Effects of Salvia miltiorrhiza extract on lung adenocarcinoma

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Abstract. Lung adenocarcinoma is the most common subtype of non-small cell lung carcinoma. Tanshinone I is an important fat-soluble component in the extract of Salvia miltiorrhiza that has been reported to inhibit lung adenocarcinoma cell proliferation. However, no studies have clearly demonstrated changes in lung adenocarcinoma gene expression and signaling pathway enrichment following Tanshinone I treatment. And it remains unclear whether salvianolate has an effect on lung adenocarcinoma. The present study downloaded the GSE9315 dataset from the Gene Expression Omnibus database to identify differentially expressed genes (DEGs) and the underlying signaling pathways involved after Tanshinone I administration in the lung adenocarcinoma cell line CL1-5. The results revealed that there were 28 and 102 DEGs in the low dosage group (0.01 and 0.10 µg/ml Tanshinone I) and medium dosage groups (1 and 10 µg/ml Tanshinone I), respectively. In the low dosage group, DEGs were mainly enriched in ‘positive regulation of T-helper cell differentiation’ and ‘protein complex’. In the medium dosage group, 102 DEGs were enriched in ‘MAPK cascade’ and ‘extracellular exosome’. Kyoto Encyclopedia of Genes and Genomes pathway analysis demonstrated enrichment of both groups in the PI3K-Akt signaling pathway. Furthermore, there were nine overlapping DEGs [ADP ribosylation factor-interacting protein 2, chemokine (C-X-C motif) ligand 6, SH2 domain-containing adaptor protein B, Src homology 2 domain-containing transforming protein1, collagen type V1 α1 chain, elastin, integrin subunit α, endoplasmic reticulum mannosyl-oligosaccharide 1,2-α-mannosidase and sterile α motif domain-containing 9 like] between the two groups, which serve to be potential targets for the treatment of lung adenocarcinoma. The present study also investigated the possible effects of salvianolate on lung adenocarcinoma in vivo using nude mouse xenograft models injected with the A549 cell line. The data revealed that salvianolate not only suppressed lung adenocarcinoma tumor growth of in nude mice, but also downregulated the expression levels of ATP7A and ATP7B, which are important proteins in the tumorigenesis and chemotherapy of lung adenocarcinoma. The present study provided evidence for the potential use of Salvia miltiorrhiza extract for treating lung adenocarcinomas in the clinic.

Introduction

Lung cancer is a type of malignant tumor that continues to be the leading cause of cancer-associated mortality worldwide (1). The estimated number of new cases and deaths from lung cancer worldwide in 2020 were 2,206,771 and 1,796,144, respectively (2). As a subtype of lung cancer, lung adenocarcinoma has the highest incidence of all types of lung cancer (3). Surgery, chemotherapy, radiotherapy, targeted therapies and combined therapies are the therapeutic strategies that are currently available (4). However, poor patient responses to therapy, individual differences, adverse side effects and resistance to chemotherapeutic agents mean that clinical difficulties remain for the treatment of lung adenocarcinoma (5-7).

Traditional Chinese medicine (TCM) has been reported to have anti-cancer effects in lung adenocarcinoma, osteosarcoma and other types of cancer by regulating the tumor microenvironment and enhancing host immune responses (8-10). In particular, the red sage Salvia miltiorrhiza (Danshen) has been reported to improve survival rate for patients with colon (11) and breast cancer (12). The chemical composition of Salvia miltiorrhiza includes two categories (13): i) Fat-soluble tanshine compounds, which are mainly comprised of Tanshinone I and Tanshinone IIA; and ii) water-soluble phenolic compounds, consisting mainly of salvianolate. Tung et al (14) found Tanshinone I to inhibit the proliferation of lung adenocarcinoma cell lines A549, CL1-0 and CL1-5, which was in turn more effective compared with Tanshinone II. However, to the best of our knowledge, no studies have clearly demonstrated changes in gene expression and pathway enrichment following Tanshinone I administration on lung adenocarcinoma. Furthermore, as a major water-soluble component of Salvia miltiorrhiza extract, it remains currently unclear whether salvianolate has an effect on lung adenocarcinoma.

ATPase copper transporting α and β (ATP7A and ATP7B) are heavy metal transporting P-type ATPases that function as...
copper efflux transporters to maintain cellular copper homeostasis (15). It has been previously shown that ATP7A/7B has effects on tumorigenesis (16), tumor cell differentiation (17) and platinum-based chemotherapy response (18,19) in breast, lung and ovarian cancer. In addition, previous studies suggest that ATP7A/7B are considered to be potential targets for the treatment of non-small cell lung cancer (NSCLC) (18,20,21). Therefore, present study investigated the effect of ATP7A/7B expression following salvianolate treatment. Numerous profiles in carcinogenesis and cancer progression have been screened following the development of microarrays and high-throughput sequencing. The present study analyzed the gene expression profile matrix file (GSE9315) (22) using a series of bioinformatics tools to identify hub genes and key pathways that are affected by Tanshinone I administration on lung adenocarcinoma. The effects of salvianolate were then investigated using a xenograft nude mouse model before the potential underlying mechanisms of action were assessed. The present study identified the underlying mechanisms of action and potential targets of the Salvia miltiorrhiza extract for the treatment of lung adenocarcinoma.

Materials and methods

Data source. The gene expression profile matrix file (GSE9315) was downloaded from the Gene Expression Omnibus (GEO) database (https://www.ncbi.nlm.nih.gov/geo/). GSE9315 contained five different concentrations of Tanshinone I (0.00, 0.01, 0.10, 1.00 and 10.00 µg/ml) in macrophage-conditioned medium (CM) and a control condition without CM in the lung adenocarcinoma cell line CL1-5. Its platform used was GPL5968 (NCHU_M&A_human 1152 cDNACHIP).

Identification of differentially expressed genes (DEGs) following different doses of Tanshinone I. GEO2R (https://www.ncbi.nlm.nih.gov/geo2r/) is an interactive web tool that was used to analyze the raw data and identify DEGs in the GSE9315 dataset. GSE9315 was divided into the following three groups: i) Control group (0 µg/ml in CM); ii) low dosage group (0.01 and 0.10 µg/ml in CM); and iii) medium dosage group (1 and 10 µg/ml in CM) (22). In the present study, log fold change (FC) >1 was used as the cut-off criteria to identify DEGs. P<0.05 was not used as a criterion for screening DEG since there was only one sample in the control group, which was not suitable for statistical analysis. A Venn diagram was used to identify overlapping DEGs in the low and medium dosage groups.

Gene Ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) enrichment analyses of DEGs. The Database for Annotation, Visualization and Integrated Discovery (DAVID; version 6.8; https://david.ncifcrf.gov/) (23) was used to perform the GO and KEGG pathway enrichment analyses. The results were visualized using ggplot2 package in R Studio (version 1.1.453; The R Foundation) (24). P<0.05 was considered to indicate a statistically significant difference.

Protein-protein interaction (PPI) network construction and module analysis. The Search Tool for the Retrieval of Interacting Gene (version 11.0; https://string-db.org/cgi/input.pl) database (25) was used to construct a functional PPI network of the DEGs. An interaction score of 0.4 was regarded as the cut-off criterion in the present study. The Molecular Complex Detection (MCODE) in Cytoscape software (version 3.6.0; https://cytoscape.org/) (26) was then used to find closely connected regions in the PPI network. The selection criteria were as follows: i) Degree Cut-off, 2; ii) node score cut-off, 0.2; iii) K-Core, 2; and iv) max depth, 100.

Cell culture. The human lung adenocarcinoma cell line A549 (Changsha Nanke Biotechnology Co., Ltd.; http://www.nkbio.cn) was cultured in RPMI-1640 medium ( Gibco; Thermo Fisher Scientific, Inc.) supplemented with 10% FBS (Gibco; Thermo Fisher Scientific, Inc.) at 37˚C with 5% CO2.

Establishment and drug administration in animal models. The present study was performed according to the guidelines of the Ethics Committee of Institutional Animal Care and Use in Central South University (27) and approved by the Medical Ethics Committee of Xiangya Hospital, Central South University (approval no. 2017121170; Changsha, China). A total of 30, 6-week-old nude mice (weight range, 16-18 g; sex, 1:1 ratio of males and females) purchased from Hunan SJIA Laboratory Animal Co., Ltd, were kept under specific-pathogen free conditions (sterile laminar flow chamber, room temperature 21-25˚C; humidity of 60-70%; 12 h light/dark cycle) with access to food and drinking water ad libitum at the Department of Laboratory Animals, Central South University. A549 cells in the logarithmic growth phase were harvested and resuspended in PBS at a density of 1x10⁶/ml. After being anesthetized by isoflurane inhalation (1.5%), the nude mice were injected subcutaneously into the left armpit in a cell suspension of 0.2 ml (2x10⁶ cells per mouse). The next day, the tumor volume of nude mice was measured, which was regarded as the first day after injection (day 1). The tumor volume was measured once every 3 days for 16 days (day 1, 4, 7, 10, 13 and 16). When the tumors became visible (day 4), 10 and 50 mg/kg salvianolate (Green Valley, Inc.) and normal saline (Sichuan Kelun Pharmaceutical Co., Ltd.) were injected into the tumor-adjacent tissue once every 3 days (day 4, 7, 10 and 13). Mice were sacrificed by cervical dislocation on day 16 following the final treatment of salvianolate and normal saline before the tumor tissues were collected for further research. The tumor volume was measured using the following equation: V (mm³) = a x b² x 0.52, where ‘a’ and ‘b’ were the longest and shortest diameters of the tumor. The tumor inhibition rate was calculated using the following equation: Inhibition rate (%) = (1-V_mean-control/V_mean-salvianolate) x100%. Lack of breathing or no basic vital signs were evaluated to ensure death after cervical dislocation. In the present study, ‘the animal exhibiting depression and hypothermia without anesthesia or sedation (dying)’ was regarded as the criterion for euthanizing the animals.

Hematoxylin and eosin (H&E) staining. The tumor tissues were fixed in 4% paraformaldehyde for 24 h at room temperature and embedded in paraffin. Tissue samples were then cut into 3-µm-thick slices and the slices were heated at 60˚C for 1.5 h. The slices were deparaffinized in xylene and rehydrated in graded ethanol solutions (100, 95, 80 and 70% ethanol.
Immunohistochemistry (IHC). Paraffin-embedded tumor tissues, as aforementioned, were cut into 4-µm-thick slices. Slices were dewaxed using xylene, rehydrated in a descending ethanol gradient and treated with 3% hydrogen peroxide for 10 min at room temperature to remove endogenous oxidases. The slices were then repaired using a citrate repairing solution (0.01 mol/l; pH 6.0; cat. no. C1010; Beijing Solarbio Science & Technology Co., Ltd.), heated at 100˚C for 1-3 sec, rinsed with water and stained with eosin for 3 min, all for 5 min at each concentration) at room temperature. After washing with water, slices were stained with hematoxylin for 2 min, rinsed with water, incubated in 0.5% HCl ethanol for 1-3 sec, rinsed with water and stained with eosin for 3 min, all at room temperature. Then the slices were dehydrated using 70, 80, 95 and 100% gradient ethanol and xylene for 5 min at room temperature, and sealed with neutral gum. The nuclei were blue-violet in color, whilst the cytoplasm and intercellular matrix were red under light microscopy when the histological changes were assessed (magnification, x400).

### Statistical analysis

All data were processed using SPSS 19.0 statistical software (IBM Corp.). One-way ANOVA with Bonferroni's correction was used for the comparison between the various groups. Data are presented as the mean ± SD. P<0.05 was considered to indicate a statistically significant difference. The number of repeats in each group is ≥3.

### Results

Identification of DEGs after low or medium dosage treatment of Tanshinone I. Based on the aforementioned threshold (|log FC| >1), 28 and 102 DEGs were identified in the low dosage group and the medium dosage groups, respectively (Table I). Notably, there were nine overlapping DEGs in the low and medium dosage groups (Fig. 1A), which were integrin subunit α (ITGA2), endoplasmic reticulum mannosyl-oligosaccharide 1,2-α-mannosidase (MAN1B1), SH2 domain containing adaptor protein B (SHB), elastin (ELN), Src homology 2 domain-containing transforming protein 1 (SHC1), collagen type VI α1 chain (COL6A1), ADP ribosylation factor interacting protein 2 (ARFIP2), chemokine (C-X-C motif) ligand 6 (CXCL6) and sterile α motif domain containing 9 like (SAMD9L). The visualized heatmaps of the DEGs in the individualized different dosage groups are presented in Fig. 1B and C, which showed that some genes were upregulated (such as CD9, IFNAR2 and PTGS2) and others were downregulated (such as MAPK1, SLC25A1 and KRAS). In the heatmaps, red and green represent upregulation and downregulation of gene expression, respectively. Analysis of overlapping DEGs in the low and medium dosage groups. There were nine overlapping DEGs in the low and medium dosage groups, which were ITGA2, MAN1B1, SHB, ELN, SHC1, COL6A1, ARFIP2, CXCL6 and SAMD9L. The visualization results of gene expression were displayed using a heatmap (Fig. 1D). Among these nine genes, eight were downregulated after four different dosages of Tanshinone I were administrated (shown as green in the heatmap), compared with the control group. Notably, the expression of these eight genes was not dose-dependent. Only one of the nine overlapping
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genes (CXCL6) was upregulated following Tanshinone I administration (shown as red in the heatmap). The GO functional enrichment and KEGG pathway analyses of each gene are presented in Table II, helping us understand the functional enrichment of each gene. For example, SHC1 was enriched in 'epidermal growth factor receptor (ErbB) signaling pathway' and COL6A1 was enriched in 'PI3K-Akt signaling pathway', indicating that Tanshinone I may treat lung adenocarcinoma by upregulating or downregulating these genes and their enriched pathways.

Figure 1. Venn diagram and heatmaps of DEGs in the different groups. (A) Venn diagram shows that there were nine overlapping DEGs in the low and middle dosage group, which contains 19 and 93 DEGs, respectively. Heatmap of DEGs (B) in the low dosage and (C) the medium dosage group. (D) Heatmap of the overlapping DEGs in the low and medium dosage groups. Red and green represent up- and downregulation of gene expression, respectively. DEGs, differentially expressed genes; Tan I, tanshinone I.
**GO functional enrichment analysis of DEGs.** Significant terms in the GO enrichment analysis are presented in Fig. 2A and B. The DEGs in the low dosage group (Fig. 2A) were particularly enriched in the biological processes (BP) including, ‘positive regulation of T-helper cell differentiation’, ‘positive regulation of smooth muscle cell proliferation’, ‘collagen catabolic process’, ‘angiogenesis’ and ‘cellular response to mechanical stimulus’. As for the cellular components (CC), the genes were enriched in ‘protein complex’, ‘extracellular region’, ‘extracellular space’ and ‘focal adhesion’. The molecular function (MF) results showed that DEGs were most enriched in ‘cysteine-type endopeptidase activator activity involved in apoptotic process’.

In the medium dosage group (Fig. 2B), BP genes were primarily enriched in the ‘MAPK cascade’, ‘in utero embryonic development’, ‘positive regulation of cell adhesion’, ‘signal transduction’ and ‘cell proliferation’. For CC, the DEGs were mainly enriched in ‘extracellular exosome’, ‘nuclear chromatin’, ‘cytoplasm’, ‘focal adhesion’ and ‘extracellular matrix’. MF genes were predominantly enriched in ‘kinase activity’, ‘identical protein binding’, ‘protein binding’, ‘double-stranded DNA binding’ and ‘chromatin binding’.

**KEGG pathway analysis of DEGs.** The KEGG pathway enrichment analysis suggested that the DEGs in the low dosage group were mainly enriched in ‘PI3K-Akt signaling pathway’, ‘Focal adhesion’, ‘Pathways in cancer’, ‘Bacterial invasion of epithelial cells’ and ‘Small cell lung cancer’ (Fig. 2C). As for the medium dosage group (Fig. 2D), the DEGs were mainly enriched in ‘PI3K-Akt signaling pathway’, ‘Pancreatic cancer’, ‘Sphingolipid signaling pathway’, ‘Pathways in cancer’ and ‘Proteoglycans in cancer’.

**PPI network construction, module analysis and hub genes identification.** As presented in Fig. 3A and B, the final PPI network of DEGs in the low dosage group was composed of 16 nodes and 22 edges after excluding the isolated nodes, whilst the PPI in the medium group contained 85 nodes and 201 edges. MCODE was used to analyze the key module (composed of genes with a relatively high degree of association) that consisted of five nodes and seven edges in the low dosage group (Fig. 3C) and two key modules in the medium dosage group (Fig. 3D). Module 1 consisted of nine nodes and 34 edges, where the genes were enriched in the KEGG pathway, including ‘Pancreatic cancer’, ‘Central carbon metabolism in cancer’ and ‘Glioma’ (data not shown). Module 2 consisted of six nodes and nine edges, where the genes were enriched in the KEGG pathways, including ‘ECM-receptor interaction’, ‘Focal adhesion’ and ‘PI3K-Akt signaling pathway’ (data not shown).

**Salvianolate attenuates xenograft tumors in nude mice.** Animal models were used to verify the effect of salvianolate on lung adenocarcinoma in vivo. In the present study, the maximum tumor diameter exhibited by any nude mouse was 10.8 mm and the specific data on the tumor volume and diameter over the entire experimental time period were shown in Table SI. The data in the present study revealed that salvianolate significantly reduced the tumor volume at
Table II. GO functional enrichment and KEGG and Genomes pathway analysis of overlapping genes in the low and medium dosage groups.

| Gene symbol | Category | ID         | Description                                                |
|-------------|----------|------------|------------------------------------------------------------|
| ARFIP2      | BP       | GO:0006928 | Movement of cell or subcellular component                  |
|             | BP       | GO:0007264 | Small GTPase mediated signal transduction                  |
| CC          | GO:0001726 | Ruffle     |
| CC          | GO:0005737 | Cytoplasm  |
| MF          | GO:0005515 | Protein binding |
| MF          | GO:0005525 | GTP binding |
| CXCL6       | BP       | GO:0002446 | Neutrophil mediated immunity                               |
|             | BP       | GO:0002690 | Positive regulation of leukocyte chemotaxis                |
| CC          | GO:0005576 | Extracellular region                                  |
| CC          | GO:0005615 | Extracellular space                                   |
| MF          | GO:0008009 | Chemokine activity                                    |
| MF          | GO:0008201 | Heparin binding                                       |
| SHB         | BP       | GO:0001525 | Angiogenesis                                              |
|             | BP       | GO:0006469 | Negative regulation of protein kinase activity             |
| CC          | GO:0005829 | Cytosol                                             |
| CC          | GO:0005886 | Plasma membrane                                       |
| MF          | GO:0001948 | Glycoprotein binding                                  |
| MF          | GO:0005070 | SH3/SH2 adaptor activity                              |
| SHC1        | BP       | GO:0000165 | MAPK cascade                                              |
|             | BP       | GO:0000187 | Activation of MAPK activity                               |
| CC          | GO:0005622 | Intracellular                                         |
| CC          | GO:0005759 | Mitochondrial matrix                                  |
| MF          | GO:0005068 | Transmembrane receptor protein tyrosine kinase adaptor activity |
| MF          | GO:0005088 | Ras guanyl-nucleotide exchange factor activity          |
| COL6A1      | BP       | GO:0001649 | Osteoblast differentiation                               |
|             | BP       | GO:0007155 | Cell adhesion                                             |
| CC          | GO:0005576 | Extracellular region                                   |
| CC          | GO:0005581 | Collagen trimer                                        |
| MF          | GO:0048407 | Platelet-derived growth factor binding                 |
| ELN         | BP       | GO:0007519 | Skeletal muscle tissue development                        |
|             | BP       | GO:0007585 | Respiratory gaseous exchange                             |
| CC          | GO:0005576 | Extracellular region                                   |
| CC          | GO:0005578 | Proteinaceous extracellular matrix                      |
| MF          | GO:0005201 | Extracellular matrix structural constituent             |
| MF          | GO:0005515 | Protein binding                                        |
| ITGA2       | BP       | GO:0001666 | Response to hypoxia                                      |
|             | BP       | GO:0002687 | Positive regulation of leukocyte migration                |
| CC          | GO:0005634 | Nucleus                                              |
| CC          | GO:0005886 | Plasma membrane                                       |
| MF          | GO:0001618 | Virus receptor activity                                |
| MF          | GO:0005178 | Integrin binding                                      |
| MAN1B1      | BP       | GO:0006491 | N-glycan processing                                     |
|             | BP       | GO:0008152 | Metabolic process                                        |
| CC          | GO:0005783 | Endoplasmic reticulum                                  |
| CC          | GO:0005789 | Endoplasmic reticulum membrane                         |
| MF          | GO:0004559 | Alpha-mannosidase activity                             |
| MF          | GO:0004571 | Mannosyl-oligosaccharide 1,2-alpha-mannosidase activity |
| SAMD9L      | BP       | GO:0034058 | Endosomal vesicle fusion                                |
|             | CC       | GO:0005769 | Early endosome                                          |
| KEGG_PATHWAY| /        | /          | /                                                          |
| KEGG_PATHWAY| /        | /          | /                                                          |

BP, biological processes; CC, cellular components; MF, molecular function; GO, Gene Ontology; KEGG, Kyoto Encyclopedia of Genes.
10, 13 and 16 days, compared with that in the control group (P<0.05; Fig. 4A). In addition, the inhibition rate of salvianolate on tumors was significantly increased in a dose-dependent manner (Fig. 4B). H&E staining showed that the salvianolate administration groups exhibited nuclear pyknosis and fragmentation, suggesting apoptosis, compared with that in the control group (Fig. 4C).

Salvianolate treatment downregulates the expression of ATP7A/7B. IHC staining was performed to detect the expression levels of ATP7A and ATP7B. IHC showed that salvianolate significantly decreased the expression levels of ATP7A and ATP7B (Fig. 5A and B).

Discussion

Salvia miltiorrhiza has been applied as a TCM since it has been reported to possess anticancer properties (11). Components of Salvia miltiorrhiza can be divided into fat-soluble compounds that are comprised of diterpenoid quinones and water-soluble compounds, such as hydrophilic phenolic acids (13). The present study mainly investigated the effects of Tanshinone I (a diterpenoid quinones compound) and salvianolate (a water-soluble compound) on lung adenocarcinoma.

Tanshinone I is a notable component of the fat-soluble compounds that can be isolated from Salvia miltiorrhiza (28). In the present study, hub genes and key pathways involved in lung adenocarcinoma following administration of different dosages of Tanshinone I were identified. In the low dosage group (0.01 and 0.10 µg/m Tanshinone I vs. control group) and the medium dosage group (1 and 10 µg/ml Tanshinone I vs. control group), 28 DEGs and 58 DEGs were identified, respectively. Notably, the KEGG pathway analysis indicated that DEGs in the two groups were both enriched in the PI3K/Akt signaling pathway. Previous studies showed that Tanshinone I induced apoptosis by inhibiting the PI3K/Akt/mTOR signaling pathway in ovarian cancer cells (29) and human breast cancer cell lines (30). Although there is some evidence to suggest that Tanshinone I can inhibit the proliferation of lung adenocarcinoma cell lines A549, CL1-0 and CL1-5 (14,22), further in vitro experiments...
are required to investigate the underlying mechanism and molecular target of Tanshinone I in lung adenocarcinoma.

Notably, nine overlapping DEGs were identified in the low and medium dosage groups, which warrant further study. ITGA2 is the α subunit of a transmembrane receptor that has been reported to regulate cell migration and differentiation (31). Previous studies have shown that ITGA2 expression was upregulated in several different types of cancer (32), including gastric (33) and breast cancer (34). The present study indicated that ITGA2 expression was downregulated by Tanshinone I in lung adenocarcinoma, which may be a potential target. MAN1B1 encodes α-1,2-mannosidase, which mediates protein glycosylation modification and glycoprotein polysaccharide hydrolysis (35). It has been reported that high expression levels of MAN1B1 were associated with poor prognosis in bladder cancer (36). However, the effects of MAN1B1 downregulation induced by Tanshinone I on responses in lung adenocarcinoma require further investigation. Other studies have shown that SHB knockdown increased the susceptibility of the SVR angiosarcoma cell line to cisplatin and staurosporine (37) whilst impairing the growth of tumors in mice injected with Lewis lung carcinoma cells or T241 fibrosarcoma cells (38). The present study demonstrated the association between Tanshinone I treatment and the downregulation of SHB in
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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions

HT, YL, JC and XL conceived and designed the study. HT and JM performed animal experiments and collected data. HT, YL and LC. JC and XL analyzed and interpreted the data. JC and XL revised the manuscript. JY and ZL interpreted the data and checked the revised version of the manuscript. All authors read and approved the manuscript.

Ethics approval and consent to participate

All animal research protocols were following with the guidelines of the Ethics Committee of Institutional Animal Care and Use in Central South University and approved by Medical Ethics Committee of Xiangya Hospital, Central South University (approval no. 2017121170; Changsha, China).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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