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Ramberg, Ingvild; Heegaard, Steffen

Published in:
Viruses

DOI:
10.3390/v13081522

Publication date:
2021

Document version
Publisher's PDF, also known as Version of record

Document license:
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Citation for published version (APA):
Ramberg, I., & Heegaard, S. (2021). Human papillomavirus related neoplasia of the ocular adnexa. Viruses, 13(8), [1522]. https://doi.org/10.3390/v13081522
Review

Human Papillomavirus Related Neoplasia of the Ocular Adnexa

Ingvild Ramberg 1,2 and Steffen Heegaard 1,2,*

1 Department of Pathology, Copenhagen University Hospital Rigshospitalet, DK-2100 Copenhagen, Denmark; Ingvild.margrethe.sellaeg.ramberg@regionh.dk
2 Department of Ophthalmology, Copenhagen University Hospital Rigshospitalet, DK-2100 Copenhagen, Denmark
* Correspondence: sthe@sund.ku.dk

Abstract: Human papillomaviruses (HPV) are a large group of DNA viruses that infect the basal cells of the stratified epithelium at different anatomic locations. In the ocular adnexal region, the mucosa of the conjunctiva and the lacrimal drainage system, as well as the eyelid skin, are potential locations for HPV-related neoplasia. The role of HPV in squamous cell neoplasia of the ocular adnexa has been debated for several decades. Due to the rarity of all these tumors, large studies are not available in the scientific literature, thereby hampering the precision of the HPV prevalence estimates and the ability to conclude. Nevertheless, increasing evidence supports that defined subsets of conjunctival papillomas, intraepithelial neoplasia, and carcinomas develop in an HPV-dependent pathway. The role of HPV in squamous cell tumors arising in the lacrimal drainage system and the eyelid is still uncertain. Further, the potential of HPV status as a diagnostic, prognostic, or predictive biomarker in these diseases is a topic for future research.

Keywords: human papillomavirus; ocular adnexa; conjunctiva; lacrimal drainage system; eyelids; squamous cell carcinoma; squamous cell papilloma; sebaceous cell carcinoma

1. Introduction

Human papillomaviruses (HPV) are a large group of DNA viruses with tropism for epithelial tissues of the skin and mucosae. Tremendous progress has been made within the field since the first report of a cell-free transmission of canine oral warts in 1898, and we are today aware of the substantial burden of neoplastic diseases caused by papillomaviruses in many anatomical locations [1]. HPV belongs to the Papillomaviridae family and is divided into five different genera (α, β, γ, μ, and ν) based on the homology of the nucleotide sequence coding for the HPV capsid protein L1. Most mucosal genotypes belong to the α-genus, whereas cutaneous genotypes predominantly belong to the α, β-, and γ-genera. Thus, in the ocular adnexa, the mucosal membranes of the conjunctiva and the lacrimal drainage system, as well as the eyelid, are potential locations for HPV-related neoplasia.

Generally, studies investigating the association between HPV and neoplasia of the ocular adnexa are hampered by small sample sizes leading to imprecise HPV prevalence estimates. The studies are mostly limited to retrospective case series, and thereby inducing inherent risks of publication and selection biases. In addition, a great diversity of applied methods for HPV detection, with different sensitivity and specificity profiles, has been applied as described by Stagner et al. [2]. With these precautions, the role of HPV in neoplasia in the ocular adnexa is discussed in the present review.

2. The Conjunctiva

The conjunctiva is the transparent mucous membrane covering the inside of the eyelids (palpebral conjunctiva) and the anterior part of the sclera (bulbar conjunctiva), where it becomes continuous with the anterior epithelium of the cornea (Figure 1). A non-keratinizing stratified columnar epithelium covers the palpebral conjunctiva that
gradually thickens toward the fornices, and at the corneoscleral junction, the epithelium becomes squamous. Being the only mucous membrane in the body exposed directly to the external environment, the conjunctiva is vulnerable to external stress stimuli, including pathogens, ultraviolet (UV) radiation, air, dust, and cigarette smoke. The role of HPV in the pathogenesis of conjunctival neoplasia has been debated in the scientific literature since the first reports of HPV in conjunctival papillomas in 1983 [3] and conjunctival dysplasia and carcinoma in 1986 [4].

2. The Conjunctiva

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Figure 1. The epithelial linings of the conjunctiva and lacrimal drainage system.

2.1. Conjunctival Papilloma

Conjunctival papilloma is a benign tumor arising from the conjunctival epithelium. Based on the growth pattern, the papillomas are histopathologically divided into exophytic and inverted papilloma. Rarely, a mixed growth pattern is present. The exophytic papilloma forms outwards projections of an acanthotic conjunctival epithelium surrounding a fibrovascular core. The papillomas can be sessile or pedunculated. Mild to moderate dysplastic changes may be present; however, severe dysplasia and malignant transformation are rarely seen [5–7]. Precise incidence estimates of conjunctival papillomas are not available in the scientific literature, but the disease accounts for 1–16% of all histopathologically verified conjunctival lesions [8]. The incidence peaks in patients aged 21–40 years and thereafter declines with a male predominance in all age groups [5,6,8].

Human papillomavirus is considered the main risk factor for developing these papillomas, consistently reported in more than 50% of the cases (Table 1) and with a reported viral load comparable to the levels in laryngeal papillomatosis [9]. The vast majority of detected genotypes are the low-risk HPV6 and HPV11, and rarely other genotypes such as
HPV13, 16, 33, and 45 [10–13]. Histopathologically, the presence of basaloid cells, intraepithelial goblet cells, a non-keratinizing squamous epithelium, and lack of solar elastosis are associated with HPV [9]. On the other hand, koilocytosis is not a valid biomarker for HPV-related conjunctival papilloma [8,11,14]. Clinically, a location of a papilloma other than at the corneal limbus is associated with HPV infection [9]. A prognostic and predictive value of HPV status in conjunctival papilloma is not yet determined.

Table 1. An overview of studies (including >5 cases) examining HPV in conjunctival exophytic papilloma. HPV; human papillomavirus, PCR; polymerase chain reaction, ISH; in-situ hybridization.

| Author, Year          | Median Age Years (Range), Gender | HPV+ (DNA PCR) | HPV+ (RNA ISH) | HPV Genotypes | HPV Detection Modality |
|-----------------------|----------------------------------|----------------|----------------|---------------|------------------------|
| Mlakar et al., 2015   | 49 (28–77), 17M/8F               | 19/25 (76%)    | NA             | 6, 11         | PCR, in-situ hybridization |
| Takamura et al., 2008 | 40 (28–76), 4M/2F                | 6/6 (100%)     | NA             | NA            | PCR, hybrid capture II  |
| Sjö et al., 2007      | 27 (18–65), (9–80), 21M/3F       | 86/106 (81%)   | NA             | 6, 11, 45     | PCR                    |
| Eng et al. 2002       | 27 (18–65), (9–80), 21M/3F       | 14/24 (58%)    | NA             | 6, 11         | PCR                    |
| Nakamura et al., 1997 | 51 (20–73), 6M/2F                | 4/8 (50%)      | NA             | 6             | PCR, in-situ hybridization |
| Saegusa et al., 1995  | 38 (14–73), 5M/11F               | 12/16 (75%)    | NA             | 16            | PCR, in-situ hybridization |
| McDonnell et al., 1987| 25 (1–71)                        | 15/23 (65%)    | NA             | 6             | In-situ hybridization   |

Opposed to the exophytic papillomas, the inverted conjunctival papillomas grow inwards, however, respecting the basement membrane, which remains intact. Inverted conjunctival papillomas are rare, with only a handful reported in the scientific literature to date [19]. An association with high-risk HPV16 and 58 genotypes has been reported [7,19,20]. Despite the limited reports, the inverted papillomas seem more aggressive than their exophytic counterparts with frequent synchronous and metachronous cancer development [7,20–22]. Although some publication bias is expected, this picture suits well with our knowledge from the adjacent sinonasal tract, where inverted papillomas often are associated with high-risk HPV genotypes and carry a significant risk of malignant transformation [23]. Thereby, close attention is required in the follow-up of these patients.

2.2. Conjunctival Intraepithelial Neoplasia and Squamous Cell Carcinoma

Conjunctival squamous cell carcinoma (SCC) is the most common malignancy arising from the conjunctiva and is the end-stage disease in the spectrum ranging from conjunctival intraepithelial neoplasia (CIN) grades I-III, carcinoma in-situ, and eventually invasive conjunctival SCC (Figure 2). While CIN encompass different degrees of epithelial involvement only, the conjunctival SCC invades the basement membrane and the underlying substantia propria. Conjunctival SCC has a locally destructive growth pattern, where the malignant cells eventually invade the adjacent structures, including the globe (American Joint Committee on Cancer (AJCC) tumor stage T3) and the orbit (AJCC tumor stage T4). Orbital exenteration is required in approximately 10% of the cases to obtain local tumor control [24,25] causing significant morbidity. However, also AJCC tumor stage T1-2 disease can cause vision loss due to limbal stem cell deficiency. Conjunctival SCC also carries a risk of lymph node- and systemic metastasis, especially in people living with human immunodeficiency virus (HIV)—a patient group that evolves more frequent and more severe disease at a younger age [26–29].
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Figure 2. (a) A limbal conjunctival squamous cell carcinoma in situ with corneal involvement. (b) Histology reveals pleomorphic squamous tumor cells with abundant, aberrant mitoses (arrows) and subepithelial inflammation (hematoxylin-eosin (HE)-stain, bar scale = 100 μm). (c) Intense positive cytoplasmatic and nuclear expression of p16INK4a in the tumor cells, a surrogate marker of human papillomavirus (HPV) infection (p16INK4a, immunohistochemistry, scale bar = 450 μm). (d) Expression of high-risk HPV oncogenes within the tumor cells. Note the clear demarcation to the underlying stromal tissue (HPV E6/E7 mRNA in-situ hybridization using a probe for detection of high-risk HPV genotypes (16, 18, 26, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, 66, 68, 73, and 82), scale bar = 350 μm).

2.2.1. Prognosis and Treatment

One of the main challenges in treating conjunctival SCC is the high recurrence rates [30]. A systematic review of the literature reported a mean risk of recurrence at 52% in cases with positive surgical margins and 11% in cases with negative surgical margins [31]. Adjuvant therapy, including topical or intralesional chemotherapy, radiotherapy, or plaque brachytherapy, further reduces the persistence of the tumors and the recurrence rates, independent of margin status, but may also cause considerable side effects [29,32]. The application of adjuvant topical interferon alpha-2b in cases with narrow, indeterminate, or negative surgical margins seems to obtain better tumor control than re-excision, incisional biopsy-guided interferon alpha-2b therapy, or empiric interferon alpha-2b treatment as monotherapy [31]. Still, non-responsiveness to interferon treatment is relatively common, with a reported pooled estimate of 12% of the cases reported in a systematic literature review [31,33]. Moreover, in a randomized controlled study, monotherapy with mitomycin C in conjunctival CIN could not be separated histologically from the placebo group after 6–8 weeks of treatment [34].

The high recurrence rates and resistance to treatment are clinical challenges, as the risk of adverse events, including corneal and conjunctival scarring, corneal stem cell deficiency, and risk of orbital exenteration, increases by repeated intervention. Therefore, in order to up- and de-escalate the treatment, also in pre-invasive tumors, there is an urge for randomized controlled studies, identification of subgroups, and incorporation of valid biomarkers.
2.2.2. Human Papillomavirus in Conjunctival CIN and SCC

As mentioned, the clinical course of HIV-associated conjunctival SCC differs substantially from those arising in immunocompetent individuals, and HIV status delineates two important subgroups to be considered in future trials. Increasing evidence also supports the role of HPV in conjunctival CIN and SCC, and HPV status may also serve as an important biomarker in these diseases. With an odds ratio (OR) 8.42 (95%CI 3.70–19.16 in 21 studies) [35] and a risk ratio (RR) 3.13 (95%CI 1.72–5.71, 16 studies) [36] in two separate meta-analyses based on observational studies, HPV has shown to be strongly associated with an increased risk of conjunctival CIN and SCC. The prevalence estimates of HPV are 26% (95%CI 13–42%) in conjunctival CIN and 18% (95%CI 7–32%) in invasive SCC compared to 1% (95%CI 0–3%) among controls [35]. An overview of studies examining HPV in conjunctival intraepithelial neoplasia and carcinoma (39 studies) is provided in a recently systematic review by our group [35].

The expression of the viral oncogenes E6 and E7 within the tumor cells, and not merely detection of viral DNA by PCR, gives additional weight to a causative role of HPV in a subgroup of conjunctival CIN and SCC (Table 2) [33,37–39]. Further, the loss of HPV E4 gene expression in the malignant cells, hence a transition from a productive to an abortive life-cycle of the HPV, is documented in a series of conjunctival CIS similar to high-risk cervical intraepithelial neoplasia [33]. The high-risk genotype HPV16 is by far the most frequent genotype in conjunctival CIN and SCC, followed by HPV18 and HPV33 [35]. Less frequent, HPV6, 11, 31, 35, 37, 39, 44, 45, 51, 52, 58, 66, 72, and 83, are detected [35].

| Author, Year   | Median Age Years (Range)/Gender | HPV+ (DNA) | HPV+ (RNA) | HPV Genotypes | HPV Detection Modality |
|----------------|---------------------------------|------------|------------|---------------|------------------------|
| Griffin et al., 2019 [33] | 61 (21–103)/25M, 16F | 17/41 (41%) | 11/13 (85%) | 16 | PCR, RNA ISH, p16INK4a |
| Nagarajan et al., 2019 [37] | 62 (36–61)/16M, 15F | NA | 8/31 (26%) | High-risk genotypes | RNA ISH |
| Ramberg et al., 2019 [38] | 65 (30–97)/81M, 31F | 24/112 (21%) | 18/19 (95%) | 6, 11, 16, 33, 39 | PCR, RNA ISH, p16INK4a |
| Scott et al., 2002 [39] | NA | 10/10 (100%) | 10/10 (100%) | 16, 18 | RT-PCR, DNA ISH |

* 13 out of 17 HPV DNA positive cases tested with RNA ISH. ** 19 out of 24 HPV DNA positive cases tested with RNA ISH. HPV; human papillomavirus, PCR; polymerase chain reaction, ISH; in-situ hybridization.

With stratification by geographical location, studies from African countries show an insignificant association to α HPV genotypes (OR 1.7, 95%CI 0.9–3.5) [35], but a significant association to β genus HPV genotypes (RR 3.52, 95%CI 1.23–10.08) [36]. The development of cancers caused by the β genus HPV is primarily seen in individuals with compromised immune surveillance, e.g., patients in systemic immunosuppressive treatment or people living with HIV [40]. The risk of conjunctival SCC is more than 10-fold in people living with HIV and is considered an early indicator of HIV infection [28]. The link between β genus HPV genotypes and conjunctival SCC in African countries may therefore be explained by the strong association between HIV and conjunctival SCC development [41], but is outside the scope of the present review.

2.2.3. Clinical Characteristics of HPV-Related Conjunctival CIN and SCC

Only a few studies have compared the clinical characteristics of HPV-positive versus HPV-negative conjunctival CIN and SCC. Due to the retrospective setup of these studies, no conclusions regarding clinical outcomes of HPV status can be drawn on this basis. Our current knowledge suggests that HPV-positive conjunctival SCC develops at an earlier age than HPV-negative conjunctival SCC [38,42] with a higher risk of extracanconjunctival extension at diagnosis [37] and a higher risk of recurrence [38]. One study has investigated the prognostic value of p16INK4a in conjunctival SCC and found a significant reduced disease-free survival in patients with p16INK4a-positive tumors. However, as HPV status was not reported in this study, this finding might be unrelated to HPV [43].
2.2.4. Histological and Immunohistological Markers of HPV-Related Conjunctival CIN and SCC

Histologically, HPV-related conjunctival CIN and SCC are associated with a basaloid, non-keratinizing morphology with foci of comedo necrosis and a dense inflammatory background [38,42]. The presence of koilocytosis, hence cellular and nuclear enlargement, hyperchromatic nuclei, and a perinuclear cytoplasmic halo, is not a valid marker of HPV-related conjunctival CIN and SCC, with reported sensitivity ranging from 0 to 100% [17,44,45] and specificity ranging from 63 to 100% [17,44,46].

Evaluated by immunohistochemistry, p16\(^{INK4a}\) expression is a surrogate marker of HPV-related carcinomas in the head-and-neck region as well as anogenital carcinomas. Under homeostatic conditions, the p16\(^{INK4a}\) acts as a tumor suppressor by inactivating the cyclin-dependent kinases that phosphorylate the retinoblastoma protein (pRB) and thereby hamper the cell cycle progression from G1 to the S phase of mitosis. Phosphorylation of pRB in turn, influences the expression of p16\(^{INK4a}\). The HPV oncogene E7 deregulates pRB leading to a compensatory increase in p16\(^{INK4a}\) expression. The expression of p16\(^{INK4a}\) in conjunctival CIN and SCC has been evaluated in several studies [11,33,38,42,45,47–52]. However, no conclusions on the applicability of p16\(^{INK4a}\) in conjunctival CIN and SCC can be taken due to the broad variability in the definition of p16\(^{INK4a}\) overexpression (percentage of tumor cells or binary definition (positive/negative), nuclear and/or cytoplasmatic staining), the immunohistochemistry staining probes used, HPV results, and HPV detection modalities used (PCR-based (HPV DNA PCR, HPV RNA RT-PCR) and/or in-situ hybridization (ISH)-based (HPV DNA ISH, HPV RNA ISH) or combination of techniques). Two studies reported p16\(^{INK4a}\) IHC along with HPV DNA PCR and E6/E7 mRNA ISH [33,38]. In these two studies, the p16\(^{INK4a}\) IHC yielded a sensitivity ranging from 85 to 90% and a specificity ranging from 66 to 90% for a prediction of high-risk HPV infection when using HPV DNA PCR as a reference, and a sensitivity of 100% and a specificity 75% of when using mRNA ISH as a reference [33,38]. Hence, the value of p16\(^{INK4a}\) overexpression as a biomarker for HPV-related conjunctival CIN and SCC is a topic for future research.

2.3. Transmission of HPV to the Conjunctiva

There are several hypotheses of how HPV reaches the conjunctiva, and most likely, there are different transmittal routes. Looking at the genotypes involved in conjunctival pathogenesis, the low-risk HPV6 (“genital type”) and HPV11 (“oral type”) predominates in the exophytic conjunctival papillomas, whereas the high-risk HPV16 constitutes the vast majority in conjunctival SCC. Likely, the HPV is transferred from the anogenital region as well as the oral and pharyngeal mucosa to the conjunctiva. Vertical transmission—hence, acquired infection during the passage of an infected birth canal—may cause neonatal or early conjunctival papillomatosis [53–55]. In a prospective cohort study of HPV-positive pregnant women, HPV was detected in the conjunctiva in 4.8% of the children at birth and/or at three months of age [56]. Auto-inoculation is another proposed transmission mode, e.g., by hand carriage of genital HPV DNA [57]. Reports of co-existent genital- and conjunctival HPV-related neoplasia support autoinoculation as a transmission route to the conjunctiva [55,58].

For the virus to infect the basal cell of the epithelium, micro-abrasions of the conjunctival surface in order to expose the basal membrane are required. Such abrasions are common in patients with vitamin A deficiency and atopy—both patient groups with increased risk of conjunctival SCC [33,59]. Micro abrasions of the conjunctiva also occur in patients with ocular prostheses and thereby making the conjunctiva susceptible to HPV infection [60].

In conclusion, the conjunctiva is a vulnerable epithelial site of HPV-related neoplasia with viral and cellular biomarkers of deregulated HPV gene expression. However, increasing evidence also supports a diagnostic and prognostic value of high-risk HPV expression in conjunctival CIN and SCC.
3. The Lacrimal Drainage System

The lacrimal drainage system (LDS) drains tears, debris, and microbes from the ocular surface to the nasal cavity. Anatomically, the LDS consists of the lacrimal canaliculi, the lacrimal sac, and the nasolacrimal duct (Figure 1). Tumors arising in the LDS are often of epithelial origin, with the vast majority being squamous cell carcinoma and non-keratinizing squamous cell carcinoma (previously transitional cell carcinoma) [61]. Of benign tumors, squamous- and transitional papillomas occur most frequently. The papillomas of the LDS can be inverted, squamous/exophytic, transitional, or mixed type. The malignant tumors are most often located in the lacrimal sac and the nasolacrimal duct, whereas the benign tumors more often appear in the upper part of the LDS [62].

Tumors of the LDS are rare; hence, our current knowledge of these tumors’ development is scarce. Suspected risk factors for developing LDS carcinoma include chronic inflammation, infection with Epstein-Barr virus, previous probing or surgery of the LDS, and HPV infection [61]. Both exophytic and inverted papillomas of the LDS can undergo malignant transformation. Thereby, squamous cell carcinomas of the LDS can develop de novo or in a pre-existing papilloma [63–65]. The role of HPV in LDS papilloma and carcinoma development is still uncertain due to the rarity of the tumors. In LDS papillomas, mostly low-risk genotypes (HPV6, 11) have been reported; however, they are limited to a handful of case reports and small case series (Table 3). One study reports the expression of HPV oncogenes in the tumor cells, suggesting that HPV is not merely an innocent bystander in the development of these tumors’ development [66].

### Table 3. An overview of studies examining HPV in papillomas of the lacrimal drainage system.

| Author, Year              | Median Age Years (Range)/Gender | HPV+ (DNA PCR) | HPV+ (RNA ISH) | HPV Genotypes | HPV Detection Modality  |
|---------------------------|---------------------------------|----------------|----------------|---------------|-------------------------|
| Jones et al., 2020 [67]   | -                               | 3/10 (30%)     | NA             | 6, 11, 16     | PCR, DNA ISH            |
| Madreperla et al., 1993 [68]| 38 (36–54)/3M                   | 2/2 (100%)     | NA             | 11            | PCR, DNA ISH            |
| Sjö et al., 2007 [66]     | 37 (30–56)                      | 4/4 (100%)     | 2/2 (100%)     | 6, 11         | PCR, DNA ISH, RNA ISH   |
| Vickers et al., 2010 [69] | 53/F                            | 1/1 (100%)     | NA             | 11            | PCR                     |
| Nakamura et al., 1997 [17]| 38 (26–50)/1F, 1M               | 1/2 (50%)      | NA             | 16            | DNA ISH, PCR            |
| Buchwald et al., 1996 [70]| -                               | 1/1 (100%)     | NA             | 6/11          | DNA ISH                 |

PCR; Polymerase chain reaction, ISH; in-situ hybridization.

Regarding LDS carcinomas, only four studies in the literature have addressed the association to HPV (Table 4) [42,66–68]. In these studies, HPV was detected in 15 out of 25 (60%) squamous cell - and transitional cell carcinomas by HPV DNA PCR, most frequently being HPV16. However, as HPV DNA detection by PCR does not prove causality, further investigations are required to decide the role of HPV in LDS carcinoma development.

### Table 4. An overview of studies examining HPV in carcinomas of the lacrimal drainage system.

| Author, Year              | Median Age Years (Range)/Gender | HPV+ (DNA PCR) | HPV+ (RNA ISH) | HPV Genotypes | HPV Detection Modality  |
|---------------------------|---------------------------------|----------------|----------------|---------------|-------------------------|
| Afroghheh et al., 2016 [42]| 60 (34–75)/4M, 5F               | 8/9 (89%)      | NA             | 16, 33, 58    | DNA ISH, PCR, p16\N\Ka |
| Madreperla et al., 1993 [68]| -                               | 1/2 (50%) *    | NA             | 18            | DNA ISH, PCR            |
| Sjö et al., 2007 [66]     | 61 (33–86)/4M, 2F               | 4/6 (67%)      | 0/4 (0%)       | 6, 11, 16 **  | PCR, DNA ISH, RNA ISH   |
| Jones et al., 2020 [67]   | -                               | 2/4 (50%)      | NA             | 16            | PCR                     |

HPV; human papillomavirus, PCR; Polymerase chain reaction, ISH; in-situ hybridization, NA; not applicable. * Only two carcinomas were available to PCR. ** Co-comitant infection with low-risk 6 or 11 and high-risk HPV16.

The treatment options of LDS carcinomas differ according to the histopathological diagnosis, the tumor location, and the stage of the disease. Due to the rarity of the diseases, the treatment regimens are often extrapolated from other head-and-neck cancers, especially sinonasal carcinomas. Surgical resection is the first-line treatment, often adjuvanted by
intensity-modulated radiotherapy [71]. The benefits of adjuvant chemotherapy in the treatment of LDS carcinomas are still uncertain. Cisplatin-based chemotherapy is frequently used in patients with inoperable tumors and metastatic disease.

4. The Eyelids

The eyelids are covered by a keratinized, stratified squamous epithelium only a few cells thick. The main functions of the eyelids are to protect the globe and secrete, distribute, and drain tears on the ocular surface. The sebaceous glands of the eyelids (the Meibomian glands, the glands of Zeis, and the glands associated with the caruncle) are a part of the epidermal appendages and are responsible for the secretion of the oily part of the tear film and cilia. The reports of HPV-associated carcinomas deriving from the skin and the sebaceous glands of the ocular adnexa are discussed in the following sections.

4.1. Sebaceous Gland Carcinoma

Carcinomas of the sebaceous glands in the ocular adnexal skin structures most commonly arise in the Meibomian glands (Figure 3). Geography impacts the incidence, being more common in Asians (one-third of all eyelid malignancies) and rare in Caucasians (0.5–5% of all eyelid malignancies) [72].

![Figure 3](image_url)

**Figure 3.** (a) A sebaceous gland carcinoma of the eyelid with origin in a meibomian gland. The clinical examination revealed a painless, firm papule of the upper eyelid. (b) Histopathologically, the tumor consists of pleomorphic sebaceous tumor cells with scattered mitoses and display pagetoid growth. The tumor is dominated by basaloid cells with only a few well-differentiated multivacuolated sebaceous tumor cells (hematoxylin-eosin (HE)-stain, scale bar = 175 μm).

Risk factors for the development of sebaceous carcinoma include UV-radiation, genetic predisposition syndromes (e.g., Muir-Torré syndrome), immunosuppression, previous radiation therapy to the head or neck, and, possibly, HPV. The genomic profile of ocular adnexal sebaceous carcinoma differs from the UV-induced and microsatellite instability (MSI) profiles that characterize sebaceous carcinoma of other anatomic locations [73]. Thereby, sebaceous gland carcinoma of the ocular adnexa represents a defined subset of sebaceous carcinomas [73].

**Human Papillomavirus in Ocular Adnexal Sebaceous Carcinoma**

Conflicting evidence exists regarding the role of HPV in ocular adnexal sebaceous carcinoma (Table 5). The highest HPV prevalence is reported in a study by Hayashi et al. using DNA ISH reporting HPV positivity in 13 out of 21 (62%) samples [72]. However, the HPV DNA signals were also present in the surrounding normal sebaceous glands and epidermis, causing doubt on the specificity of the applied method. Several other studies failed to detect HPV in their series [74–76]. Stagner et al. reported HPV16 in one out of 24 samples using PCR but did not detect viral expression by mRNA ISH [2], further
questioning the role of HPV [72]. On the other hand, recent studies by Tetzlaff et al. [77] and Moore et al. [78] detected expression of HPV oncogenes restricted to the tumor cells in 14% and 18% of their series, respectively. Tetzlaff et al. further reported mutually exclusive genetic profiles according to HPV-status; RB1/TP53 wildtype tumors harboring HPV as one group distinct to RB1/TP53 co-mutant tumors without association to HPV [77]. The high frequency of RB1 mutations in ocular adnexal sebaceous carcinoma leads to a compensatory increase in p16<sup>INK4a</sup> expression. Hence, p16<sup>INK4a</sup> expression is not a valuable surrogate marker of HPV infection in ocular adnexal sebaceous carcinoma [2,74].

Table 5. An overview of studies examining HPV in sebaceous gland carcinomas of the ocular adnexa.

| Author, Year | Median Age Years (Range)/Gender | HPV+ (DNA PCR) | HPV+ (RNA ISH) | HPV Genotypes | HPV Detection Modality |
|--------------|--------------------------------|----------------|----------------|---------------|-----------------------|
| Hayashi et al., 1994 [72] | 63 (52–83)/6M:7F | 13/21 (62%) | NA | 16, 18, 31, 33, 6, 11 | DNA ISH |
| Gonzalez-Fernandez et al., 1998 [75] | 72 (32–90)/7F | 0/7 (0%) | NA | NA | DNA ISH, PCR |
| Kwon et al., 2015 [74] | 72 (45–86)/4M:10F | 0/14 (0%) | NA | NA | HPV chip test |
| Liau et al., 2014 [79] | NA/8M:16F | 1/24 (4%) | NA | 16 | PCR |
| Stagner et al., 2016 [2] | NA | 1/24 (4%) | 0/18 (0%) | 16 | PCR, RNA ISH |
| Tetzlaff et al., 2019 [77] | 68 (44–93)/13M:16F | 4/29 (14%) | 4/29 (14%) | 16, 18 | RNA ISH, RNA sequencing |
| Chauhan et al., 2019 [76] | mean 56.8±13.9 (25–88)/16M:14F | 0/30 (0%) | NA | NA | PCR |
| Moore et al., 2021 [78] | mean 73 (27–98)/8M:10F | NA | 2/11 (18%) | High-risk HPV | RNA ISH |

HPV; Human papillomavirus, PCR; Polymerase chain reaction, ISH; in-situ hybridization, NA; not applicable.

Overall, the small sample sizes of the studies and the low prevalence of HPV in ocular adnexal sebaceous carcinoma lead to broad variations in the reported HPV prevalence. Nevertheless, the detection of transcriptionally active HPV within the tumor cells is a strong predictor of HPV involvement in a subset of ocular adnexal sebaceous carcinoma.

4.2. Cutaneous Squamous Cell Carcinoma

Cutaneous squamous cell carcinoma (SCC) is the second most common eyelid malignancy after basal cell carcinoma comprising approximately 10% of all eyelid malignancies [80,81]. Exposure to ultraviolet (UV) radiation and immunosuppression are the main risk factors for developing the disease. Moreover, β genus HPV types are suspected to promote cutaneous SCC formation, especially in immunocompromised patients [82–84]. Synergistically, the β genus HPV types are thought to interact with UV-radiation-induced genomic aberrations to promote cancer development [84]. There are several factors that question causality between HPV and cutaneous SCC. Most importantly, many HPV genotypes with cutaneous tropism are considered commensal viruses, hence a part of the human virome, causing asymptomatic infections rather than neoplastic disease [85]. Moreover, it seems as though β genus HPV is not required for cutaneous SCC maintenance and growth—different from the “classic” α-genus induced HPV-carcinogenesis [84]. Hence, the presence of HPV in apparently healthy skin, and the lack of viral expression in late stages of the disease questions the importance of HPV in this disease. The association between β genus HPV and eyelid cutaneous SCC is still uncertain.

5. Conclusions

In recent years, more sophisticated molecular techniques, including RNA ISH and high-throughput sequencing, have made it possible to demonstrate the transcription of HPV oncogenes in carcinomas of the ocular adnexa, even in older and formalin-fixed and paraffin-embedded (FFPE) tumor tissue. Today, increasing evidence supports that defined subsets of benign and malignant neoplasia in the ocular adnexa develop in an HPV-dependent pathway. The most frequently involved genotypes in both benign (HPV6, 11) and malignant (HPV16, 18) neoplasia of the ocular adnexa are covered by the L1 prophylactic HPV vaccines and are thereby potentially preventable by vaccination (Figure 4).
HPV status serves as a diagnostic biomarker a subset of head-and-neck carcinomas, and tremendous efforts are now put into the use of HPV status to up and de-escalate the treatment regimens. In the ocular adnexal region, many pieces of the puzzle are still missing in order to incorporate HPV status to the benefit of the patients. To elucidate further the role of HPV in ocular adnexal papillomas and carcinomas, the HPV status, preferentially investigated by the expression of viral oncoproteins or a combination of HPV DNA PCR and p16INK4a immunohistochemistry, should be included in future studies of all these tumors.

**Author Contributions:** Conceptualization, I.R. and S.H.; literature search, I.R.; writing—original draft preparation, I.R.; writing—review and editing, S.H.; visualization, I.R.; supervision, S.H.; funding acquisition, I.R. and S.H. Both authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by the Synoptik Foundation, Denmark; the Danish Eye Research Foundation, Denmark; the Fight for Sight Foundation, Denmark; the Faculty of Health and Medical Sciences, Copenhagen University, Denmark; the Svend Arvid Schröder and Ketty Lydia Larsen Schröder Foundation, Denmark; and Copenhagen University Hospital, Rigshospitalet, Denmark.

**Institutional Review Board Statement:** Not applicable.

**Informed Consent Statement:** Not applicable.

**Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript; or in the decision to publish the results.

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