Practical Considerations for Perforator Flap Thinning Procedures Revisited

Theddeus OH Prasetyono¹, Kristaninta Bangun¹, Frank B Buchari², Putri Rezkini¹

¹Division of Plastic Surgery, Department of Surgery, Cipto Mangunkusumo Hospital, University of Indonesia, Jakarta; ²Division of Plastic Surgery, Department of Surgery, Adam Malik Hospital, Faculty of Medicine University of North Sumatra, Medan, Indonesia

Background A thin perforator flap is one of the best methods for covering defects. This study aimed to revisit and further test the rapidly advancing field of flap thinning techniques.

Methods We performed two cadaveric studies to test the known flap thinning methods, and then applied these methods to a clinical series. In the first study, five cadavers were used to observe the anatomical relation of the perforator with the subdermal plexuses and the subcutaneous fat layer by injecting a colored latex solution. The second study was done on four cadavers independently from the first study. Last, a clinical series was performed on 15 patients.

Results The areolar fat lobules of 10 anterolateral thigh perforator (ALT), seven deep inferior epigastric artery perforator (DIEAP), and six thoracodorsal artery perforator (TAP) flaps were dissected to reduce the flap thickness guided by the colored vascular pattern. On average, the ALT, DIEAP, and TAP flaps were reduced to 32.76%±9.76%, 37.01%±9.21%, and 35.42%±9.41%, respectively. In the second study, the areolar fat lobules were directly dissected in six ALT, six TAP, and four MSAP flaps, and an average reduction in flap thickness of 53.41%±5.64%, 52.30%±2.88%, and 47.87%±6.41%, respectively, was found. In the clinical series, 13 out of the 15 cases yielded satisfactory outcomes with an average thickness reduction of 37.91%±7.15%.

Conclusions These multiple studies showed that the deep fat layer could be safely removed to obtain a thin yet viable perforator flap. This evidence suggests that the macroscopic flap thinning technique can achieve thin flaps. Surgeons should consider this technique before embracing the latest technique of supermicrosurgery.

Keywords Microsurgery / Microvessels / Perforator flap / Subcutaneous fat / Surgical flaps

INTRODUCTION

A perforator flap is one of the best modalities for covering defects to reduce donor site morbidity, particularly when compared to conventional microsurgical flaps. However, a thick and bulky skin flap does not give a favorable aesthetic outcome. To achieve a good aesthetic result, the flap should be thin and pliable yet viable [1-4]. This kind of flap is essential, particularly when covering defects in areas composed of thin skin such as the dorsal side of the hand, the head and neck regions [2,5], and the dorsal side of the foot.

Traditionally, secondary debulking procedures have been nec-

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Essary to achieve a thin yet viable flap. These secondary procedures were performed to avoid jeopardizing the flap vascularization, if the thinning procedures were carried out at the time of flap harvesting. However, a one-stage procedure that can achieve an ideal thin and pliable skin flap has been considered more beneficial. Kimura [6] suggested a one-stage flap thinning procedure that completely thinned the flap by defatting it using a microdissection technique under a microscope. This technique is thought to yield a thin yet viable flap using meticulous steps. However, flap thinning procedure has since undergone considerable advances.

The latest technique came later to achieve a thin flap without additional defatting procedures, but relies on a more distal yet smaller perforator vessel as the pedicle. This technique is challenging because it requires perforator-to-perforator vessel anastomosis, which is considered to reduce morbidities related to deep tissue dissection and the loss of major vessels. This advance technique is called the supermicrosurgery approach [7,8]. Using supermicrosurgery, surgeons are now able to harvest a thin flap just above the deeper fat layer that includes the superficial fascia [9] and anastomoses the perforator-to-perforator artery in the recipient site, including the accompanying vein. However, supermicrosurgery is relatively challenging for less experienced microsurgeons.

Therefore, we aimed to combine the flap thinning method with the latter two procedures mentioned above to enrich the repertoire of the flap thinning method. Specifically, our objective was to determine the relationship between perforator vessels, subdermal plexuses, and the subcutaneous fat layer to ease the flap thinning procedure macroscopically. Knowledge of the anatomical relationship of the subcutaneous fat layer with the perforator vessels and subdermal plexuses may allow microsurgeons to dissect the unnecessary fat layer to achieve a thin flap without having to perform an exhaustive and tedious dissection under a microscope.

METHODS

We report on two cadaveric studies and a clinical series. The first cadaveric study was performed to identify the anatomical relationship between the perforator vessels, subdermal plexuses, and the subcutaneous fat layer. The second study was a technical procedure that simulated the clinical surgical setting. It was conducted independently from the first cadaveric study to confirm the results of the first cadaveric study. Last, the subsequent clinical series was performed to test the feasibility of the macroscopic flap thinning procedures in a real clinical setting.

Cadaveric study I

In September 2006, a descriptive operational study was performed in a forensic laboratory on five fresh cadavers who were ≥ 18 years old. These cadavers were unidentifiable at their time of death and might have been victims of accidents, neglected, or suffering from chronic illnesses and/or malnutrition.

The cadavers were placed on a table in the supine position and the anterolateral thigh perforator (ALT), deep inferior epigastric artery perforator (DIEAP), and thoracodorsal artery perforator (TAP) flaps were elevated. Flap thickness was measured, and the morphology of the fat lobules was examined. After the flaps were elevated, we injected a colored latex solution through the vessels with a 3-mL syringe and cannula number 24G or 26G, depending on the vessel diameter. The colored latex solution was injected until the perforator arteries and subdermal plexuses were filled, which was indicated by the color that could be observed at the flap edges (Fig. 1). After 1 hour, the latex solution solidified. Based on the vascular pattern acquired, each of the flaps was thinned by a sharp dissection of the fat layer using scissors, while taking care to preserve the perforator vessel and subdermal plexus. The final thickness of the flaps was measured, and the morphology of the fat lobules was observed.

Cadaveric study II

To confirm the first cadaveric study, the second cadaveric study was performed in February 2007 in a forensic laboratory using four fresh cadavers with the same inclusion criteria as the first study.

**Fig. 1. Flow of the colored latex through the subdermal plexus**

The colored latex flowed from the subdermal plexus in the thoracodorsal artery perforator flap margin, as shown by the blue arrow. A, lamellar fat layer; B, the skin edge of the TAP flap; C, abdominal skin as the flap lies on the abdomen (as shown by the inset).
study. ALT, TAP, and the medial sural artery perforator (MSAP) flaps were elevated on both sides. Flap thickness was measured, and the morphology of the fat lobules was observed. Then, the thinning procedure was performed using scissors, but without the aid of a microscope. The procedure was carefully performed to preserve the superficial fat layer (areolar form) and excise the deeper fat layer (lamellar form), which was approximately 2–5 mm from the skin surface. Fig. 2 shows the different forms of the fat lobules. Additional effort was made to preserve the perforator vessels and subdermal plexus.

We injected a mixture of latex with 100 g of barium sulfate and 5 g of gelatin in 100 mL of 40°C hot water through cannula number 24G or 26G to the perforator artery using a 3-mL syringe until the perforator vessels and subdermal plexus on the flap edges were filled. As in the first study, the solution was left to solidify for one hour. Subsequently, plain X-rays were taken to assess the reliability of flap vascularization according to the distribution of the visible contrast.

Clinical series
A subsequent clinical series of the macroscopic flap thinning procedure were performed following the cadaveric studies. From April 2007 to March 2009, 15 patients who had undergone free-flap surgery were included in our series (Table 1).

RESULTS

Cadaveric study I
In this study, we elevated 23 flaps (9 cm × 15 cm; 10 ALT, 7 DIEAP, and 6 TAP flaps) on five cadavers (Table 2). Three DIEAP and four TAP flaps were not available due to previous trauma on the cadavers’ bodies.

The average thickness and reduction percentage of the ALT, DIEAP, and TAP flaps before and after the thinning procedure are listed in Table 3. On average, the ALT, DIEP, and TAP flaps were reduced to 32.76% ± 9.76%, 37.01% ± 9.21%, and 35.42% ± 9.41%, respectively.

Cadaveric study II
Twenty-one flaps (7 ALT flaps, 8 TAP flaps, and 6 MSAP flaps) were elevated on four cadavers (Table 4). However, when the preserved tissue cuff was less than 2 cm, there were errors in preserving the tissue cuff surrounding the perforator vessels. In the elevation of two MSAP and one ALT flaps, the perforator vessels were cut off. Due to these technical errors, these flaps were not assessed further.

The average thickness and reduction percentage of the ALT, TAP, and MSAP flaps before and after the thinning procedure are listed in Table 5. After the thinning procedure, which preserved 2 cm of the tissues surrounding the perforator vessels, the average thickness of the ALT, TAP, and MSAP flaps became 53.41% ± 5.64%, 52.30% ± 2.88%, and 47.87% ± 6.41%, respectively, from their origin (Table 5).

Among the six successfully elevated ALT flaps, radiological examinations were only performed on four flaps. Two of the flaps’ perforator vessels were very small (diameter < 0.5 mm); thus,
we could not insert the cannula.

From the six successfully elevated TAP flaps, only four were examined radiologically. The radiograph images showed a sufficient contrast distribution at the flap edges. Two flaps had perforator vessels that were < 0.5 mm in diameter.

All of the successfully elevated MSAP flaps had a contrast distribution at the flap edges (Fig. 3).

Clinical series
The 15 clinical series were performed at various locations, but mostly in the foot area. Table 1 presents the complete profile of these clinical cases. The average percentage of the post-thinning thickness was 37.91% ± 7.15% (range, 22.9%–52.94%). In addition, the time needed to thin the flap ranged from 20 to 35 minutes. The results of the thinning procedure are illustrated in Fig. 4, which shows the division of the two fat layers.

Thirteen out of fifteen cases yielded a satisfactory outcome as represented by the case shown in Fig. 5. One 7 cm × 5 cm ALT flap ended up being bulky, while one MSAP flap was lost due to venous thrombosis (Table 1).

DISCUSSION
The cadaveric studies were performed to determine the relationship between the perforator vessels, subdermal plexus, and the subcutaneous fat layers to facilitate the thinning of the flap. Although others have previously discussed this issue [6,7], the relationship among these structures has not been practically elucidated to provide a clear macroscopic cut-off point that can serve as a guide for the macroscopic thinning procedure of perforator flaps. Although Kimura et al. [10] also reported on the same division between the two fat layers, their flap thinning procedure was performed under a microscope.

We elevated the 9 cm × 15 cm flaps in the cadaveric studies, assuming that these flaps could be supported with one perforator. Both studies showed that the diameter of the perforator vessels varied from < 0.5 to 1.0 mm. These observations were similar to those of Koshima et al. [2] who reported that the mean diameter of the perforator vessels ranges between 0.5 mm and 0.8 mm. Hamdi et al. [3] stated that each perforator could support a DIEAP skin flap that is up to 33 cm × 13 cm or a TAP flap that

| No | Age (yr)/Sex | Problem | Perforator flap | Flap dimension (cm²) | Pre-thinning thickness (mm) | Post-thinning thickness (mm) | Post-reduction percentage (%) | Length of thinning procedure (min) | Outcome |
|----|--------------|---------|----------------|----------------------|-----------------------------|-----------------------------|-------------------------------|---------------------------------|---------|
| 1  | 25/M         | Right foot dorsum trauma defect | ALTpf | 18 × 12 | 15 | 5.2 | 34.67 | 25 | Flap survives; good aesthetic contour |
| 2  | 41/M         | Left traumatic hand defect | ALTpf | 12 × 8 | 18 | 6.7 | 37.22 | 30 | Flap survives; good aesthetic contour |
| 3  | 16/F         | Hand dorsum defect post injection induced necrosis | MSAPpf | 15 × 8.5 | 20 | 8 | 40 | 35 | Flap loss |
| 4  | 21/M         | Right foot dorsum skin avulsion | ALTpf | 12.5 × 7 | 29 | 10 | 34.48 | 35 | Flap survives; good aesthetic contour |
| 5  | 24/M         | Right foot dorsum skin avulsion | ALTpf | 14 × 8 | 15 | 5 | 33.33 | 25 | Flap survives; good aesthetic contour |
| 6  | 18/M         | Right foot dorsum skin avulsion | ALTpf | 9 × 8 | 24 | 8 | 33.33 | 25 | Flap survives; good aesthetic contour |
| 7  | 47/F         | Left diabetic foot defect | ALTpf | 18 × 8 | 16 | 5 | 31.25 | 35 | Flap survives; good aesthetic contour |
| 8  | 30/M         | Right forearm electric burn defect | ALTpf | 7 × 5 | 12 | 5 | 41.67 | 20 | Flap survives; bulky |
| 9  | 27/F         | Neck contracture | ALTpf | 13 × 15.5 | 24 | 5.5 | 22.92 | 25 | Flap survives; good aesthetic contour |
| 10 | 42/F         | Left diabetic foot defect | ALTpf | 24 × 8 | 13 | 5.5 | 42.31 | 30 | Flap survives; good aesthetic contour |
| 11 | 27/M         | Right burn hand contracture and defect | ALTpf | 13 × 9 | 23 | 11 | 47.83 | 20 | Flap survives; good aesthetic contour |
| 12 | 22/M         | Right forearm electric burn defect | ALTpf | 17 × 7 | 17 | 9 | 52.94 | 20 | Flap survives; good aesthetic contour |
| 13 | 44/M         | Diabetic foot ulcer | ALTpf | 16 × 12 | 33 | 12 | 36.36 | 25 | Flap survives; good aesthetic contour |
| 14 | 32/F         | Neck contracture | ALTpf | 18 × 8 | 26 | 10 | 38.46 | 25 | Flap survives; good aesthetic contour |
| 15 | 27/M         | Right 1st web contracture | ALTpf | 14 × 6 | 31 | 13 | 41.94 | 20 | Flap survives; good aesthetic contour |

M, male; F, female; ALT, anterolateral thigh; MSAP, medial sural artery perforator.
is up to 30 cm × 11 cm. Gedebou et al. [4] reported that one perforator can support a skin flap having the dimensions of 18 cm × 28 cm. Wolff et al. [5] concluded that in the case of the ALT flap, one perforator can support a skin flap measuring 12 cm × 30 cm, while Chen et al. [1] reported these dimensions as 15 cm × 35 cm. Alkureishi et al. [11] recommended that one perforator can support an ALT skin flap measuring 17 cm × 14 cm. All of this evidence supports the justification of the flap dimensions used in the present cadaveric studies.

Our observations revealed two morphologically different fat layers that were confined by the superficial fascia. The deeper layer had a lamellar form, and the superficial layer had an areolar form. This result is consistent with that reported by Gasperoni and Gasperoni [12]. The subdermal plexus was located superficially from the areolar fat layer. Based on this, Kimura and Satoh [13] performed fat layer thinning with the aid of a microscope, while preserving the areolar fat layer to avoid injuring the subdermal plexus. Gedebou et al. [4] and Wolff et al. [5] preserved a 1–2-cm fat layer around the perforator pedicle, while Alkureishi et al. [11] and Ross et al. [14] recommended preserving

| Cadaver | I | II | III | IV | V |
|---------|---|----|-----|----|---|
| ALT flap | L | R | L | R | L | R |
| No. of perforator | 2 | 3 | 4 | 3 | 2 | 3 |
| Perforator type | mc | mc | sc | sc | mc | mc |
| Diameter of perforator (mm) | 1 | 1 | 0.5–1 | 1 | 0.5–1 | 0.5–1 |
| Fat morphology | Deep | Lam | Lam | Lam | Lam | Lam |
| Superficial | Flap thickness (mm) | Post-thinning | 12 | 12 | 13 | 12 | 10 | 8 | 18 | 18 | 21 | 21 |
| TAP flap | Pre-thinning | 4 | 4 | 4 | 5 | 4 | 4 | 5 | 5 | 4 | 4 |
| No. of perforator | 2 | - | 5 | 6 | - | 5 | 4 | 3 | 3 | - |
| Perforator type | mc | - | mc | mc | - | mc | mc | mc | mc | - |
| Diameter of perforator (mm) | 1 | - | 1 | 0.5–1 | - | 0.5–1 | 1 | 0.5–1 | <0.5 | - |
| Fat morphology | Deep | Lam | - | Lam | - | Lam | Lam | Lam | - |
| Superficial | Flap thickness (mm) | Post-thinning | 14 | - | 12 | 12 | - | 8 | 14 | 14 | 8 | - |
| TAP flap | Pre-thinning | 5 | - | 4 | 4 | - | 4 | 4 | 4 | 4 | - |
| No. of perforator | 2 | - | 2 | 2 | - | 2 | 2 | 2 | 2 | - |

L, left; R, right; ALT, anterolateral thigh; mc, musculocutaneous; sc, septocutaneous; Lam, lamellar; Ar, areolar; DIEAP, deep inferior epigastric artery perforator; TAP, thoracodorsal artery perforator.

| Table 3. Average thickness of the elevated flaps in cadaveric study I |
|---------------------------------|-----------|-----------|---------------|------------|
| Pre-thinning thickness (mm)     | Post-thinning thickness (mm) | Post-reduction percentage (%) | Standard deviation |
| ALT flap                        | 14.5      | 4.3       | 32.76         | 9.76       |
| DIEAP flap                      | 11.7      | 4.1       | 37.01         | 9.21       |
| TAP flap                        | 9.0       | 3.0       | 35.42         | 9.41       |

ALT, anterolateral thigh; DIEAP, deep inferior epigastric artery perforator; TAP, thoracodorsal artery perforator.
2-cm of tissue surrounding the perforator pedicle to maintain the artery supply. In the second cadaveric study, we preserved a 2-cm cuff of tissues surrounding the perforator vessels as recommended above [11,14]. This tissue preservation, according to Chen et al. [1], will help to prevent the perforator vessels from becoming twisted.

The mean thicknesses of the elevated flaps listed in Table 3 were slightly different from that reported by Nakayama et al. [15] who measured flap thickness in 31 patients using ultrasonography. The resulting thickness was 7.1 ± 3.4 mm for ALT flaps and 13.7 ± 7.9 mm for the DIEAP flaps [15]. In our first cadaveric study, the flaps were thinned based on the acquired vascular pattern, but without loupe magnification, and thin flaps were achieved with a thickness ranging from 3 to 4.3 mm. This thickness is consistent with the results of previous reports about the safe limit of flap thinning [1,6,13,16-19]. However, in the second cadaveric study (Table 5), we could not produce a flap that was as thin as that produced in our first cadaveric study. The resulting thickness in the second study was relatively thicker because we were concerned about damaging the plexuses in the superficial fat layer. Nevertheless, the approach used in second cadaveric study was more similar to a real clinical setting than the first study was because we performed the thinning procedure without guidance from a contrast distribution in the second

### Table 4. Flaps profile in cadaveric study II

| Cadaver | ALT flap | TAP flap | MSAP flap |
|---------|----------|----------|-----------|
| No. of perforators | 2 | 3 | 1 |
| Perforator type | mc | mc | mc |
| Diameter of perforator (mm) | <0.5 | 0.5-1 | 0.5-1 |
| Flap thickness (mm) | Pre-thinning | 11 | 16 | 8 |
| Post-thinning | 5 | 7 | 14 | 2 |
| Pedicle thinning distance (cm) | 2 | 2 | 2 | 2 |
| Radiograph appearance | + | + | + | + |
| Distribution of contrast to the flap edges | + | + | + | + |

### Table 5. Average thickness of the elevated flaps in cadaveric study II

| Flap | Pre-thinning thickness (mm) | Post-thinning thickness (mm) | Post-reduction percentage (%) | Standard deviation |
|------|-----------------------------|------------------------------|-------------------------------|-------------------|
| ALT flap | 11.17 | 6.00 | 53.41 | 5.64 |
| TAP flap | 14.67 | 7.67 | 52.30 | 2.88 |
| MSAP flap | 8.00 | 3.83 | 47.87 | 6.41 |

ALT, anterolateral thigh; TAP, thoracodorsal artery perforator; MSAP, medial sural artery perforator; Pedicle thinning distance, tissue preservation surrounding the perforator vessels.
study. Nojima et al. [20] performed a similar cadaveric study and found the vascular territories of the ALT flaps in their cadavers to decrease when compared to their clinical cases. They stated that the vascular territories likely decreased because the cadavers were relatively old and not healthy at their time of death [20].

Rees and Taylor [21] and Bergeron et al. [22] suggest mixing lead oxide for injection into the perforator vessels of cadavers. We had difficulties obtaining a lead oxide contrast liquid, so we used a mixture of barium, latex, and gelatin. Although the contrasting appearance was not evenly distributed on the radiograph, it was sufficient for our purpose.

In the clinical series, 20–35 minutes were needed to perform the thinning procedure for the 14 ALT and 1 MSAP flaps ranging from 7 cm × 5 cm to 18 cm × 12 cm in dimension. This time required to perform these procedures is typical for flap thinning procedures. The average percentage of post-reduction thickness was 37.91% ± 7.15%, which is closer to that of the first cadaveric study (32.76% ± 9.76% for the ALT flaps) than that of the second cadaveric study (52.51% ± 5.97% for the ALT flaps).

Most of the flaps in our series yielded favorable outcomes, in which the flap survived with a good aesthetic contour. However, two cases did not have satisfactory results. Although case number 8 had the smallest flap dimensions, it had a bulky appearance. In this case, a bulky flap was unavoidable due to longstanding edema, which might have been caused by a possible decrease in venous and lymphatic back flow. As a result, unresolved edema with inherent fibrosis created a bulky appearance. In a larger series, Hong et al. [23] reported that six of 282 thin perforator flaps harvested at the superficial fascial plane required secondary debulking procedures.

Although no patient in our clinical series was considered overweight or obese, body mass index might influence the thickness of the superficial and deep fat layers; nonetheless, further study is...
needed. Body mass index might influence the thickness of the superficial and deep fat layers; nonetheless, further study is needed. Moreover, we did not investigate the potential confounding effect of body mass index, but was still able to achieve a 30%–50% reduction of flap thickness in this macroscopic thinning procedure.

The flap in case number 3 failed to survive and might have resulted from a complication during the macroscopic thinning procedure. However, many possible causes might have contributed to this failure, such as vein thrombosis. In general, the anatomy of the perforator flap vasculature is applicable to all kinds of perforator flaps; hence, the authors considered the MSAP flap failure to be a coincidence since it can also result in other types of perforator flaps.

We demonstrated that the deeper fat layer could be safely removed without interrupting the vascularization of a perforator flap, if a 2-cm cuff of fat tissue surrounding the perforator vessel is preserved. This evidence may guide surgeons to achieve thin yet viable flaps using the macroscopic flap thinning technique. We believe that this finding adds to the knowledge surrounding the fast revolution of the flap thinning technique. Our clinical series suggests that such a macroscopic flap thinning technique can be performed safely.

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