**Genome Sequence of the Uncommon *Streptococcus pyogenes* M/emm66 Strain STAB13021, Isolated from Clonal Clustered Cases in French Brittany**

Alexandra Meygret,a,b Pascal Vincent,a,b Séverine Moullec,b Jessica Nacazume,b Yahia Adnani,b Dominique Lavenier,c Samer Kayal,a,b Ahmad Failib

CHU Rennes-Service de Bactériologie et Hygiène Hospitalière, Rennes, Francea; CNRS, IGDR-UMR 6290, Team GeRMO, Rennes, Francet; National Institute for Research in Computer Science and Control, IRISA/GenScale, Team Bioinformatique, Campus de Beaulieu, Rennes, Francet

Here, we announce the complete annotated genome sequence of the invasive *Streptococcus pyogenes* strain M/emm66, isolated in 2013 from a subcutaneous abscess in new clustered cases in French Brittany.

**Group A Streptococcus pyogenes** (GAS) is a Gram-positive human pathogen that causes a broad range of invasive and non-invasive diseases (1, 2). Outbreak of infections due to GAS have long been known to be a major cause of human morbidity and mortality (3); however, the molecular basis of pathogen emergence and differences in strain virulence are still poorly understood.

Since 2009, we have been conducting a systematic survey of all GAS isolates in our geographical area based on emm-genotyping (4, 5) and have observed a clonal emergence (n = 13 isolates with the same pulsed field electrophoresis) of an unusual emm66 genotype (6–8). We sequenced and annotated the whole genome of an invasive GAS M/emm66 strain isolated in 2013 from a subcutaneous abscess, henceforth named STAB13021.

Bacterial growth and DNA extraction were performed as previously described (9, 10). Genomic DNA was sequenced using HiSeq 2000 technology (Illumina, Inc., San Diego, CA, USA), and the paired-end library was built using the MGX facility of the CNRS in Montpellier, France. There is a total of 13,785,510 high-quality reads giving an average 720-fold coverage of the genome, which was assembled using CLC Genomics Workbench version 6 software (http://www.clcbio.com). The resulting assembly consisted of 31 contigs, which were oriented on the basis of available sequences of GAS. After reassembling, 23 gaps persisted, which were filled by PCR followed by Sanger sequencing. Genome annotation was performed in parallel using the RAST server (11) and the NCBI PGAP (http://ncbi.nlm.nih.gov/genome/annotation_prok). Prophages were identified using the PHAge search tool (PHAST) (12).

Finally, strain STAB13021 harbored a single circular genome of 1,810,577 bp, with a G+C content of 38.35%. We identified 1,663 coding sequences (CDSs), 53 tRNAs genes, 18 rRNAs genes, two intact integrated prophages (38.6 kb, 61 CDSs, G+C content of 38.30% and 55.6 kb, 74 CDSs, G+C content of 38.96%), and one possible incomplete phage (inserted in the noncoding region) between the mutS and mutL genes (13.5 kb, 21 CDSs, G+C content of 36.5%). The web-based tool CRISPRFinder identified one candidate clustered regularly interspaced short palindromic repeat (CRISPR) region (13). The multilocus sequence type (14) was determined to be ST44.

Among known virulence factors the pyrogenic exotoxin speB gene and the superantigen speG and smzZ genes were identified in the chromosome, while mitogenic factors (mf2 and mf3), streptodornase (sda), and streptokinase A (ska) were not found. Interestingly, further analysis identified a single mutation in ropB coding for the stand-alone regulator RopB, resulting in a premature stop codon. This null allele may be associated with an absence of SpeB activity or, as described recently, may incur an overall fitness cost for GAS, preventing its fixation in the population (15).

The annotated sequence of this rare GAS emm66 genotype and newly emerged clone provides a genetic basis for comparison in order to enhance the understanding of clone emergence and hopefully control measures and vaccine design strategies.

**Nucleotide sequence accession number.** The complete genome sequence of strain STAB13021 has been deposited in the NCBI GenBank under the accession number CP014278, as part of Bioproject PRJNA310843.

**Funding Information**

This work was funded by University Rennes 1.

**References**

1. Sitkiewicz I, Green NM, Guo N, Bongiovanni AM, Witkin SS, Musser JM. 2009. Transcriptome adaptation of group B *Streptococcus* to growth in human amniotic fluid. PLoS One 4:e6114. http://dx.doi.org/10.1371/journal.pone.0006114.
2. Cole JN, Barnett TC, Nizet V, Walker MJ. 2011. Molecular insight into invasive group A streptococcal disease. Nat Rev Microbiol 9:724–736. http://dx.doi.org/10.1038/nrmicro2648.
3. Maruyama F, Watanabe T, Nakagawa I. 2016. *Streptococcus pyogenes* pyogenes genomics. In Ferretti JJ, Stevens DL, Fischetti VA (ed), *Streptococcus pyogenes*: basic biology to clinical Manifestations. University of Oklahoma Health Sciences Center, Oklahoma City, OK.
4. Fischetti VA. 1989. Streptococal M protein: molecular design and biological behavior. Clin Microbiol Rev 2:285–314.
5. Beall B, Facklam R, Thompson T. 1996. Sequencing emm-specific PCR products for routine and accurate typing of group A streptococci. J Clin Microbiol 34:953–958.

6. Tartof SY, Farrimond F, de Matos JA, Reis JN, Ramos RT, Andrade AN, dos Reis MG, Riley LW. 2011. Inverse association between Lancefield group G Streptococcus colonization and sore throat in slum and nonslum settings in Brazil. J Clin Microbiol 49:409–412. http://dx.doi.org/10.1128/JCM.02095-10.

7. Krucsó B, Gacs M, Libisch B, Hunyadi ZV, Molnár K, Füzi M, Pászti J. 2007. Molecular characterisation of invasive Streptococcus pyogenes isolates from Hungary obtained in 2004 and 2005. Eur J Clin Microbiol Infect Dis 26:807–811. http://dx.doi.org/10.1007/s10096-007-0359-4.

8. Menon T, Whatmore AM, Srivani S, Kumar MP, Anbumani N, Rajaji S. 2001. EMM types of Streptococcus pyogenes in Chennai. Indian J Med Microbiol 19:161–162.

9. Soriano N, Vincent P, Auger G, Cariou ME, Moullec S, Lagente V, Ygout JF, Kayal S, Faili A. 2015. Full-length genome sequence of type M1emm28 group A Streptococcus pyogenes strain STAB1101, isolated from clustered cases in Brittany. Genome Announc 3(1):e01459-14. http://dx.doi.org/10.1128/genomeA.01459-14.

10. De Andrade Barboza S, Meygret A, Vincent P, Moullec S, Soriano N, Lagente V, Minet J, Kayal S, Faili A. 2015. Complete genome sequence of noninvasive Streptococcus pyogenes M1emm28 strain STAB10015, isolated from a child with perianal dermatitis in French Brittany. Genome Announc 3(4):e00806-15. http://dx.doi.org/10.1128/genomeA.00806-15.

11. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich G, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. http://dx.doi.org/10.1186/1471-2164-9-75.

12. Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. Nucleic Acids Res 39:W347–W352. http://dx.doi.org/10.1093/nar/gkr485.

13. Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a web tool to identify clustered regularly interspaced short palindromic repeats. Nucleic Acids Res 35:W52–W57. http://dx.doi.org/10.1093/nar/gkm360.

14. Enright MC, Spratt BG, Kalia A, Cross JH, Bessen DE. 2001. Multilocus sequence typing of Streptococcus pyogenes and the relationships between emm type and clone. Infect Immun 69:2416–2427. http://dx.doi.org/10.1128/IAI.69.4.2416-2427.2001.

15. Fríaes A, Pato C, Melo-Cristino J, Ramirez M. 2015. Consequences of the variability of the CovRS and RopB regulators among Streptococcus pyogenes causing human infections. Sci Rep 5:12057. http://dx.doi.org/10.1038/srep12057.