Prenatal maternal alcohol exposure: diagnosis and prevention of fetal alcohol syndrome

Young Min Hur, MD\(^1\), Jiwon Choi, MD\(^1\), Sunwha Park, MD, PhD\(^1\), Sarah Soyeon Oh, PhD\(^2,3\), Young Ju Kim, MD, PhD\(^1,2\)

\(^1\)Department of Obstetrics and Gynecology, \(^2\)Fetal Alcohol Syndrome Prevention Center, Ewha Womans University Mokdong Hospital, \(^3\)Institute of Health Services Research, Yonsei University College of Medicine, Seoul, Korea

Fetal alcohol syndrome (FAS) is a developmental and congenital disorder characterized by neurocognitive impairment, structural defects, and growth restriction due to prenatal alcohol exposure. The estimated global prevalence of alcohol use during pregnancy is 9.8%, and the estimated prevalence of FAS in the general population is 14.6 per 10,000 people. In Korea, the estimated prevalence of alcohol use during pregnancy is 16%, and the prevalence of FAS is 18-51 per 10,000 women, which is higher than the global prevalence. Women’s alcohol consumption rates have increased, especially in women of childbearing age. This could increase the incidence of FAS, leading to higher medical expenses and burden on society. Alcohol is the single most important teratogen that causes FAS, and there is no safe trimester to drink alcohol and no known safe amount of alcohol consumption during pregnancy. Thus, physicians should assess women’s drinking patterns in detail and provide education on FAS to women by understanding its pathophysiology. Moreover, the prevention of FAS requires long-term care with a multidisciplinary approach.

Keywords: Fetal alcohol syndrome; Fetal alcohol spectrum disorders; Pregnancy; Alcohols

Introduction

Fetal alcohol syndrome (FAS) is a congenital fetal disorder caused by maternal alcohol consumption during pregnancy. Children with FAS present with characteristic facial features, growth retardation, and intellectual disability and may even have difficulties adjusting to society later by fetal programming [1-3]. Alcohol can affect fetal development in various ways, including any amount of alcohol consumed from 3 months before conception to the end of pregnancy. Some children are born with all the features of FAS, whereas others have only some malformations, especially abnormalities of the central nervous system (CNS).

The global prevalence of alcohol use during pregnancy is reported to be 9.8%, and the estimated prevalence of FAS in the general population is 14.6 per 10,000 people (Table 1) [4-7]. Moreover, it is estimated that one in every 67 women who consumed alcohol during pregnancy would have a child with FAS, which translates to approximately 119,000 children born with FAS worldwide annually [4]. In Korea, the estimated prevalence of FAS is 0.18-0.51% in general schools and 14.9% in facilities for kids with mental retardation according to an epidemiological survey in 2012 [8]. According to Lee et al. [9], approximately 16% of women were reported to drink alcohol during pregnancy, and 1.7% reported binge drinking in Korea. Moreover, the annual drinking rate among Korean women (over 19 years old) was higher in 2015 (70.8%) than in 1998 (59.3%) according to data from the Korea Health Promotion Institute [1]. The increasing rate of child-bearing-aged women who consume alcohol can be considered to increase the chance of alcohol exposure during pregnancy.
whether intentionally or unintentionally. Thus, the potential prevalence of FAS may also increase in Korea.

Alcohol consumption during pregnancy is harmful. FAS was first described in French medical literature by Lemoine et al. in 1968 [10]. This was explained based on the abnormal findings observed in 127 children of alcoholic parents at the time [10]. Five years later, Jones et al. (1973) systemically described the association between maternal alcohol abuse and certain types of birth defects and provided the first diagnostic criteria for this condition [11]. This has always been cited in basic and clinical research articles that have identified alcohol-related malformations. In 2005, Hoyme et al. [12] published practical guidelines for operating the Institute of Medicine (IOM) categories, allowing for the standardization of fetal alcohol spectrum disorder (FASD) diagnoses in clinical settings [11]. The updated clinical guidelines for the diagnosis of FASD were published in 2016 [12]. They described four distinct diagnostic categories within FASD: FAS, partial fetal alcohol syndrome (PFAS), alcohol-related neurodevelopmental disorder (ARND), and alcohol-related birth defects (ARBD) (Table 2) [12].

**Definition and diagnosis of FAS**

FAS and FASD result from prenatal alcohol exposure and are related to physical malformations and intellectual disabilities [13,14]. Children with FAS have CNS abnormalities, pre- or postnatal growth impairment, and characteristic facial abnormalities [12,13]. CNS abnormalities include microcephaly, tremors, hyperactivity, lack of motor skills, deficits in attention, learning difficulties, intellectual or cognitive impairment, and seizures [12]. Facial abnormalities include short palpebral fissures, epicanthal folds, flat midface, hypoplastic philtrum, and a thin upper vermilion border (Fig. 1) [12,13]. Children with FAS are commonly diagnosed 48.3 months after birth [14]. However, it is often missed or misdiagnosed, preventing affected children from receiving the required services promptly [15].

Clinicians should consider a detailed history of prenatal alcohol exposure to diagnose FAS. Significant prenatal alcohol exposure that can affect the fetus is defined as at least 1 of the following documented findings: (a) 6 or more drinks per week for 2 or more weeks during pregnancy; (b) 3 or more drinks per occasion on 2 or more occasions during pregnancy; (c) alcohol-related social or legal problems during pregnancy; (d) intoxication during pregnancy documented by blood, breath, or urinary alcohol tests; (e) positive test for alcohol exposure biomarkers during pregnancy (fatty acid ethyl esters, phosphatidylethanol, and ethyl glucuronide in maternal hair, fingernails, urine, or blood, or in the placenta or meconium); and (f) increased prenatal risk associated with alcohol use during pregnancy, as assessed by a validated screening tool [12]. If there was no history of prenatal alcohol exposure during the 3 months before pregnancy recognition or at the time of a positive pregnancy test, FAS could be excluded. However, FAS cannot be ruled out if there is a history of alcohol exposure during pregnancy.

FASD is a broad diagnosis that includes FAS, PFAS, ARND, and ARBD according to IOM diagnostic criteria (Table 2) [12]. FAS is the most severe form of the FASD [14]. Another diagnostic strategy is a 4-digit diagnostic code created from

| Table 1. Global prevalence of alcohol use (any amount) during pregnancy FAS in the general population in 2012, by WHO region [4] |
|---------------------------------------------------------------|
| **Alcohol use during pregnancy (%)** | **FAS (per 10,000)** |
|------------------------------------|----------------------|
| AFR 10.0 (8.5-11.8) | 14.8 (8.9-21.5) |
| AMR 11.2 (9.4-12.6) | 16.6 (11.0-24.0) |
| EMR 0.2 (0.1-0.9) | 0.2 (0.2-0.9) |
| EUR 25.2 (21.6-29.6) | 37.4 (24.7-54.2) |
| SEAR 1.8 (0.9-5.1) | 2.7 (1.3-8.1) |
| WPR 8.6 (4.5-11.6) | 12.7 (7.7-19.4) |
| Worldwide 9.8 (8.9-11.1) | 14.6 (9.4-23.3) |

Values are presented as estimates (95% confidence interval).
FAS, fetal alcohol syndrome; WHO, World Health Organization; AFR, African region; AMR, American region; EMR, Eastern-Mediterranean region; EUR, European region; SEAR, South-East Asia region; WPR, western pacific region.
Table 2. Updated Institute of Medicine (IOM) diagnostic criteria for the diagnosis of fetal alcohol spectrum disorders [12]

1. FAS
   (With or without documented prenatal alcohol exposure)
   Requires all features, A-D:
   A. A characteristic pattern of minor facial anomalies, including ≥2 of the following: 1) short palpebral fissures; 2) thin vermilion border of the upper lip; 3) smooth philtrum
   B. Prenatal and/or postnatal growth deficiency: 1) height and/or weight ≤10th percentile
   C. Deficient brain growth, abnormal morphogenesis, or abnormal neurophysiology, including ≥1 of the following: 1) head circumference ≤10th percentile; 2) structural brain anomalies; 3) recurrent nonfebrile seizures (other causes of seizures having been ruled out)
   D. Neurobehavioral impairment

2. PFAS
   (With or without documented prenatal alcohol exposure)
   Requires features, A-B:
   A. A characteristic pattern of minor facial anomalies, including ≥2 of the following: 1) short palpebral fissures; 2) thin vermilion border of the upper lip; 3) smooth philtrum
   B. Neurobehavioral impairment

3. ARND
   Requires features A-B (this diagnosis cannot be made definitively in children <3 years of age):
   A. Documented prenatal alcohol exposure
   B. Neurobehavioral impairment

4. ARBD
   Requires features A-B:
   A. Documented prenatal alcohol exposure
   B. One or more specific major malformations demonstrated in animal models and human studies to be the result of prenatal alcohol exposure: cardiac: atrial septal defects, aberrant great vessels, ventricular septal defects, conotruncal heart defects; skeletal: radioulnar synostosis, vertebral segmentation defects, large joint contractures, scoliosis; renal: aplastic/hypoplastic/dysplastic kidneys, “horseshoe” kidneys/ureteral duplications; eyes: strabismus, ptosis, retinal vascular anomalies, optic nerve hypoplasia; ears: conductive hearing loss, neurosensory hearing loss

FAS, fetal alcohol syndrome; PFAS, partial fetal alcohol syndrome; ARND, alcohol-related neurodevelopmental disorder; ARBD, alcohol-related birth defect.

![Image of fetal alcohol syndrome features]

Fig. 1. Typical appearance associated with fetal alcohol syndrome (FAS) [12].
the clinical data of the Washington State Fetal Alcohol Syndrome Diagnostic and Prevention Network [16,17]. It reflects the severity of the 4 key diagnostic features of FAS: growth deficiency, FAS facial phenotype, CNS dysfunction, and gestational exposure to alcohol (Table 3) [17]. This is objective and can be useful for surveillance and research purposes. Moreover, Canadian guidelines exist for the diagnosis of FAS [18]. This is similar to the IOM diagnostic category, but the definition of partial FAS is different [18]. There are also other guidelines and checklists for the diagnosis of FASD [19,20]. Differential diagnoses for FASD should include various chromosomal abnormalities, exposure to other teratogenic substances, and behavioral and psychiatric diagnoses, for example, Turner’s syndrome, fragile X syndrome, William’s syndrome, Noonan’s syndrome, DiGeorge syndrome, Zikavirus infection, and diseases by other teratogens (hydantoin, valproate, etc.) [14,21-23].

Clinicians should also identify maternal high-risk groups for prenatal alcohol exposure according to their drinking patterns. Different patterns of alcohol consumption during pregnancy can have various effects on the fetus. Specifically, animal and human studies have shown that binge drinking is more detrimental to fetal development than constant drinking [24-27]. Binge drinking is defined as the consumption of 5 or more drinks on a single occasion (a standard drink is defined as approximately 14 g of pure alcohol) [24,28]. This is because a higher peak blood alcohol concentration worsens fetal brain damage and leads to prolonged alcohol exposure; therefore, metabolizing all the alcohol that has been consumed takes time [24]. Recently, several studies have objectively assessed the patterns of maternal alcohol consumption and identified infants who exhibit FAS-related deficits in growth by biological analysis [29-32].

**Teratogenicity of alcohol**

Alcohol is a potent teratogen [13]. Clinicians should consider the following in determining the teratogenicity of substances: completely characteristic defects, sufficient amounts to affect embryo or fetal development, exposure to critical developmental periods, and evidence of epidemiological investigation [33]. Various processes are involved in the effects of alcohol on the fetus-teratogenic mechanism [34-36]. These processes produce variable outcomes, including stillbirth, structural anomalies in infancy, and neurobehavioral disorders in adolescence (Fig. 2).

First, drinking alcohol and hypoxia are related, and hypoxia is a primary cause of cellular damage [37]. If a mother drinks alcohol and the alcohol is metabolized in the liver, the amount of oxygen in the circulation would significantly decrease. Thus, hypoxia may affect cell damage during fetal development, and this process can explain abortions related to alcohol exposure [37]. Specifically, it can affect the developing brain, such as the hippocampus and cerebellum, which are sensitive to hypoxia and alcohol exposure [37].

Second, free radicals can cause significant damage to cells because they are unstable and reactive [37]. They cause

| Rank | Growth deficiency | FAS facial phenotype | CNS damage or dysfunction | Gestational exposure to alcohol |
|------|-------------------|---------------------|--------------------------|--------------------------------|
| 4    | Significant: height and weight below 3rd percentile | Severe: all 3 features: PFL 2 or more SDs below mean; thin lip: rank 4 or 5; smooth philtrum: rank 4 or 5 | Definite: structural or neurologic evidence | High risk: confirmed exposure to high levels |
| 3    | Moderate: height and weight below 10th percentile | Moderate: generally 2 of the 3 features | Probable: significant dysfunction across 3 or more domains | Some risk: confirmed exposure. Level of exposure unknown or less than rank 4 |
| 2    | Mild: height or weight below 10th percentile | Mild: generally 1 of the 3 features | Possible: evidence of dysfunction, but less than rank 3 | Unknown: exposure not confirmed present or absent |
| 1    | None: height and weight at or above 10th percentile | Absent: none of the 3 features | Unlikely: no structural, neurologic or functional evidence of impairment | No risk: confirmed absence of exposure from conception to birth |

FAS, fetal alcohol syndrome; CNS, central nervous system; PFL, palpebral fissure length; SD, standard deviation.
oxidative stress and disrupt stable lipids, proteins, receptors, and chromosomes [37]. In general, fetal cells are more sensitive to oxidative stress because of their lower levels of antioxidants and related enzymes [38]. Specifically, craniofacial and visceral structures are derived from neural crest cells; this sensitivity could account for the characteristic malformations associated with FAS [38]. Facial dysplasia, a prominent feature of FAS, appears to occur when peak blood alcohol levels occur during the embryonic stage of prenatal development [39]. This was also explained by the fact that mice exposed to ethanol on embryonic day 7 or 8 exhibited FAS-related facial dysplasia [39].

Brain damage in infants with FAS includes microcephaly, agenesis of the corpus callosum and anterior commissure, and anomalies in the cerebellum and brainstem [40]. According to Olney [41], the administration of a single high dose of ethanol to neonatal rats significantly reduced the thickness of the corpus callosum. Alcohol disrupts the rapid growth of the brain in the 3rd trimester of gestational age, which is characterized by glial development, synaptogenesis, and development of the cerebellum [27,42]. Thus, prenatal alcohol exposure can lead to learning and memory deficits, as well as long-term and neurobehavioral dysfunction [42,43].

Investigating the use of other potential teratogens during pregnancy is also important [12,44]. Smoking and drinking are highly correlated; therefore, alcoholics are more likely to be heavy smokers [37]. Marijuana, cocaine, and caffeine are also significantly correlated with alcohol consumption during pregnancy [37]. Marijuana smoking can increase the level of carbon monoxide in the mother’s body, causing hypoxia and thus increasing the risk of FAS [37]. Cocaine causes uterine artery vasoconstriction and exacerbates hypoxic fetal conditions [37]. Prenatal caffeine exposure can exacerbate zinc deficiency and reinforce the effects of alcohol (zinc is an essential trace mineral important for cellular replication and protein synthesis and is a cofactor for endogenous antioxidative enzymes) [37].

Genetic susceptibility to alcohol exposure

Variations in the alcohol dehydrogenase (ADH) allele related to alcohol metabolism can influence the risk of alcohol-induced malformations in the fetus [42]. According to Warren and Li [45], the more efficient ADH allele ADH1B*3
protects against FASD, and ADH1B*2 reduces the risk of FAS compared with ADH1B*1. Differences in the isoforms of aldehyde dehydrogenase (ALDH), an enzyme that converts acetaldehyde to acetate, also influence the risk of FAS [37]. If the inactive form of ALDH (ALDH2*2), acetaldehyde, is not converted to acetate in the body, it accumulates and causes flushing, tachycardia, and nausea [37]. ALDH2*2, an inactive form of ALDH, is mainly present in Asian populations. Thus, we can consider racial differences in FAS severity, and the delayed breakdown of alcohol can affect the fetus and induce malformations. Asian mothers may need to be more educated on alcohol abstinence than others. Furthermore, studies have shown differential sensitivity to FAS in dizygotic twins and high concordance for FAS in monozygotic twins [37].

**Limitations of research on FAS**

Several methods can be employed to explore the teratogenicity of alcohol, and established medical databases can be used as valuable resources for retrospective cohort studies of FAS [46]. However, cohort studies that use past medical databases have systematic errors, such as selection bias, information bias (misclassification), and confounding factors [46]. Moreover, morphological evaluation for the diagnosis of FAS has limitations such as racial differences [11]. Questionnaires are also primarily used to obtain information on pregnant women about alcohol consumption before or during pregnancy. However, this estimate of alcohol use using questionnaires is often underreported because pregnant women are concerned about social stigma [31,47]. Therefore, many studies are being conducted to identify methods to objectively evaluate alcohol exposure in pregnant women [31,32,48,49]. Furthermore, assembling prospective cohorts to observe the occurrence of FAS is expensive and inefficient in terms of time, money, and resources because of the rarity of birth defects related to FAS [46]. Thus, a case-control design is usually used to study FAS; however, the results from such studies could be limited in terms of generalization [46].

Clinical trials for FAS in humans are also difficult. Pregnant women rarely participate in randomized studies on alcohol or other drugs, so there is a lack of data on the safety or prenatal drug exposure itself [46]. Moreover, there are clear limitations to animal experiments because alcohol metabolism is species specific. Therefore, new research methodologies have been investigated recently to verify the negative effects of substances by organizing and analyzing sporadic big data through machine learning [50].

**What should obstetrician-gynecologists do?**

FAS is a disease in which differential diagnosis is important. FAS-related features that can be identified by prenatal ultrasonography include fetal growth restrictions, microcephaly, and extremity and heart malformations. If characteristic features of FAS are seen during antenatal care, other common causes (e.g., infection, genetic factors, placental insufficiency, and other teratogens) should be differentially diagnosed first. If fetal growth restriction or microcephaly appears, the patterns and degrees of growth restriction should be documented [12]. It is necessary to suspect and evaluate maternal alcohol consumption in cases of fetal growth restriction with microcephaly. During the maternal interview, we should investigate not only maternal alcohol intake but also medical history, nutrition, husband's alcohol use, and home environment. According to several studies, women who have consumed alcohol during pregnancy have a low educational level, low rates of planned pregnancy, and a low level of knowledge related to the risks of drinking alcohol during pregnancy [9,34,51]. Low educational level and unplanned pregnancy are significant risk factors for alcohol consumption in pregnant women [9]. It is also important to consider the overall drinking pattern just before pregnancy recognition, as it is common for the drinking pattern before 3 months of pregnancy to continue into the 1st trimester [12]. Once maternal alcohol consumption is confirmed, physicians should provide objective information on FAS and educate patients on neutral grounds [52]. Above all, it is important for women to be aware of their overall drinking patterns. Women should be educated about immediate alcohol discontinuation once pregnancy is confirmed and encouraged to undergo antenatal care.

FASD is a developmental disorder with a specific phenotype requiring long-term management [34]. It has a high recurrence rate, and younger siblings tend to be more severely affected [34]. FAS recurrence would reach approximately 75% if mothers continue to drink alcohol in subsequent pregnan-
cies [34]. Thus, sustainable follow-up of the mother and child is required, and a multidisciplinary approach with pediatrics and psychiatric departments is important. If a baby with suspected FAS is born, obstetricians should hand over the clinical information to the pediatrician for continuous assessment and management. Psychiatrists must also participate in treatment until neurobehavioral problems appear in adolescence. Counselors can help potential patients avoid exposure during subsequent pregnancies by intervening in alcohol abuse. Furthermore, a cohort study linked to obstetrics and gynecology, pediatrics, and psychiatry should be conducted to prevent FAS by accumulating data on FAS mothers and children; an example is the Washington State Fetal Alcohol Syndrome Diagnostic & Prevention Network, which started at the University of Washington in Seattle in 1993 [53]. It was sponsored by the centers for disease control and prevention, began diagnosing patients in 1993 and has diagnosed over 3,000 patients to date [53]. They try to prevent FAS through screening, diagnosis, intervention, education, and research [53].

In this way, we should establish a big data cohort at the national and social levels that enables accurate diagnosis and sustainable follow-up of FAS to prevent FAS through a multidisciplinary approach. This multidisciplinary team for the prevention of FAS should consist of obstetricians, gynecologists, pediatricians, psychiatrists, biologic scientists, data professionals, and policymakers. An example of such a multidisciplinary team is the FAS prevention center located at the Ewha Womans University Mokdong Hospital in Korea. It is the first FAS-specialized center in Korea, consisting of physicians, biologic scientists, data professionals, etc. They work together to conduct research on FAS, in addition to counseling and campaigning to prevent FAS, sending missions to developing countries, and publishing books for public education. Moreover, the Korean Mothersafe Counseling Center has been running a campaign regarding abstinence from alcohol and other teratogens for pregnant women.

Conclusion

Alcohol is the single most important factor in FAS, and there is no safe trimester or known safe amount to drink alcohol during pregnancy [12]. FAS is a developmental disorder that can be prevented. Physicians must accurately obtain the medical history of the mother and provide safe antenatal care and education for FAS once maternal alcohol consumption is confirmed. If the prevalence of FAS is decreased, it will be possible to reduce the financial burden on society and unrealized human suffering [12].

Conflict of interest

No potential conflict of interest relevant to this article was reported. Young Ju Kim has been an Editorial Board of Obstetrics & Gynecology Science; however, she was not involved in the peer reviewer selection, evaluation, or decision process of this article. Otherwise, no other potential conflicts of interest relevant to this article were reported.

Ethical approval

None.

Patient consent

There is no need for patient consent in this review article.

Funding information

This study was supported by funding from the National Research Foundation of Korea (NRF-2020R1A2C3011850), and BK21 FOUR (Fostering Outstanding Universities for Research) was funded by the Ministry of Education and the NRF.

References

1. Seo BA, Kim SG, Huh SY, Lee DH, An SH, Lee SY, et al. Changes of drinking behavior in Korean pregnancy women for the last 20 year. J Public Health (Oxf) 2021;43:e632-6.  
2. Öztürk HNO, Türker PF. Fetal programming: could intruterin life affect health status in adulthood? Obstet Gynecol Sci 2021;64:473-83.  
3. Lee S, Kwon EJ, You YA, Du JE, Jo I, Kim YJ. Long-term...
effects of pro-opiomelanocortin methylation induced in food-restricted dams on metabolic phenotypes in male rat offspring. Obstet Gynecol Sci 2020;63:239-50.

4. Popova S, Lange S, Probst C, Gmel G, Rehm J. Estimation of national, regional, and global prevalence of alcohol use during pregnancy and fetal alcohol syndrome: a systematic review and meta-analysis. Lancet Glob Health 2017;5:e290-99.

5. Persson A, Lindmark S, Petersson K, Gabriel E, Thorsell M, Lindström K, et al. Alcohol and illicit and non-medical prescription drug use before and during pregnancy in Stockholm, Sweden: a cross-sectional study. Sex Reprod Healthc 2021;29:100622.

6. Miyake Y, Tanaka K, Okubo H, Sasaki S, Arakawa M. Alcohol consumption during pregnancy and birth outcomes: the Kyushu Okinawa maternal and child health study. BMC Pregnancy Childbirth 2014;14:79.

7. May PA, Blankenship J, Marais AS, Gossage JP, Kalberg WO, Barnard R, et al. Approaching the prevalence of the full spectrum of fetal alcohol spectrum disorders in a South African population-based study. Alcohol Clin Exp Res 2013;37:818-30.

8. Lee HG. Development of diagnostic system for fetal alcohol spectrum disorders (2011E6100200). cheongju: Korea Disease Control and Prevention Agency; c2011. [cited 2022 Apr 24]. Available from: https://nhf.go.kr/board.es?mid=a40801000000&cbid=0050&act=view&list_no=20485.

9. Lee SH, Shin SJ, Won SD, Kim EJ, Oh DY. Alcohol use during pregnancy and related risk factors in Korea. Psychiatry Investig 2010;7:86-92.

10. Lemoine P, Harousseau H, Borteyru JP, Menuet JC. Children of alcoholic parents--observed anomalies: discussion of 127 cases. Ther Drug Monit 2003;25:132-6.

11. Calhoun F, Warren K. Fetal alcohol syndrome: historical perspectives. Neurosci Biobehav Rev 2007;31:168-71.

12. Hoyme HE, Kalberg WO, Elliott AJ, Blankenship J, Buckley D, Marais AS, et al. Updated clinical guidelines for diagnosing fetal alcohol spectrum disorders. Pediatrics 2016;138:e20154256.

13. Cunningham FG, Leveno KJ, Bloom SL, Dashe JS, Hoffman BL, Casey BM, et al. Williams obstetrics. 25th ed. New York (NY): McGraw-Hill; 2018.

14. Denny L, Coles S, Blitz R. Fetal alcohol syndrome and fetal alcohol spectrum disorders. Am Fam Physician 2017;96:515-22.

15. Moberg DP, Bowser J, Burd L, Elliott AJ, Punykio J, Wilton G. Fetal alcohol syndrome surveillance: age of syndrome manifestation in case ascertainment. Birth Defects Res A Clin Mol Teratol 2014;100:663-69.

16. Astley SJ, Clarren SK. Diagnostic guide for fetal alcohol syndrome and related conditions: the 4-digit diagnostic code. Seattle (WA): University of Washington; 1999.

17. Astley SJ, Clarren SK. Diagnosing the full spectrum of fetal alcohol-exposed individuals: introducing the 4-digit diagnostic code. Alcohol Alcohol 2000;35:400-10.

18. Chudley AE, Conry J, Cook JL, Loock C, Rosales T, LeBlanc N. Fetal alcohol spectrum disorder: Canadian guidelines for diagnosis. CMAJ 2005;172(5 suppl):S1-21.

19. Cook JL, Green CR, Lilley CM, Anderson SM, Baldwin ME, Chudley AE, et al. Fetal alcohol spectrum disorder: a guideline for diagnosis across the lifespan. CMAJ 2016;188:191-7.

20. Burd L, Klug MG, Li Q, Kerbeshian J, Martsolf JT. Diagnosis of fetal alcohol spectrum disorders: a validity study of the fetal alcohol syndrome checklist. Alcohol 2010;44:605-14.

21. Douzgou S, Breen C, Crow YJ, Chandler K, Metcalfe K, Jones E, et al. Diagnosing fetal alcohol syndrome: new insights from newer genetic technologies. Arch Dis Child 2012;97:812-17.

22. Adams DJ, Clark DA. Common genetic and epigenetic syndromes. Pediatr Clin North Am 2015;62:411-26.

23. Adibi JJ, Marques ETA Jr, Cartus A, Beigi RH. Teratogenic effects of the Zika virus and the role of the placenta. Lancet 2016;387:1587-90.

24. Maier SE, West JR. Drinking patterns and alcohol-related birth defects. Alcohol Res Health 2001;25:168-74.

25. Bonthius DJ, West JR. Alcohol-induced neuronal loss in developing rats: increased brain damage with binge exposure. Alcohol Clin Exp Res 1990;14:107-18.

26. Strandberg-Larsen K, Nielsen NR, Grønbaek M, Andersen PK, Olsen J, Andersen AM. Binge drinking in pregnancy and risk of fetal death. Obstet Gynecol 2008;111:602-9.

27. Gursky ZH, Savage LM, Klintsova AY. Executive functioning-specific behavioral impairments in a rat model of human third trimester binge drinking implicate prefrontal-thalamo-hippocampal circuitry in fetal alcohol spectrum disorders. Behav Brain Res 2021;405:113208.
28. National Institute on Alcohol Abuse and Alcoholism. What is a standard drink? [Internet]. Bethesda (MD): National Institute on Alcohol Abuse and Alcoholism; c2021 [cited 2021 Aug 26]. Available from: https://www.niaaa.nih.gov/alcohols-effects-health/overview-alcohol-consumption/what-standard-drink.

29. Breunis LJ, Wassenaar S, Sibbles BJ, Aaldriks AA, Bijma HH, Steegers EA, et al. Objective assessment of alcohol consumption in early pregnancy using phosphatidylethanol: a cross-sectional study. BMC Pregnancy Childbirth 2021;21:1-7.

30. Maugeri A, Barchitta M, Magnano San Lio R, La Rosa MC, La Mastra C, Favara G, et al. The effect of alcohol on telomere length: a systematic review of epidemiological evidence and a pilot study during pregnancy. Int J Environ Res Public Health 2021;18:5038.

31. Adler J, Rissmann A, Kropf S, Mohnicke K, Taneva E, Ansorge T, et al. Estimated prevalence of harmful alcohol consumption in pregnant and nonpregnant women in Saxony-Anhalt (NorthEast Germany) using biomarkers. Alcohol Clin Exp Res 2021;45:819-27.

32. Mahnke AH, Sideridis GD, Salem NA, Tseng AM, Carter RC, Dodge NC, et al. Infant circulating microRNAs as biomarkers of effect in fetal alcohol spectrum disorders. Sci Rep 2021;11:1429.

33. Shepard TH. Annual commentary on human teratogens. Teratology 2002;66:275.

34. Paintner A, Williams AD, Burd L. Fetal alcohol spectrum disorders--implications for child neurology, part 2: diagnosis and management. J Child Neurol 2012;27:355-62.

35. Kim JY, Lee DY, Lee YJ, Park KJ, Kim KH, Kim JW, et al. Chronic alcohol consumption potentiates the development of diabetes through pancreatic β-cell dysfunction. World J Biol Chem 2015;6:1-15.

36. Lee YJ, Kim JY, Lee DY, Park KJ, Kim GH, Kim JE, et al. Alcohol consumption before pregnancy causes detrimental fetal development and maternal metabolic disorders. Sci Rep 2020;10:10054.

37. Abel EL, Hannigan JH. Maternal risk factors in fetal alcohol syndrome: provocative and permissive influences. Neurotoxicol Teratol 1995;17:445-62.

38. Davis WL, Crawford LA, Cooper DJ, Farmer GR, Thomas DL, Freeman BL. Ethanol induces the generation of reactive free radicals by neural crest cells in vitro. J Craniofac Genet Dev Biol 1990;10:277-93.

39. Sulik KK. Genesis of alcohol-induced craniofacial dysmorphism. Exp Biol Med (Maywood) 2000;230:366-75.

40. Jones KL, Smith DW. Recognition of the fetal alcohol syndrome in early infancy. Lancet 1973;302:999-1001.

41. Olney J. Fetal alcohol syndrome at the cellular level. Addict Biol 2006;9:137-49.

42. Guerri C, Bazinet A, Riley EP. Foetal alcohol spectrum disorders and alterations in brain and behaviour. Alcohol Alcohol 2009;44:108-14.

43. Roediger DJ, Krueger AM, de Water E, Mueller BA, Boys CA, Hendrickson TJ, et al. Hippocampal subfield abnormalities and memory functioning in children with fetal alcohol spectrum disorders. Neurotoxicol Teratol 2021;83:106944.

44. Bayih WA, Belay DM, Ayalew MY, Tassew MA, Chanie ES, Feleke DG, et al. The effect of substance use during pregnancy on neonatal outcomes in Ethiopia: a systematic review and meta-analysis. Heliyon 2021;7:e06740.

45. Warren KR, Li TK. Genetic polymorphisms: impact on the risk of fetal alcohol spectrum disorders. Birth Defects Res A Clin Mol Teratol 2005;73:195-203.

46. Ehrenstein V, Sørensen HT, Bakketeig LS, Pedersen L. Medical databases in studies of drug teratogenicity: methodological issues. Clin Epidemiol 2010;2:37-43.

47. Mullally A, Cleary BJ, Barry J, Fahey TP, Murphy DJ. Prevalence, predictors and perinatal outcomes of periconceptional alcohol exposure--retrospective cohort study in an urban obstetric population in Ireland. BMC Pregnancy Childbirth 2011;11:27.

48. Auriacombe M, Moriceau S, Serre F, Denis C, Micoulaud-Franchi JA, de Sevin E, et al. Development and validation of a virtual agent to screen tobacco and alcohol use disorders. Drug Alcohol Depend 2018;193:1-6.

49. O’Keeffe LM, Kearney PM, McCarthy FP, Khashan AS, Greene RA, North RA, et al. Prevalence and predictors of alcohol use during pregnancy: findings from international multicentre cohort studies. BMJ Open 2015;5:3006323.

50. Lee JY, Lee YS, Kim DH, Lee HS, Yang BR, Kim MG. The use of social media in detecting drug safety-related new black box warnings, labeling changes, or withdrawals: scoping review. JMI R Public Health Surveill 2021;7:e30137.

51. May PA, Baete A, Russo J, Elliott AJ, Blankenship J, Kalberg WO, et al. Prevalence and characteristics of fetal
alcohol spectrum disorders. Pediatrics 2014;134:855-66.
52. Jasper JD, Goel R, Einarson A, Gallo M, Koren G. Effects of framing on teratogenic risk perception in pregnant women. Lancet 2001;358:1237-8.

53. FAS Diagnostic & Prevention Network. FAS DPN [Internet]. Seattle (WA): FAS Diagnostic & Prevention Network; c2021 [cited 2021 Aug 26]. Available from: https://depts.washington.edu/fasdpn.