RECQL5 Suppresses Oncogenic JAK2-Induced Replication Stress and Genomic Instability

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**Report**

**RECQL5 Suppresses Oncogenic JAK2-Induced Replication Stress and Genomic Instability**

**Graphical Abstract**

**Highlights**
- RECQL5 is upregulated in JAK2V617F mutant erythroblasts in MPN patients
- RECQL5 is a target of activated JAK2-PI3K signaling
- RECQL5 stabilizes stalled forks during JAK2V617F-associated replication stress
- RECQL5 depletion sensitizes JAK2V617F-expressing cells to hydroxyurea

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**In Brief**
Oncogenic JAK2 signaling in MPN patients leads to DNA damage, yet MPNs are characterized by genomic stability. Chen et al. show that the DNA helicase RECQL5 maintains genomic integrity in response to JAK2V617F-associated replication stress. Moreover, RECQL5 depletion sensitizes JAK2V617F-mutant cells to hydroxyurea, the most common treatment for MPN.

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SUMMARY

JAK2V617F is the most common oncogenic lesion in patients with myeloproliferative neoplasms (MPNs). Despite the ability of JAK2V617F to instigate DNA damage in vitro, MPNs are nevertheless characterized by genomic stability. In this study, we address this paradox by identifying the DNA helicase RECQL5 as a suppressor of genomic instability in MPNs. We report increased RECQL5 expression in JAK2V617F-expressing cells and demonstrate that RECQL5 is required to counteract JAK2V617F-induced replication stress. Moreover, RECQL5 depletion sensitizes JAK2V617F mutant cells to hydroxyurea (HU), a pharmacological inducer of replication stress and the most common treatment for MPNs. Using single-fiber chromosome combing, we show that RECQL5 depletion in JAK2V617F mutant cells impairs replication dynamics following HU treatment, resulting in increased double-stranded breaks and apoptosis. Cumulatively, these findings identify RECQL5 as a critical regulator of genome stability in MPNs and demonstrate that replication stress-associated cytotoxicity can be amplified specifically in JAK2V617F mutant cells through RECQL5-targeted synthetic lethality.

INTRODUCTION

The propensity of cancer cells to undergo clonal evolution is enabled by a heightened state of genomic instability wherein cancer cells are continuously accumulating and repairing DNA damage. This increase in genomic flux allows cancer cells to accumulate somatic mutations that can drive disease progression. However, heightened genomic instability can also activate DNA damage-associated checkpoints, which can lead to apoptosis or cellular senescence. Cancer cells continuously tread a fine balance between cell death and survival in response to DNA damage (Negrini et al., 2010).

Chronic myeloproliferative neoplasms (MPNs) encompass a spectrum of clonal hematological disorders with an inherent tendency to transform into a more aggressive disease in the form of acute myeloid leukemia (AML). MPNs provide a window into cancer early during its ontogeny and give insights into the processes that regulate genome stability during malignant clonal evolution. The most common recurrent lesion in MPN patients is an activating V617F mutation in the JAK2 non-receptor tyrosine kinase (JAK2V617F) that causes hyperactive JAK-STAT signaling and confers a capacity for cytokine-independent growth (Baxter et al., 2005; James et al., 2005; Kralovics et al., 2005; Levine et al., 2005). Recently, a growing body of work has suggested that JAK2V617F is associated with increased DNA damage: (1) increased numbers of γH2Ax-marked double-strand breaks (DSBs) have been detected in Ba/F3 pro-B cells overexpressing JAK2V617F (Marty et al., 2013) and in lineage-negative, Sca1-positive, c-Kit-positive (LSK) cells (enriched for hematopoietic stem cell [HSC] activity) from 6-month-old JAK2V617F-heterozygous knockin mice (Li et al., 2010); (2) JAK2V617F expression is associated with increased levels of DNA-damaging reactive oxygen species (Marty et al., 2013); (3) RAD51-positive foci indicative of increased DSB repair have been observed in CD34+ hematopoietic stem cell (HSC) activity from 6-month-old JAK2V617F-heterozygous knockin mice (Li et al., 2010); (2) JAK2V617F expression is associated with increased levels of DNA-damaging reactive oxygen species (Marty et al., 2013); (3) RAD51-positive foci indicative of increased DSB repair have been observed in CD34+ hematopoietic stem cells obtained from JAK2V617F-positive MPN patients (Plo et al., 2008); and (4) JAK2V617F expression in both human diploid fibroblasts and primary erythroblasts from MPN patients leads to higher rates of stalled replication forks, with improper processing of stalled replication intermediates representing a potential source of DSBs (Chen et al., 2014).
JAK2 signaling imposes a mutator phenotype on MPN cells, accelerating the accumulation of mutations and promoting clonal evolution and disease progression. However, longitudinal studies of MPN patients indicate that JAK2V617F-positive polycythaemia vera (PV) and essential thrombocythemia (ET) patients (i.e., chronic-phase MPN) typically remain clinically and cytogenetically stable over decades (Tefferi et al., 2014). A recent copy-number analysis of the genome of chronic-phase MPN patients has shown that cytogenetic abnormalities are rare (Klampfl et al., 2011), and an analysis of the mutational landscape of PV and ET patients revealed that each MPN patient harbors a modest number of mutations per exome (approximately 6.5) (Nangalia et al., 2013).

To reconcile the apparent paradox of JAK2V617F-induced DNA damage with the clinical and cytogenetic stability characteristic of chronic-phase MPN, we hypothesized that JAK2V617F, in addition to instigating a state of increased DNA damage, could also, in parallel, activate protective pathways that counteract and prevent DNA damage-induced apoptosis. In this report, we identify increased expression of the DNA repair helicase RECQL5 in JAK2V617F-expressing cells and characterize its role in constraining JAK2V617F-induced replication stress and maintaining genomic integrity in MPNs.

RESULTS

Activated JAK2 Signaling Regulates Expression of the RECQL5 Helicase in MPN Cells

We analyzed the gene expression profiles of autologous normal and JAK2V617F-heterozygous burst-forming unit erythroid (BFU-E) colonies from 40 MPN patients (Chen et al., 2010). To explore the hypothesis that JAK2V617F may activate genes that counteract DNA damage, we evaluated the expression of DNA helicases in JAK2 mutant and autologous normal cells in this dataset. Of 25 DNA helicases expressed, RECQL5 was the sole DNA helicase enzyme whose expression was elevated significantly in mutant-JAK2 BFU-Es relative to autologous BFU-Es in ET, PV, and myelofibrosis patients after multiple hypothesis testing (q < 0.05) (Figure 1A). Real-time qPCR analysis of 10 MPN patients (five PV patients and five ET patients) demonstrated increased expression of RECQL5 in HEL cells following treatment with 16 hr with INCB018424 (1 μM), the PI3K inhibitor PI103 (1 μM), or the ERK1/2 inhibitor U0126 (5 μM).

See also Figure S1.
concomitant with decreased STAT5 phosphorylation (Figure 1C). In aggregate, these data indicate that RECQL5 expression is regulated by activated JAK2 signaling in human disease-relevant contexts.

**Physiological Levels of Jak2V617F Lead to Increased Recql5 Expression**

To investigate the role of Recql5 under conditions that more closely recapitulate chronic-phase MPN in patients, we generated disease-relevant cell lines from Jak2V617F knockin mice that we developed previously and characterized extensively (Mullally et al., 2010, 2013). This mouse model closely recapitulates the features of human MPN, and Jak2V617F expression is physiologically in the model, being driven from the endogenous Jak2 promoter. To generate cell lines, we engineered bone marrow progenitors from littermate wild-type or Jak2V617F knockin mice to express a β-estradiol-regulated Hoxb8 homeodomain-containing protein. Following 3 weeks of serial passaging and antibiotic selection, immortalized myeloid progenitor cells were generated from both wild-type mice (WT-B8) and Jak2V617F knockin mice (VF-B8) (Wang et al., 2006). Both the WT-B8 and VF-B8 lines resemble cells at a granulocyte-macrophage progenitor (GMP) stage of myeloid maturation by cell surface immunophenotype analysis (Figure S1A). Immunoblotting showed equivalent levels of Jak2 expression in both cell lines but increased levels of phosphorylated Stat5 in VF-B8 cells relative to WT-B8 cells (Figure S1B). Additionally, VF-B8 cells exhibited an increased proliferative capacity and survival under reduced serum conditions (Figures S1C–S1D), indicating that VF-B8 cells recapitulate this key pathognomonic feature of MPN biology.

We next determined whether the expression of Recql5 was modulated by mutant Jak2 in these Hoxb8-immortalized cell lines. In accordance with the primary human data, VF-B8 cells exhibited elevated expression of Recql5 relative to WT-B8 cells by qPCR analysis (Figure 1D) and western immunoblots (Figure 1E). No significant differential expression of other Recq family members (Recql1, Recql4, Blm, and Wrn) was observed. These data demonstrate that mutant Jak2 increases expression of the DNA repair helicase RECQL5 in both human and murine cells.

**RECQL5 Is a Target of Jak2–PI3K Signaling**

To determine which signaling pathways were necessary for the regulation of Recql5 by Jak2V617F, we used small hairpin RNAs (shRNAs) to knock down Jak2 and downstream effector molecules of Jak2 in VF-B8 cells. Expression of shRNAs targeting Jak2 led to diminution of Recql5 levels relative to cells transduced with an empty vector control (Figure 1F), which is consistent with the finding that Recql5 is a downstream target of Jak2 signaling. Next, we assessed Recql5 levels in VF-B8 cells following transduction of shRNAs targeting Stat1, Stat5, or the p85 subunit of phosphatidylinositol 3-kinase (PI3K). The levels of Recql5 in VF-B8 cells were reduced following knockdown of PI3K but not Stat1 or Stat5 (Figure 1F). Similarly, RECQL5 levels were also attenuated following knockdown of Jak2 in HEL cells but were not attenuated following knockdown of STAT5 or STAT1 (Figure 1G), and treatment of HEL cells with the PI3K inhibitor PI103 (but not the ERK1/2 inhibitor U0126) led to attenuated levels of RECQL5, accompanied by decreased AKT phosphorylation (Figure 1H). Taken together, these data indicate that RECQL5 expression is increased upon JAK2 activation in a PI3K-dependent manner.

**Recql5 Depletion Sensitizes Jak2V617F-Expressing Cells to Replication Stress**

We next sought to clarify the function of RECQL5 upregulation in JAK2V617F-expressing cells. Previously, we had demonstrated that overexpression of JAK2V617F in human diploid fibroblasts resulted in increased replication fork stalling and replication stress (Chen et al., 2014). Because RECQL5 has been linked functionally to regulating stalled replication forks in normal cells, we hypothesized that the upregulation of RECQL5 may function to mitigate the deleterious consequences of replication stress in JAK2 mutant cells.

To test this, we knocked down Recql5 in VF-B8 cells using three independent shRNAs (Figure S2A). Densitometric analysis revealed a knockdown efficiency of 61%–95% for the hairpins. Under normal growth conditions, Recql5 knockdown in VF-B8 cells did not affect the proliferation rate (Figure S2B) or apoptosis (Figure S2C) relative to either equivalently modified WT-B8 cells or VF-B8 cells expressing empty vector alone (VA) controls. To simulate replication stress, we subjected WT-B8 and VF-B8 cells to low-serum conditions to deprive cells of deoxynucleotide triphosphate (dNTPs) (Bester et al., 2011). We observed that VA-expressing VF-B8 cells exhibited greater viability relative to WT-B8 cells under low-serum conditions (Figure 2A, left), which is in accordance with our previous data on non-genetically perturbed cells. In contrast, Recql5-depleted VF-B8 cells exhibited decreased viability relative to equivalently modified WT-B8 cells (Figure 2A, right). Hypersensitivity of Recql5-depleted VF-B8 cells to low-serum conditions was abrogated completely upon repletion of deoxynucleotide (dNTPs) into the culture medium (Figure 2B). The results shown represent the average of three independent Recql5-targeting shRNAs and are consistent with data obtained for each individual hairpin (Figure S2D). Collectively, these data demonstrate that Recql5 protects VF-B8 cells from endogenous replication stress instigated by low dNTP levels.

We next tested whether Recql5 depletion would also sensitize VF-B8 cells to exogenous instigators of replication stress, such as pharmacological agents. We tested hydroxyurea (HU) and camptothecin (CPT), which impair DNA replication by limiting production of dNTPs and inhibiting topoisomerase I activity, respectively. Strikingly, contemporaneous knockdown of Recql5 and exposure to either HU or CPT led to significantly decreased viability of VF-B8 cells compared with like-treated WT-B8 cells after 24 hr of drug treatment (Figures 2C, 2D, and 2G; Figure S2D). Decreased cell viability was noted as early as 12 hr post-drug treatment (Figures S2E and S2F). In contrast, Recql5-depleted VF-B8 cells were not hypersensitive to the double-stranded breaking agents doxorubicin (DOX) and etoposide (ETP) (Figures 2E and 2F; Figure S2D). Concordant with these data, shRNA depletion of human RECQL5 also enhanced cytotoxicity in HEL and SET2 cells, but only to pharmacological instigators of replication stress (HU, CPT, aphidicolin, and irinotecan) and not to pro-oxidants or DSB-generating drugs (Figures S2G and S2H).
Replication stressors such as HU and CPT can also induce DSBs at sufficiently high doses. We therefore validated whether the HU and CPT doses associated with preferential cytotoxicity of Recq5-depleted VF-B8 cells were causing increased replication stress or excessive formation of DSBs. To differentiate between these two phenomena, we treated parental WT-B8 and VF-B8 cells with HU (6 mM) and CPT (4 nM) and performed immunocytochemical staining for foci containing the RPA protein (which marks stalled replication forks) or 53BP1 (which localizes to DSBs) (Figures S3 A and S3B). At these HU and CPT dosages, we observed a marked increase in foci containing RPA (Figure S3C), with only a marginal increase in numbers of 53BP1-positive foci (Figure S3D). In contrast, etoposide (4 nM) generated both RPA-positive and 53BP1-positive foci (Figures S3C and S3D). In aggregate, these findings verify that the dosage of HU and CPT used to enhance the cytotoxicity of Recq5-depleted VF-B8 cells causes enhanced replication stress with only a slight elevation of DSBs.

Given the potential off-target effects of shRNAs that may potentially influence the observed phenotypes, we next confirmed the specificity of the Recq5 shRNAs. We designed a Recq5 cDNA (RQ5(res)), that was mutated at every third nucleotide to disrupt each of the shRNA-binding sites while retaining the correct amino acid encoded at each triplet codon. Expression of a wild-type Recq5 (RQ5(WT)) in VF-B8 cells together with a Recq5-targeting shRNA did not rescue Recq5 expression (Figure 2 H) and failed to abrogate the increased sensitivity of Recq5-depleted VF-B8 cells to HU (Figure 2 I). In contrast, expression of RQ5(res) resulted in high levels of Recq5 expression that were maintained despite expression of a Recq5 shRNA (Figure 2H) and successfully abrogated the hypersensitivity to HU of the VF-B8 cells co-expressing the Recq5 shRNA (Figure 2I). Moreover, a RQ5(res) cDNA harboring a K58R mutation within the helicase domain is incapable of abrogating HU hypersensitivity, revealing the essentiality of Recq5 helicase activity in protecting against replication stress (Figure 2J). Together, these data demonstrate that the effects of the shRNAs were on target and that Recq5 depletion was the critical factor for conferring increased sensitivity of VF-B8 cells to replication stress.

Recq5 Depletion Increases the Severity of Replication Fork Stalling in Jak2V617F-Expressing Cells Exposed to Exogenous Replication Stress

To understand the molecular mechanisms by which Recq5 depletion in VF-B8 cells leads to increased cytotoxicity by exogenous replication stressors (such as HU), we performed...
chromosome combing, which allows the direct visualization and analysis of replication tracts on individual, bromodeoxyuridine (BrdU)-labeled DNA fibers. We subjected WT-B8 or VF-B8 cells transduced with Recql5-targeting shRNAs or VA controls to this procedure. All cultures underwent a first labeling step with the BrdU analog iododeoxyuridine (IdU) under normal growth conditions, followed by a second labeling step with another BrdU analog, chlorodeoxyuridine (CldU), in culture medium supplemented with HU (Figure 3A). In this way, the extent of fork progression can be measured both in the absence and presence of HU to determine any differential effects of HU on the cultures.

We focused our initial analysis on fibers generated solely from the first labeling step to ascertain whether Recql5 depletion in the absence of HU altered DNA replication kinetics. For this analysis, we measured the length of these fibers and calculated the average fork rate because decreased fork processivity is a robust indicator of fork stalling. In control cells, we observed a significant decrease in the mean replication rate in VF-B8 cells compared with WT-B8 cells (1.47 ± 0.18 kb/min [n = 88] in VF-B8 cells versus 1.97 ± 0.21 kb/min [n = 94] in WT-B8 cells [p < 0.01]) (Figure 3B), consistent with reports published previously indicating a replication processivity impairment in JAK2V617F-expressing cells (Chen et al., 2014). However, the mean replication rate was not altered significantly by Recql5 knockdown in either WT-B8 or VF-B8 cells (WT-B8+sh1, 1.88 ± 0.44 kb/min [n = 80]; WT-B8+sh2, 1.91 ± 0.28 kb/min [n = 55]; VF-B8+sh1, 1.33 ± 0.31 kb/min [n = 101]; VF-B8+sh2, 1.32 ± 0.39 kb/min [n = 50]). Critically, no difference was observed in fork rate between VF-B8 cells depleted for Recql5 relative to those transduced with an empty vector (Figure 3B). These data indicate that physiological levels of Jak2V617F expression in myeloid cells gives rise to a replication stress phenotype, as evidenced by decreased fork processivity, but that Recql5 depletion alone has no additional affect.

We next explored the possibility that depletion of Recql5 in WT-B8 and VF-B8 cells could lead to altered replication dynamics in the presence of HU. Because replication fork progress in VF-B8 cells is impaired globally relative to WT-B8 cells, to facilitate comparison, the extent of fork progress during the first (HU-free) labeling step was normalized and arbitrarily designated at 1, and the second (HU-treated) labeling step was depicted relative to the normalized first label. Using this analysis, we observed that the presence of HU impaired fork progression in all cultures tested (Figure 3C). However, the impairment in fork progression caused by exposure to HU was significantly greater in Recql5-depleted VF-B8 cells relative to control VF-B8 cells and Recql5-depleted WT-B8 cells (Figure 3C). This indicates that Recql5 depletion differentially impairs replication fork progression in VF-B8 cells following HU exposure compared with WT-B8 cells.

The exacerbation in impairment of fork progression may be due to an increased frequency of fork collapse. To test for this, we measured the restart efficiency of a stalled replication fork, a process that is highly impaired following fork collapse. VF-B8 cells transduced with a control shRNA or shRNAs targeting Recql5 were initially exposed to HU for 60 min, followed by removal of HU and addition of BrdU (Figure 3D). Flow cytometric detection of BrdU-positive cells reflected cells that had restarted after HU exposure (Figure 3D). Also shown are quantification of BrdU-positive cells (E) and representative flow cytometric plots of BrdU staining (F).

See also Figure S2.
an increase in DSBs, as indicated by higher numbers of γH2Axi-positive foci (Figure 3G) and higher levels of γH2Axi by immunoblot analysis (Figure 3H). Together, these data demonstrate that RecQ5 is essential to maintain fork stability in mutant JAK2-expressing cells following exposure to HU.

**RECQL5 Depletion Increases the Sensitivity of JAK2V617F-Positive Cells from MPN Patients to HU**

Finally, we tested whether modulation of RECQL5 could also increase the sensitivity of JAK2V617F-positive cells from primary MPN patients to HU. Following depletion with RECQL5, CD34+ positive peripheral blood mononuclear cells from JAK2V617F-positive myelofibrosis patients were grown in semi-solid medium supplemented with HU for 14 days (Figure 4A). Strikingly, following RECQL5 depletion, there was a preferential eradication of total BFU-E colonies relative to a control shRNA (Figure 4B). Moreover, genotyping of the colonies for JAK2V617F positivity revealed a preferential elimination of JAK2V617F-positive BFU-E colonies compared with autologous wild-type colonies following exposure to HU (Figure 4C; Figure S4). Collectively, this indicates that RECQL5 knockdown preferentially sensitizes JAK2V617F-positive BFU-E colonies from MPN patients to pharmacological induction of replication stress with HU.

**DISCUSSION**

Chronic MPNs represent a model of the early stages of leukemogenesis and can provide insights into the balance between oncogene-associated DNA damage and the mechanisms that act to constrain it. JAK2V617F is the most common molecular driver of MPNs, and, in PV and ET, it is frequently the sole genetic driver identified. Although there is experimental evidence demonstrating that JAK2V617F induces DNA damage, clinical evidence indicates that chronic-phase MPNs follow a relatively indolent course over decades and that MPN genomes remain generally stable over time. In this study, we help resolve this apparent conundrum by demonstrating a role for the DNA repair helicase RECQL5 in regulating the balance between JAK2V617F-induced DNA replication stress and genomic integrity in patients with MPNs.

RECQ helicases are a family of highly conserved genome surveillance enzymes. Humans possess five RECQ helicases—RECQ1, BLM, WRN, RECQ4, and RECQ5—that have both unique and overlapping roles in the regulation of DNA replication, repair, and transcription (Larsen and Hickson, 2013). Strikingly, we observed that the increased expression of RECQL5 in JAK2V617F mutant cells was specific and not seen with other RECQ helicases. This finding precludes the possibility that increased RECQL5 levels are due to an excess of S phase cells in cultures of JAK2V617F-expressing cells or that they represent an epi-phenomenon of increased DNA damage load. Rather, using shRNA knockdown and various pharmacological agents to inhibit individual signaling pathways, we demonstrate that JAK2V617F increases RECQL5 expression through the PI3K-AKT signaling axis. In terms of the effectors downstream of PI3K-AKT signaling responsible for mediating RECQL5 activation, it is possible that a direct transcriptional mechanism, such as modulating the activity of the FOXO family of transcription factors, is involved. Indeed, a similar pathway has been shown previously to be active in JAK2 mutant cells to modulate expression of the antioxidant protein catalase (Marty et al., 2013).

The mechanisms by which RECQL5 maintains genomic integrity in normal and cancer cells are ongoing areas of investigation. Germline loss-of-function mutations in RECQ family members have been associated with a predisposition to developing cancer, and a key role has been described recently for RECQL4 in hematopoiesis (Smeets et al., 2014). However, RECQL5 dysregulation has not been implicated previously in human disease. Our data identify increased expression of RECQL5 as a means by which oncogene-induced replication stress is counteracted in JAK2-mutated MPNs. Concomitant induction of replication stress and RECQL5 expression downstream of JAK2-PI3K signaling potentially forms a feedback loop that regulates DNA damage accumulation. Depleting RECQL5 in JAK2 mutant cells disrupts this homeostasis and exposes a synthetic lethal vulnerability of JAK2 mutant cells with pharmacological inducers of
replication stress. Following depletion of RECQL5, JAK2V617F-expressing cells exposed to replication stressors (such as HU or CPT) exhibit more severe fork stalling and impaired fork restarting in comparison with isogenic wild-type cells. Moreover, RECQL5-depleted JAK2 mutant cells exhibited higher numbers of DSBs and underwent apoptosis at lower doses of HU compared with RECQL5-depleted wild-type cells (Figure 4D).

We envision two mutually non-exclusive scenarios to explain why JAK2 mutant cells are more sensitive than isogenic wild-type cells to contemporaneous RECQL5 depletion and exogenous replication stress. First, JAK2V617F-expressing cells are known to have more replication fork stalling and replication stress (Chen et al., 2014). Increased RECQL5 expression in these cells may be required to mitigate replication-associated genomic instability and maintain an appropriate balance of DNA damage/repair to ensure cell viability. RECQL5 depletion in JAK2V617F-expressing cells may therefore be more deleterious than to equivalently modified wild-type cells because JAK2 mutant cells may have less leeway to cope with the additional replication stress in the form of exogenous administration of HU. Second, JAK2 mutant cells may exhibit specific replication fork structures whose resolution is especially dependent on RECQL5 and, therefore, would be particularly sensitive to RECQL5 depletion. The mechanism of JAK2V617F-induced replication stress remains unresolved, but is likely to involve unscheduled euchromatination during S phase by JAK2 directly (Dawson et al., 2009) or following activation of downstream STATs (Shi et al., 2006), leading to physical collisions between the replication machinery and transcriptional apparatus on the same competing DNA template. The resulting DNA-RNA hybrid structures (called R loops) are potentially mutagenic and genome-destabilizing. RECQL5 may play an important role in resolving R loops in JAK2 mutant cells. Future studies to distinguish between these possibilities will be interesting.

Finally, our data may have broader pharmacological implications for cancer therapy. Although HU remains the frontline treatment for patients with ET and PV (Harrison et al., 2005), most studies have demonstrated that HU does not preferentially target the V617F-positive cell fraction in ET and PV patients (Antonioli et al., 2010), and, as a result, HU does not alter the natural history of MPNs. Our findings highlight a potential approach to amplify downstream STATs (Shi et al., 2006), leading to physical collisions between the replication machinery and transcriptional apparatus on the same competing DNA template. The resulting DNA-RNA hybrid structures (called R loops) are potentially mutagenic and genome-destabilizing. RECQL5 may play an important role in resolving R loops in JAK2 mutant cells. Future studies to distinguish between these possibilities will be interesting.

**Experimental Procedures**

**Generation of Myeloid Progenitor Cell Lines from Jak2V617F Knockin Mice**

Myeloid progenitor cell lines were generated from a C57Bl/6 Jak2V617F-expressing mouse (Mulally et al., 2010) and from a Jak2 wild-type littermate control by retroviral overexpression of an estrogen-dependent Hoxb8 transcription factor, as described previously (Wang et al., 2006). Conditionally immortalized myeloid progenitor cell lines from wild-type mice (WT-B6) and Jak2V617F knockout mice (VF-B8) were maintained in myeloid medium (RPMI medium + 10% fetal bovine serum (FBS) supplemented with 50 ng/ml murine stem cell factor (mSCF) and 1 μM β-estradiol (Sigma)).

**Cell Viability Assays**

WT-B6 and VF-B8 cells were counted, and 5 × 10⁴ cells were seeded in triplicate in each well of a 96-well plate in 100 μl of myeloid medium. For drug treatments, agents were added freshly. For serum deprivation studies, cells were rinsed with 1× PBS twice, and 1 × 10⁵ cells/well were seeded in a 96-well plate in RPMI medium + 0.5% FBS supplemented with 1 μM β-estradiol. Cell growth was measured using a standard Alamar blue assay (Life Technologies).

**Chromosome Combing**

Chromosome combing was performed as described previously (Chen et al., 2014). Briefly, 1 × 10⁵ WT-B6 or VF-B8 cells were seeded in a well of a 96-well plate in a volume of 100 μl. Replicating DNA was first labeled with 25 μM IdU (Sigma), followed by 250 μM CldU (Sigma) for 20 min each. Cells were then harvested, genomic DNA was extracted, and individual DNA molecules were stretched on glass slides. The slides were immunostained with fluorescently labeled anti-IdU (1:300) and anti-CldU (1:150) and analyzed under oil immersion on a Zeiss Axioscope 2 fluorescence microscope.

**Supplemental Information**

Supplemental Information includes Supplemental Experimental Procedures and four figures and can be found with this article online at http://dx.doi.org/10.1016/j.celrep.2015.11.037.

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