Genetic polymorphism in the adiponectin (*ADIPOQ*) gene and its association with the production and reproduction traits of Indian dairy cattle

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**ABSTRACT**

The present investigation was performed to explore the SNP in the promoter of the adiponectin (*ADIPOQ*) gene and its association with production and reproduction traits in Indian Sahiwal cows. A portion of the promoter (*ADIPOQ*) region of the *ADIPOQ* gene was amplified which revealed a 977 bp amplicon, and its PCR-RFLP assay with the restriction enzyme *Tas*I revealed three genotypes, in which the CT genotype was the most frequent (62.32%), followed by CC (24.64%) and TT (13.04%). The frequency of the C and T alleles was 0.558 and 0.442, respectively. The *ADIPOQ/Tas*I genotypes revealed a significant association with calving interval (CI) in the first and fourth lactations, in which the CC genotype showed a significantly (P<0.05) longer CI compared to the TT genotype, while in the fourth lactation the TT genotype showed a longer CI compared to the CC genotype. In the fourth lactation, the TT genotype showed a significantly (P<0.05) longer lactation period (LP) and greater total milk yield (TMY) as compared to CC and CT genotypes. In conclusion, the SNP identified in the promoter of the *ADIPOQ* gene and its association with production and reproduction traits suggests that this gene might serve as a candidate genetic marker for selection of dairy cattle with better milk yield. However, further studies are needed to explore these SNPs in other regions of this gene, and in other breeds and populations.

**Key words:** *ADIPOQ* gene; PCR-RFLP; SNP; reproduction traits; production traits; cattle

**Introduction**

Adiponectin is one of the most ample adipokines in the circulation that shows a negative correlation with fat mass (Kadowaki and Yamauchi, 2005) and is reported to play important role in the regulation of fatty acid oxidation, glucose metabolism, insulin sensitivity, immunity, and reproduction (Waki et al., 2003; Brochu-Gaudreau et al., 2010). The natural expression patterns of adiponectin and their receptors in follicular and luteal cells of bovine ovary have also

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been reported to affect the physiological status of the ovary (TABANDEH et al., 2010; MAILLARD et al., 2011). The information, regarding the role of adiponectin in energy metabolism and ovarian function, suggests that adiponectin could affect production and reproduction in dairy animals. Furthermore, the ADIPOQ gene has been found to be located within the bovine chromosome (BTA) 1 region, which was previously reported to harbor QTL (KIM et al., 2010) that affects carcass traits in crossbred cattle. Moreover, several single nucleotide polymorphisms (SNPs) have been identified in the promoter, intron and 3′ UTR (untranslated region) regions of the ADIPOQ gene and have revealed a significant association with meat quality traits in various breeds of beef cattle (CASAS et al., 2000; DAVIS et al., 1998; LI et al., 2004; MORSCI et al., 2006; ZHANG et al., 2009). Since genetic variability in the ADIPOQ gene was found to play a role in the phenotypic expression of carcass traits in animals, it was hypothesized that genetic variability in this gene may also play a role in the phenotypic expression of the production and reproduction traits of dairy cows. Most research into genetic polymorphism in the ADIPOQ gene has been conducted regarding their association with carcass traits in cattle, and no study related to their association with the production and reproduction traits of cattle has yet been undertaken. Thus, the present study was designed to explore the influence of genetic polymorphisms in the ADIPOQ gene and their possible association with production and reproduction traits in Sahiwal cows.

Materials and methods

Sampling and DNA extraction. Seventy Sahiwal cattle, maintained at the Instructional Livestock Farm Complex (ILFC) of the College of Veterinary Science and Animal Husbandry, Mathura, were used for the study. Approximately 5 mL of blood was collected from the jugular vein in vacutainer tubes containing EDTA as an anticoagulant, and genomic DNA was isolated as per the standard phenol-chloroform isolation protocol (SAMBROOK and RUSSELL, 2001). The quality and quantity of DNA was determined using a spectrophotometer, by measuring the optical density ratio at a wavelength of 260 and 280 nm.

PCR amplification. The following primer set (Table 1) was used to amplify the genomic sequence of the promoter region, based on the genomic sequence of the bovine ADIPOQ gene from GenBank Accession No. AH015166.2.

A total of 100-150 ng of template DNA was amplified in a total volume of 25 μL PCR mix in the thermocycler (Bio-Rad, USA). The PCR mix contained: 2.5 μL PCR dream buffer 10X, 2.5 μL of 2mM dNTPs, 0.5 μL from each primer and 1 U Taq DNA polymerase. The thermal cycling was standardized at 94 °C for 5 min for initial denaturation, followed by 35 cycles of denaturation at 94 °C for 30s, annealing at 57.5 °C for 30s and extension at 72 °C for 1 min. The final extension step was carried out for 5 min at 72 °C.

PCR-RFLP of ADIPOQ gene. Genotype analyses of ADIPOQ gene was carried out using the polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) method. For this, the PCR product of the ADIPOQ gene was digested using Fast Digest TasI (Thermo Scientific) at 65 °C for 5 minutes in a hot water bath. The digestion reaction contained 10 μL PCR product, 1 μL of 10X Buffer, 0.5 μL restriction enzyme and 8.5 μL de-ionized water. The fragments were separated by horizontal gel electrophoresis in 2% agarose gel stained with ethidium bromide (1 μg/μL) and electrophoresed in 1X TBE buffer prior to visualization under UV light.

| Gene   | Primer sequence | Region           | Reference                  |
|--------|-----------------|------------------|----------------------------|
| ADIPOQ | F:5’-TTGTGTCTTCTGATATTGGCA-3’ | Promoter (AH015166.2) | SHIN and CHUNG, 2013       |
|        | R:5’-ACTGGACAAAAATTCAAGATG-3’ |                 |                             |
**Statistical analysis.** The allelic and genotypic frequencies (FALCONER and MACKAY, 1996) of the ADIPOQ/TasI polymorphism was examined for deviation from the Hardy-Weinberg equilibrium using the χ² test (SNEDECOR and COCHRAN, 1989), and statistical significance was determined by the ANOVA test, followed by Tukey’s post-hoc multiple comparison tests using SPSS software for Windows (version 16.0). The data were presented as the mean ± SEM and a P-value < 0.05 was considered to be statistically significant.

**Results**

**ADIPOQ/TasI PCR-RFLP assay.** The amplification of the ADIPOQ gene revealed a 977 bp amplicon which was observed on 1.0% agarose gel (Fig. 1).

![Fig. 1. Agarose gel electrophoresis (1%) of the ADIPOQ PCR product showing 977bp amplicons in all lanes (1-4), L= Ladder (100bp).](image)

The TasI/PCR-RFLP assay revealed three types of banding patterns (genotypes); one of them was the CC genotype, the second was the TT genotype and the third was the CT genotype (Fig. 2). The amplicon of the CC genotype had 4 restriction sites and thus revealed 5 fragments of 607, 243, 96, 18 and 13 bp. The amplicon of the TT genotype had 5 restriction sites that revealed six fragments of 454, 243, 153, 96, 18 and 13 bp. The amplicon of the CT genotype showed all seven fragments of 607, 454, 243, 153, 96, 18 and 13 bp. For the PCR-RFLP assay, the important fragment was the fragment of 607 bp which showed mutation in the sequence. The CC genotype showed an intact fragment of 607 bp, while the TT genotype revealed a restriction site and showed two fragments of 454 and 153, and thus the CT genotype showed all these three fragments (607, 454 and 153). Other fragments were common in all three genotypes. The results revealed that the population studied of Sahiwal cattle was polymorphic in nature, with two types of alleles, C and T, with three types of genotypes: CC genotype, TT genotype and CT genotype, in the screened samples.

Further, the presence of the restriction sites for TasI in the PCR amplified ADIPOQ gene was confirmed by DNA sequencing. The sequence of ADIPOQ/TasI obtained after alignment revealed C→T substitution at nucleotide position 1431C>T. For the CC genotype, no C→T substitution was
found, for the CT genotype C→T substitution was found in only one strand, for the TT genotype C→T substitution was found in both strands (Fig. 3).

The results of this study revealed that the CT genotype was the most frequent (62.32%) in all the screened samples, followed by the CC genotype (24.64%), whereas the TT genotype was least frequent in these samples (13.04%). The frequency of the C and T alleles of ADIPOQ/TasI was 0.558 and 0.442, respectively. The \( \chi^2 \) calculated value for the ADIPOQ/TasI gene was 27.48, while \( \chi^2 \) table values were 3.84 and 6.64 at 5% and 1% level of significance, respectively for the degrees of freedom 1. These results revealed that \( \chi^2 \) (cal)>\( \chi^2 \) (tab) at 1% level of significance, which indicates that the selected population of Sahiwal cattle was not in Hardy-Weinberg equilibrium (Table 2).

Association of ADIPOQ/TasI genotypes with production and reproduction traits. The means with standard errors (Mean ± SEM) for each trait related to each genotype for four lactations are given in Tables 3 and 4, respectively. The association study of the ADIPOQ/TasI PCR-RFLP assay revealed the significant influence of genotypes on calving interval (CI), lactation period (LP), total milk yield (TMY) and days to reach peak yield (DRPY). The genetic polymorphism of ADIPOQ/TasI revealed a significant effect on LP, and TMY in the fourth lactation in the TT genotype showed significantly higher values compared to the CC and CT genotypes. The DRPY of cows showed a significant association with genotypes in the second lactation, which revealed higher values for the CT genotype compared to the TT genotype.

![Chromatograms of ADIPOQ gene sequence showing C>T nucleotide substitution in the promoter region. (A) CC genotype (B) CT genotype (C) TT genotype](image)

**Table 2. Genotypic and allelic frequencies of C>T SNP/TasI genotypes in Sahiwal**

| Breed   | Genotypic frequency (%) | Allelic Frequency | \( \chi^2 \) test |
|---------|-------------------------|-------------------|-------------------|
|         | CC (n = 17)             | CT (n = 43)       | TT (n = 9)        | G  | T  | \( \chi^2 \) cal = 27.48 |
| Sahiwal | 24.64 (n = 17)          | 62.32 (n = 43)    | 13.04 (n = 09)    | 0.558 | 0.442 | \( \chi^2 \) tab = 6.64 (P<0.01) |

\( \chi^2 \) cal >\( \chi^2 \) tab
The CI of cows showed a significant association with genotypes in the first and fourth lactations. The CC genotype showed a significantly longer CI compared to the TT genotype, while in the fourth lactation the TT genotype showed a significantly longer CI compared to the CC genotype.

Discussion

Genetic polymorphism of the *ADIPOQ* gene (1431C>T) has not been widely studied in cattle breeds and most studies have been conducted in beef cattle regarding their carcass traits. This is the first study conducted so far to elucidate the effects of genetic polymorphism in the *ADIPOQ* gene on the production and reproduction traits of dairy cattle.

In the present investigation, the *ADIPOQ*/TasI assay revealed three genotypes. SHIN and CHUNG (2013) also found similar observations in the Hanwoo (Korean) cattle population, and revealed three genotypes, where CT had the highest genotypic frequency (68.9%), followed by TT (16.0%) and CC (15.1%) genotypes, with deviations from Hardy-Weinberg equilibrium in the studied population. These findings of genotypic frequency reported by SHIN and CHUNG (2013) corroborate well with the results of the present study. Furthermore, they reported the allelic frequency of C and T alleles as 49.6 and 50.4, respectively, which is also comparable with the findings of our present study. In contrast, MORSCI et al. (2006) also reported *ADIPOQ*/TasI polymorphism in Angus cattle.

### Table 3. Association of *ADIPOQ*/TasI genotypes with reproduction traits

| Lactation | Genotype | n  | BW (kg) | AFS (days) | AFC (days) | GP (days) | DP (days) | CI (days) |
|-----------|----------|----|---------|------------|------------|-----------|-----------|-----------|
| I (n = 55) | CC       | 12 | 20.75 ± 3.75 | 986.00 ± 54.00 | 1547.00 ± 201.00 | 281.50 ± 3.50 | 223.83 ± 37.43 | 639.25 ± 49.17 |
|           | CT       | 36 | 20.43 ± 0.65 | 1077.90 ± 72.36 | 1587.38 ± 63.61 | 278.33 ± 2.71 | 175.97 ± 17.43 | 554.92 ± 23.52 |
|           | TT       | 7  | 19.88 ± 0.97 | 1306.00 ± 110.10 | 1759.25 ± 62.80 | 284.25 ± 2.56 | 141.71 ± 35.55 | 502.00 ± 39.15 |
| II (n = 40) | CC       | 10 | 284.96 ± 1.94 | 117.20 ± 21.12 | 1077.90 ± 72.36 | 1587.38 ± 63.61 | 278.33 ± 2.71 | 175.97 ± 17.43 | 554.92 ± 23.52 |
|           | CT       | 25 | 286.40 ± 2.84 | 131.80 ± 66.55 | 1759.25 ± 62.80 | 284.25 ± 2.56 | 141.71 ± 35.55 | 502.00 ± 39.15 |
|           | TT       | 5  | 289.50 ± 8.26 | 162.00 ± 38.70 | 527.75 ± 33.08 | 431.20 ± 22.93 | 480.00 ± 22.93 | 457.00 ± 40.61 |
| III (n = 24) | CC       | 4  | 282.18 ± 1.35 | 144.59 ± 19.10 | 480.00 ± 22.93 | 457.00 ± 40.61 | 480.00 ± 22.93 | 457.00 ± 40.61 |
|           | CT       | 17 | 288.67 ± 2.73 | 99.67 ± 29.24 | 480.00 ± 22.93 | 457.00 ± 40.61 | 480.00 ± 22.93 | 457.00 ± 40.61 |
|           | TT       | 3  | 286.40 ± 3.12 | 115.80 ± 27.74 | 541.20 ± 43.28 | 448.14 ± 23.46 | 480.00 ± 22.93 | 457.00 ± 40.61 |
| IV (n = 22) | CC       | 3  | 280.07 ± 2.12 | 143.86 ± 26.40 | 448.14 ± 23.46 | 480.00 ± 22.93 | 457.00 ± 40.61 | 480.00 ± 22.93 |
|           | CT       | 12 | 281.67 ± 4.41 | 134.67 ± 52.84 | 480.00 ± 22.93 | 457.00 ± 40.61 | 480.00 ± 22.93 | 457.00 ± 40.61 |
|           | TT       | 7  | 281.67 ± 4.41 | 134.67 ± 52.84 | 480.00 ± 22.93 | 457.00 ± 40.61 | 480.00 ± 22.93 | 457.00 ± 40.61 |

abc Means bearing a same superscript in a column for one lactation differ non-significantly (P>0.05); BW - Birth weight; AFS - Age at first service; AFC - Age at first calving; GP - Gestation period; DP - Dry period; CI - Calving interval
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Table 4. Association of ADIPOQ/TasI genotypes with milk production traits.

| Lactation | Genotype | n  | LP (days) | TMY (liters) | AMY (liters) | MYPP (liters) | MY300 (liters) | PY (liters/day) | DRPY (days) |
|-----------|----------|----|-----------|-------------|-------------|--------------|---------------|---------------|-------------|
| I (n = 55) | CC       | 12 | 422.50±36.98 | 2049.00±258.64 | 4.83±0.31 | 72.92±9.42 | 1449.33±92.13 | 7.04±0.51 | 41.08±3.40 |
|           | CT       | 36 | 379.22±18.58 | 1918.53±140.74 | 4.92±0.17 | 79.28±4.53 | 1477.39±51.28 | 7.53±0.29 | 42.31±3.08 |
|           | TT       | 7  | 375.43±29.35 | 2024.00±78.08  | 5.50±0.34 | 80.14±7.16 | 1657.00±102.23| 8.14±0.60 | 46.57±5.94 |
| II (n = 40)| CC       | 10 | 368.50±38.55 | 2120.80±217.85 | 5.94±0.58 | 110.05±8.60 | 1784.80±174.16| 8.15±0.53 | 39.00±5.42 |
|           | CT       | 25 | 390.84±15.46 | 2248.60±123.62 | 5.72±0.21 | 109.28±6.52 | 1717.56±63.10 | 9.12±0.45 | 42.92±2.43 |
|           | TT       | 5  | 360.00±44.55 | 2273.00±274.61 | 6.36±0.44 | 124.80±13.85| 1909.80±136.33| 9.90±0.76 | 29.00±2.14 |
| III (n = 24)| CC      | 4  | 380.50±28.66 | 1904.50±59.86  | 4.95±0.35 | 89.00±8.00 | 1483.50±109.10| 8.25±0.43 | 40.75±5.01 |
|           | CT       | 17 | 347.12±23.53 | 2092.94±94.47  | 5.91±0.35 | 104.68±11.79| 1774.53±104.04| 9.56±0.59 | 41.00±3.16 |
|           | TT       | 3  | 359.33±22.48 | 2248.33±42.22  | 6.27±0.22 | 131.33±3.86 | 1885.00±69.18 | 10.50±0.58 | 31.33±6.17 |
| IV (n = 22)| CC       | 3  | 314.80±23.15 | 1526.20±110.73 | 4.88±0.31 | 89.40±8.50 | 1464.80±90.96 | 7.90±0.71 | 60.40±19.48 |
|           | CT       | 12 | 302.86±14.67 | 1826.86±158.92 | 5.92±0.44 | 92.79±9.96 | 1780.36±130.10| 9.36±0.77 | 45.00±4.76 |
|           | TT       | 7  | 433.33±91.84 | 2658.67±201.33| 6.57±1.20 | 128.00±24.58| 1970.67±358.29|10.17±2.03| 38.00±6.66 |

abc Means bearing a same superscript in a column for one lactation differ non-significantly (P>0.05)

cattle and observed three genotypes, viz., major, minor and heterozygous genotypes, with genotypic frequencies of 65.86%, 3.57% and 30.57%, respectively. The allele frequency was 0.81 for the major allele and 0.19 for the minor allele.

Association studies of CC, CT and TT genotypes of the ADIPOQ gene by ADIPOQ/TasI PCR-RFLP assay revealed the significant association of these genotypes with CI, LP, TMY and, DRPY. The CT and TT genotypes had longer CI in the first and fourth lactations, suggesting that the T allele has a significant effect on CI. Likewise, a longer LP was observed for the TT genotype compared to the CC and CT genotypes. In addition, TMY showed higher milk yield in the TT genotype compared to the CC and CT genotype. These results suggest that the T allele of the ADIPOQ/TasI gene may be considered to be a good indicator of milk production in the Sahiwal cattle breed.

In the present study, ADIPOQ/TasI SNPs in the promoter region of the gene revealed a significant association with the reproduction and production traits of Sahiwal cows. The promoter region of the ADIPOQ gene contains several regulatory sequences which control the expression of the ADIPOQ gene, and interact with a large number of cis-acting and trans-acting factors. Therefore, the activity and variations in the ADIPOQ gene promoter region could regulate gene expression, and affect the metabolic pathways of fat and glucose metabolism. Many studies have shown that the ADIPOQ transcript is predominantly
expressed in the adipose tissues of human, rodents and porcine (DAI et al., 2006). It may suggest that bovine ADIPOQ is likely to play an important role in lipid metabolism, such as fat differentiation and deposition in cattle as in other species (SHIN and CHUNG, 2013).

This was the first report to investigate the association of genetic polymorphism in the ADIPOQ gene on the production and reproduction traits of Sahiwal dairy cattle. Most earlier studies conducted of genetic polymorphism in the ADIPOQ gene investigated their association with carcass traits in beef cattle (MORSCI et al., 2006; SHIN and CHUNG, 2013). Thus, due to the paucity of the available literature, the results could not be discussed further.

In conclusion, the SNP identified in the ADIPOQ gene and its association with production and reproduction traits suggests that this gene might serve as a candidate genetic marker for selection of dairy cattle with better milk yield. However, further studies are needed to validate this SNP of the ADIPOQ gene in other breeds and populations of dairy cattle. Moreover, it is likely to play an important role in lipid metabolism, such as fat differentiation and deposition in cattle. Finally, it is concluded that further studies are needed to validate this SNP in other breeds and populations of dairy cattle. Additionally, the results could not be discussed further.

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SAŽETAK

Cilj ovoga rada bio je istražiti polimorfizam pojedinačnog nukleotida (SNP) u promotoru gena za adiponektin (ADIPOQ) i njegovu povezanost s proizvodnim i rasplodnim svojstvima indijskog goveda Sahiwal pasmine. Dio promotora (ADIPOQ) regije gena ADIPOQ je umnožen i prikazan produkтом 977 bp, a PCR-RFLP metodom s restrikcijom enzimom TasI dobivena su tri genotipa. Među njima najčešći je bio CT genotip (62,32 %), zatim CC (24,64 %) i TT (13,04 %). Učestalost C-alela bila je 0,558, a T-alela 0,442. ADIPOQ/TasI genotipovi su pokazali znakovitu povezanost s međutelidbennim intervalom (CI) u prvoj i četvrtoj laktaciji u kojima je CC genotip pokazao znakovito duži interval između teljenja (P<0,05) u usporedbi s TT genotipom, dok je u četvrtoj laktaciji TT genotip pokazao duži interval teljenja u usporedbi s CC genotipom. U četvrtoj laktaciji TT genotip pokazao je znakovito duže trajanje laktacije (LP) (P<0,05) i ukupan prinos mlijeka (TMY) u usporedbi s CC i CT genotipovima. Zaključno, SNP identificiran u promotoru gena ADIPOQ i njegova povezanost s proizvodnim i reproduktivnim svojstvima upućuju na to da ovaj gen može poslužiti kao kandidatni genski biljeg za selekciju mliječnih krava s većim prinosom mlijeka. Potrebna su daljnja istraživanja polimorfizama pojedinačnih nukleotida u drugim regijama ovoga gena kao i u drugih pasmina i populacija.

Ključne riječi: gen ADIPOQ; PCR-RFLP; SNP; reproduktivna svojstva; proizvodna svojstva; govedo