RESEARCH ARTICLE

Combined Treatment with Static and Docetaxel Alters the Bax/Bcl-2 Gene Expression Ratio in Human Prostate Cancer Cells

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Abstract

Docetaxel, recognized as a stabilizing microtubule agent, is frequently administered as a first line treatment for prostate cancers. Due to high side effects of monotherapy, however, combinations with novel adjuvants have emerged as an alternative strategy in cancer therapy protocols. Here, we investigated the combined effects of static and docetaxel on the DU145 prostate cancer cell line. Cytotoxicity was evaluated by MTT assay. To understand molecular mechanisms of static action, apoptotic related genes including Bcl-2, Mcl-1, Survivin and Bax were evaluated by real-time RT-PCR. Alteration in the expression of pro-apoptotic Bax and anti-apoptotic Bcl-2 genes and Bax/Bcl-2 ratio were investigated via the \( \Delta \Delta CT \) method. The IC\(_{50}\) values for docetaxel and static were 3.7 ± 0.9 nM and 4.6±0.8 µM, respectively. Evaluation of key gene expression levels revealed a noticeable decrease in antiapoptotic Bcl-2 and Mcl-1 along with an increase in pro-apoptotic Bax mRNA levels (p<0.05). Our results suggest that combination of a STAT3 inhibitor with doctaxel can be considered as a potent strategy for induction of apoptosis via increasing Bax mRNA expression.

Keywords: Prostate cancer cells- apoptosis, docetaxel- monotherapy- combination chemotherapy- static

Introduction

Cancer is recognized as second leading causes of death worldwide, headed by cardiovascular and infectious diseases (Albini, DeCensi et al., 2016). Worldwide, prostate cancer is the most prevalent cause of cancer-related death in men with 27,540 deaths, while lung cancer causing death both in men and women, is responsible for 158040 deaths yearly, as on 2015 (Siegel, Miller et al., 2015, Fabre, Grosman et al., 2016). Nullification of apoptosis and continuous cell survival are critical step in the expansion of cancer (Fiandalo and Kyprianou 2012). Activation of prosurvival signal transcription factor STAT3, is among the most typical molecular alterations affecting cell survival and therapeutic susceptibility determined in cancers. Activation of STAT3 has been individually shown to appear in the majority of carcinomas such as lung and prostate. Constitutive activation of STAT3 is detected in the majority of prostate cancer cell lines, and established to promote cell survival, tumor development and resistance to apoptosis (Kamran, Patil et al., 2013, Yu, Lee et al., 2014). Apoptosis is organized by Bcl-2 family members, which consists of both anti-apoptotic genes such as Bcl-2, Bcl-xl and Mcl-1 and pro-apoptotic genes such as Bad, Bak and Bax (Pirouzpanah, Sabzichi et al., 2014). Functions of the apoptotic pathways are assisted by regulation of expression of a family of pro-apoptotic and anti-apoptotic proteins, and the ratio of expressed pro and anti-apoptotic proteins has been conveyed to establish whether cells survive or sustain apoptosis (Thomas, Quinn et al., 2013, Czabotar, Lessene et al., 2014). STAT3 can upregulate transcription of antiapoptotic protein Bcl-2, that manages cell growth and survival. Extensive studies involving the investigation of Bcl-2 and Bax expression in cultured cells have demonstrated that Bcl-2 protein can be practically characterized as an apoptosis-suppressing factor whereas the Bax protein is more practically characterized as a key apoptosis-promoting factor (Park, Kundu et al., 2014, Jahanafrooz, Motameh et al., 2015). A low relative ratio of Bax to Bcl-2 has been discovered in prostate cancer, and overexpression of Bcl-2 has been established to suppress the initiation of apoptosis and promote resistance to anticancer drugs and radiation therapy. The intracellular ratio of Bax/Bcl-2 genes can strongly control the ability of a cell to react to an apoptotic signal. Then, a cell with a high Bax/Bcl-2 ratio will be more susceptible to apoptotic stimuli when compared to a similar cell type with a relatively low Bax/Bcl-2 ratio (Huang, Yang et al., 2012, Lee, Jung et al., 2012, Del Principe, Dal Bo et al., 2016).

Multiple chemotherapeutic agents have been characterized to induce apoptosis in cancer cells (Sui, Kong et al., 2014). An adverse side effect of taxan

1Drug Applied Research Center, Department of Medical Biotechnology; 2Students’ Research Committee; 3Biotechnology Research Center; 4Immunology Research Center, Tabriz University of Medical Sciences, Tabriz, Iran. *For correspondence: drnsamadi@yahoo.com
derivatives (i.e. docetaxel) is a critical challenge that limits the efficacy of cancer therapy protocols. The innovation of novel inhibitors and also an application of combination therapy strategy shed light on chemoresistance and drug toxicity of current chemotherapeutic regiment (Fizazi, Higano et al., 2013, Swain, Baselga et al., 2015). Stattic (6-Nitrobenzo[b]thiophene-1,1-dioxide) is a small molecule inhibitor of STAT3. It selectively inhibits STAT3 activation, dimerization, and nuclear translocation by preventing the binding of tyrosine-phosphorylated peptide motifs to the STAT3 SH2 domain (Pan, Zhou et al., 2013).

In this study we applied STAT3 inhibitor, stattic concomitant with docetaxel to improve the efficacy of this chemotherapeutic agent in the induction of apoptosis. Furthermore, this study exhibits for the first time that stattic in combination with docetaxel down-regulated Survivin, Bcl-2 and Mcl-1 expression, up-regulated Bax mRNA level and altered Bax/Bcl-2 gene expression ratio in A549 and DU145 cells. Our results suggest that STAT3 inhibitors can be applied as effective adjuvants to increase docetaxel-induced cell death in prostate cancer cells.

Materials and Methods

Cell culture

Human prostate cancer (DU145) cell line was obtained from National Cell Bank of Iran (Pasteur Institute, Iran). The cells were cultured under the condition of 37 °C, 5% CO2, in RPMI (Roswell Park Memorial Institute)1640 Medium (Gibco®, Invitrogen, USA) with 10% FBS (Fetal Bovine Serum) enriched with 100 units/ml streptomycin/penicillin (Sigma, St Louis, MO, USA).

Cell viability assessment using MTT assay

Cells were seeded in the 96-well plate (2000 per well) in triplicate and treated with 1 to 16 mmol/L of docetaxel and 1 to 16 μmol/L of stattic for 24, 48 and 72 hours. The media in each well was replaced with 200 μl fresh media containing 20 μl of MTT (Methylthiazolylidiphenyl-tetrazolium bromide) solution (2 mg/ml). Then; the cells were incubated for 4 hours at 37 °C. Subsequently, media/MMT mixture was removed and 200μl of DMSO (Dimethyl sulfoxide) plus 25μl of Sorenson’s glycine buffer as solubilization solution was added to each well (Sabzichi, Samadi et al., 2016, Tupal, Sabzichi et al., 2016). The absorbance at 570 nm was read after shaking for 30 minutes, employing a microplate reader (Awareness, statfax 3200, USA). Half maximal inhibitory concentration (IC50) was calculated using Graphpad prism 6.07 software.

Cell viability analysis using trypan blue staining

Trypan Blue is a “vital stain” in which dead cells were excluded from live cells. Live cells appear colorless and bright (retractile) while dead cells stain blue and are non-retractile. Cells were re-suspended with equal volumes of 0.4% trypan blue stain for 3 min. Then, the cells were counted by hemocytometer (Wang, Zhu et al., 2016). The number of cells from the untreated control represented optimal cell survival (100%) and the relative surviving cells of each treatment were calculated using the following equation:

Relative cell viability (%) = \frac{no\; surviving\; cells\; in\; sample}{no\; surviving\; cells\; in\; control} × 100%

The experiments were repeated three times separately, to confirm reproducibility.

RNA extraction and reverse transcriptase-PCR (RT-PCR)

Cells and DU145 were treated with stattic (2μM), docetaxel (2nM) and the combination of them (docetaxel 1nM+stattic 1μM) and incubated for 24 h. RNA extraction of treated cells was carried out using TRI reagent (Sigma-Aldrich), according to the manufacturer’s instructions. The quality of RNA was evaluated by assessing the integrity of total RNA on agarose gel electrophoresis and the concentration of RNA was determined by measuring the absorbance at A260/A280 ratio with NanoDrop 1,000 spectrophotometer (Wilmington, DE, USA). After reverse transcription, cDNA was consequently amplified by regular PCR method with the primers presented in Table1. The PCR products were separated using electrophoresis through 1% agarose gels containing Ethidium bromide and PCR products were visualized under UV illumination (Minaei, Sabzichi et al., 2016).

Quantitative real-time PCR

Cells were treated with different concentrations of docetaxel and stattic and incubated for 24 h. Total RNA was isolated from cell pellets using TRI reagent according to the protocol as described previously (Pirouzpanah, Sabzichi et al., 2014, Armat, Bakhshaiesh et al., 2016). Following reverse transcription, cDNA was subsequently amplified by quantitative real-time PCR, using QPCR Green Master with low ROX (California, USA) with primers presented in Table1. The PCR reaction was performed using the Roche Light Cycler® (Germany).

Statistical analysis

All statistical analyses were carried out by Graphpad prism V6.07 (GraphPad Software Inc., San Diego, CA, USA). The statistical significance was determined using T-test for MTT assay and one-way ANOVA for Real Time PCR and p<0.05 was considered to be statistically significant.

Results

Anti-proliferative effects of docetaxel and stattic on DU145 cells (using MTT and Trypan Blue cell count assays)

Anti-proliferative activity of docetaxel and stattic was measured by MTT assay using DU145 cell. IC50 values for stattic and docetaxel in different concentration shown in Table 2. After 24 h incubation, MTT and Trypan Blue cell count assays indicated that DU145 cells exposed to stattic and docetaxel had significantly lower cell viability than the untreated control. In contrast, the two assays showed stattic and docetaxel treatment had a remarkable difference in cell viability compared the control.

On the other hand, after 48 and 72 h treatment, MTT and Trypan Blue cell count assays demonstrated that
DU145 cells exposed to static and docetaxel (Figure 1 A-D) had significantly decreased cell viability compared to 24 h treatment and the untreated control (p < 0.05).

Semi-quantitative RT-PCR
For semi-quantitative RT-PCR, the cells were incubated with docetaxel at the concentration of 2nM, static 2µM (IC50 values) and their combination for 24h. Our semi-quantitative data showed a remarkable down-regulation of anti-apoptotic Mcl-1, Bcl-2 and Survivin genes and up-regulation of pro apoptotic Bax compared to single treatment sample. Results, after normalization against β-actin, are consistent with reduction of Mcl-1, Bcl-2 and Survivin and increase of Bax transcripts in DU145 cell line but in the combinatorial treatment reduction of transcripts are more considerable (Figure 2B).

Combined effect of static and docetaxel on Bcl-2, Mcl-1, Survivin and Bax mRNA expression levels and Bax / Bcl-2 ratio
Real time RT-PCR was performed to quantitate levels of the transcripts. For DU145 cell line, the only concentration of 2nM docetaxel and 2µM static and their combination (docetaxel 1nM + static 1µM) were tested in parallel with untreated cells. Expression of Bcl-2, Mcl-1 and Survivin genes decreased in cells treated with docetaxel (2nM) and static (2µM) and remarkably decreased in combination treatment (static+docetaxel) after 24h incubation. On the other hand expression of Bax gene increased after treatment with the aforementioned condition in which in combinational treatment was increased more significantly (Figure 2A). Our results also revealed a considerable increase in Bax/Bcl-2 ratio in treatment by docetaxel (2nM) and static (2µM). The most increase in Bax/Bcl-2 ratio was seen in combinational treatment after 24h incubation (Figure 3).

Table 1. Primers Used for qRT-PCR Analysis

| Objective genes | Primer sequence |
|-----------------|----------------|
| Bcl-2           | Forward: 5’-CATCAGGAAGGCTAGATAGTTACC-3’ |
|                 | Reverse: 5’-CAGACATCGGAGACCCACAC-3’ |
| Mcl-1           | Forward: 5’-GGACACAAAGACCAATGGGCGAGT-3’ |
|                 | Reverse: 5’-GCAAAAGCCGCAGCACATCCTCGTA-3’ |
| BAX             | Forward: 5’-GATGCGGCCCAAGAGAAG-3’ |
|                 | Reverse: 5’-AGTTGAAGTTGGCCTACAG-3’ |
| Survivin        | Forward: 5’-GACCCACGCATCTCTCAATCG-3’ |
|                 | Reverse: 5’-TGCTTTTTATGTCCTTCTGAGG-3’ |
| β-actin         | Forward: 5’-TGCCCCTCTCAGGAGGATG-3’ |
|                 | Reverse: 5’-CTCCCTAATGTCACGCAGATT-3’ |

Figure 1. DU145 Cells Exposed to Docetaxel and Static Treatments for 24, 48 and 72 h Determined by: MTT (A–C); Trypan Blue Cell Count (B–D). Data are Expressed as Cell Viability (%) ± SD of Three Independent Experiments.

Table 2. IC50 Values of Docetaxel and Static in DU145 Cells

| Agents       | Incubation time | IC50     |
|--------------|----------------|----------|
| Docetaxel (nM) | 24             | 3.7 ± 0.9 |
|              | 48             | 2.4 ± 1.0 |
|              | 72             | 1.7 ± 0.6 |
| Stattic (µM)  | 24             | 4.7 ± 0.8 |
|              | 48             | 2.9 ± 0.9 |
|              | 72             | 1.7 ± 0.9 |

Table 2. IC50 Values of Docetaxel and Static in DU145 Cells

Agents       | Incubation time | IC50     |
--------------|----------------|----------|
Docetaxel (nM) | 24             | 3.7 ± 0.9 |
              | 48             | 2.4 ± 1.0 |
              | 72             | 1.7 ± 0.6 |
Stattic (µM)  | 24             | 4.7 ± 0.8 |
              | 48             | 2.9 ± 0.9 |
              | 72             | 1.7 ± 0.9 |

Figure 2. Effect of Static, Docetaxel and Their Combination on the Expression of Bcl-2, Survivin, Mcl-1 and Bax in DU145 cells. A. The levels of Bcl-2, Survivin, Mcl-1 and Bax mRNA expression, followed by treatment with docetaxel (2 nM), static (2 µM) and their combination (static 1 µM+docetaxel 1 nM) for 24h, were determined using real-time PCR. Mean ± SD was calculated from three independent experiments(*p<0.01). B. Levels of Bcl-2, Mcl-1, Survivin and Bax expression were analyzed. Bands representing Bcl-2, Mcl-1, Survivin and Bax are shown.
Jamal Mohammadian et al
Asian Pacific Journal of Cancer Prevention, Vol 17, 4322-7.

5034

Jamal Mohammadian et al

57

apoptotic pathway. Real time PCR analysis revealed the apoptotic effect of docetaxel, we performed some colon cancer cells (Lee, Jung et al., 2012). Increases the expression ratio of Bax/Bcl-2 in human with previous studies demonstrating that sanguinarine alteration in Bax/Bcl-2 ratio. This finding was consistent efficacy of docetaxel and induction of apoptosis via investigate whether stattic enhance the anti-proliferative Therefore, we carried out this study using DU145 to

line, DU145, are popular for investigating the biochemical (Zhang, Zhang et al., 2015). Human prostate cancer cells SH2 domain despite of STAT3 phosphorylation state (Zhang, Zhang et al., 2015). The mitochondrial apoptosis pathway is the most significant pathway by regulating the ratio of apoptotic proteins Bcl-2 and Bax. Exclusively, Bcl-2 suppresses apoptosis partly by preventing the release of cytochrome c from mitochondria, and the initiation of apoptosis while Bax has an apoptosis-promoting effect by antagonizing Bcl-2. The balance between the expression levels of two apoptosis regulatory genes, Bcl-2 and Bax is important for cell survival and death since the increase in Bax/Bcl-2 ratio contributes to the efflux of cytochrome C from mitochondria and activates the intrinsic mitochondrial apoptotic pathway. As a result alteration in the expression of Bax and Bcl-2 expression determines cell susceptibility to apoptosis. An increased Bax/Bcl-2 ratio can activate caspase-3 and result in cell death(Renault, Teijido et al., 2013, Sharifi, Barar et al., 2015).

In prostate cancer, it is identified that apoptosis plays a role in response to chemotherapy, recommending a relationship between drug-induced apoptosis and therapeutic efficiency (Sun, Bao et al., 2013). As a result, the analysis of mitochondrial apoptotic genes may lead to introduce a key prognostic tool to predict resistance to chemotherapeutic agents. Independently, the functions of Bcl-2 and Bax have been studied widely in prostate and other cancers, providing facts of their opposing function in cell survival and apoptosis (Zhang, Bi et al., 2014).

In this study, the alteration in Bcl-2 (anti-apoptotic), Bax (pro-apoptotic) gene expression, Bax/Bcl-2 ratio, subsequent to the stimulation of apoptosis by stattic, docetaxel and their combination in DU145 cells were reported. Our results in DU145 cells demonstrated that in vitro treatments induced a decreased expression of the antiapoptotic oncogene Bcl-2, which determines an augment in Bax/Bcl-2 ratio after treatment. Our investigation demonstrated that stattic augmented docetaxel-induced cytotoxicity and also alter expression ratio of Bax/Bcl2 in the prostate cancer cell, DU145. Our findings suggested that utilizing stattic along with docetaxel can be regarded as a novel approach to reach more effectual results in the patient with prostate cancer.

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Conflict of Interest

The authors declare that there are no conflicts of interest.

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