The Netrin-1/DCC guidance system: dopamine pathway maturation and psychiatric disorders emerging in adolescence

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Abstract
Axon guidance molecules direct growing axons toward their targets, assembling the intricate wiring of the nervous system. One of these molecules, Netrin-1, and its receptor, DCC (deleted in colorectal cancer), has profound effects, in laboratory animals, on the adolescent expansion of mesocorticolimbic pathways, particularly dopamine. Now, a rapidly growing literature suggests that (1) these same alterations could occur in humans, and (2) genetic variants in Netrin-1 and DCC are associated with depression, schizophrenia, and substance use. Together, these findings provide compelling evidence that Netrin-1 and DCC influence mesocorticolimbic-related psychopathological states that emerge during adolescence.

Objectives
The human brain is interconnected by an estimated 10 million km of neurites [1]. The specific routes taken are choreographed by a surprisingly small number of axon guidance molecules [2]. In this review, we summarize evidence that recently identified mutations and common variants of genes encoding the guidance cue Netrin-1 and its receptor, DCC (deleted in colorectal cancer), affect the adolescent expansion of mesocorticolimbic dopamine pathways and vulnerability to putative mesocorticolimbic-related psychiatric disorders.

Mesocorticolimbic dopamine anatomy and psychiatric disorders
The primate mesocorticolimbic dopamine system shares many features with the homologous pathways in rodents. As in rodents, primate dopamine cells project from the upper brainstem to the dorsal striatum and multiple cortical and subcortical limbic regions [3, 4]. These latter targets include the ventral striatum (nucleus accumbens (NAcc), olfactory tubercle), septum, hippocampus, amygdala, and cortical regions, particularly the prefrontal (PFC), cingulate, and perirhinal cortices. Primates and rodents both have descending glutamatergic and GABAergic projections from the anterior cingulate and orbital frontal cortices to several limbic and midbrain regions, including the ventral striatum and the dopamine cell body regions, the substantia nigra (SN) and ventral tegmental area (VTA) [3, 5]. In both rodents and primates, the density of mesocortical dopamine fibers increases dramatically from adolescence to adulthood [6, 7]. This process, at least in rodents, results from dopamine axons continuing to grow beyond the NAcc to the PFC across adolescence [8]. Compared to rodents, primate cortical dopamine projections are more widespread [9], innervating the entire cortical mantle, albeit more to anterior than posterior regions [10, 11]. Subcortical dopamine axons are often myelinated in primates, a feature not seen in rodents [10].

Disturbances to mesocorticolimbic development have been proposed to contribute to multiple psychiatric disorders. Consistent with this hypothesis, dopamine neurotransmission and mesocorticolimbic functional connectivity...
Fig. 1 a DCC-mediated Netrin-1 connectivity-related processes. Throughout adolescent brain development, target recognition, axon arborization, and synapse formation are ongoing including dopamine axon targeting, long distance axonal growth, and synaptogenesis by mesocorticolimbic dopamine axons. Green gradients indicate Netrin-1 and the depicted axons express DCC. b Dcc Mutation Behavioral Effects in Mice. Ages, in post-natal days (P), and periods (early adolescence, mid adolescence, and adulthood) at which stimulant drug-induced effects on reward and information processing emerge in Dcc haploinsufficient mice. The effects are in green, based on studies of Dcc haploinsufficient mice [6, 7]. c Age of Onset of DCC-implicated Psychiatric Disorders. The DCC-implicated psychiatric disorders, major depressive disorder, schizophrenia, and substance use disorder, begin to emerge in adolescence. The interquartile ranges (25th to 75th percentiles) are indicated in green. The median ages of onset for major depressive disorder, schizophrenia, and substance use disorder are 32, 23, and 20, respectively. Data to construct the figure were obtained from a U.S. survey and an international review [17, 18].

Axon guidance

The axon guidance properties of Netrin-1 and its receptor, DCC, are shared by a larger class of proteins which operate according to the following general mechanisms. During neurodevelopment, axon navigation is directed by extracellular axon guidance cues, which attract or repel growing axons by inducing molecular changes in their growth cones [2]. Growth cones are versatile structures with actin-based finger-like extensions (filopodia) and protruding sheets (lamellipodia) in their peripheral domains and microtubules in their central domains. Guidance cues induce elongation, retraction, or turning of growth cones, by altering the relative rates of polymerization and depolymerization in actin filaments as well as changes in microtubule stabilization of the growth cone’s cytoskeletal proteins, including actin filaments and microtubules [20]. These processes play a critical role in the organization of brain connectivity and are coordinated by a small number of guidance cue families, including the netrins, slits, semaphorins, and ephrins [2, 21]. There are many members of these families as well as splice...
variants [2, 21]. While the present review focusses on Netrin-1 and its receptor, DCC, there are additional Netrin-1 receptors including Neogenin [22], uncoordinated 5 homologues (UNC5-H) [23], and Down syndrome cell adhesion molecule (DSCAM) that contribute to the orchestration of neuronal networks [24]. Furthermore, in addition to guidance cue systems, cell adhesion molecules, such as neural cell adhesion molecules (NCAM) [25] and cadherins [26], as well as growth factors, also play critical roles in brain connectivity processes (i.e., synapse formation [27], axon branching [28], and axon guidance [29]).

Netrin-1 and its DCC receptor

Netrin-1 is a mammalian laminin-related diffusible molecule that interacts attractively or repulsively with several receptors, including DCC [30]. These effects of Netrin-1 play a well-documented role in axonal pathfinding, an evolutionarily conserved process demonstrated in several species, including Drosophila [31], C. elegans [32], and rodents [6]. The role of netrin and its receptors continues beyond the laying down of pathway connectivity. Indeed, once axons reach their intermediate or final targets, Netrin-1 participates in connectivity-related processes, including target recognition, axon branching, synaptogenesis, and synaptic plasticity [11, 30, 32, 33].

DCC receptors are part of the immunoglobulin superfamily and their extracellular domains are composed of four immunoglobulin domains, in addition to six fibronectin type III repeats [34]. The fourth, fifth, and sixth fibronectin type III repeats constitute the binding sites for Netrin-1 [35]. In humans and rodents, the DCC gene is located on chromosome 18 and comprises 29 exons [36]. The first demonstration of DCC’s role in axon guidance was in commissural fibers of the developing neural tube [37], but DCC’s role is not restricted to these fibers or to this early stage. Instead, DCC remains expressed across the lifespan throughout the nervous system [38–40]. DCC and Netrin-1 are highly expressed in dopamine cell bodies and terminal regions, including the SN, VTA, striatum, hippocampus, and cerebral cortex, in both rodents and humans [38–45].

Mice bred for Dcc haploinsufficiency (+/-) have altered adolescent development of mesocorticolimbic dopamine neurons affecting dopamine transmission and dopamine-related behaviors in adulthood [6, 7]. Adult Dcc+/- mice exhibit increased dopamine axon innervation, dopamine presynaptic sites, and amphetamine-induced dopamine release in the PFC. In comparison, in the NAcc, there are decreases in dopamine varicosities and amphetamine-induced dopamine release [8, 42, 45, 46]. The latter effects result from ectopic growth of mesolimbic dopamine axons to the PFC, a comitant increase in mesocortical dopamine synapses [8], function [47], and augmented cortical inhibitory control over the responsiveness of mesolimbic dopamine neurons [48]. These changes are concordant with the findings that, as adults, but not as adolescents, Dcc+/- mice display multiple alterations to dopamine-related behaviors, including diminished sensitivity to the effects of stimulant drugs (cocaine, amphetamine, methamphetamine) on locomotor activity, sensorimotor gating, conditioned place preference, and intracranial self-stimulation [6, 42, 49, 50]. These behavioral effects have been observed primarily under pharmacological challenge conditions but drug-free adult DCC-deficient mice exhibit reduced impulsivity [8] and attend less to a novel object in the presence of a familiar object [51]. These altered drug responses might be specific to stimulants, given that no differences in conditioned place preference responses are observed to either morphine or ethanol between Dcc haploinsufficient mice and controls (Personal Communication; Flores, Keifer, Darq and Nouel).

Behavioral and neurochemical effects of DCC haploinsufficiency are mirrored by Netrin-1 haploinsufficiency. In adulthood, but not in adolescence, Netrin-1 haploinsufficient mice exhibit increased medial PFC (mPFC) dopamine concentrations and reduced sensitivity to the behavioral effects of amphetamine [48]. Finally, adolescent amphetamine administration alters the expression of both DCC in dopamine neurons and Netrin-1 in the NAcc and mPFC[52].

In the following sections, we collate the evidence that similar DCC and Netrin-1 related effects occur in humans, influencing susceptibility to mood disorders, psychosis, and addictions. As described above, specifically during adolescence, DCC and Netrin-1 mediate dopamine axon targeting in rodents. These processes coincide with changing levels of a microRNA which suppresses DCC expression, miR-218 [7]. In comparison, directly testing temporal effects is generally not possible in human genetic association studies, given that the polymorphisms or mutations are present throughout the lifespan. Nonetheless, as depicted in Fig. 1, the observation that the psychiatric disorders associated with DCC and Netrin-1 polymorphisms begin to emerge in adolescence raises the tantalizing possibility that the processes follow a parallel neurodevelopmental pathway.

Human genetic investigations of DCC

Schizophrenia

Several studies have linked DCC polymorphisms with schizophrenia. In a candidate gene study, comprising 556 schizophrenia patients and 208 healthy controls, a SNP (rs2270954) in DCC was found to be nominally associated with schizophrenia. It was postulated that because this SNP
is within the regulatory 3′ untranslated region (UTR), it may alter DCC mRNA stability and consequently levels of DCC protein translation [53]. Given that the 3′UTR region contains microRNA (miRNA) binding sequences, the identified SNP may disrupt miRNA binding, potentially increasing DCC expression by preventing DCC mRNA transcript degradation and/or translation inhibition [54, 55].

A second candidate gene study of 454 patients with schizophrenia and 486 healthy controls reported a nominal association with a DCC SNP (rs2229080) on exon 3 [56]. The authors also found evidence that rs2229080 induces a protein structural change and, according to in silico analyses, alters splicing regulation. Moreover, the authors noted that rs2229080 is a known target of loss of heterozygosity (LOH) and that such LOH is associated with reduced DCC expression [57]. Thus, if the rs2229080 risk allele disrupts the LOH target site (which downregulates DCC), the authors proposed that the risk allele would result in increased DCC expression, changing mesocorticolimbic dopamine development and ultimately contributing to the schizophrenia phenotype [56]. Subsequently, a much larger genome-wide association study (GWAS), applying a false discovery rate (FDR) correction, found that an intronic locus of DCC (rs4632195) is associated with schizophrenia (n = 82,315) [58]. A mechanism regarding predicted expression outcomes remains to be determined for this SNP. Most recently, by applying next-generation sequencing, which sequences the entire genome and can detect rare variants, loci in five genes, including DCC, were shared among three family members exhibiting atypical psychosis [59].

**Depression**

Over the past six years, there has been rapidly accumulating evidence that both genetic variants and other factors that alter DCC expression also affect susceptibility to mood dysregulation and suicide. Two independent studies from our group, in a discovery and replication cohort, have demonstrated that depressed suicide completers exhibit elevated DCC mRNA expression in the PFC, and a corresponding downregulation of the DCC miRNA repressor, miR-218 [54, 60]. Moreover, a genome-wide investigation of differential gene expression in blood, applying a Bayesian approach, identified 165 differentially expressed genes in major depressive disorder, including overexpression of DCC [61]. Further strengthening these findings is a blood-derived methylyome-wide association study (MWAS) of 812 patients with depression and 320 controls, which found associations between methylation sites in DCC and depression [62]. Notably, while there is evidence of general concordance between DNA methylation across blood and brain tissue, there are exceptions [63], and this is a limitation of the study.

GWAS research has identified an intronic DCC SNP (rs4542757) associated with depressive symptoms (n = 3138) [64]. Although this effect did not achieve genome-wide significance and was not identified in a replication sample, the GWAS study was likely underpowered [64]. A larger GWAS study (n = 161,460) identified an association between depressive symptoms and an intronic DCC SNP, rs62100776 [65]. Moreover, using pathway analyses in two independent samples (n = 6455, n = 18,759), FDR-corrected associations between depression and a Netrin-1 signaling pathway were identified, comprising SNPs from multiple genes involved in Netrin-1 signaling, including DCC [66]. Additionally, gene-based tests found that depressive symptoms among participants in the UK Biobank (n = 99,057) were associated with six genes, including DCC [67]. Another UK Biobank (n = 122,935) gene-based analysis found associations between suicidality and five genes, including DCC [68]. Finally, a GWAS meta-analysis of 135,458 individuals with major depression and 344,901 controls identified 44 genomic loci significantly associated with depression, including an intronic DCC SNP (rs11663393) [69].

The above associations might reflect an effect on mood instability, a clinical feature common to numerous psychiatric disorders [70]. In a GWAS study of 60,443 controls and 53,525 mood instability cases, genome-wide significance was detected for four independent genetic loci, including an intronic DCC SNP (rs8084280) [71]. In line with this idea, genetic correlations, which assess the degree of shared heritability between phenotypes, were identified between mood instability and three psychiatric conditions: major depressive disorder, schizophrenia, and anxiety disorder [71].

Recently, a UK Biobank study (n = ~6400) found that the SNPs in the Netrin-1 signaling pathway conferring risk for major depression are associated with altered white matter microstructure in thalamic radiations, namely lower fractional anisotropy and higher mean diffusivity [72].

Strikingly, a UK Biobank meta-GWAS (n = 375,275) identified an association between anhedonia and a locus in DCC, which was the most statistically significant finding [73]. The authors also reported high genetic correlations between anhedonia and depression, as well as a moderate genetic correlation with schizophrenia [73]. Moreover, a higher anhedonia polygenic score predicted reduced brain volumes, including in the NAcc and mPFC, as well as altered white matter integrity in multiple pathways [73].

Furthermore, a genome-wide methylation study of 150 pairs of monozygotic twins (one co-twin with, and one without, early onset major depression), identified altered methylation in Netrin-1, among other genes in depression [74]. An additional meta-GWAS study reported that a Netrin-1 SNP, rs8081460, was associated with neuroticism...
DCC haploinsufficiency: personality traits and drug-related behaviors

While GWAS studies typically detect relatively subtle effects of DCC polymorphisms [77], loss-of-function haploinsufficient DCC mutation carriers were expected to exhibit larger effects, detectable with smaller sample sizes. Our group recently conducted neuroimaging and psychological studies of a large Quebec family (n = 36), half of whom possess a heterozygous frameshift mutation to DCC (NM_005215.3, c.1140 + 1 G > A). The resulting mutated allele encodes a truncated DCC protein that fails to bind to Netrin-1 [78]. As in Dcc haploinsufficient mice, robust anatomical and behavioral phenotypes are observed as a consequence of human DCC haploinsufficiency, underscoring the sensitivity of the system. The DCC haploinsufficient Quebecers have an adult behavioral phenotype that shares two striking features with adult Dcc haploinsufficient mice [79]. First, the adult DCC haploinsufficient humans exhibit reduced novelty seeking personality traits [79]. Second, compared with their unaffected relatives, the DCC haploinsufficient humans smoke less tobacco yet use similar amounts of alcohol and cannabis [79], consistent with the evidence in mice that DCC’s effects are specific to stimulant drugs. Notably, cigarette smoking increases dopamine transmission in humans [80] while lowered dopaminergic tone can decrease smoking [81], indicating that an altered smoking phenotype could reflect alterations to the dopaminergic system.

These findings are bolstered by a large meta-GWAS (n = 518,633), which identified associations between an intronic DCC SNP, rs1221976, and self-reported “ever smoker” [82]. This finding was part of a larger study on risk-tolerance, whereby DCC SNPs were also associated with “adventurousness”, defined as the propensity to be “adventurous versus cautious.” An additional UK Biobank GWAS study (n ~ 458,000) identified an intronic DCC SNP, rs12970816, associated with cigarette smoking status [83]. Finally, one more study (discovery: n = 5339, replication: n = 1682) reported that the intronic DCC SNP, rs1372626, while not genome-wide significant, was the SNP most strongly associated with cigarette smoking, and was plausibly underpowered [84].

Moreover, in a human multivariate investigation, using a powerful and sensitive alternative to traditional SNP studies, DCC was among the top genes associated with impulsivity (n = 426) [85]. Since diminished novelty seeking is associated with reduced dopamine release in the ventral striatum of humans and rodents [14, 86, 87] these behavioral alterations might reflect DCC’s effects on mesocorticolimbic dopamine development and striatal dopamine transmission [42].

DCC haploinsufficiency: dopamine mesocorticolimbic connectivity

The associations between DCC and psychiatric disorders might be a consequence of DCC-related alterations to mesocorticolimbic pathways. As predicted, our group revealed that DCC haploinsufficient members of the Quebec family, as compared with control groups without the mutation (i.e., both relatives and unrelated healthy volunteers) exhibit striking reductions in anatomical connectivity, assessed using diffusion MRI probabilistic tractography, from the SN/VTA to both the ventral striatum and ventral mPFC [79].

These effects might include changes to dopamine pathways, but some caution is warranted. First, the reduced mesocortical connectivity differs from the increased cortical dopamine innervation seen in adult Dcc haploinsufficient mice. Second, MRI methodologies do not discern the underlying neurochemistry. Indeed, since mesocorticolimbic pathways contain dopamine, gamma-aminobutyric acid (GABA) and glutamate axons [88], the anatomical connectivity findings in humans plausibly represent alterations to both dopaminergic and non-dopaminergic axons. These same considerations also raise the possibility that the dopamine focused studies in rodents have yet to identify alterations to inter-connected non-dopamine neurons.

In rodents, there is a complementary receptor to ligand expression pattern of DCC and Netrin-1 in the NAcc and PFC. While dopamine axons express high DCC levels in the NAcc, they only rarely express DCC in the PFC [45]. Conversely, the intensity of Netrin-1 expression in the NAcc is low, especially compared to the PFC, where Netrin-1 expression is substantial [45]. Indeed, dopamine axons expressing high levels of DCC target the NAcc and do not continue to grow to the PFC in adolescence [8].}

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the case of Dcc haploinsufficiency, since DCC expression is reduced, mesolimbic dopamine axons fail to recognize the NAcc as their final target and instead continue to grow ectopically into the PFC throughout adolescence [8]. In human DCC mutation carriers with reduced mesocortical anatomical connectivity, these same effects might be occurring yet the larger distances to be covered might lead the misrouted mesocortical axons to disperse more diffusely, compared to the rodents.

**Striatal brain volume**

We recently reported that both DCC haploinsufficient humans and mice exhibit reduced striatal volumes. While these effects occur in the NAcc in mice, they are localized to the putamen in humans [79]. These effects were also identified in large-scale GWAS investigations. One of these GWAS studies (n = 30,717), part of the Enhancing Neuro Imaging Genetics through Meta-Analysis (ENIGMA) initiative, investigated genetic variants associated with the volumes of subcortical structures [89]. Putamen volume was associated with four genetic loci, including an intronic DCC SNP (rs62097986), in both discovery and replication cohorts [89].

Bilateral putamen volume has also been associated with an intronic SNP in DCC (rs62098013), as identified in the UK Biobank Brain Imaging Data browser (http://big.stats.ox.ac.uk/), which comprises neuroimaging GWAS data from 9,707 participants [90]. Confidence in this finding is bolstered by a more recent GWAS study that reported an additional intronic locus of DCC (rs4632195) associated with both putamen volume (n = 11,598) and schizophrenia (n = 82,315) [58]. The risk allele for schizophrenia is associated with larger putamen volumes.

Earlier work also identified larger putamen volumes among those with schizophrenia [91], consistent with the two genetic investigations that found associations between SNPs in DCC and schizophrenia [53, 56]. This observation fits well with our earlier proposal that schizophrenia is associated with increased DCC expression [53]. Moreover, in discovery (n = 905) and replication (n = 166) cohorts, variants in genes incurring risk for schizophrenia, including DCC, were associated with alterations in gray matter volumes (putamen, thalamus, temporal gyrus), resting state functional magnetic resonance imaging (rs-fMRI) signals in the mPFC, and working-memory performance [92].

These striatal volumetric effects might have implications for mood disorders as well. In depression, there are reports of decreased putamen volume [93–95], although some other groups have failed to replicate this finding [96] potentially reflecting small sample sizes, heterogeneity within the diagnostic category, and medication effects. Indeed, there is recent MRI evidence that, in psychotropic medication-naïve participants (n = 625), elevated putamen gray matter volume is a disease risk marker across multiple diagnostic categories, namely schizophrenia, major depression, obsessive compulsive disorder, and post-traumatic stress disorder [97].

**Cortical volume**

The DCC haploinsufficient Quebecers also demonstrate modest volumetric increases in two cortical regions: the mPFC/anterior cingulate cortex and the ventral mPFC [79]. In comparison, among human carriers of another DCC mutation, a completely different phenotype is observed, such that there is a complete absence of the cingulate gyrus [98]. Volumetric cortical changes are not observed in Dcc haploinsufficient mice [79] and have not been identified in human neuroimaging GWAS investigations, to the authors’ knowledge, and therefore, the cortical findings may have limited generalizability.

![Fig. 2 Single nucleotide polymorphisms (SNPs) and associated phenotypes in the DCC gene. Depicted here is the 5’−3’ oriented DCC gene, comprising 29 exons (red), intervening introns (gray), untranslated regions (UTR; teal), transcriptional start site (TSS) region, promoter region, start codon (ATG) and stop codon. The rs ID for each SNP and associated phenotype(s) are indicated. The phenotypes are cross disorder (yellow), smoking (light green), depression (blue), schizophrenia (dark green), mood instability (orange), and putamen volume (purple). The gene structure and SNP locations were determined using the NCBI tool, Variation Viewer (https://www.ncbi.nlm.nih.gov/variation/view/), using the genome assembly, GRCh38.p12](image-url)
Identification of different DCC SNPs

The effects reported here are related to multiple SNPs (Fig. 2). This could reflect several factors, including linkage disequilibrium, the set of SNPs examined, the genetic compositions of samples, the statistical procedures, the sample sizes (i.e., statistical power), and the differing use of covariates. Of note, 85% of the discussed DCC SNPs are intronic, indicating that they plausibly influence splicing, and consequently, DCC mRNA transcription and translation; [99] such intronic SNPs may affect enhancers or repressors which may distally regulate DCC transcription [100].

Conclusions

This review highlights accumulating evidence that Netrin-1 and its receptor DCC contribute to mesocorticolimbic dopamine related psychiatric disorders that emerge during adolescence (Tables 1 and 2). The findings are strikingly convergent across heterogenous methodologies and

Table 1 Summary of human genetic studies of DCC and Netrin-1

| References          | Genetic approach                          | Gene          | Sample size          | Ancestry                  | Phenotype                                  |
|---------------------|-------------------------------------------|---------------|----------------------|----------------------------|-------------------------------------------|
| Mood disorders      |                                           | DCC mRNA      | 30 cases, 35 controls| Not available             | Depressed suicide completers             |
| Dunn et al. [64]    | Genome-wide association study (GWAS)      | DCC gene      | 3158                 | Hispanic                  | Depression                                |
| Ohkay et al. [65]   | GWAS                                      | DCC gene      | 161,460              | European                  | Depression                                |
| Smith et al. [75]   | Meta-GWAS                                 | Netrin-1      | 91,370, replication  | White, United Kingdom     | Neurocystic                               |
| Torres-Berrío et al. [54] | mRNA expression                      | DCC mRNA      | 11 cases, 12 controls| Not available             | Depressed suicide completers             |
| Zeng et al. [66]    | Pathway analysis, multilevel regional     | Neurin-1      | 25,214               | European                  | Depression                                |
| Aberg et al. [62]   | Methylome-wide association study (MWAS)   | DCC methylation sites | 812 cases, 320 controls | European | Depression |
| Leday et al. [61]   | Geno-wide gene expression                 | DCC mRNA      | 207 cases, 157 controls | Caucasian | Depression |
| Roberson-Nay et al. [74] | Genome-wide methylation study               | Neurin-1      | 150 monozygotic twin pairs | Caucasian | Depression |
| Wray et al. [69]    | GWAS                                      | DCC gene      | 135,458 cases, 344,901 controls | European | Depression |
| Amna-Soler et al. [67] | Gene-based test                         | DCC gene      | 99,057               | White, United Kingdom     | Depression                                |
| Barbu et al. [72]   | Polycigen score                           | Neurin-1      | ~6400                | Not available             | Depression                                |
| Lee et al. [76]     | Meta-GWAS                                 | DCC gene      | 232,964 cases, 494,162 controls | European | Cross-disorder |
| Strawbridge et al. [69] | Gene-based test                       | DCC gene      | 122,935              | White, United Kingdom     | Suicidality                                |
| Ward et al. [73]    | Meta-GWAS; polygenic score, genetic       | DCC gene      | 375,275              | European                  | Anhedonia                                 |
| Personality traits and substance use |                                           | DCC gene      | 406                   | Caucasian, African-American, Hispanic, other | Impulsivity                               |
| Khaleda et al. [85] | Parallel independent component analysis   | DCC gene      | 5539, replication: 1662 | African-American | Cigarette smoking |
| Ward et al. [71]    | GWAS                                      | DCC gene      | 53,525 cases, 60,443 controls | White, United Kingdom     | Mood Instability                           |
| Vosberg et al. [79] | Rare mutation cohort                      | DCC gene      | 20 cases, 36 controls | Caucasian | Novelty seeking & tobacco use |
| Khachet et al. [83] | GWAS                                      | DCC gene      | ~458,000             | European                  | Cigarette smoking                        |
| Lee et al. [76]     | Meta-GWAS                                 | DCC gene      | 232,964 cases, 494,162 controls | European | Cross-disorder |
| Limnër et al. [82]  | Meta-GWAS                                 | DCC gene      | ~518,633             | European                  | Cigarette smoking                        |
| Grant et al. [53]   | Candidate gene                            | DCC gene      | 556 cases, 208 controls | African American, Asian, Caucasian | Schizophrenia |
| Yan et al. [56]     | Candidate gene                            | DCC gene      | 454 cases, 486 controls | Han Chinese | Schizophrenia |
| Okayama et al. [59] | Next-generation sequencing                | DCC gene      | 3 cases              | Japanese                  | Atypical psychosis                        |
| Smeland et al. [58] | GWAS                                      | DCC gene      | 82,316               | European, East Asian      | Schizophrenia                             |
| Lee et al. [76]     | Meta-GWAS                                 | DCC gene      | 232,964 cases, 494,162 controls | European | Cross-disorder |
| Neurobiology        |                                           | DCC gene      | 30,717               | European                  | Putamen Volume                            |
| Hibar et al. [89]   | GWAS                                      | DCC gene      | 9707                 | White, United Kingdom     | Putamen Volume                            |
| Elliot et al. [90]  | GWAS                                      | DCC gene      | 9,659                | Han Chinese               | Putamen, thalamic, temporal gyrus volumes; mPFC rs-fMRI activity, working-memory |
| Luo et al. [92]     | Polygenic risk score                     | Schizophrenia-associated genes including DCC | Discovery (n = 905), replication (n = 166) | Han Chinese | Putamen volume |
| Smeland et al. [58] | GWAS                                      | DCC gene      | 11,598               | European                  | Putamen, thalamic, temporal gyrus volumes; mPFC rs-fMRI activity, working-memory |
| Vosberg et al. [79] | Rare mutation cohort                     | DCC gene      | 20 cases, 36 controls | Caucasian | Canadian (cases) |
| Barba et al. [72]   | Polygenic risk score                     | Neurin-1      | ~6400                | Not available             | Thalamic raditions, white matter integrity |
Table 2 Summary of DCC and Netrin-1 effects across brain regions

| Brain region | Measure | Mice | Humans |
|--------------|---------|------|--------|
| Striatum     | A. Expression | High DCC, low Netrin-1 | Both Netrin-1 and DCC are expressed and their expression decreases across the lifespan |
|              | B. Volume | Reduced in ventral striatum | Reduced in dorsal striatum |
|              | C. Connectivity | Reduced mesolimbic dopamine innervation | Reduced mesolimbic anatomical connectivity |
| Cortex       | A. Expression | Low DCC, high Netrin-1 (mPFC) | Higher DCC than Netrin-1 (prenatally) |
|              | B. Volume | No difference | Reduced |
|              | C. Connectivity | Increased mesocortical dopamine innervation | Reduced mesocortical anatomical connectivity |

(A) DCC and Netrin-1 relative expression levels in wild-type mice [7] and humans [39] (See also: http://development.psychencode.org). Group differences in (B) volumes and (C) connectivity in DCC haploinsufficient mice [7] and humans [79], relative to control groups.

samples and both between and within studies in humans and mice.

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Compliance with ethical standards

Conflict of interest The authors declare that they have no conflict of interest.

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