Global mtDNA genetic structure and hypothesized invasion history of a major pest of citrus, *Diaphorina citri* (Hemiptera: Liviidae)

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Abstract

The Asian citrus psyllid *Diaphorina citri* Kuwayama is a key pest of citrus as the vector of the bacterium causing the “huanglongbing” disease (HLB). To assess the global mtDNA population genetic structure, and possible dispersal history of the pest, we investigated genetic variation at the *COI* gene collating newly collected samples with all previously published data. Our dataset consists of 356 colonies from 106 geographic sites worldwide. High haplotype diversity (H-mean = 0.702 ± 0.017), low nucleotide diversity (π-mean = 0.003), and significant positive selection (Ka/Ks = 32.92) were observed. Forty-four haplotypes (Hap) were identified, clustered into two matrilines: Both occur in southeastern and southern Asia, North and South America, and Africa; lineages A and B also occur in eastern and western Asia, respectively. The most abundant haplotypes were Hap4 in lineage A (35.67%), and Hap9 in lineage B (41.29%). The haplotype network identified them as the ancestral haplotypes within their respective lineages. Analysis of molecular variance showed significant genetic structure (*F̂ST* = 0.62, *p* < .0001) between the lineages, and population genetic analysis suggests geographic structuring. We hypothesize a southern and/or southeastern Asia origin, three dispersal routes, and parallel expansions of two lineages. The hypothesized first route involved the expansion of lineage B from southern Asia into North America via West Asia. The second, the expansion of some lineage A individuals from Southeast Asia into East Asia, and the third involved both lineages from Southeast Asia spreading westward into Africa and subsequently into South America. To test these hypotheses and gain a deeper understanding of the global history of *D. citri*, more data-rich approaches will be necessary from the ample toolkit of next-generation sequencing (NGS). However, this study may serve to guide such sampling and in the development of biological control programs against the global pest *D. citri*.

Keywords

Asian citrus psyllid, biological control, global genetic structure, invasion history, phylogeography
1 | INTRODUCTION

Genetic structure is the nonrandom distribution of alleles or genotypes in space or time (Mahy, Vekemans, & Jacquemart, 1999). Studies on population genetic structure can estimate genetic diversity and gene flow among populations, infer ancestral populations, and population level phylogeography. Such information is essential for informed management of pest species (Porretta, Canestrelli, Bellini, Celli, & Urbanelli, 2007). In recent years, understanding of biological invasion mechanism has increasingly relied on knowledge of the genetic structure of invasive species (Lee, 2002). Such studies can help to illuminate the basic biology and ecology of invasive species, the relationships between the intrinsic genetic characteristics of biological invaders, and their successful invasions (Xu, Zhang, Lu, & Chen, 2001). Furthermore, they can aid in the identification of effective biological control agents (Hoy, 2013).

The Asian citrus psyllid Diaphorina citri Kuwayama (Hemiptera: Liviidae) (Figure 1) is a world-wild economic pest because it transmits the bacterial pathogen, Candidatus Liberibacter, that causes citrus greening disease (huanglongbing, HLB), considered one of the most serious diseases of citrus (da Graça, 2008). The psyllid was first described from Taiwan in 1907 (Halbert & Manjunath, 2004). However, the available evidence suggests that D. citri originates from the Indian subcontinent (Hollis, 1987) and has subsequently expanded to other subtropical and tropical regions of Asia and also to the Indian Ocean islands of Réunion and Mauritius (Waterhouse, 1998), and South America (Halbert & Núñez, 2004). Recently, it has reached Central American countries and southern USA (Halbert & Manjunath, 2004).

Environmental factors, such as temperature, humidity, host plants, and human activities, influence the survival and reproduction of D. citri (Wang, 2002; Waterhouse, 1998; Xu, Xia, & Ke, 1994; Yang, 1989; Zhao, 1987). However, D. citri is characterized by high reproductive output and rapid generation turnover (2–7 weeks) and may thus rapidly evolve in response to new environments after an invasion. If semi-isolated subpopulations acquire different ecological traits, such as in host preference, development time, pathogenicity, vector capacity, susceptibility to pesticides, and tolerance to heat, and other abiotic stressors (Carmichael et al., 2007; Jourdrie et al., 2010; Pinho, Harris, & Ferrand, 2007; Remais, Xiao, Akullian, Qiu, & Blair, 2011), then genetically divergence among these subpopulations may rapidly emerge. Genetic studies have the potential to trace the D. citri demographic history that determines population genetic diversity of the invasive species and thus to illuminate geographic origin and secondary translocation events. However, to date, the global genetic differentiation, phylogeographic relationships, and invasion history were not resolved mainly owing limited geographic scope and relatively small sample sizes of prior studies (Boykin et al., 2012; de León et al., 2011; Guidolin, Fresia, & Cônsoli, 2014; Lashkari et al., 2014).

To assess the mtDNA population genetic structure, and potential dispersal routes of D. citri, we study global mtDNA genetic differentiation and phylogeography. We sequenced partial fragments of mitochondrial COI gene of recently collected individuals invasive to China and augmented our sampling with sequences from GenBank covering nearly the entire geographic range of D. citri. The current dataset covers 106 distinct localities around the globe. It thus offers a more holistic insight into D. citri matriline history than have prior studies.

2 | MATERIALS AND METHODS

2.1 | Insect collection and sequencing

We sampled D. citri in China between 2015 and 2016. A total of 412 specimens were taken from 11 sampling sites (Table S1 in Appendix S1). Collected insects were stored in 95% alcohol at −20°C. In total, 48 samples representing all our sampling sites were sequenced for genetic analyses. Additional sequences of D. citri from Iran, Pakistan, Saudi Arabia, India, Vietnam, Thailand, Indonesia, China, Reunion, Mauritius, Guadeloupe, Puerto Rico, Mexico, USA, and Brazil were taken from GenBank. We analyzed the COI sequences of the pest from a total of 356 colonies from 106 geographic sites in Asia, Africa, and America (Figures 2 and 3, and Table S1 in Appendix S1).

The primers (DCITRI COI-L: 5′-AGGAGGTGGAGACCCACCT-3′; DCITRI COI-R: 5′-TCAAATGGGAGAGATTTT-3′, Boykin et al., 2012) were used to amplify the COI gene from D. citri. The 25 μl polymerase chain reaction (PCR) was run with the following temperature cycling profile: 5 min denaturation at 92°C followed by 35 cycles of 1 min at 92°C denaturation, 1 min at 53°C annealing, 1.5 min at 72°C extension and a final extension at 70°C for 10 min. The PCR reactions were composed of 0.1 mmol/L dNTPs, 2 pmol of each primer, 1.5 U Taq polymerase (Beijing TransGen Biotech Co., Ltd., China), 1× PCR buffer, 2.5 mmol/L MgCl2, and 20 ng of DNA. Amplification products were purified and sequenced with primers from the original PCR reactions on an ABI 3730 automatic sequencer (Applied Biosystems, Foster City, CA, USA), using BigDye technology. The chromatographs were interpreted with Phred and Phrap (Green, 2009; Green & Ewing, 2002) using the Chromasq module (Maddison & Maddison, 2011a) implemented in the Mesquite (Maddison & Maddison, 2011b). Quality threshold was set high, for base trimming to 49 and for calls of ambiguity minimum secondary peak fraction for ambiguity at 0.3. Sequences were checked for
errors and edited manually with the program BioEdit (Hall, 1999) and then aligned in both CLUSTALX (Jeanmougin, Thompson, Gouy, Higgins, & Gibson, 1998) under default parameters and MAFFT (http://mafft.cbrc.jp/alignment/server/) using the FFT-NS-i strategy to increase accuracy (Katoh, Kuma, Toh, & Miyata, 2005).

2.2 | Genetic diversity and differentiation
In total, 356 COI sequences of the Asian citrus psyllids from Asia, Africa, and America were included in the analyses. Pairwise genetic distances (p-distances) were calculated using the software MEGA.
v.5.05 (Tamura et al., 2011). Polymorphic sites, parsimony informative sites, and haplotype frequencies were obtained, and nucleotide (\(\pi\)) and haplotype (\(H\)) diversities were estimated as defined by Nei, Muruyama, and Chakraborty (1975) using the DnaSP ver. 5.10 software (Librado & Rozas, 2009). The genetic variability within the whole sample, and among lineages, was assessed by analysis of molecular variance (AMOVA) with the program Arlequin ver. 3.5.1.2 (Excoffier & Lischer, 2010). The estimation of nonsynonymous (\(K_a\)) and synonymous (\(K_s\)) substitution rates has been used as a way of understanding the evolutionary dynamics of protein-coding genes across populations (Fay & Wu, 2003; Ohta, 1995). \(K_s\) and \(K_a\) values were calculated in MEGA V.5 (Tamura et al., 2011) using the Kimura 2P model.

2.3 | Haplotype network construction and STRUCTURE simulation

A haplotype network was constructed to compare genetic relationships between geographic populations based on the genetic data. The unrooted minimum spanning tree (MST) was implemented in PopART (http://popart.otago.ac.nz) and was partitioned by defined seven populations from the regions: North America (NM); South America (SA); West Asia (WA); East Asia (EA); South Asia (SA); Southeast Asia (SEA); and Africa (AF).

The population substructure among the samples from the analyzed seven regions was inferred using the software STRUCTURE ver. 2.3.3 (Pritchard, Stephens, & Donnelly, 2000). We performed the analysis based on the single-nucleotide polymorphism (SNP) of COI datasets of all individuals sampled (\(K = 1–10\)) using the admixture model. Ten independent runs were evaluated for each \(k\). The total length of the Markov Chain was set to be 100,000, and the length of burn-in period was 20,000. We adopted the parsimony criterion, in which the lowest \(K\) capturing the most differentiation among populations was selected as the most likely \(K\) value (DiLeo, Row, & Lougheed, 2010).

2.4 | Demographic analysis

To test signatures of demographic changes in \(D.\ citri\), we first examined Tajima’s \(D\) (Posada, 2008) and Fu’s \(F_\text{S}\) (Clement, Posada, & Crandall, 2000) using Arlequin ver. 3.5.1.2 under the infinite site model with 10,000 coalescent simulations. Second, population demographic changes were also examined by estimating the Harpending's raggedness index (HR) based on mismatch distribution for each population with significance assessed using Arlequin by 1,000 permutations. A significant HR value (\(p < .05\)) is taken as evidence for rejecting the sudden population expansion model (Schneider & Excoffier, 1999). Third, the demographic history of whether \(D.\ citri\) experienced a range expansion was investigated by the spatial expansion model in Arlequin (Schneider & Excoffier, 1999). Mismatch distributions of pairwise differences among sequences were calculated with parametric bootstrapping (1,000 replicates). According to simulations, demographically stable or admixed populations must present a multimodal distribution, whereas populations that have undergone a recent expansion generally show a unimodal distribution (Rogers & Harpending, 1992).

3 | RESULT

3.1 | Genetic variability and differentiation

We succeeded in amplifying a 760-bp COI region for the specimens from China and found only one polymorphic site, which was nonsynonymous mutation site. Two unique haplotypes were identified

| TABLE 1 | Region, country of collections, number of individuals, haplotypes (number of individuals), and nucleotide and haplotype diversity of \(D.\ citri\) in each sampled region |
|-----------------|--------------------------|-----------------------|----------------------|----------------------|
| Region | Country of collections | Number of individuals | Haplotypes (number of individuals) | Nucleotide diversity (\(\pi\)) | Haplotype diversity ± SD (\(\sigma\)) |
| South Asia | India, Pakistan | 14 | H4(1), H5(4), H8(1), H9(8) | 0.00128 | 0.641 ± 0.097 |
| West Asia | Iran, Saudi Arabia | 15 | H9(15) | 0.00000 | 0.000 ± 0.000 |
| Southeast Asia | Vietnam, Thailand, Indonesia | 30 | H4(27), H9(3) | 0.00057 | 0.186 ± 0.088 |
| East Asia | China | 69 | H4(65), H6(1), H7(3) | 0.00040 | 0.191 ± 0.063 |
| North America | USA, Mexico, Guadeloupe, Puerto Rico | 133 | H1(1), H9(116), H12(1), H13(1), H14(11), H15(3) | 0.00169 | 0.307 ± 0.050 |
| South America | Brazil | 86 | H2(1), H3(1), H4(26), H9(4), H10(1), H11(1), H16(1), H17(4), H18(1), H19(7), H20(3), H21(1), H22(4), H23(1), H24(1), H25(1), H26(1), H27(1), H28(1), H29(4), H30(1), H31(1), H32(7), H33(1), H34(1), H35(1), H36(1), H37(1), H38(1), H39(1), H40(1), H41(1), H42(1), H43(1), H44(1) | 0.00537 | 0.892 ± 0.027 |
| Africa | Reunion, Mauritius | 9 | H4(8), H9(1) | 0.00068 | 0.222 ± 0.166 |
(GenBank Accession number: submitted), with relative frequencies of 93.75% and 6.25%, respectively. The COI gene comprised 468 bp when including data from GenBank. Forty-four haplotypes were identified with 31 polymorphic sites and 15 parsimony informative sites. The average p-distance among COI gene sequences was 0.003 (range from 0.000 to 0.017). High haplotype diversity (H-mean = 0.702 ± 0.017) and low nucleotide diversity (π-mean = 0.003) were identified within the whole sample. The average number of nucleotide differences (k) is 1.483.

Among the 44 unique haplotypes, most (95.45%) occurred only in a single locality, and the remaining 4.55% occurred in more than one locality (Table 1). Hap9 was the most abundant and widely distributed, representing 41.29% of the whole sample, and occurring in all regions except East Asia. Hap4 was the second most frequent haplotype (35.67%) and was found in four regions but not in West Asia or North America. Hap14 (3.09%) was only found in North America and the remaining haplotypes were restricted to one region (Table 1).

The overall nonhierarchical AMOVA test (Table 2) resulted in a significant \( F_{ST} \) value (\( F_{ST} = 0.50, p < .0001 \)), indicating the genetic structure among sites. The hierarchical AMOVA (Table 2) analysis between these two lineages resulted in a high \( F_{ST} = 0.62 (p < .0001) \), which demonstrates that global D. citri populations are genetically structured. The Ka/Ks ratio is 32.92 (much greater than 1) suggesting natural selection is acting to promote the fixation of advantageous mutations (positive selection).

### 3.2 Global relationships

A haplotype network analysis revealed the genetic structuring among global D. citri populations (Figure 2). All individuals of D. citri clustered into two matriline. Lineage A is comprised of 37 haplotypes from all regions expect for West Asia, forming a complex and intricate star-like connection pattern with Hap4 at the center. Hap4 is widespread in East Asia, Southeast Asia, South Asia, Africa, and South America yet absent from the North American and West Asian samples. Hap1, and Hap6 and Hap7 are from North America and East Asia, respectively. In the A lineage, other haplotypes were restricted to South America. For the lineage B, seven haplotypes were sampled from all regions except East Asia, showing a relatively simple star-like cluster pattern with Hap9 at the center. Rare haplotypes are one or two mutations from Hap9. Hap9 covered the entire range of lineage B, Hap5 and Hap8 were restricted to South Asia, while all other haplotypes came from North America. The probabilities of each individual belonging to a lineage were plotted by locality (Figure 3a), indicating geographic structuring. The STRUCTURE analysis (Figure 3b) confirmed the presence of two lineages genetically differentiated.

### 3.3 Demographic history

The whole sample and both lineages had significantly negative values of Fu’s \( F_{S} \) (for the whole sample: \( F_{S} = −27.853, p = .000 \); for the lineage A: \( F_{S} = −28.727, p = .000 \); for the lineage B: \( F_{S} = −13.020, p = .000 \)) and Tajima’s \( D \) (for the whole sample: \( D = −1.865, p = .005 \); for the lineage A: \( D = −1.713, p = .010 \); for the lineage B: \( D = −1.639, p = .017 \)). These statistical values are indicative of historical population expansions, genetic drift, and/or selection (Fu, 1997; Tajima, 1989). Furthermore, haplotype network analyses using the genetic data for D. citri further suggested rapid population expansion in that two universal haplotypes located at the central positions of two connected star-like clusters of rare haplotypes for both lineages (Figure 2). The whole sample and both lineages showed unimodal patterns of mismatch distribution curves and the strong biases toward low divergence values as distinguished by 0- and 1-nucleotide changes (Figure 4). Furthermore, none of the Harpending’s raggedness (HR) index of mismatch distribution was significant (Figure 4). These findings illustrated a relatively recent and rapid population expansion from a small number of colonizers (Rogers & Harpending, 1992) or a range expansion with high levels of migration between neighboring demes (Excoffier, 2004; Ray, Currat, & Excoffier, 2003).

### TABLE 2 Analysis of molecular variance (AMOVA) for Diaphorina citri samples using COI sequences

| Source of variation | df  | Sum of squares | Variance components | Percentage of variation |
|---------------------|-----|----------------|---------------------|------------------------|
| The whole sample    |     |                |                     |                        |
| Among populations   | 6   | 115.13         | 0.42                | 49.72                  |
| Within populations  | 349 | 148.12         | 0.42                | 50.28                  |
|                     |     |                | \( F_{ST} = 0.50, p = .00 \) |                        |
| Two lineages        |     |                |                     |                        |
| Among lineages      | 1   | 99.18          | 0.49                | 47.72                  |
| Among populations within lineages | 10 | 30.49 | 0.14 | 14.16 |
| Within lineages     | 344 | 133.58 | 0.39 | 38.12 |
|                     |     |                | \( F_{ST} = 0.27, p = .00 \) |                        |
|                     |     |                | \( F_{ST} = 0.62, p = .00 \) |                        |
|                     |     |                | \( F_{CT} = 0.48, p = .00 \) |                        |
4 | DISCUSSION

4.1 | Genetic diversity and differentiation

We present the most inclusive analysis of the mtDNA phylogeographic structure of *D. citri* to date, collating previously published COI data (Boykin et al., 2012; de León et al., 2011; Guidolin, Fresia, & Cônsoli, 2014; Lashkari et al., 2014) with novel samples from Asia. Our phylogenetic network analysis inferred by the COI dataset suggests two lineages of *D. citri* in the world, here dubbed A and B. The genetic diversity of lineage A was higher than that of lineage B. Moreover, different genetic diversity (H and \( \pi \)) levels for *D. citri* were seen in the seven analyzed regions. The psyllids from both East and West Asia, and North America showed low genetic diversity, results that could imply recent introductions events that may have consisted of as little as a single haplotype. We cannot rule out other processes such as reduction in mtDNA variation due to *Wolbachia* infections, male-biased dispersal, or other potential limitations of mtDNA data (Avise, 2000). However, our findings seem more likely to reflect colonization histories and are consistent with genetic diversity of introduced populations being low due to the small size of propagules and bottleneck effects (Nei et al., 1975). South America, we hypothesize, was colonized recently by *D. citri*, yet it displays high mtDNA diversity. This could be the result of multiple invasions and/or a relative longer expansion history, consistent with prior studies (Guidolin et al., 2014; Silva et al., 1968). The genetic diversity of *D. citri* has been studied in Iran and Pakistan (Lashkari et al., 2014), Brazil (Guidolin, Fresia, & Cônsoli, 2014), America (de León et al., 2011) and worldwide (Boykin et al., 2012) using the same COI region we have applied. However, the current study extends on prior studies by combining all existing COI data and further adds novel samples from various localities in China. Consequently, our analyses reveal a more comprehensive insight into global mtDNA diversity than prior studies. Boykin et al. (2012) identified only eight haplotypes from worldwide *D. citri* populations, compared to the 44 discussed here. Many of the haplotypes of *D. citri* we find abundant were not present in the study of Boykin et al. (2012), meaning that with the current analysis we have gained a deeper insight into the global genetic variation and distribution of the psyllid.

The results of the AMOVA analysis demonstrated significant geographic structuring of genetic variation among populations within as
well as between lineages. We also found strong genetic differentiation between the two lineages even though they do co-occur in some localities. These results indicate limited or no mtDNA gene flow between the lineages, and limited gene flow within lineages among geographically isolated populations. On this basis, we hypothesize that this species has limited natural (nonhuman mediated) dispersal capacity (at least females, Kobori, Nakata, Ohto, & Takasu, 2011), and therefore, local populations may be unique and amenable to site-specific containment strategies to reduce the citrus greening disease spread. This observed geographic structure of *D. citri* is consistent with a process of range expansion followed by isolation of populations, as demonstrated in Excoffier, Foll, and Petit (2009) studies.

The observed significant positive selection (Ka/Ks ≫ 1) here is interesting as invasion success may occur after severe population bottleneck events or even with singly mated female invasion depending on its heterozygosity level (Zayed, Constantin, & Parker, 2007). In recent years, massive pesticides were used to control *D. citri* in the citrus orchards (Lopes, Frare, Yamamoto, Ayres, & Barbosa, 2007; Yamamoto et al., 2009). Therefore, the pest has been undergoing the intense selective pressure. Additionally, rapidly changing environments in invasive regions, such as climate change, also produce strong selective pressure. These directional selection can quickly lead to the loss of haplotypes and the expansion of rare haplotypes with different fitness attributes, affecting management strategies to be adopted for controlling the pest *D. citri* and/or the citrus greening disease.

### 4.2 Dispersal history

Our mtDNA phylogeographic analysis suggests two haplotype lineages, each characterized by a common “ancestral” haplotype and several derivatives of it (Figure 2). Although several potential processes might yield such a pattern (as outlined above), we hypothesize based on these results that the two lineages have independent colonization histories from two separate ancestral haplotypes. We hypothesize three dispersal routes across the globe, in part showing parallel dispersal patterns for the two lineages in Southeast Asia, Africa, and South America. The genealogies based on network can provide insight into ancestral haplotypes, as these haplotypes are expected to be common, widely distributed and highly connected (Chen et al., 2010). In the study by Boykin et al. (2012), >60% of the analyzed individuals were sampled from the New World where *D. citri* is invasive. These sampling limitations hinder reconstruction of ancestral haplotypes and potential dispersal routes. Our analysis offers a broader and more balanced sampling of *D. citri* allowing a better prediction of ancestry and dispersal. The haplotype network analysis identified two ancestral haplotypes (Hap4, lineage A; Hap9, lineage B, Figure 2) that although genetically close are the unequivocal connection points of all other haplotypes, supporting our hypothesis of independent histories.

Mead (1977) listed the Far East as the geographic origin of the Asian citrus psyllid. More recent studies on biogeographic reconstructions inferred from origins of plant host and historical information indicate that the geographic origin of this psyllid may be southern Asia, probably India (Hall, 2008). Our findings are consistent with the latter; a southern and/or southeastern Asia origin. The Asian citrus psyllid is known to occur in China likely as an invasion within the last 80 years (Hoffmann, 1936). It colonized South America in the 1940s (Costa Lima, 1942). During the 1990s, *D. citri* invaded North America (French, Kahlke, & De Graca, 2001; Halbert & Núñez, 2004; Pluke, Qureshi, & Stansly, 2008; Tsai, Wang, & Liu, 2000).

The introductions and dispersal patterns of *D. citri* in the Americas were recently studied using COI data. de León et al. (2011) suggested that two separate invasions or founding events of the Asian citrus psyllid occurred respectively in South America and North America. Guidolin, Fresa and Cónsoli (2014) argued for two hypothetical scenarios of invasions for *D. citri* in Brazil. The first scenario is that the psyllid invaded from the eastern region of the state to São Paulo, followed by range expansion to the centralwestern regions. The second is that invasion of bordering states into the western region of São Paulo and then expanded to the central and eastern regions. But to date, no clear hypotheses exist regarding the global invasion history of *D. citri*. Based on results of this study, the haplotype frequency, the phylogeographical distribution, and the patterns of haplotype connectivity, we hypothesize three colonization scenarios: the expansion of lineage B from southern Asia into North America via West Asia, the expansion of lineage A from Southeast Asia into East Asia, and the parallel dispersals of the two lineages into Africa and South America (Figure 3a). In addition, the parallel dispersal patterns may involve multiple propagules, which might explain the high genetic diversity observed in South America, and the finding of an individual of lineage B in northern America where lineage A dominates.

Significantly negative neutrality tests (Tajima’s D and Fu’s F;) within each lineage of *D. citri*, two star-like cluster patterns in the haplotype network, and the unimodal mismatch distributions for the whole sample and both lineages all rejected the null hypothesis of neutral evolution of the partial COI gene. That low genetic diversity characterized most sampling regions, and native and invasive populations share common haplotypes (Hap4 and Hap9), is consistent with recent introductions events and could suggest a single introduction of haplotypes in most region.

### 4.3 Future directions

The continent-global scale phylogeographic history of the psyllid *D. citri* has to date exclusively been studied with mtDNA represented by the COI locus. These studies have allowed understanding of the matrilineage structure of this species and generated hypotheses regarding dispersal and colonization of this important pest. However, while mtDNA data have proven highly useful in phylogeographic studies, there are several shortcomings of this approach (e.g., Avise, 2000). Not the least of these is the potential mismatch between the dispersal history of males and females and likely mismatch between a species tree and any single gene tree. With this paper, we summarize available COI data and add new data re-analyses of global patterns, and discuss prior and novel hypotheses on dispersal and colonization history of *D. citri*. Robustly testing these hypotheses, and gaining a deeper understanding of population structure of this species, will require data drawn at random from the whole genome, utilizing methods such as RADseq or targeted sequence capture. Such data will tease apart male
and female dispersal, testing the “female-based” dispersal hypotheses proposed here. They will also allow testing of “independent lineages” hypothesis through precise measures of genetic isolation and gene flow unavailable to single-marker studies. Finally, such data will be immune to potential issues such as Wolbachia induced biases. However, for such work to be realized, resampling specimens across the globe will be necessary, and we hope our study can aid in strategical sampling reflecting known haplotype diversity.

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CONFLICT OF INTEREST

None declared.

AUTHOR CONTRIBUTIONS

Yufa Luo designed this study, collected and processed samples from all field sites, conducted data analyses, and wrote the manuscript. Ingi Agnarsson helped extensively with manuscript writing. All authors critically revised and approved the final version of the manuscript submitted.

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REFERENCES

Avise, J. C. (2000). Phylogeography: The history and formation of species. Cambridge, MA: Harvard University Press.

Boykin, L. M., De Barro, P., Hall, D. G., Hunter, W. B., McKenzie, C. L., Powell, C. A., & Shattters, R. G. (2012). Overview of worldwide diversity of Diaphorina citri Kuwayama mitochondrial cytochrome oxidase I haplotypes: Two old world lineages and a New World invasion. Bulletin of Entomological Research, 102, 573–582. https://doi.org/10.1017/S0007485312000181

Carmichael, L. E., Krizan, J., Nagy, J. A., Fuglei, E., Dumond, M., Johnson, D., & Strobeck, C. (2007). Historical and ecological determinants of genetic structure in arctic canids. Molecular Ecology, 16, 3466–3483. https://doi.org/10.1111/j.1365-294X.2007.03381.x

Chen, H., Strand, M., Norenburg, J. L., Sun, S., Kajihara, H., Chernyshev, A. V., & Sundberg, P. (2010). Statistical parsimony networks and species assemblages in Cephalotrichid nemerteans (Nemertea). PLoS One, 5, e12885. https://doi.org/10.1371/journal.pone.0012885

Clement, M., Posada, D., & Crandall, K. A. (2000). TCS: A computer program to estimate genealogies. Molecular Ecology, 9, 1567–1660. https://doi.org/10.1046/j.1365-294X.2000.01020.x

Costa Lima, A. M. (1942). Homopteros. Insetos do Brazil, 3, 1–327.

d’Agraça, J. V. (2008). Biology, history, and world status of Huanglongbing, pp. 7. Taller Internacional sobre Huanglongbing de los cítricos (Candidatus Liberibacter spp.) y el psilido asiático de los cítricos (Diaphorina citri), 7–9 May 2008, Hermosillo, Sonora, Mexico.

de León, J. H., Sétamou, M., Gastaminza, G. A., Buenahora, J., Cáceres, S., Yamamoto, P. T., & Logaro, G. A. (2011). Two separate introductions of Asian citrus psyllid populations found in the American continents. Annals of the Entomological Society of America, 104, 1392–1398. https://doi.org/10.1603/AN11086

DiLeo, M. F., Row, J. R., & Lougheed, S. C. (2010). Discordant patterns of population structure for two co-distributed snake species across a fragmented Ontario landscape. Diversity and Distribution, 16, 571–581. https://doi.org/10.1111/j.1472-6462.2010.00667.x

Excoffier, L. (2004). Patterns of DNA sequence diversity and genetic structure after a range expansion: Lessons from the infinite-island model. Molecular Ecology, 13, 853–864. https://doi.org/10.1046/j.1365-294X.2003.02004.x

Excoffier, L., Foll, M., & Petit, R. J. (2009). Genetic consequences of range expansions. Annual Review of Ecology, Evolution, and Systematics, 40, 481–501. https://doi.org/10.1146/annurev.ecolsys.39.110707.173414

Excoffier, L., & Lischer, H. E. L. (2010). Arlequin suite ver 3.5: A new series of programs to perform population genetics analyses under Linux and Windows. Molecular Ecology Resources, 10, 564–567. https://doi.org/10.1111/j.1755-0999.2010.02847.x

Fay, J. C., & Wu, C. I. (2003). Sequence divergence, functional constraint, and selection in protein evolution. Annual Review of Genomics and Human Genetics, 4, 213–235. https://doi.org/10.1146/annurev.genom.4.200303.162528

French, J. V., Kahlke, C. J., & De Graca, J. V. (2001). First record of the Asian citrus psylla Diaphorina citri Kuwayama (Homoptera: Psyllidae), in Texas. Subtropical Plant Science, 53, 14–15.

Fu, Y. X. (1997). Statistical tests of neutrality of mutations against population growth, hitchhiking and background selection. Genetics, 147, 915–925.

Green, P. (2009). PHRED, 1.090518 edn. Retrieved from http://www.phrap.org

Green, P., & Ewing, B. (2002). PHRED, 0.202425c edn. Retrieved from http://phrap.org/

Guidolin, A. S., Friesia, P., & Cònsoli, F. L. (2014). The genetic structure of an invasive pest, the Asian citrus psyllid Diaphorina citri (Hemiptera: Liviidae). PLoS One, 9, e115749. https://doi.org/10.1371/journal.pone.0115749

Halbert, S. E., & Manjunath, K. L. (2004). Asian citrus psyllids (Sternorrhyncha: Psyllidae) and greenhouseing disease of citrus: A literature review and assessment of risk in Florida. Florida Entomologist, 87, 330–353. https://doi.org/10.1653/0015-4040(2004)087[0330:DOTACP]2.0.CO;2

Hall, T. A. (1999). BioEdit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series, 41, 95–98.

Hall, D. G. (2008). Biology, history and world status of Diaphorina citri. pp. 1–11 in Proceedings of the International Workshop on Huanglongbing and Asian Citrus Psyllid, North American Plant Protection Organization, 7–9 May 2008, Hermosillo, Sonora, Mexico.

Hoffmann, W. E. (1936). Diaphorina citri Kuw. (Homoptera: Cermididae), a citrus pest in Kwangtung. Lingnan Science Journal, 15, 127–132.

Hollis, D. (1987). A new citrus-feeding psyllid from the Comoro Islands, with a review of the Diaphorina amena species group (Homoptera). Systematic Entomology, 12, 47–61. https://doi.org/10.1111/j.1365-3113.1987.tb00547.x

Hoy, A. M. (2013). Insect molecular genetics: An introduction to principles and applications, 3rd edn. Cambridge, MA: Academic Press.

Jeanninou, F., Thompson, J. D., Gouy, M., Higgins, D. G., & Gibson, T. J. (1998). Multiple sequence alignment with Clustal X. Trends in Biochemical Sciences, 23, 403–405. https://doi.org/10.1016/S0968-0004(98)01285-7

Joudrie, V., Alvarez, N., Molina-Ochoa, J., Williams, T., Bergvinson, D., Benrey, B., & Franck, P. (2010). Population genetic structure of two
primary parasitoids of Spodoptera frugiperda (Lepidoptera), Chelonus insularis and Camposepis sonorensis (Hymenoptera): To what extent is the host plant important? Molecular Ecology, 19, 2168–2179. https://doi.org/10.1111/j.1365-294X.2010.04625.x

Katoh, K., Kuma, K., Toh, H., & Miyata, T. (2005). MAFFT version 5: Improvement of accuracy and speed of multiple sequence alignment. Nucleic Acids Research, 33, 518–521. https://doi.org/10.1093/nar/gk1198

Kobori, Y., Nakata, T., Ohto, Y., & Takasu, F. (2011). Dispersal of adult Asian citrus psyllid, Diaphorina citri Kuwayama (Homoptera: Psyllidae), the vector of citrus greening disease, in artificial release experiments. Applied Entomology and Zoology, 46, 27–30. https://doi.org/10.1007/s13355-010-0004-2

Lashkari, M., Manzari, S., Sahragard, A., Malagnini, V., Boykind, L. M., & Hosseini, N. (2014). Global genetic variation in the Asian citrus psyllid, Diaphorina citri (Hemiptera: Liviidae) and the endosymbiont Wolbachia: Links between Iran and the USA detected. Pest Management Science, 70, 1033–1040. https://doi.org/10.1002/ps.3643

Lee, C. E. (2002). Evolutionary genetics of invasive species. Trends in Ecology and Evolution, 17, 386–391. https://doi.org/10.1016/S0169-5347(02)02554-5

Librado, P., & Rozas, J. (2009). DnaSP v5: A software for comprehensive analysis of DNA polymorphism data. Bioinformatics, 25, 1451–1452. https://doi.org/10.1093/bioinformatics/btp187

Lopes, S. A., Frare, G. F., Yamamoto, P. T., Ayres, A. J., & Barbosa, J. C. (2007). Ineffectiveness of pruning to control citrus huanglongbing caused by Candidatus Liberibacter americanus. European Journal of Plant Pathology, 119, 463–468. https://doi.org/10.1007/s10658-007-9173-7

Maddison, D., & Maddison, W. (2011a). Chromaseq 1.0: A Mesquite package for analyzing sequence chromatograms. Retrieved from http://www.mesquiteproject.org/packages/chromaseq

Maddison, W., & Maddison, D. (2011b). Mesquite 2.75: A modular system for evolutionary analysis. Retrieved from http://www.mesquiteproject.org

Mahy, G., Vekemans, X., & Jacquemart, A. L. (1999). Patterns of allozymic variation within Calluna vulgaris populations at seed bank and adult stages. Heredity, 82, 432–440. https://doi.org/10.1038/sj.hdy.6884990

Mead, F. W. (1977). The Asiatic citrus psyllid, Diaphorina citri Kuwayama (Homoptera: Psyllidae). Florida Department of Agriculture Conservation Service, Division of Plant Industry Entomology Circular 180, Florida Division of Plant Industry, Retrieved from http://www.freshfromflorida.com/pi/enpp/ento/entcic/ent180.pdf

Nei, M., Muruyama, T., & Chakraborty, R. (1975). The bottleneck effect and genetic variability in populations. Evolution, 29, 1–10. https://doi.org/10.1111/j.1558-5646.1975.tb00807.x

Ohta, T. (1995). Synonymous and nonsynonymous substitutions in mammalian genes and the nearly neutral theory. Journal of Molecular Evolution, 40, 56–63. https://doi.org/10.1007/bf00166595

Pinho, C., Harris, D. J., & Ferrand, N. (2007). Comparing patterns of nuclear and mitochondrial divergence in a cryptic species complex: The case of Iberian and North African wall lizards (Podarcis, Lacertidae). Biological Journal of Linnean Society, 91, 121–133. https://doi.org/10.1111/j.1095-8312.2007.00774.x

Pluke, R. W. H., Qureshi, J. A., & Stansly, P. A. (2008). Citrus flushing patterns, Diaphorina citri (Hemiptera: Psyllidae) populations and parasitism by Taraminxia radiate (Hymenoptera: Eulophidae) in Puerto Rico. Florida Entomologist, 91, 36–42. https://doi.org/10.1653/0015-4040(2008)091[0036:CFPDCH2.0.CO;2]

Porsetta, D., Canestrelli, D., Bellini, R., Celli, G., & Urbanelli, S. (2007). Improving insect pest management through population genetic data: A case study of the mosquito Ochlerotatus caspius (Pallas). Journal of Applied Ecology, 44, 682–691. https://doi.org/10.1111/j.1365-2664.2007.01301.x

Posada, D. (2008). ModelTest: Phylogenetic model averaging. Molecular Biology and Evolution, 25, 1253–1256. https://doi.org/10.1093/molbev/msn083

Pritchard, J. K., Stephens, M., & Donnelly, P. (2000). Inference of population structure using multilocus genotype data. Genetics, 155, 945–959.

Ray, N., Currat, M., & Excoffier, L. (2003). Intra-deme molecular diversity in spatially expanding populations. Molecular Biology and Evolution, 20, 76–86. https://doi.org/10.1093/molbev/msg09

Rogers, A. R., & Harpending, H. (1992). Population growth makes waves in the distribution of pairwise genetic differences. Molecular Biology and Evolution, 9, 552–569. https://doi.org/10.1093/oxfordjournals.molbev.a040727

Schneider, S., & Excoffier, L. (1999). Estimation of past demographic parameters from the distribution of pairwise differences when the mutation rates vary among sites: Application to human mitochondrial DNA. Genetics, 152, 1079–1089.

Silva, A. G. A., Gonçalves, C. R., Galvão, D. M., Gonçalves, A. J. L., Gomes, J., Silva, M. N., & Simoni, L. (1968), Quarto catálogo dos insetos que vivem nas plantas do Brasil, seus parasitos e predadores. Rio de Janeiro, Brazil: Angelo Moreira da Costa Lima.

Tajima, F. (1989). Statistical method for testing the neutral mutation hypothesis by DNA polymorphism. Genetics, 123, 585–595.

Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., & Kumar, S. (2011). MEGA5: Molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. Molecular Biology and Evolution, 28, 2731–2739. https://doi.org/10.1093/molbev/msr121

Tsi, J. H., Wang, J. J., & Liu, Y. H. (2000). Sampling of Diaphorina citri (Homoptera: Psyllidae) on orange Jasmine in southern Florida. Florida Entomologist, 83, 446–459. https://doi.org/10.1371/journal.pbio.1002013

Wang, Z. (2002). Distribution and transmission of citrus psylla in Guizhou Province. Tillage and Cultivation, 22, 62–63.

Waterhouse, D. F. (1998). Biological control of insect pests: Southeast Asian Prospects. ACIAR Monograph Series No. 51. Canberra, ACT: Australian Centre for International Agricultural Research.

Xu, C., Xia, Y., & Ke, C. (1994). Study on the biology and control of citrus psylla. Acta Phytophylacica Sinica, 21, 53–56.

Xu, C., Zhang, W., Lu, B., & Chen, J. (2001). Progress in studies on mechanisms of biological invasion. Biodiversity Science, 9, 430–438.

Yamamoto, P. T., Felipe, M. R., Sanches, A. L., Coelho, J. H. C., Garbim, L. F., & Ximenes, N. (2009). Eficácia de inseticidas para o manejo de Diaphorina citri Kuwayama (Hemiptera: Psyllidae) em citros. BioAssay, 4, 1–9.

Yang, Y. (1989). Effects on light, temperature and humidity on the development, reproduction and survival of citrus psylla. Acta Ecologica Sinica, 9, 348–354.

Zayed, A., Constantia, S. A., & Parker, L. (2007). Successful biological invasion despite a severe genetic load. PLoS One, 2, e868. https://doi.org/10.1371/journal.pone.0000868

Zhao, X. (1987). Occurrence of citrus yellow shoot (greening) disease in Guangxi and in southern Sichuan: a brief review. In: The FAO-UNDP Project Coordinator in Fuzhou, editor. Regional workshop on citrus greening huanglongbin disease, 6–12 December 1987, Fuzhou, China.

SUPPORTING INFORMATION

Additional Supporting Information may be found online in the supporting information tab for this article.

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