Improving Antimicrobial Regimens for the Treatment of Breast Tissue Expander-Related Infections

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Background: Infectious complications in tissue expander (TE) breast reconstruction can be devastating and costly. Therefore, to optimize care, we examined patient’s demographics, microbiology of TE infections, and the efficacy of empiric antimicrobial regimens and thereafter generated an algorithm for the treatment of these complex infections.

Methods: We retrospectively reviewed all patients who underwent TE breast reconstruction between 2003 and 2012 and analyzed those patients who developed a “definite” device-related infection leading to TE explantation and had a positive intraoperative culture.

Results: A total of 3,082 patients underwent immediate breast reconstruction with TE. Of these, 378 patients (12.3%) developed an infection, 189 (6.1%) eventually proceed with explantation, and 118 (3.8%) had a positive intraoperative culture. Gram-positive organisms caused 73% of infections, and Gram-negative organisms caused 27% of infections. Narrow-spectrum empiric antimicrobials with predominantly Gram-positive coverage were deemed appropriate in only 62% of cases, and those with Gram-negative coverage were appropriate in 46%. Broad-spectrum antimicrobials were used in 47% of cases, mainly recommended by infectious disease specialists, and were considered appropriate in >90% of the occasions.

Conclusions: Current empiric antibiotic regimens do not cover the vast spectrum of organisms causing TE infections. To increase the salvage rate of an infected TE, at the first sign of infection, in addition to benefiting with an infectious diseases consultation, empiric coverage with broad-spectrum antibiotics active against biofilm-embedded organisms should be administered. (Plast Reconstr Surg Glob Open 2016;4:e704; doi: 10.1097/GOX.0000000000000690; Published online 6 May 2016.)

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discontinuation of the antibiotics within 24 hours after the procedure is complete. However, these guidelines do not address antibiotic usage in breast reconstruction with TEs, permanent implants, and drainage catheters, all of which increase the risk of infection because of the presence of foreign bodies and a thin soft-tissue envelope. Survey data from the American Society of Breast Surgeons indicated that 58% of physicians prescribed prolonged postoperative antibiotics in patients undergoing breast reconstruction immediately after mastectomy. In addition, recent survey data from the American Society of Plastic Surgeons showed that 72% of physicians continued antibiotic prophylaxis in patients who had undergone breast reconstruction until the drainage catheters were removed. This practice is in part supported by studies demonstrating that microorganisms causing surgical site infections (SSIs) are most commonly those colonizing the drain tubes and that increased duration of drain tube implantation is associated with an increased rate of SSIs.

Despite the widespread practice of maintaining postoperative antibiotic prophylaxis in patients who have undergone breast reconstruction, the reported incidence of TE infection ranges from 2.5% to 24%. Risk factors associated with TE infections include tobacco use, increased body mass index, history of radiation therapy, axillary lymph node dissection, breast reconstruction immediately after mastectomy, use of surgical drains, use of acellular dermal matrices, and adjuvant chemotherapy. Despite these known risk factors for infection, many have begun questioning the extended use of perioperative antibiotics owing to the lack of evidence-based guidelines for their use and the risk of increasing the number of resistant organisms. Recently, McCullough et al identified increased rates of microorganism resistance to first-generation cephalosporins in TE SSIs in 20.5% of preoperative and 54.5% of postoperative cases. This increase in resistance to commonly used prophylactic antibiotics underscores the importance of optimizing antibiotic choice when evaluating trends of pathogens resulting in implant-based infections.

Also, many of the empiric antimicrobial regimens currently used by surgeons were selected several decades ago. As a result of recent changes in microflora antibiotic susceptibility patterns and extensive use of perioperative antimicrobial regimens, the empiric antimicrobials that are commonly used today may be outdated. Furthermore, biofilm-embedded organisms attached to the TE, living in the extracellular matrix, are much more resistant to the inhibitory and bactericidal effects of several antibiotics than are free-floating planktonic organisms. These biofilm-embedded organisms necessitate the use of antimicrobial regimens that are not only active against the microorganisms but also able to penetrate the biofilm matrix. Once established, biofilm-related infections are often impossible to eradicate without removing the associated foreign medical device. Therefore, to evaluate the efficacy of the empiric antimicrobial regimens currently used for the treatment of TE-related infections, we examined the current microbiology of TE infections resulting in removal of the TE and the efficacy of the empiric antimicrobial treatment regimens currently used at our institution once an infection is suspected. We then used these data to generate an algorithm for adequately managing infections and avoiding eventual explantation of the breast TE.

METHODS

Patient Population

We retrospectively reviewed the medical records of all patients who underwent breast reconstruction with TE immediately after mastectomy between January 2003 and December 2012 at The University of Texas MD Anderson Cancer Center, Houston, Tex. We identified all patients in this group who developed an SSI and subsequent TE-related infection. All infections fulfilled the strict definition of a deep SSI involving an implant, as determined by the Centers for Disease Control and Prevention. According to this definition, patients met at least 1 of the following criteria: purulent drainage, spontaneous dehiscence or surgically opened incision with positive cultures, evidence of abscess, or diagnosis by the surgeon of a deep infection occurring up to 1 year postoperatively. Because several patients may develop signs and symptoms that may mimic a local infection such as radiation-induced dermatitis, increased erythema and pain secondarily to rapid increase in pressure of the infused TE, superficial skin flap necrosis, or even acellular dermal matrix (ADM)-related hyperemia, to include only patients who had developed a “definite” TE infection and avoid confounding our analysis, unless a patient had their implant removed because of infection and had a positive intraoperative culture, these patients were not included in our study.

Data Collection

We reviewed data collected in a prospectively maintained database, including patient demographic information, baseline comorbidities, breast cancer characteristics, cancer treatments, timing of TE placement, operative approach, and postoperative complications. We also documented the use of perioperative and empirical antimicrobials, which
were all confirmed via electronic chart review and inpatient and outpatient pharmacy records. The empiric antibiotics used were considered appropriate if the organism causing the TE infection was sensitive to the empiric antibiotic used. A broad-spectrum antibiotic was defined as one that is active against both Gram-positive and Gram-negative bacteria, as opposed to a narrow-spectrum antibiotic that is effective against specific families of bacteria. We also gathered information on intraoperative culture results, as well as the time from TE placement to infection, explanation, and subsequent reconstruction. Moreover, to determine trends in the evolution of microorganisms over time, we evaluated the incidence and distribution of the most common causative organisms by year. Our study was approved by the MD Anderson Institutional Review Board.

Statistical Analysis

We used descriptive statistics to summarize the data, including median with range for continuous variables lacking normal distribution and percentages for categorical variables. For 3-group comparisons, we used the Kruskal-Wallis test to compare nonparametric continuous variables and the chi-square test or Fisher exact test to compare categorical variables. When a significant result ($P < 0.05$) was detected, pairwise comparisons were performed, in which Wilcoxon rank-sum tests were used for continuous variables and the chi-square or Fisher exact test was used for categorical variables. The $\alpha$ levels of the post hoc pairwise comparisons were adjusted using a sequential Bonferroni method to control for type I error. All tests except those in the pairwise comparisons were 2-sided tests with a significance level of 0.05. Statistical analyses were performed using SAS version 9.3 (SAS Institute Inc., Cary, N.C.).

RESULTS

Clinical Characteristics

A total of 3,082 patients underwent immediate breast reconstruction with TE between 2003 and 2012. Of these, 378 (12.3%) developed an SSI. TE explantation was required in 189 patients (6.1%) owing to unresponsiveness to antimicrobial treatment or advanced stage of infection. Of the 189 patients who underwent explantation, 118 (62%) had positive intraoperative cultures (Tables 1 and 2). The mean age at the time of TE infection was 52 years (range, 28–76 years). Approximately 60% of the patients were white, 31% were Hispanic, and 9% were black or Asian. A total of 36 patients (31%) had developed an infection within the first 30 days postoperatively, 47 patients (40%) within 31 to 90 days, and 35 patients (30%) developed an infection after 90 days postoperatively ($P = 0.032$). Although >70% of the patients had no metastatic lymph node disease, the involvement of lymphoid tissues or use of preoperative chemotherapy did not affect the timing of TE infections ($P = 0.51$ for lymph node involvement and $P = 0.6$ for chemotherapy). However, infections often occurred within the early postoperative period.

Table 1. Demographic and Clinical Characteristics of Patients with Tissue Expander Infections Occurring in Different Postoperative Periods

| Variable                        | Total (n = 118) | 0–30 d (n = 36) | 31–90 d (n = 47) | >90 d (n = 35) | $P$  |
|--------------------------------|----------------|-----------------|-----------------|---------------|-----|
| Age (y), median (range)        | 52 (28–76)     | 51 (32–76)      | 55 (28–76)      | 49 (28–63)    | 0.12|
| Chest circumference (inches), median (range) | 36 (32–46) | 38 (32–46) | 36 (32–44) | 36 (34–44) | 0.09 |
| Bra cup size                   | 4 (3)          | 1 (25)          | 2 (50)          | 1 (25)        | —   |
| Hypertension                   | 36 (31)        | 15 (42)         | 15 (42)         | 6 (17)        | 0.24|
| History of tobacco use         | 35 (31)        | 8 (23)          | 13 (37)         | 14 (41)       | 0.54|
| History of ethanol use         | 34 (31)        | 10 (29)         | 16 (47)         | 8 (24)        | 0.08|
| BMI (kg/m²), median (range)    | 29 (19–52)     | 32 (22–49)      | 28 (19–49)      | 29 (19–52)    | 0.06|
| Morbid obesity (BMI ≥ 40kg/m²) | 8 (7)          | 2 (25)          | 3 (38)          | 3 (38)        | 0.91|
| Diabetes                       | 18 (15)        | 8 (44)          | 8 (44)          | 2 (11)        | 0.14|
| Coronary artery disease        | 6 (5)          | 1 (17)          | 4 (67)          | 1 (17)        | 0.5 |
| Renal insufficiency            | 5 (4)          | 2 (40)          | 3 (60)          | 0             | 0.38|
| Cerebrovascular disease        | 3 (3)          | 0               | 0               | 3 (100)       | 0.025|

Data are presented as n (%), unless otherwise indicated. Percentages were rounded up and may not add to 100%. Percentages in the first column have been obtained by dividing the amount of patients in each variable by the total 118 patients, whereas in the remaining 3 columns, the percentages have been obtained by dividing the amount of patients within each postoperative variable, by the total amount of patients encountered for each given variable (column 1).

BMI, body mass index.
(<30 days) in patients with the largest chest circumferences and those with increased body mass index, although these associations were not statistically significant \((P = 0.09\) for chest circumference and \(P = 0.06\) for body mass index). Patients with relatively large tumors and those who received postoperative radiotherapy were significantly more likely to develop an infection during the late postoperative period (>90 days) than the early postoperative period \((P<0.02)\).

In addition, more than 70% of the patients had a skin-sparing mastectomy. An ADM was used in 44 cases (37%), and despite being a risk factor for seromas and persistence of the use of a drainage catheter, the use of this bioprosthetic mesh did not appear to be a risk factor for early infections \((P = 0.92)\). Similarly, initial and final expansion volumes did not predict risk of early or late TE infection \((P>0.92)\). Furthermore, almost 40% of the patients who developed a TE infection had developed some degree of skin flap necrosis, and 28% developed a postsurgical seroma or hematoma, requiring drainage catheter placement for an extended period.

### Prophylactic Antimicrobial Regimens
All patients received a standardized perioperative systemic antimicrobial therapy; the most common drugs used were cefazolin (81%), clindamycin (6%), and ampicillin-sulbactam (4%). Thereafter at the discretion of the surgeon, more than 85% of the patients underwent subpectoral TE pocket irrigation with a broad-spectrum antimicrobial solution. The most commonly used regimen was bacitracin plus polymyxin B (66%), followed by bacitracin plus cefazolin and gentamicin (9%) and vancomycin plus ciprofloxacin (4%). After surgery, oral prophylactic antibiotics were used in 78% of cases, either for a week or until the drainage catheters were removed. The most commonly prescribed postsurgical antimicrobial drugs were cephalexin (26%), cefadroxil (22%), amoxicillin/clavulanate (8%), trimethoprim/sulfamethoxazole (6%), and clindamycin (4%).

### Time to Infection and Implant Removal
Once an infection occurred, the median time from initial reconstructive surgery to infection was 47 days (range, 13–309 days, with a 25th and 75th percentile of 26 and 107 days, respectively). Overall, the median time from infection to implant removal was 4 days (range, 0–136 days, with a 25th and 75th percentile of 2 and 8 days, respectively).

### Microorganisms
Gram-positive organism infections were identified in 73% of patients, and Gram-negative infections

### Table 2. Cancer, Treatment, and Surgical Characteristics of Patients with Infections Occurring in Different Postoperative Periods

| Variable                                      | Total (n = 118) | 0–30 d (n = 36) | 31–90 d (n = 47) | >90 d (n = 35) | P       |
|------------------------------------------------|-----------------|-----------------|-----------------|---------------|---------|
| Breast cancer laterality                       |                 |                 |                 |               | 0.67    |
| Right                                          | 52 (44)         | 18 (35)         | 20 (38)         | 14 (27)       | —       |
| Left                                           | 66 (56)         | 18 (27)         | 27 (41)         | 21 (32)       | —       |
| Breast cancer involvement                      |                 |                 |                 |               | 0.022   |
| T0                                             | 25 (21)         | 11 (44)         | 8 (32)          | 6 (24)        | —       |
| T1                                             | 15 (13)         | 7 (47)          | 6 (40)          | 2 (13)        | —       |
| T2                                             | 36 (31)         | 12 (33)         | 17 (47)         | 7 (19)        | —       |
| T3                                             | 28 (24)         | 2 (7)           | 13 (46)         | 13 (46)       | —       |
| T4                                             | 11 (9)          | 3 (27)          | 3 (27)          | 5 (45)        | —       |
| T1                                             | 3 (3)           | 1 (33)          | 0               | 2 (67)        | —       |
| Lymph node involvement                        |                 |                 |                 |               | 0.51    |
| N0                                             | 86 (73)         | 29 (34)         | 36 (42)         | 21 (24)       | —       |
| N1                                             | 22 (19)         | 5 (23)          | 7 (32)          | 10 (45)       | —       |
| N2                                             | 3 (3)           | 1 (33)          | 1 (33)          | 1 (33)        | —       |
| N3                                             | 7 (6)           | 1 (14)          | 3 (43)          | 3 (43)        | —       |
| Preoperative chemotherapy                      | 36 (31)         | 10 (28)         | 13 (36)         | 13 (36)       | 0.6     |
| Skin-sparing mastectomy                       | 88 (75)         | 29 (33)         | 34 (39)         | 25 (28)       | 0.61    |
| Bioprosthetic mesh placement                   |                 |                 |                 |               | 0.92    |
| None                                           | 74 (63)         | 23 (31)         | 30 (41)         | 21 (28)       | —       |
| AlloDerm or SurgiMend                          | 44 (37)         | 13 (30)         | 17 (39)         | 14 (32)       | —       |
| Initial tissue expansion (ml), median (range)  | 250 (0–850)     | 175 (50–700)    | 250 (50–850)    | 250 (0–750)   | 0.92    |
| Final tissue expansion (ml), median (range)    | 550 (200–850)   | 550 (250–800)   | 550 (200–850)   | 550 (250–800) | 0.36    |
| Postoperative radiotherapy                     | 14 (12)         | 0               | 6 (43)          | 8 (57)        | 0.005   |
| Postsurgical seroma or hematoma                | 35 (28)         | 11 (33)         | 16 (48)         | 6 (18)        | 0.22    |
| Postsurgical skin flap necrosis                | 46 (39)         | 12 (26)         | 19 (41)         | 15 (33)       | 0.69    |

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were identified in 27% (Table 3). A polymicrobial infection was identified in 20 patients (17%). Methicillin-resistant *Staphylococcus epidermidis* (24%) was the most frequently encountered organism, followed by methicillin-sensitive *Staphylococcus aureus* (15%), *Pseudomonas* spp. (14%), and methicillin-resistant *S. aureus* (12%). The frequency of each causative microorganism remained relatively stable throughout the period studied; methicillin-resistant *S. epidermidis* was the most common microorganism throughout. These also remained stable when stratified by different postoperative time periods (Fig. 1). Of interest, *Pseudomonas* and other facultative Gram-negative rods were predominant at 30 days after surgery (accounting for 31% of TE infections), as well as at 90 days (27% of infections) and 180 days (25% of infections; Fig. 1). Furthermore, during the past 8 years of our study, the incidence of Gram-negative TE infections ranged from 24% to 44% of all TE infections (Fig. 2). Of note, similar to the proportion of patients without ADMs, of the 37 patients with an ADM, 62% had a Gram-positive infection and 35% had a Gram-negative infection with 6 (16%) had a polymicrobial infection.

**Empiric Antimicrobial Regimens**

Multiple empiric antimicrobial regimens were used to treat TE infections (Table 4). The most ineffective empiric regimens were categorized as narrow spectrum, such as the isolated use of vancomycin (12 cases, 10%), amoxicillin/clavulanate (10 cases, 8%), cephalaxin (8 cases, 7%), or ciprofloxacin (5 cases, 4%). As a group, narrow-spectrum antimicrobial regimens with predominantly Gram-negative coverage (20 cases, 17%) were deemed appropriate in only 45% of cases, and narrow-spectrum regimens with predominantly Gram-positive coverage (16 cases, 13%) were deemed appropriate in only 40% of cases. Of note, a polymicrobial infection was identified in 20 patients (17%). Methicillin-resistant *Staphylococcus epidermidis* (24%) was the most frequently encountered organism, followed by methicillin-sensitive *Staphylococcus aureus* (15%), *Pseudomonas* spp. (14%), and methicillin-resistant *S. aureus* (12%). The frequency of each causative microorganism remained relatively stable throughout the period studied; methicillin-resistant *S. epidermidis* was the most common microorganism throughout. These also remained stable when stratified by different postoperative time periods (Fig. 1). Of interest, *Pseudomonas* and other facultative Gram-negative rods were predominant at 30 days after surgery (accounting for 31% of TE infections), as well as at 90 days (27% of infections) and 180 days (25% of infections; Fig. 1). Furthermore, during the past 8 years of our study, the incidence of Gram-negative TE infections ranged from 24% to 44% of all TE infections (Fig. 2). Of note, similar to the proportion of patients without ADMs, of the 37 patients with an ADM, 62% had a Gram-positive infection and 35% had a Gram-negative infection with 6 (16%) had a polymicrobial infection.

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Fig. 2. Proportion of the most common organisms causing tissue expander infections in different years of our study. The numerator and denominator for these percentages represent the total number of patients included for that specific time period. Other Gram-negative rods include *Klebsiella*, *Serratia*, *Morganella*, *Proteus*, and *Escherichia coli*. MRSA, methicillin-resistant *Staphylococcus aureus*; MRSE, methicillin-resistant *S. epidermidis*; MSSA, methicillin-sensitive *S. aureus*.

| Antimicrobial Regimens                              | Total, n (%) | Appropriate Antibiotic, n (%)* |
|-----------------------------------------------------|--------------|--------------------------------|
| None                                                | 9 (8)        | 0                              |
| Narrow spectrum, predominantly Gram-negative coverage| 20 (17)      | 9 (46)                         |
| Amoxicillin/clavulanate                             | 10 (9)       | 330                            |
| Ciprofloxacin                                       | 5 (4)        | 4 (80)                         |
| Ampicillin/sulbactam                                | 3 (3)        | 0 (0)                          |
| Gatifloxacin                                        | 2 (2)        | 2 (100)                        |
| Narrow spectrum, predominantly Gram-positive coverage| 34 (29)      | 21 (62)                        |
| Vancomycin                                          | 12 (10)      | 9 (75)                         |
| Cephalexin                                          | 8 (7)        | 6 (75)                         |
| Linezolid                                           | 4 (3)        | 2 (50)                         |
| Cefazolin                                           | 4 (3)        | 2 (50)                         |
| Minocycline                                         | 2 (2)        | 1 (50)                         |
| TMP-SMX/taizhromycin                                | 1 (1)        | 0 (0)                          |
| TMP-SMX/clindamycin                                 | 1 (1)        | 0 (0)                          |
| Broad spectrum, for relatively sensitive organisms   | 6 (5)        | 5 (88)                         |
| TMP-SMX/ciprofloxacin                               | 3 (3)        | 3 (100)                        |
| Cephalexin/ciprofloxacin                            | 2 (2)        | 1 (50)                         |
| Minocycline/ampicillin/sulbactam/levofloxacin       | 1 (1)        | 1 (100)                        |
| Broad spectrum, for relatively resistant organisms   | 49 (42)      | 45 (92)                        |
| Vancomycin/ciprofloxacin                            | 18 (15)      | 18 (100)                       |
| Vancomycin/piperacillin/tazobactam                  | 9 (8)        | 8 (89)                         |
| Daptomycin/piperacillin/tazobactam                  | 8 (7)        | 8 (100)                        |
| Vancomycin/celepine                                 | 4 (3)        | 4 (100)                        |
| Vancomycin/levofloxacin                             | 3 (3)        | 3 (100)                        |
| Daptomycin/ciprofloxacin                            | 2 (2)        | 2 (100)                        |
| Vancomycin/cephalexin                               | 2 (2)        | 0 (0)                          |
| Daptomycin/celepine                                 | 2 (2)        | 1 (50)                         |
| Vancomycin/metopopenem                              | 1 (1)        | 1 (100)                        |
| Total                                               | 118 (100)    | 80 (68)                        |

*Empiric antibiotics were deemed appropriate if the organism was sensitive to the empiric antimicrobial regimen. TMP-SMX, trimethoprim-sulfamethoxazole.
antimicrobial regimens with predominantly Gram-positive coverage (34 cases, 29%) were appropriate in only 62% of cases. In contrast, as a group, the use of broad-spectrum antimicrobial regimens (55 cases, 47%) was considered appropriate in more than 90% of the cases in which they were used. The most commonly used regimens were the combination of vancomycin plus piperacillin and tazobactam (18 cases, 15%), vancomycin plus ciprofloxacin (9 cases, 8%), and daptomycin plus piperacillin and tazobactam (8 cases, 7%). Interestingly, we discovered that the majority of plastic surgeons were providing a narrow-spectrum empiric antibiotic regimen, compared with the infectious disease specialists whom were consulted on all of the patients in whom broad-spectrum antimicrobials were recommended.

Subsequent Breast Reconstruction

Once the infection was adequately treated, 60 patients (51%) decided to proceed with a subsequent reconstructive procedure. The median time from implant removal to subsequent reconstruction was 180 days (range, 30–1,292 days). Twenty-eight patients (47%) proceeded with an implant-based reconstruction, 22 patients (37%) underwent a flap reconstruction, and 10 patients (17%) underwent a combined implant-flap reconstructive procedure. Patients who underwent a flap reconstruction were almost equally split in terms of the type of flap; approximately one-third had an extended latissimus dorsi flap, one-third had a deep inferior epigastric perforator flap, and one-third had a muscle-sparing transverse rectus abdominis myocutaneous flap.

DISCUSSION

The rate of postoperative TE-related infections continues to remain unacceptably high despite the use of several prophylactic antimicrobial regimens. Once an infection occurs, from the very onset, it is paramount to optimize the empiric antibiotic regimen to successfully salvage the implant, decrease overall morbidity, and avoid delays in adjuvant therapy. Our results demonstrate that empiric antimicrobial regimens currently used for the treatment of TE infections are diverse and do not generally cover the spectrum of organisms that cause TE infections, including methicillin-resistant staphylococci and Gram-negative rods.

Similar to previous studies, our data show that the organisms causing TE infection and implant loss are usually resistant to commonly used postoperative prophylactic regimens that target predominantly Gram-positive bacteria, including first-generation cephalosporins. In addition, approximately two-thirds of these infections occur more than 30 days after surgery, which is often beyond when most drains have been removed and antibiotic regimens discontinued. It has been speculated that prophylactic antibiotics prevent early infections caused by nonresistant bacteria while selecting for resistant organisms that can lead to late infection.

Consensus has not been reached on either the use of postoperative prophylactic antibiotics or the appropriate overall duration needed to prevent infections. Several studies have revealed that following Surgical Care Improvement Project guidelines in prosthetic breast reconstruction and withholding postoperative prophylactic antibiotics can increase a patient’s SSI reoperation risk 4.7 times. In addition, stopping antibiotic prophylaxis after 24 hours could raise the rate of infection requiring the removal of a TE from 7.9% to 31%.

In these studies, the bacteria isolated in the groups not treated with an extended course of postoperative prophylactic antibiotics were more diverse than those seen in other studies in which patients used long-term postoperative antibiotics.

Targeting postoperative prophylaxis to patients at high risk for infection has been suggested as an alternative to universal postoperative prophylaxis in prosthetic breast reconstruction to prevent adverse outcomes from extended use, such as increasing resistance, allergic reactions, and other infections associated with antibiotic use. A heightened awareness of those patients who are at an increased risk for infections including those who are obese, smokers, those with mastectomy skin necrosis, or a need for radiation therapy should be considered for prolonged use of prophylactic postoperative antibiotics.

Once an SSI or TE infection is diagnosed, it is paramount to treat the infection with an empiric broad-spectrum antimicrobial regimen that will not only cover the most common causative organisms but also remain active among the organisms embedded in an early biofilm infection. Therefore, after reviewing the perioperative and postoperative antimicrobials used in our patient population, as well as the organisms responsible for TE infections, we have developed a comprehensive treatment algorithm (Fig. 3) to provide a more consistent approach to a patient presenting with a breast TE infection.

In our algorithm, we encourage all patients to proceed with a breast ultrasound. If any periprosthetic fluid is observed, ideally the fluid should be aspirated and submitted for microbiological analysis to determine whether bacteria, fungi, or acid-fast bacilli is present. In the meantime, while awaiting culture results, and after reviewing the patient’s medication and allergy profile, including any antimicrobial drugs used in the patient’s perioperative and postoperative
prophylactic regimen, depending on the severity of the infection, the physician should prescribe either an oral or intravenous empiric antimicrobial regimen that targets methicillin-resistant staphylococci and Gram-negative rods including *Pseudomonas*. Furthermore, the empiric antimicrobial regimen should favor antibiotics that are active against the bacteria embedded in the biofilm. For example, for Gram-positive coverage, oral minocycline plus rifampin or intravenous daptomycin plus rifampin should be favored over vancomycin or linezolid. For Gram-negative coverage, quinolone, such as ciprofloxacin, should be prioritized over a β-lactam antimicrobial regimen. If the culprit organism has been eventually recovered, usually within the first 48 hours, the empiric regimen should be transitioned to a specific antimicrobial regimen that: (a) the microorganism is sensitive to and, (b) is active against biofilm-embedded organisms. We cannot comment on the potential that withholding prophylaxis would have caused a different number of infections and TE losses. Evidence from a randomized controlled trial would be required to challenge the well-established practice patterns, which are currently based on anecdotal experience rather than evidence-based data.

**CONCLUSIONS**

To salvage an infected breast TE, based on individual hospital epidemiological data, the most common organisms causative for TE infection should be empirically covered. At our institution, methicillin-resistant...
resistant staphylococci and Gram-negative rods, including *Pseudomonas*, were responsible for more than 60% of all TE infections across different post-operative time periods and years of our study. Therefore, these organisms should be empirically covered by broad-spectrum antibiotics that can also act on biofilm-embedded organisms, rather than by traditional narrow-spectrum regimens such as first-generation cephalosporins alone. Once an organism has been identified, the empiric antimicrobial regimen should be transitioned to a specific biofilm-active antimicrobial regimen dictated by the microorganism’s antimicrobial susceptibility panel. Finally, to optimize the care of these patients, it is of benefit to have an infectious diseases consultant assist in the care of these complex medical device-related infections.

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**REFERENCES**

1. Fry DE. Surgical site infections and the surgical care improvement project (SCIP): evolution of national quality measures. *Surg Infect (Larchmt)*. 2008;9:579–584.
2. Brahmbhatt RD, Huebner M, Scow JS, et al. National practice patterns in preoperative and postoperative antibiotic prophylaxis in breast procedures requiring drains: survey of the American Society of Breast Surgeons. *Ann Surg Oncol*. 2012;19:3205–3211.
3. Phillips BT, Wang ED, Mirrer J, et al. Current practice among plastic surgeons of antibiotic prophylaxis and closed-suction drains in breast reconstruction: experience, evidence, and implications for postoperative care. *Ann Plast Surg*. 2011;66:460–465.
4. Crosby MA, Dong W, Feng L, et al. Effect of intraoperative saline fill volume on perioperative outcomes in tissue expander breast reconstruction. *Plast Reconstr Surg*. 2011;127:1065–1072.
5. Nahabedian MY, Tsangaris T, Momen B, et al. Infectious complications following breast reconstruction with expanders and implants. *Plast Reconstr Surg*. 2003;112:467–476.
6. Cordeiro PG, McCarthy CM. A single surgeon’s 12-year experience with tissue expander/implant breast reconstruction: part I. A prospective analysis of early complications. *Plast Reconstr Surg*. 2006;118:825–831.
7. Armstrong RW, Berkowitz RL, Bolding F. Infection following breast reconstruction. *Ann Plast Surg*. 1989;23:284–288.
8. Spear SL, Majidian A. Immediate breast reconstruction in two stages using textured, integrated-valve tissue expanders and breast implants: a retrospective review of 171 consecutive breast reconstructions from 1989 to 1996. *Plast Reconstr Surg*. 1998;101:53–63.
9. Francis SH, Ruberg RL, Stevenson KB, et al. Independent risk factors for infection in tissue expander breast reconstruction. *Plast Reconstr Surg*. 2009;124:1790–1796.
10. Handel N, Jensen JA, Black Q, et al. The fate of breast implants: a critical analysis of complications and outcomes. *Plast Reconstr Surg*. 1995;96:1521–1533.
11. Cordeiro PG, Pusic AL, Disa JJ, et al. Irradiation after immediate tissue expander/implant breast reconstruction: outcomes, complications, aesthetic results, and satisfaction among 156 patients. *Plast Reconstr Surg*. 2004;113:877–881.
12. Araco A, Gravante G, Araco F, et al. Infections of breast implants in aesthetic breast augmentations: a single-center review of 3,002 patients. *Aesthetic Plast Surg*. 2007;31:325–329.
13. Degnim AC, Scow JS, Hoskin TL, et al. Randomized controlled trial to reduce bacterial colonization of surgical drains after breast and axillary operations. *Ann Surg*. 2013;258:240–247.
14. Vandeweyer E, Deraemaeker R, Nogaret JM, et al. Immediate breast reconstruction with implants and adjuvant chemotherapy: a good option? *Acta Chir Belg*. 2003;103:98–101.
15. McCullough MC, Chu CK, Duggal CS, et al. Antibiotic prophylaxis and resistance in surgical site infection after immediate tissue expander reconstruction of the breast. *Ann Plast Surg*. 2014;73(2):1–5.
16. Costerton JW, Montanaro L, Arciola CR. Biofilm in implant infections: its production and regulation. *Int J Artif Organs*. 2005;28:1062–1068.
17. Horan TC, Andrus M, Dudeck MA. CDC/NHSN surveillance definition of health care-associated infection and criteria for specific types of infections in the acute care setting. *Am J Infect Control*. 2008;36:309–332.
18. Phillips BT, Bishawi M, Dagum AB, et al. A systematic review of antibiotic use and infection in breast reconstruction: what is the evidence? *Plast Reconstr Surg*. 2013;131:1–13.
19. Weichman KE, Levine SM, Wilson SC, et al. Antibiotic selection for the treatment of infectious complications of implant-based breast reconstruction. *Ann Plast Surg*. 2013;71:140–148.
20. Clayton JL, Bazakas A, Lee CN, et al. Once is not enough: withholding postoperative prophylactic antibiotics in prosthetic breast reconstruction is associated with an increased risk of infection. *Plast Reconstr Surg*. 2012;130:495–502.
21. Avashia YJ, Mohan R, Berhane C, et al. Postoperative antibiotic prophylaxis for implant-based breast reconstruction with acellular dermal matrix. *Plast Reconstr Surg*. 2013;131:453–461.
22. Ahn CY, Ko CY, Wagar EA, et al. Microbial evaluation: 139 implants removed from symptomatic patients. *Plast Reconstr Surg*. 1996;98:1225–1229.
23. Reish RG, Danjanovic B, Austen WG, Jr, et al. Infection following implant-based reconstruction in 1952 consecutive breast reconstructions: salvage rates and predictors of success. *Plast Reconstr Surg*. 2013;131:1223–1230.
24. Leyngold MM, Stutman RL, Khiani KT, et al. Contributing variables to post mastectomy tissue expander infection. *Breast J*. 2012;18:351–356.
25. McCarthy CM, Mehrara BJ, Riedel E, et al. Predicting complications following expander/implant breast reconstruction: an outcomes analysis based on preoperative clinical risk. *Plast Reconstr Surg*. 2008;121:1886–1892.
26. Peled AW, Stover AC, Foster RD, et al. Long-term reconstructive outcomes after expander-implant breast reconstruction with serious infectious or wound-healing complications. *Ann Plast Surg*. 2012;68:369–373.
27. Viola GM, Raad II, Rolston KV. Breast tissue expander-related infections: perioperative antimicrobial regimens. *Infect Control Hosp Epidemiol*. 2014;35:75–81.

28. Macadam SA, Mehling BM, Fanning A, et al. Nontuberculous mycobacterial breast implant infections. *Plast Reconstr Surg*. 2007;119:337–344.

29. Vinh DC, Rendina A, Turner R, et al. Breast implant infection with Mycobacterium fortuitum group: report of case and review. *J Infect*. 2006;52:e63–e67.

30. Singh R, Ray P, Das A, et al. Penetration of antibiotics through Staphylococcus aureus and Staphylococcus epidermidis biofilms. *J Antimicrob Chemother*. 2010;65:1955–1958.

31. Rodríguez-Martínez JM, Ballesta S, García I, et al. [Activity and penetration of linezolid and vancomycin against Staphylococcus epidermidis biofilms]. *Enferm Infecc Microbiol Clin*. 2007;25:425–428.

32. Darouiche RO, Dhir A, Miller AJ, et al. Vancomycin penetration into biofilm covering infected prostheses and effect on bacteria. *J Infect Dis*. 1994;170:720–723.

33. Jefferson KK, Goldmann DA, Pier GB. Use of confocal microscopy to analyze the rate of vancomycin penetration through Staphylococcus aureus biofilms. *Antimicrob Agents Chemother*. 2005;49:2467–2473.

34. Raad I, Hanna H, Jiang Y, et al. Comparative activities of daptomycin, linezolid, and tigecycline against catheter-related methicillin-resistant Staphylococcus bacteremic isolates embedded in biofilm. *Antimicrob Agents Chemother*. 2007;51:1656–1660.

35. Kwiecieńska-Piróg J, Bogiel T, Gospodarek E. Effects of ceftazidime and ciprofloxacin on biofilm formation in Proteus mirabilis rods. *J Antimicrob (Tokyo)* 2013;66:593–597.

36. Bowler LL, Zhanel GG, Ball TB, et al. Mature Pseudomonas aeruginosa biofilms prevail compared to young biofilms in the presence of ceftazidime. *Antimicrob Agents Chemother*. 2012;56:4976–4979.

37. Drago L, Mattina R, Legnani D, et al. Modulation of biofilm of strains isolated from patients with chronic obstructive pulmonary disease by levofloxacin, moxifloxacin, ciprofloxacin, amoxicillin/clavulanic acid and ceftriaxone. *Int J Immunopathol Pharmacol*. 2011;24:1027–1035.

38. Naves P, Del Prado G, Ponte C, et al. Differences in the in vitro susceptibility of planktonic and biofilm-associated Escherichia coli strains to antimicrobial agents. *J Chemother*. 2010;22:312–317.

39. Bret L, Di Martino P. Effect of ceftazidime, amikacin and ciprofloxacin on biofilm formation by some enterobacterial clinical isolates. *Chemotherapy* 2004;50:255–259.

40. Zimmerli W, Trampuz A, Ochsner PE. Prosthetic-joint infections. *N Engl J Med*. 2004;351:1645–1654.

41. Baddour LM, Epstein AE, Erickson CC, et al. Update on cardiovascular implantable electronic device infections and their management: a scientific statement from the American Heart Association. *Circulation* 2010;121:458–477.