CASE REPORT

Diffuse parenchymal pulmonary amyloidosis associated with multiple myeloma: a case report and systematic review of the literature

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Abstract

Background: Pulmonary is an uncommon site of extramedullary involvement in multiple myeloma (MM). Diffuse parenchymal amyloidosis as pulmonary manifestation of MM is even rarer. We report a rare case of diffuse parenchymal pulmonary amyloidosis associated with MM diagnosed by video-assisted thoracoscopic lung biopsy (VATLB).

Case presentation: A 58-year-old woman complained of cough and shortness of breath. HRCT disclosed diffuse ground-glass opacifications with interlobular septal thickening in bilateral lungs. A lung-biopsy sample obtained by VATLB revealed Congo Red-positive amorphous eosinophilic deposits in the alveolar septa. Surgical biopsy of abdominal wall skin and subcutaneous fat was also performed, which showed the apple-green birefringence with polarized light on Congo red stain was demonstrated in dermis. The serum immunoelectrophoresis showed monoclonal lambda light chains. A bone marrow biopsy specimen comprised 11.5% plasma cells. She was therefore diagnosed with diffuse parenchymal pulmonary amyloidosis accompanied by MM. The patient was referred to the hematology department for further chemotherapy.

Conclusions: It is important to recognize diffuse parenchymal pulmonary amyloidosis to avoid misdiagnosis.

Keywords: Diffuse parenchymal pulmonary amyloidosis, Multiple myeloma, Amyloidosis

Background

Amyloidosis is a rare disease characterized by the deposition of insoluble misfolded proteins in various tissues and organs. Approximately 6–10 cases occur annually per 100,000 in western Europe and the United States [1–3]. But the exact incidence is unknown.

The respiratory system is involved in 50% of patients with amyloidosis, although radiographic demonstration is much less common [4, 5]. The respiratory amyloidosis may be localized or part of systemic amyloidosis. The three main types of respiratory involvement are tracheobronchial, nodular parenchymal, and diffuse parenchymal pulmonary amyloidosis [4].

Diffuse parenchymal pulmonary amyloidosis, also known as diffuse alveolar-septal amyloidosis, is the least common type of pulmonary amyloidosis. This type is sometimes seen in patients with multiple myeloma (MM) and is associated with a poor prognosis [6]. In this article, we will describe the clinical characteristics of a patient with diffuse parenchymal pulmonary amyloidosis associated with MM diagnosed by video-assisted thoracoscopic lung biopsy (VATLB) in our hospital to improve our understanding of this disease.

Case presentation

A 58-year-old woman presented with a 2-year history of a non-productive cough and progressive shortness of breath. She had a history of renal insufficiency and persistent proteinuria, without any extra-renal involvement. She was diagnosed with IgA nephropathy for 15 years and had received immunosuppressive therapy for 6 years.
Her vital signs were stable at initial examination; the patient was afebrile and oxygen saturation was 95% in ambient air. On physical examination, auscultation of the lungs detected slight coarse crackles at the bilateral bases. The remainder of the examination was unremarkable.

Her hemoglobin was 95 g/L but renal function and calcium level were normal. The patient was negative for antinuclear and antineutrophil cytoplasmic antibodies in immunofluorescence assays. Repeated exams of sputum smear did not yield any pathogenic micro-organisms. Serum free light chain analysis showed measuring lambda light chain of 2.59 g/L. Serum protein electrophoresis revealed low IgG, IgA, and IgM levels, with reported immunoelectrophoresis (IEP) showing monoclonal lambda light chain peak with the monoclonal protein. A 24-h urine contained 5521 mg of protein. High-resolution chest CT revealed disclosed ground-glass opacities (GGOs) with interlobular septal thickening in bilateral lungs (Fig. 1a and b). Chest and abdominal CT shows soft tissue infiltration of the subcutaneous fat layer with asymmetric bulging of the chest and abdominal wall (Fig. 1c and d). Pulmonary function tests revealed a moderately restrictive ventilation disorder, with a forced vital capacity (FVC) of 1.76 L (69.0% of predicted) and severe reduction of diffusion capacity (DLCO SB 20.8% of predicted). Cardiac biomarkers, such as natriuretic peptides, particularly B-type natriuretic peptide (BNP) and cardiac troponin-T were normal. Echocardiogram showed normal left ventricular ejection fraction of 61% and there were no features of cardiac amyloidosis. A VATLB was performed, which showed marked thickening of the alveolar wall deposition of amorphous eosinophilic amyloid at the bronchial mucosa, pulmonary vessel wall and interstitium. Congo red staining display apple-green birefringence in polarised light (Fig. 2a and b). The immunohistochemical stain for protein A was negative for secondary amyloid. Following the VATLB results, bone marrow examination was performed. Bone marrow examination showed 11.5% plasma cells with lambda light chain restriction. Bone marrow cells by flow cytometry exhibited a typical phenotype for plasma cells, expressing monoclonal cytoplasmatic CD138, CD38, and cLambda instead of CD19, CD56 and CKappa, which are characteristic of the typical immunophenotype of MM. Fluorescence in situ hybridization (FISH) was carried out to check on the bone marrow aspirate. The most frequent abnormality in the patients was 1q21 amplification (25%), followed by 14q32 (IGH) translocations (29.5%), without 13q14 (RB1) deletion, 13q14.3 (D13S319) deletion and p53 deletion. No lytic bone lesions were detected with positron emission tomography/computed tomography (PET/CT). This confirmed the diagnosis of MM, λ light chain type, stage I. Surgical biopsy of abdominal wall skin and subcutaneous fat was also performed, which showed the apple-green birefringence with polarized light on Congo red stain was demonstrated in dermis (Fig. 3a and b).

She was therefore diagnosed with diffuse parenchymal pulmonary amyloidosis accompanied by MM. The patient was referred to the hematology department, where the patient started chemotherapy with bortezomib and
dexamethasone, and her symptoms relieved 2 months after the initial presentation. No adverse effects were observed and the laboratory results were stable.

**Discussion and conclusion**

MM accounts for 1% of all cancers and 10% of all hematologic malignancies [7]. Amyloidosis is a rare complication associated with MM. The most commonly affected organs are the kidneys, heart, spleen, lymph nodes and the liver [8]. Pulmonary is an uncommon site of extramedullary involvement in MM. Diffuse parenchymal amyloidosis as pulmonary manifestation of MM is even rarer; only a few cases have been reported [8–14].

Diffuse parenchymal amyloidosis is characterized by the presence of amyloid deposits in the alveolar septa and vessel walls. Pathologic examination of diffuse parenchymal pulmonary amyloidosis shows deposition of amorphous eosinophilic amyloid in the alveolar septa, especially around the capillary vessels [15]. Therefore, HRCT findings in such patients mainly comprise GGOs, interlobular septal thickening, intralobular reticular opacity, and nodules [1, 16]. Diffuse amyloidosis is sometimes accompanied by mediastinal lymphadenopathy. Pleural effusion may be present and occasionally dominate the clinical course. Multiple cysts and calcification probably resulting from fragile alveolar walls as a consequence of amyloid deposition both on alveolar walls and around capillaries have been described [17]. Differential considerations of diffuse parenchymal pulmonary amyloidosis are quite broad and include pneumonia, pneumoconiosis, interstitial lung disease and lymphangitic carcinomatosis.

Diffuse parenchymal pulmonary amyloidosis has a remarkably different, more clinical presentation. Such patients may develop symptoms of coughing and shortness of breath secondary to the amyloid deposits. It is characterized by widespread amyloid deposition involving small vessels and the interstitium [18]. This is reflected by lung function tests showing a restrictive pattern with reduced diffusion capacity of carbon monoxide and hypoxaemia upon exertion [8]. Affected individuals are more likely to progress to pulmonary hypertension and respiratory failure [1].

Tissue biopsy is the gold standard for the diagnosis and typing of amyloidosis. Diagnosis of amyloidosis is confirmed by the presence of apple-green birefringence under polarized light of a tissue biopsy stained with Congo red staining showed marked thickening of the alveolar wall deposition of amorphous eosinophilic amyloid at the bronchial mucosa, pulmonary vessel wall and interstitium. Congo red staining display apple-green birefringence in polarised light.
Lupus erythematosus and Sjögren syndrome are rarely associated with pulmonary hypertension (PH) without PH from amyloidosis secondary to systemic disease. Although transbronchial lung biopsy (TBLB) is useful in some amyloidosis cases, it may be of less efficacy in establishing diagnosis due to the limit of the amount biopsy sampling. The lung VATLB is highly specific and sensitive. The biopsy of a clinically suspected organ is an invasive procedure and may be associated with complications including hemorrhage. Because amyloidosis is a systemic disease, routine biopsies from nonsymptomatic sites including the rectal mucosa, abdominal fat pad and labial salivary glands are more commonly used [19]. But, clinical and radiological manifestations of subcutaneous amyloidosis are very rare. In our case, diffuse soft tissue infiltration of the subcutaneous fat layer with asymmetric bulging of the chest and abdominal wall were demonstrated on CT. Surgical skin biopsy including the subcutaneous fat pad can be performed safely and is useful for diagnosing amyloidosis [20, 21].

Once the diagnosis is clear, diffuse parenchymal pulmonary amyloidosis requires intervention. The therapeutic goal for patients with concurrent pulmonary amyloidosis and MM is suppression of the production of amyloid protein comprising immunoglobulin light chain [7, 22]. Monoclonal antibodies such as daratumumab (Dara, human IgG1 anti-CD38) have shown promising efficacy for the treatment of relapsed and refractory MM and heavily pretreated amyloidosis [23, 24]. Therapy was well tolerated. Prospective studies of daratumumab alone or in combination with chemotherapy are warranted. Treatment of systemic amyloidosis aims at reducing the clonal cell populations producing amyloidogenic immunoglobulins, using high-dose chemotherapy followed by autologous stem cell transplantation in carefully selected patients. Its efficacy in treating diffuse pulmonary amyloidosis has not been established. Lung transplantation for isolated pulmonary amyloidosis has been reported [25]. The AL amyloid then contributed to pulmonary hypertension (PH) with severe symptoms necessitating lung transplantation. Ellender et al. described a case with PH from amyloidosis secondary to systemic lupus erythematosus and Sjögren’s syndrome, the patient received bilateral lung transplantation and remained stable after 7 years post lung transplantation [26]. Lung transplantation for isolated pulmonary amyloidosis or combined with PH may be performed in highly selected patients with good long-term outcome.

Diffuse parenchymal amyloidosis is usually a systemic phenomenon with a poor prognosis. A median survival for untreated patients is 13 months, and with the development of heart failure the survival duration decrease to less than 4 months [18]. When compared with nodular pulmonary amyloidosis, patients with diffuse parenchymal pulmonary amyloidosis has a far worse prognosis [10]. Gradual worsening of pulmonary function and symptoms is typical.

In conclusion, diffuse parenchymal pulmonary amyloidosis is a fatal disorder that is rare and often undiagnosed. Radiologists and physicians should consider amyloid in clinically perplexing chronically ill patients, particularly those with plasma cell dyscrasias or chronic inflammatory states. Tissue biopsy is the gold standard. With the use of subcutaneous fat pad and lung biopsy, an early diagnosis can be made.

**Abbreviations**
- BNP: B-type natriuretic peptide
- FISH: Fluorescence in situ hybridization
- FVC: Forced vital capacity
- GGOs: Ground-glass opacities
- EP: Immunoelectrophoresis
- MWA: Multiple myeloma
- PET/CT: Positron emission tomography/computed tomography
- PH: Pulmonary hypertension
- TBLB: Transbronchial lung biopsy
- VATLB: Video-assisted thoracoscopic lung biopsy

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**Availability of data and materials**
The datasets used and/or analyzed during the current study available from the corresponding author on reasonable request.

**Authors’ contributions**
YL wrote the manuscript and all authors carefully revised the manuscript. YL performed a literature review and data collection to present, write, and revise the proposed manuscript. ZBJ and HYZ cared for and followed up the patient. YWZ, MKS, FQM and QS assisted with the presentation of findings and assisted with drafting and revising the manuscript. HRC managed this case and gave his expert recommendations. All authors have read and approved the final version of this manuscript.

**Ethics approval and consent to participate**
Not applicable.

**Consent for publication**
Written informed consent was obtained from the patient for publication of the case report.

**Competing interests**
The authors declare that they have no competing interests.

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References

1. Czeyda-Pommersheim F, Huang M, Chen SS, Strollo D, Fuhrman C, Bhalla S. Amyloidosis: modern cross-sectional imaging. Radiographics. 2015;35:1381–92.
2. Banyersad SM, Moon JC, Whelan C, Hawkins PN, Wechalekar AD. Updates in cardiac amyloidosis: a review. J Am Heart Assoc. 2012;1:e000364.
3. Simms RW, Prout MN, Cohen AS. The epidemiology of AL and AA amyloidosis. Baillieres Clin Rheumatol. 1994;8:627–34.
4. Urban BA, Fishman EK, Goldman SM, Scott WW Jr, Jones B, Humphrey RL, Hruban RH. CT evaluation of amyloidosis: spectrum of disease. Radiographics. 1993;13:1295–308.
5. de Almeida RR, Zanetti G, Pereira E, Silva JL, Neto CA, Gomes AC, Meirelles GS, da Silva TK, Nobre LF, Hochhegger B, Escuissato DL, Marchiori E. Respiratory tract amyloidosis. State-of-the-art review with a focus on pulmonary involvement. Lung. 2015;193:875–83.
6. Milani P, Basset M, Russo F, Foli A, Palladini G, Merlini G. The lung in amyloidosis. Eur Respir Rev. 2017;26(149):170046.
7. Rajkumar SV. Multiple myeloma: 2016 update on diagnosis, risk-stratification, and management. Am J Hematol. 2016;91:719–34.
8. Sato H, Ono A, Okada F, Maeda T, Saburi Y, Urabe S, Mori H. A case of diffuse alveolar septal amyloidosis associated with multiple myeloma. J Thorac Imaging. 2015;30:W73–5.
9. Kim SH, Han JK, Lee KH, Won HJ, Kim KW, Kim JS, Park CH, Choi BI. Abdominal amyloidosis: spectrum of radiological findings. Clin Radiol. 2003;58:610–20.
10. Ussavarungsi K, Yi ES, Maleszewski JJ, Kurtin PJ, Dasari S, Theis JD, Dispenzieri A, Ryu JH. Clinical relevance of pulmonary amyloidosis: an analysis of 76 autopsy-derived cases. Eur Respir J. 2017;49(2).
11. Kim SH, Han JK, Lee KH, Won HJ, Kim KW, Kim JS, Park CH, Choi BI. Abdominal amyloidosis: spectrum of radiological findings. Clin Radiol. 2003;58:610–20.
12. Ussavarungsi K, Yi ES, Malekzadeh JJ, Kurtin PJ, Dasari S, Theis JD, Dispenzieri A, Ryu JH. Clinical relevance of pulmonary amyloidosis: an analysis of 76 autopsy-derived cases. Eur Respir J. 2017;49(2).
13. Kim SH, Han JK, Lee KH, Won HJ, Kim KW, Kim JS, Park CH, Choi BI. Abdominal amyloidosis: spectrum of radiological findings. Clin Radiol. 2003;58:610–20.
14. Ussavarungsi K, Yi ES, Malekzadeh JJ, Kurtin PJ, Dasari S, Theis JD, Dispenzieri A, Ryu JH. Clinical relevance of pulmonary amyloidosis: an analysis of 76 autopsy-derived cases. Eur Respir J. 2017;49(2).
15. Kim SH, Han JK, Lee KH, Won HJ, Kim KW, Kim JS, Park CH, Choi BI. Abdominal amyloidosis: spectrum of radiological findings. Clin Radiol. 2003;58:610–20.
16. Ussavarungsi K, Yi ES, Malekzadeh JJ, Kurtin PJ, Dasari S, Theis JD, Dispenzieri A, Ryu JH. Clinical relevance of pulmonary amyloidosis: an analysis of 76 autopsy-derived cases. Eur Respir J. 2017;49(2).
17. Kim SH, Han JK, Lee KH, Won HJ, Kim KW, Kim JS, Park CH, Choi BI. Abdominal amyloidosis: spectrum of radiological findings. Clin Radiol. 2003;58:610–20.
18. Ussavarungsi K, Yi ES, Malekzadeh JJ, Kurtin PJ, Dasari S, Theis JD, Dispenzieri A, Ryu JH. Clinical relevance of pulmonary amyloidosis: an analysis of 76 autopsy-derived cases. Eur Respir J. 2017;49(2).
19. Kim SH, Han JK, Lee KH, Won HJ, Kim KW, Kim JS, Park CH, Choi BI. Abdominal amyloidosis: spectrum of radiological findings. Clin Radiol. 2003;58:610–20.