Activation of the AMP-Activated Protein Kinase (AMPK) by Nitrated Lipids in Endothelial Cells

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Abstract

The AMP-activated protein kinase (AMPK) is an important regulator of endothelial metabolic and functional homeostasis. Here, we examined the regulation of AMPK by nitrated oleic acid (OA-NO2) and investigated the implications in endothelial function. Treatment of bovine aortic endothelial cells (BAECs) with OA-NO2 induced a significant increase in both AMPK-Thr172 phosphorylation and AMPK activity as well as upregulation of heme oxygenase (HO)-1 and hypoxia-inducible factor (HIF)-1α. Pharmacologic inhibition or genetic ablation of HO-1 or HIF-1α abolished OA-NO2-induced AMPK phosphorylation. OA-NO2 induced a dramatic increase in extracellular signal-regulated kinase (ERK)1/2 phosphorylation that was abrogated by the HO-1 inhibitor, znic deuteroporphyrin IX 2,4,5-bis-ethylene glycol (ZnBG). Inhibition of ERK1/2 using U0126 or PD98059 reduced but did not abolish OA-NO2-induced HIF-1α upregulation, suggesting that OA-NO2/HO-1-initiated HIF-1α induction is partially dependent on ERK1/2 activity. In addition, OA-NO2 enhanced endothelial intracellular Ca2+, an effect that was inhibited by the HIF-1α inhibitor, YC-1, and by HIF-1α siRNA. These results implicate the involvement of HIF-1α. Experiments using the Ca2+/calmodulin-dependent protein kinase (CaMKK) inhibitor STO-609, the selective CaMKK inhibitor KN-93, and an isoform-specific siRNA demonstrated that OA-NO2-induced AMPK phosphorylation was dependent on CaMKKβ. Together, these results demonstrate that OA-NO2 activates AMPK in endothelial cells via an HO-1-dependent mechanism that increases HIF-1α protein expression and Ca2+/CaMKKβ activation.

Introduction

Accumulating evidence indicates that reactive nitrogen species (RNS), which are inflammatory oxidants, mediate diverse physiologic and pathologic processes in cardiovascular, pulmonary, and neurodegenerative diseases [1]. RNS, such as peroxynitrite (ONOO-), nitrogen-dioxide radical, and nitronium ion, which are formed from nitric oxide (NO), are important factors in complications of obesity and diabetes [2]. RNS react with unsaturated fatty acids to form relatively stable nitrated products (nitralkene derivatives), including the abundant and clinically important nitrated oleic and linoleic acids [3]. Although this process occurs via various mechanisms, the common denominator is a proclivity for homolytic addition of nitrogen dioxide (NO2·) to the double bond, yielding an array of regio- and stereoisomers [4]. Nitrated unsaturated fatty acids (NO2-UFAs) represent a convergence of lipid and NO signaling and have emerged as a novel class of endogenously produced vascular signaling molecules [5]. The beneficial effects of NO2-UFAs include cGMP-dependent vessel relaxation, inhibition of inflammatory cell function, adaptive and anti-inflammatory cell responses, induction of heme oxygenase-1 (HO-1) expression, inhibition of nuclear factor (NF)-κB, and activation of peroxisome proliferator activated receptor (PPAR) and Kcap1/Nrf2 [6].

Plasma free and esterified nitrated oleic acid (OA-NO2) concentrations have been reported to be 619±52 and 302±569 nM, respectively, and these levels are approximately 50% greater than the level of nitrated linoleic acid (LNO2). The combined blood levels of the free and esterified fatty acid derivatives exceeds 1 μM [4]. The plasma levels of NO2-UFA derivatives in hyperlipidemic patients are elevated relative to levels in normolipidemic subjects [7]. In addition, an increase in oxidative stress in hypercholesterolemia [8] may contribute to the formation of nitrated species in the vascular wall [9]. Thus, nitrated lipids in the plasma can be used as an indicator of the formation of nitrated species in the vascular wall [9].

The serine/threonine kinase, AMP-activated protein kinase (AMPK), is a member of the Snf1/AMPK protein kinase family that is found in all eukaryotes [11]. This kinase is thought to act as a cellular energy sensor by stimulating ATP-producing catabolic pathways and inhibiting ATP-consuming anabolic pathways [12].
AMPK is comprised of three subunits: a catalytic α-subunit and regulatory β- and γ-subunits. Activation of AMPK requires the phosphorylation of Thr172 in the activation loop of the α-subunit by an upstream AMPK kinase (AMPKK). Interestingly, the first AMPKK to be identified was LKB1, a tumor suppressor that is mutated in humans with Peutz-Jeghers syndrome [14]. Patients with this syndrome have an increased risk of developing carcinomas of the colon, stomach, and pancreas. Recently, calcium (Ca²⁺) calmodulin-dependent kinase (CaMKK) [15] was identified as an upstream AMPKK. CaMKK is activated by a rise in intracellular Ca²⁺ concentrations ([Ca²⁺]), and this kinase phosphorylates and activates AMPK in an AMP-independent manner [15]. Therefore, in addition to responding to an increase in the AMP-to-ATP ratio, AMPK may also be activated by a rise in [Ca²⁺], in response to nutrients, drugs, or physiological stimuli. While the AMPK pathway is traditionally thought to be a regulator of metabolism, recent studies have demonstrated that AMPK may also act to maintain endothelial function [16]. AMPK exerts pleiotropic effects that are believed to be beneficial for endothelial function. These effects, which are ultimately anti-atherogenic, include induction of the endothelial nitric oxide synthase (eNOS)/NO pathway and result in an increase in NO bioavailability, suppression of endothelial ROS production following exposure to deleterious stimuli, such as hyperglycemia or high free fatty acids (FFAs), and modulation of vascular tone (see review [16]). AMPK also possesses anti-apoptotic and anti-inflammatory activities [17].

Since many of the metabolic changes and endothelial-protective effects attributable to NO₂-UFAs are similar to those observed in response to AMPK activation, we hypothesized that AMPK activation is an important mediator of NO₂-UFA activity. As such, AMPK activation may explain the pleiotropic beneficial effects of NO₂-UFAs on the cardiovascular system in obesity and diabetes. Whether NO₂-UFAs activate AMPK and, if so, by what mechanism(s) has yet to be determined. In the present study, we examined the effects of NO₂-UFAs on the AMPK upstream kinases, specifically LKB1 and CaMKKβ. We also investigated whether NO₂-UFAs modulate the eNOS/NO pathway, which is known to be an indicator of endothelial function and an important property of AMPK in cardiosvascular tissue. We report that treatment with NO₂-UFAs induces CaMKKβ-dependent AMPK activation through an HO-1/HIF-1α Ca²⁺ pathway in vitro and show that NO₂-UFAs promote p-eNOS and NO production via activation of the AMPK pathway in endothelial cells.

**Results**

**OA-NO₂ induces HO-1 protein and AMPK phosphorylation/activation in bovine aortic endothelial cells (BAECs)**

To investigate activation of AMPK by OA-NO₂ in endothelial cells, we treated confluent BAECs with different concentrations of OA-NO₂ for 2–16 h. AMPK activation was indirectly assessed by western blot analysis of AMPK Thr172 phosphorylation, which is essential for AMPK activity [18]. As shown in Figure 1A and 1B, incubation of BAECs with OA-NO₂ (0.5–2.5 μM for 16 h, or 2.5 μM for 2–16 h) resulted in a dose- and time-dependent increase in AMPKα2 phosphorylation at Thr172. AMPK Thr172 phosphorylation in BAECs gradually increased beginning 6 h after incubation with 2.5 μM OA-NO₂ and reached peak levels after 16 h. This increased AMPK phosphorylation was associated with elevated AMPK activity, as measured by the SAMS peptide assay (Figure 1C). OA-NO₂ treatment did not alter total AMPK levels, suggesting that OA-NO₂-induced phosphorylation of AMPK was not due to altered expression of these proteins. Since OA-NO₂ activated AMPK in both BAECs and human umbilical vein endothelial cells (HUVECs) at similar potencies, we performed the majority of our experiments with BAECs.

The changes in AMPK phosphorylation were mirrored by changes in HO-1 protein expression, as evaluated by western blot analysis. Exposure of BAECs to a range of OA-NO₂ concentrations resulted in an upregulation of HO-1 that was initially observed at 4 h with peak induction at 16 h. HO-1 induction and AMPK activation were specifically due to the nitroalkene moiety of OA-NO₂, because oleic acid (OA) did not induce HO-1 protein expression or AMPK phosphorylation (Figure 1D). At a concentration of 2.5 μM, OA-NO₂ potently activated AMPK and induced HO-1 expression without causing cellular toxicity. On the basis of these results, we chose to stimulate BAECs with 2.5 μM OA-NO₂ for 16 h in subsequent experiments.

To determine whether OA-NO₂-induced HO-1 expression was mediated by AMPK, we used an adenovirus encoding a dominant-negative mutant form of AMPKα (Ad-DN-AMPK) to suppress AMPK activity. AICAR (5-aminimidazole-4-carboxamide-1-beta-D-ribofuranoside) and metformin, two well-characterized AMPK activators, were used as positive controls for AMPK phosphorylation. OA-NO₂ significantly upregulated AMPK phosphorylation (Figure 1E). As expected, Ad-DN-AMPK effectively suppressed AMPK phosphorylation but failed to abolish HO-1 induction by OA-NO₂, suggesting that AMPK does not act upstream of HO-1 production in this system.

**Activation of AMPK by OA-NO₂ does not require LKB1**

Two recent studies [14,19] showed that LKB1 acts as an upstream AMPK kinase. To evaluate the role of LKB1 in OA-NO₂-induced AMPK activation, we investigated whether OA-NO₂ altered LKB1 Ser 428 phosphorylation, which is essential for LKB1 activity and LKB1-dependent AMPK activation. As shown in Figure 2A, OA-NO₂ did not alter the levels of LKB1 Ser428 phosphorylation.

We next determined whether OA-NO₂ activated AMPK in Hela-S3 cells, an LKB1-deficient cell line. Similar to the results obtained in BAECs, OA-NO₂ (2.5 μM) induced AMPK phosphorylation on Thr172 in Hela-S3 cells (~3-fold, p<0.05; Figure 2B). In parallel with AMPK phosphorylation, OA-NO₂ dramatically increased phosphorylation of the known AMPK substrate, acetyl-CoA carboxylase (ACC), at Ser79 (~2.4-fold, p<0.05), providing an additional indication that AMPK was activated in the absence of LKB1. To further explore the role of LKB1, we investigated the effect of siRNA-mediated LKB1 downregulation on AMPK activation induced by OA-NO₂. As shown in Figure 2C, OA-NO₂-induced AMPK phosphorylation was not significantly altered in cells pre-treated with LKB1 siRNA. The suppression of LKB1 protein expression by LKB1 siRNA (~60%) was confirmed by western analysis. Taken together, these results strongly suggest that OA-NO₂-induced phosphorylation of AMPK Thr172 is independent of its upstream kinase, LKB1.

**Induction of HIF-1α by OA-NO₂ is dependent on HO-1**

Hypoxia-inducible factor (HIF)-1 is a dimeric protein complex that plays an integral role in the body’s response to hypoxia [20]. Like AMPK, HIF-1 is one of the primary genes involved in the homeostatic process that leads to increased vascularization in hypoxic areas, such as those within localized ischemia and in tumors [20]. HIF-1 and AMPK represent two main cellular pathways involved in coping with hypoxic stress and protecting cells against energy depletion and tissue reperfusion injury in times of metabolic crisis [21]. In addition, both HIF-1 and HO-1 are
associated with ferrous iron metabolism [21]. Given that OA-NO2 induced both HO-1 expression and AMPK activation, we determined whether OA-NO2 also influenced HIF-1 expression. As predicted, OA-NO2 induced an increase in HIF-1α in HUVECs with a time course similar to OA-NO2-induced HO-1 expression and AMPK phosphorylation (Figure 3A). To explore the potential involvement of HO-1 in OA-NO2-induced HIF-1α expression, we pre-treated cells with the selective HO inhibitor zinc deuteroporphyrin IX 2,4-bis-ethylene glycol (ZnBG) and HO-1-specific siRNA. As shown in Figure 3A and B, ZnBG and HO-1 siRNA treatment abrogated the induction of HIF-1α protein expression and AMPK phosphorylation by OA-NO2, suggesting that OA-NO2 acts through the induction of HO-1 to increase HIF-1α expression and subsequently activate AMPK. The efficacy of the HO-1 siRNA in the downregulation of HO-1 expression was confirmed by western analysis (Figure 3B).

ERK1/2 is partially responsible for OA-NO2/HO-1-mediated HIF-1α induction

Clearly, the HIF pathway plays an important role in the regulation of metabolism under hypoxic conditions; however, a variety of HIF-1 stimuli function independently of oxygen concentration. These stimuli are primarily mediated by proteins that regulate HIF-1 translation. This pathway contrasts with hypoxic stimuli, which act upon pre-existing α-subunits [20]. HIF-1α activation by non-hypoxic stimuli has been linked primarily to the phosphatidylinositol 3-kinase (PI3-K)/Akt pathway and the mitogen-activated protein kinase (MAPK) pathway [22]. Both kinase pathways are known to be intimately associated with the regulation of HIF-1α protein translation, stabilization, and transcriptional activity [23]. Our data indicate that OA-NO2 did not activate PI3-K/Akt (data not shown). To determine whether OA-NO2-mediated HIF-1α induction is dependent on the MAPK pathway, we examined p38 MAPK and extracellular signal-regulated kinase 1/2 (ERK1/2) phosphorylation following stimulation with OA-NO2. Treatment with OA-NO2 induced a dose-dependent increase in the phosphorylation of ERK1/2 and p38 MAPK (Figure 4A). ZnBG inhibited OA-NO2-induced activation of ERK1/2, suggesting a role for HO-1 in this activation (Figure 4B).

We next determined whether ERK1/2 was a potential upstream mediator of OA-NO2-induced HIF-1α upregulation using the ERK1/2 inhibitor, UO126 (10 μM), and the MAPK/ERK (MEK)-specific inhibitor PD98059 (50 μM). Pre-treatment of BAECs with UO126 or PD98059 decreased HIF-1α levels by approximately 50% relative to cells treated with OA-NO2.
alone (Figure 4C), suggesting that OA-NO2-mediated HIF-1α induction is due, at least in part, to MEK/ERK1/2 activity. UO126 and PD98059 did not affect basal HIF-1α expression. In addition to increasing HIF-1α expression, ERK1/2 has previously been reported to phosphorylate HIF-1α in vitro, and this post-translational modification can increase HIF-1α activity, presumably by impeding proteasome/von-Hippel-Lindau (VHL) recognition [20]. Although very little is known about the phosphorylation of HIF-1α, this phosphorylation event may be important for optimal activity of the HIF-1 pathway.

HIF-1α induction by OA-NO2 contributes to intracellular Ca2+ accumulation and AMPK activation

To examine the involvement of the HIF-1α pathway in OA-NO2-stimulated AMPK activation, we employed the widely used HIF-1α inhibitor, 3-(5-hydroxymethyl-2-furyl)-1-benzylindazole (YC-1) and HIF-1α siRNA. Pre-treatment of BAECs with YC-1 or HIF-1α siRNA significantly inhibited the OA-NO2-induced increase in AMPK Thr172 phosphorylation levels relative to OA-NO2 treatment alone (Figure 5A and B). Since OA-NO2-induced AMPK activation is LKB1-independent, we postulated that the Ca2+/CaMKK pathway plays a role in the activation of AMPK by OA-NO2/HIF-1α. In addition, because HIF-1α has been reported to regulate Ca2+ homeostasis in pulmonary arterial smooth muscle cells via upregulation of store-operated Ca2+ channels and enhanced Ca2+ influx [24], we examined a possible role for HIF-1α in this process. To determine whether OA-NO2 affected endothelial cell [Ca2+]i, we measured intracellular fluorescence in Fluo-4–loaded BAECs. We found that OA-NO2 treatment produced an approximately 1.4- to 2.2-fold increase in [Ca2+]i at 6 and 16 h after treatment, respectively (Figure 5C). To confirm a role for HIF-1α in this OA-NO2-mediated [Ca2+]i increase, we used pharmaceutical- and genetic-inhibition strategies to downregulate HIF-1α. Pre-treatment of BAECs with YC-1 or transfection of these cells with HIF-1α siRNA partially suppressed the OA-NO2-induced increases in [Ca2+]i (Figure 5D), indicating that this process is at least partially dependent on HIF-1α. We
further investigated whether a Ca\textsuperscript{2+} signal is involved in OA-NO\textsubscript{2}-induced AMPK activation using BAPTA-AM, a cell-permeable Ca\textsuperscript{2+} \textquoteleft sponge\textprime. Our results indicate that chelation of intracellular Ca\textsuperscript{2+} by BAPTA-AM (25 \textmu M, 30 min) significantly inhibited OA-NO\textsubscript{2}-stimulated AMPK phosphorylation (~60\%, \(p<0.05\); Figure 5E). Together, these data show that intracellular Ca\textsuperscript{2+} is necessary for OA-NO\textsubscript{2}-induced AMPK activation.

CaMKK\textbeta mediates OA-NO\textsubscript{2}-induced AMPK activation

Both the tumor suppressor LKB1 [14] and CaMKK [15] are important AMPK kinases as each activates AMPK by directly phosphorylating the AMPK \alpha subunit on Thr172. As shown above (Figure 2A–C), LKB1 is unlikely involved in the activation of AMPK by OA-NO\textsubscript{2}. Because treatment with OA-NO\textsubscript{2} elevated [Ca\textsuperscript{2+}], and OA-NO\textsubscript{2}-stimulated AMPK activation was Ca\textsuperscript{2+} sensitive, we speculate that the AMPK kinase, CaMKK\textbeta, which is activated by Ca\textsuperscript{2+}/calmodulin binding, may be involved. To test this hypothesis, we used the relatively selective CaMKK\textalpha and CaMKK\textbeta inhibitor, STO-609 (1 \textmu M) [25] and the competitive CaM inhibitor, KN-93 (3 \textmu M). Either STO-609 or KN-93 was sufficient to prevent activation of AMPK by OA-NO\textsubscript{2} (Figure 6A), suggesting the involvement of CaMKK in this activation. A specific role for CaMKK\textbeta was indicated by the results of siRNA experiments, which showed that downregulation of CaMKK\textbeta caused a significant reduction in OA-NO\textsubscript{2}-stimulated AMPK phosphorylation (Figure 6B). Downregulation of CaMKK\textbeta protein was verified by western blot analysis. These results suggest that CaMKK\textbeta is the major AMPK kinase under these conditions. To further validate this notion, we determined whether treatment with OA-NO\textsubscript{2} enhanced the association between CaMKK and AMPK as this interaction is a prerequisite for AMPK phosphorylation/activation. Indeed, following OA-NO\textsubscript{2} treatment, an enhanced CaMKK and AMPK association was detected by immunoprecipitation with either AMPK or CaMKK antibodies (Figure 6C). Moreover, pre-treatment with YC-1 dramatically inhibited the OA-NO\textsubscript{2}-induced increased CaMKK-AMPK association, further substantiating the indispensable role of CaMKK-AMPK association, further substantiating the indispensable role of CaMKK-AMPK association, further substantiating the indispensable role of CaMKK-AMPK association, further substantiating the indispensable role of CaMKK-AMPK association, further substantiating the indispensable role of CaMKK-AMPK association.

OA-NO\textsubscript{2}-induced eNOS phosphorylation is dependent on AMPK

We previously demonstrated that AMPK phosphorylates and activates eNOS in cultured endothelial cells [26]. Similarly, Zhang et al. demonstrated that infection of endothelial cells with a recombinant adenovirus expressing constitutively active AMPK resulted in eNOS activation and increased NO production [27]. Here, we used phosphorylation of Ser1177 in eNOS, which is a reported substrate of AMPK, as an indicator of AMPK activity in this system. As shown in Figure 7A, incubation of BAECs with OA-NO\textsubscript{2} increased eNOS Ser1177 phosphorylation in a dose-dependent manner that was very similar to that for AMPK phosphorylation. The time course of OA-NO\textsubscript{2}-induced eNOS phosphorylation was also similar to that for AMPK phosphorylation (Figure 7B). To confirm that OA-NO\textsubscript{2}-stimulated eNOS phosphorylation involved AMPK, we infected BAECs with adenovirus encoding a dominant-negative form of AMPK (Ad-DN-AMPK). As expected, phosphorylation of both AMPK and eNOS was increased after treatment of control BAECs (Ad-GFP-infected or non-infected BAECs) with 2.5 \textmu M OA-NO\textsubscript{2} for 16 h (Figure 7C). In contrast, overexpression of Ad-DN-AMPK completely abolished OA-NO\textsubscript{2}-induced eNOS phosphorylation. Consistent with a role for AMPK in phosphorylating eNOS, treatment with STO-609 or KN-93 to inhibit the AMPK kinase CaMKK or downregulation of CaMKK\textbeta using siRNA prevented OA-NO\textsubscript{2}-induced eNOS phosphorylation (Figure 6A and B).

Both Akt and AMPK are capable of phosphorylating eNOS at Ser1179 [28]. Thus, we determined whether Akt also contributes to OA-NO\textsubscript{2}-induced eNOS phosphorylation. OA-NO\textsubscript{2} did not increase basal Akt phosphorylation of Ser473 (data not shown), suggesting that OA-NO\textsubscript{2}-stimulated eNOS phosphorylation does not require Akt but depends on activation of AMPK.
Next, we determined whether OA-NO2-induced eNOS phosphorylation was associated with increased NO release. Treatment with OA-NO2 significantly increased NO release, an effect that was inhibited by Ad-DN-AMPK transfection (Figure 7D). Importantly, Ad-DN-AMPK transfection reduced AMPK activity in OA-NO2-treated cells to below control levels (Figure 7E). These results indicate that OA-NO2 increases NO release via AMPK activation.

Discussion

OA-UFAs are unique signaling mediators that are present in a variety of cell types, including endothelial cells [29]. Considerable evidence points to a role for circulating OA-UFAs in vascular-protective effects [9], although the mechanisms by which OA-NO2 are incompletely understood. In this study, we report that OA-NO2 activates AMPK in endothelial cells via a Ca2+ dependent pathway, and we implicate CaMKKβ as the responsible upstream kinase. Our results show that HIF-1α is an inducer of the intracellular Ca2+ mobilization that leads to AMPK activation. On the basis of the results of experiments with an HO-inhibitor and HO-1 siRNA, we also implicate HO-1 in OA-NO2-induced upregulation of endothelial HIF-1α.

HO is a rate-limiting enzyme in heme degradation and functions to convert heme to biliverdin, carbon monoxide (CO) and iron. Human HO occurs in two main isoforms, the inducible HO-1 form and the constitutive HO-2 form. Previous studies demonstrated that HO-1 exerts anti-inflammatory effects [30], including prolongation of cardiac xenograft graft survival [31] and inhibition of leukocyte transendothelial migration during complement-dependent inflammation [32] and in response to low-density lipoprotein (LDL) oxidation [33]. More importantly, once induced, HO-1 also confers vascular cytoprotection [30]. The importance of this result is demonstrated by the severe and persistent endothelial damage observed in the case of human HO-1 deficiency [34] and in gene-targeted mice deficient in HO-1 [35]. Furthermore, induction of HO-1 may directly regulate endothelial cell activation, preventing adhesion molecule expression and chronic graft rejection [36]. In vitro, HO-1 protects endothelial cells from hydrogen peroxide–mediated cell death [37] and from tumor necrosis factor α (TNFα) cytotoxicity [38]. In addition, HO-1 has been suggested to play a role in angiogenesis.

Figure 5. HIF-1α mediates OA-NO2-induced intracellular Ca2+ accumulation and AMPK activation. A) BAECs were treated with YC-1 (30 μM) for 30 min followed by incubation with OA-NO2 for the indicated times. AMPK phosphorylation and protein levels were assayed as described in Materials and Methods. The blot is representative of three blots obtained from three independent experiments. B) BAECs were incubated with HIF-1α-specific siRNA or control siRNA for 48 h and then treated with OA-NO2 or vehicle for 16 h. After treatment, cell lysates were analyzed for HIF-1α and AMPK protein levels and AMPK phosphorylation at Thr172. Lower panels: summary data (*p<0.05 vs. control; *p<0.05 vs. OA-NO2 group; n=3). C) BAECs were incubated with OA-NO2 (2.5 μM) for the indicated times. Intracellular Ca2+ was measured with Fluo-4 fluorescent dye as described in Materials and Methods. *p<0.01 vs. control (n=4). D) BAECs were pre-treated with YC-1 (30 μM) or BAPT-AM (25 μM) for 30 min or HIF-1α siRNA or control siRNA for 48 h followed by incubation with OA-NO2 for 16 h. After treatment, intracellular Ca2+ was measured in intact cells using a Fluo-4 NW kit. *p<0.05 vs. control; *p<0.05 vs. OA-NO2 group (n=3). E) BAECs were pre-loaded with 25 μM BAPT-AM for 30 min prior to incubation with 2.5 μM OA-NO2 for 16 h. AMPK protein levels and phosphorylation at Thr172 were detected as described above. Representative blots (top) and densitometric analyses (bottom) are shown. Values are means ± SD from three independent measurements. *p<0.05 vs. control; #p<0.05 vs. OA-NO2 group.

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AMPK-Dependent Angiogenesis by Nitrated Lipids

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Figure 6. CaMKK is the upstream AMPKK that mediates OA-NO2-induced AMPK activation. A) HUVECs were treated with STO-609 (1 μM) or KN-93 (3 μM) for 1 h followed by incubation with OA-NO2 for 16 h. AMPK and eNOS phosphorylation and protein expression were assayed as described in Materials and Methods. The blot is representative of three blots obtained from three independent experiments. Lower panels: summary data (*p<0.05 vs. control; **p<0.05 vs. OA-NO2 group; n=3). B) HUVECs were incubated with CaMKKβ-specific siRNA or control siRNA for 48 h and then treated with OA-NO2 for 16 h. After treatment, cell lysates were analyzed for AMPK and eNOS phosphorylation and protein levels. Lower panels: summary data (*p<0.05 vs. control; **p<0.05 vs. OA-NO2 group; n=3). C) HUVECs were pre-treated with YC-1 (30 μM) for 30 min followed by incubation with OA-NO2 for 16 h. An immunoblot of AMPK precipitated with an anti-CaMKK antibody is shown. The blot is representative of three blots obtained from three independent experiments. Lower panels: summary data (*p<0.05 vs. control; **p<0.05 vs. OA-NO2 group; n=3).

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Figure 7. AMPK mediates OA-NO2-induced eNOS phosphorylation and NO production in BAECs. BAECs were treated with (A) different concentrations of OA-NO2 for 16 h or (B) 2.5 μmol/L OA-NO2 for the indicated times. Lysates were analyzed by western blot for the indicated proteins. The blot is representative of three blots obtained from three separate experiments. C) Western blot of phosphorylated AMPK and eNOS in OA-NO2-stimulated BAECs infected with Ad-DN-AMPK (MOI = 50). Non-infected cells or cells infected with Ad-GFP served as controls. For A–C, the corresponding densitometric analyses are shown. *p<0.05 vs. control; **p<0.05 vs. OA-NO2-treated group. D) NO release by OA-NO2-stimulated BAECs infected with Ad-DN-AMPK (MOI = 50) or Ad-GFP (control). *p<0.05 vs. non-transfected, no OA-NO2 group; **p<0.05 vs. OA-NO2-treated, Ad-GFP group. E) AMPK activity corresponding to C and D above. *p<0.05 vs. no OA-NO2 treatment, Ad-GFP group; **p<0.05 vs. OA-NO2-treated, Ad-GFP group. F) The proposed signaling pathway involved in AMPK/eNOS activation in response to OA-NO2 treatment in endothelial cells.

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This supposition is supported by the observation that overexpression of HO-1 induces proliferation and formation of capillary-like structures [39]. Thus, therapeutic induction of HO-1 may be beneficial in the treatment of chronic inflammatory diseases as well as cardiovascular diseases. Our results indicate that OA-NO2 potently induced HO-1 protein expression in endothelial cells. In agreement with our studies, other studies have suggested that HO-1 expression is induced in human aortic endothelial cells (HAECs) by LNO2, which shows a higher potency in this context than other established stimuli, including oxidized fatty acids and hemin. This induction of HO-1 expression by NO2-FA is not mediated by NO, NF-oxidized fatty acids and hemin. This induction of HO-1 potency in this context than other established stimuli, including the AP-1 sequences in the 4.5 kb HO-1 promoter region [6,40]; however, the mechanisms involved in these events remain poorly characterized, and the identities of the signaling molecules downstream of HO-1 that mediate the vascular protective effects of nitrated lipids are not entirely known.

The novel finding of this study is that OA-NO2 induces HIF-1x expression via HO-1 under non-hypoxic conditions. HIF-1, which exists as a heterodimer composed of the HIF-1 subunits, has been shown to mediate numerous physiological and pathophysiological responses to hypoxia. Under normoxic conditions, however, HIF-1x is ubiquitinated and rapidly degraded [42] and is thus present at very low levels under these conditions. Under hypoxic conditions, the HIF-1x subunit is induced. Because HIF-1x is constitutively expressed, HIF-1x is responsible for conferring hypoxia sensitivity to heterodimeric HIF-1. Using specific chemical inhibitors, we demonstrated that ERK1/2 partially mediated HO-1-induced HIF-1x expression (Figure 4). In addition to inducing HIF-1x expression, ERK1/2 has been reported to phosphorylate HIF-1x and to thereby increase its activity [20]. The possibility that CO, a metabolite of HO-1 [43], also plays a role in HIF-1x induction by HO-1 cannot be excluded by our data. A recent study by Chih et al. [44] suggested that exposure of macrophages to CO resulted in rapid HIF-1x activation and stabilization, which regulates the expression of genes involved in inflammation, metabolism, and cell survival. This previous study also provided evidence that CO may serve as a signaling intermediary between HO-1 and HIF-1x.

Another important finding of the present study is that HIF-1x contributes to an increase in [Ca2+]i, which is responsible for CaMKKβ-dependent AMPK activation (Figure 5 and 6). Pharmaceutical and genetic inhibitors of HIF-1x suppressed OA-NO2-induced increases in [Ca2+]i, suggesting that HIF-1x is involved in the OA-NO2-induced mobilization of Ca2+ in endothelial cells. Previous studies demonstrated that hypoxia evokes an increase in [Ca2+]i, in endothelial cells [43], and presumably, the induction of HIF-1x contributes to this increase in [Ca2+]i, under hypoxic conditions or under normoxic conditions in the presence of induction factors that activate HIF-1x. The mechanisms that are involved in endothelial Ca2+ homeostasis within the vasculature following HIF-1x activation are also largely unknown. Although HIF-1x has been reported to regulate Ca2+ homeostasis in pulmonary arterial smooth muscle cells via upregulation of store-operated Ca2+ channels and enhanced Ca2+ influx [24], the detailed mechanisms remain to be elucidated.

Our results also showed that OA-NO2-stimulated AMPK activation was inhibited by chelation of intracellular free Ca2+, selective inhibition of CaMKK by STO-609, selective inhibition of Ca2+/calmodulin-dependent protein kinase by KN-93, and siRNA-mediated silencing of CaMKK-β expression (Figure 6). In addition, the association between AMPK and CaMKK was enhanced by OA-NO2 treatment. Taken together, these data indicate that the HIF-1x/Ca2+/CaMKK- pathway is crucial for OA-NO2-induced AMPK activation. Furthermore, CaMKK-mediated AMPK activation in endothelium has also recently been reported in response to thrombin [46], adenossine diphosphate (ADP) [47], and bradykinin [48].

Results from previous studies indicated that endothelial AMPK may play an important physiological role in the function of both endothelial cells and the cardiovascular system as a whole; thus, activation of AMPK may provide an explanation for the beneficial effects of OA-NO2 on these systems (see review [49]). Our findings show that induction of the CaMKK/AMPK pathway by OA-NO2 in endothelial cells may have both physiological and therapeutic relevance. First, endothelial AMPK activation by OA-NO2 activates nitric oxide synthase (via phosphorylation on Ser 1177) and elevates NO bioavailability, and these effects may not only protect against early events in atherogenesis, such as white cell adherence [50], but may also prevent later steps in atherogenesis, including fibrous plaque formation. Endothelial NO likely represents the most important anti-atherogenic defense molecule in the vasculature [50]. Nitrated lipids act as NO donors in vitro and are widely considered to be a possible endogenous source of NO [51]. Enhanced NO production under hyperlipidemic and hypercholesterolemic conditions, such as those that occur with obesity or insulin-resistant status, however, has been associated with low NO bioactivity [52]. Therefore, it is reasonable to postulate that the presence of these nitrated products and the related AMPK/eNOS/NO pathway in vivo may actually play a compensatory role, providing an adjustable supply of NO to compensate for the impaired NO bioactivity and endothelial-dependent vasorelaxation that is characteristic of the early steps of vascular disease. Second, AMPK signaling acts as a novel regulator of angiogenesis and is specifically required for endothelial cell migration and differentiation under conditions of hypoxia [53] or in response to adiponectin [54]. Additionally, AMPK-dependent eNOS activity is required for adequate endothelial tube formation [54]. As described above, increased levels of nitrated lipids are formed in the context of the hyperlipidemia associated with obesity and insulin resistance. Thus, the induction of AMPK and angiogenesis by nitrated lipids may also represent an adaptive defense mechanism against impaired angiogenesis and/or vascular injury caused by obesity-related dislipidemia. The role that activated AMPK plays in increasing fatty acid oxidation via phosphorylation and inhibition of ACC and leading to a decrease in the concentration of malonyl-CoA is most important [55]. In
addition, AMPK decreases fatty acid incorporation into glycerolipids in some tissues, either secondary to its effect on fatty acid oxidation or via phosphorylation and inhibition of sn-glycerophosphate acyltransferase, the first committed enzyme in diacylglycerol and triglyceride synthesis [56]. Furthermore, endothelial AMPK activity may inhibit glycerol-3-phosphate acyltransferase, which is required for de novo synthesis of diacylglycerol [56]. Thus, AMPK may lessen endothelial diacylglycerol production (and thus protein kinase C activation) both by reducing the availability of the FFA substrates required for this synthesis and by directly inhibiting the enzyme that catalyzes it.

In conclusion, we have demonstrated for the first time that NO$_2$-UFAs activate AMPK in endothelial cells by a mechanism that depends on an increase in HO-1 followed by HIF-1α protein expression and Ca$^{2+}$/CaMKK activation. Our results also indicate that AMPK activity is required for eNOS/NO production in endothelial cells (Figure 7F). The present study further suggests that AMPK activation by nitrated lipids may play an essential role in compensating or protecting vascular endothelial function against vascular injury in obesity-related dyslipidemia. AMPK might be a valid therapeutic target for treating vascular disorders in obesity and type II diabetes.

Materials and Methods

Materials

BAECs and cell culture media were obtained from Clonetics Inc. (Walkersville, MD). HUVECs and cell culture media were purchased from Cascade Biologies (Portland, OR). FFA-free bovine serum albumin (BSA) and oleic acid were obtained from Sigma (St. Louis, MO). ZnBG was purchased from Porphyrin Products, Inc. (Logan, UT). OA-NO$_2$ was obtained from Cayman Chemical (Ann Arbor, MI), and AICAR was purchased from Toronto Research Chemicals (Toronto, Canada). The MEK inhibitors PD 98059 and UO 126 were obtained from Calbiochem (La Jolla, CA). 3-(5-9-furyl)-1-benzylindazole (YC-1) was purchased from AG Scientific Inc. (San Diego, CA). Antibodies against phospho-ACC (Ser79), phospho-AMPK (Thr172), AMPK, phospho-LKB1 (Ser428), LKB1, and phospho-eNOS (Ser1177) were purchased from Cell Signaling (Beverly, MA). The antibodies against ACC were obtained from Alpha Diagnostic International (San Antonio, TX). All other chemicals and organic solvents were of the highest grade and were obtained from Sigma.

Cell culture and adenoviral infection

BAECs were grown in EBM supplemented with 2% fetal bovine serum and growth factors. HUVECs were maintained in Medium 200 supplemented with a low-serum growth supplement before use. All cell culture media were supplemented with both penicillin (100 U/ml) and streptomycin (100 µg/ml). Cells between passages 5 and 10 were used for all experiments. All cells were incubated in a humidified atmosphere of 5% CO$_2$/95% air at 37°C. BAECs were infected with adenovirus encoding a dominant-negative mutant form of AMPKα (Ad-DN-AMPK) or green fluorescence protein (Ad-GFP) as a control. Infections were performed in 80% confluent cultures of BAECs in media containing 0.1% fetal bovine serum and recombinant adenovirus at the indicated multiplicity of infection (MOI). Cultures were incubated with adenoviruses for 48 h before experimentation. Using these conditions, infection efficiency was typically at least 80%, as determined by GFP expression.

SiRNA-mediated gene silencing in endothelial cells

HUVECs or BAECs were transfected with LKB1 siRNA, HO-1 siRNA, HIF-1α siRNA, CaMKKβ, or the corresponding scrambled siRNA (negative control) for 48 h using Lipofectamine™ 2000 (Invitrogen) according to the manufacturer’s instructions. Infected cells were starved in serum-free medium for 6 h, then exposed to the indicated concentrations of OA-NO$_2$ or vehicle for 24 h.

Measurement of NO production

For NO detection, BAECs grown in 24-well plates were incubated for 30 min in the presence of 15 µM 4,5-diaminofluorescein diacetate (DAF-2 DA) in PBS or in PBS alone (control) in the dark at 37°C. Cells were then washed with PBS to remove excess DAF-2 DA, and the change in fluorescence over 15 min was measured with excitation and emission wavelengths of 485 and 530 nM, respectively, at room temperature using a microplate reader (FL 600, Bio-Tek). Changes in fluorescence were also visualized with a fluorescence microscope (Olympus IX71), and images were captured for analysis [57].

Measurement of intracellular Ca$^{2+}$

[Ca$^{2+}$], was measured using a Fluo-4 NW kit from Invitrogen according to the manufacturer’s instructions. In brief, BAECs were treated with OA-NO$_2$, control or HIF-1α siRNA, YC-1, or BAPTA-AM. The culture medium was then aspirated, cells were washed once with Hepes buffer (pH 7.4), and 1 ml of Hepes buffer with fluorescent dye was added to the cells. After the cells were incubated for 30 min, the fluorescence intensity was measured with excitation and emission wavelengths of 485 and 530 nM, respectively.

Western blot analysis

BAECs, HUVECs, or Hela-S3 cells were lysed in cold RIPA buffer. Protein concentrations were determined using a bicinchoninic acid protein assay system (Pierce, Rockford, IL). Proteins were analyzed by western blotting with ECL-Plus detection as described previously [58]. Relative PPARγ protein expression was measured in HUVEC nuclear extracts as previously detailed [59].

AMPK activity assay

AMPK activity was assayed in the presence and absence of AMP (200 µM) using the SAMS peptide, as previously described [60]. AMPK activity was calculated by determining the difference in activity between both conditions.

Statistical analysis

Statistical comparisons of vasodilation were performed using a two-way analysis of variance (ANOVA), and intergroup differences were analyzed using Bonferroni’s post-hoc test. Time-course studies were analyzed using a repeated-measure ANOVA. All other results were analyzed using a one-way ANOVA. Values are expressed as mean ± SD. P-values less than 0.05 were considered significant.

Author Contributions

Conceived and designed the experiments: YW MZ. Performed the experiments: YW YD PS. Analyzed the data: YW YD PS MZ. Wrote the paper: YW MZ.
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