Research Article

Influence of IGF2BP2, HMG20A, and HNF1B genetic polymorphisms on the susceptibility to Type 2 diabetes mellitus in Chinese Han population

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Background: The present study aimed to investigate the roles of insulin related gene IGF2BP2, HMG20A, and HNF1B variants in the susceptibility of Type 2 diabetes mellitus (T2DM), and to identify their association with age, gender, BMI, and smoking and alcohol drinking behavior among the Han Chinese population.

Methods: About 508 patients with T2DM and 503 healthy controls were enrolled. Rs11927381 and rs7640539 in IGF2BP2, rs7178572 in HMG20A, rs4430796, and rs11651052 in HNF1B were genotyped by using the Agena MassARRAY. Odds ratio (OR) and 95% confidence intervals (CI) were calculated by logistic regression.

Results: We found that HMG20A rs7178572 (OR = 1.25, P = 0.015) and HNF1B rs11651052 (OR = 1.26, P = 0.019) increased the risk of T2DM. Rs7178572, rs4430796, and rs11651052 might be related to the higher T2DM susceptibility not only by itself but also by interacting with age, gender smoking, and alcohol drinking. Rs11927381 also conferred the higher T2DM susceptibility at age ≤ 59 years. Besides, rs7178572-AA (P = 0.032) genotype and rs11651052 GG (P = 0.018) genotype were related to higher glycated hemoglobin and insulin level, respectively.

Conclusion: Specifically, we first found that rs11927381, rs7640539, and rs11651052 were associated with risk of T2DM among the Han Chinese population. We also provide evidence that age, gender, BMI, smoking, and drinking status have an interactive effect with these variants on T2DM susceptibility.

Introduction

Type 2 diabetes mellitus (T2DM) is a complex, heterogeneous and chronic metabolic disorder, and is characterized by defects in insulin secretion and/or insulin action leading to hyperglycemia [1]. It is reported that about 1 in 11 adults have diabetes mellitus worldwide, 90% of whom have T2DM [2]. International Diabetes Federation Diabetes Atlas 2015 reported that China ranked first in the world for its population of diabetic. In China, the prevalence of T2DM was 9.6% in 2013, and was predicted to reach 13.0% in 2035 [3]. Numerous risk factors have been identified as potential contributors to T2DM susceptibility, such as physical activity, poor dietary condition, increasing obesity, aging, and genetic factors [4]. Recent studies suggested that genetic variants were considered to play a key role in the genesis of T2DM [5,6].

Insulin deficiency is the main characteristic of T2DM. Expression dysregulation of IGF2BP2, HMG20A, and HNF1B genes might affect the level of insulin and lead to the development of T2DM.
IGF2BP2 regulates insulin-like growth factor 2 (IGF2) translation that participates in the growth and insulin signaling pathways [7]. HMG20A is expressed in both human and mouse islets, and the levels of HMG20A are decreased in islets of T2DM donors compared with islets from non-diabetic donors [8]. HNF1B contributes to pancreatic cell formation and controls the specification, growth, and differentiation of the embryonic pancreas [9]. Although some studies have reported the relationship between IGF2BP2, HMG20A, and HNF1B polymorphisms and T2DM risk, the study on other polymorphisms in these genes is insufficient [10–12].

In the present study, we aimed to investigate the association of IGF2BP2 rs11927381 and rs7640539, HMG20A rs7178572, HNF1B rs4430796, and rs11651052 variants with T2DM susceptibility among the Chinese Han population. Given that environment/lifestyle changes can modify the risk of T2DM [13], such as age, gender, body mass index (BMI), and smoking and alcohol drinking behavior, it is interesting to investigate whether these factors have an interactive effect with these variants on T2DM susceptibility.

**Subjects and methods**

**Study participants**

About 508 T2DM patients and 503 age- and gender-matched healthy controls were enrolled into the study from the First Affiliated Hospital of Xi’an Jiaotong University and Xizang Minzu University. All recruited subjects were genetically unrelated ethnic Han Chinese. T2DM cases were identified according to 2017 China Guideline for Type 2 Diabetes. T2DM patients were diagnosed as fasting plasma glucose ≥ 7.0 mmol/l and/or postprandial plasma glucose ≥11.1 mmol/l [14]. Patients who had Type 1 diabetes mellitus, gestational diabetes, acute or other chronic diseases, endocrine disorders, inflammatory diseases, or malignancy were excluded. The inclusion criteria for controls were with normal blood glucose levels and without family history of T2DM and no other chronic diseases. Demographic characteristics and clinical information were collected via standardized questionnaires and medical records. The data included age, sex, BMI, smoking, alcohol drinking, total cholesterol, high-density lipoprotein cholesterol (HDL-C), low-density lipoprotein cholesterol (LDL-C), serum uric acid, creatinine, glomerular filtration rate (GFR), fasting blood glucose, glycated hemoglobin, triglyceride, urea, creatinine, cystatin C, C-reactive protein, insulin, 25 hydroxy-vitamin D, ubiquitin cross-reactive protein (UCRP), and retinol-binding protein. The present study was approved by the ethics committee of Xizang Minzu University (201707) and was conducted in accordance with the declaration of Helsinki. All subjects signed an informed consent before enrolment in the study.

**Genotyping**

Peripheral blood samples were obtained from each subject in vacutainers containing disodium-EDTA anticoagulant. Genomic DNA was isolated using the GoldMag DNA Purification Kit (GoldMag Co. Ltd, Xi’an, China) according to the manufacturer’s protocol, and was stored at −20°C until further analysis. Rs11927381 and rs7640539 in IGF2BP2, rs7178572 in HMG20A, rs4430796 and rs11651052 in HNF1B were selected according to the NCBI dbSNP database (http://www.ncbi.nlm.nih.gov/projects/SNP) and the 1000 Genomes Project data (http://www.internationalgenome.org/), with minor allele frequencies (MAFs) >5% and a pairwise tagging r² of ≥0.8 in Chinese Han population. Genotyping was performed using Agena MassARRAY system (Agena, San Diego, CA, U.S.A.) [15,16], and conducted by two laboratory technicians in double-blind fashion. Primers for PCR amplification and single base extension were listed in Table 1. PCR products were sequenced with Agena MassARRAY Analyzer 4.0 software. Approximately 10% of samples were randomly selected to duplicate genotyping for quality control, and the concordance rates were 100%.

**Statistical analysis**

Statistical analyses were carried out using SPSS version 17.0 (SPSS Inc., Chicago, IL, U.S.A.) and PLINK version 1.0.7. Demographic and clinical data between patients and controls were compared using chi-square test or independent sample T test, as appropriate. Continuous variables and categorical variables were presented as means ± standard deviation (SD) or absolute number (percentage value), respectively. Hardy–Weinberg equilibrium (HWE) for each SNP in the control group was assessed using a goodness-of-fit χ² test. The frequencies of genotype and allele between healthy controls and T2DM patients were compared with χ² test. The correlation between selected SNPs and T2DM risk was estimated by odds ratios (OR) and 95% confidence intervals (CI) using logistic regression models, after adjusting for age and sex [17,18]. To explore the influence of gene–gene interactions on the risk of T2DM occurrence, multifactor dimensionality reduction (MDR) method was used [19]. Further, we stratified by gender, age, BMI, and behavioral factors (smoking and alcohol consumption) to adjust the possible cofounders. The associations of selected SNPs with the clinical parameters in T2DM patients were analyzed by one-way analysis of variance (ANOVA) test. A two-tailed P value < 0.05 was considered statistically significant.
Table 1 Primers sequence of PCR and UEP used in the present study

| Genes | SNPs     | First primer (5′-3′) | Second primer (5′-3′) | UEP_DIR | UEP SEQ (5′-3′) |
|--------|----------|----------------------|-----------------------|---------|-----------------|
| IGF2BP2| rs11927381| ACGTTGGATGAGTCTTATAGTAACTTGAG | ACGTTGGATGAGCCACAAGGAAACTTGATG | R       | cCTTGAGATATT TTGAAAGGTAAC |
| IGF2BP2| rs7640539 | ACGTTGGATGCCACCCCGAGATGATTTTGTC | ACGTTGGATGCACACCTGGCAGTGAAATTG | R       | ggggAAATAGCACTGATACATTGTG |
| HMG20A | rs7178572 | ACGTTGGATGCAACCTCATACCCAAAAATC | ACGTTGGATGGTATGGTTCAAGGTGAGTTG | R       | ACCCAAAAATCTCTTACCA |
| HNF1B  | rs4430796 | ACGTTGGATGCAAACAGAGGAGGCAAGCAC | ACGTTGGATGCAAAGACCCAACAACAGCCTTG | F       | rtGCAAGCAGACAGCTGGA |
| HNF1B  | rs11651052| ACGTTGGATGCCACCGGTGTCTCTTAAGAC | ACGTTGGATGTCTCTCTCAAGGAGGTAC | R       | ccGCGCGCTTTGAGAGTTC |

Abbreviations: DIR, direction; SEQ, sequence; SNP, single-nucleotide polymorphism; UEP, unextended mini sequencing primer.
Table 2 Characteristics of patients with T2DM and controls

| Variable                          | Cases (n = 508)       | Controls (n = 503)  | P      |
|-----------------------------------|-----------------------|---------------------|--------|
| Age, year (mean ± SD)             | 59.21 ± 11.90         | 59.34 ± 7.62        | 0.841  |
| >59                               | 263 (51.8%)           | 265 (52.7%)         |        |
| ≤59                               | 245 (48.2%)           | 238 (47.3%)         |        |
| Gender                            |                       |                     | 0.712  |
| Male                              | 277 (54.5%)           | 279 (55.5%)         |        |
| Female                            | 231 (45.5%)           | 224 (44.5%)         |        |
| BMI (kg/m²)                       |                       |                     |        |
| <24                               | 130 (25.6%)           | 173 (34.4%)         |        |
| ≥24                               | 187 (36.8%)           | 185 (36.8%)         |        |
| Unavailable                        | 191 (37.6%)           | 145 (28.8%)         |        |
| Smoking                           |                       |                     |        |
| Yes                               | 135 (26.6%)           | 115 (22.9%)         |        |
| No                                | 230 (45.3%)           | 188 (37.4%)         |        |
| Unavailable                        | 143 (28.1%)           | 200 (39.8%)         |        |
| Drinking                          |                       |                     |        |
| Yes                               | 68 (13.4%)            | 106 (21.1%)         |        |
| No                                | 277 (54.5%)           | 182 (36.2%)         |        |
| Unavailable                        | 163 (32.1%)           | 215 (42.7%)         |        |
| Total cholesterol (mmol/l)        | 4.61 ± 0.88           | 4.30 ± 1.63         | 0.029  |
| HDL-C (mmol/l)                    | 2.59 ± 0.75           | 2.49 ± 1.16         | 0.172  |
| LDL-C (mmol/l)                    | 1.12 ± 0.25           | 1.50 ± 7.45         | 0.378  |
| Serum uric acid (μmol/l)          | 6.80 ± 19.8           | 5.96 ± 3.38         | 0.396  |
| Creatinine (μmol/l)               | 67.85 ± 32.08         | 65.86 ± 32.18       | 0.390  |
| GFR (ml/min)                      | 95.95 ± 13.11         | 122.61 ± 35.88      | <0.001 |
| Fasting blood glucose             | 9.95 ± 4.70           |                     |        |
| Glycated hemoglobin               | 9.30 ± 2.48           |                     |        |
| Triglyceride                      | 2.50 ± 2.26           |                     |        |
| Urea                              | 6.25 ± 3.19           |                     |        |
| Creatinine                        | 68.97 ± 29.49         |                     |        |
| Cystatin C                        | 0.97 ± 2.18           |                     |        |
| Glomerular filtration rate        | 122.61 ± 35.88        |                     |        |
| C-reactive protein                | 1.38 ± 1.57           |                     |        |
| Insulin                           | 18.80 ± 18.65         |                     |        |
| 25 hydroxy-vitamin D              | 24.69 ± 15.37         |                     |        |
| UCRP                              | 0.54 ± 1.28           |                     |        |
| Retinol-binding protein           | 38.75 ± 11.14         |                     |        |

Abbreviations: BMI, body mass index; HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; T2DM, Type 2 diabetes mellitus; UCRP, ubiquitin cross-reactive protein.
P values were calculated by χ² test for continuous variables and Student’s t test for categorical variables. Bold indicates that P < 0.05 means the data are statistically significant.

Results

About 508 patients with T2DM (59.21 ± 11.90 years, 277 males and 231 females) and 503 healthy controls (59.34 ± 7.62 years, 279 males and 224 females) were included. There was no significant differences in the distribution of age and gender between T2DM patients and the healthy controls (P = 0.841 and P = 0.712, respectively). Demographic and clinical characteristics of participants were listed in Table 2.

Five SNPs (rs11927381 and rs7640539 in IGF2BP2, rs7178572 in HMG20A, rs4430796, and rs11651052 in HNF1B) were successfully genotyped, and all SNPs were in accordance with HWE (P > 0.05, Table 3). The MAF of all SNPs was higher than 5% in T2DM patients and healthy controls. We also found the association of HMG20A rs7178572 and HNF1B rs11651052 with the increased T2DM susceptibility.

The genotype distribution for these SNPs and their relationship with T2DM susceptibility were shown in Table 4. For HMG20A rs7178572, the higher risk of T2DM occurrence was identified in genotype, dominant, and additive
Table 3 The information about the candidate SNPs and associations with the risk of T2DM in allele model

| Genes  | SNPs ID | Chr: Position | Alleles | Frequency (MAF) | P* -value for HWE | OR (95%CI) | P† |
|--------|---------|---------------|---------|----------------|-------------------|------------|----|
| IGF2BP2 | rs11927381 | 3:185790803 | C/T | 0.285 | 1.000 | 1.17 (0.96–1.42) | 0.126 |
| IGF2BP2 | rs7640539 | 3:185795508 | A/T | 0.258 | 1.000 | 1.07 (0.88–1.31) | 0.490 |
| HMG20A | rs7178572 | 15:77454848 | G/A | 0.419 | 0.773 | 1.25 (1.04–1.49) | 0.015 |
| HNF1B | rs4430796 | 17:37738049 | G/A | 0.323 | 0.745 | 1.17 (0.97–1.42) | 0.102 |
| HNF1B | rs11651052 | 17:37742390 | A/G | 0.329 | 0.582 | 1.26 (1.04–1.52) | 0.019 |

Abbreviations: HWE, Hardy–Weinberg equilibrium; MAF, minor allele frequency; SNP, single-nucleotide polymorphism; T2DM, Type 2 diabetes mellitus. P* for HWE values were calculated by χ² test. P† values were calculated by logistic regression analysis with adjustments for age and gender. Bold indicates that P < 0.05 means the data are statistically significant.

Table 4 Relationships between HMG20A and HNF1B polymorphisms and T2DM risk

| Genes  | SNP ID | Model | Genotype | Case | Control | Adjusted by age and gender |
|--------|--------|-------|----------|------|---------|---------------------------|
| HMG20A | rs7178572 | Genotype | AA | 168 | 204 | 1.36 (1.03–1.78) | 0.028 |
|        |        |        | AG | 257 | 230 | 1.50 (1.03–2.18) | 0.037 |
|        |        |        | GG | 85 | 69 | 1.26 (0.89–1.78) | 0.192 |
|        |        | Dominant | AA | 168 | 204 | 1.39 (1.07–1.79) | 0.012 |
|        |        |        | AG-GG | 342 | 299 | 1.42 (1.05–1.87) | 0.015 |
|        |        |        | GG | 85 | 69 | 1.27 (1.04–1.54) | 0.017 |
| HNF1B  | rs11651052 | Log-additive | — | 1 | 1.27 (1.04–1.54) | 0.017 |
|        |        |        | GG | 224 | 257 | 1.30 (1.00–1.68) | 0.050 |
|        |        |        | AG | 236 | 209 | 1.55 (0.98–2.46) | 0.062 |
|        |        |        | AA | 50 | 37 | 1.37 (0.88–2.14) | 0.165 |
|        |        | Dominant | GG | 224 | 257 | 1.33 (1.04–1.71) | 0.023 |
|        |        |        | AG-GG | 286 | 246 | 1.27 (1.04–1.54) | 0.017 |
|        |        |        | AA | 50 | 37 | 1.37 (0.88–2.14) | 0.165 |

Abbreviations: 95%CI, 95% confidence interval; OR, odds ratio; SNP, single-nucleotide polymorphism; T2DM, Type 2 diabetes mellitus. P values were calculated by logistic regression analysis with adjustments for age and gender. Bold indicates that P < 0.05 means the data are statistically significant.

models. Besides, we found that HNF1B rs11651052 variant had an increased risk of T2DM in dominant and additive models.

We further analyzed whether the genotypic effects on T2DM were dependent on gender and age (Table 5). We found that HMG20A rs7178572, HNF1B rs4430796, and rs11651052 were associated with the elevated T2DM susceptibility, especially in males. We found that individuals carrying rs7178572 G allele had an increased T2DM susceptibility under allele, homozygote, heterozygote, dominant, and additive models among males. Rs4430796 polymorphism contributed the risk of T2DM occurrence under allele, homozygote, recessive, and additive models among the male population. Rs11651052 variant was also a risk factor for T2DM among males under allele, homozygote, heterozygote, dominant, recessive, and additive models. Stratified by age, rs11927381, rs7178572, rs4430796, and rs11651052 were associated with the susceptibility to T2DM at age ≤59 years under multiple genetic models (Table 5). In the allele model, rs11927381, rs7178572, and rs11651052 were related to the elevated risk for T2DM. In the homozygote model, rs11927381, rs4430796, and rs11651052 conferred T2DM susceptibility. In the dominant model, rs11927381, rs7178572, and rs11651052 increased T2DM risk. In the recessive model, rs11927381 and rs11651052 had a higher susceptibility for T2DM. In the additive model, rs11927381, rs7178572, and rs4430796, and rs11651052 contributed the developing of T2DM. However, there was a no significant relationship between these variants and T2DM among females or subjects with age ≥59 years.
Stratified analyses were also carried out to estimate the effect of these polymorphisms with BMI and behavioral factors (smoking and alcohol consumption) on T2DM risk, as shown in Table 6. For rs7178572 variant, the G allele carriers had an increased risk of T2DM occurrence among subjects with BMI > 24 kg/m² (allele, homozygote, and recessive), smokers (allele, homozygote, and recessive), or alcohol drinkers (homozygote and recessive). For rs4430796 polymorphism, GG genotype was predominantly related to a higher risk of T2DM among subjects with BMI ≤ 24 kg/m² or drinkers. Besides, rs4430796 also showed a risk-increasing effect among non-drinkers. For rs11651052

### Table 5 Relationships between IGF2BP2, HMG20A, and HNF1B polymorphisms and T2DM risk according to the stratification by gender and age

| SNP ID  | Model | Male OR (95%CI) | Male P | Female OR (95%CI) | Female P | >59 years OR (95%CI) | >59 years P | ≤59 years OR (95%CI) | ≤59 years P |
|---------|-------|----------------|--------|-------------------|----------|---------------------|-------------|---------------------|-------------|
| rs11927381 Allele | 1.29 | 0.061 | 1.03 | 0.828 | 1.00 | 0.976 | 1.41 | 0.020 |
| Homozygote | 1.66 | 0.159 | 1.30 | 0.445 | 0.96 | 0.910 | 2.65 | 0.022 |
| Heterozygote | 1.32 | 0.114 | 0.89 | 0.559 | 1.10 | 0.633 | 1.36 | 0.127 |
| Dominant | 1.36 | 0.069 | 0.96 | 0.824 | 1.07 | 0.704 | 1.50 | 0.036 |
| Recessive | 1.46 | 0.277 | 1.36 | 0.357 | 0.92 | 0.810 | 2.35 | 0.041 |
| Additive | 1.31 | 0.056 | 1.03 | 0.815 | 1.03 | 0.848 | 1.49 | 0.012 |
| rs7178572 Allele | 1.35 | 0.015 | 1.14 | 0.330 | 1.17 | 0.219 | 1.36 | 0.021 |
| Homozygote | 1.74 | 0.033 | 1.26 | 0.424 | 1.40 | 0.205 | 1.66 | 0.098 |
| Heterozygote | 1.46 | 0.044 | 1.25 | 0.284 | 1.22 | 0.314 | 1.50 | 0.050 |
| Dominant | 1.52 | 0.017 | 1.25 | 0.255 | 1.27 | 0.208 | 1.53 | 0.030 |
| Recessive | 1.42 | 0.147 | 1.11 | 0.701 | 1.25 | 0.355 | 1.32 | 0.324 |
| Additive | 1.35 | 0.016 | 1.15 | 0.318 | 1.19 | 0.176 | 1.34 | 0.040 |
| rs4430796 Allele | 1.33 | 0.027 | 0.99 | 0.943 | 1.07 | 0.600 | 1.28 | 0.079 |
| Homozygote | 2.09 | 0.021 | 0.94 | 0.848 | 1.37 | 0.336 | 2.10 | 0.034 |
| Heterozygote | 1.23 | 0.252 | 1.02 | 0.914 | 1.09 | 0.655 | 1.22 | 0.313 |
| Dominant | 1.34 | 0.087 | 1.01 | 0.977 | 1.14 | 0.484 | 1.34 | 0.124 |
| Recessive | 1.89 | 0.038 | 0.93 | 0.819 | 1.31 | 0.382 | 1.90 | 0.056 |
| Additive | 1.35 | 0.024 | 0.99 | 0.936 | 1.14 | 0.355 | 1.35 | 0.041 |
| rs11651052 Allele | 1.47 | 0.003 | 1.03 | 0.840 | 1.16 | 0.274 | 1.35 | 0.029 |
| Homozygote | 2.47 | 0.007 | 0.93 | 0.823 | 1.29 | 0.452 | 2.32 | 0.019 |
| Heterozygote | 1.43 | 0.047 | 1.14 | 0.522 | 1.25 | 0.244 | 1.58 | 0.106 |
| Dominant | 1.55 | 0.010 | 1.10 | 0.627 | 1.26 | 0.211 | 1.50 | 0.035 |
| Recessive | 2.07 | 0.023 | 0.87 | 0.684 | 1.17 | 0.639 | 1.97 | 0.049 |
| Additive | 1.50 | 0.003 | 1.03 | 0.844 | 1.18 | 0.239 | 1.46 | 0.012 |

Abbreviations: 95% CI, 95% confidence interval; OR, odds ratio; SNP, single-nucleotide polymorphism; T2DM, Type 2 diabetes mellitus. P values were calculated by logistic regression analysis with adjustments for age and gender. Bold indicates that P < 0.05 means the data are statistically significant.
Table 6 Relationships between *IGF2BP2*, *HMG20A*, and *HNF1B* polymorphisms and T2DM risk according to the stratification by BMI, smoking, and drinking

| SNP ID  | Model          | OR (95%CI) | P     | OR (95%CI) | P     | OR (95%CI) | P     | OR (95%CI) | P     | OR (95%CI) | P     |
|---------|----------------|------------|-------|------------|-------|------------|-------|------------|-------|------------|-------|
| rs7178572 | Allele        | 1.54       | 0.004 | 0.99       | 0.966 | 1.55       | 0.017 | 1.04       | 0.795 | 1.40       | 0.134 |
|         | Homozygote    | 2.71       | 0.027 | 0.92       | 0.839 | 3.32       | 0.034 | 1.12       | 0.746 | 3.37       | 0.041 |
|         | Heterozygote  | 1.09       | 0.999 | 1.41       | 0.824 | 1.07       | 0.869 | 1.22       | 0.396 | 0.71       | 0.406 |
|         | Dominant      | 1.20       | 0.499 | 1.29       | 0.348 | 1.38       | 0.387 | 1.20       | 0.418 | 1.03       | 0.948 |
|         | Recessive     | 2.71       | 0.016 | 0.76       | 0.473 | 3.19       | 0.024 | 1.00       | 1.000 | 4.12       | 0.099 |
|         | Additive      | 1.42       | 0.072 | 1.07       | 0.747 | 1.62       | 0.063 | 1.10       | 0.557 | 1.51       | 0.137 |
| rs4430796 | Allele        | 1.34       | 0.067 | 1.27       | 0.188 | 1.41       | 0.082 | 1.33       | 0.062 | 1.45       | 0.117 |
|         | Homozygote    | 1.76       | 0.221 | 4.23       | 0.034 | 4.46       | 0.076 | 2.05       | 0.093 | 7.96       | 0.025 |
|         | Heterozygote  | 1.34       | 0.274 | 1.19       | 0.518 | 0.88       | 0.720 | 1.35       | 0.179 | 0.70       | 0.368 |
|         | Dominant      | 1.41       | 0.178 | 1.35       | 0.249 | 1.07       | 0.842 | 1.44       | 0.090 | 0.97       | 0.940 |
|         | Recessive     | 1.53       | 0.335 | 3.88       | 0.042 | 4.73       | 0.060 | 1.78       | 0.165 | 9.39       | 0.013 |
|         | Additive      | 1.33       | 0.144 | 1.49       | 0.072 | 1.32       | 0.316 | 1.39       | 0.053 | 1.41       | 0.253 |
| rs11651052 | Allele       | 1.51       | 0.010 | 1.27       | 0.174 | 1.51       | 0.039 | 1.40       | 0.026 | 1.56       | 0.058 |
|          | Homozygote    | 2.44       | 0.008 | 3.41       | 0.049 | 5.01       | 0.056 | 2.35       | 0.044 | 7.65       | 0.019 |
|          | Heterozygote  | 1.54       | 0.109 | 1.44       | 0.183 | 1.14       | 0.711 | 1.48       | 0.084 | 1.04       | 0.049 |
|          | Dominant      | 1.68       | 0.043 | 1.57       | 0.087 | 1.35       | 0.376 | 1.59       | 0.032 | 1.35       | 0.426 |
|          | Recessive     | 1.99       | 0.128 | 2.83       | 0.087 | 4.7        | 0.060 | 1.95       | 0.104 | 7.49       | 0.017 |
|          | Additive      | 1.55       | 0.026 | 1.60       | 0.035 | 1.54       | 0.122 | 1.51       | 0.017 | 1.72       | 0.075 |

Abbreviations: 95%CI, 95% confidence interval; BMI, body mass index; OR, odds ratio; SNP, single nucleotide polymorphism; T2DM, Type 2 diabetes mellitus. 
P values were calculated by logistic regression analysis with adjustments for age and gender. Bold indicates that P < 0.05 means the data are statistically significant.
Figure 1. Summary of MDR SNP–SNP interaction among IGF2BP2, HMG20A, and HNF1B gene

Dark-shaded cells represent higher risk combinations compared with light-shaded cells. Each cell shows counts of “case” on left and “control” on right.

variant, BMI, smoking, and alcohol drinking status had interactive effect with selected SNPs on T2DM risk. An association of rs11651052 and T2DM risk was observed in both subjects with BMI > 24 kg/m² (allele, dominant, and additive) and subjects BMI ≤ 24 kg/m² (homozygote and additive). In non-smokers, a trend of higher risk of developing T2DM was also found for subjects with A allele, and AA, AA-AG genotypes, and in additive model. Similarly, rs11651052-A allele had a higher the incidence of T2DM in smokers. In drinkers, individuals with rs11651052 AA genotype had 7.65- and 7.49-fold increased risk of developing T2DM than drinkers who carried GG genotype and combined AG-GG, respectively. In non-drinkers, rs11651052 was also associated with T2DM occurrence (allele, dominant, and additive).

The association between higher order interactions of SNP–SNP and T2DM risk was analyzed by MDR as summarized in Figure 1. The interaction analysis revealed moderate effect between the markers HMG20A rs7178572, HNF1B rs11651052, and IGF2BP2 rs7640539, which were conferring risk toward T2DM progression. The accumulated effect of rs7178572-GG, rs11651052-AA, and rs7640539-TA conferred a higher risk for T2DM, as shown in Table 7.

The relation between selected SNPs and different clinical parameters of T2DM patients was investigated, as illustrated in Table 8. We found the significant relationship of IGF2BP2 rs11927381 and rs7640539 with the levels of retinol-binding protein ($P = 0.010$ and $P = 0.028$, respectively). The association of HMG20A rs7178572 with glycated hemoglobin was identified ($P = 0.032$). Besides, carriers of HNF1B rs11651052 GG genotype had significantly higher insulin level than AA and AG genotypes ($P = 0.018$). However, there was no relation of different genotypes with the remaining clinical parameters ($P > 0.05$).
Table 7 SNP–SNP interaction models of the IGF2BP2, HMG20A, and HNF1B genes analyzed by the MDR method

| Model | Training Bal. Acc. | Testing Bal. Acc. | CVC | OR (95%CI) | P    |
|-------|--------------------|-------------------|-----|------------|------|
| HMG20A rs7178572 | 0.539              | 0.531             | 9/10 | 1.46 (1.12–1.91) | 0.0058 |
| HMG20A rs7178572, HNF1B rs11651052 | 0.550              | 0.510             | 7/10 | 1.63 (1.25–2.15) | 0.0004 |
| HMG20A rs7178572, HNF1B rs11651052 and IGF2BP2 rs7640539 | 0.574              | 0.524             | 6/10 | 1.89 (1.45–2.48) | <0.0001 |

Bal. Acc., balanced accuracy; CI, confidence interval; CVC, cross–validation consistency; MDR, multifactor dimensionality reduction; OR, odds ratio. P values were calculated using \( \chi^2 \) tests. Bold indicates that \( P < 0.05 \) means the data are statistically significant.

Table 8 Comparisons of clinical characteristics among T2DM patients with different genotypes of SNPs in IGF2BP2, HMG20A, and HNF1B

| Characteristics | rs11927381 | rs7640539 | P    |
|----------------|-----------|-----------|------|
| Total cholesterol | 4.53 ± 1.36 | 4.49 ± 1.31 | 4.72 ± 1.05 | 0.650 |
| HDL-C | 1.19 ± 0.58 | 1.18 ± 0.50 | 1.34 ± 0.87 | 0.314 |
| LDL-C | 2.70 ± 1.14 | 2.55 ± 0.96 | 2.50 ± 0.79 | 0.306 |
| Urea | 6.35 ± 2.95 | 5.98 ± 1.91 | 6.77 ± 6.90 | 0.317 |
| Creatinine | 70.67 ± 36.71 | 68.25 ± 22.19 | 63.15 ± 18.29 | 0.336 |
| Glomerular filtration rate | 124.44 ± 36.99 | 121.03 ± 32.41 | 123.58 ± 37.93 | 0.737 |
| Fasting blood glucose | 9.51 ± 3.59 | 10.51 ± 5.92 | 9.80 ± 4.06 | 0.196 |
| Glycated hemoglobin | 9.16 ± 2.08 | 9.39 ± 2.90 | 9.49 ± 2.45 | 0.672 |
| Triglyceride | 2.80 ± 2.47 | 2.35 ± 1.86 | 2.61 ± 2.79 | 0.646 |
| Cystatin C | 0.88 ± 0.54 | 0.81 ± 0.19 | 0.80 ± 0.33 | 0.340 |
| C-reactive protein | 1.40 ± 0.97 | 1.38 ± 2.19 | 1.11 ± 0.85 | 0.708 |
| Insulin | 19.83 ± 21.28 | 17.30 ± 16.05 | 18.21 ± 10.35 | 0.553 |
| 25-Hydroxy-vitamin D | 25.77 ± 16.79 | 23.25 ± 14.28 | 24.74 ± 9.11 | 0.521 |
| UCRP | 0.46 ± 1.17 | 0.67 ± 1.49 | 0.43 ± 0.56 | 0.353 |
| Retinol-binding protein | 40.65 ± 11.13 | 36.01 ± 10.60 | 38.12 ± 9.47 | 0.010 |

| Characteristics | rs7178572 | rs11651052 | P    |
|----------------|----------|-----------|------|
| Total cholesterol | 4.48 ± 1.18 | 4.59 ± 1.46 | 4.46 ± 1.10 | 0.660 |
| HDL-C | 1.16 ± 0.51 | 1.21 ± 0.60 | 1.26 ± 0.63 | 0.501 |
| LDL-C | 2.54 ± 0.88 | 2.66 ± 1.07 | 2.65 ± 1.24 | 0.611 |
| Urea | 6.48 ± 4.15 | 6.28 ± 2.84 | 5.66 ± 1.63 | 0.237 |
| Creatinine | 67.87 ± 35.06 | 69.64 ± 27.23 | 69.15 ± 23.39 | 0.864 |
| Glomerular filtration rate | 124.6 ± 41.72 | 123.06 ± 31.73 | 119.77 ± 32.46 | 0.754 |
| Fasting blood glucose | 10.89 ± 5.79 | 9.52 ± 4.29 | 9.45 ± 3.07 | 0.051 |
| Glycated hemoglobin | 9.84 ± 3.26 | 9.05 ± 2.01 | 9.01 ± 1.82 | 0.032 |
| Triglyceride | 2.30 ± 1.70 | 2.64 ± 2.51 | 2.40 ± 2.37 | 0.470 |
| Cystatin C | 0.81 ± 0.23 | 0.88 ± 0.55 | 0.83 ± 0.17 | 0.427 |
| C-reactive protein | 1.17 ± 0.70 | 1.50 ± 2.03 | 1.41 ± 1.09 | 0.291 |
| Insulin | 20.09 ± 23.45 | 17.80 ± 16.19 | 19.30 ± 14.80 | 0.643 |
| 25-Hydroxy-vitamin D | 25.17 ± 20.70 | 24.61 ± 12.56 | 23.87 ± 8.19 | 0.915 |
| UCRP | 0.74 ± 1.73 | 0.42 ± 0.95 | 0.55 ± 1.15 | 0.162 |
| Retinol-binding protein | 38.49 ± 10.21 | 38.99 ± 11.60 | 38.54 ± 11.77 | 0.948 |

Abbreviations: HDL-C, high-density lipoprotein cholesterol; LDL-C, low-density lipoprotein cholesterol; SNP, single-nucleotide polymorphism; T2DM, Type 2 diabetes mellitus; UCRP, ubiquitin cross-reactive protein. P values were calculated by using one-way analysis of variance (ANOVA) test. Bold indicates that \( P < 0.05 \) means the data are statistically significant.

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Discussion

We performed a case–control study to investigate the association of \textit{IGF2BP2} rs11927381 and rs7640539, \textit{HMG20A} rs7178572, \textit{HNF1B} rs4430796, and rs11651052 with T2DM susceptibility. We found that \textit{HMG20A} rs7178572 and \textit{HNF1B} rs11651052 were related to an increased T2DM risk in the overall. Given that T2DM represents a complex disorder influenced by the interplay between genetic and behavioral factors, we analyzed the effect of age, gender, BMI, smoking, and alcohol drinking on the relationship of these variants with T2DM susceptibility. Our stratified analysis showed that rs7178572, rs4430796, and rs11651052 had higher risk of T2DM occurrence in males and subjects with age \( \leq 59 \) years. In addition, rs11927381 also contributed T2DM susceptibility at age \( \leq 59 \) years. We also found that these genetic variants might increase the risk of T2DM occurrence not only by itself but also by interacting with smoking and alcohol drinking. Besides, rs7178572-AA \((P = 0.032)\) genotype and rs11651052 GG \((P = 0.018)\) genotype might have higher glycated hemoglobin and insulin levels, respectively. To the best of our knowledge, this was the first to explore the effects of the relationships between \textit{IGF2BP2} rs11927381, rs7640539 and \textit{HNF1B} rs11651052 and T2DM susceptibility in the Chinese Han population.

Insulin-like growth factor 2 binding protein 2 (\textit{IGF2BP2}), located on chromosome 3q27, encodes an mRNA-binding protein associated with RNA location, stability, and translation. \textit{IGF2BP2}, highly expressed in pancreatic islets, is involved in \( \beta \)-cell function by regulating IGF2 post-translational modification [20]. IGF2 is a member of the insulin family of polypeptide growth factors, which play an important role in the development, growth, and stimulation of insulin action [21]. \textit{IGF2BP2} variations were also associated with decreased insulin secretion and hyperglycemia [22]. Several variants in \textit{IGF2BP2} were investigated for the relationship with T2DM; however, there were very few studies on rs11927381 and rs7640539. Only one study reported the association between \textit{IGF2BP2} rs11927381 and the increased T2DM risk among Slavonic population [23]. There was no report on rs7640539 polymorphism. Our results displayed rs11927381 variant had a higher risk of developing T2DM in subjects with age \( \leq 59 \) years, suggesting the association appear to be age dependent. However, the current study did not find a significant relationship between rs7640539 and T2DM susceptibility. Further studies are required to elucidate the association.

High mobility group 20 A (\textit{HMG20A}) gene, located in 15q24.3, is a member of high mobility group (HMG) box-containing genes. \textit{HMG20A} encodes a widely expressed non-histone chromosomal protein controlling gene expression by histone modification [24].

\textit{HMG20A} expression in islet is essential for metabolism-insulin secretion coupling via the coordinated regulation of key islet-enriched genes, and the depletion \textit{HMG20A} protein induces expression of genes implicated in \( \beta \) cell de-differentiation [8]. Previously, \textit{HMG20A} (rs7178572) showed an association with T2DM in European obese subjects [25]. Our results found that \( G \) allele of rs7178572, intronic SNPs within the \textit{HMG20A}, which was related to an increased T2DM susceptibility. However, a previous study showed there was no significant relationship between rs7178572 and the risk of T2DM among Han population in southern China [26], such inconsistencies in these reports might result from a different behavioral habit or sample size. As we known, genetic, environmental, behavioral, and metabolic risk factors are contributed to the development of T2DM [27]. Obesity (defined by BMI), smoking and alcohol drinking (especially heavy alcohol consumption) are known risk factors for T2DM [28–30]. Smoking increased 1.35-fold the risk of T2DM compared with non-smokers [29]. Alcohol consumption is related to glycemic control and insulin resistance [30]. Therefore, we evaluated the effects of age, gender, BMI, smoking, and alcohol consumption on the association of rs7178572 with T2DM risk. Interestingly, rs7178572 variant had a higher susceptibility to T2DM in males, smokers, drinkers, and the subjects with BMI \( > 24 \) kg/m\(^2\). These results are required to validate in larger populations.

Hepatocyte nuclear factor-1\(\beta\) (\textit{HNF1B}), located on chromosome 17q21.3, encodes a transcription factor that involved in tissue-specific regulation of gene expression and embryonic development of numerous organs [31]. \textit{HNF1B} gene played the important role in the primary pathophysiology of diabetes. It was involved in the loss of neurogenin-3 (Ngn3)-positive endocrine progenitor cells, pancreatic atrophy, and a reduced insulin sensitivity to endogenous glucose production leading to the reduction of insulin secretion [32]. Previous studies have reported that genetic variations in \textit{HNF1B} were associated with the susceptibility of T2DM. Rs4430796 \((A > G)\) in intron 2 of \textit{HNF1B} is the most frequent SNP in Chinese population. Notably, the mutant allele frequency for rs4430796 is quite different between different ethnic groups. The mutant allele \( G \) frequency in the study was 0.289, similar to the healthy Han Chinese and Asian, but significantly different from Caucasian \((0.47)\) and African \((0.67)\) [33]. Previous studies revealed the risk \( G \) allele of rs4430796 was significantly related to T2DM in a southern Chinese Han population [34], which was consistent with our results. Here, we found that rs4430796 increased the risk of T2DM occurrence, especially in males and subgroup with age \( \leq 59 \) years. Besides, the association also was observed in the subgroup with BMI \( \leq 24 \) kg/m\(^2\) and drinkers. These results indicated that gene-behavioral habit interactions might operate in the pathogenesis of
T2DM. Rs11651052 (G > A) is another SNP in HNF1B, and no study has analyzed the SNP now. In our study, we first reported that rs11651052-A allele increased 1.26-fold risk of T2DM compared with G allele. Our stratified analysis showed that rs11651052 had a higher T2DM susceptibility in males and subjects with age ≤ 59 years, suggesting the risk association of this polymorphism might be age dependent.

Inevitably, several intrinsic limitations should be considered. First, the subjects were enrolled from the identical hospitals; therefore, the selection bias could not be completely excluded. Second, some clinical characteristics were not analyzed because of missing or uncollected data in controls. Third, explicit mechanisms of these polymorphisms on the development of T2DM are still bewildered and further research is required. Therefore, further well-designed large and prospective studies and functional experiments should be conducted to verify our finding.

**Conclusion**

To sum up, our study revealed that variants in IGF2BP2, HMG20A, and HNF1B had the risk effect on T2DM occurrence among the Chinese Han population. Specifically, we first found that rs11927381, rs7640539, and rs11651052 were associated with the increased risk of T2DM occurrence. We also provided evidence that age gender, BMI, smoking, and alcohol drinking status had interactive effect with these variants on T2DM susceptibility, suggesting that gene-behavioral habit interactions might play critical roles in the risk of developing T2DM. Our study may increase the understanding of IGF2BP2, HMG20A, and HNF1B variants on the pathogenesis of T2DM.

**Competing Interests**
The authors declare that there are no competing interests associated with the manuscript.

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**Author Contribution**
The work presented here was carried out in collaboration between all authors. Ting Huang and Li Wang carried out the molecular genetic studies and drafted the manuscript. Mei Bai and Jianwen Zheng designed the methods and experiments, performed the statistical analyses and interpreted the results. Dongya Yuan, Yongjun He and Yuhe Wang designed primers and performed the SNP genotyping experiments. Tianbo Jin and Wei Cui conceived of the study, worked on associated data collection and their interpretation, participated in the design and coordination of the study, and funded the study. All authors read and approved the final manuscript.

**Ethics Approval**
This study was approved by the ethics committee of the First Affiliated Hospital of Xi’an Jiaotong University and was conducted in accordance with the declaration of Helsinki.

**Informed Consent**
All subjects signed an informed consent before enrolment in the study.

**Abbreviations**
CI, confidence interval; CVC, cross-validation consistency; HDL-C, high-density lipoprotein cholesterol; HWE, Hardy–Weinberg equilibrium; IGF2, insulin-like growth factor 2; LDL-C, low-density lipoprotein cholesterol; MDR, multifactor dimensionality reduction; OR, odds ratio; SNP, single-nucleotide polymorphism; T2DM, Type 2 diabetes mellitus.

**References**
1 Nolan, C.J., Damm, P. and Prentki, M. (2011) Type 2 diabetes across generations: from pathophysiology to prevention and management. *Lancet* (London, England) 378, 169–181, [https://doi.org/10.1016/S0140-6736(11)60614-4](https://doi.org/10.1016/S0140-6736(11)60614-4)
2 Zheng, Y., Ley, S.H. and Hu, F.B. (2018) Global aetiology and epidemiology of type 2 diabetes mellitus and its complications. *Nat. Rev. Endocrinol.* 14, 88–98, [https://doi.org/10.1038/nrendo.2017.151](https://doi.org/10.1038/nrendo.2017.151)
3 Guariguata, L., Whiting, D.R., Hambleton, I., Beagley, J., Linnenkamp, U. and Shaw, J.E. (2014) Global estimates of diabetes prevalence for 2013 and projections for 2035. *Diabetes Res. Clin. Pract.* 103, 137–149, [https://doi.org/10.1016/j.diabres.2013.11.002](https://doi.org/10.1016/j.diabres.2013.11.002)
4 Wu, Y., Ding, Y., Tanaka, Y. and Zhang, W. (2014) Risk factors contributing to type 2 diabetes and recent advances in the treatment and prevention. *Int. J. Med. Sci.* 11, 1185–1200, [https://doi.org/10.7150/ijms.10001](https://doi.org/10.7150/ijms.10001)
5 Scott, R.A., Scott, L.J., Magi, R., Marullo, L., Gaulton, K.J., Kaakinen, M. et al. (2017) An Expanded Genome-Wide Association Study of Type 2 Diabetes in Europeans. Diabetes 66, 2888–2902, https://doi.org/10.2337/db16-1253
6 Zhao, W., Rasheed, A., Tikkanen, E., Lee, J.J., Butterworth, A.S., Howson, J.M.M. et al. (2017) Identification of new susceptibility loci for type 2 diabetes and shared etiological pathways with coronary heart disease. Nat Genet 49, 1450–1457, https://doi.org/10.1038/ng.3943
7 Zeggini, E., Weedon, M.N., Lindgren, C.M., Frayling, T.M., Elliott, K.S., Lango, H. et al. (2007) Replication of genome-wide association signals in UK samples reveals risk loci for type 2 diabetes. Science 316, 1336–1341, https://doi.org/10.1126/science.1142364
8 Mellado-Gil, J.M., Fuente-Martin, E., Lorenzo, P.I., Cobo-Vuillemier, N., Lopez-Noriega, L., Martin-Montalvo, A. et al. (2018) The type 2 diabetes-associated HMG20A gene is mandatory for islet beta cell functional maturity. Cell Death Dis 9, 279, https://doi.org/10.1038/s41419-018-0272-z
9 Poll, A.V., Pierreux, C.E., Lokmane, L., Haumaitre, C., Achoury, Y., Jacquemin, P. et al. (2006) A vHNF1/TCF2-HNF6 cascade regulates the transcription factor network that controls generation of pancreatic precursor cells. Diabetes 55, 61–69, https://doi.org/10.2337/diabetes.55.01.06.db05-0681
10 Shabana, X.X., Shahid, S.U. and Hasnain, S. (2018) Identification of genetic basis of obesity and mechanistic link of genes and lipids in Pakistani populations. Biosci. Rep. 38, 1001820281, https://doi.org/10.1042/BSR20180281
11 Bayesel, S., Eyerci, N., Pinarli, F.A., Kizilgul, M., Ozcelik, O., Caliskan, M. et al. (2019) HNF1A gene p.I27L is associated with early-onset, maturity-onset diabetes of the young-like diabetes in Turkey. BMC Endocrine Disorders 19, 51, https://doi.org/10.1186/s12902-019-0375-2
12 Votsi, C., Toufekis, C., Michailidou, K., Antoniades, A., Skordis, N., Karaolis, M. et al. (2017) Type 2 Diabetes Susceptibility in the Greek-Cypriot Population: Replication of Associations with TCF7L2, FTO, HHEX, SLC30A8 and IGF2BP2 Polymorphisms. Genes 8, 16, https://doi.org/10.3390/genes8010016
13 Via, M.A. and Mechanick, J.I. (2016) Nutrition in Type 2 Diabetes and the Metabolic Syndrome. Med. Clin. North Am. 100, 1285–1302, https://doi.org/10.1016/j.mcna.2016.06.009
14 Alberti, K.G. and Zimet, P.Z. (1998) Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. Diabetic Med.: J. Br. Diabetic Assoc. 15, 539–553, https://doi.org/10.1002/(SICI)1096-9136(19980715)15:7<539::AID-DIA668>3.0.CO;2-S
15 Bai, M., Wang, R., Zhu, L., Li, G., Yuan, D., Wang, L. et al. (2018) Age-related differences in limb fat-free mass and fat mass in healthy Chinese Adults. Sci Rep 8, 8013, https://doi.org/10.1038/s41598-018-25447-z
16 Dai, Z.J., Liu, X.H., Ma, Y.F., Kang, H.F., Jin, T.B., Dai, Z.M. et al. (2016) Association Between Single Nucleotide Polymorphisms in DNA Polymerase Kappa Gene and Breast Cancer Risk in Chinese Han Population: A STROBE-Compliant Observational Study. Medicine (Baltimore) 95, e2466, https://doi.org/10.1097/MD.0000000000002466
17 Jiri, M., Zhang, L., Lan, B., He, N., Feng, T., Liu, X. et al. (2016) Genetic variation in the ABCG2 gene is associated with gut risk in the Chinese Han population. Clin. Rheumatol. 35, 159–163, https://doi.org/10.1007/s10067-016-3105-9
18 Du, J., Jin, T., Cao, Y., Chen, J., Guo, Y., Sun, M. et al. (2016) Association between genetic polymorphisms of MMP8 and the risk of steroid-induced osteonecrosis of the femoral head in the population of northern China. Medicine (Baltimore) 95, 64794, https://doi.org/10.1097/MD.0000000000004794
19 Leem, S. and Park, T. (2017) An empirical fuzzy multifactor dimensionality reduction method for detecting gene-gene interactions. BMC Genomics 18, 115
20 Christiansen, J., Kolte, A.M., Hansen, T. and Nielsen, F.C. (2009) IGF2 mRNA-binding protein 2: biological function and putative role in type 2 diabetes. J. Mol. Endocrinol. 43, 187–195, https://doi.org/10.1677/JME-09-0016
21 Parikh, H., Lyssenko, V. and Group, I.C. (2009) Prioritizing genes for follow-up from genome wide association studies using information on gene expression in tissues relevant for type 2 diabetes. BMC Med. Genet. 2, 72, https://doi.org/10.1186/1755-7900-2-72
22 Groenewoud, M.J., Dekker, J.M., Fritsche, A., Reiling, E., Nijpels, G., Heine, R.J. et al. (2008) Variants of CDKAL1 and IGF2BP2 affect first-phase insulin secretion during hyperglycaemic clamps. Diabetologia 51, 1659–1663, https://doi.org/10.1007/s00125-008-1083-z
23 Azarova, I.E., Klyosova, E.Y., Lazarenko, V.A., Konoply, I.A. and Polonikov, A.V. (2020) Associations Genome-wide association studies-derived susceptibility loci in type 2 diabetes: confirmation in a Chinese population. Clin. Invest. Med. 35, E327
24 Fletcher, B., Gulancik, M. and Lamendola, C. (2002) Risk factors for type 2 diabetes mellitus. J. Cardiovasc. Nurs. 16, 17–23, https://doi.org/10.1097/00005082-200201000-00003
25 Ganz, M.L., Wintfeld, N., Li, Q., Alas, V., Langer, J. and Hammer, M. (2014) The association of body mass index with the risk of type 2 diabetes: a case-control study nested in an electronic health records system in the United States. Diabetol. Metab. Syndrome 6, 50
26 Maddatu, J., Anderson-Baucum, E. and Evans-Molina, C. (2017) Smoking and the risk of type 2 diabetes. Transl. Res.: J. Lab. Clin. Med. 184, 101–107, https://doi.org/10.1016/j.trsl.2017.02.004
27 Steiner, J.L., Cowell, K.T. and Lang, C.H. (2015) Impact of Alcohol on Glycemic Control and Insulin Action. Biomolecules 5, 2223–2246, https://doi.org/10.3390/biom5042223
28 Tronche, F. and Yaniv, M. (1992) HNF1, a homeoprotein member of the hepatic transcription regulatory network. BioEssays: News Rev. Mol. Cell. Develop. Biol. 14, 579–587, https://doi.org/10.1002/bies.950140902
32 El-Khairi, R. and Vallier, L. (2016) The role of hepatocyte nuclear factor 1beta in disease and development. *Diabetes Obes. Metab.* **18**, 23–32
33 Winckler, W., Weedon, M.N., Graham, R.R., McCarroll, S.A., Purcell, S., Almgren, P. et al. (2007) Evaluation of common variants in the six known maturity-onset diabetes of the young (MODY) genes for association with type 2 diabetes. *Diabetes* **56**, 685–693, https://doi.org/10.2337/db06-0202
34 Wang, C., Hu, C., Zhang, R., Bao, Y., Ma, X., Lu, J. et al. (2009) Common variants of hepatocyte nuclear factor 1beta are associated with type 2 diabetes in a Chinese population. *Diabetes* **58**, 1023–1027, https://doi.org/10.2337/db08-1064