Concurrent BK polyomavirus, adenovirus and cytomegalovirus infections in a patient treated for chronic lymphocytic leukaemia

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SUMMARY
A 58-year-old woman with chronic lymphocytic leukaemia (CLL) presented with 2 weeks of fever and haematuria following chemo-immunotherapy. CT scan showed thickening of her left urethra and bladder, suggesting pyelo-ureteritis with cystitis. The patient was initially treated for suspected bacterial urinary tract infection although repeated blood and urine cultures remained negative. She then received multiple transfusions for chemotherapy-induced pancytopenia while her urinary symptoms did not improve. Due to her immunocompromised status, she was tested for viral infection, which revealed, BK polyomavirus, adenovirus and cytomegalovirus in serum and urine. Cidofovir was initially administered to treat these infections while ganciclovir was used with filgrastim due to neutropenia. The patient subsequently improved. This case represents a diagnostic and therapeutic challenge due to the multiple concurrent viral infections causing haematuria as well as the combined post-chemo-immunotherapy and antiviral myelotoxicity in a CLL patient.

BACKGROUND
Chronic lymphocytic leukaemia (CLL) is a lymphoproliferative disorder associated with mature B-cell dysfunction, hypogammaglobulinaemia and T-cell abnormalities. In Europe and North America, CLL is the most common leukaemia, with median age of diagnosis of 70 years.1 Treatment is indicated only for symptomatic disease, such as anaemia <100 g/L and/or thrombocytopenia <100×109/L, massive splenomegaly, lymphadenopathy or constitutional symptoms.2

Infection is a frequent cause of morbidity and mortality in CLL. Roughly 30%–50% of all CLL patients suffer infections, with a high subsequent mortality rate.3 While the majority of infections affect more often the respiratory track, or skin and the gastrointestinal tract.4 The most frequent organisms causing infections are bacteria like Streptococcus pneumoniae, Staphylococcus aureus, Streptococcus pyogenes and Escherichia coli. This increased susceptibility to infection stems from B-cells with impaired immunoglobulin production capability and dysfunctional T-helper cells, which also assist in antibody production. This impaired capability to produce immunoglobulins, termed hypogammaglobulinaemia, affects 85% of CLL patients, particularly in advanced disease.4

Antineoplastic agents recommended for the treatment of CLL are also immunosuppressive and further increase the risk of infections.5 These include opportunistic micro-organisms such as Candida, Pneumocystis jirovecii, listeria, nocardia and atypical mycobacteria. In addition, patients have increased risk of viral diseases such as herpes simplex, Epstein-Barr and cytomegalovirus (CMV). Additional infectious complications may include hepatitis B reactivation and progressive multifocal

Figure 1 Positron emission tomography scan (right) and corresponding CT images (left) of multiple low-grade hypermetabolic lymph nodes in 2017, 2 years after diagnosis of chronic lymphocytic leukaemia. (A) Multiple small low-grade hypermetabolic lymph nodes in bilateral neck regions predominantly in the posterior triangles. SUV of 1.9 in the left and right upper posterior triangles. (B) Hypermetabolic lymph nodes in the left axillary regions, SUV 1.8. (C) Left external iliac lymph node SUV of 3. SUV, Standardized uptake value.

Figure 2 CT urogram on admission in November 2019, demonstrating (A) poorly distended bladder and marked thickening of bladder walls (green arrow) and (B) moderate dilation of both ureters and collection ducts with marked thickening and enhancement of the urothelium associated with moderate periureteric fat stranding.
In spite of the immunosuppressive risk, chemo-immunotheapy with fludarabine, cyclophosphamide and rituximab (FCR) remains the standard risk therapy in fit patients below 65 years of age with CLL, in absence of 17p deletion, or TP53 mutation, and with immunoglobulin heavy chain V (IGHV)-mutated.6

To reduce risk of infection, immunoglobulin levels should also be routinely monitored, particularly with rituximab, which reduces the number of B-cells expressing CD20. It is postulated that monitoring immunoglobulins while being treated with rituximab may reduce the risk of serious infections.4

Herein, we present a unique case of a woman treated for CLL who experienced severe pancytopenia and immunosuppression at the completion of her chemo-immunotherapy. She subsequently experienced triple concurrent viral infections, which posed a therapeutic challenge due to her pancytopenia.

**CASE PRESENTATION**

A previously healthy 58-year-old woman was diagnosed with CLL through a routine blood test in 2015. Her physical examination was unremarkable and her lymphocyte counts were at 12.36×10^9 cells/L. Bone marrow biopsy with immunohistochemistry was suggestive of CLL and stained positive for BCL2, CD5 (aberrant T-cell marker), CD20, CD23, CD43 and CD79a (B-cell markers) while BCL6 was negative. The PCR amplification study showed immunoglobulin heavy chain gene rearrangement along with IgG kappa light chain gene arrangement. No t(14;18) BCL2/JH or t(11;14) BCL1/JH gene rearrangements were observed. As she was asymptomatic, and positron emission tomography (PET) scan showed low grade metabolic activity in lymph nodes in cervical, axillary and iliac areas, no therapy was initiated (figure 1). She was followed every 6 months.

In November 2017 the patient had symptom of a rapid bilateral enlargement of the cervical lymph nodes accompanied by persistent fatigue, night sweats and decreased weight. Lyme disease, HIV, hepatitis B and C and syphilis serologies were performed, and all tests yielded negative results. Repeat PET/CT scan, cytogenetics and bone marrow biopsy demonstrated no evidence of Richter’s transformation into a diffuse large cell lymphoma. Due to significant bulky disease and enlarged adenopathy with B constitutional symptoms, chemo-immunotherapy with FCR was considered. This was furthermore rationalised as she was a fit patient younger than 65 years of age with standard risk for CLL due to the absence of 17p deletion, or TP53 mutation, and with IGHV being mutated.7

She received six cycles of FCR which was completed uneventfully on 11 November 2019. During FCR therapy, she received oral valacyclovir 500 mg two times per day for herpes prophylaxis as well as Septra DS (160 mg trimethoprim and 800 mg sulfamethoxazole) one tablet Mondays, Wednesdays and Fridays for *P. jirovecii* prophylaxis. On 18 November, she presented to the emergency room of the McGill University Health Centre (MUHC) with haematuria, suprapubic pain and fever. She was admitted and treated for a urine infection and slowly improved after different antibiotics such as ciprofloxacin, trimethoprim-sulfamethoxazole and ceftriaxone and blood and urinary bacterial cultures were negative. A CT urogram showed moderate bilateral hydroureretonephrosis with diffuse urethral thickening (figure 2). As a whole, these findings were suggestive of an inflammatory or infectious process evoking pyleoureteritis and cystitis.
INVESTIGATIONS
On admission, she was mildly pancytopenic (table 1). As the patient partially improved and had negative urine cultures, a cyclophosphamide-induced haemorrhagic cystitis was considered, and she was placed on continuous bladder irrigation.

On 1 December 2019 she developed a pancytopenia that required platelet and packed red blood cell transfusions. She remained persistently febrile despite repeated negative bacterial blood and urine cultures. Due to her CLL, pancytopenia and the consideration of immunosuppression induced by FCR chemotherapy, viral infections were investigated. PCR for adenovirus showed 22.4 copies/mL in blood, and positive in urine; PCR for polyoma virus showed 8.81 million copies/mL blood; and PCR for CMV showed 358 copies/mL in blood, confirming viral invasive infections (figure 3). To further investigate her pancytopenia, serology for parvovirus B 19 was carried out and was negative for IgM.

On 18 December 2019 the patient underwent a double J-stent insertion for bilateral hydronephrosis. Pelvic pain persisted and she continued to pass clots in her urine. Repeat CMV PCR testing on 25 December 2019 showed 7320 copies/mL while an ophthalmological assessment revealed no evidence of CMV retinitis or keratitis. Bone marrow biopsy results showed pancytopenia without transformation (figure 4).

TREATMENT
On 1 January 2020, cidofovir was administered to treat the three viral infections of BK polyomavirus, adenovirus and CMV. Cidofovir was given at the dose of 1 mg/kg intravenously three times a week instead of a weekly dose given her kidney injury and high creatinine levels. She received probenecid 2 g orally 1 hour before cidofovir, then 2 hours and 8 hours after the end of infusion to increase cidofovir levels in the blood. For renal protection, she was hydrated with 1L of 0.9% normal saline during cidofovir infusion. Filgrastim (G-CSF) was initiated for her neutropenia, darbepoetin for her anaemia, and intravenous IgG was infused three times at 4-week intervals for her hypogammaglobulinemia.

Neutropenia worsened despite filgrastim, and CMV viral load continued to increase. Due to cidofovir myelotoxicity, a decision was made to focus treatment on CMV infection, while ignoring the BK and adenovirus. To this end, the patient was changed to ganciclovir 250mg intravenous two times per day on 23 January 2020 and her urinary symptoms improved for the first time. On 1 February 2020, she was transitioned to oral valganciclovir 450mg daily and was discharged to home 1 week later, after 101 days in hospital.

OUTCOME AND FOLLOW-UP
A bone marrow biopsy on 27 January 2020 showed a hypocellular marrow with focal area up to 20%-30% haematopoietic tissue. No evidence of lymphocytosis or lymphoma and no dysplastic changes in morphology were observed. Immunohistochemistry showed <5%CD117, and was negative for CD20, CD34 and PAX5. The histological appearance represented haematopoietic regeneration post-chemotherapy. On 3 March 2020 follow-up, CMV load was undetectable, adenovirus load was 5000 copies/mL, BK load was 3.2×10⁸ copies/mL (table 1). She was switched on CMV prophylaxis with resolved haemorrhagic cystitis and received valganciclovir 450mg p.o. daily for another 3 months and until her absolute lymphocyte count recovers above 10⁹ cells/L. She continued trimethoprim-sulfamethaxole prophylaxis for 6 months post-chemotherapy. Her double-J stents remain in place. While clinically improved, her neutropenia improved on low dose filgrastim until 4 April 2020. Her blood counts normalised 1 month after and she is followed as an outpatient with Haematology, Infectious Diseases, and Urology services at the MUHC.

DISCUSSION
FCR chemo-immunotherapy is considered the treatment for standard risk CLL patients under the age of 65, with a high overall response rate. However, the predominant complication is myelosuppression. Persistent neutropenia was reported in 19% and late infections in 9% of patients. The frequency of cytopenias was 35% at 3 months and 12% at 9 months after completion of therapy, respectively. Another side effect is prolonged lymphoid immunodeficiency.

Human adenovirus infections usually present as a keratoconjunctivitis or respiratory tract infection. In most healthy individuals the infection is usually self-limited. However, in immunosuppressed patients, infection may disseminate and

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**Figure 3** Cytomegalovirus (CMV) and BK viremia (A) and adenoviremia (B) over the course of treatment.

**Figure 4** Histology and immunochemistry of bone marrow smear (A) and biopsy (B). Bone marrow biopsy demonstrating hypocellularity. Moderate left shift of the myeloid series and mild left shift of the erythroid series. No lymphocytosis or lymphoma, no definite diagnosis of myelodysplastic syndrome (B).

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become potentially life-threatening. Disseminated adenovirus infection can result from either de novo infection or a reactivation of clinically silent infection.10

BK virus is a member of the polyomavirus family. BK virus reactivation occurs only in immunosuppressed patients and can lead to nephropathy, ureteral stenosis, tubular interstitial damage and haemorrhagic cystitis. Viremia occurs in 20%–60% of this population. There is no specific treatment for BK virus-associated nephropathy, therefore viral replication can only be controlled through modulating the immunosuppressive condition.11

CMV is a herpes virus with a large double-stranded DNA that remains latent in the host after primary infection. Immunocompromised patients can have reactivation of their latent infection. Diagnosis of viral reactivation is made through CMV PCR in blood, tissue or body fluid. Manifestations of CMV infection occur most often in the gastrointestinal tract, lungs, liver and retina.12 As we did not observe the typical owl’s eye tissue lesion in the second bone marrow biopsy, we considered that CMV reactivation was a contributing factor for the pancytopenia, and treatment may have prevented such tissue lesion to develop. First line treatment of CMV infection includes intravenous ganciclovir or its oral prodrug valganciclovir, which are DNA polymerase inhibitors. The major side effect of these drugs is myelosuppression, including neutropenia.4

In our patient, the prolonged pancytopenias secondary to FCR was associated with haematuria with an ureteral mass. To date, only two observations on the use of cidofovir to simultaneously treat BK-virus associated haemorrhagic cystitis and CMV reactivation have been reported. However, in the case report by Held et al13 the patient had received a bone marrow transplant for chronic myeloid leukaemia, whereas our patient had CLL.

Cidofovir has been suggested in some cases for the treatment of BK virus-associated haemorrhagic cystitis.13 Cidofovir and foscarnet are second-line therapies in patients with CMV that are either resistant or refractory to ganciclovir. Cidofovir is a non-cyclic analogue of cytidine monophosphate that terminates CMV DNA synthesis. In vitro studies showed effectiveness against herpes viruses, including CMV, herpes simplex viruses 1 and 2, varicella-zoster, Epstein-Barr virus, human papillomavirus and adenovirus. The main adverse effect is nephrotoxicity, causing proximal tubular injury. Serum creatinine and urine protein measurements are recommended during administration.14 Probenecid decreases intratubular secretion and maintains a sufficient plasma concentration to allow for once weekly dosing at 5 mg/kg to reduce its nephrotoxicity.13 An alternate dosing regimen, initially used in our patient, was three times per week dosing at 3 mg/kg. In the study performed by Philippe et al13 intravenous cidofovir achieve remission in 81.5% of the 22 patients.

The patient reported by Held et al recovered fully with cidofovir13; however, our patient was switched to ganciclovir due to persistent neutropenia. Bone marrow biopsy was consistent with secondary treatment-related myelosuppression syndrome, without evidence of myelodysplasia or lymphoproliferative infiltration. Myelosuppression is a major side effect of ganciclovir. However, the antiviral effects far outweigh these risks and therefore ganciclovir and its oral prodrug valganciclovir should be considered for patients with pancytopenia secondary to CMV reactivation.

In summary, this unique case of triple-viremia induced haemorrhagic cystitis in an immunosuppressed CLL patient posed both a diagnostic and therapeutic challenge as treatments are nephrotoxic and further increase immunosuppression.
Case report

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