Effects of Elevated CO2 and Nitrogen Deposition on Ecosystem Carbon Fluxes on the Sanjiang Plain Wetland in Northeast China

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Abstract

Background: Increasing atmospheric CO2 and nitrogen (N) deposition across the globe may affect ecosystem CO2 exchanges and ecosystem carbon cycles. Additionally, it remains unknown how increased N deposition and N addition will alter the effects of elevated CO2 on wetland ecosystem carbon fluxes.

Methodology/Principal Findings: Beginning in 2010, a paired, nested manipulative experimental design was used in a temperate wetland of northeastern China. The primary factor was elevated CO2, accomplished using Open Top Chambers, and N supplied as NH4NO3 was the secondary factor. Gross primary productivity (GPP) was higher than ecosystem respiration (ER), leading to net carbon uptake (measured by net ecosystem CO2 exchange, or NEE) in all four treatments over the growing season. However, their magnitude had interannual variations, which coincided with air temperature in the early growing season, with the soil temperature and with the vegetation cover. Elevated CO2 significantly enhanced GPP and ER but overall reduced NEE because the stimulation caused by the elevated CO2 had a greater impact on ER than on GPP. The addition of N stimulated ecosystem C fluxes in both years and ameliorated the negative impact of elevated CO2 on NEE.

Conclusion/Significance: In this ecosystem, future elevated CO2 may favor carbon sequestration when coupled with increasing nitrogen deposition.

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Introduction

Atmospheric CO2 concentrations are predicted to double by the end of the century [1]. Increasing CO2 concentrations usually stimulate ecosystem C uptake through photosynthesis [2,3]; however, concurrent increase in ecosystem respiration due to increasing root respiration and enhanced priming of soil organic matter may lead to ecosystem carbon losses [4,5]. The difference between photosynthetic and respiratory fluxes is called net ecosystem exchange (NEE) [6]. Thus, NEE depends on the relative changes in C uptake and release, but the extent of variance in C uptake and release in response to elevated CO2 is uncertain.

In addition, N level are expected to vary in response to global N enrichment caused by anthropogenic activities [7]. According to one estimate, by 2050, 200 Tg N yr−1 will be released and then deposited into the earth's surface [8]. In general, N is a limiting factor for plant growth and net primary production. Consequently, it has been conjectured that N deposition could increase carbon dioxide uptake from the atmosphere [7,9]. However, different ecosystem respiration responses to N addition have been reported in previous studies, including increases [10–12], decreases [13,14] and no significant changes [15]. Therefore, it remains unclear whether N deposition leads to net ecosystem C sequestration [16,17].

The progressive nitrogen limitation (PNL) hypothesis [18] suggests that N additions should enhance CO2 effects on plant productivity. Consequently, concurrent changes in atmospheric CO2 concentration and N deposition may potentially bring about complex interactive impacts on ecosystem functioning. There has been much research on the effects of elevated atmospheric CO2 concentrations and N deposition on the growth of individual plants, but relatively little research on the impact of ecosystem C fluxes on natural vegetation at ecosystem scale [4,19–21]. In particular, there has been no detailed study evaluating the interactive effect of elevated CO2 and N addition on ecosystem C fluxes on wetland ecosystems. Wetlands have globally significant stores of soil carbon and sink for CO2 [22], and consequently play an important role in the global carbon cycle.

A field experiment manipulating elevated CO2 and N deposition was conducted, beginning in May 2010, to examine the potential influences of climate change on a temperate wetland in northeast China. The objectives of this study are to address the following three questions: (1) How do elevated CO2 and N
deposition impact GPP, ER and NEE? (2) Whether elevated CO2 and N deposition interactively affect ecosystem carbon flux on the Sanjiang Plain wetland? (3) How do environmental (e.g., soil temperature, soil water content) and biotic (e.g., vegetative cover) factors regulate the responses of the ecosystem carbon flux to elevated CO2 and N deposition?

Materials and Methods

Ethics Statement

All work was undertaken with relevant permissions from the Honghe National Nature Reserve, China for our observational and field studies. The field studies did not involve endangered or protected species. Data will be made available upon request.

Study Site

This study was conducted at the Field Experimental Establishment of Institute of Natural Resources and Ecology—the Honghe National Nature Reserve (47°49’N, 135°30’E) on the Sanjiang Plain, a wetland located in the Heilongjiang Province of Northeastern China. A low floodplain was formed by the Heilongjiang River, the Songhuajiang River and the Wusulijiang River. Sanjiang Plain is approximately 1.04 km2 in area and is N deficient, as indicated by the strong stimulation of gross primary productivity after N addition [23]. Annual N deposition is 7.6 kg ha⁻² in this region [24]. The area has a temperate continental monsoon climate. The mean annual temperature ranges from 1.4 to 2.9°C, with the average maximum of 21–22°C occurring in July and the average minimum −18°C in January. The mean annual precipitation is 500–650 mm and 80% of rainfall occurs in summer. The frost-free period is 120–140 days. The growing season is limited to late April to early October. Meadow soil is the main soil type with 20% of soil organic matter content and 0.5% of total N at a 0–10 cm soil depth. On the Sanjiang Plain wetland, the native vegetation is characterized by the dominance of the perennial species Calamagrostis angustifolia, as well as the presence of Carex appendiculata, Carex lasiocarpa, Carex pseudocomaica, Glyceria spiculosa and Lathyrus quangnerusiaus.

Open Top Chamber Design

Three pairs of Open Top Chambers (OTC), each 1.8 m high with an octagonal ground surface area of 10.78 m², were set up in early May 2010. They were constructed with a 5-m-wide buffer zone between them to prevent cross contamination. Light intensity in each chamber was 95% of that in open space. The precipitation intensity was identical inside and outside of the chambers. We supplied carbon dioxide to the chambers through a pipe with pinholes connected to industrial CO2 tanks outside the chambers. We adjusted the CO2 supply in accordance with wind speed and CO2 concentrations by taking constant measurements with an infrared gas analyzer.

Experimental Design

The experiment used a paired, nested design with elevated CO2 as the primary factor and nitrogen addition as the secondary factor. Three pairs of OTC were used, and within each pair one was randomly assigned to receive the elevated CO2 treatment (700 ppm, E) and the other as the control (370 ppm, C). The CO2 enrichment began in May 2010 and continued each year for the entire growing season (from May to September) during the daytime (6:00 a.m. to 6:00 p.m.). The CO2 supply was shut off during the night. At each elevated CO2 and control OTC chamber, there were two subplots, one control and the other N addition (N). To each N addition subplot, 8 g N m⁻² yr⁻¹ (N, in the form of liquid NH4NO3) were applied once a year in early May, while the controls were sprayed with distilled water only. Thus, there were four treatments, elevated [CO2] and enriched N deposition (EN), elevated [CO2] and ambient N deposition (E), ambient CO2 and enriched N deposition (N) and ambient CO2 and ambient N deposition as a control (C). Each treatment was replicated three times. The experiments lasted two growing seasons.

Climate Data Soil Microclimate

Climate data were obtained from a Hobo meteorological station, which was approximately 250 m distant from the experimental plots. Soil temperature (Tsoil) at a 10 cm depth and soil water content (SWC) at a 0–10 cm soil depth were continuously recorded with an ECH2O dielectric aquameter (EM50/R Decagon Ltd., Pullman, WA, USA) at 2-h intervals from May 2010 to September 2011 (except when the sample plots flooded in August).

Ecosystem-Level Gas Exchange Measurement

Ecosystem gas exchange was measured on 7 sampling dates during the 2010 and 2011 growing season, respectively (May to September, except when the sample plots flooded). Measurements were performed under cloudless conditions from 9:00 a.m. to 11:00 a.m. to avoid the high air temperature and humidity at noon. In May 2010, one square aluminum frame (0.5 m×0.5 m) was inserted 3 cm into the soil of each subplot, approximately 0.5 m away from the edge. The frames provided a base between the soil surface and the CO2 sampling chamber. When ecosystem gas exchange was measured, we turned off the CO2 supply system at least 10 min prior so that the CO2 concentration was consistent in chamber. Ecosystem gas exchange was measured with a transparent chamber (0.3×0.5×1.0 m) attached to an infrared gas analyzer (IRGA; LI-6400, Li-Cor, Lincoln, NE, USA), which was placed on the frame of each subplot. Four small electric fans were running continuously to promote the mix of air within the chamber during the measurement. Twelve consecutive recordings of the CO2 concentration were taken at 10-s intervals during a 120-s period. The rates of CO2 fluxes were calculated from the time-courses of the concentrations to determine net ecosystem exchange (NEE). Using the approach of Steduto [25], we converted the concentration change to flux-per-unit of soil area. After taking the NEE measurements, the chamber was vented, replaced on each frame and covered by an opaque cloth. The ER was then measured. Gross primary productivity (GPP) was then determined by the sum of NEE and ER (GPP = NEE + ER). Positive NEE values refer to net uptake of C by the ecosystem, and negative NEE values represent a net loss of C from the ecosystem.

Green Plant Cover

For each subplot, the percent cover estimate (percentage of ground area within a subplot covered by green vegetation) was taken. These plots were scored once each year in July.

Leaf Gas Exchange Measurement

Leaf net photosynthetic rate in the dominant plant species C. angustifolia was measured with an open gas-exchange system (LI-6400; Li-Cor, Lincoln, NE, USA). In each subplot, three fully expanded leaves of C. angustifolia were selected and measured once each year in July. Leaf gas exchange was measured in the mornings between 9:00 a.m. and 11:00 a.m. on clear days. During the measurement, leaves were illuminated at 1500 μmol m⁻² s⁻¹ using an LED light system.
Statistical Analyses

The seasonal mean values used in this study were calculated from the monthly mean values, which were first averaged using all measurements from the same month. We examined soil micrometeorology (Tsoil and soil water content) and ecosystem C fluxes (NEE, ER, and GPP) for the entire 2010–2011 period and used repeated measures of ANOVAs to test for main effects and interactions, and whether these changed over time (contrasting both the measuring date and year). The between-subject effects were evaluated as elevated CO2, N addition and their interactions, and within-subject-effects were measuring date, year and their interactions with elevated CO2, and N addition. Regression with correction for autocorrelation and stepwise multiple linear analyses were used to examine the relationships of ecosystem C fluxes with soil temperature, soil water content and vegetation cover in the two growing seasons. Statistical analyses were conducted with SPSS software (SPSS Inc, Chicago, IL, USA), and figures were plotted with SigmaPlot 11.0 software.

Results

Microclimate Changes Induced by Elevated CO2 and N Addition Treatments

In comparison with the long-term averages (1981–2010) of mean air temperature (MAT, 17.3°C) during the growing season, both 2010 (18.4°C) and 2011 (17.7°C) had higher MAT. However, during the growing season, 2010 had higher precipitation (495 mm) and 2011 had lower precipitation (364 mm) than the long-term mean precipitation (MP, 459 mm; Fig. 1). Furthermore, mean air temperature was obviously higher in 2010 than that in 2011 in the early growing season—the monthly mean temperature was higher by 0.95°C in May and 3.12°C in June (Fig. 1). Moreover, the average Tsoil of 2010 was significantly higher than that of 2011 (P<0.001, Table 1, 2). N addition significantly increased Tsoil but decreased soil water contents (SWC) (P<0.05). Soil water content was higher in the elevated CO2 plots than in the control plots (P<0.001, Table 2).

Seasonal and Interannual Variations of Ecosystem CO2 Fluxes

The seasonal dynamics of ecosystem C exchange followed a parabolic-like pattern with higher values in summer and lower values in spring and autumn (Fig. 2), which was similar to the air temperature change. The rates of ecosystem CO2 flux (GPP, ER and NEE) were significantly lower in 2011 than in 2010 (Tables 1, 2; P<0.001). In the control plots, seasonal means for NEE, ER and GPP in 2010 were 56%, 79% and 68% greater than in 2011, respectively. The remarkable interannual variabilities in the ecosystem C fluxes might be partly attributed to differences in air temperature, especially in the early growing season.

Effects of Elevated CO2 on Ecosystem C Fluxes

During the two years of the study, elevated CO2 significantly decreased NEE by 15% (P<0.01) measured by using RMA-NOVA. In addition, elevated CO2-induced decline in NEE was significantly lower in the enriched N plots than in ambient N plots. Elevated CO2 even slightly increased NEE in the enriched N plots (P<0.01, Table 1, 2).

Unlike with NEE, elevated CO2 significantly enhanced ER and GPP by 20% and 4%, respectively (both P<0.01, Table 1, 2). Furthermore, there was a significant temporal shift in the ecosystem C fluxes over the entire period (P<0.05 for the CO2×D, Table 2). In addition, there were significant interactions...
between elevated CO2 and the years, which affected GPP and ER (P<0.05 for the CO2×year). An increase in GPP, induced by elevated CO2, was higher in the enriched N plots than in the ambient N plots in both years (Fig. 3 C).

Effects of N Addition on Ecosystem C Fluxes
Nitrogen addition significantly enhanced NEE, ER and GPP by 10%, 12% and 14%, respectively, (all P<0.01, Table 1, 2) during the two year study period. There was a significant temporal change in NEE, ER and GPP (P<0.001 for the N×D, Table 2) during the entire period. N-induced stimulation of seasonal average NEE and GPP was significantly higher in the elevated CO2 plots than that in the ambient CO2 plots in both years (Fig. 3B, D).

Interactive Effects of Elevated CO2 and N Addition on Ecosystem C Fluxes
NEE was significantly affected by interactions between N addition and elevated CO2, but no interactive effects on ER and GPP were detected (Table 2). In addition, there was a significant temporal shift in NEE, ER and GPP by CO2 at ambient versus enriched N (P=0.04, P=0.002 and P=0.052, respectively, for the CO2×N×D).

Vegetation Cover and Leaf -Level Gas Exchange
In agreement with the elevated CO2 and N responses of NEE, elevated CO2 decreased the vegetation cover by 22% in 2010 (P<0.05) and reduced the maximum photosynthetic rate (Pmax) of the dominant species C. angustifolia by 16% in 2010 and 21% in 2011 (P<0.05). However, N addition enhanced vegetation cover by 26% in 2010 and 28% in 2011 (P<0.05) and increased Pmax of C. angustifolia by 12% in 2010 and in 2011 (P<0.05). Moreover, the vegetation cover and Pmax of C. angustifolia were higher in 2010 than in 2011 (Fig. 4A, B).

Controlling Factors of Ecosystem CO2 Fluxes
Exponential relationships between ecosystem C fluxes and soil temperature (Tsoil) were found in both years (Fig. 5). Tsoil explained 85%, 78% and 67% the variations in NEE, ER and GPP, respectively, in 2010. In 2011, soil temperature could explain 72%, 75% and 82% of the changes in NEE, ER and GPP, respectively. However, there was no significant relationship between ecosystem C fluxes and SWC (data not shown).

We found that both NEE (R² = 0.74 and R² = 0.60, P<0.01) and GPP (R² = 0.36, P<0.01 and R² = 0.73, P<0.01) were positively correlated with the plots' vegetation cover. By contrast, there were no significant relationships between ER and vegetation cover in both years (Fig. 6B).

Discussion
Seasonal and Interannual Variability of Ecosystem C Fluxes
The seasonal dynamics of ecosystem carbon fluxes observed in this study were high in mid-summer and low in the early and late growing seasons, which reected air temperature (Fig. 1, 2). In addition, there were remarkable interannual variabilities in the ecosystem C fluxes (Fig. 2), which might be due to differences in air temperature in the early growing season. Specifically, the average air temperatures of May and of June were signiﬁcantly lower in 2011 than in 2010. Consequently, a lower monthly NEE is expected in 2011, a hypothesis that would be consistent with Welp's findings [26], who found that increases in spring air temperatures caused both GPP to increase by 74% and ER by 61% during the early part of the growing season (i.e., April, May, and June) in deciduous forest. The study by Zhou [27] indicated that in a lawn ecosystem, CO2 uptake signiﬁcantly increased because of soil warming treatment. Moreover, in our study, ecosystem C fluxes were signiﬁcantly exponentially related to soil temperature (Fig. 5). Less sensitive impact on ecosystem C fluxes to air temperature than soil temperature may be attributed to the wetland characteristics. In particular, the soil temperature is much lower than air temperature because the vegetation layer and the thick humus layer.

However, we found the C fluxes in 2010 are also higher than in 2011, when comparing fluxes recorded at the same soil temperature (Fig. 5), which implies that the vegetation cover partly accounted for the interannual differences in ecosystem C fluxes (Fig. 6).

In general, ecosystem C fluxes are closely related to SWC in grassland ecosystem [28,29]. However, we found no significant relationship between ecosystem C fluxes and SWC in our freshwater wetland. A lack of SWC impact on ecosystem C fluxes may be due to the wetland soil and plant roots, which are water-saturated during the growing seasons, thus, the SWC fluctuations have slight effects on ecosystem C fluxes. This was consistent with some previous findings [30,31].

Elevated CO2 Effects on Ecosystem CO2 Fluxes
Our study provides evidence that elevated CO2 can decrease NEE in this temperate wetland in northeast China because the changes in ER were more significant than changes in GPP for controlling the sign and magnitude of NEE (GPP and ER enhanced by 4% and 20%, respectively). This finding is consistent with a semi-arid grassland exposure to elevated CO2 [28]. In both years of our study, both CO2 treatments were net CO2 sinks, but exposure to elevated CO2 reduced CO2 sink strength by 15% over the two years. This negative effect of elevated CO2 on the net C uptake was in line with the trend observed in the Mojave Desert.

| Table 2. Results (P value) of repeat-measurement ANOVA on the effects of year (Y), elevated CO2 (CO2), N addition (N), measuring date (D), and their interactions on soil temperature (Tsoil), soil water content (SWC), net ecosystem CO2 exchange (NEE), ecosystem respiration (ER) and gross primary productivity (GPP). |
|---|---|---|---|---|---|
| Effect | Tsoil | SWC | NEE | ER | GPP |
| D | <0.001 | <0.001 | <0.001 | <0.001 | <0.001 |
| D×CO2 | 0.567 | <0.001 | <0.001 | <0.001 | 0.048 |
| D×N | 0.003 | 0.015 | <0.001 | <0.001 | <0.001 |
| D×Y×CO2 | 0.038 | <0.001 | 0.012 | <0.001 | <0.001 |
| D×Y×N | 0.709 | 0.071 | <0.001 | <0.001 | 0.016 |
| D×CO2×N | 0.796 | 0.001 | 0.04 | 0.002 | 0.052 |
| D×Y×CO2×N | 0.178 | 0.05 | 0.032 | <0.001 | 0.145 |
| Y | 0.000 | 0.001 | <0.001 | <0.001 | <0.001 |
| CO2 | 0.655 | <0.001 | 0.003 | <0.001 | 0.007 |
| N | 0.022 | 0.014 | <0.001 | 0.008 | <0.001 |
| N×CO2 | 0.447 | 0.598 | 0.007 | 0.851 | 0.196 |
| Y×CO2 | 0.247 | 0.804 | 0.972 | 0.046 | 0.13 |
| Y×N | 0.592 | 0.514 | 0.244 | 0.078 | 0.079 |
| Y×CO2×N | 0.613 | 0.792 | 0.506 | 0.869 | 0.89 |

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shrub ecosystem, which was exposed to elevated CO2 for eight years, resulting in a CO2 sink that was decreased by 30% [32]. The following two factors should, in theory, have affected the decline of NEE under elevated CO2 in our study.

(1) Leaf photosynthetic capacity and vegetation cover. Elevated CO2 decreased both the vegetation cover that was fumigated for two months and the light-saturated photosynthesis of the leaves of the dominant species (*C. angustifolia*) (Fig. 4), which might be a reason for the decline in NEE. In addition, elevated CO2 decreased the chlorophyll concentration of *C. angustifolia* in cultivation experiments [33], which provided further evidence for the decline of leaf photosynthetic capacity. These results demonstrate that photosynthetic acclimation of leaf-level under elevated CO2 may simultaneously happen at the ecosystem scale.

(2) Ecosystem C fluxes. In our study, despite the enhanced GPP response to elevated CO2 for most of experimental period (except for 23 June and 14 July 2011), elevated CO2 stimulated ER consistently to a greater extent compared to GPP during the experiment (Fig. 2, Table 1), resulting in reduced NEE, relative to control plots. Elevated CO2 led to higher ER and soil respiration (data unpublished), which might be explained by increased carbon inputs, an enhanced
turnover rate in the roots, root hairs and SOM priming [5,34–37]. Additionally, microbial biomass [38] may have also contributed to the higher respiration at elevated CO2 level.

Our findings indicated that increased ER under elevated CO2 caused lower NEE, compared to the control, which implies that increased root growth or activity could account for the much larger effect of elevated CO2 on ER. Although we could not confirm the exact source of the ecosystem respiration in this experiment, these results show that the potential for C sequestration will decreased to some extent in the next several decades at this study site.

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Figure 3. Elevated CO2-induced changes in NEE (A) and GPP (C) in the non-N-addition and N-addition plots, and the N-induced changes in NEE (B) and GPP (D) in the non-elevated CO2 and elevated CO2 plots in 2010 and 2011. See Figure 2 for abbreviations. 
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Figure 4. Responses of the vegetation cover (A) and the maximum photosynthetic rate ($P_{\text{max}}$) of the dominant species *Calamagrostis angustifolia* (B) to elevated CO2 and N addition in July of 2010 and 2011. See Figure 2 for abbreviations. 
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N Addition Effects on Ecosystem CO2 Fluxes

N addition significantly stimulated NEE in the two growing seasons in our ecosystem, largely due to N-induced increases in GPP, more so than those in ER (Table 1). The N-induced increases in canopy C uptake were partly caused by the stimulation of the leaf maximum photosynthetic rate of the dominant species (Fig. 4B), which was identical with another study [39], and by the higher vegetation cover on N addition plots (Fig. 4A). Stimulation in plant productivity provides more substrate for soil and plant respiration, leading to increases in ER under nitrogen fertilization. The positive effects have also been attributed to increased microbial biomass and activity [40] after fertilization. N stimulation of gross primary production on the Sanjiang Plain wetland (Fig. 2) is in accordance with meta-analysis results of positive N responses at global scales [9]. These results suggest that increased N deposition will enhance ecosystem carbon sequestration in the future.

Interactive Effect of Elevated CO2 and N Addition on Ecosystem CO2 Fluxes

Some research has shown that elevated CO2 and N addition interact with each other in stimulating plant growth [18,41]. Moreover, N addition has been reported to alleviate photosynthetic acclimation [42]. Thus, elevated CO2 and N addition might lead to potential interactive effects on ecosystem C fluxes. Oren [43] reported that soil fertility limits carbon sequestration in forest ecosystems with enriched CO2 level. Reich [44] found that a low
availability of N progressively suppresses the positive response of plant biomass to elevated CO2 in perennial grassland. However, Aeschlimann [45] reported that there was a long-term stimulation in the net C assimilation by elevated CO2, but high N supply led to a lower net C input than low N supply in managed grassland.

At our study sites, a significant interactive effect between elevated CO2 and N addition on ecosystem CO2 exchange including NEE (p = 0.007) was found during the experimental periods (Table 2), and their combined effect increased NEE by 13% compared to that in the control plots (Table 1). The increased response of ecosystem CO2 fluxes to N addition under elevated CO2 compared to ambient CO2 (Fig. 3), together with the interactive effects of CO2 treatment and N treatment on NEE, indicate that elevated CO2 triggers physiological responses to N supply at the ecosystem scale.

In addition, there were significant interactions between the sampling date and experimental treatment on ecosystem C fluxes (CO2×D, N×D, and CO2×N×D, Table 2), which indicated that the effect of the experimental treatment on the ecosystem C fluxes altered over time. The mechanisms responsible for the changes in the response of ecosystem C fluxes to elevated CO2 and N addition over time may involve temporal heterogeneity in plant phenology, physiological activity, soil C and N mineralization, and root and soil organic matter decomposition and turnover [44].

We must note that there were low replication levels in our experiment. Moreover, our study was concerned with instantaneous C fluxes rather than cumulative C fluxes, which might prohibit comparison with other relevant studies. Even so, these results imply that a natural, unfertilized ecosystem experienced a decreased C sink than N-rich ecosystem at this site. Moreover, a severe N limitation in natural wetland may inhibit CO2 response to assimilation [46] and, consequently, to the ecosystem C balance. Thus, this study demonstrated that the net ecosystem CO2 uptake on the Sanjiang Plain wetland depends on adequate N supply.

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Author Contributions

Conceived and designed the experiments; JBW HWN TCZ. Performed the experiments; JBW XLF HXZ JFW. Analyzed the data; JBW. Contributed reagents/materials/analysis tools; JBW HWN. Wrote the paper: JBW HWN TCZ.
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