Case Report

Neurotoxicity and Other Clinical Manifestations of a Common European Adder (Vipera berus) Bite in Romania

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Abstract: Most cases of envenomation by common European vipers (Vipera berus) have not been reported to have neurotoxic manifestations. However, these manifestations have been demonstrated in some cases of envenomation by subspecies of V. berus, found in the Carpathian Basin region of south-eastern Europe. Here, we report the case of a 5-year-old girl from the south of Romania who presented symptoms of neurotoxicity, as well as other systemic and local symptoms, after being bitten by an adder of the V. berus subspecies. Treatment consisted of monovalent antivenom, a corticosteroid, and prophylactic enoxaparin. Neurotoxic manifestations of envenomation as well as other local and systemic symptoms improved within 5 days of treatment. The presented case shows that venom from V. berus subspecies found in the Carpathian Basin can have neurotoxic effects. This case also confirmed the efficacy of monospecific antivenom treatment in bringing about rapid and complete remission, following envenomation.

Keywords: neurotoxicity; Berus Viper; common European viper; child; monospecific antivenom; south Romania

Key Contribution: V. berus subspecies from the Carpathian Basin can have neurotoxic effects. Treatment with antivenom serum proved to be efficient with complete remission after use.

1. Introduction

According to the Romanian Society of Herpetology, the only venomous snakes in Romania are adders (also known as vipers). These can be classified under three species, namely, Vipera berus, Vipera ammodytes, and Vipera ursinii [1]. Clinical manifestations of envenomation by these adders vary and may be local or systemic. Systemic manifestations of envenomation may present as gastrointestinal, cardiac, neurological, and/or respiratory symptoms, and may also include anaphylactic reactions and coagulopathy [2–6]. Neurotoxic manifestations are considered unusual; in most cases published in Europe, these manifestations were attributed to V. aspis [4,6], and not to V. berus. Nevertheless, clinical symptoms of neurotoxicity in V. berus bites were observed as far back as 2008, when Malina et al. reported a previously healthy 27-year-old man bitten by a V. berus in eastern Hungary [7] Later, in 2017, the same author also showed the possibility of neurotoxic
manifestations of *V. berus* envenomation during an experimental study of *V. berus* venom of adders from the same region. [8]. In this case report, we document the presence of symptoms of neurotoxicity and the development of these symptoms, as well as other associated clinical manifestations of envenomation, in a 5-year-old girl from south Romania, who was bitten by a common European viper (*V. berus*). The study was conducted according to the provisions of the “EU Guidelines for the Promotion and Protection of the Rights of the Child” and was approved by the Ethics Committee of the hospital (Protocol Code 15351/02.06.2022).

2. Case Report

A previously healthy 5-year-old girl was bitten on her right leg by a viper near her house, in south Romania. Twelve hours later, she was presented to our clinic, after being transferred from a local hospital.

On admission, the girl presented with a mildly altered general status. Examination revealed local signs of a snake bite on her right leg, including local pain and heat, swelling, infiltration, erythema surrounded by cyanosis and functional impairment. The snake bite mark is shown in Figure 1a. Systemic manifestations were also present, and include mild gastrointestinal and neurological symptoms. Gastrointestinal symptoms consisted of diffuse abdominal pain and nausea. Neurological manifestations included severe somnolence and axial hypotonia. Moreover, bilateral ophthalmologic disturbances (shown in Figure 1b) were noted, including palpebral ptosis and ophthalmoplegia. Diplopia was also confirmed.

The patient’s progress was clinically monitored after the initiation of the treatment. The laboratory results and symptoms over time are presented below (see Tables 1 and 3). Thirty minutes after administration of the antivenom, ocular movements reappeared and blood pressure increased to 98/66 mmHg. Somnolence, diplopia, and gastrointestinal manifestations withdrew on Day 2, while palpebral ptosis disappeared on Day 4. Local signs, after initially worsening progressively, improved and completely disappeared by Day 5.

The child was discharged on Day 6 without any clinical signs or symptoms. The laboratory test results were within normal ranges (see Table 1). No immediate or delayed adverse reactions were reported after antivenom administration, apart from a slight increase in IgE level, without clinical expression.

![Figure 1. (a) Local signs of viper bite including swelling, erythema (lower calf and ankle); (b) image showing inferior displacement of the upper eyelid with associated narrowing of the vertical palpebral fissure (bilateral palpebral ptosis).](image-url)

The initial cardiovascular assessment did not show any abnormalities; blood pressure and heart rate were within the normal ranges according to age (102/55 mmHg and 110 bpm, respectively). The results of laboratory tests were normal (see Table 1). Electrocardiogram and cardiac echography showed no abnormalities, and Doppler echography of the right lower limb showed normal results.

One hour after admission, local signs and general symptoms worsened and swelling and pain spread to the middle of the right calf, somnolence and palpebral ptosis became pronounced and blood pressure started to decrease gradually (to 80/67 mmHg, 1 h post-admission). Based on the above clinical presentation, the severity of the case was graded as 2b (regional oedema and moderate general symptoms as mild hypotension and neurotoxic signs), according to the Audebert–Boels classification, adapted to children by Marano et al. (see Table 2) [6].
Table 1. Laboratory results on days 1, 2, and 6.

| Laboratory Test                  | Normal Values | Changes by Day: |
|---------------------------------|---------------|-----------------|
|                                 |               | Day 1 (admission) | Day 2 | Day 6 (discharge) |
| Hemoglobin (g/dL)               | 12–16         | 11.8             | 13.2  |
| Leucocytes (mmc)                | 4000–12,000   | 9100             | 5200  |
| Thrombocytes (mmc)              | 150,000–400,000 | 321,000         | 293,000 |
| ESR (mm/h)                      | 7–12          | 7               |
| Fibrinogen (mg/dL)              | 150–400       | 243.9            | 253   |
| C reactive protein (mg/dL)      | 0–0.5         | 0.29             |
| ALT (IU/L)                      | 0–35          | 24              | 21    | 27               |
| AST (IU/L)                      | 0–35          | 50              | 42    | 45               |
| GGT (IU/L)                      | 15–132        | 10              | 13    |
| LDH (IU/L)                      | 110–295       | 311             | 224   | 252              |
| Amylase (IU/L)                  | 22–80         |                 | 39    |
| Direct bilirubin (mg/dL)        | 0–0.2         |                 | 0.06  |
| Indirect bilirubin (mg/dL)      | 0–1           |                 | 0.26  |
| Total bilirubin (mg/dL)         | 0.3–1.2       |                 | 0.32  |
| Iron (µg/dL)                    | 40–100        |                 | 44    |
| Chloride (mmol/L)               | 101–109       | 105             | 103   | 102              |
| Sodium (mmol/L)                 | 132–142       | 134             | 136   | 135              |
| Potassium (mmol/L)              | 3.5–5.1       | 4.1             | 3.37  | 4.71             |
| Urea (mg/dL)                    | 10.8–38.4     | 28              | 28    | 23               |
| Creatinine (mg/dL)              | 0.26–0.77     | 0.45            | 0.41  | 0.42             |
| CK (IU/L)                       | 0–145         | 120             | 106   | 135              |
| CK-MB (ng/mL)                   | <5            | <5              | <1    | <1               |
| Myoglobin (ng/mL)               | <50           | <50             | <50   | <50              |
| Troponin (ng/mL)                | <1            | <1              | <0.05 | <0.05            |
| D-dimers (ng/mL)                | <500          | <100            | <100  |
| BNP (pg/mL)                     | <100          | 15.4            | <5    |
| IgA (g/L)                       | 0.41–2.97     | 0.71            |
| IgG (g/L)                       | 5–13          | 10              |
| IgM (g/L)                       | 0.4–1.8       | 0.63            |
| IgE (IU/mL)                     | <100          | 89.53           | 233   |
| C3 fraction (g/L)               | 0.9–1.8       | 1.27            |
| C4 fraction (g/L)               | 0.1–0.4       | 0.32            |
| CIC (IU/mL)                     | <10           | 2               |
| Quick Time (s)                  | 11–14         | 14.1            | 13.1  |
| INR                             | 0.8–1.3       | 1.28            | 1.17  |
| Prothrombin activity (%)        | 70–140        | 77.9            | 84.8  |
| APTT (s)                        | 24.4–36.4     | 25.1            | 21.6  |

ESR—erythrocyte sedimentation rate, ALT—alanine aminotransferase, AST—aspartate aminotransferase, GGT—gamma-glutamyl transferase, LDH—lactate dehydrogenase, CK—creatinine kinase, CK-MB—creatinine kinase—myocardial band, BNP—B-type natriuretic peptide, IgA—immunoglobulin A; IgG—immunoglobulin G, IgM—immunoglobulin M, IgE—immunoglobulin E, C3—complement C3 fraction, C4—complement C4 fraction, CIC—circulating immune complexes, INR—international normalized ratio, APTT—activated partial thromboplastin time.
Table 2. Audebert–Boels Classification modified by Marano et al. adapted from [6].

| Grade | Description | Signs and Symptoms | Treatment |
|-------|-------------|--------------------|----------|
| 0     | No envenoming (“dry bite”) | Fang marks, No oedema, No local reaction | 6 h surveillance in the emergency room |
| 1     | Minimal envenoming | Local oedema around the bite area, No systemic symptoms | Clinical observation up to evident reduction of edema, Supportive care, including hydration and pain relief |
|       | Grade 2a    | One or both of the following: | |
|       |             | - Regional edema with progression to most of the limb | Clinical observation up to the evident reduction of edema (evaluate district perfusion and saturation) |
|       |             | - Haematoma or adenopathy | |
|       | Grade 2b    | Grade 2a + moderate general symptoms (mild hypotension, vomiting, diarrhea, neurotoxic signs), and/or biological criteria for severity: | |
|       |             | - Leukocytes > 11,000/L | Supportive care, including hydration and pain relief |
|       |             | - Neutrophils > 65% | Doppler-ultrasound of affected limb’s blood vessels |
|       |             | - INR > 1.15 | Administration of antivenom, Evaluate antibiotic therapy *, Administer LMWH ** |
| 2     | Moderate envenoming | Other or both of the following: | |
|       |             | - Edema spreading to the trunk | Same intervention as in Grade 2 |
|       |             | - Signs of hemodynamic instability (prolonged hypotension, shock, bleeding) | Admission to PICU |

* Only if clinical or laboratory signs of bacterial contamination are evident, ** Only if direct evidence of thrombophlebitis is available or in cases of extensive edema; dehydration; decreased mobility; prolonged decubitus; admission to PICU; anticipated hospitalization longer than 48 h. Do not administer in the case of overt hemorrhage or a bleeding disorder.

The viper was identified as belonging to the species *V. berus* based on the morphological description of the viper provided by the girl’s mother and the information obtained from the Romanian Association of Herpetology. The mother who accompanied the child at the moment of the incident described a completely black snake without a zigzag pattern and about 50 cm long. The last mapping of the distribution of vipers in Romania shows the presence of *V. berus* and *V. ammodytes* in the geographical location of the incident reported as the Subcarpathian area of south Romania, specifically Vâlcea county [1]. According to the mapping, the herpetologist stated that the viper belongs to the *V. berus* species, because *V. ammodytes* does not have any melanic gene and thus cannot be entirely black. Even though some authors have reported the presence of a certain subspecies of vipers, namely, *Vipera berus nikolskii* in Vâlcea [9], it is rather difficult to establish the viper in question belonging to this subspecies whose presence here has not been certified by DNA identification.

Twenty hours after the viper bite, treatment was initiated with Viper Venom Antitoxin (*Immunoserum contra venena viperarum europaearum*) manufactured by Biomed, Poland. The patient was administered 500 AU (one vial) as a bolus dose in 250 mL normal saline over 3 h. The patient was also treated with intravenous fluid therapy containing glucose and electrolytes, intravenous methylprednisolone, and subcutaneous enoxaparin (prophylactic dosage of 2000 UI/day) for 5 days.

The patient’s progress was clinically monitored after the initiation of the treatment. The laboratory results and symptoms over time are presented below (see Tables 1 and 3). Thirty minutes after administration of the antivenom, ocular movements reappeared and
blood pressure increased to 98/66 mmHg. Somnolence, diplopia, and gastrointestinal manifestations withdrew on Day 2, while palpebral ptosis disappeared on Day 4. Local signs, after initially worsening progressively, improved and completely disappeared by Day 5.

**Table 3. Changes in the clinical profile of the patient over time.**

| Day                      | Clinical Features                                                                 |
|--------------------------|----------------------------------------------------------------------------------|
| Day 1: at the time of treatment initiation | Somnolence, palpebral ptosis, ophthalmoplegia, and bilateral diplopia. Mild gastrointestinal symptoms (nausea and diffuse abdominal pain). Local manifestations (swelling, erythema surrounded by cyanosis, local heat, induration, and pain in lower half of right calf). BP = 80/67 mmHg, HR = 116 beats/min |
| Day 1: 30 min after treatment initiation | Return of ocular movements. BP = 98/58 mmHg, HR = 118 beats/min |
| Day 2: 12 h after treatment initiation | No somnolence, no diplopia, and no gastrointestinal symptoms present. Persistence of palpebral ptosis. Local inflammation reduced. BP = 103/55 mmHg; HR = 80 beats/min |
| Day 3                  | Palpebral ptosis in remission, local signs improved (decreased swelling, no local heat, modest pain); BP = 90/66 mmHg; HR = 89 beats/min |
| Day 4                  | No palpebral ptosis noted. BP = 101/61 mmHg; HR = 105 beats/min |
| Day 5                  | No local signs or symptoms. BP = 107/67 mmHg; HR = 100 beats/min |
| Day 6                  | Complete remission was noted, and patient was discharged from our clinic. |

BP = blood pressure; HR = heart rate.

The child was discharged on Day 6 without any clinical signs or symptoms. The laboratory test results were within normal ranges (see Table 1). No immediate or delayed adverse reactions were reported after antivenom administration, apart from a slight increase in IgE level, without clinical expression.

Daily lab testing was conducted using bedside assessment.

### 3. Discussion

From the venom composition, it is thought that neurotoxic effects of venom from common European adders are caused by neurotoxins with phospholipase A2 (PLA2) enzymatic activity [10,11]. A study published by Zanetti et al. in 2018 regarding the effects of *V. aspis* and *V. berus* venoms in mice showed that both types of venom have phospholipase A2 enzymes, but only *V. aspis* venom is neurotoxic. Clinical observations from various regions of France and Italy support this finding, attributing neurological manifestations exclusively to envenomation by *V. aspis* [4,6,12–14]. Nevertheless, studies and clinical observations [8,10] have shown that the venom of some *V. berus* subspecies from Eastern Europe can have neurotoxic effects, not only in animal experimental models but also in humans. Neurotoxic manifestations following *V. berus* envenomation have been sporadically communicated in the literature, with all incidents reported in the Carpathian Basin of Hungary, Romania, and Bulgaria [8,15]. Three cases out of the seven reported cases involved children aged 14, 12, and 9 years, respectively [16–18].

In our case, the incident took place in the Subcarpathian area of the south of Romania, which is in the geographical region of Oltenia (see Figure 2). Symptoms of neurotoxicity appeared 30 min after envenomation and included somnolence, palpebral ptosis, ophthalmoplegia, and bilateral diplopia. These symptoms are similar to symptoms previously described in the literature in cases of *V. berus* bites in the Carpathian Basin region [8]. Antivenom indication was established after classifying the case severity as grade 2b, using the Audebert–Boels classification adapted to children by Marano et al. as shown in Table 2 [6].
In our case, the incident took place in the Subcarpathian area of the south of Romania [1]. Red dots: records of *V. berus* after 1990; blue dots: records of *V. berus* before 1990.

Using other classification systems published in the literature, regarding severity of viper envenomation, we can classify our case as Stage 2 according to the Clinical Gradation of European Viper Envenomation system [12], and Stage 1 (ocular and mild gastrointestinal symptoms present) according to the Modified Grading Severity Score (GSS) system for peripheral neurotoxic effects, after Italian viper bites [4]. Both stages in the above classification systems are stages in which antivenom treatment is indicated.

In our case, viper Venom Antitoxin serum (Biomed, Poland) was administered. Viper Venom Antitoxin serum is a monovalent antivenom containing Fab specific equine immunoglobulins that bind the venom of *V. berus*.

Enoxaparin was administered prophylactically to prevent secondary venous thrombosis due to prolonged immobilization of the affected leg [12]. We excluded the need for antibiotic therapy since there were no local or general signs of infection. As stated in the literature, antibiotics in snakebite cases should only be administered if there is a positive history of infection, or clinical or biological signs of infection. [6,19].

Although the use of corticosteroids is controversial [6,19], the decision was made to administer intravenous methylprednisolone as a symptomatic treatment for its well-known anti-inflammatory effects. The outcome of methylprednisolone was favourable. Progressive amelioration of neurological symptoms was noted as follows: 30 min after immunotherapy administration, ocular movements reappeared; after 12 h, somnolence and diplopia withdrew; and on the fourth day, palpebral ptosis was completely absent. We believe that the complete resolution of symptoms of neurotoxicity after treatment with the monospecific antivenom could be an argument that the snake responsible for envenomation was a viper of the *V. berus* subspecies. Moreover, other systemic manifestations rapidly resolved, and local symptoms progressively improved, until complete remission was achieved on the fifth day. No adverse reactions were observed after antivenom serum administration. This is consistent with the literature [2,4,6]. In our case, however, a slight increase in the IgE level, without clinical symptoms, was noted. The limitation of our study consisted of the identification modality of the snake, which was conducted by the herpetologist based on the following arguments: the viper is the only venomous snake in our country, the geographical mapping, and the morphological description provided by the adult accompanying the child at the time of the incident.

4. Conclusions

We can conclude that the venom of *V. berus* subspecies, found in the Carpathian Basin region, appears to have neurotoxic effects. This conclusion was reached from specialist consultation in *V. berus* identification, clinical observation, and possibly because of the complete and rapid remission of the symptoms under monospecific antivenom treatment. Consequently, we must take into consideration *V. berus* when confronted with neurological symptoms in patients bitten by snakes in the Subcarpathian Basin from Eastern Europe. However, further studies are necessary to confirm it.
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References

1. Rozylowicz, L.; Cogălniceanu, D.; Székely, P.; Samoilă, C.; Stănescu, F.; Tudor, M.; Székely, D.; Iosif, R. Diversity and distribution of reptiles in Romania. *Zoologica* 2013, 341, 49–76. [CrossRef] [PubMed]
2. Jollivet, V.; Hamel, J.; de Haro, L.; Labadie, M.; Sapori, J.; Cordier, L.; Villa, A.; Nisse, P.; Puskarczyk, E.; Berthelon, L.; et al. European viper envenomation recorded by French poison control centers: A clinical assessment and management study. *Toxicon* 2015, 108, 97–103. [CrossRef] [PubMed]
3. Lamb, T.; Stewart, D.; Warrell, D.A.; Lalloo, D.G.; Jagpal, P.; Jones, D.; Thanacoody, R.; Gray, L.A.; Eddleston, M. Moderate-to-severe *Vipera berus* envenoming requiring ViperaTAb antivenom therapy in the UK. *Clin. Toxicol.* 2021, 59, 992–1001. [CrossRef] [PubMed]
4. Lonati, D.; Giampreti, A.; Rossetto, O.; Petrolini, V.M.; Vecchio, S.; Buscaglia, E.; Mazzoleni, M.; Chiara, F.; Aloise, M.; Gentilli, A.; et al. Neurotoxicity of European viperids in Italy: Pavia Poison Control Centre case series 2001–2011. *Clin. Toxicol.* 2014, 52, 269–276. [CrossRef] [PubMed]
5. Karlsson-Stibler, C.; Salmonson, H.; Persson, H. A Nationwide Study of *Vipera berus* Bites During One Year—Epidemiology and Mortality of 231 Cases. *Clin. Toxicol.* 2006, 44, 25–30. [CrossRef] [PubMed]
6. Marano, M.; Pisani, M.; Zampini, G.; Pontrelli, G.; Roversi, M. Acute Exposure to European Viper Bite in Children: Advocating for a Pediatric Approach. *Toxins* 2021, 13, 330. [CrossRef] [PubMed]
7. Malina, T.; Krecsák, L.; Warrell, D. Neurotoxicity and hypertension following European adder (*Vipera berus*) bites in Hungary: Case report and review. *QJM* 2008, 101, 801–806. [CrossRef] [PubMed]
8. Malina, T.; Krecsák, L.; Westerström, A.; Szemán-Nagy, G.; Gymánt, G.; M-Hamvas, M.; Rowan, E.G.; Harvey, A.L.; Warrell, D.A.; Pál, B.; et al. Individual variability of venom from the European adder (*Vipera berus*) from one locality in Eastern Hungary. *Toxicon* 2017, 135, 59–70. [CrossRef] [PubMed]
9. Túrancu, V.; Zinenko, O.; Strugariu, A. Distribution and morphological variation of *Vipera berus* nikolskii Vedmederja, Grubant et Rudaeva, 1986 in Western Ukraine, The Republic of Moldova and Romania. *Amphibia–Reptilia* 2010, 31, 51–67. [CrossRef]
10. Varga, C.; Malina, T.; Alfoldi, V.; Bilics, G.; Nagy, F.; Oláh, T. Extending knowledge of the clinical picture of Balkan adder (*Vipera berus bosniensis*) envenomation: The first photographically-documented neurotoxic case from South-Western Hungary. *Toxicon* 2018, 143, 29–35. [CrossRef] [PubMed]
11. Zanetti, G.; Duregotti, E.; Locatelli, C.A.; Giampreti, A.; Lonati, D.; Rossetto, O.; Pirazzini, M. Variability in venom composition of European viper subspecies limits the cross-effectiveness of antivenoms. *Sci. Rep.* 2018, 8, 9818. [CrossRef] [PubMed]
12. de Haro, L. Management of snakebites in France. *Toxicon* 2012, 60, 712–718. [CrossRef] [PubMed]
13. Ferquel, E.; De Haro, L.; Jan, V.; Guillemin, I.; Jourdain, S.; Teynié, A.; D’Alayer, J.; Choumet, V. Reappraisal of *Vipera aspis* Venom Neurotoxicity. *PLoS ONE* 2007, 2, e1194. [CrossRef] [PubMed]
14. Garrigues, T.; Dauga, C.; Ferquel, E.; Choumet, V.; Failloux, A.-B. Molecular phylogeny of *Vipera laurenti*, 1768 and the related genera *Macrovipera* (Reuss, 1927) and *Daboia* (Gray, 1842), with comments about neurotoxic *Vipera aspis* aspis populations. *Mol. Phylogenet. Evol.* 2005, 35, 35–47. [CrossRef] [PubMed]
15. Westerström, A.; Petrov, B.; Tzankov, N. Envenoming following bites by the Balkan adder *Vipera berus* bosniensis—First documented case series from Bulgaria. *Toxicon* 2010, 56, 1510–1515. [CrossRef] [PubMed]
16. Gafencu, M.; Doros, G.; Badeti, R.; Vasilie, D. Envenoming by *Vipera Berus*: A case report of neurotoxicity. Abstract no.42. Abstracts of the 2012 International Congress of the European Association of Poisons Centres and Clinical Toxicologists, 25 May–1 June 2012, London, UK. *Clin. Toxicol.* 2012, 50, 273–366. [CrossRef]
17. Strugariu, M.C.; Strugariu, A. Common Adder (*Vipera berus*) Bites in Northeastern Romania: A Retrospective Analysis. In *4th Biology of the Vipers Conference*, Athens, Greece, 10–13 October 2014, 1st ed.; Abstracts Book: Athens, Greece, 2014; p. 28.
18. Malina, T.; Babocsay, G.; Krecsák, L.; Erdész, C. Further Clinical Evidence for the Existence of Neurotoxicity in a Population of the European Adder (Vipera berus berus) in Eastern Hungary: Second Authenticated Case. Wilderness Environ. Med. 2013, 24, 378–383. [CrossRef] [PubMed]

19. Boels, D.; Hamel, J.F.; Deguigne, M.B.; Harry, P. European viper envenomings: Assessment of Viperfav™ and other symptomatic treatments. Clin. Toxicol. 2012, 50, 189–196. [CrossRef] [PubMed]