Performance, intestinal histomorphology and bone composition of broiler chickens fed diets supplemented with genistein

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Abstract
The effect of dietary genistein on performance, intestinal morphology, caecal Lactobacillus spp. count, and tibia composition in broiler chickens after 21 and 37 days of feeding was investigated. A total of 360 Cobb 500 broiler chickens (21 days old) were randomly allocated to five treatments with six replicates of 12 birds each. They were fed a basal diet (C) or a basal diet supplemented with 200 (T1), 400 (T2), 600 (T3) and 800 (T4) mg genistein/kg of feed. Genistein supplementation did not affect feed intake, but improved body weight, weight gain and feed conversion ratio (FCR) after 21 days, while 600 mg/kg led to a significant increase in FCR after 37 days of feeding. Plasma triglyceride level decreased with dietary genistein after 21 days, while increases were found in T3 and T4 groups after prolonged supplementation. Significantly improved duodenal and jejunal villus length and width, crypt depth and villus/crypt ratio were observed after the first and the second finishing periods, respectively, while adverse effects were found in the ileum for both periods. At 42 days old, greater spleen and heart weights were measured in broilers fed diets with 800 mg/kg than in other broiler groups. The shorter genistein supplementation period (21 days) of 200 and 400 mg/kg had a positive effect on tibia wet weight, ash and calcium (Ca) content, while 37 days of the higher genistein doses administered to the T2, T3 and T4 birds significantly increased caecal lactic acid bacteria (LAB) counts. Thus, recommended doses should not exceed 400 mg/kg.

Keywords: broiler performance, blood triglyceride, Lactobacillus, prolonged fattening, small intestinal morphology

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Introduction
Phytogenic compounds from plants are considered one of the most effective solutions to inefficiencies in poultry production (Wallace et al., 2010). Over the last two decades, substantial interest has arisen in bioflavonoids, a group of secondary plant metabolites, as an alternative to growth-promoting antibiotics in poultry feedstuffs (Kamboh et al., 2015; Kamboh et al., 2019).

Isoflavones are a subclass of flavonoids and are mostly in the form of glycosidic conjugates, which are widely distributed in leguminous plant foods (Rasouli & Jahanian, 2015; Zhu et al., 2015). Numerous studies have demonstrated the beneficial activities of isoflavones, including maintaining and improving bird health, enhancing performance, promoting, antioxidant, anti-inflammatory, antibacterial, and immunomodulatory effects, exhibiting estrogenic-like properties, up-regulating metabolic parameters, increasing bone mineralization, and improving meat and egg quality (Onderci et al., 2004; Jiang et al., 2007; Sahin et al., 2007; Kamboh & Zhu, 2013a; Iqbal et al., 2014; Kamboh & Zhu, 2014; Kamboh et al., 2015; Rasouli & Jahanian, 2015; Rasouli & Jahanian; 2019).

Genistein, the second soy isoflavone in importance and occurrence after daidzein, is present in soy-based food at concentrations from 0.2 to 1 mg/g (Ganaí & Farooqi, 2015). Because of the proven health-promoting effects of genistein in animals and humans, purified and combined commercial preparations are available as feed additives and nutritional supplements with up to six times greater amounts of genistein than in foods (Payne et al., 2001; Clavel et al., 2005; Markovic et al., 2015; Shiralinezhad & Shakouri, 2017).
Because genistein, unlike other isoflavonoids, has a high therapeutic index, it has become the target compound for poultry dietary investigations, although at a variety of supplement dosages (Dixon & Ferreira, 2002; Onderci et al., 2004; Kamboh & Zhu, 2013a; Kamboh & Zhu, 2014; Iqbal et al., 2014; Lv et al., 2018; Rasouli & Jahanian, 2019). Previous studies investigated the effect of genistein supplementation in broiler diets on growth and intestinal parameters during six weeks of fattening, but results were inconsistent (Kamboh & Zhu, 2013a; Iqbal et al., 2014; Kamboh & Zhu, 2014; Rasouli & Jahanian, 2015; Lv et al., 2018; Rasouli & Jahanian, 2019). Furthermore, to the best of the authors’ knowledge, data are not available on the impact of purified genistein supplement in diets of broilers older than three weeks. Considering genistein’s estrogenic activity (Sethcull & Cassidy, 1999), and the impact of dose and birds’ age on its effectiveness (Iqbal et al., 2014), the aim of this study was to examine the effect of differing genistein supplementation levels on growth performance, cholesterol and triglyceride plasma levels, small intestinal morphology, caecal LAB population, internal organ weights, and tibia Ca and phosphorus (P) levels in the late fattening phase (birds were 42 days old), and after extended fattening (birds were 58 days old).

Materials and Methods

The experiment protocols and procedures were approved by the Veterinary Directorate of the Serbian Ministry of Agriculture, Forestry and Water Management (Approval No. 119-01-51/14/2017-09) and the Institutional Animal Care and Use Committee of the Faculty of Veterinary Medicine, University of Belgrade.

Animals were fed the experimental diet supplemented with genistein as a 99.6% pure extract obtained from the roots of *Sophora subprostrata* Chun et T. Chen and *Genista tinctoria* L. (Xi’AnHuilin Bio-Tech Co., Ltd., Xian, Shaanxi, P.R. China). The purity of the extract was determined by dissolving it in methanol and using the HPLC-DAD-ESI-MS/MS technique (Orčić et al., 2014) (Figure 1).

![Figure 1](image_url)

**Figure 1** Liquid chromatography-mass spectrometry chromatogram of the genistein extract obtained in electrospray ionization negative mode

One-day-old mixed sex Cobb 500 broiler chickens with an average initial weight of 44.11 g were purchased from a commercial hatchery and raised up to 21 days old, during which time they were fed with standard starter (1 - 10 days; crumbled pellets) and grower (11 - 20 days; 1 cm pellets) diets with free access to food and water and continuous lighting. At 21 days old, a total of 360 broiler chickens were weighed and allocated so that weight distributions were similar among experimental groups and six replicates. Thirty pens at 3 m² with concrete floors were covered with wood shavings as litter to give 12 birds in each replicate. Birds were reared according to the guidelines set by the Cobb broiler management guide (Vantress, 2012). The basal corn-soybean meal diet was prepared to meet the nutrient requirements of the broilers (NRC, 1994). From 21 days to 58 days, the birds received common finisher feed mixtures (1 cm pellets) supplemented with 0 (C), 200 (T1), 400 (T2), 600 (T3), and 800 (T4) mg genistein/kg feed. The study consisted of two periods. The first lasted 21 days (until day 42 of fattening), and the second 37 days (until day 58 of fattening). The basal diets for three growth phases are reported in Table 1.

Feed samples were ground, passed through a 0.5-mm sieve, and analysed for moisture (ISO 6496:1999), total ash (ISO 5984:2002), crude protein (ISO 5983-1:2005), total lipids (ISO 6492:1999), crude fibre (ISO 6865:2000), calcium (ISO 6490-1:1985), and phosphorus content (ISO 6491:1998) (Table 1).
Bodyweights were recorded individually on days 1, 11 (average bodyweight 343.1 g), 21, 42, and 58, and total feed intake was measured on days 42 and 58 according to the discarded feed leftovers. Bodyweight gain and feed conversion ratio for these periods were calculated using this data.

**Table 1** Ingredient composition, calculated and chemical analysis of the experimental diets for broiler chickens

| Ingredients (%)          | Starter | Grower | Finisher |
|--------------------------|---------|--------|----------|
| Maize                    | 50.85   | 44.15  | 44.95    |
| Wheat                    | 10.00   | 15.00  | 20.00    |
| Soy grits                | 15.00   | 17.00  | 20.00    |
| Soybean meal (CP 44%)    | 12.40   | 1.00   | 1.00     |
| Soybean cake (CP 41.5%)  | 17.00   | 23.30  | 14.70    |
| Monocalcium phosphate    | 1.20    | 1.00   | 0.90     |
| Chalk                    | 1.60    | 1.60   | 1.60     |
| Salt                     | 0.35    | 0.35   | 0.35     |
| Premix¹                  | 1.00    | 1.00   | 1.00     |
| Lysine                   | 0.20    | 0.20   | 0.10     |
| Methionine               | 0.20    | 0.20   | 0.20     |
| Adsorbent                | 0.20    | 0.20   | 0.20     |
| Total                    | 100.0   | 100.0  | 100.0    |

**Calculated analyses (%)**

| AMEₙ (MJ/kg) | 12.64 | 12.96 | 13.22 |
| Crude protein | 22.68 | 21.80 | 19.05 |
| Calcium       | 1.08  | 1.02  | 1.08  |
| Phosphorus    | 0.61  | 0.58  | 0.71  |
| Lysine        | 1.49  | 1.42  | 1.10  |
| Methionine + cysteine | 0.84 | 0.83 | 0.82 |
| Tryptophan    | 0.31  | 0.28  | 0.22  |

**Chemical composition (%)**

| Moisture     | 12.61 | 13.32 | 10.54 |
| Total ash    | 5.32  | 5.20  | 5.11  |
| Crude protein| 22.40 | 20.85 | 18.19 |
| Total lipids | 6.89  | 7.90  | 6.48  |
| Crude fibre  | 2.74  | 3.39  | 2.59  |
| Calcium      | 0.85  | 0.77  | 0.76  |
| Phosphorus   | 0.67  | 0.58  | 0.56  |
| NFE²         | 50.04 | 49.35 | 57.09 |

¹ vitamin A 15000 IU; vitamin D₃ 5000 IU; vitamin E 80 IU; nicotinic acid 62.5 mg; pantothenic acid 10 mg; folic acid 1.25 mg; d-biotin 0.10 mg; zinc 80 mg; manganese 120 mg; iron 78.5 mg; copper 19.92 mg; cobalt 1.4 mg; selenium 2.2 mg; antioxidant 100 mg

² Nitrogen-free extract

At the end of each experimental period (birds were 42 and 58 days old), the feed was withdrawn for 12 hours. After weighing, 36 randomly selected broilers per treatment (six per replicate) were electrically stunned and slaughtered by severance of the jugular veins. Blood samples (8 mL) of fasting broilers were collected in serum non-heparinized tubes. Within one hour, these were centrifuged at 3000 × g for 15 min and serum was separated. The concentrations of triglyceride and total cholesterol in the serum samples
were analysed using colorimetric methods with an automatic biochemical analyser (RA-1000, Bayer Corp., Tarrytown, NY, USA). After carcass evisceration by trained personnel, the adhering tissue was removed from liver, spleen, heart, and these and the left tibiae were weighed. Organ weight was expressed as a percentage of live weight at the same age.

Immediately after slaughter, for both experimental periods, the gastrointestinal tracts of 18 birds per treatment (three per pen) were removed and segments approximately one centimetre long were taken from the middle part of duodenum, jejunum and ileum, flushed in physiological saline, and fixed in neutral formalin (10% formalin (37 - 40%), NaH₂PO₄·H₂O 4.0 g, and NaHPO₄·H₂O 6.5 g) solution for 24 hours. After fixation and shaping, intestinal samples (one sample for each of the three intestinal segments per bird) were dehydrated in increasing concentrations of ethyl alcohol, cleared with xylene, infiltrated with paraffin and embedded in paraffin blocks. Sections 5 µm in thickness (three cross-sections from each sample) were placed on glass slides and stained with routine Mayer’s haematoxylin and eosin procedure, and with a combination of periodic acid Schiff’s stain and Alician blue (PAS-AB) (Yamabayashi et al., 1987; Smirnov et al., 2005). Histological sections were examined using a light microscope Olympus BX53 with the objective magnifications x4 and x10. On each cross-section, 10 points were selected for microscopic investigation using Olympus cellSens software (Olympus Life Science Solutions, Tokyo, Japan). These measurements were taken: the villus height and width, and cryptal depth, which were also used to calculate villus height/crypt depth ratio (VCR) for each intestinal segment (duodenum, jejenum and ileum) (Aptekmann et al., 2001).

At the age of 42 and 58 days, one bird from each replicate was randomly selected for sampling to determine caecal LAB count. After slaughter, the intestinal tracts were excised and the contents from the cecum were immediately collected aseptically into a sterile plastic container and transported under cold conditions to the laboratory. Approximately 1 g of digesta from each sample was mixed with 9 mL of sterile physiological saline solution (0.9% NaCl) and homogenized for 3 min using an Ultra-Turrax. These homogenates were serially diluted from 10⁻² to 10⁻⁹. To enumerate *Lactobacillus* spp., 0.1 mL of appropriately diluted suspension was inoculated directly onto the surface of the agar plate (MRS, Hi Media, India) followed by incubation at 30 °C for 72 hours in anaerobic conditions (Anaerocult A, Merck, Darmstadt, Germany). Results were expressed as log colony forming unit (CFU) per gram of cecal digesta.

After separating the drumsticks with thighs from the carcasses, all soft tissues and cartilaginous caps were removed from the left tibiae (six birds per pen) and the bones were kept frozen in plastic bags at -20 °C until analysis for mineral content. The tibiae were defatted, dried at 15 °C and combusted at 600 °C for 8 hours to determine ash percentage. Calcium content was measured using a spectrophotometer (Perkin-Elmer Analyst 700 MHS, PerkinElmer Life and Analytical Science, Connecticut, United States), while P content was determined by the spectrophotometric method (ISO 6491:1998).

These data were analysed with one-way ANOVA using SPSS 20.0 (IBM, Chicago, IL, USA). The statistical differences between groups were determined with Tukey’s post hoc multiple comparisons test. Orthogonal polynomial contrasts were used to evaluate the linear and quadratic effects of graded levels of dietary genistein on the parameters. Growth performance parameters were based on replicate as the experimental unit, while all other parameters were analysed on individual broilers. All results were expressed as mean ± pooled SEM. A probability value of *P* <0.05 was statistically significant.

**Results**

Data for growth performance are presented in Table 2. On day 42, the bodyweight of broilers was increased in those that had received genistein at 400 and 600 mg/kg (linear, *P* =0.004; quadratic, *P* =0.005), while on day 58, no significant differences were found among any of the groups (*P* >0.05). From three to six weeks of fattening, weight gain and daily weight gain of T2, T3, and T4 groups were higher than C, with the linear and quadratic effects being observed, while up to day 58 in the T3 group (600 mg/kg), weight gain decreased significantly. For both experimental periods, total feed intake was not affected (*P* >0.05) by genistein. Feed conversion ratio (FCR) from day 21 to day 42 was better in all groups that received genistein compared with the control, with a significant difference in T3 and T4 (1.64, 1.65, respectively; linear, *P* =0.0001). After extended fattening, up to 58 days, FCR was best in T4 (2.37), but this did not differ from the T1 and T2. However, supplementation with 600 mg/kg genistein had an adverse effect on FCR (2.88).
Table 2 Effects of different levels of genistein supplementation on broiler performance at 42 and 58 days old

| Trait    | Age (days) | Experimental group | SEM | Contrasts, P-value |
|----------|------------|--------------------|-----|--------------------|
|          |            | C     | T1     | T2     | T3     | T4     |       | Linear | Quadratic |
| Weight   | 21         | 967   | 980    | 993    | 972    | 979    | 14.95 | 0.432  | 0.266   |
|          | 42         | 2623a | 2727a  | 2847b  | 2781b  | 2772a  | 86.37 | 0.004  | 0.005   |
|          | 58         | 4021  | 4029   | 4122   | 3835   | 4095   | 174.73| 0.851  | 0.672   |
| Gain     | 21 - 42    | 1656a | 1747ab | 1854b  | 1809b  | 1793b  | 72.26 | 0.001  | 0.003   |
|          | 43 - 58    | 1293a | 1298a  | 1323a  | 1139b  | 1407b  | 82.65 | 0.559  | 0.021   |
| ADG      | 21 - 42    | 82.81a | 87.33ab | 92.68b  | 90.45b  | 89.63b  | 3.61  | 0.001  | 0.003   |
|          | 43 - 58    | 86.19a | 86.51a | 88.21a  | 75.92b  | 93.78a  | 5.51  | 0.559  | 0.021   |
| FI       | 21 - 42    | 3236  | 3133   | 3293   | 2964   | 2959   | 223.36| 0.031  | 0.442   |
|          | 43 - 58    | 3460  | 3251   | 3484   | 3282   | 3332   | 272.89| 0.564  | 0.859   |
| FC       | 21 - 42    | 1.96a | 1.79ab | 1.78ab  | 1.64b  | 1.65b  | 0.12  | 0.0001 | 0.282   |
|          | 43 - 58    | 2.68ab | 2.50ac | 2.63abc | 2.88b  | 2.37c  | 0.16  | 0.310  | 0.051   

*ab Within a row, means with a common superscript do not differ at *P* = 0.05

1. C: control group of broilers fed the basal diet; T1: basal diet supplemented with 200 mg/kg genistein; T2: basal diet supplemented with 400 mg/kg genistein; T3: basal diet supplemented with 600 mg/kg genistein; T4: basal diet supplemented with 800 mg/kg genistein

ADG: average daily gain; FI: feed intake; FC: feed conversion ratio

Dietary genistein supplement at 800 mg/kg significantly improved the spleen (linear, quadratic, *P* = 0.0001) and heart absolute and relative weights when birds were 42 days old, while in 58-day-old birds, these differences were not observed (Table 3).

Table 3 Effect of different levels of genistein supplementation on absolute and relative organ weights of broilers

| Organ  | Age     | Experimental group | SEM | Contrasts, P-value |
|--------|---------|--------------------|-----|--------------------|
|        |         | C     | T1     | T2     | T3     | T4     |       | Linear | Quadratic |
| Liver  | 42 days | 41.60 | 44.00  | 45.07  | 46.07  | 47.87  | 3.74  | 0.007  | 0.840   |
|        | 58 days | 63.07 | 60.07  | 58.40  | 57.20  | 56.60  | 0.41  | 0.057  | 0.588   |
| Spleen | 42 days | 2.13a | 2.07a  | 2.27a  | 2.33a  | 3.87a   | 1.83   | 0.0001 | 0.0001  |
|        | 58 days | 3.80  | 3.93   | 3.87   | 3.80   | 3.40   | 0.12   | 0.303  | 0.320   |
| Heart  | 42 days | 12.27a | 12.73a | 14.67ab | 15.47ab | 17.33b  | 0.02   | 0.0001 | 0.587   |
|        | 58 days | 22.33 | 19.13  | 21.87  | 21.33  | 19.47  | 0.06   | 0.312  | 0.884   |

Relative weight (% of body weight)

| Liver  | 42 days | 1.69  | 1.63   | 1.56   | 1.63   | 1.68   | 5.78   | 0.880  | 0.061   |
|        | 58 days | 1.57  | 1.49   | 1.42   | 1.49   | 1.38   | 1.38   | 0.064  | 0.017   | 0.690   |
| Spleen | 42 days | 0.088a | 0.077a | 0.077a | 0.084a | 0.137b | 2.45   | 0.0001 | 0.0001  |
|        | 58 days | 0.096 | 0.101  | 0.094  | 0.101  | 0.083  | 0.11   | 0.312  | 0.301   |
| Heart  | 42 days | 0.50ab | 0.47a  | 0.51ab  | 0.55ab  | 0.61b  | 0.02   | 0.002  | 0.137   |
|        | 58 days | 0.55  | 0.47   | 0.53   | 0.56   | 0.47   | 0.05   | 0.302  | 0.679   |

*ab Within a row, means with a common superscript do not differ at *P* = 0.05

1. C: control group of broilers fed the basal diet; T1: basal diet supplemented with 200 mg/kg genistein; T2: basal diet supplemented with 400 mg/kg genistein; T3: basal diet supplemented with 600 mg/kg genistein; T4: basal diet supplemented with 800 mg/kg genistein
Dietary supplementation with 200, 400, 600 and 800 mg/kg genistein had no significant effect on the plasma cholesterol levels of the broilers at the end of both periods (P >0.05) (Table 4). Genistein supplementation significantly decreased (linear, P =0.0001; quadratic, P =0.022) birds’ triglyceride levels during the first finisher period (day 42) compared with the control group. In contrast, on day 58, the authors’ measured negative effects on the birds’ triglyceride levels in those animals that were supplemented with 600 and 800 mg/kg.

**Table 4** Plasma total cholesterol and triglycerides concentrations in broilers at 42 and 58 days old

| Constituent | Age   | Experimental group | SEM | Contrasts, P-value |
|-------------|-------|--------------------|-----|--------------------|
|             |       | C T1 T2 T3 T4      |     | Linear Quadratic   |
| Cholesterol | 42 days| 3.42 2.81 2.97 3.19| 3.11| 0.37 | 0.662 0.079 |
|             | 58 days| 3.36 2.77 3.10 3.22| 3.30| 0.33 | 0.528 0.062 |
| Triglycerides| 42 days| 0.50a 0.35b 0.34b 0.30b| 0.31b| 0.07 | 0.0001 0.022 |
|             | 58 days| 0.31a 0.40ab 0.39ab| 0.42b| 0.43b| 0.002 0.207 |

Within a row, means with a common superscript do not differ at P =0.05

1 C: control group of broilers fed the basal diet; T1: basal diet supplemented with 200 mg/kg genistein; T2: basal diet supplemented with 400 mg/kg genistein; T3: basal diet supplemented with 600 mg/kg genistein; T4: basal diet supplemented with 800 mg/kg genistein.

There were no differences among treatments in the numbers of *Lactobacilli* in the caeca of birds supplemented with genistein or not in the 42-day-old birds, with the exception of 200 mg/kg genistein, in which the LAB count was lower (5.80 log CFU/g). However, on day 58, the number of LAB was increased (linear, P =0.0001) in the caeca of the groups that were supplemented with genistein at concentrations of 400, 600 and 800 mg/kg (6.24, 6.42, 6.38 log CFU/g, respectively (Figure 2).

**Figure 2** Caecal *Lactobacillus* populations in broilers fed diets without (C) or with 200 mg/kg (T1), 400 mg/kg (T2), 600 mg/kg (T3), and 800 mg/kg genistein at 42 and 58 days old

Bars with a common lowercase (uppercase) superscript do not differ at P =0.05 at 42 (58) days of age.

In 42-day-old birds the villus height in the duodenum was higher in broilers fed diets with the addition of genistein (Table 5). The best effect was achieved by supplementation with 400 mg/kg (T2). Villus width was reduced significantly with the addition of 400, 600 and 800 mg/kg genistein. Also, in all treatment groups, crypt depth was shorter and the villus height: crypt depth ratio was higher than those in the control.
group. After the second fattening period (day 58), genistein supplementation did not affect villus height, but it significantly reduced the villus width of the T4 group and the crypt depth of the T2, T3 and T4, and improved villus height: crypt depth ratio in the T4 group.

At 42 days, the effect of the genistein dose on the jejunal villus volume was somewhat inconsistent up to the concentration of 600 mg/kg, while 800 mg/kg led to significantly depressed jejunal mucosa development. With prolonged genistein supplementation in the birds’ diets, at 58 days positive effects on the villus height, villus width and the villus height: crypt depth ratio was measured in the jejunum, while the crypt depths were reduced significantly only with the inclusion of 200 and 800 mg/kg genistein.

Similar inconsistencies in the effect of high doses of genistein were found on the morphometric parameters in the ileum, in which the broilers fed the diets with genistein had significantly reduced villus height and villus height: crypt depth ratio, and these were the lowest for both fattening periods in the birds receiving 600 mg/kg genistein.

Table 5 Effect of different levels of genistein supplementation on intestinal morphometric parameters in broilers at 42 and 58 days old

| Structure                  | Age (in days) | Experimental group | SEM     | Contrasts, P-value |
|----------------------------|---------------|-------------------|---------|--------------------|
|                            |               | C | T1 | T2 | T3 | T4 |       | Linear | Quadratic |
| Duodenum                   |               |   |    |    |    |    |       |        |           |
| Villus height (μm)         | 42            | 1197<sup>a</sup> | 1212<sup>a</sup> | 1284<sup>b</sup> | 1226<sup>ab</sup> | 1257<sup>ab</sup> | 36.57  | 0.011 | 0.111 |
| Villus width (μm)          | 42            | 101.5<sup>a</sup> | 101.7<sup>a</sup> | 87.01<sup>ab</sup> | 76.36<sup>c</sup> | 79.48<sup>c</sup> | 4.33   | 0.001 | 0.169 |
| Crypt depth (μm)           | 42            | 162.9<sup>a</sup> | 133.6<sup>a</sup> | 131.3<sup>b</sup> | 131.4<sup>a</sup> | 125.9<sup>b</sup> | 5.90   | 0.001 | 0.001 |
| Villus height: crypt depth ratio | 42      | 7.48<sup>a</sup> | 9.10<sup>b</sup> | 9.86<sup>b,c</sup> | 9.38<sup>b</sup> | 10.10<sup>c</sup> | 0.43   | 0.001 | 0.001 |
| Jejunum                    |               |   |    |    |    |    |       |        |           |
| Villus height (μm)         | 42            | 978.7<sup>a</sup> | 950.6<sup>bc</sup> | 1132.0<sup>b</sup> | 991.4<sup>a</sup> | 880.1<sup>c</sup> | 44.28  | 0.014 | 0.0001 |
| Villus width (μm)          | 42            | 812.9<sup>a</sup> | 965.7<sup>b</sup> | 960.4<sup>c</sup> | 938.6<sup>a</sup> | 904.0<sup>b</sup> | 46.91  | 0.021 | 0.0001 |
| Crypt depth (μm)           | 42            | 80.54<sup>a</sup> | 82.60<sup>b</sup> | 88.00<sup>c</sup> | 87.83<sup>b</sup> | 88.33<sup>b</sup> | 3.66   | 0.001 | 0.158 |
| Villus height: crypt depth ratio | 42      | 58.51<sup>a</sup> | 7.26<sup>b</sup> | 6.85<sup>bc</sup> | 6.51<sup>b</sup> | 7.13<sup>c</sup> | 0.45   | 0.001 | 0.004 |
| Ileum                      |               |   |    |    |    |    |       |        |           |
| Villus height (μm)         | 42            | 563.3<sup>a</sup> | 541.5<sup>ab</sup> | 542.3<sup>ab</sup> | 503.6<sup>b</sup> | 503.9<sup>b</sup> | 22.93  | 0.001 | 0.907 |
| Villus width (μm)          | 42            | 590.1<sup>a</sup> | 505.9<sup>b</sup> | 531.1<sup>b</sup> | 492.3<sup>b</sup> | 493.7<sup>b</sup> | 25.73  | 0.001 | 0.014 |
| Crypt depth (μm)           | 42            | 56.53<sup>a</sup> | 70.99<sup>b</sup> | 60.23<sup>c</sup> | 65.19<sup>a</sup> | 64.14<sup>ac</sup> | 2.46   | 0.003 | 0.249 |
| Villus height: crypt depth ratio | 42      | 118.5<sup>a</sup> | 135.1<sup>b</sup> | 141.8<sup>b</sup> | 139.5<sup>c</sup> | 120.1<sup>c</sup> | 4.75   | 0.249 | 0.0001 |
| a,b,c Different lowercase superscript letters after means in the same row indicate differences (P < 0.05) among groups
1 C: control group of broilers fed the basal diet; T1: basal diet supplemented with 200 mg/kg genistein; T2: basal diet supplemented with 400 mg/kg genistein; T3: basal diet supplemented with 600 mg/kg genistein; T4: basal diet supplemented with 800 mg/kg genistein
In 42-day-old birds, genistein supplementation at concentrations of 200, 400 and 600 mg/kg resulted in significantly improved absolute weight and ash contents of the birds’ tibias (Table 6). Calcium content in the tibias was significantly increased by 200 and 400 mg/kg genistein supplementation on day 42, while on day 58, these levels led to a decrease in Ca. The most pronounced decrease in P content in the birds’ tibias was observed with 600 and 800 mg/kg genistein in the 42-day-old birds, while prolonged dietary supplementation resulted in an increase in P content in the 400 mg/kg genistein group that did not differ from the control group of birds.

Table 6 Effect of different levels of genistein supplementation on absolute and relative weight and mineral content of tibia of broilers at 42 and 58 days old

| Characteristic | Age   | Experimental group | SEM | Contrasts, P-value |
|---------------|-------|--------------------|-----|-------------------|
|               |       | C | T1 | T2 | T3 | T4 | Linear | Quadratic |
| Weight (g)    | 42 days | 12.00a | 14.67bc | 15.33b | 14.00bc | 13.33bc | 0.97 | 0.153 | 0.0001 |
|               | 58 days | 17.89 | 18.89 | 19.00 | 17.56 | 20.00 | 1.83 | 0.272 | 0.666 |
| Relative weight (% BW) | 42 days | 0.507 | 0.552 | 0.550 | 0.489 | 0.469 | 0.05 | 0.077 | 0.042 |
|               | 58 days | 0.430 | 0.457 | 0.437 | 0.449 | 0.483 | 0.06 | 0.232 | 0.623 |
| Ash (%)       | 42 days | 42.15a | 49.19b | 49.27b | 44.24c | 41.62a | 0.90 | 0.0001 | 0.0001 |
|               | 58 days | 42.96bc | 36.77bc | 39.05bc | 40.94c | 41.32bc | 1.60 | 0.702 | 0.0001 |
| Ca (%)        | 42 days | 15.32a | 17.33b | 16.75 | 15.53a | 14.24d | 0.26 | 0.0001 | 0.0001 |
|               | 58 days | 14.91a | 12.95bc | 13.66 | 14.35a | 14.74bc | 0.67 | 0.272 | 0.0001 |
| P (%)         | 42 days | 8.23ab | 8.60a | 8.34a | 7.63b | 7.55b | 0.38 | 0.0001 | 0.045  |
|               | 58 days | 7.35ab | 6.68a | 8.13b | 7.28a | 7.24a | 0.43 | 0.540 | 0.163  |

a,b,c Within a row, means with a common superscript do not differ at P =0.05
1 C: basal diet; T1: basal diet supplemented with 200 mg/kg genistein; T2: basal diet supplemented with 400 mg/kg genistein; T3: basal diet supplemented with 600 mg/kg genistein; T4: basal diet supplemented with 800 mg/kg genistein
Ca: calcium, P: Phosphorus

Discussion

The effects of isoflavones, including genistein, on bodyweight, weight gain, feed intake, and feed conversion ratio in poultry are variable. Diets supplemented with low to moderate levels (20 to 80 mg/kg) of genistein positively affected weight gain and FCR values of broilers during a six-week feeding period (Rasouli & Jahanian, 2015; Lv et al., 2018). Onderci et al. (2004) observed significantly improved growth performance as a result of the addition of 400 and 800 mg/kg genistein to a quail diet. However, some studies found dietary supplementation with purified genistein did not influence the growth rate or FCR of broiler chickens (Kamboh & Zhu, 2013a; Kamboh & Zhu, 2014; Iqbal et al., 2014). The results of the present study showed improved performances after 21 days of high-dose genistein treatment, but the 600 mg genistein/kg diet for 37 days led to a significant decrease in weight gain and increase in FCR. Rasouli and Jahanian (2019) found similar discrepancies in production results during various growth periods, in which low to moderate levels (20 - 80 mg/kg) and moderate levels (40 - 80 mg/kg) of genistein significantly increased average daily feed intake and weight gain after starter and grower periods, respectively, while high genistein levels (160 - 320 mg/kg) did not cause differences in these parameters or even worsened FCR during the starter period. In agreement with the present findings, Jiang et al. (2007) obtained inconsistency in performances of 43- to 63-day-old broilers with tenfold lower doses of isoflavone. Importantly, Qian and Sun (2009) showed that soy isoflavone aglycones could exhibit potentially adverse effects if they were not well balanced in the feed. Previous studies revealed the effect of low to high supplement doses of various isoflavone preparations (Payne et al., 2001; Gjorgjevska et al., 2014; 2016; Shiralinezhad & Shakouri; 2017), combinations and purified isoflavones (Kamboh & Zhu, 2013a; 2014; Onderci et al., 2004), and isoflavone glucosidase (Qian & Sun, 2009) in broiler chicks, laying hens, quails, immune-challenged animals (Rasouli & Jahanian, 2015; Zhu et al., 2015), and animals exposed to heat stress (Onderci et al., 2004; Sahin et al., 2007). These studies indicate that various factors – including kinds and forms of isoflavones, their interactions, dose, treatment duration, and bird age, gender, individual metabolism and health status –
largely determine growth-promoting effects, which are more evident in animals that are subjected to environmental stress.

In the present study the highest level of genistein significantly increased spleen and heart absolute and relative weights, with no effect on the liver weight for both finishing periods. As immune-modulatory substances, flavonoids could cause variations in the size of the main lymphatic organs in poultry. The size of these organs is considered a good indicator of changes in the degree of lymphocyte proliferation and immune responses (Kamboh et al., 2015; Rasouli & Jahanian, 2015). Rasouli and Jahanian (2019) found that dietary genistein did not affect the relative heart weight in broilers, which is not in accordance with the present findings. By increasing relative heart weight, dietary flavonoid quercetin induced a beneficial effect on the cardiovascular system of broilers (Goliomytis et al., 2014). Unfortunately, it was confirmed that the genetic selection of broilers for weight gain led to reduced relative heart size, which resulted in insufficient oxygen supply to tissues and development of metabolic disorders such as ascites and sudden death syndrome (Havenstein et al., 1994). Further research is needed to determine by which mechanisms and to what extent genistein has the potential to prevent these syndromes.

Deficiency in oestrogen production could lead to changes in the lipid and lipoprotein profiles and consequently to increased serum cholesterol level (Setchell & Cassidy, 1999). Some studies suggested that plant isoflavones, by acting like oestrogen agonists, were involved in the cholesterol-lowering effect (Anthony, 2000; Ali et al., 2004), while this effect was not found in others (Dewell et al., 2002; Nestel et al., 1999). This variability is because of the multifactorial mechanism through which cholesterol absorption and metabolism are modulated by enzyme regulation and depends primarily on the type and dose of isoflavone, and initial serum cholesterol level (Anderson et al., 1995). Since the main enzymes that are involved in cholesterol production are 3-hydroxy-3-methylglutaryl-coenzyme A (HMG CoA) synthase and reductase (Pakalapati et al., 2009), Kamboh and Zhu (2013b) showed a dose-dependent decrease in serum cholesterol and triglyceride levels in broilers fed diets supplemented with genistein and hesperidin, indicating their inhibitory effects on HMG-CoA reductase. In addition, the cholesterol-lowering effect is observed mainly in studies with soybean extract (Anderson et al., 1995; Yilmaz et al., 2008). The results of authors who used purified phytoestrogens, along with the current findings, showed no effect on cholesterol level (Dewell et al., 2002; Onderci et al., 2004). In contrast, Iqbal et al. (2014) and Rasouli and Jahanian (2019) found decreased plasma cholesterol levels in genistein-supplemented groups after a six-week feeding period. In terms of plasma triglyceride levels, an increase was observed in ovariectomized rats fed red clover extract rich in isoflavones (Pakalapati et al., 2009). The results from the present study support earlier findings that as a selective oestrogen receptor modulator, genistein differentially influences plasma lipid levels (Pakalapati et al., 2009). After 21 days of genistein supplementation, the authors measured a lowering plasma lipid effect, while in broilers with prolonged genistein supplementation at 600 and 800 mg/kg, adverse effects were measured on serum triglycerides levels (i.e. triglycerides increased).

Changes of intestinal morphology in terms of enlargement of villus length and width along with a decrease in crypt depth and a higher villus/crypt ratio are related to improved intestinal function owing to the greater surface area, better expression of brush border enzymes, and transport systems that allow higher nutrient absorption and a lower rate of epithelial cell turnover (Awad et al., 2011; Kamboh & Zhu, 2014). The present study showed that genistein induced a positive effect on villus height, crypt depth and villus/crypt ratio in the duodenum of six-week-old broilers. Since the duodenum is the major site of nutrient digestion and absorption (Qian et al., 2012), the current observed higher growth performance indicators in this age of birds could partly be a result of hypertrophied duodenal villi. Previous studies showed that supplementation with a genistein preparation and with purified genistein for 21 and 28 days, respectively, during starter and grower periods significantly improved the jejunal mucosal morphology of broiler chickens (Shiralinezhad & Shakouri, 2017; Rasouli & Jahanian, 2019). In the present study, this effect was not observed during the finisher period after 21 days, but only after prolonged genistein administration (37 days in total). The exact mechanism by which flavonoids affect intestinal mucosal structure is still unknown, but probably it could be attributed to their antioxidative activity against intestinal apoptotic and pro-oxidant factors (Awad et al., 2011; Kamboh et al., 2015). In addition, in the present study, genistein treatment impaired ileal morphometric features for both finishing periods in comparison with the control group. This suggests that the better production results that were obtained in broilers that received genistein supplementation were not related directly to the improved gut morphology, but to mechanisms such as enhanced antioxidative status, increased nutrient digestibility and depression of harmful intestinal microbial populations, with a shift to more beneficial ones (Onderci et al., 2004; Jiang et al., 2007; Qian et al., 2012; Rasouli & Jahanian, 2015; Shiralinezhad & Shakouri, 2017; Rasouli & Jahanian, 2019).

The authors observed that longer high-level genistein treatments led to an overall significant increase in caecal LAB count by about 0.73 log CFU/g compared with the control group. It was shown that higher genistein levels were more effective in reducing the ileal number of Salmonella and E. coli in broilers.
Genistein supplementation of broilers’ diet for three weeks during the commercial finishing phase exhibited a beneficial effect, while its prolonged administration did not cause significant improvements. However, further studies are needed to rationalize the use of these high genistein doses, taking into account the immune response, the cardiovascular system, gut microbiota, and the safety, quality and sustainability of the broiler meat and the cost-effectiveness of such production.

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Authors’ Contributions
MG designed the investigation, performed the analysis, and wrote the paper. MB helped performed analyses and co-wrote the paper. AR performed histological analyses. DS analysed and interpreted the performance results. VDJ carried out chemical analysis. MR collected data and performed biochemistry analysis. MZB and RM conceived the investigation together with the first author and helped collect and analyse the data.

Conflict of Interest Declaration
The authors declare they have no potential conflicts of interests.

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