miR-98 inhibits cell proliferation and induces cell apoptosis by targeting MAPK6 in HUVECs

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Abstract. The aim of current study was to explore the role of microRNA (miR)-98 in atherosclerosis. Human vascular endothelial cells (HVECs) were isolated from the peripheral blood of healthy volunteers and patients with atherosclerosis. Compared with endothelial cells from the healthy control group, the expression level of mitogen activated protein kinase (MAPK)6 was significantly upregulated and miR‑98 was downregulated in the endothelial cells of patients with atherosclerosis. The human umbilical vein endothelial cell line (HUVEC) was adopted to perform in vitro studies. Overexpression of miR‑98 reduced the proliferation and induced the apoptosis of HUVECs, which were revealed using an MTT assay, and flow cytometry assay, respectively. The aforementioned influences of miR-98 on HUVECs were mediated by targeting MAPK6, which was verified using luciferase assays. Additionally, the overexpression of miR-98 reduced the protein level of apoptosis regulator Bcl-2 and MAPK6; however, it induced the protein expression of caspase-3 and apoptosis regulator Bax. In conclusion, these findings demonstrate that miR-98 is an important regulator of atherosclerosis, suggesting that miR-98 may be a potential therapeutic target for the treatment of atherosclerosis.

Introduction

Atherosclerosis, a unique form of arteriosclerosis, presents as arterial wall thickening due to the invasion/accumulation of white blood cells and proliferation of intimal smooth muscle cell (1,2). Vascular endothelium contributes to the development of atherosclerosis via abnormal cell proliferation and apoptosis (3). Endothelial cells maintain the homeostasis of the vascular system (4). However, the precise molecular mechanisms which underlie the contributions of the vascular endothelium to atherosclerosis remain unclear and complex.

After binding to the 3’ untranslated region (UTR) of target genes, microRNAs (miRNAs, miR) repress protein expression levels by mRNA destabilization or/and translation inhibition (5). Recent studies also found that miRNAs had the ability to reduce mRNA targets by binding to 5’UTR or protein-coding exons (6,7). Circulating miRNAs serve critical roles in multiple diseases, including atherosclerosis (8) and cancer (9), thus showing enormous potential as novel biomarkers for various diseases. Consequently, it has emerged that miRNAs function as epigenetic regulators in diverse biological processes, including cell proliferation (10) and tumorigenesis (11). In diseased vascular vessels, dysregulation of miRNAs, for instance, miR-33 (12) and miR-133 (13), has been detected.

Mitogen activated protein kinase 6 (MAPK6) is an enzyme which is encoded by the MAPK6 gene in humans (14). MAPK6 has been shown to modulate cell migration, proliferation and angiogenesis in primary human umbilical vein endothelial cells (HUVECs), and has been reported to play a key role in maintaining normal physiological conditions of vascular smooth muscle cells (VSMCs) (15). In addition, in a previous study, the knockdown of MAPK6 damaged tube formation by VSMCs (16).

However, it remains unclear whether MAPK6 could be regulated by miRNA in the functions of the vascular endothelium. The current study aimed to clarify this.

Patients and methods

Participants. All study participants provided written informed consent. The current study was approved by the Ethics Committee of the Huai’an First People’s Hospital (Jiangsu, China). Sixty atherosclerotic patients (mean age, 60.6 years; 36 male and 24 female) and 51 healthy volunteers (mean age, 56.5 years; 31 male and 20 female) were enrolled in the Huai’an First People’s Hospital. Human vascular endothelial cells (HVECs) were isolated from the peripheral blood of healthy volunteers and patients with atherosclerosis using a human vascular endothelial cell separation medium kit (Sangon Biotech, Shanghai, China). HVECs were used for detection of the differences of MAPK6/miRNAs between healthy volunteers and patients with atherosclerosis.

Cell culture and transfection. HUVECs were used for the conducting of the following in vitro experiments. Briefly, HUVEC cell dissolution was performed by keeping in 37°C
for 5 min, transferred into a centrifuge tube and centrifuged (125 g, 5-7 min). HUVEC cell retrieved was balanced by RPMI (Lonza, Allendale, NJ, USA), 10% fetal bovine serum (Sigma, St. Louis, MO, USA) and inoculated on 60×20 mm Petri dishes (1.3x10^6 cells/ml). Endothelial cells were cultivated in 5 ml culture medium, inoculated in flasks overnight at 37°C <5% CO₂ with culture media changing every other day. When cellular density reached 80%, passaging was performed at passaging rates of 1:2 or 1:3. After treatment of trypsin, endothelial cells in suspension were centrifuged, the obtained pellets were re-suspended in culture medium and inoculated on new Petri dishes.

HUVECs were seeded in Dulbecco's modified Eagle's medium containing 10% fetal bovine serum, 1% 100 U/ml penicillin and 1% 100 mg/ml streptomycin, thereafter, cultured in an incubator with 5% CO₂ at 37°C. As for cell transfection, hsa-miR-98 mimic or hsa-miR-negative control (NC) mimic was transfected into HUVECs by opti-MEM and Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA) in accordance with manufacturer's instructions.

Luciferase reporter assays. The binding site between hsa-miR-98 and MAPK6 was predicted by miRanda. For luciferase reporter assay, HUVECs were seeded in 24-well plates, afterwards, HUVECs were transfected with WT or mutant reporter plasmid and hsa-miR-98 mimic or hsa-miR-NC mimic by Lipofectamine 2000. After 24 h, dual luciferase reporter assay kit (Promega, Madison, WI, USA) was used for the determination of relative firefly and renilla luciferase activities in different groups according to the protocol provided by manufacturer. Firefly luciferase activity acted as a control.

Reverse transcription-quantitative PCR (RT-qPCR). Total RNA was extracted from cultured cells using TRIzol (Invitrogen) according to the manufacturer's instructions, miScript SYBR-Green PCR kit (Qiagen, Valencia, CA, USA) was used for the RT-qPCR analysis, which was conducted on an Applied Biosystems 7900HT fast real-time PCR system (Applied Biosystems, Foster City, CA, USA). U6 acted as a control for miR-98 and GAPDH acted as a control for MAPK6. Primers were as: MAPK6 forward, 5'-TAAAGCCAT TGACATGTGG-3' and reverse, 5'-TGCTGCCACAACAGGG ATAGA-3'; GAPDH forward, 5'-ACAAGATGTTGAAGG TCGGTGTTGA-3' and reverse, 5'-AGCCTCCCATCTCA GCCTTGACT-3'; miR-98, 5'-CCGAGGTAGTAGTTGTA TTGTT-3'; U6, 5'-ACGCAAATTGTGGACACGGT-3'. At last, relative level of hsa-miR-98 and MAPK6 in cells was quantified with the method of 2^-ΔΔCT.

Cell apoptosis assay. At 48 h after cell transfection, HUVECs were harvested and washed with PBS for three times. HUVEC cells were added with 5 µl Annexin V-FITC and 5 µl PI, stained in the dark for 15 min with Annexin V-FITC apoptosis detection kit (KeyGEN Biotech, Nanjing, China). Relative rate of Annexin V-FITC or PI-positive HUVECs in each group was analyzed by flow cytometry.

Cell proliferation assays. MTT assay (Sigma) was adopted for the analysis of cell proliferation of HUVECs in each group. HUVECs were first seeded in 96-well plates and transfected with different plasmids, at 24 h post-transfection, 25 µl MTT (5 mg/ml) was added into each well, plates were incubated at 37°C in an incubator for 3 h. Precipitates in each well were solubilized by 150 µl dimethyl sulfoxide (DMSO), assessed at 480 nm with versaMax ELISA microplate reader (Molecular Devices, Sunnyvale, CA, USA).

Western blotting. Proteins were extracted from HUVECs by RIPA lysis buffer (Invitrogen) and separated by 12% sodium dodecyl sulfate poly-acrylamide gel electrophoresis before transferring to polyvinylidene fluoride membranes (Millipore, Danvers, MA, USA). After blocking with 5% nonfat milk at 37°C for 2 h, membranes were incubated with the primary antibodies [Bax (dilution, 1:1,000), Bcl-2 (dilution, 1:1,000), caspase-3 (dilution, 1:1,000), (all from Cell Signaling Technology), MAPK6 (dilution, 1:500) and GAPDH (dilution, 1:1,000) (both from Proteintech, Rosemont, IL, USA)] over night at 4°C, then HRP-linked secondary antibodies for 2 h at room temperature. At last, protein bands were visualized by enhanced chemiluminescence (ECL) (General Electric Healthcare, Aurora, OH, USA). GAPDH was used as a control.

Statistical analysis. Data were analyzed by SPSS 17.0 (SPSS, Chicago, IL, USA) with Student's t-test and ANOVA analyses. Results were expressed as mean ± standard deviation. P<0.05 was considered as statistically significant.

Results

Different expression levels of MAPK6 in the HVECs of healthy volunteers and atherosclerotic patients. MAPK6 modulates the migration, proliferation and angiogenesis of primary HVECs (15). Therefore, the expression levels of MAPK6 in HVECs from healthy individuals and subjects with atherosclerosis were detected by RT-qPCR. The expression level of MAPK6 was found to be significantly upregulated in
the HVECs of patients with atherosclerosis when compared with HVECs from healthy volunteers (P<0.01; Fig. 1).

**Prediction of the targets of hsa-miR-98.** Mounting evidence suggests that, in cardiovascular biology, miRNAs have the ability to regulate gene transcriptional decay and repression by binding target miRNAs (17). A bioinformatic analysis of miRanda was carried out to identify potential miRNAs that target MAPK6, which exhibited a potential seed sequence of hsa-miR-98 in MAPK6-3’-UTR (Fig. 2).

**Different expression levels of miR-98 in the HVECs of healthy volunteers and atherosclerotic patients.** As acknowledged, miRNAs serve pivotal roles in atherosclerosis, and hsa-miR-98 which was reported to be aberrantly expressed in the serum of atherosclerotic patients was selected for the current research according to the results in a microarray analysis (18). The expression levels of hsa-miR-98 in the HVECs from healthy and atherosclerotic subjects were investigated by analysis using RT-qPCR. The expression of hsa-miR-98 was found to be significantly downregulated in the HVECs from atherosclerotic patients compared with the healthy volunteers (P<0.01; Fig. 3).

**Effects of overexpression of miR-98 on cell proliferation.** To investigate the potential influences of hsa-miR-98 on cell proliferation, HUVECs were treated with hsa-miR-98 mimics or miR-NC mimic at the concentration of 50 nmol/l for 48 h. Thereafter, MTT assay was used to assess the influences of hsa-miR-98 on HUVEC proliferation. The results showed that cells transfected with hsa-miR-98 mimics grew much more slowly than cells in the control group and miR-NC mimic group (P<0.01; Fig. 4).

**Effects of overexpression of miR-98 on cell apoptosis.** To explore the functions of hsa-miR-98 in HUVECs, the cells were transfected with 100 nmol/l hsa-miR-98 mimic or miR-NC mimic and cultured for 48 h. Afterwards, flow cytometry assay of cell apoptosis was carried out which demonstrated that overexpression of hsa-miR-98 in HUVECs induced an apoptosis rate higher than in those in the control group and miR-NC mimic group (P<0.01; Fig. 5).

**Effects of overexpression of miR-98 on Bax, Bel-2, caspase-3 and MAPK6 protein levels.** Western blotting was performed to explore the protein levels of Bax, Bel-2, caspase-3 and MAPK6 following the administration of hsa-miR-98 mimic or miR-NC mimic. We found that the levels of MAPK6 and Bel-2 were lower, while those of Bax and caspase-3 were higher in the hsa-miR-98 mimic group compared with the control group and miR-NC mimic group (Fig. 6).

**Verification of an interaction between miR-98 and MAPK6.** Luciferase assays showed that miR-98 repressed wild-type (WT) MAPK6-3’-UTR-luciferase activity but not mutant MAPK6-3’-UTR-luciferease reporter. This result verified that hsa-miR-98 targeted MAPK6 in HUVECs (Fig. 7).

**Discussion**

Atherosclerosis presents as arterial wall thickening (1,2). Endothelial dysfunction occurs initially during the progression of atherosclerosis (19), and is followed by the proliferation and migration of smooth muscle cells (20).

MAPK6 modulates the cell proliferation and angiogenesis of primary HUVECs, and maintains the physiological condition of VSMC (15), whereas its knockdown damages the tube formation of VSMC (16). We first examined MAPK6 mRNA levels in HVECs from healthy and atherosclerotic subjects, and found that MAPK6 was significantly higher in the HVECs of patients with atherosclerosis compared with those of healthy volunteers (Fig. 1).
miR-98 INHIBITS CELL PROGRESSION OF HUVECs

Figure 4. miR-98 mimic repressed the cell proliferation of HUVECs. MTT assay showed slower growth of HUVECs transfected with hsa-miR-98 mimic than cells in the control group and miR-NC mimic group (A and B). \( P<0.01 \) miR‑98 mimic vs. control group and miR‑NC mimic. miR, microRNA; HUVECs, human umbilical vein endothelial cells; NC, negative control.

Figure 5. miR-98 mimic induced the cell apoptosis of HUVECs. Flow cytometry assay demonstrated higher apoptosis of HUVECs transfected with hsa-miR-98 mimic than cells in control group and miR-NC mimic group (A and B). \( P<0.01 \) miR‑98 mimic vs. control group and miR‑NC mimic. miR, microRNA; HUVECs, human umbilical vein endothelial cells; NC, negative control.
mRNAs have the ability to bind to the 5’UTR or 3’UTR of target genes to inhibit their expression (5-7). Circulating miRNAs are potentially functional in the atherosclerosis associated dysfunction of gene regulatory networks (21) as well as biological processes, for instance, cell proliferation (10). However, it remains unclear whether MAPK6 could be regulated by miRNA in the functioning of the vascular endothelium.

miRanda was adopted to identify miRNAs that could target MAPK6, and a potential seed sequence of hsa-miR-98 was exhibited in the MAPK6-3’-UTR (Fig. 2). Thereafter, we identified the expression levels of hsa-miR-98 in HVECs from healthy and atherosclerotic subjects, and found that hsa-miR-98 was significantly lower in the HVECs of patients with atherosclerosis compared with healthy individuals (Fig. 3), which was consistent with a previous study (18).

The potential influences of hsa-miR-98 on HUVEC cell proliferation and apoptosis were tested by MTT assay and flow cytometry assay, respectively. Results showed slower growth (Fig. 4) and a higher apoptotic rate (Fig. 5) in HUVECs transfected with hsa-miR-98 mimic than in control group and miR-NC mimic group.

Bcl-2 and caspase families participate in the apoptotic process. Bcl-2 protein families include Bax (pro-apoptotic) and Bcl-2 (antiapoptotic) (22). Moreover, since caspase-3 is the most reliable determining factor during apoptosis (23), Bcl-2 regulates apoptosis via a caspase-3 dependent pathway (24). Western blotting results showed that MAPK6 and Bcl-2 levels were lower, while Bax and caspase-3 levels were higher in the hsa-miR-98 mimic group than in control group and miR-NC mimic group.

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Finally, the interaction between miR-98 and MAPK6 was tested by luciferase assay. The results verified that hsa-miR-98 targeted MAPK6 in HUVECs (Fig. 7).

In conclusion, miR-98 inhibited HUVEC proliferation, promoted HUVEC apoptosis, lowered MAPK6/Bcl-2 and upregulated Bax/caspase-3 via targeting MAPK6. Thus miR-98 might be a therapeutic target for atherosclerosis.
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