Sex Differences in Cancer: Epidemiology, Genetics and Therapy

Hae-In Kim¹, Hyesol Lim¹ and Aree Moon*  
Duksung Innovative Drug Center, College of Pharmacy, Duksung Women’s University, Seoul 01369, Republic of Korea

Abstract

The incidence and mortality of various cancers are associated with sex-specific disparities. Sex differences in cancer epidemiology are one of the most significant findings. Men are more prone to die from cancer, particularly hematological malignancies. Sex difference in cancer incidence is attributed to regulation at the genetic/molecular level and sex hormones such as estrogen. At the genetic/molecular level, gene polymorphism and altered enzymes involving drug metabolism generate differences in cancer incidence between men and women. Sex hormones modulate gene expression in various cancers. Genetic or hormonal differences between men and women determine the effect of chemotherapy. Until today, animal studies and clinical trials investigating chemotherapy showed sex imbalance. Chemotherapy has been used without consideration of sex differences, resulting in disparity of efficacy and toxicity between sexes. Based on accumulating evidence supporting sex differences in chemotherapy, all clinical trials in cancer must incorporate sex differences for a better understanding of biological differences between men and women. In the present review, we summarized the sex differences in (1) incidence and mortality of cancer, (2) genetic and molecular basis of cancer, (3) sex hormones in cancer incidence, and (4) efficacy and toxicity of chemotherapy. This review provides useful information for sex-based chemotherapy and development of personalized therapeutic strategies against cancer.

Key Words: Sex Difference, Cancer, Sex hormone, Chemotherapy

INTRODUCTION

Cancer represents a leading cause of death worldwide (Naghavi et al., 2016). Sex plays a crucial role in the incidence, disease prognosis and mortality in a variety of cancers (Siegel et al., 2016). The incidence of cancer was about 20% higher in men than in women and the mortality rate was 40% higher in men in the United States from 2009 to 2013 (Siegel et al., 2017). Sex differences influence cancer susceptibility at the genetic/molecular levels. Sex hormones also negatively or positively affect the development of various cancers. Biological specificities determine the outcome and response to therapy of cancer.

During the last decades, animal studies and clinical trials used males alone and excluded females (Keitt et al., 2004; Becker et al., 2005; Zucker and Beery, 2010). Altered sex hormones in the menstrual cycle affect the experimental results (Becker et al., 2005). In addition, the 1977 United States Food and Drug Administration guideline excluded women in clinical research due to the risk of birth defects (Merkatz et al., 1993). Similar doses were administered to men and women in clinical chemotherapy (Islam et al., 2017). Accumulating evidence shows sex-specific differences in toxicity and efficacy of chemotherapy (Tran et al., 1998; Rademaker, 2001; Anderson, 2005; Bren, 2005, Schmetzer and Flörcken, 2012). Especially, women experience higher adverse drug reactions to most anticancer drugs than men (Wang and Huang, 2007).

This review summarized studies involving sex differences in epidemiology, sex hormones, and genetic/molecular factors. In addition, we reviewed sex-related differences in chemotherapy using anticancer agents such as 5-fluorouracil (FU), paclitaxel, doxorubicin, cisplatin, bevacizumab and rituximab. This article provides important clues and insights for the precise understanding of sex-specific differences in cancer.

SEX DIFFERENCES IN INCIDENCE AND MORTALITY OF CANCER

Growing evidence shows sex-specific differences in the incidence and mortality associated with various cancers. Prostate, lung, and colorectal cancer occur the most in males,
while breast, lung, and colorectal cancer are predominant in females in the United States (Siegel et al., 2016). In addition to the incidence of cancer in sex-specific organs such as prostate and ovary, sex differences in cancers such as colon, lung, and liver have been reported (Dorak and Karpuzoglu, 2012; Torre et al., 2016). Thyroid cancer incidence is much higher in females than in males (Dorak and Karpuzoglu, 2012). Cancer incidence involving colorectal, stomach and liver cancer is higher in males than in females (Arnold et al., 2017). Furthermore, bladder cancer and leukemia have been predominantly diagnosed in males than in females (Fitzmaurice et al., 2017). In patients with colorectal cancer, women developed right-side ed malignancy while men manifested the disease more on the left side (Kim et al., 2015). Right-sided colon cancer is associated with a higher severity of cancer compared with left-sided disease (Kim et al., 2015). The cause of disparity in location might be due to differences in estrogen level between men and women.

The mortality of cancer is reported to be greater in men than in women (Siegel et al., 2016). Especially, lung, colorectal and stomach cancers, which are the leading causes of cancer deaths, show higher mortality in men than in women (Siegel et al., 2016; Fitzmaurice et al., 2017). Female cancers such as breast, ovarian and uterine corpus cancer result in relatively high mortality (Siegel et al., 2016; Fitzmaurice et al., 2017). Men-specific cancers such as prostate cancer also represent prominent causes for cancer death (Siegel et al., 2016). Mortality associated with esophagus, liver, and bladder cancer is higher in men than in women (Siegel et al., 2016). Men had a 34% higher risk of death due to melanoma compared with women (Crocetti et al., 2015). Therefore, mortality from various cancer types shows sex disparity.

Lung cancer was the leading cause of cancer death in men in 20th century due to smoking (Siegel et al., 2016). Lung cancer death was decreased by reduced smoking rates and early detection and treatment from 1991 to 2012 (Siegel et al., 2016). In both sexes, mortality due to gastric cancer declined rapidly in the 1930s in U.S (Siegel et al., 2016). Although the cause of dramatic reduction is not completely understood, the control of Helicobacter pylori infection, and better methods of food preservation resulted in a reduction of mortality due to stomach cancer (Bertucchio et al., 2009; Siegel et al., 2016).

SEX DIFFERENCES IN GENETIC AND MOLECULAR BASIS OF CANCER

In genetic and molecular studies, susceptibility to disease varies across the sexes. Genetic and molecular disparities between males and females contribute to differences in the incidence of a variety of cancers. Men show a higher incidence of bladder cancer than women (Siegel et al., 2016). The lower incidence of bladder cancer in women was correlated with the sulfotransferase 1A1 (SULT1A1) Histidine (His) genotype (Zheng et al., 2003). Genetic polymorphism of SULT1A1 showed alteration from Arginine to His, which was triggered by A-to-G transition (Rafatogianis et al., 1997). It was reported that the His213 allele genotype SULT1A1 significantly decreased the risk of bladder cancer exclusively in women (Zheng et al., 2003). Therefore, this gene might be associated with a protective role in women diagnosed with bladder cancer (Dorak and Karpuzoglu, 2012).

Genetic polymorphism, which is linked to drug metabolizing enzymes, influences the risk of carcinogenesis (Boluffer et al., 2007). For example, acute leukemia has a higher rate of incidence in men than in women (Boluffer et al., 2007). It was reported that men with deletion of glutathione s-methyltransferase T1 (GSTT1), which is a glutathione s-methyltransferase polymorphism, underwent phase 2 metabolism and detected frequently in acute lymphoblastic leukemia (ALL) than men with normal GSTT1 (Mannervik and Danielson, 1988; Hayes and Strange, 2000; Boluffer et al., 2007). Deletion of GSTT1 gene abrogated the enzyme activity (Arand et al., 1996).

NAD(P)H:quinone oxidoreductase 1 (NQO1) catalyzes free radical detoxification (Traver et al., 1992). NQO1 polymorphism with substitution of C to T base pair at position 609 of NQO1 decreases the activity of the enzyme (Asher et al., 2002; Fagerholm et al., 2008). NQO1 polymorphism showed a higher incidence of ALL only in males, but not in females (Boluffer et al., 2007).

Murine double minute 2 (MDM2) downregulates the expression of p53 protein, a tumor suppressor (Eliyahu et al., 1989; Bond et al., 2004). A single nucleotide polymorphism 309 (SNP 309) in MDM2 promoter increases the affinity of the transcriptional activator Sp1, which enhances the expression of MDM2 and consequently leads to attenuation of p53 pathway (Bond et al., 2004). Attenuation of p53 DNA damage response occurs in the presence of both wild-type allele of p53 and G-allele of SNP 309 (Bond et al., 2006). In addition, estrogen signaling affects the level of MDM2 (Bond et al., 2006). In the diffuse large B-cell lymphoma (DLBCL), sporadic soft-tissue sarcoma, and highly invasive estrogen receptor-positive ductal carcinoma, estrogen signaling pathway increases tumor formation directly or indirectly in women carrying the G-allele of SNP309 (Bond et al., 2006). Taken together, genetic and molecular differences might influence the disparity of the risk of cancer between men and women.

SEX DIFFERENCES IN SEX HORMONES IN CANCER INCIDENCE

Sex hormones might contribute to differences in the incidence of cancer between men and women (Do et al., 2010; Dorak and Karpuzoglu, 2012). ALL is more likely to occur in men because of limited estrogen level (Do et al., 2010). Estrogen plays a role in the inhibition of nuclear factor kappa B (NF-κB). NF-κB regulates the transcription of interferon regulatory factor 4 (IRF4) (Do et al., 2010). IRF4 is involved in the differentiation of B and T cells and is overexpressed in B-cell malignancies as a result of NF-κB hyperactivation (Do et al., 2010). IRF4 polymorphisms are related with the incidence of ALL (Do et al., 2010). Thus, a combination of intronic polymorphism of IRF4 and lack of estrogen might predispose men to leukemia.

Estrogen is linked closely to a higher rate of thyroid cancer development in women (Lee et al., 2005; Dorak and Karpuzoglu, 2012). Estrogen increases the proliferation of human thyroid papillary carcinoma cell line and promotes the expression of B-cell lymphoma-extra large (Bcl-XL), which is known as an anti-apoptotic protein (Hsu et al., 1997; Lee et al., 2005), compared with testosterone. Endogenous female sex hormone such as progesterin increase excretion of bile acid, which has been suggested as a potential inducer of colon cancer (McMi-
et al. (2016). Exogenous estrogen decreased the production of secondary bile acid which is responsible for promoting malignant change in colonic epithelium (McMichael and Potter, 1980; Everson et al., 1991; Grodstein et al., 1999). Therefore, female sex hormones may play a protective role in the development of colon cancer, by decreasing the level of bile acid (McMichael and Potter, 1980; Farhana et al., 2016).

**SEX DIFFERENCES IN EFFICACY AND TOXICITY OF CHEMOTHERAPY**

Sex-related differences at the genetic and molecular levels also affect the differences in the degree of drug response (Wang and Huang, 2007). Although sex disparities in the incidence and mortality of cancer have been observed for a variety of cancers, chemotherapy has been conducted independently of sex (Keitt et al., 2004; Becker et al., 2005; Zucker and Beery, 2010). Research involving animal model and clinical trials has been almost male-oriented. Accumulating evidence supports sex-related response to chemotherapeutic agents. Anticancer drugs, which represent sex differences in efficacy and toxicity, are summarized and listed in Table 1.

**Table 1. Anticancer drugs with sex differences in efficacy and toxicity**

| Drug          | Sex differences                                                                 | References                                                                 |
|---------------|---------------------------------------------------------------------------------|---------------------------------------------------------------------------|
| 5-fluorouracil| Clearance is higher in males than in females                                      | Milano et al., 1992                                                      |
|               | Females experienced higher toxicity (including stomatitis, leukopenia, alopecia and diarrhea) frequently and severely than males | Sloan et al., 2002                                                       |
| Paclitaxel    | Females showed lower elimination than males                                      | Joerger et al., 2006                                                     |
|               | Peripheral compartment of females is saturated at lower plasma concentrations levels compared with males | Joerger et al., 2006                                                     |
|               | Elimination is faster in males than in females                                   | Joerger et al., 2006                                                     |
| Doxorubicin   | Females experienced severe leukopenia greater than males                         | Yamamoto et al., 2008                                                    |
|               | Males have significantly higher clearance than females                           | Dobbs et al., 1995                                                      |
|               | Females experienced higher risk of early cardiotoxicity than males following treatment with doxorubicin in childhood leukemia | Lipshultz et al., 1995                                                   |
| Cisplatin     | Females experienced higher toxicities including vomiting and nausea than males   | Liaw et al., 2001                                                        |
|               | Male rats showed prolonged heat latency and slower motor nerve conduction than female rats | Wongtawatchai et al., 2009                                               |
|               | The half-maximal inhibitory concentration (IC50) of male cell lines was lower than that of females | Huang et al., 2007                                                      |
| Bevacizumab   | Clearance was higher in males than in females                                     | Lu et al., 2008                                                          |
|               | Female experienced more severe hypertension and neutropenia, and higher rate of abdominal pain than males | Brahmer et al., 2011                                                     |
| Rituximab     | Clearance was higher in males than in female                                     | Müller et al., 2012                                                     |
|               | Half-life of elimination in male was longer compared with females                | Müller et al., 2012                                                     |
|               | The better responses from treatment and outcomes were more prominent in females than in males | Riihijärvi et al., 2011                                                  |
|               | Male patients had a worse progression-free survival than female patients both in diffuse large B-cell lymphoma and follicular lymphoma | Riihijärvi et al., 2011                                                  |

5-FU

In the treatment of various cancers, 5-FU has been widely used as an effective chemotherapy (Longley et al., 2003). 5-FU as a pyrimidine antagonist inhibits thymidine synthase which is essential for DNA synthesis (Santi et al., 1974; Yoshioka et al., 1987; Longley et al., 2003). It has been observed that 5-FU might elicit different toxicities depending on sex (Stein et al., 1995; Sloan et al., 2002). Clearance of 5-FU is higher in male (179 l/h/m²) than in female (155 l/h/m²) (Milano et al., 1992). Low clearance of 5-FU in females can result in higher toxicity (Wang and Huang, 2007). Dihydropyrimidine dehydrogenase (DPD) breaks down 5-FU (Wasternack, 1980; Diasio and Harris, 1989; Longley et al., 2003). DPD plays a crucial role in toxicity associated with 5-FU chemotherapy (Harris et al., 1991). The activity of DPD was decreased in female and the difference in DPD activity between males and females is 15% (Etienne et al., 1994). A lower DPD activity associated with toxicity in women might be attributed to DPD deficiency syndrome (Milano et al., 1999). Reduced degradation of 5-FU in female influences therapeutic efficacy and toxicity (Mader, 2007). In 5-FU-based chemotherapy, women experimented higher toxicities, including stomatitis, leukopenia, alopecia, and diarrhea, more frequently and severely than men (Sloan et al., 2002).
**Paclitaxel**

Antitumor effects of paclitaxel, which inhibits depolymerization of cytoskeletal microtubules, interfere with cell division (Schiff and Horwitz, 1980; Jordan and Wilson, 2004). It was reported that female patients with solid tumors have 20% lower elimination of paclitaxel than male patients (Joerger et al., 2006). Peripheral compartments of female (0.83 mmol/l) are saturated at lower plasma concentrations compared with those of male (1.74 mmol/l) (Joerger et al., 2006). On the other hand, paclitaxel elimination is faster in male (so.5 h) than in female (51 h) (Joerger et al., 2006). In the treatment with paclitaxel combined with carboplatin, the number of female patients exhibiting toxicity such as severe leukopenia was greater than that of male patients (Yamamoto et al., 2008, Schmetzer and Flörcken, 2012). Conversely, female patients diagnosed with lung carcinoma (5.3 months) who were treated with paclitaxel combined with carboplatin showed longer median progression-free survival (PFS) rate than male patients (4.4 months) (Yamamoto et al., 2008). These sex-related differences might be explained by DNA repair, which was lower in females than in males (Wei et al., 2000). The lower DNA repair ability might affect tumor cell in female after administering cytotoxic anticancer drug and influence the prolonged PFS in female patients (Yamamoto et al., 2008).

**Doxorubicin**

Doxorubicin is an anthracycline DNA-damaging agent that targets the topoisomerase 2 activity and DNA intercalation (Gewirtz, 1999; Rivankar, 2014; Mitry and Edwards, 2016). Another predicted mechanism of doxorubicin generates free radicals resulting in damage to membrane and DNA of cancer cell (Gewirtz, 1999; Rivankar, 2014; Mitry and Edwards, 2016). In patients with breast cancer or lymphoma, male with normal liver function showed significantly higher doxorubicin clearance (59 l/h/m²) than female (27 l/h/m²) (Dobbs et al., 1995). Female might be an independent risk factor increasing the toxicity of doxorubicin such as cardiac abnormalities (Lipshultz et al., 1991, 1995). Exposure to doxorubicin in childhood leukemia in females increases the risk of early cardiotoxicity compared with males (Lipshultz et al., 1995). P-glycoprotein, which is expressed by multidrug resistance protein 1 gene, is a drug transporter that pumps many foreign substances out of cells (Schuetz et al., 1995). Expression of p-glycoprotein in males is 2-fold higher than in females (Schuetz et al., 1995). In females, doxorubicin and doxorubicinol, which is a doxorubicin metabolite, accumulate following a reduced expression of p-glycoprotein and leads to cardiotoxicity (van Asperen et al., 1999).

Disparities in drug metabolizing enzymes between men and women might have an impact on drug metabolism or volume of distribution of doxorubicin (Frisanco, 1974; Ley et al., 1992; Lipshultz et al., 1995). Doxorubicin is not accumulated at a higher concentration in adipose tissue (Rodvold et al., 1988, Lipshultz et al., 1995). The higher proportion of fat in female patients might lead to a higher concentration of doxorubicin in non-adipose tissues such as heart in women even if men and women carried the same body surface areas.

**Cisplatin**

Cisplatin as cis-diaminedichloroplatinum (II) exhibits covalent bonding with adjacent guanines located in the major groove of DNA (Bellon et al., 1991). This intra-strand cross-links with DNA result in distortion of DNA and antitumor effects. Female patients treated with cisplatin-based therapy show substantially higher toxicities such as vomiting and nausea than males (Liang et al., 2001). Male rats treated with cisplatin show general toxicity, prolonged thermal latency and slow motor nerve conduction velocity than female rats (Wongtawatchai et al., 2009). Sex-dependent difference may not be related to lower body size in females but physiological variables such as distinct body composition and metabolic activity (Wongtawatchai et al., 2009). In the Yoruban population which comprised of African descent, male-derived cell lines demonstrated higher sensitivity to cisplatin than female-derived cell lines (Huang et al., 2007). The half-maximal inhibitory concentration (IC₅₀) of cisplatin in male-derived cell lines was lower than that in female-derived cell lines (Huang et al., 2007).

**Bevacizumab**

Bevacizumab is a monoclonal antibody, which blocks vascular endothelial growth factor and inhibits angiogenesis in tumors (Ferrara, 2004; Sandler et al., 2006). It was reported that clearance of bevacizumab was 26% higher in male patients with solid tumors than in female patients, which is associated with a greater muscle mass in males than in females (Lu et al., 2008). Chemotherapy with bevacizumab for non-small cell lung cancer led to more severe hypertension and neutropenia and higher rate of abdominal pain in female patients than in male patients (Brahmer et al., 2011).

**Rituximab**

Rituximab is a monoclonal antibody, which targets CD20 on B cell surface. It is used to treat autoimmune diseases and hematological cancers such as non-Hodgkin’s lymphoma and chronic lymphocytic leukemia. The clearance of rituximab is higher in male patients (8.21 ml/h) with DLBCL than in female patients (12.68 ml/h) and the elimination half-life in male patients (t1/2=30.7 days) is longer compared with female patients (t1/2=24.7 days) (Müller et al., 2012). In the DLBCL treatment with rituximab, the better treatment outcomes were more prominent in females than in males (Riihijärvi et al., 2011). GSTT1 deletion might be related to adverse prognosis only in DLBCL male patients who are treated with rituximab (Cho et al., 2010). Also, in the treatment with rituximab, male patients had a worse PFS than female patients both in DLBCL and in follicular lymphoma (Riihijärvi et al., 2011).

**CONCLUSIONS**

The present review highlights the importance of sex differences in the epidemiology and chemotherapy of cancer. Evidence supports sex differences in efficacy and toxicity to anticancer drugs based on individual pharmacokinetics and pharmacodynamics. Data suggest that sex influences pathophysiology, clinical signs, treatment outcome and response in cancer. Sex is a crucial factor in predicting chemotherapy outcome, with implications for therapeutic efficacy and toxicity. Our review provides supporting information for appropriate chemotherapy based on sex. Based on the plethora of studies reporting potential sex differences in cancer, cancer research and therapy should be considered specifically to enhance patient outcomes. Sex difference in cancer susceptibility can be used to develop a causal hypothesis for the disease, or to
define subgroups at the highest risk for preventive action. Sex plays a crucial role in improving individual pharmacogenomics and in developing personalized therapeutic medicines. Pharmacogenomic differences between the sexes might play a significant role in chemotherapy in the future. Further studies are needed to provide greater insight into sex differences in cancer and improve treatment outcomes with anticancer agents.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGMENTS

The present study was supported by the Duksung Women’s University Research Grant 2016.

REFERENCES

Anderson, G. D. (2005) Sex and racial differences in pharmacological response: where is the evidence? Pharmacogenetics, pharmacoki

etics, and pharmacodynamics. J. Womens Health (Larchmt) 14, 19-29.

Aran, M., Mühlbauer, R., Hengstler, J., Jäger, E., Fuchs, J., Winkler, L. and Oesch, F. (1996) A multiplex polymerase chain reaction proto
col for the simultaneous analysis of the glutathione S-transferase GSTM1 and GSTT1 polymorphisms. Anal. Biochem. 236, 184-186.

Arnold, M., Sierra, M., Laversanne, M., Soerjomataram, I., Jemal, A. and Bray, F. (2017) Global patterns and trends in colorectal cancer incidence and mortality. Gut 66, 683-691.

Asher, G., Lotem, J., Kama, R., Sachs, L. and Shaul, Y. (2002) NQO1 stabilizes p53 through a distinct pathway. Proc. Natl. Acad. Sci. U.S.A. 99, 3099-3104.

Becker, J. B., Arnold, A. P., Berkley, K. J., Blaustein, J. D., Eckel, L. A., Hampson, E., Herman, J. P., Marts, S., Sadee, W., Steiner, M., Taylor, J. and Young, E. (2005) Strategies and methods for research on sex differences in brain and behavior. Endocrincine 146, 1650-1673.

Bellon, S. F., Coleman, J. H. and Lippard, S. J. (1991) DNA unwinding produced by site-specific intrastrand cross-links of the antitumor drug cis-diaminedichloroplatinum (II). Biochemistry 30, 8026-8035.

Bertiucco, P., Chatenoud, L., Levi, F., Fraud, D., Ferlay, J., Negri, E., Malvezzi, M. and La Vecchia, C. (2009) Recent patterns in gastric cancer: a global overview. Int. J. Cancer 125, 666-673.

Bond, G. L., Hu, W., Bond, E. E., Robins, H., Lutzker, S. G., Arva, N. C., Barghetti, J., Bartel, F., Taubert, H., Wuerl, P., Onel, K., Yip, L., Hwang, S. J., Strong, L. C., Lozano, G. and Levine, A. J. (2004) A single nucleotide polymorphism in the MDM2 promoter attenuates the p53 tumor suppressor pathway and accelerates tumor formation in humans. Cell 119, 591-602.

Bond, G. L., Hirshfield, K. M., Kirchhoff, T., Alexe, G., Bond, E. E., Robins, H., Bartel, F., Taubert, H., Wuerl, P., Hait, W., Toppmeyer, D., Offit, K. and Levine, A. J. (2006) MDM2 SNP309 accelerates tumor formation in a gender-specific and hormone-dependent manner. Cancer Res. 66, 5104-5110.

Boluffer, P., Collado, M., Barragán, E., Cervera, J., Calasanz, M. J., Colomer, D., Roman-Gómez, J. and Sanz, M. A. (2007) The potential effect of gender in combination with common genetic polymorphisms of drug-metabolizing enzymes on the risk of developing acute leukemia. Haematologica 92, 308-314.

Brahmer, J. R., Dahlberg, S. E., Gray, R. J., Schiller, J. H., Perry, M. C., Sandler, A. and Johnson, D. H. (2011) Sex differences in outcome on sex differences in brain and behavior. Endocrincine 146, 1650-1673.

Crocetti, E., Mallone, S., Robsahm, T. E., Gavin, A., Agius, D., Ardanaz, E., Lopez, M. C., Innos, K., Minicoczi, P., Borrogono, L., Pierrannunzio, D., Eisemann, N. and EUROCARE-5 Working Group (2015) Survival of patients with skin melanoma in Europe increases further: results of the EUROCARE-5 study. Eur. J. Cancer 51, 2179-2190.

Diafio, R. B. and Harris, B. E. (1989) Clinical pharmacology of 5-fluo
rouracil. Clin. Pharmacokinet. 16, 215-237.

Do, T. N., Ucisik-Akkaya, E., Davis, C. F., Morrison, B. A. and Dorak, M. T. (2010) An intronic polymorphism of IRF4 gene influences gene transcription in vitro and shows a risk association with childhood acute lymphoblastic leukemia in males. Biochim. Biophys. Acta 1802, 292-300.

Doobs, N. A., Twelves, C. J., Gillies, H., James, C. A., Harper, P. G. and Rubens, R. D. (1995) Gender affects doxorubicin pharmacoki

tetics in patients with normal liver biochemistry. Cancer Chemother. Pharmacol. 36, 473-476.

Dorak, M. T. and Karpuzoglu, E. (2012) Gender differences in cancer susceptibility: an inadequately addressed issue. Front. Genet. 3, 268.

Ellyahu, D., Michalovitz, D., Ellyahu, S., Pinhasi-Kimhi, O. and Oren, M. (1989) Wild-type p53 can inhibit oncogene-mediated focus formation. Proc. Natl. Acad. Sci. U.S.A. 86, 8763-8767.

Etienne, M. C., Lagrange, J. L., Dassonville, O., Fleming, R., Thyss, A., Renée, N., Schneider, M., Demard, F. and Milano, G. (1994) Population study of dihydroprymidine dehydrogenase in cancer patients. J. Clin. Oncol. 12, 2248-2253.

Everson, G. T., McKinley, C. and Kern, F. J. (1991) Mechanisms of gallstone formation in women. Effects of exogenous estrogen (Premarin) and dietary cholesterol on hepatic lipid metabolism. J. Clin. Invest. 87, 237-246.

Fagerholm, R., Hofstetter, B., Tommiska, J., Aaltonen, K., Vrtel, R., Syrijäkoski, K., Kallioniemi, A., Kilpivaara, O., Mannenmaa, A., Kosma, V. M., Uusitupa, M., Eskelinen, M., Kataja, V., Altoniemi, K., von Smitten, K., Heikkilä, P., Lukas, J., Holli, K., Barkova, J., Blomqvist, C., Bartek, J. and Nevanlinna, H. (2008) NAD(P)H:quinone oxido
deructase 1 NQO1*2 genotype (P187S) is a strong prognostic and predictive factor in breast cancer. Nat. Genet. 40, 844-853.

Farhana, L., Nangia-Makker, P., Arbit, E., Shango, K., Sarkar, S., Mahmud, H., Hadden, T., Yu, Y. and Majumdar, A. P. (2016) Bile acid: a potential inducer of colon cancer stem cells. Stem Cell Res. Ther. 7, 181.

Ferrara, N. (2004) Vascular endothelial growth factor: basic science and clinical progress. Endovcr. Rev. 25, 581-611.

Fitzmaurice, C., Allen, C., Barber, R. M., Barregard, L., Bhatta, Z. A., Brenner, H., Dicker, D. J., Chimed-Orchir, O., Dandona, R., Dandona, L., Fleming, T., Forouzanfar, M. H., Hancock, J. H., Hay, R. J., Hunter-Merritt, R., Huynh, C., Hosgood, H. D., Johnson, C. Q., Jonas, J. B., Khubchandani, J., Kumar, G. A., Kutz, M., Lan, O., Larson, H. J., Liang, X., Lim, S. S., Lopez, A. D., MacIntyre, M. F., Marczak, L., Marquez, N., Mokdad, A. H., Pinho, C., Pourmalek, F., Salomon, J. A., Sanabria, J. R., Sandar, L., Sartorius, B., Schwartz, S. M., Shackelford, K. A., Shibuya, K., Stanaway, J., Steiner, C., Sun, J., Takahashi, K., Vollett, S. E., Yos, T., Wagner, J. A., Wang, H., Westerman, R., Zeeb, H., Zockler, L., Abd-Allah, F., Ahmed, M. B., Alabed, S., Alam, N. K., Al-Raddadi, R., Almquist, A., Asayesh, H., Atnafu, N., Awasthi, A., Saleem, H. B., Barac, A., Bedi, N., Bensonor, I., Berhanie, A., Bernabé, E., Betsu, B., Binagghi, A., Boneya, D., Campos-Nanot, I., Castañeda-Ontuva, C., Catalá-López, F., Chang, P., Chibueze, C., Chilteer, A., Choi, J. Y., Cowie, B., Damtew, S., das Neves, J., Dey, S., Dharmaratne, S., Dhillon, P., Ding, E., Driscoll, T., Endries, A. Y., Ekwueme, D., Endries, A. Y., Fagerstrom, K., Frenk, J., Gakidou, E., Graetz, N., Hall, A., Haq, I., Han, S., Hattis, D., Hawken, S. J., Hay, E. L., Hellenbrand, W. E., Kassebaum, N., Khare, S., Khang, Y., Kim et al.

Kim et al. Sex Differences in Cancer
Farvid, M., Farzadfar, F., Fernandes, J., Fischer, F., Ghiwot, T. T., Gebru, A., Gopalan, S., Hallu, A., Horino, M., Horita, N., Hussein, A., Huybrechts, I., Ioune, M., Ismail, F., Jakovljevic, M., James, S., Javanbakht, M., Jee, S. H., Kasaeian, A., Kedir, M. S., Khader, Y. S., Khang, Y. H., Kim, D., Leigh, J., Linn, S., Lunevicius, R., El Razek, H. M. A., Malekzadeh, R., Malta, D. C., Marconcini, W. M., Maros, D., Melaku, Y. A., Meles, K. G., Mendoza, W., Mengistie, D. T., Meretoja, T. J., Miller, T. R., Mohammad, K. A., Mohammad, A., Mohammad, S., Moradi-Lakeh, M., Nagel, G., Nand, D., Le Nguyen, Q., Nolte, S., Ogbo, F. A., Oladimeji, K. E., Oren, E., Pa, M., Park, E. K., Pereira, D. M., Piias, D., Qorbani, M., Radfar, A., Rafay, A., Rahman, M., Rana, S. M., Sareide, K., Sapathay, M., Sawhney, M., Sepanlu, S. G., Shakiht, M. A., She, J., Shiue, I., Shore, H. R., Shritte, M. G., So, S., Sonjei, S., Stathopoulou, V., Strumpoupolis, K., Sufyan, M. B., Sykes, B. L., Tabárész-Seisdedos, R., Tadese, F., Tieda, B. A., Tessaema, G. A., Thakur, J. S., Tran, B. X., Ukwaja, K. N., Uzochukwu, B. S. C., Vlassov, V. V., Weiderpass, E., Wubshef Terefe, M., Yebyo, H. G., Yimam, H. H., Youm, E. N., Yonemoto, N., Youris, M. Z., Yu, C., Zaidi, Z., Zaki, M. E. S., Zenebe, Z. M., Murray, C. J. L. and Naghavi, M. (2017) Global, regional, and national cancer incidence, mortality, years of life lost, years lived with disability, and disability-adjusted life-years for 32 cancer groups, 1990 to 2015: a systematic analysis for the global burden of disease study. JAMA Oncol. 3, 524-548.
Frisancho, A. R. (1974) Triceps skin fold and upper arm muscle size norms for assessment of nutrition status. Am. J. Clin. Nutr. 27, 1052-1058.
Gewirtz, D. A. (1999) A critical evaluation of the mechanisms of action of fluoro-uracil. Cancer J. 5, 524-548.
Grodstein, F., Newcomb, P. A. and Stampfer, M. J. (1999) Postmenopausal hormone therapy and the risk of colorectal cancer: a review and meta-analysis. Am. J. Med. 106, 574-582.
Harris, B. E., Carpenter, J. T. and Diasio, R. B. (1991) Severe 5-fluorouracil toxicity secondary to dihydropyrimidine dehydrogenase deficiency. A potentially more common pharmacogenetic syndrome. Cancer 68, 499-501.
Hayes, J. D. and Strange, R. C. (2000) Glutathione S-transferase polymorphisms and their biological consequences. Pharmacology 61, 154-166.
Hsu, Y. T., Wolter, K. G. and Youle, R. J. (1997) Cytosol-to-membrane redistribution of Bax and Bcl-xL during apoptosis. Proc. Natl. Acad. Sci. U.S.A. 94, 3668-3672.
Huang, R. S., Kistner, E. O., Bleibel, W. K., Shukla, S. J. and Dolan, M. A. (2007) The potential of gender-specific tumor pharmacology. J. Natl. Cancer Inst. 99, 1201-1207.
Huang, R. S., Kistner, E. O., Bleibel, W. K., Shukla, S. J. and Dolan, M. A. (2007) The potential of gender-specific tumor pharmacology. J. Natl. Cancer Inst. 99, 1201-1207.
Huang, R. S., Kistner, E. O., Bleibel, W. K., Shukla, S. J. and Dolan, M. A. (2007) The potential of gender-specific tumor pharmacology. J. Natl. Cancer Inst. 99, 1201-1207.
Stein, B. N., Petrelli, N. J., Douglass, H. O., Driscoll, D. L., Arcangeli, G. and Meropol, N. J. (1995) Age and sex are independent predictors of 5-fluorouracil toxicity. Analysis of a large scale phase III trial. *Cancer* 75, 11-17.

Torre, L. A., Siegel, R. L., Ward, E. M. and Jemal, A. (2016) Global cancer incidence and mortality rates and trends—an update. *Cancer Epidemiol. Biomarkers Prev.* 25, 16-27.

Tran, C., Knowles, S. R., Liu, B. A. and Shear, N. H. (1998) Gender differences in adverse drug reactions. *J. Clin. Pharmacol.* 38, 1003-1009.

Traver, R. D., Horikoshi, T., Danenberg, K. D., Stadlbauer, T. H., Danenberg, P. V., Ross, D. and Gibson, N. W. (1992) NAD(P)H:quinone oxidoreductase gene expression in human colon carcinoma cells: characterization of a mutation which modulates DT-diaphorase activity and mitomycin sensitivity. *Cancer Res.* 52, 797-802.

van Asperen, J., van Tellingen, O., Tijssen, F., Schinkel, A. H. and Beijnen, J. H. (1999) Increased accumulation of doxorubicin and doxorubicinol in cardiac tissue of mice lacking mdr1a P-glycoprotein. *Br. J. Cancer* 79, 108-113.

Wang, J. and Huang, Y. (2007) Pharmacogenomics of sex difference in chemotherapeutic toxicity. *Curr. Drug Discov. Technol.* 4, 59-68.

Wasternack, C. (1980) Degradation of pyrimidines and pyrimidine analogs—pathways and mutual influences. *Pharmacol. Ther.* 8, 629-651.

Wei, Q., Cheng, L., Amos, C. I., Wang, L. E., Guo, Z., Hong, W. K. and Spitz, M. R. (2000) Repair of tobacco carcinogen-induced DNA adducts and lung cancer risk: a molecular epidemiologic study. *J. Natl. Cancer Inst.* 92, 1764-1772.

Wongtawatchai, T., Agthong, S., Kaewsema, A. and Chentanez, V. (2009) Sex-related differences in cisplatin-induced neuropathy in rats. *J. Med. Assoc. Thai.* 92, 1485-1491.

Yamamoto, H., Sekine, I., Yamada, K., Nokihara, H., Yamamoto, N., Kunitoh, H., Ohe, Y. and Tamura, T. (2008) Gender differences in treatment outcomes among patients with non-small cell lung cancer given a combination of carboplatin and paclitaxel. *Oncology* 75, 169-174.

Yoshioka, A., Tanaka, S., Hiraoka, O., Koyama, Y., Hirota, Y., Ayusawa, D., Seno, T., Garrett, C. and Wataya, Y. (1987) Deoxyribonucleoside triphosphate imbalance. 5-Fluorodeoxyuridine-induced DNA double strand breaks in mouse FM3A cells and the mechanism of cell death. *J. Biol. Chem.* 262, 8235-8241.

Zheng, L., Wang, Y., Schabath, M. B., Grossman, H. B. and Wu, X. (2003) Sulfotransferase 1A1 (SULT1A1) polymorphism and bladder cancer risk: a case-control study. *Cancer Lett.* 202, 61-69.

Zucker, I. and Beery, A. K. (2010) Males still dominate animal studies. *Nature* 465, 690.