Status of the Parkinson’s disease gene family expression in non-small-cell lung cancer

Quan Xing Liu1†, Hong Zheng2†, Xu Feng Deng1, Dong Zhou1 and Ji Gang Dai1*

Abstract

Background: The purpose of this study is to detect the Parkinson’s disease gene family mRNA relative expression in the non-small-cell lung cancer (NSCLC) tumor tissue and analyze the association between tumor characteristics and the Parkinson’s disease gene family.

Methods: Tumor tissue and tumor-adjacent tissue of 114 NSCLC patients were collected and SYBR quantitative analysis was used to detect the relative expression level of nine Parkinson’s disease gene mRNAs. Then, paired sample test, two-sided Student’s t-test, or two-sided Wilcoxon rank sum test was performed to analyze the mRNA relative expression level of nine Parkinson’s disease gene mRNAs in different gender, tumor histology, and tumor stage.

Results: Overexpression in the tumors was detected in 46/114 (40.35 %) PARK1/4, 74/114 (64.91 %) PARK2, 104/114 (91.23 %) PARK5, 95/114 (83.33 %) PARK6, 80/114 (70.18 %) PARK7, 100/114 (87.72 %) PARK9, 55/114 (48.25 %) PARK15, and 99/114 (86.84 %) glucocerebrosidase (GBA). Five genes PARK5 (91.23 %), PARK6 (83.33 %), PARK7 (70.18 %), PARK9 (87.72 %), and GBA (86.84 %) were supposed to be overexpressed in the lung tumor tissues compared with tumor-adjacent tissues. There was no significant difference in PARK1/4, PARK2, PARK5, PARK9, and GBA mRNA expression by different tumor stage, whereas, PARK6, PARK7, PARK8, and PARK15 mRNA expression were found to have significant difference in the comparison of different tumor stages. The expression of PARK6 (P = 0.01, P = 0.03) and PARK15 (P < 0.001, P < 0.001) were significantly higher in stages I and II when compared with stage III, respectively. NSCLC patients in stage I showed the higher expression PARK7 compared to the patients in stage II (P = 0.003).

Conclusions: The high expression of PARK6, PARK7, and PARK15 might lead to the occurrence of a primary NSCLC tumor, and the tumor with a decreasing expression of these three genes tends to be stages II and III. The results of our study indicate that the Parkinson’s disease gene family may be a potential marker for the prediction of NSCLC.

Keywords: Carcinoma, Non-small-cell lung cancer, Parkinson’s disease, Gene, mRNA

Background

Parkinson’s disease is a chronic, progressive neurological disorder caused by the degeneration and death of cells in the substantia nigra [1]. Historically, Parkinson’s disease was regarded as a sporadic disorder, with little or no contribution from hereditary factors. However, over the past 15 years, there has been a step change in our genetic understanding of the condition. SNCA the sodium channel protein para (which encodes α-synuclein) was the first gene to be discovered through linkage analysis in several large kindreds with familial Parkinson’s disease [2]. Recent genome-wide association studies have found that more than a dozen loci (including PARK1/4, PARK2, PARK5, PARK6, PARK7, PARK8, PARK9, PARK15, and glucocerebrosidase (GBA)) have been linked with familial Parkinson’s disease, and currently, upwards of 10 % of all cases of Parkinson’s disease are estimated to be associated with Parkinson’s disease gene family [3–5].

Fifteen years ago, Møller et al. [6] first reported that patients with Parkinson’s disease (PD) seem to have a lower-than-expected rate of lung cancer which has been reaffirmed by many subsequent epidemiological studies.
This association might account for the overlap function of the Parkinson’s disease gene family between these two apparently unrelated diseases. Recent genetic studies and emerging functional work show that a possible connection between Parkinson’s disease and lung cancer is alluded.

Cancer cells are prone to accumulate mutations, not all of which will necessarily contribute to cancer progression (these are known as passenger mutations, in contrast to the pivotal driver mutations that propel cancerous change). However, when the functional roles of the Parkinson’s disease gene family are considered, a picture emerges of considerable pathogenic overlap between these two apparently unrelated diseases. Currently, several studies have reported that cell systems are affected by the same dysfunctional protein in both Parkinson’s disease and cancer [11–13].

In short, the same mutations can lead either to inappropriate neuronal death in Parkinson’s disease when present in the germ line or to inappropriate cell survival in cancer when present in somatic cells. SNCA (PARK1 or PARK4) was reported overexpressed in brain [14], malignant [15], and ovary tumors [16] while low-expressed or not expressed in those normal and benign tissues. PARK7 (DJ-1) was reported overexpressed in the non-small-cell lung cancer (NSCLC) and prostate cancer cell line [17, 18]. Although many studies have reported that some Parkinson’s disease gene family are overexpressed or low-expressed in brain, malignant, ovary, and colon cancer patients and cell lines, the expression of the Parkinson’s disease gene family in NSCLC patients and the association of clinical data are still unknown.

In this article, we investigated the expression of the Parkinson’s disease gene family in NSCLC patients and further analyzed its association with different tumor histological type, gender, and tumor stage. Our analysis is at times speculative, and additional experimental research may be required to further identify the association between Parkinson’s disease gene family and NSCLC which might help us to discover the new potential oncogenes or tumor suppressors and thus how to treat NSCLC.

**Methods**

**Patients**

All the 114 NSCLC patients in the study were diagnosed and histopathologically confirmed in our hospital between Feb 2012 and Apr 2013 and without any other cancers or previous chemo- or radiotherapy. The 114 patients were aged 18–65 years old (56.4 ± 10.7) at the time of diagnosis including 50 squamous carcinomas (SQC), and 64 adenocarcinomas (ADC) and were composed of 72 males and 42 females. This study was approved by the institutional review board of Xinqiao Hospital (2012016), and all patients signed informed consent.

**Dispoze of tumor tissue and tumor-adjacent tissue**

Tumor tissue and tumor-adjacent tissue were collected at the time of surgical resection. All the samples were immediately snap-frozen in liquid nitrogen and stored at −170 °C before use. Total RNA was extracted following a TRIzol extraction protocol (Invitrogen, USA).

**Reverse transcription and SYBR quantitative real-time PCR assay**

Total RNA was transcriptioned reversely to cDNA by using PrimeScript™ RT reagent Kit with gDNA Eraser (TAKARA, Japan). SYBR (Synergy BrandsSynergy Brands) quantitative analysis of mRNA expression was used to investigate the expression of Parkinson’s disease gene family, including PARK1/4 (SCNA), PARK2 (Parkin), PARK5 (UCHL1), PARK6 (PINK1), PARK7 (DJ-1), PARK8 (LRRK2), PARK9 (ATP13A2), PARK15 (FBXO7), and GBA [19], in the tumor tissue and tumor-adjacent tissue. Primers for SYBR q-PCR were designed and synthesized by Sangon Biotech (China) (Table 1). PCR reactions (15 μl) contained 0.25 μl of each primer (10 μM), 7.5 μl SYBR® Premix Ex Taq TM II 2× (TAKARA, Japan), 6 μl ddH2O, and 1 μl cDNA. The PCR conditions consisted of an initial denaturation at 95 °C for 30 s followed by amplification for 40 cycles of 15 s at 95 °C and 50 s at 60 °C, with fluorescence acquisition at the end of each extension step. PCR using primers for β2-microglobulin was performed on each individual sample as an internal control. The optical density of each PCR band was measured semi-quantitatively using Illumina Eco software (Illumina, San Diego, USA).

**Statistical analysis**

Relative Parkinson’s disease gene family expression was calculated for each patient. The differences in mean value of Parkinson’s disease gene family expression in tumor tissue and tumor-adjacent tissue were analyzed using paired samples test. The differences in gene expression between gender, tumor histology, and tumor stage were analyzed using two-sided Student’s t-test or two-sided Wilcoxon rank sum test. All statistical analyses were performed using SPSS 18.0 software (SPSS Inc.), and $P < 0.05$ was considered to be statistically significant.

**Results**

**Relative expression level of Parkinson’s disease gene family**

Expression levels of nine Parkinson’s disease genes and a reference gene were measured in 114 matched pairs of NSCLCs/adjacent histologically normal lung tissue samples by real-time q-PCR. According to the relative mRNA
expression fold of nine different genes, patients were divided into five groups (Table 2). A relative gene expression ratio (T/N) of 1.5 was considered as positive for overexpression in the tumor when compared to the corresponding tumor-adjacent tissue. Values more than 1.5 have been used as the criterion for overexpression of genes in several studies employing proteomic analysis [20]. Therefore, overexpression in the tumors was detected in 46/114 (40.35 %) PARK1/4, 74/114 (64.91 %) PARK2, 104/114 (91.23 %) PARK5, 95/114 (83.33 %) PARK6, 80/114 (70.18 %) PARK7, 55/114 (48.25 %) PARK8, 100/114 (87.72 %) PARK9, 55/114 (48.25 %) PARK15, and 99/114 (86.84 %) GBA. PARK7 (DJ-1) was considered to be overexpressed in primary NSCLC tissues in a previous study [18], and 70.18 % of the patients were detected to have PARK7 overexpression in this study. Then, the five genes PARK5 (91.23 %), PARK6 (83.33 %), PARK7 (70.18 %), PARK9 (87.72 %), and GBA (86.84 %) were supposed to be overexpressed in the lung tumor tissue compared with the tumor-adjacent tissue. Inactivating mutations and deletions of PARK2 have been found in lung cancer [18] and overexpression of PARK15 was also detected in lung squamous cell carcinoma [21]; however, more than half (64.91 %) of the patients showed the overexpression of PARK2, and there was only 48.25 % PARK15-overexpression patients in this study. Their inconsistencies in findings may be caused by the small sample size and different research objects. The relationship between lung cancer and the expression of the other six genes PARK1/4, PARK5, PARK6, PARK8, PARK9, and GBA was first described (Fig. 1).

### Associations of tumor characteristics with Parkinson’s disease genes

Mean values of PARK1/4 (SCNA), PARK2 (Parkin), PARK5 (UCHL1), PARK6 (PINK1), PARK7 (DJ-1), PARK8 (LRRK2), PARK9 (ATP13A2), PARK15 (FBXO7), and GBA relative mRNA expression according to gender, tumor histology, and each clinical stage are summarized in Table 3. Significant statistical difference of relative mRNA expression level of these nine genes was detected in the comparison between tumor tissue and tumor-adjacent tissue. The relative expression of these nine gene mRNAs showed no significant difference between the male groups and female groups. However, the ADC group showed significantly

### Table 1 Sequence of primers used in Parkinson’s disease gene mRNA real-time q-PCR

| Primer       | GeneBank ID | Sequence forward (5′–3′)                      | Sequence reverse (5′–3′)                      | Product (bp) |
|--------------|-------------|-----------------------------------------------|-----------------------------------------------|--------------|
| PARK1/4 SCNA | NM000345.3  | AAACCAAGGGAGGAGTGGTG CTGTCTTCTGGGCTACTGCTG   | CTGCTTCTGGCTACTGCTG                          | 117          |
| PARK2 Parkin | AB009973.1  | CTGACACACGACATCTCCACG                       | CCAGTTCCCTTCACGCTCCT                        | 107          |
| PARK5 UCHL1 | BC003321.2  | GCCAATGCTGGGTAGATGAC AGCGGACTTCTCCTTGATTT    | AGCGGACTTCTTGACTTCCTG                       | 192          |
| PARK6 PINK1  | A8053323.1  | CAAGAGAGGTCCAAGACGCAAC                       | GGCAGCACATGAGGTTAGTC                       | 102          |
| PARK7 DJ-1   | D61380.2    | TGGCTAAAAAGGAGCAGGAGAA                      | ATGGACCATCAGCGGCTACA                        | 127          |
| PARK8 LRRK2  | AY792511.1  | GAGCACACGCTTCAAGTTATT                       | AGTGAAGGAAACCAACCGCTG                      | 110          |
| PARK9 ATP13A2| AL354615.1  | TGGCTGGCTTGGCACCCTACTAC                     | AGTCTGGCTTTGTCTCTGG                       | 102          |
| PARK15 FBXO7 | AF233225.1  | TACCCGACAAAGCACGTGACC                      | AAGACCGGAACGACATCCCGAC                    | 102          |
| GBA          | D13286.1    | CTCTCTGCTGGGCTTGTGTGAGT                     | TACTGTGGCGGAGGTCAGGA                       | 108          |
| β2-microglobulin | NM004048.2 | ACCCCCACTGAAAAGATGAG                       | ATCTTCAACCTCCATGAGT                      | 114          |

### Table 2 The frequency distribution of Parkinson’s disease gene mRNA relative expression

| Fold | PARK1/4 SCNA (n = 114) | PARK2 Parkin (n = 114) | PARK5 UCHL1 (n = 114) | PARK6 PINK1 (n = 114) | PARK7 DJ-1 (n = 114) | PARK8 LRRK2 (n = 114) | PARK9 ATP13A2 (n = 114) | PARK15 FBXO7 (n = 114) | GBA (n = 114) |
|------|------------------------|------------------------|----------------------|----------------------|----------------------|----------------------|------------------------|------------------------|----------------|
| ≤0.5 | 23 (20.18 %)           | 18 (15.79 %)           | 1 (0.88 %)           | 9 (7.89 %)           | 9 (7.89 %)           | 22 (19.30 %)         | 5 (4.39 %)            | 14 (12.28 %)          | 6 (5.26 %)    |
| 0.5–1.5 | 45 (39.47 %)         | 45 (39.47 %)           | 9 (7.89 %)           | 10 (8.77 %)          | 25 (21.93 %)         | 37 (32.46 %)         | 9 (7.89 %)            | 45 (39.47 %)          | 9 (7.89 %)    |
| ≥1.5 | 46 (40.35 %)           | 74 (64.91 %)           | 104 (91.23 %)        | 95 (83.33 %)         | 80 (70.18 %)         | 55 (48.25 %)         | 100 (87.72 %)         | 55 (48.25 %)          | 99 (86.84 %)  |
| ≥5  | 2 (1.75 %)             | 19 (16.67 %)           | 91 (81.23 %)         | 45 (39.47 %)         | 14 (12.28 %)         | 14 (13.16 %)         | 56 (49.12 %)          | 56 (49.12 %)          | 56 (49.12 %)  |
| ≥10 | 0 (0.00 %)             | 4 (3.51 %)             | 70 (61.40 %)         | 16 (14.04 %)         | 2 (1.75 %)           | 2 (1.75 %)           | 39 (34.21 %)          | 0 (0.00 %)            | 23 (20.18 %)  |

Fold means the fold of relative mRNA gene expression to reference gene β2-microglobulin.
higher PARK2 ($P < 0.001$) and PARK7 ($P \approx 0.005$) mRNA expression than the SQC group, and no significant difference was found in the other seven genes. There was no significant difference in PARK1/4, PARK2, PARK5, PARK9, and GBA mRNA expression by different tumor stage, whereas PARK6, PARK7, PARK8, and PARK15 mRNA expression were found to have significant difference in the comparison of different tumor stages. The expression of PARK6 ($P = 0.01$, $P = 0.03$) and PARK15 ($P < 0.001$, $P < 0.001$) were significantly higher in stages I and II when compared with stage III, respectively. NSCLC patients in stage I showed the higher expression PARK7 compared to the patients in stage II ($P = 0.003$). Interestingly, the relative expression of PARK8 for the NSCLC patients in stage II was higher than that in stages I ($P = 0.04$) and III ($P < 0.001$).

Discussion

The curious cancer pattern in certain neurological conditions has drawn increasing attention as converging evidence suggests that one family of diseases may provide protection against the other. Over 50 years ago, it was anecdotally noted that patients with Parkinson’s disease (PD) seem to have a lower-than-expected rate for most cancers [22]. Cancer is characterized by unlimited cellular proliferation, while PD is a process of premature cell death. In this sense, the diseases appear to be opposing ends of the same spectrum. In fact, they share many genes and biological pathways, and these are often regulated in different directions. The main cell components (protein degradation, cell cycle, mitochondria, PI3K–AKT–mTOR pathway and inflammation) and the main Parkinson’s-disease-associated proteins (parkin, PTEN-induced putative kinase 1 (PINK1), DJ-1, leucine-rich repeat kinase 2 (LRRK2), glucocerebrosidase (GBA), and F-box protein 7 (FBXO7)) are depicted with the multiple interactions that exist between them.

Neurons and cancer cells are fundamentally different in how they use mitochondria. Whereas neurons use oxidative phosphorylation to generate ATP, cancer cells use glycolysis to a greater extent, which is partly explained by the hypoxic tumor environment. However, this alternative metabolism persists in tumor cells even in the presence of oxygen, as originally observed by Warburg nearly a century ago [23]. So, mitochondrial dysfunction has long been implicated in the development of cancer, and this perspective is now undergoing a renaissance. Functional studies of Parkinson’s disease genes indicate that the mTOR (mammalian target of rapamycin) pathway probably has a major bearing on neurodegeneration [24–26]. The mTOR pathway is a central regulator of cell growth and proliferation that mainly functions through the modulation of protein synthesis. The central importance of this pathway for cancer biology is reflected by the fact that an mTOR inhibitor, sirolimus (also known as rapamycin), is already in use in oncology practice, and several trials of PI3K and AKT inhibitors are underway [27–29]. Inflammation promotes tumorigenesis and progression by providing growth factors that sustain proliferative signalling and survival factors that limit cell death. Chronic inflammation is regarded as an enabling characteristic in cancer, and recent work suggests that a similar mechanism might drive pathological change in PD conditions [30, 31].

To date, only one study has examined the expression of the PARK7 in primary NSCLCs [18]. However, no comparison was made between expression levels in matched tumor/normal pairs for the Parkinson’s disease gene family. The objective of the current study was to determine whether the Parkinson’s disease gene family is overexpressed in NSCLCs as compared to adjacent histologically normal lung tissue samples and further determine the relationship of the expression levels to gender, tumor sub-type, and tumor stage. One hundred and fourteen matched
Table 3 Associations of tumor characteristics with Parkinson's disease gene mRNA relative expression in NSCLC.

| Tumor characteristics | PARK1/4 (SCNA) | PARK2 (Parkin) | PARK5 (UCHL-1) | PARK6(PINK1) | PARK7(LEPI) | PARK8(LRRK2) | PARK9 (APP132) | PARK15 (FBXO7) | GBA |
|-----------------------|----------------|----------------|----------------|-------------|-------------|-------------|---------------|---------------|-----|
|                       | Means ± SD     | P              | Means ± SD     | P           | Means ± SD  | P           | Means ± SD    | P              |     |
| Overall               | 1.475 ± 1.088  | 3.177 ± 3.646  | 18.375 ± 15.551 | 5.465 ± 2.773 | 18.904 ± 2.336 | 1.277 ± 2.350 | 8.356 ± 7.402 | 1.956 ± 1.783 | 5.936 ± 4.328 |
| Gender                |               |                |                |             |             |             |               |               |     |
| Male                  | 1.499 ± 1.099  | 3.095 ± 3.974  | 18.349 ± 16.095 | 5.566 ± 2.638 | 18.904 ± 2.079 | 2.336 ± 2.357 | 8.329 ± 7.751 | 2.207 ± 1.938 | 6.472 ± 4.709 |
| Female                | 1.433 ± 1.079  | .755*± .756*   | 18.419 ± 14.760 | 5.291 ± .776* | 3.005 ± .422* | 2.161 ± .703* | 8.401 ± .960* | 1.524 ± .500* | 5.016 ± .147** |
| Histology             |               |                |                |             |             |             |               |               |     |
| Adenocarcinoma        | 1.584 ± 1.080  | 4.900 ± 4.200  | 18.383 ± 14.360 | 5.422 ± 3.085 | 18.904 ± 2.110 | 2.346 ± 2.362 | 8.904 ± 7.890 | 2.125 ± 6.857 |     |
| Squamous cell carcinoma | 1.367 ± 1.116  | .303*± .202*   | 18.784 ± 17.247 | 5.436 ± .988* | 2.410 ± .005* | 1.901 ± .274* | 7.557 ± .337* | 1.775 ± .311* | 5.642 ± .282* |
| Tumor stage           |               |                |                |             |             |             |               |               |     |
| Stage I               | 1.723 ± 1.371  | .280*± .540*   | 5.403 ± 4.102* | 22.808 ± 18.141 | 7.404 ± 3.377 | 4.074 ± 1.391 | 4.040 ± 1.391 | 9.608 ± 7.410 | 3.240 ± 2.546 |
| Stage II              | 1.457 ± 0.936  | .178*± .598*   | 2.217 ± 3.27*  | 14.625 ± 11.717 | 3.562 ± 2.880 | 1.894 ± 1.941 | 1.053 ± 1.581 | 6.626 ± 5.643 | 1.567 ± 0.818 |
| Stage III             | 2.155 ± 0.854  | .720*± .130*   | 2.001 ± 1.783  | 19.939 ± 5.382 | 2.891 ± 1.002 | 2.765 ± 1.215 | 1.002 ± 1.380 | 10.198 ± 6.880 | 1.904 ± 3.098 |

I-II means the compare between stages I and II; II-III means the compare between stages II and III; I-III means the compare between stages I and III

a: Test for equality of medians between different gene relative expression and different sex: *two-sided Student's t-test; $^b$two-sided Wilcoxon rank sum test

b: Test for equality of medians between different gene relative expression and different tumor histology: *two-sided Student's t-test; $^b$two-sided Wilcoxon rank sum test

c: Test for equality of medians between different gene relative expression and different tumor stage: *two-sided Student's t-test; $^b$two-sided Wilcoxon rank sum test
human NSCLC tumor/normal pairs were examined by real-time q-PCR analysis for expression of PARK1/4 (SCNA), PARK2 (Parkin), PARK5 (UCHL1), PARK6 (PINK1), PARK7 (DJ-1), PARK8 (LRRK2), PARK9 (ATP13A2), PARK15 (FBXO7), and GBA mRNA. Using a value of 1.5 as the criterion for mRNA overexpression, elevated levels of Parkinson’s disease gene family mRNA in the NSCLC were detected and performed in Table 2. PARK7 (DJ-1) was considered to be overexpressed in primary NSCLC tissues in a previous study, and 70.18% of the patients were detected to have PARK7 overexpression in this study. Then, the five genes PARK5 (91.23%), PARK6 (83.33%), PARK7 (70.18%), PARK9 (87.72%), and GBA (86.84%) were supposed to be overexpressed in the lung tumor tissue compared with the tumor-adjacent tissue. There was no apparent correlation between the Parkinson’s disease gene family ratio (T/N) of different genders. PARK6, PARK7, PARK8, and PARK15 mRNA expression were found to have significant difference in the comparison of different tumor stages. The high expression of PARK6, PARK7, and PARK15 might lead to the occurrence of a primary tumor, but the tumor with a decreasing expression of PARK6 and PARK15 tends to be the stages II and III tumor. Only PARK2 and PARK7 in the ADC group showed significantly higher mRNA expression than those in the SQC group, which revealed that PARK2 and PARK7 may play a more important role in normal lung cells that transform into adenocarcinomas than those that become squamous cell carcinomas. Adjacent normal lung tissue is the best control we could obtain for this study. Although the tissue was observed to be histologically normal, it is unclear what molecular changes have occurred in these cells and, therefore, how far along these cells are with respect to developing into cancer cells.

Conclusions

The data presented in this manuscript suggest that further characterization of the cells that comprise both lung tumors and adjacent histologically normal tissue is warranted to establish the molecular mechanisms resulting in overexpression of the Parkinson’s disease gene family. Overall, the results of our study indicate that the Parkinson’s disease gene family may be a potential marker for the prediction of NSCLC. In a reciprocal manner, functional studies in a Parkinson’s disease context may provide novel insights into the role of cancer-associated genes. Ultimately, perhaps there will be therapeutic agents that can target both conditions.

Competing interests

The authors declare that they have no competing interests.

Authors’ contributions

JGD contributed substantially to conception and design and gave final approval of the version to be published. QL and HZ contributed to analysis and interpretation of all data and drafted the article. XD and DZ revised the article critically for important intellectual content. All authors read and approved the final manuscript.

Acknowledgements

The study was supported by the General Program of National Natural Science Foundation of China (81172388, 81472188). Ji Gang Dai and Quan Xing Liu are supported by the Third Military Medical University and the Department of Thoracic Surgery, Xinqiao Hospital. The funders had no role in the design and conduct of the study; the collection, management, analysis, and interpretation of the data; the preparation, review, or approval of the manuscript; or the decision to submit the manuscript for peer review.

Author details

1Department of Thoracic Surgery, Xinqiao Hospital, Third Military Medical University, Chongqing 400037, China. 2Institute of Immunology of PLA, Third Military Medical University, Chongqing 400037, China.

Received: 17 May 2015 Accepted: 14 July 2015

Published online: 07 August 2015

References

1. Jankovic J. Parkinson’s disease: clinical features and diagnosis. J Neurol Neurosurg Psychiatry. 2008;79:368–76.
2. Polymenopoulou MH, Lavedan C, Leroy E, Ide SE, Dehejia A, Dutra A, Pike B, et al. Mutation in the alpha-synuclein gene identified in families with Parkinson’s disease. Science. 1997;276:2604–7.
3. Hardy J. Genetic analysis of pathways to Parkinson disease. Neuron. 2010;68:201–6.
4. Sharma JC, Vassallo M. Prognostic significance of weight changes in Parkinson’s disease: the Park-weight phenotype. Neurodegener Dis Manag. 2014;4:309–16.
5. Fiala O, Zahorakova D, Pospisilova L, Kucerova J, Matejckova M, Martasek P, et al. PARK2 (PINK1) and PARK7 mutations are rare in Czech patients with early-onset Parkinson’s disease. PLoS One. 2014;9, e107585.
6. Moller H, Mellemkjaer L, McLaughlin JK, Olsen JH. Occurrence of different cancers in patients with Parkinson’s disease. BMJ. 1995;310:1500–1.
7. Minami Y, Yamamoto R, Nishikouri M, Fukao A, Hisamichi S. Mortality and cancer incidence in patients with Parkinson’s disease. J Neurol. 2000;247:629–34.
8. Olsen JH, Friis S, Frederiksen K, Mellemkjaer L, Moller H. Atypical cancer pattern in patients with Parkinson’s disease. Br J Cancer. 2005;92:201–5.
9. Elbaz A, Peterson BJ, Bower JH, Yang P, Maraganore DM, McDonnell SK, et al. Risk of cancer after the diagnosis of Parkinson’s disease: a historical cohort study. Mov Disord. 2005;20:719–25.
10. Lo PY, Tanner CM, Van Den Eeden SK, Albers KB, Leimpeper AD, Nelson LM. Comorbid cancer in Parkinson’s disease. Mov Disord. 2010;25:1809–17.
11. O’Flanagan CH, Morais VA, Wurst W, De Strooper B, O’Neill C. The Parkinson’s gene PINK1 regulates cell cycle progression and promotes cancer-associated phenotypes. Oncogene. 2015;34:1363–74.
12. Cao J, Lou S, Ying M, Yang B. DJ-1 as a human oncogene and potential therapeutic target. Biochem Pharmacol. 2015;93:241–50.
13. Cordero Y, Foguel D, Silva JL. Pressure-temperature folding landscape in proteins involved in neurodegenerative diseases and cancer. Biophys Chem. 2013;138:19–18.
14. Kawashima M, Suzuki SO, Doh-ura K, Iwaki T. Alpha-synuclein is expressed in a variety of brain tumors showing neuronal differentiation. Acta Neuropathol. 2000;99:154–60.
15. Matsuo Y, Kamitani T. Parkinson’s disease-related protein, alpha-synuclein, in malignant melanoma. PLoS One. 2010;5, e10481.
16. Bruening W, Glasson BJ, Klein-Szanto AJ, Lee VM, Trojanowski JQ, Godwin AK. Synucleins are expressed in the majority of breast and ovarian carcinomas and in preneoplastic lesions of the ovary. Cancer. 2000;88:2154–63.
17. Hod Y. Differential control of apoptosis by DJ-1 in prostate benign and cancer cells. J Cell Biochem. 2004;92:1221–33.
18. MacKeigan JP, Clements CM, Lich JD, Pope RM, Hod Y, Ting JP. Proteomic profiling drug-induced apoptosis in non-small cell lung carcinoma: identification of RhoGDIalpha and RS/DJ-1. Cancer Res. 2003;63:928–34.
19. Devine MJ, Plun-Favreau H, Wood NW. Parkinson’s disease and cancer: two wars, one front. Nat Rev Cancer. 2011;11:812–23.
20. Kennedy CH, Pass HI, Mitchell JB. Expression of human MutT homologue (hMTH1) protein in primary non-small-cell lung carcinomas and histologically normal surrounding tissue. Free Radiat Biol Med. 2003;34:1447–57.
21. Laman H, Funke JM, Ye H, Gallina-Garcia L, Hara E, Knowles P. Transforming activity of Fbxo7 is mediated specifically through regulation of cyclin D1/Cdk4. EMBO J. 2005;24:3104–16.
22. DOSHAY LJ. Problem situations in the treatment of paralysis agitans. J Am Med Assoc. 1954;156:680–4.
23. WARBURG O. On the origin of cancer cells. Science. 1956;123:309–14.
24. Iyer AM, van Scheppingen J, Milenkovic I, Anink JJ, Adle-Biassette H, Kovacs GG. mTOR hyperactivation in down syndrome hippocampus appears early during development. J Neuropathol Exp Neurol. 2014;73:671–83.
25. Heras-Sandoval D, Perez-Rojas JM, Hernandez-Damian J, Pedraza-Chaverri J. The role of PI3K/AKT/mTOR pathway in the modulation of autophagy and the clearance of protein aggregates in neurodegeneration. Cell Signal. 2014;26:694–701.
26. O’Neill C. PI3-kinase/Akt/mTOR signaling: impaired on/off switches in aging, cognitive decline and Alzheimer’s disease. Exp Gerontol. 2013;48:647–53.
27. Liu P, Cheng H, Roberts TM, Zhao JJ. Targeting the phosphoinositide 3-kinase pathway in cancer. Nat Rev Drug Discov. 2009;8:627–44.
28. Chia S, Gandhi S, Joy AA, Edwards S, Gorl M, Hopkins S. Novel agents and associated toxicities of inhibitors of the pi3K/Akt/mTOR pathway for the treatment of breast cancer. Curr Oncol. 2015;22:33–48.
29. Safdari Y, Khalili M, Ebrahimzadeh MA, Yazdani Y, Farajnia S. Natural inhibitors of PI3K/AKT signaling in breast cancer: emphasis on newly-discovered molecular mechanisms of action. Pharmacol Res. 2015;93:1–10.
30. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144:646–74.
31. Velatyati A, Yu WH, Sidransky E. The role of glucocerebrosidase mutations in Parkinson disease and Lewy body disorders. Curr Neurol Neurosci Rep. 2010;10:190–8.