The consensus on indications, conditioning regimen, and donor selection of allogeneic hematopoietic cell transplantation for hematological diseases in China—recommendations from the Chinese Society of Hematology

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Abstract

Allogeneic hematopoietic stem cell transplantation (allo-HSCT) is widely used to treat malignant hematological neoplasms and non-malignant hematological disorders. Approximately, 5000 allo-HSCT procedures are performed in China annually. Substantial progress has been made in haploidentical HSCT (HID-HSCT), pre-transplantation risk stratification, and donor selection in allo-HSCT, especially after the establishment of the "Beijing Protocol" HID-HSCT system. Transplant indications for selected subgroups in low-risk leukemia or severe aplastic anemia (SAA) differ from those in the Western world. These unique systems developed by Chinese doctors may inspire the refining of global clinical practice. We reviewed the efficacy of allo-HSCT practice from available Chinese studies on behalf of the HSCT workgroup of the Chinese Society of Hematology, Chinese Medical Association and compared these studies to the consensus or guideline outside China. We summarized the consensus on routine practices of all-HSCT in China and focused on the recommendations of indications, conditioning regimen, and donor selection.

Keywords: Consensus, Allogeneic hematopoietic transplantation, China, Indication, Conditioning regimen, Donor selection, Standard of care

Background

Allogeneic stem cell transplantation (allo-HSCT) is widely used to treat malignant hematological neoplasms and non-malignant hematological disorders [1–4]. The Chinese Blood and Marrow Transplantation Registry (CBMTR) reported that the total number of allo-HSCT cases increased steadily from 950 cases in 2008 to over 5000 cases in 2016 [5, 6]. In contrast, 6189 allo-HSCT cases were performed in Europe and 8351 cases were performed in the USA in 2015 [3, 7]. Therefore, the standardization of allo-HSCT practices in China would provide a major global impact based on the large patient population.

The significant growth of allo-HSCT is a result of the increased availability of alternative donors and refinement of indications. First, there was a shortage of human leukocyte antigen (HLA)-matched sibling donors and unrelated donors in China, but the success of haploidentical HSCT (HID-HSCT) ushered in a new era of “everyone has a donor” [8]. A total of 99% of HID-HSCT cases followed the “Beijing Protocol”, which...
includes T-replete myeloablative HID-HSCT with granulocyte colony-stimulating factor (G-CSF) and antithymocyte globulin (ATG) [9–23]. The Baltimore group introduced HID-HSCT followed by the posttransplant cyclophosphamide (PT-CY) regimen, which was also implemented in China [24]. The number of HID-HSCT cases increased to approximately 2500 annually in 2016, which made it the largest source of allo-HSCT donors (37.6–51.5%) in China since 2013 [5, 8, 25] and take 40% of annual HID-HSCT cases worldwide. In contrast, the frequency of HID-HSCT grew steadily from 3 to 5% to approximately 10% of allo-HSCT in Europe (2000 HID-HSCT cases in 2015) and the USA (1000 HID-HSCT cases in 2015) [3], where HID remains a minor donor source compared to HLA-matched sibling donor (MSD) and matched unrelated donors (MUD). The rapid development of HID-HSCT also raised questions of “Who is the best alternative donor?” and even “Who is the best donor?” because HLA no longer plays the predominant role in donor selection [10, 16, 26–28]. Second, individualized conditioning regimens based on patients’ statuses expand the target patient population, such as a reduced-intensity regimen for older patients [29–31] or novel conditioning for severe aplastic anemia (SAA) [32–34]. Third, pre-transplant risk stratification has enabled the early identification of patients with high risk of relapse in chemotherapy, which may provide valuable information for the selection of allo-HSCT as post-remission therapy [35, 36].

The Chinese Society of Hematology (CSH) updated the recommendations from a consensus conference of the HSCT workgroup of CSH in 2017 based on differences in the practices of allo-HSCT in China and the Western world. The present guidelines focus on indications, conditioning regimen, and donor selection. Seventeen experts with recognized clinical and research expertise in allo-HSCT participated in the consensus discussion and were elected as members of the HSCT workgroup, Chinese Society of Hematology. These experts represented the most active allo-HSCT centers (approximately 60% of total allo-HSCT cases) in China. This consensus will likely contribute to the standardization of allo-HSCT practices in China and become an inspiration for further international cooperation to refine global practices.

Indication and timing of allo-HSCT

Other HSCT groups, such as the American Society for Blood and Marrow Transplantation (ASBMT) and British Society of Blood and Marrow Transplantation, have systematically summarized the indications and timing for allo-HSCT or recommended guidelines for specific diseases, such as the National Comprehensive Cancer Network (NCCN) guidelines for malignant hematological diseases and the British Committee for Standards in Haematology (BCSH) guidelines for aplastic anemia. Recent Chinese studies suggested that specific patient subgroups may benefit from allo-HSCT rather than other conventional non-allo-HSCT treatments. Therefore, the indications for allo-HSCT may be extended for these patient subgroups in China, which is not in strict accordance with the current recommendations in the Western world (Table 1).

Acute myeloid leukemia

Allo-HSCT, especially MSD-HSCT, is the standard care option for acute myeloid leukemia (AML) patients classified as an intermediate and high-risk karyotype by the NCCN stratification system [37] in any disease state (CR1/CR2 or above/active disease) in the ASBMT recommendations [38]. HID-HSCT was confirmed as an equally good option to MSD-HSCT as post-remission therapy for AML patients in CR1 who lack a matching donor following the Beijing Protocol [18]. A prospective, multicenter study investigated 450 AML patients who were assigned to undergo haplo (231 patients) or ISD HSCT (219 patients) based on donor availability. The HID and ISD HSCT groups exhibited, respectively, comparable 3-year disease-free survival (DFS) of 74 and 78% (p = 0.34), overall survival (OS) of 79 and 82% (p = 0.36), cumulative incidence of relapse (CIR) of 15 and 15% (p = 0.98), and treatment-related mortality (TRM) of 13 and 8% (p = 0.13). Therefore, we did not differentiate recommendations for transplant indications based on donor source (i.e., related donor, unrelated donor, umbilical cord blood, or haploidentical donor) (Table 2).

AML patients classified as a low-risk karyotype by the NCCN guidelines, such as patients with genetic abnormality of RUNX1-RUNX1T1 and CBFB-MYH1, may benefit from allo-HSCT in CR1 by risk-directed, minimal residual disease (MRD)-based therapy. An AML05 multicenter trial revealed that MRD status (RUNX1-RUNX1 reduction < 3 log units) after the second consolidation discriminated these patients into subgroups. The high-risk group was defined as patients in whom major molecular remission (MMR) was not achieved after the second consolidation therapy or patients who exhibited loss of MMR within 6 months of achieving it. The low-risk subgroup was defined as patients in whom MMR was achieved after the second consolidation therapy and maintained for 6 months. Allo-HSCT reduced relapse and improved survival compared to chemotherapy in these high-risk patients (HSCT vs. chemotherapy: respectively, CIR 22.1 vs 78.9%, P < 0.0001; DFS 61.7 vs. 19.6%, p = 0.001), whereas chemotherapy/auto-HSCT achieved a low relapse rate (5.3%) and high DFS (94.7%) in low-risk patients. MRD-directed pre-transplant risk stratification may improve the outcome of t(8;21) AML in CR1 [35]. Similar results were observed with inv(16) AML. Poor molecular response was defined as a CBFB-
Table 1 The clinical outcome of HID-HSCT and trials comparing allo-HSCT with chemotherapy or TKI

| Author [ref.] | Patients | No | Diagnosis | CIR | OS | LFS |
|---------------|----------|----|-----------|-----|----|-----|
| Wang et al. [27], 2013 | HID-HSCT, retrospective | 756 | Leukemia | 15% (SR), 26% (HR) at 2 years | 68% (SR) at 3 years 49% (HR) at 3 years |
| Huang et al. [15], 2012 | HID-HSCT vs CT prospective | 58 vs 74 | AML-CR1 adults IR or HR | 12 vs 57* | 7.5 vs 54.7%* at 4 years | 73.1 vs 44.2%* at 4 years |
| Zhi et al. [35], 2013 | HSCT vs CT | 58 vs 58 | ETO(+)- adults AML-CR1 | 22.1 vs 78.9% (HR)* 14.7 vs 5.3% (LR) | 71.6 vs 26.7% (HR)* 75.7 vs 100% (LR) | 61.7 vs 19.6% (HR)* 70.3 vs 94.7% (LR) |
| Qiu et al. [36], 2015 | HSCT vs CT | 57 vs 29 | In(16) adults AML-CR1 | 7.1 vs 87.7% (poor MR)* 0 vs 26.9% (good MR); 93.3 vs 40.0%, (poor MR)*; 72.9 vs 77.1% (good MR) | 67.6 vs 12.3% (poor MR)* 72.9 vs 73.1% (good MR) |
| Yan et al. [82], 2014 | HID-HSCT vs CT | 79 vs 59 | SR-ALL-CR1 adults | 29.9 vs 66.3%* at 5 years | 70.4 vs 28%* at 5 years | 54.4 vs 23.9%* at 5 years |
| Sun et al. [83], 2014 | HID-HSCT vs CT | 79 vs 104 | HR-ALL-CR1 adults | 18.7 vs 60.5%* at 3 years | 72.5 vs 26.6%; at 3 years | 63.9 vs 21.1%* at 3 years |
| Wang et al. [84], 2012 | HID-HSCT, T-ALL | 72 | T-ALL | 18.8% (CR1) 37.5% (non-CR1) | 54.0% (CR1) 12.5% (non-CR1) |
| Xu et al. [46], 2016 | HSCT | 52 | Pediatric T-ALL (HR) | 32.7% at 3 years | 55.5% at 3 years | 54.1% at 3 years |
| Jiang et al. [85], 2011 | HSCT vs IM | 87 vs 45 | CML-AP adults | 81.2 vs 100% (LR) at 6 years | 80.7 vs 80.9% (LR) at 6 years | 61.9 vs 47.1% (HR)* at 6 years |
| Xu et al. [52], 2016 | HSCT vs TKI2 | 60 vs 33 | CML-AP adults | 86.4 vs 42.9%* at 5 years | 76.1 vs 14.3%* at 5 years |

HID haploidentical donor, MRD matched-related donor, HSCT hematopoietic stem cell transplantation CT, AML acute myeloid leukemia, ALL acute lymphoblastic leukemia, MDS myelodysplastic syndrome, LR low risk, IR intermediate risk, HR high risk, T-ALL T cell ALL, MR molecular response, ph Philadelphia chromosome[t(9;22)], CML chronic myeloid leukemia, CP chronic phase, AP accelerated phase, BC blast crisis, IM imatinib, TKI2 second-generation TKI, SAA severe aplastic anemia. * The difference was significance between two groups (P<0.05)

MYH11 level of 0.2% after the second consolidation. Allo-HSCT decreased the 3-year CIR and increased the DFS and OS of patients who exhibited a poor molecular response [36]. Therefore, patients with t(8;21) or inv(16) AML, who are considered high-risk by MRD-directed risk stratification, benefited from HSCT in CR1.

The detection of some molecular markers, such as semi-quantitative assessment of FLT3-ITD allelic ratio, was not widely available in China, and the risk stratification generally followed NCCN rather than European Leukemia Net (ELN) recommendations.

**Acute lymphoblastic leukemia**

Adult patients with high Ph(+) Acute lymphoblastic leukemia (ALL) in CR1 benefit from MSD-HSCT and HID-HSCT (Table 2). Han et al. retrospectively investigated the outcomes of HID-HSCT in adults with standard-risk ALL in CR1 and compared these patients to MSD and MUD patients. A total of 127 HID, 144 MSD, and 77 MUD recipients were included in the study. There were no differences in grade III–IV acute graft-versus-host disease (aGVHD) (11.4 vs. 7.7 vs. 13.5%, p = 0.468), 5-year TRM (16.4 vs. 11.6 vs. 19.6%, p = 0.162), 5-year CIR (14.8 vs. 21.1 vs. 16.7%, p = 0.231), 5-year OS (70.1 vs. 73.7 vs. 69.8%, p = 0.525), 5-year DFS (68.7 vs. 67.3 vs. 63.7%, p = 0.606), or 3-year GVHD-relapse-free survival (GRFS; 50.8 vs. 54.9 vs. 52.2%, p = 0.847), respectively, [39]. Wang et al. compared HID and MSD for HSCT in adults with Ph(+) high-risk ALL in a biological phase III randomized multicenter study [40]. A total of 103 cases received HSCT from HID and 83 received HSCT from MSD. There were no differences in 3-year DFS (61 vs. 60%, p = 0.91) from CR, 3-year OS (68 vs. 64%, p = 0.56) from HSCT, TRM (13 vs. 11%, p = 0.84), or CIR (18 vs. 24%, p = 0.30). Therefore, HID-HSCT is a valid alternative as post-remission treatment for high- and standard-risk adult patients with ALL in CR1 who lack an identical donor [17].

Ph + ALL remains an important indication for allo-HSCT in this era of treatment with tyrosine kinase inhibitors (TKIs) [40, 41]. The results of MSD and HID-HSCTs were similar in adult and pediatric patients [42, 43]. Chen et al. investigated 50 pediatric patients with Ph + ALL who underwent HID-HSCT. The 5-year EFS was 61.0%, the OS was 70.0%, the 3-year CIR was 22.7%, and the NRM was 16.4%. Therefore, HID-HSCT for pediatric patients with Ph + ALL yielded promising long-term survival [42]. Zhang et al. analyzed the outcomes of 82 Ph + ALL patients who underwent HID-HSCT (n = 47) or MSD-HSCT (n = 35). HID-HSCT was associated with a significantly lower relapse rate than MSD-HSCT (44.8 vs. 19.1%, p < 0.05). There were no differences in NRM, LFS, or OS between the two groups [44]. Recently, Wang et al. reported for low-risk Ph + ALL patients, who were defined as WBC < 30 × 109/L at diagnosis and 3-log reduction of BCR-ABL levels from baseline after two
| Author                  | HSC type                      | Case number | Diagnosis                | TRM    | CIR    | aGVHD | OS     | LFS    |
|------------------------|-------------------------------|-------------|--------------------------|--------|--------|-------|--------|--------|
| Wang et al. [18], 2015 | HID vs MSD multicenter, prospective | 231 vs 219  | Adults IR/HR AML-CRI     | 3 years, 13 vs 8% | 3 years, 15 vs 15% | 3 years, 79 vs 82% | 3 years, 74 vs 78% |
| Liu et al. [61], 2013  | HID vs MSD                    | 212 vs 46   | Pediatric AL             | II–IV* 408 vs 20% | III–IV 143 vs 16.9% | 778 vs 65.5% | 68.9 vs 52.5% (ALL-CRI); 82.5% vs 71.7% (AML-CRI) |
| Wang et al. [17], 2016 | HID vs MSD Adults Phase II randomizes | 121 vs 89   | (Ph) ALL-CR1 (HR)        | 3 years, 18 vs 24% | 3 years, 74 vs 78% | 3 years, 61 vs 60% |
| Chen et al. [43], 2015 | HID vs MSD                    | 101 vs 38   | Ph + ALL                 | 15.6% at 5 years | 20.3% at 5 years | 74.0 vs 68% at 5 years | 65.8 vs 61% at 5 years |
| Gao et al. [44], 2015  | HID vs MSD                    | 47 vs 35    | Ph + ALL                 | 21.3 vs 17.1% | 19.1* vs 44.8% | 82.5% vs 71.7% |
| Wang et al. [19], 2016 | HID vs MSD                    | 136 (3/6) vs 90(4/6) vs 228 | MDS          |                   |                   | 58 vs 63 vs 73% | 58%, 63 vs 71% |
| Ma et al. [64], 2016   | HID vs MSD                    | 67 vs 23    | CML-BC                   |                   |                   | 3 years, 60.0 vs 55.3% | 3 years, 51.1 vs 47.8% |
| Xu et al. [33], 2016   | HID vs MSD Data base          | 101 vs 48   | SAA failure to previous IST |                   |                   | 3 years, 89.0 vs 91.0% | 3-year FFS 86.8 vs 80.3% |
| Xu et al. [34], 2017   | HID vs MSD                    | 89 vs 69    | SAA, upfront             | 97.8 vs 97.1% |                   | 3 years, 86.1 vs 91.3% | 3-year FFS 85.0 vs 89.8% |
| Sun [79], 2016         | HID vs URD, Pair-match        | 87 vs 87    | AML-CRI                  | 13.8 vs 15.7% | 127 vs 24% | 5 years, 78.2 vs 63.6% | 5 years, 73.5 vs 60.3% |
| Huang et al. [13], 2009| HID vs URD                    | 219 vs 78   | Malignant                | 2 years, 20 vs 18% | 2 years, 12 vs 18% | 4 years, 74 vs 74% | 4 years, 67 vs 61% |
| Han et al. [39], 2017  | HID vs MSD vs MUD             | 127 vs 144 vs 77 | Adult patients | 16.4 vs 11.6 vs 19.6% | 148 vs 21.1 vs 16.7% | 701 vs 73.7 vs 69.8% | 5-years DFS 68.7 vs 67.3 vs 63.7% |
| Luo et al. [23], 2014  | HRD vs URD vs MSD Phase II Prospective | 99 vs 116 vs 90 | Hematologic malignancies | 30.5 vs 22 vs 4.7%* | 142* vs 21.2 vs 34%; 15.4* vs 28.2 vs 49.9% (in HR pts) | 608 vs 635 vs 77.2%, | 58.3 vs 58.4 vs 63.6% |
| Mo et al. [81], 2016   | HID vs CBT multiple centers   | 65 vs 64    | ALL (HR) pediatric       | 2 years, 128 vs 18.8% | 2 years, 161 vs 24.1% | 2 years, 82 vs 69.6% | 2 years, 71 vs 57.2%* |

OS overall survival, RR relapse rate, LFS leukemia-free survival, HID haploidentical donor, MRD matched related donor, HSCT hematopoietic stem cell transplantation, MRD matched related donor, CBT cord blood transplantation, URD unrelated donor, AML acute myeloid leukemia, ALL acute lymphoblastic leukemia, MDS myelodysplastic syndrome, LR low risk, IR intermediate risk, HR high risk. **T-ALL** T cell ALL, MR molecular response, Ph Philadelphia chromosome[t(9;22)], CML chronic myeloid leukemia, CP chronic phase, AP accelerated phase, BC blast crisis, IM imatinib, TKI2 second-generation TKI, SAA severe aplastic anemia. * The difference was significance between two groups (P<0.05)
consolidation cycles, there was no significant difference between the allo-HSCT and non-transplant groups for CIR (8.5 vs. 7.7%, *p* = 0.671), DFS (88.2 vs. 83.9%, *p* = 0.426), and OS (96.6 vs. 83.3%, *p* = 0.128), which suggested selected low-risk patients might be free from allo-HSCT [45].

Pediatric patients with T-ALL benefit from HSCT, including HID-HSCT. Xu ZL examined 48 consecutive children with high-risk T-ALL who underwent HID-SCT in a prospective study [46]. The 3-year CIR was 30.8%, excluding HID-HSCT. Xu ZL examined 48 consecutive children with MDS who underwent HSCT from HIDs (226 patients, 136), and ISD patient groups were, respectively, at the time of SCT [52].

### Myelodysplastic syndromes (MDS)

Allo-HSCT is the standard care for advanced MDS (IPSS Intermediate-2/high-risk). Patients with lower-risk MDS (refractory anemia or refractory anemia with ringed sideroblasts) who exhibited poor prognostic features and/or signs of progression or sustained profound cytopenia (neutrophil count < 0.5 × 10^9/L and/or platelet count < 20 × 10^9/L) were also considered candidates for allo-HSCT.

Wang et al. analyzed the outcomes of 454 patients with MDS who underwent HSCT from HIDs (*n* = 226) or ISDs (*n* = 228) and reported the results to the CBMTR. The 4-year NRM values of the 3/6 HID (*n* = 136), 4–5/6 HID (*n* = 90), and ISD patient groups were, respectively, 34, 29, and 16% (*p* = 0.004); the CIR values were 6, 7, and 10% (*p* = 0.36); the 4-year OS values were 58, 63, and 73% (*p* = 0.07); and the RFS values were 58, 63, and 71% (*p* = 0.14). HLA disparity exerted no effect on survival in the HID group [19].

### Chronic myelogenous leukemia

Allo-HSCT is no longer the primary treatment option for chronic myelogenous leukemia (CML) patients in the early chronic phase because imatinib exhibited better outcomes than MSD-HSCT in young persons with newly diagnosed CML-CP [48]. The corresponding percentage of allo-HSCT for CML patients decreased from 22% in 2008 to less than 2% in 2017 in China. However, allo-HSCT was superior to first- and second-generation tyrosine kinase inhibitors (TKIs) for patients in the accelerated phase (AP) and blastic crisis (BC) [49–51]. As anti-T315i-mutated TKIs are not available in China, and allo-HSCT is an option for this group of patients in any phase. Xu et al. reported the outcome of SCT in 22 patients with T315I(+) CML, and most of whom (*n* = 16) underwent HID-SCT. The 2-year LFS were 80.0, 72.9, and 0% for the CP, AP/AP-CBn, and BP/BP-CBn groups, respectively, at the time of SCT [52].

### Severe aplastic anemia

The Guidelines of the British Society for Hematology list MSD-HSCT and MUD-HSCT as first-line standard care for severe aplastic anemia (SAA) patients (age < 50), and HID-HSCT was only considered a second-line treatment for refractory SAA after a failed immunosuppressant therapy (IST). In contrast, transplant indications for SAA are not differentiated on donor source in China. The feasibility of HID transplantation for the treatment of SAA patients after the failed immunosuppressant therapy was evaluated in a prospective multicenter clinical trial [33] (Table 2). Recipients of HID-HSCT exhibited a higher incidence of grade II–IV aGVHD than patients (*n* = 48) who received MSD-HSCT (33.7 vs. 4.2%, *p* < 0.001) but similar values of grade III–IV aGVHD (7.9 vs. 2.1%, *p* = 0.157), 3-year OS (89.0 vs. 91.0%, *p* = 0.555), and failure-free survival (FFS) (86.8 vs. 80.3%, *p* = 0.659). Furthermore, HID-HSCT was evaluated as an upfront therapy for SAA in a registry-based comparison study. Eighty-nine patients received HID-HSCT, and 69 patients received MSD-HSCT. HID recipients exhibited a similar incidences of extensive cGVHD (3.4 vs. 0%, *p* = 0.426), 3-year OS (86.1 vs. 91.3%, *p* = 0.358), and 3-year FFS (85.0 vs. 89.8%, *p* = 0.413) compared to MSD, with increased incidence of II–IV aGVHD (30.3% vs. 1.5%, *P* < 0.001) and total cGVHD (30.6 vs. 4.4%, *p* < 0.001). Xu et al. reported that the treatment of 52 children with SAA with HID-HSCT produced 3-year OS of 84.5 and FFS of 82.7% [53]. These results suggest that newly diagnosed and refractory SAA benefit from HID-HSCT as MSD or MUD-HSCT.

In summary, the published evidence suggests that allo-HSCT be recommended for intermediate- and high-risk AML-CR1, selected subgroups of low-risk AML-CR1, Ph + ALL, adult standard-risk ALL- CR1, MDS, high-risk AL, and CML-AP/CBL-BC. HID-HSCT and/or MUD-HSCT demonstrated equivalent outcomes as MSD-HSCT in China. Therefore, we did not differentiate recommendations for transplant indications based on donor source, which is different from previous recommendations from Western countries.

Systematic, standardized pre-transplant risk stratification is important for patients who are eligible for allo-HSCT. The European Group for Blood and Marrow Transplantation (EBMT) risk score and hematopoietic cell transplantation-specific comorbidity index (HCT-CI) were feasible for the predicting of patient outcomes following HID-HSCT in China [54, 55].

A modified EBMT
risk score that used the number of HLA disparity instead of donor type also predicted patient outcomes [56].

Select older patients (age > 50) with low HCT-CI (≤ 2) and good performance status tolerated myeloablative haplo-HSCT with similar outcomes as younger adults [57]. Haplo-HSCT with a reduced-intensity regimen (RIC) with substitution of cyclophosphamide with fludarabine (Flu) was feasible in patients above 60 years of age, who exhibited similar engraftment and relapse rates as myeloablative conditioning in China [29, 30]. Select adults ≥ 70 years with hematological malignancies are considered for transplant in the USA. [31]. Therefore, the present consensus does not provide specific recommendation for eligible age.

**Recommendation: indications and timing for allo-HSCT**

**Patients with non-malignant hematological diseases**

1. AML include non-APL AML and APL.
   1-1 AML (non-acute promyelocyte leukemia (APL))
   A. AML (non-APL CR1):
   a.1 Intermediate- or unfavorable-risk disease according to NCCN risk stratification.
   a.2 Patients who achieve CR1 after ≥ 2 cycles of therapy.
   a.3 Patients with AML showing myelodysplasia-related changes or therapy-related myeloid changes.
   a.4 Patients with favorable-risk diseases according to WHO risk stratification who exhibit a poor molecular response to chemotherapy. RUNX1-RUNX1 reduction < 3 log units or CBFB-MYH11 level ≥ 0.2% after two consolidation cycles.
   B. AML (non-APL) ≥ CR2
   C. AML (non-APL) not in remission: allo-HSCT as salvage therapy with individualized conditioning regimens.

1-2 APL:
   A. Patient fails to react to induction therapy.
   B. Relapsed APL patient (molecular, cytogenetic, or hematological relapse) who remains PML-RARA-positive after the second induction therapy.

2. Acute lymphocytic leukemia (ALL)
   2-1 Acute lymphocytic leukemia (ALL) included ≤ 14 and > 14 years old patients [40, 58–60].
   A. ALL-CR1: especially patients with MRD(+) 8 weeks after induction therapy or showing high-risk factors: age > 35; high WBC count at presentation (≥ 100 × 10^9/L for T lineage and ≥ 30 × 10^9/L for B lineage); t(9;22) or complicated chromosome.
   B. The decision for allo-HSCT for adolescents who receive a chemotherapy protocol for pediatric patients should be made based on the appropriate guidelines for ALL (age ≤ 14 years).
   C. ALL ≥ CR2
   D. ALL not in remission: allo-HSCT as salvage therapy with individualized conditioning regimens or refer to clinical trials with novel cellular therapies as a bridge with allo-HSCT.

2-2 Acute lymphocytic leukemia (ALL) age ≤ 14 years [40, 61].
   A. Adolescent and pediatric ALL in CR1
   a.1 Patients who fail to achieve hematological CR or MRD > 1% within 28–30 days.
   a.2 Patients achieve CR with MRD > 0.1% within 12 weeks after therapy.
   a.3 Patients who exhibit MLL rearrangements who are < 6 months old or with WBC count > 300 × 10^9 cells/L.
   a.4 Philadelphia chromosome-positive patients, especially patients exhibiting poor response to prednisone and positive MRD at any time 4–12 weeks after therapy.
   B. Allo-HSCT in ALL ≥ CR2: All patients who exhibit very early or early relapse are candidates for allo-HSCT during CR2. All patients in CR3 are also recommended for allo-HSCT.
   C. ALL not in remission: allo-HSCT as salvage therapy for refractory or relapsed ALL.

3. Chronic myeloid leukemia (CML) [48, 62–64]:
   A. Allo-HSCT could be considered if patients in CP phase fail to respond to TKIs, depending on their age, consent, and the will of the patient.
   B. Patients who are resistant to or intolerant of second-generation TKIs are recommended to receive allo-HSCT.
   C. Patients with T315I-mutated BCR-ABL should choose allo-HSCT as a first-line therapy.
   D. Patients who have progressed to the accelerated or blast crisis phase are recommended for allo-HSCT.

4. Myeloproliferative neoplasms, including MDS, MDS/MPN, CMML, atypical CML, JMML, MDS/MPN, or unclassified [19, 65].
   A. Patients with intermediate-2 or high-risk IPSS scores are recommended for early allo-HSCT [66].
B. Patients with low-risk or intermediate-1 IPSS scores but showing severe neutropenia or thrombopenia or patients who are transfusion-dependent.
C. Children with JMML.

5. Myelofibrosis (MF): Patients with primary or secondary myelofibrosis with intermediate-2 or high-risk scores are candidates for allo-HSCT. The IPSS and DIPSS scores refer to NCCN guidelines [65].

6. Multiple myeloma (MM): [67]

A. Allo-HSCT is recommended for young patients with high-risk cytogenetic changes, such as t(4;14);t(14;16);17p-. Patients exhibiting disease progression after initial auto-HSCT may also receive allo-HSCT as salvage therapy.

7. Hodgkin lymphoma (HL): Patients who are refractory or relapse after ≥1 course of auto-HSCT [68].

8. Non-Hodgkin lymphoma (NHL): [69]

A. Chronic lymphocytic leukemia/small lymphocytic lymphoma (CLL/SLL): Allo-HSCT should be considered for young patients under the following conditions, in the absence of newly available drugs.
   a.1 Patients who are refractory to purine analogs or relapse within 12 months.
   a.2 Patients who respond to auto-HSCT or purine analog-containing regimens but relapse within 24 months.
   a.3 Patients with high-risk cytogenetic or molecular factors, regardless of response to therapy or relapse.
   a.4 Patients exhibiting symptoms of Richter syndrome.

Others: Allo-HSCT is also recommended for patients with NHL, including follicular lymphoma, diffuse large B-cell lymphoma (DLBCL), mantle cell lymphoma, lymphoblastic cell lymphoma and Burkitt lymphoma, peripheral T-cell lymphoma, and NK/T-cell lymphoma who are refractory, relapsed, or in ≥ CR2. If HLA-matched donors are available, HSCT may also be considered in CR1 for adult patients with mantle cell lymphoma, lymphoblastic cell lymphoma, Burkitt lymphoma, peripheral T-cell lymphoma, and NK/T-cell lymphoma.

Patients with non-malignant hematological diseases

1. Aplastic anemia (AA): [33, 34, 70]

A. Newly diagnosed with severe aplastic anemia (SAA): Patients who are < 50 years of age with SAA or vSAA with HLA-matched sibling donors could receive allo-HSCT as first-line therapy. Pediatric SAA/vSAA patients with ≥9/10 loci-matched unrelated donors may also choose allo-HSCT as first-line therapy. HID HSCT is recommended for young patients without MSD.

B. Refractory and/or relapsed SAA:
   b.1 SAA or vSAA patients below the age of 50 years who fail to respond to immunosuppression therapy (IST) or relapse may choose to undergo HID, MUD, or CBT
   b.2 SAA or vSAA patients who are 50–60 years old and fail to respond to IST or relapse with ECOG scores ≤2 are recommended for MSD or MUD transplantation.

C. Transfusion-dependent non-SAA patients, based on the guidelines for SAA.

2. Paroxysmal nocturnal hemoglobinuria (PNH): SAA/PNH who failed in IST.

3. Thalassemia: Allo-HSCT is recommended for transfusion-dependent severe thalassemia, including severe thalassemia, hemoglobin E combined with thalassemia, and severe hemoglobin E disease. HSCT is recommended before progression to stage 3 for children (2–6 years old).

4. Fanconi anemia: HSCT should be performed before the disease progresses to MDS or leukemia and too many blood transfusions.

5. Others: Patients with congenital immune deficiencies or metabolic diseases, including severe combined immunodeficiency and mucopolysaccharidoses, should receive HSCT.

All patients eligible for allo-HSCT should be evaluated using the HCT-CI, Kanofsky or Lansky Play performance score, EBMT score, or modified EBMT score for Haplo-EBMT.

Donor selection and graft source
MSD are generally the preferred choice for allo-HSCT, and Haplo, MUD, and cord blood (CB) are alternatives. The ideal donor should be identified among alternative donor candidates based on factors such as recipient condition (refractory or relapsed status, age, and performance status), characteristics of the alternative donors, and the experience of the transplantation center (if HID-HSCT available) (Fig. 1).
HID-HSCT
HID-HSCT exhibits a similar clinical outcome as MSD and MUD in the treatment of AML, ALL, MDS, and SAA, which are supported by the multicenter prospective studies listed in Table 2. The characteristics of haplo donor: (1) almost all patients can find a haplo donor on time and often have more than one alternative candidate, (2) a haplo donor is more suitable for patients in need of urgent allo-HSCT because it is not time-consuming, with only 2–3 weeks required for HLA-typing and physical examination, (3) HID can donate enough graft cells to be stored for future cell therapy, especially for high-risk relapsed patients, (4) bone marrow and peripheral stem cells may be obtained based on the clinical condition, (5) HID-HSCT exhibits a lower incidence of relapse than MSD-HSCT or MUD-HSCT in high-risk malignant hematological patients, (6) the incidence of aGVHD is higher in HID-HSCT than MSD-HSCT, and (7) peripheral blood and/or bone marrow are feasible in HID-HSCT.

Wang et al. analyzed the outcomes of 1210 consecutive transplant cases treated with a uniform protocol and proposed an algorithm for the selection of an HID from more than one candidate. Younger donors and male donors were associated with low NRM (HR = 0.30, p = 0.008 and HR = 0.65, p = 0.002) and high OS (HR = 0.73, p = 0.033 and HR = 0.73, p = 0.005). Father donors were associated with low NRM (HR = 0.65, p = 0.02) and aGVHD (HR = 0.69, p = 0.001) and high OS (HR = 0.66, p = 0.003) compared to mother donors. Child donors were associated with lower aGVHD than sibling donors (HR = 0.57, p = 0.01). Older sister donors were inferior to father donors for NRM (HR = 1.87, p = 0.02) and OS (HR = 1.59, p = 0.03). Non-inherited maternal antigen (NIMA)-mismatched sibling donors were associated with the lowest incidence of aGVHD compared with parental and non-inherited paternal antigen (NIPA)-mismatched sibling donors. Specific HLA disparities were not significantly correlated with the outcomes. The order rank for haplo donor characteristics in this algorithm was young, male, and non-inherited maternal antigen mismatch. Transplants from older mothers and NIPA-mismatched donors should be avoided as much as possible [28, 71].

Chang et al. focused on the relationship between donor-specific anti-HLA antibodies (DSAs) and primary graft failure (GF) after HID-HSCT and designed a prospective study with randomly assigned training and validation sets. The incidence of primary GF was 6.4%, which included GR (0.9%) and PGF (5.5%). Multivariate models revealed that DSAs (median fluorescence intensity (MFI) ≥ 10,000) correlated to primary GR (p < 0.001), and DSAs (MFI ≥ 2000) strongly associated with primary PGF (p = 0.005). These results suggested that the incorporation of DSAs into the algorithm improved HID selection. Donors with DSA MFI > 10,000 should be avoided [71, 72].

Donor-recipient CMV serostatus matching was not associated with transplant outcomes in HID-HSCT following the Beijing protocol. In contrast, the Baltimore group suggested that donors should have aCMV IgG serologic status similar to that of the recipients. This discrepancy may be related to the higher incidence of CMV infections in Chinese compared to Western populations.

Cohort studies that considered the different models of NK-KIR alloreactivity (KIR ligand model, missing-ligand model, KIR-KIR model, and receptor-ligand model) reported discordant results between different HID-HSCT approaches. A high relapse rate was associated with
missing self-molecules or missing ligands in the Beijing protocol, and a benefit of using donors with KIR B haplotypes was found in the PT-CY protocol. Both of these protocols are different from the “Perfect mismatch” in T-cell-depleted HID-HSCT [73–76].

The modified G-CSF + ATG protocols with pure G-BM or G-PB were also feasible compared to the mixed grafts of G-PB and G-BM in HID-HSCT introduced by the Beijing group [21, 77]. A propensity score method-based multicenter study demonstrated that HID-HSCT with mixed grafts achieved better 3-year DFS compared to G-PB alone (59.9 vs. 44.3%) [78].

**MUD-HSCT**

MUD-HSCT exhibits similar clinical outcomes as MSD-HSCT and HID-HSCT [79]. However, there are several specific characteristics in China for MUD: (1) the probability of finding an appropriate donor is very low (approximately 11% compared to 40–70% in the Western world) [80], and bone marrow graft is not available for MUD in China, (2) searching for and preparing a MUD would require 3–6 months, (3) donors may opt out of donation at any time, (4) there is very little chance that the donor would be willing to re-donate lymphocytes or stem cells if the patient needs it, and (5) there is a decreased incidence of severe aGVHD and increased risk of relapse in MUD transplantations compared to HID transplantations.

**CB-HSCT**

The treatment outcome is similar to MUD transplantation for malignant hematological diseases [81]. The following characteristics of CBT are noted: (1) it is possible to search and prepare CB on time without delays, (2) the incidence and severity of GVHD is low, (3) blood reconstruction is generally delayed, which leads to a high risk of infection, and (4) single-dose CBT is generally used for child patients, and double-dose CBT has not been widely adopted in China.

Who is the best allo-donor for a patient with acute leukemia for transplantation in CR1? Wang et al. reported a prospective data set of 1199 consecutive subjects who received a transplant from HID (n = 685) or MSD (n = 514). The 3-year LFS values were similar (75 vs. 74%, p = 0.95). Three major risk factors for TRM were identified in the multivariate model: higher donor/recipient age ratio, female-to-male transplants, and donor-recipient ABO major-mismatch transplants. Therefore, the donor-recipient age, matching for gender, and ABO incompatibility should be considered before selecting the ideal donor for patients with acute leukemia receiving related transplants in our model [16].

HLA-matched sibling donors are generally the preferred choice for allo-HSCT. However, we found that HID may be the ideal donor for subgroups with high risk or MRD-positive patients because HID-HSCT exhibited a strong graft versus leukemia (GVL) effect. MSD-HSCT is generally considered the best allo-HSCT. Chang et al. performed a retrospective study (n = 339) and a prospective study (n = 340) to verify this relationship [82]. MRD was determined using multiparameter flow cytometry. The results indicated that haplo-SCT was associated with a lower incidence of relapse and better survival for pre-MRD-positive AML patients, which suggests a stronger anti-leukemia (GVL) effect than MSD-HSCT.

**Recommendations: donor selection and mobilization**

**General principle of donor selection**

HLA-matched sibling donors are the first choice as allo-HSCT donors. A related HID is required for patients at high risk of relapse or search for MUD when an MSD is not available. Patients with favorable-risk diseases may choose unrelated donors because post-transplant cellular therapy is not necessary. Child patients may also choose CBT.

Several factors, including conditioning regimens, donors, disease status, and performance status of the patient, affect the outcome of the transplantation. Clinical practices must be standardized, and management strategies for each patient must be individualized. The optimal procedure includes risk stratification at diagnosis, overall treatment schedule, and transplantation at the optimal times.

**Algorithm for HID**

Haploidentical donors may be chosen in this order: children, male sibling, and father, mismatched sibling with non-inherited maternal antigen (NIMA), mismatched sibling with non-inherited paternal antigen (NIPA), mother, and other collateral relatives. Donors with donor-specific anti-HLA antibodies (DSAs) median fluorescence intensity (MFI) >10,000 should also be avoided. ABO and CMV IgG serological status compatibility between donor and recipient are preferred. A KIR ligand match is preferred in HID-SCT following the Beijing protocol.

**Algorithm for MUD**

MUD requires HLA matching in high resolution. Nine to 10 loci matches are needed for HLA-A, B, C, DRB1, and DQ matches. Five to six simultaneous loci matches are needed for A, B, and DRB1 matches, or at least 8/10 loci matches.

**Algorithm for CB**

The TNC in CB is limited and should be considered based on HLA-typing, MNC, and primary disease. For malignant hematological diseases, ≥4/6 loci should be matched, with TNC > (2.5–4.0) × 10^7/kg (recipient weight) and CD34+ cells > (1.2–2.0) × 10^7/kg (recipient weight). For non-malignant hematological diseases, ≥5/6 loci should
be matched, with TNC > 3.5 × 10⁷/kg (recipient weight), and CD34+ cells > (1.7 × 10⁵/kg (recipient weight).

**General principle of mobilization**
Granulocyte colony-stimulating factor (G-CSF; 5 mg/kg of body weight per day for 5 days) was used to mobilize the BM and/or PB. The target mononuclear cell count was 6 × 10⁸/kg of recipient weight. Unmanipulated BM (harvested on day 4 after G-CSF) and/or PB stem cells (harvested on days 4 and 5 after G-CSF) were infused into the recipient on the day of collection.

**Conditioning regimens**
The myeloablative regimen (MAC) mBuCy regimen in MSD-HSCT and the mBuCy+ATG regimen in haplo-HSCT are the most popular in China and achieve remarkable results (Tables 3). RIC or intensified conditioning regimen is also used for subgroups of patients (Tables 4 and 5).

Gao et al. retrospectively analyzed the outcomes of allo-SCT in 82 patients with AML or MDS who were conditioned with BuCy or fludarabine, idarubicin, intravenous-busulfan, and cytarabine (FIBA). There was no significant difference in the 3-year OS or the relapse rate, but RIC with FIBA exhibited a lower incidence of severe aGVHD and lower NRM than the BuCy regimen [83]. Intensified conditioning for patients with refractory leukemia, introduced by the Nanfang group, may reduce the high leukemia cell burden and improve outcomes. Lie et al. used a combination of fludarabine, cytarabine, TBI, Cy, and etoposide for conditioning in the haplo-setting and demonstrated that intensified conditioning decreased the 5-year relapse rate from 33.9 to 27.3%, and it may be a better approach for refractory and acute leukemia of ambiguous lineage [84–86]. Idarubicin-intensified haplo-HSCT introduced by the Wuhan Union group improved the dismal prognosis of pre-transplant MRD and yielded a 3-year DFS of 47.3% [87].

Selected older patients (age > 50) with low HCT-CI (≤ 2) and good performance status tolerate myeloablative

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**Table 3 Traditional and modified myeloablative (MA) regimens**

| MAC          | Drug          | Dose(total)       | Schedule(d) | Donor type          |
|--------------|---------------|-------------------|-------------|---------------------|
| **Traditional** |               |                   |             |                     |
|              | Cy/TBI        | Cy 120 mg/kg      | – 6, – 5    | Allo-HSCT           |
|              |               | f-TBI 12 ~ 14 Gy  | – 3 ~ – 1   |                     |
|              | Bu/Cy         | Bu 16 mg/kg(iv) or 12.8/kg(iv) | –7 ~ – 4 | Allo-HSCT |
|              |               | Cy 120 mg/kg      | – 3, – 2   |                     |
| **Modified (PUPH)** |           |                   |             |                     |
|              | mBuCy         | Hu 80 mg/kg, divided in twice | – 10 | HLA-matched sibling HSCT |
|              |               | Ara-C 2 g/m²       | – 9         |                     |
|              |               | Bu 9.6 mg/kg(iv)   | – 8 ~ – 6   |                     |
|              |               | Cy 3.6 g/m²       | – 5, – 4   |                     |
|              |               | MeCCNU 250 mg/m²(po) | – 3   |                     |
|              | mCy/TBI       | Single TBI 770 cGy | – 6 | HLA-matched sibling HSCT |
|              |               | Cy 3.6 g/m²       | – 5, – 4   |                     |
|              |               | MeCCNU 250 mg/m²   | – 3         |                     |
|              | mBuCy+ATG     | Ara-C 4~ 8 g/m²    | – 10, – 9   | URD, CBT, HID-HSCT |
|              |               | Bu 9.6 mg/kg(iv)   | – 8 ~ – 6   |                     |
|              |               | Cy 3.6 g/m²       | – 5, – 4   |                     |
|              |               | ATG 10 mg/kg       | – 5 ~ – 2   |                     |
|              |               | Or ATG-F 40 mg/kg  | – 5 ~ – 2   |                     |
|              | mCy/TBI + ATG | TBI 770 cGy       | – 6         | URD-HSCT, HID-HSCT |
|              |               | Cy 3.6 g/m²       | – 5, – 4   |                     |
|              |               | MeCCNU 250 mg/m²   | – 3         |                     |
|              |               | ATG 10 mg/kg       | – 5 ~ – 2   |                     |
|              |               | Or ATG-F 40 mg/kg  | – 5 ~ – 2   |                     |

Cy cyclophosphamide, Bu busulfan, TBI total body irradiation, Hu hydroxyurea, Ara-C cytarabine, MeCCNU methyl CCNU, ATG anti-thymocyte globulin thymoglobuline, ATG-F rabbit anti-thymocyte globulin produced by Fresenius, allo-HSCT allogeneic hematological stem cell transplantation, URD unrelated donor, CBT cord blood transplantation, HID-HSCT haploidentical transplantation, PUPH Peking University People Hospital
| Conditioning regimen | Drug     | Dose (total) | Schedule(d) | Donor type |
|-----------------------|----------|--------------|-------------|------------|
| **International regimen** |          |              |             |            |
| Flu/Mel               | Flu      | 150 mg/m²    | − 7 − 3     | Allo-HSCT  |
|                       | Mel      | 140 mg/m²    | − 2, − 1    |            |
| Flu/Bu                | Flu      | 150 mg/m²    | − 9 − 5     | Allo-HSCT  |
|                       | Bu       | 8~ 10 mg/kg (po) | − 6 − 4   |            |
| Flu/Cy                | Flu      | 150 mg/m²    | − 7 − 3     | Allo-HSCT  |
|                       | Cy       | 140 mg/m²    | − 2, − 1    |            |
| Flu/Bu/TT             | Flu      | 150 mg/m²    | − 7 − 5     | Allo-HSCT  |
|                       | Bu       | 8 mg/kg (po) | − 6 − 4     |            |
|                       | Thiotepa | 5 mg/kg      | − 3         |            |
| TBI/Cy/ATG            | TBI      | 4 Gy         | − 5         | Flu+Ara-C+AMSA, followed by allo-HSCT |
|                       | Cy       | 120 mg/kg    | − 4, − 3    |            |
|                       | ATG      |              |             |            |
| **Modified regimen (PUPH)** |          |              |             |            |
| RIC-mBuCy             | Hu       | 80 mg/kg (divided in two) | − 10 | HLA-matched sibling HSCT |
|                       | Ara-C    | 2 g/m² (CI) | − 9         |            |
|                       | Bu       | 4.8 mg/kg (iv) | − 10, − 9 |            |
|                       | Cy       | 2.0 g/m²     | − 5, − 4    |            |
|                       | MeCCNU   | 250 mg/m² (po) | − 3     |            |
|                       | ATG      | 10 mg/m²     | − 5 − 2     |            |
|                       | or ATG-F | 40 mg/kg     | − 5 − 2     |            |
| RIC-BuFlu             | Hu       | 80 mg/kg (divided in two) | − 10 | HLA-matched sibling HSCT |
|                       | Ara-C    | 2 g/m² (CI) | − 9         |            |
|                       | Bu       | 9.6 mg/kg (iv) | − 8 − 6  |            |
|                       | Flu      | 150 mg/m²    | − 6 − 2     |            |
|                       | MeCCNU   | 250 mg/m²    | − 3         |            |
| RIC-mBuFluATG         | Ara-C    | 8 g/m³ (CI) | − 10/− 9    | HID-HSCT   |
|                       | Bu       | 9.6 mg/kg (iv) | − 8 − 6  |            |
|                       | Flu      | 150 mg/m²    | − 6 − 2     |            |
|                       | MeCCNU   | 250 mg/m²    | − 3         |            |
|                       | ATG      | 10 mg/kg     | − 5 − 2     |            |
|                       | or ATG-F | 40 mg/kg     | − 5 − 2     |            |
| RIC-mBuCyFlu+ATG      | Ara-C    | 8 g/m³ (CI) | − 10/− 9    | HID-HSCT   |
|                       | Bu       | 9.6 mg/kg (iv) | − 8 − 6  |            |
|                       | Flu      | 150 mg/m²    | − 6 − 2     |            |
|                       | Cy       | 2.0 g/m²     | − 5, − 4    |            |
|                       | MeCCNU   | 250 mg/m²    | − 3         |            |
|                       | ATG      | 10 mg/kg     | − 5 − 2     |            |
|                       | or ATG-F | 40 mg/kg     | − 5 − 2     |            |

Flu fludarabine, Mel melphan, Cy cyclophosphamide, Bu busulfan, TBI total body irradiation, Hu hydroxyurea, Ara-C cytarabine, MeCCNU methyl CCNU, ATG anti-thymocyte globulin thymoglobulin, ATG-F rabbit anti-thymocyte globulin produced by Fresenius, AMSA amacrine, allo-HSCT allogeneic hematological stem cell transplantation, URD unrelated donor, CBT cord blood transplantation, HID-HSCT haploidentical transplantation.
HID-HSCT with similar outcomes compared to younger adults [57]. HID-HSCT with reduced-intensity regimens (RIC) substituted with cyclophosphamide with Flu was feasible in patients over 60 years of age and produced similar engraftment and relapse rates as myeloablative conditioning [29].

BuCy+ATG is a novel protocol developed and verified for HID-HSCT for SAA patients. It has been used for salvage therapy and first-line therapy in pediatric and adult patients [32–34].

Recommendation: conditioning regimen
The definitions of MAC regimens and RIC are in accordance with EBMT [88].

1. MAC regimens
   a. ≥ 100 mg/kg or 3.6 mg/m² IV cyclophosphamide.
   b. ≥ 12 Gy TBI
   c. ≥ 16 mg/kg PO busulfan or 9.6 mg/kg IV busulfan

2. RIC regimens
   a. 90–160 mg/m² IV fludarabine
   b. 6–9 mg/kg oral busulfan (or equivalent dose of IV busulfan)
   c. 2–8 Gy TBI
   d. 80–140 mg/m² IV melphalan
   e. 5–10 mg/kg IV thiotepa

For patients with malignant hematological diseases

1. For patients with leukemia/MDS:
   ① Standard-intensity conditioning: MAC regimens include traditional TBICy, BuCy, and its modified regimens, and ATG is used in alternative donor transplantations at different doses. ATG (Thymoglobuline, Sanofi-Genzyme, Lyon, France) is used in a dose range of 6–10 mg/kg, and ATG-F (Grafalon, Neovii, Bad Homburg, Germany) is used in dose range of 20–40 mg/kg. ATG was recently used in MSD-HSCT to prevent GVHD. ② RIC: Fludarabine-containing regimens or ATG-included RIC regimens are used commonly (listed in Table 4). ③ Intensified regimens: Intensified regimens generally include the addition of a drug, such as Ara-C, VP16, Melphalan, TBI, Fludarabine, or Tespamin, to standard condition regimens. It is primarily used for refractory and relapsed malignant patients (Table 5).

The optimal conditioning regimen for a patient should be selected based on the type and status of the disease, comorbidities, underlying conditions, and donor source. For example, a standard-intensity conditioning regimen is used for younger patients (younger than 55 years old), and RIC regimens are used for patients older than 55 years of age and patients with poor organ function or HSCT-CI ≥ 3, regardless of age. Intensified regimens are used for young patients with refractory and relapsed diseases. Intensified regimens may reduce the incidence of relapse to some extent, but they often increase TRM. Therefore, these regimens may not significantly increase overall survival. RIC regimens are often better tolerated but require immunosuppression agents and cell therapy to reduce the relapse risk. Therefore, RIC regimens are not recommended as the initial choice for patients with malignant hematological diseases.
combined with other regimens, such as fludarabine. HSCT with standard intensification conditioning regimens followed by immunosuppression adjustment or cell therapy to enhance the graft versus leukemia (GVL) effect is also feasible for refractory and relapsed patients.

2. For patients with malignant hematological diseases other than leukemia/MDS: Conditioning protocols, such as BEAM or Flu/Mel or Flu/Bu, are generally used for patients with MM or NHL (listed in Table 6). Myeloablative (MA) regimens of leukemia may also be used for patients with MM or NLH, such as BuCy, TBICy, or the modified BuCy regimen adopted by the Peking University People’s Hospital.

For patients with non-malignant hematological diseases

1. For patients with SAA: The Cy-ATG regimen is used for HLA-matched sibling transplantation, and the FluCy-ATG regimen is used for unrelated

### Table 6 Conditioning regimens for multiple myeloma and lymphoma

| Conditioning regimen | Drug | Dose (total) | Time(d) | Transplantation type |
|----------------------|------|--------------|---------|----------------------|
| BEAM                 | BCNU | 300 mg/m²    | −6      | Allo-HSCT of lymphoma |
|                      | Vp16 | 800 mg/m²    | −5 ~ − 2|                      |
|                      | Ara-C | 800 mg/kg  | −5 ~ − 2|                      |
|                      | Mel  | 140 mg/m²    | −1      |                      |
| Flu/MEL             | Flu  | 150 mg/m2    | −7 ~ − 3| Allo-HSCT of multiple myeloma |
|                      | Mel  | 140 mg/m²    | −2, −1  |                      |
|                      | Bortizomib |         |         |                      |
| Flu/Bu              | Flu  | 150 mg/m2    | −10 ~ 6 | Allo-HSCT of multiple myeloma |
|                      | Bu   | 6.4 ~ 9.6 mg/kg (iv) | − 7 ~ − 4|                      |

BCNU carmustine, VP16 etoposide, Ara-C cytarabine, Mel melphan, Flu fludarabine, Bu busulfan, allo-HSCT allogeneic hematological stem cell transplantation

### Table 7 Conditioning regimens for severe aplastic anemia

| Conditioning regimen | Drug | Dose (total) | Time(d) | Transplantation type |
|----------------------|------|--------------|---------|----------------------|
| International regimen| Cy-ATG | Cy | 200 mg/kg | −5 ~ − 22 | HLA-matched sibling HSCT |
|                      |     | ATG | 11.25 ~ 15.00 mg/kg | −5 ~ 3, − 2 |
| FluCy-ATG           | Flu  | 120 mg/m² | −5 ~ − 2 | HLA-matched unrelated HSCT |
|                      | Cy   | 120 mg/kg | −5, −2  |                      |
|                      | ATG  | 11.25 ~ 15.00 mg/kg | −5 ~ 3, − 2 |
| Regimens in China   | mBuCyATG-SAA | Bu | 6.4 mg/kg(ivgtt) | −7, − 6 | Haploidentical-HSCT |
|                      |      | Cy | 200 mg/kg | −5, − 2 |
|                      |      | ATG | 10 mg/kg | −5 ~ − 2 |
|                      | or ATG-F | 40 mg/kg | −5 ~ − 2 |
| mBuCyFluATG         | Bu   | 6.4 mg/kg(ivgtt) | −7 ~ − 6 | Haploidentical-HSCT |
|                      | Flu  | 120 mg/m² | −10 ~ 7 |                      |
|                      | Cy   | 200 mg/kg | −6 ~ 3  |                      |
|                      | ATG-F | 20 mg/kg | −4 ~ − 1 |                      |
|                      | or ATG | 10 mg/kg | −4 ~ − 1 |                      |
| FluCy-ATG           | Flu  | 120 mg/m² | −5 ~ − 2 | Haploidentical-HSCT |
|                      | Cy   | 90 mg/kg | −3, − 2  |                      |
|                      | ATG  | 10 mg/kg | −5 ~ − 2  |                      |

Cy cyclophosphamide, ATG anti-thymocyte globulin thymoglobulin, ATG-F rabbit anti-thymocyte globulin produced by Fresenius, Flu fludarabine, Bu busulfan, allo-HSCT allogeneic hematological stem cell transplantation
transplantations. There is no standard preparative regimen of choice for haploidentical transplantations. The most commonly used regimen in China is the BuCyATG protocol followed by T-replicate HID-HSCT (Table 7).

2. For patients with thalassemia: Intensified conditioning regimens instead of standard conditioning regimens for leukemia are generally used in patients with thalassemia (Table 8).

3. Fanconi anemia: The FluCyATG regimen (Flu 150 mg/m², Cy 5–20 mg/kg/d × 4 days, and rabbit ATG 10 mg/kg) with or without low-dose TBI may be used for alternative donor transplantations.

### Conclusion and perspective

In conclusion, this consensus is based on the standard of care and available clinical evidence in China. However, we recognize the limitation that some developmental indications and treatment options are also included because prospective clinical studies demonstrate that allo-HSCT is a promising treatment option compared to other non-HSCT strategies. Randomized prospective controlled trials are absent in most conditions because transplant decisions are complex issues. We recognize the need to periodically update these recommendations to keep abreast with ongoing research. In summary, we hope these recommendations developed by Chinese doctors inspire the refining of global clinical practice.

### Abbreviations

- aGVHD: Acute graft-versus-host disease
- allo-HSCT: Allogeneic hematopoietic stem cell transplantation
- AP: Accelerated phase
- ASBMT: American Society for Blood and Marrow Transplantation
- ATG: Anti-thymocyte globulin
- BC: Blastic crisis
- BCSH: British Committee for Standards in Haematology
- CB: Cord blood
- CBMTR: Chinese Blood and Marrow Transplantation Registry
- DSA: Donor-specific anti-HLA antibodies
- EBMT: European Group for Blood and Marrow Transplantation
- FFS: Failure-free survival
- GF: Primary graft failure
- GVL: Graft versus leukemia
- HID-HSCT: Haploidentical HSCT
- HLA: Human leukocyte antigen
- MAC: Myeloablative conditioning
- MFI: Median fluorescence intensity
- MMF: Mycophenolate mofetil
- MRD: Minimal residual disease
- MSD: HLA-matched sibling donor
- MUD: HLA-matched unrelated donors
- NCCN: National Comprehensive Cancer Network
- NIMA: Non-inherited maternal antigen
- NIPA: Non-inherited paternal antigen
- PT-CY: Posttransplant cyclophosphamide
- RIC: Reduced-intensity regimens
- SAA: Severe aplastic anemia
- TKIs: Tyrosine kinase inhibitors

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### Availability of data and materials

Data sharing not applicable to this article as no datasets were generated or analyzed during the current study.

### Authors’ contributions

All authors reviewed the literature and wrote first drafts of specific sections. XLP and HXJ assembled the sections and wrote the final version of the manuscript. all authors read and approved the final manuscript.

### Table 8 Conditioning regimens for thalassemia

| Conditioning regimen | Drug | Dose | Time(d) | Transplantation |
|----------------------|------|------|---------|-----------------|
| **General intensity** | | | | |
| Regimens for leukemia | The same as leukemia | Same as leukemia | | Allo-HSCT |
| BuCy | Bu | 14 mg/kg (po) | | Allo-HSCT |
| Cy | | 200 mg/kg | | |
| Intensified regimen | | | | |
| NF0-8-TM | Cy | 110 mg/kg | −10, −9 | HLA-matched sibling transplantation |
| Flu | 200 mg/m² | −8 ~ − 4 | | Unrelated HLA-matched transplantation |
| TT | 10 mg/kg | | −5 | |
| Bu | ivgtt, QD (−8d) | Css 300 ~ 600 ng/L | −8 ~ − 6 | Haploidentical-HSCT |
| Azathioprine | 3 mg/kg, QD | | | From D ~ 45 |
| Hu | 30 mg/kg, QD | | | From D ~ 45 |
| FluBuCyATG | Flu | 150mg/m² | −12 ~ − 10 | Allo-HSCT |
| Bu | 12.8 mg/kg (ivgtt) | −9 ~ − 6 | | |
| Cy | 200 mg/kg | | −5 ~ − 2 | |
| ATG | 10 mg/kg | | −5 ~ − 2 | |
| Hu | 20 mg/kg, QD | | | 3 months before |

*Bu busulfan, Cy cyclophosphamide, Flu fludarabine, TT thiota, ATG anti-thymocyte globulin, allo-HSCT allogeneic hematological stem cell transplantation,Css steady state plasma concentration, Hu hydroxyurea, QD once a day*
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