Roles for Coincidence Detection in Coding Amplitude-Modulated Sounds

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Abstract

Many sensory neurons encode temporal information by detecting coincident arrivals of synaptic inputs. In the mammalian auditory brainstem, binaural neurons of the medial superior olive (MSO) are known to act as coincidence detectors, whereas in the lateral superior olive (LSO) roles of coincidence detection have remained unclear. LSO neurons receive excitatory and inhibitory inputs driven by ipsilateral and contralateral acoustic stimuli, respectively, and vary their output spike rates according to interaural level differences. In addition, LSO neurons are also sensitive to binaural phase differences of low-frequency tones and envelopes of amplitude-modulated (AM) sounds. Previous physiological recordings in vivo found considerable variations in monaural AM-tuning across neurons. To investigate the underlying mechanisms of the observed temporal tuning properties of LSO and their sources of variability, we used a simple coincidence counting model and examined how specific parameters of coincidence detection affect monaural and binaural AM coding. Spike rates and phase-locking of evoked excitatory and spontaneous inhibitory inputs had only minor effects on LSO output to monaural AM inputs. In contrast, the coincidence threshold of the model neuron affected both the overall spike rates and the half-peak positions of the AM-tuning curve, whereas the width of the coincidence window merely influenced the output spike rates. The duration of the refractory period affected only the low-frequency portion of the monaural AM-tuning curve. Unlike monaural AM coding, temporal factors, such as the coincidence window and the effective duration of inhibition, played a major role in determining the trough positions of simulated binaural phase-response curves. In addition, empirically-observed level-dependence of binaural phase-coding was reproduced in the framework of our minimalistic coincidence counting model. These modeling results suggest that coincidence detection of excitatory and inhibitory synaptic inputs is essential for LSO neurons to encode both monaural and binaural AM sounds.

Author Summary

Detecting coincident arrivals of synaptic inputs is a shared fundamental property of many sensory neurons. Such ‘coincidence detection’ usually refers to the detection of
synchronized excitatory inputs only. Experimental evidence, however, indicated that some auditory neurons are also sensitive to the relative timing of excitatory and inhibitory synaptic inputs. This type of sensitivity is suggested to be important for coding temporal information of amplitude-modulated sounds, such as speech and other naturalistic sounds. In this study, we used a minimal model of coincidence detection to identify the key elements for temporal information processing. Our series of simulations demonstrated that (1) the threshold and time window for coincidence detection were the major factors for determining the response properties to excitatory inputs, and that (2) timed interactions between excitatory and inhibitory synaptic inputs are responsible for determining the temporal tuning properties of the neuron. These results suggest that coincidence detection is an essential function of neurons that detect the 'anti-coincidence' of excitatory and inhibitory inputs to encode temporal information of sounds.

Introduction

Sound source localization, or the ability to determine the direction of a sound source, is a fundamental function of the auditory system. The interaural level difference (ILD), which is the difference of the sound levels between the two ears, is one of the primary cues of azimuthal sound localization (e.g., [1]). ILDs generally increase with sound frequency and source azimuth [2,3]. In the mammalian auditory brainstem, ILDs are first encoded by neurons of the lateral superior olive (LSO) [4,5] (see [6] for a review). Principal neurons in the LSO receive excitatory inputs from the bushy cells in the ipsilateral ventral cochlear nucleus (VCN) [7-11] and inhibitory inputs from the medial nucleus of the trapezoid body (MNTB) driven by contralateral sound stimuli [8,9,12,13]. Consequently, the binaural excitatory-inhibitory interaction at the LSO is the source of the ILD-dependent spike-rate coding, which has been a subject of numerous modeling studies (e.g., [14-19]). Furthermore, because of the interaction between phase-locked excitatory [20,21] and inhibitory inputs [22,23], spike rates of LSO neurons also vary with the interaural time difference (ITD) of low-frequency tones [24-26] or of the envelopes of amplitude-modulated (AM) sounds [27-29]. Importance of temporally precise excitatory-inhibitory interactions was also suggested by measured LSO responses to transient stimuli such as clicks [27,30].

Amplitude modulation is a general feature of natural stimuli including communication sounds, and hence processing of AM signals is a fundamental property of the auditory system [31]. For example, the critical importance for AM in speech perception has been demonstrated in a variety of studies where degradation of the modulation spectra results in loss of intelligibility (e.g., [32]). Moreover, physiological studies have shown that neurons at different levels of the ascending auditory neuraxis acquire sensitivities to AM stimuli that are different from those seen at more peripheral levels of the auditory system [33], and many of these neurons also exhibit sensitivity to the binaural cues to sound location [34]. It has been suggested that the sensitivities of more central auditory centers to specific ranges of AM reflect the necessity to encode ecologically relevant aspects of the acoustic environment (e.g., [35]).

LSO responses to monaural AM tones at different modulation frequencies were systematically studied by Joris and Yin [36]. Typically, spike rates showed a mild peak at modulation frequencies \( f_m \) of 100–500 Hz, and gradually decreased down to 50 spikes/sec at around \( f_m = 600–1000 \text{ Hz} \) (Fig 1A, blue lines). However, the variability of responses from neuron to neuron was unexpectedly large. Some LSO neurons showed monotonic decreases in spike rate along the modulation-frequency axis (Fig 1A, red lines) whereas spike rates of other units...
remained > 100 spikes/sec over the $f_m$ ranges tested (Fig 1A, green lines). A few LSO neurons showed low spike rates for monaural AM sounds (Fig 1A, orange lines). Joris and Yin [36] suggested that the low-pass nature of LSO rate tuning was likely not inherited from their inputs from spherical bushy cell in the VCN, because the firing rates of bushy cells were more stable (i.e., all-pass) with increasing $f_m$ than LSO neurons. The source of the variability in AM rate coding should thus originate at the synaptic and membrane levels of the LSO neuron, making a contrast to frequency tuning in the inferior colliculus that inherits and combines a variation of spectral tuning patterns of ascending projections [37,38]. Wang and Colburn [39] studied LSO responses to AM sounds, using detailed conductance-based models. Their series of simulations suggested that the membrane afterhyperpolarization, which was shown to be important for the characteristic ‘chopping’ responses of LSO to pure tone stimuli [18], was unlikely to be the primary mechanism for rate decreases in AM coding with increasing $f_m$, whereas the addition of a large amount of low-voltage-activated potassium (KLVA) conductance led to rate-$f_m$ functions that were more consistent with empirical results. Nevertheless, no combinations of parameters were able to comprehensively explain the diversity of LSO responses to monaural AM stimuli.

Previous physiological recordings in vivo revealed a number of characteristic response properties of LSO neurons to binaural AM sounds [27–29,36]. The output spike rate of an LSO neuron typically shows periodic changes with ITDs of the envelope of AM sounds, and its period equals the reciprocal of the modulation frequency $f_m$ (Fig 1B). Furthermore, ITD-tuning curves measured at different $f_m$ align at their toughts (Fig 1B), which is considered as a signature of excitatory-inhibitory interactions [27,29]. Since prior modeling approaches focused predominantly on intensity (ILD) coding (e.g., [14–18]), how LSO neurons encode binaural temporal information remained largely uncovered.
In the classical view of binaural coding in the brainstem, LSO neurons encode ILDs by simply subtracting inhibitory inputs from excitatory inputs (reviewed in [1,6]), whereas in the medial superior olive (MSO: another major nucleus in the mammalian auditory brainstem) coincidence detection of binaural excitatory inputs is essential for encoding ITDs [1,40,41]. However, monaural and binaural AM-tunings of LSO suggest that LSO neurons should also perform temporally precise processing of bilateral synaptic inputs. Since LSO neurons receive ipsilaterally driven excitatory inputs and contralaterally driven inhibitory inputs, binaural processing in the LSO is expected to be ‘anticoincidence detection’, in which the response rate of the neuron becomes maximal when its excitatory and inhibitory synaptic inputs arrive temporally out-of-phase.

In order to reveal underlying mechanisms of monaural and binaural AM coding in the LSO, we use a simple model of coincidence detection, which is based on the counting of coincident inputs [42,43]. Because of the smaller number of free parameters than previous conductance-based LSO models (e.g., [39,44,45]), our model allows us to examine the contribution of each biophysical factor in a much more simplified situation. After identifying the factors responsible for the observed unit-to-unit variability in monaural AM coding, we examine how these factors affect the temporal interaction of excitatory and inhibitory inputs and resulting response properties of the LSO neuron to binaural AM sounds.

**Materials and Methods**

**Excitatory Inputs to LSO**

Bushy cells in the VCN generally show phase-locked spiking activities to the envelope of AM sound stimuli. Excitatory phase-locked synaptic inputs to the LSO were modeled as an inhomogeneous Poisson process [46,47]. The output spike rate of each bushy cell was described as a periodic function:

\[
\lambda(t) = 2\pi \tilde{\lambda} p(2\pi f_m t),
\]

where \( t \) is time, \( \tilde{\lambda} \) is the average intensity, \( p \) is a \( 2\pi \)-periodic function, and \( f_m \) is the stimulus modulation frequency. We used a von-Mises distribution function [48] for the periodic function \( p(x) \). Assuming that \( I_n \) is the Modified Bessel function of order \( n \), the periodic function we used is written as:

\[
p_k(x) = \exp(k\cos(x))/(2\pi I_0(k)),
\]

with \( k \) being the concentration parameter that defines the steepness of the distribution. The degree of phase-locking, measured as vector strength (VS) [49], is related to the concentration parameter as:

\[
\text{VS} = I_1(k)/I_0(k).
\]

For more detail about theoretical formulations, see [47].

The average spike rate and the VS of a bushy cell to AM stimuli generally decrease with the modulation frequency (gerbil: [20]; cat: [21,36]). Based on previous physiological measurements, we modeled them as:

\[
\tilde{\lambda}(f_m) = \lambda_0 - 0.03f_m \text{ (spikes/sec)},
\]

and

\[
\text{VS}(f_m) = 0.65 (1 - \exp((f_m - 2000)/500))/(1 + \exp((f_m - 2000)/500)) \text{ for } f_m < 2000,
\]
with $f_m$ being the modulation frequency in Hz. Since there were considerable variations in bushy cell responses between animal species and between experimental conditions, we used a simple linear function for the rate (Fig 2A) and a monotonic decreasing function for the VS (Fig 2B), both of which roughly mimic the recorded response properties [20,21,36]. The default value of $\lambda_0$ (input intensity at zero modulation frequency) was 180 (spikes/sec). The non-linear decreasing function for simulating VS reaches zero at $f_m = 2000$ Hz. In our simulations, we used the above-defined rate and VS functions unless otherwise stated. As in a previous study.

Fig 2. LSO coincidence counting model. A: Modeled AM-frequency dependence of the excitatory input rates. B: Modeled AM-frequency dependence of phase-locking of excitatory inputs. C: Modeled operation of coincidence detection. Each vertical bar corresponds to a spike. An input coincidence is counted when the number of inputs in the coincidence window W (vertical gray rectangle) reaches or exceeds the threshold $\theta$. In this example, threshold $\theta$ is 3. The small arrow indicates an output spike rejected by the refractory period T. Effects of inhibitory inputs were modeled as threshold increase $\delta$ in the inhibition window $\Delta$ (dotted rectangle). D: Monaural AM-tuning curve with the default parameter set ($\theta = 8$ inputs, $W = 0.8$ ms, $T = 1.6$ ms). Peak rate = 138.3 spikes/sec. Peak frequency = 265 Hz. Baseline = 9.7 spikes/sec. Half-peak frequency = 549 Hz. E: Binaural AM-phase coding with the default parameter set ($f_m = 300$ Hz, $\delta = 2$ inputs and $\Delta = 1.6$ ms). Peak rate = 130.7 spikes/sec. Peak phase = $-137$ deg. Trough rate = 18.7 spikes/sec. Trough phase = $+46$ deg. Half-peak width = 191 deg. F: Modeled level-dependence of input spike rates. G: Modeled level-dependence of phase-locking. Both excitatory and inhibitory inputs were assumed to share the rate-level and VS-level functions. H: Effects of modulation frequency $f_m$ on coincidence detection. Depending on frequency, the length of one modulation cycle could be larger (at low $f_m$) or smaller (at high $f_m$) than width W of the coincidence window. Thick curves show the spike rates of the phase-locked inputs. Broken lines show the time-averaged (non-phase-locked) spike rates.

doi:10.1371/journal.pcbi.1004997.g002
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results). We varied these parameters independently and examined their effects on the monaural AM coding.

To test combined effects of the coincidence threshold and window, we varied these parameters together while fixing the ratio W/θ, so that the average number of required inputs within the window remains unchanged. We also covaried these parameters so that the maximum response rate was constant (see corresponding sections in Results for detail).

For simulating binaural AM coding, we assumed that Δ = 1.6 ms and δ = 2, which reproduces empirical phase-coding results of LSO (Fig 2E; see Fig 1B and [27] for experimental results). The relative widths of the coincidence window W and the inhibition window Δ also reflect previous slice recording results, in which the duration of an inhibitory postsynaptic current was roughly twice as long as that of an excitatory synaptic current [51]. In our numerical simulations, we used a time step of 2 μs; results were not affected by the choice of the time step if it was equal to or smaller than this value.

**Monaural Tuning: Peak, Half-Peak and Baseline**

Simulated output spike rates of the LSO model in response to monaural AM inputs depended on the modulation frequency. An 'AM-tuning curve' generally showed a peak at a certain frequency and decreases with increasing modulation frequency (Fig 2D). Following a notation of prior studies (e.g., [31,54]), we also refer to the rate- fm relation shown by an AM-tuning curve as the 'rate modulation transfer function', or the 'rate-MTF'.

To characterize the modulation-frequency dependence of the LSO output, we calculated the peak rate, peak AM frequency, and the baseline rate of the rate-MTF. The baseline rate is defined as the lowest spike rate for 25–1200 Hz. We also calculated the position of the half-peak rate, where the simulated spike rate reaches 50% of the peak rate measured from the baseline (Fig 2D). We refer to the frequency position of the half-peak rate as the 'corner frequency'. For each parameter set, we simulated the input spike sequences over 100 seconds and calculated the average output spike rate of the model LSO neuron. To estimate peak frequencies, we applied five-point smoothing and spline-interpolation to the rate-MTF curve.

**Phase-Locking**

Phase-locking of the LSO responses was quantified by the modulation gain, as was done in a previous experimental study [36]. The modulation gain is defined by 20log₁₀(R), where R is the VS of the LSO output spike sequence at the modulation frequency fm. Following a notation of prior studies (e.g., [54]), we also refer to the gain-fm relation as the 'synchrony modulation transfer function', or the 'synch-MTF'.

**Binaural Tuning: Peak, Trough and Half-Peak**

Simulated output spike rates of the LSO model in response to binaural AM inputs generally depended on the phase differences of excitatory and inhibitory inputs. A 'phase-tuning curve' generally showed a peak and a trough at certain phase differences (Fig 2E). As in previous experimental studies [26,27], we defined a positive phase difference as inhibitory inputs preceding excitatory inputs. To characterize the phase dependence of the model LSO output, we varied the phase differences of simulated excitatory and inhibitory inputs and calculated the average output spike rate of the model LSO neuron over 100 seconds at each phase difference. To examine how the model parameters (i.e., coincidence parameters θ, W, T and inhibition parameters Δ, δ) affect the binaural phase coding, we calculated the peak and trough rates, trough phase, and the half-peak width of the phase-tuning curve (Fig 2E). We used the above-described default parameters (summarized in Table 1) throughout our simulations unless
otherwise stated. For testing the effects of coincidence parameters, we fixed the AM frequency at \( f_m = 300 \) Hz, which roughly corresponded to the peak of the monaural AM response (Fig 2D), and varied the relative input timings of excitatory and inhibitory inputs. For testing frequency dependence of binaural tuning, we varied the AM frequency at 150, 300, 450 and 600 Hz.

Effects of Sound Levels

LSO neurons are sensitive to ILDs [4–6], and hence the phase-tuning curve of an LSO neuron is also affected by ILD [27]. To examine the ILD-dependence of binaural AM coding, we introduced level dependence to the intensity of excitatory and inhibitory inputs. Although there were noticeable variations across species, spike rates of input fibers generally increase with sound level (cat VCN: [36,55]; monkey VCN: [56]; gerbil VCN: [57]; cat MNTB: [22,50]; rodent MNTB: [23]). Based on these measurements, we used a sigmoid function to roughly approximate the level-dependence of these input fibers:

\[
\hat{\lambda}(l) = \frac{30 + 240}{1 + \exp\left(-\left(l - 20\right)/6.0\right)} \quad \text{(spikes/sec),}
\]

where \( l \) is the sound pressure level in dB (see Fig 2F for the curve shape). Since VCN bushy cells and MNTB neurons show similar level dependence, we assumed that both excitatory and inhibitory inputs share this level-dependence function. Non-monotonic, non-linear level-dependence of the degree of synchrony (VS) [21,36] was modeled as a combination of two sigmoid functions:

\[
\begin{align*}
\text{VS}(l) &= \left(\frac{1.0}{1 + \exp\left(-\left(l - 10\right)/4.0\right)}\right) \\
&\quad \times \left(0.1 + 1.0/\left(1 + \exp\left(+\left(l - 30\right)/15.0\right)\right)\right),
\end{align*}
\]

with \( l \) being the level in dB. The first and second terms correspond to the increasing (at low levels) and decreasing (at high levels) parts of the curve, respectively (Fig 2G).

The modeled ILDs were varied in our simulation. Positive ILD indicates that the contralateral sound that drives inhibitory inputs is stronger than the ipsilateral sound that drives excitatory inputs. Note that the level-dependent input rates were only used for the figure where effects of level differences are shown. For other simulation results, we assumed that the sound level was fixed and used the \( f_m \)-dependent input rates described earlier.

Pure Integrator Model

To clarify the importance of coincidence detection in LSO, we also created a ‘pure integrator’ model that lacks temporal windows but simply sums up the number of synaptic inputs irrespective of their timings. Let \( N_e(t) \) and \( N_i(t) \) be the numbers of excitatory and inhibitory inputs, respectively, that the model neuron received between the end of the last refractory period and time \( t \). When the summed excitatory spike counts subtracted by the weighted number of inhibitory inputs reached or exceeded the pre-set threshold \( \theta \), one output spike was generated and the model was then in a refractory period \( T \). Namely, a spike was generated at time \( t \), when \( N_e(t) - \delta N_i(t) \geq \theta \). The weight \( \delta \) of the inhibitory input corresponds to the threshold increase by inhibition. Thus the pure integrator model has three parameters: threshold \( \theta \), refractory period \( T \) and threshold increase \( \delta \). We used the same default values for this model (\( \theta = 8 \) inputs, \( T = 1.6 \) ms, \( \delta = 2 \) inputs) as for the coincidence counting model (Table 1).

Results

Monaural AM Coding in LSO

**Basic mechanisms.** With the default parameter set (see Materials and Methods), the simulated monaural rate-MTF curve (Fig 2D) showed a small peak around 200–300 Hz, similar to empirical in vivo results (Fig 1A, blue lines). The modeled spike rate was generally higher for
low modulation frequencies than for high. Fig 2H depicts the basic mechanism of this frequency dependence. For modulation frequencies that are slightly smaller than a half of the reciprocal of the coincidence window (Fig 2H, middle), the average number of inputs within the coincidence window varies considerably with time, resulting in a higher number of coincidences counted. For higher modulation frequencies (Fig 2H, bottom), the number of excitatory inputs stays almost constant (remains subthreshold) irrespective of the temporal location of the coincidence window, leading to a low number of output spikes. In other words, the temporal variation of the input counts within the coincidence window is not large enough to evoke a spike. For very low frequencies (Fig 2H, top), more than one coincidence may be counted within a modulation cycle, but some of them are rejected by the refractory period, resulting in slightly lower output rate. The peak frequency of the rate-MTF curve depends on the model parameters, which will be discussed in the following subsections.

Effects of excitatory inputs. First, we tested how variations in input parameters may alter the rate-MTF (Fig 3). Variations of the intensity $\lambda_0$ of excitatory input fibers slightly modified the overall output rate of the model LSO, but the shape of the rate-MTF curve was not greatly affected (Fig 3A). Therefore, in our following simulations, we assumed that all input fibers have the same rate $\lambda_0$ (see Materials and Methods for equations). The modulation-frequency dependence of input spike rate and phase-locking also did not generally affect the shape of the rate-MTF curve (Fig 3B). Fixing the input vector strength (VS) at the maximum value (Fig 3B, gray curve) had only limited effects. Additional fixation of the input rate to the maximum value (Fig 3B, thin curve) led to a spike rate increase without much altering the curve shape. In this case, the output spike rate slightly increased for $f_m > 1000$ Hz, which resulted from the combination of the fixed high degree of phase-locking and the entry of the second modulation cycle into the 0.8-ms coincidence window. The phase-locking output of the model LSO was not affected by the $f_m$-dependence on input rates or phase-locking (Fig 3C). In sum, the monaural AM-tuning of LSO was relatively insensitive to input parameters, which lends support to the suggestion that the observed low-pass properties of LSO neurons should originate from synaptic and membrane factors [36]. Possible effects of spontaneous inhibition will be examined separately in a later subsection.
Effects of coincidence threshold. Fig 4 summarizes the effects of the coincidence threshold $\theta$ on monaural AM-tuning. The choice of coincidence threshold $\theta$ greatly affected the response rates of the modeled LSO neuron to AM sounds (Fig 4A). Decreased thresholds led to higher baseline rates as well as to higher peak rates (Fig 4A and 4B). With lower thresholds, peak and half-peak frequencies shifted to higher modulation frequencies (Fig 4A and 4C). For a high threshold ($\theta = 10$ in Fig 4A), the AM-tuning curve was monotonic, with a peak frequency being close to zero Hz. Thus rate-MTF curves of LSO neurons with low thresholds tended to be shifted both to higher spike rates and to higher modulation frequencies. These observations suggest that LSO neurons with low rate-MTF (Fig 1A, orange lines) may have high coincidence thresholds, whereas those with high average spike rates over the entire frequency range (Fig 1A, green lines) may have low thresholds.

The modulation gain was higher for high coincidence thresholds (Fig 4D), consistent with the general assumption that a high threshold leads to better phase-locking with lower spike rates. For lower coincidence thresholds, we observed a small ‘dip’ in the synch-MTF (arrow in Fig 4D) associated to higher output rates, which was also seen experimentally in some LSO neurons [36].

Effects of coincidence window. We next examined possible effects of the coincidence window on monaural AM coding (Fig 5). Changing the width $W$ of the coincidence window led to variations in the shape of the rate-MTF curve (Fig 5A). Similar to the coincidence threshold, $W$ affected the peak and baseline rates of AM-tuning curves (Fig 5A and 5B). In contrast, the peak and half-peak frequencies did not greatly change if $W$ was in the range between 0.6 and 1.1 ms (Fig 5A and 5C). For a wide coincidence window ($W = 1.2$ in Fig 5A), a second peak appeared around 1000 Hz, because more than one modulation period lay in the coincidence window. In previous in vivo experiments, a few LSO units showed second peaks (Fig 1A, green lines).

The width $W$ of the coincidence window affected both the amplitude and the cut-off frequency of the modulation gain (Fig 5D). A longer coincidence window led to a lower modulation gain and a lower cut-off frequency. The rebound of the modulation gain around 1000 Hz for large values of $W$ arose because the simplified coincidence counting model did not incorporate low-pass properties of the membrane and synapses that limit high-frequency phase-locking.
In sum, variations in \( W \) could be related to the observed variations of the AM-tuning and modulation gain of LSO neurons. Low-rate units (Fig 1A, orange), for example, may have short coincidence windows, while high rate units (Fig 1A, green) may be associated with longer windows. Simulation results with coincidence windows of > 1.0 ms, however, were mostly inconsistent with empirical data, since second peaks in the rate-MTF curve were only infrequently observed and the modulation gain did not normally re-increase below 1000 Hz [36].

**Combined effects of coincidence threshold and window.** It was previously discussed that the height of the coincidence threshold \( \theta \) and the width \( W \) of the coincidence window were, to a certain extent, interchangeable [43]. Yet in our simulations, these parameters showed different effects on the rate-MTF curves (Figs 4 and 5). To test this hypothesis more directly, we covaried \( \theta \) and \( W \) while fixing their ratio constant (i.e., a higher threshold for a wider window). Simulated effects of these covaried parameters on the AM-tuning (Fig 6A) were qualitatively similar to the effects of the coincidence threshold (Fig 4A), rather than to the coincidence threshold alone.

![](image1.png)

**Fig 5. Effects of coincidence window \( W \).** A: AM-tuning curves (rate-MTFs). B: Peak and baseline spike rates. C: Peak and corner frequencies of the rate-MTF curves. D: Modulation gains (synch-MTFs). Line types in D correspond to those in A.

doi:10.1371/journal.pcbi.1004997.g005

![](image2.png)

**Fig 6. Combined effects of coincidence threshold \( \theta \) and coincidence window \( W \) with a fixed ratio of \( \theta/W \).** A: AM-tuning curves (rate-MTFs). B: Peak and baseline spike rates. C: Peak and corner frequencies of the rate-MTF curves. D: Modulation gains (synch-MTFs). Note that also for panels B and C, both the coincidence threshold and window were changed together as in panels A and D.

doi:10.1371/journal.pcbi.1004997.g006
window (Fig 5A). Namely, both peak heights (Fig 6A and 6B) and frequencies (Fig 6A and 6C) changed according to the threshold. However, this parameter dependence was slightly milder, indicating that changes in W counteract the effects of $\theta$. The synch-MTFs (Fig 6D) were almost insensitive to the combined variation of these parameters, with minor changes in the location of upswing due to the coincidence window. These results suggest that, when the ratio $\theta/W$ is fixed, the coincidence threshold would govern the overall tuning, while the width of the coincidence window may additionally modify the shape of the AM-tuning curves of LSO.

To further clarify the distinct effects of $\theta$ and $W$, we adjusted the size of $W$ so that the peak spike rates were comparable to the default parameter settings (138–139 spikes/sec: Fig 7A). In this case, the peak and baseline rates were insensitive to parameter changes except for very low thresholds (Fig 7A and 7B). However, the peak and corner frequencies were greater for a lower threshold with a narrower coincidence window (Fig 7A and 7C). This is because a smaller $\theta$ results in a higher half-peak frequency (as seen in Fig 4), while the width $W$ does not greatly affect these frequencies (as seen in Fig 5). Similar to Fig 6D, tails of the synch-MTF curves showed upswings because of large coincidence windows, whereas the low frequency part of the modulation gain did not show strong dependence on the parameter changes (Fig 7D). From these observations, we conclude that, at least in our model of LSO, the coincidence threshold $\theta$ and the coincidence window $W$ are not simply interchangeable. The peak and half-peak frequencies are primarily determined by the coincidence threshold, not by the coincidence window. The threshold is a major determinant for the frequency limit of the rate-MTF curve.

Effects of refractory period. In contrast to the coincidence threshold $\theta$ and window $W$, which affected the shape of the AM-tuning curve over the entire range, simulated effects of the refractory period $T$ were limited to low modulation frequencies (Fig 8A). Variations of $T$ affected the shape of the AM-tuning curve only below 300 Hz. For these low modulation frequencies, which are considerably lower than the half of the reciprocal of the coincidence window, two or more input coincidences could occur within the same modulation period, but because of the refractory period, not all coincidences led to output spikes. Hence a longer refractory period resulted in lower rates at low modulation frequencies without affecting the baseline (Fig 8A and 8B) and the corner frequency (Fig 8A and 8C). These observations suggest that a salient peak in the rate-MTF curve may be related to a refractory period $> \sim 1.4$ ms, and that monotonically decreasing rate-MTF (Fig 1A, red lines) may originate from a shorter
refractory periods. As expected from the rate-MTF, the synch-MTF for \( fm > 300 \text{ Hz} \) was not affected by the refractory period (Fig 8D).

A high coincidence threshold or a short coincidence window led to fewer coincidence counts (e.g., \( \theta = 10 \) in Fig 4A and \( W = 0.4 \text{ ms} \) in Fig 5A). In such a case, effects of the refractory period were generally weak, resulting in a more monotonic rate-MTF curve. Considering that the subthreshold membrane properties of LSO neurons are generally low-pass except for low CF neurons [19], the band-pass property of the rate-MTF should originate from the supra-threshold activity of the neuron. Our modeling results suggest that a relatively long refractory period combined with fairly high output rates (typically > 100–150 spikes/sec) may be relevant to the band-pass rate-MTFs, which were frequently observed in experiments (Fig 1A, blue curves).

**Effects of spontaneous inhibition.** MNTB neurons that send inhibitory projections to LSO are spontaneously active [23,50]. Effects of spontaneous inhibition on the AM-tuning curve, however, were found to be limited (Fig 9A). Removal of inhibition only resulted in a slight increase in the rate of response to monaural stimulation. Simple calculations may explain these limited effects of inhibition. Assuming that an LSO neuron receives 8 inhibitory inputs with a spike rate of 30 spikes/sec and that each inhibitory input elevates the threshold by 2 with an effective time window of 1.6 ms, the time-averaged threshold increase can be estimated as: \( 8 \times 30 \times 1.6 \times 10^{-3} \times 2 = 0.768 \) inputs. Therefore the average effect of spontaneous inhibition is expected to be smaller than the effect of changing the threshold by one.

Even when the spontaneous rates of inhibition were doubled, the overall shape of the rate-MTF did not significantly change (Fig 9A, broken line). Similarly, the modulation gain was not affected by the spontaneous inhibition (Fig 9B), whereas coding of binaural phase differences essentially relies on inhibition driven by contralateral stimulus sounds (Fig 2E). These results suggest that spontaneous inhibition should play only a marginal role (if any) in monaural coincidence detection, compared with the significant role of sound-evoked inhibition in binaural phase-coding of AM sounds.

**Binaural AM Coding in LSO**

**Coincidence parameters.** In the preceding sections, we have examined how variations of coincidence detection properties affect monaural AM-tuning in the model LSO. Since the LSO

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**Fig 8. Effects of refractory period T.** A: AM-tuning curves (rate-MTFs). B: Peak and baseline spike rates. C: Peak and corner frequencies of the rate-MTFs. D: Modulation gains (synch-MTFs). A jump in the peak frequency in C reflects transitions of AM-tuning between low-pass and band-pass. Line types in D correspond to those in A.

doi:10.1371/journal.pcbi.1004997.g008
is one of the earliest locations where inputs from the two ears converge, binaural AM coding of LSO in terms of coincidence detection should also be examined. In this and following subsections, we test how variations in the model parameters of coincidence detection may play a role in binaural phase-coding of AM sounds. We fixed the AM frequency at $f_m = 300$ Hz, where the model LSO strongly responded to monaural AM stimuli (Fig 2D). We here focus on the peak and trough spike rates as well as the half-peak width and the trough phase (see Fig 2E for their definitions), because the location and the width of the trough of a phase-response curve provide fundamental information on the temporal tuning of an LSO neuron that receives ipsilaterally driven excitatory inputs and contralaterally driven inhibitory inputs (e.g., [28,29]).

We first varied the coincidence threshold $\theta$ (Fig 10A–10C). As naturally expected, the overall spike rates were higher for low thresholds (Fig 10A and 10B), but the location of the trough phase did not change with the threshold (Fig 10A and 10C). The half-peak width tended to be narrower for deeper phase-tuning curves (Fig 10C).

Second, we examined the effects of the width $W$ of the coincidence window (Fig 10D–10F). A large $W$ resulted in both an increased spike rate (Fig 10E) and a more centered phase-response curve with a trough phase located near zero degrees (Fig 10D and 10F). Therefore, the value of $W$ was a major determinant of the trough phase (Fig 10F). This suggests that different mechanisms may determine the binaural phase-tunings (at fixed frequency) and the monaural $f_m$-tunings; i.e., the peak and corner frequencies of the monaural rate-MTF is affected primarily by the coincidence threshold $\theta$ (Figs 4–7), while the trough position of the binaural phase-tuning curve depends greatly on the coincidence window $W$.

Next, effects of the refractory period $T$ were tested (Fig 10G–10I). Simulation results indicated that, at $f_m = 300$ Hz, the length of $T$ does not affect the binaural phase-tuning curves. This suggests that possible variations of the refractory period may not be reflected to the variations of binaural phase response properties of real LSO neurons, except for very low ($< 200$Hz) modulation frequencies.

**Inhibitory inputs driven by contralateral sounds.** Our simulation results showed that spontaneous inhibitory inputs had only limited effects on monaural AM tuning (Fig 9). In binaural phase coding, however, sound-driven inhibition may play a more active role. To test this hypothesis, we varied the inhibitory model parameters; namely, the threshold increase $\delta$ and inhibition window $\Delta$ (see Materials and Methods for definitions).

The effect of the threshold increase $\delta$ was most prominent when it was varied from zero (i.e., no inhibitory effects) to one or two, resulting in a transition from a flat to standard phase-
response curve with a distinct peak and trough (Fig 11A). Further changes in threshold increase did not greatly affect the phase-coding (Fig 11A and 11B), indicating that effects of inhibition do not need to be extremely strong to create phase-sensitivity in LSO. Moreover, the threshold increase was generally ineffective on determining the half-peak width and the trough location (Fig 11C).

Variation in the width $\Delta$ of the inhibition window showed more dynamic effects on phase-response curves (Fig 11D). A narrow inhibition window led to a higher spike rate (Fig 11E) and to a trough phase closer to zero degrees (Fig 11F). Thus both the inhibition $\Delta$ (Fig 11D–11F) and the coincidence window $W$ (Fig 10D–10F) had comparable effects on determining the trough phase and width of the binaural phase-tuning curve, but the directions of their effects were opposite. In sum, our simulation results suggest that the overall rates of the phase-response curve were determined primarily by the coincidence threshold $\theta$, coincidence window $W$ and the inhibition window $\Delta$, whereas the trough phase was determined only by a combination of the latter two. Combined effects of these window parameters will be further examined in the next section.
Window sizes and trough positions. Fig 12A shows phase-tuning curves for different coincidence windows $W$, different inhibition windows $\Delta$ and at different modulation frequencies $f_m$. A high modulation frequency generally led to lower spiking rates, which was shown in previous in vivo recordings (Fig 1B). Overall response rates were higher for larger coincidence windows and shorter inhibition window. The period of each phase-tuning curve corresponded to the reciprocal of the modulation frequency, and most importantly, the trough positions did not change with $f_m$ (compare the model results of Fig 12A with experimental results of Fig 1B). Only exceptions were the phase-tuning curves for high modulation frequency combined with a wide inhibition window ($\Delta = 2.4$ ms and $f_m = 600$ Hz: right column of Fig 12A), where the inhibition window covered more than a single modulation cycle. The trough positions varied with the window widths, as expected from previous results (Figs 10F and 11F). The phase-tuning curve was shifted towards more positive time differences by smaller $W$ and larger $\Delta$. When the two window widths took the same value ($W = \Delta = 0.8$ ms: middle-left panel in Fig 12A), the trough was located at zero input time difference. In sum, all phase-tuning curves for different modulation frequencies aligned at the trough, whose position was dependent on the window widths $W$ and $\Delta$.

Our observations suggest that the trough position may be described as a function of these window widths. Therefore we plotted the trough positions with varied $W$ while the difference
Δ-W was fixed (Fig 12B). Under this condition, simulated trough positions were generally insensitive to W, except for very short coincidence windows (W < 0.5 ms). This implies that neither of the window width alone, but their relative size should determine the trough position. To test this hypothesis, we plotted the location of the trough with the window size difference Δ-W (Fig 12C). Simulated trough positions (solid lines) clearly matched the straight line of a slope 0.5. Thus the trough position of a phase-tuning curve can be predicted by the half of the size difference of the coincidence and inhibition windows, namely (Δ-W)/2. A simple explanation is given in Fig 12D. When the excitatory and inhibitory inputs arrive with a zero delay (Fig 12D, top), the total effects of the coincidence and inhibition windows are asymmetric in time, because the width of the inhibition window was generally longer. A temporal symmetry of these window functions was achieved when the delay equals half of the width difference (Fig 12D, bottom). In the real nervous system, however, effects of these window functions may not be entirely uniform in time; e.g., synaptic inputs usually show a rapid increase and a slower
decrease [51]. Nevertheless, these simulation results suggest that relative time scales of excitatory and inhibitory inputs can be a factor to determine the trough position of a phase-tuning curve.

Effects of interaural level differences. In the preceding sections, we investigated monaural and binaural AM coding in the LSO with fixed sound stimulus levels. Since LSO neurons are sensitive to binaural level differences, we here examine how ILDs affect the output of our coincidence counting model. We varied ILDs of non-modulating inputs (Fig 13A). Simulated spike rates at different sound pressure levels showed classical ILD-tuning with higher spike rates for more negative ILDs (see [5] for related in vivo data using tonal inputs).

We next varied ILDs and calculated model outputs to input phase differences (Fig 13B). More negative ILDs (i.e., higher level for excitatory inputs than for inhibitory inputs) led to higher spike rates, but the trough position and the overall phase-tuning property were not affected. Furthermore, the model LSO showed phase-tuning properties for a wide range of ILDs (Fig 13C). These results closely match previous in vivo recording data from the cat LSO (see Fig 8 of [27]), suggesting that coincidence detection is a fundamental operation in both ILD-coding and in phase-coding of AM sounds.

AM Coding of Pure Integrator

Our simulation results suggest that temporal processing of excitatory and inhibitory inputs is essential for both monaural and binaural AM-tunings. To further confirm the importance of coincidence detection in LSO, we performed additional simulations using a pure integrator model fed with the same set of inputs as used for the coincidence counting model. The perfect integrator model does not have a coincidence window or an inhibition window, since it simply sums excitatory inputs and subtracts inhibitory inputs regardless of their timings (see Materials and Methods).

Simulated rate-MTF curves of the pure integrator showed a broad tuning (Fig 14A), which resembles the all-pass property of input fibers (Fig 2A). The response rates were slightly reduced at low frequencies (< 300 Hz) where the refractory period plays a role. Simulated synch-MTFs of the pure integrator showed a high gain (> 3 dB) for only a narrow range of modulation frequencies (Fig 14B). For other frequencies, gains were generally lower than the coincidence counting model (Fig 4D) and physiological data [36]. Thus, the responses of the
pure integrator were largely inconsistent with empirical LSO responses to monaural AM sounds (Fig 1A and [36]).

For binaural inputs, the pure integrator model also showed sensitivity to time differences of excitatory and inhibitory inputs (Fig 14C). Although the pure integrator lacks explicit time windows, spike generation (and subsequent refractory period) imposes an effective time limit of synaptic integration, since the model neuron resets its state after each spike. Namely, for an output rate of 200 spike/sec, for example, integration of inputs must occur on the average time scale of 5 ms. In other words, even in the pure integrator, thresholds are more likely to be reached when excitatory inputs arrive earlier than inhibitory inputs, whereas early inhibition simply cancels late excitation which leads to fewer output spikes. A similar excitation-inhibition interaction was reported in a modeling study that used a slow integrator [58]. Nevertheless, simulated trough positions varied with modulation frequency (Fig 14C), indicating that simulated responses of the pure integrator did not match empirical binaural LSO responses (Fig 1B).

Discussion

(Anti-)Coincidence Detection in LSO

Amplitude modulation is a common feature of natural stimuli, and hence processing of AM signals is important in auditory [31] and other sensory systems (e.g., [59,60]). In the present study, we used a simple coincidence counting model of LSO neurons and investigated how monaural and binaural AM coding is affected by several biophysical factors. The model has a small number of abstract parameters for coincidence detection, which may be controlled by lower-level physiological mechanisms (i.e., detailed properties of the membrane and synapses). By changing the coincidence parameters of the model, we reproduced most of rate- and synch-MTF curve variations to monaural AM sounds observed in vivo [36]. Furthermore, the same modeling conditions were largely sufficient for explaining the binaural temporal coding in the LSO. Unlike the coincidence counting model, the pure integrator model failed to reproduce monaural and binaural AM responses of LSO. These results suggest that coincidence detection is a fundamental operation in the LSO for both monaural and binaural auditory information processing. Previous modeling results that showed possible roles of spike-timing dependent plasticity in LSO also support this suggestion [61].

Octopus cells in the posteroventral cochlear nucleus and principal cells of the MSO are also known to act as auditory coincidence detectors [62]. The integration window of the octopus
cell is ~1 ms [63], whereas the coincidence window of the MSO neuron is typically a few hundred microseconds or even shorter [43,64]. In our series of simulations, results were mostly consistent with empirical observations when we set the width of coincidence window at 1.0 ms or less. This value is comparable to the time scale of coincidence detection in octopus cells and even shorter than the integration time windows of cortical coincidence detectors, which are on the order of a few milliseconds [65].

Monaural coincidence detection has been studied frequently in relation to the improvement of temporal coding performance at the cochlear nucleus [55,66,67], which is sending excitatory afferents to the LSO. A prior modeling study showed non-monotonic AM-tuning curves and improved synchrony in auditory midbrain neurons where excitatory and inhibitory inputs converge [54]. Our LSO model also exhibited improved phase-locking for a wide range of parameters.

The parameters of excitatory inputs (i.e., rate and degree of phase-locking), which depended on the modulation-frequency, had only limited effects on monaural AM coding (Fig 3). Spontaneous inhibition also showed minor effects on the shape of monaural rate-MTFs (Fig 9). In contrast, the coincidence parameters, such as the threshold (Fig 4) and the window width (Fig 5), both affected the height and the shape of the rate-MTF curve but in different manners. Peak and corner frequencies were more strongly affected by the coincidence threshold than by the coincidence window. The refractory period (Fig 8) showed its effect only on the low-frequency portion of the rate-MTF. These results support the hypothesis [36] that synaptic and postsynaptic coincidence factors should play an essential role in determining monaural AM-tuning properties, while presynaptic input properties have only minor impacts. Increases in coincidence threshold or decreases in width of the coincidence window resulted in higher gain with lower spike rates (Figs 4 and 5). Existence of such relations in vivo needs to be tested in future experiments.

In contrast to monaural AM-tuning curves, binaural phase-response curves showed distinct dependence on the model parameters. Both the coincidence threshold and window affected the LSO output rates to simulated binaural inputs, but the trough position of a phase-response curve was not sensitive to the threshold but to the window width (Fig 10). Moreover, the inhibition window was found to be another important factor that determines the location of the trough (Fig 11). Our simulation results suggest that the size difference of these windows was a major factor for determining the trough location of a phase-tuning curve (Fig 12). Therefore, in monaural AM coding, the coincidence threshold determines the limiting modulation frequency (i.e., the half-peak frequency), whereas in binaural AM coding, the temporal factors (coincidence and inhibition windows) have more direct impact on determining the phase-tuning properties. Our observations lend support to the suggestion that the relative latency of the excitatory and inhibitory inputs directly affects the output of LSO neurons [9,27]. Furthermore, our minimalistic coincidence counting model reproduced the classical ILD-tuning curve (Fig 13A) and the empirical ILD-dependence of phase-tuning curves (Fig 13B). These results indicate that the LSO neuron acts as an 'anticoincidence detector' where the comparison of excitatory and inhibitory synaptic inputs occurs in a (sub)millisecond time scale to compute 'instantaneous ILDs'. Similar anticoincidence detection was reported in the sensory system of electric fish [68,69].

Sources of Variations

In agreement with our results, previous experimental studies reported comparable variations in coincidence detection parameters across neurons. The refractory period of cat LSO neurons in vivo, for example, ranged from 1.1–2.8 ms [52]. Similarly, mouse LSO neurons recorded in
vitro showed variations in the refractory period [70]. As previously suggested, membrane after-hyperpolarization and sodium channel inactivation would play major roles in determining the refractory period [18]. The number of subthreshold inputs of the gerbil LSO neurons measured in vitro also showed considerable variations (9.6 ± 2.8 excitatory inputs)[51]. The width of the coincidence window is determined by multiple factors [43], such as the synaptic and membrane time constants [62], inhibitory inputs (e.g., [71]) and dendritic processes [72]. Excitatory and inhibitory synaptic inputs, for example, showed neuron-to-neuron variability in both time scale and amplitude [51], which should be related to the model parameters W and θ. Empirically measured membrane time constants also showed large variations (1.1 ± 0.4 ms in gerbils [51]; 1.9–4.1 ms in mice; [70]; and 8.5 ± 4.5 ms in rats [73]). A previous theoretical study demonstrated that, under the assumption of linear summation of synaptic conductances and linear membrane responses, the width of coincidence window is linear to the membrane time constant [74].

The KLVA conductance, a major determinant of the resting membrane properties (e.g., [75]), shows graded distributions along the tonotopic axis in the LSO with neurons in the high-frequency medial limbs of the LSO showing the lowest density [76]. Increases in KLVA conductance reduce the resting input impedance and shorten the membrane time constant, which would result in a higher coincidence threshold and a shorter coincidence window [62]. Interplay between KLVA and sodium inactivation may further narrow the coincidence window [77]. In addition, KLVA conductance plays a major role in sensing the rising slope of synchronized synaptic inputs (VCN [78]; MSO [79]). Future dynamic clamp experiments in vitro, for example, might reveal more detailed relations between the higher-level coincidence parameters and the underlying membrane and synaptic factors. Previous studies showed that having non-homogeneous neuronal population is often beneficial for coding sensory information [59,80,81]. The mammalian LSO might also benefit from the observed heterogeneity of coincidence detection properties that may enhance the information capacity of the neuronal population.

**Relevant Modeling Approaches, Applications and Limitations**

Our minimalistic coincidence counting model has only a small number of parameters (Table 1), which made our analyses simple. As has been repeatedly discussed [82–84], there is always a trade-off in modeling studies between simplicity and realism. Simple models are useful for describing the operational principles of a complex system and suitable for large-scale simulations that are computationally demanding. It should also be noted that complex (~realistic) models do not always produce physiologically more accurate results compared to simpler models [85].

In our phase-tuning curves (Figs 10–12), model LSO spike rates were plotted against phase (or time) difference of simulated synaptic inputs. In the actual auditory system, external acoustic delays (i.e., interaural time differences) are combined with internal delays (such as cochlear, axonal, and other delays) to determine the relative synaptic input timings of the target binaural neuron [1,40]. Previous cat LSO studies hypothesized that, if the external delays are perfectly compensated by the internal delay, the trough position should be equal to zero [27,28]. Our simulation results showed that the relative sizes of the coincidence and inhibition windows may play an additional role in determining the trough position (Fig 12C). A similar assumption was claimed in a study of MSO that a binaural mismatch in excitatory synaptic time constants could be a further source of delays [86], although this hypothetical mechanism was later denied by their following study that reported symmetric synaptic inputs in the MSO [87]. Nevertheless, since inhibitory synaptic inputs are significantly slower than excitatory inputs in the LSO.
mismatches in temporal properties of ipsi- and contralateral synaptic inputs may still be an essential factor to determine the binaural tuning of LSO. Shifts of peak frequencies induced by timed inhibition were also suggested by a number of modeling studies of binaural neurons in the MSO (e.g., [88,89]).

A prior modeling study suggested that a large amount of KLVA conductance was necessary to reproduce empirical AM-frequency dependence of LSO spike rates [39]. The majority of experimental data on AM coding in LSO neurons, however, comes from high-frequency sensitive neurons with low expression of KLVA. Our results indicate that various other factors related to the coincidence parameters should also play an essential role in determining the rate-MTF. Temporal coding properties of the neuron may be affected by, for example, spatial distributions of sodium channels [90] and interactions between KLVA and hyperpolarization-activated cation currents [91]. Furthermore, summation of synaptic inputs along the bipolar dendrites of LSO [92] should also affect coincidence detection [72,93].

The coincidence counting model we used was originally introduced for investigating the binaural coding in the MSO [43]. This model can be regarded as an extension of the binaural 'shot-noise' model, in which excitatory inputs are added and inhibitory inputs are subtracted to calculate a model spike output [14,16]. Such shot-noise models share basic response characteristics to a leaky integrate-and-fire model with a short membrane time constant (e.g., [94,95]). Interchangeability of the coincidence threshold and window was suggested in MSO [43]. Our LSO simulation results, however, indicated that they have considerably different effects on AM tunings (Figs 6, 7, and 10). This would be related to the difference in operational time scales of MSO and LSO and may imply their functional differences as asserted by Remme et al. [19].

Although our results were consistent with previous empirical data [27,28,36], most of the limitations that Franken et al. [43] discussed also apply to our approach. For example, the model often shows better phase-locking at high modulation frequencies (> 1000 Hz) than empirical data, because it lacks additional factors that may limit the temporal precision of neuronal response (such as low-pass membrane, dynamical spike initiation, subthreshold potential fluctuation, etc.). This restricts the applicability of the model to the frequency range below 1000 Hz. Furthermore, we simply increased the coincidence threshold to mimic the effects of inhibitory inputs. The actual effect of inhibition, however, may be subtractive or divisive (e.g., [96]) depending on whether it is of hyperpolarizing and/or shunting nature (see also [1] for discussion on subtractive inhibition in LSO). Spontaneous inhibition to the LSO from the ipsilateral MNTB was suggested to result in reduced monaural LSO firing rates at high $f_m$ rates [39], but only for vastly unrealistic numbers of inhibitory inputs to LSO. Moreover, responses of LSO neurons may be modulated by ipsilaterally driven inhibitory inputs from the lateral nucleus of the trapezoid body (LNTB) and also by inhibitory inputs from the ventral nucleus of the trapezoid body (VNTB) that receives bilateral afferents from cochlear nuclei and efferents from higher auditory stages [97,98]. Time-delayed ipsilateral inhibition from LNTB is suggested to affect the discharge patterns of LSO neurons [99]. How inhibitory inputs affect the coincidence detection in the real LSO could be investigated by examining subthreshold membrane responses with whole-cell, in vivo methods, which were recently used for MSO neurons [64].

Our LSO model consisted of two parts: (1) generation of time-locked excitatory and inhibitory inputs and (2) coincidence detection of the LSO neuron. As was done in a previous modeling study [19], sound-driven synaptic inputs can be replaced with a more detailed model that closely represents activities of the auditory periphery [100]. Testing with more complex acoustic stimuli (such as vocalization or speech [19,31]) or with transient stimuli (such as clicks [27,30]) would further reveal the applicability and limitations of the coincidence counting model. Recent neurophysiological recordings in vivo showed that coincidence detection in
MSO is affected by its input history [64]. Similar dynamical threshold adaptation may also exist in the LSO. Further studies are necessary to reveal how monaural and binaural AM-tuning properties of LSO neurons eventually result in the sound localization behavior of the animal.

Acknowledgments

We thank Lena Koepcke for literally fruitful discussions.

Author Contributions

Conceived and designed the experiments: GA JK DJT. Performed the experiments: GA. Analyzed the data: GA. Contributed reagents/materials/analysis tools: GA. Wrote the paper: GA JK DJT.

References

1. Grothe B, Pecka M, McAlpine D. Mechanisms of sound localization in mammals. Physiol Rev. 2010; 90: 983–1012. doi:10.1152/physrev.00026.2009 PMID: 20664077
2. Maki K, Furukawa S. Acoustical cues for sound localization by the Mongolian gerbil, Meriones unguiculatus. J Acous Soc Am. 2006; 118: 872–886. PMID: 16158644
3. Koka K, Read HL, Tollin DJ. The acoustical cues to sound location in the rat: Measurements of directional transfer functions. J Acous Soc Am. 2008; 123: 4297–4309. doi:10.1121/1.2916587 PMID: 18537381
4. Boudreau JC, Tsuchitani C. Binaural interaction in the cat superior olive S segment. J Neurophysiol. 1988; 31: 442–454. PMID: 5667764
5. Tsai JJ, Koka K, Tollin DJ. Varying overall sound intensity to the two ears impacts interaural level difference discrimination thresholds by single neurons in the lateral superior olive. J Neurophysiol. 2010; 103: 875–886. doi: 10.1152/jn.00911.2009 PMID: 20018829
6. Tollin DJ. The lateral superior olive: a functional role in sound source localization. Neuroscience. 2003; 9: 127–143. PMID: 12708617
7. Cant NB, Casseday JH. Projections from the anteroventral cochlear nucleus to the lateral and medial superior olivary nuclei. J Comp Neurol. 1986; 247: 457–476. PMID: 3722446
8. Glendenning RB, Masterton RB, Baker BN, Wenthold RJ. Acoustic chiasm III: nature, distribution, and sources of afferents to the lateral superior olive in the cat. J Comp Neurol. 1991; 310: 377–400. PMID: 1723989
9. Wu SH, Kelly JB. Binaural interaction in the lateral superior olive: time difference sensitivities studied in mouse brain slice. J Neurophysiol. 1992; 68: 1151–1159. PMID: 1432074
10. Smith PH, Joris PX, Yin TCT. Projections of physiologically characterized spherical bushy cell axons from the cochlear nucleus of the cat: evidence for delay lines to the medial superior olive. J Comp Neurol. 1993; 331: 245–260. PMID: 8509501
11. KJ J, Kaye Yamada GH, Semple MN, Kitzes LM. Development of ventral cochlear nucleus projections to the superior olivary complex in gerbil. J Comp Neurol. 1995; 353: 317–340. PMID: 7751434
12. Smith PH, Joris PX, Carney LH, Yin TCT. Projections of physiologically characterized globular bushy cell axons from the cochlear nucleus of the cat. J Comp Neurol. 1991; 304:387–407. PMID: 2022755
13. Smith PH, Joris PX, Yin TCT. Anatomy and physiology of principal cells of the medial nucleus of the trapezoid body (MNTB) of the cat. J Neurophysiol. 1998; 79: 3127–3142. PMID: 9636113
14. Colburn HS, Moss PJJ. Binaural interaction models and mechanisms. In: Syka J, Aitkin L, editors. Neuronal mechanisms in hearing. New York: Plenum Press; 1981. pp. 283–288.
15. Johnson DH, Dabak A, Tsuchitani C. Function-based modeling of binaural processing: interaural level. Hear Res. 1990; 49: 301–320. PMID: 2292503
16. Reed MC, Blum JJ. A model for the computation and encoding of azimuthal information by the lateral superior olive. J Acous Soc Am. 1990; 88: 1442–1453. PMID: 2292677
17. Zacksenhouse M, Johnson DH, Tsuchitani C. Excitatory / inhibitory interaction in the LSO revealed by point process modeling. Hear Res. 1992; 62: 105–123. PMID: 1429246
18. Zhou Y, Colburn HS. A modeling study of the effects of membrane afterhyperpolarization on spike interval statistics and on ILD encoding in the lateral superior olive. J Neurophysiol. 2010; 103: 2355–2371. doi: 10.1152/jn.00385.2009 PMID: 20107123

19. Remme MWH, Donato R, Mikiel-Hunter J, Ballestero JA, Foster S, Rinzel J, McAlpine D. Subthreshold resonance properties contribute to the efficient coding of auditory spatial cues. Proc Natl Acad Sci USA. 2014; 111: E2339–E2348. doi: 10.1073/pnas.1316216111 PMID: 24843153

20. Frisina RD, Smith RL, Chamberlain SC. Encoding of amplitude modulation in the gerbil cochlear nucleus: I. A hierarchy of enhancement. Hear Res. 1990; 44: 99–122. PMID: 2329098

21. Rhode WS, Greenberg S. Encoding of amplitude modulation in the cochlear nucleus of the cat. J Neurophysiol. 1994; 71: 1797–1825. PMID: 8064349

22. Tschitlani C. Input from the medial nucleus of trapezoid body to an interaural level detector. Hear Res. 1997; 105: 211–224. PMID: 9083818

23. Kopp-Scheinpflug C, Tolnai S, Marmierca MS, Rübsamen R. The medial nucleus of the trapezoid body: comparative physiology. Neuroscience. 2008; 154:160–170. doi: 10.1016/j.neuroscience.2008.01.088 PMID: 18436383

24. Finlayson PG, Caspary DM. Low-frequency neurons in the lateral superior olive exhibit phase-sensitive binaural inhibition. J Neurophysiol. 1991; 65: 598–605. PMID: 2051197

25. Joris PX, Yin TCT. Envelope coding in the lateral superior olive. I. Sensitivity to interaural time differences. J Neurophysiol. 1995; 73: 1043–1062. PMID: 7608754

26. Joris PX, Yin TCT. Envelope coding in the lateral superior olive. II. Characteristic delays and comparison with responses in the medial superior olive. J Neurophysiol. 1996; 76: 2137–2156. PMID: 8999950

27. Joris PX, Yin TCT. Envelope coding in the lateral superior olive. III. Comparison with afferent pathways. J Neurophysiol. 2001; 86: 2647–2666. PMID: 11731526

28. Davis KA, Ramachandran R, May BJ. Auditory processing of spectral cues for sound localization in the inferior colliculus. J Assoc Res Otolaryngol. 2003; 4: 148–163. PMID: 12943370

29. Rabanb CF, Parthasarathy A, Venkataraman Y, Fisher ZL, Gardner SM, Bartlett EL. A computational model of inferior colliculus responses to amplitude modulated sounds in young and aged rats. Front Neural Circuits. 2012; 6: 77. doi: 10.3389/fncir.2012.00077 PMID: 23129994

30. Wang L, Colburn HS. A modeling study of the responses of the lateral superior olive to ipsilateral sinusoidally amplitude-modulated Tones. J Assoc Res Otolaryngol. 2012; 13: 249–267. doi: 10.1007/s10162-011-0300-5 PMID: 22160752

31. Joris P, Yin TCT. A matter of time: internal delays in binaural processing. Trends Neurosci. 2007; 30: 70–78. PMID: 17188761

32. Ashida G, Carr CE. Sound localization: Jeffress and beyond. Curr Opin Neurobiol. 2011; 21: 745–751. doi: 10.1016/j.conb.2011.05.008 PMID: 21646012
42. Heinz MG, Colburn HS, Carney LH. Rate and timing cues associated with the cochlear amplifier: Level discrimination based on monaural cross-frequency coincidence detection. J Acoust Soc Am. 2001; 110: 2065–2084. PMID: 11681385

43. Franken TP, Bremen P, Joris PX. Coincidence detection in the medial superior olive: mechanistic implications of an analysis of input spiking patterns. Front Neural Circuits. 2014; 8: 42. doi: 10.3389/fncir.2014.00042 PMID: 24822037

44. Zacksenhouse M, Johnson DH, Williams J, Tsuchitani C. Single-neuron modeling of LSO unit responses. J Neurophysiol. 1998; 79: 3098–3110. PMID: 9636111

45. Gai Y, Doiron B, Kotak V, Rinzel J. Noise-gated encoding of slow inputs by auditory brain stem neurons with a low-threshold K current. J Neurophysiol. 2009; 102: 3447–3460. doi: 10.1152/jn.00538.2009 PMID: 19812289

46. Gerstner W, Kistler W. Spiking neuron models. Cambridge UK: Cambridge University Press, 2003.

47. Ashida G, Funabiki K, Carr CE. Theoretical foundations of the sound analogue membrane potential that underlies coincidence detection in the barn owl. Front Comput Neurosci. 2013; 7: 151. doi: 10.3389/fncom.2013.00151 PMID: 24265616

48. Fisher NI. Statistical analysis of circular data. Cambridge, UK: Cambridge University Press, 1993.

49. Goldberger JM, Brown PB. Response of binaural neurons of dog superior olivary complex to dichotic tonal stimuli: some physiological mechanisms of sound localization. J Neurophysiol. 1989; 62: 613–636. PMID: 5810617

50. Koka K, Tollin DJ. Linear coding of complex sound spectra by discharge rate in neurons of the medial nucleus of the trapezoid body (MNTB) and its inputs. Front Neural Circuits. 2014; 8: 144. doi: 10.3389/fncom.2014.00144 PMID: 25565971

51. Sanes DH. An in vitro analysis of sound localization mechanisms in the gerbil lateral superior olive. J Neurosci. 1990; 10: 3494–3506. PMID: 2172478

52. Tsuchitani C. The inhibition of cat lateral superior olive unit excitatory responses to binaural tone bursts. II. The sustained discharges. J Neurophysiol. 1988; 59: 184–211. PMID: 3343600

53. Couchman K, Grothe B, Felmy F. Medial superior olivary neurons receive surprisingly few excitatory and inhibitory inputs with balanced strength and short-term dynamics. J Neurosci. 2010; 30: 17111–17121. doi: 10.1523/JNEUROSCI.1760-10.2010 PMID: 21159981

54. Nelson PC, Carney LH. A phenomenological model of peripheral and central neural responses to amplitude-modulated tones. J Acoust Soc Am. 2004; 116: 2173–2186. PMID: 15532650

55. Joris PX, Carney LH, Smith PH, Yin TCT. Enhancement of neural synchronization in the anteroventral cochlear nucleus. I. Responses to tones at the characteristic frequency. J Neurophysiol. 1994; 71: 1022–1036. PMID: 8201399

56. Rhode WS, Roth GL, Recio-Spinoso A. Response properties of cochlear nucleus neurons in monkeys. Hear Res. 2010; 259: 1–15. doi: 10.1016/j.heares.2009.06.004 PMID: 19531377

57. Keine C, Rübsamen R. Inhibition shapes acoustic responsiveness in spherical bushy cells. J Neurosci. 2015; 35: 8579–8592. doi: 10.1523/JNEUROSCI.0133-15.2015 PMID: 26041924

58. Vasilkov VA, Tikidji-Hamburyan RA. Accurate detection of interaural time differences by a population of slowly integrating neurons. Phys Rev Lett. 2012; 108: 138104. PMID: 22540730

59. Chacron MJ, Longtin A, Maler L. Efficient computation via sparse coding in electroreceptive neural networks. Curr Opin Neurobiol. 2011; 21: 752–760. doi: 10.1016/j.conb.2011.05.016 PMID: 21683574

60. Metzen MG, Jamali M, Carriot J, Ávila-Áckerberg O, Cullen CE, Chacron MJ (2015) Coding of envelopes by correlated but not single-neuron activity requires neural variability. Proc Natl Acad Sci USA. 2015; 112: 4791–4796. doi: 10.1073/pnas.1418224112 PMID: 25825717

61. Fontaine B, Peremans H. Tuning bat LSO neurons to interaural intensity differences through spike-timing dependent plasticity. Biol Cybern. 2007; 97: 261–267. PMID: 17989163

62. Golding NL, Oertel D. Synchronous integration in dendrites: exceptional need for speed. J Physiol. 2012; 590: 5563–5569. doi: 10.1113/jphysiol.2012.229328 PMID: 22930273

63. Oertel D, Bal R, Gardner, SM, Smith PH, Joris PX. Detection of synchrony in the activity of auditory nerve fibers by octopus cells of the mammalian cochlear nucleus. Proc Natl Acad Sci USA. 2000; 97: 11773–11779. PMID: 11050208

64. Franken TP, Roberts MT, Wei L, Golding NL, Joris PX. In vivo coincidence detection in mammalian sound localization generates phase delays. Nat Neurosci. 2015; 18: 444–452. doi: 10.1038/nn.3948 PMID: 25664914

65. König P, Engel AK, Singer W. Integrator or coincidence detector? The role of the cortical neuron revisited. Trends Neurosci. 1996; 19: 130–137. PMID: 8658595
66. Carney LH. Sensitivities of cells in anteroventral cochlear nucleus of cat to spatiotemporal discharge patterns across primary afferents. J Neurophysiol. 1990; 64: 437–456. PMID: 2213126
67. Wang GI, Delgutte B. Sensitivity of cochlear nucleus neurons to spatio-temporal changes in auditory nerve activity. J Neurophysiol. 2012; 108: 3172–3195. doi: 10.1152/jn.00160.2012 PMID: 22972956
68. Friedman MA, Hopkins CD. Neural substrates for species recognition in the time-coding electroson-ary pathway of mormyrid electric fish. J Neurosci. 1998; 18: 1171–1185. PMID: 9437037
69. Lyons-Warren AM, Kohashi T, Mennerick S, Carlson BA. Detection of submillisecond spike timing differences based on delay-line anticoincidence detection. J Neurophysiol. 2013; 110: 2295–2311. doi: 10.1152/jn.00444.2013 PMID: 23966672
70. Wu SH, Kelly JB. Response of neurons in the lateral superior olive and medial nucleus of the trape-zoid body to repetitive stimulation: intracellular and extracellular recordings from mouse barrel slice. Hear Res. 1993; 68: 189–201. PMID: 8407605
71. Burger RM, Fukui I, Ohmori H, Rubel EW. Inhibition in the balance: binaurally coupled inhibitory feedback in sound localization circuitry. J Neurophysiol. 2011; 106: 4–14. doi: 10.1152/jn.00205.2011 PMID: 21525367
72. Remme MWH, Rinzel J. Role of active dendritic conductances in subthreshold input integration. J Comput Neurosci. 2011; 31: 13–30. doi: 10.1007/s10867-010-9259-7 PMID: 21127955
73. Adam TJ, Finlayson PG, Schwartz DWF. Membrane properties of principal neurons of the lateral supe-rior olive. J Neurophysiol. 2001; 86: 922–934. PMID: 11495961
74. Toth PG, Marsalek P. Analytical description of coincidence detection synaptic mechanisms in the auditory pathway. BioSystems. 2015; 136: 90–98. doi: 10.1016/j.biosystems.2015.07.006 PMID: 26190796
75. Rothman JS, Manis PB. The roles potassium currents play in regulating the electrical activity of ventral cochlear nucleus neurons. J Neurophysiol. 2003; 89: 3097–3113. PMID: 12783953
76. Barnes-Davies M, Barker MC, Osmanu F, Forsythe ID. Kv1 currents mediate a gradient of principal neuron excitability across the tonotopic axis in the rat superior olive. Eur J Neurosci. 2004; 19: 325–333. PMID: 14725627
77. Svirsks G, Kotak V, Sanes DH, Rinzel J. Sodium along with low-threshold potassium currents enhance coincidence detection of subthreshold noisy signals in MSO neurons. J Neurophysiol. 2004; 91: 2465–2473. PMID: 14749317
78. McGinley MJ, Oertel D. Rate thresholds determine the precision of temporal integration in principal cells of the ventral cochlear nucleus. Hear Res. 2006; 216: 217–226. PMID: 16647828
79. Gai Y, Kotak VC, Sanes DH, Rinzel J. On the localization of complex sounds: temporal encoding based on input-slope coincidence detection of envelopes. J Neurophysiol. 2014; 112: 802–813. doi: 10.1152/jn.00044.2013 PMID: 24848460
80. Padmanabhan K, Urban NN. Intrinsic biophysical diversity decorrelates neuronal firing while increasing information content. Nat Neurosci. 2010; 13: 1276–1282. doi: 10.1038/nn.2630 PMID: 20802489
81. Hunsberger E, Scott M, Eliasmith C. The competing benefits of noise and heterogeneity in neural cod-ing. Neural Comput. 2014; 26: 1600–1623. doi: 10.1162/NECO_a_00621 PMID: 24877735
82. Izhikevich EM. Which model to use for cortical spiking neurons. IEEE Trans Neural Networks. 2004; 15: 1063–1070. PMID: 15484883
83. Herz AV, Gollisch T, Machens CK, Jaeger D. Modeling single-neuron dynamics and computations: a balance of detail and abstraction. Science. 2006; 314: 80–85. PMID: 17023649
84. Gerstner W, Sprekeler H, Deco G. Theory and simulation in neuroscience. Science. 2012; 338: 60–65. doi: 10.1126/science.1227356 PMID: 23042882
85. Brette R. What is the most realistic single-compartment model of spike initiation? PLoS Comput Biol 2015; 11: e1004144 doi: 10.1371/journal.pcbi.1004144 PMID: 25856629
86. Jercog PE, Svirsks G, Kotak VC, Sanes DH, Rinzel J. Asymmetric excitatory synaptic dynamics underlie interaural time difference processing in the auditory system. PLoS Biol. 2010; 8: e1000406. doi: 10.1371/journal.pbio.1000406 PMID: 20613857
87. Roberts MT, Seeman SC, Golding NL. A mechanistic understanding of the role of feedforward inhibition in the mammalian sound localization circuitry. Neuron. 2013; 78: 923–935. doi: 10.1016/j.neuron.2013.04.022 PMID: 23764291
88. Brand A, Behrend O, Marquardt T, McAlpine D, Grothe B. Precise inhibition is essential for microsec-ond interaural time difference coding. Nature. 2002; 417: 543–547. PMID: 12037566
89. Leibold C, van Hemmen JL. Spiking neurons learning phase delays: how mammals may develop auditory time-difference sensitivity. Phys Rev Lett. 2005; 94: 168102. PMID: 15904267
90. Ashida G, Abe K, Funabiki K, Konishi M. Passive soma facilitates submillisecond coincidence detection in the owl’s auditory system. J Neurophysiol. 2007; 97: 2267–2282. PMID: 17135480

91. Khurana S, Remme MWH, Rinzel J, Golding NL. Dynamic interaction of I_h and I_{K_{LVA}} during trains of synaptic potentials in principal neurons of the medial superior olive. J Neurosci. 2011; 31: 8936–8947. doi: 10.1523/JNEUROSCI.1079-11.2011 PMID: 21677177

92. Rietzel HJ, Friauf E. Neuron types in the rat lateral superior olive and developmental changes in the complexity of their dendritic arbors. J Comp Neurol. 1998; 390: 20–40. PMID: 9456173

93. Agmon-Snir H, Carr CE, Rinzel J. The role of dendrites in auditory coincidence detection. Nature. 1998; 393: 268–272. PMID: 9607764

94. Burkitt AN, Clark GM. Calculation of interspike intervals for integrate-and-fire neurons with Poisson distribution of synaptic inputs. Neural Comput. 2000; 12: 1789–1820. PMID: 10953239

95. Richardson MJE, Swarbrick R. Firing-rate response of a neuron receiving excitatory and inhibitory synaptic shot noise. Phys Rev Lett. 2010; 105: 178102. PMID: 21231083

96. Prescott SA, De Koninck Y. Gain control of firing rate by shunting inhibition: Roles of synaptic noise and dendritic saturation. Proc Natl Acad Sci USA. 2003; 100: 2076–2081. PMID: 12569169

97. Thompson AM, Schofield BR. Afferent projections of the superior olivary complex. Microscopy Res Tech. 2000; 51: 330–354. PMID: 11071718

98. Albrecht O, Donzdillo A, Mayer F, Thompson JA, Klug A. Inhibitory projections from the ventral nucleus of the trapezoid body to the medial nucleus of the trapezoid body in the mouse. Front Neural Circuits. 2014; 8: 83. doi: 10.3389/fncir.2014.00083 PMID: 25120436

99. Greene NT, Davis KA. Discharge patterns in the lateral superior olive of decerebrate cats. J Neurophysiol. 2012; 108: 1942–1953. doi: 10.1152/jn.00908.2011 PMID: 22745462

100. Zilany MS, Bruce IC, Carney LH. Updated parameters and expanded simulation options for a model of the auditory periphery. J Acoust Soc Am. 2014; 135: 283–286. doi: 10.1121/1.4837815 PMID: 24437768