserved upon ligand binding, with formation of con-
plexes characterized by binding stoichiometry of 1:3. NDI-5 preferentially bound M2 at the lower quartet and the 3′-end close residues, as well as at the upper quartet and the 5′-end close residues, in turn perturbing the stacking interactions of flanking nucleotides on the two external G-quartets of M2 G-quadruplex. The M2 loops containing two nucleobases proved to mediate the interaction between NDI-5 and the G-quadruplex. Conversely, the M2 loop containing a single residue was not involved in the ligand complexation. While direct stacking of the ligand on G-quartet surfaces appeared decisive for the interaction, the role of loops was notable in the context of interdependence of the secondary binding events: indeed, NDI-5 binding could be coupled with induced stacking interactions within A6-C7 and C15-A16, not accessible in the one-residue C11 loop. The binding of NDI-5 at the 5′-end of the parallel G-quadruplex included participation of T1 and A2 terminal residues, in agreement with previously resolved crystal structures of naphthalene diimides-G-quadruplexes complexes (25,53,54). Notably, the ligand core, with its rigid skeleton, could provide the main interface for NDI binding to a G-quadruplex via end-stacking mode, whereas the more flexible pendant groups offered the possibility of finely tuning the binding, also depending on the overall shape and nature of the loops. Interestingly, when abasic sites were introduced in A6-C7 and/or C15-A16 M2 G-quadruplex loops, the M2 G-quadruplex with abasic sites in both loops was the most similar to unmodified M2 in NDI-5 binding behaviour. This indicates that modifications at the level of a single propeller loop introduce changes in the overall symmetry of the M2 G-quadruplex structure that can be compensated upon insertion of abasic residues in two opposite propeller loops. Then, it has to be considered that not only the structural, but also the dynamic aspect of the interaction, in particular the targeted structure breathing, is crucial for the binding affinity and specificity of the ligand.

In the case of m-tel24 G-quadruplex, 1H NMR signal broadening observed for the complexes with NDI-5 suggested that multiple binding poses of the ligand on a single G-quadruplex unit and/or different complexes, comprising two or more G-quadruplex units, could be formed when targeting hybrid G-quadruplexes. DLS results confirmed these hypotheses, showing the formation of DNA/ligand higher-order species upon increasing the NDI-5 concentration.

In summary, although several binding sites and stoichiometries were observed with both M2 and m-tel24 G-
quadruplexes, both models showed preferential binding of NDI-5 at the outer G-quartets, with the initial binding events occurring at both 5′- and 3′-ends in the case of the parallel G-quadruplex and at the 5′-end for the hybrid G-quadruplex. This behaviour could be explained considering the high propensity of NDI-5 to interact with G-quadruplex targets mainly by stacking interactions. On this basis, the different degree of accessibility of the planar aromatic surfaces of G-quadruplex structures seems to be the main structural element directing NDI-5 binding preferences. Overall, this structural investigation will be helpful for the future design of optimized NDI-5 analogues as effective candidate drugs in anticancer targeted therapies, allowing the construction of reliable models for the G-quadruplex/NDI-5 complexes in docking studies aimed at the development of second-generation optimized NDI derivatives.

DATA AVAILABILITY

Biological Magnetic Resonance Data Bank ID: 50378.

SUPPLEMENTARY DATA

Supplementary Data are available at NAR Online.

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