Original Research Article

The Relationship of CD4 Cells and History of Diarrhea to Cryptosporidium sp. and Giardia lamblia Infections in HIV/AIDS Patients in West Sumatra Indonesia and its DNA Isolation

Nuzulia Irawati¹, Goldha Faroliu²* and Zahra Frizki Asty²

¹Parasitology Laboratory, Faculty of Medicine, Andalas University, Padang 25138, Indonesia
²Biomedical Sains, Faculty of Medicine, Andalas University, Padang 25138, Indonesia

*Corresponding author

A B S T R A C T

Opportunistic infections are infections that occur due to a decrease in the immune system and can cause serious conditions in people with weak immune systems such as in people with HIV / AIDS. Cryptosporidium sp. and Giardia lamblia are examples of the opportunistic intestinal protozoa that can cause chronic diarrhea so that it becomes dehydrated and malnutrition which can increase mortality and morbidity in HIV sufferers. Observational study with cross-sectional design on 50 fecal samples of HIV / AIDS patients. Cryptosporidium sp. and Giardia lamblia infections tested microscopically using Ziehl Neelsen's staining and molecular examination with DNA isolation, and looking at the relationship between CD4 cell counts and history of diarrhea with Cryptosporidium sp. and Giardia lamblia infections. The results obtained 2 respondents (4%) positive infected with Cryptosporidium sp., There is a significant relationship between CD4 cell counts with Cryptosporidium sp. infection, p-value=0.001 (p<0.05) and there is a significant relationship between Cryptosporidium sp. infections with a history of diarrhea, p-value=0.045 (p<0.05). And obtained 1 respondents (2%) positive infected with Giardia lamblia. There is no significant relationship between CD4 cell counts with Giardia lamblia infection, p-value=1.000 (p>0.05) and there is a significant relationship between Giardia lamblia infections with a history of diarrhea, p-value=0.031 (p<0.05). 18 out of 50 samples have been successfully carried out the process of isolating DNA specifically thick cell walled microorganisms.

Keywords
Cryptosporidium sp., Giardia lamblia, history of diarrhea, HIV/AIDS, CD4 cells

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Introduction

HIV is a type of virus that attacks the human immune system, especially CD4 T lymphocyte cells, with a decrease in immune status (CD4) can make various microorganisms such as bacteria, viruses, protozoa, and fungal tend to grow and multiply causing secondary infections (Mandal, 2008).
From UNAIDS (United Nation Programme on HIV/AIDS) data, cases of HIV/AIDS in the world is increase. In 2018 around 37.9 million HIV patients were discovered in the world, with 1.7 million new patients and 770,000 dying from AIDS. Asia Pacific increased from the third position in 2017 to second place in 2018 with the number of patients reaching 5.9 million, with 310,000 new patients, and 200,000 suffering from AIDS (UNAIDS, 2019). In Indonesia, the number of HIV/AIDS cases has increased compared to the last 10 years, while West Sumatra ranks eleventh (UNAIDS, 2018; Health Ministry of Indonesia, 2018).

An increase in HIV/AIDS cases is followed by an increasing number of cases of opportunistic infections that occur due to a decrease in the immune system (Agarwal, et al., 2015). One of the opportunistic infections that still causes high rates of morbidity and mortality in people with HIV/AIDS is diarrhea, in individuals with immune deficiencies often results in mild to severe symptoms resulting in chronic diarrhea and can increase in the mortality rate of HIV patients (Prasetyo, 2015).

One of the main agents causing diarrhea in HIV/AIDS patients is intestinal parasites. Intestinal protozoa that often cause disease in HIV patients include Entamoeba histolytica, Cryptosporidium sp., and Giardia lamblia (Deb et al, 2015). HIV/AIDS patients who have a CD4 cell ≤200 cells/µl become easily infected with intestinal protozoa and cause severe clinical symptoms, such as diarrhea.

Cryptosporidium sp. is the most parasite found in the stool of HIV patients (Wahdini et al, 2016). In the research of Masarat (2012) found 80% of cases of cryptosporidiosis in HIV patients. Giardia infection can also increase due to a lower CD4 cell count, especially in patients infected with HIV (Daryani, Sharif & Meigouni, 2009). In a study of 75 HIV-infected adults in India, G. lamblia was the most commonly isolated parasite, and patients with lower CD4 cell counts had more enteric disease and chronic diarrhea (Dwivedi et al., 2007).

Along with the increase in cases of HIV/AIDS and chronic diarrhea as one of the most common opportunistic infections, the incidence of Cryptosporidium sp. and Giardia lamblia needs to be investigated.

Materials and Methods

In this study, 50 faecal samples from HIV/AIDS patients from Dr. M. Djamil Padang Hospital Indonesia. Samples were examined microscopically by modified Ziehl Neelsen’s acid-resistant staining and molecular examination by isolating microorganism DNA specifically with thick cell walls using QIAamp® Fast DNA Stool Mini kits (InhibitEX Buffer, Proteinase K, Buffer AL, Buffer AW1, Buffer AW2, Buffer ATE) Qiagen®.

Results and Discussion

In this study, 50 respondents with HIV/AIDS were obtained from the Dr. M. DjamilPadang Hospital Indonesia. Respondent with the lowest CD4 cell count of 5 cells/µL and highest CD4 cell count of 350 cells/µL, and with a history of diarrhea in the last 2 weeks of 11 people (22%). In this study, respondents who were positive infected withCryptosporidium sp.and Giardia lamblia had a history of diarrhea in the past 2 weeks.

Based on the research conducted, microscopic examination (Table 1) found a total detected by Cryptosporidium sp. as many as 2 respondents (4%) and 1 respondent (2%) were detected infected with Giardia lamblia.
On microscopic examination found *Cryptosporidium sp.* oocysts with round shape almost resembles an oval measuring 4-6 micrometers. And also found an oval-shaped *Giardia lamblia* cyst that has a length of about 8-12 microns, has a thin and strong wall. The cytoplasm is fine-grained and is clearly separated from the cyst wall. Then the cyst found clearly has 2 nucleus.

*Cryptosporidium sp.* and *Giardia lamblia* microscopically tested with Ziehl Neelsen's acid-resistant staining. The Ziehl-Neelsen staining was chosen because it is a method choice that is quite simple, inexpensive, and provides quite high sensitivity. Ziehl-Neelsen's examination serves to determine the reaction of microorganism cell walls through a series of staining. Groups of acid-resistant bacteria and other microorganisms that have lipid content in cell wall are so thick that they cannot be colored with ordinary coloring reactions, but must be with acid-resistant staining and can maintain dyes when washed with a pale solution. In thick cell wall microorganisms, the cell walls are dehydrated by alcohol treatment, pores shrink, cell wall permeability and membranes decrease so that the methylene blue dye cannot enter so that microorganisms with thick cell wall will remain red in color.

In this study the results of the examination of *Cryptosporidium sp.* using Ziehl-Neelsen was obtained 2 respondents (4%) positively infected with *Cryptosporidium sp.* with average of CD4 cell count was 150±0 cells/µL, while the average CD4 cell count in respondents who were negative for *Cryptosporidium sp.* infection was 206.79±110.668 cells/µL, from the results of statistical tests using the Independent Sample T-Test to determine the relationship between CD4 cell counts and *Cryptosporidium sp.* infection with p-value=0.001 (p<0.05) showed a relationship between CD4 cell count and *Cryptosporidium sp.* infection. With a significant difference between CD4 cell counts in respondents who were positive infected with *Cryptosporidium sp.* with respondents who were negative infected with *Cryptosporidium sp.* In accordance with the research of Mitra et al (2016), respondents with CD4 counts ≤350 cells/µL were more infected with *Cryptosporidium sp.* compared to other parasites.

In a previous study, examination of *Giardia lamblia* in HIV patients with a CD4 cell count of ≤200cells/µL obtained a positive total of 3 (3%) from 100 samples (Gupta et al, 2013). Accordance with Obateru et al (2016) that from the results of his research on *Giardia lamblia* examination in HIV sufferers with a CD4 cell count of ≤200cells/µL, a total of 9 positive people (3.7%) were found, especially in individuals with HIV infection at risk of *Giardia lamblia* infection if CD4 ≤400 cells/µL (Faria et al, 2017). Statistical tests for the relationship of CD4 cell counts with *Giardia lamblia* infection obtained p-value=1,000 (p> 0.05), which showed no significant relationship between CD4 cell counts and *Giardia lamblia* infection. In accordance with the research of Samuel et al (2017) relationship between the number of CD4 cells with *Giardia lamblia* infection obtained p-value=0.852 which indicates there is no significant relationship between the number of CD4 cells with *Giardia lamblia* infection.

In this study, respondents who had a history of diarrhea in the last 2 weeks, 2 out of 50 respondents (4%) in patients with a history of diarrhea positively infected with *Cryptosporidium sp.*, and 1 out of 50 respondents (2%) in patients with a history of diarrhea positively infected with *Giardia lamblia* (Table 2), based on statistical tests showing a significant relationship between *Cryptosporidium sp.* and *Giardia lamblia* infections with a history of diarrhea.
Table 1 Relationship of CD4 Cell Counts with Cryptosporidium sp. and Giardia lamblia Infections

| Result          | N   | Mean ± SD CD4 (cells/µL) | p-value |
|-----------------|-----|--------------------------|---------|
| Cryptosporidium sp. |     |                          |         |
| Positive        | 2   | 150 ± 0                  | 0.001   |
| Negative        | 48  | 206.79 ± 10.67           |         |
| Giardia lamblia |     |                          |         |
| Positive        | 1   | 150 ± 0                  | 1.000   |
| Negative        | 49  | 205.63 ± 11.48           |         |

Table 2 Relationship of Cryptosporidium sp. and Giardia lamblia with a history of diarrhea

| Result          | History of Diarrhea | Total (%) | p       |
|-----------------|---------------------|-----------|---------|
|                 | Present             | No        |         |
| Cryptosporidium sp. | 2 (4%)             | 0 (0%)    | 2 (4%)  | 0.045   |
| Giardia lamblia  | 1 (2%)              | 0 (0%)    | 1 (2%)  | 0.031   |
| Negative (%)     | 9 (18%)             | 39 (78%)  | 48 (96%)|         |

Table 3 DNA Isolation

| No | Nucleid Acid Conc. (ng/µl) | 260/280 | 260/230 |
|----|---------------------------|---------|---------|
| 1  | 1.8                       | 0.33    | 0.01    |
| 2  | 1.2                       | 0.44    | 0.01    |
| 3  | 14.5                      | 3.59    | 0.06    |
| 4  | 1.8                       | 0.62    | 0.01    |
| 5  | 4.7                       | 2.66    | 0.03    |
| 6  | 10                        | 5.69    | 0.05    |
| 7  | 12.4                      | 2.86    | 0.09    |
| 8  | 6.8                       | 10.49   | 0.05    |
| 9  | 0.6                       | 0.11    | 0       |
| 10 | 7.2                       | 10.72   | 0.05    |
| 11 | 3.3                       | 1.98    | 0.03    |
| 12 | 2.7                       | 1.05    | 0.02    |
| 13 | 0.4                       | 0.08    | 0       |
| 14 | 1.6                       | 0.44    | 0.01    |
| 15 | 2.4                       | 0.59    | 0.02    |
| 16 | 3.6                       | 1.59    | 0.03    |
| 17 | 2.5                       | 0.87    | 0.02    |
| 18 | 5.5                       | 5.65    | 0.04    |
| 19 | 26.1                      | 4.98    | 0.1     |
| 20 | 21.4                      | 4.9     | 0.16    |
| 21 | 30.1                      | 2.86    | 0.2     |
|   | Value 1 | Value 2 | Value 3 |
|---|---------|---------|---------|
| 22 | 34.3    | 2.78    | 0.21    |
| 23 | 19.6    | 4.71    | 0.14    |
| 24 | 8.4     | 5.92    | 5.92    |
| 25 | 7.7     | 5.04    | 0.05    |
| 26 | 0.6     | 0.12    | 0       |
| 27 | 0       | 0       | 0       |
| 28 | 2.5     | 0.61    | 0.02    |
| 29 | 19.1    | 4.7     | 0.14    |
| 30 | 1.9     | 0.54    | 0.01    |
| 31 | 5.7     | 4.22    | 0.04    |
| 32 | 1.8     | 0.5     | 0.01    |
| 33 | 11.1    | 9.32    | 0.08    |
| 34 | 0.1     | 0.02    | 0       |
| 35 | 6.6     | 4.73    | 0.05    |
| 36 | 13.5    | 2.39    | 0.1     |
| 37 | 7       | 13.11   | 0.05    |
| 38 | 24.8    | 3.64    | 0.17    |
| 39 | 0       | 0       | 0       |
| 40 | 28.2    | 2.98    | 0.19    |
| 41 | 24.2    | 3.54    | 0.17    |
| 42 | 0.3     | 0.06    | 0       |
| 43 | 5.4     | 2.3     | 0.04    |
| 44 | 10.3    | 6.57    | 0.07    |
| 45 | 6.9     | 7.01    | 0.05    |
| 46 | 63.3    | 2.42    | 0.41    |
| 47 | 27.4    | 2.6     | 0.17    |
| 48 | 47.3    | 2.53    | 0.29    |
| 49 | 29.4    | 3.17    | 0.2     |
| 50 | 9.9     | 14.12   | 0.07    |

**Fig 1.** Microscopic results of *Giardia lamblia* (1000x magnification)
Fig 2. Microscopic results of Cryptosporidium sp. (1000x magnification)

In the molecular examination, DNA isolation is carried out in the sample. DNA isolation uses a special procedure to isolate DNA from faecal samples for pathogen detection based on the QIAamp Fast DNA Stool Mini Handbook protocol, where there is a difference in treatment compared to ordinary DNA isolation, i.e. with 90-95°C heating after the sample is dissolved with the InhibitEX Buffer, it aims to help lyse lysis of bacteria (Gram Positive), parasites, or other microorganisms that have thick cell walls and are difficult for lysis. Furthermore, the results of DNA isolation were examined for the purity and concentration of DNA using Nanodrop. DNA isolates with high concentrations have good purity.

In this study, 18 out of 50 samples had high DNA concentrations above 10 ng/µl, because through the DNA isolation process specifically thick cell walled microorganisms, it was suspected that the isolated DNA might originate from thick cell walled microorganisms such as intestinal parasites.

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