Genome Sequence of *Arthrobacter globiformis* Phage KeAlii from Hawai‘i

Rebecca A. Chong,a University of Hawai‘i at Mānoa SEA-PHAGES students,a Stuart P. Donachie,a Floyd A. Reed,a Megan L. Portera

School of Life Sciences, University of Hawai‘i at Mānoa, Honolulu, Hawai‘i, USA

ABSTRACT  Here, we report the genome sequence of bacteriophage KeAlii, a Siphoviridae that infects *Arthrobacter globiformis* strain B-2979, from Honolulu, Hawai‘i. The 41,850-bp genome contains 66 predicted protein-coding genes and 1 gene that encodes a tRNA for tryptophan. Genome comparisons suggest KeAlii is closely related to actinobacteriophage Adolin.

Understanding the molecular evolution of bacteriophages is critical to finding potential medical solutions for antibiotic resistance in bacteria as well as diverse applications in agricultural and biotechnological settings. Here, we present the genome sequence of the actinobacteriophage KeAlii that infects the soil bacterium *Arthrobacter globiformis* strain (str.) B-2979str, providing one of the first sequenced actinobacteriophage genomes from Hawai‘i through the SEA-PHAGES program (1, 2).

KeAlii was isolated and purified in 2020 from a surface soil sample collected at the University of Hawai‘i at Mānoa (UHM) in Honolulu, on Oahu, Hawaii (21.3265 N, 157.8024 W), on a sunny day with an ambient temperature of 30°C. Using a direct isolation method as outlined in the SEA-PHAGES manual (3), KeAlii was isolated on peptone yeast calcium agar (PYCa) medium incubated at 22°C with the host *A. globiformis* str. B-2979str. Imaging of KeAlii by transmission electron microscopy revealed an icosahedral head and a noncontractile tail that are characteristic of *Siphoviridae* phages (Fig. 1).

Phage particles were confirmed and purified via plaque assay and then amplified to a high titer for genomic DNA extraction for sequencing. Total genomic DNA extractions were performed at UHM using a Wizard DNA extraction kit (Promega) following the manufacturer’s protocols. A sequencing library was prepared at the Pittsburgh Bacteriophage Institute with an NEBNext Ultra II FS DNA Library Prep Kit with dual-indexed barcoding, and sequencing was completed using the Illumina MiSeq platform resulting in 280,361 single-end 150-bp reads. Default parameters were used for all software unless otherwise specified. Raw reads were assembled in Newbler v2.9 (Roche), resulting in a single genomic contig with 922-fold coverage. Genome completeness, accuracy, and phage genomic termini were verified using Consed29 (4–6). The complete genome sequence of KeAlii (GenBank accession OK040777.1; assembly ASM2068489v1) is 41,850 bp in size with a G+C content of 65.5% and has a characteristic physical end with an 11-bp 3’ sticky overhang.

The genome of KeAlii was annotated using Glimmer v3 (7) and GeneMark v2 (8); resulting automated annotations were verified manually using DNA Master v5.23.6 build 2701 (http://cobamide2.bio.pitt.edu/computer.html), Phage Evidence Collection and Annotation Network (PECAAN; http://pecaan.kbrinsgd.org), Phamerator (9), and Starterator (http://phages.wustl.edu/starterator/). KeAlii is predicted to have 67 genes, as follows: 1 tRNA gene, 33 genes (50%) have a putative function assigned, and the remaining 33 genes (49%) encode hypothetical proteins with unknown functions. Potential functions for predicted protein-coding genes were assigned based on top hits for searches using NCBI BLASTP (10) and HHpred (11), and putative membrane proteins were identified using TMHMM v2.0 (now DeepTMHMM https://services.healthtech.dtu.dk/service.php?DeepTMHMM).
Actinobacteriophages sharing at least 50% nucleotide identity are arranged into clusters, with KeAlii falling in the AZ cluster of actinobacteriophages. KeAlii is predicted to be a temperate phage, as a serine integrase was identified (gene 47), although no immunity repressor gene was identified. Other interesting genes identified include a VIP2-like ADP-ribosyltransferase toxin gene (gene 4) and a putative endolysin (gene 24). KeAlii is most genetically similar to Adolin (GenBank accession MN813676.1), with 81.67% nucleotide identity via BLAST alignment.

**Data availability.** The complete genome sequence of actinobacteriophage KeAlii has been deposited in GenBank with accession number OK040777, Bioproject accession number PRJNA488469, and SRA accession number SRX15105353.

**ACKNOWLEDGMENTS**

This project was generously supported by the Howard Hughes Medical Institute SEA-PHAGES program and the School of Life Sciences at University of Hawai‘i Mānoa (UHM).

We thank Graham F. Hatfull, Welkin H. Pope, Deborah Jacobs-Sera, Daniel A. Russell, and Rebecca A. Garlena for their continued technical support during the sequencing and annotation of this genome.

**REFERENCES**

1. Jordan TC, Burnett SH, Carson S, Caruso SM, Clase K, DeJong RJ, Denneyh JJ, Denver DR, Dunbar D, Elgin SCR, Findley AM, Gissendanner CR, Golebiowska UP, Guild N, Hartzog GA, Grillo WH, Hollowell GP, Hughes LE, Johnson A, King RA, Lewis LO, Li W, Rosenzweig F, Rubin MR, Saha MS, Sandz J, Shaffer CD, Taylor B, Temple L, Vazquez E, Ware VC, Barker LP, Bradley KW, Jacobs-Sera D, Pope WH, Russell DA, Cresawn SG, Lopatto D, Bailey CP, Hatfull GF. 2014. A broadly implementable research course in phage discovery and genomics for first-year undergraduate students. mBio 5:e01051-13. https://doi.org/10.1128/mBio.01051-13.

2. Hanauer DJ, Graham MJ, Betancur L, Bobrownicki A, Cresawn SG, Jacobs-Sera D, Kaufmann N, Pope WH, Russell DA, Jacobs WR, Sivanathan V, Asai DJ, Hatfull GF, SEA-PHAGES. 2017. An inclusive Research Education Community (iREC): impact of the SEA-PHAGES program on research outcomes and student learning. Proc Natl Acad Sci U S A 114:13351–13356. https://doi.org/10.1073/pnas.1710188115.

3. Powell M, Pope WH, Jacobs-Sera D, Sivanathan V, Hatfull GF. 2016. Phage discovery guide. Howard Hughes Medical Institute, Chevy Chase, MD.

4. Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen Y-J, Chen Z, Dewell SB, Du L, Fierro JM, Gomes XV, Godwin BC, He W, Helgesen S, Ho CH, Ho CH, Iryzk GP, Jando SC, Alenquer ML, Jarvie TP, Jirage KB, Kim J-B, Knight JR, Lanza JR, Leamon JH, Leftkowitz SM, Lei M, Li J, Lohman KL, Lu H, Makhijani VB, McDade KE, McKenna MP, Myers EW, Nickerson E, Nobile JR, Plant R, Puc BP, Ronan MT, Roth GT, Sarkis GJ, Simons JF, Simpson JW, Srinivasan M, Tartaro KR, Tomasz A, Vogt KA, et al. 2005. Genome sequencing in microfabricated high-density picolitre reactors. Nature 437:376–380. https://doi.org/10.1038/nature03959.

5. Gordon D, Green P. 2013. Consed: a graphical editor for next-generation sequencing. Bioinformatics 29:2936–2937. https://doi.org/10.1093/bioinformatics/bt3515.

6. Russell DA. 2018. Sequencing, assembling, and finishing complete bacteriophage genomes, p 109–125. In Clokie MRJ, Kropinski AM, Lavigne R (ed), Bacteriophages: methods and protocols, volume 3. Springer, New York, NY.

7. Decker AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with Glimmer. Bioinformatics 23:673–679. https://doi.org/10.1093/bioinformatics/btm009.

8. Besemer J, Borodovsky M. 2005. GeneMark: Web software for gene finding in
9. Cresawn SG, Bogel M, Day N, Jacobs-Sera D, Hendrix RW, Hatfull GF. 2011. Phamerator: a bioinformatic tool for comparative bacteriophage genomics. BMC Bioinformatics 12:395. https://doi.org/10.1186/1471-2105-12-395.

10. Altschul SF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. https://doi.org/10.1016/S0022-2836(05)80360-2.

11. Söding J, Biegert A, Lupas AN. 2005. The HHpred interactive server for protein homology detection and structure prediction. Nucleic Acids Res 33:W244–W248. https://doi.org/10.1093/nar/gki408.