Emodin Isolated from *Polygoni cuspidati* Radix Inhibits TNF-α and IL-6 Release by Blockading NF-κB and MAP Kinase Pathways in Mast Cells Stimulated with PMA Plus A23187

Yue Lu¹,², Yong-Tae Jeong³, Xian Li², Mi Jin Kim², Pil-Hoon Park², Seung-Lark Hwang², Jong Keun Son³,⁴* and Hyeun Wook Chang¹,²*

¹School of Pharmacy, Shanghai University of Traditional Chinese Medicine, Shanghai, 201203, China  
²College of Pharmacy, Yeungnam University, Gyeongsan 712-749, Republic of Korea

Abstract

Emodin, a naturally occurring anthraquinone derivative isolated from *Polygoni cuspidati* radix, has several beneficial pharmacologic effects, which include anti-cancer, anti-diabetic, and anti-inflammatory activities. In this study, the authors examined the effect of emodin on the production of proinflammatory cytokines, such as, tumor necrosis factor (TNF-α) and interleukin (IL)-6, in mouse bone marrow-derived mast cells (BMMCs) stimulated with phorbol 12-myristate 13-acetate (PMA) plus the calcium ionophore A23187. To investigate the mechanism responsible for the regulation of pro-inflammatory cytokine production by emodin, the authors assessed its effects on the activations of transcriptional factor nuclear factor-κB (NF-κB) and mitogen-activated protein kinases (MAPKs). Emodin attenuated the nuclear translocation of NF-κB, and of the MAPK pathway.

Key Words: Emodin, Pro-inflammatory cytokine, NF-κB, Mitogen-activated protein kinase, Bone marrow-derived mast cells, PMA plus A23187

INTRODUCTION

Mast cells are effector cells that display various functions during immune response and regulate allergic inflammation, such as, those associated with asthma, atopic dermatitis, and sinusitis. Activated mast cells release pro-inflammatory cytokines, such as, tumour necrosis factor (TNF)-α, interleukin (IL)-6, IL-8, IL-13, and inflammatory mediators, including histamine, leukotrienes, serotonin, prostaglandin (PG)E₂, and PGD₂ (Barrett and Austen, 2009; Mukherjee and Zhang, 2011). The expressions of proinflammatory cytokines depend on the activations of the transcription factor, nuclear factor-κB (NF-κB), and mitogen-activated protein kinases (MAPKs), such as, extracellular signal-regulated kinase (ERK), p38, stress-activated protein kinases (SAPK) and mitogen-activated protein kinase (JNK). Taken together, the findings of this study suggest that the anti-inflammatory effects of emodin on PMA plus A23187-stimulated BMMCs are mediated via the inhibition of NF-κB activation and of the MAPK pathway.

Received Sep 2, 2013 Revised Oct 28, 2013 Accepted Oct 31, 2013  
*Corresponding Author  
E-mail: hwchang@yu.ac.kr (Chang HW), jkoon@yu.ac.kr (Son JK)  
Tel: +82-53-810-2811 (Chang HW), +82-53-810-2817 (Son JK)  
Fax: +82-53-810-4654 (Chang HW), +82-53-810-4654 (Son JK)  
The first two authors contributed equally to this work.
binding of NF-κB to a specific sequence in the promoter region of its target genes, triggers its transcriptional activation (Amit and Ben-neriah, 2003; Takada et al., 2003; Xu et al., 2005).

Root extracts of polygono cuspidati (P. cuspidati) are used as an analgesic and to treat allergic inflammation in Korea, Japan, and China (Lim et al., 2007). Many groups have reported that P. cuspidati contains a large number of anthraquinones, naphthoquinones, flavonoids, and stilbenes. In particular, anthraquinone derivatives, such as, emodin, physcion, citreoresein, emodin-8-O-β-D-glucoside, physcion-8-O-β-D-glucoside, and cis- and trans-resveratrol have also been isolated from this plant (Hwangbo et al., 2012). Furthermore, emodin has been shown to have a number of biological activities, such as, anti-microbial, immunosuppressive, anti-inflammatory, and anti-atherosclerotic activities (Heo et al., 2008; Lee et al., 2010; Lin et al., 2010; Meng et al., 2010), and anti-diabetic and anti-obesity activities (Tzeng et al., 2012; Song et al., 2013).

In our previous studies, we described the anti-inflammatory activity of emodin and the mechanism of its action on FcεRI-mediated mouse bone marrow-derived mast cells (BMMCs) (Lu et al., 2011). Because, the effects of emodin on proinflammatory cytokine production in FcεRI independent stimulated mast cells has not been determined, we evaluated its inhibitory effect on the productions of IL-6 and TNF-α and sought to determine the mechanism responsible for its inhibitory effect in phorbol myristate acetate (PMA) plus A23187-stimulated BMMCs.

MATERIALS AND METHODS

Plant material

Emodin was isolated from the ethyl acetate fraction of a methanol extract of P. cuspidati radix as described previously (Lee et al., 2003). The emodin used in this study showed a single spot on TLC plates and had a purity exceeding 99.5% by HPLC analysis. Prior to use, emodin was dissolved in dimethyl sulfoxide (DMSO); final concentrations of DMSO in culture media were adjusted to 0.1% (v/v). DMSO alone was used as controls in all cases.

Chemicals and reagents

PMA, A23187 (calcium ionophore), and pyrrolidinedithio-carbamate (PDTC) were purchased from Sigma Chemical Co. (St. Louis, MO, USA). Enzyme-linked immunosorbent assay (ELISA) kits for TNF-α and IL-6 were purchased from R&D Systems Inc. (Minneapolis, MN, USA). Rabbit polyclonal antibodies for phospho-iKB, IKKα/β, ERK1/2, JNK, p38, β-actin, and the total forms of iKB, ERK1/2, JNK, p38 were purchased from Cell Signaling Technology, Inc. (Danvers, MA, USA). Rabbit polyclonal antibodies for p56 and IKKα/β, goat polyclonal antibodies for lamin B, rabbit anti-goat IgG-HRP and goat anti-rabbit IgG-HRP were obtained from Santa Cruz Biotechnology (Santa Cruz, CA, USA). The antibody-reactive bands were visualized with an enhanced chemiluminescence (ECL) system (Pierce Biotechnology, Rockford, IL, USA).

Preparation of mouse bone marrow-derived mast cells (BMMCs)

BMMCs isolated from male Balb/cJ mice (Sam Taco, Seoul) were cultured in RPMI 1640 media (Thermo Scientific, Utah, USA) containing 10% FBS, 100 U/ml penicillin (Thermo Scientific, Utah, USA), 10 mM HEPES buffer (Sigma-Aldrich, St. Louis, MO, USA), 100 μM MEM non-essential amino acid solution (Invitrogen, Carlsbad, USA) and 20% PWM-SCM (pokeweed mitogen-spleen cell conditioned medium) as a source of IL-3 (Lu et al., 2013). After 3 weeks, >98% of the cells were found to be BMMCs (Murakami et al., 1994).

Determinant of poinflammatory cytokine levels

BMMCs suspended in enriched medium at a cell density of 106 cells/ml were pretreated with the indicated concentrations of emodin for 1 h at 37°C and then stimulated with PMA (50 nM) plus A23187 (1 μM) for 6 h. Concentrations of IL-6 and TNF-α in supernatants were then determined using enzyme immunoassay kits (R&D Systems Inc. Minneapolis, MN, USA).

Immunoblotting

Immunoblotting was performed as previously described (Lu et al., 2011). Briefly, BMMCs were lysed in RIPA lysis buffer (50 mM Tris-HCl [pH 8.0], 150 mM NaCl, 5 mM EDTA, 1% Nonidet P-40, 1 mM PMSF, 1M DTT, 200 mM NaF, 200 mM Na3VO4, protease inhibitor cocktail). Cell debris was removed by centrifugation at 14,000 g for 15 min at 4°C and resulting supernatants were immunoblotted. Samples were separated by 8% SDS-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred to nitrocellulose membranes (Millipore, Bil-lerica, MA, USA), which were then blocked with 5% nonfat dry milk in Tris-buffered saline containing 0.1% Tween 20 and incubated with individual antibodies. Primary antibodies were diluted 1:1,000-fold (unless otherwise mentioned), and after treatment, membranes were incubated at 4°C overnight. Membranes were then washed three times for 10 minutes with TBS-T buffer, treated with HRP-coupled secondary antibodies (diluted 1:3000-fold) for 1 h at room temperature, washed three times for 3 min in TBS-T buffer, and developed using enhanced chemiluminescence (ECL) detection kits (Pierce Biotechnology, Rockford, IL, USA).

Preparation of nuclear and cytosolic extracts

Cell lysates were prepared as previously described (Lu et al., 2011). Briefly, cells were suspended in wash buffer containing 10 mM HEPES buffer (pH 8.0), 1.5 mM MgCl2, 10 mM KCl, 1 mM phenylethylsulfonyl fluoride (PMSF), and 1 nM protease inhibitor cocktail (Merck Biosciences, Darmstadt, Germany) and lysed in wash buffer containing 0.1% (v/v) NP40 by incubation on ice for 10 min. Supernatants (cyto- solic fractions) were obtained by centrifugation at 1,000 g for 4 min. Nuclear pellets were washed and re-suspended in a buffer containing 20 mM HEPES (pH 8.0), 25% (v/v) glycerol, 420 mM NaCl, 1.5 mM MgCl2, 0.2 mM EDTA, and protease inhibitor cocktail. These suspensions were then incubated for 30 min at 4°C and centrifuged at 10,000 g. The resulting supernatants were defined as nuclear fractions.

RNA isolation, reverse transcription and quantitative polymerase chain reaction (qPCR)

Real time PCR analysis was performed as described previ- ously (Lu et al., 2012). IgE-sensitized BMMCs were preincubated with emodin for 1 h, and then stimulated with DNP-HSA (100 ng/ml) for 4 h. Total RNA was extracted from BMMCs using the easy-BLUE™ Total RNA Extraction Kit (iNTRON Bio- technology, Inc, Sungnam, Korea). Five hundred nanogram of total RNA was reverse transcribed for the synthesis of cDNA

http://dx.doi.org/10.4062/biomolther.2013.068

436
using a one-step RT-PCR kit (Enzymomics, Daejon, Korea). Real time PCR amplification was then carried out over 40 cycles using the following conditions: denaturation at 95°C for 5 s, annealing at 57°C for 10 s, and elongation at 72°C for 20 s. The primers used for SYBR Green real-time PCR were as follows: for TNF-α, sense primer, 5’-AGC ACA GAA AGC ATG ATC CG-3’ and antisense primer, 5’-CTG ATG AGA GGG AGG CCA TT-3’; for IL-6, sense primer, 5’-GAG GAT ACC ACT CCC AAC AGA CC-3’; and antisense primer, 5’-AAG TGC ATC ATC GTT GTT CAT ACA-3’; and for β-actin, sense primer, 5’-ATC ACT ATT GGC AAC GAG CG-3’ and antisense primer, 5’-TCA GCA ATG CCT GGG TAG AT-3’. A dissociation curve for analysis of TNF-α, IL-6, and β-actin showed a single peak for each reaction. Mean Ct values of genes of interest were calculated from triplicate measurements and normalized versus the mean Ct of β-actin, which was used as a housekeeping gene.

NF-κB activation assay

Nuclear fractions were prepared as described previously (Lu et al., 2013). NF-κB transactivation ability was assessed using a NF-κB (p65) transcription factor assay kit (Cayman Chemicals, Ann Arbor, MI, USA). The results obtained are presented as percent activations with respect to the stimulated control.

Statistical analysis

All experiments were performed three or more times. Average values are expressed as means ± S.D. Statistical analyses were performed using SPSS 19.0 (SPSS, Chicago, IL, USA). The Student’s t-test was used to compare pairs of independent groups. Statistical significance was accepted for p values<0.05.

**Fig. 1.** Effect of emodin on PMA plus A23187-stimulated cytokine production and gene expression. BMMCs (10^6 cell/ml) were pre-treated with the indicated concentrations of emodin for 1 h and then incubated for 6 h with PMA (50 nM) plus A23187 (1 μM). TNF-α and IL-6 released into the supernatants were quantified by ELISA (A). The expression levels of IL-6 and TNF-α mRNAs were determined by real-time RT-PCR (B). Expression levels were normalized using expression of the housekeeping gene β-actin. Data are expressed as means ± S.D. of three independent experiments. *p<0.05 and **p<0.01 versus PMA plus A23187-stimulated BMMCs.

**Fig. 2.** Inhibitory Effects of emodin on PMA plus the A23187-stimulated IKK pathway in BMMCs. BMMCs were pre-incubated with emodin for 1 h and then incubated with PMA (50 nM) plus A23187 (1 μM) for 15 min. Cells were then immunoblotted for IKK, IκBα, NF-κB-p65 and β-actin. p-IKK/IKK (A), p-IκBα/β-actin (B) and p65/β-actin (C) protein level ratios were determined by quantifying the intensities of immunoblots with scanning densitometry (*p<0.05, **p<0.01 and ***p<0.001 versus PMA plus A23187-stimulated BMMCs). Results were obtained from three independent experiments, and are represented as relative ratios (%).
RESULTS

Effect of emodin on the productions of pro-inflammatory cytokines

Pro-inflammatory cytokines, such as, IL-6 and TNF-α, are produced by macrophages and mast cells, and play important roles in various inflammatory responses (Guha and Mackman, 2001). Initially, we examined the cytotoxicity of emodin on BMMCs using an MTT assay, but found that it did not affect cell viability at 50 μM (data not shown). Therefore, emodin was used at concentration below 20 μM in all experiments. To evaluate the effect of emodin on the productions of IL-6 and TNF-α, BMMCs were pretreated with the indicated concentrations of emodin for 1 h and then stimulated with PMA (50 nM) plus A23187 (1 μM) for 6 h. As shown in Fig. 1A, concentrations of IL-6 and TNF-α in media were considerably increased after stimulation with PMA plus A23187 and the release of both was dose-dependently inhibited by emodin. Next, we examined whether emodin suppressed the expressions of IL-6 and TNF-α at the transcription level. As shown in Fig. 1B, real-time RT-PCR showed that emodin significantly inhibited the inductions of both mRNAs.

Effect of emodin on the activation of the IKK/κB/α/NF-κB pathway

To evaluate the mechanism by which emodin inhibits the productions of IL-6 and TNF-α, we examined its effects on NF-κB activation. The expressions of these pro-inflammatory cytokines are regulated by the transcription factor NF-κB as mediated by IKK (Azzolina et al., 2003; Hayden and Ghosh, 2004). IKK complex includes regulatory scaffold protein NF-κB essential modulator (NEMO or IKKγ) and IKKα and IKKβ kinases. Once activated, IKK phosphorylates IκBα, and subsequent ubiquitination of phosphorylated IκBα by 26S proteasome releases NF-κB dimer from cytoplasmic IκBα/NF-κB (Peng et al., 2005). To determine whether emodin inhibits the phosphorylation of IKKα/β, the phosphorylation and degradation of IκBα, the nuclear translocation of NF-κB p65, and cytosolic NF-κB p65 activation, we pretreated BMMCs with indicated concentrations of emodin or 100 μM PDTC (a NF-κB pathway inhibitor and positive control). As shown in Fig. 2, both PDTC and emodin inhibited the phosphorylations of IKKα/β and IκBα and prevented IκBα degradation and disappearance of cytosolic p65. Furthermore, after pretreating BMMCs with emodin or PDTC for 1 h and then stimulating them with PMA plus A23187 for 30 min, the nuclear localization of the p65 subunit was dose-dependently blocked by emodin and PDTC (Fig. 3). Lamin B was used as an internal control in the nuclear fraction. Next, we examined the effect of emodin and of PDTC on the DNA binding activity of NF-κB using a DNA binding assay kit. As shown in Fig. 3B, emodin obviously suppressed the DNA binding activity of NF-κB in a dose dependent manner. PDTC was also found to inhibit NF-κB DNA binding activity strongly.

Effect of emodin on activation of MAP kinases

The MAP kinase pathway has been reported to play a major role in the regulation of proinflammatory mediator production (Beyaert et al., 1996; Baldassare et al., 1999). Thus, we examined the roles played by the MAPKs JNK, ERK1/2, and p38 on the productions of IL-6 and TNF-α in PMA plus A23187 stimulated BMMCs. As illustrated in Fig. 4, PMA plus A23187 strongly induced the expressions of phosphorylated ERK1/2, p38, and JNK without altering total protein levels. However, under the same experimental conditions, emodin significantly and dose-dependently suppressed the phosphorylations of these three MAPKs. Furthermore, the application of U0126 (an ERK1/2 inhibitor, 25 μM), SB203580 (a p38 inhibitor, 30 μM), or SP600125 (a JNK inhibitor, 25 μM) almost completely inhibited MAPK phosphorylation. These observations suggest that inhibition of the productions of IL-6 and TNF-α by emodin is mediated by blocking the phosphorylations of these three MAPKs in PMA plus A23187 stimulated BMMCs.

DISCUSSION

Emodin is an anthraquinone and is the major bioactive compound found in several herbs. Recently, several studies have shown emodin ameliorates metabolic disorder, has anti-arthritis activity, anti-allergic activity and that on molecular level, it activated AMPK, has PPARγ-agonist activity and acts...
as a potent phosphatase inhibitor (Feng et al., 2010; Chen et al., 2012; Han et al., 2012; Wang et al., 2012; Hwang et al., 2013). In this study, we investigated the effects of emodin on the productions of proinflammatory cytokine in phorbol myristate acetate (PMA) plus A23187-stimulated BMMCs and the mechanism responsible for its effects.

Mast cells are key effector cells and abundant in the airways of asthmatic patients, where they contribute to allergic inflammation by secreting an array of inflammatory mediators that exacerbate vasodilation and vascular permeability, airway smooth muscle contraction, mucus secretion, and immune cell recruitment (Galli et al., 2008; Price et al., 2013). Under inflammatory conditions, proinflammatory cytokines recruit activated immune and inflammatory cells to lesion sites, and thus, exacerbate inflammation (Church and Levi-Schaffer, 1997; Metcalfe et al., 1997). Mast cells release several cytokines, including TNF-α, IL-1β, IL-4, IL-5, IL-6, IL-8, and IL-13 (Galli et al., 2005). TNF-α is either preformed and stored in the granules of mast cells or newly synthesized following mast cell activation, and is a multifunctional cytokine that mediates a variety of immune and inflammatory responses. IL-6, another pleiotropic inflammatory cytokine produced by mast cells, also supports the biological roles of mast cells during allergy-induced inflammation (Nakae et al., 2005). In addition, MAPK signaling pathways have been implicated in the regulation of cytokine production after mast cell stimulation (Ishizuka et al., 1997; Turner and Cantrell, 1997; Boudreau et al., 2004).

It is also well known that cytokine gene expression is regulated by a combination of three different mechanisms, namely, the derepression of gene promoter, gene transcriptional activation by NF-κB, JNK, and ERK, and by the stabilization of mRNA via the p38 MAPK pathway. Furthermore, it has been reported the inductions of the gene expressions of IL-6 and TNF-α and the productions of IL-6 and TNF-α are depressed when cells are pretreated with either an inhibitor of the p38 MAPK pathway (SB203580) or an inhibitor of the ERK pathway (PD98059) (Guo et al., 2003; Glushkova et al., 2013). In this study, we investigated the effects of emodin on IL-6 production after mast cell stimulation (Ishizuka et al., 1997; Turner and Cantrell, 1997; Boudreau et al., 2004).

The expressions of various inflammatory cytokines are regulated by the NF-κB, which is strongly linked to inflammatory and immune responses. Thus, we postulate that emodin mediates the effects of NF-κB, at least partly, by suppressing its activation. The activation of NF-κB is dependent on the phosphorylation and degradation of IκB, an endogenous inhibitor that binds to NF-κB in the cytoplasm (Li et al., 2006; Moon et al., 2007). In the present study, stimulation by PMA plus A23187 led to activation of the key enzyme IκB, a complex composed of the regulatory IKKγ (NEMO) subunit and two enzymatically active subunits, IκKα and IκKβ, and IκK activation resulted in the phosphorylation of IκB and its subsequent dissociation from NF-κB (Fig. 2). IκKβ is considered primarily responsible for IκB phosphorylation, whereas IκKα is required for the processing of NF-κB (Sentfleben et al., 2001). Furthermore, when NF-κB binding subunits are released and translocated to the nucleus, they bind to specific elements in the upstream promoter regions of target genes, such as, TNF-α, IL-6, and IL-1β. (Peng et al., 2005). Since activation of the IKK-IκBζ-NF-κB axis leads to the productions of TNF-α and IL-6, we examined the effect of emodin or PDTC on the IKK/
NF-κB pathway. As illustrated in Fig. 2, 3, emodin and PDTC inhibit activation of the IKK-activated NF-κB and NF-κB DNA binding activity, which suggests that the NF-κB pathway plays an essential role in the productions of TNF-α and IL-6 in PMA plus A23187-induced BMMCs. MAPKs can also regulate the expressions of many genes via their actions on transcription factors, such as, NF-κB. In a previous study, ERK and p38 inhibitors were found to inhibit NF-κB activation and cytokine production (Sigala et al., 2011), whereas in the present study, MAPKs inhibitors significantly suppressed the productions of IL-6 and TNF-α. These findings suggest that crosstalk between MAPKs and NF-κB could mediate cytokine production.

Summarizing, this study shows that emodin, 1) regulates the productions of IL-6 and TNF-α, 2) inhibits MAPK phosphorylation, 3) suppresses the nuclear translocation of NF-κB, 4) suppresses the degradation of cytoplasmic IκBα, and 5) inhibits the phosphorylations of IκBα and IKK in PMA plus A23187-induced BMMCs. These findings and those of our previous studies (Lu et al., 2011; Alisi et al., 2012; Chu et al., 2012) suggest that emodin should be regarded as a potential means of treating mast cell-mediated allergic inflammatory diseases.

ACKNOWLEDGMENTS

This work was supported by a National Research Foundation (NRF) of Korea grant funded by the Korea government (MEST; 2012-0000283) and in part supported by Yeungnam University 2011 grant.

REFERENCES

Alisi, A., Pastore, A., Ceccarelli, S., Panera, N., Gnanì, D., Bruscalupi, G., Massimi, M., TOZZI, G., Piemonte, F. and Nobili, V. (2012) Emodin prevents interleukin-13-induced inflammation and redox status imbalance during diet-induced hepatosteatosis in rats. Int. J. Mol. Sci. 13, 2276-2289.

Amit, S. and Ben-neriah, Y. (2003) NF-kappaB activation in cancer: an essential role in the productions of TNF-α and IL-6. Semin. Cancer Biol. 13, 15-28.

Baldassare, J. J., Bi, Y. and Bellone, C. J. (1999) The role of p38 mitogen-activated protein kinase in IL-1 beta transcription. J. Immunol. 162, 5367-5373.

Barrett, N. A. and Austen, K. F. (2009) Innate cells and T helper 2 cell immunity in airway inflammation. Immunology 31, 425-437.

Beyaert, R., Cuenda, A., Vanden Berghe, W., Plaisance, S., Lee, J. C., Barrett, N. A. and Austen, K. F. (2009) Innate cells and T helper 2 cell immunity in airway inflammation and allergy. J. Immunol. 182, 1058-1068.

Beyaert, R., Cuenda, A., Vanden Berghe, W., Plaisance, S., Lee, J. C., Haegeman, G., Cohen, P. and Fiers, W. (1996) The p38/RK mitogen-activated protein kinase pathway regulates interleukin-6 synthesis and response to tumor necrosis factor. EMBO J. 15, 1914-1923.

Boudreau, R. T., Hoskin, D. W. and Lin, T. J. (2004) Phosphatase inhibition potentiates IL-6 production by mast cells in response to FcepsilonRI-mediated activation: involvement of p38 MAPK. J. Leukoc. Biol. 76, 1075-1081.

Chen, Z., Zhang, L., Li, Y., Yang, Z., Zhang, Z. and Li, Z. (2012) Promotion of apoptosis induction by emodin: a novel AMPK activator with PPARgamma-agonist activity. J. Cell. Biochem. 113, 3547-3558.

Chu, X., Wei, M., Yang, X., Cao, Q., Xie, X., Guan, M., Wang, D. and Deng, X. (2012) Effects of an anthraquinone derivative from Rheum officinale Baill, emodin, on airway responses in a murine model of asthma. Food Chem. Toxicol. 50, 2368-2375.

Church, M. K. and Levi-Schaffer, F. (1997) The human mast cell. J. Allergy Clin. Immunol. 99, 155-160.

Feng, Y., Huang, S. L., Dou, W., Zhang, S., Chen, J. H., Shen, Y., Shen, J. H. and Leng, Y. (2010) Emodin, a natural product, selectively inhibits 1beta-hydroxysteroid dehydrogenase type 1 and ameliorates metabolic disorder in diet-induced obese mice. Br. J. Pharmacol. 161, 113-126.

Galili, S. J., Grimaldil, M. and Tsai, M. (2008) Immunomodulatory mast cells: negative, as well as positive, regulators of immunity. Nat. Rev. Immunol. 8, 477-486.

Galili, S. J., Kalesnikoff, J., Grimaldi, M. A., Piliponsky, A. M., Williams, C. M. and Tsai, M. (2005) Mast cells as "tunable" effector and immunoregulatory cells: recent advances. Annu. Rev. Immunol. 23, 749-766.

Glushkovka, O. V., Parfenyuk, S. B., Khrenov, M. O., Novoselova, T. V., Lunin, S. M., Fesenko, E. E. and Novoselova, E. G. (2013) Inhibitors of TLR-4, NF-kappaB and SAPK/JNK signaling reduce the toxic effect of lipopolysaccharide on RAW 264.7 cells. J. Immunol. 286, 133-140.

Guhu, M. and Mackman, N. (2001) LPS induction of gene expression in human monocytes. Cell. Signal. 13, 85-94.

Guo, X., Gerl, R. E. and Schranner, J. W. (2003) Defining the involvement of p38alpha MAPK in the production of anti- and proinflammatory cytokines using an SB 203580-resistant form of the kinase. J. Biol. Chem. 278, 22237-22242.

Han, Y. M., Lee, S. K., Jeong, D. G., Ryu, S. E., Han, D. C., Kim, D. K. and Kwon, B. M. (2012) Emodin inhibits migration and invasion of DLD-1 (PRL-3) cells via inhibition of PRL-3 phosphatase activity. Biogro. Med. Chem. Lett. 22, 323-326.

Hayden, M. S. and Ghosh, S. (2004) Signaling to NF-kappaB. Genes. Dev. 18, 2195-2224.

Heo, S. K., Yun, H. J., Park, W. H. and Park, S. D. (2008) Emodin inhibits TNF-alpha-induced human aortic smooth muscle cell proliferation via caspase- and mitochondrial-dependent apoptosis. J. Cell. Biochem. 105, 70-80.

Hwangbo, K., Zheng, M. S., Kim, Y. J., Im, J. Y., Lee, C. S., Woo, M. H., Jahng, Y., Chang, H. W. and Son, J. K. (2012) Inhibition of DNA topoisomerases I and II of compounds from Reynoutria japonica. Arch. Pharm. Res. 35, 1583-1589.

Hwang, J. K., Noh, E. M., Moon, S. J., Kim, J. M., Kwon, K. B., Park, B. H., You, Y. O., Hwang, B. M., Kim, H. J., Kim, B. S., Lee, S. J., Kim, J. S. and Lee, Y. R. (2013) Emodin suppresses inflammatory responses and joint destruction in collagen-induced arthritic mice. Rheumatology (Oxford) 52, 1583-1591.

Ishizuka, T., Terada, N., Genxins, P., Hamelman, E., Oshiba, A., Fanger, G. R., Johnson, G. L. and Gelfand, E. W. (1997) Mast cell tumor necrosis factor alpha production is regulated by MEK kinases. Proc. Natl. Acad. Sci. USA 94, 6359-6363.

Jeong, H. J., Chung, H. S., Lee, B. R., Kim, S. J., Yoo, S. J., Hong, S. H. and Kim, H. M. (2003) Expression of proinflammatory cytokines via NF-κB activation and NF-kappaB via a natural product, se.

Jenong, G. S., Kim, Y. C., Shin, D. W. and Kwon, D. Y. (2010) Synthesis of receptor activator of NF-κB ligand (RANKL) by Fluegge. J. Mol. Biochem. Pharmacol. 1. 4, 1285-1290.

Li, Y. Q., Zhang, X. Z., Xu, Y. J., Ni, W., Chen, X. S., Yang, Z. and Ma, D. (2006) N-Acetyl-L-cysteine and pyrrolidine dithiocarbamate inhibited nuclear factor-kappaB activation in alveolar macrophages by different mechanisms. Acta Pharmacol. Sin. 27, 339-346.

Lim, B. O., Lee, K. H., Cho, S. H., Park, W. H., Kim, D. K., Kim, J. D., Kim, B. K., Kim, H. S., Her, E., Lee, H. Y. and Choi, W. S. (2007) Polyglyco cuspid in radix inhibits the activation of Syk kinase in mast cells for antiallergy activity. Exp. Biol. Med. (Maywood) 232, 1425-1431.

Lin, S. Z., Chen, K. J., Tong, H. F., Jing, H., Li, H. and Zheng, S. S. (2010) Emodin attenuates acute rejection of liver allografts by in-

http://dx.doi.org/10.4062/biomolther.2013.068
hobiting hepatocellular apoptosis and modulating the Th1/Th2 balance in rats. *Clin. Exp. Pharmacol. Physiol.* **37**, 790-794.

Lu, Y., Hwang, S. L., Son, J. K. and Chang, H. W. (2013) Manansattin B isolated from Saururus chinensis inhibits cyclooxygenase-2-dependent prostaglandin D2 generation by blocking cys-mediated nuclear factor-kappaB and mitogen activated protein kinase pathways in bone marrow derived-mast cells. *Biol. Pharm. Bull.* **36**, 1370-1374.

Lu, Y., Suh, S. J., Li, X., Hwang, S. L., Li, Y., Hwangbo, K., Park, S. J., Murakami, M., Lee, S. H., Jahng, Y. S., Son, J. K., Kim, C. H. and Chang, H. W. (2012) Citreorosein, a naturally occurring anthraquinone derivative isolated from Polygoni cuspidati radix, attenuates cyclooxygenase-2-dependent prostaglandin D2 generation by blocking Akt and JNK pathways in mouse bone marrow-derived mast cells. *Food Chem. Toxicol.* **50**, 913-919.

Lu, Y., Yang, J. H., Li, X., Hwangbo, K., Hwang, S. L., Taketomi, Y., Murakami, M., Chang, Y. C., Kim, C. H., Son, J. K. and Chang, H. W. (2011) Emodin, a naturally occurring anthraquinone derivative, suppresses IgE-mediated anaphylactic reaction and mast cell activation. *Biochem. Pharmacol.* **82**, 1700-1708.

Meng, G., Liu, Y., Lou, C. and Yang, H. (2010) Emodin suppresses lipopoly saccharide-induced pro-inflammatory responses and NF-kappaB activation by disrupting lipid rafts in CD14-negative endothelial cells. *Br. J. Pharmacol.* **161**, 1626-1644.

Metcalf, D. D., Baram, D. and Mekori, Y. A. (1997) Mast cells. *Physiol. Rev.* **77**, 1033-1079.

Moon, P. D., Jeong, H. J., Kim, S. J., An, H. J., Lee, H. J., Yang, W. M., Park, S. K., Hong, S. H., Kim, H. M. and Um, J. Y. (2007) Use of electrode configuration and a sphingosine kinase 1 inhibitor at-
tenuates airway hyperresponsiveness and inflammation in a mast cell-dependent murine model of allergic asthma. *J. Allergy Clin. Immunol.* **131**, 501-511.

Rasheed, Z., Akhtar, N., Anbazhagan, A. N., Ramamaruthy, S., Shukla, M. and Haqqi, T. M. (2009) Polyphenol-rich pomegranate fruit extract (POMx) suppresses PMACI-induced expression of pro-inflammatory cytokines by inhibiting the activation of MAP Kinases and NF-kappaB in human KUB12 cells. *J. Inflamm. (Lond)*. **6**, 1.

Senftleben, U., Cao, Y., Xiao, G., Greten, F. R., Krahm, G., Bonizzi, G., Chen, Y., Hu, Y., Fong, A., Sun, S. C. and Karin, M. (2001) Activation by IKKalpha of a second, evolutionarily conserved, NF-kappa B signaling pathway. *Science* **293**, 1495-1499.

Singal, I., Zacharatos, P., Toumpanakis, D., Michaillidou, T., Nousia, O., Theocharis, S., Roussos, C., Papapetropoulos, A. and Vassiliakopoulos, T. (2011) MAPKs and NF-kappaB differentially regu-
tulate cytokine expression in the diaphragm in response to resistive breathing: the role of oxidative stress. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* **300**, R1152-1162.

Song, P., Kim, J. H., Ghim, J., Yoon, J. H., Lee, A., Kwon, Y., Hyun, H., Moon, H. Y., Choi, H. S., Berggren, P. O., Suh, P. G. and Ryu, S. H. (2013) Emodin regulates glucose utilization by activating AMP-activated protein kinase. *J. Biol. Chem.* **288**, 5732-5742.

Suh, J. and Rabson, A. B. (2004) NF-kappaB activation in human prostate cancer: important mediator or epiphenomenon? *J. Cell. Biochem.* **91**, 100-117.

Takada, Y., Mukhopadhyay, A., Kundu, G. C., Mahabekshaw, G. H., Singh, S. and Aggarwal, B. B. (2003) Hydrogen peroxide activates NF-kappa B through tyrosine phosphorylation of I kappa B alpha and serine phosphorylation of NF-kappaB. *J. Cell. Physiol.* **201**, R1152-1162.

Tuzen, H. and Cantrell, D. A. (1997) Distinct Ras effector pathways are involved in Fe epsilon R1 regulation of the transcriptional activity of Cdk-1 and NF-kappaB in mast cells. *J. Exp. Med.* **185**, 43-53.

Tzeng, T. F., Lu, H. J., Liou, S. S., Chang, C. J. and Liu, I. M. (2012) Emodin protects against high-fat diet-induced obesity via regulation of AMP-activated protein kinase pathways in white adipose tissue. *Planta Med.* **78**, 943-950.

Wang, W., Zhou, Q., Liu, L. and Zou, K. (2012) Anti-allergic activity of emodin on IgE-mediated activation in RBL-2H3 cells. *Pharmacol. Rep.* **64**, 1216-1222.

Xu, C., Shen, G., Chen, C., Gelinas, C. and Kong, A. N. (2005) Suppression of NF-kappaB and NF-kappaB-regulated gene expression by sulforaphane and PEITC through IkappaBalpha, IKK pathway in human prostate cancer PC-3 cells. *Oncogene* **24**, 4486-4495.