Efficient Testing of Large Pools of Mycobacterium tuberculosis RD1 Peptides and Identification of Major Antigens and Immunodominant Peptides Recognized by Human Th1 Cells

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Comparative genomics has identified several regions of difference (RDs) of Mycobacterium tuberculosis that are deleted or absent in Mycobacterium bovis BCG vaccines. To determine their relevance for diagnostic and vaccine applications, it is imperative that efficient methods are developed to test the encoded proteins for immunological reactivity. In this study, we have used 220 synthetic peptides covering sequences of 12 open reading frames (ORFs) of RD1 and tested them as a single pool (RD1pool) with peripheral blood mononuclear cells obtained from pulmonary tuberculosis (TB) patients and M. bovis BCG-vaccinated healthy subjects in TH1 cell assays that measure antigen-induced proliferation and IFN-γ secretion. The results showed that RD1pool induced strong responses in both TB patients and BCG-vaccinated healthy subjects. The subsequent testing of peptide pools of individual ORFs revealed that all ORFs induced positive responses in a portion of donors, but PPE68, CFP10, and ESAT6 induced strong responses in TB patients and PPE68 induced strong responses in BCG-vaccinated healthy subjects. In addition, HLA-DR and -DQ typing of donors and HLA-DR binding prediction analysis of proteins suggested HLA-promiscuous presentation of PPE68, CFP10, and ESAT6. Further testing of individual peptides showed that a single peptide of PPE68 (121-VLTATNFFGINTIPIALT) was immunodominant. The search for sequence homology revealed that a part of this peptide, 124-ATNFFGINTIPIAL-137, was present in several PPE family proteins of M. tuberculosis and M. bovis BCG vaccines. Further experiments limited the promiscuous and immunodominant epitope region to the 10-amino-acid cross-reactive sequence 127-FFGINTIPIA-136.
ORFs (23), the genomic prediction by Gordon et al. predicted 9 ORFs (16), and Amoudy et al. predicted the presence of 20 ORFs (ORF1 to ORF20) in RD1, of which 14 ORFs (ORF2 to ORF15) were deleted in M. bovis BCG (7) (Table 1). As all of the ORFs predicted by Amoudy et al. are expressed in M. tuberculosis at the mRNA level (7), this prediction was considered appropriate to test for immunological reactivity.

To test immunological relevance of various RD1 ORFs, we have previously attempted to express six ORFs (ORF10 to ORF15) in Escherichia coli and purify the recombinant proteins (1). However, due to several problems including the inability to express the ORFs in E. coli and extensive degradation of expressed proteins, we could purify only two of the targeted proteins, i.e., ORF11 (Rv3878) and ORF14 (1 and unpublished data). When tested with human serum, ORF14 was found to be a major antigen recognized by antibodies in sera of TB patients (1). Interestingly, ORF14 was predicted only by Amoudy et al. but not by others (7) (Table 1), and several studies have confirmed that ORF14 is a real protein coding gene of M. tuberculosis that is expressed under both in vitro and in vivo growth conditions (1, 8, 11). These findings suggest that to identify additional antigens of diagnostic and/or vaccine relevance, a systematic screening of all RD1 ORF proteins should be performed for immunological reactivity. However, such a study for antibody reactivity can be done only with full-length proteins as the epitopes recognized by antibodies are mostly conformational in nature whereas, in the case of T-cell reactivity, the problem of nonavailability of full-length proteins can be overcome by using synthetic peptides because T cells recognize mostly structural epitopes (reviewed in reference 28).

A number of studies have demonstrated the validity of the synthetic peptide approach for Th1 cell reactivity by using nonselected mixtures of overlapping synthetic peptides spanning the sequences of antigens like CFP10, ESA16, Ag85B, and MPB70, for example. (3, 6, 22, 40). In these studies, it was shown that Th1 cell reactivities with the peptide mixtures were equivalent to the reactivities induced by the corresponding full-length proteins, thereby suggesting that overlapping synthetic peptides could replace complete antigens in T-cell responses. However, the tested pools have been comprised of peptides corresponding to single antigenic proteins (6, 10, 22, 40), whereas testing of RD peptides would require pools of hundreds of peptides covering the sequence of several putative proteins in each RD (16). To make T-cell assays feasible for testing peptides covering several protein regions, it is therefore essential that systems are established to test pools of hundreds of peptides in a single well of a 96-well plate. This requirement is more compelling now because the availability of the complete genome sequences of a large number of pathogenic organisms and the prediction of encoded ORFs require their functional characterization in terms of immunological reactivity for application in diagnosis and vaccine development. Furthermore, although selected ORFs of RD1, i.e., ESA16, CFP10, Rv3873, Rv3878, and Rv3879c, have been tested in the past using pools of synthetic peptides (10, 22), a comprehensive analysis of RD1 ORFs for Th1 cell reactivity, and especially the ORFs that were predicted exclusively by Amoudy et al., is lacking in humans.

The aim of the present study was to use 220 overlapping synthetic peptides covering the sequence of 12 RD1 ORFs (Table 1) as a pool (RD1 pool) to determine the possibility of using pools of large numbers of peptides in Th1 cell assays. In addition, we systematically screened the peptide pool of each RD1 ORF to identify major Th1 cell antigens recognized by peripheral blood mononuclear cells (PBMC) from HLA-heterogeneous TB patients and M. bovis BCG-vaccinated healthy subjects living in Kuwait. The promiscuous peptides of major antigens were identified by HLA-DR prediction analysis using the ProPred server, and immunodominant peptides recognized by Th1 cells were identified by testing individual peptides with PBMC from HLA-heterogeneous subjects.

**MATERIALS AND METHODS**

**Study population.** TB patients included in the study were newly diagnosed and culture-confirmed cases of pulmonary TB (n = 40) attending the Chest Diseases Hospital, Kuwait. M. bovis BCG-vaccinated healthy subjects (n = 38) were randomly selected from the blood donors at the Central Blood Bank, Kuwait. All

| Gene designation | ORF annotation by source (reference no.) | Position on RD1 (nucleotide no.) | Length of protein (aa) | No. of synthetic peptides |
|------------------|-----------------------------------------|---------------------------------|----------------------|------------------------|
| Mahairas et al. (23) | Gordon et al. (16) | Amoudy et al. (7) |
| Rv3871 | ORF1A | Rv3871 | ORF2 | 1-1776 | 591 | 39 |
| PE35 | NP | Rv3872 | ORF3 | 1919-2218 | 98 | 6 |
| orf4 | NP | NP | ORF4 | 3126-2707 | 139 | 9 |
| PPE68 | ORF1B | Rv3873 | ORF5 | 2240-3355 | 371 | 24 |
| exsB/cfp10 | NP | Rv3874 | ORF6 | 3448-3750 | 100 | 6 |
| exsA/esat6 | NP | Rv3875 | ORF7 | 3783-4070 | 95 | 6 |
| orf8 | Rv3876 | ORF1C | Rv3876 | 4184-6184 | 666 | 44 |
| Rv3877 | ORF1D | Rv3877 | ORF9 | 6058-7716 | 552 | 36 |
| Rv3878 | ORF1E | Rv3878 | ORF10 | 7867-8709 | 280 | 18 |
| orf12 | ORF1F | Rv3879 | ORF11 | 8890-10539 | 563 | ND |
| Rv3879c | ORF1G | NP | ORF12 | 10956-8767 | 746 | ND |
| orf14 | NP | Rv3879 | ORF13 | 9689-10426 | 262 | 17 |
| orf15 | NP | NP | ORF14 | 10792-11079 | 95 | 6 |

*a* NP, not predicted; ND, not determined.

*b* Positions are from reference 16.
the patients and healthy donors were serologically negative for human immunodeficiency virus, were PPD skin test positive (as determined with tuberculin PPD RT23 from the Statens Serum Institute, Copenhagen, Denmark), and included both Kuwaiti and non-Kuwaiti citizens residing in Kuwait. Informed consent was obtained from all the subjects, and the study was approved by the Ethical Committee of the Faculty of Medicine, Kuwait University, Kuwait.

**Complex mycobacterial antigens.** The complex mycobacterial antigens used in this study were irradiated whole-cell M. tuberculosis H37Rv (40), M. bovis BCG (34), and M. tuberculosis culture filtrate (provided by J. T. Belisle, Fort Collins, CO, and produced under NIH contract HHSN266200400091C/ADB contract AI40092, Tuberculosis Vaccine Testing and Research Materials Contract).

**Synthetic peptides.** Thirteen synthetic peptides (25-mers overlapping neighboring peptides by 10 amino acids [aa]) spanning the sequence of MPB70 (6) and 220 peptides spanning the sequence of 12 RD1 ORFs (Table 1) were synthesized using fluoroenylmethoxycarbonyl chemistry, as described previously (6, 39). For all peptides, identity was confirmed by mass spectroscopy, and purity was more than 70%. As an example, the sequences of peptides corresponding to PPE68 are shown in Fig. 1. The stock concentrations (5 mg/ml) of the peptides were prepared in normal saline (0.9%), as described previously (37, 39), and frozen at −70°C until used. The working concentrations were prepared by thawing the peptide pools, and individual peptides in 50 μl of complete tissue culture medium (RPMI 1640 with 10% FCS, and 2.5 μg of gentamicin per ml). The final volume of the culture in the wells was adjusted to 200 μl. The plates were incubated at 37°C in a humidified atmosphere of 5% CO₂ and 95% air. The cultures were pulsed (4 h) on day 6 with 1 μCi of [3H]thymidine (Amersham Life Sciences, Little Chalfont, United Kingdom) and harvested on filter mats with a Skatron harvester (Skatron Instruments AS, Oslo, Norway), and the radioactivity incorporated was measured by liquid scintillation counting (4, 30). The radioactivity incorporated was obtained as counts per minute (cpm). Average cpm were calculated from triplicate cultures stimulated with each antigen or peptide. Cellular proliferation results are presented as the stimulation index (SI), which is defined as follows: SI = number of cpm in antigen-stimulated cultures/number of cpm in cultures without antigen. The SI values of ≥± and ±3 were considered positive proliferative responses against complex mycobacterial antigens and peptides, respectively (6, 40).

**IFN-γ assay.** Supernatants (100 μl) were collected from cultures of PBMC (96-well plates) before they were pulsed with [3H]thymidine. The supernatants were kept frozen at −70°C until they were assayed for IFN-γ activity. The amount of IFN-γ in the supernatants was quantified by using Immunotech immunoassay kits (Immunotech SAS, Marseille, France) as specified by the manufacturer. The detection limit of the IFN-γ assay kit was 0.08 IU/ml. Secretion of IFN-γ in response to a given antigen or peptide was considered positive when the delta IFN-γ value (the IFN-γ concentration in cultures stimulated with antigen or peptide minus the IFN-γ concentration in cultures without antigen or peptide) was ≥±5 U/ml in response to complex antigens and ±±3 U/ml in response to peptides (3).

**Interpretation of antigen-induced proliferation and IFN-γ secretion results and statistical analysis.** An antigen/peptide was considered a strong, moderate, or weak stimulator of PBMC in proliferation and IFN-γ/secretion assay. The responses were considered strong when the median SI and IFN-γ value of >±5 and a percent positive value of ≥60%; moderate with a median SI and IFN-γ value of ±3 to ±5 and a percent positive value of ±40%; and weak with a median SI and IFN-γ value of ≤±3 or ±±3 and a percent positive value of <±40%. The results of proliferation and IFN-γ secretion in response to various antigens/peptides in the two donor groups were analyzed statistically for significant differences using a Pearson chi-square test for two proportions, and P values of <0.05 were considered statistically significant.

**HLA typing of PBMC.** PBMC were HLA typed genomically by isolating the high-molecular-weight genomic DNA from each PBMC by treatment of the cells with proteinase K and salting out in miniscale, as described previously (3, 39). The amount of DNA obtained was quantified by spectrophotometry. An HLA-DR “low-resolution” kit containing the primers to type for DRB1, DRB3,
DBR4, and DRB5 alleles and an HLA-DQ low-resolution kit to type for DQB1 alleles were purchased from Dynal AS (Oslo, Norway) and were used in PCR as described by the manufacturer. DNA amplification was carried out with a GeneAmp 2400 PCR system (Perkin Elmer Cetus), and the amplified products were analyzed by gel electrophoresis using standard procedures. Serologically defined HLA-DR and -DQ specificities were determined from the genotypes by following the guidelines provided by Dynal AS.

### ProPred analysis for promiscous binding regions in PPE68 protein
HLA-DR binding propensities along the primary structure of PPE68 and its individual peptides were detected by ProPred analysis at the default setting (threshold value of 3.0) using the server (http://www.imtech.res.in/raghava/propred/) (48). This server has been suggested to be a useful tool in locating the promiscuous binding regions that can bind to a total of 51 alleles belonging to nine serologically defined HLA-DR molecules (3, 30, 39, 48). These HLA-DR molecules are encoded by DRB1 and DRB5 genes including HLA-DR1 (2 alleles), DR3 (7 alleles), DR4 (9 alleles), DR7 (2 alleles), DR8 (6 alleles), DR11 (9 alleles), DR13 (11 alleles), DR15 (3 alleles), and DR51 (2 alleles). The server performs analysis for each of these alleles independently and computes the binding strength of all the peptides. The peptides predicted to bind >50% of the binding regions of the alloalleles included in the ProPred analysis were considered promiscuous for binding (39).

### RESULTS

**Antigen-induced proliferation and IFN-γ secretion by PBMC of TB patients and BCG-vaccinated healthy subjects to complex antigens, MPB70, and RD1<sub>pool</sub>.
**PBMC from 40 culture-confirmed pulmonary TB patients and 30 M. bovis BCG-vaccinated healthy subjects were tested for antigen-induced proliferation and IFN-γ secretion against complex mycobacterial antigens and the peptide pools of MPB70 and RD1<sub>pool</sub>. In response to the complex mycobacterial antigens, PBMC from each donor responded to at least one of these antigens in both assays (data not shown). Furthermore, strong antigen-induced proliferation was observed with PBMC from TB patients (median SI, 16 to 25; percent positive, 70 to 85%) as well as healthy subjects (median SI, 14 to 17; percent positive, 75 to 90%) in response to all three complex mycobacterial antigens. The antigen-induced proliferation responses to MPB70 were moderate in both TB patients (median SI, 3.1; percent positive, 50%) and healthy subjects (median SI, 3.9; percent positive, 47%), whereas in response to RD1<sub>pool</sub> the PBMC from both donor groups showed strong antigen-induced proliferation responses (median SI values of 8.2 and 5.6 and percent positive responses of 66 and 71% in TB patients and healthy subjects, respectively). When tested in IFN-γ assays, the complex mycobacterial antigens and RD1<sub>pool</sub> induced strong responses in both groups, whereas MPB70 induced moderate responses in TB patients and strong responses in healthy subjects (Table 2). Furthermore, there were no significant differences (P > 0.05) in the responses evoked by RD1<sub>pool</sub> in TB patients and healthy donors. These results demonstrate that, in addition to complex antigens, the RD1<sub>pool</sub> also has antigens that induce strong Th1 cell responses in both TB patients and M. bovis BCG-vaccinated healthy subjects.

**Antigen-induced proliferation and IFN-γ secretion by PBMC of TB patients and BCG-vaccinated healthy subjects in response to peptide pools of individual ORFs of RD1.
**To identify the major Th1 cell-stimulating antigens encoded by RD1 ORFs, pools of synthetic peptides covering the amino acid sequence of each ORF were tested with PBMC from the above group of TB patients and healthy subjects in antigen-induced proliferation and IFN-γ assays. The results showed that all ORFs induced positive responses in one or both assays in a portion of TB patients and healthy subjects tested; i.e., responses ranged from 16 to 73% and 0 to 70% in antigen-induced proliferation (data not shown) and from 18 to 80% and 17 to 73% in IFN-γ assays (Table 3), respectively. However, strong responses were induced by the peptide pools of PPE68, CFP10, and ESAT6 in TB patients and by the peptide pool of PPE68 in healthy subjects in both assays whereas other ORFs showed weak to moderate responses in both donor groups and in both assays (Table 3; data shown for IFN-γ only). To further analyze the responses of healthy subjects to individual ORFs in relation to being infected and noninfected with M. tuberculosis, the BCG-vaccinated subjects were divided into ESAT6/CFP10 responders (n = 15) and nonresponders (n = 15). The analysis of results showed that only PPE68 induced strong IFN-γ responses in both groups of healthy subjects, with median values of 23 and 13 U/ml and percent positives of 80 and 67% in ESAT6/CFP10 responders and nonresponders, respectively. HLA-DR and -DQ typing of PBMC showed that TB patients and healthy subjects included in the study were highly HLA heterogeneous and covered all the major specificities of HLA-DR and -DQ (data not shown), thus suggesting that the major antigens (PPE68, CFP10, and ESAT6) were presented promiscuously to the responding cells.

### ProPred analysis for promiscuous HLA-DR binding and identification of immunodominant peptides of PPE68 by testing PBMC from TB patients and BCG-vaccinated healthy subjects
The promiscuous nature of ESAT6 and CFP10 for HLA-DR binding and presentation to Th1 cells has been re-
ported previously (39). In this study, to determine the promis-
cuous nature of PPE68 and its peptides, the full-length protein
and the peptide sequences were analyzed for prediction to bind
HLA-DR molecules by using the ProPred server. The analysis
showed that PPE68 was predicted to bind 50/51 (98%) of the
HLA-DR specificities included in ProPred (Table 4). Furthermore,
19/24 of PPE68 peptides were predicted to bind one or
more HLA-DR specificities, with four peptides predicted to be
promiscuous binders, i.e., peptides at residues 61 to 85, 121 to
145, 136 to 160, and 301 to 325 (Table 4). To identify the
peptide(s) of PPE68 inducing antigen-induced proliferation and
IFN-γ secretion, the full-length protein and the peptide se-
quences were analyzed for prediction to bind using the ProPred server revealed that the promiscuous region
of peptide 121 to 145 was confined to the 10-aa sequence
127-FFGINTIPIA-136, which was predicted to bind the same

| Antigen | Median (U/ml [range]) | No. of positive subjects/No. of subjects tested | % Positive |
|---------|-----------------------|-----------------------------------------------|------------|
| Rv3871  | 1.3 (<0.4–47)         | 12/40                                          | 30%        |
| PE35    | 0.4 (<0.4–108)        | 11/40                                          | 28%        |
| ORF4    | 0.5 (<0.4–15)         | 7/40                                          | 18%        |
| PPE68   | 9.0 (<0.4–171)        | 27/40                                          | 68%        |
| CFP10   | 18.3 (<0.4–235)       | 32/40                                          | 80%        |
| ESAT6   | 15.4 (<0.4–361)       | 32/40                                          | 80%        |
| ORF8    | 3.0 (<0.4–41)         | 21/39                                          | 54%        |
| Rv3876  | 2.7 (<0.4–270)        | 19/40                                          | 48%        |
| Rv3877  | 1.1 (<0.4–194)        | 13/39                                          | 33%        |
| Rv3878  | 1.3 (<0.4–10.5)       | 15/37                                          | 40%        |
| ORF14   | <0.4 (<0.4–121)       | 11/37                                          | 30%        |
| ORF15   | <0.4 (<0.4–305)       | 8/37                                           | 22%        |

Table 3. Antigen-induced IFN-γ secretion by PBMC from TB patients and BCG-vaccinated healthy subjects in response to peptide pools of RD1 ORFs

| Antigen | Median (U/ml [range]) | No. of positive subjects/No. of subjects tested | % Positive |
|---------|-----------------------|-----------------------------------------------|------------|
| PPE68   | 3.8 (<0.4–84)         | 16/30                                          | 53%        |
| ORF4    | 1.0 (<0.4–72)         | 10/30                                          | 33%        |
| PPE68   | 0.9 (<0.4–17)         | 11/30                                          | 37%        |
| CFP10   | 22 (<0.4–108)         | 22/30                                          | 73%        |
| ESAT6   | 2.9 (<0.4–107)        | 14/30                                          | 47%        |
| ORF8    | 2.1 (<0.4–114)        | 11/30                                          | 37%        |
| Rv3876  | <0.4 (<0.4–51)        | 10/30                                          | 33%        |
| Rv3877  | 3.0 (<0.4–124)        | 15/30                                          | 50%        |
| ORF4    | <0.4 (<0.4–82)        | 6/30                                           | 20%        |
| Rv3878  | <0.4 (<0.4–13)        | 5/30                                           | 17%        |
| ORF14   | <0.4 (<0.4–23)        | 5/30                                           | 17%        |
| ORF15   | <0.4 (<0.4–102)       | 10/30                                          | 33%        |

a number of subjects.

b Median IFN-γ concentrations representing positive responses (delta IFN-γ of ≥3 U/ml) are given in bold.

| Antigen | HLA-DR binding prediction | Proliferation (n = 30)a |
|---------|-----------------------------|-------------------------|
| Peptide (residues) | No. of positive specificities/No. of specificities tested | % Binding | Median SI | No. of positive subjects/No. of subjects tested | % Positive |
| PPE68 | 50/51 | 98 | 5.6 | 21/30 | 70 |
| PPE68 pool | 1–25 | 2 | 1.1 | 5/30 | 17 |
| | 16–40 | 12 | 9.9 | 8/30 | 27 |
| | 31–55 | 43 | 1.1 | 5/30 | 17 |
| | 46–70 | 0 | 1.4 | 8/30 | 27 |
| | 61–85 | 69 | 1.4 | 8/30 | 27 |
| | 76–100 | 18 | 1.5 | 7/30 | 23 |
| | 91–115 | 0 | 1.1 | 8/30 | 27 |
| | 106–130 | 6 | 0.8 | 7/30 | 23 |
| | 121–145 | 65 | 3.4 | 15/30 | 50 |
| | 136–160 | 75 | 1.2 | 10/30 | 33 |
| | 151–175 | 47 | 1.4 | 11/30 | 37 |
| | 166–190 | 47 | 1.1 | 10/30 | 33 |
| | 181–205 | 45 | 1.7 | 10/30 | 33 |
| | 196–220 | 4 | 2.1 | 9/30 | 30 |
| | 211–235 | 12 | 1.9 | 10/30 | 33 |
| | 226–250 | 45 | 1.3 | 12/30 | 40 |
| | 241–265 | 22 | 1.5 | 9/30 | 30 |
| | 256–280 | 0 | 0.8 | 9/30 | 30 |
| | 271–295 | 31 | 1.3 | 9/30 | 30 |
| | 286–310 | 37 | 1.4 | 8/30 | 27 |
| | 301–325 | 57 | 1.3 | 9/30 | 30 |
| | 316–340 | 0 | 2.8 | 14/30 | 47 |
| | 331–355 | 35 | 1.7 | 9/30 | 30 |
| | 346–371 | 0 | 1.4 | 6/30 | 20 |

a The median positive responses (SI of ≥3) are given in bold. n, number of subjects.

Identification of the promiscuous epitope region in peptide 121 to 145 of PPE68 and its presence in mycobacterial species.

The search for homologies with the sequence of immunodominant peptide 121-VLTATNFFGINTIPIA-136 in the NCBI database showed that it was present in the PPE family proteins of laboratory as well as clinical stains of M. tuberculosis (H37Ra, H37Rv, C, F11, Haarlem, and CDC15151) and M. bovis AF2122/97 (data not shown). In addition, a 14-aa stretch (124-ATNFFGINTIPIA-137) of this peptide was also present in several PPE family proteins present in various mycobacterial species, including M. bovis BCG vac-

TABLE 4. HLA-DR binding prediction analysis of PPE68 and its peptides and their reactivities in antigen-induced proliferation assays with PBMC from BCG-vaccinated healthy subjects

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The table continues with similar entries as per the previous table, listing specific peptides, HLA-DR binding predictions, and proliferation responses in vivo. Each entry follows the same format, providing data on the specific peptide, its predicted binding properties, and the percentage of positive responses observed in PBMC from BCG-vaccinated healthy subjects.
number of HLA-DR alleles (33/51, or 65%) as the peptide at position 121 to 145 (Table 4). To study the recognition by Th1 cells, the peptides at positions 124 to 137 and 127 to 136 were also synthesized and tested along with the peptide at position 121 to 145 with PBMC from eight HLA-heterogeneous BCG-vaccinated healthy subjects that responded to M. tuberculosis, M. bovis BCG, a peptide pool of PPE68, and the peptide at position 121 to 145 in IFN-γ assays (Table 5). The results showed that all of the eight donors responded to peptides at positions 124 to 137 and 127 to 136, whereas two other peptides of PPE68 from the C terminus region, i.e., the peptide at positions 331 to 355 and 356 to 371, stimulated PBMC for IFN-γ secretion from only two of the six tested donors (Table 5). These results confirmed the finding that the immunodominant and promiscuous Th1 cell epitope of PPE68 was present in M. bovis BCG vaccines and was limited to the 10-aa sequence 127-FFGINTIPIA-136.

**DISCUSSION**

We have evaluated Th1 cell reactivity of 12 ORFs of the RD1 segment of M. tuberculosis by using PBMC from TB patients and M. bovis BCG-vaccinated healthy subjects. To our knowledge, this is the first study to demonstrate the feasibility of an approach using a large peptide pool for Th1 cell reactivity and the most comprehensive analysis of RD1-encoded proteins for Th1 cell reactivity in TB patients and BCG-vaccinated healthy humans. The test systems used to determine Th1 cell reactivity were antigen-induced proliferation and IFN-γ secretion assays (17, 25). IFN-γ is considered the most important Th1 cytokine involved in protective immunity against TB (2, 13, 15); therefore, in order to identify new candidates for safer subunit/peptide vaccines, it is important to identify major M. tuberculosis antigens and immunodominant peptides recognized by IFN-γ-secreting T cells. In addition, M. tuberculosis-specific antigens inducing IFN-γ production could also be useful in the specific diagnosis of TB (reviewed in reference 26).

To overcome the problems associated with recombinant protein expression and purification, as reported previously (1, 8, 11), we have used in this study pools of 220 synthetic peptides covering RD1 ORFs of various sizes (Table 1). One of the obvious advantages of this approach is the speed with which peptides can be synthesized and tested for Th1 cell reactivity. Furthermore, major antigens and immunodominant peptides could be identified, and exact T-cell epitopes could be defined by subsequent testing of peptide pools corresponding to individual ORFs, followed by single peptides included in the pools (5, 40, 41). Each synthetic peptide was 25 aa in length and overlapped with the neighboring peptides by 10 residues. The reason for the 10-residue overlap was to greatly reduce the probability of missing T-cell epitopes, which are usually ≥10 aa in length (24). Furthermore, the inclusion of peptides of CFP10 and ESAT6, the two major and widely studied antigens recognized by TB patients (9, 10, 26, 27, 38), in the RD1 pool acted as a quality control to ensure the reliability of a large peptide pool approach for determining Th1 cell reactivity.

The results of testing PBMC from TB patients for Th1 cell reactivity with the RD1 pool were highly encouraging because a large proportion of the tested patients responded to this pool (Table 2), and the responses to the RD1 pool were of magnitudes similar to peptide pools of individual ORFs like ESAT6 and CFP10, as reported previously (10, 26, 27, 38) and also found in this study (Table 3). These results suggest that large peptide pools corresponding to ORFs of various sizes could faithfully be tested for T-cell reactivity in proliferation and IFN-γ assays without having a significant inhibitory activity on the responses to individual ORFs constituting the pool. Thus, our results set the stage to use pools of synthetic peptides corresponding to other RDs of M. tuberculosis, i.e., RD4 to RD7, RD9 to RD13, and RD15, which are absent in M. bovis and M. bovis BCG and are predicted to encode antigens of diagnostic and vaccine potential (16, 43).

We further evaluated the RD1 pool for the specificity of response by testing PBMC from M. bovis BCG-vaccinated healthy subjects. Since we had included in this pool the peptides corresponding to the ORFs of RD1 that are deleted in M. bovis BCG, we had expected significantly lower responses to RD1 pool in BCG-vaccinated healthy subjects. However, contrary to our expectation, the RD1 pool induced strong cell proliferation and IFN-γ responses with PBMC from healthy subjects.

**TABLE 5. Antigen-induced IFN-γ secretion by PBMC from HLA-DR heterogeneous M. bovis BCG-vaccinated healthy subjects in response to mycobacterial antigens and peptides of PPE68**

| Organism, antigen, or peptide (residues) | Peptide sequence | Antigen-induced IFN-γ (U/ml) secretion by PBMC from the indicated donor* | No. of positive subjects/total no. of subjects tested |
|-----------------------------------------|-----------------|---------------------------|----------------------------------|
| M. bovis BCG                            |                 |                           |                                  |
| M. tuberculosis                         |                 |                           |                                  |
| PPE68pool                               |                 |                           |                                  |
| 121–145                                 | VLATNFFGINTIPIALT EMDYFIR | 43 | 44 | 27 | 18 | 40 | 43 | 15 | 30 | 8/8 |
| 124–137                                 | ATNFFGINTIPIAL  | 43 | 10 | 23 | 10 | 26 | 21 | 21 | 10 | 8/8 |
| 127–136                                 | FFGINTIPIA      | 50 | 5.0 | 19 | 5.0 | 28 | 17 | 12 | 8.0 | 8/8 |
| 331–355                                 | GAMGQGAQSGGSTRP | 5.6 | 0.0 | 2.5 | 0.6 | 7.3 | 2.0 | ND | ND | 2/6 |
| 346–371                                 | GLVAPQPLAQEEEDD EDDWDEEDDW | 0.5 | 1.0 | 0.5 | 7.0 | 23 | 2.0 | ND | ND | 2/6 |

* The following were the HLA-DR and HLA-DQ types of the healthy donors included in this study: donor 1, DR7, DR17, DR52, DR53, DQ2, and DQ6; donor 2, HLA types not determined; donor 3, DR11, DR13, DR52, and DQ7; donor 4, DR17, DR52, and DQ2; donor 5, DR1, DR18, DR52, DQ4, and DQ5; donor 6, DR14, DR15, DR51, DR52, DQ5, and DQ6; donor 7, DR11, DR13, DR52, and DQ7; donor 8, DR4, DR17, DR52, DR53, DQ2, and DQ8. IFN-γ concentrations representing positive responses (delta IFN-γ values of ≥5 U/ml with complex antigens and ≥3 U/ml with peptides) are given in bold. ND, not determined.
TB patients and healthy subjects existed at the peptide level. The major recognition of PPE68 by T cells was first reported by us in a cattle model of TB (32) and later extended by others in mice (12) and humans (22, 45). Our studies in cattle (32) as well as the studies of Demangel et al. in mouse (12) show that PPE68 was recognized by animals infected with M. bovis and M. tuberculosis, respectively, but not by animals vaccinated with M. bovis BCG. However, in humans, our present study (using Asian donors resident in Kuwait) and two previous studies (using Caucasian donors residing in the United Kingdom [22] and Denmark [45]) show that both TB patients and M. bovis BCG-vaccinated healthy humans respond to PPE68 in Th1 cell assays. Furthermore, the nonresponsiveness to PPE68 of healthy Danish subjects not vaccinated with BCG (45) suggests that the response in healthy BCG-vaccinated subjects was due to BCG vaccination and not due to exposure to environmental mycobacteria. However, differences exist in the magnitude of response; i.e., in our study 64% of ESAT6/CFP10 nonresponders responded to PPE68, whereas of tested BCG-vaccinated donors 7.9% in the United Kingdom (22) and 17.5% in Denmark (45) responded to PPE68.

It is well known that T cells recognize antigen in association with highly polymorphic HLA molecules. Therefore, among the requirements for new antigens to qualify as diagnostic reagents and/or vaccine candidates is their recognition by an HLA-heterogeneous group of donors (24, 35). The results of HLA-DR and -DQ typing showed that the TB patients and healthy donors tested in this study were HLA heterogeneous and represented all of the frequently expressed specificities of HLA-DR and -DQ molecules. Furthermore, based on the ProPred server analysis for HLA-DR binding prediction, our previous work has suggested that the antigens CFP10 and ESAT6 are promiscuous HLA-DR binders (39). The analysis of the PPE68 sequence for HLA-DR binding prediction in this study suggests that this antigen is also a promiscuous HLA-DR binder. Thus, our overall results suggest that HLA restriction will not be a problem if the above RD1 antigens are used in diagnostic applications and/or as vaccine candidates against TB.

After identifying PPE68 as the major antigen in both TB patients and BCG-vaccinated healthy subjects, our next aim was to identify the immunodominant peptide(s) of this protein recognized by Th1 cells and determine if differences between TB patients and healthy subjects existed at the peptide level. We therefore tested the individual peptides of ORF5 (PPE68) with PBMC from healthy donors and TB patients in the Th1 cell assays. The results revealed that a single peptide, peptide VLTATNFGINTIPIALTEMDYFIR, qualified to be immunodominant in both donor groups. The HLA-DR heterogeneity of the responding subjects and the prediction to bind 65% of the HLA-DR specificities included in ProPred strongly suggest that this peptide was presented to Th1 cells promiscuously. The previous studies have identified the immunodominant sequences of PPE68 as VLTATNFGINTIPIALTE in humans (45) and ATNFGINTIPIALTE in mice (12), and these sequences have been shown to have high sequence homology with other PPE proteins present in M. bovis BCG and other mycobacteria (12, 45). In this study, we identified a shorter 14-aa sequence of PPE68, i.e., ATNFGINTIPIALTE, to be completely identical in PPE family proteins present in M. tuberculosis and M. bovis BCG. Furthermore, ProPred analysis as well as testing with PBMC from BCG-vaccinated healthy subjects narrowed the promiscuous and immunodominant epitope region to the 10-aa sequence FFGINTIPIALTE.

In mice, Demangel et al. have shown that the peptide ATN FFINTIPIALTE was recognized by T cells of mice infected with M. tuberculosis H37Rv and BCG::RD1 (BCG complemented with RD1) but not by BCG-vaccinated animals or mice injected with a PPE68 knockout construct (12), suggesting the specificity of the peptide for M. tuberculosis. However, their results are contrary to our results and the results of Okkels et al. (45) in humans, showing that the peptides containing the sequence ATNFGINTIPIALTE are recognized by TB patients and BCG-vaccinated healthy subjects. The results in humans are in line with the presence of the peptide sequence in PPE68 and several other PPE proteins, which are present in both BCG and M. tuberculosis. Demangel et al. have hypothesized that the nonrecognition of the peptide sequence ATN FFINTIPIALTE in BCG-vaccinated mice, which is shared by many other PPE family proteins, may be due to the less appropriate presentation of other PPE proteins in mice (12).

Unlike the two other major Th1 cell-stimulating antigens encoded by RD1 genes (i.e., ESAT6 and CFP10), PPE68 is not related to the pathogenesis associated with RD1, as shown in SCID mice (12). M. tuberculosis specificity and association with pathogenesis suggest that the antigens ESAT6 and CFP10 should be reserved for diagnostic applications to detect latent and active TB, as reported previously (9, 10, 38, 42). However, the strong Th1 cell-stimulating ability of PPE68 in both TB patients and BCG-vaccinated healthy subjects and the nonassociation with pathogenesis suggest that PPE68 and its promiscuous immunodominant peptide sequences may be valuable as subunit and peptide-based vaccine candidates, respectively. The presence of PPE68-reactive IFN-γ-secreting T cells in healthy elderly Norwegians, who had converted to skin test PPD positivity in childhood due to natural infection but did not develop clinical disease or receive any treatment (45), further supports the usefulness of this antigen as a new vaccine candidate against TB.

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