Genome Sequences of Zika Virus Strains Recovered from Amniotic Fluid, Placenta, and Fetal Brain of a Microcephaly Patient in Thailand, 2017

Thidathip Wongsurawat,a Piroon Jenjaroenpun,a Niracha Athipanyasilp,b Bualan Kaewnapan,b Nattawat Leelahakorn,b Nasikarn Angkasekwinaic Wannee Kantakamalakul,b Ruengpung Sutthent,b @ David W. Ussery,a,d Navin Horthongkham,b Intawat Nookaewb,d

aDepartment of Biomedical Informatics, College of Medicine, University of Arkansas for Medical Sciences, Little Rock, Arkansas, USA
bDepartment of Microbiology, Faculty of Medicine, Siriraj Hospital, Mahidol University, Bangkoknoi, Bangkok, Thailand
cDepartment of Medicine, Faculty of Medicine, Siriraj Hospital, Mahidol University, Bangkoknoi, Bangkok, Thailand
dDepartment of Physiology and Biophysics, College of Medicine, University of Arkansas for Medical Sciences, Little Rock, Arkansas, USA

ABSTRACT We present here the complete genome sequences of Zika virus strains isolated from aborted fetal tissue (brain and placenta) and amniotic fluid of a microcephaly patient in Thailand in 2017. The virus genomes that were sequenced have an average length of 10,807 nucleotides.

Zika virus (ZIKV) is a linear, single-strand, positive-sense RNA virus of the family Flaviviridae. Due to an unprecedented number of infection outbreaks, ZIKV has become a global concern. The virus commonly spreads through mosquito bites and causes relatively mild symptoms in adults, but it has been shown to cause microcephaly in infants born to infected mothers. ZIKV is now spreading rapidly in Latin America, and in Asia, Thailand and the Philippines have had a high incidence of infection (1). The WHO confirmed the first microcephaly cases in Thailand in 2016, but to date, no genome sequences of ZIKV strains from Thailand have been reported. The genome sequences of ZIKV strains isolated from microcephaly cases in Asia may clarify the possible mechanisms for the transmission and pathogenesis of ZIKV. We report here the genome sequences of ZIKV strains isolated from aborted fetal tissue (brain and placenta) and amniotic fluid samples from a microcephaly patient in Thailand.

The patient was suspected of having ZIKV infection at 9 weeks of gestation. At 16 weeks, amniotic fluid tested positive for ZIKV, and the pregnancy was terminated at 17 weeks. An autopsy of the fetus demonstrated a head circumference of 12.5 cm, which does not reach the third percentile for this gestational age. RNA of ZIKV was detected in the brain and placenta. Viral RNA was extracted using a NucliSens kit (bioMérieux, France), and cDNA was synthesized using PrimeScript (TaKaRa, Japan). The whole-genome sequences of the ZIKV isolates were amplified using PrimeStar GXL (TaKaRa, Japan).

Three sequencing platforms were used for the different ZIKV isolates. The Ion Torrent platform was used for the brain isolate, and the Oxford Nanopore Technologies (ONT), Ion Torrent, and Illumina platforms were used for the amniotic fluid and placenta isolates. The read data generated for the isolates are listed in Table 1.

The Ion Torrent library was constructed using an Ion Xpress Plus fragment library kit (Life Technologies, USA), following the manufacturer’s instructions. The libraries were amplified on the OneTouch version 2 platform (Life Technologies, USA) and sequenced on an IonTorrent platform.
Torrent PGM sequencer using the 316 chip. The Ion Torrent sequencing generated 9.2 million reads with an average read length of 202 bp. Genome assembly was performed using CLC software version 9.0 with default parameters (Qiagen, Germany).

Library preparation for the ONT sequencing followed the one-dimensional genomic DNA sequencing protocol SQK-LSK108 (ONT, USA). Sequencing of the library was performed on a single R9.4/FLO-MIN106 flow cell on the MinION Mk1B system. Base calling was performed using Albacore software version 1.2.3. Last, Illumina sequences were used to obtain high-quality genome sequences.

Library preparation for Illumina sequencing was performed with a KAPA HyperPlus kit (Kapa Biosystems, USA) following the manufacturer’s protocol. The libraries were sequenced using the MiSeq platform (Illumina, USA).

Raw reads were mapped using Burrows-Wheeler alignment with the MEM algorithm (BWA-MEM) and default parameters (2). Duplicates were removed using Picard MarkDuplicates version 2.9.2 with default parameters (https://broadinstitute.github.io/picard), and reads were realigned and consensus called using GATK version 3.7 (RealignerTargetCreator, IndelRealigner, SelectVariants, CombineVariants, and FastaAlternateReferenceMaker) with default parameters, except for SelectVariants, which used a modified command (–select “ABHet < 0.4 && DP > 100”) (3).

The reads were assembled using the ZIKV genome sequence available in GenBank (accession number KY272987; 98% identity) as a reference. The total size of the assembly was 10,807 bp. Genome annotation was performed with ViPR workbench (www.viprbrc.org) (4). Using a whole-genome comparison method (5), we phylogenetically analyzed the three ZIKV genomes in our recent report (6). The tree showed that the three genomes closely resemble the local Asian strains.

**Data availability.** The complete ZIKV genome sequences reported here have been submitted to GenBank and are publicly available under the accession numbers MG548660 (amniotic fluid), MG548661 (placenta), and MF996804 (brain).

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**REFERENCES**

1. Lim S-K, Lim JK, Yoon I-K. 2017. An update on Zika virus in Asia. Infect Chemother 49:91–100. https://doi.org/10.3947/ic.2017.49.2.91.

2. Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows–Wheeler transform. Bioinformatics 25:1754–1760. https://doi.org/10.1093/bioinformatics/btp324.

3. McKenna A, Hanna M, Banks E, Sivachenko A, Cibulskis K, Kernytsky A, Garimella K, Altshuler D, Gabriel S, Daly M, DePristo MA. 2010. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. Genome Res 20:1297–1303. https://doi.org/10.1101/gr.107524.110.

4. Sun G, Larsen CN, Baumgarth N, Klem EB, Scheuermann RH. 2017. Comprehensive annotation of mature peptides and genotypes for Zika virus. PLoS One 12:e0170462. https://doi.org/10.1371/journal.pone.0170462.

5. Jun S-R, Wassenaar TM, Wanchai V, Patumcharoenpol P, Nookaew I, Ussery DW. 2017. Suggested mechanisms for Zika virus causing microcephaly: what do the genomes tell us? BMC Bioinformatics 18:471. https://doi.org/10.1186/s12859-017-1894-3.

6. Wongsurawat T, Athipanyasilp N, Jenjaroenpun P, Jun SR, Kaewnapan B, Wassenaar TM, Leelahakorn N, Angkasekwinai N, Kantakamalakul W, Ussery DW, Sutthent R, Nookaew I, Hortongkham N. 2018. Case of microcephaly after congenital infection with Asian lineage Zika virus, Thailand. Emerg Infect Dis 24:1758. https://doi.org/10.3201/eid2409.180416.

**TABLE 1 Read information generated from three sequencing technologies**

| Sequencing technology | Isolate source | Total no. of bases | No. of reads | Mean length (bp) |
|-----------------------|----------------|-------------------|--------------|-----------------|
| Ion Torrent Brain     | 9,276,530      | 45,701            | 202.98       |
| Illumina Amniotic fluid | 2,019,009,458 | 8,807,177         | 229.25       |
| Ion Torrent Amniotic fluid | 7,531,684     | 38,694            | 194.65       |
| Oxford Nanopore Amniotic fluid | 224,694,525 | 139,132           | 1,614.97     |
| Illumina Placenta     | 1,853,740,947  | 9,189,342         | 201.73       |
| Ion Torrent Placenta  | 5,639,797      | 20,933            | 269.42       |

**WEBаблиц 1 Статистика данных о чтении с трех технологий последовательного поиска**

| Технология последовательного поиска | Источник образца | Общее число байтов | Число чтений | Средняя длина (бп) |
|-----------------------------------|-----------------|-------------------|--------------|--------------------|
| Ion Torrent Brain                  | 9,276,530       | 45,701            | 202.98       |
| Illumina Amniotic fluid           | 2,019,009,458   | 8,807,177         | 229.25       |
| Ion Torrent Amniotic fluid        | 7,531,684       | 38,694            | 194.65       |
| Oxford Nanopore Amniotic fluid    | 224,694,525     | 139,132           | 1,614.97     |
| Illumina Placenta                 | 1,853,740,947   | 9,189,342         | 201.73       |
| Ion Torrent Placenta              | 5,639,797       | 20,933            | 269.42       |