Article

The Efficacy of PD-1/PD-L1 Inhibitors in Patients with Liver Metastasis of Non-Small Cell Lung Cancer: A Real-World Study

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Simple Summary: The liver is a common metastatic site of non-small cell lung cancer (NSCLC) and is associated with a poor prognosis. Immune checkpoint inhibitors (ICIs), represented by programmed death-1 (PD-1)/programmed death ligand-1 (PD-L1) inhibitors, have significantly improved efficacy in patients with advanced NSCLC, but the efficacy in patients with NSCLC and liver metastases remains controversial. We aimed to evaluate the efficacy of PD-1/PD-L1 inhibitors in patients with NSCLC and liver metastases in the real-world. In this study, we illustrated that PD-1/PD-L1 inhibitors are effective in NSCLC patients with liver metastases but were inferiorly effective in patients without liver metastases. In addition, PD-L1 expression and CD8+ T cell infiltration may be potential biomarkers for PD-1/PD-L1 inhibitor therapy in NSCLC patients with liver metastases.

Abstract: Background: A controversy exists regarding the efficacy of programmed death-1 (PD-1)/programmed death ligand-1 (PD-L1) inhibitors for patients with non-small cell lung cancer (NSCLC) and liver metastases. Our study retrospectively evaluated the efficacy of PD-1/PD-L1 inhibitors in NSCLC patients with liver metastases. Methods: This retrospective study included 1627 lung cancer patients who received immunotherapy. Among 648 patients who had advanced NSCLC and received PD-1/PD-L1 inhibitors, 61 had liver metastases and 587 did not have. We analyzed patient characteristics, progression-free survival (PFS) and overall survival (OS). An exploratory analysis of biomarkers including CD4, CD8 and CD68 for efficacy in patients with liver metastases was also performed. Results: In liver metastasis patients receiving PD-1/PD-L1 inhibitors, the objective response rate (ORR) was 29.5%, the disease control rate (DCR) was 72.1%, PFS was 6.4 months and OS was 15.2 months, which were all worse than those of patients without liver metastases (ORR: 35.8%; DCR: 81.8%; PFS: 7.9 months, \(p = 0.001\); OS: 20.6 months, \(p = 0.008\)). When compared to non-liver lesions, the ORR (26.2 vs. 39.3%) and DCR (75.4 vs. 88.5%) of liver lesions were lower. During the analysis of PD-L1 expression, 27 PD-L1-positive patients had a longer PFS than 21 patients in the negative group (\(p = 0.012\)). Being PD-L1 positive was the independent prognostic indicators for PFS (\(p = 0.006\)). Additionally, the PD-L1 and CD8 dual-positive group responded favorably to PD-1/PD-L1 inhibitors. Conclusions: PD-1/PD-L1 inhibitors are effective in liver metastasis–NSCLC patients. However, the efficacy is inferior when compared to those of patients without liver metastases. In NSCLC patients with liver metastases, PD-L1 expression and CD8+ T cell infiltration can predict the response of PD-1/PD-L1-directed immunotherapy.

Keywords: liver metastasis; non-small cell lung cancer; PD-L1; PD-1/PD-L1 inhibitor; CD8+ T cell
1. Introduction

Approximately 15–20% of patients with metastatic non-small cell lung cancer (NSCLC) present with liver metastasis, and liver metastasis correlates with an inferior prognosis [1,2]. Throughout the chemotherapy era, the median overall survival (OS) in patients with NSCLC and liver metastasis was fewer than ten months [3–5]. Metastasis to the liver is an independent negative predictor for NSCLC patients who receive chemotherapy [6,7].

For advanced NSCLC patients, immune checkpoint inhibitors (ICI) such as programmed cell death-1 (PD-1)/programmed cell death ligand-1 (PD-L1) inhibitors have been established as first-line therapy regimen [8,9]. Owing to the complicated microenvironment of metastatic liver lesions, the efficacy of ICIs in NSCLC patients with liver metastases has recently attracted much attention. Recently, preclinical mouse models discovered that liver metastasis drained activated CD8+ T cells from the systemic circulation and triggered CD8+ T cell death, resulting in a systemic immunological desert, implying that immunotherapy effectiveness for NSCLC patients with liver metastasis is diminished [10]. In clinical situations, it still remains controversial whether ICIs are effective in NSCLC patients with liver metastasis. A subgroup analysis in several phase III clinical studies has shown that NSCLC patients with liver metastases can benefit from ICI therapy [3,4]. In addition, other studies have reported that the efficacy of ICIs in NSCLC patients with liver metastases is similar to that in patients without liver metastases [11,12]. Conversely, several studies have drawn different conclusions, suggesting that ICI has limited efficiency in NSCLC patients who have liver metastasis. Data in KEYNOTE-001 showed that progression-free survival (PFS) from NSCLC patients with liver metastases after treatment with pembrolizumab was significantly shorter (p < 0.05), and the objective response rate (ORR) was lower (28.6 vs. 56.7%) than patients without liver metastases [13]. Several retrospective studies have drawn similar conclusions [14–16].

On the basis of the available evidence from subgroup analyses of clinical studies and retrospective studies, the efficacy of anti-PD-1/PD-L1 therapy in patients with NSCLC and liver metastases remains controversial. Therefore, we conducted this retrospective study in order to evaluate the effectiveness for PD-1/PD-L1 inhibitors on liver metastasis-NSCLC patients. We further enrolled NSCLC patients without liver metastases who received ICI therapy during the same period to compare their efficiency. We also explored potential prognostic factors. In addition, evaluation was carried out on the expression of PD-L1 and the infiltration of immune cells (IC).

2. Materials and Methods

2.1. Patients

A total of 1627 patients who had lung cancer and who received PD-1/PD-L1 inhibitors at Zhejiang Cancer Hospital from September 2017 to December 2020 were screened. Of these patients, 61 patients with NSCLC and liver metastasis were enrolled. Simultaneously, 587 NSCLC patients without liver metastasis were selected as comparators. The major inclusion criteria were: Eastern Corporation Oncology Group performance status (ECOG PS) of 0–2, cytologically or histologically confirmed stage IV NSCLC, at least one measurable lesion, treatment with PD-1/PD-L1 inhibitors, and complete survival data. The major exclusion criteria were: small-cell lung cancer, sensitive epidermal growth factor receptor (EGFR) mutations, anaplastic lymphoma kinase (ALK) rearrangements or ROS proto-oncogene 1 receptor tyrosine kinase (ROS1) fusions, and incomplete follow-up data. According to the Response Evaluation Criteria in Solid Tumors (RECIST) version 1.1, the response was evaluated radiographically [17]. The PFS was calculated as the time between starting PD-1/PD-L1 inhibitors and the progression of disease or the incidence of death owing to different causes. The OS was defined as the period from the initiation of PD-1/PD-L1 inhibitor treatment until death due to any cause or the latest follow-up date. These enrolled patients were followed up until June 2021. This study was approved by the Zhejiang Cancer Hospital’s Ethics Committee.
2.2. Immunohistochemistry (IHC)

We obtained tumor tissues from different origins, including the lung, liver, lymph nodes, soft tissue and bones. According to manufacturer’s recommendations, IHC was conducted using 4–5 µm formalin-fixed and paraffin-embedded (FFPE) sections. The PD-L1 clone 28-8 pharmDx kit and the Dako Automated Link 48 platform were used to measure the expression of PD-L1. CD4 (clone B468A, diluted at 1:200, Santa Cruz, Texas, USA) and CD8 (clone 144B, diluted at 1:100, Abcam, Cambridge, UK) expression in T cells, and CD68 (clone PG-M1, diluted at 1:600, Abcam, Cambridge, UK) expression on macrophages were also assessed. Two pathologists independently scored the stained tissues, and the clinical parameters were kept confidential from these two pathologists.

The tumor proportion score (TPS) was used to assess PD-L1 expression. The percentage of tumor cells stained with partial or complete membranes was the definition of TPS. The positive expression of PD-L1 was considered as TPS ≥ 1%. Among all nucleated cells in the tumor mesenchyme, the proportion of positive cells for CD4, CD8, and CD68 expression was assessed. Positivity on lymphocytes was set at 5, 5, and 20% for CD4, CD8, and CD68, respectively. Based on PD-L1 expression and CD8+ T cell infiltration, the tumor microenvironment was divided into three subgroups (both negative, both positive, or single-positive).

2.3. Statistical Analyses

SPSS (version 25.0) and GraphPad Prism (version 9.2.0) were used to perform the statistical analysis. The distribution of clinical features was assessed through the Chi-squared test. The responses were graded based on Response Evaluation Criteria in Solid Tumors (RECIST) version 1.1 [17]. Kaplan–Meier curves were applied for survival analysis of PFS and OS in various groups. Univariate and multivariate Cox regressions were used for prognostic analysis to obtain the adjusted hazard ratios (HR) and 95% confidence intervals (95% CI). The multivariate Cox regression analysis obtained variables with a p < 0.10 in the univariate analysis. Statistical significance was set of p < 0.05, and two-tailed p values were calculated in all reports.

3. Results

3.1. Patient Characteristics

Sixty-one NSCLC patients who had liver metastasis were enrolled in this study (Figure A1). They were all treated with anti-PD-1/PD-L1 therapy. The median age of these patients was 63 years (range, 34–76 years). Among them, 59.0% (n = 36) were <65 years old, 90.2% (n = 55) were male, 86.9% (n = 53) had an ECOG PS of 0–1, 82.0% (n = 50) were current or former smokers, and 11.5% (n = 7) had brain metastasis. The histology of the tumors was 27 (44.3%) adenocarcinomas, 29 (47.5%) squamous cell carcinomas, and 5 (8.2%) other NSCLCs. In 55.7% (n = 34) of the patients, PD-1/PD-L1 inhibitors were used as a first-line therapy, and 34.4% (n = 21) of the patients received them as monotherapy. Meanwhile, the control group consisted of 587 metastatic NSCLC patients who were free of liver metastasis treated with PD-1/PD-L1 inhibitors. The majority of patients (60/61, 98.4%) were treated with PD-1 inhibitors, of whom 14 (23.0%) were treated with Pembrolizumab, 8 (13.1%) with Nivolumab, 11 (18.0%) with Tislelizumab, 5 (8.2%) with Toripalimab, 11 (18.0%) with Sintilimab, and 11 (18.0%) with Camrelizumab; only one patient (1.6%) was treated with a PD-L1 inhibitor (Atezolizumab). Table 1 shows the baseline characteristics of these patients according to the liver metastasis status. Patients who had liver metastasis experienced a considerably lower positive rate of PD-L1 expression (p = 0.012). There was no statistically significant difference between the liver metastasis and non-liver metastasis groups in terms of age, gender, ECOG PS, smoking, brain metastasis, histological type, lines of ICI and ICI treatment regimens. The ICI treatment regimens included ICI monotherapy and ICI combined therapy (combined chemotherapy or combined anti-angiogenesis therapy).
Table 1. Baseline characteristics of included NSCLC patients according to the liver metastasis status.

|                        | Total (n = 648) | Liver Metastasis (n = 61) | No Live Metastasis (n = 587) | p-Value |
|------------------------|-----------------|---------------------------|-----------------------------|---------|
| Median age (range), years | 63 (32–81) | 63 (34–76) | 63 (32–81) | 0.909 |
|                        | <65 378 (58.3) | 36 (59.0) | 342 (58.3) | 0.119 |
| ≥65 270 (41.6) | 25 (41.0) | 245 (41.7) |
| Gender, n (%)          |                 |                          |                             |         |
| Male                   | 538 (83.0) | 55 (90.2) | 483 (82.3) | 0.085 |
| Female                 | 110 (17.0) | 6 (9.8) | 104 (17.7) |         |
| ECOG PS, n (%)         |                 |                          |                             |         |
| 0–1                    | 599 (92.4) | 53 (86.9) | 546 (93.0) | 0.012 |
| ≥2                     | 49 (7.6) | 8 (13.1) | 41 (7.0) |         |
| Smoking status, n (%)  |                 |                          |                             |         |
| Never                  | 179 (27.6) | 11 (18.0) | 168 (28.6) | 0.078 |
| Current or former      | 469 (72.4) | 50 (82.0) | 419 (71.4) |         |
| Histology, n (%)       |                 |                          |                             |         |
| Adenocarcinoma         | 329 (50.8) | 27 (44.3) | 300 (51.1) | 0.868 |
| Squamous               | 270 (44.7) | 29 (47.5) | 243 (41.4) |         |
| Others                 | 49 (7.6) | 5 (8.2) | 44 (7.5) |         |
| PD-L1 TPS, n (%)       |                 |                          |                             |         |
| Unknown                | 458 (70.7) | 13 (21.3) | 445 (75.8) |         |
| Know                   | 190 (29.3) | 48 (78.7) | 142 (24.2) |         |
| Negative               | 56 (29.5) | 21 (43.8) | 35 (24.6) | 0.012 |
| Positive               | 134 (70.5) | 27 (56.3) | 107 (75.4) |         |
| Brain metastases, n (%)|                 |                          |                             |         |
| Yes                    | 91 (14.0) | 7 (11.5) | 84 (14.3) | 0.544 |
| No                     | 557 (86.0) | 54 (88.5) | 503 (85.7) |         |
| Lines of ICI therapy, n (%) | 316 (48.8) | 34 (55.7) | 282 (48.0) | 0.252 |
| ≥2                     | 332 (51.2) | 27 (44.3) | 305 (52.0) |         |
| ICI treatment regimen, n (%) | 250 (38.6) | 21 (34.4) | 229 (39.0) | 0.484 |
| ICI monotherapy        | 398 (61.4) | 40 (65.6) | 358 (61.0) |         |

Abbreviations: ECOG PS, Eastern Corporation Oncology Group performance status; PD-L1, programmed cell death ligand-1; TPS, tumor proportion score; ICI, Immune checkpoint inhibitor.

3.2. Efficacy of PD-1/PD-L1 Inhibitors

Among 61 patients who had NSCLC and liver metastasis, none achieved a complete response (CR), 29.5% (n = 18) showed a partial response (PR), 42.6% (n = 26) showed stable disease (SD), and 27.9% (n = 17) had progressive disease (PD) (Table 2a). In these patients, the ORR was 29.5%, the disease control rate (DCR) was 72.1% (Table 2a), PFS was 6.4 months (Figure 1a) and OS was 15.2 months (Figure 1b). Of the non-liver metastasis patients, 0.3% (n = 2) achieved CR, 35.4% (n = 208) achieved PR, 46.0% (n = 270) showed SD, and 18.2% (n = 107) had PD. The ORR and DCR of these patients were 35.8 and 81.8%, respectively (Table 2a), the PFS was 7.9 months (Figure 1c) and OS was 20.6 months (Figure 1d). Liver metastasis patients had lower ORR and DCR, and a significantly shorter PFS (p = 0.001) and OS (p = 0.008).

To further explore whether the effectiveness in PD-1/PD-L1 inhibitors on liver metastatic lesions was distinctive from other lesions, we evaluated the response rate of liver and non-liver lesions, and we found that the ORR (26.2 vs. 39.3%) and DCR (75.4 vs. 88.5%) were lower for liver lesions (Table 2b).
Table 2. Immunotherapy efficacy of NSCLC patients. (a) Liver metastases group and non-liver metastasis group. (b) Liver and non-liver lesions in NSCLC patients with liver metastasis.

|                  | Liver Metastasis (n = 61) | Non-Liver Metastasis (n = 587) |
|------------------|---------------------------|-------------------------------|
| CR, n (%)        | 0 (0)                     | 2 (0.3)                       |
| PR, n (%)        | 18 (29.5)                 | 208 (35.4)                    |
| SD, n (%)        | 26 (42.6)                 | 270 (46.0)                    |
| PD, n (%)        | 17 (27.9)                 | 107 (18.2)                    |
| ORR, n (%)       | 18 (29.5)                 | 210 (35.8)                    |
| DCR, n (%)       | 44 (72.1)                 | 480 (81.8)                    |

|                  | Liver lesions | Non-Liver Lesions |
|------------------|---------------|-------------------|
| CR, n (%)        | 0 (0)         | 0 (0)             |
| PR, n (%)        | 16 (26.2)     | 24 (39.3)         |
| SD, n (%)        | 30 (49.2)     | 30 (49.2)         |
| PD, n (%)        | 15 (24.6)     | 7 (11.5)          |
| ORR, n (%)       | 16 (26.2)     | 24 (39.3)         |
| DCR, n (%)       | 46 (75.4)     | 54 (88.5)         |

Abbreviations: CR, complete response; PR, partial response; SD, stable disease; PD, progressive disease; ORR, objective response rate; DCR, disease control rate.

Figure 1. Kaplan–Meier analysis for progression-free survival (PFS) and overall survival (OS) in non-small cell lung cancer (NSCLC) patients treated with PD-1/PD-L1 inhibitors. (a) The median PFS of NSCLC patients with liver metastases was 6.4 months. (b) The median OS of NSCLC patients with liver metastases was 15.2 months. (c) The median PFS of NSCLC patients without liver metastases was 7.9 months, and that of patients with liver metastases was 6.4 months. (d) The median OS of NSCLC patients without liver metastases was 20.6 months, and that of patients with liver metastases was 15.2 months.
3.3. Exploratory Analysis of the Expression of Tumor Immune Microenvironment-Related Markers

In our study, 48 patients with NSCLC with liver metastasis were evaluated for PD-L1 expression (Figure A2a,b were representative images of positive and negative PD-L1 staining) and 42 patients with NSCLC with liver metastasis were tested for CD4, CD8 and CD68 infiltrates. We discovered that CD8+ T cell infiltration was considerably lower in liver lesions than that in non-liver lesions (70.0 vs. 93.8%, \( p = 0.043 \)). Among the PD-L1 expression (60.0 vs. 55.3%, \( p = 0.788 \)), CD4+ T cell infiltration (70.0 vs. 56.2%, \( p = 0.439 \)), and CD68+ T cell infiltration (90.0 vs. 78.1%, \( p = 0.404 \)), there was no statistically significant difference between liver lesions and non-liver lesions (Table 3).

### Table 3. PD-L1, CD4, CD8, CD68 expression of liver and non-liver lesions in NSCLC patients with liver metastasis.

|                | Liver Lesions | Non-Liver Lesions | \( p \)-Value |
|----------------|---------------|-------------------|--------------|
| **PD-L1**      |               |                   |              |
| Negative       | 4 (40.0)      | 17 (44.7)         | 0.788        |
| Positive       | 6 (60.0)      | 21 (55.3)         |              |
| **CD4**        |               |                   |              |
| Negative       | 3 (30.0)      | 5 (15.6)          | 0.312        |
| Positive       | 7 (70.0)      | 27 (84.4)         |              |
| **CD8**        |               |                   |              |
| Negative       | 3 (30.0)      | 2 (6.3)           | 0.043        |
| Positive       | 7 (70.0)      | 30 (93.8)         |              |
| **CD68**       |               |                   |              |
| Negative       | 1 (10.0)      | 7 (21.9)          | 0.404        |
| Positive       | 9 (90.0)      | 25 (78.1)         |              |

Abbreviations: PD-L1, programmed cell death ligand-1; TPS, tumor proportion score.

3.4. Correlation Analysis between Biomarkers of Tumor Immune Microenvironment and Efficacy

In liver metastasis patients, the PFS in the PD-L1-positive expression group was statistically substantially prolonged when compared to the negative group (8.2 vs. 4.4 months, \( p = 0.012 \)) (Figure 2a), while the OS was similar (\( p = 0.587 \)) (Figure 2b). For the tumor immune microenvironment subgroups, the PD-L1 and CD8 double-positive group had the longest PFS (8.7 months, \( p = 0.004 \)) (Figure 2e) but no difference in OS among three groups (\( p = 0.684 \)) (Figure 2f).

Between the CD8-positive and -negative groups, no statistical difference was observed in PFS (6.6 vs. 2.7 months, \( p = 0.594 \)) and OS (15.2 months vs. undefined, \( p = 0.427 \)) (Figure 2c,d). In addition, the PFS and OS in the CD4 positive group were also similar to the negative cohort (PFS: \( p = 0.852 \); OS: \( p = 0.739 \)), as well as for CD68 (PFS: \( p = 0.552 \); OS: \( p = 0.865 \)).

3.5. Prognostic Analysis of Patients with Liver Metastases

Univariate analysis indicated that brain metastases were in related to a poor PFS, while PD-L1-positive expression was in association with a superior PFS (Table 4a). Multivariate analysis found that brain metastases (HR: 2.86, 95%CI: 1.04–7.84, \( p = 0.041 \)) and PD-L1 expression (HR: 0.39, 95%CI: 0.20–0.76, \( p = 0.006 \)) were independent prognostic factors (Table 4a). No factors were associated with OS in the univariate analysis (Table 4b).
Figure 2. Kaplan–Meier analysis for progression-free survival (PFS) and overall survival (OS) in non-small cell lung cancer (NSCLC) patients with liver metastases to PD-L1, CD4+, CD8+ and CD68+ expression. (a) The median PFS of PD-L1-negative cancer was 4.4 months, and that of PD-L1-positive cancer was 8.2 months. (b) The median OS of PD-L1-negative cancer was 15.2 months, and that of PD-L1-positive cancer was 18.3 months. (c) The median PFS of CD8-negative cancer was 2.7 months, and that of CD8-positive cancer was 6.6 months. (d) The median OS of CD8-negative cancer was undefined, and that of CD8-positive cancer was 15.2 months. (e) The median PFS of PD-L1/CD8-dual-negative cancer was 2.7 months, that of PD-L1/CD8-dual-positive cancer was 8.7 months, and that of PD-L1/CD8-single-positive cancer was 4.3 months. (f) The median OS of PD-L1/CD8-dual-negative cancer was 21.2 months, that of PD-L1/CD8-dual-positive cancer was 12.2 months, and that of PD-L1/CD8-single-positive cancer was 15.2 months.

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Table 4. Univariate and multivariate analysis of progression-free survival and overall survival in relation to the baseline characteristics of included NSCLC patients with liver metastases. (a) Progression-Free Survival. (b) Overall Survival.

| Factor | Univariate Analysis | Multivariate Analysis |
|--------|---------------------|-----------------------|
|        | HR  | 95% CI   | p-Value | HR  | 95% CI   | p-Value |
| Age (≥65; <65) | 1.24 | 0.70–2.19 | 0.455 |
| Gender (female; male) | 1.04 | 0.37–2.91 | 0.935 |
| ECOG PS (2; 0–1) | 0.63 | 0.26–1.49 | 0.289 |
| Smoking status (current or former; never) | 1.17 | 0.54–2.50 | 0.695 |
| Histology | | | |
| Adenocarcinoma | | | |
| Squamous | 0.67 | 0.37–1.20 | 0.175 |
| Others | 1.17 | 0.47–2.91 | 0.742 |
| PD-L1 TPS (positive; negative) | 0.45 | 0.24–0.86 | 0.015 |
| CD8 (positive; negative) | 0.77 | 0.29–2.01 | 0.595 |
| Brain metastases (Yes; No) | 2.18 | 0.97–4.93 | 0.061 |
| Treatment line (≥2; 1) | 1.10 | 0.64–1.91 | 0.731 |
| Treatment regimen (ICI combined therapy; ICIs monotherapy) | 0.78 | 0.44–1.37 | 0.384 |

(b)

| Factor | Univariate Analysis |
|--------|---------------------|
|        | HR  | 95% CI   | p-Value |
| Age (≥65; <65) | 1.38 | 0.69–2.74 | 0.359 |
| Gender (female; male) | 0.75 | 0.23–2.50 | 0.643 |
| ECOG PS (2; 0–1) | 1.16 | 0.44–3.01 | 0.768 |
| Smoking status (current or former; never) | 1.08 | 0.42–2.82 | 0.871 |
| Histology | | | |
| Adenocarcinoma | | | |
| Squamous | 0.56 | 0.28–1.15 | 0.113 |
| Others | 0.54 | 0.12–2.39 | 0.416 |
| PD-L1 TPS (positive; negative) | 0.80 | 0.36–1.76 | 0.579 |
| CD8 (positive; negative) | 1.79 | 0.42–7.75 | 0.434 |
| Brain metastases (Yes; No) | 1.81 | 0.70–4.71 | 0.223 |
| Treatment line (≥2; 1) | 0.84 | 0.42–1.67 | 0.614 |
| Treatment regimen (ICI combined therapy; ICIs monotherapy) | 0.69 | 0.34–1.39 | 0.295 |

Abbreviations: ECOG PS, Eastern Cooperative Oncology Group performance status; PD-L1, programmed cell death ligand-1; TPS, tumor proportion score; ICI, Immune checkpoint inhibitor.

4. Discussion

The liver is an immune-tolerant organ and the liver microenvironment is highly immunosuppressive [10,18]. Hepatocytes, Kupffer cells, liver sinusoidal endothelial cells, hepatic stellate cells and liver dendritic cells can recruit T cells through antigen presentation, which also leads to PD-L1 expression and binding to PD-1 on T cells, which preferentially leads to immune tolerance [19,20]. Therefore, immunotherapy may not be effective for liver metastatic lesions. Several translational studies have revealed that liver metastasis mediates immunotherapy resistance in liver lesions and hampers systemic antitumor immunity. Meng Qiao et al. carried out IHC on the tumor tissues of liver metastasis–NSCLC patients, and IHC revealed that liver metastasis–NSCLC patients had a lower percentage of PD-L1+CD8+ ICs in comparison to non-liver metastasis patients (0 vs. 30.8%, p = 0.088) [13]. Furthermore, it was reported that liver metastasis siphoned activated CD8+ T cells, which were derived from the systemic circulation. Subsequently, monocyte-derived macrophages induced the apoptosis of CD8+ T cells, creating a systemic immune desert in preclinical mouse models [10]. In addition, Lee et al. found that liver metastases induce the accumulation of Tregs and suppressive monocytes, leading to impaired systemic antitumor immunity [21]. Accordingly, this study intended to explore the efficacy of PD-1/PD-L1 inhibitors in liver metastasis–NSCLC patients and preliminarily investigate the specific immune microenvironment of these patients.

Marina C et al. [22] found that first-line chemotherapy had the ORR of 14.3% only in non-squamous NSCLC patients with liver metastasis. In comparison to the effective-
ness of chemotherapy, we discovered that anti-PD-1/PD-L1 therapy improves outcomes among liver metastasis–NSCLC patients. The results of several recent large phase III clinical studies have shown that PD-1/PD-L1 inhibitors are effective in liver-metastatic NSCLC patients [3–5]. Our study demonstrated the advantages of PD-1/PD-L1 inhibitors for liver metastasis–NSCLC patients in the real-world situation. In addition, liver metastasis patients who were treated with PD-1/PD-L1 inhibitors had worse treatment outcomes (PFS, OS, ORR and DCR) than non-liver metastasis patients. This was subsequently confirmed in several retrospective studies [23,24]. Tumeh et al. [13] showed that when receiving pembrolizumab, liver metastasis–NSCLC patients had a significantly shorter PFS (1.8 vs. 4.0 months, \( p = 0.0094 \)) and lower ORR (28.6 vs. 56.7%) than non-liver metastasis patients. Kitadai et al. [15] carried out a real-world retrospective study. A total of 215 advanced NSCLC patients treated with ICI monotherapy were admitted, including 41 (19.1%) with liver metastasis. This study reached similar conclusions that the patients with liver metastasis had worse OS (\( p < 0.001 \)) and PFS (\( p < 0.001 \)), and liver metastasis was an independent poor prognostic factor on the multifactorial analysis. One possible explanation for the limited efficacy in NSCLC patients with liver metastasis treated with PD-1/PD-L1 inhibitors is the immune microenvironment, which is specific to liver metastasis as mentioned above, and the suppression of systemic immunity by liver metastasis. Previous studies have confirmed that liver metastasis is a poor prognostic factor in NSCLC patients, and was associated with shorter PFS and OS [7,25,26]. Therefore, the inferior survival of NSCLC patients with liver metastasis treated with PD-1/PD-L1 inhibitors may be related to the characteristic of poor prognosis in liver metastasis.

Other studies did not find significant difference in ICI efficacy between NSCLC patients with liver metastasis and without. A real-world study which included 1470 patients but only contained 11 patients with liver metastasis found that PFS (\( p = 0.955 \)) and OS (\( p = 0.968 \)) were similar in liver metastasis patients or non-liver metastasis patients in treating with ICIs [11]. Immunochemotherapy was equally effective in lung cancer patients with or without liver metastasis (the HR of PFS: 1.11, 95%CI: 0.92–1.34; the HR of OS: 1.03, 95%CI: 0.80–1.35) from a meta-analysis [12]. However, the meta-analysis included NSCLC and small cell lung cancer and lacked direct PFS or OS data. To our knowledge, this is the first largest scale retrospective study to evaluate the efficacy of PD-1/PD-L1 inhibitors in NSCLC patients with liver metastasis in the real-world. In addition, patients with positive driver genes in our study were excluded to avoid weakening the effectiveness of ICIs.

We further performed a comparison of the response among liver and non-liver lesions from liver metastasis patients who received anti-PD-1/PD-L1 therapy. It was observed that both ORR and DCR in liver lesions were less. The result was in agreement with the conclusions from a recent study, which was done to assess the efficacy among 761 lesions (58 liver metastases) in 214 NSCLC patients treated with PD-1 inhibitor monotherapy and that indicated that liver metastatic lesions showed the worst response [27].

Furthermore, we found that CD8+ T cell infiltration in liver lesions was significantly decreased when compared to non-liver lesions. CD8+ T cells are often considered the important effector cell population for ICI treatment [28,29]. With sufficient cytotoxic lymphocyte infiltration in the tumor microenvironment, blockade of the PD-1/PD-L1 pathway by ICIs can activate effective anti-tumor immunity [28,30]. Therefore, the negative efficacy of PD-1/PD-L1 inhibitors on liver lesions may be due to the lack of CD8+ T cells. As previously mentioned, a minimal proportion of PD-L1+CD8+ ICs in liver metastasis–NSCLC tumors was recorded [13]. Populations who had the limited efficiency from PD-1/PD-L1 inhibitors may owe this to the microenvironment of these populations, and be immunoinflammatory deficient [13,30]. In our data, patients in the PD-L1 and CD8 double-positive expression group had notably longer PFS than those in the single-positive or double-negative expression group. The results suggested that patients whose immune microenvironment exhibits an immunoinflammatory phenotype have better outcomes.

PD-L1 expression is related to an excellent response to anti-PD-1 therapy in NSCLC patients [29,31]. For the contained liver metastasis patients, these who positively expressed
PD-L1 had obviously longer PFS. Furthermore, univariate and multifactorial analyses showed that PD-L1 positivity was a significant predictor of PFS. This result indicated that the expression of PD-L1 may play a potentially effective predictor role among liver metastasis-NSCLC patients. This is consistent with the current belief in which PD-L1 could act as a clinically valid prognostic marker for ICIs [32–34].

It is supported by previous studies in which it was found that the liver is rich in blood vessels and overexpressed vascular endothelial growth factor (VEGF), which has an immunomodulatory effect [35–38]. In liver metastasis, anti-angiogenesis therapy such as bevacizumab can block the effect of VEGFR [35–37,39]. Thus, anti-angiogenesis therapy may have a positive effect on liver metastasis. A subgroup analysis of patients with liver metastases in Impower150, which is a phase III clinical study, found that the patients receiving atezolizumab combined with chemotherapy plus bevacizumab achieved promising efficacy [5]. Another study about the bevacizumab in NSCLC patients found that bevacizumab significantly improved PFS and OS, and that bevacizumab was more effective in patients with liver metastases than in those with non-liver metastases [40]. However, due to the limited sample among patients with liver metastases in our study, anti-angiogenesis plus PD-1/PD-L1 inhibitors did not display a beneficial advantage (Tables A1 and A2). Two liver metastasis patients who received PD-1/PD-L1 inhibitors in combination with platinum-containing chemotherapy developed PD. Notably, after received second-line anti-angiogenesis therapy plus chemotherapy, they both had PR (Figure A3). One patient was alive until the follow-up date. The above performance implies that for NSCLC patients with liver metastases, anti-angiogenesis treatment may serve as a promising scheme when they have failed with PD-1/PD-L1 inhibitors; however, further confirmation of this finding in larger scale studies is required.

Data from previous studies have confirmed the poor prognosis of NSCLC patients with liver metastasis [3–5]; the potential interventions to improve clinical outcomes are follows. The combination of local treatments such as radiofrequency ablation can be used in patients with liver oligometrics (either a single lesion smaller than 5 cm or as many as three lesions smaller than 3 cm each) [41,42]. PD-1/PD-L1 inhibitors have revealed promising activity in NSCLC patients with liver metastases. The results of the subgroup analysis of IMpower150 showed that the addition of bevacizumab was effective in NSCLC patients with liver metastases, which may be a promising treatment strategy in the future [5,43]. However, in our study, due to the small sample size of liver metastasis patients, expanding the sample size is necessary. Besides, because of the special immune microenvironment of the liver [20], it is essential to seek new immune microenvironment-related therapeutic targets to develop novel therapeutic drugs.

Several limitations in our study. First of all, the sample size of liver metastasis patients was small. Secondly, our study was single-centered and retrospective; besides, we performed PD-L1 testing in NSCLC patients with liver metastases for whom pathological tissue was available, which may account for the difference in PD-L1 expression between patients with and without liver metastases. Therefore, our findings have to be verified in future prospective studies. Finally, adverse events data were not reported because of lack of complete records.

5. Conclusions

Our findings imply that PD-1/PD-L1 inhibitors enhances effectiveness in NSCLC patients who have liver metastases, but the benefit is inferior to those in patients without liver metastasis. Furthermore, PD-L1 expression and CD8+ T cell infiltration might act as biomarkers to predict the efficacy of PD-1/PD-L1 inhibitors in patients with NSCLC and liver metastasis.
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Institutional Review Board Statement: The current study was conducted in accordance with the Declaration of Helsinki and approved by the Zhejiang Cancer Hospital’s Ethics Committee (approval number: IRB-2020-241; date: 10 October 2020).

Informed Consent Statement: Patient consent was waived due to the retrospective nature of this study.

Data Availability Statement: The data can be shared upon request.

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Appendix A

Figure A1. Study flowchart. A total of 1627 patients with lung cancer who underwent immunotherapy were screened. Major exclusion criteria were that patients had non-advanced lung cancer, small cell lung cancer, and EGFR/ALK/ROS1 mutations. 648 patients were finally enrolled, including 61 patients with liver metastasis.
Figure A2. The expression of PD-L1 within non-small cell lung cancer (NSCLC) patients with liver metastasis. (a) Positive expression of PD-L1 (PD-L1 TPS = 40%); (b) negative expression of PD-L1 (PD-L1 TPS < 1%).

Figure A3. The clinical response of two non-small cell lung cancer (NSCLC) patients with liver metastasis. The clinical response to second-line chemotherapy combined with anti-angiogenesis therapy after failure of first-line PD-1/PD-L1 inhibitors combined with chemotherapy in two NSCLC patients (a,b) with liver metastasis.

Appendix B

Table A1. Efficacy of PD-1 inhibitors combined with anti-angiogenesis therapy in NSCLC patients with liver metastasis in first-line.

|                | ICI + Anti-Angiogenesis | Non-Anti-Angiogenesis |
|----------------|-------------------------|-----------------------|
| CR, n (%)      | 0                       | 0                     |
| PR, n (%)      | 0                       | 11 (35.5%)            |
| SD, n (%)      | 3 (100.0%)              | 13 (41.9%)            |
| PD, n (%)      | 0                       | 7 (22.6%)             |
| ORR, n (%)     | 0                       | 11 (35.5%)            |
| DCR, n (%)     | 3 (100.0%)              | 24 (77.4%)            |

Abbreviations: CR, complete response; PR, partial response; SD, stable disease; PD, progressive disease; ORR, objective response rate; DCR, disease control rate; ICI, Immune checkpoint inhibitor.
Table A2. Efficacy of PD-1 inhibitors combined with anti-angiogenesis therapy in NSCLC patients with liver metastasis in ≥2 lines.

|                      | ICI + Anti-Angiogenesis | Non-Anti-Angiogenesis |
|----------------------|-------------------------|-----------------------|
| CR, n (%)            | 0                       | 0                     |
| PR, n (%)            | 0                       | 5 (27.8%)             |
| SD, n (%)            | 6 (66.7%)               | 8 (44.4%)             |
| PD, n (%)            | 3 (33.3%)               | 5 (27.8%)             |
| ORR, n (%)           | 0                       | 5 (27.8%)             |
| DCR, n (%)           | 6 (66.7%)               | 13 (72.2%)            |

Abbreviations: CR, complete response; PR, partial response; SD, stable disease; PD, progressive disease; ORR, objective response rate; DCR, disease control rate; ICI, Immune checkpoint inhibitor.

References
1. Riihimaki, M.; Hemminki, A.; Fallah, M.; Thomesen, H.; Sundquist, K.; Sundquist, J.; Hemminki, K. Metastatic sites and survival in lung cancer. *Lung Cancer* 2014, 86, 78–84. [CrossRef] [PubMed]
2. Quint, L.E.; Tummala, S.; Brisson, L.J.; Francis, I.R.; Kruenick, A.S.; Kazerooni, E.A.; Iannettoni, M.D.; Whyte, R.I.; Orringer, M.B. Distribution of distal metastases from newly diagnosed non-small cell lung cancer. *Ann. Thorac. Surg.* 1996, 62, 246–250. [CrossRef]
3. Gadgeel, S.; Rodriguez-Abreu, D.; Speranza, G.; Esteban, E.; Felip, E.; Domine, M.; Hui, R.; Hochmair, M.J.; Clingan, P.; Powell, S.F.; et al. Updated Analysis From KEYNOTE-189: Pembrolizumab or Placebo Plus Pemetrexed and Platinum for Previously Untreated Metastatic Non-Squamous Non-Small-Cell Lung Cancer. *J. Clin. Oncol.* 2020, 38, 1505–1517. [CrossRef] [PubMed]
4. Vokes, E.E.; Ready, N.; Felip, E.; Horn, L.; Burgio, M.A.; Antonia, S.J.; Gettinger, S.; Holgado, E.; Spigel, D.; et al. Nivolumab versus docetaxel in previously treated advanced non-small-cell lung cancer (CheckMate 017 and CheckMate 057): 3-year update and outcomes in patients with liver metastases. *Ann. Oncol.* 2018, 29, 959–965. [CrossRef]
5. Noğami, N.; Barlesi, F.; Socinski, M.A.; Reck, M.; Thomas, C.A.; Capuzzo, F.; Mok, T.S.K.; Finley, G.; Aerts, J.G.; Orlandi, F.; et al. IMpower150 Final Exploratory Analyses for Atezolizumab Plus Bevacizumab and Chemotherapy in Key NSCLC Patient Subgroups With EGFR Mutations or Metastases in the Liver or Brain. *J. Thorac. Oncol.* 2022, 17, 309–323. [CrossRef] [PubMed]
6. Hoang, T.; Xu, R.; Schiller, J.H.; Bonomi, P.; Johnson, D.H. Clinical model to predict survival in chemonaive patients with advanced non-small-cell lung cancer treated with third-generation chemotherapy regimens based on eastern cooperative oncology group data. *J. Clin. Oncol.* 2005, 23, 175–183. [CrossRef]
7. Hoang, T.; Dahlberg, S.E.; Sandler, A.B.; Brahmer, J.R.; Schiller, J.H.; Johnson, D.H. Prognostic models to predict survival in non-small-cell lung cancer patients treated with first-line paclitaxel and carboplatin with or without bevacizumab. *J. Thorac. Oncol.* 2012, 7, 1361–1368. [CrossRef]
8. Mariamidze, E.; Mezquita, L. ESMO20 YO for YO: Highlights on metastatic NSCLC-Keynote 024 update. *ESMO Open* 2021, 6, 100022. [CrossRef]
9. Garassino, M.C.; Gadgeel, S.; Esteban, E.; Felip, E.; Speranza, G.; Domine, M.; Hochmair, M.J.; Powell, S.; Cheng, S.Y.; Bischoff, H.G.; et al. Patient-reported outcomes following pembrolizumab or placebo plus pemetrexed and platinum in patients with previously untreated, metastatic, non-squamous non-small-cell lung cancer (KEYNOTE-189): A multicentre, double-blind, randomised, placebo-controlled, phase 3 trial. *Lancet Oncol.* 2020, 21, 387–397.
10. Yu, J.; Green, M.D.; Li, S.; Sun, Y.; Journey, S.N.; Choi, J.E.; Rizvi, S.M.; Qin, A.; Waninger, J.J.; Lang, X.; et al. Liver metastasis restrains immunotherapy efficacy via macrophage-mediated T cell elimination. *Nat. Med.* 2021, 27, 152–164. [CrossRef]
11. Choi, M.G.; Choi, C.M.; Lee, D.H.; Kim, S.W.; Yoon, S.; Kim, W.S.; Ji, W.; Lee, J.C. Different prognostic implications of hepatic metastasis according to front-line treatment in non-small cell lung cancer: A real-world retrospective study. *Transl. Lung Cancer Res.* 2021, 10, 2551–2561. [CrossRef]
12. Qin, B.D.; Jiao, X.D.; Liu, J.; Liu, K.; He, X.; Wu, Y.; Ling, Y.; Duan, X.P.; Qin, W.X.; Wang, Z.; et al. The effect of liver metastasis on efficacy of immunotherapy plus chemotherapy in advanced lung cancer. *Crit. Rev. Oncol. Hemat.* 2020, 147, 102893. [CrossRef]
13. Tumeh, P.C.; Hellmann, M.D.; Hamid, O.; Tsai, K.K.; Loo, K.L.; Gubens, M.A.; Rosenblum, M.; Harview, C.L.; Taube, J.M.; Handley, N.; et al. Liver Metastasis and Treatment Outcome with Anti-PD-1 Monoclonal Antibody in Patients with Melanoma and NSCLC. *Cancer Immunol. Res.* 2017, 5, 417–424. [CrossRef]
14. Tournoy, K.G.; Thomeer, M.; Geronpre, P.; Derijcke, S.; De Pauw, R.; Galdermans, D.; Govaert, K.; Govaerts, E.; Schildermans, R.; Declercq, I.; et al. Does nivolumab for progressed metastatic lung cancer fulfill its promises? An efficacy and safety analysis in 20 general hospitals. *Lung Cancer* 2018, 115, 49–55. [CrossRef]
15. Kitadai, R.; Okuma, Y.; Hakozaki, T.; Hosomi, Y. The efficacy of immune checkpoint inhibitors in advanced non-small-cell lung cancer with liver metastases. *J. Cancer Res. Clin.* 2020, 146, 777–785. [CrossRef]
16. Mencoboni, M.; Ceppi, M.; Bruzzone, M.; Tiveg gia, P.; Cavo, A.; Scordamaglia, F.; Gualco, M.; Filiberti, R.A. Effectiveness and Safety of Immune Checkpoint Inhibitors for Patients with Advanced Non-Small-Cell Lung Cancer in Real-World: Review and Meta-Analysis. *Cancers* 2021, 13, 1388. [CrossRef]
17. Eisenhauer, E.; Therasse, P.; Bogaerts, J.; Schwartz, L.; Sargent, D.; Ford, R.; Dancey, J.; Arbuck, S.; Gwyther, S.; Mooney, M.; et al. New response evaluation criteria in solid tumours: Revised RECIST guideline (version 1.1). Eur. J. Cancer 2009, 45, 228–247. [CrossRef]

18. Lee, J.C.; Green, M.D.; Huppert, L.A.; Chow, C.; Pierce, R.H.; Daud, A.I. The Liver-Immunity Nexus and Cancer Immunotherapy. Clin. Cancer Res. 2022, 28, 5–12. [CrossRef]

19. Tsilimigras, D.I.; Brodt, P.; Clavien, P.A.; Muschel, R.J.; D'Angelica, M.I.; Endo, I.; Parks, R.W.; Doyle, M.; de Santibanes, E.; Pawlik, T.M. Liver metastases. Nat. Rev. Dis. Primers 2021, 7, 27. [CrossRef]

20. Limmer, A.; Ohl, J.; Kurts, C.; Ljunggren, H.G.; Reiss, Y.; Grooten, M.; Momburg, F.; Arnold, B.; Knolle, P.A. Efficient presentation of exogenous antigen by liver endothelial cells to CD8+ T cells results in antigen-specific T-cell tolerance. Nat. Med. 2000, 6, 1348–1354. [CrossRef]

21. Lee, J.; Mehdizadeh, S.; Smith, J.; Young, A.; Mufazalov, I.; Mowery, C.; Daud, A.; Bluestone, J. Regulatory T cell control of systemic immunity and immunotherapy response in liver metastasis. Sci. Immunol. 2020, 5, eaba0759. [CrossRef] [PubMed]

22. Garassino, M.C.; Gadgeel, S.; Esteban, E.; Felip, E.; Speranza, G.; Flávia De Angelis, M.D.; Clingan, P.; Hochmair, M.J.; Powell, S.F.; Cheng, S.Y.-S.; et al. CT043-Outcomes among patients (pts) with metastatic non-squamous NSCLC with liver metastases or brain metastases treated with pembrolizumab (pembro) plus pembetrexed-platinum: Results from the KEYNOTE-189 study. Am. Assoc. Cancer Res. AACR 2019, 79, CT043. [CrossRef]

23. Sridhar, S.; Paz-Ares, L.; Liu, H.; Shen, K.; Morehouse, C.; Rizvi, N.; Segal, N.H.; Jin, X.; Zheng, Y.; Narwal, R.; et al. Prognostic Significance of Liver Metastasis in Durvalumab-Treated Lung Cancer Patients. Clin. Lung Cancer 2019, 20, e601–e608. [CrossRef] [PubMed]

24. Funazo, T.; Nomizo, T.; Kim, Y.H. Liver Metastasis Is Associated with Poor Progression-Free Survival in Patients with Non-Small Cell Lung Cancer Treated with Nivolumab. J. Thorac. Oncol. 2017, 12, e140–e141. [CrossRef] [PubMed]

25. Castanon, E.; Rolfo, C.; Vinal, D.; Lopez, I.; Fusco, J.P.; Santisteban, M.; Martin, P.; Zubiri, L.; Echeveste, J.I.; Gil-Bazo, I. Impact of epidermal growth factor receptor (EGFR) activating mutations and their targeted treatment in the prognosis of stage IV non-small cell lung cancer (NSCLC) patients harboring liver metastasis. J. Transl. Med. 2015, 13, 257. [CrossRef] [PubMed]

26. Tamura, T.; Kurishima, K.; Nakazawa, K.; Kagohashi, K.; Ishikawa, H.; Satoh, H.; Hizawa, N. Specific organ metastases and survival in metastatic non-small-cell lung cancer. Mol. Clin. Oncol. 2015, 3, 217–221. [CrossRef]

27. Osorio, J.; Arbou, K.; Le, D.; Durham, J.; Plodkowski, A.; Halpenny, F.; Ginsberg, M.; Savan, P.; Crompton, J.; Yu, H.; et al. Lesion-Level Response Dynamics to Programmed Cell Death Protein (PD-1) Blockade. J. Clin. Oncol. 2019, 37, 3546–3555. [CrossRef]

28. Topalian, S.L.; Drake, C.G.; Pardoll, D.M. Immune checkpoint blockade: A common denominator approach to cancer therapy. Cancer Cell 2015, 27, 450–461. [CrossRef]

29. Niemeijer, A.N.; Sahba, S.; Smit, E.F.; Lissenberg-Witte, B.I.; de Langen, A.J.; Thunnissen, E. Association of tumour and stroma PD-1, PD-L1, CD3, CD4 and CD8 expression with DCB and OS to nivolumab treatment in NSCLC patients pre-treated with chemotherapy. Br. J. Cancer 2020, 123, 392–402. [CrossRef]

30. Hashemi, S.; Fransen, M.F.; Niemeijer, A.; Ben Taleb, N.; Houda, I.; Veltman, J.; Becker-Commissaris, A.; Daniels, H.; Crombag, L.; Radonic, T.; et al. Surprising impact of stromal TIL’s on immunotherapy efficacy in a real-world lung cancer study. Lung Cancer 2021, 153, 81–89. [CrossRef]

31. Brahmer, J.; Drake, C.; Wollner, I.; Powderly, J.; Picus, J.; Sharifman, W.; Stankevich, E.; Pons, A.; Salay, T.; McMillor, T.; et al. Phase I study of single-agent anti-programmed death-1 (MDX-1106) in refractory solid tumors: Safety, clinical activity, pharmacodynamics, and immunologic correlates. J. Clin. Oncol. 2010, 28, 3167–3175. [CrossRef]

32. Hellmann, M.D.; Ciuleanu, T.E.; Pluzanski, A.; Lee, J.S.; Ottersten, G.A.; Audigier-Valette, C.; Minenza, E.; Fanucchi, A.; Burgers, S.; Salman, P.; et al. Nivolumab plus ipilimumab in Lung Cancer with a High Tumor Mutational Burden. Clin. Lung Cancer 2019, 20, 378, 2093–2104. [CrossRef]

33. Garcia, J.; Hurwitz, H.; Sandler, A.; Miles, D.; Coleman, R.; Deurloo, R.; Chion, O. Bevacizumab (Avastin®) in cancer treatment: A review of 15 years of clinical experience and future outlook. Cancer Treat. Rev. 2020, 86, 102017. [CrossRef] [PubMed]

34. Yarchoan, M.; Albacker, L.; Hopkins, A.; Montesion, M.; Murugesan, K.; Vithayathil, T.; Zaidi, N.; Azad, N.; Laheru, D.; Frampton, S.F.; Cheng, S.Y.-S.; et al. Nivolumab plus Ipilimumab in Lung Cancer with a High Tumor Mutational Burden. Mol. Clin. Oncol. 2015, 3, 217–221. [CrossRef]

35. Ramjiawan, R.; Griffioen, A.; Duda, D. Anti-angiogenesis for cancer revisited: Is there a role for combinations with immunotherapy? Angiogenesis 2017, 20, 185–204. [CrossRef]

36. Hegde, P.; Wallin, J.; Mancao, C. Predictive markers of anti-VEGF and emerging role of angiogenesis inhibitors as immunotherapeutics. Semin. Cancer Biol. 2018, 52, 117–124. [CrossRef] [PubMed]

37. Garcia, J.; Hurwitz, H.; Sandler, A.; Miles, D.; Coleman, R.; Deurloo, R.; Chion, O. Bevacizumab (Avastin®) in cancer treatment: A review of 15 years of clinical experience and future outlook. Cancer Treat. Rev. 2020, 86, 102017. [CrossRef] [PubMed]

38. Lopes-Coelho, F.; Martins, F.; Pereira, S.; Serpa, J. Anti-Angiogenic Therapy: Current Challenges and Future Perspectives. Int. J. Mol. Sci. 2021, 22, 3765. [CrossRef] [PubMed]
39. Inai, T.; Mancuso, M.; Hashizume, H.; Baffert, F.; Haskell, A.; Baluk, P.; Hu-Lowe, D.D.; Shalinsky, D.R.; Thurston, G.; Yancopoulos, G.D.; et al. Inhibition of vascular endothelial growth factor (VEGF) signaling in cancer causes loss of endothelial fenestrations, regression of tumor vessels, and appearance of basement membrane ghosts. Am. J. Pathol. 2004, 165, 35–52. [CrossRef]

40. Wong, W.B.; Wu, N.; Yang, E.; Davies, J.; Chae, Y.K. Real-World Clinical and Economic Outcomes and the Role of Bevacizumab in Patients With Non–Small-Cell Lung Cancer With Liver Metastases. J. Oncol. Pract. 2019, 15, e878–e887. [CrossRef]

41. Izzo, F.; Granata, V.; Grassi, R.; Fusco, R.; Palaia, R.; Delrio, P.; Carrafiello, G.; Azoulay, D.; Petrillo, A.; Curley, S.A. Radiofrequency Ablation and Microwave Ablation in Liver Tumors: An Update. Oncologist 2019, 24, e990–e1005. [CrossRef] [PubMed]

42. Minami, Y.; Nishida, N.; Kudo, M. Radiofrequency ablation of liver metastasis: Potential impact on immune checkpoint inhibitor therapy. Eur. Radiol. 2019, 29, 5045–5051. [CrossRef] [PubMed]

43. Reck, M.; Mok, T.S.K.; Nishio, M.; Jotte, R.M.; Cappuzzo, F.; Orlandi, F.; Stroyakovskiy, D.; Nogami, N.; Rodriguez-Abreu, D.; Moro-Sibilot, D.; et al. Atezolizumab plus bevacizumab and chemotherapy in non-small-cell lung cancer (IMpower150): Key subgroup analyses of patients with EGFR mutations or baseline liver metastases in a randomised, open-label phase 3 trial. Lancet Respir. Med. 2019, 7, 387–401. [CrossRef]