**Urogonimus turdi** (Digenea: Leucochloridiidae) from the White’s Thrush, *Zoothera aurea*, in the Republic of Korea

Hyeon Cheol Kim*1, Eui Ju Hong*1, Si Yun Ryu2, Jinho Park3, Do Hyeon Yu4, Joon Seok Chae5, Kyoung Seong Choi6, Cheolho Sim7, Bae Keun Park8,*

1College of Veterinary Medicine and Institute of Veterinary Science, Kangwon National University, Chuncheon 24341, Korea; 2College of Veterinary Medicine, Chungnam National University, Daejeon 34134, Korea; 3College of Veterinary Medicine, Chonbuk National University, Cheongju 28644, Korea; 4Laboratory of Veterinary Internal Medicine, BK21 PLUS Program for Creative Veterinary Science Research, Research Institute for Veterinary Science and College of Veterinary Medicine, Seoul National University, Seoul 08826, Korea; 5College of Ecology and Environmental Science, Kyungpook National University, Sangu 37224, Korea; 6Department of Biology, Baylor University, Waco, Texas 76798, USA

Abstract: Avian trematodes, *Urogonimus turdi* (Digenea: Leucochloridiidae), were collected from the intestine of wild birds, *Zoothera aurea*, 2013-2017 in the Daejeon Metropolitan City, Korea. The body was ellipsoidal, attenuated and/or round ends, 1,987-2,120 long and 819-831 μm wide. The oral sucker was subterminal, rounded anteriorly, and 308-425 × 351-432 μm in size; the prepharynx and esophagus were almost lacking; pharynx was well-developed, 142-179 × 78-170 μm in size; intestine narrow, bifurcating just after pharynx, ascending to the oral sucker before looping posteriorly and terminating near the posterior end; ventral sucker larger, in almost median, 536-673 × 447-605 μm and approximately 1.5 times larger than oral sucker. A phylogenetic tree constructed with 18S ribosomal RNA showed inter- and intraspecific relationships. Based on these morphological and molecular findings, we report here a *U. turdi* from White’s thrushes in Korea.

Key words: *Urogonimus turdi*, *Zoothera aurea*, White’s thrush, Korea

INTRODUCTION

Genus *Urogonimus* belongs to family Leucochloridiidae Poche, 1907 comprising 2 other genera, *Urotocus* Looss, 1899 and *Leucochloridium* Carus, 1835. Fluke members of the genus *Urogonimus* are morphologically distinguishable from those of the other 2 genera due to the location of their genital pores [1]. Records of at least 5 species, namely, *U. cardis*, *U. caroxyactis*, *U. certhiae*, *U. dryobatae*, and *U. macrostomus* have been found in literature. Their validities are still obscure and debatable except *U. macrostomus*. Locke et al. [2] have found molecular and morphological evidences from Holarctic samples that support the validity of *U. macrostomus*.

Trematodes belonging to the Leucochloridiidae were found in many wild birds including passerine and shorebirds [3-6].

In Japan, *Leucochloridium japonicum* was collected from the copper pheasant (*Syrmaticus soemmerringii*) [6]. Other similar species such as *L. sine* collected from the hawfinch (*Coccothraustes coccothraustes*) [7], and *L. cardis* and *L. turdi* collected from the Japanese thrush (*Turdus cardis*) [8]. Additionally, *U. macrostomus* was found in the rustic bunting (*Emberiza rustica*) [9]. Meanwhile, Kagan [10] classified *L. cardis* and *L. turdi* into the genus *Urogonimus* based on the differences in their reproductive organs and the arrangement of their uterine loops, which resulted in the recent revision of the species differentiation in Leucochloridiidae.

Although species belonging to the genus *Leucochloridium* are known to be common bird parasites worldwide [11-13], no species of parasites in the Leucochloridiidae has been reported among the wild bird populations in the Republic of Korea.

White’s thrush, *Zoothera aurea*, is one of the 16 species of the family Turididae recorded in Korea and belongs to the order Passeriformes [14]. White’s thrush is a carnivorous bird that usually feeds on invertebrates including insects, their larvae, small slugs, and snails [15,16]. This bird is distributed throughout Korea and often reproduces in forests and parks, there is...
little research on this. Especially, there is no study on parasitic infestation of the Z. aurea.

In this study, for the first time, White’s thrush was identified as a natural host of U. turdi in Korea. Therefore, we are going to confirm the characteristics of U. turdi by morphological and phylogenetic analysis.

**MATERIALS AND METHODS**

Sample collection and histological processing

Six dead bodies of White’s thrushes were dissected from January 2013 to August 2017 in the Daejeon city, Korea. These birds collected from the wildlife rescue center in Daejeon were immediately frozen after death and the flukes were subsequently recovered after thawing the bodies. One hundred and fifty-six flukes were harvested from the intestines of these birds. Twenty flukes were fixed with 10% formaldehyde without performing the flattening procedure for the light microscopic observation. The worms were stained with Semichon’s acetocarmine, cleared with xylene, and mounted with Canada balsam. Additionally, to observe their internal organs, 5 flukes were embedded in paraffin, sectioned into slices of 4 µm, and then stained with hematoxylin and eosin. Measurements of the specimens are presented in micrometer as the range, with the mean in parentheses.

Scanning electron microscopy (SEM)

For the SEM examination, 5 parasites were washed 5 times with 0.2 M cacodylate buffer (pH 7.3), fixed in 2.5% glutaraldehyde, and then post-fixed in 1% osmium tetroxide at 4°C. The specimens were dehydrated in a graded ethyl alcohol series, dried with a critical point CO₂ dryer, coated with gold and examined using SEM (S-4800, Hitachi) at 15 kV.

Sequencing and phylogenetic analysis

Twenty flukes were fixed in 100% ethyl alcohol for the DNA sequencing. Genomic DNA was extracted from the collected parasites according to the manufacturer’s instructions using a DNeasy® Blood & Tissue Kit (Qiagen, Alameda, California, USA). Using 18S rRNA gene sequences of Leucoclinidium, primers for common trematodes were designed using an online tool (Primer3Plus, http://www.bioinformatics.nl/cgi-bin/primer3plus/primer3plus.cgi). The oligonucleotide sequences of primers were employed to detect the L. paradoxum 18S rRNA gene (DNA) were 5’-AGGGAATGGGTTGATTT-3’ (left primer) and 5’-AGACACGACTGAAAGGTTGC-3’ (right primer). The primer sets were designed around 1,397 bp of the product, variable for all sequences aligned. Under standard conditions like 95°C for 30 sec, annealing at 60°C for 30 sec and extension at 72°C for 1 min, the polymerase chain reaction (PCR) was performed in a MyCycler Personal Thermal Cycler (Bio-Rad Laboratories, Hercules, California, USA) using the EmeraldAmp GT PCR Master Mix (Takara, Shiga, Japan) with a 1 µl aliquot of DNA. Next, the PCR products were visualized via electrophoresis on a 1.2% agarose gel and then purified using the QIAquick PCR Purification Kit (Qiagen). PCR amplicons were directly sequenced with the same primers as those used in PCR using the ABI Prism BigDye Terminator v3.0 Ready Reaction Cycle Sequencing kits (Applied Biosystems, Foster City, California, USA). The sequencing data revealed around 726 bp of DNA. Furthermore, the sequence data were aligned using the Clustal Omega program (Clustal O 1.2.1). The phylogenetic tree was constructed based on the sequence analysis conducted using the neighbor-joining method by Basic Local Alignment Search Tool (BLAST) analysis. The neighbor-joining methods were based on a guide tree that provided pairwise and multiple alignment parameters [17].

**RESULTS**

Morphological features of U. turdi

Morphological characteristics observed in the collected specimens were as follows: the bodies were elongated, relatively wide with rounded extremities that were usually widest at the acetabular levels and 1,987-2,120 long by 819-831 wide by 3.9:1 (Fig. 1A, C). The intestinal ceca were somewhat spherical (Fig. 2B). The genital pores were observed at the posterior ends of the bodies were median and...
ventro-terminal (Figs. 2, 3B). The reproductive glands occupied majority of the post-acetabular, intra-cecal areas and they were arranged in a triangle. The ovaries were spherical, 145-175 × 111-135 (161 × 129) in size, situated behind the ventral suckers and lay symmetrically to the left testes (Fig. 2). Uterine seminal receptacles were present; ootypes were surrounded by the Mehlis’ gland, present to the right of the ovaries and in the post-testicular regions. The Laurer’s canals opened posteriorly towards the dorsal side (Fig. 3C, D). The vitellaria folliculars were found to begin adjacent to the levels of the posterior...
margins of the oral suckers and end at or near the levels of the right testes (Fig. 1). The uteri were filled with eggs and confined to the intercecal areas. The eggs were oval, embryonated, and 19.6-26.0 × 12.2-15.8 (23.6 × 14.6) in size.

In SEM study, the worms were found to have flat leaf-like bodies with prominent oral and ventral suckers (Fig. 3A). The
bodies were covered with a smooth cytoplasmic layer and no tegumental spines were observed (Fig. 3D). The 2 suckers were remarkably well-developed. The ventral suckers were approximately 1.5 times larger than the oral suckers and located at the center of the bodies (Fig. 3A). The genital pores were situated at the median and ventro-terminal regions. The unspined cirri appeared from round to cylindrical (as the fully evaginated cirrus) in outline (Fig. 3B). The Laurer’s canals opened on the dorsal surfaces of the bodies (Fig. 3C).

Phylogenetic relationship

To identify the genotype of the *Urogonimus* specimens in relation to their phylogenetic relationship with Leucochloridiidae, the genomic DNA samples of the specimens were compared by phylogenetic analysis (Fig. 4). The partial sequence of the 18S rRNA in *Urogonimus* specimens was identified and compared to those of the parasites such as *Leucochloridium holostomum* (KP903629), *L. paradoxum* (KP903632), and *L. perturbatum* (KP903641). As a PCR amplicon, partial sequences (540 bp) of the *Urogonimus* specimens contained 15 variable sites of a common sequence which belonged to the clades of *L. holostomum*, *L. paradoxum*, and *L. perturbatum*, respectively. The partial sequences of the 18S rRNA gene of the *Urogonimus* specimens showed high similarity with those of the parasites *L. holostomum* (97.6%), *L. paradoxum* (97.6%), *L. perturbatum* (97.4%), *Michajlovia migrata* (99.3%), *Urotocus rossitensis* (99.3%), and *U. macrostomus* (98.5%). The results of the phylogenetic analysis validate the fact that these samples belonged to the same clades as *Urogonimus* based on their key morphological features. After the sequence of *Urogonimus* specimens was compared with several genes, the sequence data was inferred using the neighbor-joining method (Fig. 4). The sequence of *Urogonimus* specimens was also identified to be of *U. turdi*, which is genetically close to *M. migrata* (99.3%), *U. rossitensis* (99.3%), and *U. macrostomus* (98.5%). Although the morphology of *U. turdi* was also found to match with the above *Leucochloridium* sp., the identification strongly supported the fact that the samples are closer to the clades of the *Urogonimus* species. Finally, we submitted the sequence data to GenBank and received the GenBank accession number for our nucleotide sequence (BankIt MK602324).

**DISCUSSION**

Despite recent researches on Leucochloridiidae, there has been a great confusion regarding their genera, species, synonyms, nomenclature, and systematic position [1,10,18-20]. Currently, the Leucochloridiidae consists of the 3 genera *Urotoxus* Looss, 1899, *Urogonimus* Monticelli, 1888, and *Leucochloridium* Carus, 1835. These 3 genera can be distinguished based on the proportions of their suckers, position of their ventral sucker, arrangement of their gonads, position of their genital pores, and course of the uterus. *Michajlova Pojmanska, 1973* was included in Leucochloridiidae by Pojmanska (1973) and is now regarded as *Incertae sedis* [21]. The morphological characteristics of the genus *Urotoxus* among all the 3 genera include a long body and a sucker that is relatively poorly developed. On the other hand, the general shape of the other 2 genera are close to ellipsoidal and have well-developed suckers. The species belonging to the genera *Leucochloridium* and *Urogonimus* are very similar, and hence, difficult to distinguish from one another, but there are some morphological differences in the location of their genital pores, presence of a spine on the surface of their bodies and distribution of the vitelline glands. In the species belonging to the genus *Urogonimus*, the genital pores are often at the posterior end of the body and vitelline glands extend from the ceca to the region of the oral suckers and that of the gonads, but not reaching cecal extremities, and have no spines on their body surfaces. In the species belonging to the genus *Leucochloridium*, the genital pore is distinctly at the dorsal region and the vitelline glands extending from the posterior to the cecal extremities or may stretch even further posteriorly and have fine spines on their body surfaces [21].

In the genus *Urogonimus*, the vitellaria are relatively short and extend only to the level of the right testis. In a recent study, among the species of this genus, unique morphological features were observed in *U. turdi*; the genital glands were arranged in a triangle at the posterior region of the body and the acetabulum was slightly post-equatorial and this was a unique phenomenon in *U. turdi* when compared to the other species of *Urogonimus* [10]. In the present study, the vitellaria of the fluke taken from White’s thrush extend from the level of the oral sucker to that of the right testis and the position of the genital glands and the acetabulum is very similar to that of *U. turdi* reported in the previous study [10]. Moreover, our specimens are very similar to those of *L. turdi* obtained from the cloaca of *L. cardis* from Kutama near Kyoto, Japan in 1937, as described by Yamaguti [8]. It has been noticed that the size of the acetabulum was 0.34-0.68 mm in diameter which was considerably larger than the oral suckers (0.22-0.48 mm in di-
ameter). In the present study, the acetabulum is approximately 1.5 times larger than the oral sucker.

In 1979, Iskova [22] reported an infection by U. turdi in Ukrainian passerine birds and Ching [3] also reported an infection of L. turdi in varied thrushes found in British Columbia. Iwaki et al. [9] suspected that the acetabulum of U. turdi that infected Ukrainian passerine birds was U. macrostomus because it was almost in the equator. The size of that infected Ukrainian passerine birds was 1.5 times larger than the oral sucker. In the present study, the acetabulum is approximately 2017RIDIAIB06031728).

Additionally, the nuclear ribosomal DNA, the 18S rRNA gene, was used to study the inter- and intraspecific relationships, since it is highly repeated and contains variable regions flanked by more conserved regions [23,24]. Although the genetic divergence of U. turdi suggests its conspecific relationship with other Leucochloridium sp., the 18S rRNA sequences proved to be useful in distinguishing U. turdi from other parasites such as L. holostomum (KP903629), L. paradoxum (KP903632), and L. perturbatum (KP903641). In conclusion, we can determine that our specimen has been validated as U. turdi from Z. auroa in Korea.

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CONFLICT OF INTEREST

The authors declare no conflict of interest related to this study.

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