A mini review of dolphin carbohydrate metabolism and suggestions for future research using exhaled air

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INTRODUCTION

Bottlenose dolphins (Tursiops truncatus), even as neonates, have very low carbohydrate diets. Dolphin milk contains only about 1% carbohydrate. During the nursing period of 1–3 years, the offspring receive large amounts of lipid and protein, but only small amounts of lactose milk sugars. Of the 1% sugar in dolphin milk, 90% is lactose, and only minor amounts of glucose and its metabolites are found (1). Once dolphins are weaned, their diet primarily consists of whole fish of various species that yield a diet high in fat and protein and are almost devoid of carbohydrate (2). The normal dolphin diet is in many ways similar to the ketogenic diet of humans (3). Dolphin dietary protein in the whole fish diet, however, yields amino acids whose metabolic effect is primarily gluconeogenic rather than ketogenic.

Given a whole fish diet, I was surprised to find relatively high fasting blood sugar levels in dolphins (4). We collected blood samples after 14-h fasting periods. Mean ± SD glucose values for 110 samples from 10 males was 131 ± 36 mg/dL and for 121 samples from 11 females, mean glucose was 127 ± 27 mg/dL. These values for true glucose seemed high, but the findings were consistent in healthy, non-stressed dolphins. One dolphin had several fasting plasma glucose values over 200 mg/dL. Interestingly, concurrent urinalysis showed no rise in urine glucose. The animals had no symptoms of diabetes mellitus such as weight loss, fatigue, or vomiting (a colleague, Dr. David Kenney indicated that he had a diabetic dolphin that he treated with insulin. While the data were never published, Dr. Kenney’s personal observation led us to probe further into dolphin carbohydrate metabolism).

Our studies showed that dolphin glucose tolerance curves were prolonged. After we gave an oral glucose load of 1.75 g/kg body weight, plasma glucose did not return to baseline levels for many hours. Thus, several of our studies showed a prolonged glucose tolerance curve, mimicking a diabetic response. Interestingly, urine glucose values were not elevated after a glucose load. When enough glucose was given to raise plasma glucose above 300 mg/dL, urine glucose increased. The high threshold for glucose excretion is facilitated by a higher plasma osmolality, which in dolphins, is around 335 mOsm/L (5). Humans with diabetes mellitus have increased plasma osmolality (320 mOsm/L) and acid urine, more like dolphins. Thus, the tubular renal threshold for glucose in dolphins is about twice as high as in humans.

During prolonged fasting states, dolphins do not become ketogenic. When dolphins were fasted for 72 h, plasma glucose levels remained at pre-fast levels. Even after 72 h, we were not able to sniff acetone or ketone smells in the dolphin breath. Levels of acetone and acetoacetic acid were also not increased in urine. It appeared that dolphins successfully balance protein and lipid metabolism during fasting. As result, glucose is produced by gluconeogenesis in the liver as body protein is metabolized during prolonged fasting. Muscle and other body protein sources were consumed along with body fat (5). Similar lack of ketoacidosis in another marine mammal model, the northern elephant seal, occurs in more extreme circumstances, including 3-month fasting periods (6). Other animal models for diabetes were recently discussed (7).
DOLPHIN THYROID
Thyroid activity changes, including hypo- and hyper-thyroidism have been associated with diabetes in humans (8–10). Hyperthyroid states have been proposed as both the cause of a diabetes-like hyperglycemia and caused by potential injury to pancreatic islet cells. Dolphin thyroid measurements, including plasma levels of thyroxine (T4) and triiodothyronine (T3), have been previously published (5, 11–15). Dolphin T4 levels are slightly higher or similar to human normal values, depending on tests employed, while T3 levels are within the human normal range. Using iso-tope labeled T3 and T4 injected simultaneously, turnover rates of both thyroid hormones were higher in dolphins than in humans (13). Future research may focus on the potential role and reason of higher thyroid hormone turnover rates in dolphins and associations with hyperglycemia and their natural diabetes-like states. Dolphin T4 levels have been shown to increase after 24 h of fasting (11).

DOLPHIN DIVING
In our early studies, immunologically measurable insulin in the blood was low, and plasma insulin was slow to increase after an oral glucose load (5). This negligible insulin response to glucose was different than the hyperinsulinemic response after ingestion of high protein fish diets (15). The lack of increased insulin with elevated glucose in dolphins may be especially important since high insulin levels can inhibit gluconeogenesis, and gluconeogenesis is the source of nearly all glucose from the natural fish diet. Lower insulin levels may have some significance as it relates to a dolphin’s diving physiology. Considerable lactate is produced during a dive (16–18). Lactate is well used by the diabetic heart, whereas its glucose utilization is extremely low. When insulin is supplied, the heart increases its utilization of glucose but not of lactate; whereas, brain metabolism of glucose is little affected by insulin (19). Increased lactate consumption by the heart might be advantageous to an animal making frequent dives. The blood glucose level closely follows the lactate level and increases slightly after a dive (20). Trained belugas (Delphinapterus leucas) had marked increases in plasma lactate after trained dives in the open ocean (18).

In another study, trained dolphins made numerous dives in the open ocean. Upon returning from depth, animals exhaled in an underwater funnel before taking a breath at the surface (21). After dives to 200 m depth, while the dolphin was at the surface just after the dive, we conducted a short study. The animal was signaled to exhale into the funnel on the third, sixth, and ninth breath after dives. After about nine breaths the dolphin was ready to dive again. I interpreted the results to indicate that it took about 1.75 g/kg of body weight. Analysis of the exhaled breath (Figure 1) revealed that the RQ remained close to 0.70, actually lower than when they are fed fish or fasted for short periods. Future research may explore the adaptive relevance of why dolphins do not have an increased RQ in response to glucose administration, including an apparent lack of an oxidative response.

Disaccharidases are enzymes that break ingested disaccharides (i.e., sucrose, lactose, and maltose) into simpler sugars (monosaccharides, including glucose, fructose, and galactose). In humans, insulin deficiency induces an abnormal increase in intestinal disaccharidases (26). Disaccharidase in the gastrointestinal tract is absent in some species of marine mammals (27). Since dolphin intestinal disaccharidase had not been examined, I decided to test the ability of the dolphin to digest sucrose using the exhaled breath. Results from sucrose ingestion studies were slightly different from earlier studies using glucose. CO₂ in dolphin breath was measured after oral sucrose ingestion (1 kg of sugar in fish). We analyzed dolphin breath after various periods of breath hold and compared sugar-fed days with days when only fish were fed. Multi-variate 2-way analyses of variance were used to evaluate differences in CO₂ and O₂ fraction of air expired, as well as respiratory quotients, by ingestion or no ingestion of sugar. Results are shown in Figures 1B–D. Similar to my findings with ingested glucose, there were no significant differences in O₂ or RQ among dolphins that did or did not eat sugar; or by hours after eating sugar. CO₂ levels, however, were higher among dolphins that ingested sugar compared to dolphins that did not (0.06 ± 0.013 and 0.05 ± 0.005, respectively; P < 0.001). Among human patients, PaCO₂ conversely decreased when enteral-fed high fat, low carbohydrate diets, demonstrating that dolphins and humans may have similar increases and decreases in CO₂ in response to higher and lower carbohydrate intake (28). Thus, changing CO₂ levels with sucrose consumption indicates that dolphins, unlike some marine mammals, can digest complex carbohydrates. This would not
be unexpected given that dolphins originated from land and their closest terrestrial, ungulate relatives have diets with complex carbohydrates (29).

Single breath samples from dolphins has been studied to further elucidate diving physiology and carbohydrate metabolism. Recently, nitric oxide (NO) levels in dolphin breath have been determined (30).

FUTURE BREATH RESEARCH

Due to the dolphin’s unique respiratory anatomy and their diabetes-like physiology, important results will likely come from dolphin breath and breath condensate measurements (31). Analysis with modern equipment will enable more discriminate detection of compounds linked to changing metabolic states. However, I propose that the intelligence of the canine and its sensitive nose will yield useful data as well.

THE DOLPHIN BREATH

Dolphins breathe less frequently than terrestrial mammals. They compensate by taking deeper breaths, and they extract more oxygen (O\textsubscript{2}) from the breath (5, 32). In terrestrial mammals, including humans, lung volume represents about 6% of body volume irrespective of body weight (33). In bottlenose dolphins that we have studied, total lung volume on a milliliter per kilogram basis is similar to that of humans. Thus, compared to humans, dolphins take in at least four times as much air with each breath. Additionally, dolphins breathe less frequently, take much deeper breaths, and extract more O\textsubscript{2} from each lung full of air.

A total of 80% tidal air was measured in dolphins (32). Dolphins inhale in a fraction of a second, hold the breath for a considerable period then rapidly exhale and inhale again. Their blowhole and robust nasal cavities, with no turbinates or other structures for potential slowing of the air, allow for rapid inhalation through a short robust trachea to the depth of each lung. Dolphins take in air that is almost 21% O\textsubscript{2}. At the end of a long breath hold, dolphin exhaled air can contain as little as 1.5% O\textsubscript{2}. The alveoli of the dolphin lung are structured for maximum exposure of lung air to blood circulation. The inter-alveolar septa have double capillary beds separated by an elastic-tissue membrane (34). Compared to a single capillary bed in human lung, the dolphin lung may offer a more efficient exchange of O\textsubscript{2} and carbon dioxide (CO\textsubscript{2}). In addition, other metabolites can be efficiently exchanged between blood and lung air (35).

COLLECTING DOLPHIN BREATH

Wondering how to collect the dolphin breath, as I walked beside a dolphin pool, an animal approached me and released a large bubble. This was a common dolphin behavior. They sometimes exhale...
large bubbles under aggregations of fish very near the surface. The rising bubble carries some of the fish to the surface immobilizing the fish just long enough for the dolphin to easily take them. For me, the bubble release just below me appeared to be a gesture connected with food (35). Trainers used this natural bubble blowing behavior, and found that dolphins readily exhaled into a large water-filled, underwater funnel (21, 30). After an explosive exhalation, dolphin breath displaced the water in a fraction of a second and flowed through tubing for collection and analysis (Figure 1A).

**CANINE DETECTORS**

It is well known that canines (dogs) have a very sensitive sense of smell (36). They also have a very good ability to learn the names of numerous different objects (37, 38). This cognitive and sensory ability can be melded using techniques employed to train dolphins. Dolphins will exhale into an underwater funnel, the breath with flow up to the dog for detection of breath markers by smell (Figure 2). The dog then goes to pick up a specific marker that has been identified by word and by odor to match a breath marker of interest, for example acetone (Figures 2B,C). Dogs have also been trained to communicate using a keyboard (39). The keyboard capability might also be used.

**ADVANTAGE OF CANINE DETECTORS**

Although sophisticated analysis equipment is available in the laboratory setting, it is not readily adaptable for use at the dolphin locations. For example, seawater is often corrosive to complex electronic equipment. Dolphin breath has been collected in special bags and removed to the laboratory for analysis. In some cases, this may work well (30). In many cases, however, the immediate feedback from the canine detector would be invaluable. In addition, canines are quite mobile and can be posted at sea or in remote locations where laboratory support and logistics are difficult.

In summary, while the bottlenose dolphin can have a sustained postprandial hyperglycemia, mimicking diabetes in people, dolphins appear to have many adaptations that enable the benefits of readily available glucose while potentially protecting them against complications of hyperglycemia. Understanding how these mechanisms work may have profound benefits for human health, especially with regard to diabetes.

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