A role for mitochondrial oxidants in stress-induced premature senescence of human vascular smooth muscle cells

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M T H O D S

Introduction

Cell senescence prevents uncontrolled mitosis of transformed cells, but is also associated with mechanisms of age-related pathologies including cardiovascular disease [1]. Stress induced premature senescence (SIPS) is defined as the early senescence of cells resulting from repeated exposure to a variety of cellular stressors at sub-cytotoxic concentrations, e.g. \( H_2O_2 \), chemotherapeutic agents, ultraviolet and ionizing radiation [2]. Oxygen radicals and intermediates are implicated in the induction of SIPS with high concentrations of these oxidants acting through damage to DNA but there is also a role for physiological levels of \( H_2O_2 \) and \( O_2^- \) in particular, mediating effects through cell signalling pathways.

SIPS has been studied in cultured skin fibroblast models [3] and endothelial cells [4] and to a limited extent in vascular smooth muscle cells (VSMC) [5,6]. Two main types of protocol have been used to study premature senescence or SIPS. Firstly, continuous stress and secondly, a series of stresses with recovery periods in-between and following the final stress. Much lower concentrations are often used for the latter (SIPS) protocol. In the continuous model it is not possible to discriminate between immediate effects of stress and long term, irreversible effects on aging (e.g. p53 growth arrest by oxidative damage). With the repeat stress model the provision of complete growth media between stresses and following final stress allows cells to recover, thus avoiding interactions between acute effects and long term or aging effects; this paradigm has been adopted here to study angiotensin II (Ang II)-induced senescence in human VSMC.

In vivo, SIPS has been observed in chronic kidney disease patients [7] and in experimental models [8]. Notably, cells with a characteristic senescence phenotype have been observed in human arteries at sites prone to atherosclerosis [9] and their accumulation may account for the decline in tissue and organ function observed with aging in vivo.
Reports have suggested a correlation between dysfunctional or damaged mitochondria and the onset or accumulation of cellular senescence [10]. An increase in mitochondrial dysfunction has been observed in human tissue with advancing age [11], which correlates with a gradual accumulation of senescent cells. There is little evidence to determine whether mitochondria have a mechanistic role in promoting premature senescence, however, a very recent study showed that inhibition of mitochondrial superoxide promoted cell death and senescence of B16 mouse melanoma cells [12].

We and others have reported that Ang II induces senescence in human VSMC in vitro [5,13]. Ang II causes oxygen radical production in VSMC via NADPH oxidase [14,15] and oxidative DNA damage [5]. Both replicative senescence, by accelerated telomere attrition over a number of cell divisions and premature senescence have been identified as possible mechanisms for the effects of Ang II [5] but formal proof of induction of SIPS has not been demonstrated. Prevention of senescence may partly explain the beneficial effects, in terms of increasing lifespan and maintaining organ/tissue function in rodents, of drugs which inhibit Ang II synthesis and function [16]. We hypothesise that Ang II mediates senescence of VSMC by O2· generation dependent upon mitochondrial function and that inhibition of mitochondrial O2· may help delay or prevent VSMC aging in vitro.

Materials and methods

Reagents

All chemicals were supplied by Sigma unless otherwise noted. Mito-TEMPO was purchased from Enzo Life Sciences and mitoSOX from Invitrogen Ltd.

Thenoyltrifluoroacetone (TFFA) was supplied by Fisher Scientific. EXP3174 was a gift from Merck & Co Inc.

hVSMC culture and induction of SIPS

VSMC were cultured as previously described [5] and used between passage 2 and 9. Cells were cultured in RPMI 1640 media containing 10% foetal calf serum (FCS), 2.5% (v/v) smooth muscle cell growth supplement (TCS Cellworks), 100 μg/ml glutamine, 100 μg/ml penicillin/streptomycin and 20 mmol/L HEPES buffer and maintained in a 5% CO2 atmosphere. Near confluent cells were rendered quiescent in media containing 0.5% (v/v) FCS (without growth supplement) for 24 h prior to Ang II (1 × 10⁻⁸ or 1 × 10⁻⁷ mol/L) exposures.

In order to investigate induction of SIPS, the protocol of Toussaint et al. was adopted [3,2]. Sub-confluent cells were treated with successive Ang II for 2 h each day, for 3 days, followed by 24 h recovery periods after each treatment by replacing the culture media with fresh growth media. Treatment with tert-butyldihydroperoxide ( tert-BHP; 4 × 10⁻⁵ mol/L) was conducted in the same manner in parallel cultures as a positive control. The same schedule of media changes without the stress treatment was performed on control cells.

Senescence-associated β-galactosidase (SA-β-gal) staining

Senescence was determined by measurement of SA-β-gal activity [17] using the ‘Senescence Cell Staining kit’ (Sigma). Following treatments with peroxide or Ang II, hVSMC were re-plated (5 × 10⁴ per well) in 12-well plates in media containing 10% FCS for 24 h prior to staining. Senescent cells were enumerated by light microscopy in five fields of view selected at random in each well.

To investigate the effect of antioxidants and mitochondrial inhibitors on senescence, VSMC were maintained in media containing 0.5% (v/v) foetal calf serum for 24 h. Cells were then incubated in media containing catalase (300 Units/ml for 3 h), N-acetylcysteine (NAC; 0.5 × 10⁻² mol/L for 2 h), SOD (50 Units/ml for 3 h), rotenone (2 × 10⁻⁶ mol/L for 3 h), thenoyltrifluoroacetone (TFFA; 1 × 10⁻⁶ mol/L for 3 h) or the mitochondrial O2· scavenger mito-TEMPO (25 × 10⁻⁹ mol/L for 4 h) prior to Ang II exposure at 1 × 10⁻⁸ mol/L for 24 h at 37 °C. Following Ang II treatment, cells were trypsinized, counted using a haemocytometer and re-plated at 5 × 10⁴ cells per well in 12-well plates, in media containing 10% (v/v) FCS. Cells were then left to adhere for 24 h at 37 °C prior to fixation and staining for SA-β-gal.

Measurement of NADPH-dependent superoxide production

Quiescent hVSMC were stimulated with Ang II for 1 h then homogenized by sonication. The lucigenin chemiluminescence assay was used as previously described to determine NADPH-dependent superoxide production in cell lysates [5]. To assess whether Ang II-induced superoxide production was dependent upon mitochondrial activity, the effect of electron transport chain inhibitors for complex I (rotenone, 10 × 10⁻⁶ mol/L) and II (TFFA, 10 × 10⁻⁶ mol/L) was studied.

Detection of mitochondrial O2·-

MitoSOX™ red was used in live hVSMC. Near confluent cells were exposed to tert-BHP (50 × 10⁻⁴ mol/L) for 2 h or Ang II (1 × 10⁻⁷ mol/L) for 1 h at 37 °C. Following treatment, cells were loaded with 5 × 10⁻⁵ mol/L MitoSOX™ red in Hanks’ balanced salt solution (HBSS) for 10 min at 37 °C. Cells were washed 3 × in HBSS and finally 500 μL of HBSS was added to each well and fluorescence was measured using a Cary Eclipse Fluorescence Spectrophotometer plate reader (excitation: 510 nm; emission: 580 nm). Fluorescence measurements were calculated relative to untreated cells loaded with the probe.

Analysis of data

Comparison of means was performed using one way-ANOVA.

Results

Induction of SIPS in cultured hVSMC by tert-BHP

To our knowledge, the induction of SIPS under the defined protocol described by Toussaint et al. [2] has not been previously reported for hVSMC. In order to investigate this, hVSMC were exposed to mild, successive tert-BHP stresses for 2 h each over 3 days with 24 h recovery periods between each treatment followed by an extended period of recovery. Following tert-BHP, hVSMC displayed > 2.5-fold increase in cells staining for SA-β-gal compared with controls (Fig. 1A). This is the first direct evidence, using the strict definition of SIPS induction, that hVSMC in culture undergo SIPS in response to peroxide stress.

Ang II stresses induce SIPS via angiotensin II receptor

Previous work from our group showed that Ang II accelerated the onset of hVSMC senescence via telomere-dependent and -independent mechanisms [5]. To determine whether Ang II induced SIPS, hVSMC were exposed to Ang II using the same regime as for tert-BHP, namely treatment followed by recovery and consistent with other reported studies on SIPS in other cell types [3,18]. A two-fold increase in SIPS was observed at an Ang II concentration of 1 × 10⁻⁸ mol/L (Fig. 1B). This is the first report that acute, repeated Ang II treatments cause senescence of hVSMC. Induction of senescence was reduced (p < 0.05) by approximately 50% by pre-incubation of cells with the
type-1 angiotensin II receptor (AT1) antagonist, EXP3174 (E3174) prior to each Ang II exposure (Fig. 1C) confirming a true receptor-mediated mechanism for induction of SIPS by Ang II.

Contribution of mitochondria to Ang II-induced superoxide production in hVSMC

Since Ang II initiates various cell signalling pathways in VSMC via increased ROS generation [14], which is also a key trigger of cellular senescence [19], it is therefore reasonable to hypothesize that $O_2^\cdot-$ is a significant mediator of SIPS in hVSMC. Previous work has shown that Ang II elevates superoxide generation via NADPH oxidase activity in VSMC [20,15] and that, in this model, superoxide production was not detected when Complex III of the electron transport chain was inhibited by antimycin A [5]. The addition of the mitochondrial complex I inhibitor rotenone almost fully suppressed NADPH-dependent superoxide production due to Ang II over a period of 40 min (Fig. 2). There was also a trend towards decreased Ang II-induced superoxide with TTFA but this did not reach statistical significance in this model (Fig. 2). These data implicated mitochondrial function as an important determinant of NADPH-dependent $O_2^\cdot-$ production following Ang II stimulation, however, a direct scavenging of $O_2^\cdot-$ by rotenone or a direct effect on NADPH oxidase activity in lysates cannot be ruled out from these data.

The possible contribution of mitochondria to cellular $O_2^\cdot-$ following Ang II stimulation was investigated using mito-SOX as a superoxide reporter in live hVSMC (Fig. 3). Both tert-BHP and Ang II stimulated mitoSOX fluorescence by 1.5–2.0 fold compared to the control indicating that mitochondria within cells produce superoxide in response to Ang II.
mitochondrial function and specifically prevented Ang II-induced senescence, strongly implicating mitochondrial function in the induction of premature senescence of VSMC, cells were pre-incubated with mitochondrial antioxidants catalase, NAC and SOD inhibited Ang II-induced premature senescence of VSMC (Fig. 4A-C) confirming the involvement of oxidants in senescence.

To determine whether mitochondria were directly involved in the mechanism or contributed to the Ang II induced mitochondrial superoxide production of hVSMC, cells were pre-incubated with mitochondrial complex inhibitors. Ang II exposure for 24 h consistently caused a ~2-fold increase in senescent cells (Fig. 4). Pre-incubation with rotenone completely inhibited Ang II-induced senescence of hVSMC (Fig. 4D) as did incubation with the complex II inhibitor, TFFA (Fig. 4E).

Ang II-induced premature senescence is dependent upon superoxide generated by mitochondria

The data presented so far illustrate that Ang II induces NADPH-dependent 

\[ \text{O}_2^- \]

in cell lysates and mt 

\[ \text{O}_2^- \]

in living cells and that SIPS is dependent upon oxidant generation and mtETC activity. Further investigation into whether mitochondria stimulated by Ang II participate in the induction of senescence, was conducted using a mitochondrially-targeted oxygen radical scavenger, mito-TEMPO. Mito-TEMPO accumulates within mitochondria, where it scavenges 

\[ \text{O}_2^- \]

and alkyl radicals specifically [21]. Incubation of hVSMC with a relatively low concentration of mito-TEMPO (25 \times 10^{-9} \text{ mol/L}) completely prevented Ang II-induced senescence, strongly implicating mitochondrial function and specifically 

\[ \text{O}_2^- \]

in the rapid senescence response to Ang II.

Discussion

Here we show that mitochondria play a key role in Ang II-induced human VSMC senescence. Inhibition studies suggest that mitochondrial function, specifically mtETC activity, is important in the induction of premature senescence.

Our previous work showed that catalase inhibited Ang II-induced senescence [5] and this was not only confirmed here but extended with the application of SOD and NAC. Here, inhibition of 

\[ \text{O}_2^- \]

and senescence by rotenone and the effectiveness of nanomolar concentrations of mitoTEMPO at blocking senescence induced by Ang II, implicate superoxide production by mitochondria in this mechanism. This is in agreement with the suggestion that mitoTEMPO mimicks the antioxidant effects of SOD2 in mitochondria [22] and further adds weight to the hypothesis that mitochondrial superoxide is a mediator of physiological and stress response pathways in human cells [23]. Complex I is proposed as the main source of 

\[ \text{O}_2^- \]

in the mitochondrial electron transport chain, including via reverse electron transport following inhibition at complex II [24]. Therefore, the current data suggest that Complex I is an important source of superoxide generation by Ang II in hVSMC. The complex interactions between mitochondrial and NADPH oxidase 

\[ \text{O}_2^- \]

generation are discussed below.

It is well described that Ang II stimulates superoxide production via activation of NADPH oxidases in vascular cells [25,26]. A mechanism for the observed mitochondrial dysfunction caused by Ang II in endothelial cells has been described involving upstream NADPH oxidase activity [27] and redox-dependent mitochondrial K_{ATP} channels which may be activated by either 

\[ \text{O}_2^- \]

or 

\[ \text{H}_2\text{O}_2 \]

[28]. Nevertheless, there is also evidence in some cell models e.g. endothelial cells, that mitochondrial 

\[ \text{O}_2^- \]

is upstream of NADPH oxidase (NOX) activity which leads to further 

\[ \text{O}_2^- \]

production in the cytosolic phase [22]. In this endothelial cell model, in response to Ang II, SOD2 modulated NADPH oxidase activity in the cytosol, implying that mitochondrial superoxide was the signal for increased NOX activity. Moreover, mitoTEMPO effectively mimicked the activity of SOD2 in that it prevented Ang II-stimulated 

\[ \text{O}_2^- \]

production in SOD2 depleted cells [22]. Furthermore, the ability of Ang II to mediate NADPH oxidase expression and activity in rabbit VSMC was eliminated by mtDNA depletion with ethidium bromide or inhibition of complex III by antimycin A [29]. In addition to suggesting a complex mitochondrial-NOX crosstalk [30], these findings support the contention that mitochondrial redox biology plays a key biological role in regulating important cellular processes [23]. In the current model (see Fig. 5), mitochondrial 

\[ \text{O}_2^- \]

is vital for the senescence process but it is unknown whether the signal from mitochondria involves its dismutation to 

\[ \text{H}_2\text{O}_2 \]

and/or whether other intermediate signalling event(s) is/are required. In this respect, targeting catalase to mitochondria might be informative.

There are many implications of a mitochondrial pharmacology approach in biology and medicine. Mitochondrial function/signaling is a developing clinical target and mitochondria-targeted antioxidants are amongst several promising modalities [31]. Although it is some way from clinical fruition, the data presented here support the contention that this class(es) of compounds may be employed as potential therapies for targeting cells and tissues prone to aging and age-related disease with the aim of maintaining tissue/organ function and prolonging a healthy lifespan.

The present study provides the first report of SIPS in hVSMC using the specific experimental criteria developed by Toussaint et al. [2]. The current study demonstrates that short, successive exposures to either tert-BHP or Ang II, with periods of recovery between treatments, were sufficient to induce SIPS of hVSMC. Both of these mediators act via redox mechanisms, which have been intimately associated with induction of cellular senescence in several cell systems [3,13,5].

Data from the current and previous studies [5] suggest that Ang II acts via its cognate receptor, AT1, since the inhibition of the AT1 with EXP3174 prior to daily Ang II exposure blocked the senescence response in hVSMC. Complete inhibition of senescence was not attained with EXP3174 which may mean that this antagonist only partially prevents Ang II agonism under the experimental conditions employed, however an alternate mechanism for Ang II, for example via binding to the type-2 angiotensin II receptor, cannot be ruled out. Nevertheless, we favour the former explanation since in previous studies EXP3174 fully inhibited DNA damage
and telomere attrition in addition to induction of senescence by Ang II [5].

Induction of senescence by Ang II was first reported in vitro in aortic VSMC and also in vivo in apoE deficient mice [13]. These in vivo analyses showed that Ang II infusion promoted staining for SA-β-gal activity within mouse aortas and co-staining for smooth muscle actin suggested that much of the senescence staining was associated with VSMC [13]. This evidence provides an important in vivo context for the observations on Ang II-induced senescence and provides a rationale for studying mechanisms in vitro. Subsequent workers [32] have highlighted the complexity of pathways which may be involved in signal transduction and may also include a significant contribution from the cytoskeleton-extra cellular matrix axis [33].

A body of work looking at the possible role of Ang II in aging in vivo has been reported [16] with the pleiotropic actions of Ang II implying several mechanisms may be involved. One initial experimental paradigm showed that lifelong feeding of ACE inhibitors to CF1 mice resulted in lifespan extension and reduced pathology usually associated with the normal aging process, at least for laboratory animals [34]. This and subsequent work by the same group has suggested that preservation of mitochondrial function is a key determinant of anti-Ang II treatments in rodent models of aging and is not dependent on blood pressure lowering [16]. It confirms that, perhaps inadvertently, these drugs are amongst the most effective antioxidant therapies currently used in man. In this respect, blockade of the renin-angiotensin system in humans is likely to affect several biological processes/pathways and provides an avenue for further exploration particularly to mitigate cardiovascular aging.

**Conclusions**

The data presented here demonstrate that Ang II induces true stress-induced premature senescence in human VSMC and suggest that mitochondrial O$_2^-$ is necessary for this premature aging response. Furthermore, the data confirm an Ang II receptor mediated process which is likely to involve mitochondria-NADPH oxidase crosstalk by as yet undefined signalling pathways.
Cross-talk via oxidant generation; the working hypothesis is that cytoplasmic oxidants, mitochondrial superoxide/mitochondrial function are necessary for the development of SIPS. This mechanism invokes NADPH oxidase-mitochondrial cross-talk via oxidant generation; the working hypothesis is that firstly NADPH oxidase is activated by Ang II which leads to stimulation of mitochondrial superoxide resulting in a feed-forward amplification of NOX activity and subsequent downstream induction of SIPS.

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