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Permanent draft genome sequence of the gliding predator
*Saprospira grandis* strain Sa g1 (= HR1)

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**Keywords**
strictly aerobic, gliding, Gram-negative, mesophilic, chemoorganotrophic, marine, ixotrophy, *Saprospiraceae*, GEBA

**Abstract**
*Saprospira grandis* Gross *et al*. 1911 is a member to the genomically so far poorly characterized family *Saprospiraceae* in the class ‘Sphingobacteria’. The species is known for praying on other marine bacteria via ‘ixotrophy’. *S. grandis* strain Sa g1 was isolated from decaying crab carapace in France and was selected for genome sequencing due to its isolated location in the genomically sequenced part of the tree of life. Only one type strain genome has been published so far from the *Saprospiraceae*, while the sequence of strain Sa g1 is already the second genome to be published from *S. grandis* (type strain genome not yet sequenced). Here we describe the features of this organism, together with the complete genome sequence and annotation. The 4,495,250 bp long Improved-High-Quality draft of the genome with its 3,536 protein-coding and 62 RNA genes is a part of the *Genomic Encyclopedia of Bacteria and Archaea* project.

**Introduction**
Strain Sa g1 (= HR1 = DSM 2844 = ATCC 49590 = LMG 13157) belongs to the species *Saprospira grandis* \(^{[1,2]}\) in the monotypic genus *Saprospira* \(^{[2,3]}\). The type strain of the species is Lewin WH\(^1\) (= ATCC 23119 = LMG 10407) \(^{[1,3]}\) and is known for its predatory life style when capturing and praying on other bacteria via ‘ixotrophy’ \(^{[2]}\). Strain Sa g1 was isolated in 1975 from decaying crab carapace in Roscoff, France \(^{[19]}\). The genus name was derived from the Greek adjective *sapros*, meaning rotten/putrid, and the Latin *spira*, a coil/spiral, resulting in the Neo-Latin *Saprospira*, a spiral associated with decaying matter...
the species epithet was derived from the Latin adjective grandis, large [29]. Life style and ecological role of members of the species was recently summarized by Saw et al. [41] when they reported the genome sequence of strain Lewin (isolated from La Jolla beach in San Diego; not to be mixed-up with strain Lewin WH\(^1\), the type strain of the species which was also isolated by Lewin, but from a rockpool near high water, Woods Hole), the first member of the species Saprospira whose genome was deciphered. Here we present a summary classification and a set of features for S. grandis Sa g1, together with the description of the genomic sequencing and annotation.

**Classification and features**

A representative genomic 16S rRNA sequence of strain Sa g1 was compared using NCBI BLAST [7,8] under default settings (e.g., considering only the high-scoring segment pairs (HSPs) from the best 250 hits) with the most recent release of the Greengenes database [9] and the relative frequencies of taxa and keywords (reduced to their stem [10]) were determined, weighted by BLAST scores. The most frequently occurring genera were Saprospira (82.0%), Aureispira (5.4%), 'Aureospira' (4.8%), Cytophaga (3.9%) and Lewinella (3.8%) (16 hits in total). Regarding the three hits to sequences from members of the species, the average identity within HSPs was 99.4%, whereas the average coverage by HSPs was 98.6%. Among all other species, the one yielding the highest score was Aureispira maritima (AB278130), which corresponded to an identity of 87.3% and an HSP coverage of 98.0%. (Note that the Greengenes database uses the INSDC (= EMBL/NCBI/DDBJ) annotation, which is not an authoritative source for nomenclature or classification.) The highest-scoring environmental sequence was FJ792400 ('Unexpectedly archaeal species shift between rare and dominant over thousand year time scales carbonate chimney Lost City Hydrothermal Field clone SGYF672'), which showed an identity of 99.2% and an HSP coverage of 100.3%. The most frequently occurring keywords within the labels of all environmental samples which yielded hits were 'lake' (3.8%), 'sludg' (2.9%), 'microbi' (2.8%), 'mat' (2.7%) and 'activ' (2.3%) (234 hits in total) and correspond to the already known habitats for strains of this species.

Figure 1 shows the phylogenetic neighborhood of S. grandis strain Sa g1 in a 16S rRNA based tree. The sequences of the four 16S rRNA gene copies in the genome differ from each other by up to one nucleotide, and differ by up to seven nucleotides from the previously published 16S rRNA sequence (M58795), which contains 52 ambiguous base calls.
Figure 1. Phylogenetic tree highlighting the position of *S. grandis* relative to the type strains of the other species within the family *Saprospiraceae*. The tree was inferred from 1,413 aligned characters [11,12] of the 16S rRNA gene sequence under the maximum likelihood (ML) criterion [13]. Rooting was done initially using the midpoint method [14] and then checked for its agreement with the current classification (Table 1). The branches are scaled in terms of the expected number of substitutions per site. Numbers adjacent to the branches are support values from 250 ML bootstrap replicates [15] (left) and from 1,000 maximum-parsimony bootstrap replicates [16] (right) if larger than 60%. Lineages with type strain genome sequencing projects registered in GOLD [17] are labeled with one asterisk, those also listed as 'Complete and Published' with two asterisks [18].

General features of *S. grandis* were summarized in the previous issue of *Stand Genomic Sci* by Saw et al. for the short genome report of strain Lewin [41], and are therefore not repeated here. Individual features of strain *Sa g1* are largely unknown due to a lack of relevant publications, as are chemotaxonomical data. A description of the isolation and some morphological features of strain *Sa g1* are reported by Reichenbach [19]. Figure 2 shows an electron micrograph of the *S. grandis* *Sa g1* cells.

Figure 2. Scanning electron micrograph of *S. grandis* *Sa g1*
Table 1. Classification and general features of *S. grandis* Sa g1 in accordance with the MIGS recommendations [20] and the Names for Life database [2].

| MIGS ID | Property                      | Term                                | Evidence code |
|---------|-------------------------------|-------------------------------------|---------------|
|         | Domain                        | *Bacteria*                          | TAS [21]      |
|         | Phylum                         | ‘Bacteroidetes’                     | TAS [6]       |
|         | Class                          | ‘Sphingobacteria’                   | TAS [6]       |
| Current classification | Order                          | *Sphingobacteriales*                | TAS [6]       |
|         | Family                         | *Saprospiraceae*                    | TAS [22,23]   |
|         | Genus                          | *Saprospira*                        | TAS [3-5]     |
|         | Species                        | *Saprospira grandis*                | TAS [3,4]     |
|         | Strain                         | Sa g1                               | TAS [19]      |
|         | Gram stain                     | negative                            | TAS [39,40]   |
|         | Cell shape                     | helical filaments                   | TAS [39,40]   |
|         | Motility                       | via gliding                         | TAS [39,40]   |
|         | Sporulation                    | non-sporulating                     | NAS           |
|         | Temperature range              | mesophile, 6-47°C                   | TAS [39,40]   |
|         | Optimum temperature            | 25-30°C                             | TAS [19,39,40]|
|         | Salinity                       | seawater                            | TAS [39,40]   |
| MIGS-22 | Oxygen requirement             | strictly aerobe                     | TAS [39,40]   |
|         | Carbon source                  | peptides, proteins                  | TAS [39,40]   |
|         | Energy metabolism              | chemoorganotroph                    | TAS [39,40]   |
| MIGS-6  | Habitat                        | marine littoral zone                | TAS [39]      |
| MIGS-15 | Biotic relationship            | free living                         | TAS [39]      |
| MIGS-14 | Pathogenicity                  | not reported                         |               |
|         | Biosafety level                | 1                                   | TAS [24]      |
| MIGS-23.1 | Isolation                     | decaying crab carapace              | TAS [19]      |
| MIGS-4  | Geographic location            | Roscoff, France                     | TAS [19]      |
| MIGS-5  | Sample collection time         | September 1975                      | TAS [19]      |
| MIGS-4.1 | Latitude – Longitude          | 48.70 – -3.97                      | NAS           |
| MIGS-4.2 | Depth                         | not reported                         |               |
| MIGS-4.3 | Altitude                       | not reported                         |               |

Evidence codes - TAS: Traceable Author Statement (i.e., a direct report exists in the literature); NAS: Non-traceable Author Statement (i.e., not directly observed for the living, isolated sample, but based on a generally accepted property for the species, or anecdotal evidence). These evidence codes are from of the Gene Ontology project [25].

**Genome sequencing and annotation**

**Genome project history**
This organism was selected for sequencing on the basis of its phylogenetic position [26], and is part of the *Genomic Encyclopedia of Bacteria and Archaea* project [27]. The genome project is deposited in the Genomes On Line Database [17] and the complete genome sequence is deposited in GenBank. Sequencing, finishing and annotation were performed by the DOE Joint Genome Institute (JGI). A summary of the project information is shown in Table 2.

**Table 2. Genome sequencing project information**
| MIGS ID | Property                  | Term                                                                 |
|---------|---------------------------|----------------------------------------------------------------------|
| MIGS-31 | Finishing quality         | Improved-High-Quality Draft                                          |
| MIGS-28 | Libraries used            | Two genomic libraries: one 454 PE library (9 kb insert size), one Illumina library |
| MIGS-29 | Sequencing platforms      | Illumina GAii, 454 GS FLX Titanium                                   |
| MIGS-31.2 | Sequencing coverage   | 768.5 × Illumina; 8.6 × pyrosequence                                |
| MIGS-30 | Assemblers                | Newbler version 2.3, Velvet version 1.0.13, phrap version 1.080812   |
| MIGS-32 | Gene calling method       | Prodigal 1.4, GenePRIM                                             |
| INSDC ID |                           | not yet                                                            |
| GenBank Date of Release |                           | requested February 9, 2012                                         |
| GOLD ID |                           | Gi03955                                                          |
| NCBI project ID |                       | 61003                                                           |
| Database: IMG-GEBA         |                           | 2509276035                                                     |
| MIGS-13 | Source material identifier | DSM 2844                                                  |
| Project relevance         |                           | Tree of Life, GEBA                                               |

**Growth conditions and DNA isolation**

*S. grandis* strain Sa g1, DSM 2844, was grown in DSMZ medium 172 (*Cytophaga* (marine) medium) [28] at 28°C. DNA was isolated from 0.5-1 g of cell paste using Jetflex Genomic DNA Purification kit (GENOMED 600100) following the standard protocol as recommended by the manufacturer without modification.

**Genome sequencing and assembly**

The genome was sequenced using a combination of Illumina and 454 sequencing platforms. All general aspects of library construction and sequencing can be found at the JGI website [30]. Pyrosequencing reads were assembled using the Newbler assembler (Roche). The initial Newbler assembly consisting of 551 contigs in six scaffolds was converted into a phrap [31] assembly by making fake reads from the consensus, to collect the read pairs in the 454 paired end library. Illumina GAii sequencing data (3,575.7 Mb) was assembled with Velvet [32] and the consensus sequences were shredded into 1.5 kb overlapped fake reads and assembled together with the 454 data. The 454 draft assembly was based on 72.8 Mb of 454 paired end data. Newbler parameters are -consed -a 50 -i 350 -g -m -ml 20. The Phred/Phrap/Consed software package [31] was used for sequence assembly and quality assessment in the subsequent finishing process. After the shotgun stage, reads were assembled with parallel phrap (High Performance Software, LLC). Possible mis-assemblies were corrected with gapResolution [30], Dupfinisher [33], or sequencing cloned bridging PCR fragments with subcloning. Gaps between contigs were closed by editing in Consed, by PCR and by Bubble PCR primer walks (J.-F. Chang, unpublished). A total of 45 additional reactions were necessary to close gaps and to raise the quality of the final contigs. Illumina reads were also used to correct potential base errors and increase consensus quality using a software Polisher developed at JGI [34]. The final assembly consists of 84 contigs in 5 scaffolds. Together, the combination of the Illumina and 454 sequencing platforms provided 777.1 x coverage of the genome. The final assembly contained 235,183 pyrosequence and 45,502,670 Illumina reads.

**Genome annotation**

Genes were identified using Prodigal [35] as part of the Oak Ridge National Laboratory genome annotation pipeline, followed by a round of manual curation using the JGI GenePRIMP pipeline [36]. The predicted CDSs were translated and used to search the National Center for Biotechnology Information (NCBI) non-redundant database, UniProt,
TIGRFam, Pfam, PRIAM, KEGG, COG, and InterPro databases. Additional gene prediction analysis and functional annotation was performed within the Integrated Microbial Genomes - Expert Review (IMG-ER) platform [37].

**Genome properties**

The Improved-High-Quality draft assembly of the genome consists of 84 contigs in four scaffolds representing the chromosome (4,422,561 bp, 11,045 bp, 2,786 bp and 2,223 bp length, respectively) and one 56,635 bp plasmid scaffold, with an overall 46.1% G+C content (Table 3 and Figure 3). Of the 3,598 genes predicted, 3,536 were protein-coding genes, and 62 RNAs; 70 pseudogenes were also identified. The majority of the protein-coding genes (57.4%) were assigned a putative function while the remaining ones were annotated as hypothetical proteins. The distribution of genes into COGs functional categories is presented in Table 4.

**Table 3. Genome Statistics**

| Attribute                                | Value       | % of Total |
|------------------------------------------|-------------|------------|
| Genome size (bp)                         | 4,495,250   | 100.00%    |
| DNA coding region (bp)                   | 3,693,336   | 82.16%     |
| DNA G+C content (bp)                     | 2,067,067   | 46.06%     |
| Number of scaffolds                      | 5*          |            |
| Extrachromosomal elements                | 1           |            |
| Total genes                              | 3,598       | 100.00%    |
| RNA genes                                | 62          | 1.72%      |
| rRNA operons                             | 3**         |            |
| tRNA genes                               | 48          | 1.33%      |
| Protein-coding genes                     | 3,536       | 98.28%     |
| Pseudo genes                             | 70          | 1.95%      |
| Genes with function prediction (proteins)| 2,064       | 57.37%     |
| Genes in paralog clusters                | 1,575       | 43.77%     |
| Genes assigned to COGs                   | 2,064       | 57.37%     |
| Genes assigned Pfam domains              | 2,072       | 57.59%     |
| Genes with signal peptides               | 1,109       | 30.82%     |
| Genes with transmembrane helices         | 687         | 19.09%     |
| CRISPR repeats                           | 5           |            |

* four scaffolds for the chromosome and one for a plasmid  
** only two rRNA operons seem to be complete; the 3rd copy appears to be split in two incomplete fractions due to assembly problems.
Figure 3. Graphical map of the largest scaffold, SapgrDRAFT_Contig123.4, which represents >99.6% of the chromosome (plasmid scaffold not shown). From bottom to top: Genes on forward strand (colored by COG categories), Genes on reverse strand (colored by COG categories), RNA genes (tRNAs green, rRNAs red, other RNAs black), GC content, GC skew.

Table 4. Number of genes associated with the general COG functional categories

| Code | value | %age | Description                                                                 |
|------|-------|------|------------------------------------------------------------------------------|
| J    | 152   | 6.8  | Translation, ribosomal structure and biogenesis                             |
| A    | 0     | 0.0  | RNA processing and modification                                              |
| K    | 118   | 5.3  | Transcription                                                               |
| L    | 167   | 7.5  | Replication, recombination and repair                                         |
| B    | 1     | 0.0  | Chromatin structure and dynamics                                              |
| D    | 34    | 1.5  | Cell cycle control, cell division, chromosome partitioning                   |
| Y    | 0     | 0.0  | Nuclear structure                                                           |
| V    | 53    | 2.4  | Defense mechanisms                                                          |
| T    | 98    | 4.4  | Signal transduction mechanisms                                               |
| M    | 217   | 9.7  | Cell wall/membrane biogenesis                                                |
| N    | 25    | 1.1  | Cell motility                                                               |
| Z    | 1     | 0.0  | Cytoskeleton                                                                |
| W    | 0     | 0.0  | Extracellular structures                                                    |
| U    | 55    | 2.5  | Intracellular trafficking and secretion, and vesicular transport              |
| O    | 121   | 5.4  | Posttranslational modification, protein turnover, chaperones                 |
| C    | 108   | 4.9  | Energy production and conversion                                             |
| G    | 51    | 2.3  | Carbohydrate transport and metabolism                                        |
| E    | 125   | 5.6  | Amino acid transport and metabolism                                          |
| F    | 57    | 2.6  | Nucleotide transport and metabolism                                          |
| H    | 101   | 4.5  | Coenzyme transport and metabolism                                           |
Insights into the genome sequence

Comparison with the genome sequence of S. grandis strain Lewin

The two complete copies of the 16S rRNA gene in the S. g1 genome show 99.5% sequence identity with those of strain Lewin [41], but only 98.0% sequence identity with the respective sequence from in the not yet genome-sequenced type strain Lewin WH (ATCC 23119, M58795) [42]; this discrepancy is due to the huge number of ambiguous base calls in M58795, and is relativized by 99.4% identical bases within the HSPs. Given the different habitats of the two sequenced strains it appears to be interesting to compare some basic genome features and their membership to the same species.

The second largest scaffold in the Improved-High-Quality draft assembly of the S. g1 genome (SapgrDRAFT_Contig162.5) has a size of 56,635 bp, which is comparable to the size of plasmid SGRA01 in S. grandis strain Lewin, 54,948 bp (CP002832) [41]. A BLAST search against NCBI nr database revealed a full length colinearity with about 94% sequence identity between SapgrDRAFT_Contig162.5 and the plasmid of S. grandis strain Lewin. Further comparison of the two sequences with the GGDC-Genome-to-Genome Distance Calculator [43,44] revealed distances of only 0.0704 (formula 1) to 0.1342 (formula 3), corresponding to 82.7 to 88.3% DDH values. SapgrDRAFT_3602 encodes a protein involved in initiation of plasmid replication, RepB, while the largest fraction of (13) genes encoded on this scaffold belong to COG function category ‘nucleotide transport and metabolism’ (similar to SGRA01); it is therefore suggesting to consider SapgrDRAFT_Contig162.5 as a plasmid whose sequence was not circularized during the genome assembly.

The largest scaffold in the draft assembly of S. g1 (SapgrDRAFT_Contig123.4) has a size of 4,422,561 bp, which is comparable to the size of the S. grandis strain Lewin chromosome, 4,345,237 bp. The overall genome statistics (see Table 3) of the two strains is similar in some features, such as G+C content (46.1% strain Sa g1 vs. 46.4% strain Lewin), total number of genes (3,598 vs. 4,311), genes with function predictions (2,064 vs. 2,173), three rRNA operons (both), but deviates more in others, such as genes in paralog clusters (1,575 vs. 215), genes with signal peptides (1,109 vs. 589), genes with transmembrane helices (687 vs. 778), which may to a good part reflect the differences in the gene calling and annotation process (strain Lewin is not yet featured in img [37]. As for the number of genes associated with the general COG functional categories (see Table 4) there are categories with very similar content, such as transcription (118 genes, both), translation (152 vs. 160), defense mechanisms (53 vs. 52), cell motility (25 vs. 26), lipid transport (88 vs. 90) and cell wall/membrane biogenesis (217 vs. 206), while other categories deviate more significantly, such as replication (167 vs. 186), cell cycle control (34 vs. 20), intracellular trafficking (55 vs. 44), energy production and conversion (108 vs. 123), secondary metabolites biosynthesis an catabolism (35 vs. 52), which again might in parts be caused by different procedures in the annotation processes.
The sequences of SapgrDRAFT_Contig123.4 and the chromosome of strain Lewin (CP002831), which represent roughly 99% of the respective genomes, were also compared with the GGDC-Genome-to-Genome Distance Calculator [43,44]. The inferred distances from formulas 1 and 3 were 0.1139 and 0.1741, respectively, corresponding to 83.1% and 77.9% DDH values, respectively, estimated via regression-based predictions. These values indicate that both strains belong to the same species, S. grandis.

The sequence of the three smaller scaffolds (SapgrDRAFT_Contig118.2 with 11,045 bp length, SapgrDRAFT_Contig106.1 with 2,786 bp and SapgrDRAFT_Contig119.3 with 2,223 bp) were compared against the NCBI nr database and revealed significant similarities only with the chromosome of strain Lewin.

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