20 YEARS OF LEPTIN

Leptin at 20: an overview

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Abstract

Historically, adipose tissue was considered to be a passive storage vessel discharging nutrients in times of famine and accumulating fat in times of surfeit. This view changed with the identification of leptin as an adipocyte hormone. Leptin functions as an afferent signal in a negative feedback loop that regulates food intake and metabolism to maintain homeostatic control of adipose tissue mass. Before this, the existence of a system maintaining homeostatic control of energy balance was unclear. The identification of leptin has thus uncovered a new endocrine system that also links changes in nutrition to adaptive responses in most if not all other physiologic systems. Further studies have revealed a set of clinical syndromes caused by leptin deficiency, including lipodystrophy and hypothalamic amenorrhea. This work has led to new therapeutic approaches for a number of human conditions and has also established a conceptual framework for studying the pathogenesis of obesity.

Key Words

- leptin
- food intake
- energy homeostasis
- obesity

Introduction

Early one morning, slightly longer than 20 years ago, I developed a northern blotting showing changes in the levels of adipose tissue RNA detected by a probe named 2G7 (Ingalls et al. 1950, Zhang et al. 1994). The RNA in this experiment was derived from two mutant mouse strains each of which was homozygous for mutations in the mouse obese (ob) gene. ob is a fully penetrant autosomal recessive gene, which causes affected animals to eat for five times compared with a normal mouse, resulting in a profound increase in weight and adipose tissue mass. 2G7 was a clone that mapped to the region of chromosome 6 where the ob gene had been found to reside (Friedman et al. 1991, Bahary et al. 1993). The development of this film was the culmination of a 12-year odyssey that began with the demonstration that cholecystokinin, then considered to be a candidate for the ob gene, mapped to chromosome 9 and could not be ob (Friedman et al. 1985, 1989), to the identification of 2G7 from an exon-trapping experiment in which 182 different clones from an 100-kB P1 clone in the region of the ob gene on mouse chromosome 6 were arrayed on two microtiter plates (the ob gene was on plate 2, row G, column 7) (Zhang et al. 1994).

The 2G7 probe identified a 4.5-kb RNA in adipose tissue that was absent in RNA from CMC dac ob/ob mice but was increased 20-fold in adipose RNA from C57Bl/6j ob/ob mice, first characterized by Ingalls et al. (1950). CMC dac ob/ob mice were unpublished at the time but had been kindly provided by Skippy Lane at the Jackson Laboratory, as were many other mice that were critical for this study.

The demonstration that the same RNA was absent in one mutant and increased in the second mutant provided definitive proof the 2G7 was an exon from the ob gene. These data by themselves confirm that the ob gene had been isolated because, had the observed RNA changes been secondary to some other genetic defects, the expression levels would have to be similar in both ob mutant strains. Further studies revealed that a viral insertion in CMC dac ob/ob mice interfered with ob...
expression, while in C57Bl/6j ob/ob mice a point mutation in the second exon of the ob gene introduced a nonsense mutation at amino acid 105 of this 167 amino acid protein (Zhang et al. 1994, Moon & Friedman 1997). The marked overexpression of the ob gene in C57Bl/6j ob/ob adipose tissue further suggested that the gene was under feedback control with an increased level of gene expression in the obese state.

This finding that the ob gene was induced in ob/ob fat tissue was consistent with data from classic parabiosis experiments of Doug Coleman in a now-iconic set of experiments that involved stitching together the skins of living mice so that animals with different mutations shared a circulatory system (Coleman 1978). ob mice surgically joined to normal or db mice (on the same inbred strain background) ate less and lost weight. In contrast, normal mice paired to db mice starved to death. From this, Coleman concluded that ob mice normally lacked a circulating factor that was provided by the conjoined partner, which suppressed food intake and body weight. He further suggested that db mice lacked a receptor to detect the weight-suppressing factor in their blood and so overexpressed it, producing levels so high that conjoined mice sensitive to the factor stopped eating. Implicit in this hypothesis was the prediction that the ob gene was under feedback control and that obesity would be associated with increased levels of ob RNA. However, Coleman’s experiments did not predict where the hormone that was missing in ob mice was expressed, though prior experiments from Hervey predicted that the receptor would be expressed in the hypothalamus (Hervey 1969).

Putting this all together in that moment in a dark-room, it became evident in an instant that not only had the ob gene been cloned but also that the data were consistent with Coleman’s predictions. The data thus suggested the hypothesis that the ob gene encoded a novel adipocyte hormone which functioned as the afferent signal in a negative feedback loop that maintains homeostatic control of adipose tissue mass. Subsequent studies have confirmed this possibility in animals and humans, and some of the fruits of this set of observations are summarized in this timely volume and elsewhere (Halaas et al. 1995, Tartaglia et al. 1995, Lee et al. 1996, Friedman & Halaas 1998). While some of the features of this new hormonal system were predicted at the time, others were not. Science seldom proceeds in a straight line, and the field spawned by the identification of leptin and other genes that cause obesity is no exception.

This year is likely to mark the beginning of a new chapter in the biology of leptin as last February, 20 years after its discovery, leptin is now an FDA-approved human therapeutic for the treatment of severe lipodystrophy with potential for the treatment of other disorders http://www.fda.gov/newsevents/newsroom/pressannouncements/ucm387060.htm. In addition, though leptin’s utility as a monotherapy for obesity appears limited, other evidence suggests that it still has potential as part of a combination therapy for this disorder.

Thus, there have been surprises and disappointments and the passage of time now provides an opportunity to chronicle, as this timely volume does, some of what has been learned, what was surprising and where the field is headed. Some of the key lessons from the last 20 years of research and their context are summarized below.

**Wiring diagram of a complex behavior**

One can describe the phenotype of ob and db mice in several different ways. Historically, these animals have been described as obesity mutations, but one could also think of these animals as showing a behavioral phenotype. ob and db mice show abnormalities in numerous behaviors (Bray 1991). They are massively hyperphagic, show a dramatic decrease in locomotor activity, are quite gentle and non-aggressive, and are not sexually active. Thus, the identification of leptin and later the localization of the leptin receptor (Tartaglia et al. 1995, Lee et al. 1996), encoded by the db gene, have provided an entry point for delving into the neural mechanisms that control complex behaviors. Moreover, the elucidation of the pathogenesis of the obesity resulting from the yellow agouti (Ay) mutation has identified hypothalamic neurons expressing POMC, the precursor of αMSH, as a key neural target of leptin, and more generally, integrators of numerous metabolic signals (Lu et al. 1994). αMSH acts on the MC4 receptor, a GPCR, and MC4R mutations replicate the obese phenotype of Ay mice (Huszar et al. 1997). The subsequent identification of AGRP as an endogenous inhibitor in a second group of hypothalamic neurons, also expressing NPY, added another population of leptin-responsive neurons (Stephens et al. 1995, Ollmann et al. 1997). We now know that leptin acts in part by activating POMC neurons and inhibiting NPY/AGRP neurons though clearly many other neural populations also play a role either as direct neural targets or downstream of these neurons (Friedman 1997). Indeed, enormous progress has been made in defining a set of overlapping neural circuits that control food intake and body weight. With time, these findings are likely to lead to a deeper understanding
of how feeding behavior is controlled as well as our understanding of the control of other behaviors.

Thus, leptin’s neurobiologic effects are not limited to feeding circuits. Extreme weight loss in human has been shown to induce a set of emotional sequela including depression. A possible role for a reduction in leptin in mediating some portion of this was suggested by the finding that leptin injection into the hippocampus can improve the performance of animals in a forced swim test (Lu et al. 2006). This assay provides a quantitative indication of the level of depression in animals and robustly predicts the efficacy of anti-depressant drugs in human. Other studies have shown that leptin has significant effects on reward processing by dopaminergic centers in the midbrain and that it can reduce the value of a sucrose reward (Domingos et al. 2011). This is important because it shows that the pleasure we derive from eating is not fixed but rather reflects the status of metabolic signals such as leptin. Leptin also has potent effects on many other neural circuits including those controlling hormones that regulate reproduction and reproductive behaviors, activity, thermoregulation, and stress (Lu et al. 2006, Wu et al. 2009, Atasoy et al. 2012).

**Obesity has a substantial genetic component**

The identification of mutant genes that cause obesity in mice provided a molecular framework for identifying mutant genes that cause obesity in human. Thus, mutations in leptin, the leptin receptor (LepR), the MC4R as well as PCSK1, and enzymes required for the processing of POMC cause human obesity as do other components of the neural circuit that regulates food intake including brain-derived neurotrophic factor (BDNF) and Sim1. Indeed, it now appears that >10% of morbid human obesity is a result of Mendelian defects in these (and other) genes, which in the majority of cases are in MC4R and LepR (Barsh et al. 2000). This is a level of Mendelian inheritance that exceeds that for nearly every other complex trait that has been studied. The realization that obesity is often the result of genetic mutations in human provides strong evidence that this condition is a result of alterations in a neural circuit that controls the basic drive to eat as well as metabolism (and perhaps other behaviors) and provides an alternative to the widely held view that obesity develops from a failure of willpower or consequent to the modern environment.

Furthermore, it is interesting that all of the obesity genes identified thus far are expressed in the brain. This is despite the fact that there is a large body of evidence indicating that differences in metabolic rate can predict changes in weight (Ravussin et al. 1988) and that an increase in peripheral metabolism such as after treatment with thyroid hormone or uncouplers of respiration such as dinitrophenol leads to weight loss (Grundlingh et al. 2011, Pearce 2012). Moreover, while a defect in leptin signaling is associated with hyperphagia and a marked decrease in energy expenditure in mice, the principal effect in human is on appetite with little or no discernible effect on metabolism (Farooqi et al. 1999). However, while leptin does not appear to cause a net increase in energy expenditure, it does blunt the reduction of energy expenditure that is normally associated with weight loss (Farooqi et al. 2002, Galgani et al. 2010).

The hereditability of obesity has been reported to be between 0.7 and 0.8, which is higher than that for most other traits (Stunkard et al. 1990). That there is a substantial genetic contribution to obesity is also supported by adoption and familial aggregation studies (Stunkard et al. 1986, Adams et al. 1993). However, while some fraction of obesity can be attributed to the aforementioned Mendelian defects as well as variation in genes identified in GWAS studies such as FTO, there is a reason to expect that many new genes remain to be discovered (Fawcett & Barroso 2010). The use of high-throughput genomic sequencing to look for variation in patients with extreme phenotypes, as pioneered by several authors in this volume, is likely to lead to the identification of new genes (Ku et al. 2011). It will be of particular interest to learn whether these new genes also function in the neural circuit that is modulated by leptin and other metabolic signals. It is thus quite likely that future volumes on leptin will need to take into account additional as yet unidentified components of the neural circuit that regulates weight that are identified genetic analyses of human patients.

**Leptin deficiency syndromes**

Leptin-deficient ob mice show abnormalities in most, perhaps all, physiologic systems (Bray 1991). Thus, these animals show defects in the entire neuroendocrine axis and are infertile or subfertile, euthyroid sick with markedly increased corticosterone levels. In addition to these global effects on the neuroendocrine axis, ob mice are hypothermic, diabetic and have profoundly abnormal immune and hematologic function. Indeed, after they were first identified, the complex phenotype of these animals led some to question whether the identification of the ob gene would advance our understanding of how food
intake and body weight are regulated. In retrospect, the complex phenotype of these mice can be most easily understood by noting that the abnormalities they manifest are generally associated not with obesity, but rather starvation (Lord 1998, Frisch 2002). This prediction was supported by the observation that leptin administration suppresses the neuroendocrine response to fasting in mice (Ahima et al. 1996). These findings and others suggest that a key function of leptin is to communicate information to the brain and other organs that there are adequate fat stores and that the organism is not starving. In the absence of leptin, or with the reduced levels seen after fasting, a set of physiologic responses are elicited the aggregate effect of which is to reduce energy expenditure, at the same time as appetite is stimulated.

In addition to its intrinsic importance, this aspect of leptin function provides a framework for understanding the efficacy for treating a series of leptin-deficient states in human. In each case, leptin treatment improves one or more abnormalities generally associated with starvation. As outlined in this volume, lipodystrophy, the complete or partial absence of fat, is a heterogeneous disorder associated with leptin deficiency and a severe sometimes intractable insulin resistance and diabetes as well as hyperlipidemia and nonalcoholic steatohepatitis (NASH) (Ozata et al. 1999). Similarly, as also discussed in this volume, the leanness of young women who often exercise with a great avidity is also associated with leptin deficiency and hypothalamic amenorrhea (Welt et al. 2004). This condition is characterized by a failure to menstruate, infertility and also osteoporosis (Sienkiewicz et al. 2011). Leptin-replacement therapy improves the abnormalities associated with lipodystrophy and hypothalamic amenorrhea (HA) and even causes a significant improvement of bone mineral density in HA patients. Leptin confers these beneficial effects despite causing weight loss in treated patients. Similarly, patients with mutations also show extreme weight loss after leptin therapy but also show improvements in this same set of abnormalities, more typically associated with starvation. Thus, these patients show marked improvements in their metabolic profile, a restoration of fertility and improvements in immune function with leptin treatment. In aggregate, these data strongly suggest that organismal sensing of overall nutritional state (i.e. adipose tissue mass) is conveyed by leptin and not by the actual amount of fat stored in adipose tissue.

In general, the more extreme the abnormalities of patients with low leptin levels, the more significant the clinical response to leptin therapy. This raises the possibility that leptin might have potential as a treatment for other pathologies that develop in settings of leptin deficiency. For example, one female leptin-deficient patient failed to enter puberty in adolescence even though her bone age indicated they should have and leptin treatment led to the onset of menses. This suggests that leptin might be used to induce puberty in very lean young women with a delayed onset of puberty (Farooqi et al. 1999). Both leptin-deficient and starved individuals show immune abnormalities with a shift from TH1 to TH2 immunity and an increased susceptibility to infectious disease. Here again, leptin treatment of leptin-deficient humans and starved animals reverses these changes (Ahima et al. 1996, Lord 1998, Ozata et al. 1999). Thus, it is possible that leptin could be used as an immune adjuvant in settings of extreme cachexia such as starvation, cancer, or chronic inflammatory disease. It has even been proposed that leptin might be useful in patients with end-stage anorexia nervosa with the hope that low-dose leptin treatment could ameliorate some of the pathologies associated with leptin deficiency without significantly reducing food intake (further) and/or as an adjunct to parenteral nutrition (C Montzoros, personal communication).

Leptin might also be of benefit in patients who do not manifest signs or symptoms of pathologic deficiency of leptin (i.e., starvation) but who nonetheless are leptin sensitive (this would be in contrast to most obese patients who are leptin resistant, see below). Prior studies in animals have shown that leptin stimulates glucose metabolism in WT mice independent of weight loss and that it can improve the diabetes of lipodystrophic mice independent of insulin (Kamohara et al 1997, Asilmaz et al. 2004). This raised the possibility that leptin might show efficacy for the treatment of type 1 diabetes. This possibility has now been tested in streptozotocin-treated mice who are either partial or completely insulin deficient. In both cases, leptin markedly lowered blood glucose. Indeed in one study, untreated animals all died within one month, while treated animals survived as long as leptin continued to be expressed from an adenoviral vector (Wang et al. 2010).

Further evidence has suggested that leptin elicits its anti-diabetic effects by inhibiting glucagon. This has raised the possibility that leptin might also be of benefit for patients with type 1 diabetes who often present with weight loss and hyperphagia as a consequence of complete or partial insulin deficiency. In this setting, leptin could either alleviate the demands on β cells at the onset of the disease to extend the ‘honeymoon’ period and/or be used to supplement insulin at later stages of the disease as a means for smoothing glucose control with less hyperglycemia. Leptin therapy might also minimize the weight gain that is
associated with increased doses of insulin. Further studies will be necessary to evaluate these possibilities.

**Leptin resistance**

Physiologic increases in plasma leptin level in WT mice lead to a dose-dependent reduction of food intake and loss of weight (Halaas et al. 1997). While leptin has potent effects to reduce food intake and body weight in ob and WT animals, its efficacy in obese animals is variable and generally reduced (Halaas et al. 1997). Animals with mutations in the leptin receptor fail to respond to leptin treatment at all as do Ay mice that have a defect in melanocortin signaling. Diet-induced animals show only a small response, while New Zealand obese mice (NZO), a strain that develops a polygenic form of obesity, fail to respond to leptin delivered peripherally but lose significant amounts of weight when leptin is delivered through intracerebroventricular (i.c.v.) administration. Each of these strains has high plasma levels of leptin suggesting that they are leptin resistant. The most extreme case of leptin resistance is the db mouse which has a mutation in the leptin receptor (Tartaglia et al. 1995, Lee et al. 1996). In the absence of leptin action, these animals become obese and secondarily overproduce the hormone. Thus, obesity satisfies the hallmarks of a hormone resistance syndrome, with an attenuated response to exogenously administered hormone and elevated endogenous levels (Maffei et al. 1995). In addition, mutations in genes in the leptin signal transduction pathway, PTP1B and SOCS3, increase leptin signaling and lead to a resistance to obesity identifying potential biochemical mechanisms (Bjorbaek et al. 2000, Bence et al. 2006). However, leptin resistance is complex and can develop at many points in the neural circuit that regulates feeding. Thus, leptin resistance can also develop downstream of leptin target neurons as in Ay and MC4R knockout mice in which melanocortin signaling from POMC neurons is abrogated (Lu et al. 1994). Similar to other hormones, leptin resistance can also develop in response to chronically elevated hormone levels via tachyphylaxis (Knight et al. 2010). Finally, as mentioned, in NZO mice, leptin resistance can develop because of impaired leptin transport although little is known about the transcytotic mechanism (Schwartz et al. 1996).

In human, leptin is highly potent in patients with low endogenous levels though its effects in otherwise normal lean patients have never been comprehensively studied (Farooqi et al. 1999, Oral et al. 2002, Welt et al. 2004, Sienkiewicz et al. 2011). In contrast, leptin has variable effects as monotherapy for obesity in the general population. Initial studies showed encouraging effects at very high doses (0.3 mg/kg bid), but this dose was too high for general usage and a lower dose (0.1 mg/kg bid) did not show efficacy (Heymsfield et al. 1999). However, a more recent study treating obese patients with an even lower dose (0.05 mg/kg) led to ~5% weight loss equivalent with efficacy equivalent to other pharmacotherapies for obesity (Roth et al. 2008). It is thus possible that higher doses of leptin led to tachyphylaxis and that a larger study of patients treated with leptin at 0.05 mg/kg or lower could replicate the weight loss observed in the earlier study. There is also evidence that some obese patients show a greater response to leptin than others. In light of the potency of leptin in patients with low endogenous leptin levels, it is possible that the one could enrich for a responder subset by selecting obese patients with low leptin levels. Indeed, while leptin level is highly correlated with adipose tissue mass (r=0.9), plasma leptin can still vary by tenfold or more among patients of the same BMI (Maffei et al. 1995). Furthermore, mice with low levels of leptin are obese and remain leptin sensitive, while patients with heterozygous leptin mutations are obese with low leptin levels (Ioffe et al. 1998, Farooqi et al. 2001).

The efficacy of leptin for the treatment of obesity has been augmented by combining it with other agents that cause weight loss, in particular short-term signals including intestinal peptides that modulate meal pattern. For example, both leptin (0.05 mg/kg) and amylin (pramlintide), a pancreatic peptide that is approved for the treatment of diabetes, each caused ~5% weight loss in a selected group of patients. However, when the two agents were combined, a synergistic effect was observed with an average weight loss of 13% which is significant efficacy for an anti-obesity agent (Roth et al. 2008). Studies in animals also showed that pre-treatment of diet-induced obese animals with amylin restored leptin’s ability to phosphorylate Stat3 in the hypothalamus, suggesting that this gut peptide might reduce the activity of neural circuits that cause leptin resistance (Turek et al. 2010). In animals, leptin’s efficacy has also been augmented in combinations with other peptides or hormones raising the possibility that in time it could emerge as part of a combination therapy for obesity (Muller et al. 2012). Bariatric surgery is an alternative means for inducing weight loss and, while invasive, can be extremely effective. Leptin falls after this procedure, in proportion to the amount of the weight loss, and it is also possible that leptin treatment in this setting could reduce recidivism and/or mitigate some of the sequelae of the procedure that might be secondary to the relative leptin deficiency that develops (Rubino et al. 2004).
Questions for the future

While much has been learned, the road ahead is likely to lead to new advances and some surprises. Still some key questions remain unanswered including the aforementioned.

How is leptin transported into the CNS? Previous studies suggested that leptin entered the brain by transport across the vascular endothelium (Banks & Farrell 1999, Banks & Farrell 2003). However, recent evidence indicates that leptin enters the circumventricular organ in the median eminence where it is then taken up by processes from tanycytes and transcytosed into the III ventricle and then circulates through the ventricular system and accesses deeper brain structures (Langlet et al. 2013, Ballard et al. 2014).

What regulates leptin gene expression? Leptin gene expression is correlated with the intracellular lipid content of adipocytes, suggesting that its regulation might be coupled to a lipid-sensing mechanism (Maffeí et al. 1995, Fei et al. 1997). The nature of this putative lipid-sensing mechanism is unknown.

How does leptin control metabolism? What is the role of leptin signaling at peripheral tissues or are most or all of its effects mediated by the CNS? What are the physiologic and cellular mechanisms by which leptin reduces adipose deposits in fat and other tissues? What are the physiologic and cellular mechanisms by which leptin improves glucose metabolism?

Finally, how does leptin modulate a complex motivational behavior? Leptin acts directly on a number of CNS sites to reduce food intake and body weight in animals and humans and provides an entry point to study the control of feeding (Halaas et al. 1997). Feeding is a complex motivational behavior controlled by many inputs including smell, taste, hormonal state, cognitive inputs, etc. Recently, it has been shown that leptin acts in part by regulating hedonic circuitry but the anatomic site(s) responsible for initiating feeding behavior (Domingos et al. 2011). Thus, it is not known how or even when the multiple inputs are processed to formulate a ‘binary’ decision. So perhaps the biggest question is, how do we decide to eat or do not eat? Perhaps, the answer to this timeless question will be part of future volumes on leptin biology.

Declaration of interest

Leptin is now an approved drug. While Rockefeller owns the patent for leptin, I am named as an inventor on the patent for leptin and as per university policy receive a portion of the milestone payments and royalty payments from Astra Zeneca, the company that owns the license to the patents, to Rockefeller University.
Farooqi I, Keogh JM, Kamath S, Jones S, Gibson W, Trussell R, Jeffb S, Lip G & O’Rahilly S 2001 Partial leptin deficiency and human adiposity. Nature 414 34–35. (doi:10.1038/35102112)

Farooqi I, Matarese G, Lord G, Keogh J, Lawrence E, Agwu C, Sanna V, Fawcett K & Barroso I 2010 The genetics of obesity: FTO leads the way. Nature 465 632–635. (doi:10.1038/379652af)

Fawcett K & Barroso I 2010 The genetics of obesity: FTO leads the way. Nature 465 632–635. (doi:10.1038/379652af)

Friedman JM, Leibel RL & Bahary N 1991 Molecular mapping of obesity. Nature 351 726–731. (doi:10.1038/351628a0)

Friedman JM, Schneider BS, Barton DE & Francke U 1989 Level of expression of the mouse cholecystokinin gene during brain and gut development. PNAS 86 5593–5597. (doi:10.1073/pnas.86.17.5593)

Friedman JM, Schneider BS, Barton DE & Francke U 1989 Level of expression and chromosome mapping of the mouse cholecystokinin gene: implications for mutant models of genetic obesity. Genomics 5 463–469. (doi:10.1016/0888-7543(89)90010-4)

Friedman JM, Leibl RL & Bahary N 1991 Molecular mapping of obesity genes. Mammalian Genome 1 130–144. (doi:10.1002/mgg.1670010204)

Frisch R 2002 Female Fertility and the Body Fat Connection. Chicago: The University of Chicago.

Galaneri JE, Greenway FL, Caglayan S, Wong M-L, Licinio J & Ravussin E 2010 Leptin replacement prevents weight loss-induced metabolic adaptation in congenital leptin-deficient patients. Journal of Clinical Endocrinology and Metabolism 95 851–855. (doi:10.1210/jc.2009-1739)

Grundlingh J, Dargan PI, El-Zanjaly M & Wood DM 2011 2,4-Dinitrophenol (DNP): a weight loss agent with significant acute toxicity and risk of death. Journal of Medical Toxicology 7 205–212. (doi:10.1177/1556366110377893)

Halaas JL, Gajiwala KS, Maffei M, Cohen SL, Chait BT, Rabinowitz D, Lallone R, Barsh GS & Friedman JM 1995 Leptin and the regulation of body weight in mice. Science 269 540–544. (doi:10.1126/science.7624777)

Halaas JL, Boozer C, Blair-West J, Fidahusein N, Denton D & Friedman JM 1997 Physiological response to long-term peripheral and central leptin infusion in lean and obese mice. PNAS 94 8878–8883. (doi:10.1073/pnas.94.16.8878)

Hervey GR 1969 Regulation of energy balance. Nature 222 629–631. (doi:10.1038/222629a0)

Heymsfield SB, Greenway FL, Caglayan S, Wong M-L, Licinio J & Ravussin E 2010 Leptin replacement prevents weight loss-induced metabolic adaptation in congenital leptin-deficient patients. Journal of Clinical Endocrinology and Metabolism 95 851–855. (doi:10.1210/jc.2009-1739)

Hu C, Naidoo N & Pawitnan Y 2011 Revisiting Mendelian disorders through exome sequencing. Human Genetics 129 351–370. (doi:10.1007/s00439-011-0964-2)

Langelet F, Levin BE, Luquet S, Mazzone M, Messina A, Dunn-Meynell AA, Ballard E, Lacombe A, Mazur D, Carmeliet P et al. 2013 Tanyctopic VEGF-A boosts blood-hypothalamic barrier plasticity and access of metabolic signals to the arcuate nucleus in response to fasting. Cell Metabolism 17 607–617. (doi:10.1016/j払met.2013.03.004)

Lee GH, Proença RC, Montez JM, Carroll KM, Durvoshadze JG, Lee JJ & Friedman JM 1996 Abnormal splicing of the leptin receptor in diabetic mice. Nature 379 632–635. (doi:10.1038/379652af)

Lord G 1998 Leptin modulates the T-cell immune response and reverses starvation induced immunosuppression. Nature 394 897–901. (doi:10.1038/29795)

Lu D, Willard DB, Patel IR, Kadwell S, Overton L, Kost T, Luther M, Chen W, Woychik RP, Wilkinson WO et al. 1994 Agouti protein is an antagonist of the melanocyte-stimulating-hormone receptor. Nature 371 799–802. (doi:10.1038/371799a0)

LX-Y, Kim C, Frazer A & Zhang W 2006 Leptin: a potential novel antidepressant. PNAS 103 1593–1598. (doi:10.1073/pnas.0508901103)

Maffeì M, Halaas J, Ravussin E, Pratley RE, Lee GH, Zhang Y, Fei H, Kim S, Lallone R, Ranganathan S et al. 1995 Leptin levels in human and rodent: measurement of plasma leptin and ob RNA in obese and weight-reduced subjects. Nature Medicine 1 1155–1164. (doi:10.1038/nm1155)

Moon BJ & Friedman JM 1997 The molecular basis of the obese mutation in ob2J mice. Genomics 42 152–156. (doi:10.1016/j.0968-9565(96)00004-x)

Ollmann MM, Wilson BD, Yang Y, Kerss NA, Chen Y, Gantz I & Barsh GS 1997 Antagonism of central melanocortin receptors as a mechanism of obesity in congenital leptin deficiency. Nature 387 519–523. (doi:10.1038/371799a0)

Oliva EA, Simha V, Ruiz E, Andewelt A, Premkumar A, Snell P, Wagner AJ, DePaoli AM, Reitman ML, Taylor SI et al. 2002 Leptin-replacement therapy for lipodystrophy. New England Journal of Medicine 346 570–578. (doi:10.1056/NEJMoa022437)

Orata M, Ozdemir I & Licinio J 1999 Human leptin deficiency caused by a missense mutation: multiple endocrine defects, decreased sympathetic tone, and immune system dysfunction indicate new targets for leptin action, greater central than peripheral resistance to the effects of leptin, and spontaneous correction of leptin-mediated defects. Journal of Clinical Endocrinology and Metabolism 84 3686–3695. (doi:10.1210/jcem.84.10.5999)

Pearce E 2012 Thyroid hormone and obesity. Current Opinion in Endocrinology, Diabetes, and Obesity 19 408–413. (doi:10.1097/MED.0b013e328355cd6c)

Ravussin E, Lillioja S, Knowler WC, Christin L, Freymond D, Abbott WGH, Boyce V, Howard B & Bogardus C 1988 Reduced rate of energy expenditure as a risk factor for body-weight gain. New England Journal of Medicine 318 467–472. (doi:10.1056/NEJM198802253180802)

 Roth J, Roland B, Cole R, Trevisaksi J, Weyer C, Kada J, Anderson C, Parkes D & Baron A 2008 Leptin responsiveness restored by amylin agonism in diet-induced obesity: evidence from nonclinical and clinical studies. PNAS 105 7257–7262. (doi:10.1073/pnas.0706743105)

Rubino F, Gagner M, Gentileschi P, Kini S, Fukuyama S, Feng J & Diamond E 2004 The early effect of the Roux-en-Y gastric bypass on hormones involved in body weight regulation and glucose metabolism. Annals of Surgery 239 236–242. (doi:10.1016/j.ansu.2003.11.077)

Schwartz MW, Peskind E, Raskind M, Boyko EJ & Porte D Jr 1996 Adiposity in humans. Journal of Clinical Endocrinology and Metabolism 81 1103–1109. (doi:10.1210/jcem.81.4.8674006)

Tanaka M, Matsuzawa Y & Minokoshi Y 2000 Leptin signaling in the liver: role in the control of hepatic glucose metabolism. Journal of Biological Chemistry 275 534–538. (doi:10.1074/jbc.275.1.534)

Thematic Review

J FRIEDMAN

Leptin at 20

Journal of Endocrinology

223:1

T7

Published by Bioscientifica Ltd

http://joe.endocrinology-journals.org

DOI: 10.1530/EJE-14-0405

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Stenkiewicz E, Magkos F, Aronis KN, Brinkoetter M, Chamberland JP, Chou S, Arampatzi KM, Gao C, Koniaris A & Mantzoros CS 2011 Long-term metreleptin treatment increases bone mineral density and content at the lumbar spine of lean hypoleptinemic women. *Metabolism: Clinical and Experimental* **60** 1211–1221. (doi:10.1016/j.metabol.2011.05.016)

Stephens TW, Bashinski M, Bristow PK, Bue-Valleskey JM, Burgett SG, Hale H, Hoffmann J, Hsiung HM, Krauciunas A, Mackellar W *et al.* 1995 The role of neuropeptide Y in the antiobesity action of the obese gene product. *Nature* **377** 530–534. (doi:10.1038/377530a0)

Stunkard AJ, Sorensen T, Hanis C, Teasdale TW, Chakraborty R, Schull WJ & Schulsinger F 1986 An adoption study of human obesity. *New England Journal of Medicine* **314** 193–198. (doi:10.1056/NEJM198601233140401)

Stunkard AJ, Harris JR, Pedersen NL & McClearn GE 1990 The body-mass index of twins who have been reared apart. *New England Journal of Medicine* **322** 1483–1487. (doi:10.1056/NEJM198601233140401)

Tartaglia LA, Dembski M, Weng X, Deng N, Culpepper J, Devos R, Richards GJ, Campfield LA, Clark FJ, Deeds J *et al.* 1995 Identification and expression cloning of a leptin receptor, OB-R. *Cell* **83** 1263–1271. (doi:10.1016/0092-8674(95)90151-5)

Turek V, Tevaskis J, Levin B, Dunn-Meynell A, Irani B, Gu G, Wittmer C, Griffin P, Vu C, Parkes DG *et al.* 2010 Mechanisms of amylin/leptin synergy in rodent models. *Endocrinology* **151** 143–152. (doi:10.1210/en.2009-0546)

Wang M, Chen L, Clark G, Lee Y, Stevens RD, Ilkayeva OR, Wenner BR, Bain JR, Charron MJ, Newgard CB *et al.* 2010 Leptin therapy in insulin-deficient type 1 diabetes. *PNAS* **107** 4813–4819. (doi:10.1073/pnas.0909422107)

Welt CK, Chan JL, Bullen J, Murphy R, Smith P, DePaoli AM, Karalis A & Mantzoros CS 2004 Recombinant human leptin in women with hypothalamic amenorrhea. *New England Journal of Medicine* **351** 987–997. (doi:10.1056/NEJMoA040388)

Wu Q, Boyle MP & Palmiter RD 2009 Loss of GABAergic signaling by AgRP neurons to the parabrachial nucleus leads to starvation. *Cell* **137** 1225–1234. (doi:10.1016/j.cell.2009.04.022)

Zhang Y, Proenca P, Maffei M, Barone M, Leopold L & Friedman JM 1994 Positional cloning of the mouse obese gene and its human homologue. *Nature* **372** 425–432. (doi:10.1038/372425a0)

Received in final form 1 July 2014
Accepted 12 August 2014