Improvement of osteogenic potential of biphasic calcium phosphate bone substitute coated with synthetic cell binding peptide sequences

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Purpose: The aim of this study was to evaluate the improvement of osteogenic potential of biphasic calcium phosphate (BCP) bone substitute coated with synthetic cell-binding peptide sequences in a standardized rabbit sinus model.

Methods: Standardized 6-mm diameter defects were created bilaterally on the maxillary sinus of ten male New Zealand white rabbits, receiving BCP bone substitute coated with synthetic cell binding peptide sequences on one side (experimental group) and BCP bone substitute without coating (control group) on the other side. Histologic and histomorphometric analysis of bone formation was carried out after a healing period of 4 or 8 weeks.

Results: Histological analysis revealed signs of new bone formation in both experimental groups (4- and 8-week healing groups) with a statistically significant increase in bone formation in the 4-week healing group compared to the control group. However, no statistically significant difference in bone formation was found between the 8-week healing group and the control group.

Conclusions: This study found that BCP bone substitute coated with synthetic cell-binding peptide sequences enhanced osteoinductive potential in a standardized rabbit sinus model and its effectiveness was greater in the 4-week healing group than in the 8-week healing group.

Keywords: Bone regeneration, Bone substitutes, Maxillary sinus, Oligopeptides, Rabbits.

INTRODUCTION

Various bone substitutes have been used for sinus floor elevation procedures. Autologous bone is considered the gold standard [1-3]. However, the autologous bone transplanted may have an inconsistent rate of mineralization, due to the ratio of cortical to cancellous bone, and given that it also requires an additional donor-site surgery, it may not thus always be the material of choice for sinus floor elevation [3]. Recently, biphasic calcium phosphate (BCP) bone substitute has been investigated and used in sinus floor elevation procedures with favorable treatment outcomes [4-6]. BCP is a proven safe and biocompatible synthetic bone substitute that is widely used in orthopedic, maxillofacial, and periodontal
surgery [7,8]. BCP is known to have mainly osteoconductive properties [8]. Therefore, there have been various efforts to improve the osteogenic potential of this BCP with growth factor, recombinant human bone morphogenetic protein (rhBMP)-2 [9,10]. Previous studies using BCP treated with rhBMP-2 on rat calvarial defects have shown improved osteoinductive potentials when compared with untreated BCP. However, considering side effects such as the swelling of soft tissues and ectopic bone and cyst-like bone void formation, rhBMP-2 should not be applied to all periodontal defects or used in sinus floor elevation [11-13].

Biologically active peptides have been bound to the surface of inorganic bone substitutes to improve the performance of biomaterial with limited osteoinductive potentials [14-16]. PepGen (P-15) is a Food and Drug Administration approved tissue engineered product that contains a synthetic 15 amino acid sequence (766GTPGPQGIAGQRGVV780) of the α1-chain in Type I collagen, which is uniquely involved in binding of cells such as fibroblasts or osteoblasts [17-19]. In previous studies, it was shown that there was an increase in the attachment and viability of cells bound to an inorganic bone substitute with P-15 when compared with those without P-15 [20,21]. Favorable results in animal and clinical studies were also reported when using inorganic bone substitute with P-15 [14, 22]. However, binding of this biologically active peptide to BCP has not yet been attempted or investigated thoroughly. Therefore, the purpose of this study was to evaluate the improvement of osteogenic potential of BCP coated with synthetic peptide (amino acid sequence of 766GTPGPQGIAGQRGVV780).

MATERIALS AND METHODS

Study design

Standardized, bilateral, circular, transosseous, 6-mm diameter defects were created on the maxillary sinuses of the subjects. The bone graft material used in this study was BCP bone substitute, Osteon (Dentium Co., Seoul, Korea), which consists of 70% hydroxyapatite and 30% β-TCP (Table 1).

Preparation of oligopeptide-coated BCP

A peptide sequence of 766GTPGPQGIAGQRGVV780 was obtained from the manufacturer (Sigma-Aldrich Japan, Ishikari, Japan). Surface coating was then performed to combine hydroxyapatite, a constituent of BCP (Osteon, Dentium Co.), with the peptide. At first, the hydroxyl group of hydroxyapatite was linked with 3-aminopropyltriethoxysilane (Sigma-Aldrich Co., St. Louis, MO, USA), then linked with N-succinimidyl 3-maleimidopropionate (Sigma-Aldrich Co.), followed by a linkage with the peptide sequence. The BCP was soaked in the peptide solution and stored at -70°C in a deep-freezer. To sublimate the liquid phase and avoid denaturalizing the peptide, the frozen solution was then placed for a day in a vial in a lyophilizer (Ishin Lab., Seoul, Korea). The lyophilization setting was put at -45°C to -50°C with a high vacuum of 7 to 10 mTorr, and the dried granules were kept in a freezer until use. The peptide-coated BCP was sterilized with ethylene oxide (EO) gas in a gas sterilizer (Steri-Vac 400B, 3M, St. Paul, MN, USA) at 29°C for 5 hours; this is considered the ideal sterilization protocol without hampering osteoinductive activity [23].

Surgical procedure

Ten male New Zealand white rabbits (body weight, 2.5 to 3.0 kg) were used for this study. The window position on the maxillary sinus of rabbits was determined according to the instructions of Asai et al. [24]: 20 mm anterior to the nasofrontal suture line and 10 mm lateral to the midline.

Prior to all surgical procedures, the animals were anesthetized with a cocktail of ketamine hydrochloride (Ketalar, Yuhan, Seoul, Korea) and xylazine (Rumpun, Bayer Korea Ltd., Seoul, Korea) via an intramuscular injection. The surgical site was then shaved and sterilized with a povidon-iodine solution, followed by local anesthesia of 2% lidocaine with 1:100,000 epinephrine. After making a straight incision along the sagittal midline on the nasal bone, a full-thickness flap was reflected, exposing the calvarial bone. A standardized, circular, bilateral, transosseous window was created on the maxillary sinus using 6-mm-diameter trephine bur while being irrigated copiously with saline. A pin was also inserted at the mid-point between the two windows to indicate the reference point for micro-computed tomography (CT) analysis. Following removal of the trephined calvarial disk, the experimental and control treatments of grafting material were applied to the windows accordingly (Table 1). The periostium was then repositioned over the windows and all surgical sites were sutured with 4-0 Monosyn (Glyconate absorbable monofilament, B-Braun, Aesculap, Center Valley, PA, USA). The animals were euthanized 4 or 8 weeks after the surgery.

Radiographic analysis: micro-CT

The harvested block sections, including the augmented si-
nus and the surrounding bone, were fixed in 10% buffered formalin for 10 days and scanned using micro-CT (SkyScan 1076, SkyScan, Aartselaar, Belgium) at a resolution of 35 μm (100 kV and 100 μA). The area of interest was reconstructed with Ondemand 3D software (Cybermed, Seoul, Korea) and rendered visually in 3D. The CT volume was calculated from the percentage of window closure using an automated image-analysis system (Image-Pro Plus, Media Cybernetics Inc., Silver Spring, MD, USA).

**Histologic analysis**

After the micro-CT image was obtained, each of the sections was decalcified in 5% formic acid for 14 days and then embedded in paraffin. Serial 5-μm-thick sections were cut coronally along the center of the circular calvarial defects. The 2 central-most sections were selected and stained with hematoxylin and eosin (H&E) for light microscope examination (BX50, Olympus Co., Tokyo, Japan).

**Histometric analysis**

An automated image-analysis system (Image-Pro Plus, Media Cybernetics Inc.) was used for computer-assisted histometric analysis. Two parameters were measured: newly formed bone area (NB%) and remaining graft area (RG%). NB% was measured by calculating newly formed mineralized bone in the window as a percentage of total augmented bone area, measured as all tissues within the boundary of the newly formed bone. RG% was measured by calculating the area of the remaining grafting materials as a percentage of the total augmented bone area.

**Statistical analysis**

Micro-CT data and histomorphometric measurement of the samples were used to calculate group mean ± SD values. The Mann-Whitney U test was performed using SPSS ver. 15.0 (SPSS Inc., Chicago, IL, USA) to compare differences between the control group and the experimental group, as well as differences between two experimental groups according to time of healing. A P-value less than 0.05 was considered statistically significant.

**RESULTS**

**Radiographic & histometric analysis**

In the 4-week healing group, NB% was statistically significantly greater in the experimental group than in the control group (\(P<0.05\)). RG% was less in the experimental group than in the control group. However, no statistically significant difference was found in the control and experimental groups’ RG% values. In the 8-week healing group, while CT volume was statistically significantly greater in the experimental group than in the control group (\(P<0.05\)), no statistical difference was found in NB%, despite its increased value (Table 2).

**Histological findings**

Histological findings revealed that increased new bone formation was particularly evident in the experimental group (BCP coated with oligopeptide) when compared to that of the control group (BCP alone). Additionally, more osteoblasts and osteocytes in lacunae were identified in the experimental group (Figs. 1 and 2). In general, within the experimental group, more vascularization and the presence of newly formed blood vessels were identified in the 4-week healing

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**Table 2. Measurements in standardized rabbit sinus model.**

| Point in time | Groups | Materials                        | New bone (%) | Remaining graft (%) | CT volume   |
|--------------|--------|---------------------------------|--------------|---------------------|-------------|
| 4 weeks      | Control| BCP                             | 18.40 ± 4.33 | 32.61 ± 4.70        | 19.85 ± 1.33|
|              | Experimental | BCP coated with oligopeptide | 28.13 ± 6.02 | 29.23 ± 11.97      | 29.57 ± 4.68|
| 8 weeks      | Control| BCP                             | 21.57 ± 6.76 | 27.43 ± 5.96        | 16.27 ± 1.70|
|              | Experimental | BCP coated with oligopeptide | 30.53 ± 5.36 | 26.51 ± 6.17        | 24.53 ± 1.55|

Values are presented as mean ± SD.

CT: computed tomography, BCP: biphasic calcium phosphate.
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group. Immature woven bone, still undergoing the mineralization process, was found to be dominant in the 4-week healing group relative to the 8-week healing group (Fig. 3).

In contrast, the Schneiderian membrane was slightly thickened in the 8-week healing group. The shape of the graft material was also smoothened, which indicates the resorption of graft material particles (Fig. 4). In between graft material particles, Haversian systems and interstitial lamella were observed, both of which are indicative of matured lamellar bone (Fig. 5).

DISCUSSION

There have been many attempts to increase the osteoinductivity of bone graft materials by using a variety of growth factors. The use of BMPs, in particular, has shown favorable results in promoting bone formation [25] and has therefore been investigated to assess its effectiveness through various animal models [26]. However, the use of BMPs also appears to be difficult to handle due to their high molecular weight, the technical difficulties in binding them to bone graft particles, their relatively high cost, and the complexity of immunologic responses [27,28]. Moreover, there is also the limiting factor of establishing an optimum therapeutic concentration of BMPs; since various unwanted side-effects have been reported, such as ectopic bone formation, cyst-like bone void formation, and soft tissue swelling when supraphysiologic doses of BMPs were used [11,12].

Because of this unpredictability, receptor technologies, particularly the use of specific peptide sequences (oligopeptide) involved in binding to cells such as fibroblasts or osteoblasts,
have been posited as alternatives. Saito et al. [29] have reported that 73 to 92 peptide combined with the porous a-TCP scaffold almost completely repaired a 20-mm long rabbit radial bone defect 12 weeks after implantation. Suzuki et al. [30] have also shown that alginate hydrogel linked with synthetic oligopeptide derived from BMP-2 promoted ectopic bone formation. Among many peptide sequences, the use of P-15 in particular, which includes synthetic 15 peptide sequence (766GTPGPQGIAGQRGVV780) of the α1-chain in Type I collagen, has already shown favorable results in the treatment of periodontal defects and in sinus-lifting procedures [14,22]. Park et al. [14] have also reported that 15 amino acids corresponding to the BMP I and II receptor-binding domains coated on granules of bovine bone mineral significantly accelerated new bone formation in a rabbit calvarial defect model. The use of this oligopeptide corresponding to the BMP receptor-binding domains also proved effective in enhancing the bone regenerative capacity in a beagle dog’s 1-wall intrabony defect model [31].

To date, there have been many studies attempting to increase the osteoinductive potential of BCP, one of the most widely used bone graft materials, by coating it with BMPs. However, the effects of binding a biologically active peptide to BCP to increase osteoinductive potential have not been investigated thoroughly. Therefore, the purpose of the present study was to evaluate the improvement of osteogenic potential of BCP coated with synthetic peptide (amino acid sequence of 766GTPGPQGIAGQRGVV780).

In order to provide precisely the same conditions to the control and the experimental group and to minimize procedural errors, the experiment conducted in the study used the standardized rabbit sinus model with the same window shape and size introduced by De Souza Nunes et al. [32]. According to Roberts and Breznak [33], the rabbit metabolic rate is known to be three to four times faster than that of the human. Therefore healing periods of 4 and 8 weeks were chosen based on the assumption that a 4-week and an 8-week healing period for rabbits would correspond to a 3- to 4-month and a 6- to 8-month healing period in humans, respectively.

From histomorphometric analysis, it was found that in the osteogenic potential of BCP, one of the most widely used bone graft materials, by coating it with BMPs. However, the effects of binding a biologically active peptide to BCP to increase osteoinductive potential have not been investigated thoroughly. Therefore, the purpose of the present study was to evaluate the improvement of osteogenic potential of BCP coated with synthetic peptide (amino acid sequence of 766GTPGPQGIAGQRGVV780).

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From histomorphometric analysis, it was found that in the 4-week healing period, new bone formation was statistically greater in the experimental group than the control group. This result is in accordance with previous studies [34-36], as it was thought that bone graft material coated with oligopeptide would promote cell adhesion and cell differentiation, providing more favorable conditions for new bone formation. CT volume was also statistically significantly increased in the experimental group than the control group, which also supports the idea that bone graft materials with oligopeptide coating have more osteoinductive potential. However, in the 8-week healing group, though both new bone formation and CT volume in the experimental group were greater than those of the control group, there was no statistically significant difference in bone formation or CT volume between the 4-week healing group and the 8-week healing group.

Park et al. [14] also reported that oligopeptide domain-coated bovine bone mineral promoted new bone formation at 2 and 4 weeks postsurgery. The results of the current study show the effect of oligopeptide on early bone formation is clearly evident in the 4-week healing group but not as notable in the 8-week healing group. Perhaps, then, it can be suggested that oligopeptide coating may be more beneficial for bone regeneration in the early healing period. On the other hand, the current results may also be due to a high standard deviation resulting from a limited sample size. In this case, further studies with a greater sample size may help to better determine the effectiveness of oligopeptide coating according to healing time.

Within the experimental group, the 8-week healing group showed greater bone formation than did the 4-week healing group, which demonstrates that the process of bone formation continued to occur between the 4th and 8th weeks. For this reason, it may be beneficial to observe the bone formation process even after an 8-week period of time in order to accurately determine the effect of the oligopeptide coating on healing time. It is interesting to note that CT volume in the 8-week healing group was lower. One interpretation of this is that the replacement of biomaterial with newly formed bone does not necessarily occur at a 1:1 ratio, which is in agreement with Frenken et al. [5]. In other words, less bone is produced in this case, compared to the volume of biomaterials resorbed.

In conclusion, this study found that BCP bone substitute coated with biologically active peptide sequences enhanced osteoinductive potential in a standardized rabbit sinus model and, within the limitations of this study, it can be assumed that oligopeptide coating is more beneficial for bone regeneration in the early healing period.

CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

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