Molecular Characterization of *Streptococcus agalactiae* Isolated from Bovine Mastitis in Eastern China

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**Abstract**

One hundred and two *Streptococcus agalactiae* (group B streptococcus [GBS]) isolates were collected from dairy cattle with subclinical mastitis in Eastern China during 2011. Clonal groups were established by multilocus sequence typing (MLST) and pulsed-field gel electrophoresis (PFGE), respectively. Capsular polysaccharides (CPS), pilus and alpha-like-protein (Alp) family genes were also characterized by molecular techniques. MLST analysis revealed that these isolates were limited to three clonal groups and were clustered in six different lineages, i.e. ST (sequence type) 103, ST568, ST67, ST301, ST313 and ST570, of which ST568 and ST570 were new genotypes. PFGE analysis revealed that these isolates were clustered in 27 PFGE types, of which, types 7, 8, 14, 15, 16, 18, 23 and 25 were the eight major types, comprising close to 70% (71/102) of all the isolates. The most prevalent sequence types were ST103 (58% isolates) and ST568 (31% isolates), comprising capsular genotype Ia isolates without any of the detected Alp genes, suggesting the appearance of novel genomic backgrounds of prevalent strains of bovine *S. agalactiae*. All the strains possessed the pilus island 2b (Pl-2b) gene and the prevalent capsular genotypes were types Ia (89% isolates) and II (11% isolates), the conserved pilus type providing suitable data for the development of vaccines against mastitis caused by *S. agalactiae*.

**Introduction**

*Streptococcus agalactiae*, also referred to as group B streptococcus (GBS), is one of the leading causes of bovine mastitis, which has economically important implications for the dairy cattle industry throughout the world [1]. *S. agalactiae* is also an important human pathogen that can induce invasive infections in neonates, the elderly and pregnant women [2,3,4]. There is indirect evidence that *S. agalactiae* is transmitted between cattle and humans [5]. Clearly, control and prevention of *S. agalactiae* mastitis will improve the quantity and quality of milk production and have important significance for animal welfare and public health. As we know, studies of the molecular epidemiology of field *S. agalactiae* strains is vitally important in implementing efficient management practices in dairy farms. However, no information regarding the molecular characterization of *S. agalactiae* strains occurring in farms in China has previously been documented.

Multilocus sequence typing (MLST) and pulsed-field gel electrophoresis (PFGE) typing are two genotype methods used to characterize and distinguish specific clones among GBS isolates [6]. MLST is an unambiguous sequence-based and reliable typing tool, allowing comparison of the gene distribution of different isolates collected from all geographic areas and further investigation of the population structure [6,7]. Five hundred and eighty-three *S. agalactiae* sequence types (STs) have been identified and made available on the MLST website as of May 2012 (http://pubmlst.org/sagalactiae/), although information about bovine strains is still limited.

Capsular serotyping is a classical method used for *S. agalactiae* in epidemiological studies. To date, ten serotypes, based on the *S. agalactiae* capsular polysaccharides (CPS), have been identified, including Ia, Ib, II-VIII and a new serotype IX [8]. Capsular genotyping is considered more suitable for epidemiological investigation because the serotypes can be identified with or without the CPS expression.

The alpha-like protein (Alp) family play an important role in *S. agalactiae* pathogenesis and are also vaccine candidates [9]. Six members of the Alp family have been extensively studied, including Alpha-C, Rib, Alp1 (Epsilon), Alp2, Alp3 and Alp4 [10], they are encoded by genes bca, alp1 (Epsilon), alp2, Rib, and alp, respectively.
Three types of pili of S. agalactiae have been identified and designated as pilus island 1 (PI-1), PI-2a and PI-2b, of which PI-2a and PI-2b are encoded by genes located in two distinct loci in the same region of the genome, while the PI-1 gene is located in a separate region [11].

The objectives of this study were to elucidate the relationships between bovine S. agalactiae strains by molecular characterization based on capsular genotyping, pilus and Alp gene profiling and MLST and PFGE analyses.

Results

Identification of S. agalactiae in dairy cows

Milk samples were collected from a total of 619 cows with subclinical mastitis from 33 dairy farms located in six provinces (Jiangsu, Anhui, Shandong, Zhejiang, Jiangxi and Fujian) and one municipality (Shanghai). S. agalactiae was detected in milk samples from 102 cows from 21 farms located at five provinces (except Fujian province) and one municipality. In total, 102 bovine isolates (one isolate per cow) of S. agalactiae were selected and analyzed (a complete list of all isolates is provided in Table S1).

Capsular genotype and alpha-like protein

All 102 bovine S. agalactiae isolates belonged to capsular genotype Ia and II (Table 1) according to the multiplex PCR assay. Overall, type Ia was the most prevalent accounting for approximately 89% (91/102) of isolates. Type II was represented by 11% (11/102) of isolates. A large proportion (93%; n = 95/102) of the bovine S. agalactiae isolates was non-typeable (NT) to any of the detected Alp genes according to the multiple PCR assay (Table 1). Only 5% (5/102) of Alp1 gene-positive strains and 2% (2/102) Alp4 gene-positive strains were characterized. The alpha-like protein genes associated with the capsular genotype were also identified (Table 1):

Table 1. CCs, STs, capsular genotypes, pilus types, alpha-like protein genes and PFGE types of the 102 bovine S. agalactiae strains.

| CCs and STs | Allelic profile | Capsular genotype (No. of isolates) | Frequency (%) | Alp gene (No. of isolates) | PFGE type (No. of isolates) |
|-------------|----------------|-----------------------------------|---------------|---------------------------|-----------------------------|
| CC67        | 10             |                                    |               |                           |                             |
| 67          | 13 1 1 13 1 1 5 | Ia (1)                            | 1             | 2b                        | NT' (1)                     |
|             |                |                                   |               |                           |                             |
| 301         | 13 1 1 13 28 5 | II (4)                            | 4             | 2b                        | NT (4)                      |
|             |                |                                   |               |                           |                             |
| 313         | 13 1 2 1 28 5 | II (1)                            | 1             | 2b                        | alp1 (1)                    |
|             |                |                                   |               |                           |                             |
| CC64        | 2              |                                    |               |                           |                             |
| 570         | 16 1 2 1 5     | II (2)                            | 2             | 2b                        | alp4 (2)                    |
|             |                |                                   |               |                           |                             |
| CC103       | 88             |                                    |               |                           |                             |
| 103         | 16 1 6 2 9 9 2 | Ia (58)                           | 57            | 2b                        | NT (58)                     |
|             |                |                                   |               |                           |                             |
| 568         | 16 1 6 2 51 9 2 | Ia (32)                           | 31            | 2b                        | NT (32)                     |

a CC, clonal complex. b ST, sequence type. c The allelic profiles are presented in the following order: adhP, pheS, atr, gluA, sdhA, glcK, and tkt. d PI, pilus island. Minus in this column indicates the absence of a PI-1 gene. e Alp, alpha-like protein. f NT, non-typeable.

Pilus type

Each strain was detected in the three PCR assays for pilus typing. PI-1 and PI-2a genes were absent in all of the investigated bovine strains although the corresponding genes were detected in the positive controls. All 102 bovine S. agalactiae strains carried the PI-2b gene alone (Table 1).
MLST sequence types and PFGE analysis

All 102 isolates were characterized using MLST; one novel allele (sdhA allele 51, GenBank accession number is KF006268) and two new STs (ST 568 and ST570) were identified. ST568 is a single-locus variant (SLV, in which one allele differs from the ST) of ST103, ST568 and ST570 are grouped within three clonal complexes (CCs): CC103, CC67 and CC64. Both ST67 and ST64 were subgroups of CC17. ST568 was a single-locus variant (SLV) for ST103. ST301 and ST313 were SLVs and DLVs (two-locus variants), respectively of ST67. ST570 was a DLV of both ST64 and ST67, derived from ST64. The predicted founders that were not obtained in this study are marked by dashed arrows. For clarity, ST labels have been removed.

The isolates were grouped in 27 PFGE types in this study, of which 16 common (represented by dashed rectangles) and 11 unique types were identified, with a similarity between 43.9% and 100% (Figure 2). Eight major PFGE types (7, 8, 14, 15, 16, 18, 23 and 25 shown in Figure 2) comprised close to 70% (71/102) of all the isolates. A total of eight more minor types, comprising only two or three isolates, corresponded to almost 20% (20/102) of all the isolates.

Table 1 shows the 11 capsular genotype II isolates that were distributed among four STs (ST67, ST301, ST313, ST570) and eight PFGE types. The 91 type Ia isolates belonged to three
isolates of serotype II carrying the Alp genes and represented 57% (58/102) and 31% (32/102) of the isolates, respectively (Table 1). These isolates were clustered into 20 types according to the PFGE analysis. ST67, ST313 (DLV of ST67) and ST301 (SLV of ST67), were grouped in CC67 (Figure 1). ST67 represented 5% (5/102) of the isolates assigned to capsular genotype II (4 isolates) and 1a (1 isolate) associated with NT Alp genes, and were further clustered in four PFGE types. ST313 and ST301 comprised a total of 5% (5/102) of the isolates, which were assigned to capsular genotype II carrying Alp1 genes and belonged to five PFGE types. ST570, belonging to CC64, represented only two isolates of serotype II carrying the Alp4 gene and clustered in PFGE type 9 (Table 1).

The isolates clustered in the same ST were clearly divided into several distinct PFGE types. For example, ST103 and 568, were divided into nine and eleven different PFGE types, respectively (Table 1 Figure 2). It was also observed that the isolates belonging to different capsular genotypes were clustered in the same ST or PFGE type, while the same capsular genotypes were divided into different ST or PFGE types (Table 1 Figure 2).

**Distribution of S. agalactiae between and within farms**

The STs, molecular genotypes, Alp genes of S. agalactiae among the 21 S. agalactiae positive farms are presented in Figure 3. Seven different genotypes were found when combining these three types. The predominant type, ST668, a novel ST which is a SLV of ST67, combined with Ia and NT and was found exclusively among isolates from farms located in northern provinces (Jiangsu and Shandong) only. ST103, the predicted founder of ST568, combined with Ia and NT, and with few exceptions, was found exclusively among isolates in the farms located in the southern provinces (including Shanghai, Anhui and Zhejiang).

On the basis of the combination of genotypes described, the isolates were characterized into 31 types when combined with the PFGE genotypes (Figure 3). Comparison of S. agalactiae isolates between herds revealed similar genotypes (10 types) distributed in different herds, and herd specific types (21 types) were also observed (Figure 3). Comparison of S. agalactiae isolates from the same herd showed S. agalactiae isolates displaying either different types (in 12 herds) or the same type (in 9 herds) within a same herd. Clustering of multiple isolates of the same type within an individual herd was also observed on several farms.

**Discussion**

In the present study, 21 of 33 dairy farms screened positive for S. agalactiae, although control measures were managed in these farms. It was reported that the herd level prevalence of S. agalactiae increased steadily from 2000 to 2008 in Denmark, even with systematic control measures in place, which had previously been effective in preventing S. agalactiae mastitis [12]. The increasing prevalence may be due to an increase infection rate that exceeds the capacity for eradication. This demonstrates that S. agalactiae remains a significant cause of mastitis in cattle herds, and more effective management is required to control S. agalactiae mastitis.

The CPS of S. agalactiae are one of the most important virulence factors and form the main components of multivalent vaccines [13]. Capsular genotyping by multiplex PCR revealed that Ia was the most predominant capsular genotype in Eastern China. Similar findings have been reported in Germany where serotype Ia was shown to be prevalent in 19 of 79 bovine S. agalactiae isolates [14]. However, serotype III and serotypes V and IV were found to be the most prevalent in Quebec (Canada) and Norway, respectively [15,16]. The diversity in serotype distribution of mastitis S. agalactiae might be the result of divergent geographical regions, times, management practices and breeds of cow. In the present study, all the bovine S. agalactiae isolates from 21 farms of five provinces and one municipality in Eastern China belonged to serotype Ia and II.

The major surface-localized protein antigens of S. agalactiae belong to the alpha-like protein (Alp) family of surface proteins. It is reported that S. agalactiae strains usually carry at least one of the alpha-like proteins, of which proteins Epsilon (Alp1), Alp2, Alp3, Alp4, Alpha-C and Rib have been extensively studied [17], although several bovine and human strains have been reported to be negative for any of the six proteins [16,17,18]. In this study, the multiple PCR assay for detecting Alp genes showed that 92% of the tested bovine isolates were non-typeable, possibly due to mismatch of the primers for genes encoding alpha-like proteins in these isolates. Many studies have previously shown the associations of Alp genes and serotypes; for example, the Alpha-C protein gene with...
S. agalactiae strains carry at least one of the three pilus islands. Margarit et al. [20] demonstrated that all 289 human strains they investigated carried either PI-2a (73% strains) or PI-2b (27% strains), while PI-1 was missing in 28% of the strains. Following analysis of 238 isolates, Sørensen and colleagues [21] found that all the isolates carried PI-2, but PI-1 was missing in 54% of the bovine isolates and in 24% of the human isolates; therefore, it was concluded that PI-1 genes do not exist in several lineages. In this study, all of the 102 bovine S. agalactiae strains from Eastern China carried only PI-2b, while PI-1 was absent from all of the strains. Since previous studies have shown that pilus-based vaccines can be effective in preventing infections caused by homologous challenge of S. agalactiae [20], PI-2b proteins may be considered as potential vaccine candidates for formulation of subunit vaccines against bovine S. agalactiae mastitis.

Two novel sequence types, ST568 and ST570, were found in this study, which was expected because the information in the MLST database about the S. agalactiae isolates prevailing in China is severely lacking. Thus, the submission of our data to this database enriches the available data on bovine S. agalactiae isolates. The novel sequence type ST568 and its predicted founder ST103 were the predominant STs found in this study, ST103 is occasionally obtained from dairy cows according to previous reports [22]. However, a recent study shows ST103 to be a predominant ST in bovine strains from Denmark [23] and here it is shown to be prevalent in Eastern China, which contributes to a better understanding of the global epidemiology of mastitis S. agalactiae isolates. The new sequence type ST570, which is a SLV of ST 64, represented only 2% of the isolates in this study. Conversely, its predicted founder ST67 has been reported as a common ST among bovine S. agalactiae strains [21]. ST67, ST301 and ST313 were grouped in CC67, representing only a total of 12% of the isolates in this study. However, ST67 has previously been considered to be the most common ST among bovine isolates [2]. The occurrence of unique and identical STs identified between Eastern China and other countries shows that the types of ST and predominant STs differ in bovine isolates from diverse geographical regions. However, limited clonal groups appear in different region and countries according to MLST analysis. In present study, CC67 and CC64 showed a high degree clonal of relatedness, while CC103 remained as a distinct group, suggesting the S. agalactiae strains from bovine milk in Eastern China comprise two genetically distinct populations.

In this study, identical capsule genotypes belonged to different clonal groups and identical STs, PFGE types shared by various capsular genotypes were observed. This switching may be the result of horizontal transfer of capsular genes, which is likely to be driven by the host immune response and supported by the increased fitness acquired by isolates showing specific phenotype-genotype combinations [6,24].

Similar sequence types prevalent on several different farms were observed in this study. The novel ST568 is prevalent in the north, while ST103, the predicted founder of ST568, is predominant in the south of this region, thus indicating that the prevailing S. agalactiae on these related farms are from the same strain, and might be transmitted from farms in the south to those in the north. It can be speculated that this occurs as a result of the commercial movement of infected animals between farms. Further analysis using PFGE demonstrated that the isolates grouped in the same ST were divided into several different PFGE types. The combination of genotypes presented several phenomena: farm specific in several farms, homology of the isolates among several farms, heterogeneity in isolates within several individual herds and multiple cows infected by a single strain on the same farms. S. agalactiae is a well-known contagious mastitis pathogen, with transmission occurring between cows within herds [1]. This mode of transmission is thought to explain how a single strain prevails in the same herd in the present study and in previous reports [25]. Different PFGE types were found in isolates clustered in the same STs, which may be accounted for by different management practices between farms. Homology of isolates in different farms and heterogeneity in isolates within individual herds are reported for the first time in this study. As previously speculated, the homology of isolates in different farms might also be due to commercial movement of infected animals between farms; however, it is not known why and how several different genotypes of S. agalactiae strains were obtained from the same herd, because few molecular epidemiological studies of sources of infection or transmission routes have been conducted in cattle. This heterogeneity might suggest the emergence of several strains originating from several sources of infection, with infected humans as the suspected infection source [26].

In conclusion, the results of the present study demonstrate that the distribution of STs, capsular genotypes, and pilus genes among the dairy cattle in Eastern China were similar to previously reports of bovine S. agalactiae strains; however some geographic characteristics were revealed by the emergence of unique Alp profiles and prevalent novel STs. ST103 to be a predominant ST in bovine mastitis from Eastern China, which contributes to a better understanding of the global epidemiology of mastitis S. agalactiae isolates. The conserved pilus type of S. agalactiae isolates provides positive information for the development of vaccines against S. agalactiae. Further investigation is necessary to establish the epidemiology of S. agalactiae, and to determine how S. agalactiae can be best controlled and prevented.

Materials and Methods

Ethics

Milk samples were obtained with consent from animals with subclinical mastitis under the ethical approval granted by College of Veterinary Medicine, the Nanjing Agricultural
University Veterinary College. The protocol was permitted by the owners of the dairy farms under investigation. All efforts were made to minimize animal suffering.

**S. agalactiae reference strains**

Four reference strains of *S. agalactiae*: ATCC 13813 (serotype II), ATCC 12403 (Serotype III), ATCC BAA-611 (Serotype V) and A909 (Serotype Ia), used in this study were obtained from the American Type Culture Collection (ATCC) as controls.

**Identification of *S. agalactiae* from milk samples**

A total of 619 cows from 33 large-scale diary farms located in Eastern China were selected to participate in this study according to the willing of the farmers (information about the farms were provided in Table S2). Holsteins dairy cows were preferably kept on the 33 dairy farms with a mean yield at 7,539 kg of milk. For each farm, milk somatic cell counts of the subclinical mastitis cow was washed and dried with a clean face towel. Each teat was disinfected with swabs soaked in 3 ml of Tryptic soya broth (TSB, MO BIO, Laboratories. Inc.) preheating at 37°C, followed by 15 cycles of 95°C for 1 min, 54°C for 1 min, and 72°C for 2 min and then by an additional 25 cycles of 95°C for 1 min, 56°C for 1 min, and 72°C for 2 min with a final cycle of 72°C for 10 min.

**Bacterial culture and DNA extraction**

The isolates, including the reference strains, were cultured in 3 ml of Tryptic soya broth (TSB, MO BIO, Laboratories. Inc.) overnight at 37°C. The bacterial culture was centrifuged (14,000 × g for 5 min at room temperature) and the pellets were harvested and resuspended in 200 µl TE buffer (10 mM Tris-HCl, pH 8.0; 1 mM EDTA, pH 8.0) supplemented with 20 mg/ml (final concentration) of lysozyme and incubated at 37°C for 30 min. Genomic DNA was extracted using the Bacterial DNA extraction kit following manufacturer’s specifications for Gram-positive bacteria (Omega Bio-Tek, USA). The extracted DNA was used as the template for PCR.

**Capsular genotyping**

The capsular genotype Ia, Ib, II–IX of *S. agalactiae* was identified by a multiplex PCR assay as previously described [29]. The PCR system (total volume, 25 µl) contained 50 ng DNA template, 1× PCR buffer; 2 mmol/l MgCl₂, 200 µmol/l dNTPs (dATP, dTTP, dCTP, and dGTP), 400 nmol/l primers cspI-Ia-6-7-F and cspI-7-9-F, 250 nmol/l of each of the other primer and 0.3 U of HotMaster Taq DNA Polymerase (Tiangen, China). The PCR amplification conditions were as follows: preheating at 95°C for 5 min, followed by 15 cycles of 95°C for 1 min, 54°C for 1 min, and 72°C for 2 min and then by an additional 25 cycles of 95°C for 1 min, 56°C for 1 min, and 72°C for 2 min with a final cycle of 72°C for 10 min. Serotypes of strains were identified by analyzing the unique banding pattern following agarose gel (1.5% wt./vol.) electrophoresis.

**Alpha-like protein (Alp) family**

The alpha-like protein genes bca, alp1 (Epsilon), alp2/3, Rib, and alp4 in the strains were detected by a simple multiplex PCR assay described by Creti, et al. [17]. In brief, the PCR system (total volume, 25 µl) contained the following: 50 ng DNA template, 1× PCR buffer; 2 mmol/l MgCl₂, 200 µmol/l dNTPs, 400 nmol/l of each of the five pairs of primers; 0.3 U of HotMaster Taq DNA Polymerase (Tiangen, China). The amplification conditions were as follows: preheating at 96°C for 3 min, followed by 30 cycles of 95°C for 1 min, 58°C for 45 s, and 72°C for 45 s, with a final cycle of 72°C for 10 min. Amplification of the alpha-like protein genes was evaluated by agarose gel (2% wt./vol.) electrophoresis of the PCR products.

**Pilus typing**

To identify the presence of pilus islands in the strains, three PCR assays were used to detect the PI-1, PI-2a or PI-2b genes as described previously [20]. In brief, the PCR system (total volume, 25 µl) contained 10 ng DNA template, 1× PCR buffer; 2 mmol/l MgCl₂, 200 µmol/l dNTPs, 400 nmol/l of each of the six primers; and 0.25 U of TaqDNA polymerase (Tiangen, China). The amplification conditions were as follows: preheating at 94°C for 5 min, followed by 35 cycles of 94°C for 45 s, 54°C for 45 s, and 72°C for 2 min (according to the lengths of the amplicons) and concluding with a cycle of 72°C for 10 min. The identity of the PCR products was analyzed by agarose gel (1% wt./vol.) electrophoresis and deemed to be positive based on the expected size of the PCR fragment.

**MLST**

All the isolates were typed using multilocus sequence typing (MLST) in this study. The seven housekeeping genes (*adh, pheS, atr*, *glnA, sdhA, gck* and *ikd*) were amplified by PCR and internal fragments sequences were obtained as described previously [7]. For each isolate, the allele number and sequence types (STs) were defined by analysis of the alleles sequence in the MLST database (http://pubmlst.org/sagalactiae/). The allele sequences or previously undescribed ST were assigned new numbers and the data were deposited in the MLST database. CC analysis was performed using the entire *S. agalactiae* MLST database and eBURST program (http://eburst.mlst.net) [30].
PFGE

DNA was extracted and digested with the Smal restriction enzyme (Takara, China) as previously described [31,32]. The PFGE program was performed according to Chen et al. [32]. A Salmonella serotype Braenderup H9812 DNA digested with XbaI was used as the molecular size standard as recommended in PulseNet [33]. The S. agalactiae PFGE patterns were analyzed with BioNumerics (Applied Maths BVBA, Belgium) using an optimization setting of 0.00% and band position tolerance of 1.5%. Cluster analysis was performed using the Dice coefficient and UPGMA of the digitalized PFGE patterns for the 102 S. agalactiae strains. Genotypic related groups, characterized at 80% similarity or above are represented by dashed rectangles in the dendrogram.

Statistical analysis

Categorical data were analyzed by using the Pearson chi-squared test. When data were insufficient for the test demands, Fisher’s exact test and Likelihood-ratio tests were used. P values of <0.05 were considered to indicate statistical significance.

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