Visible Light-Driven Reductive Azaarylation of Coumarin-3-carboxylic Acids

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ABSTRACT: In the manuscript, reductive and decarboxylative azaarylation of coumarin-3-carboxylic acids is described. It utilizes the photocatalytic activation of (cyano)azaarenes in the presence of fac-Ir(ppy)$_3$ as a photocatalyst. The methodology is versatile and provides access to biologically relevant 4-substituted-chroman-2-ones. Visible light, photoredox catalyst, base, anhydrous solvent, and inert atmosphere constitute key parameters for the success of the described strategy. The developed methodology involves a wide range of coumarin-3-carboxylic acids as well as (cyano)-azaarenes.

INTRODUCTION

Chroman-2-one, pyridine, and their derivatives constitute privileged structural motifs present in various natural products.$^1$ Representative examples of both groups of compounds are shown in Scheme 1. Although these compounds are abundant in nature, synthetic methods for their preparation are of importance.$^2$ In this context, it is worth noting that pyridine is the second most frequent nitrogen-containing heterocyclic scaffold that is present in 62 U.S. FDA approved drugs displaying a wide range of biological activities.$^3$ Thus, the pyridine skeleton often serves as a "privileged" scaffold in drug design and discovery. Moreover, it is also a versatile building block utilized for the synthesis of chiral ligands applied in asymmetric catalysis.$^7$ As a consequence, a lot of effort has been devoted toward the development of methods for the synthesis of pyridine derivatives.$^7$ Recently, radical-based pyridylation reactions have attracted much attention, providing a flexible approach to pyridine derivatives by the application of photocatalysis. These strategies benefit from good functional group tolerance, procedural simplicity, and mild reaction conditions.$^6$

The addition of free radicals to electron-deficient olefins is known as Giese reaction (Scheme 2).$^7$ Recent advancements in this field arise from the development of photo-mediated methods allowing for the free-radical formation under mild and nontoxic conditions.$^8$ A decarboxylative Michael reaction based on nucleophilic addition to carboxylic-acid-activated olefins followed by a decarboxylation reaction constitutes a powerful synthetic tool.$^9$ Recently, we described the first photocatalytic, doubly decarboxylative Giese reaction applicable to a wide range of carboxylic acids.$^{10}$ Coumarin-3-carboxylic acids I constitute useful acceptors in this reaction, opening access to biologically relevant chroman-2-ones 3.$^{11}$ Given the interesting properties of coumarin and pyridine derivatives, the task of development of synthetic routes leading to hybrid molecules bearing both structural motifs was undertaken. Notably, the synthesis of hybrid molecules containing more than one biologically active unit constitutes an important approach in modern drug design.$^{12}$

Herein, we present our studies on the development of decarboxylative reductive arylation of coumarin-3-carboxylic acids. (Cyano)azaarenes were applied as nucleophiles in the Giese-type transformation. This methodology benefits from mild reaction conditions and a broad scope of substrates.

RESULTS AND DISCUSSION

Initially, reactions between cyanopyridine 2a and coumarin derivatives 1 bearing either no or various activating groups in the 3-position were performed (Table 1, entries 1−4). Experiments were performed in acetonitrile in the presence of 4a as a photocatalyst and triethylamine as a base under irradiation with blue light and an inert atmosphere at room temperature. When simple coumarin 5a was used, no reaction was observed. Therefore, EWG-activated coumarin derivatives 1b−e were tested. Surprisingly, derivatives 5b−d displayed no reactivity under these conditions. To our delight, the incorporation of the carboxylic acid moiety into the structure of coumarin 1a resulted in the formation of the desired product 3aa, indicating the crucial role of the carboxylic-acid-moiety of coumarin and pyridine derivatives, the task of development of synthetic routes leading to hybrid molecules bearing both structural motifs was undertaken. Notably, the synthesis of hybrid molecules containing more than one biologically active unit constitutes an important approach in modern drug design.$^{12}$

Herein, we present our studies on the development of decarboxylative reductive arylation of coumarin-3-carboxylic acids. (Cyano)azaarenes were applied as nucleophiles in the Giese-type transformation. This methodology benefits from mild reaction conditions and a broad scope of substrates.

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Further optimization studies were performed using coumarin-3-carboxylic acid 1a and 4-cyanopyridine 2a as model substrates (Table 1, entries 6−22). In the first part of the optimization studies, the catalytic activity of six different photoredox catalysts was tested (with the irradiation with the light source of suitable wavelength) (Table 1, entries 5−10). When Eosin Y 4b was used, the formation of target product 3aa was not observed (Table 1, entry 6). Catalysts 4a and 4c−f provided the desired reactivity (Table 1, entries 5 and 7−10, respectively) with the best results obtained in the presence of catalysts 4a (Table 1, entry 5). In the course of further studies, the amount of 4-cyanopyridine 2a was tested. It was shown that the reaction with a 3-fold excess of 2a gave the product 3aa with 49% yield (Table 1, entry 11). Further increasing the amount of 4-cyanopyridine 2a did not improve the result. In the next step of optimization studies, the effect of the solvent on the reaction outcome was evaluated (Table 1, entries 11−16). The use of different solvents ensured the product formation; however, the best result was obtained when dimethyl sulfoxide was employed (Table 1, entry 13). During further investigations, the amount of catalyst 4a was studied (Table 1, entries 16−18). It proved possible to be lowered to 3 mol % without a significant change of the result (Table 1, entry 17). Furthermore, the effect of base on the reaction outcome was evaluated (Table 1, entries 19−21). When DABCO was used, product 3aa was not formed (Table 1, entry 20) and the application of DIPEA and N-methyl morpholine resulted in diminished yields (Table 1, entries 19 and 21). In the course of further studies, control experiments were performed (Table 1, entries 24−26). The use of stoichiometric amount of Et₃N yielded the product 3aa with low yield (25%) (Table 1, entry 24). The reaction did not proceed in the absence of photoredox catalysts (Table 1, entry 25). A similar effect was observed when the transformation was attempted in the dark (Table 1, entry 26), thus confirming the crucial effect of photocatalyst and the source of light on the reaction outcome. Notably, the optimized reaction proved readily scalable to a 2 mmol scale and the product 3aa was obtained with a high yield (Table 1, entry 27). Finally, the experiment in the presence of TEMPO was carried out and no reaction was observed, thus confirming the radical nature of the developed reaction (Table 1, entry 28).

With the optimized reaction conditions in hand (Table 1, entry 23), the scope of the developed methodology was evaluated (Schemes 3 and 4). Initially, various coumarin-3-carboxylic acids 1a−m were tested in the reaction (Scheme 3). Acids 1b−f bearing electron-donating groups on the aromatic ring provided products 3ab−af with very good yields. For the coumarin carboxylic acid 1a with a t-butyl substituent at the 6-position of the aromatic ring, the yield was the highest despite
Table 1. Visible Light-Driven Reductive Azaarylation of Coumarin 1a and Its Derivatives 5a–d: Optimization studies

| entry | catalyst | X          | solvent | base | catalyst [mol %] | yield [%] |
|-------|----------|------------|---------|------|------------------|-----------|
| 1     | 4a       | H (5a)     | CH₂CN   | Et₃N | 10               |           |
| 2     | 4a       | CN (5b)    | CH₂CN   | Et₃N | 10               |           |
| 3     | 4a       | CO₂Et (5c) | CH₂CN   | Et₃N | 10               |           |
| 4     | 4a       | C(O)Ph (5d) | CH₂CN | Et₃N | 10               |           |
| 5     | 4a       | CO₂H (1a)  | CH₂CN   | Et₃N | 10               | 30        |
| 6     | 4b       | CO₂H (1a)  | CH₂CN   | Et₃N | 10               |           |
| 7     | 4c       | CO₂H (1a)  | CH₂CN   | Et₃N | 10               | 21        |
| 8     | 4d       | CO₂H (1a)  | CH₂CN   | Et₃N | 10               | 14        |
| 9     | 4e       | CO₂H (1a)  | CH₂CN   | Et₃N | 10               | 24        |
| 10    | 4f       | CO₂H (1a)  | CH₂CN   | Et₃N | 10               | 12        |
| 11    | 4a       | CO₂H (1a)  | CH₂CN   | Et₃N | 10               | 49        |
| 12    | 4a       | CO₂H (1a)  | CH₂Cl₂  | Et₃N | 10               | 27        |
| 13    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 10               | 61        |
| 14    | 4a       | CO₂H (1a)  | DMF     | Et₃N | 10               | 15        |
| 15    | 4a       | CO₂H (1a)  | CH₂OH   | Et₃N | 10               | 26        |
| 16    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 5                | 67        |
| 17    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 3                | 68        |
| 18    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 1                | 47        |
| 19    | 4a       | CO₂H (1a)  | DMSO    | DIPEA | 3              | 42        |
| 20    | 4a       | CO₂H (1a)  | DMSO    | DABCO | 3              |           |
| 21    | 4a       | CO₂H (1a)  | DMSO    | NMM   | 3                | 49        |
| 22    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 3                | 81        |
| 23    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 3                | 93        |
| 24    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 3                | 25        |
| 25    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 3                |           |
| 26    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 3                |           |
| 27    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 3                | 74 (333 mg) |
| 28    | 4a       | CO₂H (1a)  | DMSO    | Et₃N | 3                |           |

All reactions were performed in a 0.1 mmol scale using 1a or 5 (1.0 equiv) and 2a (2.0 equiv) in the presence of the corresponding photoredox catalyst 4 (10 mol %) and the corresponding base (2.5 equiv) in the solvent (1 mL) for 24 h at room temperature. Reaction performed under irradiation with blue light. Reaction performed under irradiation with green light. Reaction performed using 2a (3 equiv). Reaction performed for 48 h. Reaction performed in DMSO (3 mL). Reaction performed using Et₃N (1 equiv). Reaction performed in the dark. Reaction performed at a 2 mmol scale. Reaction performed in the presence of TEMPO (1 equiv).
the presence of a bulky t-butyl substituent. In the course of further studies, it was found that substrates 1 bearing electron-withdrawing groups delivered products 3 in diminished yields. Short reoptimization studies indicated that modification of a previously developed procedure (involving dropwise addition of coumarin carboxylic acids 1g−m in dry DMSO (1 mL) over 2 h to the reaction mixture, see general procedure for details) enabled the improvement of the results. Dropwise addition of coumarin carboxylic acids 1g−m suppressed its decomposition over reaction time. Under these conditions, the reaction using coumarins 1g−k bearing fluorine, bromine, or chlorine atoms at various positions provided the corresponding products 3g−k in moderate to high yields. It is only in the case of coumarin 1k with a chlorine substituent in the 8-position of the aromatic ring that the yield of the reaction dropped to 34%. Similar results were observed for doubly substituted coumarin 1l. In this context, it is worth noting that coumarin 1l was not effective in the previous decarboxylative reactions performed by our group. What is also worth emphasizing is that the reaction with doubly substituted coumarin 1m with two methoxy substituents in the aromatic ring provided the desired product 3am with very good yield.

Subsequently, the scope of the methodology with regard to different (cyano)azaarenes 2a−c was evaluated (Scheme 4). It was demonstrated that the developed protocol worked well for 4- and 2-substituted pyridines 2a and 2b as well as pyrimidine-2-carbonitrile 3c to give target products 3aa−3ca with very good yields. Disappointingly, no product formation was observed when cyanopyridines 2d and 2e were employed under optimized reaction conditions.

The postulated mechanism of the developed methodology begins with the blue light-driven excitation of the photocatalyst 4b (Scheme 5). Then, the electron transfer from the triethylamine to the photocatalyst takes place. Fluorescence quenching and cyclic voltammetry experiments confirmed the lack of quenching in the case of acids 1a as well as cyanopyridine 2a (for details, see the Supporting Information). Subsequently, the reduced Ir-catalyst acts as a reductant of the (cyano)azaarene 2a to give 7. The newly formed radical 7 undergoes the decarboxylative Giese-type reaction with the acceptor 8 to give 9 that undergoes hydrogen atom transfer to give 10. Two separate processes transform 10 into 3aa: (1) rearomatization of the pyridine ring via dehydrocyanation and (2) decarboxylative protonation to afford 3aa as the final product.

**CONCLUSIONS**

In conclusion, we have developed a decarboxylative photocatalytic reductive arylation of coumarin-3-carboxylic acids 1 that represents a unique application of free-carboxylic-acid
activated olefins in radical transformations. The reactions between coumarin-3-carboxylic acids 1a–m and (cyano)-azaarenes 2a–c were realized under photocatalytic activation in the presence of only 3 mol % of fac-Ir(ppy)$_3$. The methodology proved versatile, leading to biologically relevant 4-substituted-chroman-2-ones 3aa–ca in good to high yields under mild reaction conditions.

■ EXPERIMENTAL SECTION

General Information. NMR spectra were acquired on a Bruker Ultra Shield 700 instrument, running at 700 MHz for $^1$H and 176 MHz for $^{13}$C. Chemical shifts ($\delta$) are reported in ppm relative to residual solvent signals (CDCl$_3$: 7.26 ppm for $^1$H NMR, 77.16 ppm for $^{13}$C NMR). Mass spectra were recorded on a Bruker Maxis Impact spectrometer using electrospray (ES$^+$) ionization (referenced to the mass of the charged species). Analytical thin layer chromatography (TLC) was performed using pre-coated aluminum-backed plates (Merck Kieselgel 60 F254) and visualized by ultraviolet irradiation. Unless otherwise noted, analytical-grade solvents and commercially available reagents were used without further purification. For flash chromatography (FC), silica gel (w/ Ca, ~0.1%, 230–400 mesh), green LED (50 W, $\lambda$ = 525 nm), and blue LED (50 W, $\lambda$ = 456 nm) were purchased from commercial supplier Kessil LED Photoreactor Lightning. Fluorescence measurements were performed using a Varian Cary Eclipse spectrophotometer equipped with a thermostatted cell holder. Coumarine-3-carboxylic acids 1b–k were synthesized according to the literature procedure.$^{13}$

6-Methyl-2-oxo-2H-chromene-3-carboxylic Acid (1b). Compound 1b was synthesized according to the literature procedure$^{13}$ as a white solid in 75% yield (153.0 mg). Analytical data were in accordance with the literature.

6-Methoxy-2-oxo-2H-chromene-3-carboxylic Acid (1c). Compound 1c was synthesized according to the literature procedure$^{13}$ as a white solid in 82% yield (180.4 mg). Analytical data were in accordance with the literature.

7-Methoxy-2-oxo-2H-chromene-3-carboxylic Acid (1d). Compound 1d was synthesized according to the literature procedure$^{13}$ as a white solid in 64% yield (140.8 mg). Analytical data were in accordance with the literature.

8-Methoxy-2-oxo-2H-chromene-3-carboxylic Acid (1e). Compound 1e was synthesized according to the literature procedure$^{13}$ as a white solid in 72% yield (158.4 mg). Analytical data were in accordance with the literature.

6-(tert-Butyl)-2-oxo-2H-chromene-3-carboxylic Acid (1f). Compound 1f was synthesized according to the literature procedure$^{13}$ as a white solid in 89% yield (218.9 mg). Analytical data were in accordance with the literature.

6-Fluoro-2-oxo-2H-chromene-3-carboxylic Acid (1g). Compound 1g was synthesized according to the literature procedure$^{13}$ as a white solid in 84% yield (174.7 mg). Analytical data were in accordance with the literature.

7-Bromo-2-oxo-2H-chromene-3-carboxylic Acid (1h). Compound 1h was synthesized according to the literature procedure.$^{13}$

Scheme 4. Visible Light-Driven Reductive Arylation of Coumarin-3-carboxylic Acids 1: Reaction Involving Cyanoheteroaromatic Derivatives 2a–2c

Scheme 5. Visible Light-Driven Reductive Arylation of Coumarin-3-carboxylic Acids 1: Reaction Mechanism
as a yellow solid in 62% yield (1668 mg). Analytical data were in accordance with the literature.

8-Bromo-2-oxo-2H-chromene-3-carboxylic Acid (1i). Compound Ii was synthesized according to the literature procedure\(^\ast\) as a yellow solid in 54% yield (145.3 mg). Analytical data were in accordance with the literature.

6-Chloro-2-oxo-2H-chromene-3-carboxylic Acid (1j). Compound Ij was synthesized according to the literature procedure\(^\ast\) as a yellow solid in 89% yield (199.8 mg). Analytical data were in accordance with the literature.

8-Chloro-2-oxo-2H-chromene-3-carboxylic Acid (1k). Compound Ik was synthesized according to the literature procedure\(^\ast\) as a yellow solid in 72% yield (161.6 mg). Analytical data were in accordance with the literature.

3-Oxo-3H-benzo[f]-chromene-2-carboxylic Acid (1l). Compound Il was synthesized according to the literature procedure\(^\ast\) as a yellow solid in 89% yield (199.8 mg). Analytical data were in accordance with the literature.

5,7-Dimethoxy-2-oxo-2H-chromene-3-carboxylic Acid (3a).

The pure product was isolated by flash chromatography on silica gel (n-hexane/ethyl acetate, 2:1) as a pale yellow oil in 85% yield (21.7 mg).\(^{13}C\) NMR (700 MHz, chloroform-\(d\)) \(\delta 85.7\) (d, \(J = 5.0\) Hz, 2H), \(7.07\) (m, 2H), \(6.89\) (dd, \(J = 8.4, 0.8\) Hz, 1H), \(6.70\) (dd, \(J = 2.5, 1.7\) Hz, 1H), \(6.67\) (dd, \(J = 8.4, 2.6\) Hz, 1H), \(4.27\) (t, \(J = 6.4\) Hz, 1H), \(3.81\) (s, 3H), \(3.09\) (dd, \(J = 15.9, 6.2\) Hz, 1H), \(3.00\) (dd, \(J = 15.9, 6.7\) Hz, 1H).\(^{1}H\) NMR (176 MHz, CDCl\(_3\)) \(\delta 166.7, 160.6, 152.7, 150.5\) (2C), \(150.1, 128.9, 122.8\) (2C), \(115.6, 111.3, 103.8, 55.8, 39.7, 36.6\). HRMS (ESI-TOF) \(m/z\) [M + H\(^{+}\)] calculated for C\(_{14}\)H\(_{14}\)NO\(_{3}\): 256.0968, found: 256.0964.

8-Methoxy-(pyridin-4-yl)chroman-2-one (3a).

The pure product was isolated by flash chromatography on silica gel (n-hexane/ethyl acetate, 2:1) as a pale yellow oil in 85% yield (21.7 mg).\(^{13}C\) NMR (700 MHz, chloroform-\(d\)) \(\delta 85.7\) (d, \(J = 5.0\) Hz, 2H), \(7.12\) (ddd, \(J = 8.3, 1.3\) Hz, 3H), \(6.59\) (dd, \(J = 7.7, 4.0\) Hz, 1H), \(4.32\) (t, \(J = 6.3\) Hz, 1H), \(3.92\) (s, 3H), \(3.14\) (ddd, \(J = 7.5, 6.3\) Hz, 1H).\(^{1}H\) NMR (176 MHz, CDCl\(_3\)) \(\delta 166.7, 160.6, 152.7, 150.5\) (2C), \(128.9, 122.8\) (2C), \(115.6, 111.3, 103.8, 55.8, 39.7, 36.6\). HRMS (ESI-TOF) \(m/z\) [M + H\(^{+}\)] calculated for C\(_{13}\)H\(_{15}\)NO\(_{3}\): 256.0968, found: 256.0971.

6-tert-Butyl-(pyridin-4-yl)chroman-2-one (3af).

The pure product was isolated by flash chromatography on silica gel (n-hexane/ethyl acetate, 2:1) as a pale yellow oil in 85% yield (21.7 mg).\(^{13}C\) NMR (700 MHz, chloroform-\(d\)) \(\delta 85.7\) (d, \(J = 5.0\) Hz, 2H), \(7.12\) (ddd, \(J = 8.3, 1.3\) Hz, 3H), \(6.59\) (dd, \(J = 7.7, 4.0\) Hz, 1H), \(4.32\) (t, \(J = 6.3\) Hz, 1H), \(3.92\) (s, 3H), \(3.14\) (ddd, \(J = 7.5, 6.3\) Hz, 1H).\(^{1}H\) NMR (176 MHz, CDCl\(_3\)) \(\delta 166.7, 160.6, 152.7, 150.5\) (2C), \(128.9, 122.8\) (2C), \(115.6, 111.3, 103.8, 55.8, 39.7, 36.6\). HRMS (ESI-TOF) \(m/z\) [M + H\(^{+}\)] calculated for C\(_{13}\)H\(_{15}\)NO\(_{3}\): 256.0968, found: 256.0971.
3.12 (dd, J = 15.9, 6.1 Hz, 1H), 3.07 (dd, J = 15.9, 6.9 Hz, 1H). 13C (1H) NMR (176 MHz, CDCl3) δ 165.4, 150.9 (2C), 148.9, 148.6, 133.5, 127.4, 125.8, 125.7, 122.6 (2C), 111.5, 40.6, 36.0. HRMS (ESI-TOF) m/z [M + H+] calculated for C14H14NO2Br+: 303.9968, found: 303.9973.

6-Chloro-4-(pyridin-4-yl)chroman-2-one (3aj). The pure product was isolated by flash chromatography on silica gel (n-hexane/ethyl acetate, 2:1) as a pale yellow oil in 96% yield (24.9 mg). 1H NMR (700 MHz, chloroform-d) δ 8.63–8.61 (m, 2H), 7.32 (dd, J = 8.5, 2.5, 0.6 Hz, 1H), 7.11 (dd, J = 8.7 Hz, 1H), 7.09 (dd, J = 4.4, 1.6, 0.6 Hz, 2H), 6.98 (dd, J = 2.5, 0.8 Hz, 1H), 4.31 (t, J = 6.6 Hz, 1H), 3.10 (dd, J = 6.6, 5.5 Hz, 2H), 3.03 (dd, J = 6.8, 7.1 Hz, 1H). 13C (1H) NMR (176 MHz, CDCl3) δ 166.0, 150.9 (2C), 150.4, 148.4, 130.3, 129.7, 128.2, 125.6, 122.6 (2C), 119.0, 40.2, 35.9. HRMS (ESI-TOF) m/z [M + H+] calculated for C14H14NO2: 260.0473, found: 260.0471.

8-Chloro-4-(pyridin-4-yl)chroman-2-one (3ak). The pure product was isolated by flash chromatography on silica gel (n-hexane/ethyl acetate, 2:1) as a pale yellow oil in 34% yield (8.8 mg). 1H NMR (700 MHz, chloroform-d) δ 8.60 (m, J = 5.0 Hz, 2H), 7.42 (d, J = 8.0 Hz, 1H), 7.07 (dd, J = 21.2, 6.4 Hz, 3H), 6.91 (d, J = 7.7 Hz, 1H), 4.36 (t, J = 6.6 Hz, 1H), 3.12 (dd, J = 15.9, 6.1 Hz, 1H), 3.07 (dd, J = 15.9, 6.9 Hz, 1H). 13C (1H) NMR (176 MHz, CDCl3) δ 165.4, 150.8 (2C), 148.7, 147.8, 130.5, 125.5, 122.8, 122.7 (2C), 40.6, 36.0. HRMS (ESI-TOF) m/z [M + H+] calculated for C14H14NO2: 260.0473, found: 260.0475.

1,2-Dihydro-3H-benzof][1-(pyridin-4-yl)chromen-3-ol (3al). The pure product was isolated by flash chromatography on silica gel (n-hexane/ethyl acetate, 2:1) as a pale yellow oil in 41% yield (11.3 mg). 1H NMR (700 MHz, chloroform-d) δ 8.53–8.51 (m, 2H), 7.92 (d, J = 9.1 Hz, 1H), 7.90–7.89 (m, 1H), 7.71 (dd, J = 8.5, 0.9 Hz, 1H), 7.51 (dd, J = 8.5, 6.8, 1.4 Hz, 1H), 7.48 (dd, J = 8.0, 6.8, 1.6 Hz, 1H), 7.37 (d, J = 9.0 Hz, 1H), 7.06 (dd, J = 4.4, 1.6, 0.6 Hz, 2H), 4.94 (d, J = 6.7 Hz, 1H), 3.27 (dd, J = 16.0, 7.3 Hz, 1H), 3.18 (dd, J = 16.0, 1.8 Hz, 1H). 13C (1H) NMR (176 MHz, CDCl3) δ 166.3, 152.8 (2C), 150.1, 149.8, 131.3, 103.0, 130.7, 129.1, 128.0, 125.7, 122.7, 122.3 (2C), 117.7, 116.0, 57.1, 36.6. HRMS (ESI-TOF) m/z [M + H+] calculated for C14H14NO2: 276.1019, found: 276.1023.

5,7-Dimethoxy-4-(pyridin-4-yl)chroman-2-one (3am). The pure product was isolated by flash chromatography on silica gel (n-hexane/ethyl acetate, 2:1) as a pale yellow oil in 82% yield (23.4 mg). 1H NMR (700 MHz, chloroform-d) δ 8.63–8.29 (m, 2H), 7.17–6.90 (m, 2H), 6.31 (d, J = 2.3 Hz, 1H), 6.28 (d, J = 2.3 Hz, 1H), 4.52 (dd, J = 6.8, 2.4 Hz, 1H), 3.80 (s, 3H), 3.75 (s, 3H), 3.08–2.96 (m, 2H). 13C (1H) NMR (176 MHz, CDCl3) δ 166.9, 161.3, 157.5, 153.2, 150.6, 150.3 (2C), 122.2, 104.9, 95.3, 94.3, 56.0, 55.7, 36.0, 34.1. HRMS (ESI-TOF) m/z [M + H+] calculated for C14H14NO2: 286.1074, found: 286.1068.

General Procedure for the Synthesis of 4-(Pyridin-4-yl)chroman-2-one (3aa) in a 2 mmol Scale. In a 50 mL Schlenk tube, coumarin-3-carboxylic acid 1a (380.3 mg, 2.0 mmol, 1.0 equiv), 4-cyanopyridine 2a (624.7 mg, 6.0 mmol, 3.0 equiv), Et3N (506.0 mg, 5.0 mmol, 2.5 equiv), and catalyst fac-Ir(ppy)3 (39.3 mg, 3 mol %) were dissolved in dry DMSO (20 mL). The reaction mixture was degassed and filled three times with argon. Subsequently, the mixture was irradiated with blue LED for 48 h at room temperature. Next, the reaction was quenched with saturated solution of NaHCO3 (50 mL), extracted with CH2Cl2 (3 × 75 mL), and washed with brine (50 mL). The organic phase was dried over MgSO4 and concentrated under reduced pressure. The crude product 3aa was purified by silica gel chromatography (n-hexane/ethyl acetate, 2:1) to provide the desired product 3aa as a yellow oil in 74% yield (333 mg).

### ASSOCIATED CONTENT

**Supporting Information**

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acs.joc.2c00683.

Cyclic voltammetry, fluorescence quenching, photochemical reaction setup, and copies of 1H and 13C NMR spectra (PDF)

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**Author Contributions**

All authors have given approval to the final version of the manuscript.

**Notes**

The authors declare no competing financial interest.

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