Regulatory role of leptin in glucose and lipid metabolism in skeletal muscle

Yasuhiko Minokoshi, Chitoku Toda, Shiki Okamoto
Division of Endocrinology and Metabolism, National Institute for Physiological Sciences, Myodaiji, Okazaki, Aichi - 444-8787, Japan

ABSTRACT

Leptin is a hormone secreted by adipocytes that plays a pivotal role in regulation of food intake, energy expenditure, and neuroendocrine function. Several lines of evidences indicate that independent of the anorexic effect, leptin regulates glucose and lipid metabolism in peripheral tissues in rodents and humans. It has been shown that leptin improves the diabetes phenotype in lipodystrophic patients and rodents. Moreover, leptin suppresses the development of severe, progressive impairment of glucose metabolism in insulin-deficient diabetes in rodents. We found that leptin increases glucose uptake and fatty acid oxidation in skeletal muscle in rats and mice in vivo. Leptin increases glucose uptake in skeletal muscle via the hypothalamic–sympathetic nervous system axis and β-adrenergic mechanism, while leptin stimulates fatty acid oxidation in muscle via AMP-activated protein kinase (AMPK). Leptin-induced fatty acid oxidation results in the decrease of lipid accumulation in muscle, which can lead to functional impairments called as “lipotoxicity.” Activation of AMPK occurs by direct action of leptin on muscle and through the medial hypothalamus–sympathetic nervous system and α-adrenergic mechanism. Thus, leptin plays an important role in the regulation of glucose and fatty acid metabolism in skeletal muscle.

Key words: AMP-activated protein kinase, hypothalamus, leptin, skeletal muscle, sympathetic nervous system

INTRODUCTION

Leptin is secreted by adipocytes and signals nutritional information to regulatory centers in the hypothalamus and other brain regions.[1] Treatment with the hormone in animals suppresses food intake and increases energy expenditure, accompanied by reduction in the amount of adipose tissue and intracellular lipid in skeletal muscle, liver, and pancreatic β-cells.[1-4] Decrease in the amount of lipid in these tissues is evident at doses that do not affect weight, suggesting that the effect of leptin is not only a consequence of its ability to reduce food intake.[4]

Skeletal muscle is a principal site of glucose and fatty acid utilization, and is one of the primary tissues responsible for insulin resistance in obesity and type 2 diabetes.[6] Increased lipid stores in nonadipose tissues, such as muscle are linked to functional impairments, called “lipotoxicity,” which lead to insulin resistance and impaired insulin secretion.[6] Although the lipid factor that causes “lipotoxicity” is not identified, involvement of fatty acyl-CoA or diacylglycerol, acting through a form of protein kinase C, is suspected.[7] As well as in obesity, insulin resistance appears in lipodystrophy, in which fat tissue is absent.[8,9] In this rare human disorder, excess lipid accumulates in tissues such as the liver and skeletal muscle, and patients suffer from severe insulin-resistant diabetes. Genetically engineered experimental animals show the same pathophysiology.[8,9] It has been demonstrated that treatment with leptin decreases lipid accumulation in nonadipose tissues and improves severe diabetes in lipodystrophy patients as well as rodents.[5,8-10]

Recently, studies indicate that leptin suppresses the development of the impaired glucose metabolism in rodents treated with streptozotocin (STZ) and Akita mice with insulinopenia.[12-20] Hyperleptinemia induced by either pharmacologic leptin administration or with adenoviral infection, fully ameliorates hyperglycemia in those insulin-resistant mice.
deficient animals, even in the extremely low plasma insulin levels. While this effect is partly due to the normalization of hyperphagia and circulating levels of glucagon and corticosterone, those cannot fully explain the improvement of STZ-induced diabetes in response to hyperleptinemia. Other regulatory mechanisms are necessary to supply energy and nutrients into skeletal muscle to improve the severe diabetes phenotype in the absence of insulin.

In this review, we focus on the regulatory roles of leptin in glucose and lipid metabolism in skeletal muscle. Our results suggest that leptin stimulates glucose uptake in skeletal muscle via the hypothalamic–sympathetic nervous system and \( \beta \)-adrenergic mechanism.\(^{[21-23]} \) Furthermore, we demonstrate that leptin stimulates fatty acid oxidation in skeletal muscle via AMPK. Activation of AMPK occurs by direct action of leptin on muscle and through the medial hypothalamus–sympathetic nervous system and \( \alpha \)-adrenergic mechanism.\(^{[24]} \) Leptin-induced fatty acid oxidation results in the decrease of lipid accumulation in muscle, and suppresses functional impairments called as “lipotoxicity,” such as insulin resistance. Thus, leptin plays an important role in the regulation of glucose and fatty acid metabolism in skeletal muscle.

**LEPTIN STIMULATES GLUCOSE UPTAKE IN SKELETAL MUSCLE VIA THE HYPOTHALAMIC–SYMPATHETIC NERVOUS SYSTEM**

It is now evident that many of the effects of leptin on metabolism, as well as food intake, are exerted in the hypothalamus.\(^{[1,4]} \) We and others have shown that injection of leptin into the medial hypothalamus, including ventromedial hypothalamic nucleus (VMN) increases glucose uptake by skeletal muscle, brown adipose tissue (BAT), and heart but not in white adipose tissue (WAT) [Figure 1].\(^{[21-23,25]} \) The effect of leptin can be observed acutely (6 h after the injection) and is independent of feeding behavior. In skeletal muscle, the effect of leptin was observed in red type of muscle, such as soleus muscle, rather than white type of muscle. Leptin has little effect on glucose uptake in skeletal muscle ex vivo. Although intracerebroventricular (icv) or intravenous (iv) injection of leptin normalizes blood glucose levels in insulin-deficient diabetic animals,\(^{[12-20]} \) the leptin-induced glucose uptake is enhanced with insulin administration.\(^{[22]} \) Thus, leptin stimulates glucose uptake in muscle in both insulin-dependent and -independent manner.

Intravenous and icv injection of leptin stimulates sympathetic nerve activity in peripheral tissues.\(^{[20,27]} \) A sympathetic nerve-blocking agent (guanethidine), a \( \beta \)-adrenergic antagonist (propranolol), and surgical sympathetic denervation blunt the leptin's action,\(^{[21,22]} \) whereas the effect of leptin injection into the VMN on muscle glucose uptake is not blocked by adrenal demedulation. These results suggest that leptin increases glucose uptake in skeletal muscle via sympathetic nerve and \( \beta \)-adrenergic mechanism. In support of this, we recently reported that injection of a hypothalamic neuropeptide orexin into the VMN stimulates glucose uptake in muscle in mice, similar to that of leptin, via sympathetic nerve and \( \beta \)-adrenergic receptor.\(^{[20]} \) The orexin-induced muscle glucose uptake was blunted in \( \beta \)-adrenergic receptors-deficient mice (\( \beta \)-less mice), while the expression of \( \beta2 \)-adrenergic receptor in muscle in \( \beta \)-less mice recovered the orexin action in the tissue. Studies with catecholamine administration have suggested that the sympathetic nervous system inhibits the insulin signaling pathway in peripheral tissues. However, our results show that the preferential stimulation of sympathetic nerves and subsequent signaling of \( \beta2 \)-adrenergic receptor, which is a dominant type of \( \beta \)-adrenergic receptor, result in activation of the insulin signaling pathway in skeletal muscle *in vivo*. The effects of sympathetic nerves on muscle glucose metabolism may differ in some instances from those of catecholamine administration. Consistent with this notion, a \( \beta2 \)-adrenergic receptor-specific agonist was
previously shown to increase glucose uptake in L6 myocytes via activation of PI3-kinase, with this effect being inhibited by protein kinase A.[29] Thus, leptin increases glucose uptake and insulin sensitivity in skeletal muscle through the hypothalamic–sympathetic nervous system axis.

Leptin receptor Ob–Rb abundantly expresses in the hypothalamus, especially in the arcuate hypothalamic nucleus (ARC), VMN, and dorsomedial nucleus (DMN), with a lesser amount in the paraventricular nucleus (PVN) and lateral hypothalamic area (LHA).[1] The ARC contains two of the most well-characterized leptin-sensitive neuronal populations: one is the neuropeptide Y (NPY) and agouti-related peptide (AgRP)-coexpressing neurons, and another is the pro-opiomelanocortin (POMC)-expressing neurons. NPY and AgRP-coexpressing neurons release γ-amino butyric acid (GABA) and inhibit neuronal activity of POMC neurons in the ARC. NPY and AgRP stimulate food intake and inhibit energy expenditure. A product of POMC, α-melanocyte-stimulating hormone (α-MSH), is released in POMC neurons in the ARC and acts on melanocortin receptor (MCR) to suppress food intake and increase energy expenditure. Leptin activates POMC neurons and inhibits the activity of NPY/AgRP neurons in the ARC. It has been reported that leptin signaling in POMC neurons is important in the regulation of glucose metabolism. Expression of leptin receptors in POMC neurons in db/db mice, which lack functional leptin receptors, ameliorates hyperglycemia, hyperglucagonemia and dyslipidemia, whereas it modestly reduces body weight and hyperinsulinemia.[30]

The VMN neurons also play an important role in the leptin’s action on glucose metabolism. As mentioned, preferential injection of leptin into the VMN enhances glucose uptake in skeletal muscle, heart, and BAT, but not in WAT.[21-23] Leptin rapidly increases the firing rate of steroidogenic factor-1 (SF-1)-containing neurons in the nucleus, whereas selective deletion of leptin receptors of SF-1 neurons results in an obese, insulin-resistance phenotype.[31,32] Both SF-1 in the VMN and POMC in the ARC are necessary for the normal glucose metabolism. Severe insulin resistance occurs in mice that have neither leptin receptor in SF-1 nor POMC neurons.[8] Previous studies revealed that MCR agonist MT-II preferentially increases the expression of brain-derived neurotrophic factor in the VMN, whereas a subset of VMN neurons activates POMC neurons in the ARC.[33,34]

We recently explored the roles of the medial hypothalamic nuclei in leptin-induced glucose uptake in peripheral tissues.[23] Leptin injection into the VMN increased glucose uptake in skeletal muscle, BAT, and heart, whereas that into the ARC increased glucose uptake in BAT but not muscle or heart, and that into the DMN or PVN had no effect [Figure 2]. The icv MCR antagonist SHU9119 abolished the effects of leptin injected into the VMN on glucose uptake in muscle and other peripheral tissues.[23] Furthermore, injection of MCR agonist MT-II either into the VMN or intracerebroventricularly increased glucose uptake in muscle, BAT, and heart, whereas that into the PVN increased glucose uptake in BAT, and that into the DMN or ARC had no effect. Thus, the effect of leptin on muscle glucose uptake is dependent on MCR activation in the VMN, whereas the leptin receptor in the ARC and MCR in the PVN regulate glucose uptake in BAT.

To identify the direct neuronal circuits whereby MCR regulates peripheral tissues, retrograde tracing studies with pseudorabies virus (PRV) were conducted in combination with in situ hybridization of melanocortin 4 receptor (MC4R). Injection of PRV into interscapular BAT or WAT led to PRV co-labeling in MC4R-positive cells within hypothalamic nuclei, including the PVN, LHA, DMN, ARC, and VMN.[35,36] A recent study also used Ob-Rb-GFP reporter mice with muscle-specific injection of RFP-expressing PRV, and reported that PRV labeling is observed within these hypothalamic nuclei.[37] However, the study also indicated that significant number of double-labeled cells with Ob-Rb and PRV is found only in the brainstem nucleus of the solitary tract (NTS) and the hypothalamic retrochiasmatic area (Reh). Further studies are required to explore the role of NTS and Reh in leptin-induced muscle glucose uptake.

Figure 2: Distinct roles of the medial hypothalamic nuclei in leptin-induced glucose uptake in peripheral tissues.[23] Leptin activates VMN neurons as well as POMC neurons in the ARC, and then activates MCR in the VMN and PVN. Activation of MCR in the VMN stimulates glucose uptake in skeletal muscle, BAT, and heart, whereas that in the PVN increases glucose uptake in BAT.
AMPK is a regulator of cellular metabolism in response to changes in the energy status of the cells. It has been demonstrated that activation of AMPK participates in the contraction-induced glucose uptake and fatty acid oxidation in muscle at least in part. We examined AMPK activity in skeletal muscle after the medial hypothalamus and intravenous injection of leptin in mice. Injection of leptin into the medial hypothalamus increased the activity of α2 AMPK in red type of skeletal muscle, such as soleus muscle. The effect peaked with a threefold stimulation at 1 h and was sustained for up to 6 h [Figure 3]. In contrast, iv leptin produced a biphasic response in α2 AMPK activity, with a twofold rise at about 15 min, a return to baseline by 60 min, and a second twofold elevation by 6 h. α1 AMPK activity did not change in soleus muscle after iv or ihp injection of leptin.

A lower dose of leptin, which is closer to the physiologic response in elevation by 615 min, a return to baseline by 60 min, and a second twofold stimulation of AMPK in red muscles, more pronounced in red (slow twitch, oxidative) skeletal muscle, such as soleus, compared to white (fast twitch, glycolytic) muscles. These results indicate that intravenous and medial hypothalamic injection of leptin activate α2 AMPK in red muscle through Ob-Rb.

We explored whether leptin-induced activation of AMPK in muscle is mediated by the sympathetic nervous system, using pharmacologic adrenergic blockade and two kinds of surgical denervation of hindlimb. Denervation of the sciatic nerve, which primarily impairs motor innervation, decreased the total catecholamine content to 62% of the contralateral, intact soleus muscle. Denervation of the sciatic, femoral, and obturator nerves, which block both sympathetic and motor innervation, decreased catecholamine content to 6% of the control content.

Denervation of all three nerves blocked the ability of hypothalamic injection of leptin or iv leptin 6 h after injection to stimulate α2 AMPK in soleus muscle. But AMPK activation at 15 min after iv leptin remained intact in denervated muscle. By contrast, denervation of the sciatic nerve alone did not block leptin-induced activation of AMPK in soleus, despite a complete loss of motor function. Thus leptin's effect on AMPK is independent of motor nerve activity. These findings indicate that AMPK activation at 6 h after both iv leptin and hypothalamic injection of leptin may be mediated by the sympathetic nerves that innervate muscle. In support of the mechanism, peripheral leptin injection increases sympathetic nerve activity that innervates hindlimbs, with a time course that is consistent with the late activation of α2 AMPK in muscle after iv leptin. In contrast, activation at 15 min after iv leptin does not involve the sympathetic nervous system and may be direct. Consistent with this, leptin increased α2 AMPK activity in soleus muscle ex vivo. Thus, the early effect of iv leptin may be direct, but the latter effect depends on the sympathetic nerve innervating the muscle [Figures 3 and 4].

Interestingly, our results revealed that the α-adrenergic receptor, but not β-adrenergic receptor, is involved in the sympathetic nerve-mediated effect of leptin on AMPK. AMPK can be activated by the stimulation of Gq-coupled receptors, including α-adrenergic receptors. Gq-coupled receptors stimulate intracellular calcium signaling and activate an AMPK kinase, calcium-/calmodulin-dependent protein kinase kinase (CaMKK). The involvement of CaMKK in the activation of muscle AMPK is suggested by the recent study showing that adiponectin activates AMPK in skeletal muscle via CaMKK by increasing cytosolic Ca^{2+}-level. Thus, AMPK activation is mediated by the α-adrenergic mechanism, whereas glucose uptake is mediated by β-adrenergic receptor. Similar to that of leptin-induced glucose uptake, MCR in the brain also involves the hypothalamic effect of leptin on muscle α2 AMPK. The icv MT-II injection activated muscle AMPK, whereas MCR antagonist blunted the leptin’s effect.
Acetyl-coenzyme A (acetyl-CoA) carboxylase (ACC), which is a target of AMPK, provides a pivotal step in fuel metabolism, as it links fatty acid and glucose metabolism through the shared intermediate acetyl-CoA [Figure 4]. Phosphorylation of ACC by AMPK leads to the inhibition of ACC activity, a fall in malonyl-CoA content, and a subsequent increase in fatty acid oxidation by disinhibiting carnitine palmitoyltransferase 1 (CPT1) in skeletal muscle. To determine whether AMPK activation mediates leptin’s effects on fatty acid metabolism, we examined the activity of ACC. Intravenous and hypothalamic injection of leptin suppressed ACC activity in soleus muscle. These effects are probably caused by phosphorylation of ACC by AMPK, as ACC phosphorylation in soleus muscle increased after iv leptin, whereas concentrations of ACC protein did not change. ACC phosphorylation 6 h after iv leptin was blocked by surgical denervation of the sciatic, femoral, and obturator nerves. In db/db mice, leptin did not stimulate phosphorylation of AMPK or ACC. Moreover, leptin-induced ACC phosphorylation in cultured muscle cells expressing Ob-Rb, whereas the effect is suppressed with the expression of dominant-negative AMPK.

The decrease of ACC activity by leptin suggests that malonyl-CoA production is suppressed, leading to stimulation of fatty acid oxidation by disinhibition of carnitine palmitoyltransferase (CPT1). To elucidate this, we measured fatty acid utilization and oxidation in muscle after iv injection of leptin in vivo, by assessing the incorporation of [3H]-(R)-2-bromopalmitic acid and [14C]palmitic acid into fatty acid oxidation in muscle [24].

Recently, we and others revealed that leptin regulates food intake via changing hypothalamic AMPK activity. In contrast to the effect on skeletal muscle, leptin inhibits AMPK in the ARC and PVN. Constitutively active and dominant-negative AMPK are sufficient to change food intake, respectively. The decrease of AMPK activity in the hypothalamus is essential for the anorexic effect of leptin. Other anorexic factors, such as insulin and glucose, also suppress AMPK activity in the hypothalamus, whereas orexigenic hormone ghrelin activates the AMPK. Although it is still unclear whether hypothalamic AMPK affects peripheral metabolism, it has been shown that direct modulation of fatty acid metabolism in the hypothalamus, which is a downstream of AMPK, regulates fatty acid oxidation and mitochondrial function in muscle.

**CONCLUDING REMARKS**

Leptin administration significantly improves impaired glucose metabolism in lipodystrophy and insulin-deficient diabetes mellitus. However, the mechanism still remains elusive. Our findings indicate that leptin stimulates muscle glucose uptake via the hypothalamic–sympathetic nervous system and β-adrenergic mechanism. Both VMN neurons and POMC neurons involve the leptin’s action. Furthermore, leptin stimulates fatty acid oxidation in muscle via AMPK. The activation of muscle AMPK is mediated by two distinct mechanisms: one is a direct effect of leptin and another is mediated by the hypothalamic–sympathetic nervous system and a-adrenergic mechanism. Activation of AMPK by leptin phosphorylates and inhibits ACC, and results in potent oxidation and mitochondrial function in muscle.
stimulation of fatty acid oxidation in muscle. Although leptin likely has many functional roles in central and peripheral tissues, a physiologic role of leptin in muscle may play an important role in the regulation of glucose and fatty acid metabolism.

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