Survivin Selectively Modulates Genes Deregulated in Human Leukemia Stem Cells

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ITD-Flt3 mutations are detected in leukemia stem cells (LSCs) in acute myeloid leukemia (AML) patients. While antagonizing Survivin normalizes ITD-Flt3-induced acute leukemia, it also impairs hematopoietic stem cell (HSC) function, indicating that identification of differences in signaling pathways downstream of Survivin between LSC and HSC are crucial to develop selective Survivin-based therapeutic strategies for AML. Using a Survivin-deletion model, we identified 1,096 genes regulated by Survivin in ITD-Flt3-transformed c-kit+, Sca-1+, and lineage- (KSL) cells, of which 137 are deregulated in human LSC. Of the 137, 124 genes were regulated by Survivin exclusively in ITD-Flt3+ KSL cells but not in normal CD34neg KSL cells. Survivin-regulated genes in LSC connect through a network associated with the epidermal growth factor receptor signaling pathway and falls into various functional categories independent of effects on apoptosis. Pathways downstream of Survivin in LSC that are distinct from HSC can be potentially targeted for selective anti-LSC therapy.

1. Introduction

Survivin has been implicated in regulation of apoptosis, cell division, and cell cycle both in cancer cells and normal tissues, through caspase-dependent and -independent mechanisms [1–3]. We previously showed that Survivin is expressed and growth factor regulated in human CD34+ cells [4–6]. Antagonizing Survivin impairs production of mouse bone marrow hematopoietic progenitor cells in vitro [4, 7] and conditional Survivin gene deletion in vivo in mice leads to bone marrow ablation as a result of loss of hematopoietic stem and progenitor cells (HSPC) [8]. In contrast to tight regulation of Survivin by hematopoietic growth factors in normal CD34+ cells, deregulated expression of Survivin is frequently observed in hematological diseases, particularly those associated with hematopoietic stem cell (HSC) expansion. For instance, Survivin is aberrantly overexpressed in acute myeloid leukemia [9, 10] but downregulated in marrow cells of patients with aplastic anemia where HSPC are significantly reduced [11]. These findings suggest that Survivin regulates HSPC fate under normal and pathological conditions.

We previously reported that ITD-Flt3 mutations found in ~25–30% of patients with acute myeloid leukemia (AML) and strongly associated with poor prognosis [12–14], increase expression of Survivin. Survivin mediates aberrant hematopoietic cell proliferation induced by ITD-Flt3 and regulates development of ITD-Flt3+ acute leukemia, suggesting that antagonizing Survivin may provide therapeutic benefit for patients with AML expressing ITD-Flt3 [14]. Survivin is the fourth most highly expressed transcript in cancer [15] and is commonly associated with a higher proliferative index, reduced apoptosis, resistance to chemotherapy and increased rate of tumor recurrence in cancer cells, making anti-Survivin therapy an attractive strategy in cancer [3]. Several anti-Survivin preclinical trials in solid tumor models...
show that disrupting Survivin can reduce tumor growth [1–3]. However, studies from our group and others indicate that Survivin regulates normal HSPC [4, 7, 8], suggesting that targeting Survivin will likely result in hematopoietic toxicity. Therefore, identification of differences in signaling cascades downstream of Survivin between normal HSPC and cancer stem cells (CSC) or leukemia stem cells (LSC) are required to pinpoint targets that can effectively eradicate CSC/LSC with little toxicity on HSC. Previous reports show that ITD-Flt3 mutations are present in human LSC [13] and genes expressed in AML stem cells are deregulated [16]. The primary purpose of this study was to identify downstream Survivin signaling pathways in LSC that are distinct from normal HSC. Using ITD-Flt3 transformed c-kit+, Sca-1+, and lineage$^{-}$ (KSL) cells from conditional Survivin knockout mice as surrogates for AML stem cells and littermate controls, we identified a panel of genes that are specifically regulated by Survivin in ITD-Flt3 transformed KSL cells and known to be deregulated in human LSC. The data identify selective signaling pathways downstream of Survivin in LSC that are distinct from normal HSC that can be potentially targeted for selective anti-LSC therapy.

2. Materials and Methods

2.1. Antibodies and Cytokines. Anti-Fcy-III/II receptor antibody, APC conjugated antimouse c-kit (clone 2B8), biotin-conjugated anti-Sca-1 (E13-161.7), R-PHYcoerythrin (PE) conjugated anti-mouse CD3 (clone 143–2C11), GR-1 (clone RB6-8C5), B220 (clone RA3-6B2), Mac1 (clone M1/70), Ter119 (clone Ter1 19), rat IgG2a, rat IgG2b, hamster IgG, and Streptavidin-PE-Cy7 were purchased from BD Biosciences (San Diego, CA). Recombinant human Flt3 ligand (FL) and Thrombopoietin (Tpo) were provided by Amgen, Thousand Oaks, CA. Recombinant murine stem cell factor (rmSCF) was purchased from R&D Systems (Minneapolis, MN). Tamoxifen and 4-hydroxy (4OH) tamoxifen were from Sigma-Aldrich (St. Louis, MO).

2.2. Cell Culture, Plasmid Transfection, Retrovirus Transduction, and In Vitro Survivin Gene Deletion. Mice with the Survivin gene flanked by loxP sites and a Tamoxifen-inducible form of Cre (Cre-ER) were reported previously [14]. All mice were housed in microisolators with continuous access to rodent chow and acidified water. The Indiana University School of Medicine IACUC approved all experimental procedures. Retrovirus transduction of ITD-Flt3 into mouse bone marrow cells was carried out as described [14]. Briefly, bone marrow cells from littermate control Survivinflox/flox or CreER-Survivinflox/flox mice were transduced with ITD-Flt3 (N51) in MSCV-ires-egfp vector [17]. Seventy-two hours after sequential transduction, the medium was replaced with IMDM containing 10% FBS and the cells were incubated for 14 days without growth factors in the presence of 1uM of 4OH-Tamoxifen to induce Survivin gene deletion. All cells were cultured in triplicates in two independent experiments. Viable cells were enumerated using trypan blue exclusion and GFP$, c-kit+, Sca-1+, and lineage negative (KSL) cells were sorted on day 14 by FACSaria (BD Biosciences). Sorted cells were immediately lysed and subjected to differential mRNA microarray analysis.

2.3. In Vivo Survivin Deletion. Survivinflox/flox and CreER-Survivinflox/flox mice were treated with 5 mg Tamoxifen (5 mg/mouse i.p.) for 3 consecutive days and allowed to rest for 3 days, and Tamoxifen was administered for 3 additional days. Fourteen days after the final Tamoxifen injection, marrow cells were harvested, stained with anti-CD34, c-kit, Sca-1, and lineage markers and CD34$^{+}$ KSL cells isolated by FACSaria for mRNA microarray analysis.

2.4. mRNA Microarray. Sorted cells were immediately lysed and subjected to differential mRNA microarray analysis. Microarray analysis was performed on a fee basis by Miltenyi Biotec (Auburn, CA). Briefly, 250 nanograms of each cDNA was used as template for Cy3 and Cy5 labeling which was performed according to Miltenyi Biotec’s proprietary protocol. Equal amounts of labeled cDNAs from Tamoxifen-treated Survivinflox/flox and CreER-Survivinflox/flox mice or those cells expressing ITD-Flt3 were hybridized overnight (17 hours, 65°C) to Agilent Whole Mouse Genome Oligo Microarrays (44 K) according to the manufacturers’ protocol. Fluorescence signals of the hybridized microarrays were detected using Agilent’s DNA microarray scanner (Agilent, Palo Alto, USA). The Agilent Feature Extraction Software (FES) was used to read out and process the microarray image files. The software determines feature intensities and ratios (including background subtraction and normalization), rejects outliers, and calculates statistical confidences (P-values). For determination of differential gene expression, FES derived output data files were further analyzed using the Rosetta Resolver gene expression data analysis system (Rosetta BioSoftware, Seattle, USA). Significantly regulated genes were annotated and assigned to functional categories using the DAVID 2008 (the Database for Annotation, Visualization and Integrated Discovery; http://david.abcc.ncifcrf.gov/home.jsp) program [18]. Functional networks of the genes regulated by Survivin was visualized using Cytoscape software: http://www.cytoscape.org/ [19].

3. Results

3.1. Reduction in Proliferation ITD-Flt3 Transformed KSL Cells by Survivin Deletion Is Associated with Alteration of Expression of 1,096 Genes Classified into Various Biological Functions. While incubation of marrow cells transduced with wild-type Flt3 in IMDM containing 10% FBS without hematopoietic any growth factors failed to support KSL proliferation, overexpression of ITD-Flt3-EGFP in primary mouse bone marrow cells results in the ability of KSL cells to proliferate in the same culture condition without additional any hematopoietic growth factors as we reported, indicating that ITD-Flt3 is sufficient to transform primitive HSPC [14]. Incubation of ITD-Flt3 transduced marrow cells derived from CreER-Survivinflox/flox mice with 1uM 4OH-Tamoxifen significantly reduced factor-independent KSL cells on day 14 compared to Survivinflox/flox cells treated in exactly the same
manner (Figure 1), confirming that Survivin is required for factor independent growth of ITD-Flt3 transformed KSL cells.

To investigate the mechanism by which Survivin regulates aberrant proliferation of ITD-Flt3 transformed KSL cells, mRNA expression was compared between ITD-Flt3 transformed Survivinflox/fox and CreER-Survivinflox/fox KSL using mRNA microarrays. Although inhibition of Survivin mRNA expression in CreER-Survivinflox/fox KSL cells by 4OH-Tamoxifen was only 50% compared to control, the mRNA microarray identified 1,096 transcripts differentially regulated by Survivin. These genes were classified based on biological process and molecular function defined by Gene Ontology Term (http://www.geneontology.org/) using DAVID 2008 program (Figure 2). Representative groups for biological process include phosphate metabolic process, cell cycle, cell division, response to DNA damage stimulus, RNA biosynthetic process and transcription (Figure 2(a), \( P < .02 \)). Ion-binding, nucleotide-binding, DNA-binding, and protein kinase activity were the top 4 significantly enriched categories in molecular function defined by Gene Ontology database (Figure 2(b), \( P < .02 \)). The list of genes classified by biological process and molecular function are listed in supplementary Table S1 available at doi:10.1155/2011/946936.

3.2. Survivin Modulates Expression of Genes Deregulated in Human AML LSC. The requirement of Survivin for the self-renewing capability of ITD-Flt3 transformed KSL cells or CFU [14] and the presence of ITD-Flt3 in LSC [13] strongly suggests that Survivin is important for LSC fate decisions resulting from deregulated gene expression. We compared 1,096 differentially expressed genes in Survivin deleted ITD-Flt3+KSL cells with the existing deregulated gene expression database for human AML stem cells [16]. Out of 1,096 genes regulated by Survivin in KSL cells transformed by ITD-Flt3, 137 genes are also listed in the deregulated molecules in human AML stem cells (Tables 1(a)–1(d)) [16]. The relationship between Survivin and LSC on the modulation

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**Figure 1:** The effect of Survivin deletion on the number of KSL cells transformed by ITD-Flt3 (a) The KSL cells transformed by ITD-Flt3 were FACS sorted using GFP as a marker. Marrow cells derived from littermate control Survivinflox/fox or CreER-Survivinflox/fox mice were transduced with ITD-Flt3 (N51) in MSCV-IRES-EGFP vector. Seventy-two hours after transduction, the medium was replaced with IMDM containing 10%FBS and the cells were incubated for 14 days in the presence of 1uM of 4OH-Tamoxifen without hematopoietic growth factors. GFP⁺, c-kit⁺, Sca-1⁺, and lineage-negative (KSL) cells shown in red box were sorted on day 14 by FACSAria (BD Biosciences). (b) The number of whole cells and KSL cells derived from Survivinflox/fox or CreER-Survivinflox/fox marrow cells transduced with ITD-Flt3 cultured with 4OH-tamoxifen in the absence of any growth factors was quantitiated in triplicate wells of each sample averaged from 2 independent experiments. *\( P < .02 \) compared to control Survivinflox/fox.
of the gene expression is illustrated in Figure 3(a). Of the 137 genes identified, 79 genes were downregulated by Survivin deletion while 58 genes were upregulated, implying that these genes are conversely increased and decreased by the presence of Survivin in LSC (Figure 3(a)). In contrast, 92 genes were upregulated and 45 genes were downregulated by LSC. Among the 79 genes downregulated genes by Survivin deletion, 55 transcripts were upregulated (Figure 3(a) and Table 1(a)) whereas 24 genes were downregulated by LSC (Figure 3(a) and Table 1(c)). Similarly, out of 58 upregulated transcripts by Survivin deletion, 21 genes were downregulated (Figure 3(a) and Table 1(b)) and 37 genes were upregulated in LSC (Figure 3(a) and Table 1(d)) (Figure 3(a)).

Next, we classified the 137 genes into functional annotation groups using the DAVID program ($P < .05$, Figure 3(b)). Significantly enriched functional groups were annotated as phosphoprotein, nucleus, acetylation, cell cycle, ATP binding, regulation of EGF signaling pathway, cell adhesion and others. The size of each circle in Figure 3(b) represents the number of genes involved in each functional category and the thickness of the line indicates the number of genes shared with any function, suggesting enrichment of the gene groups with functional redundancy. Among the molecules most frequently shared within the functional groups include BGLAP, Chrac1, Hmgb1 and Smarce1. Similarly, 76 genes upregulated or downregulated both by Survivin and LSC (55 + 21 genes shown in Figure 1(a)) were functionally classified (Figure 3(c)). Phosphoprotein, acetylation, DNA binding, ATP binding and DNA replication were significantly enriched.

The 1,096 differentially Survivin-regulated genes were also searched against the list of genes expressed in mouse HSC [20]. We identified 94 differentially expressed genes in the list of genes that include HSC. There were 45 genes whose expression was selectively enriched in HSC compared to other populations (Table 2). Crem, Emp1, HmgA2, Lrrn1, Maff, Myef2, Rps4x, and Sos1 listed in HSC database are also deregulated in LSC. We next compared the human LSC associated 137 genes regulated by Survivin in ITD-Flt3+KSL cells with differentially regulated genes in normal bone marrow CD34neg KSL cells obtained from CreER-Survivinflox/flox and control Survivinflox/flox mice following Survivin deletion. Tamoxifen reduced Survivin expression by 10-fold in CD34neg KSL cells from CreER-Survivinflox/flox mice compared to control Survivinflox/flox in two independent experiments. Out of 137 genes, Arg2, Med25, Pmaip1, Pola2, Ube3b, Ephb2 and Rab18 were differentially regulated in both ITD-Flt3+KSL cells and normal CD34neg KSL cells obtained from CreER-Survivinflox/flox mice compared to control Survivinflox/flox in two independent experiments. Out of 137 genes, Arg2, Med25, Pmaip1, Pola2, Ube3b, Ephb2 and Rab18 were differentially regulated in both ITD-Flt3+KSL cells and normal CD34neg KSL cells obtained from CreER-Survivinflox/flox mice compared to control Survivinflox/flox in two independent experiments. Out of 137 genes, Arg2, Med25, Pmaip1, Pola2, Ube3b, Ephb2 and Rab18 were differentially regulated in both ITD-Flt3+KSL cells and normal CD34neg KSL cells obtained from CreER-Survivinflox/flox mice compared to control Survivinflox/flox in two independent experiments. 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Table 1
(a) Genes upregulated in LSC but downregulated in ITD-Flt3⁺ KSL cells deleted with Survivin (55 genes).

| Symbol | Name | Fold change |
|--------|------|-------------|
| Acaa2 | acetyl-Coenzyme A acyltransferase 2 (mitochondrial 3-oxoacyl-Coenzyme A thiolase) (Acaa2), [NM_177470] | −3.7 |
| Ankrd12 | Mus musculus 15 days embryo head cDNA, RIKEN full-length enriched library, clone:D930020E11 product: unknown EST, full insert sequence. [AK086311] | −5.5 |
| Ankrd28 | Mus musculus 2 days neonate thymus thymic cells cDNA, RIKEN full-length enriched library, clone:E430019N21 product: unknown EST, full insert sequence. [AK088541] | −2.9 |
| Ap1g2 | adaptor protein complex AP-1, gamma 2 subunit (Ap1g2), [NM_007455] | −5.2 |
| Arhgef18 | rho/rac guanine nucleotide exchange factor (GEF) 18 (Arhgef18), [NM_133962] | −3.6 |
| Arrdc1 | arrestin domain containing 1 (Arrdc1), [NM_178408] | −2.1 |
| Atp11a | ATPase, class VI, type 11A, [AK006628] | −2.6 |
| BGLAP | bone gamma carboxyglutamate protein 1 (Bglap1), [NM_007541] | −10.2 |
| Cars | cysteiny1-tRNA synthetase (Cars), [NM_013742] | −2.3 |
| Ccr1 | chemokine (C-C motif) receptor 1 (Ccr1), [NM_009912] | −3.5 |
| Cd96 | CD96 antigen (Cd96), [NM_032465] | −2.1 |
| Cfp | complement factor properdin (Cfp), [NM_008823] | −4.3 |
| Cyyr1 | cysteine and tyrosine-rich protein 1 (Cyyr1), [NM_144853] | −2.7 |
| Dpagt1 | dolichyl-phosphate (UDP-N-acetylglucosamine) acetylglucosaminophosphotransferase 1 (GlcNAc-1-P transferase) (Dpagt1), [NM_007875] | −2.8 |
| Dpp9 | dipetidylpeptidase 9 (Dpp9), [NM_172624] | −2.4 |
| Elov11 | elongation of very long chain fatty acids (FEN1/Elo2, SUR4/Elo3, yeast)-like 1 (Elov11), [NM_019422] | −2.5 |
| Ets2 | E26 avian leukemia oncogene 2, 3' domain (Ets2), [NM_011809] | −2.5 |
| Fgr | Mus musculus Gardner-Rasheed feline sarcoma viral (Fgr) oncogene homolog (Fgr), mRNA [NM_010208] | −2.9 |
| Gfi1 | growth factor independent 1 (Gfi1), [NM_010278] | −2.7 |
| Gga2 | golgi associated, gamma adaptin ear containing, ARF binding protein 2 (Gga2), [NM_028758] | −2.5 |
| Glt2 | G protein-coupled receptor kinase-interactor 2, [NM_178408] | −3.4 |
| Hdac10 | histone deacetylase 10 (Hdac10), [NM_199198] | −4.1 |
| Ipo13 | importin 13 (Ipo13), [NM_146152] | −2.3 |
| Isyna1 | myo-inositol 1-phosphate synthase A1 (Isyna1), [NM_023627] | −2.5 |
| Klhl17 | kelch-like 17 (Drosophila) (Klhl17), [NM_198305] | −4.8 |
| Lars2 | leucyl-tRNA synthetase, mitochondrial (Lars2), nuclear gene encoding mitochondrial protein, [NM_153168] | −3.6 |
| Lrrk1 | leucine-rich repeat kinase 1 (Lrrk1), [NM_146191] | −2.7 |
| Lztr1 | leucine-zipper-like transcriptional regulator 1 (Lztr1), [NM_025808] | −3.4 |
| Mapk8ip3 | mitogen-activated protein kinase 8 interacting protein 3 (Mapk8ip3), [NM_013931] | −2.9 |
| Mcm8 | minichromosome maintenance deficient 8 (S. cerevisiae) (Mcm8), [NM_025676] | −2.4 |
| Med25 | TCBAP0758 protein (Fragment) homolog [Homo sapiens], [AK165760] | −5.4 |
(a) Continued.

| Symbol | Name | Fold change |
|--------|------|-------------|
| Mr1    | laminin receptor 1 (ribosomal protein SA) (Lamr1), [NM_011029] | −2.6 |
| Napa   | N-ethylmaleimide sensitive fusion protein attachment protein alpha (Napa), [NM_025898] | −3.0 |
| Ndrg1  | N-myc downstream regulated gene 1 (Ndrg1), [NM_010884] | −3.3 |
| Nsfl1c | NSFL1 (p97) cofactor (p47) (Nsfl1c), [NM_198326] | −2.2 |
| Otud5  | bZ30I22.1 (Novel protein similar to human FLJ12550) (Fragment) homolog [Brachydanio rerio], [AK169921] | −3.1 |
| Pank4  | pantothen kinase 4 (Pank4), [NM_172990] | −3.0 |
| Pgl    | 6-phosphogluconolactonase, [AK158308] | −5.6 |
| Phkg2  | phosphorylase kinase, gamma 2 (testis) (Phkg2), [NM_026888] | −3.3 |
| Pmaip1 | phorbol-12-myristate-13-acetate-induced protein 1 (Pmaip1), [NM_021451] | −6.6 |
| Pola2  | polymerase (DNA directed), alpha 2 (Pola2), [NM_008893] | −2.7 |
| Polg   | polymerase (DNA directed), gamma 2, accessory subunit (Polg2), [NM_015810] | −3.2 |
| Rnf126 | ring finger protein 126 (Rnf126), [NM_144528] | −2.1 |
| Ropn1l | ropporin 1-like (Ropn1l), [NM_145852] | −6.8 |
| Rtel1  | regulator of telomere elongation helicase 1 (Rtel1), [NM_001001882] | −4.3 |
| Slc1a5 | solute carrier family 1 (neutral amino acid transporter), member 5 (Slc1a5), [NM_009201] | −2.3 |
| Slc25a28 | solute carrier family 25, member 28 (Slc25a28), [NM_145156] | −2.9 |
| Snapc4 | small nuclear RNA activating complex, polypeptide 4 (Snapc4), [NM_172339] | −2.7 |
| Solh   | small optic lobes homolog (Drosophila) (Solh), [NM_015830] | −3.9 |
| Synj1  | PREDICTED: synaptojanin 1 (Synj1), [XM_358889] | −2.8 |
| Trim21 | tripartite motif protein 21 (Trim21), [NM_009277] | −11.4 |
| Trpm2  | transient receptor potential cation channel, subfamily M, member 2 (Trpm2), [NM_138301] | −4.2 |
| Trpm7  | transient receptor potential cation channel, subfamily M, member 7 (Trpm7), [NM_021450] | −3.2 |
| Ube3b  | ubiquitin protein ligase E3B (Ube3b), [NM_054093] | −2.8 |
| Upp1   | uridine phosphorylase 1 (Upp1), [NM_009477] | −3.4 |

Fold change: ITD-Flt3+ KLS cells derived from CreER-Survivin<sup>SV</sup> compared to control Survivin<sup>SV</sup> in two experiments.

(b) Genes downregulated in LSC but upregulated in ITD-Flt3+ KSL cells deleted with Survivin (21 genes)

| Symbol | Name | Fold change |
|--------|------|-------------|
| Aebp2 | AE binding protein 2 (Aebp2), transcript variant 3, [NM_009637] | 11.4 |
| Anp32b | unspliced dna for: PROLIFERATION RELATED ACIDIC LEUCINE RICH PROTEIN PAL31 (SIMILAR TO ACIDIC PROTEIN RICH IN LEUCINES) homolog [Mus musculus], … | 2.9 |
| App    | amyloid beta protein precursor, [M18373] | 25.7 |
| Cd164  | CD164 antigen, [AK018009] | 2.5 |
| Crem   | cAMP responsive element modulator, [AK016156] | 7.3 |
(b) Continued.

| Symbol | Name | Fold change |
|--------|------|-------------|
| Emp1   | epithelial membrane protein 1 (Emp1), [NM_010128] | 2.3 |
| Fbx17  | Mus musculus 10 days neonate cerebellum cDNA, RIKEN full-length enriched library, clone:B930094M09 product: unknown EST. [AK081162] | 5.1 |
| Gas2l3 | Mus musculus 16 days embryo head cDNA, RIKEN full-length enriched library, clone:C130036O09 product: unclassifiable, [AK048137] | 12.5 |
| Myef2  | myelin basic protein expression factor 2, repressor (Myef2), [NM_010852] | 10.6 |
| Nrip1  | nuclear receptor interacting protein 1 (Nrip1), [NM_173440] | 6.2 |
| Pla2g12a | phospholipase A2, group XIA (Pla2g12a), [NM_023196] | 3.2 |
| Psmd11 | Mus musculus adult male testis cDNA, RIKEN full-length enriched library, clone: 1700089D09 product: unclassifiable. [AK007029] | 12.3 |
| Rps20  | Mus musculus bone marrow macrophage cDNA, RIKEN full-length enriched library, clone:J830013G23 product: hypothetical protein. [AK150667] | 5.5 |
| Senp7  | SUMO1/sentrin specific protease 7 (Senp7), transcript variant 3, [NM_001003972] | 12.3 |
| Sln2a16 | solute carrier family 25 (mitochondrial carrier, Graves disease autoantigen), member 16 (Slc25a16), [NM_175194] | 2.8 |
| Spire1 | spire homolog 1 (Drosophila) (Spire1), transcript variant 2, [NM_176832] | 9.5 |
| Tdrd3  | Mus musculus adult male olfactory brain cDNA, RIKEN full-length enriched library, clone:6430599B16 product: hypothetical Tudor domain containing protein. [AK078326] | 2.9 |
| Tmpo   | thymopoietin (Tmpo), [NM_011605] | 4.3 |
| Trps1  | Mus musculus adult male adrenal gland cDNA, RIKEN full-length enriched library, clone: 7330401C17 product: expressed sequence AI115454, [AK078617] | 11.4 |
| Ube2n  | ubiquitin-conjugating enzyme E2N (Ube2n), [NM_080560] | 2.4 |
| Wtap   | Mus musculus 12 days embryo embryonic body between diaphragm region and neck cDNA, RIKEN full-length enriched library, clone: 9430038B09 product: unclassifiable. [AK020459] | 14.2 |

Fold change: ITD-Flt3⁺ KLS cells derived from CreER-Survivinlox/lox compared to control Survivinlox/lox in two experiments.

(c) Genes down regulated in LSC and in ITD-Flt3⁺ KSL cells deleted with Survivin (24 genes)

| Symbol | Name | Fold change |
|--------|------|-------------|
| Arg2   | arginase type II (Arg2), [NM_009705] | −2.2 |
| Atp2c1 | CALCIUM-TRANSPORTING ATPASE TYPE 2C, MEMBER 1 (EC 3.6.3.8) (ATPASE 2C1) (ATP-DEPENDENT CA2+ PUMP PMR1) homolog | −2.2 |
| Chrac1 | chromatin accessibility complex 1., [AK031914] | −4.3 |
| Dhx30  | DEAH (Asp-Glu-Ala-His) box polypeptide 30 (Dhx30), [NM_133347] | −3.1 |
| Dsg2   | desmoglein 2, (cDNA clone IMAGE:4036406), [BC034056] | −4.1 |
| Eef1g  | eukaryotic translation elongation factor 1 gamma (Eef1g), [NM_026007] | −2.2 |
| Herc5  | hypothetical regulator of chromosome condensation (RCC1) containing protei· · · | −2.3 |
| Hmga2  | high mobility group AT-hook 2 (Hmga2), [NM_178057] | −5.0 |
| Hmgb1  | high mobility group box 1, [BC064790] | −3.4 |
| Hmmr   | hyaluronan mediated motility receptor (RHAMM) (Hmmr), [NM_013552] | −3.2 |
| Il15   | interleukin 15 (Il15), [NM_008357] | −6.1 |
| Jmy    | junction-mediating and regulatory protein (Jmy), [NM_021310] | −4.1 |
| Lrrn1  | Mus musculus 10, 11 days embryo whole body cDNA, RIKEN full-length enriched library, clone:2810047E21 product: unclassifiable, full insert sequence. [AK012914] | −4.0 |
| Symbol | Name | Fold change |
|--------|------|-------------|
| Maff   | v-maf musculoaponeurotic fibrosarcoma oncogene family, protein F (avian) (Maff), [NM_010755] | −8.4 |
| Mapk14 | mitogen activated protein kinase 14 (Mapk14), [NM_011951] | −2.4 |
| Ncoa7  | nuclear receptor coactivator 7, [BC076623] | −8.9 |
| Nedd1  | neural precursor cell expressed, developmentally downregulated gene 1 (Nedd1), [NM_008682] | −2.6 |
| Parp11 | poly (ADP-ribose) polymerase family, member 11 (Parp11), [NM_181402] | −2.5 |
| Pawr   | PRKC, apoptosis, WT1, regulator (Pawr), [NM_054056] | −2.8 |
| Rps4x  | ribosomal protein S4, X-linked (Rps4x), [NM_009094] | −2.2 |
| Smarce1| SWI [AK042961] | −2.9 |
| Tpbg   | trophoblast glycoprotein, [AK050794] | −4.5 |
| Vwf    | Von Willebrand factor homolog (Vwf), [NM_011708] | −5.8 |
| Zbtb20 | zinc finger and BTB domain containing 20 (Zbtb20), [NM_019778] | −3.1 |

Fold change: ITD-Flt3+ KLS cells derived from CreER-Survivin+/−x compared to control Survivin+/−x in two experiments.

(c) Continued.

| symbol | Name | fold change |
|--------|------|-------------|
| Cenpa  | Mus musculus 0 day neonate eyeball cDNA, RIKEN full-length enriched library, clone:E130306P06 product: hypothetical protein, [AK165029] | 10.7 |
| Clcn3  | chloride channel 3 (Clcn3), transcript variant c, [NM_173876] | 5.5 |
| Cpd    | carboxypeptidase D (Cpd), [NM_007754] | 6.2 |
| Cul4b  | cullin 4B, [AK164640] | 4.9 |
| Ddx52  | DEAD (Asp-Glu-Ala-Asp) box polypeptide 52 (Ddx52), [NM_030096] | 3.3 |
| Dnajc1 | DnaJ (Hsp40) homolog, subfamily C, member 11 (Dnajc11), [NM_172704] | 6.4 |
| Ephb2  | Eph receptor B2 (Ephb2), [NM_010142] | 3.0 |
| Gnaq   | guanine nucleotide binding protein, alpha q polypeptide (Gnaq), [NM_008139] | 17.2 |
| Gripap1| premature mRNA for mKIAA1167 protein [AK173119] | 2.2 |
| Il1rap | interleukin 1 receptor accessory protein (Il1rap), transcript variant 2, [NM_134103] | 11.0 |
| Itsn2  | SH3 domain protein 1B, [AK161743] | 4.4 |
| Lass6  | longevity assurance homolog 6 (S. cerevisiae) (Lass6), [NM_172856] | 2.3 |
| Lpp    | caseinolytic protease, ATP-dependent, proteolytic subunit homolog (E. coli) (Clpp), [NM_017393] | 2.1 |
| Mpp1   | metallophosphoesterase 1 (Mpp1), [NM_172630] | 3.4 |
| Mrps5  | mitochondrial ribosomal protein S5, [AK047438] | 9.0 |
| Myh9   | myosin heavy chain IX (Myh9), [NM_181327] | 2.8 |
| Nck2   | non-catalytic region of tyrosine kinase adaptor protein 2 (Nck2), [NM_010879] | 4.8 |
| Nin    | microtubule associated serine/threonine kinase-like (Mastl), [NM_025979] | 7.6 |
| Nmt2   | N-myristoyltransferase 2, [AK049483] | 2.3 |
| Orc5l  | origin recognition complex, subunit 5 homolog (S. cerevisiae), [AK054452] | 4.6 |

(d) Genes upregulated in LSC and in ITD-Flt3+KSL cells deleted with Survivin (37 genes).
3.3. Survivin Modulates Gene Expression in LSC That Connects Through a Functional Signaling Network Associated with Epidermal Growth Factor Receptor Signaling Pathway. Functional annotation analysis indicates that genes related with dorsoventral axis formation or epidermal growth factor receptor signaling pathway (EGFR) are significantly enriched in the shared genes associated with LSC and Survivin signaling in KEGG database (http://www.genome.jp/kegg/) (P < .03). Similarly, genes associated with regulation of EGFR signaling pathway are enriched in the Gene Ontology database (P < .02). Our analysis shows that 13 molecules shared by LSC and Survivin signaling are mapped on a functional signaling network that connects through the EGFR pathway (Figure 4).

4. Discussion

Survivin deletion in ITD-Flt3 transformed KSL cells results in a significant reduction in growth factor-independent proliferation coincident with growth inhibition. Our data shows that Survivin deletion modulates gene expression of 1,096 genes that are associated with various cellular and metabolic functions. Comparison of the 1,096 Survivin regulated genes in ITD-Flt3+KSL cells with 3,005 differentially regulated genes in the human AML stem cell database identified 137 shared genes. Functional classification of these 137 genes indicates they connect through a functional signaling network associated with EGFR signaling pathway and affect various biological and molecular processes. Comparison of the 137 Survivin-regulated genes in LSC with transcripts regulated by Survivin in normal CD34neg KSL cells show that 124 genes are regulated by Survivin exclusively in ITD-Flt3+KSL cells and not in normal HSC. These data suggest that Survivin contributes to deregulation of gene expression in AML stem cells via selective signaling pathways distinct from normal HSC that can be potentially targeted for therapeutic benefit.

There is no direct evidence that Survivin directly regulates gene transcription; however, modulation of Survivin can clearly affect transcription in cancer cells [21–23] and transgenic expression of Survivin alters the expression of multiple genes in the bladder [24].

The involvement of Survivin in transcriptional regulation is consistent with the fact that Bir1, a Caenorhabditis
Up-regulated by survivin
(down-regulated by survivin deletion)

Down-regulated in LSC
Up-regulated in LSC

Down-regulated by survivin
(up-regulated by survivin deletion)

(a)

Figure 3: Continued.
**Figure 3**: Classification of genes regulated by Survivin and LSC. (a) The 137 genes were classified based on the regulation by human LSC and Survivin in ITD-Flt3+KSL cells. Arrow bars in the quadrant indicate the direction of gene regulation by Survivin or LSC. The top-right corner shows 55 genes upregulated by both LSC and Survivin (downregulated by Survivin gene deletion), whereas bottom-left corner represents 21 genes downregulated by both LSC and Survivin (upregulated by Survivin gene deletion). The top left corner shows 24 genes downregulated by LSC but upregulated by Survivin. The bottom-right corner indicates 37 genes upregulated by LSC but downregulated by Survivin. (b) Functional group association networks for Survivin regulated genes in the list of deregulated genes in human LSC visualized using Cytoscape program. The 137 genes listed in Tables 1(a)–1(d) were functionally annotated using DAVID software (P < 0.05). The size of each circle represents the number of genes involved in each functional categories and the thickness of the line indicates the number of genes shared with any function. Functional classification was performed based on Gene Ontology Term database (biological process, molecular function and cellular component) and Swissprot Keywords. (c) The 76 genes upregulated or downregulated by Survivin and LSC (55 + 21 genes shown in Figure 3(a)) were functionally classified by DAVID software and visualized by Cytoscapse program as shown in Figure 3(b). The size of each circle represents the number of genes involved in each functional categories and the thickness of the line indicates the number of genes shared with any function. Functional classification was performed based on Swissprot Keywords. Genes annotated in each functional category are shown in the box.

**elegans** homologue of Survivin, regulates transcription, most likely through Histone phosphorylation by Aurora kinase [25]. Survivin is essential for activation of Aurora kinase that phosphorylates Histone H3 [26, 27], an event required for transcriptional regulation [28–30] and cytokinesis [31], suggesting that Survivin may regulate transcription via Aurora kinase mediated Histone H3 phosphorylation. Over expression of Survivin also leads to phosphorylation of the Sp1 transcription factor [21]. Since microarray analysis was performed on lineage marker negative viable cells that express GFP, c-kit, and Sca-1 and survived in culture, it is unlikely that the effects of Survivin on gene expression is a consequence of KSL apoptosis, although we cannot rule out indirect effects resulting from cell cycle arrest as a consequence of Survivin deletion.

Survivin is known to inhibit caspases 3, 7, and 10 that mediate apoptosis [1–3]. Our functional annotation analysis of the genes regulated by Survivin indicates that Survivin modulates a number of genes that affect multiple biological processes and molecular functions, although inhibition of Survivin mRNA expression in CreER-Survivinfloxflox KSL cells by 4OH-Tamoxifen was only 50% compared to control. While incompleteness of the gene deletion may affect the downstream signals, significant reduction of total cell number was observed. Since Survivin deletion induces cell death, the deleted cells are likely no longer present in the culture at the time of cell harvest for analyses, resulting in an underestimate of the efficiency of Survivin gene deletion and likely represents Survivin expression in cells that have escaped the Tamoxifen-induced deletion. The genes affected by Survivin deletion were classified into cell cycle, adhesion, DNA replication, transcriptional factor binding, acetylation, phosphorylation, and polymerase. We did not observe changes in genes that mediate apoptosis. These results suggest that Survivin can regulate ITD-Flt3 transformed KSL cell fate independent of its activity as a caspase inhibitor. More importantly, comparison of the 1,096 Survivin regulated genes in ITD-Flt3+KLS cells with the
Figure 4: Genes regulated by Survivin in ITD-Flt3+KSL cells and deregulated in LSC are mapped on functional signaling network associated with epidermal growth factor receptor signaling pathway. Epidermal growth factor receptor signaling pathway (EGFR) is significantly enriched in the shared genes associated with LSC and Survivin signaling by pathway enrichment analysis in KEGG and Gene Ontology database. Our analysis shows that 13 molecules shared by LSC and Survivin signaling are mapped on a functional signaling network associated with EGFR. The network was created based on KEGG database using Cytoscape software. Green circle and red circle represents downregulation and upregulation by Survivin gene deletion, respectively.

human AML stem cell database demonstrates that 137 genes (12.4%) modulated by Survivin are also deregulated in LSCs. Functional annotation analysis indicates that genes related to dorsoventral axis formation or epidermal growth factor receptor signaling pathway (EGFR) are significantly enriched in the shared genes associated with LSC and Survivin signaling, even though the number of molecules detected that associates with EGFR signaling is small. EGFR signaling activates signaling cascades involved in cell proliferation, such as Src, Sos, and MAP-kinases, and is known to be dysregulated in solid tumors [32, 33]. Our data also indicates that Survivin modulates expression of several genes that connect through the EGFR signaling pathway (Figure 4), suggesting a potential role of EGFR signaling downstream of Survivin in AML stem cells, despite the fact that EGFR is not usually upregulated in AML cells [34]. However, recent studies indicate an antineoplastic effect of an EGFR inhibitor in AML via off-target effects [34, 35]. Thus, our data would support EGFR signaling as a candidate pathway for treatment of patients with AML.

While Survivin is not listed in the LSC database [16], it is expressed at higher levels in AML cells compared to normal CD34+ cells [9, 10]. Our data clearly show that Survivin affects expression of 137 genes associated with LSC and reduces aberrant KSL proliferation induced by ITD-Flt3. This finding suggests that antagonizing Survivin in LSC may normalize expression of the deregulated genes, which in turn can inhibit their aberrant proliferation. Since Survivin
Table 2: Genes regulated by Survivin in ITD-Flt3+ KSL cells that are preferentially expressed in HSC.

| symbol | dysregulation in LSC |
|--------|----------------------|
| 2310057J16Rik | |
| 2610002J02Rik | |
| 2610042J04Rik | |
| 4930533K18Rik | |
| 9130011E15Rik | |
| Adrb3 | |
| Arl3 | |
| AW112010 | |
| Ccdn1 | |
| Cdk9 | |
| Chst1 | |
| Crem | dysregulated in LSC |
| Cyp2e1 | |
| Dnmt3b | |
| Emp1 | dysregulated in LSC |
| Erbb2ip | |
| Fmn2 | |
| Fnbp4 | |
| Foxb1 | |
| Gabpb1 | |
| Glycam1 | |
| Gpr56 | |
| Hmga2 | dysregulated in LSC |
| Ifi203 | |
| Kcnj6 | |
| Kcnmb1 | |
| Lama5 | |
| Lrrn1 | dysregulated in LSC |
| Ltbrr | |
| Maff | |
| Mup3 | dysregulated in LSC |
| Myef2 | |
| PCP4 | |
| Prox1 | |
| Rps4x | dysregulated in LSC |
| Scd2 | |
| Slc22a9 | |
| Slit2 | |
| Sos1 | dysregulated in LSC |
| Stx3 | |
| Tcf3 | |
| Traf1 | |
| Wnt6 | |
| Xrcc1 | |

is expressed and regulated in normal CD34⁺ cells [4, 6] and antagonizing Survivin reduces normal HSPC in vivo [8], it is likely that Survivin deletion will affect normal HSC in the patients. Although direct comparison of gene expression in in vitro ITD-Flt3⁺ KSL cells with in vivo CD34⁺ KSL cells may not necessarily provide definite differences in Survivin signaling between these populations in vivo, we found that 124 out of 137 Survivin associated genes shared by ITD-Flt3⁺ KSL cells and human LSC were not regulated in normal CD34⁺ KSL cells by Survivin. This disparity of downstream signaling of Survivin between normal and transformed HSC may represent as a means of identifying novel therapeutic targets against LSC that would thereby supplant direct Survivin disruption which would otherwise toxic to normal HSC.

5. Conclusion

We identified genes regulated by Survivin in ITD-Flt3 transformed KSL cells that are deregulated in human AML stem cells that are distinct from normal HSC. This study will facilitate the identification of specific therapeutic targets downstream of Survivin to eradicate LSC without affecting normal HSC.

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