Research Article

Antibacterial and Antifungal Activities of Zinc-Silicon Oxides Nanocomposite

Muhammad Arshad, Muhammad Akhyar Farrukh*, Sadia Haneef, Nabeela Aslam, Amna Afzaal

Department of Chemistry, Nano-ChemistryLab, GC University Lahore, Pakistan

*Corresponding authors: Farrukh, M.A. Department of Chemistry, Nano-ChemistryLab, GC University Lahore, 54000, Pakistan; E-mail: akhyar100@gmail.com

Abstract

Synthesis of ZnO-SiO2 nanocomposite was carried out via deposition-precipitation method by using acetonitrile as solvent. Synthesized nanocomposite was characterized by using different analytical techniques like Fournier Transformation Infrared Spectroscopy (FT-IR), Thermo-Gravimetric Analysis (TGA), Powder X-ray Diffraction (XRD), Scanning Electron Microscopy (SEM), Transmission Electron Microscopy (TEM), and Dynamic Light Scattering (DLS). Biological potential such as antibacterial and antifungal activities were also studied by using Disc Diffusion method and Agar Well Diffusion Assay respectively. The particle size of ZnO-SiO2 nanocomposite calculated by TEM was found to be 6.2 nm. The nanocomposite showed better antibacterial activity than antifungal activity.

Keywords: Nanocomposite; Deposition-precipitation; Antibacterial activity; Antifungal activity

Introduction

Several studies have suggested that nanoparticles can be used as efficient antimicrobial agents. Zinc oxide powder has been used as an active constituent in ointment, creams and lotions for skin treatments owing to its antibacterial properties. However, zinc oxide nanoparticles are much more effective in suppression of microorganism growth. Furthermore, at nanoscale the properties of particles are widely changed, enhancing their semiconducting abilities as well as the efficacy in inhibiting the growth of bacteria[1]. Microorganisms are the major constituents in the biosphere and their growth may be implement beneficial or harmful effect on the environment regarding to the human being. They are naturally symmetrical with environment and human body. So it is necessary to control their harmful effects by inhibiting their growth. Antimicrobial agents are chemical compounds which have potential to inactivate or inhibit the growth of microorganism[2]. These antimicrobial agents have vast applications in various fields including food preservation and packaging, medicine, water disinfection, textile fabrics and hospital implants[3].

Nanotechnology is also getting interest due to their attractive material properties and applications in various fields such as photo-catalysis, organic pollutants, optical devices, antibacterial coatings and sensors[4]. Nanotechnology is getting advancement particularly the potential to prepare metal oxide nanomaterials of specific shape and size which leads it in the development of new antibacterial agents[5].

Various nanoparticles of metal oxides have been synthesized and found good inhibitor of different bacterial strains. Activity of nanoparticles is directly dependent on the bacterial strains[6]. Metal oxides have wide range applications in sensors, microelectronic circuits, fuel cells and catalysis. The unique properties of metal oxides are due to their limited size and high density of edge surface sites[7]. The metal oxide nanoparticles such as magnesium oxide (MgO), gold (Au), titanium dioxide (TiO2), calcium oxide (CaO), silicon dioxide (SiO2), copper oxide (CuO) and silver oxide (Ag2O), are published as antimicrobial potential[8]. The antimicrobial potential of metal oxides nanoparticles has been recognized to their smaller size and higher surface to volume ratio, which enables them to closely bind with microbial strain but it does not due to the discharge of metal ions in solution[9].

Different methods have been used to synthesize nanoparticles like sol-gel method, surfactant mediated method[10], deposition-precipitation method, anodization method, wet-oxidation method[11], microwave-assisted combustion, thermal evaporation
Activities of Zinc-Silicon Oxides Nanocomposite

template methods\[11\], electrodeposition and sonication\[12\]. Metal oxide nanoparticles synthesized by any of these mentioned methods are used against bacterial strains by adopting various protocols\[8\]. The catalytic activity and antimicrobial properties of nanoparticles can be enhanced by doping the nanoparticles with various methods\[11\].

For a long time, scientist used antimicrobial drugs to inhibit and kill bacteria or other microbes but however they have been developed a specific microbial resistance over time. So one of the most promising strategy to get through this microbial resistance was to use nanoparticles\[13\]. Interestingly, ZnO-NPs are reported by several studies as non-toxic to human cells. This aspect necessitated their usage as antibacterial agents, noxious to microorganisms, and hold good biocompatibility to human cells. The various antibacterial mechanisms of nanomaterials are mostly attributed to their high specific surface area-to-volume ratios\[14\].

In this work we have synthesized ZnO-SiO\(_2\) nanocomposites via deposition-precipitation method by using acetonitrile solvent and tartaric acid as a stabilizing agent. Their antibacterial activity against Bacillus subtilis (gram positive) and E. Coli (gram negative) strains and antifungal activity against Candida parapsilosis and Aspergillus niger was determined. The major purpose of this study was to investigate the anti-bacterial and anti-fungal effects of nanocomposite against various strains. SiO\(_2\) was used to increase the photo-catalytic activity while ZnO was used to eliminate harmful bacteria\[15\].

**Experimental**

Synthesis of ZnO-SiO\(_2\) nanocomposite was carried out via deposition-precipitation method by using acetonitrile solvent. The chemicals used for experimental work include Zinc Nitrate Zn(NO\(_3\))\(_2\).6H\(_2\)O (Merck), Tartaric acid (Panreac), liquid Ammonia (Biom), Ethanol (Biom), Silicon dioxide (DAE-JUNG), Acetonitrile (PANREAC). The analytical instruments used for experimental work were Analytical Balance (Shimadzu), , FTIR (Shimadzu), XRD (Philips X’ Pert), FESEM (JEOL 7600), TEM (JEOL JEM 1010), TGA/SDT (G600 V8.3 Build 101), Furnace (VULCAN D-550), Oven (EV 108 Ac–Kapa), pH meter (Chemcadet 5986-62), Centrifuge machine (Sigma 1-14), DLS.

**Synthesis of ZnO-SiO\(_2\) nanocomposite**

ZnO-SiO\(_2\) nanocomposite was synthesized by using equimolar concentration of zinc nitrate Zn(NO\(_3\))\(_2\).6H\(_2\)O (0.2M) and tartaric acid (0.2M), each of them was dissolved separately in 10 mL of acetonitrile. After mixing both solutions, stirred the solution for 20 min (Ph = 2) at an elevated temperature 80°C. Now add 0.5g of SiO\(_2\) in the 20 mL of acetonitrile in a separate beaker and stirred for 20 min (pH = 7.2). Adjust the pH by drop-wise addition of NH\(_3\):H\(_2\)O in the prepared solution of SiO\(_2\) (pH = 9.3). Now mix the both solution. The suspension was refluxed for 6 h at 80°C and then centrifuged at 2500 rpm for 30 min. The precipitates were separated and washed several times with water and ethanol and dried at 100°C in the oven. The dried precipitate was annealed at 700°C for 3 h.

**Antibacterial activity**

Antibacterial activity of ZnO-SiO\(_2\) nanocomposite was studied against two bacterial strains Bacillus subtilis and Escherichia coli.

**Preparation of fresh bacterial culture**

Nutrient broth (Oxoid, UK) 100 mL was prepared in Erlenmeyer flask contained glass beads. pH was adjusted to 7.4 by addition of buffer solution and autoclaved at 121°C for 15 min. Mixture was allowed to cool in laminar air flow. Broth was inoculated with 100 µL bacteria form stored bacterial culture. Growth medium was incubated in shaker at 37°C for 24 h to get 5 × 10\(^{10}\) cells per mL.

**Disc diffusion method**

Nutrient agar was prepared by mixing agar medium in distilled water. Agar medium and 10 mm discs of wicks paper were sterilized in auto-clave and allowed to cool in laminar air flow. The agar medium was inoculated with 50 µL fresh bacterial culture and sterilized discs were poured with synthesized nanocomposite and spread in petri plates with positive control in the center. Incubate the petri dishes at 37°C for 24 h. Zones were measured with zone reader. Experiment was performed with both bacterial strains.

**Antifungal activity**

The antifungal activity was performed according to Devi, et al., 2014\[16\] with slight modification. Antifungal activity of the synthesized ZnO-SiO\(_2\) nanocomposite in acetonitrile solvent was determined using the agar well diffusion assay. Stock cultures of Candida parapsilosis and Aspergillus niger were prepared and maintained in Sabouraud Dextrose Agar (SDA) slants at 4°C. A positive control drug (Nystatin) was also done parallel. The plates were examined for evidence of zone of inhibition, which appear as area around the walls. The diameter of such zones of inhibition was measured using a meter ruler. Mean value was calculated by performing the experiments in triplicates.
Results and Discussion

Fourier Transform Infrared Spectroscopy (FT-IR)

Figure 1 shows the FT-IR spectrum of ZnO-SiO₂ nanocomposite. The broad absorption band at 3000-3600 cm⁻¹ and the peak at 1591 cm⁻¹ can be assigned to the stretching and bending vibrations of OH group of H₂O molecule respectively. The presence of these peaks in the spectrum was attributed the adsorption of atmospheric water on surface of nanocomposite during FTIR measurements. The small absorption peak at 2937 cm⁻¹ is due to the symmetric vibrations of CH group of tartaric acid which was used as a stabilizing agent[17]. The absorption peak at 2349 cm⁻¹ is the C=O stretching mode vibrations which is indicating the adsorption of atmospheric CO₂ on nanocomposite’s surface[18]. The peak at 1988 cm⁻¹ is due to stretching vibrations of C-N bond[19]. The absorption peaks at 1402 and 1876 cm⁻¹ are the stretching vibrations of C-O and C=O bonds of carboxyl group respectively. The absorptions at 1049 cm⁻¹ and 1116 cm⁻¹ are due to disymmetric vibrations of Si-O-Si[20] and the absorption peak at 788 cm⁻¹ is assigned to Si-OH. The absorption peak at 698 cm⁻¹ shows the stretching vibrations of Si-O bond[17]. The absorption peak at 437 cm⁻¹ is attributed to stretching vibrations of Zn-O[18].

Figure 1: FT-IR spectra of 0.1M ZnO-SiO₂ nanocomposite

Powder X-Ray Diffraction (XRD)

Figure 2 shows the XRD pattern of 0.1M ZnO-SiO₂ nanocomposite synthesized by deposition-precipitation method in Acetonitrile. It can be seen from the figure that all the proposed samples are well indexed to the hexagonal phase of SiO₂ with reference code (01-089-8939). ZnO-SiO₂ nanocomposite synthesized in Acetonitrile shows the characteristics peaks of SiO₂ at 2θ = 20.934°, 26.789°, 36.191°, 39.489°, 42.30°, 45.59°, 54.77°, 60.057°, 67.882°, 73.90°, 75.36°, with diffraction planes (100), (101), (110), (102), (200), (201), (211), (212), (302), and (200) which are well indexed with [ICSD PDF00-001-0649] and [ICSD PDF database 00-001-0649] with Quartz structure. The ZnO-SiO₂ in addition to above diffraction peaks, some characteristics peaks of zinc oxide (ZnO) is also shown which are all well indexed with [ICSD PDF database01-079-0208] with hexagonal structure at 2θ =31.74, 34.31, 36.31, 50.42, 56.58 and 62.25 with diffraction peaks at (002), (100), (102), (110) and (103) respectively. XRD pattern was analyzed to determine the peak intensity, position and width.

Figure 2: XRD pattern of ZnO-SiO₂ nanocomposite synthesized in Acetonitrile solvent

Scanning Electron Microscopy (SEM)

Figure 3 shows the SEM analysis of ZnO-SiO₂ nanocomposite synthesized in acetonitrile. The micrograph shows coalesced flake-like mass. From the figure it can be deduced that ZnO-SiO₂ nanocomposite has small dimension and the structure becomes more detainted. The SEM micrograph shows that the larger particles mostly consist of silicon and oxygen and the flakes are mostly made up of zinc and oxygen. The ZnO-SiO₂ nanocomposite shows the spherical morphology with highly porous structure.

Figure 3: SEM image of ZnO-SiO₂ nanocomposite

Transmission Electron Microscopy (TEM)

To elucidate the size of ZnO-SiO₂ nanocomposite, TEM image was taken with JEOL JEM 1010. The image shows that ZnO is fused on the surface of SiO₂ uniformly. Figure 4 generates some amorphicity and agglomeration. These clusters create effect on both the porosity and on surface area of SiO₂, size of image was calculated by the following formula.

Size of image = Object size × Magnification power

The average particle size calculated was 6.2 nm.
Thermo Gravimetric Analysis (TGA)

The TGA-DSC curve represents weight loss of sample as a function of temperature for ZnO-SiO$_2$ nanocomposite shown in Figure 5. Four major weight losses were clearly evident. The first step involves the weight loss of 6% at 100°C which is due to removal of physically adsorbed water molecule. The second curve indicates 9% weight loss which is due to removal of carbon monoxide from tartaric acid at 300°C. Third major weight loss was 12% due to removal of two water molecules which elucidate the decomposition of zinc tartrate to ZnO$^{[21]}$. While weight loss at 700°C elucidated the temperature above which the vicinal hydroxyl group of silica were completely condensed$^{[22]}$.

Dynamic Light Scattering (DLS)

Dynamic Light Scattering (DLS) is an important instrument for characterizing the size of nanoparticles in solution. DLS deals with the light scattered from a laser that passes through a colloidal solution and by analyzing the modulation of the scattered light intensity as a function of time, the hydrodynamic size of particles and particle cumulation can be determined. DLS analysis is used to determine the hydrodynamic diameter of synthesized nanoparticles. Hydrodynamic diameter of the nanoparticles is always larger than their size due to presence of stabilizer at their surface. Figure 6 shows the number distribution of ZnO-SiO$_2$ nanocomposite synthesized by precipitation method by using acetonitrile solvent. The result clearly shows that the Z-Average value as the mean value of the hydrodynamic diameter is 19.6 nm and Peak-Intensity is 955. A second important number is the polydispersity index which is a measure of the width of the particle size distribution. Polydispersity indices less than 0.1 are typically referred to as “monodisperse”. PDI (0.06) is calculated by using following formula$^{[23]}$.

$$\text{PDI} = \left(\frac{\sigma}{d}\right)^2$$

Biological Applications

Antibacterial activity of ZnO-SiO$_2$ nanocomposite

To confirm the antibacterial activity of ZnO-SiO$_2$ nanocomposite, the growth inhibition of the two bacterial strains i.e. Bacillus subtilis and E. coli was investigated by Disc diffusion method. The prepared nanocomposite showed 6% activity against Bacillus subtilis and 4.5% activity against E. coli, as shown in the Figure 7.

Antifungal activity of ZnO-SiO$_2$ nanocomposite

The antifungal activity of ZnO-SiO$_2$ nanocomposite was performed with an average diameter of 25 nm against the tested yeasts diluted in Mueller Hinton Broth to the final concentrations of silicon dioxide. The obtained results showed that the ZnO-SiO$_2$ nanocomposite inhibited some of the tested Candida parapsilosis and Aspergillus niger. At very low concentrations...
the inhibition was dependent on the yeast species tested. The lowest MIC of ZnO-SiO$_2$ nanocomposite showed the antifungal activity at the concentration of 0.207 mg/L against Aspergillus niger and 0.17 mg/L against Candida parapsilosis as shown in Figure 8.

![Figure 8](image-url)

**Figure 8**: Antifungal activity of ZnO-SiO$_2$ nanocomposite

**Conclusion**

The purpose of present research work was to synthesize the ZnO-SiO$_2$ nanocomposite in acetonitrile solvent by deposition precipitation method. Different characterizations were carried out to confirm the size and shape of the prepared nanocomposite. ZnO-SiO$_2$ nanocomposite showed better antibacterial activity against Bacillus subtilis than E. coli and better antifungal activity against Aspergillus niger than Candida parapsilosis. Acetonitrile solvent showed greater antibacterial activity than antifungal activity.

**Acknowledgment**: The authors would like to highly acknowledge to the Higher Education Commission Islamabad for the financial assistance to conduct this research work.

**References**

1. Arshad, M., Farrukh, M.A., Imtiaz, A., et al. Solvent Assisted Synthesis of Tin-Zinc Oxide nanoparticles: Structural Characterization and Antimicrobial Activity. (2015) Asian J Chem 27(1): 371-374.
2. Afzal, M., Shahid, M., Jamil, A., et al. Phytochemical Spectrum of Essential Oil of Paganum harmalaby GC-MS and Antimicrobial Activity using Sequential Solvents Fractions and Essential Oil. (2014) Asian J Chem 26: 574-578.
3. Li, Q., Mahendra, S., Lyon, D.Y., et al. Antimicrobial nanomaterials for water disinfection and microbial control: potential applications and implications. (2008) Water Res 42(18): 4591-4602.
4. Ahmad, R., Mohsin, M., Ahmad, T., et al. Alpha amylase assisted synthesis of TiO$_2$ nanoparticles: structural characterization and application as antibacterial agents. (2015) J Hazard Mater 283: 171-177.

5. Azam, A., Ahmad, A.S., Oves, M., et al. Antimicrobial activity of metal oxide nanoparticles against Gram-positive and Gram-negative bacteria: a comparative study. (2012) Int J Nanomed 7: 6003-6009.
6. Ahmad, S., Farrukh, M.A., Khan, M., et al. Synthesis of Iron Oxide–Tin Oxide Nanoparticles and Evaluation of their Activities against Different Bacterial Strains. (2014) Can Chem Trans 2(2): 122-1343.
7. Farrukh, M.A., Shahid, M., Muneer, I., et al. Influence of gadolinium precursor on the enhanced red shift of Gd/SnO$_2$–TiO$_2$ nanoparticles and catalytic activity. (2016) J Mater Sci: Mater El 27(3): 2994-3002.
8. Zarei, M., Jannnejad, A., Khajehali, E. Antibacterial effect of silver nanoparticles against four foodborne pathogens. (2014) Jundishapur J Microb 7(1): e8720.
9. Chwalibog, A., Sadowsz, E., Hotowy, A., et al. Visualization of interaction between inorganic nanoparticles and bacteria or fungi. (2010) Int J Nanomed 6(5): 1085-1094.
10. Muneer, I., Farrukh, M.A., Javaid, S., et al. Synthesis of Gd$_2$O$_3$/Sm$_2$O$_3$ nanocomposite via sonication and hydrothermal methods and its optical properties. (2014) Superlattice Microst 77: 256-266.
11. Perveen, H., Farrukh, A.M., Khalique-ur-Rehman, M., et al. Synthesis, Structural Properties and Catalytic Activity of MgO–SnO$_2$ Nanocatalysts. (2015) Russ J Phys Chem 89(1): 99-107.
12. Naseem, T., Farrukh, A.M. Antibacterial Activity of Green Synthesis of Iron Nanoparticles Using Lawsonia inermis and Gardenia jasminoides Leaves Extract. (2015) J Chem 2015: 1-7.
13. Spiala, A., Albu, M.G., Ficai,A., et al. The SiO$_2$/ZnO composite materials for cosmetic creams. (2014) Dig J Nanomater Bios 9(4): 1729-1737.
14. Sirelkhatim,A., Mahmud, S., Seeni, A., et al. Review on Zinc Oxide Nanoparticles: Antibacterial Activity and Toxicity Mechanism. (2015) Nano-Microlet 7(3): 219–242.
15. Molseni, A., Maleknia, L., Fazaeri, R., et al. Synthesis TiO$_2$/SiO$_2$/Ag triple nanocomposite by sonochemical method and investigation of photo-catalyst effect in wastewater treatment. (2013) Nanocon 10:16-18.
16. Devi, J.S., Bhimba, B.V. Antibacterial and Antifungal Activity of Silver nanoparticles Synthesized using Hypnea muciformis. (2014) Biosci Biotechnol Res Asia 11(1): 235-238.
17. Fanga, G., Li, H., Liu, X. Preparation and properties of lauric acid/silicon dioxide composites as form-stable phase change materials for thermal energy storage. (2010) Mater Chem Phys 122(2-3): 533–536.
18. Gondal,M., Drmosh, Q., Yamani, Z., et al. Synthesis of ZnO nanoparticles by laser ablation in liquid and their annealing transformation into ZnO nanoparticles. (2009) Appl Surf Sci 256(1): 298–304.
19. Fawcett, W., Liu, G. A Study of Ion Pairing in Acetonitrile Solutions Containing Magnesium Perchlorate Using Attenuated Total Reflection FTIR Spectroscopy. (1992) J Phys Chem 4231-4236.