Anticonvulsant effects of iridoid glycosides fraction purified from *Feretia apodanthera* Del. (Rubiaceae) in experimental mice models of generalized tonic-clonic seizures

Germain Sotoing Taiwe¹*, Bernard Dabole², Thierry Bang Tchoya³, Joseph Renaud Menanga³, Paul Désiré Djomeni Dzeufiet³ and Michel De Waard⁴,⁵,⁶

**Abstract**

**Background:** Despite the increasing number and variety of antiepileptic drugs, nearly 30% of epileptic patients who receive appropriate medical attention have persisting seizures. Anticonvulsant activity has been demonstrated for different iridoid glycoside-rich plant extracts. This study was designed to investigate the anticonvulsant effects of iridoid glycosides purified from *Feretia apodanthera* and to explore the possible mechanisms involved in antiepileptic activity.

**Methods:** The anticonvulsant effects of iridoid glycosides extracts were investigated against 2.7 mg/kg bicuculline- and 70 mg/kg pentylenetetrazole-induced convulsions. The behavioural and electroencephalographic manifestations of 50 mg/kg pentylenetetrazole-induced seizures in mice as a model of generalized tonic-clonic seizures were also evaluated. Finally, the extracts were tested on the course of kindling development, kindled-seizures and oxidative stress markers in 30 mg/kg pentylenetetrazole-kindled mice. Their effects on brain GABA content were also determined.

**Results:** The iridoid glycosides (30–90 mg/kg) protected mice against bicuculline-induced motor seizures in all pre-treated animals. Behavioural seizures- and mortality-induced by 70 mg/kg pentylenetetrazole were strongly antagonized by the extracts (60–90 mg/kg). The number of crisis (n/20 min), the cumulative duration of crisis (sec/20 min), and the mean duration of crisis (sec) recorded in 50 mg/kg pentylenetetrazole-treated mice were significantly decreased in all pre-treated mice with the extracts (60–90 mg/kg). Administration of the extracts (30–90 mg/kg) significantly increased the latency to myoclonic jerks, clonic seizures as well as generalized tonic-clonic seizures, improved the seizure mean stage and decreased the number of myoclonic jerks in 30 mg/kg pentylenetetrazole-kindled mice. Pentylenetetrazole kindling induced significant oxidative stress and brain GABA content alteration that was reversed by pretreatment with the extracts in a dose-dependent manner.

**Conclusions:** The results indicate that pretreatment with the iridoid glycosides extracts of *Feretia apodanthera* improves generalized tonic-clonic seizures-induced by chemo-convulsants, protects mice against kindling development and oxidative stress, and improves brain GABA content in pentylenetetrazole-kindled mice.

**Keywords:** Iridoid glycosides, Tonic-clonic seizures, Kindling development, Oxidative stress, Antiepileptic drugs

* Correspondence: taiwe_sotoing@yahoo.fr

¹Department of Zoology and Animal Physiology, Faculty of Science, University of Buea, Buea, Cameroon

Full list of author information is available at the end of the article

© 2016 The Author(s). Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated.
Background

Epilepsy is a prevalent neurological disorder that seriously affects the living quality of more than 65 million people worldwide [1, 2]. It is well acknowledged that epileptic seizures are the result of instant abnormal hyper-synchronous electrical activity of neuronal networks originating locally from discharges of brain regions. Imbalance between excitation and inhibition appears as the main cause of these abnormal electrical activities [3–5]. Individual generalized seizure types include absence, myoclonic, tonic-clonic, atonic, tonic, and clonic symptoms. In many studies, it is not clear that “generalized seizure” is synonymous with generalized onset seizure, leading to some ambiguity in classification by seizure type [6]. Despite the increasing number and variety of antiepileptic drugs, nearly 30 % of epileptic patients who have been taken in charge medically have persistent seizures [7]. The pharmacological resistance of epilepsy remains therefore too widespread. Also, current treatments of epilepsy are not satisfactory in terms of drug-associated deleterious effects [8]. Further, adverse effects associated with antiepileptic drugs and recurrent seizures limit their use. The development of new, affordable and accessible pharmacological agents that can overcome these limitations has become a major goal in epilepsy research [9]. The plant kingdom has become a target of great interest and value in the search of new drugs and lead compounds to treat several neurological disorders, including epilepsy for many years [7, 10].

Feretia apodanthera Del. is a member of the Rubiaceae family. This plant is found mainly in the savanna regions of the West African coast. Preparations of the plant have been used in Cameroonian folk medicine to manage epilepsy, infantile convulsions, anxiety, psychoses, pain and inflammation for many years, and their efficacies are widely acclaimed among the rural communities of Northern Cameroon [11–15]. Previous studies in our laboratory found that intra-gastric administration of an aqueous extract of Feretia apodanthera significantly reduced the progression of kindling-induced by pentylentetrazol, and attenuated the oxidative stress and cognitive impairment in kindled-mice [10]. The aqueous extracts of Feretia apodanthera contained flavonoids, alkaloids, saponins, tannins, glycosides, anthraquinones and phenols, but not lipids [10]. Many of these compounds are used widely as food additives or in traditional medicine, prompting phytochemical investigations that have in turn uncovered a variety of flavonoids, alkaloids and iridoid glycosides [16–18]. Recent study showed that the treatment with cornel iridoid glycosides isolated from Cornus officinalis Sieb. et Zucc improved neurobehavioral deficits, decreased cerebral infarct size, reduced nitric oxide and inhibited nuclear factor kappa B expression in the brain of rats 24 h after focal cerebral ischemia [19, 20]. These results indicate that cornel iridoid glycosides may improve the microenvironment of the central nervous system by increasing growth/trophic factors and decreasing inflammation-related factors. Baillieu et al. [18] showed that the iridoid glycosides-rich extracts from Feretia apodanthera decreases the spontaneous motor activity and exploratory behaviour in mice, increases hexobarbital-mediated sleeping time in rats, attenuates the intensity of amphetamine-induced stereotypies in mice, and inhibited catalepsy induced by chlorpromazine rats. It also protected rat and rabbit, respectively, against maximal electroshock- and pentylentetrazol-induced seizures [18].

In this manuscript, in order to understand the preliminary anticonvulsant mechanisms of action during epilepsy and/or epileptogenesis, we purified five iridoid glycosides from the stem barks of Feretia apodanthera using bio-guided fractionation, and we screened the effects of the mixture of these five iridoid glycosides on the central nervous system using experimental rodent models of epilepsy. The aim of this study was to assess the efficacy of iridoid glycoside extracts of Feretia apodanthera against convulsions induced by two chemoconvulsants, bicuculline and pentylentetrazole, in mice as a model of generalized tonic-clonic seizures [21, 22]. We also evaluated the effects of this extract on the course of kindling development, kindling-induced oxidative stress markers and brain GABA content in pentylentetrazol-kindled mice.

Methods

Plant material and purification of iridoid glycosides

The stem barks of Feretia apodanthera used in this study were harvested in the Mount Tenglin area of Pitoa (North Region of Cameroon, harvesting coordinates 9°25′17″ N and 13°27′2″ E). The plant collection was carried out on a private land, following the permission by the owner (Mr. Waziri Tshaolao, resident of Loumo quarter, Pitoa), to conduct the study on this site. The field studies did not involve endangered or protected species. The species was authenticated by the National Herbarium of Yaoundé (Cameroon), where a voucher was deposited (Sample Number 31225/HNC). Briefly, 500 g stem bark powder of Feretia apodanthera was decocted in 100 °C with 3 L water during 20 min and the solution was left to cool down. The procedure was repeated 3 times. The aqueous extract (2410 mL) was filtered, concentrated and subjected to XDA-16 macroporous adsorptive resins chromatographic column (250 g, 8 cm × 50 cm), which had been depurated with 95 % alcohol, with a flow rate of 2.0 mL/min. 600 mL distilled water was added to elute the iridoid glycoside extract. The end of the elution procedure was witnessed by a negative Molish reaction. Column was cleaned afterwards with 800 mL 50 % EtOH. Total iridoid glycoside extracts (27.9 g) was obtained after evaporation at 60 °C using a
rotavapor. After, the extract was dissolved in distilled water (150 mL) and chromatographed on a silica gel column prewashed with 20 % EtOH (300 mL). The iridoid glycosides of Feretia apodanthera (IGEFA; 14.9 g) were obtained by using CHCl₃-MeOH-H₂O (43/37/20, vol/vol/vol) as eluant. The purities of iridoid glycosides were determined by RP-HPLC assay. The analyses were carried out on a HPLC system (Shimadzu-LC 20AT) equipped with UV detector, pressure controlled by prominence pump and operated by spinchrom Software. The HPLC separation method was developed on an Agilent reversed-phase octyldecyl silica column (TC-C18, 4.6 mm × 150 mm, 5 μm) and for elution of the constituents, a gradient of two solvents denoted A and B was employed. The mobile phase consisted of solvent A (water) and solvent B (acetonitrile) with the following gradient program: 30 % B in 0 ~ 5 min; 30 % ~ 35 % B in 5 ~ 15 min; 35 % ~ 60 % B in 15 ~ 20 min; 60 % B in 20 ~ 30 min. The flow rate was set at 1.0 mL/min, column temperature was kept at 30 °C and the injection volume was 20 μL. The sample was dissolved in a mixture of acetonitrile and water (30:70, v/v). Iridoid glycosides were detected at a wavelength of 240 nm (Fig. 1). The structures have been successfully elucidated by using homo- and heteronuclear two-dimensional nuclear magnetic resonance (NMR) techniques such as 1H-NMR, 13C-NMR, heteronuclear multiple quantum coherence (HMQC) and heteronuclear multiple bond correlation (HMBC) spectra, and mass spectrum [16–18]. The purity of IGEFA was 78.5 %, in which feretoside (1) accounted for 29 %, gardenoside (2) 18 %, geniposidic acid (3) 14 %, apodanthoside (4) 23 % and desacetylasperolosidic acid (5) 16 %, respectively (Fig. 2). The IGEFA was dissolved in saline 0.9 % containing dimethyl sulfoxide 2 % (vehicle) at the appropriate concentrations as indicated in the various experiments and administered per os (p.o.) with a volume of 10 mL/kg.

Chemicals
Bicuculline, clonazepam, diazepam, flumazenil, methyl-β-carboline-3-carboxylate (FG7142), pentylenetetrazole, sodium valproate, reduced glutathione, thiobarbituric acid, n-butanol, pyridine, sodium dodecyl sulphate, 5’5-dithiobis (2-nitrobenzoic acid) and trichloroacetic acid were obtained from Sigma, St. Louis, MO, USA. All other chemicals and reagents used in the brain γ-aminobutyric acid (GABA) content estimation were obtained from Sigma, St. Louis, MO, USA. Diazepam was obtained from Roche (France).

Animals
Experiments were conducted on Swiss male mice (20–25 g) housed in appropriate cages with food and water ad libitum. They were kept in 12 h light/dark cycle. Each animal was used only once and was handled according to standard protocols for the use of laboratory animals. The investigation conforms to the Guide for the Care and Use of Laboratory Animal published by the US National Institutes of Health (NIH; publication No. 85-23, revised 1996) and received approval of the Cameroon National Ethical Committee (Yaounde, Cameroon) for animal handling and experimental procedure (Ref N°FW-IRB00001954). All efforts were made to minimize animal suffering and reduce the number of animals used.

Pharmacological tests
Bicuculline-induced motor seizures
Mice were divided into six groups of eight mice and received orally different doses of IGEFA (15, 30, 60 and 90 mg/kg; per os (p.o.)), diazepam (positive control; 5 mg/kg, intraperitoneal (i.p.) or vehicle (10 mL/kg; p.o.). One hour later, all animals were injected intraperitoneally with bicuculline (2.7 mg/kg; subcutaneously (s.c.)) and placed in isolated cages. Motor seizures, induced by bicuculline in mice, were visually observed by two independent investigators, unaware of the identity of the experimental groups. They were quantified in experimental and matched control mice using the following parameters: (a) the time to onset of the first seizure (either tonic or clonic); (b) the duration of the clonic seizure.
and tonic components of seizures; (c) the number of motor seizures and (d) mortality. Clonic seizures consisted of rhythmic contractions of forelimbs and/or hindlimbs and/or the back muscles [23, 24]. A tonic seizure consisted of a rigid extension of the fore and/or hind limbs with or without loss of posture. The time of observation was 120 min [24, 25].

Pentylenetetrazole-induced seizures

**Experiment 1: acute administration of pentylenetetrazole at a dose of 70 mg/kg**

Mice were divided in six groups of eight mice and received different treatments. Group I (negative control) was treated with vehicle (10 mL/kg p.o.). Groups II to V (test groups) were treated with 4 doses of IGEFA (15, 30, 60 and 90 mg/kg; p.o.). Group VI treated with clonazepam, 0.1 mg/kg i.p., was used as positive control. Tonic-clonic seizures were induced in mice by i.p. injection of 70 mg/kg pentylenetetrazole. The protective effect of the different treatments given 1 h before pentylenetetrazole injection was recorded. Animals that did not convulse within the 10 min of observation were qualified as protected [9].

**Involvement of the GABA<sub>A</sub> receptor complex in pentylenetetrazole (70 mg/kg) test**

We also studied the effects of a selective benzodiazepine receptor antagonist, flumazenil (1, 2, 3, 4, 5 and 6 mg/kg) and FG7142 (4, 6, 8, 10, 12 and 14 mg/kg), an inverse diazepam receptor agonist of the GABA<sub>A</sub> receptor complex, on the anticonvulsant activity of IGEFA in order to investigate the probable involvement of benzodiazepine receptors [26]. In these antagonistic experiments, flumazenil and FG7142 were injected 15 min prior to the IGEFA (90 mg/kg) treatments.

**Experiment 2: acute administration of pentylenetetrazole at a dose of 50 mg/kg**

**Surgery**

Animal surgery was done as previously described by Riban et al. [27]. Under general anesthesia (chloral hydrate 4 % in NaCl 0.9 %, 10 mL/kg i.p.) six groups of six mice were implanted stereotaxically with (i) two monopolar surface electrodes placed over the left and right frontoparietal cortex for pharmacological experiments or a bipolar electrode within the right frontal sensorimotor cortex (AP = +1.4 mm, ML = −1.6 mm, DV = −2 mm from Bregma) for signal analysis; (ii) a monopolar electrode placed over the cerebellum (reference electrode), and (iii) a bipolar electrode inserted into the hippocampus; with bregma as the reference [28]. The electrodes were made of stainless steel wire isolated by polyester (diameter, 0.125 mm). They were inserted in the skull above the cortex and the cerebellum. The bipolar electrode was formed of two twisted polyester insulated stainless steel wires. The implant assembly was affixed to the skull with dental acrylic. After the surgical implantation, mice were injected with antibiotics and

---

**Fig. 2** Chemical structures of five iridoid glycosides purified from the stem barks of Feretia apodantha. (1) feretoside; (2) gardenoside; (3) geniposidic acid; (4) apodanthoside; (5) desacetylasperidosidic acid.
housed individually in cages at least 14 days for recovery before the onset of experiments [27].

**Electroencephalographic recordings**

Electroencephalograms (EEGs) were recorded in awake freely moving animals using a digital acquisition system (Biopac System, MP-100, Inc.). The signals were amplified and filtered (high pass filter 1 Hz/low pass filter 97 Hz), then digitized at a sampling rate of 256 Hz and recorded using AcqKnowledge® software version 3.2 (Biopac Students Lab PRO software). During the recording and stimulation sessions, the mice were continuously watched to detect changes in their posture and behaviour. All sessions did not exceed 3 h and were performed between 9:00 a.m. and 5:00 p.m [27]. Digital video recordings were made with a webcam (Orbit Logitech Quickcams) located inside the Faraday cage, and the animals’ behaviour was simultaneously recorded (for detailed information on surgery and procedures).

**Treatments and seizures induction**

All pharmacological compounds were tested between the third and the sixth week following electrodes implantation. To test the effects of IGEFA on pentylenetetrazole-induced seizures, the animals were first divided into six groups of six mice each, and received different doses of IGEFA (15, 30, 60 and 90 mg/kg; p.o.), clonazepam (0.1 mg/kg, i.p.) or vehicle (10 mL/kg p.o.). They were then recorded for 60 min before the pentylenetetrazole injection (50 mg/kg, i.p.) and then for 120 min. The latency of first crisis, the cumulative duration of crisis, the mean duration of crisis and number of crisis were quantified.

**Histology**

Upon completion of the experiments, all mice were injected with a lethal dose of pentobarbital (100 mg/kg, i.p.). Their brains were removed, frozen and cut in 20 μm sections using a cryostat. Histological analysis was performed following Cresyl Violet staining and each implantation site was localized with reference to the atlas of Paxinos and Watson [29].

**Experiment 3: chronic administration of pentylenetetrazole at a dose of 30 mg/kg**

**Experimental design**

Animals were randomly divided into eight groups of six animals each. The first group received saline intraperitoneally while the second–seventh groups were administered pentylenetetrazole (30 mg/kg; i.p.) dissolved in saline on every second day (48 ± 2 h) [30]. One hour before administration of pentylenetetrazole, the first and second groups received vehicle (10 mL/kg), the third–sixth groups were administered IGEFA (15, 30, 60 and 90 mg/kg; p.o., respectively) orally through an intragastric feeding tube. Group seven animals were administered sodium valproate (300 mg/kg) intraperitoneally. Pentylenetetrazole and IGEFA were administered up to day 43 or until seizure stage 5 on two consecutive trials was achieved, whichever was earlier. Mice were observed for 30 min after the subconvulsant PTZ and seizure activity scored using a scoring system from 0 to 5. In group eighth and ninth, IGEFA (90 mg/kg) or sodium valproate, respectively, were administered alone to study any per se effects of the IGEFA or sodium valproate on behaviour and biochemical parameters. Group ten animals were administered vehicle and one hour later they received saline. Behavioural test were performed 24 h after the last administration of pentylenetetrazole. Following the behavioural evaluation, the animals were sacrificed and the whole brain was dissected for estimation of markers of oxidative stress.

**Kindling induction**

For pentylenetetrazole kindling, a sub-convulsant dose of pentylenetetrazole (30 mg/kg, in a volume of 10 mL/kg) was injected intraperitoneally on every second day (i.e. day 1, day 3, day 5...). The first incidence of seizure with stage five was observed between day 35 (i.e. 18th injection) and day 39 (i.e. 20th injection). Pentylenetetrazole was administered up to day 43 (22nd injection) or until seizure stage 5 on two consecutive trials was achieved, whichever was earlier. Seizure activity was evaluated for 30 min using the following scale [31]: Stage 0: no response; Stage 1: hypertonia, vibrissae twitching; Stage 2: head nodding, head clonus and myoclonic jerk; Stage 3: unilateral forelimb clonus; Stage 4: rearing with bilateral forelimb clonus; Stage 5: generalized clonic-tonic seizures (or death within 30 min) with loss of writing reflex. The number of myoclonic jerks and the latencies to myoclonic jerks and generalized clonic-tonic seizures were recorded. Animals were considered kindled if they exhibited stage 5 of seizures on two consecutive trials. Animals were also observed for 24 h mortality.

**Tissue preparation**

Following the behavioural testing, the mice were decapitated under ether anaesthesia and the brains were quickly removed, cleaned with ice-cold saline and stored at −80 °C.

The whole brain of each mouse was dissected out and divided into two cerebral hemispheres for biochemical estimations. From one half, 10 % (w/v) homogenate was prepared with ice-cold 0.1 M phosphate buffer (pH 7.4), and lipid peroxidation product, reduced glutathione and GABA concentration were assessed.
Brain lipid peroxidation
Malondialdehyde, a measure of lipid peroxidation, was measured as described by Jainkang et al. [32]. The reagents acetic acid 1.5 mL (20 %) pH 3.5, 1.5 mL thiobarbituric acid (0.8 %) and 0.2 mL sodium dodecyl sulphate (8.1 %) were added to 0.1 mL of processed tissue samples, and then heated at 100 °C for 60 min. The mixture was cooled with tap water and 5 mL of n-butanol/pyridine (15:1), 1 mL of distilled water was added. The mixture was vortexed vigorously. After centrifugation at 4000 rpm for 10 min, the organic layer was separated and absorbance was measured at 532 nm using a spectrophotometer. The concentration of malondialdehyde was expressed as nmol/g tissue.

Brain reduced glutathione
Brain reduced glutathione was measured according to the method of Ellman [33]. The homogenate was mixed with equal quantity of 10 % trichloroacetic acid (v/v) and centrifuged to separate the proteins. To 0.01 mL of this supernatant, 2 mL of phosphate buffer (pH 8.4), 0.5 mL of 5′-dithiobis (2-nitrobenzoic acid) and 0.4 mL of double distilled water were added. The mixture was vortexed and the absorbance read at 412 nm within 15 min. The concentration of reduced glutathione was expressed as μg/g tissue.

Brain GABA level
The brain GABA level was estimated in groups of mice. The measurement of GABA, based on the method of Lowe et al. [34], was carried out as follows. The brains were rapidly removed, blotted, weighed and taken in ice cold 5 mL trichloroacetic acid (10 % w/v), homogenized and centrifuged at 10,000 g for 10 min at 0 °C. A sample (0.1 mL) of tissue extract was taken in 0.2 mL of 0.14 M ninhydrin solution in 0.5 M carbonate-bicarbonate buffer (pH 9.9). This solution was kept in a water bath at 60 °C for 30 min then cooled and treated with 5 mL of copper tartrate reagent (0.16 % disodium carbonate and 0.03 % copper sulphate and 0.0329 % tartaric acid). After 10 min, the fluorescence reading was taken at 377/451 nm in a spectrofluorimeter. For GABA standards, different amounts (20, 40, 60, 80, 100 μg) mixed with 1.5 μM glutamic acid were dissolved in 0.1 mL 10 % trichloroacetic acid (w/v). GABA level was determined by the measurement of the formed fluorescent product resulting from the reaction of GABA with ninhydrin in an alkaline medium, in the presence of glutamate [35]. The GABA content in brain was expressed in μg/g of wet brain tissue.

Acute toxicity test
The acute toxicity test for the IGEFA was carried out to evaluate any possible sign of toxicity. Mice of either sex were divided into control and test groups. The first group served as a normal control treated with vehicle. The IGEFA (5, 15, 30, 90, 180, 360, 720, 1440, 2880 and 5760 mg/kg) was administered orally to different groups of mice. After administration of these extracts, mice were allowed access to food and water ad libitum and behavioural parameters including convulsion, hyperactivity, sedation, grooming, loss of righting reflex, increased or decreased respiration, food and water intake and mortality were observed for a period of 14 days [36]. The dose of extract necessary to induce mortality by 50 % relative to the control value, called the median lethal dose (LD₅₀) was estimated according to the method described by Litchfield and Wilcoxon [37].

Data analysis
Data were expressed as mean ± standard error of the mean (S.E.M.) per group. The percentages of protection against chemical-induced seizures were measured and Fisher’s exact test (two-tailed) was used to compare percentages of protection. For the behavioural seizure in the bicuculline test, the control groups were compared to the extract-treated groups by the two-way repeated measures analysis of variance, followed by Newman-Keuls post hoc test. The effects of antiepileptic drugs were assessed by counting the number of hippocampal paroxysmal discharges in 20 min period post-injections. Data were expressed as mean ± standard error mean of percentage of cumulative duration, mean duration and number of crisis per 20 min periods, compared with vehicle condition. For each extract, the number of recorded seizures during the 20 min period post-injection was compared between doses using a two-way analysis of variance with repeated measures. Post-hoc comparisons versus vehicle conditions were performed using the Newman-Keuls test. The differences were considered significant at p < 0.05. In the acute toxicity test the median lethal dose (LD₅₀) was estimated according to the method described by Litchfield and Wilcoxon [37].

Results
Effects of IGEFA on bicuculline-induced motor seizures in mice
In all mice, i.p. injection of bicuculline successfully induced motor seizures as assessed by the experimenter (Table 1). Several behavioural effects were observed after the administration of bicuculline at a dose of 2.7 mg/kg, including clonic and tonic components of seizures, ataxia and head weaving. The IGEFA antagonized bicuculline-induced motor seizures in all pretreated mice. Pretreatment with IGEFA in doses ranging from 15–90 mg/kg significantly influence the time to onset of the first seizure (either clonic [F(7, 52) = 114.62; P < 0.001] or tonic [F(7, 38) = 125.17; P < 0.001]), the duration of the
clonic \[F(7, 64) = 146.46; \ P < 0.001\] and tonic \[F(7, 29) = 132.74; \ P < 0.001\] components of seizures, the number of motor seizures \[F(7, 42) = 106.22; \ P < 0.001\] and the incidence of death \[F(7, 26) = 97.21; \ P < 0.001\]. The durations of both tonic and clonic seizures were significantly reduced. A significant delay was found in the time to onset of clonic seizures. Diazepam (5 mg/kg), an anticonvulsant, protected the mice against bicuculline-induced motor seizures. Responses to bicuculline in mice treated with IGEFA were not different from those of diazepam-treated mice (Table 1).

**Pentylenetetrazole-induced seizures**

*Effects of IGEFA on acute administration of pentylenetetrazole at a dose of 70 mg/kg*

Pentylenetetrazole (70 mg/kg) produced hind-limb tonic seizures in all the eight mice treated with vehicle. The IGEFA increased the latency to the first seizures from 3.3 ± 1.3 min to 7.4 ± 0.6 min \[F(5, 28) = 92.5; \ P < 0.01\] at the dose of 30 mg/kg, and from 3.3 ± 1.3 min to 9.9 ± 0.6 min \[F(5, 28) = 92.5; \ P < 0.001\] at the dose of 60 mg/kg (Fig. 3). The number of seizures was reduced in all the test groups as compared to negative control. The IGEFA dose-dependently protected animals against clonic seizures induced by pentylenetetrazole. At the dose of 15 mg/kg, the IGEFA protected 50 % \(P < 0.05\) of mice against seizures. The dose of 90 mg/kg provided a 100 % \(P < 0.001\) protection of mice. The percentage of mortality protection was highly significant for all tested groups when compared to negative control. Complete protection against mortality was achieved with 60 and 90 mg/kg IGEFA and clonazepam. These effects were comparable to that of clonazepam (0.1 mg/kg), a standard antiepileptic drug (Fig. 3).

**Involvement of the GABA_A receptor complex in pentylenetetrazole induced seizures**

The IGEFA, when administered alone at a dose of 90 mg/kg, protected all mice against pentylenetetrazole-induced seizures. The anticonvulsant effects of the IGEFA were strongly antagonized by flumazenil (a specific diazepam receptor antagonist at the doses of 1, 2, 3, 4 and 6 mg/kg). This effect was statistically different from the data of vehicle \[F(5, 34) = 72.4; \ P < 0.001\] but did not exhibit differences with IGEFA administered at a dose of 15 mg/kg. In a similar manner, the anticonvulsant effect of the IGEFA (90 mg/kg) was strongly antagonized and abolished by FG7142 (a specific inverse diazepam receptor agonist at the doses of 4, 6, 8, 10, 12 and 14 mg/kg) (Fig. 4).

*Effects of IGEFA on acute administration of pentylenetetrazole at a dose of 50 mg/kg*

Twenty min after pentylenetetrazole (50 mg/kg) injection in animals, high voltage sharp waves (1500–4500 μV, 3–5 Hz) followed by higher frequency and low voltage rhythmic activity (10–14 Hz, 700–1100 μV) were recorded. These hippocampal and cortical paroxysmal discharges lasted between 20 and 60 s and their rate of occurrence was variable between vehicle-treated group or tested groups, with a maximum of one discharge every other minute (Fig. 5). Behavioural seizures with head nodding could be observed concomitantly with hippocampal and cortical paroxysmal discharges. However, some animals also displayed stereotyped behaviour, such as exploration or grooming. The antiepileptic effects of IGEFA were tested in pentylenetetrazole-treated mice at the doses of 15, 30, 60 and 90 mg/kg. After pentylenetetrazole administration at a dose of 50 mg/kg, vehicle-treated mice presented a high-voltage fast ictiform activity with isolated spike-and-wave discharges (no significant difference by ANOVA). In vehicle-treated mice the number of crisis (t/20 min), the cumulative duration of crisis (sec/20 min), and the mean duration of crisis (sec) recorded were not significantly affected (Fig. 5). EEG activity was strongly impaired and animals

### Table 1 Effects of IGEFA or diazepam on bicuculline-induced motor seizures in mice

| Treatments       | Dose (mg/kg) | Behavioural seizures | Duration (min) | Number of motor seizures | Mortality |
|------------------|-------------|----------------------|----------------|--------------------------|-----------|
|                  |             | Onset (min)          | Tonus | Clonus | Tonus | Clonus |                   |                     |
| Vehicle + Saline| – –         | –                    | –    | –      | –    | –      | –                | –                   |
| Vehicle + Bic    | – + 2.7     | 2.35 ± 0.21          | 6.15 ± 1.12 | 5.32 ± 1.47 | 69.32 ± 1.38 | 24.16 ± 1.14 | 8/8               |
| IGEFA + Bic      | 15 + 2.7    | 4.12 ± 0.27          | 11.16 ± 1.13 | 4.21 ± 1.15 | 19.12 ± 1.54* | 14.57 ± 1.26 | 1/8***            |
| IGEFA + Bic      | 30 + 2.7    | 6.15 ± 1.26*         | 13.53 ± 1.04* | 3.49 ± 1.26 | 16.35 ± 1.29* | 12.32 ± 1.39* | 1/8***            |
| IGEFA + Bic      | 60 + 2.7    | 7.21 ± 1.23*         | 16.17 ± 1.18* | 3.15 ± 1.13 | 12.49 ± 1.17* | 11.49 ± 1.81* | 0/8***            |
| IGEFA + Bic      | 90 + 2.7    | 7.49 ± 1.49*         | 18.32 ± 1.36* | 2.34 ± 1.52* | 11.26 ± 1.27*** | 11.52 ± 1.16* | 0/8***            |
| Diazepam + Bic   | 5 + 2.7     | 8.27 ± 1.28**        | 21.42 ± 2.19** | 1.26 ± 1.24* | 11.32 ± 1.45** | 10.26 ± 1.28* | 0/8***            |

Results are expressed as mean ± S.E.M., for 8 animals, bicuculline (2.7 mg/kg; s.c.) was injected intraperitoneally in mice.

*P < 0.05, **P < 0.01, ***P < 0.001 significantly different compared to the vehicle, data were analysed by two-way ANOVA, followed by Newman-Keuls post hoc test.
showed signs of motor incapacitation. The acute administration of IGEFA at the dose of 90 significantly suppressed hippocampal and cortical paroxysmal discharges in a dose-dependent way (Figs. 5 and 6) and the number of crisis $[F(5, 49) = 74.2; \ p < 0.001]$, the cumulative duration of crisis $[F(5, 52) = 85.5; \ p < 0.001]$, and the mean duration of crisis $[F(5, 34) = 92.5; \ p < 0.001]$ recorded were significantly decreased or partially blocked. No mortality of mice was recorded after administration of the IGEFA at the doses of 60 and 90 mg/kg (Figs. 5 and 6).

**Effects of IGEFA on chronic administration of pentylenetetrazole at a dose of 30 mg/kg**

*Effects on the development of pentylenetetrazole kindling*

The data shown in Table 2 represent the effects of IGEFA or sodium valproate on treatment with a sub-convulsant dose (30 mg/kg) of pentylenetetrazole every 2 days. The data indicate that the development of kindled convulsions was directly proportional and cumulative with repeated exposure to pentylenetetrazole. Confirming previous studies (Taiwe et al., [36]), repeated administration of a sub-convulsant dose of pentylenetetrazole (30 mg/kg) on alternate days (for 30.00 ± 1.65 days, 15 injections) resulted in increasing convulsive activity leading to generalized clonic-tonic seizures (stage 5) $[F(7, 28) = 124.8, \ p < 0.001]$ on two consecutive trials. Pretreatment with IGEFA at the doses of 15 mg/kg or vehicle did not modify the course of kindling induced by pentylenetetrazole. However, the higher doses of IGEFA (30, 60 and 90 mg/kg) suppressed the kindled seizure significantly ($P < 0.001$), as none of the animal could achieve stage 5 with 22 injections of pentylenetetrazole. The statistical analysis showed a significant difference in the development of kindling amongst the treated groups $[F(7, 28) = 68.42, \ p < 0.001]$. Sodium valproate (300 mg/kg) significantly ($p < 0.05$) delayed the course of development of kindling by pentylenetetrazole and provided complete protection against seizures (Table 2).

*Effects of IGEFA on seizures in pentylenetetrazole kindled mice*

As shown in Table 2, IGEFA administered orally at a dose of 60–90 mg/kg suppressed significantly the expression of pentylenetetrazole-kindled seizures. Pretreatment with IGEFA caused dose-dependent increase in the latency of myoclonic jerks $[F(7, 42) = 135.3, \ p < 0.001]$ as well as the latency to generalized tonic-clonic seizure $[F(7, 38) = 54.1, \ p < 0.001]$ and a decrease in number of myoclonic jerks $[F(7, 32) = 104.8, \ p < 0.001]$ as compared to the vehicle-treated pentylenetetrazole mice. IGEFA treatment produced a significant increase in the latency to myoclonic jerks from 45.4 ± 8.6 s in the vehicle-treated pentylenetetrazole mice to 126.5 ± 8.2 s ($P < 0.001$) and 173.6 ± 9.3 s ($P < 0.001$) in the groups administered IGEFA at the doses of 60 and
Effects of the coadministration of iridoids glycosids extract of *Feretia apodanthera* receptor complex, on the convulsions induced in mice by pentylenetetrazole mice to 18.5 ± 2.1 (p < 0.01), 128.7, 102.2, and 172.3 s (p < 0.001) in the vehicle, IGEFA, valproate-treated group (Table 2). A significant difference in the number of myoclonic jerks was observed [F(7, 42) = 128.7, p < 0.001]. This decreased from 64.3 ± 2.5 in the vehicle-treated pentylenetetrazole mice to 4.9 ± 0.2 s (p < 0.001) and 3.3 ± 0.2 s (p < 0.001) in the groups administered 30, 60 and 90 mg/kg of the IGEFA, respectively (Table 2). Seizure stage amongst the groups was also significantly different [F(7, 43) = 195.3, p < 0.001]. The IGEFA significantly decreased the seizure mean stage from 4.0 ± 0.0 in the vehicle-treated pentylenetetrazole mice to 2.2 ± 0.3 (p < 0.05), 1.8 ± 0.3 (p < 0.01) and 1.3 ± 0.4 (p < 0.01), respectively in the groups administered IGEFA 30, 60 and 90 mg/kg, respectively (Table 2).

**Effects of IGEFA on brain MDA levels in pentylenetetrazole-kindled mice**

The malondialdehyde level in the vehicle-treated pentylenetetrazole mice was significantly higher than that seen in vehicle-treated saline mice. Pentylenetetrazole kindling significantly increased (p < 0.001) the brain malondialdehyde level from 142.8 ± 16.2 nmol/g wet tissue in the vehicle + saline-treated mice to 454.6 ± 18.4 nmol/g wet tissue in the distilled water-treated pentylenetetrazole mice. The IGEFA dose-dependently and significantly attenuated the increased brain malondialdehyde levels due to pentylenetetrazole kindling in a dose-dependent manner [F(7, 53) = 38.2, P < 0.001]. The MDA levels significantly decreased from 454.6 ± 18.4 nmol/g wet tissue in the vehicle-treated pentylenetetrazole mice to 195.1 ± 19.2 nmol/g (p < 0.05), 175. ± 17.4 nmol/g (p < 0.001) and 156.2 ± 16.4 nmol/g (p < 0.001) wet tissue in groups administered IGEFA 30, 60 and 90 mg/kg, respectively. IGEFA per se (90 mg/kg) caused a decrease in the oxidative stress as indicated by the significant decrease (p < 0.001) in whole brain malondialdehyde levels as compared with the vehicle-treated pentylenetetrazole mice (Table 3).

**Effects of IGEFA on brain glutathione levels in pentylenetetrazole-kindled mice**

The marker for oxidative stress, glutathione, plays an important role in protecting cells against oxidative damage by scavenging free radicals. Thus, in the present study, whole brain glutathione levels were measured in all mice groups. There were significant differences in
reduced glutathione levels between the test groups and the negative control group \( [F(7, 54) = 102.3, P < 0.001] \). In vehicle-treated pentylentetrazole animals the brain glutathione level was 101.2 ± 14.3 μg/g-wet tissue. The IGEFA (15, 30, 60 and 90 mg/kg, p.o.) administration in all doses significantly increase this levels. The glutathione levels are 185.8 ± 14.5 μg/g-wet tissue \( (p < 0.01) \) and 192.3 ± 16.2 μg/g-wet tissue \( (p < 0.01) \) for the mice pre-treated by the doses of 60 and 90 mg/kg IGEFA, respectively. IGEFA (90 mg/kg) alone also caused a significant increase \( (199.2 ± 14.2 \text{ μg/g-wet tissue}; p < 0.01) \) in the brain glutathione levels as compared to the control group (Table 3).

**Effects of IGEFA on brain GABA content**

The systemic administration of the IGEFA (15–30 mg/kg, p.o.) did not produce any significant effect \( (p > 0.05) \) in the level of brain GABA concentration in animals (Table 3). However a significant increase in the level of brain GABA concentration in animals administered with the IGEFA was observed at the doses of 60 mg/kg \( (p < 0.01) \) and 90 mg/kg \( (p < 0.01) \), and sodium valproate \( (300 \text{ mg/kg, } p < 0.001) \) the positive control, was observed 1 h after oral administration (Table 3).

**Acute toxicity**

The general behavioral changes of the mice were observed following oral administration of IGEFA at 5, 15, 30, 90, 180 and 360 mg/kg doses, which were graded through time. The IGEFA at doses lower than 360 mg/kg in mice did not produce any abnormality in fur, eye color, asthenia, anorexia, salivation, piloerection, or diarrhea in all the treated mice, and there were no deaths recorded. Doses of 5–360 mg/kg did not cause any detectable changes, whereas, the doses of 720–5760 mg/kg seemed to be lethal and caused deaths within 24–48 h. The physical signs and symptoms of toxicity, which occurred in response to IGEFA, were a decrease in motor activity and exploration. The mice that died from a high dose \( (5760 \text{ mg/kg}) \) of IGEFA showed signs of respiratory failure (decreased respiratory rate and irregular breathing), gasping and coma before death. The internal organs of both controlled and treated groups did not show any unusual signs and were found to be normal in both size and
color. The median lethal dose (LD₅₀) was calculated to be 2197.7 mg/kg (Table 4).

Discussion

The present studies were undertaken in order to examine the anticonvulsant effects of IGEFA using mice models of generalised tonic-clonic seizures induced by chemicals. IGEFA has anticonvulsant effects on seizures triggered by the GABAₐ receptor antagonists bicuculline or pentylenetetrazole. This anticonvulsant effects is demonstrated with both electroencephalographic recordings and behavioural observations.

Interestingly, the effects of IGEFA against bicuculline-induced motor seizures are comparable with the standard anticonvulsant diazepam in the case of acute administration. IGEFA (30–90 mg/kg) administered acutely by the oral route strongly protected mice against the convulsions induced by bicuculline, a selective antagonist of GABA at the GABAₐ receptors [38]. Bicuculline, acts directly on the postsynaptic GABAₐ receptor complex to induce hyperactivity behaviour and seizures [39]. GABA is the major inhibitory neurotransmitter of the mammalian nervous system and bicuculline, a competitive GABAₐ receptor antagonist is a known proconvulsant [40, 41]. The dose-dependent increase in the anticonvulsant effects of IGEFA in the bicuculline-induced motor seizures test may be explained by an action of IGEFA on the GABAₐ receptor itself in the central nervous system. However, it may also act differently on other signalling contributors to counterbalance the excessive stimulation provided GABAₐ antagonism.

It has been reported that pentylenetetrazole induces seizures by inhibiting the GABA pathway in the central nervous system [42–44]. In addition, altered excitatory, inhibitory and/or modulatory neurotransmitter receptor densities have been observed in the brains of pentylenetetrazole-treated animals [45]. IGEFA strongly protected mice against 70 mg/kg pentylenetetrazole-induced seizures. This antagonism of pentylenetetrazole-induced seizures suggested the existence of anticonvulsant activity and the interaction of the IGEFA with GABAergic neurotransmission or its functional neutralization [7, 46]. In addition, to determine which neurotransmitter system is involved in the acute anticonvulsant properties of IGEFA, a selective benzodiazepine receptor antagonist, flumazenil or FG7142, an inverse diazepam receptor agonist of the GABAₐ receptor complex, were introduced in the 70 mg/kg pentylenetetrazole-induced seizures test. Mice protection by the IGEFA administration at a dose of 90 mg/kg was indeed antagonized by flumazenil or FG7142. These results indicate that the effects of IGEFA should be mainly mediated via the GABAergic system [26, 38, 47].

Pentylenetetrazole administration produces a characteristic behavioral pattern of events: ear twitch, vibrissae twitch, straub tail, myodonic twitch, forelimb clonus, falling and tonic hind limb extension [22]. These seizure behaviors correlate with spiking activity and spike-wave discharges in the cortex as measured by electroencephalography (EEG) [22]. Pre-treatment with IGEFA significantly attenuated the 50 mg/kg pentylenetetrazole-induced seizures in mice. The pentylenetetrazole-induced epilepsy model is an effective model for screening the efficacy of anti-epileptic drugs. The behavioral and electroencephalographic manifestations of
| Treatments          | Dose (mg/kg) | Behavioural seizure | Seizure mean stage | Kindling development (day) | Number | Duration (s) | Latency to generalized clonic-tonic seizures | Duration of generalized clonic-tonic seizures |
|---------------------|--------------|---------------------|-------------------|--------------------------|--------|-------------|---------------------------------------------|---------------------------------------------|
|                     |              | Myoclonic jerks     |                   |                          | Myoclonic jerks latency | Onset of clonic seizures |                           |                                           |
| Vehicle + Saline    | – + –        | –                   | –                 | –                        | –                  | –                                  | –                                           | –                                           |
| Vehicle + PTZ       | – + 30       | 30.00 ± 1.65        | 64.32 ± 2.45      | 45.45 ± 8.55             | 61.83 ± 8.61       | 185.45 ± 15.55                     | 19.48 ± 1.32                    | 4.00 ± 0.00                               |
| IGEFA + PTZ         | 15 + 30      | 32.21 ± 2.45        | 28.23 ± 2.14**    | 68.54 ± 5.25             | 75.16 ± 7.61       | 252.27 ± 24.14*                     | 14.12 ± 1.25                    | 2.33 ± 0.45*                              |
| IGEFA + PTZ         | 30 + 30      | 44.13 ± 2.28*       | 18.48 ± 2.13**    | 85.48 ± 4.47             | 185.21 ± 9.15***   | 324.15 ± 27.12***                   | 8.24 ± 0.16**                    | 2.16 ± 0.27**                             |
| IGEFA + PTZ         | 60 + 30      | 44.36 ± 2.32*       | 14.25 ± 2.34**    | 126.53 ± 8.15***         | 285.20 ± 7.62***   | 388.22 ± 35.26***                   | 4.86 ± 0.24***                   | 1.83 ± 0.28**                             |
| IGEFA + PTZ         | 90 + 30      | 43.72 ± 2.48*       | 8.45 ± 1.22***    | 173.62 ± 9.34***         | 371.52 ± 6.27***   | 495.48 ± 57.41***                   | 3.29 ± 0.23***                   | 1.33 ± 0.44**                             |
| SVA + PTZ           | 300 + 30     | 43.58 ± 2.34***     | 7.24 ± 1.56***    | 157.47 ± 8.52***         | 245.57 ± 6.45***   | 582.54 ± 48.36***                   | 2.27 ± 0.15***                   | 1.16 ± 0.27**                             |
| SVA + Saline        | 300 + –      | –                   | –                 | –                        | –                  | –                                  | –                                           | –                                           |
| IGEFA + Saline      | 90 + –       | –                   | –                 | –                        | –                  | –                                  | –                                           | –                                           |

Results are expressed as mean ± S.E.M., for 6 animals. Data were analysis by two-way ANOVA, followed by Newman-Keuls post hoc test. *P < 0.05, **P < 0.01, ***P < 0.001, significantly different compared to the vehicle-treated pentylenetetrazole mice.
pentylentetrazole-induced seizures in rodents suggest that the studied test is a model of generalized tonic-clonic seizures [21]. The acute administration of IGEFA at the doses of 30, 60 and 90 mg/kg significantly suppressed hippocampal and cortical paroxysmal discharges in a dose-dependent way; and the number of crisis, the cumulative duration of crisis, and the mean duration of crisis recorded were significantly decreased or partially blocked. In the current study it was found that the time elapsed before the appearance of myoclonic jerk and generalized tonic-clonic seizures was

| Treatment | Dose (mg/kg) | Sex | D/T | Mortality latency (h) | Toxic symptoms |
|-----------|-------------|-----|-----|---------------------|----------------|
| Vehicle + Saline | – + – | Male | 0/5 | – | None |
| Vehicle + PTZ | – + 30 | Male | 0/5 | – | None |
| IGEFA + PTZ | 15 + 30 | Male | 0/5 | – | None |
| IGEFA + PTZ | 30 + 30 | Male | 0/5 | – | None |
| IGEFA + PTZ | 60 + 30 | Male | 0/5 | – | None |
| IGEFA + PTZ | 90 + 30 | Male | 0/5 | – | None |
| SVA + PTZ | 300 + 30 | Male | 0/5 | – | None |
| SVA + Saline | 300 + – | Male | 0/5 | – | None |
| IGEFA + Saline | 90 + – | Male | 0/5 | – | None |

Results are expressed as mean ± S.E.M., for 6 animals. Data were analyzed by two-way ANOVA, followed by Newman-Keuls post hoc test.

MDA malondialdehyde, GSH glutathione, GABA gamma-aminobutyric acid, SVA sodium valproate, IGEFA iridoid glycosides extracts of Feretia apodenthera

*P < 0.05, **P < 0.01, ***P < 0.001, significantly different compared to the vehicle-treated PTZ mice

Table 3 Effects of IGEFA on lipid peroxidation product, brain reduced glutathione and brain GABA content in whole mice brain on pentylentetrazole-induced kindling in mice

| Treatments | Dose (mg/kg) | Brain lipid peroxidation, MDA (nmol/g-wet tissue) | Brain reduced glutathione, GSH (μg/g-wet tissue) | Brain GABA level (μg/g-wet tissue) |
|------------|-------------|-----------------------------------------------|-----------------------------------------------|----------------------------------|
| Vehicle + Saline | – + – | 142.79 ± 16.24 | 196.24 ± 12.31 | 398.51 ± 12.42 |
| Vehicle + PTZ | – + 30 | 454.62 ± 18.41 | 101.17 ± 14.24 | 297.38 ± 13.59 |
| IGEFA + PTZ | 15 + 30 | 362.24 ± 16.26* | 132.15 ± 13.15* | 398.53 ± 18.37 |
| IGEFA + PTZ | 30 + 30 | 195.13 ± 19.15** | 158.47 ± 12.41* | 397.62 ± 13.85 |
| IGEFA + PTZ | 60 + 30 | 178.47 ± 17.39** | 185.75 ± 14.53** | 435.49 ± 17.52* |
| IGEFA + PTZ | 90 + 30 | 156.19 ± 16.43*** | 192.25 ± 16.24** | 455.13 ± 16.38** |
| SVA + PTZ | 300 + 30 | 149.17 ± 19.75*** | 195.62 ± 14.46** | 452.36 ± 15.46** |
| SVA + Saline | 300 + – | 147.36 ± 17.15*** | 197.37 ± 19.31** | 462.37 ± 17.25** |
| IGEFA + Saline | 90 + – | 152.61 ± 17.35*** | 199.21 ± 14.19** | 471.41 ± 14.33** |

Results are expressed as mean ± S.E.M., for 6 animals. Data were analyzed by two-way ANOVA, followed by Newman-Keuls post hoc test.

MDA malondialdehyde, GSH glutathione, GABA gamma-aminobutyric acid, SVA sodium valproate, IGEFA iridoid glycosides extracts of Feretia apodenthera

*P < 0.05, **P < 0.01, ***P < 0.001, significantly different compared to the vehicle-treated PTZ mice

Table 4 Acute toxicity of the IGEFA administered orally to different groups of mice

| Treatment | Dose (mg/kg) | Sex | D/T | Mortality latency (h) | Toxic symptoms |
|-----------|-------------|-----|-----|---------------------|----------------|
| Vehicle | – | Male | 0/5 | – | None |
| IGEFA | 5 | Male | 0/5 | – | None |
| IGEFA | 15 | Male | 0/5 | – | None |
| IGEFA | 30 | Male | 0/5 | – | None |
| IGEFA | 90 | Male | 0/5 | – | None |
| IGEFA | 180 | Male | 0/5 | – | None |
| IGEFA | 360 | Male | 0/5 | – | None |
| IGEFA | 720 | Male | 3/5 | 36–48 | Hypoactivity, piloerection, salivation |
| IGEFA | 1440 | Male | 4/5 | 24–36 | Hypoactivity, piloerection, salivation |
| IGEFA | 2880 | Male | 5/5 | 24–36 | Hypoactivity, piloerection, salivation |
| IGEFA | 5760 | Male | 5/5 | 36–48 | Asthenia, anorexia, salivation, asthenia |

D/T = Dead/Treated mice; None = No toxic symptoms during the observation period; mortality latency = time to death after the oral administration. The IGEFA was two groups of mice. Mice in each group were carefully examined for any signs of toxic (behavioral changes and mortality) for 14 days. Control group received vehicle (10 ml/kg, per os)
significantly increased in mice treated with various doses of IGEFA. Our results demonstrate that inhibition of excitatory drive in the cortex and hippocampus can significantly delay seizures onset and suppress epileptiform discharges. This provides a proof of principle that this IGEFA could be used in the future for treatment of generalized tonic-clonic seizures and for management of generalized epilepsy. The anticonvulsant effects of IGEFA against pentylentetrazole-induced seizures indicated their possible effectiveness also against absence seizures as drugs that inhibit pentylentetrazole-induced convulsions are generally effective against absence seizures [48, 49]. Pentylentetrazole induces convulsions by inhibiting the GABA pathway in the central nervous system via the inhibition of GABA\textsubscript{A} receptor-chloride channel complex [7, 49–51]. However, the effects of a lyophilized aqueous extracts stem barks extracts of Feretia apodanthera against pentylentetrazole-induced kindling, -induced kindled seizures, or prolongation of hexobarbital sleeping time have been reported [10, 18].

In the present study, sub-convulsive dose of pentylentetrazole, when given intraperitoneally, on alternate days induced kindling in the vehicle-treated mice. The groups which were administered IGEFA showed dose-dependent protection against seizures. Kindled seizures are widely accepted as an animal model of epilepsy, wherein repeated sub-threshold brain stimulation, electrical or chemical, leads to behavioral signs of tonic and clonic seizures [52]. IGEFA significantly increased the latencies to myoclonic jerks, clonic seizures and generalized tonic-clonic seizure as well as duration of generalized tonic-clonic seizure as compared to the vehicle-treated pentylentetrazole mice. The number of myoclonic jerks was also decreased by IGEFA in a dose-dependent manner. IGEFA also decreased the mean seizure stage dose-dependently as compared to the vehicletreated pentylentetrazole mice. IGEFA produced maximum seizure protective effect in the dose of 90 mg/kg. The seizure protection offered by IGEFA (90 mg/kg, p.o.) was comparable to the standard antiepileptic drug sodium valproate (300 mg/kg, p.o.). The protection offered by valproate acid on pentylentetrazole-induced kindling is well established [53]. Sodium valproate, as a standard anticonvulsant drug, has anticonvulsant effects on seizure induced by pentylentetrazole. Previous studies showed that sodium valproate is able to increase brain GABA levels via various mechanisms, including blocking GABA reuptake, inhibiting the enzymes that break down GABA, and enhanced GABA release from nerve terminals (central nervous system synapses) [54, 55]. The protective effect of sodium valproate on pentylentetrazole-induced kindling is believed to be achieved through different neural mechanisms including inhibition of the voltage-dependent sodium channels, facilitation of GABAAergic neurotransmission, reduced N-methyl-D-aspartate-receptor mediated glutamate excitation, increased serotonergic inhibition and attenuation of neurogenic inflammation [56–61].

Epilepsy has been described as a condition of excessive neuronal discharge associated with or resulting from oxidative stress [62, 63]. Free radicals are normal products of cellular aerobic metabolism involved in the development of seizures [64]. However, when the production of free radicals increases or defense mechanisms of the body decreases, they cause cellular dysfunction by attacking at the polyunsaturated sites of the biological membranes causing lipid peroxidation. Lipid peroxidation was described as a main factor in the etiology of epilepsy. The increase in levels of malondialdehyde is a marker of lipid peroxidation [65]. In the present study, pentylentetrazole kindling increased the level of malondialdehyde and decreased the glutathione level in the mice brain. Pentylentetrazole thus caused an imbalance between antioxidant and oxidant defense system which may be at least partially responsible for seizures [66]. Oral administration of IGEFA prevented the rise in brain malondialdehyde levels in a dose-dependent manner. The significant decrease in brain malondialdehyde levels with concomitant IGEFA administration as compared to pentylentetrazole alone treated mice indicates an attenuation of lipid peroxidation. IGEFA administered at a dose of 90 mg/kg alone also significantly decreased the malondialdehyde level as compared to the vehicle-treated distilled water group which supports its antioxidant property. This antioxidant effect of IGEFA is coherent with its anticonvulsant effects if one assumes indeed that excess oxidation is a consequence of convulsions.

In addition, the present study demonstrated that IGEFA has protective effects on pentylentetrazole-induced convulsions and also on oxidative damage associated with pentylentetrazole. The decrease level of glutathione was observed in the distilled water-treated pentylentetrazole mice. IGEFA administration in all doses demonstrated increase in glutathione level in kindled mice brain tissue. Glutathione is an endogenous antioxidant which gets converted to oxidized form. This oxidized form of reduced glutathione reacts with free radicals and prevent generation of most toxic hydroxyl radical [66]. Increased lipid peroxidation during kindling is independent of iron salts and excitotoxin. These results indicate that, during kindling, there is an excessive oxidative stress pertaining as a consequence of reduced glutathione levels [10, 67, 68]. These antioxidant effects of the extracts of Feretia apodanthera in pentylentetrazole-kindled mice are supported by the findings of a recent study [10] wherein oral supplementation of a lyophilized aqueous extracts of Feretia apodanthera decreased the malondialdehyde, and increased the glutathione levels in brain mouse. However,
IGEFA treatment has restored the reduced glutathione level in the brain tissues of pentylenetetrazole-kindled mice.

The main action of the pentylenetetrazole-induced seizures is reducing GABA levels [44, 69]. Reports suggest that pentylenetetrazole-induced seizures presumably by impairing GABA-mediated inhibition by an action at the GABA_A receptors [70]. GABA is one of the important endogenous inhibitory neurotransmitters widely distributed in the central nervous system. Its reduction in the brain is associated with a number of neurological disorders (such as anxiety, depression and epilepsy) [70, 71]. The present study further demonstrates that, systemic administration of IGEFA produces significant changes in behaviour, such as sedation, hypoactivity and motor incoordination in mice which is indicative of either central nervous system depressant or muscle relaxant effects [36]. The post-synaptic GABA_A receptors are implicated in the inhibitory mechanism. Drugs that enhance GABA_A receptor-mediated inhibition, such as benzodiazepines, phenobarbital, valproate and felbamate, can be used to prevent the convulsions induced by pentylenetetrazole [48, 72]. It was found that administration of IGEFA and sodium valproate significantly enhanced the brain GABA concentration which again is suggestive of an anticonvulsant action of the extracts and the reference drug. Taken together, we suggest that the anticonvulsant action of IGEFA is correlated to an increase in GABA concentrations in the brain [30, 73]. This was also evidenced in the electroclinical study. Further studies may be warranted to elucidate which exact compound is responsible for these actions although it should be noticed that all five iridoid glycosides had very resembling chemical structures. It is therefore not impossible that they possess equivalent pharmacological properties.

We did not observe any mortality case including with the highest doses of IGEFA (range studied 5–360 mg/kg). Therefore, IGEFA may be considered to be relatively safe at these doses.

Conclusions

In summary, this study illustrated that IGEFA possess potent anticonvulsant activity and significantly reduces the progression of kindling and attenuates the oxidative stress in mice. These finding provides scientific rationale for the use of Feretia apodanthera extracts for the amelioration of generalized epilepsy observed in traditional medicine in some countries of Western Africa, particularly in Cameroon. Thus, more studies are necessary to clarify the chemical structural requirements involved in the antiepileptic effects of the IGEFA components and the biophysical mechanisms underlying their pharmacological properties directly on expressed GABA_A receptors.

Abbreviations

ANOVA, analysis of variance; GABA, gamma-aminobutyric acid; HMQC, heteronuclear multiple quantum coherence; HN; Herbier National du Cameroun; IGEFA, iridoid glycosides of Feretia apodanthera; NMR, nuclear magnetic resonance; RP-HPLC, reversed-phase high performance liquid chromatography

Acknowledgments

The authors are very thankful to Smartox Biotechnologies, France, and the University of Buea, Cameroon, for supporting us by providing apparatus and reagents.

Funding

No fund was provided by any funding agency for the research reported.

Availability of data and materials

The datasets supporting the conclusions of this article are included within the article.

Authors’ contributions

Conceived and designed the experiments: GST, MDW; Performed the experiments: GST, BD, TBT, JR; Analyzed the data: GST, BD, FDDD, MDW; Contributed reagents/materials/analysis tools: GST, MDW; Wrote the paper: GST, MDW. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable. This article is not a clinical study involving human participants and this manuscript does not contain any individual clinical data.

Ethics approval and consent to participate

The investigation conforms to the Guide for the Care and Use of Laboratory Animal published by the US National Institutes of Health (NIH; publication No. 85-23, revised 1996) and received approval of the Cameroon National Ethical Committee (Yaoundé, Cameroon) for animal handling and experimental procedure (Ref N’FW/RB00001954).

Author details

1Department of Zoology and Animal Physiology, Faculty of Science, University of Buea, Buea, Cameroon. 2Department of Chemistry, Faculty of Science, University of Maroua, Maroua, Cameroon. 3Department of Animal Biology and Physiology, Faculty of Science, University of Yaoundé I, Yaoundé, Cameroon. 4Institut du Thorax, INSERM UMR 1087/CNRS UMR 6291, Nantes, France. 5University of Nantes, Nantes, France. 6Smartox Biotechnology, Saint Martin d’Hères, France.

Received: 6 August 2015 Accepted: 6 August 2016

Published online: 12 August 2016

References

1. De Boer HM, Mula M, Sander JW. The global burden and stigma of epilepsy. Epilepsy Behav. 2008;12(4):540–6. doi:10.1016/j.yebeh.2007.12.019. PubMed PMID: 18280210.
2. Thurman DJ, Beghi E, Begley CE, Berg AT, Buchhalter JR, Ding D, et al. Standards for epidemiologic studies and surveillance of epilepsy. Epilepsia. 2011;52 Suppl 7:2–26. doi:10.1111/j.1528-1167.2011.01511.x. PubMed PMID: 21899536.
3. McCormick DA, Contreras D. On the cellular and network bases of epileptic seizures. Annu Rev Physiol. 2001;63:815–46. doi:10.1146/annurev.physiol.63.1.815. 63/1/815 PubMed PMID: 11181977.
4. Duncan JS, Sander JW, Siddiqua SM, Walker MC. Adult epilepsy. Lancet. 2006;367(9516):1087–100. doi:10.1016/S0140-6736(06)66947-8. PubMed PMID: 16581409.
5. Badawy RA, Harvey AS, Macdonell RA. Cortical hyperexcitability and epileptogenesis: understanding the mechanisms of epilepsy - part 1. J Clin Neurosci. 2009;16(3):355–65. doi:10.1016/j.jocn.2008.08.026. PubMed PMID: 19124246.
6. Bancaud JHO, Rubio-Donnadieu F, Seino M, Dreiffus FE, Penny JK. Proposal for revised clinical and electroencephalographic classification of epileptic seizures. Epilepsia. 1981;22:489–501.

7. Burn EN, Taiwe GS, Nkansah LA, Moto FC, Seke Etert PF, Hiana IR, et al. Validation of anticonvulsant and sedative activity of six medicinal plants. Epilepsy Behav. 2009;14(3):454–8. doi:10.1016/j.yebeh.2008.12.022. Epub 2009/01/24. PubMed PMID: 19162225.

8. Loscher W, Nau H, Marescaux C, Vergnes M. Comparative evaluation of anticonvulsant and toxic potencies of valproic acid and J-2-en-valproic acid in different animal models of epilepsy. Eur J Pharmacol. 1984;99(2):211–8. PubMed PMID: 6429823.

9. Ngo Burn E, Taiwe GS, Moto FC, Ngoupaye GT, Nkantchoua GC, Pelanken MM, et al. Anticonvulsant, anxiolytic, and sedative properties of the roots of Nauclea latifolia Smith in mice. Epilepsy Behav. 2009;15(4):434–40. doi:10.1016/j.yebeh.2009.05.014. Epub 2009/06/30. PubMed PMID: 19560975.

10. Taiwe GS, Moto FC, Ayissi ER, Ngoupaye GT, Najdoune JS, Nkantchoua GC, et al. Effects of a lyophilized aqueous extract of Feretia apodanthera Del. (Rubiaceae) on pentylentetrazole-induced kindling, oxidative stress, and cognitive impairment in mice. Epilepsy Behav. 2015;48:100–8. doi:10.1016/j.yebeh.2014.11.022. PubMed PMID: 25601583.

11. Dzefiete PD, De Waard M. Antidepressant, myorelaxant and anti-anxiety-like activity of 1-(pyrid-3-ylsulfonamido)-2-nitroethylenediamine. J Med Chem. 1981;24(2):111–7.

12. Kerharo J, Adams JG. La pharmacopée Sénégalaise traditionnelle. Paris: Vigot Frère, 1974.

13. Adjanohoun JE, Aboukakar N, Dramane K, Ebot ME, Ekpere JA, Enow-Orock FG, et al. Effects of Nauclea latifolia Smith in mice. Phytomedicine. 1999;288(1):121–3. PubMed PMID: 113. PubMed

14. Pousset JL, Plantes médicinales africaines. Paris: ACCT, Edition Marketing; 2006. p. 345-6.

15. Arbonnier M, Arbes, arbustes et lianes des zones sèches d’Afrique de l’Ouest. Paris: Centre de coopération internationale en recherche agronomique pour le développement/Muséum national d’histoire naturelle/union mondiale pour la nature; 2000.

16. Bailleul F, Delaveau P, Rabaron A, Plat M, Koch M. Feretia et gardenoside on nervous function and neuron damage in focal cerebral ischemic rats. J Ethnopharmacol. 1989;258(3):215–32. PubMed PMID: 243.

17. Bailleul F, Delaveau P, Rabaron A, Plat M, Koch M. Apodantheroside, an iridoid glucosid from Feretia apodanthera Del. (Rubiaceae) on pentylentetrazole-induced kindling, oxidative stress, and cognitive impairment in mice. Epilepsy Behav. 2015;48:100–8. doi:10.1016/j.yebeh.2014.11.022. PubMed PMID: 25601583.

18. Bailleul F, Cathala-Jacquier F, Delaveau P, Gabriel JP. Recherches pharmacologiques préliminaires sur le Feretia apodanthera Del. Ann Pharm Fr. 1981;39(2):111–7.

19. Li CY, Li L, Li YH, Ai HX, Zhang L. Effects of extract from Cornus officinalis on nitric oxide and NF-kappaB in cortex of cerebral infarction rat model. Zhongguo Yi Xue Ke Xue Za Zhi. 2009;20:368–71. PubMed PMID: 1927430.

20. Riban V, Bouilleret V, Pharm-Le BT, Fritschy JM, Marescaux, Deapulis A. Evolution of hippocampal epileptic activity during the development of hippocampal sclerosis in a mouse model of temporal lobe epilepsy. Neuroscience. 2002;112(1):101–11.

21. Pavinos G, Franklin KBJ. The mouse brain in stereotaxic coordinates. In: press A, editor. 2nd Edition. New York: Academic Press; 2001.

22. Pavinos GFKBJ. The mouse Brain in Stereotaxic Coordinates. Second ed. New York: Academic; 2001.

23. Sotoing Taiwe G, Ngo Burn E, Talia E, Dawe A, Okomolo Moto FC, Tekmou Ngoupaye et al. Antipsychotic and sedative effects of the leaf extract of Crooscephalum bauchiangni (Hutch.) Milne-RedH (Asteraceae) in rodents. J Ethnopharmacol. 2012;143(1):213–20. doi:10.1016/j.jep.2012.06.026. PubMed PMID: 22750453.

24. Racine RJ. Modification of seizure activity by electrical stimulation. II. Motor seizure. Electroencephalogr Clin Neurophysiol. 1972;32(3):281–94. PubMed PMID: 4110937.

25. Taiwe GS, Burn EN, Talia E, Dimo T, Weiss N, Sidiki N, et al. Antipyretic and antinociceptive effects of Nauclea latifolia root. Br J Clin Pharmacol. 2014;77(6):1004–13. doi:10.1111/bcp.12530. PubMed PMID: 24540881.

26. Litchfield Jr JT, Wilcoxson F. A simplified method of evaluating dose-effect experiments. J Pharmacol Exp Ther. 1949;96(2):213–9. PubMed PMID: 1805236.

27. Low JR, Randle MP, Ritter JM. Pharmacology. London/New York: Churchill Livingstone; 1999.

28. Sperber EF, Wurpel JN, Zhao DY, Eyerman GS. The fluorometric measurement of glutamic decarboxylase and its distribution in brain. J Neurochem. 1958;31(18–18).

29. Sutton I, Simmonds MA. Effects of acute and chronic pentobarbitone on the gamma-aminobutyric acid system in rat brain. Biochem Pharmacol. 1974;23(13):1801–18. PubMed PMID: 4156468.

30. Taiwe GS, Burn EN, Talia E, Dimo T, Weiss N, Sidiki N, et al. Antipyretic and antinociceptive effects of Nauclea latifolia root. Free Radic Biol Med. 1990;9:451–4. PubMed PMID: 6428923.

31. Vicini S, Alho H, Costa E, Minervini MJ, Sarti MR, Vaccarino FM. Modulation of gamma-aminobutyric acid-mediated inhibitory synaptic currents in dissociated cortical cell cultures. Proc Natl Acad Sci U S A. 1986;83(23):3926–73. PubMed PMID: 3097650.

32. Lüscher W, Schmidt D. Which animal model should be used in the search for new antiepileptic drugs? A proposal based on experimental and clinical considerations. Epilepsia. 1988;2:145–81.

33. Corda MG, Orlandi M, Lecca D, Carboni G, Frau V, Giorgi O. Pentylenetetrazol-induced kindling in rats: effect of GABA function inhibitors. Pharmacol Biochem Behav. 1991;40(2):329–33. PubMed PMID: 1805236.

34. Petkaneval A, Schwartzkroin PA, Moshe SL. Models of seizures and epilepsy. 1st Edition. In: Press EA, editor. Burlington: Elsevier Inc.; 2006. p. 345-6.

35. Sutton I, Simmonds MA. Effects of acute and chronic pentobarbitone on the gamma-aminobutyric acid system in rat brain. Biochem Pharmacol. 1974;23(13):1801–18. PubMed PMID: 4156468.

36. Taiwe GS, Burn EN, Talia E, Dimo T, Weiss N, Sidiki N, et al. Antipyretic and antinociceptive effects of Nauclea latifolia root. Br J Clin Pharmacol. 2014;77(6):1004–13. doi:10.1111/bcp.12530. PubMed PMID: 24540881.

37. Titarenko A, Whitley K, Delfino J, Burt R, White HS. Clinical significance of animal seizure models and mechanism of action studies of potential antiepileptic drugs. Epilepsia. 1997;38 Suppl 1:S9–151. PubMed PMID: 9092952.
et al. BMC Complementary and Alternative Medicine (2016) 16:285

49. Rang HP, Dale MM, Rotter JM, Flower RJ. Rang and Dale’s Pharmacology. In: Livingstone C, editor. Anti-epileptic drugs. 2007. p. 575–86.

50. Corda MG, Giorgi O, Longoni B, Orlandi M, Biggio G. Decrease in the function of the gamma-aminobutyric acid coupled chloride channel produced by the repeated administration of pentylentetrazol to rats. J Neurochem. 1990;55(4):1216–21. PubMed PMID: 1697889.

51. Ngo Burn E, Ngah E, Ngo Mune RM, Ze Minkoulou DM, Tallé E, Moto FC, et al. Decocations of Bridélia micrantha and Croton macrostachyus may have anticonvulsant and sedative effects. Epilepsy Behav. 2012;24(3):319–23. doi:10.1016/j.yebeh.2012.03.028. PubMed PMID: 22583623.

52. Szynlinder J, Piechalc A, Blecharz-Klin K, Skorzewska A, Maciejak P, Walkowiak J, et al. Effect of kindled seizures on rat behavior in water Morris maze test and amino acid concentrations in brain structures. Pharmacol Rep. 2006;58(1):75–82. PubMed PMID: 16531633.

53. El-Azab MF, Moustafa YM. Influence of calcium channel blockers on anticonvulsant and antinociceptive activities of valproic acid in pentylenetetrazole-kindled mice. Pharmacol Rep. 2012;64(2):303–14. PubMed PMID: 22661180.

54. Davis LL, Ryan W, Adnoff B, Petty F. Comprehensive review of the psychotropic uses of valproate. J Clin Psychopharmacol. 2000;20(1 Suppl 1):15–71. PubMed PMID: 10646685.

55. Hoffmann K, Czapp M, Loscher W. Increase in antiepileptic efficacy during prolonged treatment with valproic acid: role of inhibition of histone deacetylases? Epilepsy Res. 2008;81(2–3):107–13. doi:10.1016/j.eplepsyres.2008.04.019. PubMed PMID: 18538545.

56. Chapman A, Keane PE, Meldrum BS, Simiand J, Vernieres JC. Mechanism of anticonvulsant action of valproate. Prog Neurobiol. 1982;19(4):315–59. PubMed PMID: 6134304.

57. Loscher W. Valproate: a reappraisal of its pharmacodynamic properties and mechanisms of action. Prog Neurobiol. 1999;58(1):31–59. PubMed PMID: 10321796.

58. Loscher W. Basic pharmacology of valproate: a review after 35 years of clinical use for the treatment of epilepsy. CNS Drugs. 2002;16(10):669–94. PubMed PMID: 12269861.

59. Jensen TS. Anticonvulsants in neuropathic pain: rationale and clinical evidence. Eur J Pain. 2002;6(Suppl A):61–8. doi:10.1053/euip.2001.0324. PubMed PMID: 1188243.

60. Owens MJ, Nemeroff CB. Pharmacology of valproate. Psychopharmacol Bull. 2003;37 Suppl 2:17–24. PubMed PMID: 14624230.

61. Winkler I, Blotnik S, Shimshoni J, Yagen B, Devor M, Bialer M. Efficacy of antiepileptic isomers of valproic acid and valpromide in a rat model of pentylenetetrazol-kindled cognition and oxidative stress in rats. Life Sci. 2010;87(19–20):1449–55. doi:10.1016/j.lfs.2010.09.006. PubMed PMID: 20891603.

62. Waldbaum S, Patel M. Mitochondrial dysfunction and oxidative stress: a contributing link to acquired epilepsy? J Bioenerg Biomembr. 2010;42(6):499–506. doi:10.1007/s10863-010-9320-9. PubMed PMID: 21132357.

63. Shin EJ, Jeong JH, Chung YH, Kim WK, Ko KH, Bach JH, et al. Role of oxidative stress in epileptic seizures. Neurochem Int. 2011;59(2):122–37. doi:10.1016/j.neuint.2011.03.025. PubMed PMID: 21672578.

64. Sejima H, Ito M, Kishi K, Tsuda H, Shiraishi H. Regional excitatory and inhibitory amino acid concentrations in pentylenetetrazol kindling and kindled rat brain. Brain Dev. 1997;19(3):171–5. PubMed PMID: 9134187.

65. Liu CS, Wu MH, Kao SH, Wei YH. Phenotypic-mediated oxidative stress in serum of female epileptics: a possible pathogenesis in the fetal hydantoin syndrome. Hum Exp Toxicol. 1997;16(3):177–81. PubMed PMID: 9088972.

66. Mehla J, Reeta KH, Gupta P, Gupta YK. Protective effect of curcumin against seizures and cognitive impairment in a pentylenetetrazole-kindled epileptic rat model. Life Sci. 2010;87(19–20):596–603. doi:10.1016/j.lfs.2010.09.006. PubMed PMID: 20840851.

67. Schulz JB, Lindenaus J, Seyfried J, Dichtangs J. Glutathione, oxidative stress and neurodegeneration. Eur J Biochem. 2000;267(14):4904–11. PubMed PMID: 10931172.

68. Gupta YK, Vreendranda Kumar MH, Srivastava AK. Effect of Centella asiatica on pentylenetetrazole-induced kindling, cognition and oxidative stress in rats. Pharmacol Biochem Behav. 2003;74(3):579–85. PubMed PMID: 12543222.

69. Walsh LA, Li M, Zhao TJ, Chiu TH, Rosenberg HC. Acute pentylenetetrazol injection reduces rat GABAα receptor mRNA levels and GABA stimulation of benzodiazepine binding with No effect on benzodiazepine binding site density. J Pharmacol Exp Ther. 1999;289(3):1626–33. PubMed PMID: 10336561.