Recent Advances in Macrocyclic Drugs and Microwave-Assisted and/or Solid-Supported Synthesis of Macroycles

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Abstract: Macrocycles represent attractive candidates in organic synthesis and drug discovery. Since 2014, nineteen macrocyclic drugs, including three radiopharmaceuticals, have been approved by FDA for the treatment of bacterial and viral infections, cancer, obesity, immunosuppression, etc. As such, new synthetic methodologies and high throughput chemistry (e.g., microwave-assisted and/or solid-phase synthesis) to access various macrocycle entities have attracted great interest in this chemical space. This article serves as an update on our previous review related to macrocyclic drugs and new synthetic strategies toward macrocycles (Molecules, 2013, 18, 6230). In this work, I first reviewed recent FDA-approved macrocyclic drugs since 2014, followed by new advances in macrocycle synthesis using high throughput chemistry, including microwave-assisted and/or solid-supported macrocyclization strategies. Examples and highlights of macrocyclization include macrolactonization and macrolactamization, transition-metal catalyzed olefin ring-closure metathesis, intramolecular C–C and C–heteroatom cross-coupling, copper- or ruthenium-catalyzed azide–alkyne cycloaddition, intramolecular S_N_Ar or S_N_2 nucleophilic substitution, condensation reaction, and multi-component reaction-mediated macrocyclization, and covering the literature since 2010.

Keywords: high throughput chemistry; microwave; macrocyclization; macrocycle; macrocyclic drugs; solid-phase

1. Introduction and Recently Approved Macrocyclic Drugs

Macrocycles continue to serve as an important class of compounds and have had a profound impact on chemistry, biology, and medicine [1–3]. Because of their cyclic nature, conformational and configurational characteristics, along with template-induced preorganization associated with macrocycles [4], compared to small molecule drugs and large biologics, macrocycles can offer unique drug-like profiles such as favorable pharmacokinetic and pharmacodynamic (PK/PD) parameters and improved oral bioavailability, enhanced metabolic stability and cell permeability, increased binding affinity, and desirable conformational rigidity [1,5]. As such, new synthetic strategies and structure–activity relationship (SAR) studies of macrocyclic chemical entities and related natural products remain an attractive research area in this field [6–8].

We previously reviewed macrocyclic drugs and general synthetic strategies toward macrocycles [1]. Since 2014, a number of new macrocyclic drugs have been approved by the US Food and Drug Administration (FDA), and their chemical structures, brand, and generic drug names, and the year of the approval are shown in Figures 1 and 2. These include antibacterial agents Dalvance (dalbavancin) [9], Orbactiv or Kimyrsa (oritavancin) [10,11] and Aemcolo (rifamycin SV) [12,13]; antiviral agents Olysio (simeprevir) [14], vaniprevir, Viekira Pak or Viekira XR (ombitasvir/paritaprevir/ritonavir) [15], Technivie (ombitasvir/paritaprevir/ritonavir/dasabuvir) [16], Vosevi (sofosbuvir/velpatasvir/voxilaprevir) [17], Zepatier (elbasvir/grazoprevir) [18], and Mavyret (glecaprevir/pibrentasvir) [19], for the treatment of hepatitis C virus (HCV) infection; antidote and reversal agent Bridion (sugammadex) for neuromuscular...
blockade induced by rocuronium bromide and vecuronium bromide \[20\]; guanylate cyclase-C agonist Trulance (plecanatide) for the treatment of chronic idiopathic constipation (CIC) and irritable bowel syndrome with constipation (IBS-C) \[21\]; anthelminthic moxidectin \[22\]; anticancer agent Lorbrena (lorlatinib) for the treatment of anaplastic lymphoma kinase (ALK)-positive metastatic non-small cell lung cancer (NSCLC) \[23\]; Vyleesi (bremelanotide) for the treatment of hypoactive sexual desire disorder \[24\]; Incivree (setmelanotide) for chronic weight management \[25\]; and immunosuppressant Lupkynis (voclosporin) \[26\]. In addition, several macrocyclic dotatate-based radiopharmaceuticals have also been approved, such as diagnostic imaging agents Netspot (gallium Ga 68 dotatate) \[27\] and Detectnet (copper Cu 64 dotatate) for the detection of rare neuroendocrine tumors \[28\], along with Lutathera (lutetium Lu 177 dotatate) for the treatment of somatostatin receptor-positive gastroenteropancreatic neuroendocrine tumors (GEP-NETs) \[29\]. Their detailed physiochemical properties and PK/PD parameters of these newly FDA approved macrocyclic drugs are summarized in Table 1.

Furthermore, this review discusses and highlights new advances in macrocycle synthesis using high throughput chemistry, including microwave (MW)-assisted and/or solid-supported macrocyclization, and covering the literature since 2010. Microwave and solid-phase syntheses have played an important role in enhancing chemical synthesis efficiency, such as decreasing reaction time, improving reaction yield and product purity, and streamlining the workup and purification process \[30,31\]. These enabling and accelerating technologies also play an integral role in organic chemistry, chemical biology, and drug discovery. New synthetic methods were broken down into macrolactonization and macrolactamization, transition-metal catalyzed olefin ring-closure metathesis, intramolecular C–C and C–heteroatom cross-coupling, copper- or ruthenium-catalyzed azide–alkyne cycloaddition, intramolecular $S_N 1$ or $S_N 2$ nucleophilic substitution, condensation reaction, and multi-component reaction-mediated macrocyclization.
Figure 1. Macrocyclic drugs approved by the US FDA since 2014.
Figure 2. Macrocyclic radiopharmaceuticals approved by the US FDA since 2014.
Table 1. Key attributes, physiochemical properties, and PK parameters [32,33] of FDA-approved macrocyclic drugs since 2014.

| Macrocyclic Drugs | Drug Class | MoA | MWt (g/mol) | logP | HBA | HBD | PSA (Å²) | RoA and Dosing Frequency | T_max [h] | Bioavailability (%) | Protein Binding (%) | Vd (L) | Metabolism | Excretion (%) | Developed and/or Marketed by | Initial US Approval |
|-------------------|------------|-----|-------------|------|-----|-----|----------|---------------------------|----------|---------------------|---------------------|--------|------------|---------------|----------------------|-------------------|
| Olysio (simeprevir) [DSC] | Antiviral (HCV) | NS3/4A protease inhibitor | 749.94 | 6.10 | 12 | 2 | 194 | 150 mg capsule PO q24 h with food | 4–6 h [10–13 h, healthy volunteers; [41 h, HCV-infected patients] | 62 (single dose under fed conditions) | >99.9 | <1 | 91 | Johnson & Johnson | 22 November 2013 (discontinued on 25 May 2018) [34] |
| Dalvance (dalbavancin) | Antibacterial lipoglycopeptide | Inhibition of transpeptidation and cell wall synthesis | Mixture 1816.69 | 4.39 | 38 | 21 | 573 | 1500 mg IV (a single-dose) or 1000 and 500 mg IV (two doses 1 week apart) | [346 h] | 93 | 7–13 | hydroxy-dalbavancin (minor) | 33 (P), 12 (M) | AbbVie Inc. (formerly by Allergan plc) | 23 May 2014 |
| Orbactiv (oritavancin) | Antibacterial lipoglycopeptide | Inhibition of transglycosylation and transpeptidation and disruption of bacterial membrane integrity | 1793.10 | 3.84 | 36 | 22 | 561 | 1200 mg IV (a single-dose) | [245 h] | 85 | 87.6 | Not metabolized | <5 (P) | <1 (P) | Melinta Therapeutics, Inc. | 6 August 2014 (approved in Japan) |
| Vanihep (vaniprevir) | HCV | NS3/4A protease inhibitor | 757.94 | 3.90 | 14 | 3 | 189 | 150 mg PO q12h | | | | | | | | |

NOTES: MoA = Mode of Action; MWt = Molecular Weight; logP = Logarithm of Partition Coefficient; HBA = Hydrogen Bond Acceptor; HBD = Hydrogen Bond Donor; PSA = Polar Surface Area; RoA = Route of Administration; T_max = Time to Peak; Bioavailability = Percentage of Drug Absorbed; Protein Binding = Percentage of Drug Bound to Plasma Proteins; Vd = Volume of Distribution; Metabolism = Pathway(s) of Drug Metabolism; Excretion = Percentage of Drug Excreted via Urine and/or Feces; Developed and/or Marketed by = Company(s) Developing or Marketing the Drug; Initial US Approval = Date of Initial Approval in the United States.
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Table 1. Cont.

| Macrocyclic Drugs | Drug Class | MoA | MWt (g/mol) | logP | HBA | HBD | PSA (Å²) | RoA and Dosing Frequency | Tmax \text{[t}_{1/2}] (h) | Bioavailability (%) | Protein Binding (%) | Vd (L) | Metabolism | Excretion (%) | Developed and/or Marketed by | Initial US Approval |
|--------------------|------------|-----|-------------|------|-----|-----|---------|--------------------------|-------------------|-------------------|-------------------|-------|------------|----------------|-----------------------------|-------------------|
| Viekira Pak or XR [DSC] (ombitasvir/paritaprevir/ritonavir/dasabuvir); Technivie ombitasvir/paritaprevir/ritonavir [DSC] | HCV | NS3/4A protease inhibitor | 765.88 | 5.13 | 14 | 3 | 198 | A fixed-dose combination product containing paritaprevir 50 or 75 mg PO two tablets (q24h) | 4.5 h [5.5 h] | 53 | 97-98.6 | 103 (Vss) | Metabolized by CYP3A4 and to a lesser extent CYP3A5 | 8.8 | 88 | AbbVie Inc. | 19 December 2014 (24 July 2015 for Technivie) (discontinued on 22 May 2018) [16] |
| Bridion (sugammadex) | Reversal agent for neuromuscular blockade | Antidote and selective relaxant binding agent | 2002.15 | −11.30 | 48 | 24 | 972 | 2, 4, or 16 mg/kg IV push as a single dose | [2 h] and prolonged in renal impairment | Negligible | 11–14 | Not metabolized | 95 (P) | Merck & Co., Inc. | 15 December 2015 |
| Zepatier (elbasvir/grazoprevir) | HCV | NS3/4A protease inhibitor | 766.90 | 4.37 | 15 | 3 | 204 | One tablet (50 mg/100 mg) PO (q24h) | 2 h [31 h] | 27 | 98.8 | 1250 | Hepatic (partial oxidative metabolism via CYP3A); metabolites not detected in plasma | <1 | >90 | Merck & Co. Inc. | 28 January 2016 |
| Trulance (plecanatide) | Chronic Idiopathic Constipation (CIC) and Irritable Bowel Syndrome with Constipation (IBS-C) | Guanylate cyclase-C agonist | 1681.89 | −2.81 | 44 | 26 | 819 | 3 mg PO once-daily | Minimal | Minimal | Proteolytic degradation (GI tract) | | Salix Pharmaceuticals, Inc | 19 January 2017 |
| Drug Class | MoA | MWt (g/mol) | logP | HBA | HBD | PSA (Å²) | RoA and Dosing Frequency | Tmax [t1/2] (h) | Bioavailability (%) | Protein Binding (%) | Vd (L) | Metabolism | Excretion (%) | Developed and/or Marketed by | Initial US Approval |
|------------|-----|-------------|------|-----|-----|---------|--------------------------|----------------|----------------------|-------------------|--------|------------|----------------|---------------------------------|---------------------|
| HCV        | NS3/4A protease inhibitor | 868.93 | 3.60 | 15  | 3   | 204     | A fixed-dose combination tablet (400 mg/100 mg/100 mg) PO with food q24h | 4 h [33 h] | >99 | Gilead Sciences Inc | 18 July 2017 |
| HCV        | NS3/4A protease inhibitor binds to glutamate-gated chloride channels, gamma-aminobutyric acid (GABA) receptors, and/or APT-binding cassette transporters | 838.87 | 1.19 | 15  | 3   | 204     | 300 mg/120 mg PO q24h | 5 h [6 h] | 97.5 | Secondary to CYP3A | 0.7 | 92.1 | AbbVie Inc. | 3 August 2017 |
| Anthelmintic | | 639.82 | 7.50 | 9   | 2   | 116     | 8 mg PO as a single dose | 4 h [23.3 days] | 2421 | Minimal | 2 | (P) | Pfizer Inc. | 13 June 2018 |
| Anticancer (ALK+ metastatic NSCLC) | Anaplastic lymphoma kinase (ALK) inhibitor | 406.41 | 0.78 | 8   | 2   | 110     | 100 mg PO once daily | 1.2 h [24 h] | 81 | 66 | 305 (Vss) | 48 (<1, P) | 41 (~9, P) | Pfizer Inc. | 2 November 2018 |
| Macroyclic Drugs | Drug Class | MoA | MWt (g/mol) | logP | HBA | HBD | PSA (Å²) | RoA and Dosing Frequency | Tmax [h_T/2] (h) | Bioavailability (%) | Protein Binding (%) | Vd (L) | Metabolism | Excretion (%) | Travelled and/or Marketed by | Developed and/or Marketed by | Initial US Approval |
|------------------|------------|-----|-------------|------|-----|-----|----------|--------------------------|-----------------|---------------------|-------------------|--------|------------|----------------|-----------------------------|-----------------------|------------------------|
| **Aemcolo** (rifamycin SV) | Antibacterial | Protein synthesis inhibitor by binding to the β-subunit of bacterial DNA-dependent RNA polymerase | 697.77 | 1.52 | 13 | 6 | 201 | 388 mg (two tablets) PO twice daily | <0.1 | 80 | Not expected | 86 | 86 | Aries Pharmaceuticals, Inc. | 16 November 2018 |
| **Vyleesi** (bremelanotide) | Hypoactive sexual desire disorder | Melanocortin receptor agonist 1.75 mg SubQ as one dose (maximum: 1.75 mg within 24 h). No more than 8 doses per month | 1025.16 | 1.83 | 24 | 15 | 376 | 1 h [2.7 h] | ~100 | 21 | 25 ± 5.8 | 64.8 | 22.8 | Palatin Technologies | 21 June 2019 |
| **Imcivree** (setmelanotide) | Obesity and the control of hunger associated with pro-opiomelanocortin deficiency | Melanocortin 4 (MC4) receptor agonist Starting dose 2 mg SubQ q24h for 2 weeks, then 1 or 3 mg SubQ q24h | 1117.31 | −0.08 | 27 | 20 | 495 | 8 h [11 h] | 79.1 | 48.7 | 39 (P) | Rhythm Pharmaceuticals, Inc. | 25 November 2020 |
| **Lupkynis** (voclosporin) | Immunosuppressant | Calcineurin inhibitor PO 23.7 mg (q12h) | 1214.62 | 2.89 | 23 | 5 | 279 | 1.5 h [30 h] | 97 | 2154 (Vss/F) | Primarily hepatic via CYP3A4 2 (c1, P) | 93 (5, P) | Aurinia Pharmaceuticals Inc. | 22 January 2021 |
| Macrocyclic Drugs         | Drug Class                        | MoA                                                            | MWt (g/mol) | logP | HBA | HBD | PSA (Å²) | RoA and Dosing Frequency | Tmax [h₁/₂] (h) | Bioavailability (%) | Protein Binding (%) | Vd (L) | Metabolism | Excretion (%) | Developed and/or Marketed by | Initial US Approval |
|--------------------------|----------------------------------|----------------------------------------------------------------|-------------|------|-----|-----|----------|--------------------------|----------------|---------------------|---------------------|---------|------------|--------------|--------------------------------|------------------|
| Netspot (gallium Ga 68 dotatate) | Diagnostic imaging agents       | Somatostatin type 2 (sstr2) receptor binding agent              | 1503.56     |      |     |     |          | 2 MBq/kg (0.054 mCi/kg) of body weight up to 200 MBq (5.4 mCi) IV bolus | [1.1 h]         |                     |                    |         | Extensive |              | Advanced Accelerator Applications | 1 June 2016       |
| Lutathera (lutetium Lu 177 dotatate) | Somatostatin receptor-positive gastroenteropancreatic neuroendocrine tumors (GEP-NETs). | Somatostatin type 2 (sstr2) receptor binding agent              | 1609.55     |      |     |     |          | 7.4 GBq (200 mCi) IV q8 weeks for a total of 4 doses | [71 ± 26 h] | 43 (non-radioactive form) | 460                 |         | Primarily renal |              | Advanced Accelerator Applications | 26 January 2018   |
| Detectnet (copper Cu 64 dotatate) | Diagnostic imaging agents       | Somatostatin type 2 (sstr2) receptor binding agent              | 1497.55     |      |     |     |          | 148 MBq (4 mCi) IV bolus | [12.7 h]         |                     |                    |         | Extensive |              | Curium US LLC                     | 3 September 2020   |

Lipinski properties were obtained from Scifinder and calculated using Advanced Chemistry Development (ACD/Labs) Software V11.02 © 2022–2021 ACD/Labs). MWt refers to the molecular weight of the free acid or base drug form. HBA, hydrogen bond acceptor; HBD, hydrogen bond donor; PSA, polar surface area. All PK parameters refer to adults unless otherwise noted. PO: by mouth or oral. IV: intravenous. Vd: volume of distribution. Under excretion: P refers to approximate % of total dose as parent drug and M refers to approximate % of total dose as metabolites. DSC: discontinued (brand name and/or generic product in the US).
2. Microwave-Assisted and/or Solid-Supported Synthesis of Macrocycles

2.1. Macrolactonization and Macrolactamization

Caporale et al. reported the synthesis and evaluation of a novel, cyclic parathyroid hormone fragment analog via side-chain cyclization of serine residues in 2010 [35]. On-resin diester bridge formation of 1 was performed using adipic acid via acylation of the hydroxyl groups of Serines 6 and 10 (Scheme 1). Specifically, following selective deprotection of the trityl groups from the Ser residues in 1 with 1% TFA/CH2Cl2, solid-supported peptide 2 with a cross-linked side chain was obtained via esterification with adipic acid upon in situ activation in the presence of HATU/HOAt/collidine [35].

In order to extend their previous work in the synthesis of aza crowns [36], in 2012, Rostami et al. reported the MW-assisted synthesis of new aza thia crowns [37]. Specifically, aza thia crowns 5 were synthesized from α, α-bis(bromomethyl) benzene 3, and methylthioglycolate, followed by the reaction of this diester 4 and diamines in ethyleneglycol under MW irradiation at 100 °C for 9 min in 74–91% yields (Scheme 2). In contrast, conventional reflux in methanol for 24 h gave much lower yields (9–19%) [37].

Cini et al. reported the MW-assisted synthesis of conformationally constrained peptidomimetics in 2012 [38]. The cyclopentapeptide analog 7 bearing an arginylglycylaspartic acid (RGD) motif, along with an enantiopure 7-substituted azepane-2-carboxylic acid (ACA) linker, was synthesized from a linear counterpart 6 in the presence of HATU/DIEA in dichloromethane (0.7 mM) at 75 °C under MW irradiation (25 W) for 25 min in 79% yield, followed by global side-chain deprotection in TFA/thioanisole/H2O (90/5/5) at room temperature for 3 h (Scheme 3) [38]. Macrocycle 7 showed low micromolar affinity towards ανβ3 and ανβ5 receptors with IC50 values of 1.8 and 2.9 µM, respectively [38].
Ferrie et al. reported the solid-phase synthesis and a comparative protease stability study of macrocyclic hexapeptides to mimic two endocrine hormones in 2013 [39]. Cyclohexapeptides 11 were synthesized using Rink resin and standard Fmoc amino acid coupling protocol, deprotection of the allyl group in 8, on-resin macrolactamization of 9, resin cleavage to provide 11 (Scheme 4) [39]. These endocrine peptide mimics showed improved stability profiles relative to naturally occurring vasopressin, oxytocin, and a linear control peptide.

Tao et al. reported the synthesis of azole cyclopeptide analogs via an on-resin cyclization–cleavage strategy in 2013 [40]. A selected example 13 from solid-supported linear precursor 12 is shown in Scheme 5, and using this developed solid-phase-based cyclitive–cleavage strategy, a chemical library was synthesized efficiently to produce >100 diverse azole cyclopeptide derivatives with various ring sizes, as modulators of multidrug resistance efflux pumps [40].

Scheme 3. MW-assisted synthesis of a pentapeptide RGD analog 7 [38].

Scheme 4. On-resin synthesis of cyclohexapeptides 11 [39].

Scheme 5. Synthesis of a representative azole-enriched cyclopeptide 13 via an on-resin cyclization–cleavage strategy [40].
Choi et al. reported a highly efficient pre-activation cyclization of the long peptide via succinimidyl ester-amine reaction strategy in 2015 [41]. After the formation of a pre-activated succinimidyl ester precursor, on-resin macrocyclization of the 25 AA peptide 15 was achieved effectively to yield cyclopeptide 16 (Scheme 6) [41].

Scheme 6. On-resin synthesis of cyclopeptides 16 via pre-activated N-hydroxysuccinimide (NHS) ester strategy [41].

Kumar et al. reported the MW-assisted synthesis of the Ti/Zr(IV) metal complexes of 16/18-membered macrocycles as potential antibacterial and β-lactamase inhibitors in 2017 [42]. Specifically, 19 and 22 were synthesized from 3,4-diaminopyridine 17 or 3,5-diaminobenzoic acid 21 and phthalic acid 18 in the presence of anhydrous MgSO₄ and methanol under MW irradiation for 5–10 min, followed by the formations of Ti or Zr(IV) metal complexes 20 and 23 of these macrocycle ligands (Scheme 7). The Zr(IV) macrocycle complex, 23, showed good antibacterial activities against extended-spectrum β-lactamase (ESBL)-producing E. coli strains [42].

Calisir and Çiçek reported the MW-assisted synthesis of benzo-thio crown ethers and studies of their ion chelation properties in 2017 [43]. Benzo-thio crown ethers 26 were synthesized from 2,2′-dithiodibenzoyl chloride 24 and dithiol or diol 25 in pyridine–chloroform under MW irradiation at 100 °C for 1 h in 36–87% isolated yields, compared to 18–52% yields under conventional reflux for 3–7 days (Scheme 8) [43].

Moreira et al. reported the synthesis of macrocyclic daptomycin analogs via replacing the ester with an amide in 2019 [44]. A representative example is shown in Scheme 9. Specifically, macrocyclization of peptide 27 was performed using PyAOP/HOAt/2,4,6-collidine in DMF/DCM (3/1) containing 1% Triton™ X-100, followed by simultaneous resin cleavage and global deprotection to provide daptomycin analog 28 [44]. Previously, in 2016, Lohani et al. reported an alternative solid-supported synthesis of a daptomycin analog DapE12W13 only using single α-azido amino acid and on-resin macrocyclization was performed prior to the construction of the side chain containing the decanoyl tail [45]. In 2019, Itoh and Inoue reviewed solid-supported total synthesis of macrocyclic natural peptides with branched chains (e.g., polymyxin E2 and daptomycin) via four-dimensionally orthogonal protective group strategies [46].
Arbour et al. reported the on-resin and self-cleaving head-to-tail macrolactamization of unprotected peptides via mild N-acyl urea activation in 2019 [47]. The macrocyclization of unprotected N-acyl urea-linked peptides 29 was mediated by N-terminal cysteine (Scheme 10). The S-to-N acyl transfer reaction, a robust and high-yielding chemoselective ligation methodology to form an amide bond, is widely used in organic synthesis, medicinal chemistry, and chemical biology [48]. Specifically, this efficient S-to-N acyl transfer cascade process involves the initial formation of a reactive thioester intermediate, followed by the acyl transfer to the nucleophilic NH₂ group. In this work, diverse macrocycles such as tetra- and pentapeptides 30 and 31 and the intramolecular disulfide-linked 14-AA-peptides 32 (sunflower trypsin inhibitor 1) were synthesized and demonstrated, preventing head-to-tail dimer formation and hydrolysis of most substrates [47].
Scheme 9. On-resin synthesis of macrocyclic daptomycin analog via the amide coupling strategy [44].

Scheme 10. On-resin synthesis of cyclopeptides via selective displacement of MeNbz by Cys and acyl transfer strategy [47].

Qu et al. reported the synthesis of disulfide surrogate peptides with large-span surrogate bridges via a native-chemical-ligation (NCL)-assisted diaminodiacid (DADA) approach in 2020 [49]. As an example, on-resin macrocyclization of linear peptide 33 was performed using PyAOP/HOAt/NMM at 37 °C for 3 h (2 times) to provide resin-bound 16-AA-peptide 34 with a thioether linkage as a complex mixture (Scheme 11) [49]. The failure of this on-resin DADA cyclization strategy was presumably due to its large 15AA ring size. Subsequently, a more effective NCL-assisted DADA strategy was developed to achieve large ring macrocyclization in the solution phase and with an alternative cyclization site (Trp8-Cys9) [49].
Bérubé et al. reported the solid-phase total synthesis and antimalarial evaluation of macrocyclic octapeptide dominicin, isolated from a marine sponge in 2020 [50]. The on-resin cyclization–cleavage reaction of \( \text{35} \) was performed using LiBr (5 eq) and DIPEA (2.5 eq) in \( \text{CH}_2\text{Cl}_2/\text{THF} \) (20:1), providing \( \text{36a} \) in a 58% yield as a cyclic monomer (>99%), based on the initial resin loading (Scheme 12). Subsequent catalytic Pd/C hydrogenation in acetic acid to remove the benzyl group on the side chain provided dominicin (\( \text{36b} \)) in a 53% yield after preparative HPLC purification. An alternative methodology was also developed using entirely on-resin synthesis and biorthogonal protection in \( N\)-Boc-L-Thr(OtBu)-OH [50]. In 2018, Bérubé et al. reported the on-resin synthesis of pseudacyclins A–E via a similar head-to-side chain concomitant cyclization–cleavage strategy [51].

On-resin macrocyclizations of hexapeptides were previously reviewed by Prior et al. [52]; additional cyclopeptides include those synthesized via standard macrocyclization amide coupling strategy [53–59], synthetic peptide macrocycles with two-strand forming segments [60], on-resin macrolactamization [61], dianinodiacid-based macrocyclopeptides with a 1,2,3-triazole linker to mimic the disulfide bond [62], a cyclic peptide with a side chain-to-side chain cyclization and a central warhead residue [63], and solid-phase synthesis of radiolabeled somatostatin octapeptide analogs with macrocyclic disulfide linkage and conjugated to a tetraazacyclododecane chelator motif [64]. In 2013, Kumarn et al. reported the synthesis of cycloheptapeptide integerrimide A via an on-resin tandem Fmoc-deprotection–macrocyclization strategy [65]. Moreover, in 2013, Thakkar et al. analyzed >2 million peptides to investigate on-resin cyclization efficiency with regard to various ring sizes, peptide sequence, and solvent [66].

Other on-resin natural product peptide syntheses include solid-phase synthesis of the lantibiotic lactocin S [67], on-resin macrocyclization of tetrapeptide namalide [68], cyclotheonamide analogs as human \( \beta \)-tryptase inhibitors [69], cyclotheonamide E4 and analogs as potent and selective \( \beta \)-tryptase inhibitors [70], solid-phase synthesis of the proposed struc-
ture of cyclodepsipeptide coibamide A and its O-desmethyl derivative via on-resin macrocyclization [71], side chain to side chain cyclized opioid endomorphin-2 peptide analogs [72], on-resin synthesis and evaluation of the antibiotic lysocin E and analogs [73], on-resin synthesis of rapamycin analogs [74], parallel solid-phase synthesis and anti-inflammatory evaluation of cyclic peptides cyclosquamosin D and Met-cherimolacyclopeptide B and analogs [75], and synthesis and evaluation of backbone cyclic peptides as HIV-1 protein–protein interaction inhibitors [76]. In 2021, Yoshida et al. reported the solid-phase synthesis and bioactivity evaluation of cherimolacyclopeptide E [77]. Additional on-resin MW-assisted macrocyclization of peptides via amide coupling was reported by Rashad et al. in 2015 [78] and Aneja et al. in 2019 [79].

2.2. Transition-Metal Catalyzed Olefin Ring-Closure Metathesis (RCM) Macrocyclization

Khan et al. reported the MW-assisted solid-phase synthesis of cyclic peptoids via the RCM strategy in 2011 [80]. Synthesis of 38 was performed from the model peptoid 37, a heptamer with 3-buten-1-amine at the first and last positions, using Hoveyda–Grubbs second-generation catalyst (2 mol%) in 1,2-dichlorobenzene under MW irradiation for 2 min ($4 \times 30$ s) (Scheme 13). The HPLC yield of the cyclic peptoid 38 as a mixture of $E/Z$ isomers was $\sim 80\%$ following the resin cleavage in 92% TFA [80]. Accordingly, cyclopeptoids 40 with different ring sizes were synthesized from 39 in 70–85% HPLC yields using the same MW RCM protocol. In contrast, the HPLC yields were slightly higher when the reactions were performed under conventional heating in dichloromethane at $40^\circ$C for 2 h, but with longer reaction times [80].

![Scheme 13. Solid supported MW-assisted RCM of peptoids [80].](image)

Andersson et al. reported the synthesis and biological evaluation of low nanomolar macrocyclic inhibitors of insulin-regulated aminopeptidase (IRAP) via MW-assisted solid or solution-phase olefin RCM in 2011 [81]. Macrocycles 43-($E$) and 45-($E$), the most potent IRAP inhibitors with high stability against proteolysis by metallopeptidases, were synthesized from 41 and 44 on solid and solution phase, respectively (Scheme 14). The Wang resin was used in the MW-assisted solid-phase synthesis of 41, and the on-resin RCM was performed at $140^\circ$C using the Hoveyda–Grubbs second-generation catalyst and 1,2-dichloroethane (DCE) as solvent under microwave heating [81]. Interestingly, 43 was isolated from a complex reaction mixture and obtained in 2% yield, following isomerization of 41, ring contraction, and subsequent double bond migration [81]. Accordingly, macrocycles 45-($E$) and 45-($Z$) were obtained in 28% and 6% yields, respectively.
Scheme 14. On-resin or solution phase MW-assisted RCM cyclization of 14-membered 43 and 45 [81].

Baron et al. reported the MW-assisted synthesis of macrocyclic pseudopeptides via RCM cyclization–cleavage strategy using a cis-5-aminopent-3-enoic acid (cis-Apa) linker in 2011 [82]. Macrocycle 47 with an RGD motif was synthesized from 46 using Grubbs’ second-generation catalyst in dichloromethane at 100 °C under MW irradiation in 64% purity (Scheme 15). In contrast, when this reaction was performed at 60 °C for 5 h under MW irradiation without the catalyst, 47 was obtained in a higher 73% purity. Following reverse-phase chromatography purification, 47 was isolated as two configurational E and Z-isomers (3/1 ratio) in an overall 25% yield from the resin and a 58% cyclization yield from 46. In addition, compared to conventional heating, MW heating is more efficient with higher yields and shorter reaction times [82]. Macrocyclic RGD peptides may have the potential to function as potent and/or selective α5β1 and ανβ3 integrin antagonists [83].

Scheme 15. MW-assisted RCM cyclization–cleavage of pseudopeptide [82].

Lampa et al. reported the RCM macrocyclization of the P2 phenylglycine and the alkenylc P1′ and HCV NS3 protease inhibition in 2011 [84]. As an example, the RCM of diastereomeric starting material 48 using Hoveyda–Grubbs’ second-generation catalyst in dichloromethane at 130 °C under MW irradiation is shown in Scheme 16. p-Benzoquinone was also added in the reaction to minimize the formations of ring-contracted and double bond migration side products. Four RCM products, 49-S, 49-R, 50-S, and 50-R, were obtained as stereoisomers R/S and configurational E and Z-isomers. In order to evaluate the reaction outcome, the impact of different methods (e.g., increasing temperature and MW irradiation) was measured, and it was found that the substrate appeared to play a more important role in affecting reaction outcomes relative to the cyclization method [84].
Raymond et al. reported the MW-assisted macrocyclic olefin metathesis at high concentrations using a phase-separation strategy in 2014 [85]. In this work, a protocol to promote macrocyclic olefin metathesis was developed at relatively high concentrations (up to 60 mM) under MW irradiation. As examples, diverse macrocyclic scaffolds such as 52, 54, and 56 with different alkyl, aryl, or amino acid spacers were synthesized from their respective diene substrates in the presence of the Grubbs–Hoveyda second-generation catalyst and a mixture of PEG500(OMe2)/methyl t-butyl ether (MTBE) at 100 °C under MW heating for 2 h in 60–78% yields (Scheme 17).

Qian et al. reported the synthesis of imidazolium-containing phosphopeptide macrocycles via neighbor-directed histidine N(τ)-alkylation in 2015 [86]. MW-assisted RCM reaction of 57 was performed on a solid support and by leveraging the Pro residue with a pentenyloxy side chain and the adjacent N(τ)-alkenyl group to produce various 20-, 22-, or 24-membered macrocycles 58, using second-generation Hoveyda–Grubbs catalyst in CH2Cl2/DMF (10/1) under microwave irradiation at 120 °C for 30 min (Scheme 18). For this reaction, a solvent combination of DMF and CH2Cl2 is important for desirable peptide solubility, MW absorption, and resin swelling [86]. Syntheses of charge-masked macrocyclic phosphopeptides 58, 60, and 62 and subsequent derivatives bearing the bis-alkyl-His imidazolium ring have the application potential toward biologically important protein-protein interactions.
Scheme 17. MW-assisted RCM macrocyclizations at high concentrations [85].

Scheme 18. Solid-supported MW-assisted RCM synthesis of charge-masked macrocyclic phosphopeptides [86].
Arndt et al. reported a divergent on-resin synthesis of macrocyclic depsipeptide seragamide A via relay-ring-closing metathesis (RRCM) strategy in 2015 [87]. Specifically, an on-resin RRCM reaction of 63 was performed using a Ru-based catalyst (25 mol%) in refluxing toluene at 110 °C for 3 h to provide the protected natural product 64 in 34% yield. Following sequential O-tBu and TIPS-O deprotection of individual 64 (E/Z) isomers in TFA and TBAF, seragamide A 65 (E), and its Z-olefin isomer was obtained in 65 and 59% yields, respectively (Scheme 19) [87].

Sousbie et al. reported the discovery, synthesis, and chemistry optimization of macrocyclic neurotensin analogs with potent analgesic effects in 2018 [88,89]. RCM macrocyclizations of 66 and 68 were performed under microwave heating at 50 °C using Hoveyda–Grubbs second-generation catalyst and p-benzoquinone in DCE for 1 h, followed by the acetyl deprotection for 66 using 20% piperidine in DMF and simultaneous deprotection of the nosyl and acetyl groups for 68 using 2-mercaptoethanol and DBU, and final resin cleavage and side-chain deprotection in TFA/DCM/TIPS (Scheme 20). Two promising macrocycle candidates, 67 and 69, with low nanomolar potency and/or good plasma stability, showed in vivo efficacy in two rodent pain models [88].

Morin et al. reported the synthesis and evaluation of a renewable macrocyclic musk using batch, MW, and continuous flow methodologies in 2019 [90]. At 10 mM concentration of the diene 70 in toluene, macrocycle 71 was synthesized in 32% yield under MW irradiation in the presence of Stewart–Grubbs catalyst at 70 °C for 60 min (Scheme 21). In contrast, batch synthesis at 21 °C for 5 days and continuous flow strategy at 150 °C for 5 min gave 47% and 32% yields, respectively, offering the advantage of facile scale-up (>1 g) in both cases [90].

Guo et al. reported the synthesis and evaluation of rapamycin-inspired macrocycles with new target specificity in 2019 [91]. A general synthetic route to rapafucin is shown in Scheme 22. Specifically, on-resin MW-assisted RCM cyclization–cleavage was performed from 72 using Hoveyda–Grubbs catalyst II (30 mol%) in 1,2-dichloroethane at 140 °C for 0.5 h to provide macrocycles 73 [91].
Scheme 20. Solid-supported MW-assisted RCM reactions to produce 23- and 22-membered macrocycles 67 and 69 [88].

Scheme 21. The RCM macrocyclization of musk 71 via batch, MW, and continuous flow strategies [90].

Scheme 22. Solid-phase synthesis of rapafucin via on-resin RCM/cyclative release strategy [91].
protected peptide was performed upon treatment with DIPEA to provide solid-supported 77, followed by the conversion of thiourea to the desired guanidine and subsequent resin cleavage and side-chain deprotection to yield 78 (Scheme 23) [92].

Scheme 23. On-resin synthesis of macrocyclic peptide analogs via the RCM and thiourea/guanidine strategy [92].

Yang et al. reported the synthesis and evaluation of several series of macrocyclic peptides as reversible inhibitors of LSD1 with a novel binding mode in 2020 [93]. On-resin RCM was performed from the N-terminal Fmoc and the side-chain protected 79 using the Hoveyda–Grubbs catalyst under MW irradiation at 200 °C to provide 80 (Scheme 24). Following the Fmoc and side-chain deprotection and resin cleavage, target peptide 81 was obtained. Accordingly, after the removal of allyl and allyloxycarbonyl protecting groups of 82 using Pd(PPh₃)₄, on-resin lactamization of 83 was then performed using the HCTU coupling reagent, followed by the global deprotection and simultaneous resin cleavage with TFA to provide the lactamized peptide 82 [93].
Chartier et al. reported the design, synthesis, and evaluation of the first selective macrocyclic neurotensin (NT) receptor type 2 non-opioid analgesic in 2021 [94]. Macrocycle 88, the most potent IRAP inhibitor with high stability against proteolysis by metallopeptidases, was synthesized from 86 on solid-phase (Scheme 25). The 2-chlorotrityl chloride resin was used in the MW-assisted solid-phase synthesis of 86, and the on-resin RCM was performed between two allylglycine residues at 70 °C using the second generation Grubbs–Hoveyda catalyst and DCE as solvent under microwave heating in ~70% yield, judged by UPLC-MS [94]. Based on this developed methodology, a series of new NT macrocyclic analogs with various side chain to side chain cyclizations and therapeutic potential were synthesized to probe the chemical space associated with NT receptor binding, as well as the in vivo efficacy study in rodent pain models [94].

Trần et al. reported the synthesis and evaluation of a series of apelin-13 based macrocyclic peptide analogs as modulators and pharmacological tools in the cardiovascular system in 2018 [95]. As an example, MW-assisted RCM reaction of 89 was performed on a solid support to produce a 17-membered macrocycle 90 using second-generation Hoveyda–Grubbs catalyst in DCE under microwave irradiation at 120 °C for 10 min (Scheme 26) [95]. Notably, the cleaved and side-chain deprotected compound shows a comparable binding affinity and a longer half-life in vivo relative to its linear peptide analog [95].
Scheme 25. MW-assisted on-resin RCM cyclization of 14-membered lead candidate 88 [94].

Other on-resin RCM reactions include the synthesis of a linked amino acid mimetic macrocycle under MW heating by Maxwell et al. in 2013 [96], diverse macrocyclization strategies using on-resin RCM, lactamization and thiol alkylation [97], synthesis of novel echinocandin analogs via on-resin RCM or disulfide formation strategy [98], the synthesis of
peptides with olefin crosslinks [99], synthesis of peptide thioureas and thiazole-containing macrocycles via Ru-catalyzed RCM [100], and Grb2 SH3 domain-binding peptides [101].

2.3. Intramolecular C–C, and C–Heteroatom Coupling Reactions

Bédard and Collins reported the MW-accelerated macrocyclizations of diynes via Glaser–Hay oxidative coupling reactions in 2012 [102]. Efficient macrocyclizations of diynes 91, 93, and 95 were performed at high concentrations employing MW irradiation and a phase-separation strategy. The Glaser–Hay macrocyclizations were completed in 1–6 h under microwave heating to provide macrolactones 92, 94, and 96 (Scheme 27), in contrast to conventional heating for 2 days. In addition, macrocyclization concentrations under MW heating could be increased up to 100 mM, relative to traditional concentration levels (0.2 mM) [102].

![Scheme 27. MW-assisted Glaser–Hay macrocyclizations at high concentrations [102].](image)

Godin et al. reported the MW-accelerated Glaser–Hay macrocyclizations and application toward the synthesis of antiviral agent vaniprevir in 2017 [103]. Glaser–Hay macrocyclization of 97 was performed employing MW irradiation and a phase separation strategy to provide 98, a macrocyclic core of the hepatitis C virus NS3/5A protease inhibitor vaniprevir. The Glaser–Hay cyclization was completed in 6 h under MW heating in 50% yield (Scheme 28). In contrast, the phase separation/continuous flow protocol provided 98 in 4 h in a 64% yield [103].
In order to further extend their previous work on MW-assisted synthesis of biaryl cyclopeptides via solid-supported intramolecular Suzuki–Miyaura coupling [104], in 2012, Afonso et al. reported the MW-assisted solid-phase synthesis of biaryl cyclic peptides containing a 3-aryltyrosine motif [105]. On-resin Suzuki–Miyaura macrocyclizations of 99 and 101 were performed using Pd$_2$(dba)$_3$ (0.2 eq), SPhos (0.4 eq), and KF (4 eq) in DME/EtOH/H$_2$O (9/9/2) under MW irradiation at 120 °C for 30 min, followed by simultaneous resin cleavage and side-chain deprotection in TFA/TIPS/H$_2$O to provide 100 and 102 in 16% and 22% overall yields, respectively (Scheme 29). This methodology demonstrates the synthesis of biaryl cyclopeptides with a p-Phe-m-Tyr or m-Tyr-m-Tyr linkage via solid-phase intramolecular Suzuki–Miyaura cross-coupling [105]. In the same year, Meyer et al. reported the solid-phase synthesis of 14- to 21-membered m,m-bridged biaryl macrocyclic peptides via on-resin Suzuki–Miyaura cross-coupling [106].

Mendive-Tapia et al. reported new peptide architectures and the MW-assisted macrocyclization between Trp–Phe/Tyr residues via a selective Pd-catalyzed C–H arylation strategy...
in 2015 [107]. As examples, solid-supported macrocyclization of 103 was performed using Pd(OAc)$_2$ (0.05 eq), 2-nitrobenzoic acid (1.5 eq), and AgBF$_4$ (1 eq) in DMF under MW irradiation (250 W) at 90 °C for 20 min, followed by the deFmoc, resin cleavage, and solution phase cyclization to provide bicyclopeptide 105 in an 18% unoptimized yield (Scheme 30) [107]. Accordingly, one-step double arylation to biaryl–biaryl stapled peptide 107 was synthesized from the linear peptide 106, AgBF$_4$ (6 eq), pivalic acid (1.5 eq), and Pd(OAc)$_2$ (0.4 eq) under MW irradiation (250 W) at 90 °C for 20 min, provided the bicyclopeptide 107 [107]. This developed one-step simple protocol enabled access to novel constrained cyclopeptides (e.g., dimeric macrocycles, stapled bicyclopeptides, and biaryl–biaryl analogs) from their corresponding linear precursors, facilitating further application and property studies in chemical biology and medicinal chemistry. In 2012, Dong et al. reported the peptidic macrocyclization via Pd-catalyzed chemoselective indole C-2 arylation strategy [108].

Kemker et al. reported the MW-assisted macrocyclization of RGD peptides via Suzuki–Miyaura cross-coupling strategy in 2019 [109]. Solid-supported Suzuki–Miyaura macrocyclization of 108 was performed using Pd$_2$(dba)$_3$ (20 mol %), sSPhos (40 mol %), and KF (4 eq) in DME/EtOH/H$_2$O (9/9/1) at 120 °C under MW irradiation for 30 min, followed by simultaneous resin cleavage and side-chain deprotections in TFA/TIPS/H$_2$O (95/2.5/2.5) to provide 109 in a 41% overall yield (Scheme 31) [109]. Macrocycle 109 showed low nanomolar affinity toward $\alpha_v\beta_3$ with good selectivity and plasma stability [109].
Scheme 31. MW-assisted macrocyclization of RGD peptides via Suzuki–Miyaura coupling strategy [109].

Bai et al. reported on-resin macrocyclization via Pd-catalyzed site-selective C–H olefination strategy in 2020 [110]. Specifically, backbone-directed Trp(C2) and trifluorosulfonyl-amide (Tf)-directed Trp(C4) macrocyclization was developed. For example, solid-supported cyclization of 110 was performed using Pd(OAc)$_2$ (20 mol %), AcOH (6 eq), and O$_2$ (1 atm) in p-xylene at 100 °C for 24 h, followed by resin cleavage and subsequent esterification to provide 112 in 26% overall yield (Scheme 32) [110].

Scheme 32. On-resin synthesis of cyclic peptide via Pd-catalyzed site-selective C–H olefination strategy [110].
In order to build on their previous work employing MW-assisted Suzuki–Miyaura macrocyclization reaction [111], in 2020, Ng-Choi et al. reported the MW-assisted synthesis of biaryl cyclopeptides via on-resin intramolecular Suzuki–Miyaura coupling [112]. Suzuki–Miyaura macrocyclization of 113 was performed on solid-support using Pd$_2$(dba)$_3$ (0.2 eq), SPhos (0.4 eq), and KF (4 eq) in DME/EtOH/H$_2$O (9/9/2) at 120 or 140 °C under MW irradiation for 30 min, followed by simultaneous resin cleavage and side-chain deprotections in TFA/TIS/H$_2$O to provide 114 (Scheme 33). By using this developed methodology, several other analogs, including cyclolipopeptides 115 and 116, were synthesized [112].

![Scheme 33. MW-assisted synthesis of biaryl cyclic tripeptides via Suzuki–Miyaura cross-coupling [112].](image)

Shen et al. reported the MW-assisted intramolecular Ullmann reaction to synthesize diaryl ether macrocycles 121 and 123 in 2012 (Scheme 34); the MW-assisted Ullmann cyclizations performed more efficiently with higher yields and shorter reaction times compared to sealed pressure tube as well as conventional heating [113,114]. This class of macrocyclic diarylheptanoid analogs and an extended macrocycle library, produced via MW-assisted intramolecular Ullmann coupling, were tested for antibacterial activity, and several macrocycles with phenethyl- and n-hexylamino moieties showed antibacterial activity with minimum inhibitory concentrations (MICs) of 12.5–25 µg/mL against *M. tuberculosis*, and Gram-positive organisms [115].
Scheme 34. Microwave-assisted intra- and/or bi-molecular Ullmann macrocyclizations [114].

Chávez-Riveros et al. reported the synthesis and anti-inflammatory evaluation of fourteen 15-membered diphenylamine macrocycles in 2019 [116]. By using the optimized conditions, diphenylamine macrocycle 125 was synthesized from 124 under MW irradiation at 110 °C for 4 h and using Pd(dba)₂ as the catalyst, SPhos as the ligand, cesium carbonate as the base in toluene, and in a 71% yield (Scheme 35). Macrocycle 125 with a decylamino side chain showed the most potent anti-inflammatory activity (ID₅₀ = 0.18 µM) in a 12-O-tetradecanoylphorbol-13-acetate (TPA)-induced edema model with low cytotoxicity [116].

Scheme 35. MW-assisted synthesis of a 15-membered macrocycle via Buchwald–Hartwig N-C cross-coupling reaction [116].

2.4. Click Macrocyclization via Copper- or Ruthenium-Catalyzed Azide–Alkyne Cycloaddition

Krause et al. reported the MW-assisted macrocyclization of a pseudohexapeptide containing 1,5-disubstituted 1,2,3-triazole moieties, as well as its anion-binding properties in 2011 [117]. Synthesis of 127 was performed from 126 using the [Cp*RuCl]₄ catalyst in DMF under MW irradiation at 115 °C for 30 min in 10% isolated yield (Scheme 36). In the literature, very few macrocyclizations involve Ru-catalyzed azide–alkyne cycloadditions (RuAAC) to provide a cyclic pseudopeptide derivative [117].
Pacifico et al. reported the MW-assisted on-resin click macrocyclization to synthesize novel triazole bridged Urotensin-II peptidomimetics using RuAAC chemistry in 2017 [118]. Synthesis of 1,5-triazole bridged 129 was performed from 128 using [Cp*RuCl](cod) catalyst in DMF under MW irradiation at 60 °C for 5 h in quantitative yield (Scheme 37) [118].

Ingale and Dawson reported the solid-supported side-chain macrocyclization of complex peptides using the Cu-catalyzed azide–alkyne cycloaddition (CuAAC) strategy in 2011 [119]. In this work, a panel of 21 amino acid helical peptides, such as 131, were synthesized via on-resin intramolecular CuAAC (Scheme 38). Specifically, on-resin CuAAC click macrocyclization of 130 was performed at room temperature in the presence of CuBr (1 eq), sodium ascorbate (1 eq), and 2,6-lutidine (10 eq) in DIPEA/DMSO for 18 h, followed by side chain deprotection and resin cleavage to provide 131 in 65% recovered yield [119].

Chavez-Acevedo and Miranda reported a practical synthesis of novel tryptamine-based macrocycles using sequential 4-component Ugi reaction and MW-assisted click macrocyclization methodology in 2015 [120]. Macrocycles 133, with a peptoid motif, a 1,3-
substituted indole moiety, and a triazole heterocyclic ring, were synthesized from the Ugi reaction product 132 using CuBr (0.4 eq) and DBU in toluene [5 mM] under MW irradiation at 110 °C for 6 h, providing the click chemistry 18- to 21-membered macrocycles 133 in 38–92% yields (Scheme 39) [120].

Among numerous click reactions and applications, the radical-mediated thiol–ene reaction remains an attractive and robust click strategy in organic synthesis and polymer science because of its simplicity, high efficiency, and rapid conversion rate [121]. In 2010, Aimetti et al. reported on-resin peptide photocyclization via thiol–ene click strategy [122].

2.5. Intramolecular S_NAr or S_N2 Nucleophilic Substitution

Li et al. reported the MW-assisted concise synthesis and Ca^{2+}-mobilizing activity in T-lymphocytes of new nucleobase-simplified cyclic ADP-ribose (cADPR) analogs in 2010 [123]. Synthesis of 135 was performed from 134 in pyridine and I_2 under MW irradiation at 90 °C for 15 min in 95% yield (Scheme 40) [123]. Notably, under the optimized conditions and MW heating, the yield of the intermolecular pyrophosphoration byproduct 136 decreased to < 3%. Following the acetal deprotection of 135 in 50% HCOOH, the resultant free, cyclic nucleotide bearing the 2’ and 3’-OH groups functions as a membrane-permeable cADPR mimic with calcium release activity in intact T-lymphocytes [123]. By using similar MW-assisted intramolecular pyrophosphorylation methodology, some novel cADPR structural analogs were subsequently synthesized and evaluated for Ca^{2+}-mobilizing activity [124,125].

Lee et al. reported the design and solid-phase synthesis of triazine-tethered bicyclic peptoids as conformationally constrained peptidomimetics in 2011 [126]. The resin-bound 6 to 12 peptoid residues 137 was cyclized via double nucleophilic attack to provide new bicyclic peptoids 138 in the presence of DIEA in N-methylpyrrolidone (NMP) at 65 °C (Scheme 41) [126].
Peng et al. reported the design and synthesis of the helix B surface peptide of erythropoietin and analogs with improved renoprotective effect and metabolic profile in 2013 [127]. Specifically, the peptide precursor was synthesized on a solid support, followed by global side-chain deprotection and resin cleavage to yield linear peptide 139. Next, MW-assisted macrocyclization of 139 was performed in the presence of triethylamine in water/acetonitrile at 75 °C for 20 min to produce the thioether-cyclized peptide 140 (Scheme 42) [127].

Torrejos et al. reported the MW-assisted synthesis of symmetric and asymmetric dibenzo-crown ethers in 2014 [128]. Specifically, symmetric dibenzo-crown ethers 142 (n = 1) and 146 can also be synthesized via one-pot synthesis from catechol and dibromopropane or dichloroethyl ether under basic conditions and MW irradiation with shorter reaction times and fewer purification steps, whereas asymmetric dibenzo-crown ethers 142 (n = 0), 143, and 145 were obtained from two-step MW-assisted synthesis via the diphenol intermediates 141 and 144, with satisfactory yields (Scheme 43).

Hickey et al. reported the design and synthesis of $^{99m}$Tc/Re-containing macrocyclic peptides as integrated metal-centric peptidomimetic imaging probes in 2015 [129]. For example, the macrocyclic peptide 148 was synthesized on-resin from 147 and 2,6-bis(aminomethyl) pyridine DMF with TEA via a pyridyl tridentate chelation core strategy, followed by side chain deprotection, resin cleavage, and metal chelation with $[^{99m}\text{Tc}(CO)_3(H_2O)_3]^+$ to provide 149 (Scheme 44) [129].

Wilson et al. reported the solid-phase synthesis and biological evaluation of cyclopeptide aldehydes as potent and selective 20S proteasome inhibitors in 2016 [130]. The Weinreb amide resin-bound tetrapeptides 150 were cyclized via an intramolecular $S_N$Ar mechanism, followed by the resin cleavage using lithium aluminum hydride (LiAlH$_4$) to provide peptide aldehydes 151 (Scheme 45) [130]. The biaryl ether macrocycles 151 represent the first macrocyclic peptide aldehydes with high potency (Ki = 54.5 and 241 nM), cellular stability, and specificity for the proteasome.
Scheme 43. MW-assisted synthesis of dibenzo-crown ethers [128].

Scheme 44. On-resin synthesis of $^{99m}$Tc-containing macrocyclic peptide via a pyridyl tridentate chelation core strategy [129].
Kheirabadi et al. reported the synthesis of thioether- or amine-bridged macrocyclic peptides via a “catch–release” strategy in 2018 [131]. This innovative enabling technology was highly effective, producing three macrocyclic peptide libraries containing 5–20 amino acids and thioether- or amine-based macrocyclic linkers (Scheme 46) [131]. Following resin release in 0.1 M NH₃/MeOH, 5–20 AA membered cyclopeptides 154 were provided with very good purity.

Other diverse on-resin intramolecular SₐAr or S₂ nucleophilic halide substitution reactions include synthesis of macrocyclic peptides with both α-helix and polyproline helix motifs [132], synthesis of cyclic peptides via an S₂ intramolecular thioalkylation [133], synthesis of 19-membered thioether cyclic peptidomimetics [134], synthesis of thiazole-containing cyclopeptides via on-resin intramolecular thioalkylation [135], on-resin synthesis of lipidated macrocyclic, and bicyclic peptides via convenient and intramolecular halide substitution by a diamino acid [136]. In 2018, Zhang et al. reported the on-resin MW-assisted macrocyclization and discovery of bisthioether-stapled peptides with excellent proteolytic stability and potent inhibition of PRC2 catalytic activity [137]. In 2021, Roy et al. reported a powerful, high-throughput quality control assay for on-resin synthesis and analysis of macrocyclic DNA-encoded libraries via thioalkylation strategy [138].

### 2.6. Condensation Reactions

Ahmed et al. reported the MW-assisted synthesis and antibacterial evaluation of metal complexes with Schiff base 2, 6-pyridinedicarboxaldehydehydethiosemicarbazone (PDCTC) in 2014 [139]. Specifically, macrocyclic ligand 157 was synthesized from 2, 6-pyridinedicarboxaldehyde 155 and thiosemicarbazone 156 in dry ethanol under MW irradiation for 4–5 min, followed by the
formation of Cu, Co, and Ni (II), and Cr (III) metal complexes 158 of these macrocycle ligands (Scheme 47). The metal complexes are generally more active than their parent Schiff base ligand in antibacterial testing [139].

Scheme 47. MW-assisted synthesis of 18-membered macrocycle metal complexes [139].

Hakimi et al. reported the MW-assisted template synthesis, geometry studies, and photoluminescent properties of the diazacyclam-based macrocyclic copper complex in 2014 [140]. Specifically, the macrocyclic copper complex 160 of 1,3,6,10,12,15-hexamethylenecyclocicosane (L), [CuL(NO$_3$)$_2$], was synthesized from N-(2-aminoethyl)-1,3-diaminopropane 159, formaldehyde and copper(II) nitrate in one-pot under MW irradiation for 10 min in 85% yield (Scheme 48) [140]. Several other related metal complexes including [CuL]Br$_2$, [CuL]I$_2$, CuL·H$_2$O, and [[CuL(NO$_3$)$_2$]$_2$Hg$_5$(μ-Cl)$_8$(μ$_3$-Cl)$_2$]Cl$_2$ were also synthesized from 159 by replacing the nitrate ions with bromide, iodide or by reacting 160 with HgCl$_2$, respectively, and investigated in this work [140].

Scheme 48. MW-assisted synthesis of diazacyclam-based macrocyclic copper complex [140].

Wilson et al. reported the solid-phase total synthesis of scytonemide A employing Weinreb AM resin in 2018 [141]. Following Fmoc deprotection, macrocyclization of the protected heptapeptide 161 was achieved via spontaneous imine formation upon resin cleavage using a reducing agent lithium aluminum hydride (LAH) and subsequent aqueous workup (Scheme 49). The final deprotection of 162 was performed under TBAF in methanol, providing scytonemide A (163) in a 51% yield [141].

Scheme 49. Total synthesis of scytonemide A via a biomimetic solid-phase synthesis strategy on the Weinreb AM resin [141].
Kumar and Vashistha reported the MW-assisted synthesis of Co\textsuperscript{II}HMTAA-14/16 macrocycles and their nanocomposites with highly conductive carbon black in 2019 [142]. Specifically, 166 and 168 were synthesized from acetylacetone 165, cobalt(II) chloride hexahydrate, and 2,4-/3,4-diamino toluene 164 or 167 in methanol at 70 °C for 10 min under MW irradiation (Scheme 50). These macrocycles were further used to synthesize their nanocomposites in the presence of carbon black and ethanol under MW irradiation (350 W, 4 MPa, 120 °C) for 30 min [142].

Scheme 50. Microwave-assisted synthesis of Co\textsuperscript{II}HMTAA-14/16 macrocycles [142].

Guéret et al. reported the solid-phase synthesis of macrocyclic modalities containing peptide epitopes and natural product motifs via macrocyclization by imine formation and subsequent stereoselective 1,3-dipolar cycloaddition in 2020 [143]. Specifically, a one-pot solid-phase synthesis protocol was developed to deprotect the Fmoc group in 169 and generate Schiff base 170 using trimethyl orthoformate as the dehydrating agent. Subsequent intramolecular cyclization of 170 was performed to provide 171 via azomethine ylide formation and 1,3-dipolar cycloaddition in the presence of lithium bromide (Scheme 51). Following the removal of side-chain protecting groups and simultaneous release from the resin, the major diastereomer 172 of the desired cycloadducts was produced in overall 8–14% purified yields [143].

Scheme 51. On-resin synthesis of cyclic PepNats via the imine/cycloaddition strategy [143].

2.7. Multi-Component Reaction (MCR)-Mediated Macrocyclization

Treder et al. reported the first solid-phase synthesis of functionalized, macrocyclic peptidomimetics via three-component coupling mediated by aziridine aldehyde dimers in
2015 [144]. On-resin macrocyclization of 173 was achieved using aziridine aldehyde dimer and an appropriate isocyanide, providing 174, which was subsequently functionalized by the nucleophilic opening of the aziridine ring (Scheme 52). This developed methodology enables the rapid parallel synthesis of macrocyclic peptides 175 [144].

Morejón et al. reported the solution- and solid-phase synthesis of N-aryl-bridged cyclolipopeptides via the Ugi–Smiles multicomponent methodology in 2016 [145]. Solid-supported Ugi–Smiles macrocyclization of 3-nitrotyrosine-containing peptide 176 took place effectively in the presence of an isocyanide and an aldehyde, followed by simultaneous resin cleavage and side-chain de protections in TFA/TIPS/H₂O to provide 177 in good yields (Scheme 53) [145]. Solid-supported Ugi-4-component macrocyclization methodology was subsequently reported to synthesize polycationic cyclolipopeptides with stabilized helical structures and antimicrobial properties [146], short and medium-size canonical cyclopeptides with turn-inducing moieties [147], stabilized cyclic β-hairpins, and N-alkylated peptides with various exo-cyclic functionalization such as cationic or hydrophobic tails and bioconjugation handles to install fluorescent and affinity tags, etc. [148]. In 2019, Reguera and Rivera reviewed the applications of the MCR toolbox for peptide macrocyclization and stapling [149]. In 2021, Rivera et al. reviewed solid-phase multicomponent protocols for biopolymer assembly and derivatization (e.g., the use of MCRs in the traceless on-resin synthesis of cyclic peptides) [150]. New synthetic methodology via on-resin Ugi reaction-based macrocyclization of peptide side chains to exocyclic functionalized helical peptides was subsequently developed by the same research team [151].

Yamaguchi et al. reported the synthesis of conformationally stable macrocyclic peptides via the Petasis borono–Mannich cyclization strategy in 2019 [152]. A representative example is shown in Scheme 54. Specifically, the Petasis borono–Mannich macrocyclization was carried out from 178 and glyoxylic acid in DCM/HFIP at room temperature for 24 h, followed by resin cleavage in TFA/TIPS/H₂O at room temperature for 1 h to provide the desired product 180 [152].
Scheme 54. On-resin synthesis of macrocyclic peptides via the Petasis borono–Mannich cyclization strategy [152].

Ohm et al. reported the diversity-oriented A3-macrocyclization of cyclic peptides with different substitution, ring size, and shape as CD36 receptor modulators in 2021 [153]. Specifically, cyclic azapeptides 182 were synthesized from linear precursors 181 using A3-macrocyclization strategy in aqueous formaldehyde and Cul in DMSO, followed by the resin cleavage and side-chain deprotection using a TFA/TESS/H2O cocktail (Scheme 55) [153]. Previously, the same research group studied the dynamic chirality of this cyclic azapeptide class of allosteric CD36 modulators of macrophage-driven inflammation using the same macrocyclization approach [154], aza-propargylglycine installation via aza-amino acylation strategy [155], and diverse cyclic azapeptides as CD36-modulating peptidomimetics via A3-macrocyclization approach [156].

Scheme 55. Diversity-oriented on-resin synthesis of cyclic azapeptides via A3-macrocyclization strategy [153].

3. Conclusions

Macrocycles offer some key attributes and advantages over small molecules and natural products such as favorable PK/PD parameters and improved oral bioavailability, enhanced metabolic stability, and cell permeability, and thus play an important role in chemical biology and medicinal chemistry. Since 2014, nineteen macrocyclic drugs and radiopharmaceuticals have been approved by FDA in the therapeutic areas of bacterial and viral infections, oncology, immunosuppression, etc. Remarkably, among them, five macrocyclic NS3/4A protease inhibitors have been approved for the treatment of HCV infections either alone or in combination with other viral medications. However, several of these agents were discontinued from the US due to clinical practice changes or other reasons in the arena of chronic HCV treatment, including newer, safer, and/or more effective treatment options.

Undoubtedly, macrocycles continue to represent an exciting chemical class in clinical drug development pipelines, exemplified by a tri-macrocyclic phase II clinical candidate, a proprotein convertase subtilisin/kexin type 9 (PCSK9) inhibitor, for lipid-lowering therapy [157]. This tricyclic peptide PCSK9 inhibitor has the potential to be developed as an oral once-daily lipid-lowering agent as the only other two FDA-approved PCSK9 drugs, Praluent (alirocumab) and Repatha (evolocumab), are monoclonal antibodies (biologics), which are administered subcutaneously. Notably, it remains a challenge to develop macrocyclic
peptides with oral bioavailability for systemic use. Other macrocyclic orally bioavailable peptide drugs include immunosuppressants Gengraf, Neoral, Sandimmune (cyclosporine), and Lupkynis (voclosporin). In the case of macrocyclic peptide drug Trulance (plecanatide) with two intramolecular disulfide linkages, it is taken orally but with minimal GI absorption and thus used for the treatment of GI-related disorders (e.g., CIC and IBS-C). In this context, such other recently approved macrocyclic oral drugs with minimal absorption include antibacterial Aemcolo (rifamycin SV) for managing and treating travelers’ diarrhea and Dificid (fidaxomicin) for the treatment of adult patients with C. difficile infections [13]. In addition, several advanced macrocycle clinical candidates were developed using the macrocycle–antibiotic hybrid strategy [5].

Moreover, high throughput chemistry, modular synthesis, and new macrocyclization methodology enable facile, rapid synthesis, SAR, and new advances in macrocycle drug discovery. Specifically, microwave-assisted and/or on-resin macrocyclizations have a profoundly accelerated generation of macrocycle libraries. These were highlighted via various recent MW-assisted and/or solid-supported macrocyclization strategies, including macroclonization and macroclatamation, transition-metal catalyzed olefin ring-closure metathesis, intramolecular C–C and C–heteroatom cross-coupling, copper- or ruthenium-catalyzed azide–alkyne cycloaddition, intramolecular S_N_2 or S_N_2' nucleophilic substitution, condensation reactions, and multi-component reaction-mediated macrocyclization. Depending on the nature of reaction substrates and types, both MW irradiation and solid-phase synthesis can shorten reaction times and synthetic routes, enhance reaction efficiency with higher yields and fewer side products, improve catalyst performance, consume less solvent, and/or streamline the synthesis and overall purification process. Though in some cases, significant optimization studies are required to find the most optimal reaction conditions and reagents. In multiple cases, on-resin MW-assisted macrocyclizations were effectively applied to the macroclatamation of peptide and peptidomimetics, RCM, triazole click chemistry, and intramolecular C–C coupling. Other innovative macrocycle entities and new advances in synthetic methodology development include the MCR-mediated macrocyclizations. Collectively, this work reviews and showcases some exciting advances in the macrocycle space, and its significance and impact will continue to thrive in the areas of chemistry, biology, and medicine.

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**Abbreviations**

ACA: azepane-2-carboxylic acid. cADPR: cyclic ADP-ribose mimic. ALK: anaplastic lymphoma kinase. CIC: chronic idiopathic constipation. CuAAC: copper-catalyzed azide–alkyne cycloaddition. DBU: 1,8-diazabicyclo[5.4.0]undec-7-ene. DCC: N,N'-dicyclohexylcarbodiimide. DCE: 1,2-dichloroethane. DCM: dichloromethane. DDC: sodium diethyldithiocarbamate. DIEA/DIPEA: N,N-diisopropylethylamine. DME: 1,2-dimethoxyethane. DMF: dimethylformamide. DMSO: dimethyl sulfoxide. DSC: discontinued. ESBL: extended spectrum β-lactamase. EDC: 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide. FDA: Food and Drug Administration. GI: gastrointestinal. HATU: 2-(1H-7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyl uronium hexafluorophosphate. HBA: hydrogen bond acceptor. HBD: hydrogen bond donor. HCTU: O-(1H-6-chlorobenzotriazole-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate. HCV: hepatitis C virus. HFIP: hexafluoroisopropanol. HOAt: 1-hydroxy-7-azabenzotriazole. IBS-C: irritable bowel syndrome with constipation. IRAP: insulin-regulated aminopeptidase. IV: intravenous. LAH: lithium aluminum hydride. MBHA: 4-methylbenzhydrylamine. MCR: multi-component reaction. MICs: minimum inhibitory concentrations. MW: microwave. MWt: molecular weight. NMM: N-methylmorpholine. MTBE: methyl t-butyl ether. NT: neotensin. NHS: N-hydroxysuccinimide. NMP: N-methylpyrrolidone. NSCLC: non-small cell lung cancer. PCSK9: proprotein convertase subtilisin/kexin type 9. PD: pharma-
codynamic. PK: pharmacokinetic. PO: by mouth or oral. PRC2: polycomb repressive complex 2. PSA: polar surface area. PyAOP: (7-azabenzo[1,2-b:4,3-b′]dioxepin-5-yl)methanone hexafluorophosphate. q12h: every 12 h. q24h: every 24 h. RCM: ring-closing metathesis. PDCTC: 2,6-pyridinedicarboxaldehydedihydrolactone. RGD: arginylglycylaspartic acid. RCM: relay-ring-closing metathesis. RuAAC: ruthenium-catalyzed azide alkyne cycloaddition. SAR: structure–activity relationship. S$_4$:A$_r$: nucleophilic aromatic substitution. SPhoS: 2-dicyclohexylphosphino-2′,6′-dimethoxybiphenyl. sSPhoS: sodium 2′-dicyclohexylphosphino-2,6-dimethoxy-1,1′-biphenyl-3-sulfonate hydrate. TBAF: tetra-n-butylammonium fluoride. TEA: triethylamine. TES: triethysilane. TFA: trifluoroacetic acid. TIPS/TIS: triisopropylsilane. TMDA: tetramethylethlenediamine. TPA: 12-O-tetradecanoylphorbol-13-acetate. Vd: volume of distribution.

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