Serum galectin-3 and galectin-3 binding protein levels in systemic lupus erythematosus and cutaneous lupus erythematosus

Agnieszka Kalinska-Bienias1, Emilia Kowalczyk1, Piotr Bienias2, Kamila Gala1, Pawel Jagielski4, Cezary Kowalewski3

1Department of Dermatology and Immunodermatology, Medical University of Warsaw, Warsaw, Poland
2Department of Internal Medicine and Cardiology, Medical University of Warsaw, Warsaw, Poland
3Department of Immunology, Transplantology and Internal Diseases, Medical University of Warsaw, Warsaw, Poland
4Human Nutrition Department Faculty of Health Science, Jagiellonian University Medical College, Krakow, Poland

Abstract

Introduction: The roles of galectin-3 (Gal-3) and galectin-3 binding protein (G3BP) in systemic lupus erythematosus (SLE) are of ongoing interest, but the data are insufficient due to highly limited available studies. There are no data on cutaneous lupus erythematosus (CLE).

Aim: To assess serum Gal-3 and G3BP concentrations in SLE patients with and without LE-specific skin lesions, CLE patients and to correlate levels of proteins with clinical and laboratory parameters.

Material and methods: The study included 71 SLE patients with and without LE-specific skin lesions, 23 CLE patients and 40 controls. Gal-3 and G3BP were measured by specific enzyme-linked immunosorbent assays (ELISA).

Results: Serum Gal-3 and G3BP concentrations were significantly higher in SLE with and without LE-specific lesions compared to controls, but without differences between SLE groups. Gal-3 and G3BP levels were also elevated in CLE compared to controls ($p = 0.001$, $p = 0.005$; respectively). There was a positive correlation between G3BP level and CLASI activity score in CLE ($r = 0.55$, $p = 0.006$) and in SLE patients with LE-specific lesions ($r = 0.36$, $p = 0.02$). G3BP and Gal-3 levels did not differ in SLE with LE-specific lesions and CLE. There was a positive correlation between serum G3BP level and the SLEDAI score in SLE patients ($r = 0.26$, $p = 0.03$).

Conclusions: Our findings indicate that serum G3BP and Gal-3 are elevated in CLE. Additionally, G3BP might be associated with the extent of skin lesions. There are no differences between G3BP and Gal-3 concentrations in SLE with and without LE-specific skin lesions.

Key words: galectin-3, galectin-3 binding protein, lupus erythematosus.
to now at least 15 galectins have been identified. Gal-3 plays an extracellular role, inducing the apoptosis of T-cells and monocytes, and an intracellular one with its anti-apoptotic activity [4, 5]. The expression of Gal-3 is stimulated by the hypomethylation of the gene promoter. Consequently, DNA hypomethylation is seen in various connective tissue diseases, including SLE [6].

Originally described as a macrophage surface marker binding protein (Mac-2 BP), G3BP is a scavenger cytosine-rich domain receptor that exerts its activity by binding to lectins, integrins and extracellular matrix proteins. G3BP is a ligand for galectin-3 and an important factor of cell clearance, apoptosis, intercellular adhesion and signaling [7].

Data on the serum concentration of Gal-3 and G3BP in SLE were initially derived from the study of patients with Behcet’s disease in which SLE patients represented a control group. Interestingly, serum Gal-3 and G3BP proteins were much higher in SLE than in Behcet’s disease and healthy groups [8]. The results of further studies revealed that Gal-3 and G3BP concentrations were elevated in the sera of SLE patients compared to healthy controls [9–14]. However, by reason of the pioneering character of the research as well as the small number of the study groups, their role as markers of the immune and inflammatory processes in SLE requires further investigation. In a particularly interesting study, Shi et al. showed that the expression of Gal-3 in skin biopsies obtained from LE-specific skin lesions was reduced. In addition, the authors compared serum Gal-3 data in patients with SLE with and without LE skin lesions and found no difference between the groups [9]. It indicates that the role of Gal-3 in LE skin lesions needs to be elucidated. To the best of our knowledge, there are no data on Gal-3 and G3BP in patients with CLE as well as no evaluation of G3BP level in SLE with and without LE-specific skin lesions.

Aim

The aim of this study was to assess the serum Gal-3 and G3BP concentrations in SLE patients (with and without LE-specific lesions) and in CLE patients compared to healthy subjects and to correlate the concentration of those proteins with clinical and laboratory features.

Material and methods

Patients

The study included 71 patients diagnosed with SLE, 23 patients with CLE and 40 healthy subjects as a control group. The diagnosis of SLE was determined according to the American College of Rheumatology (ACR) revised criteria if the patients fulfilled four or more than four [15]. Patients with SLE were classified into two groups based on the presence of LE-specific skin lesions at the time of serum collections: (i) SLE with LE-specific skin lesions (38 patients), (ii) SLE without LE-specific skin lesions (33 patients). The LE-specific skin lesions presented ACLE, SCLE and/or CCLE. In some patients common skin manifestations, e.g. livedo reticularis, Raynaud’s phenomenon, or periungual telangiectasia, were found, but these were not classified as LE-specific skin manifestations. The SLE activity was assessed by the Systemic Lupus Erythematous Disease Activity Index (SLEDAI) [16]. The diagnosis in patients with CLE was determined on the basis of clinical features, duration of skin lesions and histopathological study of skin lesions. All CLE patients manifested skin lesions at the time of serum collections and presented less than four ACR criteria. Additionally, immunofluorescent study of biopsies (lupus band test) taken from skin lesions was performed in 85% of CLE patients. The extent of skin lesions in patients with SLE and CLE was assessed by the Cutaneous Lupus erythematosus disease Area and Severity Index (CLASI) activity and damage scores [17]. Routine laboratory evaluation of white blood cell count (WBC), red blood count (RBC), platelet count (PLT), erythrocyte sedimentation rate (ESR), C-reactive protein (CRP) and complements 3 (C3) and 4 (C4) was assessed by standard laboratory methods. Indirect immunofluorescence on human Hep-2 cells was used to determine antinuclear antibodies (ANA). Double-stranded DNA antibodies were assessed on Crithidia luciliae. Specification of ANA was performed by immunoblot technique and/or the immunodiffusion method. Consent of the local Bioethics Committee was obtained prior to the study. All patients and controls gave informed consent and institutional approval was obtained.

Measurement of serum galectin-3 and galectin-3 binding protein concentrations

Serum samples were stored at −20°C until assessment. Specific enzyme-linked immunosorbent assay (ELISA) kits were used to measure serum Gal-3 (Wuhan Fine Biological Technology Co., Ltd., Wuhan, China) and G3BP concentrations (human 90K/Mac-2BP platinum ELISA kit, Bender MedSystems, Vienna, Austria) in patients and the control group. The procedures were performed according to the manufacturer’s instructions. Serum Gal-3 and G3BP concentrations were calculated using the standard curve. All experiments were carried out in duplicate.

Statistical analysis

All statistical analyses were performed using the statistical software packages IMAGO PRO Academic (IBM SPSS Statistics 25) and SAS Enterprise Guide 7.1. Variables with a normal distribution are presented as mean followed by standard deviation (SD). For categorical variables, the differences between groups were compared by the χ2 test. Each parameter was assessed for normality with the Shapiro-Wilk test. The Mann-Whitney-U test was used for the comparison of non-parametric continu-
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Results

The characteristics of SLE patients with and without LE-specific skin lesions and CLE patients are shown in Table 1. The mean age of SLE patients with and without LE-specific skin lesions, CLE patients and the control group was 48.0 ±16.4, 54.0 ±14.3, 52.5 ±14.4 and 48.4 ±12.9 years, respectively. There was no difference in disease duration between studied groups. Arthritis, kidney disease and neurological diseases occurred with similar frequency in groups of SLE patients. The median SLEDAI score was higher in SLE patients with LE-specific skin lesions than without LE-specific skin lesions (p = 0.02), mainly due to the presence of skin lesions. The median CLASI activity score was higher in SLE patients with LE-specific skin lesions than in CLE patients (p = 0.02) while the CLASI damage score was similar (p = 0.77).

Serum galectin-3 and galectin-3 binding protein concentrations

The mean serum Gal-3 concentration was 10.46 ±3.25 ng/ml in SLE with LE-specific skin lesions, 10.69 ±3.42 ng/ml in CLE without LE-specific skin lesions, 10.15 ±2.68 in CLE, 6.92 ±3.47 ng/ml in the control group (Figure 1 A). The mean concentration of serum Gal-3 did not differ between CLE and SLE with LE-skin lesions (p = 1.0). Similarly, there was no difference between SLE with LE-specific skin lesions vs. SLE without LE-specific skin lesions (p = 1.0). The mean serum G3BP concentration was 13714.7 ±4595.5 µg/ml in SLE with LE-specific skin lesions, 11399.6 ±3760.9 µg/ml in CLE without LE-specific skin lesions, 10964.0 ±3866.6 µg/ml in CLE and 7870.0 ±3113.8 µg/ml in the control group. The concentrations of G3BP were significantly higher in all groups of patients compared to the control group (Figure 1 B). The level of G3BP did not differ in CLE and SLE with LE-specific skin lesions. No difference in serum G3BP concentration was found between SLE patients with and without LE-skin lesions (p = 0.28).

Correlations of serum galectin-3 and galectin-3 binding protein concentrations with clinical and laboratory parameters

A positive correlation was observed between serum G3BP concentration and CLASI activity score in CLE (r = 0.55, p = 0.006) (Figure 2 A). Serum G3BP level also correlated with CLASI activity score in SLE with LE-specific skin lesions (r = 0.36, p = 0.02) (Figure 2 B). There was a positive correlation between serum G3BP concentration and the SLEDAI score in SLE patients (r = 0.26, p = 0.03) (Figure 2 C). A positive correlation was observed between serum G3BP concentration and ESR in SLE patients (r = 0.3, p = 0.01). Serum G3BP concentration did not correlate with other clinical and laboratory parameters (disease duration, CLASI damage score, serum levels of C3 and C4, ANA, arthritis, kidney and neurological diseases). No correlations were observed between serum Gal-3 concentration and disease duration, the SLEDAI score, ESR, levels of C3 and C4, ANA, arthritis, kidney and neurological diseases (precise data not shown).

Discussion

The diversity of functions performed by Gal-3 and G3BP causes an increasing interest in those proteins in the context of inflammatory and autoimmune disorders. It seems that they could contribute to a fuller understanding of lupus erythematosus pathogenesis. Available scientific reports indicate an increased serum level of Gal-3 in patients with SLE [9–11, 14, 18]. As regards G3BP, a particularly promising assessment was presented by the Nielsen et al. study (2015), in which the authors observed a higher serum G3BP concentration in SLE patients, especially in lupus nephritis [19]. Our study also showed that serum Gal-3 and G3BP concentrations were significantly higher in SLE patients.

To the best of our knowledge, we are the first to assess Gal-3 and G3BP serum concentrations in CLE patients, noting the increased levels of both proteins in this disease. Unexpectedly, the concentration of Gal-3 and G3BP was similar in patients with CLE and SLE with LE-specific skin lesions. The reason for these results is difficult to explain as CLE remains mainly a skin problem, not a systemic one. Nevertheless, some disturbances typical of systemic involvement are observed in CLE, but they are not sufficient to meet the criteria necessary for the diagnosis of SLE. We hypothesized that the high concentration of Gal-3 and G3BP in CLE may be associated with the lack of immunosuppressive therapy (usually systemic steroids) which was applied in most SLE patients as opposed to patients with CLE.

Our results suggest that elevated serum G3BP concentration may reflect the extent of skin lesions. We did not make such an observation in relation to Gal-3, which is consistent with the results of the Shi et al. study from 2018. On the other hand, there is evidence that Gal-3 may contribute to the development of lupus skin lesions by recruiting inflammatory cells for the dermis. The analysis of lupus skin biopsies in patients with SLE revealed that Gal-3 expression was reduced in the epidermis of LE-specific skin lesions compared to healthy controls (lower in chronic lesions than in acute ones) [9]. It is suggested that a decrease in Gal-3 expression in the epidermis may be associated with its involvement in lymphoid
infiltrates in the dermis as Gal-3 is known to activate many lymphoid cells and act as a chemoattractant for human monocytes and macrophages. Moreover, in vivo studies using Gal-3 deficient mice support the notion that Gal-3 promotes inflammatory responses in mouse models [20]. It has also been shown that Gal-3 knock-

### Table 1. Clinical and laboratory characteristics of patients with systemic lupus erythematosus with lupus-specific skin lesions, systemic lupus erythematosus without lupus-specific skin lesions and discoid lupus erythematosus (NA – not available). P-value in one row shows statistical differences between 2 or 3 values depending on characteristic.

| Parameter | SLE with skin lesions (n = 38) | SLE without skin lesions (n = 33) | CLE (n = 23) | P-value |
|-----------|-------------------------------|----------------------------------|-------------|---------|
| **Clinical data:** |                               |                                  |             |         |
| Age at enrolment [years] | 48.0 ±16.4                     | 54.0 ±14.3                       | 52.5 ±14.4  | 0.22    |
| Age at onset [years]     | 38.8 ±18.7                     | 38.6 ±13.8                       | 38.9 ±19.6  | 0.99    |
| Gender, n male/female    | 5/33                           | 1/32                             | 6/17        | 0.04    |
| Disease duration, (range) [years] | 6.5 (1–35)                  | 16 (1–30)                        | 7 (0–44)    | 0.07    |
| Arthritis, n (%)         | 29 (76.3)                      | 24 (72.7)                        | –           | 0.96    |
| Kidney disease, n (%)    | 10 (26.3)                      | 8 (24.2)                         | –           | 0.84    |
| Neurological diseases, n (%) | 8 (21.1)                  | 7 (21.2)                         | –           | 0.98    |
| CLASI activity score (range) | 8 (2–20)                    | –                                | 4 (2–21)    | 0.02    |
| CLASI damage score (range) | 3 (0–10)                   | –                                | 3 (0–23)    | 0.77    |
| SLEDAI score (range)     | 8 (2–18)                      | 6 (0–22)                         | –           | 0.02    |
| **Laboratory and immunologic data:** |                               |                                  |             |         |
| WBC [× 10^3/mm³]         | 5.8 ±2.7                       | 5.8 ±2.0                         | 6.1 ±1.5    | 0.82    |
| RBC [× 10^9/l]           | 4.4 ±0.5                       | 4.4 ±0.3                         | 4.6 ±0.5    | 0.19    |
| PLT [× 10^3/mm³]         | 227.0 ±96.0                    | 215.0 ±83.6                      | 228.7 ±64.8 | 0.41    |
| ESR [mm/h]               | 12.5 (3–86)                    | 18 (4–59)                        | 11.7 ±11.1  | 0.02    |
| CRP                      | 1.6 (0.2–30)                   | 2 (0.3–51)                       | 1.4 (0.1–10) | 0.29    |
| ANA mean titer ± SD      | 480 (80–5120)                  | 320 (80–2560)                    | 160 (0–2560) | < 0.0001 |
| Anti-ds-DNA, n (%)       | 8 (21.1)                      | 5 (15.2)                         | 1 (4.3)     | 0.21    |
| Anti-Ro, n (%)           | 19 (50.0)                      | 17 (51.5)                        | 0 (0)       | 0.0001  |
| Anti-La, n (%)           | 5 (13.2)                      | 5 (15.2)                         | 0 (0)       | 0.16    |
| Anti-RNP, n (%)          | 5 (13.2)                      | 3 (9.1)                          | 1 (4.3)     | 0.52    |
| Anti-Sm, n (%)           | 2 (5.3)                       | 1 (3.0)                          | 0 (0)       | 0.52    |
| Anti-histone, n (%)      | 1 (2.6)                       | 0 (0)                            | 0 (0)       | 0.47    |
| C3 complement [mg/l]     | 97.7 ±33.4                     | 99.8 ±29.2                       | 110.2 ±15.3 | 0.13    |
| C4 complement [mg/l]     | 18.4 ±10.8                     | 18.1 ±9.9                        | 25.4 ±11.7  | 0.03    |
| **Treatment, n (%):**    |                               |                                  |             |         |
| Glucocorticosteroids     | 33 (86.8)                      | 23 (69.7)                        | 2 (8.7)     | < 0.0001 |
| Antimalarial drugs       | 19 (50.0)                      | 16 (48.5)                        | 18 (78.3)   | 0.05    |
| Cyclosporine             | 5 (13.2)                      | 0 (0)                            | 0 (0)       | 0.02    |
| Methotrexate             | 1 (2.6)                       | 4 (12.1)                         | 0 (0)       | 0.09    |
| Cyclophosphamide         | 0 (0)                         | 1 (3.0)                          | 0 (0)       | 0.39    |
| Mycophenolate mofetil    | 3 (7.9)                       | 1 (3.0)                          | 0 (0)       | 0.30    |
| Azathioprine             | 7 (18.4)                      | 7 (21.2)                         | 1 (4.3)     | 0.21    |
| Dapsone                  | 3 (7.9)                       | 0 (0)                            | 0 (0)       | 0.10    |

Aggravations – CLASI Cutaneous Lupus erythematosus disease Area and Severity Index, SLEDAI – Systemic Lupus Erythematosus Disease Activity Index, WBC – white blood cell count, RBC – red blood count, PLT – platelet count, ESR – erythrocyte sedimentation rate, CRP – C-reactive protein, ANA – antinuclear antibodies.
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Downregulated resulted in the overexpression of chemokines such as CXCL1 and CCL20, and enhanced the peripheral blood mononuclear cell-attracting capacity of keratinocytes [9]. It is worth mentioning that purified anti-Gal-3 antibodies administered subcutaneously to mice resulted in LE histopathological changes (vasculitis and epidermal atrophy) [21]. Interestingly, in the present study we found that in patients with SLE there was no difference in Gal-3 concentration in groups with and without LE-specific skin lesions. Similar results were obtained by Shi et al. in 2018 [9]. It is noteworthy that in the present study we noted for the first time that patients with SLE with and without skin lesions did not differ in G3BP levels. These results suggest that the presence of skin lesions in SLE patients probably does not affect both Gal-3 and G3BP concentrations.

Nevertheless, available literature provides data on Gal-3 and G3BP as novel biomarkers of SLE activity [8–11]. Our analysis also supported a positive correlation between G3BP concentration and SLEDAI score in SLE patients; however, we did not find a correlation between Gal-3 and SLEDAI score. It is worth mentioning that SLE patients had much higher levels of Gal-3 and G3BP compared to other connective tissue diseases such as rheumatoid arthritis, Behçet’s disease or systemic sclerosis, although the concentrations of Gal-3 and G3BP were found to be elevated in those diseases [10]. It should also be underlined that the concentration of Gal-3 was

**Figure 1.** A – Serum galectin-3 (Gal-3) concentration in systemic lupus erythematosus (SLE), SLE with LE-specific skin lesions, SLE without LE-specific skin lesions, cutaneous lupus erythematosus (CLE), B – control group serum galectin-3 binding protein (G3BP) concentration in SLE, SLE with LE-specific skin lesions, SLE without LE-specific skin lesions, CLE and control group
revealed to be higher in SLE than in systemic sclerosis, although Gal-3 is widely considered as a marker of fibrosis, providing extensive proliferation of fibroblasts and stimulation of the synthesis of collagen (mainly type I) [10, 22].

The strict mechanism of Gal-3 and G3BP function in SLE is not clearly understood. It is known that typical for SLE pathogenesis is the defective clearance of apoptotic cells. Galectins are used as agents whose task is to recognize apoptotic cells and then through the glycosylation process to change their structure so that they are recognized by the immune system [5]. G3BP was found as their distinguishable biomarker.

Some data confirm that micro-particles derived from cellular apoptosis are the source of autoantigens and Gal-3 or G3BP may be among them [21]. Kang et al. analyzing renal biopsies in patients with lupus nephritis found significantly elevated glomerular expression of Gal-3 in renal tissue [11]. Regarding G3BP, it has been shown by Nielsen et al. that G3BP was present together with immune complexes in kidney tissues in SLE patients with renal involvement [19].

In addition, it was found that G3BP plasma level mirrors type I interferon (IFN) activity [13]. Increased G3BP activity is also present in α-IFN-activated neutrophils and peripheral blood mononuclear cells in SLE patients. Increased activity of IFN and G3BP in connection with an excess of circulating molecules containing fragments of apoptotic cells may be a factor triggering such a strong, systemic autoimmune reaction in SLE [7].

The practical value of Gal-3 and G3BP evaluation is an open issue. Due to numerous connections between Gal-3, G3BP and lupus erythematosus, it seems that further research will facilitate considering them as disease biomarkers in routine examinations. Moreover, some scientific reports have demonstrated that galectins have a great deal of therapeutic potential. We hope that our findings might be a proper direction for the development of galectin-based therapeutic agents.

The limitation of this study is the small representation of the patients resulting from the assessment carried out by a single university center.

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Conflict of interest

The authors declare no conflict of interest.

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