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Neurosphere Based Differentiation of Human iPSC Improves Astrocyte Differentiation

Shuling Zhou,1 Karolina Szczesna,2 Anna Ochalek,2 Julianna Kobolák,2 Eszter Varga,2 Csilla Nemes,2 Abinaya Chandrasekaran,2 Mikkel Rasmussen,4 Susanna Cirera,1 Poul Hyttel,1 András Dinnyés,2,3 Kristine K. Freude,1 and Hasan X. Avci2

1Department of Veterinary Clinical and Animal Science, Faculty of Health and Medical Sciences, University of Copenhagen, 1870 Frederiksberg, Denmark
2BioTalentum Ltd., Gödöllő 2100, Hungary
3Molecular Animal Biotechnology Laboratory, SZIE, Gödöllő 2100, Hungary
4Bioneer A/S, 2970 Hørsholm, Denmark

Correspondence should be addressed to Kristine K. Freude; kkf@sund.ku.dk and Hasan X. Avci; hasan.avci@biotalentum.hu

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1. Introduction

Neural progenitor cells (NPCs) are self-renewing and multipotent cells, which are found in the embryonic, fetal, and adult mammalian central nervous system (CNS). These cells have the capacity to differentiate into neurons as well as glia cells [1, 2]. NPCs derived from pluripotent stem cells (iPSCs), including embryonic stem cells (ESCs) and induced pluripotent stem cells (iPSCs), are an attractive in vitro model for studying the pathology of CNS disorders, for drug development and identification of novel drug targets [3, 4]. Compared with their in vivo counterparts NPCs are not as limited in regard to their proliferative status and differentiation capacity into various neural phenotypes in vitro [5–8]. NPCs are commonly isolated from different regions of murine and human brain by microdissection and subsequently cultured as free-floating aggregates using both classic and stirred suspension 3D culture system methods [9, 10]. It is common practice in the field of neuroscience and stem cell research to maintain and proliferate NPCs by using either two-dimensional (2D) adherent monolayer or three-dimensional (3D) floating neurospheres. Cells derived from the 3D culture system are thought to be more representative of the spatial cellular environment found in living organisms, including features of tissue-specific architecture, mechanical and biochemical cues, and cell-cell communication [11]. In accordance, neurospheres are widely accepted and used as in vitro assays to analyze the properties of NPCs [12]. This spatial integrity is not found in the 2D...
culturing system, which is considered to be the more artificial culturing technique [11].

A common approach of human NPCs derivation from iPSC involves neural induction by inhibition of SMAD signaling by means of two inhibitors (SB431542 and Noggin or LDN193189), followed by expansion of NPCs and subsequent terminal differentiation into neurons using the 2D culture system [8, 13, 14]. Yet, in order to model specific neurodegenerative diseases in vitro it is crucial that the culture methods display the desired regional and subtype specificity compared to the affected neurons of the patient. Consequently, disease modeling in 3D tissue culture systems has recently been successfully applied in Alzheimer’s disease [15] and Parkinson’s disease [16, 17] and to study glia cell differentiation [18, 19].

The human brain is made up of various subtypes of neurons but also by a substantial amount of glia cells (more than 50%) [20]. One subtype of glia cells is astrocytes, which play a complex and an essential role in neural maturation and homeostasis, including synaptic transmission and information processing by neural circuit functions [21]. Both neurons and glia cells, except for microglia, are derived from radial glia (RG) cells in the developing brain. RG cells are a NPC population, which originates from neuroepithelial cells the neural tube [5, 22]. During neurogenesis, 5/6 of RG cells divide asymmetrically into early bipolar intermediate progenitor (IP) cells which eventually differentiate into neurons. The remaining 1/6 of RG cells give rise to astrocyte and oligodendrocyte progenitor cells [23–25]. The differentiation from RG cells to early IP cells is accompanied by the loss of PAX6 expression [23]. Brain lipid-binding protein (BLBP) is a verified astrocyte progenitor marker, which was detected by following the expression pattern of brain BLBP in RG cells [20, 26]. Later, during development BLBP expression becomes restricted to astrocyte progenitors and downregulated in astrocytes [27]. One of the most commonly used astrocyte markers is glial fibrillary acidic protein (GFAP), which is expressed during CNS development and becomes restricted to astrocytes lineage [20]. Paired box 6 (PAX6) is restricted to astrocyte progenitors and downregulated in astrocytes [27].

Another aspect of neuronal differentiation, which may be a challenge under in vitro conditions, is the extended time frame (42–84 days) for achieving functional neuronal maturation [32, 33]. This can be accelerated by coculturing neurons with astrocytes. This makes astrocyte differentiation protocols highly desirable and needed for the neural maturation process [34, 35]. One of the main issues is that differentiation of astrocytes from fetal or adult postmortem CNS has been proven to be a difficult process with low efficiencies [36, 37]. Traditional 2D methods to generate sufficiently pure population of astrocytes derived from iPSCs and ESCs are on the other hand very time consuming (>180 days) [38]. Consequently, reliable 3D based differentiation methods, which can potentially enrich and accelerate astrocyte differentiation and maturation, would be beneficial in order to improve coculturing approaches.

In the present study, we describe a potentially efficient 3D method of astrocyte enrichment from human iPSC-derived NPCs. The method progresses through an initial phase of NPC formation with increasing expression of PAX6 and NESTIN, which are NPC markers. Furthermore, we directly established a link between the expression of BLBP, PAX6, and astrocyte differentiation efficiency [27]. Our method provides an NPCs expansion protocol in 3D, which enriches the high PAX6 expression NPCs from mixed low and high PAX6 expression NPC pool and could be beneficial for astrocyte differentiation in which the efficiency can be monitored via GFAP and AQP4 expression.

2. Materials and Methods

2.1. 2D Monolayer Culture of NPCs and Terminal Differentiation. The three human iPSC lines used in this study were generated by using the Sendai virus (CytoTune-ips 2.0 Sendai Reprogramming Kit) (Life Technologies, Carlsbad, California, USA). Three cell lines were investigated in this study: clone N S5 and N S8 derived from a healthy 34-year-old male donor and clone RT S11 derived from a 33-year-old healthy female donor. NPCs cell lines were induced from each of these human iPSCs by dual inhibition of SMAD signaling with 10 μM SB431542 (Sigma-Aldrich, St. Louis, Missouri, USA) and 100 ng/mL Noggin (R&D Systems, Minneapolis, Minnesota, USA). NPCs were expanded in neural maintenance medium and maintained on plates coated with poly-L-ornithine (Sigma-Aldrich) and laminin (Roche, Indianapolis, Indiana, USA) (POL/L). Terminal differentiation on POL/L coated plates was initiated using neural differentiation medium as described in the Media. Total time of terminal differentiation was 21 days.

2.2. 3D Neurospheres Culture of NPCs and Terminal Differentiation. Monolayer NPCs from all three lines (N S5, N S8, and RT S11) were dissociated into single cells with Accutase (Sigma-Aldrich). A total of 1 × 10^6 cells/mL were plated on the low attachment dish (Sarstedt, Newton, Massachusetts, USA) in neural maintenance medium and maintained for 7 days as neurosphere cultures. Neurospheres were visualized in an inverted microscope under phase contrast mode. Three representative pictures of neurospheres were taken from 3 independent cultures with an inverted microscope under phase contrast mode (approximately 100 neurospheres were counted for each line, 4x magnification; OLYMPUS CKX41, Tokyo, Japan). Subsequently, the neurospheres were dissociated into single cells using Accutase and next seeded on plates coated with POL/L. Finally, the cells were differentiated for 21 days in neural differentiation medium.

2.3. Media

2.3.1. Neural Maintenance Medium (NMM). NMM consisted of a 50/50 mixture of DMEM/F-12 with GlutaMAX (Life Technologies) and neurobasal medium (Life Technologies).
This basic media was supplemented with N2 (Life Technologies), B27 (Life Technologies), 2 mM L-Glutamine (Sigma-Aldrich), 10 ng/mL basic fibroblast growth factor (bFGF) (Life Technologies), 10 ng/mL epidermal growth factor (EGF) (Life Technologies), 1x nonessential amino acid (Sigma-Aldrich), and 50 U/mL penicillin/streptomycin (Life Technologies).

2.3.2. Neural Differentiation Medium (NDM). NDM was composed of a 50/50 mixture of DMEM/F-12 with GlutaMAX and neurobasal medium. This basic media was supplemented with N2, B27, 2 mM L-Glutamine, 1x nonessential amino acid, 0.2 mM ascorbic acid (Sigma-Aldrich), 25 μM 2-mercaptoethanol (Life Technologies), and 1x nonessential amino acid (Sigma-Aldrich), 10 ng/mL epidermal growth factor (EGF) (Life Technologies), 1x nonessential amino acid (Sigma-Aldrich), and 50 U/mL penicillin/streptomycin (Life Technologies).

2.4. Flow Cytometry. In order to perform flow cytometry analysis, NPCs harvested from 2D monolayer and 3D neurosphere were dissociated into single-cell suspension with Accutase. Dissociated cells were fixed with 4% paraformaldehyde (PFA) for 20 min at room temperature (RT) and subsequently permeabilized with 0.2% Triton X-100 for 20 minutes. Cells were stained for 1 hour at RT with Alexa Fluor 647 mouse anti-NESTIN and PE mouse anti-human Pax6 antibodies (BD Pharmingen, San Diego, California, USA). Flow cytometry analysis was performed using a “Flow Cytometer Cytomics FC 500” (Beckman Coulter, Pasadena, California, United States). A red solid state laser 635 nm and an argon laser 488 nm were used to detect NESTIN and Pax6 expression in NPCs. Subsequently, we set the specific gate to calculate the proportion of low Pax6 expressing NPCs from 2D and 3D culturing system. Data was analyzed using FlowJo software (version 7.6.5) from 3 independent cultures.

2.5. Immunocytochemistry (ICC), Imaging, and Quantification. To analyze neural markers cells were fixed in 4% PFA for 20 minutes at RT, washed in phosphate buffered saline (PBS), and permeabilized with 0.2% Triton X-100 diluted in PBS for 20 minutes. Afterwards, cells were blocked in 3% BSA + 0.2% Triton X-100 in PBS for 1 hour at RT. NPCs were incubated overnight at 4°C with the following primary antibodies: rabbit anti-PAX6 (Covance, Princeton, New Jersey, USA 1:250), mouse anti-NESTIN (Merck Millipore, Temecula, California, USA, 1:1000), rabbit anti-Musashi-1 (Merck Millipore, 1:400), mouse anti-SOX2 (R&D System, 1:100), and mouse anti-Ki67 (Santa Cruz, Dallas, Texas, USA 1:800). Neurospheres were incubated overnight at 4°C with rabbit anti-PAX6 (Covance, 1:250) and mouse anti-NESTIN (Merck Millipore, 1:1000). Differentiated cells were incubated overnight at 4°C with mouse anti-MAP2 (Merck Millipore, 1:1000), rabbit anti-beta-III tubulin (Covance, 1:1000), mouse anti-beta-III tubulin (Santa Cruz, 1:1000), rabbit anti-AQP4 (Santa Cruz, 1:50), and rabbit anti-GFAP (DAKO, Glostrup, Denmark 1:1000). The primary antibodies were detected using the following fluorescent labeled secondary antibodies: Alexa Fluor 488 donkey anti-rabbit IgG (H + L), Alexa Fluor 594 donkey anti-mouse IgG (H + L), Alexa Fluor 488 donkey anti-mouse IgG (H + L), and Alexa Fluor 594 donkey anti-rabbit IgG (H + L) (all from Life Technologies). Finally, the cells were incubated for 20 min at RT with 4',6-diamidino-2-phenylindole (DAPI) at a final concentration of 0.2 μg/mL diluted in PBS to detect nuclei counterstaining. Unspecific binding of the secondary antibodies was excluded by using only secondary antibody controls. Cells were analyzed under fluorescent microscope equipped with 3D imaging module (Axiolmager system with ApoTome, Carl Zeiss MicroImaging GmbH, and Jena, Germany) and controlled by AxioVision 4.8.1 Microscope software (Carl Zeiss MicroImaging GmbH, Jena, Germany). The number of proliferative cells (Ki67+) was quantified. The numbers of neurons (TUBB3+) and astrocytes (GFAP+ and AQP4+) were quantified. Five pictures per coverslip were acquired from 3 independent cultures (20x magnification). On average, approximately 300 cells were counted for each picture.

2.6. Quantitative Real-Time Polymerase Chain Reaction (qPCR). Total RNA was isolated from cultured cells using the RNasyPlus Mini kit (Qiagen, Venlo, Limburg, Netherlands). Two micrograms of total RNA was transcribed to cDNA using the SuperScript VILO cDNA Synthesis kit (Life Technologies) in accordance with the manufacturer’s instructions. qPCR reactions were performed using LightCycler 480 SYBR Green I (Roche) on a LightCycler 480 real-time PCR machine (Roche). In this study, the house-keeping gene GAPDH was used for data normalization. Samples were collected from 3 independent cultures. Data was analyzed based on 2^(-ΔΔCt) method [39]. All used primers for qPCR experiments are summarized in Table 1.

2.7. Statistics. Distribution of neurosphere diameter was tested using Kolmogorov-Smirnov normality test. A one-way analysis of variance (ANOVA) with a Tukey post hoc multiple comparison test was applied to compare the neurosphere diameter across all studied cell lines at days 4 and 7. Distribution of relative abundance of mRNA from the different genes was examined by using D’Agostino and Pearson omnibus normality test. Student’s t-test was performed to compare the difference between groups. Pearson’s correlation tests were performed to establish the correlation between the following: (1) the relative mRNA abundance of PAX6 in 3D neurosphere NPCs and neurosphere diameter at day 7; (2) the relative mRNA abundance of BLBP and PAX6 in NPCs and GFAP in differentiated cells over three studied cell lines. Distribution of proportion of Ki67+, low PAX6 expression, TUBB3+, AQP4+, and GFAP+ cells was tested using Kolmogorov-Smirnov normality test. One-way ANOVA with a Tukey post hoc multiple comparison test and Student’s t-test was performed to compare the difference between groups. A p value below 0.05 was considered significant. Results are reported as the Mean ± SEM (standard error of the mean). Data was analyzed using OriginPro Statistical software (version 9).

3. Results

3.1. Schematic Work Flow of 2D Monolayer Expansion, 3D Neurosphere Aggregation, and Terminal Differentiation of
Table 1: Human specific primers used for qPCR analysis.

| Symbol | Sequence 5'-3' | Product size (BP) | Gene name | Reference number (NCBI) | Reference |
|--------|----------------|-------------------|-----------|------------------------|-----------|
| GAPDH  | GGTGCCAGGGCTGCTTTTA GGATCTCGCTCTCGAAGATG | 195 | Glyceraldehyde-3-phosphate dehydrogenase | NM_001289746.1 | [40] |
| NESTIN | CTCCAGAAACTCAAGCACC TCTGTATCTCTTCCTCA | 145 | Nestin | NM_006617.1 | [41] |
| SOX2   | TTCATATGTCGCCAGCCTACCAGA TCACATGTGTAGAGGGAGCTGTC | 80 | SRY- (sex determining region Y-) box 2 | NM_003106.3 | [42] |
| PAX6   | TGGATATCTCCTCCCTCCT TAAGGATGGTGAAGGGGACAG | 126 | Paired box 6 | XM_00522598.3 | [43] |
| BLBP   | GCCATTTGCGAATGGGAGTCACAGGTATCTCTTTTTCGTA | 76 | Brain lipid-binding protein | XM_005266858.2 | [44] |
| TUBB3  | TCCGCTAGGGGCTTTGGGAC GCTCCGCCCCTGGTGCAG | 108 | Tubulin, beta 3 class III | NM_01197181.1 | |
| TBR2   | CACCGCCACAAACTGAGAT CGAACACAATGGGAGTCGAG | 109 | Eomesoderm | NM_001278181.1 | [45] |
| AQP4   | AGCATGTACGGGATTTCT TCTGTCCGACCCGAGTGGAT | 81 | Aquaporin 4 | XM_001278181.1 | [44] |
| MAP2   | AAGTTCAACGACCCTTCCTCCT GCCGAGGAATGGATTCCCAGA | 127 | Microtubule-associated protein 2 | XM_001278181.1 | [44] |
| GFAP   | AGGTCCATGGGGCTTTGGAC GCCATTTGCGAATGGGAGTCACAGGTATCTCTTTTTCGTA | 82 | Glial fibrillary acidic protein | NM_001242376.1 | [44] |

Human Neural Progenitor Cell Cultures (NPCs). In order to investigate whether 3D culturing methods can enhance astrocyte differentiation from NPCs in vitro, we set up a direct comparison of 2D and 3D NPC expansion methods (Figure 1).

3.2. 3D Cultured NPCs Form Proliferating Neurospheres Positive for PAX6 and NESTIN. We assessed the morphology of day 4 and 7 neurospheres by phase contrast imaging and their expression of early neural lineage markers (PAX6 and NESTIN) by immunocytochemistry. Neurosphere size was evaluated at days 4 and 7. We observed varying sizes of neurospheres within the different cell lines analyzed (Figure 2(a)). As shown in Figure 2(b), the diameter of neurospheres from NPC line N S8 (80.82 ± 6.43 μm) was larger than N S5 (74.26 ± 5.45 μm) and RT S11 (65.05 ± 6.18 μm) at day 4. Even though we observed significant differences amongst cell lines, all of them were proliferative as neurospheres and increased in diameter during culturing. At day 7 the diameter of neurospheres was significantly increased in all lines (N S5 = 129.46 ± 11.11 μm, N S8 = 168.35 ± 10.38 μm, and RT S11 = 98.57 ± 8.37 μm; p < 0.01) compared to day 4. One cell line (N S8) showed a significant larger diameter at day 7 compared to N S5 and RT S11 (p < 0.01). All lines analyzed (N S5, N S8, and RT S11) expressed NESTIN and PAX6 (Figure 2(c)), confirming NPC marker expression in the neurospheres. Additionally, to assess the proliferation capacity of NPCs expanded by the 2D method, we stained for the proliferation marker Ki-67. As shown in Figures 2(d) and 2(e), we observed significantly higher proportion of NPCs in N S8 which were positive for Ki-67 (Ki67) in comparison to N S5 and RT S11 (p < 0.01). This result demonstrates that NPCs maintain the capacity to proliferate in 2D and 3D culturing system. Furthermore, 3D suspension culture induces the formation of neurospheres with the expression of NPC markers.

3.3. Comparison of NPC Markers Expression in 2D and 3D Cultures. ICC was performed to examine the expression level of NPC markers (PAX6, NESTIN, Musashi-1, and SOX2) in 2D and 3D NPCs. Both were dissociated and plated as single cells on day 7. As expected, 2D and 3D NPCs were positive for NESTIN and PAX6 expression (Figure 3(a)). Furthermore ICC was performed using SRY- (sex determining region Y-) box 2 (SOX2) and Musashi RNA-binding protein 1 (Musashi-1), which label self-renewing NPCs in the nucleus (SOX2) and cytoplasm (Musashi-1) (Figure 3(b)). Interestingly, PAX6 expression was heterogeneous in 2D NPC cultures. Some areas showed low PAX6 expression (encircled with dashed line), whilst other areas showed high PAX6 expression (arrows) (Figure 3(a)). In contrast the expression pattern of PAX6 in 3D dissociated and plated NPCs was more uniformly distributed (Figure 3(a)). Our results show that NPCs cultured in both 2D and 3D culture system express NPC markers.

3.4. NPCs in 3D Neurospheres Showed Increased Homogenous PAX6 Expression. Flow cytometry was implemented to study the expression level of PAX6 and NESTIN in 2D and 3D NPCs. The intensity of the PAX6 signal in the FACS analyses revealed two apparently divergent populations in 2D NPCs whilst 3D NPCs showed only one distinct population.
The appearance of two individual populations was most prominent with the 2D NS8 cells (Figure 4(a)). Subsequently, specific gating was applied to compare the proportion of low PAX6 expression NPCs in 2D and 3D culturing system. Our results presented in Figure 4(b) show significant decrease of low PAX6 expression NPCs in 3D culturing system. We further analyzed all cell lines used for the two culture systems by qPCR to investigate the expression of NPC markers (SOX2, PAX6, and NESTIN). The abundance of PAX6 and NESTIN was significantly increased (p < 0.01) in 3D cultured NPCs in comparison with 2D cultured NPCs in all investigated cell lines (Figure 4(c)). However, we were not able to detect similar expression tendencies for SOX2 using the 3D culture system. Pearson's correlation was used to evaluate the relationship between the NPC marker PAX6 and neurosphere diameter. Day 7 neurosphere diameter had strongly positive (Pearson’s r = 0.68, p = 0.043) correlation with mRNA abundance of PAX6 in 3D cultured NPCs (Figure 4(d)). This indicates a direct correlation between PAX6 expression and neurosphere diameter. Taken together, we observe that within the 3D culturing system NPCs displaying low PAX6 expression become eliminated and subsequently NPC marker expression (PAX6 and NESTIN) is increased.

3.5. Neurosphere Aggregation Promotes Astrocyte Differentiation. The 2D and 3D NPCs were dissociated and differentiated on POL/L plates for 21 days. ICC analysis showed that the majority of cells derived from NPCs cultured in the two different systems were positive for neuronal markers (tubulin, beta 3 class III (TUBB3), and microtubule-associated protein 2 (MAP2); Figures 5(a) and 5(b)). A subset of cells was positive for GFAP expression, specific for astrocytes, and showed the proportion of GFAP positive cells was higher in NPCs derived from 3D versus 2D cultures, primarily observed in NS5 and RT SII lines (p < 0.01) (Figure 5(c)). We also evaluated the abundance of neuronal and astrocyte marker expression via qPCR in cells derived from 2D and 3D NPCs, which have been differentiated for 21 days. At this time point a subset of 3D derived cells expressed higher levels of GFAP compared to 2D derived cells (Figure 5(d)). Subsequently, to confirm that 3D culture system promotes astrocyte differentiation we evaluate the expression of another astrocyte marker aquaporin 4 (AQP4). As shown in Figures 6(a) and 6(b), significant higher proportion of AQP4 positive cells were observed in cells differentiated from 3D cultured NPCs (p < 0.01). Furthermore, qPCR was performed to verify the ICC quantification results. Significantly higher expression of AQP4 was observed in cells differentiated from 3D culturing method (Figure 6(c)) (p < 0.01). Together these data indicate that neurosphere aggregation could promote astrocyte differentiation. Similar expression patterns of MAP2 and TUBB3 and the population of TUBB3 positive neurons were detected in neurons derived from NPCs using both culture systems (Figures 5(c) and 5(d)). This implies that there is no apparent difference in neuronal maturation between the two methods. The correlation between the relative expression abundance of BLBP and PAX6 in NPCs and GFAP in 21-day differentiated cells was evaluated using the Pearson correlation analysis (Figures 5(e) and 5(f)). Pearson correlation analysis demonstrated that the expression of astrocyte progenitor marker BLBP and NPC marker PAX6 during NPCs stage has a significant positive correlation (r = 0.738, p = 4.59 × 10^-4 and r = 0.703, p = 1.12 × 10^-3) to the expression of astrocyte marker (GFAP) in the astrocytes derived from all studied NPCs lines. Therefore, BLBP and PAX6 expression during NPC stages can be used as an indicator for potential differentiation into the astrocyte lineage as verified by GFAP expression. Furthermore, we performed qPCR analysis to prove that the increase of PAX6 expression and the promotion of astrocyte differentiation were due to the loss of cells with low PAX6 expression during neurosphere formation. These low PAX6 expressing cells are considered early IP cells. Our qPCR results demonstrated a significant decrease of expression of the IP cell marker ceme-sodermin (TBR2) in cells differentiated from 3D cultured NPCs compared to 2D cultured NPCs (Figure 6(c)) (p < 0.01). Here it can be concluded that the 3D culture method might promote the astrocytes differentiation and provoke an inhibition of the IP cells generation.
Figure 2: 3D cultured NPCs form proliferating neurospheres positive for PAX6 and NESTIN. (a) Phase contrast images of neurospheres from all 3 lines (NS5, NS8, and RT S11) on day 4 and day 7. Scale bar = 100 μm. (b) Analyses of neurosphere diameter on day 4 and day 7 of 3D suspension culture. Significant difference was tested via one-way ANOVA analysis with a Tukey post hoc multiple comparison test. Significant differences were indicated as follows: *p < 0.05 and **p < 0.01. Results were reported as Mean ± SEM of 9 fields from 3 independent cultures. (c) ICC staining of neurospheres (NESTIN, red, and PAX6, green) on day 7 from all 3 lines. Scale bar = 100 μm. (d) Representative ICC of 2D NPCs (KI67). Scale bar = 100 μm. (e) Quantification of KI67+ NPCs for 2D culturing systems. Results were reported as Mean ± SEM of 15 fields from 3 independent cultures, tested for significant differences (**p < 0.01) via one-way ANOVA analysis with a Tukey post hoc multiple comparison test.
4. Discussion

The present study describes the fate and cellular properties of human NPCs derived from human iPSC and cultured as monolayers (2D) and neurospheres (3D). Human NPCs derived from iPSCs have traditionally been maintained and proliferated in 2D culture system, demonstrating an efficient differentiation into the neuronal lineage [13,14]. However, the 2D culture systems are poor in recapitulating the spatially well-organized intercellular relationships characteristic of neural in vivo development in the CNS. In contrast, the neurospheres generated from 3D free-floating aggregates of NPCs with a certain spatial degree of complexity better mimic the main features of brain tissue and can therefore be considered as more relevant in vitro models [46]. Thus, neurospheres as a mean to produce neurons and astrocytes could provide a reliable in vitro cellular model for understanding CNS disorders and can play a relevant role in cell-based drug screening [11, 15, 17]. Previously, other groups have described the generation of neurospheres from brain-derived NPCs [47, 48]. We have employed the same method to generate neurospheres from iPSC-derived NPCs making the methodology much more versatile for implementation for in vitro cell modeling. Our findings show that the neurosphere diameter increases from day 4 to day 7, which indicates the ability of cell proliferation within the 3D culture system. Further, ICC characterization showed a very uniform expression of NPC markers in neurospheres, such as PAX6 and NESTIN. We also monitored the proliferation capacity of 2D NPCs via ICC for KI67 and observed a higher proportion of KI67 positive cells in N S8 compared to N S5 and RT S11. The proliferative capacity of 2D NPCs is directly correlated to the size of the neurospheres in the 3D system. Therefore, highly proliferative lines in the 2D system generate larger neurospheres in the 3D system. It might be that these neurospheres are proliferative NPCs, similar to brain-derived NPCs. In the present study, we further compared the NPC identity and differentiation ability of NPCs cultured in 2D and 3D culturing systems. We chose PAX6, NESTIN, SOX2, and Musashi-1 (NPC markers) to evaluate the neural identity of our 2D and 3D NPCs. PAX6 is uniformly expressed in early neural ectoderm cells of human fetuses and in NPCs differentiated from human ESCs [49]. Pan-neuronal markers (SOX2, NESTIN, and Musashi-1) are widely expressed in all NPCs [50–53]. Some groups also found that PAX6, NESTIN, and SOX2 are expressed in iPSC-derived NPCs [8, 13, 54]. Moreover, PAX6, NESTIN, SOX2, and Musashi-1 are also the RG cell markers [55, 56]. Our results demonstrated that both 2D and 3D NPCs expressed these NPC markers, but PAX6 and NESTIN expression levels were significantly higher in 3D NPCs. This indicates that neurosphere aggregation promotes PAX6 and NESTIN expression. Furthermore, a more homogenous PAX6 expression was detected in 3D NPC populations suggesting that an uneven distributed PAX6
Figure 4: Continued.
expression pattern in 2D NPCs was avoided by neurospheres aggregation. PAX6 is an established NPC marker expressed in RG cells which plays an essential role in maintaining the NPC population, neuronal and glial differentiation [29]. Previous study has shown that neurospheres are largely composed of RG cells (high PAX6 expression) other than early IP cells (low PAX6 expression) [57]. As a consequence NPC with low expression levels of PAX6 cannot form neurospheres. Consistent with this study, our results indicate that we could enrich the high PAX6 expression cells from mixed low and high PAX6 expression NPCs pool by using 3D culturing method. Furthermore, expression of PAX6 induces production of larger neurospheres [58]. Consistent with this study, our results showed that expression of PAX6 in 3D NPCs was positively correlated to subsequent neurosphere diameters at day 7.

Herein, we demonstrated that human iPSC-derived NPCs cultured under 2D and 3D conditions differentiated into cells positive for mature neuronal markers (MAP2 and TUBB3) and the astrocyte markers (GFAP and AQP4). The overall process of directing differentiation of human iPSCs-derived NPCs to neurons and astrocytes in vitro has been...
Figure 5: Continued.

(a) DAPI, TUBB3, GFAP, Merge

(b) DAPI, MAP2, TUBB3, Merge

(c) Proportion of positive cells

Differentiated cells (derived from 2D NPCs)
Differentiated cells (derived from 3D NPCs)

Relative amount of mRNA

Figure 5: Continued.
described earlier [59]. As a novel finding, our experiments demonstrated that propagation under the 3D conditions resulted in a significantly higher yield of cells with GFAP and AQP4 expression compared to the 2D condition. Hence, we draw a conclusion that 3D NPCs culturing methods stimulate astrocyte differentiation. This observation is in agreement with previously described findings, which show efficient 3D differentiation of ESC-derived NPCs to glial cells of the peripheral nervous system, in particular Schwann cells [18]. Moreover, we demonstrated that the GFAP expression in astrocytes was significantly positively correlated to the astrocyte progenitor marker BLBP expression in NPCs, which is in accordance with observations of other groups [20, 26, 27]. Furthermore, it has been shown that abolishing the expression of PAX6 leads to the downregulation of GFAP expression, resulting in inhibition of astrocyte maturation [30, 60].
Similar to these studies, we found that the GFAP expression in astrocytes was strongly and positively associated with PAX6 expression pattern during the NPC stage. These correlation studies underlie the importance of BLBP and PAX6 expression in NPCs for differentiation towards an astrocyte fate.

Single-cell mRNA profiling indicates that neurospheres are composed of RG cells (high PAX6 expression) other than early IP cells or IP cells (low PAX6 expression) [57]. Similar to this study, we observed that there was a significant decrease of expression of the IP cell marker TBR2 in the differentiated

Figure 6: Increased astrocyte differentiation mediated by neurospheres. (a) ICC of 21-day differentiated astrocytes (AQP4) derived from 2D and 3D NPCs. (b) Quantification of AQP4+ positive astrocytes derived from 2D and 3D NPCs. Results were reported as Mean ± SEM of 15 fields from 3 independent cultures (**P < 0.01), using Student's t-test. (c) Analysis of gene expression by qPCR of AQP4 and TBR2. The expression values were normalized to GAPDH gene expression. Subsequently, the expression values were calculated as relative amount of mRNA versus expression values of N S5 (derived from 2D NPCs) 21-day differentiated cells, which was set to 1. Data was reported as Mean ± SEM of 3 independent cultures, tested for significant difference (**P < 0.01) using Student's t-test.
cells derived from 3D NPCs when compared to 2D NPCs. This indicated that neurosphere formation is associated with the loss of early IP cells (low PAX6 expression cells), resulting in the decrease of TBR2 expression in differentiated cells. During neurogenesis, IP cells asymmetrically divide into neurons and IP daughter cells [25]. Moreover, 1/6 of RG cells (high PAX6 expression cells) proceed to produce astrocyte and oligodendrocyte progenitors, resulting in the generation of astrocytes and oligodendrocytes [24, 25]. However, we do not observe the suppression of neuronal differentiation in our 2D analyzed cell lines. The reason for this may be due to the fact that only a small population of RG cells would proceed to give rise to glia cells; the remaining RG cells are still able to differentiate into neurons. Taken together, our results demonstrate that astrocyte differentiation is promoted by 3D propagation and relies on the increase of neurosphere aggregation.

5. Conclusion
In summary, we established a reliable NPC culture system in 3D that efficiently enriches GFAP and AQP4 positive astrocytes during terminal differentiation. Propagation of NPCs using the 3D culture system promotes the expression of RG cell markers (PAX6 and NESTIN). Subsequently, GFAP and AQP4 expression is increased during terminal astrocyte differentiation in the cells differentiated from 3D cultured NPCs. These results revealed an attractive method to improve the astrocyte differentiation ability from iPSC-derived NPCs in a shorter time frame and with higher efficiency compared to derivations from 2D cultured NPCs.

Conflict of Interests
The authors confirm that this paper content has no conflict of interests.

Authors’ Contribution
Shuling Zhou performed experimental design, implementation of experiments, analyses, and interpretation of the results, wrote the paper, and approved the final version. Karolina Szczesna helped with experimental design, manipulation of experiments, and writing of the paper. Anna Ochalek performed flow cytometry experiments and neural induction experiments. Julianna Koboláek was involved in the experimental design and read and approved the final version of the paper. Eszter Varga generated original iPSC lines used in this study. Csilla Nemes generated original iPSC lines used in this study. Abinaya Chandrasekaran was involved in neural induction experiments. Mikkel Rasmussen read and approved the final version of the paper. Susanna Cirera read and approved the final version of the paper. Poull Hyttel read and approved the final version of the paper. András Dinnyés helped with experimental design and implementation of experiments and read and approved the final version of the paper. Kristine K. Freude helped with analyses and interpretation of the results and writing of the paper and approved the final version. Hasan X. Avci helped with experimental design and analyses and interpretation of the results and read and approved the final version of the paper. Kristine K. Freude and Hasan X. Avci contributed equally to this paper.

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