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Review

ACE2 – From the renin–angiotensin system to gut microbiota and malnutrition

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Abstract

The renin–angiotensin system (RAS) is a complex network that regulates blood pressure, electrolyte and fluid homeostasis, as well as the function of several organs. Angiotensin-converting enzyme 2 (ACE2) was identified as an enzyme that negatively regulates the RAS by converting Ang II, the main bioactive molecule of the RAS, to Ang 1–7. Thus, ACE2 counteracts the role of angiotensin-converting enzyme (ACE) which generates Ang II from Ang I. ACE and ACE2 have been implicated in several pathologies such as cardiovascular and renal disease or acute lung injury. In addition, ACE2 has functions independent of the RAS: ACE2 is the receptor for the SARS coronavirus and ACE2 is essential for expression of neutral amino acid transporters in the gut. In this context, ACE2 modulates innate immunity and influences the composition of the gut microbiota, which can explain diarrhea and intestinal inflammation observed in Hartnup disorder, Pellagra, or under conditions of severe malnutrition. Here we review and discuss the diverse functions of ACE2 and its relevance to human pathologies.

1. An overview of the renin–angiotensin system

The renin–angiotensin system (RAS) is a complex network that regulates blood pressure, electrolyte and fluid homeostasis, as well as the function of several organs. Angiotensin-converting enzyme 2 (ACE2) was identified as an enzyme that negatively regulates the RAS by converting Ang II, the main bioactive molecule of the RAS, to Ang 1–7. Thus, ACE2 counteracts the role of angiotensin-converting enzyme (ACE) which generates Ang II from Ang I. ACE and ACE2 have been implicated in several pathologies such as cardiovascular and renal disease or acute lung injury. In addition, ACE2 has functions independent of the RAS: ACE2 is the receptor for the SARS coronavirus and ACE2 is essential for expression of neutral amino acid transporters in the gut. In this context, ACE2 modulates innate immunity and influences the composition of the gut microbiota, which can explain diarrhea and intestinal inflammation observed in Hartnup disorder, Pellagra, or under conditions of severe malnutrition. Here we review and discuss the diverse functions of ACE2 and its relevance to human pathologies.

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pathways, Ang I can be cleaved by chymase to form Ang II or by other peptidases such as prolyl-endopeptidase (PEP), neutral endopeptidase (NEP), and thymet oligopeptidase (TOP), to directly generate Ang 1–7 [10]. However, in this review we will solely focus on the ACE/ACE2 peptidase system. ACE2 can potentially also act on other peptide systems, in particular on the apelin/APJ system [11]. Apelin is produced as a 77 amino acid pre-pro-hormone which is further processed to apelin-36 and apelin-13. Apelin signals through its receptor APJ and was shown to have vasodilatory effects [12]. ACE2 can cleave the carboxyterminal phenylalanine of apelin-36 and apelin-13. Apelin signals through its receptor API and was shown to have vasodilatory effects [11]. ACE2 can cleave the carboxyterminal phenylalanine of Ang II to produce the vasodilator Ang 1–7 [14,15]. ACE2 also shows activity towards Ang I by cleaving the C-terminal leucine, which results in the supposedly biological inactive peptide Ang 1–9, which can further be processed by ACE to Ang 1–7. However, ACE2 shows a clear substrate preference for Ang II over Ang I [11].

Fig. 1. Simplified diagram of the renin—angiotensin system. Angiotensinogen is secreted by the liver and gets converted to Ang I by renin, which is mainly produced in the kidneys. Ang I gets cleaved by ACE to result in Ang II. Ang II is a ligand for the AT1 receptor and promotes vasoconstriction and hypertension. Alternatively, Ang II can bind to the AT2 receptor to inhibit vasoconstriction. ACE2 processes Ang II to Ang 1–7 which binds to the Mas receptor to induce vasodilation.

2. Angiotensin-converting enzyme 2 — ACE2

ACE2 was discovered as a homolog of ACE and mapped to the X chromosome in humans, rats, and mice [14–16]. ACE2 is expressed at high levels in kidney, heart, and testis [14], but is also found in many other tissues such as lung, small intestine, and liver [17–19]. The ace2 gene spans 18 exons and codes for an 805 amino acid type I transmembrane glycoprotein. ACE2 contains a short intracellular cytoplasmic tail and a longer extracellular domain that exhibits carboxy- monopeptidase activity [1]. The active site of ACE2 contains the HEMGH motif, characteristic for zinc-metallopeptidases and shares approximately 42% sequence homology with the amino-terminal domain of ACE [15]. The carboxy-terminal domain of ACE2 is about 48% homologous with Collectrin (also known as Tmem27) [20]. Therefore, evolutionary, ACE2 is a chimeric protein consisting of the amino-terminal carboxypeptidase domain of ACE, whereas the carboxy-terminal part of ACE2 lacking enzymatic activity shares homology with Collectrin. Another homolog of ACE was discovered in 2007 and termed ACE3 [21]. However, due to the presence of several deletions and insertions in the genomic sequence, ACE3 seems to lack catalytic activity as a metalloprotease. So far no physiologic functions could be identified and it has been suggested that ace3 is a pseudogene [21,22].

3. Physiologic functions of ACE2 within the RAS

The carboxy-peptidase domain of ACE2 efficiently cleaves the C-terminal phenylalanine of Ang II to produce the vasodilator Ang 1–7 [14,15]. ACE2 also shows activity towards Ang I by cleaving the C-terminal leucine, which results in the supposedly biological inactive peptide Ang 1–9, which can further be processed by ACE to Ang 1–7. However, ACE2 shows a clear substrate preference for Ang II over Ang I [11]. Solving the crystal structure of ACE2 revealed a hinge-bending motion upon ligand binding, characteristic for several metallopeptidases. Moreover, compared to ACE a
smaller and therefore less accommodating site within the active domain of ACE2 can explain the release of a single amino acid by ACE2 in contrast to ACE which cleaves off a dipeptide [23].

In the cardiovascular system, ACE and ACE2 are thought to be important players in blood pressure regulation through balancing the levels of the vasoconstrictor Ang II. Raised Ang II levels lead to increased blood pressure, therefore, inhibitors of ACE or antagonists of AT1 are highly effective treatment options for hypertension. Furthermore, it was reported that the ace2 gene maps to a quantitative trait locus on the X chromosome in three different rat models of hypertension and that hypertensive rats exhibit reduced ACE2 expression compared to wild type animals [16]. However, whereas targeted deletion of ACE in mice resulted in spontaneous hypertension [24], ACE2 knock-out mice showed no significant alterations in blood pressure at baseline [16,25]. Upon chronic Ang II infusion, however as expected, ACE2 deficient mice do exhibit significantly increased blood pressure compared to control animals [25]. In addition, cardiac overexpression of ACE2 in rats was found to mediate resistance against the deleterious effects of hypertension and Ang II infusion [26], and vascular ACE2 overexpression resulted in decreased blood pressure in another study using rats [27]. Thus, ACE2 acts as a negative regulator of the RAS and has an antihypertensive role in blood pressure control.

ACE2 is expressed at substantial levels in cardiomyocytes in the heart. Gene deletion of ace2 resulted in impaired cardiac contractility in two different independent targeted mouse lines [16,28]. The observed impaired heart function was associated with increased Ang II levels in heart and in the plasma. Indeed, double knock-out of ACE and ACE2 as well as treatment with AT1 antagonists rescued the cardiac malfunctions of ACE2 knock-out animals [16,28,29]. Of note, in a third ACE2 deficient mouse line aberrant cardiac contractility could not be observed [25], which turned out to be due to different mouse line backgrounds [30]. Importantly, in all mouse models generated, ACE2 protects from heart failure. In line with a beneficial effect of ACE2, treatment with Ang 1–7 has been shown to improve myocardial performance, cardiac remodeling, and survival in rodent heart failure models, including ischemia/reperfusion injury, myocardial infarction, or hypertension-induced cardiomyopathy [31,32]. Of note, besides the beneficial effects of ACE2 in cardiovascular diseases, overexpression of ACE2 can have deleterious effects resulting in cardiac fibrosis and arrhythmia [33,34]; whether this is physiologically relevant needs to be examined in future experiments.

In the lung, increased Ang II levels were shown to promote development of pulmonary hypertension [35] and pulmonary fibrosis [36] and ACE2 was reported to protect both from pulmonary hypertension and pulmonary fibrosis [37–39]. Moreover, ACE inhibitors and AT1 antagonists can attenuate the severity of experimentally induced pulmonary fibrosis [40,41], corroborating the role of Ang II in this condition. Furthermore the RAS is also involved in acute lung injury and its severest form acute respiratory distress syndrome (ARDS) [42,43]. ACE2 deficient animals develop histologically and functionally normal lungs, but upon experimentally induced acute lung injury they develop severe ARDS [44]. Importantly, ACE and ACE2 double knock-out, treatment with AT1 antagonists, or administration of recombinant ACE2 protein rescued the severe acute lung injury phenotype of ACE2 knock-out mice [44]. Therefore, therapy with recombinant ACE2 protein has the potential to be effective against acute lung injury as well as chronic lung diseases such as pulmonary hypertension or pulmonary fibrosis. Based on these data, recombinant soluble ACE2 has been developed as a possible future therapy to treat acute lung injury in humans and is currently being tested in phase 2 clinical trials.

In the kidney, ACE and ACE2 are expressed in the brush border of the proximal tubule epithelial cells and contribute to salt and fluid homeostasis as well as blood pressure control [45]. Besides systemic consequences of kidney fluid homeostasis, there is growing evidence that ACE2 protects from kidney disease by reducing local Ang II levels [46]. In this regard, ACE2 deficient mice develop late-onset nephrotic glomerulosclerosis [47] and display more severe diabetic kidney injury in experimental diabetic mice [48,49]. These phenotypes emerged via Ang II because AT1 blockers or ACE inhibitors reversed the deleterious phenotypes [47–49]. Despite this data, the renoprotective role of ACE2 is still controversial because under certain experimental conditions ACE2 and Ang 1–7 were shown to promote renal injury [50]. Therefore, the precise role of ACE2 in the kidney awaits additional experiments and needs to be translated to human disease.

### 4. ACE2 is the SARS-coronavirus receptor

Interestingly, ACE2 also plays a major role in infections with severe acute respiratory syndrome corona-virus (SARS-CoV). This virus first emerged in 2003 and caused respiratory disease which could trigger ARDS and often ended fatal [51]. In the same year ACE2 was identified to be a potential receptor for the SARS-CoV in vitro [52], which was thereafter genetically confirmed in in vivo mouse studies [17], providing definitive evidence that ACE2 is the essential SARS receptor in vivo.

SARS infections are initiated by binding of spike protein trimers of the SARS-CoV to a hydrophobic pocket of the extracellular catalytic domain of ACE2 [53]. This interaction enables endocytosis, membrane fusion, and entry of the SARS-CoV into the host cell. Following virus entry, the ACE2 protein is downregulated which leads to local increase of Ang II levels, providing a molecular explanation for the frequent development of ARDS during SARS-CoV infections [17,44], which was untypical for other coronaviruses. The significance of elevated Ang II in SARS pathogenesis was further substantiated by the fact that SARS-CoV infections of ACE2 knock-out mice do not induce lung injury, and recombinant ACE2 rescues from experimental lung injury upon SARS-CoV spike protein treatment and other triggers of lung injury [17]. Thus, the SARS-CoV virus might have shown us a novel
treatment option for a previously untreatable disease, i.e. the use of ACE2 for the treatment of acute lung injury.

It should be noted that within the family of coronaviruses only SARS-CoV and human coronavirus NL63 were shown to utilize ACE2 for cell entry [54]. In 2012, a novel coronavirus was isolated from a patient in Saudi Arabia [55]. This virus was later termed Middle East respiratory syndrome Corona Virus (MERS-CoV), and currently has an alarming mortality rate of ~60% [56]. Due to the high similarity of SARS-CoV and MERS-CoV, it was suggested that they both might enter the host via ACE2; however, recent findings have shown that MERS-CoV does not utilize ACE2 as a receptor [57], but enters the host cell via dipeptidyl peptidase 4 (DPP4 or CD26) [58].

5. ACE2 regulates amino acid transport in the intestine

Collectrin was discovered in 2001 as a protein expressed in the kidney with high sequence homology to the carboxy-terminal end of ACE2 [20]. Collectrin and ACE2 both are type I transmembrane proteins [59], but, in contrast to ACE2, Collectrin lacks catalytic activity. The collectrin gene is located on the X chromosome immediately upstream of the ace2 gene in all species examined and both genes share similar transcription factor binding sites [60]. In this regard, collectrin was shown to be under transcriptional control of hepatocyte nuclear factor α (HNF-1α) [59,61], which is involved in pancreatic insulin secretion [62]. In vitro studies and in vivo overexpression experiments supported a role for Collectrin in insulin exocytosis through interaction with the SNARE complex [61]. However, Collectrin deficient mice did not exhibit impaired insulin secretion [63].

Surprisingly, analysis of Collectrin knock-out mice revealed that Collectrin is a regulator of neutral amino acid transporters expression on brush border membranes of renal proximal tubules [64]. In this study it was observed that urine of Collectrin deficient mice forms crystals when stored at 4 °C, which turned out to be mainly composed of tyrosine and phenylalanine [64]. Additional studies showed that Collectrin non-covalently associates with the Slc6 family of neutral amino acid transporters (B⁰AT1 (Slc6a19) and B⁰AT3 (Slc6a18)), the imino transporter SIT1 (Slc6a20) and the Slc1 glutamate and aspartate transporter EAAT3 (Slc1a1) [65]. Collectrin stabilizes amino acid transporter expression on the cell surface but does not affect mRNA expression of these transporters [64,66]. These studies uncovered Collectrin as a key subunit of these amino acid transporters which controls polarized expression in the kidney required for renal reabsorption of amino acids.

B⁰AT1 is not only expressed in the kidney but also in the small intestine where Collectrin is absent. Interestingly, similar to the role of Collectrin in the kidney, we were able to show that the closest “relative” of Collectrin, i.e. ACE2, can bind and stabilize the neutral amino acid transporter B⁰AT1 in the small intestine [67]. Therefore, B⁰AT1 seems to rely on tissue specific interaction partners. In ACE2 deficient mice, B⁰AT1 is completely absent from the small intestine, but is expressed at normal levels in the kidney [67]. On the contrary, in Collectrin knock-out mice B⁰AT1 expression is lost from the kidney despite the presence of ACE2 [64]. B⁰AT1 and ACE2 colocalize on enterocytes of the small intestine, an interaction confirmed by co-immunoprecipitation experiments and overexpression in Xenopus laevis oocytes [64,67,68]. In the gut, ACE2 also interacts with SIT1 [65,69], a transporter for proline, sarcosine, or betaine [70]. Interestingly, the enzymatic activity of ACE2 is not required for B⁰AT1 surface expression or transporter function [67]. B⁰AT1 is dispensable for expression of ACE2 or Collectrin [71].

Mutations in slc6a19 encoding B⁰AT1 have been identified to cause Hartnup disorder resulting in defective amino acid uptake in the kidney and small intestine [72,73]. The main characteristic of Hartnup disorder is neutral aminoaciduria similar to the phenotype observed in Collectrin deficient mice [64]. Apart from aminoaciduria, patients with Hartnup disorder are often symptom free, but under stress conditions like malnutrition or infections develop pellagra-like symptoms such as light-sensitive rash, cerebellar ataxia, emotional instability, as well as diarrhea. More than twenty different mutations in slc6a19 have been identified so far in Hartnup disorder patients [65]. However no mutations in ace2 or collectrin have been reported in Hartnup disorder or aminoaciduria syndromes.

The aminoaciduria in Hartnup disorder is clearly caused by the impaired amino acid uptake in the kidney and small intestine, but how a defect in B⁰AT1 causes dermatitis or neurological phenotypes remains elusive. Our recent studies unraveled a novel connection between intestinal amino acid uptake and susceptibility to diarrhea and colitis [18], conditions also frequently observed in Hartnup disorder patients [74]. We assayed ACE2 deficient mice which cannot express B⁰AT1 in the small intestine and therefore exhibit dramatically reduced plasma levels of the essential amino acid tryptophan. Whereas the baseline intestinal architecture appears normal in ACE2 knock-out mice, treatment with chemical irritants revealed that ACE2 deficient mice are highly susceptible to experimentally induced colitis, while control mice only developed modest intestinal inflammation [18]. A direct involvement of the essential amino acid tryptophan was proven by administering a tryptophan free diet, which made wild-type mice highly susceptible to chemically induced colitis. Strikingly, in the reverse experiment the colitis phenotype could be rescued in ACE2 deficient mice by supplementing the diet with a glycine—tryptophan dipeptide which can be taken up by the proton-coupled peptide transporter PepT1 [75] and, therefore, circumvent the missing B⁰AT1 transporter system. Moreover, we could rescue the colitis phenotype of ACE2 mutant mice using nicotinamide, a downstream metabolite of tryptophan. Consistent with these results, nicotinamide has been used as a treatment for the vitamin B3 deficiency disease pellagra [74]. Collectrin deficient mice cannot efficiently reabsorb tryptophan from urine, but do not exhibit increased susceptibility to colitis. Therefore, only defective intestinal tryptophan uptake in ACE2 knock-out mice seems to mediate local intestinal inflammation and diarrhea [18].
ACE2 is highly expressed in the small intestine but hardly detectable in the colon. How then does the impaired amino acid uptake in the small intestine explain the colitis phenotype? This contradiction was explained by a chain of events starting with the observation that reduced tryptophan levels lead to reduced mTOR pathway activity in the small intestine [18]. Aberrant mTOR activation resulted in impaired expression of antimicrobial peptides from small intestinal Paneth cells. The impaired expression of antimicrobial peptides in turn resulted in an altered composition of the intestinal microbiota (Fig. 2). That an alteration in the intestinal microbiome mediated the increased susceptibility to colitis was finally proven by microbiota transplantation experiments. These results also provide an explanation for the effectiveness of nicotinamide treatment of pellagra patients [18,74]. It also provides an explanation for how protein malnutrition can lead to severe intestinal inflammation. Hundreds of millions of people worldwide are affected by malnutrition [76]. During famines acute diarrheal infection and intestinal inflammation are often the actual cause of death [77]. Furthermore, the observation that a reduction of mTOR activity leads to susceptibility of intestinal inflammation could provide an explanation why inhibition of mTOR with rapamycin was not successful in human clinical trials in inflammatory bowel disease [78].

Interestingly, following our initial experiments in mice, a connection between the microbiota and Kwashiorkor, a form of severe acute malnutrition, has recently been reported in humans [79]. The study was performed with twin pairs up to an age of three years in rural Malawi. Analysis of fecal microbiota revealed that gene diversity of the microbiota from children with Kwashiorkor did not develop with increasing age — in contrast to the microbiota of healthy children. To confirm a causal role of the microbiota, fecal content was transplanted to gnotobiotic mice. Only the microbiota from Kwashiorkor-affected children in combination with a Malawian diet, which lacks protein, lead to severe weight loss in recipient mice. Similarly, upon feeding a protein free diet or a tryptophan free diet to mice, we observed significant changes in the composition of gut microbiota resulting in colitis and weight loss [18]. Both studies show an intricate interaction between diets, the intestinal microbiota, and malnutrition [18,79].

In another large study, Malawian children with uncomplicated severe acute malnutrition received ready-to-use therapeutic food with or without combined antibiotic treatment. Administration of antibiotics led to reduction of dehydrating diarrhea and also significantly lowered the mortality rates [80]. This data could implicate that antibiotic treatment protects better against fatal infections or it could point towards a critical role of the patients’ microbiota in the progression of diarrhea and severe acute malnutrition. In this regard, antibiotic treatment rescued bloody diarrhea in a colitis model of ACE2 deficient mice [18].
6. Conclusions

With the discovery of ACE2, a homolog of the classic enzyme ACE, an important negative regulator of the RAS was identified. The enzymatic activity of ACE2 reduces Ang II levels and thereby counteracts the ACE/Ang II/AT1 axis. In the context of the RAS, ACE2 has vasodilator effects, regulates heart functions, exhibits a renoprotective role, counteracts fibrotic changes, and can protect from ARDS in the lung. Intriguingly, ACE2 has functions that are independent of the RAS and independent of the catalytic activity of ACE2. In this regard, ACE2 was identified as the receptor of the SARS coronavirus and recently it was discovered that ACE2 is essential for the expression of certain amino acid transporters in the small intestine. Via regulation of intestinal amino acid transport, ACE2 couples diet to the composition of the gut microbiome, also providing a molecular explanation how malnutrition in hundreds of millions of people could cause intestinal inflammation and diarrhea. How this function is linked to the regulation of the RAS needs to be further explored. Since the intestinal microbiota has been shown to influence host physiology and numerous conditions from obesity to autoimmune diseases [81], unraveling the complex network of interactions between host factors and their microbiota has the potential to uncover new therapeutic approaches.

References

[1] D.W. Lambert, N.E. Clarke, A.J. Turner, Not just angiotensinases: new roles for the angiotensin-converting enzymes, Cell Mol. Life Sci. 67 (2010) 89–98.
[2] K. Kuba, Y. Imai, J.M. Penninger, Multiple functions of angiotensin-converting enzyme 2 and its relevance in cardiovascular diseases, Circ. J. 77 (2013) 301–308.
[3] K. Kuba, Y. Imai, T. Ohto-Nakanishi, J.M. Penninger, Trigon of ACE2: a peptidase in the renin–angiotensin system, a SARS receptor, and a partner for amino acid transporters, Pharmacol. Ther. 128 (2010) 119–128.
[4] Y. Imai, K. Kuba, T. Ohto-Nakanishi, J.M. Penninger, Angiotensin-converting enzyme 2 (ACE2) in disease pathogenesis, Circ. J. 74 (2010) 405–410.
[5] R.M. Carey, S.H. Padia, Angiotensin AT2 receptors: control of renal sodium excretion and blood pressure, Trends Endocrinol. Metab. 19 (2008) 84–87.
[6] R.A. Santos, A.C. Simoes e Silva, C. Maric, D.M. Silva, R.P. Machado, I. de Buhr, S. Heringer-Walther, S.V. Pinheiro, M.T. Lopes, M. Bader, E.P. Mendes, V.S. Lemos, M.J. Campagnole-Santos, H.P. Schultheiss, R. Speth, T. Walther, Angiotensin-(1–7) is an endogenous ligand for the G protein-coupled receptor Mas, Proc. Natl. Acad. Sci. U S A 100 (2003) 8258–8263.
[7] R.A. Santos, A.J. Ferreira, T. Verano-Braga, M. Bader, Angiotensin-converting enzyme 2, angiotensin-(1–7) and Mas: new players of the renin–angiotensin system, J. Endocrinol. 216 (2013) R1–R17.
[8] M.M. Gironacci, M.P. Coba, C. Penca, Angiotensin-(1–7) binds at the type 1 angiotensin II receptors in rat renal cortex, Regul. Pept. 84 (1999) 51–54.
[9] B.P. Rowe, D.L. Saylor, R.C. Spreth, D.R. Absher, Angiotensin-(1–7) binding at angiotensin II receptors in the rat brain, Regul. Pept. 56 (1995) 139–146.
[10] D.G. Passos-Silva, T. Verano-Braga, R.A. Santos, Angiotensin-(1–7): beyond the cardio-renal actions, Clin. Sci. (Lond) 124 (2013) 443–456.
[11] C. Vickers, P. Hales, V. Kaushik, L. Dick, J. Gavin, J. Tang, K. Godbout, T. Parsons, E. Baronas, F. Hsieh, S. Acton, M. Patane, A. Nichols, P. Tummino, Hydrolysis of biological peptides by human angiotensin-converting enzyme-related carboxypeptidase, J. Biol. Chem. 277 (2002) 14838–14843.
[12] B. Chandrasekaran, O. Dar, T. McDonagh, The role of apelin in cardiovascular function and heart failure, Eur. J. Heart Fail 10 (2008) 725–732.
[13] L. Shi, C. Mao, Z. Xu, L. Zhang, Angiotensin-converting enzymes and drug discovery in cardiovascular diseases, Drug Discov. Today 15 (2010) 332–341.
[14] S.R. Tipnis, N.M. Hooper, R. Hyde, E. Karran, G. Christie, A.J. Turner, A human homolog of angiotensin-converting enzyme. Cloning and functional expression as a captorpr-insensitive carboxypeptidase, J. Biol. Chem. 275 (2000) 33238–33243.
[15] M. Donoghue, F. Hsieh, E. Baronas, K. Godbout, M. Gosselin, N. Stagliano, M. Donovan, B. Woolf, K. Robison, R. Jeyaseelan, R.E. Breitbart, S. Acton, A novel angiotensin-converting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1-9, Circ. Res. 87 (2000) E1–E9.
[16] M.A. Crackower, R. Sarao, G.Y. Oudit, C. Yagil, I. Kozieradzki, S.E. Scanga, A.J. Oliveira-dos-Santos, J. da Costa, L. Zhang, Y. Pei, J. Scholey, C.M. Ferrario, A.S. Manoukian, M.C. Chappell, P.H. Backx, Y. Yagil, J.M. Penninger, Angiotensin-converting enzyme 2 is an essential regulator of heart function, Nature 417 (2002) 822–828.
[17] K. Kuba, Y. Imai, S. Rao, H. Gao, F. Guo, B. Guan, Y. Huang, P. Yang, Y. Zhang, W. Deng, L. Bao, B. Zhang, G. Liu, Z. Wang, M. Chappell, Y. Liu, D. Zheng, A. Leibrandt, T. Wada, A.S. Slutsky, D. Liu, C. Qin, C. Jiang, J.M. Penninger, A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus-induced lung injury, Nat. Med. 11 (2005) 875–879.
[18] T. Hashimoto, T. Perlot, A. Rehan, J. Trichereau, H. Ishiguro, M. Paolino, V. Sigl, T. Hanada, R. Hanada, S. Lipinski, B. Wild, S.M. Camargo, D. Singer, A. Richter, K. Kuba, A. Fukamizu, S. Schreiber, H. Clevers, F. Verrey, P. Rosenstiel, J.M. Penninger, ACE2 links amino acid malnutrition to microbial ecology and intestinal inflammation, Nature 487 (2012) 477–481.
[19] G. Paizis, C. Tikellis, M.E. Cooper, J.M. Schembri, R.A. Lew, A.I. Smith, T. Shaw, F.J. Warner, A. Zuilli, L.M. Burrell, P.W. Angus, Chronic liver injury in rats and humans upregulates the novel enzyme angiotensin converting enzyme 2, Gut 54 (2005) 1790–1796.
[20] Y. Zhang, J. Wada, K. Hida, Y. Tsuchiyama, K. Hiragushi, K. Shikata, H. Wang, S. Lin, Y.S. Kanwar, H. Makino, Collectrin, a collecting duct-specific transmembrane glycoprotein, is a novel homolog of ACE2 and is developmentally regulated in embryonic kidneys, J. Biol. Chem. 276 (2001) 17132–17139.
[21] M. Rella, J.L. Elliot, T.J. Revett, J. Lanfear, A. Allred, T.H. Le, R. Griffiths, L. Mao, N. Philip, A.J. Turner, N.M. Hooper, Identification and characterisation of the angiotensin converting enzyme-3 (ACE3) gene: a novel mammalian homologue of ACE2, BMC Genomics 8 (2007) 194.
[22] N. Inoue, T. Kasahara, M. Ikawa, M. Okabe, Identification and disruption of sperm-specific angiotensin converting enzyme-3 (ACE3) mice, Biochim. Biophys. Acta 925 (1988) 304–310.
[23] P. Towler, B. Staker, S.G. Prasad, S. Menon, J. Tang, T. Parsons, D. Ryan, M. Fisher, D. Williams, N. Davie, M.A. Patane, M.W. Pantoliano, ACE2 X-ray structures reveal a large hinge-bending motion important for inhibitor binding and catalysis, J. Biol. Chem. 279 (2004) 17996–18007.
[24] J.H. Kregge, S.W. John, L.L. Langenbach, J.B. Hodgkin, J.R. Hagaman, E.S. Bachman, J.C. Jennette, D.A. O’Brien, E. Smithies, Male-female differences in fertility and blood pressure in ACE-deficient mice, Nature 375 (1995) 146–148.
[25] S.B. Gurley, A. Allred, T.H. Le, R. Griffiths, L. Mao, N. Philip, T.A. Haystead, M. Donoghue, R.E. Breitbart, S.L. Acton, H.A. Rockman, T.M. Coffman, Altered blood pressure responses and normal cardiac phenotype in ACE2-null mice, J. Clin. Invest. 116 (2005) 2218–2225.
[26] S. Der Sarkissian, J.L. Grobe, L. Yuan, D.R. Narielwala, G.A. Walter, M.J. Katovich, M.K. Raizada, Cardiac overexpression of angiotensin converting enzyme 2 protects the heart from ischemia-induced pathophysiology, Hypertension 51 (2008) 712–718.
K. Nakamura, N. Koibuchi, H. Nishimatsu, Y. Higashikuni, Y. Hirata, K. Yamamoto, M. Ohishi, T. Katsuya, N. Ito, M. Ikushima, M. Kaibe, B. Rentzsch, M. Todiras, R. Iliescu, E. Popova, L.A. Campos, R.A. Santos, A.J. Ferreira, A.P. Nadu, A.N. Braga, A.P. de Almeida, M. Donoghue, H. Wakimoto, C.T. Maguire, S. Acton, P. Hales, Y. Yamazato, A.J. Ferreira, K.H. Hong, S. Sriramula, J. Francis, R.P. Marshall, S. Webb, G.J. Bellingan, H.E. Montgomery, B. Chaudhari, M. Otsuka, H. Takahashi, M. Shiratori, H. Chiba, S. Abe, Reduction of cardiomyopathy in ACE2 null mice, Cardiovasc. Res. 75 (2007) 2918–2926.

Y. Imai, K. Kabuto, Y. Hishishita, H. Higashikuni, Y. Hirata, K. Kugiyama, R. Nagai, M. Sata, Candesartan ameliorates cardiac dysfunction observed in angiotensin-converting enzyme 2-deficient mice, Hypertens. Res. 31 (2008) 1953–1961.

G.Y. Oudit, Z. Kassiri, M.P. Patel, M. Chappell, J. Butany, P.H. Backx, R.G. Tsushima, J.W. Scholey, R. Khokha, J.M. Penninger, Angiotensin II-mediated oxidative stress and inflammation mediate the age-dependent cardiomyopathy in ACE2 null mice, Cardiovasc. Res. 75 (2007) 29–39.

A.J. Ferreira, R.A. Santos, A.P. Almeida, Angiotensin-(1-7)-cardioprotective effects in myocardial ischemia/reperfusion, Hypertension 38 (2001) 665–668.

R.A. Santos, A.J. Ferreira, A.P. Nadu, A.N. Braga, A.P. de Almeida, M.J. Campagnole-Santos, O. Baltatu, R. Iliasescu, T.L. Reudelhuber, M. Bader, Expression of an angiotensin-(1-7)-producing fusion protein produces cardioprotective effects in rats, Physiol. Genomics 17 (2004) 292–299.

M. Donoghue, H. Wakimoto, C.T. Maguire, S. Acton, P. Hales, N. Stagliano, V. Fairchild-Huntress, J. Xu, J.N. Lorenz, V. Kadambi, C.J. Berul, R.E. Breibtart, Heart block, ventricular tachycardia, and sudden death in ACE2 transgenic mice with downregulated connexins, J. Mol. Cell Cardiol. 35 (2003) 1043–1053.

R. Masson, S.A. Nicklin, M.A. Craig, M. McBride, K. Gilday, P. Gregorevic, J.M. Allen, J.S. Chamberlain, G. Smith, D. Graham, J.W. Scholey, Loss of angiotensin-converting enzyme-2 accelerates diabetic kidney injury, Am. J. Pathol. 171 (2007) 438–451.

D. Zimmermann, K.D. Burns, Angiotensin-(1-7) in kidney disease: a review of the controversies, Clin. Sci. (Lond.) 123 (2012) 333–346.

J.S. Peiris, K.Y. Yuen, A.D. Osterhaus, K. Stohr, The severe acute respiratory syndrome, N. Engl. J. Med. 349 (2003) 2431–2441.

W. Li, M.J. Moore, N. Vasiliou, J. Sui, S.K. Wong, M.A. Berne, M. Somasundaran, J.L. Sullivan, K. Luzuriaga, T.C. Greenough, H. Choe, M. Farzan, Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus, Nature 426 (2003) 450–454.

F. Li, W. Li, M. Farzan, S.C. Harrison, Structure of SARS coronavirus spike receptor-binding domain complexed with receptor, Science 309 (2005) 1808–1809.

H. Hofmann, K. Pyrc, L. van der Heek, M. Geier, B. Berkhourt, S. Pohlmann, Human coronavirus NL63 employs the severe acute respiratory syndrome coronavirus receptor for cellular entry, Proc. Natl. Acad. Sci. U S A 102 (2005) 7998–7993.

A.M. Zaki, S. van Boheemen, T.M. Bestebroer, A.D. Osterhaus, R.A. Fouchier, Isolation of a novel coronavirus from a man with pneumonia in Saudi Arabia, N. Engl. J. Med. 367 (2012) 1814–1820.

R.J. de Groot, S.C. Baker, R.S. Baric, C.S. Brown, C. Drosten, L. Enjuanes, R.A. Fouchier, M. Galiano, A.E. Gorbalenya, Z. Memish, S. Perlman, L.L. Poon, E.J. Snijder, G.M. Stephens, P.C. Woo, A.M. Zaki, M. Zambon, J. Ziebuhr, Middle East respiratory syndrome coronavirus (MERS-CoV); announcement of the coronavirus study group, J. Virol. 87 (2013) 7790–7792.

M.A. Muller, V.S. Raj, D. Math, B. Meyer, S. Kallies, S.L. Smits, R. Wolny, T.M. Bestebroer, S. Spiech, T. Suliman, K. Zimmermann, T. Binger, I. Eckerle, M. Tschapka, A.M. Zaki, A.D. Osterhaus, R.A. Fouchier, B.L. Haagmans, C. Drosten, Human coronavirus EMC does not require the SARS-coronavirus receptor and maintains broad replicative capability in mammalian cell lines, mBio 3 (2012) e00515-7993.

V.S. Raj, H. Mou, S.L. Smits, D.H. Dekkers, M.A. Muller, R. Dijkman, D. Muth, J.A. Demmers, A. Zaki, R.A. Fouchier, V. Thiel, C. Drosten, P.J. Rottier, A.D. Osterhaus, B.J. Bosch, B.L. Haagmans, Dipeptidyl peptidase 4 is a functional receptor for the emerging human coronavirus-EMC, Nature 495 (2013) 251–254.

P. Akpinar, S. Kuzawjima, J. Krutzfeldt, M. Stoffel, Tmem27: a cleaved and shed plasma membrane protein that stimulates pancreatic beta cell proliferation, Cell Metab. 2 (2005) 385–397.
