Occult HCV Infection (OCI) Diagnosis in Cirrhotic and Non-cirrhotic Naïve Patients by Intra-PBMC Nested Viral RNA PCR

Mohamed Darwish Ahmed Abd Alla*1, Saleh Ahmed Elibiary1, George Y. Wu2 and Mostafa Kamel El-Awady3

1Tropical Medicine Department, Faculty of Medicine, Al-Azhar University, Cairo, Egypt; 2Department of Medicine, Division of Gastroenterology-Hepatology, University of Connecticut Health Center, Farmington, CT, USA; 3Department of Microbial Biotechnology, National Research Center, Cairo, Egypt

Abstract

Background and Aims: Occult HCV infections (OCIs) include IgG antibody seronegative cryptogenic (COCIs), as well as seropositive secondary naïve (SNOCIs) and experienced (SEOCIs) cases. We used peripheral-blood-mononuclear-cell (PBMC)-PCR to evaluate COCIs and SNOCIs prevalence, serum HCV spontaneous disappearance (SCSD) in naïve cirrhotics and non-cirrhotics, intra-PBMC HCV-RNA strands in relation to cirrhosis density in naïve non-viremia cases, and HCV-RNA seroconversion after 1 year of solitary naïve intra-PBMC infection. Methods: The anti-HCV IgG antibody-positive naïve-patients (n = 785) were classified into viremic (n = 673) and non-viremic [n = 112, including non-cirrhotics (n = 55) and cirrhotics (n = 57)], and 62 controls without evidence of HCV-infection. Controls and post-HCV non-viremia cases (n = 62 + 112 = 174) were submitted to hepatic Fibroscan-Elastography evaluation. All subjects (n = 847) were screened for intra-PBMC HCV-RNA sense and antisense strands by nested-PCR. Results: Naïve-OCI cases (4.84%) that were diagnosed by PBMC-PCR significantly raised the total numbers of HCV-infection to 714 (p = 0.01). The percent positivity of SNOCIs (34.82%) was significantly higher than for asymptomatic-COCIs (3.125%, p = 0.0001). Comparing PBMC-PCR with single-step reverse-transcription (SRT)-PCR for identification of SCSD in naïve IgG antibody-positive non-viremia patients (n = 112) revealed a decline in SCSD prevalence by PBMC-PCR (from 14.27% to 9.3%), regardless of presence of hepatic cirrhosis (p = 0.03). SCSD was found to be higher by PBMC-PCR in non-cirrhotics compared to cirrhotics (p = 0.0001), with an insignificant difference when using SRT-PCR (p = 0.45). Intra-PBMC HCV-RNA infection was significantly more frequent in cirrhotics compared to both non-cirrhotics and controls (p < 0.0005). An increased hepatic fibrosis density was recognized in intra-PBMC HCV-RNA infection with sense (p = 0.0001) or antisense strand (p = 0.003). HCV-RNA seroconversion was associated with intra-PBMC infection when both sense and antisense strands were detected (p = 0.047).

Conclusions: Intraacellular HCV-RNA evaluation is crucial for diagnosing OCIs and addressing relapse probability.

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Abbreviations: COCIs, cryptogenic occult HCV infection; DAA, direct-acting antiviral; HCC, hepatocellular carcinoma; HCV, hepatitis C virus; OCIs, occult HCV infections; PBMCs, peripheral blood mononuclear cells; RAVs, resistance-associated variants; SCSD, serum HCV spontaneous disappearance; SEOCIs, secondary experienced occult HCV infections; SNOCIs, secondary naïve occult HCV infection; SRT, single-step reverse transcription; SVR, sustained virologic response.

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Introduction

In 2004, Pham et al.1 reported occult HCV infections (OCIs) in anti-HCV IgG antibody-positive naïve patients who had spontaneous recovery after self-limited HCV-infections and in experienced cases with sustained virologic response (SVR) after interferon treatment. During the same year, Castillo et al.2 described HCV-RNA existence in anti-HCV antibody-negative naïve patients who presented with active hepatocellular damage. Further refinement of the definitions of OCIs included: a) recognition of intracellular RNA strands in post-infection non-viremia naïve cases, which is termed spontaneous HCV disappearance (SCSD) and manifests as secondary naïve OCIs (SNOCIs); b) identification of post-treatment non-viremia cases, which have intracellular HCV-RNA strand infections, and manifest as secondary experienced OCIs (SEOCIs).3

Recently, baseline diagnosis and post-treatment follow-up of both SNOCIs and SEOCIs by peripheral blood mononuclear cell (PBMC) HCV-PCR in Egyptian populations addressed both validation of PBMC-PCR as a diagnostic test that can diagnose intracellular HCV when single-step reverse transcription (SRT)-PCR is negative and the association of intra-PBMC infections with liver cirrhosis in naïve non-viremia patients. The authors recommended eradication of intra-PBMC RNA-strands in SEOCIs to avoid HCV-RNA seroconversion and subsequent post-treatment relapse. A third category of OCIs is known as cryptogenic occult HCV-infection (COCIs), which is diagnosed in the IgG-seronegative population by detecting the intracellular RNA strands. Its prevalence is around 3.5% in asymptomatic populations, which might
have active replication of HCV inside PBMCs, as identified by
detection of either the viral NS3-protein or antisense-strand.5

In 1999, El-Awady et al. predicted that assay for viral replication
in PBMCs enhances sensitivity of diagnosing and
monitoring HCV associated hepatitis. Accordingly, intra-PBMC
detection of HCV-RNA by PCR is currently used to diagnose
OCIs in non-viremia patients.5 It is obvious that the immune
system is not only a part of an OCI patient’s reaction to the
infection but that it also serves as a host to the viral RNA,
thereby facilitating its replication, as indicated by antisense
HCV-RNA strands found in the PBMCs of these patients.8

Keeping in mind that HCV-infection is an important and treat-
able cause of liver disease and the fact that patients with crypto-
genetic liver diseases can potentially develop cirrhosis and
hepatocellular carcinoma, investigating the prevalence of
HCV and developing more accurate techniques to diagnose
OCIs in these patients are critically important.9

Some researchers10 have expressed doubts about using
SRT-PCR in post-treatment follow-up. They have concluded that HCV cure in individuals achieving a state of undetectable
viremia after an 8 to 12-week course of direct-acting antiviral
(DAA) therapy may not be entirely valid. The same research-
ners recommended careful longitudinal follow-up utilizing
highly sensitive assays and unique approaches to viral detec-
tion. Because PBMCs present a convenient extrahepatic home for
HCV adoption, translation, transcription, assembly and
finally release into the serum and/or other adjacent cells,6

intra cellular PCR can be used to further refine the definitions of
clinical presentations of spontaneous HCV clearance. Recogni-
tion of intra-PBMC HCV strands in SNOCIs5 should refer to
regression of using the ‘spontaneous HCV-clearance’ term. In
populations of HCV endemic areas, the SCSD is estimated to
be around 25%.11 However, cellular HCV spontaneous disap-
pearance has not been evaluated before the current report.

On the other hand, the post-treatment RNA genomic seroconversion has been attributed to either treatment failure
or new infection, and is known as HCV-relapse,12 which has not
been described in naive subjects who present with spontaneous
viremia because no temporal follow-up is recommen-
ded. Tilt now, naïve cirrhotic patients who present with non-
viremia, but have evidence of hepatocellular damage
and positive intra-PBMC HCV-RNA strand infection, have not
been considered as candidates for DAA therapy.2 However,
eradication of intra-PBMC HCV-infection is recommended in
naïve cirrhotics and experienced post-treatment patients,
respectively, because of association with cirrhotic changes
and probability of relapse in more than 18%.5

Considering the high cure rate of DAA therapy, physicians
are looking for effective treatments for COCIs and SNOCIs.
Patients with OCIs should be afforded the option of getting rid
of the intracellular HCV genomic materials, to prevent the
subsequent hepatic fibrosis.9 For this reason, the current
study was designed to shed more light on the OCI problem
among a population of cases from one of the most endemic
areas in the world for HCV-infection, in the hopes of promot-
ing attention to this unmet challenge to satisfy clinical prac-
tice demands.

The specific aims of the current study included the use of
PBMC-PCR in the evaluation of a) prevalence of asymptomatic
COCIs and SNOCIs, b) SCSD in naïve cirrhotics and non-
cirrhotics, c) intra-PBMC HCV-RNA sense and antisense
strand infections in relation to cirrhosis density in naïve
non-viremia cases, and d) rate of HCV-RNA seroconversion
after 1 year of solitary naïve intra-PBMC infection.

Methods

Study subjects

All study subjects (n = 847) were treatment-naïve and
outpatient-visitors to the Infectious Disease Clinic. All subjects
were screened by SRT-PCR between January 2015 and February
2017. Inclusion criteria were age between 18 and 70 years-old
and positivity for serum anti-HCV IgG antibodies; controls were
recruited from the population that was clinically and serologi-
cally free of HCV. Exclusion criteria included hepatocellular
carcinoma (HCC) and Child C classification. Ethical committee
approval for the study was obtained before patient enrollment
(Registration No. 10231, National Research Center).

Sample size in each group depended upon availability of
subjects that fulfill the inclusion criteria during the study
period. Study subjects (n = 847) included the following sub-
groups: anti-HCV IgG antibody-positive naïve patients (n = 7
who were classified as diagnostic cases (n = 673) and
post-HCV non-viremic cases (n = 112), with the last
group further divided into non-cirrhotic (n = 55) and cirrhotic
(n = 57) subgroups. Controls included 62 participants without
evidence of HCV-infection. Controls and post-HCV non-
viremic cases (n = 174) submitted to hepatic elastography
evaluation by Fibroscan. All subjects (n = 847) were screened
for presence of intra-PBMC HCV-RNA sense and antisense
strands by nested-PCR.

ELISA for HCV IgG antibody detection

The procedure was performed as described.13 The third-
generation ELISA contained reconfigured NS3 and core anti-
gen and in addition a newly incorporated antigen from the
NS5 region. This assay was used as a preliminary screening
test to enroll cases in the current study.

Real-time PCR for quantification of HCV RNA

The collection and transportation of specimens, RNA isolation,
SRT-PCR procedure, internal control of the isolated RNA and/
or contamination, and quantification of the HCV PCR were all
performed as described by Abd Alla and El-Awady7 in 2017.

Amplification of intracellular HCV RNA genomes by
strand-specific RT-PCR5

Extraction of RNA from PBMCs

Peripheral blood (200 μL) was diluted with 10 mL freshly pre-
pared red blood cell samples in alkaline buffer (38.8 mmol/L
NH4Cl, 2.5 mmol/L K2HCO3, 1 mmol/L EDTA; pH 8.0). After
10 min incubation at room temperature, nucleated cells were
washed with the same buffer, the cells were dissolved in
500 μL anti nuclease solution (4 mol/L guanidinium isothio-
cyanate containing 25 mmol/L sodium citrates, 0.5 sarcosyl
and 0.1 mol/L β-mercaptoethanol). A single-step method
described by14 and modified by15,16 followed to carry out the
RNA extraction.

Retrotranscription PCR of sense and antisense strands
of HCV RNA

Detection of HCV RNA strands in PBMCs was performed as
described by Lohr et al.17 The sequences of primers used in
this study are: 1CH, 5′ggt gca cgg tct acg aga cct c3′; 2CH, 5′
aac tca tgt ctc cac gca gaa3′; P2, 5′tgc tca tgg tgc acg gtc ta3′;
P3, 5′cct tgc cga ccc aac act ac3′; and P4, 5′aga gcc ata gtg
gtc tgc ggt3′. The other steps for the retrotranscription PCR
procedure for sense and antisense strands of HCV RNA were
done as previously reported by Abd Alla and El Awady.5

Elastography Fibroscan

The procedure was carried out as previously described.18
Briefly, patients were placed in supine position, with their
right arm in maximal abduction. All measurements were per-
formed in the right lobe of the liver through the costal space.
The tip of the probe transducer was covered with coupling gel
and placed on the skin between the ribs at the right liver lobe
level. The operator, assisted by ultrasound time motion and a
mode images provided by the system, located a portion of the
liver that was at least 6 cm thick and free of any large vascular
structures. Once the area of measurement was located, the
operator pressed the probe button to begin image acquisition.
The measurement depth ranged from 25 mm to 45 mm, and
10 valid measurements were obtained for each patient. The
results were expressed in units of kilopascal (kPa). All the
non-viremic subjects underwent the examination with a
normal prob. The success rate19 of the examination was calcu-
lated as the ratio between the number of measurements vali-
dated by the machine and the total number of attempted
measurements. The liver stiffness corresponds to the median
value of the validated measurements. The interquartile range
(IQR) was used as the interval around the median that con-
strained 50% of the valid measurements. To be considered int-
terpretable and valid, the examination must have included at least
10 measurements, with a SR of at least 66%; and, the IQR
must not have exceeded 33% of the results of the examination.

Statistical analysis

The diagnostic procedure (PBMC-PCR) used was compared to
the SRT-PCR (gold standard test) in a cross-sectional study
that contained two groups of selected subjects. All cases that fit
the inclusion criteria during 25 consecutive month period
were included in the study. All tests performed were two-sided
and statistical significance was considered at a p-value of 0.05.

Results

Advantages of combined screening with both SRT-PCR
and PBMC-PCR

As illustrated in Figure 1, both SRT-PCR and PBMC-PCR
were used in screening of study populations with (Fig. 1A
and 1B) and without (Fig. 1C) controls for HCV-infection.
All cases which tested positive for HCV-infection by SRT-PCR
(n = 673/847, 79.46%; Fig. 1A&B and 673/785, 85.733%;
Fig. 1C) were also positive for intra-PBMC HCV RNA strands.
Patients who presented with non-viremia, as identified by
SRT-PCR, consisted of anti-HCV IgG antibody negative
controls (n = 62) and IgG antibody-positive naïve patients
(n = 112).

Testing all RNA-seronegative subjects (controls n = 62 +
IgG-seropositive n = 112) by cellular-PCR for intra-PBMC
HCV-infection revealed 60/847 (7.1%) of negative controls,
73/847 (8.62%) of negative naïve post-HCV infection
patients, and 41 (4.84%) patients positive for intra-PBMC
RNA strands, as demonstrated in Figure 1B. Adding the extra
positive patients (n = 41) who presented with solitary
intra-PBMC HCV-RNA strand infection to the baseline RNA-
seropositive patients (n = 673/847, 79.46%) significantly
raised the overall percent positive patients for HCV-infection

Fig. 1. Advantages of combined serum and PBMCs (B and C) screening over solitary serum screening for HCV infection (A) by PCR. PBMC testing of non-
viremic subjects detected an extra 41 (4.84%) and 39 (4.97%) infections upon adding (B) or subtracting (C) controls, respectively. The higher numbers of HCV diagnosis
obtained by detecting intracellular RNA strands significantly raised the overall frequencies of HCV diagnosis by PBMCs compared to SRT-PCR (p = 0.011 for panel B and
p = 0.0029 for panel C). As noted in panel C, the frequency of SNOCIs (39/785, 4.968%) is significantly lower than the frequency among patients with negative PCR (73/785,
9.3%) (p = 0.001).
HCV RNA genomic detection within PBMCs compared to serum

| Type of HCV genomic PCR (n) | HCV RNA-negative, n (%) | HCV RNA-positive, n (%) |
|---------------------------|-------------------------|------------------------|
| SRT-PCR (785)             | 112 (14.27)             | 673 (85.73)            |
| PBMC-PCR (785)            | 73 (9.30)               | 712 (90.70)            |
| Fisher’s exact 2-tailed p | 0.00285545              |                        |

HCV diagnosis by intra-PBMCs RNA genomic detection was significantly higher compared to SRT-PCR ($p = 0.0029$). PBMC-PCR-based frequency of negative HCV infection among naïve patients (9.3%) was significantly lower than the SRT-PCR-based frequency (14.16%; $p < 0.00001$).

COCIs and SNOCIs in anti-HCV IgG antibody-positive and RNA seronegative naïve patients compared to negative controls

| Studied group (n) | COCIs, n (%) | SNOCIs, n (%) | Non-OCIs, n (%) |
|-------------------|--------------|---------------|-----------------|
| Negative controls (62) | 2 (3.125)     | 0.0 (0.0)     | 60 (96.875)     |
| IgG-positive/SRT-PCR-negative (112) | 0.0 (0.0)     | 39 (34.82)    | 73 (65.18)      |
| Fisher’s exact 2-tailed p               | 0.00000025    |               |                 |

As diagnosed by PBMC-PCR, the prevalence of SNOCIs in anti-HCV IgG antibody-positive patients (34.82%) is highly significant compared to the prevalence of COCIs (3.1%) in negative controls ($p < 0.00001$).

$n = 714/847$, 84.3%; $p = 0.012$. Figure 1C shows that prevalence of SNOCIs in the HCV IgG antibody-seropositive population (39/785, 4.968%) was significantly lower than the frequency of RNA-seronegative PBMC-PCR (73/785, 9.3%; $p = 0.001$). Table 1 demonstrates results of both SRT-PCR and PBMC-PCR regarding evaluation of SCSD of RNA genomic materials, which is significantly lower in post-HCV non-viremia patients regardless of cirrhosis compared to controls, as diagnosed by both SRT-PCR and PBMC-PCR ($p < 0.00001$). Upon comparing the diagnostic yields of SRT-PCR with PBMC-PCR regarding evaluation of SCSD, the latter procedure was associated with significantly lower SCSD of RNA ($p = 0.03$) because of higher sensitivity. Correlation of results of SRT-PCR and PBMC-PCR with hepatic cirrhosis is illustrated in Figure 2. SCSD of HCV RNA genomic materials, as diagnosed by SRT-PCR, had insignificant difference when comparing cirrhotic patients with non-cirrhotic naïve patients ($p = 0.446$). On the other hand, PBMC-PCR had a highly significant difference in SCSD of HCV strands when comparing non-cirrhotic patients with cirrhotic naïve patients ($p = 0.0001$).

Distribution of sense and antisense strands in cirrhotic versus non-cirrhotic naïve HCV infection

As demonstrated in Table 4, naïve cirrhotic patients showed significantly increased prevalence of sense and antisense strands compared to naïve non-cirrhotic and negative controls ($p < 0.001$). The naïve non-cirrhotic patients, who presented with solitary positive serum anti-HCV IgG antibodies, had a significantly increased frequency of both sense and antisense strands compared to controls ($p < 0.00001$). On the other hand, Figure 3 shows the relationship between density of hepatic cirrhosis and the intracellular HCV RNA strands’ distribution in naïve patients. Association of intra-PBMC HCV genomic materials with hepatic fibrosis was significantly lower in serum from post-HCV naïve patients regardless of cirrhotic changes ($p = 0.03$). All controls are non-cirrhotic and showed significant absence of both serum and intra-PBMC HCV RNA genomic infection than all naïve post-HCV infection ($p > 0.00001$).
found to be highly significant in those who have intracellular RNA strand infection in comparison to the post-hepatitis PCR negative patients \( (p < 0.00001 \text{ and } 0.003 \text{ for intra-PBMC sense and antisense strands, respectively}). \)

Hepatic fibrosis showed insignificant changes in distribution among the naïve HCV patients who had RNA-antisense or sense strand infections \( (p = 0.68) \). Quantitatively, the difference in hepatic fibrosis as kPa was very highly significant when HCV PCR negative, antisense and sense strand-infected patients were compared with negative controls \( (p < 0.001) \). Naïve cirrhotic patients have significantly higher frequencies of both sense and antisense strands compared to naïve non-cirrhotic and negative controls \( (p < 0.001) \). Naïve non-cirrhotic patients have a significantly high frequency of sense and antisense strands compared to controls \( (p < 0.00001) \).

HCV RNA seroconversion compared to spontaneous disappearance of intracellular infection

The fate of intra-PBMC HCV-RNA infection in 7 patients determined at 1-year follow-up is described in Table 5. At baseline, all 7 naïve patients were cirrhotic and had positive serum anti-HCV 1G antibodies, with 4 having both HCV strands inside their PBMCs and the other 3 having the intracellular RNA sense strand but lacking the antisense one. Follow-up of the 7 patients for 1 year by SRT-PCR and PBMC-PCR revealed the following: a) 3 out of 4 F4 patients \( (75\%) \) who had positive intra-PBMC sense and antisense strands at baseline showed RNA seroconversion \( (p = 0.047) \), with the fourth patient was still positive for intracellular sense and antisense RNA strands without RNA seroconversion; b) absence of antisense strand in the other 3 naïve patients who started with positive intra-PBMC HCV-RNA sense

Table 4. Distribution of sense and antisense HCV RNA strands in cirrhotic and non-cirrhotic naïve patients

| Study subjects \((n)\) | PBMC-PCR |
|-----------------------|----------|
|                       | Antisense strand \(n\) \(\%\) positive | Sense strand \(n\) \(\%\) positive | Negative \(n\) \(\%\) positive |
| Controls \((62)\)     | 0.00 (0.00) | 2.00 (3.23) | 60.00 (96.77) |
| Cirrhotic naïve \((57)\) | 11 (19.30) | 21 (36.84) | 25 (43.86) |
| Non-cirrhotic naïve \((55)\) | 3 (5.45) | 4 (7.27) | 48 (87.28) |
| Fisher’s exact 2-tailed \(p\): A vs. B | 0.000000000 | 0.000000028 | 0.0034618535 |
| : A vs. C | 0.000000000 | 0.000000000 | 0.000000000 |
| : B vs. C | 0.000000010 | 0.000371340 | 0.0026551955 |

Naïve cirrhotic patients have significantly higher frequencies of both sense and antisense strands compared to naïve non-cirrhotic and negative controls \( (p < 0.001) \). Naïve non-cirrhotic patients have a significantly high frequency of sense and antisense strands compared to controls \( (p < 0.00001) \).
Overall, the number of infected cases increased from 673 to 712. Infections that are usually recognized in clinical practice as cases with history of HCV infection (i.e., having no viremia despite positive serum IgG-antibodies) were designated as patients with SCSD in the current study. The number of cases presented with SCSD was found to be significantly lower because of the increased sensitivity of PBMC-PCR in diagnosing SNOCIs. The association of hepatic fibrosis with SNOCIs was also significant in the current data set. Furthermore, the increased density of hepatic fibrosis is accompanied by intra-PBMC infection with either sense or antisense strands. The HCV RNA seroconversion in this cohort is attributed to intra-PBMC infection with both sense and antisense strands in cirrhotic patients.

The increased sensitivity of diagnosing dormant HCV-infection addressed by a previous study adds major advantages to the management of both SNOCIs and SEOCIs in medical practice. The appreciated high predictive values of HCV-PCRs provide a trustworthy diagnostic tool to trace extra-and intra-cellular RNA replication. The current study looked for RNA strands in PBMCs because of their considerable degrees of HCV tropism and because their nuclei are essential to viral RNA genomic replication.6 The solitary intracellular recognition of HCV RNA, without viremia, has been known to occur during the incubation periods or as a post-infection presentation after successful elimination of genomic materials from sera.20

Spontaneous and drug-induced HCV RNA disappearance from sera have been collectively termed as sustained virologic disappearance; unfortunately, that naming convention doesn’t correspond to elimination of viral genomic materials from the rest of body tissues.21 RNA seroconversion in both post-treatment or naive patients is usually related to persistent intracellular infection with HCV genomic strands.9,11,22–25 We have recently evaluated PBMC-PCR of HCV-RNA as one of the most sensitive and specific tools to assess extrahepatic HCV infections.5 In current cohort, we utilized PBMC-PCR to identify HCV infections in naïve cases that tested negative by SRT-PCR (COCIs and SNOCIs). The significant increases in the numbers of the infected cases

Table 5. RNA seroconversion versus spontaneous intracellular disappearance of naïve HCV infection as recognized by concomitant SRT-PCR and PBMC-PCR

| Serial number | Baseline PCR data at entry | Follow-up by PCR after one year |
|---------------|---------------------------|---------------------------------|
|               | SRT-PCR Serum RNA         | PBMC-PCR Serum RNA (IU/ml)      | F0 = 5.5 kPa, F1 = 5.75 kPa, F2 = 7.5–10 kPa, F3 = 10–15 kPa, F4 >15.0 kPa. |
|               | Sense strand Antisense strand | Flobscan results at baseline in kPa | SRT-PCR Serum RNA Antisense strand Sense strand |
| 1             | –ve +ve +ve                | F4 = 5.14                        | 21700 +ve +ve |
| 2             | –ve +ve +ve                | F4 = 57.3                        | 242000 +ve +ve |
| 3             | –ve +ve +ve                | F4 = 28.0                        | 299000 +ve +ve |
| 4             | –ve +ve +ve                | F4 = 35.3                        | –ve +ve +ve |
| 5             | –ve +ve +ve                | F4 = 17.1                        | –ve –ve –ve |
| 6             | –ve +ve +ve                | F2 = 9.50                        | –ve –ve –ve |
| 7             | –ve +ve +ve                | F1 = 5.80                        | –ve –ve –ve |
|               |                           |                                  |                  |
|               |                           |                                  |                  |

Positive cases for both intra-PBMC HCV RNA strands at baseline are associated with RNA seroconversion in 3 out of 4 (uncorrected 2-tailed, p = 0.047) F4 naïve patients; the remaining patient failed to clear the intracellular sense and antisense RNA strands. All naïve patients who had positivity for intra-PBMC HCV-RNA sense but negativity for antisense strand cleared the intracellular infection within 1 year. kPa, Kilo Pascal; +ve, positive; –ve, negative.
after adding the newly diagnosed cases by PBMC-PCR recommends cellular screening for HCV-RNA infection to diagnose SNOCIs. On the other hand, the present results showed that OCIs prevalence is almost the same as described in other reports.2-4

It is important to remember that the studied population has a higher incidence of HCV genotype 4 and a lower incidence of genotype 1, a profile that is spreading widely in western countries. However, the current study did not aim to investigate OCIs for any particular genotype, but instead aimed to provide insight into a new use of an established diagnostic tool that would facilitate the diagnostic workup and furnish more accurate information regardless to viral genotype. The clinical application of the data analysis method that was adopted in the current study would be valid for studying of other populations and a spectrum of genotypes. However, calculation of the real prevalence of HCV infection in any population worldwide is dependent upon the combination of multiple diagnostic procedures (i.e. enzyme-linked immunosorbent assay, traditional PCR, PBMC-PCR, tissue PCR, ultracentrifugation-based PCR approach, etc.).5,20 a countless number of tissues (i.e. serum, blood cells, whole blood, bone marrow, liver tissues, etc.),20,24,26 and application in various clinical case situations (i.e. treatment-naïve, treatment-experienced, in contact with HCV patients, and with chronic active hepatic cellular damage).5,23-28

Our study concluded that SCSD is inversely proportional to the degree of hepatic cirrhosis; in addition, the increased hepatic cirrhosis is associated with diminished frequency of SCSD. These results confirm other studies regarding the association of liver cirrhosis with active HCV infection in viremic patients,10 but is considered a novel finding in SNOCIs as we reported in a previous study.5 It is convenient to consider the presence of HCV IgG antibody in naïve non-viremia (SCSD) patients as a remark for history of HCV infection, despite the belonging of dormant HCV infection in SCSD patients to the SNOCIs, as documented in the current study. Considering the clinical importance of PBMC-PCR, the significantly higher number of SCSD cases (73/112) compared to SNOCIs (39/112) supports the importance of anti-HCV IgG antibody testing as a preliminary screening approach, although it is not sufficient as a baseline diagnostic test for treatment. The numbers of SCSD cases are expected to drop down upon searching nucleated cells from other tissues (e.g., bone marrow, lymph nodes, spleen, liver, etc.) or using combined serologic diagnostic tests.20 The rising titers of anti-HCV IgG antibodies in naïve and post-treatment cases might be a good predictor for current chronic HCV-infection.

Diagnosis of hepatic fibrosis in non-viremia patients is a confusing clinical situation that might push both physician and patients to face unexpected morbidity and mortality outcomes over time. Results of intra-PBMC HCV-RNA detection in the current study concern the above-mentioned clinical situation. Fortunately, the current report addresses HCV as an inducing factor for liver cirrhosis in SNOCIs, which will cause impaired liver function. The impact of hepatic fibrosis on SCSD of infection is most probably dependent upon interactions between the host immune system and the underlying pathogenic etiology. The current data set linked density of hepatic fibrosis with the presence of intra-PBMC HCV-RNA sense and antisense strands. Cumulative data during the next few years might help in creation of effective guidelines for HCV therapy to address treatment feasibility of solitary intracellular chronic HCV infection problems in non-viremia patients. Drawbacks of neglecting progressive hepatic cirrhosis are shown by evidence of active liver cell damage, so that lack of appropriate and timely medical management is extremely hazardous. Narrowing the spectrum of the etiologic factors in anti-HCV IgG antibody-positive non-viremia patients by management of SNOCIs in cirrhotic patients should minimize the over-time morbidity and mortality rates among chronic HCV infections.

On the other hand, differential recognition of intra-PBMC sense and antisense strands by PCR in our study provided elective information regarding their relationship to HCV RNA seroconversion in naïve patients. The present results indicate that solitary identification of intra-PBMC HCV-RNA sense strand would not be enough to induce viral genomic material seroconversion in naïve patients. The co-existence of sense with antisense strands seems to be crucial for HCV-RNA genomic materials to reappear in patient’s sera. The authors believe that this is the first-report of HCV-RNA spontaneous seroconversion in naïve non-viremia cirrhotic patients occurring during the follow-up period without intervention, despite the small number (7 patients) that limits clinical significance. Contrary to these clues, the presence of intracellular antisense strand in naïve patients was reported as a good prognostic marker of response to antiviral therapy, and the post-treatment persistence of HCV-RNA antisense strands is an ominous sign because of the high probability of relapse.7

Findings of the current study include: a) diagnosis of both cryptogenic and SNOCIs by PBMC-PCR having significant impact upon the total prevalence of naïve HCV-infection; b) recognition of SNOCIs in all IgG antibody seropositive non-viremic patients being highly recommended because of their frequent detectability compared to controls; c) use of PBMC-PCR being strongly recommended to diagnose SCSD, in addition to SRT-PCR; d) cases of SCSD, as diagnosed by HCV PBMC-PCR, being more prevalent in non-cirrhotic patients than in cirrhotic patients; e) both intra-PBMC HCV-RNA sense and antisense strands being associated with increased density of hepatic fibrosis in kPa units; f) primary seroconversion of HCV-RNA in naïve patients being associated with combined intra-PBMC existence of sense and antisense strands.

In conclusion, evaluation of intra-PBMC HCV RNA infection is crucial for diagnosing OCIs, evaluating post-infection sequelae in RNA seronegative naïve patients, and addressing the probability of viral RNA serologic relapse.

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Conflict of interest

The authors have no conflict of interests related to this publication.

Author contributions

Responsible for study design, patient management, data analysis, and writing of the manuscript (MDAA), recruited
patients, revised and formatted the manuscript, and performed the virological investigation (SAE, GYW, MKEA).

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