Marine Natural Products Against Tuberculosis

Marcus Vinícius Nora De Souza
FioCruz-Fundação Oswaldo Cruz, Instituto de Tecnologia em Fármacos-Far
Manguinhos, Rua Sízenando Nabuco, 100, Manguinhos, 21041-25650 - Rio de Janeiro-RJ – Brazil

E-mail: mvndesouza@click21.com.br

Received May 8, 2006; Revised June 28, 2006; Accepted July 10, 2006; Published July 21, 2006

Natural products represent an outstanding source of compounds that play an important role in the treatment of human diseases. Due to the importance of nature as a source of new drug candidates, the aim of this review is to highlight the marine natural products, which exhibit antituberculosis activity, discovered between 2000 and 2005.

KEYWORDS: tuberculosis, marine natural products, drugs

INTRODUCTION

Tuberculosis (TB) is an infection transmitted through the air caused by the bacterium *Mycobacterium tuberculosis* (identified in 1882 by R. Koch) that mainly affects and damages the lungs, but it may spread across almost any tissue or organ of the body[1]. TB became a worldwide problem between 1985 and 1992 when the number of TB cases increased, particularly in people infected with the HIV virus. Another problem that increased TB cases is multidrug-resistant tuberculosis (MDR TB) due to inconsistent or partial treatment. Because of these problems, the World Health Organization (WHO) declared TB a global health emergency in 1993. Statistics inform us that 8 million people became sick with TB last year with almost 3 million deaths that represent, in general, four deaths each minute. It is estimated that 30 million people will die in the next 10 years[2]. At present, the accepted treatment of TB involves a combination of the first-line drugs, isoniazid 1, pyrazinamide 2, ethambutol 3, and rifampicin 4 (Fig. 1) given in combination over 6–9 months. The combinations are very important to prevent the emergence of multiple drug-resistant organisms, which would lead to an ineffective treatment[2].

In spite of the worldwide problem caused by TB, it has been nearly more than 30 years since a novel drug has been introduced to treat TB. In this context, marine natural products became a powerful source of new anti-TB compounds that will be covered in this review.

MARINE NATURAL PRODUCTS

The interest and study of marine natural products against *M. tuberculosis* started with Andersen et al.[3] and Hamman et al.[4]. Andersen and coworkers reported the *in vitro* anti-TB activity of massetolide A 5 (MIC = 5–10 μg/ml) and viscosin 6 (MIC = 10–20 μg/ml)[3], cyclic depsipeptides isolated from a marine alga and tube worm, respectively (Fig. 2).
MIC - It is defined as the minimal inhibitory concentration of an antibiotic that inhibits a bacterium.

**FIGURE 1**

**FIGURE 2**
Hamman and his group identified anti-TB activity in two known polypeptides isolated from the sacoglossan mollusk *Elysia rufescens*, Kahalalide A 7 and F 8 (Fig. 3)[4]. The first one has demonstrated important *in vitro* activity, inhibiting 83% the *M. tuberculosis* growth (H37Rv), at 12 μg/ml, and the second one, 67%, at the same concentration.

**Kahalalide A (7)** Inhibition activity = 83%
(MIC = 12.0 μg mL⁻¹)

**Kahalalide F (8)** Inhibition activity = 67%
(MIC = 12.0 μg mL⁻¹)

FIGURE 3
Hamann and coworkers also made an important contribution by testing in vitro activities against *M. tuberculosis*, marine natural and semisynthetic molecules from different classes, in the total of 48 compounds[5]. The above work has shown potential candidates for new anti-TB agents and has indicated the importance of investigating marine natural products as a new source of antimycobacterial agents. As examples of the compounds tested by Hamann and his group, the C-19 hydroxysteroids linosterol 9, nephalsterol B 10, and C 11 that were isolated from red sea *Nepthea* sp. (Fig. 4) can be mentioned[6]. The natural product linosterol 9 and nephalsterol C 11 had MICs of 3.13 and 12.5 μg/ml inhibiting 90 and 96% of the growth of *M. tuberculosis* (H37Rv). However, nephalsterol B inhibited only 69% at the same concentration, which indicated that C-7 hydroxylation could reduce the activity. When C-7 hydroxyl was blocked by an acetate group as in the case of nephalsterol C 11 or when it was absent as in litosterol 9, an improvement of the biological activity was observed. Hamann and his group were the first to identify antimycobacterial activity in this class of compounds.

![Linosterol (9)](image)

Linosterol (9)  
\(R_1\) \(R_2\)  
H \(\text{OH}\)  
Inhibition activity = 90\%  
(MIC = 3.13 \(\mu\text{g mL}^{-1}\))

![Nephalsterol (B) (10)](image)

Nephalsterol (B) (10)  
\(R_1\) \(R_2\)  
OH \(\text{OH}\)  
Inhibition activity = 69\%  
(MIC = 12.5 \(\mu\text{g mL}^{-1}\))

![Nephalsterol (C) (11)](image)

Nephalsterol (C) (11)  
\(R_1\) \(R_2\)  
\(\text{OAc}\) \(\text{OH}\)  
Inhibition activity = 96\%  
(MIC = 12.5 \(\mu\text{g mL}^{-1}\))

FIGURE 4

Hamann and coworkers have also isolated 27 diterpenes called cyanthiwigins from Jamaican sponge *Mymekioderma styx*[7], which have been tested against *M. tuberculosis*. However, the best result was the moderate activity of cyanthiwigin (C) 12[8] with 50% of inhibition at 6.25 μg/ml (Fig. 5).

![Cyanthiwigins (C) (12)](image)

Cyanthiwigins (C) (12)  
Inhibition activity = 50\%  
(MIC = 6.25 \(\mu\text{g mL}^{-1}\))

FIGURE 5
Another contribution was also done by König and coworkers who have tested 39 different marine natural products representative of different classes, such as terpenes, aliphatics, aromatics, alkaloids, and sterols[9]. In this context, 15 showed MICs of 32 μg/ml or less, and 11 had MICs of 16 μg/ml or less. The most promising compound reported in the König study was axisonitrile-3 13 (MIC = 2.0 μg/ml) (Fig. 6) isolated from the sponge *Acanthella klethra* with no toxicity to Vero cells at concentrations < 200 μg/ml[9].

![Axisonitrile-3 (13)](image)

(MIC = 2.0 μg mL⁻¹)

**FIGURE 6**

Manzamines are β-carbolines alkaloids from the Okinawan sponge genus *Haliclona* that were first reported in 1986 by Higa and coworkers[10]. This class of compounds possesses a diverse range of bioactivities, such as antibacterial[11], cytotoxic[12], and malaria activity[13]. In the search for new types of anti-TB agents, Hamman and coworkers have isolated and tested new manzanines from Indonesian sponges[14]. As examples, the two novel alkaloids called manadomanzamines A 14 and B 15 (Fig. 7) can be mentioned, isolated from Indonesian sponge *Acanthostrongylophora* sp. (Haplosclerida: Petrosiidae)[15] that exhibited strong activity against *M. tuberculosis* with MIC values of 1.9 and 1.5 μg/ml, respectively. These two alkaloids also exhibited significant activity against human immunodeficiency virus (HIV-1) with EC₅₀ values of 7.0 and 16.5 μg/ml, respectively, and moderate activity against several AIDS opportunistic infections.

![Manadomanzamine A (14): 22 β − H (MIC = 1.9 μg mL⁻¹)](image)

Manadomanzamine B (15): 22 α− H (MIC = 1.5 μg mL⁻¹)

**FIGURE 7**
Rodrigues and coworkers have been an important contribution towards the search for new natural products against TB. They have purified and identified the natural products elisapterosin B 16, elisabethin A 17[16], cumbiasin A 18 and B 19[17], and colombiasin A 20[18] (Fig. 8). These compounds have similar carbocyclic skeletons and they were isolated from an extract of the West Indian corals gorgonian octocoral *Pseudopterogorgia elisabethae* Bayer (order Gorgonacea, family Gorgoniidae, phylum Cnideria) from the waters near San Andrés Islands, Colombia, showing potent activity against *M. tuberculosis*. The elisapterosin B 16 (Fig. 8) has been found to have potent inhibitory activity against *M. tuberculosis* H37Rv at a concentration of 12.3 μg/ml[16]. In the case of cumbiasin A 18 and B 19, they display mild *in vitro* activity against H37Rv, 6.25 and 12.5 μg/ml, respectively, and both compounds caused 17% growth inhibition of *M. tuberculosis*[18]. The pharmacological properties against *M. tuberculosis* of elisabethin A 17 and colombiasin A 20 (Fig. 8) have not yet been published.

![Elisapterosin B](image1)

*Elisapterosin B*  
(MIC = 123 μg mL⁻¹)

![Cumbiasin A](image2)

*Cumbiasin A*  
(R = H, MIC = 6.25 μg mL⁻¹)

*Cumbiasin B*  
(R = OH, MIC = 12.5 μg mL⁻¹)

![Elisabethin A](image3)

*Elisabethin A*

*Colombiasin A*

MIC have not yet been determined

**FIGURE 8**

Other products that were identified by Rodrigues and coworkers from the same source of *P. elisabethae* also showed activity against *M. tuberculosis*. The three alkaloids, pseudopteroxazole 21, homopseudopteroxazole 22, and seco-pseudopteroxazole 23 (Fig. 9)[19] had potent to moderate inhibitory activity against *M. tuberculosis* H37Rv (97, 88, and 66%), respectively, at a concentration of 12.5 μg/ml. Another example is the two serrulatane-based diterpenes, erogorgiaene 24 and 7-hydroxyergorgiaene 25, which induced 96 and 77% inhibition of *M. tuberculosis* H37Rv at concentrations of 12.5 and 6.25 μg/ml, respectively. The biological results indicated that the oxazole moiety is not essential for activity[19].
Due to their biological activity, complex molecular structure, and functionalization of the compounds present in *P. elisabethae*, many organic synthetic groups have been working in various synthetic strategies to prepare compounds of this class[20]. A good example is the elegant enantiospecific total synthesis of the pseudopteroxazole 21 (Scheme 1) made by Corey and Davidson[21]. They have used (S)-(-)-limonene 26 as starting material and after cyclic hydroboration and alkaline peroxide oxidation, the diol 27 was obtained. Under selective oxidation with sodium hypochlorite in aqueous acetic acid, this intermediate furnished the diastereoisomeric mixtures of hydroxy ketones 28, which were enzymatically resolved, and after protection of the hydroxyl as the tert-butyl-diphenylsilyl ether (TBDPS), afforded the synthon 29 in 96% yield. The selective kinetic deprotonation of the intermediate using lithium diisopropylamide (LDA) and trapping chlorotrimethylsilane (TMSCl) furnished the desired silyl enol ether, which after treatment with 1-benzzyloxy-3-methyl-but-3-en-2-one and SnCl₄, gave the Mukaiyama-Michael adduct 30 in 61%. The α,β-ene 31 was obtained as a mixture of diastereoisomers after cyclization of the 1,5-diketone in dilute solution (0.01 M) of potassium hydroxide in ethanol (83%) and elimination of the tertiary hydroxyl group in presence of thionyl chloride in pyridine with 83% yield. The oxime pivalate 32 was provided in 96% yield after the oxime formation using hydroxylamine followed by acylation with pivaloyl chloride. The aromatization and N-acylation of the intermediate was carried out using an acetyl chloride at 80°C to furnish 33 in 64% yield[21]. The hydrogenolysis of the benzyl ether (99% yield), cyclization of the phenol with carbonyldimidazole, and mildly basic aqueous workup (NaHCO₃) to remove the N-acyl group afforded the cyclic carbamate 34 in 94% yield. The deprotection in presence of HF-pyridine (95% yield), followed by mild oxidation to the aldehyde with tetrapropylammonium perruthenate and 4-methylmorpholine-N-oxide (TPAP-NMO, 82% yield) and E-selective olefination, gave the diene 35 in 81% yield. The key intermediate 36 was obtained in 80% yield by cationic cyclization in the presence of three equivalents of methanesulfonic acid and acetic acid as solvent at 19°C. After N-protection using di-tert-butyl dicarbonate (BOC₂O) (96% yield) and excess of the methylmagnesium bromide (CH₃MgBr) to cleave the cyclic carbamate, followed by addition of the resulting dianion to a mixture of trifluoroacetic acid and triethylorthoformate, the natural product pseudopteroxazole 21 was achieved in 90% yield (Scheme 1)[21].
Pseudopteroxazole
Inhibition activity 97%  
(MIC = 12.5 μg mL⁻¹)

SCHEME 1
(a) Thexylboranes, alkaline peroxide oxidation, 92%; (b) NaClO, 86%; (c) AcOC(CH₃)CH₂ lipase; (d) TBDPSCl, imidazole, DMF, 96%; (e) LDA, TMSCl, –78°C, 100%; (f) 1-benzyloxy-3-methyl-but-3-en-2-one, SnCl₄, CH₂Cl₂, –78°C, 61%; (g) KOH, EtOH, –10°C, 83%; (h) SOCl₂, py, 83%; (i) NH₂OH-HCl, py, 69%; (j) pivaloyl chloride, py, 96%; (k) acetylchloride, toluene, 80°C, 64%; (l) H₂, Pd(OH)₂/C, EtOH, 99%; (m) carboxyldiimidazole, Et₃N; then, aqueous NaHCO₃, 94%; (n) HF-py, 95%; (o) TPAP, NMO, 82%; (p) Wittig olefination, 81%; (q) MeSO₃H, AcOH, 18°C, 80%; (r) BOC₂O, DMAP, 96%; (s) MeMgBr, –78 to 23°C, then (t) TFA, HC(OEt)₃, 90%.

The phytosterol saringosterol 37 (Fig. 10), isolated by Timmermann and coworkers from the Chilean brown algae Lessonia nigrescens Bory (Phaeophyta, Laminariales)[22], showed potent anti-TB activity against the H₃₇Rv strain of M. tuberculosis. The 24S epimer of saringosterol presented MIC values of 0.125 μg/ml, eight times more active than their epimer 24R (1.0 μg/ml), indicating the importance of the hydroxyl position for biological activity. Due to its specificity, activity, and low toxicity, this class of compounds could be a good target for the development of new TB drugs[22].

![Saringosterol (37)](image)

Berlinck and coworkers have isolated different compounds and the ingenamine G 38 from a cytotoxic and anti-TB, active methanol, crude extract of the marine sponge Pachychalina sp.[23]. The latter showed biological activity against M. tuberculosis (H37Rv) at 8 μg/ml (Fig. 11).

![Ingenamine G (38)](image)
Orabi and coworkers[24] identified potent anti-TB active in (+)-araguspongine C 39 (MIC = 3.94 μg/ml) (Fig. 12)[24], a natural product isolated from red sea specimens of *Xestospongia exigua*[25]. This class of compound possesses diverse important biological activities such as vasodilatation[26], cytotoxicity[27], antifungal[28], and vasoactive intestinal peptide inhibition activity[29].

![Image of (+)-Araguspongines C (39)](image)

**FIGURE 12**

Ecteinascidins 770 40 and 786 41 (Fig. 13) were isolate by Suwanborirux and coworkers from Thai tunicate *Ecteinascidia thurstoni*[30]. This class of compounds showed potent cytotoxic activity with the ecteinascidin 743 42 under phase II clinical trials in the treatment of cancer[31]. Suwanborirux and his group found potent anti-TB activity against *M. tuberculosis* (H37Rv) in ecteinascidins 770 40 and 786 41 with MIC values of 0.13 and 2.0 μg/ml, respectively (Fig. 13)[30].

![Image of Ecteinascidin](image)

**FIGURE 13**
Plubrukarn and coworkers isolated and tested different sesterpenes from Thai sponge B *Brachiaster* sp. against *M. tuberculosis* (H37Rv). As examples, the 12-deacetoxyscalarin 19-acetate 43 and the manoalide 25-acetate 44 (Fig. 14), with MIC values of 4 and 7 µg/ml, respectively, can be mentioned[32].

Bewley and coworkers isolated and identified new alkaloids 45–49 from Australian nonverongid sponge (Fig. 15)[33]. These compounds represent the first examples of inhibitors of a new mycobacterial enzyme S-conjugate amidase (MCA) that has been characterized from *M. tuberculosis* and *M. semegmatis*.

Due to the significant inhibitory activity of the alkaloid 47 against the MCA, Kende and coworkers were the first to synthesize the dibromotyrosine alkaloid 47 (Scheme 2)[34]. This synthesis used 4-hydroxyphenylpyruvic acid 50 as starting material, which was converted to the THP-oxime acid 51 in 90% yield, followed by the coupling with the protected 4-aminobutylguanidine 52 to afford the intermediate 53 in 87%. This intermediate was dibrominated to the dibromophenol amide 54 with N-bromosuccinimide (NBS) in 89% yield followed by the Mitsunobu coupling of the phenolic hydroxyl with 3-(t-butoxycarbonylamino)propanol, which furnished the compound 55 in 82% yield. The final step was carried out by removing all the protecting groups in the presence of 30% trifluoroacetic acid in dichloromethane to produce the target compound 47 as its bistrifluoroacetate salt in 82% yield[34].

![12-Deacetoxyscalarin 19-acetate (43)](image1)

(MIC = 4.0 µ g mL⁻¹)

![Manoalide 25-acetate (44)](image2)

(MIC = 7.0 µ g mL⁻¹)

FIGURE 14
De Souza: Marine Natural Products Against Tuberculosis

FIGURE 15

\[
\begin{align*}
\text{IC}_{50} &= 3 \mu M \\
\text{IC}_{50} &= 100 \mu M \\
\text{IC}_{50} &= 30 \mu M
\end{align*}
\]
**CONCLUSION**

To fight efficiently against the rapid spread of MDR TB strain against all major anti-TB drugs in the market, it is imperative have new drugs with fewer toxic side effects, improved pharmacokinetics properties, extensive and potent activity against Gram-positive and Gram-negative bacteria, including resistant strains, and drugs able to reduce the total duration of treatment. In this context, natural products, particularly from the ocean, offer unique chemical diversity with the potential to open a new era in TB treatment.

**ACKNOWLEDGMENTS**

The author thanks Solange M., S.V. Wardell, and James L. Wardell for reading this manuscript.
REFERENCES

1. Munch, R. (2003) *Microb. Infect.* 69, 74; Kaufmann, S.H.E. (2003) *Tuberculosis* 83, 86.
2. Coker, R.J. (2003) *Public Health* 117, 281; Mitscher, L.A. and Baker, W. (1998) *Med. Res. Rev.* 18, 363; Souza, M.V.N. (2006) *Curr. Opin. Palm. Med.* 3, 167; Souza, M.V.N. (2006) *Recent Patents on Anti-Infective Drug Discovery* 1, 33; Souza, M.V.N. (2005) *TheScientificWorldJournal* 5, 609.
3. Gerard, J., Lloyd, R., Barsby, T., Haden, P., Kelly, M.T., and Andersen, R.J. (1997) *J. Nat. Prod.* 60, 223; Copp, B.R. (2003) *Nat. Prod. Rep.* 20(6), 555.
4. Hamann, M.T. and Scheuer, P.J. (1993) *J. Am. Chem. Soc.* 115, 5825; Hamann, M.T., Otto, C.S., Scheuer, P.J., and Dunbar, D.C. (1996) *J. Org. Chem.* 61, 6594; Bonnard, I., Manzanares, I., and Rinehart, K.L. (2003) *J. Nat. Prod.* 66, 1466; Bourrel-Bonnet, L., Rao, K.V., Hamann, M.T., and Ganesan, A. (2005) *J. Med. Chem.* 48(5), 1330.
5. Sayed, K.A., Bartyzel, P., Shen, X., Perry, T.L., Zjawiony, J.K., and Hamann, M.T. (2000) *Tetrahedron* 56, 949.
6. Iguchi, K., Saitoh, S., and Yamada, Y. (1989) *Chem. Pharm. Bull.* 37, 2553; Liu, J., Zeng, L., and Wu, D. (1992) *Gaodeng Xuexiao Huaxue Xuebao* 13, 341; *Chem. Abstr.* 117, 23041y.
7. El Sayed, P.J., Walsh, K., Weedman, V., Bergthold, J.D., Lynch, J., Lieu, K.L., and Hamann, M.T. (2002) *Tetrahedron* 58, 7809; Luesch, H., Pangilinan, R., Yoshida, W.Y., Moore, R.E., and Paul, V.J. (2001) *J. Nat. Prod.* 64(3), 304.
8. Sennett, S.H., Pomponi, S.A., and Wright, A.E. (1992) *J. Nat. Prod.* 55, 1421.
9. Konig, G.M., Wright, A.D., and Franzblau, S.G. (2000) *Planta Med.* 66, 337.
10. Sakai, R., Higa, T., Jefford, C.W., and Bernardinelli, G. (1986) *J. Am. Chem. Soc.* 108, 6404; Nakamura, H., Deng, S., Kobayashi, J., and Ohizumi, Y. (1987) *Tetrahedron Lett.* 28, 621.
11. El Sayed, K.A., Kelly, M., Kara, U.A.K., Ang, K.K.H., Katsuyama, I., Dunbar, D.C., Khan, A.A., and Hamann, M.T. (2001) *J. Am. Chem. Soc.* 123, 1804.
12. Longley, R.E., McConnell, O.J., Essich, E., and Harmody, D. (1993) *J. Nat. Prod.* 56, 915.
13. Ang, K.K.H., Holmes, M.J., Higa, T., Hamann, M.T., and Kara, U.A.K. (2000) *Antimicrob. Agents Chemother.* 44, 1645.
14. Yousaf, M., El Sayed, K.A., Rao, K.V., Lim, C.W., Hu, J.F., Kelly, M., Frazblau, S.G., Zhang, F., Peraud, O., Hill, R., and Hamann, M.T. (2002) *Tetrahedron* 58, 7397; Hu, J., Schetz, J.A., Kelly, M., Peng, J., Ang, K.K., Flotow, H., Leong, C.Y., Ng, S.B., Buss, A.D., Wilkins, S.P., and Hamann, M.T. (2002) *J. Nat. Prod.* 65, 476; Rao, K.V., Santarsiero, B.D., Mesecar, A.D., Schinazi, R.F., Tekwani, B.L., and Hamann, M.T. (2003) *J. Nat. Prod.* 66, 823.
15. Peng, J., Hu, J.F., Kazi, A.B., Li, Z., Avery, M., Peraud, O., Hill, R.T., Franzblau, S.G., Zhang, F., Schinazi, R.F., Wirtz, S.S., Tharnish, P., Kelly, M., Wahiyoumo, S., and Hamann, M.T. (2003) *J. Am. Chem. Soc.* 125, 13382.
16. Rodriguez, A.D., Gonzalez, E., and Huang, S.D. (1998) *J. Org. Chem.* 63, 7083; Rodriguez, A.D. Ramirez, C., Rodriguez, I.I., and Barnes, C.L. (2000) *J. Org. Chem.* 65, 1390.
17. Rodriguez, A.D., Ramirez, C., and Shi, Y.P. (2000) *J. Org. Chem.* 65, 6682.
18. Rodriguez, A.D. and Ramirez, C. (2000) *Org. Lett.* 2, 507.
19. Rodriguez, A.D. and Ramirez, C. (2001) *J. Org. Chem.* 64, 100.
20. Johnson, T.W. and Corey, E.J. (2001) *J. Am. Chem. Soc.* 123, 4475; Harmata, M., Hong, X., and Barnes, C.L. (2004) *Org. Lett.* 13, 2201; Dehmel, F. and Schmalz, H.G. (2001) *Org. Lett.* 3, 3579.
21. Corey, E.J. and Davidson, J.P. (2003) *J. Am. Chem. Soc.* 125, 13486.
22. Wachter, G.A., Franzblau, S.G., Montenegro, G., Hoffmann, J.J., Maiese, W.M., and Timmermann, B.N. (2001) *J. Nat. Prod.* 64, 1463.
23. De Oliveira, J.H.H., Grube, A., Kock, M., Berlinck, R.G.S., Macedo, M.L., Ferreira, A.G., and Hajdu, E. (2004) *J. Nat. Prod.* 67, 1685.
24. Orabi, K.B., El Sayed, K.A., Hamann, M.T., Dunbar, D.C., Al Said, M.S., Higa, T., and Kelly, M. (2002) *J. Nat. Prod.* 65, 1782.
25. Rao, J., Desaiah, D., Vig, P., and Venkateswarlu, Y. (1998) *Toxicology* 129, 103.
26. Gafni, J., Munsch, J., Lam, T., Catlin, M., Costa, L., Molinski, T., and Pessah, I. (1997) *Neuron* 19, 723; De Smet, P., Parys, J., Callweaerts, G., Weidema, A., Hill, E., De Smet, H., Erneux, C., Sorrentino, V., and Missiaen, L. (1999) *Cell Calcium* 26, 9.
27. Pettit, G., Orr, B., Herald, D., Dobbek, D., Tackett, L., Schmidt, J., Boyd, M., Pettit, R., and Hooper, J. (1996) *Bioorg. Med. Chem. Lett.* 6, 1313.
28. Moon, S., MacMillan, J., Olimstead, M., Ta, T., Pessah, I., and Molinski, T. (2002) *J. Nat. Prod.* 65, 249.
29. Vassas, A., Bourdy, G., Paillard, J., Lavayer, J., Paix, M., Quirion, J., and Debitus, C. (1996) *Planta Med.* 62, 28.
30. Suwanborirux, K., Charupant, K., Amnuoypol, S., Pumangura, S., Kubo, A., and Saito, N. (2002) *J. Nat. Prod.* 65, 935.
31. Izbricka, E., Lawrence, R., Raymond, E., Eckhardt, G., Faircloth, G., Jimeno, J., Clarck, G., and Von Hoff, D.D. (1998) *Ann. Oncol.* 9, 981; Ghielmini, M., Colli, E., Erba, E., Bergamaschi, D., Pampallona, S., Jimeno, J., Faircloth, G., and Sessa, C. (1998) *Ann. Oncol.* 9, 989; Hendriks, H.R., Fiebig, H.H., Giavazzi, R., Langdon, S.P., Jimeno, J.M., and Faircloth, G.T. (1999) *Ann. Oncol.* 10, 1233; Jin, S., Gorfajn, B., Faircloth, G., and Scotto, K. (2000) *Proc. Natl. Acad. Sci. U. S. A.* 97, 6775; Takebayashi, Y., Pourquier, P., Zimonjic, D.B., Nakayama, K., Emmer, S., Ueda, T., Urasaki, Y., Kanzaki, A., Akiyama, S., Popescu, N., Kraemer, K.H., and Pommier, Y. (2001) *J. Nat. Med.* 7, 961.
32. Wonganuchitmeta, S., Yuenyongsawad, S., Keawpradub, N., and Plubrukarn, A. (2004) J. Nat. Prod. 67, 1767.
33. Nicholas, G.M., Newton, G.L., Fahey, R.C., and Bewley, C.A. (2001) Org. Lett. 3, 1543; Fetterolf, B. and Bewley, C.A. (2004) Bioorg. Med. Chem. Lett. 14, 3785; Nicholas, G.M., Eckman, L.L., Newton, G.L., Fahey, R.C., Ray, S., and Bewley, C.A. (2003) Bioorg. Med. Chem. Lett. 11, 601.
34. Kende, A.W., Lan, J., and Fan, J. (2004) Tetrahedron Lett. 45, 133; Encarnacion-Dimayuga, R., Ramirez, M.R., and Luna-Herrera, J. (2003) Pharm. Biol. 41(5), 384.

This article should be cited as follows:
De Souza, M.V.N. (2006) Marine natural products against tuberculosis. TheScientificWorldJournal 6, 847–861. DOI 10.1100/tsw.2006.174.