Immunohistochemical study on the expression of matrix metalloproteinase 2 and high-risk human papilloma virus in the malignant progression of papillomas

Ho-Jin Lee, Jin-Wook Kim
Department of Oral and Maxillofacial Surgery, School of Dentistry, Kyungpook National University, Daegu, Korea

Abstract (J Korean Assoc Oral Maxillofac Surg 2013;39:224-230)

Objectives: Papilloma frequently develops as a benign tumor of the head and neck area, but its potential for malignant transformation has yet to be studied. This study aims to provide basic information for papillomas using the immunohistochemical staining of matrix metalloproteinase 2 (MMP-2) and human papilloma virus (HPV) 16 and 18.

Materials and Methods: To evaluate the malignant transformation of papillomas, the selected tissue samples were serially diagnosed with pre-cancerous papilloma (with epithelial dysplasia, pseudo-epitheliomatous hyperplasia) or malignant lesion (squamous cell carcinoma, SCC) after the first diagnosis (squamous papilloma, inverted papilloma). The selected tissues were stained with an antibody to MMP-2 and HPV 16-E7, HPV 18-L1. A statistical analysis was performed according to each transformation step.

Results: The epithelial layer of papilloma and pre-cancerous papilloma lesions had a similar MMP-2 expression, but that of the malignant lesion had a significantly increased MMP-2 expression. HPV 16 and 18 infection rates were 28.6%, 33.3% and 63.6% in papillomas, pre-cancerous papilloma lesions, and SCC.

Conclusions: A relatively high MMP-2 expression and HPV 16 or 18 infection of papillomas may be associated with early events in the multistep processes of malignant transformation of papillomas.

Key words: Matrix metalloproteinase 2, Papilloma, Immunohistochemistry, Squamous cell carcinoma, Human papilloma virus

[paper submitted 2013. 8. 16 / revised 2013. 8. 19 / accepted 2013. 9. 23]
MMP with infection of HPV are reported rather than the case of no HPV infection; some studies report that HPV infection and over-expression of MMP-2 are related to the malignant transformation of inverted papilloma.

This study sought to identify the expression of MMP-2 and infection of high-risk HPV on malignantly mutated papilloma and such influences.

II. Materials and Methods

Selected tissue samples were serially diagnosed with precancerous papilloma (with epithelial dysplasia, pseudo-epitheliomatous hyperplasia) or malignant lesion (SCC) after the first diagnosis with squamous papilloma or inverted papilloma.

For the control group, lesions diagnosed as papilloma but did not show any recurrence or malignant transformation were used. This study was approved by the Institutional Review Board of Kyungpook National University Hospital (IRB No. KNUH-2012-07-005).

1. Immunohistochemistry

1) MMP-2 staining

Tissue sections were deparaffinized and placed in 3% hydrogen peroxide for 3 minutes to block the endogenous peroxidase activity. The step of immersing in sodium citrate buffer (pH 6.0) and subjecting to microwave irradiation for 10 minutes (110°C) was used. Blocking reagent (UltraTek HRP; ScyTek Laboratories Inc., Logan, UT, USA) was applied for 10 minutes, and primary antibody (mouse monoclonal anti-human gelatinase A, clone A-Gel VC2, 1:400 dilution; Neomarkers, Fremont, CA, USA) was reacted. Secondary antibody with biotin and peroxidase reagent with avidin were involved, and DAB chromogen was developed. Counter staining was done with haematoxylin.

As negative control, samples were reacted with phosphate-buffered saline instead of primary antibody. As positive control, we used uterine cervical cancer tissues.

2) HPV 16 and 18 staining

Tissue sections were deparaffinized and placed in 3% hydrogen peroxide for 3 minutes to block the endogenous peroxidase activity. The step of immersing in sodium citrate buffer (pH 6.0) and subjecting to microwave irradiation for 10 minutes (110°C) was used. Blocking reagent was applied for 10 minutes, and primary antibody (1:50 diluted HPV 16-E7 antibody; Zymed Laboratories Inc., San Francisco, CA, USA) was reacted. Secondary antibody with biotin and peroxidase reagent with avidin were involved, and DAB chromogen was developed. Counter staining was done with haematoxylin.

For HPV 18 staining, 1:20 diluted HPV 18-L1 antibody (Novocastra, Newcastle, UK) was used as primary antibody.

2. Analysis of immunohistochemistry

1) MMP-2

We selected 5 fields for each slide randomly and recorded them using a microscope with digital camera (×100, Olympus-Bx41; Olympus Optical, Tokyo, Japan). The expression level was described as negative, weak, moderate, and strong according to the amount of positive cells.

For the epithelium, the expression level of MMP-2 was considered as follows: (-), negative (no positively stained epithelial cells); (+), weak (mild positive, partially stained in granular or spinous layer); (++), moderate (positively stained in the overall epithelium except stratum corneum), and; (+++), strong (positively stained in the entire epithelium).

For connective tissue (CT): (-/+), weak (positively stained in some fibroblasts); (++), moderate (positively stained in most of the fibroblasts); (+++), strong (positively stained in almost all fibroblasts).

2) HPV 16 and 18

If the epithelial cell layer in lesion shows koilocytosis, and the nucleus in the epithelium is stained brown, infection of HPV 16 or 18 was assumed.

3. Statistical analysis

The different expressions of MMP-2 by location (epithelial and CT) and according to the presence of HPV infection were analyzed. Kruskal-Wallis test was used to analyze statistically significant differences in groups (P<0.05 considered statistically significant).

III. Results

1. Clinical features

A total of 21 patients (male: 15, female: 6) were included in this study. The mean age of the study group was 58.3 years (range, 30-79 years), and 39 tissues were examined (Table 1). At least 35 tissues of benign papilloma were examined as
Table 1. Distribution of age, gender, and progression of lesions

| No. of patient | Sex | Age (yr) | Progression of lesion |
|----------------|-----|----------|-----------------------|
| 1              | F   | 66       | Pa → PC → SCC         |
| 2              | M   | 62       | Pa → PC → SCC         |
| 3              | F   | 61       | Pa → PC → SCC         |
| 4              | F   | 53       | Pa → PC               |
| 5              | F   | 54       | Pa → PC               |
| 6              | M   | 58       | Pa → PC               |
| 7              | M   | 55       | Pa → PC               |
| 8              | M   | 65       | PC → SCC              |
| 9              | M   | 79       | PC → SCC              |
| 10             | M   | 74       | PC → SCC              |
| 11             | F   | 75       | PC → SCC              |
| 12             | M   | 50       | PC → SCC              |
| 13             | M   | 49       | PC → SCC              |
| 14             | M   | 52       | PC                    |
| 15             | M   | 30       | PC                    |
| 16             | F   | 72       | PC                    |
| 17             | M   | 77       | PC                    |
| 18             | M   | 65       | PC                    |
| 19             | M   | 32       | PC                    |
| 20             | M   | 32       | PC                    |
| 21             | M   | 64       | PC                    |
| Total          | M   | 15/F 6   | 58.3±14.4 7 21 11     |

(F: female, M: male, Pa: papilloma, PC: pre-cancer lesion, SCC: squamous cell carcinoma)

*Recurring lesion.

Ho-Jin Lee et al: Immunohistochemical study on the expression of matrix metalloproteinase 2 and high-risk human papilloma virus in the malignant progression of papillomas. J Korean Assoc Oral Maxillofac Surg 2013

Table 2. Distribution of anatomical location of papillomas, pre-cancerous lesions, and SCCs

| Site             | Papilloma | Pre-cancerous lesion | SCC |
|------------------|-----------|----------------------|-----|
| Paranasal sinus  | 1         | 2                    | -   |
| Pharynx          | -         | 2                    | 1   |
| Nasal cavity     | 3         | 6                    | 3   |
| Maxilla          | 1         | 1                    | 2   |
| Buccal mucosa    | 1         | 1                    | 1   |
| Tongue           | -         | 1                    | 1   |
| Larynx           | 1         | 6                    | 2   |
| Mandible         | -         | 1                    | 1   |
| Palate           | -         | 1                    | -   |
| Total            | 7         | 21                   | 11  |

(SCC: squamous cell carcinoma)

Ho-Jin Lee et al: Immunohistochemical study on the expression of matrix metalloproteinase 2 and high-risk human papilloma virus in the malignant progression of papillomas. J Korean Assoc Oral Maxillofac Surg 2013

control group. The mean age of the control group was 48.3 years (range, 8-84 years). The tumor sites of the study group are listed in Table 2.

2. Immunohistochemistry

1) Expression of MMP-2

The expression levels of MMP-2 in papilloma, pre-cancerous papilloma, and SCC are presented in Table 3. The result of immunohistochemical staining is shown in Fig. 1. These results revealed that the expression levels of MMP-2 increased in pre-cancerous papilloma and SCC than normal papilloma. The control group exhibited 6 negative, 22 weak, and 7 moderate cases in the expression levels of MMP-2.

2) Infection of HPV 16 and 18

In the papilloma study group, infection of HPV 16 was found in 1 case, and that of HPV 18 in 1 case. For pre-cancerous papilloma, infection of HPV 16 was identified in 5 cases, and that of HPV 18 in 1 case. Double infection of HPV 16 and 18 was found in 1 case of pre-cancerous papilloma. The infection rate of HPV 16 or 18 was 28.6% in the papilloma group and 33.3% in pre-cancerous papilloma.

Concerning SCC, all cases showed koilocytosis as evidence of HPV infection. In the papilloma study group, 7 cases, and that of HPV 18, in 2 cases. Double infection of HPV 16 and 18 was found in 2 cases. The infection rate of HPV 16 or 18 was 63.6% in the SCC group, which was an increased outcome compared with the pre-stage lesions. In the control group, 16 cases showed infection of HPV, and 4 cases, HPV 18 infection; double infection of HPV 16 and 18 was noted in 1 case. (Table 4)

3. Statistical analysis

For statistical analysis, we assigned scores according to the degree of expression. In the epithelium, the expression of MMP-2 was scored from 0 (for negative) to 3 (for strong). In CT, it was scored from 1 (for weak) to 3 (for strong).
Fig. 1. Immunohistochemical staining of matrix metalloproteinase 2 (MMP-2). (A) shows negative staining of the control group except keratinized layer (×100). (B) illustrates weak immunoreaction to MMP-2 in the papilloma (×100). (C) shows moderate immunoreaction to MMP-2 in the papilloma with dysplasia (×100). (D) exhibits strong immunoreaction to MMP-2 in squamous cell carcinoma, transformed from the papilloma (×100).

Ho-Jin Lee et al.: Immunohistochemical study on the expression of matrix metalloproteinase 2 and high-risk human papilloma virus in the malignant progression of papillomas. J Korean Assoc Oral Maxillofac Surg 2013

Fig. 2. Koilocytosis is noted, with the epithelial layer showing positive immunoreactivity to human papilloma virus 16 (×400).

Ho-Jin Lee et al.: Immunohistochemical study on the expression of matrix metalloproteinase 2 and high-risk human papilloma virus in the malignant progression of papillomas. J Korean Assoc Oral Maxillofac Surg 2013

Fig. 3. Epithelial cells showing koilocytosis reacting positively to human papilloma virus 18 antibody (×400).

Ho-Jin Lee et al.: Immunohistochemical study on the expression of matrix metalloproteinase 2 and high-risk human papilloma virus in the malignant progression of papillomas. J Korean Assoc Oral Maxillofac Surg 2013
lated as compound with tissue inhibitors of metalloproteinases (TIMPSs). MMP is a kind of enzyme that raises the turnover rate of the ECM proteins-related formation of CT and invasion of tumor. Thus, the degeneration of basement membrane is an essential process for the progression of tumor. Type IV collagen is the major composition of the basement membrane. MMP-2 can degenerate the basement membrane by the decomposition of type IV collagen. As a result, it plays an essential role in the invasion of tumor.

In the papilloma, there was no statistical difference in the expression of MMP-2 between the study and control groups. The epithelium of pre-cancerous lesion showed slightly higher expression of MMP rather than papilloma, but the difference was not statistically significant. Neither was there statistical difference in CT.

SCC exhibited higher expression of MMP-2 than pre-cancerous papilloma or papilloma. (Table 5)

The expression of MMP-2 in pre-cancerous papilloma and SCCs increased when there was HPV infection, and that of MMP-2 in pre-cancerous papilloma with HPV infection was significantly higher. (Figs. 4, 5)

IV. Discussion

The carcinogenesis, invasion, and metastasis of tumor cell are the result of molecular biological multi-step processes. Proper sequences are needed for tumor cells to invade and metastasize. First, tumor cells should be separated before they subsequently attach to extracellular matrix (ECM) and invade into surrounding tissues.

Most MMPs are secreted as pro-matrixin and activated by surrounding tissue, plasma proteinase, or bacterial proteinase. In tumor cells, pro-matrixin is activated in a special membrane called invadopodia. The activated MMP is regulated as compound with tissue inhibitors of metalloproteinases (TIMPSs). MMP is a kind of enzyme that raises the turnover rate of the ECM proteins-related formation of CT and invasion of tumor.

Thus, the degeneration of basement membrane is an essential process for the progression of tumor. Type IV collagen is the major composition of the basement membrane.

MMP-2 can degenerate the basement membrane by the decomposition of type IV collagen. As a result, it plays an essential role in the invasion of tumor. Because of such
destructive potential, MMP should be precisely regulated. Therefore, we can figure out the invasive growth of tumor cells by analyzing the expression of MMP-2. Davies et al. reported the enhanced activation of MMP-2 with the presence of tumor for benign or malignant breast tumors. Some studies also reported that the invasion of tumor was related to the activation of MMP-2. A model study, too, reported decreased invasion of tumor and metastasis with the suppression of MMP-2.

Kim et al. analyzed the expression of MMP-2 in irradiation fibroma, leukoplakia, and oral SCC and reported the enhanced expression of MMP-2 in those lesions compared to normal oral mucosa. Moreover, Duffy et al. reported that the activation of MMP-2 and MMP-9 increased in malignant tumor than benign tumor in general. In our study, the expression of MMP-2 in epithelium and CT was not statistically significant but increased with the progression of lesion from papilloma to pre-cancerous lesion. Likewise, the expression of MMP-2 in change of papilloma to SCC and pre-cancerous papilloma to SCC increased in epithelium and CT, and the difference was statistically significant.

MMPs are synthesized by ECM cells surrounding the tumor cells rather than tumor cells, suggesting tumor cell-stromal cell co-operation. Analyzing the expression of MMPs may be essential in predicting the prognosis of tumor, although tumor cells do not produce MMPs themselves. The result of this study showed significant difference of MMP-2 expression in papilloma--control group and papilloma--study group. Papillomas without any pre-cancerous or malignant change showed weak or decreased expression of MMP-2; in contrast, papillomas in the study group showed higher expression of MMP-2. This implies that papillomas in the study group may have higher potential to be invasive lesions.

The infection of HPV is related to several kinds of cancer. After the first study has been reported, i.e., there are relations between HPV infection and benign or malignant tumor in the head and neck area, many studies were conducted. In 1995, International Agency for Research on Cancer (IARC) reported that HPV 16 and 18 caused malignancy in humans. Arndt et al. researched the subtypes of HPV in papilloma and leukoplakia on oral and laryngeal region. They found HPV 6, 11 in most of the papilloma and reported that high-risk HPV 16 is in 22% of papilloma and 36% of pre-cancerous papilloma. Gillison et al. reported that HPV 16 caused head and neck cancer even in patients who have not been exposed to tobacco or alcohol.

About 25% of head and neck cancers were reported to be related to the infection of HPV. Miller and White inspected HPV in head and neck SCC with polymerase chain reaction (PCR) and reported the infection of HPV on 15.8-34.5% of the lesion. In 2001, Miller and Johnstone reported the result of a 15-year research (1982-1997) on the relation of oral SCC with the infection of HPV. According to them, HPV can be a risk factor of SCC, and the presence of HPV indicated 0-100%.

In our study, the infection of HPV 16 and 18 was examined with immunohistochemistry. We found one case each for infection of HPV 16 and 18 in a total of 7 cases. In pre-cancerous papilloma, 7/21 cases showed the infection of HPV 16 and double infection was found in 1 case. In SCC, 7/11 cases showed the infection of HPV 16 and double infection was found in 2 cases. In control group, infection rate of HPV 16 was 25.7% and 28.6% for HPV 18.

Although the expression levels of MMP-2 were not significantly different, pre-cancerous papilloma exhibited somewhat higher levels of MMP-2 than papilloma. The infection rate of HPV 16 and 18 was slightly higher in pre-cancerous papilloma than papilloma, but there were hardly differences. Nevertheless, the expression level of MMP-2 significantly increased as pre-cancerous lesions becoming malignant lesion and infection rate of HPV almost doubled. Concerning the study reporting that the infection of HPV is related to the expression of MMP-2, this result suggests that the infection of high-risk HPV may play an important role in the early stage of malignant transformation. As part of their research on MMP-2 and MMP-9 with regard to malignant transformation, Katori et al. also stated that the early infection of HPV must be related to malignant transformation.

Our study did not include more reliable or exact methods such as PCR analysis in HPV-type screening; only a small number of samples were analyzed. Despite such limitations, this study yielded useful results by tracking the series of mutations from the papilloma to pre-cancerous lesions and malignant lesion. Moreover, the evaluation of the expression of MMP-2 and high-risk HPV screening with immunohistochemistry is easily applicable to clinical practice because it does not require HPV gene screening. Therefore, papillomas with moderate or higher MMP-2 expression and infection of HPV should be considered to be likely to be changed into pre-cancerous or malignant lesions, in which case long-term follow-up is needed.
V. Conclusion

This study showed that the moderate or higher expression of MMP-2 and infection of high-risk HPV in papilloma may indicate potential malignancy. Consequently, in clinical diagnosis, the evaluation of the expression of MMP-2 and high-risk HPV screening can help us predict the malignant transformation of papilloma and improve patients’ prognosis.

References

1. Sapp JP, Eversole LR, Wysocki GP. Contemporary oral and maxillofacial pathology. St. Louis: Mosby; 1997.
2. Underbrink MP, Hoskins SL, Pou AM, Albrecht T. Viral interaction: a possible contributing factor in head and neck cancer progression. Acta Otolaryngol 2008;128:1361-9.
3. Atula S, Auvinen E, Grenman R, Syrjänen S. Human papillomavirus and Epstein-Barr virus in epithelial carcinomas of the head and neck region. Anticancer Res 1997;17:4427-33.
4. Umudum H, Rezanko T, Dag F, Dogruluk T. Human papillomavirus genome detection by in situ hybridization in fine-needle aspirates of metastatic lesions from head and neck squamous cell carcinomas. Cancer 2005;105:171-7.
5. Capone RB, Pai SI, Koch WM, Gillison ML, Danish BN, Westra WH, et al. Detection and quantitation of human papillomavirus (HPV) DNA in the sera of patients with HPV-associated head and neck squamous cell carcinoma. Clin Cancer Res 2000;6:4171-5.
6. Gillison ML, Koch WM, Capone RB, Spafford M, Westra WH, Wu L, et al. Evidence for a causal association between human papillomavirus and a subset of head and neck cancers. J Natl Cancer Inst 2000;92:709-20.
7. Scheckenbach K, Lieven O, Götte K, Bockmühl U, Zott R, Bier H, et al. p53 codon 72 polymorphic variants, loss of allele-specific transcription, and human papilloma virus 16 and/or 18 E6 messenger RNA expression in squamous cell carcinomas of the head and neck. Cancer Epidemiol Biomarkers Prev 2004;13:1805-9.
8. Kähäri VM, Saarialho-Kere U. Matrix metalloproteinases and their inhibitors in tumour growth and invasion. Ann Med 1999;31:34-45.
9. Mook OR, Frederiks WM, Van Noorden CJ. The role of gelatinases in colorectal cancer progression and metastasis. Biochim Biophys Acta 2004;1705:69-89.
10. Roomi MW, Ivanov V, Kalinovsky T, Niedzwiecki A, Rath M. Modulation of human renal cell carcinoma 786-0 MMP-2 and MMP-9 activity by inhibitors and inducers in vitro. Med Oncol 2006;23:245-50.
11. Thomas GT, Lewis MP, Speight PM. Matrix metalloproteinases and oral cancer. Oral Oncol 1999;35:227-33.
12. Kim MK, Lee EH, Kim J, Lee EW, Cha IH. Immunohistochemical study on expression of MMP-2 and MMP-9 in irritation fibroma, oral leukoplakia and oral squamous cell carcinoma. J Korean Oral Maxillofac Surg 2006;32:352-9.
13. da Silva Cardeal LB, Brohem CA, Corrêa TC, Winnischofer SM, Nakano F, Boccardo E, et al. Higher expression and activity of metalloproteinases in human cervical carcinoma cell lines is associated with HPV presence. Biochem Cell Biol 2006;84:713-9.
14. Katori H, Nozawa A, Tsukada M. Increased expression of matrix metalloproteinase-2 and 9 and human papilloma virus infection are associated with malignant transformation of sinonasal inverted papilloma. J Surg Oncol 2006;93:80-5.
15. Chambers AF, Matrisian LM. Changing views of the role of matrix metalloproteinases in metastasis. J Natl Cancer Inst 1997;89:1260-70.
16. Curran S, Murray GI. Matrix metalloproteinases in tumour invasion and metastasis. J Pathol 1999;189:300-8.
17. Davies B, Miles DW, Happerfield LC, Naylor MS, Bobrow LG, Rubens RD, et al. Activity of type IV collagenases in benign and malignant breast disease. Br J Cancer 1993;67:1126-31.
18. Tokuraku M, Sato H, Murakami S, Okada Y, Watanabe Y, Seiki M. Activation of the precursor of gelatinase A/72 kDa type IV collagenase/MMP-2 in lung carcinomas correlates with the expression of membrane-type matrix metalloproteinase (MT-MMP) and with lymph node metastasis. Int J Cancer 1995;64:355-9.
19. Duffy MJ, Maguire TM, Hill A, McDermott E, O’Higgins N. Metalloproteinases: role in breast carcinogenesis, invasion and metastasis. Breast Cancer Res 2000;2:252-7.
20. Jones JL, Walker RA. Control of matrix metalloproteinase activity in cancer. J Pathol 1997;183:377-9.
21. Gissmann L, Diehl V, Schultz-Coulon HJ, zur Hausen H. Molecular cloning and characterization of human papillomavirus DNA derived from a laryngeal papilloma. J Virol 1982;44:393-400.
22. IARC Working Group on the Evaluation of Carcinogenic Risks to Humans. Human papillomaviruses. IARC Monogr Eval Carcinogen Risk Hum 1995;64:1-378.
23. Arndt O, Johannes A, Zeise K, Brock J. High-risk HPV types in oral and laryngeal papilloma and leukoplakia. Laryngorhinootologie 1997;76:142-9.
24. Miller CS, White DK. Human papillomavirus expression in oral mucosa, premalignant conditions, and squamous cell carcinoma: a retrospective review of the literature. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 1996;82:57-68.
25. Miller CS, Johnstone BM. Human papillomavirus as a risk factor for oral squamous cell carcinoma: a meta-analysis, 1982-1997. Oral Surg Oral Med Oral Pathol Oral Radiol Endod 2001;91:622-35.